Zoonotic infections with avian influenza A viruses and vaccine preparedness: a game of “mix and match”

Various direct avian-to-human transmissions of influenza A virus subtypes upon exposure to infected poultry have been previously observed in the past decades. Although some of these strains caused lethal infections, the lack of sustained person-to-person transmission has been the major factor that prevented these viruses from causing new pandemics. In 2013, three (A/H7N9, A/H6N1, and A/H10N8) novel avian influenza viruses (AIVs) yet again breached the animal-human host species barrier in Asia. Notably, roughly 20% of the A/H7N9-infected patients succumbed to the zoonotic infection whereas two of three A/H10N8 human infections were also lethal. Thus, these events revived the concerns of potential pandemic threats by AIVs in the horizon. This article reviews the various human incursions with AIV variants and provides insight on how continued circulation of these viruses poses perpetual challenge to global public health. As the world anticipates for the next human pandemic, constant vigilance for newly emerging viruses in nature is highly encouraged. With the various numbers of AIVs demonstrating their capacity to breach the animal-human host interface and apparent limitations of current antivirals, there is a need to broaden the selection of pre-pandemic vaccine candidate viruses and development of novel alternative therapeutic strategies.

Keywords: Avian influenza A virus, Humans, Pathogenicity, Transmission, Pandemic, Influenza vaccines

Introduction

Influenza A (flu A) viruses belong to the Orthomyxoviridae family of RNA viruses and are the causative agent of influenza, a contagious respiratory viral disease of birds and humans [1]. They possess a negative-sense viral RNA genome packed into 8 individual gene segments which code for at least 11 functional proteins needed for infection, replication, and evasion of antiviral responses. However, recent studies suggest that there could be more viral protein products depending on the virus strain [2-4]. Flu A viruses can be further subdivided into distinct subtypes based on the genetic and antigenic properties of their hemagglutinin (HA) and neuraminidase (NA), the envelope glycoproteins express on the surface of virus particles responsible for virus binding to relevant hosts and releasing of newly formed infectious virions, respectively [5]. Sixteen HA and 9 NA subtypes have been detected in wild aquatic birds and poultry throughout the world [1] while new strains (H17-18 and N10-11) have been additionally identified in bat species [6,7].
Generally, flu A viruses are found in migratory waterfowls, the principal natural reservoirs [5]. Part of their genetic variability is due to the absence of a proof-reading mechanism of the flu A viral RNA polymerase leading to faulty replication cycles. Similarly, the segmented nature of the RNA genome is also fundamental to the genetic evolution of flu A viruses. Co-infection of a single host with two or more viruses facilitates genetic reassortment, the rearrangement of genetic material by the exchanging of gene segments, creating progeny viruses with novel gene constellation one of which may have the ability to infect new hosts [1]. Thus, even if they are generally host-specific and not readily transmissible to other species, a wide array of flu A viruses have been found in other animals which include sea mammals, domestic poultry, horses, dogs, pigs, and people [5]. Moreover, flu A viruses are responsible for the annual global influenza epidemics and occasional pandemics in human populations. In recent years, number of flu A viruses crossing the animal-human host species barrier have been increasing. This article provides a general overview of these zoonotic infections and the constant threat of a looming pandemic.

**Variant Avian Flu A Viruses Occasionally Breach the Animal-Human Interface**

Avian influenza viruses (AIVs) recognize specific sialic acid receptors of susceptible cells lining the intestinal gut of avian hosts [8]. However, specific modifications in HA through adaptation shifts receptor recognition, which in turn effectively switches binding preferences to specific receptors expressed by mammalian hosts, including humans. Further genetic alterations at the HA cleavage motif of some subtypes (e.g., H5 and H7) could make HA readily cleaved by ubiquitous cellular proteases [9]. Accumulation of polybasic residues on this viral genome could confer airborne transmission between mammals [19,20]. Various AIV subtypes have occasionally crossed the animal-human interface commonly occurring after close contact to infected birds (Table 1). Of note, humans also possess AIV-susceptible cells in the lower respiratory tract (e.g., lungs) apart from mammalian-specific receptors; such receptors are not expressed by upper respiratory tract tissues (e.g., nasal passages and trachea) [12]. Thus, it is not surprising that extensive or prolonged exposure to affected poultry increases the chances of interspecies transmission.

