Abstract
The microbial decay of stone monuments in many tropical countries has become a serious threat for their future existence. Present investigation was conducted to evaluate the status of mycobial decay of stone monuments of Dharmarajika. A total of nineteen fungal species belonging to thirteen genera were isolated from colored stones, patinas and biofilms produced on the surfaces of monuments due to mechanism of biodeterioration. The fungal species Alternaria alternata, Aspergillus fumigatus, Aspergillus flavus, Aspergillus niger, Cladosporium herbarum, Curvularia lunata, Dematium spp., Fusarium oxysporum, Mucor hiemalis, Penicillium chrysogenum, Penicillium frequentans and Rhizopus oryzae were prevalent. Some variations in the occurrence of fungal species were also recorded. The isolated fungal species were also analyzed for their organic acids production to understand the corrosive effects of their metabolic secretions on stone materials.

It was found that all prevalent fungal strains produced different organic acids including oxalic acids, citric acids, fumaric acid, acetic acid, gluconic acid and succinic acid.

Keywords: Mycobial decay; Dharmarajika; Biodeterioration; Monuments; Organic acids

Abbreviations: HPLC: High Performance Liquid Chromatography; UNESCO: United Nations Educational Scientific and Cultural Organization; SID: Serial Identification

Introduction
The weathering and decay of cultural heritage is a complex process, which is caused by the interaction of many physical, chemical and biological agents. The different biological deterioriogens such as bacteria, algae, cyanobacteria, bryophytes, mosses, fungi, insects, rodents, birds and human beings play a momentous role in the decay of historical monuments. The biological growth of microorganisms can cause staining, cracking, powdering, disfigurement and displacement of building material, which leads to the permanent loss of stone monuments. Such aesthetic damage is accompanied by the transformation of the chemical and mechanical properties of stone material and this causes the formation of surface patinas of different colors [1,2].

Among different biological agents, fungi play more dangerous role in the bio-deterioration of stone monuments because of their complex metabolic activities on stone surface. Many fungal species have been isolated from the stone monuments located in different countries. Species of Alternaria, Aspergillus, Acremonium, Arthrobotrys, Auerobasidium, Cladosporium, Curvularia, Drechslera, Fusarium, Helminthosporium, Mucor, Phoma, Penicillium, Rhizopus, Trichotheceum and Trichoderma have been reported as prevalent fungi involved in biological decay of stone monuments. The growth of these fungal genera on stone monuments was a cause of staining and structural decay of stone material of these monuments [3-8]. Fungi produce many inorganic and organic acids during their metabolic activities on monuments. These acids cause mineral dissolution and change structural configuration of stone material. The organic acids such as oxalic, lactic and gluconic acids function as chelating agents and can demineralize a variety of stone substrates including calcium, silicon, iron, magnesium and manganese [9,10]. The enzymes produce during metabolic activities of fungi are involved in transformation of complex and binding molecules of stone monuments into simple dissolvable molecules [11].

Fungal metabolites can cause solubilization of cations and produce patinas of different mineralogical composition. The discoloration of monuments due to the formation of patinas is more havoc on light colored stone monuments. The interaction between fungal hyphae and stone substrate also causes the formation of biofilms with different colors and chemical compositions. The biodeterioration of binding material of stone monuments starts by uptake of calcium and then this action leaves the monumental surface eroded and exposed it to water and frost attack [12,13]. The appearance of black crusts on lime stone monuments in two Spanish Cathedrals was studied [14]. They found that Penicillium frequentans was associated with formation of black biofilms on the surface of these monuments. In an investigation it was found that black stain on stone monuments were due to Aspergillus sydowi and Stachybotrys artro [15]. The fungal colonization of sandstone and granite from Antarctica revealed that Alternaria, Aspergillus, Auerobasidium, Candida, Cladosporium, Fusarium, Helminthosporium, Mucor and Penicillium and Sporobolomyces were found active in weathering of rock and stone monuments [16]. The black fungi such as Phoma and Alternaria were found as deteriorating agent of marble and lime stone monuments [17]. The study of different aspects of biodeterioration of monuments is always helpful for the proper preservation of this world heritage. The archaeological monuments of Taxila are under the threat of severe biological...
decay. Most monuments of Taxila have the signs of structural distress such as leans, bulges, displacement and fractures. The present investigation was carried out to understand the mechanism of biodeterioration of stone monuments of Taxila.

