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Novel approach based on GQD-PHB as anchoring platform for the development of SARS-CoV-2 electrochemical immunosensor

Gustavo Martins\textsuperscript{a}, Jeferson L. Gogola\textsuperscript{a}, Lucas H. Budni\textsuperscript{a}, Maurício A. Papi\textsuperscript{a}, Maritza A.T. Bom\textsuperscript{b}, Maria L.T. Budel\textsuperscript{b}, Emanuel M. de Souza\textsuperscript{b}, Marcelo Müller-Santos\textsuperscript{b}, Breno C.B. Beirão\textsuperscript{c}, Craig E. Banks\textsuperscript{d}, Luiz H. Marcolino-Junior\textsuperscript{a}, Mário F. Bergamini\textsuperscript{a,*}

\textsuperscript{a} Laboratório de Sensores Eletroquímicos (LabSensE), Departamento de Química, Universidade Federal do Paraná (UFPR), CP 19032, CEP, 81531-990, Curitiba, PR, Brazil

\textsuperscript{b} Núcleo de Fixação de Nitrogênio (NFX), Departamento de Bioquímica e Biologia Molecular, Universidade Federal do Paraná (UFPR), CP: 19046, CEP: 81531-980, Curitiba, PR, Brazil

\textsuperscript{c} Laboratório de Imunologia Comparada, Departamento de Patologia Básica, Universidade Federal do Paraná (UFPR), CEP: 81531-980, Curitiba, PR, Brazil

\textsuperscript{d} Faculty of Science and Engineering, Manchester Metropolitan University, Chester Street, Manchester, M1 5GD, United Kingdom

\textbf{HIGHLIGHTS}

- An innovative approach for immunosensors construction based on the anchoring of biological material at screen-printed carbon electrode (SPE) modified.
- Graphene Quantum Dots (GQD) and polyhydroxybutyric acid (PHB) as functional surface to anchor the recognition site receptor-binding domain (RBD).
- Successfully detection of Anti-S antibodies (AbS) in human serum samples.
- Feasible strategy for construction of immunosensor by exploring a single-step.

\textbf{GRAPHICAL ABSTRACT}

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\textbf{ABSTRACT}

In the present work, we report an innovative approach for immunosensors construction. The experimental strategy is based on the anchoring of biological material at screen-printed carbon electrode (SPE) modified with electrophoresed Graphene Quantum Dots (GQD) and polyhydroxybutyric acid (PHB). It was used as functional substrate basis for the recognition site receptor-binding domain (RBD) from coronavirus spike protein (SARS-CoV-2), for the detection of Anti-S antibodies (AbS). SEM images and EDS spectra suggest an interaction of the protein with GQD-PHB sites at the electrode surface. Differential pulse voltametric (DPV) measurements were performed before and after incubation, in presence of the target, shown a decrease in voltametric signal of an electrochemical probe ([Fe(CN)\textsubscript{6}\textsuperscript{3/4-}]). Using the optimal experimental conditions, analytical curves were performed in PBS and human serum spiked with AbS showing a slight matrix effect and a relationship between voltametric signal and Ab concentration in the range of 100 ng mL\textsuperscript{-1} and 10 μg mL\textsuperscript{-1}. The selectivity of the proposed sensor was tested against yellow fever antibodies (YF) and the selective layer on the electrode surface did not interact with these unspecific antibodies. Eight samples of blood serum were analyzed and 87.5% of these total investigated provided adequate results. In addition, the present approach showed better results against...
1. Introduction

The recent pandemic caused by Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) showed how unprepared we are. Diagnosis sciences still in lack of a rapid, affordable, cheap and reliable diagnostic tests. Even with the globally coordinated research to respond to COVID-19 threats, a standard procedure to deal with the virus took some time to be set [1]. The gold-standard method for viral detection is a molecular diagnostic test aiming for viral RNA, using real time reverse transcriptase-polymerase chain reaction (RT-PCR). Frequently, RT-PCR assays are complemented by serological and immunological tests, such as indirect immunofluorescence test or enzymatic immunoassay (ELISA) [2,3]. On the other side, point-of-care (POC) devices such as lateral flow tests (LFT) are often alternatives for clinical trials, as they are cheap, rapid, and easy to use [4–6].

