Digital Imaging Mass Spectrometry

Running title: Digital Imaging Mass Spectrometry

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Abstract

Methods to visualize the two-dimensional distribution of molecules by mass spectrometric imaging evolve rapidly and yield novel applications in biology, medicine, and material surface sciences. Most mass spectrometric imagers acquire high mass resolution spectra spot-by-spot and thereby scan the object’s surface. Thus, imaging is slow and image reconstruction remains cumbersome. Here we describe an imaging mass spectrometer that exploits the true imaging capabilities by ion optical means for the time of flight mass separation. The mass spectrometer is equipped with the ASIC Timepix chip as an array detector to acquire the position, mass, and intensity of ions that are imaged by MALDI directly from the target sample onto the detector. This imaging mass spectrometer has a spatial resolving power at the specimen of (84±35) µm with a mass resolution of 45 and locates atoms or organic compounds on a surface area up to ~2 cm². Extended laser spots of ~5 mm² on structured specimens allowed parallel imaging of selected masses. The digital imaging mass spectrometer proves high hit-multiplicity, straightforward image reconstruction and its potential for high-speed readout at 4 kHz or more. This device demonstrates a simple way of true image acquisition like a digital photographic camera. The technology may enable a fast analysis of biomolecular samples in near future.
Introduction

Imaging mass spectrometry is a widely used tool for detecting the molecular composition of sample surfaces [1-6] with a two-dimensional resolution in the µm range [7]. The technology has been proven valuable for many fields of application, ranging from diagnostics in microchip production to analysis of tissue samples. Mass spectrometric images visualize the spatial distribution of molecules with a set of four parameters: two for the spatial dimensions, one for molecules’ molecular mass, and one for its abundance.

The commonly used imaging methodology is scanning or micro probing, which first records mass and intensity of molecules or atoms spot-by-spot, and then reconstructs their two-dimensional distribution in a mass spectrometric image [8]. Here, an alternative approach is presented: The two-dimensional distribution of molecules or atoms is acquired in the first step, and their abundance is accumulated by subsequent measurement cycles. The advantage is the parallel acquisition of mass spectrometric data from an extended sample surface [9]. A separation of masses with the time of flight (TOF) principle favors this purpose, since the spatial information can be mapped onto the detector through ion optics. For example, this approach is realized in mass spectrometers of the TRIFT series (Physical Electronics) in the stigmatic mode: Blankers in the ion path set a narrow m/z-range and the spatial distribution of ions is derived from a read out of a phosphor screen by a CCD camera [10, 11]. Alternatively, delay-line read out may be utilized to record the time of flight of spatially separated ions in a single measurement cycle [12]. However, array detectors of high multiplicity offer an attractive and straight-forward realization.
Indeed, the recent development of a time sensitive, highly pixilated detector (Timepix), which was first conceived for 3D-reconstruction of particle tracks [13, 14] and is meanwhile used for photon detection [15, 16], provides a perfect solution. Herein we describe, for the first time to our knowledge, the application of the Timepix detector for imaging mass spectrometry. Recently, the precursor to the Timepix, the Medipix was tested for its application to mass spectrometric imaging, although lacking temporal resolving capabilities [17]. The Timepix detector covers a sensitive surface of 14 × 14 mm² with a matrix of 256 × 256 pixels. Each pixel occupies 55 × 55 µm² and detects either charge through time over threshold (TOT) or the time of arrival (TIME) of a signal.

For a proof of principle a linear time of flight mass spectrometer was assembled. Its basic elements were a MALDI driven ion source, a drift space with a focusing ion optics, a multi channel plate (MCP), and the Timepix. The electron signal generated by ions at the MCP covers several pixels of the Timepix, which allows for neighboring pixels to record TOT and TIME in an alternating fashion. As a benefit the simultaneous recording of the electron signal with several pixels allows the reconstruction of the time of arrival with higher precision [18].

The following set of experiments shows the feasibility of this digital mass imaging approach. The mass resolution and imaging properties of the Timepix are presented and the results are discussed within the scope of possible improvements for biological and biomedical applications.
Material and Methods

The Time of Flight Mass Spectrometer and one Event Cycle

The imaging mass spectrometer apparatus comprises a N\textsubscript{2} laser with optics, sample plate with MALDI analyte, drift volume, and detection system as depicted in Figure 1a and described in supplemental materials and methods. The recipient, the acceleration stage (73 mm), the drift space (117 mm) and the MCP, which is part of the detection system, have been retrieved and modified from an experimental setup used previously to analyze the dynamics of molecular interactions [19].

