The Gela Basin pockmark field in the strait of Sicily (Mediterranean Sea): chemosymbiotic faunal and carbonate signatures of postglacial to modern cold seepage

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Abstract. The geo-biological exploration of a pockmark field located at ca. 800 m below sea level in the Gela basin (Strait of Sicily, Central Mediterranean) provided a relatively diverse chemosymbiotic community and methane-imprinted carbonates. To date, this is the first occurrence of such a type of specialised deep-water cold-seep communities recorded from this key region, before documented in the Mediterranean as rather disjunct findings in its eastern and westernmost basins. The thiotrophic chemosymbiotic organisms recovered from this area include empty tubes of the vestimentiferan Lamellibrachia sp., loose and articulated shells of lucinids (Lucinoma kazani, Myrtea amorpha), vesicomyids (Isorropodon perplexum), and gastropods (Taranis moerchii). A callianassid decapod (Calliax sp.) was consistently found alive in large numbers in the pockmark mud. Their post-mortem calcified parts mixed with molluscs and subordinately milolid foraminifers form a distinct type of skeletal assemblage. Carbonate concretions display δ13C values as low as ~40‰ PDB suggesting the occurrence of light hydrocarbons in the seeping fluids. Since none of the truly chemosymbiotic organisms was found alive, although their skeletal parts appear at times very fresh, some specimens have been AMS-14C dated to shed light on the historical evolution of this site. Lamellibrachia and Lucinoma are two of the most significant chemosymbiotic taxa reported from various Mediterranean cold seep sites (Alboran Sea and Eastern basin). Specimens from station MEDCOR78 (pockmark#1, Lat. 36°46’10.18” N, Long. 14°01’31.59” E, 815 m below sea level) provided ages of 11736 ± 636 yr cal BP (Lamellibrachia sp.), and 9609.5 ± 153.5 yr cal BP (L. kazani). One shell of M. amorpha in core MEDCOR81 (pockmark#6, Lat 36°45’38.89” N, Long 14°00’07.58” E, 822 m below sea level) provided a sub-modern age of 484 ± 54 yr cal BP. These ages document that fluid seepage at this pockmark site has been episodically sustaining thiotrophic macrobenthic communities since the end of the Younger Dryas stadial up to sub-recent times.

1 Introduction

Pockmarks are circular to sub-elliptical crater-like depressions of variable size (few to many hundreds of metres in diameter) and depth (up to few tens m) typically due to degassing of fluids through fine-grained unconsolidated marine sediment under a variety of geologic scenarios (King and McLean, 1970; Hovland, 1989, 1992; Scanlon and Knebel, 1989; Kelley et al., 1994; Gay et al., 2003; Trincardi et al., 2004; Paul et al., 2007). Pockmarks often occur in clusters and cover wide areas, representing a common and widespread feature on the continental margins worldwide (Hovland and Judd, 1988). They receive much attention especially because escaping fluids are often hydrocarbon-enriched venues in the form of slow seepage, vigorous venting or even eruptions, at times representing a distinct geo-hazard for offshore constructions and navigation (Newton et al., 1980; Hovland, 1987; Curzi et al., 1988; Curzi, 2012). Such fluid releases are also considered as a potential co-actor...
in triggering mass-sediment failure along continental margins, in conjunction with sea level changes or earthquakes (Minisini and Trincardi, 2009). Pockmarks are known to host at times peculiar biota in response to nature of escaping fluids, rate of emissions, seepage resilience and water depth (Hovland and Thomsen, 1989; Hovland and Judd, 1988; Taviani, 2011, 2013). Gas charged sediments may in fact sustain microbiological communities operating on common compounds in fluids, i.e. hydrogen sulphide and hydrocarbons like methane (Cavanaugh et al., 2006; Dando, 2001; Duperron, 2010). In turn, such communities could be exploited by specialized metazoans through complex symbiotic relationships (see MacDonald et al., 1990; Olu-Le Roy et al., 2007; Taviani, 2001; Duperron, 2010, with references). Pockmarks are also sites were methane-related authigenic carbonates could be generated (Hovland et al., 1987; Hovland and Judd, 1988; Gontharet et al., 2007).

In the frame of the EU Hermes and Hermione projects, the Strait of Sicily in the central Mediterranean was targeted as a key area to study sediment instability processes that affect continental margins. During such exploration a field of pockmarks was charted and sampled in detail off the southern margin of Sicily (Fig. 1). Here we present first data on deep sea pockmarks in the Gela basin resulting in novel information about resident chemosymbiotic communities, biosedimentological processes, authigenic carbonates and seepage temporal variability in this sector of the Mediterranean Sea connecting its western and eastern basins.

2 Material and methods

The study area was surveyed during cruises HERMES05 (July 2005), MEDCOR and DECORS December 2009 and August 2011, respectively, of R/V Urania. Swath bathymetry data were acquired using a Kongsberg Simrad EM710 multibeam echosounder with a nominal sonar frequency of 70–100 kHz an angular coverage sector of 140°, a beam width of 1° × 1° and 800 soundings per ping in dual mode. Chirp-sonar profiles were obtained using a hull-mounted sixteen-transducer source with a sweep modulated 2–7 kHz outgoing signal equivalent to a 3.5 kHz profile.

Bottom samplings were performed using a 1.2 ton gravity corer, large volume grab and epibenthic hauls (Table 1).

