In vivo Magnetic Resonance Imaging of Tumor Protease Activity

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Increased expression of cathepsins has diagnostic as well as prognostic value in several types of cancer. Here, we demonstrate a novel magnetic resonance imaging (MRI) method, which uses poly-L-glutamate (PLG) as an MRI probe to map cathepsin expression in vivo, in a rat brain tumor model. This noninvasive, high-resolution and non-radioactive method exploits the differences in the CEST signals of PLG in the native form and cathepsin mediated cleaved form. The method was validated in phantoms with known physiological concentrations, in tumor cells and in an animal model of brain tumor along with immunohistochemical analysis. Potential applications in tumor diagnosis and evaluation of therapeutic response are outlined.

Many tumors have been shown to exhibit elevated levels of proteases in incipient stages, which are thought to play a crucial role in tumor angiogenesis, invasion and metastasis. Knowledge of the roles of proteases in tumor progression will potentially guide the development of novel diagnostic tools for early detection of tumors as well as disease modifying therapeutic strategies to combat tumors. Cathepsins, cysteine family proteases, are lysosomal proteases involved in cellular protein degradation and are over-expressed in many tumor cells as well as host cells associated with the tumor. Out of eleven types of cathepsins, cathepsin B (CtB) and cathepsin L (CtL) are predominantly involved in protein degradation within the lysosome. These cathepsins have been shown to be prognostic indicators of many types of tumors. The wide availability of cathepsins on tumor cells makes them attractive targets for tumor detection as well as for studying tumor evolution under therapy.

However, despite the tremendous significance, it is challenging to accomplish noninvasive detection of protease activity in vivo. Except for near infrared fluorescence (NIRF) optical imaging, there has been no known high resolution, noninvasive and non-radioactive methods to detect CtB activity in tumors in vivo. In spite of its high sensitivity, NIRF suffers from poor depth penetration (few mm) and cannot be translated into a three dimensional (3D) imaging tool either in large animal models or in humans. Furthermore, it requires synthesis and evaluation of toxicity of the injected compounds. Therefore, there is an unmet need for a noninvasive, high-resolution imaging technique for measuring the expression of cathepsins in tumors.

Here, for the first time, we demonstrate poly-L-glutamate (PLG) as a tumor protease activated magnetic resonance imaging (MRI) probe, which enables high resolution mapping of expression of cathepsins in tumors, in vivo. PLG is a nontoxic polymer that can be readily injected into animal models up to several millimoles (mM) without causing any adverse reactions. This noninvasive, nonradioactive method is based on the chemical exchange saturation transfer (CEST) between protons attached to nitrogen in amine moieties with bulk water protons. Owing to very slow exchange rates of backbone amide protons (Fig. 1a), PLG in its native form does not exhibit appreciable CEST under physiological conditions. However, cleavage by the lysosomal enzymes such as cathepsins, results in individual glutamate (Glu) moieties as well as smaller Glu peptides, which readily exhibit CEST effect from their amine protons (Fig. 1b). Determination of the relationship between the protease expression and PLG degradation via CEST can establish a potential noninvasive MRI method for quantifying tumor protease expression.
Results and Discussion

The CEST effect from Glu and PLG with and without addition of CtB is evaluated in in vitro at frequency offset, $\Delta_0 \approx 3$ ppm downfield to bulk water (Fig. 2a). Glu showed a strong CEST contrast due to its exchangeable amine protons$^{18}$ while no CEST contrast was observed from PLG due to very few amine protons. But the cleavage products of PLG showed an appreciable CEST contrast due to the exposure of amine protons interacting with bulk water after enzymatic CtB cleavage (Fig. 2a).

PLG cleavage by CtB can be monitored in vitro through proton magnetic resonance spectroscopy (1HMRS; Figs. 2 b–d). The 1HMRS spectrum from Glu showed three distinct peaks respectively at ~2.02 ppm, ~2.25 ppm and ~3.66 ppm (Fig. 2b). PLG also showed three peaks at ~2.01 ppm, ~2.28 ppm and ~4.1 ppm (Fig. 2c). Addition of CtB splits the 2.28 ppm and 4.1 ppm peaks (Fig. 2d). The splitting of PLG peaks denotes its cleavage by CtB.

Further, we assessed the expression of CtB in 9L tumor cells. CEST imaging of 9L cells was performed with and without addition of PLG. With addition of PLG, the 9L cell line showed ~20% increase in CEST contrast compared to the cells without PLG, while there was no appreciable CEST contrast observed from PLG alone (Figs. 3a–c). The increased CEST contrast can be attributed to the cleavage of PLG by the cathepsins expressed in tumor cells. Since the two cathepsins i.e. CtB and CtL are primarily responsible for cleavage of PLG, Western blot analysis of 9L cell lysates was performed to determine the presence of CtB and CtL expression (Fig. 3d). Western blot analysis showed expression of CtB both in the mature and pro form. However, mature CtB was predominately expressed than its pro form. The CtL was expressed only in the pro form. This suggests that cleavage of PLG in 9L cells is primarily due to the expression of mature CtB.

