Clinical Evaluation of an Arterial-Spin-Labeling Product Sequence in Steno-Occlusive Disease of the Brain

Matthias A. Mutke¹,5*, Vince I. Madai¹,5*, Federico C. von Samson-Himmelstjerna¹,2,5, Olivier Zaro Weber¹,4,5, Gajanan S. Revankar¹, Steve Z. Martin¹,5, Katharina L. Steng¹,5, Miriam Bauer¹,5, Stefan Hetzer³, Matthias Günther², Jan Sobesky¹,5*

1 Center for Stroke Research, Charité Universitätsmedizin, Berlin, Germany, 2 Fraunhofer/MEVIS, Bremen, Germany, 3 Berlin Center for Advanced Neuroimaging, Berlin, Germany, 4 Max-Planck-Institute for Neurological Research, Cologne, Germany, 5 Department of Neurology, Charité Universitätsmedizin, Berlin, Germany

Abstract

Introduction: In brain perfusion imaging, arterial spin labeling (ASL) is a noninvasive alternative to dynamic susceptibility contrast-magnetic resonance imaging (DSC-MRI). For clinical imaging, only product sequences can be used. We therefore analyzed the performance of a product sequence (PICORE-PASL) included in an MRI software-package compared with DSC-MRI in patients with steno-occlusion of the MCA or ICA >70%.

Methods: Images were acquired on a 3T MRI system and qualitatively analyzed by 3 raters. For a quantitative analysis, cortical ROIs were placed in co-registered ASL and DSC images. Pooled data for ASL-cerebral blood flow (CBF) and DSC-CBF were analyzed by Spearman’s correlation and the Bland-Altman (BA)-plot.

Results: In 28 patients, 11 ASL studies were uninterpretable due to patient motion. Of the remaining patients, 71% showed signs of delayed tracer arrival. A weak correlation for DSC-relCBF vs ASL-relCBF (r = 0.24) and a large spread of values in the BA-plot owing to unreliable CBF-measurement was found.

Conclusion: The PICORE ASL product sequence is sensitive for estimation of delayed tracer arrival, but cannot be recommended to measure CBF in steno-occlusive disease. ASL-sequences that are less sensitive to patient motion and correcting for delayed blood flow should be available in the clinical setting.

Introduction

Cerebral blood flow (CBF) is an important measure of brain perfusion in patients with steno-occlusive disease [1]. Using magnetic resonance imaging (MRI), dynamic susceptibility-weighted contrast-enhanced (DSC) imaging is the clinical standard of CBF measurement. It is fast and offers a reliable estimate of CBF [2]. However, the main drawback of DSC imaging is the need of a gadolinium-based contrast agent. Administration of gadolinium is invasive, may cause anaphylaxis and patients with renal insufficiency must be excluded owing to possible nephrogenic systemic sclerosis [3]. Additionally, repeated measurements are difficult to perform owing to slow clearance of gadolinium based contrast agents [4]. Arterial spin labeling (ASL) measures CBF without the need of an exogenous contrast agent [5]. Here, blood is magnetically labeled by the MR scanner and the labeled blood water protons serve as an endogenous contrast agent to calculate CBF. ASL therefore permits multiple noninvasive CBF measurement. Various ASL techniques have been successfully validated to measure CBF in neuropathologies, e.g. in brain tumors [6], epilepsy [7] and stenosis of cerebral arteries and acute stroke [8]. Most of these sequences, however, were research sequences and are not available for clinical imaging. For clinical diagnosis, in contrast, available product sequences have to be used and their validation is urgently needed [9,10]. We therefore studied the performance of a clinically available commercial ASL-sequence to measure CBF in comparison with DSC-MRI in patients with steno-occlusive disease of the brain.

Materials and Methods

Ethics Statement

All patients gave informed written consent prior to the study. The study was conducted according to the principles expressed in the Declaration of Helsinki and was approved by the authorized
Study Design

We performed an observational prospective imaging study (Perfusion imaging by arterial spin labeling for clinical use in stroke - PEGASUS, WHO international Clinical trials registry No. DRKS00003198). Patients with steno-occlusive disease were recruited at the Department of Neurology of the Charité-Universitätsmedizin Berlin or presenting at our out-patient services between September 2011 and November 2012. Inclusion criteria were: a) unilateral stenosis >70% of one internal carotid artery (ICA) or middle cerebral artery (MCA) according to the ECST (European Carotid Surgery Trial) criteria, b) age 18–80, and c) clinically and hemodynamically stable status. Grading of stenosis was confirmed prior to MRI by Duplex-Sonography and/or CT-angiography.

