Ectomyelois Heinrich, 1956 in China, with descriptions of two new species and a key (Lepidoptera, Pyralidae, Phycitinae)

Yingdang Ren¹,², Linlin Yang¹,²

¹ Institution of Plant Protection, Henan Academy of Agricultural Sciences, Henan Key Laboratory of Crop Pest Control, Key Laboratory of Integrated Pest Management on Crops in Southern Region of North China, Zhengzhou 450002, China ² College of Life Sciences, Nankai University, Tianjin 300071, China

Corresponding author: Yingdang Ren (renyd@126.com)

Academic editor: M. Nuss | Received 8 June 2015 | Accepted 22 December 2015 | Published 3 February 2016

http://zoobank.org/88EB8E24-9BCA-4A2B-B78F-2D7149BCE2B9

Citation: Ren Y, Yang L (2016) Ectomyelois Heinrich, 1956 in China, with descriptions of two new species and a key (Lepidoptera, Pyralidae, Phycitinae). ZooKeys 559: 125–137. doi: 10.3897/zookeys.559.6076

Abstract

Only three species belonging to the genus Ectomyelois Heinrich, 1956 are recorded from China, of which two species, E. bipectinalis sp. n. and E. furvivena sp. n. are described as new. We discuss the status of Ectomyelois that has been treated as a junior synonym by previous authors; we treat it as a valid genus, revised status, based on characters of the venation and female genitalia. Photographs of the adults and illustrations of the genitalia are given, along with a key to the three known Chinese species.

Keywords

Lepidoptera, Pyralidae, Phycitinae, Ectomyelois, Ectomyelois ceratoniae, new species, key, China
Introduction

_Ectomyelois_ was established by Heinrich (1956) with _Myelois decolor_ Zeller, 1881 as the type species. It is a small genus consisting of six species: _E. ceratoniae_ (Zeller, 1839), _E. decolor_ (Zeller, 1881), _E. furvidorsella_ (Ragonot, 1888), _E. muriscis_ (Dyar, 1914), _E. zeteki_ Heinrich, 1956, and _E. austrella_ Neunzig & Goodson, 1992. Most are Neotropical, but _E. ceratoniae_ also occurs in the Oriental region. All but one species was described in detail by Heinrich (1956). _Ectomyelois_ was once treated as a junior synonym of _Spectrobates_ Meyrick by Roesler (1968) and subsequently of _Apomyelois_ Heinrich by Roesler & Küppers (1981). A few authors followed these treatments (e.g. Neunzig 1979; Roesler 1983; Heppner and Inoue 1992; Leraut 2002), but most authors (Munroe 1983; Inoue 1982; Palm 1986; Sinev 1986; Goater 1986; Neunzig 1990; Balinsky 1994; Yamanaka 2013) treated _Ectomyelois_ as a valid genus. Indeed, there is little to separate _Ectomyelois_ from _Apomyelois_ in the male genitalia, but the two genera can be distinguished by the venation and the different place of inception of the ducus seminalis from the corpus bursae in the female genitalia. Here, we agree that _Ectomyelois_, revised status, does indeed represent a valid genus.

_Ectomyelois_ was only represented by the common carob moth _E. ceratoniae_ in China before this study. Herein the three species are described, including two new species: _Ectomyelois bipectinalis_ sp. n. and _E. furvivena_ sp. n.

Material and methods

Genitalia dissections were carried out following the methods described by Li (2002). The photographs of the adults and venation were taken with a Leica M205A, and photographs of the genitalia and details of head were taken with a Leica DM750, with Leica Application Suite 4.2 software to capture images. All the specimens examined are deposited in NKUM unless otherwise noted.

