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Genetic, serological and biochemical characterization of *Leishmania tropica* from foci in northern Palestine and discovery of zymodeme MON-307

Kifaya Azmi¹,²,³*, Lionel Schnur², Gabriele Schonian³, Abedelmajeed Nasreddin¹, Francine Pratlong⁴, Fouad El Baidouri⁴, Christophe Ravel⁴, Jean-Pierre Dedet⁴, Suheir Ereqat¹ and Ziad Abdeen¹

Abstract

**Background:** Many cases of cutaneous leishmaniasis (CL) have been recorded in the Jenin District based on their clinical appearance. Here, their parasites have been characterized in depth.

**Methods:** Leishmanial parasites isolated from 12 human cases of CL from the Jenin District were cultured as promastigotes, whose DNA was extracted. The ITS1 sequence and the 7SL RNA gene were analysed as was the kinetoplast minicircle DNA (kDNA) sequence. Excreted factor (EF) serotyping and multilocus enzyme electrophoresis (MLEE) were also applied.

**Results:** This extensive characterization identified the strains as *Leishmania tropica* of two very distinct sub-types that parallel the two sub-groups discerned by multilocus microsatellite typing (MLMT) done previously. A high degree of congruity was displayed among the results generated by the different analytical methods that had examined various cellular components and exposed intra-specific heterogeneity among the 12 strains. Three of the ten strains subjected to MLEE constituted a new zymodeme, zymodeme MON-307, and seven belonged to the known zymodeme MON-137. Ten of the 15 enzymes in the profile of zymodeme MON-307 displayed different electrophoretic mobilities compared with the enzyme profile of the zymodeme MON-137. The closest profile to that of zymodeme MON-307 was that of the zymodeme MON-76 known from Syria. Strains of the zymodeme MON-307 were EF sub-serotype A² and those of the zymodeme MON-137 were either A⁹ or A⁹B⁴. The sub-serotype B⁴ component appears, so far, to be unique to some strains of *L. tropica* of zymodeme MON-137. Strains of the zymodeme MON-137 displayed a distinctive fragment of 417 bp that was absent in those of zymodeme MON-307 when their kDNA was digested with the endonuclease RsaI. kDNA-RFLP after digestion with the endonuclease MboI facilitated a further level of differentiation that partially coincided with the geographical distribution of the human cases from which the strains came.

**Conclusions:** The Palestinian strains that were assigned to different genetic groups differed in their MLEE profiles and their EF types. A new zymodeme, zymodeme MON-307 was discovered that seems to be unique to the northern part of the Palestinian West Bank. What seemed to be a straightforward classical situation of *L. tropica* causing anthropic CL in the Jenin District might be a more complex situation, owing to the presence of two separate sub-types of *L. tropica* that, possibly, indicates two separate transmission cycles involving two separate types of phlebotomine sand fly vector.

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Background

Leishmania major and L. tropica cause cutaneous leishmaniasis (CL) in Palestine, and L. infantum has been known to cause CL without the clinical manifestation of VL (unpublished data from Palestine). CL caused by L. infantum was also confirmed in the Mediterranean region [1]. Cases caused by L. tropica seem to occur mainly in the northern regions [2,3] and those caused by L. major mainly in the lower Jordan Valley in the vicinity of Jericho [4-6] (Figure 1a). Often they exist in close proximity despite differences in their specific biotopes and general epidemiology. Classically, L. major was and is considered to be zoonotic, involving many different species of rodents as the animal reservoir in different Old World foci, and L. tropica was considered to be purely anthropotonic with direct transmission from person to person. However, more recently it has become clear that in some cases L. tropica is zoonotic [7]. Since people travel more widely than the sand fly vectors and reservoir animals that delineate foci of these species, it is difficult to know with certainty where human subjects contracted their CL. The clinical appearance of lesions is not indicative of the aetiology and characterization and identification of the pathogen are an integral part of the diagnosis of human CL. A single effective method often suffices for the diagnosis of an infectious disease like leishmaniasis, but several species of Leishmania, causing different clinical spectra, infect humans and these must be clearly identified and differentiated. A broader characterization incorporating several different methods is more useful in achieving this and exposing variation among the infectious agents. This might be significant in prognosis and the need of specific treatment of cases. A broader characterization is also important when what
appear to be leishmanial parasites are isolated from sand flies and animals that might prove to be the vectors and reservoirs, thus, elucidating the epidemiology. *L. tropica* is a very heterogeneous species and broad characterization of its strains assumes greater importance. All of this applies to the situation existing in Palestine. The following molecular biological characterization indicates genetic variation among strains of *L. tropica*, EF serotyping indicates antigenic variation and MLEE indicates biochemical variation between enzymes fulfilling the same function.

Schwenkenbecher [3] used 21 microsatellite markers to determine the profiles of 117 strains of *Leishmania tropica* which separated into ten genetic clusters I to X. Of the 27 strains that came from Palestinian foci, 9 were isolated from CL cases living in the Jenin District (Figure 1b). These grouped in the clusters 'I: Middle East' and 'II: Asia' whereas strains from other Palestinian foci, e.g., the Jericho area and Samaria, were all assigned to the cluster 'I: Middle East' (Figure 2). To understand why two different genetic groups of *L. tropica* exist in a small area, genotypic differences were compared with phenotypic ones. Since MLMT using 21 microsatellite markers is both time- and labour-consuming, simpler and quicker methods for genotyping strains were developed. For this, nine of the previously characterized strains from the Jenin District, representing different genetic variation among strains of *L. tropica*, were identified and characterized previously by ITS1 [9] and 7SL [10] and also MLMT analysis, which uses microsatellites that are specific for the species *L. tropica* [3].

