Task-Correlated Cortical Asymmetry and Intra- and Inter-Hemispheric Separation

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Cerebral lateralization is expressed at both the structural and functional levels, and can exist as either a stable characteristic or as a dynamic feature during behavior and development. The anatomically relatively simple olfactory system demonstrates lateralization in both human and non-human animals. Here, we explored functional lateralization in both primary olfactory cortex – a region critical for odor memory and perception-and orbitofrontal cortex (OFC) – a region involved in reversal learning- in rats performing an odor learning and reversal task. We find significant asymmetry in both olfactory and orbitofrontal cortical odor-evoked activity, which is expressed in a performance- and task-dependent manner. The emergence of learning-dependent asymmetry during reversal learning was associated with decreased functional connectivity both between the bilateral OFC and between the OFC-olfactory cortex. The results suggest an inter-hemispheric asymmetry and olfactory cortical functional separation that may allow multiple, specialized processing circuits to emerge during a reversal task requiring behavioral flexibility.

Classical examples of lateralized functions are human language and preferential handedness, though there is extensive evidence of asymmetry in cognitive and emotional processes1–5. This asymmetry is important for normal function given that deficits in hemispheric asymmetry are associated with impaired cognitive functions such as perception and memory6–11. Hemispheric specialization can enhance processing capabilities through specialization and compartmentalization, i.e., creation of new cortical space by reduction of bilateral redundancy5,12,13.

In humans, while simple odor detection does not induce asymmetrical cortical activation14, lateralization has been demonstrated in more complex olfactory cognitive functions14–17. In human subjects asked to recognize or judge the familiarity of odors, fMRI data suggest that these tasks are heavily mediated by right hemisphere structures such as the piriform cortex (PCX, primary olfactory cortex) and orbitofrontal cortex (OFC)14,18. Furthermore, damage to the right OFC, but not left OFC impairs odor recognition in humans19. In contrast, the ratings of odor hedonics (e.g., pleasant/unpleasant) are more dominated by the left OFC14,18.

Despite the potential importance of cortical asymmetry and lateralized disruption in cognitive disorders, there are few rodent models of cortical asymmetry that allow dissection of underlying mechanisms and identification of causal links between asymmetric function (or dysfunction) and cognitive/perceptual olfactory behavior. We recently demonstrated a robust, transient asymmetry in rat anterior piriform cortical (aPCX) function during specific stages of olfactory learning20. This asymmetry was expressed both as differential left-right aPCX activation over the course of odor learning, as well as performance- and context-specific changes in bilateral aPCX functional connectivity.

The aPCX receives direct input from the ipsilateral olfactory bulb and has strong reciprocal connections with the ipsilateral OFC21–23. Connections via the anterior commissure and corpus callosum allow bilateral communication between these olfactory structures, with for example cells in aPCX capable of responding to odors delivered to either the ipsilateral or contralateral naris24. While the aPCX is believed to play a crucial role in odor perception and memory25,26, the OFC has been demonstrated to be involved in a number of higher order functions, including those related to reversal learning27–29. Reversing the associations of a well learned memory requires significant behavioral flexibility, which we hypothesized might be facilitated by asymmetry in cortical function. Here, we recorded local field potential (LFP) oscillatory activity from bilateral OFC and from aPCX.

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in behaving rats to examine asymmetry in olfactory cortical function, as well as intra- and inter-hemispheric cortical functional connectivity, during initial odor learning and reversal learning. The results suggest that odor learning tasks requiring behavioral flexibility (reversal) are associated with olfactory cortical lateralization.

