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Characteristics and Outcomes of Initial Virologic Suppressors during Analytic Treatment Interruption in a Therapeutic HIV-1 gag Vaccine Trial

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Abstract

**Background:** In the placebo-controlled trial ACTG A5197, a trend favoring viral suppression was seen in the HIV-1-infected subjects who received a recombinant Ad5 HIV-1 gag vaccine.

**Objective:** To identify individuals with initial viral suppression (plasma HIV-1 RNA set point < 3.0 log10 copies/ml) during the analytic treatment interruption (ATI) and evaluate the durability and correlates of virologic control and characteristics of HIV sequence evolution.

**Methods:** HIV-1 gag and pol RNA were amplified and sequenced from plasma obtained during the ATI. Immune responses were measured by flow cytometric analysis and intracellular cytokine expression assays. Characteristics of those with and without initial viral suppression were compared using the Wilcoxon rank sum and Fisher's exact tests.

**Results:** Eleven out of 104 participants (10.6%) were classified as initial virologic suppressors, nine of whom had received the vaccine. Initial virologic suppressors had significantly less CD4+ cell decline by ATI week 16 as compared to non-suppressors (median 7 CD4+ cell gain vs. 247 CD4+ cell loss, P = 0.04). However, of the ten initial virologic suppressors with a pVL at ATI week 49, only three maintained pVL < 3.0 log10 copies/ml. HIV-1 Gag-specific CD4+ interferon-γ responses were not associated with initial virologic suppression and no evidence of vaccine-driven HIV sequence evolution was detected. Participants with initial virologic suppression were found to have a lower percentage of CD4+ CTLA-4+ cells prior to treatment interruption, but a greater proportion of HIV-1 Gag-reactive CD4+ TNF-α+ cells expressing either CTLA-4 or PD-1.

**Conclusions:** Among individuals participating in a rAd5 therapeutic HIV-1 gag vaccine trial, initial viral suppression was found in a subset of patients, but this response was not sustained. The association between CTLA-4 and PD-1 expression on CD4+ T cells and virologic outcome warrants further study in trials of other therapeutic vaccines in development.

**Trial Registration:** ClinicalTrials.gov NCT00080106

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**Competing Interests:** The authors have the following competing interests: Drs. Kuritzkes, Lederman, and Schooley are consultants to Merck; Dr. Kuritzkes has received research support from Merck; Mary Carrington is an employee of SAIC-Frederick, Inc.; Dr. Li is the recipient of a Clinical Investigator Training Program Fellowship: Harvard/MIT Health Sciences and Technology - Beth Israel Deaconess Medical Center, in collaboration with Pfizer Inc. and Merck & Co. The authors appreciate Dr. Michael Robertson’s (Merck Research Laboratories) input in the design of the study and review of the manuscript. There are no patents, products in development or marketed products to declare. This does not alter the authors’ adherence to all the PLoS ONE policies on sharing data and materials.

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Introduction

The initial optimism that antiretroviral therapy (ART) could lead to the eradication of HIV infection has been tempered by the realization that virologic control is lost with the discontinuation of ART even after an extended period on treatment [1]. Despite improved tolerability of newer ART regimens, long-term treatment carries risks of drug resistance, metabolic, and other complications of chronic ART use [2], and is constrained by limited access in resource-poor regions. Therefore, achieving drug-free remission has become a major focus of research in HIV therapeutics [3,4]. Therapeutic HIV-1 vaccines directed to the cell-mediated immune system could boost HIV-specific immune responses and improve virologic control in the absence of ART [5].

AIDS Clinical Trials Group (ACTG) protocol A5197 was a randomized, placebo-controlled trial to test the effect of a recombinant adeno-associated serotype 5 (rAd5) HIV-1 gag therapeutic vaccine on plasma viral load (pVL) in subjects undergoing an analytic treatment interruption (ATI) [6]. A total of 110 participants underwent a 16-week ATI after randomization in a 2:1 ratio to receive either three doses of vaccine or placebo. The vaccine induced significant CD4+ and CD8+ HIV-1 Gag-specific T-cell responses in a subset of participants and marginally significant decreases in the level of viremia during the analytic treatment interruption. Vaccination was associated with lower ATI viral load even after controlling for viral and host genetic factors [7]. In addition, the magnitude of detectable HIV-1 Gag-specific CD4+ and CD8+ IFN-γ-producing cells was negatively correlated with viral load set point [6].

