Glutathione S-Transferase M1 and T1 Gene Deletions and Susceptibility to Acute Lymphoblastic Leukemia (ALL) in Adults

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ABSTRACT

Objective: Biotransformation of xenobiotics is critical for their metabolism and removal from the body which is carried out by xenobiotic metabolizing enzymes. Individuals carrying variants of genes that encode these enzymes have an altered ability to metabolize xenobiotics which may lead to an increased risk of acute lymphoblastic leukemia. The current study aimed to investigate the impact of GSTM1 and GSTT1 gene deletions in causing predisposition to adult ALL.

Methods: The current case-control study involved 62 adult ALL patients and 62 age and gender matched healthy controls. Whole blood samples processed with standard phenol chloroform protocol for DNA isolation were genotyped using multiplex PCR approach for simultaneous identification of GSTM1 and GSTT1 deletions. The genotype frequency obtained for patients was compared to controls using odds ratio and chi-square.

Results: The null genotype frequency of GSTM1 and GSTT1 in a group of adult ALL patients from Pakistan were 47% and 11% respectively. Deletion of GSTM1 and GSTT1 did not show statistically significant association with adult ALL (p=0.86 and p=0.35 respectively). The combined GSTM1/GSTT1 deletion was observed in 2% patients and was not significantly associated with ALL in adults (p=0.85).

Conclusions: The results reveal that homozygous null polymorphism of GSTM1 and GSTT1 genes does not influence ALL susceptibility among adult patients. Cancer susceptibility associated with GST polymorphism varies with ethnic and geographic differences. Therefore, further investigation on different populations is needed to understand the role of these genetic variations in modifying adult ALL risk.

KEYWORDS: Acute Lymphoblastic Leukemia, ALL, GSTM1, GSTT1, Genetic, Polymorphism.

INTRODUCTION

Acute lymphoblastic leukemia (ALL) is a blood malignancy distinguished by an excessive buildup of lymphoid progenitor cells in blood and bone marrow. ALL constitutes one fourth of all cancer cases occurring in childhood making it the most common pediatric cancer.1 ALL among adults accounts for less than 1% of total cancer cases.2 According to American Cancer Society, 5960 new cases including both children and adults have been estimated to be diagnosed with ALL with over 1470 deaths within the United States in 2018.3 While exceptional progress has been made in pediatric ALL outcomes over last few decades with long term survival rates exceeding 80% in recent reports,
outcomes for adult ALL remain considerably poor.4,5

The development of ALL includes both genetic and environmental factors with DNA damage in hematopoietic precursor cells being a crucial step.6 Reactive oxygen species (ROS) generated by environmental toxins and chemical carcinogens result in DNA damage.7 Being substrates of carcinogen metabolizing enzymes, the xenobiotics influence their carcinogenic effect depending on a person’s ability to activate or inactivate them by conjugation and detoxification of these compounds.8 Variation in genes that encode carcinogen metabolizing enzymes may therefore explain the differences between the individual’s capacity to metabolize different chemical carcinogens and have thus received a considerable level of attention with respect to cancer development.

The glutathione S-transferases (GSTs) constitute an enzyme super family responsible for detoxification of carcinogens. Detoxification represents the second phase of carcinogen metabolism in human body, followed by bioactivation of procarcinogens (phase I). GSTs detoxify reactive intermediates produced during the first phase of metabolism by conjugating soluble glutathione with them.9 Furthermore, GSTs are also involved in protection of DNA against the ROS.10 Two members of these, GSTM1 and GSTT1, exhibit null polymorphism indicating homozygous deletion of the genes.11 This results in absence of the enzymatic activity. These null genetic variants of GSTs have been reported to modulate susceptibility to various cancers. The role of GST deletion has also been studied in the development of hematologic malignancies.12,13 However, few data has been reported so far for ALL in adults. Therefore, in the present study, the distribution of null genetic variants of GSTM1 and GSTT1 among adult ALL patients in comparison to controls was studied and their association with the occurrence of adult ALL was examined.

