Evaluation of Parasite Density, Plasma Total Bile Acids, Alanine Transaminase, Lactate Dehydrogenase and CD4 in Plasmodium Infected Patients Treated with Morinda lucida (Oowo)

Mathew Folaranmi Olaniyan*, Elizabeth Moyinoluwa Babatunde

Department of Medical Laboratory Science, Achievers University, Owo, Ondo state, Nigeria

Abstract Morinda Lucida has been tested to have antimalarial and antidiabetic effect among other health benefits. It is commonly used for the treatment of malaria infection in Oke-Ogun and its environs. Despite the health usefulness of this extract little has been reported about its biochemical alterations. This work was therefore designed to evaluate the parasite density, plasma total bile acids, Alanine transaminase, Lactate dehydrogenase and CD4 in Plasmodium infected patients treated with Morinda Lucida (Oowo). Sixty four (64) HIV, HBsAg and anti-HCV seronagative Plasmodium infected patients visiting traditional homes in Oke-Ogun the Northern part of Oyo state Nigeria for traditional malaria cure aged 5 to 73 years classified into Female (n=34) and Male (n=30) were investigated. Hepatitis B surface antigen (HBsAg) and anti-HCV tests were carried out by Enzyme Linked Immunoenzorbent Assay (ELIZA). HIV screening and confirmation were carried out by immuno-chromatographic and Immunobloting (Western blot) assays respectively. Fasting Plasma Total Bile Acids (TBA), Lactate Dehydrogenase (LDH) and Alanine aminotransferase (ALT) were carried out biochemically by spectrophotometry while CD4 cell count was carried out by cytoflowmetry using Partec Cyflow machine and reagent. The tests were carried out in the control and test subjects before and after the treatment with the raw liquid extract of the leaf of Morinda Lucida. The result obtained showed a significantly higher mean values of falciparum parasite density and plasma ALT in the plasmodium infected test subjects than the values of the parameters obtained in plasmodium non-infected control subjects before and after treatment of the plasmodium infected subjects with the raw liquid extract of Morinda lucida with p<0.05. There was also a higher significant difference in the value of plasma LDH in the plasmodium infected test subjects before the administration of the extract than the result obtained in the plasmodium non-infected control subjects with p<0.05. A significantly higher value of plasma LDH was obtained in the test subjects before the treatment compared to the value of this parameter obtained in the test subjects after treatment (p<0.05). There was also a significantly higher mean value of TBA in the test subjects compared with the control after treatment (p<0.05). There was no significant alteration in the mean values of the CD4 cell count in the subjects before and after treatment and also in test compared with the control subjects. This recent work showed significant alterations in the mean values of the plasmodium parasite density, ALT, LDH, TBA in the test subjects studied due to plasmodium infection and the effectiveness of the raw liquid extract of the leaf of Morinda Lucida in the treatment of malaria. Estimation of plasmodium parasite density, plasma ALT, LDH, and TBA should be considered in the treatment of malaria using the raw liquid extract of the leaf of Morinda Lucida.

Keywords Morinda Lucida, ALT, LDH, TBA, CD4, Parasite density, Raw liquid extract, Traditional, Malaria

1. Introduction

The species of Plasmodium (P) that cause malaria in human which include: P. vivax, P. ovale, P. malariae and P. falciparum are transmitted by female anopheles mosquito when the mosquito is taking a blood meal from man. The pre-erythrocytic schizogony takes place in the liver. The presentation of malaria may include headache, fever, shivering, joint pain, vomiting, hemolytic anemia, jaundice, hemoglobin in the urine, retinal damage, and convulsions [1].

Morinda Lucida known as “Oowo” in Yoruba vernacular dialect of the people of Oke-Ogun and its environs at the Northern part of Oyo state, Nigeria has been analyzed to possess anti-malaria and anti-diabetic properties. It has also been emphasized that morinda lucida is good for the treatment of hypertension, stomach pain, liver problems,
Other actions of theophylline are: lowering of blood pressure, the vagus nerve, which runs from the lungs to the brain. Theobromine acts on the central nervous system. Theobromine can be used as cough medicine. Studies indicate that theobromine acts on the bronchi muscles in the lungs. Theobromine can also affect the heart, thereby slowing down nerve cell activity. Caffeine stimulates the central nervous system, respiration and blood circulation. Caffeine also acts as a diuretic. Caffeine increases the circulation and oxidation of fatty acids. This is why caffeine is used by sportsmen to increase fatty acid metabolism. Caffeine is often used in combination with aspirin to treat headaches. Caffeine can also have negative impact on health, especially if overdosed.

