Short communication

The proteasomal and apoptotic phenotype determine bortezomib sensitivity of non-small cell lung cancer cells

Jens Voortman*1, Agnieszka Chęcińska1,2 and Giuseppe Giaccone1,3

Address: 1Department of Medical Oncology, VU University Medical Center, 1081HV Amsterdam, the Netherlands, 2Department of Dermatology, University of Michigan, 4217 Comprehensive Cancer Center, 1500 East Medical Center Drive Ann Arbor, MI 48109-0314, USA and 3Medical Oncology Branch, National Cancer Institute, National Institutes of Health, 10 Center Drive, Bethesda, MD 20892-1906, USA

Email: Jens Voortman* - j.voortman@vumc.nl; Agnieszka Chęcińska - agnieszk@med.umich.edu; Giuseppe Giaccone - giacconeg@mail.nih.gov
* Corresponding author

Abstract

Bortezomib is a novel anti-cancer agent which has shown promising activity in non-small lung cancer (NSCLC) patients. However, only a subset of patients respond to this treatment. We show that NSCLC cell lines are differentially sensitive to bortezomib, IC50 values ranging from 5 to 83 nM. The apoptosis-inducing potential of bortezomib in NSCLC cells was found to be dependent not only on the apoptotic phenotype but also on the proteasomal phenotype of individual cell lines. Upon effective proteasome inhibition, H460 cells were more susceptible to apoptosis induction by bortezomib than SW1573 cells, indicating a different apoptotic phenotype. However, exposure to a low dose of bortezomib did only result in SW1573 cells, and not in H460 cells, in inhibition of proteasomal activity and subsequent apoptosis. This suggests a different proteasomal phenotype as well. Additionally, overexpression of anti-apoptotic protein Bcl-2 in H460 cells did not affect the proteasomal phenotype of H460 cells but did result in decreased bortezomib-induced apoptosis. In conclusion, successful proteasome-inhibitor based treatment strategies in NSCLC face the challenge of having to overcome apoptosis resistance as well as proteasomal resistance of individual lung cancer cells. Further studies in NSCLC are warranted to elucidate underlying mechanisms.

Findings

Intracellular protein metabolism involves both synthesis as well as degradation of proteins. The vast majority of proteins is selectively degraded by the ubiquitin-proteasome system (UPS). The 26S proteasome complex is composed of a 20S "core", a large protein complex that harbours the proteolytically active sites, and 19S "caps", which play a role in the recognition of poly-ubiquitinated protein substrates, targeted for degradation. Poly-ubiquitination involves covalent ligation of the target protein, by the sequential action of three enzymes, to a chain of ubiquitin molecules [1].

Growing awareness of the pivotal role of the UPS in normal cell physiology as well as in (malignant) disease propelled the development of proteasome inhibitors for therapeutic applications [2-4]. Bortezomib (Velcade) is the first clinically approved small molecule proteasome inhibitor. It reversibly binds and inhibits the chymotryptic-like proteolytic activity of the proteasome, localized
within the $\beta_5$ subunit of the 20S core. This results in disturbance of intracellular protein homeostasis by accumulation of poly-ubiquitinated proteins. Among other effects, this can trigger apoptosis, with a relative selectivity for malignant as opposed to normal cells [3].

Lung cancer is the most common cause of cancer-related death in the world [5]. Non-small cell lung cancer (NSCLC) consists of epithelial tumors, accounting for approximately 80% of lung carcinomas. Clinical studies showed promising activity of bortezomib in a subset of patients with non-small cell lung cancer (NSCLC) [6,7]. So far the exact molecular mechanism of resistance of NSCLC to bortezomib remains unclear [8-11]. Recently, adaptation of leukaemia and lymphoma cells to continuous exposure of bortezomib was reported to result from increased expression and altered subunit composition of the proteasome [12,13]. Furthermore, resistance was shown to be associated with increased expression of anti-apoptotic Bcl-2 family member proteins and heat shock proteins [14-17]. In this study, we examined the effects of bortezomib in a panel of NSCLC cell lines. We observed differential activity of bortezomib in NSCLC cells regarding growth inhibition, the proteasome activity profile and apoptosis induction.

We first determined the anti-proliferative effect of a concentration range of bortezomib by MTT assay in a panel of NSCLC cell lines (Figure 1A). We observed differential sensitivity to bortezomib-induced growth inhibition, with IC$_{50}$ values ranging from 5 to 83 nM. The p53 status did not correlate with sensitivity (Figure 1B). Next, to establish the apoptosis inducing potential of bortezomib, a sensitive cell line, SW1573, and a resistant cell line, H460, were selected for treatment and subsequent PI staining-based FACS analysis to determine the subG$_1$ apoptotic fraction of the cell population. Additionally, the fractions of cells in the G$_1$, S or G$_2$M phase of the cell cycle were assessed.

