Observational Study

Immune response activation following hyperthermic intraperitoneal chemotherapy for peritoneal metastases: A pilot study

Giammaria Fiorentini, Donatella Sarti, Alberto Patriti, Emilio Eugeni, Francesco Guerra, Francesco Masedu, Andrew Reay Mackay, Stefano Guadagni

ORCID number: Giammaria Fiorentini (0000-0002-5615-889X); Donatella Sarti (0000-0002-8031-7846); Alberto Patriti (0000-0003-2414-268X); Emilio Eugeni (0000-0001-7989-6660); Francesco Guerra (0000-0003-2891-4659); Francesco Masedu (0000-0003-0290-5324); Andrew Ray Mackay (0000-0001-7096-3759); Stefano Guadagni (0000-0001-8525-084X).

Author contributions: Fiorentini G was the guarantor and designed the study; Patriti A, Eugeni E, Guerra F performed the procedures and collected the data; Guadagni S and Sarti D participated in the acquisition, analysis, and interpretation of the data, and drafted the initial manuscript; Masedu F and Mackay AR revised the article critically for important intellectual content, statistical analysis and language revision.

Institutional review board statement: The study was reviewed and approved by the Ethics Committee of L’Aquila and Teramo (Italy).

Informed consent statement: All study participants, or their legal guardian, provided informed written consent prior to study enrollment.

Conflict-of-interest statement: There are no conflicts of interest to report.

Data sharing statement: No

Abstract

BACKGROUND

Hyperthermic intraperitoneal chemotherapy (HIPEC) for peritoneal metastases (PM) is considered to be feasible, safe and to improve survival.

AIM

To investigate whether an immune response is activated following HIPEC for PM.

METHODS

Six patients were enrolled in this study. Peripheral blood samples were obtained from each patient prior to (day 0) and post-procedure (day 30), and used to evaluate the number of CD3+ total, CD3+/CD4+ T-Helper, CD3+/CD8+ cytotoxic T, CD3+/CD56+ natural killer and CD19+ B lymphocyte numbers, and CD4+: CD8+ T lymphocyte ratios.

RESULTS

The total numbers of CD3+, CD3+/CD4+ T-Helper, CD3+/CD8+ cytotoxic T, CD3+/CD56+ natural killer and CD19+ B lymphocytes, and CD4+: CD8+ T lymphocyte ratios were increased in all but one patient 30 d following the cytoreductive surgery-HIPEC procedure, and these increases were significant (P ≤ 0.05) for CD3+/CD4+ T Helper and CD3+/CD8+ cytotoxic T lymphocyte numbers.

CONCLUSION

This report provides the first evidence that HIPEC exhibits immunomodulating activity in PM patients, resulting in generalized activation of the adaptive
In this observational study, data were collected from patients with PM prospectively.

STROBE statement: The authors have read the STROBE Statement-checklist of items, and the manuscript was prepared and revised according to the STROBE Statement-checklist of items.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

Manuscript source: Invited manuscript

Received: December 24, 2019
Peer-review started: December 24, 2019
First decision: February 20, 2020
Revised: April 13, 2020
Accepted: May 12, 2020
Article in press: May 12, 2020
Published online: June 24, 2020

P-Reviewer: Cashin PH, de Melo FF, Morris DL, Zhao XP
S-Editor: Zhang L
L-Editor: Webster JR
E-Editor: Liu MY

INTRODUCTION

Cytoreductive surgery (CRS) was originally introduced for debulking peritoneal metastases (PM) in ovarian cancer patients and improved survival[1]. Despite some difficulty in being accepted by the medical oncology community, CRS was subsequently adopted for the treatment of pseudomyxoma peritonei (PMP), with beneficial results, confirming a role for CRS in the treatment of peritoneal disease[2]. CRS methodology was subsequently improved by Sugarbaker et al[3], who introduced six distinct peritonectomy procedures for complete cytoreduction. Intrapерitoneal (IP) chemotherapy was subsequently used as an adjuvant therapy to CRS. In several clinical trials, hyperthermic intrapерitoneal chemotherapy (HIPEC), in particular, resulted in survival improvements[4]. Today, CRS combined with HIPEC is considered the standard treatment for PMP and peritoneal mesothelioma[5] and, according to current Network NCCN Guidelines[6,7], is also considered for the management of colorectal PM.

