Fully homodecoupled HSQC spectra can be obtained through the use of a new pulse sequence element, "perfectBIRD". By way of illustration, we show that perfectBIRD decoupling allows one-bond residual dipolar couplings (RDCs), which provide important NMR restraints for structure elucidation, to be measured with outstanding precision, even in methylene groups.

The ever-growing drive in modern chemistry to create increasingly complex systems creates a need for improved analytical tools for their study. While it is usually considered desirable to increase the amount of information provided by a given analytical technique, there are times when information overload means that it is far more useful instead to decrease it. High-resolution solution state NMR spectroscopy, in which narrow chemical shift ranges conspire with extensive scalar coupling to give highly overlapped spectra, is a case in point. Here "pure shift" techniques can be used to suppress the effects of homonuclear couplings, collapsing multiplets to singlets, simplifying spectra and facilitating the extraction of information previously obscured.

Among the different approaches used to achieve broadband homonuclear decoupling of NMR spectra in the real-time direct dimension, in indirect dimensions, or in a pseudo-direct, interferogram, dimension, the bilinear rotation decoupling (BIRD) scheme is particularly suitable for many heteronuclear correlation experiments involving dilute heteronuclei (e.g. natural abundance $^{13}$C). In such cases isotope filtration incurs no extra sensitivity penalty; indeed the signal-to-noise ratio increases in some applications when multiplet structures collapse.

The BIRD element allows control over the effects of vicinal and long-range homonuclear couplings and both one-bond and long-range heteronuclear couplings. However it relies on the one-bond coupling to a dilute heteronucleus to distinguish between homonuclear coupling partners, and hence cannot be used to decouple geminal interactions. The latter not only cause remaining signal multiplicity due to homonuclear interactions to be present, but can also lead to significant spectral distortion. This makes BIRD pure shift methods less attractive for the study of systems containing diastereotopic methylene protons, frequently encountered in organic compounds.

To circumvent this limitation, we have incorporated BIRD decoupling into a modified perfect echo pulse sequence, to form what we refer to as a "perfectBIRD" pulse sequence element. This new sequence element provides full homonuclear broadband decoupling even in the case of diastereotopic methylene protons, at the expense of a doubling of the natural (but not instrumental) linewidth.

Here we illustrate the use of perfectBIRD decoupling in experiments to determine one-bond RDCs. RDCs have proven to be very useful for the structure determination of organic and organometallic compounds. However, RDC analysis in organic compounds is usually prone to be underdetermined, due to the small number of couplings observable. Thus it is of prime importance to obtain all possible information, including the two one-bond RDCs for diastereotopic methylene protons.

For simple AX spin systems the (original) perfect echo pulse sequence (Fig. 1a) refocuses fully both chemical shift and coupling evolution, at time $\tau$, for all $\tau$. Dropping the last pulse of the perfect echo (shown in grey) yields a sequence element which refocuses homonuclear coupling evolution in AX systems at time $\tau$, while introducing a net chemical shift evolution over a period $2\tau$. Differential chemical shift evolution however prohibits the repetitive application of perfect echoes with small $\tau$, recently used in other
methods,9 to achieve decoupling even in complex spin systems. As BIRD pulse elements are able to refocus the effects of weak coupling between protons that are bound to a 13C nucleus directly through one bond (1H–13C) and those that are remotely attached (1H–1H) replacement of the first 180° pulse in the original perfect echo sequence by a BIRDX element (inversion for 1H and 13C) leaves only geminal couplings and strong coupling contributions not refocused at the central 90° pulse in the perfect echo, enabling its use to refocus weak couplings for two geminal coupling partners 1H even if embedded in a complex spin system. To make sure that both JCH evolution and chemical shift evolution of 1H are refocused at time (III), proton inversion pulses are used at the midpoints of periods JCH/2 + τ and JCH/2 + τb. A combination of a broadband proton inversion and a BIRD element (inversion for 1H only) is then used to replace the second 180° pulse of the perfect echo, preserving chemical shift evolution and heteronuclear couplings for 1H while refocusing couplings between the prefocused diastereotopic protons, 1H and 1H, as well as heteronuclear long-range couplings, at the end of the pulse sequence element.