Materials and Methods

Study sites

Dharmarajika Stupa is one of the eight shrines constructed in the 3rd century BC during the reign of Emperor Ashoka of the Mauryan dynasty. It is earliest and largest Buddhist stupa in Taxila. The modern name of Dharmarajika is Chirtope. Dharmarajika is also famous for monastic quarters, stupas and chapels. The remains of Dharmarajika are a source of valuable data for history, local architecture and art of past times. The masonry of Dharmarajika remains is rough limestone rubble. The SID number of Dharmarajika as world heritage is 139-006 in UNESCO world heritage sites list. In present investigation fungal species were isolated from six monuments including Main central stupa of Dharmarajika, Figural decorated stupa, Wall of a small stupa, Buddhist stupa no. 12, Wall of stupa no. 11, Chapel and Stupa no.13. The archaeological monuments of Dharmarajika are under the threat of biodeterioration (Figures 1-4).

Isolation of stone mycoflora

Sterile adhesive tape was used for the isolation of mycoflora from the surface of monuments. The hard and dried stains were removed by etching with the help of sterilized sticking tape and then these small pieces of tape were dispensed into Petri dishes or in test tubes slants containing culture media. Collection of sample was done by attaching of a strip of tape to the surface being sampled. Adhesive tape was pressed firmly on the surface of compact alternations. The tape was affixed on to a sterile glass slide and was stored in dark at 4 °C for initial observation. The petri plates having sampling material were incubated at 25 °C ± 3 °C for 8-10 days and colonies were counted and isolated by digital colony counter.

Identification of fungi

The macro morphological and micro morphological features of taxonomic interest were studied at different microscopic powers i.e. X10, X40 and X100. The Identification of fungi was done with the help of reference works [18,19].

Organic acids determination by HPLC

For the Determination of organic acids produced by isolated...
fungi Malt extract broth medium was used. Fungal strains were grown and multiplied in Malt extract broth medium with three replicates in each treatment. The purified solutions were prepared. On fifth day cultures were blended having three replicates of each culture. Blended cultures were centrifuged at 2000 rpm for 20 minutes. Supernatant of each blended culture was filtered through 0.45 µm non-sterile 4mm sized micro filter syringes. Further purification was done by centrifugation of the filtered cultures (stored in up and off tip) in micro centrifuge.

The 20 µl purified solutions of each culture was injected in bio-rad ion exchange column of Aminex 87-H (25*4.6 mm). The operating conditions consisted of 0.001 N H2SO4 the mobile phase at a constant (isocratic) flow rate of 0.6 ml min-1 and column was operated at 25 ºC. Organic acid concentrations in samples were determined with the help of RI detector [20]. The software used during HPLC was a Turbochrome navigator system. RI impulse was read with the help of Turbochrome navigator programmer in gl-1. The unknown organic acids in purified solution were determined by comparing the retention times and peak areas of chromatograms with the standards of oxalic acid, citric acid, gluconic acid, fumaric acid and acetic acid.

Results

Mycoflora of stone monuments

Eighty sampling materials were isolated from the six stone monuments of Dharmarajika and 59 sampling material (73.75%) showed the presence of fungi. Nineteen fungal species belonging to thirteen different genera were identified. The isolated fungi were *Alternaria alternata*, *Alternaria solani*, *Aspergillus fumigatus*, *Aspergillus flavus*, *Aspergillus niger*, *Cladosporium herbarum*, *Cladosporium cladosporioides*, *Curvularia lunata*, *Dematium spp.*, *Fusarium oxysporum*, *Fusarium culmorum*, *Geotrichum candidum*, *Helminthosporium solani*, *Mucor hiemalis*, *Penicillium chrysogenum*, *Penicillium frequentans*, *Phoma glomerations*, *Rhizopus oryzae* and *Trichoderma spp.* [21].

The composition of fungal flora of six Archaeological sites of Dharmarajika is given in Table 1. The fungal species *Alternaria alternata*, *Aspergillus fumigatus*, *Aspergillus flavus*, *Aspergillus niger*, *Cladosporium herbarum*, *Fusarium oxysporum*, *Mucor hiemalis*, *Penicillium chrysogenum*, *Penicillium frequentans*, *Rhizopus oryzae*, *Curvularia lunata* and *Dematium spp.* were found as common and dominant fungal deteriogens of stone monuments of six archaeological sites of Dharmarajika, Taxila. These fungi were isolated from most of the sampling materials isolated from monuments. Some fungal species also showed variations in their occurrence. This variation is shown in Table 2.

