Thyroid Hormone Receptor Beta in the Ventromedial Hypothalamus Is Essential for the Physiological Regulation of Food Intake and Body Weight

Highlights
- The ventromedial hypothalamus (VMH) expresses thyroid hormone receptor beta (TRβ)
- In mice, selective knockdown of TRβ (TRβ−) in the VMH results in severe obesity
- The obesity is due to overeating (hyperphagia) and reduced energy expenditure
- TRβ− mice have altered expression of hypothalamic regulators of food intake

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In Brief
Hameed et al. report that selective knockdown of a thyroid hormone receptor in the mouse hypothalamus results in a phenotype of severe obesity, overeating, and reduced energy expenditure, which may be due to downstream changes in the expression of hypothalamic regulators of food intake.

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Thyroid Hormone Receptor Beta in the Ventromedial Hypothalamus Is Essential for the Physiological Regulation of Food Intake and Body Weight

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INTRODUCTION

Energy homeostasis is regulated by neurotransmitters and by humoral factors including thyroid hormones, which act within the hypothalamus and systemically to regulate food intake (Coppola et al., 2007; Coll et al., 2007) and energy expenditure (Kim, 2008). The effects of the active form of thyroid hormone, 3,5,3′-triiodothyronine (T3), are mediated by two thyroid hormone receptors (TRs) and TRβ, encoded by Thra and Thrb, respectively (Brent, 2012).

Metabolic phenotypes have been described in mice and humans with TR mutations. Mice with heterozygous dominant-negative mutations of TRβ display a variety of metabolic phenotypes ranging from hypermetabolism, hyperphagia, and resistance to diet-induced obesity (Sjögren et al., 2007) to increased visceral adiposity, hypophagia, and impaired cold-induced adaptive thermogenesis (Liu et al., 2003). The variation in described phenotypes is likely to be due to the differing actions of individual mutant receptors on wild-type TR function (Ortiga-Carvalho et al., 2014). Humans with heterozygous dominant-negative mutations of TRβ (resistance to thyroid hormone α [RTHα]) may be overweight or obese with reduced energy expenditure (Bochukova et al., 2012; Moran et al., 2013, 2014). Humans with RTHβ may be overweight and hyperphagic (Mitchell et al., 2010) despite features of hyperthyroidism such as tachycardia and raised energy expenditure due to T3 actions in TRβ-responsive tissues. These extensive studies demonstrate that thyroid hormone is an essential regulator of food intake and energy expenditure. Despite this, clinical and global gene targeting studies cannot differentiate between the developmental and adult, or systemic and central, effects of thyroid hormones.

The ventromedial hypothalamus (VMH) is a critical region of the brain involved in energy homeostasis. TRβ is the predominant TR isoform expressed in the VMH (Cook et al., 1992; Barrett et al., 2007), and previous studies suggest that thyroid hormones acting in the VMH regulate both food intake (Kong et al., 2004) and energy expenditure (López et al., 2010). Thus, we hypothesized that, in the VMH, TRβ physiologically regulates food intake and body weight. To investigate this hypothesis directly, we used stereotaxic Cre-lox gene targeting to generate a VMH-specific model of TRβ knockout in adult mice.

SUMMARY

The obesity epidemic is a significant global health issue. Improved understanding of the mechanisms that regulate appetite and body weight will provide the rationale for the design of anti-obesity therapies. Thyroid hormones play a key role in metabolic homeostasis through their interaction with thyroid hormone receptors (TRs), which function as ligand-inducible transcription factors. The TR-beta isoform (TRβ) is expressed in the ventromedial hypothalamus (VMH), a brain area important for control of energy homeostasis. Here, we report that selective knockdown of TRβ in the VMH of adult mice results in severe obesity due to hyperphagia and reduced energy expenditure. The observed increase in body weight is of a similar magnitude to murine models of monogenic obesity. These data identify TRβ in the VMH as a major physiological regulator of food intake and energy homeostasis.

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RESULTS

Tissue-Specific Knockdown of TRβ in the VMH in Adult Mice

We knocked down TRβ in the VMH of adult male mice using Cre-mediated excision of a floxed critical exon in the Thrb gene. This approach enabled temporally and spatially controlled reduction of TRβ expression specifically in the VMH of adult mice. This model eliminates the developmental consequences and abnormal systemic thyroid hormone levels that occur in global TRβ mutant mice (Ortiga-Carvalho et al., 2014) or in hypothyroid and thyrotoxic animals (Ishii et al., 2003; López et al., 2010).