Water-column attributes were measured with a Conductivity/Temperature/Depth profiler (CTD) using a Seabird SBE 11 PLUS employing the SEASAVE V5.33 software.
Table 1. Sampling station list of Cruises MEDCOR and DECORS in the Gela Basin.

| Station | Date       | Start Lat. GGMSS.XX | Start Long. GGMSS.XX | End Lat.  | End Long.  | Start depth | End depth | Gear          |
|---------|------------|---------------------|---------------------|-----------|-----------|-------------|-----------|---------------|
| MEDCOR-68 | 19 Dec 2009 | 36°45'38.24'' | 14°00'09.02'' | 36°45'35.00'' | 14°00'10.82'' | 820.0 | 820.0 | CTD          |
| MEDCOR-69 | 19 Dec 2009 | 36°45'37.83'' | 14°00'08.20'' | 36°45'15.69'' | 14°01'00.61'' | 840.8 | 840.6 | Gravity Core |
| MEDCOR-70 | 19 Dec 2009 | 36°45'16.42'' | 14°01'01.07'' | 36°44'30.77'' | 13°58'48.06'' | 824.0 | 850.0 | Epibenthic modified haul |
| MEDCOR-71 | 19 Dec 2009 | 36°45'16.07'' | 14°00'58.79'' | 36°45'35.00'' | 14°00'10.82'' | 840.0 | 820.0 | Epibenthic modified haul |
| MEDCOR-72 | 19 Dec 2009 | 36°45'15.11'' | 14°01'01.18'' | 36°45'10.18'' | 14°01'31.59'' | 842.0 | 811.0 | Epibenthic modified haul |
| MEDCOR-74 | 19 Dec 2009 | 36°45'38.82'' | 14°00'10.35'' | 36°44'30.77'' | 13°58'48.06'' | 824.0 | 850.0 | Epibenthic modified haul |
| MEDCOR-75 | 21 Dec 2009 | 36°45'48.41'' | 14°01'15.29'' | 36°46'10.18'' | 14°01'31.59'' | 833.0 | 811.0 | Epibenthic modified haul |
| MEDCOR-76 | 21 Dec 2009 | 36°45'34.56'' | 14°00'24.85'' | 36°46'10.18'' | 14°01'31.59'' | 820.2 | 811.0 | Epibenthic modified haul |
| MEDCOR-77 | 21 Dec 2009 | 36°45'44.57'' | 13°58'52.79'' | 36°46'10.18'' | 14°01'31.59'' | 831.7 | 811.0 | Epibenthic modified haul |
| MEDCOR-78 | 21 Dec 2009 | 36°45'40.13'' | 14°00'08.90'' | 36°46'10.18'' | 14°01'31.59'' | 815.0 | 811.0 | Epibenthic modified haul |
| MEDCOR-80 | 19 Dec 2009 | 36°45'31.18'' | 13°59'54.57'' | 36°45'40.10'' | 13°59'49.79'' | 820.0 | 815.0 | Epibenthic modified haul |
| MEDCOR-81 | 19 Dec 2009 | 36°45'37.84'' | 14°00'08.21'' | 36°45'40.10'' | 13°59'49.79'' | 832.0 | 811.0 | Gravity Core |
| MEDCOR-82 | 19 Dec 2009 | 36°45'16.28'' | 14°01'01.08'' | 36°46'16.86'' | 13°55'28.42'' | 840.9 | 811.0 | Grab         |
| DECORS-48 | 8 Aug 2011  | 36°46'16.86'' | 14°00'06.80'' | 36°46'16.86'' | 13°55'28.42'' | 816.0 | 811.0 | CTD          |
| DECORS-49 | 8 Aug 2011  | 36°45'39.24'' | 14°00'06.64'' | 36°46'16.86'' | 13°55'28.42'' | 818.0 | 811.0 | Grab         |
| DECORS-50 | 9 Aug 2011   | 36°46'16.86'' | 13°55'28.42'' | 36°46'16.86'' | 13°55'28.42'' | 827.0 | 811.0 | Grab         |

Stable isotope analyses were performed at the Mass Spectrometry Laboratory of the CNR-IGG (Institute for Geosciences and Earth Resources) Pisa, using a Finnigan MAT252 mass-spectrometer. The carbonate samples were cleansed, powdered and then analysed following standard procedures by reacting carbonate aliquots with 100% orthophosphoric acid under vacuum. Results are expressed as ‰ relative to VPDB (Vienna-Pee Dee Belemnite) standard by assigning a value of +1.95 ‰ (13C) and a value of −2.2 ‰ (18O) exactly to NBS 19 calcite; the analytical error is ±0.1 ‰ (Table 2).

14C-AMS dating was carried out at the Poznán Radiometric Laboratory, Poland (Table 3).

3 The pockmark field of Gela

Pockmarks are common features at many continental margins in the Mediterranean basin (Stefanoff, 1981; Stefanoff et al., 1983; Curzi and Veggiani, 1985; Mazzotti et al., 1987; Hovland and Curzi, 1989; Trincardi et al., 2004; Geletti et al., 2008; Coleman et al., 2012; Curzi, 2012; Somoza et al., 2012). They have been recently documented also in various locations of the Strait of Sicily, although most published studies regard relatively shallow-water (ca. 70–200 m) occurrences (Savini et al., 2009; Canigemi et al., 2010; Micaleff et al., 2011). Pockmarks do occur in the Gela basin at bathyal depths (Minisini and Trincardi, 2009, this paper).

Gela basin is the Plio-Quaternary foredeep of the Maghrebian fold-and-thrust belt and is a site of widespread and repeated mass transport. Gela Slide, the largest mass transport complex in the area, has affected the northern margin of the basin mobilising a volume of sediment in the order of 1000 km³ in mid-Pleistocene times (Trincardi and Argnani, 1990). Following this major basin-wide event, several smaller-scales mass failure events impacted the area up to the Holocene (Minisini et al., 2007) and with recurrence intervals in the order of a few thousands of years between successive events (Minisini and Trincardi, 2009). The most recent mass-transport events involved, in various mixes, two basic distinctive kinds of slope sediments: the Pleistocene clinoforms that prograde with increasing slope angle and outbuilding the shelf seaward; and upper Pleistocene to Holocene muddy contourite deposits characterised by water-saturated and poorly consolidated sediment (Verdicchio and Trincardi, 2008). In all cases where stacked MTD deposits appear sandwiched within basin-wide draped units, the latter units appear disrupted by vertical features that have been ascribed to fluid-escape features from MTDs that became rapidly buried and perhaps pressurized (Minisini and Trincardi, 2009).

