Screening for Selective Anticancer Activity of 65 Extracts of Plants Collected in Western Andalusia, Spain

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Abstract: Finding cytotoxic drugs with a high selectivity towards cancer cells is crucial to improve the low survival rates of patients diagnosed with metastatic cancers. Since plants are an important source of anticancer drugs, we have screened 65 extracts from 45 plants collected in several areas of Western Andalusia (Spain) for cytotoxic activity on lung cancer cells versus lung normal cells. An extract from the leaves of *Tetraclinis articulata* (Vahl) Mast. (Cupressaceae) showed a marked cytotoxicity (IC50 = 0.37 ± 0.03 μg/mL) and selectivity (selectivity index = 378.3) against the lung cancer cells; cisplatin, 5-fluorouracil, and an extract from the leaves of *Taxus baccata* L. (Taxaceae) were less cytotoxic and selective. Extracts from *Cascabela thevetia* (L.) Lippold (Apocynaceae), *Frangula alnus* Mill. (Rhamnaceae), *Iberis ciliata* subsp. *contracta* (Pers.) Moreno (Brassicaceae), *Juniperus macrocarpa* Sm (Cupressaceae), and *Pancratium maritimum* L. (Amaryllidaceae) also showed selective cytotoxicity (selectivity index > 10). Active extracts were also tested against a panel of cancer cell lines from a variety tissues. The plants identified in this work are potential sources of natural compounds with selective toxicity towards cancer cells.

Keywords: cancer; lung cancer; selectivity; *Tetraclinis articulata* (Vahl) Mast.; Cupressaceae

1. Introduction

Despite the recent approval of new anticancer treatments, metastasis continues to be an incurable disease for most cancer patients. The limited efficacy of the existing therapies is reflected in the poor survival rates of patients diagnosed with the most common metastatic cancers [1]. For example, distant metastases occur in 57% of patients diagnosed with lung cancer, and only 6% of them survive 5 years after diagnosis [1]. The percentages are low for other common metastatic cancers. The five-year relative survival rates for patients with distant metastases are 30% in prostate cancer, 28% in breast cancer, 14% in colorectal cancer, 27% in melanoma, 13% in renal cancer, 30% in ovarian cancer, 17% in cancers of the uterine corpus, 17% in cancers of the uterine cervix, 6% in bladder cancer, 5% in esophageal cancer, 3% in liver cancer, and 3% in pancreatic cancer [1]. Many patients with metastases do not overcome the disease despite surviving five years after diagnosis.

Understanding why the current treatments rarely cure patients with disseminated disease is important to discover better therapies. When one treats cancer cells with specific concentrations of the available anticancer drugs and examines the cells under the microscope, one generally observes a massacre. All cancer cells die in response to most treatments. However, these drugs also kill normal cells at similar concentrations. The consequence of this limited selectivity is that patients cannot receive the drug doses needed...
to kill all their cancer cells; such doses would also kill their normal body cells and would be lethal. Although oncology patients generally receive the maximum tolerated doses, these doses are usually insufficient to reach the drug concentrations required to eradicate their cancer cells. The surviving cancer cells continue to proliferate and eventually lead to a fatal outcome. Finding drugs with a high selectivity towards cancer cells is crucial to develop more effective treatments for patients with metastasis [2–4].

Several plants have provided useful drugs for the treatment of a variety of cancers, including lung cancer [5–7]. For example, the diterpene paclitaxel (isolated from the bark of *Taxus brevifolia* Nutt., Taxaceae) and its semisynthetic derivative docetaxel are FDA-approved drugs for the treatment of non-small-cell lung cancer [8,9]. Vinorelbine, a semisynthetic analog of the vinca alkaloids (isolated from *Catharanthus roseus* G. Don., Apocynaceae), is also approved for patients with this type of cancer [9]. Etoposide (a semi-synthetic analogue of the natural lignan podophyllotoxin, isolated from *Podophyllum* species, Podophyllaceae) and topotecan (an analogue of the quinoline alkaloid camptothetin, isolated from *Camptotheca acuminata* Decne, Nyssaceae) are approved for patients with small cell lung cancer. Due to the fact that several plants have provided useful anticancer agents, we recently used lung cancer cells and lung normal cells to evaluate the selective anticancer activity of 57 extracts from plants collected in Grazalema Natural Park (Andalusia, Spain) [10]. Using a similar experimental approach, we have evaluated the selective anticancer activity of 65 extracts from 45 new plants collected in several areas of Western Andalusia, and report the results in this article.

### 2. Results and Discussion

Due to the fact that patients with metastatic cancers need selective anticancer drugs, we have searched for potential sources of selective anticancer drugs in 45 plants growing in Western Andalusia, Spain. After collecting the plant material and preparing 65 extracts, we used lung cancer cells (A549) and lung normal cells (MRC-5) to evaluate their selective cytotoxicity with the MTT assay. A549 cells are epithelial cells and MRC-5 cells are fibroblastic cells. However, since normal epithelial cells are difficult to maintain and expand in culture, we selected this widely used normal cell line for our screening [11–13].

