The Roles of CCR9/CCL25 in Inflammation and Inflammation-Associated Diseases

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Chemokine is a structure-related protein with a relatively small molecular weight, which can target cells to chemotaxis and promote inflammatory response. Inflammation plays an important role in aging. C-C chemokine receptor 9 (CCR9) and its ligand C-C chemokine ligand 25 (CCL25) are involved in the regulating the occurrence and development of various diseases, which has become a research hotspot. Early research analysis of CCR9-deficient mouse models also confirmed various physiological functions of this chemokine in inflammatory responses. Moreover, CCR9/CCL25 has been shown to play an important role in a variety of inflammation-related diseases, such as cardiovascular disease (CVD), rheumatoid arthritis, hepatitis, inflammatory bowel disease, asthma, etc. Therefore, the purpose of this review gives an overview of the recent advances in understanding the roles of CCR9/CCL25 in inflammation and inflammation-associated diseases, which will contribute to the design of future experimental studies on the potential of CCR9/CCL25 and advance the research of CCR9/CCL25 as pharmacological inflammatory targets.

Keywords: chemokine, CCR9, CCL25, autoimmune, inflammatory disease

INTRODUCTION

Chemokines are a large family of small cytokines and generally have low molecular weight ranging from 7 to 15 kDa (Palomino and Marti, 2015). Chemokines can interact with different chemokine receptors that control the residence and migration of immune cells. When the body is stimulated by antigen, chemokines can be secreted by various cells such as endothelial and dendritic cells (DCs) (Vassilatis et al., 2015). Currently, more than 50 chemokines have been discovered, which can be divided into four categories according to the position of two cysteines at the N-terminus: C subfamily, CC subfamily, CXC subfamily, and CX3C subfamily (Figure 1; Bleul et al., 1996). Moreover, according to the expression of chemokines and their functions in the immune system, chemokines can be also divided into homeostatic and inflammatory chemokines (Table 1; Moser et al., 2004). Chemokines expression is regulated via a variety of factors such as interleukin (IL)-10 and glucocorticoids, which can inhibit the secretion of chemokines, while IL-1 and tumor necrosis factor (TNF)-α can increase chemokines expression (Cocchi et al., 1995; Oberlin et al., 1996;
Zlotnik and Yoshie, 2012). The chemokine receptor is a G-protein coupled receptor of the seven-transmembrane receptor family, which contains seven transmembrane segments and is widely expressed on white blood cells surface derived from bone marrow such as neutrophils and macrophages (Schulz et al., 2016). In addition, chemokine receptors have also been shown to be expressed in epithelial cells, vascular endothelial cells, and other tissue cells. The combination of chemokines and their corresponding receptors can involve in normal body development, and also participate in various pathological and physiological processes such as leukocyte activation, angiogenesis, aging, breast cancer resistance, and autoimmune diseases (Zhong et al., 2015).

C-C chemokine ligand 25 (CCL25) is also considered a thymus expressed chemokine, and the gene is located on chromosome 19p13.2 (Figure 1; Qiup ing et al., 2004). Tu et al. (2016) confirmed that CCL25 is mainly expressed in the thymus and intestinal epithelium, and also produced by vascular endothelial cells and other parenchymal cells, which can migrate immature T cells into the thymus to mature and release (Table 1). C-C chemokine receptor 9 (CCR9) was discovered in 1996 as a G protein-coupled receptors expressed on the cell membrane of dendritic cell, neutrophils, lymphocytes, monocyte macrophages, and vascular endothelial cells (Yu and Stamenkovic, 2000). However, in the 3 years after the discovery of CCR9, its ligand has not been found, so it was once called "orphan receptor." It was not until 1999 that CCL25 was found to be one-to-one combined with CCR9 (Zaballos et al., 1999). CCR9 belongs to the β-chemokine receptor family, and its gene is located on chromosome 3 p21.31 (Tu et al., 2016). CCR9 is mainly distributed in immature T lymphocytes and intestinal cells surface, and after binding to its specific ligands, plays a key role in T lymphocyte development and tissue-specific homing (Wermers et al., 2011; Wu et al., 2014). Further study found that CCL25 promoted proliferation and chemotaxis of inflammatory cells that expressed its specific receptor CCR9 (Igaki et al., 2018).

CCR9 and CCL25 are members of the CC subfamily of chemokines that are involved in a variety of inflammatory diseases and promote inflammatory responses. Over the years, the role of CCR9/CCL25 in inflammation and related diseases has become increasingly clear, including cardiovascular disease (CVD), hepatitis, arthritis (Yokoyama et al., 2014), inflammatory bowel disease (Kalindjian et al., 2016), and asthma. This study provides abundant evidence that CCR9/CCL25 can be a potential target for a variety of inflammatory diseases. Therefore, the focus of this review is to evaluate the latest findings on the role of CCR9/CCL25 in inflammatory disease. The structure and physiological function of CCR9/CCL25 were briefly introduced. The various roles of CCR9/CCL25 in the development of inflammatory disease were described (Figure 2), and some different insights were put forward concerning the CCR9/CCL25 pathway.

