Detection and phylogeny of Staphylococcus aureus sequence type 398 in Taiwan

Yhu-Chering Huang¹,²,³* and Chih-Jung Chen¹,²

Abstract

Background: Methicillin-resistant Staphylococcus aureus (MRSA) ST398 is a livestock associated-bacterium that is most prevalent in Europe. Human-adapted MRSA ST398 was recently reported from China, but there is no data available yet for Taiwan.

Methods: To identify S. aureus ST398 isolates, we examined 6413 S. aureus isolates (5632 MRSA and 781 susceptible strains) that were collected in Taiwan between 1995 and 2017. If isolates could not be typed by pulsed-field gel electrophoresis upon Sma I digestion, we performed further characterization and complete genome sequencing.

Results: We identified 18 ST398 S. aureus isolates from 16 subjects (0.28%), including 6 sensitive and 12 resistant strains. Of these, 14 were colonizing isolates, 3 were clinical (infecting) isolates and one isolate was from a pork specimen. All 3 infecting isolates were MSSA strains identified in 2015 from two children with recurrent otitis media or sinusitis. The other 3 MSSA isolates were identified from workers handling pork (2) or pork meat (1) in 2015. The first 5 MRSA colonizing isolates were identified from residents in two nursing homes in 2012. Six MRSA isolates were identified from residents and foreign employees at a nursing home in 2016 and one MRSA from a foreign worker in 2017. Phylogenetic analysis of genome sequences indicated that all 12 local ST398 MRSA strains cluster together, human-adapted and phylogenetically related to a human MRSA strain identified in China in 2002. Two local MSSA isolates could be linked to isolates from livestock. The toxin profiles were similar for the MRSA and MSSA isolates.

Conclusions: Our results demonstrate that S. aureus ST398 was present in Taiwan in 2012 and potentially earlier. Although some isolates could be linked to livestock, most ST398 S. aureus isolates identified in Taiwan, particularly MRSA, represent human-adapted strains. Local transmission of human-adapted MRSA ST398 strains has occurred in nursing homes in Taiwan, possibly after import from China. Further surveillance is needed.

Keywords: Methicillin-resistant Staphylococcus aureus, Sequence type 398, Pulsotype, Taiwan

Introduction

In addition to being a human pathogen, methicillin-resistant Staphylococcus aureus (MRSA) can also colonize and cause diseases in a variety of animals, which as known as livestock-associated MRSA (LA-MRSA) [1, 2]. In the past decade, LA-MRSA has been increasingly identified in humans and has been associated with very severe diseases and even death [1, 2]. MRSA sequence type (ST) 398 was the first identified LA-MRSA [1–4], initially identified in Europe and subsequently identified in areas outside Europe, including Asian countries, such as South Korea [5, 6], Singapore [7], Japan [8–10] and Thailand [11]. Lately, human-adapted ST398 community-associated (CA) MRSA, evolving from methicillin-susceptible S. aureus (MSSA) demonstrated by whole genome sequencing analysis, was reported from China [12–14] and caused severe human disease and even death. Human-adapted MRSA is genetically different from LA-MRSA [12, 13] and is associated with absence of the tetM resistance determinant and presence of a variant of prophage 3, containing the immune evasion complex (IEC) genes encoding the chemotaxis inhibitory protein (chp), staphylococcal complement inhibitor (scn), and staphylokinase (sak).
In Taiwan, MRSA was first documented in early 1980s and rapidly increased in 1990s [15]. In contrast to a declining trend of HA-MRSA incidence observed in 2000s, CA-MRSA infections have been increasingly reported, particularly in pediatric patients [15–17]. LA-MRSA was also identified from pigs and humans recently and caused minor to life-threatening human infections [18]. Most LA-MRSA in Taiwan belonged to ST9 while ST398 was not specifically reported yet [15–18]. Recently, we identified five MRSA ST398 isolates in a survey for MRSA carriage in nursing homes in 2012 [19], prompting our study on this strain in Taiwan to figure out the perspective of MRSA ST398 in Taiwan. Herein, we report the detection, and phylogenetic analysis of ST 398 S. aureus isolates identified in our laboratory in Taiwan as well as the epidemiologic information of these isolates.

