Novel Cytokine-independent Induction of Endothelial Adhesion Molecules Regulated by Platelet/Endothelial Cell Adhesion Molecule (CD31)

Marek Litwin,* Katherine Clark,* Leanne Noack,* Jill Furze,* Michael Berndt,§ Steven Albelda,§ Mathew Vadas,† and Jennifer Gamble*

*Division of Human Immunology, Hanson Centre for Cancer Research, Institute of Medical and Veterinary Science, Adelaide, South Australia, 5000 Australia; †Vascular Biology Laboratory, Baker Medical Research Institute, Prahran Victoria 3181; and §Director of Lung Research, Pulmonary and Critical Care Division, Department of Medicine, University of Pennsylvania Medical Center, Philadelphia, Pennsylvania 19104-4283

Abstract. Tumor necrosis factor–α, interleukin-1, and endotoxin stimulate the expression of vascular endothelial cell (EC) adhesion molecules. Here we describe a novel pathway of adhesion molecule induction that is independent of exogenous factors, but which is dependent on integrin signaling and cell–cell interactions. Cells plated onto gelatin, fibronectin, collagen or fibrinogen, or anti-integrin antibodies, expressed increased amounts of E-selectin, vascular cell adhesion molecule–1, and intercellular adhesion molecule–1. In contrast, ECs failed to express E-selectin when plated on poly-l-lysine or when plated on fibrinogen in the presence of attachment-inhibiting, cyclic Arg-Gly-Asp peptides. The duration and magnitude of adhesion molecule expression was dependent on EC density. Induction of E-selectin on ECs plated at confluent density was transient and returned to basal levels by 15 h after plating when only 7 ± 2% (n = 5) of cells were positive. In contrast, cells plated at low density displayed a 17-fold greater expression of E-selectin than did high density ECs with 57 ± 4% (n = 5) positive for E-selectin expression 15 h after plating, and significant expression still evident 72 h after plating. The confluency-dependent inhibition of expression of E-selectin was at least partly mediated through the cell junctional protein, platelet/endothelial cell adhesion molecule–1 (PECAM-1). Antibodies against PECAM-1, but not against VE-cadherin, increased E-selectin expression on confluent ECs. Culture of subconfluent ECs with PECAM-1–coated beads or with L cells transfected with PECAM-1 mutant, inhibited E-selectin expression. In contrast, untransfected L cells or L cells transfected with an adhesion-defective domain 2 deletion PECAM-1 mutant failed to regulate E-selectin expression. In an in vitro model of wounding the wound front displayed an increase in the number of E-selectin–expressing cells, and also an increase in the intensity of expression of E-selectin positive cells compared to the nonwounded monolayer. Thus we propose that the EC junction, and in particular, the junctional molecule PECAM-1, is a powerful regulator of endothelial adhesiveness.

The endothelial lining of the vascular system normally displays a nonactivated, nonadhesive phenotype. Stimulation with agents such as tumor necrosis factor–α (TNF-α), interleukin-1 (IL-1), or lipopolysaccharide (LPS) are known to induce the expression of proteins on the endothelial surface that mediate coagulation (Bevilacqua et al., 1985; Gamble et al., 1985; Poer et al., 1986b; Doherty et al., 1989), and leukocyte transendothelial migration (Furie et al., 1989; Moser et al., 1989). The endothelial antigens that are important for the adhesion of leukocytes are members of the selectin family, E- and P-selectin, and the immunoglobulin gene superfamily, vascular cell adhesion molecule–1 (VCAM-1) and intercellular adhesion molecule–1 (ICAM-1) (Carlos and Harlan, 1994; Litwin et al., 1995).

The induction of E-selectin expression on endothelial cells (ECs) in vitro after cytokine stimulation is transient and independent of the continued presence of the stimulant (Poer et al., 1986b). Previous studies have shown that
E-selectin mRNA and protein levels peak between 2 and 4 h, respectively, after treatment with an agonist, returning to near basal levels by 24 h (Bevilacqua et al., 1989; Read et al., 1994). VCAM-1 (Osborn et al., 1989) and ICAM-1 (Pober et al., 1986b) are maximal 6 and 12 h, respectively, after stimulation.

In contrast to the transiency of E-selectin and VCAM expression demonstrated by the in vitro data, these antigens have been detected on venular endothelium in chronic inflammatory lesions, such as the synovium in rheumatoid arthritis (Koch et al., 1991), and the skin in psoriasis (Pettelbauer et al., 1994). E-selectin expression is also detected on angiogenic vessels in human hemangiomata, a noninflammatory angiogenic disease (Kraling et al., 1996). Moreover, the architecture and anatomic localization of capillary loops influence the pattern of endothelial expression of E-selectin and VCAM-1, independently of the availability of cytokines (Pettelbauer et al., 1994). Thus it is likely that alternate control mechanisms exist to allow prolonged, locality-based expression of adhesion molecules on the endothelium. At least one of these alternate mechanisms may be flow, since increased shear stress has been shown to selectively modulate adhesion molecule expression, upregulating ICAM-1 but not E-selectin or VCAM-1 (Nagel et al., 1994).

