Study on the mechanism of lupenone for treating type 2 diabetes by integrating pharmacological evaluation and network pharmacology

Feng Xu a,*, Mei Zhang a,*, Hongmei Wu a, Yuanmin Wang a, Ye Yang a and Xiangpei Wang a,b

a College of Pharmacy, Guizhou University of Traditional Chinese Medicine, Guiyang, PR China; b College of Pharmacy, Guizhou Minzu University, Guiyang, PR China

ABSTRACT

Context: Lupenone (LUP) is the active ingredient of Musa basjoo Sieb. et Zucc. (Musaceae) with antidiabetes effects, but an unclear underlying mechanism of action.

Objective: Animal experiments combined with network pharmacology were used to explore the mechanism of LUP for treating diabetes.

Materials and methods: Insulin resistance (IR) in male Sprague-Dawley rats with type 2 diabetic was induced using a high-fat diet and streptozotocin. The selected rats were divided into normal group, model group, positive group and LUP (2.0, 4.0 and 8.0 mg/kg) groups, and orally administrated twice daily with Tween 80, rosiglitazone or LUP. Fasting blood glucose (FBG), oxidative stress index, blood lipids and IR-related targets were detected. A network pharmacology analysis was performed.

Results: Compared to the model group, LUP (8.0 mg/kg) significantly decreased the levels of FBG (22.3%), LEP (9.5%), HbA1c (14.9%) and MDA (12.3%), increased the ADPN (24.2%) levels and GSH-PX activity (12.4%) (p < 0.05), improved oxidative stress, lipid metabolism disorders and pancreas pathological changes, increased the mRNA and protein expression of InsR (3.7-fold and 1.3-fold), IRS-1 (3-fold and 2-fold), IRS-2 (2-fold and 1.6-fold), GLUT-4 (2-fold and 2.4-fold) in skeletal muscle and IRS-1 (6-fold and 1.6-fold), IRS-2 (5.8-fold and 1.5-fold), GLUT-4 (2.5-fold and 1.7-fold) and PPAR-γ (7-fold and 1.4-fold) in adipose tissue (p < 0.05). Network pharmacology analysis revealed that LUP improves IR by multiple targets and signal pathways.

Conclusions: The mechanism of LUP for treating diabetes is related to improving IR. LUP has the potential to be developed as a new drug for treating type 2 diabetes.

Introduction

Diabetes mellitus is a chronic disease characterized by hyperglycaemia (Ogurtsova et al. 2017). There are three main types of diabetes including type 1 diabetes, type 2 diabetes (T2DM) and gestational diabetes. Among these, T2DM is the most common. T2DM is a burden on global health-care systems (Radziuk and Pye 2001; Peiró et al. 2017). Clinical drugs currently used for the treatment of T2DM have many limitations and may result in adverse reactions, such as gastrointestinal symptoms, hypoglycaemia, oedema, osteoporosis and lactic acidosis (Donath 2014). Therefore, it is necessary to find an ideal drug for the safe and effective prevention and treatment of T2DM. Insulin resistance (IR) and its complications are one of the main factors leading to T2DM (Guo et al. 2021). IR refers to a decrease in the response of the biological target tissues, mainly in the liver, muscle and adipose tissue to normal physiological concentrations of insulin. Modern research finds that improving IR is an effective way to treat T2DM. Traditional Chinese medicine could treat T2DM through improving the IR, especially by increasing the sensitivity of the muscle, adipose tissue and liver to insulin (Tang et al. 2011; Sheng et al. 2014; Li et al. 2016). Rhizoma musae is the dried rhizome musae of Musa basjoo Sieb. et Zucc. (Musaceae), a traditional Chinese medicine. Our research team has confirmed that the Rhizoma musae, has a good antidiabetic effect, lupenone (LUP) isolated from the Rhizoma musae, has a good inhibitory effect on protein tyrosine phosphatase 1B (PTP1B) and α-glucosidase in vitro (Na et al. 2009; Wang et al. 2012; Seong et al. 2016). Furthermore, LUP could decrease the fasting blood glucose (FBG) in mice with diabetes and improve the oral glucose tolerance and insulin tolerance in rats with T2DM (Xu et al. 2014; Wu et al. 2015, 2017). These results indicate that LUP has definite antidiabetic effects. However, the mechanism of LUP in the treatment of T2DM has not been elucidated. The current study elucidates the mechanism of action of LUP to improve IR using animal experiments and network pharmacology methods.

