Hyaluronic acid in viscous malignant mesothelioma pleural effusion

Hui Min Cheah
Deirdre Fitzgerald
Amber Louw
Edith Cowan University
Jenette Creaney
Y. C. Gary Lee
Edith Cowan University

Follow this and additional works at: https://ro.ecu.edu.au/ecuworkspost2013

Part of the Diseases Commons

10.1002/rcr2.694
Cheah, H. M., Fitzgerald, D., Louw, A., Creaney, J., & Lee, Y. C. G. (2021). Hyaluronic acid in viscous malignant mesothelioma pleural effusion. Respirology Case Reports, 9(1), article e00694. https://doi.org/10.1002/rcr2.694
This Journal Article is posted at Research Online.
https://ro.ecu.edu.au/ecuworkspost2013/9319
Hyaluronic acid in viscous malignant mesothelioma pleural effusion

Hui Min Cheah1,2, Deirdre Fitzgerald1,2,3, Amber Louw4,5, Jenette Creaney2,6 & Y. C. Gary Lee1,2,3,5

1Pleural Medicine Unit, Center for Respiratory Health, University of Western Australia, Perth, WA, Australia.
2Medical School, University of Western Australia, Perth, WA, Australia.
3Department of Respiratory Medicine, Sir Charles Gairdner Hospital, Perth, WA, Australia.
4PathWest Laboratory Medicine, QEII Medical Centre, Perth, WA, Australia.
5School of Medical and Health Sciences, Edith Cowan University, Joondalup, WA, Australia.
6National Centre for Asbestos Related Diseases, Perth, WA, Australia.

Keywords
Hyaluronic acid, hyaluronidase, malignant pleural effusion, mesothelioma, viscosity.

Correspondence
Y. C. Gary Lee, Pleural Medicine Unit, Center for Respiratory Health, University of Western Australia, Room 533, Harry Perkins Building, 6 Verdun Street, Nedlands, Perth, WA 6009, Australia. E-mail: gary.lee@uwa.edu.au

Introduction
Pleural effusions affect over 90% of patients with malignant pleural mesothelioma. The malignant pleural effusion (MPE) commonly appears serosanguinous or haemorrhagic, and can be evacuated using a range of devices, such as aspiration needles, chest tube, or indwelling pleural catheter (IPC). We report two cases of extraordinarily viscous MPEs in mesothelioma patients. Our ex vivo experiments attributed the viscosity to the high hyaluronic acid (HA) content within the fluid, which could be significantly reduced with hyaluronidase.

Case Report
Patient 1
A 79-year-old man presented with a left-sided pleural effusion which was noted as viscous on the initial therapeutic aspiration. Cytology confirmed malignant mesothelioma. Upon recurrence of the MPE, an IPC (Rocket, UK) was inserted but the thick fluid prohibited spontaneous drainage. He was referred to our centre for further management. The fluid was partially evacuated by connecting the IPC via a connector to a 60-mL syringe through which manual suction could be applied. The viscosity of the fluid prohibited usual laboratory biochemical analyses of the fluid. In an attempt to reduce fluid viscosity to clear the remaining fluid, two doses of 5 mg of deoxyribonuclease (DNase) was instilled over two days via the IPC, after which ~300 and ~200 mL of fluid were drained. The underlying lung re-expanded on chest X-ray (CXR) and talc slurry pleurodesis was administered via the IPC.

Unfortunately, the fluid recurred over the next four weeks, resulting in a large multiloculated effusion associated with breathlessness and fatigue (Fig. 1A), necessitating hospitalization. Drainage via the IPC remained difficult because of the fluid viscosity. Four doses of intrapleural tissue plasminogen activator (tPA) 2.5 mg and DNase
5 mg were administered twice daily with good effect. A fluid sample was collected after the first dose (Sample 1) for viscosity and cytological analyses (Tables 1, 2 and Fig. 2, respectively). A total of 2.8 L of fluid was drained with significant radiographic improvement (Fig. 1B). However, the benefit was short-lived and fluid re-accumulated within two weeks (Fig. 1C), during which time he received one cycle of treatment in an immunotherapy trial. A further attempt with tPA 2.5 mg/DNase 5 mg given in outpatient clinic produced only marginal benefit. With strong manual suction via the IPC, 380 mL of viscous fluid was aspirated and a further 350 mL was drained the next day. Nonetheless, a massive residual effusion remained.

