Fast adaptation of tropical diatoms to increased warming with trade-offs

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Ocean warming with climate change is forcing marine organisms to shift their distributions polewards and phenology. In warm tropical seas, evolutionary adaptation by local species to warming will be crucial to avoid predicted desertification and reduction in diversity. However, little is known about the adaptation of phytoplankton in warm seas. Across the ocean, diatomic microalgae are the main primary producers in cold waters; they also contribute to tropical communities where they play a necessary role in the biological pump. Here we show that four species of diatoms isolated from the tropical Red Sea adapted to warming conditions (30 °C) after 200–600 generations by using various thermal strategies. Two of the warming adapted species increased their optimal growth temperature \( T_{\text{opt}} \) and maximum growth rate. The other two diatoms did not increase \( T_{\text{opt}} \) and growth, but shifted from specialist to generalist increasing their maximum critical thermal limit. Our data show that tropical diatoms can adapt to warming, although trade offs on photosynthetic efficiency, high irradiance stress, and lower growth rate could alter their competitive fitness. Our findings suggest that adaptive responses to warming among phytoplankton could help to arrest the sharp decline in diversity resulting from climate change that is predicted for tropical waters.

Ocean warming with climate change is occurring at an unprecedented rate as a result of increasing loads of atmospheric CO\(_2\) and other greenhouse gases. Earth system models predict that ocean warming will result in a reduction in marine primary productivity (up to 20%) throughout the twenty-first century and driven by rising temperatures that exceed the limits of thermal tolerance and nutrient limitation in a warmer, more stratified ocean. Oceanic primary production is accomplished by photosynthetic microorganisms. Their short generation times and high population densities make evolutionary responses to climate change possible. There is growing evidence of adaptation by phytoplankton through evolutionary responses to global change drivers, such as elevated CO\(_2\), pollutants, and temperature. For example, previous studies reported fast adaptation of phytoplankton to warming after 80–450 generations. Latitudinal patterns in thermal growth responses suggest that phytoplankton are adapted to environmental temperature, with tropical species showing optimum growth at the high in situ oceanic temperatures. Rising temperatures leading to poleward shifts in the thermal niches of phytoplankton has been predicted by the end of this century along with a sharp decline in tropical phytoplankton diversity in the absence of evolutionary responses to warming. Thus evolutionary responses of phytoplankton to warming will be essential to avoid a reduction in the biodiversity of the tropical ocean where replacement by temperate species is not expected. Diatomic algae are a relevant group contributing largely to oceans primary production and biogeochemical processes. Diatoms dominate primary producers in cold waters, and although are not dominant in tropical waters still play a major role in the biological pump and other biogeochemical processes. Diatoms have lower activation energies (i.e. less sensitive to increasing temperature) than other phytoplankton groups, but its adaptation to warming will be crucial to maintain an efficient biological pump in the tropical ocean.

Thermal and ecological theory describe that organisms’ adaptation to temperature could be accomplished by different strategies resulting in different performances. Organisms may adapt by showing “horizontal shifts” in thermal optimum and thermal limits, and therefore warm-adapted organisms can achieve the same maximum performance of growth as cold-adapted ones. The “Hotter is better” hypothesis assumes that organisms’ performance will increase when warm adapted because temperature activates growth and other metabolic processes, and organisms adapted to lower temperatures are predicted to have lower maximum growth performances. Then, the “Hotter is better” hypothesis predicts that organisms will increase growth together with the increase in the thermal optimum. Organisms adapting to temperature can however move from “specialist” (i.e. growth is...
maximum at the optimum temperature) to "generalist" (maximum growth could be realized in a range of temperatures) and vice versa, with associated trade-offs in growth or other metabolic processes. If such trade-offs exist, and there is selection for an increased temperature range, specialist-generalist trade-offs would result in lower maximum performance for growth in warm-adapted organisms \(^{16}\). Thermal adaptation could generate a higher and broader growth curve (i.e. reaction norm), increasing the temperature range \(^{14}\). This adaptation is referred to as "hotter is broader and better" and will represent the win-win adaptation when organisms increased maximum growth and the breadth of reaction norm without trade-offs. For warm-adapted organisms, as those from the tropical seas, it is unclear what type of thermal adaptations could experience when increasing warming will force thermal selection constraining performance.

In order to test adaptation to warming temperatures of warm-adapted tropical organisms, we conducted a long-term experiment on four diatom phytoplankton species, Chaetoceros tenuissimus, Chaetoceros sp., Thalassiosira sp. and Synedra sp., isolated from the warm surface waters of the Red Sea. The four species of diatoms were maintained in the laboratory for about six months (~200–600 generations) at 26 °C (ambient temperature control) and at 30 °C (experimental warming conditions). 26 °C represents the mean Red Sea surface temperature (SST) for the 1982–2015 period (Fig. S1). For the projected warming conditions at 30 °C (mean Red Sea temperature +4 °C), we followed the high-emission scenario (RCP 8.5) projected for the turn of the next century (2100) by IPCC in 2014, that will represent a global mean warming ranging between 2.6 and 4.8 °C. The choice of ambient control temperature is problematic when considering interannual and seasonal variability \(^{22}\) as the sea surface temperature (SST) of Red Sea range from 21 to 32 °C \(^{23}\). However, phytoplankton species can adapt their realized niches to track average increases in water temperature regardless of annual and seasonal fluctuations, so the mean SST should be a reliable experimental control value \(^{24}\).