Single isolate was cultured from each case. All were treated with Pentostam before identification of their strains and the outcome of treatment is unknown. Promastigotes were grown in culture from infected skin tissue aspirates as described by Azmi [8]. Table 1 lists the 12 strains isolated and gives their geographical origin. It also lists two Israeli reference strains of *L. tropica*, ISER/IL/98/LRC-L758 and MHOM/IL/97/P963 (=LRC-L725) and one of *L. major*, MHOM/IL/67/mericoII (=LRC-L137), for comparison in identifying the strains.

Mass cultures were grown in Schneider’s Drosophila medium supplemented with 10% heat inactivated FCS with 2 mM L-glutamine, containing penicillin at 200 IU/ml and streptomycin at 200 μg/ml. Parasite DNA was extracted using the high pure PCR template purification kit (Roche Diagnostics GmbH, Mannheim, Germany).

### Strain characterization

#### Identification of leishmanial species

The three strains LRC-L883, -L1324 and -L888 were identified here by amplification and restriction fragment length polymorphism (RFLP) analysis of either the ITS1 sequence or a partial 7SL RNA gene as described previously [9,10]. The other nine Palestinian strains had been identified and characterized previously by ITS1 [9] and 7SL [10] and also MLMT analysis, which uses microsatellites that are specific for the species *L. tropica* [3]. DNA from the reference strains *L. infantum* MHOM/TN/80/IPT1 and *L. major* MHOM/TM/73/5ASKH, and from two local strains of *L. tropica*, ISER/IL/98/LRC-L758 and MOHM/IL/97/P963 (=LRC-L725), all obtained from the WHO Reference Center for the Leishmaniases, the Hebrew University-Hadassah Medical School, Jerusalem, were amplified in each reaction as positive controls. The inclusion of control samples from *L. major* and *L. infantum* was considered necessary as strains of these two species also circulate in the overall geographical region where *L. major* also causes CL in humans and *L. infantum*, though mainly associated with human cases of infantile VL, has been known to cause CL without being associated with patent visceral manifestations [1,11]. Corresponding preparations of reaction buffer without leishmanial DNA were included as negative controls.

#### Restriction fragment length polymorphism (RFLP) analysis of the kinetoplast minicircle DNA (kDNA)

PCR amplification of the leishmanial kDNA minicircle sequence from cultured promastigotes, was done according to Anders [12], using the primer pair Uni 21 (5'-GGG GTT GGT GTA AAA TAG GCC) and LmjH4 (5'-CTA GTT TCC CGC CTC CGA G) based on a minicircle sequence of *L. major* [13] and applying the PCR conditions described by Schnur [14]. The PCR

### Methods

#### Isolation and cultivation of leishmanial parasites and extraction of their DNA

Twelve patients presenting cutaneous lesions suspected of being CL were referred to clinics of The Ministry of Health in the City of Jenin for diagnosis and treatment. None had travelled abroad. Seven had a single lesion on the face, four had a single lesion on the hand and one had a lesion on the hand and also one on the leg. A
product was digested separately with either the endonuclease \textit{RsaI} or \textit{MboI} (Promega, Madison, WI), according to the manufacturer’s instructions. kDNA of \textit{L. tropica} ISER/IL/98/LRC-L758 was included as a positive reference and that of \textit{L. major} MHOM/IL/67/JerichoII (=LRC-L137) as the out-group. kDNA-RFLP results were analyzed using the RAPDistance Package version 1.04 (http://www.anu.edu.au/Bozo/software/). All the fragments seen, represented by bands, were numbered and scored: 0 for the absence of a band; 1 for its presence. Whether fragments were identical or similar was measured according to the overall profile seen after digestion with each endonuclease and the number of shared bands among each enzyme’s profiles. A dendrogram was constructed based on the combined fragmentation of the restriction enzymes \textit{RsaI} and \textit{MboI}, using the distance matrix (Neighbor joining) option. These are not bootstrap values. The values are Jaccard distance computed as binary Jaccard, which represents the measure of dissimilarity.

**Excreted factor (EF) serotyping**

This was done according to Schnur and Zuckerman [15], using standard polyclonal anti-leishmanial serotyping sera raised in rabbits and known reference EFs for comparisons and the designation of serotypes and sub-serotypes [16]. These correlate, in most cases, with leishmanial species definitions based on other distinguishing criteria. The standard serotyping sera were: 1) anti-\textit{L. tropica} LRC-L36, which reacts only with EFs of the serotype A and, through partial cross-reactivities, also permits the designation of serotype A sub-serotypes; 2) anti-\textit{L. donovani}, LRC-L52, which reacts only with EFs of the serotype B and differentiates the sub-serotypes B2 and B4. The reference EFs were those from the strains: 1) \textit{L. tropica} LRC-L36 from Baghdad, Iraq, that indicates the sub-serotype A\textsubscript{2}, specific for one of the two known serological variants of \textit{L. tropica}; 2) \textit{L. tropica} LRC-L682 from Sanliurfa, Turkey, that indicates the sub-serotype A\textsubscript{9}, specific to the other serological variant of \textit{L. tropica}; 3) \textit{L. major} LRC-L137 from the Jordan Valley, Israel, that

