Lipid metabolic gene-wide profile and signature of lung adenocarcinoma

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Research

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Abstract

Background

Lung cancer is a worldwide cancer with high morbidity and mortality. More and more evidence shows that the disorder of lipid metabolism is the key to the development of cancer, and analysis of lipid-related genes may lead to diagnosis and prognostic biomarkers related to lung cancer.

Methods

In this study, we performed the differentially expressed analysis of 1045 lipid metabolism-related genes between LUAD tumors and normal tissues in the TCGA-LUAD cohort. Then the bioinformatic analysis of DEGs was showed. PPI networks and cytoHubba APP determine hub genes. The association between hub genes and overall survival was evaluated by Kaplan-Meier Plotter. To predict the prognosis of LUAD patients, a nomogram was built, the nomogram was validated by another cohort (GSE13213).

Results

Finally, a total of 217 lipid metabolism-related DEGs were detected in LUAD. They were significantly enriched in Glycerophospholipid metabolism, fatty acid metabolic process, and Eicosanoid Signaling. Then we identified 6 hub genes through PPI network and cytoHubba, including INS, LPL, HPGDS, DGAT1, UGT1A6, and CYP2C9. The high expression of CYP2C9, UGT1A6, and INS, whereas low expressions of DGAT1, HPGDS, and LPL, were associated with worse OS for 1925 LUAD patients. Based on the nomogram, we found that the high-risk score group had a worse OS, and the validated cohort had the same result.

Conclusion

In conclusion, we generated a lipid metabolic transcriptome-wide profile of LUAD patients and found that significant lipid metabolic pathways were correlated with the LUAD. Furthermore, we constructed a signature of six lipid metabolic genes, which significantly associated with diagnosis and prognosis of LUAD patients. The gene signature can be used as a biomarker for LUAD.

Background

Lung cancer is the most commonly diagnosed cancer (11.6% of the total cases) and the leading cause of cancer death (18.4% of the total cancer deaths) in the world [1]. Among the subtype of lung cancers, adenocarcinoma is the most common histologic subtype of lung cancer in men and women posterior to the 1990s[2]. A 2005–2014 epidemiological survey from China showed that the proportion of adenocarcinoma increased from 36.4–53.5%, while the proportion of squamous carcinoma decreased.
from 45.4–34.4%[3]. The increasing incidence of lung adenocarcinoma (LUAD) has also been reported with air pollution-related factors[4–6].

The pathways of cancer patients were various[7]. Cancer caused by different pathways may require different treatments. One of the hallmarks of cancer is metabolic reprogramming. The importance of alterations related to lipid metabolism is starting to be recognized, and the increase in de novo lipogenesis is considered a new hallmark in many aggressive cancers[8]. Epidemiological data indicated that a certain number of lung cancer patients with high high-density lipoprotein cholesterol (HDL-C) and low low-density lipoprotein (LDL) and low-density lipoprotein receptor (LDLR) level have better survival in patients[9, 10]. Compared with healthy subjects, NSCLC patients showed significant increases in phosphatidylcholine (PCs) and phosphatidylethanolamine (PEs)[11]. Other lipid metabolism indicators associated with NSCLC includes sphingomyelins, phosphatidylinositol, phosphatidylserines, phosphatidylethanolamine, phospholipids, phosphatidylcholine[12]. The cancer cells’ requirement of metabolic intermediates for macromolecule production is overwhelming. Fatty acid oxidation (FAO) can help to generate ATP to support the membranes formation, energy storage, production of signaling molecules by coordinating the activation of lipid anabolic metabolism [13]. During the process, de novo adipogenesis in cancer increases. Lipid metabolism relating genes including FASN[14], ABCA1[15], ACLY[16], FABP4[17], CD36[18], SCD1[19] have been reported to be associated with morbidity, prognosis and treatment resistance of NSCLC.

