Genetic variations in *IL1A* and *IL1RN* are associated with the risk of preeclampsia in Chinese Han population

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Preeclampsia (PE) is an excessive systemic inflammation response with dysfunction of endothelial. Our study was to investigate the association between genetic variations in IL-1 and the susceptibility to PE in Chinese Han population. 402 PE patients and 554 normal pregnant women of third trimester were enrolled. The polymorphisms of rs315952 in *IL1RN* and rs17561 in *IL1A* were genotyped by TaqMan allelic discrimination real-time PCR. Obviously statistic difference of the genotypic frequencies were found in both of IL1RN rs315952 and IL1A rs17561 between cases and controls (for rs315952, \( P < 0.001 \); for rs17561, \( P = 0.021 \).). For rs315952, the C allele was associated with development of PE (\( P = 0.003, \ OR = 1.319, 95\%CI 1.099–1.583 \)). Patients with CC or CT genotype were less likely to develop severe PE than patients carrying TT genotype (\( P < 0.001, \ OR = 0.24, 95\%CI 0.15–0.40 \)). For rs17561, the C allele was the risk factor for predisposition to PE (\( P = 0.012, \ OR = 1.496, 95\%CI 1.089–2.055 \)). Our results suggest *IL1RN* and *IL1A* may involve in the development of PE in Chinese Han population.

As a pregnancy-specific disorder, preeclampsia (PE) is diagnosed by the onset of hypertension and proteinuria after 20 gestational weeks. Because PE is often accompanied with multi-organ disorders and there is no effective treatment other than terminating pregnancy, it becomes a leading cause of perinatal morbidity and mortality. Furthermore, hypertension is the most common risk factor for the initial myocardial infarction, which may affect the hospital mortality. Therefore, the onset of PE must get our attention. Although unremitting efforts have been taken for many years, the etiology and pathogenesis of PE remain unclear. Several pathological factors are proposed to explain the development of PE, such as immune maladaptation, exaggerated inflammation, abnormal trophoblast invasion and genetic involvement. The susceptibility to PE is affected by both genetic and environment interaction. To date, a variety of candidate genes have been tested to have associations with PE, including *ERAP2*, *COMT*, *TNF-α* and so on⁶⁻⁹. Moreover, some candidate genes, such as *CYP11B2* and *ACE*, were demonstrated to be linked with hypertension⁶⁰.

PE is an excessive systemic inflammation response with dysfunction of endothelial.⁶ The serum levels of several cytokines, such as IL-1, TNF-α, IL-6, IL-8, are increased in PE patients¹¹⁻¹³. IL-1 is a critical mediator of the inflammatory response and important factor that stimulates the structural and functional alterations in endothelial cells.²⁰ It consists of three molecules namely IL-1α, IL-1β and IL-1 receptor antagonist (IL-1Ra), which is encoded by *IL-1A*, *IL-1B* and *IL-1RN*, respectively. IL-1α and IL-1β are pro-inflammatory cytokines, which bind to the IL-1 receptor (IL-R) to activate signal transduction and exert biological effects while IL1Ra is the anti-inflammatory cytokine by competing for binding sites on the IL-1R.²¹ The polymorphisms of promoter and coding regions in cytokine genes may affect the production and function of cytokines. The rs315952, located in exon 7 of *IL-1RN*, has been suggested to be associated with several autoimmune disorders, such as ankylosing spondylitis, systemic lupus erythematosus, knee osteoarthritis¹⁴⁻¹⁷. The plasma levels of IL1Ra were found to be higher with the allele C of rs315952, especially for the homozygous carriers of the C allele in acute respiratory distress syndrome.²⁸ The IL1A rs17561 located in the exon 5, is a nonsynonymous variant (Ala114Ser),²⁹ is reported to have associations with inflammatory disorders and reflect increased IL-1α production in carriers of this allele T and the allele T of rs1800587. The SNP of rs17561 is associated with the risk of endometriosis.
ovarian cancer, ankylosing spondylitis and so on\textsuperscript{21–23}. However, the mechanism of the functional role of these polymorphisms has not been identified.

