The accuracy of virtual fractional flow reserve from invasive angiography: importance of the method of reconstructing three-dimensional coronary artery anatomy.

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ABSTRACT

Background

Three dimensional (3D) coronary anatomy, reconstructed from coronary angiography (CA), is now being used as the basis to compute ‘virtual’ fractional flow reserve (vFFR), and thereby guide treatment decisions in patients with coronary artery disease (CAD). Reconstruction accuracy is therefore important. Yet these methods remain poorly validated. Furthermore, the magnitude of vFFR error arising from reconstruction is unknown. We aimed to validate a new method for 3D CA reconstruction and determine the effect this had upon the accuracy of vFFR.

Methods

Clinically realistic coronary phantom models were created (seven standard stenoses in aluminium and 15 patient-based 3D-printed) and imaged with CA, three times, according to clinical protocols, yielding 66 datasets. Each was reconstructed using epipolar line projection and intersection. All reconstructions were compared against the phantom models in terms of minimal lumen diameter, centreline and surface similarity. 3D-printed reconstructions (n=45) and the reference files from which they were printed underwent vFFR computation, and the results were compared.

Results

The average error in reconstructing minimum lumen diameter (MLD) was 0.05 (±0.03 mm) which was <1% (95%CI 0.13-1.61%) compared with caliper measurement. Overall surface similarity was excellent (Hausdorff distance 0.65 mm). Errors in 3D CA reconstruction accounted for an error in vFFR of ±0.06 (95% limits of agreement).

Conclusions
Errors arising from the epipolar line projection method used to reconstruct 3D coronary anatomy from CA are small but result in clinically relevant errors in vFFR simulation, amounting to approximately 40% of the total error associated with vFFR.

**KEYWORDS**
Invasive coronary angiography, 3D reconstruction, angiography-derived fractional flow reserve; computational modelling

**ABBREVIATION LIST**

| 2D and 3D | Two and three-dimensional |
|-----------|---------------------------|
| CA        | Coronary angiography      |
| CAD       | Coronary artery disease   |
| CFD       | Computational fluid dynamics |
| CT        | Computed tomography       |
| FFR       | Fractional flow reserve   |
| HU        | Hounsfield units          |
| IVUS      | intravascular ultrasound  |
| LCA       | Left coronary artery      |
| LAD       | Left anterior descending  |
| LCX       | Left circumflex           |
| PCI       | Percutaneous coronary intervention |
| RCA       | Right coronary artery     |
| SD        | Standard deviation        |
| STH       | Sheffield Teaching Hospitals |
| QCA       | Quantitative coronary angiography |
| vFFR      | Virtual fractional flow reserve |
INTRODUCTION

Invasive coronary angiography (CA) remains the standard method for assessing coronary artery disease (CAD) and guiding its treatment with percutaneous coronary intervention (PCI). Standard CA provides a two-dimensional (2D), dynamic, arterial ‘luminogram’ which is interpreted visually by the operator. By applying mathematical algorithms, 3D coronary anatomy can now be reconstructed using computer software from orthogonal 2D projections. 3D CA reconstructions more accurately capture lesion length, plaque eccentricity and correlate better with functional measures of disease, which is frequently overestimated using 2D QCA. Moreover, by applying the governing equations of fluid dynamics to these reconstructions, physiological parameters such as pressure and flow can be predicted.

3D CA reconstructions are therefore now being used as the basis for the computation of clinical indices of coronary physiology such as fractional flow reserve (FFR). This is a significant development, because FFR improves patient outcomes and is considered the ‘gold standard’ method for selecting appropriate cases for PCI in international guidelines. The ability to compute FFR using 3D reconstructions from CA, known as angiography-derived or ‘virtual’ FFR (vFFR), is anticipated to widen access to the benefits physiologically-guided intervention considerably, allowing the clinical benefits of FFR to become available to patients without the need for to pass an invasive pressure wire. It is, therefore, important that vFFR is accurate.

Compared with invasively measured FFR, vFFR has an error range (95%CI) of FFR ±0.14. vFFR errors arise from inaccuracies in the 3D CA reconstruction and in the assumptions and simplifications in the mathematical solution.
importance of accurate 3D CA reconstruction in applications which are used to compute vFFR, there is a paucity of published data validating their accuracy. Furthermore, the amount of vFFR error attributable to the reconstruction method is unknown.

The aims of this study were to:

1. Validate the geometric accuracy of a new method for reconstructing 3D coronary artery anatomy from standard CA.

2. Quantify how much vFFR error results from errors in the reconstruction method.
METHODS

Study design

The study was performed at the University of Sheffield. Clinical data were collected from adult patients at the South Yorkshire Cardiothoracic Centre at Sheffield Teaching Hospitals (STH) NHS Foundation Trust, as approved by the NHS regional ethics board. Participating patients provided informed consent. All methods were carried out in accordance with relevant guidelines and regulations. First, we developed a new method of 3D CA reconstruction. Next, the accuracy of this reconstruction method was validated using phantom arterial models which were imaged with CA according to standard imaging protocols and then reconstructed. The geometry (surface similarity, centreline and diameter) of the reconstruction was then compared with the known geometry of the phantom models. This step validated the geometric accuracy of the reconstruction method. We then applied computational fluid dynamics (CFD) modelling to the reference files (the files from which the phantoms were 3D printed), and to the reconstructed arteries, to predict vFFR. By comparing the vFFR results, we were able to quantify the amount of vFFR error attributable directly to the reconstruction method. A summary flowchart illustrating the study design is shown in Figure 1. A detailed explanation follows below.
Figure 1. A flowchart demonstrating the experimental phantom arterial models, clinical imaging, 3D reconstruction, validation, vFFR simulation and analysis.

