Effect of *Ricinus communis* L on Microorganisms: Advantages and Disadvantages

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**Abstract**

Castor is an important industrial and medicinal plant, as raw material for thousands of compounds can be obtained from it. Castor contains a number of toxic compounds in different parts of the plant, ricin being the most potent. Highest concentration of inhibitors is found in seeds. These compounds show antimicrobial activity against different pathogenic bacteria. So, these toxic compounds can be used to prepare drugs to treat many diseases worldwide. Antimicrobial effect of castor was also seen against soil microbial community which in turn affects soil health and fertility. However, there are many physical, chemical and biological methods which help in degradation of castor inhibitors.

**Keywords**

*Ricinus communis*, Castor

**Introduction**

Castor (*Ricinus communis* L) is an important oilseed crop belonging to the family, Euphorbiaceae. It is grown worldwide because of commercial importance of its oil. It is cultivated mainly in arid and semi arid regions in different countries, of which India, China and Brazil are major ones. Most of the global demand of castor oil is met by India, being the world’s largest producer of it. In India, the leading state in castor oil production is Gujarat, followed by Rajasthan and Andhra Pradesh (http://www.nmce.co.in/files/study/castor.pdf). Castor can be grown easily in non productive lands and provides viable income in all subtropical and tropical locations that require crops with low input costs (Gana *et al*., 2014). It can prevent desertification and erosion in marginal lands if grown there. It is generally cultivated in semiarid regions where germination and plant growth may be affected by salinity stress(Berman and Wiesman, 2011). It contains optimum level of nutrients like nitrogen, phosphorous and potassium, so it can be used as source of fertilizers for different crops (http://www.nmce.co.in/files/study/castor.pdf). The use of castor oil in manufacturing surfactants, greases, coatings,
cosmetics, pharmaceuticals and many other compounds shows its commercial relevance. Moreover, various extracts of roots, seeds and leaves possess antimicrobial activity, antidiabetic activity and anti-inflammatory activity (Jeyaseelan and Jashothan, 2012).

The castor contains many compounds that are poisonous to human beings, animals, insects and microorganisms. The major toxic protein, ricin is used as a biological weapon as a single milligram can kill an adult human being. It is mainly present in seeds (http://chemistry.about.com/cs/toxicchemicals/a/aa040403a.htm). Ricin inhibits protein synthesis by acting mainly on eukaryotic ribosomes. So, fungi are more susceptible to it than bacteria (Zartman et al., 2003). In addition, the plant contains steroids, saponins, alkaloids, flavonoids, tannins, phenols, phytates, oxalates and glycosides in different parts of the plant including roots, leaves and seeds. All these compounds are responsible for antimicrobial properties of castor. Since, these toxic compounds are present in all parts of the plant so, antimicrobial properties are also shown by whole plant (Jena and Gupta, 2012).

*Ricinus communis* possess good antimicrobial activities against pathogenic bacterial strains such as *Staphylococcus aureus*, *Streptococcus pyogenes*, *Bacillus subtilis* as well as *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Proteus vulgaris*, *Salmonella typhimurium*, *Escherichia coli* and many others. Castor also shows antimicrobial activity against some fungal pathogens i.e. *Candida albicans*, *Aspergillus niger* etc (Islam et al., 2010; Mathur et al., 2011). However, in addition to human pathogens castor inhibitors also have negative effect on soil microbial community. Soil health and productivity is directly related to soil microflora as it is an intimate part of soil organic matter and supports growth and development of different crops in many ways. So, cultivation of castor not only decline microbial count but also affects soil fertility (Zartman et al., 2003).

**Advantages**

Medicinal plants can be a good alternative to conventional medicines in combating diseases caused by infectious microorganisms. These plants contain a number of natural products which can be used to manufacture synthetic drugs. Numerous plants with medicinal properties have been gifted to mankind by nature. The interest in scientific research of *R. communis* is due to its efficacy in the alleviation of a number of diseases worldwide because of presence of toxic compounds in different parts of the plant (Mathur et al., 2011). In the recent years, a number of researchers studied the antimicrobial properties of *R. communis* throughout the world.