Electrochemical analyses are increasingly being used as rapid tests, since this methodology provides a straightforward and cheap arrangement due to the possibility of label-free detection [7–9]. In addition, the transducer can be modified with nanomaterials such as carbon nanotubes and graphene have been systematically explored as signal amplifiers [10]. Graphene Quantum Dots (GQD), a carbon-based nanomaterial, recently obtained great attention in the development of immunoensors. GQDs are small fragments of graphene sheets [11]. Thus, it shares graphene overall properties with enhancements on surface area, abundance of functional groups, ease of functionalization, higher biocompatibility and lower toxicity [11–13]. Therefore, GQD is a prosperous material for the development electrochemical immuno sensors, specially point-of-care focused ones. Since, its synthesis can be performed by incomplete pyrolysis of citric acid, an easy and cheap approach [14].

Protein anchoring on carbon functionalized materials traditionally employs crosslinking reactions to activate its surface and covalently bind the protein to the carbon surface [15]. As an alternative, polyhydroxyalkanoates (PHA) spontaneously bind to protein residues. Specifically, polyhydroxybutyrate (PHB) allows the synthesis of a core-shell nanostucture (100 nm) with a hydrophobic core and a surface exposing binding domains, that allows protein interaction and binding [16–18]. Similarly, this anchoring system is suitable for biosensors construction and application, however poorly explored in the development of electrochemical biosensors [19,20].

For this purpose, electrochemical immuno sensors combine the inherent specificity of immunoreactions with the high sensitivity of electrochemical sensors and thus sustain the development of the next generation of POC tests. Therefore, here we propose the development of an electrochemical immunosensor for the detection of SARS-CoV-2 antibodies based on bioengineered PHB and GQD as anchoring platforms on screen-printed electrodes.

2. Materials and methods

2.1. Materials and reagents

All chemicals were analytical or high-purity grade. N-(3-Dimethylaminopropyl)-N′-ethylcarbodiimidehydrochloride (EDC), N-Hydroxysuccinimide sodium salt (NHS), Potassium Ferricyanide K₃[Fe(CN)₆] and citric acid were purchased from Sigma Aldrich. Phosphate buffer saline, pH 7.4, were prepared with sodium chloride, potassium chloride, potassium phosphate monobasic and disodium hydrogen phosphate all from Sigma Aldrich. Bovine serum album (BSA) and Antibody against Yellow Fever (YF - 0.16 mg mL⁻¹) were provided by Molecular Virology Laboratory at Carlos Chagas Institute – FIOCRUZ/PR.

The screen-printed electrodes (SPE) were obtained from Professor Craig, E. Banks laboratory at Manchester Metropolitan University. Working and counter electrodes are made of graphite conductive ink, and an Ag|AgCl ink was used for the pseudo-reference electrode construction. The final set is supported on a polyester substrate with electrodes and electrical contacts delimited by an insulating ink.

Polyhydroxybutyrate particles were produced by the nanoprecipitation method [21]. 10 mg of PHB (Sigma Aldrich, #363502) was dissolved in 10 ml of TFE (2,2,2-trifluoroethanol) (Sigma Aldrich, #T63002) with stirring at room temperature. The solution was dialyzed through a 12 kDa cut-off cellulose membrane (Sigma Aldrich, #D97777) against 2 L of distilled water for 24 h.

