Nontoxic concentration of DNA-PK inhibitor NU7441 radio-sensitizes lung tumor cells with little effect on double strand break repair

Shigeaki Sunada,1,2 Hideki Kanai,1,2 Younghyun Lee,2 Takeshi Yasuda,3 Hirokazu Hirakawa,2 Cuihua Liu,2 Akira Fujimori,2 Mitsuru Uesaka1 and Ryuichi Okayasu2

1Department of Nuclear Engineering and Management, School of Engineering, University of Tokyo, Tokyo; 2Research Center for Radiation Protection, National Institute of Radiological Sciences, Chiba; 3Research Center for Radiation Emergency Medicine, National Institute of Radiological Sciences, Chiba, Japan

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Correspondence
Ryuichi Okayasu, Research Center for Radiation Protection, National Institute of Radiological Sciences, 4-9-1 Anagawa, Inage-ku, Chiba 263-8555, Japan. Tel: +81-43-362-3711; Fax: +81-43-255-0720; E-mail: rokayasu@nirs.go.jp

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Heavy-ion radiation therapy has been increasingly utilized due to its lesser side effects compared to conventional photon radiation therapy.1,2 Tumor tissue can be well targeted by physically adjusting the Bragg peak on the tumor area, and a high-linear energy transfer (LET) beam can induce more effective tumor cell killing than low-LET beams.3–5 Another strategy to improve the outcome of radiotherapy would be to use a radio-sensitizer, which enhances radiation-induced tumor cell killing by targeting DNA damage response.6–9 Although many studies are available on low-LET radiation combined with sensitizers, further biological studies are necessary in the combined regimen of radio-sensitizer and heavy ions for future clinical applications.

There are two main pathways for DNA double-strand break (DSB) repair: homologous recombination (HR) and nonhomologous end joining (NHEJ). NHEJ is a dominant pathway in DSB repair induced by low-LET radiation, such as X-rays and γ-rays,10 while NHEJ has been shown to be suppressed to some extent and the HR pathway utilized more in DSB induced by high-LET radiation.11–13

High-linear energy transfer (LET) heavy ions have been increasingly employed as a useful alternative to conventional photon radiotherapy. As recent studies suggested that high LET radiation mainly affects the nonhomologous end-joining (NHEJ) pathway of DNA double strand break (DSB) repair, we further investigated this concept by evaluating the combined effect of an NHEJ inhibitor (NU7441) at a non-toxic concentration and carbon ions. NU7441-treated non-small cell lung cancer (NSCLC) A549 and H1299 cells were irradiated with X-rays and carbon ions (290 MeV/n, 50 keV/μm). Cell survival was measured by clonogenic assay. DNA DSB repair, cell cycle distribution, DNA fragmentation and cellular senescence induction were studied using a flow cytometer. Senescence-associated protein p21 was detected by western blotting. In the present study, 0.3 μM of NU7441, nontoxic to both normal and tumor cells, caused a significant radio-sensitization in tumor cells exposed to X-rays and carbon ions. This concentration did not seem to cause inhibition of DNA DSB repair but induced a significant G2/M arrest, which was particularly emphasized in p53-null H1299 cells treated with NU7441 and carbon ions. In addition, the combined treatment induced more DNA fragmentation and a higher degree of senescence in H1299 cells than in A549 cells, indicating that DNA-PK inhibitor contributes to various modes of cell death in a p53-dependent manner. In summary, NSCLC cells irradiated with carbon ions were radio-sensitized by a low concentration of DNA-PK inhibitor NU7441 through a strong G2/M cell cycle arrest. Our findings may contribute to further effective radiotherapy using heavy ions.