We investigated the thermal adaptation strategies of the four diatoms by combining experimental evolution with measurements of fundamental physiological performance (Fig. S2). The four species showed a fast adaptation to warmer temperature after 200–600 generations. Two of the species increased their optimal growth temperature (\(T_{\text{opt}}\)) and maximum growth rate. The other two diatoms did not increase \(T_{\text{opt}}\) and growth, but shifted from specialist to generalist increasing their maximum critical thermal limit. However, none of the species was able to improve both and did not experience a "win-win" adaptation. The results suggested that despite adaptation was fast the trade-offs associated constrained growth or photophysiological performance.

### Results

After the six-month temperature selection period, Chaetoceros sp. and Thalassiosira sp. produced a similar number of generations of 439 and 440 generations and 413 and 393 generations for the experimental ambient and warming conditions, respectively. Due to its higher specific growth rate, \(C. tenuissimus\) produced nearly 600 generations (ambient: 556 generations, warming: 570 generations), whereas Synedra sp. produced only 200 generations (ambient: 188 generations, warming: 194 generations) due to its lower growth rate.

We used the mixed-effects model (Table 1) to understand the selection effect of time and temperature on the four species. Increasing temperature significantly affected the growth of all four species of diatoms. Selection time \((F = 16.45, p < 0.001)\) and selection temperature \((F = 42.61, p < 0.001)\) as well as the interaction of selection time and selection temperature \((F = 189.34, p < 0.001)\) significantly affected the growth of Chaetoceros sp. (Fig. 1a, Table S1). Increased temperature increased the growth of Chaetoceros sp. at the beginning of the selection period, while this positive effect was reversed toward the end of selection period (Fig. 1a). Among Thalassiosira sp., the growth of warming-adapted cells was significantly lower than that of ambient-adapted cells \((F = 142.56, p < 0.001)\) (Fig. 1b, Table S1). Among \(C. tenuissimus\), increasing temperature increased the growth \((F = 543.58, p < 0.001)\), and this positive effect was amplified by selection time \((F = 39.00, p < 0.001)\) (Fig. 1c). Similarly, the growth of Synedra sp. was significantly affected by selection temperature \((F = 29.85, p < 0.001)\) (Fig. 1d), with a positive effect on growth throughout the selection period \((F = 2.92, p = 0.0889)\) (Fig. 1d). While two of the phytoplankton species \((C. tenuissimus\) and Synedra sp.) benefited from the temperature increase, the other two \((Chaetoceros sp. and Thalassiosira sp.) did not.

Adaptation to increased temperature was evident in changes in the growth rates in relation to thermal performance. In Chaetoceros sp., the maximum growth rate of warming-adapted cells and \(T_{\text{opt}}\) were similar to that of the ambient-adapted strain (Fig. 2a). However, the critical thermal minimum (\(CT_{\text{min}}\)) and critical thermal maximum (\(CT_{\text{max}}\)) shifted significantly in warming-adapted cells (Table 2, Table S2). Similarly, warming-adapted Thalassiosira sp. experienced relatively higher growth rates at high temperatures (Fig. 2d). In parallel, the \(CT_{\text{max}}\) shifted in warming-adapted Thalassiosira sp. cells (ambient: 38.44 ± 0.6 °C; warming: 47.34 ± 2.32 °C) and the thermal breadth also increased (expressed as \(B_{\text{opt}}\)) (Table 2, Fig. 2 and Table S2). The \(\mu_{\text{max}}\) of warming-adapted Thalassiosira sp. cells significantly decreased when compared with that of ambient cells \((t\text{-test}, t = 2.7633, p = 0.033)\) (Table S2, Fig. 2f), indicating the cost of thermal adaptation. There were no significant changes in \(T_{\text{opt}}\) between ambient and warming-adapted Thalassiosira sp. cells (Fig. 2e). However, there were significant shifts in \(T_{\text{opt}}\) in \(C. tenuissimus\) (ambient: 29.9 ± 0.16 °C; warming: 31.1 ± 0.10 °C, Fig. 2f) and Synedra sp. (ambient: 26.6 ± 0.08 °C; warming: 29.0 ± 0.14 °C, Fig. 2g), suggesting that a different thermal adaptation occurred in these two species (Stats see in Table S2) (Fig. 2h.k). Furthermore, thermal breadth narrowed in warming-adapted \(C. tenuissimus\) cells, mostly due to the increase in \(CT_{\text{min}}\) (Table 2). The \(\mu_{\text{max}}\) of \(C. tenuissimus\) and Synedra sp. both increased significantly in warming-adapted cells (Fig. 2i,j, and Table S2).