**CCR9 AND CCL25 EXPRESSION IN TISSUE**

Vicari et al. (1997) isolated CCL25 cDNA from the thymus of recombinase activation gene-1 (RAG-1) deficient mice and were designated thymus-expressed chemokine (TECK). Moreover, they also found that CCL25 appeared weak sequence homology to other members of the CC chemokine family and located on mouse chromosome 8 (Vicari et al., 1997). Besides the thymus, mRNA encoding CCL25 was detected at substantial levels in the small intestine and at low levels in the liver (Vicari et al., 1997; Zabel et al., 1999; Kunkel et al., 2000). However, studies found that CCL25 is highly expressed in the small intestinal epithelium and thymus, which can regulate trafficking of gut-specific memory/effector T cells via upregulation of the integrin homing receptor α47 and CCR9 (Stenstad et al., 2006; Wurbel et al., 2000, 2007). Meanwhile, the source of CCL25 in the thymus was determined to be thymic dendritic cells; in contrast, bone marrow-derived dendritic cells do not express CCL25. The murine CCL25 recombinant protein showed chemotactic activity for activated macrophages, dendritic cells, and thymocytes. These research results fully demonstrated that CCL25 represents a novel thymic dendritic cell-specific CC chemokine that is possibly involved in T cell development. Within the intestines of normal mice, CCL25 expression was highest in the proximal small intestine, lowest in the distal small bowel, and no or almost no expression in the colon (Svensson et al., 2002; Rivera-Nieves et al., 2006; Stenstad et al., 2007; Wermers et al., 2011; Wurbel et al., 2011).

In circulating white blood cells, CCR9 expression is limited to activated B cells and a certain proportion of CD4 and CD8 T cells (Zabel et al., 1999; Kunkel et al., 2000; Papadakis et al., 2000, 2001, 2003). CCR9 has been identified in plasmacytoid dendritic cells in mice, but there is a lack of relevant data in humans (Wendland et al., 2007; Hadeiba et al., 2012). Similarly, despite the fact that a few papers describe the expression of CCR9 on mouse macrophages, there is a lack of human data (Nakamoto et al., 2012). In the peripheral tissues, CCR9+ cells are mostly concentrated in colon, thymus, and small intestine (Zabel et al., 1999; Kunkel et al., 2000; Papadakis et al., 2000). Intraepithelial

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**FIGURE 1 | Structure of members of the chemokine family.**
lymphocytes are mainly CD8+, most of which express CCR9 surface (Zabel et al., 1999; Kunkel et al., 2000).

THE ROLE OF CCR9/CCL25 IN INFLAMMATORY DISEASES

Inflammation underlies many physiological and pathological processes. In particular, chronic inflammation is attributed to the pathophysiological basis of various modern diseases. Of note, inflammatory diseases are characterized by wide coverage, complex pathogenesis, and large difference in prognosis (Guo et al., 2015). Therefore, we mainly focus on the effect of CCR9/CCL25 in inflammatory diseases as well as elaborate on their mechanism of action in related diseases.

Cardiovascular Disease

Cardiovascular disease, also known as circulatory disease, is a series of diseases involving the circulatory system, including the heart and blood vessels, which have always been at the forefront of human major causes of death (Jokinen, 2015). Heart failure, cardiac hypertrophy, and atherosclerosis are belong to CVD and CXC chemokines. Moreover, according to the classification of chemokines, these four types also have different functions and localizations.

In CCR9 knockout CCR9+/+ mice, surgical ligation of the left anterior descending coronary artery caused myocardial infarction (MI), and found that the expression of CCR9 in the heart of mice was significantly up-regulated after MI. Down-regulation of CCR9 expression can improve survival rate and left ventricular dysfunction, reduce infarct size, and improve cardiac function after MI. In addition, abolish of CCR9 in the mouse MI heart significantly increased Bcl-2 expression, while the expression of Bax and cleaved caspase 3 was remarkably reduced, thereby attenuating the apoptosis of cardiomyocytes. Inflammation is considered the most vital pathological response to damage and repair, and abolish of CCR9 can decrease pro-inflammatory cytokines mRNA levels (IL-6, IL-1β, and TNF-α) and suppress the inflammatory response after MI. In addition, CCR9 is involved in structural remodeling mainly by interfering with NF-κB and MAPK signaling pathways (Figure 2). These results confirmed that CCR9/CCL25 may exert a positive role in MI (Huang et al., 2016).

Heart failure is the leading cause of death in many countries, particularly in an aging population (Kumarswamy and Thum, 2013). Currently, heart failure is widely recognized as a clinical syndrome caused by cardiac hypertrophy and remodeling that involves changes of structural and functional in the left ventricle (Zhang et al., 2014). Study found that CCR9 protein levels have significantly increased in failing human hearts and mice or cardiomyocyte hypertrophy model. The loss of CCR9 in mice can reduce the hypertrophy caused by pressure overload. The results show that heart weight/body weight, lung weight, and heart weight/tibia length ratios of CCR9-deficient mice are significantly lower than those of the control group. And aortic banding treated CCR9-deficient mice were characterized by decreased Left ventricular (LV) end diastolic and systolic diameters and increased fractional shortening, demonstrating smaller LV dimensions and elevated systolic function. Moreover, the hypertrophic response of the CCR9-deficient mice was

