Microbial Quality and Safety of Bread Sold in Cafeteria, Tea and Bread Shop of Jimma Town, Oromia Regional State, Southwest Ethiopia

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Abstract

The globally popularity of the bread is could be due to it good aroma and flavor, and affordable for most of consumers and rich in carbohydrate. However, this delicious food is risk for health of people when contaminate with moulds and potential pathogenic bacteria. The aim of this study was to evaluate the microbiological quality and safety of bread sold in Jimma town. The study involved laboratory analysis for microbial quality and safety of bread. Standard methods were used for the enumeration of Enterobacteriaceae, *Staphylococcus aureus*, lactic acid bacteria, yeasts and moulds, antibiotic susceptibility patterns of the isolates. Data were analyzed using SPSS software version 16.0. A total of 90 bread samples (30 from each cafeteria, tea and bread shop) were collected. Result of the study indicated, the mean microbial counts (CFU/g) were dominated by aerobic mesophilic bacteria (5.2 ± 0.5), aerobic bacterial spore (4.6 ± 0.7), moulds (4.0 ± 0.6), *S. aureus* (3.3 ± 0.5), Yeast (3.0 ± 0.6), lactic acid bacteria (2.5 ± 0.4) and Enterobacteriaceae (1.1 ± 0.1), however, Coli form bacteria was not detected in any bread samples. Of the total of 546 isolates characterized, the most predominant were *Bacillus* spp. (40.7%) followed by *Staphylococcus* spp. (25.1%), *Micrococcus* spp. (10.6%), *Pseudomonas* spp. (9.9%), *Acinetobacter* spp. (7.7%) and *Aeromonas* spp. (6%). Totally, 68.9% of samples were positive for *S. aureus* but Salomonella spp. below detectable level in any bread sample of suppliers. *S. aureus* isolates were resistant to maximum seven antibiotics (4.8%) and highly resistant to Methicillin, Oxacillin and Penicillin G (100%). Bread contamination problems in present study could be due to poor personal hygiene, and improperly sanitized utensils. Generally, the microbial quality of bread sold in Jimma town was poor, particularly tea shop and cafeteria bread sellers needs regular inspection.

Keywords: Antibiotics resistance; Bread; Microbial quality; Safety; Pathogenic bacteria

Introduction

Bread is eaten all over the world by almost peoples of every culture. If we traveled to the other side of the planet we would probably find a culture very different from our own, yet with its own version of bread [1]. Therefore; bread is a food product that is universally accepted as a very convenient form of food that has desirability to all population rich and poor, rural and urban. Thus, none of any food types compute with bread in line with consumption in the world. In Nigeria, bread has become the second most widely consumed non-indigenous food product after rice [2]. Also in Ethiopia, next to Injera the bread is highly consumed and leaves among most people, particularly in urban area daily and on holiday.

Although bread is a prominent food for the world population, it is affected the health of the people in case of contaminated with pathogenic microorganisms. Basically, the surface of a fresh baked bread free of viable microorganisms; however, it is subject to contamination by mould spores and bacteria from the air, improperly sanitized utensils, and handlers, transporting equipments and wrapping materials. Ehavald and Estonia [3] explained that more than 90% of bread contamination occurs during cooling, transporting, slicing and wrapping operation. Moreover, it has been reported that mould spores in proofers’ cloths in bakeries can build up enough heat resistance to survive baking [4].

Ogundare and Adetuyi [5] reported from Nigeria, freshly baked bread, after ten minutes had been contained bacterial species, include *Bacillus cereus* and *Staphylococcus* spp. and after 48 and 96 h *Staphylococcus cohnii* and *Bacillus firmus* were isolated, respectively. Similarly, after 10 min mould such as *Aspergillus flavus*, *Aspergillus niger* and *Penicillium citrinum* were reported. Daniyan and Nwokwu [6] reported total aerobic bacterial count ranged from 2.85 × 10⁴ CFU/g to 6.21 × 10⁶ CFU/g. Coli form from 1.19 × 10⁴ CFU/g to 2.05 × 10⁶ CFU/g. *Staphylococcus* from 2.00 × 10⁴ CFU/g to 5.52 × 10⁵ CFU/g and fungi count ranged from 4.0 × 10⁴ to 1.40 × 10⁶ CFU/g and the highest frequency occurrence of *Staphylococcus*, *Escherichia coli* and *Bacillus* spp., respectively were recorded from the beaked bread which are currently emerging to resist various types antibiotics.

