The effect of pyrolysis temperature and feedstock on date palm waste derived biochar to remove single and multi-metals in aqueous solutions

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Abstract

In this study, leaf and frond date palm waste as feedstock was used to derive biochars. The effects of pyrolysis temperatures on their physical and chemical properties, and their capacity to remove copper, iron, nickel and zinc from single and multi-metal solutions at various pH values were investigated. Analytical and spectroscopic techniques such as scanning electron microscopy, energy dispersive X-ray, Fourier transform infrared spectroscopy, X-ray diffraction, carbon, hydrogen, nitrogen, sulfur elemental analysis, Brunauer Emmett Teller analysis were conducted for characterization. The pore volume, surface area, pH, and total carbon content of date palm leaf and frond biochar increased while functional groups and hydrogen, nitrogen and oxygen content of biochar decreased with increasing pyrolysis temperature compared to feedstock. The removal efficiencies and sorption capacity for single and mixed metal ions were found between 98 and close to 100% and 2.4 and 3.0 mg g\(^{-1}\) by leaf and frond biochar samples at pH > 6, respectively. Biochar obtained from different feedstock at different pyrolysis temperature did not show any statistically significant improvements on the removal of single or mixed metals from aqueous solutions. The date palm leaf or frond biochar obtained at low pyrolysis temperature is as effective to remove metals as the ones obtained at high pyrolysis temperatures. Therefore, to consume less energy to produce biochar at lower temperature which exhibits same effective removal efficiency will be a win-win solution in terms of sustainability and economy. As a result, date palm waste biochar can be effectively used to remove metals in water and wastewater.

Keywords: Biochar, Charcoal, Date palm, Frond, Leaf, Mixed metals

Introduction

Metals in the environment originate from a wide variety of anthropogenic sources such as battery manufacturing, power plants, fertilizers, pesticides, mining, wastewater, smelting, and sewage sludge. Metals do not biodegrade and are highly toxic due to their bioavailability [1]. When heavy metals discharged into the environment, they have a tendency to bio-accumulate and result in cancer, pancreatic damage, and stomach and intestinal distress to humans [2]. They can be phytotoxic, and limit agricultural applications such as use of wastewater sludge in fields [3]. Therefore, there is an emerging need to treat the metal contaminated wastewater with appropriate technologies. Several conventional methods are already in use such as membrane filtration, chemical precipitation, ion exchange and electrodialysis to treat heavy metal loaded waters. However, some of these methods are often not economical due to energy requirement; some of them produce toxic by-products and sludge, and in some cases may result in membrane
fouling. According to Amin et al. [4], they also tend to be ineffective in the treatment of wastewaters with high heavy metal concentrations. Adsorption, on the other hand, is simple and easy technique to operate for treatment purposes. Nevertheless, the use of commercial adsorbents for treatment systems is limited due to ensuing high cost. Therefore, development of novel adsorbent materials that are efficient, cheap, easy to grow and harvest, and abundant in nature is crucial [5]. Recently, biochar has gained popularity as a biosorbent material for the removal of a variety of pollutants from contaminated waters because of its low cost, wide availability, and desirable physical and chemical surface features as adsorbent [6].

Biochar is a black carbonaceous product of biomass such as wood, plant, or sludge when heated at temperatures greater than 250 °C in the absence of air or in limited air [7]. Biochar is considered alkaline; it is composed of mainly carbon and small amounts of minerals and volatile organics and is resistant to biodegradation [8]. It consists of 40–75% carbon, and it is not easily broken down by microorganisms [9]. Moreover, biochar is porous, and it has a large surface area and a variety of functional groups on the surface [10].

Altering pyrolysis condition affects the composition of the biochar [11]. Slow pyrolysis is often associated with the formation of higher contents of nitrogen, sulfur, calcium, magnesium and phosphorus in addition to greater surface areas and enhanced cation exchange capacities. This is due to the fact that easily decomposable and volatile components of biochar such as oxygen, hydrogen, nitrogen, total phosphorus and sulfur are lost during the slow pyrolysis [7]. Chemical changes are introduced to biochar during the pyrolysis at various temperatures. As the biomass gets dehydrated, aliphatic bonds are converted into aromatic bonds which are consolidated into stable graphene structures [12].

Sun et al. [13] have shown that the pyrolysis temperature and the type of feedstock are influential factors on the production rate, thermal stability, carbon content and elemental composition of biochar. In a similar fashion, at higher pyrolysis temperatures, the surface area and porosity increase along with the concentration of minerals including potassium, phosphorus, calcium and magnesium on the surface of the adsorbent which in turn would allow ion exchange with metals and result in higher adsorption capacities [14]. Also, as the pyrolysis temperature increases, some physical and chemical changes occur in biochar such as a decrease in cell pore diameter and an increase in the specific surface area [7].

Functional groups on the surface of adsorbents are responsible for the sorption of adsorbates [15]. According to Li et al. [16], the functional groups responsible for sorption include carboxyl, hydroxyl and amino groups, and the concentration of these groups often decreases with an increase in the pyrolysis temperature as compared to the feedstock.

Studies have demonstrated that biochars derived from a variety of sources such as paper mill waste, corn, willow, hazelnut, pine, oak, and manure have potential to remove a number of contaminants in wastewaters such as dyes, pharmaceuticals, anions, pesticides and heavy metals [7]. Table 1 reports studies that used biochar originating from a variety of sources to remove metals [12, 16–31]. From the Table 1, according to Chen et al. [17], hardwood and corn stalk biochar showed 95% of copper and 90% of zinc removals. Alkali activated hydrothermal carbonization (HTC) biochar, removed almost 100% of copper [18]. Pellera et al. [19] studied compost, rice husks, olive pomace and orange waste biochar to remove copper and found that compost biochar had the highest removal (94%) followed by rice husks (90%), orange waste (89%) and olive pomace (78%).

