Highly resolved Mn Kβ emission: a potential probe in laboratory for analysis of ligand coordination around Mn atoms in gels and solutions

Hisashi HAYASHI† and Mao TAKAISHI

Department of Chemical and Biological Sciences, Faculty of Science, Japan Women’s University, 2-8-1 Mejirodai, Bunkyo, Tokyo 112-8681, Japan

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† To whom correspondence should be addressed.

E-mail: hayashih@fc.jwu.ac.jp
Abstract

Mn Kβ spectra of Mn, MnO, MnSO₄·H₂O, KMnO₄, 0.50 M MnSO₄ aqueous solution, and the precipitation bands of Mn-Fe-based Prussian blue analogs formed in 2.4 wt. % agarose gel (“Gel”) were measured using a laboratory-use X-ray setup with ~2.6 eV instrumental resolution, which comprises a cylindrically bent Si (400) crystal monochromator and a spherically bent Ge (440) crystal analyzer. The oxidation-state dependent shift of the Mn Kβ₁,₃ peak (~1 eV) was clearly observed for Mn, MnO, and KMnO₄, confirming that the employed setup can acquire the key features of Mn Kβ spectra. The Mn Kβ spectra of MnSO₄·H₂O, the 0.50 M solution, and Gel exhibited small but distinguishable differences, whereas the spectra acquired at two positions in Gel were almost the same. These results suggest that highly resolved Mn Kβ spectra can be helpful for assessing ligand coordination around Mn atoms in gels and solutions.

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Introduction

High-resolution X-ray fluorescence (HR-XRF) spectroscopy,\textsuperscript{1} also called X-ray emission spectroscopy,\textsuperscript{2,3} is a bulk-sensitive and non-destructive method for assessing spin and ligand characteristics of a given 3d\textsuperscript{3‒13} or 4f-element\textsuperscript{14‒16} in the studied samples. HR-XRF spectroscopy, as well as its complementary technique, X-ray absorption fine structure (XAFS) spectroscopy, is advantageous in that no special sample preparation is usually required and it is applicable to crystalline materials as well as solutions, gels, or amorphous materials.

Although HR-XRF spectroscopy has a long research history of use for observing chemical effects in the laboratory,\textsuperscript{17‒19} most applications of HR-XRF spectroscopy in the last 20 years (with rare exceptions such as radiative Auger effects\textsuperscript{20}) have been performed at synchrotron radiation (SR) facilities, which can provide bright, coherent, energy-tunable and monochromatic X-rays as well as sophisticated detection systems.\textsuperscript{21} Excessive reliance on SR, however, hinders the application of HR-XRF spectroscopy (1) in more introductory, routine, high-throughput, or lengthy experiments and (2) for complicated, fragile, or unsafe (e.g., radioactive) samples. To make matters worse, at present, only a handful of SR stations are routinely available for HR-XRF measurements. Hence, HR-XRF spectroscopy is currently far less familiar to many analytical scientists compared to other common spectroscopic techniques such as IR, Raman, and UV-Vis spectroscopies.

Interestingly, interest in HR-XRF measurements in the laboratory is currently seeing a resurgence.\textsuperscript{21‒24} For example, very recently, highly resolved, high quality Kβ (3p→1s) spectra of V,\textsuperscript{23} Fe,\textsuperscript{24} Ni,\textsuperscript{23,24} Co,\textsuperscript{21,22} Cu,\textsuperscript{24} and Zn\textsuperscript{23} in several materials have been measured by different laboratory spectrometers. The spread of these ‘new-generation’ HR-XRF spectrometers may relieve the aforementioned issues related to the excessive reliance on SR, and the resultant enhancement of the versatility of HR-XRF spectroscopy may be beneficial for analytical sciences.
Meanwhile, laboratory HR-XRF applications to two chemically important systems, gels and solutions, are still scarce. Thus, in this study, we have examined the applicability of HR-XRF spectroscopy for analyzing gel and solution samples. As a test HR-XRF probe, Mn Kβ emission was selected, primarily because this emission has been particularly extensively used in determining the spin state, oxidation state, and ligand specification of many solid samples, including oxygen-evolving complexes, biomimetic complexes, and magnetic materials. Furthermore, the selection of Mn Kβ emission is also justified by the following. Recently, Mn-Fe-based Prussian blue analogs (Mn-Fe PBAs) have been found to show several interesting crystallization/precipitation phenomena in gels, and their local structure around Mn atoms (examined by XAFS spectroscopy at SR facilities) has become the subject of intense research interest. Meanwhile, the local structure during the development of Mn-Fe PBA precipitation bands (which can significantly influence the quality of Mn-Fe PBAs finally formed) has not been explored yet, because the time required for their full development (~10 days) is too long to continuously monitor their XAFS at SR facilities. Laboratory Mn Kβ spectroscopy is expected to be potentially useful also for in-situ observation of the Mn local structure during the development of the Mn-Fe PBA bands in gels.

