Value-added uses for crude glycerol—a byproduct of biodiesel production

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discussion.


to the viability of biodiesel. In this review, composition and quality factors of crude glycerol are discussed. The value-added utilization opportunities of crude glycerol are reviewed. The majority of crude glycerol is used as feedstock for production of other value-added chemicals, followed by animal feeds.

**Keywords:** Crude glycerol, Value-added utilization

**Background**

The dramatic increase in demand for transportation fuels and the increase in environmental concerns, coupled with diminishing crude oil reserves, have increased the emphasis on renewable energy. Biodiesel, one of the promising alternative and renewable fuels, has been viewed with increasing interest and its production capacity has been well developed in recent years. Although world biodiesel production was expected to reach a high capacity, in fact, it is less than the anticipated target and has increased at a slower rate [1]. The main reason is its relatively high production cost. Utilization of the glycerol co-product is one of the promising options for lowering the production cost.

Biodiesel production will generate about 10\% (w/w) glycerol as the main byproduct. In other words, every gallon of biodiesel produced generates approximately 1.05 pounds of glycerol. This indicates a 30-million-gallon-per-year plant will generate about 11,500 tonnes of 99.9 percent pure glycerin. It was projected that the world biodiesel market would reach 37 billion gallons by 2016, which implied that approximately 4 billion gallons of crude glycerol would be produced [2]. Too much surplus of crude glycerol from biodiesel production will impact the refined glycerol market. For example, in 2007, the refined glycerol’s price was painfully low, approximately $0.30 per pound (compared to $0.70 before the expansion of biodiesel production) in the United States. Accordingly, the price of crude glycerol decreased from about $0.25 per pound to $0.05 per pound [3]. Therefore, development of sustainable processes for utilizing this organic raw material is imperative.

Since purified glycerol is a high-value and commercial chemical with thousands of uses, the crude glycerol presents great opportunities for new applications. For that reason, more attention is being paid to the utilization of crude glycerol directly, review papers on crude glycerol utilization are scarce. This review mainly addresses the current and potential value-added applications of crude glycerol from biodiesel production.

**Chemical compositions of crude glycerol**

The chemical composition of crude glycerol mainly varies with the type of catalyst used to produce biodiesel, the transesterification efficiency, recovery efficiency of the biodiesel, other impurities in the feedstock, and whether the methanol and catalysts were recovered. All of these considerations contribute to the composition of the crude glycerol fraction. For instance, Hansen et al. [4] studied the chemical compositions of 11 crude glycerol collected from 7 Australian biodiesel producers and indicated that the glycerol content ranged between 38\%
and 96%, with some samples including more than 14% methanol and 29% ash. Such variations would be expected with small conversion facilities. In most cases, biodiesel production involves the use of methanol and a homogeneous alkaline catalyst, such as sodium methoxide and potassium hydroxide. Accordingly, methanol, soap, catalysts, salts, non-glycerol organic matter, and water impurities usually are contained in the crude glycerol. For example, crude glycerol from sunflower oil biodiesel production had the following composition (w/w): 30% glycerol, 50% methanol, 13% soap, 2% moisture, approximately 2-3% salts (primarily sodium and potassium), and 2-3% other impurities [5]. Moreover, while the same feedstocks were employed, the crude glycerol from alkali- and lipase-catalyzed transesterifications contained different purities of glycerol [6]. The salt content in crude glycerol, from biodiesel production via homogeneous alkaline catalysts, ranged from 5% to 7% [7] which makes the conventional purification techniques more costly. Heterogeneous processes using enzymes and solid metal-oxide catalysts have been promoted as good alternatives to homogeneous alkaline catalysts in terms of improving the quality of crude glycerol. However, even in heterogeneous transesterification processes, impurities existing in the natural raw feedstocks tend to accumulate in the glycerol phase. Therefore, purification of crude glycerol is required, in most cases, to remove impurities in order to meet the requirements of existing and emerging uses.

Value-added opportunities for crude glycerol

Worldwide, crude glycerol derived from biodiesel conversion has increased from 200, 000 tonnes in 2004 [8] to 1.224 million tonnes in 2008 [1]. Meanwhile, the global market for refined glycerol was estimated to be roughly 900, 000 tonnes in 2005 [9]. Therefore, it is of great importance for scientists to find new applications for refined and crude glycerol. Recently, numerous papers have been published on direct utilization of crude glycerol from biodiesel production. In the following sections, detailed discussions on utilization of crude glycerol are presented.

