Fibrodysplasia ossificans progressiva: current concepts from bench to bedside

Arun-Kumar Kaliya-Perumal1, Tom J. Carney1,2 and Philip W. Ingham1,2,*

ABSTRACT
Heterotopic ossification (HO) is a disorder characterised by the formation of ectopic bone in soft tissue. Acquired HO typically occurs in response to trauma and is relatively common, yet its aetiology remains poorly understood. Genetic forms, by contrast, are very rare, but provide insights into the mechanisms of HO pathobiology. Fibrodysplasia ossificans progressiva (FOP) is the most debilitating form of HO. All patients reported to date carry heterozygous gain-of-function mutations in the gene encoding activin A receptor type I (ACVR1). These mutations cause dysregulated bone morphogenetic protein (BMP) signalling, leading to HO at extraskeletal sites including, but not limited to, muscles, ligaments, tendons and fascia. Ever since the identification of the causative gene, developing a cure for FOP has been a focus of investigation, and studies have decoded the pathophysiology at the molecular and cellular levels, and explored novel management strategies. Based on the established role of BMP signalling throughout HO in FOP, therapeutic modalities that target multiple levels of the signalling cascade have been designed, and some drugs have entered clinical trials, holding out hope of a cure. A potential role of other signalling pathways that could influence the dysregulated BMP signalling and present alternative therapeutic targets remains a matter of debate. Here, we review the recent FOP literature, including pathophysiology, clinical aspects, animal models and current management strategies. We also consider how this research can inform our understanding of other types of HO and highlight some of the remaining knowledge gaps.

KEY WORDS: Bone morphogenetic protein, ACVR1, Fibrodysplasia ossificans progressiva, Heterotopic ossification, Inflammation

Introduction
Skeletal bone formation, also known as osteogenesis or ossification (see Glossary of clinical terms, Box 1), is a multi-step process involving the formation of mature mineralised bone through the differentiation of progenitor cells (Gilbert, 2000). There are two types of ossification, namely intramembranous and endochondral (Box 1), both of which are temporally and spatially regulated (Setiawati and Rahardjo, 2019). Any disruption to this regulation can lead to abnormal skeletal development or heterotopic (extraskeletal) ossification (HO; Box 1).

HO is the formation of bone, either solitary or multiple, in extraskeletal soft tissues of the body. Several genetic diseases manifest with HO at multiple sites, including fibrodysplasia ossificans progressiva (FOP; Box 1), progressive osseous heteroplasia (POH) and Albright’s hereditary osteodystrophy (AHO) (Shore and Kaplan, 2010). Among these, FOP is characterised by HO of endochondral origin, predominantly at muscles, tendons, ligaments and fascia (Kaplan et al., 2008), whereas POH and AHO are characterised by HO of intramembranous origin, predominantly at cutaneous and subcutaneous sites (Kaplan and Shore, 2000). While HO in the above conditions is driven by the underlying genetic disorder, it can also occur in response to triggering events, especially injury (Meyers et al., 2019). There are two such conditions collectively referred to as non-genetic or acquired disorders of HO, myositis ossificans traumatica (MOT) and neurogenic heterotopic ossification (NHO) (Meyers et al., 2019).

Amongst all these disorders, FOP is the most extensively studied. It is extremely rare, affecting 1 in 2,000,000 people (Baujat et al., 2017), has no ethnic predilection and is described as the most catastrophic among HO disorders in humans (Miao et al., 2019). There have been remarkable progress in understanding the pathological mechanisms of FOP; it continues to present a significant clinical challenge. This article aims to outline the current information on FOP pathophysiology, clinical aspects, animal models and current management strategies. We also consider how knowledge of FOP can inform understanding of other more common forms of HO.

Pathophysiology of FOP
Gene mutations
All FOP patients reported to date were found to carry heterozygous gain-of-function mutations in the *ACVR1* gene, located on chromosome 2 (2q23-24). *ACVR1* encodes a bone morphogenetic protein (BMP) type 1 receptor, also known as activin A receptor type 1. In most cases, a single nucleotide transition (c.617G>A) causes a missense mutation of codon 206, resulting in substitution of arginine by histidine (R206H) in the intracellular glycine-serine (GS) domain of ACVR1 (Fig. 1). Atypical missense mutations (L196P, R202I, Q207E, R258G/S, G328R/W/E, G356D and R375P) in the GS or protein kinase (PK) domains of ACVR1 have also been identified in some FOP patients (Petrie et al., 2009;
Box 1. Glossary of clinical terms

**Ankylosis**: fusion of a joint, resulting in a complete restriction of its movement.

**Endochondral ossification**: mesenchymal cells form cartilage that proliferates and matures to be replaced eventually by osteoblasts leading to bone formation.

**Fibrodysplasia ossificans progressiva (FOP)**: an extremely rare genetic disorder characterised by heterotopic endochondral ossification at multiple sites, predominantly muscles, tendons, ligaments and fascia.

**Flare-up**: onset of heterotopic ossification in FOP in which the underlying acute inflammation causes the appearance of painful and warm soft tissue swellings.

**Hallux valgus**: outward deviation of the great toe.

**Heterotopic ossification (HO)**: bone formation at extraskeletal or ectopic sites.

**Intramembranous ossification**: mesenchymal cells directly differentiate into osteoblasts that eventually form bone.

**Macrodactyly**: abnormally large fingers or toes.

**Metamorphosis**: transformation of one tissue into another.

**Osteochondromas**: benign bony protuberances with a cartilaginous cap, mostly originating from the growth plate of long bones.