Any cost of thermal adaptation could be also reflected in differences in photosynthetic parameters. The maximum quantum efficiency (\(F_{\text{v}}/F_{\text{m}}\)) of photosystem II (PSII) decreased significantly in warming-adapted Thalassiosira sp. and Synedra sp. when compared with that of ambient-adapted cells assayed in their ancestral environment (Fig. S3b,d). The maximum electron transport rate (\(ETR_{\text{max}}\)) of warming-adapted Thalassiosira sp. cells were significantly lower than the ambient-adapted cells when both assayed at warming temperature (Fig. S4b). For warming adapted \(C. tenuissimus\), \(ETR_{\text{max}}\) significantly decreased compared with that of the
ambient-adapted cells assayed at the ambient temperature (Fig. S4c). Light usage efficiency, α, showed a significant increase in the warming-adapted Chaetoceros sp. cells assayed at the warming temperature compared with that of ambient-adapted cells assayed at the ambient temperature (Fig. S5a). Moreover, the warming-adapted cells of C. tenuissimus exhibited significantly higher α than did ambient-adapted cells when both were subjected to warming assay conditions (Fig. S5c), suggesting that the warming-adapted cells could maintain sufficiently high photosynthetic capacity at warm temperatures to support higher growth rates. Costs due to thermal adaptation were also reflected in the measure of saturated light intensity (Ik), with the warming-adapted C. tenuissimus and Thalassiosira sp. exhibiting significantly lower Ik when assayed under warming conditions compared with ambient-adapted cells (Fig. S6c), suggesting a decrease in tolerance to high irradiance among warming-adapted cells. The summary of the photophysiological responses pointed to Thalassiosira sp. as the species that suffered the larger trade-offs after adaptation to warming (Fig. 3).

Discussion
Overall, our study provides evidence that phytoplankton from the tropical Red Sea demonstrated the capacity to adapt fast to ongoing warming. However, the tested diatoms exhibited differing thermal adaptation capacities. The adaptation strategies of Chaetoceros sp. and Thalassiosira sp. followed a pattern of changing from “specialist” to “generalist” (Fig. 4a) by shifting the critical thermal minimum (CTmin) and maximum (CTmax) in warming-adapted cells, with no shifts in the optimal temperature. In this trade-off model, the phytoplankton

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| Species            | Model structure                                      | df | AIC | Log Lik | I-ratio | p      |
|--------------------|------------------------------------------------------|----|-----|---------|---------|--------|
| Chaetoceros sp.    | Random effect structure                               |    |     |         |         |        |
|                    | Random = ~1|d                                              |    |     |         |         |        |
|                    | Correlation structure                                |    |     |         |         |        |
|                    | corARMA(q = 1), varPower(−temperature)                |    |     |         |         |        |
|                    | Fixed effect structure                               |    |     |         |         |        |
|                    | Growth−1 + selection day*Tem                         | 8  | −589| 303     | <0.0001 |        |
|                    | Growth−1 + selection day + Tem                       | 7  | −485| 250     | 106     | <0.0001|
|                    | Growth−1 + selection day                            | 6  | −472| 242     | 15      | 0.0001 |
| Thalassiosira sp.  | Random effect structure                               |    |     |         |         |        |
|                    | Random = −1|d                                              |    |     |         |         |        |
|                    | Correlation structure                                |    |     |         |         |        |
|                    | corrAR1(), varPower()                                |    |     |         |         |        |
|                    | Fixed effect structure                               |    |     |         |         |        |
|                    | Growth−1 + selection day*Tem                         | 8  | −564| 290     |         |        |
|                    | Growth−1 + selection day + Tem                       | 7  | −560| 287     | 5.9     | 0.0149 |
|                    | Growth−1 + selection day                            | 6  | −477| 245     | 85      | <0.0001|
| Chaetoceros tenuissimus | Random effect structure                              |    |     |         |         |        |
|                     | Random = −1|d                                              |    |     |         |         |        |
|                     | Correlation structure                                |    |     |         |         |        |
|                     | corrAR1(), varPower()                                |    |     |         |         |        |
|                     | Fixed effect structure                               |    |     |         |         |        |
|                     | Growth−1 + selection day*Tem                         | 8  | −956| 486     |         |        |
|                     | Growth−1 + selection day + Tem                       | 7  | −929| 471     | 30      | <0.0001|
|                     | Growth−1 + selection day                            | 6  | −725| 368     | 206     | <0.0001|
| Synedra sp.         | Random effect structure                               |    |     |         |         |        |
|                     | Random = −1|d                                              |    |     |         |         |        |
|                     | Correlation structure                                |    |     |         |         |        |
|                     | corrAR1(), varExp()                                  |    |     |         |         |        |
|                     | Fixed effect structure                               |    |     |         |         |        |
|                     | Growth−1 + selection day*Tem                         | 8  | −556| 286     |         |        |
|                     | Growth−1 + selection day + Tem                       | 7  | −550| 282     | 8.2     | 0.0042 |
|                     | Growth−1 + selection day                            | 6  | −532| 272     | 20      | <0.0001|

