Vitamin D binding protein polymorphisms significantly impact vitamin D status in children

Danforth A. Newton, John E. Baatz, Mark S. Kindy, Sebastiano Gattoni-Celli, Judy R. Shary, Bruce W. Hollis and Carol L. Wagner

**BACKGROUND:** Polymorphic alleles of the vitamin D (vitD)-binding protein (VDBP) gene are associated with discriminatory differences in circulating concentrations of 25-hydroxyvitamin D (25-D), the indicator of vitD status (sufficiency defined by the Endocrine Society as ≥75 nmol/L). Within a diverse group of children, we hypothesized that reaching recommended daily allowance (RDA) of vitD intake would have differential impact on vitD status depending on VDBP variability.

**METHODS:** VDBP alleles (Gc1S, Gc1F, Gc2) in 123 children (1–4 annual visits/child; ages 1–8 years) were compared for relationships with serum 25-D concentrations and daily vitD intake.

**RESULTS:** In African-American children, reaching the vitD RDA was associated with significantly higher mean serum 25-D concentrations for the 20% carrying the VDBP 1S allele than for the large majority without this allele (77 vs. 61 nmol/L 25-D; p = 0.038). Children with the Gc1S/1S homozygous genotype (30% Caucasians, 24% Hispanics, 2% African-Americans) who met RDA had 51% (39 nmol/L) greater mean serum 25-D than those below RDA (p < 0.0001).

**CONCLUSIONS:** VDBP genetic variability was a significant factor affecting childhood vitD status when following RDA guidelines. This study may inform public health policy of uniformity in recommended childhood vitD dosage, especially regarding racially/ethnically associated disparities.

**INTRODUCTION**
Vitamin D (vitD)-binding protein (VDBP), originally called Gc globulin, is one of the most abundant serum proteins and transports 85–90% of circulating vitD metabolites. VDBP is also the product of one of the most polymorphic genes in the human population. For the most common VDBP alleles, significant frequency differences can be demonstrated within even small groups of racially diverse people. VDBP variants may directly impact interpretation of clinical data, as the isoforms have different binding affinities for vitD metabolites and are associated with discriminatory serum concentrations of these metabolites, including 25-hydroxyvitamin D (25-D; 25OHD; calcidiol), the principal circulating form accepted as the indicator of vitD status. Taken together with racial/ethnic-associated differences in vitD deficiency rates, as well as other health disparities, VDBP genetic variations may have important implications that have yet to be fully addressed in public health policy.

Among VDBP polymorphic alleles, three that result in distinct protein phenotypes have been most studied in association with racial/ethnic variations and potential effects on vitD metabolite measurements. Using traditional nomenclature directly referring to the protein phenotypes, the alleles are: (a) Gc1F (rs7041-T/rs4588-C); (b) Gc1S (rs7041-G/rs4588-C, single-nucleotide polymorphism (SNP) conferring D416E amino acid change from Gc1F); and (c) Gc2 (rs7041-G/rs4588-A, SNP conferring T420K amino acid change from Gc1F). Due to the amino acid substitutions, 1S and 2 VDBP phenotypes are also distinguished from the 1F isoform by significant post-translational glycosylation differences. As Gc1S and Gc2 polymorphisms are in linkage disequilibrium (i.e., both do not occur on the same chromosome), three allelic haplotypes and six genotypes are found in humans. Thus, one or two isofoms of VDBP circulate, depending if an individual has a homozygous (1F/1F, 15/15, 2/2) or heterozygous (1F/2, 1F/1S, 1S/2) genotype. The VDBP 1F allele, likely the primordial/wild-type form, predominates in individuals of African ancestry, while 1S and 2 alleles are more frequently associated with European or Asian ancestry.

