Ethanol withdrawal-induced adaptations in prefrontal corticotropin releasing factor receptor 1-expressing neurons regulate anxiety and conditioned rewarding effects of ethanol

Reesha R. Patel¹, Sarah A. Wolfe¹, Vittoria Borgonetti¹,², Pauravi J. Gandhi¹, Larry Rodriguez¹, Angela E. Snyder¹, Shannon D’Ambrosio¹, Michal Bajo¹, Alain Domissy,² Steven Head¹, Candice Contet²,³, R. Dayne Mayfield³,⁴, Amanda J. Roberts¹ and Marisa Roberto¹

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Preclinical studies also implicate the mPFC in anxiety-like behaviors; therefore, mPFCCRF1+ neurons are poised to promote relapse to ethanol use during abstinence. Consequently, mPFCCRF1+ neurons as a critical site of enduring adaptations that may contribute to the persistent vulnerability to ethanol misuse in abstinence, and CSF1 as a novel target for therapeutic intervention for withdrawal-related negative affect.

INTRODUCTION

Alcohol use disorder (AUD) is characterized by the loss of control over ethanol intake, negative emotional states in the absence of ethanol, and a compulsion to seek and consume ethanol, which is thought to heavily involve the prefrontal cortex. Individuals with an AUD have reduced prefrontal cortex volumes [1–5], and hypofunctionality of the medial prefrontal cortex (mPFC) contributes to a loss of control over limiting intake in humans with an AUD [6]. Preclinical studies also implicate the mPFC in anxiety-like behaviors and excessive ethanol drinking [7, 8]. Identifying chronic ethanol-induced adaptations that persist into withdrawal and drive aberrant behavior will provide insight into neuronal mechanisms for more efficacious therapeutic intervention, which are currently limited for AUD.

Ethanol activates neuroendocrine stress systems, which become dysregulated following chronic exposure contributing to AUD [9, 10]. Corticotropin-releasing factor (CRF) is a major brain stress signal and plays a critical role in AUD [11–13]. CRF binds the cognate CRF receptor 1 (CRF1), CRF receptor 2 (CRF2), as well as CRF binding protein (CRF-BP) [14]. While the CRF system has been extensively studied in limbic brain regions, much less is known about its functional role in prefrontal circuits. The CRF-CRF1 system is prominently expressed in the mPFC [15, 16], while CRF2 is limited [17, 18]. Specifically, CRF and CRF1 are predominantly expressed in GABAergic and glutamatergic neurons, respectively [19–21]. Functionally, injection of CRF or CRF1 antagonists in the mPFC is anxiogenic and anxiolytic, respectively [22–25], deletion of forebrain CRF1 reduces anxiety [26], mPFC CRF-CRF1 underlies stress-induced executive dysfunction [27, 28], mPFC CRF signaling impairs working memory [29, 30], and mPFC CRF interneurons regulate behavioral selection during stress [31]. Notably, withdrawal from chronic alcohol recruits mPFC GABAergic CRF neurons [32], and binge alcohol consumption decreases mPFC CRF-BP [33], which is also predominantly expressed in mPFC GABAergic neurons [34]. Taken together, CRF1-expressing mPFC neurons (mPFCCRF1+) have the molecular components to respond to neurochemical stress signals, that are vulnerable to repeated ethanol exposure, and are also wired to mediate anxiety-like behaviors; therefore, mPFCCRF1+ neurons are poised to promote relapse to ethanol use during abstinence.
Here, we tested the hypothesis that mPFC<sup>CRF<sub>1</sub></sup> neurons comprise a distinct population that undergoes specific neuroa- daptations induced by chronic ethanol and withdrawal that underlie aberrant emotional processing and ethanol drinking. Indeed, we found that mPFC<sup>CRF<sub>1</sub></sup> neurons display reduced excitability, as well as glutamate transmission, mediated partly by basolateral amygdala (BLA) afferents. Moreover, we identified a neuroimmune mechanism, via colony-stimulating factor 1 (CSF1), underlying the observed mPFC<sup>CRF<sub>1</sub></sup> adaptations in glutamate transmission and sufficient to induce anxiety-like behavior, which may increase relapse susceptibility. These data highlight the potential of the neuroimmune mediator, CSF1, as a promising, novel target of therapeutic intervention for AUD.

METHODS AND MATERIALS

For full details see Supplementary Materials.

Animals

Adult (>10 weeks old) male and female CRF1::GFP and CRF1::Cre mice were used [35–38]. Sample sizes for each experiment are listed in Supplementary Table 1. All procedures were approved by Scripps Institutional Animal Care and Use Committee and were consistent with the National Institutes of Health Guide for the Care and Use of Laboratory Animals.

Chronic intermittent ethanol inhalation

To induce ethanol dependence, male mice were exposed to chronic intermittent ethanol (CIE) inhalation as previously described [37]. The average blood ethanol level achieved during CIE was 183.5 mg/dl. Cages were randomly assigned to naïve-control, dependent, and withdrawal groups.

In situ hybridization

In situ hybridization, image acquisition, and quantification were performed as previously described [40]. The following probes from ACD Biotechne were used: negative control (320751), Crhr1 (418011), Gad2 (439371), and Slc17a7 (416631). Note, negligible Crhr1 fluorescence was observed in the mouse septum (see Supplementary Fig. 1; modified from Wolfe et al. [35]), a region of low CRF1 expression [18], confirming the validity of the Crhr1 signal.

Immunohistochemistry

Immunohistochemistry, imaging, and quantification were performed as previously described [40]. Primary antibodies included rabbit anti-CSF1 (ab233387; Abcam), rabbit anti-Cre (69050; Novagen), or chicken anti-eGFP (ab13970; Abcam). Secondary antibodies included Alexa Fluor 488 donkey anti-rabbit (Jackson ImmunoResearch, 711-545-152), Alexa Fluor 555 donkey anti-rabbit (A22794, Invitrogen), or Alexa Fluor 488 donkey anti-chicken (703-545-155, Jackson ImmunoResearch).