Materials and methods

Reagents

LUP was isolated from the Rhizoma musae by our laboratory. Rhizoma musae was collected from Guizhou Province, and was identified as the rhizome musae of Musa basjoo Sieb. et Zucc. by Professor Xiangpei Wang of Guizhou Minzu University. A...
voucher specimen is kept in the Drug Analysis Laboratory of Guizhou University of Traditional Chinese Medicine. *Rhizoma musae* was soak with ethanol after crushing, recovering the ethanol after filtration and concentrated to the extract, then dispersed with an appropriate amount of water and extracted with petrol-eum ether, ethyl acetate and n-butanol. Further, the resulting ethyl acetate extract was separated by silica gel column, and further purified by silica gel column and identified by comparison of its spectral data (mp, MS, $^1$H NMR, $^{13}$C NMR) with the reference standards. The purity of isolated LUP is over 98% by HPLC analysis as our previous report (Wang et al. 2012; Xu et al. 2014; Wu et al. 2021). Streptozotocin (STZ) was purchased from Sigma-Aldrich (St. Louis, MO). All other chemicals used in the study were of analytical grade.

**Animals**

All experiments were performed on 6-week-old male Sprague-Dawley rats (200 ± 20 g) obtained from the Chongqing Tengxin Biological Technology Co., Ltd. (Chongqing, PR China). The colony was maintained under controlled conditions of temperature 23 ± 2°C humidity, 50 ± 5%, and 12 h light/dark cycle. All the animals in the study were cared for and treated humanely according to the National Legislations of China, as well as the local guidelines. The animal experiments were approved by the Guizhou University of Traditional Chinese Medicine and the Ethics Committee for Animal Experiments of West China Hospital of Sichuan University (under registration: 2013029A).

**Induction of type 2 diabetic rats and treatment**

After acclimatization for one week, the rats were randomly divided into normal group and high-fat diet (HFD) group. The rats in the normal group were fed with a standard diet *ad libitum* and the rats in the HFD group were fed with an HFD (15% cholesterol, 15% coconut oil, 15% granulated sugar and 55% standard diet *ad libitum*). After feeding for four weeks, all rats were fasted for 12 h, then the HFD group rats were injected twice with low-dose STZ (first with 30 mg/kg, i.p.; second with 28 mg/kg, i.p.; 24 h later) to induce partial insulin deficiency, and the normal group were intraperitoneally injected with sodium citrate buffer. Seventy-two hours after injection, the FBG levels of the rats were measured, and the rats with FBG of ≥11.1 mmol/L were categorized as those with T2DM. Furthermore, rats with T2DM were randomly divided into T2DM group, positive group and three LUP treatment groups. The normal and T2DM groups received the vehicle treatment (0.03% Tween 80; twice daily), the positive group was administered the rosiglitazone (1.00 mg/kg; twice daily) and free fatty acid (FFA) in serum were analysed using commercial enzyme-linked immunosorbent assay (ELISA) kit, according to the manufacturer’s instructions (Abcam, Cambridge, UK). The levels of total cholesterol (TC), triglyceride (TG), high-density lipoprotein-cholesterol (HDL-C) and low-density lipoprotein-cholesterol (LDL-C) were detected using automatic biochemical analyser (Olympus AU640, Tokyo, Japan).

**Biochemical parameter analysis**

The levels of serum leptin (LEP), haemoglobin A1c (HbA1c), superoxide dismutase (SOD), malondialdehyde (MDA), glutathione peroxidase (GSH-PX), adiponectin (ADPN) and free fatty acid (FFA) in serum were monitored every three days. The rest rats were fasted for 12 h every weekend to measure the FBG levels using the One-touch Ultra Blood Glucose Monitoring System (Johnson & Johnson Medical (China) Ltd., Shanghai, PR China).

