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Resting-state neural firing rate is linked to cardiac cycle duration in the human cingulate and parahippocampal cortices

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Title: Resting-state neural firing rate is linked to cardiac cycle duration in the human cingulate and parahippocampal cortices

Abbreviated title: Neural firing and cardiac cycle duration

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Abstract. Stimulation and functional imaging studies have revealed the existence of a large network of cortical regions involved in the regulation of heart rate. However, very little is known on the link between cortical neural firing and cardiac cycle duration (CCD). Here, we analyze single- and multi-unit data obtained in humans at rest, and show that firing rate co-varies with CCD in 16.7% of the sample (25/150). The link between firing rate and CCD was most prevalent in the anterior medial temporal lobe (entorhinal and perirhinal cortices, anterior hippocampus and amygdala) where 36% (18/50) of the units show the effect, and to a lesser extent in the mid- to anterior cingulate cortex (11.1%, 5/45). The variance in firing rate explained by CCD ranged from 0.5 to up to 11%. Several lines of analysis indicate that neural firing influences CCD, rather than the other way round, and that neural firing affects CCD through vagally mediated mechanisms in most cases. These results show that part of the spontaneous fluctuations in firing rate can be attributed to the cortical control of the cardiac cycle. The fine tuning of the regulation of CCD represents a novel physiological factor accounting for spontaneous variance in firing rate. It remains to be determined whether the "noise" introduced in firing rate by the regulation of CCD is detrimental, or beneficial, to the cognitive information processing carried out in the parahippocampal and cingulate regions.

Significance statement. Fluctuations in heart rate are known to be under the control of cortical structures, but spontaneous fluctuations in cortical firing rate, or "noise", have seldom been related to heart rate. Here, we analyze unit activity in humans at rest and show that spontaneous fluctuations in neural firing in the medial temporal lobe, as well as in the mid to anterior cingulate cortex, influence heart rate. This phenomenon was
particularly pronounced in the entorhinal and perirhinal cortices, where it could be observed in one neuron out of three. Our results show that part of spontaneous firing rate variability in regions best known for their cognitive role in spatial navigation and memory corresponds to precise physiological regulations.
Introduction

What does the brain at rest do? While there is no simple answer to this question, it is usually agreed that the spatio-temporal structure of spontaneous brain activity is meaningful (Arieli et al., 1996; Berkes et al., 2011) and that it can influence responses to stimuli (Greicius et al., 2004). However, the brain at rest is not only preparing to respond to future stimuli, it is also constantly engaged in monitoring and regulating bodily organs such as the heart.

Central brain regions engaged into heart rate regulation include large portions of the cingulate cortex, the ventro-medial prefrontal cortex, the insula, as well as the amygdalo-hippocampal formation. The role of those regions in heart rate regulation was established by a combination of anatomical tracing, stimulation and lesion studies in rats (Cechetto et al., 1990; Benarroch, 1993; Verbeke and Owens, 1998; Westerhaus and Loewy, 2001), cats and monkeys (Anand and Dua, 1956). There is a good agreement with the areas involved in heart rate regulation as identified in humans by direct electrical stimulation in patients (Pool and Ransohoff, 1949; Stock et al., 1978; Oppenheimer et al., 1992; Parvizi et al., 2013) or with functional magnetic resonance imaging, during tasks (for reviews, see Cechetto and Shoemaker, 2009; Thayer et al., 2012; Beissner et al., 2013; Gianaros and Wager, 2015) or at rest (Chang et al., 2013; Rebollo et al., 2018). This suggests that neural firing might be directly related to heart rate regulation, at least in so-called "limbic" regions.

Surprisingly, the link between neural activity and heart rate is mostly absent from the literature on neural variability at the single neuron level, although the link between local field potentials and respiration has recently been underlined (Tort et al., 2018). Spontaneous variations in firing rate are most often attributed to cellular machinery noise (Faisal et al., 2008), or to brain state (Poulet and Petersen, 2008; Deco
and Hugues, 2012; McGinley et al., 2015) and fluctuations in excitability (Goris et al., 2014) related to top-down factors (Gilbert and Sigman, 2007). However, some of the ongoing fluctuations in cortical firing rate might be related to heart rate. Indeed, there are two mostly unnoticed reports of a relation between the duration of the cardiac cycle and firing rate in the cat somato-sensory thalamus (Massimini et al, 2000), as well as in the human amygdala and hippocampus (Frynsinger and Harper, 1989, 1990), i.e. in structures that interact massively with cortical regions.

Here, we directly investigate the link between cardiac cycle duration (CCD) and spontaneous neural firing rate in single- and multi-unit activity (SUA and MUA) recorded in humans at rest. Recording sites were determined for diagnostic and therapeutic purposes only but happened to be often located in regions known to be related to heart rate, i.e. in the medial temporal lobe (parahippocampal gyrus, hippocampus and amygdala) and in the cingulate cortex.

Before presenting the results, we remind here the reader of some basic facts about the heart rate and its central regulation. The electrocardiogram (ECG) allows to quantify CCD, which varies spontaneously during resting-state (Figure 1). The central nervous system can affect CCD (Berntson et al., 2007; Somsen et al 2004; Thayer et al., 2012, Gianaros and Wager, 2015) by modulating the vagal output to the cardiac pacemaker, resulting in fast, beat-to-beat, but also longer-lasting changes in heart rate. Sympathetic influences affect heart rate fluctuations with a delay of several seconds and induce long lasting changes, via the modulation of cardiac contractibility and vessel resistance.

