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Safety and immunogenicity of a novel hexavalent group B streptococcus conjugate vaccine in healthy, non-pregnant adults: a phase 1/2, randomised, placebo-controlled, observer-blinded, dose-escalation trial

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Summary

Background Group B streptococcus (GBS) is a major cause of invasive disease in young infants. Infants born to women with sufficient pre-existing anti-GBS capsular IgG antibodies are at reduced risk of GBS disease, making maternal immunisation a potential strategy for prevention. We aimed to assess the safety and immunogenicity of a novel hexavalent (serotypes Ia, Ib, II, III, IV, and V) GBS conjugate vaccine (GBS6).

Methods This phase 1/2, placebo-controlled, observer-blinded, dose-escalation trial, was done at four clinical research centres in the USA (Kentucky, Georgia, and two sites in Utah). Healthy, non-pregnant adults aged 18–49 years were randomly assigned using an interactive, web-based response technology system. Within each dose group (low, medium, or high), participants in sentinel cohorts were randomly assigned 2:2:1 and expanded cohort participants were randomly assigned 4:4:1 to receive GBS6 with aluminium phosphate (AlPO₄), GBS6 without AlPO₄, or placebo (saline control). One 0.5 mL dose of either saline placebo or 5 μg capsular polysaccharide per serotype in the low-dose group, 10 μg capsular polysaccharide per serotype in the medium-dose group, or 20 μg capsular polysaccharide per serotype in the high-dose group was administered by intramuscular injection into the deltoid muscle on day 1. The primary outcome was safety up to 6 months after vaccination, including the proportion of sentinel cohort participants with clinical laboratory abnormalities at 1 week, the proportion of all participants reporting solicited local reactions, systemic events, or use of antipyretic or pain medication within 14 days, adverse events up to 1 month, and medically attended or serious adverse events up to 6 months. The secondary outcome was GBS immunogenicity (serotype-specific IgG geometric mean concentrations at 1 month). This study is registered with ClinicalTrials.gov, NCT03170609.

Findings Between June 5, 2017, and June 25, 2018, 365 participants were randomly assigned and 364 (52 in each dose group) were vaccinated and included in the safety analysis. Unsolicited adverse events were reported by 15 (29%) participants in the 5 μg with AlPO₄, group, 13 (25%) in the 5 μg without AlPO₄, group, 22 (42%) in the 10 μg with AlPO₄, group, 12 (23%) in the 10 μg without AlPO₄, group, 25 (48%) in the 20 μg with AlPO₄, group, 21 (40%) in the 20 μg without AlPO₄, group, and 20 (38%) in the placebo group. The most common unsolicited adverse events were in the system organ class of infections and infestations in any dose or formulation of GBS6 (ranging from six [12%] in the 10 μg without AlPO₄, group to 15 [29%] in the 20 μg with AlPO₄, group and placebo group). Three participants reported at least one serious adverse event during the study, one each in the 5 μg GBS6 with AlPO₄, group (diabetic ketoacidosis, two events; resolved), 10 μg GBS6 with AlPO₄, group (died by suicide), and 20 μg GBS6 with AlPO₄, group (metrorrhagia; resolved). None of these serious adverse events were considered related to the vaccine. 11 of the 363 participants were excluded from the evaluable immunogenicity population, including one participant who did not receive the vaccine, and ten who at 1 month after vaccination were withdrawn for various reasons. GBS serotype-specific IgG geometric mean concentrations increased by 1 week after vaccination for all GBS6 groups, peaked at 2 weeks, stabilised by 1 month, and declined gradually but remained higher than placebo at 6 months.

Interpretation GBS6 was well tolerated in healthy adults and elicited robust immune responses for all dose levels and formulations that persisted 6 months after vaccination. This study supports further evaluation of GBS6 in pregnant women.