**Coronary phantom models**
Fifteen 3D-printed (from patient data)  
Seven aluminum (idealised models)

Angiography in cardiac catheter laboratory  
(standard clinical protocols, all phantoms ×3)

Three dimensional reconstruction  
(epiploar intersection, two views ≥30°)

Comparison of reconstruction vs phantom

Accuracy of centerline and overall surface topography  
(3D-printed, n=45)  
Accuracy of minimum lumen diameter  
(Aluminum, n=21)

vFFR analysis  
(VIRTUheart CFD model)

How much vFFR error arises from 3D CA reconstruction?

Were phantom arteries a true representation of reference files?  
(Hi res CT scanning, n=45)

Were phantom arteries radiographically realistic?  
(Greyscale sampling vs patient angiograms, n=90)
Reconstructing 3D coronary anatomy from coronary angiography

Two angiographic projections were selected, ≥30° apart, ensuring optimal visualisation of the artery and stenosis, during end diastole, as indicated by the software’s inbuilt ECG gating. The arterial centrelines in each of two angiographic projections were computed automatically. The centreline in the first projection was discretised into a series of points and the epipolar lines of each point in the second projection were computed. Each corresponding point in the second projection was computed at the intersection of the epipolar line with the centreline in the second projection. The three-dimensional co-ordinates of each point were then computed from the corresponding projected co-ordinates. The diameter of the vessel in each projection was computed based upon automatic detection of the contrast gradient with manual correction where necessary (overlapping vessels or branch points). 3D luminal reconstruction assumed an axisymmetric vessel with the average diameter from the two projections. In the catheterisation laboratory it is normal to re-centre the region of interest by movement of the table between projections. Tracking of these movements is not universal, and there are other sources of motion, including patient cardiac and respiratory movement. Three points of correspondence in each of the two projections (usually bifurcations) were identified. Using these points and a linear least squares optimisation process table and other movements were compensated for. The epipolar lines in the second projection were then computed, incorporating the computed table shift, and the reconstruction proceeded as before. The surfaces of the arteries were computed and represented as an assembly of triangular elements in STL format.

Creating the coronary phantom models
Two types of arterial phantom models were created. To assess the method’s accuracy in reconstructing vessel diameter, particularly in the stenosis region (minimal lumen diameter), phantom models were created from 3.2 mm diameter aluminium rods, which were curved to mimic the contour of the epicardial surface. Both concentric and eccentric stenoses were fabricated by hand. Seven fabricated stenoses represented anatomically mild, moderate and severe disease, ranging from 40 to 90% diameter stenoses. The aluminium rods were chosen because, unlike the 3D printed models, they were strong enough to maintain shape and integrity even with severe stenoses. Diameter measurements were made with a high-precision digital caliper (Mitutoyo, KA, Japan) as the average of three readings. The second type of phantoms were 3D-printed using as their basis, the CAs of patients with chronic coronary syndromes in whom FFR had also been measured. These solid phantom models were used to assess the accuracy of the reconstruction’s overall surface topography. These were based on the angiograms of patients with chronic coronary syndromes, mean age 66 years, who were being assessed for PCI with FFR guidance. Three left and three right branched coronary arterial (LCA, RCA) models were printed using stereo lithography (Rep Rap X400 PRO 3D printer) in polylactic acid (PLA) doped with stainless steel to mimic the radiodensity of contrast-filled vessels under CA imaging. The six models comprised a total of 15 individual branches, which were analysed separately, as they normally would be clinically and physiologically.

**Angiographic imaging**

All phantom models underwent routine coronary angiography (multiplane 2D acquisitions, Philips Azurion, Philips Healthcare, NL), according to a standard clinical protocol. Each phantom model was positioned on the cath lab table in anatomical
orientation. A consultant cardiologist and superintendent radiographer (STH) were instructed to image the model as if performing a clinical case, ensuring table movement and panning, in x, y and z planes. Imaging protocols were in accordance with UK Ionising Radiation Medical Exposure Regulations (IRMER). Projection angles for RCAs included LAO, RAO and LAO-cranial. Projection angles for LCAs included LAO-caudal, PA-caudal, RAO-caudal, LAO-cranial, PA-cranial and RAO-cranial. Images were exported in Digital Imaging and Communications in Medicine (DICOM) format. Each phantom model underwent three separate angiogram studies. The pixel:mm ratio for the Philips angiography system was 1 pixel =0.314 mm.