Results

Rats were trained in a two-alternative forced choice task for water reward to attain a criterion performance of at least 75% correct on a peppermint vs. vanilla task. They were then implanted with bipolar stainless steel electrodes aimed at both the left and right OFC and a third bipolar electrode aimed at either the left or right aPCX. The electrodes were attached to a connector that allowed telemetry recordings (EMKA) in freely moving animals during each day of subsequent training. Following at least two weeks of recovery, rats were trained with a new odor pair. Odors included peppermint, vanilla, anise, orange (all extracts from McCormick) or overlapping 10 component mixtures of monomolecular components as previously described. Rats generated a mean of 221 ± 7 trials/daily session. Rats that successfully reached criterion (75% correct) in the new odor discrimination task then underwent reversal training, where the odor that had originally signaled a reward in the right water port now signaled water reward in the left. LFP data from animals that successfully acquired the initial learning (n = 8) or both tasks (n = 6) to criterion (Fig. 1) were analyzed. Finally, rats were tested for handedness (see Methods). Our results were independent of the handedness expressed by the recorded rats (see Methods).

Given that rats acquired the tasks at different rates (e.g., mean sessions to criterion in initial learning = 14 ± 3, mean sessions to criterion for reversal = 24 ± 4), LFP data were analyzed in a performance-based manner as previously described. For both initial acquisition and reversal, performance stages were defined as 1) initial day of training, 2) 60–70% correct, 3) > 70% correct, and 4) overtraining past criterion performance.

Asymmetry in the PCX.

Event-related LFP oscillations were analyzed with FFT (3.9 Hz bin width). Analyses were focused on the 1 s period immediately before initiation of a trial by entering the odor sampling port and the 1 s period immediately after entering the odor sampling port. Although a variety of cognitive processes are occurring during these time periods, for simplicity the pre-odor sampling period is referred to here as “spontaneous” activity and the activity during odor sampling as “odor-sampling” related activity. Odor-sampling related activity was expressed as a percent of pre-sampling activity for normalization, and the normalized data was used for statistical comparisons both across hemispheres and across performance stages. LFP oscillation data were binned from the FFT analyses into broader frequency bands, though the major effects observed were within the beta frequency band (15.6–35.11 Hz) (Fig. 2A) which is the focus of the data shown. Beta band oscillations have been widely recognized as a sensitive marker of learning in the olfactory system.

As previously reported, during initial learning the left aPCX showed significantly enhanced beta band oscillations during odor sampling compared to the right aPCX (Fig. 2B). This asymmetry was most pronounced during performance stage 3, just as the animals reach criterion (2 × 8 ANOVA, hemisphere × performance stage: main effect of hemisphere, F(1,42) = 10.95, p < 0.05; main effect of stage, F(7,42) = 3.40, p < 0.01). Post-hoc tests
Fisher, p < 0.05) revealed a significant enhancement in the left aPCX beta oscillation power compared to right aPCX during initial learning stage 3. These results replicate our previous observations. In contrast to initial learning, reversal learning resets this asymmetry and lateralized differences in aPCX beta oscillations are much less pronounced during this phase of training. Post-hoc tests revealed no significant difference between the left and right aPCX during any stage of reversal learning.

If the performance-dependent lateralized changes in aPCX function involve asymmetry in cortical plasticity, we might expect markers of synaptic plasticity to also display performance-dependent asymmetry. Long-term synaptic plasticity is dependent on a variety of receptors and molecular cascades, one of which includes the PKC.
family. In fact, Olds et al.38, demonstrated that odor learning enhances PKC translocation to the membrane in the left aPCX more than in the right PCX. Here, we examined PKC gamma levels using immunohistochemistry in the aPCX in animals killed after each stage, or in naïve rats. No LFP recordings were made from these animals. PKC gamma levels were quantified with optical densitometry of immunohistochemical staining in anterior aPCX layer I (minimum of 3 sections/hemisphere/rat), and a ratio of left-right density used to compute a laterality score. As shown in Fig. 3, PKC gamma in the left PCX was significantly enhanced relative to the right, and relative to Naïve animals during stage 2/3 of initial odor learning as the animals approached criterion performance. A 2 x 7 repeated measures ANOVA, hemisphere (repeated) X training stage revealed a significant main effect of hemisphere (F(1,21) = 6.06, p < 0.05), and a significant interaction between hemisphere and performance (F(6,21) = 4.66, p < 0.005). Post-hoc tests revealed PKC gamma labeling was significantly biased toward the left aPCX during Stage 2/3 compared to other stages (Naïve, n = 5, Stage 1, n = 5, Stage 2/3, n = 6, all other stages, n = 3). No such asymmetry was observed in aPCX during reversal learning. These results suggest that the asymmetry in aPCX evoked LFP beta oscillations that emerges during initial learning is associated with asymmetric synaptic plasticity within the aPCX.

