Analysis of the Relationship Between ADIPOR1 Variants and the Susceptibility of Chronic Metabolic Diseases in a Northeast Han Chinese Population

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Objective: Shared genetic variants in ADIPOR1 have been identified as closely related to coronary artery disease (CAD), type 2 diabetes (T2D), and T2D with CAD susceptibility, suggesting that these variants are strong candidates for the common soil hypothesis. Therefore, it is essential to analyze the relationship between ADIPOR1 variants and the susceptibility to CAD, T2D, and T2D with CAD in other populations.

Materials and Methods: A case–control study was conducted which included three case cohorts [CAD (n = 316), T2D (n = 295), T2D with CAD (n = 302)], and a control cohort (n = 268) from a population in northeast China. Six ADIPOR1 single-nucleotide polymorphisms were genotyped by high-resolution melting and polymerase chain reaction–restriction fragment length polymorphism.

Results: We confirmed that the shared variant, rs3737884*G, in ADIPOR1 is associated with CAD, T2D, and T2D with CAD (p-value range: 6.54E-6–1.82E-5, odds ratio [OR] range: 1.770–1.844) and that rs16850797*C is associated with T2D and T2D with CAD (p-value range: 0.001–0.001, OR range: 1.529–1.571). We also found that a novel shared variant, rs7514221*C, is associated with an increased susceptibility to CAD, T2D, and T2D with CAD (p-value range: 0.002–0.004, OR range: 1.194–2.382) in this population. Conclusions: ADPOR1 variants, rs3737884*G and rs7514221*C, may be shared risk factors associated with CAD, T2D, and T2D with CAD in a population of northeast China.

Introduction

Coronary artery disease (CAD) and type 2 diabetes (T2D) are both chronic metabolic diseases triggered by several common factors and shared polygenic variants with high prevalence and morbidity (Alberti and Zimmet, 1998; King et al., 1998). To our surprise, CAD is the leading cause of mortality and morbidity in patients with T2D and accounts for up to 80% of deaths in patients with this disorder (Doria, 2010). It has been hypothesized that T2D and CAD share a common genetic basis (Dong et al., 2014), and the adiponectin signaling-related gene has been postulated to play critical roles in this scenario (Tao et al., 2014).

Adiponectin is a functionally active adipokine that regulates glucose and lipid metabolism. The metabolic effects and biological function of adiponectin are mainly mediated by adiponectin receptor 1 (ADIPOR1) (Kadowaki and Yamauchi, 2011). So, ADIPOR1 plays an important role in indirectly regulating glucose and lipid metabolism in chronic metabolic diseases (Yamauchi et al., 2014). The human ADIPOR1 gene is located at chromosome 1p36.13-q41 and presents several polymorphisms (Yamauchi et al., 2014). Some of these polymorphisms have been shown to be associated with an increased risk of developing several diseases, including obesity (Lacinov et al., 2007), metabolic syndrome (Peters et al., 2013), diabetes (Qi et al., 2007), cardiovascular disease (Cox et al., 2013), gastric cancer (Shin et al., 2013), colorectal cancer (Liu et al., 2011), and prostate cancer (Kaklamani et al., 2011). However, to the best of our knowledge, there is only one report to date on the association of ADIPOR1 variants with CAD, T2D, and T2D with CAD risk among the northern Han Chinese population (Jin et al., 2014). Due to differences of genetic background, gene variation association studies may vary among populations in different populations.
allele frequencies and linkage disequilibrium (LD) structures. It is important to examine multiple ethnic populations for the identification of ethnicity-specific loci as well as common susceptibility loci (Deschamps et al., 2015).

Therefore, we conducted a study assessing the association of ADIPOR1 variants with CAD, T2D, and T2D with CAD in a northeast Han Chinese population. The study may lay a theoretical foundation for the common soil hypothesis.

Materials and Methods

Ethics statement

All participants agreed to the protocol of this study and provided written informed consent. The study protocol was approved by the local research ethics committee.

Participants

The study population included patients who self-identified as having a Han ethnic origin and permanent residents of the Jiamusi Heilongjiang area in northeast China. We enrolled a total of 1181 subjects containing 316 CAD, 295 T2D, and 302 T2D with CAD patients, as well as 268 healthy controls between October 2014 and May 2015. T2D was diagnosed according to World Health Organization criteria (Alberti and Zimmet, 1998), while classification of CAD patients was based on previous studies (Qi et al., 2013). The patients in the T2D with CAD group met both of the above inclusion criteria. The control group comprised healthy individuals who had no dyslipidemia and abnormal glucose tolerance, as well as family history of CAD or T2D in first-degree relatives. The exclusion criteria were the same as the criteria of the previous study (Jin et al., 2014).

