Identification of Genetic Variants Associated With Myocardial Infarction in Saudi Arabia

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ABSTRACT

The genetic variants associated with various genetic disorders have not been identified decisively in Saudi Arabia. Among these variants, six known for their association with coronary artery disease or myocardial infarction (MI) were studied on Saudi patients. Reference single nucleotide polymorphisms (SNPs) of these variants are rs5174, rs11591147, rs2259816, rs111245230, rs3782886 and rs2259820, referring to genes LRP8, PCSK9, HNF1A, SVEP1, BRAP and HNF1A, respectively. The analysis employed polymerase chain reaction panel coupled with mini-sequencing (SNapShot multiplex system) in order to identify these variants. A total of 100 MI patients and 103 healthy control individuals participated in this study. The six variants (SNPs) were evaluated for the risk of developing MI in the Saudi patients. Analysis of allele frequencies indicated that A allele of rs11591147 variant can be a protective allele, thus, is associated with the decreased risk of MI in Saudi individuals. Rare allele of rs111245230 variant (e.g., C allele) was extremely reduced, while rare allele of rs3782886 variant (e.g., G allele) does not exist in the ethnic signature of the Saudi population. This study elucidates the possible prediction of risk factors associated with severe diseases in Saudi population utilizing SNapShot multiplex system.

INTRODUCTION

The prevalence of coronary artery disease (CAD) is increasingly observed worldwide. The World Health Organization (WHO) anticipated that more than seven million people die from CAD each year in China [Wang 2014]. Myocardial infarction (MI) is the situation of the irreversible necrosis of the heart muscle that results from prolonged ischemia. Therefore, MI is considered the most crucial clinical manifestation of CAD [Matsouka 2015]. The inheritance of assorted genetic variants functioning in association with environmental factors for promoting the disease status characterized myocardial infarction as a complex disease [Ferreira 2018]. More than 90% of the risk of an acute MI, in a case-controlled study, is associated with the nine easily measured risk factors [Ma 2018]. The Chinese population was majorly affected by these risk factors, making a population of 89.9% at considerable risk.

The identification of several genetic loci has led to the progression of genome-wide association with CAD as well as MI risk [Isordia-Salas 2018]. Thereby, MI is a life-threatening disease as its pathogenesis is multifactorial and complex [Li 2019]. The development of MI has been demonstrated by the association between genetic factors, multiple genes, and environmental factors. Indeed, the inheritance of CAD indicates that 40-50% of the North Indian population are at risk of MI [Kashyap 2018].

The majority of the studied disease-related loci in European populations was replicated in populations of East Asian ancestry, however, this is not the case in Arab countries. Nikpay et al conducted a 1,000 genomes-based metaanalysis by including a small number of South Asian (13%) and East Asian (7%) subjects [Nikpay 2015]. However, the study did not report on ethnic-specific effects. The direct effects of CAD/MI risk alleles recognized in European populations on CAD risk in Asians generally were compatible, despite smaller data sets. On the other hand, there was a similar effect of size of CAD/MI risk alleles among the East Asian and European populations. In South Asia, the size effect apparently was weakened because of the interaction with unknown genetic or environmental factors along with the causal variants that may be inadequately tagged by markers present on available
genotyping arrays. This likely is to result in blunted genetic
effects. Until recently, genetic variants associated with various
disease-related genes have not been identified decisively in Saudi
Arabia. Among them are the variants related to CAD or MI
[Ferreira 2019; Kashyap 2018; Li 2019; Ma 2018]. Therefore,
this study attempts to examine the possible association of
genetic markers for susceptibility to MI among patients at
King Abdulaziz University Hospital (KAUH) in Jeddah, Saudi
Arabia. In addition, possible association of the ethnic back-
ground of Saudi individuals and any of these genetic markers
also was investigated. The study includes screening the vari-
ants rs5174, rs11591147, s2259816, rs111245230, rs3782886
and rs2259820, referring to LRP8, PCSK9, HNF1A, SVEP1,
BRAP and HNF1A genes, respectively.

MATERIALS AND METHODS

Blood collection: Peripheral blood samples were col-
clected from 100 MI patients and 103 healthy controls
in 2 ml EDTA-containing tubes, using a sterile syringe.
Characteristics of subjects involved in the study in terms of
numbers, gender, and smoking habits are shown in Table S1.
The collected samples were transported in an ice container
to the laboratory for further DNA isolation and stored at
-20°C. Inclusion criteria involved (i) diagnosis of MI cases
based on a history of MI, typical electrocardiographic
change, left ventricular angiography, and coronary angiogra-
phy; (ii) similar numbers of male to female, smokers to non-
smoker subjects; and (iii) subjects varied in age between 40
to 60 years old.