**Ensuring the phantom models were radiographically realistic**

To ensure the phantom models were radiographically realistic under angiographic imaging, we sampled the radiodensity of the raw CA images (pixel greyscale value, MATLAB, MathWorks Inc) at ten equally spaced intervals along the length of the main vessel of each phantom type (metal and 3D printed model angiograms). These were compared to a similar number of sample points (n=90 points in total) from the main vessel of a patient coronary angiogram to enable a comparison. Differences were compared by one-way ANOVA.

**Accuracy of the 3D printed phantom**

Our validation relied upon the 3D-printed phantom models being a true likeness of the files from which they were printed because these reference files were the gold-standard comparitor. To evaluate this, the phantom models were imaged in a 320-slice computed tomography (CT) scanner (Aquilion Genesis, Toshiba Medical Systems, Japan) at STH. The 3D rendered geometries were extracted using
Materialise Mimics software (Materialise, Belgium) and compared with the reference files they were printed from, by calculating the global Hausdorff distance within MeshLab. This was to validate the use of the 3D reference files as the gold-standard comparator.

**Validation of the reconstruction method**

**Reproducibility of the method:** It was important to understand the magnitude of error associated with the comparison method itself, i.e. the highest achievable accuracy. To assess this, we compared each 3D reference file against itself within MeshLab and measured the global Hausdorff distance between the two surfaces. Theoretically, comparing two identical surfaces should result in a Hausdorff distance close to, or equal to, zero. This analysis demonstrated the best achievable Hausdorff distance and the error inherent to the sampling method within the MeshLab software.

**Accuracy of the minimum lumen diameter:** The reconstructed surfaces of the stenosis (metal) phantoms were imported into MeshLab for 3D visualisation. The diameters of non-stenosed and stenosed regions were obtained from the raw centreline data of the reconstructions. These measurements were compared with the phantom minimum lumen diameter (MLD) which was the average of three digital caliper-measured readings at the point of maximum stenosis. 3D reconstructed MLD values were taken as the single smallest radius reading.

**Accuracy of the luminal surface topography:** Each reconstructed surface of the 3D-printed patient-specific models was imported into MeshLab (ISTI-CNR Research Centre, Italy) alongside its corresponding reference surface (from which the corresponding phantom model was 3D-printed). The reconstructed and reference geometries were optimally superimposed in 3D space. To ensure maximal overlay
and to avoid errors being introduced by imperfect manual positioning, an optimisation script (MATLAB, MathWorks) was used. The similarity between 3D surfaces was quantified and calculating the global Hausdorff distance. The Hausdorff distance objectively quantifies the difference between two geometries by measuring their mutual proximity and measuring the maximal distance between corresponding points of one geometry (reconstructed surface) relative to the other (the reference geometry). We report the minimum, maximum and RMS Hausdorff measurements. This analysis of luminal surface topography also served as a validation of the accuracy of the vessel centreline reconstruction because the latter is dependent upon the former.

**Assessing the magnitude of error in vFFR arising from the 3D CA reconstruction method:** The reconstructed and reference surfaces were imported into ANSYS Fluent (ANSYS Inc, US) for the generation of a 3D volume mesh to support computational fluid dynamics (CFD) simulation of FFR. Paired samples used identical boundary conditions to ensure a true like-for-like comparison. Virtual FFR (vFFR) was computed using VIRTUheart™ software (University of Sheffield, UK) which derives the translesional pressure ratio from a computational fluid dynamics (CFD) simulation, based upon solving the 3D form of the Navier Stokes equations. Identical boundary conditions were used. Thus, any difference in vFFR result must have arisen only from errors in the 3D CA reconstruction method. Further details of the CFD method are described in detail elsewhere. The computed FFR results for corresponding surfaces were then compared. To support this evaluation all vessels were imaged and reconstructed three times.
**Statistical analysis**

3D geometric similarity was evaluated by comparing the Hausdorff distances of the reconstructed geometries versus those of the reference standard, reported as mean, standard deviation (SD), and 95% confidence interval. Accuracy of stenosis reconstruction was assessed by comparing the measured and reconstructed stenosis measurements, expressed as distances (mm) and as a percentage of the vessel reference diameter (percentage stenosis). Agreement between the experimental and reference vFFR was assessed by analysing Lin’s concordance correlation coefficient (CCC) which (unlike Pearson’s coefficient) reports covariation and correspondence of two variables with the line of correlation passing through zero and a slope of 1.0.

Bland-Altman plots were constructed to show differences between vFFR values from reconstructed and reference files. Absolute bias and limits of agreement (±1.96 SD) were calculated. Statistical analysis was performed using SPSS (v25: IBM Analytics, Armonk, NY, USA).
RESULTS

The phantom arterial models

The aluminium stenosis models comprised four eccentric stenoses and three concentric with stenoses ranging from 44.7% to 77.2% representing the mild, moderate and severe clinical range. The seven alluminium and fifteen patient-specific 3D-printed phantom models were each imaged three times, generating 66 analyses in total (Table 1).

