All-trans retinoic acid induces GADD34 gene expression via transcriptional regulation by Six1-TLE3 and post-transcriptional regulation by p38-TTP in skeletal muscle.

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Increase of GADD34 expression in skeletal muscle by all-trans retinoic acid

MAPK / mRNA stabilization / skeletal muscle type change / transcription / unfolded protein response

All-trans retinoic acid (ATRA) increases the sensitivity to unfolded protein response (UPR) in differentiating leukemic blasts. The downstream transcriptional factors of PERK, a major arm of UPR, regulates muscle differentiation. However, the role of growth arrest and DNA damage-inducible protein 34 (GADD34), one of the downstream factors of PERK, and the effects of ATRA on GADD34 expression in muscle remain unclear. In this study, we identified ATRA increased the GADD34 expression independent of the PERK signal in the gastrocnemius muscle of mice. ATRA up-regulated GADD34 expression through the transcriptional activation of it via inhibiting the interaction of homeobox Six1 and transcription co-repressor TLE3 with the MEF3-binding site on the GADD34 gene promoter in myoblasts. ATRA also inhibited the interaction of TTP, which
induces mRNA degradation, with AU-rich element on GADD34 mRNA via p38 MAPK, resulting in the instability of GADD34 mRNA. Overexpressed GADD34 in myoblasts changes the type of myosin heavy chain in myotubes. These results suggest ATRA increases GADD34 expression via transcriptional and post-transcriptional regulation in myoblasts, which changes muscle fiber type in myotubes.

Introduction

The endoplasmic reticulum (ER) is a membranous organelle that has a central role in protein biosynthesis. Various stresses cause the accumulation of unfolded proteins in the ER, which induces ER stress (Walter & Ron, 2011; Szegezdi et al, 2006). An excessive ER stress response can result in various diseases, such as diabetes, inflammation, and cardiovascular diseases, including vascular calcification (Kim et al, 2008; Masuda et al, 2013; Masuda et al, 2015). To alleviate ER stress, the unfolded protein response (UPR) is initiated by the activation of 3 ER transmembrane sensors: PKR-like endoplasmic reticulum kinase (PERK), inositol-requiring enzyme 1 (IRE1), and activating transcription factor 6 (ATF6). PERK activation leads to the phosphorylation of a subunit of eukaryotic initiation factor 2 (eIF2\(\alpha\)), resulting in the upregulation of activating transcription factor 4 (ATF4) and C/EBP-homologous protein (CHOP).
UPR in skeletal muscle regulates muscle stem cell homeostasis and myogenic differentiation.

Especially, the PERK pathway is one of the key signals to promote the progression and commitment of satellite cells to the myogenic lineage (Rayavarapu et al, 2012; Gallot et al, 2019).

Deletion of PERK, ATF4, or CHOP in satellite cells inhibits myofiber formation via the decreased expression of myoblast determination protein 1 (MyoD) and myogenin, essential transcription factors to correctly differentiate in skeletal muscle following injury (Gallot et al, 2019; Alter & Bengal, 2011; Ebert et al, 2020). We recently reported that UPR is an important regulator of injured skeletal muscle of mice with chronic kidney disease (Niida et al, 2020). Furthermore, sine oculis homeobox homolog 1 (Six1), is a mandatory transcription factor with coactivator eyes absent (Eya) and dachshund (Dach) or corepressor Groucho/transducing-like enhancer of split (TLE) for myogenic differentiation (Jennings & Horowicz, 2008; Sakakibara et al, 2016; Maire et al, 2020). However, little is known about the relationship between Six1 and PERK-related gene in muscle differentiation.

ATF4 promotes the transcriptional activation of growth arrest and DNA damage-inducible protein 34 (GADD34/Ppp1r15a), which dephosphorylates eIF2α by interacting with the catalytic subunit of type 1 protein serine/threonine phosphatase (PP1) (Harding et al, 2000; Zebrucka et al, 2016). Therefore, ATF4-mediated induction of GADD34 functions as a negative feedback loop.
of UPR by eIF2α dephosphorylation, which is essential for cell survival. Although GADD34 plays various roles without UPR signaling in some cells, such as cytokine production in dendritic cells, inhibition of apoptosis in liver cancer cells, and the response to chronic oxidative stress in neurodegenerative diseases (Clavarino et al, 2012; Song et al, 2019; Goh et al, 2018), the specific role of GADD34 in skeletal muscle has not been revealed.

All-trans retinoic acid (ATRA), a metabolite of vitamin A, is a ligand for nuclear receptors, including retinoic acid receptors (RARα, β, and γ). RARs in the nucleus work as a transcription factor by forming a heterodimer with retinoid X receptors (RXRs) and binding to retinoic acid-response elements (RAREs) in target gene promoters (Chambon, 1996; Mangelsdorf & Evans, 1995). ATRA also activates various MAPK kinase cascades, such as p38 and ERK (p44/42), with RARα in membrane lipid rafts and RARγ in the cytosol (Eochette-Egly, 2015). In addition, activated p38 and ERK are involved in not only the transcriptional regulation of the target gene via translocation of the target gene to the nucleus but also the post-transcriptional regulation of the target mRNA by interacting with the mRNA binding proteins (RBPs), such as Tristetraprolin (TTP), which controls mRNA stability (Bhattacharyya et al, 2011; Stoecklin et al, 2004). ATRA has an important role in the differentiation and apoptosis of blood cells and the sensitivity to ER stress through the differentiation of human leukemic cells (Masciarelli et al, 2018). In contrast,
ATRA supplementation ameliorates the ethanol-induced expression of ATF4 and CHOP in the rat liver (Nair et al, 2018). Interestingly, ATRA also controls muscle differentiation via the transcriptional regulation of MyoD and myogenin (Chen et al, 2015; Lamarche et al, 2015), whereas the effects of ATRA on PERK-related genes in skeletal muscle cells is still unknown.

In this study, we investigated the effects of ATRA on the PERK pathway in various organs, especially muscle. Our results demonstrate that ATRA increases the GADD34 gene expression via the downregulation of Six1 and the stabilization of GADD34 mRNA in skeletal muscle. We also characterized the role of GADD34 in skeletal muscle fiber type change.

Results

ATRA induces GADD34 gene expression in the gastrocnemius muscle and undifferentiated skeletal muscle cells without translational activation of ATF4

To assess the response of the PERK signal to ATRA in various tissues, such as the kidney, liver, epididymal adipose (EA), gastrocnemius muscle (GM), and femur, mice were treated with ATRA (total dose: 10 mg/kg body weight). The expression of ATF4 and CHOP mRNA were not changed in all tissues (as described above) of mice treated with ATRA for 24 h. ATRA treatment significantly increased the GADD34 mRNA expression only in the GM of mice (Fig 1A). Western
blotting revealed that ATRA treatment did not increase phosphorylated eIF2α (p-eIF2α), ATF4, or CHOP downstream of the PERK signal in GM in comparison to vehicle; however, it increased GADD34 protein expression levels (Fig 1B). To determine whether the increase in the expression of GADD34 by ATRA is associated with the differentiation of skeletal muscle cells, C2C12 cells were gradually differentiated from myoblasts to myotubes, as shown in Fig 1C. In C2C12 myoblasts, myotubes, and HEK293 cells, the expression of ATF4 mRNA was comparable between DMSO and ATRA treatment, which was in line with the results of in vivo studies. Surprisingly, ATRA increased the GADD34 mRNA and protein expression levels in C2C12 undifferentiated myoblasts, but not in myotubes or HEK293 cells (Fig 1D and E). We generated C2C12 cells with the knockdown of ATF4 to confirm the effects of ATRA on the GADD34 expression independently of ATF4; with the use of ATF4-specific siRNA, the endogenous ATF4 mRNA levels of these C2C12 cells were reduced by more than 60%. ATF4-knockdown did not inhibit the induction of the expression of GADD34 mRNA by ATRA (Fig 1F).