2.2. Expression and purification of the recombinant RBD (receptor binding domain) from the SARS-CoV-2 spike protein

The amino acid sequence between the residues 319 to 541 from SARS-CoV-2 Spike protein (NCBI, Gene ID: 43740568) was codon-optimized and fused to the Substrate-Binding Domain (SBD) of PHA depolymerase of Alcaligenes faecalis [22] followed by a 6-histidines tag in the C-terminal. Twist Bioscience (South San Francisco, CA, USA) synthesized the synthetic construct on-demand and cloned it into the NdeI and Xhol sites of the pET29b(+), generating the pET29-RBD-SBD-6His. The 6-histidines tag was used to purify the protein by affinity chromatography against nickel (Ni²⁺) ions. The SBD domain interacts with PHB as previously described [22].

E. coli BL21(DE3) carrying the pET29-RBD-SBD-6His was cultivated in 80 mL of Lysogeny Broth [23]. The concentration of RBD-SBD was measured by the Bradford method using BSA as standard [24]. For the sake of clearness, hereafter, we referred to RBD-SBD as RBD.

2.3. Electrochemical measurements

Cyclic voltammetry (CV), differential pulse voltammetry (DPV) and Electrochemical impedance spectroscopy (EIS) measurements were performed with SPE on a Potentiostat/Galvanostat PGSTAT204. Experiments were conducted in PBS 0.1 mol L⁻¹, pH 7.4, with 2.0 mmol L⁻¹ of (K₃[Fe(CN)₆]) as the electrochemical probe. The progression of the proposed immunosensor was evaluated by the suppression anodic current peak after each anchoring step, since proteins are essentially nonconductors. EIS were performed with probe anodic peak potential and 10 mV amplitude.

2.4. Morphological and structural characterisation

Scan electron microscopy (SEM) images were performed in a FEI High-Resolution Scanning Electron Microscope, Felmiz ZFE - model Quanta 450, with a field emission gun (FEG) electron source, which has a resolution of 1.0 nm. All images were obtained with 10 kV voltage acceleration and secondary electron detectors. Elemental chemical analysis of EDS has resolution of 131 eV with a Apollo X SDD detector.

The Infrared spectra were obtained with a BOMEN spectrometer, 64 scans from 4000 cm⁻¹ to 500 cm⁻¹. Samples were previously homogenized with KBr pastille and dried.

2.5. Immunosensor construction

First, QGD was electrodeposited on the SPE surface. This step was performed by CV throughout –1.4 to 0.0 V, 50 mVs⁻¹, 3 until 10 cycles. Afterwards, 2.0 mL of PHB particles solution was added directly to the working electrode (WE), and it was allowed to dry in controlled
temperature \(37 \, ^{\circ}\) C). Following this step, RBD was added (2.0 \( \mu \)L) to WE and incubated at 37 \( ^{\circ}\) C in a saturated water atmosphere, every incubation was performed under this condition. All incubation times were monitored, ranging from 15 min to 60 min. Sequently, electrodes were 3 times washed by immersion on PBS solution, in order to remove excess and not binded biomolecules, this step was repeated after each incubation. Next step consisted of block PHB residual active sites, that did not bind to RBD, in order to mitigate parallel interactions. This was performed by adding 2 \( \mu \)L of BSA (1.0 mg mL\(^{-1}\)) and incubating for 15 min at 37 \( ^{\circ}\) C. Finally, antibody detection (AbS) was done by adding and incubating 2 \( \mu \)L of AbS solution. Each step involving the construction of the sensor was evaluated by CV, 0.3 to 0.6 V, 50 mV s\(^{-1}\), 3 cycles.

Scheme 1. Immunosensor step by step build-up.

2.6. Selectivity, analytical curve and sample analysis

Positive and negative (IgG) human serum samples were obtained from local hospital laboratories. Use of human samples was ethically assessed and approved by the Committee on Research Ethics, protocol number 30342520.5.0000.0008.

