Genome-wide screen for universal individual identification SNPs based on the HapMap and 1000 Genomes databases

Erwen Huang1,2, Changhui Liu1, Jingjing Zheng1,2, Xiaolong Han1,2,3, Weian Du4, Yuanjian Huang5, Chengshi Li6,2, Xiaoguang Wang1,2, Dayue Tong1,2, Xueling Ou1,2, Hongyu Sun1,2, Zhaoshu Zeng6 & Chao Liu1,2,3

Differences among SNP panels for individual identification in SNP-selecting and populations led to few common SNPs, compromising their universal applicability. To screened all universal SNPs, we performed a genome-wide SNP mining in multiple populations based on HapMap and 1000Genomes databases. SNPs with high minor allele frequencies (MAF) in 37 populations were selected. With MAF from ≥0.35 to ≥0.43, the number of selected SNPs decreased from 2769 to 0. A total of 117 SNPs with MAF ≥ 0.39 have no linkage disequilibrium with each other in every population. For 116 of the 117 SNPs, cumulative match probability (CMP) ranged from $2.01 \times 10^{-48}$ to $1.93 \times 10^{-50}$ and cumulative exclusion probability (CEP) ranged from 0.9999999996653 to 0.9999999999945. In 134 tested Han samples, 110 of the 117 SNPs remained within high MAF and conformed to Hardy-Weinberg equilibrium, with CMP = $4.70 \times 10^{-47}$ and CEP = 0.999999999862. By analyzing the same number of autosomal SNPs as in the HID-Ion AmpliSeq Identity Panel, i.e. 90 randomized out of the 110 SNPs, our panel yielded preferable CMP and CEP. Taken together, the 110-SNPs panel is advantageous for forensic test, and this study provided plenty of highly informative SNPs for compiling final universal panels.

It has been well recognized that single nucleotide polymorphisms (SNPs) are potentially valuable for DNA profiling in forensics. The advantages of SNP characteristics serving as forensic markers include: (1) much higher density interspersing in the whole genome providing more selectable loci to offset the defect of only two alleles at each locus. (2) Shorter length of amplified fragment, facilitating the amplification of degraded DNA samples. (3) Lower mutation rate, endowing them with superiority in paternity and immigration testing. (4) Faster and cheaper high-throughput approaches available for genotyping. Over the past decade, studies on population genetics and forensic application of SNPs were performed. For example, Vallone et al. investigated the allele frequencies for 70 autosomal SNPs in U.S. Caucasian, African-American, and Hispanic populations. Kidd Lab studied hundreds of SNPs in more than 40 populations. Moreover, several applicable panels were developed in recent years. In retrospect, these studies selected different candidate SNPs or populations, resulting in few common loci across these panels. For instance, there are just 4 shared SNPs in the SNP/forID 52 and IISNP panels, which are the best known and serve as the basis for several commercial kits, e.g. the HID-Ion AmpliSeq Identity Panel (Thermo Fisher Scientific) and the ForenSeq DNA Signature Prep Kit (https://support.illumina.com/downloads/forenseq-dna-signature-prep-guide-15049528.html). Among different ethnic populations, the vast majority of

---

1Faculty of Forensic Medicine, Zhongshan School of Medicine, Sun Yat-Sen University, Guangzhou, 510080, China. 2Guangdong Province Translational Forensic Medicine Engineering Technology Research Center, Sun Yat-Sen University, Guangzhou, 510080, China. 3Guangzhou Forensic Science Institute, Guangzhou, 510030, China. 4School of Forensic Medicine, Southern Medical University, Guangzhou, 510515, China. 5Homy Genetech Inc. Guangdong, Foshan, 52800, China. 6Department of Forensic Medicine, School of Basic Medical Sciences, Zhengzhou University, Zhengzhou, 450001, China. Erwen Huang, Changhui Liu, Jingjing Zheng and Xiaolong Han contributed equally to this work. Correspondence and requests for materials should be addressed to H.S. (email: sunhongyu2002@163.com) or Z.Z. (email: zs_zeng@yahoo.com) or C.L. (email: liuchaogzf@163.com)
SNPs in human genome differs in minor allele frequency (MAF) and linkage disequilibrium (LD) properties, compromising to some extent the universal applicability of the panels.

