Cytotoxicity and proliferation evaluation on fibroblast after combining calcium hydroxide and ellagic acid

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INTRODUCTION

Calcium hydroxide is a substance that is often used in endodontic therapy because it protects the pulp against thermal stimulation, stimulates the formation of reparative dentin, and has antibacterial properties.[1] The mechanism of the action of calcium hydroxide in tissues encourages the deposition of mineralized tissue, which is an important aspect because calcium hydroxide has a biological compatibility.[2] However, calcium hydroxide has many disadvantages such as having a weak bond to dentin, material reabsorption, high solubility,[3] and has a high pH of approximately 12.5–12.8.[4] Due to calcium hydroxide’s pH, it causes the necrosis of pulp tissue, chronic inflammation, and decreased viability of fibroblasts.[3,5]

One of the natural ingredients that have been investigated for reducing inflammation and cell damage due to the use of calcium hydroxide is ellagic acid. Ellagic acid has a broad spectrum of biological activities including antibacterial,
antioxidant, and anti-inflammatory properties.[6] Ellagic acid is a natural phenol antioxidant that is found in various fruits and vegetables such as extracts of pomegranate, strawberry, and blackberry. Among the natural ingredients used in traditional Chinese medicine, ellagic acid has been used to cure pain of dental caries. Ellagic acid has been shown to be a powerful antioxidant and protects against cell death because it has anti-inflammatory properties,[7] able to induce fibroblast proliferation and accelerate the healing process.[6] Previous research shows that ellagic acid used in rat wounds can accelerate the wound healing process.[8]

One of the requirements for materials used in dentistry is that it is biocompatible, which means that material must meet the requirements for use in tissues that is not harmful to the pulp and soft tissue, does not contain substances that can cause a systemic response when diffused or absorbed into the circulation system, and has no carcinogenic potential. The most frequently used method for toxicity testing is the direct counting method using the trypan blue and MTT assay method. MTT assay has long been used to analyze the cytotoxicity of dental material.[9]

This study combines calcium hydroxide with ellagic acid with ratios of 99:1, 98:2, 97:3, 96:4, and 95:5 to analyze the effect of the combination in decreasing inflammation induced by calcium hydroxide by looking at the cytotoxicity of the combination and proliferation fibroblast cells. The aim of this study is to determine the cytotoxicity and proliferation of fibroblasts after combining calcium hydroxide and ellagic acid.

**MATERIALS AND METHODS**

**Ethically approved**

This research is experimental laboratory research. It has passed ethical clearance with the serial numbers 619/HRECC.FODM/X/2019 and 649/HRECC.FODM/X/2019. The sample is a combination of calcium hydroxide and ellagic acid with five different ratios (99:1, 98:2, 97:3, 96:4, and 95:5). The size of each sample is 16 samples.

**Fibroblast cell culture**

The fibroblast cells that have been used are rat gingival fibroblasts. The four rats that were used were male, over 9 months old and weighed on average 250–300 g. Fibroblast cells were obtained from rat maxillary tissue. Rat gingival fibroblast cells that were cultured until they were homogeneous were inserted into two 96-well microplates with a density of $2 \times 10^5$ cells/ml for 50 µL and incubated for 24 h.

**Calcium hydroxide and ellagic acid preparation**

Calcium hydroxide and ellagic acid were weighed and mixed with sterile water and stirred according to Table 1.

**Toxicity test**

The combination of calcium hydroxide and ellagic acid was put into a 96-well microplate containing 50 µl Eagle’s minimum essential medium (MEM) culture media and 50 µl rat gingival fibroblasts. The division of groups on the microplate consisted of five treatment groups, one media control group, and one cell control group.

The microplate was incubated in a 37°C incubator for 24 and 72 h. The cell growth media were then removed and washed with 200 µl phosphate-buffered saline and repeated twice. 40 µl of Eagle’s MEM media were added to the well. MTT was added to every well that contained 10 µl of culture media, then re-incubated for 4 h at 37°C. For each well, 50 µl of dimethyl sulfoxide was added.

