Isolated extramedullary cutaneous relapse despite concomitant severe graft-vs.-host disease and tissue chimeraism analysis in a patient with acute lymphoblastic leukemia after allogeneic hematopoietic stem cell transplantation: A case report

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Abstract. Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is a potentially curative treatment option for patients with acute lymphoblastic leukemia (ALL). The curative potential of allo-HSCT for ALL is, in part, due to the graft-vs.-leukemia (GVL) effect, in addition to the intensive conditioning chemo-radiotherapy. However, relapse remains the major cause of treatment failure following allo-HSCT for ALL. In the allo-HSCT setting, testing for genetic markers of hematopoietic chimeraism has become a part of the routine diagnostic program. Routine chimeraism analysis is usually performed in peripheral blood or bone marrow; in fact, little is known about the value of tissue chimeraism in patients with extramedullary relapse (EMR) after the allo-HSCT setting. The present study reports on, a case of a patient with ALL who experienced isolated cutaneous EMR despite ongoing graft-vs.-host disease (GVHD), and the results of peripheral blood and skin tissue chimeraism studies using multiplex polymerase chain reaction (PCR) of short tandem repeats (STR-PCR). The present case demonstrates that, although complete remission and/or chimeraism may be achieved in the bone marrow, chimeraism achieved at the tissue level, and the subsequent GVL effect, may be limited, despite concomitant severe GVHD following allo-HSCT. Our tissue chimeraism analysis results provide a good example of how skin tissue may be a ‘sanctuary’ site for effector cells of GVL, despite active GVHD and complete hematopoietic chimeraism.

Introduction

Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is a potentially curative treatment option for patients with acute lymphoblastic leukemia (ALL). The curative potential of allo-HSCT for ALL is, in part, due to the graft-vs.-leukemia (GVL) effect, in addition to the intensive conditioning chemo-radiotherapy. However, relapse remains the major cause of treatment failure following allo-HSCT for ALL (1). ALL relapse usually occurs in the bone marrow, although a significant rate of extramedullary relapse (EMR) following allo-HSCT has been reported either alone or concomitantly with bone marrow relapse. In general, the incidence of EMR is higher in patients with ALL compared with those with acute myeloid leukemia (AML). Ge et al (2) reported that the incidence of EMR was 12.9% in patients with ALL, in which isolated EMR occurred in 7.9% of them. However, the mechanism of EMR is very poorly understood; and the prognosis of these patients is generally poor (2).

In the allo-HSCT setting, testing for genetic markers of hematopoietic chimeraism has become a part of the routine diagnostic program. Chimeraism testing permits early prediction and documentation of successful engraftment, and facilitates early detection of impending graft rejection. For patients who undergo transplantation for the treatment of malignant hematological disorders, monitoring of chimeraism can provide an early indication of incipient disease relapse, and enable an
assessment of the ability to demonstrate the GVL effect to be made (3). Routine chimerism analysis is usually performed in peripheral blood or bone marrow; in fact, little is known about the value of tissue chimerism in patients with EMR following the allo-HSCT setting.

In the present study, a case of a patient with ALL who experienced isolated cutaneous EMR despite ongoing graft-vs.-host disease (GVHD), and the results of peripheral blood and skin tissue chimerism studies using multiplex polymerase chain reaction (PCR) of short tandem repeats (STR-PCR). This case demonstrates that, although complete remission and/or chimerism may be achieved in the bone marrow, chimerism achieved at the tissue level, and the subsequent GVL effect, may be limited, despite concomitant severe GVHD following allo-HSCT.

Case report

A 52-year-old female presented to a Turkish hospital with erythematous skin nodules on her trunk, arms and face with bilateral pleural effusion and hepatosplenomegaly in October 2012. The skin and bone marrow biopsy were consistent with precursor T-cell acute lymphoblastic leukemia. The patient was started on induction chemotherapy using the German multicenter acute lymphoblastic leukemia (GMALL) 05/93 protocol (4), and remission was achieved. The treatment was subsequently continued with early consolidation, reinduction and late consolidation treatments. However, skin lesions recurred in December 2013. The patient's bone marrow examination was clean at that time. Due to progressive disease, reinduction, chemotherapy with the identical protocol and skin-directed psoralen and ultraviolet A radiation (PUVA) treatment was started in January 2014. However, progression in the skin lesions occurred with this treatment, and a relapse was also evident in the bone marrow. In March 2014, treatment with a FLAG-ida chemotherapy regimen was started. The disease was also resistant to this chemotherapy, since remission was achieved in bone marrow, but the nodular skin lesions remained. The patient was then treated with two cycles of clofarabine in combination with high-dose cytosine arabinoside in May 2014. However, the disease was also refractory to this treatment, since, although the bone marrow was kept in remission, the nodular skin lesions returned at the time of hematological recovery. A matched related donor for allo-geeneic bone marrow transplantation was not identified for this patient, and so she was referred to our clinic at the Istanbul Medipol University for a matched unrelated donor (MUD) allo-HSCT in June 2014.