**Table 1 The strains of Leishmania used in this study**

| WHO code | LRC# | Geographic origin | Microsatellite Cluster\textsuperscript{a} | Subgp | Clade kDNA\textsuperscript{b} | EFserotype\textsuperscript{c} | Zymodeme\textsuperscript{d} |
|----------|------|------------------|---------------------|-------|-----------------|-----------------|-----------------|
| MHOM/PS/2002/18JnF4* | L891 | Meselya | II | B1a | A Ltro-kD1/3 | A2 | MON-307 |
| MHOM/PS/2002/20iN3* | L884 | El-Yamoon | II | B1b | A Ltro-kD1/2 | A2 | MON-307 |
| MHOM/PS/2002/52iN18* | L890 | El-Yamoon | II | B1b | A Ltro-kD1/1 | A2 | MON-307 |
| MHOM/PS/2002/5iN5* | L893 | El-Yamoon | I | A1b | B Ltro-kD2/2 | A9 | MON-137 |
| MHOM/PS/2002/87iN5* | L887 | El-Yamoon | I | A1b | B Ltro-kD2/5 | A9B4 | MON-137 |
| MHOM/IL/1997/P963* | L725 | Tiberias, Israel | I | A1a | B Ltro-kD2/1 | A9B4 | MON-137 |
| MHOM/PS/2002/79iN4* | L885 | Silat El hartheya | I | A1a | B Ltro-kD2/6 | A9B4 | MON-137 |
| MHOM/PS/2002/89iN4* | L886 | Silat El hartheya | I | A1a | B Ltro-kD2/6 | A9B4 | MON-137 |
| MHOM/PS/2002/64iN4* | L889 | Silat El hartheya | I | A1b | B Ltro-kD2/6 | A9B4 | MON-137 |
| MHOM/PS/2002/34iN4* | L882 | Silat El hartheya | I | A1b | B Ltro-kD2/3 | A9B4 | MON-137 |
| ISER/IL/1998/LRC-L758* | L758 | Kfar Adumim, Israel | I | A1b | B Ltro-kD2/1 | A9B4 | MON-137 |
| MHOM/PS/2002/31iN17 | L883 | Tubas | nd | B | Ltro-kD2/6 | A9B4 | MON-137 |
| MHOM/PS/2002/33iN17 | L1324 | Silat El hartheya | nd | B | Ltro-kD2/4 | A9B4 | nd |
| MHOM/PS/2002/35iN45 | L888 | Jenin City | nd | B | Ltro-kD2/6 | A9 | nd |
| MHOM/IL/1967/Jericho | L137 | Near Jericho | Lmj 39 | Lmj-kD1 | A1 | MON-26 |

\textsuperscript{a} Strains previously analysed by multilocus microsatellite typing (MLMT) using 21 microsatellite markers from Schwenkenbecher [3]; \textsuperscript{b} their microsatellite cluster (I and II), Subgroup (Subgp) as delineated here (Figure 2); \textsuperscript{c} kDNA PCR-RFLP; \textsuperscript{d} excreted factor (EF) serotype done according to Schnur and Zuckerman [15]; \textsuperscript{e} MLEE done according to Rioux [18] and Pratlong [19], and of \textit{L. major} MOHM/IL/67/JerichoII (=LRC-L137) using 10 microsatellite markers and taken from Al Jawabreh [39]. The Israeli strains of \textit{L. tropica} LRC-L725 and LRC-L758 were used as reference strains of \textit{L. tropica}; nd, not done.
indicates the sub-serotype A1, 4) *L. donovani* LRC-L133 from, Humera, Ethiopia, that indicates the sub-serotype B2, associated with the species *L. donovani* and *L. infantum*.

**Multilocus enzyme electrophoresis (MLEE)**

MLEE was done according to Maazoun [17] using thick starch gel electrophoresis to study the 15 enzymes listed by Rioux [18]: Malate dehydrogenase (MDH) E.C. 1.1.1.37; Malic enzyme (ME) E.C. 1.1.1.40; Isocitrate dehydrogenase (ICD) E.C. 1.1.1.42; Phosphogluconate dehydrogenase (PGD) E.C. 1.1.1.44; Glucose-6-phosphate dehydrogenase (G6PD) E.C. 1.1.1.49; Glutamate dehydrogenase (GLUD) E.C. 1.4.1.3; Diaphorase NADH (DIA) E.C. 1.6.2.2; Nucleoside purine phosphorylase 1 (NP1) E.C. 2.4.2.1; Nucleoside purine phosphorylase 2 (NP2) E.C. 2.4.2.2; Glutamate-oxaloacetate transaminase 1 (GOT1) E.C. 2.6.1.1; Glutamate-oxaloacetate transaminase 2 (GOT2) E.C. 2.6.1.1; Phosphoglucomutase (PGM) E.C. 5.4.2.2; Fumarate hydratase (FH) E.C. 4.2.1.2; Mannose phosphate isomerase (MPI) E.C. 5.3.1.8; Glucose phosphate isomerase (GPI) E.C. 5.3.1.9).

Two Israeli strains of *L. tropica* (ISER/IL/98/LRC-L758 and MHOM/IL/97/P963 = LRC-L725) belonged to the zymodeme MON-137 and one of *L. aethiopica*, and insert them into their correct zymodemal sub-groups. (a); (b); (c); (d). With the discovery of the new zymodeme MON-307 of *L. tropica*, as of this study, the branch containing the zymodemes of the strains of *L. tropica* was reconstructed to include zymodeme MON-307 and, also, a second zymodeme encompassing an Israeli strain of *L. tropica* from the Negev [14] to determine to which other zymodemes they were closely related and insert them into their correct zymodomal sub-groups.

**Taxonomical methods**

Phenetic analysis was based on 15 isoenzyme loci, 39 zymodemes and 165 characters (electromorphs). Of the 39 zymodemes of *L. tropica*, 37 are listed in Pratlong [19] and two were the new zymodemes MON-288 from Israel and MON-307 from Palestine. On the hypothesis that *Leishmania* is ‘mainly’ diploid [20], multiband patterns were considered to be heterozygous and electromorph values were duplicated. Unweighted Pair Group Method with Arithmetic mean (UPGMA), Neighbour Joining (NJ) and Wagner analysis were used to cluster the stocks. Robustness of the nodes was statistically tested by bootstrap analysis, using the phylip3.6 package [21].

**Results**

**Isolation and species identification of leishmanial parasites from suspect cases of CL**

All 12 patients presenting cutaneous lesions were diagnosed as cases of CL and parasites were isolated from all of them (Table 1).

