Urinary interleukin-9 in youth with type 1 diabetes mellitus

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Abstract

Aims Interleukin-9 (IL-9) attenuates podocyte injury in experimental kidney disease, but its role in diabetic nephropathy is unknown. We sought to relate urinary IL-9 levels to the release of podocyte-derived extracellular vesicles (EVs) in youth with type 1 diabetes. We related urinary IL-9 levels to clinical variables and studied interactions between urinary IL-9, vascular endothelial growth factor (VEGF), tumor necrosis factor alpha (TNFα) and interleukin-6 (IL-6) on urinary albumin/creatinine ratio (ACR) a functional measure of podocyte injury.

Methods We performed an analysis of urine samples and clinical data from a cohort of youth with type 1 diabetes (n = 53). Cytokines were measured using a Luminex platform (Eve Technologies), and nanoscale flow cytometry was employed to quantify urinary podocyte-derived EVs. All urinary measures were normalized to urinary creatinine.

Results Mean age was 14.7 ± 1.6 years, and the mean time from diagnosis was 6.7 ± 2.9 years. Mean HbA1c was 70.3 ± 13.9 mmol/mol, mean ACR was 1.3 ± 1.9 mg/mmol, and mean eGFR was 140.3 ± 32.6 ml/min/1.73 m². IL-9 was inversely related to podocyte EVs (r = −0.56, p = 0.003). IL-9 was also inversely related to blood glucose, HbA1C and eGFR (r = −0.44, p = 0.002; r = −0.41, p = 0.003; r = −0.49, p < 0.001, respectively) and positively correlated with systolic BP (r = 0.30, p = 0.04). There was a significant interaction between IL-9, EVs and ACR (p = 0.0143), and the relationship between IL-9 and ACR depended on VEGF (p = 0.0083), TNFα (p = 0.0231) and IL-6 levels (p = 0.0178).

Conclusions IL-9 is associated with podocyte injury in early type 1 diabetes, and there are complex interactions between urinary IL-9, inflammatory cytokines and ACR.

Keywords Cytokines · Albuminuria · IL9 · Extracellular vesicle · Podocyte · Type 1 diabetes mellitus
Introduction

Interleukin-9 (IL-9) is a T cell-derived cytokine, generally felt to be pro-inflammatory, which has been implicated in the regulation of both innate and adaptive immunity. [1, 2] Clinically, elevated serum IL-9 levels have been observed in a number of diseases including systemic lupus erythematosus [3], asthma [4] and coronary artery disease [5]. In a recent study of experimental murine kidney disease, Xiong and coworkers reported that intra-renal IL-9 attenuated podocyte injury in mice with adriamycin-induced nephropathy [6]. IL-9 knockout mice exhibited more podocyte injury and loss of podocyte number in comparison with wildtype mice [6]. The impact of IL-9 on podocyte injury in human kidney disease remains largely unexplored.

Mesangial matrix expansion is a dominant feature of early kidney disease in type 1 diabetes mellitus (T1D), particularly in relationship to hyperfiltration and early declines in glomerular filtration rate (GFR) [7]. Ultrastructural abnormalities in glomerular podocytes also occur in T1D although the changes are subtle, especially in early kidney disease [8]. A decrease in glomerular podocyte number occurs in the Akita mouse, an experimental model of T1D, and the decrease precedes any rise in the urinary albumin excretion rate [9]. These findings suggest that early podocyte injury is an important feature of diabetic kidney disease and that podocyte injury may contribute to functional changes like a rise in albumin excretion. Identifying biomarkers that correlate with early podocyte injury may help risk stratify patients early in the course of their disease.