**Periodic acid-Schiff staining**

Distribution of glycogen particles in thin sections of liver samples was investigated using PAS staining methods. Briefly, the liver was routinely processed and embedded in paraffin. Sections of 5 μm thickness were cut. The sections of the liver tissue were deparaffinized and hydrated, immersed in 0.5% periodic acid solution for 10 min, and rinsed in distilled water for another 5 min. This was done twice. The sections were then placed in the Schiff reagent for 10 min and washed in water for 5 min before counterstained with haematoxylin for 2 min. Stained reddish dots or threadlike structures in the liver cells were considered as a positive result.

**Histological analysis**

The pancreas was fixed in 10% neutral buffered formalin for 48 h. Samples were subsequently routinely processed and embedded in paraffin. Sections of 5 μm thickness were cut and stained with haematoxylin and eosin (HE) for general histopathological examination under a light microscope, counting the number of islets in the section.

**Quantitative real-time PCR**

The total ribonucleic acid (RNA) of the skeletal muscle and adipose tissue was isolated using TRIzol reagent (Invitrogen, Carlsbad, CA), and was used to synthesize complementary deoxynucleic acid (cDNA) using the PrimeScript RT reagent kit according to the manufacturer’s protocol (TaKaRa Bio, Dalian, China). Then, the amplification reactions were carried out in 96-well reaction plates (Bio-Rad, Hercules, CA) with a 20 μL reaction volume. Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was used as an endogenous control, and the fold change in the level of the target messenger ribonucleic acid (mRNA) between the control and treatment
groups was normalized by the level of GAPDH. The change in gene expression was calculated using $2^{-\Delta\Delta Ct}$ (Xu et al. 2017).

Primer sequences for the analysis of insulin receptor (InsR), insulin receptor substrate (IRS)-1, IRS-2 and glucose transporter type 4 (GLUT-4) mRNA in the skeletal muscle and peroxisome proliferator-activated receptor (PPAR)-γ, IRS-1, IRS-2 and GLUT-4 mRNA in the adipose tissue are described in Table 1.

Table 1. PCR primers used in all quantitative PCR assays.

| Primer: gene | Forward | Reverse |
|--------------|---------|---------|
| Gapdh        | 5'-CCA AGG TCA TCC ATG ACA AC-3' | 5'-TGT CAT ACC AGG AAA TGA GC-3' |
| Insr         | 5'-GCT GGT GTT TTC CTC ATA GAT TG-3' | 5'-CTG TAG GTC AGG TTT GTT TT-3' |
| IRS1         | 5'-CCT GAC ATT GGA GGT GGG TCT T-3' | 5'-GAA TCT TCG GCA GGG GTG T-3' |
| IRS2         | 5'-TGC GAA CAG CCG TCG GTG AC-3' | 5'-GAC CGG TGA CCG CTG AAC GG-3' |
| GLUT4        | 5'-AGC CAG CCT ACG CCA CCA TA-3' | 5'-GGC GGT CTC ATG AAT AA-3' |
| Pparc        | 5'-TGG GCC TCC CTG ATG AAT AA-3' | 5'-GGC GGT CTC ATG AAT AA-3' |

Figure 1. Lupenone prevented the body weight loss (a), decreased the fasting blood glucose level (b) in rats with diabetes. Data presented as mean ± SD. $N = 9–10$ for the change in body weight change and fasting blood glucose. *$p < 0.05$ vs. the T2DM group, **$p < 0.05$ vs. the normal group. SD: standard deviation.

Figure 2. Lupenone decreased the levels of LEP (A) and HbA1c (C), increased the ADPN (B) levels, and reduced the excessive oxidative stress in the serum of rats with diabetes. Lupenone decreased the levels of MDA (D) and reduced GSH-PX (F) activity, but has no significantly effect on the SOD (E) activity. Data are presented as means ± SD. $N = 9–10$ there are nine rats in the normal group and 10 rats in the other groups. *$p < 0.05$ vs. the T2DM group, **$p < 0.05$ vs. the normal group SD: standard deviation.
Western blotting

Proteins were extracted from the skeletal muscle and adipose tissue by using lysis buffer. The proteins were then separated using sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) (10% or 15%) and electrophoretically transferred into polyvinylidene fluoride membranes. The membranes were probed with anti-InsR (1:20,000; Abcam, Cambridge, UK), anti-IRS-1 (1:500; Abcam, Cambridge, UK), anti-IRS-2 (1:5000; Abcam, Cambridge, UK), anti-GLUT-4 (1:2000; Abcam, Cambridge, UK) and anti-PPARγ (1:5000; Abcam, Cambridge, UK) antibodies overnight at 4°C, and then incubated with a horse radish peroxidase (HRP)-coupled secondary antibody. Detection was performed using a ChemiDoc XRS+ (Bio-Rad, Hercules, CA) image analysis system.

