Leaf extract of *Morinda lucida* improves pancreatic beta-cell function in alloxan-induced diabetic rats

Adam Olaitan Abdulkareem, Adedoyin Iggunnu, Adeola Adefoluke Ala and Lawrence Aderemi Olatunji

Abstract

Chemotherapy remains the utmost treatment for Type 1 diabetes (T1D) patients. This however, predisposes the patients to a wide range of serious complications, thus, the need for alternative therapeutic agents that target pancreatic β-cell function. This study investigated the effects of aqueous leaf extract of M. lucida (MLE) on β-cell dysfunction and atherogenic dyslipidaemia in alloxan-induced T1D in rats. Twenty-five Wister rats (156-168g) were randomly divided into normal, diabetic, diabetic + glibenclamide (5 mg/kg), diabetic + 120 mg/kg MLE and diabetic + 240 mg/kg MLE groups (n = 5/group). Treatments were via oral route for 14 days. Our findings revealed that, 120 mg/kg MLE significantly reduced hyperglycaemia, improved insulinaemia as well as β-cell function, and attenuated weight loss in alloxan-induced diabetic rats. The extract also attenuated (*p* < 0.05) atherogenic dyslipidaemia and malondialdehyde. The activities of the extract compared favourably with glibenclamide. This study suggested that, hypoglycaemic and mitigating effects of aqueous leaf extract of *M. lucida* on atherogenic dyslipidaemia and pancreatic β-cell dysfunction were through reduction in lipid peroxidation. The extract may therefore represent an effective source of novel drugs against TID and cardiovascular diseases. Further study is recommended, to explore the extract’s mechanism of oxidative repair.

Introduction

Type 1 diabetes (T1D) is a poorly controlled chronic metabolic disorder that is characterized by beta-cell destruction, insulin deficiency and hyperglycaemia [1,2]. It accounts for about 5–10% global incidence of diabetes mellitus, affecting approximately 20 million people worldwide, with rising incidence [3]. Approximately 542,000 children (aged 0–14 years) have T1D and 86,000 new cases are diagnosed globally each year [4].

Although, the disease is often referred to as ‘juvenile diabetes’ with symptoms such as polydypsia, polyphagia, polyuria and weight loss, alongside overt hyperglycemia [5], it has been recently shown that individuals with T1D are mostly adults and almost half of the cases of T1D were diagnosed after age 30 years [6–8]. Furthermore, it has been reported that about 5–15% of adults diagnosed with T2D may actually have T1D [9]. Hence, poor understanding of this disease has led to underestimation in adults. Pathogenetically, T1D
results from an interplay between genetic and environmental factors that induce autoimmune destruction of insulin-producing pancreatic β-cells [10]. Factors such as diet, enteroviruses and toxins have been proposed to influence the development of T-cell-dependent autoimmunity in genetically susceptible individuals [11].

Normally, hyperglycemia inhibits glucagon secretion. However, during insulin insufficiency, hyperglycemia elevates glucagon level and body then utilizes fat for energy instead of glucose [12,13]. This leads to accumulation of ketones in the blood, consequently results in diabetic ketoacidosis and therefore slows down body function [12,13]. This may consequently result in coma and eventually death [14]. Similarly, hyperglycemia induces multiple cellular changes which generate oxidative stress via several mechanisms such as non-enzymatic glycation, glucose oxidation and alterations in polyol pathway activity [15,16]. Consequently, the generated oxidative stress leads to DNA, lipid, protein and carbohydrate damage, and increases the risk of developing complications such as neuropathy, nephropathy, retinopathy and accelerated coronary artery disease [17].

Chemotherapy, including insulin treatment, remains the utmost treatment for people with T1D [18,19]. These therapies act by improving insulin secretion, decreasing glucose release from the liver or reducing gastrointestinal absorption of sugars [19]. However, while insulin and other commonly administered drugs do not provide absolute cure, they also predispose the patients to a wide range of serious complications, such as heart and kidney diseases and blindness [20,21]. To avoid these complications, production of new β-cells becomes necessary. These new β-cells should ideally be formed from already existing cell sources within an individual with diabetes to avoid immunosuppression [22]. Alternatively, exogenous sources of surrogate β-cells, such as adult human pancreases donated after death, fetal pancreas, pluripotent and multipotent stem cells have also been propagated, but not much success has been recorded so far [23].

