In Vitro Testing of Salicylanilide Derivatives Against Some Fungal and Bacterial Strains

IOANA MARIA CARMEN IENASCU1-2, DIANA OBISTIOIU1, IULIANA MARIA POPESCU1, MARIANA NELA STEFANUT3, OSSER GYONGYI3, CLAUDIA JURCA3, GABRIELA CIAVOI3, EDWIN SEVER BECHIR4*, FARAH BECHIR4, ADINA CATA4

1 National Institute of Research and Development for Electrochemistry and Condensed Matter, 144 Dr. A. P. Podeanu, 300569, Timișoara, Romania
2 Vasile Goldis Western University of Arad, Faculty of Pharmacy, Department of Pharmaceutical Sciences, 310045, Arad, Romania
3 Banat's Agricultural Science University, Faculty of Agriculture, Interdisciplinary Research Platform, 300645, Timișoara, Romania
4 University of Oradea, Faculty of Medicine and Pharmacy, 410073 Oradea

In the dental office, disinfection and sterilization are a matter of utmost importance for both the physician, the medical staff and the patient. Twelve N-(2-bromo-phenyl)-2-hydroxy-benzamide and N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives were synthesized and tested for antimicrobial activity against 6 bacterial and 2 fungal strains using the Disk diffusion method for susceptibility testing. The obtained results indicated that the N-(2-bromo-phenyl)-2-hydroxy-benzamide derivatives were more active against the tested microbes, inhibition zones of 6-12 mm being obtained, although the most effective compound against S. pyogenes proved to be N-(4-bromo-phenyl)-2-hydroxy-benzamide (14 mm inhibition zone). The tested compounds presented no effect against S. aureus, S. flexneri, S. typhimurium and C. parapsilosis at the tested concentration.

Key words: N-(2-bromo-phenyl)-2-hydroxy-benzamide derivatives, N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives, antibacterial effect, antifungal effect, disk diffusion method

In the last decades, a major intensification in the prevalence of resistance to antibacterial and antifungal agents has been observed. Antibiotic resistance occurs when a drug fails the capacity to inhibit bacterial growth efficiently. Bacteria become resistant and continue to proliferate in the presence of therapeutic concentration of the antibiotics. Prescription of broad spectrum antibiotics in hospitals to treat nosocomial infections increases antibiotic resistance [1].

The implications of the appearance of resistance to current antimicrobial agents would be the following: morbidity, mortality and high health care costs both in hospitals and in the community. Therefore, the researchers have been focused on a better understanding of the mechanisms of antimicrobial resistance, developed methods to detect resistance, new antimicrobial agents for such resistant microorganisms infections treatment, and approaches to prevent the occurrence and spread of resistance in the first place [2]. So, the development of effective and non-toxic antimicrobial agents is still a vital issue due to the increase of multi-resistant bacterial strains [3-7]. The most affected patients are those with compromised immune systems or with other associated diseases [8-14].

2-Hydroxybenzamide derivatives are a class of compounds with a large spectrum of biological activity, from which, 2-hydroxybenzanilides, as well as O-substituted 2-hydroxybenzanilides proved to be potent antimicrobial agents [15,16]. Among the currently used antibacterial agents, nitrofurazone, furazolidone and nitrofurantoin are known to contain hydrazide-hydrazine moiety [17-20]. Due to this fact, hydrazide-hydrazine derivatives represent a good source to develop new antibacterial agents.

Development of new compounds by attaching 2-hydroxybenzamides to another halogen-substituted aromatic nucleus conducted to the enhancement of the antibacterial, antiviral, anti-inflammatory and analgesic effects of these compounds. Salicylamide-O-acetic hydrazide and hydrazones displayed antiinflammatory and analgesic effect superior to salicylamide and, at the same time, inferior ulcerogenic effect [21,22].

