The representativeness of the dental calculus dietary record: insights from Taï chimpanzee faecal phytoliths

Robert C. Power 1,2 · Roman M. Wittig 3,4 · Jeffery R. Stone 5 · Kornelius Kupczik 2,6 · Ellen Schulz-Kornas 2,6

Received: 29 December 2019 / Accepted: 12 April 2021 / Published online: 29 May 2021
© The Author(s) 2021

Abstract
In recent years, new applications of microremain dietary analysis using dental calculus as a source of dietary data on ancient human subsistence and behaviours have accelerated. The dental calculus of contemporary human and non-human populations with known diets have been used as reference datasets, including the chimpanzees of Taï National Park (Côte d’Ivoire), but explaining the preservation mechanism involved is challenged by our incomplete knowledge of the microremain content within the diets of these reference populations and our rudimentary information on microremain incorporation into dental calculus. Here, we analyse phytoliths in faecal samples to assess to what extent plant phytoliths of a diet are reflected in the dental calculus as well as in the egested faeces. In this study, we identify and document the faecal phytolith assemblages as an indicator of plant consumption in two Western chimpanzees of the Taï National Park (Côte d’Ivoire) before (wet season), during (dry season) and after (dry season) a dust-rich period. Moreover, observational dietary records of these two individuals were compiled to improve the interpretability of this dental calculus phytolith dataset. The faecal phytolith assemblages vary significantly across samples in terms of abundance and diversity. The most common phytolith morphotypes were eudicot plates, single-cell and multi-cell tracheids, monocot rugulose and echinate spheroids and, to a lesser extent, unspecified thick and thin elongates. High loads of grit and other micro-remains (e.g. diatoms) are found during the dry period. Using observational dietary records as a starting point and our faecal results as a terminus, we consider how dental calculus can accumulate phytoliths. Our findings enable identification of the phytolith morphotypes that are under-represented in dental calculus, which is highly informative for future dental calculus research strategies.

Keywords Plants · Micro-remains · Rainforest · Ivory Coast · Taphonomy · Plant consumption

Introduction
In recent years, human dental calculus has received increased interest as a source of life history information. Researchers have sought to answer outstanding questions on the development and change of human subsistence using micro-remains and other dietary remains. An increasing amount of literature has examined phytolith applications in archaeological dental calculus (Deng et al. 2018; Dunseth et al. 2019; Wroth et al. 2019) and palaeoecological research (Chabot, Garnier, and Geria 2018; Criçò and Strömberg 2020; Delhon et al. 2003; Novello et al. 2018); however, researchers still largely do not understand the formation processes of dental calculus dietary records or even dental calculus itself (Radini et al. 2017). Given that only a small number of phytoliths and starches are present, and that molecular evidence of diet is generally sparse in dental calculus, it is accepted that dental calculus fails to provide a detailed picture of diet and is, at best, a
generalised picture. However, there are competing theories, and some argue that dental calculus mostly produces a random unreliable picture (Weyrich, Dobney, and Cooper 2015), while others believe it can provide valuable dietary insights (Leonard et al. 2015; Power et al. 2015). As the use of starch and phytoliths in dental calculus has accelerated, researchers have attempted to measure the reliability of starches and phytoliths in recording plant consumption and broader dietary patterns. Researchers have examined dental calculus from populations with well-documented diets, including living human forager-horticulturists from Namibia and skeletal wild chimpanzees from Côte d’Ivoire, with local reference plant collections to measure the ability of calculus to reflect diet in these two environments (Leonard et al. 2015; Power et al. 2015). These studies have set a benchmark on dental calculus’ dietary micro-remains coverage, but they also highlight discrepancies that exist between diet and dental calculus micro-remain assemblages. Some discrepancies are expected due to the high number of plant taxa that produce few or no diagnostic starches and phytoliths (Torrence and Barton 2006; Tsartsidou et al. 2007). Further variation is anticipated due to local evapotranspiration-driven differences in phytolith production (Jenkins, Janjoum, and Nuimat 2011). However, researchers have identified a disparity even when plants that do not make phytoliths and plants that make few phytoliths (low producers) are accounted for (Tsartsidou et al. 2007). Specific micro-remains from staple and non-staple foods have been found to be overrepresented, such as the starch grains of maize (a Twa staple) and the phytoliths of oil palm (a minor chimpanzee food) (Leonard et al. 2015; Power et al. 2015).

