Peptidines: glycine-amidine-based oligomers for solution- and solid-phase synthesis†

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Efforts to emulate biological oligomers have given rise to a host of useful technologies, ranging from solid-phase peptide and nucleic acid synthesis to various peptidomimetic platforms. Herein we introduce a novel class of peptide-like oligomers called “peptidines” wherein each carbonyl O-atom within poly-N-alkyl glycine oligomers is replaced with a functionalized N-atom. Compared to peptoids or peptides, the presence of this amidine N-substituent in peptidines effectively doubles the number of diversification sites per monomeric unit, and can decrease their overall conformational flexibility. We have developed iterative solution- and solid-phase protocols for the straightforward assembly of peptidines containing diverse backbone and amidine substituents, derived from readily available primary and secondary amines. We have also performed crystallographic and computational studies, which demonstrate a strong preference for the trans (E) amidine geometry. Given their straightforward synthetic preparation and high functional group density, peptidines have the potential to serve as useful tools for library generation, peptide mimicry, and the identification of biologically active small molecules.

Introduction

Oligomer-based synthesis is central to all known life processes. In particular, the structural and functional variety found in proteins is derived from the assembly of only 20 amino acid building blocks. Efforts to emulate this diversity have led to a range of oligomer-based peptidomimetic strategies, including oligopeptides, peptoids, peptidosulfonamides, sulfonylpeptides, polypyrroles and others.1–4 In turn, these strategies have given rise to numerous exciting applications, including combinatorial library synthesis,5–7 solid-supported screening,6–8 and biomarker discovery,9–11 making the development of novel oligomer-based approaches to achieve structurally-defined compound libraries a highly desirable endeavour.12,13

Herein we introduce a new class of oligomeric scaffolds that we term “peptidines”. Peptidines are oligomers composed of repeating di-substituted glycine-derived amidines (Fig. 1). Although similar to peptides and peptoids, the peptidine scaffold accommodates two substituents per monomeric unit by replacing C==O with C==NR thus doubling the accessible diversity for a given oligomer length. By varying the size and electronics of amidine N1 (backbone) and N2 (amidino) substituents, one can also modulate N-lone pair basicity, and backbone geometry.

† Electronic supplementary information (ESI) available: Experimental procedures, characterization data of new compounds. CCDC 1430515. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c5sc03882k

Fig. 1 A structural comparison of peptides, peptoids and peptidines.
Thus, we have developed a concise peptidine synthesis protocol that allows both \(N^1\) and \(N^2\) substituents to derive modularly from the large pool of commercially available primary amines. Using this route, we have been able to produce peptidines ranging in size from 2- to 4-mers, appended with sterically and electronically diverse substituents at both \(N^1\) and \(N^2\) positions. These syntheses proceed in short order, and with excellent yields in both solution and solid phases. Crystallographic and computational studies have demonstrated that amidines present within the peptidine scaffold prefer the trans-\((E)\) geometry of the \(N^2\) substituent with respect to the \(N^1\) nitrogen. Peptidines therefore adopt discrete conformations as a function of both H-bonding effects and non-bonding interactions. In light of their facile preparation and high potential for chemical conformational diversification, we envision that peptidines will provide a useful scaffold for the preparation of novel and diverse structures with a range of chemical and biological applications.

**Results and discussion**

Our strategy for preparing peptidines is outlined in Scheme 1. As shown, this protocol is composed of two separate, iteratively-applied synthetic transformations: first is an amidination step (Scheme 1A, reaction 1), wherein we amidinate a secondary amine (1) with an imidoyl chloride (2) to produce an \(\alpha\)-chloro amidine (3); second is an amination step (reaction 2), wherein the chloride atom in 3 is displaced by a primary amine (4) to form an \(\alpha\)-amino amidine (5). Iteration of reactions 1 and 2 affords peptidine oligomers (6). In turn, the \(\alpha\)-chloro imidoyl chlorides (2) used in reaction 1 are generated from commercially available primary amines (7), which are acylated to produce chloroamides (8), and then chlorinated to form imidoyl chlorides (2) (Scheme 1B).