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Materials and Methods

Table 1. The Allele and Genotypic Frequencies of ADIPOR1 SNPs

| SNP     | Control (n = 268) | CAD (n = 316) | T2D (n = 295) | T2D with CAD (n = 302) |
|---------|------------------|---------------|---------------|------------------------|
|         | n (%) p          | n (%) p/OR (95% CI) | n (%) p/OR (95% CI) | n (%) p/OR (95% CI) |
| rs7539542 |                  |               |               |                        |
| C       | 199 (37.1) 0.784 | 220 (34.8) 0.410 | 215 (36.4) 0.81 | 210 (34.8) 0.407 |
| G       | 337 (62.9)      | 412 (65.2) 0.904 | 375 (63.6) 0.970 | 394 (65.2) 0.902 |
| CC      | 45 (16.8) 0.165 | 34 (10.8) 0.711 | 36 (12.2) 0.761 | 43 (14.2) 0.708 |
| CG      | 109 (40.7)      | 152 (48.1) 0.056 | 143 (48.5) 0.115 | 124 (41.1) 0.684 |
| GG      | 114 (42.5)      | 130 (41.1)    | 116 (39.3)    | 135 (44.7)    |
| rs3737884 |                  |               |               |                        |
| G       | 362 (67.5) 9.47E-7 | 497 (78.6) 1.82E-5 | 468 (79.3) 7.24E-6 | 479 (79.3) 6.54E-6 |
| A       | 174 (32.5)      | 135 (21.4) 1.770 | 122 (20.7) 1.844 | 125 (20.7) 1.842 |
| AG      | 127 (47.4)      | 198 (62.7) 1.362 | 184 (62.4) 1.409 | 192 (63.5) 1.410 |
| AA      | 33 (12.3)       | 17 (5.3)      | 11 (36.2)     | 15 (5.0)      |
| rs1342387 |                  |               |               |                        |
| A       | 201 (37.5) 0.619 | 255 (40.6) 0.27 | 237 (40.2) 0.358 | 227 (38.0) 0.873 |
| G       | 335 (62.5)      | 373 (59.4) 1.139 | 353 (59.8) 1.118 | 371 (62.0) 1.019 |
| AG      | 115 (42.9)      | 141 (44.9) 0.057 | 135 (45.8) 0.608 | 145 (48.5) 0.394 |
| AA      | 33 (12.3)       | 17 (5.3)      | 11 (36.2)     | 15 (5.0)      |

p ≤ 0.05/3 = 0.017. n refers to number of individuals. Values are given as allele or genotype frequencies, proportions (%), and OR (95% CI); differences were compared using the Pearson’s χ² method.

aSignificance indicates difference between case and control p ≤ 0.05.
bSignificance indicates CAD versus control.
cSignificance indicates T2D versus control.
dSignificance indicates T2D with CAD versus control.

CAD, coronary artery disease; CI, confidence interval; OR, odds ratio; SNP, single-nucleotide polymorphism; T2D, type 2 diabetes.
Six single-nucleotide polymorphisms (SNPs) (rs7539542, rs3737884, rs1342387, rs16850797, rs12045862, and rs7514221) on chromosome 1q32 were selected for genotyping in the present study. Genomic DNA was extracted from frozen EDTA whole peripheral blood using a salting-out procedure. The SNPs were genotyped using the polymerase chain reaction–restriction fragment length polymorphism method, with the exception of rs12045862, which was genotyped using high-resolution melting curves–unlabeled probe genotyping analysis. To ensure the quality of genotyping, we selected randomly three samples of each genotype to be directly sequenced. No discrepancies were observed.

Statistical analyses

The statistical analyses were performed using SPSS version 17.0 software. Continuous parameters were presented as mean ± standard deviation (SD) and were compared using one-way analysis of variance (ANOVA). The normality of distributions of the continuous variables was assessed with the Kolmogorov–Smirnov test. Categorical data were given as proportions of all the samples and were compared using the Pearson’s chi-square (\( \chi^2 \)) test. The odds ratios (ORs) and 95% confidence interval (CI) were calculated to evaluate the strength of association between variables. Hardy–Weinberg equilibrium (HWE) was verified using a chi-square goodness-fit test. Pairwise LD and haplotype analysis were confirmed using the open-source software, SHEsis. Bonferroni correction was conducted for multiple comparisons. A two-tailed value of \( p \leq 0.05 \) was considered to be statistically significant.