The laser pulse set the reference time and triggered the shutter gate for the Timepix chip (see supplemental material and methods for detailed description of the Timepix). The logic for each pixel provided optionally two modes of operation TIME and TOT (Figure 1b). A shutter gate of fixed length (~10 µs) was applied at the time the laser had fired accepting a charge signal above threshold. From this moment on the 13.5 bit register stored the number of clock cycles (clock speed between 70 and 80 MHz) until either the end of the shutter gate was reached (TIME) or the charge signal dropped below threshold (TOT).

In the final experiment with the extended laser area the grid terminating the drift space was removed to avoid possible interference of micro-lensing with experiments using a highly structured target, thereby slightly modifying the ion optics (Supplemental materials and methods, and supplemental Figure 1).
Data Analysis

The events were processed with MATLAB 7 (The Mathworks) separating the two modes TIME and TOT and searching for contiguous areas of hits originating from ion signals, subsequently called clusters (see also supplemental materials and methods). The intensity of the ion signal was determined from the sum of the TOT counts in the cluster and the spatial position from the centre of gravity. All clusters, above a predetermined cluster size, were stored along with its spatial position, signal intensity, and time of arrival data for further analysis.

Results

In order to show the applicability of the Timepix detector for biomolecular imaging mass spectrometry, the imaging mass spectrometer was equipped with focusing ion optics for imaging purposes. An initial set of mass spectra was acquired without activation of the Einzel lens, i.e. in a non-focusing mode and without the diaphragm. MALDI-matrix embedded Cesium (m/z ~133) or Caffeine (m/z ~194), or a mixture of both was deposited on the target sample plate. Depending on the energy of the UV-laser pulse, multiple Caffeine or Cesium ions were extracted simultaneously by MALDI from the target sample in one event.

Cluster Shapes and Multiplicity

As expected for MALDI ionization, the number of ions detected varied depending on laser intensity and location on the target sample plate. Events recording up to 200 ions resulted in mostly well-separated clusters generated from individual ions as shown in Figure 2a. It demonstrates the capability of the device to record events with high ion multiplicity, which is limited only by excessive overlap of
clusters. The separation of clusters depends on the cluster size and the multiplicity (see supplemental results). Small cluster sizes are preferred in order to achieve high multiplicities resulting from increased laser intensities. Because the events shown are recorded in non-focusing mode of the mass spectrometer, the spatial distribution of clusters reflects the dispersion of ions caused by the initial transverse momentum obtained during the MALDI ionization process.

For each recorded event the number of observed clusters was determined. Here more than five adjacent, recording pixels were required for a cluster. Figure 2b depicts a close-up of the matrix TIME for several clusters recorded. The clusters in the TIME matrix have column-like shape with a flat plateau extending over several pixels due to the simultaneous arrival time of the electrons. The corresponding clusters in the TOT matrix are shaped in Gaussian profiles reflecting the two-dimensional projection of the cloud of electrons leaving the MPC (Figure 2c). The event of Figure 2a is displayed in a three-dimensional representation in Figure 2d, which visualizes that the Caffeine signal is well separated from background.

**Acquisition of Mass Spectra**

The m/z value for each detected ion was calculated from the corrected TIME signal (“time walk” correction, see supplemental results and supplemental Figure 3a) and all m/z values were plotted in a histogram to obtain the final mass spectrum for a series of events recorded (Figure 3a). In this example, two significant peaks appear, which correspond to single-charged Caffeine (m/z ~195) and Iron (m/z ~56), which represents an ionization by-product from the stainless steel backing of the sample.
In a TOF mass spectrometer, the resolution and the accuracy of mass
determination are dependent on (1) the dimensions of the mass analyzer and (2) the
precision of the time to digital converter. In order to assess the influence of these
parameters for the small time of flight mass spectrometer presented, the shape and
position of the Caffeine peak was determined by fitting a Gaussian to the data points
(Figure 3b). The average time of flight for Caffeine was 9.610±0.138 µs and a signal
dispersion of $\Gamma = 115$ ns. Based on the full width at half maximum (FWHM) for
Caffeine, a mass resolution of 45 m/Δm was determined at m/z 195 (Figure 3b). It
should be mentioned that the resolution was not limited by the clock speed (see
supplemental results) and no delayed extraction has been applied which is well known
to increase mass precision. The overall calibration of the mass spectrometer is
demonstrated using three different ions, namely Iron, Cesium and Caffeine
(supplemental Figure 3b).