The Gela basin pockmark field (GBPF) is located in the Strait of Sicily, ca. 20 nm offshore the southern coast of Sicily in a depth range of 800–900 m (Fig. 1). Firstly detected upon TOBI side scan sonar in 2003 and by low-resolution multibeam bathymetric data in 2005, the GBPF was then surveyed more in detail in winter 2009 and again in summer 2011 during Hermione cruise DECORS when high resolution multibeam data were acquired, and additional pockmark fields further west, identified.
The resulting detailed morpho-bathymetric map documents the existence of discrete clusters of pockmarks in a restricted sector of the Gela basin (Figs. 1–4). Our data do not show any clear preferential orientation of these features. All pockmarks in the studied field are clustered in seven seafloor patches for a total area of about 18 km². The pockmarks present generally a sub-circular shape, with the exception of the pockmark #1 (Fig. 2) which is elliptical shaped, and range in diameter from 40 to 310 m, although most of them are between 40 and 100 m and only 11 (Fig. 2) are between 200 m and 310 m. The bottom of some of these is as much as 20 m below the surrounding seafloor; most however are less than 6 m deep. In a cross section, extracted from the bathymetric profile, the pockmarks in most cases appear to be U-shaped; but four of them are V-shaped and (Fig. 2) show a positive relief ranging from 4 m to 7 m along their rim. The side slope ranges from 7° to 13° reaching a maximum of 16°. The regional surrounding slope is about 2–3°. Width to depth ratio were calculated for 11 pockmarks from bathymetric profiles crossed near the feature centres. The ratio fell between 11 : 1 and 40 : 1 with most near 18 : 1. There is not consistent relation between this ratio and relative size of the pockmark.

Noticeably, acoustic signals relatable to the presence of gas entrapped in the sediment (such as BSR) or to active degassing (plumes) were not detected during both surveys (Fig. 5). Furthermore, CTD profiles show no T–S anomalies.

The largest depression with an internal bulge, resulting from the coalescence of two pockmarks, was selected as the prime target for bottom sampling. The entire area is a fishing ground subjected for many years to a peculiar local fishing (mal)practice known as canizzi that litters the bottom by leaving behind scores of dead weights with long plastic threads attached (Taviani, 2010). Before proceeding with grab and core sampling, and CTD casting, it was therefore necessary to clear first the bottom by removing this litter. This goal was achieved by using a small modified haul that in fact successfully entangled some such canizzi remains (threads and some weights), the former intensely over grown by live cold water corals. A second trawl on the same trajectory intercepted the bottom collecting many dead shells of Lucinoma clams and tubeworms, living ghost shrimps and a few carbonate concretions amidst other benthic organisms and litter (Fig. 7a). Only at this stage, this large pockmark remained free of debris and ready for deeper sampling. By leaving behind scores of dead weights with long plastic threads attached (Taviani, 2010).

### Table 2. Stable Isotopes Analysis expressed in the conventional δ notation relative to the Vienna-Pee Dee Belemnite (VPDB) reference standard.

| Sample   | Material             | Species                  | $^{13}\text{C} \delta_{\text{PDB}}^{\text{vs}}$ | σ$^{13}\text{C}$ | $^{18}\text{O} \delta_{\text{PDB}}^{\text{vs}}$ | σ$^{18}\text{O}$ |
|----------|----------------------|--------------------------|-----------------------------------------------|-----------------|-----------------------------------------------|-----------------|
| MEDCOR70 | Shell                | Lucinoma kazani          | 0.82 ± 0.1                                   | 2.71 ± 0.1      | 2.66 ± 0.1                                    | 2.48 ± 0.1      |
| MEDCOR78 | Shell                | Isorropodon perplexum    | −1.23 ± 0.1                                  | 2.33 ± 0.1      | 2.21 ± 0.1                                    | 2.1 ± 0.1       |
| MEDCOR81 | Shell                | Myrtea amorpha           | 0.16 ± 0.1                                   | 2.33 ± 0.1      | −1.84 ± 0.1                                   | 3.23 ± 0.1      |
| MEDCOR81 | Claw                 | Calliax sp.              | −40.77 ± 0.1                                 | 4.98 ± 0.1      | −4.05 ± 0.1                                   | 3.18 ± 0.1      |
| MEDCOR70 | Mudstone             |                          | −37.57 ± 0.1                                 | 2.86 ± 0.1      | 6.75 ± 0.1                                    | 2.86 ± 0.1      |

### Table 3. Radiocarbon data from the Poznán Radiocarbon Laboratory in Poznán, Poland. Radiocarbon years ($^{14}\text{C}$-yr) correspond to the true half-life. Calibrated age ranges reflect the 95.4 % probability window (2σ-error) and were calculated with Calib6.0.1, using the surface marine calibration curve MARINE09 by Reimer et al. (2009).

| Code #   | Sample   | Species                  | Radiocarbon Age ($^{14}\text{C}$-yr BP) | Delta R (yr) | Two-σ ranges (yr cal BP) | Age cal BP (yr cal BP) | Calibration data |
|---------|----------|--------------------------|-----------------------------------------|-------------|--------------------------|------------------------|------------------|
| Poz-38025| MEDCOR78 | Lamellibrachia sp.       | 10500 ± 200                             | 41 ± 7      | 11 121–12 343             | 11 732 ± 611           | Marine09         |
| Poz-37995| MEDCOR78 | Lucinoma kazani          | 8960 ± 60                               | 41 ± 7      | 9456–9763                | 9610 ± 154             | Marine09         |
| Poz-55055| MEDCOR78 | Isorropodon perplexum    | 7920 ± 50                               | 41 ± 7      | 8082–8583                | 8333 ± 250             | Marine09         |
| Poz-55053| MEDCOR69 | Taranis moerchii         | 7820 ± 130                              | 41 ± 7      | 7901–8589                | 8245 ± 344             | Marine09         |
| Poz-37994| MEDCOR81 | Myrtea amorpha           | 1090 ± 30                               | 41 ± 7      | 547–669                  | 486 ± 54               | Marine09         |

The largest depression with an internal bulge, resulting from the coalescence of two pockmarks, was selected as the prime target for bottom sampling. The entire area is a fishing ground subjected for many years to a peculiar local fishing (mal)practice known as canizzi that litters the bottom by leaving behind scores of dead weights with long plastic threads attached (Taviani, 2010). Before proceeding with grab and core sampling, and CTD casting, it was therefore necessary to clear first the bottom by removing this litter. This goal was achieved by using a small modified haul that in fact successfully entangled some such canizzi remains (threads and some weights), the former intensely over grown by live cold water corals. A second trawl on the same trajectory intercepted the bottom collecting many dead shells of Lucinoma clams and tubeworms, living ghost shrimps and a few carbonate concretions amidst other benthic organisms and litter (Fig. 7a). Only at this stage, this large pockmark remained free of debris and ready for deeper sampling. By leaving behind scores of dead weights with long plastic threads attached (Taviani, 2010).

