Constitutive expression of murine c-FLIP<sub>R</sub> causes autoimmunity in aged mice

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Death receptor-mediated apoptosis is a key mechanism for the control of immune responses and dysregulation of this pathway may lead to autoimmunity. Cellular FLICE-inhibitory proteins (c-FLIPs) are known as inhibitors of death receptor-mediated apoptosis. The only short murine c-FLIP splice variant is c-FLIP<sub>Raji</sub> (c-FLIP<sub>R</sub>). To investigate the functional role of c-FLIP<sub>R</sub> in the immune system, we used the vavFLIP<sub>R</sub> mouse model constitutively expressing murine c-FLIP<sub>R</sub> in all hematopoietic compartments. Lymphocytes from these mice are protected against CD95-mediated apoptosis and activation-induced cell death. Young vavFLIP<sub>R</sub> mice display normal lymphocyte compartments, but the lymphocyte populations alter with age. We identified reduced levels of T cells and slightly higher levels of B cells in 1-year-old vavFLIP<sub>R</sub> mice compared with wild-type (WT) littermates. Moreover, both B and T cells from aged vavFLIP<sub>R</sub> animals show activated phenotypes. Sera from 1-year-old WT and transgenic animals were analysed for anti-nuclear antibodies. Notably, elevated titres of these autoantibodies were detected in vavFLIP<sub>R</sub> sera. Furthermore, tissue damage in kidneys and lungs from aged vavFLIP<sub>R</sub> animals was observed, indicating that vavFLIP<sub>R</sub> mice develop a systemic lupus erythematosus-like phenotype with age. Taken together, these data suggest that c-FLIP<sub>R</sub> is an important modulator of apoptosis and enforced expression leads to autoimmunity.

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It is crucial that excessive lymphocytes are deleted by apoptosis after efficient antigen clearance to maintain cell homeostasis. Moreover, insufficient apoptosis of autoreactive immune cells may result in autoimmune diseases. The extrinsic apoptosis pathway is triggered by ligand binding of death receptors, such as CD95 (Fas/APO-1), and regulates the removal of unwanted lymphocytes. Elimination of T cells is achieved by activation-induced cell death (AICD), which is triggered upon restimulation of T cells without co-stimulation. Notably, the CD95 death receptor pathway was shown to be important for AICD. Furthermore, dysregulation of the CD95 signalling pathway was reported to result in severely altered lymphoproliferation (Lpr) and autoimmunity in both lpr and generalised lymphoproliferative disorder (gld) mice, which have mutated CD95 receptor or ligand, respectively, as well as in patients suffering from the homologous human disease autoimmune lymphoproliferative syndrome. The c-FLIPs inhibit apoptosis by competing with procaspase-8 for further substrates, eventually leading to apoptosis. The c-FLIPs inhibit apoptosis by competing with procaspase-8 for binding sites at the DISC and additionally interfere with caspase-3 processing. So far, three c-FLIP isoforms expressed on the protein level have been reported: c-FLIP<sub>Long</sub> (c-FLIP<sub>L</sub>), c-FLIP<sub>Short</sub> (c-FLIP<sub>S</sub>) and c-FLIP<sub>Raji</sub> (c-FLIP<sub>R</sub>). The long isoform resembles caspase-8, but lacks catalytic function because of substitution of amino-acid residues critical for enzymatic activity. The two short isoforms are distinguished by a functional nucleotide polymorphism and mainly consist of the death effector domains with unique C-terminal tails. The structure of the murine c-FLIP protein, Raji isoform; CNS, central nervous system; DC, dendritic cell; DISC, death-inducing signalling complex; DN, double negative; EAE, experimental autoimmune encephalomyelitis; FADD, Fas-associated death domain-containing protein; Gld, generalised lymphoproliferative disorder; IFN-γ, interferon-γ; Lpr, lymphoproliferation; MOG, myelin oligodendrocyte glycoprotein; MS, multiple sclerosis; SLE, systemic lupus erythematosus; WT, wild type

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Cflar (caspase-8 and FADD-like apoptosis regulator) gene differs from the human gene locus, in that only c-FLIP L and c-FLIP R can be expressed. Although the short c-FLIP isoforms have been described as anti-apoptotic, c-FLIP L was reported to act both pro- and anti-apoptotic depending on expression level and strength of receptor stimulation. In contrast, mice overexpressing human c-FLIP S did not develop autoimmune disease. Not much is known about the physiological function of c-FLIP R. To investigate the functional role of c-FLIP R in the immune system, we used a mouse model, called vavFLIP R, with constitutive expression of murine c-FLIP R in all hematopoietic compartments. We previously reported that these transgenic mice have a better clearance of the Gram-positive bacteria *Listeria monocytogenes* compared with infected wild-type (WT) mice. Here we show that aged mice constitutively expressing murine c-FLIP R have altered lymphocyte populations with higher levels of activated B and T cells compared with WT littermates. Moreover, vavFLIP R animals spontaneously develop autoimmunity with age, showing features of systemic lupus erythematosus (SLE).

