Impact of endophyte inoculation on the morphological identity of cultivars of *Lolium perenne* (L) and *Festuca arundinacea* (Schreb.)

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Grass endophytes have been shown to confer enhanced environmental resilience to symbiont cultivars with reports of modified growth. If inoculating with an endophyte (E+) made an accession morphologically distinct from its registered endophyte free (E−) accession, there could be protection and ownership issues for testing authorities and breeders. This study investigated if, in official Plant Breeders Rights (PBR) field trials, the morphological characteristics of E+ and E− accessions of perennial ryegrass and tall fescue cultivars were sufficiently modified to designate them as mutually distinct and also distinct from their definitive accessions (Def), held by the testing authorities. Testing perennial ryegrass on 17 characters at 2 sites generated 48,960 observations and for tall fescue on 9 characters at 1 site, 12,960 observations (each for 3 accessions of 4 cultivars × 60 plants × 2 growing cycles). Distinctness required a p < 0.01 difference in a single character from the combined over years analysis (COYD). A few significant differences were recorded between E− and E+ accessions. Cultivar Carn E+ was smaller than Carn E− for Inflorescence Length (p < 0.01) in both years but COYD analysis (p < 0.05) was insufficient to declare distinctiveness. Overall, the number of observed differences between E−/E+ accessions was less or similar to the number expected purely by chance. In contrast, comparisons between Def and E− or E+ accessions showed a number of significant differences that were substantially more numerous than expected by chance. These results showed no conclusive evidence of endophyte inclusion creating false PBR distinctions but unexpectedly, several E− and E+ accessions were distinguished from their official definitive stock.

Vertically transmitting endophytes (*Epichloë* spp.) are non-sporulating asexually reproducing fungi of the *Clavicipitaceae* family, with no known soil borne resting spores. They are indigenous in many soils and naturally occurring in many of the cool-season grass species of the Pooidae subfamily. Endophytes are present in approximately 20% of wild populations of European ryegrass and tall fescue and at higher levels in meadow fescue. The relatively low levels of endophyte occurrence is probably because the fungus–grass relationship is not a pure symbiotic one since the endophyte removes photosynthate resources from the grass and in times of shortage or stress can even deny the plant its requirements in a parasitic fashion. However, evidence from New Zealand shows that selective increases in endophyte prevalence occurs in pastures where insect attack is a problem, but largely for those genotypes that confer an advantage to the host grass through the production of an insect toxin.

Following the 2014 taxonomic revision of the genus *Epichloë*, the two species of endophyte that infect ryegrass are *E. festucae* var. *lolii* in perennial ryegrass (*Lolium perenne* L.) and *E. occultans* in Italian ryegrass (*Lolium multiflorum* Lam.). A single species (*E. coenophialum*), infects tall fescue (*Festuca arundinacea* Schreb.), each as an intercellular fungus. There are four major groups of secondary metabolites that these fungi can produce: ergot alkaloids such as ergovaline and chanoclavine; indole diterpenes including lolitrem B and epoxy-janthitrem; pyrrolizidines which include lolines; the pyrrolopyrazine metabolite peramine. Most strains of *E. festucae* var. *lolii* produce the alkaloid Lolitrem B, which is a neurotoxin involved in the neuromuscular disorder ‘Ryegrass 1Agri-Food & Biosciences Institute, AFBI Hillsborough, Northern Ireland, UK. 2GEVES, Anjouère Experimental Unit, Domaine de l’Anjouère, La Pouëze, 49370, Erde en Anjou, France. 3Bundessortenamt Prüfstelle Scharnhorst, 2, 31535, Neustadt am Rübenberge, Germany. 4DLF Trifolium, Ny Oestergade 9, 4000, Roskilde, Denmark. 5Barenbrug Holland BV, Stationsstraat 40, 6515, AB, Nijmegen, Netherlands. 6Institute of Global Food Security, Queens University Belfast, Belfast, UK. ✉e-mail: david.patterson@afbini.gov.uk
Staggers\(^11\). In addition, is the metabolite Peramine, which deets feeding of insect adults and larvae at 10 ppm in artificial diets\(^12\). Likewise, in tall fescue, strains of the single infecting species, can produce the alkaloid ergovaline that causes ‘Fescue Toxicosis’ in livestock\(^13\). This toxin production capability is not an obligatory condition and strains that produce only one compound or none also exist, albeit these non-toxic strains are very rare in ryegrass and tall fescue.

As it has not been possible to induce a reproductive cycle in vitro it has not been possible to breed new endophyte variants. So all existing strains have been ‘discovered’ by screening existing populations and clonally multiplying selected isolates (GM and gene editing variants are now also possible but is beyond the scope of this study). Grass breeders have been able to find and incorporate endophyte strains that, for example, only carry the insect toxin and so do not impair the grazing stock and also ‘double zero’ strains that produce no toxins. It is for these ‘animal safe’ strains that plant breeders have more recently claimed agronomic advantages for farmers. This is due to these double-zero strains conferring greater environmental resilience and productivity to their host grass\(^14\).