We exposed both cell lines for 72 h to at least five concentrations of the extracts and the anticancer drugs cisplatin and 5-fluorouracil (5-FU). Table 1 shows the botanical names in alphabetical order, the plant families and other pertinent information on the 45 plant species. It also shows an identification number for each extract, the IC50 value for each cell line, and the selectivity index. Dose–response curves for the 65 extracts and positive controls are provided in Figures 1–6; these curves allow for the visualization of their cytotoxic profile over a wide concentration range.
Table 1. Cytotoxic activity of plant extracts on lung cancer cells (A549) versus lung normal cells (MRC-5).

| Extract | Plant Name (Family) | Part Used           | Voucher Number (SEV) | Origin | IC50 (MTT) (Mean ± SEM, μg/mL) | S.I. (Mean ± SEM) |
|---------|---------------------|---------------------|----------------------|--------|-------------------------------|-------------------|
|         |                     |                     |                      |        | A549 (Cancer)                 | MRC-5 (Normal)    |
| 1       | Acis autumnalis (L.) Sweet (Amaryllidaceae) | Whole plant | 284654              | Seville | 32.9 ± 9.6 | 59.1 ± 18.0 | 1.9 ± 0.6 |
| 2       | Anagallis monelli L. (Primulaceae) | Aerial Parts | 284675              | Cádiz | 3.2 ± 1.4 | 0.6 ± 0.2 | 0.3 ± 0.2 |
| 3       | Anagallis monelli L. (Primulaceae) | Root | 284675              | Cádiz | 7.4 ± 5.2 | 0.9 ± 0.5 | 0.3 ± 0.3 |
| 4       | Anthyllis hamosa Desf. (Leguminosae) | Whole plant | 284685              | Huelva | 260.9 ± 14.7 | 317.7 ± 3.1 | 1.2 ± 0.1 |
| 5       | Aristolochia baetica L. (Aristolochiaceae) | Fruits | 284674              | Seville | 249.1 ± 2.5 | 649.9 ± 345.0 | 2.6 ± 1.4 |
| 6       | Aristolochia baetica L. (Aristolochiaceae) | Leaves | 284674              | Seville | 91.8 ± 70.5 | 66.0 ± 39.9 | 0.9 ± 0.3 |
| 7       | Armeria pungens (Link) Hoffmanns. & Link (Plumbaginaceae) | Flowering aerial parts | 284687              | Huelva | 143.4 ± 64.3 | 550.7 ± 352.2 | 3.6 ± 1.1 |
| 8       | Armeria velutina Welw. ex Boiss. & Reut. (Plumbaginaceae) | Whole plant | 284689              | Huelva | 25.7 ± 1.9 | 36.7 ± 0.9 | 1.4 ± 0.1 |
| 9       | Campanula lasticanica L. (Campanulaceae) | Whole plant | 284667              | Seville | 223.4 ± 51.2 | 263.7 ± 34.2 | 1.3 ± 0.4 |
| 10      | Cascabela thevetia (L.) Lippold (Apocynaceae) | Leaves | 284662              | Seville | 0.14 ± 0.02 | 1.6 ± 0.5 | 26.6 ± 11.6 |
| 11      | Centaurea sphaerocephala L. (Compositae) | Whole plant | 284683              | Huelva | 134.2 ± 82.4 | 160.4 ± 33.8 | 1.7 ± 0.8 |
| 12      | Centaurea sphaerocephala L. (Compositae) | Flowers | 284676              | Cádiz | 139.1 ± 40.1 | 108.7 ± 7.7 | 1.7 ± 1.1 |
| 13      | Centaurea sphaerocephala L. (Compositae) | Leaves | 284676              | Cádiz | 43.1 ± 11.1 | 114.7 ± 14.4 | 3.0 ± 0.8 |
| 14      | Cistus crispus L. (Cistaceae) | Leaves | 284660              | Seville | 52.4 ± 5.1 | 126.4 ± 34.3 | 2.5 ± 0.7 |
| 15      | Cistus crispus L. (Cistaceae) | Root | 284660              | Seville | 24.9 ± 0.7 | 58.8 ± 14.8 | 2.1 ± 0.7 |
| 16      | Cistus salviifolius L. (Cistaceae) | Leaves | 284653              | Seville | 56.6 ± 12.6 | 142.7 ± 20.4 | 3.0 ± 0.9 |
| 17      | Cleome violacea L. (Cleomaceae) | Aerial Parts | 284668              | Seville | 253.9 ± 39.9 | 261.1 ± 3.4 | 1.1 ± 0.2 |
| 18      | Digitalis purpurea L. (Plantaginaceae) | Leaves | 284691              | Huelva | 0.17 ± 0.15 | 1.06 ± 0.69 | 9.3 ± 0.5 |
| 19      | Dorycnium rectum (L.) Ser. (Leguminosae) | Flowers | 284690              | Seville | 102.1 ± 40.2 | 182.9 ± 46.1 | 1.7 ± 3.2 |
| 20      | Dorycnium rectum (L.) Ser. (Leguminosae) | Leaves | 284690              | Seville | 293.6 ± 21.2 | 322.1 ± 27.4 | 1.1 ± 0.2 |
| 21      | Echiium gaditanum Boiss. (Boraginaceae) | Aerial Parts | 284684              | Huelva | 196.1 ± 107.2 | 325.6 ± 44.0 | 1.1 ± 0.1 |
| 22      | Elaeoceliniun foetidum (L.) Boiss. (Apioaceae) | Flowers | 284670              | Seville | 119.0 ± 16.2 | 262.1 ± 8.7 | 2.3 ± 0.3 |
| 23      | Elaeoceliniun foetidum (L.) Boiss. (Apioaceae) | Leaves | 284670              | Seville | 246.5 ± 25.5 | 295.0 ± 36.4 | 1.2 ± 0.0 |
| 24      | Erica arborea L. (Ericaceae) | Bark | 284657              | Seville | 32.2 ± 5.6 | 62.1 ± 28.9 | 1.8 ± 0.7 |