Abbreviations

- **BMNH** Natural History Museum, London, UK.
- **HAASM** Insect Collection, Institution of Plant Protection, Henan Academy of Agricultural Sciences, Zhengzhou, China.
- **NKUM** Insect Collection, College of Life Sciences, Nankai University, Tianjin, China.
- **USNM** National Museum of Natural History, Smithsonian Institution, Washington, D.C. 20560, U.S.A.
- **ZMHB** Museum für Naturkunde, Universität Humboldt, Invalidenstrasse 43, 104 Berlin, Germany.
- **TD** Type depository.
- **TL** Type locality.
Systematic part

_Ectomyelois_ Heinrich, 1956

_Ectomyelois_ Heinrich, 1956: 43. Type species: _Myelois decolor_ Zeller, 1881, by original designation.

**Diagnosis.** Antenna of male usually shortly ciliate (bipectinate in _E. bipectinalis_ sp. n.), basal shaft without notch or other modifications, of female simple. Labial palpus upturned, nearly reaching apex, third segment distinctly shorter than second. Forewing with _R₂_ closely approximate to the stalk of _R₃₊₄₊R₅_, _M₂_ and _M₃_ stalked for less than half of their length. Hindwing with Sc + _R₁_ and _Rs_ strongly anastomosed for most of their lengths, _M₂_ and _M₃_ stalked for not over half of their length. Male genitalia with uncus subtriangular to bell-shaped, apical projection of gnathos simple, slightly bent and furcated at apex, transtilla well developed, juxta U-shaped, with lateral lobes stout, vinculum U-shaped, more truncate and less tapering, phallus without cornutus. Female genitalia with signum consisting of an elongate patch of scobinations (absent in _E. furvidorsella_) and ductus seminalis from corpus bursae near junction of corpus bursae and ductus bursae.

_Ectomyelois_ is similar to _Apomyelois_, but can be distinguished from the latter by the forewing with _R₂_ closely approximate to the stalk of _R₃₊₄₊R₅_, and the female genitalia with the ductus seminalis arising from the corpus bursae near junction of the corpus bursae and ductus bursae. In _Apomyelois_, the forewing with _R₂_ shortly stalked with _R₃₊₄₊R₅_, and the female genitalia with the ductus seminalis arising from anterior end of the corpus bursae.

**Distribution.** China (Gansu, Guangdong, Guangxi, Hainan, Taiwan, Yunnan), India, Sri Lanka, Sikkim, Israel, Mediterranean, Central Europe, Norway, United Kingdom, North Africa, Australia, Argentina, United States, Cuba, Haiti, Puerto Rico, Jamaica, Bahamas, Guatemala, Costa Rica, Colombia, Panama, Venezuela, Guiana, Surinam, Bolivia, Guyana, French Guiana, Brazil.

**Key to species of Ectomyelois from China**

1 Forewing with narrow grayish white, distinctly notched antemedial line (Fig. 1) .......................................................... _E. ceratoniae_
   – Forewing with antemedial line invisible .................................................. 2

2 Male flagellum bipectinate (Fig. 3b); transtilla trefoiled (Fig. 7); corpus bursae three times as long as wide (Fig. 11) ................................. _E. bipectinalis_ sp. n.
   – Male flagellum simple, not bipectinate (Fig. 5b); transtilla inverse-goblet shape (Fig. 9); corpus bursae twice as long as wide (Fig. 12) ......................... 

..................................................................................................................................................... _E. furvivena_ sp. n.
Ectomyelois ceratoniae (Zeller, 1839)
Figs 1, 2, 7, 10

Myelois ceratoniae Zeller, 1839: 176. TL: Laibach, Austria. TD: BMNH.
Phycis ceratoniella Fischer von Roeslerstamm, 1839: 147. TL: Laibach, Austria. TD: unknown.
Trachonitis pryerella Vaughan, 1870: 130. TL: London, England. TD: BMNH.
Myelois tuerckheimiella Sorhagen, 1881: 103. TL: Berlin, Germany. TD: ZMHB.
Euzophera zellerella Sorhagen, 1881: 104. TL: Berlin, Germany. TD: unknown.
Phycita dentilinella Hampson, 1896: 91. TL: Manipur, India. TD: BMNH.
Hypsipyla psarella Hampson, 1903: 30. TL: Sikkim, India. TD: BMNH.
Heterographis rivularis Warren & Rothschild, 1905: 31. TL: Nakheila, Sudan. TD: unknown.
Myelois oporedestella Dyar, 1911: 30. TL: Florida, USA. TD: USNM.
Myelois phoenicis Durrant, 1915: 303. TL: Constantine, Algeria. TD: BMNM.
Laodamia durandi Lucas, 1950: 142. TL: Tunisia. TD: unknown.
Apomyelois ceratoniae (Zeller): Roesler and Kuppers 1981: 80.
Ectomyelois ceratoniae (Zeller): Heinrich 1956: 44.