**ATRA upregulates the transcriptional activity of the GADD34 gene in C2C12 cells**

To investigate the molecular mechanisms underlying the undifferentiated muscle-specific regulation of the GADD34 gene expression by ATRA, we examined the responsiveness of human
GADD34 gene promoters to ATRA using a luciferase assay with undifferentiated C2C12 myoblasts and HEK293 cells. The overexpression of RARs increased the luciferase activity of pGADD34-0.5k in both C2C12 myoblasts and HEK293 cells. ATRA increased the luciferase activity of pGADD34-0.5k in C2C12 myoblasts expressing RARα and RARγ, but not RARβ (Fig 2A). ATRA dose-dependently increased the GADD34 gene promoter activity in C2C12 myoblasts, unlike in HEK293 cells (Fig 2B). Surprisingly, TTNPB, a major agonist of RAR, did not increase the GADD34 gene promoter activity, although TTNPB increased the mRNA expression of the target gene (Appendix Fig S1A and B) (Thé et al, 1990). Unlike TTNPB, ATRA can activate the non-genomic p38 and ERK MAPK signals via extranuclear RARα/γ and RARγ, respectively (Tanoury et al, 2013; Bouchard & Paquin, 2013; Khatib et al, 2019). Next, we tested whether these MAPK signals are involved in the regulation of the GADD34 expression by ATRA. As a result, the ATRA-induced GADD34 gene promoter activity was blocked by p38 inhibitor SB203580 or ERK inhibitor FR180204 via RARα/γ or RARγ, respectively (Appendix Fig S1C). Next, to explore the muscle-specific regulatory element of ATRA on the human GADD34 gene promoter, several reporter constructs lacking portions of the 5’-promoter region of the human GADD34 genes were tested in C2C12 myoblasts overexpressing RAR/RXR, with or without ATRA. These
deletion analyses suggest that the sequence from -162 to -131 is responsible for ATRA-dependent activation of the human GADD34 gene promoter activity in C2C12 myoblasts (Fig 2C).

Six1 downregulates the transcriptional activity of the GADD34 gene via MEF3-binding site in C2C12 cells

To explore the molecular mechanism underlying the responsiveness of the GADD34 gene to ATRA in C2C12 myoblasts, we searched for undifferentiated muscle-specific transcription factor binding sites from -162 to -131 on the human GADD34 gene promoter with MatInspector-Genomatix. A search for transcription factor binding motifs with this region suggested five potential binding sites: three MEF3 sites, Pax3, and Sox6 (Fig 3A). Based on this search, we overexpressed Six1, a major transcription factor for the MEF3-binding site, Pax3, and Sox6 with GADD34 gene promoter constructs in C2C12 myoblasts. Although each transcription factor decreased the luciferase activity of pGADD34-0.16k, Six1 did not affect the activity of pGADD34-0.13k, unlike Pax3 and Sox6 (Fig 3B). The overexpression of Six1 reduced the GADD34 mRNA and protein expression levels in C2C12 myoblasts (Fig 3C and D). To investigate which MEF3-binding sites are responsible for the decrease in GADD34 gene promoter activity induced by Six1, we made luciferase reporter vectors of three mutated MEF3-binding sites in the pGADD34-0.13k,
as shown in Fig 3E. These mutation analyses showed that the MEF3-2 sequence in the MEF3-binding sites is essential for the repression of the GADD34 gene promoter activity by Six1 (Fig 3F). Next, we examined whether Six1 binds to the MEF3-2 sequence on the GADD34 gene promoter with an EMSA analysis. A radiolabeled oligonucleotide containing a GADD34-MEF3 probe, but not a mutated GADD34-MEF3 probe (GADD34 Mut-M2), detected a band in nuclear extracts prepared from C2C12 cells overexpressing Six1 (Fig 3G and H). Although these complexes were susceptible to competition with unlabeled GADD34-MEF3, an unlabeled mutated oligonucleotide (M2: GADD34-Mut-M2) and consensus C/EBP did not compete with these complexes (Fig 3G). These results suggest that Six1 may be involved in the induction of the GADD34 gene promoter activity by ATRA.

ATRA upregulates the human GADD34 gene expression through the reduction of the Six1 expression in C2C12 cells

Next, we investigated the effects of ATRA on the Six1 expression in several tissues of mice. The Six1 mRNA levels were undetectable in the kidney, liver, and EA. ATRA treatment suppressed the mRNA expression of Six1 (which is highly expressed only in the GM of mice, as well as the protein expression of Six1 (Fig 4A and B). The mRNA expression levels of Pax3 and Sox6, which
affect \textit{GADD34} gene promoter activity, were not changed in the GM of ATRA-treated mice in comparison to mice treated with DMSO (Appendix Fig S2A). Likewise, ATRA treatment downregulated the of Six1 mRNA and protein expression in C2C12 myoblasts, but not myotubes (Fig 4C and D). We also examined the time-dependent effects of ATRA on the mRNA expression of \textit{Six1} and \textit{GADD34} for up to 24 h in C2C12 myoblasts. In comparison to DMSO, ATRA transiently decreased the expression of \textit{Six1} mRNA only after 24 h of treatment. It also increased the expression of \textit{GADD34} mRNA; this was observed at both 3 h and 24 h (Fig 4E). The mutated promoter constructs shown in Fig 3E were transfected in C2C12 myoblasts to examine the effects of ATRA on MEF3-binding sites on the \textit{GADD34} gene promoter. These mutation analyses suggested that the binding of Six1 to the MEF3-2 sequence is essential for the activation of the \textit{GADD34} gene promoter by ATRA (Fig 4F). An EMSA assay showed that the band detected by a contact with a radiolabeled oligonucleotide containing a GADD34-MEF3 probe and nuclear extracts prepared from C2C12 cells treated with ATRA was decreased in comparison to treatment with DMSO (Fig 4G). Moreover, Six1-knockdown increased the GADD34 protein expression and inhibited ATRA-induced GADD34 protein expression in C2C12 myoblasts (Fig 4H). As shown in Appendix Fig S1C, since p38 and ERK are involved in the increase of the \textit{GADD34} gene promoter activity by ATRA, we tested whether these MAPK signals regulate the mRNA expression of \textit{Six1}
and GADD34 in myoblasts. As a result, p38 and ERK inhibitors blocked the decrease of the Six1 mRNA expression and the increase of the GADD34 mRNA expression induced by ATRA (Appendix Fig S1D). These results indicated that ATRA increases the GADD34 expression via suppression of the binding of Six1 with the MEF3-2 sequence on the human GADD34 gene promoter by decreasing the expression of Six1.