Amplification of the 300 bp ITS1 sequence and the 185 bp 7SL RNA gene gave the same-sized products and RFLP patterns, respectively, for all 12 Palestinian strains and the two Israeli reference strains of *L. tropica* (ISER/IL/98/LRC-L758 and MHOM/IL/97/P963 = LRC-L725). These differed from those of the reference strains of *L. infantum* (MHOM/ TN/80/IPT1) and *L. major* (MHOM/ TM/73/5ASKH) (data not shown).

**Characterization of strains of *L. tropica***

**PCR-RFLP analysis of kDNA**

Amplification of the kDNA minicircle sequence of the 12 Palestinian and two Israeli reference strains of *L. tropica*, (ISER/IL/98/LRC-L758 and MHOM/IL/97/P963 = LRC-L725), gave the same-sized (872 bp) PCR product (Figure 3a), which differed from the smaller-sized (650 bp) PCR product of *L. major* (MHOM/IL/ 67/ jericho1 = LRC-L137). RFLP analysis of all fourteen PCR products from the strains of *L. tropica* after their digestion with RsaI did produce different kDNA RFLP profiles that consisted of seven to nine bands ranging in size from 800 bp to 150 bp but of two basic kinds: *Ltro*-kD1 and *Ltro*-kD2, that differed mainly by the presence of a 417 bp component in only the *Ltro*-kD2 profiles (Figure 3b). These two types of profile were reproducible in different gels and separated the strains into two clusters (Table 1 and Figure 3b and 3c).

Digestion of the PCR products from the strains of *L. tropica* with MboI also produced different kDNA RFLP profiles, consisting of three to nine bands that were of two basic kinds and congruent with the separation seen for digestion with RsaI. The RAPDistance Package version 1.04 (http://www.anu.edu.au/Bozo/software/) used to analyze the kDNA RFLP fragments seen in the profiles after digestion separately with RsaI and MboI exposed nine different genetic variants, three, i.e., *Ltro*-kD1/1-3, falling into kDNA Clade A, and six, i. e., *Ltro*-kD2/1-6, falling into kDNA Clade B (Table 1 and Figure 3c). It is not surprising that the distances shown in the dendrogram are ‘low’ since the strains are all strains of a single species, *L. tropica*.
Figure 3 (See legend on next page.)
However, the distance is greater between the two subgroups of *L. tropica* seen in the dendrogram, and the distance between both subgroups of *L. tropica* and the strain of *L. major* used as the out-group is much greater.

**Excreted factor (EF) serotyping**

Table 1 and Figure 3c list the EF sub-serotypes of the 12 Palestinian strains. Five reacted only with the anti-serotype A serum and were, therefore, serotype A strains, three being sub-serotype A$_2$ and two sub-serotype A$_9$ revealed by the partial cross-reactivity disclosing partial identity of the two types of EF and indicated by the spur forming at the junction of the two adjacent EF-antibody precipitation bands developing in the diffusion gel. The other seven reacted with, both, the anti-serotype A and the anti-serotype B sera and, therefore, produced EFs with an A and a B component, and were of the mixed sub-serotype A$_B$B$_4$ (Figure 4). The presence of either a sub-serotype A$_2$ or a sub-serotype A$_9$ component in an EF indicates that a strain is one of *L. tropica*. Also, the presence of a B$_4$ component in an EF seems to indicate that the strain is one of *L. tropica*, and, in fact, the presence of a B$_4$ component in an EF has, so far, always been accompanied by the presence of an A$_9$ component, conferring the mixed sub-serotype A$_B$B$_4$ on strains producing this type of EF, in both, this study (Table 1) and in all the other strains with the B$_4$ component that have been serotyped.

**Multilocus enzyme electrophoresis (MLEE)**

Seven (LRC-L882, -L883, -L885, -L886, -L887, -L889, -L893) of the ten Palestinian strains examined by MLEE had enzyme profiles identical to those of the two Israeli reference strains (ISER/IL/98/LRC-L758 and MHOM/IL/97/P963 = LRC-L725), showing that they belonged to the zymodeme MON-137 [14]. A novel finding was that the other three strains (LRC-L884, -L890, -L891) were identical in their enzyme profiles and constituted a new zymodeme, zymodeme MON-307. Ten of the electrophoretic mobilities of the 15 enzymes in their enzyme profile were different from those of the strains in the

![Figure 3 The kDNA PCR and RFLP products of the 12 Palestinian *L. tropica* strains. a. The kDNA PCR products of three of the 12 Palestinian strains: LRC-L890, -L891, -L887, and reference strains of *L. tropica*, -L758, and *L. major*, -L137; MW: molecular weight marker ΦX174 DNA/Hinf. b. The RFLP patterns of the six different genotypes resulting from digestion of kDNA PCR products with RsaI. MW: molecular weight marker ΦX174 DNA/HaeIII Markers; c. Dendrogram based on the kDNA restriction fragment length polymorphism (RFLP) of the strains of *L. tropica* after digestion of their PCR products, separately, with the endonucleases RsaI and MboI, which yielded ten sub-genotypes that segregated into the Clade A, with kDNA sub-type *L. tropica* Ltro-kD1 and the Clade B, with kDNA sub-type *L. tropica* Ltro-kD2. It was constructed using RAPDistance Package (version 1.04) and was based on the presence or absence of the bands in each sample. The numbers on the branches represent the % of difference. The lengths of the branches indicate the degree of similarity in fingerprints among the strains of *L. tropica* relative to the strain of *L. major*, MHOM/IL/67/Jericho II (=LRC-L137), which served as an out-group in this analysis: Ts = Tubas; Ma = Meselya; Yn = El-Yamoon; Jn = Jenin; S-h = Silat El hartheiya; Tb = Tiberias; KA = Kfar Adumim. The zymodomal designations and EF serotypes are given for complete comparison.