Material examined. CHINA: Guangdong: 9 ♂♂, 1 ♀, Mt. He (22°45′N, 112°57′E), 09–10-X-2002, coll. Guilin Liu & Binglan Zhang, gen. slide nos. RYD04529m, RYD04530f; 1 ♂, same data as former except dated 6-XI-2002. Guangxi: 1 ♀, Milv (21°59′N, 107°52′E), Nanping, Shangsi, 770 m, 3-IV-2002, coll. Shulian Hao & Huaijun Xue, gen. slide no. RYD04658; 1 ♂, Yachang Yard, Leye County (24°47′N, 106°33′E), 665 m, 24-VII-2004, coll. Jiasheng Xu, gen. slide no. KDH05263; 1 ♀, Longrui (22°45′N, 110°55′E), 18-VIII-2011, coll. Muchun Cheng, gen. slide no. RYD20120185 (deposited in HAASM); 1 ♂, 1 ♀, Nonggang (23°14′N, 108°10′E), 20-VIII-2011, coll. Dandan Zhang, gen. slide no. RYD2014237 (deposited in HAASM). Hainan: 1 ♂, Mt. Diaoluo (18°39′N, 109°54′E), 29-V-2007, 80 m, coll. Zhiwei Zhang & Weichun Li, gen. slide no. LJY10595. Yunnan: 1 ♂, 1 ♀, Ganlanba (22°45′N, 101°08′E), Xishuangbanna, 19-IV-1995, coll. Guangyun Yan, gen. slide nos. RYD04615m, LJY10107f; 1 ♂, 4 ♀, Mt. Yunpan (23°44′N, 100°39′E), Puer, 1600 m, 6-VII-2013, coll. Linlin Yang, gen. slide nos. RYD2014219m, RYD2014221f (deposited in HAASM).

Diagnosis. Wingspan 15.0–22.00 mm (Fig. 1). Ectomyelois ceratoniae can be recognized by the following characters: the forewing with a narrow, distinctly notched antemedial line, the hindwing with free element of Sc+Ry very short (Fig. 2); the uncus is bell-shaped, basally protruded on both sides, the apical projection of gnathos is stout, gently curved, about same length of the uncus, the trefoiled transtilla includes a pair of inflated bases and a more constricted central projection, the basally rectangular bursae with signum is an elongate patch of scobinations, the ductus seminalis from junction of corpus and ductus bursae in the female genitalia (Fig. 10). It is quite similar to E. bipectinalis sp. n., but with differences as mentioned in the diagnosis of the latter.
Ectomyelois Heinrich, 1956 in China, with descriptions of two new species and a key...

**Figures 1−6.** Adults of Ectomyelois spp. 1 *E. ceratoniae*, male (1a, head; 1b, antenna) 2 *E. ceratoniae*, venation, slide No. RYD04529w 3 *E. bipectinalis* sp. n., paratype, male (3a head 3b antenna) 4 *E. bipectinalis* sp. n., venation, slide No. RYD04718w 5 *E. furvivena* sp. n., paratype, male (5a head 5b antenna) 6 *E. furvivena* sp. n., venation, slide No. RYD04529w. Scale bars: 2.0 mm.
Distribution. China (Guangdong, Guangxi, Hainan, Taiwan, Yunnan), Japan, India, Sri Lanka, Sikkim, Israel, Mediterranean, Central Europe, Norway, United Kingdom, North Africa, Australia, Argentina, United States, Puerto Rico, Jamaica.