Results

Clinical characteristics

The prevalence of known chronic metabolic risk factors, including SBP, DBP, FBG, TG, TC, and LDL-C, appeared to be higher in each subgroup (CAD group, T2D group, and...
T2D with CAD group), while HDL-C was lower compared with the normal control ($p > 0.05$).

**SNP association analysis**

We analyzed all SNPs and found a week pairwise LD for those SNPs. The observed genotype frequencies of the six polymorphisms conformed to the HWE in the control ($p > 0.05$). The allele and genotypic frequencies are shown in Table 1. The G allele frequency for rs3737884 of each case subgroup (CAD, T2D, and T2D with CAD patients) was higher ($p = 1.82E-5$, OR = 1.770, 95% CI (1.362–2.301)); $p = 1.745E-4$, OR = 1.652, 95% CI (1.217–2.028)) compared with the control. However, the C allele frequency for rs16850797 only conferred risk for the T2D and T2D with CAD groups ($p = 0.001$, OR = 1.529, 95% CI (1.181–1.980); $p = 0.001$, OR = 1.571, 95% CI (1.217–2.028)). In addition, the allele C of rs7514221 was also positively associated with the three diseases [$p = 0.004$, OR = 1.629, 95% CI (1.165–2.276); $p = 0.002$, OR = 1.674, 95% CI (1.194–2.347); $p = 0.002$, OR = 1.702, 95% CI (1.217–2.382)].

Through genetic model analysis, we found that rs3737884*G, rs16850797*C, and rs7514221*C are risk variants for CAD, T2D, and T2D with CAD in at least one of the three genetic models (Table 2, $p < 0.017$).

Haplotype GCT, which contains 3737884*G, rs16850797*C, and rs7514221*C, was the most prevalent risk haplotype for patients with CAD, T2D, and T2D with CAD (Table 3, $p < 0.017$).

**Discussion**

**ADIPOR1** variant, rs3737884*G, was associated with the susceptibility of CAD, T2D, and T2D with CAD, but rs16850797*C was associated with T2D and T2D with CAD, which has only been reported in a northern Han Chinese population (Jin et al., 2014). There are different effects of genetics backgrounds on the disease in different ethnic groups, so an estimate of genetic effect size from one ethnic background is frequently biased and more precise estimates can be obtained in independent replication study. Therefore, we conducted a replication study to assess the association of **ADIPOR1** variants with T2D, CAD, and T2D with CAD in a northeast Han Chinese population.

Consistent with the above findings, we also identified a novel shared variant rs7514221*C in **ADIPOR1** that is associated with CAD, T2D, and T2D with CAD susceptibility in our study. Our finding confirms that **ADIPOR1** variant, rs3737884*G, is a strong shared candidate for the three diseases in a Chinese Han population. SNP rs7514221*C seems to be also a shared genetic variant of CAD, T2D, and T2D with CAD in our future study population. Considering that this is the first study on the shared variant rs7514221, it should be verified in other populations.

Although rs16850797*C was just associated with T2D and T2D with CAD, our haplotype analysis showed that GCT containing two risk variants (rs3737884*G and rs16850797*C) was the most prevalent at-risk haplotype for the three diseases. It seems to suggest that we should pay attention to the integrity of the gene when we try to identify the genetic variation associated with diseases (Hannou et al., 2015).

The mechanisms through which variations in the **ADIPOR1** could influence CAD and T2D are only hypothetical at the moment. We suppose the two polymorphisms (rs3737884 and rs7514221) located inside intron of the **ADIPOR1** locus could influence adiponectin receptor expression levels and further physiologically modulate adiponectin metabolic activities in distal metabolically active tissues (Luo et al., 2013).

However, we focused only on SNPs; numerous factors act individually and together to influence risk of the three diseases. So, we should involve more factors in our future work. Besides, further physiological and functional studies are also needed to reveal the molecular mechanisms and pathways underlying the associations with the three diseases (Ghanbari et al., 2015).
Conclusion

Taken together, our study not only confirmed that ADPOR1 variant, rs3737884*G, was a strong shared risk variant for CAD, T2D, and T2D with CAD but also identified a novel risk factor, rs7514221*C, shared with the three diseases in a population of northeast China.

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Author Disclosure Statement

No competing financial interests exist.

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