Utilizing waste materials to produce biochar is a sustainable solution in terms of cost effectiveness and impact on global warming potential (GWP). Alhashimi and Aktas [32] compared the cost of biochar and activated carbon and they found that granular activated carbon and powdered activated carbon cost 6.4 and $1.2–2 kg$^{-1}$ respectively as compared to coconut shell biochar, pine wood biochar and forestry debris biochar which cost 0.8, 0.9 and $1.5 kg^{-1}$ respectively. In terms of environmental impact, Alhashimi and Aktas [32] deducted that biochar had a negative impact on GWP with a value of −0.9 kg CO$_2$-eq kg$^{-1}$ as compared to activated carbon (6.6 kg CO$_2$-eq kg$^{-1}$). Similarly, according to Roberts et al. [33], the net greenhouse gas emissions for biochar from stover and yard waste were also negative, at −864 and −885 kg CO$_2$-eq emissions reductions per ton of dry feedstock. This is due to the ability of biochar to remove carbon as compared to activated carbon. The energy requirements for biochar were lower requiring only 6.1 MJ kg$^{-1}$, whereas, activated carbon required 97 MJ kg$^{-1}$ [32].

Therefore, utilizing date palm waste as a low-cost adsorbent will be a sustainable waste utilization solution, since the date palm waste is abundant in the Gulf Cooperation Council Countries. In fact, annually, 50 kt of wastes are generated from date palm trees which end up in landfills in the United Arab Emirates [34]. Similarly, wastes from 22 million date palms are disposed directly in the landfills or burns in open fields in the Kingdom of Saudi Arabia [35]. Accordingly, the objective of this study is to bring attention to utilize date palm leaf and palm frond as biochar/biomass feedstock at different pyrolysis temperature to remove single and mixed metals in wastewater. Date palm leaf and palm frond as
biochar were not utilized widely in wastewater treatment applications. In addition, the available biochar studies mostly deal with single solute removal. There are a few studies dealing with mixed ion solutes removal. As a matter of fact, multiple ions are present in wastewater; therefore, it is vital to study removal of single as well as mixed ions.

| Study | Adsorbents | Adsorbates | Removal efficiency [adsorption capacity] | Reference |
|-------|------------|------------|-----------------------------------------|-----------|
| 1     | Buck wheat, corn cobs, mulberry wood, poultry manure and peanut shells BC pyrolyzed at 350, 450, 550 and 650 °C | (1) Lead (2) Cadmium (3) Arsenic | (1) 97.5% [2.5 mg g⁻¹] (2) < 90% [1.5 mg g⁻¹] (3) 42% [0.5 mg g⁻¹] | [12] |
| 2     | Rape straw BC-NaOH | Cadmium | 90% [72 mg g⁻¹] | [16] |
|       | Rape straw BC-MnOₓ | Arsenic | 47% [67 mg g⁻¹] | |
| 3     | Hard wood and corn stalk BC | Copper | 95% | [17] |
|       | | Zinc | 90% | |
| 4     | Activated HTC BC | (1) Copper (2) Cadmium | (1) 100% [31 mg g⁻¹] (2) 100% [34 mg g⁻¹] | [18] |
|       | HTC BC | Cadmium | (1) 16% [4.0 mg g⁻¹] (2) 5.6% [1.5 mg g⁻¹] | |
| 5     | Compost BC | Copper | 93.6% [3.6 mg g⁻¹] | [19] |
|       | Rice husks BC | Copper | 90.1% [2.5 mg g⁻¹] | |
|       | Olive pomace BC | Zinc | 77.8% [2.2 mg g⁻¹] | |
|       | Orange waste BC | Arsenic | 88.7% [3.3 mg g⁻¹] | |
| 6     | Non-impregnated rice husk BC | Arsenic | 25% | [20] |
|       | Ca impregnated rice husk BC | Arsenic | 95% | |
|       | Fe⁰ impregnated rice husk BC | Arsenic | 58% | |
| 7     | Soybean stalk BC | Mercury | 86.4% [0.7 mg g⁻¹] | [21] |
| 8     | Oak bark BC | Arsenic | 70% [7.4 mg g⁻¹] | [22] |
|       | | Cadmium | 50% [5.4 mg g⁻¹] | |
|       | | Lead | Nearly 100% [13.1 mg g⁻¹] | |
| 9     | Sugar beet tailings BC | Chromium (VI) | 98% [123 mg g⁻¹] | [23] |
| 10    | Pinewood (P) BC | Lead | P: Nearly 80% [4.1 mg g⁻¹] | [24] |
|       | Rice husk (R) BC | R: Nearly 60% [2.4 mg g⁻¹] | |
| 11    | Chitosan modified bamboo BC | Lead | 50% [25.1 mg g⁻¹] | [25] |
|       | | Cadmium | 45% [14.3 mg g⁻¹] | |
|       | | Copper | Close to 55% | |
| 12    | Olive mill solid waste BC | Lead | > 80% | [26] |
|       | | Copper | > 80% | |
|       | | Selenium | < 20% | |
| 13    | Groundnut shell BC | Cadmium | 99.93% [0.14 mg g⁻¹] | [27] |
|       | | Mercury | 99.99% [0.14 mg g⁻¹] | |
|       | | Lead | 100% [0.18 mg g⁻¹] | |
| 14    | Groundnut shell BC | Cadmium | 99.20% [29.0 mg g⁻¹] | [28] |
| 15    | Corn straw BC | Cadmium | 99.24% [38.9 mg g⁻¹] | [29] |
| 16    | Dairy manure derived BC | Lead | 92.8~99.8% | [30] |
| 17    | Carbon microspheres derived from walnut shell | Chromium | 792 mg g⁻¹ | [31] |
|       | | Lead | 638 mg g⁻¹ | |
|       | | Cadmium | 574 mg g⁻¹ | |
|       | | Copper | 345 mg g⁻¹ | |

BC Biochar
multi-ion solutes in wastewater [36]. Moreover, studies dealing with different feedstock and pyrolysis effect on biochar to remove mixed metals using statistical analysis are needed. In this study, physical and chemical features of date palm leaf/frond biochar obtained at different pyrolysis temperatures (400, 500 and 600 °C) were characterized, and the effects of feedstock and pyrolysis temperatures on the removal efficiencies of heavy metals such as copper, zinc, nickel and iron in mixed metal and single metal solutions at different pH values were investigated statistically. The result of this study may help and enable end-users to apply date palm-derived biochar as a low-cost adsorbent in their treatment systems.