ATRA downregulates the transcriptional activity of the GADD34 gene via co-repressor TLE3 with Six1 in C2C12 cells

Six1 requires the Eya family as the co-activator for transcriptional activation but requires the TLE family as the co-repressor for transcriptional repression (Jennings & Horowicz, 2008). To define a co-repressor that is responsible for the Six1-mediated reduction of the GADD34 gene expression, we first analyzed the mRNA expression levels of the TLE family (TLE1-6) by qPCR with the absolute standard curve method. Among the TLE family, the TLE1, TLE3, and TLE5 genes were expressed in C2C12 myoblasts (Fig 5A), we generated C2C12 cells with the knockdown of these genes using specific siRNA for each of these genes. As shown in Fig 5B, TLE1, TLE3, and TLE5 siRNAs reduced the expression of these genes by 50%, 80%, and 99%, respectively. Although pGADD34-0.16k significantly increased the luciferase activity in response
to TLE3-knockdown—but not TLE1- and TLE5-knockdown—pGADD34-0.13k exhibited no increase in response to these knockdowns (Fig 5C). The mutated promoter constructs shown in Fig 3E revealed that the MEF3-2 sequence is essential for the activation of the GADD34 promoter by TLE3-knockdown (Fig 5D). TLE3-knockdown blocked the effects of the luciferase activity of pGADD34-0.16k by the overexpression of Six1 or ATRA treatment (Fig 5E). On the other hand, ATRA did not change the mRNA expression of TLE1, TLE3, and TLE5 in C2C12 myoblasts (Appendix Fig S2B). These results indicate that ATRA induces the transcriptional activity of the GADD34 gene through the inhibition of the collaborative work in TLE3 and Six1 through the decreased expression of Six1.

ATRA increases the stability of GADD34 mRNA via TTP in a p38 dependent manner

Thus far, we have demonstrated that ATRA increases the GADD34 expression by transcriptional regulation through the reduced expression of Six1. However, in comparison to vehicle, ATRA transiently increased the GADD34 mRNA expression after 3 h of treatment, unlike the Six1 mRNA expression (Fig 4E). Similarly to the mRNA expression, Western blotting revealed that the GADD34 protein expression was increased by ATRA treatment at 3 h in myoblasts (Fig 6A). These results imply the presence of the regulation of GADD34 expression by ATRA,
independently of the action of Six1. Next, we tested the stabilization of GADD34 mRNA at early time points using actinomycin D, a transcriptional inhibitor. As a result, ATRA promoted GADD34 mRNA stability at 3 h in C2C12 myoblasts that were treated with actinomycin D (Fig 6B). Generally, mRNA stability is regulated through an AU-rich element (ARE: AUUUA motif) in the 3’-untranslated region (3’UTR) of mRNA. To investigate the molecular mechanism underlying the GADD34 mRNA stabilization in response to ATRA via the 3’UTR of its mRNA, we created a pGL3-GADD34-3’UTR construct (GADD34-3’UTR) by replacing the 3’UTR of the pGL3-basic with the 3’UTR of the GADD34 gene, which contained two AREs, as illustrated in Fig 6C. ATRA dose-dependently stimulated the luciferase activity of the GADD34-3’UTR, but not pGL3-basic empty (control), in C2C12 myoblasts (Fig 6C).

Since the mRNA stability requires signal transduction pathways of MAPK, such as p38, ERK, and JNK in lung cancer cells (Bhattacharyya et al, 2011), we examined which MAPK pathway is responsible for the stabilization of GADD34 mRNA by ATRA using p38 and ERK inhibitors. Unlike ERK1/2 inhibitor, p38 inhibitor completely suppressed the ATRA-induced luciferase activity of pGADD34-3’UTR (Fig 6D; Appendix Fig S3A and B). The phosphorylation of p38 inhibits the TTP contact with ARE, resulting in the induction of target mRNA stabilization (Bhattacharyya et al, 2011). Although ATRA and TNFα, a p38 phosphorylation inducer, stimulated the luciferase
activity of GADD34-3'UTR, the effects of these treatments on its activity in C2C12 cells were
canceled by TTP-knockdown using TTP-specific siRNA (Fig 6E; Appendix Fig S3C–E). In addition
to TTP, human antigen R (HuR), one of the major RBPs, inhibits mRNA degradation (Stoecklin
et al, 2004). Unlike TTP-knockdown, HuR-knockdown did not change the luciferase activity of
GADD34-3UTR induced by ATRA (Appendix Fig S3F and G). To further test whether the ARE1
and ARE2 sequences are responsible for the regulation of GADD34 mRNA stability by ATRA, we
determined the luciferase activity of the mutated ARE1 (Mut-A1) and mutated ARE2 (Mut-A2) in
the GADD34-3'UTR (Fig 6F). These mutation analyses showed that both of AREs are important
for the enhancement of the stabilization of the GADD34 mRNA by ATRA (Fig 6F). Using an RNA
ChIP assay, we confirmed that p38 inhibitor treatment recovered the decreased binding between
GADD34 mRNA and TTP that was induced by ATRA (Fig 6G). These results suggest that ATRA
suppress TTP-induced GADD34 mRNA degradation through the binding of ARE on the 3'UTR of
human GADD34 mRNA in a p38-dependent manner.

GADD34 decreases the expression of MYHC2a and changes the muscle fiber type.

Finally, C2C12 myoblasts transfected with human GADD34 expression vector or empty (control)
at Day 0 were collected at Days 0, 1, 2, and 4 to examine the effects of GADD34 on the C2C12
phenotype (Fig 7A). Unexpectedly, the overexpression of GADD34 did not affect the
differentiation speed, thickness of myotubes, the mRNA expression of major muscle
differentiation marker genes (Pax7, Myf5, MyoD, and Myogenin) or muscle-specific E3 ubiquitin
ligase (MuRF1 and Atrogin1), or the synthesis of proteins in C2C12 cells (Fig 7B–D; Appendix
Fig S4A–C). Although the GADD34 overexpression did not change the total MYHC protein
expression in C2C12 cells from Day 0 to 4, the protein expression of MYHC1 (type 1 slow fibers)
was slightly increased in C2C12 cells that overexpressed GADD34 on Day 4. In contrast, the
overexpressed GADD34 slightly decreased the protein expression of MYHC2 (type 2 fast fibers)
and MYHC2 isoforms, such as MYHC2a and MYHC2x, but not MYHC2b, in C2C12 cells on Day
4 (Fig 7E). In addition, the overexpression of GADD34 downregulated the mRNA expression of
MYH2 and MYH1, which respectively encode MYHC2a and MYHC2x proteins (Fig 7F). These
results suggest that GADD34 regulates muscle fiber types, such as MYHC2a and MYHC2x, by
transcriptional control.

Discussion

In the present study, we showed the molecular mechanisms by which ATRA increases the
GADD34 expression independently of ATF4 (known to be a potent inducer of GADD34) in
myoblasts. Likewise, although a recent study reported that GADD34 levels were decreased in satellite cells collected from injured muscle with increased ATF4 (Xiong et al, 2017), the mechanism of this paradoxical result is unknown. Since muscle atrophy can occur with differential sensitivity due to selective skeletal muscle fiber subtypes in various pathological conditions, muscle fiber type change is suggested to be an important factor for muscle wasting (Wanga and Pessinn 2013). Muscle atrophy is a frequent complication in CKD patients (Carrero et al, 2013). Interestingly, the plasma levels of vitamin A (retinol or ATRA) increase in CKD patients (Gueguen et al, 2005; Jing et al, 2016). It has been reported that the percentage of type 1 slow fiber in type 2 fast fiber increases in CKD mice through a decrease in the expression of MYHC2a (Tamaki et al, 2014). However, the molecular mechanisms underlying the muscle fiber type change in CKD are largely unknown. In the present study, we revealed that the overexpression of GADD34 increases the MYHC1 protein expression, which is expressed in type 1 slow fibers, and decreases the MYHC2 protein expression, which is expressed in type 2 fast fibers, in myotubes. Based on these results and reports, our findings may provide clues in relation to the unknown mechanisms of pathogenesis, such as muscle atrophy, in CKD patients.