Extracellular vesicles (EVs), including microparticles, exosomes, and apoptotic bodies, are cell-membrane-derived vesicles released by a variety of cell types in response to injury [10–12]. Cultured podocytes release EVs when subjected to mechanical strain or high glucose concentrations [13]. In accord with these in vitro observations, podocyte-derived EVs are also present in the urine of diabetic mice, and we found increased podocyte-derived EVs in the urine of adults with T1D [13, 14]. More recently, we reported that there was a significant correlation between podocyte-derived EVs and both blood glucose levels and eGFR in youth with T1D [15]. However, the relationship between podocyte-derived EVs and IL-9 in T1D is unknown.

Accordingly, our aim in this exploratory study was to examine the relationship between urinary IL-9 and urinary podocyte-derived EV numbers in the urine, reflecting less podocyte injury and a protective effect of IL-9. We evaluated the relationships between IL-9 and HbA1C, blood glucose (BG), eGFR and blood pressure (BP). We hypothesized there would be negative correlations with each, supporting a reno-protective effect. In an exploratory analysis, we also studied interactions between IL-9, the albumin/creatinine ratio (ACR) and urinary cytokines linked to the progression of diabetic nephropathy: vascular endothelial growth factor (VEGF), interleukin-6 (IL-6) and tumor necrosis factor alpha (TNFα) [16–18].

Methods

Ethical statement and cohort description

We examined urine samples from a Cohort created from Canadian participants recruited from the observational arm of the Adolescent Type 1 Diabetes Cardio-Renal Intervention Trial (AdDIT) [19]. In brief, eligible individuals were 10 to 16 years of age with a diagnosis of Type 1 diabetes. The size of the cohort was based on the exploratory analysis that showed significant relationships between EVs and the clinical variables BG and eGFR [15]. Exclusion criteria included other forms of diabetes, pregnancy or unwillingness to adhere to contraceptive advice and pregnancy testing, severe hyperlipidemia or a family history suggesting familial hypercholesterolemia, the presence of coexisting conditions (excluding treated hypothyroidism and celiac disease), proliferative retinopathy, and the presence of non-diabetic renal disease. Study participants and their guardians gave informed consent. Studies were approved by The Hospital for Sick Children, Research Ethics Board at the University of Toronto (ID# 1,000,012,240).

Serum and urine samples

Fasting first morning midstream urine samples were collected for all participants in the cohort. Urine samples were centrifuged at 2000 g at 4 ºC for 6 min before storage in −80 ºC. All analyses were conducted from a single urine sample from each participant. A single venous blood sample was also taken at the time of urine collection for the analysis of blood glucose levels and HbA1c. HbA1c was measured by enzymatic assay (Architect Analyzer; Abbott Diagnostics, USA).

Urinary cytokines

Urinary levels of cytokines/chemokines were measured with Cytokine/Chemokine Panel Luminex Assay [19] from the urine samples. The urine samples were centrifuged at
1500 g for 15 min, separated into 1 ml aliquots and frozen at −80 °C. The urine was then thawed at 4 °C one day prior to use [19]. We limited our cytokine/chemokine analysis to IL-9, VEGF, TNFα and IL-6 to maintain statistical power. The accuracy and precision of the urinary cytokine/chemokine assay is available from the vendor at [http://www.millipore.com/userguides/tech1/proto_mpzcytol60k](http://www.millipore.com/userguides/tech1/proto_mpzcytol60k). We have previously published the detection limits of the assays [20].

**Podocyte extracellular vesicles**

Quantification of podocyte EVs was performed using our previously described nanoscale flow cytometry approach [13–15]. Briefly, stored urine samples were thawed, and EVs were isolated by sequential centrifugation first 2500 × g for 10 min at 4 °C, followed by 20,000 × g for 20 min at 4 °C. [9] The pellet was re-suspended in Annexin V Binding buffer with Annexin V FITC (1:50, Biologend, California, USA). Podocyte origin was confirmed by co-staining with an anti-podoplanin APC-conjugated antibody (1:100, Biologend, California, USA). Samples were analyzed by nanoscale flow cytometry (CytoFLEX S, Beckman Coulter, USA), and ApogeeMix beads (Cat. 1493–Apogee Flow Systems, UK) were used for size calibration. EVs were defined as particles sized between ~100–1000 nm. Samples labeled with isotype controls, antibodies alone in buffer, and unlabeled samples were analyzed as controls. FlowJo ver 7.6.5 was used for analysis. Levels of urinary EVs were normalized to urinary creatinine levels and podocyte EV levels were expressed as number/umolCr.