Novel 2-hydroxy-benzamide derivatives, esters, hydrazide and hydrazones were designed [23-27] and investigated for their biological activity [28, 29] in our laboratory. Antimicrobial activity of salicylanilide derivatives, esters, hydrazides and hydrazones, was evaluated in vitro against some bacteria and fungi. The minimum inhibitory concentrations obtained for esters were generally comparable to those for salicylanilide, but the hydrazides and hydrazones proved to possess superior antibacterial and antifungal effect compared to the control substance [29].

The aim of this work was to synthetize some novel 2-hydroxy-benzamide derivatives and to evaluate their biological activity. The relationship between the antimicrobial activity and the structure of these synthesized compounds was also established.

Experimental part

Chemicals and strains

The reagents used for the synthesis are: 2-bromoaniline, 4-bromoaniline, phosphorus trichloride (Acros Organics, for synthesis); salicylic acid, ethyl chloroacetate, methyl...
chloroacetate (Sigma-Aldrich, for synthesis); hydrazine monohydrate, 4-dimethylaminobenzaldehyde, 5-bromosalicylaldehyde (Merck, for synthesis). Solvents, chlorobenzene (Scharlau, analytical purity), absolute ethanol, dimethylformamide, 2-butanone (Merck, analytical purity), were used as supplied.

For microbiological tests, Petri plates were used, containing microcomprimates (comercial dishes with Gentamicin (10 mg-Ref E110712, BioMaxima, Polonia) and Nystatin (100 mg-Ref SD 025, Himedia, India). Sabouraud Dextrose Agar (Oxoid, CM0041), Sheep Blood Agar Base and Brain Heart Infusion Agar (Oxoid, CM1136) were used as culture media. Bacterial strains, S. aureus (ATCC 25923), E. coli (ATCC 25922), P. aeruginosa (ATCC 27853), S. pyogenes (ATCC 19615), S. flexneri (ATCC 12022), S. typhimurium (ATCC 14028) and fungal strains, C. albicans (ATCC 10231), C. parapsilopsis (ATCC 22019) were used.

Conventional heating synthesis

Anilide - To a mixture of salicylic acid, 2-bromoaniline and phosphorus trichloride 1:1:0.3, chlorobenzene was added. The mixture was stirred and heated to reflux for 5 h. After being cooled at room temperature, the obtained, crude product was filtered off, washed three times successively with hot water and then three times successively with 10% sodium carbonate solution. The solid product was then filtered off, dried and recrystallized from dimethylformamide.

Ethyl/Methyl esters were added dropwise to a mixture of appropriate salicylamide and anhydrous K₂CO₃, refluxed in 2-butanone, ethyl/methylchloro-acetate. The Optimum molar ratio was amide:ester:K₂CO₃ = 1:1:1. The mixture was stirred and heated to reflux for 5 h. After cooling at room temperature, the mixture was poured into water and shaken intensively. The organic phase was dried over MgSO₄. After filtration and evaporation of solvent in vacuum, the esters were obtained in crystalline form and were recrystallized from absolute ethanol.

Hydrazide - A mixture of ethyl ester and hydrazine hydrate 1:1 was refluxed in ethanol for 3 h. The reaction mixture was cooled, the separated solid filtered off, then subjected to recrystallization from absolute ethanol.

Hydrazones - were added to a solution of hydrazide in ethanol, the appropriate benzaldehyde was 1:1. The reaction mixture was refluxed for 5 h. The solid obtained after cooling was filtered off, washed with water and recrystallized from dimethylformamide.