Even though, the bread is highly consumed as daily meal in home, cafeteria, and tea shops in Ethiopia [7], the microbial safety of this delicious food is still not documented. On the other hand, the bread is eaten by nearly all of the world population but some groups of the consumers are criticized about the microbial safety of the bread because of unhygienic of transporting material, handlers and storages place of bread [8], which is presently appeared in Jimma town. Hence, having this scenario insight, the present study was designed to assess the microbial load of shop and cafeterias sold bread, and evaluate their antibiotic susceptibility.

Materials and Methods

Description of the study area

The study was conducted in Jimma town, Oromia regional state, located at 353 km southwest of Addis Ababa. The microbial analysis was carried out at Jimma University, Biology department, research...
and Post graduate Laboratory. The geographical location of the town is 7°41’N latitude, 36°50’E longitude, and an average altitude of 1,780 m above sea level. The average of minimum and maximum temperature of the town is of 14 and 30°C, respectively with the annual rainfall ranges from 1138 to 1690 mm [9].

Sample collection

A total of 90 samples were collected from baked bread selling shops of Jimma town for past two years. The food samples were purchased from bread sellers at time between 8:00 and 12 am. The purchased food samples were added into sterile polyethylene bag by bread sellers and transported to Research and Postgraduate Lab. The microbial analysis was conducted within one to three hours of collection. The food samples were kept in the refrigerator at 4°C until microbial analysis was conducted.

Sample preparation and microbial enumeration

Sample preparation: A 25 g of bread samples were suspended in 225 mL of buffered peptone water (BPW), and homogenized in Erlenmeyer flasks for 5 min using shaker at 160 rpm. A 1 mL of homogenized sample was transferred into 9 mL of BPW, and mixed thoroughly by using vortex mixer. The homogenized food sample was serially diluted from 10^{-1} to 10^{-6} and 0.1 mL aliquot of appropriate dilution was plated on pre-solidified plates and incubated at appropriate temperature and period. The colonies was counted from plate containing microbial colonies between 30 and 300 and expressed in colony forming units per gram (CFU/g).

Microbial enumeration: From appropriate serial dilutions, 0.1 mL of the aliquot was plated on Plate Count Agar (PCA), Violet Red Bile Agar (VRBA) plates [10], MacConkey agar [11], Mannitol Salt Agar (MSA) [12] and incubated at 32°C for 48 h for count Aerobic mesophilic bacteria, Coliform, Enterobacteriacaeae and Staphylococci, respectively, and also for count of Aerobic bacterial spore forms, appropriate serially diluted sample was heat treated in a water bath adjusted at 80°C for at least 10 min. Thereafter, 0.1 mL aliquot was plated on pre-solidified surface of PCA and incubated at 35°C for 48 h [12]. Moreover, after, 0.1 mL aliquot was plated on pre-solidified surfaces of Potato Dextrose Agar supplemented with 0.1 g chloramphenicol and incubated at 25°C for 5-7 days [11]. Smooth (non-hairy) colonies without extension at periphery were counted as yeasts whereas hairy colonies with extension at periphery were counted as mould.

Microbial analysis: From appropriate aerobic mesophilic countable plate, 10 to 15 colonies with distinct morphological differences were randomly picked from plates and aseptically transferred in to a test tube containing 5 mL of nutrient broth. Then, incubated at 32°C for 24 h. The repeatedly sub-culturing isolates were characterized up to genus level based on John [13] bacterial classification manual.

The cell morphology of the bacterial pure isolates was examined microscopically after gram staining, and also the motility of isolates were checked using motility medium. Moreover, after repeatedly purified the agar block of mold culture on PDA, the morphology of hyphal culture was stained using lacto phenol cotton blue and observed under the microscope for identification of fungal genera. The biochemical test including KOH (potassium hydroxide), Catalase, Oxidation fermentation (O/F), Cytochrome oxidase tests were conducted, accordingly.

Catalase test: After plate contains young colony had flooded with 1 mL of 3% H2O2, and the formation of gas bubbles was observed. The occurrence of gas bubbles was taken as positive for catalase test [14].

Oxidation fermentation (O/F) test: Ingredients (g/L): Peptone, 2 g; yeast extract, 1 g; NaCl, 5 g; K2HPO4, 0.2 g; glucose,10 g; bromothymol blue, 0.08 g; agar, 2.5 g; distilled water, 1000 mL; pH, 7.10 were prepared. Accordingly, test tubes containing 15 mL of freshly prepared medium for O/F test were autoclaved and immediately cooled under tap water to avoid dissolution of oxygen in the medium. Then, the broth cultures were inoculated into the medium by stabbing with a sterile straight wire to the bottom. An organism with oxidative metabolism displayed yellow in the upper half of the tube and green in the lower half. An organism with fermentative metabolism displayed yellow in both halves of the tube. Acid formation and growth regions were interpreted after 2 to 5 days of incubation at 32°C.