![Figure 4 Leishmanial excreted factor (EF)-antibody precipitation patterns. Leishmanial excreted factor (EF)-antibody precipitation patterns between serotype A and serotype B anti-leishmanial specific sera and serotype A and serotype B EFs of different sub-serotype: 1 and 2, EF sub-serotype A reactions; 3, EF sub-serotype B reactions. Serotype A EFs react only with anti-serotype A sera. Serotype B EFs react only with anti-serotype B sera. EFs that react with, both, anti-serotype A and anti-serotype B sera are of mixed AB serotype where both the A and the B component can be of different sub-serotype, giving various mixed AB sub-serotypes, e.g., A$_B$B$_4$ as in some of the strains of *L. tropica* studied. Variation in the style of banding patterns and the formation of spurs (s) at the junctions of adjacent precipitation bands indicate incomplete identity of EFs of the same serotype, relegating them to different sub-serotypes of the serotype. EFs of the sub-serotypes A$_1$ and A$_4$ are produced by *L. major*. The diffusion preparations were stained with Coomassie blue and dried before photographing.](image-url)
zymodeme MON-137 (Table 2). For strains of the zymodeme MON-307, the enzyme MPI occurs as two electromorphs MPI110 and MPI100, only one of which, MPI110, is shared by the enzyme profile of strains of the zymodeme MON-137. This can be explained by the existence of two alleles each of which determines an isoform of the enzyme. These are identical in some cases, giving just one band, and in others not, giving two bands.

In the reconstructed branch of the zymodemes of *L. tropica* (see under Methods for MLEE), the new zymodeme of *L. tropica*, zymodeme MON-307, fell into the subgroup (b) (Figure 5); and, within the subgroup (b) itself, the strains in zymodeme MON-307 were closest to those in the zymodeme MON-76, which were isolated in Syria. The electrophoretic mobilities of only three of the 15 enzymes, MDH, GLUD and FH, differed between the profiles of the two zymodemes (Table 2).

### Table 2 The zymodemaal profiles of the strains of *Leishmania tropica*, indicating the specific differences between zymodemes encountered in Israel and the Palestinian authority

| Enzyme | Country | MDH | ME | ICD | PGD | G6PD | GLUD | DIA | NP1 | NP2 | GOT1 | GOT2 | PGM | FH | MPI | GPI |
|--------|---------|-----|----|-----|-----|------|------|-----|-----|-----|------|------|-----|----|-----|-----|
| MON    | Iraq    | 100 | 95 | 100 | 93  | 82   | 95   | 110 | 450 | 100 | 135  | 90   | 100 | 100| 76  |
|        | Lebanon |     |    |     |     |      |      |     |     |     |      |      |     |    |     |
| MON    | Syria   | 112 | 95 | 100 | 93  | 82   | 95   | 110 | 450 | 100 | 135  | 90   | 108 | 110| 76  |
|        | Palestine| 100 | 95 | 100 | 93  | 82   | 80   | 110 | 450 | 100 | 135  | 90   | 108 | 105| 76  |
| MON    | Israel  | 116 | 95 | 100 | 95  | 82   | 95   | 100 | 450 | 100 | 135  | 90   | 108 | 100| 76  |
|        | -275    | 100 |
| MON    | Israel  | 112 | 95 | 100 | 94  | 82   | 95   | 100 | 450 | 100 | 135  | 90   | 108 | 100| 76  |
|        | -288    | 100 |
| MON    | Israel  | 112 | 95 | 100 | 100 | 82   | 80   | 120 | 450 | 90  | 135  | 90   | 100 | 110| 76  |
|        | -54     | 100 |
| MON    | Jordan | 100 | 100 | 100 | 98  | 85   | 80   | 100 | 460 | 110 | 140  | 85   | 108 | 100| 76  |
|        | -265    | 100 |
| MON    | Palestine| 100 | 110 | 100 | 98  | 85   | 80   | 100 | 450 | 110 | 140  | 85   | 88  | 100| 76  |
|        | -137    |    |
|        | Jordan  | 130 | 108 | 100 | 140 | 82   | 95   | 100 | 700 | 120 | 127  | 110  | 123 | 105| 53  |
|        | Egypt   |     |
| MON    | *L. aethiopica* | 160 | 88  | 100 | 122 | 94   | 200  | 100 | 400 | 90  | 110  | 110  | 118 | 79 | 150 |
|        | -14     |     |
| MON    | *L. major* | 130 | 108 | 100 | 140 | 82   | 95   | 100 | 700 | 120 | 127  | 110  | 123 | 105| 53  |
|        | -26*    |     |

This is modified from Schnur [14] to include the new zymodeme of *L. tropica* MON-307 and the zymodeme MON-265 associated with strains of *L. tropica* from the focus just north of the Sea of Galilee [32], but previously found associated with strains of *L. tropica* from Qadisieh in the Tafila area south-east of the Dead Sea in Jordan [36]. The enzyme profiles associated with the zymodemes MON-6, MON-76, MON-275, MON-288, MON-54 are those of other strains of *L. tropica* from the Middle East for comparison when viewing the dendrogram of the zymodemes of *L. tropica* in Figure 5. The profile representing the species *L. aethiopica* is that of the WHO reference strain *L. aethiopica* MHOM/ET/72/L100. The enzyme profile of *L. major* MHOM/IL/67/JerichoII from the Jordan Valley belonging to the zymodeme MON-26 was used as an out-group. (Note: There are three errors in the electrophoretic mobilities quoted in Table 4 in Schnur et al. (2004) [14]: in the profile given for MON-275, the mobility of FH should be 100 not 110; in the profile given for MON-26, the mobility of FH should be 79 not 72, and the mobility of MPI should be 150 not 50). They are corrected here.

*The out-group is a geographically widely distributed zymodeme of *L. major* [19].