Methods

For surveillance of molecular epidemiology of S. aureus, we collected and molecularly characterized (at least by pulsed-field gel electrophoresis (PFGE)) a total of 6413 S. aureus isolates (MRSA, 5632 and MSSA, 781) island-wide in Taiwan between 1995 and 2017 and molecularly characterized in Chang Gung Memorial Hospital (CGMH). Identification of Staphylococcus aureus was confirmed by the conventional methods according to Clinical and Laboratory Standards Institute (CLSI) guidelines [20]. MRSA was screened by cefoxitin disc first and confirmed by the detection of mecA gene. The distribution of these MSSA and MRSA isolates stratified by identified years and sources is shown in Table 1. In addition to CGMH, the isolates were also collected from at least 10 hospitals in Taiwan. Of 105 non-human MRSA isolates, 93 isolates were from pigs, 9 from the environmental objects, 2 from the bus and one from the pork. Of 105 non-human MSSA isolates, 10 isolates were from the bus and 5 from the pork. The clinical (infecting) isolates were identified from any body site, and by epidemiologic criteria, around 460 isolates were community-associated (without risk factors for MRSA acquisition) while 90% of the remaining 2592 isolates were estimated to be healthcare-associated. Among the 2580 MRSA colonizing isolates, 1632 isolates (63%) were identified from children < 18 years of age, including infants hospitalized in neonatal intensive care units (NICUs), neonates in nurseries, children for well baby clinic visits, school children and case patients, while 838 isolates were from adult patients, parturient mothers and elderly stayed in nursing homes. Forty-five percent of the 2580 MRSA colonizing isolates were identified from the subjects without risk factors for MRSA acquisition.

Identification and characterization of S. aureus ST398 isolates

Because ST398 S. aureus cannot be digested by Smal on pulsed-field gel electrophoresis (PFGE), we first examined the isolates which could not be digested by Smal to identify potentially ST398 S. aureus isolates. For those not typeable by Smal digested PFGE, we performed multilocus sequence typing (MLST) to confirm the isolates to be ST398 or not. For ST398 S. aureus isolates, we performed further characterizations, including staphylococcal cassette chromosomal typing (SCCmec), spa typing, and detection of Panton-Valentine leukocidin (PVL) genes and some other toxin genes. All the methods were performed according to the procedures described elsewhere [19, 21, 22]. The antibiotic susceptibility test was performed on Mueller–Hinton agar with disk-diffusion method according to the CLSI guidelines [20] and included teicoplanin, linezolid, fusidic acid, trimethoprim/sulfamethoxazole, ciprofloxacin, doxycycline, erythromycin, clindamycin and penicillin. The susceptibility to vancomycin was determined by minimal inhibition concentration.

Whole genome sequencing (WGS) and phylogenetic analysis

All the ST398 S. aureus isolates identified were subjected to whole genome sequencing (WGS), which was

Table 1 Distribution of methicillin-resistant Staphylococcus aureus and methicillin-susceptible Staphylococcus aureus collected island-wide between 1995 and 2017 and molecularly characterized in Chang Gung Memorial Hospital, stratified by identified years and sources

| Period/population | MRSA No. (%) | MSSA No. (%) | Total No. (%) |
|-------------------|--------------|--------------|---------------|
|                   | Clinical | Colonizing | Subtotal    | Clinical | Colonizing | Subtotal |
| 1995–2004         | 1307     | 502        | 1809 (32)   | 51       | 0          | 51 (6.5) |
| 2005–2010         | 1102     | 1435       | 2537 (45)   | 6        | 311        | 317 (41) |
| 2011–2017         | 643      | 643        | 1286 (23)   | 190      | 223        | 413 (53) |
| Total             | 3052     | 2580 (46)  | 5632        | 247      | 534 (68)   | 781      |
| Pediatric         | 980      | 1632       | 2612 (46)   | 184      | 218        | 402      |
| Adults            | 334      | 838        | 1172 (21)   | 12       | 301        | 313      |
| Unclassified      | 1738     | 5          | 1743 (31)   | 51       | 0          | 51       |
| Non-human         | 0        | 105        | 105 (1.9)   | 0        | 15         | 15       |

*We estimated more than 90% of the isolates were from adult patients
performed on an Illumina MiSeq sequencer (Illumina, San Diego, CA, USA) [23] and the results were deposited at DDBJ/ENA/GenBank under the accession PRJNA554199. The multi-alignment of the core genomes was conducted with the Mauve procedure and the multi-alignment file was subsequently used for maximum likelihood phylogeny construction of the ST398 strains based on the single nucleotide polymorphisms (SNPs) outside of the potential recombination regions with the gubbins procedure [24, 25]. The length of the alignment was 2,561,981 bp and the SNP numbers for the isolates are shown in Fig. 2. The resistance and the virulence genes carried by the local ST398 strains were detected by the srrst2 software using the ARG-ANNOT database and VFDB (http://www.mgc.ac.cn/VFs/), respectively [26, 27]. The final phylogenetic tree and the data of carried resistance and virulence genes were input into Interactive Tree Of Life (http://itol.embl.de) for further annotation.