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Introduction

Streptococcus agalactiae, also known as group B streptococcus (GBS), is a leading cause of invasive bacterial infection in young infants and causes substantial infant morbidity and mortality worldwide, including in the USA.14 GBS can cause serious disease, such as sepsis,

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Although largely effective in the prevention of early birth, preterm delivery, and puerperal sepsis, infections to chorioamnionitis, which can result in stillbirth with ascending infections ranging from urinary tract infections in the USA to 2·59 cases per 1000 livebirths in South Africa; antibiotic prophylaxis for those who were colonised.13,14

In pregnant women, GBS can be associated with invasive GBS disease in infants and the limitations of prevention and treatment.11,12 As a result, in the late 1990s, US guidelines were broadened to include recommendations for screening of pregnant women for GBS carriage and the use of intrapartum antibiotic prophylaxis for those who were colonised.13,14

Although largely effective in the prevention of early onset disease, intrapartum antibiotic prophylaxis is not optimal because some cases occur even when adequate screening is available (eg, in the setting of preterm births). Additionally, screening failures or incomplete intrapartum antibiotic prophylaxis coverage can occur even in high-income countries that have resources for broad implementation. Intrapartum antibiotic prophylaxis has no effect on late-onset disease (ie, disease onset at 7–90 days of life) and prevents neither GBS-associated stillbirth nor GBS infection of the mother in the peripartum period.11,12 Finally, intrapartum antibiotic prophylaxis might be too late to affect disease in the setting of preterm deliveries, might not be available to women with little access to routine antenatal care, might contribute to antimicrobial resistance, and might affect the neonatal microbiome.13,14 Maternal immunisation could address these shortcomings of intrapartum antibiotic prophylaxis and reduce the global burden of invasive GBS infection in infants and pregnant women. Monovalent, bivalent, and trivalent GBS vaccines against serotypes Ia, Ib, II, III, and V have been previously studied in non-pregnant and pregnant women. These vaccines were shown to be safe and effective in the prevention of GBS disease.
elicited postvaccination increases in GBS antibodies in infants and in vaccinees.\textsuperscript{17} There are no licensed vaccines against GBS disease, and no vaccine has targeted immunity against serotype IV, which is becoming an important contributor to invasive GBS disease.\textsuperscript{18}

This study aimed to evaluate the safety and immunogenicity of a GBS-containing vaccine that includes the six most prevalent capsular polysaccharide serotypes (Ia, Ib, II, III, IV, and V; GBS6), accounting for approximately 98% of disease-causing isolates worldwide.\textsuperscript{19}

Methods
Study design and participants
This phase 1/2, randomised, placebo-controlled, observer-blinded, dose-escalation trial was done at four clinical research centres in the USA (in Kentucky, Georgia, and two sites in Utah). This study used a sentinel cohort design with progression and dose escalation taking place after review of the sentinel cohort at each dose. The sentinel cohorts functioned as a first stage in the study and an expanded cohort served as a second stage to provide an increased number of participants for immunogenicity and further safety assessment.

Healthy, non-pregnant adults aged 18–49 years were eligible for enrolment. Participants were excluded if they were of childbearing potential and were unwilling or unable to use a highly effective method of contraception for at least 3 months after study vaccination; if they had a history of microbiologically proven invasive disease caused by GBS; or if they had any previous vaccination with any licensed or investigational GBS vaccine or vaccination with diphtheria-containing or cross-reactive material 197 (CRM\textsubscript{197})-containing vaccines from 6 months before investigational product administration (or planned receipt of these vaccines up to 6 months after vaccination with the study product).

The final protocol and informed consent document were approved by institutional review boards for each of the investigational centres participating in this study, and this study was done in compliance with International Council for Harmonisation Good Clinical Practice guidelines and the ethical principles of the Declaration of Helsinki. Signed and dated written informed consent was required from each participant before any study-specific activity was done. The protocol for this study is included in the appendix (p 23).