**Experimental**

**Chemicals**

Analytical reagent grade powders of Mn, MnO, MnSO₄·H₂O, KMnO₄, and K₃[Fe(CN)₆] were obtained from Wako Pure Chemical Industries (Osaka, Japan). Agarose for electrophoresis (gel strength: 1800–2300 g/cm³) was purchased from Kanto Chemical (Tokyo, Japan). All chemicals were used without further purification. All aqueous solutions were prepared using deionized water, which was purified from tap water by a cartridge water purifier (G-10, Organo, Tokyo, Japan).
Preparation of samples

The powders of Mn, MnO, MnSO₄·H₂O, and KMnO₄, as well as 0.50 M MnSO₄ solution, were packed in sample cells prepared from plastic straws (straw cells) for their Mn Kβ emission measurements.

A sample, named “Gel”, was prepared in accordance with a previous study as follows. In this sample, two different electrolytes were loaded in separate gel columns in a straw cell. The longer gel column (~30 mm) was placed at the bottom, where the periodic precipitation bands subsequently form. The shorter column (~20 mm), on top, had a higher electrolyte concentration. To prepare the longer gel column, an appropriate amount of agarose was dissolved at ~98 °C in deionized water (30 mL) with vigorous stirring to prepare a sol with 2.4 wt.%. A predetermined amount of K₃[Fe(CN)₆] was added to the sol, and the mixture was stirred continuously for 30 s to prepare the 0.10 M [Fe(CN)₆]³⁻ sol. This sol was transferred to a straw cell using a Pasteur pipette. The hot sol was allowed to cool to 25 °C, and a solidified gel was formed within 20 min. Moreover, to prepare the shorter column, an appropriate amount of agarose was dissolved at ~98 °C in deionized water (30 mL) with vigorous stirring to prepare a sol with 0.5 wt.%. A predetermined amount of MnSO₄·H₂O was added to the agarose sol, and the mixture was stirred continuously for 30 s to prepare the 0.50 M Mn²⁺ sol. The resultant Mn²⁺ sol was poured over the longer [Fe(CN)₆]³⁻ gel column using a Pasteur pipette, and the Mn²⁺ sol also solidified into a gel within 20 min. The straw cell was subsequently closed using styrene-resin stoppers covered by Teflon and was allowed to stand at 25 °C for ~8 days to allow precipitation bands to form in the gels. The unreacted electrolytes in Gel were then removed by suction and addition of deionized water (~40 mm in height) to the longer gel column using a Pasteur pipette. This removal procedure was repeated three times every 2 days.
Apparatus and measurements

To measure highly resolved Mn Kβ emission spectra, a wavelength-dispersive, laboratory-use HR-XRF setup was arranged (refer to Fig. 1). A previously reported setup\textsuperscript{32,33} was modified to form the setup for this study. The Cu Kα\textsubscript{1} X-rays from an 18 kW generator (RU-300, Rigaku, Tokyo, Japan), operated at 40 kV and 150 mA, were focused within 0.5 mm in the horizontal direction by a Si (400) Johansson-type crystal monochromator (Saint-Gobain Crystals, Nemours, France). This crystal was asymmetrically cut and had a radius of curvature of 450 mm. The focal lengths for Cu Kα\textsubscript{1} were 187 mm (generator–monochromator) and 316 mm (monochromator–sample). The monochromator was aligned to obtain the maximal output current from an ion chamber (S-1194B1, Ohyo Koken Kogyo, Tokyo, Japan). A slit (Cross slit screen 3013, Huber, Rimsting, Germany) of 5 × 7 mm\textsuperscript{2} (width × height) was placed nearly 100 mm in front of the sample to guard from parasitic emissions from the monochromator. Before and after each Mn Kβ measurement, the intensity of the incident X-rays was monitored by the ion chamber, with the variation being within 2%. The straw cell containing samples was mounted vertical to the line-shape X-ray beams on the focal position. No automatic positioning stages for the sample are currently available for this setup, so the irradiation position was changed manually.

