Abstract: Graft versus host disease (GVHD) remains one of the leading causes of morbidity and mortality associated with conventional allogeneic hematopoietic stem cell transplantation (HCT). The use of T-cell depletion significantly reduces this complication. Recent prospective and retrospective data suggest that, in patients with AML in first complete remission, CD34+ selected grafts afford overall and relapse-free survival comparable to those observed in recipients of conventional grafts, while significantly decreasing GVHD. In addition, CD34+ selected grafts allow older patients, and those with medical comorbidities or with only HLA-mismatched donors to successfully undergo transplantation. Prospective data are needed to further define which groups of patients with AML are most likely to benefit from CD34+ selected grafts. Here we review the history of T-cell depletion in AML, and techniques used. We then summarize the contemporary literature using CD34+ selection in recipients of matched or partially mismatched donors (7/8 or 8/8 HLA-matched), and provide a summary of the risks and benefits of using T-cell depletion.

Keywords: AML; CD34+ selection; T-cell depletion; graft-versus-host disease; hematopoietic stem cell transplantation
1. Introduction

Cytogenetic risk stratification in acute myelogenous leukemia (AML) allows clinicians to determine which patients are most likely to benefit from allogeneic hematopoietic stem cell transplantation (HCT), with evidence to support a survival advantage in patients with intermediate or high-risk cytogenetics [1]. In addition to more accurate patient selection based on cytogenetic risk factors, over the past decades, transplantation outcomes have also improved as a result of more accurate patient selection tools, such as the HCT Sorror comorbidity index [2], improvements in HLA matching techniques and supportive care.

Despite these improvements, graft-versus-host disease (GVHD) remains a leading cause of post-transplant morbidity and mortality. A variety of T-cell depletion (TCD) techniques have been developed and used over the years in an effort to reduce transplant-related mortality (TRM) due to GVHD (Table 1). In this chapter we will focus on outcomes of HCT using T cell depletion for the treatment of AML in recipients of matched or partially mismatched donors (7/8 or 8/8 HLA-matched), with a primary focus on ex vivo CD34+ selection of the graft. Other methods of T-cell depletion will be mentioned for historic context only.

2. T-Cell Depletion Techniques

The goal of T-cell depleting a graft is to reduce GVHD while maintaining the graft-versus-leukemia or lymphoma (GVL) effect. A variety of TCD techniques have been used with mixed results. When reviewing reports utilizing TCD for transplantation it is critical to determine the following: (1) Which technique is being used? (2) Which cell population is being removed (i.e., T, B, NK or all non-hematopoietic cells) and to what extent? Different techniques lead to both quantitative and qualitative differences in the cells being depleted with important clinical implications; (3) Is post-transplant GVHD prophylaxis utilized? and (4) What is the graft source and what is the degree of HLA matching? All of these factors significantly affect outcomes and therefore must be considered when interpreting published reports.

Early TCD techniques differed in the use of negative vs. positive selection. Negative selection can be achieved either through physical methods such as counterflow elutriation [3–5] or soybean lectin agglutination (SBA) and sheep red blood cell (sRBC)-rosette depletion (E-rosetting) [6–10], or immunological methods using monoclonal antibodies [5,11–29]. Monoclonal antibodies can be used with or without complement, or conjugated to toxins. Antibodies vary in their specificities, which can be narrow, such as T10B9 targeting the α/β T cell receptor (TCR), or broad, such as combination of antibodies targeting CD2, CD4 and CD8 [30].
Table 1. Results of T-cell depletion (TCD)-PBSCT in patients with acute myelogenous leukemia (AML) 1.

| Method                  | Number of Patients | Patients With AML | Donor                  | Degree of Depletion | GVHD Prophylaxis                  | Acute GVHD | Graft Failure | EFS/DFS 2 | OS 2 | Reference |
|-------------------------|--------------------|-------------------|------------------------|---------------------|-----------------------------------|------------|---------------|-----------|------|-----------|
| CD34+                   | 50                 | 29                | HLA-MRD               | NR                  | Cyclosporine or Cyclosporine + Steroids | 16%        | 0             | DFS 65%   | Not reported | [31]     |
| CD34+E−                 | 52                 | 21                | HLA-MRD               | 5 logs              | None                             | 8%         | 0             | NR        | 17% 1 year   | [32]     |
| CD34+E−                 | 29                 | 16                | HLA-MUD or HLA-MMUD   | 5 logs              | None                             | 9%         | 3%            | 57% at 4 years | 59% at 4 years | [33]     |
| CD34+                   | 47                 | 47                | HLA-MRD               | 4.9 logs            | None                             | 22.7%      | 0             | EFS 63% at 4 years | 71% 4 years   | [34]     |
| CD3/CD19 depletion      | 29                 | 16                | Haplo                  | 4.4 logs            | None                             | 48%        | 0             | 35% at 1 year | 31% at 241 days | [35]     |
| CD34+E− or CD34+        | 115                | 115               | HLA-MUD or MRD        | NR                  | None                             | 5%         | RFS 58% at 3 years | 57% at 3 years | [36]     |

1 Abbreviations: CD34+: CD34-selection; CD34+E−: CD34-selection and E-rosetting; NR: not reported; 2 DFS and OS are reported for the entire patient population included in the studies.
Currently, the most common method for T cell depletion relies on positive selection of CD34+ hematopoietic stem cells from the graft [32–34,36,37]. CD34+ selection of peripheral blood stem cells (PBSCs) was performed in initial studies with the ISOLEX 300i magnetic cell selection system (Baxter, Deerfield, IL, USA), followed by E-rosetting [32,33,37]. The ISOLEX device is no longer being manufactured, and the most commonly used method in current studies uses immunomagnetic beads with the CliniMACS CD34 Reagent System (Miltenyi Biotech, Gladbach, Germany) for CD34+ selection [31,34,37]. The CliniMACS system can also be used to negatively select grafts through depletion of CD3+ and CD19+ cells or depletion of TCRαβ+ T cells [35,38–41]. The two CD34+ selection methods differ in the degree of TCD; for example, the CliniMACS CD34 Reagent System can achieve a 5-log reduction in T cells, whereas the ISOLEX 300i system achieves a 3.5-log reduction, requiring additional T-cell depletion through E-rosetting.

The cell dose and graft source also impact outcomes. In recipients of T-cell depleted marrow grafts from HLA-identical donors, the risk of GVHD was shown to increase if the graft contained >1 × 10^5 T cells/kg [42]. Differences between TCD methods can have a significant impact on clinical outcomes, including the risk of graft failure, GVHD and relapse.