Organic acids of isolated fungal species

The organic acid produced by fungal isolates in broth medium were analyzed. The results indicated that the highest acid producing strains were *Aspergillus niger*, *Aspergillus flavus*, *Fusarium oxysporum*, *Mucor hiemalis*, *Penicillium frequentans*, *Penicillium chrysogenum*, *Trichoderma spp.* and *Rhizopus oryzae*. The lowest acid producing fungi were *Alternaria alternata*, *Cladosporium herbarum*, *Cladosporium cladosporioides*. Most common acids produced by the fungal strains in broth medium were citric acid, oxalic acid, gluconic acid, succinic acid, fumaric acid and acetic acid. The lowest and highest values of these acids recorded are shown in Table 3.

| Serial No | Fungal Genera | Fungal Species |
|-----------|---------------|----------------|
| 1         | *Alternaria*  | *Alternaria alternata*, *Alternaria solani* |
| 2         | *Aspergillus* | *Aspergillus fumigatus*, *Aspergillus flavus*, *Aspergillus niger* |
| 3         | *Cladosporium*| *Cladosporium herbarum*, *Cladosporium cladosporioides* |
| 4         | *Curvularia*  | *Curvularia lunata* |
| 5         | *Dematium*    | *Dematium spp.* |
| 6         | *Fusarium*    | *Fusarium oxysporum*, *Fusarium culmorum* |
| 7         | *Geotrichum*  | *Geotrichum candidum*, |
| 8         | *Helminthosporium* | *Helminthosporium solani* |
| 9         | *Mucor*       | *Mucor hiemalis* |
| 10        | *Penicillium* | *Penicillium chrysogenum*, *Penicillium frequentans* |
| 11        | *Phoma*       | *Phoma glomerata* |
| 12        | *Rhizopus*    | *Rhizopus oryzae* |
| 13        | *Trichoderma* | *Trichoderma spp.* |
| Total     | 13            | 19             |

Table 1: Composition of Fungal flora isolated from Archaeological sites of Dharmarajika.
### Table 2: Variations in occurrence of fungal flora of archaeological sites of Dharmarajika.

| Fungi                          | S1 | S2 | S3 | S4 | S5 | S6 |
|-------------------------------|----|----|----|----|----|----|
| Alternaria alternata          | +  | +  | +  | +  | +  | +  |
| Alternaria solani             | +  | -  | -  | +  | +  | -  |
| Aspergillus fumigatus         | +  | +  | +  | +  | +  | +  |
| Aspergillus flavus            | +  | +  | +  | +  | +  | +  |
| Aspergillus niger             | +  | +  | +  | +  | +  | +  |
| Cladosporium herbarum         | +  | +  | +  | +  | +  | +  |
| Cladosporium cladosporioides  | -  | -  | +  | -  | -  | +  |
| Curvularia lunata             | +  | +  | +  | +  | +  | +  |
| Dematiuim spp.                | +  | +  | -  | +  | +  | +  |
| Fusarium oxysporum            | +  | +  | +  | +  | +  | +  |
| Fusarium culmorum             | +  | -  | +  | -  | -  | -  |
| Geotrichum candidum           | +  | +  | -  | -  | -  | +  |
| Helmintothorium solani        | -  | -  | +  | +  | +  | +  |
| Macr hyemalis                 | +  | +  | +  | +  | +  | +  |
| Penicillium chrysogenum       | +  | +  | +  | +  | +  | +  |
| Penicillium frequentans       | +  | +  | -  | +  | +  | +  |
| Phoma glomerata               | -  | -  | -  | +  | +  | +  |
| Rhizopus oryzae               | +  | +  | +  | +  | +  | +  |
| Trichoderma spp               | -  | +  | -  | +  | -  | +  |

### Table 3: Organic acids produced by fungi in broth medium (g/L).

| Fungi                          | Total acids | Oxalic acid | Citric acid | Gluconic acid | Fumaric acid | Succinic acid | Acetic acid |
|-------------------------------|-------------|-------------|-------------|---------------|--------------|---------------|-------------|
| Alternaria alternata          | 0.409       | 0.409       |             |               |              |               |             |
| Aspergillus niger             | 1.76        | 0.76        | 0.53        | 0.04          | 0.25         | 0.18          |             |
| Aspergillus flavus            | 0.71        | 0.21        | 0.14        | 0.12          | 0.24         |               |             |
| Aspergillus fumigates         | 0.98        | 0.41        | 0.32        | 0.11          | 0.14         |               |             |
| Cladosporium herbarum         | 0.70        | 0.47        | 0.23        |               |              |               |             |
| Dematiuim spp.                | 0.77        | 0.32        | 0.45        |               |              |               |             |
| Fusarium oxysporum            | 0.99        | 0.52        | 0.21        |               |              | 0.26          |             |
| Macr hyemalis                 | 0.86        | 0.49        | 0.24        | 0.13          |              |               |             |
| Penicillium chrysogenum       | 1.45        | 0.40        | 0.24        |               | 0.38         | 0.27          |             |
| Penicillium frequentans       | 1.02        | 0.36        | 0.21        |               | 0.28         | 0.17          |             |
| Rhizopus oryzae               | 0.46        | 0.31        | 0.15        |               |              |               |             |
| Curvularia lunata             | 0.12        | 0.12        |             |               |              |               |             |