Among the vast pool of AIVs in nature, HPAI A/H5N1 viruses have been considered to present the greatest concern to cause the next flu pandemic (Table 1). Since its first appearance in Guangdong, China in 1996, it has already spawned various clusters of antigenically divergent HPAI H5 lineages that spread widely in Asia towards Europe and Africa [13], causing devastating poultry outbreaks along its path. In humans, more than 640 HPAI A/H5N1 infections worldwide have been recorded since 1997 by the World Health Organization wherein about 60% were fatal [14]. Aberrant innate immune responses have been shown to be a hallmark of the atypical virulence of the HPAI A/H5N1 virus that results in the high incidence of mortality [15-17]. In 2014 alone, new cases have been already reported in Asia with Vietnam and Cambodia recording fatalities for the year [14]. The first imported case in North America was also recently confirmed in a resident of Alberta, Canada who returned from Beijing, China in December 27, 2013, was hospitalized, and died after few days due to viral infection [18]. At present, there are no imminent indications that contemporary HPAI A/H5N1 viruses have the capacity to support efficient viral transmission in humans. However, two independent studies have simultaneously demonstrated how few changes in the HPAI A/H5N1 viral genome could confer airborne transmission between mammals [19,20].

Human infection with A/H9N2 AIVs was first documented in Hong Kong in 1999 [21]. Retrospectively, it was postulated that the HPAI A/H5N1 virus, which caused the 1997 human outbreaks also in Hong Kong, acquired most of its gene segments from avian A/H9N2 subtypes that were circulating in 1997 [22]. Sporadic cases of AIV A/H9N2 zoonoses followed ever since with at least 16 human total infections as shown in the influenza virus sequence database (http://www.ncbi.nlm.nih.gov/genomes/FLU/FLU.html) [23], including a recent Hong Kong case in late December 2013, the first in four years [24]. Serologic studies also showed that some poultry workers in China and India were positive for A/H9N2 virus infection through HA inhibition and virus micro-neutralization assays [25-27]. Notably, avian A/H9N2 infections only induced rela-
Table 1. Reported cases of human infections with avian influenza A viruses

| Nation            | H5N1| H6N1| H7N2| H7N3 | H7N7 | H7N9 | H9N2 | H10N7 | H10N8 |
|-------------------|-----|-----|-----|------|------|------|------|-------|-------|
| Africa            |     |     |     |      |      |      |      |       |       |
| Djibouti          | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Egypt             | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Nigeria           | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| America (North)   |     |     |     |      |      |      |      |       |       |
| Canada            | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| New York          | -   | -   | √   | -    | -    | -    | -    | -     | -     |
| Virginia          | -   | -   | √   | -    | -    | -    | -    | -     | -     |
| America (South)   |     |     |     |      |      |      |      |       |       |
| Mexico            | -   | -   | -   | -    | -    | -    | -    | -     | -     |
| Asia              |     |     |     |      |      |      |      |       |       |
| Azerbaijan        | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Bangladesh        | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Cambodia          | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| China             | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Hong Kong         | √   | -   | -   | -    | √    | √    | -    | -     | √     |
| Indonesia         | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| India             | -   | -   | -   | -    | -    | √    | -    | -     | -     |
| Iraq              | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| LPDR              | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Malaysia          | -   | -   | -   | -    | -    | -    | -    | -     | -     |
| Myanmar           | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Pakistan          | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Taiwan            | -   | -   | √   | -    | -    | -    | -    | -     | -     |
| Thailand          | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Turkey            | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Vietnam           | √   | -   | -   | -    | -    | -    | -    | -     | -     |
| Europe            |     |     |     |      |      |      |      |       |       |
| Italy             | -   | -   | -   | √    | -    | -    | -    | -     | -     |
| Netherlands       | -   | -   | -   | -    | √    | -    | -    | -     | -     |
| United Kingdom    | -   | -   | √   | √    | √    | -    | -    | -     | -     |
| Australia         |     |     |     |      |      |      |      |       |       |
| Australia         | -   | -   | -   | -    | -    | -    | -    | √     | -     |

Total cases  
≥ 650  
1  
≥ 4  
≥ 9  
90  
≥ 350  
≥ 16  
4  
3

Most of these cases, obtained from the World Health Organization, US Center for Disease Control and Prevention, and in PubMed, have been associated with close contact or prolonged exposure to infected birds. To date, HPAI A/H5N1 infections remain the highest in occurrence with about 60% of the cases lethal. However, A/H7N9 human infections have been constantly growing since its first detection in March 2013. From about 350 cases recorded, over 70 (roughly 20%) reportedly succumbed to virus infection. Amid the H7N9 infections still being reported, another novel avian H10N8 strain also killed two of its three victims since December 2013. LPDR, Lao People’s Democratic Republic.