**Spatial Resolution of the Detector System**

As shown in Figure 1a the field-free section for the ion drift is terminated by a
grounded grid, followed by the post acceleration stage of ~2.5 kV. As a result of the
field gradient, micro-lensing occurred at the detector plane causing the Cesium ion
intensities to peak in a periodic pattern when observed over a large number of events
(Figure 3c). Using this image of the grid structure, it was possible to estimate the
spatial resolution of the combined MCP-Timepix detector system. The analysis of the
Cs$^+$ ion intensity pattern with properly chosen angle of projection revealed periodic
intensity maxima with a step size of 0.22 mm, which closely matched the periodicity
of the grid. Based on the lateral discrimination capability determined from the Cesium
intensity pattern, the detector provides an estimated spatial resolution of $\sigma = 36 \, \mu m$.

Indeed, the spatial resolution is smaller than the lateral extension of a single detector
pixel (55 $\mu m$), which is due to the fact that the center of gravity is determined with
cluster sizes being spread over several pixels.

The imaging capability of the detector section only was visualized in an
additional experiment: A copper-coated PC board with 0.2 mm pinholes separated by
$\geq 0.6$ mm depicting the letters of the word “IMAGE” was positioned near the grid at
the end of the drift tube. The passage of mass selected Cs ions through the pinholes
reveals the word IMAGE on the detector plane with $10^3$ events (Figure 3d).

This first set of experiments showed that a mass spectrometric detector based
on the Timepix is capable of spatially resolving a distribution of ions while
determining m/z values at the same time. The analysis of individual clusters recorded
in large numbers with each event allowed a fast data acquisition and collection of
mass spectra. The simultaneous measurement of TIME and TOT can improve the
precision in the m/z-measurement introduced by the finite rise time of the input
circuit.

**Imaging of Laser Spots on the Target Surface**

The instrument provided also a two-dimensional resolution at the target side,
which is a prerequisite for imaging. As mentioned above an Einzel lens mapped the
ion-optical image of the target plate onto the detector surface. A SIMION 7 simulation
verified that the ion optical image was de-magnified by a factor of 2.2 as a
consequence of the acceleration stage and the Einzel lens being positioned near the
front end of the drift volume. A diaphragm (Ø 6 mm) was introduced near the
entrance grid of the drift tube to keep astigmatic distortions small. Following activation of the Einzel lens, Caffeine-ions generated with a laser spot of small size were focused by the ion optical system onto the detector system in a point like spot (supplemental Figures 4a, b).

To show the imaging capabilities of the mass spectrometer Cs$^+$ ions were generated by MALDI at two distinct positions on the target plate by displacing the laser spot. First, the optical focusing of the laser was checked for its quality by estimating its visual transverse extension at the target plate. It was found to be less than $0.3 \times 0.3$ mm$^2$. After recording the mass spectrum at the first position, the laser spot was moved by displacing the lens vertically along the y-axis of the target plate by $\sim 1.0$ mm. Due to geometric constraints for the mounting of the Timepix chip its axes are rotated by $135^\circ$ relative to the target reference system. Therefore, the imaged spot was displaced in both axes of the Timepix reference frame, – by $0.3$ mm along the x-axis (Figure 4a) and $0.2$ mm along the y-axis (Figure 4b). The resulting ion optical image for Cs$^+$-ions was shifted diagonally by about $0.37$ mm on the Timepix detector with an average standard deviation of the Gaussian fits for both spots of $38.1$ µm. Concluding from the distinct positions of two spots the imaging mass spectrometer can resolve two spatially separated positions at the sample surface.

Subsequently we determined the spatial resolution and its error with respect to the sample plate. To this end, a more refined estimation of the average standard deviation was performed taking into account, that micro-lensing occurs at the detector stage as described above. By considering the maximum contribution of micro-lensing to the change of the spot width, the resolution with a systematic error was determined
to $(38.1\pm16.0) \, \mu m$. Due to the demagnification of the system the resolution at the sample plate achieved in this experiment is $(84\pm35) \, \mu m$. A possible contribution due to the MPCs’ finite channel diameter of $<10 \, \mu m$ is neglected. Whereas a spatial resolution of the digital imaging mass spectrometer is demonstrated in this experiment, the precision of the spatial resolution might be affected by additional factors such as the extension and spatial intensity variations of the laser spot on the target sample plate.

