Prospects of using nanotechnology for food preservation, safety, and security

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A R T I C L E   I N F O

Article history:
Received 14 January 2018
Received in revised form 6 June 2018
Accepted 11 June 2018
Available online 3 July 2018

Keywords:
Nanostructured materials (NSMs)
Food safety
Food preservation
Functional food
Packaging
Human health

A B S T R A C T

The rapid development of nanotechnology has transformed many domains of food science, especially those that involve the processing, packaging, storage, transportation, functionality, and other safety aspects of food. A wide range of nanostructured materials (NSMs), from inorganic metal, metal oxides, and their nanocomposites to nano-organic materials with bioactive agents, has been applied to the food industry. Despite the huge benefits nanotechnology has to offer, there are emerging concerns regarding the use of nanotechnology, as the accumulation of NSMs in human bodies and in the environment can cause several health and safety hazards. Therefore, safety and health concerns as well as regulatory policies must be considered while manufacturing, processing, intelligently and actively packaging, and consuming nano-processed food products. This review aims to provide a basic understanding regarding the applications of nanotechnology in the food packaging and processing industries and to identify the future prospects and potential risks associated with the use of NSMs.

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1. Introduction

Nanotechnology integrates several disciplines, including physics, chemistry, biotechnology, and engineering, and refers to the use of nanomaterials whose nanoscale structures range from 1 to 100 nm [3]. In these nano-sizes, materials adopt unique properties that were not present when the materials were in their original form. Understanding these unique properties in order to develop new and improved products through green processes is the goal of nanoscientists across the globe [2].

Nanomaterials or nanostructure materials exhibit different dimensions for their structural elements, clusters, molecules, or crystallites, such as zero dimensions (nanoparticles, nanoclusters, and quantum dots), one dimension (nanorods and nanotubes), two dimensions (nano-thin films), and three dimensions (nanomaterials) in the 1–100 nm range [3,4]. Combining nanostructure materials with other polymer, biomolecule, and other nanostructure material or existing in the aggregate form can result relatively in a larger particle size material (>100 nm) leading to the formation of nanocomposite [5]. These nanomaterials having high surface volume ratio exhibit inimitable physio-chemical characteristics, such as solubility, toxicity, strength, magnetism, diffusivity, optics, color, and thermodynamics [6,7].

The applications of these nanomaterials are growing in various sectors, including agriculture, medicine, clothing, cosmetics, food, and public health due to their unique ability to increase solubility and bioavailability and to protect bioactive components while they are being processed and stored [4,8,9]. Due to the excellent physiochemical nature and the antimicrobial potential of nanomaterials, they are widely used against various pathogenic microbes and in healthcare, crop protection, water treatment, food safety, and food preservation [8,10]. In addition, nanostructured materials (NSMs) are being applied in the food industry as a nanosensor, new packaging material, and encapsulated food component. In general, this review concerns the use of nanotechnology, specifically nanoparticles and constructed nanostructures, in the food sector.

Nanotechnology has a wide range of food-related applications. In these applications, a specific type of nanomaterial is incorporated into a specific food product in order for that food product to develop certain desired properties [2]. The field of nanotechnology has also been an integral part of research and development for the large-scale manufacturing of agricultural products and processed foods and drinks, as well as for food packaging across the world [3]. Several reports confirm that these nanomaterials can successfully improve food safety by enhancing the efficacy of food packaging, shelf-life, and nutritional value as additives without changing the taste and physical characteristics of food products [13,14]. Although they have much potential to frame innovative products and production processes in the food industry, nanotechniques are facing a major challenge in using cost-effective processing operations to create edible and non-toxic nano-delivery systems and to develop effective formulations that are safe for human consumption. Therefore, due to the increased use of these NSMs, there have been growing concerns regarding the development of biocompatible, safe, and non-toxic nanostructures from food-grade ingredients using simple, green, and cost-effective strategies, including the layer-by-layer technique [15].

United states-based safety evaluation agencies such as FDA and EPA, including the Directorate of European Health and Consumer Protection, and many other regulatory authorities, have released several guidelines on potential human health risks associated with the use of nanostructure materials in food [16]. These potential risks are determined by nanoscientists using nanotoxicology, which is a branch of toxicology and an interdisciplinary field that concerns varied toxicity aspects of nanomaterials [15]. Nanotoxicity mediated via generation of reactive oxygen species (ROS) is considered a crucial mechanism leading to successive human oxidative stress [8]. Also, there is a huge interest in using nanomaterials in various fields, including their safety concerns. Consequently, there is a compelling demand to address these issues so as to expand our knowledge on the use of nanomaterials in terms of their biocompatibility, safety, and toxicity in the food sector.

Although nanotechnology has applied green-synthesized NSMs in several ways in the food sector, on a few occasions, the use of NSMs has been controversial, as NSMs are scientifically uncertain and can impart long-term detrimental effects on human health and the environment [17]. In this regard, the complexity and limitations in the field of nanotechnology in terms of toxicity and accumulation (in appropriate doses) can be overcome only by elucidating the physiochemical and biological properties of NSMs through extensive research [18]. In light of the effects of nanomaterials on the food industry, this review has been formulated as a “snapshot” to address the current status of the techniques and implementations of nanotechnology in food preservation, safety, and security criteria and to identify the underlying mechanistic actions regarding safety or packaging issues. In addition, the review focuses on the current and future prospects of nanotechnology and discusses the strengths and weaknesses (i.e. the risks related to toxicological effects) of existing regulatory authorities. Moreover, the review discusses the current efforts to address these weaknesses and other issues related to the development, understanding, and promotion of nanotechnology. Fig. 1 illustrates an overview of the important usages of nanoparticles/nanomaterials and nanostructures in the food sector.

2. Natural and synthetic nanostructures in the food system

The food system naturally contains a variety of nano-sized elements that include self-assembled higher order structures, such as carbohydrates and fats [19]. These ingredients are different from synthetically manufactured nanomaterials/ nanostructures thus can be used to make nanoemulsion, nano-encapsulate, and food-grade polymer [19]. During food processing, manufacturing, or thermal treatments, nanostructures are used to produce, which are not associated with modern nanotechnology (e.g. coagulation, emulsifying, or homogenizing). In brief, food proteins, including polysaccharides and lipids, are globular particles whose size can vary between 10 and several hundred nanometers. Coagulation and emulsification are based on the formation of reticular, zero, one,
two and three-dimensional nanostructures. When corn starch is boiled to make custard-like food, small, three-dimensional crystalline structures that are only tens of nanometers in thickness are melted [20]. Fresh milk and milk-based products naturally contain nanostructures, such as milk proteins and casein. When milk is homogenized, fat globules that are about 100 nm in size are produced [20].

Nanotechniques play an essential role in the functional and nutraceutical food sectors. Encapsulation of effective molecules or components in NSMs can enhance the bioavailability and solubility of coloring agents and nutritional ingredients such as minerals and vitamins, thus facilitating controlled release and protecting biologically active substances and micro-nutrients during their processing. Currently, the most usable NSMs are nanocapsules (e.g. micelles and liposomes) and nanoemulsions, including a few carbon-based, green-synthesized, ecofriendly nanomaterials.

