Review

Current knowledge about the connection between health status and gut microbiota from birth to elderly. A narrative review

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1. Abstract

The human body is colonized from the birth by a large number of microorganisms. This will constitute a real “functional microbial organ” that is fundamental for homeostasis and therefore for health in humans. Those microorganisms. The microbial populations that colonize humans creating a specific ecosystem they have been collectively referred to as “human microbiota” or “human normal microflora”. The microbiota play an important pathophysiological role in the various locations of the human body. This article focuses on one of the most important, that is the enteric microbiota. The composition (quantitative and qualitative) of microbes is analyzed in relation to age and environment during the course of human life. It also highlights eubiosis and dysbiosis as key terms for its role in health and disease. Finally, it analyzes its bi-directional relationship with the microbiota of the lungs, skin and that of the brain, consequently for the whole central and peripheral nervous system for the maintenance of health in the human body.

2. Introduction

The gastrointestinal tract is undoubtedly the main place for the growth of microorganisms in the human body and according to more recent estimates about $3.8 \times 10^{13}$ bacteria colonize the large intestine of a 70 kg human being aged between 20 and 30 years, representing 0.3% of the total body weight. The composition (species/population) of microbes that “live” in the intestine is unique for each person. Analysis of the 16s rDNA gene in fecal samples is often used to study the gut microbiota. The development of DNA sequencing technologies along with the development of bioinformatics have contributed to the study of the gut
The various actions/effects of the human gastrointestinal microbiota in the large intestine.

3. Human gut microbiota composition

The amount of germs can be different along the digestive system, from low concentrations for the stomach, duodenum, jejunum and ileum to high concentrations in the colon. In fact, in the colon stand 1011–1013 microorganisms that for the most part belong to various genes of bacteria. About 90% of germs are found in the colon, the last part of the digestive tract that serves as an anaerobic bioreactor. There are various reasons that can help great development for bacterial quantity and variability in the colon. The main factors then are (a) the pH around the neutrality, (b) the decrease of bile, (c) the absence of pancreatic juice and (d) the slow transit in the intestinal lumen of the colon that helps the multiplication and the metabolism through the fermentation of the existing nutrient substrates, and that come by the diet and/or exudates produced by the intestinal epithelium (Fig. 1) [1, 3–5].

The microbiota with its components (mainly bacterial) plays a specific role in normal development in the structures of the immune system network to perform its functions efficiently. To achieve this, the organism with its immune system tolerates at the intestinal level all antigens that can help it (such as food, Simbiotic bacteria) and recognizes its action against pathogenic microorganisms. However, the organism benefits from the microbiota through some of the main functions such as: (a) mucus-protective and trophic effects in the epithelium, (b) formation of organic compounds such as the naphthoquinone (Vit. K), energy source by producing SCFAs from unabsorbed food residues, inhibition of pathogen growth, (c) preservation of the integrity of the intestinal lumen epithelium barrier, (d) help in metabolism during the presence of xenobiotics, and (e) the proper function of the immune response [1, 5].

In the human gastrointestinal tract, the composition of each microbiota for each person constitutes an organic “fingerprint”. Thus, the number and location along the lumen of bacterial components would be similar in subjects without specific pathologies. The genera Bacteroides, Firmicutes (genus Clostridium, Eubacterium) are predominant (represent at least ¾ of the microbiota), and after them Verrucomicrobia and Actinobacteria. The colon being a mostly microaerophilic or oxygen-free environment, most microbes are anaerobic. Within this microbiota are mostly overrepresented the Bacteroides, Gram-positive sporing (such as Ruminococcus, Eubacterium, Bifidobacterium, Peptostreptococcus, and others) and Gram-positive bacilli are mainly represented by the genus Clostridium. Instead, in the large intestine environmental anaerobic or aerotoxic conditions are present. Gram-positive bacilli are mainly represented by the genus Clostridium. To a lesser extent, anaerobic bacteria such as Enterobacteriaceae, Enterococci, Lactobacilli and Streptococci, necessary for microbial homeostasis, appear in the large intestine. But there are other species that have been cultivated (it is estimated that only 30% of the species can be cultivated with the available techniques), other species whose presence was only indicated by their characteristic DNA sequences and therefore the complexity of the gastrointestinal microbiota was revealed (Fig. 2) [1, 3, 4, 6].

Among the populations of the enteric microbiota we can also distinguish the category of ancient (methanogens), various eukaryotic species, viruses and especially bacteriophages, fungi (mainly yeasts). The composition between the lumen of the intestinal tract associated to the mucous membranes and that of the lumen have differences not only in the microbial composition (Fig. 3) [1, 7–9].
Fig. 2. The main genera of bacteria that have been identified in specific locations of the gastrointestinal tract and their corresponding number.

| Location          | Number of CFU/ml (Sample Genera)                                                                 |
|-------------------|-------------------------------------------------------------------------------------------------|
| Duodenum/jejunum  | $10^5$ - $10^6$ CFU/ml (Lactobacillus, Streptococcus, Enterobacteriaceae, Staphylococcus, yeasts) |
| Ileum/Caecum      | $10^3$ - $10^5$ CFU/ml (Bifidobacterium, Bacteroides, Lactobacillus, Streptococcus, Enterobacteriaceae, Staphylococcus, Clostridium, yeasts) |
| Colon             | $10^9$ - $10^{12}$ CFU/ml (Bacteroides, Eubacterium, Clostridium, Peptostreptococcus, Bifidobacterium, Lactobacillus, Enterobacteriaceae, Staphylococcus, yeasts) |

Fig. 3. Intestinal microbiota’s differences between the mucosa (where bacteria are attached) and the intestinal lumen (where bacteria lean on).

### 3.1 The evolution of microbiota from birth to old age

The germ-free model (GF) concept, is based on the fact that during the perinatal life the fetus lives in a sterile context and subsequently with the procedure of a non-vaginal but surgical delivery no longer has the opportunity the newborn to have a colonization of the microorganisms present in the mother that constitute the first microbiota development. Subsequently, the growth of intestinal germs begins immediately after birth and depends on the bacteria of the mother and the environment in which the child grows. In fact, these exposures to the microbiota belong to the first initial development of the newborn microbiota. This will have its importance in the health its subsequent, because from a bleached colonization of species and microbial number have been observed the development of diseases such as various allergic forms, bronchial asthma, increase of the corporeal fat up to infantile obesity, type 1 diabetes mellitus, neurological disorders and more [10, 11]. The composition of the microflora is therefore influenced mainly by age, environmental factors and the homeostasis of the immune system. After two years and in adulthood the microbiota remains almost constant and is characteristic of every individual. Several studies show that the composition of the microbiota is not similar to that of young age. There are no limits of time or age in which the composition of the microbiota changes, changes occur gradually over time. Perhaps the most important but also changing factor are the eating habits that shape the microbiota, which in turn affects the health of the elderly. Bacteroidetes predominate in newborns, while over time the composition changes gradually and in the elderly the species of the genus Firmicutes predominate (Fig. 4) [6, 12–14].

