Bioequivalence Studies and Pharmacokinetic Properties of Atorvastatin 40Mg Tablet in Healthy Bengali Subjects

Abstract
The present study is aimed to investigate the single-dosing pharmacokinetics of two pharmaceutical formulations of atorvastatin 40 mg tablet, test (T) sample (Stacor®) and reference (R) sample (Lipitor®) in Bengali subjects. An open-label, randomized-sequence, two-way two-period crossover study was designed with 24 healthy Bengali male volunteers. Plasma samples collected before and over 0.5-72.0h after a single oral dose per subject. Samples were analyzed by solid phase extraction method and a validated LCMS/MS method. The non-compartmental pharmacokinetic model was used to measure pharmacokinetic parameters of peak plasma concentration (C_{max}), time to reach the peak (T_{max}), elimination half-life (T_{1/2}), elimination constant (k_e), and area under the curve to 72h (AUC_{0-72}) and to infinity (AUC_{∞}). The mean ± SD values of pharmacokinetic parameters for the test and reference products with minor variation were C_{max}: 27.617 ± 2.632ng/mL and 26.166 ± 2.948ng/mL; T_{max}: 2.479 ± 0.634h and 2.396 ± 0.589h; AUC_{0-72}: 256.799 ± 52.318ng-h/mL and 271.562 ± 52.188ng-h/mL; AUC_{0-∞}: 289.172 ± 56.431ng-h/mL and 309.975 ± 59.594ng-h/mL; k_e: 0.055 ± 0.005h⁻¹ and 0.053 ± 0.006h⁻¹; T_{1/2}: 12.591±1.325h and 13.178 ± 1.555h respectively. No significant differences (p > 0.05) for these parameters in the paired t-test and no period or sequence effect was observed in ANOVA analysis. The point estimates (90% CI) of ln-transformed values of C_{max}, AUC_{0-72} and AUC_{0-∞} were 0.97238 - 1.04110, 0.99296 - 1.00641 and 0.99113 - 1.00255 respectively which fell within the acceptable range of bioequivalence.

Keywords: Atorvastatin; Pharmacokinetics; Bioequivalence; LCMS/MS; Bengali subjects

Abbreviations: t_{max}: Time to Reach the Maximum Concentration; C_{max}: Maximum Plasma Concentration; t_{1/2}: Half-Life; AUC_{0-72}: Area Under the Plasma Time Curve Up to Last Quantifiable Time; AUC_{0-∞}: Area Under the Plasma Time Curve Up to Time Infinity; k_e: Elimination Rate Constant; NCDs: Non-Communicable Diseases; DGDA: Directorate General of Drug Administration; TPR: Total Peripheral Resistance

Introduction
There is an emerging importance of bioequivalence and pharmacokinetics analysis of highly prescribed brand drugs to judge their therapeutic effectiveness, toxic city, and quality to ensure therapeutic validity and patient’s safety. Statin classes of lipid-lowering agents are most commonly used treatment intervention for Non-Communicable Diseases (NCDs) like diabetes, obesity, hypertension, and dyslipidemia [1]. Atorvastatin is an FDA approved synthetic lipid reducing agent under statin class. This drug, also known as 2-[(4-fluorophenyl) -3R,8R-dihydroxy-5-(1-methyl ethyl)-3-phenyl-4[phenylamino] carbonyl]-1H-pyrole-1-heptanoic acid (Figure 1), is a second generation 3-hydroxy-3-methylglutaryl-CoA reductase inhibitor [2]. This synthetic HMG-CoA reductase inhibitor significantly lowers total cholesterol, low-density lipoprotein cholesterol, and plasma triglycerides in clinical studies [3]. According to IMS Health, Lipitor, the pioneer brand of atorvastatin is the world’s one of the biggest selling prescription medicines with cumulative sales in excess of $130 billion, but most of the principal brands are now prescribed worldwide as far cheaper generic medicines. In Bangladesh, about 50 atorvastatin brands with different strengths and dosage forms are available and their average sales volume is nearly 1.17 crore taka monthly in the national local market according to Bangladesh Directorate General of Drug Administration (DGDA). This indicates widespread prescribing of this cholesterol-lowering drug in Bangladesh. However, there is no bioequivalence data for any of existing 50 brands of atorvastatin.

