Effect of rhG-CSF Combined With Decitabine Prophylaxis on Relapse of Patients With High-Risk MRD-Negative AML After HSCT: An Open-Label, Multicenter, Randomized Controlled Trial

Lei Gao, MD, PhD1; Yanqi Zhang, PhD2; Sanbin Wang, MD, PhD2; Peiyan Kong, MD, PhD1; Yi Su, MM1; Jiong Hu, MD7; Ming Jiang, MD6; Hai Bai, MD2; Tao Lang, MD2; Jishi Wang, MD, PhD1; Li Liu, MD, PhD15; Tonghua Yang, MD12; Fang Liu, MD4; Shifeng Lou, MD13; Yao Liu, MD, PhD1; Cheng Zhang, MD, PhD1; Hong Liu, MM1; Li Gao, MD, PhD1; Jia Liu, MM1; Lidan Zhu, MM1; Qin Wen, PhD1; Ting Chen, MM1; Ping Wang, MM1; Jun Rao, MD1; Min Mao, MD1; Cunbang Wang, MD7; Xiantin Duan, MD6; Le Luo, MD, MM, MS3; Xiangui Peng, MM1; Kaniel Cassady, PhD14; Jiang F. Zhong, PhD15; and Xi Zhang, MD, PhD1

Abstract

PURPOSE Relapse is a major cause of treatment failure after allogeneic hematopoietic stem-cell transplantation (allo-HSCT) for high-risk acute myeloid leukemia (HR-AML). The aim of this study was to explore the effect of recombinant human granulocyte colony-stimulating factor (rhG-CSF) combined with minimal-dose decitabine (Dec) on the prevention of HR-AML relapse after allo-HSCT.

PATIENTS AND METHODS We conducted a phase II, open-label, multicenter, randomized controlled trial. Two hundred four patients with HR-AML who had received allo-HSCT 60-100 days before randomization and who were minimal residual disease negative were randomly assigned 1:1 to either rhG-CSF combined with minimal-dose Dec (G-Dec group: 100 μg/m2 of rhG-CSF on days 0-5 and 5 mg/m2 of Dec on days 1-5) or no intervention (non-G-Dec group). The primary outcome was relapse after transplantation, and the secondary outcomes were chronic graft-versus-host disease (cGVHD), safety of the treatment, and survival.

RESULTS The estimated 2-year cumulative incidence of relapse in the G-Dec group was 15.0% (95% CI, 8.0% to 22.1%), compared with 38.3% (95% CI, 28.8% to 47.9%) in the non-G-Dec group (P = .01), with a hazard ratio (HR) of 0.32 (95% CI, 0.18 to 0.57; P < .01). There was no statistically significant difference between the G-Dec and non-G-Dec groups in the 2-year cumulative incidence of cGVHD without relapse (23.0% [95% CI, 14.7% to 31.3%]) and 21.7% [95% CI, 13.6% to 29.7%], respectively; P = .82), with an HR of 1.07 (95% CI, 0.60 to 1.92; P = .81). After rhG-CSF combined with minimal-dose Dec maintenance, increasing numbers of natural killer, CD8 T, and regulatory T cells were observed.

CONCLUSION Our findings suggest that rhG-CSF combined with minimal-dose Dec maintenance after allo-HSCT can reduce the incidence of relapse, accompanied by changes in the number of lymphocyte subtypes.

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Introduction

The prognosis of patients with high-risk (HR) acute myeloid leukemia (AML) continues to worsen after treatment. One potentially curative regimen for HR-AML is allogeneic hematopoietic stem-cell transplantation (allo-HSCT). Over the past decade, although considerable advances have been achieved in therapeutic approaches for allo-HSCT, the best survival rate among adults with HR-AML who have undergone allo-HSCT after achieving complete remission (CR) is only approximately 55%, and relapse remains the leading cause of death in patients after allo-HSCT, accounting for 20%-50% of deaths. Donor lymphocyte infusion (DLI) is one of the most common interventions for AML relapse because donor lymphocytes are expected to promote the graft-versus-leukemia (GVL) effect. However, DLI treatment success in AML relapse has been limited, and the reported overall survival (OS) rates at 3 years are only 10%-20%. Accordingly, the development of novel regimens to prevent leukemia relapse remains the highest priority in the treatment of HR-AML after allo-HSCT.

