Ertapenem non-susceptibility and independent predictors of the carbapenemase production among the Enterobacteriaceae isolates causing intra-abdominal infections in the Asia-Pacific region: results from the Study for Monitoring Antimicrobial Resistance Trends (SMART)

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Objectives: This study investigated the prevalence rates of carbapenemase positivity, antibiotic susceptibility, and independent predictors of carbapenemase producers among the Enterobacteriaceae isolates recovered from patients with intra-abdominal infections (IAI) in the Asia-Pacific region between 2008 and 2014.

Materials and methods: Multiplex PCR was used for the detection of specific β-lactamases, while the broth microdilution method was used to determine the minimum inhibitory concentrations (MICs) of antibiotics among the IAI-related Enterobacteriaceae isolates. We studied the abovementioned parameters in 484 ertapenem-non-susceptible (Erta-NS) isolates and explored the independent predictors of carbapenemase-producing Enterobacteriaceae (CPE) isolates.

Results: Eighty (16.5%) Ert-NS-IAI Enterobacteriaceae isolates were found to be CPE. In Vietnam and the Philippines had the highest CPE prevalence rates. The IAI isolates of Enterobacter species and Klebsiella pneumoniae followed by Escherichia coli were the three major pathogens with 77.4%, 40.9%, and 11.7% Ert-NS prevalence rates, respectively. Furthermore, the highest CPE prevalence (35%) was noted among the Ert-NS-K. pneumoniae isolates. The CPE isolates harboring the blaNDM, blaKPC, or blaOXA-48-like alleles had higher imipenem MIC levels than those harboring the blaTEM alleles. Using multivariate logistic regression analysis, we concluded that Ert-NS-IAI isolates with an imipenem non-susceptible phenotype (OR, 56.4), with cefepime MIC >8 µg/mL (OR, 4.4), cultured from the peritoneal space samples (tissue or abscess; OR, 3.3), and harboring the extended-spectrum β-lactamase encoding allele (OR, 11.5) are independent predictors of CPE.

Conclusion: Imipenem non-susceptibility, cefepime MIC >8 µg/mL, and the peritoneal space as a culture site are independent clinical predictors of CPE among the Ert-NS-IAI Enterobacteriaceae isolates in the Asia-Pacific region.

Keywords: intra-abdominal infection, antimicrobial susceptibility, ertapenem-non-susceptible, carbapenemase-producing Enterobacteriaceae

Introduction

Infections with septicemia usually pose a threat to the survival of many inpatients. Among the diverse infection categories, intra-abdominal infections (IAI), mostly caused
by Enterobacteriaceae species,1 are diseases with high fatality risks.2,3 In the last decade, resistance to carbapenem drug(s) among Enterobacteriaceae isolates has been gradually growing to be a critical concern for hospitalized patients worldwide.4–12 An Asia-Pacific IAI study conducted prior to 2010 by Sheng et al13 showed that the carbapenemase-producing Enterobacteriaceae (CPE) isolates accounted for up to 11.0% of the Enterobacteriaceae isolates having positive phenotypes for extended-spectrum β-lactamase (ESBL) production or ertapenem non-susceptibility (minimum inhibitory concentration [MIC] of ertapenem ≥1 µg/mL). In that study, the CPE predominantly harbored the alleles encoding New Delhi metallo-β-lactamases (NDM) and were mostly identified in India.13 However, this differs from the data in another recent study on Asia-Pacific IAI Enterobacteriaceae.14 We observed the emergence of NDM producing Enterobacteriaceae in many other Asian countries and Australia, as this worrisome situation challenges infection control.

