Evaluation of the Neurobehavioural Toxic Effects of Taurine, Glucuronolactone, and Gluconolactone Used in Energy Drinks in Young Rats

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ABSTRACT

Objectives: The neurotoxic effects of food additives used in energy drinks have been investigated since the 1900s but safety concerns are rising and reassurance via safety testing in animals is demanded by the public. Rigorous safety testing is performed for dose optimisation and duration of treatment and to detect the methods to assess changes in mood and behaviour. Hence, we studied the neurobehavioral effects of selected food additives used in energy drinks and their combination in rats when consumed in high doses.

Materials and Methods: Young Sprague Dawley rats were divided into six groups. Group 1 was treated with the vehicle, group 2 was treated with 25 mg/kg p.o. caffeine, group 3 was treated with 5 mg/kg p.o. glucuronolactone, group 4 was treated with 8 mg/kg p.o. taurine, group 5 was treated with 84 mg/kg p.o. gluconolactone, and group 6 was treated with a combination of the three food additives. Neurobehavioral changes were evaluated on days 7, 14, and 21 using behavioural parameters. Neurobehavioral scoring and neurotransmitter estimation in rat brain tissue was performed on day 21.

Results: Significant changes were observed in the neurobehavioral parameters and neurobehavioural scoring in group 4 and group 6, compared with the control group (p<0.001). Furthermore, the significant decreases in neurotransmitter levels in the brains of rats that were treated with food additives indicated the neurotoxic effects of these substances.

Conclusion: This study elaborated the neurobehavioral effects of selected food additives, namely glucuronolactone, taurine, and gluconolactone, when administered orally for 21 days in young rats. The highest toxic effects, including alterations in neurotransmitter levels, were observed in animals treated with a combination of food additives at high doses.

Key words: Energy drinks, food additives, taurine, glucuronolactone, gluconolactone

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Genç Sıçanlarda Enerji İçeceklerinde Kullanılan Taurin, Glukuronolakton ve Glukonolaktonun Nörodavranışsal Etkilerinin Değerlendirilmesi

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ÖZ

Amaç: Enerji içeceklerinde kullanılan gıda katkı maddelerinin nörotoksik etkileri 1900′lardan bu yana incelenmektedir; ancak, güvenlilik endişeleri artmaktadır ve hayvanlarda güvenliliklerinin test edilerek onaylanması halk tarafından talep edilmektedir. Sıkı güvenlilik testleri doz ve uygulama süresi optimizasyonu ve ruh hali ve davranışsal değişiklikleri belirlemek için yapılmaktadır. Bu nedenle, biz enerji içeceklerinde kullanılan seçilmiş gıda katkı maddelerinin ve kombinasyonlarının sıçanlarda nörodavranışsal etkilerini yüksek dozda araştırdık.

Gereç ve Yöntemler: Genç Sıçanlara enerji içeceklerinin katkı maddeleri ve kombinasyonu 21 gün süren studya dahil edildi. Group 1’te ve 2’de kafein, group 3’te ve 4’te glucuronolakton, group 5’te ve 6’da gluconolakton uygulandı. Davranışsal ve davranışsal değişiklikler 7, 14 ve 21. günlerde değerlendirildi. Bu şekilde, gıda katkı maddelerinin ve kombinasyonlarının sıçanlarda nörodavranışsal etkilerini yüksek dozda araştırıldı.

Bulgular: Bu çalışma, enerji içeceklerinde kullanılan gıda katkı maddelerinin nörotoksik etkilerini araştırdı. Gıda katkı maddelerinin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı. Örneğin, taurin, glucuronolakton ve gluconolakton, sıçanlarda nörodavranışsal etkileri saptandı.
INTRODUCTION