**Ectomyelois bipectinalis** sp. n.
http://zoobank.org/BF7291BA-0081-45D5-B1F4-6980247E5C59
Figs 3, 4, 8, 11

Type material. **Holotype** ♂, CHINA: Guanping (22°14’N, 100°53’E), Xishuangbanna, Yunnan, 1200 m, 19-VII-2005, coll. Yingdang Ren. **Paratypes:** Fujian: 2 ♀♀, Mt. Tianzhu (24°35’N, 117°55’E), 220 m, 8,14-IX-2010, coll. Yinghui Sun & Jing Zhang. Gansu: 1 ♂, Fanba (32°44’N, 105°07’E), Wenzian, 718 m, 18-VII-2005, coll. Haili Yu, gen. slide no. RYD04745. **Guangxi:** 1 ♂, Yachang Yard, Leye County (24°47’N, 106°33’E), 665 m, 24-VII-2004, coll. Jiasheng Xu; 1 ♂, Mt. Yuanbao (25°14’N, 109°07’E), 500 m, 10-VIII-2006, coll. Weichun Li, gen. slide no. LKY10112; 1 ♂, Shaoping Yard, Pingxiang (22°03’N, 106°55’E), 190 m, 24,28-VII-2011, coll. Bingbing Hu; 1 ♀, Qingshan Yard, Pingxiang (22°03’N, 106°55’E), 300 m, 20-VII-2011, coll. Bingbing Hu. **Hainan:** 1 ♂, Mt. Diaoluo (18°39’N, 109°54’E), 70 m, 27-V-2007, coll. Zhiwei Zhang & Weichun Li, gen. slide no. LJY10104; 1 ♀, Mt. Duowen (19°48’N, 109°45’E), 120 m, 2-V-2009, coll. Qin Jin & Bingbing Hu, gen. slide no. LJY10105; 1 ♀, Wuzhishan (18°46’N, 109°30’E), 740 m, 14-IV-2009, coll. Qin Jin & Bingbing Hu, gen. slide no. LJY10097; 1 ♂, Shuimanxiang (18°53’N, 109°40’E), Wuzhishan, 620 m, 19-IV-2014, coll. Tengteng Liu, Wei Guan & Xue mei Hu; 1 ♂, Wuzhishan (18°40’N, 109°29’E), 500 m, 12-IV-2013, coll. Yingdang Ren & Xiaoguang Liu, gen. slide nos. RYD2013046 (deposited in HAASM). **Yunnan:** 4 ♂♂, Rare Botanical Garden, Ruili (24°00’N, 97°50’E), 1000 m, 5-8-VII-2005, leg. Yingdang Ren, gen. slide nos. RYD04718, RYD04718w; 1 ♂, Guanping (22°15’N, 100°53’E), Xishuangbanna, 1200 m, 17-VIII-2005, coll. Yingdang Ren, gen. slide no. RYD04717; 1 ♂, Botanical Garden, Menglu (21°52’N, 101°18’E), 570 m, 13-VIII-2005, coll. Yingdang Ren, gen. slide no. RYD04721; 4 ♂♂, Bubang (21°36’N, 101°35’E), Xisuiangbanna, 650 m, 22-24-VIII-2005, coll. Yingdang Ren, gen. slide nos. LHX14081, LHX14081w; 1 ♂, Botanical Garden (21°55’N 101°16’E), Xishuangbanna, 560 m, 1-VIII-2010, coll. Yinghui Sun & Lixia Li; 2 ♂♂, Bakaxiaozhai (21°58’N, 101°12’E), Mengla, Xisuiangban na, 630 m, 7-VIII-2010, coll. Yinghui Sun & Lixia Li; 7 ♂♂, 1 ♀, Mengyuan (21°42’N, 101°23’E), Mengla, Xishuangbanna, 640 m, 10-13-VIII-2010, coll. Yinghui Sun & Lixia Li, gen. slide nos. LJY10351, LJY10352; 20 ♂♂, 16 ♀♀, Bubang (21°36’N, 101°35’E), Mengla, 650 m, 12-14-VII-2013, coll. Linlin Yang, gen. slide nos. RYD20120106m, RYD20120107m, RYD20120167f, RYD20120168m (deposited in HAASM).