We first focused on converting \(\alpha\)-chloro-amides (8a-i) to the corresponding imidoyl chlorides (2a-i, Table 1, reaction 1). This transformation was accomplished with substantial generality using \(\text{PCI}_3\) in refluxing benzene.\(^{14,15}\) Although the poor lytic stability of intermediate imidoyl chlorides precluded traditional work-up and purification protocols, \(^1\)H-NMR analysis of crude reaction mixtures indicated \(\approx 95%\) purity, alleviating the need for further purification. These intermediates were stable at room temperature for months without observable decomposition upon storage under moisture-free conditions as 1 M stock solutions in DCM.

We next focused on amine amidination with imidoyl chlorides (Table 1, reaction 2). Thus, treatment of intermediates 2a-i with diisopropylamine (intended to serve as a solution-phase model for sterically-hindered, resin-bound amine) successfully afforded sulfonfyl- \((3a-d)\), aryl \((3e-f)\), carbamoyl-\(^{16}\), and urea-derived \((3g-h)\) amidines. These yields were universally high (>70%) throughout a range of \(N^2\)-substituents and as expected, reactions with sulfonamides proceeded faster (14 h \textit{versus} 2 h), and in higher yields, than aryl and acyl derivatives. Furthermore, despite the possibility for reaction at the alkyl chloride position, we observed complete chemoselectivity for displacement at the acylimino carbon for all substrates examined, with no evidence of double addition. Similar chemoselectivities have been observed previously in condensation reactions between bis-electrophiles such as \(\alpha\)-chloro-acid chlorides and secondary amines.\(^{16}\) Although alkyl imidoyl chloride 2i did appear to provide the corresponding \(\alpha\)-chloro-amidine (3i) following diisopropylamine treatment, attempts to purify this compound were hampered by the presence of inseparable amounts of diisopropylammonium chloride and diisopropylamine. However, by carrying out the chlorination reaction at room temperature to avoid decomposition (likely via polymerization), followed by trapping with diethylamine, we were able to access alkyl amide 3j (ESI Fig. 1†). Despite this one example, we found \(N^2\)-alkyl-substituted intermediates to be highly unstable, leading to difficulties in isolation and purification. We therefore elected not to pursue this electron-rich substrate class further.

Having demonstrated the ability to produce a wide array of \(\alpha\)-chloro amides (3a-h), we next focused our efforts on displacing the pendant chloride with amine nucleophiles (Table 1, reaction 3). We conducted these reactions using benzylamine as a model primary amine nucleophile and used an excess of this reagent to mimic the conditions that we would be using to produce these oligomers on solid phase. We found that treatment of \(\alpha\)-chloro amides with both primary and secondary amines in the presence of iodide afforded the corresponding \(\alpha\)-amino amides (5a-h) in excellent yields.\(^{17}\) Reactions employing benzylamine as the nucleophilic component proceeded smoothly with electron poor sulfonamides (Table 1, entries 1-4) and \(p\)-nitrophenyl (entry 5) derivatives to give \(\alpha\)-amino amides (5a-e) in near quantitative yields. Interestingly, the phenyl derivative 5f was found to decompose rapidly as the free base, however addition of hydrochloric acid in ether to this compound immediately after silica gel purification facilitated its isolation as the stable HCl salt. Furthermore, the
reaction of 3h with benzylamine produced cyclic amidine 5h (ESI Fig. 2†) in nearly quantitative yield, with no acyclic product observed by LCMS.

We next analyzed how varying the structure of the amine nucleophile would affect the chloride displacement process using substrate 3a as the α-chloro amidine component (Table 1 entries 10–15). Overall, this reaction proceeded in greater than 90% yield using cyclohexylamine and aniline as nucleophiles to yield 9a and 9b respectively, although the latter substrate required heating. Similarly high yields were observed for secondary amine nucleophiles, such as diethylamine and pyrrolidine, affording 9c and 9d respectively. Decreases in yield were observed for the sterically bulky amines shown in entries 14 and 15. Benzhydrylamidine provided 9e in 72% yield and t-butylamine was incapable of reacting with 3a, and afforded none of the desired product (9f). Taken together, these results confirm that the peptidine core can be constructed through a three-step procedure involving: (1) amide chlorination, (2) amidination of a secondary amine, and (3) α-halide displacement with a primary or secondary amine.

