Inductive effects in amino acids and peptides: Ionization constants and tryptophan fluorescence

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ABSTRACT

Although inductive effects in organic compounds are known to influence chemical properties such as ionization constants, their specific contribution to the properties/behavior of amino acids and functional groups in peptides remains largely unexplored. In this study we developed a computationally economical algorithm for ab initio calculation of the magnitude of inductive effects for non-aromatic molecules. The value obtained by the algorithm is called the Inductive Index and we observed a high correlation ($R^2 = 0.9427$) between our calculations and the $pK_a$ values of the alpha-amino groups of amino acids with non-aromatic side-chains. Using a series of modified amino acids, we also found similarly high correlations ($R^2 > 0.9600$) between Inductive Indexes and two wholly independent chemical properties: i) the $pK_a$ values of ionicizable side-chains and, ii) the fluorescence response of the indole group of tryptophan. After assessing the applicability of the method of calculation at the amino acid level, we extended our study to tryptophan-containing peptides and established that inductive contributions of neighboring side-chains are transmitted through peptide bonds. We discuss possible contributions to the study of proteins.

1. Introduction

Inductive effects are defined as the polarization of a bond by electronegativity differentials between the atoms of adjacent bonds. These effects propagate through the bond structure of the molecule and are distinct from electrostatic or field effects, which are by definition not transmitted through the chemical bonds. Inductive effects were described in detail, albeit empirically, already in the 1930’s and what was then established was incorporated into advanced organic chemistry textbooks [1]. Inductive effects have been long recognized to play a central role in the ionization of chemical groups and in many other chemical properties. Although the measurement, and more importantly, prediction of ionization constants of functional groups in peptides and proteins is an essential aspect of protein research, the importance of inductive effects remains largely unexplored.

The development of formal methods to calculate the magnitude of inductive effects was attempted most notably in the classical works of Hammett [2–4] and Taft [5–7]. Taft studied the variation of acid dissociation constants of simple non-aromatic carboxylic acids and amines substituted with different atoms. These careful studies established that electron-withdrawing substituents had the effect of favoring acid dissociation (lowering $pK_a$) of the reporter groups even in non-aromatic (non resonant) molecules. Taft’s investigations were semiempirical, that is, they could not provide an ab initio method of calculation of the magnitude of the effect on the measured property but they established two general features of inductive effects: i) they depend on the electronegativity of the substituent and ii) they depend on its distance -in number of intervening bonds-from the group being measured. Hammett had applied a similar outlook to the study of some aromatic molecules, most particularly substituted benzoic acids. He demonstrated that substituents of varying electronegativity influenced both the ionization constants and reactivities of the reporter groups in a
manner much like the one described by Taft, although the relatively simple concept of distance (in number of bonds from the reporter group) necessarily incorporated the notion of position of the substituent in the phenyl ring due to the resonance or mesomeric effect.

A significant step was taken in the early 1960s, when a general ab initio method for calculating the magnitude of inductive effects in non-ionicizing molecules of, in principle, any size, was reported by Chiang and Tai [8]. Their calculation method took into account the electronegativity of all atoms in the molecule (in the Pauling scale), bond lengths, formal charges and the number of intervening bonds between the polarizing bond and the (inductively) polarized bond under study (e.g., the O-H bond in a carboxylic acid). They called the number provided by their method the Inductive Index and tested it in a very large number of molecules and chemical properties, finding a very good agreement between their calculations and the measured chemical parameters. Since then, however, the concept of electronegativity in its relation to the state of hybridization of the atoms in the bond has been much improved [9,10] introducing the possibility of further refinement of Chiang and Tai’s work.

From a theoretical perspective, all α-amino acids fit the general scheme of the classic works of Taft and Hammett, that is, they all have an identical basic structure (the backbone) and a substituent (the side-chain) which confers to each its chemical identity. Therefore, if inductive effects are relevant at this level of description, it is not unreasonable to expect that differences in the ionization of the α-carboxyl and α-amino groups of amino acids are a function of the inductive contributions -as approached by Chiang and Tai- of the side-chain groups. In fact, if this is true, there should exist a direct correlation between the ionization constants of the α-carboxyl and α-amino groups of α-amino acids with non-ionizable side-chains, i.e. the side-chain does not ionize in the pH interval between the ionizations of the α-carboxyl (−2) and the α-amino groups (−9). This correlation would not be expected in pK_a values of the α-groups of those amino acids in which the side-chain ionizes and therefore its inductive contribution changes between both α-group ionizations. However, in the course of our research we found a surprising level of inconsistency in the pK_a values reported through the years, probably reflecting different experimental conditions used to measure them (titrants, amino acid concentration) and methods of calculation [11-14].