Following the approach used in Frynsinger and Harper (1989) and Massimini et al (2000), we first examine the correlation between neuronal firing rate and CCD, and
further characterize the firing properties of the neurons that exhibit a strong correlation. We then investigate the temporal aspects of the link between firing rate and CCD, to provide indications on the underlying mechanisms. Last, we analyze the directionality of interactions, from neuron to heart and from heart to neuron.
Materials & Methods

Subjects

Twelve patients with pharmacoresistant focal epilepsy (5 male, 7 female, mean age 31.9 ± 8.83 years) were stereotactically implanted with depth electrodes to determine the seizure onset-zone for potential surgical resection. Implantation sites were selected for diagnostic and therapeutic purposes only. During the following seizure monitoring period, participants provided informed and written consent to participate to the present study. At the time of recordings, all patients received at least one medication that could interfere with cardiac activity (most often, carbamazepine, lamotrigine, or lacosamide, for details see Table 1). All procedures were approved by the local Ethics committee (CPP Paris VI, INSERM C11-16).

Recordings

The depth electrodes (AdTech®, Wisconsin, Behnke-Fried type) consisted of 8 macro contacts (platinum, diameter of 1.3mm) embedded on the surface of a polyurethane tube with a hollow lumen. Eight 40 μm diameter platinum microwires, including one used as a reference, protruded 3-6mm into the cerebral tissue beyond the tip of the deepest macrocontact. The electrocardiogram (ECG) was recorded from two cutaneous electrodes on the upper chest, but the exact positioning was not standardized. All signals were collected using a 160 channels recording system (ATLAS, Neuralynx®, Inc, Bozeman, MO) with a 32 kHz sampling rate and a 0.1-8000Hz bandpass filter for microwire data and a 4 kHz sampling rate and 0.1-1000Hz bandpass filter for the ECG.

Experimental design

During data collection subjects were seated in their bed and fixated a central point on a
gray background displayed on a laptop monitor. The extracellular neural signals analyzed in the current study come from either one continuous recording during resting-state fixation lasting 5 to 6 minutes, or from shorter blocks of 13 to 30 seconds of resting-state fixation embedded within a task (Babo-Rebelo et al., 2016).

**Electrode localization**

Anatomical localization was based on individual postoperative MRI warped to the MNI brain using the EpiLoc toolbox developed by STIM: Stereotaxy, Techniques, Images, Models (http://pf-stim.cricm.upmc.fr), as well as on the CT scan aligned with MRI data. Note that the microelectrode bundle opens at the tip of the macroelectrode, with a distance of a few mm between microwires. The localization described in Table 1 provides an anatomical description of all 8 microwires that in some cases could span different regions.

**Spike detection and spike time series**

An adaptive filtering procedure (Keshtkaran and Yang, 2014) was applied to the raw data to limit power line interference as well as harmonics. Spike detection and waveform sorting were performed using the semi-automatic procedure implemented in the software waveclus (Quiroga et al., 2004). Data were first band-pass filtered between 300 and 3000 Hz (elliptic band pass filter, 4th order) and spikes were detected using the automatic amplitude threshold algorithm. Thresholds for spike detection ranged between 3.5 and 5.5 standard deviations. Waveforms were then clustered according to the super-paramagnetic clustering algorithm. Typically one to three clusters were isolated from each microelectrode where spikes had been identified. A number of additional steps were applied for selecting valid units after cluster isolation.
First, we visually investigated waveforms for each cluster and discarded clusters displaying multiple peaks (n=5). Second, we discarded clusters without any spike for at least 45 consecutive cardiac cycles (n=7). We then removed the 5 clusters with the lowest firing rate (below 0.2 spikes/s). All remaining clusters (150) were considered as valid units. Units were further identified as single or multi-unit based on the percentage of very short inter-spike intervals. Units with less than 1% of inter-spike intervals shorter than 3ms were classified as single-units, else they were classified as multi-units.

After applying all the steps described above, 108 single- and 42 multi-units were selected for further analysis.

Spike times were downsampled from 32 kHz to 1 kHz. Spike density, a continuous estimate of instantaneous firing rate, was estimated by convoluting the spike time series with a Gaussian kernel (20 ms standard deviation). The spike density time-series were used for the spectral analysis and for the computation of coherence with heart rate variability, as well as to search for transient changes in firing rate in response to heartbeats.

Cardiac cycle duration (CCD) and CCD time series

We detected R peaks in the ECG using a semi-automatic procedure, that involved correlating the ECG with a template QRS complex defined on a subject-by-subject basis, and the manual verification of all R peaks separated from their neighbours by very short or very long intervals. R-peak timings were downsampled from 4 kHz to 1 kHz. Cardiac cycle duration (CCD), or inter-beat interval, was defined as the latency difference between two successive R peaks (Figure 1A,B). In the 4 patients with at least 5 minutes of continuous recordings, CCD time series were created by assigning each CCD to the central time point between the heartbeats corresponding to the cardiac cycle, and
interpolating with a cubic spline function (Figure 1C). These time-series were used for the spectral analysis of heart rate variability as well as for cross-correlation between inter-spike interval and CCD time series.

**Linear correlation analysis between firing rate and cardiac cycle duration.** To quantify the correlation between firing rate and CCD in each unit, we computed the mean firing rate at each cardiac cycle (number of spikes during a cardiac cycle divided by cycle duration, spike/s) and computed for each unit the Pearson correlation coefficient between the firing rate and CCD, across all cardiac cycles. Statistical significance was evaluated using a permutation-based procedure, where the original order of CCDs was shuffled 10,000 times. The resulting Monte-Carlo p-values were corrected for multiple comparisons across recording sites using the false discovery rate (FDR) procedure (Hochberg and Benjamini, 1990).

**Spectral analysis of heart rate variability and spike density**

We analyzed both heart rate variability (HRV) and spike density time series in the frequency domain in the 4 patients in whom we recorded 5 minutes of continuous data. Spectral analysis was performed using the FieldTrip toolbox for Matlab (Oostenveld et al., 2011) using a multitaper frequency transformation in sliding 60-s time windows with 6-s overlap. Coherence between the HRV and spike density spectral estimates was computed using the FieldTrip function ft_connectivityanalysis.m.