### TABLE 1 | Classification, structural features, function, and location of the chemokine family members.

| Family | Structural features | Function | Location | Chemokines |
|--------|---------------------|----------|----------|------------|
| CXC    | The CXC family has a single amino acid residue in between the first two cysteines. | CXC chemokines have the highest ability to attract neutrophils and monocytes. | 4q12-13 | CXCL1-8 |
|        |                     |          | 4q21,21  | CXCL9-11   |
|        |                     |          | 10q11.1  | CXCL12     |
|        |                     |          | 4q21     | CXCL13     |
|        |                     |          | 5p31     | CXCL14     |
|        |                     |          | 17p13    | CXCL1      |
| CC     | The CC chemokine family has the first two cysteine residues adjacent to each other. | CC chemokines are mainly responsible for the recruitment of lymphocytes. | 17q11.2   | CCL1-15    |
|        |                     |          | 7q11.23  | CCL16      |
|        |                     |          | 16q13    | CCL17      |
|        |                     |          | 17q11.2  | CCL18      |
|        |                     |          | 9q13     | CCL19      |
|        |                     |          | 9q13     | CCL21      |
|        |                     |          | 16q13    | CCL22, CCL23 |
|        |                     |          | 11q11.2  | CCL24      |
|        |                     |          | 7q11.23  | CCL25      |
|        |                     |          | 19p13.2  | CCL26      |
|        |                     |          | 7q11.23  | CCL27      |
|        |                     |          | 9q13     | CCL28      |
|        |                     |          | 5q       |            |
| C      | The C family there is only one cysteine residue and only one disulfide chain. | C chemokine has chemotactic activity for lymphocytes, CD4+ and CD8+ T cells, and NK cells. | 1q23      | XCL1-2     |
| CX3C   | The CX3C chemokine family three amino acid residues separate the first two cysteines. | CX3C chemokines can induce chemotaxis of monocytes and cytotoxic T cells. | 16q13     | CX3CL1     |

The spacing of two conserved cysteine residues at the N-terminus determines the nomenclature and classification of the chemokine family members; in CXC, CC, C, and CX3C chemokines.
FIGURE 2 | Schematic diagram illustrating the regulation of molecular mechanisms of CCR9/CCL25 in inflammatory diseases. CCR9/CCL25 exerts multiple effects in a variety of inflammatory diseases by regulating molecular mechanisms, such as molecules associated with apoptosis and inflammation. CCR9, C-C chemokine receptor 9; CCL25, C-C chemokine ligand 25; Bcl-2, B-cell lymphoma-2; Bax, Bcl-2 Associated X-protein; IL-6, interleukin-6; IL-1β, interleukin-1β; TNF-α, tumor necrosis factor-α; mTOR, mechanistic target of rapamycin; GSK3β, glycogen synthase kinase 3-beta; p70S6K, p70 S6 kinase; IL-10, interleukin-10; MMP3, matrix metalloproteinase-3; TNF-β, tumor necrosis factor-β; IL-17, interleukin-17; CCL11, C-C chemokine ligand 11; Th1/Th17, helper T1/helper T17; IFN-γ, interferon-gamma; IL-2, interleukin-2; IL-17A, interleukin-17A.

Remarkably attenuated compared with controls, and these animals manifested decreased heart size and cardiomyocyte cross sectional area as well as alleviation of interstitial fibrosis. The mRNA levels of hypertrophic and fibrotic markers were also reduced significantly in the CCR9 deficient mice. Notably, overexpression of CCR9 in mice reversed the result of the appeal (Xu et al., 2016). In terms of mechanism, they further found that the expression levels of MAPK lack difference between the two groups, while the phosphorylation levels of AKT/protein kinase B and downstream effectors (mTOR, GSK3b, and p70S6K) were remarkably reduced in CCR9 knockout mice and increased in CCR9 transgenic mice after aortic surgery (Figure 2). These results suggested that CCR9 could promote hypertrophy mainly due to the protein kinase B mammalian target of the rapamycin GSK-3β signaling cascade, rather than through the MAPK signaling pathway, which indicates that CCR9 can be used as a new regulator of myocardial hypertrophy and may provide a novel therapeutic target for suppressing myocardial hypertrophy in the future (Xu et al., 2016).

In conclusion, the knockout of CCR9/CCL25 serves as a novel modulator of pathological progression for inhibiting the development of MI and heart failure (Table 2). However, the role of CCR9/CCL25 in other CVD has not been reported, so further research is needed.

Hepatitis
Hepatitis usually refers to various pathogenic factors, including parasites, viruses, drugs, bacteria, and autoimmune factors, which cause damage to liver cells and functions, thereby causing a series of uncomfortable symptoms of the body and abnormal liver function indicators (Crispe, 2003; Thomson and Knolle, 2010; Linder and Malani, 2017).