Since sites of inflammation are often associated with morphological changes including cell retraction of the endothelium (Schumacher, 1973), we hypothesized that cell contacts may be important in the regulation of endothelial phenotype. We describe here the central role of the juxtional protein, platelet/endothelial cell adhesion molecule–1 (PECAM-1), through the formation of cell–cell interactions, in the maintenance of the functional integrity of the endothelial monolayer. Furthermore, we demonstrate a novel pathway for the induction of adhesion molecules on endothelial cells that is independent of exogenous addition of cytokines, but is related to integrin- and cell shape–associated signaling events.

Materials and Methods

Cell Culture

ECs were extracted from human umbilical veins by collagenase treatment, according to a modified method of Wall et al. (1978). Cells were grown in 25-cm², gelatin-coated Costar flasks (Costar Corp., Cambridge, MA), maintained with endothox-free medium 199 (Cytosystems, Sydney, Australia), 20% FCS (Commonwealth Serum Laboratories, Melbourne, Australia), 20 mM Hepes, 2 mM glutamine, 1 mM sodium pyruvate, 1% non-essential amino acids (Cytosystems), 0.225% sodium bicarbonate and antibiotics in the absence of exogenously added heparin or basic fibroblast growth factor, at 37°C in a humidified, 5% CO₂ (in air) atmosphere. After 2–5 d of culture, the cells were harvested by trypsin-EDTA treatment (Cytosystems) and replated at specified cell densities on 1.9-cm², gelatin- and fibronection-coated multiwell dishes (Nunc, Roskilde, Denmark) for flow cytometry experiments, or on fibronection-coated glass or Permanox® chamber slides (Labtek, Nunc, Roskilde, Denmark) for immunofluorescence and adhesion assays. The preparation of PECAM-1–transfected L cells has been reported previously (DeLisser et al., 1993). The cells were maintained in RPMI 1640 (GIBCO BRL, Gaithersburg, MD) 10% FCS with 0.5 mg/ml G418.

Antibodies

Mouse mAbs directed against E-selectin (49-IB11), VCAM-1 (51-10C9), PECAM-1 (51-9H6, 51-6F6, 55-3D2), VE-cadherin (55-7H1), integrin β₃, (61-2C4), colony-stimulating factor (CSF), common β chain (SD7, supplied by Q. Sun, Hanson Centre for Cancer Research [HCCR], Adelaide, Australia), and Keyhole Limpet Hemaeycin (25-1F11) were raised at the HCCR. The anti–PECAM-1 mAbs were functional in an assay of neutrophil transendothelial migration and anti–VE-cadherin antibody was functional in an assay of EC aggregation. For coating of wells with antibody, plates were coated with rabbit anti-mouse Ig (50 μg/ml) for 18 h, blocked with 1% BSA, and then anti-integrin or anti-CSF, chain–purified antibodies added for 1 h at 37°C. Wells were then washed and further blocked before addition of cells.

Flow Cytometry

Flow cytometric analysis of in situ endothelial monolayers was performed as previously described (Gamble et al., 1993). EC monolayers were blocked in 5% sheep serum, and then were stained with primary antibody for 30 min at 37°C, washed twice with RPMI 1640 containing 2.5% FCS, and then stained with FITC-conjugated, anti-mouse Ig (Fab2, DAF; Silenus Laboratories, Hawthorn, Australia) for 30 min at room temperature. Cells were washed twice, removed by trypsin-EDTA treatment, and then fixed in 1% formaldehyde, 0.02% azide, and 0.02% glucose. In experiments involving endothelial pretreatment with mAbs, E-selectin was detected using a single layer, FITC-conjugated, anti-E-selectin mAb (49-1B11). A minimum of 1,000 events per test was analyzed using an EPICS Profile II (Coulter Immunology, Hialeah, FL). Results of individual EC lines are expressed either as a plot of frequency versus log fluorescence, or as the mean fluorescence channel number, subtracting the accompanying value for the negative control Ig. When results from multiple EC lines have been pooled, the mean fluorescence intensity (MFI) represents n cell lines.

In cocultures of ECs and L cells prepared for flow cytometric analysis, the cells were stained with anti–VE-cadherin, detected with phycoerythrin (PE)-conjugated anti-mouse F(ab')2 (DDAPE; Silenus Laboratories, Hawthorn, Australia) and simultaneously stained with goat anti–E-selectin detected with FITC–conjugated, anti–goat antibody (Silenus Laboratories). L cells were negative for VE-cadherin, EC were 100% positive for VE-cadherin and were selected for analysis of E-selectin (FITC staining) using a second fluorescence detector. The flow cytometer was calibrated using single PE- or FITC-stained cells.