**Cross-correlation and coupling between inter-spike interval and CCD time series**

To assess whether the fluctuations of neuronal spiking and heartbeat activity are correlated, we computed cross-correlograms between smoothed neuronal interspike
interval (ISI) and CCD time series. Similarly to CCD time series, a neuronal ISI time series was generated by assigning to each ISI center time point the ISI duration, and interpolation using cubic splines. We estimated the coupling kernel, $\alpha(t)$, which expresses the systematic interaction between the two smoothed time series over the whole recording duration: $\text{CCD}(t) = \sum_l \alpha(l) \cdot \text{ISI}(t-l)$. Specifically, we calculated this kernel using (inverse) Fourier transforms, $\alpha(t) = \text{ifft} \{ \text{fft( CCD(t) )} / \text{fft( CCD(t) )} \}$.

Computations were performed for each continuous recording block. Results were averaged across the available blocks, the contribution of each block being weighted by its duration.

Detection of transient changes in instantaneous firing rate in response to heartbeats

Data were epoched from one R peak of the electrocardiogram to the next. Data analysis was performed either by analyzing the data locked to the onset of the epoch, end of the epoch, or by normalizing epoch duration from the current heartbeat to the next, latencies being then expressed in percentage of CCD. Results were similar in all three cases and the results reported here correspond to the normalized epoch duration. The existence of transient increases or decreases in firing rate in the spike density time series was then assessed by a cluster-based permutation procedure (Maris and Oostenveld, 2007) that intrinsically corrects for multiple comparisons. For each unit, we created 10,000 surrogate datasets where R peak latencies were shuffled but the CCD distribution was preserved. We compared at each time point the observed spike density value with the distribution of the 10,000 surrogate spike density value to derive a Monte Carlo p value. Adjacent time points with a Monte-Carlo p-value < 0.01 defined candidate clusters. Monte-Carlo p values were then converted into z-scores. In a second step, we summed over time the z-scores of each candidate cluster. We repeated this procedure on the
10,000 surrogate datasets, retaining for each surrogate dataset the largest z-sum, to obtain the max (z-sum) distribution that could be obtained under the null hypothesis. The comparison between the observed z-sum and distribution of surrogate z-sum identifies clusters of significantly increased or decreased firing in the original data, with a Monte Carlo p <0.05, two-tailed. The resulting Monte-Carlo p-values were corrected for multiple comparisons across recording sites using the false discovery rate (FDR) procedure (Hochberg and Benjamini, 1990).

**Phase response analysis**

To quantify the effect of occurrence and timing of spikes on CCD we calculated phase response curves using a method adopted from Blot et al. (2016) (where these curves are referred to as delayed spike curves due to the specific context). The phase response curve (PRC) measures how much a spike within a cardiac cycle shortens or lengthens the current CCD on average. In particular, defining $T_-$ as the time duration since the previous heartbeat and $T_+(T_-)$ the time duration to the next heartbeat (which naturally depends on $T_-$) the PRC is expressed as the averaged difference of the duration $T_+(T_-)$ observed (using heartbeat and spike times) and the duration $\langle T_+(T_-) \rangle_{\text{exp}}$ that is expected based on knowledge of the CCD distribution only:

$$\text{PRC}(T_-) = \langle T_+(T_-) - \langle T_+(T_-) \rangle_{\text{exp}} \rangle$$

where the latter term is calculated by

$$\langle T_+(T_-) \rangle_{\text{exp}} = \frac{\int_{t>T_-} t \cdot p_{\text{CCD}}(t) \, dt}{\int_{t>T_-} p_{\text{CCD}}(t) \, dt} - T_-$$

and the (outer) average $\langle \rangle$ is calculated using a suitable binning of CCDs. We considered two types of binning: only one bin corresponding to the whole cardiac cycle, and two bins of different lengths corresponding to systole and diastole. Systole duration
was determined in each patient according to the formula proposed by Fridericia (2003):

\[ systole = 8.22 \sqrt[3]{RR} \]

were RR is the mean CCD of the patient. Diastole was defined, at each cardiac cycle, as the remaining part of the cardiac cycle. Note that this method cannot easily be extended to the analysis of adjacent cycles in a meaningful way.

To test for significance we applied a permutation-based procedure using a large number (10,000) of surrogate spike trains generated by shuffling the interspike intervals of the original data. We used the Hochberg and Benjamini procedure (Hochberg and Benjamini, 1990) to control for multiple comparisons across recording sites, by maintaining the false discovery rate (FDR) at \( p < 0.05 \).

PRC values computed over one cardiac cycle are positive in cells showing a positive correlation between firing rate and CCD (R+ cells) and negative in R- cells. Considering the two-bin (systole, diastole) PRC, we tested for the existence of a significant reduction in PRC from systole to diastole (i.e., positive or negative values getting closer to zero) by multiplying systolic and diastolic PRC values in R- cells by -1, and comparing the resulting PRC values in systole and diastole using a two-tailed paired t-test.

**Statistical analysis.**

As described above, all statistical analysis relied first on the estimation of a Monte-Carlo p-value at each recording site, obtained by comparing the empirical result with the distribution under the null hypothesis computed on data in which CCDs have been shuffled, and then on a control for multiple comparisons across recordings sites using the Hochberg and Benjamini procedure (Hochberg and Benjamini, 1990) to maintain the false discovery rate at \( p<0.05 \).
Bayes factors were computed using the online tool (http://pcl.missouri.edu/bayesfactor), which is based on Liang et al. (2008). Here, a Bayes factor larger than 10 indicates strong evidence for the null hypothesis, between 3.2 and 10 indicates substantial evidence for the null, and smaller than 3.2 indicates inconclusive evidence for or against the null hypothesis (Kass and Raftery, 1995).