**Diagnosis.** This new species is notable superficially for its bipectinate male flagellum. It is much more similar to *E. ceratoniae* in genitalic structures, but can be distin-
Figures 7–9. Male genitalia of *Ectomyelois* spp. 7 *E. ceratoniae*, slide No. LJJ10595 8 *E. bipectinalis* sp. n., paratype, slide No. RYD04718 9 *E. furvivena* sp. n., paratype, slide No. LHX14084. Scale bars: 0.5 mm.

guished from the latter by the narrower uncus with width almost equaling length, the widest part is at basal 2/5, the apical projection of gnathos approximately 3/5 length of uncus in male, and the elongate corpus bursae three times as long as wide in female. In *E. ceratoniae*, the uncus is more wide than long, the widest part at base, the apical projection of gnathos nearly the same length as uncus in male, and the corpus bursae is twice as long as wide in female.
Description. Wingspan 19.5−28.0 mm (Figs 3, 4). Vertex brown, with individual scales tipped with grayish white. Antenna (Fig. 3a, b) brown; bipectinate, with pecten about twice length of width of shaft in male, shortly ciliate in female. Labial palpus brown, individual scales white-tipped. Maxillary palpus brown. Occiput, patagium, tegula and thorax brown. Forewing dark grayish fuscous, blackish brown along veins; antemedial line invisible; discal spots blackish brown, separated; postmedial line faint, grayish white, serrated; terminal line black, interrupted; cilia brown. Hindwing grayish white, pale brown along costa, termen and veins; cilia grayish white. Legs brown, mottled with grayish white, spurs grayish white. Abdomen yellowish brown.

Male genitalia (Fig. 8). Uncus bell-shaped, width almost equals length, rounded at apex, triangularly protruded laterally at basal 2/5, arched on basal margin. Apical projection of gnathos about 3/5 length of uncus, clubbed, slightly bent and furcated at apex. Transtilla trefoiled, including a pair of triangularly inflated bases, and a tongue-shaped central projection with rounded apex posteriorly. Valva three times as long as wide, evenly curved toward rounded apex, costal margin almost parallel with ventral margin except slightly convex at basal 2/3; costa strongly sclerotized, broad at base, slightly narrowed and extending to end of valva, without process apically; sacculus strongly sclerotized, stout, 2/5 length of valva. Juxta U-shaped, base an arched belt, with a pair of wide, stout, incurved lateral lobes, expanded and bearing sparse setae apically. Vinculum U-shaped, length almost equal to the widest posterior margin, slightly concave at middle of anterior margin. Phallus slightly shorter than valva, with membranous crimples internally; cornutus absent. Eighth tergite fan-shaped, 4/5 length than width, with a pair of spoon-like sclerites anteriorly; eighth sternite with a pair of triangular plates narrowly connected anteriorly. Culcita (sensu Amsel 1956) simple, one pair of fine scale tufts.

Female genitalia (Fig. 11). Anal papillae triangular, with a few setae, blunt apically. Eighth tergite slightly concave on posterior margin, trapezoidally convex on anterior margin; eighth sternite with membranous part inverse-funneled. Antrum twice as wide as length. Ductus bursae membranous, twice length of apophyses posteriores. Corpus bursae membranous, elongate, slightly shorter than ductus bursae, three times as long as wide; signum an elongate patch of microspines, placed at posterior 2/5. Ductus seminalis from junction of corpus bursae and ductus bursae.

Distribution. China (Fujian, Gansu, Guangxi, Hainan, Yunnan).

Etymology. The specific name is derived from the Latin prefix bi-, meaning two, and the Latin pectinalis, meaning pectinate, referring to the bipectinate male flagellum.