Antimicrobial activity evaluation

Antimicrobial activity of the 2-hydroxy-benzamide derivatives was tested against 6 bacterial strains, and 2 fungal strains, using the Disk diffusion method for susceptibility testing, according to the Standard Rules for Antimicrobial Susceptibility Testing using Impregnated Disks [30,31]. In vitro testing was performed in plates, containing microcomprimates with Nystatin for the antifungal activity and Gentamicin for the antimicrobial activity as positives controls, alongside blank filter papers impregnated with DMSO as negatives controls and filter papers impregnated with 10 µL of 20 g/L stock solution of each compound. A 10⁻² dilution of the fresh Candida cultures and a 10⁻² fresh bacteria culture was used to perform the assay, an inoculum equivalent to a 0.5 McFarland standard. The Petri plates so seeded and the respective specimens containing the tested compounds were incubated at 30 °C for Candida species and 37 °C in case of the bacterial strains, for 24-48 h. Tests were performed in triplicate. Finally, the interpretation of the result, the ratio of the antymycotic and antibacterial effect of the tested compounds, was achieved by measuring the diameter of the analyzed culture inhibition zones (including the diameter of the disc – 5mm) in millimeters. The results are presented as the average of two determinations and a percentage representation of the efficacy of the compounds in relation to the effectiveness of the positive control.

Results and discussions

Twelve N-(2-bromo-phenyl)-2-hydroxy-benzamide and N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives were synthesized, except anilides, the rest of the compounds were synthesized by our team, probably for the first time, were obtained in good yields, between 72-92%. The synthesis pathway is presented in figure 1. The compounds were characterized using modern analytical methods, IR, NMR, elemental analysis, some of the data has already been published [28]. The synthesized compounds, yields and melting points are presented in table 1.

Table 2 presents the diameters of inhibition in mm and the compound efficacy expressed in percentages. The results of the microbiological tests indicated that the N-(2-bromo-phenyl)-2-hydroxy-benzamide derivatives were more active against the tested microbes compared to N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives, inhibition zones of 6-12 mm being obtained.
The most susceptible microorganisms to the action of the N-(2-bromo-phenyl)-2-hydroxy-benzamide derivatives are Candida albicans and Streptococcus pyogenes. Anilide, ethyl ester and hydrazide inhibited both microbes, moreover, C. albicans is also inhibited by hydrazide. The most active compound against Escherichia coli and Pseudomonas aeruginosa is hydrazide which exhibited a percentage of efficacy in relation to the effectiveness of Gentamicin of 57.7% and 46.7, respectively. The highest efficacy (57.1%) against Streptococcus pyogenes was exhibited by ethyl ester, and in case of Candida albicans, the most active compounds were anilide and ethyl ester (36.9%).

Among N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives, N-(4-bromo-phenyl)-2-hydroxy-benzamide itself is the only one which exhibited antimicrobial activity. This compound is active only against Pseudomonas aeruginosa and Streptococcus pyogenes, with a percentage of efficacy in relation to the effectiveness of Gentamicin as positive control of 40 and 66.7%, respectively, being the most effective compound (14 mm inhibition zone). However, anilide presented no effect against fungal strains. So, the 4-substitution of aniline ring with bromine conducted to zero efficiency in terms of antifungal activity. Moreover, the substitution of phenolic hydroxyl, apparently, brings also no benefit.

The tested compounds presented no effect against S. aureus, S. flexneri, S. typhimurium and C. parapsilopsis at the tested concentration. C. albicans and S. pyogenes were very susceptible to the action of the N-(2-bromo-phenyl)-2-hydroxy-benzamide derivatives. The most active compound against E. coli and P. aeruginosa is hydrazide. Anilide exhibited the highest efficacy against Streptococcus pyogenes. The most potent compounds