Code accessibility. Matlab/Python code is freely available at https://github.com/neuromethods/neural-firing-and-cardiac-cycle-duration.
Results

Data summary

We recorded unit activity together with the electrocardiogram in 12 epileptic patients (Table 1) with normal heart rate (mean 72.5 ± sem 2.8 beats per minute; range 61.4 to 91.8) during passive fixation. We isolated a total of 150 units (108 SUA, 42 MUA), recorded from 112 microelectrodes at 22 recording sites. The average firing rate across all units was 2.54 ± sem 0.10 spikes/s (SUA, 2.28 ±0.09; MUA, 3.19 ±0.12 spikes/s). 14 recording sites (Figure 2A), corresponding to 88 units, were located in the medial temporal lobe (parahippocampal gyrus, hippocampus and amygdala). 5 recording sites (45 units) were located in the mid- and anterior cingulate regions while the ventral visual pathway (2 recording sites in the fusiform gyrus, 15 units) or the subgenual anterior cingulate cortex (1 recording site, 2 units) were only occasionally sampled.

Correlation between spontaneous firing rate and cardiac cycle duration

As in Frysinger and Harper (1989) and Massimini et al (2000), we first computed the mean firing rate during each cardiac cycle, and observed a correlation between spontaneous firing rate and CCD. Figure 3A, B shows an example of a cell in the parahippocampal gyrus whose spontaneous firing rate is negatively correlated with CCD, and Figure 3C, D an example of a cell in the anterior cingulate gyrus whose firing rate is positively correlated with CCD.

Across all units, the correlation coefficient shows a bias toward negative values (Figure 3E, mean Pearson r=−0.027 ±0.008, t-test against 0 on Fischer-Z transformed correlation coefficients, t(149)= -3.51, p<10⁻³). This result indicates that at the population level, the firing rate is higher when the heart is beating faster. However, this effect was not evenly distributed amongst units.
We therefore tested the significance of the correlation between firing rate and CCD at the level of each individual unit. We found that 25 out of 150 units (16.7%, from 8 different patients and 9 recording sites) showed a significant correlation between firing rate and CCD (Figure 3E, Pearson correlation, all FDR corrected p<0.05). In those 25 neurons, CCD explained on average 4.25% ±0.53 of spontaneous firing rate variance, ranging from 0.5% up to 11%.

In 17 of these neurons (7 SUA, 10 MUA), the correlation was negative, i.e. elevated firing rates corresponded to shorter cardiac cycles (mean r=-0.22 ±0.014) and in 8 neurons (7 SUA, 1 MUA), the correlation was positive, i.e. elevated firing rates corresponded to longer cardiac cycles (mean r=0.14 ±0.014). Those neurons will be hereafter referred to as R- and R+ neurons respectively. The firing rate variance explained by CCD was significantly larger for R- units (5.26 % ±0.62) than for R+ units (2.1 % ±0.4; Mann-Whitney U test p=0.002).

The correlation between spontaneous firing rate and cardiac cycle duration is most prominent in the anterior part of the medial temporal lobe and cingulate cortex

The medial temporal lobe (MTL) was well represented in our dataset. In the MTL, 22.7% of the units (20 out of 88) showed a significant correlation with CCD. The most anterior MTL regions, corresponding to the anterior parahippocampal cortex, a region that encompasses the entorhinal and perirhinal cortices, anterior hippocampus and amygdala, seemed to be particularly involved (Figure 2). To quantify the difference between posterior and anterior MTL regions, we labelled MTL regions anterior if y≥-7 and posterior if y≤-10. The anterior MTL regions contained the highest proportion of units (36%, 18 out of 50 units) with a significant CCD effect (Figure 2B and Table 1). This percentage fell to 5.3% (2 units out of 38) for posterior (y≤-10) MTL recording.
sites. In the mid and anterior cingulate sites, 11.1% of the units showed a significant CCD effect (5 out of 45 units; proportion lower but not significantly different from that found in MTL, Chi-square test, $\chi^2 = 2.63$, df =1, p=0.10). Last, no CCD effect was observed in less well sampled regions (fusiform gyrus, 0/15 units, 2 patients; subgenual anterior cingulate gyrus, 0/2 units, one patient). All the contacts showing a significant correlation between firing rate and CCD broadly belong to the mid to anterior cingulate cortex, and to the amygdalo-hippocampal formation and adjacent cortices.

Positive and negative correlations between firing rate and CCD could be found in the same region (Figure 2A) in different patients. In two instances we even found positive and negative correlations at the same location in the same patient, in neurons recorded by different microwires, i.e, separated by a few millimeters only (patient #4, amygdala / parahippocampal gyrus, 1 R+, 2 R-; patient #12, mid-cingulate cortex, 1 R+, 2 R-).

Neurons showing a significant CCD effect are more variable

Does firing rate in neurons with a significant CCD effect differ from firing rate in other neurons? We first compared the firing rate in neurons with a significant CCD effect (n=25, mean firing rate=2.14 ± 1.28 spike/s) to the firing rate of all other neurons (n=125, mean firing rate=2.62 ±2.21 spike/s) but found no significant difference (unpaired t-test, $t_{(148)}$=-1.03, p=0.30, BF=2.79, inconclusive). The firing of R- neurons (2.13 ±1.07 spike/s) was similar to that of R+ neurons (2.17 ±1.74 spikes/s; unpaired t-test, $t_{(23)}$=0.08, p=0.94, BF = 2.58, inconclusive).

We then computed a compact measure of firing variability, the Fano factor (Figure 4A), corresponding to the mean number of spikes divided by the variance in number of spikes, computed over 1s time windows. The Fano factor of neurons showing a significant CCD effect (n=25, FF= 3.78 ±3.90) was significantly larger than the Fano...
factor of non-significant neurons (n=125, FF= 1.62±1.18; unpaired t-test, t (148)=5.18, p<10^{-6}). The Fano factor was increasing with cardiac-related variance (Figure 4B), with a significant correlation between the two (n=150, Pearson correlation, r=0.51, p<10^{-9}).