METHODS

The current case control study was assessed and ethically approved by the Institutional Ethical Committee of the Karachi Institute of Biotechnology and Genetic Engineering (KIBGE), University of Karachi. The study comprised 62 adult ALL cases and an equal number of controls. Patients were recruited from Department of Oncology, Jinnah Postgraduate Medical Centre (JPMC) Karachi, Pakistan after clinical diagnosis. Questionnaire was filled to collect personal information of patients while medical records were checked to extract their clinical history. Patients with history of other cancers were excluded. Healthy individuals from the population qualified as controls and were matched to cases on age and gender. All participants were provided with an information sheet explaining study objective along with potential risks and benefits associated with their participation prior to their enrolment in this study. A written informed consent declaring their voluntary participation was obtained from them.

Sample Collection: Five ml of whole blood was withdrawn from patients as well as controls in sterile acid citrate dextrose (ACD) vacutainer for genotype analysis. Collected blood samples were stored at -20°C till further use. They were treated with standard phenol chloroform procedure for genomic DNA isolation.14 DNA quality was evaluated by gel electrophoresis and the concentration and purity was analyzed by spectrophotometer respectively.

Genotype Analysis: Multiplex PCR assay was used to screen out homozygous deletions in GSTM1 and GSTT1 genes.15 The sense and antisense primers used in the reaction were 5′ GAACCTCCCTGAAAGCTAAGC 3′ and 5′ GTTGGGCTCAAATATACGGTG 3′ for GSTM1, 5′ TTCTTACTGGTCTTACATCTC 3′ and 5′ TCACCGGATCATGGCCAGCA 3′ for GSTT1 which resulted in 219 bp and 459 bp fragments respectively. β-globin gene was indicative of successful PCR. Primers used for its amplification were 5′ CAACCTCATCCACGGTCACC 3′ and 5′ GAAGGCCAACAGGACTAC 3′ that gave a 268 bp amplified product. Multiplex PCR was carried out with 200 ng extracted DNA in total reaction volume of 20 µl. The other PCR components included 1x (NH4)2SO4 buffer, 1.5 mM MgCl₂, 200 µM dNTPs mix, 1.5 units of Taq DNA polymerase, 0.6 µM of sense and antisense primer for GSTM1, 0.3 µM of each primer for GSTT1 and 0.4 µM of each primer for β globin. PCR cycle conditions included an initial melting step at 94°C for four minutes, 35 PCR cycles, each including 30 s denaturation at 94°C, 35 s annealing at 58°C and 30 s elongation at 72°C, followed by a seven minutes long final elongation step at 72°C. Eight microliters of the PCR products were loaded on 2.5% agarose gel and electrophoresis was carried out for 40 min at 100 V. The ethidium bromide stained gel containing amplified fragments was visualized under gel documentation system.

Statistical Analysis: Odds ratio (OR) with 95% confidence interval (CI) was calculated to measure
Table-I: Frequency distribution and odds ratio (95% CI) of GSTM1 and GSTT1 deletion among ALL patients and controls.

| Genes  | Genotype   | Cases (%) | Controls (%) | OR          | 95% CI       |
|--------|------------|-----------|--------------|-------------|--------------|
| Total (n=62) |            |           |              |             |              |
| GSTM1  | Present    | 33 (53.2) | 34 (54.8)    | Ref         |              |
|        | Null       | 29 (46.8) | 28 (45.2)    | 1.0671      | 0.5265 to 2.1628 |
| GSTT1  | Present    | 55 (88.7) | 58 (93.6)    | Ref         |              |
|        | Null       | 7 (11.3)  | 4 (6.4)      | 1.8455      | 0.5117 to 6.6553  |

Fig.1: Agarose gel electrophoresis of PCR products for the detection of null polymorphism of GSTM1 and GSTT1

the risk conferred by null allele of GSTM1 and GSTT1 for the disease. Chi-square test was performed to test association of homozygous deletion in GSTM1 and GSTT1 genes with development of adult ALL using VassarStats, a freely available online tool, under the null hypothesis of “no association”.16

RESULTS

The present study involved 62 adult ALL patients ranging from 15 to 55 years of age. The mean age (±SD) of patients at diagnosis was 24.7 ± 1.1 years. Patients under 25 years of age constituted 53% of the total number of cases while 89% of patients were less than 35 years. Patients included both genders but male cases predominated female cases with a ratio of 2.4:1.

Multiplex PCR Analysis: Multiplex PCR assay for co-identification of GSTM1 and GSTT1 null polymorphism resulted in banding pattern shown in Fig. 1. Absence of a band indicated absence of the respective gene. GSTM1 deletion resulted in the absence of 219bp amplified product while absence of 459bp fragment meant homozygous deletion of GSTT1 gene. Amplification of β-globin gene resulting in 268bp fragment confirmed the success of PCR when both GST genes were deleted.