Though synthesized endogenously, vitD deficiencies are common due to the fact that de novo biosynthesis of parent compound cholecalciferol (vitD3) involves a non-enzymatic, photolysis reaction requiring skin exposure to ultraviolet B radiation, the efficiency of which is affected by factors such as geography, skin tone, body mass index (BMI), and human behavior. Thus, because natural food sources are uncommon, vitD (D3 or D2, ergocalciferol) is commonly taken as a dietary supplement. Currently, the recommended daily allowance (RDA) of vitD for all children over 12 months old is 600 IU (15 μg), raised from 400 IU (10 μg) by the Institute of Medicine (IOM) in 2010. Most health policy agencies, including the IOM, World Health Organization, and Endocrine Society, agree that maintenance of serum 25-D >50 nmol/L (20 ng/mL) prevents most bone development and calcium homeostasis disorders associated with severe vitD deficiency.

This 25-D concentration can generally be achieved by daily vitD intake of 400–600 IU (10–15 μg). However, as understanding of the extent of vitD functioning in the human body has greatly expanded in recent years, including knowledge of numerous intracrine/paracrine roles, the Endocrine Society now recommends that serum 25-D ≥75 nmol/L (30 ng/mL) be considered vitD
METHODS

Study design

Participants were 123 children born to mothers previously enrolled in two completed pregnancy vitD supplementation trials (Carol L. Wagner, Stuart A. Hamilton, Bruce W. Hollis, P.I.s). This was part of a prescribed follow-up study conducted at the Medical University of South Carolina (MUSC). The subjects themselves were not part of a vitD supplementation protocol and not subject to any exclusion criteria. Parents gave their written informed consent for their child’s participation (MUSC Institutional Review Board protocol HR# 19641), completed sociodemographic questionnaires, and chose the racial/ethnic group to which their child belonged. All subjects were identified as African-American, Hispanic, or non-Hispanic Caucasian.

Participating children returned for 1 to 4 annual clinical visits, totaling 333 available data collection points. Eighty-five percent of children had their first visit at ages 3–5 years. For 333 total visits, represented ages, rounded off to year, were: 1 (1%), 2 (4%), 3 (16%), 4 (22%), 5 (26%), 6 (17%), 7 (11%), and 8 (3%) years old (subsequent visits were scheduled as close to 12 months later as possible). Participants were 48% male and 52% female.

At each annual visit, blood was collected for measurement of total serum 25-D concentration. Blood cells from one visit per subject were used for VDBP genotyping. Other data obtained from most visits included weight, skin colorimetry (IMS SmartProbe, Berkeley, CA, USA) and visualized by GelRed stain (Biotium, Carlsbad, CA, USA) and oligonucleotide primers: forward, 5′-TATGATCTCAGAAGGACATG-3′; reverse, 5′-AATCACAG-TAAAGAGGAGGT-3′ (synthesized by Integrated DNA Technologies, Coralville, IA, USA; GenBank L10641.1). PCR amplicons were then treated with either Haelll or StyI restriction enzymes that cleave at Gc1S or Gc2 polymorphic sites, respectively. Resulting fragments were separated on 4–12% TBE polyacrylamide gels (Invitrogen, Carlsbad, CA, USA) and visualized by GelRed stain (Biotium, Fremont, CA, USA) to assign one of six possible genotypes based on fragment sizes. RFLP analyses were repeated for any samples with unclear results; also, random samples were repeated for quality control. PCR reagents and enzymes were from New England Biolabs (Ipswich, MA, USA).

Data analyses and statistics

Comparisons of circulating 25-D concentrations to VDBP variants or race/ethnicity were made using Student’s t tests or analysis of variance, as appropriate. If numbers were sufficient for statistical power, 25-D values were compared in numerous ways: (a) mean of all subjects’ samples per grouping; (b) if subject had more than one annual serum sample, 25-D values associated with vitD intake above or below RDA were averaged before using in group mean calculations; (c) the 25-D value of each subject’s sample taken at the time of highest or lowest daily vitD intake; or (d) for individual subjects who provided different serum samples associated with both intake groups (above and below vitD RDA), paired analyses was performed using 25-D values from same subjects (values averaged if subject had more than one such sample per intake group). Linear relationships between vitD intake and circulating 25-D concentrations were determined by calculating Pearson’s correlation coefficient (r). Multiple regression analyses were used to determine correlations of 25-D with both vitD intake and sun exposure. For comparison, no groups with n ≤ 5 were used for calculations. P values <0.05 were considered significant.

