Recent Updates on the Melanin-Concentrating Hormone (MCH) and Its Receptor System: Lessons from MCH1R Antagonists

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Abstract Melanin-concentrating hormone (MCH) is a 19-amino-acid cyclic peptide which was originally found to lighten skin color in fish that is highly conserved among many species. MCH interacts with two G-protein-coupled receptors, MCH1R and MCH2R, but only MCH1R is expressed in rodents. MCH is mainly synthesized in the lateral hypothalamus and zona incerta, while MCH1R is widely expressed throughout the brain. Thus, MCH signaling is implicated in the regulation of many physiological functions. The identification of MCH1R has led to the development of small-molecule MCH1R antagonists that can block MCH signaling. MCH1R antagonists are useful not only for their potential therapeutic value, but also for understanding the physiological functions of the endogenous MCH system. Here, we review the physiological functions of the MCH system which have been investigated using MCH1R antagonists such as food intake, anxiety, depression, reward, and sleep. This will help us understand the physiological functions of the MCH system and suggest some of the potential applications of MCH1R antagonists in human disorders.

Keywords Melanin-concentrating hormone (MCH) · G-protein-coupled receptors (GPCRs) · Antagonists · Energy homeostasis

Introduction

Melanin-concentrating hormone (MCH) was initially discovered in 1983 to be a 17-amino-acid cyclic peptide that induces paling of the skin in fish (Kawauchi et al. 1983). In 1996, MCH was discovered to be overexpressed in rodents upon fasting and in ob/ob mice (Qu et al. 1996). Central MCH injection increases food intake, suggesting that MCH is an orexigenic peptide. Mice lacking precursor MCH (ppMCH) have been shown to be hypophagic and lean (Shimada et al. 1998). These lines of evidence suggest that the MCH system modulates energy homeostasis. However, its site of action was not known until 1999 when we and other groups discovered a receptor for MCH (Bachner et al. 1999; Chambers et al. 1999; Lembo et al. 1999; Saito et al. 1999; Shimomura et al. 1999). MCH1 receptor is a G-protein-coupled receptor that couples to Gi and Gq proteins (Hawes et al. 2000). MCH1R is exclusively expressed in the hypothalamus and zona incerta whereas MCH1R is widely expressed in central and peripheral tissues. MCH1R distribution is particularly widespread throughout the brain, being particularly concentrated in areas such as the cortex, hippocampus, amygdala, and nucleus accumbens shell (Saito et al. 2001), suggesting that the MCH system may be involved in a wide range of physiological functions in the central nervous system. MCH1R is also expressed in many peripheral tissues such as pituitary, intestine, lymphocytes, and adipose tissue (Hill et al. 2001), suggesting peripheral...
sites of MCH action and involvement in peripheral physiological functions. In 2001, another receptor for MCH, MCH2R, which exists in humans but not in rodents, was discovered (Hill et al. 2001; Sailer et al. 2001).

Studies using ppMCH knockout mice or MCH1R knockout mice suggested that the MCH system might be involved in regulating energy homeostasis (Marsh et al. 2002; Shimada et al. 1998). Genetic knockout mice are useful animal models to understand the physiological functions of the MCH system. However, significant developmental compensation can occur as a result of germ line gene disruption leading to discordant results between genetic and pharmacological studies. Examples of discrepancies between results obtained through genetic versus pharmacological studies have been seen with MCH and other signaling systems. MCH1R knockout mice are hyperphagic (Marsh et al. 2002) whereas central MCH injection increases food intake (Qu et al. 1996). Another example of this phenomenon was seen when neuropeptide Y (NPY)5R knockout mice exhibit a mildly obese phenotype (Marsh et al. 1998) while NPY5R antagonists cause mild weight loss (Erondu et al. 2006; Ishihara et al. 2006).

Therefore, the use of pharmacological antagonists to understand the biological functions of a system is crucial to avoid the confounding effects of developmental compensations which are often present in knockout and transgenic animals.

Here, we review what has been learned about the physiological functions of the MCH system by blocking endogenous MCH signaling with MCH1R antagonists. In particular, we will focus on the role of MCH in food intake, anxiety, depression, reward, and sleep.

