Mechanical properties, *in vitro* corrosion and biocompatibility of newly developed biodegradable Mg-Zr-Sr-Ho alloys for biomedical applications

Yunfei Ding¹, Jixing Lin², Cuie Wen¹, Dongmei Zhang³ & Yuncang Li¹

Our previous studies have demonstrated that Mg-Zr-Sr alloys can be anticipated as excellent biodegradable implant materials for load-bearing applications. In general, rare earth elements (REEs) are widely used in magnesium (Mg) alloys with the aim of enhancing the mechanical properties of Mg-based alloys. In this study, the REE holmium (Ho) was added to an Mg-1Zr-2Sr alloy at different concentrations of Mg1Zr2SrXHo alloys (x = 0, 1, 3, 5 wt. %) and the microstructure, mechanical properties, degradation behaviour and biocompatibility of the alloys were systematically investigated. The results indicate that the addition of Ho to Mg1Zr2Sr led to the formation of the intermetallic phases MgHo₃, Mg₂Ho and Mg₁₇Sr₂ which resulted in enhanced mechanical strength and decreased degradation rates of the Mg-Zr-Sr-Ho alloys. Furthermore, Ho addition (≤5 wt. %) to Mg-Zr-Sr alloys led to enhancement of cell adhesion and proliferation of osteoblast cells on the Mg-Zr-Sr-Ho alloys. The *in vitro* biodegradation and the biocompatibility of the Mg-Zr-Sr-Ho alloys were both influenced by the Ho concentration in the Mg alloys; Mg1Zr2Sr3Ho exhibited lower degradation rates than Mg1Zr2Sr and displayed the best biocompatibility compared with the other alloys.

Biodegradable implants used in the human body are essential to provide adequate mechanical integrity and a suitable corrosion rate before sufficient tissue healing. Biomaterials should also have acceptable biocompatibility and minimal deleterious effects on organisms. Importantly, the components of biomaterials should be not only biocompatible, but also promote the growth and healing of tissue. Among the metallic biomaterials for hard tissue engineering, magnesium (Mg) alloys are receiving increasing attention as promising biodegradable materials for orthopaedic applications because of their remarkable advantages. Although significant progress has been achieved in the development of biodegradable Mg alloys, some issues, such as rapid corrosion with generated hydrogen gas, weakened mechanical integrity over time and potential toxicity of their components, still restrict their application. Hence, it is crucial to develop Mg alloys with bio-friendly alloying elements and enhanced biocorrosion resistance.

Recent studies have found that rare earth elements (REEs) exhibit many desirable advantages in Mg alloys, such as improved corrosion resistance and enhanced mechanical properties. REE-containing Mg alloys are the most successful biodegradable alloys for biomedical applications; e.g. WE43 has been successfully used in clinical application. Among REEs, Ho (>0.5 wt. % hereafter) has been used to enhance the tensile properties of Mg-Zn-Al due to the reduction of the petal-like phase and Ho also displays potential effects on grain refinement, resulting in significant improvement in strength and plasticity. However, in view of the performance of Ho in relation to biocompatibility, a limited number of studies made comparisons under identical experimental conditions. Although Mg-REE-based alloys for cardiovascular applications have been used in clinical trials, for the bulk of Mg-REE alloys in orthopaedic applications, concerns about the biosafety and usage of REEs have...
been be raised\(^1\(^8\)\), as there is no consensus on their safe dosage\(^4\). Therefore, it is essential to understand the roles of REEs in the microstructure, mechanical properties, corrosion and biocompatibility of Mg alloys.

A recent breakthrough in the development of Mg alloys for orthopaedic applications was achieved by a group led by Li\(^1\(^9\)\), leading to a series of Mg-Zr-Sr alloys with enhanced corrosion resistance compared to cast pure Mg and excellent biocompatibility in the form of support of cell adhesion and spreading. In detail, the addition of zirconium (Zr) to Mg alloys can significantly refine the grain size, which benefits the mechanical properties and corrosion resistance\(^2\(^0\)-\(^2\(^1\)\). The alloying element strontium (Sr) significantly enhances the replication of preosteoblastic cells, and even stimulates bone formation\(^2\(^2\)-\(^2\(^4\)\). Considering the benefits of Zr, Sr and Ho in enhancing biocompatibility, corrosion and mechanical properties, a new series of Mg\(_{1}\)Zr\(_{2}\)Sr\(_{x}\)Ho alloys (\(x = 1, 3, 5\%\)) have been developed in this study to satisfy the urgent and exclusive requirements for hard tissue engineering.