### 4. Microbial balance against colony development by pathogens

Several studies show that the composition of the microbiota is not like that of young age. The enteric microbiota, under conditions of normal interaction and operation (eubiosis), provokes a continuous stimulus to the immune system and this has because of a condition of “light normal intestinal inflammation”. This creates a directed organized activity barrier against “bad” germs that is opportunistic pathogens. In addition, the “good” bacteria of enteric biomass release both molecules that block proliferation and consuming the nutrients necessary for the survival of pathogenic germs, thus playing a second protective role [15]. In fact, it has been shown that in the presence of the some bacterial species (such as rectal *Eubacterium rectale*), these are able to harden the secretion to the level of the intestinal mucosa of the β-glucans (that are suitable for these bacteria) and that metabolizing them (eutrophic effect) and can avoid the overpopulation of pathogenic germs. We conclude that diet and other factors can determine changes in microbial composition hence the balance of both species and populations (Fig. 5) [1, 14, 16].
Fig. 4. The development of the gut microbiota (main genus and populations%) after birth.

Fig. 5. Factors affecting the composition of the gut microbiota: the intestinal microbiota is of particular importance for the maintenance of human health and vice versa for the mutually beneficial dynamic interaction between host microbes in the intestine. This happens with a cross-talk between the immune system and the microbial biomass that is recognized not “dangerous” (not pathogenic). In addition, diet, and the environment, especially after birth, played an important and precise role in the growth and selection of “good” microorganisms. The interaction takes place as well as through intestinal metabolites and the nervous system (through the secretion of neurotransmitters) which can modulate the normal microbial enteric biomass.

Immune system works by learning, that is, at the beginning of life it has the necessary components (cells, intercellular mediators, etc.). But it does not have available data from the environment that acquires the newborn during the first period of life by contact with the mother’s and other persons microorganisms and the surrounding habitat. Indeed, on that first period years an inadequate data, the mechanisms of regulation of the host immunity defense could be inadequate. Hence, the immune system will be against not only pathogenic microorganisms but also other factors such as pollen, various types of dust or food and more, causing acute and chronic allergic reactions. Microorganisms along with digestive enzymes, mucus layer, intestinal peristalsis and epithelial barrier help the body’s immune response. The activity of the microorganisms of the microbiota in eubiosis is to defend the organism are on the one hand to influence in a determining way the development of the intestinal immune mechanism (for which we have made reference to the trophic role) and on the other hand to prevent the possible invasion of pathogens by effect on them and/or “activating” the immune mechanism of the host [16–18].

In terms of natural immunity, it can distinguish “bad” from the “good” germs by recognizing the molecular models associated with PAMPs (Pathogen Associated Molecular Patterns) on microorganisms. More specifically, natural immunity cells using PRP (Pattern Recognition Receptors) detect PAMPs. PRPs (Pattern Recognition Receptors) are also involved in the activation of acquired immunity and release of cytokines. It is worth noting that there are many species PRPs, with Toll-like (TLR) receptors on the front line, are found in macrophages, neutrophils, dendritic cells, and epithelial cells of the intestinal mucosa. PAMPs recognized by PRP receptors are nothing more than, microorganism’s polysaccharides or monosaccharides, structural peptides, nucleic acids, lipoic acid (present in gram + bacteria) or fungal lipoproteins and glucans. However, since these molecules are also found in symbiotic microbes, we characterize them as MAMP (Microbe-Associated Molecular Patterns). So, through these MAMPs, it seems that symbiotic germs change the expression of Toll-like receptors (TLRs) in non-specific, immune response cells. Therefore, recognition of MAMPs triggers the activation of the Nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) pathway leading to increased cytokine secretion. It also sparks the activation of other helper and necessary molecules on APCs (Antigen-presenting cells), which will activate the T cells, that’s in the acquired immunity [16, 19]. Another mechanism of natural immunity that germs can modify is the amount of mucus produced by the goblet or caliciform cells in the intestine. Mucus can reduce infection by pathogens, by directly binding to them, to protect epithelial cells from secreted secretions of acid and lithic enzymes and to be the means through which to accumulate products of bacterial metabolism and activate the body’s defenses. As for the immunity acquired in the gastrointestinal tract, it is “based” mainly on the intestine’s lymphoid structure tissue (Peyer patches and lymph nodes) [16, 19]. It should be remembered that the intestine is the most important immune func-
tion organ of the individual, so much so that about 60% of the immune cells of the entire organism are located in the intestinal mucosa. Thus, the evolution and homeostasis of the intestinal immune system will mostly depend on the microorganisms present, through the development of lymphoid structures, the modulation of the differentiation of subpopulations of immune cells. Finally, mast cells forming a necessary core, having a fundamental role in nonspecific immunity (innate) because are at the forefront of the interface between the host and the intestinal lumen [16, 20].

4.1 The intestinal microbiota in health

The human body as host and the gut microbiota have evolved so that there is benefit to both sides. On the one hand, the host provides space, adequate conditions, and food to the microbiota to grow and this in turn generally contributes to the supply of useful substances and induces resistance to various infections. The enteric active microorganisms flora have a great value for the human’s health preservation. It carries out processes that the human body has not evolved and therefore does not have the ability to act autonomously. The “supreme organism” theory has generally been formulated to note the mutually beneficial interaction between host microbes in the gut. Microorganisms have protective and trophic roles and are also involved in host metabolic pathways and immune functions. Several studies have shown that, in order for the intestinal mucosa to assume its complete structure, it needs to be colonized by microorganisms. For example, mice raised in sterile environments have developed fewer vessels in their intestinal villi. Growth in a sterile environment has also shown that defective growth occurs in lymphoid tissue associated with the intestine and in the production of antibodies. In addition, fewer Peyer patches develop in the sterile environment and less plasma cells in the lymph nodes germinal centers (GC) of mesentery organ than growth data in a non-sterile environment [1, 16, 21].

The gut microbiota acts as a bio-metabolic interconnecting “network” which interconnects with the human body to perform many of the primary functions necessary to maintain its health. The breakdown of many food ingredients that are not digested—fibers, some lipids and proteins, bile acids, cholesterol, endogenous mucus—are some of the most important actions of the intestinal microbiota (7–10% of the daily energy requirement of the host). In this way bacteria provide energy but also produce main metabolites such as the short-chain fatty acids (SCFAs) which are another additional source of energy for the host. Some species synthesize and secrete vitamins such as K, B12, folate (vit.B9), thiamine, biotin as well as amino acids. Indeed, the Bacteroides thetaiotaomicron, is an important member responsible for the breakdown of polysaccharides that end up being indigestible in the large intestine. It has many enzymes such as the polysaccharide glycosyl hydrolases that break down pectin, arabinoce, etc. The archaea such as the Methanobrevibacter smithii, develop cooperative relationships with bacteria by removing the hydrogen (H2) they produce, facilitating the release of Adenosine triphosphate (ATP) [3, 5, 21–23].

