Effect of Cow Urine and Plant Growth Promoting Rhizobacteria (PGPR) on Seed Germination, Growth and Survival of Karonda (Carissa carandas L.) Seedlings

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ABSTRACT

The present investigation entitled “Effect of cow urine and Plant Growth Promoting Rhizobacteria (PGPR) on seed germination, growth and survival of Karonda (Carissa carandas L.) seedlings” was conducted at Fruit Research Station, Imalia, Department of Horticulture, College of Agriculture, J.N.K.V.V., Jabalpur (M.P.) during the year 2018 - 2019. The experiment was laid out in Factorial Randomized Block Design with three replications. The experiment consist of 8 treatments T0 (Control), T1 (25% cow urine), T2 (50% cow urine), T3 (100% cow urine), T4 (Pseudomonas 6ml/kg seeds), T5 (25% cow urine + Pseudomonas 6ml/kg seeds), T6 (50% cow urine + Pseudomonas 6ml/kg seeds), T7 (100% cow urine + Pseudomonas 6ml/kg seeds). All the seed and soil inoculation treatments showed better germination, growth and survival when compared to uninoculated control. Among various combinations, the combined effect of seed treatment with 25% cow urine and soil inoculation with 6ml/kg pseudomonas were more superior over the other combinations with highest seed germination (5.33 days), 50% germination (14.50 days), germination at 60 DAS (79.63%), coefficient of germination (6.10), speed of germination (9.15). Growth parameters at 60, 90 and 120 DAS with seedling height (8.75, 11.20 and 12.60 cm), stem girth (1.98, 2.36 and 2.64 mm), number of leaves per seedling (29.3, 72.2 and 89.4) respectively, other growth parameters at 120 DAS such as length of seedling (28.39 cm), number of roots per seedling (118.3), fresh weight of seedlings (3.41 g), dry weight of seedlings (1.43 g), seedling vigor index I (2269.7 cm), seedling vigor index II (106.8 g), survival of seedlings (81.4%), leaf area index (4.09), leaf area duration (677798 cm².days), light transmission ratio (34.52%) and energy interception (0.44 cal cm² min⁻¹).

KEYWORDS
Karonda (Carissa carandas L.), Cow urine, Plant growth promoting rhizobacteria, Pseudomonas

INTRODUCTION

Carissa carandas is native plant of India while 30 species of genus Carissa have been reported by Singh et al., (1967). It is cultivated throughout India in tropical and subtropical areas. The plants of Carissa are found in plains and hills and wild form in Deccan - Peninsula as well as in West Coast, Punjab, Kashmir, U.P., Mount Abu (Rajasthan), parts of Gujarat and West Bengal (Rai and Gupta, 1996). Karonda is a fruit of
dry areas and flourishes well on lands with high temperature. At present, it is grown on a limited scale in Rajasthan, Gujarat, Bihar and Uttar Pradesh. It is also grown in other countries viz. Bangladesh, South Africa, U.S.A., Denmark, Ghana, Israel, Pakistan, Nepal and Afghanistan. It is also found in some parts of Sri Lanka in the lowland rain forests eco-region. Karonda fruit is a rich source of iron (3.91°per on dry weight basis) and contains a fair amount of Vitamin C. Therefore, it is very useful for cure of anemia and has antiscorbutic properties. Vitamin A content of fruit is 1619 IU 100 g of edible portion as well as 87 to 90 per cent pulp, 13 to 14 per cent total soluble solids and 4 to 6 per cent acidity. Under the changing world trade scenario, it can be exploited on a commercial scale as a fruit for the processing industries.