Cytochrome oxidase test: Accordingly, freshly prepared reagent A and B were mixed in the ratio of 2:3 immediately before use. Reagents: A, 1% α-naphthalene dissolved in absolute ethanol, B, 1%N, N-dimethyl-p-phenylenediammonium chloride in distilled water. Three drops of the oxidase reagent were added on to the surface of the growth of isolated colonies of test bacterium. The presence or absence of appearance of a blue color on the colonies was observed within 30 s Kovacs [15].

Isolation of Salmonella spp.: A 25 g of food samples was mixed with 225 mL of BPW and incubate at 37°C for 24 h, then 1 mL pre-enrichment broth culture was added to 10 mL of selenite cystine broth and incubate at 37°C for 24 h. Next, a loopful of suspension from secondary enrichment broth was streaked onto Xylose Lysine Deoxycholate Agar (XLD). The presumptive Salmonella colonies was picked off and transfer to 5 mL nutrient broth and incubate at 37°C for 24 h, then streak onto Nutrient Agar for purity and incubate at 37°C for 24 h. For conformation the isolates were Salmonella spp., the biochemical tests was done according to the procedure of Johnson and Case [16].

Isolation of Staphylococcus aureus: The golden yellow colony shown on MSA and Gram-positive cocci with clustered arrangement under the microscope were subjected to preliminary biochemical tests coagulase tests. The pure colony of isolates were emulsified with distilled water on the duplicate clean slide (i.e., control and test slide), then, a loopful of human blood plasma was added on the emulsified suspension and mixed. The formation of clumping with in 10 s was suggested isolates as Coagulase positive [17].

Antimicrobial susceptibility testing for some pathogens: The antimicrobial susceptibility testing for pathogens isolated from bread samples was tested using the disk diffusion method and the microbial cell concentration was adjusted to the standardize turbidity of 0.5 McFarland which is equivalent to 10^{8}- 10^{9} CFU/g [18]. The sterilized cotton swab moistened with bacterial cell suspension had been swabbed on to the Muller-Hinton Agar and allows to drying. Thereafter, the antibiotic discs were dispensed on the medium and incubate at 37°C for 18 h and the zones of inhibition were measured using vernal caliper. The results of the antimicrobial susceptibility were interpreted based on the guidance of National Committee for Clinical Laboratory Standards NCCLS [19]. Finally, the isolates were classified as sensitive, intermediate, or resistant. Intermediates were considered as resistant for purpose of analysis. The following standard drug discs (Oxoid) and their potency (μg/mL) were used. As a result chloramphenicol (30), Norflaxacin (10), Amoxicillin (10), Erythromycin (15), Oxacillin (5), Vancomycin (30), penicillin G (10) and Methicillin (5) were used for Staphylococcus aureus. The reference strains were Staphylococcus aureus (ATCC25923).
Data analysis: The Percentage of Coefficient of variation (%CV) was calculated to see if there is significant variation in counts within the bread samples analyzed. Mean values of bread samples from different source were compared using one way ANOVA and the significance of differences were considered at 95% confidence interval (P<0.05).

Results

The result of this study indicated highest mean count of Enterobacteriaceae (2.0 log CFUg⁻¹) and S. aureus (3.7 log CFUg⁻¹) were recorded from cafeteria and tea shop, respectively, while considerable number of lactic acid bacteria (LAB 2.8 log CFUg⁻¹) was observed in bread shop. On the other hand, the mean counts of Coliform were below detectable level in all bread samples suppliers (Table 1). There was statistically significant difference (p<0.05) among the mean counts of Aerobic mesophilic bacteria (AMB), Enterobacteriaceae, coliform, Aerobic bacterial spore count (ABS), Staphylococci, LAB, Yeasts and Moulds in all bread samples sources.

Microbial analysis of bread samples

From the total of 90 bread samples analyzed, 546 bacterial isolates were obtained. The isolates were grouped into six genera based on John’s [13] bacterial classification system. Among the identified genera, the predominant bacterial group was Bacillus spp. (40.7%) followed by Staphylococcus spp. (25.1%), Micrococcus spp. (10.6%) and Pseudomonas spp. (9.9%) (Table 2).

Prevalence of S. aureus and Salmonella spp.

