Dynamics of absorption, metabolism, and excretion of 5-aminolevulinic acid in human intestinal Caco-2 cells

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5-Aminolevulinic acid (ALA) is a precursor for the biosynthesis of porphyrins and heme. Although the oral administration of ALA has been widely applied in clinical settings, the dynamics of its absorption, metabolism, and excretion within enterocytes remain unknown. In this study, after enterocytic differentiation, Caco-2 cells were incubated with 200 µM ALA and/or 100 µM sodium ferrous citrate (SFC) for up to 72 h. Both ALA and the combination of ALA and SFC promoted the synthesis of heme, without affecting the expression of genes involved in intestinal iron transport, such as DMT1 and FPN. The enhanced heme synthesis in Caco-2 cells was more pronounced under the effect of the combination of ALA and SFC than under the effect of ALA alone, as reflected by the induced expression of heme oxygenase 1 (HO-1), as well as a reduced protein level of the transcriptional corepressor Bach1. Chromatin immunoprecipitation analysis confirmed Bach1 chromatin occupancy at the enhancer regions of HO-1, which were significantly decreased by the addition of ALA and SFC. Finally, Transwell culture of Caco-2 cells suggested that the administered ALA to the intestinal lumen was partially transported into vasolateral space. These findings enhance our understanding of the absorption and metabolism of ALA in enterocytes, which could aid in the development of a treatment strategy for various conditions such as anemia.

1. Introduction

5-Aminolevulinic acid (ALA) is an important precursor of heme. This compound is synthesized from glycine and succinyl-CoA in mitochondria; this process is catalyzed by two different ALA synthases (ALAS): one expressed ubiquitously (ALAS1) and the other expressed only by erythroid precursors (ALAS2) [1]. During synthesis, ALA is converted to protoporphyrin IX, and heme is generated by the insertion of ferrous iron into protoporphyrin IX. The oral administration of ALA has recently been widely used in various clinical settings. For example, because porphyrin is known to be a strong photosensitizer, ALA has been used to diagnose and treat various cancers [2]. Furthermore, ALA was expected to have therapeutic effects on some types of anemia. It has been suggested that ALA may represent a novel therapeutic option for congenital sideroblastic anemia (CSA), attributable to the mutation of ALAS2, which converts glycine and acetylcoenzyme A to generate ALA [3]. In addition to ALA, some studies have suggested the efficacy of orally administering the combination of ALA and iron for pre-diabetic subjects in reducing serum glucose levels [4,5]. The increase in the heme synthesis in the body by the combination of ALA and iron, rather than ALA alone, was estimated to have a beneficial effect on the serum glucose levels [4,5]. Whereas pharmacokinetic studies of ALA have demonstrated good oral bioavailability [6–9], other reports have suggested that the administration could cause an excessive accumulation of ALA in enterocytes [9,10]. In addition, the detailed differences of the combination of ALA and iron versus ALA only on the enterocytes remain unknown. Thus, there is a need to clarify the dynamics of absorption, metabolism, and excretion of ALA and/or iron in enterocytes.

Herein, based on a model of enterocytic differentiation using human intestinal Caco-2 cells, we evaluated the effects of ALA on enterocytes.
2. Materials and methods

2.1. Cell culture and reagents

Caco-2 cells were obtained from American Type Culture Collection (ATCC; Manassas, VA). Cells were grown in a humidified incubator at 37 °C with 5% carbon dioxide, and maintained in a DMEM medium (Sigma-Aldrich, St. Louis, MO) containing 10% fetal bovine serum (Biowest, Miami, FL). ALA hydrochloride and sodium ferrous citrate (SFC) (SBI Pharmaceuticals Co., Ltd., Tokyo, Japan) were prepared using distilled water. Cells were seeded at a density of 1 × 105 cells/cm² onto 60-mm plastic flasks or 6-well Transwell permeable support plates (Costar, Corning Inc., New York, USA), and cultured for 21 days. The medium was refreshed every 2–3 days. Cells were incubated with a medium containing 200 µM ALA and/or 100 µM SFC for a culture period of up to 72 h. ALA and/or SFC were added to the upper chamber to the Transwell inserts, corresponding to the intestinal lumen. The cells used for all experiments are described here in passages 24–35.