Radiodensity of the phantoms

The pixel greyscale contrast values of the coronary arteries in the clinical angiograms (n=30 sample points) were similar to those of the aluminium (n=30) and 3D-printed (n=30) phantom models (76.3 ±15.0u; 60.0 ±27.7u; and 71.7 ±28.7u). One-way ANOVA analysis between the samples from each group demonstrated no statistical difference (F(2,27)=1.18, P=0.32). From this we concluded that the phantom model imaging studies were radiographically realistic compared to the clinical imaging studies.

Accuracy of the phantom models

When comparing the CT imaging results against the reference files, the Hausdorff distance was 0.61 mm (±0.16 mm, 95% CI 0.45 - 0.78 mm). We concluded that the printed phantoms were an accurate embodiment of the reference files from which they were printed from and, therefore, that the reference files could be used as the ‘gold standard’ reference comparator for the validation analyses.
Table 1. Accuracy of diameter measurement the 3D CA method. The absolute (mm) and percentage error are reported for each of the seven stenosis models. Percentage stenosis was calculated as the ratio between reference (unstenosed) vessel diameter to diameter at the point of maximum stenosis.

| Model | Stenosis  | Caliper measured min diameter (mm) | Min diameter from 3D recon (mm) | Delta (mm) | Percentage Diameter stenosis, (caliper) | Percentage Diameter stenosis (recon) | Delta (%) |
|-------|-----------|------------------------------------|--------------------------------|------------|---------------------------------------|-------------------------------------|----------|
| 1     | Concentric| 0.91                               | 0.90                           | 0.01       | 71.9%                                 | 71.5%                               | 0.4%     |
| 2     | Concentric| 1.43                               | 1.50                           | -0.07      | 55.4%                                 | 53.1%                               | 2.3%     |
| 3     | Concentric| 1.65                               | 1.60                           | 0.05       | 48.6%                                 | 49.9%                               | 1.3%     |
| 4     | Eccentric | 1.77                               | 1.84                           | -0.07      | 44.7%                                 | 44.2%                               | 0.5%     |
| 5     | Eccentric | 0.88                               | 0.83                           | 0.05       | 72.8%                                 | 74.1%                               | 1.3%     |
| 6     | Eccentric | 0.74                               | 0.72                           | 0.02       | 77.2%                                 | 77.5%                               | 0.3%     |
| 7     | Eccentric | 1.76                               | 1.84                           | -0.08      | 44.7%                                 | 44.7%                               | 0.0%     |
Reproducibility of the printed phantoms

When the fifteen 3D arterial files were superimposed and compared against themselves (i.e. the same arterial 3D file opened twice and overlayed in 3D space), the Hausdorff distance was 0.28 mm (±0.20 mm). This defined the best achievable accuracy with this method and therefore the inherent error associated with the sampling method itself. This provided context for assessing the validated accuracy of the reconstruction method.

Accuracy of the method in reconstructing minimal lumen diameter

For the aluminium stenosis phantom models, compared with the digital caliper measurements, the average error in minimum diameter measurement was 0.05 mm (±0.03). The error as a percentage of the minimum diameter measurement (i.e. max stenosis) was <1% (±0.87%, 95% CI 0.13% to 1.61%). There was no statistical difference between the caliper measurements made on the phantom models and those from the reconstructions created by the novel method (P=0.93). Model analysis is detailed in Table 1.