We are exposed to neurotoxins naively through food products. Today the evaluation of the effects of food additives on behaviour and mood in adults is of great concern. Various regulatory bodies are encouraging scrutiny of the use of food additives rigorously for safety and reassurance. The Food and Drug Administration (FDA) and European Food Safety Authority have been evaluating and supporting risk assessment and safety in the use of appropriate doses of acceptable daily intake (ADI). The food additives used in many products like baby foods, cool drinks, energy drinks, and soft drinks are approved by the FDA after safety evaluation. However, various food additives like antioxidants, stabilisers, sweeteners, thickeners, preservatives, and flavouring agents have effects on behaviour when taken in high doses that are listed under the safety margin. As of 2006, FDA guidelines on food additives are classified based on level of concern and safety margin into Low concern level I (12-50 ppb), Intermediate concern level II (50-250 ppb), and High concern level III (250-1000 ppb) based on primary toxicological data.¹ The maximum level of additive that has no demonstrable toxic effect, called the "no-observed-adverse-effect level", and ADI are the check parameters for each food additive. Chronic consumption per day more than the ADI leads to toxicity. The risk to human health varies depending upon the type and time of exposure. Specific studies such as for neurotoxicity, immunotoxicity, and allergenicity are rigorously performed repeatedly to ensure the safety of food additives.²

Common food additives used in energy drinks like taurine, glucuronolactone, and gluconolactone are considered elevated risk. The daily exposure to taurine, glucuronolactone, and gluconolactone from energy drinks in young generations is higher than the mean daily exposure (1420 mL/day of energy drink or 2.6 cans/day). In adults, chronic habitual intake of energy drinks was reported to cause several neurological disorders including migraine, seizures, endocrine disorders, and neuropsychiatric disorders.³ Hence, excessive consumption of energy drinks has toxic effects on the nervous system.

The safety of these food additives used in energy drinks was not documented by the Scientific Committee on Food. According to EFSA 2009 data, the stimulatory effect of taurine on the central nervous system was not clearly documented. The major constituents of energy drinks are taurine, glucuronolactone, and gluconolactone.⁴ Based on this background, a research protocol was elaborated to assess systematically possible neurobehavioural toxic effects in animals of individual food additives and the combination of the food additives taurine, glucuronolactone, and gluconolactone used in energy drinks at high doses. The study included an evaluation of neurobehavioural effects, neurobehavioural scoring, and neurotransmitter estimation in the brain tissue of young rats to show possible neurobehavioural effects and ensure the safety level of food additives used in energy drinks, which are listed under the safety margin.

MATERIALS AND METHODS

Chemicals and reagents

Chemicals

Glucuronolactone, gluconolactone, and caffeine (food grade 99.5%) were procured from Srinelima Labs, Hyderabad, India. Taurine (food grade 99.6%) was obtained from Nutrijar Lifesciences, Nagda, Madhya Pradesh, India. All other chemicals (analytical grade) were from Himedia Pvt Ltd., India.

Reagents

Hydrochloric acid ([HCl]-butanol solution (0.85 mL of 37% HCl in 1 L), 0.4 M HCl (3.4 mL of concentrated HCl and made up to 100 mL with water), 0.1 M HCl (0.85 mL of concentrated HCl made up to 100 mL with water), 5 M NaOH (20 g of sodium hydroxide pellets dissolved in distilled water and volume made up to 100 mL with distilled water), and 10 M acetic acid (57 mL of glacial acetic acid and made up to 100 mL with distilled water) were used. Reagents and buffers like sodium acetate buffer (EDTA pH 6.9), heptane, sodium sulphite solution, and O-phthaldialdehyde (OPT) reagent were obtained from Sigma Aldrich, Hyderabad, India.

Equipment

A morris water maze (MWM), version 5.0, was obtained from Orchid Scientific. A wooden arena with 64 squares was prepared by Wood Works, Hyderabad. A tissue homogeniser 160 W, a refrigerated centrifuge from Gravity Labs, and a spectrofluorometer model, ALT 2380 (wavelength range 200 to 700), were also used.

Animals

Sprague Dawley albino rats of both male and female in equal ratio weighing 150-200 g were obtained from the animal house of MLR Institute of Pharmacy, Hyderabad. The animals were divided into four groups and housed under standard laboratory conditions (temperature 25±10°C, relative humidity 55±5%, and 12:00:12.00 h dark:light cycle) with standard pellet diet and water ad libitum. The experimental procedure was approved by the Institutional Animals Ethics Committee (IAEC) as required by the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), MLR Institute of Pharmacy, Hyderabad (CPCSEA/IAEC/PR3/2019).

