Expression and Purification of SARS Coronavirus Membrane Protein

DAI Wuxing (戴五星), LEI Mingjun (雷明军), WU Shouting (吴少庭), CHEN Zhihao (陈智浩), LIANG Liang (梁 李), PAN Hui (潘晖), QIN Li (秦 利), GAO Shitong (高士同), YUAN Shishan (袁仕善), ZHANG Renli (张仁利)  

1Department of Biochemistry and Molecular Biology, School of Basic Medical Science, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430070, China  
2Shenzhen Municipal Center for Disease Control and Prevention, Shenzhen 518020, China

Summary: To construct a recombinant plasmid Pet23a-M, the gene encoding severe acute respiratory syndrome (SARS) coronavirus membrane protein was amplified by RT-PCR and cloned into the expression plasmid Pet23a. Results of restriction endonuclease analysis, PCR detection and DNA sequencing analysis revealed that the cloned DNA sequence was the same as that reported. The recombinants were transformed into Escherichia coli (E. coli) BL21 (DE3) and induced by Isopropyl-1-thiogalactopyranoside (IPTG). The expression of 27 kD (1 kD=0.992 1 ku) protein was detected by SDS-PAGE and purified by metal chelated chromatography. Results of Western-blot showed that this expressed protein could react with antibodies in sera of SARS patients during convalescence. This provided the basis for the further study on SARS virus vaccine and diagnostic agents.

Key words: SARS; membrane protein; gene expression; protein purification; Western-blot

A novel coronavirus (SARS-CoV) was discovered in association with cases of severe acute respiratory syndrome (SARS). The sequence of the complete genome of SARS-CoV was determined [1,2]. It is not closely related to any of the previously characterized coronavirus. The analysis of open reading frames of SARS-CoV showed that there were similar major structural proteins that may have played important roles in causing the infection. They are spike protein (S protein), nucleocapsid protein (N protein), membrane protein (M protein), and small envelope protein (E protein). To screen and prepare effective SARS virus vaccine and diagnostic antigens, we designed a pair of primers to amplify the gene encoding SARS coronavirus membrane protein and cloned it into an expression plasmid Pet23a.

1 MATERIALS AND METHODS

1.1 Materials

1.1.1 Bacterial Strains and Plasmids  
SARS Coronavirus was kindly provided by Vice-director He (Division of Microbiological Testing, Center for Disease Control and Prevention, Shenzhen, China). Escherichia coli (E. Coli) DH5α, BL21 (DE3) were kept in our laboratory. Vector pMD18-T was obtained from the TaKaRa Biotechnology (Dalian) Co., Ltd. Expression plasmid Pet23-a was from EMD. Bioscience, Inc (USA).  

1.1.2 Enzyme and Reagents  
Taq DNA polymerase, dNTP, protein marker were purchased from Promega Company (USA). Virus RNA purification kit was purchased from Qiagen Company (Germany). RT-PCR kit, T4 DNA ligase, restriction endonuclease SalI, BamHI, DNA marker were from TaKaRa Biotechnology Co (Dalian), Plasmid DNA isolation kit was obtained from Shanghai DNA Biotechnologies Co, Ltd. Isopropyl-β-D-thiogalactopyranoside (IPTG), SDS-Na, EDTA-Na, Tris base, Ampicillin were purchased from Shanghai Sangon Biological Engineering & Technology Service Co, Ltd. Sheep anti-human IgG labeled by horseradish peroxidase was purchased from Pierce Company (USA). Sera of SARS patients during convalescence were offered by Shenzhen East Lake Hospital.

1.2 Methods

1.2.1 Preparation and Purification of Virus RNA  
The preparation and purification of virus RNA was performed according to the manual provided by the virus RNA purification kit.