Whole-cell patch-clamp electrophysiology and optogenetics

Whole-cell voltage-clamp and current-clamp recordings were collected, in an unblinded fashion, from pyramidal neurons, morphologically confirmed visually and with a cell capacitance criterion of >80 pF, and analyzed as previously described [40, 41]. A K-glutonate internal solution was used to record spontaneous excitatory postsynaptic currents (sEPSCs) in artificial cerebrospinal fluid (ACSF), miniature excitatory postsynaptic currents (mEPSCs) in the presence of 30 µM bicuculline (BIC; Tocris) and 0.5 µM tetrodotoxin (TTX; Sigma Aldrich), and excitability in ACSF.

For optogenetic experiments, channelrhodopsin-2 (ChR2)-photocur- rents were measured using wide-field illumination. Mono-synaptic connectivity was measured in ACSF containing 30 µM BIC, 0.5 µM TTX, and 100 µM 4-aminopyridine (4-AP). AMPA and NMDA currents were recorded from a holding potential of −80 mV and −40 mV, respectively, using an Cs+-methanesulfonate internal solution. Data analysis was conducted in a blind fashion until the final step of grouping cells and statistics.

Site-specific viral injection surgery

Viral injections were performed as previously described [42]. Cages were randomly assigned into control and treatment groups for each experiment.

RESULTS

mPFC<sup>CRF<sub>1</sub></sup> neurons comprise a distinct glutamatergic population that regulates anxiety and ethanol reinforcement

To determine the identity and electrophysiological properties of mPFC<sup>CRF<sub>1</sub></sup> neurons, we used male, CRF1::GFP reporter mice for in situ hybridization and ex vivo whole-cell patch-clamp electrophysiology. mPFC<sup>CRF<sub>1</sub></sup> neurons are densely distributed in layer 2/3 of the prelimbic (PrL) subdivision (Fig. 1A). Nuclei expressing Crhr1 mRNA predominantly co-express Slc17a7 compared to Gad2 mRNA, suggesting that mPFC<sup>CRF<sub>1</sub></sup> neurons are primarily glutamatergic (Fig. 1B). mPFC<sup>CRF<sub>1</sub></sup> pyramidal neurons showed a distinct electrophysiological signature compared to neighboring mPFC prL CRF1 non-expressing pyramidal neurons (mPFC<sup>CRF<sub>1</sub></sup>−) including reduced excitability, less voltage sag, greater initial action potential amplitude, and a more depolarized resting membrane potential (Fig. 1C, Supplementary Fig. 2). Moreover, mPFC<sup>CRF<sub>1</sub></sup> neurons had reduced spontaneous excitatory post-synaptic current (sEPSC) frequency and amplitude, suggesting less glutamatergic input and post-synaptic transmission in this population (Fig. 1D–F). No effect on sEPSC kinetics were observed (data not shown). In addition, we found that acute CRF application (200 nM) decreases excitability of mPFC<sup>CRF<sub>1</sub></sup> and mPFC<sup>CRF<sub>1</sub></sup>− neurons (Supplementary Fig. 3 and Supplementary Table 3). These data suggest that mPFC<sup>CRF<sub>1</sub></sup> neurons comprise a unique glutamatergic population with distinct electrophysiological properties.

To assess the behavioral relevance of this population, we selectively ablated mPFC<sup>CRF<sub>1</sub></sup> neurons in male, CRF1::Cre mice and examined the behavioral consequence (Fig. 1G–K, Supplementary Fig. 4). We measured anxiety-like behavior using the novelty suppressed feeding test [45]. Ablation of mPFC<sup>CRF<sub>1</sub></sup>− neurons decreased latency to feed in the arena, suggesting a decrease in...
Fig. 1 mPFC\textsuperscript{CRF1}\textsuperscript{+} neurons comprise a distinct glutamatergic population that regulates anxiety and conditioned rewarding effects of ethanol. A: Coronal mouse brain atlas \cite{80} image of medial prefrontal cortex (mPFC), corresponding 4X magnification, brightfield and GFP fluorescence images of boxed region depicting the mPFC and 40X magnification, brightfield and GFP fluorescence images depicting neighboring GFP positive (green asterisk; mPFC\textsuperscript{CRF1}\textsuperscript{−}) and negative (white asterisk; mPFC\textsuperscript{CRF1}\textsuperscript{+}) mPFC prelimbic (PrL) neurons. B: Representative traces of spontaneous excitatory post-synaptic currents (sEPSCs) recorded with a hold potential of \(-70\) mV. 

Representative in situ hybridization images of colocalization of Crhr1 (red), Slc17a7 (green), Gad2 (yellow), and DAPI (blue) in the mPFC PrL layer 2/3 subregion and summary graph of percent nuclei expressing. Scale bar = 10 µM. 4 images from \(N = 2\) male mice. C: Representative voltage traces of action potential firing elicited by increasing current injections (−120, 40, 60 and 200 pA, 1 s) in mPFC PrL CRF1 non-expressing (mPFC\textsuperscript{CRF1}\textsuperscript{−}) and CRF1 expressing (mPFC\textsuperscript{CRF1}\textsuperscript{+}) pyramidal neurons. Average number of action potentials elicited by increasing current injections in mPFC\textsuperscript{CRF1}\textsuperscript{−} and mPFC\textsuperscript{CRF1}\textsuperscript{+} neurons with significant effects of current injection \(F(29,1972) = 70.78, p < 0.0001; \text{group} F(1,68) = 4.26, p = 0.04; \) and interaction between current injection and group \(F(29,1972) = 1.99, p = 0.001\) by two-way ANOVA and post hoc significance #\(p < 0.05\) represented in panel. 