3. Outcomes in AML with T-Cell Depletion

One of the primary benefits of allogeneic HCT is derived from the GVL effect, driven by the recognition of tumor cells by donor T cells. Therefore, a concern in the utilization of TCD is the potentially negative impact on relapse resulting from reduced T-cell doses included in the graft. However, it is clear that certain diseases rely more on the GVL effect than others. For example, early studies with TCD in chronic myelogenous leukemia (CML) were associated with a significantly increased risk of relapse. In a retrospective study of 46 patients who received TCD grafts from HLA-identical siblings, relapse at 3-years was significantly higher in the TCD group (62% vs. 24%, p = 0.0003). However, a significant proportion of these patients were then salvaged with donor lymphocyte infusion (DLI), supporting the role of GVL for this disease [43].

On the other hand, studies utilizing TCD in AML and ALL have shown favorable outcomes in recipients of 7/8 or 8/8 HLA matched related or unrelated donors that are comparable to those seen in unmodified, conventional grafts, calling into question the contribution of GVL in those diseases [34,36,44,45].

4. TCD in AML—Early Studies

Studies using TCD done in the 1980’s and 1990’s mostly used bone marrow from sibling donors as the graft source and used a variety of techniques (lectin separation, elutriation, E-rosettes, antibodies against T and NK cells, antibodies with or without complement). These techniques were associated with a 1–2 log reduction in T cells in the graft, and patients were often given post-transplant cyclosporine for additional GVHD prophylaxis. These studies were also associated with increased graft failure [8,46]. The addition of antithymocyte globulin (ATG) and thiotepa to the conditioning regimens, which had traditionally included cyclophosphamide and total body irradiation (TBI) or busulfan, decreased the rate of graft failure and eliminated the need for post transplant GVHD prophylaxis. Favorable results were reported in recipients of HLA-identical donors using this approach along with sequential soybean
lectin agglutination and sheep red blood cell-rosette depletion. A study by Papadopoulos et al. included 31 patients with AML in CR1 or CR2 and showed a disease-free survival (DFS) at 4 years of 70% with no GVHD or graft rejection [9]. Similar results were reported by Aversa et al. using the same regimen in 54 patients with acute leukemia, including 2 with HLA-DR mismatched donors [10]. No GVHD or graft rejection was observed and the event-free survival at 4.9 years was 74% in 30 patients with AML.

These early studies demonstrated the feasibility of TCD, and overcame graft rejection with the utilization of ATG, at the expense of added immunosuppression and ensuing delayed immune recovery.

5. TCD in AML—Contemporary Studies

Contemporary studies have utilized two main approaches of TCD by CD34+ selection described above, either the ISOLEX 300i Magnetic Cell Separator followed by sRBC-rosette depletion or, more recently, the Miltenyi CliniMACS CD34 Reagent System. In 2011, Devine et al. [34] reported the results of a Blood and Marrow Clinical Trials Network study (BMT CTN 0303) utilizing CD34+ selection with the Miltenyi CliniMACS CD34 Reagent System. The study included 44 patients with AML in CR1 or CR2 (excluding patients with t(15;17), and core binding factor leukemia) who received T cell depleted PBSCT from HLA-identical siblings after conditioning with hyperfractionated TBI (HFTBI), thiotepa, cyclophosphamide and ATG. No additional GVHD prophylaxis was given. All patients engrafted; the incidence of aGVHD (grades II–IV) was 22.7% and extensive cGVHD was 6.8% at 2 years. The relapse rate at three years for patients in CR1 was 17.4%, with a DFS for all patients at 6 months of 82% and a 2-year OS of 60%. Rates of infection were comparable to other studies; however EBV reactivation occurred in 18% of patients leading to 1 death from Epstein-Barr Virus (EBV) post-transplant lymphoproliferative disease (PTLD) [36,47,48]. In a second report, the outcomes of patients from the BMT CTN 0303 study were compared to a similar cohort of patients (AML in either CR1 or CR2, PBSCT from HLA-identical siblings) who received a conventional HCT on BMT CTN 0101 study (a study comparing fluconazole with voriconazole as antifungal prophylaxis after HCT) [44]. There were no differences in leukemia relapse (23% vs. 27% in the TCD and conventional graft groups, respectively) and 2-year OS (65% vs. 59% in the TCD and conventional graft groups, respectively). However rates of GVHD were higher in the conventional graft group (aGVHD 23% vs. 39%, p = 0.07 and cGVHD 19% vs. 50%, p < 0.001).

More recently, a retrospective study compared the use of TCD HCT to conventional grafts in patients with AML in CR1 by examining outcomes of 115 patients who received TCD grafts at Memorial Sloan Kettering Cancer Center (MSKCC) with a cohort of 181 patients at MD Anderson Cancer Center (MDACC) [36]. A hundred and seven patients in the MSKCC cohort received PBSC grafts, including 85 that were CD34-selected with the ISOLEX 300i Magnetic Cell Separator followed by sRBC-rosette depletion, and 22 with the Miltenyi CliniMACS CD34 Reagent System. Patients at both centers received myeloablative conditioning (MAC) and both cohorts included recipients of matched related, matched unrelated and mismatched donors. Patients at MSKCC were more likely to be recipients of a mismatched graft (27% vs. 14%, p < 0.001). Patients at MSKCC did not receive additional GVHD prophylaxis. Patients at MDACC received tacrolimus and mini-methotrexate for GVHD prophylaxis, and ATG for HLA-mismatched donors. There were no significant differences in
the rate of relapse at 3 years between groups (18% vs. 25%, in the TCD vs. conventional grafts, respectively, \( p = 0.3 \)). However, rates of GVHD were significantly lower in the TCD group (5% vs. 18% for aGVHD, \( p = 0.005 \), and 13% vs. 53% for cGVHD, \( p < 0.001 \)).

Although contemporary studies that compare outcomes of conventional to TCD grafts are retrospective, the results suggest that TCD transplants offer similar DFS and OS with significantly lower rates of GVHD.

As noted above additional TCD approaches are being investigated beyond CD34 selection [35,38–41]. The potential advantages of negative selection by depletion of CD3+/CD19+ or TCRαβ+ T cells include the presence of additional cells in the graft such as natural killer (NK) cells or TCRγδ+ T cells, which may play a role in relapse or infection prevention. To date, the published studies of these approaches have been in recipients of haplo-identical grafts and there has been limited data on patients with AML. In a study by Bethge et al. [35], EFS was 35% at one year in 16 patients with AML who received a CD3/CD19 depleted transplant from a haplo-identical donor.

