Review Article

Direct Control of Stem Cell Behavior Using Biomaterials and Genetic Factors

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Stem cells have recently emerged as an important candidate for cell therapy. However, some major limitations still exist such as a small quantity of cell supply, senescence, and insufficient differentiation efficiency. Therefore, there is an unmet need to control stem cell behavior for better clinical performance. Since native microenvironment factors including stem cell niche, genetic factors, and growth factors direct stem cell fate cooperatively, user-specific in vitro settings are required to understand the regulatory roles and effects of each factor, thereby applying the factors for improved cell therapy. Among others, various types of biomaterials and transfection method have been employed as key tools for development of the in vitro settings. This review focuses on the current strategies to improve stemness maintenance, direct differentiation, and reprogramming using biomaterials and genetic factors without any aids from additional biochemicals and growth factors.

1. Introduction

Stem cell therapy possesses significant advantages compared to conventional cell therapy using mature cells, as stem cells are more accessible and obtainable, are easy to culture and expand, and enable avoiding graft-versus-host rejection [1–3]. With such merits, stem cells have emerged as a candidate for cell therapy since 1968 when bone marrow transplantation surgery was conducted. Stem cells can self-renew and further differentiate into specific lineages upon stimulation. Among many kinds of stem cells, adult stem cells, represented by mesenchymal stem cells (MSCs), can be isolated or derived from many kinds of tissues and thus possess similar but different properties from each other. In a native microenvironment, MSCs are surrounded by stem cell niches composed of extracellular matrix (ECM) and growth factors. These microenvironment factors play instructive roles in directing stem cell behavior such as growth, lineage commitment, and stemness maintenance.

For clinical applications, stem cells have to be expanded because only a limited number of cells can be extracted from a tissue source. Moreover, when stem cells are expanded in a series of exhausted in vitro culture, the efficacy of their proliferation and differentiation decreases due to a progressive loss of stemness driven by senescence. To overcome such problems, state-of-the-art technologies using biomaterials, genetic factors, and growth factors which can mimic a native microenvironment or improve stem cell behavior have been employed recently. In conventional studies, various types of biomaterials and transfection method have been employed as key tools for development of the in vitro settings. This review focuses on the current strategies to improve stemness maintenance, direct differentiation, and reprogramming using biomaterials and genetic factors without any aids from additional biochemicals and growth factors.
growth factors, such as bone morphogenetic proteins (BMPs) or transforming growth factor β (TGF-β), can promote MSCs to differentiate into osteoblast or chondrocyte in vitro and induce efficient bone formation and cartilage regeneration compared to no treatment control [7–10]. However, FGF2 treatment is not able to overcome cellular senescence and the loss of differentiation potential of MSCs [11]. Moreover, because of the short half-life of growth factors, a large amount of growth factors is required to achieve the goal, resulting in high cost. Also, direct injection of growth factors may cause serious side effects such as osteophyte formation, swelling, and synovial hyperplasia [9]. Because of such disadvantages of growth factor treatment, applying biomaterials (e.g., natural, synthetic), biophysical factors (e.g., ultrasound), or biochemical factors (e.g., gene transfection) have emerged as alternative encouraging strategies to control stem cell fate.

Here, we review the current strategies to control stem cell fate using biomaterials, physiochemical factors, and genetic factors (Figure 1) in the absence of growth factor treatment. We first reviewed the strategies for stemness maintenance of adult stem cells using physiochemical factors (Table 1) and biomaterials (Table 2). Next, we introduced various types of biomaterials which can help adult stem cells to induce differentiation into specific lineages (Table 3). Finally, we reviewed genetic reprogramming methods for induced pluripotent stem cells (iPSCs) (Tables 4 and 5).

2. Improvement of MSC Stemness Using Biophysical Stimulation, Organic Compounds, and Biomaterials

Adult stem cells, represented by MSCs, are considered as an attractive agent for cell therapy because of their ability to self-renew and differentiate into various tissue cell types [12, 13]. However, the cell number when isolated is not usually sufficient for clinics. Therefore, a series of in vitro expansion of stem cell is indispensable. As MSCs lose their self-renewing ability and differentiation capacity during subculturing, maintenance of stemness has become an essential requirement for a successful stem cell therapy [14, 15]. Here, we review biophysical stimulation (Table 1), organic compound treatment (Table 1), and biomaterials (Table 2) as major methodological factors to maintain mature and homogeneous differentiation of stem cells [16, 17].

