Stem cell-derived biofactors fight against coronavirus infection

Mohammadreza Ardalan, Leila Chodari, Sepideh Zununi Vahed, Seyed Mahdi Hosseiniyan Khatibi, Aziz Eftekhari, Soodabeh Davaran, Magali Cucchiarini, Leila Roshangar, Elham Ahmadian

Author contributions: Ahmadian E conceived and designed the study; Eftekhari A and Chodari L acquired the data; Zununi Vahed S prepared the first draft of the manuscript; Hosseiniyan Khatibi SM designed and drew the figures in the manuscript; Cucciarini M and Ardalan M proofread the manuscript and made critical revisions; Davaran S and Roshangar L revised the manuscript; all authors read and approved the final version of the manuscript.

Conflict-of-interest statement: The authors declare that there is no conflict of interest.

Country/Territory of origin: Iran

Specialty type: Cell and tissue engineering

Provenance and peer review: Invited article; Externally peer reviewed.

Abstract

Despite various treatment protocols and newly recognized therapeutics, there are no effective treatment approaches against coronavirus disease. New therapeutic strategies including the use of stem cells-derived secretome as a cell-free therapy have been recommended for patients with critical illness. The pro-regenerative, pro-angiogenic, anti-inflammatory, anti-apoptotic, immunomodulatory, and trophic properties of stem cells-derived secretome, extracellular vesicles (EVs), and bioactive factors have made them suitable candidates for respiratory tract regeneration in coronavirus disease 2019 (COVID-19) patients. EVs including microvesicles and exosomes can be applied for communication at the intercellular level due to their abilities in the long-distance transfer of biological messages such as mRNAs, growth factors, transcription factors, microRNAs, and cytokines, and therefore, simulate the specifications of the parent cell, influencing target cells upon internalization and/or binding. EVs exhibit both anti-inflammatory and
tolerogenic immune responses by regulation of proliferation, polarization, activation, and migration of different immune cells. Due to effective immunomodulatory and high safety including a minimum risk of immunogenicity and tumorigenicity, mesenchymal stem cell (MSC)-EVs are more preferable to MSC-based therapies. Thus, as an endogenous repair and inflammation-reducing agent, MSC-EVs could be used against COVID-19 induced morbidity and mortality after further mechanistic and preclinical/clinical investigations. This review is focused on the therapeutic perspective of the secretome of stem cells in alleviating the cytokine storm and organ injury in COVID-19 patients.

Key Words: COVID-19; Secretome; Mesenchymal stem cell; Exosome; Stem cell; Biofactors

Citation: Ardalan M, Chodari L, Zumuni Vahed S, Hosseiniyan Khatibi SM, Eftekhar A, Davaran S, Cucchiarini M, Roshangar L, Ahmadian E. Stem cell-derived biofactors fight against coronavirus infection. World J Stem Cells 2021; 13(12): 1813-1825

URL: https://www.wjgnet.com/1948-0210/full/v13/i12/1813.htm

DOI: https://dx.doi.org/10.4252/wjsc.v13.i12.1813

INTRODUCTION

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is designated as the etiology of coronavirus disease 2019 (COVID-19), which is a pandemic viral disease. The symptoms from death or acute respiratory distress syndrome (ARDS) to mild gastrointestinal symptoms have gained considerable attention as a probable therapeutic option in COVID-19. Stem cell and stem cell-derived secretome-related therapies have gained increasing momentum in the treatment of a broad range of diseases in the past decade. In particular, the immunomodulatory properties of stem cell-derived biofactors could be a new avenue in the treatment of COVID-19 patients.

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is designated as the etiology of coronavirus disease 2019 (COVID-19), which is a pandemic viral disease. The symptoms from death or acute respiratory distress syndrome (ARDS) to mild respiratory symptoms[1]. Excessive systemic immune activation of patients generates a cytokine storm, which is found in severely ill COVID-19 patients. Recent evidence indicates that the cytokine storm could play a key role in disease progression, resulting in the failure of various organs or even death. Hence, approaches which prevent the cytokine storm may be significant in mitigating COVID-19[2,3].

Complications such as cardiovascular system dysfunction, primarily acute myocardial injury, arrhythmia, or heart failure[4], neurological complications[5], gastrointestinal symptoms[6], and acute kidney injury[7] have been identified in a substantial proportion of COVID-19 patients. The unprecedented COVID-19 pandemic demands urgent therapies. Currently, multiple medicines involving anti-viral, anti-malarial, and anti-inflammatory agents are being investigated. Regardless of the patient’s recovery and survival due to various therapeutics, lung damage in these patients does not fully recover. Recently, promising stem cell therapies and importantly secreted extracellular vesicles (EVs) have been shown to exhibit anti-inflammatory effects and attenuate COVID-19-related lung injury.

A new therapeutic approach which involves cellular therapies is promising in treating chronic and acute lung diseases due to their anti-inflammatory, immunomodulatory, regenerative, pro-angiogenic, and anti-fibrotic features. Mesenchymal stem cell (MSC)-secretome (a paracrine mechanism) composed of EVs and free soluble proteins mediate those therapeutic impacts[8]. The remarkable properties of exosomes have gained considerable attention as a probable therapeutic option in COVID-19. In vivo and in vitro studies have been conducted to determine the various therapeutic effects of MSC-secretome in tissue regeneration, heart, and lung diseases. The inflam-
mation suppressing effects of MSC-secretome are due to the prevention of monocyte differentiation into dendritic cells, prohibiting natural killer (NK) cells proliferation and cytotoxicity, stimulating macrophage polarization from the pro-inflammatory (M1) to anti-inflammatory (M2) phenotype, regulating the inflammatory characteristics of T helper cells, and inhibiting T cell proliferation. Growth factors found in the MSC-secretome regenerate the damaged lung tissue by increasing proliferation and reducing apoptosis of resident lung epithelial and endothelial cells. Additionally, antimicrobial peptides (AMPs) have been observed in MSC-secretomes and demonstrate antimicrobial properties, whereas protease inhibitors reduce extra protease function in the lung, preserving the protease/anti-protease equilibrium[9]. Exosomes extracted from MSC act as multitargeting agents. Therefore, they diminish the cytokine storm and prevent the inhibition of COVID-19-related anti-viral defenses in hosts[10]. Exosomes may hamper the cytokine storm and inflammatory process due to their reparative properties and thus induce endogenous repair. Hence, MSC-secretome might be a valuable cell-free substitute to cell-based therapies alone or in combination with pharmacological agents. In this review, the therapeutic potential of the secretome of stem cells in mitigating COVID-19-induced cytokine storm and organ damage is presented.

STEM CELL-BASED THERAPEUTICS

Evidence has shown the promising role of MSCs in COVID-19 pneumonia treatment. Human umbilical cord MSCs given to a 65-year-old female with severe COVID-19 induced substantial recovery through modulation of the immune system and regeneration of damaged tissue with high safety[11]. Every three days, MSCs were administered intravenously by clinicians three times ($5 \times 10^7$ cells each time). Leng et al [12] demonstrated the enhancement of pulmonary function and symptom improvement in seven COVID-19 patients with pneumonia in only two days after administration of MSCs. In their study, only one administration per kilogram of weight containing $1 \times 10^6$ cells was performed. The authors proposed that the therapeutic impact mainly occurred based on the immunoregulating features of MSCs. Remarkably, MSCs are not virus infectable as they are angiotensin-converting enzyme 2 negative. Hence, for treating seriously ill COVID-19 patients under certain protocols, MSCs can be considered a potential treatment option[13]. As the severity of this viral infection is closely associated with the host’s immune response, the immunomodulatory effects of MSCs can efficiently prohibit the cytokine storm and thus treat severe cases of COVID-19. Indeed the outcomes of COVID-19 patients can be enhanced by the transplantation of MSCs using various methods, including “as a result of their immunoregulatory impact, as a result of inducing regeneration and repairing tissue, and as a result of their antimicrobial, antifibrotic, and angiogenic properties”.