Finally, an alternative GVHD prophylaxis approach that will be compared to CD34-selection in an upcoming phase 3 trial (BMT CTN 1301) relies on the use of post-transplant high dose cyclophosphamide after a T-replete bone marrow graft from a matched donor [49–51]. In a recently published study using this approach in 138 patients with AML, the 3-year DFS and OS were 43% (95% CI, 35% to 52%) and 53% (95% CI, 45% to 62%), respectively [50]. DFS (48% vs. 29% at 3 years) and OS (55% vs. 50% at 3 years) were higher in patients with AML in morphologic CR compared to those with active disease. The approach was associated with low rates of grade III to IV acute GVHD (11% at 100 days) and chronic GVHD (13% at 2 years).

6. Impact of T-Cell Depletion on Engraftment and Immune Reconstitution

Hematopoietic stem cell transplantation, regardless of donor source and manipulation, is associated with significant and prolonged immunosuppression and risk of severe and fatal infections [48,52], disease relapse and secondary malignancies [20,53]. The use of TCD, whether in vivo with agents such as alemtuzumab or ATG, or ex vivo with T-cell depletion considerably affects immune recovery. In early studies comparing immune reconstitution in TCD and unmodified grafts the rate of CD3+, CD4+ and CD8+ T cell reconstitution was significantly delayed in TCD recipients, correlating with increased risk of infections, including EBV-PTLD [48,54]. T-cell receptor (TCR) studies using 5′-RACE PCR with deep sequencing have confirmed these findings by showing more rapid recovery of TCR diversity in conventional graft recipients compared to TCD grafts [55]. Lower T-cell levels result from decreased thymic output, which can be quantified via measurements of T-cell receptor excision circles (TRECs). Studies have shown that older patients and recipients of TCD grafts have lower TRECs than unmodified graft recipients; however this difference abates beyond 9 months [56].

ATG plays an important role in TCD by reducing the risk of graft rejection. However it is associated with delayed immune recovery and an increased risk of opportunistic infections (OI), with approximately 15% of the patients in early TCD studies dying from OIs [9,10]. Furthermore, a study of immune reconstitution in patients receiving TCD grafts with or without ATG found delayed immune reconstitution with ATG, which was associated with increased OIs [54]. Although studies of TCD performed without ATG by substituting cyclophosphamide with fludarabine have demonstrated
durable engraftment in recipients of matched related donors, there did not appear to be a significant effect on immune recovery or the risk of OIs [32].

It is important to note however that, in addition to age and TCD, the presence of GVHD also significantly hampers immune reconstitution via direct effects on the thymus [57–59], as well as the immunosuppressive drugs required for treatment of GVHD [60–64]. Although TCD impacts immune reconstitution leading to higher infection related deaths, GVHD in conventional grafts similarly leads to increased mortality. This is a potential explanation for the similar RFS and OS outcomes observed in patients with AML in the MSKCC/MDACC retrospective study and the BMT CTN study [36,44].

7. Strategies to Enhance Immune Recovery Post HCT

HCT affords a curative treatment option to many patients with otherwise incurable malignancies. However, the benefit of this therapy comes at the risk of significant complications, including infection, GVHD, relapse and secondary malignancies [20,48,52,53,65,66]. The rationale for TCD is to mitigate GVHD while preserving the benefit of GVL. TCD and HCT in general, are associated with prolonged immunosuppression. Therefore, strategies to optimize post transplant immunity, enhance GVL, while minimizing infectious complications are needed.

The addition of T cells post transplantation represents one such strategy. In one recent study, 19 pediatric patients (13 transplanted for malignant disease) received CD34+ selected matched unrelated donor (MUD) HCT with CD3+ T cells added back, at a dose of 1.0–2.5 × 10^5 CD3+/kg, and tacrolimus for GVHD prevention [67]. Rates of aGVHD, cGVHD and extensive cGVHD were 15.8%, 23.3% and 0%, respectively, which are low compared to conventional HCT. All patients on this study had neutrophil engraftment, and infection-related mortality at one year was 5%–6%, showing the feasibility of this approach. This approach has also been used in recipients of CD34-selected haplo-identical grafts [68]. The same group previously reported low rates of acute and chronic GVHD in a retrospective series of 16 patients who received DLI (up to 6 × 10^4 CD3+/kg) from haplo-identical donors to enhance immune recovery and/or treat infections [69]. One ongoing trial is evaluating the effect of serial DLI post TCD in patients with advanced multiple myeloma (NCT01131169).

Another strategy to boost immune reconstitution post-transplant is the use of Keratinocyte Growth Factor (KGF). KGF has been shown in pre-clinical models to play an important role in T cell homeostasis and immune recovery, as well as in thymic regeneration after radiation injury [70–72]. Based on these data, KGF along with sex steroid ablation is being studied in an ongoing phase II clinical trial (NCT01746849).

We recently published the results of a phase I study using recombinant human IL-7 (rhIL-7, CYT107, Cytheris) in recipients of TCD HCT and demonstrated enhanced immune recovery, with significant increases in CD4+ and CD8+ T cells along with increased T cell function, without causing significant GVHD or other serious toxicity [73].

8. Benefits of TCD HCT in AML

Although reduced intensity conditioning (RIC) regimens allow older patients to undergo HCT, they are associated with higher relapse rates in AML [74,75]. However, the combined toxicity of GVHD prophylaxis that usually includes a calcineurin inhibitor (CNI) and methotrexate with myeloablative
conditioning (MAC), makes this approach prohibitive in older patients. Unlike conventional grafts, CD34+ selected grafts do not require post-transplant GVHD prophylaxis, and as a result, older patients can be treated with MAC. In addition, patients with renal insufficiency can also successfully undergo transplantation by avoiding the use of CNIs.

The use of CD34+ selected grafts is associated with significant reductions in both acute and chronic GVHD. In addition to the obvious advantage of lowering GVHD, patients without fully matched donors are also able to undergo transplantation, therefore expanding the pool of potential donors.

Finally, CD34+ selected grafts are an ideal platform for post-transplant immunotherapy with adoptive cell therapy targeting both minimal residual disease and viral reactivation by CMV and EBV, among others [76–79]. The administration T cells specific for tumor or viral antigens post-transplant has the potential advantage of overcoming any loss of GVL or increased infectious risk associated with TCD without affecting the benefit of reduced GVHD.

9. Conclusions

After three decades of investigation, it is reasonable to consider CD34+ selected allografts for patients with AML in CR1 based on prospective data [34], and well conducted retrospective studies [36,44]. These contemporary studies are significantly more homogeneous in their methodology and patient inclusion than prior studies, mostly using the CliniMACS CD34 Reagent System for CD34+ selection, and reporting consistent favorable outcomes for patients with AML in CR. The use of CD34+ selected grafts overcomes the morbidity and mortality associated with GVHD, a significant contributor of transplant-related complications, without compromising the benefit of transplantation and affording the same overall survival as conventional transplantation.