Since, Karonda is very hardy and drought tolerant fruit crop but heavy rainfall and water logged conditions are not desirable. The roots of the plant are heavily branched and make it suitable for stabilizing eroding slopes. It can be grown on a wide range of soils including saline and sodic soils. Propagation by seed is the common method by which plant reproduces in nature and one of the most efficient and widely used propagation methods for raising seedling. Sowing of seed is the physical beginning of seedling propagation. Specific physical and chemical changes that take place during maturation and ripening of the fruit lead to fruit senescence and dissemination of the seed. One of the most obvious changes is the drying of the pericarp tissues. In certain species, this lead to dehiscence and the discharge of the seeds from the fruit. Seeds of most species dehydrate at ripening and prior to dissemination, when moisture content drop to 30 per cent or less. The seed dries further during harvest, usually to about 4-6 per cent for storage. Germination cannot take place at this level of dryness which is important basis for maintaining viability and controlling germination. In certain species, seed must not dry below which 30 to 50 per cent or they will lose their ability to germinate (Chin 1981). Plant species can be separate as recalcitrant or orthodox seed based on their genetic potential to tolerate storage. Most recalcitrant seed cannot tolerate moisture below 25 per cent and some species are also sensitive to chilling temperature. Karonda is commonly propagated by seed and fresh seeds are sown for raising seedlings in the month of August and September. Cow urine contains iron, urea, uric acid, estrogen and progesterone which affect the inhibitory response to seed germination, shoot growth and seedling vigor (Dilrukshi, 2009). Plant growth promoting rhizobacteria are the soil bacteria inhabiting around and, on the surface and are directly or indirectly involved in promoting plant growth and development via production and secretion of various regulatory chemicals in the vicinity of rhizosphere. They stimulate plant growth through mobilizing nutrients in soils, producing numerous plant growth regulators, protecting plants from phytopathogens by controlling or inhibiting them, improving soil structure and bioremediating the polluted soils by sequestering toxic heavy metal species and degrading xenobiotic compounds (like pesticides) (Ahemad, 2012; Hayat et al., 2010; Rajkumar et al., 2010; Braud et al., 2009).

Materials and Methods

Experimental site

The experiment was conducted at Fruit Research Station, Imalia, Department of Horticulture, College of Agriculture, J.N.K.V.V., Jabalpur, Madhya Pradesh, India. The experiment was conducted during October 2018 to February 2019. The experimental site was situated at 23.9ºN latitude and 79.58ºE longitude and an altitude of 411.78 meters above Mean Sea Level (MSL).
Seed materials

The seeds of Karonda (*Carissa carandas* L.) were obtained from Fruit Research Station of Jawaharlal Nehru Krishi Vishwa Vidyalaya, Imalia, Jabalpur District, Madhya Pradesh, India.

Plant Growth Promoting Rhizobacteria

Plant growth promoting rhizobacteria (*Pseudomonas*) was obtained from the Department of Soil Microbiology, Faculty of Agriculture, JNKVV, Jabalpur, Madhya Pradesh.

Preparation of seed soaking media

25 ml and 50 ml fresh cow urine was measured with measuring cylinder and poured into 100 ml beaker each.

The volume was made up by addition of distilled water to prepare 25 per cent and 50 per cent solution of cow urine respectively. 100 ml fresh cow urine in 100 ml beaker gave 100 per cent solution of cow urine.

Seed Treatment

Freshly harvested seeds of Karonda were soaked in cow urine solutions of different concentrations (25%, 50% and 100%) for a period of 24 hours and sown in polybags under the poly house.

Soil Inoculation

*Pseudomonas* solution was measured with measuring cylinder and applied to soil @ 6 ml per kg.

Filling of polybags

Polybags of 200 gauge thickness having length of 15 cm and diameter of 10 cm were used. Bags were filled as per treatment i.e. with and without *Pseudomonas*.

Seed sowing

Treated seeds of Karonda were sown in polythene bags of 15 X 10 cm size filled with and without *Pseudomonas* treated soil.

One seed per poly bag was sown at 2-2.5 cm depth.

Seed vigor parameters

Initiation of germination

The days taken to initiate germination in each treatment after sowing were observed and recorded.

50% germination

The days taken for 50 per cent germination in each treatment after sowing were observed and recorded.

Germination at 60 days after sowing (%)

The germination in each treatment was recorded at 60 days after sowing. Number of germinated seeds were counted and expressed as germination percentage.

Coefficient of germination

The germination in each treatment was recorded after sowing. Number of seedlings was counted and expressed the coefficient of germination.