Randomisation and masking
Study participants were randomly assigned using an interactive, web-based response technology system following an allocation schedule that was created before the study start. Randomisation was done at each dose level. Participants were enrolled into the lowest dose group, within which participants in a sentinel cohort were randomly assigned 2:2:1 to receive GBS6 with aluminium phosphate (AlPO\textsubscript{4}), GBS6 without AlPO\textsubscript{4}, or placebo (saline control) at a rate limited to a maximum of four participants per day. Expanded cohort participants within the lowest dose group were then randomly assigned in a 4:4:1 ratio to receive GBS6 with AlPO\textsubscript{4}, GBS6 without AlPO\textsubscript{4}, or placebo, with no limit on the rate of enrolment. The same randomisation process was then repeated with participants being enrolled into the medium dose group after the review of safety data from the low dose group; enrolment started into the high dose group after review of safety data from the medium dose group. The appearance of GBS6 and placebo was not matched, which required an unmasked vaccine administrator at each study site. The unmasked administrators were not involved in other aspects of the study. Internal review committee and external data monitoring committee members reviewed unmasked safety data throughout the study. After a prespecified interim analysis at 1-month postvaccination, the study was unmasked to the sponsor’s study team. Sponsor laboratory personnel responsible for doing immunogenicity testing remained masked until study completion. Participants and all other study staff were masked to the allocation throughout the study. The staff who observed the patients were masked.

Procedures
Three doses (5 µg capsular polysaccharide per serotype in the low-dose group, 10 µg capsular polysaccharide per serotype in the medium-dose group, or 20 µg capsular polysaccharide per serotype in the high-dose group) of the GBS6 vaccine candidate were formulated with or without AlPO\textsubscript{4}, and participants received one 0.5 mL dose of either GBS6 or placebo, administered by intramuscular injection into the deltoid muscle. GBS6 is composed of equal proportions of serotypes Ia, Ib, II, III, IV, and V capsular polysaccharide individually conjugated to the CRM\textsubscript{197} carrier protein. The placebo was a sterile saline solution for injection (0.9% sodium chloride injection, in a 0.5 mL dose). Participants received a single vaccine or placebo dose on day 1.

Adverse events were recorded in the following categories: any event (all categories combined); serious adverse event (untoward medical occurrence at any dose that results in death, is life threatening, requires inpatient hospitalisation, results in persistent or clinically significant disability or incapacity, results in congenital anomaly or birth defect, or is considered to be an important medical event, or any combination of these results); immediate adverse events (occurs within the first 30 min after study vaccination); severe adverse events (interferes substantially with the participant’s usual function); related adverse events (deemed related to study vaccination by the study investigator); medically attended adverse events (non-serious adverse events that result in an evaluation at a medical facility); and adverse events leading to withdrawal (the primary reason for a participant’s study discontinuation). Safety assessments for all participants included 30 min observation after vaccination for immediate
adverse events and self-reporting of solicited local reactions (pain at injection site, erythema or redness, and induration or swelling), systemic events (fever, nausea or vomiting, diarrhoea, headache, fatigue, muscle pain, or joint pain), and the use of antipyretic or pain medication by the patient in an electronic diary for 14 days after vaccination (appendix p 9). Unsolicited adverse events were recorded for up to 1 month after vaccination and serious adverse events and medically attended adverse events were recorded until 6 months after vaccination. Adverse events related to study procedures were recorded within 48 h after the 3-month and 6-month blood draws. For the sentinel cohort participants only, blood was collected at screening and 1 week after vaccination for haematology and chemistry assessments (details are included in the protocol; appendix p 23).

The protocol specified safety stopping rules (appendix p 10) for GBS6-vaccinated participants. An internal review committee reviewed the 14-day safety data from each sentinel cohort to determine if expanded cohort enrolment should begin at that dose and if sentinel cohort enrolment for the next higher dose should begin. This study used an external data monitoring committee that provided regular review of cumulative safety data and ad-hoc review if a stopping rule was met.