There is evidence of endophyte presence conferring increased abiotic stress tolerance to the host plant. A 50% higher growth has been reported\(^15\) for endophyte infected plants at higher N levels, as well as increased tillering, plus greater tolerance and regrowth recovery from mild to severe moisture stress. An endophyte induced amelioration of drought stress has been shown in perennial ryegrass\(^16\) and recorded differences in tiller number, tiller length and shoot mass compared to endophyte free plants. Regarding the effect of endophyte presence on cultivar competitiveness\(^17\) found that endophyte carrying turf grass cultivars differed in measures of composition, structure and nutrient cycling from endophyte free cultivars. Recent work\(^18\) concluded that endophyte infection in perennial ryegrass could significantly increase days to heading and number of seeds per head, and decrease leaf length and the number of spikes per plant. This study further indicated that there were genetic differences between the cultivars that could be attributed to the level of endophyte infection, other work\(^19\) has found endophyte effects on grass cultivars to be variable.

Although not bred, endophyte strains are considered the property of the discoverer and in some cases patents have been taken out to protect this ownership. The equivalent protection system for the grass cultivars is the Plant Breeders Rights (PBR) statutory schemes (conforming to guidelines of the International Union for the Protection of New Varieties of Plants\(^20\)) or as in some countries such as USA, similar ‘Plant Variety Protection’ schemes. This has introduced a potential for conflicting ownership and principles concerning how to describe and protect a new candidate grass cultivar that has been inoculated with an endophyte, whether patented or not. Furthermore, if the presence of endophyte can impact on a plant’s growth and morphology then it is possible that this could create a difference in the morphological plant characters used to assess the Distinctness, Uniformity and Stability (DUS) in PBR trials. The concern is that this could create an apparent distinctness between two accessions of the same cultivar, one with and the other without an endophyte inoculation. This would let an unscrupulous breeder circumvent the PBR protection on a competitor’s existing registered variety, by inoculating it with an endophyte. There is no consensus among testing authorities on how to manage this. Within the EU, submitting endophyte-carrying seed for testing is not permitted. However, New Zealand accepts such seed but requires full details of the endophyte strain and level of infection\(^21\).

The registration authorities could avoid any risk of plagiarism by requiring all seed submissions for PBR testing to be endophyte free. However, such a requirement would incur costly cleaning and seed stock management procedures, which the grass breeders’ representatives regard as unacceptable. Similarly, the breeders have concerns that they may incur seed certification problems when later seed lots containing endophyte were compared to definitive PBR stocks that would be endophyte free.

The current study compared example endophyte free, endophyte infected and official (endophyte free) ‘definitive’ seed stocks of cultivars of perennial ryegrass and tall fescue using official DUS spaced plant trials. The objective was to determine if the expression of the DUS characteristics of plants grown from endophyte infected seed lots was sufficiently modified to designate them as distinct from endophyte-free plants taken from the same seed lot of that cultivar or from its definitive stock.

**Materials and Methods**

The plant material comprised of four cultivars of diploid amenity perennial ryegrass and four cultivars of hexaploid tall fescue. These were from different breeding programmes in the EU, NZ and USA and from known European/Australasia germplasm origins, to give a wide genetic and geographic base. All were EU registered cultivars and so previously tested and proven to be distinct, uniform and stable. For commercial in confidence reasons the registered names were replaced by codenames derived from Irish mountain names. The codenames were selected to have the same first letter as the original cultivar names as follows: Perennial ryegrass: Binnian, Carn, Croob and Gullion; Tall Fescue: Benbaun, Beann, Cove and Eagle.

The endophytes are defined as ‘wild type’ as they were not selectively screened to identify and isolate individual strain toxicity profiles. Hence all the ryegrasses were infected with the same *E. festucae* var. lolii inoculant producing Lolitrem-B and Peramine and the tall fescue with *E. coenophialum* producing Ergovaline and Lolitrem.

Each cultivar was represented by two accessions, one containing endophyte (*E* +) and the other not containing endophyte (*E* −). These were produced by germinating out a large number of individual grass plants from each endophyte infected cultivar. As none of the seed lots had close to 100% of plants infected it was possible to screen for the presence or absence of endophyte and compile two sub-populations of plants from within each cultivar that were 100% endophyte infected and 100% endophyte free. This was done by agrinostic immunoblot SKU Number: Endo797-3 [http://www.agrinostics.com/shop/](http://www.agrinostics.com/shop/). As per this previously published detection method\(^22\), the presence of *Epichloë* endophytes was assessed in two tiller sections from each plant, 6 weeks after transplanting, using the immunoblot assay according to the manufacturer’s description. The presence of endophyte was tested with an *in planta* assay using microsatellite markers B10 and B11\(^23\) on up to 200 tillers per accession. Tillers where no amplifications were detected were considered endophyte-free. From these resources, sixty plants were
For the perennial ryegrasses, each E− were then established at the Examination Office (EO) test sites as part of the official the DUS trials. In total 1440 plants (8 cultivars × 60 plants/accession × maximum of 17 characters. Similarly, for tall fescue, a total of 12,960 observations were taken, comprising 1 location × 2 growing cycles × 4 varieties × 3 accessions × 60 plants/accession × maximum of 17 characters. Statistical analyses were conducted according to the DUS testing guidelines of UPOV using the specifications for allogamous grasses. This involved planting the 60 plants of each accession as individual spaced plants in the early summer and then recording on each plant the morphological characters listed in Tables 1 and 2. These recordings were conducted at the specified timings given in the CPVO protocol during the autumn of the year of sowing through to the summer of the following year. For the purpose of this paper shortened character names have been created for use in the data tables and text, as shown in Tables 1 and 2. The complete experimentation on perennial ryegrasses generated 48,960 observations, comprising 2 locations × 2 growing cycles × 4 varieties × 3 accessions × 60 plants/accession × maximum of 17 characters. Similarly, for tall fescue a total of 12,960 observations were taken, comprising 1 location × 2 growing cycles × 4 varieties × 3 accessions × 60 plants/accession × maximum of 9 characters.

