Tetrandrine inhibits migration and invasion of human renal cell carcinoma by regulating Akt/NF-κB/MMP-9 signaling

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Abstract

Renal cell carcinoma (RCC) is known as one of the most lethal malignancies in the urological system because of its high incidence of metastasis. Tetrandrine (Tet), a traditional Chinese herbal medicine, exerts a potent anti-cancer effect in a variety of cancer cells. However, the anti-metastatic effect of Tet and its possible mechanism in RCC is still unclear. The present study revealed that Tet significantly suppressed the migration and invasion of RCC 786-O and 769-P cells in vitro. Mechanistically, the protein levels of matrix metalloproteinases 9 (MMP-9), phosphorylated PI3K, PDK1, Akt and NF-κB were markedly reduced after Tet treatment. Moreover, co-treatment with LY294002 (PI3K inhibitor) could further enhance the Tet-inhibited migration and invasion, and the NF-κB and MMP-9 protein levels were further decreased. Similar results were observed after PDTC (NF-κB inhibitor) co-treatment. Conversely, SC79, an Akt activator, could partially reverse the anti-metastatic effects of Tet, accompanied by the restoration of NF-κB and MMP-9 protein levels. In conclusion, the current results indicated that Tet inhibited migration and invasion of RCC partially by regulating Akt/NF-κB/MMP-9 signaling pathway, suggesting that Tet may be a potential therapeutic candidate against metastatic RCC.

Introduction

Renal cell carcinoma (RCC) is the third most common urologic malignancy, accounting for approximately 3% of all cancers and 90% of kidney cancers in adults [1]. The advanced diagnostic methods and systemic treatment (chemotherapy and radiotherapy), benefits a remarkable improvement in 5-year survival and recurrence-free survival. Despite that, 30% patients with RCC continue to progress into the metastatic disease, resulting in a median overall survival of metastatic RCC of only 12 months [2]. Furthermore, it is insensitive to chemotherapy, and drugs for immunotherapy and targeted therapy have a low effect on the treatment for metastatic RCC [3]. Thus, there is an urgent need to explore novel therapeutic agents against metastatic RCC.
Tumor metastasis has been identified as a leading cause of cancer-related deaths [4]. Moreover, the degradation of the extracellular matrix (ECM) by proteinases is a vital step in the metastatic process [5]. Matrix metalloproteinases (MMPs), members of the zinc-dependent endopeptidases, could modulate the breakdown of the ECM and facilitate tumor invasion [6, 7]. Among the MMPs, MMP-9 is closely correlated with tumor metastasis in various cancers, including gastric cancer [8], prostate cancer [9], and cervical cancer [10]. Therefore, MMP-9 and other MMPs may be considered as effective targets for anti-cancer drugs [11].

Tetrandrine (Tet), a bisbenzylisoquinoline alkaloid, is isolated from traditional Chinese medicine Stephaniae [12]. Tet has been widely used as anti-hypertension, anti-arrhythmic, and anti-rheumatism agent [13–15]. Recently, accumulating evidence suggested the anti-cancer effects on various cancers, which were associated with growth inhibition, induction of apoptosis and cell cycle arrest, and suppression of angiogenesis in vitro and in vivo [16, 17]. Our previous study had demonstrated the anti-cancer effects of Tet on the bladder and prostate cancers [18, 19]. Despite its potential of anti-proliferation in solid tumors, whether Tet inhibits cell migration and invasion of RCC has not yet been elucidated. Also, the underlying mechanism of Tet on cell migration and invasion is unknown. Hence, our study aimed to explore the effects of Tet on RCC cell lines and to investigate its possible mechanisms.

Material and methods

Cell culture

Human RCC cell lines 786-O and 769-P were obtained from the American Type Culture Collection (ATCC; Manassas, VA, USA) and cultured in RPMI 1640 medium, which contains 10% fetal bovine serum (FBS; Gibco, Grand Island, NY, USA) and 1% penicillin-streptomycin (Invitrogen, Carlsbad, CA, USA), in a humidified incubator with 5% CO₂ at 37°C.