In some archaeological contexts, phytolith assemblages are mostly composed of digested phytoliths (Albert et al. 2008). Though phytoliths that pass through the gastrointestinal tract have been studied for their potential for reconstructing human behaviour (Gur-Arie et al. 2014; Phillips and Lancelotti 2014; Shahack-Gross 2011), few studies have examined the impact of this process on the assemblages. It is not clear how mammalian digestion affects phytoliths, despite the fact that phytoliths are frequently encountered damaged, and we need to understand the damage patterns that often affect them (Madella and Lancelotti 2012). The inconsistency between dietary records and dental calculus data in chimpanzees and humans indicates unexplained biases exist in dental calculus assemblages of starch grains and phytoliths (Leonard et al. 2015; Power et al. 2015). These undermine the validity of dietary interpretations of calculus data. These problems cannot be resolved simply by using reference plants of possible diets alone. One factor in this discrepancy that is characteristic of Taï chimpanzees is that they frequently absorb plant nutrients by chewing plant material into wadage without swallowing the food bolus. Presumably, this causes a reduction in exposure to phytoliths. Additionally, most chimpanzee feeding records do not specify the plant part consumed or give weights of consumed foods. Lastly, local factors can influence phytolith content in plants. Phytolith content can vary seasonally in response to moisture availability and evapotranspiration, meaning that no reference collection can replicate true chimpanzee phytolith exposure (Jenkins et al. 2011; Rosen and Weiner 1994).

Also, other micro-remains can be found in dental calculus, including the microscopic skeletons of algae and sponges (diatoms and sponge spicules), both of which are linked to aquatic environments. They have been interpreted as potentially offering an alternative method to reconstruct the use of water (Dudgeon and Tromp 2014). How these micro-remains enter dental calculus is understood poorly, because experimental studies are lacking.

To rectify these problems, we examined phytolith assemblages from faeces of Western chimpanzees of the Taï National Park (Côte d’Ivoire) (Schulz-Kornas et al. 2019) to document the phytolith types that are missing from the published chimpanzee dental calculus (Power et al. 2015) and possibly systematically underrepresented across dietary studies. This will generate the first complete model of a mammalian interface with the phytoliths in their natural environment. Chimpanzees of the Taï National Park have a highly diverse diet composed of hundreds of plant and other species and are an ideal model for the dietary reconstruction of historic human and fossil hominin populations (Boesch and Boesch-Achermann 2000).

Annually, Taï chimpanzees are exposed to high levels of airborne dust, which accumulates on plants eaten by these animals during dry periods. Between December and February, this Aeolian dust is transported by north-easterly surface winds (Harmattan) blowing from the Sahara Desert over West Africa into the Gulf of Guinea (Engelstaedter and Washington 2007; Stoorvogel, Van Breeemen, and Jassen 1997). The dust-laden Harmattan reaches the Taï National Park for a few weeks, depositing 60–100 kg/ha of uniformly graded materials in the canopy of the Taï forest ecosystem (Elipe and Lopez-Querol 2014; Stoorvogel et al. 1997; Wittig 2018). The dust particles have a size distribution of fine sand (50–125 µm), silt (2–50 µm) and clay (below 2 µm); approximately 69% of the dust typically is in the silt range (Enete et al. 2011).

Here, we present a unique case study where phytolith representation and feeding data of a well-known chimpanzee population are available (Boesch and Boesch-Achermann 2000; Falchi et al. 2014; Power et al. 2015). We aim to determine and quantify the phytolith, diatom and mineral particle abundance and morphotypes in the faecal samples and compare these results to published dental calculus phytolith data of the same chimpanzee population (Power et al. 2015). Additionally, we consider the effects of digestion on phytoliths. This allows us to use the faecal phytolith record to establish the usefulness of dental calculus assemblages.
Methods

Fourteen faecal samples were collected from two habituated male chimpanzees, the individuals Poseidon (Pos) and Athos (Ath), living in the East Group of the chimpanzee population of the Taï National Park in southwest Côte d’Ivoire (Wittig and Boesch 2019). This chimpanzee population has detailed behavioural and dietary records from 1992 until the present. The faecal samples were collected from the two target chimpanzees as part of the study on feeding ecology and chewing efficiency at intervals between December 2014 and March 2015 (Schulz-Kornas et al. 2019). Each sample was weighed, preserved in 96% ethanol and later freeze-dried following the procedure for faecal particle size analysis as described in Schulz-Kornas et al. (2019).

These samples were collected in the field over 19 weeks, from December to March, primarily as part of a faecal particle size analysis study. We made use of 11 dry-sieved size fractions, each of which were analysed and weighed in the labs of the Max Planck Weizmann Center for Integrative Archaeology and Anthropology, Max Planck Institute of Evolutionary Anthropology (Leipzig, Germany). We selected the three smallest fractions (20, 40 and 63 μm) for phytolith analysis following Power, Rosen and Nadel (2014). These sizes were selected because they represent the size range of phytoliths recovered from dental calculus. We then processed each of the three samples by dry-ashing in a muffle oven at 500 °C for 4 h in the labs of the Department of Human Evolution, Max Planck Institute for Evolutionary Anthropology (Leipzig, Germany). After cooling, calcites were removed from each sample by gradually adding 10% HCl for 10 min in 1.5-ml centrifuge tubes. A weighed subsample of the residue was mounted in Entellan New (Merck) with a 24×24-mm or 22×22-mm coverslip, depending on mounted mass.