MCH1R Antagonists and Energy Homeostasis

Central MCH injection increases food intake (Qu et al. 1996) while mice lacking ppMCH are hypophagic and lean (Shimada et al. 1998), and MCH1R KO mice are hyperphagic, hyperactive, and lean (Marsh et al. 2002). This strongly suggests that MCH signaling is involved in energy balance and food intake.

In 2002, two papers were published indicating that small-molecule MCH1R antagonists, SNAP-7941 and T-226296, may be useful in the treatment of obesity (Borowsky et al. 2002; Takekawa et al. 2002). Systemic injections of both MCH1R antagonists decreased MCH-induced food intake, and chronic SNAP-7941 administration decreased body weight in rats with diet-induced obesity (DIO).

Numerous small-molecule MCH1R antagonists have since been developed and have consistently been effective in animal models of DIO (Borowsky et al. 2002; Eric Hu et al. 2008; Gehlert et al. 2009; Ito et al. 2010; Kowalski et al. 2006; Luthin 2007; Mashiko et al. 2005), reviewed in (Luthin 2007; McBriar 2006). Chronic administration of MCH1R antagonists successfully decreased food intake and body weight gain induced by high caloric food consumption. The central MCH system seems to play an important role in regulating energy balance and metabolism since Mashiko et al. (2005) showed that chronic central infusions of a peptide MCH1R antagonist into DIO mice only slightly decreased food intake but significantly reduced body weight. Another study reported that only MCH1R antagonists which can cross the blood brain barrier (BBB) are able to decrease food intake and body weight, while those that do not are ineffective (Eric Hu et al. 2008). However, the BBB-crossing antagonist used in this study also induced a modest conditioned taste aversion (Eric Hu et al. 2008), which complicates interpretation of these results. Additional studies found that central MCH1R antagonist injection reduced MCH-induced food intake but failed to decrease spontaneous food intake when it was injected alone (Audinot et al. 2009; Morens et al. 2005).

Although there is strong evidence that the central MCH system plays an important role in modulating food intake and body weight gain, many studies suggest that peripheral MCH1R signaling may also be involved in modulating these processes since MCH1Rs are expressed in a variety of peripheral tissues. It has been shown that the MCH system modulates leptin secretion and insulin release (Bradley et al. 2000; Tadayyon et al. 2000), suggesting that systemic injections of MCH1R antagonist may modulate food intake and body weight through both peripheral and central mechanisms. Thus, peripheral MCH1R antagonist administration will affect not only the brain, but also peripheral tissues such as brown adipose tissues, liver, pancreas, small intestines, and white adipose tissues.

Some studies suggest that MCH1R antagonist-induced weight loss may not be exclusively due to inhibition of feeding but may also involve an increase in metabolism (Huang et al. 2005; Ito et al. 2010). It has been reported that administration of a MCH1R antagonist only moderately suppressed feeding but led to a significant reduction in body weight (Ito et al. 2010). Pair feeding resulted in a significant weight loss, suggesting that MCH1R antagonism can modulate energy expenditure which leads to weight loss. Therefore, antiobesity effects of the MCH1R antagonists may be caused not only by a suppression of feeding, but also by an increase in energy expenditure.

These studies strongly indicate that pharmacological blockade of MCH signaling leads to a reduction in body weight and demonstrate that the MCH system is a promising target for the treatment of obesity.

MCH1R Antagonists and Anxiety

MCH1 receptors are widely distributed throughout the limbic brain regions that regulate stress and anxiety. Central MCH injection has been shown to be anxiogenic (Smith et al. 2006)
or anxiolytic (Monzon and De Barioglio 1999; Monzon et al.
2001), and MCH1R KO mice exhibit less anxiety-like
behaviors than wild-type mice (Roy et al. 2007; Smith et al.
2006), suggesting that the MCH system plays a critical
role in anxiety-like behaviors. In order to avoid potentially
confounding effects inherent in the injection and knockout
mouse studies, several groups have used MCH1R antag-
onsists to investigate the role of endogenous MCH signaling in
anxiety (summarized in Table 1).