Exclusion criteria were a) contralateral stenosis >50% of the CCA (common carotid artery), ICA or MCA b) magnetic implants, c) claustrophobia, d) aphasia or reduced level of consciousness, e) severe allergic reactions in the previous medical history, f) allergic reactions against Gadolinium-based contrast agents in the past, g) renal insufficiency (defined by a GFR ≤30 ml/min/1.73 m²), h) pregnancy and i) unstable clinical status. For each patient National Institute of Health Stroke Scale (NIHSS), modified Rankin Scale (mRS) and relevant clinical data was assessed before imaging.

Magnetic Resonance Imaging Hardware

MR-imaging was performed on a 3 T whole-body system (Magnetom Trio, Siemens Healthcare, Erlangen, Germany) using a 12-channel receive RF coil (Siemens Healthcare, Erlangen, Germany) tailored for head imaging.

Magnetic Resonance Imaging Sequences

We used a commercially available pulsed arterial spin labeling (PASL) PICORE Q2TIPS product sequence [11,12]: 50 pairs of label/control ASL images were acquired in axial direction at a single inversion time of 1800 ms (EPI-readout, TR = 2600 ms, TE = 13 ms, TI1 = 700 ms, label thickness = 100 mm, PICORE Q2T perfusion mode, voxel size: 3x3x3 mm³, no crusher gradients, enabled prospective motion correction [3D-PACE]).

The DSC-MRI protocol consisted of 80 series of whole-brain images using a single-shot FID-EPI sequence (TR = 1390 ms, TE = 29 ms, voxel size: 1.8x1.8x5 mm³) after injection of 5 ml Gadovist® (Gadobutrol, 1 M, Bayer Schering Pharma AG, Berlin) followed by 25 ml saline flush by a power injector (Spectris, Medrad Inc., Warrendale PA, USA) at a rate of 5 ml/s. Acquisition time for ASL was 4 min:18 s, for DSC-MRI 1 min:54 s. DSC-MRI was performed immediately after ASL.

Data Postprocessing

ASL images were acquired directly from the console and were used without further post-processing. DSC-MRI images were postprocessed offline with PMA (Perfusion Mismatch Analyzer, ASIST Japan, Iwate, Japan). Two maps were generated from the raw data: 1) DSC-time-to-peak (TTP)-maps - indicating regional delayed blood flow- and 2) DSC-CBF-maps – indicating regional blood flow - with automatic placement of the arterial input function using sSVD (standard singular value decomposition) deconvolution.

Figure 1. Rating of quality. The figure demonstrates the different image qualities. Rows show examples for good, medium, sufficient and uninterpretable quality. The first column demonstrates DSC-CBF images. All DSC-CBF maps were of either “good” or “medium” quality. The second column shows ASL-CBF images. No ASL-CBF map was rated “good”. doi:10.1371/journal.pone.0087143.g001
Figure 2. Quantitative analysis of DSC-CBF, DSC-TTP and ASL-relCBF. 20 mm circular regions of interest (ROIs) were placed on the FLAIR image and copied onto coregistered DSC-CBF, DSC-TTP and ASL images. For relative values, the ratio of each ipsilateral ROI to the arithmetic mean of all contralateral ROIs was calculated. In case of DSC-TTP the values were subtracted to obtain relative TTP-delay. DSC-perfusion maps were automatically masked by the used software. Therefore, the contour of DSC maps was used to define the cortical rim (contour, arrows).

doi:10.1371/journal.pone.0087143.g002

Table 1. Clinical data of all patients.