| 25      | Erica arborea L. (Ericaceae) | Leaves | 284657              | Seville | 45.7 ± 7.4 | 158.4 ± 43.6 | 3.9 ± 1.4 |
| 26      | Erophaca baetica (L.) Boiss. (Leguminosae) | Leaves | 284673              | Seville | >1000 | >1000 | N.D. |
| 27      | Frangula alius Mill. (Rhamnaceae) | Bark | 284680              | Huelva | 32.8 ± 7.4 | 339.3 ± 74.1 | 12.4 ± 3.3 |
| 28      | Frangula alius Mill. (Rhamnaceae) | Leaves | 284680              | Huelva | 28.6 ± 3.5 | 74.0 ± 38.5 | 2.6 ± 1.0 |
| 29      | Genista hispida M.Vahl (Leguminosae) | Aerial Parts | 284671              | Seville | 273.6 ± 5.8 | 363.9 ± 27.9 | 1.3 ± 0.1 |
| 30      | Halimium calycinum (L.) K.Koch (Cistaceae) | Leaves | 284656              | Seville | 47.4 ± 10.0 | 135.4 ± 23.6 | 4.3 ± 1.9 |
| Extract | Plant Name (Family) | Part Used | Voucher Number (SEV) | Origin | IC50 (MTT) (Mean ± SEM, µg/mL) | S.I. (Mean ± SEM) | A549 (Cancer) | MRC-5 (Normal) |
|---------|---------------------|-----------|---------------------|--------|-------------------------------|-----------------|--------------|-------------|
| 31      | *Halimium calycinum* (L.) K.Koch (Cistaceae) | Root | 284656 | Seville | 60.1 ± 17.7 | 2.2 ± 0.8 | 101.1 ± 13.6 |
| 32      | *Halimium halimifolium* (L.) Willk. (Cistaceae) | Leaves | 284659 | Seville | 47.9 ± 10.9 | 2.7 ± 0.9 | 105.9 ± 21.4 |
| 33      | *Halimium halimifolium* (L.) Willk. (Cistaceae) | Root | 284659 | Seville | 67.5 ± 16.0 | 1.8 ± 0.8 | 110.2 ± 33.4 |
| 34      | *Hedysarum coronarium* L. (Leguminosae) | Flowers | 284677 | Cádiz | 230.5 ± 54.6 | 1.4 ± 0.1 | 306.9 ± 55.1 |
| 35      | *Hedysarum coronarium* L. (Leguminosae) | Fruits | 284677 | Cádiz | 247.0 ± 3.2 | 1.2 ± 0.0 | 294.5 ± 0.4 |
| 36      | *Hedysarum coronarium* L. (Leguminosae) | Leaves | 284677 | Cádiz | 173.9 ± 60.6 | 2.1 ± 0.9 | 262.0 ± 32.3 |
| 37      | *Iberis cilata* subsp. *contracta* (Pers.) Moreno (Brassicaceae) | Whole plant | 284688 | Huelva | 0.31 ± 0.06 | 13.0 ± 4.7 | 2.31 ± 0.88 |
| 38      | *Jasione montana* L. (Campanulaceae) | Whole plant | 284666 | Seville | 145.0 ± 26.0 | 2.0 ± 0.1 | 301.6 ± 69.2 |
| 39      | *Juniperus macrocarpa* Sm. (Cupressaceae) | Monosperma cones | 284682 | Huelva | 146.1 ± 125.9 | >20 | >1000 |
| 40      | *Juniperus macrocarpa* Sm. (Cupressaceae) | Aerial Parts | 284682 | Huelva | 3.7 ± 1.9 | 0.8 ± 0.1 | 2.8 ± 1.0 |
| 41      | *Malcolmia lacera* (L.) DC. (Brassicaceae) | Whole plant | 284664 | Seville | 322.3 ± 86.7 | 1.0 ± 0.3 | 295.0 ± 5.9 |
| 42      | *Malva hispanica* L. (Malvaceae) | Aerial Parts | 284669 | Seville | >1000 | N.D. | 703.1 ± 51.1 |
| 43      | *Orobanche lutea* L. (Orobanchaceae) | Whole plant | 284695 | Huelva | 63.9 ± 23.6 | 4.6 ± 1.4 | 232.3 ± 29.5 |
| 44      | *Ornithopus sativus* Brot. (Leguminosae) | Whole plant | 284693 | Seville | 340.7 ± 28.4 | 1.8 ± 0.6 | 583.8 ± 150.0 |
| 45      | *Pancratium maritimum* L. (Amaryllidaceae) | Whole plant | 284692 | Seville | 326.6 ± 37.5 | 2.3 ± 1.1 | 699.0 ± 262.9 |
| 46      | *Pycnoconium rutifolium* (Vahl) Hoffmanns. & Link (Caprifoliaceae) | Whole plant | 284681 | Huelva | 3.4 ± 0.2 | 19.7 ± 14.6 | 74.1 ± 56.1 |
| 47      | *Ranunculus peltatus* Schrank (Ranunculaceae) | Whole plant | 284672 | Seville | 186.7 ± 54.6 | 1.1 ± 0.3 | 224.9 ± 56.7 |
| 48      | *Rhamnus lycioides subsp. oleoides* (L.) Jahand. & Maire (Rhamnaceae) | Root | 284678 | Cádiz | 262.5 ± 131.9 | 2.2 ± 1.4 | 70.6 ± 18.1 |
| 49      | *Rhamnus lycioides subsp. oleoides* (L.) Jahand. & Maire (Rhamnaceae) | Whole plant | 284672 | Seville | 186.7 ± 54.6 | 0.9 ± 0.1 | 170.1 ± 58.8 |
| 50      | *Scrophularia frutescens* L. (Scrophulariaceae) | Bark | 284655 | Seville | 332.2 ± 61.9 | 2.1 ± 0.8 | 599.4 ± 122.1 |
| 51      | *Scleranthus annuus* L. (Asteraceae) | Leaves | 284655 | Seville | 151.5 ± 16.7 | 3.1 ± 1.6 | 421.3 ± 176.5 |
| 52      | *Scleranthus annuus* L. (Asteraceae) | Root | 284655 | Seville | 257.0 ± 76.6 | 2.3 ± 0.8 | 442.3 ± 35.1 |
| 53      | *Scleranthus annuus* L. (Asteraceae) | Whole plant | 284686 | Huelva | 281.5 ± 9.2 | 1.1 ± 0.1 | 297.6 ± 8.6 |
| 54      | *Stenophyllum canariense* Willd. (Tamaricaceae) | Aerial Parts | 284679 | Huelva | 278.4 ± 32.9 | 2.0 ± 0.4 | 546.5 ± 53.5 |
| 55      | *Tamarix canariensis* Willd. (Tamaricaceae) | Flowers | 284650 | Seville | 212.8 ± 10.2 | 1.6 ± 0.1 | 336.3 ± 3.6 |
Table 1. Cont.