Animal feedstuff

Using glycerol as a feed ingredient for animals dates back to the 1970s [10]. However, glycerol’s utilization in feeds has been limited by the availability of glycerol [3]. Recently, the possibilities of using crude glycerol from biodiesel in feeds have been investigated because of the increase in the price of corn and the surplus of crude glycerol.

Crude glycerol in non-ruminant diets

Glycerol has high absorption rates and is good energy source. Once absorbed, it can be converted to glucose for energy production in the liver of animals by the enzyme glycerol kinase [3]. Crude glycerol samples, from different biodiesel producers, were analyzed as energy sources. The digestible energy (DE) values for 85% of the crude glycerol samples were in the range of 14.9-15.3 MJ/kg with metabolizable energy (ME) values in the range of 13.9-14.7 MJ/kg [11]. Crude glycerol was an excellent source of calories for non-ruminants, for example, the ME determined in broilers, laying hens and swine were 15.2, 15.9 and 13.4 MJ ME/kg, respectively [3]. In growing pigs and laying hens, 14.0 MJ/kg apparent DE [12] and 15.9 MJ/kg nitrogen-corrected apparent ME (AMEn) [13] were reported, respectively, which implied that crude glycerol was used efficiently. The AMEn of crude glycerol was metabolized efficiently by broiler chickens with an AMEn of 14.4 MJ/kg. That was very similar to the general energy (GE) of 15.2 MJ/kg [14]. In nursery pigs, the GE concentration of crude glycerol depended on the concentration of glycerol, methanol, and fatty acids, with ME as a percent of GE averaging 85.4% [15].

Although crude glycerol can be added to animal feed, excess glycerol in the animal diet may affect normal physiological metabolism. A few manuscripts have been published that focused on the levels of crude glycerol fed and the performance of crude glycerol in animal feeds. The improvement of daily gains by pigs depended on the actual intake of glycerol during the growing period but not on the finishing period. The dietary treatments had no significant effects on meat quality [16,17]. When crude glycerol was added to the diets of weaned pigs, at levels up to 10%, the feed performance was enhanced [18]. Up to 9% crude glycerol could be added to the diets of lactating sows with performances similar to sows fed standard corn-soybean meal control diets [19]. No detrimental effects, with respect to egg performance, egg quality, nutrient retention, and metabolizable energy, were found when crude glycerol was incorporated at a level of 6% in the diet of laying hens [20]. In broiler diets, increasing the intake level of crude glycerol increased feed conversion ratio but did not affect growth performance and nutrient digestibility [21]. Crude glycerol was used effectively at levels of 2.5 or 5%. But the use of 10% crude glycerol resulted in poor feed flow. The influences of levels and quality of crude glycerol on pellet quality needs further study [22].

Crude glycerol in ruminant diets

Crude glycerol, added at up to 15% dry matter in the diets of finishing lambs, could improve feedlot performance, especially during the first 14 d, and had no associated effect on carcass characteristics [23]. Compared to medium-quality hay, diets for meat goats with up to 5% crude glycerol proved to be beneficial [24]. In addition, the inclusion of purified glycerol at up to 15% of the dry
matter of lactating dairy cows was possible, without deleterious effects on feed intake, milk production, and yield [25,26]. When crude glycerol was added at levels of 8% or less, based on dry matter in cattle finishing diets, it improved weight gain and feed efficiency [27]. Apart from the above mentioned investigations, a patent described approaches for using or incorporating crude glycerol into animal feeds as well as feeding recommendations [28].

In all, the use of crude glycerol as an animal feed component has great potential for replacing corn in diets, and is gaining increasing attention. However, one must be aware of the presence of potential hazardous impurities in crude glycerol from biodiesel. For example, residual levels of potassium may result in wet litter or imbalances in dietary electrolyte balance in broilers [22]. The levels of methanol must be minimized because of its toxicity [3,12,22,26]. More attention should be paid to the crude glycerol from small scale biodiesel facilities that use simple batch distillation or evaporation techniques.