*Positive detection through serologic assays only.

- The report indicates a total of 650 cases, with 1 case reported in H6N1, 4 cases in H7N2, 9 cases in H7N3, 90 cases in H7N7, 350 cases in H7N9, 16 cases in H10N7, and 4 cases in H10N8.

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- HPAI A/H5N1 infections remain the highest in occurrence with about 60% of the cases lethal.

- A/H7N9 human infections have been constantly growing since its first detection in March 2013.

- Around 70 cases (20%) of A/H7N9 reported the virus infection.

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- The report highlights the difference in mortality rates between H7 and H5 subtypes, with H7 strains being less virulent.

- Prior to 1996, human infections with the H7 AIV subtype were primarily due to laboratory or occupational exposures.[28]. Direct avian-to-human transmission of a low pathogenic avian influenza (LPAI) A/H7N7 was first confirmed in 1996 on an English woman while tending to her apparently infected pet ducks.[29]. In 2002, LPAI A/H7N2 virus was detected in Virginia.[30] and isolated in an immunocompromised New York resident.[31]; this subtype was also found among poultry workers in Wales, England.[32]. Similarly, an LPAI A/H7N3 was recovered from a poultry attendant in the UK.[33] whereas retrospective serologic analysis of workers who responded during the 2002-2003 H7N3 Italian poultry outbreak identified 7 persons seropositive.[34]. In contrast to these LPAI H7 strains, HPAI A/H7N3 variants were responsible for zoonotic infections among poultry workers in British Columbia, Canada.[35] and Jialisco, Mexico.[36]. Before 2013, the largest human H7 outbreak occurred in the Netherlands.[37] caused by an HPAI A/H7N7 tallying over 80 cases includ-
ing one fatality. Up to that point, all of the H7 patients have only presented generally mild respiratory disease commonly accompanied by conjunctivitis, except for the United States cases [28]. However, pathobiology of H7 AIVs changed when human cases caused by an A/H7N9 virus emerged since late March 2013 killing a quarter of those infected [38]. Unlike its A/H5N1 and A/H9N2 relatives, A/H7N9 does not induce remarkable influenza-like disease signs in afflicted poultry albeit efficient replication [39,40]. The pathogen, a product of multiple reassortment between strains found in migratory birds and prevailing A/H9N2 viruses in Chinese poultry [41], does not contain the pathotypic HA cleavage motif but show indications that it can infect and replicate well in mammalian species [39,40]. As of writing, more than 350 cases have been confirmed with over 70 casualties mostly from mainland China. Imported cases in Hong Kong, Taiwan and a Chinese visitor in Malaysia are the only other reports outside the Chinese territory [42], indicating that it hasn’t yet gone global dissemination. Notably though, human infections with A/H7N9 viruses have been characterized by rapid progression to severe pulmonary disease and acute respiratory distress syndrome, reminiscent of human HPAI A/H5N1 disease [39-41]. The scale of these zoonotic infections also highlights the need for enhanced surveillance in neighboring countries. The geographic location of South Korea puts it well within the East Asian flyway of migratory waterfowls. In addition, exchange of tourists between China and Korea are constantly growing due to close proximity and improved air travel. Thus, A/H7N9 could be carried over by infected migratory wild birds and/or human traveler/visitors as what happened in the imported case of Malaysia.

Apart from the more popular strains above, H6 and H10 AIV infections have been also recorded. In May 2013, the first human case of A/H6N1 AIV infection was reported [43]. The woman patient, who in worked in a deli shop and has not visited or had any contact with poultry, was hospitalized after

Fig. 1. Transmission and spread of avian influenza viruses from wild to domestic birds and potential spill to humans. Wild aquatic birds could carry avian influenza viruses (AIVs) over long distances. Ducks play important role in the transmission of AIVs from migratory waterfowls to domestic poultry. Newly introduced AIVs cause occasional poultry outbreaks, as in the case of highly pathogenic avian influenza H5 and H7 viruses, resulting to mass culling that leads to huge economic losses. Some AIVs become established in domestic poultry where they co-circulate and reassort with locally endemic strains producing novel virus progenies. Similarly, these newly formed virus variants find their way back to migratory birds through ducks. Direct human infections with AIVs typically occur during outbreak settings due to extensive exposure to infected poultry usually. These cases, although sporadic, could provide an opportunity for the virus to evolve into one that could sustain efficient human-to-human transmission. Meanwhile, introduction of AIVs into pig herds could further hasten up the genetic evolution of AIVs through adaptation and/or reassortment which could generate strains with potential threat to animal and human health.
developing high fever, cough and shortness of breath, but fully recovered upon early medication. Occurrences of A/H10N7 infections were also separately recorded in Egypt [44] and Australia [45]. Although those patients had minor symptoms and eventually recovered, two of three A/H10N8 AIV infections in China since December 2013 succumbed to death due to infection [46,47]. Like its A/H7N9 predecessor, A/H10N8 does not cause remarkable disease in birds and harbor segments from A/H9N2 virus segments in Chinese poultry [46].