In 2002, the MCH1R antagonist SNAP-7941 was shown to
exhibit anxiolytic properties in the separation-induced vocal-
ization test in guinea pig pups and to increase the interaction
time of paired unfamiliar rats in a social interaction test
(Borowsky et al. 2002). Following this initial discovery,
another MCH1R antagonist, SNAP-94847, was found to
display anxiolytic properties in the light–dark transition test
following both acute and chronic administration (David et al.
2007). Additionally, chronic administration of SNAP-94847
was found to be anxiolytic in the rat novelty-suppressed
feeding test (David et al. 2007).

Two other MCH1R antagonists, ATC0065 and ATC0175,
significantly reverse swim stress-induced anxiety in the
elevated plus maze test in rats and stress-induced hyperther-
mia in mice (Chaki et al. 2005). ATC0175 also increased
social interaction between unfamiliar rats and reduced
separation-induced vocalizations in guinea pig pups. Inter-
estingly, both of these compounds were found to be
ineffective in the marble burying task.

Another MCH1R antagonist, GW803430 (also referred
to as GW3430), exhibits anxiolytic activity in a diverse
array of behavioral models including the marble burying
task, elevated plus maze, and stress-induced hyperther-
mia without affecting motor activity in mice (Gehlert et al.
2009; Smith et al. 2006). GW803430, along with SNAP-
7941, has also been shown to have anxiolytic properties in
a Vogel conflict test and ultrasonic vocalization test (Mi-
Ian et al. 2008). Stressful stimuli are known to elevate plasma
adrenocorticotropic hormone (ACTH) and corticosterone
levels by activating the hypothalamic–pituitary–adrenal
(HPA) axis (Herman et al. 1996). MCH administration
increases ACTH and corticosterone levels, an effect that is
reversed by pretreatment with GW803430 (Smith et al.
2006). This suggests that the anxiolytic activity of MCH1R
antagonists may involve regulation of the HPA axis.

These studies consistently show that both acute and
chronic MCH antagonisms have anxiolytic effects and
strongly support a role for the MCH system in the
modulation of stress and anxiety.

MCH1R Antagonists and Depression

Due to the dense expression of MCH1R in areas of the
brain involved in stress, reward, and emotional regulation
(Saito et al. 2001), it was surmised that MCH signaling may
regulate depression-like behaviors. Pharmacological sup-
port for this hypothesis was found when the MCH1R
antagonist SNAP-7941 was discovered to reduce immobi-
lity time in the rat forced-swim test with similar efficacy as
fluoxetine (Borowsky et al. 2002). Following this initial
discovery, additional MCH1R antagonists, ATC0065
(Chaki et al. 2005), ATC0175 (Chaki et al. 2005), and
GW803430 (Gehlert et al. 2009), were found efficacious in
standard acute rodent depression models (McBriar 2006).
In addition to high affinity for MCH1R, ATC0175 showed
a moderate to high affinity for both 5-HT2A and 5-HT1A
receptors, and ATC0065 showed a moderate affinity for 5-
HT1A receptors (Chaki et al. 2005). This nonspecific
activity of ATC0065 and ATC0175 on other receptors
could make it difficult to interpret the specificity of
MCH1R involvement in the antidepressant effects of
ATC0065 and ATC0175. Additionally, the MCH1R antag-
onist SNAP-94847 has recently been reported to be
effective in reversing decreased sucrose intake in the
chronic mild stress anhedonia model (Smith et al. 2009)
and to decrease latency to novelty-suppressed feeding
(David et al. 2007) although it was not effective in the
forced-swim test. This indicates that MCH1R antagonists
are effective in both chronic and acute models of
depression.

