IgA Nephropathy After SARS-CoV-2 Vaccination

Matthew Abramson,* Samuel Mon-Wei Yu,* Kirk N. Campbell, Miriam Chung, and Fadi Salem

Here we present the first case of newly diagnosed IgA nephropathy (IgAN) after a SARS-CoV-2 vaccination. A 30-year-old man with no known past medical history presented with gross hematuria and subnephrotic proteinuria 24 hours after the second dose of the mRNA-1273 SARS-CoV-2 vaccine. A kidney biopsy showed IgAN. He was started on an angiotensin receptor blocker, resulting in proteinuria reduction. Similar to natural infection of SARS-CoV-2, persons who receive 2 mRNA-based vaccines demonstrate robust antibodies against the receptor-binding domain (RBD) of the S1 protein. Given the uniqueness of glycosylation of RBD and potent stimulation of immune response from mRNA-based vaccine compared to other vaccines, we hypothesize that our patient developed de novo antibodies, leading to IgA-containing immune-complex deposits. This case highlights the urgency of understanding the immunological responses to novel mRNA-based SARS-CoV-2 vaccines in more diverse populations. Despite the lack of clear causality, nephrologists should be alerted if any new-onset hematuria or proteinuria is observed.

INTRODUCTION

While IgA nephropathy (IgAN) is typically triggered by upper respiratory infections, relapse of IgAN has been reported in patients receiving vaccinations. More recently, there have been 3 case reports of patients who presented with IgAN relapse following SARS-CoV-2 mRNA vaccination.1-3 Here, we present the first case of newly diagnosed IgAN after receiving the Moderna SARS-CoV-2 mRNA vaccine.

CASE REPORT

A 30-year-old man of Western European and South American ancestry presented with new-onset hematuria and proteinuria. He had no known past medical history and had never been tested for COVID-19 infection. He did not report any known COVID-19 exposures and had not had any flu-like illness throughout the COVID-19 pandemic. He reported no family history of kidney disease, including IgAN. He received the first dose of mRNA-1273 SARS-CoV-2 vaccine, manufactured by Moderna, and remained asymptomatic during the 28-day interval between doses. However, 1 day after receiving the second vaccine, he developed fevers, chills, headache, and brown-colored urine. He presented to his primary care physician, where urinalysis showed 4+ protein, >30 red blood cells per high-power field (ref: 0-3), 11-30 white blood cells per high-power field (ref: 0-4), and 3+ blood (ref: negative). Creatinine was 1.02 mg/dL (Ref: 0.76-1.27 mg/dL), and estimated glomerular filtration rate was 98 cc/min/1.73 m². Gross hematuria resolved after 48 hours, but a repeat urinalysis 10 days later showed persistent microscopic hematuria and proteinuria. He was sent to nephrology for consultation. Physical examination was normal, and blood pressure was 125/73 mm Hg. Pertinent negatives included lack of lower extremity edema, rash, lymphadenopathy, and throat erythema. Random urine protein-creatinine ratio was 0.8 g/g (ref: 0.0-0.2 g/g), estimating 24-hour urine protein excretion of 800 mg. Urinalysis after centrifugation revealed numerous acanthocytes, but no red blood cell casts. Kidney ultrasound showed mildly increased echogenicity of normal size and cortical thickness. Additional serological work-up for glomerulonephritis was negative, including hepatitis B and C, HIV, and antinuclear and antineutrophil cytoplasmic antibodies. Erythrocyte sedimentation rate and C-reactive protein were normal. Complements C3 (105, ref: 82-167 mg/dL) and C4 (19, ref: 12-38 mg/dL) were normal. Creatinine phosphokinase was 254 U/L (ref: 49-439 U/L). Immunoglobulin A levels were elevated at 860 mg/dL (ref: 90-386 mg/dL). Given the unclear diagnosis, a kidney biopsy was performed. Light microscopy revealed 9 glomeruli with mild mesangial expansion and hypercellularity without endocapillary hypercellularity (Fig 1A), 1 of which showed segmental adhesion of a capillary loop to the Bowman capsule. Immunofluorescence revealed 3+ diffuse granular mesangial staining for IgA (Fig 1B). Staining was weakly positive for C3 and negative for IgG and other immunoglobulins/complement antibodies. Ultrastructural examination revealed scattered immune-type electron-dense deposits in the mesangium and mild podocyte foot process effacement (Fig 1C). Pathologic features were consistent with IgAN with Oxford MEST-C classification as M1-E0-S1-T0-C0, and his risk of a 50% decline in estimated glomerular filtration rate or progression to kidney failure within 5 years was approximately 3.9%, as per a recent risk prediction model by the International IgA Nephropathy Network. He was started on losartan 25 mg daily, which was well tolerated. After 6 weeks of therapy, urine protein-creatinine ratio
improved to 0.43 g/g and creatinine remained stable at 1.03 mg/dL.