Asymmetry in a higher order olfactory area (OFC). LFP beta oscillations recorded bilaterally in the OFC also demonstrated asymmetry, though in contrast to the aPCX which showed a left bias, it was the right OFC that demonstrated significantly greater learning associated change than the left (Fig. 2C). A 2 x 8 ANOVA (hemisphere x performance stage) revealed a main effect of hemisphere, F(1,70) = 5.92, p < 0.05; main effect of stage, F(7,70) = 3.83, p < 0.01, and a significant hemisphere x stage interaction, F(7,70) = 3.15, p < 0.01). Post-hoc tests revealed PKC gamma labeling was significantly biased toward the left aPCX during Stage 2/3 compared to other stages (Naïve, n = 5, Stage 1, n = 5, Stage 2/3, n = 6, all other stages, n = 3). No such asymmetry was observed in aPCX during reversal learning. These results suggest that the asymmetry in aPCX evoked LFP beta oscillations that emerges during initial learning is associated with asymmetric synaptic plasticity within the aPCX.

Figure 3. (A) Representative PKC gamma immunohistochemistry in bilateral aPCX at Stage 3. (B) Side-by-side comparison of Stage 3 aPCX PKC gamma immunohistochemistry. (C) Mean (±SEM) asymmetry across initial and reversal learning shows elevated left PKC gamma staining during initial stages 2/3 compared to all other stages. Staining density was quantified in Layer I with optical densitometry (darker staining produced lower values) and compared across hemispheres within each animal (right aPCX/left aPCX, higher ratios signify great staining in the left aPCX). Asterisk signifies significant difference from all other stages, p < 0.05.
Asymmetry in the functional connectivity either between hemispheres or between OFC and aPCX. LFP coherence in the beta frequency band was quantified over the entire training session (initial learning: s1-s4; reversal learning r1-r4). (A) Pseudocolor summary of beta band coherence in each tested pathway. (B) Bilateral OFC beta band coherence across each performance stage. Reversal training induced a significant decrease in bilateral coherence compared to initial learning in the same animals (asterisk signifies significant decrease from initial Stage 1, p < 0.05). (C) Coherence between the left OFC and the ipsilateral and contralateral aPCX. During initial learning there was significantly greater coherence between the left OFC and ipsilateral aPCX than the left OFC and the contralateral aPCX (significant difference between pathways, p < 0.05). This enhanced ipsilateral coherence decreased during reversal conditioning (asterisk signifies significant decrease from initial Stage 1, p < 0.05). No learning associated change was observed in the left OFC to right aPCX pathway at any stage. (D) Coherence between the right OFC and the ipsilateral and contralateral aPCX. There was no change in beta coherence between the right OFC and the ipsilateral aPCX at any performance stage. In contrast, the right OFC displayed significantly greater coherence with the contralateral aPCX during initial learning (significant difference between pathways, p < 0.05), which then significantly declined during reversal learning compared to the ipsilateral pathway (p < 0.05) and compared to the first stage of initial learning (*p < 0.05).