Human serum samples were 1000 \( \times \) diluted on PBS solution. The diluted serum samples were added to W.E. (2 \( \mu \)L), incubated for 60 min at 37 \( ^{\circ}\) C and sequentially washed. Electrochemical signals were measured before and after incubation. Selectivity assay were performed with 1000 \( \times \) diluted negative serum samples. Negative samples were spiked with AbS and YF antibodies (0.47 and 1.6 \( \mu \)g mL\(^{-1}\) final concentration, respectively). Two analytical curves were performed one on 0.1 mol L\(^{-1}\) PBS and another on 1000 \( \times \) diluted negative human serum sample spiked with AbS (ranging from 25 \( \mu \)g mL\(^{-1}\) to 50 ng mL\(^{-1}\), final concentration).

3. Results and discussions

3.1. GQD synthesis, characterization and immunosensor construction

PHB is a polymer produced by microorganisms as a form of energy storage [25]. PHB particles has shown stable interaction with biomolecules and also can be bioengineered to express specific ligands to allow a specific interaction with biostuctures. Here, PHB particles were used to modify the electrode surface and interact with SARS-CoV-2 RBD domain from the Spike protein (S1+S2 structure) one step later to create the selective layer.

GQD synthesis was carried out as described by Gevaerd et al. [26]. Synthesized nanoparticles were evaluated through FTIR to verify functional groups. Each step of immunosensor assemble was characterized by CV, SEM and EDS to evaluate material morphology and electrode surface after each modification. FTIR, SEM/EDS images and electrochemical behavior of the electrode with GQD, PHB and proteins are presented in Fig. S1A (FTIR) and S1C (SEM/EDS) and discussed in supplementary material (Section S1).

Fig. S1B shows K\(_3\)[Fe(CN)]\(_6\) voltammetric behavior recorded after each modification. After GQD electrodeposition, the redox signal intensified from 7.98 \( \mu \)A to 12.09 \( \mu \)A (\(|\Delta I| = 4.11 \, \mu A \pm 3.88\%\), n = 3) as a result of GQD properties. GQD use in this system resides as current
amplifier and also creates an anchoring interface between the SPE surface and PHB, through interaction with its highly functionalized surface. QGD PHB was subsequently added by drop-casting 2.0 μL (8.33 μg mL⁻¹) and dried at 37 °C. Comparing PHB behavior with QGD modified electrode, I_p increased from 12.09 to 15.15 μA (ΔI_p = 3.06 ± 6.5%, n = 3), suggesting that PHB was successfully immobilized on the electrode surface. Observed ΔI_p may be associated to QGD rearrangement due to an interaction with PHB, thus a larger electroactive area is available and a higher I_p is registered. Next step consisted of anchoring RBD upon PHB, since the polymer particles self-assemble to proteins expressed with the SBD domain, already described in the literature [22], the incubation step can be performed. Therefore, 2.0 μL of RBD solution (0.47 μg mL⁻¹) was added to the WE surface and incubated in a water-saturated atmosphere at 37 °C. This condition allows the incubation to be carried out at 37 °C as it prevents the protein solution to dry and favors binding kinetics without compromising protein conformation. After incubating, the electrode was washed 3 times by immersion in PBS solution and CV measurements were performed. CV recorded after RBD anchoring displays a decrease in I_p (ΔI_p = -2.36 ± 4.7%, n = 3), due to the hindering effect caused by protein binding over the PHB. Thus, RBD compromises electroactive area, as it is an insulator.

Last step consisted of blocking remaining active sites that RBD did not occupy. For this purpose, BSA (2.0 μL, 1.0 mg mL⁻¹) was anchored. BSA binds to such active sites, ensuring that the interaction of antibodies with the device may only occur by binding with RBD. The anchoring process was evaluated by the decrease of redox probe signals, since BSA causes an insulating effect, as can be seen on the CV.

