Deep neck abscess is a life-threatening infection that causes laryngeal edema and upper airway occlusion. The predominant bacterial species involved in this disorder is group A streptococcus. Group G streptococcus (GGS) constitutes the normal commensal flora of the human upper airway. Although rarely, it can cause pharyngitis, tonsillitis, and peritonsillar abscess. Here, we report a case of a woman with parapharyngeal abscess caused by GGS. A 56-year-old woman presented to the emergency department with complaints of sore throat and cervical swelling, and a diagnosis of parapharyngeal abscess was established. She had upper airway occlusion, requiring urgent tracheostomy. Endoscopic incision and drainage of the abscess using a specially designed, rigid curved laryngoscope was successfully performed. Since a rigid curved laryngoscope creates a wide viewing field and working space, it was useful for incision and drainage of the parapharyngeal abscess.

1. Introduction

Deep neck abscess, including parapharyngeal and retropharyngeal abscesses, is a life-threatening infection that can cause laryngeal edema and upper airway occlusion. Several case reports showed that group G streptococcus (GGS) bacteria are normal commensal flora of the human upper airway that can cause severe pharyngitis, tonsillitis, and, rarely, peritonsillar abscess [1–3]. Here, we present a case of parapharyngeal abscess with upper airway occlusion caused by GGS. We successfully performed endoscopic surgery for incision and drainage of the abscess using a rigid curved laryngoscope that is a specially designed laryngoscope developed by Satou (Satou’s Curved Laryngo-Pharyngo Scope®; Nagashima Medical Instruments Company, Tokyo, Japan).

1.1. Case Presentation. A 56-year-old woman presented to our emergency department with complaints of sore throat and cervical swelling. Her medical history included hypertension, habitual smoking, and occasional alcohol consumption. She developed a sore throat and visited a local clinic 3 days before presenting to our emergency department. She was diagnosed with tonsillitis. Group A antigen test was negative, and she was administered amoxicillin 750 mg/day. However, 2 days later, she developed dyspnea, dysphagia, and neck stiffness. On arrival at our hospital, she had a severe sore throat and muffled voice and was drooling. Laryngeal fiberscopy revealed swelling of the caudal oropharyngeal mucosa on the right side and a severely swollen epiglottis and arytenoid region that caused upper airway occlusion. Her SpO₂ was 97% on 2 L oxygen, and her body temperature was 37.4°C. Blood test results suggested strong inflammation (white blood cell count, 15.3 × 10⁹/L; C-reactive protein, 27.6 mg/L).

We established a diagnosis of parapharyngeal abscess. Because of a high risk of suffocation, we first performed tracheostomy with the patient under local anesthesia. Enhanced computed tomography after tracheostomy revealed hypodense lesions at the left lateral and posterior pharyngeal walls (Figure 1). Incision and drainage of the abscess was performed with the patient under general anesthesia using a rigid curved laryngoscope.

Peritonsillitis containing mucus and pus from the posterior pillar was observed (Figure 2(a)). We incised and opened a part of the swollen posterior pillar and lateral and
posterior pharyngeal walls, draining pus from these regions (Figure 2(b)). The operation was completed without any adverse events.

The patient was administered 3 g/d meropenem as empiric therapy. On postoperative day 4, culture for aerobes and anaerobes revealed GGS and *Parvimonas micra*, respectively. Therefore, the antibiotics were changed to 4 g/d piperacillin and 1.2 g/d clindamycin. The recovery course was uneventful.

2. Discussion

GAS is a major microbial pathogen causing pharyngitis, peritonsillar abscess, and deep neck infection. In our case, GGS and *Parvimonas micra* were isolated from the pus. It is often the case that mixed aerobic and anaerobic bacteria can be identified through pus culturing. Tsai et al. reported that polymicrobial growth was observed in 57.39% of pus cultures [4]. Empirical antibiotics targeting both aerobes and anaerobes would be appropriate. However, considering GGS’s aggressive characteristic like GAS, GGS appeared to be an important pathogen of the parapharyngeal and concomitant peritonsillar abscesses that could cause potentially fatal upper airway occlusion. GGS is frequently present in the human pharynx and tonsils. Group C and G streptococci are two antigenic variants of the same organism, *Streptococcus dysgalactiae* subspecies *equisimilis* (SDSE). Genomic sequence homology analysis of GGS revealed that GGS was closest in sequence to GAS, with 72% similarity [5]. Virulence profile analysis of SDSE revealed that its genetic basis of disease propensity is shared with GAS, including the antiphagocytic M protein, streptolysin O, streptolysin S, streptokinase, and one or more pyrogenic exotoxins [6]. The burden of SDSE infection is comparable to that caused by invasive GAS infection [7]. We reviewed the bacteriology of the peritonsillar abscess from previous studies searching the PubMed database. There were 9 studies from 2014 to 2018. Two studies were excluded as the detailed *Streptococcus* sp. was not mentioned. As shown in Table 1, GGS or GCS has a prevalence of less than 5% [8–14]. Notably, GGS was
not detected using the rapid antigen test because of the lack of group A antigen, which is the target of these tests. Current pharyngitis guidelines focus only on group A streptococci and only recommend antibiotics. However, as we demonstrated, GGS also causes life-threatening diseases, such as a deep neck abscess. It is important for the primary care physician to consider carefully negative results.