Gene expression profiling is an essential part of bioinformatics, which has broad application prospects and powerful functions in oncology medicine. It has excellent clinical application potential in molecular diagnosis, tumor molecular screening, new target discovery, tumor response prediction, patient classification, and prognosis prediction [20]. To explore the further lipid mentalism relating regulation network and pathway, we conducted an integrated bioinformatic method to construct the gene-wide expression profile and a signature of lipid metabolism on LUAD. We explored the potential biomarkers for diagnosis and prognostic guidance of LUAD caused by lipid metabolism disorder.

**Materials And Methods**

**Patients and datasets**

We downloaded 519 lung adenocarcinoma (LUAD) tissues and 58 normal tissues with mRNA expression data from The Cancer Genome Atlas (TCGA, https://cancergenome.nih.gov/) database using the R package TCGAbiolinks [21]. The ensemble ID of TCGA samples was annotated with human genes GRCh38/hg38.

**Identification of lipid metabolism-related differentially expressed genes**

21 lipid metabolism-related pathways and five lipid metabolism-related gene sets were collected from the Kyoto Encyclopedia of Genes and Genomes (KEGG) web site (http://www.kegg.jp/blastkoala/) [22] and
the Molecular Signatures Database (MisDB) web site (https://www.gsea-msigdb.org/gsea/msigdb/index.jsp) [23], respectively. After removing the overlapped genes, a total of 1045 lipid metabolism-related genes were obtained. Lipid metabolism-related differentially expressed genes (DEGs) between LUAD tissues and normal tissues were screened through R package edgeR [24]. The parameters set for differential expression analysis were FDR < 0.05 and |log2 fold change| (logFC) > 1.

**Bioinformatic analysis**

We used the R package clusterProfiler to furtherly explore the biological significance of lipid metabolism-related DEGs [25]. In GO and KEGG analysis, FDR < 0.05 was considered a significant enrichment. Then we uploaded the DEGs that containing gene identifiers and corresponding FDR values and log2FC values into the IPA software (Qiagen). The “core analysis” function included in the software was used to interpret the DEGs.

**Protein-protein interaction network generation and hub genes analysis**

We built a protein-protein interaction (PPI) network of differentially expressed lipid metabolism-related genes using the Search Tool for the Retrieval of Interacting Genes (STRING, http://string-db.org/) database [26]. The combined score of ≥ 0.4 was the cut-off value. Cytoscape software (version 3.6.0) helped to visualize PPI networks [27]. According to 12 ranking methods in cytoHubba [28], an APP in Cytoscape, the top ten genes of each method were selected for overlap analysis, and the genes with the highest number of overlaps were used as hub genes.

**Survival analysis**

The overall survival (OS) analysis of hub genes was shown by Kaplan-Meier Plotter (http://kmplot.com/analysis/), which includes clinical data and gene expression information for 1925 lung cancer patients [29]. Then, information on the number of cases along with median values of mRNA expression levels, hazard ratios (HR) with 95% confidence intervals (CI), and log-rank P-values were extracted from the KM plotter webpage. Log-rank P-values < 0.05 were considered significant.

**Prediction model**

Based on the selected hub genes, we use the nomogram package of R (“rms“)[30] to develop a model to evaluate the prognosis of LUAD patients. Using the formula of the nomogram, we calculated the prognosis score of each patient. According to the score, patients are divided into a low-risk score group and a high-risk score group using the median classification method. The prognosis score was validated by the patients’ actual prognosis outcome. Then we did the same analyses on the external set (117 LUAD patients from GSE13213) to validate the availability of this 6-gene-based risk model.