Among the Six family of homeobox (Six1/Six2, Six3/Six6, and Six4/Six5), Six1, Six2, and Six4 are expressed in myoblasts (Kumar, 2009; Grand et al, 2012). MyoD reprogramming ability
(whereby the MyoD expression turns extra muscle cells into muscle) is impaired in mouse embryonic fibroblasts in Six1/4 double mutant mice because Six1 and MyoD interact with the Myogenin gene promoter to differentiate muscle (Santolini et al, 2016). On the other hand, TLE3 downregulates myogenic differentiation via the suppression of MyoD activity (Kokabu et al, 2017).

Six1 requires the Eya family as the co-activator for transcriptional activation but the TLE family as the co-repressor for transcriptional suppression. In the present study, we suggested that—among the TLE family—TLE3 is the most abundant gene in myoblasts and is an important co-repressor of Six1 for the regulation of GADD34 gene transcriptional activity, which consequently changes the ratio of muscle fiber type. In addition, we confirmed that other undifferentiated muscle-specific transcriptional factors, Pax3 and Sox6, also suppress the GADD34 gene promoter activity. These findings suggest that the expression of GADD34 may constantly be kept low by not only Six1-TLE3 but also by undifferentiated muscle-specific transcriptional factors. However, the reason why the expression of GADD34 is constantly low in myoblasts is unknown.

The expression of genes is determined by transcriptional regulation and post-transcriptional regulation via mRNA stability. The 3'UTR works as one of the regulatory components of mRNA degradation by promoting or inhibiting the deadenylation via interaction with RBP (Mayr, 2019).

Although the classic target mRNAs that interact with RBPs for mRNA degradation are associated
with inflammatory-related genes, a recent study revealed that other target mRNAs are related to
cancer, apoptosis, and other conditions (Kaempfer, 2003; Schuster & Hsieh, 2019; Lal et al, 2005).
However, UPR-related mRNA has not previously been reported as an ARE-dependent target for
stabilization. TTP and HuR, the most famous RBP interacted with ARE (5'-AUUUA-3') on the
3'UTR, to induce mRNA degradation and stabilization, respectively. However, unlike TTP, HuR
did not regulate GADD34 mRNA stability in the present study. The distinction between HuR and
TTP binding has been reported to involve subtle content features: TTP: 5'-
AUA[A/U/G/C][A/U][A/U]-3' and HuR: 5'-[A/C/U]UUUU[U/A/C][U/A]-3' (Bhandare et al, 2017). In
other words, HuR strongly prefers U-rich sequences, whereas TTP prefers AU-rich with
increasing A content. This may explain why HuR did not interact with the ARE sequences of
GADD34 mRNA in the present study. ATRA activates the non-genomic p38 MAPK via
extracellular RARα and RARγ (Tanoury et al, 2013). Furthermore, phosphorylated p38 inhibits
the TTP interaction with ARE on target mRNA (Bhattacharyya et al, 2011). Our results indicated
that ATRA stabilizes GADD34 mRNA by inhibiting the interaction between its mRNA and TTP via
p38 MAPK. Although it has been reported that ATRA regulates the stabilization of mRNA via
interaction between apo-cellular retinoic acid-binding protein 2 (apo-CRABP2) and HuR
(Vreeland et al, 2014), this is the first study to report that ATRA-dependent MAPK activation
increases—via ARE—the mRNA stability of the 3'UTR of the target mRNA. Because p38 MAPK signaling regulates a large number of cellular processes, the mechanism underlying mRNA stabilization by ATRA may also be involved in many mRNAs other than GADD34 mRNA.

Although the Six genes, especially Six1, are expressed in multiple organs during mammalian development, their expression in global tissues decreases as the individual grows to adulthood (Kumar, 2009). Six1 also controls muscle physiology not only in embryogenesis but also in the adult (Maire et al., 2020). In soleus, Six1 deficiency reduced MYHC2a fiber in 3-week-old C57BL6 mice and caused the complete disappearance of the expression of MYHC2a fiber in 12-week-old C57BL6 mice (Sakakibara et al., 2016). These results imply that Six1 regulates fast-fiber type acquisition and maintenance in adult mice. Although the effects of GADD34 on skeletal muscle in vivo are unknown, our study revealed that GADD34, which is downregulated by Six1, suppressed the MYHC2a expression in myoblasts. Thus, GADD34 may mediate the regulation of the expression of MYHC2a by Six1 in myotubes. Aside from this, GADD34 induces cellular senescence via the regulation of the p21 expression in MEF and 7EJ-Ras cells (Minami et al., 2007). Cellular senescence, a permanent state of replicative arrest in otherwise proliferating cells, is a hallmark of aging and has been linked to aging-related diseases (Childs et al., 2015). Since the plasma levels of vitamin A (retinol or ATRA) increase with aging (Gueguen et al., 2005), the
increase of ATRA that occurs with aging may accelerate cellular senescence in the muscle via
the induction of GADD34, resulting in muscle atrophy in elderly patients. It has been reported that
aged mice exhibit an increase of slow muscle fibers but a decrease of fast muscle fibers (Shang
et al, 2020). These results are in line with the effects of GADD34 on myotubes that were observed
in our study.

In conclusion, as shown in Fig 8, our findings revealed that the upregulation of the GADD34
gene expression by ATRA contributes to muscle fiber type change in myoblasts. Furthermore,
ATRA and its receptors can increase the transcriptional activity of the GADD34 gene by blocking
the interaction of Six1-TLE3 with the GADD34 gene promoter and stabilize GADD34 mRNA by
inhibiting the interaction of p38-TTP with the 3'UTR of GADD34 mRNA.