To clarify the genetic relatedness of the strains in Taiwan and global isolates, 87 published whole genome sequences data of ST398 S. aureus strains were downloaded from the GenBank database and included for analysis. In addition to the maximum likelihood phylogeny construction mentioned above, the Bayesian analysis of the molecular sequences using Markov chain Monte Carlo was performed with the BEAST procedure (v1.10.4) to estimate the time to most common recent ancestor [28]. A HKY model with estimated base frequencies, strict clock type and constant size of tree prior was used. Three independent chains were run for 100 million generations and sampling every 1000 generations. The effect sample size values were checked by tracer (v1.7.1) and were greater than 200. A burn-in of 10 million states was removed from each of the three independent runs before combining the results from those runs with the logcombiner program (v1.10.4) from the BEAST package. The final tree was output with Figtree (https://github.com/rambaut/figtree/releases). The resistance and the virulence genes carried by the local ST398 strains were detected by the srrst2 software using the ARG-ANNOT database and VFDB (http://www.mgc.ac.cn/VFs/), respectively [26, 27].

Clinical features and sources of the ST398 isolates identified in 2012 possessed PVL genes while the other seven isolates did not. The six MSSA isolates belonged to four spa types. Both MSSA isolates from patient 10 with recurrent acute otitis media belonged to spa type t1250, and both isolates from workers handling pork belonged to spa type t571.

All 18 isolates were susceptible to vancomycin, teicoplanin, linezolid, fusidic acid while 17 were susceptible to doxycycline, and trimethoprim/sulfamethoxazole. One MRSA (RR82) and MSSA (ZS13) isolate was resistant to ciprofloxacin. Of the six MSSA isolates, one isolate (ZS13) was non-susceptible to doxycycline, and four were resistant to penicillin (Table 3).

Whole genome sequencing and phylogenetic analysis of ST398 S. aureus

The core genome-based phylogeny of the 18 local strains and 87 global strains of ST398 is shown in Fig. 1a. Multiple likelihood analysis of SNPs in core genomes disclosed all twelve local ST398 MRSA strains were clustered together and were genetically related to a human MRSA strain (strain 36_CN_2002) isolated from China in 2002. The BEAST analysis (Fig. 1b) suggested that all of the ST398 MRSA human isolates from Taiwan shared the
common ancestor in 1986 with a human strain (strain 36_CN_2002) isolated from China in 2002 and then were diverged in 2006. These findings suggested that the local ST398 MRSA shared the common ancestor of the isolate from China and had caused local transmission in some long-term care facilities in Taiwan after 2012. Of the six ST398 MSSA isolates, except for two strains (OSSA1096 and OSSA1105) isolated from the child with recurrent otitis media in different time points, the phylogeny of the local ST398 MSSA strains, though all of them were identified in the same year (2015), was very diverse. The number of SNPs between these MSSA isolates could be greater than 550, suggesting different origins of the isolates (Fig. 1a).

The BEAST analysis suggested that all the ST398 MSSA isolates but the strain of ZS13 were derived from another ancestor in 1974, which was shared by some human isolates from France, United States and China between 2007 and 2009. The details of the comparative genomics of the ST398 strains were provided (Fig. 1).

The distributions of the virulence and resistance genes carried by these strains are displayed in Fig. 2. The resistant profile was distinct between MRSA and MSSA isolates while the toxin profile was relatively similar between both groups of the isolates. All 12 MRSA isolates were found to lack the tetM resistance gene and contain the immune evasion complex (IEC) genes, including chp, scn, and sak, which are genetic determinants previously associated with human origin and adaptation, thus suggesting a non-livestock-associated

### Table 3

| Strain name | Subject No. | CC | E | P | SXT | D | CIP |
|-------------|-------------|----|---|---|-----|---|-----|
| RR70        | 1           | S  | S | R | S   | S | S   |
| RR69        | 2           | S  | S | R | S   | S | S   |
| RR74        | 3           | S  | S | R | S   | S | S   |
| RR82        | 4           | S  | S | R | S   | S | S   |
| RR95        | 5           | S  | S | S | S   | S | S   |
| AAS1#       | 6           | S  | S | S | S   | S | S   |
| AAS9#       | 7           | S  | R | S | S   | S | S   |
| ZS13#       | 8           | R  | R | R | S   | I | R   |
| OSSA1032#   | 9           | R  | R | R | S   | S | S   |
| OSSA1096#   | 10          | R  | R | R | S   | S | S   |
| OSSA1105#   | 10          | R  | R | R | S   | S | S   |
| ABR26       | 11          | S  | S | R | S   | S | S   |
| ABR171      | 11          | R  | R | R | S   | S | S   |
| ABR112      | 12          | R  | R | R | I   | S | S   |
| ABR121      | 13          | S  | S | R | S   | S | S   |
| ABR134      | 14          | S  | S | R | S   | S | S   |
| ABR180      | 15          | S  | S | R | S   | S | S   |
| ACR49       | 16          | S  | S | R | S   | S | S   |