To screen such common SNPs as many as possible, in this study we performed a genome-wide screen through 25,580,678 SNPs based on the databases of HapMap r28 (released in Aug 2010) and 1000 Genomes Phase 3 (released in May 2015). We collected all of the SNPs with different threshold values of MAF, and evaluated those with MAF ≥ 0.39 for their forensic genetic parameters. We also experimentally analyzed 117 independent SNPs with MAF ≥ 0.39 in two other populations — Chinese Han in Guangzhou (CHG, N = 96) and Chinese Han in Zhengzhou (CHZ, N = 38).

### Results

#### Selection of universal highly informative SNPs.

Based on the HapMap bulk data r28, we screened 4,402,208,440 genotypes at 25,580,678 SNPs in the whole genome. At the criterion of MAF ≥ 0.35, the number of SNPs shared by the 11 populations was 13466. When the threshold of MAF was set at ≥ 0.38–0.44, the number of shared SNPs ranged from 3881 to 21. When MAF was elevated up to 0.45, no shared SNP was retained (Table 1). After further screening through the 1000 Genomes populations (26 totally), the number of shared SNPs in the 37 populations was: 2769, 433, 169, 62, 12, 1, at the criterion of MAF ≥ 0.35, 0.38, 0.39, 0.40, 0.41 or 0.42, respectively; and no SNP was retained when the MAF was raised to 0.43. (Tables 2, S1). A few SNPs (rs3125289, ...
rs311870 and rs7176637) are triallelic and the MAF is lower than 0.38, but both of the high-frequency alleles have a frequency not lower than 0.38. Such SNPs were also included in the panel of MAF ≥0.38 (Table S1). It caught our attention that in the panel of MAF ≥0.35, only one SNP (rs891700) was shared by the SNP for ID52 12, and only seven (rs13218440, rs1872575, rs1554472, rs1498553, rs891700, rs1019029 and rs2291395) were shared by the IISNP panel composed of 92 SNPs developed in the Kidd Lab 13, 17 (Table S1).

Genetic investigation of the SNPs with MAF ≥0.39. We studied the population genetic profiles of the SNPs in the panel of MAF ≥0.39, which includes 169 SNPs. Significant deviation of genotype frequency from expectations was observed in 366 out of 6253 HWE tests in the 37 populations (P < 0.05, Table S2). After the Bonferroni’s correction, the deviation remained in 34 tests (P < 0.0003, Table S2). There was no LD between at least 117 out of the 169 SNPs in each of the populations (r² < 0.05, Table S3). We selected 117 independent SNPs (including one X-linked SNP rs722847, and only one of those with r² ≥ 0.05 between each other in any of the populations was selected) to evaluate forensic parameters (Table S3). For the 116 autosomal SNPs, MP ranged from 0.333 (rs7127767 in HapMap-MEX) to 0.529 (rs7561460 in 1000 Genomes-PEL), and CMP ranged from 2.01 × 10⁻⁴⁸ in HapMap-ASW to 1.93 × 10⁻⁵⁰ in 1000 Genomes-STU (Table S4). EP ranged from 0.012 (rs2624459 in HapMap-IPT) to 0.419 (rs7561460 in 1000 Genomes-PEL), and CEP ranged from 0.9999999996653 in 1000 Genomes-STU to 0.9999999999945 in 1000 Genomes-ASW (Table S4). For the X-linked SNP rs722847, calculated from the genotypes in females, MP ranged from 0.336 in HapMap-MKK to 0.445 in 1000 Genomes-CHS, and EP was between 0.030 in HapMap-CHB and 0.295 in 1000 Genomes-CHS (Table S4).

