HAX1 enhances the survival and metastasis of non-small cell lung cancer through the AKT/mTOR and MDM2/p53 signaling pathway

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Keywords
AKT; HS-1-associated protein-1; non-small cell lung cancer.

Abstract

Background: HS-1-associated protein-1 (HAX1) has been reported to be overexpressed in non-small cell lung cancer (NSCLC) tissues. However, the underlying mechanism of HAX1 in NSCLC has not previously been demonstrated. The present study investigated the role and underlying mechanism of HAX1 in NSCLC.

Methods: The HAX1 expression were confirmed in NSCLC tissues through TCGA database and qRT-PCR. Moreover, we performed qRT-PCR, Western blotting, Transwell assays, TUNEL assays and so on to evaluate the role of HAX1 in A549 and H1299 cell lines.

Results: mRNA expression of HAX1 was overexpressed in NSCLC tissues compared to adjacent normal tissues according to The Cancer Genome Atlas (TCGA) database. QRT-PCR assays showed that HAX1 mRNA expression was upregulated in NSCLC tissues. The high HAX1 mRNA levels were found to be positively associated with tumor size, TNM stage and lymphatic metastasis. Silencing of HAX1 promoted apoptosis and reduced invasion of A549 and H1299 cells by inhibiting the AKT/mTOR and MDM2/P53 signal pathway. AKT agonist SC79 could inhibit apoptosis and promote proliferation, migration and invasion of A549 and H1299 cells transfected with si-HAX1.

Conclusions: The present study provided a better understanding of HAX1 mechanism in NSCLC and potential therapeutic target for NSCLC.

Introduction

Lung cancer is one of the most lethal cancers in the world, which causes a heavy socioeconomic burden.1–3 There are two major histological types including small cell lung cancer (SCLC) and non-small cell lung cancer (NSCLC) and NSCLC accounts for more than 80% of all lung cancer cases.4 NSCLC includes adenocarcinoma, squamous cell carcinoma, adenosquamous cell carcinoma, and large cell carcinoma. Despite huge efforts in the fight against the disease, the prognosis for NSCLC is still poor and a five-year survival rate is no more than 15%.5–7 The development of lung cancer is a multistep process, including changes in oncogenes and tumor suppressor genes, leading to gradual dysregulation of pathways related to cell proliferation and survival.8 Therefore, the discovery that new molecular targets play a regulatory role in the uncontrolled proliferation of tumor cells is of great significance for the diagnosis and treatment of NSCLC.9
HS-1 related protein X-1 (HAX1), a 35 kDa protein, is an important regulator of apoptosis and was first discovered to interact with HS-1, a substrate of serine/tyrosine kinases. Its gene is located on human chromosome lq21.3 and encodes 279 amino acid residues. HAX1 plays an anti-apoptotic function in many tissues due to a homology domain of the BCL-2 family and a weak homology with the apoptosis-related protein NIP3. Recent studies have shown that HAX1 can bind to a variety of intracellular apoptosis-related proteins (such as caspase 9, OM1/HtrA2, VPR, K15, AKT) and exert antiapoptotic effects. HAX1 may also participate in the cell migration process through molecules such as Galphal3, integrin αVβ6, IL-1α, GRB7, NF-kB. In addition, some studies have confirmed that HAX1 also binds 3' translation region mRNAs and acts as a new trans-activating factor in the post-transcriptional regulation of Pol β expression. HAX1 is upregulated in lung cancer, hematoma, lymphoma, melanoma, leukemia and myeloma according to Oncomine, a cancer microarray database. In lung cancer tissues, upregulation has been observed in the most advanced stages of the disease (stages III–IV, 0.049 for lung cancer, respectively). However, the role of HAX1 in NSCLC metastasis and recurrence as well as the underlying mechanism have not been previously reported.

Here, we explored the relationship between HAX1 expression and the apoptosis and metastasis of NSCLC. Our study confirmed HAX1 expression in NSCLC samples, and revealed that HAX1 expression is significantly elevated in NSCLC tissues, suggesting that HAX1 might play an important role in EMT in NSCLC cells and in the promotion of NSCLC metastasis.