3. Nanotechnology in food packaging and security

The packaging of food is one of the most critical steps in terms of food safety. Natural substances, atmospheric gases and water vapors are absolutely impermeable to no packing materials [21,22]. However, completing blocking the migration and permeability of gases is not desirable in the case of packaging fresh fruits and vegetables that undergo cellular respiration [21]. In contrast, the packaging of carbonated beverages should eliminate the flow of oxygen and carbon dioxide (CO₂) to prevent oxidation and de-carbonation [21]. The flow of CO₂, oxygen, and water vapors varies according to the food matrices and the packaging materials that are used. Hence, these complexities in food packaging can be addressed and overcome by employing various nanocomposite materials, including polymers [24]. Nanoparticle having a diameter of less than 100 nm is thousand-times thinner than about 100,000 nm thick book page or hundred-times thinner than about 10,000 nm thick human hair. With an appropriate regulatory processing approval, different structures of nanoparticles can be of significant use in various fields of food and pharmaceutical industries, especially for the research programs associated with food science-based research (Fig. 2).

Recently, the use of nano-biocomposites in food packaging has enhanced the ability of food packaging to act as a barrier against gases [25]. The applications of various nanoparticles in food packaging are briefly shown in Table 1. Recent trends in food packaging are encouraging the use of biodegradable polymers reinforced with nanofillers, which are eco-friendly [26]. However, there is a major concern regarding the

![Fig. 1 – Systematic representation of application of nanoparticles in various areas of food industry.](image)

![Fig. 2 – Steps for processing and utilizing of nanomaterials in food sectors.](image)
ingestion of these nano-compounds during consumption of the food. Hence, it is essential to investigate how these nanoparticles migrate within the human body as well as these nanoparticles’ toxic and immunogenic effects [27]. Another concern is related to the biodegradability of these nanofilled, biodegradable polymers [28]. These concerns are seriously considered by researchers across the world who are searching for human and environmentally friendly nanomaterials [29].

3.1. Nanocoatings as intelligent packaging for surfaces

In food packaging, NSMs, with biopolymers, can either enhance the property characteristics of neat polymer or improve the functional properties of active and intelligent packaging. Packaging can be categorized as “Improved”, “Active,” and “Intelligent” packaging, which represent which kind of packaging material could be used for what applications [30]. However, official restrictions have been imposed by the European Union (EU) using “active” and “intelligent” packaging materials in foods except using titanium nitride in plastic bottles [31].

For in-packaging detection of oxygen, Mill [32] developed nano-sized titanium dioxide (TiO₂) or tin dioxide particle-based favorable photo-indicator intelligent ink using a redox-activate methylene blue dye, and this detector was gradually able to change the color in response to minor changes in oxygen quantity. In recent years, the packaging industry has focused on the various types of nanostructures and specified nanomaterials, including nano-clay particulates due to their easy accessibility, low-cost, easy process ability, and great performance. In addition, graphene nano-sheets and carbon nano-tubes are also encouragingly developed as a carbon-based nanomaterial [33]. Polymeric substrate-based flexible or static films and bottles (various materials, such as glass and plastic etc.) in combination with vacuum-coated thin inorganic layers have great demand in material packaging due to their ability to block out oxygen and aromas. While aluminum foil and aluminum-metallization used to be the material and method of choice, respectively, new materials are being used more frequently.

Thick and uniform layers (10–100 nm coating thickness) are broadly referred to as “nanocoatings” [34]. In recent years, few researchers have developed an intelligent type of nanocoating film that can indicate whether any contamination has occurred during storage. Gas content and non-invasive detection methods have also shown great ability to continually and easily monitor the gas content, excess moisture, and oxygen content of a package-headspace thus providing effective means to evaluate the quality and safety of food even after the production process [35,36]. Presence of oxygen inside the packaging can cause shelf-life threatening of food due to the ability of oxygen to create healthy environment for microbial growth [37]. Yu et al. [38] developed polynvinyl alcohol/chitosan polymer-based biodegradable and cost-effective films using a nanocomposite of silica in-situ which caused significant reduction in the permeability of oxygen and moisture by 25.6% and 10.2%, respectively, suggesting that the films extended the preservation time of cherries by three-fold compared to normal packaging. Swaroop and Shukla [39] developed a food packaging material using a combination of nanostructures of magnesium oxide (MgO) and polyactic acid biopolymer, and found that the material effectively protected against bacterial biofilms. Furthermore, Foltynowicz et al. [40] synthesized zero-valent iron particles to act as oxygen scavengers in food packaging.

3.2. The mechanisms of nanoparticles/nanocapsules as antimicrobial agents for active packaging

The outstanding use of nanoparticles with antimicrobial properties has, to some extent, protected our food products from foodborne illness outbreaks due to the consumption of spoiled packaged food. When compared with conventional food-packagings, an active food-packaging not only has the ability of being a passive barrier, but also helps to release antioxidant and antimicrobial compounds through the direct interaction with the food as well as helps to remove some negative factors, such as oxygen or water vapor. These interactions usually improve food stability [41].

Active packaging contains specific molecules which have the ability to absorb or release the components into or from the surrounding environment of the packaged food. Today, active polymer nanomaterials in food packaging have mainly been developed for antimicrobial packaging applications [42]. Overall, in order to improve the functionality and effectiveness of food packages, various bioactive substances can be incorporated into the packaging material by capsulation, capping the material with nanomaterials, or employing other nanotechniques [43]. To effectively apply antimicrobial active packaging to food products, food providers should select effective antimicrobial substances-based unique methodology to ensure the quality of the packaged products, such as their visual appearance and sensory levels [44].

The encapsulation of nutraceuticals and functional antimicrobial ingredients is essential for the preservation and

| Types of nanoparticles | Matrix | Application | Reference |
|------------------------|--------|-------------|-----------|
| Silver                 | Asparagus, Orange juice, Poultry meat, Fresh-cut melon, Beef meat exudates | Retards the growth of aerobic psychrotrophics, yeasts and molds; antimicrobial effect against Escherichia coli and Staphylococcus aureus | [165–169] |
| Zinc oxide            | Orange juice, Liquid egg albumen | Effectively reduces Lactobacillus plantarum, Salmonella, yeast and mold counts without changes in quality parameters | [169–171] |
| Titanium oxide        | Chinese jujube, Strawberry | Reduces browning, slow-down ripening, senescence and decay | [172] |
| Silver oxide          | Apple slice | Retards microbial spoilage | [173] |
bioavailability of bioactive ingredients, as it is extremely important in food processing, food storage, and passage through the gastrointestinal tract [5]. The macromolecule-based nanoparticles in food not only improve the bioavailability of bioactive polyphenolics, such as resveratrol, epigallocatechin-3-gallate, and curcumin, but also enhance the solubility of these polyphenols and thus prevent their degradation in the gastrointestinal environment [9]. There are various encapsulation techniques that have been used to produce nano or micro-particulate systems, such as nano-emulsion [45], coacervation [46,47], the extrusion method [48], fluidized bed coating [49], spray cooling [50], and spray drying [51].

Nano-encapsulation allows for the direct contact of nanomaterials during the consumption of food. In a commonly known example, silicon dioxide (SiO2) is widely applied as a fragrance carrier in various food products [52]. Several lipid-based, nano-encapsulation systems have been developed to increase the activity of antioxidant components by improving their bio-availability and solubility [53] as well as their targeted, site-specific delivery with efficient absorption [54]. Studies have found that nano-sized edible coating is an effective alternative for preservation and shelf-life extension of food, as well as prevention of food from microbial spoilage [55]. Efficacy of coatings made with gelatin incorporating nanocrystals of cellulose [56], chitosan or nano-silica [57], chitosan with nano-silica [58], and alginate or lysozyme nano-laminate has been found very effective for the preservation of the quality of fresh foods during prolonged storage [59]. Additionally, the nano-packaging-based method that incorporates blended polyethylene with nano-powders such as Ag, kaolin, anatase TiO2, and rutile TiO2 has been considered to be a novel and facile technique to preserve fruits, such as strawberries (Fragaria ananassa Duch.) [60]. Despite the tremendous research in nano-encapsulation using various materials [61], the precise delivery of foods and their safety has not been studied in detail. Therefore, future research should examine the long-term toxicity of nano-encapsulated foods [62].