All these methods improve lung repair and prevent multiple organs from exaggerated immune response-derived damage. The ongoing COVID-19 clinical trials based on MSCs have been reviewed recently[14]. It now seems that MSCs convey their therapeutic effects through the paracrine pathway. As these cells can discharge secretome (active biological substances), therefore they can be potentially addressed as drug stores[15].

THE SECRETOME OF STEM CELLS: A CELL-FREE ALTERNATIVE TO CELL-BASED THERAPEUTICS

The secretome is defined as a stem cell secretion composed of regulatory factors and various soluble molecules, including AMPs, angiogenic growth factors, lipid mediators, and anti-inflammatory cytokines. Evidence has shown that these molecules are packed into EVs, also known as cell-secreted vesicles[16,17].

Common secretory mechanisms are involved in the excretion of secretomes by stem cells (Figure 1). Following the administration of the secretome or the culture medium in patients, through a paracrine signaling pathway, neighboring cells assimilate them[18]. Two important secreted EVs, are exosomes and MVs, also known as microvesicles alongside the apoptotic bodies secreted by stem cells[19]. The fusing of plasma and multi-vesicular bodies facilitates exosome (30-100 nm) elicitation. However, cellular membrane budding generates MVs (100-1000 nm), which possess cellular cytoplasm. EVs are discharged into the extracellular microenvironment and act like soluble
Figure 1 The stem cell secretome. After the interaction between host angiotensin-converting enzyme 2 (ACE2) receptor and the spike S protein of severe acute respiratory syndrome coronavirus 2, the membrane is fused, and the viral genome is released into the cell or the virus enters the cell by clathrin-dependent/independent endocytosis. Most of the cells in human organs including the intestine, heart, kidney, and alveolar type II (AT2) cells of the lungs express ACE2 receptors. Extracellular matrix metalloproteinase inhibitor (EMMPRIN or CD147) and two proteases transmembrane serine protease 2 (TMPRSS2) are required for virus entry into the host cell. The lungs are damaged after direct destruction of capillary endothelial cells and AT2, the renin-angiotensin system is disrupted or the immune response is diminished indirectly. Following virus-induced infection, pathogen-associated molecular patterns lead to recognition of the virus by the innate immune system, activation of nuclear factor-κappa B and IRF3 pathways, type I interferon (IFN) expression and consequently activation of the JAK/STAT pathway and finally the expression of IFN-stimulated genes (ISG) which have anti-viral activity. The abovementioned effective immune response is required for successful virus clearance and clinical disappearance of the disease. Nonetheless, the IFN response may be delayed due to evasion of IFN and ISG mediated killing by the virus which in turns leads to hyper-inflammatory neutrophils and macrophage infiltration at the pulmonary site accompanied by pro-inflammatory cytokines including granulocyte-colony stimulating factor, tumor necrosis factor, MCP1, and interleukin-1β/2/6/7/8/17. This hyper-activation of T lymphocytes and the innate response is called the ‘cytokine storm’ and is responsible for lung disorders including acute respiratory distress syndrome, pneumonitis, viral sepsis, respiratory and organ failure. A high number of pro-inflammatory cytokines leads to hyaluronan synthase 2 induction which elevates hyaluronan production and fluid accumulation in the lungs. In critical cases of coronavirus disease 2019, the virus enters the peripheral blood and translocates to various target organs including kidney, heart, and intestine and can cause multiple organ failure. ACE2: Angiotensin-converting enzyme 2; SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2; TNF: Tumor necrosis factor; IL: Interleukin; DC: Dendritic cell.

 MSC-DERIVED EVS

Upon secretion, proteins, and EVs, through ligand-receptor interactions or internalization, engage with the target cells and regulate cellular responses. The secretome stimulates endogenous stem cells and progenitor cells, prevents apoptosis, attenuates the inflammatory response, triggers extracellular matrix remodeling and angiogenesis, prevents fibrosis, and regulates chemoattraction. It was revealed that following the MSCs’ functional mitochondria or mitochondrial DNA transfer to target cells, they protect cellular aerobic respiration with non-healthy mitochondria or modulate T cell functions. By resembling their parent cells, EVs from MSCs have characteristics such as immunoregulatory, anti-oxidative, regenerative, pro-metabolic, anti-apoptotic, and anti-inflammatory, in the microenvironment of damaged tissue. In components and through endocrine and paracrine methods, they express their biological effects. In a more general definition, MSC-secretome contains all the secreted bioactive factors of MSCs, with both extravesicular and the soluble elements.
MSC-based therapy, extracted EVs from MSCs are considered a substantial alternative to cell-based treatments[22]. Their efficacy is presently under in vitro examination for lung damage treatment utilizing MSC-derived EVs in various preclinical experiments [22]. EVs extracted from MSCs showed efficacy in lung injury due to influenza in a pig model. Studies have revealed that extracted EVs from MSCs are available 12 h after virus infection and diminish the levels of pro-inflammatory cytokines as well as viral replication[23]. Ang-1 mRNA (an angiogenic trophic factor) is found in EVs from MSCs. Due to this factor’s role in limiting leukocytes and vessel endothelial cells' interaction and sustaining the vascular barrier integrity, Ang-1 mRNA is considered substantial in endothelial cell stabilization during the injury process. The impact of EVs extracted from MSCs on lipopolysaccharide-induced acute lung damage in an experimental mouse model sheds light on the contributions of Ang-1 mRNA transfer by EVs to restore pulmonary capillary permeability[24]. Furthermore, EVs extracted from MSCs influence inflammation by inhibiting the expression of tumor necrosis factor alpha (TNF-α) and stimulating interleukin (IL)-10 secretion. Upon Ang-1 mRNA transfer and the internalization of EVs extracted from MSCs into injured endothelial cells, within the damaged lung microvascular endothelium, Ang-1 mRNA partially preserves protein permeability[25]. EVs are increasingly regarded as a substantial alternative to cell-based therapy. 

Secretome administration is associated with multiple advantages in comparison with complete MSCs therapy. The secretome lacks self-replication and is not involved in endogenous tumor development, as it has low immunogenicity, and upon intravenous injection, it contributes to low emboli formation. Therefore, the secretome is deemed more advantageous than cells[26]. The MSC-secretome can be altered and preserved more conveniently than cells with fewer costs regarding technological advances. It also suits emergency interventions as the product is ready-to-use[27]. With regard to monoclonal antibody therapy, the costs of the MSC-secretome are lower, which is vital in the management of a pandemic. Nonetheless, many concerns regarding EVs should be resolved before clinical application, and the delivery route (intravenous or inhalation), purification, bio-distribution, production, and characterization should be determined.

**EXOSOMES**

Exosomes are nanoparticles (40-150 nm) which have various bioactive components, including proteins, growth factors, lipids, microRNAs (miRNA), long noncoding RNAs, and transfer RNAs. The lipid contents of exosomes provide the platform for their infusion with neighboring cells and plasma membrane[28]. Following internalization of the secretome components, neighboring cells alter several downstream pathways, such as fibrosis inhibition, immunoregulation, damaged tissue remodeling, and apoptosis suppression[8].

