A new species of *Longitarsus* Latreille, 1829 (Coleoptera, Chrysomelidae, Chrysomelidae) pupating inside stem aerenchyma of the hydrophyte host from the Oriental Region

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Abstract

A new species of subaquatic *Longitarsus* pupating inside the stem aerenchyma of its hydrophyte host plant is described. Eggs are laid on tender leaves and buds and the larvae are open feeders. This is the first report of an Oriental flea beetle pupating inside the stem of its hydrophyte host. A key to the species of southern Indian *Longitarsus* is provided.

Keywords

Chrysomelidae, subaquatic *Longitarsus*, new species, key, stem aerenchyma pupation, *Limnophila*

Introduction

Larvae of flea beetles, in general, are subterranean root feeders. A few of them mine the leaves or feed exposed on it, while fruit borers and stem borers are extremely rare (Jolivet and Hawkeswood 1995, Konstantinov and Vandenberg 1996). As a rule,
flea beetle larvae associated with terrestrial plants, including leaf feeders, pupate in soil or litter. However, root and soil under submergence are inaccessible to all life stages of flea beetles due to lack of adaptation for a true aquatic life. Still a few of them have adopted strategies to circumvent this limitation to harness aquatic and subaquatic plants. Jolivet (2003) reviewed the biology of subaquatic Chrysomelidae, including subaquatic flea beetles. In many such flea beetles, larvae feed on leaves and pupation occurs in the soil on shore or facultatively on the plant itself when soil is not available. In the case of beetles living on rooted emergents and floating plants, inaccessibility to soil force all life stages, including pupae, to adapt to the portions of the plant above water level. For example, *Pseudolampsis guttata* (LeConte) that feeds on the floating water fern *Azolla caroliniana* Willd. pupates on leaf surface (Buckingham and Buckingham 1981). Species of *Agasicles* Jacoby, 1904 are unique among the subaquatic flea beetles as they pupate only inside the stem internode cavity of their amaranthaceous hosts (Vogt et al. 1979). A pair of modified T-shaped urogomphi secures the pupa from falling into the wedged position inside the internode cavity, from which emergence might be impossible (Vogt et al. 1979; Cox 1998). A new species of *Longitarsus* Latreille, 1829 from the Oriental Region, similarly adapted for pupation in stem aerenchyma is described here. Aerenchyma is a large intercellular space that acts as a mediator of internal gas exchange and maintains strength with the least tissue (Jung et al. 2008).

The cosmopolitan *Longitarsus* is the most speciose genus of flea beetles with about 700 species. Though about 100 species are known from the Oriental Region, only eight named species of *Longitarsus* occur in south India (Maulik 1926; Scherer 1969; Gruev and Askevold 1988; Prathapan et al. 2005) and several of them still await naming and description. Known host plants of the genus belong to at least a dozen families with marked preference for members of Boraginaceae, Lamiaceae, Asteraceae, and Verbenaceae (Jolivet and Hawkeswood 1995). Species of *Longitarsus* are small to medium sized flea beetles easily identified by a very long first metatarsomere which is at least half as long as the metatibia. Other salient features of the genus are the lack of transverse or longitudinal impressions on the pronotum, open procoxal cavities and the confused elytral punctures rarely forming regular striae. Members of *Longitarsus* are generally terrestrial and their larvae feed on the roots (Furth 1980, Ireson et al. 1991, Schwarzländer 2000, Baars 2001 and Simelane 2010). Exceptions include the European *Longitarsus nigerrimus* (Gryllenhal) that lives in peat bogs and swamps. Booth (2000) reviewed its records from the British Isles and Jolivet (2003) summarized information on its biology.

**Methods**

Natural populations of the beetle under field conditions were observed during 2009–2011 at Vembayam, Trivandrum, Kerala, India during several visits. The host plant, *Limnophila aquatica* (Roxb.) Alston was grown in a concrete tank partially filled with
soil and water at Vellayani and live beetles were released on to these plants to confirm the biology observed in the field.

Descriptive terminology follows Konstantinov (1998). The holotype of the new species is deposited in the Natural History Museum, London (BMNH). Paratypes will be deposited in the National Pusa Collection, Indian Agricultural Research Institute, New Delhi (NPC), University of Agricultural Sciences, Bangalore (UASB), National Museum of Natural History, Smithsonian Institution, Washington DC (USNM), and in the personal collection of the first author (PKDC). Plant vouchers are deposited in the Calicut University Herbarium (Accession nos 6426, 6427, 113059, 113060). The immature stages are being retained by the first author for further studies.

Systematics

Longitarsus limnophilae Prathapan & Viraktamath, sp. n.
urn:lsid:zoobank.org:act:A24AE7BC-6B6D-4880-994E-BB8D05667079
Figs 1–8, 12–15

Holotype ♂, with labels as follows: “INDIA Kerala / Vembayam / 12. ix. 2009 Prathapan Coll.” “Longitarsus limnophilae sp. nov. / Prathapan & Viraktamath” “HOL-OTYPE [red printed label]” (BMNH).