Ferreira et al., (2002) found that antimicrobial activity of castor oil plant detergent on different anaerobic bacteria (*Fusobacterium nucleatum* ATCC 25586, *Clostridium perfringens* ATCC 13124, *Prevotella nigrescens* ATCC 33563 and *Bacteroides fragilis* ATCC 25285) was different. Jombo et al., (2008) reported that water and alcohol extracts of dry seeds and leaves of *R. communis* have significant antibacterial activity against *K. pneumoniae*, *E. coli*, *P. vulgaris* and *S. aureus*. Antibacterial activity of various leaf extracts of castor was reported by Islam et al., (2010) against dermatophytic and pathogenic bacteria such as *Staphylococcus aureus*, *Escherichia coli*, *K. pneumoniae* and *Streptococcus pyogenes*. These findings established the potential of the leaves of *Ricinus communis* as an effective antibacterial agent. Khan and Yadav (2011) evaluated cold and hot aqueous and different organic solvent extracts of leaves, stem and roots of castor for their antifungal properties against *Trychophyton rubrum*, *Candida*
albican and Microsporum sp. They found that in case of leaves cold aqueous, methanolic and acetone extracts were effective while in case of stem, only cold aqueous extracts were effective. Cold aqueous extracts of roots were most effective followed by acetone, ethyl acetate and hexane extracts.

Al-kuraishy et al., (2012) reported that Ricinus communis produces significant antimicrobial activity particularly against Gram negative bacteria, in comparison with standard antibiotics. They selected four bacterial genera - two Gram negative (Escherichia coli and Pseudomonas aeruginosa); and two Gram positive (Staphylococcus aureus, Enterococcus fecalis) and found that the minimum inhibitory concentration (MIC) of aqueous extracts ranged between 8-32 mg ml\(^{-1}\) for all. Iqbal et al., (2012) studied the antibacterial activity of aerial parts of R. communis against two Gram-positive bacteria, namely Staphylococcus aureus and Bacillus subtilis and two Gram-negative bacteria, namely Escherichia coli and Shigella flexneri. Ethyl acetate and chloroform extracts were showing more effective MIC values against bacterial strains. The least active fraction among all the extracts was n-butanol showing 0.625 and 2.50 µg ml\(^{-1}\), antibacterial activity against S. aureus and S. flexneri, respectively.

Jeyaseelan and Jashothan (2012) studied antibacterial activity of different leaf extracts of Ricinus communis L against pathogenic bacteria Staphylococcus aureus (NCTC 6571) and Escherichia coli (ATCC 25922). They found that all the four test extracts showed inhibition on both S. aureus and E. coli due to presence of inhibitory compounds like saponins, tannins, flavonoids, cardiac glycosides and terpenoids in all test extracts. Naz and Bano (2012) studied the antibacterial and antifungal activity of methanol, ethanol and aqueous leaf extracts of the plant Ricinus communis. Both Gram positive (Bacillus subtilis: ATCC 6059 and Staphylococcus aureus: ATCC 6538) and Gram negative bacteria (Pseudomonas aeruginosa: ATCC 7221 and Klebsiella pneumoniae) were found to be more sensitive to methanolic leaf extracts. Methanolic and aqueous leaf extracts also showed antifungal activity against selected fungal strains as Aspergillus fumigatus and Aspergillus flavus.

Momoh et al., (2012) studied the antimicrobial activity of the essential oil of castor (Ricinus communis) seeds extracted using soxhlet extractor in 98% n - hexane against fourteen bacteria and six fungi. Comparatively, fungi were found to be less susceptible than bacteria. Poonam and Pratap (2012) studied the antimicrobial activity of seed extracts of castor against some human pathogenic bacteria namely Bacillus subtilis, Bacillus cereus, Staphylococcus aureus, Escherichia coli and two fungal strains namely Candida albicans and Candida glabrata. Bhaumik et al., (2014) screened the in vitro antimicrobial activity of various extracts of fruit-seeds of R. communis using bacterial cultures Staphylococcus aureus (ATCC 9144), Bacillus subtilis (ATCC 6633), Pseudomonas aeruginosa (ATCC 27853) Escherichia coli (ATCC 25922) and fungal cultures Aspergillus niger (ATCC 9029), Aspergillus flavus (ATCC 204304), Candida albicans (ATCC 10231).