**Osteogenesis/ossification**: the process of bone formation.

**Thoracic insufficiency syndrome**: inability of the thorax to carry out normal breathing functions.

Katagiri et al., 2018a,b; Furuya et al., 2008; Huning and Gillessen-Kaesbach, 2014; Kaplan et al., 2009). In most cases, the mutations arise spontaneously *de novo*; however, a small number of FOP patients showing autosomal dominant inheritance from a symptomatic parent have also been reported (Kaplan et al., 1993; Connor et al., 1993; Shore et al., 2006).

**Dysregulated BMP signalling**

BMPs are required for multiple developmental processes (Wang et al., 2014), including bone and cartilage formation. Secreted BMPs bind to complexes of type I and type II serine/threonine kinase BMP receptors on the cell surface, such as ACVR1, to activate the intracellular signal transduction pathway (Fig. 2). In the absence of BMP ligands, the FK506-binding protein 1A (FKBP1A) binds to the GS domain of ACVR1 and inhibits the binding of effector molecules (Shen et al., 2009). Upon ligand binding, the type II receptor phosphorylates the type I receptor within its GS domain, releasing FKBP1A and thus allowing ACVR1 to bind and phosphorylate intracellular BMP-responsive transcription factors, the receptor-regulated SMADs (R-SMADs) SMAD1/5/9(8) (Wang et al., 2014). Phosphorylated SMAD1/5/9(8) forms a complex with the co-mediator SMAD4 that translocates into the nucleus, where it associates with co-activators or co-repressors to regulate transcription involved in endochondral ossification (Wang et al., 2014). Whereas SMAD1 and SMAD5 activate transcription in this context, SMAD9 acts as a transcriptional repressor (Tsukamoto et al., 2014). Notably, SMAD9 loss-of-function mutations lead to increased bone mineral density and cortical thickness, resulting in greater bone strength but not HO, as in FOP (Gregson et al., 2020).

Earlier studies of FOP suggested the activation of ACVR1R206H in a ligand-independent manner, especially due to impaired binding of FKBP1A and thus inappropriate binding and phosphorylation of SMAD proteins, to be the predominant cause of HO (Shen et al., 2009). More recent work, however, established the role of inflammation in HO genesis and propagation in FOP patients (Alessi Wolken et al., 2018; Meyers et al., 2019). Alessi Wolken et al. suggested that the process by which HO occurs in FOP is likely to be ligand dependent and to involve ligands that activate ACVR1R206H, which are themselves regulated by inflammation (Alessi Wolken et al., 2018). Specifically, they showed that activin A, which is expressed by innate immune system cells and plays an important role in both promoting and resolving inflammation, is effectively perceived as a BMP ligand by ACVR1R206H, leading to downstream BMP signalling via SMAD1/5/9(8) (Alessi Wolken et al., 2018). Similarly, Hatsell et al. found that, as well as showing increased sensitivity to its ligands BMP2, BMP4, BMP7, BMP9 and BMP10, the mutant ACVR1R206H also responded to activins A, AB, AC and B, to which the wild-type ACVR1 is unresponsive (Hatsell et al., 2015).

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**Fig. 1. Schematic representation of human ACVR1, its various domains and locations of the mutations that have been causally linked to FOP.** GS, glycine-serine.
Activins, unlike BMPs, do not normally promote chondrogenesis or osteogenesis and generally signal via different SMADs, namely SMAD2/3 (Macias-Silva et al., 1998). In FOP, however, the mutated receptor erroneously perceives activin as a BMP, eliciting phosphorylation of SMAD1/5/9(8) as in normal BMP signalling (Sanchez-Duffhues et al., 2016). In addition to this mechanism, Wang et al. suggested that increased basal phosphorylated SMAD1/5/9(8) activity and local hypoxia occurring during tissue damage and inflammation can induce BMP signalling in a ligand-independent manner (Wang et al., 2018, 2016). Thus, tissue damage and inflammation in FOP patients can result in both ligand-dependent and -independent aberrant activation of the BMP signalling cascade, leading to activation of the endochondral ossification transcription programme by phosphorylated SMAD1/5/9(8).

The cell of origin

The cells exhibiting dysregulated BMP signalling and osteogenic differentiation in FOP were originally thought to belong to the myogenic lineage (Katagiri et al., 2018a). However, the mesenchymal stem cell population at the site of inflammation has been found to be a more relevant source of progenitor cells that differentiate into chondrocytes and osteoblasts in response to aberrant signalling (Billings et al., 2008). The origin of these mesenchymal stem cells has been traced to local stromal/fibroblastic cells, endothelial cells (via endothelial-mesenchymal transition), Scx⁺ tendon progenitor cells, bone marrow-derived muscle-resident Mx1⁺ cells, glutamate transporter (Glast; also known as SLC1A3)-expressing progenitor cells and some circulating osteogenic precursor cells that can access bone-forming sites (Pignolo and Kassem, 2011; Ranganathan et al., 2015; Dey et al., 2016; Wosczyana et al., 2012; Pulik et al., 2020). This remains an area for further exploration and there are several ongoing studies in animal models to identify progenitor cell populations contributing to the development of HO (Lees-Shepard and Goldhamer, 2018).