Materials and methods

Preparation of biochar and stock solutions

The biochar was composed of date palm waste, namely fronds and leaves. The fronds and leaves were dried under the sun. The fronds were chopped to 1 cm and the leaves were chopped to 0.5 cm length. The moisture and volatility contents were determined according to The American Society for Testing and Materials D1762–84 (Standard Test Method for Chemical Analysis of Wood Charcoal). The date palm fronds/leaves were placed in a stainless-steel container and packed tightly to ensure no room for air, and covered with aluminum foil. The container was placed in an electrical muffle furnace (Vulcan 3–550) and pyrolyzed as shown in Fig. 1 at 400, 500 and 600 °C at a rate of 8 °C min⁻¹ temperature increment. After completion of pyrolysis, the biochar samples were left to cool for 3 to 4 h inside the furnace. The date palm leaf and frond biochar samples were labeled as Frond 400 (frond was pyrolyzed at 400 °C), Frond 500 (pyrolyzed at 500 °C), Frond 600 (pyrolyzed at 600 °C), Leaf 400 (leaf was pyrolyzed at 400 °C), Leaf 500 (pyrolyzed at 500 °C) and Leaf 600 (pyrolyzed at 600 °C). Later, the date palm frond and leaf biochar samples were ground and sieved to obtain a particle size of about 0.15 mm. The sieved biochar samples were washed with distilled water and dried at 105 °C for 2 h and placed inside air-tight containers in desiccators. Stock solutions of copper, nickel, zinc and iron of 5 mg L⁻¹ were prepared using copper sulfate pentahydrate, nickel chloride hexahydrate, zinc chloride and iron sulfate heptahydrate for the single solute and mixed metal solutes. All chemicals were of analytical grade and provided by Sigma Aldrich and Merck. The pH of the solutions was adjusted using 0.1 M HCl and 0.1 M NaOH.

Characterization of the adsorbent

The characterization of the date palm frond and leaf biochar samples at different pyrolysis temperatures (400, 500, 600 °C) were shown in Fig. 1. The characterization included physical and chemical features of the biochar.
500 and 600 °C) was performed using a variety of analytical techniques. Scanning electron microscopy (SEM) with a JEOL JSM-7610F was used to determine the morphology of the adsorbent. An accelerating voltage between 10 and 12 kV with low probe current and a working distance of 8–15 mm was maintained during the analysis. Energy dispersive X-Ray (EDX) was used to obtain the elemental analysis of the biochar surface. X-Ray diffraction (XRD) spectroscopy with a D2 Phaser Bruker dieractometer with CuKα radiation of wavelength 0.154 nm was conducted to obtain the mineral composition of the samples. Fourier transform infrared spectroscopy (FT-IR) with a Bruker Vertex 80v was used to identify the different functional groups present in the samples represented by their vibrational modes. Surface area was analyzed using nitrogen adsorption at 77 K with a Quantachrome Autosob-1. The Brunauer–Emmett–Teller (BET) method was used to calculate the specific surface area. The Carbon, Hydrogen, Nitrogen, Sulfur (CHNS) content was analyzed using an elemental analyzer with a EuroEA Elemental Analyzer in which the samples were combusted at 980 °C, and the resultant gaseous species separated quickly allowing the detection of thermal conductivity. The pH of biochar samples was measured with a pH meter, HANNA instruments 2210, using biochar/water ratio of 1/10 (v:v).

Removal studies
The effect of pyrolysis temperature and the effect of using either date palm frond or leaf derived biochar samples on the removal efficiencies of metal ions from the single and multi-metal solutions were studied in batch studies. In the experiments, 0.1 g of date palm frond or leaf biochar samples (namely, Frond 400, Frond 500, Frond 600, Leaf 400, Leaf 500 and Leaf 600) were added to solutions of 50 mL of 5 mg L⁻¹ of iron, nickel, copper and zinc of single or mixed metal solutions. 0.1 M of NaOH and 0.1 M of HCl were used to adjust the pH of the solutions at 2, 4, 6, 7, 8 and 10 using a pH meter. 50 mL of solutions were shaken at 200 rpm for 20 h using a shaker, Bench Mark OrbiShaker. All samples were filtered using 0.45 μm Whatman filter paper. The filtrates were analyzed to determine the concentrations of the metals in solutions using an inductively coupled plasma mass spectrometer (ICP-MS), NexION 350 X dual channel. Removal efficiency was calculated as follows:

\[
\% \text{removal} = \frac{C_0 - C_e}{C_0} \times 100
\]  

(1)

where, \(C_0\) = initial concentration, \(C_e\) = final concentration

To calculate the metal sorption capacity \(q_e\) (mg g⁻¹) of the biochar, the following formula was used:

\[
q_e = \frac{(C_0 - C_e) \times V}{m}
\]

(2)

where, \(V\) is the sample volume and \(m\) the weight of the adsorbent.

Data analysis
Student’s t-test, two-tailed with a 95% confidence interval, and one-way analysis of variance (ANOVA) tests with 95% confidence interval were used to determine whether the increase of the pyrolysis temperature for the preparations of date palm frond or leaf derived biochar samples showed any significance on the metal ion removal efficiencies. T-test analysis was also used to investigate if different feedstocks namely date palm frond or leaf at different pyrolysis temperatures made any significance on the removal efficiencies of the metals.

Results and discussion
Characterization of the samples
The moisture contents of the date palm frond and leaf as feedstock were found to be 2.5 and 1.8% for date palm frond and leaf feedstock respectively. The volatile matter and lignin. Furthermore, the leaf and frond feedstocks had an integrated structure without considerable pores as compared to the pyrolyzed biochar [38]. In date palm frond feedstock, the surfaces were curly indicating the presence of cellulose, hemicelluloses and lignin. Furthermore, the leaf and frond feedstocks had an integrated structure without considerable pores as compared to the pyrolyzed biochar [38]. In date palm frond feedstock, CuKα Ray diffraction (XRD) spectroscopy with a D2 Phaser Bruker diffractometer with CuKα radiation of CuKα 1.54 nm was conducted to obtain the mineral composition of the samples. Fourier transform infrared spectroscopy (FT-IR) with a Bruker Vertex 80v was used to identify the different functional groups present in the samples represented by their vibrational modes. Surface area was analyzed using nitrogen adsorption at 77 K with a Quantachrome Autosob-1. The Brunauer–Emmett–Teller (BET) method was used to calculate the specific surface area. The Carbon, Hydrogen, Nitrogen, Sulfur (CHNS) content was analyzed using an elemental analyzer with a EuroEA Elemental Analyzer in which the samples were combusted at 980 °C, and the resultant gaseous species separated quickly allowing the detection of thermal conductivity. The pH of biochar samples was measured with a pH meter, HANNA instruments 2210, using biochar/water ratio of 1/10 (v:v).
Table 2  Elemental analysis for leaf and frond feedstock and biochar samples