**Results**

**vavFLIP R mice do not recapitulate the lpr/gld phenotype.** Insufficient cell death can lead to autoimmunity as a result of altered immune cell populations. Lpr and gld mice, with mutations in the CD95 receptor and ligand, respectively, develop Lpr and autoimmune disease. Moreover, these mice accumulate unusual CD4−CD8− double negative (DN) B220+/β2 T cells. c-FLIP R inhibits apoptosis by competing with caspase-8 for binding sites at the DISC. The mouse model vavFLIP R, in which murine c-FLIP R is under control of the vav-promoter to ensure expression in all hematopoietic compartments, was used to study the functional role of murine c-FLIP R in the immune system. As thymocytes as well as B and T cells from vavFLIP R mice were protected against CD95-induced apoptosis, we investigated if the impaired lymphocyte apoptosis in vavFLIP R mice would lead to elevated numbers of lymphocytes and altered lymphocyte populations with age. No Lpr was identified in 12–14 months old vavFLIP R animals. In fact, the number of cells in the peripheral lymph nodes (pLNs) from vavFLIP R mice was reduced compared with WT littermates (Figure 1a). Furthermore, pLN cells of WT and vavFLIP R mice were analysed for DN B220+ cells compared with 3 months old MRL/lpr mice as controls. The characteristic DN B220+ population was identified in MRL/lpr mice, but was not observed in either WT or vavFLIP R animals (Figure 1b). The lack of DN B220+ T cells in vavFLIP R mice is similar to the phenotype of c-FLIP L transgenic mice and mice with transgenic expression of human c-FLIP S. Thus, vavFLIP R mice do not develop an lpr/gld-like phenotype.

**Altered lymphocyte populations in aged vavFLIP R mice.** Young vavFLIP R mice displayed normal lymphocyte populations. We analysed aged WT and vavFLIP R mice to investigate if the constitutive expression of c-FLIP R alters lymphocyte populations over time. Dendritic cells (DCs), macrophages and granulocytes have been reported to be regulated through death receptors and c-FLIP proteins. We therefore analysed the frequencies of CD11c+ DCs,
CD49b<sup>+</sup> natural killer (NK) cells, F4/80<sup>+</sup> macrophages and Gr1<sup>+</sup> granulocytes in 1-year-old mice. The populations of CD11c<sup>+</sup>, CD49b<sup>+</sup>, F4/80<sup>+</sup> and Gr1<sup>+</sup> cells from WT and vavFLIP<sub>R</sub> were comparable (Supplementary Table 1). Flow cytometry analyses of lymphocyte sub-populations in spleen and pLNs from 12 to 14 months old mice revealed reduced frequencies of vavFLIP<sub>R</sub> CD3<sup>+</sup> T cells compared with WT CD3<sup>+</sup> T cells (Figures 2a and b), consistent with reduced absolute CD3<sup>+</sup> T-cell number in pLNs from vavFLIP<sub>R</sub> animals (Figure 2c). Furthermore, an increased percentage of vavFLIP<sub>R</sub> CD19<sup>+</sup> B cells in pLNs compared with WT was observed (Figures 2a and d). Frequencies and absolute numbers of CD19<sup>+</sup> B cells were not significantly altered in the spleen (Figure 2e). Neither did we detect alterations in germinal centre formation in aged transgenic mice (data not shown). The CD4<sup>+</sup>/CD8<sup>+</sup> cell profiles within the CD3<sup>+</sup> T-cell compartment were comparable between WT and vavFLIP<sub>R</sub> mice (Supplementary Figure 1). Nevertheless, lower frequencies of splenic CD8<sup>+</sup> T cells were observed as well as reduced absolute cell numbers of CD8<sup>+</sup> cells in pLNs (Supplementary Figure 1b). As we previously reported that thymocyte development in vavFLIP<sub>R</sub> mice is normal, we suggest that c-FLIP<sub>R</sub> expression regulates homeostasis of peripheral CD8<sup>+</sup> T cells. Taken together, the B- and T-cell populations are altered in aged vavFLIP<sub>R</sub> mice, whereas other haematopoietic cells are unaffected.

