Risk of Prostate Cancer and Cyclin D1 A870G polymorphism; a Study of Correlation

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Abstract

Cyclin D1 (CCND1) is a critical gene in regulating the progression of cell cycle from G1 to S phases. Like other cyclins, cyclin D1 is frequently dysregulated in multiple cancers. Various clinical and epidemiological studies have suggested the possible association of cyclin D1 A870G polymorphism with the development of various cancers. Hence, we investigated the role of cyclin D1 A870G polymorphism in modulating the risk of prostate cancer (CaP) in a Kashmiri population. We examined a case-control study in which 129 CaP cases were studied for cyclin D1 A870G polymorphism against 221 controls taken from the general population by employing the polymerase chain reaction-restriction fragment length polymorphism technique. We observed the cyclin D1 A allele was more frequently present in the CaP group than the control group. Furthermore, men with AA genotype have an increased risk for developing CaP as compared to the control groups. We found AA genotype statistically significantly associated with dwelling, lymph node metastases, histopathological grade, and PSA levels. Therefore, our findings suggest that A870G polymorphism is a risk factor for CaP development. Moreover, men with AA genotype have an increased risk of developing CaP.

Keywords: CaP; Cyclin D1; Genetic polymorphism

Introduction

Prostate cancer (CaP) is a complex disease caused by multiple factors, and is one of the most frequent malignant diseases among men [1]. Accumulating studies have revealed the association of various genetic elements in the development of sporadic CaP [2,3], which might be used as potential therapeutic targets for CaP therapy. Hence, studying the role of genetic variants including single nucleotide polymorphic is of great significance in understanding the susceptibility CaP.

An unbalance in the cell cycle regulation has been found to play a key role in a various type of cancers, and is often associated with the onset of metastasis, in part, by negatively modulating the cell's ability to respond appropriately to DNA damage [4,5]. Reports have suggested that a strong relation between the alleles of these genes and CaP susceptibility [6-10]. Cyclin D1, also known as CCND1, is confined to the nucleus during G1 phase, and plays a pivotal role in the progression from the G1 to S phase of cell cycle [11]. During the CaP development, the expression of cyclin D1 gene has often been seen overexpressed and is associated to poor prognosis of CaP [12-14]. These findings indicate the importance of cyclin D1 (as a cell-cycle regulator) in carcinogenesis and progression of CaP [15].

Cyclin D1 mRNA is alternatively spliced into two different transcripts, which get translated into two different proteins [16,17]. The A870G polymorphism appears to modulate the splicing at codon 242 within conserved donor site of exon 4 of the gene [16-18]. Recent studies have suggested that the variant allele encoding A is a major source of variant transcript b in several kinds of cancer cells [16-18]. Moreover, cyclin D1 genotypes have been frequently found associated to variety of cancers [16,18,19-21]. Therefore, considering the importance of cyclin D1 in the development of CaP, the present study was conducted to investigate the association between the Cyclin D1 A870G polymorphism and the susceptibility to CaP or its disease status.

Materials and Methods

Prostate cancer patients

A cohort of 129 CaP tissue samples were collected, with the histopathological diagnosis of the CaP. All the samples included in this study consisted of tumor and adjacent normal tissues. Only the tissue samples confirmed by histopathological studies to be cancerous were included in the study. The patient participation was obtained through informed consent and after approval from the Ethics Committee of Sher-i-Kashmir Institute of Medical Sciences.

The patients underwent histopathological diagnosis in the Department of Histopathology of our institution. Fifty prostate tumor samples and 45 benign hyperplasia (BHP) samples were collected. Samples from 80 healthy males over 50 years of age served as the controls. We also obtained prostate sextant biopsy specimens from 10 patients with elevated levels of serum PSA. Prostate cancer tissue samples consisting of tumor tissues and adjacent normal tissue were collected. Only histo-pathologically confirmed tumors were included in the study. The study was approved by the Ethical Committee of the Sher-i-Kashmir Institute of Medical Sciences.
controls

Patients attending the Department of general medicine at Sher-I-Kashmir Institute of Medical Sciences (SKIMS) for general checkup were screened. A total of 260 patients visited the SKIMS, out of 260 patients only 221 agreed to take part in the present study, henceforth written informed consent was obtained from all patients for their participation.

A870G gene polymorphism

The A870G polymorphism lies within the conserved splice donor site of exon 4 of cyclin D1 gene. This SNP was detected by restriction fragment length polymorphism of PCR-amplified fragments. PCR reactions were carried out in a final volume of 25 μL containing 50 ng genomic DNA template, 1X PCR buffer (Biotools) with 2 mM MgCl₂, 0.4 μM of each primer (Genescript), 50 μM dNTPs (Biotools), and 0.5 U DNA polymerase (Biotools). For PCR amplification, the standard program was used as follows: After an initial denaturation step at 94°C for 7 min, followed by 35 denaturation cycles of 30s at 94°C, 30s of annealing at 54°C, and 30s of extension at 72°C, followed by a final elongation cycle at 72°C for 5 min. The amplified product was digested overnight with the ScrFI enzyme at 37°C and electrophoresed on a 3.0% agarose gel. Restriction fragments obtained were 146 and 22 bp for the G homozygote, and the unrestricted fragment was 167 bp for the A homozygote, while as the heterozygote displayed both the two bands.