**Pigs as Intermediate Hosts that Could Hasten Virus Evolution**

With the inherent capacity of flu A viruses to evolve and adapt, it is only a matter of time before novel AIV variants capable of sustaining person-to-person transmission arise. The role of pigs as a focal point in the generation of novel flu A viruses with pandemic potential has been constantly highlighted. The presence of mixed populations of sialic acid viral receptors that line the pig respiratory tract epithelia make pigs susceptible to avian and human flu strains, apart from their own viruses [8,48]. Hence, infection of pigs with AIVs could facilitate adaptation to mammalian hosts particularly humans via accumulation of adaptive mutations (Fig. 1). Meanwhile, simultaneous infection of a pig with at least two viruses from different sources and lineage could promote genetic reassortment.

Through natural and experimental conditions, some of the notable AIVs mentioned above have the capacity to infect and become established in swine populations. Although domestic pigs appear to have low susceptibility [49], HPAI A/H5N1 viruses have been isolated from pig herds in China [50-53] and Indonesia [54,55]. Serologic exposure to HPAI A/H5N1 viruses was also detected in asymptomatic pigs in Vietnam [56]. Since 1998, several A/H9N2 viruses have been also isolated from pigs in Hong Kong and other provinces of China [56-59]. Somehow, a more recent study predicted that A/H9N2 viruses may have become endemic among domestic pigs [59]. Variants of the A/H7N9 virus have been also shown to productively infect pigs in experimental settings but were unable to transmit to other animals [60]. It is also noteworthy that the internal gene constellations of the newly emergent A/H7N9 and A/H10N8 viruses contain corresponding segments from A/H9N2 virus subtypes prevailing in Chinese poultry [38,40,46]. Given the general reputation of pigs with regards to virus ecology and evolution, their role in the genetics in either or both of these AIVs that are making ripples for pandemic concerns cannot be ruled out. Regardless, introduction of these AIVs in swine populations could provide alternative means to hasten genetic evolution (Fig. 1).

**Pre-pandemic Vaccine Development and the Need for Novel Antiviral Drugs**

Vaccination remains the most effective countermeasure against the spread of novel and potentially pandemic flu A viruses [61]. For the past decade, several vaccine strategies and preparations for humans have been developed and are being tested ranging from inactivated, live-attenuated, recombinant protein-based, virus-like particles to DNA-based and adenovirus-vectored vaccines [62]. Since the world's population is expected to have no pre-existing immunity against A/H5N1 strains, the use of efficacious and safe adjuvants to maximize vaccine coverage is also being considered and evaluated. There are also increasing efforts in developing universal vaccines that target conserved domains of various viral proteins [62]. Acknowledging the probability of a future A/H5N1 pandemic, the World Health Organization Global Influenza Programme for its part has recommended the development of representative pre-pandemic H5N1 vaccine candidate viruses that represent antigenically and genetically distinct H5 strains, particularly those that have infected humans through contact with ill or dead infected birds, for stockpiling [63]. Such pre-pandemic vaccines have been prepared by reverse genetics, and safety tested prior to distribution for pilot vaccine production that may be used for experimental studies and for stockpiling [64].

Because A/H7N9 infection of humans is very rare, it is not surprising to find that no pre-existing immunity was detected against the A/H7N9 virus in any of the age groups tested in serologic assays [65]. Apparently, receipt of current seasonal human flu vaccines did not also induce detectable cross-reactive antibodies suggesting that even current seasonal vaccination will not afford protection. In a more promising note though, few groups have shown that some H7-specific vaccine preparations, developed by applying strategies learned from the H5 and 2009 pandemic vaccine development efforts, could offer protection from A/H7N9 virus challenge in animal models [66-69]. However, their efficacy in human subjects remains to be evaluated. Although pre-pandemic H5 and H7 vaccines are in the pipeline for development, there is also a dire need for other subtype-specific candidate vaccines.
(e.g., H6, H9, and H10). Furthermore, strategies that can augment vaccine coverage will be invaluable in the event of a pandemic to meet global demands.