Genomic DNA isolation and quantification: The QIAamp
DNA mini kit was used to extract DNA from the leukocytes
of blood samples, while nanodrop-2000 spectrophotometer
(Thermo Scientific, USA) was used to determine the isolated
genomic DNA concentration.

SNaPshot Multiplex System for SNP genotyping: The
DNA sequences encompassing six SNPs were selected from
the genes LRP8, PCSK9, HNF1A (two SNPs), SVEP1, and
BRAP. The ensemble (http://www.ensembl.org/index) and
dbSNP (http://www.ncbi.nlm.nih.gov/ projects/SNP/) data-
bases were used to select candidate polymorphic sites. Genes
and reference SNP numbers along with nucleotide sequences
of primers flanking the polymorphic sites and amplicon sizes
used during PCR are described in Table 1.

Single base extension (SBE) primers subsequently were
constructed to have lengths of 30 to 78 nucleotides by adding
poly dC tails with varying lengths to secure the occurrence
of six nucleotide differences in the SBE primer lengths,
while avoiding regions of which other SNPs already had
been described. SBE primer sequences and sizes along with
the target variants of the consequent downstream SNPs are
Agarose gel electrophoresis: Gel electrophoresis was performed for the recovered PCR products using 3% agarose gel with SYBR-safe DNA gel stain added to the gel to analyze the size and quality of the PCR products. Moreover, the gel was placed into UV Trans-illuminator to visualize the successive products.

SNaPshot reaction preparation: The SNaPshot reaction was performed by adding the six different single base extension (SBE) primers to be hybridized with the different corresponding PCR products and extended at the 3' ends by only one fluorescent labeled ddNTP, each with a different color. This reaction is called single base extension that is used in order to detect SNP alleles of the different six genes for participating individuals. SNaPshot analysis was conducted using the ABI Prism SNaPshot Multiplex kit (Applied Biosystems, Life Technologies, CA). An amount of 1 pmol of SBE primers (1.5 µl) was added to an amount of 3.5 µl of purified multiplexed PCR products (30 ng) and 5 µl of the ABI Prism SNaPshot™ Multiplex kit (Applied Biosystems, Life Technologies, CA) to reach a final reaction volume of 10 µl. The SNaPshot reaction was placed on a Veriti™ 96 well thermal cycler (Applied Biosystems Inc., Life Technologies, CA) after mixing with 1 µl of 9.5 µl of Hi-Di formamide and 0.5 µl of GeneScan™ LIZ120™ internal size standard (Applied Biosystems Inc., Life Technologies, CA). Then, fragments were analyzed in a 3500 Genetic Analyzer (Applied Biosystems Inc., Life Technologies, CA) using a POP-7™ Polymer (Applied Biosystems Inc., Life Technologies, CA). Conditions during fragment analysis were as follows: injection (15 sec), pre-run (3 min), run time (15 min), and data delay (4 min). The resulted patterns were analyzed using GeneMapper Software v4.1.

SNaPshot validation: DNA samples of 20 patients and 20 control individuals were genotyped for the six SNPs by Sanger sequencing. The results of direct sequencing should match those of SNaPshot analyses in order to confirm 100% compatibility of the results.

Statistical analysis: The sequence variants were identified using the Bioedit software 6 editor version. The allele and genotype frequencies among the samples were estimated from the results of SNaPShot Assay. Hardy-Weinberg Equilibrium (HWE) was used to detect allelic frequencies and χ2 was used to detect significance of data among genotypic frequencies. A compact tool package for analysis and conversion of genotype data and SNP Tools for Microsoft Excel was used.

**RESULTS**

The aim of the study was to detect genetic markers related to MI among Saudi patients and the possible association of any of these markers with the ethnic signature shown in Table 2.

Agarose gel electrophoresis: Gel electrophoresis was performed for the recovered PCR products using 3% agarose gel with SYBR-safe DNA gel stain added to the gel to analyze the size and quality of the PCR products. Moreover, the gel was placed into UV Trans-illuminator to visualize the successive products.

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**RESULTS**

The aim of the study was to detect genetic markers related to MI among Saudi patients and the possible association of any of these markers with the ethnic signature
of Saudi participants. These markers included the variants rs5174, rs11591147, rs2259816, rs111245230, rs3782886 and rs2259820, referring to LRP8, PCSK9, HNF1A, SVEP1, BRAP and HNF1A genes, respectively. The study included a total of 203 individuals, among which 100 were MI patients, while 103 were healthy controls. The mean age of MI patients was 57 years old, whereas that of the control group was 55 years old. Among the group of MI patients, 89 were male and 11 were female of whom 43% were smokers and 57% non-smokers.