TCD represents an important step in graft manipulation, allowing older patients, and those with comorbidities to successfully undergo transplantation. Ongoing research aims to continue to decrease morbidity and mortality associated with transplantation by improving immune reconstitution and the GVL effect.

As mentioned above, an ongoing national phase 3 trial will compare TCD with the CliniMACS CD34 Reagent System to post-transplant cyclophosphamide [49], and a control arm (tacrolimus and methotrexate) in patients with acute leukemia and MDS who are eligible for a MAC transplant from a matched related or unrelated donor (BMT CTN 1301).

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Author Contributions

Gabriela Soriano Hobbs and Miguel-Angel Perales wrote reviewed the literature, wrote the manuscript and edited its contents.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Koreth, J.; Schlenk, R.; Kopecky, K.J.; Honda, S.; Sierra, J.; Djulbegovic, B.J.; Wadleigh, M.; DeAngelo, D.J.; Stone, R.M.; Sakamaki, H.; et al. Allogeneic stem cell transplantation for acute myeloid leukemia in first complete remission: Systematic review and meta-analysis of prospective clinical trials. *JAMA* 2009, 301, 2349–2361.

2. Sorror, M.L.; Maris, M.B.; Storb, R.; Baron, F.; Sandmaier, B.M.; Maloney, D.G.; Storer, B. Hematopoietic cell transplantation (HCT)-specific comorbidity index: A new tool for risk assessment before allogeneic HCT. *Blood* 2005, 106, 2912–2919.

3. De Witte, T.; Hoogenhout, J.; de Pauw, B.; Holdrinet, R.; Janssen, J.; Wessels, J.; van Daal, W.; Hustinx, T.; Haanen, C. Depletion of donor lymphocytes by counterflow centrifugation successfully prevents acute graft-versus-host disease in matched allogeneic marrow transplantation. *Blood* 1986, 67, 1302–1308.

4. Wagner, J.E.; Donnenberg, A.D.; Noga, S.J.; Cremo, C.A.; Gao, I.K.; Yin, H.J.; Vogelsang, G.B.; Rowley, S.; Saral, R.; Santos, G.W. Lymphocyte depletion of donor bone marrow by counterflow centrifugal elutriation: Results of a phase I clinical trial. *Blood* 1988, 72, 1168–1176.

5. Wagner, J.E.; Thompson, J.S.; Carter, S.L.; Kernan, N.A. Effect of graft-versus-host disease prophylaxis on 3-year disease-free survival in recipients of unrelated donor bone marrow (T-cell Depletion Trial): A multi-centre, randomised phase II-III trial. *Lancet* 2005, 366, 733–741.

6. Reisner, Y.; Kapoor, N.; Kirkpatrick, D.; Pollack, M.S.; Cunningham-Rundles, S.; Dupont, B.; Hodes, M.Z.; Good, R.A.; O’Reilly, R.J. Transplantation for severe combined immunodeficiency with HLA-A,B,D,DR incompatible parental marrow cells fractionated by soybean agglutinin and sheep red blood cells. *Blood* 1983, 61, 341–348.

7. Reisner, Y.; Kapoor, N.; Kirkpatrick, D.; Pollack, M.S.; Dupont, B.; Good, R.A.; O’Reilly, R.J. Transplantation for acute leukaemia with HLA-A and B nonidentical parental marrow cells fractionated with soybean agglutinin and sheep red blood cells. *Lancet* 1981, 2, 327–331.

8. Young, J.W.; Papadopoulos, E.B.; Cunningham, I.; Castro-Malaspina, H.; Flomenberg, N.; Carabasi, M.H.; Gulati, S.C.; Brochstein, J.A.; Heller, G.; Black, P.; et al. T-cell-depleted allogeneic bone marrow transplantation in adults with acute nonlymphocytic leukemia in first remission. *Blood* 1992, 79, 3380–3387.

9. Papadopoulos, E.B.; Carabasi, M.H.; Castro-Malaspina, H.; Childs, B.H.; Mackinnon, S.; Boulad, F.; Gillio, A.P.; Kernan, N.A.; Small, T.N.; Szabolcs, P.; et al. T-cell-depleted allogeneic bone marrow transplantation as postremission therapy for acute myelogenous leukemia: Freedom from relapse in the absence of graft-versus-host disease. *Blood* 1998, 91, 1083–1090.
10. Aversa, F.; Terenzi, A.; Carotti, A.; Felicini, R.; Jacucci, R.; Zei, T.; Latini, P.; Aristei, C.; Santucci, A.; Martelli, M.P.; et al. Improved outcome with T-cell-depleted bone marrow transplantation for acute leukemia. J. Clin. Oncol. 1999, 17, 1545–1550.

11. Prentice, H.G.; Blacklock, H.A.; Janossy, G.; Bradstock, K.F.; Skeggs, D.; Goldstein, G.; Hoffbrand, A.V. Use of anti-T-cell monoclonal antibody OKT3 to prevent acute graft-versus-host disease in allogeneic bone-marrow transplantation for acute leukaemia. Lancet 1982, 1, 700–703.

12. Filipovich, A.H.; McGlave, P.B.; Ramsay, N.K.; Goldstein, G.; Warkentin, P.I.; Kesey, J.H. Pretreatment of donor bone marrow with monoclonal antibody OKT3 for prevention of acute graft-versus-host disease in allogeneic histocompatible bone-marrow transplantation. Lancet 1982, 1, 1266–1269.

13. Prentice, H.G.; Blacklock, H.A.; Janossy, G.; Gilmore, M.J.; Price-Jones, L.; Tidman, N.; Trejdosiewicz, L.K.; Skeggs, D.B.; Panjwani, D.; Ball, S.; et al. Depletion of T lymphocytes in donor marrow prevents significant graft-versus-host disease in matched allogeneic leukaemic marrow transplant recipients. Lancet 1984, 1, 472–476.

14. Martin, P.J.; Hansen, J.A.; Thomas, E.D. Preincubation of donor bone marrow cells with a combination of murine monoclonal anti-T-cell antibodies without complement does not prevent graft-versus-host disease after allogeneic marrow transplantation. J. Clin. Immunol. 1984, 4, 18–22.

15. Martin, P.J.; Hansen, J.A.; Buckner, C.D.; Sanders, J.E.; Deeg, H.J.; Stewart, P.; Appelbaum, F.R.; Clift, R.; Fefer, A.; Witherspoon, R.P.; et al. Effects of in vitro depletion of T cells in HLA-identical allogeneic marrow grafts. Blood 1985, 66, 664–672.