**Table 2** Detailed clinical features of the 16 subjects with 18 ST398 *Staphylococcus aureus* isolates identified in Taiwan

| Strain name | Subject No. | Year of isolation | Age (years) | Status | Remarks |
|-------------|-------------|-------------------|-------------|--------|---------|
| RR70        | 1           | 2012              | 69          | Colonized | Survey in 14 nursing homes |
| RR69        | 2           | 2012              | 53          | Colonized | Survey in 14 nursing homes |
| RR74        | 3           | 2012              | 63          | Colonized | Survey in 14 nursing homes |
| RR82        | 4           | 2012              | 82          | Colonized | Survey in 14 nursing homes |
| RR95        | 5           | 2012              | 72          | Colonized | Survey in 14 nursing homes |
| AAS1#       | 6           | 2015              | –           | Non-human | Pork meat in a supermarket |
| AAS9#       | 7           | 2015              | Unknown     | Colonized | Survey for pork processing workers |
| ZS13#       | 8           | 2015              | Unknown     | Colonized | Survey for pork processing workers |
| OSSA1032#   | 9           | 2015              | 2           | Sinusitis | Cured |
| OSSA1096#   | 10          | 2015              | 4           | Otitis media | Combined immunodeficiency, Recurrent otitis |
| OSSA1105#   | 10          | 2015              | 4           | Otitis media | Combined immunodeficiency, Recurrent otitis |
| ABR26       | 11          | 2016              | 86          | Colonized (resident) | 3rd Longitudinal survey in a nursing home (1st survey) |
| ABR171      | 11          | 2016              | 86          | Colonized (resident) | (4th survey) |
| ABR112      | 12          | 2016              | 93          | Colonized (resident) | (3rd survey) |
| ABR121      | 13          | 2016              | 29          | Colonized (worker, Philippines) | (3rd survey) |
| ABR134      | 14          | 2016              | 30          | Colonized (worker, Philippines) | (3rd survey) |
| ABR180      | 15          | 2016              | 26          | Colonized (worker, Philippines) | (4th survey) |
| ACR49       | 16          | 2017              | 27          | Colonized (worker, Indonesia) | Survey for foreign workers recruited to Taiwan |

### Table 3

| Strain name | Subject No. | CC | E | P | SXT | D | CIP |
|-------------|-------------|----|---|---|-----|---|-----|
| RR70        | 1           | S  | S | S | R   | S | S   |
| RR69        | 2           | S  | S | S | R   | S | S   |
| RR74        | 3           | S  | S | S | S   | S | S   |
| RR82        | 4           | S  | S | S | S   | S | S   |
| RR95        | 5           | S  | S | S | S   | S | S   |
| AAS1#       | 6           | S  | S | S | S   | S | S   |
| AAS9#       | 7           | S  | R | S | S   | S | S   |
| ZS13#       | 8           | R  | R | R | S   | S | S   |
| OSSA1032#   | 9           | R  | R | R | S   | S | S   |
| OSSA1096#   | 10          | R  | R | R | S   | S | S   |
| OSSA1105#   | 10          | R  | R | R | S   | S | S   |
| ABR26       | 11          | S  | S | R | S   | S | S   |
| ABR171      | 11          | R  | R | R | S   | S | S   |
| ABR112      | 12          | R  | R | R | I   | S | S   |
| ABR121      | 13          | S  | S | R | S   | S | S   |
| ABR134      | 14          | S  | S | R | S   | S | S   |
| ABR180      | 15          | S  | S | R | S   | S | S   |
| ACR49       | 16          | S  | S | R | S   | S | S   |

**Table 3** Antibiograms of the 16 patients with 18 ST398 *Staphylococcus aureus* isolates identified in Taiwan

- CC clindamycin, E erythromycin, P penicillin, SXT trimethoprim-sulfamethoxazole, D doxycycline, CIP ciprofloxacin
- All 18 isolates were susceptible to vancomycin, teicoplanin, fusidic acid and linezolid
- These six isolates were methicillin-sensitive *Staphylococcus aureus*; others were MRSA
- The frequency of sampling in this longitudinal study was bi-monthly

Huang and Chen *Journal of Biomedical Science* (2020) 27:15
Fig. 1 (See legend on next page.)
origin. All but 2 MSSA isolates (AAS1 and ZS13) were found to be human adapted. Strain ZS13, which was identified from a worker handling pork in a traditional wet market, had abundant resistant genes but was completely absent for the IEC genes, suggesting a livestock-associated MSSA isolate. Strain AAS1, identified from the pork specimen, had sak gene but absent for tetM determinant, scn gene and chp gene, implying a potential livestock-associated.