The tall fescue accessions were planted at the official GEVES Examination Office (EO) site at L’Anjouère, France. For the perennial ryegrasses, each E− and E+ plant was split into two and grown on in multi-pots to provide two identical matching sets of plants. One set was planted into the official DUS trials at the Agri-Food and Biosciences Institute (AFBI), Crossnaacrevey, Northern Ireland, UK and the other at the Federal Plant Variety Office (Bundessortenamt), Prüfstelle, Scharnhorst, Germany. This planting was repeated in the following year using new plants from the E−/E+ breeders’ resource and a new set of 60 seeds of each definitive accession, to give a full two-year DUS examination as routinely conducted by the EOs.

All test accessions were integrated into the regular DUS trials at each EO and examined using the Community Plant Varieties Office Technical Protocols. This involved planting the 60 plants of each accession as individual spaced plants in the early summer and then recording on each plant the morphological characters listed in Tables 1 and 2. These recordings were conducted at the specified timings given in the CPVO protocol during the summer of the year of sowing through to the summer of the following year. For the purpose of this paper shortened character names have been created for use in the data tables and text, as shown in Tables 1 and 2.

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Statistical analyses were conducted according to the DUS testing guidelines of UPOV using the specifications for allogamous grasses. This involved the UPOV-approved statistical methods for determining the distinctness, uniformity and stability of candidate varieties, as used by all UPOV member state EOs. This is a Combined Over-Years Distinctness (COYD) analysis based on reported methods. This incorporated a modified joint regression analysis (MJRA) model, which took account of systematic annual increases or decreases in character expression across all varieties by fitting extra terms, one for each year, in the analysis of variance. Each term represented the linear regression of the observations for the year against the cultivar means over both years. This is standard practice among workers in official plant registration schemes, for example across Europe, New Zealand and Canada.

As the objective of the study was to determine if the presence and absence of endophyte can create a PBR ‘distinction’ between accessions of the same cultivar, it is important to define the magnitude of difference required to designate them as ‘distinct’, independent varieties in these three official testing schemes. This is defined as the product of the COYD analysis tool and requires a combined over-two years difference at the 1% (P < 0.01) level in at least one character (assuming that the expression of that character is within a uniform range of expression and was not greater in one year and lesser in the other year, relative to the comparison accession).

Table 1. Morphological characters examined in Perennial Ryegrass. CPVO -EU Community Plant Variety Office, Angers France – controller of PBR test schemes in EU.

| CPVO Code | Character Name (UK)                                                                 | Shortened Character Name |
|-----------|-----------------------------------------------------------------------------------|-------------------------|
| 2         | Plant: vegetative growth habit (without vernalization)                            | Veg Habit – Vern.       |
| 4         | Plant: width (after vernalization)                                                | Width + Vern.           |
| 5         | Growth of habit after vernalization                                               | Habit + Vern.           |
| 6         | Height of plant after vernalization                                               | Height + Vern.          |
| 10        | Plant: time of inflorescence emergence (after vernalization)                      | Time of Emerge          |
| 11        | Plant: natural height at inflorescence emergence                                  | Height @ Emerge         |
| 12        | Plant: growth habit at inflorescence emergence                                    | Habit @ Emerge          |
| 13        | Flag leaf: length                                                                | Flag Length             |
| 14        | Flag leaf: width                                                                  | Flag Width              |
| 15        | Flag leaf: length/width ratio                                                     | Flag L/W Ratio          |
| 16        | Plant: length of longest stem, + inflorescence                                    | Stem + Inflor. Length   |
| 17        | Plant: length of upper internode                                                  | Internode Length        |
| 18        | Inflorescence: length                                                            | Inflor. Length          |
| 19        | Inflorescence: number of spikelets                                               | Spiklet Number          |
| 20        | Inflorescence: dens Inflorescence: density                                        | Spiklet Density         |
| 21        | Inflorescence: length of outer glume on basal spikelet                           | Glume Length            |
| 22        | Inflorescence: length of basal spikelet excluding awn                             | Spiklet Length          |
Results