Controls on recording stability and epileptic activity

We examined if the link between spontaneous spiking activity and CCD could be due to some cardiac-related recording instability. We tested whether spike waveform differed depending on CCD, as proposed by Frysinger & Harper (1989). We measured waveform peak amplitude in spikes occurring in the 20% shortest versus 20% longest cardiac cycles and tested for possible differences. Spike waveform was stable and did not depend on CCD, neither in the 25 neurons showing a significant cardiac effect (Figure 3F; waveform peak amplitude, paired t-test between spikes occurring in short and long cardiac cycles, t (24) = 0.043, p=0.97, Bayes Factor BF =4.65 indicating substantial evidence for the null hypothesis), nor across the 150 neurons tested (paired t-test, t (149) =0.099, p=0.92, BF=10.9 indicating strong evidence for the null hypothesis).

Thus, the relationship between spontaneous firing rate and CCD does not arise from cardiac-related recording instability.

We also verified that spike waveform peak amplitude was stable across recording time. The waveform peak amplitude in the first 20% of the recordings was not different from the last 20%, neither in the 25 neurons showing significant CCD effect (paired t-test, t (24)= 0.13, p=0.90, BF = 4.62 indicating substantial evidence for the null hypothesis), nor in the 150 neurons analyzed (paired t-test, t (149) = 0.098, p=0.92, BF = 10.91 indicating strong evidence for the null hypothesis).

Last, we verified that the correlation between firing rate and CCD was not confined to the seizure onset zone, which could be determined for all patients but one.
(Table 1). Four out of the 9 recording sites showing the correlation were located outside the seizure onset zone (2 in the anterior parahippocampal cortex, 2 in the cingulate cortex).

**Temporal extension of the link between firing rate and cardiac cycle duration**

So far, we have analyzed neural firing within the current cardiac cycle. We further investigated whether the link between spontaneous firing rate and CCD extends over a few cycles only, indicating a vagally mediated process, or over longer time periods, compatible with sympathetic influences. This analysis was performed in the subset of 4 patients where we recorded continuous segments of data of at least 5 minutes, i.e. the data length recommended to properly evaluate fluctuations in heart rate (Task Force, 1996). In those 4 patients, 10 cells showed a significant CCD effect (6 R+, 4 R-).

We first computed the correlation between firing rate in a given cardiac cycle and the duration of subsequent cardiac cycles. As can be seen in Figure 5 in 6 cells, located either in the medial temporal lobe or the cingulate cortex, the correlation was maximal for the current cardiac cycle duration, and decreased over time, a pattern suggesting vagal influences. In the remaining 4 cells, all located in the medial temporal lobe, the correlation increased after a few cycles and was maintained for much longer durations, up to 12 cardiac cycles later, compatible with either sympathetic or vagal influences.

In the same 10 cells, we then investigated whether the correlation between firing rate and CCD was modulated by respiration, as observed in the cat, in somatosensory thalamic neurons (Massimini et al, 2000). Indeed, CCD typically decreases during inspiration and increases during expiration. This modulation can be quantified by the spectral analysis of fluctuations in CCD over time, also known as heart-rate variability (HRV), where a peak between 0.15-0.4 Hz captures cardiac rate changes locked to
respiration. All patients showed normal HRV spectra (see examples in Figure 6 A,D), but the coherence between firing rate and HRV spectra did not reveal any distinctive peak, neither in the low nor high-frequency range, in any of the 10 cells analyzed (see examples in Figure 6 B,E). The coupling between firing rate and CCD that extends over several cycles is thus not related to respiration.

Fast, beat-to-beat fluctuations

The analysis across several cycles shows that the coupling between firing rate and CCD can persist over several seconds, but that it is not linked to cardio-respiratory modulation. To test whether the coupling also has a fast component related to beat-to-beat changes that would be a hallmark of vagal influences, we used a different method to assess short-range temporal correlations between variations in firing rate and CCD. In the 25 neurons showing a significant correlation, we extracted the effective coupling kernel between smoothed interspike interval and CCD time series, which is independent from the auto-correlation structure of each time series. All coupling kernels showed a marked narrow peak at 0 time-lag and very limited slower modulations (see examples in Figure 6 C,F). The interaction between the two time series occurred at a fast time-scale, suggesting that the link between firing rate and CCD is partly explained, at least in part, by short-lived, beat-to-beat fluctuations.

Directionality of the interaction between firing rate and cardiac cycle duration

The correlation between firing rate and CCD does not inform us on the directionality of the effect. We first searched for evidence of directed interactions going from the heart to spiking activity, by testing whether heartbeats trigger a transient change in firing rate. We analyzed the spike density function locked to heartbeats and
tested whether a heartbeat would trigger a transient increase or decrease in firing rate in any of the 25 neurons with a significant CCD effect. Out of the 25 neurons, only two neurons showed a transient increase in firing rate and one neuron a transient decrease, and none of those observations survived correction for multiple comparisons. We then used a model-based approach to test whether heartbeats elicit transient changes of neuronal firing activity. Each cell was described by a simple spiking neuron model with noisy background inputs and an additional input current triggered by heartbeats. Parameters were fitted using the observed spike trains and heartbeat times (Ladenbauer et al., 2018), and significance was assessed using a permutation-based procedure based on shuffled data. None of the 25 neurons showed a significant modulation of firing rate caused by heartbeats according to this analysis. Thus, there is little evidence that heartbeats trigger a transient neural response in the 25 neurons showing a CCD effect.

We then searched for evidence of directed interactions in the other direction, i.e. an influence of neural firing on CCD. To test whether the spiking activity of any of the 25 significant neurons exerts an effect on CCD, we used a phase-response analysis, that quantifies how much cardiac cycles are shortened or lengthened depending on the occurrence and timing of spikes within the cardiac cycle (Figure 7A; for details see Methods). In all 25 neurons we found that neuronal spikes were significantly associated with a lengthening or shortening of the cardiac cycle (all FDR-corrected Monte-Carlo p<0.05). Depending on neurons, the mean cardiac cycle shortening or lengthening induced by neural firing varied from -87.2 to +25.3 ms (Figure 7B). Cardiac cycle lengthening or shortening revealed by the phase-response analysis was directly related to the strength and sign of the correlation between firing rate and CCD (Figure 7B; correlation between cardiac cycle lengthening and Pearson correlation coefficient...
between firing rate and CCD, Pearson $r = 0.75$, $p<10^{-4}$). The correlation between firing rate and CCD can thus mostly be accounted for by the influence of individual spikes on CCD.