Percent time spent in the ethanol-paired context during the pre-conditioning phase (open bars) and following ethanol conditioning (filled bars) in the ethanol place conditioning test in control (black) and mPFC\textsuperscript{CRF1}\textsuperscript{+} ablated (red) mice with significant main effects of condition \(F(1,23) = 70.78, p < 0.0001\) and group \(F(1,23) = 4.66, p = 0.04\) by two-way ANOVA, and post hoc significance \(p < 0.05\) represented in panel. J: Percent time spent in the ethanol-paired context during the pre-conditioning phase (open bars) and following ethanol conditioning (filled bars) in the ethanol place conditioning test in control (black) and mPFC\textsuperscript{CRF1}\textsuperscript{+} ablated (red) mice with a significant interaction effect \(F(1,24) = 11.05, p = 0.002\) by two-way ANOVA, and post hoc significance \(p < 0.05\) represented in panel. K: Pre-conditioning normalized percent time spent in the ethanol-paired side with significant \(p < 0.05\) effects by one-sample t-test and unpaired t-test. \(N = 12\) to 15 male mice.

Anxiety-like behavior (Fig. 1I). No difference in latency to feed in the home cage was observed, suggesting a similar motivation for food. We also measured ethanol place conditioning to assess conditioned ethanol reward. Control mice preferred the ethanol-associated context, highlighting the conditioned reinforcing properties of ethanol (Fig. 1J, K). Remarkably, ablation of mPFC\textsuperscript{CRF1}\textsuperscript{+} neurons induced a conditioned place aversion to the ethanol-associated context, suggesting conditioned aversion to ethanol. Together, these findings demonstrate that mPFC\textsuperscript{CRF1}\textsuperscript{+} neurons regulate anxiety-like behaviors and conditioned rewarding effects of ethanol, supporting the potential role of this population in aberrant emotional processing in abstinence.

Withdrawal selectively decreases mPFC\textsuperscript{CRF1}\textsuperscript{+} excitability

Since mPFC\textsuperscript{CRF1}\textsuperscript{+} neurons regulate negative affective behaviors and reinforcing properties of ethanol that can contribute to relapse, we asked if this population undergoes specific neuroadaptations following withdrawal from chronic ethanol. We exposed male CRF1:GFP mice to chronic intermittent ethanol (CIE) inhalation to generate dependent and withdrawn mice that were 5–8 days into forced abstinence. We found that mPFC\textsuperscript{CRF1}\textsuperscript{+} pyramidal neuron excitability is increased in dependent and withdrawn mice (Fig. 2A, B). Electrophysiological properties are summarized in Supplementary Table 2. Overall, mPFC\textsuperscript{CRF1}\textsuperscript{+} neuronal excitability is increased by ethanol dependence and withdrawal.

We then selectively recorded from neighboring mPFC\textsuperscript{CRF1}\textsuperscript{+} pyramidal neurons and found no significant effect of ethanol dependence on neuronal excitability (Fig. 2C, D). Notably, withdrawal markedly decreased mPFC\textsuperscript{CRF1}\textsuperscript{+} excitability compared to naïve, pointing to the selective sensitivity of mPFC\textsuperscript{CRF1}\textsuperscript{+} neurons to withdrawal. Interestingly, there was a significant reduction in
Withdrawal selectively decreases post-synaptic glutamate transmission in mPFC^CRF1+ neurons, which is partly mediated by the basolateral amygdala

Given that CRF signaling remodels spine density contributing to anxiety-like behaviors [46, 47], we next asked whether chronic ethanol alters glutamatergic signaling in mPFC^CRF1+ pyramidal neurons in male CRF:GFP mice. We found that mPFC^CRF1+ neurons displayed a trend toward and a significant increase in sEPSCs frequency in dependent and withdrawn mice, respectively, indicative of increased presynaptic glutamate release (Fig. 3A, B). mPFC^CRF1+ neurons also displayed increased sEPSC amplitude and current kinetics in dependent as well as withdrawn mice, suggesting enhanced postsynaptic glutamatergic signaling (Fig. 3C, Supplementary Fig. 5). Similar to mPFC^CRF1+ neurons, dependence increased sEPSC frequency, amplitude, and kinetics in mPFC^CRF1+ pyramidal neurons compared to naive (Fig. 3D–F, Supplementary Fig. 5). However, the increase in sEPSC frequency was lost in withdrawn mice and significant decreases in sEPSC amplitude and current kinetics were observed, suggesting a decrease in postsynaptic glutamate transmission in mPFC^CRF1+ neurons in withdrawal. Together, these findings further highlight the unique sensitivity of mPFC^CRF1+ neurons to withdrawal and point to reduced glutamatergic signaling in this population as a potential mechanism underlying heightened anxiety-like behavior in withdrawal.