In addition to this, Johnston [63] employed nanostructured calcium silicate (NCS) for the absorption of Ag⁺ ions from a solution, which formed an NCS–Ag complex that was successfully used as an antimicrobial agent for food packaging purposes. Similarly, TiO2 is commonly utilized in surface-coating as a photocatalytic disinfecting material [64] and to inactivate several pathogenic bacteria in food [65]. The antibacterial potential of chitosan-capsulated nanoparticles could be mediated through the interaction between the positively-charged chitosan and the negatively-charged cell membranes. These interactions lead to enhanced membrane permeability and cause the rupture and eventual leakage of the intracellular material [66].

Sarwar et al. [67] developed polyvinyl alcohol (PVA), nanocellulose and Ag nanocomposite films and applied them to antimicrobial food packaging. These films profound antimicrobial effects against both Methicillin Resistant Staphylococcus aureus (MRSA) and DHS-alpha strain of Escherichia coli. Valerini et al. [68] also developed nanostructured, and aluminum-doped zinc oxide-based antimicrobial coatings functionalized with polyactic acid using sputtering power. The coatings exhibited strong antibacterial potential against E. coli and were proposed as promising materials for environment-friendly active packaging. Moreover, Lu et al. [69] prepared an antimicrobial nano-emulsion that was 100 nm by encapsulating citral essential oil using ultrasonic power, and as a result, the designed nano-emulsion was significantly effective in delivering an antimicrobial agent (citral essential oil) in the food system.

Metal and metal oxide nanoparticles (NPs)-based active packagings could be effective tools of nano-composites in terms of their antimicrobial potential. Among all metal nanoparticles, silver (Ag⁺) nanoparticles are widely used in commercial sectors. Silver nanoparticles are toxic for various food pathogens [70] because silver nanoparticles increase cell membrane permeability via cell surface adherence and by degrading lipopolysaccharide [71]; in this way, the silver nanoparticles can penetrate the bacterial cell, damage its DNA [72], and release antimicrobial Ag⁺ ions [73], which bind to sulfur, oxygen, or nitrogen containing electron donor groups in molecules and thus inhibiting synthesis of adenosine triphosphate (ATP) and DNA replication, eventually leading the cell to die [74]. Millimolar concentrations of Ag⁺ ions can easily cause cytoplasm shrinkage; separation of the cell wall membrane; the devastation of the peptidoglycan in the cell wall; denaturation of ribosomes; and DNA compression, which inhibits DNA synthesis, ruptures the cell membrane, and causes cell death [74]. Basically, the three major mechanisms of the bacterial toxicity of metal-containing NPs that are widely accepted include the uptake of metal ions, which causes the intracellular ATP depletion; ROS production, which causes oxidative cellular damage, and disruption of bacterial membrane [4].

Similarly, reports have confirmed incorporation, coating, immobilization and surface modification of antimicrobial components onto packaging material [75]. Also, the higher antimicrobial effect can be achieved via direct incorporation of antimicrobial components into packaging films. Similarly, films coated with various effective antimicrobial components can result in higher antimicrobial potential. While incorporating these antimicrobial agents (volatile and non-volatile) into film packaging, these molecules can be dispersed onto the food surfaces via adopting migration, evaporation or diffusion abilities [76]. Recently, Arfat et al. [77] designed a biocomposite film based on fish skin gelatin and silver-copper bimetallic NPs. The film showed significant antibacterial potential against Listeria monocytogenes and Salmonella enterica. Several silver-based, antimicrobial master batches, such as Bactiblock®, Aglon®, Surfacine®, d2p®, IonPure®, Irgaguard®, and Biomaster® have been used to retard the growth of Campylobacter, Salmonella, and E. coli.

4. Nanotechnology in food functionality

4.1. Nanoparticles for protection from chemical deterioration

The main reason behind deterioration in the quality of food lies in an array of chemical reactions that occur between various components present in food and the external environment. Researchers have identified several nanomaterials
that can control these unwanted reactions in various food matrices. However, some of these metal and metal oxide nanomaterials cause nano-toxicity, primarily because they can form ROS and cause oxidative stress, which leads to an imbalanced redox state in the cell [78]. Therefore, relatively less reactive nanomaterials are employed as an antioxidant carrier [18, 79, 80]. Polymeric nanoparticles are suitable for the encapsulation of bioactive compounds, such as vitamins and flavonoids, and for the release of these compounds in an acidic environment, such as in the stomach [81]. The application of antioxidants and edible coatings control the browning, the conversion of phenolics into dark-colored pigments in the presence of oxygen of fresh-cut fruits [82]. Although few nanomaterials have been used directly as anti-browning agents, nano-ZnO has been used as coated active packaging to improve the shelf-life of fresh-cut Fuji apples [83].

Nanotechnology can enhance food functionality either by eliminating chemical toxicants or by enhancing nano-sized nutritional supplements. For instance, some of the nutraceuticals incorporated in the carriers consist of lycopene, β-carotenes, and phytoestrogens in order to avoid cholesterol accumulation in body [84]. It is well-known that food itself has few nanostructures that are responsible for self-mediated nano-effects. For example, green tea has many health benefits because of its nano-selenium content, which facilitates the effective uptake of selenium.

Nano-encapsulation involves a nanoscale process of packaging material by using nanocapsules, and it ensures the functionality of the final product, which includes the controlled release of the core. Therefore, encapsulated forms of ingredients exhibit numerous advantages, including consecutive delivery of multiple active ingredients, shelf-life extension, increased stability, and pH-triggered controlled release [85]. Further, Liang et al. [86] encapsulated epigallocatechin gallate (EGCG in zein/chitosan) nanoparticles for controlled, effective, functional applications in the food system. It was observed that the release of EGCG from zein/chitosan (CS) NPs and the DPPH scavenging ability of the zein/CS NPs was relatively higher than that of the nanoparticles that did not have the zein/CS coating in fatty simulant of 95% ethanol. These findings confirmed that the antioxidant activity and controlled-release of EGCG from zein/CS NPs in fatty simulant of 95% ethanol protect fatty foods from chemical deteriorations via enhancing antioxidant efficiencies and thus may help fatty foods to protect from oxidation for longer period.

4.2. Nanoparticles for enhancing the physical properties of food and packaging materials

Nanomaterials developed have shown enormous ability to increase the physical properties of both food and packaging materials [87]. In the 1990s, polymer nanocomposites with layered silicates that had several important characteristics, such as high flame resistance [88] and protection from UV rays, were developed [89]. Moreover, a number of NPs have been developed to increase the physical attributes of food, such as color and appearance. TiO₂ was approved as a coloring food additive by the USFDA with the agreement that the limit of TiO₂ should not exceed 1% (w/w) as an additive ingredient [90]. Besides TiO₂, mixture of color additives may contain SiO₂ and/or Al₂O₃, but the use of carbon black is strictly prohibited by the USFDA as a food coloring additive [91]. On the other hand, SiO₂ is used as an anti-caking agent to maintain the flow properties in powdered products and as a carrier of aroma in food and non-food products. Nano-sized SiO₂ is widely employed in food products and has been registered within the EU as a food additive, as per Directive E551 [52].