Furthermore, Takahashi et al.14 reported that when using HR and NHEJ pathway-deficient mouse embryonic fibroblasts, high-LET radiation caused significant radio-sensitization even in NHEJ-defective cells. Their study suggests that the NHEJ pathway is still dominant in DSB repair induced by high-LET radiation. Therefore, it is important to study how cells with reduced or inhibited NHEJ respond to high-LET heavy-ion radiation. DNA-PKcs is one of the key proteins in the NHEJ pathway, and many studies report radio-sensitization as a consequence of DNA-PK inhibition.15–18 In particular, NU7441 has often been studied as an effective specific DNA-PK inhibitor; this specificity was confirmed by the use of DNA-PKcs defective cells, which did not show any radio-sensitization with NU7441 treatment.9

In this study, we investigate the combined effect of NU7441 and carbon ion irradiation in human non-small cell lung cancer (NSCLC) cells. We find that the nontoxic concentration of NU7441 causes significant radio-sensitization with X-rays and carbon ions in NSCLC cells, with little effect on DSB repair; strong G2/M arrest is found to be associated with this sensitization.
Materials and Methods

Cell culture. Non-small cell lung cancer cell lines A549 and H1299 cells were obtained from Riken BRC (Tsukuba, Japan) and the American Type Culture Collection (Manassas, VA, USA), respectively. A549 and H1299 were grown in MEMx and RPMI-1640, respectively, and supplemented with 10% (v/v) FBS. Normal human fibroblast cell line HFL1 cells were obtained from Riken BRC and were grown in MEMx supplemented with 15% FBS. All cells were maintained in a 37°C incubator with 5% CO2. Exponentially growing cells were used in the present study.

Irradiation and drug treatment. Cells were irradiated with 290 MeV/n carbon ions (LET: 50 keV/m) at the Heavy Ion Medical Accelerator in Chiba (HIMAC). The dose rate for carbon ions was 1 Gy/min. X-ray irradiation was performed using a TITAN-320 (200 kV, 20 mA, Shimadzu, Kyoto, Japan) at a dose rate of 1 Gy/min. NU7441, a specific DNA-PK inhibitor, was purchased from Cayman Chemical (Ann Arbor, MI, USA). NU7441 was dissolved in DMSO and stored at −80°C. NU7441 was reconstituted in DMSO and was used to adjust the final concentration. NU7441 was added to the cell culture medium 1 h before irradiation, and the drug was kept throughout the experiment.

Clonogenic survival assay. Cells were seeded at concentrations that may yield approximately 50–100 colonies per dish after 10–12 days. The colonies were fixed with 100% ethanol and stained with 0.1% crystal violet solution. The colonies containing more than 50 cells were scored.

Flow cytometry. γ-H2AX (a DSB marker), cell cycle distribution and senescence-associated beta-galactosidase (SA-βGal) signals were analyzed using a FACS Calibur (BD Biosciences, Franklin Lakes, NJ, USA) flow cytometer. For detection of DSB, the γ-H2AX signal was measured according to protocol described elsewhere. Cells were trypsinized, washed with cold PBS twice and stained with γ-H2AX antibody FITC conjugate (0.6 μg/mL; Merck Millipore, Billerica, MA, USA) on ice overnight. The cells were then suspended and analyzed. For cell cycle analysis, cells were trypsinized, washed with cold PBS, fixed in 70% ethanol and stored at −20°C. Cells were stained with propidium iodide (PI) at 50 μg/mL in the presence of RNase (1 mg/mL) for 30 min on ice and then analyzed.

Results

Non-toxic concentration of NU7441 induces radio-sensitization in non-small cell lung cancer cells irradiated with low-LET and high-LET radiation. Cellular toxicity of NU7441 was evaluated in normal HFL1 cells and NSCLC cells using a clonogenic survival assay. HFL1 cells showed a concentration-dependent decrease in plating efficiency, and this was more distinct for the concentrations higher than 1 μM (Fig. 1a). NSCLC cells showed a clear drop in plating efficiency at 3 μM (Fig. 1b). These results indicate that 0.3 μM of NU7441 is nontoxic in both normal and cancer cells. A clonogenic survival assay was performed with this nontoxic concentration of NU7441. Significant radio-sensitization was confirmed in NU7441-treated NSCLC cells, not only with X-rays but also with carbon ions (Fig. 1c,d). The relative biological effectiveness (RBE) of carbon ions (50 keV/μm) compared to X-rays and the
sensitization enhancement ratio (SER) of NU7441 (0.3 μM) were calculated based on the $D_{10}$ values. Carbon ions showed an RBE of 2.15 in A549 cells and 1.37 in H1299 cells. NU7441 gave an SER of 1.77 (X-rays) and 1.55 (carbon ions) in A549 cells and 1.94 (X-ray) and 1.58 (carbon ions) in H1299 cells. Judging from these analyses, carbon ions caused more effective cell killing in A549 than H1299 cells, while NU7441 induced higher radio-sensitization with X-rays than carbon ions in H1299 cells, and the degree of sensitization was smaller for A549 cells.