Immunofluorescence Confocal Microscopy

Confocal microscopy was performed on ECs cultured on fibronectin-coated glass or Permanox® chamber slides. A staining three layer method was used in winding assays and involved initial fixation in ice-cold methanol for 5 min and then acetone 1 min and washing in M199-containing 2.5% FCS. Cells were stained with saturating amounts of mAb for 30 min at 25°C. After two washes, cells were incubated with biotin-conjugated, affinity-purified anti-mouse Ig (Vector Labs, Burlingame, CA), washed twice, and incubated with avidin–FITC (Dako Corp., Carpinteria, CA). Slides were mounted using 2% propylgallate in glycerol as an anti-fade agent. Images were captured with a laser scanning confocal microscope (MRC600; Bio Rad Laboratories, Hercules, CA). Comparison images were subjected to equivalent amounts of contrast enhancement.

Neutrophil–Endothelial Adhesion

Peripheral blood from normal volunteers was sedimented on dextran, followed by density-gradient centrifugation on Lymphoprep (Nycomed, Oslo, Norway) at 450 g. Contaminating erythrocytes were then lysed by hypotonic 0.2% sodium chloride. Cells were resuspended in RPMI-1640 with 2.5% FCS and yielded a purity of >98%. 5 × 10⁵ neutrophils were added in 125 μl medium to human umbilical vein endothelial cells (HU-VECs), which had been plated 16 h earlier onto fibronectin-coated chamber slides at cobblestone and subconfluent densities. After 25 min at 37°C in a humidified 5% CO₂ in air atmosphere, the wells were washed three times removing unattached neutrophils. The slides were fixed in 0.5% glutaraldehyde, examined by confocal microscopy, and the number of neutrophils detected with FITC–conjugated, anti–VE-cadherin antibody was functional in an assay of EC aggregation. For coating of wells with antibody, plates were coated with rabbit anti-mouse Ig (50 μg/ml) for 18 h, blocked with 1% BSA, and then anti-integrin or anti-CSF, chain–purified antibodies added for 1 h at 37°C. Wells were then washed and further blocked before addition of cells.

Preparation of Protein-coupled Beads

Tosyl-activated paramagnetic beads (Dynabeads M-450; DYNAL A.S., Oslo, Norway) at 450 g. Contaminating erythrocytes were then lysed by hypotonic 0.2% sodium chloride. Cells were resuspended in RPMI-1640 with 2.5% FCS and yielded a purity of >98%. 5 × 10⁵ neutrophils were added in 125 μl medium to human umbilical vein endothelial cells (HU-VECs), which had been plated 16 h earlier onto fibronectin-coated chamber slides at cobblestone and subconfluent densities. After 25 min at 37°C in a humidified 5% CO₂ in air atmosphere, the wells were washed three times removing unattached neutrophils. The slides were fixed in 0.5% glutaraldehyde, examined by confocal microscopy, and the number of neutrophils detected with FITC–conjugated, anti–VE-cadherin antibody was functional in an assay of EC aggregation. For coating of wells with antibody, plates were coated with rabbit anti-mouse Ig (50 μg/ml) for 18 h, blocked with 1% BSA, and then anti-integrin or anti-CSF, chain–purified antibodies added for 1 h at 37°C. Wells were then washed and further blocked before addition of cells.

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Oslo, Norway) were coated with purified platelet PECAM-1 as previously described (Plopper and Ingber, 1993). Essentially 99% of beads were coated with PECAM-1 as assessed by flow cytometry using polyclonal anti-PECAM-1 antibody staining.

**Endotoxin Assay**

A quantitative, photometric assay (Coatest; Kabi Diagnostica, Stockholm, Sweden) based upon activation of a proenzyme in limulus amoebocyte lysate was used, which detected endotoxin at 0.1–1.2 EU/ml.

**Cytokines and Cytokine Antagonists**

TNF-α (lot S9010AX; sp act 6.27 × 10^2 U/mg), TGF-β (lot 8987-53), and a monoclonal anti-TNF were gifts from Genentech, Inc. (South San Francisco, CA) IL-1β (10^6 thymocyte mitogenesis U/mg) was kindly supplied by Immunix (Seattle, WA). IL-1ra was a gift from Syenrogen (Boulder, CO). All cytokines contained <3 U/ml of LPS.

**Reagents and Peptides**

Polymyxin B sulfate (Sigma Chemical Co., St. Louis, MO) was used at 10 µg/ml. When added at plating, it effectively abolished induction of E-selectin by LPS on ECs. Soluble PECAM-1 protein was purified from platelet and was used at 0.01–100 µg/ml. Cyclic RGD and RAD peptides (EMD66203, 67679, 69601) were kindly supplied by A. Jonczyk from Merck KGaA (Darmstadt, Germany). These peptides were identical to those used by Brooks et al. (1994) in inhibiting αβ₂-dependent angiogenesis.

**Enumeration of EC Contacts**

Multiple photomicrographs of low power, phase contrast fields (see Fig. 1) were obtained of ECs plated at 0.25 and 10^5 cells per cm^2. The number of cell contacts made with adjoining ECs were counted for 10 ECs per field.