Metamorphosis

In FOP, soft tissue metamorphosis (Box 1) is the process by which skeletal muscle, tendons and ligaments transform into mature bone at various extraskeletal sites. There are two phases in this process: catabolic and anabolic. Inflammation represents the catabolic phase and is characterised by amplified infiltration of lymphocytes, macrophages and mast cells into the affected tissues, leading to necrosis (Kaplan et al., 2011). Since FOP patients have a pro-inflammatory baseline state characterised by amplified circulating pro-inflammatory monocytes, they naturally predispose to flare-ups (Box 1) (Barruet et al., 2018). In addition, experimental studies have shown that inflammatory triggers cause prolonged activation of the NF-κB signalling pathway in FOP monocytes and abnormal cytokine/chemokine secretion in both FOP monocytes and FOP primary monocyte-derived macrophages, likely mediated by NF-κB or p38 MAPK (also known as MAPK14) activity (Barruet et al., 2018). These factors seem sufficient to drive the catabolic phase in FOP. Following this, an anabolic phase supervenes, characterised by fibroproliferation, neovascularization and angiogenesis (Pignolo et al., 2005). The pool of mesenchymal cells at the site of inflammation undergoes differentiation in response to the activation of the BMP signalling cascade, leading to transformation of the fibroproliferative tissue into cartilage, which

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**Fig. 2. BMP signalling.** (A) BMPs bind to complexes of type I and type II serine/threonine kinase BMP receptors, such as ACVR1, on the cell surface to activate intracellular signal transduction via R-SMADs SMAD1/5/9(8). Phosphorylated SMAD1/5/9(8) forms a complex with co-mediator SMAD4 and translocates into the nucleus, where it regulates transcription that drives endochondral ossification. (B) On binding activin A, complexes of type I and type II BMP receptors activate intracellular signal transduction via SMAD2/3, which activates a transcription programme that regulates inflammation. (C) ACVR1 carrying a FOP mutation (most frequently the R206H substitution) in the intracellular glycine-serine domain not only yields enhanced response to various BMP ligands by initiating downstream signalling via SMAD1/5/9(8), but also responds to various activin ligands, thereby favouring endochondral ossification by triggering an osteogenic gene expression programme. BMP, bone morphogenetic protein; Co-SMAD, common partner SMAD; P, phosphorylation; R-SMAD, receptor-regulated SMAD.
in turn matures into bone through an endochondral process, thus
completing the process of metamorphosis (Kaplan et al., 2011).

**Diagnosis and management**

Identification of the congenital and episodic signs and correlating them to FOP is of utmost importance (Box 2). This is sufficient for a working diagnosis of FOP. However, a high index of suspicion is needed to make a diagnosis based solely on clinical presentation. A lack of suspicion leads to delayed diagnosis or misdiagnosis, which potentiates inappropriate and unnecessary testing, especially invasive biopsies that may cause flare-ups and actually promote HO. Hence, clinicians, especially paediatricians, who are typically the first to come across children with FOP, need to be aware of this condition, its consequences and effective mitigating strategies. Even though biochemical and radiological investigations can provide useful information on the disease process, any kind of invasive procedure is contraindicated as it induces local inflammation and eventually HO; hence, biopsy of lesions should never be attempted (Trigui et al., 2011). Ultimately, the diagnosis can only be confirmed by DNA sequence analysis to trace the underlying mutation.

As the entire cascade of events leading to HO is triggered by tissue damage and subsequent inflammation, prevention and control of inflammation are the basis of clinical management. However, preventing a triggering event is extremely challenging, as such events could be anything from significant trauma to trivial occurrences such as intramuscular injections, blunt muscle trauma from bumps, bruises and falls, influenza-like illnesses and, in some cases, mere muscle fatigue (Kaplan et al., 2008). How each of these events influence HO is not fully understood, mainly because they remain unnoticed until a flare-up occurs. However, since inflammation follows every trigger, clinical management is mainly focused on mitigating inflammation, thereby alleviating symptoms. As yet, there is no definitive treatment that can alter the natural course of FOP.

According to Kaplan et al. (2019) the pharmacological agents for managing FOP based on research findings and anecdotal experience can be divided into three classes (Table 1). Class I medications are those that contain acute inflammation flare-ups. These include high-dose corticosteroids and non-steroidal anti-inflammatory drugs (Kaplan et al., 2019; Pignolo et al., 2013; Qi et al., 2017). Class II medications are used for management of other conditions but have a theoretical application in FOP (Kaplan et al., 2019; Schaper et al., 2011; Convente et al., 2018; Werner et al., 2013; Brantus and Meunier, 1998; Pennanen et al., 1995; Pabst et al., 2014; Brennan et al., 2018). These are being used anecdotally as adjunctive therapy for refractory flare-ups in FOP (Kaplan et al., 2019). However, there is as yet no solid evidence to substantiate the use of these drugs. Finally, class III medications are those that are currently under clinical investigation as holding the key for an effective treatment (Kaplan et al., 2019; IFOPA, 2020). These drugs are mainly focused on inhibiting the canonical and non-canonical BMP signalling from ACVR1R206H at both the extracellular and intracellular levels. Other proposed pre-clinical approaches to inhibit mutant ACVR1-dependent bone induction include the use of nucleic acid-based inhibitors, such as allele-specific RNA interference or exon-skipping oligonucleotides, BMP receptor kinase inhibitors (such as dorsomorphin), downstream BMP signalling inhibitors (for example, Fendiline and Perhexiline) and fungal metabolite osteoblast differentiation inhibitors (such as NG-391, NG-393 and Trichocyalide A/B) (Katagiri et al., 2018a; Yamamoto et al., 2013; Sanvitale et al., 2013; Fukuda et al., 2012; Cippato et al., 2018).