Among all carbapenem agents, ertapenem is most vulnerable to hydrolysis of the advanced β-lactamases (including ESBL plus AmpC or most carbapenemases) in carbapenem-resistant Enterobacteriaceae (CRE) isolates. Therefore, non-susceptibility to ertapenem is deemed a sensitive indicator for the initial screening for potential carbapenemase production in the Enterobacteriaceae isolates.15 However, when compared to the high case fatality rates (44%–67%) in patients with sepsis due to CRE (producers of Klebsiella pneumoniae carbapenemase [KPC]) showing in vitro non-susceptibility to imipenem,16,17,18 patients infected with Enterobacteriaceae isolates having ertapenem resistance (MIC >2 µg/mL, in accordance with the criteria in 2009) but relative susceptibility to imipenem correlated with lower mortality rates (10%–31%).17,18 Therefore, the infection entities related to ertapenem-non-susceptible (Erta-NS) Enterobacteriaceae with or without susceptibility to imipenem may have different clinical outcomes. In addition, in a study by Tamma et al,19 the MIC values for imipenem and meropenem in the subgroup of CPE isolates (>90% were KPC producers) were significantly higher than those of the non-CPE subgroup. Furthermore, the overall 14-day mortality rate was four-fold higher in patients with CPE bacteremia than that seen in the non-CPE group. Consequently, discrimination of clinical infections caused by carbapenem-NS Enterobacteriaceae isolates by non-CPE and CPE is useful in predicting the outcomes of hospitalized patients. Using our existing database documenting the demographic factors, abdominal sampling sites for culture, health care settings for sample collection, as well as the detailed in vitro susceptibility testing results on bacterial isolates from Asia-Pacific IAI patients, we attempted to identify the independent predictors of CPE among the clinical Erta-NS-IAI Enterobacteriaceae isolates that produced important plasmidic β-lactamase(s) (including AmpC, ESBL, and carbapenemases). In addition, because new susceptibility break points of cefepime against Enterobacteriaceae species were proposed in Clinical and Laboratory Standards Institute (CLSI),20 we used different MIC levels for cefepime (8 and 16 µg/mL) to explore the independent predictors of CPE among the Erta-NS-IAI Enterobacteriaceae strains.

Patients and methods

Ethical considerations
In this survey, all Enterobacteriaceae isolates were collected as part of the routine hospital laboratory procedure and were not specially isolated for this study from the participating hospitals. This study was approved by the Institutional Review Boards and Ethical Committees of the participating centers, including the National Taiwan University Hospital, Taipei, Taiwan (NTUH 9561709108). The Ethical Committees waived the need for informed consent because limited private health information was collected and this research involved minimal risk to the subjects.

Study countries, isolates, and definitions of demographic variables
Thirty-seven medical centers from 12 countries/regions across the Asia-Pacific region participated in the Study for Monitoring Antimicrobial Resistance Trends (SMART) program during 2008–2014,14 which included Australia (n=5), the Hong Kong Special Administrative Region of China (n=2), Japan (n=3), Kazakhstan (n=1), Malaysia (n=2), New Zealand (n=1), Singapore (n=2), South Korea (n=2), Taiwan (n=8), Thailand (n=2), the Philippines (n=2), and Vietnam (n=4). Isolates of Enterobacteriaceae that were cultured from the first clinical samples of respective patients with IAI were obtained. Clinical samples from intra-abdominal sites related to IAI comprised tissue, fluid, or deep wound cultures obtained intraoperatively, or fluid obtained from percutaneous aspiration of abscesses. Duplicate isolates (the same species from the same patient within 30 days of the first positive culture) were excluded. Aside from the gender of IAI patients and the hospital settings where the IAI isolates were collected (intensive care units [ICUs] vs non-ICU, medical general ward vs surgical general ward), we also classified the ages of patients into three subgroups for analysis, including pediatric (age ≤14 years), adult (age between 15 and 64 years), and geriatric (age ≥65 years) subgroups.
Antimicrobial susceptibility testing and molecular detection of ESBLs, AmpC β-lactamases, and carbapenemases

Antibiotic susceptibility testing (MIC of the following antibiotics: cefoxitin, ceftriaxone, piperacillin/tazobactam, cefepmine, ertapenem, imipenem, amikacin, ciprofloxacin, and levofloxacin) determined by the broth microdilution method, quality control testing, and molecular analyses (by multiplex PCR) for all Enterobacteriaceae isolates were performed at the Central Laboratory (International Health Management Associates, Inc., Schaumburg, IL, USA). Antibiotic susceptibility results were interpreted according to the MIC break points recommended by the CLSI 2015. In this study, we excluded the IAI Enterobacteriaceae strains from the analysis if they did not harbor β-lactamase encoding genes based on the multiplex PCR results (see below).

Multiplex PCR was used to detect genes encoding ESBLs, AmpC β-lactamase, and carbapenemases in all isolates with ertapenem MIC >0.5 µg/mL. Whole genomic DNA of the isolates was extracted using the QIAamp DNA Mini kit and the QIAcube instrument (Qiagen NV, Venlo, the Netherlands) from the colonies grown overnight in blood agar plates (Thermo Fisher Scientific, Waltham, MA, USA). Specific primers for detection of the ESBL alleles (blaCTX-M, blaTEM, blaSHV, blaVER, blaOES, and blaPER), plasmid-borne AmpC genes (blaACC, blaCMY, blaMOX, blaFOX, blaDHA, blaACT, and blaMBR), and carbapenemase encoding genes (including blaSPM, blaIMI, blaKPC, blaVIM, blaNDM, blaIMP, and blaOXA-48-like) were used as previously described. If the isolates tested positive for the abovementioned allele(s) encoding carbapenemase, they were categorized as CPE.