| Patient number | sex | Age (y) | mRS (p) | NIHSS (p) | Barthel (p) | Stroke | TIA | GoS (%) | GoS (%) | GoS (%) | GoS (%) |
|---------------|-----|---------|---------|-----------|------------|--------|-----|---------|---------|---------|---------|
|               |     |         |         |           |            |        |     |         |         |         |         |
| 1             | f   | 66      | 0       | 0         | 100        |        |     | 100     |         |         |         |
| 2             | m   | 63      | 0       | 0         | 100        |        |     | 70–80   |         |         |         |
| 3             | f   | 70      | 0       | 0         | 100        | x      |     |         | 80      |         |         |
| 4             | f   | 79      | 2       | 3         | 100        | x      |     |         | 70–80   |         |         |
| 5             | m   | 45      | 0       | 0         | 100        | x      |     |         | 100     |         |         |
| 6             | f   | 45      | 1       | 1         | 100        | x      |     |         | 100     |         |         |
| 7             | f   | 56      | 0       | 0         | 100        | x      |     |         | 100     |         |         |
| 8             | m   | 42      | 1       | 1         | 100        | x      |     |         | 100     |         |         |
| 9             | f   | 50      | 0       | 0         | 100        |        |     | 100     |         |         |         |
| 10            | m   | 59      | 1       | 2         | 100        | x      |     | 90      |         |         |         |
| 11            | f   | 57      | 0       | 0         | 100        | x      |     | 100     |         |         |         |
| 12            | f   | 45      | 0       | 0         | 100        | x      |     | 70      |         |         |         |
| 13            | m   | 63      | 2       | 4         | 100        | x      |     | 80      |         |         |         |
| 14            | f   | 49      | 0       | 0         | 100        | x      |     | 90      |         |         |         |
| 15            | f   | 36      | 2       | 2         | 100        | x      |     | 100     |         |         |         |
| 16            | m   | 76      | 0       | 0         | 100        | x      |     | 70      |         |         |         |
| 17            | m   | 31      | 0       | 0         | 100        | x      |     | 100     |         |         |         |
| 18            | m   | 47      | 2       | 4         | 75         | x      |     | 100     |         |         |         |
| 19            | f   | 63      | 0       | 0         | 100        | x      |     | 70      |         |         |         |
| 20            | f   | 37      | 0       | 0         | 100        |        |     | 100     |         |         |         |
| 21            | f   | 73      | 0       | 0         | 100        |        |     | 70      | 30      |         |         |
| 22            | m   | 48      | 0       | 0         | 100        |        |     | 100     |         |         |         |
| 23            | m   | 46      | 0       | 0         | 100        | x      |     | 100     |         |         |         |
| 24            | f   | 73      | 0       | 0         | 100        | x      |     | 70–80   |         |         |         |
| 25            | m   | 65      | 0       | 0         | 100        |        |     | 100     |         |         |         |
| 26            | f   | 74      | 0       | 0         | 100        | x      |     | 80      |         |         |         |
| 27            | m   | 58      | 0       | 0         | 100        | x      |     | >80     |         |         |         |
| 28            | m   | 68      | 0       | 0         | 100        | x      |     | 70–80   |         |         |         |

y: years; p: points; mRS: Modified Rankin Scale, NIHSS: National Institute of Health Stroke Scale; GoS: Grade of Stenosis; ICA: Internal Carotid Artery; MCA: Middle Cerebral Artery.
hypoperfusion adjacent to the hyperintense areas [13]. (Both signs are are addressed in detail in the discussion).

Quantitative Analysis

For the quantitative analysis, ASL-images rated as “uninterpretable” were excluded. Coregistration and region of interest (ROI) placement was performed with VINCI 3.93 (Max-Planck Institute for Neurological Research, Cologne, Germany) [14].

Maps of ASL-CBF, DSC-CBF and DSC-TTP were coregistered with FLAIR as the reference image. Cortical ROIs (diameter: 20 mm) were placed on 7 consecutive axial slices located at the height of the lateral ventricles. Slices above the lateral ventricles and slices including the cerebellum were not included. ROIs were placed on FLAIR images and copied on all perfusion maps (Figure 2). ROIs were labeled according to the side of steno-occlusion (ipsi- or contralateral). Relative CBF values (relCBF) were calculated using the mean value of each ROI: (ipsilateral ROI value/mean of all contralateral ROIs on the same slice) [%]. The relative time to peak (relTTP) for ipsilateral ROIs was obtained by subtraction from the mean TTP of all contralateral ROIs. Negative voxel values were not included in the analysis.