In the present study, the overall 68.9% bread samples were positive for S. aureus. However, the frequency distribution varied among the bread suppliers. Accordingly, it was as prevalent as 93.3% of S. aureus in tea shop and 70% in cafeteria bread whereas the lowest prevalence was observed in bread shop (43.3%) (Figure 1); on the other hand, no Salmonella spp. were detected in any bread samples collected from aforementioned suppliers.

Methicillin-resistant patterns of Staphylococcus aureus (MRSA) isolates

The MRSA patterns of S. aureus revealed that, 40.3% of the isolates were resistant to 4 antibiotics followed by 14.5% to 4 and 5 antibiotics and fewer isolates (4.8%) resistant to 7 antibiotics (Table 3).

Discussion

In the present study, the mean total counts of AMB (5.2 CFUg⁻¹) was observed which is in agreement with Daniany and Nwokwu [6] who reported between 4.5 to 6.8 log CFUg⁻¹ from beaked bread. Unsanitary handling of bread sellers might cause for cross contamination [20]. Some bread seller is carriers for pathogenic microbes and transfer food borne pathogens to the consumers during bare handling of bread. Salmonella, Campylobacter and E. coli can survive on fingertips for long periods of time. In generally, the mean count of AMB in all bread suppliers shop were 4.7 and above which is belonged to unsatisfactory level [21]. The over loaded of AMB in bread samples could be due to poor hygienic of bakery, bread sellers and transporting materials. Ehaltov and Estonia [3] also reported more than 90% of bread contamination occurs during cooling, transporting, slicing and wrapping processes. AMB loaded food regarded as harmful even if the organisms are not known to be pathogens [22].

The mean count of Enterobacteriaceae in the present study was 1.1 logs CFUg⁻¹ which is very low compare to the earlier report of Mustafa and Abdul [23] who reported the counts between 2.3 and 4.4 log CFUg⁻¹ in traditional foods. Moreover, none of Enterobacteriaceae detected from bread shop. Hence, according to the guideline the Enterobacteriaceae count fit to the acceptable level. The turn down of Enterobacteriaceae count implies that due to low water activities of bread.

In the present study, in any bread suppliers samples the coliform bacteria were below the detectable level in contrast to the report of Danyian and Nwokwu [6], who reported counts between 4.1 and 6.3 CFUg⁻¹ from beaked bread. The absence of Coliform in the present study could be due to the bread samples less exposed to fecal contamination.

The aerobic bacterial spore (ABS) count (4.6 log CFUg⁻¹) of the present study is higher compared to report by Mosupye and Holy [24] where the counts ranged between 1.2 and 2.0 log CFUg⁻¹ in ready to eat food samples from Johannesburg, South Africa, however, comparable to the finding of Ismail [25], who reported between 2.3 and 4.7 log CFUg⁻¹ from ready to eat food. According to the guideline, in all bread suppliers the mean count of ABS were ranged in unsatisfactory level (>4 log CFUg⁻¹). The higher counts in the present study implies that due to the contamination of bread by the heat resistant spore forming bacteria that survive baking and germinate in the bread after baking, from air during transportation, improperly sanitized utensils, and handlers, transporting equipments and wrapping materials.

Table 1: Mean of microbial counts (log CFUg⁻¹) from bread sold in Jimma town.

| Sample source | No AMB | % CV | Entero | % CV | ABS | % CV | S. aureus | % CV | LAB | % CV | Yeast | % CV | Mould | % CV |
|---------------|--------|------|--------|------|------|-------|-----------|------|-----|------|-------|------|-------|------|
| Tea shop      | 30     | 5.9 ± 0.4 | 8     | 1.3 ± 0.2 | 15 | 4.5 ± 0.8 | 17 | 3.7 ± 0.8 | 20 | 2.7 ± 0.7 | 25 | 3.4 ± 0.6 | 17 | 4.6 ± 0.7 | 15 |
| Bread shop    | 30     | 4.7 ± 0.4 | 8.5 | 0.0 ± 0.0 | 15 | 4.5 ± 0.9 | 17 | 3.7 ± 0.9 | 20 | 2.7 ± 0.8 | 25 | 3.4 ± 0.7 | 17 | 4.6 ± 0.8 | 15 |
| Cafeteria     | 30     | 4.9 ± 0.7 | 14 | 2.0 ± 0.1 | 15 | 4.5 ± 0.10 | 17 | 3.7 ± 0.10 | 20 | 2.7 ± 0.9 | 25 | 3.4 ± 0.8 | 17 | 4.6 ± 0.9 | 15 |
| Total         | 90     | 5.2 ± 0.5 | 11.1 | 0.1 | 4.5 ± 0.11 | 17 | 3.7 ± 0.11 | 20 | 2.7 ± 0.10 | 25 | 3.4 ± 0.9 | 17 | 4.6 ± 0.10 | 15 |

AM: Aerobic Mesophilic Bacteria; Entero: Enterobacteriaceae; ABS: Aerobic Bacterial Spore, S. aureus: Staphylococcus aureus; LAB: Lactic Acid Bacteria; CV: Coefficient of Variance; SD: Standard Deviation; CFU: Colony Forming Unit.