Statistical analysis

Continuous variables were presented as mean ± SD or median with an IQR and were compared using Student’s t-test or Wilcoxon’s rank sum test for the two indicated groups depending on the normality of distribution. In addition, the Kruskal–Wallis test was employed for more than two subsets if the assumption of normality was invalid. By contrast, the categorical variables were expressed as absolute numbers and their respective percentages in a subgroup were analyzed and evaluated for differences by using Pearson chi-squared test or Fisher’s exact test as appropriate. To explore the independent predictors of CPE among the Erta-NS-IAI isolates, all variables with P<0.10 between different subgroups in the univariate analysis were included in a multivariate logistic regression model with a backward conditional method.

Overall goodness of fit was analyzed by the Hosmer and Lemeshow test with Nagelkerke’s R-square. Data with a P-value <0.05 were considered statistically significant. All the tests were two-tailed and performed analyses using the SPSS statistical software for Windows (version 17.0; SPSS Inc., Chicago, IL, USA).

Results

Distribution of IAI isolates, including those with an Erta-NS phenotype

In this Asia-Pacific Enterobacteriaceae survey from 2008 through 2014, a total of 1,564 isolates harboring alleles encoding β-lactamase(s) were collected from abdomen. There were 484 (30.9%) IAI Enterobacteriaceae isolates with in vitro non-susceptibility to ertapenem. These 484 IAI Enterobacteriaceae isolates accounted for ~2.6% of the overall 18,689 Asia-Pacific IAI Enterobacteriaceae isolates over the 7-year period. Of note, the isolates of Enterobacter species (n=223), followed by K. pneumoniae (n=140) and Escherichia coli (n=98) accounted for the majority (95.2%) of the Erta-NS-IAI isolates. The remainder were 23 Erta-NS isolates that comprised Citrobacter species (n=12), Serratia species (n=8), Klebsiella oxytoca (n=1), Cronobacter sakazakii (n=1), and Pantoea agglomerans (n=1). Among the Erta-NS-IAI isolates, 80 (16.5%) isolates were confirmed to harbor genes encoding carbapenemases, while the remaining 404 Erta-NS isolates did not. All IAI-CPE isolates harbored not more than one carbapenemase encoding gene.

High Erta-NS prevalence rates among the IAI isolates of K. pneumoniae as well as Enterobacter species (40.9% and 77.4%, respectively), and a notably higher CPE prevalence rate (35%) among the Erta-NS-K. pneumoniae isolates (the NDM producers predominantly, followed by the imipenemase [IMP] producers) were noted relative to the Erta-NS group of E. coli and Enterobacter species (P<0.001; Figure 1). In addition, the NDM producers were also the main CPE among the isolates of E. coli and Enterobacter spp.

Distribution of Erta-NS-IAI and CPE-IAI isolates collected by geographic regions, patients’ age, and hospital settings

Except for Japan and Kazakhstan with <10 Erta-NS-IAI isolates (Figure 2), the highest Erta-NS prevalence rates were observed in samples obtained from Australia, Taiwan, and South Korea, followed by the Philippines, New Zealand, the Hong Kong region, Singapore, and Vietnam during the 2008–2014 period. In stark contrast, samples from the
Philippines and Vietnam had the highest CPE prevalence rates (58.9% and 45.8%) among their Erta-NS-IAI isolates, respectively. Among the CPE isolates, IMP-26 producers (n=14) and NDM-1 producers (n=12) accounted for the majority of the CPE isolates in the Philippines, while NDM-1 producers (n=16) followed by OXA-48-like (n=7) dominated the CPE isolates collected from Vietnam.

When the IAI patients were stratified by age, a trend toward higher Erta-NS prevalence was observed in the IAI isolates from the geriatric (≥65 years old) subgroup (P<0.0001).
Nevertheless, the pediatric subgroup had the highest CPE prevalence among the Erta-NS isolates followed by the adult and geriatric subgroups (Figure 3). Additionally, significantly higher Erta-NS prevalence rates among the IAI isolates identified by culturing samples from patients hospitalized in the ICU (40.1%) or medical general ward (41.1%) were noted when compared with that of the non-ICU (29.7%) or surgical general ward (30.4%) settings, respectively (Figure 4).