The goals of this analysis were to characterize study participants with initial virologic suppression, evaluate viral and immunologic correlates of such a response, and determine the durability of virologic control 49 weeks after treatment interruption.

Methods

Patients and Study Design

Study design and patient inclusion criteria for ACTG A5197 have been described in detail [6]. Eligible participants were on ART with CD4+ cell counts ≥500/mm³, plasma HIV-1 RNA levels of ≤50 copies/mL at screening with a history of pVL ≥500 copies/mL for 24 months prior to enrollment. Participants received a rAd5 vaccine containing an HIV-1 gag insert or placebo at weeks 0, 4, and 26 (Step I). Participants were randomized to receive vaccine or placebo in a 1:1 ratio and were followed for 16 weeks (Step II). Participants underwent a 16-week ATI after randomization in a 2:1 ratio to receive either three doses of vaccine or placebo. The vaccine induced significant CD4+ and CD8+ HIV-1 Gag-specific T-cell responses in a subset of participants and marginally significant decreases in the level of viremia during the analytic treatment interruption. Vaccination was associated with lower ATI viral load even after controlling for viral and host genetic factors [7]. In addition, the magnitude of detectable HIV-1 Gag-specific CD4+ and CD8+ IFN-γ-producing cells was negatively correlated with viral load set point [6].

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Results

Characteristics of Initial Virologic Suppression

Of the 104 participants with evaluable pVL set point, 11 were found to have a pVL set point <3.0 log_{10} copies/mL and were categorized as an initial virologic suppressor (Table 1). Eighty-two percent (9/11) of initial virologic suppressors received the vaccine as compared to 66% (61/93) of the non-suppressors (P = 0.5). Initial virologic suppressors had a median pVL set point of 2.6 log_{10} RNA copies/mL versus a pVL set point of 4.2 log_{10} RNA copies/mL for virologic non-suppressors. The time on ART and distribution of HLA allele groups (protective, neutral, unfavorable) was not significantly different between the initial virologic

Intracellular Cytokine Staining Assays (ICS) and Flow Cytometric Analysis

Cell-mediated immune responses at study weeks 0, 8, and 38 were evaluated by an intracellular cytokine staining assay for interferon-gamma (IFN-γ), tumor necrosis factor-alpha (TNF-α), and interleukin-2 (IL-2) as previously described [6,14,15]. Peripheral blood lymphocytes were first exposed to a Gag peptide pool for 18 hours. CD4+ and CD8+ T cells producing cytokines and expressing cytotoxic T lymphocyte antigen 4 (CTLA-4) or programmed death-1 (PD-1) were detected by multiparameter flow cytometry. A “positive change” in the numbers of CD4+ or CD8+ IFN-γ-producing T cells in response to Gag stimulation was defined as at least a two-fold increase from baseline to week 38.

Viral sequencing from Plasma and Sequence Analysis

Most participants had plasma viral sequences analyzed at three time points: at the time of initial detectable viremia (in general, ATI weeks 2–7), ATI week 16 (study week 54), and ATI week 49 (study week 87). The Gag and reverse transcriptase (RT) coding sequences were amplified from plasma HIV-1 RNA by nested RT-PCR using gene-specific primers. Population (“bulk”) sequencing was performed on an ABI 3730 automated DNA sequencer. Chromatograms were analyzed using Sequencher (Gene Codes). We calculated the number of amino acid mismatches between the vaccine or consensus HIV-1 subtype B sequence and the patient-derived sequence. HLA-associated polymorphisms in patient HIV-1 sequences were determined based on a published list [16].

Statistical Analysis

A comparison of host genetic, immunologic, and viral sequence characteristics of those with and without initial viral suppression was performed using the 2-sample Wilcoxon rank sum test. Fisher’s exact tests were used to compare dichotomous outcomes between initial viral suppressors and the non-suppressors. All of the p-values are exact 2-sided p-values. No adjustments were performed for multiple comparisons. Note that the comparisons presented here were not originally planned in the protocol.

Characterization of Initial Virologic Suppression

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suppressor and non-suppressor groups. When compared to CD4+ cell counts at study entry, participants with initial virologic suppression had a median gain of 7 CD4+ cells/mm³ at ATI week 16. This was in contrast to a median loss of 247 CD4+ cells/mm³ among non-suppressors (P = 0.04).