The detection of IgG antibodies is based on the interaction between Abs and RBD, anchored at the electrode surface, thus forming the immunocomplex RBD-Abs. The RBD-Abs complex creates not just an insulating effect, but even a steric impediment, which affects the interaction between the redox probe and the electrode surface, leading to a decrease in the faradic signal. Therefore, CV measurements showed a reduction of I_p (ΔI_p = -2.66 ± 16.7%, n = 3), as shown in Fig. S1B –BSA-Abs. Each step was also evaluated through EIS, Fig. S2, and shows ΔR_a concordant with CV data. Through the fit of equivalent circuit shown at Fig. S2, R_a values were calculated and found to be 3.43 kΩ for bare electrode; 2.29 kΩ after QGD electrodeposition; 1.58 kΩ for PHB adsorption; 3.5 kΩ for anchoring RBD process; 4.7 kΩ after BSA and 6.1 kΩ to Abs detection and immunocomplex formation.

Alternatively, it was evaluated the construction of the immunosensor by exploring a single-step approach after QGD electrodeposition. In order to demonstrate this feature, a mixture of PHB, RBD and BSA was prepared under the same conditions previously applied. The device was step by step evaluated by CV (Fig. S3A). QGD was electrodeposited, followed by drop-casting PHB-RBD-BSA mixture and dried at 37 °C. A decrease of I_p is observed from the comparison of QGD voltammetric behavior and the mixture, better evidenced in Fig. S3B. The decrease is attributed to the mixture immobilization and its protein constituents, which partially blocks the electroactive surface. Abs detection was also evaluated and showed a decrease in I_p due to the immunocomplex formation, as the main factor of steric impediment and resistance to charge transfer.

For comparison purposes, the same electrochemical experiments and anchoring were performed with EDC/NHS, a well-known crosslinking agent [27] for binding proteins (Fig. S3C). The overall build-up of the device shows similar electrochemical behavior to PHB (Fig. S3D). The proposed method was optimized, aiming to enhance the sensitivity, reproducibility and feasibility of the sensor. Parameters involving the immunosensor assemble like QGD electrodeposition, modification with PHB and RBD immobilization (time and concentration), were evaluated. Tested and optimized parameters are summarized and discussed in supplementary material (Section S2).

3.2. Analytical performance, effect of matrix, selectivity and sample analysis

To evaluate how the optimized sensor responds to different target antibody concentrations, the proposed immunosensor was subjected to incubation with SARS-Cov-2 antibodies between 25 μg mL⁻¹ and 50 ng mL⁻¹ in PBS solution (Fig. 1). In order to verify the matrix effect on voltammetric response, a similar experiment was carried out using 1000x diluted human serum sample (negative) spiked with different Abs concentrations (Fig. 1).

For both studies, a relationship was observed between the decrease of faradic signal and Abs concentration from 100 ng mL⁻¹ to 1 μg mL⁻¹ ΔI_p (%), n = 5, ±SD) obtained on 0.1 mol L⁻¹ PBS medium and 1000x diluted negative serum sample. LOD was set as the lowest detectable value for both curves, 100 ng mL⁻¹. In addition, measurements recorded with Abs 25 μg mL⁻¹ did not show variation in the voltammetric signal. This behavior can be related to the hook effect, which is a phenomenon whereby the effectiveness of antibodies to interact and form an immunocomplex is impaired when the concentration of an antibody or antigen is very high. Thus, the formation of immunocomplexes is lowered with higher concentrations [28,29]. The correlation curve obtained by fortified PBS solution showed a higher sensitivity to Abs detection compared to the human serum sample (Fig. 1). A sensitivity around 10% lower using human serum was observed when compared to PBS solution. A crucial step for diagnostics through electrochemical measurements is the response in sample conditions since the complexity of the matrix can lead to unspecific interactions resulting in a false-positive diagnosis. The results suggested a slight effect on the response, which was not considered a drawback for the proposed immunosensor since the biological sample used is a very complex matrix.