Figure 3 The prevalence rates (%) of Erta-NS isolates among the isolates that caused IAI (n=1,564) and were positive for harboring gene(s) encoding β-lactamase(s) (black bars), and the prevalence rates of carbapenemase producing CPE among the Erta-NS isolates (gray bars) in the various age populations of patients hospitalized in the Asia-Pacific countries (or regions) participating in this IAI study, from 2008 through 2014.

Abbreviations: CPE, carbapenemase-producing Enterobacteriaceae; Erta-NS, ertapenem-non-susceptible; IAI, intra-abdominal infections.

Figure 4 The prevalence rates of Erta-NS isolates among those causing IAI (n=1,564) and those that were confirmed to possess gene(s) encoding β-lactamase(s) (black bars), and the percentages of carbapenemase producing CPE among the Erta-NS isolates (gray bars) in the ICU and various ward settings in the Asia-Pacific countries (or regions) participating in this IAI study, from 2008 through 2014.

Abbreviations: CPE, carbapenemase-producing Enterobacteriaceae; Erta-NS, ertapenem-non-susceptible; IAI, intra-abdominal infections; ICU, intensive care unit.
In vitro susceptibility and MIC distributions of the Erta-NS- and CPE-IAI isolates

The majority (>80%) of the Erta-NS-non-CPE isolates exhibited in vitro susceptibility to imipenem. Furthermore, a significantly lower rate of non-susceptibility to imipenem was observed in the non-CP subgroup than in the CP subgroup among the Erta-NS-IAI isolates of Enterobacter species (4.3% vs 61.5%; P<0.001). A similar scenario was also detected among the isolates of Erta-NS-E. coli as well as Erta-NS-K. pneumoniae. Geographically, among the Erta-NS-IAI isolates, 56 isolates collected from the Philippines as well as 59 isolates collected from Vietnam had the highest MICs (average in distributions) for imipenem (median [IQR] MIC value, 2 [1–8] and 1 [0.25–8] µg/mL; the MIC<sub>50</sub> and MIC<sub>90</sub> values of imipenem, 2 and 32 µg/mL, as well as 1 and 32 µg/mL, respectively) when the imipenem MIC data of all of the 10 countries were evaluated by the Kruskal–Wallis test. Nevertheless, the Erta-NS-IAI isolates from New Zealand (n=19), Taiwan (n=166), Australia (n=49), and Thailand (n=16) had relatively higher imipenem MIC ranges (overall median [IQR] MIC value, 0.5 [0.5–1.0] µg/mL, with a MIC<sub>50</sub> and MIC<sub>90</sub> of 0.5 and 4.0 µg/mL for imipenem in 250 IAI isolates) than did the following four (Hong Kong region, South Korea, Malaysia, and Singapore) as seen from the nonparametric analysis.

As for the CPE isolates, 95% harbored the ESBL encoding allele(s). The imipenem MICs (average in the distributions) among those harboring NDM-, KPC-, or OXA-48-like encoding genes were notably higher than those of the IMP producing CPE according to the Wilcoxon’s rank sum test (Table 1).

Univariate and multivariate logistic analyses of CPE in the Erta-NS-IAI isolates

The differences among the various demographic factors, hospital settings of collection, intra-abdominal cultured sites, types of leading species, and the non-susceptibility percentages against the evaluated antimicrobial agents in the CP and non-CP subgroups of Erta-NS-IAI Enterobacteriaceae isolates, are illustrated in Table 2. As all CPE isolates had cefepime MICs >8 µg/mL, we chose the cefepime MIC of >8 µg/mL as the threshold for the univariate analysis of CPE. According to the univariate analysis, isolates of K. pneumoniae, IAI isolates from the peritoneal cavity (identified by culturing), patients aged >15 years old, IAI isolates acquired from the overall ICU, surgical ICU alone, or the medical general ward settings, and those showing non-susceptibility to the antibiotics except cefoxitin, the molecular confirmation for ESBL positive and plasmidic AmpC negative were likely to be CPE, whereas the reverse was true for the Erta-NS isolates of Enterobacter spp., or isolates identified by culturing samples from the hepatobiliary system. Furthermore, the multivariate analysis for Erta-NS-CPE revealed that the Erta-NS strains with imipenem non-susceptibility (OR, 56.4), with cefepime MIC >8 µg/mL (OR, 4.4), identified by culturing of samples from the peritoneal space (tissue or abscess; OR, 3.3), or carrying the ESBL encoding gene(s) according to PCR test (OR, 11.5) were found to be independent factors predicting CPE in the Erta-NS-IAI isolates, whereas the Erta-NS-IAI isolates that lacked the gene(s) encoding plasmid-borne AmpC enzyme(s) were an independent predictor of non-CPE (OR, 0.26; Table 3). This model has an acceptable goodness of fit (P=0.363) with a pseudo-R-square (Nagelkerke) of 0.731.