**Animal models in FOP research**

One of the first animal models to provide insights into the role of BMP signalling in HO was established prior to the discovery that FOP patients carry activating mutations in ACVR1. Transgenic mice overexpressing BMP4 under the control of neuron-specific enolase were initially developed to study the role of BMP signalling in brain development (Gomes et al., 2003). However, by 2 months of age, these mice started showing phenotypes such as stiffness and gait abnormalities (Kan et al., 2004). Histological and immunohistochemical analysis of limb tissue showed muscle degradation and proliferation of subcutaneous fibroblast-like cells (Kan et al., 2004). Subsequently, HO of endochondral origin was found at multiple locations, such as hind limbs, fore limbs,

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**Box 2. Clinical case presentation – a consolidated summary from the literature**

**Classical signs**

Most patients with FOP are typically born with congenital great toe malformations such as hallux valgus and macrodactyly (Box 1). Signs of HO start to occur episodically from the first decade of life. Usually, painful and warm soft tissue swellings are the first to appear, a stage referred to as a ‘flare-up’ (Alessi Wolken et al., 2018). Flare-ups are generally sporadic and unpredictable, and are caused by the underlying inflammation in the ligaments, tendons or skeletal muscle occurring upon pro-inflammatory insults such as muscle fatigue, tissue damage, intramuscular injections or viral illness. Over time, owing to the repeated flare-ups at different sites, progressive and cumulative ossification of soft tissues occurs, leading to the debilitating effects of FOP (Kaplan et al., 2012; Pignolo et al., 2013).

**Atypical features**

In addition to the two classical signs (hallux valgus and macrodactyly), some FOP patients present with one or more atypical signs and are categorised as FOP-plus. These atypical signs include tibial osteochondromas (Box 1), spinal malformations and broad femoral neck, which are usually reported among patients with R206H and Q207E missense mutations, and thumb malformations, cognitive impairment and diffuse scalp thinning, which are reported among patients with other atypical missense mutations in ACVR1 (Qi et al., 2017). Several recent reports have also documented delayed-onset HO and absence of characteristic great toe malformations among patients with atypical mutations, who are thus categorised as FOP variants (Jiao et al., 2013).

**Debilitating effects**

Starting with neck stiffness and a local flare-up episode, the cervical spine becomes involved early during the course of the disease (Pignolo et al., 2011). At this stage, radiographs of the cervical spine might reveal large posterior elements, tall and narrow vertebral bodies and fused facet joints. Eventually, neck movements become completely restricted due to bridging bone formation across segments, referred to as bony ankylosis (Box 1) (Katagiri et al., 2018b). Similar episodes occur in no particular order throughout the body, resulting in ankylosis of various joints, and most patients become wheelchair bound by the end of the second decade of life (Kaplan et al., 2010). Some patients may experience hearing loss due to middle ear ossification (Levy et al., 1999). The involvement of the jaw can lead to feeding difficulties, resulting in malnourishment and gradual weight loss, which can be addressed with feeding assistance (Pignolo et al., 2011). However, the debilitating effects of FOP start to become life threatening with the involvement of intercostal muscles, costovertebral joints and thoracic paravertebral soft tissues, which, when ossified, result in thoracic insufficiency syndrome (Box 1) (Kaplan and Glaser, 2005).

Patients eventually die due to resulting complications such as pneumonia or right-sided heart failure. FOP patients have a median life span of around 40 years (Kaplan et al., 2010).
Table 1. Current treatment concepts (Kaplan et al., 2019)

| Class | Status | Drug family | Examples clinically used | Mechanism of action | Notes | References |
|-------|--------|-------------|--------------------------|---------------------|-------|------------|
| 1     | In use for restraining acute-phase inflammation in FOP | Corticosteroids | Prednisolone | Activation of glucocorticoid receptor; decreases recruitment of inflammatory mediators, and inhibits vasodilation and increased vascular permeability during inflammation. | • Highly potent against initial inflammatory response. • Effective when administered within 24 h of the onset of a flare-up and for up to 4 days. • Systemic side effects should be considered. • Treatment should be restricted to early stages of flare-up. | (Pignolo et al., 2013, Kaplan et al., 2019) |
|       |        | Non-steroidal anti-inflammatory drugs (NSAIDs) | Ibuprofen, Indomethacin | Non-specific inhibition of COX-1 and COX-2 (also known as PTGS1 and PTGS2), involved in the synthesis of prostaglandins (responsible for pain, inflammation and fever); these drugs possess analgesic, anti-inflammatory and antipyretic properties. | • NSAIDs can be started after discontinuation of corticosteroids. | (Qi et al., 2017, Kaplan et al., 2019) |
| 2     | Approved for other indications but with theoretical application to FOP | Leucotriene receptor antagonist (LRA) | Montelukast | Inhibits cysteinyl leukotriene-1 (CysLT1; also known as CYSLTR1) receptor involved in inflammation; commonly used for treating asthmatic patients. | • Should be used with caution in patients with cardiovascular or renal dysfunctions. | (Schaper et al., 2011) |
|       |        | Mast cell stabilisers | Cromolyn | Inhibits degranulation of mast cells and prevents release of histamine and related mediators; normally used for treating allergies. | • Could be of use to prevent spread of inflammation in FOP. • Prolonged use might be needed to achieve therapeutic effects. | (Convente et al., 2018, Brennan et al., 2018) |
|       |        | Selective thysosine kinase inhibitors | Imatinib | Inhibits HIF1-α, PDGFRα, c-KIT (also known as KIT) and multiple MAP kinases; induces mast cell apoptosis; potent chemotherapy agent. | • Attacks multiple targets during the early hypoxic stages of inflammatory flare-ups. | (Kaplan et al., 2019, Werner et al., 2013) |
|       |        | Aminobisphosphonates | Pamidronate | Immunomodulating effect; inhibits cytokine secretion by macrophages, decreases micro-vessel density, affects bone mineralisation and, at high doses, suppresses calcification. | • Should not be used in those with renal dysfunctions or hypocalcaemia. | (Kaplan et al., 2019, Brantus and Meunier, 1998, Pennanen et al., 1995, Pabst et al., 2014) |
| 3     | Under clinical trials for FOP | Mammalian target of rapamycin (mTOR) inhibitors | Rapamycin | Inhibits non-canonical ACVR1 signal transduction in chondrocytes. | • Under phase II clinical trial. | (Hino et al., 2017, IFOPA, 2020) |
|       |        | Monoclonal antibody against activin A | REGN 2477 (Garetosmab) | Binds and blocks activin A. | • Under phase III clinical trial. | (IFOA, 2020, Hatsell et al., 2015) |
|       |        | Retinoic acid receptor gamma (RARγ) agonists | Palovarotene | Inhibits downstream SMAD signalling, promotes SMAD destruction and prevents chondrogenesis. | • Under phase III clinical trial. | (IFOA, 2020, Sheng et al., 2010, Chakkalakal et al., 2016) |
abdominal wall and paravertebral regions, with concomitant skeletal deformities and restriction of movements (Kan et al., 2004; Kan and Kessler, 2011). Other approaches to inducing ectopic BMP activity, such as knocking out BMP inhibitors or overexpressing BMP target genes, however, failed to phenocopy FOP, and subsequent models have focused on the expression of mutant ACVR1 (Kan and Kessler, 2011).

