Can we find a good biochemical marker of early cardiotoxicity in children treated with haematopoietic stem cell transplantation?

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Introduction

Haematopoietic stem cell transplantation (HSCT) is widely used in the treatment of malignant and non-malignant disorders in paediatric patients. The procedure can be associated with severe toxicities, including cardiotoxicity [1]. Toxicities may be connected with radio- and chemotherapy used in previous treatment as well as in conditioning regimens and other potentially serious complications that may appear in the early post-transplant period. There are many different methods used to assess cardiac failure: echocardiography (ECHO), electrocardiography, radionuclide ventriculography, or biochemical markers [2–9]. Echocardiography is one of the most commonly used non-invasive methods for imaging cardiac structure and function. Standard M-mode echocardiography allows the evaluation of motion of cardiac structures and provides accurate assessment of left ventricular systolic and diastolic parameters, which define cardiac efficiency [4]. Biochemical markers may be useful for early detection of cardiotoxicity. Atrial natriuretic peptide (ANP) is secreted by the atria in response to increased stretch as a result of an increased left atrial pressure. N-terminal fragment of brain natriuretic peptide (NT-proBNP) is produced by ventricles in response to ventricular dilatation and increased wall stress. The levels of natriuretic peptides are inversely correlated with cardiac function [2, 7, 10–13]. Cardiac isoforms of troponin I and T (TnI, TnT) are released into the circulation rapidly after myocardial injury and may be sensitive markers of cardiac necrosis [10, 14]. Endothelin 1 (ET-1) is a vasoconstrictor peptide synthesised in the vasculature and by the endothelial cells. Elevated plasma levels of ET-1 have been reported in association with heart failure [11, 15].

The aim of the study was to assess the incidence of cardiac complications and the significance of determination of NT-proBNP, ANP, ET-1, and TnI plasma levels in the early post-transplant period in children treated with HSCT.

Material and methods

Patients

A total of 30 consecutive children (22 boys and 8 girls) with a median age of 9.6 years (range 0.2–18), in which 31 transplants were performed, were included in the study. The control group consisted of 14 healthy children with a median age of 10.9 years (range 1.6–17 years). In the analysed group 12/31 (38.7%) autologous (auto-HSCT), 8/31 (25.8%) matched sibling donors (MSD), and 11/31 (35.5%) alternative transplantations (ALT HSCT: 9 matched unrelat-
ed donor – MUD and 2 mismatched family donor – MMFD) were performed. 23/30 (76.7%) patients were transplanted because of malignant diseases; 11 had acute leukaemias (ALL-4, AML-7); 9 – solid tumours (neuroblastoma – 2, Ewing sarcoma – 4, rhabdomyosarcoma – 2, germ cell tumour – 1) and 3 children were treated for lymphomas. Seven transplantations (23.3%) were performed in children with non-malignant disorders (severe aplastic anaemia – 4, myelodysplastic syndrome – 1, Wiscott-Aldrich Syndrome – 1, Omenn syndrome – 1, adrenoleukodystrophy – 1). Conditioning regimens differed according to the diagnosis and are presented in Table 1.

Patients transplanted for malignant diseases were previously treated with anthracyclines, and standard chemotherapy protocols, in a median dose of 240 mg/m² (180–360 mg/m²). All patients were managed according to the institutional protocol for HSCT. Fluid balance was carefully monitored and compensated with fluids or diuretics as clinically indicated. Patients undergoing allogeneic HSCT received graft versus host disease (GvHD) prophylaxis with cyclosporine +/- short motretexate. Treatment of aGvHD followed EBMT guidelines [16].

Methods

The plasma levels of biochemical markers: NT-proBNP, ANP, TnI, and ET-1 were assessed immediately in the Central Lab by Immunoassay (Abbot). Blood samples for assessing NT-proBNP, ANP, and ET-1 were collected from the patients on the analysed days (on days –7, +7, +14, and +21). The values of serum cardiac troponin I were assessed immediately in the Central Lab by Immunoassay (Abbot). Blood samples for assessing NT-proBNP, ANP, and ET-1 were collected from the patients on the analysed days then immediately centrifuged and separated and the plasma was stored at –80°C until the analysis was performed. The samples were thawed and analyzed at the same time. The values of NT-proBNP, ANP, and ET-1 were estimated by Electrochemiluminescent Immunoassay (Biomedica). Left ventricular systolic parameters [fractional shortening (FS) and ejection fraction (EF)] and diastolic parameters [early peak flow velocity/atrial velocity (E/A ratio) and isovolumic relaxation time (IVRT)] were measured using M-mode standard echocardiography prior to HSCT and approximately day +30 and +100 after transplantation. The same biochemical markers, as well as echocardiographic parameters, were assessed once in the control group.