16. Herve, P.; Flesch, M.; Cahn, J.Y.; Racadot, E.; Plouvier, E.; Lamy, B.; Rozenbaum, A.; Noir, A.; Des Floris, R.L.; Peters, A. Removal of marrow T cells with OKT3-OKT11 monoclonal antibodies and complement to prevent acute graft-versus-host disease. A pilot study in ten patients. Transplantation 1985, 39, 138–143.

17. Trigg, M.E.; Billing, R.; Sondel, P.M.; Extent, R.; Hong, R.; Bozdech, M.J.; Horowitz, S.D.; Finlay, J.L.; Moen, R.; Longo, W.; et al. Clinical trial depleting T lymphocytes from donor marrow for matched and mismatched allogeneic bone marrow transplants. Cancer Treat. Rep. 1985, 69, 377–386.

18. Mitsuyasu, R.T.; Champlin, R.E.; Gale, R.P.; Ho, W.G.; Lenarsky, C.; Winston, D.; Selch, M.; Elashoff, R.; Giorgi, J.V.; Wells, J.; et al. Treatment of donor bone marrow with monoclonal anti-T-cell antibody and complement for the prevention of graft-versus-host disease. A prospective, randomized, double-blind trial. Ann. Intern. Med. 1986, 105, 20–26.

19. Patterson, J.; Prentice, H.G.; Brenner, M.K.; Gilmore, M.; Janossy, G.; Ivory, K.; Skeggs, D.; Morgan, H.; Lord, J.; Blacklock, H.A.; et al. Graft rejection following HLA matched T-lymphocyte depleted bone marrow transplantation. Br. J. Haematol. 1986, 63, 221–230.

20. Maraninchi, D.; Gluckman, E.; Blaise, D.; Guyotat, D.; Rio, B.; Pico, J.L.; Leblond, V.; Michallet, M.; Dreyfus, F.; Ifrah, N.; et al. Impact of T-cell depletion on outcome of allogeneic bone-marrow transplantation for standard-risk leukaemias. Lancet 1987, 2, 175–178.
21. Cahn, J.Y.; Herve, P.; Flesch, M.; Plouvier, E.; Racadot, E.; Vuillier, J.; Montcuquet, P.; Noir, A.; Rozenbaum, A.; Leconte des Floris, R. Marrow transplantation from HLA non-identical family donors for the treatment of leukaemia: A pilot study of 15 patients using additional immunosuppression and T-cell depletion. Br. J. Haematol. 1988, 69, 345–349.

22. Martin, P.J.; Hansen, J.A.; Torok-Storb, B.; Moretti, L.; Press, O.; Storb, R.; Thomas, E.D.; Weiden, P.L.; Vitetta, E.S. Effects of treating marrow with a CD3-specific immunotoxin for prevention of acute graft-versus-host disease. Bone Marrow Transplant. 1988, 3, 437–444.

23. Laurent, G.; Maraninchi, D.; Gluckman, E.; Vernant, J.P.; Deroq, J.M.; Gaspard, M.H.; Rio, B.; Michaelet, M.; Reiffers, J.; Dreyfus, F.; et al. Donor bone marrow treatment with T101 Fab fragment-ricin A-chain immunotoxin prevents graft-versus-host disease. Bone Marrow Transplant. 1989, 4, 367–371.

24. Filipovich, A.H.; Vallera, D.; McGlave, P.; Polich, D.; Gajl-Peczalska, K.; Haake, R.; Lasky, L.; Blazar, B.; Ramsay, N.K.; Kersey, J.; et al. T cell depletion with anti-CD5 immunotoxin in histocompatible bone marrow transplantation. The correlation between residual CD5 negative T cells and subsequent graft-versus-host disease. Transplantation 1990, 50, 410–415.

25. Antin, J.H.; Bierer, B.E.; Smith, B.R.; Ferrara, J.; Guinan, E.C.; Sieff, C.; Golan, D.E.; Macklis, R.M.; Tarbell, N.J.; Lynch, E.; et al. Selective depletion of bone marrow T lymphocytes with anti-CD5 monoclonal antibodies: Effective prophylaxis for graft-versus-host disease in patients with hematologic malignancies. Blood 1991, 78, 2139–2149.

26. Soiffer, R.J.; Fairclough, D.; Robertson, M.; Alyea, E.; Anderson, K.; Freedman, A.; Bartlett-Pandite, L.; Fisher, D.; Schlossman, R.L.; Stone, R.; et al. CD6-depleted allogeneic bone marrow transplantation for acute leukemia in first complete remission. Blood 1997, 89, 3039–3047.

27. Soiffer, R.J.; Freedman, A.S.; Neuberg, D.; Fisher, D.C.; Alyea, E.P.; Gribben, J.; Schlossman, R.L.; Bartlett-Pandite, L.; Kuhlman, C.; Murray, C.; Freeman, A.; et al. CD6+ T cell-depleted allogeneic bone marrow transplantation for non-Hodgkin’s lymphoma. Bone Marrow Transplant. 1998, 21, 1177–1181.

28. Alyea, E.P.; Weller, E.; Fisher, D.C.; Freedman, A.S.; Gribben, J.G.; Lee, S.; Schlossman, R.L.; Stone, R.M.; Friedberg, J.; DeAngelo, D.; et al. Comparable outcome with T-cell-depleted unrelated-donor versus related-donor allogeneic bone marrow transplantation. Biol. Blood Marrow Transplant. 2002, 8, 601–607.

29. Lee, S.J.; Zahrieh, D.; Alyea, E.P.; Weller, E.; Ho, V.T.; Antin, J.H.; Soiffer, R.J. Comparison of T-cell-depleted and non-T-cell-depleted unrelated donor transplantation for hematologic diseases: Clinical outcomes, quality of life, and costs. Blood 2002, 100, 2697–2702.

30. Champlin, R.E.; Passweg, J.R.; Zhang, M.J.; Rowlings, P.A.; Pelz, C.J.; Atkinson, K.A.; Barrett, A.J.; Cahn, J.Y.; Drobyski, W.R.; Gale, R.P.; et al. T-cell depletion of bone marrow transplants for leukemia from donors other than HLA-identical siblings: Advantage of T-cell antibodies with narrow specificities. Blood 2000, 95, 3996–4003.