The feasibility of mapping CtB expression in in vivo was evaluated on a 9L rat brain gliosarcoma model (Figs. 3 e–g, Figs. 4 a–e). PLG (160 mg/kg) was injected in these rats through the tail vein and CEST imaging was performed up to 90 minutes with a 30 minutes time interval. A continuous increase in CEST contrast was observed in the tumor region due to the cleavage of PLG by protease enzyme (primarily CtB) expressed in tumor. We further performed immunohistochemistry of rat brain tumor tissues for CtB expression. A strong positive expression of CtB was observed in the tumor region while the normal appearing brain (NAB) showed negligible expression of CtB. The same tumor region showed higher nuclear density and mitosis reflecting increased cellular proliferation than NAB region on hematoxylin and eosin stain (Figs. 4f–i). The graph (Fig. 4j) shows the mean (±SD) CEST contrast from tumor (n=6) as well as NAB region (n=6). A progressive increase in CEST contrast from tumor region was observed over the period of 90 minutes, while the CEST contrast from NAB did not change appreciably over the time course of 90 minutes. An average of 19% increase in CEST contrast was observed from tumor region after 90 minutes injection of PLG (Fig. 4k).

Since PLG mediated CEST contrast directly depends on the expression of cathepsin proteases, monitoring changes in the CEST contrast in vivo will enable the quantitation of cathepsin expression in different tumor types.

Conjugation of polymers to anticancer drugs is an emerging approach for clinical development of combination therapy$^{19}$. Such studies have the potential for advancing polymer therapeutics as a novel class of anticancer agents. These techniques primarily exploit the degradation of polymers by the lysosomal enzymes such as cysteine proteases, particularly CtB and CtL$^{16,19-22}$. In particular, PLG has been used as a macromolecular carrier for drug delivery$^{17}$. The results from this study suggest the potential of PLG as a CEST contrast agent for in vivo imaging.
and PLG in combination with paclitaxel (PG-TXL) has advanced to clinical trials\textsuperscript{23,24}. Since the development and application of targeted delivery of anticancer agents using PLG as the drug carrier is emerging\textsuperscript{23,24}, we envisage that, in the long term, the proposed CEST imaging technology has the potential to monitor PLG conjugated drug delivery and therapeutic response of tumors.

This is the first study to show that PLG can be used as a protease activated MRI probe for tumor mapping in vivo and PLG degradation by cathepsins in tumors can be imaged with high resolution MRI. We expect that the current method will have numerous applications in cancer research including differentiation of radiation necrosis from tumor growth as well evaluating anti-autophagy therapies.

![Figure 2](image-url)

**Figure 2** | PLG as a marker of cathepsin B (CtB) enzyme activity. (a). Anatomical proton density weighted MR image showing three different solutions (Glu, PLG, and PLG + CtB) in NMR tubes immersed in a PBS phantom. (b). CEST contrast map at 3 ppm downfield to the bulk water from glutamate (Glu). PLG and PLG with CtB. Glu shows a strong CEST contrast. No CEST contrast was observed from PLG alone. After addition of CtB an appreciable CEST contrast was observed, this is due to the cleavage of PLG into smaller fragments by CtB. This cleavage exposes significant number of labile $-\text{NH}_3$ protons, which can be easily detected using CEST MRI. (c-e). $^1$H MR spectra from phantoms containing Glu, PLG, and PLG + CtB. Spectrum from Glu shows multiple peaks related to aliphatic protons at $\sim 2.02, \sim 2.25$, and 3.66 ppm. Spectrum from PLG shows 3 peaks respectively at $\sim 2.02, \sim 2.28$, and 4.1 ppm. The spectra are broad due to reduced mobility of PLG molecule and hence the fine structure is masked. Phantom with PLG and CtB is showing splitting of 2.28 and 4.1 ppm peaks, due to cleavage of PLG by CtB.