Accuracy of the method in reconstructing luminal surface topography

For all 45 3D CA reconstructions (reconstructed from the 3D-printed phantoms), the Hausdorff distance error, relative to the phantom reference files, was 0.65 mm (±0.30 mm, 95% CI 0.56 mm to 0.74 mm). To help interpret this result, the best achievable Hausdorff distance (from the previous analysis) was was 0.28 mm (±0.20 mm). The following section reports how this small error in surface reconstruction translates into error when computing vFFR. Per-vessel accuracy data are presented in Table 2. Figure 2 demonstrates the Hausdorff distance analysis.
| Model, Artery | Minimum (mm) | Maximum (mm) | Mean (mm) | Root Mean Square (mm) |
|--------------|--------------|--------------|-----------|----------------------|
| RCA1 (Main)  | 0            | 2.80         | 0.63      | 0.84                 |
| RCA1 (Main)  | 0            | 4.03         | 0.73      | 1.01                 |
| RCA1 (Main)  | 0            | 2.70         | 0.60      | 0.83                 |
| RCA2 (Main)  | 0            | 8.67         | 1.48      | 1.94                 |
| RCA2 (Main)  | 0            | 3.94         | 0.76      | 1.00                 |
| RCA2 (Main)  | 0            | 3.45         | 0.66      | 0.86                 |
| RCA3 (Main)  | 0            | 2.73         | 0.75      | 0.98                 |
| RCA3 (Main)  | 0            | 3.24         | 0.85      | 1.10                 |
| RCA3 (Main)  | 0            | 2.65         | 0.65      | 0.82                 |
| LCA1 (LAD)   | 0            | 3.75         | 0.61      | 0.78                 |
| LCA1 (LAD)   | 0            | 3.12         | 0.58      | 0.74                 |
| LCA1 (LAD)   | 0            | 2.20         | 0.34      | 0.44                 |
| LCA1 (Diagonal) | 0     | 2.12         | 0.54      | 0.68                 |
| LCA1 (Diagonal) | 0     | 2.02         | 0.39      | 0.52                 |
| LCA1 (Diagonal) | 0     | 2.12         | 0.41      | 0.55                 |
| LCA1 (Marginal) | 0     | 2.69         | 0.65      | 0.79                 |
| LCA1 (Marginal) | 0     | 3.14         | 0.77      | 0.97                 |
| LCA1 (Marginal) | 0     | 4.15         | 0.47      | 0.67                 |
| LCA1 (Circumflex) | 0    | 1.65         | 0.28      | 0.36                 |
| LCA1 (Circumflex) | 0    | 0.77         | 0.17      | 0.22                 |
| LCA1 (Circumflex) | 0    | 1.98         | 0.29      | 0.37                 |
| LCA2 (LAD)   | 0            | 2.88         | 0.44      | 0.57                 |
| LCA2 (LAD)   | 0            | 2.02         | 0.26      | 0.33                 |
| LCA2 (LAD)   | 0            | 2.53         | 0.41      | 0.54                 |
| LCA2 (Diagonal) | 0     | 2.19         | 0.38      | 0.49                 |
| LCA2 (Diagonal) | 0     | 2.70         | 0.40      | 0.45                 |
| LCA2 (Diagonal) | 0     | 1.48         | 0.32      | 0.41                 |
| LCA2 (Marginal) | 0    | 1.74         | 0.24      | 0.30                 |
| LCA2 (Marginal) | 0    | 1.84         | 0.22      | 0.30                 |
| LCA2 (Marginal) | 0    | 2.65         | 0.59      | 0.75                 |
| LCA2 (Circumflex) | 0   | 1.82         | 0.29      | 0.35                 |
| LCA2 (Circumflex) | 0   | 2.95         | 0.30      | 0.43                 |
| LCA2 (Circumflex) | 0   | 1.80         | 0.28      | 0.34                 |
| LCA3 (LAD)   | 0            | 2.01         | 0.44      | 0.57                 |
| LCA3 (LAD)   | 0            | 2.88         | 0.45      | 0.61                 |
| LCA3 (LAD)   | 0            | 1.58         | 0.35      | 0.46                 |
| LCA3 (Diagonal) | 0     | 3.00         | 0.63      | 0.88                 |
| LCA3 (Diagonal) | 0     | 2.63         | 0.59      | 0.77                 |
| LCA3 (Diagonal) | 0     | 1.44         | 0.37      | 0.46                 |
| LCA3 (Marginal) | 0    | 1.99         | 0.35      | 0.48                 |
| LCA3 (Marginal) | 0    | 2.02         | 0.59      | 0.71                 |
| LCA3 (Marginal) | 0    | 1.95         | 0.59      | 0.72                 |
| LCA3 (Circumflex) | 0   | 4.04         | 0.44      | 0.65                 |
| LCA3 (Circumflex) | 0   | 2.02         | 0.59      | 0.71                 |
| LCA3 (Circumflex) | 0   | 1.63         | 0.30      | 0.37                 |

**Table 1.** Hausdorff distance analysis of surface similarity, presented on a per-vessel basis for all 45 comparisons. Measurements include minimum, maximum and mean distances between sampled points on reference and reconstructed 3D vessels when aligned and superimposed. Root mean square is the most important result, providing an overall value of error in reconstruction of each vessel in millimetres.
Figure 2. 3D-printed left coronary artery phantom (A) underwent coronary angiography according to standard clinical protocol in the cardiac catheter laboratory (B). Paired images ≥30° apart (C and D) were used to reconstruct the arterial anatomy (E). Panel E is a screen shot from the MeshLab software: the artery reconstructed from imaged phantom is superimposed on top of the artery from the reference file (used to 3D the phantom in panel A) in 3D virtual space. Visual inspection demonstrates excellent agreement but the colour map demonstrates areas of perfect agreement (green) through to areas without perfect correspondence (red).
Figure 3. vFFR analysis. Individual branches of the reference file (A) and the 3D CA reconstructed artery (B) were subjected to vFFR analysis under identical conditions. C and D show the obtuse marginal branch from the reference file (C) and 3D CA (D). Excellent anatomical reconstruction means that both fluid dynamics analyses result in vFFR of 0.72. Panels E and F demonstrate the analysis for the left anterior descending branch. In this branch both the reference and 3D CA reconstructed vessels have a vFFR of 0.83.
**Accuracy of the 3D CA reconstruction method when computing vFFR**

