Assessment of Anti-nutritive Activity of Tannins in Tea By-products Based on In vitro Rumen Fermentation

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ABSTRACT: Nutritive values of green and black tea by-products and anti-nutritive activity of their tannins were evaluated in an in vitro rumen fermentation using various molecular weights of polyethylene glycols (PEG), polyvinyl pyrrolidone (PVP) and polyvinyl polypyrrolidone as tannin-binding agents. Significant improvement in gas production by addition of PEG4000, 6000 and 20000 and PVP was observed only from black tea by-product, but not from green tea by-product. All tannin binding agents increased NH3-N concentration from both green and black tea by-products in the fermentation medium, and the PEG6000 and 20000 showed relatively higher improvement in the NH3-N concentration. The PEG6000 and 20000 also improved in vitro organic matter digestibility and metabolizable energy contents of both tea by-products. It was concluded that high molecular PEG would be suitable to assess the suppressive activity of tannins in tea by-products by in vitro fermentation. Higher responses to gas production and NH3-N concentration from black tea by-product than green tea by-product due to PEG indicate that tannins in black tea by-product could suppress rumen fermentation more strongly than that in green tea by-product. (Key Words: Tea By-Product, In vitro Rumen Fermentation, Tannin, Tannin Binding Agent)

INTRODUCTION

A large quantity of nitrogen fertilizer has been applied for production of tea leaf rich in amino acids (Oh et al., 2006). Nitrogenous compounds in the leaf are partially extracted by hot water and consumed as tea, but most of the compounds still remain in the residue of tea leaf (Kondo et al., 2004a). As concern increases about environmental issues and material recycling, the utilization of tea by-product as feedstuff would be useful for both economic and environmental reasons. In Japan, and East Asian and Southeast Asian countries, the consumption of ready-made green and black tea in cans, packs or bottles has increased remarkably in recent years. Beverage companies that manufacture tea drinks produce many tons of tea by-products annually and most of them are composted. Previous reports showed that tea by-products produced by the beverage industry contained high crude protein (CP, 220 to 350 g/kg dry matter; DM) and tannins (50 to 80 g/kg DM) (Kondo et al., 2004a,b). In order to evaluate the potential use of tea by-products as feedstuffs, the suppressive activity of tea tannins should be assessed.

The suppressive activity of tannins on rumen degradability has been assessed mainly in tropical leguminous leaves and fruit pods (Tolera et al., 1997; Ruhanza et al., 2005; Wambui et al., 2012) and agro-industrial by-products (Abarghuei et al., 2010; Al-Masri, 2012; Bhatta et al., 2012). Since polyethylene glycol (PEG) binds tannins and deactivates the anti-nutritive activity, an increase of the in vitro gas production from tannin-containing feeds due to PEG addition means that the tannins lowered the feed degradability in the rumen (Makkar et al., 1995). The higher increment of gas production due to PEG...
administration indicates the higher suppressive activity of tannin in the feeds (Jayanegara et al., 2009). Therefore the \textit{in vitro} gas production test in conjunction with tannin-binding agents has been used as a useful method for assessing tannin related suppressive activity on ruminant feedstuffs (Rubanza et al., 2005; Osuga et al., 2007). These tannin binding agents can be also useful as dietary supplements when tannin-rich feedstuffs are fed to animals (Ben Salem et al., 1999). However, to date, there has been no information regarding the effects of different molecular weights (MW) of PEG and other tannin binding agents on the \textit{in vitro} fermentation of tea by-products.

Therefore, the objective of this present study was to assess the anti-nutritive activity of tannins in green and black tea by-products by the \textit{in vitro} gas production method coupled with ammonia nitrogen (NH$_3$-N) measurement, organic matter (OM) digestibility and metabolizable energy (ME) content. Experiments were conducted to compare the different types of tannin-binding agents on the \textit{in vitro} test when green or black tea by-products were incubated with buffered rumen fluid.