Phytoliths and all other encountered micro-remains were counted at ×400 magnification (Num. Aperture = 0.95) using a field of view counting method with a light microscope (Axioskop A1, Zeiss). We counted single cells until we documented 250 or more examples (in some cases up to 500 cells) per sample, including unidentified phytoliths (Supplementary Table 1). We classified phytoliths into as many categories as were deemed useful to the project (Supplementary Table 2).

Samples for scanning electron microscope analyses were mounted on carbon tape or dried onto aluminium specimen stubs and then mounted to stubs with carbon tape. All samples were sputter-coated with gold on a Denton Desk V sputter coater for 1.5 min at 50 amps. Images were collected by a secondary detector at 10 kV with working distances ranging from 21 to 5 mm on a TESCAN Vega 3 Scanning Electron Microscope.

Multi-celled phytoliths were counted both as multi-celled entities and as single cells by individually recording each cell. For single-celled monocot phytoliths, we followed the ICPN classification system when possible ((ICPT) 2019; Madella, Alexandre, and Ball 2005). The number of phytoliths on the slide was calculated using the following algorithm:

\[ n \text{ phytoliths per slide} = \frac{n \text{ counted/fields counted}}{\text{total fields on slide}} \times \text{total amount of sediment mounted (mg)} \times 1000 \]

This value was used to derive a comparable unit of quantification—the number of phytoliths per gram of sediment. This value was calculated with the following formula:

\[ n \text{ phytoliths/gram} = \frac{n \text{ phytoliths per slide}}{\text{total amount of sediment mounted (mg)}} \times \frac{\text{total initial sediment (mg)}}{\text{total phytolith amount (mg)}} \times 1000 \]

The three values from each sample were combined, and the resulting information was compared with existing information on phytoliths in Taï Chimpanzee dental calculus (Power et al. 2015, 2014).

Observational dietary records taken by researchers standing no less than 7 m from a subject were obtained from the East Group Chimpanzees for comparison (Supplementary Fig. 1). Observational data were not available for Pos and Ath from all of the dates, so we sourced data from the relevant dates from across the East Group population from October 2007 to August 2017 to generate a reliable overview. The total observation time was approximately 10,000 hours of feeding data from 27,000 h of behavioural data. The data includes observation of 41 chimpanzees (16 males and 21 females). Taï chimpanzees primarily consume ripe eudicot fruit, although leaves, piths, nuts from eudicots, monocots and magnoliads are also used (Supplementary Figs. 1, 2, 3). We choose to sample 10 years of data because it averages out yearly fluctuations in fruiting phenology, which commonly occurs in the Tai Forest. We analysed feeding data based on its taxonomic families, genera and plant part, rather than based on the properties of each food (hard versus tough foods) or based on being chewed wadage or swallowed foods because this information is not available. Taxonomic information and the plant parts consumed were specifically marked in 104,000 entries of the record set.

Although surveys have been conducted of Western and Central African phytoliths, particularly savannahs, surveys of forests are incomplete, and they lack detailed coverage of the ‘Upper Guinea’ rainforest habitats that comprise the Tai Forest (Barboni and Bremond 2009; Collura 2017; Runge 1999). Additionally detailed phytolith classification that exceeded Power et al. (2015) in specificity was unnecessary. Because of this, we took a minimalist approach to
identification, and we classified phytoliths principally into the 15 groupings used in Power et al. (2015), which was based on a reference collection of 113 species. To support this information, we also prepared 52 phytolith samples of 45 unstudied plant species of Tai food plants using nitric acid-based wet ashing to assess their morphotypes (Supplementary Table 3) (Piperno 2006).

**Results**

**Phytolith abundance**

The phytolith concentration (measured as the number of phytoliths/g of dried faeces within the 20–63 µm fraction) varies significantly across different samples (Fig. 1). While some samples had only hundreds/g or no phytoliths, others contained high numbers (47,000/g) with a mean of 18,906/g. Generally, samples with a higher per cent of acid-insoluble fractions also had a higher number of phytoliths, but in many cases, this pattern was not present (Fig. 1; Table 1). In most samples, the acid-insoluble fraction varied from 0.9 to 5.6%. However, in two samples, it far exceeds this, reaching over 19%. These samples did not exhibit a corresponding increase in the number of phytoliths. As a whole, across the samples, there was no relationship evident between the acid-insoluble fraction and the number of multi-celled phytoliths (Table 1).