4.1.1 The relationship of the enteric microbiota on diseases

According to clinical studies conducted in recent years, changes in the composition of gut microbiota is associated with a number changes in the normal microbiological synthesis of the enteric biomass can present a number of serious pathological conditions such as low immune system health, allergy, irritable bowel syndrome (IBS), autoimmune diseases such as idiopathic inflammatory disease, diabetes (type 2), weight gain (which can lead to obese) and obesity, asthma, and chronic sinusitis, gastroesophageal reflux disease, constipation or diarrhea, dermatological problems, mental health disorders, Alzheimer’s disease and other [1, 16, 24].

In irritable bowel syndrome (IBS), several studies have shown a change and imbalance in the composition of the gut microbiota and suggest that dysbiosis can lead to impaired immune activity, which could lead to a continuous slight intestinal inflammation. The alleged cause of this could be caused by exogenous or endogenous trigger factors; however, the pattern of immune activity in IBS is complex and most likely involves both inborn and acquired immunity mechanisms. On the one hand, there is a reduced diversity of the community with a characteristic decrease of the strains of the genus Firmicutes (such as Bifidobacteria, Lactobacilli and Faecalibacterium prausnitzii) and on the other an increase of the microbes that are attached to the mucus such as the Bacteroidetes, while the remaining patients had a normal composition of the intestinal microbiota. Firmicutes strains are the main SCFAs producers, such as the butanoic acid, which has immunomodulatory properties. Individuals with IBS with diarrhea (IBS-D) appear to have less Lactobacillus spp. instead, those suffering from irritable bowel syndrome with constipation (IBS-C) have more Veillonella spp. in the environment of the intestinal flora the quality (the presence of different species) and the quantity allow a complete functioning of the am bio-network in patients with IBS seems to have a reduced settlement of “good” microorganisms. About innate immunity, that of macrophages plays an important role. Currently, that relationship between an increase or a decrease in the number of macrophages observed in IBS is still the subject of various observations. In fact, the presence of reduced chemokines produced such as CXC Motif Chemokine Ligand 9 (CXCL-9) and monocyte chemoattractant protein-1 (MCP-1/CCL2) has been discovered that engage some immune cells (such as dendritic cells). However, an increase in MCP-1 was noted, thus also questioning the data on the expression of intestinal chemotactic factors. Furthermore, an increase in serum of the cytokines such as the tumor necrosis factor alfa (TNFα), IL-6 and IL-8 tends to be found in patients.
with IBS, and this once again evokes the idea that the alterations caused to the intestinal microbiota following an immune response, evoked by any pathogen, causes systemic perturbations. Toll-like receptor (TLR) pattern recognition receptors play a key role and are extensively explicit in numerous cells, including epithelial cells and macrophages. TLRs intervene in recognizing warning signs such as structures found on microorganisms. These recognition mechanisms are nothing more than the key to the first pass for the secretion of cytokines [16, 24–27].

“Idiopathic” inflammatory bowel disease (IBD) is a chronic immunologically direct disorder disease and is represented by Crohn’s disease and Ulcerative colitis. The IBD is currently thought pathophysiologic derive from a hostile immune reaction to endogenous intestinal symbiotic microbes with or without involvement of the autoimmune process. In non-pathological conditions intestine enclose a vast number of immune cells that are in a state of activation such that they do not have a complete immune response to normal microbiota microbes and food antigens. This is achieved through very powerful regulatory pathways, such as that of suppressor T cells (Tregs) that express the transcription of the forkhead box P3 factor (Foxp3) and suppress inflammation. However, when there is a real infection or other environmental stimuli, in a normal organism, there is a complete activation of the intestinal immune system, but it is quickly suppressed [16, 24, 25]. However, in the case of patients with IBD, this process of extinction of the immune response may not be adequately regulated. The creation and maintenance of the composition and function of intestinal microbes is under the control of the host (e.g., immune, and epithelial responses), the environment (e.g., through diet and the use of antibiotics) and possibly genetic. In turn microbes, through their structural components and their metabolic activity, have notable repercussions on both epithelial and immune function of the host; through epigenetic efforts, these functions can be permanent. From an early age, when the gut microbial community is established, these microbial effects on the host can be important in determining the risk of developing IBD in the distant future. In most studies the rate of idiopathic inflammatory bowel disease increases particularly in the second to fourth decade of life, while some studies report even a second high increase in the third age. In particular, therefore, the components of microbes can promote or protect against diseases [22, 25, 26]. In patients suffering from ulcerative colitis or Crohn’s disease, the germs community have been shown to be different from those without infection, a condition of dysbiosis: the presence of pathogenic microorganisms (e.g., directs the immune response and/or the loss of microorganisms that inhibit inflammation (e.g., Firmicutes such as Faecalibacterium prausnitzii). But many changes and inflammations cause changes in the microbial community. In addition, antibiotics such as metronidazole, ciprofloxacin and some di-
mucosa more frequently in patients with Crohn’s disease [27–29]. However, the Carcinomebryonic antigen cell adhesion molecule 6 (CAECAM6) and the cellular heat shock protein Gp96 there are two important receptors on the enteric epithelial (particularly in the final ileum tract) that are responsible for selective colonization, penetration and retention of E. coli (AIEC). Also noteworthy is the ability of E. coli (AIEC) to survive and multiply in macrophages. It has been found that the E.coli (AIEC) series has the ability to cause chronic inflammation in genetically sensitive hosts [29–32]. Dysbiosis of the intestinal microbiota in situations of infections such as viral ones can lose its eu­biosis. Even in the case of the current SARS-CoV-2 pandemic, the infection has led to an alteration of the normal intestinal flora causing dysbiosis conditions. This means that the infection leads to extensive inflammatory reactions which can worsen the symptoms and therefore the prognosis of the patients. Changes in the fecal microbiota were observed compared to healthy controls. Indeed, in these patients they found that some genera were overrepresented such as Streptococcus, Clostridium, Morganella morganii and others. Instead, Coprococcus, Parabacteroides merdae, Fimbicutes, Bacteroidetes were less present and others. It has also been noted that Bacteroidetes and Firmicutes with their action during viral infection is protective for the host. Therefore, further alterations in the enteric microbiota leads to worsening of the patient’s immune homeostasis [33–35].