The patient was readmitted two weeks later for ongoing difficulty with IPC drainage (<100 mL each attempt) (Fig. 1D). Sample 2 was collected for viscosity analyses (Table 1). During the hospitalization, he received intrapleural tPA 5 mg/DNase 5 mg and suction via the IPC with little effect (60 and 275 mL after the first two doses). A further attempt using tPA 10 mg and DNase 10 mg yielded 200 mL and minimal radiographic improvement. He was discharged and died seven months later from his mesothelioma.

Patient 2

A 79-year-old female presenting with a right pleural effusion underwent video-assisted thoracoscopic pleural biopsy (which showed mesothelioma) and talc poudrage. The fluid recurred after surgery necessitating a therapeutic...
aspirate which revealed extremely thick exudative pleural fluid. She underwent regular therapeutic aspiration in an outside hospital for her MPE over the next four years (during which time she received various systemic therapy) before she was referred to our centre for IPC insertion. The fluid was described as thick and “jelly-like” and failed to drain via the IPC. Manual suction via the IPC (Rocket) was required for complete evacuation of the fluid. Talc slurry was administered after lung re-expansion but the pleurodesis failed.

For the following 12 months, drainage was performed (roughly once weekly) via IPC. Sample 3 showed a typical specimen. Standard IPC suction bottles were ineffective to clear the thick fluid. Hence, the patient’s carers were taught to perform IPC drainage using manual suction.

After one year, IPC drainage failed despite the above-mentioned measures causing significant breathlessness necessitating a hospital admission. A trial of intrapleural tPA 2.5 mg/DNase 5 mg was successful and drained 1.5 L with significant radiographic clearance. Regular IPC drainage was re-established. Her mesothelioma continued to progress and she died 10 months later.

Fluid Viscosity and HA Analyses

Pleural fluid viscosity was determined as described by Simpson et al. [1]. HA concentrations were determined in the MPE supernatant using a commercially available HA binding assay (Corgenix, USA). To test the contribution of HA to the viscosity of the effusion, samples were incubated at room temperature for 10 min with hyaluronidase type V (0–100 U/mL) diluted in phosphate-buffered saline (both Sigma-Aldrich, USA). To model the effects of recombinant human DNase (rhDNase) treatment, the original pleural effusion sample was incubated in the laboratory with rhDNase (Pulmozyme®; Roche, USA) diluted in normal saline (Baxter, Australia) for 4 h before viscosity measurement.

We found that MPE from both patients had very high pleural fluid HA concentrations. Hyaluronidase, but not rhDNase, was effective in reducing the viscosity of the fluid in a dose-dependent manner (Tables 1, 2, Video S1).

Discussion

We described two mesothelioma patients with HA-rich and extremely viscous MPE, limiting fluid drainage. Our

| Table 1. Pleural fluid HA concentrations and viscosity levels (mPa s) of pleural fluid after incubation with hyaluronidase (U/mL). |
|-------------------------------------------------|
| HA concentration in pleural fluid | Fluid viscosity (mPa s) after treatment with different concentrations of hyaluronidase (U/mL) |
|-----------------------------------|---------------------------------|
| 2.66×10^6 ng/mL                  | Patient 1 (Sample 1)            |
| 5.39×10^6 ng/mL                  | Patient 1 (Sample 2)            |
| 2.31×10^6 ng/mL                  | Patient 2 (Sample 3)            |
| 0                                | 800.21                          |
| 1                                | 54.62                           |
| 10                               | 21.91                           |
| 100                              | 2.94                            |
| Too viscous                      | Patient 1 (Sample 2)            |
| Too viscous                      | Patient 2 (Sample 3)            |

The two samples from Patient 1 were taken about 38 days apart.

HA, hyaluronic acid.

| Table 2. Pleural fluid viscosity levels (mPa s) of pleural fluid after incubation with rhDNase. |
|-------------------------------------------------|
| rhDNase (μg/mL) | 0 | 2.5 | 5 | 10 |
|-------------------------------------------------|
| Patient 1 (Sample 1) | 780.56 | 690.93 | 627.61 | 567.38 |

Sample 1 was incubated with rhDNase at 2.5, 5, and 10 μg/mL at room temperature for 4 h before viscosity was measured. The range of concentration selected was to approximate the concentration of enzyme in a patient with a 1.5-L pleural effusion treated with 5 mg of rhDNase. rhDNase, recombinant human deoxyribonuclease.
laboratory work confirmed that HA, present in high concentrations in these fluids, likely contributed to the high viscosity which could be significantly reduced using hyaluronidase.