In the case of ionizable side-chains, the same reasoning should hold true: their pK_a values should respond to the inductive contribution of the rest of the molecule. This includes the effects of both the state of protonation of the α-groups and their modification by, for example, amidation, esterification, acetylation or even peptide bond formation, which should alter their inductive contributions. This behavior should also extend to side-chains in peptides, that is, specific inductive contributions of formal charges present in terminal α-groups and, more generally, to the contribution of neighboring residues with particular side-chains, to the chemical properties of any given side-chain in a sequence.

Since the pioneering work of Gregorio Weber on the fluorescent properties of aromatic amino acids, most notably tryptophan and tyrosine [15], it has been known that their fluorescence closely follows the state of ionization of the α-groups. It increases when these groups deprotonate and decreases when they protonate so closely that their ionization constants can be determined from changes in fluorescence intensity upon titration [16]. The mechanisms proposed, however, have not generally given careful consideration to inductive effects, save passing references [17-19]. When the fluorescent group is part of a peptide, similar effects of ionization on fluorescence have been reported.

In glycy-tryptophan peptides in which the terminal amino group is progressively removed from the tryptophan by addition of an increasing number of glycyl residues two things are apparent: First, the effect of protonation/deprotonation of the terminal amino group on fluorescence intensity decreases with distance. Second, the change in fluorescence that accompanies terminal amino group deprotonation is considerably smaller and follows the altered pK_a of the terminal group [20]. In this particular reported case, the effect of a progressively more removed electron withdrawing group (a protonated amino) was studied by the authors. Although field effects have been generally favored as an explanation for these observations, an inductive interpretation as described above is also consistent with the data. Inductive effects, albeit less marked, should also be exerted by side-chains with different inductive contributions on neighboring amino acids. In other words, changing the amino acid adjacent to the tryptophan with a different residue -all other things equal-should be reflected in its fluorescence properties due to changes in inductive contributions.

In this study we endeavored to: i) refine the method of ab initio calculation of Inductive Indexes originally proposed by Chiang and Tai to extend it to amino acids and peptides; ii) correlate the calculated values with measured differences in the pK_a of ionizable groups of amino acids and some modified derivatives; iii) establish whether the results of the calculations correlate adequately with differences in fluorescence properties of the indole group in tryptophan and some model peptides containing tryptophan. The implications of our findings are discussed in the context of the current understanding of peptides and proteins.

2. Materials and methods

2.1. Amino acids and potentiometric titration reagents

All standard L-α-amino acids were analytical standards from Sigma-Aldrich. Modified amino acids were the highest available purity from Sigma-Aldrich and Research Organics. Water was twice-distilled crystallization grade. Titrants (0.1 N NaOH or 0.1 N HCl) were standardized solutions from JT Baker. Standard pH buffers for electrode calibration from Hannah (HI 6000 series: pH 1.679, pH 4.010, pH 6.862, pH 7.010, pH 10.010).

2.2. Peptides and reagents for fluorescence measurements

Peptides were obtained from JPT Peptide Technologies (Berlin, Germany) and were of a purity > 95%. All peptides had Trp as fluorophore, the N terminus acetylated and the C terminus amidated. The tetrapeptides had Gly as the linking amino acid.

2.3. Determination of ionization constants of α-groups and ionizable side-chains of unmodified and modified amino acids by potentiometry

Titrations were conducted under rigorously identical conditions in a Metrohm 853 automatic titration unit equipped with a water jacketed titration vessel for temperature control (25 °C ± 0.1 °C), magnetic stirring and a Metrohm Biotrode pH electrode. Initial volume was 10 ml in all cases; titrations were performed under a stream of humid nitrogen and using a CO_2 trap. Electrode drift was stabilized to <0.5 mV/min and the electrode was calibrated each day with five buffer standards. For titration of α-amino groups and ionizable side-chains, amino acids were initially dissolved at 20 mM in 0.1 N HCl and then titrated with 0.1 N NaOH; ionization constants (expressed as pK_a) were determined by analysis of the titration curve by standard second derivative calculations implemented in the software supplied with the titration system. In the case of ionization constants of α-carboxyl groups, amino acids were initially dissolved in distilled water at 20 mM and titrated by sequential addition of 0.1 equivalents of titrant, in this case 0.1 N HCl. The experimental methods and calculations were performed as described in detail in Albert and Serjeant [21]. All titrations were replicated at least three times on different days and with two independent batches for most standard amino acids.