Human tumors can be caused not only by genetic factors, food (excess of red meat, fat, etc.), lifestyle (smoking, alcohol, drug abuse etc.) and the environment (radiation, etc.), but also by chronic inflammation and persistent infections. In fact, the infections caused by Helicobacter pylori can cause gastric carcinoma instead a chronic inflammatory bowel disease such as ulcerative colitis which for about 5% can evolve into cancer. In fact, a quantitative and qualitative alteration in the microbes of the normal intestinal bacterial flora can lead to carcinogenesis (both through the diet and through its anti-inflammatory action on the intestinal mucosa). Indeed, the intestinal microbiota in patients with colorectal cancer (CRC) is characterized by an increased variety of Clostridium spp. As well as enrichment of the intestine with Bacteroides and Bifidobacterium spp. acid, such as Lactobacillus spp. and Eubacterium aerofaciens [33, 36]. Animal studies led them to propose a carcinogenicity model. These scientists found that mutations in the intestinal epithelial cells cause the intracellular ligaments to relax and the mucus to shrink, affecting the integrity of the intestinal mucosa. This results in the transfer of bacteria from the lumen to the skin where microbial products bind to tumor-associated macrophage receptors and release Interleukine-1, Interleukine-6 and especially Interleukin-23 which in turn stimulates T-helper lymphocytes that secrete interleukin IL-17 [37–39]. The latter activates the transcription factor STAT3 in epithelial cells, which increases the survival and proliferation of epithelial cells with the consequent addition of additional mutations that lead to dysplasia and possible carcinoma [39–44]. These changes in the epithelium aggravate the already disturbed integrity of the epithelium, worsening the bacterial alteration and contribute to the vicious pathogenic cycle between bacterial translocation-inflammation-dysbiosis that can lead to oncogenesis. It is therefore understood that “good” bacteria can be the cause of carcinogenesis only if there is an alteration of the host’s immune response, indicating the special role of the mucosa in preventing the development of cancerogenesis and its complications [38, 45–48].

There are several studies in which the microbiota has been associated with the development of diabetes (particularly, type 2), which is characterized by a decrease in concentrations of Clostridial bacteria (genera Roseburia and Faecalis). Variations in the number of Bifidobacterium, Lactobacillus, Clostridium, Firmicutes, Bacteroidetes have also been observed in the gut microbiota of young patients with type 1 diabetes [49, 50].

The increase in energy accumulation in obese individuals is related to the transport of hydrogen between taxonomic groups of micro-organisms, since they observed a simultaneous increase of the Prevotellaceae producing hydrogen (H2) and archaea methanogens using H2. In a study that compared to normal weight and obese and gastric bypass groups, the obese group showed a significant increase and a relative abundance of Gammaproteobacteria and proportionally less Clostridia. In addition, the group with gastric bypass patients showed a different intestinal microbiota but rarely in the obese group. Obesity in the previous normal weight subjects showed the same that is an alteration of their intestinal microbiota. Obese people harbor groups of H2-producing bacteria (such as Prevotellaceae and Firmicutes). In obese individuals in the gastrointestinal tract are found over these bacteria, also a high population of H2-oxidative methanogens (they are 10% of all anaerobes in the fatty intestine) [51, 52]. The various plant fibers and polysaccharides are metabolized by intestinal bacteria to produce various metabolites (mainly SCFAs), including acetate, propionate, butyrate, and lactate. In addition, the high presence of H2-oxidative methanogens facilitates this process, which results in the production of more SCFAs with the production of more acetate and hydrogen. These metabolites are absorbed through the human intestinal epithelium, while H2 is an energy exchange factor within microbial communities [52, 53].

5. The Gut-brain, Gut-pulmonary and Gut-skin axes

The interaction between the gut microbiota and the brain constitutes the so-called crosstalk Gut-brain axis. This interaction is therefore bidirectional and occurs through endocrine, neural, immune, and humoral signaling
connections pathways from the gut microbiota to the brain and from the brain to the gut microbiota. This communication network includes the central nervous system (CNS) and the peripheral nervous system (PNS), and the hypothalamic–pituitary–adrenal (HPA or HTPA) axis. Through the pneumogastric nerve (component of the parasympathetic nervous system) the interaction takes place in a bidirectional world. This neuro-intestinal system includes the nerve part with its motor neurons, primary intrinsic afferent neurons and glial cells contained in the Auerbach's plexus and tela submucosa that extend along the intestine. The CNS through the autonomic nervous system (ANS) and the HPA axis affects the gastrointestinal tract to its functions such as through motility its secretory capacity and its permeability (such as motility and secretion and more). In clinical practice, the evidence for GBA-microbial interactions stems from a dysbiosis condition between nervous system disorders (such as depression, autism and others) and functional gastrointestinal disorders [53–55]. The central nervous system through the autonomic nervous system but also through the HPA axis affects its function gastrointestinal tract through the motility of the digestive tract, its secretory capacity and its permeability. These effects in turn affect its microbiota. On the other hand, germ products can reach the brain and affect its function. In addition, it has been observed in experimental animals that the intestinal microbiota affects the formation of synapses in the brain and the production of neurotransmitters. So, we notice that there is a substantial interaction between the microbiota and the nervous system. It is believed that changes in the bidirectional interaction between the brain and the gut may be the cause but also for its evolution over time of irritable bowel syndrome. The pathogenesis of functional gastrointestinal disorders therefore, as we have previously mentioned, beyond the pathogenesis of functional gastrointestinal disorders, neurological disorders are involved (such as Parkinson's disease, disorders on the affective sphere, mood and emotions, chronic pain and others). Intestinal microbes with their metabolites influence the permeability of the intestinal barrier, the immune system, motility and activity of the intestinal nervous system. Preclinical data also show that they can regulate brain behaviors and functions, including responses to stress, emotions, pain control, eating behavior and generally brain biomolecular functions [56, 57]. The gut microbiota therefore it can interfere with the neural pathways and behavior in the face of a stressful condition. Social stresses increase the risk of inflammatory diseases, promoting the expression of pro-inflammatory genes and the differentiation of monocytes. Therefore, the alterations that cause inflammatory processes induce the alteration of the intestinal microbiota and thus can further favor the ability of pathogens to colonize the intestine. It has also been shown that a condition of continuous stress can influence the secretion of IgA, the inflammatory response subsequently leading to dysbiosis and thus disturbing intestinal homeostasis. Several on animals studies are conducted for research on the interaction on stress-inducing stimuli between the intestinal microbiota and the HPA axis. These provide us with information on the importance of the gut microbiota in the development of the HPA axis. It has been noticed, in the germ-free (GF) mice, in front of a mild contentious stress stimulus there is an increase in the secretion of ACTH and corticosterone compared to mice lacking for particular pathogens. It was noted that this condition of increased hormone release had been partially normalized in these animals by the introduction of the fecal microbiota from lacking for particular pathogens animals. It was also noted that everything normalized with the introduction of the Bifidobacterium infantis strain, during their early childhood. On the other hand, in another study the administration of B. infantis improved the response to the stressful stimulus in gf mice. Therefore, for a response to the same stimuli in adult life, the development of an effective intestinal microbiota from birth is important. This is what will ensure the correct development of the HPA axis in the future [52]. In fact, in addition to the increase in hormones during stress in these animals, it was also noted the reduction in the levels of the Brain-derived neurotrophic factor (BDNF), which is a neurotrophin involved in both growth and neuronal survival. Finally, in many studies on gf animals, alterations in the expression in the hippocampus of the serotonin 1A receptor (5-HT1A) and N-methyl-D-aspartate (NMDA) receptor have been found. By influencing the release of corticotropin-releasing hormone (CRH) from the hypothalamus and therefore a modified response of action of HPA. Stress also leads to high levels of the secretion of Interleukin-6 and of the chemotactic monocyte protein-1 (MCP-1), which were associated with changes caused by the stress-induced stimulus in certain bacteria such as Co- prococcus and other [53, 57]. In another study, observed the changes with B. infantis treatment, but not a reduction in corticosterone, while later with the use of a similar model found that adding Lactobacillus to the diet reduced corticosterone levels [58–60]. It is also unclear whether changes in the gut microbiota in patients with such disorders as irritable bowel syndrome are due to primary changes involving only germs and/or changes associated with bowel-brain communication. In addition, although there are rare cases of patients who develop psychotic symptoms after using a wide range of antibiotics, there are not enough clinical data showing that a sudden change in the gut microbes has a clinically obvious effect on the individual’s symptoms. The first study in experimental animals has already shown that the absence of a normal gut microbiota can significantly affect the response to stress, and this can be reversed by re-colonization of the gut. The (GF) mice are thinner than the free specific pathogen mice (SPF), although they consume more calories. The metabolic changes that occur in them can affect the brain development and alter the activity of neural circuits associated with eating behavior and
metabolism. Changes in barrier permeability in germ-free mice can lead to significantly different access of microbial metabolites to the brain. There are therefore two-way interactions of the gut microbes with the central nervous system. The CNS regulates the intestinal tract and the intestinal nervous system through sympathetic and parasympathetic pathways of the ANS as well as with the HPA axis (Fig. 6). The CNS effects now can affect the intestinal microbiota indirectly by changing their environment and directly through a variety of signaling molecules. The ANS regulates functions such as motility, acid secretion, production of bicarbonate and mucus, retention of epithelial fluids, intestinal permeability, and mucosal immune response. Most of these functions are under the influence of sympathetic and parasympathetic in the intestinal nerve circuits systemic [53, 61, 62].