The patient underwent unmanipulated peripheral blood SCT from a 10/10 MUD on July 1, 2014. The myeloablative conditioning (MAC) regimen was with total body irradiation (TBI: 12 Gy in six fractions) and cyclophosphamide (120 mg/kg). Cyclosporin A, short-course methotrexate and standard dose anti-thymocyte globulin (ATG) were used for acute GVHD prophylaxis. Allo-HSCT was performed without any difficulties, and neutrophil and thrombocyte engraftment was achieved on day +16 of the transplantation. However, a diffuse erythematous rash appeared on the patient's trunk and bilateral extremities at the time of engraftment. The results of the liver function tests were also abnormal. Acute GVHD of the skin and liver was suspected, and a treatment with prednisolone started at a dose of 2 mg/kg/day. The skin lesions disappeared, and results of the liver function tests were normalized with this treatment. At day +63 of the transplantation, acute GVHD progressed, with a tapering of steroid doses. In view of this development, three doses of pulse steroid treatment (10 mg/kg/day) and mycophenolate mofetil (MMF) were added to the treatment. In the follow-up, acute GVHD had progressed again, despite pulse steroid and MMF treatment, and so β-human chorionic gonadotropin (β-hCG; pregnyl) was used at a dose of 187.5 mg/day for 2 weeks. However, on day +118, the skin lesions of GVHD had progressed, and the levels of the liver enzymes were again increased. Furthermore, diarrhea had been added to the patient's symptoms. The results of the previous skin and liver biopsies were consistent with GVHD. Consequently, extracorporeal photopheresis (ECPP) in combination with mesenchymal stem cell infusion was performed for the patient. With this treatment, the GVHD was able to be controlled successfully. At day +143 of the transplantation, new nodular lesions were appearing on the chest and torso of the patient (Fig. 1). The biopsy of these lesions was consistent with T-cell ALL infiltration. Notably, GVHD was still ongoing, and the bone marrow was still in remission at that time. Isolated EMR was considered, presenting with skin involvement. Nlerabine treatment was planned for the patient. However, the condition of the patient deteriorated with the infection, and she succumbed to hospital-acquired bactemia with sepsis at +200 days of allo-HSCT.

During the patient's follow-up, no sign of relapse in the bone marrow biopsy examinations of the patient was observed. The peripheral blood STR-PCR chimerism results were all consistent with complete chimerism at days +30, +90 and +180, which supported the bone marrow remission (Table I). STR-PCR chimerism analysis in paraffin-embedded skin biopsies at days +16, +63, +143 and +198 were also retrospectively performed. The skin biopsies that were taken at days +16, +63 and +198 were from the areas of skin GVHD, whereas the biopsy at day +143 was taken directly from the nodular skin lesion of leukemia relapse. It is noteworthy that the chimerism status was not chimeric (recipient type) at days +16 and +63, but it was mixed chimeric at days +143 and +198 (Table II).

Discussion

The case presented in the present study details a heavily pretreated patient with high-risk features, who underwent an allo-HSCT and suffered from a very aggressive course of disease that progressed rapidly, to be controlled by a very active immune response.

The vast majority (up to 90%) of adult patients with ALL are able to achieve remission with the use of intensive induction chemotherapy. However, relapse eventually occurs, and the 5 year overall survival (OS) is 42-63% for adolescents and young adults, 24.1% for patients between the ages of 40-59, and 17.7% for patients between the ages of 60-69 (5). Historically, age, a high leukocyte count at presentation, poor response to treatment, T-cell immunophenotype, cytogenetic profile of the disease and extranodal presentation of the disease have been considered adverse clinical prognostic factors in adult ALL. In relapsed patients, long-term disease-free survival and cure...
may be obtained in 7-24% of patients with only allo-HSCT (6). Cutaneous involvement with ALL occurs rarely, with an incidence between 0.5-3% (7).