Results

The microstructures of the Mg-Zr-Sr-Ho alloys are shown in Fig. 1a–d. The grain size of Mg\(_{1}\)Zr\(_{2}\)Sr\(_{1}\)Ho (200–1000 \(\mu\)m) is relatively larger than that of Mg\(_{1}\)Zr\(_{2}\)Sr (20–50 \(\mu\)m) and the other Mg-Zr-Sr-Ho alloys (20–80 \(\mu\)m). However, with the increasing addition of Ho (from 1\% to 5\%), the grain size dramatically decreased in Mg\(_{1}\)Zr\(_{2}\)Sr\(_{3}\)Ho and Mg\(_{1}\)Zr\(_{2}\)Sr\(_{5}\)Ho, compared to Mg\(_{1}\)Zr\(_{2}\)Sr\(_{1}\)Ho. Also, some tiny black particles can be seen in Mg\(_{1}\)Zr\(_{2}\)Sr and Mg\(_{1}\)Zr\(_{2}\)Sr\(_{1}\)Ho, while Mg\(_{1}\)Zr\(_{2}\)Sr\(_{3}\)Ho and Mg\(_{1}\)Zr\(_{2}\)Sr\(_{5}\)Ho showed homogeneous structures without the appearance of black particles. The XRD patterns of the Mg-Zr-Sr-Ho alloys are shown in Fig. S1, displaying the characteristic crystalline structure of Mg-Zr-Sr-Ho alloys, including Mg, Zr, Mg\(_{17}\)Sr\(_{2}\) and Mg-Ho phases. It is evident that the intensity of the peaks for the Mg\(_{17}\)Sr\(_{2}\) phase slightly decreased with increasing Ho addition in the Mg\(_{1}\)Zr\(_{2}\)Sr\(_{x}\)Ho alloys. Two different Ho-containing phases, that is, MgHo\(_3\) (JCPDS No. 03-065-7200) and Mg\(_2\)Ho (JCPDS No. 04-002-0737), were detected in Mg\(_{1}\)Zr\(_{2}\)Sr\(_{3}\)Ho and Mg\(_{1}\)Zr\(_{2}\)Sr\(_{5}\)Ho, suggesting that the Ho addition to Mg\(_{1}\)Zr\(_{2}\)Sr led to the formation of the intermetallic Mg-Ho phases. The volume fraction of Mg\(_{1}\)Zr\(_{2}\)Sr\(_{3}\)Ho and Mg\(_{1}\)Zr\(_{2}\)Sr\(_{5}\)Ho, suggesting that the Ho addition to Mg\(_{1}\)Zr\(_{2}\)Sr led to the formation of the intermetallic Mg-Ho phases. In addition, the Mg\(_{1}\)Ho\(_{3}\) and Mg\(_{1}\)Ho\(_{5}\) phases exhibited different intensities in the Mg-Zr-Sr-Ho alloys. The volume fraction of Mg\(_{1}\)Ho in Mg\(_{1}\)Zr\(_{2}\)Sr\(_{3}\)Ho was much higher than in Mg\(_{1}\)Zr\(_{2}\)Sr\(_{1}\)Ho and Mg\(_{1}\)Zr\(_{2}\)Sr\(_{5}\)Ho (Fig. 1 and Fig. S1); whereas the intensity of Mg\(_{1}\)Ho increased dramatically with the increase in Ho addition to Mg\(_{1}\)Zr\(_{2}\)Sr\(_{5}\)Ho, indicating a higher quantity of Mg\(_{1}\)Ho than in Mg\(_{1}\)Zr\(_{2}\)Sr\(_{3}\)Ho and Mg\(_{1}\)Zr\(_{2}\)Sr\(_{1}\)Ho. Energy-dispersive X-ray spectroscopy (EDX) mapping of the elemental distribution over the Mg-Zr-Sr-Ho alloys (Fig. 1e,f) reveals that the intermetallic phases of Mg\(_{1}\), Sr\(_{2}\), Mg\(_{1}\)Ho\(_{3}\) and Mg\(_{1}\)Ho\(_{5}\) were mainly distributed along grain boundaries, and the signals for Sr were surpassed by those of the Mg-Ho phases, in agreement with the XRD analysis and confirming that the Ho additions to the Mg-Zr-Sr-Ho alloys influenced the microstructures of the alloys.
Figure 2a shows the Tafel curves of the Mg-Zr-Sr-Ho alloys after immersion in simulated body fluid (SBF) for 2 h at 37 ± 0.5 °C. As can be seen, Mg1Zr2Sr3Ho presented the lowest current density compared with Mg1Zr2Sr and the other Mg-Zr-Sr-Ho alloys. In contrast, Mg1Zr2Sr1Ho exhibited the highest current density among the Mg-Zr-Sr-Ho alloys, indicating the lowest corrosion resistance in SBF. Compared to Mg1Zr2Sr1Ho and Mg1Zr2Sr3Ho, Mg1Zr2Sr5Ho displayed a higher potential, indicating that MgHo3 and Mg2Ho phases (XRD, Fig. S1) have a different potential compared with the Mg matrix, and the increased volume fraction of the intermetallic phases created a higher open-circuit potential for the Mg-Zr-Sr-Ho alloys.

Analogous to the polarisation test, which represents corrosion behaviour at a select point of time, hydrogen evolution is a reliable method to assess the average corrosion rates of Mg alloys25. The rate of generated hydrogen gas against immersion time in SBF is shown in Fig. 2b. After washing using a 200 g L−1 chromic acid solution, the weight loss rate was measured and its results are listed in Fig. 2c. The fitting results of polarisation tests, and the corrosion rates based on polarization test, hydrogen evolution and weight loss are summarised in Table 1 and Fig. 2d, respectively. As can be seen, although the values of the corrosion rate based on these various assessment methods are different, the results reveal that Mg alloys with different Ho additions suffered different attacks from SBF in the increasing order of Mg1Zr2Sr3Ho < Mg1Zr2Sr1Ho < Mg1Zr2Sr3Ho < Mg1Zr2Sr, which suggests that the addition of Ho to Mg1Zr2Sr is beneficial to corrosion resistance and that Mg1Zr2Sr3Ho is relatively stable compared to the other Mg-Zr-Sr-Ho alloys.

The dissolution of Mg in a physiological environment generally involves an electrochemical reaction with water and produces magnesium hydroxide (Mg(OH)2) and hydrogen gas (H2): Mg + 2H2O → Mg(OH)2 + H2. The generated Mg(OH)2 at the initial stage provides temporary protection for the Mg alloys, retarding further corrosion26. Thus, it is essential to identify the corrosion products on the surfaces of the Mg alloys. It can be seen that the corrosion products on the Mg-Zr-Sr-Ho alloys mainly contained MgO and Mg(OH)2 (Fig. S2).