_Ectomyelois furvivena_ sp. n.
http://zoobank.org/72178A69-8258-4F83-A8C0-D1153B7E86CE
Figs. 5, 6, 9, 12

Type material. Holotype ♂, CHINA: Rare Botanical Garden, Ruili (24°00’N, 97°50’E), Yunnan, 1000 m, 8-VII-2005, leg. Yingdang Ren, gen. slide no. RYD04737.
Figures 10−12. Female genitalia of Ectomyelois spp. 10 E. ceratoniae, slide No. RYD04658 11 E. bipectinalis sp. n., paratype, slide No. IJY10107 12 E. furvivena sp. n., paratype, slide No. RYD04744. Scale bars: 0.5 mm.

Paratypes: Gansu: 1 ♂, Fanba (32°44′N, 105°07′E), Wenxian, 718 m, 18-VII-2005, coll. Haili Yu, gen. slide no. RYD04744. Yunnan: 2 ♀♀, same data as for holotype, gen. slide nos. LHX14084, LHX14084w, LHX14085; 1 ♀, Botanical Garden, Menglun (21°52′N, 101°18′E), 570 m, 13-VIII-2005, coll. Yingdang Ren, gen. slide no. RYD04720; 1 ♂, Baihualing, Mt. Gaoligong (25°31′N, 98°32′E), 1470 m, 30-VII-2013, coll. Linlin Yang, gen. slide no. RYD20120181 (deposited in HAASM).
Diagnosis. This new species is similar to *E. bipectinalis* sp. n., but can be recognized by the male antenna is not bipectinate, the uncus is rather abruptly narrowed beyond its broad base, tapered apical projection of the gnathos is about half length of the uncus and the inverse-goblet transtilla in the male genitalia. In *E. bipectinalis* sp. n., the antenna is bipectinate, the uncus protrudes triangularly at basal 2/5, the apical projection of gnathos is about 3/5 length of the uncus and the transtilla is trefoiled in the male genitalia. There is little difference in the female genitalia except the corpus bursae is much broader and the signum is smaller than in *E. bipectinalis* sp. n.

Description. Wingspan 25.0–30.0 mm (Figs 5, 6). Vertex brown, with individual scales grayish white-tipped. Antenna (Fig. 5a, b) brown, scales dark-tipped. Labial palpus brown, first segment with scales grayish white-tipped. Maxillary palpus brown. Occiput, patagium, tegula and thorax grayish brown, with scales tipped with grayish white. Forewing dark grayish brown with some white powdering, black along veins; antemedial line invisible; discal spots blackish brown, separated; postmedial line faint, grayish white, serrated, gently curved inwardly from costal 1/5 to dorsum 1/5; terminal line black, interrupted; cilia brown. Hindwing grayish white, light brown along costa and veins; cilia white. Foreleg blackish brown; mid- and hind legs brown with grayish white powdering, spurs yellowish brown. Abdomen with each tergite gray basally and grayish white distally, sternite yellowish brown.

Male genitalia (Fig. 9). Uncus bell-shaped, length longer than wide, abruptly narrowed beyond its broad base, rounded at apex. Apical projection of gnathos about half length of uncus, tapered, slightly bent and furcated at apex. Transtilla inverse-goblet shaped; deeply concaved in U shape on anterior margin, a rounded plate protruding on posterior margin. Valva three times as long as wide, evenly curved toward rounded apex, costal margin almost parallel with ventral margin, ventral margin concave at basal 1/3; costa strongly sclerotized, broad at base, narrowed and extending to near end of valva, without process apically; sacculus strongly sclerotized, stout, 2/5 length of valva. Juxta a broad, quadrate plate; lateral lobes ovate, 1.5 times as long as wide, bearing sparse setae in distal half. Vinculum trapezoid, widest posterior margin about 1.6 times of its length, straight on anterior margin. Phallus about 2/3 length of valva, smooth inside; cornutus absent. Eighth tergite cupped, with a pair of triangular sclerites anteriorly; eighth sternite with a pair of boot-like sclerites narrowly connected anteriorly. Culcita simple, one pair of fine scale tufts.