For this reason, allo-HSCT, despite its limitations and toxicities, is an accepted therapy for adults. In addition to historical prognostic factors, certain factors, including the performance status of the patient, availability and type of a transplant donor, persistence of residual disease at transplantation, the cytomegalovirus seropositivity of the recipient, the type of conditioning regimen, using a T-cell-depleted graft, and the presence and grade of GVHD during transplantation, were defined as the predictors of allo-HSCT outcome (8-12). In our patient, the age at presentation, T-cell phenotype of the disease, extranodal involvement, refractoriness to conventional chemotherapy and absence of a donor in first complete remission (CR1) were the most important predictors of relapse. In contrast, the performance status was good in our patient, therefore it was possible to perform a MAC allo-HSCT without a T-cell-depleted graft.

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Table I. Results of peripheral blood chimerism studies using PCR-STR analysis.

| STR locus | Donor (pre-Tx) | Recipient (post-Tx +30) | Recipient (post-Tx +90) | Recipient (post-Tx +180) |
|-----------|----------------|-------------------------|-------------------------|-------------------------|
| D21S11    | 28,32.2        | 28,32.2                 | 28,32.2                 | 28,32.2                 |
| D7S820    | 10,10          | 10,10                   | 10,10                   | 10,10                   |
| CSF1PO    | 9,10           | 9,10                    | 9,10                    | 9,10                    |
| D3S1358   | 15,18          | 15,18                   | 15,18                   | 15,18                   |
| TH01      | 6,9            | 6,9                     | 6,9                     | 6,9                     |
| D13S317   | 12,14          | 12,14                   | 12,14                   | 12,14                   |
| D16S539   | 9,11           | 9,11                    | 9,11                    | 9,11                    |
| D2S1338   | 24,24          | 24,24                   | 24,24                   | 24,24                   |
| D19S433   | 13,2,14        | 13,2,14                 | 13,2,14                 | 13,2,14                 |
| VWA       | 14,17          | 14,17                   | 14,17                   | 14,17                   |
| TPOX      | 11,12          | 11,12                   | 11,12                   | 11,12                   |
| D18S51    | 12,14          | 12,14                   | 12,14                   | 12,14                   |
| D5S818    | 11,11          | 11,11                   | 11,11                   | 11,11                   |
| FGA       | 19,21          | 19,21                   | 19,21                   | 19,21                   |

Result of PCR-STR analysis: Complete chimeric, Complete chimeric, Complete chimeric, Complete chimeric.

Pre-Tx, pre-transplant; post-Tx, post-transplant; PCR-STR, polymerase chain reaction-short tandem repeats.

Table II. Result of skin tissue chimerism studies by PCR-STR analysis.

| STR locus | Donor (pre-Tx) | Recipient (post-Tx +16) | Recipient (post-Tx +63) | Recipient (post-Tx +143) | Recipient (post-Tx +198) |
|-----------|----------------|-------------------------|-------------------------|-------------------------|-------------------------|
| D3S1358   | 15,18          | 16,18                   | 16,18                   | 16,18,15,18             | 16,18,15,18             |
| TH01      | 6,9            | 7,7                     | 7,7                     | 7,7,6,9                 | 7,7,6,9                 |
| D21S11    | 28,29          | 29,31                   | 29,31                   | 29,31,28,29             | 29,31,28,29             |
| D18S51    | 22,22          | 25,26                   | 25,26                   | 25,26,22,22             | 25,26,22,22             |
| PENTA-E   | 15,17          | 7,15                    | 7,15                    | 7,15,15,17              | 7,15,15,17              |
| D55S18    | 11,11          | 11,12                   | 11,12                   | 11,12,11,11             | 11,12,11,11             |
| D13S317   | 11,11          | 11,12                   | 11,12                   | 11,12,11,11             | 11,12,11,11             |
| D7S820    | 10,10          | 9,10                    | 9,10                    | 9,10,10,10              | 9,10,10,10              |
| D16S539   | 9,11           | 8,13                    | 8,13                    | 8,13,9,11               | 8,13,9,11               |
| CSF1PO    | 6,13           | 7,7                     | 7,7                     | 7,7,6,13                | 7,7,6,13                |
| PENTA-D   | 10,10          | 10,13                   | 10,13                   | 10,13,10,10             | 10,13,10,10             |
| VWA       | 14,14          | 16,18                   | 16,18                   | 16,18,14,14             | 16,18,14,14             |
| TPOX      | 11,11          | 8,10                    | 8,10                    | 8,10,11,11              | 8,10,11,11              |
| FGA       | 21,21          | 24,25                   | 24,25                   | 24,25,21,21             | 24,25,21,21             |

Result of PCR-STR analysis: Not chimeric, Not chimeric, Mixed chimeric, Mixed chimeric.