The mean values for each character is presented for perennial ryegrass in Table 3 (Crossnacreevy) and Table 4 (Scharnhorst) and for tall fescue at L’Anjouère in Table 5. Each EO fully complied with the CPVO examination protocols for each species however for some characters alternative recording methods are permitted. Therefore, the Crossnacreevy data (Table 3) and the Scharnhorst data (Table 4) do not always report the observations in the same units. For example in Crossnacreevy, ‘Veg Habit – Vern’ is recorded as a scored estimate of the degree angle from the vertical and ‘Habit @ Emerge’ as the plant height:width ratio at ear emergence, while in Scharnhorst these two characters are recorded as notes on a 0–9 scale. As it was important to the objective of the study to comply with the DUS test procedures at each EO, the different recording values were retained so as to accurately emulate the normal tests employed at each EO. However, while Crossnacreevy records date of ear emergence as the number of days after the 1st March and Scharnhorst records from the 1st April, these data were standardised to 1st March, to make direct comparisons of the data possible. Tall fescue ear emergence is recorded at L’Anjouère from 1st January and was also standardized to 1st March. Similarly, length units were standardised across all three EOs.

As the expression of most of the measured characters are responsive to growing conditions there are notable differences in the magnitude of the results for the same cultivar at Crossnacreevy and Scharnhorst. So when the definitive stocks of the four perennial ryegrass varieties are compared between EOs, ear emergence was approximately 4.6 days later at Crossnacreevy than at Scharnhorst (approximately day 85 and 80.4 respectively). In making greatest to least comparisons across all twelve accessions, the differences in ear emergence was 20.6 days at Crossnacreevy and 20.8 days at Scharnhorst. On average these accessions had shorter vegetative plants (9.58 cm) and were wider (1.07 mm) and longer (0.9 cm) than at Scharnhorst. The range in plant size was greatest at Crossnacreevy (13.37 cm and 4.54 cm for Width and Height) than Scharnhorst (1.07 mm and 0.9 cm). The range in plant size was greater at Crossnacreevy than Scharnhorst

| CPVO Code | Character Name (FR) | Shortened Character Name |
|-----------|---------------------|-------------------------|
| 6         | Plant: natural height in autumn without vernalization | Height – Vern. |
| 7         | Plant: tendency to form inflorescences (without vernalization) | Inflor. – Vern. |
| 8         | Plant: natural height after vernalization (4 weeks after beginning of vegetative growth) | Height + Vern. |
| 9         | Plant: time of inflorescence emergence (after vernalization) | Time of Emerge |
| 10        | Plant: growth habit at inflorescence emergence | Habit @ Emerge |
| 11        | Plant: natural height at inflorescence emergence | Height @ Emerge |
| 12        | Flag leaf: length | Flag Length |
| 13        | Flag leaf: width (same flag leaf as that used for 12) | Flag Width |
| 14        | Stem: length of longest stem including inflorescence (fully expanded) | Stem + Inflor. Length |
| 15        | Plant: length of upper internode (as for 14) | Internode Length |
| 16        | Inflorescence: length (as for 14) | Inflor. Length |

Table 2. Morphological characters examined in Tall Fescue. CPVO - EU Community Plant Variety Office, Angers France – controller of PBR test schemes in EU.
### Table 3. Two year mean recordings of morphological characteristics of accessions of perennial ryegrass cultivars recorded at AFBI Crossnacreevy. Def, definitive accessions; E−, endophyte free accessions; E+, endophyte inoculated accessions; * number of days after 1st March.