Statistical analysis was performed using computer software Statistica 10.0 Stat Soft Inc. (USA). The statistical difference between biochemical marker values in patients and controls was compared using non-parametric Mann-Whitney U test. For the comparison of the values of chosen parameters in more than two groups the Kruskal-Wallis test was used. The correlation between assessed parameters was estimated using Pearson’s test. The statistical significance level of p < 0.05 was assumed.

The study was approved by the Ethical Committee of the Medical University of Lublin, Poland.

Results

None of the children from the analysed group developed clinical cardiotoxicity in the post-transplant period.

Baseline echocardiographic systolic parameters were within the normal range in all patients included in the study (median FS – 40.2%, median EF – 73.3%). Concerning diastolic parameters, the median E/A ratio assessed before HSCT was statistically lower in the transplanted patients (1.34 vs. 1.73, p < 0.05) as well as median E/A on day +30 (1.37 vs. 1.73, p < 0.05) and on day +100 (1.42 vs. 1.73, p < 0.05) post-transplant. Median SF and EF values on day +30 in transplanted children and in the control group did not differ significantly. The results are presented in Table 2.

Median ET-1 values in the transplanted children were significantly higher than estimated in controls in all analysed time points, and median NT-proBNP values differed significantly at three analysed time points: before HSCT (day –7) and on days +7 and +14 post-transplant. Comparing median ANP values in transplanted patients and controls, a statistically significant difference was found only on day +14 post-transplant. No difference was found between the median TnI values in all analysed time points. Median values of all analysed biochemical parameters (ET-1, ANP, NT-proBNP, TNI) are presented in Table 3.

Table 1. Conditioning regimens in transplanted patients according to the type of transplant and diagnosis

| Type of transplant | n (%) | Diagnosis (n) | Conditioning regimens (n) |
|--------------------|-------|---------------|--------------------------|
| Auto n = 12 (38.7%) | RMS (2), GCT (1) | Mel, Eto, Carbo (3) |
| Auto  alternative | Sa Ewing (4), AML (1), NBL (2) | ALL, AML | TBI, Eto (1), Treo (1) |
| Allo MSD n = 8 (25.8%) | lymphoma | TBI, Thiost, Eto (2) |
| Allo alternative (MUD, MMFD) n = 11 (35.5%) | ALL (1), lymphoma (1) | TBI, Eto (2) |
| | AML | TBI, Flu (1) |
| | AML | Treo, Cy, Mel (3) |
| | SAA | Flu, Cy (2) |
| | SAA II | TBI (1) |
| | MDS | Bu, Cy, Mel (1) |
| Ommens syndrome | MUD | Flu, Mel (1) |

Auto – autologous transplantation; Allo alternative – transplantation from alternative donors; MSD – matched sibling donor; MUD – matched unrelated donor; MMFD – mismatched family donor; TBI – total body irradiation; Thiost – thiotepa; Eto – etoposide; Flu – fludarabine; Bu – busulfan; Mel – melphalan; Treo – treosulfan; CY – cyclophosphamide; Carbo – carboplatinum; AML – acute myeloblastic leukaemia; ALL – acute lymphoblastic leukaemia; SAA – severe aplastic anaemia; Sa Ewing – Ewing sarcoma; NBL = neuroblastoma; WAS – Wiscott-Aldrich syndrome; ALD-X – adrenoleukodystrophy; RMS – rhabdomyosarcoma; GCT – germ cell tumour
Since median ET-1 and NT-proBNP plasma levels were elevated in at least 3 out of 4 analysed time points in the transplanted patients when compared to controls, the analysis was extended according to the type of transplant. No difference was found in median ET-1 and NT-proBNP values depending on the particular type of transplant when compared with each other, while median EF-1 and NT-proBNP values in auto, MSD, and especially in alternative transplant recipients differed significantly in most analysed time points when compared to controls. Analyses and values are presented in Table 4.

Dependences between median biochemical marker values and echocardiographic systolic and diastolic parameters were analysed. No correlation was found, except for median ET-1 level assessed on day +14 and SF and EF assessed on day +30 (r = −0.374 and −0.446, respectively) and on day +100 (r = −0.420 and −0.477, respectively).