Figure 4. Mean beta band LFP coherence between bilateral OFC and OFC-aPCX as quantified across each training session (initial learning: s1-s4; reversal learning r1-r4). (A) Pseudocolor summary of beta band coherence in each tested pathway. (B) Bilateral OFC beta band coherence across each performance stage. Reversal training induced a significant decrease in bilateral coherence compared to initial learning in the same animals (asterisk signifies significant decrease from initial Stage 1, p < 0.05). (C) Coherence between the left OFC and the ipsilateral and contralateral aPCX. During initial learning there was significantly greater coherence between the left OFC and ipsilateral aPCX than the left OFC and the contralateral aPCX (significant difference between pathways, p < 0.05). This enhanced ipsilateral coherence decreased during reversal conditioning (asterisk signifies significant decrease from initial Stage 1, p < 0.05). No learning associated change was observed in the left OFC to right aPCX pathway at any stage. (D) Coherence between the right OFC and the ipsilateral and contralateral aPCX. There was no change in beta coherence between the right OFC and the ipsilateral aPCX at any performance stage. In contrast, the right OFC displayed significantly greater coherence with the contralateral aPCX during initial learning (significant difference between pathways, p < 0.05), which then significantly declined during reversal learning compared to the ipsilateral pathway (p < 0.05) and compared to the first stage of initial learning (*p < 0.05).
During reversal learning, while OFC-aPCX connectivity is maintained in the right hemisphere, the rest of the network is disrupted, resulting in lateralized, relatively separated, cortical components.

**Discussion**

The present data suggest a robust performance and task-dependent asymmetry in the rat OFC and PCX, as well as in the functional connectivity between these cortical structures. These results replicate and extend our previous work on asymmetry in the aPCX to show that, within the same animals, OFC and aPCX display distinct patterns of asymmetry, with a left aPCX bias during initial odor discrimination learning, and a right OFC bias during initial learning and especially reversal learning. The left aPCX bias is associated with enhancement in PKC gamma staining in the left aPCX, suggesting lateralized synaptic plasticity may underlie this form of asymmetry. In addition to the region- and task-specific lateralization, functional connectivity between these structures also varied with task, with significant decreases in bilateral OFC and in OFC-aPCX coherence during the reversal task. Interestingly, our previous work demonstrated a similar reduction in aPCX bilateral coherence during initial learning. By analyzing the data from individual rats, we argue that intra- and inter-olfactory cortical separation is not simply correlated to the difficult nature the task per se but rather to the flexibility required for task. Reversing of the initial acquired information requires significant behavioral flexibility which expresses in extensive cortical separation. This model reflects non-holistic dynamic computation during flexible olfactory tasks. Overall, functional lateralization can enhance circuit specialization and allow for parallel processing, improving speed and efficiency, especially during tasks requiring behavioral flexibility.

There are a variety of mechanisms that could account for the lateralization and functional isolation of these olfactory circuits, including differential effects of neuromodulators and/or synaptic/cellular plasticity. For example, recent work has demonstrated lateralization in hippocampal contributions to memory in rodents mediated by asymmetrical synaptic plasticity. There may also be evidence of asymmetry in plasticity in the olfactory system. For example, our results here demonstrate an asymmetry in aPCX PKC gamma, a marker sensitive to neural plasticity that aligns well with aPCX LFP changes. The PKC changes observed here support an earlier study of odor memory associated PCX lateralization in PKC. Furthermore, two separate studies that investigated learning-induced synaptic strength in the OFC - aPCX pathway found distinct patterns of learned changes depending on the hemisphere investigated. Following odor-rule learning, there is a potentiation of OFC evoked responses in the ipsilateral aPCX in the right hemisphere, but a depression of OFC evoked responses in the ipsilateral aPCX in the left hemisphere. Importantly, we find no differences in the physiological modifications between left and right handed rats (see Methods). This further supports a behaviorally dependent source for cortical olfactory lateralization in rodents. Further work is required to identify specific mechanisms of dynamic laterality, though these studies suggest asymmetry in neural plasticity may be an important factor in odor processing and memory.