**Phytolith morphotype representation**

The number of phytoliths that can be readily identified closely followed the total number of phytoliths/g (Supplementary Table 1; Table 2). The most common types were monocot rugulose phytoliths and echinate spheroids (21% of all phytoliths), unspecific thick and thin elongates (14%) and eudicot plates, single-cell and multi-cell tracheids (11%) (Table 3; Fig. 2). Rarer types that were present include grass short-cells, which were less than 1% including reniforms, wide-lobe narrow-shank bilobates, flat bilobates, square trapezoid, ovals, rondels, and saddles. We also found occasional grass bulliforms, hairs, psilate and sinuate long-cell as well as eudicot multi-cellular honeycomb polygons, jigsaws, sclereids and thick decorated elongates, blocks and more ambiguous psilate thick oblongs and some other types (Supplementary Table 2). Cylindroid, hair bases and eudicot-type hairs were rare (below 2%). We did not find cystoliths.

**Diatoms and sponge spicules**

Diatoms occurred in low levels and in most samples were rare (<2% of all micro-remains) (Table 3). Diatoms were not systematically identified but observed species present in samples included the following genera: Orthoseira, Rhopalodia, Gomphonema, Calonesis and Nitzschia (Supplementary Fig. 4). Orthoseira are chain-forming aerophytic taxa that are typically found on wet surfaces, mosses and tree-bark (Spaulding and Kociolek 1998). The other diatom genera observed are all representative of shallow aquatic or semi-aquatic systems, typically found living attached to macrophytes, rocky or muddy substrates (Wehr, Sheath and Kociolek 2015). Diatoms were most abundant in the sample of Pos from the 10th of Dec. Sponge spicules were considerably rarer than diatoms (< 0.2% of all micro-remains) and were absent from most samples.

**Table 1** Summary of analysed samples and phytolith extraction information

| ID  | Birth year | Sampling date | Weight (mg) of residue | % acid insoluble | N. phytoliths/g | Multi-cell % | Fresh matter (g) | % 20–63 µm sieves |
|-----|------------|---------------|------------------------|------------------|----------------|--------------|----------------|-----------------|
| ATH | 1994       | 08 Dec 2014   | 3.807                  | 2.76             | 212            | 2.67         | 15            | 4.7             |
| ATH | 1994       | 24 Dec 2014   | 61.106                 | 18.03            | 34,235         | 2.3          | 15            | 11.1            |
| ATH | 1994       | 06 Jan 2015   | 51.99                  | 19.84            | 13,191         | 0.8          | 15            | 6.4             |
| ATH | 1994       | 06 Jan 2015   | 4.580                  | 4.31             | 29,638         | 2.13         | 15            | 13.9            |
| ATH | 1994       | 19 Jan 2015   | 0.719                  | 0.43             | 5,268          | 0            | 15            | 2.3             |
| ATH | 1994       | 05 Feb 2015   | 7.116                  | 3.79             | 7,911          | 2.41         | 15            | 11.2            |
| ATH | 1994       | 02 Mar 2015   | 5.886                  | 1.21             | 47,820         | 4.7          | 15            | 7.8             |
| ATH | 1994       | 16 Mar 2015   | 3.500                  | 0.97             | 0              | 5.27         | 15            | 2.2             |
| POS | 1998       | 10 Dec 2014   | 4.121                  | 3.6              | 640            | 0.89         | 20            | 5.7             |
| POS | 1998       | 23 Dec 2014   | 10.177                 | 2.62             | 24,604         | 1.12         | 17            | 11.3            |
| POS | 1998       | 06 Jan 2015   | 9.529                  | 5.78             | 19,638         | 4.29         | 21            | 32.1            |
| POS | 1998       | 02 Jan 2015   | 8.429                  | 4.11             | 33,495         | 4.69         | 19            | 22.7            |
| POS | 1998       | 02 Mar 2015   | 5.886                  | 5.38             | 3,005          | 2.58         | 19.1          | 24.1            |
| POS | 1998       | 16 Mar 2015   | 16.374                 | 3.89             | 20,392         | 7.11         | 19            | 27.5            |
Geological inclusions (grit content)

Very large angular (5–3000 μm, but mostly 40–200 μm) birefringent mineral particles from dust were common in many samples (Supplementary Fig. 5). Some examples exhibited considerable wear. In Ath 24 Dec 2014 and Ath 6 Jan 2015, these particles were extremely common, greatly outnumbering phytoliths on the slides and in some cases made counting phytoliths challenging. The acid-insoluble fraction of the faecal samples showed that on occasion, these grit particles were extremely common and far more abundant than phytoliths. Peaks in grit content were apparent around the period associated with the Harmattan (Fig. 1).