Feedstocks for chemicals

Chemicals produced via biological conversions

1, 3-propanediol The anaerobic fermentative production of 1,3-propanediol is the most promising option for biological conversion of glycerol. Mu et al. [6] demonstrated that crude glycerol could be used directly for the production of 1,3-propanediol in fed-batch cultures of Klebsiella pneumoniae. The differences between the final 1,3-propanediol concentrations were small for crude glycerol from the methanolation of soybean oil by alkali (51.3 g/L) and lipase-catalysis (53 g/L). This implied that the composition of crude glycerol had little effect on the biological conversion and a low fermentation cost could be expected. Further, the production of 1,3-propanediol by K.pneumoniae was optimized using response surface methodology. The maximum yield of 1,3-propanediol was 13.8 g/L [29]. More recently, the production of 1,3-propanediol, from crude glycerol from Jatropha biodiesel by K.pneumoniae ATCC 15380, was optimized. The obtained 1,3-propanediol yield, purity and recovery were 56 g/L, 99.7% and 34%, respectively [30]. Additionally, an incorporated bioprocess that combined biodiesel production by lipase with microbial production of 1,3-propanediol by K.pneumoniae was developed in a hollow fiber membrane. The bioprocess avoided glycerol inhibition on lipase and reduced the production cost [31].

Clostridium butyricum also could be used to produce 1,3-propanediol from crude glycerol. For example, C. butyricum VPI 3266 was able to produce 1,3-propanediol from crude glycerol on a synthetic medium. Trivial differences were found between commercial glycerol and crude glycerol [32]. C. butyricum strain F2b and C. butyricum VPI 1718 potentially could convert crude glycerol to 1,3-propanediol [33-35]. In order to avoid the isolation of 1,3-propanediol from crude glycerol fermentation media, a one vessel bio- and chemo-catalytic process was developed to convert crude glycerol to secondary amines directly in a biphasic system without intermediate separation of 1,3-propanediol [36]. Additionally, Chatzifragkou et al. [37] studied the effects of different impurities in crude glycerol on 1,3-propanediol production by C. butyricum. The double bond from long-chain fatty acids or methyl esters might influence the growth performance of the microorganism, methanol did not affect the microbial conversion and the presence of NaCl had certain effect during a continuous process but not a batch process.

Citric acid A few reports are available on the use of crude glycerol for citric acid biosynthesis. The production of citric acid from crude glycerol by Yarrowia lipolytica ACA-DC 50109 was not only similar to that obtained from sugar-based conventional media [33] but also single-cell oil and citric acid were produced simultaneously [34,38]. When a fed-batch fermentation by acetate-negative mutants of Y. lipolytica Wratislavia AWG7 strain was used to ferment crude glycerol, the final concentration of citric acid was 131.5 g/L, similar to that obtained from pure glycerol (139 g/L). On the other hand, when Y. lipolytica Wratislavia K1 was used, a lower concentration of citric acid (about 87-89 g/L) and a high concentration of erythritol (up to 47 g/L) were obtained [39]. It was in line with the results shown by Rymowicz et al. [40]. Further, Y. lipolytica Wratislavia K1 proved to be superior to other strains by producing erythritol and not citric acid from crude glycerol under optimal conversion conditions, which may be a valuable development [41]. Y. lipolytica LGAM S (7)1 also showed potential for converting crude glycerol to citric acid [42]. More recently, it was reported that Y. lipolytica N15 could produce citric acid in high amounts, specifically, up to 98 g/L of citric acid and 71 g/L of citric acid were produced from pure glycerol medium and crude glycerol medium, respectively [43].

Hydrogen and other lower molecule fuels The bacterium Rhodopseudomonas palustris was capable of photofermentative conversion of crude glycerol to hydrogen. Nearly equal productions were obtained from crude glycerol and pure glycerol. Up to 6 moles H2 per mole glycerol were obtained, which was 75% of theoretical. Both rates and yields of hydrogen production could be modified by changing the concentration of added nitrogen. However, some technical obstacles, such as enhancing the efficiency of light utilization by the organisms and developing effective photobioreactors, still need to be solved during development of a practical process [44]. When Enterobacter aerogenes HU-101 was employed,
hydrogen and ethanol were produced at high yields and with high production rates. But the crude glycerol should be diluted with a synthetic medium in order to increase the rate of glycerol utilization [45]. For maximizing hydrogen production, Jitrwung and Yargeau [46] optimized some media compositions of *E. aerogenes* ATCC 35029 fermented crude glycerol process. More recently, it was reported that *K.pneumoniae mutant* strain and nonpathogenic *Kluvera cryocrescens* S26 were promising for producing ethanol from crude glycerol [47,48]. In addition, crude glycerol, as a co-substrate, could be used to enhance hydrogen and especially methane production during the anaerobic treatment of different feedstocks including the organic fraction of municipal solid wastes, sewage sludge and slaughterhouse wastes [49-51].