31. Urbano-Ispizua, A.; Brunet, S.; Solano, C.; Moraleda, J.M.; Rovira, M.; Zuazu, J.; de La Rubia, J.; Bargay, J.; Caballero, D.; Diez-Martin, J.L.; et al. Allogeneic transplantation of CD34+-selected cells from peripheral blood in patients with myeloid malignancies in early phase: A case control comparison with unmodified peripheral blood transplantation. Bone Marrow Transplant. 2001, 28, 349–354.
32. Jakubowski, A.A.; Small, T.N.; Young, J.W.; Kernan, N.A.; Castro-Malaspina, H.; Hsu, K.C.; Perales, M.A.; Collins, N.; Cisek, C.; Chiu, M.; et al. T cell depleted stem-cell transplantation for adults with hematologic malignancies: Sustained engraftment of HLA-matched related donor grafts without the use of antithymocyte globulin. *Blood* 2007, 110, 4552–4559.

33. Jakubowski, A.A.; Small, T.N.; Kernan, N.A.; Castro-Malaspina, H.; Collins, N.; Koehne, G.; Hsu, K.C.; Perales, M.A.; Papanicolaou, G.; van den Brink, M.R.; et al. T Cell-Depleted Unrelated Donor Stem Cell Transplantation Provides Favorable Disease-Free Survival for Adults with Hematologic Malignancies. *Biol. Blood Marrow Transplant.* 2011, 17, 1335–1342.

34. Devine, S.M.; Carter, S.; Soiffer, R.J.; Pasquini, M.C.; Hari, P.N.; Stein, A.; Lazarus, H.M.; Linker, C.; Stadtmauer, E.A.; Alyea, E.P., III; et al. Low risk of chronic graft-versus-host disease and relapse associated with T cell-depleted peripheral blood stem cell transplantation for acute myelogenous leukemia in first remission: Results of the blood and marrow transplant clinical trials network protocol 0303. *Biol. Blood Marrow Transplant.* 2011, 17, 1343–1351.

35. Bethge, W.A.; Faul, C.; Bornhauser, M.; Stuhler, G.; Beelen, D.W.; Lang, P.; Stelljes, M.; Vogel, W.; Hagele, M.; Handgretinger, R.; et al. Haploidentical allogeneic hematopoietic cell transplantation in adults using CD3/CD19 depletion and reduced intensity conditioning: An update. *Blood Cells Mol. Dis.* 2008, 40, 13–19.

36. Bayraktar, U.D.; de Lima, M.; Saliba, R.M.; Maloy, M.; Castro-Malaspina, H.R.; Chen, J.; Rondon, G.; Chiattone, A.; Jakubowski, A.A.; Boulad, F.; et al. Ex Vivo T Cell Depleted versus Unmodified Allografts in Patients with Acute Myeloid Leukemia in First Complete Remission. *Biol. Blood Marrow Transplant.* 2013, 19, 898–903.

37. Aversa, F.; Terenzi, A.; Tabilio, A.; Falzetti, F.; Carotti, A.; Ballanti, S.; Felicini, R.; Falcinelli, F.; Velardi, A.; Ruggeri, L.; et al. Full haplotype-mismatched hematopoietic stem-cell transplantation: A phase II study in patients with acute leukemia at high risk of relapse. *J. Clin. Oncol.* 2005, 23, 3447–3454.

38. Bertaina, A.; Merli, P.; Rutella, S.; Pagliara, D.; Bernardo, M.E.; Masetti, R.; Pende, D.; Falco, M.; Handgretinger, R.; Moretta, F.; et al. HLA-haploidentical stem cell transplantation after removal of alphabeta+ T and B cells in children with nonmalignant disorders. *Blood* 2014, 124, 822–826.

39. Zecca, M.; Strocchio, L.; Pagliara, D.; Comoli, P.; Bertaina, A.; Giorgiani, G.; Perotti, C.; Corbella, F.; Brescia, L.; Locatelli, F. HLA-haploidentical T cell-depleted allogeneic hematopoietic stem cell transplantation in children with Fanconi anemia. *Biol. Blood Marrow Transplant.* 2014, 20, 571–576.

40. Lang, P.; Teltschik, H.M.; Feuchttinger, T.; Muller, I.; Pfieffer, M.; Schumm, M.; Ebinger, M.; Schwarze, C.P.; Gruhn, B.; Schrauder, A.; et al. Transplantation of CD3/CD19 depleted allografts from haploidentical family donors in paediatric leukaemia. *Br. J. Haematol.* 2014, 165, 688–698.

41. Gonzalez-Llano, O.; Rodriguez-Romo, L.N.; Mancias-Guerra Mdel, C.; Tarin-Arzaga, L.; Jaime-Perez, J.C.; Herrera-Garza, J.L.; Cantu-Rodriguez, O.G.; Gutierrez-Aguirre, C.H.; Garcia-Sepulveda, R.D.; Garcia-Marin, A.Y.; et al. Feasibility of an outpatient HLA haploidentical stem cell transplantation program in children using a reduced-intensity conditioning regimen and CD3-CD19 depletion. *Hematology* 2014, 19, 10–17.
42. Kernan, N.A.; Collins, N.H.; Juliano, L.; Cartagena, T.; Dupont, B.; O’Reilly, R.J. Clonable T lymphocytes in T cell-depleted bone marrow transplants correlate with development of graft-v-host disease. *Blood* 1986, 68, 770–773.

43. Sehn, L.H.; Alyea, E.P.; Weller, E.; Canning, C.; Lee, S.; Ritz, J.; Antin, J.H.; Soiffer, R.J. Comparative outcomes of T-cell-depleted and non-T-cell-depleted allogeneic bone marrow transplantation for chronic myelogenous leukemia: Impact of donor lymphocyte infusion. *J. Clin. Oncol.* 1999, 17, 561–568.

44. Pasquini, M.C.; Devine, S.; Mendizabal, A.; Baden, L.R.; Wingard, J.R.; Lazarus, H.M.; Appelbaum, F.R.; Keever-Taylor, C.A.; Horowitz, M.M.; Carter, S.; et al. Comparative outcomes of donor graft CD34+ selection and immune suppressive therapy as graft-versus-host disease prophylaxis for patients with acute myeloid leukemia in complete remission undergoing HLA-matched sibling allogeneic hematopoietic cell transplantation. *J. Clin. Oncol.* 2012, 30, 3194–3201.

45. Hobbs, G.S.; Hilden, P.; Hamdi, A.; Goldberg, J.D.; Poon, M.; Ledesma, C.; Devlin, S.; Rondon, G.; Papadopoulos, E.B.; Jakubowski, A.A.; et al. Outcomes in Patients with Acute Lymphoblastic Leukemia in First or Second Complete Remission Receiving Ex-Vivo T-Cell Depleted or Unmodified Allografts: Comparison of Results At Two Institutions. *Blood* 2013, 122, 3370.