With regard to exosome isolation and production, MSCs discharge exosomes under circumstances such as cytokine treatment, serum starvation, or hypoxia[29]. Purification and the introduction of exosomes into the body can then take place. It has been shown that exosomes derived from MSCs generate an impact resembling that of MSCs[30]. The multiple proteins, miRNAs, and mRNAs transported from secretory cells to the exosomes’ target cells exhibit anti-inflammatory traits[31]. Exosomes can stimulate regulatory cytokines, decrease the production of inflammatory cytokines, and prevent inflammation[32]. Impeding NK cells, CD4+ and CD8+ T cells can occur with MSC-exosomes[33]. They induce T cell IL-7 expression and stimulate the expression of IL-10 by regulatory cells, which are implicated in the suppression of inflammation. Furthermore, MSC-exosomes, by secreting transforming growth factor β (TGF-β) prevent CD4+ and CD8+ T cell differentiation and suppress inflammation in vivo[34]. MSC-exosomes treatment inhibits the activation and proliferation of NK cells[35]. MSC-exosomes engage in prohibiting pro-inflammatory states by shifting M1 macrophages to M2 phenotypes[36]. Moreover, MSC-exosomes prevent the secretion of pro-inflammatory factors including IL-17, interferon (IFN)-γ, IL-1β, TNF-α, and IL-6[37], and stimulate the secretion of anti-inflammatory factors including TGF-β, IL-4, and IL-10[38]. Also, their function decreases the serum chemokine level[39]. The immunoregulatory features of MSC-exosomes are associated with their anti-inflammatory components, including PD-L1, HLA-G, Galectin-1, and IDO[40-42]. Furthermore, MSC-exosomes, by escalating ATP levels in alveolar type II cells, increase their survivability[43]. In addition, exosomes possess adhesion molecules which accurately guide them to the damaged site. The exosome components then cross the
blood-brain barrier. They are low-cost and do not undergo independent self-renewal processes. Thus, they impede serious consequences involving tumor development and other adversities.

**THE ADVANTAGEOUS THERAPEUTIC IMPACT OF STEM CELL-DERIVED SECRETOME**

The therapeutic effects of MSC-derived EVs on lung and heart injuries have been demonstrated. There are also studies on the impact of MSC-EVs on hemagglutination of swine, avian, and human influenza viruses[23]. In addition, MSC-exosomes reduced mortality in H7N9 patients with no concomitant toxic complications during the follow-up period[44]. One study revealed that S proteins within exosomes can be considered a novel vaccine for countering SARS-CoV infections[45]. In a test of S-containing exosomes immunogenicity in mice, the results showed amplified titers of neutralizing antibody. Moreover, with regard to economics, MSC-exosomes therapy was a less expensive treatment than maintaining and extending individualized clonal cell populations[46]. In the following section, we provide a general review of the present findings on stem-cell extracted secretomes in preclinical studies for lung and heart injuries (the organs most damaged by SARS-CoV-2).

**STEM CELL-DERIVED SECRETOME IN THE PATHOGENESIS OF ORGSANS**

EVs extracted from MSCs have been shown to have an impressive impact on ARDS and acute lung injury. This is the result of the immunoregulatory and anti-inflammatory features of MSC-EVs[24], which induces shrinkage of the permeability of the endothelium and epithelium of alveoli[25], enhancing alveolar fluid clearance[43], macrophage phagocytosis improvement[47], and direct mitochondrial transfer with host cells promoting tissue repair.

Secretome in the blood has impressive stability, subsequent to MSC intravenous administration, and reaches the lungs via blood flow. It is then distributed in the tissues and promotes bacterial clearance, resolution of inflammation, enhances immune regulation, and preserves capillary barrier function[19]. The soluble components of MSCs inhibit inflammation, and EVs induce tissue repair. EVs secreted from MSCs, in particular, lung injuries, provide metabolites, DNA, miRNA, mRNA, and proteins to cells thus enhancing lung repair, and restoring and regenerating lung function[48].

Lung accumulation of MSCs occurs after systematic administration. Following secretion of their components, they enhance the pulmonary microenvironment, preserve the epithelial cells of alveoli, inhibit pulmonary fibrosis, and strengthen lung function[13]. Furthermore, distant affected organs (for example, the cardiovascular system) can take advantage of MSCs due to the secretory characteristics of these cells. Various studies have focused on the circulation of the cellular cargo, and preclinical trials revealed their capacity to manipulate diverse pathways to promote cellular communication. miRNA (a composition of exosomes) has been demonstrated to have a significant role in physiological functions, including immune modulation, development, epigenetic modifications, and so on[49]. The manufacture and isolation of EVs could be a beneficial therapy in pulmonary injuries[50].

An experimental mouse model of neonatal hyperoxia showed that MSCs from human bone marrow and Wharton’s jelly inhibited lung fibrosis, enhanced pulmonary vascular remodeling, and stimulated lung development in bronchopulmonary dysplasia. MSC-derived exosomes exhibited anti-inflammatory activity and altered the pro-inflammatory M1 pulmonary macrophages to anti-inflammatory M2 macrophages followed by inhibition of lung inflammation and the immune response facilitating organ development[51]. Exosomes derived from MSCs have demonstrated mitigating effects in an asthma and ARDS model of lung damage[52]. The potential role of exosomes in alveoli fluid clearance was identified during an ex vivo experiment involving human donor lung (not suitable for transplantation) perfusion. This was assisted by exosome CD-44 which was involved in the internalization mechanism in damaged host cells[53]. In addition, exosomes extracted from MSCs have been indicated in the direct inhibition of viral multiplication.

Overall, exosomes extracted from MSCs show promising effects on decreasing pulmonary edema and protein permeability, reversing lung inflammation, the prolif-
eration of lung epithelium, and the polarization of lung macrophages[54]. Additionally, MSC-derived exosomes are effective in the treatment of cardiovascular [55] and renal disease[56].

The therapeutic impact of MSC-EVs on acute myocardial infarction has been reported. This has been shown to involve the following underlying mechanisms: Reduction of the inflammatory response[29], reduction of cardiac fibrosis, mitigation of cardiomyocyte apoptosis[57], induction of angiogenesis[58] and promotion of cardiomyocyte autophagy[59]. It was also shown that, fibroblast growth factor, composed of MSC-secretome (derived from adipose tissue), inhibited viral replication processes[60].

**STEM CELL-DERIVED EXOSOMES; A NANO-PLATFORM FOR COMBATING COVID-19**

COVID-19 patients may develop multiorgan damage. In the initial phases of infection, mainly pneumocyte type II cells are infected, and other target cells may be bronchial cells, monocytes, macrophages, and enteric cells. Moreover, the principal SARS-CoV-2 cardiovascular complication is acute myocardial injury[61]. Heart tissue biopsies from COVID-19 patients revealed mononuclear inflammatory infiltration, more commonly found in cardiomyocyte necrosis sites[62]. Applying stem cell-derived secretome to organs damaged by COVID-19 is possible, according to extracted data. Also, the survival rate of septic mice increased following MSC-derived exosome treatment[63]. With regard to MSC-derived exosomes as supportive therapy in the current pandemic, this can be beneficial in inhibiting the effects of COVID-19 [42] and healing organ damage.