**Imaging of a Two-dimensional Structure on the Target by Extended Illumination**

The following set of experiments addresses the simultaneous imaging of an extended target plate area in macroscopic scale. For an illumination of an extended surface, the Nitromite N$_2$-laser was replaced by a high power N$_2$-laser providing effective beam energy up to 2 mJ. This allowed for an increase of the illuminated area to an approximate rectangle of $\sim(5 \times 2) \, \text{mm}^2$ (see supplemental Figure 5b).

The first experiment assessed the general imaging capability of the mass spectrometer with the grid at the end of the drift volume in place. The target consisted of Cesium embedded in a MALDI matrix deposited on a circular plateau of 3 mm $\varnothing$, which is separated from the remainder of the flat target plate by a circular groove of 1 mm width (Figure 5a, inset). During target preparation the matrix crystallized preferentially close to the rims of the groove. Imaging of the macroscopic structure on the target plate was achieved along the major axis of the defocused UV-laser beam directly, and along the minor axis by step-wise displacement of the beam between each measurement cycle while the number of measured events was kept equal and the
ion optics unaltered. All data acquired was merged to one dataset, which finally reproduced the original ring-like deposition of Cs on the sample plate for the Cs$^+$ line in the mass spectrum (Figure 5a). The same experimental procedure revealed an even distribution of Iron background from an ion image taken from the untreated flat stainless steel plate (supplemental Figure 5a). The spatial frequency distribution of Cs$^+$ signals correlates with the density of Cs$^+$/MALDI-matrix as observed by a light image of the target plate surface. Note that the signal of Cs$^+$ shows a rarified intensity in the center of the disk similar to the Cesium deposit seen in the photograph of the target. The correlation observed between the initial deposition of Cesium and the mass spectrometric image indicates that the digital imaging mass spectrometer can be used to collect information about the spatial distribution of molecules or atoms in a quantitative manner.

In a second experiment, the spatial resolution was tested by imaging isolated spots of analyte deposited onto a stainless steel plate. The setup was modified so micro-lensing was suppressed (See supplemental material and methods, and supplemental Figure 1b). A MALDI target with a regular spacing of spots was produced by ink jet printing [20] of Cesium-containing MALDI matrix onto the stainless steel target plate. Dots were deposited in a rectangular grid with a pitch of 1.6 mm onto the Fe backing (Figure 5b, inset). Figure 5b shows the spatial distribution of the Cesium-MALDI-matrix initially deposited by the printer (white spots). The nominal regular structure of Cesium depositions (yellow disks) and the spatial distribution of Cesium signal recorded by the Timepix detector (red dots) is superimposed on the photograph of the sample plate. The laser image was displaced on the target once by ~0.5 mm during
image acquisition and the Iron background signal for one of the two laser positions is shown in supplemental Figure 5b. Regions of high Cesium content closely match the original depositions of the Cesium-MALDI-matrix mixture.

Both sets of experiments show the feasibility to image two-dimensional samples with an extended laser spot and the mass spectrometric images reproduce the initial spatial distributions of the analyte on the target sample plate. The beam energy per pulse limited the sampled surface to about 5-10 mm$^2$. Therefore, we employed the scanning method in order to cover still larger surfaces, but ionization sources with higher energy fluxes are readily available from other laser sources e.g. tripled NdYAG–lasers.

**Discussion and Conclusion**

In summary, the Timepix chip was successfully operated as an array detector in a small linear TOF mass spectrometer. The mass spectrometer imaged the spatial distribution of atoms and small organic molecules with sufficiently high mass resolution. It demonstrated that the acquisition of mass spectrometric images is possible with ion detection using front-end digitization and highly pixilated sensors. The Timepix offers the capability of high event rates up to 4 kHz [21] and allows very fast data acquisition with an appropriate interface for data transfer. The experimental setup presented here was not optimized for fast data acquisition, since more advanced USB-based readout modules are currently being developed [22].