| Extract | Plant Name (Family) | Part Used | Voucher Number (SEV) | Origin | IC50 (MTT) (Mean ± SEM, µg/mL) | S.I. (Mean ± SEM) |
|---------|---------------------|-----------|----------------------|--------|-------------------------------|-------------------|
| 56      | *Tamarix canariensis* Willd. (Tamaricaceae) | Leaves    | 284650               | Seville | 92.7 ± 24.6                   | 192.1 ± 33.0     | 2.2 ± 0.3         |
| 57      | *Taxus baccata* L. (Taxaceae) | Leaves    | 284621               | Seville | 0.86 ± 0.27                   | 146.9 ± 87.8     | 157.3 ± 110.6    |
| 58      | *Tetraclinis articulata* (Vahl) Mast. (Cupressaceae) | Leaves    | 284663               | Seville | 0.37 ± 0.03                   | 129.5 ± 64.0     | 378.3 ± 178.1    |
| 59      | *Teucrium fruticans* L. (Lamiaceae) | Leaves    | 284658               | Seville | 157.1 ± 30.5                  | 433.0 ± 112.3    | 2.8 ± 0.7         |
| 60      | *Thymus mastichina* (L.) L. (Lamiaceae) | Whole plant | 284694              | Huelva  | 36.8 ± 7.7                    | 277.7 ± 40.6     | 8.2 ± 2.0         |
| 61      | *Tolpis barbata* (L.) Gaertn. (Compositae) | Whole plant | 284665              | Seville | 86.5 ± 28.3                   | 43.7 ± 12.2      | 0.5 ± 0.0         |
| 62      | *Ulex parviflorus* Pourr. subsp. *parviflorus* (Leguminosae) | Flowers   | 284652               | Seville | 361.7 ± 89.6                  | 876.1 ± 426.1    | 4.0 ± 2.9         |
| 63      | *Ulex parviflorus* Pourr. subsp. *parviflorus* (Leguminosae) | Leaves    | 284652               | Seville | 99.6 ± 34.8                   | 270.4 ± 85.3     | 3.4 ± 1.2         |
| 64      | *Viburnum tinus* L. (Adoxaceae) | Fruits    | 284651               | Seville | 26.6 ± 6.5                    | 65.4 ± 8.6       | 2.6 ± 0.3         |
| 65      | *Viburnum tinus* L. (Adoxaceae) | Leaves    | 284651               | Seville | 234.8 ± 58.7                  | 568.2 ± 73.0     | 2.6 ± 0.3         |
|         | Cisplatin (Standard anticancer drug) |           |                      |        | 10.5 ± 5.5 (µM)              | 25.4 ± 7.4 (µM)  | 4.2 ± 2.2         |
|         | 5-Fluorouracil (Standard anticancer drug) |           |                      |        | 101.8 ± 7.7 (µM)             | >1000 (µM)       | >9.9              |

S.I.: selectivity index (calculated as the average of the IC50 value in the MRC-5 normal cell line divided by the IC50 value in the A549 cancer cell line obtained in each independent experiment); Nd: not determined.
Figure 1. Screening for selective cytotoxic activity of plant extracts 1–12 on A549 lung cancer cells and MRC-5 lung normal cells. The cells were exposed for 72 h to the extracts and cell viability was determined with the MTT assay.
Figure 2. Screening for selective cytotoxic activity of plant extracts 13–24 on A549 lung cancer cells and MRC-5 lung normal cells. The cells were exposed for 72 h to the extracts and cell viability was determined with the MTT assay.
Figure 3. Screening for selective cytotoxic activity of plant extracts 25–36 on A549 lung cancer cells and MRC-5 lung normal cells. The cells were exposed for 72 h to the extracts and cell viability was determined with the MTT assay.
Figure 4. Screening for selective cytotoxic activity of plant extracts 37–48 on A549 lung cancer cells and MRC-5 lung normal cells. The cells were exposed for 72 h to the extracts and cell viability was determined with the MTT assay.
Figure 5. Screening for selective cytotoxic activity of plant extracts 49–60 on A549 lung cancer cells and MRC-5 lung normal cells. The cells were exposed for 72 h to the extracts and cell viability was determined with the MTT assay.
Results show that several extracts induced selective cytotoxicity towards the cancer cell line, including the extract from the leaves of *Cascabela thevetia* (L.) Lippold (Apocynaceae) (10), the extract from the leaves of *Digitalis purpurea* L. (Plantaginaceae) (18), the extract from the bark of *Frangula alnus* Mill. (Rhamnaceae) (27), the extract from the whole plant *Iberis ciliata subsp. contracta* (Pers.) Moreno (Brassicaceae) (37), the extract from the seeds of *Juniperus macrocarpa* Sm. (Cupressaceae) (39), the extract from the bulb of *Pancratium maritimum* L. (Amaryllidaceae) (46), and the extract from the leaves of *Tetraclinis articulata* (Vahl) Mast. (Cupressaceae) (58). The anticancer drugs cisplatin and 5-fluorouracil, and an extract from the leaves of *Taxus baccata* L. (Taxaceae) (57) also showed selectivity against the cancer cells (Table 1). The extract from the leaves of *Tetraclinis articulata* (Vahl) Mast. (58) (Figure 5) showed the most relevant activ-
Plants 2021, 10, 2193