**Results**
Identification and functional analysis of lipid metabolism-related DEGs

A total of 217 lipid metabolism-related DEGs were identified from the TCGA-LUAD cohort. A volcano plot was constructed to reveal the significant DEGs (Fig. 1A), and a heatmap was performed to show the hierarchical clustering analysis of the DEGs (Fig. 1B). To get an overall understanding of 217 lipid metabolism-related DEGs, we conducted GO terms and KEGG pathway enrichment by clusterProfiler package, while canonical pathways analysis by IPA. The results of KEGG pathway enrichment showed that DEGs were significantly enriched in glycerophospholipid metabolism, arachidonic acid metabolism, and metabolism of xenobiotics by cytochrome P450. In contrast, they were significantly enriched in fatty acid metabolic process, glycerolipid metabolic process, and steroid metabolic process from GO terms (Fig. 1C). IPA identified significant canonical networks associated with the DEGs. IPA showed that the top canonical pathways associated with common DEGs were Eicosanoid Signaling, FXR/RXR Activation, and Atherosclerosis Signaling (Fig. 1D).

PPI network construction and cytoHubba analysis

Lipid metabolism-related DEGs were analyzed by the STRING tool. Ultimately, a PPI network with 216 nodes and 1140 edges was established and visualized in Cytoscape (Fig. 2). Then a total of 6 hub genes were identified by the overlap of the top 10 genes according to 12 ranked methods in cytoHubba (Table S1). Moreover, these genes were related to Insulin (INS), Lipoprotein Lipase (LPL), Hematopoietic Prostaglandin D Synthase (HPGDS), Diacylglycerol O-Acyltransferase 1 (DGAT1), UDP Glucuronosyltransferase Family 1 Member A6 (UGT1A6), and Cytochrome P450 Family 2 Subfamily C Member 9 (CYP2C9).

Survival analysis of hub genes

We studied the relationship between mRNA expression of hub genes and clinical outcome using the Kaplan-Meier plotter. It was found that high expression of CYP2C9 [HR = 1.14 (1.00–1.29), P = 0.042], UGT1A6 [HR = 1.58 (1.34–1.87), P = 5.2e-08], and INS [HR = 1.16 (1.02–1.31), P = 0.026], whereas low expression of DGAT1 [HR = 0.75 (0.66–0.85), P = 6e-06], HPGDS [HR = 0.64 (0.56–0.73), P = 7.6e-12], and LPL [HR = 0.62 (0.55–0.71), P = 2.4e-13], were associated with worse OS for 1925 LUAD patients (Fig. 3).

Prediction model based on survival-related hub genes and validation

Based on the Cox regression model, a nomogram was built to predict the prognosis of LUAD patients, using the mRNA expression of the six survival-related hub genes (Fig. 4A). Then we calculated the prognosis score of each patient, and found that high-risk score group had worse OS of 3 years [HR = 1.51
We validated the model that high-risk score group had worse OS [HR = 1.84 (1.00–3.37), P = 0.047] (Fig. 4C).

**Discussion**

Metabolic change has been widely observed in cancer cells[31]. Among those metabolisms, lipid metabolism widely participates in the regulation of many cellular processes such as cell growth, proliferation, differentiation, survival, apoptosis, inflammation, motility, membrane homeostasis, chemotherapy response, and drug resistance[32]. Some recent researches have reported some component of PM2.5, promotes pulmonary injury by modifying lipid metabolism[33]. However, there are less researches regarding the association between transcriptome-wide lipid metabolism and lung cancer. Therefore, this study using the LUAD cohort to generate the transcriptome-wide profile of lipid-related.