Female genitalia (Fig. 12). Anal papillae triangular, with a few setae, blunt apically. Eighth tergite slightly concave on posterior margin, convex W-shaped on anterior margin; eighth sternite with membranous part inverse-funneled. Antrum somewhat quadrate. Ductus bursae membranous, 1.5 times length of apophyses posteriores. Corpus bursae membranous, about same length as ductus bursae, twice as wide; signum a spindle-like patch of scobinations, at middle of corpus bursae. Ductus seminalis from junction of corpus bursae and ductus bursae.

Distribution. China (Gansu, Yunnan).

Etymology. The specific name is derived from the Latin prefix *furv-*-, meaning black, and the Latin *vena*, vein, referring to the forewing with black scales along its veins in this species.
Discussion

The genus *Ectomyelois* is characterized by the wing venation, a signum with a patch of microspines and the inception of the ductus seminalis in the female genitalia. Two new species are assigned to this genus based on these characters. *Ectomyelois bipectinalis* sp. n. is unique for its bipectinate male flagellum, but the other characters, especially the genitalia, are in accord with the generic characters.

Neunzig and Goodson (1992) described one new species, *Ectomyelois austrella* Neunzig & Goodson, 1992, from Argentina. However, the male genitalia has a basal protuberance on the valva not found in *Ectomyelois* species, and the female genitalia bears a narrowly and deeply invaginated signum on the corpus bursae also not found in *Ectomyelois* species. Although we retain this species in *Ectomyelois*, the characters indicate that *austrella* might not be suitably placed in this genus.

Acknowledgments

We express our cordial thanks to Prof. H.H. Li and Prof. S.X. Wang (Nankai University, Tianjin) for providing precious references and specimens, to those who participated in the field collection for their hard work, to Dr. H.X. Liu (Kaili University, Guizhou) and Dr. J.Y. Liu (Guiyang Meical University, Guizhou) for dissecting some specimens. We also sincerely thank anonymous referees for their useful comments and suggestions. This study was supported by the National Natural Science Foundation of China (No. 31172141 and No. 31093430) and partly funded by the Basic Scientific research project of Henan Academy of Agricultural Sciences (No. 2015JC19).

