Histological observation of the effect of aqueous extract of *sorghum bicolor* leaf sheath on paracetamol-induced liver damage in rats

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**Abstract**

Introduction: Acetaminophen is a severe hepatotoxic drug. This study was undertaken to examine the protective and ameliorative effects of aqueous extract of Sorghum Bicolor stem bark on acetaminophen induced hepatotoxicity in male albino rats. Materials and Methods: Thirty five adult wistar rats weighing 150 to 200g were randomized into seven groups of five animals each. Group A (control) received 200mg/kg body weight (bw) of distilled water for 7 days, Group B received 200mg/kg bw aqueous extract of *Sorghum bicolor* stem bark for 7 days, Group C received 300mg/kg bw paracetamol for 1 day, Group D received 300mg/kg bw paracetamol for 1 day plus aqueous extract of *Sorghum bicolor* stem bark (200mg/kg bw) for 7 days, Group E received 300mg/kg bw paracetamol for 1 day plus silymarin (100mg/kg bw) for 7 days, Group F received 200mg/kg bw aqueous extract of *Sorghum bicolor* stem bark for 7 days plus paracetamol (300mg/kg bw) for 1 day and Group G received silymarin (100mg/kg bw) and paracetamol (300mg/kg bw). All solutions were administered orally. At the end of the administration, animals were sacrificed under chloroform anesthesia, blood samples collected via cardiac puncture. Paraffin sections of liver were stained for histology using Hematoxylin and Eosin, and for histochemistry using Masson’s trichome, Gordon & Sweets, and Periodic Acid Schiff’s staining techniques. Results: Results from this study showed that there was congestion of hepatic portal triad, dilated sinusoids, and there was necrosis of hepatocytes nuclei in group C, compare with group A and group B, which showed normal liver architecture and hepatocytes, group D and G showed mild portal triad congestion, mild dilated sinusoids, presence of inflammatory cells, hepatocytes showed dysplastic changes, Group E an F showed mild portal triad congestion, mild dilated sinusoids, reduced inflammatory cells, and dysplastic changes of the hepatocytes. Conclusion: The results of this study indicated that 300mg/kg bw of paracetamol was hepatotoxic, and *Sorghum bicolor* stem bark extract had ameliorative and protective effect on the Paracetamol-induced liver damage.

**Keywords:** Sorghum bicolor, Acetaminophen, Hepatotoxicity.

**INTRODUCTION**

Exposure to drugs and chemicals often induce toxicity to living organisms. Factors determining the toxicity include the pharmacokinetics of the compound, the metabolic fate of the compound and the target organ ability to respond to the toxic insult. During the last decades considerable attention has been focused on the involvement of oxygen free radical (OFR) in various diseases. Active oxygen molecules such as superoxide and hydroxyl radicals have been demonstrated to play important role in the inflammation process produced by ethanol, carbon tetrachloride or paracetamol (Rippka et. al. 1979). Despite the presence of strong antioxidant defense mechanism to counteract the OFR and to minimize the plausible oxidative damage, OFR dependent damage to DNA and other biomolecules accumulate during the life time of organism. Liver is the
most important organ concerned with the biochemical activities in the human body.

It has great capacity to detoxicate toxic substances and synthesize useful principles. Therefore, damage to the liver inflicted by hepatotoxic agents is of grave consequences. (Subramoniam and Pushpangadan, 1999). There is an ever increasing need of an agent which could protect it from such damage.

Acetaminophen (Paracetamol, N-acetyl-p-aminophenol; APAP) is also known as paracetamol, is widely used as prescription and over the counter analgesic and antipyretic agent (Trumper et. al. 2004). It is a safe drug when given in therapeutic doses but its overdose is fairly common since it has narrow therapeutic index. Acute overdoses of paracetamol can cause potentially fatal liver damage and, in rare individuals, a normal dose can do the same; Paracetamol toxicity is the foremost cause of acute liver failure. (Walker et. al., 1981). In view of severe undesirable side effects of synthetic agents, there is growing focus to follow systematic research methodology and to evaluate scientific basis for the natural compounds which are claimed to possess hepatoprotective activity, Sorghum bicolor is an indigenous African cereal and traditional plant that belongs to the grass family, the Gramineae. It is a plant that has been used as an anti-toxicant, antiabortive, cyanogenetic, demulcent (providing relief for cough), diuretics (drugs that help in discharge of urine) and emollient (skin soften and smoothen substance). The parts that are commonly used for herbal remedies include leaves, whole plant or grains. The root is used for the treatment of malaria in southern Rhodesia; the seeds for the treatment of breast diseases and diarrhoea; the stem for tubercular swellings.