| Sample          | EDX analysis | CHNS analysis |
|-----------------|--------------|---------------|
|                 | %C | %O | %Mg | %Si | %S | %Cl | %Ca | %P | %Na | %P | %Al | %N | %C | %H | %S | %O |
| Leaf feedstock  | 37.7 | 57.9 | 0.5 | 1.5 | 0.7 | 0.4 | 1.0 | 0 | 0 | 0 | 0.3 | 0.6 | 31.2 | 4.5 | 0 | 63.7 |
| Leaf 400        | 44.7 | 39.7 | 0.4 | 10.3 | 0.4 | 0.4 | 3.9 | 0.3 | 0 | 0 | 0 | 0.6 | 47.8 | 3.0 | 0 | 48.6 |
| Leaf 500        | 49.1 | 35.7 | 0.4 | 10.5 | 0.5 | 3.8 | 0.3 | 0.4 | 0 | 0 | 0 | 0.6 | 49.2 | 2.1 | 0 | 48.1 |
| Leaf 600        | 67.3 | 25.0 | 0.5 | 3.4 | 0 | 0 | 3.9 | 0 | 0 | 0 | 0 | 0.6 | 57.7 | 1.0 | 0 | 29.1 |
| Frond feedstock | 45.5 | 47.6 | 0 | 0 | 2.4 | 4.6 | 0 | 0 | 0 | 0 | 0 | 0.2 | 41.9 | 5.8 | 0 | 52.1 |
| Frond 400       | 66.9 | 23.3 | 0.8 | 1.5 | 0.8 | 1.0 | 4.6 | 0.7 | 0.5 | 0 | 0 | 0.1 | 46.9 | 3.5 | 0 | 49.8 |
| Frond 500       | 72.0 | 17.8 | 0.9 | 1.6 | 0.6 | 1.3 | 4.2 | 1.0 | 0.7 | 0 | 0 | 0.1 | 50.9 | 3.0 | 0 | 45.6 |
| Frond 600       | 74.8 | 11.5 | 0.4 | 1.0 | 4.3 | 5.0 | 1.0 | 1.6 | 0 | 0 | 0.1 | 66.7 | 2.3 | 0 | 30.9 |
calcination of calcium carbonate to soluble calcium oxide [35]. For Frond 400 and 500 samples, magnesium content increased with an increase in the pyrolysis temperature. This is mainly due to the presence of insoluble periclase, magnesium oxide [7, 35]. The presence of the other elements such as silicon and aluminum can be attributed to metallic oxides present in the biochar samples [39]. Both CHNS and EDX analyses shown in Table 2 showed that as the pyrolysis temperature increased, the carbon content of leaf and frond biochar samples increased compared to feedstock due to the carbonization and the thermochemical decomposition of the biomass in which cellulose, hemicellulose and lignin break down, and new resistant graphitic bonds form [39]. Carbon content of leaf feedstock was 31% and increased to 47–57% in leaf biochar samples. Similarly, the carbon content of frond feedstock was 42% and increased to 47–67% for frond biochar samples. It was observed that oxygen content decreased in the leaf and frond biochar samples compared to leaf and frond feedstock with an increase in the pyrolysis temperature due to the volatilization [7]. Oxygen content of leaf feedstock was 64% and decreased to 29–49% for leaf biochar samples. The oxygen content of frond feedstock was 52% and decreased to 30–50% for leaf biochar samples. Comparably, hydrogen content decreased in leaf and frond biochar samples compared to leaf and frond feedstock with an increase in the pyrolysis temperature. The hydrogen content of leaf feedstock was 4.5% and decreased to 1.0–3.0% for leaf biochar samples. The hydrogen content of frond feedstock was 5.8% and decreased to 2.3–3.5% for frond biochar samples. Nitrogen content relatively unchanged between leaf/frond feedstock and leaf/frond biochar samples, which is consistent with the findings of other studies [13, 40].

Figures 3a, b show the FT-IR spectra of leaf and frond feedstock and frond and leaf biochar pyrolyzed at 400, 500 and 600 °C. The band at 1620 cm\(^{-1}\) represented alkanyl C=C stretching vibrations. It can also represent H-O-H bending band of water which tends to decrease with an increase in pyrolysis temperature. The band at 1701 cm\(^{-1}\) represented C=O from -COOH, amides, esters and ketones [39]. The band at 1724 cm\(^{-1}\) represented esters and aldehydes. The furthest bands at 1764 and 1811 cm\(^{-1}\) represented alkyl carbonate and aryl carbonate respectively [41]. It can be noticed that the alkyl carbonate and aryl carbonate bands for the frond feedstock are slightly lower than those of Frond 400. According to Usman et al. [35], biochar pyrolyzed at low temperatures have some functional properties similar to their feedstock. FT-IR analysis showed that chemical changes occurred in biochar during pyrolysis at various temperatures. The leaf and frond feedstocks were dehydrated upon increases in the pyrolysis temperature, whereby aliphatic bonds were converted to aromatic bonds which become stable graphene [12]. The content of carboxyl, hydroxyl and amino groups which are responsible for the sorption in leaf and frond biochar samples [42], was decreased with an increase in the pyrolysis temperature as compared to the feedstocks as shown in Fig. 3. Various functional groups such as oxygen containing functional groups present in the feedstocks and frond and leaf biochar samples can impact reactions on the surface, hydrophilicity, and electrical and catalytic properties of the samples [43].