In most laboratory-use HR-XRF spectrometers developed recently,\textsuperscript{21-24} X-rays from the generator are directly irradiated onto the sample to avoid the loss of the incident X-ray intensity upon introducing a monochromator. In fact, the intensity of the incident X-rays (Cu Kα\textsubscript{1}) in the current setup, which was estimated by the output from the ion chamber (~5 × 10\textsuperscript{9} /s),\textsuperscript{33} is considerably lower than the intensities of modern HR-XRF spectrometers (e.g., ~8 × 10\textsuperscript{11} /s).\textsuperscript{21} Meanwhile, the crystal monochromator is very effective in eliminating the background in the HR-XRF spectra. Such elimination compensates, at least partially, for the intensity loss of the incident X-rays (or the intensity loss of the HR-XRF signals). Note also that line-focused X-ray beams, which are produced by the curved monochromator, are preferable to examine position dependence.
of HR-XRF in (pseudo-) one-dimensional systems, such as Liesegang bands in gels.\textsuperscript{25-27,29} In addition, line-focused, monochromatic X-ray incidence allows us to easily incorporate the HR-XRF setup with other X-ray instruments, such as small-angle X-ray scattering (SAXS) diffractometers\textsuperscript{34} (SAXS is a standard technique to study structural properties of gels and solutions\textsuperscript{35}).

Mn K$\beta$ emission from the sample was analyzed and focused by a Ge (440) spherically bent Johansson-type crystal (Saint-Gobain Crystals; bending radius, 820 mm; diameter, 75 mm). The crystal was placed so that the detection angles were 140°. To achieve high-resolution, the analyzer crystal was operated close to back-scattering geometry, covering the Bragg angles ranging from 71.62° to 72.60°, corresponding to an energy range of 6464–6500 eV. The instrumental resolution of this setup will be discussed later.

Highly resolved Mn K$\beta$ spectra were measured by rotating the analyzer crystal on a computer-controlled $\theta$ stage (RA07A-W, Kohzu Precision, Kawasaki, Japan) and linearly moving a silicon PIN detector (XR-100CR, Amptek, Bedford, MA) on an $\chi$ stage (XA05A-L2, Kohzu Precision) simultaneously. Signals from the detector were fed into conventional photon-counting electronics: a timing single-channel analyzer (Model 553, Ortec, Atlanta, GA) and a dual counter/timer (Model 994, Ortec). The beam path has 50 $\mu$m-thick Kapton windows, and it was evacuated using a rotary pump (VR16, Koki Holdings, Tokyo, Japan) to avoid the absorption and scattering of X-rays by air. In current measurements, the Mn K$\beta$ spectra were collected over the emission energy range of 6464–6500 eV in 0.5 eV intervals. All measurements were performed at 25 °C.

The energies of the measured Mn K$\beta_{1,3}$ peaks were determined by fitting the peaks with a Voigt function using a software for data analysis and graphing, Origin 2019 (OriginLab, Northampton, MA). The fitting errors in the calculated peak energies using Origin 2019 were within 0.05 eV. The reproducibility of the K$\beta$ spectra was checked by repeating the above-
mentioned measurements at least 3 times. The differences in the Kβ1,3 peak energies obtained by the repeatedly measured spectra were within 0.06 eV. The spectra obtained for each sample were added using Origin 2019 to reduce their statistical uncertainties. A typical collection of the Mn Kβ spectra took 6 (Mn) – 24 h (Gel), depending on the number of repetitions of the measurements.

