Chapter

IncRNAs in Hallmarks of Cancer and Clinical Applications

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Abstract

Long noncoding RNAs (IncRNAs) are transcripts longer than 200 nucleotides in length that, in general, do not appear to have protein-coding potential. IncRNAs act in gene regulation involved with several biological processes. Furthermore, IncRNAs have been associated with a significant number of cancers, suggesting a potential role in tumorigenesis and progression. For example, HOTAIR regulates proliferation processes and other IncRNAs like highly upregulated in liver cancer (HULC), H19, PTENP1, HEIH, and antisense noncoding RNA in the INK4 locus (ANRIL). Other IncRNAs as AFAP1-AS1 and lincRNA-p21 can interact with BCL-2 and TP53, acting in apoptosis. Moreover, NORAD plays a vital role in genomic stability. Additionally, due to deregulated expression and high tissue specificity level, IncRNAs exhibit great potential as prognostic markers. In this chapter, we review the most highlighted IncRNAs acting in hallmarks of cancer and clinical application.

Keywords: cancer, hallmarks, IncRNAs, ncRNAs, tumor

1. Introduction

The landscape of human transcriptome is more complicated than was imagined. In the last decade, the technology of RNA sequencing reveals more than 100,000 different RNA molecules produced by mammalian organisms [1, 2], most of them, without protein-coding potential, named as noncoding RNAs (ncRNAs). These molecules called the attention for their multiple roles in cell physiology. NcRNAs are classified, by the size, as small (microRNAs 22~25 bp) or long, with more than 200 nucleotides [3]. Previously, it was considered that IncRNAs were “dark matter” or “transcriptional noise” of the human transcriptome, with no biological functions. Recently, IncRNAs were found in all the branches of the tree of life, and their amount and diversity are more correlated with organismal complexity than protein-coding genes [4].

The majority of IncRNAs is transcribed by RNA polymerase II, capped and polyadenylated with some IncRNAs being also spliced. They are described as noncoding RNAs. New studies have shown functional micropeptides derived from some of the IncRNAs [5]. Until now, 16,000 IncRNAs were identified in the human genome with approximately 30,000 distinct IncRNA transcripts according to the Encyclopedia of DNA Elements (ENCODE) Project Consortium (GENCODE release 30). This number continues to increase, mainly through sensitive RNA sequencing and advanced
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bioinformatics pipelines. IncRNAs have a lower expression level than other RNAs, and they show specific expression in tissues [6, 7], cell types, and subcellular compartments [8]. IncRNAs are classified according to their relative position to protein-coding genes in a sense, antisense, bidirectional, intronic, and intergenic [9]. Also, IncRNAs can be regulated by well-established transcription factors and associated with epigenetic signatures that modify chromatin states, making the IncRNA loci more accessible in the cell [10].

The current knowledge about IncRNAs is essential to understand cell biology, especially in cancer cells. Cancer is a complex disease characterized by extreme genetic and epigenetic changes that can fundamentally alter cell homeostasis to promote uncontrolled cell growth. Emerging evidence suggests that IncRNAs are involved with cancer-associated phenotypes like resisting cell death, invasion, proliferation, gene deregulation, and genomic instability and evade growth suppressors [11]. IncRNAs also interact with transcriptional regulation of tumor suppressors or oncogenes [12, 13]. One example is lincRNA-p21 that acts as a repressor in p53-dependent transcriptional responses [12]. Alternatively, HOTAIR can increase metastasis in primary breast tumors and hepatocellular carcinomas [14].

IncRNAs can participate in gene regulation at transcriptional and posttranscriptional levels [15]. For example, related to epigenetic mechanisms, IncRNAs can recruit methyltransferases [16] and polycomb complex [17] to prevent DNA accessibility through histone modification. IncRNAs are also involved in several posttranscriptional processes, such as splicing and nuclear export, mRNAs localization and stability, and in protein translation process [18–22].

IncRNAs can serve as a molecular scaffold, enhancing the interactions between protein–protein, protein–RNA, and protein–DNA, by base complementarity or interaction by secondary structures [23]. Alternatively, IncRNAs can function as a decoy when they titrate transcription molecules and other proteins away from the target [24]. Additionally, IncRNAs can work as binding platforms regulating miRNAs competing with mRNAs for miRNA response elements, known as competitive endogenous [25].