**Poly (hydroxyalkanoates)** Poly(hydroxyalkanoates) (PHA) represent a complex class of naturally occurring bacterial polyesters and have been recognized as good substitutes for non-biodegradable petrochemically produced polymers. Ashby et al. [52] reported that crude glycerol could be used to produce PHA polymer. PHB is the most-studied example of biodegradable polyesters belonging to the group of PHA. The study of the feasibility of using crude glycerol for PHB production, with *Paracoccus denitrificans* and *Cupriavidus necator* JMP 134, showed that the resulting polymers were very similar to those obtained from glucose. But the PHB production decreased significantly when NaCl-contaminated crude glycerol was used. The authors suggested that the harmful effect of the NaCl-contaminant could be reduced by mixing crude glycerol from different manufacturers [53]. Further, a process based on the *Cupriavidus necator* DSM 545 fermentation of crude glycerol was designed for the large-scale production of PHB. However, sodium still hindered the cell growth [54]. *Zobellella denitrificans* MW1 could utilize crude glycerol for growth and PHB production to high concentration, especially in the presence of NaCl. Therefore, it was recommended as an attractive option for large-scale production of PHB with crude glycerol [55].

Additionally, when mixed microbial consortia (MMC) was used for PHA production from crude glycerol, it was found that methanol in the crude glycerol was transformed to PHB by MMC. Further, it was estimated that a 10 million gallon per year biodiesel plant would have the potential of producing 20.9 ton PHB [56]. More recent report showed that *Pseudomonas oleovorans* NRRL B-14682 could also be used for PHB production from crude glycerol [57].

**Docosahexaenoic acid** A series of papers on the production of docosahexaenoic acid (DHA)-rich algae were published, using crude glycerol, by fermentation of the alga *Schizochytrium limacinum*. For supporting alga growth and DHA production, 75-100 g/L concentration of crude glycerol was recommended as the optimal range. The algal DHA yield was influenced significantly by temperature and ammonium acetate concentration. The optimal amounts for temperature and ammonium acetate were 19.2°C and 1.0 g/L, respectively. The highest DHA yield obtained was 4.91 g/L under the optimized culture conditions [58]. Different sources of crude glycerol did not result in significant variations in algal biomass compositions. The resulting algae had a similar content of DHA and a comparable nutritional profile to commercial algal biomass. That proposed good potential for using crude glycerol-derived algae in omega-3-fortified foods or feeds [59]. Further, DHA-containing algae have been developed as replacements for fish oil for omega-3 fatty acids [60]. Crude glycerol was used to produce fungal biomass that served as eicosapentaenoic acid (EPA)-fortified foods or feeds through fungal fermentation with fungus *Pythium irregulare*. Growing in medium containing 30 g/L crude glycerol and 1.0 g/L yeast extract, the EPA yield and productivity could reach 90 mg/L and 14.9 mg/L per day, respectively. The resulting EPA content was low compared to microalgae for EPA. Optimizing culture conditions and developing high cell density culture techniques are imperative in future work [60]. Recently, it was reported that continuous culture was an effective approach for studying the growth kinetics and behaviors of the algae on crude glycerol [61].

**Lipids** As the sole carbon source, crude glycerol could be used to produce lipids which might be a sustainable biodiesel feedstock. For example, crude glycerol could be used for culturing *Schizochytrium limacinum* SR21 and *Cryptococcus curvatus*. *S. limacinum* algal growth and lipid production were affected by the concentrations of glycerol. Higher concentrations of glycerol had negative effects on cell growth. For batch culturing of crude glycerol derived from yellow grease, the optimal glycerol concentrations for untreated and treated crude glycerol were 25 and 35 g/L, respectively. With 35 g/L, the obtained highest cellular lipid content was 73.3%. Methanol remaining in crude glycerol could harm *S. limacinum* SR21 growth [62]. For *C. curvatus* yeast, fed-batch was a better process than batch for lipid production. Culturing for 12 days, the lipid content from one-stage fed-batch operation and two-stage fed-batch process were 44.2% and 52%, respectively. Methanol did not have significant inhibitory effect on cell growth. The produced lipid had high concentration of monounsaturated fatty acid and was good biodiesel feedstock [63].