HIPEC, pressurized intrapерitoneal aerosol chemotherapy, hepatic artery infusion, trans-arterial chemo-embolization and thermal ablation are all under investigation as approaches for locoregional treatment[8,9]. However, only thermal tumor ablation has been reported to induce a potential immunomodulatory effect, illustrated by the intense inflammatory cell response that is associated with thermally ablated tumors[10,11], and characterized by ablated tissue transitional zone infiltration by immune and inflammatory cell populations, including: Dendritic cells, neutrophils, macrophages, B and T lymphocytes and natural killer cells. In the present study, we evaluated and confirm that hyperthermic intraperitoneal chemotherapy for peritoneal metastases also exhibits immunomodulatory activity, inducing the general activation of an adaptive immune response.

MATERIALS AND METHODS

Study population and data collection

In this observational study, data were collected from patients with PM prospectively...
submitted for CRS and HIPEC, from October 2018 to August 2019, in accordance with Research Ethics Committee regulations. The inclusion criteria were: Age > 18 and < 75 years, diagnosis of PC with T4 stage (including cases with bowel perforation), mucinous subtype, or positive cytology of peritoneal lavage and written informed consent. Exclusion criteria were: Pre-operative chemotherapy or oncology medical treatment, concomitant tumor and autoimmune or rheumatologic disorders.

**Cytoreductive surgery and HIPEC**
CRS and HIPEC were performed according to learning curve and training program recommendations[11]. HIPEC, performed immediately after R0 primary tumor debulking (intraoperative administration), involved 60-min infusion/lavage with Oxaliplatin 400 mg/m² and Mitomycin C 15 mg/m², with drugs added to perfusion solutions at temperatures ≥ 42°C.

**Immunological parameters**
Peripheral blood (PB) samples were collected in ethylenediaminetetraacetic acid, prior to (day 0) and following (day 30) CRS-HIPEC procedures, and immunophenotypic analysis was performed within 24 h (Figure 1). Single-cell suspensions were labelled with fluorochrome-conjugated anti-human CD3, CD4, CD8, CD19 and CD56 monoclonal antibodies or matched isotypes, and CD3+ total, CD3+/CD4+ T-Helper, CD3+/CD8+ cytotoxic T, CD3+/CD56+ NK and CD19+ B lymphocytes, were quantified by fluorescence-activated cell sorting, an antibody-based cell sorting method for heterogeneous mixtures based upon the specific light scattering and fluorescent characteristics of each antibody-labelled cell-type[12]. Additional immunological parameters included the CD4+:CD8+ T lymphocyte ratio. Immunological populations were also analyzed in stained samples, after exclusion of debris and doublets, as previously described[13].

**Outcome measures**
The primary outcome was to assess adaptive immune response activation and decreased immunosuppression, and the secondary outcome was to monitor recurrence-free survival, by computed tomography scan or magnetic resonance imaging.

**Statistical analysis**
The characteristics of recruited immune/inflammatory cell populations are described qualitatively and by quantitative variables. Unilateral paired Student’s t-tests were used to demonstrate intra-patient and intra-cohort treatment response variability, trend, and statistical significance associated with a P value of ≤ 0.05.

**RESULTS**

**Overall characteristics of the patients**
The 6 patients in this study comprised 3 males (50%) and 3 females (50%), with a median age of 55 years (range 48-71 years). Two patients (33%) presented with PM from colorectal cancer and the other 4 (77%) with PMP (Table 1).