### Discussion

*Leishmania tropica* is a very heterogeneous species and its intraspecific micro-heterogeneity has been demonstrated by several investigators in different ways and examining various parasite constituents: EF serotyping using rabbit polyclonal anti-leishmanial sera [15] and anti-leishmanial specific mouse monoclonal antibodies [16]; ITS genotyping [9]; single strand conformation polymorphism (SSCP) of the ITS1 sequence [14,22]; DNA fingerprinting [14,22]; MLEE [18,19,23]; kDNA [12]; PPIP-PCR [24]; MLMT [3]. In the dendrogram constructed, based on MLMT data, all of the Palestinian strains of *L. tropica* fell into the sub-groups ‘I: Middle East’, designated A1 here, and ‘II: Asia’, designated B1 here (see Figure 2 for derivation of the terms A1 and B1 and their sub-groups) [3].
The dendrogram shows the geographical distribution of the known zymodemes of *L. tropica* incorporating two new zymodemes of the species, zymodemes MON-288, comprising one strain from the L. tropica absence of enzyme characters. 39 zymodemes representative of the genetic were analyzed. Two new zymodemes were included in the data set; zymodeme MON-288, comprising one stock from the Page 10 of 14 presented by Pratlong [19] in Figure 3 of their article. Unweighted Pair Group Method with Arithmetic mean (UPGMA) phenogram obtained from the matrix of presence / absence of enzyme characters. 39 zymodemes representative of the genetic diversity of *L. tropica* were analyzed. Two new zymodemes were included in the data set; zymodeme MON-288, comprising one stock from the Negev, Israel, [14] and MON-307 (at arrow), comprising the three strains from the Jenin District. This is based on the branch encompassing the strains and zymodemes of *L. tropica* presented by Pratlong [19] in Figure 3 of their article. Unweighted Pair Group Method with Arithmetic mean

The three methods used in this study for the characterization of strains of *L. tropica*: EF serotyping; kDNA analysis; and MLEE, confirmed the main subdivision detected by MLMT. Two of these methods, EF serotyping and kDNA RFLP are however, much easier and quicker to perform. Furthermore, EF serotyping and MLEE revealed that the strains assigned to different genetic groups present different phenotypes which, possibly, reflect differences in leishmanial parasite – sand fly vector interrelationships.

Amplification of the kDNA minicircle sequence and RFLP analysis of the PCR products of the 12 Palestinian strains after their digestion with either *RsaI* or *MboI* also identified them as strains of *L. tropica*, and enabled their separation into the kDNA Clades Ltro-kD1 and Ltro-kD2, respectively. The analysis of the kDNA RFLP profiles also exposed a further level of micro-heterogeneity and differentiation whereby three different sub-types were detected in Clade Ltro-kD1 and six in Clade Ltro-kD2 (Figure 3c). Furthermore, this micro-heterogeneity among the mini-circle kDNAs showed partial congruity with the local geographical distribution of the cases from which these strains of *L. tropica* were isolated (Table 1).

The results of EF serotyping correlated with those of the molecular biological analyses just mentioned, showing that the 12 Palestinian strains were strains of *L. tropica* of two antigenic types as indicated by the serological reactions of the strains and subsequently determined EF sub-serotypes; this exposed two serological sub-groups that coincided exactly with the two sub-groups delineated by amplification of the leishmanial kDNA minicircle sequence and RFLP analysis. This serological sub-division was based on the A component of strains’ sub-serotypes: one sub-group possessing the EF sub-serotype A₀; the other the EF sub-serotype A⁴. In fact, also the presence of the B₄ component in the EF sub-serotype of some of the strains in the latter sub-group but not in other strains in this sub-group indicates a further antigenic sub-division. It is interesting to note that antiserum raised in rabbits against living promastigotes of the strain *L. donovani*, LRC-L52, according to the method of Adler [25], has never before

![Figure 5 The taxonomic relationship of the zymodemes of *Leishmania tropica*.](http://www.parasitesandvectors.com/content/5/1/121)
differentiated serotype B components of EFs into B sub-serotypes except in the case of strains of *L. tropica* of the mixed sub-serotype A\(_9\)B\(_4\) as shown in this study. Where enzyme profiles have also been determined, the B\(_4\) component seems to be associated solely with strains of *L. tropica* belonging to the zymodeme MON-137 and the B\(_4\) component, it seems, is always present together with the A\(_9\) component (Table 2 and Figure 3c).