Our next goal was to iterate the above three-step sequence to access longer oligomeric peptidines. We therefore chose to target a simple 3-mer composed of identical repeating monomeric units (Scheme 2). Starting from α-chloro amidine 5b, and using imidoyl chloride 2b and benzylamine as nucleophile, we were able to access 2-mer 10 and 11, and 3-mer 12 in only three steps and 80% overall yield by simply repeating chloride displacement and amidination steps. Notably, none of these...

**Table 1 Modular synthesis of α-amino-amidines**

| Entry | Amide | R   | Product | Yield 3a–h (%) (2-step) | R¹ | R² | Product | Yield 5a–h/9a–f |
|-------|-------|-----|---------|-------------------------|----|----|---------|-----------------|
| 1     | 8a    | TorSO₃⁻ | 3a       | 93%                     | H  | Bn | 5a       | 96%             |
| 2     | 8b    | MeSO₃⁻ | 3b       | 86%                     | H  | Bn | 5b       | 95%             |
| 3     | 8c    | (IPr)₂SO₃⁻ | 3c       | 88%                     | H  | Bn | 5c       | 92%             |
| 4     | 8d    |       | 3d       | 88%                     | H  | Bn | 5d       | 99%             |
| 5     | 8e    |       | 3e       | 72%                     | H  | Bn | 5e       | 90%             |
| 6⁶     | 8f    |       | 3f       | 70%                     | H  | Bn | 5f       | 91%             |
| 7     | 8g    |       | 3g       | 83%                     | H  | Bn | 5g       | 96%             |
| 8     | 8h    |       | 3h       | 79%                     | H  | Bn | 5h       | 92%             |
| 9     | 8i    | Ph(CH₂)³⁻ | 3i       | —                      | —  | —  | —       | —               |
| 10    | 8a    | TorSO₃⁻ | —       | —                      | H  | Cy | 9a       | 96%             |
| 11⁷     | 8a    | TorSO₃⁻ | —       | —                      | H  | Ph | 9b       | 90%             |
| 12    | 8a    | TorSO₃⁻ | —       | —                      | Et | Et | 9c       | 99%             |
| 13    | 8a    | TorSO₃⁻ | —       | —                      | -(CH₂)₃⁻ | 9d | 92%       |
| 14    | 8a    | TorSO₃⁻ | —       | —                      | H  | CHP₃⁻ | 9e | 72%       |
| 15⁸     | 8a | TorSO₃⁻ | —       | —                      | H  | ‘Bu | 9f | 0%           |

⁶ DIPA = diisopropylamine. Cy = cyclohexyl. ⁷ Isolated yield from 8a–h. ⁸ Isolated yield from 3a–h. ⁹ Reaction 1 was run at 25 °C; reaction 2 was run for 14 hours. ¹ Product was isolated as a hydrochloride salt. See ESI for details. ² Reaction 3 product rapidly cyclizes to form 1-benzyl-4-(diisopropylamino)-1,5-dihydro-2H-imidazol-2-one (ESI Fig. 2). ³ We were unable to isolate 3i due to difficulties in purification, as described in the text. The corresponding N²-diethylelamidine (37) was prepared in 78% yield, as detailed in ESI Fig. 1†. ⁴ Reaction 3 was carried out in the absence of NaI at 80 °C for 12 hours. ⁵ No product was observed by LCMS.
transformations proceeded in lower than 90% yield. Furthermore, using this protocol, we have been able to prepare 12 in quantities greater than 1 g demonstrating the scalability of this procedure.

Efforts to advance 3-mer 12 into longer oligomers gave rise to several notable findings. For example, treatment of 12 with benzylamine led exclusively to cyclic product 13 (Table 2, entry 1), as characterized by NMR and LCMS. Because formation of 12 proceeds without evidence of cyclic byproducts, we hypothesize that the diisopropyl groups in 11 protect the C-terminal amidine from internal nucleophilic attack by the appended secondary amine. Although 13 is incapable of further elongation, this observation demonstrates that the peptidine platform is capable of giving rise to nonlinear molecular architectures. Replacement of benzylamine with (S)-methylbenzylamine afforded no cyclic product upon reaction with 12, but instead exclusively provided the expected linear product 14 (Table 2, entry 2) in 99% yield. We hypothesize that the additional methyl group in (S)-methylbenzylamine provides sufficient steric encumbrance to prevent internal cyclization. Switching the nucleophile to Bn₂NH provided linear 3-mer 15. We also succeeded in producing a linear 4-mer peptidine (16) via our two-step elongation protocol (Table 2, entry 4); however unlike other acylation reactions, formation of 16 required heating to 60 °C to reach completion, most likely due to steric hindrance surrounding the amine terminus in the starting material (14). These results demonstrate that by regulating the steric environment around backbone N₁ substituents, we can access both linear and cyclic peptidines.