**ACR measurement**

ACR was measured in 2 sets of 3 of the collected urine samples. A normal albumin excretion rate was defined this as an ACR < 2.0 mg/mmol [21]. Microalbuminuria was defined as an ACR of 2.0 to 20 mg/mmol.

**Estimated glomerular filtration rate (eGFR)**

eGFR was calculated using the Larsson equation (cystatin C) as previously described [19]. Serum cystatin C was measured by a single operator using an immunoassay (Dade Behring Diagnostics, Newark, DE, USA) conducted on a BN Prospec System nephelometer [19]. Hyperfiltration was defined as an eGFR > 135 ml/min/1.73m2. [19].

**Blood pressure**

BP values were the mean of two separate in-clinic measurements using an automated and validated BP cuff. Normal BP, elevated BP, stage one and stage two hypertension were defined as per American Academy of Pediatric Guidelines [22]. We calculated the BP percentiles using [https://apps.cpec-gcep.net/BPz_cpec//](https://apps.cpec-gcep.net/BPz_cpec//), accessed July 2020). [15].

**Statistical analysis**

Descriptive statistics were used to describe the clinical characteristics of the sample, including mean and standard deviation for continuous measures, frequency and percentages for categorical variables. Spearman correlations were used to assess the strength of the association among continuous measures. In all parts of the analysis, p values were corrected for multiple comparisons using false discovery rate method. Our primary outcome of interest was the relationship between IL-9 and podocyte EVs. Data were disaggregated by sex to confirm the relationship. Secondary outcomes included the relationship between IL-9 and BP, glycemic control, eGFR and the cytokines VEGF, TNFα and IL-6. We then looked for interactions between IL-9, podocyte EVs and ACR. Finally, we looked for interactions between IL-9, the cytokines VEGF, TNFα and IL-6, and ACR. We log-transformed the continuous measures and used linear regression to assess the significance of the interactions.

**Results**

**Clinical characteristics of the study cohort**

The mean age was 14.7 ± 1.6 years, and the mean duration of time from diagnosis of diabetes was 6.7 ± 2.9 years. There were 26 males and 27 females. Ethnicity of the T1D group was as follows: 32/53 (60.4%) were white, 6/53 (11.3%) were black, 3/53 (5.7%) were South Asian, 4/53 (7.5%) were South East Asian, and 8/53 (15.1%) were classified as other. HbA1C levels averaged 70.3(8.6%) ± 13.9(3.1%) mmol/mol (Table 1), and the average BMI (kg/m2) was 22.1 ± 3.7 kg/m2. The mean eGFR (Larsson) was 140.3 ± 32.6 ml/min/1.73m2, and 47% of the cohort exhibited hyperfiltration. The mean ACR was 1.3 ± 1.9. Only 13% of the cohort exhibited an ACR > 2 mg/mmol (Table 1).

**Urinary IL-9 and urinary podocyte extracellular vesicles**

The median value for the podocyte-derived EVs normalized to urinary creatinine (EV/UCr) was 7.88 in the cohort, and as we previously reported there was no differences in the median urinary EV values between male and female subjects [15]. There was a negative correlation between IL-9 and EV (r = −0.56, p < 0.0003) such that higher values of IL-9 were associated with lower EV shown in Fig. 1A. The relationship
was similar in female \((r = -0.56, p = 0.003)\) and male subjects \((r = -0.58, p = 0.002)\) as seen in Fig. 1B and C.