Discussion

Representativeness of morphotypes

The phytolith morphotypes of the faecal samples of the Taï chimpanzees are largely comparable to published Kibale chimpanzee faecal phytolith datasets (Phillips and Lancelotti 2014). The abundance of eudicot plate, tracheid phytoliths is similar to Kibale chimpanzees (Phillips and Lancelotti 2014), reflecting the eudicot-rich forest world and eudicot-based diet of both chimpanzee groups. The principal differences relate to the relative abundance of monocot rugulose and echinate spheroids in the Taï (Côte d’Ivoire) samples versus the Kanyawara, Kibale National Park (Uganda). We assume that this reflects underlying dietary differences between the two on a geographic and on a taxonomic level, i.e. subspecies of Pan troglodytes and distinct localities. The decorated rugulose/globular/spheroid phytoliths are produced by Zingiberaceae, which are commonly consumed at both localities. However, sub-variants are also produced by Marantaceae and especially Arecaceae, which are families that are consumed by chimpanzees in the Tai forest but not Kibale (Phillips and Lancelotti 2014; Power et al. 2015; Watts et al. 2012). There are other differences, too; the phytoliths consumed by the chimpanzees in Kibale appear to show more stomata, hair-base and hair morphotypes, although the reason for this is not clear. In addition, morphotypes were generally ubiquitous amongst Taï samples compared to those from Uganda, although this may relate to the shorter sampling window at Taï, 100 days from December to March, compared to 162 days over June to
December at Kibale. Contrasting these two signals is challenging because of our incomplete botanical knowledge of these two environments. Although surveys have been conducted of phytolith assemblages of Western and Central Africa, particularly of the savannahs, surveys of forests are incomplete and lack detailed coverage of the ‘Upper Guinea’ rainforests that comprise the habitats of the Taï Forest (Collura 2017; Runge1999).

**Representativeness of inferred diet**

The phytolith record has varying levels of representation. Although eudicots are known to produce few phytoliths, they are well represented in all samples, particularly by tracheids types (Table 3). The types of eudicot foods that are rare in chimpanzee diet are also rare in the samples, such as wood and sedge. Grasses occur in the samples, although grasses are not consumed at Taï, nor do they use grass as tools (Whiten et al. 1999), and grass is not abundant in their habitat. The presence of grasses plausibly could be linked to accidental inhalation of wind-blown grass phytoliths, or via accumulated dust adhering to foods. This explanation is plausible given that grasses produce very high numbers of phytoliths. Palms (monocots) are overrepresented but this is unsurprising given the high production in palm and other monocot plants, which are important chimpanzee staples (Supplementary Figs. 1 and 2). Marantaceae is an important food including the staple Halopegia pith; however, we were not able to identify any phytoliths from this family (Fig. 3). Overall, the faecal phytoliths more strongly reflect the Taï diet over the represented period than the dental calculus record (Table 3). However, it also represents their annual diet much better than dental calculus.

**Phytolith and dental calculus records**

The faecal samples present rich assemblages, even though chimpanzees engage in dietary behaviours that reduce the consumption of phytoliths. For example, chimpanzees consume a high amount of tree and shrub fruit and leaves (Boesch and Boesch-Achermann 2000). Taï forest fruit and leaves are thick and leathery, and chimpanzees chew them into wadage, for example, fig fruit (Boesch and Boesch-Achermann 2000), to absorb leaf nutrients without swallowing the fibrous and phytolith-rich wadage. The composition of the faecal phytolith assemblages is diverse, even though only two individuals were represented over a few months. Samples with abundant phytoliths also produced the most diverse assemblages. This reflects the diverse Taï diet, which includes at least 48 plant genera according to feeding observations (Schulz-Kornas et al. 2019) and at least 43 families during the seasonal window represented by the faecal samples of the present study (Supplementary Table 4). Although we do not have personalised feeding records from the immediate period prior to faecal sampling, analysis of the feeding data shows that several species may be consumed in 1 day (Supplementary Fig. 3). Although dental calculus represents years of life (Silverstone et al. 1981), the dental calculus assemblages were not as diverse as expected (compare Figs. 1 and 3). Therefore, we suggest that this indicates that dental calculus phytoliths omit a great deal of dietary diversity. In faecal samples, palms and gingers were moderately overrepresented. However, this overrepresentation contrasts with the dental calculus assemblages, which had substantial overrepresentation of these types. In contrast, the morphotypes, such as parallelepipedal and tracheids, seem to be disproportionately rare in the dental calculus even though they occur in greater abundance in their diet.
The reasons for this may relate to the toughness of palm and ginger phytolith types. Palm rugulate and spherical echinate morphotypes are known to resist decay, due to chemical or mechanical properties. This may relate to their bulkiness; they have a large volume to surface area ratios (Cabanes and Shahack-Gross 2015). In addition, their high abundance in dental calculus may relate to the fact that these echinate spheroids possess spines that may more securely embed to form dental plaque. In contrast, many phytolith types are smooth (e.g. parallelepipedals) and may be less likely to attach to dental plaque. In addition, their high abundance in dental calculus may relate to the fact that these echinate spheroids possess spines that may more securely embed to form dental plaque. In contrast, many phytolith types are smooth (e.g. parallelepipedals) and may be less likely to attach to dental plaque. In addition, overrepresentation may relate to the phytoliths of palms and gingers being less firmly in their parent plant cells than other phytolith types. Nevertheless, these findings highlight that researchers need to employ caution when using these morphotypes to study palms and gingers. This is important because palms are of key importance in African dietary history and agricultural expansions, as well as for ecological reconstruction (Neumann et al. 2012). Although palms are a key crop of agricultural expansions of Bantu peoples, researchers need to allow for the fact that palm is overrepresented (Maley and Chep 2001; Neumann et al. 2012). When only using phytoliths, the role of tropical eudicots can be overlooked easily; these phytoliths are substantially underrepresented in dental calculus and to a lesser extent in faecal samples.