---

**Table 1**

| No. | Compound name | Molecular formula / weight | Yield (%) | Melting point (°C) |
|-----|---------------|---------------------------|-----------|------------------|
| 1.  | N-(2-bromo-phenyl)-2-hydroxy-benzamide | C₁₆H₁₂BrN₂O₂ | 72 | 158-161 |
| 2.  | [2-(2-bromo-phenylcarbamoyl)-phenoxyl]-acetic acid ethyl ester | C₁₆H₁₂BrN₂O₄ | 81 | 87-88 |
| 3.  | [2-(2-bromo-phenylcarbamoyl)-phenoxyl]-acetic acid methyl ester | C₁₆H₁₂BrN₂O₄ | 80 | 119-120 |
| 4.  | N-(2-bromo-phenyl)-2-hydrazinocarbonylmethoxy-benzamide | C₁₆H₁₂BrN₂O₄ | 87 | 156-157 |
| 5.  | 2-(2-hydroxy-phenyl)-2-hydroxybenzamide | C₁₆H₁₂BrN₂O₄ | 85 | 272-273 |
| 6.  | N-(2-bromo-phenyl)-2-(4-dimethylamino-phenylcarbamoyl)-2-hydrazinocarbonylmethoxy-benzamide | C₁₆H₁₂BrN₂O₄ | 60 | 220-221 |
| 7.  | N-(4-bromo-phenyl)-2-hydroxy-benzamide | C₁₆H₁₂BrN₂O₂ | 75 | 169-172 |
| 8.  | [2-(4-bromo-phenylcarbamoyl)-phenoxyl]-acetic acid ethyl ester | C₁₆H₁₂BrN₂O₄ | 84 | 168-170 |
| 9.  | [2-(4-bromo-phenylcarbamoyl)-phenoxyl]-acetic acid methyl ester | C₁₆H₁₂BrN₂O₄ | 81 | 158-160 |
| 10. | N-(4-bromo-phenyl)-2-hydrazinocarbonylmethoxy-benzamide | C₁₆H₁₂BrN₂O₄ | 88 | 187-188 |
| 11. | 2-(2-hydroxy-phenyl)-2-hydroxybenzamide-N-(4-bromo-phenyl)-benzamide | C₁₆H₁₂BrN₂O₄ | 87 | 276-279 |
| 12. | N-(4-bromo-phenyl)-2-(4-dimethylamino-phenylcarbamoyl)-2-hydrazinocarbonylmethoxy-benzamide | C₁₆H₁₂BrN₂O₄ | 92 | 271-274 |

**Table 2**

| Tested compounds | Diameter of inhibition (mm) | Compound efficacy (%) |
|------------------|----------------------------|-----------------------|
|                  | S. a. | E. c. | P. a. | S. f. | S. p. | S. i. | C. p. | C. a. |
| 1.                |       |       |       | 8/40  |       | 8/38  |       |       |
| 2.                |       |       |       | 7/30  |       | 12/57.1|       |       |
| 3.                |       |       |       |       |       |       |       |       |
| 4.                |       |       |       | 7/57/7| 7/46.1| 10/35.1| 6/30.8| 6/30.8|
| 5.                |       |       |       |       |       |       |       |       |
| 6.                |       |       |       |       |       |       |       |       |
| 7.                |       |       |       | 6/40  |       | 11/65.7|       |       |
| 8.                |       |       |       |       |       |       |       |       |
| 9.                |       |       |       |       |       |       |       |       |
| 10.               |       |       |       |       |       |       |       |       |
| 11.               |       |       |       |       |       |       |       |       |
| 12.               |       |       |       |       |       |       |       |       |
| Gentamicin        | 14    | 13    | 15    | 13    | 21    | 15    |       |       |
| Nystatin          |       |       |       |       |       |       |       | 16/19.5|

Abreviations: S. a. - Staphylococcus aureus; E. c. - Escherichia coli; P. a. - Pseudomonas aeruginosa; S. f. - Shigella flexneri; S. p. - Streptococcus pyogenes; S. t. - Salmonella typhimurium; C. p. - Candida parapsilosis; C. a. - Candida albicans.
against Candida albicans were anilide and ethyl ester. The low efficiency of some of the tested compounds may be due to the low solubility of the compounds which act like a barrier against diffusion in the culture medium or due to the tested concentration. Thus, filter papers were impregnated with 10 µL of 20 g/L stock solution of compounds, meaning that the tested concentrations were 50 folds lower than gentamicin and 500 folds lower than nystatin.

Interactions between oral microbes and dental materials may also occur, although today not very numerous information are available regarding this possibility [32].

Under current circumstances, the issue of safety in terms of sterilization and disinfection in the dental office is of utmost importance, wherefore the study of substances used in disinfection and sterilization should be a constant concern for the specialists in the field.