EFs are glyco-conjugates, bearing antigenic carbohydrate moieties that confer serological specificity on the different antigenic types of leishmanial parasite, which, approximately, coincides with leishmanial species designations [16]. They are copiously produced and released by promastigotes to their environment, which is the cultural medium surrounding them during culture in *vitro* [15,26]. In nature, promastigotes grow in the mid- and foregut, and in some Latin American species also the hindgut, of female sand flies where they release EF into the lumen of the vector’s gut. This serves to protect the promastigotes from harmful effects of the sand fly’s digestive enzymes [27]. Amastigotes also produce EF but much less so [26]. In addition to producing EF, promastigotes generate lipophosphoglycan (LPG), bearing similar and, possibly, even the same antigenic carbohydrate moieties conferring serological specificity on EFs. It is anchored in the cell membrane; though some is also released and is, supposedly, one of the antigenic components in EFs. LPG appears to promote promastigote survival in the sand fly vector by facilitating attachment to the wall of the sand fly’s midgut, preventing the voiding of the promastigotes with the remnants of the digested blood meal [7,28]. LPGs extracted from the promastigotes of different leishmanial species have different molecular structures [29,30] that appear to govern leishmanial species - sand fly vector species specific attachment to the midgut wall and survival in the vector [31] as well as defining serological specificities accounting for different EF serotypes. For example, a strain of *L. tropica* from a focus just north of the Sea of Galilee, characterized as belonging to zymodeme MON-265 and EF sub-serotype A\(_4\), survived and grew in female sand flies of the species *Phlebotomus (Adlerius) arabis*icus [7], a proven vector in that focus [32], but did not survive in female sand flies of the species *Ph. (Paraphlebotomus) sergenti*, the proven vector in other Israeli foci [14]. Conversely, the same study showed that a strain of *L. tropica* from a more southerly focus near Tiberias, producing EF of the sub-serotype A\(_9\)B\(_4\) and, therefore, probably belonging to the zymodeme MON-137 (compare with Table 1) survived and grew well in females of both sand fly species. In this case, two very different vectors were involved in the transmission of the two different sub-types of *L. tropica*, supposedly, associated with ecological differences in the two habitats from which the two different sand fly species and two sub-types of *L. tropica* came. While the experimental cross-infectivity study [7] mentioned above does not impinge directly on the leishmanial strains used here, it does raise an interesting point. Since the strains of *L. tropica* of both EF sub-serotypes A\(_4\) and A\(_9\)B\(_4\) survived in the sand fly species *Ph. (A.) arabis*icus; but only the strain producing EF of the sub-serotype A\(_9\)B\(_4\) survived in *Ph. (P.) sergenti*, one might assume that the lack of survival of the strain of the EF sub-serotype A\(_4\) in *Ph. (P.) sergenti* is a parasite-based effect rather than a sand fly vector-based one. It would be interesting to identify the sand fly vector(s) transmitting the two sub-types of the Palestinian strains of *L. tropica* described in this study, to establish if they are transmitted by the same vector or if each sub-type of *L. tropica* has its own species or even just sub-species of the same vector. However, despite the strain of *L. tropica* belonging to the zymodeme MON-265 and producing EF of the sub-serotype A\(_4\) and LPG like that produced by *L. major*, it did not survive and grow in female sand flies of the species *Ph. papatasi*, the proven sand fly vector of *L. major* (Svobodova, unpublished data mentioned in Svobodova [7]). It seems, therefore, that other factor(s), in addition to producing the ‘correct’ type of EF and LPG, is or are required for infection of female *Ph. papatasi* to succeed. Dobson [31] also came to this conclusion after completing their refined study on the effect of genetically manipulating the LPG structure of strains of *L. major* and checking their survival in their “selective” vector species, *Ph. papatasi*, and “permissive” vector species, *Ph. duboscqi*. In the study, they also engineered a strain of *L. donovani* to produce LPG of *L. major* that gave maximal survival of *L. major* in *Ph. papatasi*, but despite this manipulation the promastigotes failed to survive, indicating that another factor or factors besides the ‘correct’ LPG is or are required for their survival. Since the promastigotes of the 12 Palestinian strains of *L. tropica* were of two serological/antigenic types: EF sub-serotype A\(_2\), congruent with zymodeme MON-307 and the minicircle DNA sequence *Ltro*-kD1, and serotype A\(_9\)B\(_4\), congruent with zymodeme MON-137 and the minicircle sequence DNA *Ltro*-kD2, with each type, supposedly, also bearing a surface LPG with the same antigenic determinants that enables successful attachment in their specific natural vector, there is the possibility of two sand fly species or just sub-species transmitting the two Palestinian types of *L. tropica* in the foci of CL in the Jenin District.

Despite the introduction of many new methods for characterizing and identifying strains of *Leishmania*, MLEE is still the widely accepted standard means for differentiating leishmanial species and exposing biochemical micro-heterogeneity and interrelationships among strains, within the species [18,19,33,34]. The
report given here is the first on the enzyme analysis of Palestinian strains of L. tropica and the zymo-
demes to which they belong. Ten of the 12 Palestin-
ian strains were subjected to MLEE that also showed
that the strains were strains of L. tropica, according
to the electrophoretic mobilities of certain enzymes
in their enzyme profiles, notably NP1 and GOT1. The
electrophoretic mobilities of other enzymes in
their enzyme profiles separated the ten strains into
two sub-groups: seven belonged to the zymodeme
MON-137; three to the new zymodeme MON-307,
which are the reference strains of this zymodeme
(Table 2). These sub-groups were fully congruent
with those identified by kDNA analysis, EF serotyp-
ing and the MLMT done by Schwenkenbecher [3]
(Table 1). Interestingly, the strains of L. tropica
belonging to zymodemes MON-137 and MON-307
overlapped geographically within the limited region
from where the strains came (Figure 1a and b).

Of the 15 enzymes used in formulating the enzyme
profiles, one, fumarate hydratase had an electrophoretic mobility (FH105) different from the electrophoretic mobi-

lities of all the known variants of FH found so far among
strains of L. tropica examined by MLEE in the way
described above (Table 2). However, FH of this mobility
(FH105), while being unique to the strains of L. tropica
belonging to the zymodeme MON-307, has been found
in all the strains of L. aethiopica analyzed by this method
of MLEE ([18,19]; Table 2). The seven Palestinian strains
belonging to the zymodeme MON-137 were definitely L.
tro

ica and strains of L. tropica belonging to this
zymo
deme also occur in Israel [14]; the Sinai Peninsula, Egypt,
19] and Jordan [35,36]. The UPGMA dendrogram
(Figure 5) indicates that strains belonging to zymodeme
MON-137 have a more marginal enzyme profile than
most other strains of L. tropica and are closer to strains
of L. tropica (syn. L. killicki), from Tunisia in the zymo-
deme MON-8 and strains of L. tropica from Algeria,
Yemen and Kenya in the zymodemes MON-301, MON-
71 and MON-119, respectively, that are all found in clus-
ter (a) in Figure 5. Pratlong [19] showed that strains of L.
tropica belonging to the zymodemes MON-137 and
MON-265, also found in Israel and Jordan [35,36], clus-
tered more closely with one another than they did with
any of the other strains falling into their ‘L. tropica com-
p lex’, despite significant differences in geographical
distribution, sand fly vector specificity, EF serotypes,
enzyme profiles and some of their molecular biological
criteria [3,7,14,32]. Correspondingly, strains of the zymo-
demes MON-137 and MON-265 subjected to MLMT
clustered together in the sub-population I: Middle East,
but those of zymodeme MON-137 belonged to the
microsatellite sub-sub-groups A1a and A1b and those of
zymodeme MON-265 belonged to the sub-sub-group
A2b, a grouping into which no Palestinian strains fell.