**Low concentration of NU7441 does not seem to show double strand break-repair inhibition in irradiated cells.** Double strand break repair after irradiation was evaluated by analyzing γ-H2AX signals. Three micromolar of NU7441 showed significantly increased persistent γ-H2AX signals compared to the control, even in carbon-irradiated cells showing clear DSB repair inhibition. In contrast, 0.3 μM of NU7441 did not seem to show a significant shift from the control in γ-H2AX signals, except for a very slight increase in the 24-h sample in some cases (Fig. 2a-d). Our results suggest that the low concentration of NU7441 (0.3 μM) did not induce obvious DSB repair inhibition in spite of its marked radio-sensitization effect at the cell survival level (Fig. 1c,d).

**Low concentration of NU7441 causes significant G2/M arrest in irradiated H1299 cells.** To study the mechanism of radio-sensitization at the low concentration (0.3 μM) of NU7441, we investigated the cell cycle distribution in tumor cells 24 h after the combined treatment. As shown in Figure 3, both NU7441 and X-rays alone did not seem to cause a significant change in cell cycle distribution compared to the control. However, G2/M arrest was clearly observed in the combined treatment of NU7441 and radiation. The most intense G2/M arrest was induced by the combination of NU7441 and carbon ions in both cell lines. Interestingly, with carbon irradiation alone, A549 cells showed more G2/M arrest than H1299 cells, but after NU7441 and carbon irradiation, G2/M arrest was more distinct in H1299 cells than A549 cells.

**Irradiated H1299 cells were more effectively killed by NU7441.** To investigate the mode of cell death, DNA fragmentation and cellular senescence were studied. For DNA fragmentation analysis, the sub-G1 population in NSCLC cells was evaluated for 3 days. A very small portion of X-irradiated A549 cells became sub-G1 phase even with NU7441, while carbon irradiation increased the sub-G1 population regardless of NU7441 presence 3 days after treatment (Fig. 4a). We were not able to confirm a difference between A549 and H1299 cells 1 day after irradiation; however, H1299 cells treated with NU7441 (0.3 μM) and exposed to X-rays and carbon ions showed a remarkable increase in the sub-G1 population after day 2 (Fig. 4b). For cellular senescence analysis, an SA-βGal assay in NSCLC cells was performed 3 days after irradiation. SA-βGal-positive cells increased significantly in carbon-irradiated A549 cells and in H1299 cells treated with NU7441 and carbon irradiation (Fig. 5a,b). Because the p53-p21 signal pathway is an important senescence pathway, and the p53 status in A549 and H1299 cells are wild type and null, respectively, p21 protein expression was measured in these cells. As shown in Figure 5(c), p21 expression in A549 cells was consistent with the result of the SA-βGal assay; in contrast, no activation was observed in H1299 cells. These results indicate that cellular senescence was observed in p53-null H1299 cells, suggesting that this process does not depend on p53 status.