The mechanism through which MCH1R antagonists
exert antidepressant activity has thus far not been
ecluciated, but current studies suggest that it likely occurs
through a mechanism distinct from existing antidepres-
sants. MCH1R antagonists enhance time spent swimming
without altering climbing behavior in the forced-swim test
(Chaki et al. 2005), an effect typically seen with
antidepressant compounds possessing primarily serotonin-
gic activity (Detke et al. 1995). Chronic treatment with
either selective serotonin reuptake inhibitors (SSRIs) or
MCH1R antagonists induce hippocampal neurogenesis
(David et al. 2007; Santarelli et al. 2003); however,
neurogenesis is not required for chronic MCH1R antago-
nist effects (David et al. 2007), indicating a mechanism of
action differing from SSRIs. Interestingly, it has also been
shown that MCH1R antagonists may enhance adaptation
to stressful stimuli (Smith et al. 2006) at least partially by
modulating HPA axis response. Additionally, the MCH1R
antagonist SNAP-94847 has a more rapid onset of action
in the novelty-suppressed feeding task than a traditional
antidepressant (David et al. 2007), highlighting a potential
advantage of MCH1R as a target for the treatment of
depression.

Despite a limited understanding of the role of endogenous
MCH signaling in the regulation of emotion and the absence
of detailed mechanistic studies into the nature of the
antidepressant effects observed with MCH1R antagonism,
the MCH1 receptor appears to be a promising new target for the treatment of depression.

MCH1R Antagonists and Reward

MCH1Rs are highly expressed throughout the mesolimbic dopamine system (Saito et al. 2001), particularly in the nucleus accumbens shell (Chung et al. 2009), a region important in the regulation of rewarding behaviors. To study the role of the MCH system in cocaine reward behavior, the MCH1R antagonist TPI 1361-17 (Nagasaki et al. 2009) was acutely injected into rats undergoing cocaine self-administration (Chung et al. 2009). Acute blockade of the MCH system decreased cocaine self-administration and cue- and cocaine-induced reinstatement. The role of the MCH system in food reward has also been studied. Systemic injections of a MCH1R antagonist, SNAP 94847, decreased high-fat food-reinforced operant responding and MCH-induced reinstatement of food-seeking behaviors in rats (Nair et al. 2009), suggesting that the MCH system is also involved in food reward.

MCH1R Antagonists and Sleep

It has recently been shown that MCH-producing neurons are primarily active during paradoxical (rapid eye movement (REM)) sleep and inactive during slow wave and waking states (Hassani et al. 2009; Verret et al. 2003), suggesting that MCH may regulate some aspects of REM sleep. ICV injection of MCH significantly increases time spent in paradoxical sleep (Verret et al. 2003), and mice lacking the MCH peptide (Willie et al. 2008) or receptor (Adamantidis et al. 2008) display altered sleep patterns. Several other studies have found that microinjection of MCH into brain nuclei with known involvement in sleep has modest effects on slow wave sleep but significantly increases paradoxical sleep time (Lagos et al. 2009). Somewhat surprisingly, relatively few studies have focused on how MCH antagonism affects sleep. One study found that two MCH1R antagonists, compound A and compound B, reduced both deep sleep and REM sleep while increasing time spent awake (Ahnaou et al. 2008), though it should be noted that both compounds possess affinity for the muscarinic M1 receptor and compound A for the dopamine transporter, targets known to affect sleep parameters. Conversely, another group reported that high occupancy of MCH1R with another MCH1R antagonist also named compound A does not affect sleep (Able et al. 2009).

While MCH injection, neuroanatomical, and knockout mouse studies all point towards the involvement of MCH signaling in the regulation of REM sleep, reports of the effects of MCH1R antagonism on sleep are limited and
conflicting. Further antagonist studies are needed to elucidate how endogenous MCH signaling is involved in sleep.

Conclusion

Here, we have reviewed studies into physiological functions regulated by the MCH system using MCH1R antagonists. Most MCH1R antagonists have been shown to be effective in reducing body weight in rats with DIO. However, the expansive distribution pattern of MCH1Rs indicates that MCH signaling is likely to be involved in a wide range of physiological functions, suggesting that MCH1R antagonists may be useful in human disorders other than obesity. The current scientific literature has shown that MCH1R antagonists are efficacious in animal models of depression, anxiety, sleep, and reward. The neuroanatomical and peripheral distribution of MCH1 receptors suggest that MCH is involved in more functions than are currently known. MCH1R antagonists will be invaluable tools for discovering additional functions of MCH signaling and will likely have therapeutic value in more disorders than are reviewed here.

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