The development of several discoveries about the role of IncRNAs, especially in cancer, highlighted the importance of gene regulation in cellular functions. The evidence that we show here supports the idea that IncRNAs have an essential role in tumorigenesis and are associated with several cellular processes. Here, we review the current knowledge about IncRNAs in hallmarks of cancer and their potential for clinical application.

2. IncRNAs act in hallmarks of cancer

The transformation of a regular cell into cancer involves several processes, including molecular and environmental alterations [26]. The healthy cells must acquire different abilities to change the cell physiology and dictate malignant growth. The hallmarks of cancer comprise the biological capabilities acquired during the multistep development of human tumors. These changes include sustaining proliferative signaling, evading growth suppressors, resisting cell death, enabling replicative immortality, inducing angiogenesis, and activating invasion/metastasis [27]. Additionally, genomic instability, inflammation, reprogramming of energy metabolism, and evading immune destruction were also included [28]. There are many IncRNAs well described associated with cancer [29], and, herein, we highlighted some IncRNAs with strong evidence for cancer process association and with molecular details, for example (Figure 1). The influence of those long noncoding RNAs in hallmarks of cancer is due to the regulation of different pathways.
lncRNAs have virtual association in all the hallmarks of cancer, some of them linked with more than one hallmark. For example, the expression of ANRIL, growth arrest-specific transcript 5 (GAS5), urothelial cancer-associated 1 (UCA1), and HULC regulates the resistance to cell death. Also, ANRIL, HULC, and H19 are associated with the evasion of growth suppressors, GAS5, and H19, sustaining cell proliferation. UCA1 and H19 are also associated with invasion and metastasis. Moreover, UCA1 is associated with metabolism reprogramming and GAS5 with an inflammatory process.

In more detail, ANRIL may inhibit apoptosis by silencing \textit{KFL2} and \textit{p21} genes, involved in cell proliferation. Moreover, regulating MET and MMP3 proteins that facilitate cell migration and invasion \cite{30–34}.

The lncRNA GAS5 can sustain proliferation by regulating glucocorticoid receptors (GR), with strong influences in cell growth and repression of anti-apoptosis genes. lncRNA binds to the DNA-binding domain of GR, competing with glucocorticoid response element \cite{35}.

lncRNA is also observed in cell cycle regulation, whereas it arrests cells in the G0/G1 phase. In stomach cancer, GAS5 binds to YBX1 to regulate \textit{p21} expression, enhancing G1 arrest. In bladder cancer, GAS5 associates with CDK6 and reduces both \textit{CDK6} mRNA and protein levels, resulting in the inhibition of cell proliferation. Besides, GAS5 acts in the immune response by NF-\textit{kB} and ERK1/2 pathways. The knockdown of GAS5 increases expression and secretion of interleukin-10 (IL-10) and vascular endothelial growth factor (VEGF-A), the essential cytokines involved in inflammation \cite{36}.

Due to its oncogenic regulatory potential, the lncRNA UCA1 regulates the proliferation and migration targeting the transcription factor KLF832 in pancreatic cancer. UCA1 also promotes invasion and metastasis through activation of \textit{MMP14}, \textit{FGFR1/ERK}, and \textit{ZEB1/2-FSCN1} \cite{37–40}. In hypoxic conditions, \textit{HIF-1\alpha} effectively activates UCA1 transcription by binding in the \textit{UCA1} promoter, inducing proliferation, migration, invasion, and apoptosis resistance of cancer cells \cite{41}.

Several pathways regulate the switch of oxidative phosphorylation to aerobic glycolysis, such as the PI3K, AMPK, p53, and HIF-1 pathways. These pathways
are involved in metabolism deregulation associated with the Warburg effect, which consists in the special metabolism of glucose in the cytosol even in the presence of oxygen \[42, 43\], and UCA1 acts indirectly in genes involved in this process \[44, 45\].