Further, Saenge et al. [64] presented that oleaginous red yeast *Rhodotorula glutinis* TISTR 5159, cultured on crude glycerol, produced lipids and carotenoids. The addition of ammonium sulfate and Tween 20 increased the accumulation of lipids and carotenoids. When
fed-batch fermentation was employed, the highest lipid content, lipid yield and carotenoids production were 10.05 g/L, 60.7% and 6.10 g/L, respectively. *Chlorella protothecoides* also converted crude glycerol to lipids. The lipids yield was 0.31 g lipids/g substrate [65]. Similarly, with *C. protothecoides* and crude glycerol (62% purity), Chen and Walker [66] demonstrated that the maximum lipid productivity of 3 g/L per day was obtained in a fed-batch operation, which was higher than that produced by batch process. Additionally, Chatzifragkou *et al.* [67] studied the potential of fifteen eukaryotic microorganisms to convert crude glycerol to metabolic products. The results showed that yeast accumulated limited lipids (up to 22 wt.%, wt/wt, in the case of *Rhodotorula* *sp*), while fungi accumulated higher amounts of lipids in their mycelia (ranging between 18.1 and 42.6%, wt/wt, of dry biomass).

**Other chemicals** Beyond the chemicals mentioned above, several other processes for producing useful chemicals from crude glycerol via biotransformations have been developed. A continuous cultivation process and a recently isolated bacterium *Basfia succiniciproducens* DD1 were identified for succinic acid production. The process was characterized as having great process stability, attractive production cost, and impossible pathogenicity of the production strain, but the final production strain needs to be examined further for commercial succinic acid production [68]. Via simulation method, Vlysidis *et al.* [69] showed that the succinic acid co-production from crude glycerol, for a 20 years biodiesel plant, would improve the profit of the overall biorefinery by 60%.

Further, crude glycerol, as the sole carbon source, had the potential of producing phytase in industrial scale in high cell density fermentations with recombinant *Pichia pastoris* possessing a pGAP-based constitutive expression vector [70] and producing butanol with *Clostridium pasteurianum*. The highest yield of butanol was 0.30 g/g, which was significantly higher than the 0.15-0.20 g/g butanol yield typically obtained during the fermentation of glucose using *Clostridium acetobutylicum*. However, further understanding and optimizing of the process are still needed. It remained unclear what impact the impurities in crude glycerol would have on the solvent formation [71]. Similarly, crude glycerol could be used in a bioprocess with *P. pastoris* without any purification. Canola oil-derived crude glycerol was the most favorable carbon source and showed great potential for the production of additional value-added products such as the recombinant human erythropoietin and cell growth [72]. Crude glycerol also could be economic carbon and nutrient sources for bacterial cellulose (BC) production. The BC amount obtained was about 0.1 g/L after 96 h incubation. The addition of other nutrient sources (yeast extract, nitrogen and phosphate) to crude glycerol culture media increased the BC production by ~200% [73]. Additionally, *Gluconobacter* sp. NBRC3259 could be used to produce glyceric acid from crude glycerol with an activated charcoal pretreatment. 49.5 g/L of glyceric acid and 28.2 g/L dihydroxyacetone were produced from 174 g/L of glycerol [74]. When *Staphyloccocus caseolyticus* EX17 was employed, crude glycerol could be used for solvent tolerant lipase production [75]. More recently, it was reported that *Ustilago maydis* was a good biocatalyst for converting crude glycerol to glycolipid-type biosurfactants and other useful products [76]. Fungal protein, *Rhizopus microsporus var. oligosporus*, production on crude glycerol was another potential use of crude glycerol. The obtained fungal biomass contained high amounts of threonine and could be co-fed with commercial sources. But feeding formulation need be further studied [77].