Considering the exaggerated inflammation response and the changes in serum levels of inflammatory cytokines of PE patients, we hypothesized polymorphisms of IL-1 may influence the susceptibility and clinical symptoms of PE. The aim of this study is to investigate the association between polymorphisms of IL1RN rs315952 and IL1A rs17561 and susceptibility to PE in Chinese Han population.

Results

Demographic and clinical characteristics. The clinical characteristics of people enrolled and \textit{p}-value for comparison between cases and controls were summarized in table 1. The mean age of cases and controls was 30.74 ± 5.70 and 30.67 ± 4.48 years old, respectively. As compared with controls, PE patients had earlier gestational weeks at delivery (36.31 ± 2.93 weeks vs. 39.23 ± 1.35 weeks, \(P < 0.001\)), lower birth weight of offspring (2684 ± 909 g vs. 3382 ± 394 g, \(P < 0.001\)) and higher blood pressure (\(P < 0.001\)). In addition, there was higher serum levels of the count of white blood cell and neutrophil, triglycerides, ALT, AST, urea nitrogen, creatinine in PE patients (\(P < 0.05\)). There is no statically significant difference in weight increased during pregnancy, number of abortion, age of menarche, serum level of total cholesterol between the two groups (\(P > 0.05\)).

Genetic analysis. The population of controls was in Hardy–Weinberg equilibrium for both SNPs (for rs315952, \(\chi^2 = 0.024, P = 0.877\); for rs17561, \(\chi^2 = 0.0005, P = 0.983\)). Table 2 showed the genotypic and allelic frequencies of rs315952 and rs17561 in cases and controls. For rs315952, there was significant difference of genotypic and allelic frequencies between two groups (\(\chi^2 = 13.741, P = 0.001\) by genotype; \(\chi^2 = 8.869, P = 0.003\) by allele). When subdividing these samples into CC/TT/TT groups or TT/CT/CC groups, we found a significant difference between cases and controls in TT/CT/CC groups (\(P < 0.001\), OR = 1.775, 95\%CI 1.308–2.411). The C allele of rs315952 was associated with development of PE (OR = 1.319, 95\%CI 1.099–1.583). Similarly, for rs17561, the distribution of genotypes differed significantly between cases and controls (\(\chi^2 = 7.725, P = 0.021\), and there is a statistical difference in AA + AC and CC groups (\(P = 0.007, OR = 1.605, 95\%CI 1.137–2.266\)). The C allele was the risk allele for predisposition to PE (\(P = 0.012, OR = 1.496, 95\%CI 1.089–2.055\)).

Analysis of genotype-phenotype relationship. The results were present in table 3, 4. For rs17561, patients who had genotype CC showed higher urea nitrogen than those who had genotype AA and AC (4.81 ± 2.09 mmol/L vs. 4.10 ± 1.36 mmol/L, \(P = 0.002\)). Similarly, the level of serum creatinine was higher in patients carrying CC genotype than patients carrying AA and AC genotype (69.64 ± 19.89 umol/L vs. 63.45 ± 20.51 umol/L, \(P = 0.041\)). We did not found associations between rs315952 and demographic characteristics and results of serum biochemistry. However, the distribution of the severe preeclampsia in this three genotypes of rs315952 was significantly different (\(P < 0.001\)). Patients with CC or CT genotype were less likely to develop severe PE than patients carrying TT genotype (\(P < 0.001, OR = 0.24, 95\%CI 0.15–0.40\)).