**Effect of digestion**

Of the many processes that can affect phytoliths, digestion by plant-eating species is an important example and may affect the archaeological and palaeoecological phytolith record. Mastication and digestion expose phytoliths to harsh mechanical and chemical processes in the gastrointestinal tract. This has resulted in reports of fractured phytoliths and possibly deform phytoliths (Baker, Jones, and Wardrop 1959; Power and Berthaume 2016; Walker, Hoeck, and Perez 1978). As we only have generalised information on the frequencies of different phytolith morphotypes in each plant food, we cannot

---

**Table 3** Comparing consumed plant taxa and taxa inferred from faecal phytolith assemblage and dental calculus. As identified foods and as phytolith morphotypes, with time spent consuming in feeding data; excludes unidentified phytoliths

| Identified foods       | Count | Percentage of plant diet | Time consumed in feeding data | Morphotypes          | Count | Percentage of diet |
|------------------------|-------|--------------------------|-------------------------------|----------------------|-------|-------------------|
|                        | Calculus | Faecal | Calculus | Faecal                  | (h/min/s) | Percentage of plant diet |
| Palms and gingers      | 1369  | 41510 | 92 | 34 | 21:01:00 | 0.63 |
| Eudicots               | 70 | 66109 | 5 | 54 | 3296:26:00 | 99 |
| Grasses                | 28 | 6205 | 2 | 5 | 01:07:00 | 0.03 |
| Woods                  | 0 | 3005 | 0.00 | 2.45 | 11:20:00 | 0.33a |
| Diatoms                | 16 | 3593 | 1 | 3 | 01:20:00 | 0.3b |
| Sedges                 | 0 | 700 | 0.00 | 0.57 | Unavailable | 0.57 |
| Feathers               | 8 | 700 | 0.54 | 0.57 | | |
| Spicules               | 0 | 700 | 0.00 | 0.57 | | |

| Morphotypes            | Count | Percentage of diet | Time consumed in feeding data |
|------------------------|-------|-------------------|-------------------------------|
|                        | Calculus | Faecal | Calculus | Faecal                  | (h/min/s) | Percentage of plant diet |
| Unidentified phytoliths | 89 | 66772 | 4.88 | 37.43 | | |
| Spheroid echinates     | 1369 | 37459 | 75.10 | 21.00 | | |
| Parallelepipedals      | 57 | 25914 | 3.13 | 14.53 | | |
| Tracheids              | 4 | 10663 | 0.22 | 5.98 | | |
| Plates                 | 13 | 10253 | 0.71 | 5.75 | | |
| Long-cells             | 106 | 9322 | 5.81 | 5.23 | | |
| Short-cells            | 28 | 6205 | 1.54 | 3.48 | | |
| Hair cells             | 65 | 3876 | 3.57 | 2.17 | | |
| Bulliforms             | 53 | 3503 | 2.91 | 1.96 | | |
| Smooth spheroids/ovals | 0 | 3005 | 0.00 | 1.68 | | |
| Acicular hair cells    | 12 | 897 | 0.66 | 0.50 | | |
| Ellipsoids             | 7 | 507 | 0.38 | 0.28 | | |
| Cylindroids            | 20 | 3 | 1.10 | 0.00 | | |

a Based on water consumption
b Based on Boesch and Boesch-Acherman (2000)
precisely determine if digestion breaks down or dissolves some morphotypes. However, we do note the assemblages underrepresent some phytolith types, such as plates. Other types, such as stoma, hair bases, polygons, irregular and thick types also appear underrepresented. It is not clear if the existing Tai faecal phytoliths are eroded or fractured by digestion, because the morphotypes that occur in the samples are types that do not readily exhibit damage. Although phytoliths such as grass long-cells easily show damage, it is less clear on tracheids and spheroids. However, multi-cellular phytolith forms were less common than expected. For example, multi-celled polyhedral forms were infrequent, which is inconsistent with their eudicot-rich diet. This may reflect fracturing of fragile multi-cellular types. Morphotype representation is greater in samples with more phytoliths per gram, possibly suggesting poor preservation (Madella and Lancelotti 2012).