During surgery, it is important to visualize the abscess lesion and create a sufficient working space. The intraoral approach using a self-retaining mouth gag with ipsilateral tonsillectomy can be used to access diseases in the parapharyngeal space [15]. However, a Davis gag can be used to visualize oropharyngeal structures around the oral cavity and tonsils but cannot be used to visualize the caudal oropharynx. Therefore, physicians should perform invasive surgery to remove the ipsilateral tonsil and approach the parapharyngeal space. We describe a novel method using a rigid curved laryngoscope for incision and drainage of a parapharyngeal abscess. We demonstrate that this instrument was designed for laryngopharyngeal surgery under endoscopic vision [16, 17] (Figure 3). Recent case reports have described successful removal of a fish bone in the hypopharynx and drainage of retropharyngeal abscesses using a rigid curved laryngoscope [18, 19], which is useful for hypopharyngeal and oropharyngeal surgery. The blade is inserted into the pharynx and lifted forward. The oropharynx is well visualized. After exposing the whole oropharynx, the handle is attached to a holder fixed to the operating table. Because of a crooked line path, we used devices such as malleable high-frequency knives (KD-600®; Olympus, Tokyo, Japan) and malleable forceps (Laryngo FIT®; Karl Storz, Tuttlingen, Germany). In our procedure, the rigid curved laryngoscope exposed the whole oropharynx including the caudal oropharynx. A conventional straight laryngoscope can be used to visualize the hypopharynx; however, it provides only a small visual and working space; therefore, it is useful for incision and drainage of a parapharyngeal abscess.

3. Conclusion

GGS is an important pathogen of deep neck abscesses. A rigid curved laryngoscope enables a wide viewing field and working space; therefore, it is useful for incision and drainage of a parapharyngeal abscess.

Ethical Approval

This case report was written in accordance with the Declaration of Helsinki.

Consent

Written informed consent was obtained from the patient for publication of this case.

Conflicts of Interest

The authors declare that there are no conflicts of interest associated with this manuscript.

Authors’ Contributions

ST, TA, and KI prepared and edited this manuscript. RM, HO, TI, MF SK, and SI contributed to the collection of data. TA and KI gave final approval for this version of the manuscript. All authors read and approved the final manuscript.

References

[1] N. Gupta, J. Lovvorn, and R. M. Centor, "Peritonsilar abscess requiring intensive care unit admission caused by group C and G Streptococcus: a case report," *Cases Journal*, vol. 2, no. 1, p. 6808, 2009.

| Investigator            | Country | Year     | Number of samples, n | GGS or GCS, n (%) |
|-------------------------|---------|----------|----------------------|-------------------|
| J. Wiksten et al. [8]   | Finland | 2010-2011| 149                  | 2 (1.3)           |
| Vaikjarv et al. [9]     | Estonia | 2011-2012| 22                   | 1 (4.5)           |
| Tachibana et al. [10]   | Japan   | 2008-2013| 100                  | 1 (1.0)           |
| Lepelletier et al. [11] | French  | 2009-2012| 412                  | 11 (2.6)          |
| Plum et al. [12]        | USA     | 2002-2012| 69                   | 1 (1.4)           |
| Mazur et al. [13]       | Poland  | 2003-2013| 45                   | 1 (2.2)           |
| Gavriel et al. [14]     | Israel  | 1996-2003| 132                  | 1 (0.76)          |

GGS, group G streptococcus; GCS, group C streptococcus.
[2] M. Shah, R. M. Centor, and M. Jennings, “Severe acute pharyngitis caused by group C streptococcus,” *Journal of General Internal Medicine*, vol. 22, no. 2, pp. 272–274, 2007.

[3] H. Faden, “Group C and G streptococcal disease among children,” *Pediatric Infectious Disease Journal*, vol. 24, no. 11, pp. 1014–1015, 2005.

