Cross-packaging of genetically distinct mouse and primate retroviral RNAs

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Abstract

Background: The mouse mammary tumor virus (MMTV) is unique from other retroviruses in having multiple viral promoters, which can be regulated by hormones in a tissue specific manner. This unique property has lead to increased interest in studying MMTV replication with the hope of developing MMTV based vectors for human gene therapy. However, it has recently been reported that related as well as unrelated retroviruses can cross-package each other’s genome raising safety concerns towards the use of candidate retroviral vectors for human gene therapy. Therefore, using a trans complementation assay, we looked at the ability of MMTV RNA to be cross-packaged and propagated by an unrelated primate Mason-Pfizer monkey virus (MPMV) that has intracellular assembly process similar to that of MMTV.

Results: Our results revealed that MMTV and MPMV RNAs could be cross-packaged by the heterologous virus particles reciprocally suggesting that pseudotyping between two genetically distinct retroviruses can take place at the RNA level. However, the cross-packaged RNAs could not be propagated further indicating a block at post-packaging events in the retroviral life cycle. To further confirm that the specificity of cross-packaging was conferred by the packaging sequences (ψ), we cloned the packaging sequences of these viruses on expression plasmids that generated non-viral RNAs. Test of these non-viral RNAs confirmed that the reciprocal cross-packaging was primarily due to the recognition of ψ by the heterologous virus proteins.

Conclusion: The results presented in this study strongly argue that MPMV and MMTV are promiscuous in their ability to cross-package each other’s genome suggesting potential RNA-protein interactions among divergent retroviral RNAs proposing that these interactions are more complicated than originally thought. Furthermore, these observations raise the possibility that MMTV and MPMV genomes could also co-package providing substrates for exchanging genetic information.
Background
The mouse mammary tumor virus (MMTV) is a betaretrovirus that has been primarily implicated in causing breast cancer and, to some extent, T-cell lymphomas in mice, reviewed in Mustafa et al., [1,2]. Classically, MMTV has been categorized as a simple retrovirus containing only the structural and regulatory genes needed to complete the virus life cycle. However, recently, it has been proposed that MMTV be reclassified as a more complex murine retrovirus because of the presence of accessory/regulatory factors such as sag [3,4], Naf [5], and the recently identified Rem/RmRE regulatory pathway of MMTV [6-8]. In addition, it has recently been shown that Rev and Rex proteins of human complex retroviruses can interact with MMTV Rem responsive element [9]. Furthermore, MMTV is unique from other simple and complex retroviruses in harboring several promoters for the expression of its various gene products [10-12]. Of these several promoters, two have been identified in the long terminal repeats (LTR) and two in the envelope region, reviewed in Mustafa et al., [1]. The LTR promoters are under the influence of steroid hormones in a tissue-specific manner due to the presence of hormone responsive elements (HREs), making them inducible [13]. Therefore, the tissue-specific and inducible MMTV promoters have lead to increased interest in studying MMTV replication with the ultimate goal of developing MMTV based vectors for their potential use in targeted gene therapy. In one recent study, MMTV promoters have been utilized in murine leukemia virus (MLV) based vectors used for targeted enzyme prodrug therapy both in vivo and in vitro [14]. The use of MMTV based vectors would not only provide tissue-specific and inducible expression of the therapeutic gene, but also its non-primate nature may circumvent potential safety concerns. Such concerns include cross-packaging and co-packaging of the transfer vector RNA genome by related primate retroviruses or retrovirus-like elements resulting in the generation of recombinant variants with unknown pathogenic potential.

Packaging of retroviral genomic RNA by the assembling virus particle is a crucial step in the virus life cycle. RNA packaging among retroviruses is unique and a highly specific phenomenon during which two copies of full length "unspliced" genomic RNA are preferentially packaged from amongst a wide pool of cellular and other spliced viral RNAs, reviewed by D'Souza and Summers [15] and Lever [16]. The specificity towards RNA packaging by the newly assembling viral particles is conferred by the recognition of specific cis-acting sequences, the packaging signal (ψ), present at the 5' end of the viral genome, and the nucleocapsid (NC) protein is responsible for discriminating between spliced and unspliced viral RNA, reviewed by D'Souza and Summers [15] and Lever [16]. Despite this specificity, in some cases, it has been shown that evolutionary related, however molecularly different, retroviruses can cross- and co-package each other's genome suggesting that phylogenetically related retroviruses are capable of using similar protein-and RNA-packaging elements, reviewed by D'Souza and Summers [15] and Lever [16]. Such cross- and co-packaging among retroviruses have been shown to exchange genetic information resulting in recombinant variants [17,18] undermining many advantages of using retroviral vectors in human gene therapy studies. The use of phylogenetically distant non-human retroviral vectors, such as those based on MMTV, should minimize the chances of recombination with unrelated primate exo-and/or endogenous retroviruses.

In spite of the advantages of using non-human retroviral vectors with inducible tissue-specific promoters as in the case of MMTV, so far, no detailed studies have been conducted to investigate the ability of MMTV RNA to be cross-packaged in human cells by heterologous primate retroviral proteins. However, in one earlier report, Günzburg and Salmons have reported that MMTV RNA could not be packaged by Moloney murine leukemia virus (MoMLV) packaging cell lines [19]. Such limited information regarding MMTV RNA packaging and cross-packaging studies in the literature can be attributed to 1) a trans complementation assay for MMTV was not developed until recently, and 2) MMTV is not expressed very efficiently in human cells due to the promoter's low transcriptional activity. In order to overcome these drawbacks, we have successfully replaced the U3 region of MMTV 5'LTR with the human cytomegalovirus (hCMV) promoter in MMTV based vectors to allow for its efficient expression in human cells; and we have also developed a three-plasmid trans complementation assay for MMTV to study its RNA packaging and propagation [20]. Using this in vivo packaging and transduction assay, we investigated the ability of MMTV RNA to be cross-packaged by a primate retrovirus, the Mason-Pfizer monkey virus (MPMV) that, like MMTV, also preassembles in the cytoplasm before budding. Our results showed that both of these viruses could cross-package each other's RNAs. However, the cross-packaged RNA could not be propagated further and therefore failed to transduce the target cells suggesting a block at post RNA packaging events of the retroviral life cycle such as reverse transcription and/or integration. Our results further demonstrated that this cross-packaging specificity was conferred specifically by the packaging sequences, which were in turn recognized by the heterologous proteins; since cloning of these sequences in plasmids, which generate non-viral RNAs, resulted in the encapsidation of these non-viral RNAs by MMTV and MPMV proteins reciprocally. The results presented in this study strongly suggest that MPMV and MMTV are promiscuous in their ability to cross-package each other's genome and that interactions between retroviral RNAs.
and Gag polyprogenitors are more complicated than originally thought.