**Spearman correlation coefficients relating urinary IL-9, clinical variables and the cytokines VEGF, TNFα and IL-6**

There was a negative correlation between IL-9 and eGFR \((r = -0.49, p < 0.0005)\) such that higher values of eGFR were associated with lower IL-9 levels (Table 2). There was a negative correlation between IL-9 and BG \((r = -0.44, p < 0.002)\) such that higher BG values were associated with lower IL-9 levels. Similarly, there was a negative correlation between IL-9 and HbA1C \((r = -0.41, p < 0.004)\) with higher HbA1C measurements associated with lower IL-9 levels. In contrast, there was a positive correlation between IL-9 and systolic blood pressure.
There were positive correlations between IL-9 and VEGF (r = 0.72, p < 0.0003), IL-9 and TNFα (r = 0.55, p < 0.0003) and IL-9 and IL-6 (r = 0.47, p = 0.001).

**Spearman correlation coefficients relating urinary IL-9, EV and ACR**

The correlations between IL-9 and ACR (r = −0.26, p = 0.073) and between EV and ACR (r = 0.20, p = 0.16) were more modest and did not reach statistical significance (Table 3). However, there was a significant interaction between IL-9, EV and ACR (p = 0.0143). As shown in Fig. 2A, the relationship between EV and ACR is dependent on the IL-9 levels such that the steepness and direction of the relationship varies depending on the IL-9 percentile (10th to 90th) in the cohort. The 3-D plot in Fig. 2B is a visual representation of the interaction between these three variables.

**Spearman correlation coefficients relating urinary IL-9, urinary VEGF and ACR**

There was a strong positive correlation between IL-9 and VEGF (r = 0.72, p < 0.001) such that higher values of IL-9 were associated with higher values of VEGF (Table 3). The modest correlation between VEGF and ACR (r = −0.23, p = 0.13) did not reach statistical significance. There was, however, a significant interaction between IL-9, VEGF and ACR (p = 0.0083). As shown in Fig. 3A, the relationship between IL-9 and ACR is dependent on the VEGF levels such that the steepness and direction of the relationship varies depending on the VEGF percentile (10th to 90th).

**Spearman correlation coefficients relating urinary IL-9, urinary TNFα and ACR**

There was a positive correlation between IL-9 and TNFα (r = 0.55, p < 0.001), but the correlation between TNFα and ACR (r = −0.24, p = 0.13) did not reach statistical significance (Table 3). There was a significant interaction between IL-9, TNFα and ACR (p = 0.0231). As shown in Fig. 3B, the relationship between IL-9 and ACR is also dependent on the TNFα levels. Again, the steepness and direction of the relationship varies depending on the TNFα percentile (10th to 90th).

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**Table 2** Spearman correlations coefficients for observed relationships with urinary interleukin-9

| Clinical variable     | Spearman correlation | p value  |
|-----------------------|----------------------|----------|
| eGFR                  | −0.49401             | 0.0005   |
| Blood glucose         | −0.43534             | 0.0023   |
| ACR                   | 0.2601               | 0.0752   |
| Systolic blood pressure | 0.29846           | 0.0417   |
| Diastolic blood pressure | 0.04467           | 0.7556   |
| HbA1C                 | −0.40896             | 0.0041   |
| Podocyte EV           | −0.55692             | 0.0003   |
| Cytokines             |                      |          |
| VEGF                  | 0.71701              | 0.0003   |
| TNFα                  | 0.55391              | 0.0003   |
| IL-6                  | 0.46874              | 0.0014   |

*eGFR* estimated glomerular filtration rate, *ACR* albumin/creatinine ratio, *EV* extracellular vesicle, *VEGF* vascular endothelial growth factor, *TNFα* tumor necrosis factor alpha, *IL-6* interleukin-6

(SBP) (r = 0.30, p < 0.04) but not for IL-9 and diastolic blood pressure (DBP) (r = 0.05, p = 0.76).