Here, the combination of QGD and PHB as platforms to anchor protein and/or other biological components should be highlighted since it is a quick and feasible way to construct immunosensors. Overall performance and assembly time of QGD-PHB system is comparable with literature, which highlights the competitiveness of application when comparing to well-established methodologies. Table 1 summarizes other reports aimed at detecting COVID using well-known strategies for sensor preparation.

Selectivity is one of the top requirements for immunosensors, with false positives being a common hurdle to overcome in most diagnostic methods. The selectivity of the proposed sensor was tested against YF antibodies, as a control, and compared with the detection of the
intended Anti-S antibodies, with YF antibodies as an interferent as well. The signal obtained before and after the interaction with each antibody and the comparison between the two procedures is shown at Fig. 2 A.

As observed in Fig. 2 A, there is no statistical difference between the signal obtained before and after incubation of control antibodies (YF) ($t_{\text{calculated}} - 1.243 \leq t_{\text{critical}}$. This result indicates that the selective layer on the electrode surface did not interact with unspecific antibodies or YF. Fig. 2 A also shows the signal before and after the incubation with both antibodies (AbS and YF - control), which AbS has provide a shift in the signal, implying an interaction with the device (RBD). In contrast, YF antibodies show no competition or interference for the detection.

The proposed sensor showed low interaction with the sample matrix, that paired with the good selectivity tested beforehand, allowed to differentiate 0.50 $\mu$g mL$^{-1}$ of specific SARS-CoV-2 antibodies from a negative serum sample. Despite the number of antibodies in samples being dependent on the patient immune system and the chronology of the disease, varying through a wide range, the use of the GQD-PHB-RBD sensor can help diagnose the early onset of the Covid-19 disease.

Once the proposed sensor showed good selectivity, it was tested against positive and negative human serum samples (1000x diluted). Fig. 2B shows representative DPV measurements carried out using the proposed immunosensor. Overall negative serum samples measurements did not alter the redox signal, while COVID-19 positive serum samples decreased $I_{p}$. An overview of 8 serum samples analysis is summarized in Fig. 2 C. Negative serum samples did not show a statistical difference from the blank signal ($t_{\text{calculated}} < t_{\text{critical}}$. However, negative serum sample 2- has shown a slight statistical difference ($t_{\text{calculated}} > t_{\text{critical}}$), indicating a false positive test. Meanwhile, positive serum samples showed a difference ($t_{\text{calculated}} > t_{\text{critical}}$) for all samples analyzed, clearly differentiating positives from negatives. Using eight serum samples, only one false positive and no false negatives were obtained. Thus, around 87.5% of samples investigated provided adequate results, but a more significant sample set should be evaluated to determine the device’s effectiveness and its reliability as a point-of-care technique for diagnosing COVID-19. In addition, stability of response was evaluated considering immunosensor built and RBD-PHB dispersion. It was observed good performance for at least one week with the sensor “ready-to-use” and one month for solution.

### Table 1

| Anchoring System       | Target                  | LOD         | Sample Assembly | Ref.   |
|------------------------|-------------------------|-------------|-----------------|--------|
| SPAuE-Peptide          | S-Protein               | 18.2 ng mL$^{-1}$ | Nasopharyngeal swab | 120 min. | [30] |
| Au-Anti-RBD            | S-Protein               | 0.1 mg mL$^{-1}$ | Human serum     | 1 - 3 days | [31] |
| Graphene – Anti-S-RBD  | Anti-S (IgG/IgM)        | ND          | Blood and saliva| 315 min. | [32] |
| NCM                    | Biotinylated Antibody (SARS-COV-2) | 0.77 ng mL$^{-1}$ | Buffer solution | 240 min. | [33] |
| SPCE-CB                | Anti-N (IgG)           | 2 ng mL$^{-1}$ | Human blood     | 60 min  | [34] |
| Co-TiO$_2$ nanotubes   | S-protein               | 14 nM       | Nasopharyngeal and saliva | N.D. | [36] |
| ePAD-GO                | Anti-S (IgG/IgM)       | 0.11 ng mL$^{-1}$ | Human serum     | 180 min. | [37] |
| SPE-GQD-PHB            | Anti-S (IgG)           | 100 ng mL$^{-1}$ | Human serum     | 120 min. | This Work |