**Wounding EC Monolayers**

ECs plated at cobblestone density on fibronectin-coated chamber slides were wounded by scraping with the tip of a 1,000 µl pipette. The wells were washed three times with medium. All wounds consisted of a clearly demarcated cross in the center of a monolayer and healed as an advancing front of elongated, flattened cells. At specified times after wounding, cells were stained for immunofluorescence confocal microscopy as described above. Phase contrast microscopy with ×100 magnification was used to select fields at wound fronts and on areas of cellular monolayer, at least two fields were removed from a wound. Selected fields were then examined with the laser confocal microscope at ×200 magnification and the images stored. Images were retrieved in SETCOL format (COMOS 7.0; Bio Rad Laboratories, Hercules, CA), which displays fluorescence intensity on a color scale (green being minimal, red maximal), and were then examined for the number of ECs positive and negative for E-selectin per ×200 field. An EC was determined to be positive if any part of its surface, >5 mm in diam was red. At least 1,700 ECs were counted at wounds or monolayers per well and positivity was expressed as a percentage of all cells counted. The proportion of ECs expressing E-selectin in the top 50% range of expression intensity was assessed by analysis of pixel intensity in areas of monolayer and wound front and calculated using the histogram format.

**Statistics**

The statistical significance of results was assessed using the two-tailed Student’s t test with either paired or unpaired groups of data as indicated. The frequency histograms of neutrophil adherence to ECs were compared by the Kolmogorov-Smirnov test and the effects of PECAM-1–coated beads on E-selectin expression by the analysis of variance (ANOVA) test.

**Results**

**Confluency-dependent Expression of Endothelial Adhesion Molecules**

HUVECs were seeded onto gelatin-coated tissue culture plates at varying numbers (Fig. 1). 15 h after plating, the EC surface expression of E-selectin was measured by flow cytometry. Confluent, cobblestone cultures as seen in Fig. 1 a, showed negligible E-selectin expression (Fig. 2 a). This is in agreement with published work demonstrating a lack of E-selectin on unstimulated ECs (Pober et al., 1986b; Bevilacqua et al., 1989). However, subconfluent ECs (Fig. 1 b) displayed substantial expression of E-selectin (Fig. 2 a). A comparison between subconfluent and confluent density cells using five separate EC lines showed a 17-fold greater induction in subconfluent ECs, (6.4 ± 1.0 vs. 0.37 ± 0.19 mean fluorescence intensity units ± SEM, P = 0.0003, unpaired t test). In these five experiments, 57 ± 4% of ECs at subconfluent density were positive for E-selectin, as opposed to 7 ± 2% of cells in a cobblestone monolayer. The confluency-dependent expression of E-selectin was evident over a range of EC densities (Fig. 2 b) and was seen on HUVECs extracted from their original monolayer culture by treatment with trypsin-EDTA or EDTA alone (data not shown).

**Time Course of E-selectin Expression**

The expression of E-selectin was measured on ECs at varying times after plating. Fig. 3 shows that cells plated at high and low densities both displayed significant levels of E-selectin early after plating that is within 4 to 12 h. However, expression of E-selectin on high density ECs was fourfold less at its maximum 8 h after plating, was transient and returned to basal levels by 24 h. In contrast, E-selectin expression on ECs plated at low density peaked at 12 h after plating and was still evident at 32 h. ECs at very sparse densities, such that single cells were maintained over the course of the experiment, displayed a persistent and significant expression of E-selectin even 72 h after plating (data not shown). There was considerable variation in the absolute levels of E-selectin between different EC preparations, the reason for which is not known. However, the fold induction calculated between high and low density cells was relatively consistent (21.1 ± 3.6%, n = 30) when measured 16–20 h after plating.

VCAM-1 and ICAM-1 (as well as E-selectin) demonstrated EC density–dependent expression, but the expression of PECAM-1 (or CD31), a non-inducible adhesion protein was not altered (Fig. 4). Increased expression of VCAM-1, ICAM-1, and E-selectin was independent of cell size, as identical forward scatter gates of high and low
density cells were always used in the FACS® analysis. Furthermore, the change in adhesion molecule expression was observed, whether the cells were stained in situ before detachment, or after extraction from matrix (data not shown).

**E-selectin Expression by Subconfluent ECs Is Independent of Cytokines and Supports Neutrophil Adhesion**

The time course of expression seen in Fig. 2 suggested two phases in the regulation of E-selectin expression: an induction phase, and a maintenance phase. Induction of the expression of adhesion molecules by subconfluent ECs occurred in the absence of exogenous cytokines. As shown in Fig. 5a, IL-1 receptor antagonist (IL-1ra) or blocking anti–TNF-α antibody potently and specifically inhibited IL-1 or TNF-α-mediated induction of E-selectin, respectively, but these agents were ineffective on the induction of E-selectin by subconfluent ECs (Fig. 5b). Furthermore, conditioned medium taken from subconfluent ECs or from cells multiply wounded such that the majority of cells were undergoing migration, did not induce E-selectin expression on confluent density ECs (Table I). Thus a role for endogenous endothelial cytokine production, or the release of some other stimulant from the EC, appeared unlikely.