Given the suitability of oligomer-based synthetic strategies for the preparation of one-bead-one-compound libraries, our next effort was focused on adapting our platform to the solid-

Table 2  Tetramer synthesis and cyclization

| Entry | Reactant | Condition(s) | Product | Yield |
|-------|----------|--------------|---------|-------|
| 1⁵⁺ᵇ | 12       | Benzylamine, NaI, CH₃CN, 25 °C, 1 h | 13       | 79%   |
| 2ᵇ    | 12       | (S)-PhCH(Me)NH₂, CH₃CN, 80 °C, 1 h | 14       | 99%   |
| 3ᵇ    | 12       | Benzylamine, CH₃CN, 80 °C, 16 h | 15       | 99%   |
| 4     | 12       | 2b, NMM, CH₃CN, 60 °C, 3 h, 1) Entry 2, 2) Two-step yield from 12 | 16       | 82%⁴⁺ᵈ |

⁵ 5 equivalents of NaI were used. ⁶ 5 equivalents of amine were used. ⁷ 1.5 equivalents of imidoyl chloride and 3 equivalents of NMM were added to the corresponding amine in CH₃CN at 60 °C for 3 hours. ⁸ Two-step yield from 12.
phase. After developing a protocol for converting commercially-available Rink resin (38) to the corresponding cyclohexylamine derivative (39, ESI Fig. 3A†), we applied our iterative acylation-amination sequence (17 → 21) to the synthesis of peptidines containing between one and four monomeric units (Fig. 2). After optimizing the coupling of tosyl imidoyl chloride 3a to the resin bead (ESI Fig. 3B†), we were then able to access linear peptidines 22–26 in good to excellent crude purities. Microcleavage of each resin-bound intermediate during the production of these peptidines followed by LC/MS analysis demonstrated that each synthetic step was capable of fully consuming each resin-bound starting material (for both amination and amidination steps).

Interestingly, the steric bulk of the resin-bound amine plays an important role in solid-phase syntheses by preventing on-resin cyclization and cleavage. Whereas peptidines containing N-terminal secondary amines tend to undergo spontaneous cyclo-deamination (e.g. 12 → 13, Table 2), analogous solid-supported peptidines do not, likely due to steric effects of the cyclohexyl and Rink-benzhydryl functional groups. This feature proved fortuitous in enabling the synthesis of 3- and 4-mers 24–26. Upon resin cleavage, however, linear intermediates

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**Fig. 2** Solid phase synthesis of peptidine oligomers. Crude purity assessed by integration of the crude LC/MS spectrum at 254 nm (peptidine maximum absorbance). Values represent product peak area over total peak area. Rink MBHA resin (peptides international, 0.62 mmol g⁻¹ or 0.51 mmol g⁻¹) was used for synthesis.
underwent rapid cyclization. Compound 27 is an example of such a situation, and was initially observed as a mixture with its uncyclized counterpart 43 (ESI Fig. 4†), accounting for the relatively low crude purity of this compound. Further cyclization occurred during purification upon exposure to silica, leading to an acceptable isolated yield.

After establishing the solid-phase elongation protocol, we explored the scope of $N^1$ and $N^2$ amidine substituents (Fig. 2B). In general, peptidines exclusively containing $N^2$-sulfonyl substituents were produced in high purities and moderate isolated yields, even with widely varying $N^1$-substituents (28–32, Fig. 2B). Notably, the synthesis of 32 was carried out on large scale (1 g of Rink resin), and provided 228 mg of pure product, demonstrating the scalability of our solid-phase platform. $N^2$-carbamoyl and carbonyl substituents could also be incorporated into our solid-phase platform (33 and 34, respectively); however, as observed in analogous solution phase experiments, $N^2$-carbamate derivative 34 readily underwent intramolecular cyclization to give an amidazolone substructure, similar to compound 5h. Amines containing acid-labile side-chain protecting groups, similar to those employed in Fmoc SPPS (e.g., Boc, tert-Bu), were readily incorporated into peptidines, and unmasked after resin cleavage to give carboxylic and amine functional groups (as demonstrated by the synthesis of 35). Employing homo-β-alanine as the C-terminal group allowed us to access 2-mer 36 in good yield and purity (Fig. 2B). Significant efforts to incorporate $N^2$-arylamidines into solid-phase oligomers proved unsuccessful, and yielded either unreacted, resin-bound starting material or complex mixtures. Taken together, these results indicate that a diverse range of $N^1$ substituents, and electron-withdrawing $N^2$ substituents are compatible with the solid-phase peptidine synthesis platform.