Theobromine a constituent of Morinda Lucida has a similar effect than caffeine, but about 10 times weaker. Theobromine has diuretic, stimulant and relaxing effects. Theobromine can lower the blood pressure because it can to dilate blood vessels. Theobromine has stimulant properties, similar to caffeine. Unlike caffeine theobromine does not affect the central nervous system. Theobromine can also relax bronchi muscles in the lungs. Theobromine can be used as cough medicine. Studies indicate that theobromine acts on the vagus nerve, which runs from the lungs to the brain. Other actions of theophylline are: lowering of blood pressure, anti-inflammatory effect and chronotropic effect.

Anthraquinone a phytochemical of Morinda Lucida include many important drugs (collectively called anthracenediones) and could be used as laxatives such as dantron, emodin, and aloe emodin, and some of the senna glycosides; antimalarials such as rufagillo; antineoplastics used in the treatment of cancer, such as mitoxantrone, pixantrone, and the anthracyclines; DNA dyes / nuclear counterstains for flow cytometry and fluorescence microscopy. Cardenolide glycosides are often toxic; specifically, they are heart-arresting. organic compounds with a characteristic ring structure many of which are found in plants (as some milkweeds), have an effect on the vertebrate heart like that of digitalis, and cause vomiting. Most alkaloids have a bitter taste or are poisonous when ingested.

Lactate dehydrogenase (LDH) catalyzes the interconversion of lactic acid and pyruvic acid. The enzyme is composed of 4 peptide chains and exists in 5 isomeric forms. LDH is widely distributed throughout the body; highest concentrations are found in the liver, heart and skeletal muscle. LDH is a general marker of tissue damage and is often used to determine the root cause and location of damage. LD activity is significantly elevated during myocardial infarction. Maximum levels are reached 24 to 48 hours after the onset of chest pain and may remain elevated for 7 to 12 days postinfarction. Increased activity is also associated with stroke, kidney disease, liver disease, progressive muscular dystrophy, cancer, intestinal and pulmonary infarction.

Bile acids are synthesised in the liver as a breakdown product of cholesterol and secreted into the gall bladder. They are released into the small intestine where they solubilise dietary lipids such as cholesterol, aiding their absorption. Bile acids are reabsorbed from the portal blood by hepatocyte extraction and re-excreted into bile, passing through the enterohepatic circulation several times before final excretion. The measurement of Total bile acids (TBA) in serum is a sensitive indicator of liver function. Fasting serum bile acids can be used in the diagnosis and prognosis of liver disease. Levels rise in many liver diseases, for example hepatitis and liver sclerosis. Abnormal levels in fasting patients or immediately after a meal can be used to detect liver disease and damage, impaired liver function, intestinal dysfunction and perhaps a gall bladder blockage. Bile acid measurement may detect some forms of liver disease earlier than standard liver tests because bile acids levels correspond to liver function, rather than liver damage. In veterinary medicine, bile acid measurement is considered to be a superior indicator of liver disease.

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Aspartate aminotransferase (AST) or Alanine Aminotransferase (ALT) is or are tested to give an indication of the degree of inflammation. They are present in hepatocytes and leak into the bloodstream if liver cells are damaged or injured. Perhaps ALT is the most specific marker as AST can also be elevated in other disease states. The aminotransferases are a group of enzymes that catalyze the
inter-conversion of amino acids and a oxoacids by transferring amino groups. The enzyme ALT (alanine aminotransferase or glutamate pyruvate transaminase) has been found to be in highest concentrations in the liver, with decreasing concentrations found in the kidneys, heart, skeletal muscle, pancreas, spleen and lung tissue respectively [12]. ALT measurements are used in the diagnosis and treatment of certain liver diseases (e.g. viral hepatitis and cirrhosis) and heart diseases. Extremely elevated levels of ALT can indicate acute hepatitis with moderately high levels indicating chronic hepatitis. Elevated levels of the transaminases can also be indicative of myocardial infarction, hepatic disease, muscular dystrophy and organ damage. Elevated levels of ALT in serum is rare except in parenchymal liver disease, since ALT is a more liver specific enzyme. ALT is often tested in combination with AST as part of a liver panel with ALT levels being higher in most types of liver disease [12].