Reagents

Tetrandrine (Tet) (C₃₈H₄₂N₂O₆) and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were obtained from Sigma Chemical Co. (St. Louis, MO, USA). Tet was solubilized in 0.1 M HCl to a concentration of 25 mg/mL as the stock solution and then diluted to the desired concentrations before use. Antibodies against PI3K, phosphor-PI3K, PDK1, phospho-PDK1, Akt, phospho-Akt, NF-κB, and MMP-9 were purchased from Cell Signaling Technology, Inc. (Beverly, MA, USA). LY294002 (PI3K inhibitor), PDTC (NF-κB inhibitor), TNF-κB (NF-κB activator) were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA). SC79 were purchased from abcam. Inc.(Cambridge, Britain).

MTT assay

786-O and 769-P cells were seeded in 96-well plates (1×10⁴ cells/well, 90% density) and exposed to different doses of Tet. Then, 0.5 mg/mL MTT dye solution was added to each well, and the cells were incubated at 37°C for 4 h. Subsequently, the culture medium was discarded, and dimethyl sulfoxide (DMSO) was added to solubilize the precipitate. A 96-well microplate reader (Bio-Rad, Hercules, CA, USA) was used to estimate the absorbance at 490 nm.

Wound healing assay

RCC 786-O or 769-P cells were seeded in 6-well plates. When the cell density reached up to 90% confluency, the cell monolayer was scratched using a 200-μL pipette tip. Then, the wounded cells were treated with Tet at different times and visualized in six randomly chosen fields by microscopy to evaluate the ability of cell migration.
Transwell migration assay

Transwell migration assays were performed to detect the anti-migratory ability of Tet on human RCC 786-O and 769-P cells. The cells (786-O: 2×10^4 or 769-P: 3×10^4 per chamber, respectively) treated with or without Tet were seeded into the upper chamber, while 800 μL of the medium containing 10% fetal calf serum was added to the lower chamber. After incubation in a humidified atmosphere at 37˚C for 24 h, the non-migrated cells in the upper chamber were removed with a cotton swab. The migrated cells in the bottom chamber were fixed with 4% paraformaldehyde for 10 min, stained with 0.1% crystal violet for 10 min, and images captured with a microscope five randomly chosen fields at 100× magnification.

Matrigel invasion assay

The upper parts of the transwell apparatus (polycarbonate membrane, 6.5 mm diameter, 8 μm pore size) were coated with 50 μL of 1:5 mixture of Matrigel: RPMI1640 medium. Cells (786-O: 8×10^4 or 769-P: 8×10^4 per chamber, respectively) treated with Tet were seeded into the upper chamber and incubated at 37˚C for 4 h. The subsequent procedure was similar to the transwell migration assay.

Western blotting

The RCC cells were collected and lysed after the Tet treatment. The clarified protein lysates (approximately 30–60 μg) were electrophoresed on SDS-polyacrylamide gel (10%) and transferred to polyvinylidene fluoride membranes. Western blotting was performed with primary indicated antibodies at 4˚C, overnight. Then, the membranes were washed with TBST buffer and incubated with horseradish peroxidase-conjugated secondary antibody for 1 h. Subsequently, the proteins of interest were analyzed by enhanced chemoluminescence and quantified using Image Lab software (Bio-Rad, Hercules, CA, USA).

Statistical analysis

GraphPad Prism 5.2 software (GraphPad Software Inc., La Jolla, CA, USA) was used for all statistical analyses. Statistical differences in different groups were compared by Student’s t-test (two-sided) or one-way analysis of variance (ANOVA). Significant differences were represented as P<0.05.

Results

Effects of Tet on cell proliferation of RCC 786-O and 769-P cells

Before we explored the biological effects of Tet (Fig 1A) on cell mobility ability, we first investigated the effects of Tet on cell growth using a modified MTT assay. The 786-O and 769-P cells with 90% density were seeded and treated with various concentrations of Tet from 0.05 to 15.0 μM (Fig 1B and 1D). And the effects of Tet on 786-O cell growth inhibition, was shown in Fig 1C, with the 24h Tet treatments at 0.1 μM, 0.25 μM and 0.5 μM resulting in 2.2%, 6.3%, and 9.5% of inhibitory rate, respectively. Similar effects were also exhibited in 769-P cells (Fig 1E). The results indicated that Tet markedly inhibited cell growth at the concentration of ≥0.5 μM in these two cell lines, while Tet at 0.5 μM represented for a less than 10% inhibitory rate of cell proliferation. Furthermore, a time-dependent manner of cell growth inhibition was observed in 786-O and 769-P cells after Tet treatment (Fig 1F). Based the results from the MTT assay, Tet at 0.5 μM was chosen to conduct the subsequent experiment.
Effects of Tet on migration and invasion of RCC 786-O and 769-P cells