2.1. Biophysical Stimulation. Biophysical stimuli are one of important factors to enhance the differentiation capability of MSCs, for example, when a normal human cartilage was continuously exposed to physical pressure, such as joint loading. This stimulus went through cell membranes, thereby playing a pivotal role in structural maturation of cartilage. As another example, when MSCs were subjected to low-intensity pulsed ultrasound (LIPUS) stimuli in vitro, the cells differentiated into chondrocytes. Furthermore, when chondrogenic differentiation was induced in alginate, LIPUS-stimulated MSCs were not dedifferentiated even though the culture environment was not suitable for chondrogenic differentiation [18]. Along the same line, when MSCs were transplanted with PGA scaffold in a defect site post-LIPUS exposure for a week, the tissue morphology was maintained like an intact cartilage [19]. Another study demonstrated that when MSCs were stimulated with ultrasound, osteogenic differentiation was reduced compared to control MSCs [20, 21]. This result suggests that biophysical stimulation has significant effects on MSCs to keep the undifferentiated or differentiated status.

2.2. Biochemical Stimulation. MSCs express important pluripotent factors including Oct4, Sox2, Nanog, and cMyc, and these factors have been widely studied. However, expression of these factors reduces when MSCs undergo cell senescence during a series of subculture [22–24]. To address this issue, overexpression of pluripotent factors through lentiviral transfection was studied, thereby enhancing the self-renewal and differentiation potential of MSCs [25]. In addition to pluripotent factors, the telomere activity was found to reduce
Table 1: Maintenance of stemness using biophysical and biochemical stimulations.

| Type of stimulation                  | Details of condition | Type of cells | Observation                                                                 | Ref. |
|--------------------------------------|----------------------|---------------|------------------------------------------------------------------------------|------|
| Biophysical stimulation              | Low-intensity pulsed ultrasound (LIPUS) | hMSCs         | hMSCs differentiated into chondrocyte without dedifferentiation in nonchondrogenic differentiation environments. | [18] |
|                                      | LIPUS                | hMSCs         | The transplanted cells differentiated into chondrocytes and regenerated defect sites of recipient cartilage. | [19] |
|                                      | Ultrasound           | hMSCs         | Ultrasound treatment enhanced fracture healing by promoting osteogenic differentiation of hMSCs. | [20] |
|                                      | Fluid flow           | Osteocyte, osteoblast, and hMSCs | Flow stimulation promoted recruitment, proliferation, and differentiation of osteoprogenitor cells. | [21] |
| Overexpression of genetical factor    | SRY- (sex-determining region Y-) box 2 (SOX2) Sirtuin 1 (SIRT1) | hMSCs         | Overexpression of Sox2 enhanced stemness of MSCs during in vitro cultivation. | [23] |
|                                      | Octamer-binding transcription factor 4 (Oct4) or pron. nanOg (Nanog) | hMSC          | Overexpression of SirT1 prevented age-associated senescence of MSCs via Sox2 regulation. | [26, 27] |
|                                      |                      |               | Viral transfection of Oct4 or Nanog enhanced the self-renewal and differentiation potential of MSCs. | [24, 25] |
| Treatment of organic compound        | Resveratrol          | hMSCs         | Resveratrol treatment enhanced maintenance of the self-renewal and differentiation capacity of MSCs during ex vivo cultivation. | [28] |
|                                      | Nuclear factor erythroid-derived 2-like 2 (NRF2) | hMSCs         | Treatment of t-BHQ, the activator of NRF2, promoted self-renewal ability and osteogenic differentiation via inhibition of p53 expression. | [35] |

During in vitro cultivation. Thus, sirtuin 1 (SIRT1: a class III histone deacetylase protein) was treated to induce expression of telomerase reverse transcriptase (TERT) [26]. Sirt1 is also known as an important factor which regulates the lifespan, aging, metabolic homeostasis, and age-associated senescence of MSCs by controlling Sox2 acetylation [27].

In order to develop a better strategy to reduce cell senescence or to improve stemness, organic compounds are treated to MSCs to prevent the decrease of pluripotent marker expression. For example, since resveratrol is an antioxidant as well as Sirt1 activator, its treatment improved the stability of Sox2 by preventing acetylation and degradation of Sox2 [27]. Moreover, sustained treatment of resveratrol during ex vivo expansion maintained self-renewal and differentiation capacities from an early passage until a late passage [28]. These results suggest that treating stem cells with antioxidants can be a reliable option to maintain MSC stemness during subculture.

Another major cause of cell senescence is intracellular accumulation of reactive oxygen species (ROS) [29] ("oxidative stress") which results in aging with end point apoptosis of MSCs [30, 31]. Nuclear factor (erythroid-derived 2)-like 2 (NRF2) plays a vital role in defending against oxidative stress at the cellular level. Therefore, conservation of NRF2 nuclear localization is important to overcome MSC aging during subculture [32, 33]. A previous study reported that treatment of t-BHQ, an antioxidant, increased translocation of NRF2 into the nucleus and prevented cellular senescence by regulation of the p53-Sirt1 axis, as p53 can suppress the transcriptional activity of Sirt1 by binding to the Sirt1 promoter [34]. Due to such changes in cellular behaviors under t-BHQ treatment, aged cells elevated the abilities for self-renewal and osteogenic differentiation [35]. Together, the results suggest that antioxidant treatment is a promising approach to reduce cell senescence especially in long-term culture for a successful MSC therapy in vivo.