**Activated phenotypes of T and B cells from vavFLIP<sub>R</sub> mice.** The activation status of T cells in young vavFLIP<sub>R</sub> mice was comparable to WT littermates. As AICD is impaired in T cells from vavFLIP<sub>R</sub> mice, we investigated if the activation status changed with age. We observed higher percentages of vavFLIP<sub>R</sub> CD4<sup>+</sup> cells expressing the activation marker CD25 in comparison with WT CD4<sup>+</sup> cells (Figures 3a and b), with absolute cell numbers being similar between 12 and 14 months old WT and vavFLIP<sub>R</sub> mice (Figure 3c). As CD25 is not only a marker for activated T cells, but is also highly expressed on regulatory T (T<sub>reg</sub>) cells, we analysed the expression of the master transcription factor of T<sub>reg</sub> cells, Foxp3. Indeed, most of the CD25<sup>+</sup>CD4<sup>+</sup>T cells co-expressed Foxp3 indicating that 1-year-old vavFLIP<sub>R</sub> mice have higher levels of T<sub>reg</sub> cells in both pLNs (Figure 3d) and spleen (data not shown) compared with WT littermates. Interestingly, we also observed an increase in CD25<sup>+</sup> T cells that were negative for Foxp3 suggesting that

![Figure 2](#)

**Figure 2** Altered lymphocyte populations in 12–14 months old vavFLIP<sub>R</sub> mice. (a) Representative dot plots of CD3- and CD19-stained WT and vavFLIP<sub>R</sub> pLN cells. Frequency and absolute cell number of CD3<sup>+</sup> cells (b and c) and CD19<sup>+</sup> cells (d and e) in freshly isolated pLNs and spleens from WT (n = 13) and vavFLIP<sub>R</sub> (n = 15) littermates. Individual mice are represented as separate symbols. Horizontal lines show the mean pooled from five independent experiments; error bars display the S.E.M. Statistical analyses were performed with two-tailed nonparametric Mann–Whitney U-tests.
conventional T cells may indeed be more activated (Figure 3d). Therefore, the memory/naive T-cell profile was analysed in 12–14 months old WT and vavFLIP<sub>R</sub> animals. Strikingly, elevated frequencies of CD4<sup>+</sup> antigen-experienced cells within the CD3<sup>+</sup> compartment were identified in both pLNs and spleen from vavFLIP<sub>R</sub> mice (Figures 4a and b). We also observed reduced numbers of CD3<sup>+</sup>CD62L<sup>+</sup> naive T cells in vavFLIP<sub>R</sub> animals (Supplementary Figure 2a). Notably, both within the CD4<sup>+</sup> helper T-cell population and the CD8<sup>+</sup> cytotoxic T-cell population from vavFLIP<sub>R</sub> pLNs and spleens, higher levels of antigen-experienced (CD44<sup>+</sup>) cells and lower levels of naive (CD62L<sup>+</sup>) cells were identified compared with WT (Figures 4c and d). The absolute cell numbers of CD62L<sup>+</sup> cells were reduced for both CD4<sup>+</sup> and CD8<sup>+</sup> T-cell populations (Supplementary Figures 2b and c). These data indicate that T cells from vavFLIP<sub>R</sub> mice are highly activated in comparison with WT T cells.

Similar to T cells, B cells from vavFLIP<sub>R</sub> mice are protected against apoptosis. The activation status of B cells was examined by flow cytometry. CD40, CD80 and CD95 had similar expression levels in vavFLIP<sub>R</sub> and WT animals (data not shown), whereas analysis of the activation markers CD69, CD86 and major histocompatibility complex class II molecule

**Figure 3** Increased levels of CD25<sup>+</sup> T cells and T<sub>reg</sub> cells in 1-year-old vavFLIP<sub>R</sub> animals. (a) Representative dot plots of WT (upper panel) and vavFLIP<sub>R</sub> (lower panel) CD3<sup>+</sup> cells in pLNs. Frequency (b) and absolute cell number (c) of CD4<sup>+</sup>CD25<sup>+</sup> cells in WT (n = 13) and vavFLIP<sub>R</sub> (n = 15) pLN and spleens. Symbols represent individual mice. Horizontal lines display the mean ± S.E.M. Statistical analyses were performed with two-tailed nonparametric Mann–Whitney U-tests. (d) Dot plots showing CD25<sup>+</sup> and Foxp3<sup>+</sup> T cells within the CD4<sup>+</sup> compartment representative for three WT (left) and four vavFLIP<sub>R</sub> (right) mice.
I-A/I-E revealed an increased expression of these markers in vavFLIP<sub>R</sub> B cells compared with WT B cells (Figure 5). This indicates that B cells from 12 to 14 months old vavFLIP<sub>R</sub> mice are more activated in comparison with B cells from WT animals at 1 year of age. The activated phenotypes of B and T cells identified in 1-year-old vavFLIP<sub>R</sub> mice are consistent with a previous study, which reported that B and T cells from c-FLIP<sub>R</sub> transgenic mice on a Balb/c background were highly activated.25