Discussion

To our knowledge, this is the first report focusing on MRSA ST398 in Taiwan. In this study, the first ST398 isolate was identified in 2012 from a resident in one nursing home in a survey conducted in 2012 for MRSA carriage [19], which suggests that ST398 existed in Taiwan since 2012 or earlier. Also, though the S. aureus isolates collected between 2012 and 2017 in our laboratory only accounted for 15.7% of the pool of 6413 S. aureus isolates which were collected between 1995 and 2017, all the 18 ST398 isolates in this study were identified since 2012, suggesting the identification increased markedly recently in Taiwan.

MRSA ST398 was first identified as a LA-MRSA and prevailed in Europe, then in the North America but rarely reported from Asian countries [1–4]. Human infections of LA-MRSA ST398 have been reported, particularly from Europe, since first identified from pigs [3, 4, 29–31] and may manifest from a minor infection of skin and soft tissue infection, to life-threatening events of necrotizing pneumonia, bloodstream infection and endocarditis, even resulting in subsequent lethality. LA-MRSA has become an important cause of skin and soft tissue infections (SSTIs) in previously healthy individuals in European countries with a low prevalence of MRSA [29, 30]. The strains can also cause several difficult-to-treat infections and even death in patients with or without exposure to livestock [32–34].

The prevalence of LA-MRSA among pigs in Asian countries varied widely by geographical locations [1, 2]. In most Asian countries, LA-MRSA isolates belong to

![Fig. 1](image1.png)

**Fig. 1** a Maximum likelihood phylogenetic analysis of 18 ST398 Staphylococcus aureus isolates from Taiwan based on single nucleotide polymorphisms in the core genomes. 87 global ST398 isolates were included for comparison. All 12 local ST398 human MRSA strains were clustered and were genetically related to a human MRSA strain (36_CN_2002) isolated from China in 2002. The phylogenetic tree was produced by gubbins procedure using the multi-alignment file of all ST398 genomes generated by Mauve procedure and finally annotated in the tree of life website (Methods for detailed description). b The estimated time to most common recent ancestor among the ST398 strains with the BEAST program. The phylogenetic tree of isolates from Taiwan and China were amplified for more clear display. All of the ST398 MRSA human isolates from Taiwan shared the most common recent ancestor in 1986 with the strain of 36_CN_2002 from China in 2002. All ST398 MSSA isolates but the strain of ZS13 shared the most common recent ancestor in 1974 with some isolates from France, United States and China identified between 2007 and 2009. Abbreviation: AT, Austria; BE, Belgium; CA, Canada; CH, Switzerland; CN, China; DE, Germany; DK, Denmark; ES, Spain; FI, Finland; FR, France; GF, French Guiana; HU, Hungary; IT, Italy; NL, The Netherlands; PE, Peru; PL, Poland; PT, Portugal; SI, Slovenia; TW, Taiwan; US, United States.

![Fig. 2](image2.png)

**Fig. 2** The distributions of the virulence and resistance genes identified in the 18 ST398 Staphylococcus aureus isolates from Taiwan. Human-adapted immune evasion complex genes (sak, chp and scn genes) were identified in all human ST398 isolates except Strain ZS13 but mostly missing in the isolate identified from the pork meat (strain AAS1). The figure was produced using the tree of life website (http://itol.embl.de).
ST9, but with different molecular characteristics [1, 2]. LA-MRSA ST9 was rarely reported to be associated with human diseases at present [35, 36]. In a lately report from Taiwan [18], LA-MRSA ST9 was identified in 0.24% of 3328 human MRSA clinical (infecting) isolates collected from 1998 to 2012 islandwide. However, LA-MRSA ST398 was rarely identified from Asian countries and only reported from Korea (in commercial pigs) [5, 6], Singapore (in experimental pigs in a hospital research) [7], Japan [8–10] and Thailand [11].

Human-adapted ST398 isolates, genetically different from LA clones, often cause life-threatening infections and share some genetic features for the adaptation to humans as hosts [12–14, 34]. Almost all previously reported human-adapted ST398 isolates are MSSA [12, 37]. Lately, eight cases, of which six were severe and two were fatal, infected with ST398 CA-MRSA without connection to livestock were reported from China [12]. Genomic analysis revealed that these highly infectious ST398 CA-MRSA isolates evolved from human-adapted methicillin-susceptible clones. Similar severe, even fatal, cases were also reported from Japan [8] and Australia [38]. This strain seemed to be spreading internationally and needs further surveillance and monitoring.