In order to meet up with the current challenge, therefore, there is a need to search for effective and safe alternative sources of treatment through herbal therapy, to provide improved treatment for diabetes and associated complications [23]. Morinda lucida is one of the widely used medicinal plants in West Africa (especially Nigeria), for the management/treatment of various types of ailments [24]. Antidiabetic effects of M. lucida extracts have been previously reported [25–28]. However, there is insufficient information on the effect of these extracts on pancreatic β-cell function and atherogenic dyslipidaemia. In this study, we investigated the ameliorative effects of M. lucida aqueous leaf extract on pancreatic β-cell function and atherogenic dyslipidaemia in alloxan-induced T1D.

**Materials and method**

**Sample collection and preparation of extract**

Fresh leaves of M. lucida were harvested from Rot-Tund farms Nigeria investment limited, Ila-Orangun, Osun State, Nigeria in December, 2017 and authenticated at the herbarium department, Forest Research Institute of Nigeria, Ibadan, Nigeria (Registration number FHI.110162). The leaves were properly cleaned, air dried and macerated. The macerated samples (100 g) were dissolved in 1000 ml of distilled water for 24 h and then filtered. The filtrate was concentrated under reduced pressure, using a rotary evaporator at 55°C to yield a crude semisolid mass, which was stored in a refrigerator at 4°C until required.

**Experimental animals**

Twenty-five male Wistar rats, weighing 156–168 g were procured from the Animal House, Department of Biochemistry, University of Ilorin,
They had unrestricted access to standard feed and water. The animals were main-
tained under standard environmental conditions of temperature, relative humidity and dark/light
cycle, in accordance with the guidelines of National Institutes of Health Guide for the Care
and Use of Laboratory Animals and the Institutional Ethical Review Board of University of
Ilorin. Body weight, food consumption and water intake were monitored throughout the period of
administration.

**Induction of diabetes**

The rats were fasted overnight and then injected, intra-peritoneally, with a single dose
of 0.5 ml of 160 mg/kg body weight (bw) of Alloxan monohydrate (a product of Mekphar
Chemical Pharmaceutical Joint-Stock Company, Chiminh City, Vietnam), dissolved
in freshly prepared normal saline, to induce T1D [29]. The control animals (nondiabetic)
were injected with 0.5 ml of the vehicle (nor-
mal saline). Stable hyperglycemia was con-
firmed on ninth day [29], using glucometer
(ACCU-Chek, Roche Diagnostics) and this was
taken as week 0. Rats with fasting blood glu-
cose level greater than 200 mg/dl were con-
sidered diabetic and used for this study.

**Animal grouping and treatment**

After 2 weeks of acclimatization, animals
were randomly assigned to five groups (n
= 5/group). Groups 1 (normal control) and
2 (diabetic control): received distilled water (vehicle) daily; Group 3: (diabetic + gliben-
clamide): received 5 mg/kg bw glibencla-
mide; Groups 4 (diabetic + 120 mg/kg
MLE): received 120 mg/kg bw aqueous *M.
lucida* leaf extract; while Group 5 (diabetic + 240 mg/kg MLE): received 240 mg/kg bw
aqueous *M. lucida* leaf extract. The MLE
doses used were in accordance with pre-
vious study [25], with slight modification.

All treatments were through oral administra-
tion and lasted for 14 days.

**Blood collection**

At the end of experiment, four (4) rats, from each
group, were sacrificed by cervical dislocation and
blood was then collected by cardiac puncture into plain bottle. Blood samples were centrifuged
at 3000 rpm for 5 min and serum was stored
frozen until needed for biochemical assays.

**Biochemical assays**

Insulin was determined using ELISA kit from
Ray Biotech, Inc. (Georgia, USA), while malon-
dialdehyde (MDA), a marker of lipid peroxida-
tion, was measured by standardized method
using kit obtained from Oxford Biomedical
Research Ltd (Rochester Hills, MI). Estimation
of total cholesterol (TC) and triglyceride (TG)
was done by standardized enzymatic colori-
metric methods using assay kit obtained from
Fortress Diagnostics Ltd. (Antrim, United
Kingdom). High-density lipoprotein-cholesterol
(HDL-C) was measured by enzymatic clearance
assay (Daiichi Pure Chemicals Co., Ltd., Tokyo,
Japan) while low-density lipoprotein-choles-
terol (LDL-C) was estimated using modified
Friedewald’s formula [30]. TC/HDL-C and TG/
HDL-C ratios were estimated as markers of
atherogenic lipid indices [31].

