Short stature with low insulin-like growth factor 1 availability due to pregnancy-associated plasma protein A2 deficiency in a Saudi family

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Abstract
In 2016 a new syndrome with postnatal short stature and low IGF1 bioavailability caused by biallelic loss-of-function mutations in the gene encoding the metalloproteinase pregnancy-associated plasma protein A2 (PAPP-A2) was described in two families. Here we report two siblings of a third family from Saudi Arabia with postnatal growth retardation and decreased IGF1 availability due to a new homozygous nonsense mutation (p.Glu886* in exon 7) in PAPPA2. The two affected males showed progressively severe short stature starting around 8 years of age, moderate microcephaly, decreased bone mineral density, and high circulating levels of total IGF1, IGFBP3, and the IGF acid-labile subunit (IGFALS), with decreased free IGF1 concentrations. Interestingly, circulating IGF2 and IGFBP5 were not increased. An increase in growth velocity and height was seen in the prepuberal patient in response to rhIGF1. These patients contribute to the confirmation of the clinical picture associated with PAPP-A2 deficiency and that the PAPPA2 gene should be studied in all...
patients with short stature with this characteristic phenotype. Hence, pediatric endocrinologists should measure circulating PAPP-A2 levels in the study of short stature as very low or undetectable levels of this protein can help to focus the diagnosis and treatment.

**KEYWORDS**
free, IGF1, bone mineral density, IGF1, IGFALS, PAPP-A2, PAPP-A2, rhIGF1, short stature

1 | **INTRODUCTION**

Short stature can be due to simple or complex etiology affecting hormonal status and/or the growth plate. Over the last decade, we have learned much regarding monogenic causes of short stature due to affection of the growth hormone (GH)/insulin-like growth factor (IGF) axis, which controls linear growth. Understanding the physiology, genetics, and etiology of growth defects is important for implementing correct treatment. Regulation of the growth plate is influenced by genes such as GH1, GHR, IGF1, IGF2, and IGFLS. In 2016, biallelic loss-of-function mutations in the PAPP-A2 were reported to cause a new syndrome with postnatal short stature due to low IGF1 bioavailability.

GH stimulates IGF, IGFBP, and IGFLS production by the liver, with IGF1 being the main factor stimulating linear growth. IGF1 is also produced in most other tissues and exerts a paracrine effect. Approximately 95% of IGF1 circulates in ternary complexes (TC) that consist of IGF1, IGFBP3 or IGFBP5 and ALS. As free IGF1 (fIGF1) is the active molecule that promotes linear growth, it must be released from these complexes. To liberate IGF1 from the ternary complex PAPP-A2 cleaves IGFBP3 and IGFBP5.

Mutations in the PAPPA2 gene have been reported in five members of two unrelated families with short stature and high total IGF1, but low fIGF1 levels. Here we report two siblings of a family from Saudi Arabia carrying a new loss-of-function mutation in homozygosis in the PAPPA2 gene associated with postnatal short stature. To our knowledge, these represent two of the seven cases reported to date worldwide.

2 | **MATERIALS AND METHODS**

ELISA was used to determine serum levels of ALS and insulin (BioVendor), IGF1, free IGF1, IGF2, total IGFBP4, and IGFBP5, intact IGFBP3 and IGFBP4, PAPP-A and PAPP-A2 (Ansh Labs, Webster). GH and total IGFBP3 were measured by chemiluminiscence immunosays (DiaSorin). All assays were performed according to the manufacturers’ instructions and all intra- and inter-assay coefficients of variation were lower than 10%.

Whole-genome sequencing was performed in blood DNA from the index case by CENTOGENE. DNA sequencing and validations were performed by CAP/CLIA, commercially accredited laboratories. The average coverage depth was ~30X. A complete illustration of the pipeline, filtration processes, and prioritization steps have been previously published [https://pubmed.ncbi.nlm.nih.gov/30202406/](https://pubmed.ncbi.nlm.nih.gov/30202406/). The variant plus 150 bp on either side of each variant in the genomic region were amplified using specific primers PAPPA2 c.2656 FWD, ACTCTCTCTACCTCCCCATCTC. PAPPA2 c.2656 REV CAGTGTCACCCAGGTAATAC, and bidirectionally sequenced using Sanger sequencing.

2.1 | **Ethical statement**

Written consent for these studies was obtained from all adults and parents of the index patient.

The laboratory procedures employed were reviewed and approved by the institutional review committee of the Hospital Infantil Universitario Niño Jesús (C.I: R-0017/19) and conform to the Helsinki Declaration.