Initial Virologic Suppression and Immune Preservation was not Sustained

Of the 10 participants with initial virologic suppression and a measured pVL at ATI week 49 off of ART, only 3 subjects continued to have a pVL <3.0 log₁₀ copies/ml (Table 2, Figure 1). Two of the individuals with sustained virologic control had protective HLA alleles: One participant was found to have HLA B*27 and B*57, while another had B*27. For participants with initial virologic suppression, the median CD4+ cell decline between ATI weeks 16 and 49 was 82 cells/mm³ (N = 9) as compared to a decline of 99 cells/mm³ for initial non-suppressors (N = 52, P = 0.4).
available ATI time point (median 0.44 versus 0.45, respectively, \( P = 0.4 \)) or at ATI week 49 (median 0.48 versus 0.47, respectively, \( P = 0.9 \)). This finding suggests that the lack of sustained pVL suppression in the latter group could not be explained by the accumulation of HLA-associated escape mutations in Gag.

The number of predicted amino acid mismatches was calculated between the vaccine *gag* sequence and the earliest available patient-derived HIV-1 sequence after ATI (median 4 weeks). These early sequences represent viral populations least likely to have been shaped by significant vaccine-driven immune responses. There was no significant difference in the number of Gag amino acid mismatches between the vaccine and patient HIV-1 sequences among initial virologic suppressors and non-suppressors who received the vaccine (median 33 versus 29 amino acid mismatches, respectively, \( P = 0.19 \)). The change in the number of mismatched amino acids between the two time points and no significant differences between initial virologic suppressors and non-suppressors (mean 0 vs. 0.24 amino acid changes, respectively, \( P = 0.59 \)).

### Immunologic Factors Associated with Initial Virologic Suppression

There was no significant difference between initial virologic suppressors and non-suppressors in the CD4\(^+\) T-cell count at study entry (median 846 versus 839 cells/mm\(^3\), \( P = 0.58 \)). In the initial A5197 analysis, an inverse association was seen between the ATI set point viral load and the number of HIV-1 Gag-specific CD4\(^+\) IFN-\(\gamma\)-producing CD4\(^+\) T cells at study weeks 8 and 38 [6].

![Figure 1. Viral load trends for individuals with ATI set point viral load <3.0 log10 copies/ml.](image)

The light yellow highlighted region encompass the HIV-1 viral load measurements used to calculate the viral set point (mean of the ATI weeks 12 and 16 HIV-1 RNA copies/ml). Pre-ARV, pre-antiretroviral therapy; PID, patient identification number. doi:10.1371/journal.pone.0034134.g001

### Table 2. Characteristics of initial viral suppressors.

| PID | Study Arm | HLA group | Pre-ARV RNA copies/ml (log\(_{10}\)) | ATI Set Point RNA copies/ml (log\(_{10}\)) | ATI week 49 RNA copies/ml (log\(_{10}\)) |
|-----|-----------|-----------|-------------------------------------|------------------------------------------|----------------------------------------|
| 1   | Vaccine   | Protective| 3.3                                 | 2.9                                      | 2.7                                    |
| 2   | Vaccine   | Protective| 2.4                                 | 4.8                                      |                                        |
| 3   | Vaccine   | Protective| 5.1                                 | 2.7                                      | 2.6                                    |
| 4   | Vaccine   | Neutral   | 3.7                                 | 2.4                                      | 3.5                                    |
| 5   | Vaccine   | Neutral   | 1.7                                 | 1.7                                      |                                        |
| 6   | Vaccine   | Neutral   | 4.0                                 | 2.9                                      | 3.9                                    |
| 7   | Vaccine   | Neutral   | 3.8                                 | 2.6                                      | 3.6                                    |
| 8   | Vaccine   | Neutral   | 4.8                                 | 2.99                                     | 3.7                                    |
| 9   | Vaccine   | unfavorable| 2.8                                 | 4.1                                      |                                        |
| 10  | Placebo   | Neutral   | 5.7                                 | 2.1                                      | 4.8                                    |
| 11  | Placebo   | Neutral   | 2.6                                 | 1.7                                      |                                        |

PID, patient identification number.
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Viral Suppression after Therapeutic Vaccination