Statistical analysis

Observed frequencies of genotypes in CaP patients were compared to controls using chi-square or Fisher exact tests when expected frequencies were small. The chi-square test was used to verify whether genotype distributions were in Hardy-Weinberg equilibrium. Statistical significance was set at P<0.05. Statistical analyses were performed with PASW version 18 Software.

Results

We determined the A870G polymorphism in 129 CaP patients; 22.4% (29 of 129) of the patients were homozygous for G/G variant, 27.9% (36 of 129) were heterozygous for A/G variant, 49.61% (64 of 129) were homozygous for A/A variant. Whereas the pattern of A870G polymorphism in 221 healthy controls is 11.31% (25 of 221) of the patients were homozygous for A/A variant. Whereas the pattern of A870G gene polymorphism in 221 healthy controls is 11.31% (25 of 221) of the patients were homozygous for A/A variant. Whereas the pattern of A870G polymorphism in 221 healthy controls is 11.31% (25 of 221) of the patients were homozygous for A/A variant. Whereas the pattern of A870G polymorphism in 221 healthy controls is 11.31% (25 of 221).

The present study consisted of 129 CaP patients, out of which 116 patients were above or equal to 65 years of age and 13 were of 50 years of age. Most of our patients (72) had grade I & II tumor, and 57 had grade III & IV tumor status. 104 CaP patients had widely or moderately differentiated histo-pathological grade and 25 had poorly differentiated histo-pathological grade. Men aged 65 years or over formed the greater part of the CaP cases. Table 1 show clinicopathological characteristics related to the GG, AG and AA variants. A larger number of patients included in our study were having tumor stage I and II, whereas the patients having tumor stage III and IV were relatively small.

In the cyclin D1 A870G polymorphism, the AA genotype was statistically significantly associated with the dwelling, PSA levels, tumor stage, and histopathological grade (Table 1). The genotype was not statistically significantly associated with the mean age at the time of diagnosis.

| Variables          | Cases (n=129) | GG   | A/G  | AA   | P Value ; χ² |
|--------------------|--------------|------|------|------|--------------|
| Age                | N=129 (%)    |      |      |      |              |
| >50                | 13 (10.0%)   | 03 (23.07%) | 04 (30.7%) | 06 (46.15%) | 0.56; 1.12   |
| ≤ 65               | 116 (90.0%)  | 25 (15.55%) | 32 (27.5%)  | 59 (50.86%) |              |
| Dwelling           |              |      |      |      |              |
| Rural              | 45 (34.6%)   | 08 (17.77%) | 12 (26.6%)  | 25 (55.5%)  | 0.52; 1.21   |
| Urban              | 84 (65.4%)   | 23 (27.38%) | 28 (33.33%) | 33 (39.28%) |              |
| Pesticide Exposure |              |      |      |      |              |
| Low                | 44 (34.6%)   | 05 (11.3%)  | 09 (20.4%)  | 30 (68.1%)  | 0.06; 4.98   |
| High               | 85 (65.4%)   | 20 (23.5%)  | 29 (34.1%)  | 36(42.3%)   |              |
| PSA Levels         |              |      |      |      |              |
| Low (4-8)          | 35 (27.7%)   | 05 (14.2%)  | 06 (17.1%)  | 24 (68.5%)  | 0.04; 6.02   |
| High (8-13)        | 94 (72.3%)   | 23 (24.4%)  | 32 (34.0%)  | 39 (41.4%)  |              |
| Tumor Stage        |              |      |      |      |              |
| I +II (a+b)        | 72 (55.4%)   | 19 (26.3%)  | 20 (27.7%)  | 33(45.8%)   | 0.05; 4.96   |
| III (a+b)+ IV      | 57 (44.6%)   | 09 (16.7%)  | 19 (33.3%)  | 29 (50.8%)  |              |
| Histopathological Grade |      |      |      |      |              |
| PD                 |              |      |      |      |              |
| MD+WD              | 25 (96.0%)   | 07 (28.0%)  | 10 (40.0%)  | 08 (32.0%)  | 0.04; 5.32   |
| 57+47              | 21 (62.1%)   | 07 (28.0%)  | 10 (40.0%)  | 08 (32.0%)  |              |

Table 1: Association between Cyclin D1 A870G and clinicopathologic characteristics.

In this study, we observed genotype frequencies in cases and controls were in Hardy-Weinberg equilibrium. The genotype frequencies of Cyclin D1 A870G in cases and controls were observed and it was found that AA genotype is significantly associated with the CaP cases (p value=0.001) (Table 2).
A significant correlation was found between the AA variant and poorly differentiated histopathological grade as has been reported by various earlier studies, suggesting that cyclin D1 A870G SNP may be involved in different mechanisms and possibly also various stages of tumor development.

We also found that the genotype frequencies of A870G in cases and controls, where AA genotype is significantly associated with the CaP cases (p value=0.001), suggesting that the CaP patients carrying a AA allele are at much higher risk for CaP.

Conclusion

In conclusion, our results support the idea of cyclin D1 A870G genotypic variance do have a strong association with the CaP development. The AA genotype was associated with increased of CaP in Kashmiri men as compare to GG genotype.

Conflict of Interest

The contributing Authors have no financial or any non-financial competing interests.

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