In a following ROI-analysis, TTP-ROIs were divided in two groups: First, TTP-ROIs showing an ATDA in the corresponding ASL-ROI and second TTP-ROIs without ATDA in the corresponding ASL-ROI. For comparison, the relative TTP delay was calculated as described above.

In a further volumetric analysis, we segmented grey matter on MPRAGE (T1) maps using Mevislab (MeVis Medical Solutions AG, Bremen, Germany). The grey matter map was divided into three flow territories: ACA (anterior cerebral artery), MCA (middle cerebral artery), PCA (posterior cerebral artery) based on a brain perfusion atlas [15]. The grey matter maps were applied to co-registered ASL-CBF and DSC-CBF maps. The average value for a specific flow territory volume was normalized to the mean perfusion of the whole contralateral grey matter perfusion.

Statistical Analysis

Study data are given in median and interquartile range owing to skewed distribution of some variables if not indicated otherwise. In the ROI-analysis, values of ASL-relCBF were plotted against relTTP and DSC-relCBF and correlated using Spearman’s rho.
relCBF and high differences in high average relCBF, suggesting a proportional error. This was, however, not confirmed in a logarithmic transformation of the plot (data not shown). The Bland Altman Plot is shown in Figure 4.

TTP-ROIs with a corresponding ATDA on ASL had a significantly (p < 0.001) longer median delay (1.9 sec) than those without a corresponding ATDA (0.6 sec).

In the volumetric grey matter analysis, for ASL-CBF and DSC-CBF no significant correlation was found for all three arterial flow territories (ACA: r = 0.01 [p = 0.97], MCA: r = 0.09 [p = 0.72], PCA: r = 0.05 [p = 0.85]).

**Discussion**

We report on the performance of an arterial spin labeling product sequence to measure CBF in patients with steno-occlusive disease in comparison with the clinical standard DSC-MRI. The tested ASL sequence is the only commercial, approved product sequence currently available for this system [10]. Sensitivity to patient motion made 39% of obtained ASL-images uninterpretable. In the visual analysis, ASL identified blood flow delay as defined by DSC-MRI with fair sensitivity, but led to unreliable measurement of CBF.

There is emerging consensus that ASL is a reliable alternative to exogenous contrast agent based perfusion measurements. Its use in routine clinical imaging has been encouraged [16]. However, most of the ASL techniques used for scientific neuroimaging are research sequences. This is important to note, as these sequences are usually not available in the clinical setting. Clinicians, who implement ASL in routine clinical imaging, currently are obliged to use commercially available product sequences. Therefore, the need for a validation of those sequences for CBF measurements in specific neurovascular pathologies is demanded [9,10]. For cerebrovascular disease, only a few clinical validation studies have been conducted with product sequences [17–22]. Particularly for steno-occlusive disease, validation studies have not been performed for all available product sequences. Further, ASL in steno-occlusive disease is of high interest as it can be used in patients with contraindications for gadolinium and facilitates a noninvasive estimation of the cerebrovascular reactivity (CVR) [23]. In this respect, we aimed to define the performance of a commercially available ASL sequence (PICORE, pulsed ASL [PASL] technique) in patients with unilateral hemodynamically relevant steno-occlusive disease on a widely available 3T MRI system.

