THE HYDROPHOBICITY AND THE ANTIBACTERIAL ACTIVITY OF POLYESTER MODIFIED WITH SILVER NANOPARTICLE AND HEXADECYLTRIMETHOXYSILANE

HIDROFOBISITAS DAN AKTIVITAS ANTIBAKTERI POLIESTER HASIL MODIFIKASI DENGAN NANOPARTIKEL PERAK DAN HEKSADESILTRIMETOKSISILAN

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ABSTRACT

The objective of this research was to study hydrophobicity and antibacterial activity of polyester fibers before and after modification by using silver nanoparticles as antibacterial agent and hexadecyltrimethoxysilane (HDTMS) as a self-cleaning agent. The research was conducted in stages, preparation of silver nanoparticles, deposition of silver nanoparticles on polyester fiber, modification of polyester fiber through the addition HDTMS, and characterization. Modification of polyester fibers with silver nanoparticles and HDTMS was conducted through dipping method, followed by curing. Characterization was performed by testing the contact angle and antibacterial activity against Staphylococcus aureus ATCC 25923 and Escherichia coli ATCC 35218. Silver nanoparticles have been prepared by adding sodium citrate and polyvinylalcohol as a stabilizer. The addition of silver nanoparticles decreased hydrophobicity of polyester fibers without and with modification. Modification with HDTMS increased the hydrophobicity of polyester fibers. Modification with silver nanoparticles and HDTMS increased antibacterial activity of polyester fabrics. Statistic analysis showed that there were significant differences in the antibacterial activity of polyester fibers against Staphylococcus aureus ATCC 25923 and Escherichia coli ATCC 35218.

Keywords: antibacterial activity, hydrophobicity, silver nanoparticles, polyester fabric.

INTRODUCTION

Pollution is one of the causes of contamination of the natural environment in Indonesia. Air pollution makes the air temperature is becoming increasingly high. The hot air causes the body to sweat more and more dust stick. This condition speeds the growth of many types of bacteria that are pathogenic for the human body. These bacteria can cause various diseases to the human body.

Clothing is one of the outer protection for the body from environmental including the bacteria that grow in a skin as well as clothing was worn. Therefore, it needs fabrics that can...
protect the human body from bacteria effectively. One effort to improve the quality and quantity of textile materials is to conduct modifications on textiles with properties hydrophobic and antibacterial.

Textile applications with hydrophobic and antibacterial properties have been widely used in the medical field as a base for dressings, protective clothing for patients, physicians, and as well as in the military was used for soldiers’ uniforms (El-Khatib, 2012). However, until now the textile material with antibacterial and hydrophobic properties is not found on the market. Demand for textile products with antibacteria and hydrophobic properties still dependent on foreign material with a high price. The need for textiles with antibacterial and hydrophobic properties that encourage increased import of the material of the cloth.

The background of this research is dependence on imported products, high-value import, high prices, and the demand for textile with antibacterial and hydrophobic properties. Production of textile materials with antibacterial and hydrophobic properties in the country is expected to reduce import value of textile materials Indonesia and can meet the demand for clothing, especially textile materials with antibacterial and hydrophobic properties.

Nanoparticle technology can be applied in the textile industry to modify various textile fibers that have antibacterial properties. The silver nanoparticle is one of nanostructured particle which has effective antibacterial activity (Haryono and Harmami, 2010). Silver nanoparticles can inhibit the growth of bacteria Escherichia coli, Bacillus subtilis, Staphylococcus aureus as disclosed by Ariyanta, Wahyuni, and Priatmoko (2014), and Pseudomonas aeruginosa (Guzman, Dille, and Godet, 2012).