Functional orthologues of human ACVR1 are found throughout the animal kingdom, and expression of the classical or atypical FOP mutant forms of the receptor has been shown to cause dysregulated BMP signalling in several laboratory animals, including Drosophila, mouse, chick and zebrafish (Twombly et al., 2009; Chakkalakal et al., 2012; Haupt et al., 2014; LaBonty and Yelick, 2018) (Table 2). Among these commonly used animals, the mouse ACVR1 is 98.4% identical to its human orthologue, whereas the zebrafish ACVR1 (Acvr1l) shows only 69% identity. However, the GS and kinase domains of the zebrafish receptor are more similar (85% identity) (Yelick et al., 1998; LaBonty and Yelick, 2018), validating the use of both these model organisms for studying FOP. Because normal ACVR1 activity is required for gastrulation, neural crest differentiation and germ cell development in mouse, and for dorsal-ventral patterning in zebrafish, embryonic expression of the FOP-associated mutation results in lethality (Chakkalakal et al., 2012; LaBonty et al., 2017). To circumvent this problem, conditionally expressed mutant forms of ACVR1 have been developed to model FOP in both mouse and zebrafish (Table 2).

### Table 2. Established mouse and zebrafish models conditionally expressing the FOP-associated mutant ACVR1 receptor

| Animal       | Model characteristics                                                                 | Phenotypes                                                                 | References                                    |
|--------------|----------------------------------------------------------------------------------------|-----------------------------------------------------------------------------|-----------------------------------------------|
| Mouse        | Localised expression of ACVR1<sup>Q207D</sup>, driven by a ubiquitous CAG promoter, was induced by removal of an intervening floxed-lacZ expression cassette via injection of adenoviral Cre recombinase into the hindlimbs of transgenic mice on postnatal day 7. | • Initially, mononuclear infiltrates and myofibre oedema were seen in the injected side gastrocnemius, soleus and hamstring muscles, indicating myositis.  
  • On postnatal day 30, radiographs revealed bony calluses circumferentially encasing the tibia and fibula, often fused with the pelvis and femur, causing restriction of movement in the injected limb. | (Yu et al., 2008, Fukuda et al., 2006) |
| Mouse        | Global expression of ACVR1<sup>Q207D</sup> driven by a ubiquitous CAG promoter was induced by removal of an intervening floxed-lacZ expression cassette via ubiquitously expressed Cre<sup>ER</sup>, activated by tamoxifen injection on postnatal day 7. | • Did not lead to detectable HO by postnatal day 60.  
  • However, injection of a control adenovirus on postnatal day 8, following tamoxifen on day 7, resulted in ectopic calcifications by day 14, leading to decreased range of movement. This suggests that inflammation or injury from viral immunogenicity or cytotoxicity might stimulate bone formation. | (Yu et al., 2008) |
| Knock-in mouse model expressing the classical FOP mutation p.R206H generated by precise homologous recombination to replace one of the two normal copies of the mouse Acvr1 gene by the mutant allele. | • Heterozygotes for this mutant allele are perinatal lethal, but chimeric animals with 70-90% mutant cells survive and exhibit features of FOP.  
  • Almost 50% of them demonstrate first digit malformations in the hind limbs.  
  • By 6-8 weeks of age, most of the animals had extensive HO of endochondral origin, causing ankylosis of major joints of both the axial and appendicular skeleton.  
  • In addition to spontaneous HO, the animals also developed HO noted at 6 weeks post intramuscular cardiotoxin injection, thus substantiating the role of tissue injury and inflammation in FOP.  
  • Early stages of HO showed muscle destruction, inflammatory infiltration and fibroblast proliferation at the involved sites.  
  • From as early as 2 weeks, spontaneous and progressive HO (attached with normal bone) was seen in the regions of sternum, caudal vertebra, hip joint and hind limbs, causing restriction of movement. | (Chakkalakal et al., 2012) |
| Mouse        | Mouse carrying a conditional-on knock-in allele of Acvr1<sup>R206H</sup> in which the mutant-coding exon is cloned in the antisense strand. Expression of the mutant allele is achieved through Cre-mediated transposition of the mutant exon into the sense strand and simultaneous deletion of the corresponding wild-type exon, by crossing to appropriate Cre drivers. | • All of the heat-shocked transgenic fish displayed some degree of spinal lordosis, in distinct contrast to the slightly kyphotic normal spinal curvature of zebrafish.  
  • Some of the fish developed small HO lesions just behind the dorsal fin and single vertebral fusions.  
  • Distinct malformation of both pelvic fins was also absorbed in one fish.  
  • However, inflammatory triggers such as activin A injection, cardiotoxin injection or caudal fin clip injury did not result in HO at the site of injury; rather, these fish developed HO at distant sites, such as the body cavity and along the spine, which could have occurred irrespective of the trigger.  