HULC contributes to malignant phenotypes by, at least, three mechanisms. The first is regulating the expression of P18, an auxiliary protein of cell cycle that inhibits the tumor suppressors CDK4 and CDK6. Also, hepatitis B virus infection activates the HULC promoter and induces cell cycle progression by downregulation of P18 \[46\]. The other two mechanisms are related to angiogenesis. In breast and liver cancer, HULC sequesters miR-107 and regulates the transcription factor E2F1/SHPK1 \[47, 48\]. In glioma, HULC acts through PI3K/Akt/mTOR signaling pathway, inducing ESM-1/VEGF-A and affecting vascular permeability and cell mobilization \[49\].

Some lncRNAs have been described more exclusively in only one hallmark, for example, telomeric repeat-containing RNA (TERRA) associated with replicative immortality. We know that cells have a limited survival rate that can be explained by telomere end loss, which generate a waste of genetic conservation, restricting the number of mitosis in a tissue. Thus, neoplastic cells can escape this telomere process with the help of telomerase. This enzyme can increase the size of telomere adding repeats on the edge 3’ in chromosomes \[50\]. TERRA comprises a heterogeneous class of lncRNAs transcribed from telomeric regions \[51\]. TERRA transcripts negatively regulate the activity of telomerase, acting as a tumor suppressor. Besides acting as telomere maintenance and genome stability, TERRA is also regulated by genes, such as TP53 and RB, highlighting that TERRA transcripts can be crucially involved in tumorigenesis \[52–54\].

Another critical feature of neoplastic cells is in genomic instability, which can be related to defects in the DNA repair machinery. Activation of telomerase, following by individual DNA variations, activating proto-oncogenes or deactivating tumor suppressor genes, conferring a selective advantage on subclones of tumor cells, enabling their survival and outgrowth \[28\].

One lncRNA that is important in this tumor cell feature is NORAD. lncRNA protects the cells against aneuploidy by binding to PUM1/PUM2 proteins and suppressing their binding to other targets, including those that maintain genomic stability \[55, 56\]. An alternative mechanism to define the relationship among NORAD and genomic stability is that a nuclear ribonucleoprotein complex, named NORAD-activated ribonucleoprotein complex 1 (NARC1), is joined by NORAD, recruiting proteins known to act as suppressors of genomic instability, such as topoisomerase I (TOP1), ALYREF, and the PRPF19-CDC5L \[57\].

The implication of lncRNAs in cancer development and progression has been proved in the last decade and indicates that those new class of RNAs has a great potential as biomarkers on cancer and a future perspective to targeted specific therapies.

3. Clinical application for lncRNAs

As we discussed in earlier topics, we have examples of lncRNAs that participate in essential processes in tumor development. Many of them have great potential as diagnostic/prognostic markers and therapeutic targets. A great example is the lncRNA prostate cancer antigen 3 (PCA3), already used as a molecular marker in prostate cancer \[58, 59\]. PCA3 is a prostate-specific lncRNA overexpressed in 95% of prostate cancer cases. PCA3 may be detected by in vitro nucleic acid amplification
in urine specimens, and the US Food and Drug Administration approved the test in 2012 [60].

In cases of suspicion of prostate cancer, the PCA3 test is recommended, based on prostate-specific antigen (PSA) level and post-digital rectal examination with biopsy results. PCA3 has a high expression in prostate cancer without any correlation to prostatic volume and other prostatic diseases. This feature makes a PCA3 an attractive biomarker [61], but some recent studies question the use of this isolated biomarker and propose that the test should be carried out in association with another test, like TMPRSS2:ERG quantification [58].

Although there are few lncRNAs used in medical practice, many are being discovered and tested. For example, a treatment protocol for triple-negative high-risk breast cancer predicted by the integrated mRNA-lncRNA signatures is initiated in the clinical trials evaluated, to validate the efficacy of lncRNA signature [62].

Also, in clinical trials, there is an early phase study to evaluate the HOTAI R as a potential lncRNA biomarker in thyroid cancer. Many lncRNAs have a different expression in tumors when compared to healthy tissues and are strongly associated with clinical parameters, making them a candidate for tumor markers or even therapeutic targets [63].