Figure 1: Chemical structure of atorvastatin.
Dose-dependent lowering of total cholesterol, LDL cholesterol and triglyceride levels have been noticed after oral administration of atorvastatin to patients with hypercholesterolemia and to patients who have hypertriglyceridemia [4,5]. The usual starting dose is 10mg atorvastatin daily. The dose should be adjusted in accordance with plasma lipid levels and can be increased to up to 80mg daily [6,7]. Pharmacokinetically, atorvastatin has rapid absorption after oral administration due to its high membrane permeability and solubility and maximum plasma concentrations are obtained within 1 to 3 hour [8]. Multiple daily doses of 2.5 to 80mg atorvastatin produce steady-state maximum plasma concentrations (Cmax) of 1.95 to 252µg/L within 2 to 4 hours after administration and area under the plasma concentration-time curve (AUC) values of 25.2 to 1293µg/L [9].

The bioavailability of atorvastatin is only ~14% because of extensive pre-systemic extraction in the gastrointestinal tract and/or liver [7,10]. This drug is ≥ 98% protein-bound in plasma. The mean volume of distribution of atorvastatin is reported to be 381L after intravenous infusion of 5mg which clearly indicates its extensive binding to peripheral tissues [10]. This drug has a mean elimination half-life of about 14 hours, but due to the presence of active metabolites, the half-life of HMG-CoA reductase inhibition becomes 20 to 30 hours. The active acid form of atorvastatin is mainly used for an administration which undergoes extensive first-pass metabolism, predominantly by cytochrome P450 3A4 (CYP3A4) in the liver [7]. Less than 2% of the parent drug and metabolites follow renal excretion [11]. Like other HMG-CoA reductase inhibitors, the gastrointestinal disturbance is the most frequently reported adverse event associated with atorvastatin [12]. It depends on pharmacokinetic variation, subject variation and even metabolic rate from the context of racial and ethnic variation. Consequently, its pharmacokinetics should be well established as country and population specific to predict the toxicity of atorvastatin and to optimize dose, which would likely to ensure patient safety.

In fact, tolerability and bioequivalence of different preparations containing same active ingredient have gained considerable importance over the past few years because of increasing generic substitution and hence cheaper price. However, as the development of generic drugs is mainly based on pharmacological properties of the original brand, one would expect the same quality and almost similar effects like that of the original brand [13]. There is insufficient pharmacokinetics and bioequivalence study on widely used drugs in Bangladesh and atorvastatin is one of them. Similarly, no pharmacokinetics and bioequivalence evaluation of atorvastatin 40mg tablet have been established yet in Bangladesh despite enormous prescribing of atorvastatin from primary to tertiary care facilities. Therefore, the aim of this study was to evaluate pharmacokinetics and bioequivalence of two pharmaceutical products of atorvastatin; Stacor® (T) and Lipitor® (R) tablet in healthy Bengali subjects in order to establish quality, safety and even tolerability of this generic drug.

Materials and Methods

Drugs and reagents

Two commercially available brands of 40mg atorvastatin, Stacor® tablet (T) (Batch No: MH 51, DAR No: 225-291-25 manufactured at 08/2011 by Unimed-Unihealth Pharmaceutical Ltd, Dhaka, Bangladesh) and Lipitor® tablet (R) (Batch No: 0744010 U manufactured at 09/2011 by Pfizer Ireland Pharmaceuticals, Dublin, Ireland) were obtained. Preparation of deionized water was done using Milli-Q system (Continental Water Systems, El Paso, TX, U.S.A.). HPLC grade formic acid and methanol, and analytical grade ethyl acetate were procured from Merck, India.