Further, the induction of E-selectin is not related to cell proliferation. Rapamycin, an inhibitor of the G1-S transition (Kato et al., 1994) showed no effects on E-selectin induction in subconfluent ECs (data not shown) although TGF-β, another inhibitor of cell proliferation (Heimark et al., 1986), which we have shown previously to inhibit cytokine-induced E-selectin expression (Gamble et al., 1993) did inhibit the cytokine-independent induction of E-selectin (data not shown). Moreover, E-selectin induction was independent of cell cycling since no difference in the level of E-selectin expression was seen in cells in G0/G1 with respect to S/G2/M at either cell density, both up to 20 h and even at 72 h after EC plating (data not shown).

Low doses of TNF-α were able to increase E-selectin expression on both low and high density ECs (Fig. 6a) suggesting that the cytokine and non-cytokine pathways of induction are at least additive. Although the level of E-selectin expression induced by the cytokine-independent mechanism is less than that induced by TNF or IL-1 (Figs. 5 and 6), it is however, functionally relevant. Assessment of the number of neutrophils adherent per EC showed that there

**Figure 3.** Time course of E-selectin expression after EC plating. ECs were plated at confluent (∆, 10⁵ cells per cm²) and subconfluent (■, 0.25 × 10⁵ cells per cm²) densities. The expression of E-selectin was assayed by flow cytometry at specified times after plating. The MFI (± SEM) of three to five cell lines is shown but values at 2, 32, and 72 h after plating are singlicate. Asterisks denote values significantly different between the two cell densities (P < 0.03) by paired t test.

**Figure 4.** Expression of E-selectin, VCAM-1, and ICAM-1 are confluency dependent. ECs plated at subconfluent density (stripes, 0.25 × 10⁵ cells per cm²) and confluent density (solid, 10⁵ cells per cm²) were stained 20 h later for a, E-selectin, b, VCAM-1, c, ICAM-1, or d, PECAM-1. Expression is shown as the MFI of 23, 11, 7, and 7 EC lines, respectively. Error bars represent the SEM. Asterisks denote values significantly different from cells plated at cobblestone density (P < 0.03) by paired t test.

**Figure 5.** The induction of E-selectin on subconfluent ECs is not mediated through TNF-α or IL-1. (a) ECs plated at subconfluent density (0.25 × 10⁵ cells per cm²) were stimulated with 10 U/ml TNF-α (TNF) or 1 ng/ml IL-1β (IL-1) in the presence of anti–TNF-α (TNF + anti-TNF) (1:1,000), or IL-1ra (IL-1 + IL-1ra) (100 ng/ml), respectively. Inhibitors and agonists were added immediately after EC plating and E-selectin expression was assessed 12 h later. (b) ECs were plated at subconfluent density (stripes, 0.25 × 10⁵ cells per cm²) and confluent density (solid, 10⁵ cells per cm²). Anti–TNF-α or IL-1ra were added at plating. E-selectin expression was assessed 12 h later. Results shown are of one representative experiment of four performed.
was an increase in the number of neutrophils attached per EC in cells plated at low versus high density EC (Fig. 6b). In three experiments, the percentage of EC supporting the adhesion of two or more neutrophils was 24.3\% ± 14\% for low density EC and 1.4\% ± 1.0\% for high density EC (mean ± SEM; P = 0.02).

**E-selectin Induction Is Dependent on Integrin Engagement**

ECs plated on collagen I, gelatin, fibronectin, fibrinogen, and laminin in BSA-containing media resulted in an up-regulation of E-selectin when measured 6 h after plating (Fig. 7a). The level of induction on these matrices in the absence of FCS in the media was never as high as in the presence of FCS. E-selectin expression was not induced on ECs plated on poly-L-lysine (Fig. 7a) although they responded to TNF stimulation, suggesting that the poly-L-lysine was not toxic (data not shown). The induction was seen whether cells were plated at subconfluent density (Fig. 7) or at high density (data not shown). Cells plated on anti-β1 integrin-coated surfaces, attached, spread, and induced E-selectin. In contrast, cells plated on an antibody to the common β chain (β,) of GM-CSF, IL-3, and IL-5 receptor, which is also expressed on EC (Korpelainen et al., 1993), attached but failed to spread and also did not result in E-selectin induction (data not shown). Cells plated on fibrinogen in the presence of cyclic RGD (Arg-Gly-Asp) or RAD (Arg-Ala-Asp) peptides (final concentration 10 \( \mu \)M) in 1\% BSA-containing media. E-selectin expression was measured 6 h after plating and is given as the mean ± SEM of a representative experiment where each group was performed in triplicate. Asterisks denote values significantly different from control peptide (P < 0.01).

**PECAM-1 Regulates the Maintenance of Endothelial Adhesion Molecules**

Although E-selectin is induced on all cells, only subconfluent EC maintain this expression. The chief difference between these density phenotypes lies in the number of cell-cell contacts and their establishment rate. ECs plated at confluent density (10^5 cells per cm^2) were rapidly sur-
rounded by other cells. Within 1 h they had established 5 ± 1 contacts with adjoining cells (mean ± SEM; n = 3). This number remained constant over the following 24 h. By contrast, subconfluent ECs (0.25 × 10^6 cells per cm^2) had an average of only 1 ± 1 contact (n = 3) at 1 h after plating and continued to form new associations such that 24 h later they had 4 ± 1 contacts (n = 6).