Experimental protocol

All the animals were divided into six separate groups and each group consisted of six animals with equal ratios of males and
females, 3:3 (n=6, 3 males + 3 females). All the doses were calculated based on the human dose available in the literature and were converted to animal dose. High doses of food additives were administered and the animals were observed for neurotoxic effects. Group I animals served as controls, treated with water administered orally. Group II animals acted as the working standard treated with caffeine 25 mg/kg p.o. Group III animals were treated with glucuronolactone 5 mg/kg p.o., group IV animals were treated with taurine 8 mg/kg p.o., group V animals were treated with glucuronolactone 84 mg/kg p.o., and group VI animals were treated with a combination of the three food additives (glucuronolactone 5 mg/kg p.o., taurine 8 mg/kg p.o., and glucuronolactone 84 mg/kg p.o.). All the animals were treated with freshly prepared doses dissolved in water and administered through an oral gauge every day until day 21.

**Assessment of neurobehavioural effects**

Neurobehavioural changes were observed in animals treated with the respective doses for 21 days. On days 7, 14, and 21 the animals were screened for neurobehavioural effects by functional observational battery (FOB) and the Irwin protocol. These include studies of behavioural alterations, the MWM test, a locomotor activity test, and the Katz protocol as described below.

**Behavioural alterations**

Behavioural changes were evaluated by measuring rearing and paw licking behaviour for 5 min. The observations were noted by three blind observers.

**Morris water maze test**

Cognitive changes such as in learning, conditioning, memory, and attention were evaluated by MWM test in rats. The maze was a round grey tank (0.45 m radius, 0.5 m tall) filled with water (22°C) to a depth of 0.15 m. An adjustable platform of size 0.06 m x 0.06 m made of steel was placed 0.01 m under the water level and 0.13 m from the edge. Milk (1 mL) was added to make the water cloudy and thus the platform was hidden. On the edge of the tank the four letters nominated as north (N), south (S), east (E), and west (W) divided the tank into four portions (N-W, N-E, S-E, and S-W). On day 1, the rats were allowed to swim in the tank for 1 min without the hidden platform. Thus, they were trained for swimming in the tank. On day 2 they were trained to identify and move onto the submerged platform for 6 trials per day until day 5. In each trial the rats were released into the tank with their faces pointing towards the water to confirm immersion. The latency from immersion in the tank to escape onto the hidden platform (maximum duration of trial 2 min) was noted. In 2 min, if the animal could not identify the platform it was physically directed to climb by using a glass rod. Then the score of 2 min was noted for these trials. The number of such unsuccessful trials was calculated. For learning and memorising the spatial cues each animal was given an interval of 0.5 min on climbing onto the platform.

**Locomotor activity test**

Locomotor changes such as coordination and equilibrium were assessed by locomotor activity test. This test consists of a square wooden field measuring 0.8x0.8x0.3 m and the flooring was divided into 64 squares of equal dimensions. Duration of immobility and locomotion in 5 min for each animal was recorded.

**Katz protocol (neurobehavioural scoring)**

Neurobehavioural scores were calculated for the animals after 21 days’ treatment with high doses of food additives and they were evaluated for neurobehavioural toxic effects (Table 1). The observations were noted by three blind observers.

**Estimation of neurotransmitters**

**Preparation of tissue extract**

On day 21 the rats were sacrificed, the whole brain was dissected out, and the subcortical region was separated and weighed. The weighed tissue was homogenised in a homogeniser with 5 mL of HCl butanol for about 1 min. The homogenised tissue was then centrifuged for 10 min at 2000 rpm. The supernatant layer