46. Marmont, A.M.; Horowitz, M.M.; Gale, R.P.; Sobocinski, K.; Ash, R.C.; van Bekkum, D.W.; Champlin, R.E.; Dicke, K.A.; Goldman, J.M.; Good, R.A.; et al. T-cell depletion of HLA-identical transplants in leukemia. *Blood* 1991, 78, 2120–2130.

47. Small, T.N. Immunologic reconstitution following stem cell transplantation. *Curr. Opin. Hematol.* 1996, 3, 461–465.

48. Small, T.N.; Papadopoulos, E.B.; Boulad, F.; Black, P.; Castro-Malaspina, H.; Childs, B.H.; Collins, N.; Gillio, A.; George, D.; Jakubowski, A.; et al. Comparison of immune reconstitution after unrelated and related T-cell-depleted bone marrow transplantation: Effect of patient age and donor leukocyte infusions. *Blood* 1999, 93, 467–480.

49. Luznik, L.; Bolanos-Meade, J.; Zahurak, M.; Chen, A.R.; Smith, B.D.; Brodsky, R.; Huff, C.A.; Borrello, I.; Matsui, W.; Powell, J.D.; et al. High-dose cyclophosphamide as single-agent, short-course prophylaxis of graft-versus-host disease. *Blood* 2010, 115, 3224–3230.

50. Kanakry, C.G.; Tsai, H.L.; Bolanos-Meade, J.; Smith, B.D.; Gojo, I.; Kanakry, J.A.; Kasamon, Y.L.; Gladstone, D.E.; Matsui, W.; Borrello, I.; et al. Single-agent GVHD prophylaxis with posttransplantation cyclophosphamide after myeloablative, HLA-matched BMT for AML, ALL, and MDS. *Blood* 2014, 124, 3817–3827.

51. Kanakry, C.G.; O’Donnell, P.V.; Furlong, T.; de Lima, M.J.; Wei, W.; Medeot, M.; Mielcarek, M.; Champlin, R.E.; Jones, R.J.; Thall, P.F.; et al. Multi-institutional study of post-transplantation cyclophosphamide as single-agent graft-versus-host disease prophylaxis after allogeneic bone marrow transplantation using myeloablative busulfan and fludarabine conditioning. *J. Clin. Oncol.* 2014, 32, 3497–3505.

52. Storek, J.; Gooley, T.; Witherspoon, R.P.; Sullivan, K.M.; Storb, R. Infectious morbidity in long-term survivors of allogeneic marrow transplantation is associated with low CD4 T cell counts. *Am. J. Hematol.* 1997, 54, 131–138.
53. Curtis, R.E.; Rowlings, P.A.; Deeg, H.J.; Shriner, D.A.; Socie, G.; Travis, L.B.; Horowitz, M.M.; Witherspoon, R.P.; Hoover, R.N.; Sobocinski, K.A.; et al. Solid cancers after bone marrow transplantation. *N. Engl. J. Med.* 1997, 336, 897–904.

54. Small, T.N.; Avigan, D.; Dupont, B.; Smith, K.; Black, P.; Heller, G.; Polya, T.; O’Reilly, R.J. Immune reconstitution following T-cell depleted bone marrow transplantation: Effect of age and posttransplant graft rejection prophylaxis. *Biol. Blood Marrow Transplant.* 1997, 3, 65–75.

55. Van Heijst, J.W.; Ceberio, I.; Lipuma, L.B.; Samilo, D.W.; Wasilewski, G.D.; Gonzales, A.M.; Nieves, J.L.; van den Brink, M.R.; Perales, M.A.; Pamer, E.G. Quantitative assessment of T cell repertoire recovery after hematopoietic stem cell transplantation. *Nat. Med.* 2013, 19, 372–377.

56. Lewin, S.R.; Heller, G.; Zhang, L.; Rodrigues, E.; Skulsky, E.; van den Brink, M.R.; Small, T.N.; Kernan, N.A.; O’Reilly, R.J.; Ho, D.D.; et al. Direct evidence for new T-cell generation by patients after either T-cell-depleted or unmodified allogeneic hematopoietic stem cell transplantations. *Blood* 2002, 100, 2235–2242.

57. Weinberg, K.; Blazar, B.R.; Wagner, J.E.; Agura, E.; Hill, B.J.; Smogorzewska, M.; Koup, R.A.; Betts, M.R.; Collins, R.H.; Douek, D.C. Factors affecting thymic function after allogeneic hematopoietic stem cell transplantation. *Blood* 2001, 97, 1458–1466.

58. Clave, E.; Busson, M.; Douay, C.; Peffault de Latour, R.; Berrou, J.; Rabian, C.; Carmagnat, M.; Rocha, V.; Charron, D.; Socie, G.; et al. Acute graft-versus-host disease transiently impairs thymic output in young patients after allogeneic hematopoietic stem cell transplantation. *Blood* 2009, 113, 6477–6484.

59. Olkinuora, H.; von Willebrand, E.; Kantele, J.M.; Vainio, O.; Talvensaari, K.; Saarinen-Pihkala, U.; Siitonen, S.; Vettenranta, K. The impact of early viral infections and graft-versus-host disease on immune reconstitution following paediatric stem cell transplantation. *Scand. J. Immunol.* 2011, 73, 586–593.

60. Perales, M.A.; Ishill, N.; Lomazow, W.A.; Weinstock, D.M.; Papadopoulos, E.B.; Dastigir, H.; Chiu, M.; Boulad, F.; Castro-Malaspina, H.R.; Heller, G.; et al. Long-term follow-up of patients treated with daclizumab for steroid-refractory acute graft-vs.-host disease. *Bone Marrow Transplant.* 2007, 40, 481–486.

61. Willenbacher, W.; Basara, N.; Blau, I.W.; Fauser, A.A.; Kiehl, M.G. Treatment of steroid refractory acute and chronic graft-versus-host disease with daclizumab. *Br. J. Haematol.* 2001, 112, 820–823.

62. Arai, S.; Margolis, J.; Zahurak, M.; Anders, V.; Vogelsang, G.B. Poor outcome in steroid-refractory graft-versus-host disease with antithymocyte globulin treatment. *Biol. Blood Marrow Transplant.* 2002, 8, 155–160.

63. McCaul, K.G.; Nevill, T.J.; Barnett, M.J.; Toze, C.L.; Currie, C.J.; Sutherland, H.J.; Conneally, E.A.; Shepherd, J.D.; Nantel, S.H.; Hogge, D.E.; et al. Treatment of steroid-resistant acute graft-versus-host disease with rabbit antithymocyte globulin. *J. Hematother. Stem Cell Res.* 2000, 9, 367–374.