Table 1 | The demographic and clinical characteristics of cases and controls

| Characteristic                   | Cases(n = 402) | Controls(n = 554) | t   | p-value |
|----------------------------------|---------------|------------------|-----|---------|
| Age(years)                       | 30.7 ± 5.7    | 30.7 ± 4.5       | 0.197 | 0.844   |
| Gestational age(weeks)           | 34.5 ± 4.4    | 38.7 ± 1.8       | -17.46 | <0.001  |
| Gestational age at delivery(weeks) | 36.3 ± 2.9    | 39.2 ± 1.4       | -16.68 | <0.001  |
| Weight increased during pregnancy(kg) | 18.15 ± 6.65  | 17.68 ± 4.42     | 0.593   | 0.554   |
| Birth weight of offspring(g)     | 2684 ± 909    | 3382 ± 394       | -12.78 | <0.001  |
| Number of abortion               | 0.9 ± 1.1     | 0.8 ± 0.9        | 1.69  | 0.092   |
| Age of menarche(years)           | 14.1 ± 1.3    | 14.2 ± 1.4       | -1.21  | 0.226   |
| Systolic blood pressure(mmHg)    | 157 ± 22      | 113 ± 10         | 35.65  | <0.001  |
| Diastolic blood pressure(mmHg)   | 103 ± 16      | 74 ± 8           | 33.52  | <0.001  |
| White blood cell(\times 10^12/L) | 9.66 ± 3.11   | 9.10 ± 3.44      | 2.527  | 0.012   |
| Neutrophil(\times 10^9/L)        | 7.41 ± 2.93   | 6.78 ± 2.37      | 3.175  | 0.002   |
| Triglycerides(mmol/L)            | 3.81 ± 1.86   | 2.94 ± 1.39      | 2.187  | 0.030   |
| Total cholesterol(mmol/L)        | 6.56 ± 1.98   | 6.34 ± 1.66      | 0.521  | 0.603   |
| ALT(IU/L)                        | 27.29 ± 45.04 | 16.53 ± 19.48    | 4.287  | <0.001  |
| AST(IU/L)                        | 31.01 ± 38.56 | 20.60 ± 16.96    | 4.842  | <0.001  |
| Urea nitrogen(mmol/L)            | 4.71 ± 2.01   | 3.15 ± 1.11      | 13.38  | <0.001  |
| Creatinine(umol/L)               | 68.77 ± 20.07 | 55.38 ± 15.49    | 10.68  | <0.001  |

Table 2 | The genotypic and allelic frequencies of rs315952 and rs17561 in two groups

| Genotypes | Cases | Controls | \(\chi^2\) | p-value* | OR(95\%CI) |
|-----------|------|----------|------------|----------|------------|
| rs315952  |      |          |            |          |            |
| CT        | 169  | 269      | 13.741     | 0.001    |            |
| CC        | 114  | 101      |            |          |            |
| TT        | 119  | 184      |            |          |            |
| CC + CT   | 283  | 370      | 1.403      | 0.236    | 1.183(0.896–1.561) |
| TT        | 119  | 184      |            |          |            |
| CC        | 114  | 101      |            |          |            |
| TT + CT   | 288  | 453      |            |          |            |
| Alleles   |      |          |            |          |            |
| C         | 397  | 471      | 8.869      | 0.003    | 1.319(1.099–1.583) |
| T         | 407  | 637      |            |          |            |
| rs17561   |      |          |            |          |            |
| Genotypes |      |          |            |          |            |
| AC        | 53   | 111      | 7.725      | 0.021    |            |
| CC        | 344  | 436      |            |          |            |
| AA        | 5    | 7        |            |          |            |
| AA + AC   | 397  | 547      | 0.001      | 0.978    | 1.016(0.320–3.225) |
| CC        | 5    | 7        |            |          |            |
| AA + AC   | 57   | 118      | 7.324      | 0.007    | 1.605(1.137–2.266) |
| Alleles   |      |          |            |          |            |
| C         | 741  | 983      | 6.240      | 0.012    | 1.496(1.089–2.055) |
| A         | 63   | 125      |            |          |            |

*p-value < 0.025 was considered significant after Bonferroni’s correction.
Table 3 | Associations between genotypes of rs315952 and characteristics among PE patients

