Franseen Needles May Be Promising for Improving the Sampling Adequacy of EUS-FNA for Subepithelial Lesions

Noriki Kasuga 1,2, Yusuke Kurita 1,*, Emiko Tanida 2, Shin Yagi 1, Ko Suzuki 1, Sho Hasegawa 1, Takamitsu Sato 1, Kunihiro Hosono 1, Shingo Kato 1, Yusuke Sekino 3, Noritoshi Kobayashi 4, Itaru Endo 5, Kensuke Kubota 1 and Atsushi Nakajima 1

1 Department of Gastroenterology and Hepatology, Yokohama City University Hospital, Yokohama 236-0004, Japan; kasuganoriki1992@gmail.com (N.K.); e103083b@gmail.com (S.Y.); kosuzuki@yokohama-cu.ac.jp (K.S.); pinicom-6@hotmail.co.jp (S.H.); tkmtsato@yokohama-cu.ac.jp (T.S.); hiro1017@yokohama-cu.ac.jp (K.H.); shin800m@yokohama-cu.ac.jp (S.K.); kubotak@yokohama-cu.ac.jp (K.K.); nrk20799@nifty.com (A.N.)
2 Department of Gastroenterology, Machida Municipal Hospital, Tokyo 194-0023, Japan; et0505polepole@gmail.com
3 Department of Gastroenterology, Yokohama Rosai Hospital, Yokohama 222-0036, Japan; ysekino1978@gmail.com
4 Department of Oncology, Yokohama City University Hospital, Yokohama 236-0004, Japan; norikoba@yokohama-cu.ac.jp
5 Department of Gastroenterological Surgery, Yokohama City University Hospital, Yokohama 236-0004, Japan; endoit@yokohama-cu.ac.jp
* Correspondence: kuritay@yokohama-cu.ac.jp; Tel.: +81-45-787-2640; Fax: +81-45-784-3546

Abstract: Endoscopic ultrasound-guided fine needle aspiration (EUS-FNA) is useful in diagnosing subepithelial lesions (SELS), and adequate tissue sampling is necessary to differentiate between benign and malignant diseases to determine therapeutic strategies. This study aimed to evaluate sampling adequacy and diagnostic performance of EUS-FNA for SELS with Franseen needles. This retrospective study enrolled 130 patients who underwent EUS-FNA with a 22-gauge needle for SELS from January 2010 to March 2021. We compared sampling adequacy and predictive factors influencing the sampling adequacy of EUS-FNA for SELS between Franseen and conventional needles. The sampling adequacy rates were 95.0% (38/40) with Franseen needles and 76.7% (69/90) with conventional needles (p = 0.011). The mean number of punctures with Franseen needles (2.80) was significantly less than that with conventional needles (3.42) (p < 0.001). In the multivariate analysis, the use of Franseen needles (p = 0.029; odds ratio [OR], 5.37; 95% confidence interval [CI], 1.18–23.36) was an independent factor influencing the sampling adequacy. Compared to conventional needles, the Franseen needle could play a vital role in accurately diagnosing SELS by yielding better sampling adequacy and reducing the number of passes.

Keywords: endoscopic ultrasound-guided fine needle aspiration; SEL; sampling adequacy rate; multivariate analysis

1. Introduction

Subepithelial lesions (SELS) of the gastrointestinal tract are often encountered during endoscopy [1]. A gastrointestinal stromal tumor (GIST), which is an SEL with the potential to vary from benign to malignant, is found incidentally in endoscopy; thus, pathological evaluations of SELS are important to determine a treatment strategy [2–4]. It is crucial for endoscopists to make a differential diagnosis of SELS for potential malignancy, which can lead to further procedures such as resection. Since imaging detection sometimes results in an inaccurate diagnosis and endoscopic biopsy is challenging, it is essential for patients with SELS to receive adequate tissue sampling by endoscopic ultrasound-guided fine needle aspiration (EUS-FNA) [5,6].
The sampling adequacy rate of EUS-FNA for SELs [7–10] has been reported, and EUS-FNA has been found to be useful for diagnosis. Various instrument innovations have been developed to enhance the sampling adequacy of EUS-FNA. The usefulness of a Franseen needle for EUS-FNA, which was developed with three novel, symmetric heels on the tip of the needle (Figure 1), has been established [7,11–14]. Although the sampling adequacy rate of the Franseen needle tends to be higher than that of the conventional needle in SELs <20 mm in diameter in a study with few cases [14,15], the superiority of Franseen needles for the sampling adequacy and diagnostic ability for SELs is also unclear [7]. There are no reports that clearly indicate that Franseen needles are an independent factor that contribute to sampling adequacy for SELs. Therefore, the present study aimed to compare the sampling adequacy rate, factors contributing to sampling adequacy, and diagnostic performance of EUS-FNA with the Franseen and conventional needles for SELs in the gastrointestinal tract.

**Figure 1.** A Franseen needle, which has three points and heels on the tip. © 2022 Boston Scientific Corporation. All rights reserved. Reprinted with permission from [https://www.bostonscientific.com/pt-BR/produtos/agulhas/acquire.html]. 2022, Boston Scientific Corporation.

### 2. Materials and Methods

#### 2.1. Patients

This retrospective study involved 130 patients who underwent EUS-FNA with 22-gauge needles for SELs at Yokohama City University Hospital and Machida Municipal Hospital from January 2010 to March 2021.