2.3. Biomaterials. The extracellular matrix (ECM) controls stem cell fate (e.g., proliferation and differentiation) through integrin-receptor binding [36, 37]. Therefore, a series of biomaterials have been employed due to user-defined tunability of cell-matrix interaction (e.g., cell adhesion and cytoskeletal tension) as an artificial matrix platform. In this part, we introduce major types of biomaterials which have been used to maintain or to enhance stemness of adult stem cells, especially MSCs.

Biomaterials can be categorized into natural or synthetic materials in general. Among natural biomaterials, decellularized ECMs have been studied to control stem cell behavior recently. For example, the ECM where naive human MSCs (hMSCs) resided was decellularized and used for in vitro culture. This culture substrate was found to maintain stemness of human or mouse-derived adult stem cells most likely because it provided an in vivo-like stem cell niche [38–41]. Also, decellularized tendon tissue was found to maintain stemness and thereby promoted tenocyte differentiation of human tendon stem cells (hTSCs) by providing an amiable niche [42].

In addition to natural biomaterials, synthetic biomaterials have been recently designed to maintain or enhance stemness. For example, encapsulation of MSCs into hydrogels can mimic the three-dimensional microenvironment of...
native tissues. Polyacrylamide, alginate/GelMA, or pullulan-collagen hydrogels with low stiffness maintained stemness because they helped with the maintenance of low cytoskeletal tension [43–45]. Also, decreasing the cell matrix-binding affinity by reducing the Arg-Gly-Asp (RGD) density is revealed to enhance stemness in poly(carboxybetaine) hydrogels [46].

Besides, surface topography is a common method to control cell behavior related to stemness. While MSCs were cultured on nanopatterned poly(ε-caprolactone) (PCL) substrates with 120 nm pits in a square arrangement with a centre-centre spacing of 300 nm, stemness was enhanced [47]. On the other hand, nanotopography with an aligned shape (polydimethylsiloxane, 250 nm in depth, 350 nm in width, and with 700 nm pitch) did not enhance MSC stemness compared to a nontopographic surface [48].

Polymeric surface coating serves as another promising option to control stemness. Poly-L-lysine (PLL) is a widely used coating to improve cell adhesion and proliferation. PLL-coated surface improved proliferation but retarded the replicative senescence of hBMSCs by increasing the S-phase [49]. PCL nanofiber matrices increased the expression of self-renewal factors and cell-cell interaction markers in hMSCs. PEG-PCL copolymer exhibited moderate surface repellency and induced aggregation of hMSCs, which promoted stemness and lowered intracellular ROS accumulation [51]. Emu oil-loaded nanofibers with higher tensile strength enhanced the expression of stemness, proliferation, and cell adhesion markers in hASCs compared to unloaded nanofibers. The 3D culture of hMSCs in a nanostructured electrospun gelatin patch maintained stemness of hMSCs for 3 weeks [54].

Chitosan film induced spheroid formation of hASCs with higher activities of self-renewal and colony formation, as well as significant upregulation of pluripotency marker expression [56]. The chitosan film promoted spheroid formation of hUCBMSC under hypoxia than normoxia. HIF-1 additionally induced expression of stemness genes [57].

| Type of biomaterials | Details of materials | Type of cells | Observation | Ref. |
|----------------------|----------------------|---------------|-------------|-----|
| Natural (nonsynthetic) | Decellularized ECM of undifferentiated hMSCs | hMSCs mASCs | Decellularized ECM of undifferentiated MSCs promoted self-renewal, colony formation, and stemness maintenance of hMSCs. | [38–41] |
| | Decellularized tendon tissue | hTSCs | Decellularized tendon tissues enhanced self-renewal and stemness maintenance of hTSCs. | [42] |
| Hydrogel | Polyacrylamide gels and PDMS stamps | hMSCs | Low cytoskeletal tension was maintained by controlling substrate stiffness as cell spreading was restricted, thereby enhancing stemness. Polyacrylamide gels and PDMS stamps were used to regulate biophysical parameters. | [43] |
| | Alginate/GelMA hydrogels | hBMSCs and GMSCs | Compared to alginate hydrogels, alginate/GelMA hydrogels maintained stemness due to decreased hydrogel stiffness. | [44] |
| | Pullulan-collagen hydrogel | mBMSCs | Biomimetic hydrogel maintained stemness of mouse bone marrow-derived MSCs (mBMSCs) compared to tissue plate culture, resulting in enhanced viability after in vivo injection. | [45] |
| | RGD-modified poly(carboxybetaine) hydrogel | hBMSCs | hMSCs formed 3D spheroids on the 5 μM RGD substrate, and the stemness was well maintained compared to 5 mM RGD substrate, which enhanced osteogenic differentiation. | [46] |
| Topography | PLL-coated surface | hBMSCs | PLL-coated surface improved proliferation but retarded the replicative senescence of hBMSCs by increasing the S-phase. | [49] |
| | PCL nanofiber | hMSCs | PLL substrates increased the total number of hMSCs while stemness was maintained. | [50] |
| | PEG-PCL copolymer | hMSCs | Bone marrow collagen-mimetic PCL nanofiber matrices increased the expression of self-renewal and cell-cell interaction markers in hMSCs. | [51] |
| | Emu oil-loaded PCL/Coll nanofiber | hASCs | Emu oil-loaded nanofibers with higher tensile strength enhanced the expression of stemness, proliferation, and cell adhesion markers in hASCs compared to unloaded nanofibers. | [53] |
| Polymers | Gelatin nanofiber | hMSCs | 3D culture of hMSCs in a nanostructured electrospun gelatin patch maintained stemness of hMSCs for 3 weeks. | [54] |
| | Chitosan film | hASCs | The chitosan film induced spheroid formation of hASCs with higher activities of self-renewal and colony formation, as well as significant upregulation of pluripotency marker expression. | [56] |
| | Chitosan film + hypoxia | hUCBMSC | The chitosan film promoted spheroid formation of hUCBMSC under hypoxia than normoxia. HIF-1 additionally induced expression of stemness genes. | [57] |
### Table 3: Direct differentiation using biomaterials.