The XRD analysis confirmed the organic mineral melilite, \(\text{Al}_2[\text{C}_6(\text{COO})_6] \cdot 16\text{H}_2\text{O}\), at 21.2° for the leaf feedstock and 22° for the frond feedstock. However, melilite was lost during the pyrolysis and it could not be identified in any of the leaf and frond biochar samples. The peaks at 39 and 42° in Frond 600 were identified as ZnS. In Frond 500 and 400, the peaks at 41° were identified as \(\text{CaCO}_3\) as well as the peak at 48° in Leaf 500. At higher temperatures, calcium concentration decreases due to the calcination of calcium carbonate to soluble calcium oxide [22]. Hence, no \(\text{CaCO}_3\) peaks were identified in Leaf 600 and Frond 600. Sr\(_2\)FeNbO\(_6\) was identified at peak 47° in Frond 500. NaBr was found at peak 28° in Leaf 500 and 600. Broad peaks at 30° in Leaf 400 was indicative of amorphous carbon. The list of minerals identified along
with their peak positions for leaf and frond feedstock is given in Fig. 4a and that of frond and leaf biochar in Fig. 4b. The presence of carbonate, calcite, and quartz, in the samples was probably due to the soil particles mixed with the leaf and frond feedstocks during the picking date palm waste from source or from the entrapment of carbon dioxide gas during the pyrolysis. In the Emirate of Abu Dhabi, soils are broadly classified into sandy, sandy-calcareous, gypsiferous, saline, saline-gypsiferous and hard pan soils. Calcareous or CaCO$_3$ containing soils originate from either the weathering of the parent material or dust rich with CaCO$_3$ brought over from the desert [44]. The source of calcium oxalate on leaf and frond biochar samples is phytolite found in leaves, wood and the bark of plants, and as the pyrolysis temperature increases, phytolite is converted into calcite [39]. Sr$_2$FeNbO$_6$ and NaBr originate from date palm leaf and frond. According to the previous study [45], the mineral composition of palm leaves includes a significant concentration of Na and Fe.

The BET surface areas of frond feedstock, Frond 400, 500 and 600 were 1.1, 3.8, 208 and 246 m$^2$ g$^{-1}$, respectively and the BET surface areas for leaf feedstock, Leaf 400, 500 and 600 were 1.9, 3.5, 152 and 232 m$^2$ g$^{-1}$, respectively as shown in Table 3. The BET results of frond and leaf biochar samples were in parallel with the other studies as shown in Table 3 [34, 37, 46–48]. The BET surface area for date palm feedstock was reported to be in the range of 1-4 m$^2$ g$^{-1}$ [34, 37, 47]. The date palm biochar pyrolyzed between 300-700 °C showed BET surface areas of 5.5-249 m$^2$ g$^{-1}$ [46]. BET analysis reveals that as pyrolysis temperature increases from 400 to 600 °C, date palm leaf and frond biochar BET surface area increases as well. At higher pyrolysis temperature, aliphatic alkyls and ester functional groups disappeared upon exposing to the aromatic lignin core [49].

![Fig. 4 XRD spectra of a leaf-frond feedstocks and b leaf-frond biochar samples](image-url)
The pore size distribution of leaf and frond biochar pyrolyzed at 400, 500 and 600 °C is shown in Fig. 5. It can be deduced that as the pyrolysis temperature increases, the pore volume increases and the pore diameter decreases. With increasing pyrolysis temperature, the organic matter decomposes resulting in the formation of micropores [49]. These results are in parallel with other reported studies. According to Yuan et al. [50], when the pyrolysis temperature of biochar made from sewage sludge increased from 300 to 700 °C, the pore volume increased from 0.08 – 0.13 mL g\(^{-1}\). According to Zhao et al. [51], the pore diameter for biochar pyrolyzed at 500 and 650 °C was 4.0 and 2.2 nm, respectively. As more labile material is removed, smaller diameter pores begin to form [51].

**Removal studies**

Date palm leaf and frond biochar samples pyrolyzed at different temperatures were tested to remove single copper, iron, zinc and nickel, and mixed metal solution at pH 2, 4, 6, 7, 8 and 10.

**Single metal removal/sorption capacity studies**

It was observed that as the pH increased, copper, zinc, nickel and iron removal efficiencies and sorption capacity (qe) in single metal solution by leaf and frond biochar samples increased as well. Copper removal efficiency and sorption capacity increased from pH 2–6 then the increase in the efficiency/sorption capacity was moderate at pH 7, 8 and 10. The highest average copper removal efficiencies were all similar, or close to 99% (qe: 2.7 mg g\(^{-1}\)) for all pyrolyzed leaf and frond biochars at three temperatures at pH 7 as shown in Fig. 6a and Table 4. Similarly, iron removal efficiencies increased from pH 2 to 6, and then a slight increment was observed at pH 7, 8 and 10. The highest average iron removal efficiencies were between 97 and 99% (qe: 2.7–2.9 mg g\(^{-1}\)) all for all pyrolyzed leaf and frond biochars at three temperatures at pH 7 and pH 6 as shown in Fig. 6b and Table 4. In parallel to iron and copper, nickel removal efficiency increased as the pH was increased. The highest average removal efficiencies for nickel were 82% for Leaf 400 (qe: 2.0 mg g\(^{-1}\)), 98% for Leaf 500 (qe: 2.4 mg g\(^{-1}\)), 99% for Leaf 600 (qe: 2.4 mg g\(^{-1}\)), 96% for Frond 400 (qe: 2.4 mg g\(^{-1}\)), 53% for Frond 500 (qe: 1.3 mg g\(^{-1}\)) and 96% for Frond 600 (qe: 2.4 mg g\(^{-1}\)) at pH 10 as shown in Fig. 6c and Table 4. For zinc, the removal efficiencies increased as the pH was increased to pH 7, 8 and 10. The highest average removal efficiencies were 97% (2.7 mg g\(^{-1}\)) at pH 10, 90% for Frond 400 (qe: 2.7 mg g\(^{-1}\)) and 98% for Frond 600 (qe: 3.0 mg g\(^{-1}\)) at pH 7 as shown in Fig. 6d and Table 4.