Disease progression in MOG-EAE is comparable between WT and vavFLIP<sub>R</sub> mice. As constitutive expression of c-FLIP<sub>R</sub> alters lymphocyte populations and activation status with age, we investigated if this has functional relevance in T-cell-mediated autoimmune disease. Experimental autoimmune encephalomyelitis (EAE) is a widely accepted animal model for studying the human demyelinating inflammatory disorder multiple sclerosis (MS).36 T<sub>H</sub>1 and T<sub>H</sub>17 effector T cells have been shown to

Figure 4 T cells from vavFLIP<sub>R</sub> mice at 12–14 months of age are highly activated. (a) Representative dot plots of pLN cells (left panel) and splenocytes (right panel) from WT (upper panel) and vavFLIP<sub>R</sub> animals (lower panel). Frequencies of CD4<sup>+</sup>, CD4<sup>+</sup>CD62L<sup>+<sup> and CD62L<sup>-</sup> cells within CD3<sup>+</sup> cells (b), CD4<sup>+</sup> cells (c) and CD8<sup>+</sup> cells (d) from pLN and spleen. (b-d) Symbols represent individual WT (b: n = 13, c and d: n = 6) and vavFLIP<sub>R</sub> (b: n = 15, c and d: n = 6) mice. Horizontal lines represent the mean pooled from five independent experiments or the mean of one experiment representative for two independent experiments; error bars display S.E.M. Statistical analyses were performed with two-tailed nonparametric Mann–Whitney U-tests.
contribute to the pathology in EAE\textsuperscript{37} and c-FLIP\textsubscript{R} transgenic mice were reported to be resistant to myelin oligodendrocyte glycoprotein (MOG)-EAE because of enhanced T\textsubscript{H}2, and thus diminished T\textsubscript{H}1, effector responses.\textsuperscript{38} To assess if c-FLIP\textsubscript{R} influences the differentiation of T\textsubscript{H}0, T\textsubscript{H}1, T\textsubscript{H}2 or T\textsubscript{H}17 cells, we cultured naive WT and vavFLIP\textsubscript{R} T cells under T\textsubscript{H}1, T\textsubscript{H}2 or T\textsubscript{H}17 polarising conditions in vitro. The frequencies of interferon-\gamma (IFN-\gamma)-producing T\textsubscript{H}1 cells were comparable between WT and vavFLIP\textsubscript{R} differentiated T cells (Figures 6a and b). Interestingly, frequencies in IL-17A-producing T\textsubscript{H}17 cells and IL-4 mRNA induction in T\textsubscript{H}2 cells were lower in vavFLIP\textsubscript{R} T cells cultured under the respective polarising conditions (Figures 6a, b and c).

WT and vavFLIP\textsubscript{R} mice were immunised with the MOG\textsubscript{35-55} peptide in complete Freund's adjuvant in conjunction with pertussis toxin to investigate if constitutive murine c-FLIP\textsubscript{R} expression in T cells, macrophages and DCs influences EAE disease. However, both WT and vavFLIP\textsubscript{R} animals developed disease with neurological defects resulting in paralysis of the tail and muscle weakness with comparable severity of disease. Only a slightly slower disease progression was observed in vavFLIP\textsubscript{R} mice (Figure 6d). To investigate whether T-cell effector responses were affected by the transgenic expression of c-FLIP\textsubscript{R} in vivo, we analysed cytokine production in MOG-peptide restimulated T cells from MOG-immunised mice. We observed slightly reduced frequencies of IFN-\gamma\textsuperscript{+} and IL-17A\textsuperscript{+} cells from vavFLIP\textsubscript{R} pLNs in comparison with WT 11 days after immunisation (Figure 6e). This effect was levelled out at higher MOG peptide concentrations on day 14 (Figure 6e). The frequencies of IL-4-producing T cells in pLNs from vavFLIP\textsubscript{R} and WT mice were comparable (Figure 6e). Similar results were obtained when T cells were restimulated in a polyclonal manner, that is, with anti-CD3 and anti-CD28 (Supplementary Figure 3). In summary, our data indicate a minor defect in vavFLIP\textsubscript{R} effector T-cell responses that results in a slight delay in EAE disease progression in vavFLIP\textsubscript{R} mice.

Elevated ANA-titres and tissue damage of kidneys and lungs in aged vavFLIP\textsubscript{R} mice. B-cell-mediated autoimmunity is characterised by autoantibodies targeting the body's own tissues. As vavFLIP\textsubscript{R} mice had slightly higher B-cell levels with activated phenotype, we analysed sera from 1-year-old WT and vavFLIP\textsubscript{R} mice for anti-nuclear antibodies (ANAs) using an indirect immunofluorescence assay with HEp-2 cells as source of nuclear antigens. Negative fluorescent pattern was most often observed when sera from WT mice were incubated on HEp-2 cells (Figure 7a), whereas homogenous fluorescent pattern was detected for the majority of sera from vavFLIP\textsubscript{R} mice even at high serum dilution (Figure 7b). Hence, significantly more