Methods

Clinical specimen collection

A total of 35 paired tumor tissues and their matched adjacent normal tissues were obtained from March 2015 to June 2019 in Xinhua Hospital (Shanghai, China). Each fresh resected specimen was snap-frozen in liquid nitrogen and stored at −80°C prior to RNA isolation. This study was approved by the Medical Ethics Committee of the Xinhua Hospital and all experiments were performed in accordance with the Declaration of Helsinki.

Cell culture

NSCLC cell lines H1299 and A549 were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA). All cells were cultured in DMEM high-glucose medium (Gibco, USA) with 10% fetal bovine serum (Gibco, USA) at 37°C in a humidified environment with 5% CO2.

RNAi and transfection

HAX1-siRNAs (si-HAX1) were designed and purchased from Sangon Biotech, China. The target sequences for HAX1 were as follows: Sense: GGAUACGUUUCCCGAUAA dTdT; Antisense: UUAUCUGGGAGAAGUACGdTdT. Cells were seeded on six-well plates for 70% confluency and transfected with si-HAX1 using Lipofectamine 2000 (Invitrogen, USA) according to the manufacturer’s instructions. Cells were obtained for the following experiments after 48 hours.

Quantitative real-time polymerase chain reaction (qRT-PCR)

Total RNA was obtained from tissue samples or cells by using Trizol reagent (Beyotime, China) according to the manufacturer’s instruction. Subsequently, cDNA synthesis and qRT-PCR were performed by one-step RT qPCR SYBRGreen Kit (DBI, Germany) and the ABI 7500 Fast Real-Time PCR System (Life Tech, Carlsbad, California, USA). 2−ΔΔCt method was used to calculate the relative expression of genes, which were normalized to β-actin.

Western blotting

Cells were lysed using RIPA buffer containing 10% PMSF (Beyotime, China) and total protein obtained. Then, 30 μg protein lysate were separated by 8% to 12% SDS-PAGE (sodium dodecyl sulfate polyacrylamide gel electrophoresis) and electro-transferred to PVDF (polyvinylidene difluoride). PVDF were blocked in 5% skimmed milk for 2 hours. The membranes were then incubated with primary antibodies at 4°C overnight. Primary antibodies are listed as follows: β-actin (Beyotime, China, 1:1000), Cytochrome C (Abcam, UK, 1:1000), Caspase 9 (Proteintech, USA, 1:1000), Bcl-2 (Proteintech, USA, 1:1000), Bax (Proteintech, USA, 1:1000), Caspase 3 (Abcam, UK, 1:1000), AKT (CST, USA, 1:1000), p-AKT (CST, USA, 1:1000), mTOR (CST, USA, 1:1000), p-mTOR (CST, USA, 1:1000), MDM2 (CST, USA, 1:1000), p-MDM2 (CST, USA, 1000), p53 (CST, USA, 1:1000), and HAX1 (Proteintech, USA, 1:1000). Membranes were washed with three times and subsequently incubated with horseradish peroxidase (HRP)-conjugated IgG secondary antibodies (Beyotime, China, 1:2000) at room temperature for 1 hour. The target protein was visualized using an ECL kit (Millipore, Billerica, MA, USA). GAPDH was regarded as an endogenous reference.

Transwell invasion assay

Cell invasion assay was carried out using 24-well transwell plates (Corning, USA) containing Matrigel...
(BD, USA). Approximately $2 \times 10^5$ cells were planted in the upper chamber with medium in triplicate without serum. Medium supplemented with 10% FBS was added to the lower chamber. After incubation for 24 hours, the cells above the Matrigel layer were removed using a cotton swab, and then cells below the membrane were fixed using 4% paraformaldehyde, and stained with crystal violet for 10 minutes. Finally, five randomly chosen fields of each well were counted.

**Figure 1** Information of HAX1 expression in TCGA database. (a) Expression of HAX1 in LUAD based on sample type. (b) Expression of HAX1 in LUAD based on patients' smoking habits. (c) Expression of HAX1 in LUAD based on individual cancer stages. (d) Expression of HAX1 in LUAD based on patients’ gender. (e) Expression of HAX1 in LUAD based on patients’ age. (f) Expression of HAX1 in LUAD based on nodal metastasis status. (g) Expression of HAX1 in LUSC based on sample type. (h) Expression of HAX1 in LUSC based on patients’ smoking habits. (i) Expression of HAX1 in LUSC based on individual cancer stages. (j) Expression of HAX1 in LUAD based on patients’ gender. (k) Expression of HAX1 in LUSC based on patients’ age. (l) Expression of HAX1 in LUSC based on nodal metastasis status. LUAD, lung adenocarcinoma; LUSC, lung squamous cell carcinoma; N0, no regional lymph node metastasis; N1, metastases in 1–3 axillary lymph nodes; N2, metastases in 4–9 axillary lymph nodes; N3, metastases in 10 or more axillary lymph nodes.
Annexin V-FITC/PI apoptosis assay