2.4. Fluorescence measurements

Fluorescence lifetime measurements were performed in a Chronos...
ISS (Champaign, IL, USA) Multifrequency phase shift and modulation fluorimeter, using a 280 nm LED (ISS model 90101) for excitation and p-terphenyl in ETOH (τ = 1.05 ns) as a reference. To eliminate scatter from excitation light the fluorescence emission was collected through a WG320 long-pass filter. Data were collected until the standard deviation from each measurement of phase delay (ϕ) and modulation (M) were at most 0.2° and 0.004 respectively. Diode modulation frequency was ranged logarithmically from below to above the crossover of ϕ and modulation M in at least 8 steps (usually between 20 MHz and 200 MHz), and analyzed for each peptide using the Vinci software (ISS) assuming a single lifetime with a Lorentzian distribution (both lifetime and width being variables). χ² scores were all below 1.40 and generally below 1 (see Table S3). A special case is given for tryptophan amide which was the only compound which showed two lifetimes (1.05 ns) as a reference. To eliminate scatter from multiphoton excitation light the fluorescence emission was collected through a δn multiple frequencies phase shift and modulation.

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2.5. Calculation of inductive indexes

2.5.1. Chiang and Tai’s algorithm

The original algorithm depends on three parameters: the electronegativity (in the Pauling scale) of all atoms involved in the calculation, the bond lengths between them and an arbitrary parameter 1/α called the transmission factor (see below). For a hypothetical molecule.

\[
i = \frac{1}{\alpha} \sum (\delta_i) + \left(\frac{1}{\alpha}\right)^2 \sum (\delta_i)^2 + \left(\frac{1}{\alpha}\right)^3 \sum (\delta_i)^3 + \ldots + \left(\frac{1}{\alpha}\right)^n \sum (\delta_i)^n
\]

where \(\frac{1}{\alpha}\) is called the transmission factor and α is equal to 2.7; n is the bond order, that is, the distance in terms of covalent bonds from A (for example, B-C4 has order 1, C1-D1 has order 2 and A-B has order zero), \(\sum (\frac{1}{\alpha})^n\) is the sum of all the \(\frac{1}{\alpha}\) of order n.

Finally, equation (3) defines the sum of the contributions of formal charges as

\[
i_\pm = (\pm N_i/r_\pm) + \frac{1}{\alpha} \sum (\pm N_i/r_\pm)_1 + \left(\frac{1}{\alpha}\right)^2 \sum (\pm N_i/r_\pm)_2 + \left(\frac{1}{\alpha}\right)^3 \sum (\pm N_i/r_\pm)_3 + \ldots
\]

\[+ \left(\frac{1}{\alpha}\right)^n \sum (\pm N_i/r_\pm)_n\]

where \(r_\pm\) is the bond length in the ionized group, and \(\pm N_i\) can be equal to 1, -1 or 0.

2.5.2. Modified CT algorithm (MCT)

Our method of calculation modified Chiang and Tai’s algorithm in several ways. First, we did not include bond distances in the calculations. Second, while we maintained the concept of polarity index (δ), we used values of electronegativity not of atoms in their ground state, but as they appear in molecules, that is, considering their valence state based on the concept of orbital electronegativity as developed by Hinze and Jaffé [9], which, unlike the Pauling scale of electronegativity, is contemporaneous to the publication of Chiang and Tai’s opus. Third, the transmission factor \(\left(\frac{1}{\alpha}\right)\) was 0.5263 (α = 1.9). Fourth, formal charges were uniform for all calculations (amino acids and peptides) but were not unitary: positive charges had a value of +1.2 and negative charges of -0.9 (see Supplementary Information for a detailed discussion of value selection). Thus, the final expression of inductive index (i), or MCT, is as follows:

\[ii = i_0 + i + i_\pm\]

Where

\[i_0 = \delta_{AB},\]

\[i = \frac{1}{\alpha} \sum (\delta_1) + \left(\frac{1}{\alpha}\right)^2 \sum (\delta_1)^2 + \left(\frac{1}{\alpha}\right)^3 \sum (\delta_1)^3 + \ldots + \left(\frac{1}{\alpha}\right)^n \sum (\delta_1)^n,\]

and

\[i_\pm = (\pm N_i)_0 + \frac{1}{\alpha} \sum (\pm N_i)_1 + \left(\frac{1}{\alpha}\right)^2 \sum (\pm N_i)_2 + \left(\frac{1}{\alpha}\right)^3 \sum (\pm N_i)_3 + \ldots + \left(\frac{1}{\alpha}\right)^n \sum (\pm N_i)_n.\]

