Differential expression of microRNA-411 and 376c is associated with hypertension in pregnancy

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Abstract

Preeclampsia is a major reason of morbidity and mortality in pregnant women and perinatal fetus. Hence, it is of prime importance that diagnostic markers are defined to predict chances of preeclampsia in pregnant women. It has been previously shown that microRNA (miRNA)-376c expression is decreased in the placenta of preeclampsia patients at term. Even though this decrease was not mimicked in the placenta at the pre-term stage, miR-376c expression was decreased in the plasma of these patients as early as the second trimester. Plasma and placenta specimens were obtained from pregnant women having unifetal gestation undergoing perinatal care between January 2014 and December 2016 (n=49). Early trimester placentas were collected from patients undergoing terminated pregnancies through dilation and curettage procedure. Our results showed that in addition to miR-376c, miR-441 levels were decreased in the placenta of preeclampsia patients, and this decrease occurred both at pre-term and at term. This decrease is also mimicked in the plasma levels at both early and late weeks of pregnancy, highlighting that miR-441 levels can serve as a diagnostic marker of risk of preeclampsia in pregnant women. Overexpression of the miR-441, as well as miR-376c, promoted cell viability, migration, and invasion in the human immortalized cytotrophoblast cell line HTR8/ SVneo, indicating that their decrease in pregnant women would result in anomalous apoptosis and functional imbalance resulting in premature abortion and other complications. MiR-441 level can thus potentially serve as diagnostic marker of preeclampsia in pregnant women.

Key words: Preeclampsia; Pregnancy; Hypertension; miR-411; miR-376c

Introduction

Pregnancy-associated hypertension is a major cause of perinatal and maternal mortality (1,2). Hypertension can be transient or occur within 20 weeks of gestation and is associated with proteinuria. This later form used to be called preeclampsia (3). However, according to the current guidelines of The American College of Obstetricians and Gynecologists, preeclampsia is now diagnosed by persistent high blood pressure that develops during pregnancy or during the postpartum period that is associated with proteinuria or the new development of decreased blood platelets, problems with the kidney or liver, fluid in the lungs, or signs of brain problems such as seizures and/or visual disturbances (4). Even though anomalous placentation is widely believed to be the reason for preeclampsia, the exact etiology is unknown (3–5).

Cardinal features of preeclampsia include, but are not limited to, decreased trophoblast proliferation and increased apoptosis, impeded differentiation of trophoblast, and impaired migration and invasiveness of the trophoblast in the uterus, resulting in defective spiral artery remodeling (6–10). However, the precise mechanism initiating or supporting preeclampsia has not been elucidated.

Micro-RNAs (miRNA) are evolutionarily conserved 21–23 nucleotides RNAs that regulate post-transcriptional gene expression either by blocking translation or degrading target messenger RNAs (mRNAs) and have been increasingly shown to function as inhibitor or promoter of many diseases (11,12). miRNAs can function in both normal and transformed cells and have even been shown to play a role in cardiovascular diseases, including hypertension (13–18).

Importantly, anomalous miRNA expression in placenta has been shown to be involved in pregnancy problems, including preeclampsia (19,20). Placental miRNAs can also be detected in the plasma, providing a great opportunity of studying their changing expression level and determining if they can be used for prognostic purposes in predicting complications during pregnancy (21). Indeed, it has been
shown that miR-376c levels decrease in the placenta of preeclampsia patients at term and this decrease can be observed in the plasma as early as the second trimester (22). However, it is not known why the decrease in the plasma levels is not mimicked in the placenta of these patients. Hence, it is important to discover additional miRNAs, which are decreased in preeclampsia patients’ plasma and placenta at the pre-term stage.

Material and Methods

Patients, clinical samples, and ethical considerations

The Institutional Review Board of the Jinan Central Hospital Affiliated to Shandong University approved the protocol of the study (#2016185). Plasma and placenta specimens were obtained from pregnant women having unifetal gestation undergoing perinatal care at the Department of Gynecology and Obstetrics, Jinan Central Hospital Affiliated to Shandong University, between January 2014 and December 2016 (n=49). All patients provided written consent. All experiments conformed to the principles set out in the World Medical Association Declaration of Helsinki and the NIH Belmont Report. Patients with gestational diabetes, transient hypertension, spontaneous abortion, congenital abnormalities in the fetus, and intrauterine fetal death were excluded from the current study (n=18). Early trimester placentas were collected from patients undergoing terminated pregnancies through dilation and curettage procedure (n=9). Heparin containing DNase/RNase-free tubes were used for blood sample collection. Cell culture and transfection

The immortalized cell line from first trimester human cytotrophoblast cells HTR-8/SVneo was obtained from ATCC and cultured in RPMI-1640 supplemented with 5% FBS and penicillin/streptomycin (ThermoFisher Scientific, USA). Where indicated, cells were mock-transfected or transfected with miR-376c and miR-411 mimic (ThermoFisher Scientific) using lipofectamine 3000 (ThermoFisher Scientific) following the manufacturer’s recommended protocol.

miRNA extraction and quantitative real time polymerase chain reaction (qRT-PCR)

miRNA was extracted by the TRiZol LS isolation kit (ThermoFisher Scientific) according to the manufacturer’s instructions. The expression levels of miR-376c, miR-377, miR-411, and RNU6B were detected by TaqMan miRNA assays (ThermoFisher Scientific). Data were analyzed by the -ΔΔCt method and normalized to RNU6B expression.