Antiviral drugs provide an important alternative public health intervention for preventing and treating influenza when a vaccine is not available. Currently, two major classes of anti-flu drugs are available which inhibit the release of infectious viruses (NA inhibitors) and block the matrix protein 2 ion-channel ( adamantane derivatives), respectively [70,71]. However, given the plasticity of the flu A viral genome, the emergence of drug-resistant strains against these classes of prophylactic agents could arise. Apart from that, the timing of treatment is also significant on their efficacy. Notably, emerging variants of the human-infecting A/H7N9 virus already gained NA inhibitor resistance without losing their viability and pathogenicity [72]. Altogether, these underscore the need for the development of new antiviral drugs and other novel preventive and therapeutic measures.

Conclusion

At present, none of the zoonotic AIVs above indicated efficient human-to-human dissemination beyond index cases. However, the growing number of cases sparks continuing worries that they could eventually evolve into variants that will cause global concern. Recent evidence suggest that after the initial outbreaks, the human-infecting flu A (H7N9) and local avian A/H9N2 strains had undergone further reassortment which may be responsible for the sudden increase in human cases in early 2014 [73]. It should be noted that HPAI A/H5N1 took about five years to attain the number of cases A/H7N9 is currently racking up. Such impressive disease burden catapults A/H7N9 alongside HPAI A/H5N1, if not ahead, as the next potential human pandemic contender. Unlike HPAI A/H5N1 viruses which cause massive die-offs in affected poultries, A/H7N9 and A/H10N8 do not induce noticeable clinical disease indicators. Such feature would make virus tracking and surveillance more difficult and challenging. In another light, the constant inclusion of A/H9N2 viral components into the zoonotic A/H5N1, A/H7N9, and A/H10N8 strains raises the question whether A/H9N2 viruses predominantly circulating in Chinese poultry could have adapted to support human infections when mixed with the right combination of segments (e.g., HA and NA).

Overall, these zoonotic transmissions validate the potential of AIVs to cause the next human pandemic. With the continued circulation of these viruses, the generation of more virulent and human-transmissible variants, however remote at the moment, remains a possibility given the right conditions and appropriate hosts. Hence, these concerns collectively call for continued virus monitoring and surveillance in humans and animal populations alike, worldwide. Coupled with virus characterization and pathobiologic assessment, these will help identify which strains we should be particularly worried about and set focus on in the future. Finally, the recent incursions of variant flu strains in humans, apart from H5 and H7 viruses, also require the need for appropriate virus-specific or efficacious universal vaccines that can induce broad cross-protective responses which can be made available and administered in the event of or even before a pandemic happens.

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References

1. Palese P, Shaw ML. Orthomyxoviridae: the virus and their replication. In: Knipe DM, Howley PM, editors. Fields virology. 5th ed. Philadelphia: Lippincott Williams & Wilkins; 2007. p.1647-90.
2. Jagger BW, Wise HM, Kash JC, et al. An overlapping protein-coding region in influenza A virus segment 3 modulates the host response. Science 2012;337:199-204.
3. Muramoto Y, Noda T, Kawakami E, Akkina R, Kawaoka Y. Identification of novel influenza A virus proteins translated from PA mRNA. J Virol 2013;87:2455-62.
4. Wise HM, Foeglein A, Sun J, et al. A complicated message: Identification of a novel PB1-related protein translated from influenza A virus segment 2 mRNA. J Virol 2009;83:8021-31.
5. Webster RG, Bean WJ, Gorman OT, Chambers TM, Kawaoka Y. Evolution and ecology of influenza A viruses. Microbiol Rev 1992;56:152-79.
6. Tong S, Li Y, Rivailier P, et al. A distinct lineage of influenza A virus from bats. Proc Natl Acad Sci U S A 2012;109:4269-74.
7. Tong S, Zhu X, Li Y, et al. New world bats harbor diverse influenza A viruses. PLoS Pathog 2013;9:e1003657.
8. Ito T, Couceiro JN, Kelm S, et al. Molecular basis for the...
generation in pigs of influenza A viruses with pandemic potential. J Virol 1998;72:7367-73.

9. Steneke-Grober A, Vey M, Angliker H, et al. Influenza virus hemagglutinin with multibasic cleavage site is activated by furin, a subtilisin-like endoprotease. EMBO J 1992; 11:2407-14.