The study utilized multiplex SNAPSHOT assay to detect SNPs (or variants) that might be associated with MI. This early detection assay is needed to screen the individuals at high risk of the disease, considering the time factor and sensitivity of molecular diagnostic tools. The genotypes referring to the six genes were identified across the 203 participants and data is shown in Table 3. As a model, two typical GeneMapper electropherograms describing the most

### Table 3. Genotypes and the numbers of the MI patients along with the healthy controls and the frequencies of the rare alleles of variants used in the study

| Gene     | Variant   | SNP-based genotype | Total no. individuals | Rare allele frequency |
|----------|-----------|--------------------|-----------------------|----------------------|
|          |           |                    | MI patient | Healthy control | MI patient | Healthy control |
| LRP8     | rs5174    | AA                 | 1          | 2                | 0.01       | 0.03             |
|          |           | GA                 | 0          | 1                |            |                  |
|          |           | GG                 | 93         | 95               |            |                  |
| PCSK9    | rs11591147| AA                 | 16         | 19               | 0.28       | 0.41             |
|          |           | CA                 | 20         | 44               |            |                  |
|          |           | GT                 | 56         | 36               |            |                  |
| HNF1A    | rs2259816 | AA                 | 20         | 28               | 0.48       | 0.55             |
|          |           | CA                 | 49         | 52               |            |                  |
|          |           | CC                 | 24         | 19               |            |                  |
|          |           | CC                 | 0          | 0                | 0.03       | 0.04             |
| SVEP1    | rs111245230| TC                 | 6          | 8                |            |                  |
|          |           | TT                 | 86         | 91               |            |                  |
| BRAP     | rs3782886 | GG                 | 0          | 0                | 0.00       | 0.00             |
|          |           | CC                 | 0          | 0                |            |                  |
| HNF1A    | rs2259820 | TT                 | 13         | 21               | 0.41       | 0.47             |
|          |           | CT                 | 51         | 50               |            |                  |
|          |           | CC                 | 29         | 27               |            |                  |

*Rare allele, **Complementary rare allele

### Table 4. Results of chi square ($\chi^2$) and odd ratio (OR) within the 95% confidence interval (CI) of six selected variants

| Variant   | OR   | 95% CI   | $\chi^2$ value | $P$  |
|-----------|------|----------|----------------|------|
| rs5174    | 1.96 | 0.17     | 21.96          | 1.272|.53   |
| rs11591147| 1.85 | 0.84     | 4.05           | 13.366|.0013 |
| rs2259816 | 1.77 | 0.77     | 4.06           | 1.818|.403  |
| rs111245230| 0.95 | 0.06    | 15.35          | N/A* | N/A   |
| rs3782886 | 0.95 | 0.06    | 15.40          | N/A† | N/A   |
| rs2259820 | 1.74 | 0.73     | 4.13           | 1.834|.40   |

*As calculation of one column is zero, $\chi^2$ cannot be computed
†As calculations of two columns are zero, $\chi^2$ cannot be computed
Table S1. Characteristics of subjects involved in the study in terms of numbers, gender, smoking habits

| Group         | No. | Mean | Percentage |
|---------------|-----|------|------------|
| Age Control   | 103 | 55   | 54%        |
| Patients      | 100 | 57   | 57%        |
| Gender        |     |      |            |
| Male Patients | 89  | 57   | 89%        |
| Female Patients | 11 | 11   | 11%        |
| Smoker        |     |      |            |
| Male Smoker   | 43  | 57   | 43%        |
| Female Smoker | 11  | 11   | 11%        |
| Nationality   |     |      |            |
| Patients      |     |      |            |
| Male Patients | 89  | 57   | 89%        |
| Female Patients | 11 | 11   | 11%        |
| Non-smoker    |     |      |            |
| Male Non-smoker | 43 | 57   | 43%        |
| Female Non-smoker | 11 | 11   | 11%        |