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| Table 1. Neurobehavioural scores by Katz protocol to evaluate neurotoxic effects |
|-----------------------------------------------|
| Neurobehavioural effect | Scores |
|---------------------------|--------|
| General behavioural deficit |  |
| Consciousness | Present | 0 |
| | No attempt (coma) | 20 |
| Respiration | Normal | 0 |
| | Abnormal | 20 |
| Cranial nerve reflexes |  |
| Olfactory (sniffing food) | Present | 0 |
| | Absent | 4 |
| Vision (follow hand) | Present | 0 |
| | Absent | 4 |
| Corneal reflex | Present | 0 |
| | Absent | 4 |
| Whisker (movement) | Present | 0 |
| | Absent | 4 |
| Hearing (turning to clapped hands) | Present | 0 |
| | Absent | 4 |
| Motor deficit: |  |
| (Leg/tail movement) | Normal | 0 |
| | Stiff | 5 |
| | Paralysed | 10 |
| Sensory deficit |  |
| Leg/tail (on pinching) | Present | 0 |
| | Absent | 10 |
| Coordination: |  |
| Beam walking (1.5 cm) | Present | 0 |
| | Absent | 5 |
| Placing test | Present | 0 |
| | Absent | 5 |
| Righting reflex | Present | 0 |
| | Absent | 5 |
| Stopping at edge of table | Present | 0 |
| | Absent | 5 |
| Neurobehavioural toxicity | Total | 100 |
(1 mL) was separated and added to a centrifuge tube containing 2.5 mL of heptane and 0.3 mL of 0.1 M HCl. After 10 min of shaking vigorously the tube was centrifuged under identical conditions. Two layers were separated, the supernatant layer (organic layer) was discarded, and the remaining aqueous extract was used to estimate noradrenaline, dopamine, and serotonin. All the steps were carried out at 0°C. The brain extracts were stored at -20°C until further experimentation.

**Estimation of noradrenaline**

First 0.2 mL of the aqueous layer was taken from tissue extract stored at ice cool temperature after preparation of extract. Then 0.05 mL of 0.4 M HCl and 0.1 mL of EDTA (pH 6-9) were added to the aqueous extract accompanied by 0.1 mL of iodine solution for oxidation. The reaction was stopped after 2 min by adding 0.1 mL of Na₂SO₃ solution. Next, 0.1 mL of acetic acid was added after 1.5 min. The solution was heated to 100°C for 6 min. The sample was allowed to cool and excitation and emission spectra were noted from the spectrofluorometer. These interpretations were measured at 395-485 nm for noradrenaline.

**Estimation of dopamine**

To 0.2 mL of aqueous phase extract were added 0.5 mL of HCl and 1 mL of EDTA (pH 6.9) accompanied by 0.1 mL of iodine solution for oxidation. The reaction was stopped after 2 min by adding 0.1 mL of Na₂SO₃ solution. Then 0.1 mL of acetic acid was added after 1.5 min. The solution was heated to 100°C for 6 min. The sample was allowed to cool and excitation and emission spectra were noted from the spectrofluorometer. These interpretations were measured at 330-375 nm for dopamine.

**Estimation of serotonin**

First, 0.2 mL of aqueous tissue extract was added with 0.25 mL of OPT reagent. Then it was heated for 100°C for 10 min. After the sample reached ambient temperature, the readings were taken at 360-470 nm in the spectrofluorometer for the estimation of serotonin.

Tissue blanks for dopamine and noradrenaline were prepared by adding the reagents of the oxidation step in reverse order (sodium sulphite before iodine). For the serotonin tissue blank, 0.25 mL of concentrated HCl without OPT was added. Internal standard was prepared by taking 500 µg/mL each of noradrenaline, dopamine, and serotonin prepared in distilled water: HCl butanol in 1:2 ratio. The concentration of the neurotransmitters expressed in µg per gram wet weight of tissue was calculated by using the formula:

\[
\text{Concentration of unknown (Cu) = } \frac{\text{Sample O.D-Blank OD}}{\text{Standard O.D-Blank OD}} \times \text{Cs}
\]

Cs: Concentration of standard (500 µg/mL)
OD: Optical density

**RESULTS**

**Neurobehavioural changes**

**Behavioural alterations**

Alterations in behavioural effects were observed in animals treated with high doses of individual food additives, with successive increases in the behavioural effects with increases in treatment duration on days 7, 14, and 21. Animals treated with taurine and the combination of food additives showed significant increases in rearing and hind paw licking (p<0.001) compared with the experimental group. Group VI, given the combination of food additives, showed a high significant difference (p<0.05) in behavioural activity compared with the group given caffeine as shown in Figure 1.

**Morris water maze test**

Taurine treated animals showed longer escape latency onto the submerged platform in the water maze compared with the controls. With an increase in the duration of treatment the increase in escape latency was significant (p<0.001). Animals treated with the combination of food additives showed significantly (p<0.05) longer escape latency on day 21, indicating altered cognitive effects compared with the caffeine treated animals (Figure 2).

