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Doxycline and Eradication of Microfilaria in Patients with Loiasis

To the Editor: Wolbachia are intracellular symbionts found in 20% of insects and in several nematodes, including filarial worms. Because tetracycline eradicates Wolbachia in nematodes, this drug has been proposed for chemotherapy in filariasis (1). We report two patients with loiasis in whom no Wolbachial DNA was detected by PCR in Brugian and Bancroftian microfilariae by polymerase chain reaction (PCR), and for whom 6 weeks of doxycycline failed to eradicate the microfilaria. We conclude that doxycycline may not be an efficient therapy for loiasis.

Filariae are responsible for 150 million infections worldwide, some of them devastating diseases such as elephantiasis (caused by Bancroftian and Brugian filariasis) and blindness (caused by onchocerciasis). There is no satisfactory treatment for filariasis: although diethycarbamazine citrate has been used for 50 years to treat the disease, this drug is not efficient in most adults (2). Ivermectin has been found to be efficient in treating microfilaria and possibly for viability and fertility of adult Onchocerca volvulus worms if therapy is prolonged for 2 to 3 years (2). However, the microfilarialic effect of ivermectin in Wuchereria, Brugia, and Loa loa is similar to diethycarbamazine: ultrasonography shows that it has no effect on adult Bancroftian worms, and microfilariae often reappear after a few months (2).

Intracellular bacteria have been observed in the lateral cords of adult female worms as well as in microfilariae of W. bancrofti, and these bacteria have recently been identified as belonging to the Wolbachia genogroup (3). Wolbachia have been detected by PCR in Brugian and Bancroftian filariae, dirofilariae, and most species of Onchocerca including O. volvulus, but have never been detected in L. loa worms (4). Wolbachia are causative agents of a variety of modifications in host development and reproduction, including cytoplasmic incompatibility and pathogenesis. Consequently, it has been proposed that antibiotic eradication of Wolbachia from infected filarial worms would reduce microfilariaemia. This has been demonstrated in an animal model with Litomosoides sigmodontis and recently confirmed in patients infected with O. volvulus (1-5). In experimental L. sigmodontis infection, after 41 days of therapy, microfilariaemia in tetracycline-treated animals was one-tenth that in normally infected animals (5).

We report the failure of tetracycline to reduce microfilaria in two patients with L. loa filariasis. Patient no. 1 was a 58-year-old man who worked in Gabon for many years and had cutaneous larva migrants. L. loa microfilariaemia was detected (4x10^9/mL). Patient no. 2, a 15-year-old boy living in Cameroon, was diagnosed with Calabar swelling with L. loa microfilariaemia (1x10^9/mL). After giving informed consent, both patients were treated with doxycycline 200 mg daily for 6 weeks as previously described in O. volvulus-infected patients (1). We observed patients for microfilariaemia every week for 6 weeks and then every 2 weeks for 2 months. The presence of adult worms was detected by physical examinations.

Microfilariaemia was detected in both patients at the completion of treatment and at day 120 of follow-up. In patient no. 1, the frequency of migrating adult worms seemed to diminish during therapy, but they never disappeared. For Wolbachia detection in worms, blood samples were collected both in Dupont-Isolator and EDTA-containing tubes. After centrifugation at 5000 x g for 30 min, the worm-enriched pellet was resuspended in 1 mL of sterile deionized water for erythrocyte lysis. DNA was extracted from the suspension by using the QIAmp-blood kit (Qiagen, Hilden, Germany) following manufacturer’s recommendations. W. pipientis DNA was used as a positive control. Control of DNA extraction was performed by amplifying microfilarial DNA using the nematode-specific 18S rDNA-derived primers 18SF (5'-GAT-ACC-GGC-CCA-TGA-TTG-ACC-3') and 18SR (5'-ACC-AAC-TAA-GAA-CGG-CCA-TG-3'). Wolbachia detection was attempted with the FD1 (5'-AGA-GTT-TGA-TCC-TGG-CTC-AG-3') and Rp2 (5'-ACG-GTC-ACC-TGG-CAA-GGA-3') eubacterial primers, with the Ehrlichia genus-specific 16S rDNA primers EHR16SD (5'-GGT-ACC-YAC-AGA-AGT-CC-3') and EHR16SR (5'-TAG-CAC-TCA-TGG-TTT-ACA-GC-3'), and with primers specific for the 16S rDNA of B. malayi endosymbiont, Bsymbf (5'-ACG-TAT-ATA-ACT-3'), and Bsymbr (5'-CCT-TGG-AAT-AGA-AGT-AAT-3') (3-6). PCR reactions were performed on PTC-200 thermocycler (MJ-Research, USA) by using 45 cycles of denaturation at 94°C for 30 sec, hybridization for 45 sec, and elongation at 72°C for 1 min. Hybridization temperatures were 55°C for FD1/Rp2, 53°C for EHR16SD/EHR16SR, 42°C for Bsymbf/Bsymbr, and 57°C for 18SPE16S. Experiments were repeated three times.