A generalized pulse scheme for CLean In-/Anti-Phase (CLIP/CLAP) HSQC experiments, widely employed in the measurement of one-bond scalar and total couplings, that uses the perfect-BIRD homonuclear decoupling element is given in Fig. 1b. The perfect echo period spans times (I) to (V), with its central mixing pulse positioned at (III). In contrast to the CLIP/CLAP HSQC experiments without homodecoupling, the direct acquisition period normally found after (II) is replaced by the perfect-BIRD element described above.

Construction of a free induction decay with negligible homonuclear coupling modulation is achieved using the interferogram-based approach, recently employed in F2-heterocoupled CLIP/CLAP HSQC spectra with BIRD decoupling in the proton dimension.16 Data are collected between times (IV) and (VI), for time 1/sw2 equal to the time increment in τ2 and centred on the point of full coupling refocusing. Keeping 1/sw2 ≈ 1/(2JH), where JH is of the order of typical proton–proton couplings, restricts data collection to times over which proton–proton coupling evolution can be neglected. A full 3D time domain signal s(t1, t2, t3) is collected, from which a 2D signal s(t1, t2*) is constructed such that t2* = t2 + τ2. This leads to a signal sampled uniformly in t2* for the total time (1/sw2) TD2, where TD2 is the number of points sampled in t2. This data treatment requires 1/sw2 to be an integer multiple of the dwell time used for F2. Construction of the 2D time signal from the 3D dataset is performed conveniently using a Bruker AU program available at http://nmr.chemistry.manchester.ac.uk/. Afterwards s(t1, t2*) can be subjected to double Fourier transformation as usual.

To illustrate the potential of perfect-BIRD decoupled HSQC experiments, we determined one-bond 1H–13C-RDCs for (+)-isopinocampheol (IPC, structure shown in Fig. 2) along the pure shift dimension (F2*) of the experiments. This compound is frequently used for method

Fig. 1 (a) Perfect echo pulse sequence as proposed by Takegoshi, Ogura and Hikichi.6 The pulse shown in grey can be dropped to introduce a net chemical shift evolution during 2τ. (b) A generalized pulse sequence for perfect-BIRD homodecoupled HSQC experiments. Hard 90°-pulses are shown as narrow filled bars and 180°-pulses as wide filled bars, broadband inversion and refocusing pulses used on 13C are shown as open symbols. All experiments shown in the main text use τ2 = τ2 to achieve decoupling for diastereotopic protons. In contrast, setting τ2 = 0 for all τ2 allows the acquisition of clean absorptive doublets even for protons with non-negligible geminal coupling during d2 (see the ESII). The delays d1 and d2 are adjusted to match (4 JCH)−1 and (2 JCH)−1, respectively. In CLIP (CLean In-Phase) experiments d2 = d1 and the pulses marked with an asterisk are used, while in CLAP (CLean Anti-Phase) experiments these pulses are omitted and d2 = δ 10. δ equals the length of the gradients plus a recovery delay, τ4 = (4 sw2)−1 + τ + τ1 − 2 d2 − δ 2, τ3 = (4 sw2)−1 + τ + τ1 − 2 d2 − δ 1, and τ2 = (4 sw2)−1 + τ + τ1 = δ + 350 μs, where τ1 and τ2 are the lengths of the hard 90° pulse on proton and carbon respectively and τ3 is the length of the broadband inversion pulse on 13C. G2 and G3 are set according to the ratio of gyromagnetic ratios and G2 is inverted in alternating experiments to achieve the frequency sign encoding along τ1 according to the echo/antiecho procedure. The pulse phases used are: Φ1 = 1, Φ2 = 0, Φ3 = 0 0 0 0 2 2 2 2, Φ4 = 1 1 3 3, Φmuc = 0 2 0 2 2 2 0 2 0.

Fig. 2 F2-heterocoupled CLIP HSQC spectra without homonuclear decoupling (black), and with BIRD (blue) and with perfect-BIRD (red) homonuclear decoupling during acquisition, collected for (+)-IPC in isotropic CD2Cl2 solution at 600 MHz proton frequency. Experiment durations were 10.5 min, 7.1 h and 9.4 h, respectively. The structure of the analyte is shown in the figure, with the numbering used. The corresponding proton spectrum is displayed at the top. For selected protons, traces along the proton dimension are shown. The decoupled spectra are shifted in the carbon dimension for easier comparison.
BIRD particularly attractive for measurements on aligned samples. The constant-time approach necessar-
yly limits the range of couplings accessible, while the perfectBIRD method can accommodate a wide range of \( J_{HH} \) making perfectBIRD particularly attractive for measurements on aligned samples.

