P4HB knockdown induces human HT29 colon cancer cell apoptosis through the generation of reactive oxygen species and inactivation of STAT3 signaling

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Abstract. Colon cancer is the second most lethal malignancy worldwide. A better understanding of colon cancer at the molecular level may increase overall survival rates. Previous studies have indicated that prolyl 4-hydroxylase, β polypeptide (P4HB) is associated with tumorigenesis in colon cancer; however, its role and molecular mechanisms in colon cancer remain unclear. In the present study, the cellular responses to P4HB in human colon cancer cell lines were investigated by proliferation and apoptosis assays, western blotting, and immunohistochemistry. The results showed that expression of P4HB was higher in colon cancer tissues compared with adjacent normal tissues. P4HB knockdown increased the apoptosis of human HT29 cells. Furthermore, P4HB knockdown reduced the activation of signal transducer and activator of transcription 3 (STAT3) and promoted accumulation of reactive oxygen species (ROS). Inhibiting the accumulation of ROS abrogated the increased cell apoptosis induced by P4HB knockdown. Notably, decreased ROS levels effectively antagonized the effects of P4HB on STAT3 inactivation. In conclusion, these findings suggested that P4HB knockdown may induce HT29 human colon cancer cell apoptosis through the generation of ROS and inactivation of the STAT3 signaling pathway.

Introduction

Colon cancer is the second most lethal malignancy worldwide, which is associated with >600,000 cases of mortality per year (1). It is estimated that there are >1.4 million people living in the United States with this disease, and an additional 134,490 cases are diagnosed annually (2). Clinicians are faced with a great challenge with regards to colon cancer treatment and targeting the molecular features of this disorder are critical for therapeutic success (3); therefore, understanding the molecular pathogenesis of colorectal cancer is crucial for disease management.

Prolyl 4-hydroxylase, β polypeptide (P4HB) is the β-subunit of prolyl 4-hydroxylase, which acts as an endoplasmic reticulum chaperone to suppress aggregation of misfolded proteins (4). P4HB expression in lung cancer tissues is increased compared with adjacent tissues and normal lung epithelium, and it has been suggested to induce the growth of lung carcinoma (5). P4HB is also upregulated in high-grade glioma (6). Downregulation of P4HB decreases temozolomide resistance in malignant glioma via endoplasmic reticulum stress response pathways (7). Our previous study revealed that P4HB is upregulated in human hepatocellular carcinoma (HCC) tissues, and it promotes HCC cell proliferation and migration (8). These findings indicate that P4HB may serve an important role in tumorigenesis; however, little is currently known about the effects of P4HB and its underlying molecular mechanisms in colon cancer.

It is believed that reactive oxygen species (ROS) serve a crucial role in cell apoptosis, and increasing evidence has indicated that ROS regulate the apoptosis of cancer cells (9). ROS function as ‘redox messengers’ in intracellular signaling and regulation, whereas excessive ROS accelerate cell death (9,10). ROS result in a cellular redox imbalance in various cancer cells, which may be associated with oncogenic stimulation (11,12). Notably, the regulation of ROS may be a promising therapeutic approach against colon cancer. Chen et al (13) demonstrated...
that a novel benzimidazole acridine derivative induced human colon cancer cell apoptosis via the upregulation of ROS. ROS has also been revealed to enhance cisplatin-induced colon cancer cell apoptosis (14). Furthermore, Wang et al (15) revealed that dihydrotanshinone induces p53-independent but ROS-dependent apoptosis of colon cancer cells.

ROS may activate downstream signaling pathways to regulate the phosphorylation status of transcription factors, including signal transducer and activator of transcription (STAT) (16). STAT proteins are recruited to receptors by binding to phosphotyrosine residues in the Src homology 2 domain of the STAT protein (17). STAT is subsequently phosphorylated by Janus kinase 2 and translocated into the nucleus. The STAT family is a group of latent cytoplasmic proteins that regulate various metabolic processes (18,19). The STAT family includes seven structurally and functionally associated proteins: STAT1-4, STAT5a and b, and STAT6 (18). STAT3 is constitutively aberrantly activated in ~70% of human solid tumors, and it modulates the expression of oncogenes controlling the proliferation and metastasis of tumor cells (20,21). STAT3 is often a downstream effector of numerous oncogenic mutations (22). Substantive evidence has indicated that downregulating STAT3 may mitigate the malignant behavior of cancer cells (23,24).