| db SNP ID (T/C) | (1)1/1 | (2)1/2 | (3)2/2 | (1) vs. (2) vs. (3) | (1) vs. (2) | (1) vs. (3) | (2) vs. (3) | (1) + (2) vs. (3) | (1) vs. (2) + (3) |
|-----------------|------|------|------|-------------------|----------|-------------|---------|-----------------|-----------------|
| (allele1/allele2) | (n)  | (n)  | (n)  | p-value | p-value | p-value | p-value | p-value | p-value |
| rs315952        | 119  | 169  | 114  | 0.819   | 0.562  | 0.922   | 0.642   | 0.803  | 0.669   |
| Age (years)     | 30.6 ± 5.3 | 31.0 ± 5.8 | 30.6 ± 5.9 | 0.819   | 0.562  | 0.922   | 0.642   | 0.803  | 0.669   |
| Gestational age at diagnosis (weeks) | 32.8 ± 5.6 | 31.6 ± 6.0 | 32.7 ± 5.2 | 0.180   | 0.109  | 0.944   | 0.134   | 0.356  | 0.277   |
| Age of menarche (years) | 14.0 ± 1.4 | 14.1 ± 1.3 | 14.2 ± 1.3 | 0.775   | 0.871  | 0.500   | 0.569   | 0.486  | 0.665   |
| Gestational age at delivery (weeks) | 36.7 ± 2.9 | 36.0 ± 3.0 | 36.4 ± 2.8 | 0.256   | 0.103  | 0.496   | 0.376   | 0.798  | 0.164   |
| Serum biochemistry (Mean ± SD) | | | | | | | | | |
| White blood cell (×10^9/L) | 9.62 ± 3.37 | 9.62 ± 2.85 | 9.77 ± 3.21 | 0.913   | 0.979  | 0.728   | 0.689   | 0.668  | 0.878   |
| Neutrophil (×10^9/L) | 7.56 ± 3.32 | 7.22 ± 2.60 | 7.53 ± 3.01 | 0.615   | 0.389  | 0.943   | 0.443   | 0.631  | 0.551   |
| ALT (IU/L) | 31.94 ± 66.71 | 25.80 ± 36.03 | 24.65 ± 24.30 | 0.428   | 0.274  | 0.237   | 0.841   | 0.480  | 0.323   |
| AST (IU/L) | 34.35 ± 46.83 | 29.47 ± 29.72 | 29.72 ± 29.72 | 0.545   | 0.306  | 0.376   | 0.959   | 0.685  | 0.271   |
| Urea nitrogen (mmol/L) | 4.62 ± 1.51 | 4.79 ± 2.39 | 4.69 ± 1.87 | 0.776   | 0.485  | 0.795   | 0.683   | 0.895  | 0.560   |
| Creatinine (μmol/L) | 67.51 ± 16.92 | 69.63 ± 18.91 | 68.83 ± 24.43 | 0.704   | 0.402  | 0.632   | 0.757   | 0.971  | 0.436   |
| Clinical characteristics | | | | | | | | | |
| Systolic pressure (mmHg) | 153.6 ± 20.7 | 158.5 ± 22.2 | 157.5 ± 23.1 | 0.175   | 0.069  | 0.187   | 0.717   | 0.686  | 0.067   |
| Diastolic pressure (mmHg) | 101.5 ± 15.7 | 104.2 ± 15.4 | 104.0 ± 16.0 | 0.326   | 0.158  | 0.239   | 0.911   | 0.620  | 0.135   |
| Severe preeclampsia | 70/119 | 149/169 | 93/114 | <0.001 | <0.001 | <0.001 | 0.123 | <0.001 |<0.001 |
| OR(95%CI) | 0.19(0.11–0.35) | 0.32(0.18–0.59) | 1.68(0.87–3.27) | 0.72(0.42–1.24) | 0.24(0.15–0.40) |
### Table 4 | Associations between genotypes of rs17561 and characteristics among PE patients