Conclusions

Twelve N-(2-bromo-phenyl)-2-hydroxy-benzamide derivatives and N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives, anilides, esters, hydrazides and hydrazones, were obtained in higher yields and purity using conventional heating synthesis. Except anilides, the rest of compounds are new. The antimicrobial activity of the compounds was assessed against 6 bacterial and 2 fungal strains using the disk diffusion method. The N-(2-bromo-phenyl)-2-hydroxy-benzamide derivatives were more active against the tested microorganisms, inhibition zones of 6-12 mm being obtained.

Among N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives, only anilide exhibited antimicrobial effect, being active against Pseudomonas aeruginosa and Streptococcus pyogenes; no effect against fungal strains was obtained. In case of N-(4-bromo-phenyl)-2-hydroxy-benzamide derivatives, the substitution of phenolic hydroxyl, apparently, is not favorable for antimicrobial effect, but this may also be due to the low solubility/diffusion of the compounds in the culture medium or due to the tested concentration. These results demonstrated once more the great potential of 2-hydroxy-benzamide derivatives as antimicrobial agents.

References

1.ZAMAN, S. B., HUSSAIN, M. A., NYE, R., MEHTA, V., MAMUN, K. T., HOSSAIN, N. Cureus., 9, no. 6, 2017, e1403.
2.DIACONU, C.C., DRAGOI, C.M., BRATU, O.G., NEAGU, T.P., STOIAN, A.P., COBELSCHI, P.C., NICOLAE, A.C., IANCU, M.A., HAINAROSIE, R., STANESCU, A.M.A., SOCEA, B., Farmacia, 66, no. 3, p. 408.
3.MOELLERING, J. R. C., Int. J. Antimicrob. Agents, 37. 2011, p. 2.
4.PALLAG, A., JURCA, T., SIRBU, V., HONIGES, A., JURCA, C., Rev. Chim. (Bucharest), 69, no. 2, 2018, p. 445.
5.PALLAG, A., FILIP, G.A., OLTEANU, D., CLICHIICI, S., BALDEA, I., JURCA, T., MICLE, O., VICAS, L., MARIAN, E., SORIUA, O., CENARIU, M., MURESAN, M., Oxid. Med. Cell. Longev., 2018, ID 3060525, 14 pages, https://doi.org/10.1155/2018/3060525.
6.SPINU, D., BRATU, O., MARCU, D., MISCHIANU, D., HUICA, R., SURCEL, M., MUNTEANU, A., SOCEA, B., BODEAN, O., URSACIU, C., Rev. Chim. (Bucharest), 69, no. 3, 2018, p. 645.
7.MEHEDITIU, C., ANTONOVICI, M., BRINDUSE, L., BRATILA, E., STANCULESCU, R., BERCEANU, C., BRATU, O., PITIRU, S., ONOFRESCU, M., MATASARIU, D.R., Rev. Chim. (Bucharest), 69, no. 3, 2018, p. 581.
8.COATES, A., HU, Y., BAX, R., PAGE, C., Nat. Rev. Drug Discov., 1, 2002, p. 895.
9.TIT, D.M., PALLAG, A., IOVAN, C., FURAU, G., FURAU, C., BUNGUA, S., Iran. J. Public Health, 46, no. 11, 2017, p. 1128.
10.PALLAG, A., ROSCA, E., TIT, D.M., MUTIU, G., BUNGUA, S.G., POP, O.L., Rom. J. Morphol. Embriol., 56, no. 3, 2015, p. 1103.