**Fasting blood glucose and pancreatic β-
cells**

Weekly fasting blood glucose was assessed using
 gluometer (ACCU-Chek, Roche Diagnostics). The pancreatic β-cell function was estimated using
HOMA-β formula [31].

**Statistical analysis**

Data were analyzed and presented as mean ±
SEM. One-way analysis of variance (ANOVA) fol-
lowed, by Bonferroni post hoc test was performed,
using PRISM 5.0 (GraphPad Software, USA). Values were considered statistically significant at $p < 0.05$.

Results

**Effect of M. lucida leaf extract (MLE) on fasting blood glucose (FBG) and fasting insulinemia in alloxan-induced diabetic rats**

Fasting blood glucose was higher than 200 mg/dl in diabetic control as well as 240 mg/kg bw MLE-treated group (postinduction), throughout the experimental period (Figure 1(a)), while treatment with glibenclamide and 120 mg/kg MLE progressively reduced FBG ($p < 0.05$). There was decrease ($p < 0.05$) in fasting insuline mia in diabetic control, compared with normal control (Figure 1(b)). However, glibenclamide and 120 mg/kg MLE treatments significantly improved fasting insulinemia.

**Effect of M. lucida leaf extract (MLE) on body weight and pancreatic $\beta$-cell function in alloxan-induced diabetic rats**

Alloxan treatment resulted in gradual loss in body weight (Figure 2), while glibenclamide and MLE increased weight loss. Similarly, pancreatic $\beta$-cell function was reduced ($p < 0.05$) in diabetic control when compared with normal control, whereas glibenclamide and 120 mg/kg MLE significantly improved $\beta$-cell function (Figure 3).

**Effect of M. lucida leaf extract (MLE) on lipid profile, atherogenic lipids and malondialdehyde (MDA) in alloxan-induced diabetic rats**

Table 1 shows the effect of glibenclamide and MLE on lipid profile in alloxan-induced diabetic rats. There was increase ($p < 0.05$) in total cholesterol (TC) and LDL-C in diabetic control.

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**Figure 1.** Effect of *M. lucida* leaf extract (MLE) on fasting glycemia and insulinemia in alloxan-induced diabetic rats. Fasting glycemia was kept above 200 mg/dl in both diabetic control and 240 mg/kg MLE group, while glibenclamide and 120 mg/kg MLE treatments lowered fasting glycemia. Meanwhile, alloxan reduced circulating insulin, while administration of glibenclamide and 120 mg/kg MLE elevated blood insulin level (*$p < 0.05$ vs. normal control; $*p < 0.05$ vs. diabetic control; $n = 4$).

**Figure 2.** Effect of glibenclamide and *M. lucida* leaf extract (MLE) on body weight in alloxan-induced diabetic rats. Alloxan treatment led to decrease in body weight, while alloxan + glibenclamide and alloxan + MLE (120 and 240 mg/kg) treatments improved body weight ($n = 4$).
compared with normal control, whereas, HDL-C was decreased ($p < 0.05$) in diabetic control. Upon treatment with glibenclamide and MLE, alloxan-induced dyslipidemia was attenuated. As shown in Figure 4, the atherogenic indices (TC/HDL-C and TG/HDL-C) were increased ($p < 0.05$) in diabetic control when compared with normal control. However, treatment with glibenclamide and MLE led to attenuation in atherogenic indices. Similarly, alloxan treatment elevated MDA in diabetic control ($p < 0.05$) as compared with normal control, while glibenclamide and MLE lowered MDA (Figure 5).

**Discussion**

This study investigated the effect of aqueous leaf extract of *M. lucida* on pancreatic β-cell function and dyslipidemia in type 1 diabetic rats, using alloxan induction. Alloxan is a cytotoxic diabetogenic compound, widely used in experimental diabetes research [32]. It stimulates insulin-dependent diabetes (T1D) by inducing selective necrosis of the beta-cells of pancreatic islets, thus, destroying β-cells and reducing their function [33, 34]. Our results showed that *M. lucida* aqueous leaf extract lowered alloxan-induced hyperglycemia at low dose (120 mg/kg), which compared favorably with glibenclamide. The main etiology of T1D is destruction of pancreatic β-cells [35]. In this study, treatment of diabetic rats with the extract improved pancreatic β-cell function and thus, insulin production, better than the standard drug (glibenclamide) used. This may imply that, the leaf extract of *M. lucida* induced regeneration of pancreatic β-cell, and hence, enhanced its function. Our observation agrees with an earlier study, which reported that leaf extracts of *M. lucida* ameliorated alloxan-induced pancreatic damage [36].