For immunogenicity assessments, approximately 30 mL of blood was collected from each participant before study vaccination and at 1 and 2 weeks and 1, 3, and 6 months after vaccination. Immunogenicity was measured by IgG antibody concentrations using a direct-binding Luminex (Luminex Corporation, Austin, TX, USA) assay platform. Each of the six GBS capsular polysaccharides included in GBS6 was individually conjugated to poly-L-lysine and the conjugates were chemically coupled to spectrally distinct magnetic Luminex microspheres. The six-plex pool of microspheres was incubated with diluted serum samples to allow serotype-specific primary antibodies to bind to the immobilised polysaccharide antigens. Antibodies that bound to the polysaccharide-coated beads were detected with R-phycocerythrin-conjugated goat antihuman IgG secondary antibodies. Data were captured as median fluorescent intensities using a Luminex reader and converted to µg/mL antibody concentrations using a reference standard curve and accounting for the serum dilution factor. Assay results are reported in weight-based µg/mL units of IgG antibodies. Pfizer’s six-plex direct-binding Luminex assay was qualified to measure serotype-specific serum IgG antibodies against GBS6 serotypes Ia, Ib, II, III, IV, and V with appropriate accuracy, precision, and specificity.

**Outcomes**

The primary outcomes were the proportion of sentinel cohort participants with clinical laboratory abnormalities.
1 week after vaccination; the proportion of sentinel and expanded cohort participants reporting solicited local reactions, systemic events, and use of antipyretic or pain medication within 14 days after vaccination; adverse events up to 1 month after vaccination; and medically attended adverse events and serious adverse events up to 6 months after vaccination. The secondary outcome was GBS serotype-specific IgG geometric mean concentrations 1 month after vaccination. Exploratory outcomes were GBS serotype-specific antibody (IgG) geometric mean concentrations measured at 1 and 2 weeks and 1, 3, and 6 months; and the proportion of participants with serotype-specific IgG concentrations ≥1.0 µg/mL at baseline, 1 week, 2 weeks, and 1 month.

The immunogenicity endpoints (secondary and exploratory outcomes) were analysed in the evaluable immunogenicity population, which included participants who were eligible, received vaccination as randomly assigned, had at least one valid and determinate assay result for the 1-month-after-vaccination visit within a prespecified window (within 27–45 days, inclusive, after vaccination), and had no potentially important protocol violations.

Descriptive summary statistics were calculated for all immunogenicity endpoints. Antcapsular IgG antibody concentrations for the six serotypes at all immunogenicity assessment timepoints were reported as GBS serotype-specific concentrations and were logarithmically transformed for analysis. For each serotype, geometric mean concentrations and the associated two-sided 95% CI from each vaccine group were calculated by back transformation of the mean and CI of the logarithmically transformed assay results computed using Student’s t distribution. In addition, mixed-effects models with repeated measurements were used to analyse the GBS serotype-specific IgG antibody concentrations and assess the effect of specific covariates. The proportions of participants achieving serotype-specific IgG concentrations of 1·0 µg/mL or more were also descriptively summarised with two-sided 95% exact CIs. Post-hoc analyses were done for proportions of participants achieving serotype-specific IgG concentrations of 0·5 µg/mL or more in a similar manner. This study is registered with ClinicalTrials.gov, NCT03170609.