#### 2.2. EUS-FNA Indication and Procedure

In this study, EUS-FNA for SEL was performed in symptomatic cases and in cases with high-risk features such as ulceration, irregular borders, internal heterogeneity in EUS, and an increase in size during follow-up in accordance with the guidelines [4].

We intravenously administered 2–10 mg midazolam (Astellas, Tokyo, Japan), 5 mg diazepam (Takeda Pharma, Tokyo, Japan), and 15 mg pentazocine (Maruishi Pharma, Osaka, Japan) to patients to perform EUS-FNA with sedation. EUS-FNA procedures were performed with two different convex types of ultrasound endoscopes: a forward-oblique viewing endoscope (GF-UCT240 and GF-UCT260; Olympus Corporation, Tokyo, Japan) and a forward-viewing endoscope (TGF-UC260J; Olympus Corporation, Tokyo, Japan). These were connected to an ultrasound scanning system (EU-ME1 and EU-ME2 Premier Plus; Olympus Corporation, Tokyo, Japan). After the lesions were punctured, EUS-FNA was performed by stroking 20–30 times using negative suction with a 20-mL syringe (Figure 2A). These processes were carried out by experienced endoscopists who had performed more than 50 EUS-FNA procedures prior to this study. All EUS-FNA procedures were performed with 22-gauge needles. We used either a Franseen needle (Acquire; Boston Scientific Corporation, Tokyo, Japan) or conventional needles (EZ-Shot3
Diagnostics 2022, 12, x FOR PEER REVIEW 3 of 11

and a forward-viewing endoscope (TGF-UC260J; Olympus; Tokyo, Japan). The protocols are outlined as follows: for deparaffinization, the slides were heated to 72 °C for 16 min, each diluted at 1:50 to 1:500, depending on the type of antibody. Experienced pathologists performed all analyses. Immunohistochemical staining was performed using an automated immunohistochemistry system (Ventana BenchMark ULTRA; Roche Diagnostics, Basel, Switzerland). The two institutions did not implement rapid on-site cytologic evaluation (ROSE) by cytopathologists or cytotechnologists. The EUS-FNA procedure was terminated after each endoscopist visually confirmed the presence of thin, whitish specimens (Figure 2B). The specimens were fixed in 10% formalin and embedded in paraffin. EUS-FNA specimens were stained with hematoxylin and eosin (H&E) for histologic examination (Figure 2C) and, if necessary for diagnosis, immunohistochemical staining was performed (Figure 2D,E). A diagnosis of mesenchymal tumors was made by comprehensive evaluation with immunohistochemical staining with c-kit, CD34, desmin, S-100 protein, and DOG-1. A positive c-kit staining with or without positive CD34 staining was diagnosed as GIST, positive desmin staining was diagnosed as leiomyoma, and positive S-100 staining was diagnosed as schwannoma. Immunohistochemical staining with other agents like cytokeratin, vimentin, Ki-67, and p53 was used to diagnose carcinoma or sarcoma, and staining with chromogranin A, synaptophysin, and CD56 was used to diagnose neuroendocrine neoplasm (NEN). Immunohistochemical staining was performed using an automated immunohistochemistry system (Ventana BenchMark ULTRA; Roche Diagnostics, Basel, Switzerland). The protocols are outlined as follows: for deparaffinization, the slides were heated to 72 °C; for antigen retrieval, the slides were heated to 95 °C and incubated for 8 min; the required antibodies were titrated onto the slides and incubated for 16 min, each diluted at 1:50 to 1:500, depending on the type of antibody. Experienced pathologists performed all analyses.

2.3. Histological Evaluation

The two institutions did not implement rapid on-site cytologic evaluation (ROSE) by cytopathologists or cytotechnologists. The EUS-FNA procedure was terminated after each endoscopist visually confirmed the presence of thin, whitish specimens (Figure 2B). The specimens were fixed in 10% formalin and embedded in paraffin. EUS-FNA specimens were stained with hematoxylin and eosin (H&E) for histologic examination (Figure 2C) and, if necessary for diagnosis, immunohistochemical staining was performed (Figure 2D,E). A diagnosis of mesenchymal tumors was made by comprehensive evaluation with immunohistochemical staining with c-kit, CD34, desmin, S-100 protein, and DOG-1. A positive c-kit staining with or without positive CD34 staining was diagnosed as GIST, positive desmin staining was diagnosed as leiomyoma, and positive S-100 staining was diagnosed as schwannoma. Immunohistochemical staining with other agents like cytokeratin, vimentin, Ki-67, and p53 was used to diagnose carcinoma or sarcoma, and staining with chromogranin A, synaptophysin, and CD56 was used to diagnose neuroendocrine neoplasm (NEN). Immunohistochemical staining was performed using an automated immunohistochemistry system (Ventana BenchMark ULTRA; Roche Diagnostics, Basel, Switzerland). The protocols are outlined as follows: for deparaffinization, the slides were heated to 72 °C; for antigen retrieval, the slides were heated to 95 °C and incubated for 8 min; the required antibodies were titrated onto the slides and incubated for 16 min, each diluted at 1:50 to 1:500, depending on the type of antibody. Experienced pathologists performed all analyses.

2.4. Definitions of Sampling Adequacy and Diagnostic Ability

Adequate sampling was defined as samples sufficient for histopathologic evaluations and immunohistochemical analyses.