**Locomotor activity test**

A significant increase in immobility duration was seen in animals treated with individual food additives and the combination of food additives (p<0.001) and with an increase in...
in duration of treatment compared to the control group when placed in the wooden arena. The combination of food additives caused a significant increase in immobility duration (p<0.05), indicating a decrease in locomotion compared to the caffeine treated animals (Figure 3).

*Katz protocol of neurobehavioural scoring*

In the Katz protocol animals treated with high doses of individual food additives showed high neurobehavioural scores on day 21. All the experimental groups showed significantly higher (p<0.001) scores than the control animals (Figure 4). The combination group exhibited the highest neurobehavioural scoring (p<0.05), indicating an increase in neurobehavioural toxic effects compared with the caffeine treated animals.

*Estimation of neurotransmitters*

On day 21, tissue extract was prepared and neurotransmitters were estimated. The noradrenaline and serotonin levels were pointedly (p<0.001) lower in the taurine and combination of food additives treated animals than in the controls. The combination group animals showed high significance compared with the caffeine treated group (p<0.05) (Figures 5 and 6).

Figure 2. Assessment of neurobehavioural effects of food additives on exposure to high doses for 7, 14, and 21 days in Sprague Dawley rats by escape latency in seconds using the Morris water maze test. Data were represented as mean ± SEM (n=6). *p<0.001 showed significant differences between the experimental group and the control group. *p<0.05 showed significant differences between the experimental group and the caffeine treated group.

SEM: Standard error of the mean, GLUR: Glucuronolactane, TAU: Taurine, GLU: Glucanolactane

Figure 3. Assessment of neurobehavioural effects of food additives on exposure to high doses for 7, 14, and 21 days in Sprague Dawley rats by duration of immobility using a locomotor activity test. Data were represented as mean ± SEM (n=6). *p<0.001 showed significant differences between the experimental group and the control group. *p<0.05 showed significant differences between the food additives treated group and the caffeine treated group.

SEM: Standard error of the mean, GLUR: Glucuronolactane, TAU: Taurine, GLU: Glucanolactane

Figure 4. Assessment of food additives on exposure to high doses for 21 days in Sprague Dawley rats for neurobehavioural effects by neurobehavioural scoring using the Katz protocol. Data were represented as mean ± SEM (n=6). *p<0.001 showed significant differences between the experimental group and the control group. *p<0.05 showed significant differences between the experimental group and the caffeine treated group.

GBD: General behavioural deficits (score 40), CNR: Cranial nerve reflexes (score 20), MD: Motor deficit (score 10), SD: Sensory deficit (score 10, CD: Coordination (score 20), SEM: Standard error of the mean, GLUR: Glucuronolactane, TAU: Taurine, GLU: Glucanolactane

Figure 5. Effect of food additives on Nor-adrenaline levels in whole brain tissue of rats exposed to high doses for 21 days. Data were represented as mean ± SEM (n=6). *p<0.001 showed significant differences between the experimental group and the control group. *p<0.05 showed significant differences between the food additives treated group and the caffeine treated group.

SEM: Standard error of the mean, GLUR: Glucuronolactane, TAU: Taurine, GLU: Glucanolactane
Decreases in dopamine levels were observed in the taurine and combination group animals compared to the control (p<0.001). The combination animals showed highly significant results when compared with the caffeine treated animals, indicating altered neurotransmission in the brain (p<0.05) (Figure 7).

DISCUSSION

The food additives used in energy drinks, when consumed above the acceptable level, were reported to produce toxic effects, as stated by the EFSA. However, the exact ingredients and the dose responsible for toxic effects were not evaluated or documented clearly. The present research provides evidence for neurobehavioural toxic effects for the selected FDA approved food additives used in energy drinks when consumed above the ADI. The neurobehavioural toxic effects of food additives when administered orally at doses of glucuronolactone 5 mg/kg p.o., taurine 8 mg/kg p.o., gluconolactone 84 mg/kg p.o., and a combination of the three food additives were evaluated and documented over 21 days of treatment in young rats.

Earlier studies suggested that the Irwin protocol (FOB test) explains many parameters and provides a multidimensional method for the explanation of neurobehavioural effects. Based on the Irwin protocol, the Sprague Dawley rats were treated with food additives and neurobehavioural changes were evaluated using behavioural alterations test, the MWM test, and a locomotor test for clarification of neurobehavioural toxic effects.