Compound and IR related targets

The targets of LUP were obtained from the Traditional Chinese Medicine Systems Pharmacology Database and Analysis Platform (TCMSP, http://tcmspw.com/tcmsp.php), Swiss Target Prediction database (http://www.swisstargetprediction.ch/) and BATMAN-TCM (http://bionet.ncpsb.org/batman-tcm/index.php). The genes targets associated with IR were screened using comprehensive database of human genes and gene phenotypes (OMIM, http://www.omim.org/), DrugBank database (https://www.drugbank.ca/) and GeneCards (https://www.genecards.org/).

Construction of ‘component-target-disease’ network

The LUP-related targets and IR-related targets were linked into a ‘component-target-disease’ network, and the interaction between the proteins were introduced into the STRING database (http://string-db.org/). The above network was visualized using Cytoscape ver. 3.7.1 software (http://www.cytoscape.org/).

Biological process analysis and pathway analysis

Gene Ontology (GO) functional annotation and the KEGG pathway analysis were performed to explore related biological processes and signalling pathways.

PPI network analysis and screening of the core targets

Protein–protein interaction (PPI) network analysis was performed using the STRING database. In ‘component-target-disease’ network, there are three key topology parameters in the network including betweenness centrality, closeness centrality and degree, which can be used to screen the more important nodes (Huang et al. 2021). The nodes with degree and betweenness centrality value were two times higher than the median value of all nodes, and the closeness centrality value was greater than the median value of nodes; thus, were chosen as the core targets.

Molecular docking

The *mol2 format of LUP was downloaded at the TCMSP database, and the 3D structure (*PDB format) format of the targets was obtained from the protein data bank (PDB) database (https://www.rcsb.org/). PyMol software was used to perform dehydration and hydrogenation of the proteins, and docking was
Figure 4. Effects of lupenone on synthesis of hepatic glycogen and pathological changes of the pancreas. (A) Stained reddish dots or threadlike structures in the liver cells are considered positive result; rosiglitazone and lupenone increased the synthesis of the hepatic glycogen in rats with diabetes (×100 magnification; n = 6). (B) Acinar cell steatosis (short arrows) and small irregular-shaped islet are observed in the pancreas, rosiglitazone and lupenone protected the structure of the exocrine part of the pancreatic tissues and decreased fat degeneration (×400 magnification; n = 6). (C) Quantitative analysis of the synthesis of hepatic glycogen in rats with T2DM. (D) The number of islets.

Figure 5. Lupenone increased the mRNA expression in the skeletal muscle and adipose tissue related to insulin resistance of rats with diabetes. Lupenone increased the mRNA expression of InsR (a), IRS-1 (b), IRS-2 (c) and GLUT-4 (d) in the skeletal muscle and PPAR-γ (e), IRS-1 (f), IRS-2 (g) and GLUT-4 (h) in the adipose tissue of rats with diabetes. Data are presented as means ± SD. n = 6. *p < 0.05 vs. the T2DM group, #p < 0.05 vs. the normal group; SD: standard deviation.
performed using the AutoDock Vina software. The results were visualized and analysed.

**Statistical analysis**

The data were presented as the mean ± standard deviation (SD). The SPSS 20.0 software (SPSS Inc., Chicago, IL) was used, and the means were assessed using a one-way analysis of variance (ANOVA) and Fisher's protected least significant difference (LSD) post hoc test was used to determine the significant for multiple comparisons. $p < 0.05$ was considered statistically significant.

**Results**

**LUP improves body weight loss and decreases the FBG in rats with T2DM**

LUP inhibited the body weight loss induced by STZ. The results are shown in Figure 1(a). After 2 weeks of treatment, the LUP and rosiglitazone significantly decreased the FBG level ($p < 0.05$). After 3 weeks of treatment, LUP (2.0 and 8.0 mg/kg) decreased the FBG level ($p < 0.05$). After 4 weeks of treatment, LUP (4.0 and 8.0 mg/kg) significantly decreased the FBG level of rats ($p < 0.05$). The results showed that LUP has a stable hypoglycaemic effect in rats with T2DM (Figure 1(b)).