Experimental studies on the biological activity of MSC secretomes have demonstrated the possibility of applying MSC-derived secretomes for seriously ill COVID-19 patients as a cell-free therapy. EVs and proteins contained in MSCs affect endogenous stem and progenitor lung cells. They promote cell differentiation and proliferation, inhibit the inflammatory response, prevent apoptosis, reduce fibrosis, and recover capillary barrier function. Due to their similarity to parental MSCs, they are also effective in the management of chronic and acute lung injuries[64]. Exosomes (ExoFlo™) derived from MSCs of allogeneic bone marrow have been proposed as a treatment for seriously ill COVID-19 patients according to the first prospective nonrandomized open-cohort study conducted. ExoFlo is considered to be a hopeful therapeutic candidate for COVID-19 due to its capacity to restore oxygenation, immunity reconstitution, safety traits, and downregulation of the cytokine storm[10]. This study included twenty-four patients suffering from ARDS with severe and moderate-to-severe symptoms. Exolfo was administered intravenously in a single dose, with no harmful effects identified 72 h after administration. The study showed an 83% survival rate, cytokine storm downregulation, substantial hypoxia recovery, and immune restoration. Exosome-involved COVID-19 clinical trials are ongoing in the United States, China, Turkey, and Russia (NCT04276987, NCT04493242, NCT04991240, ChiCTR2000030484, ChiCTR2000030261, NCT04384445, NCT04389385).

The advantage of MSC-derived secretome therapy is its two forms (inhalable and injectable formulation) for potential clinical applications[9]. Both forms exist as freeze-dried powder and can be used in patients with a severe COVID-19 lung infection. An examination of the inhalable secretome form for COVID-19 pneumonia was conducted in a clinical trial (NCT04276987) in China, and its tolerance was examined in healthy individuals (NCT04313647)[8]. Assessment of its therapeutic efficacy demands further randomized controlled trials with comprehensive delivery of the exosomes. Moreover, in addition to MSC-derived secretomes, the secretome of oral tissue stem cells is also considered to have a therapeutic impact in infected cases due to their immunoregulatory and anti-inflammatory characteristics. Non-invasive therapy is superior to invasive therapy in prophylaxis and results in minimum risk of the treatment process, and prevents COVID-19, offering a novel immunoregulatory pathway for COVID-19 therapy[65].

The potential role of exosomes in treating COVID-19 can be classified into three general divisions. First, instead of cell therapy, the exosomes derived from multiple MSCs are utilized. Second, particular mRNAs and miRNAs are incorporated into the exosomes. Third, exosomes could be used as drug carriers in the treatment of COVID-19[66]. The efficacy of stem cell-derived secretome therapies is the main focus of continuing clinical trials. Nonetheless, the effectiveness, safety, and long-term consequences of these therapies require further study.
CHALLENGES IN TREATING COVID-19 USING STEM CELL-DERIVED SECRETOME

Administering stem-cell EVs as a possible treatment for COVID-19 is supported by initial examinations. However, for the sake of scientific rationale, further understanding and justification of MSC-EVs and other EV treatment effects on COVID-19 are required. MSCs have demonstrated promising effects on COVID-19 pathogenesis. However, their resemblance to exosomes is uncertain. Regulation of the immune response is the main impact of MSC-EVs, rather than impeding it (Figure 2). By regulating the response, they moderate it. They also strengthen tolerance and improve homeostasis[67]. Although stimulating tolerance in graft-vs-host and other non-infectious diseases is effective, it sometimes has an adverse effect on replicating pathogens. Albeit in selected models, Escherichia coli and influenza infection did not escalate but other replicating bacteria and viruses can experience augmented uncontrollable infection due to tolerance stimulation[68].

Prior to MSC-EVs application in COVID-19 patients, particular concerns must be resolved. The EV isolation method, purification, and characterization must be determined. These have a meaningful impact on the examination results, and in clinical trials can generate obscure conclusions. These parameters involve the origin of MSC-EVs. MSCs (as a heterogeneous cell essence) are derivable from various tissues. Even derivatives of the same tissue vary in inter-individual and clone-specific functions[69]. In fact, comparing four MSC-EV samples harvested from separate donor-derived bone marrow placed in conditioned media revealed substantial cytokine component differences[70]. Moreover, in one study MSCs from young individuals (suffering from acute lung injury), but not elderly individuals, attenuated lipopolysaccharide-induced acute lung injury[71].

The problem of EV heterogeneity from dissimilar resources, preparations, and other issues can be solved by manufacturing immortal MSC colonies that can be deliberately examined for potency and production of EVs[72]. Besides the problem of derived MSC-EVs from different origins, another consideration is their various responses to different disease conditions. Therefore, it is unclear whether the divergent immune response regulation of exosomes is due to tissue specificity. Regardless of the immunoregulatory properties, MSC-EVs seem to influence additional biological mechanisms with therapeutic functions[73] and other probable unanticipated effects. Recently conducted studies demonstrated that adipose-derived MSC-EVs had higher thrombogenic traits than bone marrow-derived MSC-EVs[74]. Accordingly, the origin of parental cells can potentially result in a higher thrombosis risk. The complement pathway stimulation accompanied by the procoagulant condition in a fraction of serious COVID-19 cases can result in the devastating microvascular injury syndrome[49], and the application of MSC-EVs may be ineffective.

A further hurdle is sustaining their stability and productivity with time[75]. Exosomes, from MSCs at -80°C, are viable for an extended period. Nevertheless, exosome clustering appears after storage due to freeze-thaw cycles. Moreover, preservation at low temperature during transportation and handling contributes to translational application impediments[76].

The manufacture of EVs requires living cells that are cultured under GMP-compliant (good manufacturing practice-compliant) processes to conserve safety and quality standards criteria. Hence, EV manufacture resembles the ethical and scientific guidelines used for MSCs. Also, basing any therapy on EVs extracted from MSCs requires the approval of national regulatory agencies to ensure their safety and productivity. The International Society for Cellular and Gene Therapies and the International Society for Extracellular Vesicles decrease the risk of critical side effects by deliberately weighing the probable benefits and risks of MSC-EVs for COVID-19 and have already provided preclinical data in connection with animal models and relevant MSC clinical trial-derived data. Additionally, they urge deliberate EV use evaluation by rational clinical trial design, applying well-characterized EV preparations generated according to strict GMP conditions and under proper regulatory oversight[77].

CONCLUSION

MSC-derived secretome demonstrates beneficial results as a cell-free therapy for acute and chronic lung diseases. It exhibits immunoregulatory, pro-angiogenic, anti-inflammatory, regenerative, and anti-protease characteristics. Due to the prominent role of
Figure 2 Immunomodulation by the stem cell secretome. The behavior of immune cells is altered by the secretion of various immunomodulatory factors. Mesenchymal stem cells (MSCs) secrete SOD3 which inhibits neutrophil activation and infiltration of leukocytes. Also, MSCs secrete PGE2, SDF-1, and interleukin (IL)-10 and lead to macrophage polarization to the M2 phenotype. Furthermore, MSCs induce HLAG5 secretion (shift to CD4[+]/CD25[+]/T reg population), IL-4 secretion (shift to Th2 population), constitutive expression of transforming growth factor β1 (TGF-β1), HGF, COX2, IL-10, IDO, and PGE2 which in turn inhibit T-cell proliferation. The secretion of IL-2 prevents inactivated NK cell proliferation, and secretion of HLAG5, TGF-β1, IDO, and PGE2 prevent cytokine secretion. In addition, MSCs cause B cells arrest in the G0/G1 phase and reduce the level of circulating immunoglobulins and secretion of CXCR4, CXCR5, CXCR7 by B cells. Due to the high amount of ISG gene expression, MSCs induce an antiviral response in the lungs. After secretion of immunomodulatory, anti-inflammatory, and microRNAs mediators and their extracellular vesicles-mediated transfer, MSCs lead to regulatory lymphocyte and M2 macrophage production. Differentiation of MSCs to various lung epithelial cells or differentiation of host tissue-resident stem cells lead to the secretion of numerous growth factors and angiogenic factors to stimulate revascularization, and consequently repair structural injury. Recovery of alveolar cell functions, their ATP stores, and metabolic capacity is feasible by direct transfer of functional mitochondria. Anti-fibrotic cytokines in high amounts reduce collagen fibers and subsequently hyper-inflammation and oxidative stress. A combination of anti-viral drugs with the immunomodulatory cargo of MSCs exosomes is a promising intervention tool in disease treatment [54]. Remdesivir is the best example of drug loading on exosomes [79]. Nevertheless, more studies are needed to clarify the exact mechanism of the specificity, targeted delivery and safety of exosomes.