Another important systemic axis is the communication between the lung microbiota and the intestinal microbiota (Gut-lung axis). This is another type of crosstalk with the exchange of immunological information between the two systems. This will result in the possibility of influencing the functional behavior of the lung microbiota under certain conditions. Thus, the gut microbiota can regulate and modify the immunological activity of the lung via bacterial lipopolysaccharides (LPS) and various bacterial metabolites (such as SCFAs and other). Subsequently, after stimulation with the production of dendritic cells that they cause the activation of various T lymphocytes (particularly T-reg, Th17, Th1) migrating to the lower respiratory tract through the circulatory flow. Instead, bacterial metabolites cause tumor necrosis factor (TNF-α) to decrease via activation of the activated B cell light chain kappa-enhancer nuclear factor (NFκB). This will result in a downregulation of pattern recognition receptors (PRRs) which will decrease the production of cytokines (Interleukin-1, Interleukin-12, Interleukin-18, TNF-α, IFN-γ and GM-CSF). This important interconnection maintains homeostasis of the immune system in the lungs and vice versa, and microbial dysbiosis is avoided. In addition, the environment and the appropriate lifestyle, nutrition with the integration of probiotics, can be a protective factor against dysbiosis that can lead to the regulation of the Gut-lung axis. The onset and progression of respiratory diseases such as these from the SARS-CoV-2 virus can significantly alter the lung microbiota by increasing potential pathogenic species thus affecting the course and severity of disease (Fig. 7) [63–69].

Finally, we reported that the interconnection between enteric microbes and host immune functions promotes the proper functioning of the intestinal immune system, but a condition of severe intestinal dysbiosis that leads to not only intestinal inflammation with the integumentary system involvement. This is the crosstalk between the Gut-skin axis. Consequently, we can have various pathological manifestations of the skin such as eczema, atopic dermatitis, acne, and others. It is reported that in some cases of dysbiosis conditions of the microbiota, there is an increase in the final products of the metabolism of aromatic amino acids (i.e., free phenol and p-cresol from the Clostridium difficile. This can lead to changes in the immune response (modification of the production between Teff and Treg lymphocytes) in the intestine which may involve the skin. Finally, most likely during SARS-CoV-2 skin manifestations may be due
6. Probiotics, prebiotics and gut microbiota

Probiotics (Greek = πρό + βιοτικά (proviotika) = for the life), were live microorganisms that are intended to have health benefits when consumed or applied to the body (Lilly and Stillwell first introduced the term in 1965). The original idea that some bacteria can benefit human health is attributed to Ilya Metchnikoff, who worked at the Pasteur Institute in the early 20th century [70]. Scientific research supporting the purported benefits of probiotics was limited, mainly due to the complexity of the gut ecosystem. Later studies showed that the act of directly modifying the composition of the intestinal microbiota through the use of probiotics, such as Bifidobacterium spp. (B. breve and other), Lactobacillus spp. (L. Acidophilus and other) and Saccharomyces spp. (Saccharomyces cerevisiae and other) has been shown to have a positive effect (not always) on many gastrointestinal and non-gastrointestinal diseases (Fig. 8) [3, 69, 71, 72].

The choice of strains depends on their safety, efficacy for health and their ability to benefit humans. Indeed, e.g., probiotics that act on the large intestine must be resistant to sialic enzymes, acid secretion of stomach (changes of pH), bile secretions, small intestine enzymes such as those of the pancreatic secretions (lipase and amylase) and the environment of other foods and drinks encountered during their passage along the gastrointestinal tract. In addition, they must be able to compete with microbial flora. The manipulation of probiotics as oral therapeutics has also been shown to be useful in reducing the small intestinal bacterial overgrowth linked with anxious-depressive disorders. In one study, probiotics from 2 strains L. helveticus and B. longum in healthy volunteers were involved in the therapy for anxiety and depression. After two weeks the improvement of the symptoms was noticed. The same happens with the use of Lactobacillus rhamnosus which it was noticed that it acts on GABA (gamma-aminobutyric acid) which in cases of anxiety and depression its production is modified (Fig. 9).

Therefore, some studies report that as adjuvant therapy and to prevent psychiatric disorders (such as addiction to substances of abuse, bipolar disorder and others) it would be appropriate to use some selected bacterial strains such as Bifidobacterium infantis (increase in the levels of the 5-HT precursor tryptophan) has been studied in its efficacy in relieving IBS-associated depression and anxiety. Indeed, researchers now refer to these living organisms as “psychobiotics” which, if administered in well-established quantities they can have beneficial effects on the
Fig. 8. The main probiotics in relation to their main use (1 mainly for animal use, 2 mainly for drugs development. Adapted from Santacroce et al. A successful history: probiotics and their potential as antimicrobials, Expert Rev Anti Infect Ther 2019, 17, 8, 635–645).