**Results and discussion**

**In vivo packaging and transduction assay for MMTV and MPMV**

To study cross-packaging between MMTV and MPMV, we used three-plasmid trans complementation assays developed earlier by our laboratory [20,21]. These MMTV and MPMV trans complementation assays consist of a packaging construct, pA10 or pTR301, which expresses either MMTV or MPMV gag/pol genes, respectively, which results in the production of viral particles, which are capable of encapsidating viral RNA containing $\psi$. The source of the packageable RNA is provided by MMTV (pDA024, pSS013) and MPMV (pKAL11, pSS015) transfer vectors (Figure 1A and 1B). These transfer vectors contain the sequences responsible for RNA packaging, in addition to the cis-acting sequences needed for viral replication, which include Primer Binding Site (PBS), Poly Purine Tract (PPT) (needed for reverse transcription), and U3 and U5 attachment (att) sites (required for integration). In addition, these transfer vectors express hygromycin resistance and/or enhanced green fluorescence protein (EGFP) gene from an internal simian virus (SV40) promoter (SV-hyg/SV-EGFP), which allows for the monitoring of the successful propagation of the transfer vector RNAs by transducing the target cells with these marker genes. An envelope expression plasmid (MD.G) based on vesicular stomatitis virus envelope G (VSV-G) was also used to enable the study of steps involved in both packaging and propagation of the transfer vector RNAs [22].

Briefly, in these assays, the three plasmids are co-transfected into 293T producer cells, which will generate virus particles containing the encapsidated RNA, the replication of which is limited to a single round because reinfection of the target cells cannot take place. These virus particles can be used to: 1) directly examine the viral RNA content in the virus particles using reverse transcriptase polymerase chain reaction (RT-PCR) and 2) to infect target cells resulting in the transduction of these cells with the marker gene, thus allowing for monitoring the propagation of the transfer vector RNA. The number of Hygromycin-resistant (Hygr) colonies and/or EGFP positive cells obtained should be directly proportional to the amount of RNA that is packaged into the virus particles providing an indirect estimate of RNA packaging.

**MMTV RNA can be cross-packaged but cannot be propagated by MPMV proteins**

To determine whether MMTV RNA can be cross-packaged by the heterologous primate retrovirus (MPMV) proteins, we co-transfected MMTV transfer vectors (pDA024 and

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**Figure 1**

**Schematic representation of MMTV and MPMV based vectors.** (A) MMTV genome, MMTV transfer vectors, and expression plasmids containing MMTV or MPMV packaging signal. (B) MPMV genome, MPMV transfer vectors, and expression plasmids containing MPMV or MMTV packaging signal. The design and construction of these vectors is described in materials and methods and can be further obtained from authors upon request. CMV, human cytomegalovirus promoter; SV, Simian virus 40 promoter; hygr, hygromycin resistance gene; CTE, constitutive transport element from Mason-Pfizer monkey virus (MPMV); EGFP, enhanced green fluorescence protein gene; BGH, bovine growth hormone.
pSS013) with MPMV packaging construct (pTR301) along with the envelope expression plasmid (MD.G) into 293T cells. As a control, MMTV transfer vectors were also co-transfected with the homologous packaging construct (pA10) along with the envelope expression plasmid. Supernatants from the transfected cultures were used to isolate the viral RNA to determine vector RNA packaging and to infect human HeLa CD4+ cells in order to study vector RNA propagation.

To ensure that the transfer vector RNAs are efficiently and stably expressed and properly transported from the nucleus to the cytoplasm, RNAs from the transfected cells were fractionated into cytoplasmic and nuclear fractions. To verify that there was no contaminating plasmid DNA in our cytoplasmic RNA preparations, which may confound the interpretation of our results, cytoplasmic RNAs were treated with RNase free DNase and were PCR amplified. The lack of a positive PCR signal, following 30 cycles of amplification, indicated that the contamination in our cytoplasmic RNA preparations is below the detection level of amplification, indicated that the contamination in our cytoplasmic RNA preparations is below the detectable range. Since our vectors contained a hygromycin resistance marker or an EGFP gene, we examined the propagation of these vectors by infecting the target cells with supernatants produced by the transfected cells. Following infection, if the vectors were properly propagated, these marker genes (hygromycin resistance gene in the case of pDA024 and EGFP gene in the case of pSS013) will transduce the target cells resulting in Hygr colonies or EGFP positive cells suggesting the successful completion of crucial steps of the viral life cycle such as reverse transcription and integration following RNA packaging. As expected, the propagation of the packaged MMTV transfer vector RNAs by homologous MMTV proteins was readily observed as evidenced by the presence of Hygr colonies or EGFP positive cells when MMTV vector (pDA024 and pSS013) RNAs were cross-packaged by MPMV proteins (pTR301) suggested that the cross-packaged RNAs could not be propagated (Table 1).

