Molecular Identification of *Brucella abortus* Collected from Whole Blood Samples of Seronegative Dairy Cattle with Reproductive Disorders in Central Java, Indonesia

Hastari Wuryastuty¹, Raden Wasito² and Sugiyono²

¹Department of Internal Medicine; ²Department of Pathology, Faculty of Veterinary Medicine, Universitas Gadjah Mada, Yogyakarta 55281, Indonesia

*Corresponding author: hastari@ugm.ac.id*

**ARTICLE HISTORY** (18-323)

Received: August 31, 2018

Revised: November 26, 2018

Accepted: December 19, 2018

Published online: January 24, 2019

**Key words:**

Brucellosis

IS711 gene fragment

PCR

Whole blood

Zoonotic disease

**ABSTRACT**

Brucellosis is a zoonotic disease that led to economic losses to cattle industries worldwide including Indonesia. A rapid, precise and accurate diagnosis technique is required for early detection and for the control of brucellosis in animals and humans. The objective of this study was to molecularly identify and characterize *B. abortus* in the blood samples obtained from seronegative dairy cattle that have had reproductive disorders in Central Java, Indonesia, using PCR, phylogenetic and nucleotide sequence analyses of the IS711 regions. Results showed that in seronegatively dairy cattle with reproductive disorders, *B. abortus* was successfully detected from the whole blood and serum samples without separation of the buffy coat and without isolation of the organism by conventional PCR. The result also showed high homology level (up to 100%) and close phylogenetic relationships between nucleotide sequences of the IS711 gene fragment of *B. abortus* local isolates compare to the isolates that have been accessed in GenBank (CP 009099, CP 023242, CP 023242, CP 023308 and LT 671513). We concluded that PCR technique is useful for routine diagnosis of brucellosis in seronegative dairy cattle with reproductive disorders.

©2018 PVJ. All rights reserved

**INTRODUCTION**

Reproductive efficiency is an important component of a successful cattle operation. Factor that induces poor reproductive performances is a major concern to the economic consequences. *Brucella* is facultative intracellular gram negative bacteria that cause brucellosis in several species of animals and human (Gupta et al., 2014). Even though is not pathognomonic, the clinical consequences caused by *Brucella spp* in infected animals usually correlated with reproductive problems such as abortion, infertility, retained placenta and birth of weak calves that produce large economic losses to dairy and beef farmers. Identification of infected animals among cattle population is one of the most important but challenging strategy to control the disease from spreading. In Indonesia, brucellosis is categorized as a strategic animal disease that rapidly spread among animals in the herd. Like most other ASEAN countries, the presence of brucellosis in the herd is mostly detected serologically. Serological method for brucellosis are not always sensitive and repeatedly been reported to cross-react with antigens other than *Brucella spp* (Al-Garadi et al., 2011; Priyadarshini et al., 2013). Other than that, the methods may also produce false-positive results, especially in cattle that has been vaccinated using *B. abortus* strain 19 or may cause false-negative results in cattle with no antibodies to *B. abortus*. False-negative animal can be a source of disease transmission that interferes the brucellosis control program (Silva Moi et al., 2014; El-Diasty et al., 2018). The gold standard for the diagnosis of brucellosis is isolation. However, the standard procedure for *Brucella* isolation has many disadvantages such as false-negative results, time consuming, and requires biosafety level 3 facilities and highly skilled technical personnel to handle samples and live bacteria for eventual identification and characterization (Perez-Sancho et al., 2013; Silva Moi et al., 2014). Therefore, polymerase chain reaction (PCR)-based method to detect the occurrence of the *Brucella* infection have become prevalent in the last decade because of their specificity, technical ease and lowering the costs (Mahajan et al.,
DNA purification: Positive control (live freeze-dried vaccine, Pusat Veteriner Farma, Surabaya, Indonesia that has been reconstituted with distilled water), whole blood and serum samples were extracted using GeneJET Genomic DNA Purification Kits described in the protocol provided by the manufacturer (Thermo Fisher Scientific, Waltham, Massachusetts, America). Briefly, 200 µL of either positive control solution, whole blood or serum were pipetted into a sterile 1.5 ml eppendorf tube and then 400 µL of lysis solution and 20 µL of Proteinase K solution were added into it, the solution was mixed thoroughly by vortexing and incubated the sample mix at 56°C for 10 min. After incubation, the prepared lysate was transferred into a GeneJET Genomic DNA Purification Column inserted in a collection tube and centrifuge for 1 min at 6000 x g. After centrifugation, discard the flow-through, place back the purification column into a new collection tube, add 500 µL of wash buffer I and centrifuge for 1 min at 8000 x g. After centrifugation, discard the flow-through, place back the purification column into a new collection tube, add 500 µL of wash buffer II and centrifuge for 3 min at 12000xg. Discard the collection tube and insert the GeneJET Genomic DNA Purification Column into a sterile 1.5 ml eppendorf tube, add 200 µL of elution buffer to the center of the GeneJET Genomic DNA Purification Column membrane to elute genomic DNA. Incubate for 2 min at room temperature and centrifuge for 1 min at 8000 x g.