There were no significant differences between initial virologic suppressors and non-suppressors in the number of HIV-1 Gag-specific IFN-γ-producing CD4+ T-cells at week 8 or 38 (week 8 median 2.4 versus 2.2 log_{10} cells/10^6 lymphocytes, \( P = 0.21 \); week 38 median 2.3 versus 2.2 log_{10} cells/10^6 lymphocytes, \( P = 0.20 \)), or Gag-specific IFN-γ-producing CD8+ T-cells (week 8 median 2.8 versus 2.9 log_{10} cells/10^6 lymphocytes, \( P = 0.35 \); week 30 median 2.9 versus 2.9 log_{10} cells/10^6 lymphocytes, \( P = 0.55 \)). Similarly, no significant differences were detected between initial virologic suppressors and non-suppressors in the number of subjects who had an increase in HIV-1 Gag-specific IFN-γ-producing CD4+ and CD8+ T-cells from study entry to week 38. Thirty percent (3/10) of initial virologic suppressors had a significant increase in the number of CD4+ IFN-γ-producing cells between baseline and week 38 as compared with 35% (31/89) non-suppressors (\( P = 1.0 \)). Fifty percent (5/10) of initial virologic suppressors had a significant increase in the number of CD8+ IFN-γ-producing cells as compared with 38% (34/89) of non-suppressors (\( P = 0.31 \)).

The number of HIV-1 Gag-specific CD4+ IFN-γ-producing cells detected was associated with vaccination status, but not with status of initial virologic suppression. There were no differences in the magnitude of HIV-1 Gag-specific CD4+ IFN-γ-producing cells between vaccinated participants with or without initial virologic suppression (median 2.4 versus 2.3 log_{10} cells/10^6 lymphocytes, \( P = 0.68 \) at week 8, and median 2.5 versus 2.3 log_{10} cells/10^6 lymphocytes, \( P = 0.15 \) at week 38, Figure 2). In addition, vaccinated participants regardless of status of initial virologic suppression were found to have higher levels of HIV-1 Gag-specific IFN-γ-producing cells at week 8 compared to initial non-suppressors who had received placebo (Figure 2). These results indicate that the magnitude of in vitro CD4+ IFN-γ responses to HIV-1 Gag peptides may have been influenced by therapeutic vaccination, but was not clearly correlated with initial virologic suppression.

The association of initial virologic suppression with expression of the immunomodulatory molecules CTLA-4 and PD-1 on CD4+ and CD8+ cells expressing either TNF-α, IFN-γ, or IL-2 were evaluated in a subset of participants at both study entry and study week 38. At week 38, participants with initial virologic suppression had significantly lower proportions of CD4+ T cells expressing CTLA-4 (median 8.9% [N = 6] versus 14.1% [N = 46], \( P = 0.02 \), Figure 3). No significant differences were seen in the expression of CD4+ PD-1+ T cells between those with and without initial virologic suppression (median 10.4% [N = 6] versus 9.1% [N = 46], \( P = 0.67 \)). However, participants with initial virologic suppression had a significantly higher percentage of Gag-specific CD4+ TNF-α+ cells expressing either CTLA-4 or PD-1 (median CTLA-4 expression 54% [N = 5] for initial suppressors versus 33% [N = 44] for non-suppressors, \( P = 0.01 \); and median PD-1 expression 36% [N = 5] for suppressors versus 14% [N = 44] for non-suppressors, \( P = 0.04 \); Figure 4). No significant differences were seen between the groups in either the expression of other cytokines in CD4+ T cells or in any CD8+ T cell populations.

**Discussion**

In ACTG A5197, vaccination with a rAd5 HIV-1 gag therapeutic vaccine was associated with increased HIV-specific T-cell activation and a trend towards improved virologic control during the analytic treatment interruption. In this follow-up study, we describe 11 subjects with viral load set point under 3.0 log_{10} copies/ml including 9 subjects who received the vaccine. No virologic differences were identified in participants with and without initial virologic suppression, but those with initial virologic suppression were found to have a lower proportion of CD4+ T cells expressing CTLA-4 prior to treatment interruption, and a greater proportion of HIV-1 gag-reactive CD4+ TNF-α+ cells expressing either CTLA-4 or PD-1. Virologic suppression, however, was not sustained in the majority of subjects with initial virologic control.