Speed of germination

When seeds started germinating their number was counted daily till maximum germination was obtained.
**Growth parameters**

**Seedling height**

Five plants were randomly selected for recording the shoot length of seedlings. They were measured by using centimeter scale and recorded at 60, 90 and 120 DAS.

**Stem girth (mm)**

The diameter of shoot above the root collar region was recorded using Vernier caliper at 60, 90 and 120 days after sowing for five randomly selected tagged plants in each treatment and the average of girth was computed.

**Number of leaves per seedling**

The total number of leaves was counted from five randomly selected plants in each treatment at 60, 90 and 120 days after sowing and the average of number of leaves was computed.

**Length of seedling (cm)**

The height of seedling is measured from root tip to the shoot tip and expressed in centimeter at 120 days after sowing.

**Number of roots per seedling**

The number of roots per seedling, primary, secondary and tertiary and rootlets were counted.

**Fresh weight of seedling (g)**

The plants were carefully washed to remove the soil adhering to their roots and shoots. The weight was taken with the help of electronic balance and average value was computed.

**Dry weight of seedling (g)**

For dry weight plants were chopped and oven dried at 60 ± 20°C temperature till a constant weight. The weight was taken with the help of electronic balance and average value was computed.

**Seedling vigor index - I (cm)**

It was calculated by adding the values of root length and shoot length which was randomly selected and multiplying with their corresponding germination per cent and the values were recorded.

Seedling vigor index I = germination per cent x [root length (cm) + shoot length (cm)]

**Seedling vigor index - II (g)**

It was calculated by multiplying dry weight of seedlings with their corresponding germination per cent.

Seedling vigor index II = dry weight of seedlings (g) x germination per cent

**Survival percentage of seedlings**

The survival percentage of each treatment was recorded at 120 days after seed sowing.

**Physiological parameters**

**Leaf Area Index**

LAI expresses the ratio of leaf surface considerably to the ground area occupied by the plant or a crop stand worked out as per specification of Gardner et al., (1985)

**Leaf Area Duration (LAD) (cm².days)**

Leaf Area Duration expresses the magnitude and persistence of leaf area of leafiness during...
the period of crop growth. It reflects the extent of period of crop growth. It reflects the extent of seasonal integral of light interaction and corrected with yield.

**Light Transmission Ratio (LTR) (%)**

It was given by Golingai and Mabbayad in 1969.

**Energy Interception (EI) (cal cm\(^{-2}\) min\(^{-1}\))**

It can be calculated by lux meter. Its values were converted in terms of energy as per constants. The values were recorded at 90 and 120 DAS and then the mean value was worked out.

71 K Lux =1 calorie per cm\(^2\) per min

\[ EI = \text{total incident energy} - \text{transmitted energy} \]

**Results and Discussion**

**Seed vigor parameters**

**Initiation of germination**

The results on the effect of cow urine and Plant Growth Promoting Rhizobacteria on initiation of germination of Karonda seeds is shown in Table 1. Earliest germination (5.33 days) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. Soaking of tamarind seeds in 10 per cent cow urine or cow dung solution for 24 h increased the germination and vigor index as compared to that of untreated seeds (Shankaranarayanan et al., 1994). Inoculation of seeds of *Phragmites australis* with *Pseudomonas asplenii* improved germination (Bashan et al., 2008).

**50% germination**

The results on the effect of cow urine and Plant Growth Promoting Rhizobacteria on initiation of germination of Karonda seeds is shown in Table 1. Minimum days taken to 50% germination (14.50) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. 10 per cent cow urine 24 hours soaking treatment is effective for breaking dormancy and increasing germination and seedling growth tamarind (Parameswari et al., 2001).

Plant Growth Promoting Rhizobacteria significantly enhanced seed germination and seedling vigor of maize (Gholami et al., 2009).

**Germination at 60 DAS (%)**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on germination percentage at 60 days after sowing of Karonda seedlings is represented in Table 1.

The highest percentage of germination (79.63%) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. Khirni seeds soaked in 20 per cent cow urine was effective to get early and maximum germination per cent (Shinde et al., 2015).

**Coefficient of germination**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on coefficient of germination of Karonda seedlings is represented in Table 1.