Table 1. Corrosion of Mg1Zr2Sr and Mg-Zr-Sr-Ho alloys in SBF. *Calculated based on H2 evolution; **calculated based on weight loss; ***calculated based on potentiodynamic polarisation.

| Materials         | Vψ (ml cm−2 day−1) | Pψ (mm year−1) | ΔW (mg cm−2 day−1) | Pi** (mm year−1) | Δm (mg cm−2 day−1) | Pw*** (mm year−1) |
|-------------------|---------------------|-----------------|--------------------|-----------------|---------------------|-------------------|
| Mg1Zr2Sr1Ho       | 6.13 ± 0.14         | 13.96 ± 0.38    | 6.67 ± 0.14        | 14.02 ± 0.29    | 6.74 ± 0.14        | 14.02 ± 0.29      |
| Mg1Zr2Sr3Ho       | 4.01 ± 0.09         | 9.15 ± 0.26     | 5.19 ± 0.16        | 10.90 ± 0.35    | 5.75 ± 0.17        | 10.90 ± 0.35      |
| Mg1Zr2Sr5Ho       | 4.86 ± 0.14         | 11.08 ± 0.38    | 6.55 ± 0.15        | 13.76 ± 0.33    | 7.22 ± 0.19        | 13.76 ± 0.33      |
| Mg1Zr2Sr          | 3.31 ± 0.11         | 14.39 ± 0.31    | 7.05 ± 0.17        | 14.80 ± 0.36    | 7.69 ± 0.18        | 14.80 ± 0.36      |

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Specifically, the intensities of the MgO and Mg(OH)₂ peaks in Mg1Zr2Sr3Ho are higher than those of the other alloys, indicating better protection provided by the corrosion products (Fig. S2b).

Figure 3 shows the corrosion morphologies of the Mg-Zr-Sr-Ho alloys after immersion in SBF for various time periods (24 h, 72 h, and 120 h). It can be seen that the surface of each alloy was covered with a layer of corrosion product after immersion in SBF for 24 h (Fig. 3a–c). The corrosion products on Mg1Zr2Sr3Ho and Mg1Zr2Sr5Ho exhibited a similar corrosive pattern and a loose structure, indicating a lower corrosion rate than that of Mg1Zr2Sr1Ho. EDX of the selected region of Mg1Zr2Sr5Ho (Fig. 3c) revealed that the corrosion products were mainly composed of magnesium, oxygen, chlorine and calcium. However, chlorine- and calcium-containing phases were not identified in XRD patterns (Fig. S2), which might be explained by the low quantity on the surfaces.

The corrosion morphologies of the Mg-Zr-Sr-Ho alloys after immersion in SBF for 72 h are shown in Fig. 3d–f. Compared to Mg1Zr2Sr1Ho and Mg1Zr2Sr5Ho, Mg1Zr2Sr3Ho still maintained its surface integrity and a relatively dense structure (Fig. 3e). EDX investigation of selected region of Mg1Zr2Sr5Ho (Fig. 3f) revealed that the corrosion products consisted of chlorine, oxygen and magnesium (Fig. 3f). It is evident that the chlorine from SBF deteriorated the corrosion products (MgO and Mg(OH)₂)²⁷, and thus the corrosion rate of Mg1Zr2Sr5Ho subsequently increased due to the absence of temporary protection by the corrosion products.

After 120 h of immersion in SBF, the Mg-Zr-Sr-Ho alloys were further deteriorated. At highly magnified SEM image of Mg1Zr2Sr3Ho demonstrates that the corrosion products broke down into a network structure (Fig. 3h). In contrast, a rough surface can be observed on Mg1Zr2Sr1Ho, indicating large quantities of the Mg matrix dissolved into SBF (Fig. 3g). For the Mg1Zr2Sr5Ho alloy the surface was further deteriorated, as indicated by the increased signal of Cl⁻ (Fig. 3i).

Figure 3. Corrosion morphologies of Mg-Zr-Sr-Ho alloys after immersion in SBF for: (a–c) 24 h, (d–f) 72 h, (g–i) 120 h; and (j–l) overall corrosion morphologies of Mg alloys after 168 h immersion in SBF.
The overall surfaces of the Mg-Zr-Sr-Ho alloys after immersion in SBF for 168 h exhibited differently corroded morphologies, indicating different corrosion behaviours of the alloys (Fig. 3j–l). The surface of Mg1Zr2Sr3Ho still retained its overall integrity and relatively uniform corrosion, demonstrating the highest corrosion resistance in SBF (Fig. 3k), while grooves were formed on the surface of Mg1Zr2Sr1Ho by severe attacks from SBF ions, and the regions undergoing severe attacks were much wider and deeper than in other Mg-Zr-Sr-Ho alloys (Fig. 3j). As for Mg1Zr2Sr5Ho, the majority of the surface was covered with white corrosion products; pits are also visible in some regions and a long corroded crack extends to the border (Fig. 3l).

Compressive testing has been widely used to evaluate the mechanical properties of Mg alloys, as it possesses the advantage of simple operation by loading circular cylinder samples between rigid platens which effectively constrain the movement of the samples at the loaded interfaces. Figure 4 shows the yield strength, the ultimate compressive strength, and the strain of the Mg-Zr-Sr-Ho alloys. It can be seen that the yield strength of Mg1Zr2Sr1Ho was lower than that of Mg1Zr2Sr (Fig. 4a), however, it increased with the increase in the addition of Ho. Similarly, Mg1Zr2Sr3Ho and Mg1Zr2Sr5Ho exhibited higher compressive yield strengths than those of Mg1Zr2Sr and Mg1Zr2Sr1Ho (Fig. 4b). The ultimate compressive strength of the Mg-Zr-Sr-Ho alloys increased with the increase in Ho addition, suggesting that the addition of Ho enhances compressive strength (Fig. 4b). It is worth noting that Mg1Zr2Sr1Ho showed the highest ultimate strain (~34%) and the ultimate strain decreased with the increase in the addition of Ho. It can be concluded that the concentration of Ho in Mg-Zr-Sr-Ho alloys has a significant effect on their mechanical properties.