Given the unique structures of peptidines we next sought to evaluate their chemical and spectral properties. First, we observed that numerous peptidines (11–12, 14–15, 23–26, and 28–35) tended to exhibit broadened peaks in $^1$H and $^{13}$C NMR spectra during routine characterization. This phenomenon was not surprising, as other oligomeric scaffolds, such as peptoids, can be very flexible and often exist in multiple different conformations at room temperature. Indeed, variable temperature NMR experiments performed on compound 12 (d$_6$-DMSO, 25–125 °C) demonstrated peak sharpening with increasing temperatures (ESI Fig. 5†). These data – coupled with analytical HPLC results (ESI Appendix†) – support both the chemical purity of peptidines, while also indicating some degree of conformational flexibility at room temperature.

We sought to gain additional insights into peptidine structure using X-ray crystallography. To this end, we obtained a crystal structure of 28 as the hydrochloride salt. As shown in Fig. 3A, 28 exists in a turn-like conformation, wherein the chloride counter-ion (green) interacts with atoms at both termini of the molecule. Additionally, a hydrogen bond between the terminal ammonium N–H group and the adjacent sulfonamide oxygen atom (Fig. 3A, right most dotted green line, 2.1 Å) locks the N-terminal residue into a H-bonded seven-membered ring. Examining 28 from an alternate perspective reveals that all $N^2$-substituents are located opposite the chloride ion (ESI Fig. 6f) while the $N^1$-substituents project perpendicularly with respect to the peptidine backbone. This data suggests that the unique arrangement of H-bond donors and acceptors within peptidines exerts a significant impact on conformation and may impart distinct structural features that could prove useful for the design of peptidomimetics in future studies.

To gain further insight into peptidine structure, we analyzed crystallographic data in terms of standard peptide dihedral angles phi ($\Phi$), psi ($\Psi$), and omega ($\omega$) (Fig. 3B). Additionally we also defined a new dihedral angle, called tau ($\tau$), referring to the position of the $N^2$-substituent relative to the $N^1$-nitrogen (Fig. 3B). Perhaps most notably, all three $\tau$ angles exist in a trans conformation ($\tau = 180^0 \pm 15^0$), despite each being in a different steric environment. This observation is consistent with previous literature reports, as well as computational studies (see below). The $\omega$ angles in 28, on the other hand, proved slightly more variable, demonstrating a preference for either cis or trans conformations ($\omega = -10.5^0 \pm 174.9^0, 177.3^0$), reminiscent of disubstituted amides found in peptides and peptoids. Due to these structural constraints ($\omega \approx 180^0$ or $0^0$; $\tau \approx 180^0$), there is a high degree of co-planarity surrounding each amidine in 28 (Fig. 3A and B). The most flexible dihedrals in 28 were $\Phi$ and $\psi$, once again reminiscent of peptides and peptoids. Although in the case of 28, we believe these angles are driven to accommodate the organizing N–H–Cl interactions, we are currently conducing investigations on the intrinsic preferences of these angles as a function of $N^1$ and $N^2$ substituent properties.