Table 2: Frequency distribution of dominant bacteria in bread collecting from selling area, Jimma town, southwestern Ethiopia.
In the present study, the mean count of LAB was 2.5 log CFU/g. This result is lower than the earlier work of Omemoo and Omeike [27] that ranging from 4.5 to 9.2 log CFU/g from baked bread. The higher S. aureus in the present study could be due to unhygienic handling of the bread sellers, particularly tea shop and cafeteria workers frequently used bare hand that is why the amount of S. aureus was increased in these bread suppliers. Moreover, they did not use special cloth while processing and selling. According to Mensah et al. [26] increased in these bread suppliers. Moreover, they did not use special S. aureus was frequently used bare hand that is why the amount of handling of the bread sellers, particularly tea shop and cafeteria workers.

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In present study, the antibiotic resistance patterns of the isolates revealed that all of S. aureus isolates resistant to three Methicillin group bacteria including Methicillin, Oxacillin and Penicillin G, and also most of isolates resistance to Amoxicillin (85.5%) and Erythromycin (40.3%). This was in agreement with Alexandra et al. [35] who reported that, 100% of the isolates were resistant to most Methicillin group bacteria whereas highly resistance than Temilade [36] isolates in which out of 106 isolates of S. aureus, 40.6% were resistant to erythromycin, 63.2% to penicillin G, and 20.7% resistant to oxacillin. This could be, presently isolates S.aureus carry mecA gene that encodes a variant Penicillin binding protein (PBP2a or due to the production of penicillinase enzyme that hydrolyzed the beta-lactam ring of penicillin derivatives antibiotics [37]. The alarming emerging of MRSA could be due to integration of genetic mobile elements such as plasmids, transposons, and insertion sequence in case of inappropriate or uncontrolled use of antibiotics [38,39]. Therefore, it is necessary to pay more attention to food hygiene practices to reduce or eliminate the risk from resistance to antibiotics and pathogenic bacteria originating from food [40].

Conclusion

In concluded, the most predominant microbial genera were Bacillus spp., Staphylococcus spp. and Micrococcus spp. hence, the presence of these microorganisms could be forecast for the presence of potential pathogens. Although Salmonella spp. and coliform were

Table 3: MRSA patterns of S. aureus isolated from bread in Jimma town.

| No. of patterns | Antimicrobial resistance patterns | No. of isolates (%) | Total (%) |
|-----------------|-----------------------------------|---------------------|-----------|
| Two             | MET/OX                           | 6 (9.7)             | 6 (9.7)   |
| Three           | MET/OX/VAN                       | 3 (4.8)             | 6 (9.6)   |
| Four            | MET/E/OX/AM                      | 3 (4.8)             | 9 (14.5)  |
| Five            | MET/P/OX/AM                      | 22 (35.5)           |           |
| Six             | MET/E/OX/AM/VAN                  | 3 (4.8)             | 9 (14.5)  |
| Seven           | MET/E/OX/AM/VAN/VAN              | 3 (4.8)             | 3 (4.8)   |

MET: Methicillin, OX: Oxacillin; VAN: Vancomycin; AM: Amoxicillin; P: Penicillin; E: Erythromycin; C: Chloramphenicol.
not detected in present study, the presence of high number *S. aureus* cause food intoxication’ that lead to food born diseases. All of *S. aureus* isolates were resistant to three Meticillin group antibiotics including Meticillin, Oxacillin and Penicillin G. Thus, the *S. aureus* isolates considered as MRSA. The overall microbial quality of bread samples collected from suppliers was poor as compared to the guidelines set by other regulatory bodies. This could be due to poor personal hygienic, transporting materials and exposing to air microorganisms. Thus, the concerned bodies like the municipal and health official of Jimma town should give attention to improve the safety of baked bread by providing training to bakers and bread sellers to keep their personal hygiene, clean the transporting materials and how to manage the hygienity of bread accordingly.

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