Decitabine (Dec) is a hypomethylating agent that is currently approved for the treatment of high-risk
myelodysplastic syndrome, chronic myelomonocytic leukemia, and AML. Recently, retrospective case series studies and small-sample, prospective, single-arm studies have found that the primary effects of hypomethylating treatment for the prevention of AML relapse after allo-HSCT consist of an increase in the number of T regulatory (Treg) cells and the induction of cytotoxic CD8+ T-cell responsiveness to tumor antigens. Furthermore, these studies explored the appropriate doses and optimal treatment courses of hypomethylating agents. However, further optimization of the anti-AML effect while reducing the hematologic toxicities of hypomethylating agents must still be addressed.

Importantly, the demethylating effect of Dec has been shown to be cyclin dependent, indicating that the combination of Dec with an agent that promotes cell cycle entry could synergistically promote the elimination of AML and prevent relapse. Notably, granulocyte colony-stimulating factor (G-CSF) is a soluble growth factor that interacts with the G-CSF receptor to promote cell entry into the cell cycle. Moreover, G-CSF and Dec have been shown to promote the production and function of cytotoxic CD8+ T cells, natural killer (NK) cells, and Treg cells, which may promote the GVL effect and reduce graft-versus-host disease (GVHD). We hypothesized that recombinant human G-CSF (rhG-CSF) followed by minimal-dose Dec could effectively prevent AML relapse. To test this hypothesis, we conducted a multicenter, open-label, randomized controlled study to investigate the efficacy, safety, and tolerability of G-CSF combined with minimal-dose Dec for prophylaxis against relapse in patients who underwent allo-HSCT.

PATIENTS AND METHODS

Study Design and Participants

This study was a phase II, open-label, multicenter, randomized controlled trial (ChiCTR-IIR-16008182) conducted by the 12 hospitals in Hematopoietic Stem Cell Transplantation–Western China Group. The protocol and all amendments were approved by the China Registered Clinical Laboratory Ethics Committee and the institutional review boards at each of the 12 participating institutions. All of the patients or their legal guardians signed informed consent forms in accordance with the Declaration of Helsinki.

Eligible patients with HR-AML included in the trial were categorized as having AML with poor genetic abnormalities, primary refractory AML, relapsed AML, or secondary AML. Eligible patients met the following criteria: established stable donor hematopoiesis; CR and minimal residual disease (MRD) negative; no acute GVHD (aGVHD) or controlled aGVHD for > 60 days after transplantation; and no uncontrolled infections or severe liver, renal, lung, or heart disease. The exclusion criteria were serious organ dysfunction, leukemia combined with other cancers, active systemic infection, GVHD requiring treatment, inability to schedule follow-up, brain dysfunction or severe mental illness preventing compliance with the study protocol, and hypersensitivity to any components of rhG-CSF and/or Dec.

Sample Size and Power Analysis

Our primary outcome variable was the cumulative incidence of relapse. The sample size was determined using the log-rank test comparing two cumulative incidences. In the preliminary research, the incidences of relapse in the patients who received rhG-CSF plus Dec (G-Dec group) and the patients who received no treatment (non-G-Dec group) were 12.6% and 32.1%, respectively. We assumed 1 year of enrollment and 2 years of follow-up. With a power of 90% and a two-sided test of \( \alpha = 0.05 \) type I error, a total sample size of 204 patients would be needed, with 102 patients in each arm. With this sample size, if there were no deaths without relapses, 46 relapses would be observed, obtaining a 90% power to test a treatment difference between 67.9% and 87.4% with a hazard ratio (HR) of 0.35. See Data Supplement (online only) for details.
Randomization and Procedure

A stratified block randomization method was used in this study. At each research center, consenting eligible participants were stratified by disease status before HSCT (two levels: CR and MRD negative [CRMRD−] and CR and MRD positive [CRMRD+]/partial remission [PR]/no remission [NR]) and randomly assigned in a 1:1 ratio to the G-Dec group or non–G-Dec group using a computer-generated block randomization schedule. The block size was randomly generated among the numbers 4, 6, 8, and 10.

In the G-Dec group, patients received six courses of rhG-CSF combined with minimal-dose Dec (rhG-CSF 100 μg/m² by subcutaneous injection on days 0-5, and Dec at 5 mg/m² by infusion on days 1-5). No treatment was given to the patients in non–G-Dec group. rhG-CSF and Dec were administered every 6-8 weeks for up to six courses. The maintenance treatment was discontinued if leukemia relapsed, severe chronic GVHD (cGVHD) occurred, or severe adverse effects of the infusion occurred or upon completion of six cycles of infusion. The observation period extended from enrollment to March 21, 2019 or to the time of the patient’s death, whichever occurred first. Both patients and clinicians were aware of the treatment allocation, but the outcome assessors were blinded to the treatment allocation when assessing all primary and secondary end points.