The BLA and mPFC show excitatory coupling and enhanced activity during anxiety, and activation of BLA inputs to the mPFC induces anxiety-like behaviors [48, 49], suggesting the BLA-mPFC circuit is a candidate in underlying aberrant emotional processing following withdrawal. To measure BLA-mPFC^CRF1+ connectivity, we expressed ChR2-mCherry in the BLA of CRF:GFP mice (Fig. 3G) and measured light-evoked post-synaptic potentials in mPFCCRF1+ neurons. Blue light reliably elicited time-locked photocurrents in mCherry-expressing BLA neurons, demonstrating precise control of BLA neuronal activity (Fig. 3H). To assess presynaptic BLA-mediated glutamate release onto mPFC^CRF1+ neurons, we measured responses in mPFC^CRF1+ neurons to optical paired pulse stimulation of BLA terminals in the mPFC (Fig. 3I, J). The paired pulse ratio was significantly increased in dependent and withdrawn mice, suggesting a decrease in glutamate release onto mPFC^CRF1+ neurons following chronic ethanol. To assess postsynaptic alterations in BLA-mPFC^CRF1+ connectivity, we measured optically evoked mono-synaptic AMPA and NMDA currents in mPFCCRF1+ neurons (Fig. 3K, L). NMDA amplitude was significantly decreased, and correspondingly the AMPA/NMDA ratio was significantly increased in withdrawn mice compared to naive, indicating a selective decrease in postsynaptic glutamate transmission in the BLA-mPFC^CRF1+ pathway in withdrawn mice. Together, these findings demonstrate that the upstream BLA, which directly innervates mPFC^CRF1+ neurons, is dysregulated by dependence and that this dysregulation persists into withdrawal. Dysregulation of the BLA temporally precedes the selective dysregulation of mPFC^CRF1+ neurons in withdrawal, suggesting that the BLA partly drives aberrant activity of mPFC^CRF1+ neurons in withdrawal.
Since CRF-CRF1 signaling can potently modulate neuronal activity and synaptic plasticity [11, 50, 51], we also examined the impact of CRF on the BLA-mPFC CRF1+ pathway. CRF (200 nM) potentiated BLA-mediated AMPA currents in mPFC CRF1+ neurons in naïve mice (Fig. 3M, N), which was abolished by dependence, but recovers in withdrawal, suggesting a transient neuroadaptation to CRF signaling in mPFC CRF1+ neurons. CRF did not significantly alter BLA-mediated NMDA currents or the paired pulse ratio (Supplementary Fig. 6). Of note, CRF does not significantly alter global miniature EPSCs (mEPSCs) in mPFC CRF1+
Fig. 3 Withdrawal selectively decreases post-synaptic glutamate transmission in mPFC^{CRF1+} neurons, which is partly mediated by the basolateral amygdala. A Representative traces of average spontaneous excitatory post-synaptic currents (sEPSC) recorded with a holding potential of −70 mV in mPFC^{CRF1+} neurons from naive, dependent, and withdrawn mice. B, C Average sEPSC frequency and amplitude in mPFC^{CRF1+} neurons from naive (white), dependent (black), and withdrawn (grey) mice. n = 12–22 cells from N = 6–10 male mice; *p < 0.05, **p < 0.01 by one-way ANOVA and post hoc multiple comparisons compared to naive. D Representative traces of average sEPSCs recorded in a holding potential of −70 mV in mPFC^{CRF1+} neurons from naive, dependent, and withdrawn mice. E, F Average sEPSC frequency and amplitude in mPFC^{CRF1+} neurons from naive (green), dependent (white), and withdrawn (grey) mice. n = 10–21 cells from N = 6–10 male mice; *p < 0.05, **p < 0.01 by one-way ANOVA and post hoc multiple comparisons compared to naive. G Schematic of viral strategy for ex vivo optogenetic circuit dissection of glutamate transmission in the basolateral amygdala (BLA) to mPFC^{CRF1+} pathway. Representative 4X magnification, brightfield and mCherry fluorescence images of injection site expression of channelrhodopsin-2 (ChR2)-mCherry in the BLA (right, top) and corresponding ChR2-mCherry BLA terminal in the mPFC (right, bottom). H ChR2-mediated photocurrents in mPFC expressing BLA neurons elicited by pulsed trains of blue light stimulation from a holding potential of −70 mV. I Representative BLA-mediated AMPA currents in mPFC^{CRF1+} neurons (scale bar = 300 ms). J Representative traces of average sEPSCs recorded with a holding potential of 200 ms from a holding potential of −80 mV. K Average sEPSC frequency (i.e., amplitude of the second response normalized to that of the first) in mPFC^{CRF1+} neurons from naive, dependent, and withdrawn mice with a significant F(2,57) = 4.27, p = 0.01 effect by one-way ANOVA, and post hoc significance *p < 0.05 compared to naive is represented in panel. n = 12–27 cells from N = 5–6 male mice. L Average ± SEM BLA-mediated NMDA (top row) and AMPA (bottom row) currents in mPFC^{CRF1+} neurons from naive, dependent, and withdrawn mice with a significant F(2,86) = 5.50, p = 0.001 effect by one-way ANOVA, and post hoc significance *p < 0.05 compared to naive is represented in panel. n = 17–39 cells from N = 5–6 male mice. M Average ± SEM BLA-mediated AMPA currents elicited by blue light pulses during baseline and following 200 nM CRF from a holding potential of −80 mV in mPFC^{CRF1+} neurons from naive, dependent, and withdrawn mice. N Average baseline normalized AMPA amplitude elicited by optical stimulation of BLA terminals with a significant F(2,16) = 3.55, p = 0.05 effect of CRF across groups by one-way ANOVA and post hoc multiple comparisons, and significant *p < 0.05 and **p < 0.01 effects of CRF by one sample t-test. n = 4–11 cells from N = 3–6 male mice per group.

neurons from naive or dependent mice, but selectively potentiates mEPSCs in withdrawal (Supplementary Fig. 7). Together these findings demonstrate that CRF can potentiate BLA-mPFC^{CRF1+} connectivity.

Withdrawal from chronic ethanol alters the transcriptome of mPFC^{CRF1+} neurons – upregulating CSF1

To gain mechanistic insight into the observed electrophysiological neuroadaptations, we sequenced the transcriptome of mPFC^{CRF1+} neurons from naïve and withdrawn mice. Using male CRF1:GFP mice, we sequenced RNA from mPFC^{CRF1+} neurons isolated using fluorescence-activated cell sorting (Fig. 4A). We detected over forty-thousand genes and found 344 significant differentially expressed genes (DEGs) (Fig. 4B). We then identified pathways, hub genes, and functional processes most impacted by withdrawal from chronic ethanol in mPFC^{CRF1+} neurons (Fig. 4C-E). From these analyses and a literature review, we identified an intriguing candidate, colony stimulating factor 1 (CSF1) for further in-depth analysis. CSF1 is highlighted in yellow throughout Fig. 4. We first identified the most significantly impacted pathways based on the DEGs (Fig. 4, Supplementary Table 4). CSF1 is included in the gene list comprising the PI3K-Akt signaling pathway, which was one of the top ten impacted pathways (Fig. 4C inset). In addition, several other signaling, peptide, and immune pathways were identified along with a specific alcoholism pathway, highlighting a central role of mPFC^{CRF1+} neurons in an AUD phenotype (Supplementary Table 4). Network analysis of DEGs revealed the known interactions and relationships between genes (Fig. 4D). Notably, CSF1 is a hub gene, indicated by its centrality in the network showing a high degree of connectivity among the DEGs. Lastly, to identify the functional processes in mPFC^{CRF1+} neurons that are most impacted by withdrawal from chronic ethanol, we used gene ontology (GO) analysis. The top ten GO terms within the three domains including: biological processes, molecular functions, and cellular components that are significantly over-represented in the set of DEGs were identified (Fig. 4E). CSF1 is among the top ten GO terms for biological processes and many of the GO terms for molecular components as well. These data highlight the fundamental changes observed in mPFC^{CRF1+} biological adaptations induced by withdrawal from chronic ethanol in mPFC^{CRF1+} neurons, and recruitment of CSF1 as a candidate gene that could underlie critical neuroadaptations driving aberrant behavior.