The diagnostic ability of EUS-FNA was evaluated in terms of sensitivity, specificity, positive predictive value (PPV), negative predictive value (NPV), and accuracy. In this
study of diagnostic ability, all surgery cases were included. Of the nonsurgical cases, those diagnosed adequately by EUS-FNA as malignant or benign with at least 6 months of clinical follow-up were included. Of the nonsurgical cases, those without sufficient EUS-FNA samples for histopathologic evaluation and immunohistochemical analysis and/or those without clinical follow-up for at least 6 months were excluded. The final diagnostic criteria were determined either by the surgical histopathological results of the cases of surgical resection or by EUS-FNA diagnosis and clinical follow-up of at least 6 months. The diagnostic ability of EUS-FNA was defined as follows: (1) the EUS-FNA histopathological examination was consistent with the diagnoses of the surgical resection specimens, and (2) the clinical follow-up course of at least 6 months without surgical resection corresponded to that of the EUS-FNA diagnoses.

We categorized the patients into two groups: (1) malignant group, including all GIST, carcinomas such as adenocarcinoma or squamous cell carcinoma, sarcoma, NEN, and malignant lymphoma, and (2) benign group, including leiomyoma, schwannoma, spindle cell tumor (containing mesenchymal tumors for which immunostaining was not performed), aberrant pancreas, inflammatory granuloma, lymph node, hematoma, mucosal prolapse syndrome, gastritis, and lipoma. Non-diagnosis cases were defined as cases excluded from this study of diagnostic ability, nonsurgical cases without adequate sampling of EUS-FNA, and/or cases without clinical follow-up for at least 6 months.

2.5. Factors Influencing the Sampling Adequacy of EUS-FNA

Variables employed for univariate and multivariate analysis were gender (females vs. males), tumor size (<20 mm vs. ≥20 mm), location of the lesion (esophagus, stomach, duodenum, and rectum), EUS-FNA procedure period (2017–2021 vs. 2010–2016), needle shape (Franseen needle vs. conventional needle), and the endoscope (forward-viewing ultrasound endoscope: TGF-UC260J vs. forward-oblique viewing ultrasound endoscope: GF-UCT240 and GF-UCT260).

2.6. Endpoints

The primary endpoint was the sampling adequacy rate of EUS-FNA for SELs using Franseen and conventional needles. Secondary endpoints were factors influencing the sampling adequacy rate. We also assessed the sensitivity, specificity, PPV, NPV, and accuracy of EUS-FNA for SELs.

2.7. Statistical Analysis

Continuous variables were presented as median (range) or mean ± standard deviation. Statistical analysis was conducted using the chi-squared test (or Fisher’s exact, if appropriate) for categorical variables and the Student’s t-test or Mann–Whitney U test for continuous variables for univariate analyses. The Student’s t-test was used if a normal distribution was likely, and the Mann–Whitney U test was used if normality could not be demonstrated. Logistic regression analysis was used for multivariate analysis. Values of p < 0.05 were regarded as statistically significant. Multivariable analysis was performed by selecting variables with p < 0.05 in univariate analysis. SPSS version 28 software (IBM Corp, Armonk, NY, USA) was used for statistical analysis.

3. Results

3.1. Patient and Lesion Characteristics

All 130 patients who underwent EUS-FNA for SELs at the two institutions were enrolled. The patient and lesion characteristics of EUS-FNA for SELs are shown in Table 1. The median age of the patients was 65.0 years (range, 23–90 years), 46.2% (60/130) were females, and the median diameter of the SELs was 25.0 mm (range, 8.0–90.5 mm). Tumor locations were the stomach in 96 cases (73.8%), the duodenum in 21 cases (16.2%), the rectum in 10 cases (7.7%), and the esophagus in 3 cases (2.3%). Patient and lesion characteristics showed no significant difference between the Franseen and conventional needles.
Table 1. Patient and lesion characteristics of cases with EUS-FNA for SELs (n = 130).

| Variables                      | Total n = 130 | Franseen n = 40 | Conventional n = 90 | p-Value |
|-------------------------------|---------------|-----------------|---------------------|---------|
| Age (years)                   |               |                 |                     |         |
| Median (range)                | 65.0 (23–90)  | 65.0 (44–84)    | 64.5 (23–90)        | 0.772   |
| Gender, n (%)                 |               |                 |                     |         |
| Females                       | 60 (46.2)     | 16 (40.0)       | 44 (48.9)           | 0.348   |
| Males                         | 70 (53.8)     | 24 (60.0)       | 46 (51.1)           |         |
| Lesion size (mm)              |               |                 |                     |         |
| Median (range)                | 25.0 (8.0–90.5)| 24.0 (10.0–80.0)| 25.0 (8.0–90.5)     | 0.739   |
| Lesion location, n (%)        |               |                 |                     | 0.145   |
| Esophagus                     | 3 (2.3)       | 1 (2.5)         | 2 (2.2)             |         |
| Stomach                       | 96 (73.8)     | 29 (72.5)       | 67 (74.4)           |         |
| Duodenum                      | 21 (16.2)     | 4 (10.0)        | 17 (18.9)           |         |
| Rectum                        | 10 (7.7)      | 6 (15.0)        | 4 (4.4)             |         |

EUS-FNA, endoscopic ultrasound-guided fine needle aspiration; SELs, subepithelial lesions.

Among 130 cases, 72 (55.4%) were surgically resected. The final diagnosis of all SELs for which EUS-FNA was performed is shown in Table 2. The most frequent diseases were GIST (52.3%, 68/130), followed by leiomyoma (7.7%, 10/130), carcinoma (7.7%, 10/130), and schwannoma (4.6%, 6/130). All non-diagnosis cases were nonsurgical; 17 cases did not have adequate sampling for EUS-FNA, and 2 cases were not followed-up for more than 6 months.