As shown in Figure 2, upon exposure of SW1573 cells to a low concentration of bortezomib, 10 nM, cells went into G$_2$M cell cycle arrest, whereas H460 cells remained predominantly in the G$_1$ phase of the cell cycle. This finding suggested that the proteasome activity in H460 cells is not inhibited at 10 nM of bortezomib, as proteasome inhibition typically results in G$_2$M arrest [18,19]. Furthermore, G$_2$M arrest in SW1573 and H460 cells was associated with increased expression of Mcl-1, which is known to be upregulated upon proteasome inhibition [15,20]. As shown in Figure 2, Mcl-1 is only upregulated in H460 cells when exposed to bortezomib at a high concentration (100 nM), whereas Mcl-1 is upregulated in SW1573 cells also at a low concentration of bortezomib (10 nM). Concordantly, treatment with bortezomib 10 nM for 48 h led to significant apoptosis in SW1573 cells (18%), but not in H460 cells. However, at a higher dose of bortezomib (50 to 100 nM), both SW1573 and H460 cells went into G$_2$M arrest, resulting after 48 h in more apoptotic H460 cells (49%) compared to SW1573 cells (32%). Thus, upon effective induction of G$_2$M cell cycle arrest, the apoptosis inducing potential of bortezomib was more pronounced.
in H460 cells compared to SW1573 cells, indicating a different apoptotic phenotype. In contrast, SW1573 cells went into G2M arrest at a lower threshold concentration of bortezomib than H460 cells, also indicating a different proteasomal phenotype of these two cell lines.

In order to further evaluate the mechanism of resistance to bortezomib, proteasome activity was measured upon treatment with various concentrations of bortezomib in H460, SW1573 and H460-Bcl-2 cells. H460-Bcl-2 cells, overexpressing anti-apoptotic factor Bcl-2, have a more apoptosis resistant phenotype compared to the parental H460 cells [15].

As shown in Figure 3, the basal level of proteasome activity was about 10% lower in SW1573 cells compared to H460 and H460-Bcl-2 cells. Furthermore, at 10 nM of bortezomib, proteasome inhibition occurred exclusively in SW1573 cells. In contrast, at 100 nM of bortezomib the level of proteasome inhibition was similar in all cell lines. Overexpression of Bcl-2 did not alter the basal proteasome activity nor its inhibition induced by bortezomib [15].

These observations confirmed a differential proteasomal phenotype comparing H460 and SW1573 cells and indicate that the proteasomal phenotype is independent of the apoptotic phenotype.

Bortezomib predominantly inhibits the chymotryptic-like activity of the 20S proteasome, harboured in the $\beta_5$ subunit, and to a lesser extent the caspase-like activity harboured in the $\beta_1$ subunit [21,22]. We therefore next evaluated whether the relative sensitivity for proteasome
inhibition of SW1573 cells correlated with a decreased expression level of the 20S proteasome and its individual catalytically active subunits, β1, β2 and β5. As shown in Figure 3B, the expression level of the β1, β2 and β5 subunits was slightly increased in H460 cells compared to SW1573 after 24 h exposure to bortezomib (10, 100 nM). Additionally, exposure to bortezomib induced expression of subunit β1 in H460 cells as compared to untreated control. In agreement with our results, increased expression of proteasome subunits, as well increased (basal) proteasome activity, was recently related to resistance to proteasome inhibitor treatment [12].

To examine the possibility that apoptotic factors might explain the differential sensitivity for apoptosis induction once the proteasome has been effectively inhibited, we assessed the expression of heat shock proteins, such as Hsp27, Hsp70 and Hsp90 upon exposure of H460 and SW1573 cells to bortezomib. Notably the expression of Hsp27, but also Hsp90, was higher in SW1573 cells than in H460 cells, correlating with less pronounced apoptosis in SW1573 cells. The expression levels of Hsp70 was similar in both cell lines.

NSCLC cell lines display differential sensitivity towards the proteasome inhibitor bortezomib. For effective induction and execution of apoptosis at least two conditions must be met. First, the dose of bortezomib must be above the threshold necessary to inhibit the proteasome activity in individual NSCLC cells. We showed that this threshold concentration differs among non-small cell lung cancer cells. Typically in clinical studies, proteasome inhibition...
in peripheral blood mononuclear cells is taken as a surrogate pharmacodynamic marker. These results indicate however that this might not be a fair estimate of what is occurring in the actual tumour tissue, as the proteasomal phenotype of individual tumours is subject to significant variability [23,24].