64. Khoury, H.; Kashyap, A.; Adkins, D.R.; Brown, R.A.; Miller, G.; Vij, R.; Westervelt, P.; Trinkaus, K.; Goodnough, L.T.; Hayashi, R.J.; et al. Treatment of steroid-resistant acute graft-versus-host disease with anti-thymocyte globulin. *Bone Marrow Transplant.* 2001, 27, 1059–1064.
65. Storek, J.; Joseph, A.; Espino, G.; Dawson, M.A.; Douek, D.C.; Sullivan, K.M.; Flowers, M.E.; Martin, P.; Mathioudakis, G.; Nash, R.A.; et al. Immunity of patients surviving 20 to 30 years after allogeneic or syngeneic bone marrow transplantation. Blood 2001, 98, 3505–3512.

66. Storek, J.; Witherspoon, R.P.; Storb, R. T cell reconstitution after bone marrow transplantation into adult patients does not resemble T cell development in early life. Bone Marrow Transplant. 1995, 16, 413–425.

67. Geyer, M.B.; Ricci, A.M.; Jacobson, J.S.; Majzner, R.; Duffy, D.; Van de Ven, C.; Ayello, J.; Bhatia, M.; Garvin, J.H., Jr.; George, D.; et al. T cell depletion utilizing CD34(+) stem cell selection and CD3(+) addback from unrelated adult donors in paediatric allogeneic stem cell transplantation recipients. Br. J. Haematol. 2012, 157, 205–219.

68. Dvorak, C.C.; Gilman, A.L.; Horn, B.; Oon, C.Y.; Dunn, E.A.; Baxter-Lowe, L.A.; Cowan, M.J. Haploidentical related-donor hematopoietic cell transplantation in children using megadoses of ClinIMACs-selected CD34(+) cells and a fixed CD3(+) dose. Bone Marrow Transplant. 2013, 48, 508–513.

69. Dvorak, C.C.; Gilman, A.L.; Horn, B.; Jaroscak, J.; Dunn, E.A.; Baxter-Lowe, L.A.; Cowan, M.J. Clinical and immunologic outcomes following haplocompatible donor lymphocyte infusions. Bone Marrow Transplant. 2009, 44, 805–812.

70. Alpdogan, O.; Hubbard, V.M.; Smith, O.M.; Patel, N.; Lu, S.; Goldberg, G.L.; Gray, D.H.; Feinman, J.; Kochman, A.A.; Eng, J.M.; et al. Keratinocyte growth factor (KGF) is required for postnatal thymic regeneration. Blood 2006, 107, 2453–2460.

71. Jenq, R.R.; King, C.G.; Volk, C.; Suh, D.; Smith, O.M.; Rao, U.K.; Yim, N.L.; Holland, A.M.; Lu, S.X.; Zakrzewski, J.L.; et al. Keratinocyte growth factor enhances DNA plasmid tumor vaccine responses after murine allogeneic bone marrow transplantation. Blood 2009, 113, 1574–1580.

72. Vadhan-Raj, S.; Goldberg, J.D.; Perales, M.A.; Berger, D.P.; van den Brink, M.R. Clinical applications of palifermin: Amelioration of oral mucositis and other potential indications. J. Cell Mol. Med. 2013, 17, 1371–1384.

73. Perales, M.A.; Goldberg, J.D.; Yuan, J.; Koehne, G.; Lechner, L.; Papadopoulos, E.B.; Young, J.W.; Jakubowski, A.A.; Zaidi, B.; Gallardo, H.; et al. Recombinant human interleukin-7 (CYT107) promotes T-cell recovery after allogeneic stem cell transplantation. Blood 2012, 120, 4882–4891.

74. Hegenbart, U.; Niederwieser, D.; Sandmaier, B.M.; Maris, M.B.; Shizuru, J.A.; Greinix, H.; Cordonnier, C.; Rio, B.; Gratwohl, A.; Lange, T.; et al. Treatment for acute myelogenous leukemia by low-dose, total-body, irradiation-based conditioning and hematopoietic cell transplantation from related and unrelated donors. J. Clin. Oncol. 2006, 24, 444–453.

75. Aoudjhane, M.; Labopin, M.; Gorin, N.C.; Shimoni, A.; Ruutu, T.; Kolb, H.J.; Frassoni, F.; Boiron, J.M.; Yin, J.L.; Finke, J.; et al. Comparative outcome of reduced intensity and myeloablative conditioning regimen in HLA identical sibling allogeneic haematopoietic stem cell transplantation for patients older than 50 years of age with acute myeloblastic leukaemia: A retrospective survey from the Acute Leukemia Working Party (ALWP) of the European group for Blood and Marrow Transplantation (EBMT). Leukemia 2005, 19, 2304–2312.

76. O’Reilly, R.J.; Doubrovina, E.; Trivedi, D.; Hasan, A.; Kollen, W.; Koehne, G. Adoptive transfer of antigen-specific T-cells of donor type for immunotherapy of viral infections following allogeneic hematopoietic cell transplants. Immunol. Res. 2007, 38, 237–250.
77. Koehne, G.; Doubrovina, E.; Hasan, A.; Barker, J.N.; Castro-Malaspina, H.; Perales, M.A.; Jakubowski, A.; Papadopoulos, E.; Young, J.W.; Boulad, F.; et al. A Phase I Dose Escalation Trial of Donor T Cells Sensitized with Pentadecapeptides of the CMV-pp65 Protein for the Treatment of CMV Infections Following Allogeneic Hematopoietic Stem Cell Transplants. *Blood* 2009, 114, 2262.

78. Doubrovina, E.; Oflaz-Sozmen, B.; Prockop, S.E.; Kernan, N.A.; Abramson, S.; Teruya-Feldstein, J.; Hedvat, C.; Chou, J.F.; Heller, G.; Barker, J.N.; et al. Adoptive immunotherapy with unselected or EBV-specific T cells for biopsy-proven EBV+ lymphomas after allogeneic hematopoietic cell transplantation. *Blood* 2012, 119, 2644–2656.

79. Blyth, E.; Clancy, L.; Simms, R.; Ma, C.K.; Burgess, J.; Deo, S.; Byth, K.; Dubosq, M.C.; Shaw, P.J.; Micklethwaite, K.P.; et al. Donor-derived CMV-specific T cells reduce the requirement for CMV-directed pharmacotherapy after allogeneic stem cell transplantation. *Blood* 2013, 121, 3745–3758.

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