To understand the tendency of the amidine $N^2$-substituent to prefer the trans geometry ($\tau = 180^0 \pm 15^0$), we performed quantum chemical energy calculations (Table 3). Previous analysis of amidine geometries suggest that the trans geometry is favored in $N^2$-alkyl and $N^2$-aryl substituted amides, with several examples of substituents preferring the cis geometry. However, to our knowledge, no calculations have been carried out to evaluate the conformational preferences of strongly electron withdrawing substituents at the $N^2$ position such as sulfonyl or carbonyl groups. In evaluating the geometric preferences of the $N^2$-substituent we utilized a simple monomer amidine as a model system. Thus, we employed 2D Sketcher and MacroModel software packages using the OPLS 2005 force field to construct and optimize six model structures including sulfonyl-, carbomethoxy-, carbamoyl-, and aryl-functionalized $N,N$-dimethylacetamide derivatives (Table 3, entries 1–6). For each compound, we set initial $\tau$ angles to 0° and 180°, and performed geometry optimizations using the program Jaguar. Density functional theory (DFT) with the B3LYP functional and 6–31G** basis set were used in this process. Energies from minimized structures are presented in Fig. 3C. As expected, for each amidine tested, calculations predict a significant energetic preference for the trans geometry, ranging from 2–12 kcal mol$^{-1}$. These results are consistent with crystallographic data for compound 28, and suggest that, in general, the peptidines reported herein are all likely to exhibit a substantial preference for the trans geometry. Interestingly, other authors have observed that amidines containing electron-donating $N^2$
substituents such as alkoxy substituents readily exist as cis/trans mixtures with preferences for the cis geometry.\textsuperscript{27,28} Although our synthesis platform is currently incompatible with such electron-rich groups on $N^2$, the possibility exists that the $\tau$ geometry can be modulated through substituent effects. Additionally, we tested the hydrolytic stability of several compounds in PBS and observed minimal hydrolysis suggesting that peptidines may be suitable for use in biological contexts (ESI Fig. 7†).

### Conclusions

Herein we have introduced a novel class of glycine-amidine-based oligomers, which we term “peptidines” readily afforded through modular synthetic approaches using both solid and solution phase chemistry. These synthetic protocols are high yielding, readily scalable, and enable straightforward installation of a variety of substituents onto backbone ($N^1$) and amidine ($N^2$) nitrogen atoms. Also, we have obtained a crystal structure of 3-mer 28, which demonstrates all amidine motifs to adopt a trans geometry about the $\tau$ angle. This geometry conforms to structural constraints predicted by computational studies, and suggests that peptidines have the potential to project functionality in an ordered array. Furthermore, because peptidines possess two sites of diversity per monomeric unit (as opposed to only one present in peptides and peptoids), they may prove useful for library generation as greater diversity could be generated with shorter length oligomers. Peptidines therefore have significant potential to serve as useful tools for small molecule synthesis, peptidomimicry, and library generation.\textsuperscript{29–31} Further studies to explore peptidine structure and

### Table 3 Calculations on tau angle preference

| Entry | R        | Minimized $\tau$ geometry (degrees) | $\Delta$Energy $\text{trans-cis}$ (kcal mol$^{-1}$) |
|-------|----------|-------------------------------------|-----------------------------------------------|
| 1     | S(O)$_2$Tol | trans: $-177.9$ cis: $-8.9$         | $-11.6$                                       |
| 2     | S(O)$_2$Me | trans: $178.2$ cis: $-16.2$         | $-4.4$                                        |
| 3     | C(O)OMe   | trans: $179.1$ cis: $27.9$          | $-2.0$                                        |
| 4     | C(O)NBn$_2$ | trans: $-171.3$ cis: $-27.4$      | $-4.1$                                        |
| 5     | Ph        | trans: $-179.4$ cis: $17.4$         | $-5.2$                                        |
| 6     | p-NO$_2$Ph | trans: $177.0$ cis: $-20.8$         | $-4.7$                                        |

Fig. 3 Peptidine structure and angle preferences. (A) Crystal structure of 28 in ORTEP model at the 50% confidence interval. Grey represents carbon, blue represents nitrogen, yellow represents sulfur, red represents oxygen, green represents chlorine, white represents hydrogen. Dashed lines represent hydrogen bonds or ionic interactions observed in crystal. (B) List of peptidine dihedral angles taken from the crystal structure of 28. Atoms 6, 11, and 16 referenced to the sulfur atom.
stability, and to apply this platform to the synthesis of a wide variety of functional molecules, are currently ongoing.

Acknowledgements

We thank Dr Michael Takase for completing crystallography studies and Dr Brandon Mercado for assistance with crystallography data analysis and presentation. We thank Dr Dianna Bartel for assistance in writing of the manuscript.

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