Experimental studies on population genetics of the 117 SNPs with a MAF ≥0.39. We further investigated the 117 SNPs in two Chinese Han groups including CHG and CHZ. In the groups, 112 out of the 117 SNPs were successfully genotyped among all the samples (genotyping rate: 100%), 3 SNPs (rs508485, rs530913, and rs10451160) were with a genotyping rate of 99.3%, and 2 SNPs (rs6136874 and rs10503926) were 97.8%. A total of 89 SNPs remained a MAF of ≥0.39 in both groups (Table S5), as did 109 when the two groups were pooled (CHP, Table S5). Four SNPs (rs6431272, rs431951, rs4469483, rs4487849) exhibited significant deviation of genotype frequency from HWE expectations after Bonferroni’s correction in CHG, CHZ and CHP (P < 0.000428, Table S6), as did the other two SNPs (rs10451160 and rs7710223) in CHG and CHP, respectively. The observed range of heterozygosities was 0.39583–0.68750, 0.34211–0.65789, 0.41791–0.65672, respectively in CHG, CHZ and CHP, except for the SNPs deviating significantly from HWE expectations (Table S6). Out of 40716 pairwise comparisons, 2040 pairs showed significant LD (data not shown). All of the LDs were most likely due to chance, because the paired SNPs locate on different chromosomes, or on the same chromosomes but there are other SNPs showing no LD between them.

Results of AMOVA analysis showed that the global genetic variation in CHG and CHZ could be explained by individual variability (Table S7), suggesting the reasonability of combining CHG and CHZ. Moreover, the analysis of genetic distance for the 110 autosomal SNPs without significant deviation from HWE revealed tiny variations between all of the studied populations (Table S8, Fig. 1), proving the universality of these SNPs.

We further selected the 110 autosomal SNPs for calculating forensic parameters. MP ranged from 0.334 (rs12819145 and rs1293288 in CHZ) to 0.494 (rs2624459 in CHZ) (Table S9), and CMP ranged from 4.70 × 10⁻⁴⁷ in CHP to 8.37 × 10⁻⁴⁶ in CHZ (Tables 3 and S9). EP ranged from 0.082 (rs2895309, rs12819145, rs1293288...
and rs2380437 in CHZ) to 0.366 (rs2624459 and rs4607417 in CHZ) (Table S9), and CEP ranged from 0.99999999862 in CHP to 0.99999999939 in CHZ (Tables 3 and S9). The commercialized HID-Ion AmpliSeq Identity Panel (currently named Precision ID Identity Panel) includes 90 autosomal SNPs and was recently evaluated in Chinese Han population16. CMP of this panel and the PowerPlex Fusion STR System were 1.318 × 10⁻³³ and 2.147 × 10⁻²⁵, respectively; and CEP were 0.999999724 and 0.9999999673, respectively (Table 4)16. Here, in CHP, CMP calculated for 90 randomized out of the 110 SNPs with a MAF of ≥0.39 ranged from 3.071 × 10⁻³⁹ to 5.368 × 10⁻³⁹, at least 10⁻⁵ lower than that achieved with these two commercial panels; CEP ranged from 0.9999999728 to 0.99999999736, higher than that of HID-Ion AmpliSeq Identity Panel and lower than that of PowerPlex Fusion STR System (Table 4). These results suggest that the MAF ≥0.39 panel would have obvious advantages in forensic application over HID-Ion AmpliSeq Identity Panel. When the 110 SNPs are all included, it would have overall advantage than PowerPlex Fusion STR System.

### Discussion

An expanded genome-wide search in this study contributes to the identification of more and probably better forensic markers. Our results indicated that high-throughput SNPs databases can provide convenient, efficient and cost-saving approaches to select highly informative SNPs for forensic purposes. With these approaches, this study obtained several panels with such candidate SNPs, which have potential to be used to develop final forensic SNP panels with universal applicability in a variety of ethnic populations. Until now, three individual identification SNP kits are commercially available, including the HID-Ion AmpliSeq Identity Panel16,18, the ForenSeq DNA Signature Prep Kit19 and the Qiagen SNP-ID Kit20. They are all mainly based on the SNP Kidd IISNP panels13,17, which have no SNP in common with the MAF ≥0.39 panel, and just 8 with the MAF ≥0.35 panel (Tables S1 and S9). Comparison shown in Table 4 indicated that the MAF ≥0.39 panel is potentially advantageous in forensic application.