RESULTS

Subjects and samples available for this study

VDBP genotyping was performed for three well-characterized alleles (Gc1F, 15 and 2) using blood from 123 children of diverse
racial/ethnic backgrounds born to mothers previously enrolled in two completed pregnancy vitD supplementation trials. The participating children in this follow-up study were not part of a vitD supplementation protocol themselves. The children returned for one to four annual clinical visits, for a total of 333 data collection points that were eligible for subsequent analyses; that is, samples for which VDBP genotype, 25-D measurements, and detailed dietary/supplement information were available (Table 1). The majority of these serum samples came from subjects 3–6 years old. To compare children of significantly different sizes over the study period, total daily vitD intake (from estimated dietary and supplemental contribution) was normalized either by body weight (as IU/kg) or calculated BMI. Though both methods obtained similar results, normalization by weight was slightly more linearly correlated to observed effects on 25-D concentration; thus, those values are reported herein.

Factors affecting study design and approach
The use of all available samples from the participating subjects was necessary for the statistical power to uncover some differences among the six VDBP genotypes due to numerous variables in this study population:

1. Frequencies of the three VDBP alleles were dramatically different among the subjects in the study (Table 1). For example, 75% of the African-American children, but no non-

---

**Table 1.** VDBP genetic variability in subject population by race/ethnicity and samples available for this study

| VDBP genotype or haplotype | A. Number of individual subjects (% of total individuals within group) | B. Number of annually collected serum samples (% of represented genotypes within group) | C. Samples collected when subjects were meeting the vitD RDA (normalized to >25 IU/kg) |
|---------------------------|--------------------------------------------------------------------------|----------------------------------------------------------------------------------|----------------------------------------------------------------------------------|
|                           | All subjects | African-American | Caucasian | Hispanic | All subjects | African-American | Caucasian | Hispanic |
| 1F/1F                     | A. 33 (27%)  | 30 (75%)         | 0 (0%)   | 3 (6%)   | 333          | 101              | 85         | 147       |
|                           | B. 84 (25%)  | 75 (74%)         | –        | 9 (6%)   | 333          | 101              | 85         | 147       |
|                           | C. 32        | 30               | –        | 2        | 333          | 101              | 85         | 147       |
| 1S/1F                     | A. 25 (20%)  | 6 (15%)          | 6 (18%)  | 13 (26%) | 25 (19%)     | 7                | 11         | 6         |
|                           | B. 72 (22%)  | 17 (17%)         | 15 (17.6%) | 40 (27%) | 60 (18%)     | 2                | 23 (27%)   | 35 (24%)  |
|                           | C. 24        | 7                | 11        | 6        | 24           | 7                | 11         | 6         |
| 1S/1S                     | A. 23 (19%)  | 1 (2.5%)         | 10 (30.5%) | 12 (24%) | 60 (18%)     | 2                | 23 (27%)   | 35 (24%)  |
|                           | B. 60 (18%)  | 2 (2%)           | 27 (31.8%) | 40 (27%) | 60 (18%)     | 2                | 23 (27%)   | 35 (24%)  |
|                           | C. 18        | 0                | 14        | 4        | 18           | 0                | 14         | 4         |
| 1S/2                      | A. 24 (19.5%) | 1 (2.5%)       | 10 (30.5%) | 13 (26%) | 68 (20%)     | 1                | 27 (31.8%) | 40 (27%)  |
|                           | B. 68 (20%)  | 1 (1%)           | 27 (31.8%) | 40 (27%) | 68 (20%)     | 1                | 27 (31.8%) | 40 (27%)  |
|                           | C. 25        | 0                | 13        | 12       | 25           | 0                | 13         | 12        |
| 1F/2                      | A. 13 (10.5%) | 2 (5%)          | 3 (9%)   | 8 (16%)  | 15 (12%)     | 3                | 4          | 7         |
|                           | B. 36 (11%)  | 6 (6%)           | 20 (14%) | 20 (14%) | 36 (11%)     | 6                | 20 (14%)   | 20 (14%)  |
|                           | C. 14        | 3                | 4         | 7        | 14           | 3                | 4          | 7         |
| 2/2                       | A. 5 (4%)    | 0 (0%)           | 4 (12%)  | 1 (2%)   | 5 (4%)       | 0                | 4 (12%)    | 1 (2%)    |
|                           | B. 13 (4%)   | –                | 10 (11.8%) | 3 (2%)   | 13 (4%)      | –                | 10 (11.8%) | 3 (2%)    |
|                           | C. 1         | –                | 1         | 0        | 1           | –                | 1          | 0         |
| All genotypes             | A. 123 subjects | 40              | 33        | 50        | 114 (34%)     | 40 (40%)         | 43 (51%)   | 31 (21%)  |
|                           | B. 333       | 101              | 85        | 147       |
|                           | C. 114 (34%) | 40 (40%)         | 43 (51%) | 31 (21%)  |