As well known, various immune cells in the liver involve in the pathogenesis of liver diseases. Inflammatory macrophages play a key role in liver injury and then liver fibrosis and canceration. Nakamoto et al. (2012) previous research suggests that CCR9+ inflammatory macrophage triggers acute liver injury by interacting with helper T 1 (Th1) cells in the inflamed liver. Subsequently, they found that macrophages expressing CCR9+CD11b+ aggravated liver damage through production of inflammatory cytokines and the promotion of Th1 cell development during the concanavalin A-induced murine acute liver injury and human acute hepatitis (Nakamoto, 2016). Further analysis using liver-shielded radiation and
TABLE 2 | The role of CCR9/CCL25 in inflammatory-associated diseases.

| Inflammation disease            | Expression of CCR9 or CCL25 | Model                                                                 | Effect of CCR9/CCL25                                                                 | Outcome                                                                 | References                  |
|--------------------------------|-----------------------------|----------------------------------------------------------------------|-------------------------------------------------------------------------------------|-------------------------------------------------------------------------|-----------------------------|
| Myocardial infarction           | Increased CCR9              | Model of myocardial infarction induced by ligation of left anterior descending coronary artery of CCR9+/− and CCR9+/− mice | Down-regulation of CCR9 expression can improve survival rate, left ventricular dysfunction, cardiac function, reduce infarct size, meanwhile, significantly attenuate the apoptosis of cardiomyocytes, and inhibit the inflammatory response after myocardial infarction. | CCR9/CCL25 axis may play an important role in inflammatory cell infiltration and cardiac remodeling after myocardial infarction. | Huang et al., 2016          |
| Hepatitis                       | –                           | Concavalin A-induced murine acute liver injury and human acute hepatitis | Macrophages expressing CCR9+CD11b+ aggregated liver damage through production of inflammatory cytokines and the promotion of Th1 cell development. | Inflammatory macrophages originated from bone marrow and became locally differentiated and proliferated by interaction with hepatic stellate cells via knocking out CCR9 gene during acute liver injury. | Amiya et al., 2016; Nakamoto, 2016 |
| Rheumatoid arthritis            | Increased CCR9 and CCL25    | In the synovium of RA and non-RA patients                             | Stimulation with CCL25 increased IL-6 and MMP-3 production from RA fibroblast-like synoviocytes, and also increased IL-6 and TNF-α production from peripheral blood monocytes. Collagen-induced arthritis was suppressed in CCR9-deficient mice. | The interaction between CCL25 and CCR9 may promote cell infiltration and production of inflammatory mediator in RA synovial tissues. | Yokoyama et al., 2014       |
| Inflammatory bowel disease      | Increased CCR9 and CCL25    | DSS-induced acute colitis                                             | CCR9−/− mice are more susceptible to DSS colitis than WT littersmate controls as shown by higher mortality, increased IBD score, and delayed recovery. During recovery, the CCR9−/− colonic mucosa is characterized by the accumulation of activated macrophages and elevated levels of Th1/Th17 inflammatory cytokines. | CCR9/CCL25 interaction regulates the inflammatory immune response of the intestinal mucosa by balancing different dendritic cell subsets. | Wurbel et al., 2011         |
| Inflammatory bowel disease      | Increased CCR9 and CCL25    | DSS-induced acute colitis                                             | The mice lacking the interaction of CCR9/CCL25 showed enhanced adhesion promoting ability of cell after the transfer of naive T cells, and regulated the pro-inflammatory and anti-inflammatory subgroups of cDC. | CCR9/CCL25 interactions have protection against large intestinal inflammation in chronic colitis. | Wurbel et al., 2014         |
| Asthma                          | Increased CCL25             | OVA-induced model of allergic inflammation within CCR9 deficient mice | In challenged CCR9-deficient mice, cell recruitment was impaired at peribronchial and perivenular levels. OVA administration in CCR9-deficient mice leads to a less inflammatory cell recruitment, which modifies the expression of Il-10, CCL11, and CCL25 after OVA challenge. | CCR9 and CCL25 expressions are induced in the early stages of airway inflammation and they have an important role in modulating eosinophils and lymphocytes recruitment at the first stages of inflammatory process. | Lopez-Pacheco et al., 2016  |
| Asthma                          | Increased CCR9              | NKT cells                                                            | In patients with allergic asthma, the CCR9/CCL25 binding mechanism selectively induces chemotaxis of NKT cells. And NKT cells may be in direct contact with CD3+ T cells, polarizing them to a Th2 bias. | CCR9/CCL25 signal can interact with CD226 signaling to activate asymptomatic NKT cells, leading to airway hyperresponsiveness and inflammation, aggravating asthma. | Sen et al., 2005            |

Compelling evidence has indicated that alterations of CCR9 or CCL25 expression are also involved in inflammatory-associated diseases, including myocardial infarction, hepatitis, rheumatoid arthritis, inflammatory bowel disease, and asthma.

CCR9, C-C chemokine receptor 9; CCL25, C-C chemokine ligand 25; RA, rheumatoid arthritis; IBD, inflammatory bowel disease; DSS, dextran sulfate sodium; DCs, dendritic cells; IL-6, interleukin-6; IL-10, interleukin-10; MMP3, matrix metalloproteinase-3; TNF-α, tumor necrosis factor-α; NKT, natural killer T; DCs, dendritic cells; Th, helper T.

Bone marrow transplantation mouse models revealed that these CCR9+CD11b+ macrophages were originated from bone marrow-derived monocytes, but not liver resident macrophages. In addition, in contact with hepatic stellate cells, these CD11b+ inflammatory macrophages participated in the pathogenesis of experimental liver fibrosis by knocking out CCR9 gene. The results with further verification in human samples clarified the pathogenic role of CCR9/CCL25 axis as therapeutic target of a variety of liver diseases (Nakamoto, 2016). Similarly, Amiya et al. (2016) also demonstrated that in acute liver injury, inflammatory macrophages originated from bone marrow and became locally differentiated and proliferation through interaction with hepatic stellate cells by knocking out CCR9 gene (Figure 2).