Table 1. Linear mixed effects analysis for trajectories of the specific growth rate. Random effects on the slope and intercept were determined at the level of replicates nested within the selection temperatures. Analyses show that growth rates changed significantly over time and that growth trajectories were significantly different between selection temperatures. The most parsimonious model is highlighted in bold.
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improved their growth rates at higher temperatures and functioned over a wider range of high temperatures, whereas they reduced their growth capacity at the lowest temperatures. Furthermore, costs are likely to be associated with shifts of critical thermal limits\(^25\) because adaptations of enzymes and membranes to high temperatures may result in maladaptations to low temperatures\(^26,27\). The “hotter is better” hypothesis assumes that low temperature slows rates of biochemical reactions and organisms adapted to lower temperatures are predicted to have lower maximum performances (e.g., sprinting speeds, fitness, and growth rates) than those adapted to higher temperatures\(^18\). Then organisms adapted to warming therefore should have higher maximum growth rates and optimal growth temperatures than those adapted to low temperatures\(^18,19\). C. tenuissimus and Synedra sp. utilized a “hotter is better” strategy\(^25,27\) (Fig. 4) to adapt to warming by shifting their \(T_{\text{opt}}\) and increasing their \(\mu_{\text{max}}\) under warming conditions. However, C. tenuissimus showed trade-offs in their photosynthetic performance increasing \(\alpha\) and decreasing the light intensity to saturate photosynthesis, reducing tolerance to high light. Successful long time (one decade) adaptation to warming of the microalgae Chlamydomonas reinardtii, resulted in a significant increase in \(T_{\text{opt}}\) and maximum growth rate, accompanied by an increase in the photosynthetic capacity\(^15\). In fact, C. reinardtii showed the “hotter is better and broader” model, which implied no trade-offs\(^18\). Furthermore, C. tenuissimus and Synedra sp. benefitted from the temperature increase by increasing their growth, while the other two species of Chaetoceros sp. and Thalassiosira sp. did not. Warming adapted Thalassiosira sp. also showed a reduction on the photosynthetic performance at high light. Although the underlying mechanisms for the interspecific differences are unclear based on the present study, the various thermal adaptation strategies implied different trade-offs. Furthermore, costs are likely to associate with shifts of critical thermal limits\(^25\).

Diatoms become the predominant phytoplankton in the Red Sea when nutrient availability increases\(^28,29\). Diatoms proportion increase in the North during winter, in the cyclonic eddies and largely in the South, the warmest area in the Red Sea where temperature ranges between 26 and 32 °C and nutrient availability increases through the influx of Gulf of Aden Intermediate Water\(^28–30\). Their adaptation capacities to warming, on one hand, may help them to maintain their presence and distribution in the tropical ocean, maintaining the biogeochemical processes sustained by diatoms\(^16,31,32\). On the other hand, the trade-offs associated with the thermal adaptations may decrease their competitive fitness, and could reshape the communities.

A recent comparative analysis coupled with an eco-evolutionary species distribution model demonstrated that the geographic variation in thermal niches of phytoplankton closely matched the temperature regime of their natal environment\(^14\). This study suggests that rising temperatures will cause poleward shifts in the thermal niches of phytoplankton and a sharp decline in trophic phytoplankton diversity in the absence of evolutionary adaptation. Our long-term experiments on diatoms from a tropical sea suggested that ~200–600 generations (6 months) were sufficient to evoke adaptation to ocean warming. Our observations that the Red Sea diatoms could shift their optimal growth temperatures or critical thermal limits provide an empirical basis for parameterizing thermal evolution in eco-evolutionary models of phytoplankton dynamics. Our study suggests that rapid thermal adaptation may help to reduce the sharp decline in tropical phytoplankton diversity as projected in the absence of evolutionary responses.
Several recent studies have shown that adaptation to warming becomes evident after 80–100 generations\(^{12,13}\). However, experiments investigating evolutionary responses to environmental change in phytoplankton have thus far focused on phytoplankton from natal environments in the temperate and sub-tropical oceans. Our study builds on this knowledge by investigating adaptive responses in organisms isolated from one of the warmest seas in the world\(^{23}\). Our findings demonstrate that photosynthetic microorganisms living in oceanic extreme temperatures can adapt to future warming conditions. The tropical diatoms tested showed contrasting adaptation capacities with different strengths and weaknesses. Specialization to increased optimal temperatures optimizes responses against competitors at higher water temperatures, but the thermal niche allows a generalist species to persist in a changing thermal environment. However, these adaptation strategies involve costs for some species, associated with trade-offs, including reduction in the maximum growth or the photosynthetic capacity, that should be considered for projections (Fig. S7). Since the oceans are now undergoing multiple environmental changes including ocean acidification, deoxygenation, increased light exposure and decreased nutrient availability within the upper mixing layers\(^{33}\), the adaptation associated with such trade-offs may alter the resistance of a phytoplankton species to other stressors. At the same time, the thermal adaptation of phytoplankton under...

Figure 2. Patterns of thermal adaptation in Chaetoceros sp. (a–c), Thalassiosira sp. (d–f), Chaetoceros tenuissimus (g–i) and Synedra sp. (j–l) at ambient (black symbol) and warming (red symbol) selection temperatures. a, d, g and j, Thermal reaction norms for growth rates across a wide range of temperatures. Solid lines show the thermal reaction norms based on the thermal model. b, e, h and k, Box and whisker plots of the replicate-level (n = 4) optimal temperature for growth (\(T_{\text{opt}}\)) at ambient and warming selection temperatures. c, f, i and l, Box and whisker plots of the replicate-level (n = 4) maximum growth rate (\(\mu_{\text{max}}\)) at ambient and warming selection temperatures. Asterisks indicate significant differences between ambient and warming treatments based on the Students’ t-test.
natural conditions might also be amplified or retarded by other environmental changes and it might also depend on co-evolutionary interactions with other species that remain difficult to predict.