Materials and Methods

Chemicals and reagents

ATRA, DMSO, FR180204, anti-GAPDH antibody, anti-Six1 antibody, goat anti-mouse IgG (H +
L)-HRP conjugate, goat anti-rabbit IgG (H + L)-HRP conjugate, MISSION siRNA oligos were
purchased from Sigma-Aldrich (St. Louis, MO, USA). Buprenorphine hydrochloride was
purchased from Otsuka Pharmaceutical Co., Ltd. (Tokyo, Japan). Pentobarbital sodium salt was
purchased from Tokyo Kasei Co., Ltd. (Tokyo, Japan). Anti-phosphorylated eIF2α (p-eIF2α) (#9721, Ser51), anti-eIF2α (#9722), anti-ATF4 (#11815), anti-CHOP (#2895), anti-TTP (#71632), anti-phosphorylated ERK (p-ERK) (#9101, Thr202/Try204), and anti-ERK (#9102) antibody were purchased from Cell Signaling Technology (Danvers, MA, USA). Anti-phosphorylated p38 (p-p38) (sc-166182, Tyr182), anti-p-38 (sc-7972), and anti-HuR (sc-5261) antibody, and C/EBP consensus oligonucleotide (cebp; catalog number sc-2525) were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Anti-GADD34 antibody (10449-1-AP) was purchased from Proteintech (Rosemont, IL, USA). Anti-MYHC1 (BA-D5), anti-MYHC2 (F59), anti-MYHC2a (SC-71), anti-MYHC2x (6H1), and anti-MYHC2b (BF-F3) antibody were purchased from Developmental Studies Hybridoma Bank (Iowa city, IA, USA). Anti-puromycin antibody was purchased from Cosmo Bio (Tokyo, Japan). [γ-32P] ATP was purchased from ICN (Costa Mesa, CA, USA). T4 polynucleotide kinase and TransIT®-LT1 Reagent were purchased from TAKARA (Shiga, Japan). SB203580 was purchased from Adipogen Life Science (San Diego, CA, USA). Human recombinant TNFα was purchased from Invitrogen (Carlsbad, CA, USA). Puromycin was purchased from Funakoshi (Tokyo, Japan). Chemi-Lumi One Super was purchased from Nacalai Tesque (Kyoto, Japan). QIAzo® Lysis Reagent was purchased from QIAGEN (Venlo, Nederland).
TOYOBO KOD one™ PCR Master Mix -Blue- was purchased from TOYOBO (Osaka, Japan).

RNA ChIP-IT® was purchased from Active Motif (Tokyo, Japan).

Animal experiments

The animal work took place in Division for Animal Research and Genetic Engineering Support Center for Advanced Medical Sciences, Institute of Biomedical Sciences, Tokushima University Graduate School. Eight-week-old male C57BL/6J mice were purchased from Japan SLC (Shizuoka, Japan) and individually caged in a climate-controlled room (22 ± 2°C) with a 12-h light-dark cycle. These mice were randomly divided into two groups and a total of 1 mg/kg body weight of ATRA or 1% DMSO (control) prepared in sterile saline was intraperitoneally administered, then the mice were sacrificed 24 h later. Before sacrifice, mice were anesthetized with a total of 0.1 mg/kg body weight of buprenorphine hydrochloride and a total of 50 mg/kg body weight of pentobarbital sodium salt, and tissues were removed. The present study was approved by the Animal Experimentation Committee of Tokushima University School of Medicine (animal ethical clearance No. T30-66) and was carried out in accordance with guidelines for the Animal Care and use Committee of Tokushima University School of Medicine.
Cell culture

C2C12 myoblast cells were cultured as described previously (Niida et al, 2020). Briefly, C2C12 myoblast cells were cultured in DMEM (Sigma, St. Louis, MO) containing 10% FBS (Sigma), 100 units/ml penicillin and 100 µg/ml streptomycin at 37°C with 5% CO₂. At 100% confluence, C2C12 myoblast cells were fused by shifting the medium to DMEM containing 2% horse serum (HS; Moregate Biotech, Bulimba, Australia). These cells were maintained in 2% HS/DMEM (differentiation medium) for 4 days prior to experiments. HEK293 cells were cultured in DMEM containing 10% FBS, 100 units/ml penicillin and 100 µg/ml streptomycin at 37°C with 5% CO₂.

Western blotting

Protein samples were heated to 95°C for 5 min in sample buffer in the presence of 5% 2-mercaptoethanol and subjected to SDS-PAGE. The separated proteins were transferred by electrophoresis to polyvinylidene difluoride transfer membranes (Immobilon-P, Millipore, MA, USA). The membranes were treated with diluted affinity-purified anti-p-eIF2α (p-eIF2α) (1:1000), anti-eIF2α (1:1000), anti-ATF4 (1:3000), anti-CHOP (1:1000), anti-GADD34 (1:1000), anti-Six1 (1:1000), anti-TTP (1:1000), anti-HuR (1:1000), anti-p-p38 (p-p38) (1:1000), anti-p38 (1:1000), anti-p-ERK (p-ERK) (1:1000), anti-ERK (1:1000), anti-MYHC1 (1:1000), anti-MYHC2 (1:1000), anti-p-p38 (p-p38) (1:1000), anti-p38 (1:1000), anti-p-ERK (p-ERK) (1:1000), anti-ERK (1:1000), anti-MYHC1 (1:1000), anti-MYHC2 (1:1000),
anti-MYHC2a (1:1000), anti-MYHC2x (1:500), anti-MYHC2b (1:500), and anti-puromycin (1:2000) antibody. Mouse anti-GAPDH monoclonal antibody was used as an internal control. Goat anti-
mouse IgG (H + L)-HRP conjugate (1:3000) and Goat anti-rabbit IgG (H + L)-HRP conjugate
(1:3000) was utilized as the secondary antibody, and signals were detected using the Chemi-
Lumi One Super.

Quantitative PCR

Total RNA was isolated from kidney, liver, EA, GM, femur, C2C12 myoblasts, C2C12 myotubes,
and HEK293 cells using an QIAzol® Lysis Reagent. Real-time quantitative PCR assays were
performed using an Applied Biosystems StepOne qPCR instrument. In brief, the cDNA was
synthesized from 1 µg of total RNA using a reverse transcriptase kit (Invitrogen, Carlsbad, CA)
with an oligo-dT primer. After cDNA synthesis, quantitative real-time PCR was performed in 5 µl
of SYBR Green PCR master mix using a real-time PCR system (Applied Biosystems).

Amplification products were then analyzed by a melting curve, which confirmed the presence of
a single PCR product in all reactions (apart from negative controls). The quantification of given
genes was expressed as the mRNA level normalized to a ribosomal 18S housekeeping gene
using the ΔΔCt method. Quantitative expression values were calculated from an absolute
standard curve method using the plasmid template for each target gene. The primer sequences used for real-time PCR analysis are shown in Appendix Table S1.

4 Reporter plasmid construction

The promoter fragment of luciferase reporter plasmid pGADD34-0.5k generated by Eurofins Japan (Tokyo, Japan) was subcloned into a pGL-3 vector (Promega, Madison, WI, USA) by restriction enzyme cutting site KpnI/HindIII. Luciferase reporter plasmids GADD34-3’UTR was constructed by PCR amplification of human genomic cDNA as a template using gene-specific primers (Appendix Table S2). These products were subcloned into a pGL-3 vector. Deleted reporter plasmid pGADD34-0.3k was cloned by the digestion of pGADD34-0.5k using DpnI/HindIII. Deleted reporter plasmids pGADD34-0.16k, pGADD34-0.13k, and mutated reporter plasmids pGADD34-0.16k-Mut-MEF3-1 (Mut-M1), pGADD34-0.16k-Mut-MEF3-2 (Mut-M2), pGADD34-0.16k-Mut-MEF3-3 (Mut-M3), GADD34-3’UTR-Mut-ARE-1 (Mut-A1), GADD34-3’UTR-Mut-ARE-2 (Mut-A2) were constructed with TOYOBO KOD one™ PCR Master Mix -Blue- using the oligonucleotides shown in Appendix Table S2. The β-galactosidase expression vector pCMV-β (CLONTECH, Palo Alto, CA, USA) was used as an internal control. Each plasmid was purified with a FavorPrep™ Plasmid DNA Extraction Midi Kit (Favorgen, Ping-Tung, Taiwan).
Transfection and luciferase assay

Mouse Six1, Pax3, and Sox6 expression vector (pCR3-Six1, pCR3-Pax3, and pCR3-Sox6) were kindly provided by Dr. P. Maire (Santolini et al, 2016). Human GADD34 expression vector (pRP-Neo-CMV-hPPP1R15A) was designed by Vector Builder. Cells were transfected by TransIT®-LT1 Reagent, then treated with several concentrations of ATRA, SB203580, FR180204, TNFα, or DMSO as vehicle control for an additional 16 h. A luciferase assay was performed as described previously (Masuda et al, 2010).