Cell proliferation assay

The MTT assay kit (Sigma-Aldrich, China) was used to quantitatively determine cell proliferation. Results of relative absorbance are reported as means ± SD of three biological replicates, each performed with three technical replicates.

Cell migration and cell invasion assays

The Culturex 96-well cell migration and Culturex 96-well BME cell invasion assay kits (R&D Systems, USA) were used to detect the in vitro ability of cell migration and invasion, according to the manufacturer’s recommendation. The results were used to analyze the percent of cell migration and invasion and data are reported as means ± SD.

Statistical analyses

Statistical analyses were performed using SPSS statistics software 20.0 (IBM, USA). Differences between groups were determined by one-way ANOVA. Student’s t-test or Mann-Whitney U-test (non-normal data distribution) was used for comparison between two groups. Adjustment for both gestational and maternal age was done by performing multiple regression analysis. P<0.05 was considered statistically significant.

Results

The included cases (preeclamptic, n=31 and control, n=9) (Figure 1A and B) were not significantly different in age (P>0.05). However, there were significantly more cesarean sections than normal vaginal deliveries in patients with preeclampsia compared to controls (P<0.05). Additionally, patients with preeclampsia had higher blood pressure compared to controls and more patients were on antihypertensive medication (methyldopa or labetalol) (P<0.05 in each case) (Table 1).

miRNA-376c, -377, and -411 levels were determined in normal and preeclamptic pregnancies by qRT-PCR. No significant change was noted for miR-377 expression between the first (1 ± 0.34), second (0.81 ± 0.62), and third trimesters (1.09 ± 0.37) (P=0.735) in normal pregnancy. However, both miR-376c (1 ± 0.29; 4.82 ± 0.43; and 6.21 ± 0.41, respectively, in first, second, and third trimesters) and miR-411 (1 ± 0.20; 5.11 ± 0.11; and 7.94 ± 0.36, respectively, in first, second, and third trimesters) significantly increased in the second compared to the first trimester, and third compared to the first or second trimester (Figure 1C; P<0.05 in each case) in normal pregnancy. This indicated that miR-411 and miR-376c expressions were induced during the progression of normal pregnancy.
We next determined miR-376c, miR-377, and miR-411 expression in preeclamptic pregnancy compared to age-matched controls, both pre-term (26–35 weeks) and at term (36–40 weeks). In at-term placentas, both miR-376c and miR-411 levels were significantly lower in patients with severe preeclampsia (miR-376c: 2.38 ± 0.34 and miR-411: 2.13 ± 0.02) compared to control patients (miR-376c: 5.39 ± 0.44 and miR-411: 6.34 ± 0.46) (Figure 2; P < 0.05 in each case). There was no significant difference in miR-376c expression between severe preeclampsia (1.03 ± 0.04) and normal pre-term patients (1.01 ± 0.02) (Figure 2; P > 0.05). However, miR-411 expression was significantly lower in pre-term placenta obtained from patients with severe preeclampsia (0.23 ± 0.05) compared with normal subjects (1.04 ± 0.06) (Figure 2; P < 0.05), indicating that miR-411 expression was regulated at an earlier time point than miR-376c.

We next determined miR-376c, miR-377, and miR-411 expression levels in plasma of patients who later developed preeclampsia compared to control patients. In contrast to the observation in pre-term placentas, miR-376c was significantly lower in plasma (0.19 ± 0.04) of patients who later developed preeclampsia compared to control (1.01 ± 0.03) patients (Figure 3; P < 0.05). A similar pattern of decreased expression was observed for miR-411 in patients who later developed preeclampsia (0.15 ± 0.06).