| Shortened Character Name | Units | Binnian (def) | Binnian (E−) | Binnian (E+) | Carn (def) | Carn (E−) | Carn (E+) | Croob (def) | Croob (E−) | Croob (E+) | Gullion (def) | Gullion (E−) | Gullion (E+) |
|--------------------------|-------|---------------|--------------|--------------|------------|------------|------------|------------|------------|------------|--------------|--------------|--------------|
| Veg Habit - Vern.        | degree | 45.18         | 44.88        | 44.88        | 43.38       | 44.53      | 44.92      | 40.04      | 41.44      | 41.28      | 44.00        | 46.00        | 46.14        |
| Width - Vern.            | cm     | 32.98         | 34.19        | 31.86        | 32.68       | 33.15      | 32.77      | 49.57      | 40.22      | 41.98      | 33.86        | 35.62        | 34.14        |
| Habitat - Vern.          | degree | 49.05         | 48.89        | 48.86        | 47.61       | 48.69      | 49.72      | 50.35      | 48.21      | 48.27      | 51.47        | 53.00        | 57.59        |
| Height - Vern.           | cm     | 25.98         | 25.55        | 25.17        | 23.39       | 24.77      | 25.44      | 37.85      | 31.08      | 31.49      | 29.52        | 30.83        | 32.60        |
| Time of Emerge           | *Day no | 90.2         | 91.6         | 91.2         | 89.4        | 88.0       | 88.0       | 71.0       | 75.6       | 74.1       | 87.3         | 86.9         | 86.5         |
| Height @ Emerge          | cm     | 34.38         | 34.96        | 33.70        | 31.29       | 34.66      | 33.93      | 37.38      | 34.73      | 33.33      | 40.50        | 40.21        | 41.49        |
| Habitat @ Emerge         | ratio  | 1.63          | 1.65         | 1.69         | 1.81        | 1.61       | 1.70       | 1.72       | 1.67       | 1.70       | 1.29         | 1.35         | 1.27         |
| Flag Length              | cm     | 13.80         | 14.64        | 14.30        | 14.15       | 14.53      | 14.84      | 15.02      | 14.10      | 14.04      | 15.29        | 16.34        | 15.66        |
| Flag Width               | mm     | 4.93          | 5.06         | 5.04         | 4.61        | 5.02       | 4.88       | 5.42       | 5.19       | 5.09       | 4.81         | 4.93         | 4.92         |
| Flag L/W Ratio           | ratio  | 2.93          | 3.20         | 2.89         | 3.16        | 3.02       | 3.23       | 2.85       | 2.86       | 2.82       | 3.30         | 3.40         | 3.40         |
| Stem + Inflor. Length    | cm     | 73.94         | 74.53        | 71.83        | 63.62       | 66.28      | 69.62      | 73.66      | 69.07      | 72.58      | 70.48        | 69.91        | 70.63        |
| Internode Length         | cm     | 27.32         | 28.04        | 26.40        | 23.85       | 24.54      | 24.95      | 20.20      | 20.88      | 20.77      | 23.56        | 22.36        | 21.94        |
| Inflor. Length           | cm     | 20.03         | 19.95        | 19.58        | 19.85       | 20.00      | 21.36      | 20.41      | 19.98      | 20.51      | 20.45        | 20.82        | 21.10        |
| Spiklet Number           | number | 22.79         | 23.09        | 22.09        | 21.57       | 22.16      | 24.26      | 23.48      | 24.26      | 24.28      | 24.24        | 24.33        | 24.48        |
| Spiklet Density          | ratio  | 0.88          | 0.87         | 0.85         | 0.94        | 0.91       | 0.92       | 0.87       | 0.85       | 0.86       | 0.89         | 0.87         | 0.87         |
| Glume Length             | mm     | 8.36          | 8.34         | 8.30         | 8.91        | 8.62       | 9.22       | 9.18       | 8.98       | 8.87       | 8.95         | 9.00         | 8.83         |
| Spiklet Length           | mm     | 15.69         | 15.54        | 15.46        | 17.20       | 16.47      | 17.52      | 14.50      | 14.77      | 14.18      | 16.78        | 16.80        | 17.37        |

at p < 0.05, which is insufficient to designate these accessions as ‘distinct cultivars.’ Similarly for Spiklet Length there was no significant difference in either test year but when these differences were combined over two years a p < 0.05 was achieved. It is notable that there were also significant differences in the first test year in Stem + Inflor. at p < 0.01 and Spiklet Length at p < 0.02 and in the second test year for Habitat @ Emerge at p < 0.05 and Spiklet Number at p < 0.01. However, none of these resulted in an over-years’ significance. Similarly, at Scharnhorst, Carn E− and Carn E+ were also found to differ over the two years at p < 0.05, but in this case in Width + Vern. (Table 7). In contrast the comparison between the E− and E+ accessions of the tall fescue cultivar Cove provided no significant differences either within individual years or in the combined over-years analysis, and so both versions of the cultivar were found to have remained indistinguishable and thus ‘not distinct’ in the DUS test requirement (Table 8).

Tables 9 and 10 provide the overall summaries of significances for each ryegrass pair-wise comparison (Def/ E−; Def/E+; E−/E+). This shows that only one E−/E+ comparison was DUS distinct (p < 0.01, Gullion at Scharnhorst). The other, below threshold (p < 0.05), significant differences were three of the four E−/E+
Table 5. Two year mean recordings of morphological characteristics of accessions of tall fescue varieties recorded at GEVES, L’Anjouère. Def, definitive accessions; E−, endophyte free accessions; E+, endophyte inoculated accessions; *number of days after 1st March.

| CPVO Code | Shortened Character Name | 2yr Mean Difference | T Value | Probability | Difference Year1 | Year2 |
|-----------|--------------------------|---------------------|---------|-------------|-----------------|-------|
| 2         | Veg Habit - Vern.        | 0.39                | -0.46   | 64.277      | ns              |       |
| 4         | Width + Vern.            | -0.38               | 0.24    | 81.385      | ns              | -     |
| 5         | Habit + Vern.            | 1.03                | -1.01   | 31.196      | ns              | +     |
| 6         | Height + Vern.           | 0.67                | -0.61   | 54.291      | ns              |       |
| 10        | Time of Emerge           | 0.08                | -0.07   | 94.201      | ns              | +     |
| 11        | Height @ Emerge          | -0.73               | 0.21    | 83.236      | ns              |       |
| 12        | Habit @ Emerge           | 0.09                | -0.54   | 58.686      | ns              | +     |
| 13        | Flag Length              | 0.31                | -0.56   | 57.587      | ns              |       |
| 14        | Flag Width               | -0.14               | 0.89    | 37.254      | ns              | +     |
| 15        | Flag L/W Ratio           | 0.21                | -1.24   | 21.615      | ns              | -     |
| 16        | Stem + Inflor. Length    | 3.34                | -1.71   | 8.931       | ns              | -     |
| 17        | Internode Length         | 0.41                | -0.49   | 62.178      | ns              |       |
| 18        | Inflor. Length           | 1.36                | -2.38   | 1.814       | *              | -1     |
| 19        | Spiklet Number           | 1.29                | -1.66   | 9.801       | ns              | -     |
| 20        | Spiklet Density          | 0.01                | -0.12   | 90.524      | ns              | +     |
| 21        | Glume Length             | 0.60                | -1.91   | 5.75        | ns              | -2    |
| 22        | Spiklet Length           | 1.05                | -2.00   | 4.652       | *              | -     |