In this study, only three MSSA isolates from two children were infecting isolates and all three isolates were genetically human-adapted by WGS analysis, as reported from China [12, 37]. Both pediatric patients presented with respiratory tract infections, including acute otitis media and sinusitis. In contrast, all 12 MRSA ST398 isolates in the present study were colonizing isolates. No invasive diseases caused by these ST398 strains were identified yet in Taiwan.

In this study, toxin profiles showed that all 12 MRSA ST398 isolates and four of six MSSA ST398 isolates were human-adapted while only two MSSA ST398 isolates from a worker handling pork in a traditional wet market and a pork meat, respectively, were potentially livestock-associated, suggesting that LA-S. aureus ST398 isolates were still uncommonly circulating but potentially existed in Taiwan. Multiple likelihood analysis of SNPs in core genomes revealed that all twelve local ST398 MRSA strains were clustered together and when compared with 87 global ST398 strains, they were phylogenetically related to a strain identified from China in 2002. The further BEAST analysis (Fig. 1b) showed that all of the ST398 MRSA human isolates from Taiwan shared the common ancestor in 1986 with the human strain isolated from China in 2002 and then were diverged in 2006.

Of noting, five MRSA isolates were identified from five residents and three health care workers in one NH in a longitudinal survey conducted in 2016 in northern Taiwan. These findings suggest that MRSA ST398 might have been circulating in some NHs recently in Taiwan. In contrast, the phylogeny of the six local ST398 MSSA strains, though all isolated in the same year (2015), was very diverse except for two isolates from a single subject in different time points, which suggest that these isolates might have different origins and might be unrelated to each other (no local transmission). These findings were supported by the BEAST analysis.

There are several limitations of this study. Firstly, since all the ST398 S. aureus isolates identified and characterized in this study were only from our research collection, further studies are needed to figure out the whole perspective of ST398 S. aureus in Taiwan. Secondly, the nature of present study is retrospective, so in addition to four foreign workers (three from the Philippines and one from Indonesia), the detailed traveling history of going abroad could not be identified exactly from the case patients as well as their household members. Thus, whether the isolates of ST398 from these patients were imported or not cannot be traced clearly.

Conclusions
Though still uncommon, ST398 S. aureus strain was identified in Taiwan since 2012. Among the ST398 S. aureus isolates identified in Taiwan, most isolates, particularly MRSA, were human-adapted, though few isolates were potentially livestock-associated. Local transmission of human-adapted MRSA ST398 strains have occurred in some nursing homes in Taiwan after possibly imported from China. Further surveillance of local MRSA molecular epidemiology is needed.

Abbreviations
CA: Community-associated; HCW: Health care worker; IEC: Immune evasion complex; LA: Livestock-associated; MRSA: Methicillin-resistant Staphylococcus aureus; NH: Nursing home; PFGE: Pulsed-field gel electrophoresis; PVL: Panton-Valentine leucocidin; SCC: Staphylococcal cassette chromosome; ST: Sequence type

Acknowledgements
Not applicable.

Authors’ contributions
YCH designed the study, applied the grants, providing the laboratory works, analyzed and interpreted the data and writing the manuscript. CJC analyzed and interpreted the data. All authors read and approved the final manuscript.

Funding
This study was supported by a grant from Ministry of Science and Technology, Republic of China (MOST 105-2314-B-182A-138) and a grant from Chang Gung Memorial Hospital (CMRPG3F1852).

Availability of data and materials
The data that support the findings of this study are available from the corresponding author upon reasonable request.

Ethics approval and consent to participate
This study was approved by the Institutional Reviewing Board of Chang Gung Memorial Hospital.
Consent for publication
Not available.

Competing interests
The authors declare that they have no competing interests.

Author details
1Division of Pediatric Infectious Diseases, Chang Gung Memorial Hospital, Taoyuan, Taiwan. 2Chang Gung University College of Medicine, Taoyuan, Taiwan. 3Department of Pediatrics, Chang Gung Memorial Hospital, No. 5, Fu-Shin Street, Gueishan, 333 Taoyuan, Taiwan.