Genotype Distribution: Frequency distribution of homozygous null polymorphism in GSTM1 and GSTT1 genes among patients and controls and the odds of their association with ALL are shown in Table-I. The overall frequency of GSTM1 and GSTT1 deletion among study participants (n=124) was 46% and 8.7% respectively. GSTM1 was deleted in 47% adult ALL patients (n=62) and in 45% controls (n=62). The null genotype of GSTT1 was 11% for cases and 6% for healthy individuals. The Pearson chi-square test revealed neither GSTM1 deletion nor GSTT1 null genotype was significantly associated with adult ALL (χ²=0.03, p=0.86 and χ²=0.9, p=0.35 respectively). GSTM1 and GSTT1 were simultaneously deleted in 1.6% cases as compared to 3.2% controls (Table-II). This difference in genotype frequencies, however, could not attain statistical significance (χ²=0.03, p=0.85).

DISCUSSION

Glutathione S-transferases take part in detoxification of different carcinogens, environmental toxins and chemotherapeutic drugs. The null genotypes of GSTM1 and GSTT1 resulting from homozygous deletion of the respective genes lead to the lack of active enzymes. This loss of enzyme...
activity affects metabolism of several carcinogens and may therefore influence an individual’s risk of cancer development. GST genetic variants are also good candidates for association studies in leukemia as they have a potential to alter metabolism of leukemogens and to cause lack of protection against ROS leading to cellular DNA damage.17

The current study evaluating the impact of GSTM1 and GSTT1 deletions as predisposing factor of adult ALL revealed that the genotype frequencies of both genes were not significantly higher in ALL patients as compared to healthy controls (47% vs. 45% for GSTM1 and 11% vs. 6% for GSTT1). The use of healthy controls over hospital based controls was intentionally preferred to reduce possible bias in results due to the risk conferred by deletion polymorphism of GSTM1 and GSTT1 to non-cancer diseases.

Distribution of GST alleles is not uniform across human population and ethnic as well as geographic variations have been observed. In Asians, the frequency of GSTM1 deletion has been reported to vary between 42% and 54% (n=1511) while 35% to 52% for GSTT1 null polymorphism (n=575).18 However, these genotype frequencies may not be a true representation of Asian population as the study included data from only 3 countries: Japan, Korea and Singapore. Studies from neighboring countries have reported frequency of GSTM1 and GSTT1 deletion to be 45% and 21% in Iranian (n=229) while 33% and 18% in North Indian (n=198) population respectively.19,20

The current study reports no association of either GSTM1 null polymorphism (OR, 1.07; 95% CI, 0.53-2.16) or GSTT1 gene deletion (OR, 1.85; 95% CI, 0.51-6.66) with adult ALL but no association of GSTM1 null genotype. The participants of this study were Caucasians.21 Results from another study involving 36 adult ALL patients from Turkey were similar for GSTM1 null genotype but were contradictory for GSTT1 as they showed negative association of GSTT1 deletion with ALL (OR, 0.2; 95% CI, 0.05-0.9).22 However, the sample size of these studies was not large enough to define the frequency of these alleles among ALL patients in population-specific manner. A study with a larger sample size, 192 ALL patients, from India revealed no association of either GSTM1 or GSTT1 null genotype with ALL. The results of this study are comparable to our findings, however, the patients included in the study were all under 25 years of age.23 A meta-analysis comprising 23 studies from different Asian countries illustrated significant association of GST null polymorphisms with acute leukemia risk in children while no association with adults.24 This difference in results depict geographic variations in GST null allele distribution and its association with ALL.

**CONCLUSIONS**

Results from the current study suggest no statistically significant association of GSTM1 null, GSTT1 null, and GSTM1/GSTT1 double null genotypes with ALL in adult patients from Pakistan. ALL risk may not only be affected by genetic factors but also with environmental factors. Therefore, studies with a better study design and a larger sample size including environmental exposure data in addition to genetic variants of xenobiotic metabolism are required to provide more insight into the etiology of adult ALL.

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16. Authors’ Contribution:
AZ, SZ & MI: Concept and study design.
AZ: Experimental work, statistical analysis and manuscript writing.
SZ & MI: Review of manuscript.
AA & SZ: Final approval of manuscript.

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