Fast organisms’ adaptation will be required in fast changing environments. The Red Sea is warming at $0.17 \pm 0.07 ^\circ \text{C} \text{ decade}^{-1}$, exceeding the global rate. Our results confirmed fast adaptation capacity to warming (i.e. within months) for Red Sea diatoms. found that phytoplankton species in the CARIACO station in a 15-years time series adapted to a change of 1 °C over a decade. Our study was conducted at fixed ambient and warming scenarios ignoring the temperature fluctuations in the natural environment and its influence on the rate of adaptation. Future studies with fluctuating treatment mimicking temperature variation in the region would contribute to help identify more accurate adaptation rates as well as examine thermal adaptation strategies and trade-offs.

In conclusion, our results suggest organisms from the warmest sea on earth can adapt fast to ongoing warming, on top of already experiencing thermal extremes, with trade-offs. Experiments examining evolutionary responses to increasing temperature in the ocean, like those performed in this study, provide important insights into responses of organisms to climate change. They also help to identify potential evolutionary winners and losers in the context of climate change. The thermal adaptation strategies reported in the present study and associated constrains to performance, may improve our knowledge on the ecological evolutionary responses to warming.

| Species          | Treatment  | $\text{CT}_{\text{min}}$ | $\text{CT}_{\text{max}}$ | $B_{\text{80}}$ |
|------------------|------------|--------------------------|--------------------------|-----------------|
| Chaetoceros sp.  | Ambient    | 13.66 ± 2.41a            | 36.76 ± 0.56a            | 8.93 ± 0.70a    |
|                  | Warming    | 17.27 ± 0.83b            | 39.02 ± 0.19b            | 9.72 ± 0.26b    |
| Thalassiosira sp.| Ambient    | 13.47 ± 1.53b            | 38.44 ± 0.60b            | 10.90 ± 0.70b   |
|                  | Warming    | 15.28 ± 0.93b            | 47.34 ± 2.32b            | 13.22 ± 1.00b   |
| Chaetoceros tenuissimus | Ambient | 10.40 ± 0.28a            | 41.31 ± 0.10b            | 12.66 ± 0.16b   |
|                  | Warming    | 13.37 ± 0.93b            | 41.66 ± 0.12b            | 11.71 ± 0.19b   |
| Synedra sp.      | Ambient    | 17.50 ± 0.58a            | 36.31 ± 0.56a            | 8.40 ± 0.45a    |
|                  | Warming    | 17.05 ± 0.66a            | 36.12 ± 0.68a            | 7.80 ± 0.23a    |

Table 2. The critical thermal minimum ($\text{CT}_{\text{min}}$, °C), critical thermal maximum ($\text{CT}_{\text{max}}$, °C) and 80% performance breadth ($B_{\text{80}}$, °C) of Chaetoceros sp., Thalassiosira sp., Chaetoceros tenuissimus and Synedra sp. under ambient and warming conditions. These three thermal reaction traits were derived from the nonlinear curve fitting according to equation (4) based on their thermal reaction norms (Fig. 2). Data are presented as means ± SE. The superscripts represent the significant difference between ambient and warming treatments in each species based on a Student’s $t$-test.
Methods

Species isolation and culture conditions. Four diatom species, *Chaetoceros* sp., *Thalassiosira* sp., *Chaetoceros tenuissimus* and *Synedra* sp., were isolated from coastal Red Sea waters near the Al Fahal Reef (22.2528°N, 38.9612°E). Surface water samples were collected and then were passed through a 45-μm filter. Approximately ten clonal cultures were established by single-cell isolation under a microscope. All cultures were maintained as batch cultures in filtered seawater that was taken from the same location and enriched with f/4 medium and silicate. The cultures incubated at 24 °C in a precise temperature-controlled incubator (Percival, United States). The cultures grew with a light: dark cycle of 12 h: 12 h under 50 μmol photons m⁻² s⁻¹.

After pure cultures of each species were established, mono-specific cultures of *Chaetoceros* sp., *Thalassiosira* sp., *Chaetoceros tenuissimus* and *Synedra* sp. were grown in 200 mL Erlenmeyer flasks at 26 ± 0.1 °C.  

Figure 4. Thermal adaptation optimality models for tropical diatoms. (a) Change from specialist to generalist trade-off. *Chaetoceros* sp. and *Thalassiosira* sp. conformed to this adaptation model in which the optimal temperature for growth did not shift, but the thermal niche width in warming-adapted cells (red lines) was wider than that of ambient-adapted cells (black line). The adaptation trade-offs of *Thalassiosira* sp. (dashed red line) imply decreasing maximum growth rates. (b) The “hotter is better” model predicts that populations adapted to higher temperatures (red line) increased their optimal temperature for growth and their maximum growth rate with respect to ancestors (black line), as observed in *C. tenuissimus* and *Synedra* sp. (c) The “hotter is better and broader” model was not utilized by diatoms in the present study. This model implies no trade-offs for adaptation to higher temperatures, resulting in increased maximum growth rate and optimal temperature and a wider thermal niche.
ambient, termed as ambient hereafter) and 30 ± 0.1 °C (experimental warming, termed as warming hereafter).