Two molecules known to be concentrated in cell–cell contacts and implicated in establishment of some of the junctional properties of endothelial cell monolayers are PECAM-1 (Albelda et al., 1991; DeLisser et al., 1994) and VE-cadherin (Lampugnani et al., 1992; Ayalon et al., 1994). To determine whether PECAM-1 was involved in the cytokine-independent regulation of E-selectin, three independent methods were used. Firstly, confluent density EC monolayers were exposed to functional antibodies directed to PECAM-1 or VE-cadherin. Monoclonal anti-VE-cadherin antibody (antibody 7H1) had no effect. Polyclonal anti-PECAM-1 resulted in a twofold increase in E-selectin expression (Fig. 8, a and b). Addition of both anti–PECAM-1 and anti–VE-cadherin antibodies produced no further increase than with anti–PECAM-1 antibody alone (data not shown). Although two mAb directed to domain one of PECAM-1 consistently and significantly enhanced E-selectin expression their activity was always less than that seen with the polyvalent anti–PECAM-1 antibody (Fig. 8 b) suggesting the involvement of multiple domains. 55-3D2, an mAb directed to domain two-thirds of PECAM-1 was without function in these assays although it inhibits neutrophil transendothelial cell migration (Yan et al., 1995). The anti–PECAM-1 antibody effect was dose dependent (maximal efficacy at 5 μg/ml) and not due to contaminating endotoxin as polymyxin B did not abrogate the enhancement, the antibodies did not

Figure 8. Anti–PECAM-1 antibody upregulates E-selectin on ECs plated at cobblestone density. (a) ECs at cobblestone density (10^6 cells per cm^2) were treated with Ig-purified rabbit polyclonal antibody to PECAM-1 (5 μg/ml) or a control nonimmune rabbit Ig at the time of plating. E-selectin expression was assessed by flow cytometry 12 h later. The MFI (± SEM) of two to six experiments is given where untreated confluent density ECs have been normalized to 1.0. Subconfluent ECs showed a 5.1 ± 1.4-fold increase in E-selectin expression relative to confluent EC. Asterisks denote a significant difference (P < 0.05) compared with control rabbit Ig by unpaired t test. (b) ECs at confluent density were treated with 5 μg/ml Ig-purified mAbs to PECAM-1 or VE-cadherin at the time of plating or polyvalent anti–PECAM-1 antibody. E-selectin expression was measured 12 h later. The MFI (± SEM) of three experiments is given where untreated confluent density ECs have been normalized to 1.0. Subconfluent ECs showed a 4.0 ± 0.7-fold relative increase in E-selectin expression relative to confluent EC. Asterisks denote a significant difference (P < 0.05) compared with cobblestone EC group.
have detectable endotoxin and boiling the antibody abolished its potency (Fig. 8a). VCAM-1 was also upregulated on cobblestone ECs by polyclonal anti–PECAM-1 antibody by 1.7 ± 0.19-fold (mean ± SEM, n = 3, P = 0.03, paired t test).

The second approach used platelet purified PECAM-1 immobilized on beads. E-selectin expression on EC plated at low density in the presence of PECAM-1–coated beads was inhibited when measured 18 h after plating (inhibition was 61 ± 20%; n = 3). BSA-coated beads had no effect (Fig. 9). Interestingly, neither purified, soluble PECAM-1, nor its immobilization on plastic was able to regulate E-selectin expression (data not shown) suggesting that valency, concentration, or microenvironment problems may be operating.

The third approach used PECAM-1 transfectants. ECs were plated at sub confluent density in the presence of L cells expressing full-length PECAM-1, PECAM-1 lacking domain 2, or lacking the cytoplasmic tail. Both the full length and the truncation of the cytoplasmic tail inhibited E-selectin expression, whereas the mutant lacking domain 2 had no effect (Fig. 10a). Untransfected L cells (Fig. 10b) or L cells expressing the L1 adhesion glycoprotein (a member of the Ig superfamily expressed by neural cells and lymphocytes (Hubbe et al., 1993) failed to regulate E-selectin expression.

E-selectin Is Upregulated at Endothelial Wound Edges Whereas the Cell Junctional Molecule PECAM-1 Is Diminished

An in vitro wound assay was established as an in vivo correlate of EC migration (Schimmenti et al., 1992; Taylor and Alexander, 1993). ECs were plated and allowed to come to confluence. At variable times thereafter wounds were made and the cells stained 11 and 27 h later for E-selectin and PECAM-1. As seen in Fig. 11b, the cells at the wound front displayed a spread, motile, morphology, and had advanced beyond the wound edge. These migrating cells had less PECAM-1 staining at the cell-to-cell borders. Significantly more ECs at the wound front and immediately behind the front expressed E-selectin in comparison...
rate experiments at 16 h after wounding; 6
sion was 1.7
ing E-selectin in the top 50% range of intensity of expres-
expressing cells (Table II). The proportion of ECs express-
substantiated by direct counts of the number of E-selectin
positive cells at either the wound front or in the monolayer in a single experiment (n = 1)
is shown. Also shown is the mean increase in the number of positive cells at the
wound front compared to the monolayer for three experiments where all experiments
showed the same trend.