### Table 2. Echocardiographic systolic and diastolic parameters at analysed time points in patients and controls

| ECHO parameter | Time point | Analysed patients n = 31 | Controls n = 14 | p-value |
|----------------|------------|---------------------------|----------------|---------|
|                | Median (range) | Median (range) |               |         |
| LVSF (%)       | Before HSCT | 40.2 (28.6–50.5) | 40.85 (36.0–57.2) | NS |
| LVEF (%)       | Before HSCT | 73.3 (55.5–82.0) | 71.0 (67.1–83.7) | NS |
| LVSF (%)       | Day +30    | 37.0 (27.9–49.5) | 40.85 (36.0–57.2) | NS |
| LVEF (%)       | Day +30    | 69.2 (55.2–82.0) | 71.0 (67.1–83.7) | NS |
| LVSF (%)       | Day +100   | 42.1 (30.3–51.2) | 40.85 (36.0–57.2) | NS |
| LVEF (%)       | Day +100   | 74.1 (60.0–83.7) | 71.0 (67.1–83.7) | NS |
| E/A            | Before HSCT | 1.34 (0.76–2.62) | 1.73 (1.47–2.57) | 0.00015 |
| IVRT (ms)      | Before HSCT | 63 (49–83) | 65 (50–77) | NS |
| E/A            | Day +30    | 1.37 (0.76–2.62) | 1.73 (1.47–2.57) | 0.0005 |
| IVRT (ms)      | Day +30    | 63 (48–83) | 65 (50–77) | NS |
| E/A            | Day +100   | 1.42 (1.26–1.72) | 1.73 (1.47–2.57) | 0.0002 |
| IVRT (ms)      | Day +100   | 62 (47–82) | 65 (50–77) | NS |

Significant p value < 0.05

HSCT – haematopoietic stem cell transplantation; LVEF – left ventricular ejection fraction; LVSF – left ventricular shortening fraction; E/A ratio – early peak flow velocity/atrial velocity; IVRT – isovolumic relaxation time

### Table 3. Biochemical parameters at analysed time points in patients and controls

| Biochemical parameter | Time point | Analysed patients n = 31 | Controls n = 14 | p-value |
|-----------------------|------------|---------------------------|----------------|---------|
|                       | Median (range) | Median (range) |               |         |
| ET-1 [fmol/ml]        | –7         | 0.43 (0.07–2817) | 0.22 (0.09–0.63) | 0.002 |
|                       | +7         | 0.52 (0.15–424) | < 0.001 |         |
|                       | +14        | 0.40 (0.07–1481) | 0.001 |         |
|                       | +21        | 0.48 (0.49–1635) | 0.005 |         |
| NT-proBNP [fmol/ml]   | –7         | 13.27 (0.55–128.8) | 3.31 (0.74–19.06) | 0.016 |
|                       | +7         | 10.3 (0.78–107.4) | 0.015 |         |
|                       | +14        | 12.39 (0.63–564.2) | 0.006 |         |
|                       | +21        | 7.78 (0.35–85.9) | NS |         |
| ANP [fmol/l]          | –7         | 1110 (280–5001) | 1064 (298–2705) | NS |
|                       | +7         | 1287 (307–6589) | NS |         |
|                       | +14        | 1700 (101–7325) | 0.022 |         |
|                       | +21        | 1203 (266–7310) | NS |         |
| TnI [ng/ml]           | –7         | < 0.01 (0.00–2.14) | 0.01 (0.0001–0.02) | NS |
|                       | +7         | < 0.01 (0.00–1.64) | NS |         |
|                       | +14        | < 0.01 (0.00–1.11) | NS |         |
|                       | +21        | < 0.01 (0.00–0.07) | NS |         |

Significant p-value < 0.05

ET-1 – endothelin 1; NT-proBNP – brain natriuretic peptide; ANP – atrial natriuretic peptide; TnI – troponin I
Table 4. The median values of endothelin-1 and NT-pro BNP at all analysed time points according to the type of transplant

| Parameter     | Time point | Type of transplant | Controls | p-value |
|---------------|------------|--------------------|----------|---------|
| Endothelin-1 (fmol/ml) median, (range) |  | Auto | Allo MUD, MMFD | Allo MSD |         |
| −7 | 0.54 (0.07–1.82) | 0.31 (0.15–28.15) | 0.43 (0.21–1.12) | 0.22 (0.09–0.63) | 0.03 |
| +7 | 0.48 (0.22–1.15) | 0.52 (0.15–42.4) | 0.49 (0.25–11.12) | 0.03 |
| +14 | 0.49 (0.26–2.5) | 0.40 (0.07–1481) | 0.39 (0.15–10.39) | 0.007 |
| +21 | 0.49 (0.199–1.71) | 0.58 (0.05–1635) | 0.399 (0.21–1.12) | NS |
| NT-proBNP (fmol/ml) median, (range) |  |  |  |  |  |  |  |  |  |
| −7 | 6.49 (0.65–63.69) | 16.06 (0.55–128.8) | 12.78 (3.17–59.77) | 3.31 (0.74–19.06) | 0.0007 |
| +7 | 4.13 (0.78–67.22) | 59.35 (6.35–107.4) | 16.86 (0.99–64.12) | 0.22 |
| +14 | 7.93 (0.64–32.97) | 24.26 (7.38–564.2) | 14.45 (4.26–46.65) | 0.008 |
| +21 | 7.57 (0.15–17.08) | 10.58 (2.32–85.9) | 6.37 (2.13–19.6) | 0.26 |