Primary sclerosing cholangitis (PSC), as a chronic inflammatory liver disease, is characterized by progressive bile duct destruction and is also a parenteral complication of inflammatory bowel disease (IBD). In addition, PSC is a non-suppurative autoimmune granulomatous inflammation of the small bile ducts in the liver that can lead to chronic progressive cholestasis and eventually cirrhosis and liver failure (Chapman, 1991). In the healthy liver, the expression of CCL25 is very low or even undetected (Vicari et al., 1997; Zabel et al., 1999; Eksteen et al., 2004). However, in PSC, the expression of CCL25 was increased in the liver, mainly through hepatic sinusoidal endothelial cells and portal DCs expression (Eksteen et al., 2004). The hepatic inflammation in PSC was related to the increase
of CCR9 positive T cells. Other study proposed that long-lived memory gut homing cells that expressed CCR9 and α4β7 and were early activated during episodes of IBD could exacerbate PSC through interactions with hepatic endothelial that ectopically expressed CCL25 (Eksteen et al., 2004). Notably, they also demonstrated for the first time that T cells activated in the intestine can be recruited to extra-intestinal site of disease in humans and provide basic research to explain the pathogenesis of extra-intestinal complications of IBD (Eksteen et al., 2004; Table 2).

Rheumatoid Arthritis

Rheumatoid arthritis (RA) is a chronic inflammatory disease that can cause a large number of macrophages, T cells, and B cells to accumulate in the synovium (Firestein, 1991; Kinne et al., 2000), and the accumulation of these cells can further participate in the development of inflammation, joint destruction, and pain. However, biological agents (TNF blockers and IL-6 receptor antagonists) are effective in RA patients (Feldmann, 2002; Nishimoto et al., 2004; Smolen et al., 2010).

A large number of mononuclear/macrophages accumulate in the rheumatoid synovium, which play a promotive role in inflammation and joint destruction. Identifying the molecules involved in its accumulation and differentiation is essential for the development of therapeutic strategies (Mulherin et al., 1996; Haringman et al., 2005; Nishimoto and Takagi, 2010). Cluster of differentiation (CD) 14 and CD68 co-localize with CCR9 or CCL25 to identify macrophages. Other studies have confirmed that the augment CD68+ cells in the substratum of RA are one of the characteristic pathological changes of synovial membrane early RA (Singh et al., 2004). Other study demonstrated that CCR9 was expressed by PB monocytes/macrophages in RA and healthy donors, and increased in RA. Moreover, in the synovium of RA and non-RA patients, CCR9 colocalizes with CD14+ and CD68+ macrophages and was more abundant in RA synovium. Therefore, the percentage of CCR9+ monocytes in the synovium of RA (81%) is increased compared to that in blood (40%), and the percentage of CCR9+ monocytes in the synovium of non-RA (66%) is greater than that in blood (16%). Meanwhile, CCL25 could be detected in both RA and non-RA synovia, which can be co-localized within CD14+ and CD68+ cells (Schmutz et al., 2010). Furthermore, CCL25 induced stronger monocyte differentiation in RA. CCL25 induced significant chemotaxis of peripheral blood monocytes, however, which was inconsistent between different individuals. These results indicate that monocyte-induced CCR9/CCL25 expression is significantly increased in RA. CCL25 may be involved in the differentiation of monocytes to macrophages particularly in RA (Haringman et al., 2005). Moreover, CCR9 and CCL25 are expressed at higher levels in RA synovial tissues compared to osteoarthritis synovial tissues. Most CD68+ macrophages in RA synovial tissue express CCR9 and CCL25 revealed by immunohistochemical. CCR9 was expressed in macrophage, fibroblast-like synoviocytes, and DCs of synovial tissues. CCL25 stimulated the production of IL-6 and MMP-3 in RA fibroblast-like synoviocytes, and also increased the production of IL-6 and TNF-α in peripheral blood monocytes (Figure 2). Collagen-induced arthritis was inhibited in CCR9−/− mice. CCR9 antagonist (CCX8037) also suppressed collagen-induced arthritis and decreased the migration of CD11b+ splenic cells to synovial tissues. These results suggested that the interaction between CCL25 and CCR9 may promote cell infiltration and production of inflammatory mediator in RA synovial tissues. Blocking CCL25 or CCR9 may represent a strange new safety therapy for RA, and provide theoretical basis for further research a novel safe therapy for RA (Yokoyama et al., 2014; Table 2).

Inflammatory Bowel Disease

Inflammatory bowel disease is a group of chronic, non-specific inflammatory bowel diseases whose etiology has not yet been clarified, mainly including ulcerative colitis (UC) and Crohn's disease (CD). UC is a continuous inflammation of the colonic mucosa and submucosa that first affects the rectum and gradually spreads to the entire colon. CD can affect the entire digestive tract and is a discontinuous full-layer inflammation, most commonly involving terminal ileum, colon, and perianal (Loftus, 2004; Molodecky et al., 2012).