### Table 1: Baseline characteristics in the safety population

|                          | GBS6 5 µg with APO4 (n=52) | GBS6 5 µg without APO4 (n=52) | GBS6 10 µg with APO4 (n=52) | GBS6 10 µg without APO4 (n=52) | GBS6 20 µg with APO4 (n=52) | GBS6 20 µg without APO4 (n=52) | Placebo (n=52) | Total (n=364) |
|--------------------------|-----------------------------|--------------------------------|-----------------------------|--------------------------------|-----------------------------|--------------------------------|----------------|---------------|
| **Sex**                  |                             |                                |                             |                                |                             |                                |                 |               |
| Female                   | 35 (67%)                    | 41 (79%)                       | 32 (62%)                    | 33 (64%)                       | 35 (67%)                    | 37 (71%)                       | 43 (83%)        | 256 (70%)     |
| Male                     | 17 (33%)                    | 11 (21%)                       | 20 (39%)                    | 19 (37%)                       | 17 (33%)                    | 15 (29%)                       | 9 (17%)         | 108 (30%)     |
| **Race**                 |                             |                                |                             |                                |                             |                                |                 |               |
| White                    | 44 (85%)                    | 41 (79%)                       | 46 (89%)                    | 41 (79%)                       | 45 (87%)                    | 46 (89%)                       | 44 (85%)        | 307 (84%)     |
| Black or African American| 5 (10%)                     | 9 (17%)                        | 5 (10%)                     | 8 (15%)                        | 4 (8%)                      | 4 (8%)                         | 7 (14%)         | 42 (12%)      |
| Other                    | 2 (4%)                      | 1 (2%)                         | 0                           | 2 (4%)                         | 2 (4%)                      | 2 (4%)                         | 0               | 9 (3%)        |
| Asian                    | 1 (2%)                      | 1 (2%)                         | 1 (2%)                      | 1 (2%)                         | 0                           | 1 (2%)                         | 0               | 5 (1%)        |
| Native Hawaiian or other Pacific Islander | 0 | 1 (2%) | 0 | 0 | 0 | 0 | 1 (<1%) | 1 (<1%) |
| **Ethnicity**            |                             |                                |                             |                                |                             |                                |                 |               |
| Non-Hispanic or non-Latino| 50 (96%)                    | 47 (90%)                       | 50 (96%)                    | 49 (94%)                       | 47 (90%)                    | 51 (98%)                       | 47 (90%)        | 341 (94%)     |
| Hispanic or Latino       | 2 (4%)                      | 4 (8%)                         | 2 (4%)                      | 2 (4%)                         | 5 (10%)                     | 1 (2%)                         | 5 (10%)         | 21 (6%)       |
| Not reported             | 0                           | 1 (2%)                         | 0                           | 0                              | 0                           | 0                              | 0               | 1 (<1%)       |
| Unknown                  | 0                           | 0                              | 1 (2%)                      | 1 (2%)                         | 0                           | 0                              | 0               | 1 (<1%)       |
| **Age at vaccination, years** |                          |                                |                             |                                |                             |                                |                 |               |
| Mean (SD)                | 32·9 (8·43)                 | 34·7 (9·35)                    | 32·8 (8·31)                 | 31·9 (8·10)                    | 33·2 (8·67)                 | 30·5 (8·25)                    | 32·8 (8·74)     | 32·7 (8·57)   |
| Median                   | 31·0                        | 36·5                           | 31·0                        | 30·0                           | 32·0                        | 30·0                           | 32·0                        | 32·0           |
| Range                    | 20–49                       | 18–49                          | 18–49                       | 19–45                          | 18–49                       | 18–49                          | 18–49           | 18–49         |

Data are n (%) unless otherwise stated. Due to rounding, percentages might not add up to 100%. Race and ethnic group were reported by participants. APO4 =aluminium phosphate. GBS6=group B streptococcus-containing vaccine with serotypes Ia, Ib, II, III, IV, and V.
Role of the funding source
Pfizer was responsible for the study design, data collection, data analysis, data interpretation, and writing of the report. The corresponding author had full access to all the data in the study and had final responsibility for the decision to submit for publication. All study data were available to all authors.

Results
Between June 5, 2017, and June 25, 2018, 485 individuals were screened, 365 of whom were enrolled into the study. 53 participants were randomly assigned to the GBS6 5 µg with AlPO₄ group, and 52 participants were randomly assigned to each of the six other study groups (five GBS6 groups and one placebo group; figure 1). One patient in the GBS6 5 µg with AlPO₄ group was subsequently found to be ineligible and did not receive the study vaccination. 348 (95%) of 365 participants completed the study. The most common reasons for not completing the study included being lost to follow-up (ten [3%] of 365 participants) or withdrawal (four [1%]). No participant was withdrawn because of an adverse event. A specific target for participant enrolment by sex was not pre-specified; however, the intention was for each site to enrol at least 50% women within each dose group. The majority of participants were women (256 [70%]), White (307 [84%]), and non-Hispanic (341 [94%]), with a mean age of 32·7 years (SD 8·57), and within each of the study groups, demographic characteristics were fairly similar (table 1).