| 1F allele\(a\) (rs7041-T/rs4588-C) | A. 71 (58%) | 38 (95%) | 9 (27%)   | 24 (48%)  |
|                                     | B. 192 (58%) | 98 (97%) | 25 (29%)  | 69 (47%)  |
|                                     | C. 70        | 40      | 15        | 15        |

| 1S allele\(b\) (rs7041-G/rs4588-C) | A. 72 (59%) | 8 (20%)   | 26 (79%)  | 38 (76%)  |
|                                     | B. 200 (60%) | 20 (20%) | 65 (76%)  | 115 (78%) |
|                                     | C. 67        | 7       | 38        | 22        |

| 2 allele\(c\) (rs7041-T/rs4588-A) | A. 42 (34%) | 3 (8%)    | 17 (52%)  | 22 (44%)  |
|                                     | B. 117 (35%) | 7 (7%)   | 47 (55%)  | 63 (43%)  |
|                                     | C. 40        | 3       | 18        | 19        |

\(a\)1F/1F, 1S/1F, and 1F/2 genotypes
\(b\)1S/1F, 1S/1S, and 1S/2 genotypes
\(c\)1S/2, 1F/2, and 2/2 genotypes
Hispanic Caucasians, had the homozygous 1F/1F genotype. Conversely, nearly one-third of Caucasians, but only a single African-American child, had the homozygous 1S/1S genotype. Overall, only 20% of African-Americans in the study carried the 1S allele, most with the 1S/1F genotype. Comparing percentages in lines a and b of Table 1, the 333 available samples were extremely representative of the allelic frequencies within the population of individual subjects.

2. Total daily vitD intake between children also differed between racial/ethnic groups, with Hispanic children having much lower average daily intake than African-Americans or Caucasians (Table 1, line c). These differences in intake were largely attributed to the varying percentages of children taking supplements containing vitD at the time of the clinical visit: Caucasians (71%), African-Americans (47%), and Hispanics (25%). Consequently, only 21% of Hispanics in this study were receiving >25 IU/kg (>625 ng/kg) vitD daily (similar to 400–600 IU total vitD for most subjects), within the range of the RDA. All children had total daily vitD intakes of <1000 IU at the time of their annual clinical visits.

3. At the time of serum collection, a SmartProbe skin colorimeter was used as the primary estimate of sun exposure (i.e., by measuring the differences in skin pigmentation at 450 nm between areas normally exposed or covered by clothing), but other information was not available (e.g., hours of outside activity, sunscreen usage, etc.), nor were all clinic visits during the same time of year. These additional data are considered important for accurate estimates, especially for darker-skinned individuals. For children with low vitD intakes (below the RDA), significant linear correlations were calculated between sun exposure and circulating 25-D; however, this relationship was obscured in children vitD intakes above the RDA (data not shown). As the focus of this study was the relationship to vitD status between different VDBP genotypes and vitD exposure (i.e., not endogenous biosynthesis), these estimates of sun exposure were not considered further.