The Thrb<sup>lox/</sup> allele contains loxP sites flanking exon 5 of Thrb (Winter et al., 2009) (Figure S1A). Cre recombinase-mediated excision of this critical exon results in inactivation of Thrb (Winter et al., 2009). Cre recombinase was introduced into the VMH of adult male Thrb<sup>lox/lox</sup> mice by stereotaxic injection of recombinant adeno-associated virus (rAAV) expressing a Cre-GFP fusion protein to generate mice with reduced TRβ expression in the VMH (VMH-TR<sup>β</sup>FA<sup>C<sup>0</sup></sup>) mice. Thrb<sup>lox/lox</sup> mice injected with rAAV encoding GFP into the VMH (VMH-GFP) were used as controls. Cre-mediated excision of the Thrb<sup>lox</sup> allele was confirmed by PCR of DNA from whole hypothalami of VMH-TRβ<sup>C<sup>0</sup></sup> mice (Figure S1B). The Thrb<sup>lox</sup> allele was not excised in either the cerebellum or brainstem, indicating rAAV did not enter the ventricular system following stereotaxic injection (Figure S1B). Fluorescence microscopy and in situ hybridization (ISH) both confirmed transgene expression localized to the VMH in both groups of mice (Figures S2A and S2B). ISH using a probe specific for the floxed exon of Thrb demonstrated reduced expression within the VMH of VMH-TRβ<sup>C<sup>0</sup></sup> mice compared with controls (Figures S2C and S2D).

Selective Knockdown of TRβ in the VMH in Adult Mice

Results in Hyperphagia and Obesity

VMH-TRβ<sup>C<sup>0</sup></sup> mice consumed more food and gained more weight than controls (Figures 1A and 1B). Weight gain in VMH-TRβ<sup>C<sup>0</sup></sup>
mice was three times greater than that of control mice by the end of the study (Figures 1C and 1D).

Whole hypothalami for RNA-sequencing (RNA-seq) analysis were collected from mice before significant changes in body weight had occurred. This was so that changes in expression are likely to be due to changes in thyroid hormone signaling rather than secondary effects of the increase in body weight and food intake. Differential expression analysis was performed (Table S1). Pathway analysis of differentially expressed genes revealed an over-representation of genes involved in dopamine, growth hormone, and leptin signaling pathways, as well as genes that are involved in neuronal activity regulation including long-term potentiation (LTP) and long-term depression (LTD); these results were qualitatively the same when the false discovery rate (FDR) for analysis was set between 0.001 and 0.1 (Table S2). Among the genes differentially expressed, Pomc expression was decreased (log Fc 0.7, p = 9.33 × 10^-6) (Figure 1F), whereas that of Thrβ was not altered at the level of the whole hypothalamus (Table S1). Expression of steroidogenic factor 1 (Nr5a1), and uncoupling protein-2 (Ucp2), both of which are implicated in hypothalamic control of energy homeostasis (Majdic et al., 2002; Coppola et al., 2007), were unchanged. The differentially expressed genes were compared to those previously reported to be T3 responsive or directly regulated by T3 in cerebrocortical cells (Tables S1 and S2 and Figure S3)(Gil-Ibáñez et al., 2017). Of the genes directly regulated by T3 in cerebrocortical cells, we identified 89 (15%) were also significantly changed in our samples, among which was hairless (Hr). For genes regulated indirectly by T3, we identified 133 that were also changed (9%).

Total, visceral, subcutaneous, and epididymal fat mass were all increased in VMH-TRβ− mice compared to controls (Figures 2A–2E). In keeping with the increased adiposity, VMH-TRβ− mice had a higher plasma leptin concentration than controls (Figure 2F).
Vmh-TRβ Mice Are Systemically Euthyroid
Alterations in circulating thyroid hormones affect food intake and body weight (Pijl et al., 2001). Measurement of plasma TSH, thyroxine (T4), and T3 confirmed that both VMH-TRβ- and control mice were euthyroid (Figures S4A–S4C).