CSF1 overexpression in mPFC^{CRF1+} decreases post-synaptic glutamate transmission and is sufficient to increase anxiety-like behavior while abolishing conditioned ethanol reward

Notably, CSF1 is upregulated by mPFC neurons following stress leading to microglia-mediated spine pruning underlying anxiety- and depressive-like behaviors [52]. Therefore, we hypothesized that upregulation of CSF1, as revealed by our transcriptomic analysis, may be a mechanism underlying the reduced post-synaptic glutamate transmission observed selectively in mPFC^{CRF1+} pyramidal neurons following withdrawal (Fig. 3D; Supplementary Fig. 5). To test this, we selectively overexpressed CSF1 in mPFC^{CRF1+} neurons, using female CRF1:Cre mice, and used whole-cell patch-clamp electrophysiology to assess mEPSCs (Fig. 5A; Supplementary Fig. 8). We found that CSF1 overexpression in mPFC^{CRF1+} neurons decreased mEPSC amplitude and decay kinetics compared to control but did not alter mEPSC frequency or rise kinetics (Fig. 5B–E). CSF1 overexpression in mPFC^{CRF1+} neurons did not significantly alter excitability of mPFC^{CRF1+} or mPFC^{CRF2+} neurons (Supplementary Table 5; Supplementary Fig. 9). These data show that CSF1 overexpression in mPFC^{CRF1+} neurons reduce post-synaptic glutamate transmission, identifying a potential mechanism underlying the reduced glutamatergic signaling selectively observed in this population following withdrawal.

We then asked if CSF1 overexpression in mPFC^{CRF1+} neurons is sufficient to alter behavior (Fig. 5F–I; Supplementary Fig. 10). Immunohistochemical staining confirmed mPFC CSF1 overexpression in CSF1-mCherry compared to mCherry control mice (Fig. 5F). We found that CSF1 overexpression significantly increased the latency to feed in an open arena in the novelty-suppressed feeding test, suggesting an increase in anxiety-like behavior (Fig. 5G). CSF1 overexpression in mPFC^{CRF1+} neurons abolished ethanol place preference, suggesting a loss in the reinforcing properties of ethanol (Fig. 5H, I). These findings demonstrate that CSF1 overexpression in mPFC^{CRF1+} neurons is sufficient to induce anxiety-like behavior and abolish the rewarding properties of ethanol, providing mechanistic insight into the regulation of these aberrant behaviors following ethanol withdrawal.
DISCUSSION

This study pinpoints behaviorally relevant molecular-, cell-type-, and circuit-specific adaptations selectively induced by withdrawal from chronic ethanol which underlie negative affective behavior and conditioned ethanol reward. We found that mPFC\textsuperscript{CRF1$^+$} neurons display decreased excitability and glutamatergic transmission in withdrawal, suggesting that they comprise a distinct population highly vulnerable to ethanol withdrawal. Moreover, we found that the BLA in part drives the downstream dysregulation of mPFC\textsuperscript{CRF1$^+$} neurons in withdrawal. Ablation of mPFC\textsuperscript{CRF1$^+$} neurons decreased anxiety-like behavior and induced conditioned aversion to ethanol, supporting a unique role of mPFC\textsuperscript{CRF1$^+$} neurons in AUD-related behaviors. Notably, we found that ethanol withdrawal fundamentally alters the neurobiology of mPFC\textsuperscript{CRF1$^+$} neurons including overexpression of colony stimulating factor 1. Selective overexpression of CSF1 in mPFC\textsuperscript{CRF1$^+$} neurons was sufficient to decrease postsynaptic glutamate transmission and induce behavioral deficits in anxiety and conditioned ethanol.
reward, providing mechanistic insight into the observed withdrawal-associated neuroadaptations in glutamate transmission and aberrant behavior. Taken together, we have identified a distinct mPFC CRF1-expressing subpopulation in the BLA-mPFC circuit that undergoes specific neuroadaptations following ethanol withdrawal, potentially underlying increased vulnerability to relapse during abstinence.

The CRF-CRF1 system is prominently expressed in the mPFC and serves as a potent regulator of neuronal activity, structural and functional plasticity, and emotional and cognitive behaviors. Stress-induced CRF-CRF1 signaling results in mPFC and hippocampal dendritic atrophy underlying anxiety and memory deficits [46, 47]. Consistent with CRF’s role in synaptic remodeling [53], we found that mPFC CRF1+ neurons have reduced basal postsynaptic glutamatergic transmission, possibly due to ongoing basal CRF-CRF1 signaling regulating spine density. CRF also increases mPFC neuronal excitability [19, 20]. Accordingly, mPFC CRF1+ neurons displayed a depolarized resting membrane potential, potentially due to basal CRF-CRF1 signaling induced persistent sodium or Ih current [19, 54–57]. Though, mPFC CRF1+ neurons exhibited overall reduced excitability, voltage sag, and glutamate transmission under basal conditions. Moreover, CRF modulates mPFC glutamate transmission. We found that CRF enhances BLA-mPFC CRF1+ connectivity. Notably, we didn’t observe a significant impact of CRF on global mEPSCs in mPFC CRF1+ neurons, suggesting that CRF may impact select circuits comprising mPFC CRF1+ neurons. Indeed, CRF-induced mPFC excitatory post-synaptic currents requires BLA input [58], suggesting that CRF may bias toward a greater influence of the BLA on mPFC activity. Given CRF’s neuromodulatory role, it is predictable that mPFC CRF-CRF1 signaling impacts behavior. Our findings further support a role for mPFC CRF-CRF1 neurons in anxiety-like behavior and conditioned rewarding effects of ethanol, although it is possible that compensatory changes may contribute to these behavioral effects. The pleiotropic effects of mPFC CRF-CRF1 signaling on neuronal physiology and AUD-related behaviors positions mPFC CRF1+ neurons to mediate aberrant behaviors underlying AUD.