Together, the results suggest that the rodent olfactory cortex displays task- and performance-dependent asymmetry, similar to that observed in the human olfactory system. The lateralization of aPCX corresponds to changes in expression of the plasticity associated PKC gamma. OFC lateralization appears most pronounced during reversal learning, a task known to be OFC-dependent. As this area-specific lateralization emerges during reversal learning, there is a concomitant functional de-coupling of these regions, allowing hemispheric and regional

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**Figure 5.** Schematic diagram summarizing learning induced changes in bilateral OFC/aPCX trial-evoked beta oscillations and functional connectivity. The elevation in beta power occurred selectively during stage 3 of either initial or reversal learning. The decrease in functional connectivity was expressed across all stages of reversal learning relative to initial learning.
specialization. These findings open the opportunity for future studies of olfactory cortical asymmetry contributions to odor memory in condensed non-spatial paradigms, for example as in odor-fear conditioning.

Methods

Subjects. Male Long-Evans hooded rats (250–450 g) were used as subjects. Animals were housed individually on a 12 hr light/dark cycle, with food and water available ad libitum unless specifically noted. Tests were performed during the light portion of the day/night cycle. All handling, housing and experimental procedures were approved by, and performed in accordance with the Nathan Kline Institutional Animal Care and Use Committee guidelines (IACUC protocol number AP2014–489).

Paw preference test. Previous research indicates that there is an equal distribution of left or right handedness in rodents while Tang and Verstynen mention earlier studies suggesting a slight right-handedness preference. Here, handedness was tested using a standard reaching task. 10cc syringe with tip cut off to 4 ½ cm, syringe plunger, Froot Loops cereal pieces (about 0.05 to.08 grams each). Testing occurred in the rats’ homecage. With each rat, the first few cereal bits were pushed to the end of the syringe to allow the rat to get the treat easily with their mouth. Gradually the cereal was kept further back to 3 cm so that the rat would have to use his paw to grab it, allowing us to determine their handedness. A rat was classified as right or left handed if it reached at least 75% with one hand, a criterion which all animals attained (see Supplemental Fig. 1).

Chronic electrophysiology. Bipolar stainless steel electrodes (127 μm diameter wire, minimal distance between the two tips of approximately 20 μ) were implanted bilaterally in the Ventralis lateralis pars oralis (VLO; coordinates from Bregma - Anterior: ±3.2 mm, Medial: ±2 mm, Ventral: ±4 mm) and in layer III of the left or right anterior PCX (aPCX; coordinates from Bregma - Anterior: 2 mm, Medial: ±4 mm, Ventral: 6 mm) under isoflurane anesthesia. Electrodes and a ground lead were placed posterior to the skull with dental acrylic and attached to a connector. Following at least 2 weeks of recovery, the head connector was attached to a telemetry transmitter (EMKA Technologies) which allowed free movement and performance in the odor discrimination task while recording LFP’s. The bipolar differential recording signals were amplified (4000x) and transmitted at 1 KHz to the telemetry receiver, and then fed to an analog to digital converter and analyzed with Spike2 software (Cambridge Electronic Design). LFP recordings were synchronized with simultaneously recorded behavioral markers (sampling and water port entries and exits) in the operant chamber (Vulintus). Following the termination of testing, animals were overdosed with anesthetic, transcervically perfused with 10% formalin, and brains sectioned to confirm electrode placements.

Behavioral training and testing. Animals were given limited access to water during behavioral training and body weight monitored to ensure no animal lost more than 15% of initial body weight. All animals gained weight over the course of training. Odor discrimination was assessed in a two-alternative choice, Go-Left, Go-Right odor discrimination task for water reward. Animals received 30 min training sessions, 5 days/week. Individual animals received a single training session/day and generated >100 trials/session. Trials were initiated by a nose poke in the odor port (at least 350 ms, with odor onset beginning 100 ms after poke onset). Odor delivery terminated on nose withdrawal from the port. Water reward was delivered, depending on odor identity, upon a correct choice of the left or right reward port. All the animals were first trained on a vanilla versus peppermint discrimination until criterion (performance >75%) was attained. Animals reached this criterion in (mean ± SD) 14 ± 3 d.