Vmh-TRβ Mice Are Insulin Resistant but Do Not Show Changes in the Expression of Genes Involved in Hypothalamic Glucose Sensing
Obese VMH-TRβ- mice had high levels of fasting insulin (Figure S4D), as expected. However, when glucose tolerance and insulin tolerance were tested before the development of obesity in the VMH-TRβ- mice, there were no differences between the VMH-TRβ- and VMH-GFP mice (Figures S4E and S4F). RNA-seq analysis did not identify changes in expression of hypothalamic glucose-sensing genes.

Vmh-TRβ Mice Fail to Mount an Orexigenic Response to Administered T3
In order to validate loss of T3 signaling following TRβ inactivation in the VMH, we administered T3 to VMH-TRβ- and VMH-GFP mice by subcutaneous injection. Over the 24-hr study period, T3 significantly increased food intake in VMH-GFP mice but VMH-TRβ- mice failed to mount an orexigenic response to the administered T3 (Figure S4I).

VMH-TRβ Mice Do Not Become Obese When Pair-Fed to the Food Intake of Lean Controls
To investigate whether the hyperphagia contributed to, or was a consequence of, the development of the obese phenotype, VMH-TRβ- mice were pair-fed to the food intake of a weight-matched VMH-GFP littermate for 5 weeks. During pair-feeding, there was no difference in cumulative body weight change or food intake (Figures 3A and 3B) or locomotor activity between the two groups. After 5 weeks of pair-feeding, ad libitum access to food was restored for 4 weeks. Following restoration of free feeding, VMH-TRβ- mice gained significantly more weight and consumed significantly more food than controls (Figures 3A and 3C).

VMH-TRβ Mice Have Reduced Energy Expenditure and Reduced Locomotor Activity
The contribution of changes in energy expenditure to the obese phenotype was investigated. Oxygen consumption (VO2), carbon dioxide production (VCO2), and locomotor activity were all decreased during the dark phase in ad libitum-fed VMH-TRβ- mice both before and after the onset of obesity (Figures 4A–4C). By contrast, there was no difference in VO2, VCO2, or locomotor activity during the light phase (Figures 4A–4C). The decrease in nocturnal locomotion in VMH-TRβ- mice was confirmed by behavioral analysis (Table S3). There was no difference in respiratory exchange ratio (RER) (Figure 4D) and no difference in brown adipose tissue (BAT) uncoupling protein-1 (Ucp1) expression (Figure 4E) between VMH-TRβ- and control mice. In addition, VMH-TRβ- mice have a normal body temperature (Figure S4J).

Discussion
These studies identify hypothalamic TRβ as an important physiological regulator of appetite and body weight. Reduced TRβ expression in the VMH resulted in marked weight gain,
comparable to severe forms of monogenic obesity (Tecott et al., 1995; Yaswen et al., 1999). The weight gain was a consequence of increased total body fat, and in particular a marked increase in subcutaneous and visceral white adipose tissue, the latter being an important risk factor for cardiovascular disease and diabetes (Montague and O’Rahilly, 2000).

VMH-TRβ_mice ate more than control animals, and pair-feeding studies indicated that hyperphagia contributed directly to the obesity. Thus, VMH-TRβ_mice remained lean when food intake was restricted but rapidly became obese when ad libitum feeding was restored. Selective TRβ knockdown specifically in the VMH was confirmed by ISH and fluorescence microscopy. Although expression of Thrb was not reduced in the RNA-seq analysis, these samples are derived from whole hypothalami, and therefore the decrease in the level in the VMH is likely masked by the expression of Thrb throughout the rest of the sample. Indeed, the loss of TRβ function in the VMH was demonstrated by the failure of the expected orexigenic response to administered T3 in VMH-TRβ_mice and further supported by the appropriate changes in genes directly regulated by T3. The possibility of the phenotype arising through virus spread to other CNS areas was excluded by rAAV-Cre injection into the lateral ventricles, which did not result in hyperphagia or obesity.

Previous work in rats has reported the acute orexigenic effect of exogenously administered T3 (Kong et al., 2004). Here, we show the endogenous effect of thyroid hormone action following selective TRβ knockdown. We suggest that our current work describes a local circuit within the VMH that physiologically regulates food intake as distinct from the feeding response to administered pharmacological doses of T3 analogous to the contrasting effects of NPY and PYY.