β-Carotene is a nutraceutical component that acts as a coloring agent and as provitamin A, but its incorporation into foods is limited because of its low chemical stability and hydrophobicity. To improve the physicochemical stability of β-carotene, Mehrad et al. [92] synthesized nanomaterial by encapsulating it into solid lipid nanoparticles (SLNPs) that contained palmitic acid and corn oil and stabilized the material using a whey protein isolate (WPI). In this nanostructure, the encapsulated β-carotene was protected by a solid shell made by palmitic acid crystals covering the surface of the oil droplets. Corn oil was able to decrease the exclusion of β-carotene from the solid lipid matrix to the surface of the SLNPs, while WPI increased the stability of the colloidal system, thus improving the oxidative stability of the β-carotene.

5. Nanotechnology in food safety

Globally, food safety is a growing public health concern. The primary goal of food safety is to ensure that food will not cause any harm to the consumer during preparation and consumption [93]. Food must be protected from any type of physical, chemical, and biological contamination during processing, handling, and distribution [94]. Recent developments in nanotechnology have revolutionized the food industry with its various applications in food processing, safety, and security, as well as its strides in enhancing nutraceutical value, extending shelf-life, and reducing packaging waste [94]. Today, food safety is a major concern due to rapid changes in food recipes and food habits. Foodborne pathogens, toxins, and other contaminants can cause serious threats to human health. The conventional methods for the detection of pathogens and their toxins are labor intensive and time consuming. Advances in nanotechnology have expedited the addressing of food safety issues in microbial contaminants and have improved toxin detection, shelf-life, and packaging strategies [84]. In addition, nanomaterials, including metal nanoparticles, carbon nanotubes, quantum dots, and other active nanomaterials can be used to develop biosensors for the quantification of microbes and other tests for food safety applications [84, 94].

5.1. Nanoparticles for the detection of foodborne pathogens

Nano-biosensors are bioanalytical devices that are developed using various NSMs and biological receptors in an integrated system design [96]. Various types of biosensors have been developed to detect foodborne pathogens and food spoiling materials [97, 98]. Surface enhanced Raman scattering (SERS) is employed as a nano-biosensing tool to rapidly and accurately detect microbial pathogens [99, 100]. Silver nano-
Nanoparticles employed for the detection of foodborne pathogens. Besides silver nanoparticles, graphene oxide [102], magnetic beads [103], carbon nanotubes [104], plasmonic gold [44], and silver nanoparticles [24] are commonly used to detect food-associated bacterial pathogens. In addition, synthetic DNA molecular beacons tagged with color-coded probes are used as nano-barcodes to identify pathogenic bacteria [72].

The direct detection of E. coli in food samples has also become possible by the measurement and detection of light that is scattered by the cells. This type of sensor operates on the basis of binding with a known protein and is characterized as a bacterium onto a silicon chip, which can further bind with any other E. coli bacteria present in the food sample [105]. Chen and Durst [106] devised an array-based immunosorbent assay to detect E. coli O157:H7, Salmonella spp., and L. monocytogenes using protein G-liposomal nano-vesicles in pure and mixed cultures. The researchers showed that protein G-liposomal nano-vesicles can be successfully used in immunoassays for simultaneous detection of foodborne pathogens and demonstrated the effectiveness of protein G-liposomal nano-vesicles as universal immunoassay reagents. Further, DeCory et al. [107] developed an immunomagnetic bead-immunoliposome fluorescence assay to rapidly detect E. coli O157:H7 in aqueous samples. The findings demonstrated the feasibility of using immunomagnetic beads in combination with sulfodihyamine B encapsulated in immunoliposomes to rapidly detect E. coli O157:H7 in aqueous samples. Additionally, liposome-based methods have also been investigated by other researchers for pathogen detection [108, 109]. Nano-sensors, such as nano-cantilevers, employ silicon-based materials to recognize proteins and detect pathogens that vibrate at different frequencies depending on their biomass [110].

In recent years, various new nanoparticle-based detection platforms have been developed. Tominaga [111] developed lateral-flow immune test strips with palladium nanoparticles against Klebsiella, which allowed for the specific binding and visualized detection of specific bacteria. Further, Thakur et al. [112] detected single E. coli bacterial cell using a reduced graphene, nanoparticle-based, field-effect transistor device.

Recently, our research group developed an electrochemical sensing platform based on graphene oxide–gold nanoparticles to detect Cronobacter sakazakii, which is a bacterium that is hazardous to infants, in infant formula powder with a detection limit of $2.0 \times 10^7$ cfu/mL [113]. Also, Song et al. [114] developed a fluorescence sensing platform using immunomagnetic nanoparticles coupled with liposome nanoparticles to detect Cronobacter sp. at the genus level with a detection limit of $5.9 \times 10^3$ cfu/mL. Additionally, our research team developed an aptamer and gold nanoparticle-based optical detection platform to detect Salmonella in contaminated pork samples [115]. The various types of nanoparticles for the detection of various foodborne pathogens and their detection limits are shown in Table 2.

### 5.2. Nanoparticles for protection from allergens

Nanotechnology has been employed as a basic tool to control and manage food allergens [116, 117]. Despite these efforts, certain nanomaterials cause allergic pulmonary inflammation [118, 119] in humans. For instance, SiO$_2$ nanoparticles have been reported to cause allergen-specific, Th2-type immune responses in vivo in female BALB/c mice [118]. The immunotherapy of allergies using aluminum hydroxide (alum) as an adjuvant displayed several side effects, including swelling, indurations, erythema, cutaneous nodules, and granulomas at the injection site [119]. To overcome this, researchers have investigated the use of polymeric nanoparticles [120] and substances that target toll-like receptors (TLR) as alternative adjuvants [121]. For instance, the protamine-based nanoparticles with TLR-9 ligand cytosine phosphate guanine (CpG)-oligodeoxynucleotides (ODNs) can be used as an adjuvant. Protamines are arginine-rich peptides of approximately 4 kDa found in the sperm of salmon and are isolated from the mature testicles of salmons. Protamines are biodegradable and efficiently utilized for the reversal of heparin activity during surgical operations [122] or as an insulin-additive [123]. Additionally, protamine-based nanoparticles with CpG-ODN counteract the Th2-dominated immune response induced by an allergen, thus have shown remarkable potential in allergen immunotherapy as a novel carrier system [124].