**MPMV RNA can be cross-packaged but cannot be propagated by MMTV proteins**

To determine whether MMTV proteins can cross-package MPMV RNA or not, MPMV transfer vectors (pKAL011 and pSS015) were co-transfected along with MMTV packaging construct (pA10) and the envelope expression plasmid MD.G. In parallel, as a control, MPMV transfer vectors were also co-transfected with their homologous packaging construct (pTR301) and the envelope expression plasmid. After confirming the absence of any contaminating plasmid DNA in our cytoplasmic and viral RNA preparations (Figure 3A and 3D), we confirmed that all MPMV transfer vector RNAs were efficiently transported to the cytoplasm and were stably expressed (Figure 3B and 3C). Next, we investigated the ability of MPMV RNA to be cross-packaged by MMTV proteins by directly examining the viral RNA content in MMTV particles. RT-PCR results in figure...
3E demonstrated that MPMV transfer vector (pKAL011 and pSS015) RNAs were cross-packaged by MMTV proteins (pJA10). Consistent with the results obtained for MMTV RNA cross-packing, the efficiency of the cross-packaged MPMV vector RNAs was lower when compared to the homologous MPMV vector (pKAL011 and pSS015) RNAs being packaged by its own proteins (pTR301) (Figure 3E). The cross-packaging efficiency of MPMV RNA by MMTV proteins appeared to be lower when compared to MMTV RNAs being cross-packaged by MPMV proteins since Southern blotting was needed to appreciate cross-packaging after 20 cycles of PCR instead of the 15 cycles needed to demonstrate the cross-packaging of MMTV RNA by MPMV proteins (Figures 2E and 3E).

Similar to the results we obtained for MMTV cross-packaged RNA, MPMV cross-packaged transfer vector (pKAL011 and pSS015) RNAs could not be propagated further as evidenced by the lack of Hyg' colonies or EGFP positive cells in the infected cultures (Table 1). The presence of Hyg' colonies or EGFP positive cells in the infected cultures when MPMV transfer vector (pKAL011 and pSS015) RNAs were packaged by its own proteins (pTR301) assured that the marker genes were efficiently
Table 1: Propagation of MMTV and MPMV transfer vectors RNA by homologous and heterologous proteins.

| Transfer Vector | Description of the Transfer Vector                                                                 | Titers (CFU/ml)a, b | % EGFP Positive Cellsc |
|-----------------|----------------------------------------------------------------------------------------------------------------|---------------------|------------------------|
| pDA024          | Chimeric LTR, 5’ region upto 400 bp of MMTV Gag and SV-hyg’                                                | 3,676 ± 196         | < 1                    |
| pSS013          | Chimeric LTR, 5’ region upto 400 bp of MMTV Gag and SV-EGFP                                               | -                   | 19                     |
| pND015          | Same as DA024 but the putative MMTV ψ has been replaced with that of MPMV (5’ UTR + 282 bp of Gag)           | 2                   | -                      |
| pND016          | Same as DA024 but the putative MMTV ψ and PBS have been replaced with that of MPMV                           | < 1                 | 7                      |
| pND017          | Control MMTV vector in which putative MMTV ψ has been cloned back after creating NotI site at PBS/UTR junction | 187 ± 33            | ND                     |
| pND018          | Control MMTV vector in which putative MMTV ψ and PBS have been cloned back after creating NotI site at U5/PBS junction | 30 ± 4             | ND                     |
| pKAL011         | MPMV 5’ region upto 282 bp Gag and SV-hyg’                                                                  | < 1                 | 36,373 ± 3,972         |
| pSS015          | Same as KAL011 but has SV-EGFP instead of SV-hyg’                                                            | -                   | < 1                    |
| pND011          | Same as KAL011 but MPMV ψ has been replaced with that of MMTV (5’ UTR + 400 bp of Gag)                      | < 1                 | 3                      |
| pND012          | Same as KAL011 but MPMV ψ and PBS have been replaced with that of MMTV                                        | < 1                 | 23                     |
| pND013          | Control MPMV vector in which MPMV ψ has been cloned back after creating NotI site at PBS/UTR junction         | ND                  | 2813 ± 99              |
| pND014          | Control MPMV vector in which MPMV ψ and PBS have been cloned back after creating NotI site at U5/PBS junction | ND                  | 1660 ± 111             |
expressed and that there was no compromise on the integrity of our transduction assay. Therefore, it is reasonable to conclude that the absence of Hyg or EGFP positive cells reflected a true lack of propagation of MPMV cross-packaged vector RNAs. A similar situation has been reported earlier when MPMV RNA could be cross-packaged by heterologous feline immunodeficiency virus (FIV) proteins but these proteins could not further propagate the cross-packaged MPMV RNA [21]. Although not quantitative, the detection levels of the RT-PCR were consistent with the titters obtained in the transduction assay. Using homologous proteins, MPMV transfer vectors (pKAL011 and pSS015) were propagated at much higher efficiency (36,373 CFU/ml and 41% EGFP positive cells) when compared to MMTV transfer vectors (pDA024 and pSS013) (3,676 CFU/ml and 19% EGFP positive cells). Corroborating this observation, MPMV transfer vector RNAs were packaged more efficiently when compared to the same cycle of PCR amplification for MMTV transfer vector RNAs being packaged by its own proteins (Table 1, Figure 2E and 3E).

| Table 1: Propagation of MMTV and MPMV transfer vectors RNA by homologous and heterologous proteins. (Continued) |
|--------------------------------------------------|
| pTR174 | Control vector containing SV-hyg cassette and SIV 3'LTR as poly(A) | < 1 | < 1 | - | - |
| pAG001 | Control vector containing SV-hyg cassette and FIV 3'LTR as poly(A) | < 1 | < 1 | - | - |
| Mock | No DNA (control) | < 1 | < 1 | < 1 | < 1 |

* CFU/ml, colony forming units per milliliter of non-concentrated supernatant from the transfected cultures. * Each value represents a mean of three experiments performed in duplicates. * Percentage of EGFP positive cells represent the phenotypic analysis of the transduced target cells. Single cell suspensions of the infected HeLa CD4+ cells were prepared, and 10,000 events (cells) per group were counted and analyzed using Becton Dickinson FACS. < 1 indicates < 1 Hyg colonies in the target cells or < 1 EGFP positive cells out of 10,000 events (cells) analyzed.