Initial calculations of inductive indexes, graphs and linear regressions for individual amino acids were done using the LibreOffice Calc spreadsheet. Both graphs and regressions were later repeated using QtiPlot (GPLv2). The Inductive Index calculated was always that exerted on the Cα of the amino acid under scrutiny. When examining ionization of α-NH₃⁺ groups, the value was calculated by the structure of the side-chain alone, considering that α-COOH is already deprotonated and, therefore, its inductive contribution is fixed for all α-amino acids. When examining the behavior of ionizable side-chains, the inductive index on Cα calculated was that exerted by the terminal α-groups, whether ionized or covalently modified.

Calculations of inductive index on the Cα of Trp in peptides incorporate: i) the contributions of all backbone elements in the chain; ii) the contribution of all side-chains except the Trp side-chain, where the state
of ionization of ionizable side-chains is specified, and; iii) terminal
groups, whose state of ionization or covalent modification (N-acetyla-
tion, C-amination) is specified as well. All values calculated in
the appropriate bond order with respect to the corresponding Cα (See Sup-
plementary Information).

3. Results

3.1. Inductive index and ionization constants

If inductive effects of the side-chains are indeed the main factor
determining the differences in the ionization constants (as pKα) of
ionizable α-groups of α-amino acids, then it is necessarily the case that
there must be a positive correlation between the pKα values of the
α-amino and α-carboxyl groups of all twelve α-amino acids with non-
ionizable side-chains. To test this prediction, we carefully and consis-
tently determined their pKα values, which are summarized in Table 1.

Using this self-consistent dataset of ionization constants we found a
very significant correlation (R² = 0.9320) between ionization of the
α-amino and α-carboxyl groups in each amino acid, as shown in Fig. 1.

The outstanding correlation between the pKα values of α-groups of
amino acids lent tentative support to an explanation considering
inductive effects as a primary cause.

In fact, if these are not considered there is no hint of an explanation of
why this proportionality should exist at all (or why there are differ-
ences in the ionization constants of α-groups in the first place). We then
proceeded to calculate Inductive Index values (as a measure of the
magnitude of inductive effects exerted on the Cα by the side-chain) for
all non-aromatic α-amino acids using our modified Chiang and Tai al-
gorithm (MCT). The results are shown in Table 2.

When the calculated values are plotted against the measured pKα
values of the α-amino α-values of non-ionizable
α-amino acids using our modified Chiang and Tai al-

Table 1

| Amino Acid | α-NH₂ | α-COOH |
|------------|------|-------|
| Ala        | 9.80 ± 0.02 | 2.40 ± 0.04 |
| Val        | 9.55 ± 0.01 | 2.33 ± 0.05 |
| Gly        | 9.68 ± 0.01 | 2.40 ± 0.04 |
| Ile        | 9.69 ± 0.01 | 2.38 ± 0.04 |
| Leu        | 9.64 ± 0.01 | 2.35 ± 0.04 |
| Glu        | 9.15 ± 0.01 | 2.22 ± 0.05 |
| Met        | 9.15 ± 0.02 | 2.24 ± 0.05 |
| Ser        | 9.11 ± 0.02 | 2.25 ± 0.05 |
| Thr        | 9.01 ± 0.01 | 2.23 ± 0.05 |
| Asn        | 8.80 ± 0.02 | 2.19 ± 0.05 |
| Phe        | 9.15 ± 0.03 | 2.23 ± 0.05 |
| Trp        | 9.42 ± 0.02 | 2.35 ± 0.04 |

Fig. 1. Plot of pKα (±SD) values of α-amino (ordinate) against α-carboxyl (abscissa) groups for all α-amino acids with non-ionizable side-chains and linear regression line. The larger error bars of the pKα values of the α-carboxyl groups are a consequence of the greater concentration of hydrogen ions (and thus total ionic strength) at the point of ionization (discussed at length in Albert and Sergeant [21], whose method we followed).