| Nanoparticles | Pathogens                                    | Detection limit | Reference |
|--------------|----------------------------------------------|-----------------|-----------|
| Gold nanoparticle | Salmonella enterica serotype Typhi           | 98.9 CFU/mL     | [174]     |
| Gold/silicon nanorod | Salmonella enterica serotype Typhi; Respiratory syncytial virus | Not reported | [175]     |
| Gold nanorod | Escherichia coli O157:H7 | 1–10 CFU/mL | [159]     |
| Quantum dot | Salmonella enterica serotype Typhi, E. coli O157:H7, Listeria monocytogenes | $10^3$–$10^6$ cells/mL | [160–163] |
| Magnetic bead/quantum dot | E. coli O157:H7 | $10^3$ CFU/mL | [176]     |
| RuBpy doped silica | E. coli O157:H7 | 1 cell/mL | [164, 176] |
| Single walled carbon nanotube | E. coli | Not reported | [177]     |
| Magnetic nanoparticle | E. coli O157:H7, S. aureus, S. epidermidis | $10^6$ CFU/mL, 8 CFU/mL, 10 CFU/mL | [177]     |
| Immunomagnetic liposome nanoparticle | Cronobacter sakazakii | $10^7$ CFU/mL | [108]     |
| Aptamer conjugated gold nanoparticles | Salmonella typhimurium | $10^4$ CFU/mL | [115, 120] |
| Liposome nanoparticles | Salmonella typhimurium | $10^2$ CFU/mL | [173]     |
Similarly, Gamazo et al. [125] reported immunotherapy that employed nanoparticles as an allergen delivery system, which facilitated the administration, reduced the dose, and diminished the allergen exposure to the immunoglobulin E that was bound to mast cells and/or basophils. Moreover, in recent years, bioinspired NSMs are being explored that are known to have little to no toxic effects and can thereby be potentially exploited in the food industry [126]. Localized surface plasmon resonance (LSPR)-based label-free biosensing methods have gained huge attention due to their high sensitivity, simplicity, and relatively low cost efficacy. However, the challenges have been observed using these methods for in situ analysis of real samples due to the instability of colloidal nanoparticles at certain levels of pH and salt concentration. Recently, Lee et al. [127] developed a simple and cost-effective optical fiber coated with aptamer-modified gold nanorods for easy detection of ochratoxin A in grape juice samples (food mycotoxin causing allergy) and subjected the fiber to LSPR analysis, which demonstrated a significant detection limit of 12.0 pM. Zhang et al. [128] reported a magnetic, nanoparticle-assisted, aptamer-based fluorescence assay to detect allergens in food matrices. As a result, under optimal conditions, the linear range was recorded as 0.4–5 g/mL (R² = 0.996), and a low limit of detection was calculated as 77 ng/mL with efficient selectivity. Moreover, Brotons-Canto et al. [129] evaluated the positive effects of poly-(anhydride) nanoparticles (150 nm) as oral vehicles for immunotherapy against experimental peanut allergies. Several other studies demonstrated the promising applications of nanotechnologies in vaccinology and in allergen immunotherapy [130]. In the near future, nanoparticle-based formulations can be applied to allergen immunotherapy to solve its unmet needs [131].

5.3. Nanoparticles for preventing heavy metal reduction

The release of heavy metals from nanomaterials poses a high risk of toxic outbreaks [80]. The release of these metals in food products has adverse effects in the case of long-term accumulation. Metal and metal oxide-based nanomaterials, such as ZnO [132], Ag [133], and CuO [134], increase the intracellular ROS level [132] and ultimately cause lipid peroxidation and DNA damage. Cationic surfactant coated silica-modified magnetite nanoparticles act as adsorbents for the micro-extraction and have ability to determine trace amount of Cu, Ni, Co, Cd, Pb, and Mn from environmental samples. The silica-coated NPs synthesized using cetylpyridinium bromide, can solubilize metal ions after being complexed with 8-hydroxyquinoline [135]. While many nanomaterials have tremendous potential for the remediation of contaminants, magnetite nanoparticles have proven to be the most attractive and inexpensive substrates for the recovery of heavy metals [136] from various sources. Zhang et al. [137] fabricated Fe₃O₄ core/shells, nanowires, and nano-necklaces to remove chromium from aqueous solutions.

Recently, Lin et al. [138] demonstrated the ability of amine-functionalized magnetic iron oxide nanoparticles, especially Cu²⁺, Ni²⁺, Pb²⁺, and Zn²⁺, to act as adsorbents to remove aqueous heavy metal ions. The results of the study displayed that the increase in the degree of amination had a positive effect with a simultaneous increase in the adsorption capacity and the initial rate of adsorption of heavy metal ions. MgO nanoparticles synthesized via sol-gel and calcination processes showed great potential against bacterial infections and were able to remove heavy metal ions from contaminated water samples [139]. These findings anticipated that nano-sized MgO particles may have potential for the treatment of bacterial and heavy metal contaminated wastewater due to their high removal efficiency, low cost, facile preparation, and environmentally friendly characteristics.

In recent years, Lingamminne et al. [140] demonstrated how synthesized, reusable, recoverable iron oxide nanoparticles could reduce heavy metals without losing their stability. In addition, over the years, various carbon nanoparticles with high fluorescence properties such as carbon dots (C-dots) with a particle size of smaller than 10 nm and carbon nanoparticles (CNPs) of about 10 nm size or larger have shown a broad range of biological properties with low toxicity and good biocompatibility. Recently, through a thermal process in the presence of H₃PO₄, Simpson et al. [141] fabricated, characterized, and approved carbon nanoparticles (66 nm) with glycerol for the detection of heavy metal ions, with a detection limit as low as 0.30 ppm.

5.4. Nanoparticles for the inhibition of biofilm formation

Biofilms are tightly-packed bunch of bacterial cells that adhere to many substrates and produce a polymeric extracellular matrix that is extremely difficult to penetrate [142]. The biofilm formation begins with the attachment of free floating microorganisms to the surface by employing van der Waals forces that create problems such as biocorrosion, biofouling, and accumulation in the food processing industries [143]. Glycerol monolaurate (GML), which is recognized as safe by the USFDA, acts as an antimicrobial agent against many Gram-positive bacteria, including Bacillus anthracis [144]. GML has been shown to inhibit the biofilm formation of three different strains of S. aureus and of MRSA [145]. Besides this, the use of antiinfectials throughout the nano-fibers of filter membranes is employed to prevent biofilm formation [137]. Moreover, Shahrorkh and Emtiazi [146] demonstrated that nanosilver particles at low concentrations (0.2 ppm) enhanced bacterial metabolism and thus suggested the use of an optimum concentration of nanosilver particles for various nanomaterials to prevent biofilm formation.

Further, nickel oxide nanoparticles (NiO NPs) have been proposed as prospective antibacterial and anti-tumor agents. Through a green approach and using Eucalyptus globulus leaf extract, Saleem et al. [147] synthesized NiO-NPs that were 10–20 nm in size and assessed their anti-biofilm activity. In addition, Gambino et al. [148] demonstrated the bio-applicability of zinc oxide nanoparticles for hindering fungal biofilm (Alternaria alternata, Penicillium chrysogenum, P. pinophilum, and Aspergillus niger) that had developed in an ancient Egyptian tomb. Chlorhexidine-conjugated gold nanoparticles exhibited biofilm inhibitory effects against Klebsiella pneumonia [149].

Bacillus subtilis has shown enormous potential in various industrial processes for the production of value-added products. Fermentation process of B. subtilis is directly related to its ability to form biofilm resulting in operational issues and major process. Ranmadugala et al. [150] evaluated the effects of two types of superparamagnetic iron oxide (IONs), such as naked
IONs and IONs coated with 3-aminopropyltriethoxylsilane (IONs@APTES) against B. subtilis in terms of their ability to inhibit its biofilm formation, bacterial growth, and cell viabilities. The results showed that a significant reduction in the total biomass of the bacterial biofilm was observed without affecting the cell viabilities. These results suggested that these nanoparticles could be applied in several industries to combat the growth of bacterial biofilm. Moreover, Thuptumdang et al. [151] suggested that controlling the growth conditions to alter the biofilm physical structure was a possible approach to reduce the impact of silver nanoparticles (AgNPs) on biofilms in engineered and natural systems. These recent findings are increasing the demand for researchers to explore the use of nanoparticles in several other industries, including medicine.

6. Are nanotechnology and big data effective enough for next industrial revolution for securing “Smart Food”?

The engineering applications of nanotechnology will herald a new digital future characterized by improvements in food safety, shelf-life performance, and food reliability. These advancements are frequently referred to as big data information and open new fields of research and applications that will increasingly affect all parts of society. There is a huge complicated response to outbreaks associated with foodborne diseases due to the globalization of food supply chains. Generation of big data in the food industry may help to facilitate encouraging advancement in food safety and security at the global scale [156]. Big data is represented as high volume, high velocity, high veracity, and/or high variety information assets requiring new processing formats so as to enhance the process of decision making, process optimization, and insight discovery [157]. Although the big data analysis and large datasets in food safety and quality have focused on improving root cause and retrospective analyses, there is a huge growth in the advancement and utilization of predictive analytics in food security sector [157]. For example, whole genome sequencing data for L. monocytogenes isolates obtained from ice cream in Kansas became publicly available soon after a listeriosis outbreak linked to ice cream was reported in early 2015. Other omics datasets, such as metagenomics data, have also been used to identify and characterize food spoilage issues. It is likely that these types of data sources will become increasingly available to personnel in the food industry.