**MATERIALS AND METHODS**

Green and black tea by-products were acquired from a local ready-made tea drink factory. Both green and black tea by-products were freeze-dried and ground to pass through 1 mm screen. Standard methods as described in AOAC International (2002) were used for determination of DM, OM, and CP. Neutral detergent fiber (NDF), acid detergent fiber (ADF) and acid detergent lignin (ADL) were analyzed as outlined by Van Soest et al. (1991). Acid detergent insoluble protein (ADIP) was determined by the method of Licitra et al. (1996). Total extractable phenolics (TEPH), total extractable tannins (TET), condensed tannins (CT) and protein precipitating capacity (PPC) were determined by the method of Makkar and Goodchild (1996). The PPC was expressed as the amount of bovine serum albumin (BSA) bound with tea by-products. NH$_3$-N concentration in the incubated medium was determined using the indophenol reaction (Weatherburn, 1967).

\textit{In vitro} rumen gas production from tea by-products was determined according to Menke and Steingass (1988). Two hundreds mg of freeze-dried green tea and black tea by-products passed through 1 mm screen were weighed each into 100 mL calibrated glass syringe. Four hundred mg of various MW of PEG (MW: 1,000, 4,000, 6,000, 20,000), polyvinyl pyrrolidone (PVP, MW: 10,000) or insoluble polyvinyl polypyrrolidone (PVPP) were added into the syringes. The ratio of feedstuff and PEG, PVP or PVPP were referred to Makkar et al. (1995) and Kondo et al. (2014). The syringes were filled with 30 mL medium consisting of 10 mL of rumen fluid and 20 mL of buffer solution, then incubated in a water bath at 39°C in three consecutive runs (n = 3 replicates) and gas production was measured at a series of incubation time for 24 h. The incubated medium was transferred to 50 mL centrifuge tube and the supernatant collected after centrifuging (1,090×g) for 10 min at 4°C. NH$_3$-N concentration of the supernatant was measured. The data “Increase %” of gas production or NH$_3$-N was calculated as Increase % = (data with additives – data without additives)×100/data without additives. \textit{In vitro} organic matter digestibility (IVOMD) and ME were estimated from the equation of Makkar (2004) using a gas production data.

The data of gas production, NH$_3$-N concentration and estimated IVOMD and ME were analyzed by a factorial analysis of variance, in which the two fixed factors were tea by-products and tannin-binding agents. Rumen content for each incubation batch served as block in the statistical model. When a factor or the interaction showed statistically significant at $p<0.05$, the differences among treatments means were further tested using Tukey’s test. All statistical analyses were performed by using SAS software version 9.3 (SAS Institute, Cary, NC, USA).

**RESULTS AND DISCUSSION**

Green and black tea by-products contained 336 CP and 283 g/kg DM, respectively (Table 1). The ADIP content, a lower digestible protein fraction, was relatively higher in black tea by-product than the green one. The NDF, ADF and ADL contents of both tea by-products were similar. The TEPH, including low molecular phenolics and tannins, was present in similar amounts in both tea by-products, approximately 84 g/kg DM. The TET content was high in both tea by-products and the values were 76.5 and 66.6 g/kg DM in green and black tea by-products, respectively. The

| Table 1. Chemical composition of green and black tea by-products |
|---------------------------------|--------|--------|
| **Items**                        | **Green tea** | **Black tea** |
| Dry matter (DM, g/kg)            | 196    | 201    |
| Organic matter (g/kg DM)         | 970    | 979    |
| Crude protein (CP, g/kg DM)      | 336    | 283    |
| Acid detergent insoluble protein (g/kg CP) | 70      | 92      |
| Neutral detergent fiber (g/kg DM) | 388    | 404    |
| Acid detergent fiber (g/kg DM)   | 294    | 271    |
| Acid detergent lignin (g/kg DM)  | 96.3   | 94.1   |
| Total extractable phenolics (g/kg DM) | 83.4  | 84.8   |
| Total extractable tannins (g/kg DM) | 76.5  | 66.6   |
| Condensed tannins (g/kg DM)      | 20.9   | 5.7    |
| Protein precipitating capacity (mg BSA/g DM) | 18.1 | 65.3 |