Computed tomography scans, performed 3, 6 and 12 mo following CRS-HIPEC, demonstrated complete responses in all 6 patients, associated with a median progression-free survival of 12 mo. No complications were observed and the mean hospitalization-time, following CRS-HIPEC, was 10 ± 2 d. One patient however, was hospitalized for 58 d with severe oxaliplatin toxicity, characterized by grade 4 hematopoietic toxicity, according to National Cancer Institute, Common Terminology Criteria for Adverse Events (CTCAE v4.03), and grade 3 hematomas in the abdominal wall and pelvis. The other patients did not exhibit any HIPEC drug-associated adverse events.

Evaluation of system lymphocyte populations in patient PB samples (Tables 2 and 3), revealed increases in systemic CD3+ total, CD3+/CD4+ T Helper, CD3+/CD8+ cytotoxic T, CD3+/CD56+ NK, CD19+ B lymphocyte numbers and increased CD4+:CD8+ T lymphocyte ratios, statistically significant for CD3+/CD4+ T Helper and CD3+/CD8+ Cytotoxic T lymphocyte populations (unilateral paired Student’s t test, P < 0.05) (Figure 2). These increases were observed in all but one patient, who exhibited reduced systemic lymphocyte counts post-HIPEC, consistent with severe oxaliplatin toxicity.
DISCUSSION

The peritoneum is the second most common site of colon cancer recurrence\(^\text{[14]}\). However, early imaging detection of peritoneal metastases is difficult and adjuvant systemic treatment does not reduce peritoneal dissemination. Therefore, current international clinical practice guidelines suggest HIPEC for treatment of patients at high risk of PM, as this procedure has been shown to reduce peritoneal recurrences in the long-term and to prolong overall survival\(^\text{[14-18]}\).

Hyperthermia induces tissue necrosis immediately adjacent to the thermal source and induces apoptosis in more peripheral or transitional zone tissues, surrounded by tissues unaffected by thermal ablation\(^\text{[10]}\). Tumor cell necrosis and apoptosis result in the release of tumor-debris, tumor antigens and damage-associated molecular patterns, all of which could theoretically activate the immune system. For this reason, we evaluated systemic immune cell profiles in PM patients treated with CRS-HIPEC, as a potential index of immune response activation by this procedure. The results obtained provide the first evidence that HIPEC induces immune system activation in PM patients, characterized by induction of a generalized adaptive immune response and decrease in immunosuppression. Furthermore, this CRS-HIPEC-induced immunological effect was long-lived and lasted for several weeks, consistent with authentic immunomodulation, rather than a normal inflammatory response.

Figure 1 Representative example of fluorescence-activated cell sorting immunophenotypic analysis.
Table 1  Sample characteristics

|                        | n  | %   |
|------------------------|----|-----|
| Males                  | 3  | 50  |
| Females                | 3  | 50  |
| Appendiceal mucinous neoplasms - pseudomyxoma peritonei | 4  | 67  |
| Colon cancers          | 2  | 33  |
| Age (yr)               | 55 (median) | 48-71 (range) |

In our opinion, therefore, HIPEC is not only a useful chemotherapeutic procedure but also stimulates the immune system. In fact, 60-min of HIPEC appears to promote not only efficacious cytotoxicity but also sufficient release of tumor debris, antigens and damage-associated molecules to activate the immune system and promote cytotoxic T lymphocyte maturation.