**LUP decreases the levels of LEP, HbA1c, MDA and increases the ADPN levels and GSH-PX activity in rats with T2DM**

After treatment with LUP for 4 weeks, LUP (8.0 mg/kg) and rosiglitazone significantly decreased the levels of LEP in the serum (Figure 2(A)). Additionally, rosiglitazone and LUP (4.0 and 8.0 mg/kg) significantly decreased the serum HbA1c levels in rats with T2DM (Figure 2(C)). Rosiglitazone and LUP (8.0 mg/kg) also significantly decreased the serum MDA levels and increased the serum GSH-PX activity (Figure 2(D,F)). Furthermore, rosiglitazone and LUP (2.0 and 8.0 mg/kg) increased the serum ADPN levels (Figure 2(B)). However, LUP only had a slight effect in the serum SOD activity (Figure 2(E)).

**LUP improves the lipid metabolism disorder in rats with T2DM**

Compared to the normal group, the levels of FFA, TG and TC of the T2DM group were significantly increased, suggesting that the lipid metabolism disorder occurred in the rats with T2DM.
LUP (8.0 mg/kg) significantly decreased the levels of FFA, TG and TC in serum \((p < 0.05)\). In addition, LUP (2.0 and 4.0 mg/kg) significantly decreased the TC levels and LUP (4.0 mg/kg) reduced the levels of TG in serum (Figure 3(A–C)). Furthermore, rosiglitazone and LUP (2.0 mg/kg) significantly reduced the ratio of TC/HDL-C (Figure 3(F)). However, LUP had no significant effect on LDL-C and HDL-C levels in the serum (Figure 3(D,E)).

**LUP increases the synthesis of hepatic glycogen in rats with T2DM**

The liver cell morphology of T2DM group was complete, but the interstitial had hyperplasia. The cell gaps of the hepatocytes were large, cells were irregularly arranged, the intercellular and intracellular PAS staining was low, and the relative homogeneity was poor. However, in the normal, rosiglitazone and the LUP (8.0 mg/kg) groups, the structure of the hepatocytes was tight and neat with no cell gaps. Furthermore, the intercellular and the intracellular PAS staining were diffusely coloured and had good homogeneity. Compared to the T2DM group, the interstitial hyperplasia was less, cells gap was smaller, the cells were arranged regularly, and PAS-positive cells were significantly increased in the LUP (2.0 and 4.0 mg/kg) group. The rosiglitazone and LUP could increase the synthesis of hepatic glycogen in rats with T2DM (Figure 4(A,C)).

**LUP improves the pathological changes of the pancreas in rats with T2DM**

The structure of the exocrine part of the pancreatic tissue in rosiglitazone and LUP (2.0, 4.0 and 8.0 mg/kg) group was
normal, the endocrine part of the pancreatic tissue had a small amount of fatty degeneration and no obvious inflammatory cell infiltration was observed (Figure 4(B)), and 8.0 mg/kg LUP could significantly inhibit the number of islets reduction (Figure 4(D)).

**LUP increases the mRNA and protein expression of InsR, IRS-1, IRS-2, GLUT-4 in skeletal muscle and PPAR-γ, IRS-1, IRS-2, GLUT-4 in the adipose tissue of T2DM rats with T2DM**

The mRNA expression of InsR, IRS-1, IRS-2 and GLUT-4 in the skeletal muscle and PPAR-γ, IRS-1, IRS-2 and GLUT-4 in the...
adipose of rats with T2DM is decreased (Figure 5(A–H)). LUP (4.0 and 8.0 mg/kg) and rosiglitazone significantly increased the expression of InsR, IRS-1, IRS-2 and GLUT-4 mRNA in the skeletal muscle \((p < 0.05)\) and PPAR-\(\gamma\), IRS-1, IRS-2 and GLUT-4 mRNA in the adipose tissue \((p < 0.05)\). In addition, LUP (2.0 mg/kg) significantly increased the expression of IRS-1 mRNA in the skeletal muscle \((p < 0.05)\). In addition, LUP (2.0 and 4.0 mg/kg) significantly increased the expression of IRS-1 and GLUT-4 mRNA in the adipose tissue, and LUP (4.0 mg/kg) significantly increased the expression of PPAR-\(\gamma\) mRNA in the adipose tissue \((p < 0.05)\).