Table 2. Cytotoxicity as IC₅₀ values of selected extracts, cisplatin, and 5-FU on a panel of human cancer cell lines of different tissue origin and three genetically modified skin cell lines. Values are expressed as mean ± SEM.

| Cell Line | 10  | 18  | 37  | 46  | 57  | 58  | Cisplatin (Mean ± SEM, µg/mL) | 5-FU (Mean ± SEM, µM) |
|-----------|-----|-----|-----|-----|-----|-----|-------------------------------|----------------------|
| HepG2     | 24.6 ± 4.1 | 13.2 ± 3.0 | 19.4 ± 10.4 | 7.0 ± 1.3 | 632.6 ± 106.1 | 18.4 ± 17.1 | 3.5 ± 0.8 | 0.3 ± 0.2 |
| SW480     | 20.7 ± 5.6 | 18.3 ± 1.2 | 1.2 ± 0.3 | 2.1 ± 0.4 | 2.2 ± 0.7 | 2.1 ± 0.8 | 2.0 ± 0.5 | 3.4 ± 1.0 |
| U2OS      | 9.2 ± 1.5 | 8.8 ± 0.8 | 1.9 ± 0.2 | 2.8 ± 0.7 | 4.0 ± 1.2 | 2.1 ± 0.9 | 4.0 ± 1.0 | 7.2 ± 1.0 |
| HeLa      | 4.7 ± 0.1 | 4.9 ± 0.2 | 3.5 ± 0.5 | 5.4 ± 0.8 | 2.5 ± 0.4 | 1.3 ± 0.6 | 4.8 ± 0.1 | 66.5 ± 27.0 |
| PC3       | 4.7 ± 1.5 | 2.6 ± 0.4 | 5.6 ± 2.1 | 5.4 ± 1.2 | 123.8 ± 42.7 | 0.50 ± 0.04 | 7.3 ± 4.6 | 1.5 ± 0.8 |
| MCF7      | 10.0 ± 0.5 | 6.5 ± 1.0 | 4.0 ± 0.9 | 5.9 ± 0.8 | 37.1 ± 4.9 | 0.7 ± 0.1 | 8.0 ± 3.0 | 1.1 ± 0.8 |
| NB4       | 2.9 ± 0.7 | 1.8 ± 0.5 | 0.5 ± 0.2 | 1.2 ± 0.2 | 2.8 ± 0.2 | 0.2 ± 0.1 | 0.08 ± 0.04 | 2.7 ± 0.3 |
| HL-60     | 3.5 ± 0.4 | 3.6 ± 0.3 | 0.8 ± 0.2 | 1.0 ± 0.3 | 3.3 ± 0.2 | 0.7 ± 0.3 | 0.2 ± 0.1 | 0.6 ± 0.5 |
| BJ-hTERT  | 3.7 ± 0.4 | 2.8 ± 0.3 | 3.4 ± 0.5 | 3.4 ± 0.1 | 4.0 ± 0.6 | 0.8 ± 0.3 | 1.2 ± 0.4 | 2.6 ± 0.6 |
| BJ-SV40T  | 2.9 ± 0.4 | 2.1 ± 0.4 | 2.9 ± 0.4 | 3.2 ± 0.4 | 2.8 ± 0.2 | 1.0 ± 0.4 | 0.7 ± 0.2 | 5.8 ± 0.7 |
| BJ-RASV12 | 2.9 ± 0.4 | 1.8 ± 0.7 | 4.0 ± 1.4 | 2.0 ± 0.7 | 3.3 ± 0.2 | 0.6 ± 0.2 | 1.1 ± 0.1 | 1.7 ± 0.8 |

HepG2: human hepatocellular carcinoma; SW480: human colon adenocarcinoma; U2OS: human osteosarcoma; HeLa: human cervical carcinoma; PC3: human prostate adenocarcinoma; MCF7: human breast adenocarcinoma; NB4: human acute promyelocytic leukemia; HL-60: human acute promyelocytic leukemia; BJ-hTERT: hTERT-immortalized skin non-malignant BJ; BJ-SV40T: SV40T-transformed BJ-hTERT; BJ-RASV12: H-RAS V12-transformed BJ-SV40T. Extract numbers (10, 18, 37, 46, 57, and 58) can be identified from Table 1.

Due to the fact that Tetraclinis articulata (Vahl) Mast. extract (58) showed a high cytotoxicity against several cancer cell lines, we acquired additional cancer cell lines from a variety of tissues to further evaluate the activity of the extract. Fourteen human cancer cell lines were treated with several concentrations of 58 for 96 h and cell viability was estimated with the resazurin assay. Results show that this extract was cytotoxic against all the cancer cell lines, with IC₅₀ values in the range 4.3–4.9 µg/mL (Table 3).
Several types of phytochemicals may be involved in the selective anticancer activity shown by the most active extracts. Cardiac glycosides may be responsible for the selective cytotoxicity shown by the extracts from the leaves of *Cascabela thevetia* (L.) Lippold (10) and the leaves of *Digitalis purpurea* L. (18). We, and others, have previously observed that plants with cardiac glycosides, and several cardiac glycosides (e.g., digitoxin), induce potent and selective cytotoxic effects against several types of cancer cells, including lung cancer cells [15–23]. Isoquinoline alkaloids may participate in the cytotoxicity and selectivity observed for the extract from the bulb of *Pancratium maritimum* L. (46) [24,25]. The cytotoxicity and selectivity of our extract from the leaves of *Taxus baccata* L. (57) were probably mediated by different taxane-type diterpenes, including paclitaxel [26,27]. Diterpenes (e.g.,

Figure 7. Cytotoxicity of selected plant extracts (10, 18, 37, 46, 57, and 58), cisplatin, and 5-FU against six cancer cell lines derived from solid tumors and two acute promyelocytic leukemia cell lines. Cells were treated for 72 h, and cell viability was evaluated with the resazurin assay.
ferruginol and sandaracopimaric acid), monoterpenes (e.g., limonene and carveol), and sesquiterpenes (e.g., β-caryophyllene and humulene) may participate in the cytotoxicity of the extract from the leaves of *Tetraclinis articulata* (Vahl) Mast. [28–33].