Received: 6 August 2019 Accepted: 19 December 2019
Published online: 03 January 2020

References
1. Chuang YY, Huang YC. Livestock-associated meticillin-resistant Staphylococcus aureus in Asia: an emerging issue? Int J Antimicrob Agents. 2015;45:334–40.
2. Chen CJ, Huang YC. Emergence of livestock-associated meticillin-resistant Staphylococcus aureus: should it be a concern? J Formos Med Assoc. 2018;117:658–61.
3. Lewis HC, Mølbak K, Reese C, Aarestrup FM, Selchau M, Sørum M, et al. Pigs as source of meticillin-resistant Staphylococcus aureus CC398 infections in humans. Denmark. Emerg Infect Dis. 2008;14:1383–9.
4. Larsen J, Petersen A, Larsen AR, Sieber RN, Stegger M, Koch A, et al. Emergence of livestock-associated meticillin-resistant Staphylococcus aureus bloodstream infections in Denmark. Clin Infect Dis. 2017;65:1072–6.
5. Lim SK, Nam HM, Jang GC, Lee HS, Jung SC, Kwak HS. The first detection of meticillin-resistant Staphylococcus aureus ST398 in pigs in Korea. Vet Microbiol. 2012;155:88–92.
6. Moon DC, Tamang MD, Nam HM, Jeong JH, Jang GC, Jung SC, et al. Identification of livestock-associated meticillin-resistant Staphylococcus aureus isolates in Korea and molecular comparison between isolates from animal carcasses and slaughterhouse workers. Foodborne Pathog Dis. 2015;12:327–34.
7. Sergio DM, Kho TH, Hsu LY, Ogden BE, Goh AL, Chow PK. Investigation of meticillin-resistant Staphylococcus aureus in pigs used for research. J Med Microbiol. 2007;56:1107–9.
8. Koyama H, Sanui M, Saga T, Harada S, Ishii Y, Tateda K, et al. A fatal infection caused by sequence type 398 meticillin-resistant Staphylococcus aureus carrying the Panton-valentine leukocidin gene: A case report in Japan. J Infect Chemother. 2015;21:541–3.
9. Furuno M, Uchiyama M, Nakahara Y, Fujihara H, Morino S, et al. A Japanese trial to monitor meticillin-resistant Staphylococcus aureus (MRSA) in imported swine during the quarantine period. J Glob Antimicrob Resist. 2018;14:182–4.
10. Aung MS, Urushibara N, Kawaguchiya M, Sumi A, Shinagawa M, Takahashi S, et al. Clonal diversity and genetic characteristics of meticillin-resistant Staphylococcus aureus isolates from a tertiary care hospital in Japan. Microb Drug Resist. 2019. https://doi.org/10.1089/mdr.2018.0468.
11. Chanchaitong P, Pereret V, Arm-In N, Lugisonya K, Tummaruk P, Praparaksak N. Molecular characterization and antimicrobial resistance of livestock-associated meticillin-resistant Staphylococcus aureus isolates from pigs and swine workers in Central Thailand. Microb Drug Resist. 2019. https://doi.org/10.1089/mdr.2019.0011.
12. He L, ZhengHX, Wang Y, Le KY, Liu Q, Shang J, et al. Detection and analysis of meticillin-resistant human-adapted sequence type 398 allows insight into community-associated meticillin-resistant Staphylococcus aureus evolution. Genome Med. 2018;10:5.
13. Price LB, Stegger M, Hasman H, Aziz M, Larsen J, Andersen PS, et al. Staphylococcus aureus CC398: host adaptation and emergence of meticillin resistance in livestock. MBio. 2012;3:e00305–11.
14. Sun L, Chen Y, Wang D, Wang H, Wu D, Shi K, et al. Surgical site infections caused by highly virulent meticillin-resistant Staphylococcus aureus sequence type 398. China Emerg Infect Dis. 2019;25:157–60.
15. Huang YC, Chen CJ. Community-associated meticillin-resistant Staphylococcus aureus in children in Taiwan, 2000s. Int J Antimicrob Agents. 2011;38:2–8.
16. Chuang YY, Huang YC. Molecular epidemiology of community-associated meticillin-resistant Staphylococcus aureus in Asia. Lancet Infect Dis. 2013;13:698–708.
17. Chen CJ, Huang YC. New epidemiology of Staphylococcus aureus in Asia. Clin Microbiol Infect. 2014;20:605–23.
18. Chen CJ, Lauderdale TY, Lu CT, Chuang YJ, Yang CC, Wu TS, Lee CY, Lu MC, Ko WC, Huang YC. Clinical and molecular features of MRD livestock-associated MRSA ST9 with staphylococcal cassette chromosome mecVIII in humans. J Antimicrob Chemother. 2018;73:334–40.