Subjects

Twenty-four healthy Bengali male volunteers participated in the study (Table 1). No abnormalities were detected in the routine laboratory (hematological and biochemical) and clinical tests that were pre-requisite as inclusion criteria. All the subjects avoided alcohol consumption, drug abuse, concomitant medication use and smoking during the period of the study.

| Parameter                  | Sequence Group          | P-Value |
|----------------------------|-------------------------|---------|
| Age, y                     | RT (n = 12)             | TR (n = 12) | Total (N = 24) |         |
| Height, cm                 | 26.91 (2.34)            | 26.23 (2.04) | 26.46 (3.88) | 0.786   |
| Weight, kg                 | 164.66 (4.30)           | 163.29 (4.54) | 163.38 (4.43) | 0.376   |
| Body Mass Index, kg/m²     | 59.55 (5.43)            | 58.64 (5.35) | 59.25 (5.3)   | 0.645   |
| Male Sex                   | 12                      | 12        | 24           |         |
| Smoke (Yes/No)             | 4/8                     | 5/7       | 9/15         | 0.201   |
| Alcohol (Yes/No)           | 0/12                    | 0/12      | 0/24         | 0.986   |
| Caffeine (Yes/No)          | 6/6                     | 5/7       | 11/13        | 0.658   |

*Values are given as mean (SD) or number.

RT: Reference tablet first then test tablet administered; TR: Test tablet first then reference tablet administered.

P< 0.05 indicated significant difference among two sequence group RT and TR.
Ethical Review

The protocol of the whole research was approved by the clinical review committee, Faculty of Pharmacy, University of Dhaka. Demographic data were collected from every participant who gave written consent after reading the protocol.

Ethical review and consent procedure

Guidelines, as drawn up by the Bangladesh Medical Research Council (BMRC), were followed regarding the treatment of human volunteers in the study. These guidelines were in accordance with the U.S. Code of Federal Regulations (Title 21, Part 56), the Declarations of Helsinki and the Canadian MRC Guidelines. The study was initiated after approval of the protocol, ethical clearance and the informed consent form by BMRC, Bangladesh. The ethical clearance number was BMRC/NERC/DO 2013-2016/1020 date 08.09.2015.

Confidentiality

Every prospective candidate was given a full explanation of the study. Once all the necessary information was provided to them and the physician in charge was confident enough that he understood the implications of participation in the study, the subject was asked to sign the consent form. It was agreed that the protocol information and the results of the study would remain undisclosed to others, without written authorization from the principal investigator.

Study Design

An open-label, randomized-sequence, two-way, two-period crossover study with single dose was designed with 24 healthy Bengali male volunteers. All the volunteers participated in two dosing sessions with a respite of 15 days washout period. As per the randomization code at a fixed time, volunteers received either the test preparations or reference preparations as a single dose in each dosing session on that particular study day. Volunteers were given code numbers by the bio-statistician and were allocated to the treatment in accordance with the randomization code. Neither the personnel in charge of the determination of plasma levels nor the physician and nursing staff in charge of the clinical aspects was informed of the sequence of administration.

Blood Sampling

After overnight fasting for 10 hours, all the volunteers assembled in the medical center, the University of Dhaka at 6.00 a.m. on the study day of each session. Total peripheral resistance (TPR), blood pressure (BP) of every volunteer was recorded and an indwelling intravenous catheter was inserted in the suitable vein having a flow rate of 0.5000mL/min. An anticoagulant, was validated.

LCMS/MS Analysis

Identification and quantification of the atorvastatin by using LCMS/MS (Shimadzu Prominence, Kyoto, Japan) detection were done for determination of the pharmacokinetic parameters. The method, used for analysis of atorvastatin in a plasma containing an anticoagulant, was validated.

Chromatographic analysis

Atorvastatin and internal standard rosuvastatin plasma levels were determined by Shimadzu Prominence (Kyoto, Japan), an LC-MS system which consists of Integrated System Shimadzu Controller SIL-20AC, prominence, autosampler with two pumps Shimadzu LC-20AD, prominence, and liquid chromatography; Pump A Model: LC-20AD, Pump B Model: LC-20AD. Pumping models were of binary flow. Model SIL-20AC was used as the auto-sampler in which rinsing volume, needle stroke, rinsing speed, sampling speed, purge time, rinse dip time and the cooler temperature was fixed at 200µL, 52mm, 35µL/sec, 15.0µL/sec, 25.0min, 0 sec and 15°C respectively. Separation of compounds was carried out by column (C18 50 x 4.5mm, 5µ particle size) (Phenomenox, Gemini, Torrance, USA) eluted with 9:1(v/v) of MeOH: H2O (Containing 1% formic acid) at room temperature having a flow rate of 0.5000mL/min.