RNAi experiments

C2C12 myoblasts were transfected with siRNA directed against ATF4 (SASI_Mm02_00316863), Six1 (SASI_Mm01_00198104), TLE1 (SASI_Mm01_00069933), TLE3 (SASI_Mm02_00300046), TLE5 (SASI_Mm01_00139428), TTP (SASI_Mm01_00178605), HuR (SASI_Mm02_0031872), or negative control (SIC001) using TransIT®-LT1 Reagent, according to the manufacturer's instructions.

EMSA
EMSA was performed as described previously (Masuda et al, 2020). Double-stranded nucleotides for GADD34-MEF3 and GADD34-MEF3-2-Mutant (GADD34-Mut-M2) were synthesized (Appendix Table S3). Purified DNA fragments were radiolabeled with [γ-32P] ATP (110 TBq/mmol) using T4 polynucleotide kinase. Nuclear extracts (pCR3-Six1) were prepared as described previously (Masuda et al, 2010). Briefly, the C2C12 myoblast cells were cultured in 10-cm dishes to 60% confluence and transfected with pCR3-Six1 or treated with 1 µM ATRA. Prepared nuclear extracts (15 µg) were incubated with the radiolabeled probe in binding buffer [10 mM (Tris–HCl), pH7.5, 1 mM DTT, 1 mM EDTA, 10% Glycerol, 1 mM MgCl2, 0.25 mg/ml bovine serum albumin, 2.5 µg/ml salmon sperm DNA and 2 µg poly(dI-dC)] in a final volume of 20 µl for 30 min at room temperature. The specificity of the binding reaction was determined with a 100-fold molar excess of the indicated cold competitor oligonucleotide. The reaction mixture was then subjected to electrophoresis on a 5% polyacrylamide gel with 0.25× TBE running buffer for 2 h at 150 V. The gel was dried and analyzed with an image scanner (FLA-9000 Starion, Tokyo, Japan).

RNA ChIP assay
Using the RNA ChIP-IT®, an RNA ChIP assay was performed according to the manufacturer's instructions. Briefly, the C2C12 myoblast cells were cultured to 80% confluence in 10-cm dishes and treated with 1 µM ATRA. The C2C12 cells were collected and lysed in lysis buffer. The cell extract was prepared and incubated with RNA ChIP buffer pre-conjugated with TTP antibodies or control mouse IgG at 4°C for 16 h. The complexes were treated with Proteinase K for 1 h at 45°C and 1.5 h at 65°C. Immunoprecipitated RNA in the precipitates was purified using QIAzol® Lysis reagent and analyzed for GADD34 by RT-qPCR.

Measurement of C2C12 diameters

The myotube diameters were determined as previously reported (Niida et al, 2020). Under a fluorescence microscope (BIOREVO BZ-9000, Keyence), three photographs per cell-culture well were obtained in a high-power field. We measured the diameter at the middle portion of the myotube with the built-in BZ-II analyzer software program. We measured the diameters of 100 myotubes/group.

Surface Sensing of Translation (SUnSET) assay in C2C12 cells
C2C12 was incubated with or without 1 μM puromycin for 30 min before collecting the cells and washed with PBS as previously described (Lim et al, 2017). Puromycin-labeled proteins were detected by Western blotting, as shown above.

Statistical analysis

Data were collected from more than two independent experiments and were reported as the mean and SEM. The statistical analysis for two-group comparison was performed using a two-tailed Student’s t-test, or one-way ANOVA with Tukey-Kramer post-hoc test for multi-group comparison. All data analyses were performed using the GraphPad Prism 5 software program (GraphPad Software, San Diego, CA, USA). P values of < 0.05 were considered to indicate statistical significance.

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Figure legends

Figure 1. Effects of ATRA treatment on PERK signaling in skeletal muscle.

A, B  Eight-week-old male C57BL/6J mice were given an intraperitoneal injection of 0.1% DMSO (vehicle) and ATRA (10 mg/kg) for 24h. (A) The mRNA expression levels of ATF4, CHOP, and GADD34 in kidney, liver, epididymal adipose (EA), gastrocnemius muscle (GM), and femur of mice were determined by real-time PCR (n = 4 mice per group). (B) Western blotting of phosphorylated-eIF2α (p-eIF2α), total eIF2α (eIF2α), ATF4, CHOP, and GADD34 in GM (n = 4 mice per group).

C  Schematic illustration of the experimental timeline in C2C12.

D  The mRNA expression levels of ATF4 and GADD34 in C2C12 cells (myoblasts and myotubes) and HEK293 cells treated with the indicated concentrations of ATRA (n = 3–4).

E  Western blotting of GADD34 in C2C12 cells myoblasts with the indicated concentrations of ATRA.

F  The mRNA expression levels of GADD34 in C2C12 myoblasts transfected with ATF4 siRNA (siATF4) or control and incubated in the presence of 1 μM ATRA or DMSO as a vehicle for 24 h (n = 3–4).
Data information: In (A–B), data are presented as the mean ± SEM. *p < 0.05 (two-tailed unpaired Student’s t-test). In (D, F), data are presented as the mean ± SEM. *p < 0.05 vs. DMSO (NT) (one-way ANOVA with a Student-Newman post-hoc test). N.S. = not significant.

Source data for this figure are available online.

Figure 2. Activation of human GADD34 gene promoter by ATRA and its receptors in C2C12 cells.

A A schematic illustration of the human GADD34 gene promoter in the upper panels. pGADD34-0.5k and pCMV-β were transfected with pSG5-RAR (α, β, γ) and pSG5-RXRα, or empty vector and incubated in the presence of 1 μM ATRA or DMSO (NT) as a vehicle for 24 h in C2C12 and HEK293 cells (n = 3–4).

B C2C12 (black circles) and HEK293 (white circle) cells were transfected with pGADD34-0.5k, pSG5-RARα, and pSG5-RXRα, and treated with the indicated concentrations of ATRA for 24 h (n = 3–4).

C Transcriptional activity of deletion constructs of GADD34 gene promoters (pGADD34-0.5k, pGADD34-0.3k, pGADD34-0.16k, and pGADD34-0.13k). Deletion constructs are illustrated in the panels on the left. C2C12 cells were transfected with the indicated GADD34 gene reporter
constructs, pSG5-RARα and pSG5-RXRα and incubated in the presence of 1 μM ATRA or DMSO (NT) as a vehicle for 24 h (n = 3–4).

Data information: In (A), data are presented as the mean ± SEM. *p < 0.05 vs. empty vector. #p < 0.05 (one-way ANOVA with a Student-Newman post-hoc test). In (B), data are presented as the mean ± SEM. *p < 0.05 vs. NT (two-tailed unpaired Student’s t-test). In (C), data are presented as the mean ± SEM. *p < 0.05 (two-tailed unpaired Student’s t-test). Similar results were obtained from independent experiments.