The present study aimed to investigate the effects of P4HB on colon cancer. The results showed that P4HB was significantly upregulated in colon cancer tissues, whereas P4HB knockdown significantly increased cell apoptosis. Furthermore, P4HB knockdown reduced the activation of STAT3 and increased ROS accumulation. These data indicated that P4HB may inhibit colon cancer cell apoptosis via the ROS/STAT3 signaling pathway.

Materials and methods

Reagents. N-acetyl cysteine (NAC), an antioxidant, was purchased from Beyotime Institute of Biotechnology (Haimen, China). As described previously (15), colon cancer cells (5x10^5 cells/cm^2) were pretreated with NAC (10 mM) for 1 h at 37°C and 5% CO2.

Clinical tissues. The present study was approved by the Medical Ethics Committee of the Seventh People's Hospital of Shanghai University of Traditional Chinese Medicine (Shanghai, China). All subjects provided written informed consent, and none of them received chemoradiotherapy prior to surgery. From May 2015 to July 2017, 9 patients (5 males and 4 females) with an average age of 52.4 years were enrolled in the present study. Patients that underwent chemoradiotherapy prior to surgery were excluded. Colon cancer tissues and adjacent normal colon tissues were obtained during surgery.

Immunohistochemistry (IHC). According to the manufacturer's protocol, P4HB IHC staining was performed manually using a Bosterbio IHC kit (cat. no. RC1865; Boster Biological Technology, Pleasanton, CA, USA). Briefly, the clinical samples were fixed in 10% neutral formalin for 2 days at room temperature and embedded in paraffin, after which 3-μm sections were cut and mounted onto slides. Slides were incubated at 56°C, deparaffinized in xylene and dehydrated in a graded series of alcohol. Heat-induced (121°C) antigen retrieval was conducted with sodium citrate (pH 6.7) in a pressure-cooker for 30 min at room temperature. Following washing in wash buffer, peroxidase-blocking reagent (included in IHC kit) was applied for 15 min at room temperature. Subsequently, sections were incubated overnight at 4°C with rabbit anti-P4HB monoclonal antibody (1:100; ab137110; Abcam, Cambridge, UK). Subsequently, a secondary antibody (included in IHC kit) was applied for 30 min at room temperature. Horseradish peroxidase-streptavidin (included in IHC kit) was used to detect immunoactivity, followed by counterstaining with hematoxylin for 1 min at room temperature. Under a light microscope (magnifications, x100 and 200; Olympus Corporation, Tokyo, Japan), each section was imaged and semi-quantitatively analyzed according to a previously published method (25).

Cell culture. The HT29 human colon cancer cell line was purchased from the Cell Bank of Shanghai Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China). The cells were cultured in Dulbecco's modified Eagle's medium (DMEM; Sigma-Aldrich; Darmstadt, Germany) supplemented with 10% fetal bovine serum (FBS; HyClone; GE Healthcare, Chicago, IL, USA) at 37°C and 5% CO2.

Lentiviral infection. The HT29 human colon cancer cell line was used for lentiviral infection. A lentiviral short hairpin RNA (shRNA) construct targeting P4HB (SHCLNG-NM_000918) was obtained from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany) supplemented with 10% fetal bovine serum (FBS; HyClone; GE Healthcare, Chicago, IL, USA) at 37°C and 5% CO2.

Assessment of apoptosis. Apoptosis was evaluated using an Annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) dual staining kit (C1052; Beyotime Institute of Biotechnology). Following treatment (shRNA transfection or NAC pretreatment), the HT29 cells (1x10^5 cells) were trypsinized and centrifuged at 400 x g for 5 min at 4°C. Cells were then dissolved in 100 μl Annexin V-FITC binding buffer, and were...
incubated with 5 µl Annexin V-FITC and 5 µl PI for 15 min at room temperature in the dark. Harvested cells were analyzed by fluorescence-activated cell sorting using a flow cytometer.

Detection of ROS. According to a previous study (26), intracellular ROS can be detected using the peroxide-sensitive fluorophore 2',7'-dichlorofluorescein diacetate (DCF-DA; Beyotime Institute of Biotechnology). Briefly, the cells were plated in six-well plates, washed with DMEM without FBS, and incubated with 10 µM DCF-DA at 37˚C for 20-30 min. Fluorescence distribution was detected using a fluorescence spectrophotometer at an excitation wavelength of 488 nm.