| db SNP ID      | (1)(1)/1 | (2)(1)/2 | (3)(2)/2 | (1)(vs.(2))vs.(3) | (1)(vs.(2)) | (1)(vs.(3)) | (2)(vs.(3)) | (1) + (2)(vs.(3)) | (1)(vs.(2)) + (3) |
|---------------|----------|----------|----------|-------------------|-------------|-------------|-------------|-------------------|-------------------|
| (allele1/allele2) | (n)      | (n)      | (n)      | p-value           | p-value     | p-value     | p-value     | p-value           | p-value           |
| rs17561 (C/A)  | 344      | 53       | 5        |                   |             |             |             |                   |                   |
| Demographic characteristics (Mean ± SD) |          |          |          |                   |             |             |             |                   |                   |
| Age (years)    | 30.6 ± 5.7 | 31.5 ± 5.6 | 30.8 ± 9.9 | 0.574             | 0.292       | 0.945       | 0.790       | 0.990             | 0.307             |
| Gestational age at diagnosis (weeks) | 32.2 ± 5.8 | 32.8 ± 4.7 | 35.0 ± 2.6 | 0.565             | 0.503       | 0.393       | 0.519       | 0.405             | 0.394             |
| Age of menarche (years) | 14.1 ± 1.3 | 14.1 ± 1.4 | 14.2 ± 0.4 | 0.979             | 0.997       | 0.838       | 0.843       | 0.837             | 0.958             |
| Gestational age at delivery (weeks) | 36.2 ± 3.0 | 36.9 ± 2.2 | 36.7 ± 3.1 | 0.399             | 0.186       | 0.727       | 0.881       | 0.773             | 0.177             |
| Serum biochemistry (Mean ± SD) |          |          |          |                   |             |             |             |                   |                   |
| White blood cell (×10^9/L) | 9.63 ± 3.08 | 9.87 ± 3.41 | 9.35 ± 2.25 | 0.860             | 0.617       | 0.840       | 0.722       | 0.823             | 0.676             |
| Neutrophil (×10^9/L) | 7.37 ± 2.91 | 7.72 ± 3.19 | 6.96 ± 2.03 | 0.726             | 0.469       | 0.760       | 0.588       | 0.734             | 0.556             |
| ALT (IU/L)     | 28.18 ± 47.84 | 22.64 ± 21.74 | 13.70 ± 4.84 | 0.604             | 0.424       | 0.524       | 0.703       | 0.545             | 0.333             |
| AST (IU/L)     | 31.91 ± 41.21 | 26.10 ± 15.84 | 23.16 ± 11.85 | 0.552             | 0.323       | 0.615       | 0.871       | 0.647             | 0.281             |
| Urea nitrogen (mmol/L) | 4.81 ± 2.09 | 4.08 ± 1.32 | 4.32 ± 1.93 | 0.054             | 0.017       | 0.583       | 0.801       | 0.661             | 0.002             |
| Creatinine (μmol/L) | 69.64 ± 19.89 | 63.68 ± 21.49 | 61.39 ± 7.51 | 0.120             | 0.060       | 0.356       | 0.800       | 0.403             | 0.041             |
| Clinical characteristics |          |          |          |                   |             |             |             |                   |                   |
| Systolic pressure (mmHg) | 156.6 ± 22.6 | 156.6 ± 17.6 | 168.0 ± 25.9 | 0.519             | 0.980       | 0.254       | 0.270       | 0.252             | 0.763             |
| Diastolic pressure (mmHg) | 103.3 ± 15.9 | 103.4 ± 14.2 | 106.0 ± 19.5 | 0.929             | 0.956       | 0.703       | 0.728       | 0.704             | 0.872             |
| Severe preeclampsia | 266/344 | 42/53 | 4/5 | 0.945 | 0.755 | 1.000 | 1.000 | 1.000 | 0.737 |

0.89 (0.44–1.82) 0.85 (0.09–7.74) 0.96 (0.10–9.42) 0.87 (0.10–7.84) 0.89 (0.45–1.76)
Discussion

The basic pathological change of PE is the spasm of systemic small vessel\(^\text{38}\)\(^\text{39}\), thus resulting a reduction of blood flow in various organs. Other than the most common symptoms hypertension and proteinuria, PE is often accompanied with additional disturbance of multi-organs, such as activation of the clotting system, impaired liver and renal function, pulmonary edema in cases of severe PE particularly. Due to unclear pathogenesis, there are no effectively preventive methods and the treatments of PE is limited to deal with clinical symptoms and terminate pregnancy\(^\text{3}\). In our study, PE patients had earlier gestational weeks at delivery and lower birth weight of offspring, which demonstrated the fact that PE patients usually have to choose induced preterm delivery to relieve the clinical symptoms. What’s more, the PE patients enrolled in our study had higher serum level of ALT, AST, urea nitrogen and creatinine compared with controls. Moreover, the count of white blood cells and neutrophil were higher in cases than controls, which revealed a status of exaggerated inflammation in PE.