We detected Wolbachia in the positive control and 18S rRNA of the nematode in the sample, but no signal compatible with Wolbachial DNA was obtained with the sets of primers used. In fact, four species of filariae (Dipetalonema setariosum, Acanthochelionema vitae, O. flexuosa, and L. loa) tested for intracellular bacteria by electron microscopy, immunohistochemistry, or PCR had no bacteria (4). The absence of Wolbachia in L. loa microfilariae may explain the failure of tetracycline therapy in our patients.

More work is needed to determine the prevalence of Wolbachia in filariae, their impact on fertility in each species, and the use of antibacterial agents for eradicating these pathogens.

Acknowledgment
We thank MJ Taylor for providing W. pipientis DNA for controls.
To the Editor: The article by Brown et al. (1) contains the statement “it appears likely that changes in the rendering process that had taken place around 1980 allowed the etiologic agent in infected carcasses to survive.” If that is the case, why not revert to the rendering methods used before 1980? That measure would seem more cost-effective than trade embargoes and mass killing of cattle. Meal made from meat and bone was used as a livestock feed additive in many countries without the apparent disastrous effect seen in the United Kingdom. Did the rendering methods remain unchanged in these other countries? Historically, rendering was viewed somewhat differently in continental Europe; the primary purpose was not to make animal feed but to destroy infectious agents, as indicated by the very name of the facility: destruction plant (Destruktionsanstalt). The plants were under governmental inspection and there were mandatory time-temperature requirements for processing meat, bones, and other offal. Temperature requirements varied from 120°C to 140°C, which presumably would reduce if not eliminate prions. It would be of interest to see a description and objective analysis of rendering methods and regulations in various countries before and after 1980. In these times when the concept of hazard analysis and critical control points (HACCP) is gaining popularity, it would seem natural to extend the principle of process control to rendering. One might easily get the impression that policies to control bovine spongiform encephalitis are dominated by some of the competitors’ beef kept off the market.

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Mad Cow Disease

To the Editor: In regards to “Bovine Spongiform Encephalopathy and Variant Creutzfeldt-Jakob Disease: Background, Evolution, and Current Concerns” (1), the use of the name “variant Creutzfeldt-Jacob disease” (vCJD) for human cases of bovine spongiform encephalopathy (BSE) is regrettable. The disease that occurs in humans exposed to BSE is zoonotic and CJD is not; in addition, the human form of BSE has important clinical, pathologic, and epidemiologic differences from CJD. Continued use of this terminology perpetuates the error.

The fact that 12 years after the feed ban bovine cases continue to occur in the United Kingdom at a much higher rate than in any other country could have two possible causes: inefficient controls or additional routes of transmission. Data on alternative routes of transmission must be evaluated, and important gaps in our understanding of BSE in cattle must be addressed.

Extensive epidemiologic data on BSE in the United Kingdom seemed to clearly implicate the practice of feeding cattle bovine offal as the primary, if not the sole, cause of the spread of BSE. Alternative theories for the origin and spread of BSE, e.g., use of insecticides on bovines or the practice of artificial insemination, appear to have been ruled out quickly on the basis of early epidemiologic data. Confidence in the reliability of these data seems to have been so great as to unduly delay transmission experiments to assess the role of alternative pathways (e.g., artificial insemination) in the propagation of BSE. Is prion protein present in semen or is it not? What if it were present in semen? Would this route lead to shorter incubation than the alimentary route? There seems to be no experimental information on the effect of freezing mutated prion protein to -196°C and placing it, after thawing, directly on the stimulated uterine mucosa.

Although the United Kingdom has acknowledged that compliance with the feed ban improved after 1996, we should not be too eager to accept lack of compliance as the only possible reason for the persistence of BSE. Fifteen years after the BSE epidemic began, it cannot be disputed that the ban on offal feed has interrupted spread of the disease. However, if continued spread by the alimentary route can be excluded as the cause of more recent cases, each of these cases should be carefully evaluated to uncover heretofore unknown or underappreciated routes of transmission.

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