| Property            | Type of materials | Differentiation | Details of materials | Comments                                                                                                                                                                                                 | Ref. |
|---------------------|-------------------|-----------------|----------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
|                     |                   |                 |                      |                                                                                                                                                                                                          |      |
| Chondrogenesis      | Cellulose/silk blend | Chondrogenesis | Cellulose/silk blend | Growing MSCs on a specific blend combination of cellulose and silk in a 75:25 ratio significantly upregulated expression of chondrogenic markers.                                                      | [120]|
|                     |                   |                 |                      |                                                                                                                                                                                                          |      |
| Scaffold            |                   | Chondrogenesis  | ECM-like porous scaffold of poly(3-hydroxybutyric acid-co-3-hydroxyvaleric acid) (PHBHV)/gelatin blends | PHBHV/gelatin constructs mimicking myocardial structural properties.                                                                                                                                     | [121]|
|                     |                   | Myogenesis      | Collagen-glycosaminoglycan | Collagen-chondroitin sulphate (CCS) scaffolds enhanced osteogenesis while collagen-hyaluronic acid (CHyA) scaffolds enhanced chondrogenesis.                                                             | [122]|
|                     |                   | Cardiomyogenesis | Carbon nanotube/poly-L-lactide acid (PLA) nanofiber | The two-pronged carbon nanotube template provided a biomimetic electroactive cue, thereby directing MSC differentiation.                                                                               | [13] |
| Composition         |                   | Chondrogenesis  | Cartilage extracellular matrix-derived particles (CEDPs) | Microtissue aggregates (BMSCs and CEDPs (263 ± 48 μm) cocultured in a rotary cell culture system) showed a more rapid restoration of joint functions with superior cartilage repair compared to the control groups in vivo. | [3]  |
| Decellularized      |                   | Chondrogenesis  | Calcium phosphate nanoparticles and demineralized bone matrix (DBM) particles incorporated into injectable polyHIPE | PolyHIPE compositions with BMSCs promoted osteogenic differentiation through upregulation of bone-specific marker expression compared to a time zero control.                                           | [4]  |
| tissues             |                   | Osteogenesis    | 3D graphene foams (GFs) | 3D GF culture platforms maintained stem cell viability and promoted osteogenic differentiation.                                                                                                        | [123]|
| Bioinorganics       | Osteogenesis      | Polyacrylate substrate functionalized with RGD peptide | Biomimetic polyacrylate substrates can direct chondrogenic differentiation of mMSCs, hMSCs, and mouse KSCs in the absence of exogenous TGF-bs.                                                   | [124]|
| Biomimetics         | Chondrogenesis    | Osteogenesis/ | Polyacrylamide (0.5–40 kPa) hydrogel substrate | MSCs on soft (~0.5 kPa) gels promoted expression of neurogenesis markers while MSCs on stiff (~40 kPa) substrates elevated expression of osteogenesis markers. Transfer of MSCs from soft to stiff or stiff to soft substrates led to a switch in the lineage specification. Both chondrogenic and osteogenic markers were elevated when MSCs were grown on substrates with stiffness < 10 MPa. MSCs on lower stiffness gels express elevated chondrogenic differentiation markers while MSCs on the higher stiff substrates express elevated osteogenesis markers. GHPA as a promising soluble factor-free cell delivery template induced endothelial differentiation of MSCs with robust neovascularization formation with favorable host responses. | [60] |
| Hydrogels           |                   | Osteogenesis/ | Methyl acrylate/methyl methacrylate (18–72 MPa) hydrogel substrate |                                                                                                                                                                                                          | [61] |
|                     |                   | Chondrogenesis  | Gelatin hydrogel conjugating enzymatically cross linkable hydroxyphenyl propionic acid (GHPA) |                                                                                                                                                                                                          | [63] |
used polymer as a surface coating material, because it improves proliferation but retards replicative senescence by increasing the S-phase population of MSCs [49] and hematopoietic stem cells (HSCs) [50] in the cell cycle. Also, coating the culture substrate with PCL nanofibers which can mimic the size and shape of collagen fibers of ECM is also known to maintain stemness [51]. As another approach, poly(ethylene glycol)-poly(ε-caprolactone) (PEG-PCL) copolymers were used to control surface repellency by altering the molar percentage or chain length of PEG. This surface repellency induced aggregation of hMSCs by upregulation of cell-cell interaction proteins such as connexin-43, which further increased stemness with a significant decrease in intracellular ROS accumulation (Figure 2) [52].