While the CRF-CRF1 system has been strongly implicated in preclinical models of AUD particularly in limbic brain regions [11], considerably less is known about its role in the mPFC. Here we hypothesized that mPFC CRF1+ neurons are uniquely sensitive to chronic ethanol and undergo selective dysregulation underlying AUD-related behaviors. Our previous work demonstrated differential sensitivity in GABAergic signaling in CeACRF1+ neurons compared to CeACRF1− neurons to acute and chronic ethanol [36, 37]. In this study, mPFC CRF1+ neurons exhibited decreases in excitability and glutamate transmission selectively in withdrawal. In contrast, mPFC CRF1− displayed increases in excitability and glutamate transmission following dependence and withdrawal. This finding is consistent with previous work showing an increase in mPFC neuronal excitability following chronic intermittent ethanol exposure [59, 60], which may contribute to CIE-induced cognitive deficits [60–63]. This highlights the selective sensitivity of mPFC CRF1+ to withdrawal and its potential in uniquely contributing to AUD-related behaviors. Decreased mPFC CRF1+ excitability may be driven by changes in glutamatergic drive as well as in intrinsic properties. Indeed, transcriptomic analysis identified an increase in SCNS5A, encoding the sodium channel Nav1.5, which may underlie the reduced action potential amplitude of mPFC CRF1+ neurons in withdrawal. Notably, withdrawal leads to the activation of mPFC CRF GABAergic neurons [32], which presumably innervate mPFC CRF1+ neurons and may
also contribute to the reduced mPFCC\textsubscript{CF1+} excitability in withdrawal, as we observed CRF decreases mPFCC\textsubscript{CF1+} excitability. Further identification of the circuitry comprising mPFCC\textsubscript{CF1+} neurons will provide a framework to understand the role of this population in AUD.

The BLA sends dense glutamatergic projections to the mPFC and contributes to anxiety-related behaviors and addiction [64]. We found that BLA-mPFCC\textsubscript{CF1+} synapses undergo adaptations, suggesting a reduced probability of glutamate release from BLA terminals following chronic ethanol which persists into withdrawal. Post-synaptic BLA connections to mPFCC\textsubscript{CF1+} neurons exclusively undergo adaptations in withdrawal, specifically a decrease in NMDA current leading to increased AMPA/NMDA ratio. The temporal sequence of these pathological synaptic adaptations suggests that the BLA partly drives the aberrant glutamatergic transmission observed in mPFCC\textsubscript{CF1+} neurons following withdrawal. mPFC NMDA hypofunction is associated with cognitive impairment [65, 66] and increased opiate reward sensitivity, which is dependent on the BLA [67]. While not directly tested here, it is possible that reduced mPFCC\textsubscript{CF1+} NMDA current following withdrawal may increase the rewarding properties of ethanol, given the role of mPFCC\textsubscript{CF1+} neurons in conditioned ethanol reward. A global understanding of the neurobiological changes will provide insight into concurrent mechanisms driving aberrant behavior.

To examine global molecular changes associated with withdrawal selectively in mPFCC\textsubscript{CF1+} neurons, we used cell-type specific transcriptomic analysis. Notably, overrepresented transcriptomic changes in this single, specific mPFCC\textsubscript{CF1+} population were sufficient to identify a significant perturbation in biological pathways associated with ‘alcoholism’, supporting the role of this population in AUD. Interestingly, the PI3K-Akt pathway, a highly impacted pathway by withdrawal, has previously been identified as a mPFCC\textsubscript{CF1+} molecular mechanism underlying ethanol intake and anxiety in withdrawal [68]. In addition, our bioinformatic analysis identified CSF1, a neuroimmune mediator important for the development and maintenance of microglia, as a hub gene [69]. Indeed, stress-induced increases in mPFC CSF1 expression leads to microglia-mediated synaptic pruning, and CSF1 knockdown was sufficient to reverse stress-induced anxiety and depressive behaviors [52]. We found that CSF1 overexpression in mPFCC\textsubscript{CF1+} neurons was sufficient to induce decreased postsynaptic glutamate transmission in this population, supporting a microglia-mediated mechanism underlying synaptic adaptations. In line with this, withdrawal from chronic ethanol increases mPFC microglial reactivity [70, 71]. Indeed, mPFCC\textsubscript{CF1+} CSF1 overexpression increases anxiety-like behavior, highlighting CSF1 overexpression as a critical neuroadaptation that may contribute to negative affect in withdrawal. Note, CSF1 overexpression also induced a loss of the conditioned rewarding effects of ethanol. While this may appear counterintuitive with the anxiogenic effects of CSF1 overexpression, this finding is consistent with a shift in negative reinforcement-driven drinking, rather than the rewarding effects of ethanol itself, following chronic ethanol [11, 72, 73]. Thus, CSF1 overexpression in mPFCC\textsubscript{CF1+} neurons induced by withdrawal from chronic ethanol may drive escalated drinking by increasing negative affective states, contributing to negative reinforcement mechanisms motivating drinking in AUD.