Mean vFFR of the reference vessels was 0.75 (±0.12) and the range was 0.43 to 0.98. Twenty-five cases (56%) were below the threshold value for PCI (0.80) and twenty-one (47%) were within the zone 0.75-0.85. Mean vFFR of the vessels reconstructed from the 3D-printed phantom models (n=45) was 0.73 (±0.12) and the range was 0.49 to 0.96. Relative to the reference vessels, the reconstructions had a small but statistically significant negative bias in vFFR results (mean delta -0.01 ±0.03, P<0.05) (Figure 2). In addition, the concordance correlation coefficient between reference and 3D CA reconstructed vFFR results was excellent (CCC=0.96, 95% CI 0.92 to 0.98) (Figure 4). Case-specific comparison of the reconstructed and reference vessels is displayed in Table 3. Figure 3 demonstrates an example of vFFR analysis of the reconstructed and reference surfaces. Relative to the vFFR results computed from the reference surfaces, the Bland Altman 95% limits of agreement of the vFFR results computed from the reconstructed surfaces were ±0.06.
Figure 4. Panel A) Scatter plot demonstrating high concordance correlation coefficient (CCC) between angiography-derived vFFR from the 3D reconstruction method and the gold-standard reference file-derived vFFR. Panel B) Bland-Altman Plot demonstrating agreement and differences between angiography-derived vFFR from the 3D CA reconstruction method and the gold-standard reference file-derived vFFR. Mean delta (bias) is shown as a solid line (-0.01) and the upper and lower limits of agreement (SD: ±1.96) are shown with the interrupted line (-0.07 to +0.05).
| Vessel | Model name | vFFR (print) | vFFR (recon) | Absolute Error (Delta vFFR) | Bias (Delta vFFR) |
|--------|------------|--------------|--------------|-----------------------------|------------------|
| LAD    | LCA1A      | 0.76         | 0.76         | 0.00                        | 0.00             |
| Diagonal | LCA1A      | 0.72         | 0.71         | 0.01                        | -0.01            |
| Marginal | LCA1A     | 0.82         | 0.73         | 0.09                        | -0.09            |
| Circumflex | LCA1A     | 0.72         | 0.70         | 0.02                        | -0.02            |
| LAD    | LCA2A      | 0.84         | 0.84         | 0.00                        | 0.00             |
| Diagonal | LCA2A      | 0.86         | 0.80         | 0.06                        | -0.06            |
| Marginal | LCA2A     | 0.70         | 0.65         | 0.05                        | -0.05            |
| Circumflex | LCA2A     | 0.63         | 0.62         | 0.01                        | -0.01            |
| LAD    | LCA3A      | 0.67         | 0.66         | 0.01                        | -0.01            |
| Diagonal | LCA3A      | 0.65         | 0.64         | 0.01                        | -0.01            |
| Marginal | LCA3A     | 0.79         | 0.79         | 0.00                        | 0.00             |
| Circumflex | LCA3A     | 0.52         | 0.51         | 0.01                        | -0.01            |
| Main   | RCA1A      | 0.81         | 0.85         | 0.04                        | 0.04             |
| Main   | RCA2A      | 0.55         | 0.51         | 0.04                        | -0.04            |
| Main   | RCA3A      | 0.82         | 0.79         | 0.03                        | -0.03            |
| LAD    | LCA1B      | 0.83         | 0.84         | 0.01                        | 0.01             |
| Diagonal | LCA1B      | 0.89         | 0.90         | 0.01                        | 0.01             |
| Marginal | LCA1B     | 0.98         | 0.96         | 0.02                        | -0.02            |
| Circumflex | LCA1B     | 0.77         | 0.76         | 0.01                        | -0.01            |
| LAD    | LCA2B      | 0.83         | 0.83         | 0.00                        | 0.00             |
| Diagonal | LCA2B      | 0.97         | 0.96         | 0.01                        | -0.01            |
| Marginal | LCA2B     | 0.72         | 0.72         | 0.00                        | 0.00             |
| Circumflex | LCA2B     | 0.76         | 0.76         | 0.00                        | 0.00             |
| LAD    | LCA3B      | 0.76         | 0.76         | 0.00                        | 0.00             |
| Diagonal | LCA3B      | 0.94         | 0.92         | 0.02                        | -0.02            |
| Marginal | LCA3B     | 0.79         | 0.79         | 0.00                        | 0.00             |
| Circumflex | LCA3B     | 0.67         | 0.67         | 0.00                        | 0.00             |
| Main   | RCA1B      | 0.85         | 0.84         | 0.01                        | -0.01            |
| Main   | RCA2B      | 0.58         | 0.64         | 0.06                        | 0.06             |
| Main   | RCA3B      | 0.84         | 0.86         | 0.02                        | 0.02             |
| LAD    | LCA1C      | 0.71         | 0.69         | 0.02                        | -0.02            |
| Diagonal | LCA1C      | 0.65         | 0.64         | 0.01                        | -0.01            |
| Marginal | LCA1C     | 0.82         | 0.80         | 0.02                        | -0.02            |
| Circumflex | LCA1C     | 0.78         | 0.76         | 0.02                        | -0.02            |
| LAD    | LCA2C      | 0.81         | 0.80         | 0.01                        | -0.01            |
| Diagonal | LCA2C      | 0.78         | 0.74         | 0.04                        | -0.04            |
| Marginal | LCA2C     | 0.81         | 0.77         | 0.04                        | -0.04            |
| Circumflex | LCA2C     | 0.63         | 0.56         | 0.07                        | -0.07            |
| LAD    | LCA3C      | 0.67         | 0.66         | 0.01                        | -0.01            |
| Diagonal | LCA3C      | 0.66         | 0.66         | 0.00                        | 0.00             |
| Marginal | LCA3C     | 0.43         | 0.52         | 0.09                        | 0.09             |
| Circumflex | LCA3C     | 0.54         | 0.53         | 0.01                        | -0.01            |
| Main   | RCA1C      | 0.84         | 0.76         | 0.08                        | -0.08            |
| Main   | RCA2C      | 0.55         | 0.49         | 0.06                        | -0.06            |
| Main   | RCA3C      | 0.82         | 0.78         | 0.04                        | -0.04            |