Pharmacokinetic analysis

Plasma drug concentrations for both test and reference product at specified time points were used in pharmacokinetic calculations of the study. For estimating pharmacokinetic parameters, the dataset was prepared by using program kinetic (version 4.4, Adept Scientific, UK) followed by the non-compartmental method of analysis. The pharmacokinetic parameters, included in the study protocol, comprised: time to reach the maximum concentration (tmax), maximum plasma concentration (Cmax), half-life (t1/2), area under the plasma time curve up to last quantifiable time (AUC0→t), area under the plasma time curve up to time infinity (AUC0→∞), elimination rate constant (k) and ratio Cmax/AUC0→∞.

Recovery experiment

Determination of the percentage recoveries was done by measuring the peak areas of the drug from the prepared plasma low, medium and high-quality control samples. The peak areas of the plasma low, medium and high-quality control samples were then compared to the absolute peak area of the un-extracted
standards containing the same concentrations of the atorvastatin. The results are presented in Table 2. Recovery after extraction was 81.82% - 85.77%.

Table 2: Absolute recovery of analyte from plasma drug samples.

| Inj No. | QCL 3ng/mL | QCM 30ng/mL | QCH 40ng/mL | QCL 3ng/mL | QCM 30ng/mL | QCH 40ng/mL |
|---------|------------|-------------|-------------|------------|-------------|-------------|
| 1       | 2.839      | 27.359      | 39.864      | 2.697      | 25.554      | 34.229      |
| 2       | 2.708      | 28.588      | 36.604      | 2.194      | 24.3        | 35.44       |
| 3       | 2.912      | 26.936      | 39.779      | 1.976      | 21.961      | 31.615      |
| 4       | 2.707      | 29.444      | 37.242      | 2.17       | 23.752      | 32.536      |
| 5       | 2.97       | 28.742      | 38.523      | 2.529      | 22.49       | 30.878      |
| Mean    | 2.827      | 28.214      | 38.403      | 2.313      | 23.612      | 32.94       |

% Recovery 81.82 83.69 85.77

Statistical Analysis

The primary variables C\textsubscript{\text{max}}, AUC\textsubscript{0-t}, and AUC\textsubscript{0-\infty} were taken into consideration to evaluate the bioequivalence between the test and reference formulations. Two-way ANOVA with crossover design (2x2) was used to determine the effects of formulation, period, sequence, and subjects on log-transformed C\textsubscript{\text{max}}, AUC\textsubscript{0-t}, and AUC\textsubscript{0-\infty}. The ratios of the log-transformed C\textsubscript{\text{max}}, AUC\textsubscript{0-t}, and AUC\textsubscript{0-\infty} values were calculated for both formulations and 90% CIs were obtained. The two formulations were considered bioequivalent based on 90% CI of the test/reference ratios of AUC within 0.80 to 1.25.

Results

The mean plasma concentrations of atorvastatin observed in the 24 healthy volunteers studied with two oral pharmaceutical formulations: Lipitor® and Stacor® were shown in Figure 2. A total of 4 mild adverse events including 1 abdominal discomfort and 3 venipuncture syncope were reported by three subjects (6.5%). These were not considered to be associated with the administration of the atorvastatin. The primary variables C\textsubscript{\text{max}}, AUC\textsubscript{0-t}, and AUC\textsubscript{0-\infty} values of the pharmacokinetic parameters were found out to be: 0.257 for C\textsubscript{\text{max}}; 0.657 for t\textsubscript{\text{max}}; 0.879 for AUC\textsubscript{0-24}; 0.856 for AUC\textsubscript{0-\infty}; 0.763 for MRT; 0.451 for AUMC\textsubscript{0-24}; 0.485 for AUMC\textsubscript{0-\infty}; 0.640 for k\textsubscript{el}. These high P-values indicate a lack of significant difference in each biochemical parameter. Based on a comparison of the AUC\textsubscript{\text{max}} values of the test and reference preparation, the relative bioavailability of the test preparation was 94.56% of that of the reference preparation. The subject, period and treatment were applied to the ln AUC\textsubscript{\text{max}}, ln C\textsubscript{\text{max}}, ln AUC\textsubscript{0-24}, ln AUC\textsubscript{0-\infty}, ln k\textsubscript{el}, and ln C\textsubscript{\text{max}} values. The 90% confidence interval of ratios of test and reference preparation for C\textsubscript{\text{max}}, AUC\textsubscript{0-t}, and AUC\textsubscript{0-\infty} values are stated in Table 5. No statistically significant difference was found for the treatment values of AUC\textsubscript{\text{max}} and ln C\textsubscript{\text{max}}.