Some of the literature revealed that the antibacterial properties of textiles can be developed by coating of silver nanoparticles in textile materials, for example on cotton (Shateri-Khalilabad and Yazdanshenas, 2013), silk (Zhang, Liu, Gao, and Chen, 2014), wool (Boroumand, Montazer, Simon, Liesiene, Šaponjic, and Dutschik, 2015), polyester (Kavitha and Dasan, 2013). The antibacterial activity is influenced by the size of the silver particles (Haryono and Harmami, 2010). The smaller the size of the nanoparticle, its activity increases. The optimal size of silver nanoparticles is small size with a distribution is very narrow (Crespo, García-Barrasa, López-de-Luzuriaga, and Monge, 2012). Also, the low concentration of silver nanoparticles is safe to use on the human body because the silver nanoparticles are not toxic to humans (Rai, Yadav, and Gade 2009). Preparation of silver nanoparticles can be done by various methods including: sputtering (Jiang, Qin, and Zhang, 2010), reduction with a compound organic (Ahmad, Tay, Shameli, Hussein, and Lim, 2011), reduction with a fungi (Duran and Marcato, 2007), and reduction with a sodium citrate solution (Ariyanta, Wahyuni, and Priatmoko, 2014).

Hydrophobicity of the textile affect the properties of the textile. Textiles with hydrophobic properties are often referred to as self-cleaning textiles. The hydrophobic properties of textile materials can be developed by using a silane-based compound. Fiber materials are coated by the silane with low surface energy, are proven to increase the hydrophobicity of the fiber (Khalil-Abad and Yazdanshenas, 2010; Xue, Chen, Yin, Jia, Jian-Zhong, 2012). This coating is inspired from the nature of the plant leaf surface Nelumbo nucifera or lotus is often called hydrophobic. Some types of silane-based compounds have been used to modify textiles such as γ-methacryloxypropyl trimethoxy silane (MAPS) and hexamethylidisilazene (HMDS) (Gao, Watanabe, Nakane, and Zhao, 2016); perfluorooctylated quaternary ammonium silane coupling agent (PFSC) (Yu, Gu, Meng, and Qing, 2007); polymethylhydroxydisiloxane (PMHS) and tetraethoxy-silane (TEOS) (Guo, Zhai, Xiao, and An, 2015); and Hexadecyltrimethoxysilane (HDTMS) (Xue et al., 2012).

Polyester is known as Dacron or Terelene in the trade. Polyester fiber is one kind of synthetic fiber which strong, durable, and not wrinkled. Polysters are also often used as a mixture of natural fibers such as cotton, wool or rayon to obtain fabrics with superior quality. Naturally polyester has hydrophobic properties that are often used as a sports clothing, underwear, bed linen, and medical garments. Textiles that are resistant to bacteria, viruses, fungi, and other harmful microorganisms are also often needed as raw materials for sports clothing, underwear, bed linen, and medical garments (Hassan, Qashqary, Hassan, Shady, and Alansary, 2012).
Based on the background of the problem, the research on the modification of polyester fibers to improve the antibacterial properties and hydrophobic has been conducted. The use of silver nanoparticles and silane-based compound to coat the polyester fiber is conducted to improve the antibacterial properties and hydrophobic of polyester fiber. Preparation of silver nanoparticles has been performed by the reduction method using AgNO₃ solution. The polyester fiber was coated with HDTMS compound by the immersion method. The formation of silver nanoparticles was analyzed by using UV-Vis spectrophotometer and the hydrophobic properties by measuring the angle of contact, and the antibacterial activity of the polyester fiber by measuring clear zone. The objective of this study was to determine the characteristics of polyester fibers with and without modification using silver nanoparticles and Hexadecyltrimethoxysilane.

MATERIALS AND METHODS

Equipment and Materials

The UV-Vis spectrophotometer (Shimadzu UV-2400PC Series), Laminar Air Flow (LAF), incubator, oven, autoclaves, regulators of nitrogen gas, shaker, hot plate - magnetic stirrer, analytical balance, camera, glasswears, Bunsen, calipers, tip pipet, thermometer, Drigalsky, and ose have been used in this research.