The scanning method and delay line readout approach realized in conventional mass spectrometric imaging obtains positional information indirectly. In contrast, highly pixilated detectors directly record all four parameters necessary to acquire mass
spectrometric images (position, m/z value, and intensity) thereby omitting cumbersome reconstruction means. Key advantages are (1) multi-hit capability up to 200 ions per event with (2) a lateral resolution of 36 µm of the detector stage. We achieved a lateral resolution of 84 µm at the target sample plate, which is mainly constrained by the de-magnifying ion optical system presently used. It should be stressed that the ion optical demagnification in this setup is unfavorable for demonstrating a high resolution power on the sample due to both the extended acceleration stage and as well as the small drift distance. There are ion optical focusing systems with multiple lenses available [23], which would provide higher magnifications thereby increasing the spatial resolution at the target.

Timepix chips can be seamlessly attached at three sides to expand the detector surface (e.g. quadruplets or higher) thereby increasing either the overall multi-hit capability of the system or the spatial resolution at the target depending on the setting of the ion optics. Although the multi-hit capability of a pixel detector clearly offers the advantage of parallel ion detection, it seems to be limited by the need of a finite cluster size in the mixed mode operation to reconstruct an accurate m/z value and position of the ion signal. However, data acquisition in purely TIME sensitive mode offers a spatial reconstruction marginally worse than with a mixed mode operation. In order to get a time walk correction, the number of pixels in TOT-mode may be reduced in favor of an increased time precision. Moreover, variations in cluster size at small MPC signals are not negligible and more sophisticated algorithms for the separation of partially overlapping clusters are developed.
We estimated the speed and limits of data taking based on the multiplicities of registered ions, which might well be in the region of several hundred per laser shot as shown. If only one dominant ion species would be analyzed, the data rate increases approximately 200 fold per laser shot, provided space-like overlap of ion-induced clusters would be acceptable. Taking into account a maximum laser repetition rate of ~4 kHz, the total improvement in data acquisition speed would be a factor 1000 when compared with conventional micro probing. For the Timepix the data volume is 2 Gb/s, which can be handled easily for transfer and storage. The data volume to be processed is sizable, since the parameters of all clusters per readout cycle have to be determined. Typically such a task can be handled by computing farms using optimized algorithms.

In fact, the data volume would be higher when allowing also the detection of ions with different arrival times impinging at the same position of the detector. However, the Timepix is designed in principle with a single hit feature. Of course a Timepix with multi-hit capability for all pixels would eventually avoid the application of narrow mass windows (see also supplemental discussion).

Modern MALDI scanning mass spectrometric imagers achieve a spatial resolution of typically ~150 \( \mu \)m [24] at the sample-stage dictated by the step size. Using a differential scanning method a resolution of less than 10 \( \mu \)m has been demonstrated [7]. With appropriate sample ionization, the digital imaging mass spectrometer presented here can achieve comparable spatial resolution. The spatial resolution at the sample depends on the ion optics of the imaging TOF mass spectrometer. Indeed, time of flight mass spectrometers with a magnification up to 300
are realized in commercially available instruments of the TRIFT-series. The combination of appropriate ion optics with the highly parallel detection principle will promote digital imaging mass spectrometry as high throughput instrumentation in the future.

The digital imaging TOF as presented here is limited in spectral resolution \((m/\Delta m\text{ at 195 m/z})\). There are two essential reasons: the short drift path (0.12 m) and the lack of velocity focusing. The digital imaging mass spectrometer was not equipped to correct for initial differences in kinetic energy with the delayed extraction principle, which in general improves spectral resolution. In addition, the time to digital converter (TDC) in the Timepix is restricted to \(~100\text{ MHz}\), a factor 100 lower than the time resolution of modern TDCs used in non-imaging TOF mass spectrometers. Therefore, spatial resolution rather than spectral resolution using the Timepix opens an interesting field of mass spectrometric applications.

The MALDI process was adopted to generate ions on an extended surface because MALDI efficiently ionizes biomolecules and is well employed together with the time of flight mass spectrometry for imaging applications [25]. It is not excluded that the MALDI process has some intrinsic limit for the spatial resolution [26] as well as a transformation of the analyte by crystallization seems to be necessary. With respect to imaging applications, the efficiency and reproducibility of the sample ionization is dependent on many variables introduced during sample preparation and ionization (see supplemental discussion). Another field of application is SIMS using fullerenes, where an improvement of spatial resolution with ion optical imaging and digital readout with the Timepix is expected.
In conclusion, the digital mass spectrometer presented here proved the principle of highly parallel mass spectrometric image acquisition. Thereby it opens the door for high throughput sample analysis by imaging ions extracted from extended sample surfaces. Based on the achieved spatial ion multiplicity and the potential readout rates the true mass spectrometric imaging principle presented here allows an increase of data taking speed of several orders of magnitude over micro probing. Moreover, the technology may enable a fast analysis of samples spotted in arrays, eventually illuminated with a bundle of laser beams. Although with limited spectral resolution, the digital imaging mass spectrometer may have broad range of applications ranging from biological and medical experimentation, material sciences to forensic analysis.