Apoptotic cells were detected with an annexin V-FACS apoptosis detection kit (BD, USA) according to the manufacturer’s instructions. Briefly, after transfection with si-HAX1 for 48 hours, cells were washed with ice PBS twice and incubated in 300 μL 1× binding buffer supplemented with 5 μL annexin V-FITC for 10 minutes and then resuspended with PI (propidium iodide) in the dark for 5 minutes at 25°C. The stained cells were detected using an FC 500 flow cytometer (Beckman Coulter, Brea, CA, USA) within 1 hour according to the manufacturer’s protocol.

TUNEL assays

Cells were seeded on the coverslips and then transfected with si-HAX1 for 48 hours. After being fixed using 4% paraformaldehyde, cells were penetrated with 0.3% Triton X-100 for 5 minutes. Then, 50 μL TUNEL was added to each coverslip and incubated for 1 hour. Finally, images were photographed by microscopy (Olympus BX51).

Statistical analysis

All data are presented as mean ± standard deviation (SD). Paired sample t-test was used to calculate the expression differences of HAX1 between NSCLC tissues and adjacent noncancerous tissues. Independent sample Student’s t-test was used to calculate the differences between groups. Pearson’s correlation coefficient was used to analyze the expression correlation of HAX1. All experiments were performed in triplicate. The difference was considered to be statistically significant when P-values were less than 0.05. ***P < 0.001, **P < 0.01, *P < 0.05. Statistical analysis was carried out in SPSS 20.0 (SPSS, USA).

| Patients’ smoking habit | Comparison | P-value | Cancer stages | Comparison | P-value |
|-------------------------|------------|---------|---------------|------------|---------|
| Normal vs. non-smoker   | <0.05      | Normal vs. stage 1 | <0.05 |
| Normal vs. smoker       | <0.05      | Normal vs. stage 2 | <0.05 |
| Normal vs. reformed smoker 1 | <0.05 | Normal vs. stage 3 | <0.05 |
| Normal vs. reformed smoker 2 | <0.05 | Normal vs. stage 4 | <0.05 |
| Non-smoker vs. smoker   | >0.05      | Stage 1 vs. stage 2 | >0.05 |
| Non-smoker vs. reformed smoker 1 | >0.05 | Stage 1 vs. stage 3 | >0.05 |
| Non-smoker vs. reformed smoker 2 | >0.05 | Stage 1 vs. stage 4 | >0.05 |
| Smoker vs. reformed smoker 1 | >0.05 | Stage 2 vs. stage 3 | >0.05 |
| Smoker vs. reformed smoker 2 | >0.05 | Stage 2 vs. stage 4 | >0.05 |
| Reformed smoker 1 vs. reformed smoker 2 | >0.05 | Stage 3 vs. stage 4 | >0.05 |

| Patients’ gender | Comparison | P-value | Cancer stages | Comparison | P-value |
|------------------|------------|---------|---------------|------------|---------|
| Normal vs. male  | <0.05      | Normal vs. N0 | <0.05 |
| Normal vs. female| <0.05      | Normal vs. N1 | <0.05 |
| Male vs. female  | >0.05      | Normal vs. N2 | <0.05 |