Because of transmission delays between the recorded structures and the heart, spikes occurring early in the cardiac cycle (systole) should have more influence on the timing of the next heartbeat than spikes occurring late in the cardiac cycle (diastole). We found that it was indeed the case (Figure 7C) in 20 neurons out of 25, with a significantly larger influence of spikes in systole than in diastole (paired t-test, $t(24)=3.13$, $p=0.0045$). Altogether, these results speak in favor of a directionality of interaction flowing from spiking activity to the heart.

Last, we found that R+ and R- neurons could be observed in the same region, from microwires only a few mm apart. We tested whether co-localized R+ and R- neurons systematically display a negative correlation between their firing rate, i.e. whether when a R- neuron fires and shortens the cardiac cycle, the R+ neuron remains relatively silent. The data do not speak in favor of such a simple mechanism. We correlated the firing rate per cardiac cycle in the 4 pairs of (R+, R-) neurons available in the recordings (Table 1) and observed all possible patterns: two negative correlations (one non significant; one significant $p=0.0026$ uncorrected), two positive correlations (one non significant; one significant $p=0.0007$ uncorrected).
Discussion

We recorded single and multi-unit activity in humans and found that spontaneous firing rate is directly related to cardiac cycle duration in more than a third of the neurons in the anterior parahippocampal gyrus (36%, 18/50), a region that encompasses both entorhinal and perirhinal cortices. We observed the same phenomenon, but in smaller proportion, in the mid to anterior cingulate cortex (11%, 5/45), and verified that this effect was not due to cardiac-related recording instability. Up to 11% of the variance of spontaneous firing rate was linked to fast, beat-to-beat changes in CCD, and neurons with a significant correlation between firing rate and CCD displayed an increased temporal variability in mean firing rate. The analysis of directed interactions pointed toward a direction of information flow from neurons to heart. The detailed analysis of the temporal delays in neuron-heart coupling further reveals the involvement of a vagally-mediated influence.

Cardiac cycle duration and firing rate in anterior parahippocampal regions and cingulate cortex

The regions where we find the correlation between firing rate and CCD are known to be related to heart-rate variability. Both cingulate regions (Ter Horst et al., 1996) and the amygdala and surrounding parahippocampal structures (Price and Amaral, 1981; Schwaber et al., 1982) project to autonomic nuclei, and the electrical stimulation of those regions alters CCD both in monkeys (Smith, 1945) and humans (Selimbeyoglu, 2010). Moreover, the BOLD signal in both cingulate and medial temporal lobe regions fluctuate with heart-rate variability during tasks (Thayer et al., 2012; Beissner et al., 2013) as well as during resting state (Chang et al 2013; Rebollo et al., 2018). Our results further show that the link between neural activity and CCD in both cingulate and medial temporal regions is directly reflected in fluctuations of spontaneous firing rate.
Perhaps the most striking feature of our results is the very large prevalence (36%, 18/50) of cardiac-related units in the anterior part of the parahippocampal gyrus, confirming earlier findings (Frysinger and Harper, 1989). The anterior parahippocampal gyrus contains the entorhinal and perirhinal cortices, located at the interface between the neocortex and the hippocampus, that play an important role in spatial navigation and memory (Fyhn et al., 2004). Neuronal loss in this region is associated with cognitive impairments (Braak and Braak, 1991; Gómez-Isla et al., 1996). Spatial navigation, a behavior that strongly engages the entorhinal cortex (Fyhn et al., 2004), could benefit from a fast and precise regulation of heart rate to anticipate, and react to, the metabolic demands of walking and running.

The data presented here come from epileptic patients, which raises the question of whether the present results extend to healthy organisms. Epileptic patients tend to show a moderate reduction in heart rate variability (Sevcencu and Struijk, 2010) that may be related to medication or epilepsy (Tomson et al., 1998). The data analyzed here corresponded to a quiet resting state, devoid of any seizure. Several arguments suggest that the results presented here are not specific to epileptic patients. First, the regions where we find a link between firing rate and CCD show no systematic relationship with the seizure onset zone. Second, as detailed above, the regions where we find the CCD effect are known to be involved in cardiovascular regulation, either in animals or healthy human participants. Last, while all patients were under medication at the time of recordings, there was no consistent pattern between the presence of any given drug and the presence of the CCD effect.

**Cortical influence on heart rate**

We found that changes in firing rate influenced heart rate, with several
characteristics suggesting a vagal pathway: 1) the correlation between firing rate at time $t$ and subsequent CCDs broke down after a few cycles in a number of cells, 2) the coupling between firing and CCD included a fast component, acting within a single cardiac cycle, and 3) the directed interaction from neuron to heart was most pronounced at systole. Note that slower modulations could also be observed, with a link between firing rate and heart rate spanning several cycles. The slower mechanism could co-exist with the fast, vagal influence in the same neuron. A putative anatomical pathway for such modulation relies on the projections of the parahippocampal areas (Pitkänen et al., 2006) and cingulate regions (Pandya et al., 1981) to the amygdala, that in turn projects directly to the vagal nuclei in the medulla, the nucleus ambiguus and dorsal motor nucleus of the vagus nerve (Hopkins and Holstege, 1978; Schwaber et al., 1982; Spyer, 1994). Still, the analysis we performed cannot rule out that the link between firing rate and CCD is mediated by another structure, not investigated in the present dataset, that would influence both firing in entorhinal and cingulate cortices on the one hand, and CCD on the other hand. It is also worth underlying the fact that in line with previous results from Frysinger and Harper (1989), the correlation between firing rate and CCD did not appear to be linked to the respiratory-related changes in heart rate (Eckberg, 2009).