To further analyze whether Tet could inhibit the migration and invasion of human RCC, we performed wound healing assay and transwell assay. Results from wound healing assay indicated that 0.5 μM Tet-treated 786-O cells migrated much more slowly compared with control cells at 24 h (Fig 2A). Similarly, 769-P cells treated with Tet also greatly impeded cell migration (Fig 2B). Additionally, we also found that Tet markedly attenuated the migration and invasion of 786-O and 769-P cells using a transwell assay (Fig 2C and 2D). Altogether, these results confirmed the anti-metastatic effect of Tet on RCC cells, as determined by wound healing assay and transwell assay.

Tet significantly decreased the expressions of NF-κB, MMP-9, phospho-AKT, phospho-PI3K and phospho-PDK1 in human RCC

Previous studies have reported that Akt/NF-κB signaling pathway was positively correlated with tumor metastasis [20, 21]. To explore whether Akt/NF-κB signaling was also necessary for Tet-induced anti-metastatic effects on 786-O and 769-O cells, Western blotting was then performed. As expected, the protein level of NF-κB was markedly reduced by Tet in a concentration-dependent manner (Fig 3A–3D). Additionally, exposing 786-O and 769-P cells to Tet resulted in a significant suppression of Akt phosphorylation. Intriguingly, the levels of phospho-PI3K and phospho-PDK1 protein, which were upstream regulators of Akt, were also decreased after Tet treatment, suggesting a role of Tet in inactivation of Akt/NF-κB signaling pathway (Fig 3A–3D and S1 Fig). Previous studies have reported that MMP-9, an indicator of tumor metastasis, was regulated by NF-κB [9]. Our results exhibited that there was a dramatic decrease of MMP-9 expression in Tet-treated 786-O and 786-P cells (Fig 3A–3D).
together, these results indicated that Akt/NF-κB/MMP-9 signaling was inhibited in Tet-treated 786-O and 769-P cells.

**Akt phosphorylation was involved in Tet-induced anti-metastatic effects in RCC**

To further explore the role of Akt in Tet-inhibited cell migration and invasion, LY294002 (PI3K inhibitor), which was widely used to inhibit the activation of Akt, was used to combine with Tet for the subsequent experiment. Our results revealed that co-treatment with LY294002 could further reinforce the anti-metastatic effects of Tet in 786-O and 769-P cells (Fig 4A and 4B), as well as the protein levels of NF-κB and MMP-9 compared with the Tet alone and LY294002 alone treatment cells (Fig 4C and 4D). Additionally, the Akt activator SC79, could remarkably reverse the inhibitory effect of migration and invasion by Tet in RCC, accompanied with the restoration of NF-κB and MMP-9 protein levels (S2A–S2D Fig). These data strongly supported that Tet exhibited anti-metastasis effect by inhibition of Akt phosphorylation.

**Tet inhibited human RCC migration and invasion via negatively regulating NF-κB expression**

To gain deeper insights into the involvement of Akt/ NF-κB signaling, PDTC, a NF-κB inhibitor, was then used. As expected, the anti-metastatic effect of Tet on RCC could be further enhanced by PDTC (Fig 5A and 5B), and a dramatic decline of MMP-9 protein levels was observed (Fig 5C and 5D), suggesting that NF-κB might function as a downstream regulator of Akt in Tet-inhibited cell migration and invasion.
Discussion

Metastasis is a vast obstacle for systemic treatment in patients with RCC [22]. Metastasis is the hallmark of cancer, which spreads the tumors from the original site to the distant organs. Moreover, it is a complicated process implicated in local invasion, transfer, extravasation, and
tumor deposit [23]. Of the metastatic process, the degradation of ECM is a key step in initiating tumor invasion, and interference with this step may be an efficient strategy for tumor treatment [24]. For tumor invasion and metastasis, MMPs are the crucial proteolytic enzymes to degrade the ECM and initiate the metastatic process [7]. Although more than one MMPs have been identified, MMP-9 is the enzyme most closely correlated with the degradation of ECM and is highly expressed in metastatic cancer [25].