The highest coefficient of germination (6.10) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. Sweet flag seed treated with mixture of 50g of powdered rhizomes 2.5 I of water 1 I of cow urine showed germination high, seedling growth fast and also increasing tolerance against insect pest (Vijayalakshmi et al., 1998).
Speed of germination

The results on the effect of cow urine and Plant Growth Promoting Rhizobacteria on speed of germination of Karonda seeds is shown in Table 1. The highest speed of germination (9.15) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

Growth Parameters

Seedling height (cm)

The results on the effect of cow urine and Plant Growth Promoting Rhizobacteria on seedling height at various intervals (60, 90 and 120 DAS) is represented in Table 2. The maximum seedling height (8.75, 11.20 and 12.60 cm) was obtained in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. There is increase P uptake, biomass and yield of gram due to seed treatment with Pseudomonas striata (Krishna Murthy et al., 1991)

Stem girth (mm)

The results on the effect of cow urine and Plant Growth Promoting Rhizobacteria on stem girth at various intervals (60, 90 and 120 DAS) is represented in Table 2. The maximum stem girth (1.98, 2.36 and 2.64 mm) was obtained in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

Number of leaves per seedling

The results on the effect of cow urine and Plant Growth Promoting Rhizobacteria on number of leaves per seedling at various intervals (60, 90 and 120 DAS) is represented in Table 2. The maximum number of leaves (29.3, 72.2 and 89.4) were obtained in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

Fresh weight of seedling (g)

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on fresh weight of seedlings at 120 days after sowing of Karonda seeds is represented in Table 2. The maximum fresh weight (3.41 g) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

Dry weight of seedlings (g)

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on dry weight of seedlings at 120 days after sowing of Karonda seeds is represented in Table 2.
**Table 1** Effect of cow urine and plant growth promoting rhizobacteria (PGPR) on seed vigor and growth parameters

| Treatments                        | Initiation of germination (days) | 50% germination (days) | Germination 60 DAS (%) | Speed of germination | Coefficient of germination | Seedling height (cm) |
|-----------------------------------|----------------------------------|------------------------|------------------------|----------------------|---------------------------|----------------------|
|                                   |                                  |                        |                        |                      |                           | 60 DAS | 90 DAS | 120 DAS |
| T0 - control (tap water)          | 9.83                             | 24.50                  | 63.10                  | 5.10                 | 3.95                      | 4.91    | 7.35    | 9.36    |
| T1 - 25% cow urine                | 7.83                             | 18.33                  | 70.68                  | 6.51                 | 4.63                      | 6.74    | 9.02    | 11.09   |
| T2 – 50% cow urine                | 8.50                             | 19.00                  | 70.15                  | 6.43                 | 4.60                      | 6.70    | 9.10    | 10.55   |
| T3 – 100% cow urine               | 8.50                             | 19.38                  | 69.56                  | 6.01                 | 4.50                      | 6.42    | 8.74    | 10.51   |
| T4 – *Pseudomonas* 6ml/kg         | 9.00                             | 22.83                  | 65.18                  | 5.61                 | 4.30                      | 6.16    | 8.07    | 9.95    |
| T5 – 25% cow urine + *Pseudomonas* 6ml/kg | 5.33                             | 14.50                  | 79.63                  | 9.15                 | 6.10                      | 8.75    | 11.20   | 12.60   |
| T6 – 50% cow urine + *Pseudomonas* 6ml/kg | 6.33                             | 15.00                  | 78.71                  | 8.80                 | 5.55                      | 7.85    | 10.23   | 11.70   |
| T7 – 100% cow urine + *Pseudomonas* 6ml/kg | 6.33                             | 15.50                  | 72.91                  | 8.23                 | 5.35                      | 7.60    | 9.78    | 11.65   |
| SEm±                              | 0.15                             | 0.20                   | 0.24                   | 0.02                 | 0.019                     | 0.040   | 0.056   | 0.091   |
| CD at 5% level                    | 0.44                             | 0.59                   | 0.71                   | 0.07                 | 0.055                     | 0.115   | 0.162   | 0.263   |
Table 2 Effect of cow urine and plant growth promoting rhizobacteria (PGPR) on growth parameters at different intervals