A most recent study by Li et al21 also performed a genome-wide screening by selecting SNPs with Fst ≤0.01 and heterozygosity ≥0.40, based on HapMap r24, which includes 4 populations. It proposed a final panel encompassing 175 SNPs, compiled based on 26 populations (four in HapMap r24, thirteen in 1000 Genomes phase 1, nine Chinese populations including five Han). A certain MAF/heterozygosity value was not set as a screening criterion after the initial screening in HapMap r24. As a result, MAF values of these 175 SNPs differed greatly: in the range of 0.08–0.50 in 1000 Genomes phase 3, 0.20–0.50 in the nine Chinese populations, and allele frequencies for 69 out of the 175 SNPs were not available in HapMap r28. In our study, MAF ≥0.35, 0.38, 0.39, 0.40, 0.41, 0.42, … was set as a screening criterion in all of the HapMap r28 (includes all of the 4 populations in r24) and 1000 Genomes phase 3 populations. The selected loci were all high polymorphic in all of the studied populations. Consequently, the final panel in Li’s study21 includes just 7 SNPs in common with our MAF ≥0.35 panel.

In conclusion, our study provided several semi-finished panels, which are convenient for researchers to select candidate high polymorphic SNPs to further test in more ethnic lines for the purpose of compiling final universal panels.

### Methods

#### Genome-wide screening for highly informative SNPs.

Bulk data of the whole genome-genotyped SNP allele frequencies of the HapMap Public Release #28 were downloaded from the website [http://hapmap.ncbi.nlm.nih.gov/downloads/frequencies/2010-08_phaseII+III/](http://hapmap.ncbi.nlm.nih.gov/downloads/frequencies/2010-08_phaseII+III/). These data include genotyping results of the whole genome-wide SNPs in 1301 individuals from 11 ethnic populations. A method described in our previous studies was used to select candidate SNPs13,15. Briefly, frequency files in.gz format were unzipped and read by Microsoft Excel 2007 (Microsoft Corp., Redmond, WA). Rs numbers corresponding to SNPs at certain MAF criterion (i.e., MAF ≥0.45, 0.44, 0.43, 0.42, 0.41, 0.40, 0.39, 0.38, 0.35) were listed in a new.xlsx file containing one column for each population. Rs numbers shared by all 11 columns were screened out by a program (for program codes, please refer to Supplementary materials in Table S10).

---

**Table 3.** Forensic parameters calculated for 110 SNPs in CHG, CHZ and CHP. CHG, Chinese Han in Guangzhou; CHZ, Chinese Han in Zhengzhou; CHP, the pooled population with CHG and CHZ.

| Population | CHG | CHZ | CHP |
|------------|-----|-----|-----|
| CMP        | 9.220 × 10⁻⁵⁻ | 8.366 × 10⁻⁴⁻ | 4.697 × 10⁻⁵⁻ |
| CEP        | 0.999999999896 | 0.99999999939 | 0.999999999862 |

**Table 4.** Forensic parameters calculated for two commercialized panels and 90 randomized SNPs included in the MAF ≥0.39 panel in Chinese Han population.

| Panel MAF ≥0.39 panel | HID-Ion AmpliSeq Identity Panel16 | PowerPlex Fusion STR System16 |
|-----------------------|-----------------------------------|-----------------------------|
| Number of autosomal loci | 90 | 90 | 22 |
| Number of individuals | 134 | 108 | 108 |
| CMP | 3.071 × 10⁻³⁹⁻⁵⁻⁻, 5.368 × 10⁻³⁹⁻ | 1.318 × 10⁻³³⁻ | 2.147 × 10⁻²⁵⁻ |
| CEP | 0.9999999728⁻⁻⁻, 0.9999999736⁻⁻⁻ | 0.999999724⁻⁻⁻ | 0.9999999673⁻⁻⁻ |
All of the selected SNPs were further screened with the same MAF value in the database of 1000 Genomes Phase 3 at http://browser.1000genomes.org/index.html. The database includes genotyping results of the whole genome-wide SNPs in 2504 individuals from 26 ethnical populations. All of the populations studied were listed in Table S10. SNPs shared by all of the 26 populations at a MAF criterion were selected.