Our findings suggest that mPFCC\textsubscript{CF1+} regulate anxiety-like behavior, due to the observed decreases in anxiety-like behavior following ablation of this population. These findings suggest that synaptic adaptations in mPFCC\textsubscript{CF1+} neurons may contribute to heightened anxiety following withdrawal. Indeed, we demonstrated that withdrawal-induced molecular adaptations in CSF1 expression in mPFCC\textsubscript{CF1+} neurons are sufficient to heighten anxiety and recapitulate the reduced postsynaptic glutamate transmission seen in this population in withdrawal. Together, mPFCC\textsubscript{CF1+} neurons regulate withdrawal-related ethanol drinking and anxiety-like behaviors and undergo unique adaptations following withdrawal from chronic ethanol exposure that may increase vulnerability to relapse.

Of note, while both male and female mice were used, all electrophysiological data and mPFCC\textsubscript{CF1+} ablation behavioral experiment were conducted in male mice, while CSF1 overexpression behavioral experiments were conducted in female mice. We found that mPFCC\textsubscript{CF1+} ablation and CSF1 overexpression studies both altered behavior in the novelty-suppressed feeding test, suggesting this population regulates anxiety-like behavior in both male and female mice. Although, there are sex differences in mPFC circuits and anxiety-like behavior [20, 74, 75], which may contribute to the findings in this study. Future studies are warranted to explore potential sex differences in the role of mPFCC\textsubscript{CF1+} population.

While preclinical studies suggest that the CRF-CRF1 system is critical in AUD-related behavior, it has not yet been successful in a clinical setting [76, 77]. Therefore, it is imperative to find novel targets that may be translated in the clinic. Here, we rationalized the mPFCC\textsubscript{CF1+} neurons are responsive to the brain stress signal CRF and used transcriptomics to identify other potential molecular targets for therapeutic intervention. Our findings suggest that CSF1, which has been a target of interest for cancers, Alzheimer’s disease, and other disorders [78, 79], may be a novel target to alleviate negative affect during withdrawal from chronic alcohol exposure contributing to relapse-like behavior.

REFERENCES

1. Jernigan TL, Butters N, DiTraglia G, Schafer K, Smith T, Irwin M, et al. Reduced cerebral grey matter observed in alcoholics using magnetic resonance imaging. Alcohol Clin. Exp. Res. 1991;15:418–27. https://doi.org/10.1111/j.1530-0277.1991.tb00540.x
2. de la Monte SM. Disproportionate atrophy of cerebral white matter in chronic alcoholics. Arch. Neurol. 1988;45:990–2. https://doi.org/10.1001/archneur.1988.00200700513
3. Pfeifferbaum A, Sullivan EV, Mathalon DH, Lim KO. Frontal lobe volume loss observed with magnetic resonance imaging in older chronic alcoholics. Alcohol Clin. Exp. Res. 1997;21:521–9. https://doi.org/10.1111/j.1530-0277.1997.tb03798.x
4. Harris GJ, Jaffin SK, Hodge SM, Kennedy D, Caviness VS, Marinkovic K, et al. Frontal white matter and cingulum diffusion tensor imaging deficits in alcoholism. Alcohol Clin. Exp. Res. 2008;32:1001–13. https://doi.org/10.1111/j.1530-0277.2008.00661.x
5. Kri JJ, Gundlach AL, Dodd PR, Johnston GA, Harper CG. Cortical dihydropyridine binding sites are unaltered in human alcoholic brain. Ann. Neurol. 1989;26:395–7. https://doi.org/10.1002/ana.1041260315
6. Goldstein RZ, Volkow ND. Dysfunction of the prefrontal cortex in addiction: neuroimaging findings and clinical implications. Nat. Rev. Neurosci. 2011;12:652–69. https://doi.org/10.1038/nrn3119
7. Hare BD, Duman RS. Prefrontal cortex circuits in depression and anxiety: distribution of discrete neuronal populations and target regions. Mol. Psychiatry. 2020;25:2742–58. https://doi.org/10.1038/s41380-020-0685-9
8. Klenowski PM. Emerging role for the medial prefrontal cortex in alcohol-seeking behaviors. Addict. Behav. 2018;77:102–6. https://doi.org/10.1016/j.addbeh.2017.09.024
9. Blaine SK, Sinha R. Alcohol, stress, and glucocorticoids: From risk to dependence and relapse in alcohol use disorders. Neuropsychopharmacology. 2017;12:136–47. https://doi.org/10.1038/nj.neuropsychopharm.2017.01.037
10. Cui C, Noronha A, Warren RR, Koob GF, Sinha R, Thakkar M, et al. Brain pathways to recovery from alcohol dependence. Alcohol. 2015;49:435–52. https://doi.org/10.1016/j.alcohol.2015.04.006
11. Koob GF, Buck CL, Cohen A, Edwards S, Park PE, Schlosburg JE, et al. Addiction as a stress surfet disease. Neuropsychopharmacology. 2014;67:370–82. https://doi.org/10.1016/j.neuropharm.2013.05.024
12. Roberto M, Spierling SR, Kirson D, Zornella EP. Corticotropin-Releasing Factor (CRF) and Addictive Behaviors. Int. Rev. Neurobiol. 2017;136:51–51. https://doi.org/10.1016/bs.irn.2017.06.004
13. Roberto M, Cruz MT, Gilpin NW, Sabino V, Schweitzer P, Bajo M, et al. Corticotropin-releasing factor-induced amygdala gamma-aminobutyric Acid release plays a key role in alcohol dependence. Biol. Psychiatry. 2010;67:831–9. https://doi.org/10.1016/j.biopsych.2009.11.007
34. Wolfe, SA, Sidhu, H, Patel, RR, Kreifeldt, M, D’Ambrosio, SR, Contet, C, et al. Molecular, morphological, and functional characterization of corticotropin-releasing factor receptor 1-expressing neurons in the central nucleus of the amygdala. eNeuro 6, https://doi.org/10.1523/EJNNEURO.0087-19.2019 (2019).