MPE from mesothelioma contains a vast number of important mediators. Mesothelioma has been known to express HA for over 80 years and HA pleural fluid concentrations are a valuable ancillary test for diagnosis. In our previous study including 96 mesothelioma patients, the pleural fluid HA level has an area under the curve of 0.89 in its diagnostic value of mesothelioma, highly comparable with mesothelin. Pleural fluid HA has also prognostic values [2]. Our experiments provide proof to the long suspicion that HA is involved in generating high viscosity in MPE.

HA is a long polymer of disaccharides up to 20,000 kDa in size produced by hyaluronan synthases, present in abundance in extracellular matrix. It can promote inflammation and angiogenesis—processes involved in MPE formation. HA is approved as intra-articular treatment for osteoarthritis and for use in ophthalmologic conditions. It is also a common ingredient in skin care products [3].

HA can be degraded by hyaluronidases, a family of enzymes, and by various non-enzymatic reactions (e.g. acidic/alkaline hydrolysis) [4]. Hyaluronidases have been used in low doses in various conditions, mainly to breakdown extracellular matrix and enhance drug distribution [5,6]. Whether it can be safely applied intrapleurally and in sufficient therapeutic quantity to lyse HA in human MPE will need to be investigated.

At present, clinically available options for viscous MPE are limited. Use of tPA/DNase enhanced the evacuation of thick infected pleural fluid in clinical trials. The regime provided some short-term benefits in Patient 1, but subsequent administration did not prove effective. Our ex vivo experiments did not find a major role of rhDNase in reducing the viscosity. As tPA is known to stimulate significant production of pleural fluid in healthy and diseased pleura, it is possible that the dilutional effect of this fluid formation helped with initial drainages. Whether tPA itself affects viscosity of MPE is unknown.

The composition of pleural fluid may be important to provide an informed clinical decision to ascertain an effective therapy for pleural effusion management. In cases of viscous fluid and high HA levels, strategies to counteract HA production or enhance its breakdown warrant exploration.

**Disclosure Statement**

Appropriate written informed consent was obtained for publication of this case report and accompanying images.

**Acknowledgments**

Y. C. Gary Lee is a recipient of the National Health and Medical Research Council (NHMRC)/Medical Research Future Fund Next Generation Practitioner Fellowship. Y. C. Gary Lee and J. Creaney have received project grant funding from NHMRC, iCare New South Wales Dust Diseases Authority, Sir Charles Gairdner Research Advisory Committee, Institute for Respiratory Health, Cancer Council of Western Australia, and Cancer Australia.
References
1. Simpson G, Roomes D, and Heron M. 2000. Effects of streptokinase and deoxyribonuclease on viscosity of human surgical and empyema pus. Chest 117(6):1728–1733.
2. Creaney J, Dick IM, Segal A, et al. 2013. Pleural effusion hyaluronic acid as a prognostic marker in pleural malignant mesothelioma. Lung Cancer 82(3):491–498.
3. Abatangelo G, Vindigni V, Avruscio G, et al. 2020. Hyaluronic acid: redefining its role. Cells 9(7):1743.
4. Stern R, and Jedrzejas MJ. 2006. Hyaluronidases: their genomics, structures, and mechanisms of action. Chem. Rev. 106 (3):818–839.
5. Weber GC, Buhren BA, Schrumpf H, et al. 2019. Clinical applications of hyaluronidase. Adv. Exp. Med. Biol. 1148:255–277.
6. Buhren BA, Schrumpf H, Hoff NP, et al. 2016. Hyaluronidase: from clinical applications to molecular and cellular mechanisms. Eur. J. Med. Res. 21:5.

Supporting information
Additional Supporting Information may be found in the online version of this article at the publisher’s web-site: http://onlinelibrary.wiley.com/doi/10.1002/rcr2.694/supinfo.

Video S1. Highly viscous pleural fluid from Patient 1 (Sample 2) in the tube labelled untreated is shown. Treatment of the same sample with hyaluronidase (100 U/mL for 10 min) significantly reduced the fluid viscosity as shown in the tube labelled treated.