Wurbel et al. (2011) study the acute inflammation and recovery in WT and CCR9−/− mice in a model of dextran sulfate sodium (DSS)-induced colitis. The results show that CCL25 and CCR9 are both expressed in the large intestine and are upregulated during DSS colitis. CCR9−/− mice are more susceptible to DSS colitis than WT littermate controls as shown by higher mortality, increased IBD score, and delayed recovery. During recovery, the CCR9−/− colonic mucosa is characterized by the accumulation of activated macrophages and added Th1/Th17 inflammatory cytokines levels. Activated plasmacytoid dendritic cells (pDC) accumulate in the mesenteric lymph nodes of CCR9−/− animals, changing the local proportion of DC subsets. The T cells separate from these mesenteric lymph nodes were stimulated again, which were significantly higher levels of TNF-α, IFN-γ, IL-2, IL-6, and IL-17A while down-regulating IL-10 production (Figure 2). These results suggest that the CCR9/CCL25 interaction regulates the inflammatory immune response of the intestinal mucosa by balancing different dendritic cell subsets (Wurbel et al., 2011).

As mentioned before, CCR9−/− and CCL25−/− mice are more susceptible to acute DSS colitis than WT controls. As human ulcerative colitis is associated with signs of chronic colonic inflammation, they investigated whether the increased susceptibility to acute inflammation associated with defective CCL25/CCR9 interactions would also translate into increased susceptibility to chronic inflammation, and found that chronic DSS exposure results in exacerbated colitis in mice deficient for either CCR9 or CCL25 when compared with WT control mice. Although CCR9−/− T cells traffic to the colon and induce severe colitis similar to WT T cells, naive WT T cells induce more severe disease in recipient animals devoid of CCL25 expression. Moreover, compared with WT control mice, there was no significant difference in the total number of pDC and conventional dendritic cells (cDC), and the pro-inflammatory and anti-inflammatory subgroups of cDC were regulated by the CCR9/CCL25 interaction. In the end, the...
CCR9/CCL25-dependent innate immune cell lineage specificity and lineage-dependent function will be remarkably assisted via targeting the loss of CCR9/CCL25 in innate immune cell and/or epithelial compartment. These results suggested that CCR9/CCL25 interactions have protection against large intestinal inflammation in chronic colitis (Wurbel et al., 2014). Moreover, Papadakis et al. (2001) demonstrated that the proportion of peripheral blood CCR9+ CD4+ cells rose remarkably in active small bowel CD compared with normal groups, but not in patients with purely colonic Crohn’s. It is worth noting that GSK/Chemocentryx has advanced its CCR9 antagonist CCX282-B (also known as vercirnon) to a pivotal phase III clinical trial. It was unfortunate that due to poor efficacy of IBD, the clinical project was terminated at the end of 2013 (Zhang et al., 2015).

The pathophysiological characteristics of necrotizing enterocolitis (NEC) include excessive inflammation and necrosis, which can affect any part of the gastrointestinal tract (especially the small intestine) (Neu and Walker, 2011; Nino et al., 2016). The imbalance caused by the decrease in tolerogenic Foxp3+ regulatory T (Treg) cells and the increase in the production of Th17 cells in the lamina propria pro-inflammatory IL-17 leads to the NEC-induced excessive inflammatory response (Weitkamp et al., 2013; Liu et al., 2014; Egan et al., 2016). Moreover, another study has shown that in the peripheral blood of NEC patients and mice, the proportion of CCR9+ CD4+ T cells was significantly elevated. Increased CCR9+ CD4+ T cells were mainly CCR9+ IL-17-producing Treg cells, which are characteristic of common Treg cells, but their inhibitory activities were seriously damaged and passively correlated with the serious of intestinal tissue injury. Treg cells that produce CCR9+ IL-17 may be an important biomarker, improving NEC by regulating the balance of lymphocytes (Ma et al., 2019). In summary, these results demonstrated that CCR9+ IL-17-producing Treg cells could be an important biomarker, but further research is needed for clinical use (Table 2).

**Asthma**

Asthma, as a lung disease, is characterized by reversible airway obstruction, airway inflammation, and increased airway responsiveness to a variety of stimuli (Hahn, 2009; Mims, 2015). Chemokine receptors have been confirmed to be involved in leukocyte recruitment, and are closely related to asthma pathology (DeKruyff et al., 2014; Griffith et al., 2014). An ovalbumin (OVA)-induced allergic inflammation model was established in CCR9−/− mice, and the expression of CCR9 and CCL25 in eosinophils and T lymphocytes was found 6 h post-OVA challenge. Meanwhile, compared with wild-type mice, the peribronchial infiltration in CCR9-deficient mice was significantly reduced (nearly 50%), while the total number of eosinophils recruited in bronchoalveolar fluid (BALF) was decreased (30%). Moreover, in CCR9−/− mice, OVA administration can reduce the recruitment of inflammatory cell, which modifies IL-10, CCL11, and CCL25 expression at 24 h after OVA (Lopez-Pacheco et al., 2016). Interestingly, expression of TGF-β and IL-17 was elevated in OVA-stimulated CCR9-deficient mice compared to WT mice, whereas IL-10, as an anti-inflammatory chemokine, was significantly reduced (more than 60%). In addition, there was an increase in the expression of CCL25 in the WT mice as early as 6 h after ova-challenge. Meanwhile, CCL25 expression was decreased at all time test in CCR9−/− mice (Figure 2). The above results confirm that CCR9 deficiency has a positive regulatory effect on eosinophils and lymphocytes in the early stage of inflammation induction, suggesting that they may be potential targets for regulating asthma inflammation in asthma (Lopez-Pacheco et al., 2016).