The limitation of this pilot study, however, is that it only provides preliminary evidence that this methodological approach is appropriate for evaluating immunological changes following CRS and HIPEC, and is, therefore, a forerunner to a future confirmatory large cohort study. Furthermore, this study was not designed to evaluate treatment safety, efficacy or effectiveness, justifying the small sample size. The data should also be interpreted with some care, since HIPEC is performed immediately following tumor debulking, implicating surgical trauma as an alternative source of physiological and immunological alteration. Indeed, the normal physiological response to tissue injury involves a complex integration of inflammatory, immunological, neuroendocrine and metabolic mechanisms, and incision, dissection, organ manipulation and vascular alterations all induce acute inflammation for the purpose of host defense and tissue repair, with negative immunosuppressive feedback activated to avert an exaggerated immunological/inflammatory response. Notwithstanding this, the difference in the characteristics of acute inflammation due to surgical stress-induced and HIPEC-induced long-lived increases in systemic CD3⁺, CD3⁺/CD4⁺ T Helper, CD3⁺/CD8⁺ cytotoxic T, CD3⁺/CD56⁺ NK, CD19⁺ B lymphocytes, and CD4⁺ : CD8⁺ T lymphocyte ratio, statistically significant for CD3⁺/CD4⁺ T Helper and CD3⁺/CD8⁺ cytotoxic T lymphocyte populations, supports the hypothesis that HIPEC activates the immune system, with differences consistent with generalized activation of an adaptive immune response.

In conclusion, this pilot study provides the first evidence that HIPEC activates the immune response in PM patients, supporting an additional immunomodulatory function for this procedure. Of course, results from this small patient cohort pilot study must await confirmation in a larger patient cohort, which could also benefit from comparing the CRS-HIPEC-induced immune response activation to that induced by HIPEC combined with minimally-invasive surgical procedures.
Table 2  Immunological analysis: absolute values, cells/L

| ID | Tumor type | CD3⁺ | CD3⁺/CD4⁺ T helper | CD3⁺/CD8⁺ T cytotoxic | CD19⁺ B lymphocytes | CD4⁺/CD8⁺ ratio | CD3⁺ | CD3⁺/CD4⁺ T helper | CD3⁺/CD8⁺ T cytotoxic | CD19⁺ B lymphocytes | CD4⁺/CD8⁺ ratio |
|----|-------------|------|---------------------|----------------------|---------------------|----------------|------|-----------------|-----------------|-----------------|----------------|
| 1  | Colon       | 842  | 602                 | 188                  | 234                 | 61             | 3    | 1335            | 835             | 409             | 501            | 78             | 2 |
| 2  | PMP         | 970  | 505                 | 473                  | 238                 | 129            | 1    | 487             | 300             | 187             | 57             | 68             | 2 |
| 3  | PMP         | 1044 | 1500                | 1700                 | 475                 | 150            | 1    | 4477            | 2248            | 2179            | 576            | 488            | 1 |
| 4  | Colon       | 1244 | 1075                | 158                  | 44                  | 230            | 7    | 1409            | 1095            | 290             | 116            | 148            | 4 |
| 5  | PMP         | 1228 | 874                 | 340                  | 141                 | 134            | 3    | 2304            | 1443            | 846             | 421            | 141            | 2 |
| 6  | PMP         | 912  | 776                 | 147                  | 173                 | 152            | 5    | 2002            | 1184            | 782             | 334            | 184            | 2 |

PMP: Pseudomyxoma peritonei; HIPEC: Hyperthermic intraperitoneal chemotherapy; NK: Natural killer.