The Polyester cloth was purchased from the fabric store in Yogyakarta. Silver nitrate, trisodium citrate, polyvinyl alcohol (PVA), ethanol, acetone, and hexadecyltrimethoxysilane (HDTMS) were purchased as commercial products and used as they were without any further purification. Nutrient Agar (NA) and Nutrient Broth (NB) were purchased from Oxoid. Staphylococcus aureus ATCC 25923 and Eschericia coli 32518 were obtained from a collection of Faculty of Medicine, Gadjah Mada University.

Procedures

Preparation of Silver Nanoparticle (N)

Silver nanoparticle was prepared by using 0.001M silver nitrate solution, 10% trisodium citrate solution as a reductor, and 0.2% PVA solution as a stabilizer (Haryono, Sondari, Harmami, dan Randy, 2008). PVA solution and silver nitrate solution were added into three neck flask then refluxed for 5 hours (Saputra, Haryono, Laksmono, and Anshari, 2011). Trisodium citrate solution was added dropwise. Gas N₂ was flowing until reflux process finished. Heating and flowing of N₂ gas were stopped if already transformed solution into a yellow, but stirring was still done until room temperature reached. Then, silver nanoparticles were characterized using UV-Vis spectrophotometer.

Application of Silver Nanoparticles on Polyester Fiber (Polyester-Ag)

The polyester fabric was cut to the size of 10 cm x 10 cm. Polyester fiber was immersed in colloidal of silver nanoparticle then twisted around using a shaker at 150 rpm for 24 hours and dried at 70 °C.

Modification Surface Polyester Fiber with Compound HDTMS (Polyester-HDTMS)

The polyester and the polyester-Ag were immersed into the 4% of HDTMS solution. The reacting process between HDTMS and ethanol solution was carried out at room temperature for 6 hours. The polyester and polyester - Ag which immersed in silane solution were twisted at 150 rpm for 60 minutes. Then, polyester fiber before and after modification were analyzed including antibacterial activity test and contact angle test.

The polyesters which prepared in this study were polyester fiber, polyester-Ag, polyester-HDTMS, and polyester-Ag-HDTMS (Table 1).

| No | Type of Sample | Code |
|----|----------------|------|
| 1  | Polyester without modification | P    |
| 2  | Polyester with adding silver nanoparticles | P-Ag |
| 3  | Polyester with adding HDTMS | P-HDTMS |
| 4  | Polyester with adding silver nanoparticles and HDTMS | P-Ag-HDTMS |

Tabel 1. The variation of the polyester fiber without and with modification
The characteristic of silver nanoparticle was performed using UV-Vis spectrophotometer (Shimadzu UV-2400PC series, Japan). An absorbance of silver nitrate solution $1 \times 10^{-3}$ M and silver nanoparticle were measured using a reference solution of distilled water. The properties anti-dirty (hydrophobic) of the samples were determined by measuring the water contact angle ($\theta$) between the fluid and the sample surface. Samples were placed on the surface of a table or a flat board and micropipette is placed on the top then paired with the upright. By using a pipette, liquid dripped from a height of 1 cm of the sample. Once the liquid dropped, the contact angle shooting was done. The images processed using software to determine the contact angle between the liquid surface of the sample.

Antibacterial Activity of the sample of P, P-Ag, P-HDTMS, and P-Ag-HDTMS performed against *Escherichia coli* ATCC 35218 and *Staphylococcus aureus* ATCC 25923 by measuring the clear zone is formed around the sample. Antibacterial activity was
performed by preparing bacterial growth media such as Nutrient Agar (NA) and Nutrient Broth (NB) by dissolving NA and NB in distilled water. All the tools and media for growing bacteria were sterilized in an autoclave. Rejuvenation of Staphylococcus aureus ATCC 25923 and Escherichia coli ATCC 32518 was conducted on an agar medium NA and incubated for 24 hours at room temperature. Staphylococcus aureus 25923 and Escherichia coli ATCC 32518 which has been rejuvenated for 24 hours and then inoculated into a liquid medium NB in the culture bottles and incubated for 24 hours at a temperature of 37 °C. Meanwhile, NA poured into each petri dish and wait ed about 24 hours anyway. Petridish had been ascertained that no contamination is then coated NB which had been overgrown with bacteria and leveled by using Drygalsky. Each sample was cut with a diameter of 0.6 cm, inserted into the petri dish and allowed in the incubator for 24 hours, then observed a clear zone every three hours for 96 hours. The inhibition zone was measured by using calipers.