Western blot analysis. Cells were lysed in radioimmunoprecipitation assay lysis buffer supplemented with a protease inhibitor (Beyotime Institute of Biotechnology). The concentration of total protein was detected by the BCA method. Whole cell extracts containing equal quantities of proteins (50 µg) were separated by 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis and were then transferred onto a polyvinylidene fluoride membrane. Following blocking in 5% bovine serum albumin (cat. no. BA7019, Boster Biological Technology) for 1 h at room temperature, the membranes were incubated overnight at 4˚C with antibodies specific to β-actin (1:1,000; cat. no. 4970), phosphorylated (p)-STAT3 (1:1,000; cat. no. 12640), B-cell lymphoma (Bcl)-2 (1:1,000; cat. no. 3498S; all Cell Signaling Technology, Inc., Danvers, MA, USA), cleaved caspase-3 (1:1,000; cat. no. 9661; Abcam), c-Myc (1:1,000; cat. no. 13987) and p53 (1:1,000; cat. no. 2527; both Cell Signaling Technology, Inc.). Horseradish peroxidase-conjugated goat anti-rabbit IgG (1:5,000; cat. no. BA1099; Boster Biological Technology) was applied as a secondary antibody for 1 h at 37˚C. For all western blots, β-actin served as an internal control. Protein expression was semi-quantified using Bio-Rad Quantity One software 3.76 (Bio-Rad Laboratories, Inc., Hercules, CA, USA).

Statistical analysis. Statistical analysis was performed using SPSS 18.0 software (SPSS, Inc., Chicago, IL, USA). All experiments were performed at least in triplicate. Data are presented as the means ± standard deviation. Statistical significance was determined using a two-tailed Student's t-test. Comparisons among multiple groups were analyzed by a one-way analysis of variance, followed by Tukey’s post hoc test. P<0.05 was considered to indicate a statistically significant difference.

Results

P4HB levels are lower in normal colon tissues than in colon cancer tissues. To assess the expression levels of P4HB in normal and cancerous colon tissues, IHC was performed on

Table I. Sequences interfering with P4HB.

| shRNA     | Sequence (5’-3’)                      |
|-----------|--------------------------------------|
| shRNA1    | CCGGGCTCCCATTTTGGGATAAACTGCTCGAGCAGTTTATCCCAAATGGGAGCTTTTTG |
| shRNA2    | CCGGAGGTGAAATCAGAATCTGATGAGATGTCTTGTTCACCTTTTTTTT |
| shRNA3    | CCGGGTGTGGTCACGTGAAACAGTGTTCAGAGAACCTTGGCAGTACCACACTTTTTT |
| ShCtrl    | CCTTCTCCGAACGTGTCACGT                |

P4HB, prolyl 4-hydroxylase, β polypeptide; shRNA, short hairpin RNA.
nine clinical specimens. Significantly higher P4HB levels were identified in colon cancer tissues compared with in normal colon tissues (Fig. 1; P<0.05).

**Knockdown of P4HB in colon cancer cells by lentiviral infection.** To investigate the effects of P4HB, lentiviral vectors were used to downregulate P4HB expression in colon cancer HT29 cells. As shown in Fig. 2, P4HB levels were knocked down using three shRNAs. shRNA3 was chosen for subsequent experiments, as it most effectively downregulated P4HB.

**Knockdown of P4HB inhibits proliferation and promotes apoptosis of human HT29 cells.** The effects of P4HB on cell proliferation were identified using the CCK8 assay, which revealed that proliferation was significantly reduced in the P4HB-knockdown group compared within the control group (Fig. 3A).

Cell apoptosis was evaluated by Annexin V/PI staining. The percentage of cells undergoing early or late apoptosis was demonstrated in Fig. 3B. Compared with in the control group, apoptosis was significantly increased by P4HB knockdown (P<0.05). As illustrated in Fig. 4A, the P4HB-knockdown group exhibited markedly higher protein expression levels of p53 and cleaved caspase-3 compared with in the control group. Furthermore, Bcl-2 levels were lower in the P4HB-knockdown group than in the control group.

**P4HB knockdown regulates p-STAT3 expression and promotes the accumulation of ROS.** To explore the molecular mechanism underlying the effects of P4HB on colon cancer cells, ROS generation and STAT3 levels, which are key regulators of colon cancer cell proliferation and apoptosis, were evaluated. P4HB knockdown markedly inhibited activation of p-STAT3 (Fig. 4A) and significantly increased the accumulation of ROS (Fig. 4B). Furthermore, a previous study demonstrated that ROS downregulates p-STAT3 by downregulating c-Myc (27); therefore, the expression of c-Myc was also examined. However, no significant difference was detected in the levels of c-Myc (Fig. 4A), which indicated that ROS may downregulate p-STAT3 levels via other signaling pathways.