Fig. 9. The effect after oral administration of Lactobacillus rhamnosus in the brain in relation to the variation of the expression of GABA mRNA.

health of patients suffering from psychiatric diseases; and it has already been proposed as an additional treatment in patients suffering from depression or in mild depressive states in general. In addition to these that we have mentioned, other psychobiotics can be E. Coli, Bacillus and Saccharomyces (facilitate the increase of norepinephrine), Candida, Streptococcus and Enterococcus (increase the production of 5-HT) Bacillus and Serratia (stimulate the secretion of dopamine) [69, 72–75]. The Anandamide and 2-Arachidonoylglycerol (2-AG) of the Endocannabinoid System (ECS) which have neurotransmitter and neuromodulatory capabilities, a strain of Lactobacillus acidophilus can modify the function of their receptors in the spinal cord [76–78]. Thus, probiotics in general can now be effective in therapy and prevention as an adjunct in various infectious and non-infectious diseases. Probiotics in general can now be effective in various infectious and non-infectious diseases [79–83]. Prebiotics are foods that the body is unable to digest but which lead to a selective and specific function leading to the development and/or activity of one or a limited number of bacterial species already present in the large intestine. Therefore, they are mainly made up of indigestible carbohydrates and the most used may be inulin, fructo-galacto- and xyloligosaccharides. These prebiotics once found in the large intestine undergo fermentation by the SCAFs of the local bacterial flora. Furthermore, it has been noted through several studies that they also have benefits in comparison with similar probiotic strains (such as Bifidobacterium spp. and Lactobacillus spp.). In chronic inflammatory bowel disease. The benefit of this treatment has been shown to be highly dose-dependent. In fact, high levels of prebiotics can often intensify problems like bloating and flatulence. Finally, the prebiotics could be useful in combination with probiotics [69, 84].

7. Conclusions

Actually, much “light” has been made about the microbial biomass that colonize our body and particularly for the intestinal microbiota. Its pathophysiological involvement in various diseases such as non-gastrointestinal has been extensively studied and continues to be studied. The composition of the gut microbiota differs in health and disease conditions. It contributes to the pathophysiological processes associated with the three axes: Gut/Brain, Gut/Pulmonary and Gut/Skin. Therefore, in conditions of eubiosis of the intestinal microbiota it acts in favor of human health thus preventing the development of “bad” germs (pathogens). Moreover, by modulating the local immune defenses, it puts the immune system in an equilibrium for an effective and adequate response against the pathogenesis of various diseases. Finally, due to the intake of antibiotics, diet, infections such as the viral pandemic SARS-CoV-2 potential pathogens favoring the growth and/or transmission in the various other systems of pathogenic germs that will lead to various local or systemic diseases. Furthermore, this microbial imbalance can compromise local intestinal health such as allergies, inflammatory diseases, pre-cancer conditions and more. Thus, probiotics and prebiotics restore the balance and the functional homeostasis of the intestinal microbiota which with its cross-talking axes not only creating effective immune defenses against various diseases but can also be an additional help for mental ones (Psychobiotics).

8. Author contributions

All the authors equally contributed to conceiving the study, literature investigation and project management. Dr. Charitos wrote the manuscript and all the authors re-
viewed it and agreed to its publication.

9. Ethics approval and consent to participate

Not applicable.

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12. Conflict of interest

The authors declare that there is no conflict of interest.

13. References

[1] Thursby E, Juge N. Introduction to the human gut microbiota. Biochemical Journal. 2017; 474: 1823–1836.
[2] Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. PLoS Biology. 2016; 14: e1002533.
[3] Bottalico L, Castellaneta F, Charitos IA. From hydrotherapy to the discovery of the gut microbiota: the historical gastrointestinal health concept. Pharmcophore. 2020; 11: 82–90.
[4] Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargent M, et al. Diversity of the human intestinal microbial microbiota. Science. 2005; 308: 1635–1638.
[5] Hill MJ. Intestinal microbiota and endogenous vitamin synthesis. European Journal of Cancer Prevention. 1997; 6: S43–S45.
[6] Thomas LV, Ockhuizen T, New insights into the impact of the intestinal microbiota on health and disease: a symposium report. British Journal of Nutrition. 2012; 107: S1–S13.
[7] Dominguez-Bello MG, Godoy-Vitorino F, Knight R, Blaser MJ. Role of the microbiota in human development. Gut. 2019; 68: 1108–1114.
[8] Palmer C, Bik EM, DiGiulio DB, Relman DA, Brown PO. Development of the human infant intestinal microbiota. PLoS Biology. 2007; 5: e177.
[9] Schloissnig S, Arumugam M, Sunagawa S, Mitreva M, Tap J, Zhu A, et al. Genomic variation landscape of the human gut microbiota. Nature. 2013; 493: 45–50.
[10] Cabrera-Rubio R, Collado MC, Laitinen K, Salminen S, Isolauri E, Mira A. The human milk microbiota changes over lactation and is shaped by maternal weight and mode of delivery. The American Journal of Clinical Nutrition. 2012; 96: 544–551.
[11] Moya-Pérez A, Luczynski P, Renes IB, Wang S, Borre Y, Anthony Ryan C, et al. Intervention strategies for cesarean section-induced alterations in the microbiota-gut-brain axis. Nutrition Reviews. 2017; 75: 225–240.
[12] Mangiola F, Ianiro G, Franceschi F, Fagiuoli S, Gasbarrini G, Gasbarrini A. Gut microbiota in autism and mood disorders. World Journal of Gastroenterology. 2016; 22: 361–368.
[13] O’Toole PW, Jeffery IB. Gut microbiota and aging. Science. 2015; 350: 1214–1215.
[14] Penders J, Thijs C, Vink C, Stelma FF, Snijders B, Kummeling I, et al. Factors influencing the composition of the intestinal microbiota in early infancy. Pediatrics. 2006; 118: 511–521.
[15] Claesson MJ, Jeffery IB, Conde S, Power SE, O’Connor EM, Casack S, et al. Gut microbiota composition correlates with diet and health in the elderly. Nature. 2012; 488: 178–184.
[16] Purchiaroni F, Tortora A, Gabrielli M, Bertucci F, Gigante G, Ianiro G, et al. The role of intestinal microbiota and the immune system. European Review for Medical and Pharmacological Sciences. 2013; 17: 323–333.
[17] Tannock GW, Savage DC. Influences of dietary and environmental stress on microbial populations in the murine gastrointestinal tract. Infection and Immunity. 1974; 9: 591–598.
[18] Liu Z, Li N, Neu J. Tight junctions, leaky intestines, and pediatric diseases. Acta Paediatrica. 2005; 94: 386–393.
[19] Bloomfield SF, Rook GA, Scott EA, Shanahan F, Stanwell-Smith R, Turner R. Time to abandon the hygiene hypothesis: new perspectives on allergic disease, the human microbiota, infectious disease prevention and the role of targeted hygiene. Perspect Public Health. 2016; 136: 213–224.
[20] Santacroce L, Bufo P, Latorre V, Losacco T. Role of mast cells in the physiopathology of gastric lesions caused by Helicobacter pylori. Chirurgia Italiana. 2000; 52: 527–531. (In Italian)
[21] Srikanth CV, McCormick BA. Interactions of the intestinal epithelium with the pathogen and the indigenous microbiota: a three-way crosstalk. Interdisciplinary Perspectives on Infectious Diseases. 2008; 2008: 626827.
[22] Tremaroli V, Bäckhed F. Functional interactions between the gut microbiota and host metabolism. Nature. 2012; 489: 242–249.
[23] Macfarlane S, Macfarlane GT. Regulation of short-chain fatty acid production. The Proceedings of the Nutrition Society. 2003; 62: 67–72.
[24] Lankelma JM, Nieuworp M, de Vos WM, Wiersinga WJ. The gut microbiota in internal medicine: implications for health and disease. The Netherlands Journal of Medicine. 2015; 73: 61–68.
[25] Stasi C, Rosselli M, Bellini M, Laffi G, Milani S. Altered neuroendocrine-immune pathways in the irritable bowel syndrome: the top-down and the bottom-up model. Journal of Gastroenterology. 2012; 47: 1177–1185.
[26] Natividad JMM, Verdu EF. Modulation of intestinal barrier by intestinal microbiota: pathological and therapeutic implications. Pharmacological Research. 2013; 69: 42–51.
[27] Santacroce L, Mavaddati S, Hamedi J, Zeinali B, Ballini A, Bellini M, et al. Role of the microbiota in human development. Gut. 2019; 68: 150–165.
[28] Khan I, Ullah N, Zha L, Bai Y, Khan A, Zhao T, Che T, Zhang C. Alteration of gut microbiota in inflammatory bowel disease (IBD): cause or consequence? IBD treatment targeting the gut microbiota. Pathogens. 2019; 8: 126.
[29] Schippa S, Conte MP. Dysbiotic events in gut microbiota: impact on human health. Nutrients. 2014; 6: 5786–5805.
[30] Ananthakrishnan AN. Epidemiology and risk factors for IBD. Nature reviews. Gastroenterology & Hepatology. 2015; 12: 205–217.
[31] Bennet SMP, Ohman L, Simren M. Gut microbiota as potential orchestrators of irritable bowel syndrome. Gut and Liver. 2015; 9: 318–331.
[32] Di Serio F, Lovero R, D’Argostino D, Nisi L, Miraglia G, Contino R, et al. Evaluation of procalcitonin, vitamin D and C-reactive protein levels in septic patients with positive emoclures. Our preliminary experience. Acta Medica Mediterranea. 2016; 32: 1911–1914.
[33] Carding S, Verbeke K, Vipond DT, Corfe BM, Owen LJ. Dysbiosis of the gut microbiota in disease. Micobacterial Ecology in Health and Disease. 2015; 26: 26191.