Over expressed non-specific RNAs cannot be packaged by MMTV or MPMV proteins

As an additional control to rule out the possibility of non-specific packaging and possible propagation of random RNAs, we transfected control vectors pTR174 and pAG001 (lacking all the viral sequences at the 5’end) (Figure 4A) separately with MPMV and MMTV packaging constructs (pTR301 and pIA10) along with the envelope expression plasmid MD.G. After taking into consideration all the necessary controls for plasmid DNA contamination, RNA fractionation, and stability of the control transfer vector RNAs (data not shown), we investigated the possibility of packaging and propagation of these non-specific RNAs by both MMTV and MPMV proteins using RT-PCR. Amplification was conducted using control vector specific primers for 30 cycles to ensure the amplification of any control vector RNAs that may have been non-specifically packaged. As shown in figure 4B, neither MPMV nor MMTV proteins could package the control transfer vector RNA (or, if packaged, the level of packaging was below the threshold level of detection), and consequently the vector RNA could not be propagated (Table 1). The lack of packaging of these non-specific RNAs further confirmed our results that MMTV and MPMV proteins show relative specificity in recognizing the heterologous ψ and can efficiently encapsidate the heterologous RNA.

Comparison between MMTV and MPMV cis-acting nucleotide sequences and amino acid sequences important for reverse transcription and integration

The reciprocal cross-packaging of MPMV and MMTV RNAs suggests that the ψ of these viruses are readily recognized by each other’s NC domain of Gag protein facilitating their efficient cross-encapsidation. However, the absence of Hyg colonies or EGFP positive cells in the infected cultures suggests a block in post-packaging steps of the viral life cycle such as reverse transcription and/or integration. The successful propagation of MMTV and MPMV transfer vector RNAs would require the recognition of a number of cis-acting sequences present on their RNAs by the heterologous proteins following cross-packaging. Of these, reverse transcription and integration would require specific enzymes namely reverse transcriptase (RT) and integrase to work on targets such as PBS/PPT and att sites, respectively. Therefore, it is conceivable that MPMV and MMTV enzymes were not able to function on the respective targets of the cross-packaged RNAs resulting in the lack of propagation of these cross-packaged RNAs. With this rationale in mind, we compared the cis-acting sequences present on MMTV and MPMV transfer vector RNAs. Our sequence analysis shown in figure 5A revealed that PPT sequences between the two viruses have 90% sequence homology, while PBS showed a sequence homology of approximately 72%.

Comparison of att sites between the two viruses indicated less than 50% sequence homology (U3 att 40% whereas the U5 att 45%).

Regulatory enzymes working on the above-mentioned cis-acting sequences may also have played an important role in the inability of the cross-packaged RNAs to propagate because of the sequence heterogeneity between these two
viruses. Therefore, RNaseH and integrase were good candidates to be examined carefully since RNaseH plays a pivotal role in hydrolyzing the RNA-DNA hybrid [25] and its inactivation leads to production of non-infectious virus [26]. Similarly, integrase is essential for the integration of the linear retroviral DNA, a crucial step for the completion of the virus life cycle and has also been shown to promote reverse transcription through interactions with the nucleoprotein reverse transcription complex [27]. Thus, using the sequence alignment program, CLUSTAL W [28], we compared the amino acid sequences of MPMV and MMTV RNaseH and integrase. Because of the lack of any published sequences of MMTV integrase and RNaseH, we took the amino acid sequences of MPMV integrase [29] and RNaseH [30] and aligned them with the entire Gag-Pro-Pol polyprotein amino acid sequence of exogenous MMTV(C3H) (accession number AF228552) [31]. As expected, the MPMV RNaseH and integrase sequences aligned with those of the 3'end of Gag-Pro-Pol polyprotein of MMTV revealing varying degrees of heterogeneity.

The sequence homology between MPMV RNaseH and integrase with the corresponding sequences of MMTV was found to be 32% and 49%, respectively (Figure 5B).

Based on the sequence analyses of MPMV and MMTV cis-acting sequences and those of the regulatory enzymes acting on them (and the fact that both of these viruses utilize Lys tRNA primers; Lys-1, 2 for MPMV and Lys-3 for MMTV), it is plausible to propose that the cross-packaged RNAs have the potential to successfully initiate the process of reverse transcription; however, they may not have been able to efficiently complete this event. In addition to this, the possibility of a fully reverse transcribed unintegrated provirus cannot be ruled out as has been reported earlier [32]. The inability of the cross-packaged RNAs to reverse transcribe and/or to integrate should result in a failed transduction of the target cells because our read out assay is based on the expression of the marker genes from an integrated provirus. This postulation stems from the fact that inadequate integration has been implicated in
The results presented so far clearly indicate that MMTV and MPMV proteins were able to recognize the \( \psi \) sequence on each other’s RNAs. If the recognition of the packaging sequences by the heterologous proteins was sufficient to encapsidate the RNAs, we would argue that any RNA containing these sequences should also be able to be cross-packaged into the homologous as well as the heterologous viral particles. Therefore, the region encompassing sequences containing MPMV [35] and the putative MMTV \( \psi \) (unpublished observations) were cloned into expression plasmids that can generate non-viral RNAs, which could act as a substrate for packaging into the assembling virus particles. In a two-plasmid trans-complementation assay, the expression plasmids containing the putative MMTV \( \psi \) (5’UTR + 400 bp of Gag in the case of pNF007 and R/U5 + 5’UTR + 400 bp of Gag in the case of pNF008; Figure 1A) were transfected along with MMTV (pJA10) as well as MPMV (pTR301) packaging constructs into the 293T producer cells. Similarly, the expression plasmids containing MPMV \( \psi \) (R/U5 + 5’UTR + 282 bp of Gag in pND001 and 5’UTR + 282 bp of Gag in pND002; Figure 1B) were also transfected with the homologous MPMV (pTR301) or the heterologous MMTV (pJA10) packaging constructs.