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setup and data taking were performed by A.B. and C.B., manuscript was prepared by A.B. and C.B.

Legends for Figures

**Figure 1**

Schematic of a time-of-flight mass spectrometer with high multiplicity detector array and representation of one measurement cycle. (a) The major components of the time-of-flight mass spectrometer are the acceleration stage (horizontal black, thick, long-dashed lines), the drift volume with the Einzel lens between the grids (vertical dotted lines) and the detector section. The black dashed line indicates the path of ions generated off-axis and focused on the Timepix chip. The N₂-laser beam (grey arrow) releases ions by MALDI on the sample plate with a focused spot or with an extended area by adjusting the laser optics (Einzel lens). (b) The time sequence starts with opening the shutter gate triggered by the laser pulse (vertical arrowhead). TOT determines approximately the charge of the signal and TIME serves to determine the time of flight after correcting for the gate length. The black dashed line indicates the minimal charge necessary to trigger detection whereas black bars represent the opening time of the shutter, the stored number of cycles (TIME or TOT) and deduced time of flight (TOF). Numbers given to the right exemplify the measured value based on the clock cycle schematic visualized at the bottom of the graph.

**Figure 2**

Recordings by the Timepix visualize MALDI events. (a) The two-dimensional representation of the detector surface shows the TOF matrix (TIME
converted to time of arrival) for one event of high multiplicity recorded from a Caffeine sample with the Einzel lens switched off. Clusters of pixels are visualized on the detector surface according to a rainbow-colored time code. Red clusters represent Caffeine and light blue clusters indicate Iron. (b) Close up of a selected x-y surface of the detector for a single laser shot. The time stamp (TIME) visualizes plateau-like shaped clusters in units of clock cycles. (c) The charge distribution (TOT) of the same close up shows Gaussian shaped clusters. (d) Three-dimensional representation of 129 individually reconstructed clusters based on (a) in terms of time walk corrected, color-coded clock cycles.

**Figure 3**

**Digital imaging mass spectra, mass and spatial resolution, and image generating capability of the Timepix.** (a) The mass spectrum ($10^3$ events) shows two peaks (Caffeine and Iron) above background noise. Solid red curves represent Gaussian fits. (b) A Gaussian curve was fitted to the frequency histogram of the drift times measured for Caffeine. The mass resolution measured was $m/\Delta m = 45$ at $m/z \sim 195$ (Caffeine+H$^+$). (c) The frequency histogram for the Cesium ion signal visualizes the effect of micro-lensing caused by the grid at the end of the drift volume. The y-coordinate was rotated with respect to the Timepix to match the grid orientation. The peaks are fitted by multiple Gaussians (solid lines) having a resolution of 36 µm. (d) The (x,y)-plot displays the lateral distribution of Cesium ions that were acquired with a pinhole mask depicting the word "IMAGE". The mask was positioned at the end of the drift volume.
**Figure 4**

Two distant spots on the target sample plate are imaged to distinct positions onto the Timepix. Frequency histograms of Cesium ions show the displacement along the x-axis (a) and along the y-axis (b) of the Timepix reference coordinate system as the laser focus was shifted by 1.0 mm at the target. Gaussian fits represent the signal intensity (rate of events) before (red) and after (violet) displacement (arrow) of the laser focus.

**Figure 5**

Mass spectrometric images of a ring-shaped Cesium deposition and single spots containing Cesium were acquired with an extended laser spot. (a) The mass spectrometric image visualizes the distribution of Cesium deposits on the target plate (inset). The dotted black lines indicate the projection of the target surface area onto the mass spectrometric image. Each color shows one of the 8 consecutive measurements. Between each measurement, the laser spot was shifted vertically with respect to the target plate reference system. (b) A Cesium-containing MALDI sample was spotted by an inkjet printer in a regular pattern (white spots) onto a stainless steel sample plate as shown in the photograph (inset). A scaled photographic image of this sample plate was overlaid with (1) the nominal position of sample spots on the sample plate (yellow) and (2) the respective mass spectrometric image of Cesium ions (red) which were acquired from two adjacent positions consecutively illuminated with the laser beam.
Literature