Table 3  Immunological response: statistical analysis

|                     | Mean (standard deviation) baseline | Median (interquartile range) baseline | Mean (standard deviation) 30 d after HIPEC | Median (interquartile range) 30 d after HIPEC | Unilateral paired Student t test | P value |
|---------------------|-----------------------------------|--------------------------------------|--------------------------------------------|--------------------------------------------|---------------------------------|---------|
| CD3⁺                | 1040.0 (± 165.8)                  | 1007.0 (912.0–1228.0)                | 2002.3 (± 1364.4)                          | 1705.5 (± 1335.0–2304.0)                 | -1.75                           | 0.07    |
| CD3⁺/CD4⁺ T helper  | 888.7 (± 360.8)                   | 825.0 (602.0–1075.0)                 | 1184.2 (± 649.7)                           | 1139.5 (835.0–1443.0)                    | -2.05                           | 0.048   |
| CD3⁺/CD8⁺ T cytotoxic | 501.0 (± 600.9)                 | 264.0 (158.0–473.0)                 | 782.2 (± 733.6)                            | 595.5 (290.0–846.0)                     | -2.05                           | 0.048   |
| CD3⁺/CD56⁺ NK       | 217.5 (± 144.9)                   | 203.5 (141.0–238.0)                 | 334.2 (± 208.9)                            | 377.5 (116.0–501.0)                     | -1.69                           | 0.07    |
| CD19⁺ B lymphocytes | 142.7 (± 54.2)                    | 142.0 (129.0–152.0)                 | 184.5 (± 155.1)                            | 144.5 (78.0–184.0)                      | -0.67                           | 0.26    |
| CD4⁺/CD8⁺ ratio     | 3.3 (± 2.3)                       | 3.0 (1.0–5.0)                       | 2.2 (± 1.0)                                | 2.0 (2.0–2.0)                           | 1.78                            | 0.07    |

HIPEC: Hyperthermic intraperitoneal chemotherapy.

Figure 2  Median values (absolute value cells/mL) of T helper and T cytotoxic levels at baseline and 30 d after hyperthermic intraperitoneal chemotherapy. HIPEC: Hyperthermic intraperitoneal chemotherapy.
immune response activation following HIPEC

ARTICLE HIGHLIGHTS

Research background
According to current Network NCCN Guidelines, cytoreductive surgery (CRS) combined with hyperthermic intraperitoneal chemotherapy (HIPEC) is considered the standard treatment for pseudomyxoma peritonei and peritoneal mesothelioma. In several clinical trials, HIPEC, in particular, resulted in survival improvements.

Research motivation
Thermal tumor ablation has been reported to induce a potential immunomodulatory effect, illustrated by the intense inflammatory cell response, including: Dendritic cells, neutrophils, macrophages, B and T lymphocytes and natural killer cells. The induction of this immune response might be important in increasing tumor response and improving survival.

Research objectives
The aim of this study was to investigate whether HIPEC for peritoneal metastases induces an immune response, which was determined by analyzing the immune population before and 30 d after HIPEC.

Research methods
Peripheral blood samples were collected prior to (day 0) and following (day 30) CRS-HIPEC procedures, and immunophenotypic analysis was performed within 24 h. The levels of immunological populations were quantified by fluorescence-activated cell sorting, an antibody-based cell sorting method for heterogeneous mixtures based upon the specific light scattering and fluorescent characteristics of each antibody-labelled cell-type as reported in the literature. Immunological populations were also analyzed in stained samples, after exclusion of debris and doublets, as previously described.

Research results
The evaluation of systemic lymphocyte populations revealed an increase in systemic CD3^+ total, CD3^+CD4^+ T Helper, CD3^+CD8^+ cytotoxic T, CD3^+CD56^- natural killer, CD19^- B lymphocyte levels and CD4^+CD8^- T lymphocyte CD3^+CD8^- Cytotoxic T lymphocyte ratios. Statistical significance was observed for CD3^+CD4^+ T Helper and CD3^+CD8^- Cytotoxic T lymphocyte populations (unilateral paired Student’s t test, P < 0.05). These increases were observed in all but one patient, who exhibited reduced systemic lymphocyte counts post-HIPEC, consistent with severe oxaliplatin toxicity.

Research conclusions
The results obtained provide new evidence that HIPEC induces immune system activation in pseudomyxoma peritonei patients, characterized by induction of a generalized adaptive immune response and decrease in immunosuppression. Another new finding was that the CRS-HIPEC-induced immunological effect was long-lived and lasted for several weeks, consistently with authentic immunomodulation, rather than a normal inflammatory response.

Research perspectives
This pilot study provides the first evidence that HIPEC activates the immune response, supporting an additional immunomodulatory function for this procedure. Further studies are required to confirm these results in a large cohort study and to evaluate treatment safety, efficacy or effectiveness.