[4] Y. W. Tsai, Y. H. Liu, and H. H. Su, “Bacteriology of peritonsillar abscess: the changing trend and predisposing factors,” *Brazilian Journal of Otorhinolaryngology*, 2017.

[5] Y. Shimomura, K. Okumura, S. Y. Murayama et al., “Complete genome sequencing and analysis of a Lancefield group G streptococcus dysgalactiae subsp. equisimilis strain causing streptococcal toxic shock syndrome (STSS),” *BMC Genomics*, vol. 12, no. 1, p. 17, 2011.

[6] M. R. Davies, D. J. McMillan, R. G. Beiko et al., “Virulence profiling of streptococcus dysgalactiae subspecies equisimilis isolated from infected humans reveals 2 distinct genetic lineages that do not segregate with their phenotypes or propensity to cause diseases,” *Clinical Infectious Diseases*, vol. 44, no. 11, pp. 1442–1454, 2007.

[7] L. N. Broyles, C. Van Beneden, B. Beall et al., “Population-based study of invasive disease due to beta-hemolytic streptococci of groups other than A and B,” *Clinical Infectious Diseases*, vol. 48, no. 6, pp. 706–712, 2009.

[8] J. Wiksten, E. Kaltiainen, A. Pitkaranta, and K. Blomgren, “Renewal of peritonsillar abscess: impact of the bacterial species of the infection and clinical features of the patient—a prospective comparative aetiological study,” *Clinical Otolaryngology*, vol. 42, no. 6, pp. 1358–1362, 2017.

[9] R. Vaikjarv, P. Kasenomm, L. Jaanimae et al., “Microbiology of peritonsillar abscess in the South Estonian population,” *Microbial Ecology in Health & Disease*, vol. 27, p. 27787, 2016.

[10] T. Tachibana, Y. Orita, S. Takao et al., “The role of bacteriological studies in the management of peritonsillar abscess,” *Auris Nasus Larynx*, vol. 43, no. 6, pp. 648–653, 2016.

[11] D. Lepelletier, V. Pinaud, P. Le Conte et al., “Peritonsillar abscess (PTA): clinical characteristics, microbiology, drug exposures and outcomes of a large multicenter cohort survey of 412 patients hospitalized in 13 French university hospitals,” *European Journal of Clinical Microbiology & Infectious Diseases*, vol. 35, no. 5, pp. 867–873, 2016.

[12] A. W. Plum, A. J. Mortelliti, and R. E. Walsh, “Microbial flora and antibiotic resistance in peritonsillar abscesses in upstate New York,” *Annals of Otolaryngology, Rhinology & Laryngology*, vol. 124, no. 11, pp. 875–880, 2015.

[13] E. Mazur, E. Czerwinska, I. Korona-Glowniak, A. Grochowalska, and M. Koziol-Montewka, “Epidemiology, clinical history and microbiology of peritonsillar abscess,” *European Journal of Clinical Microbiology & Infectious Diseases*, vol. 34, no. 3, pp. 549–554, 2015.

[14] H. Gavriel, Y. Golan, T. Lazarovitch, and E. Eviatar, “Bacteriology of peritonsillar abscess in patients over 40 years—a neglected age group,” *European Archives of Oto-Rhino-Laryngology*, vol. 272, no. 4, pp. 981–984, 2015.

[15] Y. Okumura, H. Hidaka, N. Noguchi, and Y. Katori, “Intraoral drainage under surgical microscopy with tonsillectomy for parapharyngeal abscesses,” *Journal of Laryngology & Otology*, vol. 129, no. 6, pp. 595–597, 2015.

[16] Y. Satou, T. Omori, and M. Tagawa, “Treatment of superficial carcinoma in the hypopharynx,” *Nippon Jibiinkoka Gakkai Kaiho*, vol. 109, no. 7, pp. 581–586, 2006.

[17] I. Tateya, M. Muto, S. Morita et al., “Endoscopic laryngopharyngeal surgery for superficial laryngo-pharyngeal cancer,” *Surgical Endoscopy*, vol. 30, no. 1, pp. 323–329, 2016.

[18] H. Sakaida, K. Chiyonobu, H. Ishinaga, and K. Takeuchi, “Use of a rigid curved laryngoscope for removal of a fish bone in the hypopharynx,” *Case Reports in Otolaryngology*, vol. 2016, Article ID 9689521, 3 pages, 2016.

[19] K. Watanabe, D. Yamauchi, Y. Takanashi, H. Hidaka, and Y. Katori, “Novel application of a rigid curved laryngopharyngoscope for examination and treatment of hypopharyngeal lesions,” *Auris Nasus Larynx*, vol. 45, no. 2, pp. 367–370, 2018.