Following trans-complementation, both cytoplasmic and viral RNAs were isolated and after consideration of the necessary controls (data not shown), the RNAs were reverse transcribed and the RT-PCR was conducted to determine the encapsidation of non-viral RNAs that contain either the MPMV or the putative MMTV \( \psi \) by the homologous or the heterologous proteins. RT-PCR analysis conducted on the RNAs demonstrated that non-viral (pNF007 and pNF008) RNAs were packaged by the homologous (MMTV; pJA10) and the heterologous (MPMV; pTR301) proteins (Figure 6A). These results further confirmed our findings of the ability of MPMV proteins to cross-package MMTV transfer vector RNAs in both viral (pDA024 and pSS013) and non-viral (pNF007 and pNF008) context (Figures 2E and 6A). In a reciprocal fashion, MMTV proteins (pJA10) cross-packaged non-viral (pND001 and pND002) RNAs containing MPMV \( \psi \) (Figure 6B) confirming our initial observation of the cross-packaging of MPMV transfer vector (pKAL011 and pSS015) RNAs by MMTV proteins in the viral context (Figure 3E). The cross-packaging of non-viral RNA containing \( \psi \) further confirmed that the specificity towards RNA packaging was conferred by \( \psi \). Packaging of non-viral RNAs containing the packaging sequences has also been observed for other retroviruses such as bovine leukemia virus (BLV) [36], FIV [24], and MoMLV [37].

**Substitution of the packaging signal in MMTV and MPMV transfer vectors resulted in efficient packaging but drastically reduced vector RNA propagation**

The results of the cross-packing experiments between MMTV and MPMV suggested that the reciprocal cross-packaging was due to the cross-recognition of packaging sequences by the heterologous proteins. The lack of propagation of these cross-packaged transfer vector RNAs, on the other hand, suggested that following packaging some of the events imperative for the transduction of the target
Figure 5
Nucleotides and amino acid sequences comparison between MMTV and MPMV. (A) Comparison between MMTV and MPMV cis-acting sequences needed for successful reverse transcription and integration. PBS, primer binding site; PPT poly purine tract; U3 att, attachment site at 3'LTR; U5 att, attachment site at 5'LTR. The boxed areas represent the canonical TG and CA dinucleotides in the U3 and U5 att sequences. The "-" represents homologous sequences, the differences are represented by the actual nucleotides. (B) Amino acid sequence alignment of MPMV RNaseH and integrase with the corresponding region of MMTV using the sequence alignment program, CLUSTAL W. Identical amino acids are boxed.
cells were not carried out. Therefore, we rationalized that since MPMV proteins are able to recognize the putative MMTV ψ, we should be able to exchange successfully the putative MMTV ψ with that of MPMV in the presence of either MMTV or MPMV PBS (pND015 and pND016 respectively) or vice versa (pND011 and pND012) (Figure 1A and 1B). The chimeric transfer vectors (pND015 and pND016) when co-transfected with MMTV packaging constructs, in addition to being cross-packaged, should also be able to propagate its RNA. Similar strategy has been used to propagate chimeric lentiviral transfer vector RNAs; for example, exchanging the packaging signal in FIV transfer vectors with those of human and simian immunodeficiency virus (HIV and SIV, respectively) resulted not only in packaging but also in the propagation of the chimeric FIV transfer vector RNA by FIV, HIV, and SIV proteins, albeit at a much reduced efficiency [21].

A test of MMTV (pND015 & pND016) and MPMV (pND011 & pND012) chimeric transfer vectors in our in vivo packaging and transduction assay using either MMTV (pJA10) or MPMV (pTR301) packaging constructs revealed that the chimeric transfer vector RNAs were successfully packaged and cross-packaged by the respective homologous and heterologous proteins (Figure 6C). However, contrary to our expectations, the packaged RNAs by the homologous or by the heterologous proteins either could not be propagated at all or there was a drastic drop in propagation as evidenced by very few Hyg+ colonies (< 25 CFU/ml compared to 3,676 CFU/ml when pDA024 was packaged by pJA10 or 36,373 CFU/ml when pKAL011 was packaged by pTR301; Table 1). Taken together, these rather unexpected results pointed in the direction that we may have disrupted the region around the U5/PBS/UTR during the course of exchanging sequences encompassing ψ, and that this disruption proved to be detrimental for vector RNA propagation and not RNA packaging. During the course of cloning these chimeric transfer vectors, an artificial NotI site either at PBS/UTR (pND015 and pND011) or at U5/PBS junctions (pND016 and pND012) was created (Figure 1A and 1B). Thus, it is conceivable that such non-natural sequences at these crucial junctions may have affected essential steps in the viral life cycle such as reverse transcription thereby hindering the propagation of these RNAs.

To confirm whether the introduction of such artificial sequences could limit the ability of the transfer vector RNAs to propagate, we created control vectors for both MMTV (pND017 and pND018) and MPMV (pND013 and pND014). In these control transfer vectors, regions encompassing MMTV and MPMV ψ were amplified and cloned back at the same location after artificially creating a NotI site as in the case of MMTV (pND015 and pND016) and MPMV (pND011 and pND012) chimeric transfer vectors (Figures 1A and 1B). Test of these control transfer vectors (pND013 and pND017) in which the NotI site was created at the PBS/UTR junction revealed that their RNAs were propagated to a much reduced level (2 fold reduction in the case of pND017 when compared to pDA024 and 13 folds reduction in the case of pND013 when compared to pKAL011; Table 1). On the other hand, when NotI site was created at the U5/PBS junction (pND014 and pND018), the propagation of these control transfer vectors was further hindered (120 fold reduction in pND018 when compared to pDA024 and 22 fold reduction in pND014 when compared to pKAL011; Table 1). We did not directly investigate the encapsidation of these control transfer vector RNAs, because when similar sequences were exchanged by creating artificial NotI site on MMTV and MPMV chimeric transfer vectors (pND015, pND016, pND011, and pND012), their RNAs were shown to be efficiently packaged by both homologous and heterologous proteins (Figure 6C).