DNA amplification and detection of PCR products: Molecular detection of B. abortus was done using gene-specific bscp31 primer (Bailey et al., 1992) and species-specific IS711 primer (Doust et al., 2007). PCR was carried out in a total volume of 50 µL, the DNA amplification was obtained by adding 25 µL of GoTaq®Green Master Mix 2x (Promega, Madison, WI 53771-5399 USA), 2.5 µL of 10 pMol primer forward and 2.5 µL of 10 pMol primer reverse (Table 1), 2.5 µL of 50 nMol MgCl2, 2, 5 µL of Template DNA and Nuclease free water to 50 µL. The PCR mixtures were then subjected to the cycling condition as in Table 2 in Personal Combi Thermocycler Biometra (37079 Goettingen Germany). The PCR amplification product(s) were separated on a 1.5% agarose gel, stained after electrophoresis with ethidium bromide and visualized using ultraviolet transillumination. For sequencing, PCR products were purified using High Pure PCR Product Purification Kit (Roche Life Science, Mannheim, Germany). Forward and reverse sequences for each sample were aligned and used in phylogenetic analysis. The sequences were compared to other previously published sequences. The sequence identities of nucleotide, as well as the estimation of the evolutionary divergence between sequences were analyzed using DNA Baser and Mega7 software, respectively (Kumar et al., 2016). The same tool was used to perform Neighbor-Joining analysis.

RESULTS

In this study, the presence of B. abortus was detected by conventional PCR from four out of 50 dairy cattle. Sample either in the form of whole blood or sera from the same animal produce the same result. Gel electrophoresis of amplicon confirmed that 2 pairs of primers used, specifically amplified bscp31 and IS711 brucella fragment genes in the position of 223 bp and 498 bp respectively (Fig. 1 and 2).

The clinical status of the dairy cattle at the time of sampling varied from abortion (10), repeat breeding (32), retention of placenta (6) and uterine prolapsed (2). Three of dairy cattle that showed positive by PCR had experienced abortion and one cattle suffered from sub-infarertility. The incidence of abortion that give positive results of brucellosis occur at different term of pregnancy (2.5 months, 4 months and 7.5 months).

The results of homology analysis in this study (Fig. 3) showed high degree of homology (98-100%) between IS711 gene nucleotide sequence of B. abortus local isolates and B. abortus from foreign countries. Whereas based on phylogenetic analysis, multiple nucleotide alignment among B. abortus isolates based on IS711 regions has close relationships with isolates from Zimbabwe (CP 009099) and water buffalo from Italy (CP 023242, CP 023308 and LT 671513).
**DISCUSSION**

*Brucellosis* is a zoonotic disease that has been classified in a group of high-risk pathogens. A rapid, precise and accurate diagnosis technique is needed for early detection of infection and for the control of *brucellosis* in animals and humans (Mahajan et al., 2017). PCR is a very potential and reliable molecular diagnosis method that has been widely used for detecting many infectious diseases. According to Ducrototy et al. (2017), in the case of detecting brucella infection at various stages, a combination between types of biological samples and appropriate diagnostic methods must be considered. The results of this study have revealed that using PCR both whole blood without buffy-coat separation and serum samples are suitable for direct testing without previous isolation of the organism (*B. abortus*). The use of commercially available kits in this study help improve efficiency in *Brucella* DNA extraction by reducing sample processing time and reducing the risk of cross contamination between samples. This finding support previous research done by AL-Garadia et al. (2011), Khamesipour et al. (2013) and Karthik et al. (2014).