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Discussion

Biodeterioration of archaeological monuments due to fungi has also been studied in different countries and according to the prevailing climatic conditions and related physico-chemical factors, varied genera have been reported to be dominating. The fungal genera Alternaria, Aspergillus, Acremonium, Cladosporium, Curvularia, Dematium, Helminthosporium, Mucor, Penicillium, Phoma, Rhizopus, Fusarium and Trichoderma isolated in present study were also encountered in the findings of many previous investigations carried out in different parts of the world [22]. The investigations from other countries showed almost the same genera as prevalent fungi active as biological agents in weathering of many monuments. In China the fungal strains Penicillium, Aspergillus, Cladosporium and Alternaria as dominant fungal genera during his research on the biodeterioration of Chinese Tong Tomb [4]. Forty-five fungi from Japanese paintings and he reported Aspergillus, Trichoderma, Cladosporium and Fusarium as deteriorating agents of Japanese paintings [6]. In Poland a research was conducted to isolate fungi from old ruins. He encountered Alternaria alternata, Aspergillus fumigatus, Cladosporium herbarum, Fusarium solani, Chrysosporium pannorum, Cunninghaemelia elegans, Epicocum purpurascens, Fusarium culmorum, Fusarium equiseti, Monocillium indium, Mortierella acaminata, Penicillium cyclopium, Penicillium martesic and Tririraxhium sousem as dominant fungi [23].

The analysis of chromatograms obtained by HPLC analysis also indicated different concentrations of glucose used as carbon source for the production of organic acids. The presence of organic and inorganic residues on mineral surface or within cracks and fissures can encourage the growth of fungi. It also proliferate on the waste products of algae and bacteria, dead cells and decaying plant material, dust particles, aerosol and animal faeces [24]. The isolated fungal strains and their organic acids clearly indicated the mechanism of patinas and stains formation on the surface of archaeological monuments of Dharmarajika. The organic acids analysis in the present investigation can be correlate with the formation of different colored hard stains on lime stone based monuments of Taxila as previously it has been revealed that colored patinas are formed on the surface of stone monuments due to the formation of oxalate films. These patinas are more noticeable on light colored stone monuments. The formation of yellow, brown and red patinas on marble buildings and monuments has been studied in Italy. The chemical composition of these patinas was calcium oxalate [25]. The Oxalic acid is considered as one of the most common acid exerted by fungi. This acid is involved in the formation of calcium oxalate films in the forms of whewellite and weddellite minerals [26]. Fungi can also form other metallic oxalates with different metals and metal bearing minerals like cadmium, cobalt, copper, zinc and nickel. The precipitation of carbonates on fungal hyphae particularly calcite occurs in form of calcareous salts on the surface of lime stone [27].

Weathering of stone monuments is based on many physical, chemical and biological processes which includes dissolution of carbonates and sulfates, solubilization by leaching elements from silicates, carbonates etc. weathering due to crystallization and hydration pressure microbial attack by inorganic and organic acids [25]. The study of fungal deterioration of archaeological monuments is significant. It helps not only to understand the effects of rapidly changing environmental conditions on the monuments but also helpful to find out the process of physical and chemical weathering of monuments. The importance of such studies has taken priorities in many European countries. In last decades many effective researches on different aspects of biodeterioration have been carried out in different regions of the world. The results of such investigations revealed that most of the world fame archaeological monuments are under severe biodeterioration especially in tropical countries like Pakistan due to changing environmental conditions, air pollution, industrialization, general negligence and so many other factors.

Conclusion

The results of present investigations revealed that the stone monuments of world fame archaeological sites of Taxila are under the severe biodeterioration. The fungal species isolated from the surface of monuments are found biodeterioragens as many thick biofilms, hard colored stains and patinas are very frequent on these monuments. The presence of some fungal species like Alternaria, Aspergillus, Cladosporium, Rhizopus and Penicillium can cause many aesthetical damages of world fame archaeological remains of Taxila because these fungi have been reported as biodeteriogens of cultural heritage especially stone based monuments. The stone monuments of Taxila are a sign of human civilization and concerned with Gandhara civilization so it is very important to preserve these world heritage for coming generation. The conservation authorities are recommended to take prompt action to eradicate fungal species from these monuments and also take preventive methods to avoid any future mechanism of decay.

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