  • This effect was attributed to the functional differences between the artificial p.Q204D Acvr1l variant and the naturally occurring human FOP mutations; alternatively, it could reflect the biology of wound healing in zebrafish. | (Hatsell et al., 2015) |
| Zebrafish    | Transgenic zebrafish in which the acvr1<sup>Q204D</sup> allele can be conditionally expressed using a heat-inducible promoter. Such fish complete embryogenesis normally, allowing expression of the mutant receptor to be induced in juveniles or adults by heat shocking once daily for 1 h at 38°C. | •Almost 50% of them demonstrate first digit malformations in the hind limbs.  
  • By 6-8 weeks of age, most of the animals had extensive HO of endochondral origin, causing ankylosis of major joints of both the axial and appendicular skeleton.  
  • In addition to spontaneous HO, the animals also developed HO noted at 6 weeks post intramuscular cardiotoxin injection, thus substantiating the role of tissue injury and inflammation in FOP.  
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  • This effect was attributed to the functional differences between the artificial p.Q204D Acvr1l variant and the naturally occurring human FOP mutations; alternatively, it could reflect the biology of wound healing in zebrafish. | (LaBonty et al., 2017, 2018) |
These transgenic animal models have paved the way not only to identifying the cell of origin and the pathomechanisms of FOP, but also to implementing further pre-clinical testing of novel medical interventions. The discovery that retinoic acid receptor (RAR) agonists can prevent the stimulatory effect of RARs on SMAD-mediated transcription (Lees-Shepard et al., 2018; Chakkalakal et al., 2016; Shimono et al., 2011; Sheng et al., 2010) prompted testing of palovarotene, a RAR agonist, in animal models. Its efficacy in repressing chondrogenesis, cartilage formation and subsequent HO was initially demonstrated in vivo in two of the FOP models described above, the Cre-inducible constitutively active ACVR1Q207D mouse model and the genetically humanised conditional-on knock-in mouse harbouring the classical ACVR1R206H mutation (Fukuda et al., 2006; Chakkalakal et al., 2016). Work in juvenile FOP mice also showed that daily dosing with palovarotene prior to skeletal maturity could result in long bone growth plate ablation, suggesting that the developmental stage, duration of exposure and dosing interval need to be optimised for safe and effective use of palovarotene without complications (Lees-Shepard et al., 2018). Another concept of inhibiting activin A with a blocking antibody, the basis of the clinical candidate REGN2477 (garetosmab), was first demonstrated in the genetically humanised conditional-on knock-in mouse model of FOP that showed neither spontaneous nor injury-mediated HO (Hatsell et al., 2015). Furthermore, Rapamycin, which is currently the subject of a clinical trial, was tested in ACVR1R206H mice and in a FOP-induced pluripotent stem cell-based HO model in which ectopic bones derived from FOP patient-derived cells are formed in mice. In both models, treatment with Rapamycin reduced HO (Agarwal et al., 2016; Hino et al., 2017). Overall, animal studies have contributed significantly to the understanding and management of FOP. Moreover, the need for accurate animal models remains, as more therapeutic modalities that target and regulate multiple mechanisms of the BMP signalling cascade in FOP are constantly being designed.

Future directions

FOP research is progressing towards translational success. Animal models have helped unravel its pathobiology: it is now evident that inflammation, dysregulation of BMP signalling and endochondral ossification are key processes contributing to HO in FOP patients. One focus of current research is on discovering more ways to redirect the progenitor cells in the inflammatory environment away from adopting an osteogenic fate towards more of a soft tissue fate. However, there is also a debate as to whether aberrant BMP signalling is solely responsible for HO in FOP (Kan et al., 2018). Given the role of Hedgehog (Hh) signalling, mediated predominantly via Indian Hh in normal osteogenesis, especially the differentiation of chondrocytes during endochondral ossification (Martelli and Santos, 2014; Lai and Mitchell, 2005; St-Jacques et al., 1999; Long et al., 2004), it is plausible that this pathway may also contribute to HO in FOP, which has yet to be explored in detail. Similarly, the Wnt/β-catenin signalling pathway, which is thought to influence the differentiation and function of mesenchymal stem cells, chondrocytes, osteoblasts and osteoclasts during normal bone formation, may also have a role (Regard et al., 2012). Although mutations in these pathways have not been found in patients with FOP, it is possible that they could influence HO through crosstalk with aberrant BMP signalling seen in FOP. Indeed, a recent study demonstrated that genetic removal of Hh can abolish HO in mouse models, not only POH, but also FOP and acquired HO. The authors identified ‘Hedgehog-driven, self-amplifying osteoblast differentiation as a common cellular and molecular mechanism underlying HO initiation and expansion’, suggesting a new therapeutic focus (Yang, 2020). The possible involvement of Hh and Wnt signalling pathways in FOP and other forms of HO certainly warrants further investigation.