3 | **RESULTS**

The index case, a Saudi boy presented to our endocrine clinic at the age of 14 years complaining of short stature. Poor growth was noticed around 8 years of age and it became progressively more apparent over time (Figure 1A). His delivery was at term (39 weeks) and uneventful. His parents are first cousins with a mid-parental height of 172 cm, which is above average in the Saudi society. Among his three surviving siblings, a 22-year-old brother had short stature and similar features (Figure 1B), the others are of normal height.

He and his elder brother, with the same genetic condition, had microcephaly, triangular face, small chin, and anterior pointed ears (Table 1), with mild learning difficulties. The index’s height was 138.8 cm (~3.3 SDS). Growth velocity (GV) initially was 1.2 cm/year and no abnormalities in the sequence of chromosome 11. A genome sequence revealed a homozygous loss-of-function variant in the PAPPA2 gene on chromosome 1 (GRCh37):g.176664905G > T; NM_020318.2:c.2656G > T; p.Glu886* (Figure 2A). This variant, presumably resulting in a null allele undergoing nonsense-mediated mRNA decay and coding for a truncated 86/1791 amino acid protein, has not been reported previously in gnomAD, ESP, or 1000 Genomes. This novel homozygous mutation in PAPPA2 was highlighted as likely pathogenic using ACMG (American College of Medical Genetics) criteria, given its loss-of-function nature and the
clinical presentation in these patients similar to those previously reported with PAPPA2 mutations.\textsuperscript{3} Family segregation analysis revealed that the older brother with similar clinical features (Figure 2B, Table 1) is also homozygous for the mutation, while the parents and the two other living siblings are heterozygous carriers (Figure 2B). Subsequent biochemical analysis showed significantly increased total IGF1, total IGFBP3, intact IGFBP3, and IGFALS levels in both affected siblings. PAPP-A2 levels were undetectable, while PAPP-A levels were in the normal range (Table 1). Normal values have been previously reported.\textsuperscript{3} Before the genetic diagnosis, the patient received GH (0.035 mg/kg/day) and testosterone (50 mg IM monthly for 3 months due to pubertal delay). After testosterone treatment, he attained Tanner stage II. His GV was 6 cm after 1 year in response to rhGH and the advancement of puberty. He started treatment with rhIGF-1 (40 $\mu$g/kg twice daily gradually increasing to 80 $\mu$g/kg twice daily based on response) and after 7 months, he gained 4.9 cm of height (8.4 cm/year).

4 | DISCUSSION

To date short stature related to PAPPA2 gene defects has been reported in three different families, including that reported here. The first report was of a 9-year-old girl and a 5-year-old boy from a Spanish family of non-consanguineous parents that presented with growth retardation compared with their target heights.\textsuperscript{3} In the other family of consanguineous parents (cousins) from Palestine an adolescent girl with two younger brothers had growth retardation that warranted medical evaluation at different ages.\textsuperscript{3} The cases reported here share clinical and laboratory data with those previously reported. Furthermore, the observed likely pathogenic variant in the PAPPA2 gene (p.Glu886*), presumably a null allele undergoing nonsense-mediated mRNA decay, is consistent with the clinical observations and the diagnosis of PAPP-A2 deficiency, as well as with undetectable levels of PAPP-A2 in serum.\textsuperscript{3}