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\caption{vavFLIP\textsubscript{R} B cells from 12 to 14 months old mice show an activated phenotype. (a-c) pLN cells and splenocytes from WT and vavFLIP\textsubscript{R} mice (n = 6) were isolated and analysed by flow cytometry. The activation status of CD19\textsuperscript{+} B cells was assayed by the markers CD69 (a), CD86 (b) and I-A/I-E (c). Representative histograms with fluorescent minus one (FMO) as control (left panel) and diagrams displaying the mean fluorescent intensity (MFI; right panel) ± S.E.M. are shown. Statistical analyses were performed with two-tailed nonparametric Mann–Whitney U-tests.}
\end{figure}
autoantibodies were identified in sera from vavFLIP<sup>R</sup> mice compared with WT mice (Figure 7c).

Multiple organs can be affected in systemic autoimmune diseases. We therefore examined several organs from 1-year-old WT and vavFLIP<sup>R</sup> animals by histology. No differences between WT and vavFLIP<sup>R</sup> mice could be observed in brains, spleens and pancreas stained with haematoxylin and eosin (H&E) and in spleens stained for B cells and macrophages (data not shown). Kidneys from aged mice were stained with periodic acid–schiff (PAS) followed by histological scoring. The majority of glomeruli in kidneys from WT mice were normal (Figure 7d). In contrast, thickening of the Bowman’s capsule and protein deposition in glomeruli were observed in kidneys from vavFLIP<sup>R</sup> mice, indicating nephropathy (Figure 7e). Glomerular deposition of PAS-positive material is common in aging mice. Significantly more tissue damage was detected in vavFLIP<sup>R</sup> kidneys with 63.6% of the kidney sections displaying a...
histological score of three or higher, implying mild-to-
moderate alterations (Figure 7f). Thus, vavFLIPR mice have a greater degree of nephropathy compared with WT animals at 12–14 months of age.

The histological analyses of lungs from aged mice revealed that three of six vavFLIPR lungs stained with H&E displayed minor effect on the progression of EAE. Yu et al.

Taken together, the elevated levels of ANAs and tissue damage in kidneys and lungs are symptoms similar to human SLE. Hence, the kidney damage may be related to the ANAs and represent a mild form of SLE nephropathy.

Discussion

CD95-mediated apoptosis has a crucial role in controlling immune responses and preventing autoimmune diseases. c-FLIP proteins protect cells against CD95-induced apoptosis at the level of the DISC. Although the isoforms c-FLIPR and c-FLIPS have been thoroughly characterised, not much is known about the functional role of c-FLIPR in the immune system. In this study, we report that constitutive expression of murine c-FLIPR results in altered lymphocyte populations and lupus-like symptoms with age.

Lpr and gld mice with mutations in the CD95 receptor and ligand, respectively, develop lymphoproliferative disease and autoimmunity with accumulation of unusual DN B220+ T cells. c-FLIP proteins inhibit the same apoptosis pathway by preventing caspase-8 cleavage at the DISC. Indeed, T-cell-specific c-FLIP transgenic mice were reported to develop autoimmunity when bred on Balb/c, but not C57BL/6, background. However, neither altered Lpr nor accumulation of DN B220+ T cells was identified in up to 1-year-old vavFLIPR mice, consistent with transgenic mice with T-cell-specific expression of murine c-FLIPR or human c-FLIPS in the C57BL/6 background. Hence, neither constitutive expression nor overexpression of c-FLIP proteins, at least in the C57BL/6 background, is sufficient for recapitulation of the lpr/gld phenotype.