![Figure 3](image-url)

**Figure 3** | PLG as a marker of protease enzyme expression in cancer 9L cells. (a). Cancer cells cultured without any PLG. This image shows $\sim 22\%$ CEST contrast, which is due to endogenous Glu in cultured cells. (b). Same cell line (as in a) cultured in the presence of PLG (20 mg/ml) for 1 hour shows an further elevation of $\sim 17\%$ of the CEST contrast over the baseline indicating that the CtB present in the tumor cells cleaved the PLG. (c). PLG alone in saline does not show any appreciable CEST effect owing to the slow exchange of amide protons present on the PLG. (d). Western blot analysis of tumor cell lysates shows both mature and pro form of CtB while cathepsin L (CtL) was expressed only in the pro form. The cleavage of PLG in this tumor cell line is predominantly due to CtB. (e), (f). Baseline anatomical CEST weighted image and CEST map from rat brain with a 9L tumor. (g). At 60 minutes post intra-venous injection of PLG (160 mg/kg), a $\sim 32\%$ increase in GluCEST contrast was observed in the tumor region.
Figure 4 | In vivo imaging of cathepsin expression by monitoring PLG cleavage mediated CEST. (a). Anatomical CEST weighted image showing the tumor as a hyper-intense region. (b). the baseline CEST contrast map from the same slice. (c–e). CEST contrast maps of the same imaging slice respectively at 30, 60 and 90 minutes following PLG administration through tail vein. Following PLG injection a continuous increase in CEST contrast up to 90 minutes was observed mainly in tumor region due to cleavage of PLG into its monomer and/or smaller peptide fragments by cathepsins expressed in tumor. (f). (g). Immunohistochemistry of CtB from normal appearing brain (NAB) region and tumor region showing the expression of CtB (40X). NAB region shows very few cells positive for CtB expression while the tumor cells showed strong positivity for CtB. (h). (i). Hematoxylin and eosin staining from NAB and tumor regions show higher cell proliferation in tumor. (j). Graph showing PLG kinetics in the tumor (n = 6) and NAB (n = 6) for a period of 90 minutes following the injection of PLG. (k). The graph shows the mean percentage change in the CEST contrast following PLG injection. While no appreciable change in NAB is observed, tumor shows 19% increase in CEST contrast in about 90 minutes. One way analysis of variance showed a significant change (p = 0.048) in CEST contrast over the time period of 90 minutes.

Methods

All animal experiments were performed according to an approved University of Pennsylvania Institutional Animal Care and Use Committee Protocol.

In vitro Study. Phantom Experiment. Solutions of 20 mg/ml PLG were prepared with and without addition of 0.5 mg/ml CtB enzyme (Sigma Aldrich, USA) in 10 mm NMR tubes. In addition, 10 mM sodium glutamate solution was also prepared. These phantoms were placed in a larger bottle containing phosphate buffered saline (PBS) and the CEST imaging of these phantoms was performed at room temperature on a 7T whole body MRI scanner (Siemens Medical Systems, Malvern, PA, USA).

For CEST imaging a frequency selective saturation pulse followed by a segmented radiofrequency spoiled gradient echo (GRE) readout sequence was used. The sequence parameters were: slice thickness = 10 mm, flip angle = 10°, TR = 5.6 ms, TE = 2.7 ms, field of view = 100 × 100 mm², matrix size = 192 × 192, and one saturation pulse and 64 segments acquired every 10 s. CEST images were collected at peak B1max of 3.64 μT and 1 s saturation duration for the frequencies (+2.4 to +3.6 ppm and −2.4 to −3.6 ppm in 0.2 ppm steps) from bulk water resonance. Radiofrequency field inhomogeneity (B1) and static magnetic field inhomogeneity (B0) maps were also obtained to correct for any inhomogeneities as described previously46. Single voxel spectra (SVS) with point-resolved-spectroscopy (PRESS) acquisition mode were performed from each phantom using the following parameters: voxel size = 5 mm × 5 mm × 20 mm (Voxel volume 500 μL), spectral width = 4 kHz, number of points = 2048, averages = 64, TE = 20 ms, and TR = 3 s. Water suppression was achieved using the variable pulse power and optimized relaxation delays method (VAPOR).

Cell experiments. Rat 9L gliosarcoma cells were cultured in DMEM containing 10% Fetal Bovine Serum, containing Penicillin-streptomycin. 9L cells growing in log phase were trypsinized and used for in vitro and in vivo studies.

Ten million 9L cells were trypsinized and washed with sterile PBS and transferred to a 0.5 ml microfuge tube. Six tubes with equal volume of 9L cells were obtained for MR imaging. These tubes were categorized in two groups. In the first group (n = 3), 0.1 ml of normal saline was added, while in the 2nd group (n = 3) 0.1 ml of PLG solution was added making a total concentration of 20 mg/ml. CEST imaging was performed on a 9.4 T horizontal bore small animal MR scanner (Varian, Palo Alto, CA). CEST imaging was performed using a custom-programmed segmented radio-frequency GRE readout pulse sequence with a frequency selective continuous wave saturation preparation pulse. The sequence parameters were: field of view = 30 × 30 mm², slice thickness = 2 mm, flip angle = 15 degree, TR = 6.2 ms, TE = 2.9 ms, matrix size = 128 × 128. For every 8 s one saturation pulse and 128 acquisition segments were applied. CEST images were collected using a 1 second saturation pulse at B1max of 5.87 μT for multiple frequencies (+2.4 to +3.6 ppm and −2.4 to −3.6 ppm in 0.2 ppm steps) from bulk water. B1 and B0 field maps were also acquired and used to correct the CEST maps as described previously46.