### 4 Fauna

The living component in the benthic macro-and megafauna inside the pockmarks at the time of sampling was rather scarce and mostly represented by sparse bivalves...
Fig. 2. Shaded relief DTM of the study area (5 m resolution and 10 × vertical exaggeration) showing the complex, rough topography punctuated by many sub-circular depressions interpreted as pockmarks. The 11 major pockmarks, with diameters greater than 200 m, are numbered from maximum to minimum dimension. Their geo-morphometric characteristics are listed in the table; the bathymetric profiles show the cross section of two main pockmarks showing the different internal geometry, respectively U-shaped and V-shaped with internal positive relief. Furthermore, the image reveals that the major pockmark (Number 1) is formed by two partly coalescent depressions.

Table 4. List of major established (chemosymbiotic bivalves, and tubeworm) and putative (gastropods, callianassid) metazoans associated with pockmark cold seepage at the GBPF.

| MEDCOR | MEDCOR | MEDCOR | MEDCOR | MEDCOR | MEDCOR | MEDCOR | DECORS | DECORS |
|--------|--------|--------|--------|--------|--------|--------|--------|--------|
| 69     | 71     | 76     | 78     | 80     | 82     | 49     | 50     |
| Lucinoma kazani | x | x | x | | | |
| Salas and Woodside, 2002 | | | | | | |
| Myrtea amorpha | x | | | | | | |
| (Sturany, 1896) | | | | | | |
| Isorropodon perplexum | x | x | x | x | x | x | x | x |
| Sturany, 1896 | | | | | | |
| Taranis moerchii | x | | x | x | x | x | x | x |
| (Malm, 1861) | | | | | | |
| Lamellibrachia sp. | x | x | x | x | x | x | x | x |
| Calliix sp. | x | x | x | x | x | x | x | x |

(Ennucula corbuloides, Cardiomya costellata, Kelliella miliaris), caprellid amphipods and ghost shrimps. Numerous specimens of a callianassid ghost shrimp (Calliix sp.: Fig. 7f, Table 4) were found in the pockmark area. In the quantitative grab samples its density ranged from 1 up to 7 live animals/0.2 square metre whereas number of loose main claws ranged between 23 and 128. This is likely a still undescribed species morphologically close to an elusive callianassid taxon reported from shallower settings in the Western Mediterranean (de Gaillande and Lagardere, 1966). This species is consistently associated with such type of environments since a few specimens of this same shrimp have been...
also found in active pockmarks in the central Adriatic (Taviani et al., 2010). Members of the family Callianassidae are known to inhabit deep sea reducing habitats worldwide (summarised by Komai and Fujiwara, 2012). In the eastern Atlantic sector close to the Mediterranean, a new subfamily (Vulcanocalliacinae) and a new species (Vulcanocalliax ayrtyunovi) have been recently described by Dworschak and Cunha (2007) from a mud volcano in the deep Gulf of Cádiz.

The macrofaunal death assemblages recovered from the GBPF show a predominance of benthic molluscs over other components, predominantly decapods, pelagic molluscs, echinoids, serpulids and octocorals, Non-seep organisms are more diverse in terms of number of species (Appendix A), but chemosymbiotic bivalves in the families Lucinidae, and Vescicomyidae are at places noticeably abundant (Table 4). Lucinids are represented by the large Mediterranean endemic taxon Lucinoma kazani Salas and Woods, 2002 (Fig. 7c), and subordinately by Myrtea amorpha (Sturany, 1896) (Fig. 7e). Vescicomyidae are represented by the Mediterranean endemic Isorpododon perplexum Sturany, 1896 (Fig. 7b), the sole species recorded that far from the recent Mediterranean basin. Such bivalves have been recorded from deep sea reducing habitats in the Mediterranean basin (Corselli and Basso, 1996; Cosel and Salas, 2001; Olu-Le Roy et al., 2004; Werne et al., 2004; Duperron et al., 2007; Duperron, 2010; Ivanov et al., 2010a, b; Ritt et al., 2010; Brissac et al., 2011; Shank et al., 2011; Taviani, 2011; Rodríguez et al., 2012).

Another remarkable megabenthic organism found in the GBPF is a siboglinid tubeworm of which only empty tubes have been collected by us (Fig. 7a). The morphology of the tube resembles Lamellibrachia, a genus recorded from deep water cold seep sites (Olu-Le Roy et al., 2004; Werne et al., 2004; Duperron et al., 2009; Ritt et al., 2010; Hilário et al., 2011; Taviani, 2011), sunken wrecks (Hughes and Crawford, 2006; Gambi et al., 2011), and hydrothermal vents (Lott and Zimmerman, 2012) in the basin. Tubes might belong to Lamellibrachia anaximandri (Southward, Andersen and Hourdea, 2011), described from the Anaximander mud volcano in the eastern Mediterranean (Southward et al., 2011), and possibly much widespread in the entire Mediterranean basin (references in Taviani, 2013). In addition to these well known deep-water chemosymbiotic metazoans, we can further list a few other taxa preferentially coping with reducing environments, i.e. thyasirid (Thyasira, Mendicula, Axinulus, Axinus) and lucinid (Myrtea spinifera)
Fig. 4. Slope map of the study areas, calculated from the DTM, showing clearly the difference between the side slope of the pockmarks reaching the maximum of $16^\circ$ and the regional surrounding slope about $2-3^\circ$.