| Treatments                     | Stem girth (mm) | Number of leaves per seedling | Length of seedling at 120 DAS (cm) | Number of roots per seedling at 120 DAS | Fresh weight of shoots at 120 DAS (g) | Dry weight of shoots at 120 DAS (g) | Fresh weight of roots at 120 DAS (g) | Dry weight of roots at 120 DAS (g) |
|--------------------------------|----------------|-------------------------------|-----------------------------------|------------------------------------------|--------------------------------------|------------------------------------|----------------------------------|----------------------------------|
|                                | 60 DAS  | 90 DAS  | 120 DAS | 60 DAS  | 90 DAS  | 120 DAS | 60 DAS  | 90 DAS  | 120 DAS | 60 DAS  | 90 DAS  | 120 DAS | 60 DAS  | 90 DAS  | 120 DAS | 60 DAS  | 90 DAS  | 120 DAS |
| T0 – control (tap water)       | 1.42    | 1.86    | 2.13    | 13.13   | 32.06   | 49.36   | 19.42   | 40.23   | 1.11    | 0.35    | 0.35    | 0.14    | 1.11    | 0.35    | 0.35    | 0.14    |           |
| T1 - 25% cow urine             | 1.78    | 2.07    | 2.28    | 21.73   | 46.43   | 57.50   | 21.92   | 70.41   | 1.99    | 0.68    | 0.50    | 0.20    | 1.99    | 0.68    | 0.50    | 0.20    |           |
| T2 – 50% cow urine             | 1.74    | 2.00    | 2.25    | 21.03   | 45.30   | 57.90   | 20.90   | 63.63   | 1.87    | 0.65    | 0.47    | 0.18    | 1.87    | 0.65    | 0.47    | 0.18    |           |
| T3 – 100% cow urine            | 1.73    | 1.99    | 2.24    | 20.70   | 44.60   | 56.50   | 20.44   | 55.76   | 1.76    | 0.59    | 0.46    | 0.18    | 1.76    | 0.59    | 0.46    | 0.18    |           |
| T4 – pseudomonas 6ml/kg        | 1.65    | 1.92    | 2.15    | 19.26   | 40.73   | 50.93   | 19.97   | 46.80   | 1.32    | 0.43    | 0.38    | 0.15    | 1.32    | 0.43    | 0.38    | 0.15    |           |
| T5 – 25% cow urine + pseudomonas 6ml/kg | 1.98 | 2.36 | 2.64 | 29.33 | 72.20 | 89.43 | 28.39 | 118.38 | 2.63 | 1.09 | 0.78 | 0.34 | 2.63 | 1.09 | 0.78 | 0.34 |           |
| T6 – 50% cow urine + pseudomonas 6ml/kg | 1.91 | 2.28 | 2.50 | 28.10 | 68.70 | 78.00 | 27.23 | 115.23 | 2.50 | 0.86 | 0.70 | 0.26 | 2.50 | 0.86 | 0.70 | 0.26 |           |
| T7 – 100% cow urine + pseudomonas 6ml/kg | 1.88 | 2.24 | 2.44 | 24.03 | 62.60 | 73.76 | 25.67 | 101.83 | 2.40 | 0.82 | 0.59 | 0.24 | 2.40 | 0.82 | 0.59 | 0.24 |           |
|SEM±                          | 0.007   | 0.003   | 0.006   | 0.291   | 0.755   | 0.458   | 0.17    | 1.18    | 0.03    | 0.04    | 0.001   | 0.003  | 0.03    | 0.04    | 0.001   | 0.003  |
| CD at 5% level               | 0.021   | 0.009   | 0.017   | 0.841   | 2.182   | 1.324   | 0.50    | 3.41    | 0.11    | 0.11    | 0.002   | 0.008  | 0.11    | 0.11    | 0.002   | 0.008  |
### Table 3: Effect of cow urine and plant growth promoting rhizobacteria (PGPR) on growth and physiological parameters at 120 days after sowing