Table 3. Cytotoxic activity of *Tetraclinis articulata* (Vahl) Mast. (58) against 14 cancer cell lines from a variety of tissues.

| Cell Line | IC50 (Resazurin) (Media ± SEM, µg/mL) |
|-----------|-------------------------------------|
| A64-CLS   | 4.7 ± 0.3                           |
| AN3Ca     | 4.5 ± 0.9                           |
| Calu-1    | 4.7 ± 0.2                           |
| GAMG      | 4.5 ± 0.6                           |
| HNO97     | 4.5 ± 0.3                           |
| HT29      | 4.4 ± 0.4                           |
| KATO III  | 4.4 ± 0.7                           |
| MDA-MB-231| 4.7 ± 0.2                           |
| MeWo      | 4.3 ± 0.2                           |
| PC-3      | 4.5 ± 1.0                           |
| Sk-Br-3   | 4.9 ± 0.0                           |
| Sk-OV-3   | 4.7 ± 0.1                           |
| T24       | 4.5 ± 0.3                           |
| UACC-62   | 4.5 ± 0.2                           |

A64-CLS: submaxillary gland adenoma; AN3Ca: endometrial adenocarcinoma; Calu-1: squamous lung cancer; GAMG: glioblastoma; HNO97: tongue cancer; HT29: colorectal cancer; KATO III: gastric cancer; MDA-MB-231: triple-negative breast cancer; MeWo: BRAF wild-type melanoma; PC-3: prostate cancer; Sk-Br-3: HER2-positive breast cancer; Sk-OV-3: ovarian cancer; UACC-62: BRAF mutant melanoma; and T24: bladder cancer.

In summary, because patients with advanced cancers need selective anticancer drugs, we have searched for potential sources of selective anticancer drugs in a variety of plants. After collecting 45 plants and preparing 65 extracts, we used a cancer cell line and a normal cell line from the same tissue to detect selective cytotoxic activity [2–4]. Several extracts induced selective cytotoxicity towards the cancer cell line, including extracts from *Cascabela thevetia* (L.) Lippold (Apocynaceae), *Frangula alnus* Mill. (Rhamnaceae), *Iberis ciliata* subsp. *contracta* (Pers.) Moreno (Brassicaceae), *Juniperus macrocarpa* Sm (Cupressaceae), *Pancratium maritimum* L. (Amaryllidaceae), and *Tetraclinis articulata* (Vahl) Mast. (Cupressaceae). Due to the fact that the selective anticancer activity of *Tetraclinis articulata* (Vahl) Mast was high, we evaluated and observed cytotoxic activity in an additional 22 cancer cell lines, representing the most common types of cancer. Our study shows that several plants are promising sources for the isolation and the development of new anticancer drugs with cancer-selective toxicity.

3. Materials and Methods

3.1. Plant Material

All plants were collected by F. García between November 2012 and April 2013 in several areas of Sevilla, Cadiz, and Huelva (Andalusia, Spain). Collection was non-destructive and plant specimens (5–110 g) were carefully selected to avoid any damage that could affect the conservation of any species. A voucher specimen was deposited in the herbarium at the CITIUS II Celestino Mutis (Center for Research, Technology, and Innovation of the University of Seville). The botanical names, plant parts, and voucher specimen numbers are listed in Table 1. Collection coordinates are provided in Table 4.
Table 4. Collection coordinates of plants used in this work.