Fig. 5. (A) Chirp profile showing the subsurface stratigraphy of the pockmark area; (B) 3-D perspective view of the DTM shows the location of the chirp profile and the sample locations (yellow); red numbers are pockmarks.
Fig. 6. Chirp sonar profiles across the pockmark-hosting morphologic bulge at the base of the slope showing the top of the last basin-wide mass-transport event (Gela slide?) and the thinning of plane-parallel reflectors (basin-plain fine-grained turbidites and hemipelagites) toward the bulge. Where thinner, this unit is likely easier to breach by the ascending fluids than in other areas.

bivalves (Dufour, 2005; Löfler et al., 2005; Taylor et al., 2007; Taylor and Glover, 2010; Brissac et al., 2011; Rodrigues and Duperron, 2011), and the turrid gastropod *Taranis moerchii* (Fig. 7d) that has been observed to be particularly common at such seep sites (Olu-Le Roy et al., 2004; M. Taviani, unpublished data). Only some pockmarks were found to contain spoils of truly seep communities that are instead totally absent from the sea bottom outside the pockmarks (Appendix A). Furthermore, the more emblematic cold seep species (e.g. *L. kazani, I. perplexum, Lamellibrachia sp.*) are not co-occurring in all investigated pockmarks in the Gela basin, showing instead a spatial variability. On the other hand, spatiotemporal variability in analogue habitats responding to variability through time and space of fluids’ composition and flow intensity is a well documented trait at modern and past seepage sites (Olu et al., 1997;
Fig. 7. Chemosymbiotic metazoans recovered from the GBPF: (A) haul whole catch of station MEDCOR 78 (pockmark #1) showing chemosymbiotic bivalves, tubeworms, and carbonate concretions mixed with non-seep fauna and anthropogenic litter; (B) *Isorropodon perplexum* from st. MEDCOR71, (C) *Lucinoma kazani*, theratological shell from st. MEDCOR78, (D) *Taranis moerchii* from st. MEDCOR69, (E) *Myrtea amorpha* from st. MEDCOR78, (F) *Calliax* sp. from st. MEDCOR71.

Jenkins et al., 2007; Olu-Le Roy et al., 2007; Lessard-Pilon et al., 2010). As it is often the case, the sustainability of such chemosymbiotic bivalves and siboglinids relies upon high sulphide concentrations since their metabolic pathway is thiotrophic. Thus, they cannot be considered as direct fingerprints of hydrocarbon availability in the seeping fluids at the GBPF.

5 Seep carbonates

- **Sediments.** The pockmark field affects a predominantly siliciclastic-muddy sector of the southern Sicilian margin. Sampling however reveals the occurrence of biogenic carbonate production in the form of skeletal components shed primarily by calcified benthic organisms and subordinately by pelagics. A distinct type of carbonate biogenic sediment is produced here through cold seep factories in the form of medium to coarse skeletal assemblages enriched in shells of chemosymbiotic bivalves, mainly vesicomyids (20–40%), and calcified parts of callianassid decapods (30–50%) in the sand to silt fraction (Fig. 8). Secondary components in order of abundance are miliolid foraminifers (5–10%), followed by echinoids, serpulids, ostracods and octocorals and pelagic shells. This remarkable sediment represents a novelty since its genesis is in fact related to chemical carbonate factories.

- **Authigenic carbonates.** Besides shells of chemosymbiotic bivalves and tubeworms, sampling from the GBPF produced also carbonate concretions, indurated burrows (Figs. 8 and 9), and cemented mudstones (Fig. 10). Microcrystalline pyrite druses were noticed on small darker carbonate concretions. Some carbonates were tested for their stable carbon and oxygen isotopic composition (Table 2, Fig. 12). The $\delta^{13}C$ composition of chemosymbiotic bivalve shells display isotopic values around between $-1.23$ and $+2.66\% \text{PDB}$ reflecting the seawater carbonate reservoir as expected (Cobabe, 1998; Lietard and Pierre, 2009). The stable carbon composition of a poorly cemented layer in core MEDCOR70, provided instead a $\delta^{13}C$ value of $-40.77\% \text{PDB}$. A significantly carbon-depleted composition was also measured for concretioned mudstone encrusting a dead shell of the chemosymbiotic *Lucinoma kazani* which provided a $\delta^{13}C$ value of $-37.57\% \text{PDB}$. Both very negative carbon signatures document unquestionably that at sometime hydrocarbons were present in the seeping fluids of this pockmark field in the Gela basin. On the other hand, this region is known to be highly productive for hydrocarbons (Etiope et al., 2002; Holland et al., 2003).

The stable carbon range is consistent with authigenic carbonates formed through the incorporation of carbon derived from anaerobic methane oxidation (Hovland et al., 1987; Roberts et al., 1987; Kulm and Suess, 1990; Aharon, 1994; Kelly et al., 1995; von Rad et al., 1996; Aloisi et al., 2000, 2002a, b; Elvert et al., 2000; Naehr et al., 2000; Greinert et al., 2001; Peckmann et al., 2001; Taviani, 2001, Gontharet et al., 2007; Campbell et al., 2008; Cangemi et al., 2010; Feng and Roberts, 2010; Ivanov et al., 2010a, b; Himmler et al., 2011; Capozzi et al., 2012). The presence of pyrite might represent a legacy of sulphate reduction in the formative environment of such carbonates (e.g. Gontharet et al., 2007). Interestingly, the GBPF concretions represent a case of incipient embedding of chemosymbiotic infaunal and epifaunal organisms in authigenic carbonates (Lalou et al., 1992), eventually turning into macrofossiliferous cold seep rocks as those diffused in the Neogene Apennine chain (Taviani, 1994, 2001, 2011). Finally, the stable oxygen composition
of bivalves ($\delta^{18}$O values comprised between +2.21 and −2.71 ‰ PDB) are interpretable as indicative of bottom cold temperatures (cf. Sakai et al., 1992), thus supporting the late glacial ages obtained by radiocarbon dating (see below).

