Study on the Extraction of Polyphenols from Blueberry Leaves and their antioxidant properties

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Abstract. Blueberry leaf as materials, ethanol ultrasonic assisted extraction of blueberry polyphenols, ethanol concentration and solid-liquid ratio on blueberry ultrasonic time leaves influence the yield of polyphenols were researched, and the orthogonal experiment to optimize the ethanol ultrasonic assisted extraction technological conditions of blueberry leaf polyphenol polyphenols was used. The results showed that the optimal extraction conditions of polyphenols from blueberry leaves were as follows, 9.06% with 4min ultrasonic time, 80% ethanol concentration and 1:45 (g/mL) ratio. Ultrasonic assisted extraction is a convenient and high yield method, which is suitable for the extraction of polyphenols from blueberry leaves.

1 Introduction

Blueberry leaves contain polysaccharides, polyphenols, amino acids, fatty acids, crude protein, crude fat, crude fiber, organic acids, vitamins, sugars and a large number of polyphenols. Polyphenol, also known as tannins and casuarinin, is a kind of mixture of secondary metabolites widely existing in plants. They are the general names of plant components with multiple hydroxy phenols in molecules[1]. Studies such as Li showed that blueberry leaf polyphenols could effectively prolong the shelf life of perch during cold storage, which is an effective food biological preservative[2]. In addition, plant polyphenols have strong antioxidant, scavenging free radicals, anticancer, anti-aging, antiviral and other effects, known as 'seventh nutrients'[3].

However, in blueberry machining processing, blueberry leaves are often discarded as waste, resulting in huge waste of resources. Turhan et al.[4] confirmed that the polyphenol content in blueberry fruits was significantly lower than that in blueberry leaves, and the antioxidant activity and free radical scavenging ability of blueberry leaves were stronger than those of blueberry fruits.

In the paper, blueberry leaf as materials, ethanol ultrasonic assisted extraction of blueberry polyphenols, ethanol concentration and solid-liquid ratio on blueberry research ultrasonic time leaves influence the yield of polyphenols, and using the orthogonal experiment to optimize the ethanol ultrasonic assisted extraction technological conditions of blueberry leaf polyphenol polyphenols.

2 Materials and methods

2.1 Materials
Blueberry leaf (cultivar 'Lanfeng')

2.2 Methods

2.2.1 Blueberry pretreatment
A certain amount of blueberry leaves was taken to dry at 50℃, and screen after crushing 100 mesh sieve.

2.2.2 Standard curve drawing
The 250mg gallic acid standard was weighed, and the volume was fixed to 250mL volumetric flask with deionized water. The standard solution of 1mg/mL was prepared, and 5mL was removed from the standard solution of 1mg/mL. The volume was fixed to 100mL volumetric flask, and the standard solution of 50μg/mL was prepared. Accurately absorb 0, 0.5, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5mL standard solution in 25mL volumetric flask, add 1mL Folin-Ciocalteu reagent 0~8min, add 3mL 10% sodium carbonate solution, deionized water constant volume to 25mL, shake, prepare gallic acid mass concentration were 0, 1.0, 2.0, 3.0, 4.0, 5.0, 6.0, 7.0μg/mL, 30℃ placed 1.5h, at the wavelength of 750nm determination of absorbance, draw mass concentration-absorption curve [3].
2.3 Experimental treatment

2.3.1 Effect of ultrasonic time on polyphenol yield
Reference Li et al.[6] slightly changed. 1.0g blueberry leaf powder was weighed, and the effects of extraction time on the yield of polyphenols from blueberry leaves were investigated at the solid-liquid ratio of 1:25 (g/mL), ethanol concentration of 60%, and ultrasonic power of 250W.

2.3.2 Effect of ethanol concentration on polyphenol yield
The 1.00 g blueberry leaf powder was accurately weighed. Under the conditions of ultrasonic time of 6 min, ultrasonic power of 250 W, and solid-liquid ratio of 1:35 (g/mL), the effects of ethanol concentration of 20%, 40%, 60%, 80%, and 100% on the yield of polyphenols from blueberry leaves were investigated.

2.3.3 Effect of solid-liquid ratio on polyphenol yield
The 1.00g blueberry leaf powder was accurately weighed. Under the conditions of ultrasonic time of 6 min, ultrasonic power of 250W and ethanol concentration of 60%, the effects of solid-liquid ratio of 1:15, 1:25, 1:35, 1:45 and 1:55( g/mL)on the yield of polyphenols from blueberry leaves were investigated.

2.3.4 Determination of polyphenol content
Folin phenol colorimetric method was used to determine the content of polyphenols. The method of Li[7] was slightly modified. The regression curve equation of polyphenol absorbance and polyphenol concentration is $y=15.511x-0.0032\,(R^2=0.9992)$, where $y$ is the absorbance of polyphenol at 752nm and $x$ is the concentration of polyphenol (mg/mL). The absorbance of polyphenols in blueberry leaves was determined at 752nm, the concentration of polyphenols was calculated according to the regression curve equation, and the yield of polyphenols was calculated according to the following formula.

\[ \text{Polyphenol yield(%)=} \frac{C \times V \times n \times 100}{1000 \times m} \quad (I) \]

Among them: C is the blueberry leaf polyphenol concentration (mg/mL), V is the extraction sample volume (mL), n is the dilution ratio, m is the blueberry leaf mass (g).