2.2. Real-time quantitative reverse transcription polymerase chain reaction (RT-PCR)

Total RNA was purified using TRIzol (Invitrogen) and 1 µg of the purified total RNA was used to synthesize complementary DNA (cDNA) with ReverTra Ace qPCR RT Master Mix (Toyobo). Reaction mixtures (20 µL) for real-time quantitative RT-PCR comprised 2 µL of cDNA, 10 µL of Quantitect SYBR Green PCR Master Mix (Qiagen), and 8 µL of the appropriate primers. Product accumulation was monitored by measuring SYBR Green fluorescence and normalized relative to GAPDH messenger RNA (mRNA). We used the following primers: DMT1, AGCAGGCTTAAAGATGCTYA and ATTATATGTGGTGCTGGTGTT; FPN, CCTGTTTAAACAGCACCTCAGC and TTGCAAGATGGCTAGTGTCG; ALAS1, GCCGCTCAGCTGATCACTCA and CCACTCATGGGTCTTCAAATCT; heme oxygenase 1 (HO-1), ATGAACTCCCTGGAGATGACTC and ATACGTTTGTGGCTGCTGTG; GAPDH, GAAGGTCGGAGTCAACGGATTT and GATTTGCCATGGTGGGAAT; solute carrier family 36 (proton/amino acid symporter), member 1 (SLC36A1), ATAGCTTTGTGGCTCTTTCGTTTGCTGTA and TTACTGAGGTGACTTGCTTGC. To evaluate absolute expression levels of human ALA transporters (SLC36A1 and SLC15A1), ATACGTTTGTGGCTCTCCA and CCTTTTAACAGGTGGATCAAGG; solute carrier family 15 (oligopeptide transporter), member (SLC15A1), ATACGTTTGTGGCTCTTTCGTTTGCTGTA and TTACTGAGGTGACTTGCTTGC. Expression levels of human ALA transporters (SLC36A1 and SLC15A1), were measured with the appropriate primers. Product accumulation was monitored by measuring SYBR Green fluorescence and normalized relative to GAPDH messenger RNA (mRNA). We used the following primers: DMT1, AGCAGGCTTAAAGATGCTYA and ATTATATGTGGTGCTGGTGTT; FPN, CCTGTTTAAACAGCACCTCAGC and TTGCAAGATGGCTAGTGTCG; ALAS1, GCCGCTCAGCTGATCACTCA and CCACTCATGGGTCTTCAAATCT; heme oxygenase 1 (HO-1), ATGAACTCCCTGGAGATGACTC and ATACGTTTGTGGCTGCTGTG; GAPDH, GAAGGTCGGAGTCAACGGATTT and GATTTGCCATGGTGGGAAT; solute carrier family 36 (proton/amino acid symporter), member 1 (SLC36A1), ATAGCTTTGTGGCTCTTTCGTTTGCTGTA and TTACTGAGGTGACTTGCTTGC. To evaluate absolute expression levels of human ALA transporters (SLC36A1 and SLC15A1), an amplified CDNA fragment of each gene was cloned into the pGEM™-T Easy Vector (Promega, Madison, WI), and was used as an internal standard in quantitative RT-PCR. The plasmid copy number was calculated as follows: copy number (copy/µL) = 6.02 × 1023 × [plasmid DNA concentration (µg/µL)] × 10−6/(total plasmid size (base pair)) × 660, as described previously [3].

2.3. Intracellular heme and protoporphyrin IX content

Intracellular heme content was determined fluorometrically, as described previously [11]. In brief, cell pellets were suspended in 2 M oxalic acid and boiled (100 °C) for 30 min to dissociate protoporphyrin IX and iron from heme. The fluorescence for protoporphyrin IX was then measured at 400 nm (excitation) and 662 nm (emission). To exclude endogenous levels of protoporphyrin IX, the fluorescence of un-boiled samples was subtracted.