Postnatal short stature is the main complaint for which these patients are clinically evaluated. To date, growth is described to be appropriate prenatally and during infancy, but with a progressive decline after ~6 years of age. Indeed, late detection of gene defects in this entity leads to more severe short stature as found here and in the previously reported Palestinian girl.\textsuperscript{3} Three of the reported cases were born SGA, with one achieving good catch-up during the first 2 years of life. In all cases, a typical phenotype included small chin, long thin fingers, and modest microcephaly, Anterior rotated ears (33.3%) and delayed dental eruption (50%) have also been observed.\textsuperscript{3} Four of the seven reported patients are postpubertal.\textsuperscript{2} Two girls had spontaneous onset of puberty at around 11 years of age with normal progression, while two males, including our index patient, had
| Clinical characteristics                                      | Index case | Affected brother |
|---------------------------------------------------------------|------------|-----------------|
| **Growth data**                                              |            |                 |
| Sex               | M          | M               |
| Age at examination (years)                                   | 14         | 22              |
| Gestational age (weeks)                                     | 39         | 36              |
| Birth weight (kg)                                          | N/A (SGA) | N/A (SGA)       |
| Height (cm)                                                | 138.8 (−3.3) | 153.7 (−3.6) |
| BMI                                     | 14.68      | 17.84           |
| Head circumference (SDS)                                    | −3.4       | N/A             |
| **Clinical features**                                    |            |                 |
| Small chin                                               | Yes        | Yes             |
| Thin long fingers                                      | Yes        | Yes             |
| Anterior rotated ears                                      | Yes        | No              |
| Delayed dental eruption                                   | No         | Yes             |
| Onset of puberty (years)                                  | 14.5       | 15              |
| Growth spurt                                          | Still growing | N/A             |
| **Radiological data**                                    |            |                 |
| BMD                                                | Low        | Not done        |
| Skeletal survey                                        | Normal     | Normal          |
| **Genetic results**                              |            |                 |
| PAPP-A2 gene variant                                      | (c.2656G > T, p.Glu886*) | (c.2656G > T, p.Glu886*) |
| **Therapeutic data**                               |            |                 |
| Treatment                                        | GH and rhIGF1 | N/A             |
| Complications                                | None        | N/A             |
| **Biochemical parameters**                               |            |                 |
| Subjects                                      | IGF1 (µg/L) | rIGF1 (µg/L) | IGF2 (µg/L) | Total IGFBP3 (µg/L) | Intact IGFBP3 (µg/L) | Total IGFBP4 (µg/L) | Intact IGFBP4 (µg/L) | IGFBP5 (µg/L) | ALS (U/L) | Insulin (mU/L) | PAPP-A (µg/L) | PAPP-A2 (µg/L) | GH (CLIA) (µg/L) |
| Index                                        | 3193        | 2.37            | 95        | 8510       | 4347       | 37.52       | 7.19       | 466        | 4422       | 13.44       | 0.48          | Undetectable          | 0.53      |
| Affected brother                                 | 1751        | 0.40            | 124       | 5620       | 2535       | 51.94       | 6.91       | 409        | 3266       | 3.00        | 0.72          | Undetectable          | 0.16      |
| Father                                        | 342         | 0.21            | 135       | 3420       | 175        | 165.80      | 2.14       | 205        | 1925       | 1.30         | 0.95          | 0.04          | 0.13      |
| Mother                                        | 778         | 3.20            | 189       | 4680       | 691        | 68.24       | 4.43       | 629        | 2673       | 2.55         | 1.27          | 0.02          | 0.01      |
| Healthy brother                                | 260         | 5.13            | 393       | 4010       | 216        | 96.67       | 6.94       | 446        | 2386       | 13.45        | 1.41          | 0.04          | 0.10      |
| Healthy sister                                | 301         | 0.86            | 346       | 5020       | 735        | 159.18      | 10.51      | 367        | 1257       | 4.97         | 0.67          | 0.02          | 0.38      |

Note: Biochemical parameters related to the family members: Index, affected brother, father, mother, and two healthy siblings.

Abbreviations: BMD, body mass index; GH, growth hormone; PAPP-A2, pregnancy-associated plasma protein A2; rhIGF-1, recombinant human insulin-like growth factor-1; SGA, small for gestational age.
delayed puberty. In two cases where pubertal onset and progression were normal, a pubertal growth spurt was not observed. The Spanish female patient began puberty while receiving rhIGF1 and was immediately treated with GnRH to stop pubertal development. Thus, it is possible that pubertal delay is more common in males than in female PAPP-A2 deficient patients. The question regarding male fertility might be raised later as fertility is affected in PAPP-A2 deficient male mice but not females.

All cases reported to date have high total IGF1, IGFBP3, IGFALS, and stimulated GH levels, very low free IGF1 and very low or undetectable levels of PAPP-A2; however, the cases reported here did not have elevated IGF2 or IGFBP5. Fasting hyperinsulinism and impaired fasting glucose is reported in some patients. Our index case had a modest basal hyperinsulinemia, but his affected brother did not.

Skeletal surveys demonstrated thin long bones in all previous cases, which was not observed in our patient. Densitometric scans in the two Spanish cases were in the osteoporotic range of decreased lumbar density (Z-score –2 to –2.3 SD). One Palestinian subject had a significantly low BMD (–2.5). Our patient had a severely decreased lumbar density (Z-score –3.4).

The Spanish patients had a good response to rhIGF1 injections improving linear growth, BMD, and glucose metabolism, with no hypoglycaemia reported. The two patients from Palestinian origin were subsequently treated with rhIGF1; however, to date the youngest patient has been reported to experience a modest growth response with rhIGF1. Treatment was discontinued in the second brother due to development of pseudotumor cerebri. In the index case reported here, GH in combination with testosterone treatment was used before the genetic diagnosis. As expected testosterone improved growth; however, after 7 months of rhIGF1 treatment, growth velocity improved more dramatically (8.4 cm/year). Long-term outcomes will need further follow-up.