Table 2

| Side chain | α-NH₂ | pKα |
|------------|------|-----|
| Cys (thiol ionized) | −339.6 | 10.33 ± 0.02 |
| Asp (β-carboxyl ionized) | −162.5 | 9.76 ± 0.02 |
| Ala | −134.0 | 9.80 ± 0.02 |
| Val | −127.1 | 9.55 ± 0.01 |
| Ile | −96.8 | 9.69 ± 0.01 |
| Glu (γ-carboxyl ionized) | −91.3 | 9.74 ± 0.01 |
| Leu | −72.8 | 9.64 ± 0.01 |
| Gly | −53.2 | 9.68 ± 0.01 |
| Met | 19.7 | 9.15 ± 0.02 |
| Arg (guanidinium ionized) | 46.5 | 9.08 ± 0.01 |
| Lys (ε-amino ionized) | 62.8 | 9.16 ± 0.01 |
| Ser | 81.5 | 9.11 ± 0.02 |
| Gln | 89.3 | 9.15 ± 0.01 |
| Thr | 92.3 | 9.01 ± 0.01 |
| Asn | 180.8 | 8.80 ± 0.02 |

Table 3

| Side chain | pKα (±SD) |
|------------|----------|
| His (R² = 0.9978) | 3.42 ± 0.02 |
| Asp (R² = 0.9869) | 3.23 ± 0.01 |
| Cys (R² = 0.9670) | 3.35 ± 0.03 |

removal of both charges on the pKα of the side-chain. The Inductive
Index calculations incorporated the structures of the covalent modifi-
cations. The results of the calculations and measurement of side-chain
pKα values are listed in Table 3.

A plot of measured side-chain pKα values against calculated Induc-
tive Index on the Cα of each modified or zwitterionic amino acid is
shown in Fig. 3.

Several things are noteworthy. First, the linear regressions show very
significant correlations between calculated α and measured pKα values:
His (R² = 0.9978), Asp (R² = 0.9869), Gln (R² = 0.9802) and Cys (R² =
0.9670). Second, removal of the positive charge in the α-amino group by
N-acetylation results in the greatest side-chain pKα in all amino acids,
and removal of the negative charge in the $\alpha$-carboxyl group in the lowest, with the zwitterion and the doubly modified amino acids giving intermediate values, with the zwitterion giving consistently a slightly lower pK$_a$ than the doubly-modified amino acid. Third, the effect of the modifications is variable, that is, the change in pK$_a$ with $ii$ value (the slope) varies for each amino acid side-chain. Fourth, there is a difference in slope between the $\beta$-carboxyl group of Asp and the $\gamma$-carboxyl group of Glu (see Discussion). Finally, in the case of His, where two different modifications of the $\alpha$-carboxyl group were available (amidation and methyl esterification), the calculation of Inductive Index discriminates between the two values of measured pK$_a$. These points will be taken up in more detail in the Discussion.

| Side chain pK$_a$ | $ii$ on C$_\alpha$ (Asp, Glu, Cys, and His) | $\tau$ (ns) of Trp |
|------------------|-----------------|------------------|
| Zwitterion        | 653.0           | 3.81 ± 4.27 ± 6.02 ± 8.27 ± 2.55 |
| N-Ac              | –338.2          | 4.72 ± 4.83 ± 6.98 ± 9.66 ± 4.24 |
| (α-COO)           | 1305.2          | 2.94 ± 3.88 ± 5.35 ± 1.44 |
| C-NH$_2$ (α-NH$_2$) | – –             | 5.26 ± 6.58 ± – |
| C-OMe (α-NH$_2$)  | 314.0           | 4.01 ± 4.34 ± 0.01 ± 0.01 |
| N-Ac/C-NH$_2$     | 394.6           | 4.01 ± 4.34 ± – |
| N-Ac/C-OMe        | 304.9           | 4.01 ± 4.34 ± – |
| N-Ac/C-NHMe       | – –             | 8.78 ± – |

Fig. 2. Plot of the pK$_a$ (±SD) values of the $\alpha$-amino groups of all non-aromatic $\alpha$-amino acids against calculated Inductive Index value ($ii$, on C$_\alpha$). In the case of amino acids with ionizable side-chains, the state of ionization of the side-chain (as considered for calculation of $ii$) at the pK$_a$ of the $\alpha$-amino group is indicated.