The design of the warming temperature that ignoring the regional temperature variation (both annual and seasonal) in the Red Sea rather serves as a temperature increase by average as projected by high-emission scenario (RCP 8.5, IPCC 2014) for the turn of next century, to explore the extent the adaptive capacity. The cultures grew under 400 μmol photons m⁻² s⁻¹ with a light: dark cycle of 12 h: 12 h. Four independent replicated cultures (n = 4) were run semi-continuously for about 6 months under ambient and warming conditions by renewing the medium every 3 days for Chaetoceros sp., Thalassiosira sp. and C. tenuissimus and every 7 days for Synedra sp. due to their lower growth rate. This long-term nature of the experiments allowed the diatoms to adapt to the experimental temperature environment. The initial cell concentration was set at 1000 cells mL⁻¹, and the medium was partially renewed every 3 or 7 days to restore the cell density to the initial level (i.e., growth batch cycle). Nutrients were not limiting as the cell abundances achieved at the end of the batch cycles were far from those expected at the stationary phase. Cell abundance was quantified every 3 days for Chaetoceros sp., Thalassiosira sp. and C. tenuissimus and every 7 days for Synedra sp., by examining the samples under an optical microscope (LEICA DMI 3000B-Germany) by hemocytometer. The specific growth rate, μ (d⁻¹), during each batch growth cycle was calculated as

\[ \mu = \frac{\ln(C_t - C_0)}{(t_1 - t_0)}, \]  

where \( C_t \) and \( C_0 \) are the cell densities at times \( t_1 \) and \( t_0 \) (\( t_1 - t_0 = 3 \) or \( 7 \) days), respectively. The number of generations per transfer (\( g \)) is equivalent to the number of doubles and was calculated as follows:

\[ g = \frac{t_1 - t_0}{\ln(2)/\mu}, \]  

where \( t_1 - t_0 \) is the time interval of the transfer (d), \( \ln(2)/\mu \) is the doubling time (d) and \( \mu \) is the specific growth rate (d⁻¹).

### Thermal growth curves

The thermal growth responses for all four diatoms adapted to ambient and warming conditions for about 6 months were determined at ten (Chaetoceros sp., Thalassiosira sp. and Synedra sp.) or twelve (C. tenuissimus, depending on its high thermal capacity) assay temperatures. At the end of the long-term selection period, cultures of ambient and warming conditions were inoculated into 200 mL flasks at an initial cell density of 1000 cells mL⁻¹ and then incubated at 18, 20, 22, 24, 26, 28, 30, 32, 34 and 36 °C (Chaetoceros sp., Thalassiosira sp. and Synedra sp.) and 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38 and 40 °C for species C. tenuissimus. To accommodate the different growth rates, batch cycles lasted 3 days for Chaetoceros sp. (~6-8 generations), Thalassiosira sp. (~5-7 generations) and C. tenuissimus (~9-11 generations) and 7 days for Synedra sp. (~7-8 generations) of the different temperatures. At the end of each growth cycle, the cell densities were determined by counting cell abundances under an optical microscope, and specific growth rates (μ) for each assay temperature were calculated from cell abundances versus time.

### Photosynthetic responses to temperature

Photosynthetic responses were measured at the end of each growth cycle using Pulse-Amplitude-Modulation (PAM) fluorometry (Phyto-PAM, Walz, Germany) at the ambient and warming assay temperatures. At the end of the long-term selection period, cultures of ambient and warming conditions were inoculated into 200 mL flasks at an initial cell density of 1000 cells mL⁻¹ and then incubated at 18, 20, 22, 24, 26, 28, 30, 32, 34 and 36 °C (Chaetoceros sp., Thalassiosira sp. and Synedra sp.) and 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38 and 40 °C for species C. tenuissimus. To accommodate the different growth rates, batch cycles lasted 3 days for Chaetoceros sp. (~6-8 generations), Thalassiosira sp. (~5-7 generations) and C. tenuissimus (~9-11 generations) and 7 days for Synedra sp. (~7-8 generations) of the different temperatures. At the end of each growth cycle, the cell densities were determined by counting cell abundances under an optical microscope, and specific growth rates (μ) for each assay temperature were calculated from cell abundances versus time.

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Photosynthetic responses were measured at the end of each growth cycle using Pulse-Amplitude-Modulation (PAM) fluorometry (Phyto-PAM, Walz, Germany) at the ambient and warming assay temperatures. At the end of the long-term selection period, cultures of ambient and warming conditions were inoculated into 200 mL flasks at an initial cell density of 1000 cells mL⁻¹ and then incubated at 18, 20, 22, 24, 26, 28, 30, 32, 34 and 36 °C (Chaetoceros sp., Thalassiosira sp. and Synedra sp.) and 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38 and 40 °C for species C. tenuissimus. To accommodate the different growth rates, batch cycles lasted 3 days for Chaetoceros sp. (~6-8 generations), Thalassiosira sp. (~5-7 generations) and C. tenuissimus (~9-11 generations) and 7 days for Synedra sp. (~7-8 generations) of the different temperatures. At the end of each growth cycle, the cell densities were determined by counting cell abundances under an optical microscope, and specific growth rates (μ) for each assay temperature were calculated from cell abundances versus time.

### Statistical analyses

We used the linear mixed-effects model to quantify trajectories of specific growth rates under experimental ambient and warming temperatures. For the analysis, μ was considered the dependent variable, time (in days) and treatment temperature were the fixed effects, while slopes and intercepts were treated as random effects at the level of replicates nested within the selection temperature35. The significance of the parameters was assessed using likelihood ratio tests, comparing models with common slopes and intercepts for each selection temperature (Table 1). Comparisons of models were done using Akaike’s information criterion (AIC) and likelihood ratio tests on models fitted with restricted maximum likelihood (REML) for comparison of random effects, and maximum likelihood (ML) for comparison of fixed effects36.