Figure 3. Effects of Six1 on the MEF3 site and the mutation analysis of the GADD34 gene promoter.

A Myoblast-specific transcription factor binding sites (Pax3 and Sox6 sequences: underline, MEF3 sequences: box) in the human GADD34 gene promoter region.

B Each GADD34 reporter plasmid (pGADD34-0.16k and pGADD34-0.13k) was transfected with each pCR3 plasmid (empty, Six1, Pax3, or Sox6) into C2C12 myoblasts (n = 3–4).

C The mRNA expression levels of GADD34 in C2C12 myoblasts transfected with each pCR3 plasmid (empty or Six1) (n = 3–4).
D Western blotting of Six1 and GADD34 in C2C12 myoblasts transfected with each pCR3 plasmid (empty or Six1).

E Three MEF3 sites mutated in the GADD34 gene reporter region are underlined. Mut-M1, Mut-M2, and Mut-M3 targeted MEF3-1, MEF3-2, and MEF3-3 respectively.

F Each GADD34 reporter plasmid (WT, Mut-M1, Mut-M2, and Mut-M3) was transfected with pcDNA3 empty or Six1 into C2C12 myoblasts (n = 3–4).

G, H EMSAs using $^{32}$P-labelled (G) GADD34-MEF3 and (H) mutated GADD34-MEF3-2 (GADD34-Mut-M2) as probes. EMSAs were performed with nuclear extracts (N.E.) from C2C12 myoblasts overexpressing Six1 or empty with the addition of unlabeled competitor oligonucleotides, as indicated. A 100-fold molar excess of each competitor was used. The location of the DNA-protein complex band is indicated by an arrowhead. cold, WT GADD34-Six1; M2, GADD34-Mut-M2; C/EBP, C/EBP-binding sequence.

Data information: In (B), data are presented as the mean ± SEM. *p < 0.05 vs. empty vector (one-way ANOVA with a Student-Newman post-hoc test). In (C, F), data are presented as the mean ± SEM. *p < 0.05 (two-tailed unpaired Student’s t-test). N.S. = not significant. Similar results were obtained from independent experiments.

Source data for this figure are available online.
Figure 4. Effects of ATRA treatment on Six1 and GADD34 expression in skeletal muscle.

A  The mRNA expression levels of Six1 in kidney, liver, epididymal adipose (EA), gastrocnemius muscle (GM), and femur of ATRA-treated mice (n = 4 mice per group).

B  Western blotting of Six1 in GM of ATRA-treated mice (n = 4 mice per group).

C  The mRNA expression levels of Six1 in C2C12 cells (myoblasts, myocytes, and myotubes) treated with the indicated concentrations of ATRA (n = 3–4).

D  Western blotting of Six1 in C2C12 myoblasts with the indicated concentrations of ATRA.

E  The mRNA expression levels of GADD34 and Six1 in each C2C12 cells (0, 3, 6, 12, and 24 h) incubated in the presence of 1 µM ATRA or DMSO as a vehicle (n=3–4).

F  Each GADD34 reporter plasmid (WT, Mut-M1, Mut-M2, and Mut-M3) was transfected and incubated in the presence of 1 µM ATRA or DMSO (NT) as a vehicle for 24 h in C2C12 myoblasts (n = 3–4).

G  EMSAs using ³²P-labelled GADD34-MEF3 as probes. EMSAs were performed with nuclear extracts (N.E.) from C2C12 myoblasts treated with 1 µM ATRA or DMSO as a vehicle for 24 h. The location of the DNA-protein complex band is indicated by an arrowhead.
Western blotting of Six1 and GADD34 in C2C12 myoblasts transfected with Six1 siRNA (siSix1) or control (siCont.) and incubated in the presence of 1 μM ATRA or DMSO as a vehicle for 24 h.

Data information: In (A, B, F), data are presented as the mean ± SEM. *p < 0.05 (two-tailed unpaired Student’s t-test). In (C), data are presented as the mean ± SEM. *p < 0.05 vs. NT (DMSO) (one-way ANOVA with a Student-Newman post-hoc test). In (E), data are presented as the mean ± SEM. *p < 0.05 vs. DMSO (two-tailed unpaired Student’s t-test). U.D. = undetectable. N.S. = not significant. Similar results were obtained from independent experiments.

Source data for this figure are available online.

**Figure 5. Effects of the TLE family on GADD34 gene promoter activity in C2C12 myoblasts.**

A The quantitative expression values of the TLE family (TLE1–6) were calculated using the absolute standard curve method of real-time PCR with plasmid templates containing each target gene (n = 3).

B The mRNA expression levels of TLE1, TLE3, and TLE5 in C2C12 myoblasts transfected with each siRNA (siCont., siTLE1, siTLE3, or siTLE5) (n = 3–4).
C Each GADD34 reporter plasmid (pGADD34-0.16k and pGADD34-0.13k) was transfected with each siRNA (siCont., siTLE1, siTLE3, or siTLE5) into C2C12 myoblasts (n = 3–4).

D Each GADD34 reporter plasmid (WT, Mut-M1, Mut-M2, and Mut-M3) was transfected with TLE3 siRNA (siTLE3) or control into C2C12 myoblasts (n = 3–4).

E phGADD34-0.13k was transfected with TLE3 siRNA (siTLE3) or control and incubated in the presence of 1 μM ATRA or DMSO as a vehicle for 24 h in C2C12 cells (n = 3–4).

Data information: In (B, D), data are presented as the mean ± SEM. *p < 0.05 (two-tailed unpaired Student’s t-test). In (C), data are presented as the mean ± SEM. *p < 0.05 vs. siCont. (one-way ANOVA with a Student-Newman post-hoc test). In (E), data are presented as the mean ± SEM. *p < 0.05 vs. DMSO (NT) (one-way ANOVA with a Student-Newman post-hoc test). U.D. = undetectable. N.S. = not significant.

Figure 6. Effects of ATRA on GADD34 mRNA stability in C2C12 myoblasts.

A Western blotting of GADD34 in C2C12 myoblasts incubated in the presence of 1 μM ATRA or DMSO (NT) as a vehicle for 3 h (n = 3–4).

B The mRNA expression levels of GADD34 in C2C12 myoblasts treated with the indicated concentrations of ATRA and 2.5 μg/ml Actinomycin D (n = 3–4).
C A schematic illustration of human GADD34 mRNA sequence in the upper panels. pGL3-basic plasmid empty (control) or GADD34-3’UTR was transfected and incubated with the indicated concentrations of ATRA for 3 h in C2C12 myoblasts (n = 3–4).

D, E Human GADD34 mRNA 3’UTR reporter plasmid was transfected (E) with TTP siRNA (siTTP) or control (siCont.) and incubated in the presence of (E) 1 μM ATRA or DMSO (D) and 10 μM SB203580 (p-p38 MAPK inhibitor) for 24 h (n = 3–4).

F A schematic illustration of the mutated human GADD34 mRNA sequence in the upper panels. Mut-A1 and Mut-A2 targeted the binding sites AREs for RNA binding protein respectively. Each human GADD34 mRNA 3’UTR reporter plasmid (WT, Mut-A1, and Mut-A2) was transfected and incubated in the presence of 1 μM ATRA or DMSO as a vehicle for 24 h (n = 3–4).