To investigate the underlying cause of hyperphagia in VMH-TRβ_mice, hypothalamic gene expression patterns were determined by RNA-seq. The expression of Pomc and Fto were downregulated in the hypothalamus, whereas Npy was upregulated. POMC and FTO are thought to inhibit food intake, whereas NPY simulates food intake; therefore, these changes in expression may explain in part the phenotype observed.

Energy expenditure in VMH-TRβ_mice was reduced both before and after the onset of obesity. There was no difference in BAT Ucp1 expression between VMH-TRβ− and control mice, suggesting that adaptive thermogenesis was unaffected. It is likely that changes in energy expenditure in VMH-TRβ_mice resulted from decreased locomotor activity. The reduced locomotor activity is not a consequence of the obesity because it occurred before differences in body weight. In addition, during pair-feeding studies, the reduction in locomotor activity was lost.

Figure 4. Energy Expenditure and Locomotor Activity in Mice with Reduced Expression of TRβ in the VMH
(A) Oxygen consumption.
(B) Carbon dioxide production.
(C) Locomotor activity.
(D) RER.
(E) Ucp1 expression in BAT (n = 7 VMH-GFP and 11 VMH-TRβ−).
L, light phase; D, dark phase; 1, 1 week, and 6, 6 weeks, after recovery. Data are mean ± SEM (n = 5 VMH-GFP; n = 6 VMH-TRβ−); ANOVA with Student-Newman-Keuls analysis, **p < 0.01. See also Table S3.
possibly due to food-seeking behavior. This is likely to explain why body weight gain did not differ between the two groups before the restoration of ad libitum feeding. The energy expenditure and pair-feeding data indicate that both increased food intake and reduced locomotor activity contribute to obesity in VMH-TRβ−/− mice.

In contrast to VMH-TRβ−/− mice, global heterozygous TRβ−/− mice do not have an obese phenotype (Ortega-Carvalho et al., 2014). This may be explained by the peripheral hyperthyroidism of these mice. In addition, the appetite circuits within the hypothalamus are subject to developmental plasticity and compensatory redundancy (Bouret et al., 2004; Horvath, 2005). For example, neither global deletion of Agrp and/or Npy nor ablation of arcuate AgRP/NPY neurons in neonatal mice results in a metabolic phenotype (Erickson et al., 1996; Qian et al., 2002; Luquet et al., 2005), whereas ablation of these neurons in adult mice produces profound hypophagia and starvation (Luquet et al., 2005; Gardiner et al., 2005; Bewick et al., 2005). Similar developmental compensation may occur in global TRβ−/− knockout mice.

Studies using adenovirus-mediated expression of a dominant-negative TR (DN-TR) in the rat VMH have been reported (López et al., 2010). Although VMH DN-TR expression did not affect food intake or body weight in euthyroid animals, it prevented weight loss in thyrotoxic rats and resulted in reduced hypothalamic AMP-activated protein kinase (AMPK) expression (López et al., 2010). AMPK expression was unchanged in our model. DN-TR interferes with the actions of both TRα and TRβ and exerts a marked repressive effect on gene transcription (Ortega-Carvalho et al., 2014; Ferrara et al., 2012). By contrast, VMH-TRβ−/− mice have only reduced TRβ activity rather than the pathological repression of TR target genes that is present in animals expressing a dominant-negative receptor. This fundamental difference is likely to explain the contrasting phenotypes observed in these two models.

In summary, we have shown that hypothalamic TRβ is an important physiological regulator of energy homeostasis because TRβ knockdown in the VMH results in a phenotype of hyperphagia and severe obesity that is comparable to some of the most extreme forms of monogenic obesity (Tecott et al., 1995; Yaswen et al., 1999). Our findings provide insights into the central regulation of energy homeostasis by TRβ that could be a target for anti-obesity therapies.

**EXPERIMENTAL PROCEDURES**

**Animals**

VMH-TRβ−/− mice (Winter et al., 2009) were genotyped by PCR using specific oligonucleotide primers (Figure S1). Mice were housed in single cages and maintained under a controlled environment (temperature, 21–23°C; 12-h light–dark cycle, lights on at 07:00) with ad libitum access to chow and water (RM1; SDS Diets), except where stated. Male mice that were 8 weeks old at the start of procedures were used in all experiments. All animal studies were approved under the Animals (Scientific Procedures) Act (1986) (Project Licence Number 70/7229) and approved by the Animal Welfare and Ethical Review Body, Imperial College London, which is signed up to the ARRIVE (Animal Research: Reporting of In Vivo Experiments) guidelines.