Furthermore, nanofibrous scaffolds provided a 3D microenvironment to stem cells and thus enhanced stemness. For example, ASCs displayed an improved adhesion capacity with high rates of bioactivity and proliferation when cultured on emu oil-loaded nanofibers [53]. Mesenchymal stem cells (MSCs) showed a superior differentiation capacity towards typical mesenchymal lineages when grown in a nanostructured electrospun gelatin patch [54]. Especially, emu oil exhibited a free radical scavenging activity, thereby enhancing stemness [55].

Finally, a chitosan film induced spheroid formation and triggered a cell-cell interaction of hASCs, thereby enhancing stemness. After spheroid formation, the spheroid-forming hASCs expanded efficiently, formed a colony, and upregulated the expression of pluripotency marker genes compared to the monolayer-cultured control condition [56, 57].

In conclusion, the aforementioned types and formats of biomaterials were found to enhance or maintain stemness. We summarize the three major mechanisms by which the biomaterials were found to enhance or maintain stemness as follows: (1) reduction of cytoskeletal tension by reducing matrix stiffness, (2) spheroid formation by reduction of integrin-binding sites and consequent promotion of cell-cell interaction, and (3) antioxidative effects by radical scavenging activity. These strategies have potential to effectively improve stemness of MSCs in various biomaterial formats.

### 3. Direct Differentiation of MSCs Using Biomaterials

While most methods known to induce mesenchymal lineage differentiation of MSC depend on exposure to one or more soluble growth factors, a growing body of evidence suggests that it is possible to control MSC differentiation in the absence of soluble factors. MSCs exhibit the ability to differentiate towards specific lineages through biomaterials with modification of mechanical or biochemical properties, matrix composition, topography, and surface stiffness. This
approach would simplify the tissue engineering procedure and be cost-effective. Here, we summarize recent studies that employed such approaches to induce direct differentiation of MSC via biomaterial technologies (Table 3).

3.1. Composition. Collagen and glycosaminoglycan (GAG), major components of natural ECM, play a key role in osteochondral regeneration. Hence, their combined (CG) scaffolds have been used successfully in tissue engineering applications for regeneration of cartilage and bone [57, 58]. A previous study reported the effect of the composition and stiffness of collagen and GAG scaffolds composed of chondroitin sulphate (CS) and hyaluronic acid (HA) on MSC differentiation [59]. The study showed that the lowest stiffness (0.5 kPa) of the CG scaffold facilitated chondrogenesis, while the stiffest (1.5 kPa) scaffold induced osteogenesis. This was the first evidence proving that osteochondral differentiation of MSC could be directed via scaffold composition using CG and further enhanced by the GAG type. When cellulose, another abundant natural polymer, was blended with silk at different compositions, growth and chondrogenesis of MSC were

| Type | Advantages | Disadvantages | Transgene expression | Efficiency | Ref. |
|------|------------|---------------|-----------------------|------------|------|
| Virus | Nonintegrative; infects dividing and nondividing cells | Low efficiency | No | 0.0001–0.01% | [84, 85] |
| Adenovirus | Ease of handling with experience; medium–high efficacy | Integration of foreign DNA into genome; residual expression of reprogramming factors; controversy regarding tumor formation | Yes | 0.1–1% | [73, 125] |
| Lenti/retrovirus | Medium–high efficiency; nonintegrating; robust protein-expressing property; wide host range | Involve viral transduction | No | 0.5–1.0% | [88, 89] |
| Sendai virus | Very low efficiency; the use of potent viral oncoprotein (SV40LT antigen) | Very low efficiency | No | 0.01% | [127] |
| Episomal | More persistent transgene expression; lack bacterial origin | Requires high gene dosages and multiple transfections; daily transfection; controversy in reproducibility and mitigating cost effectiveness | No | 1.4–2% | [128, 129] |
| Minicircle | Relative high efficiency; nonintegration; easily automated, making it an exciting candidate for routine biomanufacture. | Inefficient excision, potential for genomic toxicity | Excision with transposase | 0.1–1% | [80] |
| miRNA | More persistent transgene expression; lack bacterial origin | Very low efficiency | No | 3–6 × 10 − 6 | [87, 126] |
| PiggyBac transposons | Elimination of insertional mutagenesis; no footprint upon excision; higher genome integration efficiency | Slow kinetics, low efficiency; difficulties in generation and purification of reprogramming protein | No | 0.005–0.001% | [130] |
| Protein | Free of genetic materials; direct delivery of reprogramming factor proteins | Inefficient excision, potential for genomic toxicity | Excision with transposase | 0.1–1% | [80] |
| Small molecules | Ease of handling; no requirements for reprogramming factors | More than one target, toxicity | No | 0.3–0.5% | [86] |