**Chemicals produced through conventional catalytic conversions**

**Oxygcnated chemicals** As fuel additive, the oxygenate synthesized compound, (2,2-dimethyl-1,3-dioxolan-4-yl) methyl acetate, could be produced from crude glycerol and used as a biodiesel additive. It could improve biodiesel viscosity and could meet the requirements established for diesel and biodiesel fuels by the American and European Standards (ASTM D6751 and EN 14214, respectively) for flash point and oxidation stability. This new compound could compete with other biodiesel additives [78]. Further, acrolein is an important starting chemical for producing detergents, acrylic acid ester and super absorber polymers. Sereshki *et al.* [79] reported a process involving adding liquid crude glycerol directly into a fluidized bed reactor, vaporizing it, and then reacting them to produce acrolein over a tungsten doped zirconia catalyst. In this process, glycerol evaporated in the fluidized bed reactor leaving behind salt crystals which were only loosely bound to the surface and could be separated from the catalyst using mechanical agitation. This process has the potential to reduce the accumulation of salt in the reactor.

**Hydrogen or syngas** Crude glycerol was proven to be a viable alternative for producing hydrogen or syngas [80,81]. Gasification was the main employed technique. Thermo-gravimetry coupled with FTIR spectroscopy analysis proved that the thermal decomposition mechanism of crude glycerol mainly involved four phases and CO₂, H₂, CH₄, and CO were the main gas products [82]. Gasification with in situ CO₂ removal was effective and had high energy efficiency [83]. Supercritical water gasification of crude glycerol was performed under uncatalyzed and alkaline catalyzed conditions. The reaction temperature determined the decomposition degree. When NaOH was employed as catalyst nearly 90 vol.%
of the product gas was \( \text{H}_2 \) and no char was produced [84]. Additionally, co-gasification of crude glycerol and hardwood chips, in a downdraft gasifier, was another promising option for utilizing crude glycerol. The loading amount of crude glycerol had significant influence on \( \text{CO} \) and \( \text{CH}_4 \) concentration, while having no effect on \( \text{H}_2 \) and \( \text{CO}_2 \) yield. The study suggested that the co-gasification could perform well in downdraft gasifiers with hardwood chips mixing with liquid crude glycerol up to 20 wt.% [85].

**Other chemicals from conventional catalytic conversions** Addition to the previous mentioned chemicals produced from crude glycerol, several other chemicals from conventional catalytic conversion processes were reported. Monoglycerides could be produced via glycerolysis of triglyceride with crude glycerol. A two-step process, involving the removal of residual glycerol and crystallization, was employed for purification of the monoglycerides produced from glycerolysis. An approximate 99% purity monopalmitin was achieved [86]. More recently, the glycerolysis of soybean oil, using crude glycerol, was investigated. The amount of inorganic alkaline catalyst in the crude glycerol affected the concentration of the produced monoglycerides. Crude glycerol containing lower content of inorganic catalyst (about 2.7 wt.% \( \text{NaOH} \)) produced the highest concentration of monoglycerides, which was about 42% [87].

**Other uses**

Beyond the aforementioned uses, a few other potential applications of crude glycerol have been reported. Crude glycerol was used as a high-boiling-point organic solvent to enhance enzymatic hydrolysis of lignocellulosic biomass during atmospheric autocatalytic organosolv pretreatment [88]. Crude glycerol (without any purification) also could be used as a green solvent for organic reactions. Two representative reactions were base catalyzed aldol condensation and palladium catalyzed Heck carbon-carbon coupling [89]. Functioning as an organic carbon source for the removal of nitrate in the wastewater denitrification process, the denitrification efficiency could be increased by 2.0-5.0 mg \( \text{NO}_3^- \) L\(^{-1}\) per crude glycerol dose of 100 L [90]. Additionally, crude glycerol could be used as a fuel for generating electricity from microbial fuel cells [91]. Both co-hydrothermal pyrolysis and co-liquefaction of manure with crude glycerol could improve the production yield of bio-oil. However, for the co-liquefaction, too much addition of crude glycerol affected the carbon content and heat value of the bio-oil [92,93].

**Conclusions**

Effective utilization of crude glycerol is very crucial to the commercialization and further development of biodiesel production. In the long term, utilization of biomass-derived glycerol will not only contribute to reducing society’s dependence on nonrenewable resources but also will promote the development of integrated biorefineries.