Paratypes (30 specimens): 7 ♂, 3 ♀. The same labels as holotype; 5 ♀. same data as for holotype except dating 3.x.2009; 5 ♂. same data except dating 24.x.2009; 9 ♂, 1 ♀. same data except dating 16.i.2010 (5 BMNH, 5 USNM, 5 UASB, 12 NPC, 3 PKDC).

Etymology. This unique species is named after its host plant. The name is a noun in the genitive case.

Description. Length 1.89 – 2.15 mm; width 0.91 – 1.08 mm; female (2.09 – 2.15 mm) slightly larger than male (1.89 – 2.12 mm). General color brown (Fig. 1). Fore- and middle legs, hind tibia and tarsi light brown. Antenna piceous with proximal three to five antennomeres gradually turning brown. Labrum dark brown to piceous, suture narrowly piceous. Ventrites lighter than dorsum.

Vertex shiny, impunctate, minutely wrinkled. Ommatidia fully developed. Postcallinal sulcus weak but distinct. Frontal ridge unusual, broad and not sharply raised, anteriorly widening towards frontoclypeal suture, anteriorly forming ill-defined denticle in middle of flat, poorly developed anterofrontal ridge. Maxillary palpus with last palpomere longest. Antenna extends well beyond apex of elytra over pronotum. Second antennomere longer than half of third; second and third together longer than first, subequal to fourth; fifth longer than fourth; fifth to seventh subequal, eighth to tenth progressively shorter than previous antennomere. Pronotum anteriorly wider than posteriorly; 1.27 – 1.34 times wider than long; anterolateral callosity posteriorly lower than anteriorly, not forming denticle at pore; posterolateral callosity protrudes beyond lateral margin; lateral margin weakly curved,
Figure 1. *Longitarsus limnophilae* sp. n., dorsal habitus

anteriorly broader than posteriorly; disc shiny with minute punctures more evident posteriorly. Elytra with well developed humeral calli, punctures distinct, width of interstices smaller than diameter of one puncture in middle of elytron. Elytral apex convex, with one long seta. Hind wings well developed. Scutellum triangular. First male protarsomere 1.60 – 1.67 times longer than wide; first female protarsomere 2.00 – 2.43 times longer than wide. Metatibia strongly curved in dorsal view, slightly curved in lateral view. Number of spinules on dorsolateral margin of metatibia, proximal to row of stiff bristles, vary from seven to ten. In lateral view, first metatarsomere 0.55 – 0.57 times as long as metatibia. Proximal end of first metatarsomere ventrally with thick characteristic patch of short pointed and capitate setae in both sexes. Last male ventrite internally with longitudinal ridge along middle (Fig. 5); posterior margin bisinuate.

Aedeagus in lateral view gently curved, apex acutely pointed and slightly recurved (Fig. 4); ventral side depressed with transparent window, lateral edges raised (Fig. 2); dorsal opening covered with lamina not extending to apex (Fig. 3). Arms of tegmen subequal to or slightly shorter than stem.

Spermatheca with receptacle widest in middle, internal side strongly convex, external side weakly concave; pump with horizontal part longer than vertical; spermathecal duct curved towards receptacle, coiled thrice proximally, not reaching half of receptacle...
A new species of Longitarsus Latreille, 1829 (Coleoptera: Chrysomelidae: Galerucinae)...

Figures 2–8. *Longitarsus limnophilae* sp. n. 2 median lobe of aedeagus, ventral view; 3 median lobe of aedeagus, dorsal view; 4 median lobe of aedeagus, lateral view; 5 last abdominal ventrite of male (macerated specimen); 6 spermatheca; 7 tignum; 8 vaginal palpi.
Figures 9–12. *Longitarsus limnophilae* sp. n. 9 habitat; 10 *Limnophila aquatica*; 11 *L. repens*; 12 adult feeding scars on leaf.
A new species of *Longitarsus* Latreille, 1829 (Coleoptera: Chrysomelidae: Galerucinae)...

Figures 13–16. *Longitarsus limnophilae* sp. n. 13 egg; 14 larva; 15, 16 pupa inside stem aerenchyma.

(Vig. 6). Vaginal palpus narrow at distal 1/3, widest at proximal 1/4; distal sclerotization shorter than proximal sclerotization or lateral membranous area (Fig. 8). Tignum nearly straight, anterior sclerotization slightly wider than posterior (Fig. 7).