Table 2. Details of final diagnosis of all SELs (n = 130).

| Disease, n (%)         | Total n = 130 | No. Surgical n = 72 | Nonsurgical n = 58 |
|------------------------|---------------|---------------------|-------------------|
| GIST 68 (52.3)         | 55 (76.4)     | 13 (22.4)           |                   |
| Leiomyoma 10 (7.7)     | 3 (4.2)       | 7 (12.1)            |                   |
| Carcinoma 10 (7.7)     | 5 (6.9)       | 5 (8.6)             |                   |
| Schwannoma 6 (4.6)     | 3 (4.2)       | 3 (5.2)             |                   |
| Sarcoma 3 (2.3)        | 0             | 3 (5.2)             |                   |
| NEN 2 (1.5)            | 1 (1.4)       | 1 (1.7)             |                   |
| Aberrant pancreas 2 (1.5)| 0         | 2 (3.4)             |                   |
| Hematoma 2 (1.5)       | 2 (2.8)       | 0                   |                   |
| Lymph node 2 (1.5)     | 0             | 2 (3.4)             |                   |
| Inflammatory granuloma 1 (0.8)| 1 (1.4) | 0                   |                   |
| MALT lymphoma 1 (0.8)  | 0             | 1 (1.7)             |                   |
| MPS 1 (0.8)            | 0             | 1 (1.7)             |                   |
| Gastritis 1 (0.8)      | 0             | 1 (1.7)             |                   |
| Hyperplasia 1 (0.8)    | 1 (1.4)       | 0                   |                   |
| Lipoma 1 (0.8)         | 1 (1.4)       | 0                   |                   |
| Non-diagnosis 19 (14.6)| 0             | 19 (32.8)           |                   |

SELs, subepithelial lesions; GIST, gastrointestinal stromal tumor; NEN, neuroendocrine neoplasm; MALT lymphoma, mucosa-associated lymphoid tissue lymphoma; MPS, mucosal prolapse syndrome.

3.2. Sampling Adequacy Rates and Outcomes

The comparison of the outcomes of EUS-FNA performed with either Franseen or conventional needles is shown in Table 3. The puncture technique was successful in all cases (130/130). The overall sampling adequacy rate of EUS-FNA for SELs was 82.3% (107/130). The Franseen needles had a significantly superior sampling adequacy rate compared to conventional needles (95.0% vs. 76.7%; p = 0.011). Endoscopists visually confirmed the presence of thin, whitish specimens, which could be attained with one to six punctures. The mean overall number of punctures was 3.23 ± 0.95, and the mean number of punctures for Franseen needles was significantly lower than that of conventional...
needles (2.80 ± 0.87 vs. 3.42 ± 0.92; p < 0.001). Two cases (1.5%; 2/130) experienced bleeding adverse events with conventional needles, but none of them required blood transfusions. There were no deaths or serious adverse events related to EUS-FNA in this study.

Table 3. Comparison of technical EUS-FNA outcomes for SELs (n = 130).

| Variables                        | Total n = 130 | Franseen n = 40 | Conventional n = 90 | p-Value |
|----------------------------------|---------------|-----------------|---------------------|---------|
| Puncture success                 | 100% (130/130)| 100% (40/40)    | 100% (90/90)        | N.S.    |
| Adverse events                   |               |                 |                     |         |
| Bleeding                         | 1.5% (2/130)  | 0               | 2.2% (2/90)         | 1.000   |
| Perforation                      | 0             | 0               | 0                   | N.S.    |
| Infection                        | 0             | 0               | 0                   | N.S.    |
| Death                            | 0             | 0               | 0                   | N.S.    |
| Number of punctures              |               |                 |                     |         |
| Mean ± SD                        | 3.23 ± 0.95   | 2.80 ± 0.87     | 3.42 ± 0.92         | <0.001  |
| Sampling adequacy rate           | 82.3% (107/130)| 95.0% (38/40)  | 76.7% (69/90)       | 0.011   |

EUS-FNA, endoscopic ultrasound-guided fine needle aspiration; SELs, subepithelial lesions; SD, standard deviation; N.S., not significant.

3.3. Analysis of Factors influencing the Sampling Adequacy

Univariate and multivariate analyses were performed to identify the factors influencing the sampling adequacy rate of EUS-FNA for SELs (Table 4). Univariate analysis revealed duodenal lesions (p = 0.047) and the use of Franseen needles (p = 0.011) as significant factors. However, gender, tumor size, procedure period, and type of endoscope were found to be non-significant factors. In multivariate analysis, the use of Franseen needles (p = 0.029; odds ratio [OR], 5.37; 95% confidence interval [CI], 1.18–23.36) was an independent factor influencing the sampling adequacy of EUS-FNA for SELs.