In general, a polygonal shape for cells represents an improvement of cell conditions. Once in contact with an implant, cells explore it using filopodia, lamellipodia and peripheral ruffles, which contribute to polarised edges. After cell seeding and culturing for 120 h, SaOS2 cells displayed full spreading with a spindle-shaped morphology on the surfaces of Mg1Zr2Sr3Ho (Fig. 5e). Compared to the other Mg-Zr-Sr-Ho alloys, Mg1Zr2Sr3Ho exhibited desirable morphology, attachment and spreading of the cultured SaOS2, as indicated by flourishing growths of filopodia with widely extended microspikes solidly attached to the surface of the alloy (highly magnified SEM, Fig. 5h). In contrast to Mg1Zr2Sr3Ho with high proliferation of SaOS2 cells, Mg1Zr2Sr1Ho displayed the lowest cell viability after culturing for 120 h (Fig. 5d,g). In the case of Mg1Zr2Sr5Ho, although some SaOS2 cells on its surface displayed shrinking-shaped rounds without long filopodia after seeding and culturing for 120 h (Fig. 5f), Mg1Zr2Sr5Ho still exhibited superior cell proliferation to Mg1Zr2Sr1Ho in terms of cell spread and viability.

The in vitro cytotoxicity of the Mg-Zr-Sr-Ho alloys was assessed using osteoblast-like cells (SaOS2). The optical densities in the extract of each alloy were measured using MTS assay and are listed in Fig. 6. The optical density measured by MTS assay is proportional to the number of cell. The control group was regarded as biocompatible. A higher optical density of an Mg alloy compared with that of control group represents a lower cytotoxicity. After culturing for 24 h, the cytotoxicity of Mg1Zr2Sr3Ho was slightly lower than that of the other Mg-Zr-Sr-Ho alloys, which exhibited a similar level of cytotoxicity to cast Mg and the control group. After culturing for 72 h, a remarkable difference in the cytotoxicity of the alloys was characterized. Specifically, the Mg-Zr-Sr-Ho alloys displayed relatively lower cytotoxicity compared to pure Mg, Mg1Zr2Sr and the control group, indicating that the addition of Ho (≤5%) to Mg alloys enhanced the growth of SaOS2 cells. After culturing for 120 h, Mg1Zr2Sr3Ho and Mg1Zr2Sr5Ho still maintained lower cytotoxicity than the other groups. The results of the in vitro cytotoxicity assessment of Mg-Zr-Sr-Ho alloys offer evidence that their cytotoxicity is associated with their corrosion rate: a lower corrosion rate always denotes a lower cytotoxicity.

Discussion

In this study, an increase in Ho addition significantly enhanced the compressive yield strength (Fig. 4a) and the ultimate compressive strength, but reduced the ultimate strain (Fig. 4b) of the Mg-Zr-Sr-Ho alloys, which is in agreement with the conclusions of previous studies on the effects of REEs on the mechanical properties of Mg alloys. The results of this study indicate that the ultimate compressive strength was significantly improved and the ultimate strain decreased with increasing Ho addition from 1% to 5% (Fig. 4). This is due to the following two reasons. First, the volume fraction of the Ho-containing intermetallic phases (MgHo3 and Mg2Ho) increased with an increase in the Ho of the Mg alloys. The intermetallic phases have inherent high hardness and strength, so high volumes of Mg-Ho phases will cause high strength. Furthermore, in this study the MgHo3 and Mg2Ho...
phases were mainly distributed along the grain boundaries. This may restrain the progress of the deformation during loading-deformation and can lead to an increase in ultimate compressive strength but decreased ultimate strain. Second, in the Mg-Zr-Sr-Ho alloys, Ho distributed not only along the grain boundaries, but also in the interiors of the Mg matrix (EDX mapping, Fig. 1). As a result, Ho addition might contribute to the solid solution strengthening because of the significant difference in atomic radius between Mg (1.73 Å) and Ho (2.3 Å), as a large difference in atomic radius would lead to substantial lattice distortion and enhance the solid solution strengthening of Mg alloys.

In this study, the Mg-Ho phases were identified as MgHo₃ and Mg₂Ho, which differs from the reported Mg-Zn-Ho phases and the Mg₂4Ho₅ phase along the grain boundaries. An isolation between Sr and the Mg matrix was introduced with the increased addition of Ho (EDX mapping, Fig. 1), which weakened the galvanic effects, and reduced the corrosion rates of the Mg-Zr-Sr-Ho alloys. The effect of Ho on the corrosion behaviour of Mg-Zr-Sr-Ho alloys in most cases depends on the composition of the alloys, and the volume fraction and the distribution of the intermetallic phases. This can also be seen in the changes in the corrosion potential of the alloys after immersion in SBF for 2 h (Fig. 2a). Due to the addition of Ho, the corrosion potential of Mg₁Zr₂Sr₂

Figure 5. Cell adhesion and cell numbers on the Mg-Zr-Sr-Ho alloys after cell culture for various periods: (a–c) 24 h, (d–f) 120 h; and (g–i) highly magnified SEM images of cells after culturing for 120 h.

Figure 6. Cell numbers on Mg-Zr-Sr-Ho alloys after incubation for various periods.

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[Image of cell culture results]

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1.6345 $V_{\text{SCE}}$ decreased to $-1.6401 V_{\text{SCE}}$ for Mg1Zr2Sr1Ho. This suggests that the Mg$_{17}$Sr$_3$ and Mg-Ho phases have different thermodynamic stability. The addition of Ho to Mg alloys may eliminate the galvanic effects between the Mg matrix and the other intermetallic phases, and thus can enhance the corrosion resistance.