Our results further emphasizes the co-localization of neurons that increases or decreases heart rate within the same region. This finding is in line with the observation that the stimulation of the central nucleus of the rat amygdala (Iwata et al., 1987) or of the monkey entorhinal cortex (Reis and Oliphant, 1964) can trigger either increases or decreases in heart rate. Similarly in humans, both heart rate increases and decreases are related to activity in the cingulate cortex, as measured with fMRI (Gianaros and Wager, 2015).
A novel factor accounting for the spontaneous variations in firing rate

So far, spontaneous variations in firing rate have been attributed to cellular machinery noise (Faisal et al., 2008), to brain state (Poulet and Petersen, 2008; Deco and Hugues, 2012; McGinley et al., 2015), and to fluctuations in excitability (Gilbert and Sigman, 2007; Goris et al., 2014). Our results suggest that in the parahippocampal region, and to a lesser extent in the cingulate cortex, the fine tuning of the CCD represents a novel, physiological factor accounting for a non negligible part of the spontaneous variance in firing rate.

Changes in heart rate are often thought to be confined to emotions, pain, stress and physical effort. It is worth underlining that non-emotional standard tasks, such as visual or auditory detection tasks (Lacey and Lacey, 1978; Bradley, 2009; Park et al., 2014; Raimondo et al., 2017) are associated with a precise and reproducible change in CCD. The functional imaging literature in humans has underlined the convergence of cognitive function and cardiac regulation in both the cingulate cortex (Critchley et al., 2003) and in the amygdalo-hippocampal region (Gianaros et al., 2004). It remains to be determined whether the variance in firing rate related to cardiac cycle regulation is detrimental, or beneficial (Harris and Wolpert, 1998; McDonnell and Ward, 2011), to the cognitive information processing carried out in the parahippocampal and cingulate regions.
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Table 1. Patients and recording sites

| Patient# | Age | Gender | Medication          | Heart rate bpm | MNI mm xyz | Anatomical description        | #units | Cardio cyclic effect | Contact in SoZ |
|----------|-----|--------|---------------------|----------------|------------|--------------------------------|--------|---------------------|----------------|
| 1        | 30  | M      | OXC, LCM, CLBZ      | 61             | -24 -3 -30 | Parahip                        | 8      | 1R-                 | no             |
|          |     |        |                     |                | -38 -29 -23| Parahip                        |        | 2                   | no             |
| 2        | 26  | M      | CBZ, VPA            | 68             | 30 -5 -27 | Parahip                        | 1      | 0                   | no             |
|          |     |        |                     |                | 41 -54 -15| Fusiform gyrus                 |        | 6                   | no             |
| 3        | 23  | F      | CBZ, TPM, LCM       | 80             | -25 -14 -16| Parahip                        | 5      | 2R+                 | no             |
|          |     |        |                     |                | -20 -20 -12| Parahip                        |        | 2                   | no             |
| 4        | 32  | M      | LEV, LMT, CBZ       | 76             | -17 -5 -21| Amygdala/ Parahip              | 7      | 1R+, 2R-            | yes            |
|          |     |        |                     |                | -18 -14 -21| Hippocampus                    |        | 9                   | yes            |
| 5        | 22  | F      | VPA, LMT            | 88             | 5 -4 49   | Mid Cingulate sulcus           | 8      | 0                   | no             |
|          |     |        |                     |                | 10 20 27  | Anterior Cingulate sulcus      |        | 13                  | no             |
| 6        | 40  | F      | LCM, PHT, Venti ne, | 71             | -16 -10 -26| Hippocampus / Parahip          | 6      | 0                   | no             |
|          |     |        | Piascledine         |                | -9 38 -20 | Subgenual anterior cingulate   |        | 2                   | no             |
|          |     |        |                     |                | -27 -7 -46| Parahip                        |        | 7                   | yes            |
| 7        | 48  | F      | LMT, ZNS, PER       | 92             | -20 -2 -31| Parahip                        | 7      | 2R+                 | yes            |
|          |     |        |                     |                | -24 -20 -16| Hippocampus / Parahip          |        | 3                   | yes            |
| 8        | 34  | M      | CBZ, LMT            | 65             | 24 -14 -26| Hippocampus / Parahip          | 11     | 0                   | no             |
|          |     |        |                     |                | 21 -5 -39 | Parahip                        |        | 7                   | no             |
| 9        | 26  | F      | LEV, LCM, VPA, Fluoxe tine, Zolpid em, DZP, Propan olol | 63 | 22 -3 -34 | Parahip | 13 | 11 R- | unknown* |
| No. | Age | Gender | Medications | X,Y,Z Coords | Location | No. of Units | Side | Status |
|-----|-----|--------|-------------|--------------|----------|--------------|------|--------|
| 10  | 25  | F      | LMT, LCM, CLBZ | 68 | 10 18 30 | Anterior Cingulate gyrus | 7 | 0 | no |
| 11  | 47  | M      | ESL, ZNS     | 69 | -18 -65 -10 | Fusiform gyrus | 9 | 0 | no |
| 12  | 30  | F      | LMT, LEV, CBZ, PER | 69 | -2 30 21 -2 5 41 | Anterior Cingulate gyrus, Mid Cingulate gyrus | 7 | 10 | 1R+, 2R- | no |

Abbreviations: Parahip: parahippocampal gyrus. R+: unit showing a positive correlation between firing rate and cardiac cycle duration (p<0.05, FDR corrected); R-: unit showing a negative correlation. SoZ: seizure onset zone. Medication, CBZ: carbamazepine; CLBZ: clobazam; DZP: diazepam; ESL: eslicarbazepine; LCM: lacosamide; LEV: levetiracetam; LMT: lamotrigine; OXC: oxcarbazepine; PER: perampanel; PHT: phenytoine; VPA: valproate; ZNS: zonisamide. *: premature interruption of the clinical procedure, no seizure recorded.
Figure legends

Figure 1. Measuring the cardiac cycle. A. An example ECG trace showing the QRS complex. Cardiac cycle duration (CCD) is defined as the time between two successive R peaks. B. Histogram of CCD distribution in one patient. C. The CCD time series (bottom) is constructed by assigning each CCD (top) to the central time point (black filled circles) between two heartbeats, then interpolating the points with a cubic spline function.