Auto – autologous transplantation; MUD – matched unrelated donor; MMFD – mismatched family donor; MSD – matched sibling donor

In 9 out of 19 allogeneic transplant recipients (47.4%) aGvHD was diagnosed (1st grade in two patients, 2nd grade in six patients, and 3rd grade in one patient). No difference in echocardiographic parameters and in biochemical marker levels was found between patients with and without aGvHD. None of the patients developed acute kidney injury. Of all the analysed group, 10 patients died (33.3%): seven because of disease relapse/progression, one patient developed multiorgan failure, one because of graft rejection, and one because of infectious complications.

Discussion

Clinically significant cardiotoxicity after HSCT has been estimated in about 5% of adult patients [7]. In patients following HSCT, myocardial function can be impaired by cardiotoxic drugs used in conditioning regimens, which can cause toxic endothelial damage [3, 5]. Early diagnosis of cardiac failure in patients following HSCT may be difficult because they may have multiple medical problems: veno-occlusive disease (VOD), acute renal failure, GvHD, infections [17]. Heart failure may also be due to the use of hyperhydration regimens, transfused blood components, impaired renal function, sepsis, or electrolyte abnormalities [18]. Previous treatment of malignancy with chemotherapeutic agents, including anthracyclines, may explain our results, in which the median E/A ratio was lower and median ET-1 and NT-proBNP values were elevated before HSCT in the analysed group. NT-proBNP elevations may reflect chemotherapy-associated diastolic abnormalities, and the level may be increased by myocardial stress without necrosis. ET-1 is a biomarker of vascular injury and is increased in many conditions in which vascular endothelium is vulnerable. Endothelial cells upregulate ET-1 secretion in response to hypoxia, oxidized LDL, pro-inflammatory cytokines, and bacterial toxins [9, 12]. Several studies confirm the usefulness of ET-1 and NT-proBNP as predictors of chemotherapy related cardiotoxicity [14, 19–24]. The study by Hakayawa, performed on a group of 34 children receiving doxorubicin-containing chemotherapy, showed natriuretic peptides to be useful markers of cardiotoxicity in children [19]. In studies by Nouisiainen, conducted in adult patients with Hodgkin’s lymphoma receiving high doses of doxorubicin (400–500 mg/m²), increased concentrations of plasma natriuretic peptides were found [23, 24]. ET-1 may also contribute to the prediction of doxorubicin-induced cardiotoxicity, which was shown in Yamashita’s study, performed on patients with breast and lung cancer, in which ET-1 plasma concentrations rose progressively during doxorubicin treatment [25]. Measurement of natriuretic peptides plasma levels has already been used to monitor cardiotoxicity following HSCT. Snowden confirmed the utility of plasma BNP assessment as a ventricular dysfunction indicator in adult patients [2]. Kultinnen studied patients with non-Hodgkin lymphoma and observed symptoms of cardiotoxicity diagnosed after conditioning treatment, and a correlation was found between the changes in NT-proBNP levels and LVEF; and more strongly with diastolic function parameters [18]. Morandi et al., in their study, concluded that use of high-dose cyclophosphamide, but also cytarabine, mitoxantrone, melphalan, and fludarabine may induce cardiac injury [7]. In our analysis median NT-proBNP levels were statistically higher on days +7 and +14 post-transplant, as well as when assessed before HSCT. According to our study, only plasma levels of ET-1 assessed in the early post-transplant period inversely correlated with systolic echocardiographic parameters.

Our results concerning the diagnostic values of ANP and TnI as predictors of cardiotoxicity do not confirm their usefulness. In previous studies, e.g. in the Herman et al. study on rats, in which doxorubicin was given, troponin was proven to be useful in early detection of cardiotoxicity [14]. In our study no difference was found between the median TnI values at all analysed time points in transplanted patients compared to controls. The results are similar to those obtained by Mavinkurve-Grothus et al. in their analysis conducted on childhood cancer survivors, in which abnormal troponin levels were not found in any of the patients [21]. Also in the study by Romano et al., performed on 34 breast cancer patients, TnI was abnormal only occasionally in four of them, and in the retrospective Sherief et al. study none of the analysed survivors of acute leukaemia had abnormal troponin levels [20, 22].

In our cohort of patients, clinical symptoms of cardiac dysfunction were not observed, but abnormalities in biochemical marker levels, especially ET-1 and NT-proBNP, as well as in diastolic echocardiographic parameters, were found. This confirms the need for careful follow-up in pa-
tients who have received chemotherapy and have been treated with haematopoietic stem cell transplantation.

The authors declare no conflict of interest.

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