In addition to the effect of the Mg-Ho phases, the impurities of Mg-Zr-Sr-Ho alloys may also influence the corrosion rates. Zhou et al. found that the addition of Ho to AZ91D alloys resulted in the formation of an AlMnHo phase; this newly formed phase not only reduced the volume fractions of Mg-Al phases, but also eliminated the effects of impurities on the corrosion of Mg alloys. As can be seen from Table S1, the Mg alloys contained Fe, Al, Mn and Si. Although Mn has been considered to be beneficial in relation to the transformation of Fe and other impurities into harmless intermetallic phases, the relatively higher concentrations of Fe (0.05 wt. %) in Mg1Zr2Sr1Ho can still deteriorate the corrosion resistance, as traces of Fe (≥0.003 wt. %) are detrimental to the corrosion resistance of Mg alloys.

The utilisation of Ho in clinical applications mainly relates to the controllable radioactivity of the isotope element and the wide application of Ho in the treatment of tumors and has also made contributions to the treatment of cancer in human using Ho-doped microspheres with acceptable biocompatibility for the desired curative effects by controlling the concentration of Ho in vivo. Given the popularity of using REEs in Mg alloys, researchers have explored Mg-Ho alloys in order to obtain desirable qualities of acceptable corrosion resistance, suitable mechanical properties and excellent curative effects. Nevertheless, the biocompatibility of Ho, and in particular its biological role in Mg-Ho based alloys as hard tissue implants, has not been covered in these studies.

In this study, the biocompatibility of Ho has been explored by comparing the cytotoxicity and the performance of SBF seeded on the Mg-Zr-Sr-Ho alloys. Compared with the control group, Ho addition to Mg1Zr2Sr boosted cell growth and proliferation. In particular, the cytotoxicity and cell densities of Mg-Zr-Sr-Ho alloys were dependent on Ho concentration. More specifically, the cell on Mg1Zr2Sr3Ho which exhibited the slowest corrosion rate among the Mg-Zr-Sr-Ho alloys were much higher in number than those of the other alloys after culturing for 120 h. This suggests that the dose of Ho is critical for designing and preparing Ho-containing Mg alloys, as a high dose of Ho has been demonstrated to have negative effects on biocompatibility. In another study, rats fed with Ho chloride in their diet at doses up to 500 mg (kg day)$^{-1}$ for more than 2 months showed no adverse effect. Although the present study does not provide accurate Ho concentrations in relation to toxicity, it suggests that Ho is a biocompatible element and can be used as one of the elements in Mg alloys for hard tissue engineering by carefully selecting its concentration (≤5 wt. %) and monitoring the corrosion rates of Mg-Ho based alloys, because the dissolution of Mg alloys is always accompanied by the release of ions into the body fluid. Rapid corrosion rate of an Mg alloy may release an overdose of metallic ions which simultaneously affects tissue growth and cell proliferation.

Rapid corrosion of Mg alloys always gives rise to more corrosion products. Thus, Mg alloys with different corrosion rates exhibit various surface conditions in terms of topography and roughness. A rough surface with defects, e.g., cracks, on the surface of Mg alloys, does not provide an ideal substrate for cell growth and adhesion. As can be seen from Fig. 5a,d, Mg1Zr2Sr1Ho initially had a rough surface with many cracks after cell culturing for 24 h, and these cracks were dramatically enlarged after cell culturing for 120 h in extracts. Larger and deeper cracks allow more attack from the components of the extracts, leading to accelerated corrosion of the Mg matrix. Thus, the generated hydrogen gas and the volatile pH value adjacent to the surface of Mg alloys, along with the peeled-off substrate, further deteriorated the cell growth and adhesion (Fig. 5d,g).

The corrosion process of Mg-Zr-Sr-Ho alloys in a physiological environment can be illustrated in five stages, as shown in Fig. 7 based on the results obtained in the present study.

**Stage I.** Hydrogen evolution. Hydrogen evolution initiates at the interface between the Mg alloy and SBF due to the attack of components such as cations, organic substances and anions of the SBF, and Mg ions are released into the SBF so that an MgO/Mg(OH)$_2$ layer is formed on the surface of the Mg alloys.

**Stage II.** Mg degradation. As prolonged immersion time in SBF, some regions of the MgO/Mg(OH)$_2$ layer convert to Mg$^{2+}$ due to further attack from SBF (Fig. 5a,c). Consequently, the Mg substrate in these regions is exposed to the medium directly, leading to further degradation.

**Stage III.** Interface degradation. As the degradation proceeds, more regions of the MgO/Mg(OH)$_2$ layer are corroded constantly and more Mg substrates are exposed (Fig. S2c). Mg$^{2+}$ may pass through the loosened MgO/Mg(OH)$_2$ layer and form a new MgO/Mg(OH)$_2$ layer on the exterior surface (Fig. 3g,h). Various components can also penetrate the loosened surface and attack the interior Mg substrate, leading to interface degradation between the Mg matrix and the components of the SBF beneath the surface layer. Meanwhile, the galvanic effects between Mg$_7$Sr$_3$ and the Mg matrix will be weakened due to the presence of the Mg$_2$O$_3$ and Mg$_2$O phases, which have a corrosion potential closer to that of the Mg matrix and are distributed along the grain boundaries.

**Stage IV.** Degradation shift. The newly formed MgO/Mg(OH)$_2$ layer cannot resist dissolution. Hence, uniform dissolution occurs and some regions are inevitably exposed to the SBF. The outer MgO/Mg(OH)$_2$ layer is dissolved continuously and the corrosion extends to the interior Mg substrate and more regions would be exposed to the SBF.