Recently, the World Health Organization (WHO) has enfold the big data information approach to deal with the food-safety process of decision making, which has brought about a new food-safety platform known as “FOSCOLLAB,” which integrates different sources from various disciplines [158]. On this platform, organized and nano-structured information from numerous areas, for example, food, horticulture, animals, public health, and economic index, are incorporated and accessible to the users by means of a few committed dashboards [158]. The information sources associated in FOSCOLLAB are assessments of the Joint FAO/WHO Expert Committee on Food Additives and the Joint FAO/WHO Meeting on Pesticide Residues databases on chemical risk assessment, the WHO database on Collaborating Centers, and the Global Environment Monitoring System (GEMS) food databases on chemical occurrence and food consumption. There is no restriction of utilizing big data information in the fields of food safety and food quality to facilitate improved methodologies.

Finally, input food industries often work worldwide, are R&D intensive, and deal with new technologies, including genetics, robotics, informatics, and nanotechnology. Besides the challenge of feeding the world, there is the issue of sustainability, which is mainly an issue of farming and transport. Together with the relationship between food and health (e.g., obesity concerns), food safety directly affects food processors, retailers, and consumers. The ultimate objective is to estimate risks to farms and to build up a preventive way to deal with food safety issues. The use of geographical information system technology (GIST) to create food security has demonstrated incredible practices, for example, helping producers settle on more informed choices about field practices and establish the programs of targeted pathogen surveillances. Overall, the utilization of GIST to yield big data information in food safety investigations has created remarkable measures of new data with respect to the ecology of various life forms in the food environment and data on different contamination episodes. In the era of big data information, GIST is one of the data tools that enables scientists to store, analyze, process, capture, and visualize extensive datasets. Additionally, the use of GIST to complex food safety/security issues is being demonstrated for further integration of various large data sets. Essential components associated with big data requirements in nano-food safety are presented in Fig. 3.

7. Future perspectives and potential risks of nanotechnology

There have been tremendous developments in the application of nanotechnology in food science and research. Nanotechnology helps in the detection of pesticides [159], pathogens [160], and toxins [161], and tracking-tracing-monitoring helps
in the maintenance of food quality. A massive amount of interest is being on using carbon nano-tubes for incorporating them into packaging materials for the detection of toxic proteins, microorganisms, and food spoilage [162]. Furthermore, carbon nanotube can transform our future food packaging materials to active and intelligent packaging systems [163]. Along with the incessant research on the application of nanomaterials, several potential risks and toxicity issues associated with the application of nanomaterials have been revealed, and these concerns must be addressed [104,161]. The effect of these nanoscale particles on human beings, animals, and the environment are unpredictable due to changes over time in their properties. Some nanoparticles can even cross biological barriers, such as the blood–brain barrier, and enter various cells and organs [164].

The evidences for the health risks associated with the inhalation of ultrafine and nanoparticles have been increasing every day [60]. In addition, bioaccumulation of nanomaterials, such as nano-silver, derived from either nanopackaging or plants and animals has been confirmed in food and human being [62]. The use of nanomaterials as activation catalysts, pesticides, and microbicides always pose an unknown associated risk, and hence risk assessment procedures must be strictly followed while processing food [57,58]. Even with the advent of nanotechnology, the challenges to the development of a healthy and sustainable food industry remain. With the introduction and development of nanotechnology in the food system, the public should be educated regarding the associated health, safety, and environmental effects of nanotechnology.

Acknowledgements

We acknowledge financial support from the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT & Future Planning (2016R1A2B4013374 and 2014R1A5A1009799).

References

[1] Chandra P. Nanobiosensors for personalized and onsite biomedical diagnosis. The Institution of Engineering and Technology; 2016. https://doi.org/10.1049/PBHE001E.
[2] Sandoval B. Perspectives on FDA’s regulation of nanotechnology: emerging challenges and potential solutions. Compr Rev Food Sci Food Saf 2009;8:375–93.
[3] Moriarty P. Nanostructured materials. Rep Prog Phys 2001;64:297.
[4] Pathkoti K, Manubolu M, Hwang HM. Nanostructures: current uses and future applications in food science. J Food Drug Anal 2017;25(2):245–53.
[5] Bhushan B. Springer handbook of nanotechnology. Berlin; Heidelberg: Springer Science & Business Media; 2010.
[6] Gupta A, Eral HB, Hatton TA, Doyle PS. Nanoemulsions: formation, properties and applications. Soft Matter 2016;12:2826–41.
[7] Singh T, Shukla S, Kumar P, Wahlia V, Bajpai VK, Rather IA. Application of nanotechnology in food Science: perception and overview. Front Microbiol 2017;8:e1501.
[8] Fu PP. Introduction to the special issue: nanomaterials – toxicology and medical applications. J Food Drug Anal 2014;22:1–2.
[9] Hu B, Liu X, Zhang C, Zeng X. Food macromolecule based nanodelivery systems for enhancing the bioavailability of polyphenols. J Food Drug Anal 2017;25:3–15.
[10] Baranwal A, Srivastava A, Kumar P, Bajpai VK, Maurya PK, Chandra P. Prospects of nanostructure materials and their composites as antimicrobial agents. Front Microbiol 2018:9:e422.
[11] Das MP, Livingstone JR, Veluswamy P, Das J. Exploration of Wedelia chinensis leaf-assisted silver nanoparticles for antioxidant, antibacterial and in vitro cytotoxic applications. J Food Drug Anal 2017;26:917–25.
[12] He X, Hwang HM. Nanotechnology in food science: functionality, applicability, and safety assessment. J Food Drug Anal 2016;24:671–81.
[13] Adabi M, Naghibzadeh M, Adabi M, Ali Zarrinfard M, Esnaashari SS, Seifalian AM, et al. Biocompatibility and nanostructured materials: applications in nanomedicine. Artif Cells Nanomed Biotechnol 2017;45:833–42.
[14] He X, Aker WG, Pelaez M, Lin Y, Dionysiou DD, Hwang HM. Assessment of nitrogenfluorine-codoped TiO2 under visible light for degradation of BPA: implication for field remediation. J Photochem Photobiol Chem 2016;314:81–92.
[15] Wang B, Feng WY, Wang TC, Jia G, Wang M, Shi JW, et al. Acute toxicity of nano- and micro-scale zinc powder in healthy adult mice. Toxicol Lett 2006;16:115–23.
[16] He X, Aker WG, Hwang HM. An in vivo study on the photoenhanced toxicities of S-doped TiO2 nanoparticles to zebra fish embryos (Danio rerio) in terms of malformation, mortality, rheotaxis dysfunction, and DNA damage. Nanotoxicology 2014;8(S1):185–95.
[17] Peters RJ, Bouwmeester H, Gottardo S, Arena M, Brandhoff P. Nanomaterials for products and application in agriculture, feed and food. Trends Food Sci Technol 2016;54:155–64.
[18] Nanotechnology information statement. IFST – Institute of Food Science & Technology; 2006.
[19] Robertson GL. Food packaging: principles and practice. 2nd ed. New York: CRC Press, Taylor & Francis Group; 2006. ISBN 9781439862414.
[20] Finnigan B. Barrier polymers. In: Yam KL, editor. The wiley encyclopedia of packaging technology. New York: John Wiley and Sons Inc; 2009. p. 103–9.
[21] Abbaspour A, Nerouz-Sarvestani F, Noori A, Soltani N. Aptamer-conjugated silver nanoparticles for electrochemical dual-aptamer-based sandwich detection of Staphylococcus aureus. Biosens Bioelectron 2015;68:149–55.
[22] Ghannarazadeh B, Oleyaie SA, Almasi H. Nano-structured materials utilized in biopolymer based plastics for food packaging applications. Crit Rev Food Sci Nutr 2014;7:37–41.
[23] Abdollahi M, Rezaei M, Farzi G. A novel active biocomposite film incorporating rosemary essential oil and nanoclay into chitosan. J Food Eng 2012;111(2):343–50.
[24] Azeredo HMC, Mattoso LHC, McHugh TH. Nanocomposites in food packaging—a review. In: Reddy B, editor. Advances in diverse industrial applications of nanocomposites. InTech; 2011. p. 550. https://doi.org/10.5772/1931.
[25] Klaine SJ. Koelmans AA, Horne N, Carley S, Handy RD, Kapustka L. Paradigms to assess the environmental impact of manufactured nanomaterials. Environ Toxicol Chem 2012;31(1):3–14.
[26] Klaine SJ. Considerations for research on the environmental fate and effects of nanoparticles. Environ Toxicol 2009;28(9):1787–8.
of the assay with a standard microbiological method. Appl Environ Microbiol 2005;71:1856–64.