RESULTS AND DISCUSSION

The Characteristics of Silver Nanoparticles and P-Ag

The silver nanoparticles have been successfully prepared in the colloidal form

$$4Ag^+_{(aq)} + C_6H_5O_7Na_3(aq) + 2H_2O(l) \rightarrow 4Ag_0(s) + C_6H_5O_7H_3(aq) + 3Na^+_{(aq)} + H^+_{(aq)} + O_2(g) \quad (1)$$

with yellow color. The Equation 1 shows reduction reaction that occurs in preparation silver nanoparticle (Haryono and Sri Budi Harmami, 2010). Figure 1 shows the UV-Vis spectra of silver nitrate solution and colloidal of silver nanoparticles. The UV-Vis spectrum of colloidal of silver nanoparticles shows absorption peak at wavelength 429 nm and the absorbance of 0.333 which suggests that the Ag⁺ has reduced into Ag₀ (Junaidi, Wahyudi, and Umaningrum 2015). The polyester fiber which has been deposited by silver nanoparticles (P-Ag) turns into more brown color than the pure polyester fiber (P) (Figure 2).

Hydrophobic Properties of Polyester

Hydrophobic properties of samples was determined by measuring contact angle (Figure 3 dan Table 2). The hydrophobic surface has a contact angle more of 90° (Arkles, 2006). Based on Table 2 can be seen that P-HDTMS has the highest contact angle. Thus, HDTMS compound can increase contact angle of polyester. It can be caused HDTMS compounds interact with a material surface through the formation of covalent bonding and lowering the critic surface tension until smaller than critic surface tension of the water. Consequently, surface of polyester fiber becomes more hydrophobic.
The contact angle test also showed that the contact angle of polyester fibers decreased. It was caused by the addition of silver nanoparticles as an antibacterial material. Deposit of silver nanoparticles into the polyester can cause a decreasing of a contact area between HDTMS and polyester. Thus HDTMS can not coat on surface of polyester. Modification of silver nanoparticles can cause a decreasing of the contact angle of polyester fibers without and with modification by the compound of HDTMS. Thus the presence of silver nanoparticles can reduce hydrophobic properties of polyester fibers.

**Antibacterial Activities of Polyester Fiber**  

**Figure 4a** shows antibacterial activities of polyester fibers without and with modification against *E. coli* and **Figure 4b** shows antibacterial activities of polyester fibers without and with modification against *S. aureus*. All samples show antibacterial activities except P and P-HDTMS. Samples polyesters almost do not perform antibacterial activities against *E. coli* but show significant antibacterial activities against *S. aureus*. Antibacterial activities of samples P-HDTMSs against *S. aureus* are very low but are more against *E. coli*.
Figure 5a. The Clear Zone of Polyester Fiber Before and After Modification against *Staphylococcus aureus* ATCC 25923

Figure 5b. The Clear Zone of Polyester Fiber Before and After Modification against *Escherichia coli* ATCC 35218
Figure 4 shows that the samples which deposited with silver nanoparticles have the highest antibacterial activity. The silver nanoparticles are attached to the bacterial cell membrane is very possible. Silver nanoparticles react with sulfur protein and phosphorus-containing DNA in a bacterial cell. The reaction causes a changing morphological in bacterial cells, DNA damage, and respiratory problems, so the bacteria die (Zhang et al., 2014).