Following criterion performance in the vanilla–peppermint discrimination, animals were anesthetized and bilateral electrodes were implanted as described above. The animals were allowed at least 2 weeks to recover, and then began training with a new odor pair. Odors included anise vs. orange or an odor mixture discrimination. The 10 component odorant mixtures which have been described in detail elsewhere included: isovaleric acid (100PPM), nonane (100PPM), neryl valerate (100PPM), 5-methyl-2-hexenone (100PPM), isopropylbenzene (100PPM), 1-pentanol (100PPM), 1,7-octadiene (400PPM), 2-heptanone (100PPM), heptanal (100PPM), 4-methyl-3-penten-2-one (100PPM). For reversal mixture discrimination training, we used the same pair of odors but in a reversed direction from the initial learning. Performance on the task was divided into stages based on individual ability. For both initial acquisition and reversal, performance stages were defined as 1) initial day of training, 2) 60–70% correct, 3) > 70% correct, and 4) overtraining past criterion performance.

LFP data analyses. Bilateral OFC and aPCX LFP data were analyzed for each animal from a single training session within each stage of behavioral performance as previously described. Briefly, analyses included the 1s period immediately before entering the odor sampling port and the 1s period immediately after entering the odor sampling port regardless of the outcome of the trial – correct, error or aborted trial. Although a variety of cognitive processes are occurring during these time periods, for simplicity the pre-odor sampling period is referred to here as “spontaneous” and the activity during odor sampling as “odor-sampling” related activity. The 1 second sampling periods were chosen to allow sufficient data of even low frequency oscillations for accurate Fast Fourier Transform (FFT) analyses (3.9 Hz resolution). Odor-sampling evoked activity was expressed as a percent of spontaneous activity for normalization, and the normalized data was used for statistical comparisons both across hemispheres and across learning stages. LFP oscillation data were binned from the FFT analyses into broader frequency bands, including theta (3.9–11.7 Hz), beta (15.6–35.11 Hz), low gamma (39.06–50.78 Hz) and high gamma (70.31–101.56 Hz) bands.

LFP coherence between the left and right OFC and between the OFC and ipsilateral and contralateral aPCX was also assessed over the entire sessions (30 min). The coherence was determined using a “cohere” script function within Spike2 at 5-Hz resolution.
PKCgamma Immunohistochemistry and quantification. A group of rats separate from those used for electrophysiology were trained as described above and killed after attaining a specific performance stages, or were killed as naïve rats. Rats were transcardially perfused with 4% paraformaldehyde and post-fixed in glycerol-formaldehyde before sectioning at 40µm. Standard immunohistochemistry was performed for PKCgamma (primary antibody AbCam, ab71558) and visualized with Vector VIP peroxidase substrate. Images were taken of bilateral aPCX and optical density of staining in Layer I was determined using ImageJ image analysis software. Low values represented higher levels of staining. The right/left ratio was determined as a metric of PKCgamma asymmetry in PCX at each performance stage.