Natural killer T (NKT) cells are the main operators in the development of asthma. Multiple NKT cells migrate and aggregate in the airway, producing a Th2 bias effect that directly or indirectly promotes asthma. Studies have found that the recruitment of NKT cells depends on the high expression of CCR9 and the connection of CCR9/CCL25 (DeKruyff et al., 2014; Griffith et al., 2014). In patients with allergic asthma, the CCR9/CCL25 binding method selectively induces chemotaxis of NKT cells, whereas in healthy volunteers, normal NKT cells are not able to induce. Further studies have confirmed that the pathways for NKT cells regulate the development of asthma. In the process of migrating from blood vessels to airway bronchial mucosa, NKT cells may directly contact CD3+ T cells, making them polarized toward Th2. This regulatory function depends on DC participation and CCR9/CD226 coordinated activation. The adhesion molecule CD226 is overexpressed in asthmatic NKT cells. The CCR9/CCL25 linkage can directly phosphorylate CD226, and the lack of CD226 can block the Th2 bias effect induced by NKT cells. These results indicate that CCR9/CCL25 signaling pathway could interact with CD226 signals to activate asthmatic NKT cells, leading to airway hyperresponsiveness and inflammation, aggravating asthma (Sen et al., 2005). In addition, Castan et al. (2018) demonstrated that CCR9 knock-out mice eliminated the aggravation of lung symptoms in consecutive food and respiratory allergies, which were featured by a rise in lung resistance and a higher Th17/Treg ratio in solely asthmatic-like mice. Moreover, to better understand the mechanism underlying the food allergy-induced aggravation of asthma and the role of CCR9, they performed adoptive transfer of CD4+ T cells from food-allergic mice into naïve mice, which were subsequently sensitized to house dust mites. They found that asthmatic mice received food-sensitized CD4+ T cells and had more severe inflammation than asthmatic mice that received non-sensitized CD4+ T cells. Interestingly, when mice received food allergen-sensitized lymphocytes from CCR9−/− mice, the exacerbation of asthma caused by food allergen CD4+ lymphocytes was eliminated. These results show that CCR9 is a driving factor for the worsening of lung inflammation in food allergic mice (Castan et al., 2018), and confirm its application potential in the development of treatment strategies for allergic diseases (Table 2).

**Autoimmune Diseases**

Toll-like receptor 4 (TLR4) is an important part of innate immunity and has been linked to central nervous system (CNS) inflammatory diseases (Ureña-Peralta et al., 2020). Zhang et al. (2019) have shown that TLR4−/− mice were inadequate to induce experimental autoimmune encephalomyelitis (EAE),
which is featured by reducing low clinic score, weight loss, demyelinating, and spinal cord inflammatory cell infiltration. The deletion of TLR4 in the lesion area of EAE mice can decrease inflammatory cytokines and CCL25 secretion. After transformation, CCR9 expression is reduced in TLR4−/− Th17 cells, and the chemotaxis and migration ability are weakened. In summary, TLR4 may play an important role in the infiltration of Th17 during the pathogenesis of EAE by CCL25/CCR9 signaling pathway (Zhang et al., 2019). Kadowaki et al. (2019) also demonstrate that in the mice model of experimental autoimmune encephalomyelitis, CCR9+ memory T cells preferentially infiltrate the inflammatory CNS and express lymphocyte activation 3 gene (LAG3) in the late stage. Antibiotic treatment can alleviate the symptoms of experimental autoimmune encephalomyelitis, which is supported by a significant rise in peripheral blood CCR9+ memory T cells. Generally speaking, they postulate that the alterations in CCR9+ memory T cells observed, caused by either the gut microbiota changes or ageing, may lead to the development of secondary progressive multiple sclerosis (Kadowaki et al., 2019). Moreover, the CCL25/CCR9 was also powerful to authorize effector/memory T cells to access anatomic sites (Evans-Marin et al., 2015). Binding of CCR9 to CCL25 inhibits CD4 T cells differentiating to Tregs. In autoimmunity, CCR9+ T helper cells induce diabetes by secreting IL-21 and promote the expansion and survival of CD8+ T cells (McGuire et al., 2011).

Omenn syndrome is resulted from a mild rag mutation, which is featured by severe immunodeficiency and similar manifestations of autoimmune (Villa et al., 1998; Khiong et al., 2007). Rigoni et al. (2016) confirmed that long-term use of broad-spectrum antibiotics in Rag2 (R229Q) mice can ameliorate intestinal and systemic autoimmunity by reducing mucosal and circulating gut-tropic CCR9+ Th1 and Th17 T cells frequency. In addition, during the allergic reaction, CCL25 drives the mobilization of IL-17+ γδ T cells to inflamed tissues through α4β7 integrin, and regulates the level of IL-17 (Costa et al., 2012). The above studies have proved that CCR9/CCL25 plays an important role in autoimmune diseases.