Fig. 3. Measured side-chain pK$_a$ (±SD) against calculated Inductive Index value ($ii$) on the C$_\alpha$ of each modified or zwitterionic amino acid: Cys (circles), His (squares), Glu (triangles) and Asp (inverted triangles). Z is the unmodified amino acid in its zwitterionic form and N-Ac indicates $\alpha$-amino acetylation. C-NH$_2$ and C-OMe indicate $\alpha$-carboxyl amidation and methyl esterification, respectively. Doubly modified amino acids are indicated as a combination of modifications (Ac/NH$_2$, Ac/OMe). In the case of doubly modified His, the $\alpha$-carboxyl group was N-methyl amidated (C-NHMe). When an $\alpha$-group was not modified, it was assumed to be charged at the pH of deprotonation of the side-chain groups, indicated in parentheses. Note that the calculated Inductive Indexes on C$_\alpha$ do not include the side-chain itself, therefore, they are identical for all identical modifications and in the zwitterion form. (–) indicates modification not available for that amino acid.

Fig. 4. Correlation of Mean Fluorescence Lifetime ($\tau$) of the indole group of Trp with Inductive Index ($ii$) values for the zwitterion (Z) and covalently modified $\alpha$-groups ($R^2 = 0.9678$). Modifications, as well as $ii$ and lifetime values are those listed in Table 3. N-Ac indicates N-acetyl-Trp, C-NH$_2$ indicates Trp-amide and Ac/NH$_2$ indicates N-acetyl-Trp-amide (NATA).
3.2. Inductive index and tryptophan fluorescence

Fluorescence of the indole group of tryptophan is known to be strongly affected by the state of protonation of its α-groups. Its intensity increases with increasing pH during titration as each of the α-groups deprotonate. We used the same covalent α-group modifications in tryptophan to examine the correlation between Inductive Indexes and Trp fluorescence as Mean Fluorescence Lifetime (denoted by τ). We chose to measure fluorescence lifetime because it is a more robust measure than fluorescence intensity alone, as it is independent of fluorophore concentration. Measured values are listed in the last column of Table 3 and a plot of τ against Inductive Index is shown in Fig. 4.

Our next step was to examine whether inductive effects, as calculated by us, propagate through peptide bonds to a neighboring amino acid. To this end we examined a series of peptides containing tryptophan. We calculated the Inductive Index value on the Cα of the Trp residue of each peptide and measured their fluorescence (as τ). The values are listed in Table 4.

Two sets of peptides were used: the first consisted of dipeptides in which we varied the amino acid adjacent to the Trp residue (Asn, Val, Met and Ala). The second set consisted of tetrapeptides in which the distance of either aspartic acid (a strongly electron-donating residue when ionized) or asparagine (a much weaker electron donor) was varied by increasing the number of intervening glycyl residues. The plot of Mean Fluorescence Lifetime against inductive index of all peptides is shown in Fig. 5, top panel.

4. Discussion

4.1. Inductive index and the behavior of ionizable groups of amino acids

Surprisingly (to us), reported pKa data for many molecules—including amino acids—are quite inconsistent across standard reference sources [22], even when reportedly obtained at the same temperature and in comparable experimental setups [11–14]. Therefore, when we decided to explore the relevance of inductive polarization of bonds on the behavior of ionizable groups of amino acids, we found ourselves in need of obtaining a consistent dataset of pKα values to work with. Obtaining the data consistently involved an experimental component and a calculational one. Experimentally, we made use of carefully controlled conditions, electrodes and reagents for potentiometric titration of ionizable groups. For calculational consistency, we avoided making any suppositions about the samples, staying well below the limits of ionic strength that would be likely to exert a measurable influence on the ionization of the measured groups [21].

α-amino acids are well suited to an approach such as the classical one originally developed by Hammett: they have a constant structure with two ionizable sites (the α-amino and α-carboxyl groups) and a variable site (equivalent to Hammett’s “substituent”), namely, the side-chain. Therefore, a straightforward consequence of the effect of polarization (induction) by a non-ionizable side-chain on the ionizable α-groups is that it should affect both of them equally, that is, an electron-donating side-chain will lower the pKα of both ionizable α-groups, and an electron-donating one should do the opposite. Fig. 1 shows that this is indeed the case for all α-amino acids with non-ionizable side-chains: those with more acidic α-carboxyls have proportionally more acidic α-amino groups. This correlation has not been hitherto apparent from reported values in the literature, probably due to the inconsistencies mentioned above. Using four independent references correlation (as R2) between the pKα values of α-amino and α-carboxyl groups are 0.42 [14], 0.64 [12], 0.697 [11] and 0.72 [13].