In PASL, the spins of protons of passing blood are inverted by a radiofrequency (RF)-pulse at a labeling slice usually at the neck,
covering the internal and external carotid and the vertebral arteries. By this technique, a single bolus of labeled blood is generated. The time this blood bolus needs to reach the brain capillaries is termed arterial transit time (ATT). In the brain capillary bed, labeled blood water then exchanges with residual tissue water. ASL images are acquired at one predefined time point after labeling, the inflow time (TI). It is important to notice that the TI has to be longer than the ATT to allow imaging in the capillary phase of blood transit. A second (control) image is then obtained without labeling. Subtraction of the two images yields perfusion weighted maps providing CBF-values. All ASL sequences in steno-occlusive disease, however, faces major obstacles owing to the prolonged ATT caused by stenosis/occlusion of brain feeding arteries. In this case, two signs may be observed: First, at the predefined inflow time, the blood bolus has not yet reached the capillary bed in the regions affected by steno-occlusion. Therefore, labeled blood is found mainly in precapillary arterioles. On ASL images, these vessels appear as hyperintense dots or lines depending on the image orientation [10,13,24]. Second, if the delay leads to very long arterial transit times, the blood bolus has not yet reached the perfusion territory at the predefined inflow time at all. In these regions, the signal is very low or lacks completely and thus severely underestimates real perfusion [10,25]. These signs represent the arterial transit delay artifact (ATDA) [10,13,24]. The origin of the ATDA is shown in figure 5, examples for both effects are shown in figure 6. Both effects may appear simultaneously when areas with different prolonged ATTs are present in one patient. Therefore, a patient with normal cerebral blood flow but delayed arrival time, might have a falsely elevated or a falsely decreased CBF in the ASL image (Figure 6). In line with pathophysiology, these effects were present in our study since ipsilateral ATDA was found in 12 of 17 patients (71%). On one hand, the presence of ATDAs clearly identified the site of DSC-TTP-delay and/or DSC-hypoperfusion in the qualitative analysis and areas with ATDA in ASL showed a longer median TTP-delay on TTP-maps. On the other hand, however, the hyperintense nature of ATDA led to an unreliable quantification of ASL-CBF. This causes a weak correlation between ASL-CBF and DSC CBF-values as indicated by BA-plot analysis. These results at 3 T are supported by previous results obtained at 1.5 T, Wolf et al. found no correlation between ASL-CBF and DSC-CBF at 1.5 T in patients with steno-occlusive disease [15]. In our ROI-based approach we chose large ROIs to account for spatial distortion of the different imaging modalities. As a consequence, our ROIs included white matter, in which ASL might not allow for reliable quantification of CBF. However, our volumetric analysis of gray matter perfusion alone did not lead to an improved correlation.

To account for the prolonged ATT in patients with steno-occlusive disease it would be possible to choose longer TIs [26], but most of the signal would be lost at that time [27]. This intrinsic problem can only be solved by multiple averaging but leads to unacceptable scanning times.

Out of 28, 11 data sets (39%) were uninterpretable owing to patient motion artifacts despite online motion correction (performed by 3D-PACE) and, generally, image quality in ASL was worse in comparison with DSC-MRI. Patients were asked to refrain from motion, and small pads were placed between the patient head and the coil to limit head movement. The high frequency of motion artifacts in our study - despite the mentioned measures, - might thus reflect a limitation of this method in our specific patient sample, as increased patient motion reflects clinical reality. This is supported by a validation of the same product sequence in acute stroke, where the authors also identified patient motion and imperfect motion correction as a major factor for limited image quality [18]. Offline motion correction by in-house
Figure 6. Exemplary patients showing blood arrival delay effects in ASL imaging. A) 66-year-old female, occlusion of the right ICA. The TTP image shows a visible ipsilateral delay (relTTP = 0.8 sec). In the ASL image, hyperintense arterial transit delay artifacts are seen (small white arrows). On the reference DSC-CBF map, a hypoperfusion in the affected hemisphere is present. B) 45-year-old female, occlusion of the left middle cerebral artery. In the ASL image, a hypointensity is seen in the area affected by delay (large white arrow, relTTP = 1.5 sec). In contrast, no apparent changes are present in the DSC-CBF map. In patient A, a moderate blood transit delay leads to the presence of hyperintense ADTA. The more severe delay in patient B might explain the hypointense ADTA surrounded by hyperintense areas in its borderzone as a sign of collateral macrovessels. All slices are coregistered. The scales do not represent absolute values.

doi:10.1371/journal.pone.0087143.g006
solutions might be preferable [28] but is not available in the clinical setting. The described PICORE sequence offers the option of crusher gradients, that suppress intravascular blood signal [29] and reduce hyperintense vessel ATDAs. However, for reasons of methodology we chose this option: First, it does not eliminate the cause of the artifact, but its visibility. Second, the ATDA indicates the underlying transit delay and the lack of this information would render image analysis more difficult. Third, the use of crusher gradients further reduces the signal-to-noise ratio in ASL, which is already intrinsically low [10] especially in sequences without background suppression, e.g. the PICORE sequence.