Combining results from MLEE and MLMT showed that
the zymodeme MON-137 encompassed strains in the
microsatellite sub-sub-groups A1a and A1b and zymodeme
MON-307 encompassed strains in the sub-sub-groups
B1a and B1b, demonstrating the greater discrimination
of MLMT (Figure 2).

A similar degree of discrimination was previously
shown for strains of the zymodemes of L. infantum,
which were studied using MLMT [37] and L. donovani,
which were studied using multilocus sequence typing
(MLST) [38]. Strains with the same enzyme profile are
not necessarily a genetically monolithic group. In fact,
Mauricio [38] showed that enzymes of identical electro-
phoretic mobility can have different amino acid
sequences, and that the resulting changes in molecular
mass and charge can in some cases balance one another,
conferring the same electrophoretic mobility, and indistin-
guishable zymodema phenotypes can result from dis-
tinct genotypes.

Strains of zymodeme MON-137 seem to be restricted to
the western part of the Middle East and those of zymo-
deme MON-307 appear unique to the northern part of
the Palestinian West Bank but, on comparing enzyme
profiles and microsatellite profiles of strains of L. tropica,
those of the new zymodeme MON-307 are taxonomically
closer to strains of L. tropica that are geographically much
distant from the foci in the Jenin District (Table 2
and sub-group (b) in Figure 2 and Figure 5). Only three
enzymes had different mobilities (MDH112, GLUD95
and FH110) in the 15 enzyme profile of Syrian strains of zymo-
deme MON-307; and only three enzymes had differ-
ent mobilities (GLUD95, PGM100 and FH105) in the 15 en-
zyme profile of some Lebanese and Iraqi strains, compared
with these enzymes (GLUD80, PGM108 and FH105) in the 15
enzyme profile of some strains belonging to the zymodeme
MON-307. This might suggest importation of this type of
L. tropica but it is sufficiently different from its geographic-
ally distant relatives to think otherwise.

kDNA analysis, EF serotyping and MLEE, were able to
split the 12 Palestinian strains into the same two sub-
groups to the same degree as could MLMT (Tables 1
and 2; Figures 2 and 3b). The strains displaying the en-
zyme profile associated with the zymodeme MON-307
also possessed kDNA minicircles with the sequence
Ltro-kD1, a microsatellite profile that fell into the sub-
group B1 (type II: Asia) and produced EF of the sub-
serotype A2 while the strains displaying the enzyme profile
of zymodeme MON-137 also possessed kDNA minicir-
cles with the sequence Ltro-kD2, a microsatellite profile
that fell into the sub-group A1 (I: Middle East) and EF
of the sub-serotype A9 or A9B.
The enzyme and microsatellite profiles of the strains LRC-L888 and -L1324 in kDNA Clade B are not known. However, since the nine strains LRC-L725, -L758, -L882, -L883, -L885, -L886, -L887, -L889, -L893 belonging to the zymodeme MON-137 fell into the kDNA Clade B, had microsatellite profiles that fell into the sub-group A1, and either the EF serotype, A9 or A9B10, one can surmise that the strains LRC-L888 and -L1324 would belong to the zymodeme MON-137.

The presence of two separate sub-types of *L. tropica*, possibly, indicates two separate transmission cycles involving two separate types of phlebotomine sand fly vector. Further research should be directed at collecting potential vectors, checking for infections and isolating and characterizing leishmanial isolates from them to clarify this and to see if these leishmaniases are anthropomous or zoonomous by searching for infected domestic, e.g., dogs, and wild, e.g., hyraxes, animals.

**Conclusions**

A high degree of congruity was seen among the results of the three methods used here to characterize and differentiate Palestinian strains of *L. tropica*: EF serotyping; MLME; and analysis of kDNA mini-circle sequences, and also with those from MLMT that was applied in a previous study [3], even though the types of criteria examined were very different from one another, having been derived from different cellular components with different intra-cellular origins within the parasites. The results from the kDNA analysis corresponded well with those of MLMT and could, possibly, replace the more complicated and labour intensive application of MLMT to leishmanial parasites from this area. The Palestinian strains that were assigned to different genetic groups also differed in their MLEE profiles and their EF types.

A novel outcome of this study was the discovery of a new zymodeme, MON-307 that, at this time, seems to be unique to the northern part of the Palestinian West Bank. As two sub-types of *L. tropica* are circulating in the JD, future studies could investigate whether they are transmitted by the same sand fly species or indicate the presence of two different transmission cycles, involving separate sand fly vectors species; and whether the two different phenotypes reflect differences in the biological functions of the two parasitic sub-types, such as virulence, in different human hosts and attachment and survival in sand fly vectors.

**Competing interests**

The authors declare that they have no competing interests.

**Authors’ contributions**

KA: conception and design of the study, involved in the collection of samples, extraction of DNA, PCR-RFLP, drafting the paper, analysis and interpretation of data, revising. LS. carried out all of the serotyping and its analysis, wrote his part of the Methods and also helped with the overall analysis of the results, the presentation of the data, including the figures and tables, and the writing of the manuscript. GS: analysis and interpretation of data, drafting the paper and revising. AN: Providing the reference samples, revising and drafting the paper. JD and FP: carried out all of the MLEE work, wrote their part of the Methods and reviewed the manuscript, lending their expertise on the analysis of the results. CR and FB: analysed the zymodermal data of the Centre National de référence des Leishmanias, Montpellier, and constructed the dendrogram displaying the geographical distribution of the zymodemes of *L. tropica* and their interrelationship, and were responsible for writing their part of the Methods and reviewing the manuscript and its analyses of the various data presented. SE: Revising and drafting the paper. ZA: conception and design of the study, providing the laboratory facilities, supervision of the work, analysis and interpretation of data. All authors read and approved the final version of the manuscript.

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