Table 4. Factors influencing the sampling adequacy of EUS-FNA for SELs (n = 130).

| Factors                        | Sampling Adequacy | Univariate Analysis p-Value | OR  | Multivariate Analysis 95% CI | p-Value |
|--------------------------------|-------------------|-----------------------------|-----|------------------------------|---------|
| Gender                         |                   |                             |     |                              |         |
| Females                        | 83.3% (50/60)     | 0.777                       | 0.77 | 0.006                        | 0.006   |
| Males                          | 81.4% (57/70)     |                             |     |                              |         |
| Tumor size                     |                   |                             |     |                              |         |
| <20 mm                         | 73.3% (22/30)     | 0.142                       | 0.48 | 0.001                        | 0.001   |
| ≥20 mm                         | 85.0% (85/100)    |                             |     |                              |         |
| Location                       |                   |                             |     |                              |         |
| Esophagus                      | 66.7% (2/3)       | 0.485                       | 0.485 | 0.003                        | 0.003   |
| Stomach                        | 85.4% (82/96)     | 0.124                       | 0.124 | 0.006                        | 0.006   |
| Duodenum                       | 63.6% (14/22)     | 0.047                       | 0.397 | 0.13–1.15                    | 0.088   |
| Rectum                         | 90.0% (9/10)      | 0.515                       | 0.515 | 0.001                        | 0.001   |
| Period                         |                   |                             |     |                              |         |
| 2010–2016                      | 78.4% (58/74)     | 0.177                       | 0.177 | 0.003                        | 0.003   |
| 2017–2021                      | 87.5% (49/56)     |                             |     |                              |         |
| Shape of needle                |                   |                             |     |                              |         |
| Franseen                       | 95.0% (38/40)     | 0.011                       | 5.37 | 1.18–23.36                   | 0.029   |
| Conventional                   | 76.7% (69/90)     |                             |     |                              |         |
| Field of view                  |                   |                             |     |                              |         |
| Forward                        | 100% (7/7)        | 0.352                       | 0.352 | 0.001                        | 0.001   |
| Forward-oblique                | 81.3% (100/123)   |                             |     |                              |         |

EUS-FNA, endoscopic ultrasound-guided fine needle aspiration; SELs, subepithelial lesions; OR, odds ratio; CI, confidence interval; –, not analyzed.

3.4. Comparison of Diagnostic Ability

The diagnostic ability was studied in 111 cases, including all 72 surgical cases and 39 nonsurgical cases excluded without adequate sampling of EUS-FNA and/or without clinical follow-up for at least 6 months. The overall sensitivity, specificity, PPV, NPV, and accuracy of EUS-FNA for SELs were 96.4% (81/84), 96.3% (26/27), 98.8% (81/82), 89.7% (26/29), and 96.4% (107/111), respectively, as shown in Table 5. Comparing Franseen
and conventional needles, the accuracy was higher for Franseen needles (100% vs. 94.7%; \( p = 0.302 \)), but there was no significant difference.

Table 5. Diagnostic ability of EUS-FNA for SELs compared with Franseen and conventional needles \((n = 111)\).

| Needle                  | Cases \((n)\) | Total \(n = 111\) | Franseen \(n = 36\) | Conventional \(n = 84\) | \( p\)-Value |
|-------------------------|---------------|-------------------|---------------------|-------------------------|------------|
| Sensitivity             |              | 96.4% (81/84)     | 100% (29/29)        | 94.5% (52/55)           | 0.548      |
| Specificity             |              | 96.3% (26/27)     | 100% (7/7)          | 95.0% (19/20)           | 1.000      |
| PPV                     |              | 98.8% (81/82)     | 100% (29/29)        | 98.1% (52/53)           | 1.000      |
| NPV                     |              | 89.7% (26/29)     | 100% (7/7)          | 86.4% (19/22)           | 0.557      |
| Accuracy                |              | 96.4% (107/111)   | 100% (36/36)        | 94.7% (71/75)           | 0.302      |

EUS-FNA, endoscopic ultrasound-guided fine needle aspiration; SELs, subepithelial lesions; PPV, positive predictive value; NPV, negative predictive value.

4. Discussion

The current study conducted in the two institutions showed that the Franseen needle, as an independent factor in multivariate analysis, was associated with a better sampling adequacy rate than the conventional needle. For needle penetration into the tissue, a smaller included angle of the cutting edge radius and a smaller inclination angle of cutting edge relative to the cutting direction are advantageous because they reduce the insertion force. As a result, the Franseen needle, with a symmetrical three-heeled geometry, reduces the included and inclination angles and enables easier and more secure tissue acquisitions than the lancet shape of the conventional needle [17]. Regarding EUS-FNA for SELs, the Franseen needle would be preferable for endoscopists in terms of sampling adequacy and high diagnostic ability compared to the conventional needle. The lack of a significant difference in the accuracy between Franseen and conventional needles may be because many cases were excluded from the diagnostic ability study due to inadequate sampling adequacy with the conventional needle.

Furthermore, this study showed that the Franseen needle could reduce unnecessary punctures. Although the use of Franseen needles has been shown to reduce the number of punctures in pancreatic lesions [7,18], there have been no such clear reports for SELs, which was novel in this study. Despite the few punctures, the Franseen needle provided a sufficient number of samples for pathology in the present study. As a result, the Franseen needle yields sufficient specimens to enable visual assessment by endoscopists.

ROSE is an effective method to reduce the number of EUS-FNA punctures [19,20], but not all endoscopic units can perform ROSE due to lack of manpower; in this study, ROSE was not performed. Evidence demonstrating that the Franseen needle reduces the number of EUS-FNA punctures for SELs, as in the present study, might be valuable for endoscopic units where ROSE is unavailable.