BSA, bovine serum albumin.
CT content in green and black tea by-product was 20.9 and 5.7 g/kg DM, respectively. The PPC of black tea by-product was more than three times higher than that of green tea. Singh et al. (2005) showed PPC had a high positive correlation with TEPH and CT content in 12 kinds of tropical tree forages. This capacity was not related to the contents of tannins in our study; it is supposed that molecular structures of tannins in tea by-products would be important for their PPC (Yamamoto et al., 1997). Further, tannins have a great structural diversity among various plant sources, and even slight changes in tannin molecular structures could lead to a significant alteration in their biological effects (Mueller-Harvey, 2006).

In vitro gas production from green tea by-product was significantly higher than black tea by-product (p<0.05) (Table 2), indicating that green tea by-product contained more fermentable substrates. Contrary to expectation, the gas produced from green tea by-product did not increase with the addition of any tannin-binding agents. Significant improvement in gas production by addition of PEG40000, 6000 and 20000 and PVP10000 were observed only in black tea by-product (p<0.05), but not by PEG10000 and PVPP. The highest increment (18.9%) in gas production was observed in PEG6000 treatment, but the extent of the increment was not significantly different (p>0.05). Many studies have shown that PEG increased in vitro gas production from most tannin rich feeds (Rubanza et al., 2005; Singh et al., 2005; Osuga et al., 2007). Although green tea by-product contained high tannins, the gas volume did not increase by PEG, PVP or PVPP addition. Apparently tannin biological activity as measured by the capacity of tannins to precipitate protein (BSA) is more related to the increase of gas production as compared to the tannin contents. This is in agreement with Silanikove et al. (2001) who stated that protein precipitation assay is more useful than that of quantification of tannins by colorimetric methods for the evaluation of anti-nutritional effects of the substances. Apart from that, gas production is a result of nutrient fermentation of feed in the rumen, mainly carbohydrate (Menke and Steingass, 1988). It may also indicate that there was no contribution to gas production from fermented substrate in green tea by-product due to supplemental tannin binding agents.

Large differences of response in gas production and NH3-N concentration to tannin binding agents were observed in the incubation of green tea by-product (Table 3). Contrary to the result of gas production, all kinds of PEG, PVP, and PVPP increased NH3-N concentration from green tea by-product. Higher response was obtained in PEG 6000, 20000 and PVPP, but it was not significantly different from other agents. NH3-N concentration from black tea by-product was also increased by addition of any of the agents with the higher response observed with PEG6000, 20000 and PVP10000, while PEG1000 and PVPP showed a relatively lower response. NH3 is the main end-product of protein fermentation in the rumen. The PEG also recovers feed proteins that are bound by tannins. An increased increment of NH3-N resulting from the addition of PEG can be attributed to a reduction in the opportunity of tannin binding to feed protein, which makes tannin-bound protein available to microbial degradation and metabolism (Getachew et al., 2000). The greater increase in NH3-N concentration means a greater suppressive effect of tannins on protein degradation. According to the greater NH3-N improvement from black tea by-product compared with green by tannin binding agents, protein degradation of black tea by-product would be highly suppressed by its tannins. This result could be reflected in the higher PPC of black tea by-product. The PPC was negatively correlated to NH3-N concentration more clearly than gas production because the

| Agents        | Green tea by-product | Black tea by-product |
|---------------|----------------------|----------------------|
|               | Gas (mL) | Increase (%) | Gas (mL) | Increase (%) |
| Control       | 38.9     | -0.1a       | 29.9a    |
| PEG1000*      | 38.8     | -0.4a       | 31.3bcd  | 4.8b       |
| PEG4000       | 38.7     | -0.5a       | 34.0ab   | 13.6a      |
| PEG6000       | 40.3     | 3.7a        | 35.6a    | 18.9a      |
| PEG20000      | 40.2     | 3.6a        | 34.6a    | 15.7a      |
| PVP10000      | 38.3     | -1.4ab      | 33.4abc  | 11.7ab     |
| PVPP          | 36.0     | -7.4b       | 31.2cd   | 4.4b       |
| SEM           | 1.09     | 1.42        | 0.56     | 1.75       |
| p-value       | <0.001   | <0.001      | <0.001   | <0.001     |
| Interaction   | 0.110    | 0.053       |          |            |