REFERENCES

1. Neuwirth MG, Alexander HR, Karakousis GC. Then and now: cytoreductive surgery with hyperthermic intraperitoneal chemotherapy (HIPEC), a historical perspective. J Gastrointest Oncol 2016; 7: 18-28 [PMID: 26941981 DOI: 10.3978/j.issn.2078-6891.2015.106]

2. Sugarbaker PH. Peritoneectomy procedures. Ann Surg 1995; 221: 29-42 [PMID: 7826158 DOI: 10.1097/00000658-199501000-00004]

3. Morano WF, Khalili M, Chi DS, Bowen WB, Esquivel JL. Clinical studies in CRS and HIPEC: Trials, tribulations, and future directions—A systematic review. J Surg Oncol 2018; 117: 245-259 [PMID: 29120491 DOI: 10.1002/jso.24813]

4. Morano WF, Aggarwal A, Love P, Richard SD, Esquivel JL, Bowen WB. Intrapertioneal immunotherapy: historical perspectives and modern therapy. Cancer Gene Ther 2016; 23: 373-381 [PMID: 27834358 DOI: 10.1038/cgt.2016.49]

5. Verwaal VJ, van Ruth S, de Bree E, van Sloothen GW, van Tinteren H, Boot H, Zoutmulder FA. Randomized trial of cytoreduction and hyperthermic intraperitoneal chemotherapy versus systemic chemotherapy and palliative surgery in patients with peritoneal carcinomatosis of colorectal cancer. J Clin Oncol 2003; 21: 3737-3743 [PMID: 14551293 DOI: 10.1038/sj.jco.3001094]

6. Network NCCN. 2016 Colon Cancer. NCCN Guidelines Version 2. Available from: https://www.nccn.org/professionals/physician_gls/pdf/colon.pdf

7. Guadagni S, Clementi M, Mackay AR, Ricevuto E, Fiorentini G, Sarti D, Palumbo P, Apostolou P, Papasotiriou I, Masedu F, Valenti M, Giordano AV, Bruera G. Real-life multidisciplinary treatment for unresectable colorectal cancer liver metastases including hepatic artery infusion with chemo-filtration and liquid biopsy precision oncology: observational cohort study. J Cancer Res Clin Oncol 2020; 146: 1273-1290 [PMID: 32088781 DOI: 10.1007/s00432-020-03156-3]
Fiorentini G et al. Immune response activation following HIPEC

8 Fiorentini G, Sarti D, Nardella M, Inchingolo R, Nesta M, Rebomato A, Guadagni S. Chemoembolization Alone or Associated With Bevacizumab for Therapy of Colorectal Cancer Metastases: Preliminary Results of a Randomized Study. In Vivo 2020; 34: 683-686 [PMID: 32111770 DOI: 10.21873/invivo.11824]

9 Chu KF, Dupuy DE. Thermal ablation of tumours: biological mechanisms and advances in therapy. Nat Rev Cancer 2014; 14: 199-208 [PMID: 24561446 DOI: 10.1038/nrc3672]

10 Giardino A, Innamorati G, Ugol S, Perbellini O, Girelli R, Frigerio I, Regi P, Scopelliti F, Butturini G, Paletta S, Baccioni M, Bassi C. Immunomodulation after radiofrequency ablation of locally advanced pancreatic cancer by monitoring the immune response in 10 patients. Pancreatology 2017; 17: 962-966 [PMID: 29037917 DOI: 10.1016/j.pan.2017.09.008]

11 Kusamura S, González-Moreno S, Nizzi E, Baratti D, Guadagni S, Guaglio M, Battaglia L, Deraco M. Learning Curve, Training Program, and Monitoring of Surgical Performance of Peritoneal Surface Malignancies Centers. Surg Oncol Clin N Am 2018; 27: 507-517 [PMID: 29355686 DOI: 10.1016/j.soc.2018.02.009]