EZ-shot3 (Olympus Corporation, Tokyo, Japan), EchoTip Ultra (Cook Medical Japan, Tokyo, Japan), Expect (Boston Scientific Corporation, Tokyo, Japan), and SonoTip (Medico’s Hirata, Osaka, Japan), which were used as the conventional needles, were respectively priced at JPY 27,000, JPY 27,000, JPY 27,000, and JPY 26,000, while Acquire (Boston Scientific Corporation, Tokyo, Japan), which was used as the Franseen needle in this study, was priced at JPY 32,000. The Franseen needle was more expensive than conventional needles; however, the sampling adequacy rate of conventional needles was 76.7%, and the sampling adequacy rate of the Franseen needle was 95.0% in this study. The sampling adequacy rate of conventional needles was 18.3% lower than that of the Franseen needle and is therefore associated with the risk of increasing the number of patients who need to perform a repeat EUS-FNA, thus increasing the medical costs. Therefore, using Franseen needles in EUS-FNA for SELs may reduce total costs compared to using conventional needles.
In univariate analysis, the use of a Franseen needle and a duodenal lesion were factors affecting the sampling adequacy rate of EUS-FNA for SELs. There was no significant difference in patient age, gender, tumor size, or scope type. The size of the SEL did not affect the sampling adequacy in our study, indicating that EUS-FNA for SELs with a Franseen needle, if successful, provides sampling adequacy whether or not the tumor diameter was <20 mm. The significantly lower sampling adequacy rate for duodenal lesions compared to other gastrointestinal tracts may be related to the fact that puncture resistance is very high in positions that require an upward EUS angle, as described by Itoi et al. [21].

The impact of the Franseen needle, which was mainly used in this study, has been shown for SELs [7,14], and other new needles, such as needles with side holes [22] and fork-tips [23,24], have been developed and show a good sampling adequacy rate for SELs. Despite this increase in the number of puncture needle options, there is no one particular needle used uniformly. Regarding needle selection, needle size has also been discussed; needle size may be a confounder, as the potential usefulness of 19-gauge [25–27] and 25-gauge [28] puncture needles has been shown. Although a 19-gauge needle can provide sufficient tissue, the strong puncture resistance makes it very difficult to use a 19-gauge needle, especially for small lesions. Furthermore, the specimens may be too small for a 25-gauge needle. Therefore, 22-gauge needles are more commonly used in EUS-FNA. In the present study, only 22-gauge puncture needles were included, and a 22-gauge Franseen needle showed a good sample collection rate. In the case of SELs, a 22-gauge Franseen needle is considered sufficient to obtain sufficient samples for cell block preparation and subsequent immunohistochemical staining, which is very useful for diagnosing SEL subtypes, and sampling of these lesions should be done routinely.

Recently, EUS-FNA has been shown to have utility not only for diagnosing lesions but also for genomic characterization to develop molecular profiling to find personalized anti-cancer therapies [29]. Genomic characterization for GIST is also attracting attention [30,31], but surgical specimens are often used since genomic characterization requires sufficient tissue sampling. The Franseen needle, with a high sampling adequacy rate, may have a role in the potential genomic characterization for GIST. Consequently, the Franseen needle would meet future medical needs.

We had several limitations in this study: first, the retrospective nature of the study had inherent limitations; second, one type of Franseen needle was compared to several types of conventional needles in this study; and third, the current study included the possibility of needle selection bias by the endoscopists.

5. Conclusions

While the sampling adequacy rate for duodenal lesions was significantly lower and should be treated with caution, the Franseen needle was also an independent factor in improving the sampling adequacy rate of EUS-FNA for SELs. EUS-FNA for SELs with the Franseen needle had a significantly better sampling adequacy rate with fewer punctures than the conventional needles.

Author Contributions: N.K. (Noriki Kasuga); manuscript writing and study conception and design. Y.K.; study conception and design, performing the endoscopic procedures, and assisting with manuscript writing. E.T., S.Y., K.S., S.H., T.S., K.H. and K.K.; performing the endoscopic procedures, analyzing data, and providing clinical advice. S.K., Y.S., N.K. (Noritoshi Kobayashi) and A.N.; providing clinical advice. I.E.; performing the surgery and providing clinical advice. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Institutional Review Board of the Yokohama City University Hospital (B200700001, approved on 15 July 2020) and Machida Municipal Hospital (CT-59, approved on 15 September 2021).
Informed Consent Statement: Patient consent was waived because only medical information was used without invasion of the participants’ privacy in this multicenter retrospective observational study. Instead, all patients received an opt-out form for informed consent, and those who did not consent were excluded.

Data Availability Statement: Data available on request because of restrictions, e.g., privacy or ethics.

Acknowledgments: We would like to thank the endoscopic medical staff of the Yokohama City University Hospital and Machida Municipal Hospital for their cooperation and help with EUS-FNA.

Conflicts of Interest: Noritoshi Kobayashi received an honorarium from FUJIFILM Toyama Chemical Co., Ltd.