| Donor cell type | Ref. |
|-----------------|------|
| Adipose-derived stem cells | [131] |
| Amniotic fluid | [132] |
| Blood cell cord blood stem cells | [104] |
| B lymphocytes | [133] |
| Bone marrow cells | [134] |
| Cardiac myocytes | [135] |
| Dental pulp | [136] |
| Dermal fibroblasts | [137] |
| Endometrial stromal fibroblasts | [100] |
| Hematopoietic progenitor cells | [138] |
| Hepatocytes | [139] |
| Keratinocytes (from hair pluck) | [101] |
| Pancreatic β-cells | [140] |
| Peripheral blood mononuclear cell | [126] |
This was also the first report demonstrating the potential of cellulose to induce chondrogenic differentiation of MSC.

Recent studies investigated whether chondrogenic differentiation of MSC could be directed by biomimetic or decellularized tissue-derived materials. Biomimetic polyacrylate substrates functionalized with the RGD integrin-binding peptide promoted chondrogenesis of MSC in the absence of any soluble growth factors [61]. They suggested that the amount of surface amine residues from the RGD peptide was a key regulator to inducing differentiation. Several studies demonstrated that ECM components derived from the cartilage promoted chondrogenesis of MSC [62–64]. In all cases, however, the use of chondrogenic growth factors was found to be essential for MSC chondrogenesis with deposition of necessary matrix components. In addition, a study reported that novel particles derived from natural cartilage ECM induced chondrogenic differentiation of MSC even in the absence of TGF-β when the particles were used as a cell carrier [65].

Calcium phosphate- (CaP-) based ceramics play a significant role in bone repair due to their osteoconductive properties.
potential for filling lost bone volumes, when CaP nanoparticles and demineralized bone matrix (DBM) were fabricated as an injectable bone graft by incorporating polymerized high internal phase emulsions (polyHIPEs). This injectable bone graft was found to induce direct osteogenesis of MSC [58]. On the other hand, our recent study investigated the potential of 3D graphene substrates to induce spontaneous osteogenesis of MSC without additional stimuli [59]. These reports revealed that material-derived cues were able to guide MSC differentiation to osteogenesis in the absence of extrinsic biochemical inputs.

Proper regeneration of the myocardium is dependent on scaffold properties and thus can be enhanced by mimicking features of the myocardial ECM. A three-hydroxybutyrate and 3-hydroxyvalerate (PHBV)/gelatin construct mimicking the myocardial ECM structure was developed to promote cardiac differentiation of MSC and cardiac resident cells without any chemical stimulation [66]. This study demonstrated that when specific physicochemical properties with a microtopograph were produced to mimic the structural and mechanical properties of myocardium in the PHBV/gelatin construct, myogenesis of the stem cells was promoted. Electroconductive carbon nanotubes (CNT) demonstrated an ability to induce myogenic differentiation of MSC in the absence of additional stimuli [67]. Although the exact mechanism of this result is unclear, it was suggested that electrical stimulation of MSC by culturing on the CNT-based polylactic acid scaffold was a key factor to enhancing differentiation to cardiomyocytes.

3.2. Substrate Stiffness. Among the biophysical cues that were identified to regulate cell fate in static in vitro cultures, stiffness of culture substrates was suggested first as a key property in several important studies [60–64]. An increased stiffness of the culture substrate was found to induce osteogenic differentiation of MSCs [60, 61]. The role of matrix stiffness in directing lineage specification of MSCs was examined on the surface-charged methyl acrylate/methyl methacrylate

Figure 3: Gelation of GHPA by H₂O₂ and horseradish peroxidase-catalyzed cross linking. In vitro endothelial differentiation of hMSCs in GHPA hydrogels. Figure 3 is reproduced with permission from [63], John Wiley and Sons. * indicates p < 0.05 in comparison to the control MSCs on tissue culture plate.
(MA/MMA) polymer substrate with varying elastic modulus [60]. This study demonstrated that the substrate group with lower stiffness induced chondrogenesis of MSCs while the substrate with rigid stiffness induced osteogenic specification of MSCs. Although its specific mechanism is unclear, this study revealed that integrin β1 played a critical role in this process. Cells sense their mechanical microenvironment via integrin-ligand interactions which form focal adhesions and thereby regulate intracellular signaling [65]. Another study supported this finding [61] by showing that the soft (~0.5 kPa) substrate was effective in promoting neurogenesis of MSCs whereas the stiff (~40 kPa) one was effective in promoting their osteogenesis. Switching the biophysical microenvironment of MSCs from soft to stiff or stiff to soft substrates led to rewiring the two directions of MSC lineage specification.