Previous literature indicated that behaviour is a measure of the integration of neural function and alteration in behaviour was used to evaluate neurobehavioural toxic effect. In the present study, alteration in behavioural activity was assessed by considering behavioural parameters like paw licking and rearing behaviours, which were considered indicators of grooming. An increased anxiety level due to any stimulus was reported to change paw licking and rearing behaviour. Similar alterations in paw licking and rearing behaviour were caused by taurine and the combination, which clearly indicates the alteration in neuronal functioning with the selected food additives. Previous studies that evaluated cognitive effects in rats using a water maze test reported an increase in duration of escape latencies, indicating a decrease in cognition. In the present study a significant increase in duration of escape latency to the submerged platform was observed in the taurine and combination groups. The decreased cognition may be due to the decrease in cyclic GMP levels as reported with cognitive impairment and neurobehavioural deficit reported in aluminium toxicity studies. A similar decrease in cGMP levels was reported with taurine in cardiomyocytes. Our study indicated that neurotoxicity caused by food additives progressively increased with days of exposure from day 7 to day 21. Earlier studies stated that locomotor activity indicates attentiveness. In the present study, the decrease in locomotor activity indicated by an increase in duration of immobility in the taurine and combination groups affirms that a decrease in attentiveness leads to altered neurological functioning.
Previous studies reported that taurine showed dose correlated behavioural changes in rats. Chewing of limbs after treatment with taurine indicated its central pharmacological and neuromodulator effects. In the present study, the taurine treated group also showed altered behavioural activity, which confirmed its potent neuromodulator effect on neurotransmitters of the brain. In a subacute toxicity study for 14 days in rats, gluconolactone showed mortality, abnormal clinical signs, body-weight changes (on days 1, 2, 3, 7, 10, and 14), and gross pathological changes in the brain but was not focused on neurobehavioural symptoms. Our study for the first time showed changes in behavioural activity in gluconolactone treated animals and may hint at neurotoxicity when consumed higher than the acceptable doses. These changes were high when given in combination with taurine.

Neurological scales/scores for motor, sensory and reflex functions in rats, mice, and dogs were used to detect effects on brain injury. In the present research work, the Katz protocol of neurobehavioural scores was used considering various parameters like general behavioural deficits, cranial nerve reflexes, motor deficit, sensory deficit, and coordination to evaluate the neuronal damage in animals. High scores for neurobehavioural deficits were observed in animals receiving the food additives in combination, rather than individually. This indicates chances of increases in brain neuronal damage and can be correlated with the decrease in neurotransmitters.

Selected food additives were hypothesised to enhance neurotransmitter activity concentrated in the subcortical regions according to the literature. Therefore, subcortical regions of whole brain extracts were used to estimate neurotransmitters. Decreases in noradrenaline, serotonin, and dopamine levels indicating neurochemical alterations and neurotoxic effects on subchronic drug administration were suggested previously. In addition, earlier studies also focused on the participation of serotonin in cognition and memory, and altered serotonergic neurotransmission by toxic substances was reported. The neurotransmitter modulatory effect of these selected food additives was mentioned. Corroborating the earlier studies, noradrenaline, serotonin, and dopamine were decreased prominently in the current study. The combination of taurine, gluconolactone, and glucuronolactone caused more noticeable changes in neurotransmitter levels than when given alone, which indicates a risk of more neuronal damage, modulation, and toxicity. These changes support the observed neurobehavioural deficits caused by food additives.

The present study raises concern about the safety of the mentioned food additives at the doses studied considering the aspect of simultaneous consumption of these food additives via energy drinks, although the safety of these additives was established and approved but individually and at a different exposure level. Furthermore, histopathological studies are needed for correlation of neurobehavioural toxic effects.

CONCLUSION

This study elaborated the neurotoxic effects of glucuronolactone, taurine, and gluconolactone used in energy drinks when consumed above the ADI. It showed significant neurobehavioural toxic effects accompanied by altered neurotransmitter levels in rats treated with a combination of selected food additives. Furthermore, investigation is required to understand the mechanism and interaction between food additives. Appraisal of the developmental neurotoxic effects of these food additives in combination will also be valuable.

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