**Table 3:** BET analysis results and comparison with biochar derived from palm material

| Biochar derived from palm material | BET surface area (m\(^2\) g\(^{-1}\)) | Reference |
|-----------------------------------|-------------------------------------|-----------|
| Feedstock-palm frond              | 1.0                                 | [34]      |
| Biochar-palm frond                | 2.0                                 |           |
| Feedstock-palm frond              | 4                                   | [37]      |
| Biochar-palm frond                | 9                                   |           |
| Activated carbon                  | 322                                 |           |
| Feedstock-date palm waste         | 1.0                                 | [46]      |
| Date palm biochar at 300 °C        | 2.0                                 |           |
| Date palm biochar at 400 °C        | 5.5                                 |           |
| Date palm biochar at 500 °C        | 124                                 |           |
| Date palm biochar at 600 °C        | 221                                 |           |
| Date palm biochar at 700 °C        | 249                                 |           |
| Feedstock-date palm fibers         | 1.5                                 | [47]      |
| Date palm rachis biochar at 300 °C | 15.8                                | [48]      |
| Date palm rachis biochar at 700 °C | 268                                 |           |
| Date palm frond feedstock          | 1.1                                 | This study|
| Biochar Frond 400                  | 3.8                                 | This study|
| Biochar Frond 500                  | 152                                 | This study|
| Biochar Frond 600                  | 246                                 | This study|
| Biochar Leaf 400                   | 3.5                                 | This study|
| Biochar Leaf 500                   | 152                                 | This study|
| Biochar Leaf 600                   | 289                                 | This study|
| Date palm leaf feedstock           | 1.9                                 | This study|
Mixed metal solutions

In a similar fashion, it was observed that when pH was increased, copper, zinc, nickel and iron removal efficiencies in mixed-metal solutions increased by leaf and frond biochar samples. Copper removal efficiencies were comparably lower between pH 2 and 4. However, a slight increase was observed beyond pH 6. The highest average removal efficiencies were close to 100% for all leaf and frond biochars (qₑ: 2.51–2.7 mg g⁻¹) at pH 7, 8 and 10 as shown in Fig. 6e and Table 4. Iron removal efficiencies increased from pH 2 to 6, and slight increments were observed beyond pH 6. The highest average removal efficiencies were 94–95% for all pyrolyzed leaf and frond biochars at pH 6 and 7 as shown in Fig. 6f and Table 4. Nickel removal efficiencies were low at low pH values (pH 2–4) similar to copper, and removal efficiencies increased when pH was raised beyond 6. The highest average removal efficiencies were close to 100%
The removal efficiencies of a copper, b iron, c nickel, d zinc, in single metal solutions and e copper f iron, g nickel and h zinc in mixed metal solution by biochar samples.
| pH 2 | pH 4 | pH 6 | pH 7 | pH 8 | pH 10 |
|------|------|------|------|------|-------|
| Copper | Iron | Nickel | Zinc | Copper | Iron | Nickel | Zinc | Copper | Iron | Nickel | Zinc | Copper | Iron | Nickel | Zinc | Copper | Iron | Nickel | Zinc |
| Leaf 400 | 0.52 | 0.29 | 1.34 | 0.30 | 2.06 | 1.76 | 2.09 | 2.08 | 2.56 | 2.90 | 1.95 | 2.58 | 2.72 | 2.89 | 2.08 | 2.56 | 2.88 | 1.92 | 2.82 | 2.70 | 2.79 | 2.03 | 2.90 |
| Leaf 500 | 2.44 | 0.29 | 1.55 | 041 | 2.61 | 0.29 | 2.29 | 1.83 | 2.60 | 2.79 | 2.21 | 2.54 | 2.71 | 2.87 | 2.35 | 2.64 | 2.69 | 2.88 | 2.42 | 2.87 | 2.70 | 2.83 | 2.43 | 2.96 |
| Leaf 600 | 2.16 | 0.29 | 0.20 | 061 | 2.55 | 0.44 | 0.93 | 1.27 | 2.70 | 2.87 | 1.57 | 2.62 | 2.70 | 2.80 | 1.86 | 2.85 | 2.71 | 2.86 | 1.92 | 2.80 | 2.71 | 2.61 | 2.44 | 2.97 |
| Frond 400 | 0.33 | 0.45 | 0.24 | 0.50 | 2.58 | 2.67 | 1.19 | 1.49 | 2.47 | 1.51 | 1.37 | 0.45 | 2.72 | 2.83 | 1.59 | 2.69 | 2.54 | 2.74 | 1.60 | 2.28 | 2.70 | 2.85 | 2.38 | 1.61 |
| Frond 500 | 2.42 | 0.29 | 0.52 | 2.24 | 2.60 | 0.44 | 0.86 | 2.50 | 1.82 | 2.84 | 0.98 | 1.23 | 2.73 | 2.81 | 1.13 | 2.60 | 2.01 | 2.84 | 1.73 | 0.93 | 2.72 | 2.84 | 1.31 | 2.71 |
| Frond 600 | 2.42 | 0.29 | 0.47 | 2.44 | 2.53 | 2.77 | 0.69 | 2.86 | 2.71 | 2.84 | 0.72 | 2.36 | 2.73 | 2.85 | 0.79 | 2.95 | 2.59 | 2.84 | 0.95 | 1.63 | 2.72 | 2.85 | 2.38 | 2.91 |
| Mixed metal solutions |
| Leaf 400 | 0.27 | 0.29 | 0.25 | 0.30 | 0.42 | 0.29 | 0.25 | 0.30 | 0.88 | 1.89 | 0.12 | 0.19 | 2.69 | 2.77 | 1.90 | 2.58 | 2.74 | 2.46 | 2.98 | 2.74 | 2.77 | 2.47 | 2.99 |
| Leaf 500 | 0.27 | 0.29 | 0.25 | 0.30 | 0.37 | 1.36 | 0.25 | 0.30 | 1.46 | 2.76 | 0.25 | 0.30 | 2.73 | 2.76 | 2.13 | 2.64 | 2.74 | 2.46 | 2.98 | 0.30 | 2.74 | 2.77 | 2.47 | 2.98 |
| Leaf 600 | 0.27 | 0.29 | 0.25 | 0.30 | 2.73 | 2.76 | 1.25 | 2.41 | 2.74 | 2.76 | 2.24 | 2.73 | 2.73 | 2.76 | 2.47 | 2.99 | 2.73 | 2.76 | 2.19 | 2.73 | 0.29 | 2.73 | 0.28 | 0.42 |
| Frond 400 | 0.27 | 0.29 | 0.25 | 0.30 | 0.32 | 1.08 | 0.25 | 0.30 | 2.65 | 2.76 | 0.93 | 1.12 | 2.73 | 2.75 | 2.05 | 2.61 | 2.72 | 2.75 | 1.92 | 2.37 | 2.74 | 2.75 | 2.47 | 2.98 |
| Frond 500 | 0.27 | 0.29 | 0.25 | 0.30 | 0.27 | 0.68 | 0.25 | 0.30 | 2.74 | 2.76 | 2.34 | 2.77 | 2.74 | 2.75 | 2.47 | 2.99 | 2.74 | 2.75 | 2.38 | 2.86 | 0.27 | 0.29 | 0.25 | 0.30 |
| Frond 600 | 0.27 | 0.29 | 0.25 | 0.30 | 0.27 | 2.51 | 0.25 | 0.30 | 2.42 | 2.75 | 0.30 | 0.18 | 2.73 | 2.75 | 1.75 | 2.18 | 2.51 | 2.75 | 0.79 | 0.62 | 2.73 | 2.75 | 2.47 | 2.98 |
higher pH values because of lower H⁺ ion concentration and more available binding sites for metal cations at pH > 6. At higher pH values, deprotonation of hydroxyl and carboxylic groups on the biochar creates negatively charged surface complexation with oxygen-containing groups on the biochar. Furthermore, at lower pH values protonation of the acidic functional groups on biochar occurs leaving a positively charged surface which can repel the metal ions, thereby decreasing the removal efficiencies [35, 43]. At higher pH values, deprotonation of hydroxyl and carboxylic groups on the biochar creates negatively charged surfaces leading to removal of the metal cations through electrostatic attraction. Generally, there are more available binding sites for metal cations at higher pH values because of lower H⁺ ion concentration causing less competition for the binding sites, hence increasing the removal efficiencies. Kolodynska et al. [54] and Usman et al. [55] have also reported that heavy metal ion adsorption on biochar increased as the pH was increased. When the pH of the solution increases, metals ions become less soluble and start to precipitate as metal hydroxide. As the pH of solution increases, zinc presents as Zn²⁺ at pH < 8.8 and presents as zinc hydroxides at pH > 8.8 [56]. Copper presents as Cu²⁺ at pH < 5.8 and presents as copper hydroxide at pH > 5.8 [57]. Nickel presents as nickel hydroxide at basic solutions [58]. Fe (II) ions form iron hydroxide at alkaline ranges [58, 59]. It has to be noted that at higher pH values, both the adsorption of metal ions through electrostatic attraction and the precipitation process contribute to the removal efficiencies.