Participants with initial virologic suppression were found to have an initial immunologic benefit whereas non-suppressors had substantial CD4+ cell declines over the initial 16 weeks of the analytic treatment interruption. However, this initial virologic control was not sustained in the majority of initial suppressors and was associated with a CD4+ cell decline over the subsequent 33 weeks. Potential explanations for the loss of virologic control include the waning of vaccine-induced immune function over time (cellular immune responses were only evaluated early in the ATI), and differing characteristics (e.g. size, heterogeneity) of the latent HIV-1 reservoir from which the rebounding virus originated. Although we detected no significant differences between initial virologic suppressors and non-suppressors in the number of accumulated HLA-associated HIV-1 polymorphisms, we cannot rule-out the possibility that qualitative differences exist between escape mutations with differential impact on viral fitness.

Three participants were able to maintain virologic suppression 49 weeks after treatment interruption. All three individuals had received study vaccine and two of the three had CD4+ cell counts at ATI week 49 above that found at study entry. In evaluating which virologic or immunologic characteristics may be predictive of initial virologic suppression, we found a trend toward lower pre-ART viral load for participants with initial virologic suppression. Pre-ART viral load has also been seen in other studies to be associated with the extent of viral rebound [17–19] and validates the use of stratification by viral load at randomization in A5197.

Two of the three participants who maintained virologic suppression were also found to have protective HLA alleles. HLA class I molecules mediate the cell-mediated immune response to HIV and play a crucial role in the immune control of HIV. Certain HLA alleles, termed “protective”, have been associated with decreased viral load set point [20–23] and delayed disease progression [24]. In addition, in the STEP trial of the rAd5 Gag-Pol-Nef vaccine, participants in the vaccine arm with protective HLA alleles were found to have significantly lower viral load set point after infection [25]. However, the impact of HLA alleles on initial virologic rebound during treatment interruption is far less clear. In this study, the prevalence of individuals with protective HLA alleles was no higher in the initial suppressor group as compared to the non-suppressor group (27% vs. 36%). One explanation is that in these chronically infected individuals, HIV has had an opportunity to adapt to the host immune response as evidenced by the fact that almost all individuals in the study were found to have accumulated HLA-associated polymorphisms in HIV-1 Gag.

The magnitude of HIV-1 gag-specific IFN-γ-producing CD4+ T cells has been previously associated with virologic control [26–28]. It has been hoped that a therapeutic vaccine-induced augmentation of such a response would lead to improved viral control. However, in a recent study of a recombinant canarypox HIV-1 vaccine (vCP1452), patients exposed to the vaccine had a worse outcome including higher levels of viral replication [29]. A subsequent analysis suggested that the extent of vaccine-induced activation of HIV-specific CD4+ T cells was associated with the detrimental outcome [30]. In contrast, we found no evidence of an adverse effect of HIV-specific CD4+ T-cell activation on plasma viremia. In addition, the initial analysis of A5197 found that a greater number of gag-specific IFN-γ-producing CD4+ T cells were associated with lower viral rebound [6].
CTLA-4 and PD-1 are inhibitory immunoregulatory molecules that regulate T cell activation and peripheral immune tolerance [31]. Their expression on HIV-specific CD4+ T cells has been associated with increased viral load and disease progression [32,33]. As might be expected, we found that CD4+ T cells from initial virologic suppressors had a lower expression of CTLA-4 immediately prior to the ATI. We found, however, that CTLA-4+ and PD-1+ cells from initial virologic suppressors made up a greater proportion of HIV-1 Gag-specific CD4+ TNF-α+ T cells than those from non-suppressors. One potential explanation may be that the subpopulation of CD4+ T cells expressing TNF-α and CTLA-4 or PD-1 may be less readily able to support productive HIV-1 infection despite evidence that CTLA-4 signaling may be associated with increased CCR5 expression and enhanced

Figure 2. HIV-1 Gag-specific CD4+ IFN-γ-producing T cells at weeks 8 and 38 categorized by initial virologic suppression and treatment arm. Panel A, CD4+ IFN-γ response 8 weeks after receiving the first vaccine dose. Number of cells quantified by the intracellular cytokine staining assay in response to pooled HIV Gag peptides. Panel B, CD4+ IFN-γ response 38 weeks after receiving the first vaccine dose and immediately prior to initiating the analytic treatment interruption. Statistical comparison not provided for participants in the “Suppressors (Placebo)” group as only two participants fell into this category.

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Figure 3. Expression of HIV-1 CD4+ T cells expressing either CTLA-4+ or PD-1+ by participants with and without initial virologic suppression. Panel A, Percentage of CD4+ T cells expressing CTLA-4 at week 38 detected using flow cytometry. Panel B, Percentage of CD4+ T cells expressing PD-1 at week 38 detected using flow cytometry.