In order to estimate the distribution of Mn-Fe PBAs precipitates in Gel, energy-dispersive X-ray fluorescence (ED-XRF) intensity distribution of Gel was also measured using an experimental setup, the details of which are provided elsewhere. Briefly, Cu Kα X-rays from RU-300, operated at 40 kV and 60 mA, were used as the excitation source and were focused within 0.5 mm in the horizontal direction by an SiO2 (10 1̅ 1) Johansson-type crystal monochromator (Saint-Gobain Crystals). Gel was placed vertically with respect to the incident X-ray beam with line-shape on a computer-controlled X-Z stage (the combination of two XA05A-L2 stages). The X-ray fluorescence signals from Gel were detected using an XR-100CR detector, and the signals were collected for 90 s using a multichannel analyzer (MCA8000A, Amptek).

The ED-XRF intensity distribution of Gel in the X-direction (the X-dependence of the ED-XRF intensity) was monitored at 25 °C by moving the straw cell containing Gel in the X-direction in 1 mm increments. Following the subtraction of the constant background from the ED-XRF peaks (Mn Kα and Fe Kα peaks), the integrated intensities of the peaks were used to determine the ED-XRF distributions: Mn Kα (5691–6124 eV) and Fe Kα (6214–6647 eV). The overlap of the Mn Kβ–Fe Kα lines in the region containing both of these elements was estimated by taking the Mn Kβ (Kβ1,3 + Kβ3,5) components in the Fe Kα line to be 11.8% of the observed Mn Kα (Kα1 + Kα2) intensity, as described in the literature.

Results and Discussion

The instrumental resolution of the current setup and the confirmation of the oxidation-state dependence of Mn Kβ emission
Fig. 2(a) compares the Mn Kβ spectrum of MnO (“Lab”) acquired from the laboratory setup (see Fig. 1) with the corresponding spectrum (“SPring-8”) acquired from multi-crystal, von Hamos-type setup at BL47XU, SPring-8, Japan. Note that the two spectra were obtained under different experimental conditions (different X-ray sources, different optics, different instrumental resolutions, different analyzer crystals, and different detectors). To better overlay with the laboratory data, the SPring-8 spectrum was additionally broadened by convolution with a Gaussian function of FWHM = 1.2 eV using Origin 2019. The satisfactorily good agreement found in Fig. 2(a) suggests that the instrumental resolution of the current setup is (1) lower than that of the SPring-8 setup (~1.4 eV, which was estimated from the FWHM of the elastic scattering peak for 6.4 keV X-rays), and (2) estimated to be ~1.4 eV + 1.2 eV ≈ 2.6 eV. The lower resolution of the current setup is considered to be mostly due to: (1) the different width of the irradiation area on the sample (~0.2 mm in SPring-8 versus ~0.5 mm in the current setup) and (2) the astigmatism of the spherically bent Ge(440) crystal.

Fig. 2(b) shows Mn Kβ spectra of Mn, MnO, and KMnO₄ acquired from the laboratory setup. Despite a lower instrumental resolution, the shift of the Mn Kβ₁,3 peak of their compounds with the oxidation state change is clearly observable, as reported in the literature. The shift values from the peak energy of Mn (so-called, chemical shift) were estimated to be +1.1 eV (MnO) and −1.0 eV (KMnO₄) through fitting using a Voigt function. These values agree well with the values available in the literatures: +1.1 eV (MnO) and −1.0 eV (KMnO₄). These agreements indicate that the instrumental error of the current setup is within 0.1 eV. Moreover, considering the fitting errors (< 0.05 eV) and the precision of the spectral data (i.e., the random errors, < 0.06 eV), we estimate the overall experimental error in the obtained Mn Kβ₁,3 peak energies to be within \( \sqrt{0.1^2 + 0.05^2 + 0.06^2} \approx 0.13 \) eV.

In the lower energy side of the Kβ₁,3 peak of MnO, a relatively intense satellite peak, i.e., Mn Kβ’ peak, which is split from the Kβ₁,3 peak by (3p, 3d) exchange, can be seen, whereas the
Kβ′ peak for Mn and KMnO₄ is not very evident. These Kβ′ results are also consistent with previous high-resolution Mn Kβ studies,¹⁷,¹⁸ including modern SR studies.³⁷,³⁸ Thus, Fig. 2(b) demonstrates that the instrumental resolution of the current setup (2.6 eV), as well as the relatively small experimental error (< 0.13 eV), is sufficient to resolve the key features of Mn Kβ spectra without severe loss in scientific information.