Taken together, some relationships of VDBP genotypes to a child’s vitD status and daily intake were more apparent if the subjects were analyzed separately by racial/ethnic groups, and that is the approach followed to clarify some of the data reported herein. However, due to the low percentage of Hispanic children meeting the vitD RDA at the time of serum collection, this group could not be analyzed individually.
Effects of vitD intake on children’s circulating 25-D concentrations

Without considering VDBP genotype, the children in this study differed significantly in vitD status, as measured by mean serum 25-D concentration (Fig. 1). As a group, Caucasians had much higher average 25-D concentrations than did African-Americans or Hispanics and met the ≥75 nmol/L threshold recommended by the Endocrine Society, even with low daily vitD intake.

If all subjects were considered together, the RDA of vitD intake (normalized to ≥25 IU/kg) did raise the children’s mean 25-D levels above 75 nmol/L (Fig. 1a), from an average of 64L to 81 nmol/L 25-D in children regularly receiving below or above the vitD RDA, respectively. However, as groups, African-American and Hispanic children on average showed minor effects of higher amounts of daily vitD intake on measured vitD status and had <75 nmol/L mean circulating 25-D, even with reaching the RDA (all subjects in this study had daily vitD intake of <000 IU) (Fig. 1a, b). As illustrated in Fig. 1, these relationships between vitD intake and vitD status were consistent in this population regardless of the method of sample analyses used, including paired analyses within the same children comparing 25-D values from different annual samples collected when those subjects were either below or above regular RDA intake (Fig. 1c).

![Image](image1.png)

Fig. 2 The VDBP 1S allele was associated with the highest circulating 25-D concentrations in children reaching the vitD RDA. a Comparisons of mean 25-D between children of different VDBP genotypes who regularly reached vitD RDA (normalized as >25 IU/kg) or not at the time of annual serum collection (308 total samples). No group of n≤5 compared (affected only 2/2 genotype). b Mean 25-D in children reaching the vitD RDA compared between subjects who carried the 1S allele and those who did not (47 and 33, respectively). If an individual child had more than one such sample, 25-D values were averaged. c Association of VDBP genotypes to mean 25-D within the same children who were both below and above vitD RDA at different annual serum collections (55 total subjects). Also shown are children grouped by carriers of the 1S allele per se. Dashed line in each graph represents threshold serum concentration of ≥75 nmol/L 25-D, vitD, Vitamin D, VDBP, vitamin D binding protein; 25-D, 25-hydroxyvitamin D, RDA recommended daily allowance.
relationships between VDBP genotype and 25-D concentration (data not shown).

The VDBP 1S/1S genotype is associated with the highest serum 25-D concentrations in non-Hispanic Caucasian children. As shown above, the majority of Caucasian children in this study had circulating 25-D \(\geq 75 \text{ nmol/L} \) at the time of their annual clinical visits (Fig. 1), and these children were most likely to be meeting the vitD RDA intake (Table 1). Thus, statistical comparisons of all individual VDBP genotypes in subjects with low vitD intake were not possible. However, within this non-Hispanic Caucasian population, VDBP 1S/1S individuals consistently had the highest circulating 25-D concentrations and the largest mean difference (40 nmol/L) between those either below or above vitD RDA intake (Fig. 3). Also, the linear correlations between vitD intake and 25-D concentrations were strongest between 1S/1S individuals (\( r = 0.55; p = 0.011 \)), than with other genotypes (\( r = 0.35, p = 0.0068 \)) (data not shown).

The VDBP 1S allele is associated with higher circulating 25-D concentrations in African-American children. Seventy-five percent of African-American children in this study were homozygous for the VDBP 1F allele. By comparison, only three Hispanic and no Caucasian children in this study had this 1F/1F genotype. Twenty percent of the remaining African-American children carried the VDBP 1S allele (1S/1F, 1S/2 or 1S/1S genotypes). At the time of their clinical visits, 40% of African-American children in this study were regularly meeting the vitD RDA (Table 1).