They found that most of the extracts executed moderate to good antimicrobial activity against all the tested micro-organisms. Javaid et al., (2015) screened antimicrobial activity of various seeds extracts of castor against Rhodococcus spp, Escherichia coli, Bacillus subtilis, Aspergillus flavus, Aspergillus niger and Trichoderma harzianum. Chloroform and Methanol extracts showed maximum zone of inhibition against bacterial and fungal strains, while Acetone extracts showed significant antifungal activity than antibacterial activity.
Disadvantages

Soil microflora is one of the most important factors which enhance soil fertility in many ways. Microbes are the only source in soil which covert nutrients into forms that can be utilized by the plants and play important role in nutrient cycling (Gaddeyya, 2012). Microbes can be used as biofertilizers in concern with the negative aspect of chemical fertilizers. Biofertilizers promote vegetative growth and yield of crops by providing required nutrients viz. nitrogen, phosphorus, iron, zinc, copper, etc. Moreover, they produce plant growth promoting hormones, vitamins and amino acids and control plant pathogens, thus increase crop production and help in the improvement of soil health (Glick, 1995; Saharan and Nehra, 2011; Mishra et al., 2013). Kumar and Kanjana (2009) concluded that the application of specific bacterial strains can enhance nutrients availability by accelerating the mineralization processes of organic matter in soil, which in turn encourages the vegetative growth and yields of castor. Moreover, many researchers reported that using bio-fertilizers in addition to organic fertilizer led to improvement of vegetative growth and productivity of castor (Patel et al., 2010). Application of biofertilizers to castor can achieve the merits including increasing the soil fertility, saving N-fertilizers, increasing the availability of various nutrients to plant absorption and led to improvement of plant growth and yield (Hussein et al., 2013; Aruna et al., 2015).

As discussed, castor possessed antimicrobial properties due to presence of different toxic compounds. In addition to human pathogens castor inhibitors also affect fungal and bacterial functional diversity in soils after bean maturation. Fungal diversity declined in soils cultivated with castor as compared to fields cultivated with cotton (Zartman et al., 2003). A significant decline in the population of Bradyrhizobium sp. was observed when castor was cultivated in preceding year. This might indicate sensitivity of certain fungal and bacterial species to residual inhibitors in the soil which, in turn, affects soil health and growth of plants (Venkateswarlu et al., 1997). However, there are certain microorganisms which can survive at high concentrations of inhibitors and can effectively degrade them. Actinomycetes concentrations as high as 30,000/g of soil identified in castor field soils might decompose ricin. Though, in vitro assays indicate that this group of bacteria is not effective at degrading ricin. On the other hand, two other bacterial genera, Pseudomonas and Erwinia can effectively degrade the protein in in vitro assay. So these bacteria may be used as biofertilizers for castor so that soil health can be retained in castor grown fields in addition to promoting plant growth and yield (Zartman et al., 2003). Streptomyces thermophilus, Str. Diacetilotis and Lactobacillus acidophilus are also used for the detoxification of castor cake in in vitro assays (Ulanova and Kravchenko, 2013). Many physical, chemical and biological methods are used for the detoxification of castor toxins. Ricin can be detoxified by treatment with proteolytic enzymes, autolysed yeast or Azotobacter, sodium ricinoleate, H_2O_2, KMnO_4, halogens, ethanol extraction, heat and UV irradiation. Another important toxin ricinine can be inactivated by steam treatment, or by heat treatment with lime, Ca (OH)_2, NaCl, formaldehyde, NH_4OH, (NH_4)_2SO_4, KMnO_4 or urea (Rao, 1970).