Conclusion

The aetiology of FOP has long been an unsolved puzzle; however, years of extensive effort are bringing us closer to a full understanding of this distressing and debilitating condition. Exploration of FOP has informed our understanding of the BMP signalling cascade, cellular response to inflammation and connective tissue metamorphosis, leading to the development of promising new therapeutic strategies that are the subject of ongoing clinical trials. Since lesions in acquired HO disorders such as MOT and NHO often present a combination of endochondral ossification, as in FOP, and intramembranous ossification, as in POH, progress in understanding and treating these rare diseases could hold the key for developing management strategies and effective treatment for the far more prevalent, yet clinically puzzling, acquired forms of HO. Progress in understanding FOP has been made possible through the growing FOP research network that has overcome the fundamental challenges of rare diseases: creating awareness, maintaining patient registries, providing education and support for patients and families, conducting clinical trials and sharing information. In this way, FOP research provides a model for other rare disease communities to overcome these challenges through active collaboration between patients, researchers and advocates.

Competing interests

The authors declare no competing or financial interests.

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References

Agarwal, S., Loder, S., Brownley, C., Cholok, D., Mangiavini, L., Li, J., Breuler, C., Sung, H. H., Li, S., Ranganathan, K. et al. (2016). Inhibition of Hif1alpha prevents both trauma-induced and genetic heterotopic ossification. Proc. Natl. Acad. Sci. USA 113, E338-E347. doi:10.1073/pnas.1515397113
Alessi Wolken, D. M., Idone, V., Hatsell, S. J., Yu, P. B. and Economides, A. N. (2018). The obligatory role of activin a in the formation of heterotopic bone in fibrodysplasia ossificans progressiva. Bone 109, 210-217. doi:10.1016/j.bone.2017.06.011
Barruet, E., Morales, B. M., Cain, C. J., Ton, A. N., Wentworth, K. L., Chan, T. V., Moody, T. A., Haks, M. C., Ottenhoff, T. H., Hellman, J. et al. (2018). NF-kappaB/MAPK activation underlies ACVR1-mediated inflammation in human heterotopic ossification. JCI Insight 3, e122958. doi:10.1172/jci.insight.122958
Bouajt, G., Choquet, R., Bouée, S., Jeanbart, V., Couroule, L., Ruel, A., Michot, C., Le Quan Sang, K.-H., Lapidus, D., Messiain, C. et al. (2017). Prevalence of fibrodysplasia ossificans progressiva (FOP) in France: an estimate based on a record linkage of two national databases. Orphanet J. Rare Dis. 12, 123. doi:10.1186/s13023-017-0674-5
Billings, P. C., Fiori, J. L., Bentwood, J. L., O’Connell, M. P., Jiao, X., Russbaum, B., Caron, R. J., Shore, E. M. and Kaplan, F. S. (2008). Dysregulated BMP signalling and enhanced osteogenic differentiation of connective tissue progenitor cells from patients with fibrodysplasia ossificans progressiva (FOP). J. Bone Miner. Res. 23, 305-313. doi:10.1359/jbmr.071030
Brantus, J. F. and Meunier, P. J. (1998). Effects of intravenous etidronate and oral corticosteroids in fibrodysplasia ossificans progressiva. Clin. Orthop. Relat. Res. 117-120. doi:10.1097/00002183-199810000-00017
Brennan, T. A., Lindborg, C. M., Bergbauer, C. R., Wang, H., Kaplan, F. S. and Pignolo, R. J. (2018). Mast cell inhibition as a therapeutic approach in fibrodysplasia ossificans progressiva (FOP). Bone 109, 259-266. doi:10.1016/j.bone.2017.08.023
associated osteonecrosis of the jaws: influence on microvessel sprouting in an in vivo 3D Matrigel assay. *Clin. Oral Investig.* 18, 1015-1022. doi:10.1007/s00784-013-1060-x

Pennanen, N., Lapinjoki, S., Urtti, A. and Monkkonen, J. (1995). Effect of liposomal and free bisphosphonates on the IL-1 beta, IL-6 and TNF alpha secretion from RAW 264 cells in vitro. *Pharm. Res.* 12, 916-922. doi:10.1023/ A:1016821608773

Petrie, K. A., Lee, W. H., Bullock, A. N., Pointon, J. J., Smith, R. R. G., Brown, M. A., Wordsworth, B. P. and Triffitt, J. T. (2007). Novel mutations in ACVR1 result in atypical features in two fibrodysplasia ossificans progressiva patients. *PLoS ONE* 4, e5005. doi:10.1371/journal.pone.0005005

Pignolo, R. J. and Kassem, M. (2011). Circulating osteogenic cells: implications for injury, repair, and regeneration. *J. Bone Miner. Res.* 26, 1685-1693. doi:10.1002/jbmr.370

Pignolo, R. J., Suda, R. K. and Kaplan, F. S. (2005). The fibrodysplasia ossificans progressiva lesion. *Clinic. Rev. Bone Miner Metab.* 3, 195-200. doi:10.1385/BMM:3:3-4:195

Pignolo, R. J., Shore, E. M. and Kaplan, F. S. (2011). Fibrodysplasia ossificans progressiva: clinical and genetic aspects. *Orphanet J. Rare Dis.* 6, 80. doi:10.1186/1750-1172-6-80

Pignolo, R. J., Shore, E. M. and Kaplan, F. S. (2013). Fibrodysplasia ossificans progressiva: diagnosis, management, and therapeutic horizons. *Pediatr. Endocrinol. Rev.* 10 Suppl. 2, 437-448.