[108] Shukla S, Lee G, Song X, Park S, Kim M. Immunoliposome-based immunomagnetic concentration and separation assay for rapid detection of Cronobacter sakazakii. Biosens Bioelectron 2016;77:986–94.

[109] Shukla S, Leem H, Kim M. Development of a liposome-based immunochromatographic strip assay for the detection of Salmonella. Anal Bioanal Chem 2011;401:2581–90.

[110] Jain KK. Nanodiagnostics: application of nanotechnology in molecular diagnostics. Expert Rev Mol Diagn 2003;3(2):153–61.

[111] Tominaga T. Rapid detection of Klebsiella pneumoniae, Klebsiella oxytoca, Raoultella ornithinolytica and other related bacteria in food by lateral-flow test strip immunassays. J Microbiol Meth 2018;147:43–9.

[112] Thakur B, Zhou G, Chang J, Pu H, Jin B, Sui X, et al. Rapid detection of single E. coli bacteria using a graphene-based field-effect transistor device. Biosens Bioelectron 2018. https://doi.org/10.1016/j.bios.2018.03.014.

[113] Shukla S, Haldorai Y, Bajpai VK, Rengaraj A, Hwang SK, Song X, et al. Electrochemical coupled immunosensing platform based on graphene oxide/gold composite for sensitive detection of Cronobacter sakazakii in powdered infant formula. Biosens Bioelectron 2018;30:139–49.

[114] Song X, Shukla S, Kim M. Detection of Cronobacter species in powdered infant formula using immunoliposome-based immunomagnetic concentration and separation assay. Food Microbiol 2018;72:23–30.

[115] Oh SY, Heo NS, Shukla S, Cho HY, Ezhil Vilian AT, Kim J, et al. Development of gold nanoparticle aptamer-based LSPR sensing chips for the rapid detection of Salmonella typhimurium in pork meat. Sci Rep 2017;7:e10130.

[116] Kumar S, Dilbaghi N, Barnela M, Bhanjana G, Kumar R. Biosensors as novel platforms for detection of food pathogens and allergens. BioNano Sci 2012;2:196–217.

[117] Pilolli R, Monaci L, Visconti A. Advances in biosensor development based on integrating nanotechnology and applied to food-allergen management. Trends Anal Chem 2013;47:12–26.

[118] Yoshida T, Yoshioka Y, Fujimura M, Yamashita K, Higashisaka K, Morishita Y, et al. Promotion of allergic immune responses by intranasal-administrated nanosilica particles in mice. Nanoscale Res Lett 2011;6:1–6.

[119] Vogelbruch M, Nuss B, Korner M, Kapp A, Kiehl P, Bohm W. Aluminium induced granulomas after inaccurate intradermal hyposensitization injections of aluminium-adsorbed depot preparations. Allergy 2000;55:883–7.

[120] De Souza Reboucas J, Espana I, Ferrer M, Sanz ML, Irache JM, Gamazo C. Nanoparticulate adjuvants and delivery systems for allergen immunotherapy. J Biomed Biotechnol 2012:e474605.

[121] Hedayat M, Takeda K, Rezaei N. Prophylactic and therapeutic implications of toll-like receptor ligands. Med Res Rev 2012;32:294–325.

[122] Schulman S, Bijsterveld NR. Anticoagulants and their reversal. Transfus Med Rev 2007;21:37–48.

[123] Owens DR. Insulin preparations with prolonged effect. Diabetes Technol Therapeut 2011;13(51):S5–14.

[124] Pali-Scholl I, Szollosi H, Starkl P, Scheiber B, Streumntzer C, Hofmeister A. Protamine nanoparticles with CpG-oligodeoxynucleotide prevent an allergen-induced Th2-response in BALB/c mice. Eur J Pharm Biopharm 2013;85:656–64.

[125] Gamazo C, Gastaminza G, Ferrer M, Sanz ML, Irache JM. Nanoparticle-based-immunotherapy against allergy. Immunotherapy 2014;6(7):885–97.

[126] Prasad A, Mahato K, Chandra P, Srivastava A, Joshi SN, Maurya PK. Bioinspired composite materials: applications in diagnostics and therapeutics. J Mol Eng Mater 2016;4:e164004.

[127] Lee B, Park JH, Byun YJ, Kim JH, Kim MG. An optical fiber-based LSPR aptasensor for simple and rapid in-situ detection of ochratoxin A. Biosens Bioelectron 2018;102:504–9.

[128] Zhang Y, Wu Q, Sun M, Zhang J, Mo S, Wang J, et al. Magnetic-assisted aptamer-based fluorescent assay for allergen detection in food matrix. Sensor Actuator B Chem 2018;263:43–9.

[129] Brotons-Cantoa A, Gamazo C, Martin-Arbella N, Abdulkarim M, Matias J, Gumbleton M, et al. Evaluation of nanoparticles as oral vehicles for immunotherapy against experimental peanut allergy. Int J Biol Macromol 2018;110:325–35.

[130] Di Felice G, Barletta B, Bonura A, Butteroni C, Corinti S, Colombo P. Nanoparticles adjuvants in allergy: new challenges and pitfalls. Curr Pharmaceut Des 2015;21:4229–39.

[131] Di Felice G, Colombo P. Nanoparticle–allergen complexes for allergen immunotherapy. Int J Nanomed 2017;12:4493–504.

[132] Fuku H, Horie M, Endoh S, Kato H, Fujita K, Nishio K. Association of zinc ion release and oxidative stress induced by intratracheal instillation of ZnO nanoparticles to rat lung. Chem Biol Interact 2012;198:29–37.

[133] McShan D, Ray PC, Yu H. Molecular toxicity mechanism of nanosilver. J Food Drug Anal 2014;22:116–27.

[134] Karlsson HL, Cronholm P, Hedberg Y, Tornberg M, Battie LD, Svedhem S, et al. Cell membrane damage and protein interaction induced by copper containing nanoparticles-importance of the metal release process. Toxicology 2013;313:59–69.