2.4. Measurement of ALA, total porphyrin, and heme

Heme content in medium was determined using a chemiluminescence assay as described previously [12]. In brief, the same amounts of medium and chemiluminescence detection reagents (Pierce western blotting substrate plus; ThermoFisher, USA) were mixed and incubated at room temperature for 5 min. Chemiluminescence intensities were then measured using a luminometer (GloMax 20/20, Luminometer, Promega, USA).

2.7. Statistics

Statistical significance was assessed by one-way ANOVA followed by Tukey's post hoc test. In all analyses, differences were considered significant at p < 0.05.

Fig. 1. Differentiation of human intestinal Caco-2 cells. Quantitative RT-PCR analysis of DMT1, FPN, and HO-1 in Caco-2 cells during days 7–21 after seeding. Data are expressed as mean ± standard error (SE). n = 3; *, p < 0.05. Asterisks indicate levels that are statistically significantly different from those on day 7.

ALA levels in medium were analyzed by synthesizing a fluorescent ALA derivative and measuring its concentration with a fluorometric HPLC system, as described previously [13]. Porphyrin levels in medium were also measured with the HPLC system, as described previously [14].

2.5. Western blotting

Western blotting was conducted as described previously [15,16]. Antibodies for ferritin (ab75973) and HO-1 (ab 13248) were purchased from Abcam (Cambridge, UK). The antibody for Bach1 was a generous gift from Prof. Kazuhiro Igarashi (Tohoku University, Japan). A densitometric analysis of Western blot was conducted with ImageJ software (http://rsbweb.nih.gov/ij/). For calculating relative intensity, control samples were set to 1.

2.6. Quantitative ChIP analysis

Real-time PCR-based quantitative chromatin immunoprecipitation (ChIP) analysis was conducted as described previously [17]. We used the following primers: HO-1 E1, ATGAACTCCCTGGAGATGACTC and ATACGTTTGTGGCTCTCCA and CCTTTTAACAGGTGGATCAAGG; solute carrier family 15 (oligopeptide transporter), member (SLC15A1), ATACGTTTGTGGCTCTTTCGTTTGCTGTA and TTACTGAGGTGACTTGCTTGC. To evaluate absolute expression levels of human ALA transporters (SLC36A1 and SLC15A1), an amplified CDNA fragment of each gene was cloned into the pGEM™-T Easy Vector (Promega, Madison, WI), and was used as an internal standard in quantitative RT-PCR. The plasmid copy number was calculated as follows: copy number (copy/µL) = 6.02 × 1023 × [plasmid DNA concentration (µg/µL)] × 10−6/(total plasmid size (base pair)) × 660, as described previously [3].
3. Results and discussion

3.1. Enterocytic differentiation of human intestinal Caco-2 cells

To confirm the enterocytic differentiation of Caco-2 cells, we examined the expressions for DMT1 and FPN, which are known to be an intestinal iron importer and exporter, respectively [18,19]. As shown in Fig. 1, the levels of DMT1 mRNA gradually increased to reach a maximum at day 14, whereas those of FPN mRNA steadily increased until day 21, which were in line with a previous study demonstrating the enterocytic differentiation of Caco-2 cells [20]. On the other hand, the levels of HO-1 mRNA remained the same until day 14 and then decreased until day 21 during Caco-2 cell differentiation. The decrease of HO-1 expression during Caco-2 cell differentiation was similar to that found in a previous study, demonstrating that the HO-1 protein is undetectable after its confluence [21]. Taken together, Caco-2 cells that are subjected to our culture conditions are the representatives of intestinal epithelial cells.