Similar to the previous studies regarding other kinds of cancers[34], the fatty acid, glycerolipid, and glycerophospholipids were also the primary driven enrichment biological function. Besides, arachidonic acid metabolism, PPAR signaling pathway, insulin resistance, eicosanoids signaling, and other pathways and GO terms are also reported to find in cancer[35–37]. From the network of those biological function modules, which were connected with genes shared between modules, the lipid metabolic of LUAD was associated with nicotine, estrogen biosynthesis, melatonin, and atherosclerosis. Nicotine may promote LUAD development by regulating lipid metabolism. The interaction between estrogen biosynthesis and lipid metabolic is one of the high-risk factors of LUAD, which is consistent with the trend that lung cancer incidence is rising in women and has, in fact, more than doubled since the mid1970s[38]. Atherosclerosis and cancer have many similarities[39]. Patients with atherosclerotic disease are prone to repeated episodes of ischemia/reperfusion, which induces oxidative stress through the formation of oxygen free radicals. Endogenous exposure to free radicals increases the risk of cancer in individuals with atherosclerotic diseases[40]. Besides, retinoic acid inducing RAR-beta /RXR activation to promote tumor progression should be a potential way to promote LUAD. RXR can activate FXR/RXR and LXR/RXR, and the two activations also overlap. FXR has been reported as a tumor suppressor[41]. A possible mechanism is that FXR activates CCND1 expression and promotes cell proliferation. In order to activate the expression of its target genes, FXR is a heterodimer with retinoid X receptor alpha (RXRα). It binds to FXR response element (FXRE) after activation by a specific agonist, mainly IR-1[42]. Besides, FXR has been recently found related to the microenvironment of immunotherapy closely. Through FXR/RXR and LXR/RXR, lipid metabolic may influence the development of LUAD by regulating the immunity system.

To find the potential interventional target of LUAD patients, we constructed the network of those genes that are related to lipid and LUAD and find six hub genes. CYP2C9, which is a drug target of lung cancer, can be slowed by cytochrome P450, the tumorigenesis was regulated[43, 44]. LUAD patients with a lower expression of CYP2C9 have a better prognosis. UGT1A variants may play only a minor role in other lung cancer risk[45]. LUAD patients with a lower expression of UGT1A have a better prognosis (Fig. 3). DGAT1 and LPL are lipid metabolic genes. Both of them are involved in fatty acid synthesis. HPGDS has the therapeutic potential in allergic inflammation[46]. Those three genes were positively related to survival time. INS encodes insulin and plays a vital role in the regulation of carbohydrate and lipid metabolism.
LUAD patients with a lower expression of INS have a better prognosis. The regulation of fatty acid synthesis and insulin and the inflammation control may be the treatment of LUAD patients. Based on those six genes, a risk model was constructed. LUAD patients from two cohorts with the lower risk score had a better prognosis.

**Conclusions**

In summary, we generated a lipid metabolic transcriptome-wide profile of LUAD patients and found that significant lipid metabolic pathways were correlated with the LUAD. Our findings suggest that lipid metabolic is a way through which exogenous substances can affect the development of cancer. A signature of six lipid metabolic genes was significantly associated with diagnosis and prognosis of LUAD patients. The gene signature can be used as a biomarker for LUAD.

**Abbreviations**

LUAD: lung adenocarcinoma; HDL-C: high-density lipoprotein cholesterol; LDL: low-density lipoprotein; LDLR: low-density lipoprotein receptor; PCs: phosphatidylcholine; PEs: phosphatidylethanolamine; FAO: Fatty acid oxidation; NSCLC: non-small-cell lung carcinoma; TCGA: the Cancer Genome Atlas; KEGG: Kyoto Encyclopedia of Genes and Genomes; MisDB: Molecular Signatures Database; DEGs: differentially expressed genes; logFC: log2 fold change; PPI: protein-protein interaction; STRING: Search Tool for the Retrieval of Interacting Genes; OS: overall survival; HR: hazard ratios; CI: confidence intervals; INS: Insulin; LPL: Lipoprotein Lipase; HPGDS: Hematopoietic Prostaglandin D Synthase; DGAT1: Diacylglycerol O-Acyltransferase 1; UGT1A6: UDP Glucuronosyltransferase Family 1 Member A6; CYP2C9: Cytochrome P450 Family 2 Subfamily C Member 9; RXRα: retinoid X receptor alpha; FXRE: FXR response element.

**Declarations**

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Availability of data and materials**

The datasets generated and/or analysed during the current study are available in the TCGA & GEO databases, [https://cancergenome.nih.gov/](https://cancergenome.nih.gov/) & [https://www.ncbi.nlm.nih.gov/geo/].
Competing interests

The authors declare that they have no competing interests.