The normal pregnancy is suggested to be a condition of controlled mild maternal systemic inflammation. And exaggerated inflammation is proposed to play an important role in the development of PE\(^\text{\text{10–13}}\). There is Th1/Th2 imbalance in PE patients, and Th1 immunity is predominant in the immune and inflammatory response\(^\text{29}\). Among the Th1-type pro-inflammatory cytokines, IL-1 initiates and perpetuates inflammatory response. The levels of IL-1 synthesized both from decidual lymphocytes and peripheral blood mononuclear cells are higher in PE patients\(^\text{8,29,30}\). IL-1 can stimulate expression and activity of matrix metalloproteinase (MMP) 9 and MMP2, thus regulate trophoblast differentiation along the invasive pathway, which may affect the process of placentation\(^\text{29,37}\). Moreover, IL-1 can alter the structure and function of endothelial cells. In vitro, IL-1 produced by placenta altered the proliferation of umbilical vein endothelial cell and induced the secretion of soluble ICAM and IL-6\(^\text{6}\), whose serum levels were increased in PE patients. Therefore, IL-1 is a potential mediator of endothelial dysfunction and may involve in the development of PE.

Previously, genetic researches, including the polymorphic and functional studies, have been carried out in the investigations of PE or hypertension\(^\text{29–30}\). Santulli et al. found that CaMK4 deletion induced hypertension through the influence on the synthease activity of endothelial nitric oxide and further confirmed the association between rs10491334 variant and a reduction in the expression levels of CaMKIV in hypertensive patients. Among G-protein-coupled receptor kinase (GRK) family, the GRK5 Leu41 allele was reported to decrease the risk for adverse cardiovascular outcomes in treated hypertensive patients\(^\text{29}\). What’s more, GRK2 abundance was related with hypertension through impairment of β-adrenergic mediated vasodilatation, which was also present in PE\(^\text{30}\). Furthermore, the associations between polymorphisms of IL-1 and the risk of PE have also been investigated. However, because of mutations, genetic recombination, human mobility and natural selection, the frequencies of genotypes and alleles are different in the population from different race or region, therefore the results of these studies are controversial\(^\text{29–34}\). Goddard et al. analyzed 775 SNPs in 190 candidate genes, founding that rs3783550 in IL1A had a statistically significant association with PE in Chile (P = 0.0014)\(^\text{37}\). Faisal et al. studied on 135 PE patients and 112 controls from Finland, suggesting an association between the variable copy number of 86-bp tandem repeats (VNTR) of the IILRN polymorphism and PE\(^\text{33}\). However, Valenza et al. did not find the associations between polymorphism of IILRN VNTR and PE in Mexican-Mestizo and Maya-Mestizo women\(^\text{34}\). And polymorphism of rs16944 and rs1143634 in IL-1β showed no associations with the risk of PE in Taiwanese\(^\text{35}\).

In our study, we selected IL-1RN rs315952 and IL1A rs17561 to investigate the associations with PE. Both rs315952 and rs17561 polymorphisms had a significant association with PE in Chinese Han population. For rs315952, the C allele was the risk allele for the development of PE. Patients with CC or CT genotype were more inclined to develop severe PE than patients with TT genotype. For rs17561, the frequency of the C allele was higher in PE patients. The levels of serum urea nitrogen and creatinine in patients carrying CC genotype were higher than those carrying AA or AC genotype. Our results suggested that IILRN and IL1A may involve in the development of PE, which is consistent with the studies of Goddard and Faisal\(^\text{33}–\text{35}\). To our knowledge, it is the first time to examine whether the SNPs of rs315952 and rs17561 in IL-1 are associated with the risk of PE.