6 Age constraints

The absence of distinct signals documenting either active degassing or gas charged sediment close to surface and the lack of live chemosymbiotic macro-and megafauna was taken as an indication that seepage backing the genesis of the GBPF is at present either dormant/episodic or ended. In order to unravel this fundamental aspect we examined the core record, and carbon-dated faunas and carbonate concretions. Superficial bottom samples and two sedimentary cores were taken from pockmark #1 and #6 (respectively MEDCOR81 and MEDCOR70) to reconstruct the recent history of fluid seepage at this place (see Fig. 11). Regarding pockmark #1, a tube of *Lamellibrachia* produced an AMS-$^{14}$C age of 11732 ± 611 yr cal BP, while a valve of *Lucinoma kazani* yielded an AMS-$^{14}$C age of 9610 ± 154 yr cal BP. *Isorropodon perplexum* and *Taranis moerchii* shells from the same pockmark yielded slightly younger but roughly comparable ages of 8333 ± 250 yr cal BP and 8245 ± 344 yr cal BP respectively. A valve of *Myrtea amorpha* from sediment core MEDCOR70 (pockmark #6) provided an AMS-$^{14}$C age of 484 ± 54 yr cal BP. Although potentially somewhat influenced by carbon sourced from seeping fluids, our radiocarbon ages clearly indicate that seepage activity in the pockmarks is lasting, albeit not necessarily continuously, since the end of the last glacial epoch (Younger Dryas stadial) up to sub-recent times.

7 Discussion

The pockmark area investigated in this study is located at the base of the northern slope of Gela basin where Chirp-sonar profile detect the top of the last basin-wide MTD at a depth of few tens of metres under a succession of basin-floor muddy turbidites. The pockmarks area are not randomly distributed but corresponds to a bulge at the base of slope with a positive relief of few km$^2$ (Figs. 1 and 3) and the overlying succession, characterised by draped and onlapping plain-parallel reflectors (muddy turbidites?) thins toward the slope and are breached by the pockmarks (Fig. 6). The perfect matching of the pockmark field with the slide-related base-of-slope bulge is suggestive of a possible causal relationship. In this view, the fluids emanating through the pockmarks would be fed from the MTD, acoustically-transparent and possibly gas charged deposit. However, in the lack of deeper-penetration MCS profiles we cannot rule out the possibility that fluids come, at least in part, from beneath the MTD rather than from it.

The genesis of the GBPF under study is thus not fully understood yet but a potential link between defluidization (pockmarks) and a main motif of this part of the continental margin (mass sediment failure) merits it to be considered. In fact, this cluster of pockmarks in the Gela basin are just located distally in front of a complex made up by stacked multiple submarine mass-transport deposits of late Pleistocene age (Minisini et al., 2007). These composite units are by large buried and only a few are exposed at present on the seabottom, the best examples of which are the “twin slides” whose multi-phase emplacement occurred since the last Post-Glacial (Minisini et al., 2007; Minisini and Trincardi, 2009). Pore fluid gradients, resulting in fluid migration and eventual expulsion onto the sea bottom, are suggested to be a potential co-driving factor, although not necessarily the dominant, in triggering episodic mass-transport
Fig. 9. Carbonates from the GBPF: (A–D) authigenic concretions (cemented matrix) embedding skeletal biosomes of seep-related metazoans, all from st. MEDCOR78: (A–B) Calliax, (C–D) Isorropodon; (E) indurated callianassid? burrow, armoured with chemosymbiotic-organisms bioclasts; (F) cemented matrix embedding a reworked black-stained shallow-water foraminifer (Elphidium crispum); (G) cemented mudstone layer at 210–230 cm in core MEDCOR70.

Fig. 10. Incipient methane-driven cementation of matrix on valves of chemosymbiotic bivalves from GBPF, precursors of seep-limestones: (A) Lucinoma kazani encrusted by carbon-depleted concretion ($\delta^{13}$C – 37.57‰ PDB, see text), (B) Isorropodon perplexum, almost completely embedded in mudstone.
Fig. 11. Sedimentary cores MEDCOR81 (pockmark #1), and MEDCOR70 (pockmark #6): photographic (left) and (right) lithostratigraphic logs reporting main chronological, petrographic, isotopic and faunal features.
Fig. 12. Stable isotope compositions of bivalve shells (core MEDCOR 81), callianassid claw and authigenic carbonates all from st. MEDCOR70 and MEDCOR78 expressed in the conventional δ notation relative to the PDB reference standard.

events that shape this region (Minisini et al., 2007; Minisini and Trincardi, 2009). The GBPF under scrutiny here lies ca. 5 nm away to the west of the “twin slides” and does not appear to have contiguity with such features, suggesting that no causative link between the two phenomena (twin-slide-slumping and pockmark-fluid expulsion) does exist. A series of still-unexplored pockmarks has been, however, identified at the toe of the “twin slides” (Minisini and Trincardi, 2009). It is predictable that they may in principle contain similar faunas and carbonates as their counterparts. The multiswath topographic map suggests the subsurface existence of now-draped lobate bodies northwards of the pockmarks, that in all likeness represent buried mass-transport deposits comparable to those described by Minisini et al. (2007) from this general area. The mise-en-place process of the slumping units may in theory cause squeezing out of entrapped fluids or, otherwise, the under pressuring of fluids related to sliding off sediment that may be conducive to gas release pochmarking the bottom. Release of hydrocarbons has been invoked as a likely mechanism to trigger submarine slumps (Carpenter, 1981 and many others). On-land, similar causative scenarios have been suggested to account for localized Miocene chemosymbiotic faunal assemblages and/or authigenic carbonates associated with slumped bodies in the Apennine chain (Berti el al., 1994; Conti and Fontana, 2002; Lucente and Taviani, 2005).

The GBPF shows a certain degree of homogeneity regarding the chemosymbiotic assemblages inhabiting individual pockmarks. This is especially true for small vesti-

comyids (Isorropodon), gastropods (Taranis) and callianassid decapods (Calliax) that are almost ubiquitous in our samples. Such homogeneity is not at all surprising once we consider the overall reduced variability of cold seepage at the GBPF when compared to larger and heterogeneous seep sites as those observable elsewhere in the ocean (Olu et al., 1997; Sibuet and Olu-Le Roy, 2002; Olu-Le Roy et al., 2007; Cordes et al., 2010), or even in the Mediterranean itself (Olu-Le Roy et al., 2004). Such variability often reflects primarily variations in the fluid regimes (Teichert et al., 2003). Nevertheless, minor faunal differences are equally observable at the GBPF such as, for example, the comparable rarity of the large chemosymbiotic clam L. kazani or of the large vestibimentiferan Lamellibrachia which are not recorded thus far from more than two pockmarks.