Example 1: Mn²⁺ aqueous solution

Fig. 3 compares the Mn Kβ spectrum of the 0.50 M MnSO₄ solution with the spectrum of MnSO₄·H₂O powder acquired from the laboratory setup in Fig. 1. Fig. 3 shows that there is a small but measurable shift (~0.4 eV) between the Kβ₁,₃ peaks of the two samples. The local structure around Mn atoms in the two samples is different: the octahedral coordination of O atoms with a distance of 0.220 nm in MnSO₄ aqueous solution differs from the distorted octahedral coordination of O atoms with three Mn–O distances of 0.212, 0.214, and 0.228 nm in MnSO₄·H₂O powder.⁴⁰ Generally, the Mn Kβ₁,₃ peak is less dependent on the ligand environment of Mn atoms than on the oxidation state.⁵,⁸ Nevertheless, the 0.4 eV shift observed here suggests that the Kβ₁,₃ peak energy is, in some cases, sensitive enough to detect the changes in the ligand coordination around Mn atoms. The suggested Mn Kβ₁,₃ sensitivity is expected to be helpful for addressing issues in solution chemistry, e.g., the ligand changes of Mn atoms in Mn²⁺ aqueous solutions (including MnCl₂ and MnBr₂ solutions⁴¹,⁴²) as the solute concentration increases.

Example 2: Mn–Fe PBA precipitation bands formed in agarose gel

The ED-XRF (Mn Ka and Fe Ka) intensity distributions of Gel obtained after 16 days from the sample preparation are shown in Fig. 4(a). The horizontal axis in Fig. 4(a) represents the distance from the gel boundary in Gel (X), and its value is positive at the bottom of Gel. The vertical axis represents the relative ED-XRF intensity. These ED-XRF distributions, which
provide a good approximation of the Mn and Fe distributions, are compared with the positions of the precipitation bands shown in the corresponding image at the top of the panel.

The image and ED-XRF distributions shown in Fig. 4(a) are basically similar to the results reported in previous studies. This image confirms that Mn-Fe PBAs form discrete precipitation bands in agarose gel, although their number is minimal, and their bandwidths are broad. In accordance with this observation, the ED-XRF distributions in Fig. 4(a) exhibit a rich structure. As also found in previous studies, the peak positions of the Mn and Fe Kα distributions coincided well with one another, suggesting that these peaks could be attributed to the presence of Mn-Fe PBAs. It is worth noting that both ED-XRF distributions fluctuated not only at the periodic band zone (X > 15 mm) but also in the turbid zone (0 < X < 15 mm), suggesting that the Mn-Fe PBA concentration fluctuated significantly over a wide region in agarose gel, including the turbid zone.

The inset in Fig. 4(b) shows raw Mn Kβ spectra (where the vertical axis represents count rates) obtained at the turbid zone (X = 3 mm) and the periodic band zone (X = 20 mm). These measuring points are marked by arrows in the top image in Fig. 4(a). For comparison, the spectrum of the 0.50 M MnSO₄ solution is also shown in the inset of Fig. 4(b). The Mn Kβ₁,3 peak intensity at 3 mm is approximately 4-fold higher than the intensity at 20 mm, which shows good correspondence with the ED-XRF results in Fig. 4(a). The Mn Kβ₁,3 peak intensity of the 0.50 M solution is 1.2-fold higher than the 3 mm intensity. Meanwhile, even for the 0.50 M solution, the peak intensity is considerably low (~1.5 /s). Such weak signals might disappoint researchers expecting HR-XRF to be a potential experimental probe in laboratory. Nevertheless, the normalized Mn Kβ spectra in the main panel of Fig. 4(b) could offer interesting information about Mn local structure in Gel as follows.

Comparison of the normalized Mn Kβ spectra of Gel at X = 3 mm and 20 mm reveal agreement between the two spectra within experimental uncertainties (e.g., the difference of the
Kβ_{1,3} peak energies is \(~0.07\text{ eV} < 0.13\text{ eV}\), indicating that local structures around Mn atoms in the Mn-Fe PBA precipitation bands are mostly position-independent.