Therefore, the PICORE product sequence available for the MRI system used in our study is not suited to reliably depict CBF in steno-occlusive disease at 3 T owing to the sensitivity to delayed blood arrival.

Currently, new ASL sequences address many methodological issues by correction for the delayed blood arrival time employing measurements at multiple inflow times. A very promising approach is a 3D ASL sequence (3D-GRASE) [30]. The benefit of this sequence in patients with steno-occlusive disease has been shown recently [31]. Another promising approach is the "look-locker approach" (ITS-FAIR [32], QUASAR [33]). Using this sequence in steno-occlusive disease at 3 T, recent studies found a high correlation with SPECT data [17] and a fair [34] to high [35] correlation with H2O-PET. These sequences, however, are not yet commercially available for all MRI systems.

Our study has some limitations: First, perfusion changes cannot be ruled out completely given the heterogeneity of perfusion alterations in patients with steno-occlusive disease. However, DSC-imaging followed ASL immediately and the clinical status of the measured patients was stable. Second, the methods of ASL and DSC differ considerably: While ASL uses blood as a freely diffusible and endogenous tracer, gadolinium in DSC mainly remains intravascular. Such methodological differences are a well-known difficulty of multimodal imaging and must be kept in mind when interpreting the results of comparative studies. While DSC-MRI can be considered a clinical standard for perfusion imaging, it is not a gold-standard. Methodological shortcomings include overemphasis of large vessels intrinsic to GRE-imaging [36] and the influence of the deconvolution method in patients with transit delay [37]. While a block circulant deconvolution (sSVD) might be preferable to sSVD from a technical point of view, this has been shown in direct comparison in only few pilot studies [37,38]. Additionally, sSVD requires much more computing power limiting its use in the clinical setting. Therefore, sSVD, as the current clinical standard, was chosen for the comparison with a clinically available ASL sequence in this study.

In conclusion, the commercially available ASL-sequence used in our study can indicate delay of blood arrival in steno-occlusive disease, but cannot be recommended for CBF measurement in these patients. New ASL-sequences present promising alternatives, but are not yet available for clinical use on all MRI systems.

Acknowledgments

We express our gratitude to the staff of the MRI unit at the Berlin Center for Advanced Neuroimaging (BCAN). We would like to thank especially Mrs. Andrea Hassenpflug for her excellent technical assistance.

Author Contributions

Conceived and designed the experiments: MM VM FS OZW MB JS. Performed the experiments: MM VM FS OZW SM KS SH JS. Analyzed the data: MM VM FS OZW GR MG JS. Contributed reagents/materials/analysis tools: MM VM FS OZW SH MG JS. Wrote the paper: MM VM FS JS.