Figure 2. Anatomical description of recording sites. A. All 22 recording sites from 12 patients projected on the medial view of an inflated hemisphere, with left and right hemispheres collapsed for visualization purposes. Each recording site is represented by a dot, with size indicating the number of cells isolated. Dot color indicates sites with at least one unit with a significant increase in firing rate for slower heart rate (positive correlation between firing rate and CCD, pink), with at least one unit with a significant decrease in firing rate for slower heart rate (negative correlation, green), or both (red). Sites where no unit showed a significant link between firing rate and heart rate are presented in gray. B. Coronal brain views with a projection of medial temporal recording sites with y comprised between -10 and -30 (black contour, left) and sites with y comprised between -2 and -7 (yellow contour, right), corresponding to the black and yellow brackets in A.

Figure 3. Correlation between firing rate and cardiac cycle duration. A. Example cell (#128 patient #9, parahippocampal gyrus) with a negative correlation between firing rate (spikes/s) and CCD. The left panel is the spike raster plot where the y-axis
corresponds to 30 consecutive cardiac cycles, organized by order of occurrence from first (bottom) to last (top), and the x-axis corresponds to the time from current R-peak to the next R-peak marked by the orange triangle. The middle panel corresponds to the same raster as the left one, but the data are sorted according to increasing CCDs, from short (bottom) to long (top) cardiac cycles. B. Mean firing rate (±SEM) as a function of CCD duration (quintiles, from short to long CCD), in the same cell as in A, for the full dataset (365 cardiac cycles). The inset illustrates the distribution of correlation coefficients between firing rate and CCD derived from surrogate data. The green arrow indicates the correlation coefficient of the empirical data. C, D. Example cell (#162, patient #12, anterior cingulate), showing a positive correlation between firing rate and CCD, same conventions as in A and B. E. Population histogram (n=150 units) of the correlation coefficient between firing rate and CCD. The colored bars indicate cells with significant positive (pink) and negative (green) correlation between firing rate and CCD (FDR corrected, p<0.05). F. Comparison of spike waveform amplitude peak between first and last CCD in quintile (n=25). Horizontal black bars indicate mean of waveform peak amplitude within first and last CCD in quintile over 25 significant units. The above two insets show example waveforms averaged over first and last CCD in quintile and the peak amplitude is indicated by the arrow.

Figure 4. Fano factor and cardiac related variance. A. Violin plot indicating mean (red) and median (black) of Fano factor of units showing no significant correlation (NS) and units showing significant correlation with CCD (R+, R-). B. The Fano factor (on a log₁₀ scale for graphical purpose only) is plotted as a function of variance in spontaneous firing rate explained by CCD, for units showing no significant correlation (gray), a
negative correlation (green), and a positive correlation (pink) between firing rate and CCD.

**Figure 5. Correlation between firing rate and CCD over time.** Firing rate at a given cardiac cycle is correlated with duration of this cardiac cycle (CCD₀), or with CCD occurring 4, 8 or 12 cardiac cycles later (resp. CCD₄, CCD₈ and CCD₁₂). All correlation coefficients are normalized to 1 at CCD₀. Colors indicate units showing significant positive (pink) and negative (green) correlation between firing rate and CCD₀.

**Figure 6. A, D.** Power spectrum of heart-variability (HRV) in patient #6 (A) and patient #12 (D). B, E. Coherence between HRV and firing rate in two units. No distinctive peak corresponding to high or low frequency HRV could be observed. C, F. Coupling kernel (α) between the smoothed interspike interval and CCD time series for two units.

**Figure 7. Effects of spikes and their timing on cardiac cycle duration.** A. Change of current CCD caused by spikes depending on their timing since the last heartbeat in comparison to the expected CCD (gray dots) and mean impact on CCD averaged across all spikes irrespective of their timings (green line for shortening, pink line for lengthening) for 3 example units. B. Mean impact on CCD vs. correlation coefficient between firing rate and CCD for the 25 units with significant correlation. C. Average change of CCD caused by spikes during systole and diastole separately for the 25 units from B (green or pink dots indicate a significant difference from 0). Lines connect average impact on CCD unit-wise. The influence of spikes on CCD was larger during systole in 20 out of 25 units.
Figure 1

A

B

C

CCD | 946 | 912 | 800 | 735 | 732 | 996 | 1149 | 1054
---|---|---|---|---|---|---|---|---
CCD (ms) | 1200 | 1000 | 800 | 600 | 400 | 200 | 0 | 200 | 400 | 600 | 800 | 1000 | 1200

time (ms) | 2500 | 4500 | 6500 | 8500

# of R-peaks

0.5 | 1 | 1.5

500 ms
Figure 2

A

Firing rate and cardiac cycle duration
- R+ & R-
- R+
- R-
- n.s

Number of cells isolated
- 1-5
- 6-10
- 11-15

B

y = 17

y = 4
Figure 4

(A) Fano (variance/mean) vs Correlation for NS, R+ & R-. The red line represents the mean, and the black line represents the median.

(B) Scatter plot showing Fano against Cardiac related variance (%). The data points are color-coded for R+ and R-.
Figure 5
Figure 6
Figure 7

A

B

C

Time from previous heartbeat (ms)

Correlation coefficient

CCD change (ms)

-300 -200 -100 0 100 200 300

-300 -200 -100 0 100 200 300

Systole Diastole

Shorter Longer

-100 -80 -60 -40 -20 0 20 40

CCD change (ms)