10. Garten W, Klentz HD. Cleavage activation of the influenza virus hemagglutinin and its role in pathogenesis. In: Klentz HD, Matrosovich MN, Stech J, editors. Avian influenza. Vol. 27. Basel: Karger; 2008. p.156-67.

11. Medina RA, Garcia-Sastre A. Influenza A viruses: new research developments. Nat Rev Microbiol 2011;9:590-603.

12. Shinya K, Ebina M, Yamada S, Ono M, Kasai N, Kawaoka Y. Avian flu: influenza virus receptors in the human airway. Nature 2006;440:435-6.

13. Sonnberg S, Webby RJ, Webster RG. Natural history of highly pathogenic avian influenza H5N1. Virus Res 2013;178: 63-77.

14. World Health Organization (WHO). Cumulative number of confirmed human cases of avian influenza A(H5N1) reported to WHO, 2003-2014 [Internet]. Geneva: WHO; c2014 [cited 2014 Feb 28]. Available from: http://www.who.int/influenza/human_animal_interface/EN_GIP_20140124CumulativeNumberH5N1cases.pdf.

15. de Jong MD, Simmons CP, Thanh TT, et al. Fatal outcome of human influenza A (H5N1) is associated with high viral load and hypercytokinemia. Nat Med 2006;12:1203-7.

16. Perrone LA, Plowden JK, Garcia-Sastre A, Katz JM, Tumpey TM. H5N1 and 1918 pandemic influenza virus infection results in early and excessive infiltration of macrophages and neutrophils in the lungs of mice. PLoS Pathog 2008; 4:e1000115.

17. Szretter KJ, Gangappa S, Lu X, et al. Role of host cytokine responses in the pathogenesis of avian H5N1 influenza viruses in mice. J Virol 2007;81:2736-44.

18. Center for Infectious Disease Research and Policy (CIDRAP). Fatal H5N1 case in Canada is North America's first [Internet]. Minneapolis: CIDRAP; c2014 [cited 2014 Feb 28]. Available from: http://www.cidrap.umn.edu/news-perspective/2014/01/fatal-h5n1-case-canada-north-americas-first.

19. Herfst S, Schrauwen EJ, Linster M, et al. Airborne transmission of influenza A/H5N1 virus between ferrets. Science 2012;336:1534-41.

20. Imai M, Watanabe T, Hatta M, et al. Experimental adaptation of an influenza H5 HA confers respiratory droplet transmission to a reassortant H5 HA/H1N1 virus in ferrets. Nature 2012;486:420-8.

21. Peiris M, Yuen KY, Leung CW, et al. Human infection with influenza H9N2. Lancet 1999;354:916-7.

22. Guan Y, Shortridge KF, Krauss S, Webster RG. Molecular characterization of H9N2 influenza viruses: were they the donors of the “internal” genes of H5N1 viruses in Hong Kong? Proc Natl Acad Sci U S A 1999;96:9363-7.

23. Bao Y, Bolotov P, Dernovoy D, et al. The influenza virus resource at the National Center for Biotechnology Information. J Virol 2008;82:596-601.

24. Wei L, Kao E. Hong Kong sees first case of H9N2 avian flu in four years [Internet]. Hong Kong: South China Morning Post; c2013 [cited 2014 Feb 28]. Available from: http://www.scmp.com/news/hong-kong/article/1393266/86-year-old-man-infected-h9n2-avian-flu.

25. Huang R, Wang AR, Liu ZH, et al. Seroprevalence of avian influenza H9N2 among poultry workers in Shandong Province, China. Eur J Clin Microbiol Infect Dis 2013;32:1347-51.

26. Pawar SD, Tandale BV, Raut CG, et al. Avian influenza H9N2 seroprevalence among poultry workers in Pune, India, 2010. PLoS One 2012;7:e36374.

27. Zhou P, Zhu W, Gu H, et al. Avian influenza H9N2 seroprevalence among swine farm residents in China. J Med Virol 2014;86:597-600.

28. Belser JA, Bridges CB, Katz JM, Tumpey TM. Past, present, and possible future human infection with influenza virus A subtype H7. Emerg Infect Dis 2009;15:859-65.

29. Kurtz J, Manvell RJ, Banks J. Avian influenza virus isolated from a woman with conjunctivitis. Lancet 1996;348:901-2.

30. Centers for Disease Control and Prevention (CDC). Update: influenza activity: United States and worldwide, 2003-04 season, and composition of the 2004-05 influenza vaccine. MMWR Mortal Wkly Rep 2004;53:547-52.