The protein expression of InsR, IRS-1, IRS-2 and GLUT-4 in the skeletal muscle and PPAR-\(\gamma\), IRS-1, IRS-2 and GLUT-4 in the adipose tissue of rats with diabetes is significantly decreased (Figure 6(A–H)). Rosiglitazone significantly increased the expression of IRS-1, IRS-2 and GLUT-4 in the skeletal muscle and PPAR-\(\gamma\), IRS-2 and GLUT-4 in the adipose tissue \((p < 0.05)\). LUP (8.0 mg/kg) significantly increased the protein expression of InsR, IRS-1, IRS-2 and GLUT-4 in the skeletal muscle and PPAR-\(\gamma\), IRS-1, IRS-2 and GLUT-4 in the adipose tissue \((p < 0.05)\). In addition, LUP (4.0 mg/kg) increased the protein expression of PPAR-\(\gamma\) in the adipose tissue \((p < 0.05)\).

Network analysis

An interactive network of LUP to improve IR is constructed and visualized using different colour and shape patterns (Figure 7). Among these, the red triangle is the chemical component, the pink diamond is the disease and the green circle is the common target genes of the component and the disease.

GO functional annotation and KEGG pathway analysis

GO functional annotation was performed. Potential targets (139) were used to perform the biological functional enrichment using the R packages ggplot2 and cluster profiler. A total of 2724 biological processes with a total \(p\) value of \(\leq 0.01\) were enriched. A biological process is most strongly correlated with response to steroid hormone, protein kinase B signalling and gland development, etc. (Figure 8(A)); In the cellular component, there are 110 biological processes with \(p\) value \(\leq 0.01\), including the neuronal cell body, dendritic spine, neuron spine and Golgi lumen, etc. (Figure 8(B)); In the molecular function, there are 150 biological processes with \(p\) value \(\leq 0.01\), among which G protein-coupled receptor binding, histone deacetylase binding and 1-phosphatidylinositol-3-kinase activity are the strongest (Figure 8(C)).

A total of 164 pathways were screened using the KEGG enrichment analysis \((p \leq 0.01)\), and some of the signal pathways are shown in Figure 9. The LUP might improve IR by acting on prostate cancer, endocrine resistance, advanced glycation end products/receptor for advanced glycation end products (AGE-RAGE) signalling pathway in diabetic complications, proteoglycans in cancer, and kaposi sarcoma-associated herpesvirus infection. The targets of the effect of LUP on IR are shown in Figure 10.

PPI network analysis and the core targets results

The targets are introduced into the STRING database, and the specie was ‘Homo sapiens’, and then the interaction network of the protein targets was obtained, as shown in Figure 11. There was a total of 139 nodes and 1357 edges in the network. Additionally, the results of the topological analysis in the

Figure 9. The KEGG pathway analysis of the targets.
network showed that the median value of the betweenness centrality, closeness centrality and degree was 0.000577, 0.534351 and 18, respectively. According to the screening criteria, 24 hub targets of LUP were obtained. The results are shown in Table 2.

**Molecular docking results**
The conformation with the highest score (lowest affinity value) was selected, and the docking conformation was visualized using the PyMol software. The results showed that the LUP had good affinity with the proteins corresponding to the genes including INS, tumour protein p53 (TP53), tumour necrosis factor (TNF), SRC.oestrogen receptor 1 (ESR1) and Jun Proto-Oncogene, AP-1 Transcription Factor Subunit (JUN) (Figure 12). These six genes were the most relevant genes for LUP to improve IR according to the network analysis.

**Discussion**
In this study, we found that LUP could inhibit the body weight loss and decrease the FBG levels. Furthermore, LUP improved the lipid metabolic disorders and reduced the excessive oxidative...
stress in rats with T2DM. Moreover, LUP promoted the synthesis of hepatic glycogen and improved the pathological changes of the pancreas. LUP could improve the IR via regulation of gene and protein expression in rats with T2DM. In addition, network pharmacology analysis revealed that LUP improves IR by multiple targets and signal pathways. These results indicate that LUP has the potential to be developed as a new drug for the treatment of T2DM.