**Inhibiting ROS accumulation rescues the increased apoptosis induced by P4HB knockdown.** NAC was used to scavenge intracellular ROS. According to a previous study (15), the cells were pretreated with NAC (10 mM) for 1 h. As demonstrated in Fig. 5A, intracellular ROS levels were decreased by NAC. The levels of apoptosis and apoptosis-associated proteins (Bcl-2, cleaved-caspase-3) were markedly decreased, whereas proliferation significantly increased in response to NAC compared with in cells without NAC (Fig. 5B-D). The increased apoptosis induced by P4HB knockdown was also reduced by NAC. Finally, NAC effectively reversed the effects of P4HB knockdown on the suppression of p-STAT3 (Fig. 5B).

**Discussion**

Colon cancer is one of most lethal malignancies, and there are still great challenges concerning its treatment. According to previous studies, P4HB is associated with tumorigenesis (5,6). The present study aimed to investigate the effects of P4HB on human colon cancer. In the present study, compared with in normal colon tissues, P4HB levels were significantly upregulated in colon cancer tissues. P4HB knockdown significantly decreased cell proliferation and increased cell apoptosis in a human cancer cell line. Additionally, downregulation of P4HB suppressed p-STAT3 expression and accelerated the accumulation of ROS. Notably, the increased apoptosis observed after P4HB knockdown was rescued by inhibition of ROS. Notably, the downregulation of ROS could also upregulate p-STAT3 levels. Taken together, these findings suggested that P4HB knockdown may decrease cell proliferation and increase cell apoptosis in a human cancer cell line.

A considerable amount of evidence has indicated that P4HB is associated with tumorigenesis (7,8). Our previous study reported that P4HB overexpression promotes HCC cell growth, migration, invasion and epithelial-to-mesenchymal transition, and the knockdown of P4HB expression by small molecules or small interfering RNA may be used as a therapeutic target in HCC (8). Apoptosis is a gene-directed program that eliminates excess, abnormal cells in vivo. A hallmark of cancer is the capability of cancer cells to resist cell death (28). Therefore, disturbed regulation of apoptosis is a vital factor in tumorigenesis (29). Evasion of apoptosis may have an important role in tumor initiation and therapy resistance (30). p53 is a tumor suppressor, which serves a key role in apoptosis and is frequently inactivated in colon cancer cells (31). In addition, Bcl-2 functions as a suppressor and is a key regulator of apoptosis. Alterations in the Bcl-2/Bax ratio may have an anti-apoptotic effect and may result in caspase activation, thereby inducing apoptosis (27). In the present study, depletion of P4HB upregulated p53 and cleaved caspase-3 expression, and inactivated the expression of Bcl-2. These data indicated that P4HB knockdown increased the apoptosis of colon cancer cells in vitro.

Elevated ROS levels may promote tumor onset and progression by increasing DNA damage and genomic instability (32). ROS-induced oxidative stress can also directly provoke programmed cell death, including apoptosis and autophagy (33). Ding et al. (34) revealed that ROS overload prompts apoptosis of colon cancer cells; therefore, targeting ROS in colon cancer cells may be exploited as an anticancer strategy. ROS may activate downstream signaling pathways to regulate the phosphorylation status of transcription factors, including STAT3 (16). Inappropriate activation of STAT3 serves a crucial role in the apoptosis of tumor cells. Chae et al. (35) demonstrated that CAY10598 induced apoptosis...
of colon cancer cells by activating ROS/STAT3 signaling. Kasiappan et al (27) also revealed that the ROS/STAT3 signaling pathway serves an important role in regulating the behaviors of tumor cells. Considering the critical role of ROS/STAT3 in cancer cells, the expression levels of ROS/STAT3 signaling pathway members were investigated in a colon cancer cell line. The present results were consistent with previous reports (27,35). The results demonstrated that P4HB knockdown induced colon cancer cell apoptosis by increasing ROS generation and downregulating STAT3, which may be associated with subsequent activation of the intrinsic apoptosis pathway. Furthermore, blocking the generation of ROS by NAC rescued the increased cell apoptosis induced by P4HB knockdown. Notably, the decreased ROS levels effectively antagonized the effects of P4HB on the activation of p-STAT3. Levels of ROS are controlled at both the level of production and by degradation. The predominant transcriptional response that increases the production of antioxidant proteins in cancer cells is through the activation of nuclear factor (erythroid-derived 2)-like 2 (NRF2). The levels of ROS have been reported to be regulated by antioxidant proteins, such as NRF2 (36). Notably, P4HB may be associated with the expression of NRF2 (36).