A study found the downregulation of expression of downregulated in liver cancer (DILC) in colorectal cancer tissues compared to their adjacent healthy tissues and the normal colorectal tissues. The downregulation of DILC was associated with aggressive clinical characteristics, including depth of invasion and advanced TNM stage, and the lower expression of DILC was associated with more reduced survival and disease-free survival. With multivariate analyses, the authors confirmed that the expression of DILC was an independent prognostic factor in colorectal cancer [64]. Most of lncRNA papers characterize biomarkers that are specific to one type of cancer, such as those cited above. However, some lncRNAs are found differentially expressed in several types of cancer compared with healthy tissues, like the loc285194 lncRNA [65].

In addition to the association of lncRNAs with stage and prognosis, their association with drug resistance is also possible [66]. Several works have tried to reallocate the lncRNAs in the mechanism of resistance to the primary drugs used in the treatment of cancer. For example, Campos-Parra et al. [67] have identified several lncRNAs that participate in resistance mechanisms to several drugs utilized in the therapy of breast cancer. Most studies with lncRNAs measure their expression in tissues, but it is possible to detect and quantify their presence in other types of samples, like whole blood, plasma, urine, gastric acid, and saliva [68].

Within the use of lncRNAs as biomarkers in cancer, these molecules have applications as therapeutic targets in the development of new treatments and drugs. New therapeutic strategies are already focusing on noncoding RNAs, such as silencing via small interfering RNA (siRNA), an antisense oligonucleotide (ASO)-based strategies and other molecular inhibitors further modulating lncRNA expression by gene editing [69, 70]. An experimental model that aims to modulate lncRNAs in cancer cells is the use of siRNAs, which can decrease the amount of a target lncRNA, since they are complementary molecules to the sequence of lncRNA, promoting lncRNA binding and subsequent degradation. Although this methodology may be functional in many studies, some lncRNAs are not efficiently reduced by siRNA [70]. This methodology is efficient for cytoplasmic lncRNAs since the siRNA mechanism is located predominantly in the cytoplasm. In this case, siRNA does not silence nuclear lncRNAs [11, 71].

Another mechanism used to block lncRNAs activity relies on the ability of lncRNAs to bend and create secondary structures and on the ability of protein
interactions in lncRNAs. RNAi molecule can compete with the protein for the binding site; or when it binds to the target lncRNA, it changes the structure of the RNA, disrupting the binding site of the protein [72, 73].

For nuclear lncRNAs, an alternative strategy is the use of antisense oligonucleotides, which function predominantly in the nucleus [71]. ASOs modulate gene expression by inducing ribonuclease H cleavage of the duplex DNA-RNA. A limitation of the ASO and siRNA strategies is the possibility of non-specific targets, as well as the inconvenience of incomplete knockdown and transient modulation [74]. One method that has shed light on ncRNA-based cancer therapy and solves the problem of target specificity is genome editing by clustered regularly interspaced short palindromic repeats-associated endonuclease 9 (CRISPR/Cas9). The Cas9 nuclease can act guided to generate site-specific DNA cleavage in the genome, by an optimized equivalent single-guide RNA (sgRNA) [75]. That is, it can delete lncRNA genes or introduce RNA-destabilizing elements into their locus.

A limitation to apply CRISPR/Cas9 system to noncoding genes is that tiny indels may not necessarily generate a functional loss of a specific noncoding gene and the most protocols can perform small point mutations; plus not all lncRNAs CRISPR can be applied. Another limitation is that, although it is more specific than other systems, this technique may still have off-target effects [76, 77].

Currently, many studies performed lncRNA modulation technologies, both in vitro and in vivo. Moreover, while these models may resemble reality, the clinical use of modulation technology has a barrier that still needs to be broken: an efficient delivery to the target. There are some techniques for delivering lncRNA modulation systems to live cells, based on viral and non-viral methods, but both ways have limitations and problems to be solved before the utilization in clinical practice.

The main advantage of viral vectors is their innate ability to efficiently transfer the genetic material into the cell and the possibility of infecting specific cells. However, this technique also has a significant disadvantage, which is relatively high immunogenicity and toxicity. The possibility of generating an immune response is the main challenge for the use of this tool [78, 79].