Common patterns of healthy controls and MI patients are shown in Figure 1. The genotypes based on SNaPshot patterns of these two participants are GG/CA/CC/TT/A/A/CC and GG/CC/CA/TT/A/A/GT, referring to SNPs of the six genes LRP8, PCSK9, HNF1A, SVEP1, BRAP and HNF1A genes, respectively. Four out of the six synthesized SBE primers are reverse primers of the six genes. These four primers target sequences of the LRP8, PCSK9, HNF1A, and BRAP genes. Then, the actual genotypes of the two individuals in Figure 1 are CC/GT/GG/TT/TT/CC and CC/GG/TT/TT/CT. Based on the information available in the gene bank (https://www.ncbi.nlm.nih.gov), rare alleles of the six studied genes are complementary allele T (e.g., A allele), complementary allele T (e.g., A allele), complementary allele T (e.g., A allele), complementary allele C (e.g., G allele), and T allele, respectively. Interestingly, rare allele of variant rs111245230 (e.g., complementary allele T) was almost non-existent in the Saudi population, indicating this variant is di-allelic or harbor only two alleles (common and rare) that are conserved as an ethnic signature of the Saudi population, while harbor multiple alleles in populations with other ethnic backgrounds. Additionally, the variant rs3782886 was shown to be mono-allelic (e.g., complementary allele T) in the Saudi population that displayed only the homozygous genotype of complementary T allele (e.g., AA genotype) indicating complete absence of the rare allele in both homozygous or heterozygous conditions (Table 3). As per the association of the different variants and MI patients, the statistical analysis indicated that the rare allele of the rs11591147 variant (e.g., A allele) significantly is reduced ($P = .0013$) in MI patients (rare allele frequency = 0.28) as compared with that of the healthy controls (rare allele frequency = 0.41) (Table 4). The odd ratio (OR), as a relative measure of effect, of the common allele was estimated as 1.85 with a confidence interval (CI) of 0.84-4.05 at 95% significance level. The OR value indicates that MI patients are much more likely to carry the common allele of PCSK9 gene than the healthy control. The confidence interval (CI) indicates the level of uncertainty around the measure of effect. CI was measured in the present study because we recruited a small sample of the overall Saudi population. Therefore, by having an upper and lower confidence limit, we can infer that the true population effect lies between these two points. The results of the two variants rs2259816 and rs2259820 also indicated reductions in the rare alleles (e.g., A and T, respectively), but not at the statistical level. Further analysis of the two variants with larger number of participants might indicate otherwise. Thus, we can confidently conclude that the nonappearance of the complementary allele T (e.g., A allele) of variant rs11591147 in a given individual is a strong risk factor of the occurrence of the MI disease.

**DISCUSSION**

Six SNPs or variants in the present study were selected on the basis of multiple recent large-scale genome wide association studies showing strong statistically significant association with MI risk. The main focus of the study was on investigating the association between the genetic markers rs5174, rs11591147, rs2259816, rs111245230, rs3782886 and rs2259820, referring to LRP8, CSK9, HNF1A, SVEP1, BRAP and HNF1A genes, respectively, and MI among Saudi patients. The mean age of MI patients in the present study was 57 years old. A similar study conducted by Kathiresan et al showed that the majority of the MI cases were observed among the individuals who are > 65 years old [Kathiresan 2009]. Another study conducted by Maas et al showed that cardiovascular disease develops 7-10 years early among men as compared to women [Maas 2010]; however, it still is the major cause of death in women over the age of 65 years. The genotyping method used in the present study is SnapShot assay that assisted in screening the six SNPs, simultaneously with single reaction in individual samples. Similar to the present study, Ben et al determined the genotyping of two genes, CYP2D6 and ADRB1, by using SNaPshot assay [Ben 2016]. The results showed that this technique is simple, efficient, and accurate in detecting the genotypes based on these two genes in significantly reduced costs. Another study conducted by Ghosh et al found that SNaPshot technique is an informative approach deeply helpful in detecting parent- and yet stage-of-origin of nondisjunction and underscores the need for characterizing additional markers [Ghosh 2012].

The output of 40 random samples genotyped by SNaPshot method was validated by Sanger sequencing (data not shown). Moreover, the study achieved an agreement of 100% with SNaPshot for the six SNP analyzed in these 40 samples. In a similar context, significant association with CAD and MI was
The present study has examined the association between MI and SNPs of six genes related to the disease. It has also evaluated the gene variants as genetic risk factors to MI among patients in Saudi Arabia. The principal finding of the study is SNP rs11591147 working as a protective allele against the MI disease in Saudi Arabia, which means that the rare allele (e.g., A) frequency of this SNP was significantly higher in the healthy control individuals and lower among the patients group. The results also showed that Multiplex SNaPShot assay is a rapid, reliable, flexible, accurate technique and cost-effective to detect risk alleles of any complex diseases in the Kingdom of Saudi Arabia or in any other ethnic groups. However, the study results recommend conducting future studies with a larger sample size and multi centers for reliable statistical analysis and more significant results. It also recommends the creation of local genomic variants database from a healthy Saudi population that will give local distribution for the selection of variants in such association studies for any complex diseases such as MI, celiac, obesity, or diabetes.

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