At 24 and 72 h after incubation, the microplate was shaken using a plate shaker for 5 min until the formazan crystals had dissolved. Fibroblast living cells were colored with formazan purplish blue, as the dead cells do not turn the purplish-blue color. The formazan absorbance was read using an ELISA reader with a wavelength of 540 nm. The more concentrated the color, the higher the absorbance value, and the higher the number of living cells. The percentage of living fibroblasts cells was calculated using the following formula:[10]

$$% \text{living cells} = \frac{\text{OD treatment} - \text{OD media}}{\text{OD cell control} - \text{OD media}} \times 100\%$$

- % living cells: percentage of the number of fibroblasts living after testing
- Optical density (OD) treatment: the OD values in each sample after testing
- OD media: the OD values in the control media
- OD cell control: the OD values in the cell control group.

**Table 1: The calcium hydroxide and ellagic acid combinations**

| Group                  | CH powder (g) | EA powder (g) | Sterile water (ml) | Total (g) |
|------------------------|---------------|---------------|--------------------|-----------|
| Pure CH (control)      | 0.2           | -             | 0.2                | 0.2       |
| Combination A          | 0.198         | 0.002         | 0.2                | 0.2       |
| Combination B          | 0.196         | 0.004         | 0.2                | 0.2       |
| Combination C          | 0.194         | 0.006         | 0.2                | 0.2       |
| Combination D          | 0.192         | 0.008         | 0.2                | 0.2       |
| Combination E          | 0.190         | 0.010         | 0.2                | 0.2       |

Combination A: CH and EA ratio 99:1, Combination B: CH and EA ratio 98:2, Combination C: CH and EA ratio 97:3, Combination D: CH and EA ratio 96:4, Combination E: CH and EA ratio 95:5. CH: Calcium hydroxide, EA: Ellagic acid
**Statistical analysis**

The data obtained were tabulated, and the one-way ANOVA test was performed using the SPSS Statistics for Macintosh Version 23.0 (IBM, New York, USA) and posthoc Tukey’s honestly significant difference test to find out the significant differences between the groups with a significance of $P < 0.05$.

**RESULTS**

**Toxicity test**

Twenty-four hours obtained the highest percentage of living cells with combination E (calcium hydroxide and ellagic acid with a ratio of 95:5). Seventy-two hours after treatment, the highest percentage of living cells was combination A (calcium hydroxide and ellagic acid with a ratio of 99:1) [Table 2].

Within 24 h after treatment, the percentage of living cells in combinations D and E (calcium hydroxide and ellagic acid with ratios of 96:4 and 95:5) increased, whereas the other groups decreased. Within 72 h after treatment, only combination A (calcium hydroxide and ellagic acid with a ratio of 99:1) showed an increase in the percentage of living cells [Table 2].

Twenty-four hours after exposure, combination A (calcium hydroxide and ellagic acid with a ratio of 99:1), C (calcium hydroxide and ellagic acid with a ratio of 97:3), and E (calcium hydroxide and ellagic acid with a ratio of 95:5) showed a lower percentage of living cells compared to the control cell ($P = 0.001$, $P = 0.000$, and $P = 0.003$, respectively). Combination E (calcium hydroxide and ellagic acid with a ratio of 95:5) showed a higher percentage of living cells compared to combination D (calcium hydroxide and ellagic acid with a ratio of 96:4) ($P = 0.005$), combination C (calcium hydroxide and ellagic acid with a ratio of 97:3) ($P = 0.000$), combination B (calcium hydroxide and ellagic acid with a ratio of 98:2) ($P = 0.001$), and combination A (calcium hydroxide and ellagic acid with a ratio of 99:1) ($P = 0.000$) [Figure 1].

**Table 2: The fibroblast cells after treatment with the combination of calcium hydroxide and ellagic acid**

| Groups       | Absorbance     | Percentage of living cells (%) |
|--------------|----------------|--------------------------------|
|              | 24 h           | 72 h                          |
| Control cell | 0.278±0.023    | 0.280±0.011                   |
| Combination A| 0.234±0.014    | 0.282±0.042                   |
| Combination B| 0.273±0.017    | 0.234±0.021                   |
| Combination C| 0.218±0.045    | 0.264±0.014                   |
| Combination D| 0.279±0.012    | 0.270±0.012                   |
| Combination E| 0.318±0.043    | 0.273±0.015                   |

Combination A: CH and EA ratio 99:1, Combination B: CH and EA ratio 98:2, Combination C: CH and EA ratio 97:3, Combination D: CH and EA ratio 96:4, Combination E: CH and EA ratio 95:5. CH: Calcium hydroxide, EA: Ellagic acid

At 72 h after exposure, combination B (calcium hydroxide and ellagic acid with a ratio of 98:2) showed a lower percentage of living cells compared to the control ($P = 0.000$). Combination A (calcium hydroxide and ellagic acid with a ratio of 99:1), combination C (calcium hydroxide and ellagic acid with a ratio of 97:3), combination D (calcium hydroxide and ellagic acid with a ratio of 96:4), and combination E (calcium hydroxide and ellagic acid with a ratio of 95:5) were higher than combination B (calcium hydroxide and ellagic acid with a ratio of 98:2) ($P = 0.006$, $P = 0.000$, and $P = 0.000$, respectively) [Figure 2].