| Treatments                              | Seedling vigor Index I (cm) | Seedling vigor index II (g) | Survival of seedlings (%) | Leaf Area Index | Leaf Area Duration (cm².days) | Light Transmission Ratio (%) | Energy Interception (cal cm⁻² min⁻¹) |
|-----------------------------------------|----------------------------|-----------------------------|---------------------------|-----------------|-------------------------------|-----------------------------|-------------------------------------|
| T0 - control (tap water)                | 1207.00                    | 31.71                       | 64.28                     | 1.13            | 1611.55                       | 81.39                       | 0.12                                |
| T1 - 25% cow urine                      | 1475.09                    | 60.61                       | 68.85                     | 1.89            | 3036.18                       | 48.95                       | 0.18                                |
| T2 – 50% cow urine                      | 1346.53                    | 53.59                       | 68.15                     | 1.87            | 2974.03                       | 59.58                       | 0.17                                |
| T3 – 100% cow urine                     | 1321.56                    | 51.31                       | 67.15                     | 1.79            | 2814.00                       | 61.02                       | 0.15                                |
| T4 – *Pseudomonas* 6ml/kg               | 1250.10                    | 38.58                       | 65.28                     | 1.43            | 2222.03                       | 70.91                       | 0.12                                |
| T5 – 25% cow urine + *Pseudomonas* 6ml/kg | 2269.79                   | 106.85                      | 81.46                     | 4.09            | 6777.98                       | 34.52                       | 0.44                                |
| T6 – 50% cow urine + *Pseudomonas* 6ml/kg | 2121.61                   | 88.14                       | 80.18                     | 2.95            | 4912.13                       | 38.66                       | 0.36                                |
| T7 – 100% cow urine + *Pseudomonas* 6ml/kg | 1866.81                   | 77.54                       | 74.13                     | 2.62            | 4327.35                       | 39.91                       | 0.26                                |
| SEm±                                    | 21.71                      | 0.88                        | 0.37                      | 0.02            | 41.50                         | 0.40                        | 0.003                               |
| CD at 5% level                          | 62.73                      | 2.54                        | 1.07                      | 0.06            | 119.91                        | 1.18                        | 0.009                               |
The maximum dry weight (1.43 g) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

**Seedling vigor index – I (cm)**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on seedling vigor index – I at 120 days after sowing of Karonda seeds is represented in Table 3. The maximum seedling vigor index-I (2269.7cm) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

**Seedling vigor index – II (g)**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on seedling vigor index – II at 120 days after sowing of Karonda seeds is represented in Table 3. The maximum seedling vigor index-II (106.8 g) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

**Survival percentage of seedlings**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on survival percentage of seedlings at 120 days after sowing of Karonda seeds is represented in Table 3. The maximum survival (81.46%) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg.

**Physiological parameters**

**Leaf Area Index**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on leaf area index at 120 days after sowing of Karonda seedlings is represented in Table 3. The maximum LAI (4.09) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. *Albizia lebbeck* seeds soaked with cow urine showed significant increase in shoot length, root length, total leaf area and total dry weight of seedlings (Illango et al., 1999).

**Leaf Area Duration (cm².days)**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on leaf area duration at 120 days after sowing of Karonda seedlings is represented in Table 3. The maximum LAD (677798 cm².days) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. Pseudomonas ACC-deaminase-producing strains can improve the growth, physiology, and quality of mung beans under salt-affected environments (Ahmad 2013)

**Light transmission ratio (%)**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on light transmission ratio at 120 days after sowing of Karonda seedlings is represented in Table 3. The lowest LTR (34.52%) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. Plant Growth Promoting Rhizobacteria enhanced yield and growth of wheat (Ozturk et al., 2003)

**Energy interception (cal cm² min⁻¹)**

The results regarding the effect of cow urine and Plant Growth Promoting Rhizobacteria on energy interception at 120 days after sowing of Karonda seedlings is represented in Table 3. The highest EI (0.44 cal cm⁻² min⁻¹) was recorded in seeds treated with 25% cow urine and sown in soil inoculated with pseudomonas 6ml/kg. Increase in seed yield and N uptake by crop when pigeon pea seeds were
inoculated with *Pseudomonas striata* before sowing (Pawar and Pawar, 1998).

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