MSC-derived exosomes in inhibiting the inflammatory response and in injured tissue regeneration, they may be a valuable therapeutic option in COVID-19-related pneumonia. Additionally, exosomes are considered potential nanocarriers, biomarkers, and vaccines for COVID-19 treatment. Regarding the concerns on their outcome, it is important to assess the risk when utilizing MSC-exosomes for COVID-19 by determining applicable preclinical findings in vivo models.

Upon reaching the lungs, the inhalable administered secretome encounters three main hurdles (anatomical, pathological, and immunological) and exerts their therapeutic effects. Although, the advantages of cell-free therapy are significant, it is considered a novel approach and requires secretome optimization and standardization via a comprehensive investigation of its components, dosing conditions, quality control, formulation and preparation process, and long-term storage strategies. Further data on the therapeutic mechanism, new formulation strategies, scalable and GMP-compliant isolation processes, and the capability to convey EVs and soluble proteins through non-invasive pathways of administration are required. These challenges will be groundbreaking, and will provide impressive clinical outcomes in the treatment of acute and chronic lung diseases.

ACKNOWLEDGEMENTS

The authors are thankful to Tabriz University of Medical Sciences, Tabriz, Iran.
REFERENCES

1 Velavan TP, Meyer CG. The COVID-19 epidemic. Trop Med Int Health 2020; 25: 278-280 [PMID: 32052514 DOI: 10.1111/tmi.13383]

2 Yang L, Liu S, Liu J, Zhang Z, Wan X, Huang B, Chen Y, Zhang Y. COVID-19: immunopathogenesis and Immunotherapeutics. Signal Transduct Target Ther 2020; 5: 128 [PMID: 32712629 DOI: 10.1038/s41992-020-00243-2]

3 Cao X. COVID-19: immunopathology and its implications for therapy. Nat Rev Immunol 2020; 20: 269-270 [PMID: 32275594 DOI: 10.1038/s41577-020-0306-3]

4 Nishiga M, Wang DW, Han Y, Lewis DB, Wu JC. COVID-19 and cardiovascular disease: from basic mechanisms to clinical perspectives. Nat Rev Cardiol 2020; 17: 543-558 [PMID: 32690910 DOI: 10.1038/s41569-020-0413-9]

5 Verma K, Amitabhi, Prasad DN, Kumar B, Kohli E. Brain and COVID-19 Crosstalk: Pathophysiological and Psychological Manifestations. ACS Chem Neurosci 2020; 11: 3194-3203 [PMID: 33006881 DOI: 10.1021/acschemneuro.0c00446]

6 Galanopoulos M, Gikers F, Doukatas A, Karianakis G, Pontas C, Tsoukalas N, Viazis N, Liatsos C, Mantzaris GJ. COVID-19 pandemic: Pathophysiology and manifestations from the gastrointestinal tract. World J Gastroenterol 2020; 26: 4579-4588 [PMID: 32884218 DOI: 10.3748/wjg.v26.i31.4579]

7 Ahmadian E, Hosseiniyan Khaiti SM, Razi Soofiyan S, AbediAzar S, Shoja MM, Ardalan M, Zununi Vahedi S. Covid-19 and kidney injury: Pathophysiology and molecular mechanisms. Rev Med Virol 2021; 31: e2176 [PMID: 33022818 DOI: 10.1002/rmv.2176]

8 Bari E, Ferrarotti I, Saracino L, Perteghella S, Torre ML, Corsico AG. Mesenchymal Stromal Cell Secretome for Severe COVID-19 Infections: Premises for the Therapeutic Use. Cells 2020; 9 [PMID: 32283815 DOI: 10.3390/cells9040924]

9 Bari E, Ferrarotti I, Torre ML, Corsico AG, Perteghella S. Mesenchymal stem/stromal cell secretome for lung regeneration: The long way through “pharmaceuticalization” for the best formulation. J Control Release 2019; 309: 11-24 [PMID: 31326462 DOI: 10.1016/j.jconrel.2019.07.022]

10 Sengupta V, Sengupta S, Lazo A, Woods P, Nolan A, Bremer N. Exosomes Derived from Bone Marrow Mesenchymal Stem Cells as Treatment for Severe COVID-19. Stem Cells Dev 2020; 29: 747-754 [PMID: 32380908 DOI: 10.1089/scd.2020.0080]

11 Liang B, Chen J, Li T, Wu H, Yang W, Li Y, Li J, Yu C, Nie F, Ma Z, Yang M, Xiao M, Nie P, Gao Y, Qian C, Hu M. Clinical remission of a critically ill COVID-19 patient treated by human umbilical cord mesenchymal stem cells: A case report. Medicine (Baltimore) 2020; 99: e21429 [PMID: 32756149 DOI: 10.1097/MD.0000000000021429]

12 Leng Z, Zhu R, Hou W, Feng Y, Yang Y, Han Q, Shan G, Meng F, Du D, Wang S, Fan J, Wang W, Deng L, Shi H, Li H, Hu Z, Zhang F, Gao J, Liu H, Li X, Zhao Y, Yin K, He X, Gao Z, Yang Y, Yang B, Jin R, Stambler I, Lim LW, Su H, Moskalev A, Cano A, Chakrabarti S, Min KJ, Ellison-Hughes G, Caruso C, Jin J, Zhao RC. Transplantation of ACE2 Mesenchymal Stem Cells Improves the Outcome of Patients with COVID-19 Pneumonia. Aging Dis 2020; 11: 216-228 [PMID: 32257537 DOI: 10.14356/AD.2020.0228]

13 Meng F, Xu R, Wang S, Xu Z, Zhang C, Li Y, Yang T, Shi L, Fu J, Jiang T, Huang L, Zhao P, Yuan X, Fan X, Zhang JY, Song J, Zhang D, Jiao Y, Liu L, Zhou C, Mauerer M, Zunila A, Shi M, Yang FS. Human umbilical cord-derived mesenchymal stem cell therapy in patients with COVID-19: a phase 1 clinical trial. Signal Transduct Target Ther 2020; 5: 172 [PMID: 32855385 DOI: 10.1038/s41392-020-00286-5]

14 Sadeghi S, Soudi S, Shafiee A, Hashemi SM. Mesenchymal stem cell therapies for COVID-19: Current status and mechanism of action. Life Sci 2020; 262: 118493 [PMID: 32979360 DOI: 10.1016/j.lifesci.2020.118493]

15 Caplan AI. Mesenchymal Stem Cells: Time to Change the Name! Stem Cells Transl Med 2017; 6: 1445-1451 [PMID: 28452204 DOI: 10.1002/sctm.17-0051]

16 Krasnodembskaya A, Song Y, Fang X, Gupta N, Serikov V, Lee JW, Matthay MA. Antibacterial effect of human mesenchymal stem cells is mediated in part from secretion of the antimicrobial peptide LL-37. Stem Cells 2010; 28: 2229-2238 [PMID: 20945332 DOI: 10.1002/sctm.2044]

17 Ranganath SH, Levy O, Inamdar MS, Karp JM. Harnessing the mesenchymal stem cell secretome for the treatment of cardiovascular disease. Cell Stem Cell 2012; 10: 244-258 [PMID: 22385653 DOI: 10.1016/j.stem.2012.02.002]