SEM, standard error of the mean. * Molecular weight.

a,b,c Means with different letters within a column are significantly different (p<0.05).
increase ratio to NH$_3$-N concentration by tannin binding agents was relatively higher than that to the gas production. Although in vitro rumen gas test with tannin-binding agents has been well-accepted method to identify anti-nutritional activity of tannin for rumen fermentation (Tolera et al., 1997; Rubanza et al., 2005; Osuga et al., 2007), measurement of gas production cannot fully assess the effects of tannin on feed degradation according to the response of PEG, PVP, and PVPP to green tea by-product. Additional to measurement of gas production, monitoring NH$_3$-N concentration in the incubated medium is recommended for tannin bioassay by in vitro rumen gas test. According to the present data, PEG6000 and 20000 would be suitable to assess the suppressive activity of tea tannins on protein degradability in the rumen. Makkar et al. (1995) supports this observation as they also reported that PEG6000 and PEG10000 showed relatively higher improvements by the gas production test from tannin-rich feeds. This could be due the binding of PEG and phenols takes place via H bonding, therefore, highly polymerized PEG contains contains more effective sites (oxygen of ether linkage) which bind to the hydroxyl group of tannins (Jones, 1965). PEG1000 and PVPP were less effective on gas production and NH$_3$-N concentration from black tea by-product. These results indicate that the low MW (~1,000) of PEG and insoluble PVPP would be least effective on binding to black tea tannin.

The estimated IVOMD and ME content were higher in green by-product than in black tea (Tables 4 and 5), and these values were comparable or higher than several ruminant browse forages containing tannin (Rubanza et al, 2005; Osuga et al., 2006;2007). None of the tannin binding

### Table 3. Effect of addition of polyethylene glycol (PEG), polyvinyl pyrrolidone (PVP) and polyvinyl polypyrrolidone (PVPP) on ammonia nitrogen (NH$_3$-N) at 24 h from green and black tea by-products

| Agents          | Green tea by-product | Black tea by-product |
|-----------------|----------------------|----------------------|
|                 | NH$_3$-N (mg/dL)     | Increase (%)         | NH$_3$-N (mg/dL)     | Increase (%)         |
| Control         | 25.2$^a$             | 15.1                 | 16.9$^a$             |                      |
| PEG1000*        | 29.0$^b$             | 13.8                 | 18.3$^{bc}$          | 8.4$^b$              |
| PEG4000         | 28.7$^b$             | 17.0                 | 21.7$^a$             | 33.2$^a$             |
| PEG6000         | 29.4$^a$             | 16.4                 | 22.5$^a$             | 33.7$^a$             |
| PEG20000        | 29.3$^a$             |                      | 22.6$^a$             |                      |
| PVPP            | 29.0$^a$             |                      | 22.5$^a$             |                      |
| SEM             | 0.62                 | 2.60                 | 0.38                 | 2.28                 |

p-value

|                     | Green tea by-product | Black tea by-product |
|---------------------|----------------------|----------------------|
| Tea by-products     | <0.001               | <0.001               |
| Tannin binding agents | <0.001              | <0.001              |
| Interaction         | 0.107                | 0.072                |

DM, dry matter; SEM, standard error of the mean. * Molecular weight.