**Stage V.** Massive degradation. Some corroded residues may fall off the surface of Mg alloys, causing local pits in the Mg substrate and deeper cavities. Accordingly, the Mg matrix will be further attacked due to the galvanic effects and finally the Mg alloys will degrade completely in the SBF, resulting in the massive degradation.

This study has explicated a promising Mg alloy system (Mg1Zr2Sr3Ho) with excellent corrosion resistance compared to recently developed Mg alloys, such as Mg1Zr2Sr$^{39}$, Mg-O alloys$^{34}$ and coated Mg-Al based alloys$^{35}$. 

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**References:**

1. Zhou et al. (2017) J. Biomed. Mater. Res. A, 105(11), 2552-2561.
2. Zhang et al. (2018) Biomaterials, 183, 137-149.
3. Li et al. (2019) Acta Biomater., 81, 169-180.
4. Chen et al. (2020) J. Mater. Chem. B, 8(48), 8382-8393.
5. Wang et al. (2021) Adv. Funct. Mater., 31(40), 2105218.
Although the Mg1Zr2Sr3Ho alloy showed excellent performance in the assessment of cytotoxicity, there are still challenges in fully understanding its biocompatibility. Particularly from the aspect of the development of biodegradable Mg alloy implants, it is essential to carry out thorough experimentation in vivo to provide accurate data on the safe dosage and biological properties, through taking advantage of new techniques such as the newly developed 3D printing of Mg alloys, adjustment the phase constitutions and creative methods such as “materials smartness” and “bridge the gap between in vitro and in vivo”.

Methods

Mg1Zr2Sr3xHo alloys (x = 1, 3, 5 wt. %) were fabricated by melting of pure Mg, Mg-30Zr, Mg30Sr and Mg10Ho alloys (Hunan Rare Earth Metal Material Research Institute, China) with the designated compositions. The chemical compositions of the Mg alloys (Table S1) were determined by wavelength dispersion X-ray fluorescence (WDXRF, S4 Pioneer, Bruker, Germany). Samples with a diameter of 10 mm and thickness of 3 mm were further machined using electrical discharge machining (EDM) along the long axis of the Mg alloy ingots for microstructure characterisation, corrosion measurement and biocompatibility assessment. Cylindrical samples with a diameter of 5 mm and length of 10 mm were processed in the same way from the Mg alloy ingots for the compressive test.

Basic characteristics, including microstructure analysis, were investigated using optical microscopy (Olympus, DP70), scanning electron microscopy (SEM, Supra 55, Zeiss) equipped with energy-dispersive X-ray analysis (EDX) mapping technique, and X-ray diffraction (XRD-X’pert, Philips, The Netherlands) employing Cu-Kα radiation at an accelerating voltage of 40 kV and a current of 40 mA. To evaluate the effects of Ho addition on the mechanical properties of Mg alloys, compressive test was conducted on the Mg1Zr2Sr3xHo samples at an initial strain rate of 10^−3 s^−1 using an Instron universal tester equipped with a video extensometer (Instron 5567, USA). The degradation behaviour of the Mg1Zr2Sr3xHo alloys was investigated using polarization measurement with a scan scope ±250 mv against open-circuit potential using a multichannel potentiostat (VSP 300, Bio-logic France), and immersion tests in SBF (the compositions are listed in Table S2), including hydrogen evolution, weight loss and characterization of degradation morphologies after immersion in SBF for various periods. For hydrogen evolution and weight loss, the SBF was refreshed every 4 h in order to maintain a stable pH value around 7.4. The hydrogen evolution rate V_h (ml cm⁻² day⁻¹) can be converted into the degradation rate P_h (mm year⁻¹) using the equation given by P_h = 2.279V_h 59, and the corrosion rate (P_w, mm per year) based on weight loss was calculated from the weight loss rate ΔW (mg cm⁻² day⁻¹) using the equation P_w = 2.10 ΔW 60.

To assess biocompatibility, SaOS2 cells were used to evaluate the cytotoxicity of the Mg alloys. These cytotoxicity assessments were carried out using extracts which were prepared using MEM α to extract Mg alloy specimens in a humidified atmosphere of 5% CO₂ at 37°C for 72 h (the details of the biocompatibility assessment are provided in the supporting information).

References

1. Witte, F. et al. Degradable biomaterials based on magnesium corrosion. Curr. Opin. Solid. State. Mater. Sci. 12, 63–72 (2008).
2. Staiger, M. P., Pietak, A. M., Huadmai, J. & Dias, G. Magnesium and its alloys as orthopedic biomaterials: A review. Biomaterials 27, 1728–1734 (2006).
3. Fischer, J. et al. Interference of magnesium corrosion with tetrazolium-based cytotoxicity assays. *Acta Biomater.* 6, 1813–1823 (2010).
4. Ding, Y., Wen, C., Hodgson, P. & Li, Y. Effects of alloying elements on the corrosion behavior and biocompatibility of biodegradable magnesium alloys: a review. *J. Mater. Chem. B* 2, 1912–1933 (2014).
5. Zheng, Y. F., Gu, X. N. & Witte, F. Biodegradable metals. *Mater. Sci. Eng. R: Rep.* 77, 1–34 (2014).
6. Li, H. F. et al. Development of biodegradable Zn–I-X binary alloys with nutrient alloying elements Mg, Ca and Sr. *Sci. Rep.* 5, doi: 10.1038/srep10719 (2015).
7. Birbilis, N. et al. A combined neural network and mechanistic approach for the prediction of corrosion rate and yield strength of magnesium–rare earth alloys. *Corros. Sci.* 53, 168–176 (2011).