**DISCUSSION**

Ellagic acid is a natural flavonoid in fruits and plant extract. It has therapeutic effects such as anti-inflammatory, antibacterial properties, and antioxidant activity.\(^8\) Calcium hydroxide has a high pH, which leads to pulp tissue necrosis and chronic inflammation.\(^9\) Therefore, researchers are trying to investigate the effect of ellagic acid when it is combined with calcium hydroxide to see whether this combination can reduce inflammation and speed up the wound healing process.

This experiment was conducted to investigate the cytotoxicity and proliferative effects of combining calcium hydroxide and ellagic acid, which was given in five different ratios to fibroblast cells after 24 h and 72 h. The parameter used in this study is the inhibitory concentration 50% (IC\(_{50}\)). IC\(_{50}\) represents the concentration that is required to inhibit 50% of the proliferation.\(^{11}\) The percentage of living rat gingival fibroblast indicates the level of toxicity for the sample groups. If the percentage of living fibroblasts increases, it indicates high cell viability and fibroblast cell proliferation. If the material used is not toxic, the dehydrogenase enzyme will be active, and formazan crystals will form.\(^{12}\)

According to the experiment’s results after 24 h, the percentage of living fibroblast cells was more than 50% for...
all combinations. The combination of calcium hydroxide and ellagic acid with a ratio of 95:5 showed a significant increase in cell viability compared to the control group. This might be due to the different ratios of ellagic acid that was used in every treatment group. Therefore, it can be concluded that the combination of calcium hydroxide and ellagic acid is not toxic when it is applied to fibroblast cells. At 72 h after exposure, there was a significant decrease in fibroblast cell viability after combining calcium hydroxide and ellagic acid with a ratio of 98:2. On the other hand, a comparison of fibroblast cell viability at 24 h and 72 h showed that there was a significant fibroblast cell proliferation after combining calcium hydroxide and ellagic acid with a ratio of 99:1 and 97:3.

Calcium hydroxide has a high pH, which can lead to mitochondria dysfunction and increase the cell respiration rate. This will increase the radical superoxide. Under these conditions, superoxide diffuses from cytosol to the mitochondrial membrane and causes the de-energization of mitochondria or cell apoptosis. As ellagic acid is an antioxidant, it can reduce the free radicals that are produced due to calcium hydroxide and prevent oxidative stress damage to fibroblasts. Ellagic acid can reduce the intracellular ROS level on the gingival fibroblast undergoing oxidative stress due to ultraviolet-B radiation.

Moreover, combining calcium hydroxide and ellagic acid can increase cell viability and fibroblast cell proliferation because ellagic acid is an anti-inflammatory agent. Calcium hydroxide activates nuclear factor kappa beta, which promotes inflammation. Ellagic acid can inhibit COX; therefore, it is a strong anti-inflammatory agent. Ellagic acid inhibits COX-2, tumor necrosis factor-α (TNF-α), and interleukin (IL)-6. Furthermore, ellagic acid can also reduce the NF-kB expression and pro-inflammatory cytokines production such as TNF-α and IL-1β. Therefore, ellagic acid can reduce inflammation induced by calcium hydroxide, which can increase fibroblast cell viability. When cell viability increases, it means that combining calcium hydroxide and ellagic acid is not toxic and promotes fibroblast cell proliferation.

CONCLUSION

In summary, fibroblast cells that are given a combination of calcium hydroxide and ellagic acid with ratios of 99:1 and 97:3 show high cell viability and cell proliferation. It can be concluded that combining calcium hydroxide and ellagic acid is not toxic. This research confirmed that the combination has nontoxic properties and able to stimulate the proliferation of dental pulp. This combination may also have ideal characteristics as pulp capping materials.

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Conflicts of interest

There are no conflicts of interest.

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