Data from the 364 participants who received study vaccine or placebo are included in the safety analyses. Among participants in the sentinel cohorts, two (8%) of 24 placebo recipients and 20 (21%) of 96 GBS6 recipients had grade 1 or higher abnormal laboratory values at the 1-week follow-up visit. One (6%) of 16 participants in the 5 µg GBS6 without AlPO₄ group had a grade 3 aspartate aminotransferase abnormality (193 U/L; reference range 11–36 U/L); all other abnormalities were classified as grade 1 or 2, with the majority being grade 1 (appendix p 15). Of the participants whose testing was repeated 1 week after the
Figure 3: Participants reporting systemic events, by maximum severity, within 14 days after study vaccination

Grading of solicited systemic events is described in the appendix (p 9). AlPO₄ = aluminium phosphate. GBS6 = group B streptococcus-containing vaccine with serotypes Ia, Ib, II, III, IV, and V.
abnormal result, all laboratory abnormalities were found to have normalised after retesting. None of the changes in laboratory values after vaccination were associated with clinical findings.

In the 14 days after vaccination, pain at the injection site was the most frequently reported solicited local reaction, ranging from 13 (25%) of 52 participants in the GBS6 20 µg without AlPO4 group to 28 (54%) of 52 participants in the GBS6 5 µg with AlPO4 group. Injection-site pain was reported slightly more frequently among participants in the AlPO4-containing GBS6 groups (figure 2). Most local reactions were mild and no severe or grade 4 local reactions were reported (figure 2). The median duration of any local reaction ranged from 1·0 to 3·5 days among GBS6 recipients and was generally similar across groups.

Solicited systemic events reported within 14 days after vaccination for GBS6 and placebo groups are shown in figure 3. Most systemic events reported among GBS6 recipients were mild or moderate in nature, and similar frequencies and severities were observed across all doses and formulations (ranging from 26 (50%) participants in the GBS6 10 µg with AlPO4 group to 36 (69%) in the GBS6 5 µg with AlPO4 group). No grade 4 systemic events were reported. The most frequently reported systemic events among GBS6 recipients and placebo recipients were fatigue or tiredness and headache. No more than one (2%) of 52 participants at any dose reported a fever greater than 102·1°F (38·9°C). The median duration of any reported systemic reaction ranged from 1·0 to 3·5 days among GBS6 recipients and was generally similar across groups.

In the 14 days after vaccination, the most common of which were injection-site reactions and headache (one participant in the GBS6 20 µg without AlPO4, group had injection site discoloration; one participant in the GBS6 10 µg with AlPO4, group and one participant in the GBS6 20 µg with AlPO4, group had headache). No related adverse events

![Table 2: Unsolicited adverse events in participants in the safety population](https://www.thelancet.com/infection)
were reported in the placebo group. No immediate adverse events were reported during the study (table 2).

Of the 365 participants enrolled into the study, 11 were excluded from the evaluable immunogenicity population, including one participant who did not receive the vaccine and ten who at 1 month after vaccination were withdrawn for one or more of the following reasons: invalid or indeterminate serological results (five participants), blood samples drawn outside of the blood draw window (three), or a potentially important protocol deviation (three).

Overall, few study participants had prevaccination antibody concentrations below the lower limit of quantitation (data not shown). IgG geometric mean concentrations after vaccination were generally lower in these participants than in those with pre-existing antibodies. However, geometric mean fold rises stratified by baseline IgG concentration were similar between groups irrespective of baseline concentration (data not shown).