Figure 5a and Figure 5b shows the clear zone of polyester fiber before and after modification. ANOVA test to the diameter of the clear zone of sample types (treatment) in inhibiting the growth of Escherichia coli and Staphylococcus aureus showed sig 0.00 (P<0.05), as well as on further testing LSD showed the differences significant in the two types of bacteria (Table 3).

Further LSD test was conducted to determine the differences among the four types of samples. Data to be tested using LSD should be declared significantly different at ANOVA test to obtain valid test results. Table 3 shows the results of LSD test for antibacterial activity against E. coli and S. aureus. The sixth combination of sample type showed a significant difference regarding antibacterial activity against E. coli and S. aureus.

Based on analysis using Post Host Duncan indicated that all samples have differences antibacterial activity in inhibiting the growth of bacteria Escherichia coli ATCC 35218 and Staphylococcus aureus ATCC 25923 significantly. T-test results showed that all types of samples showed a significance value of less than 5% error level (0.05), indicating a significant difference in antibacterial activity against Escherichia coli ATCC 35218 and Staphylococcus aureus ATCC 25923.

Thus, there is a difference in the antibacterial activity between polyester fiber with and without modification against E. coli and S. aureus. This difference is influenced by the characteristics of a cell wall of Gram-positive bacteria and gram-negative. S. aureus as gram-positive bacteria have a few layers of peptidoglycan join to form a thick and rigid structure, while E. coli is a gram-negative bacteria have a thin layer of peptidoglycan and outer membrane protected (Shagam, 2006 and Wheelis, 2007).

The cell walls of gram-positive bacteria also contain acids teichoic, consisting of -OH groups (such as ribitol and alcohol) and phosphate. A variety of functional groups here that have an important role in the interaction of silver nanoparticles with bacteria. It leads to more polyester effectively as an antibacterial against Staphylococcus aureus (Wheelis, 2007).

The mechanism of antibacterial activity can be explained by the interaction of silver with the bacteria that make up the protein corona (Figure 6). When the silver nanoparticles are mixed with bacteria in the culture medium, in the first phase of silver nanoparticles join with salt and proteins to form Protein Corona. Protein Corona will be close and attached to the bacterial cell wall and then destroyed it so that the penetration could occur. Once inside, the nanoparticles are still in the form of protein corona immediate release of silver ions. Silver ions will destroy the membrane of bacteria and cause the death of bacteria (Rai, Yadav, and Gade, 2009; Jin, Xu, Dong, Lan, Jiang, and Liu, 2015).

Table 3. Interpretation of LSD Test: Diameter of Clear Zone of Type Samples against Staphylococcus aureus ATCC 25923 and Escherichia coli ATCC 35218

| Variable (Type of Sample)                  | Conclusion | E.coli | S.aureus |
|--------------------------------------------|------------|--------|----------|
| (P) – (P-Ag)                               | Significant|        |          |
| (P) – (P-HDTMS)                            | Significant|        |          |
| (P) – (P-Ag-HDTMS)                         | Significant|        |          |
| (P-Ag) - (P-HDTMS)                         | Significant|        |          |
| (P-Ag) – (P-HDTMS)                         | Significant|        |          |
| (P-HDTMS) – (P-Ag-HDTMS)                   | Significant|        |          |

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CONCLUSION
Silver nanoparticles decrease hydrophobicity polyester fiber without and with the addition of HDTMS. The addition of HDTMS compounds increases the hydrophobicity properties of polyester fibers without and with the addition of silver nanoparticles. Modifications with silver nanoparticles and HDTMS increase antibacterial activity of polyester fiber against Staphylococcus aureus ATCC 25923 and Escherichia coli ATCC 35218. There are significant differences regarding the antibacterial activity of polyester fibers with and without modifications against Staphylococcus aureus ATCC 25923 and Escherichia coli ATCC 35218.

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