8. Al-Samman, T. & Li, X. Sheet texture modification in magnesium-based alloys by selective rare earth alloying. *Mater. Sci. Eng. A* 528, 3809–3822 (2011).
9. Bayani, H. & Saebooroo, E. Effect of rare earth elements addition on thermal fatigue behaviors of AZ91 magnesium alloy. *J. Rare Earth* 27, 255–258 (2009).
10. Zhang, J. et al. New horizon for high performance Mg-based biomaterial with uniform degradation behavior: Formation of stacking faults. *Sci. Rep.* 5, 13933, doi: 10.1038/srep13933 (2015).
11. Zeng, R. C., Li, X. T., Li, S. Q., Zhang, F. & Han, E.-H. *In vitro* degradation of pure Mg in response to glucose. *Sci. Rep.* 5, 13026, doi: 10.1038/srep13026 (2015).
12. Cha, P.-R. et al. Biodegradability engineering of biodegradable Mg alloys: Tailoring the electrochemical properties and microstructure of constituent phases. *Sci. Rep.* 3, 2367, doi: 10.1038/srep02367 (2013).
13. Waksman, R. et al. Safety and efficacy of bioabsorbable magnesium alloy stents in porcine coronary arteries. *Catheter. Cardiovasc. Interv.* 66, 607–617 (2006).
14. Xu, C. X., Zhang, Z. W., Ju, H. & Jia, Y. B. Effect of Ho on microstructure and mechanical properties of ZA52 magnesium alloy. *Foundry* 61, 661–665 (2012).
15. Li, D., Wang, Q. & Ding, W. Effects of Ho on the microstructure and mechanical properties of Mg–Zn–Ho–Zr magnesium alloys. *Rare Metals* 30, 131–136 (2011).
16. Drynda, A., Deinet, N., Braun, N. & Peuster, M. Rare earth metals used in biodegradable magnesium-based stents do not interfere with proliferation of smooth muscle cells but do induce the upregulation of inflammatory genes. *J. Biomed. Mater. Res. A* 91A, 360–369 (2009).
17. Qu, A., Wang, C. R. & Bo, J. Research on the cytotoxic and genotoxic effects of rare-earth element holmium to Vicia faba. *Hereditas.* 26, 195–201 (2004).
18. Li, N. & Zheng, Y. Novel Magnesium Alloys Developed for Biomedical Application: A Review. *J. Mater. Sci. Technol.* 29, 489–502 (2013).
19. Li, Y. et al. Mg–Zr–Sr alloys as biodegradable implant materials. *Acta Biomater.* 8, 3177–3188 (2012).
20. Li, Y. C., Wong, C. S., Wen, C. & Hodgson, P. D. Biodegradable Mg-Zr-Ca alloys for bone implant materials. *Mater Technol.* 27, 49–51 (2012).
21. Lee, Y. C., Dahle, A. K. & StJohn, D. H. The role of solute in grain refinement of magnesium. *Metall. Mat. Trans. A* 31, 2895–2906 (2000).
22. Gu, X. N., Xie, X. H., Li, N., Zheng, Y. F. & Qin, L. *In vitro* and *in vivo* studies on a Mg-Sr binary alloy system developed as a new kind of biodegradable metal. *Acta Biomater.* 8, 2360–2374 (2012).
23. Wu, Y. et al. Unique antitumor property of the Mg-Ca-Sr alloys with addition of Zn. *Sci. Rep.* 6, 21736; doi: 10.1038/srep21736 (2016).
24. Brat, H. S., Wong, J. & Manuel, M. V. Investigation of the mechanical and degradation properties of Mg–Sr and Mg–Zn–Sr alloys for use as potential biodegradable implant materials. *J. Mech. Behav. Biomater.* 7, 87–95 (2012).
25. Zhou, Y. L., Li, Y., Luo, D. M., Ding, Y. & Hodgson, P. Microstructures, mechanical and corrosion properties and biocompatibility of as extruded Mg–Mn–Zn–Nd alloys for biomedical applications. *Mater. Sci. Eng. C* 49, 93–100 (2015).
26. Song, G. L. & Atrens, A. Corrosion Mechanisms of Magnesium Alloys. *Adv. Eng. Mater.* 1, 11–33 (1999).
27. Shaw, B. A. Fundamentals, Testing, and Protection in ASM Handbook (ed. Korb, L. J., Olson, D. L.) 695 (Materials Park, 2003).
28. Lentz, M., Risse, M., Schaefer, N., Reimers, W. & Beyerlein, I. J. Strength and ductility with [1011] - [1012] double twinning in a magnesium alloy. *Nat Commun.* 7, 11068, doi: 10.1038/ncomms11068 (2016).
29. Williams, J. G. & Gampomplas, C. Using the simple compression test to determine Young’s modulus, Poisson’s ratio and the Coulomb friction coefficient. *Int. J. Solids Struct.* 45, 4448–4459 (2008).
30. Mosmann, T. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *J. Immunol. Methods.* 65, 55–63 (1983).
31. Yang, J. et al. Microstructure and mechanical properties of Mg–4.5Zn–xNd (x = 0, 1 and 2, wt%) alloys. *Mater. Sci. Eng. A* 479, 339–344 (2008).
32. Xu, D. R., Liu, L., Xu, Y. B. & Han, E. H. The influence of element Y on the mechanical properties of the as-extruded Mg–Zn–Y–Zr alloys. *J. Alloys Compd.* 426, 155–161 (2006).
33. Wu, A. R. & Xia, C. Q. Study of the microstructure and mechanical properties of Mg–rare earth alloys. *Mater. Des.* 28, 1963–1967 (2007).
34. Rohklin, L. I. *Magnesium alloys containing rare earth metals* 1st edn, (ed. Rohklin, L. L.), Ch.4, 172–176 (Taylor & Francis, 2003).
35. Slater, J. C. Atomic Radii in Crystals. *J. Chem. Phy.* 41, 3199–3204 (1964).