Molecular weight

| Agents          | IVOMD (%)     | Increase (%) |
|-----------------|---------------|--------------|
| Control         | 64.8          |              |
| PEG1000*        | 64.7          | 0.0$^a$      |
| PEG4000         | 64.6          | −0.2$^a$     |
| PEG6000         | 66.0          | 2.0$^a$      |
| PEG20000        | 66.0          | 1.9$^a$      |
| PVPP10000       | 64.3          | −0.7$_{ab}$  |
| PVPP            | 62.2          | −4.0$^b$     |
| SEM             | 0.96          | 0.76         |

p-value

|                     | Green tea by-product | Black tea by-product |
|---------------------|----------------------|----------------------|
| Tea by-products     | <0.001               | <0.001               |
| Tannin binding agents | <0.001              | <0.001              |
| Interaction         | 0.107                | 0.072                |

DM, dry matter; SEM, standard error of the mean. * Molecular weight.

Molecular weight

| Agents          | Green tea by-product | Black tea by-product |
|-----------------|----------------------|----------------------|
| Control         |                       |                      |
| PEG1000*        |                       |                      |
| PEG4000         |                       |                      |
| PEG6000         |                       |                      |
| PEG20000        |                       |                      |
| PVPP10000       |                       |                      |
| PVPP            |                       |                      |
| SEM             |                       |                      |

p-value

|                     | Green tea by-product | Black tea by-product |
|---------------------|----------------------|----------------------|
| Tea by-products     | <0.001               | <0.001               |
| Tannin binding agents | <0.001              | <0.001              |
| Interaction         | 0.107                | 0.072                |

DM, dry matter; SEM, standard error of the mean. * Molecular weight.
agents affected IVOMD and ME content of green tea by-product, but PEG40000, 6000, and 20000 significantly increased IVOMD and ME content of black tea by-products (p<0.05). These parameters of both tea by-products were numerically high when PEG6000 and 20000 were used; indicating that highly polymerized PEG can improve nutritional value of tea by-product.

Min et al. (2003) reviewed that lower CT contents (20 to 45 g/kg DM) in temperate legumes improve nitrogen utilization, and milk and wool production in ruminants; in contrast, higher CT contents (>50 g/kg DM) reduce voluntary feed intake and digestibility, and depress rates of body and wool growth. Compared to these studies, CT content in both tea by-products were low; however, the in vitro fermentation was suppressed by their tannins. These different observations could be due to the effects produced are not the same for all CT, but rather depend upon the concentration, molecular structure, and biological activity of the CT (Min et al., 2003; Waghorn, 2008). It can also be expected that phenolic compounds other than CT present in green and black tea by-products could possess PPC and therefore may suppress the in vitro fermentation, such as epigallocatechin-3-gallate, epicatechin-3-gallate in green tea, and the aflavins in black tea (Yang et al., 2000). In the present study, despite of lower CT in the black tea by-product than that in green, PPC and increment of in vitro fermentation with PEG in black tea by-product were higher than those in green one. These findings suggest that CT measured by colorimetric assay for tea by-products might not be related to tannin’s biological activity.

CONCLUSION

The present study confirms the potentiality of green and black tea by-products to be used as protein-rich feedstuffs as indicated by their high CP contents. However, the concomitantly high tannin contents in the by-products may limit their utilization by ruminant livestock. Addition of tannin binding agents, especially PEG with higher MW (6,000 and 2,0000), is apparently suitable to deactivate the suppressive activity of the tannins as shown by the elevated values of total gas production, NH3-N concentration, IVOMD and ME contents in vitro in comparison with the control groups. Between the two tea by-products, greater responses in gas production and NH3-N concentration from the black tea by-product than from the green tea by-product due to PEG addition indicate that tannins in black tea by-product could suppress rumen fermentation more strongly than that in green tea by-product. Such responses seem to be more correlated with the PPC properties rather than the concentrations of tannins (either TET or CT) in the by-products. Thus, tannin bioassay (like the PPC) is apparently better for predicting nutrient utilization in the rumen than the colorimetric assay. Further studies are required for assessment of the nutritional values of these tea by-products as protein supplements with PEG addition in in vivo animal feeding experiments.

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