CD4 + T helper cells are white blood cells that are an essential part of the human immune system. They are often referred to as CD4 cells, T-helper cells or T4 cells. They are called helper cells because one of their main roles is to send signals to other types of immune cells, including CD8 killer cells. CD4 cells send the signal and CD8 cells destroy the infectious particle. If CD4 cells become depleted, for example in untreated HIV infection, or following immune suppression prior to a transplant, the body is left vulnerable to a wide range of infections that it would otherwise have been able to fight [13] [14].

Total bile acids, Alanine transaminase, and Lactate dehydrogenase are liver markers that increase in plasma concentration in liver injury/diseases. The pathophysiology of Plasmodium infection involves liver which could cause a significant alteration in the plasma level of liver markers [1]. It has been said that most of the medicinal plants used for traditional treatment are hepatotoxic due to either the phytochemical constituents or dosage [1] [2] [3]. This recent work was therefore designed to evaluate the parasite density, plasma total bile acids, Alanine transaminase, Lactate dehydrogenase and CD4 in Plasmodium infected patients treated with Morinda Lucida (Oowo).

2. Materials and Methods

2.1. Materials

Study area

The study was carried out in ATISBO, Saki-East and Saki-West Local government areas of Oke-Ogun – the Northern part of Oyo state-Nigeria. The three local governments constitute the former Ifedapo Local government area of Oyo state and presently a Nigeria Federal Constituency. The three local governments share border with Kwara state-Nigeria, Ogun state-Nigeria and the Republic of Benin.

Study design

Experimental research design.

Study population

a. Sixty four (64) Plasmodium infected HIV, HBsAg and anti-HCV seronegative patients aged 5 – 73 years that have not been treated with any malaria medication but have decided to be treated traditionally using Morinda Lucida leaf extract were recruited from fifteen (15) traditional homes in ATISBO, Saki-East and Saki-West Local government areas of Oke-Ogun – the Northern part of Oyo state-Nigeria. None of the subject was jaundiced as at the time of sample collection.

Sample size

Sixty four (64)(84.2%) out of the seventy six (76) Plasmodium infected patients (Female: n= 34; Male: n=30) that visited the traditional healers between September, 2013 and April, 2014 for treatment that volunteered themselves for this study were recruited based on the inclusion and the exclusion criteria.

Case selection procedure/s

Inclusion criteria

Anicteric Plasmodium infected HIV, HBsAg and anti-HCV seronegative patients aged 5 -73 years that have not been treated with any antimalarial medication but have volunteered to be treated with raw liquid extract of the leaf of Morinda Lucida in the traditional homes were recruited.

Exclusion criteria

1. Plasmodium infected patients that have been treated or being treated with antimalarial drugs were not recruited for the study.
2. Icteric Plasmodium infected HIV, HBsAg and anti-HCV seronegative patients aged 5 -73 years that have not been treated with any antimalarial medication were not included for the study.
3. Anicteric Plasmodium infected HIV, HBsAg and anti-HCV seropositive patients aged 5 -73 years that have not been treated with any antimalarial medication were not included in the study.
4. Anicteric Plasmodium infected HIV, HBsAg and anti-HCV seropositive patients aged 5 -73 years that have not been treated with any antimalarial medication were not included in the study but on drugs such as paracetamol, contraceptives, alcohol and have been taking cigarette were not included in the study.