Tet, a Chinese herbal medicine, exerts anti-metastasis activity in a variety of cancers. Our previous study revealed that Tet could inhibit the expression of MMP-9 via the inactivation of Akt/NF-κB signaling [9]. Additionally, Tet inhibited the metastasis of breast cancer in vivo, partially by regulating the endothelial cell-specific molecule-1 (ESM-1) and intercellular cell adhesion molecule-1 (ICAM-1) [26]. In the present study, our findings revealed that Tet significantly suppressed the migratory and invasive abilities of human RCC 786-O and 769-P cells. Moreover, our results demonstrated that Tet could markedly inhibit the protein level of MMP-9 in both 786-O and 769-P cells. The anti-invasive capacity of Tet may be of great importance in decreasing cancer mortality in RCC patients. Further studies are needed to explore the underlying mechanism of Tet on the regulation of MMP-9.

NF-κB, a pivotal transcriptional factor in cancer cells, participates in the tumor progression by regulating angiogenesis and suppressing apoptosis [27, 28]. Also, it has been proven as a regulator of MMP-9 to promote tumor invasion and metastasis [29, 30]. In addition, Akt signaling regulates the expression of NF-κB as well as MMP-9 in several cancer cells [20, 21]. Thus, to elucidate the effect of Tet on activation, Akt/NF-κB signaling was assessed by western blotting. In our study, Akt, PI3K, PDK1 phosphorylation and NF-κB activation were found to be inhibited by Tet treatment, whereas, no remarkable effect was observed on total Akt, PI3K and PDK1, indicating that Tet could inhibit Akt/NF-κB signaling pathway in 786-O and 769-P cells.

Moreover, PI3K inhibitor LY294002 further reinforced the decrease of NF-κB and MMP-9 protein levels induced by Tet. Similar results were observed after NF-κB inhibitor PDTC treatment. In contrast, the decreased NF-κB and MMP-9 protein levels by Tet were partially
reversed after SC79 treatment. These findings strongly suggested that Tet exerted an anti-metastatic effect at least partially by Akt/NF-κB/MMP-9 axis.

However, our present study has several limitations. Although, we confirmed that Akt/NF-κB signaling pathway played an essential role in Tet-inhibited cell migration and invasion, the effect of the downstream molecules of Akt, such as activator protein-1 (AP-1) and STAT3 were still unknown. On the other hand, we did not establish a lung metastasis in vivo model of nude mice to validate our in vitro results. Therefore, further investigations are imperative to resolve these issues.

Our results concluded that Tet suppressed cell migration and invasion of human RCC by negatively regulating Akt/NF-κB signaling and MMP-9 expression. Importantly, our findings suggest a potentially significant correlation of Tet and Akt/NF-κB signaling in the metastatic progression of human RCC; therefore, Tet may be a potential chemotherapeutic candidate against metastatic RCC.

Supporting information

S1 Fig. Effects of Tet on the phosphorylated levels of PI3K and PDK1. 786-O (A) and 769-P (B) cells treated with vehicle or Tet (0.1μM to 0.5μM) for 24 h were immunoblotted for PI3K, phospho-PI3K, PDK1 and phospho-PDK1. β-actin was used for a loading control. Representative results from three independent experiments were shown. (TIF)

S2 Fig. Akt activator SC79 reverses Tet-induced anti-metastatic effects. In vitro cell mobility of 786-O (A) and 769-P (B) cells were determined by transwell assay under tetrandrine (0.5μM), SC79 (10μM), or the combined treatment for 24 h. The protein levels of Akt, phospho-Akt, NF-κB and MMP-9 of 786-O (C) and 769-P (D) cells were detected as the indicated treatments by western blotting after SC79 treatment. Representative results from three independent experiments were shown. (TIF)

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