Ethnics. Blood samples were collected from the CHZ and CHG populations upon approval of the Ethics Committee at Zhongshan School of Medicine, Sun Yat-Sen University. Informed consent was obtained from all participants. All the experimental procedures were carried out in accordance with the approved guidelines of Zhongshan School of Medicine, Sun Yat-Sen University. This study was approved by the Ethics Committee of Zhongshan School of Medicine, Sun Yat-Sen University.

Genotyping assay. Genotypes of the 117 SNPs without LD with each other in the panel of MAF ≥0.39 were analyzed using the MassARRAY Genetic Analysis System (Sequenom, San Diego, California, USA). Biochemical reactions were performed in four wells. Primers were designed using the software AssayDesigner version 3.1. Reaction reagents and program were used as we previously described34. Briefly, the polymerase chain reaction (PCR) volume was composed of 0.5 μL of 10 × PCR buffer, 0.4 μL of 25 mM MgCl2, 1 μL of 0.5 μM primer Mix, 0.1 μL of 25 mM dNTP Mix, 1 μL of 10 ng/μL DNA template, 0.2 μL of 5 U/μL HotStar Taq DNA polymerase and 1.8 μL of water. The SAP reaction volume contained 0.17 μL of SAP buffer, 0.3 μL of 1.7 U/μL SAP enzyme, 1.5 μL of nanopure water. The extension reaction reagents included 0.2 μL of iPLEX Buffer Plus, 0.2 μL of iPLEX Termination Mix, 0.94 μL of iPLEX Extend Primer Mix, 0.041 μL of iPLEX Enzyme, 0.619 μL of nanopure water, 7 μL of PCR + SAP product. The extension reaction program was set as: initial denaturing in 94 °C for 30 s, then denaturing in 94 °C for 5 s, five cycles (we termed mini-cycle) of annealing in 52 °C for 5 s and extension in 80 °C for 5 s. Totally 40 cycles of the denaturing and mini-cycle, followed by final extension in 72 °C for 3 min. Twenty samples were randomly selected for Sanger sequencing to verify the genotyping results. All of the sequencing results were consistent with that of the MassARRAY method.

Statistical analyses. Results of LD analysis of SNPs in the 1000 Genomes populations were online searched at http://www.ensembl.org/Homo_sapiens/Variation. Deviations of genotype frequencies from Hardy-Weinberg equilibrium (HWE) expectations for intra-population were tested using the modified PowerStats (Promega, Madison, WI, USA). Forensic parameters such as random match probability (MP), cumulative random match probability (CMP), exclusion probability (EP), cumulative exclusion probability (CEP) were estimated also using the modified PowerStats. Similar statistical analyses for experimental populations were performed using the Arlequin version 3.525. A two-hierarchical AMOVA analysis was performed to study the degree of genetic heterogeneity between the experimental populations. A multidimensional scaling analysis was performed from genetic distance using the SPSS version 22.0.