There are several limitations in this study that should be noted. Firstly, the sample size was relatively small and all participants were ethnic Han Chinese. Because the results are affected by ethnic or region, our study could not represent other human races. Secondly, other variants of IL-1A and IL-1RN were not genotyped for the associations with PE, thus genetic linkage analysis could not be conducted. Further studies on the associations of genetic variants of IL-1 with PE will be necessary. Thirdly, the serum level of IL-1 were not measured in our population, thus the relationships between the SNPs and IL-1 levels were not observed. In spite of some limitations, our study suggested that IILRN and IL1A may involve in the development of PE in Chinese Han population. Our results need to be validated in a larger sample and in other races with functional analyses to clarify the potential mechanisms underlying the links between SNPs of IL-1 and susceptibility to PE.

Methods

Subjects. 482 PE patients and 554 normal pregnant women of third trimester admitted to the Affiliated Hospital of Qingdao University, Linyi People’s Hospital and Heze Municipal Hospital were enrolled in our study. Age was matched in the two groups. The mean age of PE patients was 30.74 ± 5.70 years old and the controls were 30.67 ± 4.88 years old. The controls requested pregnant women without multiple pregnancy or any pathological states, such as premature rupture of membrane, placenta previa, poly- or oligo-hydramnios, threatened abortion, diabetes mellitus, hypertension, autoimmune disease and so on. If the fetus was congenital malformations or macrosomia, the maternal sample was removed from the control group. The demographic and clinical characteristics of participants, such as pregnancy and family history, clinical symptoms, the results of blood routine test, placenta previa, poly- or oligo-hydramnios, threatened abortion, diabetes mellitus, pulmonary edema or cyanosis, impaired liver functions, epigastric or right upper-quadrant pain, and fetal growth restriction\(^\text{36}\).

Extraction of DNA and genotyping of IL-1. Genomic DNA was extracted from 300 ml peripheral blood by using Qiagen DNA extraction kit. The polymorphisms of rs315952 in IILRN and rs17561 in IL1A were genotyped by TaqMan allelic discrimination real-time PCR. The Taqman probes and primers were designed by Applied Biosystems of Life Technologies. For rs315952, the sequence of forward and reverse primer is 5′-GCTCGGCTTCTATCGCGCTCTCAGGACG-3′ and 5′-GGCCCAAGCAACCAGGTGGAGTCG-3′ respectively. For rs17561, the forward primer is 5′-CACT TTGCTCAGGAAGCTAAAGGTTG-3′, and reverse primer is 5′-TACGTTCCGTTGATGATT TTTAA-3′. The polymerase chain reaction (PCR) was conducted in 25 ul reaction mixture, containing 20 × SNP Genotyping Assay 1.25 ul, 2 × PCR Master Mix 12.5 ul, DNA and DNA-free water 11.25 ul. The amplifications were carried out by C1000™ thermal cycler system with the following conditions: 95°C for 3 min, followed by 45 cycles at 95°C for 15 sec and 60°C for 1 min. For each cycle, the fluorescent signal from the VIC- or FAM-labeled probes was determined. The discrimination of genotypes was conducted with Bio-Rad CFX Manager 3.0 software.

All experiments were carried out in accordance with relevant regulations and guidelines.

Statistical analysis. All analyses were performed by statistical software package SPSS19.0. Hardy-Weinberg equilibrium was examined in control group with goodness-of-fit χ² test. Student’s t test was used to test the comparison of demographic and clinical characteristics between cases and controls. An analysis of variance (ANOVA) was used to conduct genotype-phenotype analysis. The level of
statistical significance was defined as p-value < 0.05. The allelic and genotypic distributions of cases and controls were compared by Pearson’s χ² test (if expected values were below 5, Fisher’s exact test was used) and P-values < 0.025 were considered significant when Bonferroni’s correction was made.

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Author contributions
J.L. carried out the initial analyses, reviewed and revised the manuscript. M.L. carried out the initial analyses, and approved the final manuscript as submitted. J.L.* and M.L.* contribute equally to this work. J.Z. and P.T. designed the data collection instruments. J.W. coordinated and supervised data collection. X.L. approved the final manuscript as submitted. J.L. drafted the initial manuscript, and wrote manuscript. X.L. conceptualized and designed the study, drafted the initial analyses, and approved the final manuscript as submitted. J.Z.

Additional information
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