Blood Sample

Five (5) milliliter of blood was collected into lithium heparinized bottle from each of the control and the test subjects before and after the administration of the raw liquid of extract Morinda Lucida (Oowo) after an overnight fasting for the estimation of Plasmodium parasite density, CD4 cell count, HIV, HBsAg and anti-HCV tests, plasma Alanine transaminase, Lactate Dehydrogenase Total Bile Acids, LDL
and Total Cholesterol. The post treatment blood sample was
collected after one week of administration.

**Preparation of the raw Liquid extract of *Morinda Lucida* (Oowo)**

The leaves of the *Morinda Lucida* were plucked and
washed in water. The water was drained and the leaves were
crushed or squished for the extraction of the liquid content
into a container. A plastic cup with a capacity of 70ml was
dedicated by the healers for the measurement of the extract to
be administered into the patients. The liquid extract is freshly
prepared on daily bases prior to administration. The raw
liquid content of the leaf is extracted without the addition of
water and also administered undiluted.

2.2. Methods

a. Based on the information obtained from the fifteen
traditional homes visited in the three local governments
about 70ml of the raw undiluted liquid extract of the
*Morinda Lucida* leaf is administered to the patients on
daily bases for at least one week.

b. Estimation of plasma Alanine Transaminase was
carried out by Cobas C111 Auto-Chemistry Analyzer
using the Reagent Kit of Roche Diagnostics,
GmbhSandhoferstrasse, 116, D-68305, Mannheim.
www.roche.com.

c. Estimation of Total Bile Acids was carried out on the
plasma samples of the subjects using Randox reagent kit.
The manufacturer’s instruction was strictly followed.

Principle: Two reactions are combined in this kinetic
enzyme cycling method. In the first reaction bile acids are
oxidised by 3-α hydroxysteroid dehydrogenase with
the subsequent reduction of Thio-NAD to Thio-NADH. In the
second reaction the oxidised bile acids are reduced by the
same enzyme with the subsequent oxidation of NADH to
NAD. The rate of formation of Thio-NADH is determined
by measuring the specific absorbance change at 405nm.
(Abbreviations: NADH, NAD, Thio -NADH,
Thio-NAD)  

d. Screening for HIV Antibodies HIV screening were
carried out using Immuno chromatographic kit (Chembio
HIV 1 and 2 STAT-PAK). Positive samples were further
confirmed by Western blot/Immunoblotting using
Immunogenetics Qualicode TM HIV 1 and 2 kit.

e. Screening for HbsAg by Enzyme-Linked
Immunosorbent Assay (ELISA) The ELISA kit from
BIORAD Monolisa HBsAg ULTRA EIA92430
Marnes-La-Coquette-France was used. ELISA was done
according to the manufactures instruction. The Optical
density OD was read at 450/620 to 700 nanometre. The cut
off value was determined by the mean of negative
control + 0.05 (0.08). The test is valid if all values of
negative control are lower or equal to 0.08 and Positive
control was over 0.08 or equal to 1.0. A test sample is
considered negative if the ratio value of sample: cut off
value is lower than 1.0 and positive if equal to or greater
than 1.0.

f. Screening for HCV Antibody by ELISA ELISA kit
from DIA PRO Diagnostic Bioprobes 20099 Sesto San
Giovanni (Milano)-Italy was used. ELISA was done
according to the manufactures instruction The Optical
density OD is read at 450/620 to 700 nanometre. The cut
–off value is calculated as follows: NC (negative control)
+350= cut-off (C), Calibrator mean value=0.540, S/C=1.4
(where S= sample and C- cut off), S/C = higher than 1.1.
Any sample with a ratio value of sample /cut off less than
0.9 was considered negative and if higher than 1.1 is
positive.

g. CD4 Count was Carried out by Cytoflometry Using
the Reagent Kit of Partec and Partec CD4 Machine
h. Identification and the estimation of the density of
Plasmodium parasite in the subjects were carried out as
follows: Plasmodium spp was determined in the blood of
the control and the test subjects using Giemsha thick
blood staining technique described by Cheesbrough
(2002). Estimation of parasite number was carried out by
multiplying average number of parasites per high power
field (100x objective) by 500 within 10 fields [15] [16].

i. Plasma Lactate Dehydrogenase was estimated in the
subjects using reagent kit of Randox. The manufacturer’s
instruction was followed strictly.