Statistical Analysis

The primary end point was cumulative incidence of relapse. Secondary end points included the cumulative incidence of cGVHD, cGVHD without relapse, and transplantation-related mortality (TRM); safety of rhG-CSF combined with Dec; OS; and leukemia-free survival (LFS). Exploratory end points were the cell numbers of T, B, NK, and Treg cell subsets during the G-Dec treatment courses. The Mann-Whitney U test, χ² test, and Fisher’s exact probability test were used to compare the baseline characteristics between the G-Dec and non–G-Dec groups. The competing risk model (Fine and Gray model) was used to estimate 2-year cumulative incidences and HRs with 95% CIs of relapse, cGVHD, and cGVHD without relapse. The Kaplan-Meier method, the log-rank test, and Cox proportional hazard models were used to analyze OS and LFS. Subgroups analysis and interaction P values are shown in a forest plot. In the analysis of T, B, NK, and Treg cell subsets, an analysis of

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**FIG 1.** Flowchart of the study participants. (*) Observation was stopped according to the visit schedule, and only disease status and survival of the patients were followed. cGVHD, chronic graft-versus-host disease; Dec, decitabine; rhG-CSF, recombinant human granulocyte colony-stimulating factor.
| Variable                                      | G-Dec     | Non-G-Dec  | P    |
|-----------------------------------------------|-----------|------------|------|
| No. of patients                               | 100       | 102        |      |
| Age, years                                    | .62a      |            |      |
| Median age (range)                            | 30 (3-62) | 28 (2-52)  |      |
| < 20                                          | 25 (25.0) | 27 (26.5)  |      |
| 20-40                                         | 52 (52.0) | 55 (53.9)  |      |
| > 40                                          | 23 (23.0) | 20 (19.6)  |      |
| Patient sex                                   | .58b      |            |      |
| Male                                          | 56 (56.0) | 61 (59.8)  |      |
| Female                                        | 44 (44.0) | 41 (40.2)  |      |
| Prognostic risk category                       | .48a      |            |      |
| Favorable risk                                | 7 (7.0)   | 3 (2.9)    |      |
| Intermediate risk                             | 19 (19.0) | 20 (19.6)  |      |
| Poor risk                                     | 74 (74.0) | 79 (77.5)  |      |
| Disease status before HSCT                    | .62b      |            |      |
| CRMRD+                                        | 68 (68.0) | 68 (66.7)  |      |
| CRMRD-                                        | 24 (24.0) | 29 (28.4)  |      |
| PR                                            | 1 (1.0)   | 0 (0)      |      |
| NR                                            | 7 (7.0)   | 5 (4.9)    |      |
| Donor sex                                     | .98b      |            |      |
| Male                                          | 58 (58.0) | 59 (57.8)  |      |
| Female                                        | 42 (42.0) | 43 (42.2)  |      |
| Donor/patient sex combination                 | .58b      |            |      |
| Male for male                                 | 36 (36.0) | 33 (32.4)  |      |
| Male for female                               | 22 (22.0) | 25 (24.5)  |      |
| Female for male                               | 20 (20.0) | 27 (26.5)  |      |
| Female for female                             | 22 (22.0) | 17 (16.7)  |      |
| Transplantation schemes                       | .14b      |            |      |
| HLA-matched sibling                           | 20 (20.0) | 16 (15.7)  |      |
| HLA-matched unrelated                         | 5 (5.0)   | 13 (12.7)  |      |
| HLA-haplo                                     | 75 (75.0) | 73 (71.6)  |      |
| Conditioning regimen                          | .08b      |            |      |
| BU+CY                                         | 21 (21.0) | 16 (15.7)  |      |
| BU+CY+ATG                                     | 5 (5.0)   | 14 (13.7)  |      |
| ACNU+Ara-c+BU+CY+ATG                          | 74 (74.0) | 72 (70.6)  |      |
| CD34+ cells, 10^6/kg, median (range)          | 8.175 (3.56-26.70) | 8.33 (2.94-13.00) | .83a |
| ANC engraftment, days, median (range)          | 14 (10-26) | 14 (10-55) | .94a |
| PLT engraftment, days, median (range)          | 16 (11-48) | 16 (10-40) | .90a |
| Acute GVHD                                    | .76a      |            |      |
| Non                                           | 67 (67.0) | 70 (68.6)  |      |
| I                                              | 12 (12.0) | 13 (12.7)  |      |
| II                                             | 21 (21.0) | 19 (18.6)  |      |

NOTE. Data presented as No. (%) unless otherwise indicated.