Table 3: Plasma pharmacokinetic parameters of 24 volunteers with the test and reference formulations.

| Pharmacokinetic Parameters | Mean | Geometric Mean | CV (%) | SD | Max | Min |
|----------------------------|------|---------------|--------|----|-----|-----|
| Reference Formulation (R)  | C\textsubscript{\text{max}} (ng/mL) | 26.166 | 26.001 | 11.268 | 2.948 | 32.163 | 19.106 |
|                            | T\textsubscript{\text{max}} (h)     | 2.396  | 2.325  | 24.6  | 0.589 | 3.5  | 1.5  |
|                            | AUC\textsubscript{0-24} (h.ng/mL)  | 271.562 | 266.69 | 19.266 | 52.318 | 379.243 | 156.634 |
|                            | AUC\textsubscript{0-\infty} (h.ng/mL) | 309.975 | 304.519 | 19.226 | 59.594 | 460.19 | 179.305 |
|                            | k\textsubscript{el}                | 0.053  | 0.053  | 11.327 | 0.006 | 0.063 | 0.043 |
|                            | t\textsubscript{1/2} (h)           | 13.178 | 13.0928 | 11.907 | 1.555 | 16.159 | 10.955 |

Test Formulation (T)

| C\textsubscript{\text{max}} (ng/mL) | 27.617 | 27.495 | 9.53 | 2.632 | 32.373 | 22.915 |
| T\textsubscript{\text{max}} (h)     | 2.479  | 2.4    | 25.566 | 0.634 | 3.5   | 1.5   |
| AUC\textsubscript{0-24} (h.ng/mL)  | 256.799 | 251.503 | 21.029 | 54.002 | 387.678 | 153.488 |

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Discussion

The Food and Drug Administration (FDA) highlights the same quality, effectiveness, and performance of generic drugs as brand-name drugs and it does not allow the difference in the effectiveness of generic drug product about 45 percent. A recent FDA’s review of 2,070 human studies between 1996 and 2007 stated that average difference in absorption between the generic and the brand was 3.5 percent [14]. Another study compared cardiovascular generic drugs to their brand name counterparts taking 38 published clinical trial results and found no evidence that brand-name cardiovascular drugs worked any better than generic cardiovascular drugs [15]. Statin drugs are one of the drug classes among them. However, these trials are rare among low and middle-income countries although many of these countries have huge drug market. Bangladesh is one of them.

To ensure the clinical effectiveness and safety of the generic product to that of innovator brand and due to the unsuitability of repetition of clinical studies for generic products, pharmacokinetics and bioequivalence studies are considered as ‘second to none’. Determination of parent drug release from dosage forms over metabolites is a general recommendation for bioequivalence studies from US FDA guidelines [16]. The considerable variability between individuals and substantial differences between the two formulations within each individual are two striking observations under important consideration in any comparative pharmacokinetics study. Considerable interindividual variability, particularly in $C_{max}$, $T_{max}$, and AUC is evidenced for many drugs. This variability might be due to genetic polymorphism of the cytochrome P450 (CYP) isoform CYP2C19 [17-20]. Additionally, demographic data of subjects; age, weight, height and concomitant medication could add to the high inter-individual variability.