Rats with induced HFD/STZ T2DM are often accompanied by weight loss (Sharma et al. 2011; Wei et al. 2017), but there are conflicting reports on the changes in body weight of rats with T2DM after drug treatment (Gheibi et al. 2017). In this study, LUP could inhibit the body weight loss of rats T2DM. In addition, LUP decreased the LEP level, which is closely related to the appetite, body weight, neuroendocrine functions and glycaemia (Coppari and Bjørbaek 2012). The above results

Figure 11. Interaction network of targets, PPI enrichment $p$ value: $p<1.0e^{-16}$.
suggested that LUP has a good effect on weight maintenance of rats with T2DM. As we know, HbA1c was formally included in the diagnostic index of T2DM by the American Diabetes Association (ADA) in 2010. LUP could reduce the FBG and HbA1c levels in rats with T2DM, and the result is consistent with our previous study (Xu et al. 2014). Furthermore, the IR is closely related to the hepatic glycogen metabolism disorders; LUP could increase the synthesis of the hepatic glycogen; thus, LUP might improve the IR by regulating gluconeogenesis in rats with T2DM.

**Table 2.** Topological analysis of core targets in ‘lupenone-target-disease’ network.

| Gene name | Betweenness centrality | Closeness centrality | Degree |
|-----------|------------------------|----------------------|--------|
| INS       | 0.068121               | 0.756757             | 95     |
| TP53      | 0.02875                | 0.679612             | 74     |
| TNF       | 0.023754               | 0.669856             | 71     |
| SRC       | 0.022054               | 0.660377             | 68     |
| ESR1      | 0.013141               | 0.633484             | 59     |
| JUN       | 0.009126               | 0.625                | 56     |
| PTGS2     | 0.008946               | 0.611354             | 51     |
| HSP90AA1  | 0.01071                | 0.608696             | 50     |
| CTNNB1    | 0.009412               | 0.606061             | 49     |
| MTK       | 0.008296               | 0.606061             | 49     |
| AR         | 0.007686               | 0.598291             | 46     |
| MAPK14    | 0.005711               | 0.595745             | 45     |
| IL10      | 0.006232               | 0.59322              | 44     |
| SIRT1     | 0.005683               | 0.59322              | 44     |
| PIK3CA    | 0.005147               | 0.588235             | 42     |
| IL4       | 0.005161               | 0.585774             | 41     |
| KIT       | 0.004878               | 0.585774             | 41     |
| RELA      | 0.003582               | 0.585774             | 41     |
| HIF1A     | 0.004726               | 0.583333             | 40     |
| EDN1      | 0.004736               | 0.578512             | 38     |
| PIK3R1    | 0.004544               | 0.578512             | 38     |
| MDM2      | 0.004322               | 0.578512             | 38     |
| AGT       | 0.006154               | 0.576132             | 37     |
| CRP       | 0.005963               | 0.576132             | 37     |

**Figure 12.** Molecular docking results of lupenone and the corresponding protein of the gene targets.
Lipid metabolism disorders are closely related to the cardiovascular diseases and T2DM (Loza-Rodriguez et al. 2020), and TC/HDL-C ratio. TC/HDL-C ratio is a reasonable blood lipid index in predicting the cardiovascular risk of patients with T2DM. Our results showed that LUP significantly decreases the levels of TC, TG and TC/HDL-C ratio, which indicates that LUP can improve the lipid metabolism disorders and may reduce cardiovascular risk patients with T2DM.