Fungal and bacterial functional diversity declined in soils after bean maturation. Fungal taxonomic diversity declined in soils cultivated with castor relative to fields cultivated with cotton. This might indicate sensitivity of certain fungal species to residual ricin in the soil. Fungi being eukaryotic are expected to be susceptible to ricin. Bacteria, which have a different ribosome structure from eukaryotes are resistant to ricin and
therefore any observed changes in populations, cannot be directly ascribed to ricin levels. However, since bacteria may be dependent on fungi for degradation of certain components of soil organic matter, fluctuations in bacterial numbers might reflect fluctuations in fungal populations. Actinomycete concentration as high as 30,000/g of soil identified in castor field soils might effectively decompose ricin. In vitro assays indicate that this group of bacteria was not effective at degrading ricin. On the other hand, two other bacteria genera, Pseudomonas and Erwinia were observed to effectively degrade the protein in in vitro assays. Fungal and bacterial functional diversity declined in soils after bean maturation. Fungal taxonomic diversity declined in soils cultivated with castor relative to fields cultivated with cotton. This might indicate sensitivity of certain fungal species to residual ricin in the soil. Fungi being eukaryotic are expected to be susceptible to ricin. Bacteria, which have a different ribosome structure from eukaryotes are resistant to ricin and therefore any observed changes in populations, cannot be directly ascribed to ricin levels. However, since bacteria may be dependent on fungi for degradation of certain components of soil organic matter, fluctuations in bacterial numbers might reflect fluctuations in fungal populations. Actinomycete concentration as high as 30,000/g of soil identified in castor field soils might effectively decompose ricin. In vitro assays indicate that this group of bacteria was not effective at degrading ricin. On the other hand, two other bacteria genera, Pseudomonas and Erwinia were observed to effectively degrade the protein in in vitro assays.

In conclusion, castor possesses good antimicrobial activity against many pathogenic microorganisms due to presence of a number of different inhibitory compounds. Therefore, various extracts of castor can be used to manufacture drugs which may be used to treat a number of diseases instead of conventional antibiotics. Moreover, negative effect of castor on soil microorganisms and soil health can also be minimized if resistant strains of bacteria are used as biofertilizers in castor grown fields.

**References**

Al-kuraishy, H.M.K., Al-weendy, S.M., Al-buhadilly, A.K., Al-bajajy, I.N.A., Al-gareeb, A.I. and Al-hafied, A.A.A. 2012. Antibacterial activity of *Ricinus communis*: In vitro study. Iraqi Journal of Science. 53, 524 – 529.

Anonymous. (2012) National Multi-commodity Exchange of India Limited. http://www.nmce.co.in/files/study/castor.pdf.

Anonymous. (2013) Waterloo researcher
discovers sustainable way to protect crops from drought and disease. http://chemistry.about.com/cs/toxicchemicals/a/aa040403a.htm.

Aruna, B., Bhadraiah, B. and Pindi, P.K. 2015. Effect of different AM fungi on biomass productivity of castor (Ricinus communis). Bioinfolet.12, 176–179.

Berman, P.S.N. and Wiesman, Z. 2011. Castor oil biodiesel and its blends as alternative fuel. Biomass Bioenergy. 35, 2861-2866.

Bhaumik, A., Swapna, M., Sucharitha, M., Sravya, S., Lavanya, M. and Ahmed, S.M. 2014. Phytochemical and antimicrobial screening of various extracts of castor fruit-seeds (Ricinus communis L.). International Journal of Information Research and Review. 1, 124-127.

Ferreira, C.M., Rosa, O.P.S., Torres, S.A., Ferrira, F.B.A. and Bernardino, N. 2002. Activity of endodontic antibacterial agents against selected anaerobic bacteria. Brazilian Dental Journal.13, 118-122.

Gaddeyya, G., Niharika, G.P.S., Bharathi, P. and Kumar, P.K.R. 2012. Isolation and identification of soil mycoflora in different crop fields at Salur Mandal. Advances in Applied Science Research. 3(4): 2020-2026.

Gana, A.K., Amosun, A. and Alhaji, B.B. 2014. Determination of number of manual hoe weeding for optimal yield of castor (Ricinus communis L., Euphorbiaceae) in Nigeria. Global Journal of Botanical Science.2, 21-25.

Glick, B.R. 1995. The enhancement of plant growth by free-living bacteria. Canadian Journal of Microbiolog. 41, 109-117.

Hussein, M.M., ElHabbasha, T.S.F. and Mekki, B.B. 2013. Prospect of Bacterial Inoculants and Organic Fertilizers for Improving Growth, Productivity and Quality of Castor Bean (Ricinus communis L.) Plants in Newly Reclaimed Sandy Soils. World Journal of Agricultural Sciences.9, 421-428.