to the nonwounded areas (Fig. 11, c and d), and this was
substantiated by direct counts of the number of E-selectin
expressing cells (Table II). The proportion of ECs express-
ing E-selectin in the top 50% range of intensity of expres-
sion was 1.7 ± 0.7% in the cell monolayer compared to 16
± 5.0% at the wound front (mean ± SEM of three sepa-
rate experiments at 16 h after wounding; P = 0.05).

Discussion

In this article, we have demonstrated a novel mechanism
for the induction and expression of adhesion molecules on
ECs. The induction is initiated through an integrin-depen-
dent process and is associated with cell spreading. The in-
tensity and duration of expression of the adhesion mole-
cules is inversely proportional to cell density, and is
mediated at least in part through the junctional protein,
PECAM-1. This phenomenon does not involve the known
cytokines and differs from the cytokine-dependent path-
ways in a number of ways: (a) it is dependent on integrin
engagement and cell shape; (b) it is regulated by cell junc-
tions; and (c) it can result in prolonged cell surface expres-
sion of adhesion molecules (Table III).

Since the phenomena described in this paper represent
a fundamentally new mechanism underlying important
pathological processes, we suggest nomenclature where
type I induction refers to the classic cytokine-dependent
process, and type II induction refers to the new cytokine-
independent pathway. Although the level of E-selectin
induced in type II conditions is less than that seen for maxi-
mal doses of cytokine-induced E-selectin, it is functional
as evidenced by increased neutrophil attachment, and is at
least additive with low doses of TNF. Such situations of
low cytokine concentrations may be relevant during early
phases of inflammatory lesions, suggesting the potential
importance of both these induction processes in determin-
ing endothelial phenotype.

Type II induction of E-selectin described herein is inde-
pendent of cell size, cell proliferation, and cell cycle, but is
initially integrin mediated and seen in cells plated at both
high and low density. Cells plated onto ligands that facili-
tated integrin-mediated attachment induce E-selectin ex-
pression (Fig. 7). However if the initial attachment is
blocked, as with the addition of cyclic RGD peptides to
cells on fibrinogen, E-selectin induction is inhibited. Bind-

| Time after | E-selectin–positive ECs | Mean increase in number of |
| wounding   | (n = 3) | E-selectin–positive ECs |
|           | %     | (n = 3) |
| Monolayer  | Wound front |
| 11 h       | 4.1    | 13.5    | 2.2 ± 0.5 |
| 27 h       | 4.3    | 10.4    | 2.4 ± 0.5 |

Confluent EC monolayers were wounded by scraping with a pipette tip at either 13 or
29 h after plating. The cells were stained for E-selectin 27 or 11 h thereafter, respec-
tively (i.e., 40 h after plating). An average of 2,600 cells was counted in the confluent
monolayer, and 1,700 at the wound edge for each experiment, and the E-selectin–pos-
itive cells was calculated as described in Materials and Methods. The percent of posi-
tive cells at either the wound front or in the monolayer in a single experiment (n = 1)
is shown. Also shown is the mean increase in the number of positive cells at the
wound front compared to the monolayer for three experiments where all experiments
showed the same trend.

The intensity and duration of induction of type II re-
sponses was related to the density of cell plating. Cells
plated at confluent density expressed less E-selectin when
measured 8–48 h after plating, compared to cells plated at
subconfluent density (Fig. 3). Morphologically, cells plated
at high density form rapid cell–cell interactions in contrast
to low density cells. Thus, junctional control of EC adhe-
sion molecule expression was considered. Two molecules
have been described that are known to be important in en-
thodelial junctional integrity (Albelda et al., 1991; Lam-
pugnani et al., 1992; Ayalon et al., 1994; DeLisser et al.,
1994); PECAM-1, a transmembrane glycoprotein belong-
ing to the Ig gene superfamily containing six extracellular,
Ig-like domains; and VE-cadherin, an endothelium-spe-
cific member of the cadherin family of cell junctional mol-
ecules. The involvement of PECAM-1 in the control of
E-selectin expression was shown by three independent
methods. Firstly, anti–PECAM-1 but not anti–VE-cad-
herin antibodies induced E-selectin expression on conflu-
ent ECs (Fig. 8). Although the increases achieved with the
antibodies were not to the level seen on low density cells,
they were consistent. This may suggest that molecules
other than PECAM-1 could exert secondary events that
influence the level of expression of E-selectin. Secondly,
PECAM-coated beads, when added at plating, inhibited
the level of E-selectin expression on low density cells com-
pared to BSA-coated beads (Fig. 9). Thirdly, transfectants
expressing full-length PECAM-1, or those expressing the
extracellular domains but lacking the cytoplasmic domain,
inhibited E-selectin expression on subconfluent ECs (Fig.
10). In contrast, L cells expressing the domain 2 deletion
mutant failed to effect E-selectin expression, suggesting a
critical involvement of domain 2 in this regulation. These
results are in agreement with other studies showing the
importance of domain 2 in cell aggregation (Sun et al.,
1996). Together with our antibody studies showing that
antibodies to domain 1 increased the level of E-selectin
expression, suggesting a secondary effect that may be
regulated by antibodies to domain 1, we suggest that both
domains 1 and 2 of PECAM-1 are critical in regula-
ing of RGD to cells per se is insufficient to inhibit E-selec-
tin induction since RGD peptides have no effect on cells
plated onto collagen.