There are several limitations in the present study. Firstly, these in vitro results need to be verified in other colon cancer cell lines and in animal models. In addition, it is well known
that P4HB and its downstream targets may induce ROS accumulation; however, the mechanisms by which P4HB regulates ROS levels remain unclear. Therefore, further studies are required.

In conclusion, the present data suggested that P4HB knockdown may induce apoptosis of human colon cancer HT29 cells through the generation of ROS and inactivation of the STAT3 signaling pathway; however, these results require further investigation.

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Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Author's contributions
WX and JW designed the experiments. YZ, JY, QZ, QX and LL conducted the experiments. YZ and JY analyzed the data. WX wrote and revised the manuscript.

Ethics approval and consent to participate
The present study was approved by the Medical Ethics Committee of the Seventh People's Hospital of Shanghai University of Traditional Chinese Medicine (reference no. 2013876). All subjects provided written informed consent.
Inhibition of prolyl 4-hydroxylase, New benzimidazole acridine derivatives: Protein alterations associated with cancer treatment. Dihydrotanshinone induces gp53-independent but ROS-dependent apoptosis in colon cancer. gp53-independent but ROS-dependent apoptosis in colon cancer treated with Chinese medicines. Chin J Integr Med 21: 689-696, 2015.

6. Sun S, Wong TS, Zhang XQ, Pu JK, Lee NP, Day PJ, Ng GK, Lui WM and Leung GK: Protein alterations associated with temozolomide resistance in subclones of human glioblastoma cell lines. J Neurooncol 107: 89-100, 2012.

7. Sun S, Lee D, Ho AS, Pu JK, Zhang XQ, Lee NP, Day PJ, Lui WM, Fung CF and Leung GK: Inhibition of prolyl 4-hydroxylase, beta-polypeptide (P4HB) attenuates temozolomide resistance in malignant glioma via the endoplasmic reticulum stress response (ERSR) pathways. Neuro Oncol 15: 562-577, 2013.

8. Xia W, Zhuang J, Wang G, Ni J, Wang J and Ye Y: P4HB promotes HCC tumorigenesis through downregulation of GRP78 and repression of STAT3 protein by down-regulation of specificity protein 1 (SFP) in human colon cancer cells. Oncotarget 8: 689-696, 2017.

9. Circu ML and Aw TY: Reactive oxygen species, cellular redox systems, and apoptosis. Free Radic Biol Med 48: 749-762, 2010.

10. Lee JE, Park JH, Shin IC and Koh HC: Reactive oxygen species regulated mitochondria-mediated apoptosis in PC12 cells exposed to chlorpyrifos. Toxicol Appl Pharmacol 263: 148-162, 2012.

11. Matés JM and Sánchez-Jiménez FM: Role of reactive oxygen species in apoptosis: Implications for cancer therapy. Int J Biochem Cell Biol 32: 157-170, 2000.

12. Valko M, Rhodes CJ, Moncol J, Izakovic M and Mazur M: Free radicals, metals and antioxidants in oxidative stress-induced cancer. Chem Biol Interact 160: 1-40, 2006.

13. Chen K, Chu BZ, Liu F, Li B, Gao CM, Li LL, Sun QS, Shen ZF and Jiang YY: New benzimidazole acridine derivative induces human colon cancer cell apoptosis in vitro via the ROS-JNK signaling pathway. Acta Pharmacol Sin 36: 1074-1084, 2015.

14. He G, He G, Zhou R, Pi Z, Zhu T, Jiang L and Xie Y: Enhancement of cisplatin-induced colon cancer cells apoptosis by shikonin, a natural inducer of ROS in vitro and in vivo. Biochem Biophys Res Commun 469: 1075-1082, 2016.

15. Wang L, Yeung JH, Hu T, Lee WY, Lu L, Zhang L, Shen J, Chan RL, Wu WK and Cho CH: Dihydrotanshinione induces p53-independent but ROS-dependent apoptosis in colon cancer cells. Life Sci 93: 344-351, 2013.

16. Gao X and Schöttler B: Reduction-oxidation pathways involved in cancer development: A systematic review of literature reviews. Oncotarget 8: 51888-51906, 2017.