Table 1. Clinicopathological characteristics of the patients included in the current study.

| Characteristics         | Control (n=9) | Preeclampsia (n=31) | P     |
|-------------------------|--------------|---------------------|-------|
| Age (years)             | 32.83 ± 3.19 | 31.57 ± 2.98        | 0.45  |
| BMI (kg/m²)             | 20.97 ± 2.12 | 23.97 ± 3.12        | 0.039 |
| Parity (n)              | 2 ± 1        | 2 ± 1               | 1.00  |
| Type of deliveries      |              |                     |       |
| NVD                     | 8 (88.9%)    | 22 (70.96%)         | 0.001 |
| CS                      | 1 (11.1%)    | 9 (29.04%)          | 0.001 |
| Gestational age (weeks) | 29.66 ± 2.38 | 29.13 ± 1.37        | 0.23  |
| Medications             |              |                     |       |
| Methyldopa              | 0 (0%)       | 30 (96.77%)         | NA    |
| Labetalol               | 0 (0%)       | 1 (3.23%)           | NA    |
| Tonics                  | 9 (100%)     | 31 (100%)           | 1.00  |
| Blood Pressure          |              |                     |       |
| SBP (mmHg)              | 102.12 ± 12.19 | 143.57 ± 7.21     | 0.001 |
| DBP (mmHg)              | 76.83 ± 2.39 | 95.42 ± 1.34        | 0.001 |
| Mean BP (mmHg)          | 86.23 ± 4.39 | 111.29 ± 8.43       | 0.001 |
| Pulse BP (mmHg)         | 26.21 ± 3.22 | 51.49 ± 5.28        | 0.001 |

Results are reported as means ± SD or number and percentage. BMI: body mass index; NVD: normal vaginal delivery; CS: caesarian section; SBP: systolic blood pressure; DBP: diastolic blood pressure; BP: blood pressure; SD: standard deviation; NA: not applicable. Data were analyzed with Student’s t-test or chi-squared test.
compared to controls (1.04 ± 0.04) (Figure 3; P < 0.05). Both miR-411 (6.34 ± 0.46 in controls vs 1.13 ± 0.02 in preeclampsia) and miR-376c (5.39 ± 0.44 in controls vs 1.24 ± 0.35 in preeclampsia), but not miR-377 (1.04 ± 0.02 in controls vs 1.01 ± 0.06 in preeclampsia), were significantly less expressed in plasma of preeclampsia patients at 36 to 40 weeks of gestation (Figure 3; P < 0.05 in each case).

It has been previously shown that miR-376c overexpression in the human trophoblast cell line, HTR8/SVneo, induces cell proliferation, migration, and invasion (22). Hence, we finally determined if transient expression
of a miR-376c or miR-441 mimic in the HTR8/SVneo cells would induce increased proliferation, migration, and invasion. Ectopic expression of both miR-441 and miR-376c significantly increased cell proliferation by 2.01 ± 0.34 and 1.91 ± 0.08, respectively, as assessed by the MTT assay (Figure 4A; P < 0.05), migration (miR-376c by 24 ± 2%; and miR-411 by 25 ± 3%) (Figure 4B; P < 0.05), and invasion (miR-376c by 37 ± 2%; and miR-411 by 39 ± 3%) (Figure 4C; P < 0.05).

Discussion

A significant reduction of placental and plasma miR-411 was observed in preeclamptic patients, compared with their gestational age-matched normal subjects. Our results also demonstrated that miR-411, like miR-376c (shown before (22) and here), can promote trophoblast cell proliferation, migration, and invasion. In comparison to miR-376c, miR-411 levels decreased even in pre-term placenta indicating that an miR-411 decrease was an earlier event compared to miR-376c downregulation.

miR-411 belongs to the miR-379 family and is located in the imprinted DLK1/DIO3 region within the miR-379/miR-656 cluster on human chromosome 14 (23), which is highly conserved in placental mammals (22,23). miR-376c is also a member of the miR-379/miR-656 cluster (22). In fact, expression of miR-154 and miR-377, both members of the miR-379/miR-656 cluster, has been previously shown to be decreased in preeclamptic placentas (24). However, in the current study, we did not find any difference in expression in the placenta or plasma of patients with preeclampsia. On the other hand, our results are in line with previous findings on miR-376c in preeclamptic placenta and plasma, both pre-term and at term (22).

Given our and previous (22) findings that miR-411 and miR-376c expressions were associated with cell proliferation, migration, and invasion and their continued increase during normal pregnancy, it can be inferred that these miRNAs are associated with normal placental development, and hence their decrease results in anomalous placental development and ultimately premature abortion. How hypertension causes this decrease in miR-411 and miR-376c needs to be determined. Given that all findings including ours suggest that miRNAs associated with the miR-379/miR-656 clusters are downregulated in preeclampsia patients, it needs to be determined which miRNAs in this cluster are expressed less and if there is a pattern to their decrease, as some are decreased early whereas others, late during pregnancy. Furthermore, how their transcription affects hypertension and how this leads to proteinuria need to be identified as they have the potential of providing novel therapeutic targets for treating preeclamptic patients.

Given that miR-441 and miR-376c expression were down-regulated in the plasma of these patients by the 15th week of pregnancy, determining their levels in incoming pregnant patients can serve as indicators of potential risk of preeclampsia. However, such observations should be validated in a larger cohort of patients in a multi-center study design.
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