| Patients’ age | Comparison | P-value | Nodal metastasis status | Comparison | P-value |
|---------------|------------|---------|-------------------------|------------|---------|
| Normal vs. age (21–40 years) | >0.05 | Normal vs. N0 | <0.05 |
| Normal vs. age (41–60 years) | <0.05 | Normal vs. N1 | <0.05 |
| Normal vs. age (61–80 years) | <0.05 | Normal vs. N2 | <0.05 |
| Normal vs. age (81–100 years) | <0.05 | Normal vs. N3 | >0.05 |
| Age (21–40 years) vs. age (41–60 years) | >0.05 | N0 vs. N1 | >0.05 |
| Age (21–40 years) vs. age (61–80 years) | >0.05 | N0 vs. N2 | >0.05 |
| Age (21–40 years) vs. age (81–100 years) | >0.05 | N0 vs. N3 | >0.05 |
| Age (41–60 years) vs. age (61–80 years) | >0.05 | N1 vs. N2 | <0.05 |
| Age (41–60 years) vs. age (81–100 years) | >0.05 | N1 vs. N3 | <0.05 |
| Age (61–80 years) vs. age (81–100 years) | >0.05 | N2 vs. N3 | <0.05 |

N0, no regional lymph node metastasis; N1, metastases in 1 to 3 axillary lymph nodes; N2, metastases in 4 to 9 axillary lymph nodes; N3, metastases in 10 or more axillary lymph nodes.
Results

HAX1 upregulated in NSCLC tissues according to TCGA database

HAX1 has been reported to be overexpressed in lung cancer. To confirm whether HAX1 was overexpressed in NSCLC, we searched the express information of HAX1 in the Cancer Genome Atlas (TCGA) database. We found that expression of HAX1 was upregulated in LUAD based on sample types (Fig 1a). The P-value of each group is shown in Tables 1 and 2. There was no significant difference in HAX1 expression in LUAD on cancer stage, patients’ smoking habits, gender, age and nodal metastasis status (Fig 1b–f). In addition, expression of HAX1 was also overexpressed in LUSC based on sample types (Fig 1g). HAX1 expression was not significantly different in LUSC on cancer stage, patients’ smoking habits, gender, age and nodal metastasis status (Fig 1h–l). The HAX1 expression was explored in TCGA database and revealed that HAX1 was overexpressed in NSCLC and high levels might be positively related to the clinicopathological characteristics of NSCLC patients.

Upregulation of HAX1 observed in NSCLC tissues compared to adjacent normal tissues

We performed qRT-PCR to measure the mRNA expression of HAX1 in NSCLC and adjacent normal tissues. The results showed that HAX1 mRNA expression was higher in NSCLC tissues than adjacent normal tissues (Fig 2a). Furthermore, we observed that the expression level of HAX1 was positively associated with tumor size (P = 0.027) (Table 3). HAX1 expression level was not significantly associated with gender, age, tumor status progression or lymphatic metastasis. Together, our data suggested that increased HAX1 expression might be involved in NSCLC progression.

| Patients’ smoking habit | P-value | Cancer stages | P-value |
|------------------------|---------|--------------|---------|
| Normal vs. non-smoker   | <0.05   | Normal vs. stage 1 | <0.05   |
| Normal vs. smoker       | <0.05   | Normal vs. stage 2 | <0.05   |
| Normal vs. reformed smoker 1 | <0.05   | Normal vs. stage 3 | <0.05   |
| Normal vs. reformed smoker 2 | <0.05   | Normal vs. stage 4 | <0.05   |
| Non-smoker vs. smoker   | >0.05   | Stage 1 vs. stage 2 | >0.05   |
| Non-smoker vs. reformed smoker 1 | >0.05   | Stage 1 vs. stage 3 | >0.05   |
| Non-smoker vs. reformed smoker 2 | >0.05   | Stage 1 vs. stage 4 | >0.05   |
| Smoker vs. reformed smoker 1 | >0.05   | Stage 2 vs. stage 3 | >0.05   |
| Smoker vs. reformed smoker 2 | >0.05   | Stage 2 vs. stage 4 | >0.05   |
| Reformed smoker 1 vs. reformed smoker 2 | >0.05   | Stage 3 vs. stage 4 | >0.05   |

| Patients’ gender | P-value | Cancer stages | P-value |
|------------------|---------|--------------|---------|
| Normal vs. male  | <0.05   | Normal vs. N0 | <0.05   |
| Normal vs. female| <0.05   | Normal vs. N1 | <0.05   |
| Male vs. female  | >0.05   | Normal vs. N2 | <0.05   |