The thermal reaction norms of the four diatoms adapted to the two treatment temperatures were assessed by applying the equation described in refs14,37:

\[ rETR = \Phi_{PSII} \times 0.5 \times PAR, \]  

where \( \Phi_{PSII} \) is the photochemical quantum yield of PSII in light, \( PAR \) is the actinic light intensity (μmol photons m⁻² s⁻¹), and the factor 0.5 accounts for approximately 50% of all the absorbed energy allocated to PSII.

The photosynthetic parameters maximum electron transport rate (ETRmax), light usage efficiency (α) and saturated light intensity (Iκ) were derived from the fitted curves (Supplementary Methods).

### Statistical analyses

We used the linear mixed-effects model to quantify trajectories of specific growth rates under experimental ambient and warming temperatures. For the analysis, μ was considered the dependent variable, time (in days) and treatment temperature were the fixed effects, while slopes and intercepts were treated as random effects at the level of replicates nested within the selection temperature35. The significance of the parameters was assessed using likelihood ratio tests, comparing models with common slopes and intercepts for each selection temperature (Table 1). Comparisons of models were done using Akaike’s information criterion (AIC) and likelihood ratio tests on models fitted with restricted maximum likelihood (REML) for comparison of random effects, and maximum likelihood (ML) for comparison of fixed effects36.

The thermal reaction norms of the four diatoms adapted to the two treatment temperatures were assessed by applying the equation described in refs14,37:
where specific growth rate, $f$, depends on temperature, $T$, and is defined as a function of the parameters $z$, $w$, $a$ and $b$. $w$ is the temperature niche width (the range of temperatures over which the growth rate is positive), while the other three ($z$, $a$, $b$) possess no explicit biological meaning but interact to influence the rate of increase in growth with temperature, the maximum growth rate and the optimum temperature for growth. Specifically, $z$ determines the location of the maximum of the quadratic portion of this function, $a$ and $b$ are the Epply curve coefficient and Epply curve exponent, respectively. We estimated the critical thermal minimum ($CT_{\text{min}}$, the lowest temperature at which the growth of phytoplankton is zero), the critical thermal maximum ($CT_{\text{max}}$, the highest temperature at which the growth of phytoplankton is zero), the maximum growth rate ($\mu_{\text{max}}$), the optimal temperature for growth ($T^{\text{opt}}$), identified as the temperature at which the growth rate is maximal, and thermal breadth (expressed as $B_N$, 80% performance of the maximum growth rate breadth) by numerically maximizing the equation after estimating the parameter values for each replicate. We note that the estimated $\mu_{\text{max}}$ in ambient- and warming-adapted C. tenuissimus showed followed opposite trends with that of measured $\mu_{\text{max}}$. The measured $\mu_{\text{max}}$ values are presented in Fig. 2. For the other three species, $\mu_{\text{max}}$ is given in the values estimated by equation (4). Student’s $t$-test was used to test the differences in $CT_{\text{min}}$, $CT_{\text{max}}$, $T^{\text{opt}}$, $\mu_{\text{max}}$ and $B_N$ between two temperature-adapted populations.