*P from Mann-Whitney test

bP from Chi-square test

cAccording to the NCCN guidelines (2015)
variance of repeated measurement data and t test with Bonferroni corrections were used to assess the differences in the repeated measurement data based on lymphocyte numbers during the G-Dec treatment courses. For details of the statistical analysis, see the Data Supplement.

All reported P values are two-sided. The sample size and power calculations were performed using PASS software (version 11; NCSS Statistical Software, Kaysville, UT). The statistical analyses were performed using IBM SPSS statistics software version 22 (IBM China, Beijing, China) and the R software package version 2.5.0 (http://www.r-project.org).

RESULTS
A total of 220 patients from 12 transplantation centers were included in the clinical study from April 5, 2016 to January 16, 2017 (Fig 1). The patient characteristics are provided in Table 1, and other patient characteristics are described in the Data Supplement.

The median follow-up time was 28.0 months (range, 2.8-35.9 months) for the G-Dec group and 26.4 months (range, 2.9-35.7 months) for the non–G-Dec group. We found that relapse occurred in 15 (15.0%) of 100 patients in the G-Dec group and 39 (38.2%) of 102 patients in the control group; the estimated 2-year cumulative incidence rates of relapse were 15.0% (95% CI, 8.0% to 22.1%) and 38.3% (95% CI, 28.8% to 47.9%), respectively, with an HR of 0.32 (95% CI, 0.18 to 0.57; \( P < .01 \); Fig 2A). Both in the univariable competing model and the multivariable model after adjustment for possible confounders, treatment and disease status before HSCT were screened out as influencing.

FIG 2. Cumulative incidence of (A) relapse, (B) total chronic graft-versus-host disease (cGVHD), and (C) cGVHD without relapse. G-Dec, recombinant human granulocyte colony-stimulating factor plus decitabine; HR, hazard ratio.
factors of relapse. The HRs were 0.31 (95% CI, 0.18 to 0.56; $P < .01$) and 2.77 (95% CI, 1.60 to 4.79; $P < .01$) in the multivariable model (Data Supplement). We also performed subgroup analyses based on different clinical characteristics. Among the patients with CRMRD, the 2-year cumulative incidence of relapse in the G-Dec group was lower than that in the non-G-Dec group (5.9% [95% CI, 0.2% to 11.6%] vs. 31.0% [95% CI, 19.9% to 42.1%], respectively; $P < .01$), with an HR of 0.16 (95% CI, 0.06 to 0.48; $P < .01$). For patients with CRMRD–/PR/NR before transplantation, G-Dec maintenance therapy showed a tendency to reduce the 2-year cumulative incidence of relapse after transplantation compared with no G-Dec (34.5% [95% CI, 17.7% to 51.4%] vs. 52.9% [95% CI, 35.8% to 70.1%], respectively; $P = .05$), with an HR of 0.48 (95% CI, 0.23 to 0.99; $P = .05$). There was no interaction between treatment and disease status before HSCT ($P = .10$). The other clinical characteristics also showed no interaction with the treatment (Appendix Fig A1, online only).

All patients who survived longer than 100 days after HSCT were evaluated to determine the incidence of cGVHD. In the G-Dec group, cGVHD developed in 34 patients (34.0%) as follows: 30 (30.0%) exhibited mild or moderate cGVHD, whereas four (4.0%) exhibited severe cGVHD. In the non-G-Dec group, cGVHD occurred in 49 patients (48.0%) as follows: 45 (44.1%) exhibited mild or moderate cGVHD, and four (3.9%) exhibited severe cGVHD. The 2-year cumulative incidence rates of cGVHD in the G-Dec and non-G-Dec groups were 34.0% (95% CI, 24.7% to 43.3%) and 48.2% (95% CI, 38.4% to 58.0%), respectively (HR, 0.62; 95% CI, 0.40 to 0.96; $P = .03$; Data Supplement and Fig 2B). Eleven patients in the G-Dec group and 27 patients in the non-G-Dec group developed cGVHD as a result of the discontinuation of immunosuppressants and DLI during recurrence of the disease. Accordingly, no difference was found between the two groups in the incidence of cGVHD without relapse intervention (23.0% [95% CI, 14.7% to 31.3%] in G-Dec group vs. 21.7% [95% CI, 13.6% to 29.7%] in non-G-Dec group; HR, 1.07 [95% CI, 0.6 to 1.92]; $P = .81$; Data Supplement and Fig 2C). The clinical manifestations of cGVHD are described in the Data Supplement.