Non-viral vectors are becoming recognized as an alternative to the immunogenicity of viral vectors, although their transfection capabilities usually do not reach such levels. Besides its main strengths are the low capacity to generate an immune response and the relatively easy and inexpensive synthesis with large-scale production and safety, which make them very attractive delivery systems for in vivo application. However, the target specification still needs to be better developed [79].

In order to choose the best delivery system, it is necessary to consider the type of cell/tissue, since some tissues are more accessible than others. When the target of therapy is a difficult-to-access tissue, some strategies may improve delivery efficiencies, such as binding to specific targeting elements like antibodies, carbohydrates, and synthetic peptides. These molecules also have their advantages and limitations; for example, although antibodies have a high recognition specificity and interaction with different receptors on target cells, they may be immunogenic and chemically unstable. Peptides and carbohydrates, on the other hand, demonstrate low immunogenicity, but the binding affinity to the target is lower [79, 80].

All the techniques cited in this topic have their limitations. Although in vitro studies present satisfactory results that have the potential to use in clinical aspects. However, these techniques require better improvements, especially in the delivery of drugs in specific targets. Despite this, our knowledge of lncRNAs linked with clinical applications creates hope for the development of better biomarkers and therapeutic targets. In Table 1, we list the main lncRNAs that have excellent potential for biomarker and therapeutic target.
| Cancer                | Refs. | AFAP1-AS1 | ANRIL | CCAT1/CCAT2 | CRNDE | DANCER | GAS5 | H19 | HOTAIR | HULC | LINC00152 | LincRNA-p21 | MALAT1 | MALAT2 | MEG3 | MIAT | NEAT1 | PANDA |
|-----------------------|-------|-----------|-------|-------------|-------|--------|------|-----|--------|------|-----------|------------|---------|---------|------|------|-------|-------|
| Head/neck             |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Gastric               |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Lung                  |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Breast                |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Colon                 |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Liver                 |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Uterine               |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Ovarian               |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Prostate              |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Bladder               |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Renal                 |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| SNC                   |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Leukemia              |       | X         | X     | X           | X     | X      | X    | X   | X      | X    | X         | X          | X       | X       |       |      |       |       |
| Refs.                 |       | [81]      | [82]  | [83]        | [84]  | [85]   | [86] | [87]| [88]   | [89] | [90]      | [91]         | [92]    | [93]    | [94] | [95] | [96]  | [97]  | [98]  |
## Table 1.

*IncRNAs with potential to be used as biomarkers, in several types of cancer.*

| IncRNAs | Head/neck | Gastric | Lung | Breast | Pancreas | Liver | Colon | Uterine | Ovarian | Osteosarcoma | Prostate | Bladder | Renal | SNC | Leukemia | Refs. |
|---------|-----------|---------|------|--------|----------|-------|-------|---------|---------|-------------|---------|---------|-------|-----|----------|-------|
| PCA3    |           |         |      |        |          |       |       |         |         |             |         |         |       |     |          | [99, 100] |
| PCAT-1  | x         | x       | x    | x      |          | x     | x     | x       |         |             | x       |         |       |     |          | [101, 102] |
| TUG1    | x         | x       | x    | x      |          | x     | x     | x       | x       |             | x       | x       |       |     |          | [103, 104] |
| XIST    | x         | x       | x    | x      |          | x     | x     | x       | x       |             | x       | x       |       |     |          | [105] |
4. Conclusion

The vast number of studies describing lncRNAs associated with several tumor types and regulating several processes of cancer cells is shown here. The great advance in RNA sequencing technology allows us to identify new molecules and characterized better lncRNAs. From the discovery of these molecules, in the beginning, they appeared to have no important functions; however, today many researches in this area propose that more information about these molecules may help us understand numerous characters of tumor cells that are still unknown. Some lncRNAs are associated with several hallmarks of cancer demonstrating the importance of these molecules in the mechanism of disease, like MALAT and HOTAIR. Other are already utilized as biomarker in prostate cancer like PCA3. Considering the challenges for in vivo experimental designs, lncRNAs continue to be promising as biomarkers and potential therapeutic targets.

Conflict of interest

The authors declare no conflict of interest.

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Non-Coding RNAs

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