A corollary is that side-chains that are in a different chemical state (protonated/deprotonated) between α-carboxyl and α-amino ionizations would be expected to exert different inductive contributions for each chemical state, and therefore result in non-proportional pKα values. This is indeed the case, as illustrated by Cys (α-carboxyl pKα of ~1.8 -one of the lowest- and α-amino pKα of ~10.3, the highest) but also for aspartic acid, glutamic acid and histidine.

While inductive effects have been understood, or at least considered, for small organic molecules for a long time, it is remarkable that they have not, to the best of our knowledge, been incorporated into the mainstream of current thinking about amino acids, peptides and, more importantly, proteins. Although the classic semi-empirical work of Hammett remains relevant and is used for pKα prediction in small molecules [22], in peptides and proteins pKα prediction is dominated by the use of models that are both semi-empirical and computationally demanding, and statistical models that need large data sets for training [23].

The work of Chiang and Tai [8] provided, for the first time, an ab initio method to calculate a quantity (the Inductive Index) that correlated very well with ionization constants of a wide array of compounds and also with a range of observed chemical properties. It was also a crucial first step, from a theoretical standpoint, to discriminate the effect of one chemical group or moiety on another in terms of its inductive
that for equal structure the values calculated are equal (Table 3), but nonetheless correlated extremely well with changes in the pKa of very different chemical groups (carboxyl, thiol and imidazole) in widely different ranges of pH. It is also of note that the response (slope) of each ionizable group to inductive input is characteristic. This property probably reflects both the polarizability of the ionizable group (greatest for the thiol group in cysteine) and the number of bonds between the ionizable group and the backbone (the slope is almost twice for the β-carboxyl of aspartic acid than for the γ-carboxyl of glutamic acid), independently supporting our choice of transmissivity factor (1/α).

4.2. Inductive index and peptides

Having found significant correlations between Inductive Index values and ionization constants (as pKa) in amino acids, it was natural to explore whether these correlations extended to peptides, as one aim of this investigation was to extend the ab initio calculation of Inductive Indexes to peptides to establish whether they are useful to understand some of their properties. However, many peptides have a very limited solubility in water, especially if they are composed of relatively hydrophobic residues and termini are covalently modified, constraining our ability to perform accurate potentiometric titrations to determine ionization constants. To circumvent this experimental limitation, we took advantage of the well-established fact that the pKa of α-groups in fluorescent amino acids can be determined by the changes in fluorescence upon titration [15,16]. The possible contribution of inductive effects to their fluorescence has not, however, been quantitatively examined. To ascertain if there is a correlation between Inductive Index and tryptophan fluorescence we used the same strategy of Fig. 3, that is, we measured the Mean Fluorescence Lifetime (τ) in tryptophan as a zwiterion and in N- and C-covalently modified derivatives (N-acetyl-Trp, Trp-amide and N-acetyl-Trp-amide, Fig. 4). The correlation with Inductive Index values (which were the same calculated for identically modified amino acids with ionizable side-chains) was very significant (R² = 0.9678), indicating that electron donating - or withdrawing-groups, and therefore bond polarization, determine τ to a significant extent at the amino acid level.

We then calculated Inductive Indexes on the Cα of tryptophan in a series of peptides to determine whether our observations at the amino acid level were applicable to larger, and more relevant, molecules. First, we chose a series of dipetides and changed the amino acid adjacent to Trp (WA, WV, WM and WN). All peptides were N-acetylated and C-amidated for two reasons: i) to avoid a pH dependence of fluorescence and ii) to determine whether the inductive contribution of the side-chain of an adjacent amino acid sufficed to measurably change the fluorescence response of the Trp residue. Our results show that this is clearly the case, as shown in Fig. 5 (bottom panel). It is important to add that, in the case of the dipetides, the only difference in value on the Cα of tryptophan is due to the side-chain of the amino acid being varied since all other factors are constant. Therefore, the graph of μ value of each side-chain (A, V, M and N, Table 2) against τ should give an identical correlation to that shown in Fig. 5, which is indeed the case.

We next studied the effect of distance, in number of residues, from the amino acid being varied on the fluorescence of the Trp residue. To this end we designed a series of tetrapeptides in which either electron-donating (Asp) or withdrawing (Asn) residues were progressively moved away from the Trp residue (GGWX, GWGX and WGGX, where X is either Asp or Asn). The correlations for these tetrapeptides were also very significant as shown in Fig. 5 (bottom panel). For comparison purposes, the peptide GGWG was measured and included in the correlation of all the peptides against our calculations, which was also significant as shown in Fig. 5 (top panel).