Our previous study suggested that in the turbid zone of Mn-Fe PBAs in agarose gels, the Mn atoms are surrounded by one O atom (Mn–O length: 0.220 nm) and five N atoms (Mn–O length: 0.221 nm) with pseudo octahedral configuration.\(^{30}\) The good agreement of the two Gel spectra in Fig. 4(b) strongly suggests that the aforementioned local geometry around Mn atoms essentially holds in the periodic precipitation band zone in Gel as well. The uniformity observed in Gel (the agarose gel sample) can be useful in its future applications as radioactive Cs adsorbents,\(^{30}\) and interestingly, such uniformity has not observed in water-glass gel samples studied previously.\(^{25,26}\)

Fig. 4(b) also shows small but distinguishable shifts of Mn Kβ peaks (\(~0.3\text{ eV in } Kβ_{1,3} \text{ peak and } ~1\text{ eV in } Kβ^{'}, \text{peak}\)) between Gel and the 0.50 M solution. Because the bond lengths of Mn–N (0.221 nm) and Mn–O (0.220 nm) in Gel are almost the same as the Mn–O length in the solution (0.220 nm), the observed difference can be attributed to the ligand difference (i.e., one O and five N atoms in Gel versus six O atoms in the 0.50 M solution). Thus, Fig. 4(b) suggests that the high-resolution Mn Kβ spectra are applicable to determine the N to O ratio in the Mn-Fe PBAs. This potential applicability is important, because (1) Mn-Fe PBAs have, more or less, some defects due to the absence of the Fe(CN)_6 moiety (causing a low number of N atoms around Mn atoms) and the presence of connecting H_2O molecules (causing a high number of O atoms around Mn atoms), and (2) the number of such defects (hence, the ratio of N-to-O around the Mn atoms in Mn-Fe PBAs) strongly influence their magnetic\(^{31}\) and electronic properties.\(^{43}\) Note also that the suggested relation between the Mn Kβ shifts and the N-to-O ratio around the Mn atoms provides a significant basis for future applications of high-resolution Mn Kβ spectroscopy to \textit{in-situ} observation of the Mn states in Mn-Fe PBA precipitation bands in gels, which was mentioned in the introduction.
Conclusion

By using a laboratory-use HR-XRF setup with ~2.6 eV instrumental resolution, comprised of a cylindrically bent monochromator and a spherically-bent analyzer, the small shifts in Mn Kβ₁,3 and Mn Kβ´ peaks with changes in octahedral coordination around Mn atoms in Gel and the 0.50 M Mn²⁺ solution are substantially measurable. Thus, the chemical effects on Mn Kβ spectra of gels and solutions are measurable in the laboratory and are expected to be potentially usable for studying ligand characteristics around Mn atoms in the above media, thus making them useful for the defect analysis of Mn-Fe PBAs in gels/solutions.

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Figure Captions

Fig. 1 Schematic representation of a laboratory-use setup to measure highly resolved Mn Kβ spectra.

Fig. 2 (a) High-resolution Mn Kβ spectra of MnO powder. Black squares (Lab) and red line (SPring-8) show the spectrum acquired by the laboratory-use setup and the spectrum taken at SPring-8, respectively. (b) High-resolution Mn Kβ spectra of Mn, MnO, and KMnO4 powders. In both panels, the peak energy of the Mn powder is set to zero, and each peak height is normalized to one.

Fig. 3 High-resolution Mn Kβ spectra of MnSO4·H2O powder and 0.50 M MnSO4 solution. The peak energy of the Mn powder is set to zero, and each peak height is normalized to one.

Fig. 4 (a) Mn and Fe Kα intensity distribution of Gel. Captured image is displayed at the top of this figure to facilitate a comparison of the Mn and Fe Kα distributions with the position of the precipitation bands. The positions at which the high-resolution Mn Kβ spectra (“3 mm” and “20 mm”) were recorded are also shown by arrows. (b) The high-resolution Mn Kβ spectra acquired at these positions. For comparison, the spectrum of 0.50 M MnSO4 solution is also shown. The peak energy of the Mn powder is set to zero. Each peak height is normalized to one, and the count rates of these samples are plotted in the inset.
Fig. 1
Fig. 2
Fig. 3
Fig. 4