**Dietary dust richness**

Phytoliths are a major component of plant resistance to herbivory, and phytolith production is increased by above-ground herbivory (Hartley and DeGabriel 2016; Hodson et al. 2005; Power et al. 2016). Because phytoliths are hard and gritty, they serve as an anti-feedant that actively discourages herbivory by inhibiting nutrient absorption (Moore and Johnson 2017; Sanson, Kerr, and Gross 2007). It has been argued that phytoliths may cause dental wear, which is a ubiquitous feature of ageing teeth (Piperno 1988). Although it is debated if phytolith material properties and ultrastructural morphology are sufficient to drive dental wear (for review (Kaiser et al. 2018; Schulz-Kornas et al. 2017)), hard grit, probably composed of quartz, was common in the faecal samples during the dry season (Harmattan). These particles were often large, often far greater in size (5–200 μm) than phytoliths and appeared to be Saharan Aeolian dust. Grass short-cells were also found, which we believe were transported to Tai by the Harmattan, helping to verify Aeolian dust as the likely source. Given that, these indicate a variety of environments, likely including savannahs (Supplementary Fig. 6). In contrast to phytoliths, these particles undoubtedly wear down dental enamel (van Casteren et al. 2018; Kaiser et al. 2015; Lucas et al. 2014). The phytolith content of Tai plant foods vary, but in nearly all foods, it is below 5% of the dry matter (Supplementary Table 3). Only in seed cases of the *Palisota* genus have we identified a very high acid-insoluble fraction. However, this plant is a minor food, and *Palisota* seed cases are reported to be spat out by chimpanzees, so the high phytolith content in this plant cannot explain the high acid-insoluble fraction (e.g. in Ath 24 Dec 2014 and Ath 6 Jan 2015). Nevertheless, it shows that phytoliths are found in seedcases, and eudicotyledoneous seeds have been suggested to be the main reason for tooth wear.

![Mosaic selection of recovered phytoliths. a Large spheroid echinate from a palm, probably oil palm fruit (Pos 2 March 2015). b Large prolate echinate from a palm (Pos 23 Dec 2014). c Short-celled rondel from a grass (Pos 6 Jan 2015). d Dentate thick block (Pos 2 Feb 2015). e Decorated-margin elongate (Ath 6 Jan 2015). f Multi-cellular polyhedral honeycomb from a eudicot leaf (Pos 10 Dec 2014). g Hair base from a eudicot leaf (Pos 10 Dec 2014). h Multi-cellular tracheid, from a eudicot leaf (Pos 2 Feb 2015). i Surface decorated elongate from a eudicot (Pos 23 Dec 2014). The black scale bar is 10 μm.](image)
wear creating large pits (Ragni, Teaford, and Ungar 2017; Ungar and Peter 2008) and valleys (Schulz-Kornas et al. 2019) on primate tooth facets.

Conclusions

Bringing faecal and dental calculus datasets together addresses some of the methodological concerns researchers have raised about inferring diet from dental calculus. Although this study highlights biases that exist with using dental calculus and faecal samples for reconstructing diet, it also helps to validate that these studies in these regions have produced a signal that is dietary in origin. This study provides important data, crucial for interpreting diet in the African tropics, but further studies with a similar research design are needed to assess the representativeness of dental calculus in other environments and amongst other species.

Acknowledgements

We thank the Ministère de l’Enseignement Supérieur et de la Recherche Scientifique, the Ministère de l’Environnement et des Eaux et Forêts of Côte d’Ivoire and the Office Ivorien des Parcs et Reserves for permission to conduct this research. We are grateful for the support by the Centre Suisse de Recherches Scientifiques at Abidjan, Côte d’Ivoire. We would like to express our gratitude to Amanda G. Henry, Jean-Jacques Hublin, Róisín Murtagh and Tobias Deschner for the access to the research facilities. We thank the field assistants at Tâ National Park for the collection of faecal samples. We are thankful to Liran Samuni for helping provide Tâ data files and Nicholas Ting for help with programming. We thank Mallory Wilson for assistance with preparation of samples for SEM analyses at Indiana State University. Tina Kottek and Ramona Beuth (Max Planck Weizmann Center for Integrative Archaeology and Anthropology) are thanked for assisting with sieving the faeces.

Funding

Open Access funding enabled and organized by Projekt DEAL. Core funding for the Tâ Chimpanzee Project is provided by the Max Planck Society since 1997. This research was supported by the Max Planck Society.