18 Deffune E, Prudenciatti A, Moroz A. Mesenchymal stem cell (MSC) secretome: A possible therapeutic strategy for intensive-care COVID-19 patients. Med Hypotheses 2020; 142: 109769 [PMID: 32371362 DOI: 10.1016/j.mehy.2020.109769]

19 Crippelli B, Chlapanidas T, Perteghella S, Lucarelli E, Pasucci L, Brini AT, Ferrero R, Marazzi M, Pessina A, Torre ML, Italian Mesenchymal Stem Cell Group (GISM). Mesenchymal stem/stromal cell extracellular vesicles: From active principle to next generation drug delivery system. J Control Release 2017; 262: 104-117 [PMID: 28736264 DOI: 10.1016/j.jconrel.2017.07.023]

20 Court AC, Le-Gatt A, Luz-Crawford P, Parra E, Aliaga-Tobar B, Bátiz LF, Contreras RA, Ortizúzar MI, Kurte M, Elizondo-Vega R, Maracaia-Coutinho V, Pino-Lagos K, Figueroa FE, Khoury M. Mitochondrial transfer from MSCs to T cells induces Treg differentiation and restricts inflammatory response. EMBO Rep 2020; 21: e48052 [PMID: 31984629 DOI: 10.15252/embr.201948052]

21 Spees JL, Olson SD, Whitney MJ, Prockop DJ. Mitochondrial transfer between cells can rescue
aerobic respiration. Proc Natl Acad Sci USA 2006; 103: 1283-1288 [PMID: 16432190 DOI: 10.1073/pnas.0510511103]

22 Worthington EN, Hagood JS. Therapeutic Use of Extracellular Vesicles for Acute and Chronic Lung Disease. Int J Mol Sci 2020; 21 [PMID: 32230828 DOI: 10.3390/ijms211072318]

23 Khatri M, Richardson LA, Meulia T. Mesenchymal stem cell-derived extracellular vesicles attenuate influenza virus-induced acute lung injury in a pig model. Stem Cell Res Ther 2018; 9: 17 [PMID: 29376639 DOI: 10.1186/s13287-018-0774-8]

24 Tang XD, Shi L, MONSEL A, LI XJ, ZHU HL, ZHU YG, QU JM. Mesenchymal Stem Cell Microvesicles Attenuate Acute Lung Injury in Mice Parly Mediated by Ang-1 mRNA. Stem Cells 2017; 35: 1849-1859 [PMID: 28376568 DOI: 10.1002/stem.2619]

25 Hu S, Park J, LEE A, Lee J, Zhang X, Tao Q, Lee JW. Mesenchymal Stem Cell Microvesicles Restore Protein Permeability Across Primary Cultures of Injured Human Lung Microvascular Endothelial Cells. Stem Cells Transl Med 2018; 7: 615-624 [PMID: 29737632 DOI: 10.1002/sctm.17-0278]

26 Zhu X, Badawi M, Pomeroy S, Sutarisa DS, Xie Z, Baek A, Jiang J, Elgamal OA, MO X, Perle K, Chalmers J, Schnittgen TD, Phelps MA. Comprehensive toxicity and immunogenicity studies reveal minimal effects in mice following sustained dosing of extracellular vesicles derived from HEK293T cells. J Extracell Vesicles 2017; 6: 1324730 [PMID: 28717420 DOI: 10.1080/20013078.2017.1324730]

27 Lopes-Pacheco M, Robba C, Rocco PRM, Pelosi P. Current understanding of the therapeutic benefits of mesenchymal stem cells in acute respiratory distress syndrome. Cell Biol Toxicol 2020; 36: 83-102 [PMID: 31485828 DOI: 10.1007/s40651-019-09493-5]

28 Keshkatkar S, Azarpina N, Halhameani MH. Mesenchymal stem cell-derived extracellular vesicles: novel frontiers in regenerative medicine. Stem Cell Res Ther 2018; 9: 63 [PMID: 29523213 DOI: 10.1186/s13287-018-0791-7]

29 Lou G, Chen Z, Zheng M, Liu Y. Mesenchymal stem cell-derived exosomes as a new therapeutic strategy for liver diseases. Exp Mol Med 2017; 49: e346 [PMID: 28620221 DOI: 10.1038/emmm.2017.63]

30 Mahmoudi M, Taghavi Farahabadi M, Hashemi SM. Exosomes: Mediators of Immune Regulation. Immunoregulation 2019; 2: 3-8

31 EL. Andaloussi S, Mäger I, Breakfeld XO, Wood MJ. Extracellular vesicles: biology and emerging therapeutic opportunities. Nat Rev Drug Discov 2013; 12: 347-357 [PMID: 23584393 DOI: 10.1038/nrd3978]

32 Shah TG, Predescu D, Predescu S. Mesenchymal stem cells-derived extracellular vesicles in acute respiratory distress syndrome: a review of current literature and potential future treatment options. Clin Transl Med 2019; 8: 25 [PMID: 31512000 DOI: 10.1186/s40169-019-0242-9]

33 Lai P, Chen X, Guo L, Wang Y, Liu X, Liu Y, Zhou T, Huang T, Geng S, Luo C, Huang X, Wu S, Ling W, Du X, He C, Weng J. A potent immunomodulatory role of exosomes derived from mesenchymal stromal cells in preventing cGVHD. J Hematol Oncol 2018; 11: 135 [PMID: 30526632 DOI: 10.1186/s13045-018-0680-7]

34 Álvarez V, Sánchez-Margallo FM, Macías-García B, Gómez-Serrano M, Jorge I, Vázquez J, Blázquez R, Casado JG. The immunomodulatory activity of extracellular vesicles derived from endometrial mesenchymal stem cells on CD4+ T cells is partially mediated by TGFbeta. Immunoregulation 2018; 12: 398-408 [PMID: 30382799 DOI: 10.1089/scd.2018.0015]

35 Domenis R, Cifí A, Quaglia S, Pistics C, Moretti M, Vicario A, Parodi PC, Fabris M, Niazi KR, Soon-Shiong P, Curcio F. Pro inflammatory stimuli enhance the immunosuppressive functions of adipose mesenchymal stem cells-derived exosomes. Sci Rep 2018; 8: 13325 [PMID: 30190615 DOI: 10.1038/s41598-018-31707-9]

36 Yang JH, LIO FX, Wang JH, CHENG M, Wang SF, Xu DH. Mesenchymal stem cells and mesenchymal stem cell-derived extracellular vesicles: Potential roles in rheumatic diseases. World J Stem Cells 2020; 12: 688-705 [PMID: 32843922 DOI: 10.4252/wjsc.v12.i7.688]

37 Ji L, Bao L, Gu Z, ZHOU Q, LIANG Y, ZHENG Y, Xu Y, ZHANG X, Feng X. Comparison of immunomodulatory properties of exosomes derived from bone marrow mesenchymal stem cells and dental pulp stem cells. ImmunoL Res 2019; 67: 432-442 [PMID: 31407157 DOI: 10.1007/s12026-019-09085-6]

38 He P. At0291e the effect of human umbilical cord mesenchymal stem cells-derived exosomes on chemokine production in collagen-induced arthritis rats. Ann Rheum Dis 2019; 78: 1606

39 Mardpour S, Hamidieh AA, Taleahmad S, Sharifzad F, Taghkhani A, Baharvand H. Interaction between mesenchymal stromal cell-derived extracellular vesicles and immune cells by distinct protein content. J Cell Physiol 2019; 234: 8249-8258 [PMID: 30078105 DOI: 10.1002/jcp.27669]