The antibody level and the number of the bacteria in the circulation were affected by the infection status of the animals at the time of specimen collection (Moussa et al., 2011; Raghava et al., 2017). *Brucella* is a facultative intracellular microorganism. In the early stage of infection, high concentrations of *brucella* were in the blood circulation. As a consequences, a good quantity of DNA can be easily extracted and give positive result for PCR analyses. While during chronic condition most *brucella* will hide intracellularly which make the diagnosis more challenging and make the whole blood become sample of choice (Islam et al., 2018). All dairy cattle in this study were seronegative but were positive molecularly. This condition was probably related to the number of *brucella* in the circulation was not enough to trigger off the antibody level that can be detected serologically but still enough to be amplified by PCR.

In this study, time difference between the disease incidence and the time of blood collections for each brucellosis positive animals were 1 week to 6 weeks. The degree of *brucella* infection in the blood was affected by the susceptibility of the animal against the organism (Manthei et al., 2015). A high persistence of bacteremia was demonstrated in highly susceptible animal. Whereas, low persistence of bacteremia correlated with animals that have varying degree of resistance was observed. In
experimentally infected cattle with virulent \textit{B. abortus}, the bacteria could be found in the blood circulation for as long as ninety-seven weeks (Manthei and Carter, 2010).

Based on phylogenetic analysis (Fig. 3) 1 positive sample in this study (BRU_AB_1018) shared high similarity (98\% homology) with the representation of the \	extit{Brucella abortus} genome strain accession number CP000999 isolated from bovine in Zimbabwe (Ledwaba et al., 2014). Two other positive samples (BRU_AB_2118 and BRU_AB_2218) has 100\% homology with \textit{Brucella abortus} strain 9810 chromosome 1 and \textit{B. abortus} strain 84573 chromosome 2 isolated from water buffalo in Italy (Paradiso et al., 2017). Whereas another positive sample (BRU_AB_0818) was related to \textit{Brucella abortus} strain Wisconsin genome assembly, chromosome 2 (Anonymous NCBI, 2018). Previous research has reported that based on omp2a gene, \textit{B. abortus} isolated from beef cattle in South Sulawesi and East Nusa Tenggara, Indonesia, were closely related to isolates from France (AY008719) and India (FN552417, FN552432, FN552430) (Ratnasari et al., 2014). This difference is likely to be due to the amplification of different nucleotide gene fragments. In order to prove the existence of genetic diversity among \textit{Brucella} sp, in Indonesia further research involving higher amounts of \textit{Brucella} positive samples from different regions of Indonesia needs to be carried out immediately, so that the economic losses incurred can be minimized and public health can be guaranteed.

Conclusions: It could be concluded that PCR has successfully confirmed the presumptive diagnosis of brucellosis based upon the clinical consequences of having reproductive disorders in seronegative dairy cattle using whole blood samples. The application of the PCR technique for routine diagnosis of brucellosis in seronegative animals with reproductive disorders was therefore advisable.

Acknowledgments: The authors greatly acknowledge to the Direktorat Riset and Pengabdian Masyarakat Direktorat Jenderal Penguatan Riset and Pengembangan, Kementrian Riset, Teknologi, dan Pendidikan Tinggi and Gadjah Mada University for financial support through Penelitian Dasar Unggulan Perguruan Tinggi, Gadjah Mada University No.195/UN1/DITLIT/DIT-LIT/2018 Tanggal 5 Maret, 2018. The authors also very grateful for the thoughtful advice given by Prof. Dr. Roger K. Maes. Michigan State University, East Lansing, MI, USA during the preparation of this manuscript.

Authors contribution: S played a valuable role in blood sampling in the field. RW had responsibility in preparing and extracting the DNA from whole blood and serum samples. HW had role in doing the PCR analysis and sequencing. All researchers together discussed and compiled manuscripts for journal publications.

REFERENCES

Al-Garadi MA, Khairani-Bejo S, Zuniga Z, et al., 2011. Detection of \textit{Brucella melitensis} in blood samples collected from goats. J Anim Vet Adv 10:1437-44.

Anonymous OIE, 2009. OIE Territorial Manual: Bovine Brucellosis. version adopted by the world assembly of delegates of the OIE in May 2009.