References

1. Wintermark M, Sesay M, Barbier E, Borbely K, Dillon WP, et al. (2005) Comparative Overview of Brain Perfusion Imaging Techniques. Stroke 36: e83–e99. doi:10.1161/01.STR.0000177847.26703.7b.
2. Sobesky J (2012) Refining the mismatch concept in acute stroke: lessons learned from PET and MRI. J Cereb Blood Flow Metab 32: 1416–1425. doi:10.1038/jcbfm.2011.54.
3. Kaewri L, Abuhelal H (2012) Nephrogenic systemic fibrosis. AJR Am J Roentgenol 199: W17–23. doi:10.2214/AJR.11.14114.
4. Levin JM, Kaufman MJ, Ross MH, Mendelow JH, Maas LC, et al. (1995) Sequential dynamic susceptibility contrast MR experiments in human brain: residual contrast agent effect, steady state, and hemodynamic perturbation. Magn Reson Med 34: 653–663.
5. Detre JA, Leigh JS, Williams DS, Koretsky AP (1992) Perfusion imaging. Magn Reson Med 23: 37–45.
6. Lehmann P, Monet P, de Marco G, Salou G, Perrin M, et al. (2010) A comparative study of perfusion measurement in brain tumors at 3 Tesla MR: Arterial spin labeling versus dynamic susceptibility contrast-enhanced MRI. Eur Radiol 20: 21–26. doi:10.1007/s00330-009-1137-0.
7. Lim YM, Cho YW, Shamin S, Solomon J, Birn R, et al. (2008) Usefulness of pulsed arterial spin labeling MR imaging in mesial temporal lobe epilepsy. Epilepsy Res 82: 103–109. doi:10.1016/j.eplepsyres.2008.08.001.
8. Donohue MJ, Strother MK, Hendrikse J (2012) Novel MRI approaches for assessing cerebral hemodynamics in ischemic cerebrovascular disease. Stroke 43: 903–915. doi:10.1161/STROKEAHA.111.635995.
9. Golay X, Gruetter M (2012) Arterial spin labelling: final steps to make it a clinical reality. Magnetic Resonance Materials in Physics, Biology and Medicine 25: 53–62. doi:10.1007/s10334-012-0309-9.
10. Zahrachak G (2012) Arterial Spin Labeling for Acute Stroke: Practical Considerations. Transl Stroke Res 3: 228–235. doi:10.1007/s12973-012-0159-9.
11. Uhl W, Wong EC, Bandettini PA, Hyde JS (1999) QUIPSS II with thinslice TI1 periodic saturation: A method for improving accuracy of quantitative perfusion imaging using pulsed arterial spin labeling. Magnetic Resonance in Medicine 41: 1246–1254. doi:10.1002/1527-1583(199909)41:6<1246::AID-MRM222>3.0.CO;2-N.
12. Wong EC, Buxton RB, Frank LR (1998) Quantitative imaging of perfusion using a single subtraction (QUIPSS and QUIPSS II). Magn Reson Med 39: 762–768.
13. Wolf RL, Alsop DC, McGarvey ML, Maddijian JA, Wang J, et al. (2003) Susceptibility Contrast and Arterial Spin Labeled Perfusion MRI in Cerebrovascular Disease. Contrast and Molecular Imaging of Human Brain. Front Neurol Neurosci 30: 99–110. doi:10.1159/000333692.
14. Cizek J, Hertle K, Vollmar S, Schrader R, Klein J, et al. (2004) Fast and robust registration of PET and MR images of human brain. Neuroimage 22: 434–442. doi:10.1016/j.neuroimage.2004.01.016.
15. Tatu L, Moulin T, Vuillier F, Bogousslavsky J (2012) Arterial territories of the human brain. Front Neurol Neurosci 30: 99–110. doi:10.1159/000333692.
16. Detre JA, Rao H, Wang DJJ, Chen YF, Wang Z (2012) Applications of arterial spin labeled MRI in the brain. J Magn Reson Imaging 35: 1026–1037. doi:10.1002/jmri.22581.
17. Uchiha Y, Hosoda K, Zimino I, Fujita A, Fujiy M, et al. (2011) Clinical application of arterial spin-labeling MR imaging in patients with carotid stenosis: quantitative comparative study with single-photon emission CT. AJNR Am J Neuroradiol 32: 1545–1551. doi:10.3174/ajnr.A2525.
18. Hock S, Kod HH, Al-Zghoul M, Groden C, Nolte I (2012) Arterial spin labeling at 3.0 Tesla in subacute ischemia: comparison to dynamic susceptibility perfusion. Clin Neuroradiol 22: 29–37. doi:10.1007/s00062-011-0126-x.
19. Zahrachak G, Mogy ISE, Fischbein NJ, Albers GW (2012) Comparison of Arterial Spin Labeling and Bolus-Tracking Perfusion Imaging for Detecting Mismatch in Acute Stroke. Stroke 43: 1843–1848. doi:10.1161/STROKEAHA.111.639773.
20. Bivard A, Krishnamurthy V, Swannell P, Levi C, Spratt NJ, et al. (2013) Arterial Spin Labeling Versus Bolus-Tracking Perfusion in Hyperacute Stroke. Stroke 44: 2601–2603. doi:10.1161/STROKEAHA.113.002997.