Before the study, 20 patients were randomly identified in each group to examine the effect of the G-Dec treatment on lymphocyte subpopulations. The baseline characteristics and representative analysis of 20 patients in each group are provided in the Data Supplement. We monitored the immune cell subtypes, including T cells, B cells, NK cells, and Treg lymphocytes. No significant changes were observed in the numbers of B cells or CD4+ T cells in either the G-Dec group or the non-G-Dec group ($P > .05$, Figs 3A and 3B). In contrast, the numbers of CD8+ T, NK, and Treg cells increased by the second to third course of G-Dec treatment ($P < .05$, Figs 3C-3E). Next, for 20 patients in each group, the numbers of CD8+ T, NK, and Treg cells at the end of the last course of G-Dec treatment were used as covariates in a competing risk model to assess the association between increased effector cell numbers and relapse. The univariable competing model revealed that the increase in CD8+ T, NK, and Treg cells reduced the risk of relapse, and their HRs were 0.99 (95% CI, 0.98 to 1.00; $P = .44$), 0.97 (95% CI, 0.95 to 0.99; $P < .01$), and 0.96 (95% CI, 0.92 to 1.00; $P = .44$), respectively. In the multivariable model, the number of NK cells was found to be an independent factor influencing relapse, with an HR of 0.96 (95% CI, 0.94 to 0.99; $P < .01$; Data Supplement).

Eighty-three patients in the G-Dec group and 70 patients in the non-G-Dec group were still alive on January 16, 2019. The 2-year cumulative incidence rates of TRM in the G-Dec and non-G-Dec groups were 3.4% (95% CI, 1.1% to 10.3%) and 1.6% (95% CI, 0.2% to 11.1%), respectively, with an HR of 2.4 (95% CI, 0.25 to 22.97; $P = .44$). The causes of TRM included severe infections in three patients and recurrent episodes of serious GVHD in two patients. The 2-year rates of LFS in the G-Dec and non-G-Dec groups were 81.9% (95% CI, 72.8% to 88.2%) and 60.7% (95% CI, 50.5% to 69.4%), respectively, with an HR of 0.38 (95% CI, 0.22 to 0.66; $P < .01$; Fig 4A). The 2-year rates of OS in the G-Dec and non-G-Dec groups were 85.8% (95% CI, 77.1% to 91.3%) and 69.7% (95% CI, 59.6% to 77.8%), respectively, with an HR of 0.45 (95% CI, 0.24 to 0.83; $P = .01$; Fig 4B). Adverse events of any grade regardless of attribution to a trial regimen by an investigator occurred in 93.0% of the patients in the G-Dec group and 83.3% of the patients in the non-G-Dec group. The most common treatment-emergent adverse event was hypoleukocytosis, which mainly occurred during the first two courses of G-Dec treatment (Data Supplement). The nonhematologic toxicities included nausea, vomiting, diarrhea, peripheral edema, abnormal liver function, and abnormal renal function. Most adverse events were grade 1 or 2 and improved after symptomatic treatment (Table 2). No deaths resulted from lethal organ toxicities as a result of G-Dec maintenance therapy in the study.

**DISCUSSION**

In this study, we investigated rhG-CSF combined with minimal-dose Dec as a treatment strategy for HR-AML relapse prophylaxis. The incidence of relapse in the G-Dec group was significantly lower than that in the non-G-Dec group. In addition, the relapse rate in patients with CRMRD– before transplantation was significantly decreased after rhG-CSF combined with minimal-dose Dec treatment, indicating that maintenance treatment has a greater benefit for patients with HR-AML who are MRD negative before transplantation.

Selecting the optimal dose is important when administering hypomethylating agents after allo-HSCT. In our study, approximately 97.0% of patients tolerated at least four
cycles, and 96.0% of patients completed all six cycles. Although the main dose-limiting toxicity was myelosuppression, only 13.0% of the patients (13 of 100 patients) experienced grade 3 or 4 neutropenia or thrombocytopenia requiring rhG-CSF or recombinant human thrombopoietin support, and 16.0% of patients (16 of 100 patients) experienced grade 3 or 4 anemia requiring an RBC infusion.