Western blot analysis. 9L cells were lysed in radioimmunoprecipitation assay buffer containing complete mini protease inhibitor cocktail (Roche Diagnostics, city, state, country) and protein lysate was quantitated using Pierce BCA protein assay kit (Thermo scientific). 9L cell lysate (40 μg) was boiled in sample buffer and loaded onto single lane of a 10% SDS-PAGE gel under reducing conditions. The separated proteins were blotted onto a PVDF membrane, which was blocked with 5% non-fat dry milk for 1 hour and then probed with rabbit anti-rat CtB polyclonal IgG (4 μg/ml; cross reacts with human and mouse; Abcam) or rabbit anti-human Ct L monoclonal...
Intraperitoneal injection of a ketamine (91 mg kg

Scientific Study. Rat brain tumor. To develop intracranial tumors, rat gliosarcoma cells (9L) were used. Syngeneic female Fisher rats (F344/NCR, four-six weeks old) weighing 30–150 g were used to generate tumor-bearing rats as described previously. Briefly, general anesthesia was induced by isoflurane followed by intraperitoneal injection of a ketamine (91 mg kg

In vivo study. Rat MR imaging. MRI imaging was performed on tumor bearing rats three weeks after 9L cell implantation. Rats were transferred to a 9.4T horizontal bore small animal MR scanner (Varian, Palo Alto, CA) and placed in a 20-mm diameter commercial quadrature proton coil (m2m Imaging Corp., Cleveland, OH). Animals were kept under anesthesia (1.5% isoflurane in 1 liters/min oxygen) and their body temperature maintained with the air generated and blowing through a heater (SA Instruments, Inc., Stony Brook, NY). Respiration and body temperature were continuously monitored using a MRI compatible small animal monitoring system (SA Instruments, Inc., Stony Brook, NY). The Institutional Animal Care and Use Committees of the University of Pennsylvania approved experimental protocols.

CEST imaging of rat brain tumor was performed using the pulse sequence and parameters as described in case of 9L cell line imaging except field of view 35 × 35 mm². After baseline imaging the rats were injected with 1 ml PLG (160 mg/kg) solution at a rate of 0.1 ml per minute through the catheter inserted in a tail vein. After PLG administration, CEST imaging was performed at each 30 minute time interval for a total period of 90 minutes.

At the end of the study the rats were euthanized by CO₂ asphyxiation and brain tissue was collected for immunohistochemical studies.

Immunohistochemistry. All the excised rat brains were fixed in 10% buffered formalin and embedded in paraffin. The whole brain specimen was blocked and 6 μm sections were taken for immunohistochemistry analysis. The slides were deparaffinized in Xylene (Surgipath, 03672) followed by rehydration with 100%, 95%, 70% ethanol.

Slides were taken for immunohistochemical analysis. The slides were deparaffinized in Xylene (Surgipath, 03672) followed by rehydration with 100%, 95%, 70% ethanol. Immunohistochemistry.

These slides were digitized with a 40× objective using Olympus camera.

Image Processing. All image processing and data analysis were performed using software routines written in MATLAB (version 7.5, R2007b). Acquired CEST images were corrected for B₀ and used to generate CEST contrast map using Equation [1].

\[
\text{CEST}(\%) = 100 \times \left( \frac{S_{\text{ve}} - S_{\text{ve}} - S_{\text{ve}}}{S_{\text{ve}}} \right)
\]

where \(S_{\text{ve}}\) and \(S_{\text{ve}}\) are the \(B₀\) corrected MR signals at −3 ppm and +3 ppm respectively. CEST contrast was further corrected for \(B₀\) and mapped as false-colors onto anatomical proton image. To compute the CEST contrast, region of interests were manually drawn on tumor and NAB regions.

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Author contributions M.H. designed and performed experiments, analyzed data and wrote the manuscript. A.S. helped with experiments, image processing, data analysis and contributed to manuscript editing. M.I. helped with cell line Western blotting. R.I. helped with rat brain tumor model development. K.N., R.P.R.N., C.D. and F.K. helped with phantom imaging, animal handling and manuscript editing. K.C. provided technical support with cell lines and animal imaging. H.P. advised on tumor model development and contributed to manuscript editing. D.R. provided technical support with handling the cells line and tumor model development; H.H. provided technical guidance and contributed to the manuscript editing; R.R. provided the concept, experimental design and contributed to manuscript writing and editing. All authors reviewed the manuscript.

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