**Table 3** Spearman correlations coefficients and the interaction terms for three variable analyses with interleukin-9

| Variables                      | Correlation | P value  |
|--------------------------------|-------------|----------|
| IL-9 and ACR                   | −0.26011    | 0.0943   |
| Podocyte extracellular vesicle  |              |          |
| analysis                       |             |          |
| Podocyte EV and IL-9            | −0.55692    | 0.0004   |
| Podocyte EV and ACR             | 0.20433     | 0.1629   |
| Interaction term                |             |          |
| IL-9, podocyte EV, ACR          | 0.0143      |          |
| VEGF analysis                   |             |          |
| VEGF and IL-9                   | 0.71701     | 0.0004   |
| VEGF and ACR                    | −0.22874    | 0.1258   |
| Interaction term                |             |          |
| IL-9, VEGF, ACR                 | 0.0083      |          |
| TNFα analysis                   |             |          |
| TNFα and IL-9                   | 0.55391     | 0.0004   |
| TNFα and ACR                    | −0.23531    | 0.1258   |
| Interaction term                |             |          |
| IL-9, TNFα, ACR                 | 0.0231      |          |
| IL-6 analysis                   |             |          |
| IL-6 and IL-9                   | 0.46874     | 0.0023   |
| IL-6 and ACR                    | −0.09641    | 0.5099   |
| Interaction term                |             |          |
| IL-9, IL-6, ACR                 | 0.0178      |          |

*IL-9* interleukin-9, *ACR* albumin/creatinine ratio, *EV* extracellular vesicle, *VEGF* vascular endothelial growth factor, *TNFα* tumor necrosis factor alpha, *IL-6* interleukin-6
Spearman correlation coefficients relating urinary IL-9, urinary IL-6 and ACR

There was a positive correlation between IL-9 and IL-6 ($r = 0.47$, $p = 0.002$), but the correlation between IL-6 and ACR ($r = -0.10$, $p = 0.51$) did not reach statistical significance (Table 3). There was a significant interaction between IL-9, IL-6 and ACR ($p = 0.018$). As shown in Fig. 3C, the relationship between IL-9 and ACR is also dependent on the IL-6 levels. The steepness and direction of the relationship varies depending on the IL-6 percentile (10th to 90th).

Discussion

In this study, we sought to characterize relationships between IL-9 and early podocyte injury in youth with T1D. The rationale was based on recent studies that revealed a protective effect of IL-9 on glomerular podocyte injury in experimental kidney disease. [6] Our first major observation was that there was a negative correlation between urinary IL-9 and podocyte-derived EVs, a relationship that was similar in females and males. Although correlation does not establish causation, the current study is the first to relate urinary IL-9 to measures of glomerular podocyte injury in humans with T1D, and the finding is consistent with the hypothesis that IL-9 may limit podocyte injury in diabetic subjects, which is reflected by the release of EVs into the urine.

We have reported that urinary IL-9 levels are higher in adolescents with T1D compared to healthy age-matched subjects [23], but there is a paucity of data on urinary levels of IL-9 in diabetes. Vasanthakumar and coworkers reported that serum levels of IL-9 were lower in adults with type 2 diabetes and normal kidney function compared to healthy subjects [24]. They did find that IL-9 levels increased in participants with diabetic kidney disease [24]. Although IL-9 may limit podocyte injury in early diabetic kidney disease, its role in advanced disease is less clear.

Hyperglycemia, hyperfiltration and hypertension are risk factors for the onset and progression of diabetic nephropathy [25–28]. As part of our exploratory analysis, we therefore sought to relate IL-9 to glycemia (using both the blood glucose level at the time of urine collection and HbA1C), eGFR and blood pressure. There were negative correlations seen between urinary IL-9 with both HbA1C and BG at the time of urine collection. We similarly found lower values of IL-9 were associated with higher values of eGFR. These correlations support the hypothesis that IL-9 is reno-protective. In contrast, IL-9 was positively correlated with SBP but not with DBP. It is important to note that few of the study participants were hypertensive.