**rAAV Preparation**

rAAV was produced (Grimm et al., 1998) and isolated (Zolotukhin et al., 1999), as previously described.

**Confirmation of rAAV Transgene Expression, Thrb Excision, and Reduced TRβ Expression in the VMH**

Excision of the Thrb allele within the hypothalamus was confirmed by PCR (Figure S1). ISH using a probe specific to the excised portion of TRβ was performed to confirm reduced TRβ expression within the VMH (Smith et al., 2008).

**Measurement of Energy Expenditure**

Metabolic parameters were measured by indirect calorimetry using an open-circuit Oxymax system of the Comprehensive Lab Animal Monitoring System (Columbus Instruments) (Gardiner et al., 2010).

**RNA-Seq Analysis**

RNA-seq analysis was performed using hypothalamic RNA from VMH-GFP (n = 3) and VMH-TRβ−/− (n = 4) mice using next-generation sequencing (NGS) technologies (Imperial BRC Genomics Laboratory, Imperial College London). For further details, see Supplemental Experimental Procedures.

**Statistical Analyses**

Cumulative food intake and body weight data were analyzed using generalized estimating equations with exchangeable correlation matrix and robust SEs. Differences between two groups at individual time points were analyzed by unpaired t tests; for multiple comparisons, a Bonferroni correction was applied. Values from the behavioral study were analyzed using a one-way ANOVA followed by Kruskal-Wallis test. Data from the energy expenditure test were analyzed using a one-way ANOVA followed by Newman-Keuls test. Plasma thyroid hormones were compared using Mann-Whitney U test. Differences between groups were considered statistically significant at the 95% confidence level (p < 0.05).

**ACCESSION NUMBERS**

The accession number for the RNA-seq data reported in this paper is GEO: GSE988690.

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes Supplemental Experimental Procedures, four figures, and three tables and can be found with this article online at http://dx.doi.org/10.1016/j.celrep.2017.05.066.

**AUTHOR CONTRIBUTIONS**

W.S.D., S.R.B., J.H.D.B., G.R.W., and J.V.G. conceived of and supervised the project. S.H., M.P., W.S.D., S.A.R., Y.M., C.H., W.F., and J.V.G. conducted the majority of the experiments. S.A.R. and A.G. maintained the mice. A.G. and J.H.D.B. prepared the TRβ probe. J.B. and J.A. performed the MRI study. G.S.H.Y., B.Y.H.L., and J.P.-W. performed the RNA-seq experiments and analysis. J.S. generated the Thrbflx mice. S.H., W.S.D., S.R.B., J.H.D.B., G.R.W., and J.V.G. wrote the manuscript. All authors discussed the results and commented on the manuscript.

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REFERENCES

Barrett, P., Eblinger, F.J., Schuhler, S., Wilson, D., Ross, A.W., Warner, A., Jet-hwa, P., Boelen, A., Visser, T.J., Ozanne, D.M., et al. (2007). Hypothalamic thy-roid hormone catabolism acts as a gatekeeper for the seasonal control of body weight and reproduction. Endocrinology 148, 3608–3617.

Bewick, G.A., Gardiner, J.V., Dhillon, W.S., Kent, A.S., White, N.E., Webster, Z., Ghatel, M.A., and Bloom, S.R. (2005). Post-embryonic ablation of AgRP neu-rons in mice leads to a lean, hypophagic phenotype. FASEB J. 19, 1680–1682.

Bochukova, E., Schoenmakers, N., Agostini, M., Schoenmakers, E., Rajanay-gam, O., Keogh, J.M., Henning, E., Reinemund, J., Gevers, E., Sari, M., et al. (2012). A mutation in the hypothalamic hormone receptor alpha gene. N. Engl. J. Med. 366, 243–249.

Bouret, S.G., Draper, S.J., and Simerly, R.B. (2004). Trophic action of leptin on hypothalamic neurons that regulate feeding. Science 304, 108–110.

Brent, G.A. (2012). Mechanisms of thyroid hormone action. J. Clin. Invest. 122, 3035–3043.