Both our study design and findings differ from those of previous studies using hypomethylation agent maintenance therapy after allo-HSCT. We found that rhG-CSF combined with minimal-dose Dec treatment demonstrated a prominent advantage in the prevention of AML recurrence. rhG-CSF is known to stimulate the production, maturation, and effector functions of granulocytes and

FIG 3. Changes in the mean absolute counts of lymphocyte subsets after recombinant human granulocyte colony-stimulating factor (rhG-CSF) plus decitabine (Dec) maintenance treatment (G-Dec group). The number of patients in both the G-Dec group and the non–G-Dec (no treatment) control group was 20. (A) Natural killer (NK) cells. (B) CD19+ B cells. (C) CD3+CD4+ T cells. (D) CD3+CD8+ T cells. (E) CD4+CD25+FOXP3+ T cells. (*) After Bonferroni correction, $P$ < .001 vs non–G-Dec group. C, cycle; C1D1, first day in the first cycle before medication; D, day; K, thousand; Treg, regulatory T cell.
co-stimulate early progenitor cells synergistically with several other cytokines. Although the use of rhG-CSF in AML has been controversial because it stimulates the in vitro proliferation of leukemic blast cells from most patients with AML, a large number of clinical studies show that rhG-CSF does not reduce the remission rate of AML chemotherapy. Furthermore, rhG-CSF combined with Dec and chemotherapy have been proven to improve

![Graph A](image1)

**FIG 4.** (A) Leukemia-free survival and (B) overall survival among the patients in the two groups. G-Dec, recombinant human granulocyte colony-stimulating factor plus decitabine; HR, hazard ratio.

### TABLE 2. Adverse Events of Any Cause in the Two Groups

| Event                                           | Any Grade (G-Dec Group n = 100) | Grade ≥ 3 (G-Dec Group n = 100) | Any Grade (Non-G-Dec Group n = 102) | Grade ≥ 3 (Non-G-Dec Group n = 102) |
|------------------------------------------------|---------------------------------|---------------------------------|-------------------------------------|-------------------------------------|
| Any adverse event                               | 93 (93.0)                      | 38 (38.0)                      | 85 (83.3)                           | 45 (44.1)                           |
| Neutropenia                                     | 78 (78.0)                      | 8 (8.0)                        | 48 (47.1)                           | 7 (6.9)                             |
| Anemia                                          | 70 (70.0)                      | 18 (18.0)                      | 65 (63.7)                           | 23 (22.5)                           |
| Decreased appetite                              | 63 (63.0)                      | 8 (8.0)                        | 70 (68.6)                           | 12 (11.8)                           |
| Thrombocytopenia                                | 42 (42.0)                      | 15 (15.0)                      | 45 (44.1)                           | 10 (9.8)                            |
| Increased blood uric acid level                 | 38 (38.0)                      | 3 (3.0)                        | 32 (31.4)                           | 2 (2.0)                             |
| Nausea                                          | 35 (35.0)                      | 2 (2.0)                        | 30 (29.4)                           | 5 (4.9)                             |
| Vomiting                                        | 22 (22.0)                      | 3 (3.0)                        | 27 (26.5)                           | 5 (4.9)                             |
| Diarrhea                                        | 18 (18.0)                      | 3 (3.0)                        | 22 (21.6)                           | 2 (2.0)                             |
| Peripheral edema                                | 17 (17.0)                      | 0 (0.0)                        | 12 (11.8)                           | 0 (0.0)                             |
| Increased γ-glutamyl transferase level          | 15 (15.0)                      | 5 (5.0)                        | 18 (17.6)                           | 3 (2.9)                             |
| Constipation                                     | 13 (13.0)                      | 0 (0.0)                        | 8 (7.8)                             | 0 (0.0)                             |
| Increased alanine aminotransferase level         | 12 (12.0)                      | 0 (0.0)                        | 12 (11.8)                           | 0 (0.0)                             |
| Rash                                            | 12 (12.0)                      | 0 (0.0)                        | 15 (14.7)                           | 3 (2.9)                             |
| Increased blood creatinine level                | 12 (12.0)                      | 0 (0.0)                        | 13 (12.7)                           | 0 (0.0)                             |
| Epistaxis                                       | 8 (8.0)                        | 0 (0.0)                        | 12 (11.7)                           | 0 (0.0)                             |
| Pleural effusion                                | 5 (5.0)                        | 0 (0.0)                        | 8 (7.8)                             | 0 (0.0)                             |