Anonymous, NCBI, 2018.

Bailey GG, Krahm JB, Drasar BS, et al., 1992. Detection of \textit{Brucella melitensis} and \textit{Brucella abortus} by DNA amplification. J Trop Med Hyg 95:271-5.

Doust 3RH, Ahmadi Z, Ahamdi A, et al., 2007. Detection of \textit{Brucella abortus} by alkB and IS711 based primers. J Res Med Sci 12:62-7.

Ducrotoy M, Bertu WJ, Matope G, et al., 2017. Brucellosis in Sub-Saharan Africa: Current challenges for management, diagnosis and control. Acta Tropica 165:179-93.

El-Diasty M, Wareth G, Melzer F, et al., 2018. Isolation of \textit{Brucella abortus} and \textit{Brucella melitensis} from seronegative cows is a serious impediment in brucellosis control. Vet Sci 5:23-31.

Geresu MA and Kassa GM, 2016. A review on diagnostic methods of \textit{Brucellosis}. J Vet Sci Techno 7:1-8.

Gupta VK, Nayakwadi S, Kumar A, et al., 2014. Markers for the molecular diagnosis of brucellosis in animals. Adv Anim Sci 2:31-9.

Islam Md S, Islam Md A, Khazun Mst M, et al., 2018. Molecular detection of \textit{Brucella} spp. from milk of seronegative cows from some selected area in Bangladesh. J Pathogen Article ID 9378976 1-8.

Karshik K, Rathore R, Thomas P, et al., 2014. Serological and molecular detection of \textit{Brucella abortus} from cattle by RBPT, STAT and PCR and sample suitability of whole blood for PCR. Asian J Animal Vet Adv 9:262-9.

Khamesipour F, Doost A and Taheri H, 2013. Molecular detection of \textit{Brucella} spp. in the semen, tests and blood samples of cattle and sheep. J Pure Appl Microbiol 7:495-500.

Kumar S, Steetch G, Tamura K, et al., 2016. Mega 7. Molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol 3:1870-4.

Ledwaba B, Mafolo J, vanHeerden H, et al., 2017. Genome sequences of \textit{Brucella abortus} and \textit{Brucellussuis} strains isolated from bovine in Zimbabwe. Genome Announc 5:e00106-13.

Mahajan V, Banga HS, Fila G, et al., 2017. Comparison of diagnostic tests for the detection of bovine brucellosis in the natural cases of abortion. Iran J Vet Res 18:183-9.

Manthei KA and Carter RW, 2010. Persistence of \textit{Brucella abortus} infection in cattle. Vet Res 21:183-90.

Manthei KA, Hill MC, Burke JE, et al., 2015. Persistence of \textit{Brucella abortus} infection in cattle. Front Natl Acad Sci USA 112:4292-7.

Moussa IM, Omnia ME, Amin AS, et al., 2011. Evaluation of the currently used polymerase chain reaction assays for molecular detection of \textit{Brucellos} species. Afr J Microbiol Res 5:1511-20.

Paradiso R, Orsini M, Camma C, et al., 2017. Complete genome sequencing of eight \textit{Brucella abortus} biovar 1 strains isolated from water buffalo. Genome Announc 6:e00179-18.

Perez-Sanco M, Garcia-Seco T, Arrogante L, et al., 2013. Development and evaluation of an IS711-based loop mediated isothermal amplification method (LAMP) for detection of \textit{Brucella} spp on clinical samples. Res Vet Sci 95:489-94.

Priyadarshini A, Sarangi LN, Palai TK, et al., 2013. Brucellosis in cattle and occupationally exposed human beings: A serosurveys in Odisha, India. J Pure Applied Microbiol 7:3255-60.

Raghava S, Gowda MM, Shome R, et al., 2017. Epidemiological and molecular characterization of \textit{Brucella} species in cattle. Asian J Anim Sci 11:123-31.

Ratnasari R, Suwarno S, Rantam FA, et al., 2014. Homology and phylogenetic analysis of nucleotide sequence of \textit{omp2a} gene of \textit{Brucella abortus} local isolates. J Vet Adv 4:374-80.

Silva Moi JP, Araujo Franca S, Paixoa TA, et al., 2012. Laboratory diagnosis of animal brucellosis. R Bras Ci Vet 19:117-26.