3.2. The combination of ALA and SFC enhanced heme and protoporphyrin IX synthesis in Caco-2 cells

As shown in Fig. 2A, we found that cell pellets treated with ALA were reddish, implying that the ALA-treated cells may contain excessive porphyrin as observed in a patient with porphyria in which reddish urine was exhibited [22]. Notably, whereas the ALA treatment induced heme synthesis, the combination of ALA and SFC promoted heme synthesis (Fig. 2A). We next conducted a western blot analysis to confirm the changes in intracellular heme/iron status. Whereas SFC increased the level of ferritin protein, the combined use of ALA and SFC did not increase the level of ferritin protein, suggesting that iron was efficiently utilized to generate heme (Fig. 2B).
Heme induced the expressions of globins and HO-1, by repressing the activity of Bach1, which is known as a transcriptional repressor that binds the maf-recognition element located at the regulatory region of globin and HO-1 [23]. Induced heme accumulation and HO-1 expression in Caco-2 cells have recently been demonstrated [24]. Noticeably, we demonstrated that the combination of ALA and SFC strongly increased the level of HO-1 protein (Fig. 2B). We further confirmed that ALA reduced the level of Bach1 protein (Fig. 2B), which

**Fig. 3.** ALA did not affect the expression of intestinal iron transporters and minimally changed ALA transporters. A: Quantitative RT-PCR analysis of DMT1, FPN, HO-1, and ALAS1 in Caco-2 cells treated with ALA (200 μM) and/or SFC (100 μM) for 72 h. Data are expressed as mean ± standard error (SE). n = 3; *, p < 0.05. Asterisks indicate levels that are statistically significantly different from those of the control. B: Comparison of quantitative RT-PCR analysis of SLC36A1 and SLC15A1 on day 21. C: Quantitative RT-PCR analysis of SLC36A1 and SLC15A1 in Caco-2 cells treated with ALA (200 μM) and/or SFC (100 μM) for 72 h. Data are expressed as mean ± standard error (SE). n = 3; *, p < 0.05. Asterisks indicate levels that are statistically significantly different from those of the control.
was confirmed by densitrometric analysis (Relative Bach1/tubulin intensity for ALA, SFC, ALA/SFC, and control were 0.48, 1.09, 0.61 and 1, respectively). Furthermore, we conducted quantitative ChIP analysis to assess whether Bach1 directly binds to the regulatory element of HO-1 in Caco-2 cells. The analysis revealed Bach1 occupancy at the HO-1 enhancer region, which was significantly decreased by the addition of ALA/SFC (Fig. 2C). As a reference, we also examined Bach1 occupancy at the NECDIN and RPII215 promoter regions, which are considered to be negative control sites. Taken together, the enhanced heme synthesis in Caco-2 cells was more pronounced under the effect of the combination of ALA and SFC than under ALA alone.

Heme synthesis mostly occurs in developing erythroblasts located in the bone marrow to produce hemoglobin; however, approximately 15% of daily synthesis occurs in the liver to form heme-containing enzymes such as cytochrome P450 [25]. Previous study showed that the induction of HO-1 expression by ALA and SFC occurred in a macrophage cell line (RAW264 cells). The authors demonstrated that the exposure of RAW264 cells to the combination of ALA and SFC increases the HO-1 expression via MAPK activation with the negative regulation of Bach1 [26]. Additional studies, based on a human hepatoma cell line [27] and mouse kidney cells [28], reported ALA-mediated upregulation of HO-1 mRNA levels. Heme has been reported to inhibit the transcriptional repressor activity of Bach1, resulting in the derepression of its target genes such as globin in erythroid cells and HO-1 in diverse cell types [23]. These results are in line with our findings that the combination of ALA and SFC efficiently induces heme biosynthesis in enterocytes.