**DISCUSSION**

To our knowledge, this is the first reported case of newly diagnosed IgAN in a kidney biopsy related to a COVID-19 vaccine, after a very recent report of exacerbated IgAN in 2 patients who received the second dose of mRNA-1273 SARS-CoV-2 vaccine. Although correlation does not inherently imply causation, the timing of symptom onset shortly after the vaccine should be considered as the inciting event.

IgAN is an immune-complex disease characterized by mesangial IgA deposition with or without concurrent IgG and C3 deposits. Despite IgAN being the most common primary glomerulonephritis worldwide, its underlying pathogenesis was unclear until the recognition of circulating, poorly galactosylated IgA alone is not sufficient to trigger IgAN. Immune complexes formed by pathogenic IgA and glycan-specific IgG or IgA targeted at these aberrantly galactosylated areas are usually required, followed by impaired liver removal and an increased affinity for mesangial cells. Thus, IgAN is proposed as a multi-hit disease. It may involve genetic predispositions (gene variants encoding galactosylation) and subsequent triggering events such as infection, environmental exposure, or dietary influences that lead to anti-glycan IgG/IgA production. Although the occurrence of postvaccination IgAN remains scarce, the interplay between vaccination and IgA vasculitis/Henoch–Schönlein purpura has been postulated. Intramuscular inactivated influenza vaccine elicits hyper-responsiveness in a cohort of pre-existing IgAN patients, with excessive productions of IgA monomers (but not polymers). Recurrence of IgAN following influenza vaccine has also been reported in kidney transplant recipients.

Our patient completed 2 doses of mRNA-1273 SARS-CoV-2 vaccine, a lipid nanoparticle–encapsulated mRNA.
vaccine.\textsuperscript{13} Once injected, mRNA encoding prefusion-stabilized spike glycoprotein is translated by host cells, and major histocompatibility complex molecules subsequently present viral protein segments to elicit immune responses with antibody production, including IgG/A/M. Importantly, the receptor-binding domain (RBD) of the S1 subunit is the immunodominant target of neutralizing antibodies in both infected patients\textsuperscript{14} and vaccinated persons.\textsuperscript{15} In contrast to other S protein areas, RBD has limited glycosylation sites, likely to allow stable and persistent entry of SARS-CoV-2 into host cells. This unique property of RBD, on the other hand, serves as the ideal epitope for vaccine development.\textsuperscript{16} Since vaccination does not elicit mucosal immune responses, one possible explanation for the IgAN seen in our patient is the production of antiglycan antibodies that cross-react with pre-existing under-galactosylated IgA1. In addition, mRNA-based vaccine may induce stronger T follicular helper and subsequent B-cell responses in the germinal center, which may lead to more robust antibody productions. Another possibility might be an increase of pathogenic IgA production similar to the influenza vaccine, given his elevated IgA level. A recent preprint article also suggested robust spike-specific IgA responses after receiving mRNA vaccines in healthy individuals.\textsuperscript{17} Nevertheless, the diagnostic value of circulating total IgA level in IgAN remains very limited. Additionally, there is the possibility that the patient had underlying subclinical “lanthanic” IgAN that was exacerbated by the vaccine.

According to Centers for Disease Control and Prevention data, there was no reported hematuria/proteinuria except for 1 patient who developed acute kidney injury owing to obstructive nephrolithiasis, resulting in death.\textsuperscript{18} It is beyond the scope of the discussion on the causality of IgAN in our patient. Still, emerging data suggest the importance of analyzing pre-existing antibodies to self-carbohydrates in either infected or vaccinated persons.\textsuperscript{19} Another intranasal vaccine candidate against SARS-CoV-2 is currently under investigation,\textsuperscript{20} which might directly perturb dysregulated mucosal immune system in genetically predisposed persons. Of note, local mucosal responses to natural SARS-CoV-2 in the nasopharynx are shown to be dominated by early IgA-neutralizing antibodies compared to IgG and IgM and were associated with a peripheral expansion of IgA-secreting plasmablasts.\textsuperscript{21} It is therefore essential to monitor any de novo IgAN or exacerbations of pre-existing IgAN following intranasal vaccinations. Lastly, in the first published mRNA-1273 trial data, there were only 4.6% Asians among all participants. Since IgAN is more common in the Asian population, larger clinical trial data, particularly with more diverse participants, are urgently needed to fully assess the possible correlation between IgAN and vaccination against SARS-CoV-2. While providers and patients should be aware of this adverse effect, it should be emphasized that this is likely a rare occurrence and should not deter others from seeking SARS-CoV-2 vaccination. Furthermore, SARS-CoV-2 infection confers a much higher risk than IgAN, and our finding is unlikely to change the risk-benefit ratio of vaccination.