Discussion

Until recently, only few studies have specifically addressed IAI related to carbapenem-NS Enterobacteriaceae species in the PubMed literature. In addition, with a significant difference to previous surveys pertaining to carbapenem-NS Enterobacteriaceae isolates related to KPC production, the Erta-NS-CPE isolates in this Asia-Pacific IAI investigation mostly harbored the bla<sub>NDM</sub> or bla<sub>IMP</sub> alleles. Among the

| Carbenpenemases (isolate number) | Imipenem MIC range (µg/mL) | Imipenem MIC<sub>50</sub> value (µg/mL) | Imipenem MIC<sub>90</sub> value (µg/mL) |
|--------------------------------|-----------------------------|----------------------------------------|----------------------------------------|
| NDM (37)                      | 0.5–32                      | 8                                      | 16                                     |
| KPC (7)                       | 2–32                        | 16                                     | 32                                     |
| OXA-48-like (8)               | 1–16                        | 4                                      | 16                                     |
| IMP (28)                      | 0.5–16                      | 2                                      | 8                                      |

Abbreviations: IMP, imipenemases; KPC, Klebsiella pneumoniae carbapenemases; MIC, minimum inhibitory concentration; NDM, New Delhi metallo-β-lactamases; OXA, oxacillinases.
Table 2 Comparison of the variables of demographics of Asia-Pacific patients with IAI, the hospital sampling settings from where the Erta-NS IAI Enterobacteriaceae isolates were collected, types of species (comprising Escherichia coli, Klebsiella pneumoniae, and overall Enterobacter species), detection of the Erta-NS-IAI isolates by culturing samples from the abdominal space, the non-susceptibility rates toward different antimicrobial agents for three leading Erta-NS-IAI Enterobacteriaceae species between the subgroups of cPe and non-cPe isolates collected from 2008 through 2014