1.2.2 RT-PCR Amplification of M Gene and T/A Clone  
RT-PCR amplification was performed with the SARS-CoV genomic RNA used as the template by the manual of RT-PCR kit. 1.5 μl cDNA after reverse transcription was used as PCR template. A pair of primers used for the amplification of the whole gene sequences encoding membrane protein were upstream primer, 5'-GGA TCC ATG GCA GAC AAC GGT AC-3' and downstream primer, 5'-GTC GAC CTG TAC TAG CAA AGC AAT AT-3'. The reaction was performed with a Perkin-Elmer 9700 thermal cycle. PCR was carried out in 100 μl reaction volume containing 50 pmol of each primer, 0.2 mmol/L of dNTP, 2.0 mmol/L. MgCl2, 5 IU of Taq polymerase and 1/10 total volume of 10× buffer. The cDNA was then transferred to 2 min at 94℃ and followed by 30 amplification cycles of 94℃ for 30 s, 52℃ for 30 s and 72℃ for 45 s and a 5 min terminal extension at 72℃. The PCR products were electrophoresed on 1.5 % agarose gel, got extracted and were cloned into vector pMD18-T.

1.2.3 Construction of the Recombinant Expression Plasmid Pet23a-M  
The sequence of gene enco-
ding M protein was obtained from recombinant plasmid pMD18-T-M which was digested by BamHI and SalI and purified as the target gene fragment. The plasmid Pet23a was digested by the same enzymes and the larger fragment was purified as the vector. The target gene fragment and the vector were joined to construct the plasmid Pet23a-M. Then the plasmid Pet23a-M was transformed into E. Coli DH5a and the screening of positive clones was performed by colony PCR method. The recombinant plasmid DNA was identified by restriction enzyme digestion and transferred into E. Coli BL21(DE3).

1.2.4 Induced Expression of Recombinant Gene Encoding Membrane Protein

Above constructed plasmid was transformed into the E. Coli strain, BL21(DE3), and signal colony was grown in LB medium with ampicillin (200 µg/ml) and glucose (10 mg/ml). When the culture was saturated, the culture was transferred to LB medium at a dilution of 1:50. The protein expression was induced with 0.2 mmol/L of IPTG after the culture medium was around 0.6. After IPTG induction at 25 °C for 10 h, the cells were harvested by centrifugation. The cell pellet was suspended by PBS buffer and incubated in a 100 °C water bath for 5 min. The suspension was centrifuged at 10,000 g for 10 min and the supernatants were collected for the SDS-PAGE analysis.

1.2.5 The Purification of the M Protein

The constructed plasmid was transformed into the E. Coli strain, BL21(DE3), and signal colony was grown in LB medium. The culture was incubated overnight at 37 °C. Then 6 ml of culture was transferred to 300 ml of LB medium at 37 °C. The protein expression was induced with l mmol/L of IPTG after the culture medium was around 0.6. After IPTG induction at 25 °C for 5 h, cells were harvested by centrifugation. The cell pellets were resuspended by buffer (20 mmol/L Tris-HCl pH 7.9, 500 mmol/L NaCl, 10 % glycerin). The cells were sonicated in ice bath. The lysed cells was centrifuged at 20,000 g at 4 °C for 10 min and pellets were discarded. Then the M protein was purified according to the manual of Pet Vector DNA from Novagen and SDS-PAGE was performed.

1.2.6 Western-blot of the Recombinant Membrane Protein

Protein samples were electrophoresed on SDS-PAGE and then transferred to nitrocellulose membrane, which blocked with phosphate buffered saline (PBS), 3 % non fat-dried milk and 0.05 % Tween 20. The membrane was incubated with mixed sera of 10 SARS patients during convalescence at a dilution of 1:100 at 4 °C overnight. After being washed by PBS-T (PBS and 0.05 % Tween 20) for 4 times, the blot was incubated with sheep anti-human immunoglobulin (G) labeled by HRP at a dilution of 1:5000 at 37 °C for 2 h. The blots were then washed and incubated with diaminobenzidine (DAB) until a brown precipitate formed in the protein band. The membrane was transferred to the PBS to stop the reaction.

2 RESULTS

2.1 Identification of the Recombinant Plasmids

2.1.1 PCR Analysis

PCR was performed by employing the SARS-CoV genomic RNA as templates. DNA fragment of about 700 bp in length was amplified and was the same as expected (fig. 1). Then PCR was performed by employing the positive plasmids as templates. DNA fragment of about 700 bp length was amplified and the same as reported.

![Fig. 1 RT-PCR of M gene](image)

| 1: RT-PCR products of M gene; M: DNA marker |

2.1.2 Identification by Restriction Endonuclease Analysis

The recombinant plasmid pMD18-T-M was treated with BamHI and SalI. The restriction fragments were conformed by agarose gel electrophoresis. The result showed that digestion with BamHI and SalI generated a 700 bp fragment. The restriction map was the same as expected (fig. 2).