References
1. Hugdahl, K. Symmetry and asymmetry in the human brain. European Review 13(Supp. 2), 119–133 (2005).
2. Kaveic, V., Zhong, J., Yoshiura, T. & Doty, R. W. Frontal cortex, laterality, and memory: encoding versus retrieval. Acta Neurobiologiae Experimentalis 63, 337–350 (2003).
3. Lowell, S. Y., Reynolds, R. C., Chen, G., Horwitz, B. & Ludlow, C. L. Functional connectivity and laterality of the motor and sensory components in the volitional swallowing network. Experimental Brain Research 219, 85–96 (2012).
4. Smaers, J. B., Mulveny, P. L., Soligo, C., Zilles, K. & Amunts, K. Sexual dimorphism and laterality in the evolution of the primate prefrontal cortex. Brain, Behavior and Evolution 79, 205–212 (2012).
5. Habib, R., Nyberg, L. & Tulving, E. Hemispheric asymmetries of memory: the HERA model revisited. Trends Cogn Sci 7, 241–245 (2003).
6. Kelley, W. M. et al. Hemispheric specialization in human dorsal frontal cortex and medial temporal lobe for verbal and nonverbal memory encoding. Neuron 20, 927–936 (1998).
7. Dennis, N. A., Kim, H. & Cabeza, R. Effects of aging on true and false memory formation: an fMRI study. Neuropsychologia 45, 3157–3166 (2007).
8. Jacobs, H. I. et al. Relevance of parahippocampal-tempo-coeruleus connectivity to memory in early dementia. Neurobiol Aging (2014).
9. Liu, Q. et al. Decreased coherence between the two olfactory bulbs in Alzheimer's disease model mice. Neuroscience Letters 545, 81–85 (2013).
10. Thompson, P. M. et al. Cortical variability and asymmetry in normal aging and Alzheimer's disease. Cereb Cortex 8, 492–509 (1998).
11. Pailly, J., d'Amato, T., Saoud, M. & Royet, J. P. Left tempo-temporal-occipital and orbital dysfunction in schizophrenia during odor familiarity and hedonicity judgments. Neuroimage 29, 302–313 (2006).
12. Vallortigara, G., Rogers, L. J. & Busizza, A. Possible evolutionary origins of cognitive brain lateralization. Brain Res Brain Res Rev 30, 164–175 (1999).
13. Tommasi, I. Mechanisms and functions of brain and behavioural asymmetries. Philos Trans R Soc Lond B Biol Sci 364, 855–859 (2009).
14. Royet, J. P. & Pailly, J. Lateralization of olfactory processes. Chemical Senses 29, 731–745 (2004).
15. Thierauf, N. et al. Olfactory lateralization: odor intensity but not the hedonic estimation is lateralized. Neuroscience Letters 438, 228–232 (2008).
16. Zatorre, R. J., Jones-Gotman, M., Evans, A. C. & Meyer, E. Functional localization and lateralization of human olfactory cortex. Nature 360, 339–340 (1992).
17. Zucco, G. M. & Tressoldi, P. E. Hemispheric differences in odour recognition. Cortex; a journal devoted to the study of the nervous system and behavior 25, 607–615 (1989).
18. Royet, J. P. et al. Functional neuroanatomy of different olfactory judgments. Neuroimage 13, 506–519 (2001).
19. Hudry, J. et al. Lateralization of olfactory processing: differential impact of right and left temporal lobe epilepsies. Epilepsy Behav 37, 184–190 (2014).
20. Cohen, Y., Putrino, D. & Wilson, D. A. Dynamic cortical lateralization during olfactory discrimination learning. J Physiol 593, 1701–1714 (2015).
21. Chen, C. F. et al. Nonsensory target-dependent organization of piriform cortex. Proceedings of the National Academy of Sciences of the United States of America 111, 16931–16936 (2014).
22. Illig, K. R. Projections from orbitofrontal cortex to anterior piriform cortex in the rat suggest a role in olfactory information processing. The Journal of Comparative Neurology 488, 224–231 (2005).
23. Cleland, T. A. & Linster, C. Central olfactory structures. In: Handbook of olfaction and gustation (ed. (eds Doty R. L.). 2nd edn. Marcel Dekker (2003).
24. Wilson, D. A. Binaral interactions in the rat piriform cortex. Journal of Neurophysiology 78, 160–169 (1997).
25. Gottfried, J. A. Central mechanisms of odour object perception. Nat Rev Neurosci 11, 628–641 (2010).