References
1. Genomes Project, C. et al. An integrated map of genetic variation from 1,092 human genomes. Nature 491, 56–65 (2012).
2. Genomes Project, C. et al. A global reference for human genetic variation. Nature 526, 68–74 (2015).
3. Sanchez, J. J. & Endicott, P. Developing multiplexed SNP assays with special reference to degraded DNA templates. Nat. Protoc. 1, 1370–1378 (2006).
4. Nachman, M. W. & Crowell, S. L. Estimate of the mutation rate per nucleotide in humans. Genetics 156, 297–304 (2000).
5. Kondrashov, A. S. Direct estimates of human per nucleotide mutation rates at 20 loci causing Mendelian diseases. Hum. Mutat. 21, 12–27 (2003).
6. Hakenberg, J. et al. A SNPshot of PubMed to associate genetic variants with drugs, diseases, and adverse reactions. J. Biomed. Inform. 45, 842–850 (2012).
7. Lopes, J. S. et al. SNP typing reveals similarity in Mycobacterium tuberculosis genetic diversity between Portugal and Northeast Brazil. Infect. Genet. Evol. 18, 238–246 (2013).
8. Pereira, V., Mogensen, H. S., Borsting, C. & Morling, N. Evaluation of the Precision ID Ancestry Panel for crime case work: A SNP typing assay developed for typing of 165 ancestral informative markers. Forensic Sci. Int. Genet. 28, 138–145 (2017).
9. Vallone, P. M., Decker, A. E. & Butler, J. M. Allele frequencies for 70 autosomal SNP loci with U.S. Caucasian, African-American, and Hispanic samples. Forensic Sci. Int. 149, 279–286 (2005).
10. Kud, K. K. et al. Developing a SNP panel for forensic identification of individuals. Forensic Sci. Int. 164, 20–32 (2006).
11. Pakstis, A. J., Speed, W. C., Kidd, J. R. & Kud, K. K. Candidate SNPs for a universal individual identification panel. Hum. Genet. 121, 305–317 (2007).
12. Sanchez, J. J. et al. A multiplex assay with 52 single nucleotide polymorphisms for human identification. Electrophoresis 27, 1713–1724 (2006).
13. Pakstis, A. J. et al. SNPs for a universal individual identification panel. Hum. Genet. 127, 315–324 (2010).
14. Borsting, C., Fordyce, S. L., Olafsson, J., Mogensen, H. S. & Morling, N. Evaluation of the Ion Torrent HD SNP 169-plex: A SNP typing assay developed for human identification by second generation sequencing. Forensic Sci. Int. Genet. 12, 144–154 (2014).
15. Jager, A. C. et al. Developmental validation of the MiSeq FGx Forensic Genomics System for Targeted Next Generation Sequencing in Forensic DNA Casework and Database Laboratories. Forensic Sci. Int. Genet. 28, 52–70 (2017).
16. Guo, F. et al. Next generation sequencing of SNPs using the HID-Ion AmpliSeq Identity Panel on the Ion Torrent PGM platform. Forensic Sci. Int. Genet. 25, 73–84 (2016).
17. Kud, K. K. et al. Expanding data and resources for forensic use of SNPs in individual identification. Forensic Sci. Int. Genet. 6, 646–652 (2012).
18. Zhang, S. et al. Parallel Analysis of 124 Universal SNPs for Human Identification by Targeted Semiconductor Sequencing. Sci. Rep. 22, 18683 (2015).
19. Churchill, J. D., Schmedes, S. E., King, J. L. & Budowle, B. Evaluation of the Illumina Beta Version ForenSeq DNA Signature Prep Kit for use in genetic profiling. Forensic Sci. Int. Genet. 20, 20–29 (2016).
20. de la Puente, M. et al. Evaluation of the Quagen 140-SNP forensic identification multiplex for massively parallel sequencing. Forensic Sci. Int. Genet. 28, 35–43 (2017).
21. Li, L. et al. Genome-wide screening for highly discriminative SNPs for personal identification and their assessment in world populations. Forensic Sci. Int. Genet. 28, 118–127 (2017).
22. Zeng, Z. et al. Evaluation of 96 SNPs in 14 populations for worldwide individual identification. J. Forensic Sci. 57, 1031–1035 (2012).
23. Zeng, Z. S. et al. Genome-wide Screen for Individual Identification SNPs (IISNPs) and the Confirmation of Six of Them in Han Chinese with Pyrosequencing Technology. J. Forensic Sci. 55, 901–907 (2010).
24. Huang, E. W. et al. Investigation of associations between ten polymorphisms and the risk of coronary artery disease in Southern Han Chinese. J. Hum. Genet. 61, 389–393 (2016).
25. Excoffier, L. & Lischer, H. É. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. Mol. Ecol. Resour. 10, 564–567 (2010).

Acknowledgements
We thank professor Jiangwei Yan for excellent direction of statistical analysis. This study was supported by grants (No. 2014A030313127) from the Natural Science Foundation of Guangdong Province.

Author Contributions
E.H., C.L. and H.S. designed the study. C.L. and X.H. performed the statistic analyses. E.H., J.Z., W.D. and Y.H. did the experiments. C.L. and X.W. collected the 1000 Genomes data. D.T. and X.O. provided the samples of CHZ and CHG. Z.Z. did the programing and used it to screen the HapMap database. All authors reviewed the manuscript.

Additional Information
Supplementary information accompanies this paper at https://doi.org/10.1038/s41598-018-23888-0.

Competing Interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2018