**Principle**

The LDH method measures the oxidation of L-lactate to
pyruvate with simultaneous reduction of nicotinamide
adenine dinucleotide (NAD). The change in absorbance at
340 nm due to the appearance of reduced NAD (NADH) is
directly proportional to the LDH activity, since other
reactants are present in non-rate limiting quantities and is
measured using a bichromatic (340, 383 nm) rate technique.

**Ethical Consideration**

The proposal was reviewed and approved by the Research
and Ethical Committee of Baptist Medical Centre, Saki-Oyo
state-Nigeria before the commencement of the work. This is
to protect the interest of patients to ensure that the patients
and the community are not harmed in any form by the
procedure. Only plasmodium infected patients that
volunteered themselves for the study were recruited.

**Statistical Analysis**

the values of the biochemical parameters obtained in the
patients before and after treatment with the raw liquid extract
were subjected to statistical analysis to determine the mean
values, standard deviation and students ‘t’ test, for t value, p
value and level of significant at 0.01(99%) using online
Student T-Test Calculator for 2 Independent Means on line
at. http://www.socscistatistics.com/tests/studentttest
3. Result

Table 1. The value of the Falciparum parasite density, Plasma LDH, ALT, TBA, and CD4 cell count

| Parameter Description | Plasmodium parasite density/µL | Plasma LDH (U/L) | Plasma ALT (U/L) | Plasma TBA (µmol/L) | CD4 (cells/µL) |
|-----------------------|--------------------------------|------------------|------------------|---------------------|----------------|
| Mean and Standard deviation of the parameters in Normal Control subjects n = 64 | 0 ± 170 | 25 ± 10.0 | 6.0 ± 1.5 | 700 ± 20.5 |
| Mean and Standard deviation of the parameters in the test subjects before treatment n = 64 | 895 ± 30.1 | 301 ± 10.1 | 46 ± 3.0 | 14 ± 2.0 | 645 ± 25.2 |
| Mean and Standard deviation of the parameters in the test subjects after treatment n = 64 | 65 ± 16.1 | 215 ± 10 | 49 ± 2.5 | 16 ± 1.2 | 676 ± 27.3 |

Table 2. Comparative analysis of the values of the Falciparum parasite density, Plasma LDH, ALT, TBA, and CD4 cell count in the subjects

| Parameter Description | Falciparum parasite density/µL | Plasma LDH (U/L) | Plasma ALT (U/L) | Plasma TBA (µmol/L) | CD4 (cells/µL) |
|-----------------------|--------------------------------|------------------|------------------|---------------------|----------------|
| Values obtained in Control/values in test subjects before treatment | 't' value | 29.833333 | 10.99651 | 5.824352 | 3.2 | 1.874085 |
| 'p' value | 0.000561 | 0.008169 | 0.028236 | 0.085341 | 0.201772 |
| comment | P < 0.05 | P < 0.05 | P < 0.05 | P > 0.05 | P > 0.05 |
| Values obtained in Control/values in test subjects after treatment | 't' value | 4.333333 | 3.181981 | 7.652514 | 5.547002 | 0.762138 |
| 'p' value | 0.024673 | 0.043094 | 0.008325 | 0.015498 | 0.262796 |
| comment | P < 0.05 | P < 0.05 | P < 0.05 | P < 0.05 | P < 0.05 |
| Values obtained in test subjects before treatment/values in test subjects after treatment | 't' value | 24.754819 | 6.081118 | 0.707107 | 0.894427 | 0.970352 |
| 'p' value | 0.000815 | 0.012996 | 0.276393 | 0.232739 | 0.217116 |
| comment | P < 0.05 | P < 0.05 | P < 0.05 | P < 0.05 | P < 0.05 |