![Fig. 2 The restriction enzyme analysis](image)

| 1: Pet23a/BamHI + SalI; 2: Pet23a-M; BamHI + SalI; M1, M2; DNA marker |

2.1.3 DNA Sequencing

The sequencing result showed that the sequence of M gene from recombinant plasmid pMD18-T-M was the same as reported in NCBI GenBank.

2.2 Expression of Recombinant M Protein

The cell lysate supernatants of recombinant E. Coli BL21 (DE3) were detected by SDS-PAGE and the expression of the 27 kDa protein (containing M protein and His6 • Tag at carboxy terminal tail and T7 • Tag at nitrol terminal tail), which was the same as expected, was detected (fig. 3).
2.3 Purification of Recombinant M Protein in pMD18-T-M

The fused protein containing M protein in the cell lysate supernatants was purified by Ni²⁺ chelated chromatography (fig. 3).

2.4 Western-blot Analysis of Recombinant M Protein

Results of Western-blot revealed that the recombinant protein could be recognized specifically by the antibodies of M protein in the sera of SARS patients, but could not be recognized by the sera of people without SARS. It provided the evidence that the recombinant M protein possessed the biological character of membrane protein of Coronavirus (fig. 4).

3 DISCUSSION

Coronavirus is enveloped positive-strand RNA viruses that contain 4 structural proteins and its genome is about 30 kb[32]. The S, M and N proteins have been broadly studied for their important roles in receptor binding and virion budding[4-7]. The M protein is the most conserved protein and is the major determinant of virion morphogenesis. It also interacts with the N protein presumably to assemble the nucleocapsid at the surface of the mature virion[51]. The M gene encodes a protein of 221 amino acids and its isoelectric point is 9.3. The sequence analysis of amino acids shows that the M protein contains three transmembrane domains but without signal peptide. It has a large carboxyl terminal tail with 121 amino acids, and this tail presumably interacts with the virus nucleocapsid. In this research, we designed a pair of primer according to the whole sequence of M gene in NCBI GenBank. Then the whole sequence of the gene was obtained by RT-PCR and was cloned into expression plasmid Pet23a. Restriction endonuclease analysis, PCR detection and DNA sequencing analysis revealed that the gene cloned was identical to the gene sequences from the reported strain. After it was transformed into E. Coli BL21 (DE3) and induced by IPTG, Western-blot showed that the expressed 27 kD protein which was characterized by SDS-PAGE and purified by metal chelated chromatography could react with antibodies in sera of SARS patients during convalescence, demonstrating the recombinant protein possessed the biological activity of membrane protein. It provided the evidence that M protein could be employed as antigen to prepare SARS virus vaccine and diagnostic agents. But the immunogenicity, the effect of anti-infection and the prospect in clinical application of the recombinant M protein need further research.

REFERENCES

1 Rota P A, Oberste M S, Monroe S S et al. Characterization of a novel coronavirus associated with severe acute respiratory syndrome. Science, 2003, 300(5624): 1394
2 Marr A M, Jones S J M, Astell C R et al. The complete sequence of the SARS-associated coronavirus. Science, 2003, 300(5624): 1399
3 Siddell S G, The coronavirusiae: an introduction, In: Siddell S G, editor. The coronavirusiae. New York: Plenum Press, 1995, 1-10.
4 Bosch B J, van der Zee R, de Haan C A et al. The coronavirus spike protein is a class I virus fusion protein: structural characterization of the fusion core complex. J Virol, 2003, 77(16): 8801
5 Kuo L, Masters P S. Genetic evidence for a structural interaction between the carboxy termini of the membrane and nucleocapsid protein of mouse hepatitis virus. J Virol, 2002, 76(10): 4987
6 Nguyen V P, Hogue B G. Protein interactions during coronavirus assembly. J Virol, 1997, 71(12): 9278
7 Narayanan K, Maeda A, Maeda J et al. Characterization of coronavirus M protein and nucleocapsid interaction in infected cells. J Virol, 2000, 74(17): 8127

(Received Dec. 26, 2003)