References

Amsel HG (1956) Microlepidoptera Venezolana I. Boletin de Entomologia Venezolana, Maracay 10(1954) (1–2): 1–336.
Balinsky BI (1994) A study of African Phycitinae in the Transvaal Museum. Johannesburg, South Africa, 208 pp.
Durrant JH (1915) *Myelois neophanes* sp. n., an addition to the British list. The Entomologist’s Monthly Magazine 51: 302–303.
Dyar HG (1911) Two species of Phycitinae new to our fauna. Proceedings of the Entomological Society of Washington 13: 30.
Dyar HG (1914) Report on the Lepidoptera of the Smithsonian Biological Survey of the Panama Canal Zone. Proceedings of the United States National Museum 47(2050): 139–350. doi: 10.5479/si.00963801.47-2050.139
Fischer von Röslerstamm JE (1834–1843) Abbildungen zur Berichtigung und Ergänzung der Schmetterlingskunde, Besonders der Microlepidopterologie als Supplement zu Treitschke’s und Hübner’s europeischen Schmetterlingen, mit erläuterndem Text. Hinrichs, Leipzig, 304 pp.
Goater B (1986) British Pyralid Moths. A Guide to their Identification. Harley Books, Colchester, 175 pp.
Hampson GF (1896) Moths. The Fauna of British India, Including Ceylon and Burma (London) 4: 1–594.
Hampson GF (1903) The moths of India. Supplementary paper to the volumes in “The fauna of British India.” Series II. Part IX, X. The Journal of the Bombay Natural History Society 15: 19–37 [part IX], 206–226 [part X].
Heinrich C (1956) American moths of the subfamily Phycitinae. Bulletin of the United States National Museum (Washington, DC) 207: 1–581.
Heppner JB, Inoue H (1992) Checklist. Lepidoptera of Taiwan. Scientific Publishers, Gainesville, Florida 1(2): 1–276.
Inoue H, Sugi S, Kuroko H, Moriuti S, Kawabe A, Owada M (Eds) (1982) Moths of Japan. Kodansha, Tokyo, 968 pp. [Vol. 1], 556 pp. [Vol. 2].
Leraut PJA (2002) Contribution à l’étude des Phycitinae (Lepidoptera, Pyralidae). Nouvelle Revue d’Entomologie (Paris) 19(2): 141–177.
Lucas D (1950) Contribution à l'étude des Lépidoptères Nord-Africains. Bulletin de la Société entomologique de France (Paris) 55(1): 141–144.
Mansbridge W (1907) Notes and Observations. Entomologist 40: 8–11.
Munroe EG (1983) Pyralidae (except Crambinae). In: Hodges RW (Ed.) Check List of the Lepidoptera of America north of Mexico including Greenland. EW Classey & Wedge Entomological Research Foundation, London, 67–76, 78–85.
Neunzig HH (1979) Systematics of immature Phycitines (Lepidoptera: Pyralidae) associated with leguminous plants in the southern United States. United States Department of Agriculture Technical Bulletin 1589: 1–119.
Neunzig HH (1990) Pyraloidea: Pyralidae: Phycitinae (part). In: Dominick RB (Ed.) The Moths of America North of Mexico including Greenland (15.3). The Wedge Entomological Research Foundation, Washington, DC, 165 pp.
Neunzig HH, Goodson RL (1992) New genera and species of southern South American Phycitinae (Lepidoptera: Pyralidae). Proceedings of the Entomological Society of Washington 94(2): 189–222.
Palm E (1986) Nordeuropas Pyralider - med saerligt henblik paa den danske fauna (Lepidoptera: Pyralidae). In: Lyneborg L (Ed.) Danmarks Dyreliv. Fauna Bøger 33. National-Trykkeriet, Copenhagen, Denmark, 1–287.
Ragonot EL (1888) Nouveaux genres et espèces de Phycitidae & Galleriidae. Publié par l’auteur, Paris, 1–52.
Roesler RU (1968) Das neue systematische Verzeichnis der deutschen Phycitinae (Lepidoptera, Pyralidae). Nachrichtenblatt der Bayerischen Entomologen, München 17(1–2): 1–9, 25–28.
Roesler RU (1973) Phycitinae. Trifine Acrobasiina. In: Amsel HG, Gregor F, Reisser H (Eds) Microlepidoptera Palaearctica 4(1–2). Georg Fromme & Co., Wien, 752 pp. [part 1], 137 [part 2].
Roesler RU (1983) Die Phycitinae von Sumatra (Lepidoptera: Pyralidae). Heterocera Sumatrana, Keltern 3: 1–136.
Roesler RU, Küppers PV (1981) Beiträge zur Kenntnis der Insektenfauna Sumatras. Teil 9. Die Phycitinae (Lepidoptera: Pyralidae) von Sumatra; Taxonomie Teil B, Ökologie und Geobiologie. Beiträge zur Naturkundlichen Forschung in Südwestdeutschland, Karlsruhe Beih. 4: 1–282, 4 Text Abbildung, 42 Tafel.

Sinev SJu (1986) Phycitidae. In: Medwedjewa GS (Ed.) Opredelitel Nasekomych Evropejskoj Tschasti SSSR [Russ] 4 (3). Nauka, Leningrad, 251–340.

Sorhagen L (1881) Zwei neue Pyraliden. Berliner Entomologische Zeitschrift, Berlin 25(1–2): 103–104.

Vaughan H (1870) Descriptions of three species of Phycidae (from Britain) new to science. The Entomologist’s Monthly Magazine, London 7: 130–132, 160.

Warren W, Rothschild LW (1905) Lepidoptera from the Sudan. Novitates Zoologicae, London 12: 21–33.

Yamanaka H (2013) The Standard of Moths in Japan IV. Gakken Education Publishing, Tokyo, Japan, 551 pp.

Zeller PC (1839) Versuch einer naturgemäßen Eintheilung der Schaben. Isis von Oken, Leipzig, [32](3): 167–219.

Zeller PC (1881) Columbische Chiloniden, Crambiden und Phycideen. Horae Societatis entomologicae Rossicae, St. Petersbourg 16: 154–256.