Fig. 3 also illustrates a limitation of the perfectBIRD decoupling element: even on anisotropic samples, the triplets observed using BIRD decoupling are only partially collapsed using perfectBIRD decoupling (see inset 10). In isotropic solution this problem does not arise.

In small organic molecules, strong coupling effects are quite common, though not present in the case studied. Neither the perfect echo nor the BIRD element will fully refocus the effects of strong coupling, and complete decoupling of strongly coupled protons remains an unsolved challenge in pure shift NMR (as in many other methods). The precise measurement of RDCs from strongly coupled spins is an issue best addressed using specialized approaches as illustrated in the ESI, strong coupling can be identified in homodecoupled spectra through characteristic changes in signal shapes.

The spectra shown for (+)-IPC, and additional experiments on chloroform, representing a simple AX test system, were used to test the influence of perfectBIRD homonuclear decoupling on the accuracy and precision of coupling constant measurements (see ESI†). Considering accuracy first, under the experimental conditions used, systematic errors in homodecoupled measurements of coupling constants were less than 0.05 Hz, greater than those for measurements by some conventional methods but negligible in the context of RDC measurements that typically have uncertainties of several tenths of a Hz. In contrast, the precision of \( J_{CH} \) measurements was significantly improved by homodecoupling in
The practical example of (+)-IPC, because of the simplification of line shapes and the avoidance of signal overlap caused by homonuclear couplings.

The confidence intervals shown in Tables 1 and 2 have two contributions: a very conservative estimate of the possible effects of the systematic errors noted (±0.1 Hz, double the observed uncertainty range) and the results of confidence interval estimation performed according to the procedure of Kummerlöwe et al. In many cases these confidence intervals show a significant improvement with BIRD and perfectBIRD, particularly for methylene signals in the latter case. Couplings extracted from the CLAP HSQC spectra are given in Table S5 (ESI). From the values obtained we conclude that homonuclear decoupling can indeed improve the precision of coupling constant measurements in the high-resolution proton dimension, which is particularly beneficial for RDC-based structure analysis in the case of diastereotopic methylene protons.

In this communication, we have introduced a homonuclear decoupling element, based on the BIRD and perfect echo techniques, which is able to collapse splittings due to geminal couplings between diastereotopic methylene protons. Pure shift $f_2$-heteronuclear HSQC spectra of exceptional quality can be obtained, allowing highly precise measurements of one-bond couplings in the high-resolution proton dimension, even in weakly aligned media. We expect that the extended measurement times needed for these experiments will prove to be well justified, by the higher precision of the coupling constants extracted and the improved ease of analysis, when complex structures are to be solved. Modifications of the technique that also achieve heteronuclear decoupling in the high-resolution dimension are under development, and could be used to collect HSQC spectra with full homo- and heteronuclear decouplings in both dimensions as well as very high resolution in the proton dimension.

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Table 1 Scalar couplings extracted from the CLIP HSQC spectra of (+)-IPC in an isotropic CD$_2$Cl$_2$ solution shown in Fig. 2.

| No decoupling | BIRD decoupling | PerfectBIRD decoupling |
|---------------|-----------------|------------------------|
| $J_{CH}$ [Hz] | $J_{CH}$ [Hz]   | $J_{CH}$ [Hz]          |
| FID-res:      |                 |                        |
| 1             | 1.02 Hz         | 1.02 Hz                |
| 10            | 0.76 Hz         | 0.76 Hz                |

Table 2 Total couplings extracted from the CLIP HSQC spectra of (+)-IPC in an isotropic CD$_2$Cl$_2$/PBDG solution ($\Delta_{CH} = 107.6$ Hz) shown in Fig. S2 (ESI).

| No decoupling | BIRD decoupling | PerfectBIRD decoupling |
|---------------|-----------------|------------------------|
| $J_{CH}$ [Hz] | $J_{CH}$ [Hz]   | $J_{CH}$ [Hz]          |
| FID-res:      |                 |                        |
| 1             | 1.02 Hz         | 1.02 Hz                |
| 10            | 0.76 Hz         | 0.76 Hz                |

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