Advances in our understanding of short stature emphasize the importance of performing genetic studies, as well as taking into consideration the genetic potential of growth, or the patient’s target height in the evaluation of short stature. Identification of genetic causes of short stature provides guidance in treatment selection, helps in predicting prognosis and in indicating possible associated conditions that might need treatment, and supports genetic counseling. Indications of genetic studies in cases of short stature should include

FIGURE 2  PAPPA2 gene and family pedigree. (A) Schematic representation of the genomic structure of the PAPPA2 gene on chromosome band 1q24, the encoded mRNA, and the protein with its relevant functional domains. The red asterisk shows the position of the variant identified in the present study. (B) Pedigree and representative Sanger sequencing trace of the mutation identified in an affected individual (top), a heterozygous carrier (middle), and a control (bottom) [Colour figure can be viewed at wileyonlinelibrary.com]
not only severe familial forms of isolated GH deficiency or specific syndromic forms of multiple pituitary hormone deficiencies, severe short stature (>−3SDS for the population or >3 SD lower than MPH), but also patients that are significantly far from their predicted target height.\textsuperscript{2,9} Identification of the genetic bases in patients with idiopathic short stature should provide a more precise approach to treatment, as demonstrated with the discovery of PAPP-A2 deficiency.\textsuperscript{31}

In conclusion, it is probable that PAPP-A2 deficiency is under-diagnosed as some patients may present normal height, or the physician does not correctly interpret the high IGF1 levels or the decline in growth velocity that these patients present during childhood. Thus, the PAPP-A2 gene, as well as circulating levels of PAPP-A2, should be studied in all patients with short stature with these characteristics.

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CONFLICT OF INTEREST
The authors declare no conflicts of interest.

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DATA AVAILABILITY STATEMENT
Data available on request due to privacy/ethical restrictions.

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REFERENCES
1. Argente J, Chowen JA, Pérez-Jurado LA, Frystyk J, Oxvig C. One level up: abnormal proteolytic regulation of IGF activity plays a role in human pathophysiology. EMBO Mol Med. 2017;9(10):1338-1345.
2. Argente J, Pérez-Jurado LA. Genetic causes of proportionate short stature. Best Pract Res Clin Endocrinol Metab. 2018;32(4):499-522.
3. Dauber A, Muñoz-Calvo MT, Barrios V, et al. Mutations in pregnancy-associated plasma protein A2 cause short stature due to low IGF-I availability. EMBO Mol Med. 2016;8(4):363-374.
4. Yakar S, Werner H, Rosen CJ. Insulin-like growth factors: actions on the skeleton. J Mol Endocrinol. 2018;61(1):T115-T137.
5. Muñoz-Calvo MT, Barrios V, Pozo J, et al. Treatment with recombinant human insulin-like growth factor-1 improves growth in patients with PAPP-A2 deficiency. J Clin Endocrinol Metab. 2016;101(11):3879-3883.
6. Christians JK, King AY, Rogowska MD, Hessels SM. Pappa2 deletion in mice affects male but not female fertility. Reprod Biol Endocrinol. 2015;13:109.
7. Hawkins-Carranza FG, Muñoz-Calvo MT, Martos-Moreno GÁ, et al. rhIGF-1 treatment increases bone mineral density and trabecular bone structure in children with PAPP-A2 deficiency. Horm Res Paediatr. 2018;89(3):200-204.
8. Cabrera-Salcedo C, Mizuno T, Tzyzinski L, et al. Pharmacokinetics of IGF-1 in PAPP-A2-deficient patients, growth response, and effects on glucose and bone density. J Clin Endocrinol Metab. 2017;102(12):4568-4577.
9. Lango Allen H, Estrada K, Lettre G, et al. Hundreds of variants clustered in genomic loci and biological pathways affect human height. Nature. 2010;467(7317):832-838.
10. Collett-Solberg PF, Ambler G, Backeljauw PF, et al. Diagnosis, genetics, and therapy of short stature in children: a growth hormone research society international perspective. Horm Res Paediatr. 2019;92(1):1-14.
11. Argente J, Pérez-Jurado LA. History and clinical implications of PAPP-A2 in human growth: when reflexions on idiopathic short stature lead to a specific and new diagnosis. Growth Horm IGF Res. 2018;40:17-19.

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