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**REFERENCES**

1. Gul Rahim SE, Lin J, Wang JC. A case of gross hematuria and IgA nephropathy flare-up following SARS-CoV-2 vaccination. *Kidney Int*. 2021;100(1):238.

2. Negrea L, Rovin BH. Gross hematuria following SARS-CoV-2 vaccination in patients with IgA nephropathy. *Kidney Int*. 2021;100(2):466-468.

3. Trimarchi H, Barratt J, Cattrant DC, et al. Oxford Classification of IgA nephropathy 2016: an update from the IgA Nephropathy Classification Working Group. *Kidney Int*. 2017;91(5):1014-1021.

4. Barbour SJ, Coppo R, Zhang H, et al. Evaluating a new international risk-prediction tool in IgA nephropathy. *JAMA Intern Med*. 2019;179(7):942-952.

5. Schena FP, Nistor I. Epidemiology of IgA nephropathy: a global perspective. *Semin Nephrol*. 2018;38(5):435-442.

6. Hiki Y, Odani H, Takahashi M, et al. Mass spectrometry proves under-O-glycosylation of glomerular IgA1 in IgA nephropathy. *Kidney Int*. 2001;59(3):1077-1085.

7. Novak J, Julian BA, Tomana M, Mestecky J. IgA glycosylation and IgA immune complexes in the pathogenesis of IgA nephropathy. *Semin Nephrol*. 2008;28(1):78-87.

8. Lai KN. Pathogenesis of IgA nephropathy. *Nat Rev Nephrol*. 2012;8(5):275-283.

9. Patel C, Shah HH. Vaccine-associated kidney diseases: a narrative review of the literature. *Saudi J Kidney Dis Transpl*. 2019;30(5):1002-1009.

10. van den Wall Bake AW, Beyer WE, Evers-Schouten JH, et al. Humoral immune response to influenza vaccination in patients with primary immunoglobulin A nephropathy. An analysis of isotype distribution and size of the influenza-specific antibodies. *J Clin Invest*. 1989;84(4):1070-1075.

11. McNally A, McGregor D, Searle M, Irvine J, Cross N. Henoch-Schönlein purpura in a renal transplant recipient with prior IgA nephropathy following influenza vaccination. *Clin Kidney J*. 2013;6(3):313-315.
12. Fischer AS, Moller BK, Krag S, Jespersen B. Influenza virus vaccination and kidney graft rejection: causality or coincidence. *Clin Kidney J*. 2015;8(3):325-328.

13. Baden LR, El Sahly HM, Essink B, et al. Efficacy and safety of the mRNA-1273 SARS-CoV-2 vaccine. *N Engl J Med*. 2021;384(5):403-416.

14. Premkumar L, Segovia-Chumbez B, Jadi R, et al. The receptor binding domain of the viral spike protein is an immunodominant and highly specific target of antibodies in SARS-CoV-2 patients. *Sci Immunol*. 2020;5(48):eabc8413.

15. Stamatatos L, Czartoski J, Wan YH, et al. mRNA vaccination boosts cross-variant neutralizing antibodies elicited by SARS-CoV-2 infection. *Science*. 2021; https://doi.org/10.1126/science.abg9175

16. Fernandez A. Glycosylation of SARS-CoV-2 steers evolutionary outcomes in the postvaccination phase. *ACS Pharmacol Transl Sci*. 2021;4(1):410-412.

17. Campillo-Luna J, Wisnewski AV, Redlich CA. Human IgG and IgA responses to COVID-19 mRNA vaccines. *medRxiv*. 2021:2021:2003.2023.21254060.

18. Vaccines and Related Biological Products Advisory Committee Meeting. December 17, 2020. Vol 2021. Accessed August 3, 2021. https://www.fda.gov/media/144434/download

19. Butler DL, Gildersleeve JC. Abnormal antibodies to self-carbohydrates in SARS-CoV-2 infected patients. *bioRxiv*. 2020;2020. 10.15.341479.

20. Hassan AO, Kafai NM, Dmitriev IP, et al. A single-dose intranasal ChAd vaccine protects upper and lower respiratory tracts against SARS-CoV-2. *Cell*. 2020;183(1):169-184 e113.

21. Sterlin D, Mathian A, Miyara M, et al. IgA dominates the early neutralizing antibody response to SARS-CoV-2. *Sci Transl Med*. 2021;13(577):eabd2223.