1. Goodwin, R. J. A.; Pennington, S. R.; Pitt, A. R., Protein and peptides in pictures: Imaging with MALDI mass spectrometry. *Proteomics* 2008, 8 (18), 3785-3800.
2. Walch, A.; Rauser, S.; Deininger, S. O.; Hofler, H., MALDI imaging mass spectrometry for direct tissue analysis: a new frontier for molecular histology. *Histochem Cell Biol* 2008, 130 (3), 421-434.
3. Hardesty, W. M.; Caprioli, R. M., In situ molecular imaging of proteins in tissues using mass spectrometry. *Anal Bioanal Chem* 2008, 391 (3), 899-903.
4. Cornett, D. S.; Reyzer, M. L.; Chaurand, P.; Caprioli, R. M., MALDI imaging mass spectrometry: molecular snapshots of biochemical systems. *Nat Methods* 2007, 4 (10), 828-833.
5. Cooks, R. G.; Ouyang, Z.; Takats, Z.; Wiseman, J. M., Detection Technologies. Ambient mass spectrometry. *Science* 2006, 311 (5767), 1566-1570.
6. Roggli, V. L.; Ingram, P.; Linton, R. W.; Gutknecht, W. F.; Mastin, P.; Shelburne, J. D., New techniques for imaging and analyzing lung tissue. *Environ Health Perspect* 1984, 56, 163-183.
7. Spengler, B.; Hubert, M., Scanning microprobe matrix-assisted laser desorption ionization (SMALDI) mass spectrometry: instrumentation for sub-micrometer resolved LDI and MALDI surface analysis. *J Am Soc Mass Spectrom* 2002, 13 (6), 735-748.
8. Caprioli, R. M.; Farmer, T. B.; Gile, J., Molecular imaging of biological samples: localization of peptides and proteins using MALDI-TOF MS. *Anal Chem* 1997, 69 (23), 4751-4760.
9. Luxembourg, S. L.; Mize, T. H.; McDonnell, L. A.; Heeren, R. M., High-spatial resolution mass spectrometric imaging of peptide and protein distributions on a surface. *Anal Chem* 2004, 76 (18), 5339-5344.
10. Altelaar, A. F.; Luxembourg, S. L.; McDonnell, L. A.; Piersma, S. R.; Heeren, R. M., Imaging mass spectrometry at cellular length scales. *Nat Protoc* 2007, 2 (5), 1185-1196.
11. Luxembourg, S. L.; Vaezaddeh, A. R.; Amstalden, E. R.; Zimmermann-Ivol, C. G.; Hochstrasser, D. F.; Heeren, R. M., The molecular scanner in microscope mode. *Rapid Commun Mass Spectrom* 2006, 20 (22), 3435-3442.
12. Yoon, O. K.; Robbins, M. D.; Zuleta, I. A.; Barbula, G. K.; Zare, R. N., Continuous Time-of-Flight Ion Imaging: Application to Fragmentation. *Analytical Chemistry* 2008, 80 (21), 8299-8307.
13. Llopart, X. B.; R. Campbell, M. Tlustos, L. Wong, W., Timepix, a 65k programmable pixel readout chip for arrival time, energy and/or photon counting measurements. *Nuclear Instruments and Methods in Physics Research, Section A* 2007, 581 (1-2), 485-494.
14. Ballabriga, R.; Campbell, M.; Heijne, E. H. M.; Llopart, X.; Tlustos, L., The medipix3 prototype, a pixel readout chip working in single photon counting mode with improved spectrometric performance. *Ieee Transactions on Nuclear Science* 2007, 54 (5), 1824-1829.
15. Vallerga, J.; McPhate, J.; Tremsin, A.; Siegmund, O., High-resolution UV, alpha and neutron imaging with the Timepix CMOS readout. *Nuclear Instruments & Methods
in Physics Research Section a-Accelerators Spectrometers Detectors and Associated Equipment 2008, 591 (1), 151-154.
16. Anton, G.; Gebert, U.; Michel, T.; Rugheimer, T. K., A hybrid photodetector using the Timepix semiconductor assembly for photoelectron detection. Nuclear Instruments & Methods in Physics Research Section a-Accelerators Spectrometers Detectors and Associated Equipment 2009, 602 (1), 205-208.
17. Jungmann, J. H.; Macaleese, L.; Buijs, R.; Giskes, F.; de Snajer, A.; Visser, J.; Visschers, J.; Vrakking, M. J.; Heeren, R. M., Fast, High Resolution Mass Spectrometry Imaging Using a Medipix Pixelated Detector. J Am Soc Mass Spectrom 2010.
18. Bamberger, A.; Desch, K.; Renz, U.; Titov, M.; Vlasov, N.; Wienemann, P.; Zwerger, A., Resolution studies on 5 GeV electron tracks observed with triple-GEM and MediPix2/TimePix-readout. Nuclear Instruments & Methods in Physics Research Section a-Accelerators Spectrometers Detectors and Associated Equipment 2007, 581 (1-2), 274-278.
19. Lange, M.; Pfaff, O.; Muller, U.; Brenn, R., Projectile fragment-ion fragment-ion coincidences (PFIFICO) following fast ion impact on SF6. Chemical Physics 1998, 230 (1), 117-141.
20. Baluya, D. L.; Garrett, T. J.; Yost, R. A., Automated MALDI matrix deposition method with inkjet printing for imaging mass spectrometry. Anal Chem 2007, 79 (17), 6862-6867.
21. Mikulec, B.; Clark, A. G.; Ferrere, D.; La Marra, D.; McPhate, J. B.; Siegmund, O. H. W.; Tremsin, A. S.; Vallerga, J. V.; Clement, J.; Ponchut, C.; Rigal, J. M., A noiseless kilohertz frame rate imaging detector based on microchannel plates read out with the Medipix2 CMOS pixel chip. Nuclear Instruments & Methods in Physics Research Section a-Accelerators Spectrometers Detectors and Associated Equipment 2006, 567 (1), 110-113.
22. Platkevic, M.; Bocarov, V.; Jakubek, J.; Pospisil, S.; Tichy, V.; Vykydal, Z., Signal processor controlled USB2.0 interface for Medipix2 detector. Nuclear Instruments & Methods in Physics Research Section a-Accelerators Spectrometers Detectors and Associated Equipment 2008, 591 (1), 245-247.
23. Harting, E.; Read, F. H.; Brunt, J. N. H., Electrostatic lenses. Elsevier Scientific Pub. Co.: Amsterdam ; New York, 1976; p viii, 322 p.
24. McDonnell, L. A.; Heeren, R. M. A., Imaging mass spectrometry. Mass Spectrometry Reviews 2007, 26 (4), 606-643.
25. Caldwell, R. L.; Caprioli, R. M., Tissue profiling by mass spectrometry - A review of methodology and applications. Molecular & Cellular Proteomics 2005, 4 (4), 394-401.
26. Knochenmuss, R., A quantitative model of ultraviolet matrix-assisted laser desorption/ionization. Journal of Mass Spectrometry 2002, 37 (8), 867-877.
Figures