2.3.5 Nitrite removal ability
It was slightly modified by referring to the method of Zhao[8]. Take three test tubes and number them A, B and C respectively. Add 2mL of extract to tube A, 2mL of extract to tube B, and 2mL of deionized water to tube C. Then 2mL 10μg/mL NaNO₂ solution was added to tube A and tube C, and 2mL deionized water was added to tube B. After shaking the three tubes separately, the tubes were left standing at room temperature for 5min. Then, 2mL 0.4% p-aminobenzene sulfonate solution was added to test tubes A, B and C respectively, and mixed. After standing for 15min, 1mL 0.2% naphthalene ethylenediamine hydrochloride solution was added to the three test tubes respectively, mixed, and stood for 15min in the dark. The absorbance value was measured with a 2cm colorimetric cup at the maximum absorption wavelength of 538nm. Taking deionized water as reference, the absorbance value $A_0$ of C tube without extraction solution was measured, and the absorbance value $A$ of A tube with extraction solution was measured with B tube as reference.

The following formula was used to calculate the nitrite clearance rate:

\[ S\% = \left(1 - \frac{A_i - A_j}{A_0}\right) \times 100 \]

3 Results and analysis

3.1 Effects of different ultrasonic time on the yield of polyphenols from blueberry leaves

It can be seen from Fig.1 that the extraction rate of polyphenols increased first and then decreased with the ultrasonic time, and the optimum extraction time was 6min.
3.2 Effect of ethanol concentration on the yield of polyphenols from blueberry leaves

It can be seen from Fig.2 that the extraction efficiency is the highest when the concentration of ethanol is 60%, and either the concentration is too high or too low, which is not conducive to extraction. According to the principle of similar phase solution, the polyphenols of blueberry leaves can be fully dissolved in the solvent with similar polarity so as to be extracted.

![Fig.2 Effect of ethanol concentration on the yield of polyphenols from blueberry leaves](image)

3.3 Effect of material liquid ratio on the yield of polyphenols from blueberry leaves

From Fig.3, solid-liquid ratio 1:35 had the highest extraction efficiency. Too much ethanol was not conducive to the extraction of polyphenols, so the solid-liquid ratio was 1:35 is the best extraction ratio.

![Fig.3 Effect of material liquid ratio on the yield of polyphenols from blueberry leaves](image)

3.4 orthogonal experiment

| material liquid ratio | ethanol concentration (%) | ultrasonic time(min) | the yield of polyphenols(%) |
|-----------------------|---------------------------|----------------------|-----------------------------|
| 1:25                  | 40                        | 4.00                 | 5.18                        |
| 1:25                  | 80                        | 6.00                 | 3.75                        |
| 1:25                  | 60                        | 8.00                 | 5.38                        |
| 1:35                  | 60                        | 4.00                 | 5.85                        |
| 1:35                  | 40                        | 6.00                 | 5.70                        |
| 1:35                  | 80                        | 8.00                 | 6.11                        |
| 1:45                  | 80                        | 4.00                 | 9.06                        |
| 1:45                  | 60                        | 6.00                 | 6.09                        |
| 1:45                  | 40                        | 8.00                 | 6.02                        |

According to the results of orthogonal experiment, the optimal extraction efficiency was obtained under the conditions of 80% ethanol concentration, 4min ultrasonic time and 1:45 solid-liquid ratio, which was different from the optimal extraction conditions of 60% ethanol concentration, 6min ultrasonic time and 1:35 solid-liquid ratio under the single factor condition, indicating the optimization of the experimental results by orthogonal experiment.

3.5 Nitrite removal rate

According to the Fig.4, the nitrite removal rate of blueberry leaves in May was the highest at the same concentration. For blueberry leaves in different months, the higher the concentration of extract, the higher the clearance rate.

![Fig.4 Nitrite removal rate of blueberry leaf extracts with different concentrations in May, June and July](image)
3.6 DPPH scavenging rates

According to the Fig.5, under the same concentration, the scavenging rate of DPPH of blueberry leaves in July was the highest. For blueberry leaves in different months, the lower the concentration of extract, the higher the scavenging rate.

4 Conclusion

Studies have shown that blueberry leaves contain relatively rich total phenols, and have strong free radical scavenging ability and reducing power[10]. The results of single factor experiment showed that the optimal extraction conditions existed regardless of ultrasonic time, ethanol concentration and solid-liquid ratio, and too high or too low would affect the extraction rate. The optimal extraction conditions of polyphenols from blueberry leaves were as follows, 9.06% with 4 min ultrasonic time, 80% ethanol concentration and 1:45 (g/mL) ratio. Ultrasonic assisted extraction is a convenient and high yield method, which is suitable for the extraction of polyphenols from blueberry leaves. However, the orthogonal experiment showed that the interaction of different extraction conditions was different from the accumulation of single factor optimal conditions.

Acknowledgements

Research on comprehensive utilization and pilot production of secondary fruit of Wudi jujube, provincial key R & D plan in 2019 (tackling key scientific and technological problems for public welfare), (Project No. 2019GSF107083)

Research on the theory and practice of the deep integration course of specialty and innovation based on the cultivation of students' innovation and entrepreneurship--Taking bioengineering as an example(JG201918)

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