Coll, A.P., Farooqi, I.S., and O’Rahilly, S. (2007). The hormonal control of food intake. Cell 129, 251–262.

Cook, C.B., Kakucska, I., Lechan, R.M., and Koenig, R.J. (1992). Expression of thyroid hormone receptor beta 2 in rat hypothalamus. Endocrinology 130, 1077–1079.

Coppola, A., Liu, Z.W., Andrews, Z.B., Paradis, E., Roy, M.C., Friedman, J.M., Ricquier, D., Richard, D., Horvath, T.L., Gao, X.B., and Diano, S. (2007). A cen-tral thermogenic-like mechanism in feeding regulation: an interplay between arcuate nucleus T3 and UCP2. Cell Metab. 5, 21–33.

Erickson, J.C., Clegg, K.E., and Palmer, R.D. (1996). Sensitivity to leptin and susceptibility to seizures of mice lacking neuropeptide Y. Nature 381, 415–421.

Ferrara, A.M., Onigata, K., Ercan, O., Woodhead, H., Weiss, R.E., and Retefoff, S. (2012). Homozygous thyroid hormone receptor beta gene mutations in resis-tance to thyroid hormone: three new cases and review of the literature. J. Clin. Endocrinol. Metab. 97, 1328–1336.

Gardiner, J.V., Kong, W.M., Ward, H., Murphy, K.G., Dhillon, W.S., and Bloom, S.R. (2005). AAV mediated expression of anti-sense neuropeptide Y cRNA in the arcuate nucleus T3 and UCP2. Cell Metab. 5, 415–421.

Gardiner, J.V., Bataveljic, A., Patel, N.A., Bewick, G.A., Roy, D., Campbell, D., Greenwood, H.C., Murphy, K.G., Hameed, S., Jetinha, P.H., et al. (2010). Pro-kineticin 2 is a hypothalamic neuropeptide that potently inhibits food intake. Diabetes 59, 397–406.

Gil-Ibañez, P., Garcia-Garcia, F., Dopazo, J., Bernal, J., and Morte, B. (2017). Global transcriptome analysis of primary cerebrocortical cells: identification of genes regulated by triiodothyronine in specific cell types. Cereb. Cortex 27, 706–717.

Grimm, D., Kem, A., Rittner, K., and Kleinschmidt, J.A. (1998). Novel tools for production and purification of recombinant adenovirus vectors. Hum. Gene Ther. 9, 2745–2760.

Horvath, T.L. (2005). The hardship of obesity: a soft-wired hypothalamus. Nat. Neurosci. 8, 561–565.

Ishii, S., Kamegai, J., Tamura, H., Shimizu, T., Sugihara, H., and Oikawa, S. (2003). Hypothalamic neuropeptide Y/Y1 receptor pathway activated by a reduction in circulating leptin, but not by an increase in circulating ghrelin, con-

tributes to hyperphagia associated with triiodothyronine-induced thyrotoxi-
cosis. Neuroendocrinology 78, 321–330.

Kim, B. (2008). Thyroid hormone as a determinant of energy expenditure and the basal metabolic rate. Thyroid 18, 141–144.

Kong, W.M., Martin, N.M., Smith, K.L., Gardiner, J.V., Connoley, I.P., Stephens, D.A., Dhillon, W.S., Ghatel, M.A., Small, C.J., and Bloom, S.R. (2004). Triiodothyronine stimulates food intake via the hypothalamic ventromedially-nucleus independent of changes in energy expenditure. Endocrinology 145, 5252–5258.

Liu, Y.Y., Schultz, J.J., and Brent, G.A. (2003). A thyroid hormone receptor alpha gene mutation (P398H) is associated with visceral adiposity and impaired catecholamine-stimulated lipolysis in mice. J. Biol. Chem. 278, 38913–38920.

López, M., Varela, L., Vázquez, M.J., Rodríguez-Cuenca, S., González, C.R., Velagapudi, V.R., Morgan, D.A., Schoenmakers, E., Agassandian, K., Lage, R., et al. (2010). Hypothalamic AMPK and fatty acid metabolism mediate thy-
roid regulation of energy balance. Nat. Med. 16, 1001–1008.