In India, the plant is considered antihelminthic and insecticidal (Duke and Wain, 1981). Recently focus has been on the leaf sheath of sorghum bicolor being used as herbal remedy for anaemia and having a boosting effect on blood concentration hematinic potentials (Ogwumike, 2002). Aqueous extracts of Sorghum bicolor leaf – sheaths has been shown to contain phenolic compounds and to possess hemopoietic and antioxidant properties in the rat (Akande et al. 2010). The presence of antioxidants in sorghum is believed to have reduced the risk of cancer, diabetes, heart disease, nerve disorders among others. Its health potentials, experts say reside in its rich nutrients essential for healthy living (Sofowora, 2008). This study investigated the ameliorative and protective potential of Sorghum bicolor stem bark on PCM-induced liver damage in wistar rats.

MATERIALS AND METHODS

Drugs and chemicals

Silymarin (Micro Labs Limited, 92, Sipcot Hosur – 635 126, India) Batch number SYFH0005) is marketed as a hepatotonic and it basically contains phospholipids with vitamins. Paracetamol powder was purchased from (M and B pharmaceutical Nigeria Plc.). All chemicals used were of analytic grade. Assay kits for Biochemical parameters carried out were purchased from Randox Laboratories Limited, UK.

Drug preparations

Paracetamol

Each capsule contains 500 mg of Paracetamol. One capsule was dissolved in 1 ml of distilled water. Then the equivalent of 300mg/kg of the drug in the solution will be administered to the animal

Silymarin

Silymarin (100mg) will be weighed also using sensitive weighing balance and dissolved in 5mL of water and stirred thoroughly (Jibrin. et al., 2012)

Extraction of sorghum bicolor

Dry stem bark leave sheath was obtained from herb sellers at Ile Ife, Osun State, Nigeria. The plant specimen was identified and authenticated by a Taxonomist at the Department of Botany, Obafemi Awolowo University, Ile-Ife and a voucher number was given. The leave sheath was air dried at room temperature for weeks, weighed every three days to ascertain the dryness of the leaves. The air dried leave sheath weighing 750g was grounded into fine powder in an electric blender and the powdered. The powder weighing 500g was extracted in aqueous solution (50%) by percolation for 24hrs. The mixture was filtered and the filtrate was allowed to evaporate at 40ºC using a vacuum Rotary evaporator. The wet residue was freeze- dried using a vacuum freeze drier and stored in a desiccator. An aliquot portion of the crude extract residue was dissolved in distilled water for use on each day of the experiment.

Animal care and management

Thirty- five adult Male wistar rats weighing 150g - 200 g that were used in this study were obtained from the Animal House of the College of Health Sciences, Obafemi Awolowo University, Ile-Ife. The male rats were housed in separate cages. The animals were kept under normal environmental conditions with a 12 hrs light/dark cycle and had free access to standard rat pellet diet (Ladokun Feed mill PLC Ibadan, Nigeria) and water ad
Table 1: Dose Regimen

| Animal groups       | Treatments                                      |
|---------------------|-------------------------------------------------|
| Group I (Control)   | distilled water                                 |
| Group II 200mg/ Kg b.w. | AESBLS                                        |
| Group III 300mg/ Kg b.w. | PCM                      |
| Group IV 300mg/ Kg b.w. | PCM + 200 mg /Kg b.w. AESBLS                   |
| Group V 300mg/ Kg b.w. | PCM + 100 mg /Kg b.w. Silymarin               |
| Group VI 200mg/ Kg b.w. | AESBLS + 300 mg /Kg b.w. PCM                   |
| Group VII 100mg/ Kg b.w. | Silymarin + 300 mg /Kg b.w. PCM                |

b.w. (body weight), AESBLS (Aqueous extract of Sorghum bicolor leaf sheath), PCM (paracetamol)

libitum. They were allowed to acclimatize in the laboratory for 2 weeks before the commencement of the study. The experimental procedures adopted in this study were in strict compliance with Experimental Animal Care and Use of Laboratory Animals in Biomedical Research, College of Health Sciences, Obafemi Awolowo University, Ile-Ife.

Histopathological studies

The method of Baker and Silverton was employed for the processing of liver for histopathological studies. (Baker and Silverton 1985)

Photomicrograph

The sections were examined under a LEICA research microscope (LEICA DM750, Switzerland) interfaced with digital camera (LEICA ICC50). Digital photomicrographs of stained sections of the kidney was taken at various magnifications.

RESULTS AND DISCUSSION

The use of plants for remedies has long been in existence and is among the most attractive sources for developing drugs. Any part of plant can be considered as herbs including leaves, leaf sheath and bark. Research has shown that these practices have produced results of proven efficacies comparable to conventional modern medicine (Nwogu, 2008). This study investigated the effect of *sorghum bicolor* on histoarchitecture of the liver in rats with PCM-induced liver damage.