The drastic drop in the propagation of the chimeric and the control transfer vectors can be explained on the basis that the introduction of NotI site in these vectors may have disrupted RNA structural elements encompassing sequences such as PBS (needed for reverse transcription) and/or sequences encompassing or overlapping ψ (needed for RNA dimerization) and these manipulations in turn may have diminished the chances of viral RNA propagation. Alterations/mutations in dimerization region have been shown to affect multiple steps in the viral life cycle, excluding packaging [38]. For example, in the case of HIV-1, a stem loop structure downstream of the PBS called dimerization initiation site (DIS) is able to dimerize via a "kissing loop" model [39], and further reviewed in [16]. In addition, the apical loop of the "kissing loop" hairpin contains a GC-rich autocomplementary sequence called palindrome (pal) [40,41]. Mutations in DIS region and in the palindrome have shown pronounced effects on viral infectivity, minor or no effects on RNA packaging, and variable effects on RNA dimerization [41,42], and further reviewed in [43]. Although the existence of a "kissing loop" model and its potential role in the life cycle of MPMV and MMTV has not been established yet, we have observed in close vicinity of the PBS a GC-rich pal sequence in both MPMV (5' UGGCGGCGCCGGC 3') and MMTV (5' UUCGGGCCGCAC 3') with 100% auto-complementarity (unpublished observations). In addition, using RNA structure prediction program, we observed putative interaction(s) between NotI site (5' GGGCCGCG 3') and PBS (MMTV PBS: 5' UGGCGGCGCCGGCG 3'; MPMV PBS: 5' UGGCGGCGCCGGCG 3') or pal in some of the control transfer vectors showing no or much reduced vector RNA propagation (data not shown). The complementarity between these sequences and their GC-rich nature might have disrupted the RNA structural elements in these regions, which compromised the role of these biologically
Figure 6
Heterologous and chimeric transfer vector RNAs cross-packaging. (A) Reciprocal cross-packaging of heterologous RNAs containing MMTV packaging signal. RT-PCR of cytoplasmic (upper panel) and viral (lower panel) cDNAs amplified using MMTV specific primers and probed with the PCR product amplified using the same set of primers and HYB MTV as a template. For this set of experiment, primers OTR567 and OTR560 were used and should amplify 149 bp fragment. (B) Reciprocal cross-packaging of heterologous RNAs containing MPMV packaging signals. RT-PCR of cytoplasmic (upper panel) and viral (lower panel) cDNAs amplified using MPMV specific primers and probed with the PCR product amplified using the same set of primers and pKAL011 as template. For this set of experiments, primers OTR112 and OTR197 were used and should amplify 154 bp fragment. (C) MMTV and MPMV chimeric transfer vectors RNA containing each other’s packaging sequences can be packaged in a reciprocal fashion. Upper panel represents RT-PCR of cytoplasmic cDNAs using vector RNA specific primers to ensure their stable expression. Lower panel represents RT-PCR of viral cDNAs amplified using chimeric vector specific primers. For this set of experiment, primers OTR567 and OTR560 were used for pND011 and pND012 and should amplify 149 bp. For pND015 and pND016, primers OTR730 and OTR197 were used and should amplify 268 bp. However, for the sake of clarity and to follow cross-packaging, the PCR products are shown next to each other's in this figure.
important sequences. Furthermore, it has earlier been reported that the interaction between the PBS and the tRNA 3' terminus is crucial for primer selection in MLV reverse transcription and that mutations in PBS of both MLV and HIV-1 involving aberrant reverse transcription have been reported to hinder viral replication [44]; consequently, the interaction between the NotI site and PBS may have hindered the accessibility of tRNA to PBS in our control and chimeric transfer vectors. Therefore it is reasonable to propose that the substitution and/or introduction of the heterologous sequences, in addition to the creation of the NotI site, may have disrupted the yet to be identified "kissing loop" model in the case of MPMV and MMTV, which thwarted post RNA packaging steps of retroviral life cycle, resulting in the abrogation and/or much reduced propagation of the packaged RNAs in the control as well as chimeric transfer vectors.

Owing to the wide use of retroviral vectors in human gene therapy and the safety issues related with cross- and co-packaging among retroviruses, these areas have been extensively investigated and have revealed that reciprocal as well as non-reciprocal cross-packaging among genetically distinct, simple and complex, retroviruses can take place [17,21,45-54]. Not only cross-packaging but also co-packaging, resulting in the exchange of genetic information, has been reported in genetically distant retroviruses such as SNV and MLV [17] and HIV-1 and HIV-2 [18]. One of the most important consequences of exchanging genetic information is the generation of viral variants with unknown pathogenic potential. This has brought the vector safety concerns to the forefront especially in the light of the mobilization of HIV-1 based vectors from the transduced cells following infection with the wild type virus [55,56]. In addition, the generation of a replication competent retrovirus through the process of recombination between the vector, the packaging construct, and endogenous retrovirus-like elements has also been reported [57]. In light of these safety concerns, we tested whether another non-primate retrovirus candidate for human gene therapy, MMTV, would be prone to cross-packaging with a primate retrovirus as has previously been shown that a phylogenetically distant non-primate lentivirus (FIV) could be packaged by primate lentiviruses [21]. Our study showed that MMTV and MPMV packaging sequences are promiscuous and could be recognized by each other’s proteins suggesting potential RNA-protein interactions among divergent retroviruses. It would be interesting to determine how frequently MMTV RNA can be cross-packaged by other retroviral proteins and whether there are obstacles to these RNA-protein interactions. These observations raise the possibility that MMTV and MPMV RNAs could also co-package and exchange genetic information. Therefore, it would be interesting to investigate whether co-packaging between MMTV and MPMV could occur and if so, at what frequency and what are the properties of the resulting recombinant variants, if at all.