**NOTE.** Data presented as No. (%). All adverse events that occurred during the trial period are listed regardless of the attribution to any trial regimen by an investigator. The G-Dec population included all patients who underwent randomization and received at least one cycle of the assigned combination treatment.
the remission rate in elderly or refractory patients with AML.\textsuperscript{26,27} Interestingly, Xiong et al\textsuperscript{28} found that NK cell populations were expanded in G-CSF–mobilized sources and could also exhibit improved functionality, demonstrating increased GVL capacity. In addition, Dec can enhance NK cell–mediated antibody-dependent cellular cytotoxicity against AML blasts and decrease relapse in patients with AML.\textsuperscript{29} In our study, the number of NK cells was increased after rhG-CSF combined with Dec treatment. Furthermore, the number of NK cells was found to be an independent factor influencing relapse; with an increasing number of NK cells, the risk of relapse was reduced. This finding suggests that the G-Dec group had a reduced cumulative incidence of relapse as a result of an increase in NK cells. Concomitantly, we found that the number of CD8\textsuperscript{+} T cells was also increased in patients who received the G-Dec treatment compared with controls. Recently, Ghoneim H.E. et al\textsuperscript{30} reported that blocking de novo DNA methylation with Dec in activated CD8\textsuperscript{+} T cells promoted their retention of effector function and inhibited exhaustion. Furthermore, the GVL effects exerted by donor T cells against leukemic-associated antigens also play a crucial role in disease eradication and relapse prevention.\textsuperscript{31,32} Therefore, we postulate that the effect of G-CSF combined with the Dec maintenance regimen to increase the numbers of NK and CD8\textsuperscript{+} T cells might be the primary mechanism that reduces relapse in patients with HR-AML after allo-HSCT. In the current study, G-CSF therapy combined with a Dec maintenance regimen did not increase the incidence of cGVHD in patients with HR-AML after allo-HSCT. The lymphocyte subset analysis showed that the number of Treg cells significantly increased after the second cycle of treatment. This finding is in agreement with preclinical animal models, in which Treg cells have been shown to suppress GVHD without decreasing the GVL effect.\textsuperscript{33} Dec and azacytidine (another demethylation drug) have been shown to expand immunomodulatory Treg cells in animal models and in phase I and II clinical trials.\textsuperscript{34} Recently, in vitro experiments have shown that the exposure of T cells to azacytidine leads to demethylation of the FOXP3 promoter, FOXP3 overexpression, and expansion of Treg cells.\textsuperscript{35} Moreover, G-CSF can promote the expansion and function of Treg cells without diminishing their function, cytokine profiles, or phenotypic characteristics.\textsuperscript{36-38}

In terms of limitations, the small sample size in some subgroups might have resulted in low test power, and the open-label design might have exaggerated the treatment effect in the G-Dec group. The results require validation in a multicenter, randomized, controlled, double-blind trial with a larger sample size. In addition, only 20 patients were randomly identified in each group to examine the effect of G-Dec treatment on lymphocyte subpopulations and the relationship between lymphocyte subpopulations and relapse. The representativeness of 20 patients is relatively limited. Further studies of lymphocyte subpopulations and functional studies should be performed to show the allorreactivity or anti-AML efficacy in the entire cohort.

Altogether, to our knowledge, our findings demonstrate that rhG-CSF combined with minimal-dose Dec treatment after allo-HSCT could significantly reduce the incidence of HR-AML relapse. The reduction in relapse is associated with increased numbers of NK and CD8\textsuperscript{+} T cells, which likely promote the GVL effect, and increased numbers of Treg cells, which likely control the development of GVHD. Future studies are required to investigate combinations of other drugs or treatment regimens with minimal-dose Dec for AML relapse prophylaxis in patients who are MRD positive before transplantation.

\textbf{AFFILIATIONS}

\textsuperscript{1}Medical Center of Hematology, Xinqiao Hospital, Army Medical University, Chongqing, China
\textsuperscript{2}Department of Health Statistics, College of Military Preventive Medicine, Army Medical University, Chongqing, China
\textsuperscript{3}Department of Hematology, General Hospital of Kunming Military Region of the People's Liberation Army (PLA), Kunming, China
\textsuperscript{4}Department of Hematology, General Hospital of Chengdu Military Region of the PLA, Chengdu, China
\textsuperscript{5}Department of Hematology, Ruijin Hospital, Shanghai Jiao Tong University, Shanghai, China
\textsuperscript{6}Department of Hematology, the Affiliated Hospital of Xinjiang Medical University, Urumqi, China
\textsuperscript{7}Department of Hematology, General Hospital of Lanzhou Military Region of the PLA, Lanzhou, China
\textsuperscript{8}Department of Hematology, Xinjiang Provincial People's Hospital, Urumqi, China
\textsuperscript{9}Department of Hematology, Affiliated Hospital of Guiyang Medical University, Guiyang, China
\textsuperscript{10}Department of Hematology, Tangdu Hospital, Forth Military Medical University (Air Force Medical University), Xi'an, China
\textsuperscript{11}Department of Hematology, Yunnan Provincial People's Hospital, Kunming, China