For African-American children with the majority VDBP 1F/1F genotype, meeting the RDA was not associated with mean serum 25-D \(\geq 75 \text{ nmol/L} \), regardless of the method of sample analysis (Fig. 4). Even in the subset of 1F/1F children obtaining substantially above RDA (between 600 and 1000 IU daily), mean serum 25-D did not reach 75 nmol/L (data not shown).

Conversely, the presence of the VDBP 1S isoform was associated with mean 25-D above the 75 nmol/L threshold in African-American children reaching the RDA (77–83 nmol/L mean serum 25-D, depending on the type of sample analysis used) (Fig. 4a, b). Though only 20% of these children carried the 1S allele (i.e., 1S/1F, 1S/2 and 1S/1S genotypes), mean 25-D concentration was still.
Vitamin D binding protein polymorphisms significantly impact vitamin D... DA. Newton et al.

significantly higher when meeting the RDA (p value = 0.038) than in those children who did not carry the 1S allele (i.e., 1F/1F and 1F/2 genotypes) (Fig. 4a). Also, linear correlations between vitD intake and 25-D were stronger for those children who carried the 1S allele (r = 0.41; p = 0.04), compared to those who did not (r = 0.24; p = 0.03) (data not shown).

DISCUSSION

This study analyzed the association of vitD intake to circulating 25-D concentration (i.e., vitD status) according to VDBP genetic diversity in a group of 123 children. These subjects returned for up to four annual follow-up clinical visits after their mothers completed clinical studies involving vitD supplementation during pregnancy.18–20 The results reported herein clearly demonstrate that the VDBP 1S allele is associated with the highest vitD status in children who regularly receive the RDA of vitD intake (currently set 600 IU by the IOM for children older than 12 months). The Endocrine Society currently defines "vitD sufficiency" as serum 25-D ≥75 nmol/L (≥30 ng/mL).15 Most strikingly, in those subjects who reached the RDA, the VDBP 1S allele was associated with mean circulating 25-D above this concentration even in in African-American children, a population that regularly falls below this threshold of vitD sufficiency. However, only 20% of African-American children in this study carried the 1S allele (most with the 15/1F genotype), which is consistent with the frequency seen other studies.4,6,10,24

Conversely, children homozygous for the VDBP 1F allele, including 75% of the African-Americans in this study, were much more likely to be vitD “insufficient” by standard definition (i.e., circulating 25-D ≤50 to <75 nmol/L), even when reaching the RDA. Thus, though the data here suggest further investigation through a controlled clinical study, in this population of African-American children, only the cohort that carried the VDBP 1S allele reached mean vitD sufficiency with RDA intake as currently defined. Though previous studies have also found a general association with the VDBP 1S allele and higher circulating 25-D concentrations,10,12,13,25 to our knowledge, this is the first time that the vitD intake within the range of the RDA has been so conclusively linked to differential effects on childhood vitD status due to VDBP genetic variability.

Probably originating as humans radiated out of Africa, the Gc1F, 1S, and 2 polymorphic alleles of VDBP have likely been maintained for millennia, and their frequent variations are well known to be associated with racial ancestry.5 Speculation as to the reasons for maintenance of this genetic diversity has varied widely from an association with skin tone, geographic differences in diet or sun exposure, and immunological pressure from pathogens.5,30,31 In this study of children from the southeastern United States, the frequencies of these VDBP alleles among racial/ethnic groups were very similar to those found in previous publications.4,6,10,24 Over 100 other VDBP polymorphisms have been found in the human population, but their frequencies and relationships to vitD intake within the range of the RDA have been less explored.