| Extract | Plant Name | Collection Coordinates |
|---------|------------|------------------------|
| 1       | Acis autumnalis (L.) Sweet | 37°14′22.06″ N 6°11′37.85″ W |
| 2–3     | Anagallis monelli L. | 36°36′15.24″ N 6°16′2.76″ W |
| 4       | Anthyllis hamosa Desf. | 37°04′25.2″ N 6°41′19.68″ W |
| 5–6     | Aristolochia baetica L. | 37°14′16.68″ N 6°11′48.38″ W |
| 7       | Armeria pungens (Link) Hoffmarns. & Link | 37°04′13.73″ N 6°41′16.97″ W |
| 8       | Armeria velutina Welw. ex Boiss. & Reut. | 37°02′33.33″ N 6°35′53.87″ W |
| 9       | Campanula lasioccia L. | 37°14′14.46″ N 6°11′55.8″ W |
| 10      | Castaleta thecata (L.) Lippold | 37°22′59.8″ N 5°59′27.36″ W |
| 11–13   | Centaura sphaerocephala L. | 37°05′40.18″ N 6°43′37.9″ W |
| 14–15   | Cistus crispus L. | 37°14′22.06″ N 6°11′37.85″ W |
| 16      | Cistus salviolus L. | 37°14′19.74″ N 6°11′40.71″ W |
| 17      | Cleome violaca L. | 37°14′23.37″ N 6°11′52.72″ W |
| 18      | Digitalis purpurea L. | 37°27′30.78″ N 6°41′20.3″ W |
| 19–20   | Dorycnium rectum (L.) Ser. | 37°20′14.71″ N 5°51′27.13″ W |
| 21      | Echium gadianum Boiss. | 37°04′11.69″ N 6°41′17.16″ W |
| 22–23   | Elaeoselium foetidum (L.) Boiss. | 37°14′15.72″ N 6°11′50.86″ W |
| 24–25   | Erica arborea L. | 37°14′38.35″ N 6°11′49.92″ W |
| 26      | Erophaca baetica (L.) Boiss. | 37°14′17.72″ N 6°11′46.41″ W |
| 27–28   | Frangula alnus Mill. | 37°05′40.46″ N 6°43′34.47″ W |
| 29      | Genista hissuta M. Vahl | 37°14′16.70″ N 6°11′49.05″ W |
| 30–31   | Halimium calycinum (L.) K. Koch | 37°14′21.27″ N 6°11′38.12″ W |
| 32–33   | Halimium halimifolium (L.) Willk. | 37°14′23.43″ N 6°11′38.77″ W |
| 34–36   | Hedysarum coronarium L. | 36°36′39.92″ N 6°16′46.6″ W |
| 37      | Iberis ciliata subsp. contracta (Pers.) Moreno | 37°04′47.25″ N 6°41′13.82″ W |
|         | Jason montana L. | 37°13′45.76″ N 6°9′16.08″ W |
| 39–40   | Juniperus macrocarpa Sm. | 37°04′13.53″ N 6°41′16.34″ W |
| 41      | Malcomia lacer (L.) DC. | 37°13′45.76″ N 6°9′16.08″ W |
| 42      | Malva hispanica L. | 37°14′16.09″ N 6°11′51.89″ W |
| 43      | Ononis subspicata Lag. | 37°04′45.90″ N 6°41′14.14″ W |
| 44      | Ornithopus compressus L. | 37°13′45.76″ N 6°9′16.08″ W |
| 45      | Ornithopus sativus Broit. | 37°13′45.76″ N 6°9′16.08″ W |
| 46      | Pancratium maritimum L. | 37°04′11.14″ N 6°41′12.9″ W |
| 47–48   | Pycnoncom rutifolium (Vahl) Hoffmanns. & Link | 36°36′12.69″ N 6°15′35.45″ W |
| 49      | Ranunculus pelatus Schrank | 37°13′45.76″ N 6°9′16.08″ W |
| 50–52   | Rhamnus lycioides subsp. oleoides (L.) Jahand. & Maire | 37°14′25.18″ N 6°11′38.35″ W |
| 53      | Scrophularia frutescens L. | 37°04′47.25″ N 6°41′13.82″ W |
| 54      | Stauracanthus genistoides (Brot.) G. Sampaio | 37°04′44.55″ N 6°41′16.05″ W |
| 55–56   | Tamaxir canariensis Willd. | 37°15′45.64″ N 5°59′50.9″ W |
| 57      | Taxus baccata L. | 37°22′27″ N 5°59′19″ W |
| 58      | Tetraclinis articulata (Vahl) Mast. | 37°22′22.18″ N 5°59′10.75″ W |
| 59      | Teucrium fruticans L. | 37°14′29″ N 6°11′52.78″ W |
| 60      | Thymus mastichina (L.) L. | 37°02′33.33″ N 6°35′53.85″ W |
| 61      | Tolpis barba (L.) Gaertn. | 37°13′45.76″ N 6°9′16.08″ W |
| 62–63   | Ulmus parviflorus Poutr. subsp. parviflorus | 37°13′48.06″ N 6°1′31.4″ W |
| 64–65   | Viburnum tinus L. | 37°22′27″ N 5°59′19″ W |

3.2. Preparation of the Extracts

Extracts were prepared within several hours after collecting the plants. Fresh plant material (5–110 g) was extracted with 100–200 mL of ethanol/ethyl acetate/water (1:1:1) at 60 °C for 1 h by using an ultrasound water bath apparatus. After vacuum filtration, ethanol and ethyl acetate were eliminated in a rotary vacuum evaporator at 60 °C. Finally, the remaining water solution was lyophilized to yield dried extracts. The extraction yield (%) for each extract (see identification number in Table 1) was: 1 (1.9%), 2 (3.0%), 3 (5.6%), 4 (4.5%), 5 (0.9%), 6 (2.9%), 7 (5.9%), 8 (4.0%), 9 (2.7%), 10 (5.5%), 11 (2.0%), 12 (2.9%), 13 (3.5%), 14 (8.5%), 15 (5.2%), 16 (7.4%), 17 (8.9%), 18 (Nd), 19 (6.7%), 20 (7.2%), 21 (2.4%), 22 (1.9%), 23 (2.4%), 24 (6.0%), 25 (14.8%), 26 (3.3%), 27 (1.0%), 28 (4.4%), 29 (6.8%), 30 (10.6%),
31 (4.9%), 32 (9.6%), 33 (4.6%), 34 (4.2%), 35 (4.0%), 36 (3.3%), 37 (4.2%), 38 (3.2%), 39 (5.9%),
40 (7.5%), 41 (5.3%), 42 (3.4%), 43 (2.7%), 44 (3.5%), 45 (2.4%), 46 (3.7%), 47 (2.6%), 48 (5.5%),
49 (0.9%), 50 (4.4%), 51 (5.5%), 52 (4.3%), 53 (5.8%), 54 (5.9%), 55 (Nd), 56 (8.4%), 57 (7.8%),
58 (5.5%), 59 (8.1%), 60 (6.5%), 61 (3.6%), 62 (10.7%), 63 (6.6%), 64 (9.0%), and 65 (9.9%). The
extracts were stored in dark glass bottles and kept in a cool dark place. The first cytotoxicity
experiment of the screening was carried out within the first month after preparing the
extracts to avoid the possible degradation of active compounds. In the first cytotoxicity
experiment, a stock solution of each extract was prepared in DMSO (100 mg/mL); a part of
this solution was diluted in culture medium and immediately used to treat the cells. The
stock solutions were aliquoted and frozen at −80 °C. The rest of independent cytotoxicity
experiments were carried out using different aliquots to avoid freeze–thaw cycles.