The GBPF is midway between the Eastern Mediterranean and Alboran basins where a variety of cold seep habitats is exploited by macro- and megabenthic organisms as those found in the study area (e.g., Lucinoma, Isorropodon and Lamellibrachia). Thus, it represents a relevant site to be considered in the crucial and poorly-known issue of connectivity patterns among geographically disjunct but compositionally similar deep sea chemosynthetic sites (e.g., Young, 2003; Tyler and Young, 2003; Olu-Le Roy et al., 2007; Marcus et al., 2009; Mullineaux et al., 2010).

8 Conclusions

1. The Gela basin deep-water pockmark field hosting Mediterranean-emblematic thiotrophic chemosymbiotic macro- and mega fauna (Lucinoma, Isorropodon, Lamellibrachia) is relevant as it is the first such site in this key-region of the Mediterranean Sea transitional to its eastern and western basins;

2. the episodic occurrence of hydrocarbons in the seeping fluids is demonstrated by the occurrence of carbonate concretions and mudstones with very depleted stable carbon isotope composition;

3. the quite different ages provided by organisms from individual pockmarks suggest that seepage activity in the area is not a uniformly-distributed and steady event but differs in space and time;

4. dating of chemosymbiotic Lamellibrachia tubeworms documents that these biota were thriving since the end of the last glacial epoch (Younger Dryas stadial) and up to very recent times;

5. a possible causative link between seepage with related pockmark genesis, and mass-transport processes is tentatively suggested but still undemonstrated.
### Table A1. Benthic and pelagic molluscs recovered from the GBPF during cruises MEDCOR and DECORS: taxa of proven or putative chemosymbiotic type are bold.

| Benthic Mollusc | MEDCOR 69 | MEDCOR 71 | MEDCOR 76 | MEDCOR 78 | MEDCOR 80 | MEDCOR 82 | DECORS 49 | DECORS 50 |
|----------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| Bivalvia       |           |           |           |           |           |           |           |           |
| Cardiomya costellata (Deshayes, 1833) | x         |           | x         | x         | x         | x         | x         | x         |
| Cuspidaria rostrata (Spengler, 1793) | x         | x         |           |           |           |           |           |           |
| Trachia sp.    |           |           |           |           |           | x         |           |           |
| Lacinoma kazani Salas and Woodside, 2002 | x         |           | x         | x         |           | x         |           |           |
| Myrtea sp.     |           |           |           |           |           |           |           |           |
| Myrtea amorpha (Sturany, 1896) | x         |           |           |           |           |           |           |           |
| Myrtea spinifera (Montagu, 1803) | x         | x         |           |           |           |           |           |           |
| Axinulus croulensis (Jeffreys, 1847) | x         | x         |           |           |           |           |           |           |
| Axinulus eumyarus (M. Sars, 1870) | x         |           |           |           |           |           |           |           |
| Axinus grandis (Verrill and Smith [in Verrill], 1885) | x         | x         | x         | x         | x         | x         |           |           |
| Mendoicula ferruginosa (Forbes, 1844) | x         |           |           |           | x         | x         | x         |           |
| Thyasira sp.   |           |           |           |           |           |           |           |           |
| Thyasira succisa (Jeffreys, 1876) | x         |           |           |           |           |           |           |           |
| Cortula gibba (Olivi, 1792) | x         |           |           |           | x         |           |           |           |
| Xylophaga sp.  |           |           |           |           |           |           |           |           |
| Acanthocardia (Gray, 1851) | x         |           |           |           |           |           |           |           |
| Parvicardium sp. |           |           |           |           |           |           |           |           |
| Kuriella cf. indentata (Montagu, 1803) | x         |           |           |           |           |           |           |           |
| Mellia obtusa (G. O. Sars, 1872) | x         |           |           |           |           |           |           |           |
| Saccella commutata (Philippi, 1844) | x         |           |           |           |           |           |           |           |
| Microcoeloma sp. |           |           |           |           |           |           |           |           |
| Yoldiella curta Verrill and Bush, 1898 | x         |           |           |           |           |           |           |           |
| Yoldiella lucida (Lovén, 1846) | x         |           |           |           |           |           |           |           |
| Yoldiella nana (M. Sars, 1865) | x         |           |           |           |           |           |           |           |
| Yoldiella sequenzae Bondito and Sabelli, 1995 | x         | x         | x         | x         | x         | x         | x         | x         |
| Yoldia minima G. Seguenza, 1877 | x         |           |           |           |           |           |           |           |
| Ennucula sp.   |           |           |           |           |           |           |           |           |
| Ennucula aegerensis (Forbes, 1844) | x         |           | x         | x         |           |           |           |           |
| Ennucula corbuloidea (Seguenza, 1877) | x         | x         | x         | x         |           |           |           |           |
| Nucula sulcata Bronn, 1831 | x         |           |           |           |           |           |           |           |
| Nucula tumidula Malm, 1861 | x         | x         |           |           |           |           |           |           |
| Asperarca nodulosa (O. F. Müller, 1776) | x         |           |           |           |           |           |           |           |
| Bathysarcapectunculoides (Scacchi, 1835) | x         | x         | x         | x         |           |           |           |           |
| Limatula gwyni (Sykes, 1903) | x         |           |           |           |           |           |           |           |
| Limatula subauriculata (Montagu, 1808) | x         | x         |           |           |           |           |           |           |
| Notolima crassa (Forbes, 1844) | x         | x         |           |           |           |           |           |           |
| Parvamussium sp. | x         |           |           |           |           |           |           |           |
| Pectinidae sp. |           |           |           |           |           |           |           |           |
| Gastropoda     |           |           |           |           |           |           |           |           |
| Bittium sp.    |           |           |           |           |           |           |           |           |
| Turritella communis Risso, 1826 | x         |           | x         | x         |           |           |           |           |
| Epitonium sp.  |           |           |           |           |           |           |           |           |
| Halisella stenostoma (A. Adams, 1861) | x         |           |           |           |           |           |           |           |
| Triphoridae sp. |           |           |           |           |           |           |           |           |
| Actis attenuatus Jeffreys, 1883 | x         |           |           |           |           |           |           |           |
| Melanella sp.  |           |           |           |           |           |           |           |           |
| Naticidae sp.  |           |           |           |           |           |           |           |           |
| Euspira fusca (Blainville, 1825) | x         |           |           |           |           |           |           |           |
| Alvania sp.1   |           |           |           |           |           |           |           |           |
| Alvania sp.2   |           |           |           |           |           |           |           |           |
| Alvania cimicoides (Forbes, 1844) | x         | x         | x         | x         | x         | x         | x         | x         |
| Alvania elegansmissa (Monterosato, 1875) | x         | x         | x         |           |           |           |           |           |
| Alvania subsulcata (Aradas, 1847) | x         | x         | x         | x         |           |           |           |           |
| Alvania testae (Aradas and Maggiore, 1844) | x         |           |           |           |           |           |           |           |
Table A1. Continued.