[34] Dhar D, Mohanty A. Gut microbiota and COVID-19: possible link and implications. Virus Research. 2020; 285: 198018.

[35] Zuo T, Zhang F, Lui GYC, Yeo YK, Li AYL, Zhan H, et al. Alterations in Gut microbiota of patients with COVID-19 during time of hospitalization. Gastroenterology. 2020; 159: 944–955.e8.

[36] Santacroce L, Cagiano R, Del Prete R, Bottalico L, Sabatini R, Carrai L, et al. Helicobacter pylori infection and gastric MALTomas: an up-to-date and therapy highlight. Clinical Therapeutics. 2008; 159: 457–462.

[37] Grivennikov SI, Wang K, Muñica D, Stewart CA, Schnabl B, Jauch D, et al. Adenoma-linked barrier defects and microbial products drive IL-23/IL-17-mediated tumour growth. Nature. 2012; 491: 254–258.

[38] Farhana L, Banerjee NN, Verma M, Majumdar APN. Role of microbiome in carcinogenesis process and epigenetic regulation of colorectal cancer. Methods in Molecular Biology. 2018; 1856: 35–55.

[39] Polimen L, Barone M, Mosca A, Viggiani MT, Joukar F, Mansour-Ghanai F, et al. Soy metabolism by gut microbiota from patients with precancerous intestinal lesions. Microorganisms. 2020; 8: E469.

[40] Nistal E, Fernández-Fernández N, Vivas S, Olcoz JL. Factors determining colorectal cancer: the role of the intestinal microbiota. Frontiers in Oncology. 2015; 5: 220.

[41] Santacroce L, Buonfantino M, Santacroce S. Surgical treatment and prognostic factors in colon cancer. Journal of Chemotherapy. 1997; 9: 144–145.

[42] Santacroce L, Bufo P, Gagliardi S, Mastropasqua MG, Losacco T. Argyrophilic nucleolar organizer regions (AgNORs) as malignancy biomarkers in colorectal neoplasms. La Clinica Terapeutica. 2001; 152: 91–93. (In Italian)

[43] Bufo P, Losacco T, Greco L, Gagliardi S, Logrieco S, Santacroce L. Expression of epithelial oncoproteins in large intestine neoplasms. La Clinica Terapeutica. 2002; 153: 243–245. (In Italian)

[44] Santacroce L, Losacco T. Abdominal sepsis in surgical patients. Pathophysiology and prevention. Recenti Progressi in Medicina. 2006; 97: 411–416. (In Italian)

[45] Losacco T, Santacroce L. Treatment of bowel obstruction in our own experience. La Clinica Terapeutica. 2005; 156: 89–92. (In Italian)

[46] Moore WE, Moore LH. Intestinal microbiotas of populations that have a high risk of colon cancer. Applied and Environmental Microbiology. 1995; 61: 3202–3207.

[47] Man A, Mare A, Toma F, Curticăpean A, Santacroce L. Health threats from contamination of spices commercialized in Romania: risks of fungal and bacterial infections. Endocrine, Metabolic & Immune Disorders Drug Targets. 2016; 16: 197–204.

[48] Polimen L, Barone M, Mosca A, Viggiani MT, Di Leo A, Debellis L, et al. Gut microbiota imbalance is related to sporadic colorectal neoplasms. A pilot study. Applied Sciences. 2019; 9: 5491.

[49] Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg L, et al. Gut metagenome in European women with normal, impaired and diabetic glucose control. Nature. 2013; 498: 99–103.

[50] Murn M, Leiva I, Gomez-Zumaquero JM, Tinafones FJ, Cardona F, Surigué F, et al. Gut microbiota in children with type 1 diabetes differs from that in healthy children: a case-control study. BMC Medicine. 2013; 11: 46.

[51] Zhang H, DiBaise JK, Zuccolo A, Kudrna D, Braiddy M, Yu Y, et al. Human gut microbiota in obesity and after gastric bypass. Proceedings of the National Academy of Sciences of the United States of America. 2009; 106: 2365–2370.

[52] Schink B. Energetics of syntrophic cooperation in methanogenic degradation. Microbiology and Molecular Biology Reviews. 1997; 61: 262–280.