| Variables | CPE (n=80), | Non-CPE (n=404), | OR | 95% CI | P-value |
|-----------|-------------|------------------|-----|--------|---------|
|           | yes or no   | yes or no        |     |        |         |
| Gender, male | Yes | 45 (56.3) | 240 (59.4) | 0.879 | 0.541, 1.426 | 0.620 |
|           | No | 35 (43.8) | 164 (40.6) |     |        |         |
| Age (years) | ≤14 years (pediatric) | Yes | 4 (5.0) | 9 (2.2) | 2.244 | 0.708, 7.110 | 0.244 |
|           | No | 76 (95.0) | 395 (97.8) |     |        |         |
|           | 15–64 years (adult) | Yes | 47 (58.8) | 162 (40.1) | 1.465 | 1.177, 1.824 | 0.003 |
|           | No | 33 (41.3) | 242 (59.9) |     |        |         |
|           | ≥65 years (geriatric) | Yes | 29 (36.3) | 233 (57.7) | 0.417 | 0.254, 0.686 | 0.001 |
| Sites of acquisition of isolates | ICU setting acquisition | Yes | 18 (22.5) | 59 (14.6) | 1.698 | 0.938, 3.072 | 0.08 |
|           | No | 62 (77.5) | 345 (85.4) |     |        |         |
|           | Surgical ICU patients | Yes | 14 (17.5) | 35 (8.7) | 2.236 | 1.141, 4.383 | 0.024 |
|           | No | 66 (82.5) | 369 (91.3) |     |        |         |
|           | Medical ICU patients | Yes | 3 (3.8) | 22 (5.4) | 0.677 | 0.198, 2.316 | 0.782 |
|           | No | 77 (96.3) | 382 (94.6) |     |        |         |
|           | Surgical ward patients | Yes | 34 (42.5) | 140 (34.7) | 1.116 | 0.686, 1.814 | 0.659 |
|           | No | 46 (57.5) | 264 (65.3) |     |        |         |
|           | Medical ward patients | Yes | 22 (27.5) | 157 (38.9) | 0.597 | 0.351, 1.014 | 0.058 |
|           | No | 58 (72.5) | 247 (61.1) |     |        |         |
| Bacterial species | Escherichia coli | Yes | 15 (18.8) | 83 (20.5) | 0.892 | 0.484, 1.644 | 0.763 |
|           | No | 65 (81.3) | 321 (79.5) |     |        |         |
|           | Klebsiella pneumoniae | Yes | 49 (61.3) | 91 (22.5) | 5.437 | 3.275, 9.024 | <0.001 |
|           | No | 31 (38.8) | 313 (77.5) |     |        |         |
|           | Enterobacter spp. | Yes | 13 (16.3) | 210 (52.0) | 0.179 | 0.096, 0.335 | <0.001 |
|           | No | 67 (83.8) | 194 (48.0) |     |        |         |
| Source of isolates | Peritoneal space | Yes | 54 (67.5) | 145 (35.9) | 3.710 | 2.227, 6.179 | <0.001 |
|           | No | 26 (32.5) | 259 (64.1) |     |        |         |
|           | Hepatobiliary system | Yes | 14 (17.5) | 193 (47.8) | 0.232 | 0.126, 0.426 | <0.001 |
|           | No | 66 (82.5) | 211 (52.2) |     |        |         |
|           | LGI tract | Yes | 3 (3.8) | 16 (4.0) | 0.945 | 0.269, 3.321 | 1.000 |
|           | No | 77 (96.3) | 388 (96.0) |     |        |         |
| Phenotype of isolates (μg/mL) | Imipenem NS (MIC >1) | Yes | 74 (92.5) | 62 (15.3) | 68.032 | 28.365, 163.175 | <0.001 |
|           | No | 6 (7.5) | 342 (84.7) |     |        |         |
|           | Amikacin NS (MIC >16) | Yes | 51 (63.8) | 91 (22.5) | 6.049 | 3.625, 10.095 | <0.001 |
|           | No | 29 (36.3) | 313 (77.5) |     |        |         |
|           | Cefepime MIC >8 | Yes | 77 (96.3) | 150 (37.1) | 43.462 | 13.477, 140.167 | <0.001 |
|           | No | 3 (3.8) | 254 (62.9) |     |        |         |
|           | Cefepime MIC >16 | Yes | 69 (86.3) | 107 (26.5) | 17.411 | 8.877, 34.148 | 0.003 |
|           | No | 11 (13.8) | 297 (73.5) |     |        |         |
|           | Piperacillin–tazobactam NS | Yes | 74 (92.5) | 327 (80.9) | 2.904 | 1.219, 6.920 | 0.010 |
|           | No | 6 (7.5) | 77 (19.1) |     |        |         |
|           | Ciprofloxacin NS (MIC >1) | Yes | 67 (83.8) | 193 (47.8) | 5.635 | 3.015, 10.529 | <0.001 |
|           | No | 13 (16.3) | 211 (52.2) |     |        |         |
|           | Levofloxacin NS (MIC >2) | Yes | 55 (68.8) | 155 (38.4) | 3.534 | 2.115, 5.906 | <0.001 |
|           | No | 25 (31.3) | 249 (61.6) |     |        |         |

(Continued)
Table 2 (Continued)

| Variables                        | Yes or no | CPE (n=80), no (%) | Non-CPE (n=404), no (%) | OR     | 95% CI   | P-value |
|----------------------------------|-----------|--------------------|-------------------------|--------|----------|---------|
| Ceftriaxone NS (MIC >1)          | Yes       | 80 (100)           | 388 (96.0)              | 1.041  | 1.021, 1.062 | 0.007   |
|                                  | No        | 0 (0)              | 16 (4.0)                | 0.889  | 0.188, 4.192 | 1.000   |
| Cefoxitin NS (MIC >8)            | Yes       | 78 (97.5)          | 395 (97.8)              | 0.889  | 0.188, 4.192 | <0.001  |
|                                  | No        | 2 (2.5)            | 9 (2.2)                 | 24.614 | 8.836, 68.566 | <0.001  |
| ESBL positivity                  | Yes       | 76 (95)            | 176 (43.6)              | 0.222  | 0.130, 0.377 | <0.001  |
|                                  | No        | 4 (5)              | 228 (56.4)              |        |           |         |
| AmpC positivity                  | Yes       | 22 (27.5)          | 255 (63.1)              |        |           |         |
|                                  | No        | 58 (72.5)          | 149 (36.9)              |        |           |         |

Abbreviations: LGI, lower gastrointestinal tract; NS, non-susceptibility; CPE, carbapenemase-producing Enterobacteriaceae; IAI, intra-abdominal infections; Erta-NS, ertapenem-non-susceptible; ICU, intensive care unit; ESBL, extended-spectrum β-lactamase; MIC, minimum inhibitory concentration.