Table 3. Per-vessel vFFR measurements for reconstructed and reference meshes. The observed error in physiological simulation for 3D reconstructed vessels when compared to their reference (print mesh) counterparts is also reported.
DISCUSSION

We have developed and validated a method for reconstructing 3D coronary artery anatomy from standard CA suitable for using as the basis for the computation of vFFR. The method uses a simple protocol of two standard 2D angiographic image projections separated by at least 30 degrees. The method compensates for inter-acquisition patient and table movement in any plane, and for intra-acquisition movement or 'panning'. Using both idealised metallic, and patient-specific 3D-printed phantom coronary models, we have demonstrated good geometric accuracy of the reconstruction method in terms of both minimum lumen diameter capture and overall surface similarity. The reconstruction was sufficient for reliable physiological simulation (computing vFFR). vFFR agreement with the reference results was excellent. Relative to the results computed from the reference surfaces, the 95% Bland Altman limits of agreement of the vFFR results computed from the reconstructed surfaces were ±0.06 and the concordance correlation coefficient was 0.96.

Reconstructing 3D coronary anatomy from conventional (2D) CA is challenging. Epicardial coronary arteries are only 2-5 mm in diameter, smaller in stenosed regions, tortuous, and constantly moving due to cardiac, ventilatory and patient activity. In addition, the x-ray table is moved between and during image acquisitions, arteries overlie each other, and an individual projection may not adequately show all regions of interest. Early innovations attempted to overcome some of these challenges by rotating the C-arm while acquiring images, or by simultaneous biplane acquisition but these systems are not widely available, are cumbersome, and associated with significant practical shortcomings.\textsuperscript{10,11} A strength of the methods
described this study is that a standard multiple single-plane CA is all that is required to reconstruct the coronary anatomy.

Uniquely, but importantly, our study associated geometric with computed physiological error. The physiological (vFFR) error arising purely from errors in the reconstruction was FFR ±0.06 (Bland-Altman 95% limits of agreement). Whilst this is clinically relevant, it needs placing in context. A recent meta-analysis of thirteen studies of vFFR reported average error of vFFR ±0.14. The previously reported error of the current vFFR method was FFR ±0.16. Similarly, a study of vFFR computed from CT angiography reported an error of ±0.15 (24). Thus, our data suggest that approximately 40% of the error associated with vFFR is attributable to errors in the reconstruction process. This corresponds with the findings of published vFFR sensitivity analyses which highlight that the major source of vFFR error arises from the selection and tuning of boundary conditions to represent the microvascular resistance and flow, and not the anatomical reconstruction per se. Thus, the present study supports the notion that, in the context of the computation of vFFR from coronary arterial reconstructions, the greatest scientific challenge remains the selection and application of appropriate boundary conditions, because these are the main contributor to the remaining error.

In the context of the MLD, which is important when computing vFFR, our method was associated with <1% error compared with digital caliper measurement. In terms of surface similarity, the lowest possible achievable error (when we compared identical surfaces) was 0.28 mm (±0.20 mm). The error of our reconstruction method (0.65 mm ±0.30 mm) was therefore only 0.37 mm above the best achievable. Given that the pixel sizing of the Philips angiography system was 1 pixel = 0.314 mm (at 512 by
512 pixel resolution), our model was accurate, at worst, to two pixels; and, at best, to a single pixel. This is reassuring because no system can be more accurate than a single pixel. It would therefore be interesting to test the method on higher resolution detectors (1024 by 1024). Furthermore, the error we observed in our method was almost identical to that of high-resolution CT reconstruction (0.65 mm ± 0.30 novel vs 0.61 mm, ± 0.16 CT), an imaging modality used routinely in clinical practice for coronary angiographic reconstructions and vFFR computation.

Our analysis was more detailed and pragmatic than previously published studies. Shechter et al (2003), Movassaghi et al (2004), and Yang et al (2009) all used physical coronary phantom models to assess the accuracy of their 3D arterial reconstructions. However, these studies considered only the accuracy of reconstructed centreline data, using Euclidian distance measurement. This is different to our study because we also integrated an assessment of the luminal surface topography (which incorporates centreline accuracy), stenosis capture, and physiological accuracy. Furthermore, the models used in these studies were relatively simple, contrast-filled tubes with narrowings made to mimic stenoses, not reflecting actual patient anatomy. One of the challenges of reconstructing 3D models from 2D angiographic images is in capturing vessel curvature and dealing with images that foreshorten the arterial anatomy. It would not be possible to assess this in these phantom types. In their comparison of centreline data, Yang et al, described their average positional accuracy (distance between reconstructed and true phantom centrelines) to be 0.665 mm, similar to the average error in surface reconstruction reported in our study. Centreline comparison provides an analysis of the system’s ability to capture the curvature of the vessel, however it gives no information on the quality of surface topography or diameter accuracy. Shechter et al subjected their
phantoms to magnetic resonance imaging and used the resultant reconstructed centreline data as their comparator.\textsuperscript{17} Arguably, using this method, errors in the process of obtaining centreline data for the imaged phantoms may influence the results; it is more a comparison of reconstructions rather than validation against a ground truth comparator.