References

1. Hedenbro, J.L.; Ekelund, M.; Wetterberg, P. Endoscopic diagnosis of submucosal gastric lesions. The results after routine endoscopy. Surg. Endosc. 1991, 5, 20–23. [CrossRef] [PubMed]
2. Miettinen, M.; Lasota, J. Gastrointestinal stromal tumors—Definition, clinical, histological, immunohistochemical, and molecular genetic features and differential diagnosis. Virchows Arch. 2001, 438, 1–12. [CrossRef] [PubMed]
3. Ando, N.; Goto, H.; Niwa, Y.; Hirooka, Y.; Ohmiya, N.; Nagasaka, T.; Hayakawa, T. The diagnosis of GI stromal tumors with EUS-guided fine needle aspiration with immunohistochemical analysis. Gastrointest. Endosc. 2002, 55, 37–43. [CrossRef] [PubMed]
4. Nishida, T.; Blay, J.Y.; Hirota, S.; Kitagawa, Y.; Kang, Y.K. The standard diagnosis, treatment, and follow-up of gastrointestinal stromal tumors based on guidelines. Gastric Cancer 2016, 19, 3–14. [CrossRef]
5. Iglesias-Garcia, J.; Larino-Noia, J.; Abdulkader, I.; Dominguez-Munoz, J.E. Rapid on-site evaluation of endoscopic-ultrasound-guided fine-needle aspiration diagnosis of pancreatic masses. World J. Gastroenterol. 2014, 20, 9451–9457. [CrossRef]
6. Okasha, H.H.; Naguib, M.; El Nady, M.; Ezzat, R.; Al-Gemeie, E.; Al-Nabawy, W.; Aref, W.; Abdel-Moaty, A.; Essam, K.; Hamdy, A. Role of endoscopic ultrasound and endoscopic-ultrasound-guided fine-needle aspiration in endoscopic biopsy negative gastrointestinal lesions. Endosc. Ultrasound 2017, 6, 156–161. [CrossRef]
7. Facciorusso, A.; del Prete, V.; Buccino, V.R.; Purohit, P.; Setia, P.; Muscatiello, N. Diagnostic yield of Franseen and Fork-Tip biopsy needles for endoscopic cutting edge tissue acquisition: A meta-analysis. Endosc. Int. Open 2019, 7, E1221–E1230. [CrossRef]
8. Mekky, M.A.; Yamao, K.; Sawaki, A.; Mizuno, N.; Hara, K.; Nafeh, M.A.; Osman, A.M.; Koshikawa, T.; Yatabe, Y.; Bhatia, V. Diagnostic utility of EUS-guided FNA in patients with gastric submucosal tumors. Gastrointest. Endosc. 2010, 71, 913–919. [CrossRef]
9. Niimi, K.; Goto, O.; Kawakubo, K.; Nakai, Y.; Minatsuki, C.; Asada-Hirayama, I.; Mochizuki, S.; Ono, S.; Kodashima, S.; Yamamichi, N.; et al. Endoscopic ultrasound-guided fine-needle aspiration skill acquisition of gastrointestinal stromal tumour by trainee endoscopists: A pilot study. Endosc. Ultrasound 2016, 5, 157–164. [CrossRef]
10. Akahoshi, K.; Sumida, Y.; Matsui, N.; Oya, M.; Akinaga, R.; Kubokawa, M.; Motomura, Y.; Honda, K.; Watanabe, M.; Nagai, T. Preoperative diagnosis of gastrointestinal stromal tumor by endoscopic ultrasound-guided fine needle aspiration. World J. Gastroenterol. 2007, 13, 2077–2082. [CrossRef]
11. Bang, J.Y.; Hebert-Magee, S.; Hasan, M.K.; Navaneethan, U.; Hawes, R.; Varadarajulu, S. Endoscopic ultrasonography-guided biopsy using a Franseen needle design: Initial assessment. Dig. Endosc. 2017, 29, 338–346. [CrossRef]
12. Bang, J.Y.; Hebert-Magee, S.; Navaneethan, U.; Hasan, M.K.; Hawes, R.; Varadarajulu, S. Randomized trial comparing the Franseen and Fork-tip needles for EUS-guided fine-needle biopsy sampling of endoscopic pancreatic masses. Gastrointest. Endosc. 2018, 87, 1432–1438. [CrossRef] [PubMed]
13. Mukai, S.; Itoi, T.; Yamaguchi, H.; Sofuni, A.; Tsuchiya, T.; Tanaka, R.; Tonozuka, R.; Honjo, M.; Fujita, M.; Yamamoto, K.; et al. A retrospective histological comparison of EUS-guided fine-needle biopsy using a novel franseen needle and a conventional end-cut type needle. Endosc. Ultrasound 2019, 8, 50–57. [CrossRef] [PubMed]
14. Fujita, A.; Ryozawa, S.; Kobayashi, M.; Araki, R.; Nagata, K.; Minami, K.; Tanisaka, Y.; Kobatake, T.; Mizuide, M. Diagnostic ability of a 22G Franseen needle in endoscopic ultrasound-guided fine needle aspiration of subepithelial lesions. Mol. Clin. Oncol. 2018, 9, 527–531. [CrossRef] [PubMed]
15. Sekine, M.; Miura, T.; Fujisawa, J.; Uehara, T.; Araki, N.; Matsumoto, S.; Miyatani, H.; Mashima, H. Utility of endoscopic ultrasonography-guided fine-needle biopsy (EUS-FNB) for diagnosing small subepithelial lesions (<20 mm). J. Ultrason. 2021, 25, 35–40. [CrossRef]
16. Cotton, P.B.; Eisen, G.M.; Aabakken, L.; Baron, T.H.; Hutter, M.M.; Jacobson, B.C.; Mergener, K.; Nemec, A., Jr.; Petersen, B.T.; Pettrini, J.L.; et al. A lexicon for endoscopic adverse events: Report of an ASGE workshop. Gastrointest. Endosc. 2010, 71, 446–454. [CrossRef]
17. Han, P.; Che, D.; Pallav, K.; Ehmann, K. Models of the cutting edge geometry of medical needles with applications to needle design. Int. J. Mech. Sci. 2012, 65, 157–167. [CrossRef]
18. Facciorusso, A.; Bajwa, H.S.; Menon, K.; Buccino, V.R.; Muscatiello, N. Comparison between 22G aspiration and 22G biopsy needles for EUS-guided sampling of pancreatic lesions: A meta-analysis. Endosc. Ultrasound 2020, 9, 167–174. [CrossRef]
19. Wani, S.; Mullady, D.; Early, D.S.; Rastogi, A.; Collins, B.; Wang, J.F.; Marshall, C.; Sams, S.B.; Yen, R.; Rizeq, M.; et al. The clinical impact of immediate on-site cytopathology evaluation during endoscopic ultrasound-guided fine needle aspiration of pancreatic masses: A prospective multicenter randomized controlled trial. *Am. J. Gastroenterol.* **2015**, 110, 1429–1439. [CrossRef]