Luquet, S., Perez, F.A., Hnasko, T.S., and Palmer, R.D. (2005). NPY/Agrp knock-out neurons are essential for feeding in adult mice but can be ablated in neonates. Science 310, 683–685.

Madjic, G., Young, M., Gomez-Sanchez, E., Anderson, P., Szczepaniak, L.S., Dobbins, R.L., McGarry, J.D., and Parker, K.L. (2002). Knockout mice lacking steroidogenic factor 1 are a novel genetic model of hypothalamic obesity. Endocrinology 143, 607–614.

Mitchell, C.S., Savage, D.B., Dufour, S., Schoenmakers, N., Murgatroyd, P., Befroy, D., Hallsal, S., Northcott, S., Raymond-Barker, P., Curran, S., et al. (2010). Resistance to thyroid hormone is associated with raised energy expendi-
ture, muscle mitochondrial uncoupling, and hyperphagia. J. Clin. Invest. 120, 1345–1354.

Montague, C.T., and O’Rahilly, S. (2000). The perils of portliness: causes and consequences of visceral adiposity. Diabetes 49, 883–888.

Moran, C., Schoenmakers, N., Agostini, M., Schoenmakers, E., Offiah, A., Kydd, A., Kahaly, G., Mohr-Kahaly, S., Rajanayagam, O., Lyons, G., et al. (2013). An adult female with resistance to thyroid hormone mediated by defective thyroid hormone receptor α. J. Clin. Endocrinol. Metab. 98, 4254–4261.

Ortega-Carvalho, T.M., Sidhaye, A.R., and Wondisford, F.E. (2014). Thyroid hormone receptors and resistance to thyroid hormone disorders. Nat. Rev. En-
docrinol. 10, 582–591.

Pijl, H., de Meijer, P.H., Langius, J., Coenegracht, C.I., van den Berk, A.H., Chandie Shaw, P.K., Boom, H., Schoenmakers, R.C., Cohen, A.F., Burggraaf, J., and Meinders, A.E. (2001). Food choice in hyperthyroidism: potential influence of the autonomic nervous system and brain serotonin precursor availability. J. Clin. Endocrinol. Metab. 86, 5848–5853.

Qian, S., Chen, H., Weingarth, D., Trumbauer, M.E., Novi, D.E., Guan, X., Yu, H., Shen, Z., Feng, Y., Frazier, E., et al. (2002). Neither agouti-related protein nor neuropeptide Y is critically required for the regulation of energy homeosta-
sis in mice. Mol. Cell. Biol. 22, 5027–5035.

Sjogren, M., Alkemade, A., Mittag, J., Nordstro¨m, K., Katz, A., Rozell, B., Westerblad, H., Arner, A., and Vennstrom, B. (2007). Hypermetabolism in mice caused by the central action of an unliganded thyroid hormone receptor α1. EMBO J. 26, 4535–4545.

Smith, K.L., Gardiner, J.V., Ward, H.L., Kong, W.M., Murphy, K.G., Martin, N.M., Ghatel, M.A., and Bloom, S.R. (2008). Overexpression of CART in the PVN increases food intake and weight gain in rats. Obesity (Silver Spring) 16, 2239–2244.


Tecott, L.H., Sun, L.M., Akana, S.F., Strack, A.M., Lowenstein, D.H., Dallman, M.F., and Julius, D. (1995). Eating disorder and epilepsy in mice lacking 5-HT2c serotonin receptors. Nature 374, 542–546.

Winter, H., Rüttiger, L., Müller, M., Kuhn, S., Brandt, N., Zimmermann, U., Hirt, B., Bress, A., Sausbier, M., Conscience, A., et al. (2009). Deafness in TRbeta mutants is caused by malformation of the tectorial membrane. J. Neurosci. 29, 2581–2587.

Yaswen, L., Diehl, N., Brennan, M.B., and Hochgeschwender, U. (1999). Obesity in the mouse model of pro-opiomelanocortin deficiency responds to peripheral melanocortin. Nat. Med. 5, 1066–1070.

Zolotukhin, S., Byrne, B.J., Mason, E., Zolotukhin, I., Potter, M., Chesnut, K., Summerford, C., Samulski, R.J., and Muzyczka, N. (1999). Recombinant adeno-associated virus purification using novel methods improves infectious titer and yield. Gene Ther. 6, 973–985.