Insulin deficiency in T1D results in stimulation of lipase, which hydrolyzes stored

![Figure 3](image1.png)

**Table 1. Effect of glibenclamide and *M. lucida* leaf extract (MLE) on lipid profile in alloxan-induced diabetic rats.**

| Parameters (mg/dl)       | Normal control | Diabetic control | Diabetic + 120 mg/kg MLE | Diabetic + 240 mg/kg MLE |
|--------------------------|----------------|------------------|--------------------------|--------------------------|
| Total cholesterol        | 202.70 ± 0.17  | 208.20 ± 0.15*   | 159.90 ± 0.73*           | 197.8 ± 2.30             |
| Triglyceride             | 114.40 ± 2.95  | 118.50 ± 7.65    | 96.77 ± 4.17             | 96.77 ± 0.77             |
| HDL cholesterol          | 70.30 ± 2.86   | 55.05 ± 3.20*    | 60.14 ± 3.49             | 73.12 ± 3.61             |
| LDL cholesterol          | 109.60 ± 0.20  | 125.3 ± 2.85*    | 85.39 ± 0.27*            | 100.50 ± 1.84            |

All data are presented as mean of four replicates ± standard error of mean (SEM).

(*$p < 0.05$ vs. normal control.*)
adipocyte triglyceride to fatty acids and glycerol, and consequently leads to loss of body weight [35]. In conformity with previous report [24], this study showed that *M. lucida* leaf extract improved alloxan-induced weight loss following increase in insulinemia. Our results thus imply that, the extract is capable of increasing metabolic efficiency via stimulation of insulin secretion. It may, therefore, be a good therapeutic agent for alleviating metabolic inequity in T1D and similar conditions.

Atherogenic indices (TC/HDL-C and TG/HDL-C ratios) have been identified as successful markers for predicting individuals at the risk of atherosclerosis [36,37], which begins early in life [38]. Therefore, our results which showed that alloxan increased TC/HDL-C and TG/HDL-C ratios and that treatment with leaf extract of *M. lucida* attenuated rise in the ratios are very prominent. The results suggest that, monitoring TC/HDL-C and TG/HDL-C ratios may assist in early detection of associated atherogenic dyslipidemia in T1D and that, leaf extract of *M. lucida* could serve as a good therapeutic agent for early intervention against cardiometabolic disorders.

It has been reported that reactive oxygen species produced by oxidative stress can penetrate through cell membranes, causing damage to the pancreatic β-cells and reducing insulin sensitivity [39,40]. Hence, oxidative stress plays a vital role in the pathophysiology of diabetes and pancreatic β-cell dysfunction [40,41]. Our result which shows elevated MDA level in diabetic control indicates increased oxidative stress (lipid peroxidation), and possibly increased tissue damage, by free radicals, in alloxan-induced diabetes [17]. This is in consonance with a previous human study [42] as well as previous observation which showed that alloxan-induced beta-cell destruction is mediated by lipid peroxidation [32]. Upon treatment with the extract, MDA was
significantly reduced. This suggests that aqueous leaf extract of *M. lucida* can improve body antioxidant defense. Meanwhile, earlier study has reported antioxidant activity of *M. lucida* extracts, attributed to the presence of bioactive components such as phenols, flavonoids and tannins [43]. We therefore suggest that antidiabetic effect of *M. lucida* leaf extract is through inhibition of lipid peroxidation, which consequently improves β-cell function and attenuates atherogenic dyslipidaemia.

**Conclusion**

This present study demonstrated that aqueous leaf extract of *M. lucida* improved pancreatic β-cell function and attenuated dyslipidemia in T1D condition by inhibiting lipid peroxidation. It may therefore represent an effective alternative source of novel drugs, for better management/treatment of T1D and cardiovascular diseases. Further study to explore the mechanism of oxidative repair is recommended.

**Author contributions**

All the authors have accepted responsibility for the entire content of this submitted manuscript and approved submission.

**Disclosure statement**

No potential conflict of interest was reported by the authors.

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