Though beyond the scope of this study, there is an on-going debate concerning the meaning of vitD status as determined by circulating 25-D, and if estimated calculations of free (unbound to protein) or "bioavailable" (free plus fraction bound to albumin, rather than to VDBP) metabolite would be better assessments of vitD sufficiency.32–35 Determination of vitD metabolite bioavailability is further complicated by the polymorphic forms of VDBP, which may have different steady-state circulating concentrations (e.g., resulting from protein synthesis and/or turnover rates) and 25-D-binding affinities.2,5,26 Taken together, the known associations of VDBP polymorphisms with quantitative differences in circulating 25-D concentrations among individuals could have important implications for vitD-related physiology.11 This would be especially true concerning the intracrine and paracrine functions of vitD, such as in immune cells, which likely compete with circulating VDBP for binding 25-D (these cells, like many others, can synthesize the active metabolite, 1,25(OH)2D, from 25-D for intracellular use).36–38 For example, in vitro assays clearly demonstrate that increasing VDBP concentrations can abrogate monocyte/macrophage vitD signaling, an effect that can be discriminated by using different VDBP allelic forms.38

The meanings of vitD "sufficiency" or "deficiency" are still controversial. If referring solely to circulating 25-D concentrations associated with the endocrine role of vitD, then serum calcium and PTH concentration, bone density, and growth are likely good clinical correlates of deficiency. Even most current studies addressing the relationship of free or bioavailable vitD metabolites to VDBP genetic diversity are solely focused on this endocrine role of vitD (e.g., are African-Americans truly vitD deficient with low circulating 25-D if they have high bone density?). However, a large body of research over the past few decades has now elucidated hundreds of other genes and physiologic processes affected by vitD, many of which are associated with subtle, homeostatic regulation that could potentially influence life-long health, and are much more difficult to correlate to simple clinical/pathological measurements. This accumulated research is what primarily led the Endocrine Society to distinguish between deficiency, insufficiency, and sufficiency in circulating 25-D. Although the official recommendation of sufficient in that report was for circulating 25-D of ≥75 nmol/L, even a range of 100–150 nmol/L (40–60 ng/ml) was considered safe and would likely guarantee full sufficiency in most children and adults.15

Though future research may indeed change interpretations of vitD sufficiency, the primary objective of our study was to determine if potential relationships existed between VDBP polymorphisms and the currently accepted clinical determination of vitD sufficiency in children as defined by the Endocrine Society, and if these data would impact the current uniformity of vitD RDA as determined by the IOM. There were several limitations to this study, most associated with the moderate sample size (123 subjects, 333 data collection points) and the fact that the children were not part of a vitD supplementation protocol. Most of these limitations were incorporated into the study design as discussed in detail in Results section. Since this population of children were not part of a clinical trial of vitD supplementation themselves, most factors normally controlled in such a study (e.g., equivalent vitD intake cohorts) were highly variable. In fact, this prevented the separate analysis of Hispanic children here, since the large majority of the subjects in this group fell far below the RDA of vitD intake. Though beyond the scope of this report, many other measurements needed to further elucidate the physiological effects associated with VDBP diversity were also unavailable, including circulating PTH, Ca++, 1,25-dihydroxyvitamin D, and free or bioavailable 25-D. Thus, interpretations here were limited to the primary data available: serum 25-D concentration and detailed dietary information. Again, however, these are the only two measurements currently accepted as indicators of vitD status/sufficiency and RDA of vitD intake, respectively.

In summary, these results clearly indicate that VDBP genotypic variability is a significant factor in determining vitD status associated with recommended vitD intake in children. Furthermore, these data suggest additional studies addressing recommended standards as they relate specifically to African-American children, the majority of whom lack the 1S allele associated with higher circulating 25-D. Along with recent research into the significance of vitD metabolite bioavailability, these data may challenge public health policy related to current uniformity in recommended vitD daily allowance, especially with regard to racially/ethnically associated disparities in children.
Acknowledgements

We would like to thank Renée Washington and Dr. Kaleena Dezi for assistance in completing these studies. This study was supported by Thrasher Research Fund and NIH/NCRR UL1 RR029882, UL1 TR000062.

Author Contributions

Conception/design/data acquisition and analyses: All authors. Drafting/revision: D.A.N., J.E.B., C.L.W. Final approval: All authors.