20. Khan, M.A.; Grimm, I.S.; Ali, B.; Nollan, R.; Tombazzi, C.; Ismail, M.K.; Baron, T.H. A meta-analysis of endoscopic ultrasound-fine-needle aspiration compared to endoscopic ultrasound-fine-needle biopsy: Diagnostic yield and the value of onsite cytopathological assessment. *Endosc. Int. Open* **2017**, 5, E363–E375. [CrossRef]

21. Itoi, T.; Iokawa, F.; Kurihara, T.; Sofuni, A.; Tsuchiya, T.; Ishii, K.; Tsuji, S.; Ikeuchi, N.; Kawai, T.; Moriyasu, F. Experimental endoscopy: Objective evaluation of EUS needles. *Gastrointest. Endosc.* **2009**, 69, 509–516. [CrossRef] [PubMed]

22. Inoue, T.; Okumura, F.; Mizushima, T.; Nishie, H.; Iwasaki, H.; Anbe, K.; Ozeki, T.; Kachi, K.; Fukusada, S.; Suzuki, Y.; et al. Assessment of Factors Affecting the Usefulness and Diagnostic Yield of Core Biopsy Needles with a Side Hole in Endoscopic Ultrasound-Guided Fine-Needle Aspiration. *Gut Liver* **2016**, 10, 51–57. [CrossRef] [PubMed]

23. El Chafic, A.H.; Loren, D.; Siddiqui, A.; Mounzer, R.; Cosgrove, N.; Kowalski, T. Comparison of FNA and fine-needle biopsy for EUS-guided sampling of suspected GI stromal tumors. *Gastrointest. Endosc.* **2017**, 86, 510–515. [CrossRef] [PubMed]

24. De Moura, D.T.H.; McCarty, T.R.; Jirapinyo, P.; Ribeiro, I.B.; Flumignan, V.K.; Najdawai, F.; Ryou, M.; Lee, L.S.; Thompson, C.C. EUS-guided fine-needle biopsy sampling versus FNA in the diagnosis of subepithelial lesions: A large multicenter study. *Gastrointest. Endosc.* **2020**, 92, 108–119. [CrossRef]

25. Larghi, A.; Verna, E.C.; Ricci, R.; Seerden, T.C.; Galasso, D.; Carnuccio, A.; Uchida, N.; Rindi, G.; Costamagna, G. EUS-guided fine-needle tissue acquisition by using a 19-gauge needle in a selected patient population: A prospective study. *Gastrointest. Endosc.* **2011**, 74, 504–510. [CrossRef]

26. Yasuda, I.; Goto, N.; Tsurumi, H.; Nakashima, M.; Doi, S.; Iwashita, T.; Kanemura, N.; Kasahara, S.; Adachi, S.; Hara, T.; et al. Endoscopic ultrasound-guided fine needle aspiration biopsy for diagnosis of lymphoproliferative disorders: Feasibility of immunohistological, flow cytometric, and cytogenetic assessments. *Am. J. Gastroenterol.* **2012**, 107, 397–404. [CrossRef]

27. Lopes, C.V.; Hartmann, A.A.; Artifon, E.L.A. Eus-Fna with 19 or 22 Gauges Needles for Gastric Subepithelial Lesions of the Muscle Layer. *Arq. Bras. Cir. Dig.* **2018**, 31, e1350. [CrossRef]

28. Camellini, L.; Carlinfante, G.; Azzolini, F.; Iori, V.; Cavina, M.; Seneri, G.; Decembrino, F.; Gallo, C.; Tamagnini, I.; Valli, R.; et al. A randomized clinical trial comparing 22G and 25G needles in endoscopic ultrasound-guided fine-needle aspiration of solid lesions. *Endoscopy* **2011**, 43, 709–715. [CrossRef]

29. Gleeson, F.C.; Kipp, B.R.; Voss, J.S.; Campion, M.B.; Minot, D.M.; Tu, Z.J.; Klee, E.W.; Graham, R.P.; Lazaridis, K.N.; Henry, M.R.; et al. Frequency of mitogen-activated protein kinase and phosphoinositide 3-kinase signaling pathway pathogenic alterations in EUS-FNA sampled malignant lymph nodes in rectal cancer with theranostic potential. *Gastrointest. Endosc.* **2015**, 82, 550–556. [CrossRef]

30. Nishida, T.; Yoshinaga, S.; Takahashi, T.; Naito, Y. Recent Progress and Challenges in the Diagnosis and Treatment of Gastrointestinal Stromal Tumors. *Cancers* **2021**, 13, 318. [CrossRef]

31. Park, J.; Yoo, H.M.; Sul, H.J.; Shin, S.; Lee, S.W.; Kim, J.G. Genetic Characterization of Molecular Targets in Korean Patients with Gastrointestinal Stromal Tumors. *J. Gastric Cancer* **2020**, 20, 29–40. [CrossRef] [PubMed]