Histological and histochemical results of this study clearly support earlier reports that PCM has hepatotoxic effect on the liver cells when taken at high doses (Ita et al., 2009), table 1 show Dose Regimen. In group C (paracetamol only) as shown in figures 1C, and 2C shows Portal triad congestion, hepatic sinuses are dilated, the endothelia cells lining the vascular channels shows dysplastic changes and their was necrotic hepatocytes, also the presence of mild fibrosis (demonstration of young collagen fibres). Stroma with young collagen fibre in paracetamol alone treated group when compared with the control group. The photomicrograph of the group A (Normal control) and group B (Extract control) show normal histology of the liver with the hepatocytes, portal triad, the endothelia cell linings, the vascular channels showed no dysplastic changes, and hepatic sinuses showing normal liver histoarchitecture. This is an indication that aqueous extract of *Sorghum bicolor* does not have any adverse effects on liver structure and function and this is in conformity with Akande et al, (2009); Olayinka et al (2010) who reported that *Sorghum bicolor* has no toxic effects or undesired properties on the liver. Improvement in the histological lesions, which was characterized by mild central venous congestion and few necrotic hepatocytes with many normal and surviving hepatocytes and few necrotic hepatocytes indicating mild hepatic necrosis when compared to group C rat livers was evident in the groups treated with *Sorghum bicolor* extract and silymarin after paracetamol and before induced hepatotoxicity significantly attenuated the effect of paracetamol on the liver cells and the histological architecture of the liver as shown in figures 1D, 2D, 1E and 2E. A similar finding was reported by Olayinka et al. 2010 that *Sorghum bicolor* was able to ameliorate hepatotoxicity.

Pre-treatment with SB extract and silymarin before paracetamol administration significantly prevented the damage caused by the effect of paracetamol on the liver cells and the histological architecture of the liver as shown in figure 1F, 2F, 1G and 2G based on the histological observations.

Masson trichrome

Masson’s trichrome staining techniques were used to demonstrate collagen fibre in this study. In the result, Collagen fibers stained green with Masson’s trichrome
Figure 1: Representative Photomicrographs of the liver of control (A) and treated, (B, C, D, E, F, G) groups. Arrow represents (hepatocytes – yellow), (sinusoids – white), (central vein – blue), (inflammatory cells – black), (bile ducts – green), (portal vein – red), (hepatic artery – brown) (H and E X400).

Figure 2: Representative Photomicrographs of the liver of control (A) and treated, (B, C, D, E, F, G) groups. Observe the presence of mild fibrosis following PCM alone treatment as shown in figure 4C (MT X 400).

stain, and the nuclei stained black. The collagen fibre in the A normal control group, B (sorghum bicolor only), D (PCM+ SB), E (PCM+ SILY), F (SB+PCM), and G (SILY+PCM) all appear normal with no fibrosis and
Figure 3: Representative photomicrographs of the liver of control (A) and treated, (B, C, D, E, F, G) groups. Observe normal reticular fibre appearance in all the groups (GS X 400)

Figure 4: Representative photomicrographs of the liver of control (A) and treated, (B, C, D, E, F, and G) groups. Observe all groups contains PAS (+) glycogen granules (PAS X 100)

presence of prominent collagen fibre as shown in figure 3A, 3B, 3D,3E, 3F and 3G, except in the negative control group where there was presence of mild fibrosis, this is demonstrated by presence of stroma with young collagen fibre as shown in figure 4C.

Gordon and sweet staining

Gordon and Sweet staining techniques were used to demonstrate reticular fibre in this study. In the result, the reticular fibre stained black and the nuclei stained black
or unstained. The reticular fibre in the control group, B (sorghum bicolor), C (PCM), D (PCM+ SB), E (PCM+ SILY), F (SB+PCM), and G (SILY+PCM) as shown in figure 4A, 4B, 4C, 4D, 4E, 4F, and 4G all appear normal and visible with normal appearance

**Periodic acid schiff’s staining**

Periodic Acid Schiff’s staining techniques were used to demonstrate glycogen in the liver. PAS stains glycogen magenta in this study. In the result, glycogen stained magenta and the nuclei stained blue. The glycogen in the control group, B (sorghum bicolor only), C (PCM), D (PCM+ SB), E (PCM+ SILY), F (SB+PCM), and G (SILY+PCM) all contains PAS (+) glycogen granules in hepatocytes cytoplasm as shown in figures 5A, 5B, 5C, 5D, 5E, 5F and 5G.

**CONCLUSION**

The results of this study indicated that 300mg/kg b w of paracetamol was hepatotoxic, and *Sorghum bicolor* stem bark extract had ameliorative and protective effect on the Paracetamol-induced liver damage.

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