The autoimmune inflammation in EAE is mainly caused by autoreactive T11 and T17 effector cells. Lpr mice showed a prolonged and enhanced inflammation in the central nervous system (CNS) upon EAE induction. Hence, CD95/CD95L interactions are of importance to delete disease-initiating, autoreactive T cells in the CNS in the course of EAE. Both long and short c-FLIP isoforms have been reported to be upregulated in activated T cells from the cerebral spinal fluid in patients with active MS. Interestingly, transgenic mice overexpressing human c-FLIPR were protected against EAE because of enhanced T12 effector responses and consequently reduced T11 effector responses. Transgenic expression of murine c-FLIPR did not alter the cytokine production of T cells in a similar way, which may be one reason why the enforced expression of c-FLIPR only had a minor effect on the progression of EAE. Yu et al. reported high expression levels of c-FLIPS in T17 cells, which protected these cells against AICD. This was proposed to be a mechanism for the high pathogenicity of T17 cells in autoimmune diseases. Expression levels of c-FLIPR in T17 cells was not investigated in this study, nevertheless, the increased expression of c-FLIPR in this cell population may override the effect of the constitutive c-FLIPR expression in vavFLIPR mice in the course of EAE. The impaired ability of naive vavFLIPR T cells to differentiate into the T17 subset in vitro and the minor reduction of effector T-cell responses observed in MOG-immunised vavFLIPR mice could possibly explain the slightly slower disease progression in vavFLIPR mice compared with WT littermates.
B- and T-cell numbers and distribution were normal in young vavFLIP\textsubscript{R} mice\textsuperscript{28} and no differences in DCs, macrophages, granulocytes or NK cells could be identified even in mice at 1 year of age. Interestingly, aged vavFLIP\textsubscript{R} mice had altered lymphocyte populations with lower levels of T cells and higher frequencies of B cells compared with WT littermates. Moreover, reduced levels of CD8\textsuperscript{+} cells were observed, which is consistent with a report of Wu \textit{et al.}\textsuperscript{48} where reduced peripheral CD8\textsuperscript{+} cell numbers were described in mice with transgenic expression of the viral FLIP MC159 under control of the CD2 enhancer cassette. Furthermore, alterations in the ratio of antigen experienced to naive T cells were identified, most probably because of defective AICD. Human c-FLIP\textsc{S} is upregulated in the activation and expansion phase of an immune response, but is thereafter downregulated to enable AICD and elimination of effector T cells.\textsuperscript{49} We previously described a similar kinetic for murine c-FLIP\textsc{R}, which explains the defect in AICD upon constitutive c-FLIP\textsc{R} expression.\textsuperscript{28} In this study, increased frequencies of activated T helper cells and antigen-experienced T cells as well as lower frequencies and numbers of naive T cells in vavFLIP\textsubscript{R} mice compared with WT mice were observed. This is consistent with a previous report where an increased memory T-cell pool was identified in c-FLIP\textsc{S} transgenic mice compared with WT animals after immunisation with the superantigen staphylococcal enterotoxin B.\textsuperscript{27} Similarly to the activated T-cell phenotype, vavFLIP\textsubscript{R} B cells expressed activation markers to a higher extent than WT B cells. Strikingly, histological analyses of kidneys and lungs from 12 to 14 months old mice revealed a higher degree of nephropathy in vavFLIP\textsubscript{R} mice and a larger proportion of vavFLIP\textsubscript{R} animals displayed focal interstitial pneumonia and an increased amount of BALT compared with WT littermates. Moreover, elevated levels of anti-nuclear autoantibodies were identified in vavFLIP\textsubscript{R} sera, indicating that vavFLIP\textsubscript{R} mice develop a lupus-like phenotype with age. Hence, our findings imply that the altered lymphocyte populations with activated phenotypes in aged vavFLIP\textsubscript{R} mice are of functional relevance. Most probably the autoimmune disease observed in vavFLIP\textsubscript{R} animals is caused by a combination of autoreactive B and T cells, with persisting CD4\textsuperscript{+} helper T cells, which may prime B cells to produce autoantibodies, and increased levels of B cells. In support of this notion, lupus-like disease was described in c-FLIP\textsc{L} transgenic mice on the Balb/c background and the development of autoimmunity in this study was shown to require CD4\textsuperscript{+} T cells, which were proposed to result from impaired thymic selection.\textsuperscript{25} Notably, elevated gene expression of CLFAR in CD33\textsuperscript{+} myeloid cells, CD4\textsuperscript{+} T cells and CD19\textsuperscript{+} B cells was reported in SLE patients.\textsuperscript{50} Furthermore, Xu \textit{et al.}\textsuperscript{51} observed that human lupus T cells did not downregulate c-FLIP\textsc{S} after initial activation, which suggests that c-FLIP proteins are of importance in the development of SLE disease. The higher level of T\textsubscript{reg} cells in aged vavFLIP\textsubscript{R} mice is presumably a compensatory mechanism because of the observed autoimmunity. Imbalance of T\textsubscript{reg} cells has been described in various autoimmune diseases.\textsuperscript{52} Reduced levels or conflicting data have been reported in patients with, for example, juvenile idiopathic arthritis, autoimmune liver disease and myasthenia gravis.\textsuperscript{53} Consistent with our findings, increased levels of T\textsubscript{reg} cells were identified in patients with

**Figure 8** Histological alterations in lungs of aged vavFLIP\textsubscript{R} mice. Lungs were prepared from 1-year-old WT (\(n = 5\)) and vavFLIP\textsubscript{R} (\(n = 6\)) mice and paraffin embedded. Sections were stained with H&E, the B-cell marker B220, the T-cell marker CD3 and the macrophage marker MAC2, respectively. Deposition of IgG-containing immune complexes was detected with goat-anti-mouse antibodies. Representative tissue sections are shown. Arrows indicate increased BALT and interstitial pneumonia, respectively. Scale bars represent 200 \(\mu\text{m}\) for H&E section and 50 \(\mu\text{m}\) for all other sections.
primary Sjögren’s syndrome and in experimental arthritis in mice. The expanded Treg cell population observed in 1-year-old vavFLIPR animals most probably dampens the severity of the SLE-like phenotype.