Pulik, L., Mierzewijski, B., Ciemerych, M. A., Brzoska, E. and Legosz, P. (2020). The survey of cells responsible for heterotopic ossification development in skeletal muscles-human and mouse models. Cells 9, 1324. doi:10.3390/cells9061324

Qi, Z., Luan, J., Zhou, X., Cui, Y. and Han, J. (2014). Smad9 is a new type of transcriptional regulator in bone morphogenetic protein signaling. *Sci. Rep.* 4, 7596. doi:10.1038/srep07596

Tombowy, V., Bangi, E., Le, V., Malnic, B., Singer, M. A. and Wharton, K. A. (2009). Functional analysis of saxosine, the Drosophila gene encoding the BMP type I receptor ortholog of human ALK1/ACVR1 and ACVR1/LAK2. *Genetics* 183, 563-579. 191-851. doi:10.1534/genetics.109.105585

Wang, R. N., Green, J., Wang, Z., Deng, Y., Qiao, M., Peabody, M., Zhang, Q., Ye, J., Yan, Z., Denduluri, S. et al. (2014). Bone Morphogenetic Protein (BMP) signaling in development and human diseases. *Genes Dis.* 1, 87-105. doi:10.21037/gd.2014.07.005

Wang, H., Lindborg, C., Louvne, V., Kim, J. H., Mccarrick-Walmsley, R., Xu, M., Mangiavini, L., Groppe, J. C., Shore, E. M., Schipani, E. et al. (2016). Cellular hypoxia promotes heterotopic ossification by Amplifying BMP signaling. *J. Bone Miner. Res.* 31, 1652-1665. doi:10.1002/jbmr.2848

Trugil, M., Ayadi, K., Zribi, M., Triki, Z. and Keskes, H. (2011). Fibrodysplasia ossificans progressiva: diagnosis and surgical management. *Acta Orthop. Belg.* 77, 139-144.

Tsukamoto, S., Mizuta, T., Fujimoto, M., Ohke, S., Osawa, K. and Miyamoto, A. (2010). Yoneyama, K., Murata, E., Machiya, A., Jimi, E. et al. (2014). Smad9 is a new type of transcriptional regulator in bone morphogenetic protein signaling. *Sci. Rep.* 4, 7596. doi:10.1038/srep07596

Turner, J. B., Zhong, Z., Williams, B. O. and Yang, Y. (2012). Wnt signaling in bone development and disease: making stronger bone with Wnts. *J. Bone Miner. Res.* 27, 1004-1017. doi:10.1002/jbmr.1562

Yang, Y. (2020). Genetic and acquired heterotopic ossification are driven by a self-amplifying positive feedback loop of Hedgehog signaling. *FASEB J.* 34, 1-1.

Yelick, P. C., Abduljabbar, T. S. and Stashenko, P. (2011). Anti-inflammatory properties of montelukast, a leukotriene receptor inhibitor reduces heterotopic ossification. *J. Bone Miner. Res.* 26, 1652-1665. doi:10.1002/jbmr.2848

Zou, Q., Luan, J., Cai, L., Zhou, X., Deng, Y., Qiao, M., Peabody, M., Zhang, Q., Ye, J., Yan, Z., Denduluri, S. et al. (2014). Bone Morphogenetic Protein (BMP) signaling in development and human diseases. *Genes Dis.* 1, 87-105. doi:10.21037/gd.2014.07.005

Werner, C. M., Zimmermann, S. M., Wurgler-Hauri, C. C., Lane, J. M., Wanner, G. A. and Simmen, H. P. (2013). Use of imatinib in the prevention of heterotopic ossification. *HSS J.* 9, 166-170. doi:10.1142/s1174-0139-9335-y

Wosczyna, M. N., Biswas, A. A., Cogswell, C. A. and Goldhamer, D. J. (2012). Multipotent progenitors resident in the skeletal muscle interstitium exhibit robust BMP-dependent osteogenic activity and mediate heterotopic ossification. *J. Bone Miner. Res.* 27, 1004-1017. doi:10.1002/jbmr.1562

Yamamoto, R., Matsushita, M., Kitoh, H., Masuda, A., Ito, M., Katagiri, T., Kawai, T., Ishiguro, N. and Ohno, K. (2013). Clinically applicable antianginal agents suppress osteoblastic transformation of myogenic cells and heterotopic ossifications in mice. *J. Bone Miner. Metab.* 31, 26-33. doi:10.1007/s00774-012-0380-2

Yang, Y. (2020). Genetic and acquired heterotopic ossification are driven by a self-amplifying positive feedback loop of Hedgehog signaling. *FASEB J.* 34, 1-1.

Yelick, P. C., Abduljabbar, T. S. and Stashenko, P. (2011). Anti-inflammatory properties of montelukast, a leukotriene receptor inhibitor reduces heterotopic ossification. *J. Bone Miner. Res.* 26, 1652-1665. doi:10.1002/jbmr.2848

Yelick, P. C., Abduljabbar, T. S. and Stashenko, P. (1998). zALK-8, a novel type I serine/threonine kinase receptor, is expressed throughout early zebrafish development. *Dev. Dyn.* 211, 352-361. doi:10.1002/sdj.1079804211

Yu, R. B., Deng, D. Y., Lai, C. S., Hong, C. C., Cuny, G. D., Bouxein, M. L., Hong, D. W., Mcmanus, P. M., Katagiri, T., Sachidanandan, C. et al. (2008). BMP type I receptor inhibition reduces heterotopic ossification. *Nat. Med.* 14, 1363-1369. doi:10.1038/nm.1888