(a) Schematic diagram showing the setup with the laser pulse interacting with the acceleration stage, drift tube, and detection system.

(b) Graph showing the voltage (V(q)) over time with markers for shutter, TOT, TIME, and TOF, along with the number of clock cycles.
Figure 2

(a)

(b)

(c)

(d)
Figure 3

(a) Drift time vs. rate for Fe and caffeine.
(b) Drift time vs. rate for caffeine with M/ΔM = 45.
(c) Number of clusters/bin vs. y coord in units of pixels for Cs ions with resolution better than 36 μm and step width 0.22 mm.
(d) Cs ions image with x and y coordinates in pixels.
Figure 4

(a) Lens with diaphragm laser spot displaced by \(-1\) mm in \(y_{\text{chip}}\)-direction distance 0.2 mm on the chip.

(b) Lens with diaphragm laser spot displaced by \(-1\) mm in \(y_{\text{chip}}\)-direction distance 0.3 mm on the chip.
Supplemental Figure 1

(a) First grid at the end of the drift volume
Field gradient 0 to 2.5 kV/cm

(b) No structure without grid
detecting electrode → analog
CSA → comparator
threshold
4 bit threshold adj.
digital
synchronization memory
reference clock
mode: TIME or TOT
Supplemental Figure 3

(a) function used for time walk correction

(b) atomic number [amu]

S

Fe

Cs

caffeine

drift time [μs]
Supplemental Figure 5

(a) Fe backing w/o groove

(b) Fe background