The statistical analysis on feedstock and pyrolysis effect on removal

The removal efficiencies for single and mixed metal ion solutions were between 98% and close to 100% for all metals by leaf and frond biochar samples at pH > 6. The two-tailed t-test and one-way ANOVA tests were found to be not statistically significant (at the 0.05 level) among date palm frond and leaf biochar samples at different pyrolysis temperatures at the same pH values shown in Table 5. The only exception was nickel removal efficiencies in single metal solutions between Leaf 400 and 500 and between Frond 400, 500 and 600. According to these results, it can be inferred that the increase of the pyrolysis temperature does not make any significance in terms of copper, iron, nickel and zinc percent removal in both single and mixed metal solutions. Although higher pyrolysis temperatures increased the surface area of the biochar, this feature is not the sole factor affecting the removal capacity of adsorbent. Other surface characteristics such as cation exchange, surface precipitation of carbonate, and surface complexation with oxygen-containing groups also affect the removal efficiency [13].

The significance to use date palm frond or leaf as biochar feedstock to remove single metal ion or mixed metal ions was analyzed using two-tailed t test as shown in Table 6. According to the t test results, only using Leaf 500 to Frond 500 to remove the nickel was statistically significant in single ion solutions. As a conclusion, using different date palm feedstocks at different temperatures did not make any significance to remove single or mixed metals from aqueous solutions.

Using different feedstock of date palm or different pyrolysis temperatures did not make any statistical significance to remove single or mixed metal ions, and this observation is in alignment with the other reported studies as well. According to Lou et al. [60], the pyrolysis temperature did not affect the phosphate adsorption on the biochar. In the same manner, biochar samples

(qe: 2.47 mg g⁻¹) for all pyrolyzed leaf and frond biochars at pH 7 as shown in Fig. 6g. Zinc removal increased when pH was increased beyond pH 6, and the highest average removal efficiencies were close to 100% (qe: 3.0 mg g⁻¹) for all pyrolyzed leaf and frond biochars at pH 7, 8 and 10 as shown in Fig. 6h and Table 4.

As a result, at the pH values beyond 6, the removal efficiencies and qe of metal ions in single and mixed metals solutions were higher, and this observation is in parallel with the other reported studies. Chen et al. [17] reported that when using hardwood and corn stalks biochar, the removal efficiencies of copper and zinc increased when the pH increased from 2 to 5. Similarly, it was found that copper removal was higher at pH 6.0 as compared to pH 3.5 using three different biochar [52]. Kilic et al. [53] investigated the effect of pH on the adsorption of nickel and cobalt onto almond shell biochar, and it was found that the adsorption capacity increased from 0 to 10 mg g⁻¹ for nickel and from 10 to 30 mg g⁻¹ for cobalt as the pH was increased from 2 to 6. When the pH is further increased beyond 6, the adsorption capacity reduced to nearly 0 mg g⁻¹ for nickel and 10 mg g⁻¹ for cobalt. The effect of the pH on the adsorption of copper onto biochar derived from compost, rice husks, orange waste and olive pomace was studied over the pH range of 2 to 7. Removal efficiencies increased at higher pH values, and the optimum value was found to be pH 7 for all samples with removal efficiencies of 94% with compost, 90% with rice husks, 89% with orange waste, and 78% with olive pomace [19].