12 He M, Huang H, Wang M, Chen A, Ning X, Yu K, Li Q, Li W, Ma L, Chen Z, Wang X, Sun Q. Fluorescence-Activated Cell Sorting Analysis of Heterotypic Cell-in-Cell Structures. Sci Rep 2015; 5: 9588 [PMID: 25913613 DOI: 10.1038/srep09588]

13 Ito F, Ka AW, Buscek MJ, Vardam-Kaur T, Kim M, Fisher DT, Camoriano M, Khoury T, Skitzki J, Golfinck SQ, Evans SS. Immune Adjuvant Activity of Pre-Resectional Radiofrequency Ablation Protects against Local and Systemic Recurrence in Aggressive Murine Colorectal Cancer. PLoS One 2015; 10: e0143370 [PMID: 26599402 DOI: 10.1371/journal.pone.0143370]

14 Klaver CE, Musters GD, Bemelman WA, Punt CJ, Verwaal VJ, Dijkgraaf MG, Aalbers AG, van der Bilt JD, Schuitemaker IC, Meijerink WJ, Nienhuis SW, Bosjes WI, Radema SA, van Ramshorst B, Snaebjornsson P, Tuyman JB, Te Velde EA, Wierse MJ, de Witt HJ, Tanis PJ. Adjunctive hyperthermic intraperitoneal chemotherapy (HIPEC) in patients with colon cancer at high risk of peritoneal carcinomatosis: the COLOPEC randomized multicentre trial. BMC Cancer 2015; 15: 428 [PMID: 26903840 DOI: 10.1186/s12885-015-1430-7]

15 Klaver CEL, Stamm R, Sloopaun DAM, Crezee J, Bemelman WA, Punt CJA, Tanis PJ. Colorectal cancer at high risk of peritoneal metastases: long term outcomes of a pilot study on adjuvant laparoscopic HIPEC and future perspectives. Oncotarget 2017; 8: 51200-51209 [PMID: 28881641 DOI: 10.18632/oncotarget.17153]

16 Pelt JO, Chua TC, Esquivel J, Stojadinovic A, Doerfer J, Morris DL, Gershenkron K, van der Velden J, de Groot T, Verhaegen F, Evans SS. Evaluation of best supportive care and systemic chemotherapy as treatment stratified according to the retrospective peritoneal surface disease severity score (PSDSS) for peritoneal carcinomatosis of colorectal origin. BMC Cancer 2010; 10: 689 [PMID: 21176206 DOI: 10.1186/1471-2407-10-689]

17 Klaver YL, Lemmens VE, Nienhuis SW, Luyer MD, de Hingh IH. Peritoneal carcinomatosis of colorectal origin: Incidence, prognosis and treatment options. World J Gastroenterol 2012; 18: 5489-5494 [PMID: 23112540 DOI: 10.3748/wjg.v18.i39.5489]

18 Guaglio M, Simukumar S, Kusamura S, Milione M, Pietrantonio F, Battaglia L, Guadagni S, Baratti D, Deraco M. Correction to: Clinical Surveillance After Macroscopically Complete Surgery for Low-Grade Appendiceal Mucinous Neoplasms (AMN) with or Without Limited Peritoneal Spread: Long-Term Results in a Prospective Series. Ann Surg Oncol 2018; 25: 987 [PMID: 29352430 DOI: 10.1245/s10434-018-6341-9]

19 Leon AC, Davis LL, Kraemer HC. The role and interpretation of pilot studies in clinical research. J Psychosom Res 2011; 45: 626-629 [PMID: 21035130 DOI: 10.1016/j.jpsychores.2010.10.008]

20 Novitsky YW, Litwin DE, Callery MP. The net immunologic advantage of laparoscopic surgery. Surg Endosc 2004; 18: 1411-1419 [PMID: 15791361 DOI: 10.1007/s00464-003-8275-x]