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Authors’ contributions

JYL and QL analyzed the data and helped draft the manuscript. FZ and HTM supervised this work and edited and revised manuscript. ZYS, QS, YZ, TNF and JYJ prepared figures and contributed to the drafting of the manuscript. The authors read and approved the final manuscript.

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References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68:394–424.
2. Barta JA, Powell CA, Wisnivesky JP. Global Epidemiology of Lung Cancer. Annals of global health. 2019;85:8.
3. Shi J-F, Wang L, Wu N, Li J-L, Hui Z-G, Liu S-M, Yang B-Y, Gao S-G, Ren J-S, Huang H-Y, et al. Clinical characteristics and medical service utilization of lung cancer in China, 2005–2014: Overall design and results from a multicenter retrospective epidemiologic survey. Lung Cancer. 2019;128:91–100.
4. Tseng C-H, Tsuang B-J, Chiang C-J, Ku K-C, Tseng J-S, Yang T-Y, Hsu K-H, Chen K-C, Yu S-L, Lee W-C, et al. The Relationship Between Air Pollution and Lung Cancer in Nonsmokers in Taiwan. Journal of Thoracic Oncology. 2019;14:784–92.
5. Groen HJM, Hiltemann TJN. Air Pollution and Adenocarcinoma in Never-Smokers. Journal of Thoracic Oncology. 2019;14:761–3.
6. Chen F, Jackson H, Bina WF. Lung adenocarcinoma incidence rates and their relation to motor vehicle density. Cancer Epidemiology Prevention Biomarkers. 2009;18:760–4.
7. Whittaker S, Marais R, Zhu A. The role of signaling pathways in the development and treatment of hepatocellular carcinoma. Oncogene. 2010;29:4989–5005.
8. Salvador MM, de Cedrón MG, Rubio JM, Martínez SF, Martínez RS, Casado E, de Molina AR, Sereno M. Lipid metabolism and lung cancer. Crit Rev Oncol/Hematol. 2017;112:31–40.

9. Zhou T, Zhan J, Fang W, Zhao Y, Yang Y, Hou X, Zhang Z, He X, Zhang Y, Huang Y. Serum low-density lipoprotein and low-density lipoprotein expression level at diagnosis are favorable prognostic factors in patients with small-cell lung cancer (SCLC). BMC Cancer. 2017;17:269.

10. Chi P-D, Liu W, Chen H, Zhang J-P, Lin Y, Zheng X, Liu W, Dai S. High-density lipoprotein cholesterol is a favorable prognostic factor and negatively correlated with C-reactive protein level in non-small cell lung carcinoma. PloS one 2014, 9.

11. Chen Y, Ma Z, Shen X, Li L, Zhong J, Min LS, Xu L, Li H, Zhang J, Dai L: Serum lipidomics profiling to identify biomarkers for non-small cell lung cancer. BioMed research international 2018, 2018.

12. Marien E, Meister M, Muley T, Fieuws S, Bordel S, Derua R, Spraggins J, Van de Plas R, Dehairs J, Wouters J. Non-small cell lung cancer is characterized by dramatic changes in phospholipid profiles. Int J Cancer. 2015;137:1539–48.

13. Luo X, Cheng C, Tan Z, Li N, Tang M, Yang L, Cao Y. Emerging roles of lipid metabolism in cancer metastasis. Mol Cancer. 2017;16:76.

14. Wang Y, Zhang X, Tan W, Fu J, Zhang W. Significance of fatty acid synthase expression in non-small cell lung cancer. Zhonghua zhong liu za zhi [Chinese journal of oncology]. 2002;24:271–3.

15. Troost J, Lindenmaier H, Haefeli WE, Weiss J. Modulation of cellular cholesterol alters P-glycoprotein activity in multidrug-resistant cells. Mol Pharmacol. 2004;66:1332–9.