Our next major observation involved a functional correlate of podocyte injury, ACR. We discovered that there was a significant interaction between IL-9, podocyte-derived EVs and ACR. At low levels of urinary IL-9, there is a positive relationship between EVs and ACR, suggesting that as podocyte injury worsens, reflected by increasing EVs, there is a rise in the ACR. As IL-9 levels (represented as quintiles in Fig. 2A) rise, the association between EVs and ACR flattens, suggesting that urinary levels of IL-9 may change the relationship between podocyte injury reflected by EVs and ACR.
IL-9 was also strongly correlated with VEGF, TNFα and IL-6 (Table 2) all of which have all been previously implicated in diabetic nephropathy [16–18]. VEGF is primarily produced by podocytes and plays a critical role in maintenance of the fenestrated endothelium in the glomerular basement membrane [29]. IL-9 may limit podocyte injury and accordingly preserve VEGF synthesis. Members of the TNFα superfamily have been identified as biomarkers of kidney risk in several clinical studies of both T1D and type 2 diabetes [18, 30]. IL-6 expression is increased in diabetic nephropathy, and levels correlate with glomerular basement membrane thickening [17, 31]. Interestingly, both TNFα and IL-6 regulate IL-9 expression, which may explain the positive correlation [6, 32, 33].

ACR is a functional outcome of podocyte injury, and we found that there were significant interactions between IL-9 with VEGF, TNFα, and IL-6 and ACR (Table 3). The direction of the relationship between IL-9 and ACR was dependent on each of these cytokines (Table 3), and each cytokine exhibited a similar pattern. At low levels of VEGF, TNFα and IL-6 (10th percentile line in Fig. 3A, B, C), there is a negative relationship between IL-9 and ACR. At high levels (90th percentile line in Fig. 3A, B, C), there is a positive relationship between IL-9 and ACR. This suggests that kidney inflammation impacts the protective effect of IL-9 on podocyte function.

The strengths of this study include evaluation of a well-characterized cohort of adolescents with T1D with standardized collections and analysis of bio-specimens. However, there are also some important limitations. First, we did not identify the source of IL-9 in the urine. IL-9 is primarily produced by T helper cells recruited by IL-6 [34, 35] and by T-regulatory cells [36]. Animal models of diabetic nephropathy have shown helper T cells (CD4+), cytotoxic (CD8+) T cells and small numbers of T-regulatory cells in kidney [34, 37]. As inflammation in the kidney increases, infiltration of T cells may result in increased levels of IL-9. Second, the data are cross-sectional and therefore the implications of our findings with respect to the progression of ACR and declining GFR are unknown. Establishing an IL-9 threshold for high-risk patients could make it a clinically useful biomarker and establish its role as a potential treatment target. Long-term studies of larger cohorts will be required to address the limitation of our current study. Thirdly, the analyses are limited to correlations and interactions and do not establish causation. In vitro studies of cultured podocytes and studies of IL-9 gene deletion in murine models
of diabetes can address these limitations. Finally, validation in other populations of T1D will strengthen our conclusions and allow for generalizability.

Conclusions

In summary, there is a strong relationship between urinary IL-9 and measures of glomerular podocyte injury (the release of EVs) in youth with T1D. Complex interactions between IL-9, cytokines and ACR support the hypothesis that inflammation is an important determinant of both glomerular injury and function in the diabetic kidney.

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Authors’ contributions All authors made substantial contributions to the conception and design, acquisition of data, analysis and interpretation of data, drafting the article and revising it critically for important intellectual content and were involved in the final approval prior to publication. Julie Semenchuk is the guarantor of this work.

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Availability of data and materials The datasets used and/or analyzed during the current study are available from the corresponding authors on reasonable request.

Declarations

Conflict of interest The authors declare that they have no competing interests.

Consent for publication Not applicable.

Ethics approval and consent to participate Informed consent from each individual and their guardian was obtained for all participants. This research received approval from The Hospital for Sick Children, Research Ethics Board at the University of Toronto (ID# 1000012240).

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