Taken together, our results show a significant correlation between Inductive Index, a magnitude calculated solely from covalent structure and electronegativities, and two independent properties of amino acid side-chains: pKa of ionizable groups and fluorescence lifetime of the
5. Conclusions

The successful prediction of ionization constants of functional groups of biomolecules, small and large, remains one of the most important areas of biochemical interest, both from basic and applied perspectives [12,17]. Currently it is generally accepted that, in the case of proteins, the two key amino acid properties studied here, ionization (as $pK_a$) and Trp fluorescence (as $\tau$), are largely determined by the specific environments generated by the three dimensional structure the protein acquires in the folding process. It is reasonable to suppose, however, that other factors, hitherto relatively overlooked, exert significant influence over them. One such factor, as our results show for Trp fluorescence, is the propagation of polarization of bonds in groups of atoms of different electronegativity, which in the context of the constant backbone structure of peptides (and proteins) are located on side-chains.

Although our modified CT (MCT) algorithm is not a $pK_a$ predicting algorithm per se, the fact that there is a linear relation between $pK_a$ and Trp fluorescence with Inductive Index values implies that the latter capture an intrinsic property of molecules in terms of two of the most basic chemical parameters, namely, covalent structure and electronegativity. Inductive Index calculations are simple and straightforward; they require neither steric arguments nor the use of statistical models that rely on many observations/data points for training.

If inductive effects through the peptide bond are, as our study suggests, a factor to take into account in understanding the complex behavior of functional groups in peptides, and maybe proteins, there are important implications for our understanding of the factors that determine the functionality of groups. For example, active sites of enzymes or binding sites of peptide/protein ligands, many of which are highly pH dependent. To illustrate, Fig. 3 shows that the thiol group of cysteine is very sensitive to inductive effects, a fact that may comport consequences, for example, for the catalytic properties of cysteine proteinases or the formation of disulfide bonds during oxidative folding.

Polarization of the electron clouds of side-chain functional groups is a consequence of the inductive effects of the rest of the molecule ($\alpha$-groups in individual amino acids, or the sum of side-chains and backbone elements in peptides). Consequently, our results show that the amino acid sequence of a peptide determines a set of initial physicochemical conditions independent of steric effects and microenvironments that have a direct relation to the functional group. Our results suggest that the inductive contributions from the amino acid sequence of a protein modify the density of the electron cloud of the atoms responsible for the formation of intramolecular hydrogen bonding, in a way that is particular to each atom depending on the sequence of neighboring amino acids. This could have further implications on backbone-based models of protein folding, such as the one developed by Rose et al. [28].

5.1. Necessary refinements and future directions

The value of the transmission factor is a heuristically selected number. We believe that $\alpha$ is likely different for every bond and probably a function of $\alpha$ and $\beta$ depending on the electronegativity of the bonded atoms and the nature of the bond (as considered in the Hinze & Jaffe electronegativity scale). Nonetheless, our selected transmission factor constitutes an average that seems to capture the overall effect of distance on inductive polarization but a more detailed exploration is clearly necessary to provide a more accurate method of calculation and, eventually, a more complete explanatory theory.

Chiang and Tai stated that their method of calculation did not include aromatic molecules. It is important to extend the capacity of the modified algorithm to incorporate aromatic groups. The Inductive Index value for aromatic amino acid side-chains on their $C_6$, and therefore on vicinal amino acids, can as of now be obtained by interpolation of their measured $\alpha$-amino $pK_a$ in Fig. 2.

Proline presents a particular calculation challenge for two reasons: i) it is not clear to us at this point how to adapt the algorithm to accurately reflect its cyclic structure, (i.e. it does not have a basic-structure/substrat form like the $\alpha$-amino acids) and, ii) the $pK_a$ values of its ionizable groups are not proportional, as is the case of all non-ionizable $\alpha$-amino acids.

CRediT authorship contribution statement

Jesús Lara-Popoca: Software, Investigation, Methodology, Visualization, Writing - original draft. Henrik S. Thoke: Investigation, Methodology. Roberto P. Stock: Conceptualization, Methodology, Investigation, Resources, Supervision, Visualization, Writing - original draft, Writing - review & editing. Enrique Rudino-Pinera: Funding acquisition, Resources. Luis A. Bagatolli: Resources, Methodology, Supervision, Writing - review & editing.

Declaration of competing interest

No interests declared.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.bbrep.2020.100802.

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