This review addresses the value-added opportunities for crude glycerol from biodiesel production, mainly as a feedstuff for animal feed and as feedstocks for chemicals. Promising results have been achieved by researchers, especially for non-ruminant animals such as pigs, laying hens, and broilers. However, there still are some considerations that need to be taken into account for broad-scale use of this biomass-derived chemical in animal feeds. First of all, the chemical composition of crude glycerol varies significantly with the methods and feedstocks used to produce biodiesel. Since different researchers have used different qualities of crude glycerol for their studies, animal producers need to pay attention when they decide to include crude glycerol as a component of animal feed rations. Secondly, to some extent the impurities in crude glycerol affect feed performance. Finally, the amount of crude glycerol included in feed formulations needs to be considered. The establishment of a crude glycerol feed specification is recommended. The resulting “standard” crude glycerol would have a higher value simply because it would be consistent from all producers.

Conventional catalysis and biotransformation are the two main routes available for converting crude glycerol to a variety of value-added chemicals. In more recent years, there have been extensive studies, and some encouraging results, on processes for crude glycerol conversion. For example, the productions of 1,3-propanediol, citric acid, poly (hydroxyalkanoates), butanol, hydrogen, docosahexaenoic acid-rich algae, monoglycerides, lipids and syngas from crude glycerol are promising. However, many of the technologies offered still need further development to make them cost-effective and operationally feasible for incorporation into biorefineries.

Impurities in crude glycerol can greatly influence the conversion of glycerol into other products. In some biological conversion processes, pollutants in crude glycerol, inhibit cell and fungal growth and result in lower production rates and product yields (compared with pure or commercial glycerol under the same culture conditions). On the other hand, for conventional catalytic conversions, impurities poison the catalysts, increasing char production and influencing product yield. Many of the technologies need to be more fully understood and optimized, such as optimizing reaction parameters, production yields, and fermentation conditions, developing mutant strains and efficient bioreactors for stable cultures, and improving the activity and selectivity of catalysts. Additionally, cell growth has been shown to be inhibited during fermentation when the initial crude glycerol concentration was high. Conversely, higher crude
| Product                        | Pathway                                                                 | Product productivity | Reference |
|-------------------------------|-------------------------------------------------------------------------|----------------------|------------|
| 1, 3-propanediol              | Fed-batch cultures of *Klebsiella pneumoniae* strain                    | 1.7 g/L/h            | [6]        |
|                               | Maximum 1,3-propanediol production from *K. pneumonia*                  | 13.8 g/L             | [29]       |
|                               | Optimize 1,3-propanediol production from *K. pneumoniae* ATCC 15380     | 56 g/L               | [30]       |
|                               | Integrated bioprocess combining biodiesel production by lipase with microbial production of 1, 3-propanediol by *K. pneumoniae* strain | 1.7 g/L/h            | [31]       |
|                               | *Clostridium butyricum* strain VPI 3266 on a synthetic medium           | 0.60 mol/mol glycerol| [32]       |
|                               | *C. butyricum* strain F2b (process modelling)                           | NAa                  | [33]       |
|                               | *C. butyricum* VPI 1718; fed-batch operation under non-sterile culture conditions | 67.9 g/L             | [35]       |
|                               | One vessel bio- and chemocatalytic process; in a biphasic system without intermediate separation of 1, 3-propanediol; *C. butyricum* DSM10703 | 134 m mol/L          | [36]       |
|                               | *Citric acid* *Y. lipolytica* strain ACA-DC 50109 (process modelling)  | NAa                  | [33]       |
|                               | Acetate Mutants of *Y. lipolytica* Wratislavia AWG7 strain; Fed-batch operation | 139 g/L              | [39]       |
|                               | *Yarrowia lipolytica* strain LGAM S (71)                               | 35 g/L               | [42]       |
|                               | *Y. lipolytica* N15                                                   | 71 g/L               | [43]       |
| Erythritol                    | Fed-batch cultures of *Y. lipolytica* Wratislavia K1                    | 1 g/L/h              | [41]       |
| Hydrogen                      | Photofermentative conversion process; *Rhodopsseudomonas palustris* strain | 6 mol/mol glycerol   | [44]       |
|                               | *Enterobacter aerogenes* strain HU-101; continuous culture; porous ceramics as a support material to fix cells | 63 mmol/L/h          | [45]       |
|                               | Optimize some media compositions of *E. aerogenes* ATCC 35029           | 0.85 mol/mol glycerol| [46]       |
|                               | Anaerobic treatment process; crude glycerol was a co-substrate         | 2.9 mmol/g glycerol  | [49]       |
| Poly (hydroxyalkanoates) (PHAs) | *Pseudomonas oleovorans* NRRL B-14682 and *P. corrugata* 388 grew and synthesized PHB and mcl-PHA, respectively | NAa                  | [52]       |
|                               | Producing PHB; *Paracoccus denitrificans* and *Cupriavidus necator* JMP 134 strains | 48%                  | [53]       |
|                               | Producing PHB; *Cupriavidus necator* strain DSM 545                     | 50%                  | [54]       |
|                               | Producing PHB; *Zobellella denitrificans* MW1; fed-batch cultivation   | 66.9% ± 7.6%         | [55]       |
|                               | Producing PHB; Mixed microbial consortia (MWC)                         | > 50%                | [56]       |
|                               | *Pseudomonas oleovorans* NRRL B-14682, batch culture                   | 30%                  | [57]       |
| Phytase                       | High cell density fermentations, recombinant *Pichia pastoris*         | 1125 U/mL            | [70]       |
| Lipase                        | *Staphylococcus caseolyticus* EX17                                     | 127.3 U/L            | [75]       |
| Succinic acid                 | *Bastia succiniciproducens* DD1; continuous cultivation process         | 1.02 g/g glycerol    | [68]       |
| Docosahexaenoic acid-rich algae | Fermentation of the alga, *Schizochytrium limacium* strain, continuous culture | 0.52 g/L-day         | [61]       |
| Eicosapentaenoic acid         | *Fungus Pythium* irregular                                             | 90 mg/L              | [60]       |
| Lipid                         | *Schizochytrium limacium* SR21; batch culture                          | 73.3%                | [62]       |
|                               | *Cryptococcus curvatus*; two-stage fed-batch process                   | 52%                  | [63]       |
|                               | Oleaginous red yeast *Rhodotorula glutinis* TISTR 5159; fed-batch fermentation | 60.7%                | [65]       |
|                               | *Chlorella protothecoides*                                            | 3 g/L per day        | [66]       |
|                               | *Fungi*                                                                | 42.6%                | [67]       |
| Recombinant human erythropoietin etc. | *Pichia pastoris* medium                                               | 31 mg/L              | [72]       |
| Ethanol                       | Nonpathogenic *Kluyvera cryocrescens* S26; batch fermentation            | 27 g/L               | [47]       |
|                               | *Klebsiella pneumoniae* mutant strain (GEM167)                         | 21.5 g/L             | [48]       |
| Methane                       | Anaerobic digestion                                                    | 0.306 m^3/kg glycerol| [51]       |
| Butanol                       | *Clostridium pasteurianum* (ATCC® 6013™)                               | 0.30 g/g glycerol    | [71]       |
| Fungal protein                | *Rhizopus microsporus var. oligosporus*                                | 0.83 ± 0.02 g/g glycerol| [77]       |