**Remarks.** *Longitarsus limnophilae* can easily be separated from all other south Indian species of *Longitarsus* by the anteriorly widening, flat frontal ridge (in the other species, the frontal ridge is more or less narrowly raised). *Longitarsus belgaumensis* Jacoby closely resembles *Longitarsus limnophilae* in having narrowly piceous elytral suture and dark distal antennomeres. But *Longitarsus limnophilae* can be separated from *Longitarsus belgaumensis* based on the antenna extending slightly beyond the apex of elytra over pronotum (in *Longitarsus belgaumensis*, antenna does not extend beyond the apex of elytra over pronotum), pronotum anteriorly wider than posteriorly (in *Longitarsus belgaumensis*, pronotum is an-
teriorly as wide as posteriorly with the maximum width in the middle), structure of frontal ridge (frontal ridge sharply raised along middle in *Longitarsus belgaumensis*) and genitalia.

**Host plants.** *Limnophila aquatica* (Roxb.) Alston (Scrophulariaceae) (Fig. 10) is a rooted emergent hydrophyte growing in shallow streams, marshes and rice fields (Fig. 9). Species of *Limnophila* R. Br., 1810 are widely distributed in the tropics and subtropics of the Old World and also occur as weeds. Stem aerenchyma in *L. aquatica* resembles the same in *L. sessiliflora* (Vahl) Blume termed as “wheel-type” by Jung et al. (2008). *Longitarsus limnophilae* was also found to feed on *L. repens* (Benth.) Benth. (Fig. 11).

**Biology.** Eggs (Fig. 13) are laid on tender leaves and buds and the larvae are open feeders (Fig. 14). The closely oriented tender leaves provide sufficient cover for the larva. Mature larva enters the stem aerenchyma in the internode by boring a tiny hole and pupation occurs in it, a little above the entry hole (Figs 15, 16). Adult emerges through an exit hole nearly circular in shape with irregular margin. Adult feeds on both adaxial and abaxial surface of, mostly, tender leaves by scraping, often resulting in holes on the lamina (Fig. 12). Adults when thrown in water floated initially and then swam with raised antennae held back over the sides of the pronotum. After swimming for a while, some performed a short jump on to the shore.

**Distribution.** The types were collected from a single locality only. India (Kerala, Vembayam, 8°38′28″N; 76°56′39″E).

**Discussion.** Jolivet and Hawkeswood (1995) have listed *Bacopa* Aublet, 1775 (Scrophulariaceae), a genus of hydrophytes, among host plants of *Longitarsus*. But no further information is available on its biology on *Bacopa*. *Longitarsus limnophilae* and species of *Agasicles* represent two independently evolved lineages in Alticini adapted for larval leaf feeding and pupation inside the stem. *Agasicles* and *Longitarsus limnophilae* are the only flea beetles known to pupate inside the stem of their aquatic host plant above the water level. Wheel shaped stem aerenchyma of *Limnophila aquatica* serve as a safe abode for the pupa offering protection against natural enemies. Species of *Limnophila* being widely distributed aquatic weeds, *Longitarsus limnophilae* could be a potential biocontrol agent for them.

This is the first report of an Oriental flea beetle pupating inside the stem of its hydrophyte host.

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**Key to the southern Indian species of *Longitarsus***

1. Vertex with conspicuous deep punctures..................*L. rangoonensis* Jacoby
   – Vertex impunctate to minutely punctuate .......................................................2

2. Elytra laterally with prominent longitudinal ridge extending backwards from the humerus.................................................................*L. liratus* Maulik
   – Elytra laterally without longitudinal ridge extending from humerus........3

3. Metatibial spur minutely serrulated on either side dorsally....................
   ...............................................................*L. serrulatus* Prathapan, Faizal & Anith
   – Metatibial spur not serrulated..........................................................4
A new species of *Longitarsus* Latreille, 1829 (Coleoptera: Chrysomelidae: Galerucinae)...

4 Elytral suture narrowly piceous compared to rest of elytra..........................5
- Elytral suture not distinctly darker than rest of elytra...............................6
5 Frontal ridge broad, widening towards frontoclypeal suture, not sharply
raised; antenna extends well beyond apex of elytra over pronotum..............
L. limnophilae Prathapan & Viraktamath, sp. n.
- Frontal ridge sharply raised, narrow; antenna hardly reaches apex of elytra
over pronotum ........................................................................... L. belgaumensis Jacoby
6 Dorsum red ............................................................ L. rufipennis Jacoby
- Dorsum yellow brown or dark brown .............................................................7
7 Three basal and four apical antennomeres light brown, four intermediate an-
tennomeres dark brown to piceous.............................................. L. gilli Gruev & Askevold
- Middle antennomeres not distinctly darker than distal or basal antennomeres...........................................................................................................8
8 Dorsum uniform light brown; hind wings well developed; elytral punctures
tend to be regular........................................................................... L. sari Maulik
- Elytra dark brown with lighter margins; hind wings absent; punctures con-
fused......................................................................................... L. fumidus Maulik

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