\textbf{CORRESPONDING AUTHOR}

Xi Zhang, MD, PhD, Medical Center of Hematology, Xinqiao Hospital, The Army Medical University, Chongqing 400037, China; e-mail: zhangxxi@sina.com.

\textbf{EQUAL CONTRIBUTION}

Lei Gao, Yanqi Zhang, and Sanbin Wang contributed equally to this work.

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AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Effect of rhG-CSF Combined With Decitabine Prophylaxis on Relapse of Patients With High-Risk MRD-Negative AML After HSCT: An Open-Label, Multicenter, Randomized Controlled Trial

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Kaniel Cassady
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### FIG A1.
Forest plot summarizing hazard ratios for the G-Dec group versus the non-G-Dec group in the subgroup analyses with a test for interaction. ACNU, nimustine; Ara-C, cytarabine; ATG, thymoglobulin; BU, busulfan; CRMRD, complete remission and minimal residual disease negative; CY, cyclophosphamide; HSCT, hematopoietic stem-cell transplantation.

| Subgroup                                      | No. of Patients | Hazard Ratio (95% CI) | P   |
|-----------------------------------------------|-----------------|-----------------------|-----|
| **Age, years**                                |                 |                       |     |
| < 20                                          | 52              | 0.37 (0.12 to 1.19)   | .38 |
| 20-40                                         | 107             | 0.40 (0.18 to 0.85)   |     |
| > 40                                          | 43              | 0.15 (0.03 to 0.63)   |     |
| **Patient sex**                               |                 |                       | .21 |
| Male                                          | 117             | 0.21 (0.09 to 0.50)   |     |
| Female                                        | 85              | 0.48 (0.21 to 1.08)   |     |
| **Prognostic risk category**                  |                 |                       | .40 |
| Favorable                                     | 10              | 0.13 (0.01 to 1.14)   |     |
| Intermediate                                  | 39              | 0.39 (0.08 to 1.94)   |     |
| Poor                                          | 153             | 0.32 (0.17 to 0.61)   |     |
| **Disease status before HSCT**                |                 |                       | .10 |
| CRMRD                                         | 136             | 0.16 (0.06 to 0.48)   |     |
| Non-CRMRD                                     | 66              | 0.48 (0.23 to 0.99)   |     |
| **Donor sex**                                 |                 |                       | .62 |
| Male                                          | 117             | 0.36 (0.18 to 0.75)   |     |
| Female                                        | 85              | 0.26 (0.10 to 0.68)   |     |
| **Donor/patient sex combination**             |                 |                       | .34 |
| Male for male                                 | 69              | 0.25 (0.09 to 0.67)   |     |
| Male for female                               | 47              | 0.56 (0.19 to 1.61)   |     |
| Female for male                               | 47              | 0.12 (0.02 to 0.91)   |     |
| Female for female                             | 39              | 0.36 (0.11 to 1.18)   |     |
| **Transplantation schemes**                   |                 |                       | .62 |
| HLA-matched sibling                           | 36              | 0.33 (0.17 to 0.64)   |     |
| HLA-matched unrelated                         | 18              | 0.49 (0.06 to 4.26)   |     |
| HLA haplo                                     | 148             | 0.23 (0.05 to 1.10)   |     |
| **Conditioning regimen**                      |                 |                       | .55 |
| BU + CY                                       | 37              | 0.21 (0.04 to 1.04)   |     |
| BU + CY + ATG                                 | 19              | 0.41 (0.05 to 3.42)   |     |
| ACNU + Ara-c + BU + CY + ATG                  | 146             | 0.35 (0.18 to 0.67)   |     |
| **Acute GVHD**                                |                 |                       | .61 |
| No                                            | 137             | 0.35 (0.18 to 0.68)   |     |
| Yes                                           | 65              | 0.24 (0.07 to 0.81)   |     |