**Discussion**

In this report, we found that a nontoxic concentration of DNA-PK inhibitor, NU7441, caused significant radio-sensitization both in X-irradiated and carbon-irradiated NSCLC cells; for X-rays, SER are 1.77 and 1.94, and for carbon ions SER are 1.55 and 1.58 depending on cell lines (Fig. 1). In previous reports,(9,21–24) much higher concentrations of this drug were used to induce radio-sensitization. The drug concentration of 0.3 μM used in this study, 1/3 to 1/30 the concentrations used in the abovementioned publications, did not affect cell viability. Therefore, 0.3 μM of NU7441 is a useful strategy for radio-sensitization. Our strategy is particularly beneficial for particle radiation therapy including heavy ions as its irradiation beams mostly hit the tumor tissue with little sensitizing effect on surrounding normal tissues. The sensitization ratio is lower in cells irradiated with carbon ions than X-rays. Our results may not be in conflict with a partial diminution of the role for NHEJ in high-LET irradiated cells.(14,25) Our results also indicate that p53-deficient cells, which may be resistant to carbon ions, might become a good candidate for the combination regimen of NU7441 and heavy ions.
Inhibition of DNA-PK, a key protein for NHEJ pathway, is known to impair DSB repair.(9,21) The nontoxic concentration (0.3 μM) of NU7441 did not seem to cause DNA repair inhibition in irradiated tumor cells, whereas 3 μM of NU7441, toxic to normal cells, clearly showed DSB repair inhibition (Fig. 2). We can suggest that a low and nontoxic concentration of DNA-PK inhibitor might cause radio-sensitization via a pathway other than inhibition of DSB repair. Our results somewhat reflect a previous study indicating that DNA-PKcs knockdown cells induced radio-sensitization, while there was no significant difference in DSB repair kinetics.(24) These authors also reported that smaller amounts of DNA-PKcs protein were accumulated at DSB sites in DNS-PKcs knockdown cells and that this amount might be sufficient for DSB repair function. The detailed mechanism needs to be further examined, but DNA-PK inhibition might have another role leading to an increase in cell death.

One of the remarkable effects that the low concentration of NU7441 caused was the marked G2/M cell cycle arrest following irradiation. It is known that DNA-PK inhibition increases the percentage of cells in the G2/M phase with low-LET radiation.(26) There is a possibility that DNA-PK inhibition partially regulates the G2/M checkpoint pathway without affecting NHEJ. G2/M delay was more clearly manifested in p53-null H1299 cells. G1 arrest is not normally observed in p53-deficient cells(27) and, thus, we think that with NU7441 treatment, more cells can be accumulated at G2/M in irradiated H1299 cells. In particular, this phenomenon was noted in...
carbon-irradiated cells. These results suggest high-LET heavy-ion-irradiated cells with a low concentration of NU7441 might be arrested in the G2/M phase regardless of NHEJ suppression.

We also evaluated DNA fragmentation and cellular senescence. Irradiated H1299 cells with NU7441 showed a dramatic increase in the sub-G1 population, especially 2 days after irradiation and beyond (Fig. 4). p53-deficient H1299 cells induced a higher number of sub-G1 cells than p53 wild-type A549 cells in our case. Interestingly, these tendencies are similar to those of G2/M arrest and subsequent DNA fragmentation. We also believe that the marked G2/M arrest induced by DNA-PK inhibition might cause DNA fragmentation. In addition, we found that NU7441 also promoted cellular senescence in irradiated H1299 cells (Fig. 5). Cellular senescence is a form of cellular growth control and this has been suggested as a strategy to deter tumor cell proliferation. The p53-p21 pathway is the most essential senescence pathway, and we showed no p21 expression in H1299 cells. Several studies also indicated p53 independent senescence. Thus, we also investigated p16 levels in H1299 cells as an important senescence pathway; however, the levels had no correlation with senescence induction (data not shown). Therefore, the enhanced senescence in H1299 cells via DNA-PK inhibition might be caused by telomere dysfunction and mitosis skipping. In this senescence analysis, H1299 cells showed resistance to carbon ions compared to A549 cells (Fig. 5); however, NU7441 treatment caused more effective cell deaths in carbon-irradiated H1299 cells. These results are consistent with cell survival, and indicate that p53 status would be an important factor when treatment options are considered.

In summary, we found that a low concentration of DNA-PK inhibitor NU7441, nontoxic to normal cells, caused a significant radio-sensitization in irradiated tumor cells, independent of their p53 status. This is probably not from DSB repair inhibition but rather a consequence of strong G2/M arrest. A more detailed mechanism of this sensitization may be determined in the future. NU7441 caused effective radio-sensitization in irradiated H1299 cells, which showed a slight radio-resistant phenotype. These findings would provide a basis for future clinical application of a DNA-PK inhibitor in radiation therapy including heavy ion treatment.

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Disclosure Statement

The authors have no conflict of interest to declare.

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