16. Migita T, Narita T, Nomura K, Miyagi E, Inazuka F, Matsuura M, Ushijima M, Mashima T, Seimiya H, Satoh Y. ATP citrate lyase: Activation and therapeutic implications in non–small cell lung cancer. Cancer research. 2008;68:8547–54.

17. Uehara H, Takahashi T, Oha M, Ogawa H, Izumi K. Exogenous fatty acid binding protein 4 promotes human prostate cancer cell progression. International journal of cancer. 2014;135:2558–68.

18. DeFilippis RA, Chang H, Dumont N, Rabban JT, Chen Y-Y, Fontenay GV, Berman HK, Gauthier ML, Zhao J, Hu D. CD36 repression activates a multicellular stromal program shared by high mammographic density and tumor tissues. Cancer discovery. 2012;2:826–39.

19. Huang J, Fan X-X, He J, Pan H, Li R-Z, Huang L, Jiang Z, Yao X-J, Liu L, Leung EL-H. SCD1 is associated with tumor promotion, late stage and poor survival in lung adenocarcinoma. Oncotarget. 2016;7:39970.

20. Wadlow R, Ramaswamy S. DNA microarrays in clinical cancer research. Curr Mol Med. 2005;5:111–20.

21. Colaprico A, Silva TC, Olsen C, Garofano L, Cava C, Garolini D, Sabedot TS, Malta TM, Pagnotta SM, Castiglioni I, et al. TCGAbiolinks: an R/Bioconductor package for integrative analysis of TCGA data. Nucleic Acids Res. 2016;44:e71.

22. Kanehisa M, Furumichi M, Tanabe M, Sato Y, Morishima K. KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic Acids Res. 2017;45:D353-d361.
23. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, Paulovich A, Pomeroy SL, Golub TR, Lander ES, Mesirov JP. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. Proc Natl Acad Sci U S A. 2005;102:15545–50.

24. Robinson MD, McCarthy DJ, Smyth GK. edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. Bioinformatics. 2010;26:139–40.

25. Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. Omics. 2012;16:284–7.

26. Szklarczyk D, Franceschini A, Wyder S, Forslund K, Heller D, Huerta-Cepas J, Simonovic M, Roth A, Santos A, Tsafou KP, et al. STRING v10: protein-protein interaction networks, integrated over the tree of life. Nucleic Acids Res. 2015;43:D447–52.

27. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 2003;13:2498–504.

28. Chin CH, Chen SH, Wu HH, Ho CW, Ko MT, Lin CY. cytoHubba: identifying hub objects and sub-networks from complex interactome. BMC Syst Biol. 2014;8(Suppl 4):11.

29. Győrffy B, Surowiak P, Budczies J, Lánczky A. Online survival analysis software to assess the prognostic value of biomarkers using transcriptomic data in non-small-cell lung cancer. PLoS One. 2013;8:e82241.

30. Zhang Z, Kattan MW. Drawing Nomograms with R: applications to categorical outcome and survival data. Annals of translational medicine. 2017;5:211–1.

31. Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. Cell Metabol. 2016;23:27–47.

32. Santos CR, Schulze A. Lipid metabolism in cancer. FEBS J. 2012;279:2610–23.

33. Zhang S-Y, Shao D, Liu H, Feng J, Feng B, Song X, Zhao Q, Chu M, Jiang C, Huang W, Wang X. Metabolomics analysis reveals that benzo[a]pyrene, a component of PM2.5, promotes pulmonary injury by modifying lipid metabolism in a phospholipase A2-dependent manner in vivo and in vitro. Redox Biology. 2017;13:459–69.

34. Huang C, Freter C. Lipid metabolism, apoptosis and cancer therapy. Int J Mol Sci. 2015;16:924–49.

35. Marks F, Fürstenberger G, Müller-Decker K: Metabolic targets of cancer chemoprevention: interruption of tumor development by inhibitors of arachidonic acid metabolism. In Chemoprevention of Cancer. Springer; 1999: 45–67.