[53] Carabotti M, Scirocco A, Maselli MA, Severi C. The gut-brain axis: interactions between enteric microbiota, central and enteric nervous systems. Annals of Gastroenterology. 2015; 28: 203–209.

[54] Moloney RD, Johnson AC, O’Mahony SM, Dinan TG, Greenwood-Van Meerveld B, Cryan JF. Stress and the microbiota-gut-brain axis in visceral pain: relevance to irritable bowel syndrome. CNS Neuroscience & Therapeutics. 2016; 22: 102–117.

[55] Sudo N. Stress and gut microbiota: does postnatal microbial colonization program the hypothalamic-pituitary-adrenal system for stress response? International Congress Series. 2006; 1287: 350–354.

[56] Neufeld KM, Kang N, Bienenstock J, Foster JA. Reduced anxiety-like behavior and central neurochemical change in germ-free mice. Neurogastroenterology and Motility. 2011; 23: 255–64. e119.

[57] Dinan TG, Cryan JF. Gut-brain-microbiota axis and mental health. Psychosomatic Medicine. 2017; 79: 920–926.

[58] Desbonnet L, Clarke G, Toplán A, O’Sullivan O, Crispe F, Moloney RD, et al. Gut microbiota depletion from early adolecence in mice: implications for brain and behaviour. Brain, Behavior, and Immunity. 2015; 48: 165–173.

[59] Gareau MG, Silva MA, Perdue MH. Pathophysiological mechanisms of stress-induced intestinal damage. Current Molecular Medicine. 2008; 8: 274–281.

[60] Zhang H, Wang Y. Gut microbiota-brain axis. Chinese Medical Journal. 2016; 129: 2373–2380.

[61] Rhee SH, Pothoulakis C, Mayer EA. Principles and clinical implications of the brain-gut-enteric microbiota axis. Nature Reviews Gastroenterology & Hepatology. 2009; 6: 306–314.

[62] Mayer EA, Tillisch K, Gupta A. Gut-brain axis and the microbiota. The Journal of Clinical Investigation. 2015; 125: 926–938.

[63] Samuelsdon DR, Welsh DA, Shellito JE. Regulation of lung immunity and host defense by the intestinal microbiota. Frontiers in Microbiology. 2015; 6: 1085.

[64] Bassis CM, Erb-Downward JR, Dickson RP, Freeman CM, Schmidt TM, Young VB, et al. Analysis of the upper respiratory tract microbiotas as the source of the lung and gastric microbioitas in healthy individuals. MBio. 2015; 6: e00037.

[65] Enaud R, Preevl R, Ciono E, Beaufils F, Weers G, Guery B, et al. The gut-lung axis in health and respiratory diseases: a place for inter-organ and inter-kingdom crosstalks. Frontiers in Cellular and Infection Microbiology. 2020; 10: 9.

[66] Santacroce L. Letter in response to the article “Enhancing immunity in viral infections, with special emphasis on COVID-19: a review” (Jayawardena et al.), Diabetes & Metabolic Syndrome: Clinical Research & Reviews. 2020; 14: 927.

[67] Santacroce L, Charitos IA, Ballini A, Inchingolo F, Luperto P, De Nitto E, et al. The human respiratory system and its microbiome at a glance. Biology. 2020; 9: 318.

[68] Santacroce L, Charitos IA, Carretta DM, De Nitto E, Lovero R. The human coronaviruses (HCoVs) and the molecular mechanisms of SARS-CoV-2 infection. Journal of Molecular Medicine. 2021; 99: 93–106.

[69] Santacroce L, Inchingolo F, Topi S, Del Prete R, Di Cosola M, Charitos IA, et al. Potential beneficial role of probiotics on the outcome of COVID-19 patients: an evolving perspective. Diabetes & Metabolic Syndrome: Clinical Research & Reviews. 2021; 15: 295–301.

[70] Santacroce L, Charitos IA, Bottalico L. A successful history: probiotics and their potential as antimicrobials. Expert Review of Anti-Infective Therapy. 2019; 17: 635–645.

[71] Leone D, Valenzano A, Grande G, Santacroce L. Drug/food interactions: an actual therapeutic outcome. La Clinica Terapeutica. 2004; 155: 139–147. (In Italian)
Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, et al. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. Proceedings of the National Academy of Sciences of the United States of America. 2011; 108: 16050–16055.

Desbonnet L, Garrett L, Clarke G, Kiely B, Cryan JF, Dinan TG. Effects of the probiotic Bifidobacterium infantis in the maternal separation model of depression. Neuroscience. 2010; 170: 1179–1188.

Dinan TG, Stanton C, Cryan JF. Psychobiotics: a novel class of psychotrophic. Biological Psychiatry. 2013; 74: 720–726.

McKernan DP, Fitzgerald P, Dinan TG, Cryan JF. The probiotic Bifidobacterium infantis 35624 displays visceral antinociceptive effects in the rat. Neurogastroenterology and Motility. 2010; 22: 1029–35. e268.

Muccioli GG, Naslain D, Bäckhed F, Reigstad CS, Lambert DM, Delzenne NM, et al. The endocannabinoid system links gut microbiota to adipogenesis. Molecular Systems Biology. 2010; 6: 392.

Charitos IA, Gagliano-Candela R, Santacroce L, Bottalico L. The Cannabis spread throughout the continents and its therapeutic use in history. Endocrine, Metabolic & Immune Disorders - Drug Targets. 2020.

Vermesan D, Vermesan H, Dragulescu SI, Bera I, Di Giovanni A, Sabatini R, et al. Secondary pathologic fractures in osteosarcoma: prognosis and evolution. European Review for Medical and Pharmacological Sciences. 2009; 13: 71–76.

Prejbeanu R, Vermesan H, Dragulescu SI, Vermesan D, Motoc A, Sabatini R, et al. Thromboembolic risk after knee endoprosthesis. European Review for Medical and Pharmacological Sciences. 2007; 11: 297–300.

Giudice G, Cutrignelli DA, Sportelli P, Limongelli L, Tempesta A, Gioia GD, et al. Rhinocerebral mucormycosis with orosinus involvement: diagnostic and surgical treatment guidelines. Endocrine, Metabolic & Immune Disorders Drug Targets. 2016; 16: 264–269.

Mangini F, Santacroce L, Bottalico L. Periodontitis and systemic diseases. La Clinica Terapeutica. 2006; 157: 541–548. (In Italian)

Ballini A, Santacroce L, Cantore S, Bottalico L, Dipalma G, Vito DD, et al. Probiotics improve urogenital health in women. Open Access Macedonian Journal of Medical Sciences. 2018; 6: 1845–1850.

Charitos IA, Topi S, Castellaneta F, D’Agostino D. Current issues and perspectives in patients with possible sepsis at emergency departments. Antibiotics. 2019; 8: 56.

Davani-Davari D, Negahdaripour M, Karimzadeh I, Seifan M, Mohkam M, Masoumi SJ, et al. Prebiotics: definition, types, sources, mechanisms, and clinical applications. Foods. 2019; 8: 92.

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