Secondly, upon inhibition of the proteasome, an intrinsic or acquired resistance to apoptosis must be overcome, the apoptotic phenotype being variable in different NSCLC cell lines. In this regard, apoptosis resistance does not directly correlate with resistance resulting from a differential proteasomal phenotype, as was shown in SW1573 cells, which were more sensitive to proteasome inhibition, but more resistant to apoptosis than H460 cells. Additionally, H460 cells overexpressing anti-apoptotic Bcl-2 showed a similar proteasomal phenotype as did wild type H460 cells. Increased apoptosis resistance of SW1573 cells compared to H460 cells might result from increased expression of Hsp27 in SW1573 cells, which has been postulated to be a resistance factor for bortezomib-induced cell death [14].

It was recently suggested in a mice model of colon carcinoma, that potentiation of the anticancer activity of bortezomib by combination with TNF-α resulted from reduced expression of proteasome subunits and inhibition of Hsp27 [25]. Breast carcinoma cells were shown to be more resistant to treatment with bortezomib than normal breast cells, despite a higher level of feedback proteasome upregulation in the normal cells. The authors suggested resistance of the cancer cells compared to the normal cells was primarily due to a defective pathway of bortezomib-induced apoptosis in the cancer cells [11]. Another report suggested resistance to bortezomib can occur in lymphoma cells through an altered proteasomal phenotype coinciding with increased expression and altered subunit composition of proteasome [12]. Our preliminary results show that the proteasomal as well as apoptotic phenotype determines bortezomib sensitivity in NSCLC cells. There is a preclinical rationale to combine proteasome inhibition with pro-apoptotic agents as well as agents promoting a more favourable proteasomal phenotype to overcome this resistance [20,26].

**Materials and methods**

**Cell lines and drugs**

NSCLC cell lines H292, H460, H1299, H322, A549, H1703 and SW1573 were obtained from the American Type Culture Collection (Manassas, VA, USA) and cultured in RPMI1640 or, SW1573 only, DMEM (Cambrex Bioscience, Verviers, Belgium). H460 cells stably overexpressing Bcl-2 (H460-Bcl-2) were generated previously [27]. Culture mediums were supplemented with 10% FCS, 100-units/ml penicillin, and 100-μg/ml streptomycin (Invitrogen, Breda, The Netherlands). Bortezomib (Velcade™) (Millennium Pharmaceuticals Inc. Cambridge, MA, USA) was dissolved in DMSO.

**Growth inhibition assay**

Growth inhibition was determined 72 h after treatment with a concentration range of bortezomib, by MTT assay, as described previously [15]. Results are presented as percentage of survival taking the control (untreated cells) as 100% survival. The concentration resulting in 50% of cell-growth inhibition (IC$_{50}$) was calculated using SigmaPlot version 8.0 software (SPSS Inc., Chicago, IL, USA).

**Flow cytometric analysis of PI-stained cells**

Propidium iodide (PI) staining and flow cytometry analysis were performed as described previously [15]. The fraction of cells with hypodiploid DNA content was considered as the apoptotic cell population. Cell cycle fractions were estimated using WinMDI Version 2.9 (The Scripps Research Institute, La Jolla, CA, USA) and Cylchred (Cardiff University, Cardiff, UK) version 1.0.2 software.

**Proteasome activity assay**

The chymotryptic activity of the proteasome was estimated as described previously [15], using succinyl-Leu-Leu-Val-Tyr-AMC substrate (Bachem, King of Prussia, PA, USA). Fluorescence of the released 7-amido-4-methylcoumarin dye was measured on a SpectraFluor multwell plate reader (Tecan, Salzburg, Austria), set at an excitation wavelength 380 nm and emission wavelength 460 nm.

**Western blot analysis**

Western blot analysis was performed as described before [15]. Rabbit polyclonal antibodies used were: anti-Hsp70 (Affinity Bioreagents, Golden, CO, USA), anti-Hsp90 (Cell Signaling, Danvers, MA, USA), anti-20S proteasome core, anti-subunit β$_1$, β$_2$, β$_5$ (Biomol, Plymouth Meeting,
PA, USA), anti-Mcl-1 (clone 22, BD PharMingen, San Diego, CA, USA), mouse monoclonal antibodies used were: anti-Hsp27 (Cell Signaling), anti-β-actin (Sigma-Aldrich, St Louis, MO, USA).

Competing interests The author(s) declare that they have no competing interests.

Authors' contributions JV conceived of the studies, carried out experimental assays, analyzed data and drafted the manuscript. AC carried out experimental assays and drafted the manuscript. GG initiated the studies and drafted the manuscript. All authors read and approved the final manuscript.

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