| Patients’ age | P-value | Nodal metastasis status | P-value |
|---------------|---------|-------------------------|---------|
| Normal- vs. age (21–40 years) | >0.05   | Normal vs. N0 | <0.05   |
| Normal vs. age (41–60 years) | <0.05   | Normal vs. N1 | <0.05   |
| Normal vs. age (61–80 years) | <0.05   | Normal vs. N2 | <0.05   |
| Normal vs. age (81–100 years) | <0.05   | Normal vs. N3 | <0.05   |
| Age (21–40 years) vs. age (41–60 years) | >0.05   | N0 vs. N1 | >0.05   |
| Age (21–40 years) vs. age (61–80 years) | >0.05   | N0 vs. N2 | >0.05   |
| Age (21–40 years) vs. age (81–100 years) | >0.05   | N0 vs. N3 | >0.05   |
| Age (41–60 years) vs. age (61–80 years) | >0.05   | N1 vs. N2 | <0.05   |
| Age (41–60 years) vs. age (81–100 years) | >0.05   | N1 vs. N3 | >0.05   |
| Age (61–80 years vs. age (81–100 years) | >0.05   | N2 vs. N3 | >0.05   |

N0, no regional lymph node metastasis; N1, metastases in 1 to 3 axillary lymph nodes; N2, metastases in 4 to 9 axillary lymph nodes; N3, metastases in 10 or more axillary lymph nodes.
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**Figure 2** HAX1 knockdown increases apoptosis of A549 and H1299 cells. (a) HAX1 mRNA expression was increased in NSCLC tissue samples. (b) Adjacent tissues and (c) cancer tissues. (b) HAX1 mRNA expression was reduced remarkably by si-HAX1 in A549 and H1299 cells. (d) Quantitative analysis of protein HAX1 expression in A549 and H1299 cells. (e) Western blotting for cleaved-caspase 9, cleaved-caspase 3, Bcl-2, Bax and β-actin in A549 and H1299 cells. (f) Quantitative analysis of protein cleaved-caspase 9, cleaved-caspase 3, Bcl-2, Bax in A549 and H1299. (g) Western blotting for cleaved-caspase 3, Bcl-2, Bax expression in A549 and H1299. (h) Quantitative analysis of protein cleaved-caspase 9, cleaved-caspase 3, Bcl-2, Bax in A549 and H1299. (i) Western blotting for cytochrome C and β-actin in A549 and H1299 cells. (j) Quantitative analysis of protein cytochrome C expression in A549 and H1299. (k) TUNEL assays for apoptosis of A549 and H1299 cells. ***P < 0.001, **P < 0.01, *P < 0.05.

**HAX1 knockdown increases apoptosis of NSCLC cell lines**

To explore the role and underlying mechanism of HAX1, we constructed the siRNA to silence the HAX1 expression in A549 and H1299 cell lines. First, qRT-PCR was used to measure the mRNA expression in A549 and H1299 cell lines transfected with si-HAX1. The results of qRT-PCR confirmed that si-HAX1 caused a significant decrease of
HAX1 in A549 and H1299 cells (Fig 2b). Western blotting also confirmed si-HAX1 worked in A549 and H1299 cells (Fig 2c,d). Second, we wanted to investigate whether HAX1 knockdown caused A549 and H1299 cells apoptosis. Thus, western blotting and TUNEL assays were used to investigate the apoptosis of A549 and H1299 cells. HAX1 knockdown increased the protein levels of cleaved-caspase 9 and cleaved-caspase 3 and decreased the ratio of Bcl-2/Bax (Fig 2e–h). HAX1 knockdown also promoted the flux of cytochrome 3 from mitochondria to cytoplasm in A549 and H1299 cells (Fig 2i,j). TUNEL assay results showed that HAX1 knockdown caused the apoptosis of A549 and H1299 cells (Fig 2k). These results showed that HAX1 knockdown could cause NSCLC cell apoptosis.

**Silencing of HAX1 reduces invasion and epithelial–mesenchymal transition of NSCLC cells**

To investigate whether HAX1 regulates the metastasis of NSCLC cells, si-HAX1 was transfected into A549 and H1299 cell lines. As shown in Fig 3a–f, HAX1 knockdown reduced the protein expression of Vimentin, Slug, Snail and Twist, whereas E-cadherin protein expression was increased. Silencing of HAX1 remarkably reduced the epithelial-mesenchymal transition of NSCLC cells. In addition, transwell assay results showed that HAX1 knockdown caused a significant decrease of invasion in A549 and H1299 cells transfected with si-HAX1 (Fig 3g,h). Silencing of HAX1 reduced the invasion and epithelial–mesenchymal transition of NSCLC cells.