Other methods of validation have been used. In an \textit{in vivo} assessment of the accuracy of its 3D vessel reconstruction, the Cardiovascular Angiographic Analysis Systems (CAAS) QCA-3D (Pie Medical Imaging, BV, NL), ten patients underwent coronary angiography and intravascular ultrasound (IVUS). Reconstructed arterial anatomy was compared with the IVUS data.\textsuperscript{20} Although the CAAS QCA-3D system underestimated luminal area and lesion length, it did show a strong correlation with IVUS derived dimensions. IVUS itself has multiple drawbacks in vessel analysis: probe placement and choice of imaging plane can introduce geometrical inaccuracies, image quality is low and most importantly in this case, it produces a straight tube and cannot represent the 3D shape or curvature of vessels.\textsuperscript{21} The CAAS workstation is now providing commercially available \textit{vFFR} simulation, using 3D arterial anatomy.

The physiological significance of the cases included in this study was clinically relevant. Sixty percent were physiologically significant and nearly half (47\%) were in the borderline range (0.75-0.85). One of the greatest challenges for reconstructing coronary anatomy from angiography, particularly when computing physiology, is the ability to accurately capture small diameters at the point of maximum stenosis (minimum vessel diameter).\textsuperscript{22} Small changes in diameter have a relatively large influence on CFD analysis of computed pressure gradient and \textit{vFFR}. A strength of
the current study was the use of clinically realistic stenotic phantoms. The phantoms comprised of curved and linear geometries, concentric and eccentric stenoses and minimum diameters ranging from 0.74-1.77 mm (45-77%). These combinations reflect real-world CAD, investigated and treated in the cardiac catheter laboratory within the anatomically mild, moderate and significant range.\textsuperscript{23} Despite the tightest stenosis being just over two pixels wide on angiography (0.74 mm), our tool captured this to within 0.02 mm of error. This same value also defined the average error across all stenosis analyses. Most x-ray angiography-based modelling tools generate axisymmetric reconstructions based upon average radius calculation. Although this is ideal for concentric lesions, it is not representative of real-world atherosclerotic plaque that is usually eccentric, at least to a small degree.\textsuperscript{24} It is, therefore, possible that most tools for this type of 3D reconstruction are introducing a small magnitude of error as a result of circular diameter approximation. Galassi et al refined their 3D reconstruction algorithm to better account for more complex luminal contours by using a Non-Uniform Rational B Spline contour, allowing for flexible freeform diameter reconstruction.\textsuperscript{15} A comparison of luminal diameter was made against intravascular imaging using optical coherence tomography. In a sample of vessels with true diameters ranging between 0.72-1.03mm, the average error was 0.29mm despite the use of ‘flexible’ luminal modelling. Our method still supports the analysis of eccentric stenoses, including those in the current study, because the centreline deviates accordingly. Nevertheless, this is a potential limitation, and there are algorithms that compute non-circular cross-sections: we would observe that there is simply not enough information in two projections to identify the shape of the cross-section uniquely, although the more sophisticated processes certainly produce better approximations than axisymmetric.\textsuperscript{15} However, the primary purpose of this process was to produce geometries to support physiological computation, and separate
exploration of the sensitivity of this parameter to the assumption of circular cross-sections had indicated that this is a minor limitation in comparison to other sources of error, especially estimation of the distal myocardial resistance.\textsuperscript{13}

The sample size in our study was small. However, it was larger than those used in other validation studies in this area. It also utilised patient-specific 3D-printed models.\textsuperscript{17-19} This enabled a unique, clinically relevant analysis. Although the phantom models were static, this is unlikely to affect results because our tool incorporates a correction for movement in any plane (x,y or z) alongside ECG-gating which was designed and implemented to compensate for cardiac and respiratory motion and for radiographic panning.

**CONCLUSIONS**

We have developed and validated a tool that reconstructs 3D coronary arterial anatomy from standard CA. The method was simple to use and compensated effectively for inter- and intra-acquisition movement. Errors in reconstruction were small and led to small but clinically relevant errors in vFFR accounting for around 40% of the total error associated with vFFR.

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Author contribution: PDM and JPG initiated the project and supervised RS who performed the experiments, simulations and data analysis. DRH and VR developed and implemented the reconstruction method. Technical help with analysis of the 3D CA reconstructions was provided by AJN and JF. JH led on the clinical imaging. RCG, PVL, GP and PG and all co-authors helped to draft and critically appraise the final manuscript.
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