**Cancer**

In microenvironment, tumor formation and immune system are mutually restricted, and chemokines also play an important role between tumor formation and immune system (Fan et al., 2019; Rossi et al., 2019; Lei et al., 2020). T cells are one of the main functional cells of the anti-cancer immune response. CD4+ T cells have an important regulatory effect, and CD8+ T cell stem cell-like subgroups have the potential to self-proliferate and differentiate toward effector cells, which can produce durable anti-tumor immune response and specifically kill tumor cells (Borst et al., 2018; Farhood et al., 2019; He et al., 2019; Brightman et al., 2020).

Studies have found that T cell function depends on the expression of CCR9. CD4+ CCR9+ T helper cells express high amounts of interleukin 21 and induce T cell costimulatory factor, transcription factor B cell CLL/lymphoma 6 (Bcl-6), and Maf, which are local features of autoimmune diseases that affect the accessory organs of the digestive system (McGuire et al., 2011). The CCR9 signal transmitted during the startup of naïve T cells promotes the differentiation of memory CD4+ T cells that produce α 4 β + 7 IFN-γ, increasing the immune microenvironment in gastrointestinal tissues, thereby affecting effector immunity in infections and cancer (Fu et al., 2019). Notably, Chen et al. (2020) used tumor acidity-responsive nanoparticle delivery system (NP-siCD47/CCL25) to discharge CCL25 protein and CD47 siRNA in tumors to enhance CD47 targeted immunotherapy.

In the study, NP-siCD47/CCL25 remarkably increased the invasiveness of CCR9+CD8+ T cells, inhibited the expression of CD47 in tumor cells, and inhibited tumor cells metastasis and growth by T-cell-dependent immunity (Chen et al., 2020). In addition, Khandelwal et al. (2015) pointed out that CCR9 regulates the STAT signal in T cells and inhibits the secretion and cytotoxicity of T lymphocytes type 1 cytokines. This study shows that inhibiting CCR9 expression may promote tumor-specific T cell immunotherapy (Khandelwal et al., 2015). In conclusion, targeting CCR9/CCL25 is expected to be a new approach for treating tumors, by suppressing or treating tumor patients with an autoimmune system that produces a lasting antitumor response.

**CONCLUSION AND POTENTIAL FUTURE DIRECTIONS**

Inflammatory diseases have become a serious threat to the health of the global population. Inflammatory diseases involve lesions of multiple tissues and organs in the body, which are characterized by tissue damage caused by excessive or persistent inflammation, such as CVD, hepatitis, asthma, RA, and inflammatory bowel diseases. However, the common features of these diseases are abnormal regulation of inflammatory response and imbalance of immune pathways. Chemokines are a family of small molecule cytokines whose receptors are seven-transmembrane glycoproteins, mainly expressed on the surface of inflammatory cells. Previous studies have shown that as members of the chemokine CC subfamily, CCR9 and CCL25 have been related to various inflammatory diseases as well as could promote inflammatory responses. Recently, the roles of CCR9/CCL25 in inflammation and related diseases have become clearer. CCR9/CCL25 is participated in many inflammatory diseases, including CVD, hepatitis, RA, IBD, and asthma (Table 2). Although the functional studies regarding CCR9/CCL25 in inflammatory diseases have gradually deepened, the understanding of CCR9/CCL25 has become more comprehensive. However, studies on the mechanisms of CCR9/CCL25 underlying inflammatory diseases are mostly restricted to the theoretical basic, and their clinical application is few.

Although several lines of evidence have clarified the specific mechanism of CCR9/CCL25 signaling in inflammation-associated diseases, the development of successful treatment of CCR9/CCL25 is still in further study stage.
CCX282-B is bioavailable in the circulation following oral administration, and which has high specificity for CCR9 with negligible binding to any other chemokine receptor, have been evaluated in murine models of IBD as well as in clinical trials. Meanwhile, related studies have confirmed that the use of an oral antagonist for CCR9 (CCX282-B) was also evaluated in phase II and phase III clinical trials with mixed results, but the results were not as expected. Therefore, to make further progress in delineating the potential clinical role of CCR9 in human disease, detailed mechanistic studies and well-designed proof-of-concept human trials are now required. Further investigation is likely to focus on (1) studying the effects of blocking CCR9 on the activation state of circulating and tissue-infiltrating lymphocytes, (2) investigating disease remission in CD remains a strong indication for CCR9 antagonist treatment, and any future clinical trials should include patients with UC and with PSC, (3) investigating whether the clinical application of CCR9 antagonism has any suboptimum efficacy, and (4) continuing to study CCR9/CCL25 upstream and downstream molecules, which are possible future potential therapeutic targets for inflammatory diseases. In conclusion, this review presents a comprehensive picture of the role of CCR9/CCL25 in inflammatory diseases and proposes a new underlying therapeutic mechanisms and target for the future treatment of inflammatory diseases.

AUTHOR CONTRIBUTIONS

XW and MS: conceptualization, literature search, and original draft writing and editing. ZY, CL, QW, HW, and CD: original draft writing and editing.YL and YY: conceptualization, original draft writing and editing, supervision, and funding acquisition. All authors contributed to the article and approved the submitted version.

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Wu et al. Roles of CCR9/CCL25 in Inflammation
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