Conclusion

The results presented in this study raise important questions about the design and use of candidate vectors for human gene therapy and suggest that sequences, which could contribute to such cross-packaging, should be eliminated from the transfer vectors to allow the establishment of safer and more effective vectors for human gene transfer studies. The retrovirus life cycle may be exploited to render the cis-acting sequences of an integrated transfer vector inactive through a process called self-inactivation (SIN) [45,55,58-61]. In addition to SIN vectors, a novel super-split packaging system has also been recently developed, which incorporates many new safety features. In such a system, gag was separated from pol and the protease was provided in trans by a less cytotoxic mutant, while maintaining high titers [62]. Surprisingly, despite taking into consideration several safety aspects in modified packaging systems and transfer vectors, vector mobilization as well as leaky transcriptional termination have been reported [63-65]. Therefore, in order to construct efficient and safe vectors, a thorough understanding of RNA packaging and cross packaging among retroviruses is imperative, and the understanding we will gain from such studies should allow employing multiple strategies to prevent viral replication through packaging and/or recombination after delivery of the gene of interest into the target cells.

Methods

Numbering system

Nucleotide designation for MMTV is based on HYB MTV molecular clone sequence [66]. Nucleotide designation for MPMV is based on GenBank accession number M12349 [67].

Plasmid Constructions

MMTV packaging constructs and transfer vectors

The MMTV packaging construct, pJA10, has been described previously [20]. Briefly, pJA10 expresses MMTV gag/pol gene from hCMV promoter and contains constitutive transport element (CTE) from MPMV downstream of MMTV gag/pol. pDA024 and pSS013 are MMTV based transfer vectors that have been described previously [20] and contain 5' chimeric LTR in which MMTV promoter sequences have been replaced with that of hCMV (Figure 1A).

To assess whether the presence of regions encompassing the putative MMTV ψ on non-viral RNAs would facilitate the packaging of these RNAs, sequences containing the putative MMTV ψ (5'UTR and 400 bp of Gag) and (R/U5/ 5'UTR and 400 bp of Gag) were amplified using sense primers OTR680 and OTR617 and an anti-sense primer
OTR552 using HYB MTV as a template. The PCR products were cleaved with the artificially created HindIII and SpeI sites and cloned into HindIII and XbaI sites of pNF003 (a derivative of pcDNA3) generating pNF007 and pNF008, respectively (Figure 1A).

pND015 and pND016 are pDA024 derivatives containing MPMV ψ either in the presence (pND015) or absence (pND016) of MMTV PBS and were generated through series of cloning steps. Briefly, the 3' end of MMTV genome containing SV-hyg cassette and MPMV CTE along with 3'LTR was obtained by digesting pDA024 with SpeI and KpnI. The 5' region of MMTV genome containing 5'LTR with and without PBS was amplified using sense primer OTR726 and anti-sense primer OTR727 and pDA024 as a template. The resulting PCR products were digested with the artificially created Sacl and SpeI sites. The resulting Sacl-Spel PCR fragment containing 5'end of the genome (without the packaging signal) and SpeI-KpnI fragment containing the 3'end of the genome were ligated in a three way ligation to a pIC based cloning vector [68], which had already been cleaved with Sacl and KpnI resulting in pND005 and pND006, respectively. Such a cloning strategy created an artificial NotI site at the junction of PBS/UTR in the case of pND005 and at the U5/PBS junction in the case of pND006. Next, sequences encompassing MPMV packaging determinants (UTR and 282 bp of Gag) were amplified using sense primers OTR723 and OTR727 and OTR731 as anti-sense primers. The resulting PCR products were cleaved by the flanking artificially created NotI sites and were cloned into the NotI site of pND005 and pND006 resulting in the final clones pND015 and pND016, respectively (Figure 1A).

To ensure that the creation of an artificial NotI site in the region encompassing MMTV packaging sequences will not affect RNA packaging or propagation adversely, we created pND017 and pND018 as control transfer vectors. Putative sequences that are involved in MMTV RNA packaging with and without the MMTV PBS were amplified using sense primers OTR724 and OTR725 and anti-sense primer OTR726 and pDA024 as a template. The combination of these primers created flanking artificial NotI sites in the PCR products facilitating their cloning into NotI site of pND005 and pND006 to generate pND017 and pND018, respectively (Figure 1A).

**MPMV packaging constructs and transfer vectors**

The MPMV packaging construct, pTR301, has already been described [21]. Briefly, it expresses MPMV gag/pol genes from hCMV intron A promoter/enhancer and contains MPMV CTE between Pol termination codon and bovine growth hormone (BGH) Poly (A) sequences. The MPMV transfer vector, pKAL011, has also been described previously [21] and contains all cis-acting sequences necessary for RNA packaging and propagation. pSS015 is similar to pKAL011 except it contains SV-EGFP cassette instead of SV-hyg cassette (Figure 1B).