**HAX1 suppresses NSCLC cell apoptosis through inhibiting AKT/mTOR signal pathway**

A recent study reported that HAX1 prevents glioblastoma cells from apoptosis through the Akt1 pathway, and it also has been reported that the AKT/mTOR signal pathway plays an important role in NSCLC. AKT/mTOR signal pathway induces gefitinib-resistant of NSCLC. FAM83D has been reported to improve epithelial-mesenchymal transition, invasion and cisplatin resistance through activating the AKT/mTOR pathway in NSCLC. The oncogene IARS2 promotes the occurrence of NSCLC by activating the AKT/mTOR pathway. Therefore, we wanted to explore whether HAX1 protected the apoptotic NSCLC cells through inhibiting AKT/mTOR signal pathway. We found that HAX1 knockdown could decrease the levels of p-AKT, p-mTOR and p-70S6K in A549 and H1299 cells (Fig 4a,b). Furthermore, we performed AKT agonist SC79 (5 μg/mL for 1 hour) to treat A549 and H1299 cells after silencing of HAX1. As shown in Fig 4c,d, SC79 was not able to change the protein expression of HAX1, which suggested the AKT/mTOR signal pathway was not able to regulate HAX1. However, SC79 was capable of promoting the levels of p-AKT and p-mTOR in A549 and H1299 cells after silencing of HAX1. To investigate whether SC79 could attenuate A549 and H1299 cell apoptosis, we performed a TUNEL assay to detect the apoptosis of A549 and H1299 cells. As shown in Fig 4f, we determined that SC79 could promote the survival of A549 and H1299 cells after HAX1 knockdown. These results showed that HAX1 increases the apoptotic NSCLC cells through inhibiting the AKT/mTOR signal pathway.

As a downstream of AKT, MDM2/p53 signal pathway has been reported to play an important role in glioblastoma cells and endothelial progenitor cells. We further investigated whether the MDM2/p53 signal pathway participates in NSCLC cells transfected with si-HAX1. We observed the protein level of p-MDM2 was reduced and p53 was increased in A549 and H1299 cells after si-HAX1 transfection (Fig 5a,b). MDM2, as a negative regulator of p53, can bind to p53 to promote degradation. To investigate whether HAX1 inhibits NSCLC cells apoptosis through the AKT/MDM2/p53 signal pathway, we detected the protein levels of p-MDM2 and p53 after AKT agonist SC79 treatments in NSCLC cells. SC79 increased the protein expression of MDM2 but not p53.
levels of p-MDM2 in A549 and H1299 cells (Fig 5c,d). Although HAX1 was silenced, SC79 caused an increased expression of p-MDM2 in A549 and H1299 cells (Fig 5c,d). SC79 did not change the expression of p53 in A549 and H1299 cell (Fig 5c,d). However, SC79 could reduce the p53 protein expression in A549 and H1299 cells transfected with si-HAX1 (Fig 5c,d). These results confirmed that HAX1 also prevented NSCLC cells apoptosis through promoting AKT/MDM2/p53 signal pathway.

**Discussion**

NSCLC is one of the most prevalent cancers in the world with a high risk of patients developing subsequent
metastasis. Patients with local or distant metastasis have a poor prognosis. Therefore, early diagnosis and treatment are important for NSCLC patients.

HAX1 serves as a multifunctional protein and exerts an important role in many metabolisms through various cellular proteins and viral proteins. Many studies have reported that HAX1 participated in multiple cellular processes and signaling pathways, such as cancer cell apoptosis, invasion, proliferation, and migration. Upregulation of HAX1 is found in many tumors. In breast cancer, HAX1 mRNA expression has been reported to be higher in tumor tissues than paired adjacent noncancerous tissues.

Figure 4 HAX1 promotes the survival of NSCLC cells via AKT/mTOR signaling pathway. (a) Phosphorylation of AKT, mTOR and 70S6K was inhibited in A549 and H1299 cells transfected with si-HAX1. (b) Quantitative analysis of protein p-AKT, p-mTOR and p-70S6K expression in A549 and H1299 cells. (c) SC79 promoted the phosphorylation of AKT and mTOR in A549 and H1299 cells transfected with si-HAX1. (d, e) Quantitative analysis of protein HAX1, p-AKT, and p-mTOR expression in A549 and H1299 cells. (f) TUNEL assays for apoptosis of A549 and H1299 cells transfected with si-HAX1 after treatment of SC79.