11.MRAZ, C., MURESAN, M., MICLE, O., VICAS, L., PALLAG, A., COLTAU, M., PUSCAS, I., Farmacia, 60, no. 2, 2012, p. 264.
12.FODOR, K., TIT, D.M., PASCA, B., BUSTEA, C., UIVAROSAN, D., ENDRES, L., IOVAN, C., ABDEL-DAIM, M., BUNGUA, S., Oxid. Med. Cell. Longev., 2018, ID 4147320, 10 pages. https://doi.org/10.1155/2018/4147320.
13.TIT, D.M., BUNGUA, S., IOVAN, C., NISTOR CSEPENTO, D.C., ENDRES, L., SAVA, C., SABAU, A.M., FURAU, G., FURAU, C., JCM, 7, no. 10, 2018, p. 297. doi: 10.3390/jcm7100297
14.ABDEL-DAIM, M.M., ZAKHARY, N.I., ALEYA, L., BUNGUA, S.G., BOHARA, R.A., SIDDHIQI, N.J., Oxid. Med. Cell. Longev., 2018, ID 2098123, 2 pages. https://doi.org/10.1155/2018/2098123
15.MATYK, J., WAISSER, K., DRAZKOVA, K., KUNES, J., KLIMENSOVA, V., PALAT, K.R., KAUSTOVA, J., Farmaco., 60, no. 5, 2005, p. 399.
16.WAISSER, K., MATYK, J., DIVISOVA, H., HUSAKOVA, P., KUNES, J., KLIMENSOVA, V., KAUSTOVA, J., MOLLMANN, U., DAHSE, H.M., MIKO, M., Arch Pharm., 339, no. 11, 2006, p. 616.
17.MCCALLA, D.R., REUVERS, A., KAISER, C., J. Bacteriol., 190, 2017, p. 1126.
18.CHATTERJEE, S.N., GHOSH, S., Indian J. Biochem. Biophys., 16, no. 3, 1979, p. 125.
19.ALI, B.H., Vet. Res. Commun., 6, 1983, p. 1.
20.MUNDOZ-DAVILA, M.J., Antibiotics, 3, 2014, p. 39.
21.FAHMY, H.H., EL-ERAKY, W., Arch. Pharm. Res., 24, no. 3, 2001, p. 171.
22.FAHMY, H.H., SOLIMAN, G.A., Arch. Pharm. Res., 24, no. 3, 2001, p. 180.
23.IENASCU, I.M.C., LUPEA, A.X., POPESCU, I.M., PADURE, M.A., ZAMFIR, A.D., J. Serb. Chem. Soc., 74, no. 8-9, 2009, p. 847.
24.LUPEA, A.X., POPESCU, I., TARABASANU, C., IENASCU, I., PADURE, M., Rev. Roum. Chem., 51, no. 6, 2006, p. 517.
25.IENASCU, I.M.C., PADURE, M.A., LUPEA, A.X., BALCU, I., POPESCU, I.M., Rev. Chim. (Bucharest), 61, no. 4, 2010, p. 368.
26.IENASCU, I.M.C., LUPEA, A.X., POPESCU, I., TOMAS, S., ZAMFIR, A.D., Rev. Chim. (Bucharest), 59, no. 1, 2008, p. 56.
27.IENASCU, I.M.C., LUPEA, A.X., POPESCU, I., MOSOARCA, C., ZAMFIR, A.D., Rev. Chim. (Bucharest), 53, no. 4, 2008, p. 273.
28.IENASCU, I.M.C., BALAES, T., PETRE, C.V., POP, R.O., CATA, A., STEFANUT, M. N., ALBU, P., POENARU, M., Rev. Chim. (Bucharest), 69, no. 7, 2018, p. 1876.
29.IENASCU, I.M.C., LUPEA, A.X., HADARUGA, D., HADARUGA, N., POPESCU, I., Rev. Chim. (Bucharest), 59, no. 2, 2008, p. 247.
30.CAVALIERI, I., STEPHEN, J., Manual of Antimicrobial Susceptibility Testing, M.B. Coyle (Coord. Editor), American Society for Microbiology, 2005, p. 39.
31.SIRBU, V., PALLAG, A., HIENIGES, A., COJOCARU S.I., Rev. Chim. (Bucharest), 68, no. 2, 2017, p.390.
32.BECHIR, A., PACURAR, M., BECHIR, E.S., COMANEAU, R.M., CHIBBELEAN CIRES, M., MARIS, M., BARBU, H.B., Mat. Plast., 51, no. 1, 2014, p. 54-61.

Manuscript received: 21.10.2018