26. Wilson, D. A. & Sullivan, R. M. Cortical processing of odor objects. Neuron 72, 506–519 (2011).
27. Feistner, C. E., Quirk, M. C., Uchina, N., Sosuiki, D. L. & Mainen, Z. F. Representation of spatial goals in rat orbitofrontal cortex. Neuron 51, 495–507 (2006).
28. Schoenbaum, G., Gottfried, J. A., Murray, E. A. & Ramus, S. J. Critical contributions of the orbitofrontal cortex to behavior. In: Annals of the New York Academy of Sciences (ed. (eds). New York Academy of Sciences (2011).
29. Schoenbaum, G., Nugent, S. L., Suddoris, M. P. & Selkow, B. Orbitofrontal lesions in rats impair reversal but not acquisition of go/no-go odor discriminations. Neuropsychologia 13, 885–890 (2002).
30. Schoenbaum, G. & Roesch, M. Orbitofrontal Cortex, Associative Learning, and Expectancies. Neuron 47, 633–636 (2005).
31. Barnes, D. C., Hofacer, R. D., Zaman, A. R., Rennaker, R. L. & Wilson, D. A. Olfactory perceptual stability and discrimination. Nature Neuroscience 11, 1378–1380 (2008).
32. Lovitz, A. M., Sloan, A. M., Rennaker, R. L. & Wilson, D. A. Complex mixture discrimination and the role of contaminants. Chemical Senses 37, 533–540 (2012).
33. Chen, C. E., Barnes, D. C. & Wilson, D. A. Generalized versus stimulus-specific learned fear differentially modifies stimulus encoding in primary sensory cortex of awake rats. Journal of Neurophysiology 106, 3136–3144 (2011).
34. Chapuis, J. & Wilson, D. A. Bidirectional plasticity of cortical pattern recognition and behavioral sensory acuity. Nature Neuroscience 15, 155–161 (2012).
35. Kay, L. M. & Beshel, J. A beta oscillation network in the rat olfactory system during a 2-alternative choice odor discrimination task. Journal of Neurophysiology 104, 829–839 (2010).
36. Martin, C., Gervais, R., Messaoudi, B. & Ravel, N. Learning-induced oscillatory activities correlated to odor recognition: a network activity. Eur J Neurosci 23, 1801–1810 (2006).
37. Martin, C. & Ravel, N. Beta and gamma oscillatory activities associated with olfactory memory tasks: different rhythms for different functional networks? Frontiers in behavioral neuroscience 8, 218 (2014).
38. Olds, J. L. et al. Lateralization of membrane-associated protein kinase C in rat piriform cortex: specific to operant training cues in the olfactory modality. Behavioural Brain Research 61, 37–46 (1994).
39. Sakaguchi, Y. & Sakurai, Y. Left-right functional asymmetry of ventral hippocampus depends on aversiveness of situations. Behavioural Brain Research 325, 25–33 (2017).
40. Shipton, O. A. et al. Left-right dissociation of hippocampal memory processes in mice. *Proceedings of the National Academy of Sciences of the United States of America* **111**, 15238–15243 (2014).

41. Cohen, Y., Reuveni, I., Barkai, E. & Maroun, M. Olfactory learning-induced long-lasting enhancement of descending and ascending synaptic transmission to the piriform cortex. *The Journal of Neuroscience* **28**, 6664–6669 (2008).

42. Cohen, Y., Wilson, D. A. & Barkai, E. Differential modifications of synaptic weights during odor rule learning: dynamics of interaction between the piriform cortex with lower and higher brain areas. *Cereb Cortex* **25**, 180–191 (2015).

43. Biddle, E. G. & Eales, B. A. The degree of lateralization of paw usage (handedness) in the mouse is defined by three major phenotypes. *Behav Genet* **26**, 391–406 (1996).

44. Vyazovskiy, V. V. & Tobler, I. Handedness leads to interhemispheric EEG asymmetry during sleep in the rat. *Journal of Neurophysiology* **99**, 969–975 (2008).

45. Tang, A. C. & Verstynen, T. Early life environment modulates ‘handedness’ in rats. *Behavioural Brain Research* **131**, 1–7 (2002).

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**Author Contributions**

Y.C. and D.A.W. designed the research, Y.C. collected the data, Y.C. and D.A.W. analyzed the data, Y.C. and D.A.W. prepared the manuscript.

**Additional Information**

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