17. Silver DL, Naora H, Liu J, Cheng W and Montell DF: Activated signal transducer and activator of transcription (STAT) 3: Localization in focal adhesions and function in ovarian cancer cell motility. Cancer Res 64: 3550-3558, 2004.

18. Darnell JE Jr: STATs and gene regulation. Science 277: 1630-1635, 1997.

19. Levy DE and Darnell JE Jr: Stats: Transcriptional control and biological impact. Nat Rev Mol Cell Biol 3: 651-662, 2002.

20. Yao X, Liu H, Zhang X, Zhang L, Li X, Wang C and Sun S: Cell surface GRP78 accelerated breast cancer cell proliferation and migration by activating STAT3. PLoS One 10: e0125634, 2015.

21. Yeh JE and Frank DA: STAT3-interacting proteins as modulators of transcription factor function: Implications to targeted cancer therapy. ChemMedChem 11: 795-801, 2016.

22. Chai EZ, Shannumugam MK, Arluso F, Dharmarajan A, Wang C, Kumar AP, Samy RP, Lim LH, Wang L, Goh BC, et al: Targeting transcription factor STAT3 for cancer prevention and therapy. Pharmacol Ther 162: 86-97, 2016.

23. Furtek SL, Backos DS, Matheson CJ and Reigan P: Strategies and approaches of targeting STAT3 for cancer treatment. ACS Chem Biol 11: 308-316, 2016.

24. Nelson EA, Walker SR, Kepich A, Gashin LB, Hideshima T, Ikeda H, Chauhan D, Anderson KC and Frank DA: Nifuroxazide inhibits survival of multiple myeloma cells by directly inhibiting STAT3. Blood 112: 5095-5102, 2008.

25. Hu K and Olsen BR: Osteostatin-derived VEGF regulates osteoblast differentiation and bone formation during bone repair. J Clin Invest 126: 509-526, 2016.

26. Chen MF, Li YJ, Yang TL, Lou B and Xie XM: Losartan inhibits myocardin expression induced by ADMA via downregulation of chemokine receptors in monocytes. Eur J Clin Pharmacol 65: 457-464, 2009.

27. Kasiappan R, Jutooru I, Karki K, Hedrick E and Safe S: Benzy1 isothiocyanate (BITC) induces reactive oxygen species-dependent repression of STAT3 protein by down-regulation of specificity proteins in pancreatic cancer. J Biol Chem 291: 27122-27133, 2016.

28. Hanahan D and Weinberg RA: Hallmarks of cancer: The next generation. Cell 144: 646-674, 2011.

29. Zhang L, Cai Q, Lin J, Fang Y, Zhan Y, Shen A, Wei L, Wang L and Peng J: Chloroform fraction of Scutellaria barbata D. don promotes apoptosis and suppresses proliferation in human colon cancer cells. Mol Med Rep 9: 701-706, 2014.

30. Yaacoub K, Pedeux R, Tärte K and Guillaudeux T: Role of the tumor microenvironment in regulating apoptosis and cancer progression. Cancer Lett 378: 150-159, 2016.

31. Pavlou D and Kirimizis A: Depletion of histone N-terminal-acetyltransferase Naa40 induces p53-independent apoptosis in colorectal cancer cells via the mitochondrial pathway. Apoptosis 21: 298-311, 2016.

32. Teppo HR, Soini Y and Karhitala P: Reactive oxygen species-mediated mechanisms of action of targeted cancer therapy. Oncotarget 8: 14852-14863, 2017.

33. Zhang Y, Du Y, Yu W, Wang J, Kieffer N and Zhang J: Redox control of the survival of healthy and diseased cells. Antioxid Redox Signal 15: 2867-2908, 2011.

34. Ding Y, Wang H, Niu J, Luo M, Gou Y, Miao L, Zou Z and Cheng Y: Induction of ROS overload by alantolactone prompts oxidative DNA damage and apoptosis in colorectal cancer cells. Int J Mol Sci 17: 558, 2016.

35. Chae IG, Kim DH, Kundu J, Jeong CH, Kundu JK and Chun KS: Generation of ROS by CAY10598 leads to inactivation of STAT3 signaling and induction of apoptosis in human colon cancer HT29 cells. Free Radic Res 46: 1311-1321, 2014.

36. Lee LC, Weng YT, Wu YR, Soong BW, Tseng YC, Chen CM and Lee-Chen GF: Downregulation of proteins involved in the endoplasmic reticulum stress response and NFκB-ARE signaling in lymphoblastoid cells of spinocerebellar ataxia type 17. J Neural Transm (Vienna) 121: 601-610, 2014.