21. Huang YC, Liu HL, Lee JD, Yang JT, Weng HH, et al. (2013) Comparison of Arterial Spin Labeling and Dynamic Susceptibility Contrast perfusion MRI in patients with acute stroke. PLoS ONE 8: e69085. doi:10.1371/journal.pone.0069085.
22. Bolke RPH, van Osch MJP, van der Woop HB, de Boer GJ, Mali WPMT, et al. (2010) Symptomatic Carotid Artery Stenosis: Impairment of Cerebral Autoregulation Measured at the Brain Tissue Level with Arterial Spin-labeling MR Imaging. Radiology 256: 201–208. doi:10.1148/rad.201091262.
24. Golay X, Hendrikse J, Lim TCC (2004) Perfusion imaging using arterial spin labeling. Top Magn Reson Imaging 15: 10–27.
25. Chng SM, Petersen ET, Zimine I, Sitoh YY, Lim CCT, et al. (2008) Territorial Arterial Spin Labeling in the Assessment of Collateral Circulation Comparison With Digital Subtraction Angiography. Stroke 39: 3246–3254. doi:10.1161/STROKEAHA.108.520593.
26. Alsop DC, Detre JA (1996) Reduced transit-time sensitivity in noninvasive magnetic resonance imaging of human cerebral blood flow. J Cereb Blood Flow Metab 16: 1236–1249. doi:10.1097/00004647–199611000–00019.
27. Qiu D, Straka M, Zun Z, Bammer R, Moseley ME, et al. (2012) CBF measurements using multidelay pseudocontinuous and velocity-selective arterial spin labeling in patients with long arterial transit delays. Comparison with xenon CT CBF. Journal of Magnetic Resonance Imaging 36: 110–119. doi:10.1002/jmri.23613.
28. Bivard A, Stanwell P, Levi G, Parsons M (2013) Arterial spin labeling identifies tissue salvage and good clinical recovery after acute ischemic stroke. J Neuroimaging 23: 391–396. doi:10.1111/j.1552–6569.2012.00728.x.
29. Ye FQ, Mattay VS, Jezzard P, Frank JA, Weinberger DR, et al. (1997) Correction for vascular artifacts in cerebral blood flow values measured by using arterial spin tagging techniques. Magn Reson Med 37: 226–235.
30. Günther M, Oshio K, Feinberg DA (2005) Single-shot 3D imaging techniques improve arterial spin labeling perfusion measurements. Magnetic Resonance in Medicine 54: 491–498. doi:10.1002/mrm.20580.
31. MacIntosh BJ, Lindsay AC, Kylintireas I, Kuker W, Günther M, et al. (2010) Multiple inflow pulsed arterial spin-labeling reveals delays in the arterial arrival time in minor stroke and transient ischemic attack. AJNR Am J Neuroradiol 31: 1892–1894. doi:10.3174/ajnr.A2008.
32. Günther M, Bock M, Schad LR (2001) Arterial spin labeling in combination with a look-locker sampling strategy: inflow turbo-sampling EPI-FAIR (ITS- FAIR). Magn Reson Med 46: 974–984.
33. Petersen ET, Lim T, Golay X (2006) Model-free arterial spin labeling quantification approach for perfusion MRI. Magn Reson Med 55: 219–232. doi:10.1002/mrm.20784.
34. Bokkers RP, Bremmer JP, van Berckel BN, Lammertsma AA, Hendrikse J, et al. (2009) Arterial spin labeling perfusion MRI at multiple delay times: a correlative study with H215O positron emission tomography in patients with symptomatic carotid artery occlusion. J Cereb Blood Flow Metab 30: 222–229.
35. Kamano H, Yoshiura T, Hiwatashi A, Abe K, Togao O, et al. (2013) Arterial spin labeling in patients with chronic cerebral artery steno-occlusive disease: correlation with (13)O-PET. Acta Radiol 54: 99–106. doi:10.1258/ar.2012.120450.
36. Boxerman JL, Hamborg LM, Rosen BR, Weisskoff RM (1995) MR contrast due to intravascular magnetic susceptibility perturbations. Magn Reson Med 34: 555–566.
37. Wu O, Østergaard L, Weisskoff RM, Benner T, Rosen BR, et al. (2003) Tracer arrival timing-insensitive technique for estimating flow in MR perfusion-weighted imaging using singular value decomposition with a block-circulant deconvolution matrix. Magn Reson Med 50: 164–174. doi:10.1002/mrm.10522.
38. Matsushima S, Kubota T, Yamada K, Akazawa K, Masunami T, et al. (2008) Effect of vascular stenosis on perfusion-weighted imaging: differences between calculation algorithms. J Magn Reson Imaging 27: 1103–1108. doi:10.1002/jmri.21362.