31. Ostrowsky B, Huang A, Terry W, et al. Low pathogenic avian influenza A (H7N2) virus infection in immunocompromised adult, New York, USA, 2003. Emerg Infect Dis 2012; 18:1128-31.

32. Avian influenza A/(H7N2) outbreak in the United Kingdom. Euro Surveill 2007;12:E070531.2.

33. Nguyen-Van-Tam JS, Nair P, Acheson P, et al. Outbreak of low pathogenicity H7N3 avian influenza in UK, including associated case of human conjunctivitis. Euro Surveill 2006; 11:E060504.2.

34. Puzelli S, Di Trani L, Fabiani C, et al. Serological analysis
of serum samples from humans exposed to avian H7 influenza viruses in Italy between 1999 and 2003. J Infect Dis 2005;192:1318-22.

35. Tweed SA, Skowronski DM, David ST, et al. Human illness from avian influenza H7N3, British Columbia. Emerg Infect Dis 2004;10:2196-9.

36. Centers for Disease Control and Prevention (CDC). Notes from the field: Highly pathogenic avian influenza A (H7N3) virus infection in two poultry workers: Jalisco, Mexico, July 2012. MMWR Morb Mortal Wkly Rep 2012;61:726-7.

37. Fouchier RA, Schneeberger PM, Rozendaal FW, et al. Avian influenza A virus (H7N7) associated with human conjunctivitis and a fatal case of acute respiratory distress syndrome. Proc Natl Acad Sci U S A 2004;101:1356-61.

38. Gao R, Cao B, Hu Y, et al. Human infection with a novel avian-origin influenza A (H7N9) virus. N Engl J Med 2013;368:1888-97.

39. Zhang Q, Shi J, Deng G, et al. H7N9 influenza viruses are transmissible in ferrets by respiratory droplet. Science 2013;341:410-4.

40. Watanabe T, Kiso M, Fukuyama S, et al. Characterization of H7N9 influenza A viruses isolated from humans. Nature 2013;501:551-5.

41. Liu D, Shi W, Shi Y, et al. Origin and diversity of novel avian influenza A H7N9 viruses causing human infection: phylogenetic, structural, and coalescent analyses. Lancet 2013;381:1926-32.

42. World Health Organization (WHO). Human infection with avian influenza A(H7N9) virus: update [Internet]. Geneva: WHO; c2014 [cited 2014 Feb 28]. Available from: http://www.who.int/csr/don/2014_02_27b/en/.

43. Yuan J, Zhang L, Kan X, et al. Origin and molecular characteristics of a novel 2013 avian influenza A(H6N1) virus causing human infection in Taiwan. Clin Infect Dis 2013;57:1367-8.

44. Pan American Health Organization (PAHO). Avian influenza virus A(H10N7) circulating among humans in Egypt [Internet]. Washington, DC: PAHO; c2007 [cited 2014 Feb 28]. Available from: http://www1.paho.org/English/AD/DPC/CD/eid-eer-07-may-2004.htm.

45. Arzey GG, Kirkland PD, Arzey KE, et al. Influenza virus A (H10N7) in chickens and poultry abattoir workers, Australia. Emerg Infect Dis 2012;18:814-6.

46. Chen H, Yuan H, Gao R, et al. Clinical and epidemiological characteristics of a fatal case of avian influenza A H10N8 virus infection: a descriptive study. Lancet 2014;383:714-21.

47. Health and Family Planning Commision of Jiangxi Province. Jiangxi 1 new confirmed cases of human infection with avian influenza H10N8 [Internet]. Jiangxi: JXWST; c2014 [cite 2014 Feb 28]. Available from: http://www.jxwst.gov.cn/gzdt/201402/t20140213_308109.htm.

48. Rogers GN, Paulson JC. Receptor determinants of human and animal influenza virus isolates: differences in receptor specificity of the H3 hemagglutinin based on species of origin. Virology 1983;127:361-73.

49. Lipatov AS, Kwon YK, Sarmento LV, et al. Domestic pigs have low susceptibility to H5N1 highly pathogenic avian influenza viruses. PLoS Pathog 2008;4:e1000102.

50. He L, Zhao G, Zhong L, et al. Isolation and characterization of two H5N1 influenza viruses from swine in Jiangsu Province of China. Arch Virol 2013;158:2531-41.

51. Li H, Yu K, Yang H, et al. Isolation and characterization of H5N1 and H9N2 influenza viruses from pigs in China. Chin J Prev Vet Med 2004;26:1-6.