Pre-Tx, pre-transplant; post-Tx, post-transplant; PCR-STR, polymerase chain reaction-short tandem repeats.
These observations suggested that the GVL effect at extra-medullary sites may be less prominent compared with that in the bone marrow. However, other factors, including the nature of the leukemic blasts, status of acute leukemia at hematopoietic cell transplantation, and the conditioning regimen, may also influence the frequency of EMR. In the literature, central nervous system (CNS) relapse is the most common subtype of EMR after allo-HSCT. Patients with high-risk cytogenetics, an advanced disease status, a history of EM leukemia prior to allo-HSCT, hyperleukocytosis at diagnosis, receipt of peripheral blood stem cells (PBSCs) and the male gender are reported risk factors for EMR following allo-HSCT (2). Accordingly, the cutaneous leukemic involvement of our patient was problematical from the beginning of the disease. Relapse occurred despite intensive chemotherapy and ongoing GVHD. Unfortunately, the prognosis for EMR following allo-HSCT is poor, and efficient treatment strategies are lacking in this setting. Radiotherapy, salvage chemotherapy, second transplantation and donor lymphocyte infusion (DLI) have all been utilized, but the choice of the therapeutic strategy that is optimal remains controversial. DLI was not a treatment option for our patient due to the active GVHD.

Among recipients of allo-HSCT, donor stem cell engraftment in non-hematopoietic tissues has been observed by several groups. Graft-derived cells with an epithelial phenotype have been described in skin, the gastrointestinal tract, and in liver of human allo-HSCT recipients (20). However, the exact origin of these cells and their pattern of engraftment in response to injury have yet to be elucidated. In this regard, Willemze et al (21) investigated the occurrence of endothelial and epithelial cell chimerism skin biopsies of allo-HSCT recipients using the fluorescence in situ hybridization (FISH) method. Endothelial cell chimerism was found in 25% of the biopsies, and increased in time, particularly in patients with acute GVHD. Epithelial cell chimerism was found in 85% of the biopsies, and was not correlated with the time interval following SCT or with tissue damage caused by GVHD (21). These results contrasted with observations made by Murata et al (22), who observed endothelial cell chimerism (using FISH method) shortly after the start of acute GVHD. However, in the study by Imanishi et al (23) published during the same year, donor-derived DNA in the fingernails of allo-HSCT recipients was identified only in 9 of 21 cases using the STR-PCR method. The time from transplantation to sampling was in excess of 300 days in that study (23). In support of this study, Pearce et al (24) also reported donor chimerism in four of eight cases in the fingernails of reduced intensity conditioning regimen (RIC) allo-HSCT patients using the STR-PCR method. It should be emphasized that the latter two studies investigated the contribution of donor-derived cells in fingernails, i.e. a tissue without blood cells. In our patient, STR-PCR chimerism analysis was performed in paraffin-embedded skin biopsies that contained blood cells. The chimerism status was not chimeric (recipient type) at days +16 or +63 following the transplantation, until cutaneous EMR occurred on day +143. This finding emphasized that complete chimerism was achieved in the hematopoietic tissue: The skin of our patient was a ‘sanctuary’ site for GVL effector cells up to day +63. This result supports a previously published report by our group (25). In the present study, the mixed chimerism obtained at day +143 may be associated with either an immune booster reaction secondary to the leukemia relapse, or it may
be associated with the natural course of chimerism achieved at the tissue level. In neither situation was it sufficient to control the disease in our patient.

In conclusion, the present case study has presented a patient with ALL with isolated cutaneous EMR despite active severe GVHD. Our tissue chimerism analysis results have illustrated a good example that skin tissue may be a ‘sanctuary’ site for effector cells of GVL, despite active GVHD and complete hematopoietic chimerism.

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