In the present study, atorvastatin 40mg tablet was detected in plasma over 0.5 hours to 48.0 hours in the reference and test preparations after administration. Peak plasma levels of atorvastatin 40mg for both the preparations were achieved between 2.0 and 3.5 hours (Figure 2). The mean peak plasma levels of reference formulation ranged between 19.106-32.163ng/mL compared to test preparation ranged between 22.915-32.373ng/mL (Table 3). The ratios of $C_{max}$ between these two formulations were ranged between 97%-104% (Table 5). Similarly, other main pharmacokinetic parameters $T_{max}$ and AUC for both formulations were compared by analysis of variance and no statistically significant difference was observed (Table 4). In fasting state, $C_{max}$, $t_{max}$, and $AUC_{0.4}$ values were comparable for the reference and the test preparation. Moreover, 90% confidence limits of ratios for $C_{max}$, $AUC_{0-72}$ and $AUC_{0-\infty}$ were found to be within the limits of bioequivalence acceptance ranges. The $C_{max}$, $AUC_{0-72}$ and $AUC_{0-\infty}$ ratios were 0.97238 - 1.04110, 0.99296 - 1.00641 and 0.99113 - 1.00255 respectively, which falls within the accepted limit to that of the reference preparation (i.e. 0.8 - 1.2). In this study, we selected these limits of acceptance, based on the variability of atorvastatin pharmacokinetics and to the fact that atorvastatin shows double peaks or major shouldering characteristics. It has been recommended that, in the case of drugs with a wide variability in absorption, these limits are adequate and are accepted in Europe [21,22]. Based on these study derived results, it can be inferred that the tested formulation of atorvastatin is bioequivalent with innovator formulation.

Table 4: P-values for sources of variations in plasma pharmacokinetic parameters obtained from Analysis of Variance (ANOVA).

| Pharmacokinetic Parameters | Formulation | Period | Sequence | Subject |
|----------------------------|-------------|--------|----------|---------|
| $C_{max}$ (ng/mL)          | 0.18        | 0.57   | 0.36     | <0.01   |
| $t_{max}$ (h)              | 0.47        | 0.67   | 0.54     | 0.07    |
| $AUC_{0-72}$ (h,ng/mL)     | 0.78        | 0.78   | 0.46     | <0.01   |
| $AUC_{0-\infty}$ (h,ng/mL) | 0.76        | 0.89   | 0.85     | <0.01   |
| $K_{el}$                   | 0.64        | 0.34   | 0.71     | 0.13    |
| $t_{1/2}$ (h)              | 0.76        | 0.42   | 0.83     | 0.17    |
**Bioequivalence Studies and Pharmacokinetic Properties of Atorvastatin 40Mg Tablet in Healthy Bengali Subjects**

### Table 5: 90% confidence interval of the ratios of the Test and Reference preparation.

|                | Untransformed Data | In-Transformed Data |
|----------------|--------------------|---------------------|
| $C_{\text{max}}$ | 0.89461 - 1.15268  | 0.97238 - 1.04110   |
| AUC$_{0-\infty}$ | 0.95754 - 1.03991  | 0.99296 - 1.00641   |
| AUC$_{0-t}$     | 0.94615 - 1.01599  | 0.99113 - 1.00255   |

However, this study had some limitations. Firstly, the study was open labeled which might not address tolerability of two formulations. Secondly, as the study was conducted solely on healthy Bengali subjects who are free from hypercholesterolemia and hypertriglyceridaemia, pharmacokinetics might be different for patients and/or other targeted populations. Thirdly, the study was limited only to male volunteers. Thus, gender variation in the pharmacokinetics of atorvastatin in Bengali subjects was not reflected in the study. Fourthly, the study didn’t observe the effect of food on pharmacokinetics. Further research regarding gender variation and effect of food on the pharmacokinetics of atorvastatin on Bengali population is recommended.

### Conclusion

The study found that the reference and test formulations of atorvastatin 40mg tablet are pharmacokinetically safe and met the regulatory benchmark for bioequivalence in healthy Bengali subjects. The test product Stacor® may be supplanted for the regulatory benchmark for bioequivalence in healthy Bengali subjects who are free from hypercholesterolaemia and hypertriglyceridaemia.

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### Declaration of Interest Statement

The authors have no conflict of interest.

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