3.3. ALA did not affect the expression of intestinal ALA/iron transporters

To test whether the ALA-mediated increase in heme concentration in enterocytes could be accompanied by the changes of iron absorption as well as excretion in intestinal cells, quantitative RT-PCR was conducted for DMT1 and FPN. As shown in Fig. 3A, there were no significant changes in the expression of genes involved in iron transport, including DMT1 and FPN (Fig. 3A), suggesting that the ALA-mediated increase in heme concentration may not affect iron absorption/excretion in intestinal cells. Heme has been reported to induce HO-1 expression and suppress ALAS1 expression; both contribute to a negative feedback loop in non-erythroid cells [1]. Similar to the western blot analysis, we demonstrated that HO-1 expression was significantly increased by ALA treatment and that this increase was more pronounced under the combination of ALA and SFC (Figs. 2B and 3A). Furthermore, whereas ALAS1 expression was not affected by ALA, its expression was significantly reduced by the co-administration of SFC (Fig. 3A). We also examined the expression levels of ALA transporters, SLC36A1 and SLC15A1, which were considered to be important ALA transporters in intestinal cells [29,30]. As shown in Fig. 3B, we demonstrated that the absolute mRNA expression level of SLC36A1 was clearly low (by more than 40 times) as compared to that of SLC36A1, suggesting that in Caco-2 cells SLC36A1 may be the main ALA transporter (Fig. 3B). As shown in Fig. 3C, ALA, SFC, and the combination of ALA and SFC did not affect the expression of SLC36A1. On the other hand, whereas SFC and the combination of ALA and SFC did not affect the expression of SLC15A1,
we noticed that ALA administration slightly induced its expression (Fig. 3C). According to the markedly low expression of SLC15A1 as compared to that of SLC36A1 (Fig. 3B), the contribution of the ALA-mediated increase of SLC15A1 may be negligible and thus we consider that ALA did not significantly affect the ALA import into enterocytes.

3.4. Subsets of ALA and porphyrin in Caco-2 cells were transported into vasolateral space

Finally, to demonstrate the dynamics of absorption, metabolism, and excretion of ALA as well as SFC within enterocytes, we measured the concentrations of ALA and porphyrin based on the Transwell culture of Caco-2 cells. The Transwell has two compartments that are separated by one microporous membrane insert. The Transwell insert system can represent the human intestinal environment, i.e., the upper and lower compartments represent the intestinal lumen and vasolateral space, respectively. We demonstrated that a majority of ALA existed in the upper compartment, and the co-administration of SFC did not affect their concentrations (Fig. 4A and B). Among the administered ALA, 35.4% remained in the intestinal lumen and 7.2% of the ALA moved to the vasolateral space in the ALA only group, whereas 42.4% and 8.0% existed in the intestinal and vasolateral lumen, respectively, in ALA and SFC group. In addition, based on the evidence that eight molecules of ALA are used to form one porphyrin ring [22], we calculated the percentage of ALA converted into porphyrins. We demonstrated that 10.3% and 1.6% of administered ALA was converted into porphyrins and existed in the intestinal and vasolateral lumen, respectively (Fig. 4B). Similar to the ALA, the addition of SFC did not affect the distribution of porphyrins. These results suggested that a subset of the administered ALA was transported to vasolateral space across the Caco-2 cell layer. Also, synthesized porphyrins in the Caco-2 cells were mainly excreted into intestinal lumen, whereas a subset of the porphyrins was moved to vasolateral lumen.

The effect of ALA on heme synthesis has been examined in human erythroid cells such as K562 cells and human-induced pluripotent stem cell-derived erythroid progenitor cells [3]. The study demonstrated that ALA resulted in a significant dose-dependent accumulation of heme in the K562 cell line, implying that ALA may represent a novel therapeutic option for the treatment of CSA, particularly for cases involving ALAS2 mutations. Based on the finding that subsets of ALA and porphyrin were transported into vasolateral space, it may be reasonable to develop the ALA treatment for CSA. However, since Caco-2 cells are derived from colorectal adenocarcinoma, which could exhibit reduced mitochondrial respiratory function as a result of Warburg effect, it is possible that heme biosynthesis is compromised in Caco-2 cells [31]. Thus, further studies in vivo should be required to address the potential effect of iron co-administration in the absorption, metabolism, and excretion of ALA and porphyrin.

In conclusion, our findings improve the understanding of absorption, metabolism, and excretion of ALA in enterocytes, which would be translated into clinical settings such as treatment as well as diagnosis.

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Appendix A. Transparency document

Transparency document associated with this article can be found in the online version at http://dx.doi.org/10.1016/j.bbrep.2017.07.006.
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