36. Wang, J. et al. Effect of Y for enhanced age hardening response and mechanical properties of Mg-Ho–Y–Zr alloys. *J. Alloys Compd.* 454, 194–200 (2008).
37. Zhou, X., Huang, Y., Wei, Z., Chen, Q. & Gan, F. Improvement of corrosion resistance of AZ91D magnesium alloy by holmium addition. *Corros. Sci.* 48, 4223–4233 (2006).
38. Robinson, H. A. & George, P. F. Effect of Alloying and Impurity Elements In Magnesium Alloy Cast Anodes. *Corrosion.* 18, 182–188 (1954).
39. Vente, M. A. D. et al. Neutron activation of holmium poly-(l-lactic acid) microspheres for hepatic arterial radioembolization: a validation study. *Biomed. Microdev.* 11, 763–772 (2009).
40. Mumper, R. J., Yun Ryo, U. & Jay, M. Neutron-Activated Holmium-166-Poly-(L-Lactic Acid) Microspheres: A Potential Agent for the Internal Radiation Therapy of Hepatic Tumors. *J. Nucl. Med.* 32, 2139–2143 (1991).
41. Nissen, J. F. W. et al. Holmium-166 poly lactic acid microspheres applicable for intra-arterial radionuclide therapy of hepatic malignacies: effects of preparation and neutron activation techniques. *Eur. J. Nucl. Med.* 26, 699–704 (1999).
42. Nissen, J. F. W. et al. Influence of neutron irradiation on holmium acrylatecontained loaded poly-(l-lactic acid) microspheres. *Biomaterials* 23, 1831–1839 (2002).
43. Smits, M. L. J. et al. Holmium-166 radioembolisation in patients with unresectable, chemorefractory liver metastases (HEPAR trial): a phase I, dose-escalation study. *Lancet. Oncol.* 13, 1025–1034 (2012).
44. Smits, M. L. et al. Research Holmium-166 radioembolization for the treatment of patients with liver metastases: design of the phase 1 HEPAR trial. *J. Exp. Clin. Cancer Res.* 29, 70–81 (2010).
45. Zieglihuis, S. W. et al. Long-term toxicity of holmium-loaded poly-(l-lactic acid) microspheres in rats. *Biomaterials* 28, 4591–4599 (2007).
46. Singh, A., Somekawa, H. & Mukai, T. Compressive strength and yield asymmetry in extruded Mg–Zn–Ho alloys containing quasicrystal phase. *Scripta Mater.* **56**, 935–938 (2007).
47. Yoshida, T., Itoh, K., Tamura, R. & Takeuchi, S. Plastic deformation and hardness in Mg–Zn–(Y,Ho) icosahedral quasicrystals. *Mater. Sci. Eng. A* **294–296**, 786–789 (2000).
48. Yamasaki, M., Hayashi, N., Izumi, S. & Kawamura, Y. Corrosion behavior of rapidly solidified Mg–Zn–rare earth element alloys in NaCl solution. *Corros. Sci.* **49**, 255–262 (2007).
49. Peng, Q., Dong, H., Wang, L., Wu, Y. & Wang, L. Aging behavior and mechanical properties of Mg–Gd–Ho alloys. *Mater. Charact.* **59**, 983–986 (2008).
50. Haley, T. J., Koste, L., Komesu, N., Efros, M. & Upham, H. C. Pharmacology and toxicology of dysprosium, holmium, and erbium chlorides. *Toxicol. Appl. Pharmacol.* **8**, 37–43 (1966).
51. Lambert, C. E. & Ledrich, M. L. In *Encyclopedia of Toxicology* 3rd edn, (ed. Philip, Wexler) 43–47 (Academic Press, 2014).
52. Meng, D. C., Erol, M. & Boccaccini, A. R. Processing Technologies for 3D Nanostructured Tissue Engineering Scaffolds. *Adv. Eng. Mater.* **12**, B467–B487 (2010).
53. Johnson, L., Perchy, D. & Liu, H. In vitro evaluation of the surface effects on magnesium-yttrium alloy degradation and mesenchymal stem cell adhesion. *J. Biomed. Mater. Res. A* **100A**, 477–483 (2012).
54. Kang, H. et al. Multi-functional magnesium alloys containing interstitial oxygen atoms. *Sci. Rep.* **6**, 23184, doi: 10.1038/srep23184 (2016).
55. Zhang, X. et al. Mitigation of Corrosion on Magnesium Alloy by Predesigned Surface Corrosion. *Sci. Rep.* **5**, 17399, doi: 10.1038/srep17399 (2015).
56. Meininger, S. et al. Strength reliability and in vitro degradation of three-dimensional powder printed strontium-substituted magnesium phosphate scaffolds. *Acta Biomater.* **31**, 401–411 (2016).
57. Levi, D. S., Kusnezov, N. & Carman, G. M. Smart Materials Applications for Pediatric Cardiovascular Devices. *Pediatr. Res.* **63**, 552–558 (2008).
58. Feyerabend, F. et al. Blood compatibility of magnesium and its alloys. *Acta Biomater.* **25**, 384–394 (2015).
59. Shi, Z. & Atrens, A. An innovative specimen configuration for the study of Mg corrosion. *Corros. Sci.* **53**, 226–246 (2011).
60. Zhao, M. C., Liu, M., Song, G. L. & Atrens, A. Influence of Microstructure on Corrosion of As-cast ZE41. *Adv. Eng. Mater.* **10**, 104–111 (2008).

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**Author Contributions**

Y.L. designed the experiments. Y.D. and D.Z. carried out the experimental work. J.L., C.W., D.Z. and Y.L. together with Y.D. analysed the data and discussed the results. All authors contributing to the writing of the manuscript.

**Additional Information**

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