Stencil printed carbon electrode – SPCE; Screen-printed Au Electrode – SPAuE; Nitrocelullose membrane – NCM; Screen printed electrode – SPE; Polyhydroxibutirate – PHB; Graphene Quantum Dots – GQD; Carbon Black – CB; Graphene oxide – GO; electrochemical paper-based analytical device – ePAD.

Fig. 2. (A) DPV data summarized for selectivity assay on negative serum sample against YF antibodies (n = 5, ±SD). (B) Representative DPV measurements obtained for positive and negative serum sample assays. (C) Summarized DPV assays for negative and positive human serum assays (n = 5, ±SD).
4. Conclusion

The use of PHB showed potential for developing biosensors, as it is cheap, easy to obtain and can be scaled to industrial production. Its application relies on the self-assembly of proteins over its structure and the synergistic effect of biocompatibility, with no toxicity. In the present study, PHB showed better results than the traditional EDC/NHS protein coupling reaction. There were enhancements in electrode preparation time and the possibility of developing an immunosensor in a single drop since the proteins can be anchored prior to the electrode modification step. The proposed sensor showed selectivity against Yellow Fever antibodies and could differentiate COVID-19 negative from positive serum samples with results comparable with the literature. Thus, PHB grants a simpler array and has great potential for developing electrochemical biosensors, as demonstrated here for SARS-CoV-2.

CRediT authorship contribution statement

Gustavo Martins: Conceptualization, Formal analysis, Investigation, Methodology, Validation, Writing – original draft. Jefferson L. Gogola: Conceptualization, Methodology, Investigation, Lucas H. Budni: Investigation. Maurício A. Papi: Investigation. Mariza A.T. Bom: Methodology, Investigation. Maria L.T. Budel: Methodology, Investigation. Emanuel M. de Souza: Supervision, Resources, Funding acquisition, Writing – review & editing. Marcelo Müller-Santos: Supervision, Resources, Funding acquisition, Writing – review & editing. Breno C.B. Beirão: Supervision, Resources, Funding acquisition, Writing – review & editing. Craig E. Banks: Resources, Writing – review & editing. Luiz H. Marcolino-Junior: Conceptualization, Resources, Funding acquisition, Supervision, Writing – review & editing. Márcio F. Bergamin: Conceptualization, Resources, Funding acquisition, Writing – review & editing.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

No data was used for the research described in the article.

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Appendix A. Supplementary data

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M.Z. Rashed, J.A. Kopechek, M.C. Priddy, K.T. Hamorsky, K.E. Palmer, N. Mittal, M. Daccord, E. Regalbuto, P. Salvatori, G. Terova, D. Moscone, F. Lista, F. Arduini, L.P.S. Alves, E.M. De Souza, D. Jendrossek, A. Amore, C. B. Beirão, W. Khamcharoen, I. Jang, W. Siangproh, Mauricio A. Papi: received his Ph. D. degree at Federal University of Paraná (UFPR), Brazil, in 2007. He is a Professor in the Chemistry Department at Federal University of Paraná (DQ-UFPR), Curitiba-PR, Brazil. His current research interests are development of electrochemical sensors using nanostructured materials and development of microfluidic devices with electrochemical detection.

Marcio Fernando Bergamini received his Ph.D. degree in Analytical Chemistry from Federal University of São Carlos (UFSCar), Brazil, in 2007. He is a Professor of Chemistry at Federal University of Paraná (UFPR). His principal research interest comprises the development of new electrochemical sensor for the determination of inorganic and organic compounds in pharmaceutical, biological or environmental samples.