Iqbal, J., Zaib, S., Farooq, U., Khan, A., Bibi, I. and Suleman, S. 2012. Antioxidant, antimicrobial, and free radical scavenging potential of aerial parts of periplocaaphylla and Ricinus communis. International Scholarly Research Network. 2012, 1-6.

Islam, T., Bakshi, H., Sam, S., Sharma, E., Hameed, B., Rathore, B., Gupta, A., Ahirwar, S. and Sharma, M. 2010. Assessment of antibacterial potential of leaves of Ricinus communis against pathogenic and dermatophytic bacteria. International Journal of Pharma Research and Development. 1, 1-7.

Javaid, B., Rana, N. and Javed, K. 2015. Antimicrobial studies of Ricinus communis seeds extracts. International Journal of Scientific Research and Management. 3, 2752-2759.

Jena, J. and Gupta, A.K. 2012. Ricinus communis linn: A phytopharmacological review. International Journal of Pharmacy and Pharmaceutical Sciences. 4, 25-29.

Jeyaseelan, E.C. and Jashothan, P.T.J. 2012. In vitro control of Staphylococcus aureus (NCTC 6571) and Escherichia coli (ATCC 25922) by Ricinus communis L. Asian Pacific Journal of Tropical Biomedicine. 2, 717–721

Jombo, G.T.A. and Enenebeaku, M.N.O. 2008. Antibacterial profile of fermented seed extracts of Ricinus communis: findings from a preliminary analysis. Nigerian Journal of Physiological Sciences.23, 55-59.

Khan, J.A. and Yadav, K.P. 2011. Assessment of antifungal properties of Ricinus
communis. Journal of Pharmaceutical and Biomedical Sciences. 11, 1-3.

Kumar, N.S. and Kanjana, D. 2009. Influence of integrated nutrient management practices on yield attributes, seed yield, oil yield and nutrient uptake of castor under irrigated conditions. Indian Journal of Agricultural Research. 43, 200-205.

Mathur, A., Verma, S.K., Yousuf, S., Singh, S.K., Prasad, G. and Dua, V.K. 2011. Antimicrobial potential of roots of Ricinus communis against pathogenic microorganisms. International Journal of Pharma and Bio Sciences. 2, 545-548.

Mishra D.J., Singh, R., Mishra U.K. and Kumar, S.S. 2013. Role of Bio-Fertilizer in Organic Agriculture: A Review. Research Journal of Recent Sciences. 2, 39-41.

Momoh, A.O., Oladunmoye, M.K. and Adebolu, T.T. 2012. Evaluation of the antimicrobial and phytochemical properties of oil from castor seeds (Ricinus communis Linn). Bulletin of Environment, Pharmacology and Life Sciences, 1, 21-27.

Naz, R. and Bano, A. 2012. Antimicrobial potential of Ricinus communis leaf extracts in different solvents against pathogenic bacterial and fungal strains. Asian Pacific Journal of Tropical Biomedicine. 2, 944-947.

Patel, H.M., Bafna, A.M. and Patel, Z.N. 2010. Yield and quality of castor affected by INM. Green Farming. 1, 263-265.

Poonam, K. and Pratap, S.K. 2012. Antimicrobial activities of Ricinus communis against some human pathogens. International Research Journal of Pharmacy. 3, 209-210.

Rao, K.H. 1970. Toxic factors and their detoxification in castor. Journal of Food Science and Technology. 7, 77-82.

Saharan, B.S. and Nehara, V. 2011. Plant Growth Promoting Rhizobacteria: A Critical Review. Life Sciences and Medicine Research. 21, 1-30.

Ulanova, R. and Kravchenko, I. 2013. Lactic acid bacteria fermentation for detoxification of castor bean meal and processing of novel protein feeds supplement. International Journal of Engineering Science and Innovative Technology. 2, 618-624.

Venkateswarlu, B., Hari, K. and Katyal, J.C. 1997. Influence of soil and crop factors on the native rhizobial populations in soils under dryland farming. Applied Soil Ecology. 7, 1-10.

Zartman, R., Green, C., Francisco, M.S., Zak, J., Jaynes, W. and Boroda, E. 2003. Mitigation of ricin contamination in soils: sorption and degradation. DTIC documents. http://www.dtic.mil/cgibin/GetTRDoc?AD=ADA482765.

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