Note- ns: not significant  s: significant

The result obtained in this study is as shown in tables 1 & 2 above. The result showed a significantly higher mean values of Plasmodium parasite density and plasma ALT in the plasmodium infected test subjects than the values of the parameters obtained in plasmodium non-infected control subjects before and after treatment of the plasmodium infected subjects with the raw liquid extract of *Morinda lucida* with p < 0.05. There was also a higher significant difference in the value of plasma LDH in the plasmodium infected test subjects before the administration of the extract than the plasmodium non-infected control subjects with p < 0.05. A significantly higher value of plasma LDH was obtained in the test subjects before the treatment compared to the value of this parameter obtained in the test subjects after treatment (p < 0.05). There was no significant difference in the mean values of the CD4 cell count in the subjects before and after treatment and also in test compared with the control subjects (p > 0.05). There was also a significantly higher mean value of TBA in the test subjects compared with the control after treatment (p < 0.05). Also, no significant difference was obtained in the mean value of plasma LDH in the test and control subjects and also in the plasmas value of ALT and LDH in the test subjects before and after treatment with the liquid extract (p > 0.05).

4. Discussion, Conclusions and Recommendations

This work has been used to determine the alterations in the level of plasmodium parasite density, CD4 cell, LDH, ALT and TBA in the blood of Plasmodium infected patients treated with the raw liquid extract of *Morinda lucida*. A significantly higher mean values of plasmodium parasite density and plasma ALT obtained in the Plasmodium infected test subjects than the values of the parameters in plasmodium non-infected control subjects before and after treatment of the plasmodium infected subjects with the raw liquid extract of *Morinda lucida*. The mean values of the parasite density obtained before the treatment indicates plasmodium parasitaemia. The density however decreases after treatment as a result of the antimalarial effect of raw liquid extract of *Morinda lucida*. Raised plasma ALT in Plasmodium infected patients could be as a result of the liver involvement in the pathophysiology of malaria convulsions [1]. Alanine Aminotransferase (ALT) is tested to give an indication of the degree of inflammation. They are present in hepatocytes and leak into the bloodstream if liver cells are damaged or injured [12].
Significantly raised plasma LDH level in plasmidium infected subjects than the control subjects and also in the test subjects before treatment than the value obtained in the test subjects after treatment with the raw liquid extract of *Morinda lucida*. These findings could be attributed to the fact that LDH is widely distributed throughout the body; highest concentrations are found in the liver, heart and skeletal muscle. LDH is a general marker of tissue damage and is often used to determine the root cause and location of damage. Plasmodium infection is also associated with haemolysis/tissue destruction. The liver is also involved at the pre-erythrocytic stage of the life cycle of Plasmodium which may affect its normal metabolic activities may also account for these findings. Decrease in plasma LDH after treatment could also be associated with the effectiveness of *Morinda lucida* extract as an antimalarial leading to the reduction in plasmidium parasite density that may bring about less tissue destructions [7].

There was also a significantly higher mean value of TBA in the test subjects compared with the control after treatment. This could be explained by the fact that there could be liver dysfunction / hepatopathy in malaria which may not be conveniently associated with the hepatotocxic effect of raw liquid extract of *Morinda lucida*. This is because there was no significant difference in the plasmidium infected patients when the plasma ALT and TBA levels before the administration of the extract were compared with the result obtained in the patients after treatment which may rule out hepatotoxicity as this also agrees with the findings of Oduola et al., [18] that evaluated hepatotoxicity and nephrotoxicity in Wistar albino rats exposed to Morinda lucida leaf extract and found that ingestion of Morinda lucida leaf extract has no toxic effect on liver and kidney functions.

The findings of this study could also be generally be attributed to the reports of Uzuegbu and Emeka [19] and Bhatta et al., [20] that there is evidence of liver dysfunction among the malaria infected patients.

**4.1. Conclusions**

This work has been used to evaluate parasite density, plasma total bile acids, Alanine transaminase, Lactate dehydrogenase and CD4 in Plasmodium infected patients treated with *Morinda Lucida* (Oowo). The findings of the study showed a significant alteration in the plasmodium parasite density, ALT, LDH and TBA in the test subjects due to plasmodium infection and the effectiveness of the raw liquid extract of the leaf of *Morinda Lucida* in the treatment of plasmodium infection.

**4.2. Recommendations**

Estimation of Plasmodium parasite density, Plasma TBA, ALT and LDH should be considered in routine monitoring of the treatment of malaria using the leaf of *Morinda Lucida*.

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