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susceptibility to infection [34]. An alternative explanation is that these cells may serve to augment the immune suppression of viral replication or may reflect a more active antiviral response in other compartments such as lymphoid or mucosal tissue. Characterization of these T cell subsets in other HIV-infected populations is needed to investigate further the importance of this exploratory finding.

This post-hoc analysis has several limitations. Only a subset of participants were initial virologic suppressors, which limited our ability to identify significant viral and immunologic predictors of virologic control. The analysis of viral load and CD4+ cell counts between ATI weeks 16 and 49 may be confounded by selection bias, especially in the non-suppressor group. After the ATI week 16 time point, participants had the option of restarting ART and those with particularly high viral loads or low CD4+ cell counts were encouraged to do so. Therefore, the viral load increases and CD4+ cell declines in the non-suppressor group are likely to be underestimated, which may have obscured continued viral load and CD4+ T cell benefits in the initial suppressor group. Immunologic studies on the magnitude of CTLA-4 and PD-1 expression on CD4+ and PD-1 expression of CD4+ cells were performed on a subset of participants, which may have limited our ability to detect a significant difference between groups.

An effective HIV-1 therapeutic vaccine would be a significant advance in our efforts at HIV eradication and could provide insight into the optimal preventative vaccine strategy. In ACTG A5197, we found that participants with initial virologic suppression had a lower proportion of CD4+ T cells expressing CTLA-4 and a higher percentage of Gag-specific CD4+ TNF-α+ T cells expressing either CTLA-4 or PD-1. Further studies are needed to determine whether optimizing CTLA-4 and PD-1 expression on CD4+ T cells will improve virologic control in recipients of other therapeutic vaccines in development.

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Author Contributions

Conceived and designed the experiments: JZL MML ZLB DRK. Performed the experiments: JZL CJB MML MC KM. Analyzed the data: JZL CJB ZLB HW JS. Contributed reagents/materials/analysis tools: MML MNR MC BDW RTS DRK. Wrote the paper: JZL CJB MML ZLB HW JS MNR MC RTS DRK.

References

1. Richman DD, Margolis DM, Delaney M, Greene WC, Hazuda D, et al. (2009) The challenge of finding a cure for HIV infection. Science 323: 1304–1307.
2. Thompson MA, Aberg JA, Cahn P, Montaner JS, Rizzardi G, et al. (2010) Antiretroviral treatment of adult HIV infection: 2010 recommendations of the International AIDS Society-USA panel. JAMA 304: 321–333.
3. Johnston R (2010) HIV cure: controversy, consensus, and a consortium. AIDS 24: 1007–1012.
4. Folkers GK, Fauci AS (2010) Controlling and ultimately ending the HIV/AIDS pandemic: a feasible goal. JAMA 304: 350–351.
5. Lederman MM, Penen-Nicholson A, Stone SF, Sieg SF, Rodriguez B (2007) Monitoring clinical trials of therapeutic vaccines in HIV infection: role of treatment interruption. Curr Opin HIV AIDS 2: 56–61.
6. Schooley RT, Spritzler J, Wang H, Lederman MM, Hadler D, et al. (2010) AIDS clinical trials group 5197: a placebo-controlled trial of immunization of HIV-1-infected persons with a replication-deficient adenovirus type 5 vaccine expressing the HIV-1 core protein. J Infect Dis 202: 705–716.
7. Li JZ, Brunme ZL, Brunme CJ, Wang H, Spritzer J, et al. (2011) Factors associated with viral rebound in HIV-1-infected individuals enrolled in a therapeutic HIV-1 gag vaccine trial. J Infect Dis 203: 976–983.
8. van den Berg-Wolf M, Hulsker KI, Peng G, Kozal MJ, Novak RM, et al. (2008) Virologic, immunologic, clinical, safety, and resistance outcomes from a long-term comparison of efavirenz-based versus nevirapine-based antiretroviral regimens as initial therapy in HIV-1-infected persons. HIV Clin Trials 9: 324–336.
9. Murphy RA, Sunpath H, Lu Z, Chelin N, Losina E, et al. (2010) Outcomes after virologic failure of first-line ART in South Africa. AIDS 24: 1007–1012.
10. Babiker A, Castro née Green H, Compagnucci A, Fiscus S, Giampaio C, et al. (2011) First-line antiretroviral therapy with a protease inhibitor versus non-nucleoside reverse transcriptase inhibitor and switch at higher versus low viral