Table 6. Example comparison of E− and E+ accessions of Carn perennial ryegrass by COYD analysis of two growing cycles at Crossnacreevy, N. Ireland. Values positive if Carn E+ larger than Carn E−. Values positive if Carn E− larger than Carn E+. Values positive if Carn E− larger than Carn E−. Differences at p = 0.05 (Table 11). In stark contrast, the vast majority of comparisons between the definitive stocks and either their E+ or E− version produced significant differences. At Crossnacreevy, Carn, Croob and Gullion were all significantly (p < 0.01) different and so above the DUS pass threshold, with only the Binnian E+ and E− accessions remaining indistinguishable from their definitive stocks. This was partially replicated at Scharnhorst as Carn E− and Gullion E− were distinct from their respective definitive stocks. At L’Anjouère, a high frequency of significant differences between the fescue definitive stocks and their E− or E+ equivalents was similarly observed, with Benbaun E− and E+, Beann E− and Cove E− all distinct from their definitive stocks at p < 0.01, with Cove E+ and Eagle E+ at p < 0.05. This meant that half of the fescue definitive versus E− or E+ comparisons were distinguished above the DUS threshold.

A further understanding of the nature of these differences can be derived from the number of characters that provided these differences at each probability level (Tables 9–11). For the E−/E+ comparisons, only Carn involved more than one character and only at p < 0.05 at Crossnacreevy, with the other five occurrences involving a single character. In stark contrast, there were up to six ryegrass characters showing significant differences in pair comparisons of definitive versus E− or E+ (Carn def to E−, 3 at p < 0.01 and 3 at p < 0.05). Equally notable were the definitive stock comparisons with Croob E− and with Croob E+, both of which recorded three character differences at p < 0.001. Among the fescue comparisons, differences from the definitive stocks were similarly strong, with occurrences in 4–6 of the nine characters measured in four of the eight Def/E− or Def/E+ comparisons.
In order to interpret these observations appropriately requires a consideration of the likelihood of these distinctions occurring purely by chance or whether they represent a cause and effect. For ryegrass, the total number of comparisons (character \times accession pairs (Def/E− and Def/E+) \times varieties) was 136 at Crossnacreevy and

Table 7. Example comparison of E− and E+ accessions of Carn perennial ryegrass by COYD analysis of two growing cycles at Scharnhorst, Germany. Values positive if Carn E+ larger than Carn E−, +/- if larger/smaller/equal in each year, +5 = p < 0.05.

Table 8. Example comparison of E− and E+ accessions of Cove tall fescue as COYD analysis of two growing cycles at L'Anjouère, France. Values positive if Cove E+ larger than Cove E−, +/- if larger/smaller in each year.

Table 9. Number of characters showing a significant difference in each accession in pair-wise comparisons for perennial ryegrass varieties at two sites after two growing cycles at Crossnacreevy, N. Ireland. Significances based on COYD analysis of two growing cycles, values show the number of characters expressing a difference at the denoted probability level. ns = number of characters showing no significant difference.
be reported at around 97%\(^27\), incorrect assignment of tillers to \(E^{+}\) differed in a number of comparisons with their official definitive stock. No conclusive evidence of endophyte inclusion creating DUS distinctions, but that these test accessions (\(E^{-}\) and \(E^{+}\)) substantially greater than that expected by chance, in both ryegrass and fescue. Therefore, overall the results show of perennial ryegrass growth\(^30\). Furthermore, under mild to severe drought there is evidence of increased tillering number of characters showing no significant difference.\(^6\)

| Accession | Definitive Accession (Def) | Endophyte Free Accession (E−) |
|-----------|---------------------------|-------------------------------|
|           | \(P < 0.001\) | \(P < 0.01\) | \(P < 0.05\) | ns | \(P < 0.001\) | \(P < 0.01\) | \(P < 0.05\) | ns |
| Binnian (E−) | 0 | 0 | 0 | 15 | – | – | – | – |
| Binnian (E+) | 0 | 0 | 0 | 15 | 0 | 0 | 0 | 15 |
| Carn (E−) | 0 | 0 | 1 | 14 | – | – | – | – |
| Carn (E+) | 0 | 0 | 0 | 15 | 0 | 0 | 1 | 14 |
| Croob (E−) | 0 | 0 | 0 | 15 | – | – | – | – |
| Croob (E+) | 0 | 0 | 0 | 15 | 0 | 0 | 0 | 15 |
| Guillon (E−) | 0 | 1 | 2 | 12 | – | – | – | – |
| Guillon (E+) | 0 | 0 | 0 | 15 | 0 | 1 | 0 | 14 |