Figure 4: Pluripotency reprogramming of human endometrial cells (hEMC). hEMC-derived iPS (heiPS) showed higher expression of pluripotent markers compared to neonatal fibroblasts. Figure 4 is reproduced with permission from [100], Oxford University Press.
Although the differentiation potential of MSCs into endothelial cells (ECs) remains unclear [68, 69], some studies reported possible approaches to differentiate MSCs into ECs [63, 64]. A previous study demonstrated that a 3D matrix with tunable properties directed the differentiation of MSC towards vascular cell types [64]. We also applied in situ cross-linkable gelatin hydrogels by conjugating enzymatically cross-linkable hydroxyphenyl propionic acid (GHPA) (Figure 3) [63]. The 3D culture of MSCs in these hydrogels induced vasculogenesis both in vitro and in vivo. Our results showed that GHPA hydrogels induced spontaneous endothelial differentiation of MSC without any soluble factors.

3.3. Surface Topography. When cells are cultured on biomaterial substrates, surface topography is known as a key regulator of cell behavior. Several previous studies reported that surface topographical cues induced direct lineage specification of MSCs [66, 67, 70–72]. Microfeatures (40 μm line, 20 μm spacing, and 1 μm height) of fibronectin strips printed on a poly(lactic-co-glycolic acid) (PLGA) thin film were found to direct lineage commitment of MSCs [66]. In this study, modification of MSC morphology and cytoskeletal arrangement on the patterned film resulted in both neuronal and myogenic lineages, even if myogenic differentiation was dominant when the expression of functional proteins was examined. Along the same line to direct MSC differentiation, Lee et al. fabricated pseudo-3D microwells by templating a hydrazine-immobilized polyacrylamide gel displaying inverse features of circular surface topography via PDMS stamps (circular) [67]. As a result, small circular islands induced an adipogenic phenotype of MSCs while anisotropic geometries induced neurogenic differentiation. Micro- and nanostructured titanium surfaces were used as potential topographical cues to induce osteogenic differentiation of MSCs [71, 72]. The nanotube-shaped titanium oxide surface structures, which have 70 to 100 nm titanium oxide nanotube arrays on them, induced cytoskeletal stress and thereby directed osteogenic differentiation of MSCs [71]. It was also reported that osteoblastic differentiation of MSCs was induced on the microstructured titanium surface (Ra = 3.22 μm) through α2β1 integrin-mediated interactions with cocultured osteoblasts [72]. Our recent study determined causative roles of topographical cues in directing lineage specification of MSC via patterned graphene surfaces with additional evaluation of electrical stimulation as another cue [70]. Our result showed that expression of osteoprogenitor markers was increased by either (un)patterned graphene substrate or electrical stimulation while the expression of osteoblast markers was increased only when electrical stimulation was applied together with the surface patterns.

4. Selection of Genetic Factor and Source Cell Type for iPSC Reprogramming

Induced pluripotent stem cells (iPSCs) were introduced in 2006, which opened a new avenue for stem cell research and regenerative medicine [73]. Obtaining an adequate amount of stem cells is a major limitation for stem cell therapy and research. Previously, classical methods employed to induce pluripotency of somatic cells include somatic cell nuclear transfer (SCNT) and cell fusion. However, limitations associated with oocyte supply, low reprogramming efficiency, and phenotypic abnormalities of the produced animal offspring still hamper the widespread distribution of these classical methods [74].

Resident stem cells in various tissues were also studied as a promising source of stem cells, but the lack of appropriate markers to define their phenotypes and their low differentiation potential were considered as major hurdles for using these cell sources. Hence, the breakthrough idea of reprogramming somatic cells with ectopic pluripotent markers (Sox2, Oct3/4, Klf4, c-Myc, and Lin28) to eventually represent embryonic stem cell (ESC) characteristics was undisputedly attractive [73]. However, it is still controversial whether iPSCs possess the same pluripotency and differentiation ability to ESCs. Donor cell-specific epigenetic signatures remain even after reprogramming and thus generate problematic variations from the expected quality and characteristics of iPSCs in terms of homogeneity and the potential for maturation in stem cell therapy [75, 76]. Moreover, abnormalities created during the process of iPSC reprogramming, such as establishing aberrant DNA methylation patterns, were found to increase the heterogeneity in iPSCs [77, 78]. Additionally, donor-specific genetic variations further increase the heterogeneity of iPSC genetic profiling, such as establishing aberrant DNA methylation patterns.