36. Go R-E, Hwang K-A, Choi K-C. Cytochrome P450 1 family and cancers. J Steroid Biochem Mol Biol. 2015;147:24–30.

37. Ashton KA, Proietto A, Otton G, Symonds I, McEvoy M, Attia J, Gilbert M, Hamann U, Scott RJ. Polymorphisms in genes of the steroid hormone biosynthesis and metabolism pathways and endometrial cancer risk. Cancer Epidemiol. 2010;34:328–37.
38. Barta JA, Powell CA, Wisnivesky JP. Global epidemiology of lung cancer. Annals of global health 2019, 85.

39. Tapia-Vieyra JV, Delgado-Coello B, Mas-Oliva J. Atherosclerosis and Cancer; A Resemblance with Far-reaching Implications. Arch Med Res. 2017;48:12–26.

40. Dreyer L, Prescott E, Gyntelberg F. Association between atherosclerosis and female lung cancer—a Danish cohort study. Lung Cancer. 2003;42:247–54.

41. Fu T, Coulter S, Yoshihara E, Oh TG, Fang S, Cayabyab F, Zhu Q, Zhang T, Leblanc M, Liu S. FXR regulates intestinal cancer stem cell proliferation. Cell. 2019;176:1098–112. e1018.

42. You W, Chen B, Liu X, Xue S, Qin H, Jiang H. Farnesoid X receptor, a novel proto-oncogene in non-small cell lung cancer, promotes tumor growth via directly transactivating CCND1. Sci Rep. 2017;7:591.

43. Chen J, Bearz A, Kim D, Mamdani H, Bauman J, Chiari R, Ou S, Solomon B, Soo R, Felip E. P1. 01–84 Interaction of Lorlatinib with CYP2B6, CYP2C9, UGT, and P-gp Probe Drugs in Patients with Advanced Non-Small Cell Lung Cancer. Journal of Thoracic Oncology. 2019;14:392–3.

44. Sausville LN, Gangadhariah MH, Chiusa M, Mei S, Wei S, Zent R, Luther JM, Shuey MM, Capdevila JH, Falck JR, et al. The Cytochrome P450 Slow Metabolizers CYP2C9*2 and CYP2C9*3 Directly Regulate Tumorigenesis via Reduced Epoxyeicosatrienoic Acid Production. Can Res. 2018;78:4865–77.

45. Schieffer KM, Angstadt AY, Zhu J, Lazarus P, Gallagher CJ. Associations between polymorphisms and haplotypes in the UDP-glucuronosyl transferase 1A gene family with lung cancer risk. Next Generat Sequenc Applic. 2016;3:124.

46. Rittchen S, Heinemann A. Therapeutic Potential of Hematopoietic Prostaglandin D2 Synthase in Allergic Inflammation. Cells. 2019;8:619.

Figures
Figure 1

Identification and functional analysis of lipid metabolism-related DEGs. (A) Volcano plot of lipid metabolism-related genes, (B) Heatmap analysis of lipid metabolism-related DEGs, (C) GO and KEGG pathway enrichment analysis by clusterProfiler, (D) functional and signaling pathway enrichment by IPA. In (A) and (B), red, white, and blue represent higher expression levels, no expression differences, and lower expression levels, respectively.
Figure 2

The PPI network of lipid metabolism-related DEGs. Red, white, and blue nodes represent upregulated genes, no expression differences genes, and downregulated genes, respectively. The magnitude of the degree is positively correlated with the size of a node.
Figure 3

Survival analysis of hub genes. LUAD patients were subdivided into high/low gene expression groups based on the median expression level of each gene in LUAD tissues. (A) OS analysis of CYP2C9, (B) OS analysis of UGT1A6, (C) OS analysis of INS, (D) OS analysis of DGAT1, (E) OS analysis of HPGDS, and (F) OS analysis of LPL.
Figure 4

Prediction model based on survival-related hub genes and validation. (A) The nomogram of 6 survival-related genes, (B) survival analysis between high-risk score group and low-risk score group, and (C) validation of the model.

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