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Upregulation of HAX1 has also been observed in hepatocellular carcinoma and was associated positively with grade of differentiation, tumor thrombus and tumor satellite. A recent study showed that HAX1 is upregulated in lung cancer. However, the role and underlying mechanism of HAX1 are still unclear. In this study, we searched the expression information of HAX1 in TCGA database and found that HAX1 was overexpressed in NSCLC tissues. We then analyzed HAX1 expression in NSCLC tissues and paired adjacent normal tissues by using qRT-PCR and western blotting. The results showed that HAX1 mRNA expression was higher in NSCLC tissues than paired adjacent normal tissues. We also found that high HAX1 levels were related positively with tumor status progression, tumor size and lymphatic metastasis.

HAX1 acts as antiapoptotic protein and maintains various cell survival. Many studies have previously reported that HAX1 protects various cells from apoptosis.
including cardiomyocytes, neurons as well as many cancer cells.34–37 Therefore, we investigated the role of HAX1 in NSCLC apoptosis. Si-HAX1 was designed to silence the expression of HAX1 in A549 and H1299 cells. Silencing of HAX1 caused an increased expression of cleaved-caspase 9, cleaved-caspase 3 and decreased ratio of Bcl-2/Bax. Importantly, HAX1 knockdown increased the flux of cytochrome C from mitochondria to cytoplasm. TUNEL assays confirmed that apoptotic cells were observed in A549 and H1299 cells. Furthermore, our results demonstrated that HAX1 could reduce the invasion and epithelial-mesenchymal transition of NSCLC cells. These results suggested that HAX1 plays an important role in maintaining the survival and metastasis of NSCLC cells.

Emerging evidence has revealed that HAX1 protects glioblastoma cells from apoptosis through the AKT pathway.15 The AKT signal pathway exerts an important role in protecting cells from apoptosis. In osteoblasts, PMSC6 has been reported to increase osteoblast apoptosis via inhibiting AKT signaling pathway activation in an OVX mouse model.38 MiR-208 has also been reported to protect neonatal rat cardiomyocyte apoptosis in simulated ischemia-reperfusion injury via AKT signaling pathway.39 However, inhibition of the AKT signaling pathway can promote the apoptosis of gastric cancer cells, breast cancer, lung cancer and so on.40–42 Importantly, we found that HAX1 knockdown suppressed the phosphorylation of AKT, mTOR and S6K. Subsequently, we performed AKT agonist SC79 to treat A549 and H1299 cells transfected with si-HAX1. The results of western blotting showed that SC79 could promote the phosphorylation of AKT and mTOR, whereas there was no change in HAX1 expression. SC70 also reduced the apoptosis of A549 and H1299 cells caused by HAX1 knockdown. These results indicated that HAX1 protected NSCLC cells from apoptosis by regulating the AKT signaling pathway.

MDM2, as a negative regulator of p53, can bind to p53 to limit its function, and ubiquitinate p53 to promote degradation.22–24 MDM2 maintains the tumor characteristics of lung cancer via suppressing the function and expression levels of p53.43–46 For example, Cai et al. reported miR-305 restricts the progression of NSCLC through inhibiting MDM2.45 Ning et al. also reported that ZCCHC10 inhibits lung cancer progression and cisplatin resistance by attenuating MDM2-mediated p53 ubiquitination and degradation.46 It has also been reported that P53, a famous tumor suppressor gene, inhibits NSCLC cell proliferation, invasion and epithelial-mesenchymal transition.47–49 AKT also regulates the tumor characteristics of tumor by MDM2/p53 signal pathway.50–52 Our results indicated that AKT/MDM2/p53 pathway participates in the protective effect of HAX1 in lung cancer.

In conclusion, our findings indicate that HAX1 may be an essential factor in the development of NSCLC. Therefore, the present study provided a potential target for NSCLC therapy and a better understanding for the mechanism of NSCLC.

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Disclosure
The authors confirm that there are no conflicts of interest.

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