Additional Information

Competing interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

References

1. Cooke, N. E. & David, E. V. Serum vitamin D-binding protein is a third member of the albumin and alpha fetoprotein gene family. J. Clin. Invest. 76, 2420-2424 (1985).

2. White, P. & Cooke, N. The multifunctional properties and characteristics of vitamin D-binding protein. Trends Endocrinol. Metab. 11, 320-327 (2000).

3. Cleve, H. & Constans, J. The mutants of the vitamin-D-binding protein: more than 120 variants of the GC/DBP system. Vox. Sang. 54, 215-225 (1988).

4. Speeckaert, M., Huang, G., Delanghe, J. R. & Taes, Y. E. Biological and clinical aspects of the vitamin D binding protein (Gc-globulin) and its polymorphism. Clin. Chim. Acta 372, 33-42 (2006).

5. Cooke, N. E. & Haddad, J. G. Vitamin D binding protein (Gc-globulin). Endocr. Rev. 10, 294–307 (1989).

6. Kamboh, M. I. & Ferrell, R. E. Ethnic variation in vitamin D-binding protein (GC): a review of isoelectric focusing studies in human populations. Hum. Genet. 72, 281–293 (1986).

7. Braun, A., Bichlmaier, R. & Cleve, H. Molecular analysis of the gene for the human vitamin-D-binding protein (group-specific component): allelic differences of the common genetic GC types. Hum. Genet. 89, 401–406 (1992).

8. Holick, M. F. Vitamin D status: measurement, interpretation, and clinical application. Ann. Epidemiol. 19, 73–78 (2009).

9. Arnaud, J. & Constans, J. Affinity differences for vitamin D metabolites associated with the genetic isoforms of the serum carrier protein (DBP). Hum. Genet. 92, 183–188 (1993).

10. Lauridsen, A. L. et al. Plasma concentrations of 25-hydroxy-vitamin D and 1,25-dihydroxy-vitamin D levels in Hispanic and African Americans. J. Clin. Endocrinol. Metab. 93, 3381–3388 (2008).

11. Fu, L. et al. Common genetic variants of the vitamin D-binding protein (DBP) predict differences in response of serum 25-hydroxy-vitamin D (25(OH)D) to vitamin D supplementation. Clin. Biochem. 42, 1174–1177 (2009).

12. Yuen, A. W. & Jablonski, N. G. Vitamin D in the evolution of human skin colour. Med. Hypotheses 74, 39–44 (2010).

13. Kappelman, J. et al. First Homo erectus from Turkey and implications for migrations into temperate Eurasia. J. Hum. Evol. 135, 110–116 (2008).

14. Bilek, D., Bouillon, R., Thadhani R. & Schoenmakers I. Vitamin D metabolites in captivity? Should we measure free or total 25(OH)D to assess vitamin D status? J. Steroid Biochem. Mol. Biol. 137, 105–116 (2017).

15. Powe, C. E. et al. Vitamin D-binding protein and vitamin D status of black Americans and white Americans. N. Engl. J. Med. 369, 1991–2000 (2013).

16. Chun, R. F. et al. Vitamin D and DBP: the free hormone hypothesis revisited. J. Steroid Biochem. Mol. Biol. 144, Pt A. 132–137 (2014).

17. Sheih, A. et al. Effects of cholecalciferol vs calcifediol on total and free 25-hydroxyvitamin D and parathyroid hormone. J. Clin. Endocrinol. Metab. 102, 1133–1140 (2017).

18. Jeffery, L. E. et al. Availability of 25-hydroxyvitamin D(3) to APCs controls the balance between regulatory and inflammatory T cell responses. J. Immunol. 189, 5155–5164 (2012).

19. Liu, P. T. et al. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. Science 311, 1770–1773 (2006).

20. Chun, R. F. et al. Vitamin D-binding protein directs monocyte responses to 25-hydroxy- and 1,25-dihydroxyvitamin D. J. Clin. Endocrinol. Metab. 95, 3368–3376 (2010).

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, and provide a link to the Creative Commons license. You do not have permission under this license to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.

© The Author(s) 2019