To determine if the presence of regions encompassing MPMV packaging sequences on non-viral RNAs would facilitate the packaging of these RNAs, two different regions encompassing MPMV ψ were cloned into pNF003. A region encompassing MPMV ψ (R/U5, 5'UTR, and 282 bp of Gag) was amplified using a sense primer OTR615, an anti-sense primer OTR616, and pKAL011 as a template. The PCR product was cleaved with the artificially created HindIII and NheI sites and was cloned into HindIII and XbaI sites of pNF003 generating pND001. pND002 is another version of pcDNA3 based clone containing MPMV ψ and was created by isolating Sfo-Hpal fragment containing MPMV PBS, 5'UTR and 282 bp of Gag from pKAL011 and was cloned into the blunted HindIII and XbaI sites of NF003 (Figure 1B).

pND011 and pND012 are MPMV based vectors in which MPMV ψ has been replaced with that of MMTV either in the presence (pND011) or absence (pND012) of MPMV PBS and were created through series of cloning steps. As a first step, the 3'end of MPMV genome containing 3'LTR was obtained by digesting pKAL011 with NheI and BamHI. The MPMV 5'LTR along with or without PBS were amplified using sense primer OTR721 and anti-sense primers OTR722 and OTR723 using pKAL011 as a template. The resulting Xhol-Nhel PCR fragments containing 5'end of the genome (without the packaging signal) and Nhel and BamHI fragment containing the 3'end of the genome were ligated in a three way ligation at the Xhol and BamHI sites of a pIC based cloning vector resulting in pND003 and pND004, respectively. Similar to the cloning scheme of MMTV chimeric vectors, such a cloning strategy created an artificial NotI site at the PBS/UTR and US/PBS junctions in the case of pND003 and pND004, respectively. Next, the putative MMTV ψ (5'UTR and 400 bp of Gag) with and without MMTV PBS were amplified from pDA024 using sense primers OTR724 and OTR725 and anti-sense primer OTR726 as the anti-sense primer. The resulting PCR products were cleaved by the flanking artificially created NotI sites and were cloned into the NotI site of pND003 and pND004 resulting in pND007 and pND008. Finally, SV-hyg cassette containing flanking Nhel sites was cloned into the Nhel site of pND007 and pND008 generating pND011 and pND012 (Figure 1B).

pND013 and pND014 were created as controls to see the effects of an artificially created NotI site within MPMV ψ. Regions containing MPMV ψ (with and without the PBS) were amplified using sense primers OTR729 and OTR730 and anti-sense primer OTR731 and pKAL011 as a tem-
plate. The PCR products were digested with the flanking NotI site and cloned into the NotI site of pND003 and pND004, respectively generating pND009 and pND010. Finally, SV-hyg cassette containing flanking NheI sites was cloned into NheI site of pND009 and pND010 generating the final clones pND013 and pND014 (Figure 1B).

Plasmids for nonspecific RNA packaging
pT1R174 has been described previously [49] and was created to test the possibility of non-specific packaging of an over expressed RNA containing the SV-hyg cassette through "retrofection" [69,70] and uses SIV 3’LTR for transcript termination. pAG001 is similar to pT1R174 except that at the 3’end it contains FIV 3’LTR for transcript termination. Both of these control vectors lack all of the viral sequences at the 5’end (Figure 4A).

Envelope expression construct
All these transfer vectors were pseudotyped in a three plasmid trans-complementation assay using vesicular stomatitis virus envelope G (VSV-G) expression plasmid (MD.G) that has been previously described [22].

Transfection and infection of cells
The producer 293T cells were maintained and transfected by the calcium phosphate method as described earlier [71]. Seventy two hours post-transfection, viral supernatants were harvested for pelleting virus particles to isolate viral RNA and some portion was used to infect HeLa CD4+ cells as described previously [21]. Forty-eight hours following infection, cells that were infected with transfer vectors containing hygromycin resistance gene were selected with media containing hygromycin B phosphotransferase. Hyg colonies that appeared were stained and counted with media containing hygromycin B phosphotransferase.

Ultracentrifugation of virus particles
Viral supernatants collected seventy-two hours post-transfection were subjected to a low-speed centrifugation (4000 rpm for 10 minutes) for removal of cellular debris. Following which, viral particles were filtered using 0.2-micron syringe filters and ultracentrifuged using 20% sucrose cushion (using SW41 rotor at 26,000 rpm for 2 hours at 4°C) as described previously [24].

RNA isolation and Reverse Transcriptase Polymerase Chain Reaction (RT-PCR)
The transfected cells were taken off from the culture plates without trypsinization and fractionated into cytoplasmic and nuclear RNA fractions as described earlier [71]. For viral RNA isolation, pelleted virions resuspended in TNE buffer were lysed in 500 µl of Trizol LS reagent containing 8 µl of polyacryl (used as a carrier) as described earlier [24]. Following RNA isolation, both cytoplasmic and viral RNAs were DNase treated and tested by PCR to confirm the absence of any contaminating plasmid DNA that may have been carried over from the transfected cultures. After confirming that RNA preparations were devoid of any contaminating DNA, cytoplasmic and viral RNAs were reverse transcribed and amplified. cDNA prepared from the cytoplasmic RNA fractions were amplified using vector specific primers to monitor the expression of the transfer vector RNA in the cells. In addition, cytoplasmic fractions were also amplified to confirm the integrity of the fractionation process in a multiplex PCR in the presence of primers/competimers for 18S ribosomal RNA as a control for the presence of amplifiable cDNA in the PCR reactions (18S Quantum competitor control, Ambion, Austin, TX). Viral cDNAs were amplified using vector specific primer to determine the packaging of the transfer vector RNA. Amplified PCR products were analyzed on 2–4% agarose gels and, in some cases, the gels were further processed for Southern blot analysis as described previously [24]. Details of the primers used in cloning, RT-PCR, and Southern hybridization can be obtained from authors upon request.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
NSD, PSP, AG, JA, EB, and TAR performed the experiments (cloning, trans complementation assay, RNA extraction, RT-PCR, Southern blotting). SAJ performed the FACS analysis. We would also like to thank Ms. Suraiya Jahan Akhtar (United Arab Emirates University, Al Ain, UAE) for stimulating discussions and critical reading of the manuscript. The authors also wish to thank Ms. Noura Al Shamsi and Ms. Fatima Al Awadi for their assistance in cloning some of the vectors while volunteering in the laboratory.

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