Open Access

This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1007/s12520-021-01342-z.
References

(ICPT), International Committee for Phytolith Taxonomy (2019) International Code for Phytolith Nomenclature 2.0 (ICPN 2.0). Ann Bot 124(2):iii
Albert RM, Shahack-Gross R, Cabanes D, Gilboa A, Lev-Yadun S, Portillo M, Sharon I, Boaretto E, Weiner S (2008) Phytolith-rich layers from the Late Bronze and Iron Ages at Tel Dor (Israel): mode of formation and archaeological significance. J Archaeol Sci 35(1):57–75
Baker G, Jones LHP, Wardrop ID (1959) Cause of wear in sheep’s teeth. Nature 184(4698):1583–1584
Barboni D, Bremond L (2009) Phytoliths of East African Grasses: an assessment of their environmental and taxonomic significance based on floristic data. Rev Palaeobot Palynol 158(1–2):29–41
Boesch C, Boesch-Achermann H (2000) The Chimpanzees of the Taï Forest: Behavioural Ecology and Evolution. Oxford University Press, Oxford
Cabanes D, Shahack-Gross R (2015) Understanding fossil phytolith preservation: the role of partial dissolution in paleoecology and archaeology. PLoS One 10(5):e0125532
van Casteren A, Oelze VM, Angedakin S, Kalan AK, Kambi M, Boesch C, Kühn HS, Langenbraber KE, Piel AK, Stewart FA, Kuczki K (2018) Food mechanical properties and isotopic signatures in forest versus savannah dwelling eastern chimpanzees. Commun Biol 1(1):109
Chabot Y, Garnier A, Made Geria I (2018) Phytolith analysis from the Tigrinya-Afar region, Eritrea: a contribution to the state-of-the-art. Constr Build Mater 73(30):728–739
Enete IC, Obienusi EA, Igu IN, Ayadiulo R (2011) Harmattan dust: composition, characteristics and effects on soil fertility in Enugu, Nigeria. Curr J Appl Sci Technol 2(1):72–81
Engelstaedter S, Washington R (2007) Atmospheric controls on the annual cycle of North African dust. J Geophys Res 112(D3):D03103
Falchi M, Moustafa JSE-S, Takousis P, Pesce F, Bonnefond A, Anderssson-Assarsson JC, Sudhant PM, Dorajoo R, Al-Shafai MN, Bottolo L, Ozdemir E, So H-C, Davies RW, Patrice A, Dent R, Mangino M, Hysi PG, Dechaume A, Huyvaert M, Skinner J, Pigenet M, Caiuzzo R, Raverdy V, Vaillant E, Field S, Balkau B, Marre M, Visvikis-Siest S, Well J, Pouliain-Godefroy O, Jacobson P, Sjoestrom L, Hammond CJ, Deloukas P, Sham PC, McPherson R, Lee J, Shyong Tai E, Sladek R, Lena M., S., Carlson. Andrew Walley, Evan E, Eichler, Francois Pattou, Timothy D, Spector, and Philippe Froguel. (2014) Low copy number of the salivary amylase gene predisposes to obesity. Nat Genet 46(5):492–497
Gur-Arieh S, Shahack-Gross R, Mair AM, Lehmann G, Hitchcock LA, Boaretto E (2014) The Taphonomy and preservation of wood and dung ashes found in archaeological cooking installations: case studies from Iron Age Israel. J Archaeol Sci 46:50–67
Hartley SE, DeGabriel JL (2016) The ecology of herbivore-induced silicon defences in grasses edited by J. Cooke Func Ecol 30(8):1311–1322
Hodson MJ, White PJ, Mead A, Broadley MR (2005) Phylogenetic variation in the silicon composition of plants. Ann Bot 96(6):1027–1046
Jenkins E, Jamjoum K, Al Nuimat S (2011) Irrigation and phytolith formation: an experimental study. In: Mithen S, Black E (eds) Water, Life and Civilisation: Climate, Environment and Society in the Jordan Valley. Cambridge University Press, Cambridge, pp 347–372
Kaiser TM, Braune C, Kalinka G, Schulz-Kornas E (2018) Nanoindentation of native phytoliths and dental tissues: implications for herbivore-plant combat and dental wear proxies. Evol Syst 26(1):55–63
Kaiser TM, Clauss M, Schulz-Kornas E (2015) A set of hypotheses on tribology of mammalian herbivore teeth. Surf Topogr: Metrol Prop 4(1):014003
Leonard CA, Vashro L, O’Connell JF, AG, Henry (2015) Plant microremains in dental calculus as a record of plant consumption: a test with two forager-horticulturalists. J Archaeol Sci Rep 2:449–457
Lucas PW, van Casteren A, Al-Fadhalah K, Almusallam AS, Henry AG, Michael S, Watzke J, Reed DA, Diekwisch TGH, Strait DS, Atkins AG (2014) The role of dust, grit and phytoliths in tooth wear. Ann Zool Fenn 51(1–2):143–152
Madella M, Alexandre A, Ball T (2005) International Code for Phytolith Nomenclature 1.0. Ann Bot 96(2):253–260
Madella M, Lanceccotti C (2012) Taphonomy and phytoliths: a user manual. Quat Int 260:75–120
Maley J, Chep A (2001) Elaeis Guineensis Jacq. (oil palm) fluctuations in the state-of-the-art. Constr Build Mater 73(30):728–739
Springer

made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.