The intensity and duration of induction of type II re-
sponses was related to the density of cell plating. Cells
plated at confluent density expressed less E-selectin when
measured 8–48 h after plating, compared to cells plated at
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antibodies to domain 1 increased the level of E-selectin
expression, suggesting a secondary effect that may be
regulated by antibodies to domain 1, we suggest that both
domains 1 and 2 of PECAM-1 are critical in regula-
ing of E-selectin expression.

Cell–cell interactions are known to regulate a number of
events, including cell proliferation (Gradl et al., 1995), re-
lease of bFGF from astrocytes (Murphy et al., 1988), re-
sponsiveness of fibroblasts to TGF-β (Paulsson et al.,
1988), and the regulation of intracellular pH (Galkina et
al., 1995). The mechanisms that underlie such regulation
however have not been elucidated, although autocrine re-
lease of growth inhibitory factors (Antonelli-Orlidge et al.,
1989; Gradl et al., 1995) and gap junction changes (Chen et

Table II. E-selectin Expression by ECs at Wound Edges

| Time after wounding | E-selectin–positive ECs | Mean increase in number of E-selectin–positive ECs (n = 3) |
|---------------------|-------------------------|----------------------------------------------------------|
| 11 h                | 4.1                     | 13.5                                                     | 2.2 ± 0.5 |
| 27 h                | 4.3                     | 10.4                                                     | 2.4 ± 0.5 |

Table III. Mechanisms Controlling Expression of Endothelial Adhesion Molecules

| Type I                  | Type II                  |
|-------------------------|--------------------------|
| Induction               | Cytokine, endotoxin      |
| Duration                | Transient                |
| Peak                    | 4–6 h                   |
| Regulation              | Prolonged                |
|                        |                          |

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al., 1995) have been proposed. As described here, a cell surface junctional protein, PECAM-1, plays a central role in the density-dependent regulation of endothelial E-selectin expression. The possibility of a signaling role for PECAM-1 has been raised previously in a number of different systems. These include activation of integrins on T cells (Tanaka et al., 1992), natural killer cells (Berman et al., 1996), monocytes and neutrophils (Berman and Muller, 1995), and on CD34+ hematopoietic progenitor cells (Leavelsley et al., 1994), inhibition of EC proliferation (Fawcett et al., 1995), and platelet aggregation (Newman et al., 1992). The association of the tyrosine phosphatase-SHP2 with aggregated platelets (demonstrated recently by Jackson et al. [1997]) has put credence to a signaling pathway associated with PECAM-1 itself. Of interest to our study here is the recent observation by Lu et al. (1996) that integrin engagement and cell spreading results in PECAM-1 dephosphorylation in EC. Thus, the likely cross-talk between integrin and PECAM-1 is further strengthened.

Our in vitro model of cell wounding displays similar features to that described in the density-dependent regulation of adhesion molecules. Firstly, the induction of E-selectin expression is restricted to the migrating front and is associated with a change in cell shape and PECAM-1 redistribution away from the cell junction. Secondly, the E-selectin expression on these migrating cells is maintained at least up to 30 h after wounding. Thus we would suggest that this model may reflect identical signaling pathways as those operating in our plating experiments, and may therefore be appropriate as a model for in vivo endothelial regulation at wound sites.

Pathological tissue inflammation is characterized by increased and chronic expression of endothelial adhesion molecules (Koch et al., 1991; Petzeltbauer et al., 1994; Kraling et al., 1996), changes in endothelial morphology and angiogenesis (Fitzgerald et al., 1991). These morphological changes include the formation of high endothelial venules (Fremont et al., 1983) and cell retraction with associated intercellular gaps (Nagel et al., 1994). In balloon angioplasty, expression of adhesion molecules at the wound front associated with morphological changes has been observed in contrast to the lack of expression in the area behind the wound edge (Tanaka et al., 1993). More recently, a role for E-selectin in angiogenesis has been postulated (Nguyen et al., 1993; Koch et al., 1995; Kraling et al., 1996). The results reported here, demonstrate: (a) a cyto-kine-independent mechanism of adhesion molecule expression, and (b) that cell–cell interactions influence the duration and magnitude of this expression, may explain some of these in vivo observations. Furthermore, the observation that restoration of PECAM-1 interactions can downregulate adhesion molecule expression on ECs offers the promise that manipulation of EC junctional molecules may permit the development of novel therapeutics.

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