We previously reported that vavFLIPR mice are better protected against Listeria monocytogenes infection compared with WT littermates. Hence, c-FLIPR expression is beneficial in an acute infection, whereas we in this study demonstrate that chronic expression results in autoimmunity. To conclude, aged vavFLIPR mice have altered lymphocyte populations with increased levels of antigen-experienced T cells and higher activation status of both B and T cells. Moreover, these animals develop lupus-like symptoms with age. Our results show that c-FLIPR has an important role in the control of autoimmunity.

Materials and Methods
Mice. vavFLIPR mice were previously described. WT littermates were used as control animals. All animals were kept in a specific pathogen-free environment in the animal facility of Helmholtz Centre for Infection Research, Braunschweig. C57BL/6 mice were purchased from Harlan Laboratories (Indianapolis, IN, USA) and Charles River (Wilmington, MA, USA). MLR/er mice were a kind gift from Dr Delphi Neumann, Hannover Medical School, Hannover, Germany. All breeding and experiments were performed in accordance with the guidelines of national and local authorities.

Cell preparation. Lymphoid organs (pLNs, mesenteric lymph nodes, spleen and thymus) were isolated from mice killed via CO2. Organs were homogenised in RPMI 1640 (Gibco – Life technologies), 1% non-essential amino acids (Gibco – Life technologies, Grand Island, NY, USA) supplemented with 10% FCS (PAA, Pasching, Austria), 50 mM EDTA, pH adjusted to 7.3 with NaOH) at room temperature followed by a further washing step. Primary murine cells were cultured in RPMI 1640 (Gibco – Life technologies) by incubation through a 70 μm nylon mesh and washed with PBS. Erythrocytes were removed by 2-min incubation in ACK lysis buffer (0.15 M NH4Cl, 1 mM KHCO3, 0.1 mM EDTA, pH adjusted to 7.3 with NaOH) at room temperature followed by a further washing step. Primary murine cells were cultured in RPMI 1640 (Gibco – Life technologies, Grand Island, NY, USA) supplemented with 10% FCS (PAA, Pasching, Austria), 50 μg/ml of penicillin and streptomycin (Gibco – Life technologies), 1% non-essential amino acids (Gibco – Life technologies), 2 mM L-glutamine (Gibco – Life technologies) and 1 mM sodium pyruvate (Gibco – Life technologies).

In vitro generation of T helper cell subsets. For differentiation of T helper subsets, CD4+ CD62L+CD25− naive T cells were sorted by using a FACS Aria II (BD Biosciences, Heidelberg, Germany) or MoFlo (Beckman and Coulter, Indianapolis, IN, USA). Cells were seeded directly after sorting with 2 × 10^5 cells per well in 96-well plates and activated with plate-bound anti-CD3 and anti-CD28 in the presence of priming cytokines and inhibitory antibodies according to the respective T helper subset: Th0: 2 μg/ml anti-CD3 (145-2C11, BioLegend, San Diego, CA, USA), 2 μg/ml anti-CD28 (37.51, BioLegend), 10 μg/ml anti-IL-4 (11B11, self-purified), and 10 μg/ml anti-IFN-γ (XMG1.2, self-purified); Th1: 2 μg/ml anti-CD3 (145-2C11, BioLegend, San Diego, CA, USA), 2 μg/ml anti-CD28 (37.51, BioLegend, 10 μg/ml anti-IL-2 (JE56-1A12, BioLegend), 10 μg/ml anti-IFN-γ (XMG1.2, self-purified), and 10 μg/ml anti-IL-12 (11B11, self-purified); Th0 restimulated with plate-bound anti-CD3 (2 μg/ml; 145-2C11, BioLegend) and anti-CD28 (2 μg/ml; 37.51, BioLegend) or up to 50 μg/ml MOG35-55 peptide for 24 h with addition of 10 μg/ml Brefeldin A (Sigma Aldrich, Munich, Germany) for 30 min in the dark. Thereafter, slides were washed twice 5 min in PBS followed by incubation with FITC-conjugated donkey anti-mouse IgG (Dianova, Hamburg, Germany) for 30 min in the dark. Subsequently, cells were stained with Live/Dead near IR or blue fluorescent reactive dyes (Life Technologies) by incubation for 30 min in PBS at 4 °C. Thereafter, cells were washed and Fcγ IIIIR receptors were blocked by incubation with anti-CD16/CD32 (2.4G2, BD Biosciences). Subsequently, cells were washed and stained with antibodies in PBS containing 2% BSA for 20 min at 4 °C. The following antibodies were used: CD3-CD345-V500 (17A2), CD3-IFN-γ (14-4-4S, BD Biosciences), CD25-APC (Cytomx1.2, BD Biosciences), PerCP-Cy5.5 (ME-14, BD Biosciences), FITC-PE-Cy7 (H1.2F3), F4/80-PE (BM8) and Gr-1-Pacific Blue (RB6-8C5) (all from eBioscences, San Diego, CA, USA). Samples were analysed by LSRII or LSRFoRr flow cytometers (BD Biosciences). The FlowJo software (TreeStar, Ashland, OR, USA) was used to analyse the data.