Table 10. Number of characters showing a significant difference in each accession in pair-wise comparisons for perennial ryegrass varieties at two sites after two growing cycles at Scharnhorst, Germany. Significances based on COYD analysis of two growing cycles, values show the number of characters expressing a difference at the denoted probability level. ns = number of characters showing no significant difference.

| Accession | Definitive Accession (Def) | Endophyte Free Accession (E−) |
|-----------|---------------------------|-------------------------------|
|           | \(P < 0.001\) | \(P < 0.01\) | \(P < 0.05\) | ns | \(P < 0.001\) | \(P < 0.01\) | \(P < 0.05\) | ns |
| Benbaun (E−) | 1 | 1 | 3 | 5 | – | – | – | – |
| Benbaun (E+) | 1 | 3 | 1 | 4 | 0 | 0 | 0 | 9 |
| Beann (E−) | 1 | 2 | 3 | 3 | – | – | – | – |
| Beann (E+) | 0 | 0 | 0 | 9 | 0 | 0 | 1 | 8 |
| Cove (E−) | 1 | 0 | 3 | 5 | – | – | – | – |
| Cove (E+) | 0 | 0 | 1 | 8 | 0 | 0 | 0 | 9 |
| Eagle (E−) | 0 | 0 | 0 | 9 | – | – | – | – |
| Eagle (E+) | 0 | 0 | 2 | 7 | 0 | 0 | 0 | 9 |

Table 11. Summary of significant difference occurrences in pair-wise comparisons between different accessions of the same tall fescue cultivar at L’Anjouère, France. Significances based on COYD analysis of two growing cycles, values show the number of characters expressing a difference at the denoted probability level. ns = number of characters showing no significant difference.

120 at Scharnhorst and for the E−/E+ comparisons were 68 and 60 respectively (Table 12). For fescue, the total number of comparisons were 72 (Def/E− and Def/E+) and 36 (E−/E+). It can be seen from Table 12, that the number of observed differences between E−/E+ in ryegrass was less or similar to the number expected purely by chance. The same outcome was found for the comparisons between the fescue E−/E+ accessions. For the comparisons between the definitive samples and the E− or E+ accessions, the number of observed differences was substantially greater than that expected by chance, in both ryegrass and fescue. Therefore, overall the results show no conclusive evidence of endophyte inclusion creating DUS distinctions, but that these test accessions (E−/E+) differed in a number of comparisons with their official definitive stock.

**Discussion**

The absence of distinguishing differences between E−/E+ accessions raises two initial questions. Firstly whether the E−/E+ accessions were correctly formulated. Given that the repeatability of the immunoblot assay used has been reported at around 97%\(^27\), incorrect assignment of tillers to E−/E+ accessions is a highly unlikely cause. Furthermore, by using the immunoblot approach it was possible to avoid any side effects of treating seed lots to remove endophyte, such as by heat treatment, that might impair germination or vigour or might incur a fitness sub-selection that could create apparent phenotypic divergences. Secondly it could be questioned whether it was reasonable to expect endophyte inoculation to cause any such effects. There is however, a considerable body of previously published studies that report notable differences between E− and E+ grasses. The symbiotic relationship between endophyte and grass is not strictly mutualistic as there can be negative implications for the host plant. For example, it has been found\(^28\) that some novel cultivar-endophyte associations could incur a yield disadvantage compared with endophyte free plants in the absence of insect herbivores. Similarly, infected tall fescue seed required more moisture to germinate and their seedlings more nutrients\(^29\), presumably due to the photosynthetic cost of supporting the fungus. In contrast there are also some reports of direct benefits of endophyte infection for the host plant. For example, an increased efficiency under low soil nitrogen for infected tall fescue has also been reported\(^28\), possibly through raised glutamine synthetase activity, while others have recorded enhancement of perennial ryegrass growth\(^30\). Furthermore, under mild to severe drought there is evidence of increased tillering and regrowth rates\(^31\), and changes in tiller number, tiller length and shoot mass\(^32\). Most dramatically of all, it has been discovered that endophyte infection triggered reprogramming of host metabolism\(^33\), favouring secondary metabolism at a cost to primary metabolism. It also induced changes in host development, particularly trichome formation and cell wall biogenesis. Therefore, it was reasonable to question whether inoculation with an endophyte might change a cultivars DUS identity.
The three EO sites represented a relatively wide geographic and climatic spread. Crossnacreevy (north of Ireland) is exposed to cool moist air from the maritime polar air mass. L’Anjouère (west coast of France) has a temperate maritime climate with a narrow annual temperature range. Scharnhorst (northern Germany) has a very cold winters and rain spread evenly throughout the year. Despite these contrasting conditions there was no detectable evidence of climatic stress, such as drought, in either test year (nor of any insect attack). Therefore, any protective properties of the endophytes could not have benefited the grasses during the study. Furthermore, this null response is not in conflict with other published work. No significant differences were found in DM yield, DMD, WSC or CP between E− and E+ accessions of the perennial ryegrass cultivars AberDart and AberMagic32, while others also reported similar absence of responses in ryegrass and tall fescue33,34.