[135] Karatapans AE, Fiamigos Y, Stalikas CD. Silica-modified magnetic nanoparticles functionalized with cetylpyridinium bromide for the preconcentration of metals after complexation with 8-hydroxy-quinoline. Talanta 2011;84:834–9.

[136] Amin MT, Alazba AA, Manzoor U. A review of removal of pollutants from water/wastewater using different types of nanomaterials. Adv Mater Sci Eng 2014:1–24.

[137] Zhang L, Luo J, Menkhaus TJ, Varadaraju H, Sun Y, Fong H. Antimicrobial nano-fibrous membranes developed from electropun polycrylonitrile nanofibers. J Membr Sci 2011;369:499–505.

[138] Lin S, Xu M, Zhang W, Hu X, Lin K. Quantitative effects of amination degree on the magnetic iron oxide nanoparticles (MIONPs) using as adsorbents to remove aqueous heavy metal ions. J Hazard Mater 2017;335:47–55.

[139] Cai Y, Li C, Wu D, Wang W, Tan F, Wang X, et al. Highly active MgO nanoparticles for simultaneous bacterial inactivation and heavy metal removal from aqueous solution. Chem Eng J 2017;312:158–66.

[140] LinghamJNP,ChangYY,YangJK,SinghJ,ChoiEH, Shiratani M, et al. Biogenic reductive preparation of active MgO nanoparticles for simultaneous bacterial inactivation and heavy metal removal from aqueous heavy metal ions in aqueous media. Carbon 2018;127:122–30.

[141] Forier K, Raemdonck K, De Smedt SC, Demeester J, Coenye T, Braeckmans K. Lipid and polymer nanoparticles
for drug delivery to bacterial biofilms. J Control Release 2014;190:607–23.
[143] Shakeri S, Kermanshahi R, Moghaddam M, Emtiaz G. Assessment of biofilm cell removal and killing and biocide efficacy using the microtiter plate test. Biofouling 2007;23:79–86.
[144] Vetter SM, Schlievert PM. Glycerol monolaurate inhibits virulence factor production in Bacillus anthracis. Antimicrob Agents Chemother 2005;49:1302–5.
[145] Schlievert PM, Peterson ML. Glycerol monolaurate antibacterial activity in broth and biofilm cultures. PLoS One 2012;7:e40350.
[146] Shahrokh S, Emtiaz G. Toxicity and unusual biological behavior of nanosilver on Gram positive and negative bacteria assayed by microtiter-plate. Eur J Biol Sci 2009;1:28–31.
[147] Saleem S, Ahmed B, Khan MS, Al-Shaeri S, Musarrat J. Toxicity and unusual biological behavior of nanosilver on Gram positive and negative bacteria assayed by microtiter-plate. Eur J Biol Sci 2009;1:28–31.
[148] Saleem S, Ahmed B, Khan MS, Al-Shaeri S, Musarrat J. Toxicity and unusual biological behavior of nanosilver on Gram positive and negative bacteria assayed by microtiter-plate. Eur J Biol Sci 2009;1:28–31.
[149] King T, Cole M, Farber JM, Eisenbrand G, Zabaras D, Fox EM, Wiedmann M. Can big data revolutionize food safety? Food Quality and Safety 2015. Retrieved from: http://www.who.int/foodsafety/foscollab_dashboards/en/.
[150] WHO – World Health Organization, FOSCOLLAB. Global platform for food safety data and information. 2015. http://www.who.int/foodsafety/foscollab_dashboards/en/.
[151] Fu J, Park B, Siragusa G, Jones L, Tripp R, Zhao Y. An Au/Si hetero-nanorod-based biosensor for Salmonella detection. Nanotechnology 2008;19:155502.
[152] Hahn MA, Keng PC, Krauss TD. Flow cytometric analysis to detect pathogens in bacterial cell mixtures using semiconductor quantum dots. Anal Chem 2008;80:3864–72.
[153] Yang L, Li Y. Quantum dots as fluorescent labels for quantitative detection of S. typhimurium in chicken carcass wash water. J Food Protect 2005;6:1241–5.
[154] Tully E, Hearty S, Leonard P, O’Kennedy R. The development of rapid fluorescence-based immunoassays, using quantum dot-labelled antibodies for the detection of L. monocytogenes cell surface proteins. Int J Biol Macromol 2006;1:3;127–34.
[155] Wang C, Irudayaraj J. Gold nanorod probes for the detection of multiple pathogens. Small 2008;12:2204–8.
[156] Su SL, Li Y. Quantum dot biolabeling coupled with immunomagnetic separation for detection of Escherichia coli O157:H7. Anal Chem 2004;76(16):4806–10.
[157] An J, Zhang M, Wang S, Tang J. Physical, chemical and microbiological changes in stored green asparagus spears as affected by coating of silver nanoparticles-PVP. LWT – Food Sci Technol 2008;41(6):1100–7.
[158] Fernandez A, Soriano E, Lopez-Carballo G, Picouet P, Lloret E, Gavara R. Preservation of aseptic conditions in absorbent pads by using silver nanotechnology. Food Res Int 2009;42(8):1105–12.
[159] Fernandez A, Picouet P, Lloret E. Cellulose-silver nanoparticle hybrid materials to control spoilage-related microflora in absorbent pads located in trays of fresh-cut melon. Int J Food Microbiol 2010a;142(1–2):222–8.
[160] Fernandez A, Picouet P, Lloret E. Reduction of the spoilage-related microflora in absorbent pads by silver nanotechnology during modified atmosphere packaging of beef meat. J Food Protect 2010b;73(12):2263–9.
[161] Emamifar A, Kadivar M, Shahedi M, Soleimanian-Zad S. Evaluation of nanocomposite packaging containing Ag and ZnO on shelf life of fresh orange juice. Innovat Food Sci Emerg Technol 2010;11(4):742–8.
[162] Emamifar A, Kadivar M, Shahedi M, Soleimanian-Zad S. Effect of nanocomposite packaging containing Ag and ZnO on inactivation of Lactobacillus plantarum in orange juice. Food Control 2011;22(3–4):408–13.
[163] Jin T, Gurtler JB. Inactivation of Salmonella in liquid egg albumen by antimicrobial bottle coatings infused with allyl isothiocyanate, nisin and zinc oxide nanoparticles. J Appl Microbiol 2011;110(3):704–12.
[164] Li H, Li F, Wang L, Sheng J, Xin Z, Zhao L, et al. Effect of nanopack on preservation quality of Chinese jujube (Ziziphus jujuba Mill. var. inermis (Bunge) Rehd). Food Chem 2009;114(2):547–52.
[165] Zhou L, Lv S, He G, He Q, Shi B. Effect of PE/AG2O nanopackaging on the quality of apple slices. J Food Qual 2011;34(3):171–6.
[166] Vikesland PJ, Wigginsen KR. Nanomaterial enabled biosensors for pathogen monitoring – a review. Environ Sci Technol 2010;10:3656–69.
[167] Dungchai W, Sangroh W, Chaicumpa T, Tongtawe P, Chailapakul O. S. typhi determination using voltammetric amplification of nanoparticles: a highly sensitive strategy for metalloimmunoassay based on a copper-enhanced gold label. Talanta 2008;2:727–32.
[168] Yang L, Li Y. Quantum dot bioconjugates for simultaneous detection of Escherichia coli O157:H7 and Salmonella Typhimurium. Analyst 2006;131:394–401.
[169] Zhao X, Hilliard L, Mechery S, Wang Y, Bagwe R, Jin S. A rapid bioassay for single bacterial cell quantitation using bioconjugated nanoparticles. Proc Natl Acad Sci USA 2004;42:15027–32.