References
1. IPCC Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part A: Global and Sectoral Aspects (eds Field, C. B. et al.) (Cambridge Univ. Press, 2014).
2. Bopp, L. et al. Potential impact of climate change on marine export production. Global Biogeochem. Cycles 15, 81–99 (2001).
3. Steinacher, M. et al. Projected 21st century decrease in marine productivity: a multi-model analysis. Biogeosciences. 7, 979–1005 (2010).
4. Laufkötter et al. Drivers and uncertainties of future global marine primary production in marine ecosystem models. Biogeosciences. 12, 6955–6984 (2015).
5. Falkowski, P. G., Fenchel, T. & Delong, E. F. The microbial engines that drive earth’s biogeochemical cycles. Science 320, 1034–1039 (2008).
6. Collins, S. Many possible worlds: Expanding the ecological scenarios in experimental evolution. Evol. Biol. 38, 3–14 (2011).
7. Lobbeck, K. T., Riebesell, U. & Reusch, T. B. H. Adaptive evolution of a key phytoplankton species to ocean acidification. Nature Geosci. 5, 346–351 (2012).
8. Jin, P., Gao, K. & Beardall, J. Evolutionary responses of a coccolithophorid Gephyrocapsa oceanica to ocean acidification. Evolution. 67, 1689–1878 (2013).
9. Schlüter, L. et al. (2014). Adaptation of a globally important coccolithophore to ocean warming and acidification. Nat. Clim. Chang. 4, 1024–1030 (2014).
10. Schlüter, L., Lobbecke, K. T., Gröger, J. P., Riebesell, U. & Reusch, T. B. H. Long-term dynamics of adaptive evolution in a globally important phytoplankton species to ocean acidification. Science Advances 2, e1501600 (2016).
11. Stachowski-Haberkorn, S. et al. Multigenerational exposure of the microalga Tetraselmis suecica to diuron leads to spontaneous long-term strain adaptation. Aquat. Toxicol. 140–141, 380–388 (2013).
12. Padfield, D., Yvon-Durocher, G., Buckling, A., Jennings, S. & Yvon-Durocher, G. Rapid evolution of metabolic traits explains thermal adaptation in phytoplankton. Ecol. Lett. 19, 133–142 (2015).
13. Stoude, E. et al. Adaptation of phytoplankton to a decade of experimental warming linked to increased photosynthesis. Nature Ecol & Evol. 1, 6994 (2017).
14. Thomas, M. K., Kremer, C. T., Klausmeier, C. A. & Litchman, E. A global pattern of thermal adaptation in marine phytoplankton. Science. 338, 1085–1088 (2012).
15. Smetacek, V. S. Role of sinking in diatom life-history cycles: ecological, evolutionary and geological significance. Mar Biol. 84, 239–251 (1985).
16. Agustí, S. et al. Ubiquitous healthy diatoms in the deep sea confirms deep carbon injection by the biological pump. Nat. Commun. 6, 7608, https://doi.org/10.1038/ncomms8608 (2015).
17. Chen, B. & Laws, E. A. Is there a difference of temperature sensitivity between marine phytoplankton and heterotrophs? Limnol. Oceanogr. 62, 806–817 (2017).
18. Frazier, M., Huey, R. B. & Berrigan, D. Thermodynamics constrains the evolution of insect population growth rates: “warmer is better”. Am. Nat. 168, 512–520 (2006).
19. Knies, J. L., Kingsolver, J. G. & Burch, C. L. Hotter is better and broader: thermal sensitivity of fitness in a population of bacteriophages. Am. Nat. 173, 419–430 (2009).
20. Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M. & Charnov, E. L. Effects of size and temperature on metabolic rate. Science. 293, 2248–2251 (2001).
21. Savage, V. M., Gillooly, J. F., Brown, J. H., West, G. B. & Charnov, E. L. Effects of body size and temperature on population growth. Am. Nat. 163, 429–441 (2004).
22. Boyd, P. W. et al. Physiological responses of a Southern Ocean diatom to complex future ocean conditions. Nat. Clim. Change. 6, 207–213 (2016).
23. Chait, V., Dreano, D., Agusti, S., Duarte, C. M. & Hoteit, I. Decadal trends in Red Sea maximum surface temperature. Sci. Rep. 7, 8144 (2017).
24. Irwin, A. J., Finkel, Z. V., Müller-Karger, F. E. & Ghinaglia, L. T. Phytoplankton adapt to changing ocean environments. Proc. Natl Acad. Sci. 112, 5762–5766 (2015).
25. Angilletta, M. J. Jr. Thermal Adaptation: A Theoretical and Empirical Synthesis (Oxford Univ. Press, 2009).
26. Portner, H. O., Van, Dijk P. L. M., Hardewig, J. & Sommer, A. Levels of metabolic cold adaption: tradeoffs in eurythermal and stenothermal ecototherms-Antarctic ecosystems: models for wider ecological understanding. (New Zealand Natural Sciences, 102–122, 2000).
27. Padfield, D. et al. Metabolic compensation constrains the temperature dependence of gross primary production. Ecol. Lett. 20, 1250–1260 (2017).
28. Pearman, J. K., Kürtén, S., Sarma, Y. V. B., Jones, B. & Carvalho, S. Biodiversity patterns of plankton assemblages at the extremes of the Red Sea. FEMS Microbiol Ecol. 92, fiw002 (2016).
29. Kheireddine, M. et al. Assessing Pigment-Based Phytoplankton Community Distributions in the Red Sea. *Front in Mar. Sci.* 4(32), https://doi.org/10.3389/fmars.2017.00132 (2017).
30. Ismael, A. A. Phytoplankton of the Red Sea. In *The Red Sea* (eds Rasul N. M. A & Stewart I. C. F) (Springer, 2015).
31. Nelson, D. M., Treguer, P., Brzezinski, M. A., Leinaert, A. & Queguiner, B. Production and dissolution of biogenic silica in the ocean: Revised global estimates, comparison with regional data and relationship to biogenic sedimentation. *Glob. Biogeochem. Cycles.* 9, 359–372 (1995).
32. Smetacek, V. Diatoms and the ocean carbon cycle. *Protists.* 150, 25–32 (1999).
33. Doney, S. C. et al. Climate change impacts on marine ecosystems. *Ann. Rev. Mar. Sci.* 4, 11–37 (2012).
34. Eilers, P. & Peeters, J. A model for the relationship between light intensity and the rate of photosynthesis in phytoplankton. *Ecol. Model.* 42, 199–215 (1988).
35. Pinheiro, J. & Bates, D. Mixed-Effects Models in S and S-PLUS. Springer Science & Business Media, New York (2006).
36. Zuur, A. F., Ieno, E. N., Walker, N. J., Sevělíková, A. A. & Smith, G. M. Mixed-Effect Models and Extensions in Ecology with R. Springer, New York (2009).
37. Boyd, P. W. et al. Marine phytoplankton temperature versus growth responses from polar to tropical waters-outcome of a scientific community-wide study. *PLoS ONE.* 8, e63091 (2013).
38. Huey, R. B. & Stevenson, R. D. Integrating thermal physiology and ecology of ectotherms: a discussion of approaches. *Am. Zoo.* 19, 357–366 (1979).

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Author Contributions

PJ. and S.A. conceived and designed the study. PJ. acquired the data and both authors analysed and interpreted the data. Both authors wrote the manuscript.

Additional Information

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