40 Baharlouei H, Azimi M, Salehi Z, Izad M. Mesenchymal Stem Cell-Derived Exosomes: A Promising Therapeutic Ace Card to Address Autoimmune Diseases. Int J Stem Cells 2020; 13: 13-23 [PMID: 31887849 DOI: 10.15283/jsc.19108]

41 Jayaramaaya K, Mahalaxmi I, Subramaniam MD, Raj N, Dayem AA, Lin KM, Kim SJ, An JY, Lee Y, Choi Y, Raj A, Cho SG. Vellingiri B. Immunomodulatory effect of mesenchymal stem cells and mesenchymal stem-cell-derived exosomes for COVID-19 treatment. BMJ Rep 2020; 53: 400-412 [PMID: 32731913 DOI: 10.5483/BMRep.2020.53.8.121]
Ardalan M et al. Stem-cell secretomes and COVID-19

43 Monsel A, Zhu YG, Gennai S, Hao Q, Hu S, Rouby JJ, Rosenzwaig M, Matthy MA, Lee JW. Therapeutic Effects of Human Mesenchymal Stem-cell Derived Microvesicles in Severe Pneumonia in Mice. Am J Respir Crit Care Med 2015; 192: 324-336 [PMID: 26067592 DOI: 10.1164/rccm.201410-1766OC]

44 Chen J, Hu C, Chen L, Tang L, Zhu Y, Xu X, Gao H, Lu X, Yu L, Dai X, Xiang C, Li L. Clinical Study of Mesenchymal Stem Cell Treatment for Acute Respiratory Distress Syndrome Induced by Epidemic Influenza A (H7N9) Infection: A Hint for COVID-19 Treatment. Engineering (Beijing) 2020; 6: 1153-1161 [PMID: 32292627 DOI: 10.1016/j.eng.2020.02.006]

45 Kuate S, Cinaat J, Doerri HW, Uberal K. Exosomal vaccines containing the S protein of the SARS coronavirus induce high levels of neutralizing antibodies. Virology 2007; 362: 26-37 [PMID: 17258782 DOI: 10.1016/j.virol.2006.12.011]

46 Sun DZ, Abelson B, Babbar P, Danuser MS. Harnessing the mesenchymal stem cell secretome for regenerative urology. Nat Rev Urol 2019; 16: 363-375 [PMID: 30923338 DOI: 10.1038/s41585-019-0169-3]

47 Morrison TJ, Jackson MV, Cunningham EK, Kissmغنigen A, McAlayef DF, O’Kane CM, Krasnoodembskaya AD. Mesenchymal Stromal Cells Mediate Corticosteroids in Clinically Relevant Lung Injury Models by Extracellular Vesicle Mitochondrial Transfer. Am J Respir Crit Care Med 2017; 196: 1275-1286 [PMID: 28598224 DOI: 10.1164/rccm.201701-0170OC]

48 Ji F, Li L, Li Z, Jin Y, Liu W. Mesenchymal stem cells as a potential treatment for critically ill patients with coronavirus disease 2019. Stem Cells Transl Med 2020; 9: 813-814 [PMID: 32320535 DOI: 10.1002/stem.20083]

49 Marginol S, Sadovsky Y. The biology of extracellular vesicles: The known unknowns. PLoS Biol 2019; 17: e3000363 [PMID: 31318874 DOI: 10.1371/journal.pbio.3000363]

50 Benuke J, Kremer S, Shahzad T, Chao CM, Böttcher-Friebertshäuser E, Morty RE, Bellussi S, Ehrhardt H. MSC Based Therapies-New Perspectives for the Injured Lung. J Clin Med 2020; 9 [PMID: 32138309 DOI: 10.3390/jcm9030682]

51 Willis GR, Fernandez-Gonzalez A, Anastas J, Vitali SH, Liu X, Ericsson M, Kwong A, Mitsialis SA, Kouremanis A, Mesenchymal Stromal Cell Exosomes Ameliorate Experimental Bronchopulmonary Dysplasia and Restore Lung Function through Macrophage Immunomodulation. Am J Respir Crit Care Med 2018; 197: 104-116 [PMID: 28853608 DOI: 10.1164/rccm.201705-0925OC]

52 Fujita Y, Kosaka N, Araya J, Kuwano K, Ochiya T. Extracellular vesicles in lung microenvironment and pathogenesis. Trends Mol Med 2015; 21: 533-542 [PMID: 26231094 DOI: 10.1016/j.molmed.2015.07.004]

53 Gennai S, Monsel A, Hao Q, Park J, Matthy MA, Lee JW. Microvesicles Derived From Human Mesenchymal Stem Cells Restore Alveolar Fluid Clearance in Human Lungs Rejected for Transplantation. Am J Transplant 2015; 15: 2404-2412 [PMID: 25847030 DOI: 10.1111/ajt.13271]

54 Pinky, Gupta S, Krishnakumar V, Sharma Y, Dinda AK, Mohanty S. Mesenchymal Stem Cell Derived Exosomes: a Nano Platform for Therapeutics and Drug Delivery in Combating COVID-19. Stem Cell Rev Rep 2021; 17: 33-43 [PMID: 32661867 DOI: 10.1007/s12015-020-10002-z]

55 Barani B, Rajasingh S, Rajasingh J. Exosomes: Outlook for Future Cell-Free Cardiovascular Disease Therapy. Adv Exp Med Biol 2017; 998: 285-307 [PMID: 28936747 DOI: 10.1007/978-981-10-4397-0_19]

56 Tsuji K, Kitamura S, Wada J. Secretomes from Mesenchymal Stem Cells against Acute Kidney Injury: Possible Heterogeneity. Stem Cells Int 2018; 2018: 8693137 DOI: 10.1155/2018/8693137

57 Liu L, Jin X, Hu CF, Li R, Zhou Z, Shen CX. Exosomes Derived from Mesenchymal Stem Cells Rescue Myocardial Ischaemia/Reperfusion Injury by Inducing Cardiomyocyte Autophagy Via AMPK and Akt Pathways. Cell Physiol Biochem 2017; 43: 52-68 [PMID: 28848091 DOI: 10.1155/900480317]

58 Xuan W, Khan M, Ashraf M. Extracellular Vesicles From Notch Activated Cardiac Mesenchymal Stem Cells Promote Myocyte Proliferation and Neoangiogenesis. Front Cell Dev Biol 2020; 8: 11 [PMID: 32154243 DOI: 10.3389/fcell.2020.00011]

59 Zou L, Ma X, Lin S, Wu B, Chen Y, Peng C. Bone marrow mesenchymal stem cell-derived exosomes protect against myocardial infarction by promoting autophagy. Exp Ther Med 2019; 18: 2574-2582 [PMID: 31555366 DOI: 10.3892/etm.2019.7873]

60 Kjellberg G, Holm M, Lindvall G, van der Linden J. Calculation Algorithm Reduces Probe Toning Doses Without Increasing Blood Loss or the Transfusion Rate in Cardiac Surgery: Results of a Randomized Controlled Trial. J Cardiothorac Vasc Anesth 2019; 33: 985-992 [PMID: 30206011 DOI: 10.1053/j.jvca.2018.07.044]

61 Bansal M. Cardiovascular disease and COVID-19. Diabetes Metab Syndr 2020; 14: 247-250 [PMID: 32247212 DOI: 10.1016/j.dsx.2020.03.013]