G The mRNA expression levels of GADD34 by RNA ChIP assay in C2C12 cells treated with 1 μM ATRA and 10 μM SB203580 for 24 h (n = 3–4).

Data information: In (B, C, D, E), data are presented as the mean ± SEM. *p < 0.05 vs. NT (DMSO) (one-way ANOVA with a Student-Newman post-hoc test). In (F), data are presented as the mean ± SEM. *p < 0.05 (two-tailed unpaired Student’s t-test). N.S. = not significant. Similar results were obtained from independent experiments.

Source data for this figure are available online.
Figure 7. Effects of the overexpression of GADD34 on the muscle fibertype in C2C12 myotubes.

A  A schematic illustration showing the experimental timeline in C2C12.

B  Representative images of C2C12 treated as indicated times (Days 0, 1, 2, and 4). Scale bars = 100 µm.

C  The measurement of diameters of C2C12 myotubes at Day 4, as described in MATERIALS and METHODS (n = 100).

D  The mRNA expression levels of Pax7, Myf5, MyoD, and Myogenin in C2C12 cells on each day (Days 0, 1, 2, and 4) (n = 3–4).

E  Western blotting of total MYHC, MYHC1 (Slow), MYHC2 (fast), MYHC2a, MYHC2x, and MYHC2b in C2C12 cells on each day (Days 0, 1, 2, and 4).

F  The mRNA expression levels of MYH7, MYH2, MYH1, and MYH4 in Day 4 C2C12 cells (n = 3–4).

Data information: In (C, D, F), data are presented as the mean ± SEM. *p < 0.05 (two-tailed unpaired Student’s t-test). N.S. = not significant.

Source data for this figure are available online.
Figure 8. Schematic illustration of muscle specific GADD34 induction through ATRA dependent transcriptional regulation and post-transcriptional regulation.

ATRA decreases the expression of Six1, which decreases the transcriptional activity of GADD34 with TLE3 in myoblasts. ATRA also increases GADD34 mRNA stability via p38 phosphorylation, which leads to the blocking of interaction between TTP and ARE on GADD34 mRNA. Increased GADD34 proteins change the type of myosin heavy chain in myotubes.
Figure 1.

A) Relative mRNA levels of various genes in different tissues:
- DMSO
- ATRA

B) Western blot analysis of GADD34 protein levels in DMSO and ATRA-treated HEK293 cells.

C) Schematic diagram of myoblast differentiation:
- Seeding
- 1 day (10% FBS)
- 2 days (10% FBS)
- 4 days (2% HS)
- Myotubes

D) Relative mRNA levels of ATF4 and GADD34 in myoblasts, myotubes, HEK293 cells.

E) ATRA (logM) treatment effects on GADD34 expression in HEK293 cells.

F) Relative GADD34 mRNA levels upon ATRA and ATF4 siRNA treatment.
Figure 2.

A

human GADD34 promoter
-474 | +7  human GADD34
5' | 3'

-474 | +7  pGADD34-0.5k

C2C12  HEK293

Relative luciferase activity

empty RARα RARβ RARγ empty RARα RARβ RARγ

RXRα

B

C2C12  HEK293

Relative luciferase activity

ATRA (logM)  NT -8 -7 -6 -5

C

-474 +7  Luc pGADD34-0.5k

-329 +7  Luc pGADD34-0.3k

-162 +7  Luc pGADD34-0.16k

-131 +7  Luc pGADD34-0.13k

Relative luciferase activity

0  0.5  1  1.5  2  2.5

NT  ATRA
Figure 3.

A

-162 AGGGCGTGGTCACGCTCGGAATCTCCG -131

Pax3, Sox6

B

-162 +7 Luc pGADD34-0.16k

-131 +7 Luc pGADD34-0.13k

Relative luciferase activity

0 0.2 0.4 0.6 0.8 1 1.2

C

1.2

1

0.8

0.6

0.4

0.2

0

1

Six1

GADD34

GAPDH

D

Six1 - +

(kDa)

31.3

72.8

31.3

E

-162 MEF3-1 MEF3-2 MEF3-3 -131

WT: AGGGCGTGGTCACGCTCGGAATCTCCG

Mut-M1: AGGGACGTGGTCACGCTCGGAATCTCCG

Mut-M2: AGGGCCGAGGTGACGCTCGGAATCTCCG

Mut-M3: AGGGCGTGTCAGCTCGGAATCTCCG

F

1.4

1.2

1

0.8

0.6

0.4

0.2

0

WT Mut-M1 Mut-M2 Mut-M3

G

N.E. (empty) + - - -

N.E. (Six1) - + + +

Competitor - - cold M2 C/EBP

H

Probe GADD34-MEF3

N.E. (empty) + -

N.E. (Six1) - +
The gene expression of 6 TLE family genes in myoblast C2C12 cells

| Gene Name | (pg)/18S(ng) |
|-----------|--------------|
| TLE1      | 0.567        |
| TLE2      | U.D.         |
| TLE3      | 34.401       |
| TLE4      | U.D.         |
| TLE5      | 23.736       |
| TLE6      | U.D.         |

Relative mRNA levels

- siCont.
- siTLE1
- siTLE3
- siTLE5

Relative luciferase activity

- siCont.
- siTLE1
- siTLE3
- siTLE5

N.S.

* * *
Figure 6.

A. ATRA (logM) NT -8 -7 -6 (kDa)
   GADD34 72.8
   GAPDH 31.3

B. Relative mRNA levels
   2
   2.5
   1.5
   0.5
   1
   2
   NT
   ATRA (logM)

C. Human GADD34-3'UTR
   5' UTR CDS 3' ARE (AUUU)
   2295 2399
   5' Luc 2265 2350
   ARE-1 ARE-2
   Relative luciferase activity
   pGL3 basic empty GADD34-3'UTR

D. Relative luciferase activity
   2.5
   2
   1.5
   1
   0.5
   0
   ATRA - + +
   SB203580 - - +

E. Relative luciferase activity
   2.5
   2
   1.5
   1
   0.5
   0
   ATRA - + +
   siCont. - + -
   siTPP - - +

F. Relative luciferase activity
   3
   2.5
   2
   1.5
   1
   0.5
   0
   WT Mut-A1 Mut-A2
   DMSO ATRA

G. Input logA
   anti-TTP
Figure 7.

A (Transfection) Day 0 △ seeding △ myoblasts △ myotubes
Day 1 (10% FBS) Day 2 Day 4 (2% HS)

B Day 0 1 2 4
empty
GADD34

C Thickness of myotubes (µm)
empty
GADD34

D empty GADD34

E Day 0 1 2 4 (kDa)
GADD34 - - + - - + +
total MYHC MYHC1 (slow) MYHC2 (fast) MYHC2a MYHC2x MYHC2b GAPDH

F Day 4

| MYHC type | 1 | 2a | 2x | 2b |
|-----------|---|----|----|----|
| Relative mRNA levels | * | * | * | |
In myoblasts

All-trans retinoic acid: ATRA

GADD34 mRNA

Six1

TLE3

(transcriptional regulation)

TTP

p38

ARE 3’

GADD34 protein

MHC Type1 Type2

(transcriptional regulation)

(post-transcriptional regulation)