Real-time detection PCR. cDNA was used as a template for real-time PCR using SYBR Green (Roche, Mannheim, Germany). Ubiquitin C (UBC) was used as a reference gene for normalisation of gene expression. The primers used in the LightCycler 96 system (Roche) were used as follows: IFN-γ forward (5′-AGAGATCCACAGAGAAGTATGG-3′), IFN-γ reverse (5′-CGAGCAGGCTTGAACACAGC-3′); IL4 forward (5′-CATCGGCACTTACGACGGAC-3′); IL4 reverse (5′-GCAGCTTCATGTCATGTCG-3′).

Experimental autoimmune encephalomyelitis. EAE was induced in 10–12 weeks old WT and vavFLIPR mice by injecting 200 μg MOG35-55 Peptide (MEVGYVRPRPSFVRHLVRNL) together with 4 mg/ml mycobacteria in complete Freund’s adjuvant subcutaneously at four sites. In all, 200 ng pertussis toxin was injected intraperitoneally to open the blood–brain barrier. The pertussis toxin injection was repeated 2 days later. Mice were monitored daily for clinical signs of EAE from days 3 to 4 with 0 – no symptoms, 0.5 – partial limp tail, 1 – limp tail, 2 – delayed rotation from dorsal position, 2.5 – hindleg weakness, 3 – complete hindleg paralysis, 3.5 – starting foreleg weakness, 4 – paralysis of one foreleg, 5 – moribund, death. All mice with a score higher than 3 were killed.

For analysis of cytokine-producing T cells, mice were immunised with MOG35-55 peptide as above, but without pertussis toxin to prevent the migration of primed T cells into the brain. pLNs were isolated from mice killed 11 and 14 days after injection. In all, 5 × 10^5 cells were seeded per well in 96-well plates and either left untreated or restimulated with plate-bound anti-CD3 (2 μg/ml; 145-2C11, BioLegend) and anti-CD28 (2 μg/ml; 37.51, BioLegend) or up to 50 μg/ml MOG35-55 peptide for 24 h with addition of 10 μg/ml Brefeldin A (Sigma Aldrich) for the last 2 h. IFN-γ, IL-4 and IL-17A-producing T cells were analysed by flow cytometry.

Analysis of ANAs. ANAs in sera from 1-year-old WT and vavFLIPR mice were analysed semiquantitatively by incubating HEP-2 cells seeded on microscope slides (Geuder Assays, Dahlewitz, Germany) with sera diluted 1:50–1:1280 for 30 min at room temperature. Slides were washed twice 5 min in PBS followed by incubation with FITC-conjugated donkey ant-mouse IgG (Dianova, Hamburg, Germany) for 30 min in the dark. Thereafter, slides were washed two times 5 min in PBS, transferred to cover slips and sealed. Slides were analysed by fluorescence microscopy where a homogenous pattern was considered as positive for ANAs.

Histology. Kidneys, lungs, liver, spleens, pancreas and brains were isolated from 1-year-old WT and vavFLIPR mice. Organs were fixed in 4% neutral buffered formaldehyde and embedded in paraffin for histological analysis. In all, 5 μm sections were stained with H&E. The degree of focal interstitial pneumonia and BALT was determined in H&E-stained lung sections. Lungs were immunohistochemically stained with B220 (RA3-6B2, BD Biosciences), CD3 (SP7; Neo Markers, Fremont, CA, USA) and MAC3 (M3/38; CEDARLANE, Burlington, ON, Canada) to determine lymphocyte and macrophage infiltration. IgG complexes in lung sections were stained with goat-anti-mouse (TM-060-BN, Thermo Fisher Scientific, Waltham, MA, USA). Kidney sections were stained with PAS staining and the sections were analysed for thickening of the Bowman’s capsule and c-FLIPR in autoimmunity
protein casts in glomeruli. Scoring was performed in a blinded manner for each criterion as follows: 0 – no alteration, 1 – low grade (slightly visible), 2 – moderate (clearly visible), 3 – severe (dominant finding). Scores were added to a total score for each organ.

Statistical analyses. Statistical analyses were performed by nonparametric Mann–Whitney U-test or Fisher’s exact test using GraphPad Prism software (GraphPad Software, La Jolla, CA, USA). Data are presented as the mean with S.E.M. and S.D. as error bars.

Conflict of Interest
The authors declare no conflict of interest.

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