19. Tsao FY, Kuo HW, Huang YC. Dissemination of meticillin-resistant Staphylococcus aureus ST45 among nursing home residents and staff in Taiwan. Clin Microbiol Infect. 2015;21:451–8.
20. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing, 20st informational supplement; CLSI document M100-S20. Wayne: CLSI, 2010.
21. Huang YC, Chen CJ. Nasal carriage of meticillin-resistant Staphylococcus aureus during the first two years life in children in northern Taiwan. Pediatr Infect Dis J. 2013;34:131–5.
22. Huang YC, Su LH, Wu TL, Lin YT. Meticillin-resistant Staphylococcus aureus nasal carriage in international medical conference attendees. J Med Microbiol Immunol Infect. 2019;52:242–7.
23. Bankevich A, Nurk S, Antipov D, et al. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol. 2012;19:455–77.
24. Darling ACE, Mao B, Blattner FR, Perna NT. Mauve: multiple alignment of conserved genomic sequence with rearrangements. Genome Res. 2004;14:1394–403.
25. Croucher NJ, Page AJ, Connor TR, et al. Rapid phylogenetic analysis of large samples of recombinant bacterial whole genome sequences using Gubbins. Nucleic Acids Res. 2015;43:e11.
26. Inouye M, Dashnow H, Raven LA, et al. SRST2: rapid genomic surveillance for public health and hospital microbiology labs. Genome Med. 2014;6:90.
27. Gupta SK, Padmanabhan BR, Diene SM, et al. ARG-ANNOT, a new bioinformatic tool to discover antibiotic resistance genes in bacterial genomes. Antimicrob Agents Chemother. 2014;58:212–20.
28. Suchard MA, Lemey P, Baele DL, Drummond AJ, Rambaut A. Bayesian phylogenetic and phylodynamic data integration using BEAST 1. 10. Virus Evol. 2013:evy016.
29. Lozano C, Aspizio C, Ana M, Gómez-Sanz E, Zaragoza M, Torres C. Meticillin-resistant Staphylococcus aureus (MRSA) ST398 in a farmer with skin lesions and in pigs of his farm: clonal relationship and detection of Imm (A) gene. Clin Microbiol Infect. 2011;17:923–7.
30. van de Sande-Bruinsma NA, Leerverstein van Hall MA, Janssen M, Nagzaan N, Leenders S, de Grief SC, et al. Impact of livestock-associated MRSA in a hospital setting. Antimicrob Resist Infect Control. 2015;4:41.
31. Spoor LE, McAdam PR, Weinstert LA, Rambaut A, Hasman H, Aarestrup FM, Keams AM, Larsen AR, Skov RL, Fitzgerald JR. Livestock origin for a human pandemic clone of community-associated meticillin-resistant Staphylococcus aureus. MBio. 2013;4:e00356–13.
32. Larsen J, Petersen A, Sorum M, Stegger M, Van Alphen L, Valentiner-Branth P, et al. Meticillin-resistant Staphylococcus aureus CC398 is an increasing cause of disease in people with no livestock contact in Denmark, 1999 to 2011. Euro Surveill. 2015;20:3711.
33. Nielsen RT, Kemp M, Holm A, Skov MN, Detlefsen M, Hasman H, et al. Fatal septicemia linked to transmission of MRSA clonal complex 398 in hospital and nursing home. Denmark Emerg Infect Dis. 2016;22:900–2.
34. Larsen J, Stegger M, Andersen PS, Petersen A, Larsen AR, Westh H, Agero Y, F Jetsch A, Kraushaar B, Kasbohrer A, et al. Evidence for human adaptation and foodborne transmission of livestock-associated meticillin-resistant Staphylococcus aureus. Clin Infect Dis. 2016;63:1349–52.
35. van Loo I, Huijsdens X, Tiemersma E, de Neeling A, van de Sande-Bruinsma NA, Beujiang D, et al. Emergence of meticillin-resistant Staphylococcus aureus of animal origin in humans. Emerg Infect Dis. 2007;13:1894–9.
36. Liu Y, Wang H, Du N, Shen E, Chen H, Niu J, et al. J Med Microbiol. Molecular evidence for spread of two major meticillin-resistant Staphylococcus aureus clones with a unique geographic distribution in Chinese hospitals. Antimicrob Agents Chemother. 2009;53:12–8.
37. Zhao C, Liu Y, Zhao M, Liu Y, Yu Y, Chen H, et al. Characterization of community-acquired Staphylococcus aureus associated with skin and soft tissue infections in Beijing: high prevalence of PVL+ ST398. PLoS One. 2012;7:e38577.
38. Coombs GW, Pang S, Daley DA, Lee Y, Abraham S, Leroi M. Severe disease caused by community-associated MRSA ST398 type V, Australia, 2017. Emerg Infect Dis. 2019;25:190–2.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.