*a* represents data are not available
glycerol concentrations are necessary for improving production efficiency.

In brief, it has been shown that biological conversions of crude glycerol can produce higher value chemicals. However, it is impossible for the current state of the technologies to convert glycerol at rates necessary to prevent a large accumulation of glycerol. Both improved biological and conventional techniques will be needed in future biodiesel plants. Other promising uses, starting with crude glycerol, also have been disclosed. Although hopeful results have been achieved, there are still aspects that need to be improved.

In summary, in recent years, scores of utilization opportunities of crude glycerol have been presented and promising results have been achieved. However, it is imperative to point out that there are still more technical hurdles to jump for developing practical processes to directly utilize crude glycerol from biodiesel production on a large scale. Quick overviews of what has been investigated, with respect to potential chemicals from biological and conventional catalytic conversions of crude glycerol, are summarized in Tables 1 and 2, respectively.

### Abbreviations
DE: Digestible energy; ME: Metabolizable energy; AMEn: nitrogen-corrected apparent ME; GE: General energy; PHA: Poly(hydroxyalkanoates); MMC: Mixed microbial consortia; DHA: Docosahexaenoic acid; EPA: Eicosapentaenoic acid; BC: Bacterial cellulose.

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## Authors' contributions
MAH conceived the study and participated in the design of this review manuscript, and revised the manuscript critically for important intellectual content and language. RS participated in the coordination and helped draft the manuscript. FY carried out the collecting of the references and the drafting of the manuscript. All authors read and approved the final manuscript.

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### Competing interests
The authors declare that they have no competing interests.

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