35. Herman MA, Contet, C, Justice NJ, Vale, W, Roberto M. Novel subunit-specific tonic GABA currents and differential effects of ethanol in the central amygdala of CRF receptor-1 reporter mice. Neurosci. 2013;33:3284–98. https://doi.org/10.1523/JNEUROSCI.2490-12.2013

36. Herman MA, Contet, C, Roberto M. A functional switch in tonic GABA currents alters the output of central Amygdala corticotropin releasing factor receptor-1 neurons following chronic ethanol exposure. J Neurosci. 2016;36:10729–41. https://doi.org/10.1523/JNEUROSCI.1267-16.2016

37. Justice NJ, Yuan ZF, Vale W. Type 1 corticotropin-releasing factor receptor expression reported in BAC transgenic mice: implications for reconciling ligand-receptor mismatch in the central corticotropin-releasing factor system. J Compr Neurol. 2008;511:479–96. https://doi.org/10.1002/cne.21848

38. Agoglia, AE, Zhu, M, Ying, R, Sidhu, H, Natividad, LA, Wolfe, SA, et al. Corticotropin-releasing factor receptor-1 neurons in the lateral amygdala display selective sensitivity to acute and chronic ethanol exposure. eNeuro 7, https://doi.org/10.1523/EJNNEURO.0429-19.2020 (2020).

39. Warden AS, Wolfe SA, Khom S, Varodayan FP, Patel RR, Steinman MQ, et al. Microglia control escalation of drinking in alcohol-dependent mice: Genomic and synaptic drivers. Biol Psychiatry. 2020. https://doi.org/10.1016/j.biopsych.2020.05.011

40. Patel RR, Khom S, Steinman MQ, Varodayan FP, Kiooses WB, Hedges DM, et al. IL-1beta expression is increased and regulates GABA transmission following chronic ethanol in mouse central amygdala. Brain Behav. Immun. 2019;75:2028–19. https://doi.org/10.1016/j.bbi.2018.10.009

41. Patel, RR, Wolfe, SA, Bajo, M, Abeynaike, S, Pahng, A, Borghetti, V, et al. IL-10 normalizes aberrant amygdala GABA transmission and reverses anxiety-like behavior and dependence-induced escalation of alcohol intake. Prog. Neurobiol. 2019,10152. https://doi.org/10.1016/j.progneurobiol.2020.101952 (2020).

42. Tsumeto R, Lee S, Szucs A, Chubukov P, Sokolova I, Blanchard J, et al. J. Diverse reprogrammng codes for neuronal identity. Nature. 2018;557:375–80. https://doi.org/10.1038/s41586-018-0103-5

43. Love MI, Huber W, and Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. Genome Biol. 15, https://doi.org/10.1186/s13059-014-0550-8 (2014).

44. Daluza SC, Hen R. Recent advances in animal models of chronic antidepressive effects: The novelty-induced hypophagia test. Neurosci Biobehav Rev. 2005;29:771–83. https://doi.org/10.1016/j.neubiorev.2005.03.017

45. Dong H, Keegan JM, Hong E, Gallardo C, Montalvo-Ortiz J, Wang B, et al. Corticotrophic releasing factor receptor 1 antagonists prevent chronic stress-induced behavioral changes and synapse loss in aged rats. Psychoneuroendocrinology. 2018;90:92–101. https://doi.org/10.1016/j.psyneuen.2018.02.013

46. Chen Y, Rex CS, Rice CJ, Dubé CM, Gall CM, Lynch G, et al. Correlated memory defects and hippocampal dendritic spine loss after acute stress involve corticotropin-releasing hormone signaling. Proc. Natl. Acad. Sci. USA 2010;107:13123–8. https://doi.org/10.1073/pnas.1003825107

47. Felix-Ortiz AC, Burgos-Robles A, Bhagat ND, Leppä CA, Tyé KM. Bidirectional modulation of anxiety-related and social behaviors by amygdala projections to the medial prefrontal cortex. J Neurosci. 2016;36:219–209. https://doi.org/10.1523/NEUROSCI.1323-15.2016

48. Marcus DJ, Bede G, Gaulden AD, Ryan JD, Kondiev V, Winters ND, et al. Endocannabinoid signaling collapse mediates stress-induced amygdalo-cortical strengthening. Neuron. 2020;105:762–761066. https://doi.org/10.1016/j.neuron.2019.12.024

49. Varodayan FP, Correa D, Kirson D, Khom S, Olesta CS, Luu G, et al. CRF modulates glutamate transmission in the central amygdala of naive and ethanol-dependent rats. Neuropearmacology. 2017;125:418–28. https://doi.org/10.1016/j.neuropharm.2017.08.009

50. Natividad LA, Buczynski MW, Herman MA, Kirson D, Olesta CS, Irinia C, et al. Constitutive increases in amygdalar corticotropin-releasing factor and fatty acid amide hydrolase drive an anxiolysis phenotype. Biol. Psychology. 2017;125:500–10. https://doi.org/10.1016/j.biopsycho.2017.01.005

51. Wohleb ES, Tewriggerill R, Duman CH, Duman RS. Stress-induced neuronal colony formation. Front. Brain Sci. 2019;8:124. https://doi.org/10.3389/fnbeh.2019.00124

52. Regen L, Baram TZ. Corticotropin releasing factor in neuroplasticity. Front. Neuropsychopharmacology. 2014;35:171–9. https://doi.org/10.3389/fnbeh.2013.00101

53. Libster AM, Title B, Yarom Y. Corticotropin-releasing factor increases Purkinje neuron excitability by modulating sodium, potassium, and Ih currents. J Neurophysiol. 2015;114:3339–50. https://doi.org/10.1152/jn.00745.2015

54. Giesbrecht CJ, Mackay JP, Silveira HB, Urban JH, Colmers WF. Countervailing modulation of Ih by neuropeptide Y and corticotrophin-releasing factor in basolateral