The pH of aqueous solutions influences the adsorbent’s surface charge and ionization as well as the ionization and speciation of the adsorbate [18]. Low removal efficiencies at lower pH values can be attributed to excess H⁺ ion concentration resulting in competition between H⁺ and metals for available sites on the biochar. Furthermore, at lower pH values protonation of the acidic functional groups on biochar occurs leaving a positively charged surface which can repel the metal ions, thereby decreasing the removal efficiencies [35, 43]. At higher pH values, deprotonation of hydroxyl and carboxylic groups on the biochar creates negatively charged surfaces leading to removal of the metal cations through electrostatic attraction. Generally, there are more available binding sites for metal cations at higher pH values because of lower H⁺ ion concentration causing less competition for the binding sites, hence increasing the removal efficiencies. Kolodynska et al. [54] and Usman et al. [55] have also reported that heavy metal ion adsorption on biochar increased as the pH was increased. When the pH of the
derived from orange peel pyrolyzed at 400, 500, 600, 700 and 800 °C did not make any statistically significant difference in the adsorption of cadmium [61]. Biochar derived from giant Miscanthus pyrolyzed at the ranges 300–600 °C did not increase the adsorption of cadmium to a significant extent [62]. Similarly, biochar derived from mulberry pyrolyzed at 350, 450, and 550 °C produced similar adsorption capacities for Pb (II) [12]. Therefore, it was concluded that changing pyrolysis temperature or changing feedstock for palm-derived biochar will not make any significant improvement to remove metals; therefore, the energy requirement to produce palm-derived biochar at lower temperature will be less for mass production. In fact, biochar can be pyrolyzed at site with available energy sources, and it can be utilized effectively to remove contaminants.

**Conclusions**

Our study investigated the effect of pH, feedstock type, and pyrolysis temperature at 400, 500 and 600 °C on the capacity of date palm waste derived biochar to remove copper, iron, nickel and zinc in single and mixed metal solutions. It was found that biochar pyrolyzed at higher temperatures did not produce statistically significant removal efficiencies for the heavy metal ions when compared to biochar pyrolyzed at lower temperatures. In economical perspective, it was concluded that the energy consumption to produce palm-derived biochar at lower temperature will be less and cost effective.

However, pyrolysis temperature was found to influence both surface chemistry and physical characteristics. Biochar pyrolyzed at high temperatures lost most of their functional properties as compared to the feedstock. From FT-IR spectra, it was observed that -OH stretching and other volatile groups decreased in biochar samples. As for the surface structure, surface area and pore volume increased with increased pyrolysis temperature and they became more porous, and showed deep channels due to the volatilization of organic matter. With increasing pyrolysis temperature, hydrogen, oxygen, nitrogen and sulfur content decreased while carbon content increased due to carbonization. Feedstock type was shown not be a statistically significant criterion for the metal removal. The removal efficiencies for single metal and mixed metal ions were between 98% and close to 100% for nickel, iron, zinc and copper by leaf and frond biochar. The leaf or frond date palm biochar are effective low-cost absorbers for removing metals. Utilizing date palm waste materials to produce biochar is a sustainable solution due to its negative impact on GWP, cost effectiveness and waste management perspective.

**Table 5** T test and one-way ANOVA p values comparing different pyrolysis temperatures for biochar samples

| Ion        | T test | One-way ANOVA |
|------------|--------|---------------|
|            | Leaf 400 and 500 | Leaf 500 and 600 | Frond 400 and 500 | Frond 500 and 600 | Frond 400 and 600 | Leaf 400, 500 and 600 | Frond 400, 500 and 600 |
| Single metal ion | Copper 0.4 | 0.31 | 0.16 | 0.27 | 0.17 | 0.31 | 0.94 | 0.16 |
|             | Iron 0.52 | 0.38 | 0.25 | 0.32 | 0.16 | 0.31 | 0.89 | 0.98 |
|             | Nickel 0.0019 | 0.07 | 0.74 | 0.22 | 0.97 | 0.44 | 0.13 | 0.001 |
| Mixed metal ions | Zinc 0.68 | 0.67 | 0.38 | 0.86 | 0.59 | 0.60 | 0.22 | 0.73 |
| Copper 0.79 | 0.68 | 0.37 | 0.38 | 0.47 | 0.22 | 0.91 | 0.84 |
| Iron 0.37 | 0.11 | 0.52 | 0.35 | 0.35 | 0.034 | 0.69 | 0.55 |
| Nickel 0.41 | 0.97 | 0.42 | 0.58 | 0.97 | 0.40 | 0.85 | 0.79 |
| Zinc 0.36 | 0.95 | 0.38 | 0.53 | 0.91 | 0.36 | 0.53 | 0.73 |

**Table 6** T test p values to compare different feedstocks as biochar material

| Ion | Single metal ions removal | Mixed metal ions removal |
|-----|---------------------------|--------------------------|
|     | Leaf 400 and Frond 400    | Leaf 500 and Frond 500   | Leaf 600 and Frond 600 |
|     |                            | Frond 400                | Leaf 500                | Frond 600                |
| Copper 0.74 | 0.18 | 0.59 | 0.36 | 0.68 | 0.47 |
| Iron 0.28 | 0.63 | 0.35 | 0.38 | 0.35 | 0.85 |
| Nickel 0.29 | 0.0008 | 0.14 | 0.87 | 0.49 | 0.53 |
| Zinc 0.16 | 0.27 | 0.43 | 0.91 | 0.51 | 0.44 |
Acknowledgments
The authors wish to thank Dr. Agathe Talarmin for conducting CHNS Analysis and Dr. Grzegorz Brudecki for helping ICP-MS analysis.

Authors’ contributions
Banu Sziricz contributed: Conceptualization, Methodology, Investigation, Writing - Review & Editing, Yohanna Fseha contributed: Investigation, Writing, Ibrahim Yildiz contributed: Validation, Writing - Review & Editing. Thomas Delclos contributed: Resources, Abbas Khaleel contributed: Resources. The author(s) read and approved the final manuscript.

Funding
This research was supported by Khalifa University and Dubai Electric and Water Authority, UAE through the Mohammed bin Rashid Al Maktoum Global Water Award 2019 Innovative Research and Development, National Institutions program (Grant number: 8434000361).

Availability of data and materials
All data generated or analyzed during this study are available upon request.

Competing interests
The authors declare they have no competing interests.

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Received: 11 August 2020 Accepted: 9 February 2021

Published online: 25 February 2021

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