4.1. Choice of Vectors for iPSC Reprogramming

Substantial progress has been made in the methodologies to improve the efficiency and efficacy of reprogramming somatic cells to iPSCs in the past decade (Table 4). The type of vectors used to overexpress ectopic pluripotency factors within the target cells are classified into integrating DNA vectors and nonintegrating DNA free vectors. Integrating vectors are further subclassified into insertional vectors including viral and linear DNA delivery systems whereas insertion-free transgene vectors include PiggyBac transposon [79, 80] and plasmid/episomal vectors. Recently, nonintegrating systems involving direct protein or microRNA vectors as well as various small molecules are used for reprogramming of somatic cells into iPSCs [81, 82]. Integrating DNA vectors represent an early generation tool for reprogramming and are still commonly used in experiments owing to their high efficiency. Combinations of retroviral or lentiviral Sox2, Oct4, Klf4, c-Myc, or Lin28 were most popularly used with or without the use of transgene selection markers. Especially, these first-generation viral vectors possess the potential for random insertional mutagenesis, but their undeniable high efficiency still renders them useful for a wide range of iPSC research. Such random insertional mutagenesis contributes to the unpredictability of iPSCs upon in vivo transplantation. Viral promoter-driven fluorescence and cre-LOX expression systems have been used to track and control ectopic gene expression but still generate insertional mutagenesis [83]. In order to overcome problems associated with mutagenesis resulting from ectopic gene insertion, adenoviral, episomal, or plasmid vectors were used as alternatives in the course of developing the next generation of reprogramming methods.
Donor somatic cell-specific transcriptional and epigenetic signatures significantly contribute to the heterogeneity of efficiency and efficacy in reprogramming and differentiation of iPSCs (Table 5). The most widely used cell source for reprogramming into iPSCs is dermal fibroblasts, most frequently harvested from neonates as well as adults [73]. Keratinocytes (a type of bone marrow cells), peripheral blood cells (a type of CD34+ peripheral blood mononuclear cells), amniotic fluid cells, cord blood stem cells, endometrial stromal fibroblasts, and dental pulp cells have been reported as reliable sources of somatic cells for reprogramming (Figure 4) [100–106]. Moreover, it has been reported, even in cells which were originated from the same donor but from different organs, that the tissue-specific epigenetic signatures affect the heterogeneity of efficiency and efficacy in reprogramming and the differentiation potential of iPSC lines. Such observations were prominent in the early passages when the reprogramming process is not yet complete [107–109].

If stemness is enhanced, the negative effect of tissue-specific epigenetic signature may be attenuated during the process of serial passaging while losing the epigenetic memory sequentially. Characteristic DNA methylation patterns are originated from the donor cells and thus can be tracked in specific iPSC clones. Consequently, limitations in stemness of iPSCs as opposed to the full pluripotency of embryonic stem cells are inevitable. Experimental techniques to overcome the gap between the epigenetic memory of the donor cell and the stemness of the derived iPSC lines have been described in previous studies [110, 111]. One strategy is to increase the iPSC passage number while another approach is to introduce chromatin-modifying substances, which diminishes the epigenetic memory and enhances stemness. The process of acquiring pluripotency may not be complete upon immediate silencing expression of exogenous pluripotency factors but may continue for several rounds of cell passaging. iPSCs exhibit considerable differences in their telomere length and the global pattern of transcription and DNA methylation [110, 112, 113]. On the other hand, transgenes are usually silenced in the process of reprogramming by de novo methylation. When this process is not fully accomplished, gaining the pluripotency of the reprogrammed cells primarily relies on the exogenously introduced factors. When the endogenous pluripotent genes are halted from being fully expressed, these colonies are defined as “partially reprogrammed.” Within such colonies, pluripotency is frequently not fully acquired even after the exogenous factors are eventually turned off [114, 115]. Conversely, when ectopic transgenes are not silenced and exposed to residual activities or reactivation of the viral transgenes in the iPSC cells, tumor formation occurs as demonstrated in chimera experiments [73]. Other potential causes of epigenetic differences have also been attributed to either aberrant or incomplete reprogramming or even by various cell culture conditions [77, 116–119].

5. Conclusion

Although a growing body of evidence suggests stem cells as a promising candidate for cell therapy in the position of
replacing somatic cells, the aforementioned issues regarding senescence and low differentiation efficiency must be addressed for successful clinical applications. In this review, we introduced state-of-the-art methods which are currently approached to improve efficiency and efficacy of stemness maintenance, direct differentiation, and iPSC reprogramming, with the minimal use of expensive and side-effect occurring growth factors. Biophysical stimulation, organic compound treatment, genetic transfection, and various types of biomaterials were employed to achieve the purposes. In particular, the effects of matrix stiffness, improving cell-cell interaction, and antioxidant treatment became a major part of interest. Additionally, biomaterials with specific composition, stiffness, and topography can serve as a promising toolbox to guide direct differentiation of stem cells. Finally, several combinations or individual uses of genetic factors to induce reprogramming of somatic cells were introduced as a means of generating iPSCs. Pros and cons of major reprogramming methods were discussed as well. Taken together, selection of biomaterials or other external factors needs to be customized for target-specific developments and application of stem cell therapy towards successful clinical applications.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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