A further factor in fully understanding the implications of the current study is that infection responses appear to be more than endophyte strain specific, as detailed earlier, but can also depend on the genotype of the host plant. For example, host plant genotype specific differences in phenotypic responses have been reported36,38. While breeding for effective host-endophyte associations, changes in endophyte metabolite expression associated with host genotype and evidence of co-adaptation between plant and fungus have been reported39. This has been carried through to the commercial scale in fescues by selecting host genotypes that reduced the animal toxic ergovaline production of the endophyte37. Likewise, as the host genotype determines host growth and reproduction, this interacts with the endophyte biology38. Consequently, the findings of the current study cannot be deemed as a fundamental rule applicable to all grass-endophyte associations. Rather, it must be regarded to some extent as specific to these cultivar/strain symbionts. Nonetheless, the evidence does show that inoculating a registered cultivar with an endophyte that is novel to the host does not guarantee that the new association will be distinct from the original cultivar. This is more likely to be the case in DUS tests conducted under conditions where there are no detectable biotic or abiotic stress responses. In these circumstances, the endophyte cannot provide a resilience benefit to the host and so is less likely to induce any change in plant growth or productivity. This should allay, to some extent, the concern that an apparent PBR distinctness could occur between two accessions of the same cultivar, one with and the other without endophyte. However, to fully ensure that PBR protection can be retained for existing registered cultivars, further investigation of whether these findings can be repeated with a wider range of cultivar/strain symbionts, would be additionally informative. As would using test sites where biotic and abiotic stresses ranged from absent to prevalent.

The surprising result of the current study was the high frequency of significant differences and DUS distinctions between individual E− or E+ accessions and their equivalent definitive stocks. It is difficult to discount this as an aberration, given that all three EOs largely observed the same outcome. Each EO provided its own definitive stocks and so those from Crossnacreevy and Scharnhorst were not directly compared to confirm identicalness. However, they had been stored in ideal conditions from the time they were submitted for registration by the breeder. It is also a basic premise of the EU PBR scheme that definitive stocks at each EO are identical. This is to ensure the same protection exists across the entire EU, with validation tests and audit inspections conducted to impose this standardization. Therefore, this eliminates any concerns regarding the authenticity of the definitive stocks, particularly as any fault would have had to reoccur in several cultivars and two species at three sites, independently. The search for a cause therefore, switches to the E−/E+ samples provided by the breeders. The generation of the seed lots used in the trials was considered as a possible factor but as all lots were from certified seed stocks they were validated as complying with their definitive stocks. So this also seems an unlikely cause, particularly as any drift would have also had to occur in several cultivars and two species independently. The individual E−/E+ plants were produced by the breeders and posted to the EOs. This means that they had been treated differently to the definitive stocks, which were grown directly from seed. However, all plants were grown on in multi-pots before planting out. All centres reported successful establishment of all the measured plants, however,
the German EO subsequently reported that the E−/E+ plants were much smaller when planted than those in the rest of the DUS trial, including their definitive stocks. While this might have affected development, particularly for the autumn recorded characters, it would be less expected to significantly affect growth the following year, when most characters were measured. Furthermore, as this difference did not occur at the other test centres, this does not provide a full or adequate explanation. The only other obvious step in the experimentation where accession differences could have been artificially created was during the compilation of the E− and E+ accessions. There is evidence that this was a risk to the study as the vertical transmission of endophytes has been shown to vary with host genotype9,40. Co adaptation of host and endophyte has also been reported31 and in breeding for successful grass/endophyte combinations56. This could mean that in subdividing each accession into E− and E+ plants, this had also divided the cultivar into two different types, one more conducive and one less conducive to endophyte transmission, with associated phenotypic differences. However, if this was the causal mechanism, then the E− and E+ accessions should have been distinct from each other, which they were not. So, at time of publication, the cause of the observed definitive stock differences remains unexplained.

Conclusion

The potential for creating differences in a grass cultivar’s morphological identity by endophyte inoculation has implications for cultivar distinctiveness and for plant variety protection. The evidence from this study shows that inoculating a registered grass cultivar with an endophyte that is novel to the host does not ensure that the new association will be distinct from the original cultivar i.e. endophyte presence is not a reliable mechanism to change the DUS traits of a cultivar. This has importance for those bodies responsible for regulating and testing grass cultivars across Europe as well as the grass breeding industry. However, this cannot be interpreted as a universal rule for all grass-endophyte associations, rather the findings should be regarded as being new evidence, which at the very least, confirms that PBR distinguishing morphological changes are not an inevitable consequence of endophyte inoculation. It must also be stressed that in the three experimental sites there were no acute biotic or abiotic stresses, which reduced the opportunity for the endophyte to benefit the host grass in terms of growth and/or productivity. These implications must be taken into account when evaluating any other cultivar-endophyte associations and particularly when considering how to manage the DUS testing of cultivars that are commercialised as a cultivar/endophyte symbiont.

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Author contributions
Prof Gilliland led this collaborative study in which N. Roulland and S. Charrier prepared and distributed the E+/E− accessions to the three DUS Examination Offices. The authors from these centres, (Lafaillette, Patterson, Gilliland and Wöster), took varying responsibilities for conducting the taxonomy studies at their site, analysing the data and preparing the scientific paper, which was reviewed and agreed by all authors.

Competing interests
The authors declare no competing interests.

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