The oxidative stress caused by obesity, hyperglycaemia, FFA and inflammation could lead to the islet \( \beta \) cell damage (Halim and Halim 2019). As reported, long-term exposure to high concentrations of FFA will damage the islet \( \beta \) cell function that reduces the secretion of insulin, increased the gluconeogenesis, and eventually lead to IR (Zhande et al. 2002). Moreover, ADPN can increase the muscle and liver sensitivity to insulin by increasing the FFA oxidation (Wang et al. 2009); hence, ADPN can increase the muscle and liver sensitivity to insulin by regulating IR in the skeletal muscle and adipose tissue of patients with T2DM. Our results showed that LUP significantly decreases the level is an important marker for improvement of IR (Yukihiro et al. 2001). LUP increased the LD/HDL-C increase in rats with T2DM. This study indicates that LUP could improve the IR in rats with T2DM. Furthermore, levels of SOD, MDA and GSH-PX are closely related to the islet \( \beta \) cells damage (Coskun et al. 2005; Sun et al. 2007). LUP increased the GSH-PX activity and decreased MDA levels in rats with T2DM. Interestingly, LUP also inhibited the mild inflammatory response in the pancreatic tissue. Taken together, LUP might protect the patient with T2DM via inhibiting the inflammation, and the result is consistent with our previous study (Xu et al. 2020). IGF-1/insulin receptor (IR) and IRS-1/insulin receptor substrate-1 (IRS-1) are important receptors for signal transduction in the insulin pathway, which are closely related to the secretion of insulin. The results in this study showed that LUP could increase the gene and protein expression of IGF-1 and IRS-1 in the skeletal muscle and adipose tissue. Network pharmacology analysis results showed that LUP can improve IR by acting on PI3K (Figure 10). These results indicate that LUP might increase the sensitivity of the skeletal muscle and adipose tissue to insulin, at least partly, by activating the IGF-1/PI3K/GLUT-4 signalling pathway. According to the report, the expression of IRS-1 in the skeletal muscle and adipose tissue of patients with T2DM is significantly decreased (Ducluzeau et al. 2001; Hammarstedt et al. 2003). Both IRS-1 and IRS-2 function in the peripheral carbohydrate metabolism, and IRS-2 has the major role in the \( \beta \)-cell development and compensation for peripheral IR (Withers et al. 1999). In the present study, LUP up-regulated the IGF-1 and IRS-2 function in the peripheral carbohydrate metabolism, and IRS-2 has the major role in the \( \beta \)-cell development and compensation for peripheral IR (Withers et al. 1999). In the present study, LUP up-regulated the gene and protein expression of IRS-1 and IRS-2 in the skeletal muscle and adipose tissue, suggesting that LUP might improve the IR via acting on the family of insulin substrates in T2DM. In addition, peroxisome proliferator-activated receptor gamma (PPAR-\( \gamma \)) is known to play a major role in the improvement of insulin sensitivity through stimulating differentiation of small adipocytes from preadipocyte (Kadowaki et al. 2003). Here, LUP might improve the IR by up-regulating the expression of PPAR-\( \gamma \). Therefore, the above results showed that LUP improves the skeletal muscle and adipose tissue sensitivity to insulin through increasing the gene and protein transcription related to IR in T2DM.

Combining the drug-target network with the biological system network can provide new approaches and strategies for new drug development (Hopkins 2007). Network pharmacological studies have found that LUP improves IR through multiple targets and multiple pathways. The biological process is mainly related to the regulation of cell proliferation, signal transduction, metabolism and cell differentiation. The results suggested that LUP could improve IR by regulating multiple biological processes. The KEGG pathway analysis also showed that LUP improves IR involving multiple signalling pathways, like AGE-RAGE signalling pathway in diabetic complications, IR and type II diabetes mellitus. AGEs play a significant role in the pathogenesis of multiple diabetic complications (Sergi et al. 2021). These results indicated that LUP might be used for the prevention and treatment of T2DM-associated complications. Of course, many signalling pathways related to cancer, tumour, atherosclerosis and infection were predicted, indicating that LUP might have various modes of pharmacological activities, which deserves further research.

Conclusions

LUP could improve the lipid metabolism disorder and alleviate the excessive oxidative stress in T2DM. Furthermore, as expected, LUP improved the IR via increasing the sensitivity of the skeletal muscle and adipose tissue to insulin by regulating related gene and protein expression in T2DM. Our preliminary results indicate that LUP has the potential to be developed as a new drug for the treatment of T2DM.

Authors contributions

Feng Xu and Mei Zhang contributed equally to this work. Xianggei Wang, Feng Xu, and Hongmei Wu contributed to the study conception and design, acquisition, analysis and interpretation of data, and writing of the article. Mei Zhang, Yuanmin Wang and Ye Yang contributed to the acquisition and interpretation of data and to the final critical revision of the article. All authors read and approved the manuscript.

Disclosure statement

No potential conflict of interest was reported by the author(s).

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