3.3. Chemicals and Cell Lines

Cisplatin, MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide), and
resazurin were bought from Sigma. MRC-5 (human fetal lung fibroblastic cells) and A549
(human non-small-cell lung cancer cells) were purchased from the European Collection of
Cell Cultures. MDA-MB-231 (human triple-negative breast cancer cells) was purchased
from the American Type Culture Collection (ATCC). UACC-62 (human BRAF mutant
melanoma cells) was obtained from National Cancer Institute. HepG2 (human hepatocel-
lar carcinoma cells), PC3 (human prostate cancer cells), HT29 (human colorectal cancer
cells), MCF7 (human breast cancer cells), HeLa (human cervical carcinoma cells), NB4
(human acute promyelocytic leukemia cells), HL-60 (human acute promyelocytic leukemia
cells), SW480 (human colon adenocarcinoma cells), and U2OS (human osteosarcoma cells)
were kindly provided by Dr. Helleday (Karolinska Institute, Sweden). HNO97 (human
tongue cancer cells), A64-CLS (human submaxillary gland adenoma cells), AN3Ca (hu-
man endometrial adenocarcinoma cells), Sk-OV-3 (human ovarian cancer cells), KATO III
(human gastric cancer cells), Sk-Br-3 (HER2-positive breast cancer cells), T24 (human blad-
er cancer cells), Calu-1 (human squamous lung cancer cells), and MeWo (human BRAF
wild-type melanoma cells) were purchased from Cell Lines Service (CLS). GAMG cells
(human glioblastoma cells) were provided by Dr. A. Ayala (University of Seville). BJ-hTERT
(hTERT-immortalized foreskin fibroblast BJ cells), BJ-SV40T (SV40T-transformed BJ-hTERT
cells), and BJ RASV12 (H-RAS V12-transformed BJ-SV40T cells) were provided by Dr. Hahn
(Dana-Farber Cancer Institute, USA) [14]. Cells were maintained in appropriate medium
and propagated according to standard protocols. MRC-5, A549, MDA-MB-231, HepG2,
HT29, MCF7, HeLa, SW48, U2OS, HNO97, A64-CLS, AN3Ca, SK-OV-3, Sk-Br-3, MeWo,
GAMG, BJ-hTERT, BJ-SV40T, and BJ-RASV12, were maintained in Dulbecco’s modified
Eagle’s medium (DMEM) high-glucose medium. PC3 and T24 were grown in DMEM-
F12. UACC-62, NB4, HL60, and Calu-1 were maintained in RPMI 1640. KATO III were
grown in Ham’s F12. All media were supplemented with 100 U/mL penicillin, 100 µg/mL
streptomycin, and 10% fetal bovine serum. All cells were kept at 37 °C in a humidified
atmosphere containing 5% CO2. Cell culture reagents were purchased from Biowest.

3.4. Cell Viability Assays

Exponentially growing cells (3000–5000 cells per well) were seeded in 96-well plates
and were allowed to grow for 24 h. The cells were then exposed to several concentrations
of the extracts or the positive controls cisplatin and 5-fluorouracil. After a treatment period
of 72 h, cell viability was estimated with the MTT assay or the resazurin assay [2,15]. Due
to the fact that plant extracts contain compounds (e.g., polyphenols) that may interfere
with these assays, cell culture medium was removed before conducting these assays in all
adherent cell lines.

The MTT assay is based on the ability of viable cells to convert the MTT compound
(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) into an insoluble and purple
formazan product. After an incubation period of the cells with the MTT and a solubilization
step, the quantity of the colored product was measured with a plate reading spectropho-
Plants 2021, 10, 2193

tometer. Dead cells are metabolically inactive and cannot produce the colored product. Briefly, after the 72 h treatment period, the medium was removed, the cells were washed with PBS, and 125 µL MTT (1 mg/mL in medium) was added to each well. The plates were incubated for 4 h to allow viable cells to transform the MTT compound into an insoluble formazan product. This insoluble compound was solubilized by adding 80 µL 20% SDS in 0.02M HCl to each well and by incubating the plates overnight at 37 °C. Finally, optical densities were measured at 540 nm on a multi-well plate spectrophotometer reader (Multiskan EX Labsystems, Vantaa, Finland).

The resazurin assay is a redox-based colorimetric or fluorometric assay based on the capacity of viable cells to reduce the blue compound resazurin into the pink, fluorescent and soluble product resorufin. The quantity of resorufin produced is proportional to the number of viable cells. After treatment period, medium was removed and 150 µL of resazurin (20 µg/mL in medium) was added to each well for 5–6 h. The absorbance was measured at 540 nm and 620 nm on a multi-well plate spectrophotometer reader (Multiskan EX Labsystems, Vantaa, Finland).

In both assays, cell viability was calculated as a percentage in relation to untreated cells from at least two independent experiments. IC50 values were calculated by linear regression analysis after selecting the two concentrations leading to cell viability values immediately above and below 50%. With this approach, IC50 values always are within the experimentally observed concentration range [15,34–37]. After estimating cell viability and calculating IC50 values, results were expressed as mean ± standard error of the mean (SEM), and a t-test (paired, two tailed) was used for statistical analysis. A p-value > 0.05 was not considered statistically significant and was not represented by any symbol. A p-value ≤ 0.05 was considered statistically significant and was represented with an asterisk, two asterisks (p ≤ 0.01), or three asterisks (p ≤ 0.001). Since selectivity is the most relevant parameter to detect anticancer potential in vitro [3,4], selectivity indices were used to quantify this parameter. The selectivity index (S.I.) was calculated as the average of the IC50 value in the normal cell line (MRC-5) divided by the IC50 value in the cancer cell line (A549) obtained in each independent experiment [3].

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10.3390/plants10102193/s1. Figure S1: Comparison of cytotoxicity of Tetraclinis articulata (Vahl) Mast. extract (58) assessed by MTT and resazurin assays. A549 and MRC-5 cells were treated for 72 h. Figure S2: Tetraclinis articulata (Vahl) Mast. (Cupressaceae). General view of the plant (a) and the plant material (b) used in the present study.

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