52. Shi WF, Gibbs MJ, Zhang YZ, et al. Genetic analysis of four porcine avian influenza viruses isolated from Shandong, China. Arch Virol 2008;153:211-7.

53. Zhu Q, Yang H, Chen W, et al. A naturally occurring deletion in its NS gene contributes to the attenuation of an H5N1 swine influenza virus in chickens. J Virol 2008;82:220-8.

54. Takano R, Nidom CA, Kiso M, et al. A comparison of the pathogenicity of avian and swine H5N1 influenza viruses in Indonesia. Arch Virol 2009;154:677-81.

55. Nidom CA, Takano R, Yamada S, et al. Influenza A (H5N1) viruses from pigs, Indonesia. Emerg Infect Dis 2010;16:1515-23.

56. Choi YK, Nguyen TD, Ozaki H, et al. Studies of H5N1 influenza virus infection of pigs by using viruses isolated in Vietnam and Thailand in 2004. J Virol 2005;79:10821-5.

57. Peiris JS, Guan Y, Markwell D, Ghose P, Webster RG, Shortridge KF. Cocirculation of avian H9N2 and contemporary “human” H3N2 influenza A viruses in pigs in southeastern China: potential for genetic reassortment? J Virol 2001;75:9679-86.

58. Cong YL, Pu J, Liu QF, et al. Antigenic and genetic characterization of H9N2 swine influenza viruses in China. J Gen Virol 2007;88(Pt 7):2035-41.

59. Yu H, Zhou YJ, Li GX, et al. Genetic diversity of H9N2 influenza viruses from pigs in China: a potential threat to human health? Vet Microbiol 2011;149:254-61.

60. Zhu H, Wang D, Kelvin DJ, et al. Infectivity, transmission,
and pathology of human-isolated H7N9 influenza virus in ferrets and pigs. Science 2013;341:183-6.
61. Nichol KL, Treanor JJ. Vaccines for seasonal and pandemic influenza. J Infect Dis 2006;194 Suppl 2:S111-8.
62. Baz M, Luke CJ, Cheng X, Jin H, Subbarao K. H5N1 vaccines in humans. Virus Res 2013;178:78-98.
63. World Health Organization (WHO). Pandemic influenza preparedness framework for the sharing of influenza viruses and access to vaccines and other benefits [Internet]. Geneva; WHO; c2011 [cited 2014 Feb 28]. Available from: http://apps.who.int/gb/pip/pdf_files/pandemic-influenza-preparedness-en.pdf.
64. Lillie PJ, Berthoud TK, Powell TJ, et al. Preliminary assessment of the efficacy of a T-cell-based influenza vaccine, MVA-NP+M1, in humans. Clin Infect Dis 2012;55:19-25.
65. Zhou J, Wang D, Gao R, et al. Biological features of novel avian influenza A (H7N9) virus. Nature 2013;499:500-3.
66. Klausberger M, Wilde M, Palmberger D, et al. One-shot vaccination with an insect cell-derived low-dose influenza A H7 virus-like particle preparation protects mice against H7N9 challenge. Vaccine 2014;32:355-62.
67. Krammer F, Albrecht RA, Tan GS, et al. Divergent H7 immunogens offer protection from H7N9 virus challenge. J Virol 2014;88:3976-85.
68. Rudenko L, Isakova-Sivak I, Donina S. H7N3 live attenuated influenza vaccine has a potential to protect against new H7N9 avian influenza virus. Vaccine 2013;31:4702-5.
69. Smith GE, Flyer DC, Raghunandan R, et al. Development of influenza H7N9 virus like particle (VLP) vaccine: homologous A/Anhui/1/2013 (H7N9) protection and heterologous A/chicken/Jalisco/CPA1/2012 (H7N3) cross-protection in vaccinated mice challenged with H7N9 virus. Vaccine 2013;31:4305-13.
70. Englund JA. Antiviral therapy of influenza. Semin Pediatr Infect Dis 2002;13:120-8.
71. Wang C, Takeuchi K, Pinto LH, Lamb RA. Ion channel activity of influenza A virus M2 protein: characterization of the amantadine block. J Virol 1993;67:5585-94.
72. Hai R, Schmolke M, Leyva-Grado VH, et al. Influenza A(H7N9) virus gains neuraminidase inhibitor resistance without loss of in vivo virulence or transmissibility. Nat Commun 2013;4:2854.
73. Meng Z, Han R, Hu Y, et al. Possible pandemic threat from new reassortment of influenza A(H7N9) virus in China. Euro Surveill 2014;19:pii 20699.