62 Yao XH, Li TY, He ZC, Ping YF, Liu HW, Yu SC, Mou HM, Wang LH, Zhang HR, Fu WJ, Luo T, Liu F, Guo QN, Chen C, Xiao HL, Guo HT, Lin S, Xiang DF, Shi Y, Pan QG, Li QR, Huang X, Cui Y, Liu XZ, Tang W, Pan PF, Huang XQ, Ding YQ, Bian YC. [A pathological report of three COVID-19 cases by minimal invasive autopsies]. Zhonghua Bing Li Xue Za Zhi 2020; 49: 411-417 [PMID: 32172546 DOI: 10.3760/cma.j.cn12151-20200312-00193]

63 Song Y, Dou H, Li X, Zhao X, Li Y, Liu D, Ji J, Liu F, Ding L, Ni Y, Hou Y. Exosomal miR-146a Contributes to the Enhanced Therapeutic Efficacy of Interleukin-1β-Primed Mesenchymal Stem Cells Against Sepsis. Stem Cells 2017; 35: 1208-1221 [PMID: 28090688 DOI: 10.1002/stem.2564]
64 **MohammadiPOOR A**, Antebi B, Batchinsky AI, Cancio LC. Therapeutic potential of products derived from mesenchymal stem/stromal cells in pulmonary disease. *Respir Res* 2018; 19: 218 [PMID: 30413155 DOI: 10.1186/s12931-018-0921-x]

65 **DiOMede F**, Marconi GD, Fonticoli L, Pizzicannella J, Trubiani O. Stem Cells Secretome from Oral Tissue Could Represent a Promising Therapeutic Approach in COVID-19-Disease? *Int J Mol Sci* 2020; 21 [PMID: 32957696 DOI: 10.3390/ijms21186833]

66 **Rezakhani L**, Kelishadrokh AF, Soleimaniandezeh A, Rahmati S. Mesenchymal stem cell (MSC)-derived exosomes as a cell-free therapy for patients infected with COVID-19: Real opportunities and range of promises. *Chem Phys Lipids* 2021; 234: 105009 [PMID: 33189639 DOI: 10.1016/j.chemphyslip.2020.105009]

67 **Giebel B**, Herrmann DM. Identification of the right cell sources for the production of therapeutically active extracellular vesicles in ischemic stroke. *Ann Transl Med* 2019; 7: 188 [PMID: 31205906 DOI: 10.21037/atm.2019.03.49]

68 **Hao Q**, Gudapati V, Monsel A, Park JH, Hu S, Kato H, Lee JH, Zhou L, He H, Lee JW. Mesenchymal Stem Cell-Derived Extracellular Vesicles Decrease Lung Injury in Mice. *J Immunol* 2019; 203: 1961-1972 [PMID: 31451675 DOI: 10.4049/jimmunol.1801554]

69 **Phimney DG**. Functional heterogeneity of mesenchymal stem cells: implications for cell therapy. *J Cell Biochem* 2012; 113: 2806-2812 [PMID: 22511358 DOI: 10.1002/jcb.24166]

70 **Kordelas L**, Rebmann V, Ludwig AK, Radlje S, Ruesing J, Doepnner TR, Epple M, Horn PA, Beelen DW, Giebel B. MSC-derived exosomes: a novel tool to treat therapy-refractory graft-versus-host disease. *Leukemia* 2014; 28: 970-973 [PMID: 24445666 DOI: 10.1038/leu.2014.41]

71 **Huang R**, Qin C, Wang J, Hu Y, Zheng G, Qiu G, Ge M, Tao H, Shu Q, Xu J. Differential effects of extracellular vesicles from aging and young mesenchymal stem cells in acute lung injury. *Aging (Albany NY)* 2019; 11: 796-8014 [PMID: 31575829 DOI: 10.18632/aging.102314]

72 **Chen TS**, Arslan F, Yin Y, Tan SS, Lai RC, Choo AB, Padmanabhan J, Lee CN, de Kleijn DP, Lim SK. Enabling a robust scalable manufacturing process for therapeutic exosomes through oncogenic immortalization of human ESC-derived MSCs. *J Transl Med* 2011; 9: 47 [PMID: 21513579 DOI: 10.1186/1479-5876-9-47]

73 **Arslan F**, Lai RC, Snets MB, Akerooy L, Choo A, Aguor EN, Timmers L, van Rijen HV, Doevendans PA, Pasterkamp G, Lim SK, de Kleijn DP. Mesenchymal stem cell-derived exosomes increase ATP levels, decrease oxidative stress and activate PI3K/Akt pathway to enhance myocardial viability and prevent adverse remodeling after myocardial ischemia/reperfusion injury. *Stem Cell Res* 2013; 10: 301-312 [PMID: 23399448 DOI: 10.1016/j.scr.2013.01.002]

74 **Chance TC**, Rathbone CR, Kamucheka RM, Peltier GC, Cap AP, Bynum JA. The effects of cell type and culture condition on the procoagulant activity of human mesenchymal stromal cell-derived extracellular vesicles. *J Trauma Acute Care Surg* 2019; 87: S74-S82 [PMID: 31246910 DOI: 10.1097/TA.0000000000002223]

75 **Hood JL**. Post isolation modification of exosomes for nanomedicine applications. *Nanomedicine (Lond)* 2016; 11: 1745-1756 [PMID: 27348448 DOI: 10.2217/nnm-2016-0102]

76 **Jeyaram A**, Jay SM. Preservation and Storage Stability of Extracellular Vesicles for Therapeutic Applications. *AAPS J* 2017; 20: 1 [PMID: 29181730 DOI: 10.1208/s12248-017-0160-y]

77 **Bürger V**, Weiss DJ, Anderson JD, Borriès FE, Bussolati B, Carter DRF, Dominici M, Falcón-Pérez JM, Gimona M, Hill AF, Hoffman AM, de Kleijn D, Levine BL, Lim R, Lötvall J, Mitsialis SA, Mongiu-Tortajada M, Muraca M, Nieuwland R, Nowocin A, O’Driscoll L, Ortiz LA, Phinney DG, Reischl I, Rohde E, Sanzenbacher R, Théry C, Toh WS, Witwer KW, Lim SK, Giebel B. International Society for Extracellular Vesicles and International Society for Cell and Gene Therapy statement on extracellular vesicles from aging and young mesenchymal stem cells in pulmonary disease. *Ann Transl Med* 2019; 7: 2806-2812 [PMID: 31451675 DOI: 10.4049/jimmunol.1801554]

78 **Shi Y**, Wang Y, Shao C, Huang J, Gan J, Huang X, Bucci E, Piacentini M, Ippolito G, Melino G. COVID-19 infection: the perspectives on immune responses. *Cell Death Differ* 2020; 27: 1451-1454 [PMID: 32205856 DOI: 10.1038/s41418-020-0530-3]

79 **Grein J**, Ohmagari N, Shin D, Diaz G, Asperges E, Castagna A, Feldt T, Green G, Green ML, Lescure FX, Nicastri E, Oda R, Yo K, Quiros-Roldan E, Studemeister A, Redinski J, Ahmed S, Bennett J, Chelliah D, Ching D, Chihara S, Cohen SH, Cunningham J, D’Arminio Monforte A, Ismail S, Kato H, Lapadula G, L’Her E, Maeno T, Majumder S, Massari M, Mora-Rillo M, Mutoh Y, Nguyen D, Vervij E, Zoufaly A, Otsuji AO, DeZure A, Zhao Y, Zhong L, Chakkalingam A, Elboudwarej E, Telep L, Timbs L, Henne I, Sellers S, Cao H, Tan SK, Winterbourne L, Desai P, Mera R, Gaggar A, Myers RP, Brainard DM, Childs R, Flanagan T. Compassionate Use of Remdesivir for Patients with Severe Covid-19. *N Engl J Med* 2020; 382: 2327-2336 [PMID: 32275812 DOI: 10.1056/NEJMoa2007016]