|                        | MEDCOR 69 | MEDCOR 71 | MEDCOR 76 | MEDCOR 78 | MEDCOR 80 | MEDCOR 82 | DECORS 49 | DECORS 50 |
|------------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| Benthonella tenella (Jeffreys, 1869) | x | x | x | x | x | x | x | x |
| Aporhais serresianus (Michaud, 1828) | x | x | x | x | x | x | x | x |
| Galeodona echinophora (Linnaeus, 1758) | x | x | x | x | x | x | x | x |
| Amphitrite acutecostata (Philippi, 1844) | x | x | x | x | x | x | x | x |
| Fusinus rostratus (Olivieri, 1792) | x | x | x | x | x | x | x | x |
| Drilolosa emendata (Monterosato, 1872) | x | x | x | x | x | x | x | x |
| Drilolosa loprestiana (Calcara, 1841) | x | x | x | x | x | x | x | x |
| Spirotropis modiolus (de Cristofori and Jan, 1832) | x | x | x | x | x | x | x | x |
| Mangelia serga (Dall, 1881) | x | x | x | x | x | x | x | x |
| Pleurotomella gibbura Bouchet and Warén, 1980 | x | x | x | x | x | x | x | x |
| Taronis moerchi (Malm, 1861) | x | x | x | x | x | x | x | x |
| Teretia tenes (Reeve, 1844) | x | x | x | x | x | x | x | x |
| Granulina gastro Smiglio & Mariotti, 1996 | x | x | x | x | x | x | x | x |
| Pagodula echinata (Kiener, 1840) | x | x | x | x | x | x | x | x |
| Trophonopsis barvicensis (Johnston, 1825) | x | x | x | x | x | x | x | x |
| Bathysciadium cyclophagum Warén and Carrozza in Warén, 1997 | x | x | x | x | x | x | x | x |
| Acteon pusillus (Forbes, 1844) | x | x | x | x | x | x | x | x |
| Crenilabium exile (Jeffreys, 1870) | x | x | x | x | x | x | x | x |
| Eulimella sp. | x | x | x | x | x | x | x | x |
| Syrion sp. | x | x | x | x | x | x | x | x |
| Colpodaspis sp. | x | x | x | x | x | x | x | x |
| Diaphana lactea (Jeffreys, 1877) | x | x | x | x | x | x | x | x |
| Philine sp. | x | x | x | x | x | x | x | x |
| Philine scabra (Müller, 1784) | x | x | x | x | x | x | x | x |
| Philine monterosato (Sykes, 1905) | x | x | x | x | x | x | x | x |
| Rosania monterosato (Dautzenberg and H. Fischer, 1896) | x | x | x | x | x | x | x | x |
| Fissurisepta granulosa Jeffreys, 1883 | x | x | x | x | x | x | x | x |
| Patuzesia wiseri (Calcara, 1842) | x | x | x | x | x | x | x | x |
| Ditolepis sp. | x | x | x | x | x | x | x | x |
| Polyplacophora | x | x | x | x | x | x | x | x |
| Hanleya sp. | x | x | x | x | x | x | x | x |
| Scaphopoda | x | x | x | x | x | x | x | x |
| Antalis agilis (M. Sars in G.O. Sars, 1872) | x | x | x | x | x | x | x | x |
| Dentalium sp. | x | x | x | x | x | x | x | x |
| Entalina tetragona (Brocchi, 1814) | x | x | x | x | x | x | x | x |
| Pelagic Mollusc | x | x | x | x | x | x | x | x |
| Caeno gastropoda | x | x | x | x | x | x | x | x |
| Caninaria lamarkii Blainville, 1817 | x | x | x | x | x | x | x | x |
| Atlanta brunnea J. E. Gray, 1850 | x | x | x | x | x | x | x | x |
| Atlanta peronii Lesueur, 1817 | x | x | x | x | x | x | x | x |
| Heterobranchia | x | x | x | x | x | x | x | x |
| Cavolinia infesa (Lesueur, 1813) | x | x | x | x | x | x | x | x |
| Cavolinia tridentata (Forsskål in Niebuk, 1775) | x | x | x | x | x | x | x | x |
| Cavolinia uncinita (Rang, 1829) | x | x | x | x | x | x | x | x |
| Diacria trispinosa (Blainville, 1821) | x | x | x | x | x | x | x | x |
| Clio pyramidalis Linnaeus, 1767 | x | x | x | x | x | x | x | x |
| Cereis clava (Rang, 1828) | x | x | x | x | x | x | x | x |
| Hyaloxyris siroto (Rang, 1828) | x | x | x | x | x | x | x | x |
| Styliola subula (Quoy and Gaimard, 1827) | x | x | x | x | x | x | x | x |
| Helicocoides inflatus (d'Orbigny, 1834) | x | x | x | x | x | x | x | x |
| Limacina sp. | x | x | x | x | x | x | x | x |
| Limacina bulimoides (d'Orbigny, 1834) | x | x | x | x | x | x | x | x |
| Limacina trochoforma (d'Orbigny, 1834) | x | x | x | x | x | x | x | x |
| Peracca diversa (Monterosato, 1875) | x | x | x | x | x | x | x | x |
| Peracca reticulata (d'Orbigny, 1834) | x | x | x | x | x | x | x | x |

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