Table 3 Independent predictors of carbapenemase producers among the ertapenem-non-susceptible Enterobacteriaceae isolates causing intra-abdominal infections in patients hospitalized in the Asia-Pacific region from 2008 through 2014

| Variables                        | OR     | 95% CI      | P-value |
|----------------------------------|--------|-------------|---------|
| Imipenem non-susceptibility      | 56.4   | 19.8, 161.1 | <0.0001 |
| Cefepime MIC >8 µg/mL            | 4.4    | 1.0, 19.2   | 0.047   |
| Cultured from the peritoneal space (tissue, abscess) | 3.3 | 1.4, 7.9 | 0.008  |
| Positivity of the ESBL encoding allele(s) by multiplex PCR test | 11.5 | 2.8, 47.1 | 0.001   |
| Positivity of the AmpC encoding allele(s) by multiplex PCR test | 0.26 | 0.10, 0.66 | 0.004   |

Abbreviations: ESBL, extended-spectrum β-lactamase; MIC, minimum inhibitory concentration.

NDM-1 producers of IAI-CPE in this survey, 38.7% and 51.6% were collected from the Philippines and Vietnam, respectively. Therefore, the Philippines and Vietnam will likely face a huge therapeutic challenge in treating substantial CPE burden among clinical IAI isolates in the future. Of the five independent CPE predictors for the Erta-NS-IAI Enterobacteriaceae isolates (not including samples collected in ICU settings), imipenem non-susceptibility, cefepime MIC >8 µg/mL, and Erta-NS-IAI strains detected by culturing samples from the peritoneal space could be conveniently applied to identify isolates likely to be CPE in the Asia-Pacific region.

In this Asia-Pacific Enterobacteriaceae IAI study, a significantly higher CPE percentage (35%) was observed in the Erta-NS-K. pneumoniae isolates than the other two species (E. coli [15.3%] and Enterobacter species [5.8%]; P-values, 0.0007 and <0.0001, respectively; Figure 1). The diverse carbapenemases on CP K. pneumoniae isolates were dominated by NDM (42.9%; especially NDM-1), followed by IMP (38.8%; especially IMP-26) and OXA-48-like (8.2%). Spread of the mobile NDM-1 encoding genes which are primarily located on an easily transferable, 180-kb plasmid (with high horizontal transfer potential) in K. pneumoniae is facilitated by population mobility.21 In addition, in a CRE study reported by Gupta et al26 issuing the clinical US cases with IMP or NDM producing Enterobacteriaceae infections (reported to the Centers for Disease Control and Prevention, USA), K. pneumoniae isolates accounted for the majority (70%) of implicated Enterobacteriaceae strains. Other surveys also showed that clinical isolates of K. pneumoniae predominated the NDM producing Enterobacteriaceae strains.27-28 The findings of the above studies suggest that clinical isolates of K. pneumoniae indeed have a higher likelihood of harboring plasmid-mediated carbapenemases than the other Enterobacteriaceae spp. These results corresponded with our Erta-NS-IAI Enterobacteriaceae study as well. Nevertheless, isolates of carbapenem-resistant K. pneumoniae were surprisingly found not to be an independent predictor of CPE in the multivariate logistic regression analysis in this IAI study. The reason why IAI-Erta-NS-K. pneumoniae was not an independent CPE predictor might be related to the possibility that the CPE proportion among Erta-NS-K. pneumoniae isolates was not high enough.