Figure 4. Expression of HIV-1 Gag-specific CD4+ TNF-α+ T cells expressing either CTLA-4+ or PD-1+ by participants with and without initial virologic suppression. Panel A, Percentage of CD4+TNF-α+ T cells expressing CTLA-4 at week 38. Panel B, Percentage of CD4+TNF-α+ T cells expressing PD-1 at week 38. doi:10.1371/journal.pone.0034134.g004
lead in HIV-infected children: an open-label, randomised phase 2/3 trial. Lancet Infect Dis 11: 273–283.
11. Miller LG, Golim CI, Liu H, Rady RD, Hua J, et al. (2004) No evidence of an association between transient HIV viremia ("Blips") and lower adherence to the antiretroviral medication regimen. J Infect Dis 189: 1467–1496.
12. Sungkanuparph S, Overton ET, Sefried W, Groger RK, Fraser VJ, et al. (2005) Intermittent episodes of detectable HIV viremia in patients receiving nonnucleoside reverse-transcriptase inhibitor-based or protease inhibitor-based highly active antiretroviral therapy regimens are equal in incidence and prognosis. Clin Infect Dis 41: 1326–1332.
13. Gao X, Nelson GW, Karacki P, Martin MP, Phair J, et al. (2001) Effect of a single amino acid change in HLA class I molecules on the rate of progression to AIDS. N Engl J Med 344: 1668–1675.
14. Tobery TW, Dubey SA, Anderson K, Freed DG, Cox KS, et al. (2006) A comparison of standard immunogenicity assays for monitoring HIV type 1 gag-specific T cell responses in Ad5 HIV Type 1 gag vaccinated human subjects. AIDS Res Hum Retroviruses 22: 1081–1090.
15. Trigona WL, Clair JH, Persaud N, Punt K, Bachinsky M, et al. (2003) Intracellular staining for HIV-specific IFN-gamma production: statistical analyses establish reproducibility and criteria for distinguishing positive responses. J Interferon Cytokine Res 23: 369–377.
16. Brumme ZL, John M, Carlsson JM, Brumme C, Chan D, et al. (2009) HLA-associated immune escape pathways in HIV-1 subtype B Gag, Pol and Nef proteins. PLoS ONE 4: e6687.
17. Le Moing V, Chene G, Carrier MP, Alioun A, Brun-Beuzin F, et al. (2002) Predictors of virological rebound in HIV-1-infected patients initiating a protease inhibitor-containing regimen. AIDS 16: 21–29.
18. Ioannidou JP, Havlir DV, Tchus P, Hirsch MS, Collier AC, et al. (2000) Dynamics of HIV-1 viral load rebound among patients with previous suppression of viral replication. AIDS 14: 1481–1488.
19. Fagard C, Orenius A, Gutthard H, Garcia F, Le Braz M, et al. (2003) A prospective trial of structured treatment interruptions in human immunodeficiency virus infection. Arch Intern Med 163: 1220–1226.
20. Alfie M, Adoko MM, Rosenberg ES, Hecht FM, Lee PK, et al. (2005) Influence of HLA-B57 on clinical presentation and viral control during acute HIV-1 infection. AIDS 17: 2581–2591.
21. Kiepiela P, Leslie A, Pereyra F, Zaunders J, Mackey EW, et al. (2007) Upregulation of CTLA-4 by HIV-specific CD4+ T cells correlates with disease progression and defines a reversible immune dysfunction. Nat Immunol 8: 1246–1254.
22. Riley JL, Schildner K, Blair PJ, Carreno B, Craighead N, et al. (2000) Modulation of susceptibility to HIV-1 infection by the cytotoxic T lymphocyte antigen 4 costimulatory molecule. J Exp Med 191: 1987–1997.
23. Honeyborne I, Prendergast A, Pereyra F, Leslie A, Caostrad H, et al. (2007) Control of human immunodeficiency virus type 1 is associated with HLA-B*13 and targeting of multiple gag-specific CD8+ T-cell epitopes. J Virol 81: 3667–3672.
24. Pereyra F, Jia X, McLernon P, Telenii A, de Bakker PJ, et al. (2010) The major genetic determinants of HIV-1 control affect HLA class I peptide presentation. Science 330: 1551–1557.