Prior research into CPE infections has indicated that most (>75%) of these patients are overexposed to some antimicrobials.5,9 We did not investigate this aspect in our IAI study. Of note, this Asia-Pacific IAI Enterobacteriaceae study indicates that the Erta-NS strains isolated from the peritoneal cavity were independently predicted to be CPE. In addition, the molecular confirmation of ESBL encoding genetic determinants is another predictor of Erta-NS-CPE among the IAI-cultured isolates. With respect to these associations, we consider that extended-spectrum cephalosporin in conjunction with metronidazole is often prescribed as the
first-line treatment for most mild-to-moderate IAI episodes during the initial therapy. Usually a prolonged duration of treatment with antimicrobials targeting the bacteria in the peritoneal space is needed, especially for patients who are too vulnerable to undergo surgery. For these debilitated patients, such a therapeutic maneuver is highly likely to predispose them to acquiring clinical CPE infections in nosocomial settings. In addition, Ben-David et al suggested that such a situation may simultaneously cause the originally susceptible Enterobacteriaceae isolates to acquire plasmidic ESBL encoding gene(s) from other bacteria residing in the same microbial environment. As for the higher Erta-NS prevalence rates among the IAI Enterobacteriaceae isolates for two age groups, whether there were significant differences in the past antimicrobial exposure between them also needs to be clarified.

In this Asia-Pacific IAI study, the majority of Erta-NS-non-CP Enterobacteriaceae isolates virtually displayed in vitro non-susceptibility to imipenem. A UK study conducted by Doumith et al investigated 16 Erta-NS-non-CPE isolates (including 10 K. pneumoniae and six Enterobacter species) that were mostly non-susceptible to imipenem (MICs range, 2–8 µg/mL among 11 strains). Apart from ESBL and/or AmpC production, they found that most of them lacked or had a significantly poor expression of major porin(s) encoding genes as well (OmpK35 and OmpK36 for K. pneumoniae; OmpC and OmpF for Enterobacter species). The isolates of Enterobacter species are known to be constitutive hyper-producers of inducible AmpC β-lactamase(s). Furthermore, in this Asia-Pacific IAI survey, ~70% of the 223 isolates of Erta-NS Enterobacter species (more than three-fourths of which were Enterobacter cloacae; of note, the overall imipenem non-susceptibility rate was only 7.6%) were proven to be plasmidic AmpC producers, whereas only 16% and 5.8% of them were ESBL producers and CPE, respectively. In addition, among the Erta-NS-non-CP K. pneumoniae isolates in our IAI survey, the prevalence rate of ESBL plus concomitantly plasmidic AmpC was 44.0% (not shown in the “Results” section), disproportionally higher than the imipenem non-susceptibility rate (7.7%; P<0.01). These results imply that most Erta-NS-non-CPE IAI isolates in the Asia-Pacific region likely have intact outer-membrane porins. The above mechanisms might plausibly explain why imipenem non-susceptibility (MIC >2 µg/mL) is identified as an independent predictor of CPE among these Asia-Pacific Erta-NS-IAI Enterobacteriaceae isolates.

There are some limitations to this study. Firstly, although we designed the study to collect clinical IAI isolates from many Asia-Pacific countries for 7 years, the total number of Erta-NS-CPE isolates is relatively small (n=80). Secondly, with respect to the detection of accurate variants among the diverse blaOXA-48-like alleles, the PCR test failed to differentiate them among the Enterobacteriaceae isolates. The OXA-163 hydrolyzes cephalosporins as well as aztreonam more effectively than carbapenems, while the OXA-232 enzyme also hydrolyzes the carbapenem agents less efficiently than the other OXA-48-like carbapenemases. Thirdly, the genetic determinants encoding carbapenemases among the Erta-NS-CPE isolates of this Asia-Pacific IAI study are much different from those in most Western countries where KPC producers are endemic. Consequently, the conclusions of our IAI survey cannot be generalized to other geographic regions. Fourthly, the genetic relatedness and clones of special sequence types (eg, ST131 E. coli and ST11 K. pneumoniae) of the enrolled IAI isolates in this study were not determined. Lastly, we did not analyze the major porin(s) encoding genes and emergence of efflux pump(s) in the Erta-NS-IAI Enterobacteriaceae isolates investigated in this study.

**Conclusion**

Many countries in the Asia-Pacific region have high (>20%) Erta-NS prevalence rates among the IAI isolates. The isolates of Enterobacter species followed by K. pneumoniae have higher prevalence rates of the Erta-NS phenotype (77.4%, 40.9%) than E. coli. Of note, the Erta-NS-IAI K. pneumoniae isolates have an alarmingly high prevalence rate (35%) of CPE. The Erta-NS Enterobacteriaceae isolates detected by culturing samples from the peritoneal space (fluid, tissue), with in vitro cefepime MIC >8 µg/mL, with imipenem non-susceptibility, or carrying the ESBL encoding gene(s) proven by the PCR test, are independent predictors of CPE.

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