GBS serotype-specific IgG geometric mean concentrations increased rapidly from baseline to 1 week after vaccination across all doses and formulations of GBS6 compared with placebo, peaking by approximately 2 weeks after vaccination for most serotypes and remaining stable up to 1 month after vaccination (figure 4; appendix p 11). Although IgG geometric mean concentrations waned by 6 months after vaccination, they remained markedly higher than baseline concentrations and compared with placebo for all doses and formulations of GBS6. No difference in the GBS serotype-specific IgG geometric mean concentration immune response was observed between the GBS6 doses and formulations (figure 4). GBS serotype-specific IgG geometric mean fold rises ranged from approximately 25 to more than 200 for each serotype 1 month after vaccination (appendix p 8). Additionally, GBS serotype-specific IgG geometric mean fold rises remained substantially high for all doses and formulations of GBS6 at 6 months after vaccination (approximately 10-fold to 56-fold rise at 6 months after vaccination) compared with placebo (appendix p 8).

The proportion of participants with an IgG antibody concentration of 1·0 μg/mL or more 1 month after vaccination varied by serotype and by dose and formulation. However, at least 75% of participants for at least one dose and formulation group had an antibody concentration of 1·0 μg/mL or more for five of the six vaccine serotypes (Ia, II, III, IV, and V). For serotype Ib, 40–57% of GBS6 participants had reached this threshold at 1 month after vaccination. Geometric mean fold rises from baseline to
1 month after vaccination for serotype Ib were similar to the other serotypes (a 40-fold to 56-fold rise; appendix pp 2, 8). Similar results were observed in post-hoc analyses, using GBS IgG concentration threshold of 0·5 μg/mL for serotypes Ia, II, III, IV, and V (appendix p 5).

Effects of blood sampling timepoint by vaccination group, sex, age, serotype-specific baseline colonisation status, and baseline IgG concentration (on a log scale) were assessed using a mixed-effects model with repeated measurement for serotype-specific IgG concentrations (data not shown). The model-based GBS serotype-specific IgG geometric mean concentrations show similar patterns as the unadjusted GBS serotype-specific IgG geometric mean concentrations. Of the terms evaluated in the model, baseline IgG concentration was strongly correlated with immune response for all six serotypes. Sex, age at vaccination, and baseline colonisation did not considerably or consistently affect the IgG immune response.

**Discussion**

Overall, GBS6 was safe and well tolerated across all doses and formulations. The vaccine’s safety profile was characterised primarily by mild injection-site pain of short duration. There was no evidence of a worsening of reactogenicity with higher doses. The occurrence of local reactions and systemic events with GBS6 was similar to those reported for vaccines that are licensed for use in adults, as reported by Bryant and colleagues,21 and for a different investigational GBS–CRM197 conjugate vaccine, as reported by Madhi and colleagues,9 as well as other vaccines that might be recommended in pregnancy.22-25

GBS6 induced robust immune responses (as measured by serotype-specific IgG) among vaccine recipients, regardless of dose or formulation. Although we did not observe differences in this non-pregnant population, it is possible that in pregnant women, dose or formulation differences could be observed that directly affect antibody concentrations in the infant. Multiple doses as well as adjuvanted and non-adjuvanted formulations will need to be evaluated in initial studies of GBS6 in pregnant women. For all GBS6-vaccinated groups, a rapid rise in IgG antibody concentration was observed at 1 week after vaccination. Peak responses occurred by 2 weeks and were maintained up to 1 month after vaccination. The rapid responses observed within the first 2 weeks after vaccination might offer an important advantage for pregnant women who might deliver sooner than expected in their third trimester. Geometric mean fold rises at 1 month after vaccination ranged from approximately 25 to more than 200 compared with baseline concentrations. There was a slow decline in IgG concentrations starting at 3 months; however, IgG antibody concentrations and geometric mean fold rises remained substantially elevated from baseline at 6 months after vaccination. This persistent immune response might be important for protection from disease caused by GBS in the mother (eg, post-partum caesarean wound infections)26 and might be an indicator that protection in the infant persists up to at least 3 months of age, with the potential to affect not only early onset disease, but also late-onset disease, for which current prevention approaches are not effective.

A general correlation between greater maternal GBS antibody concentrations and reduced likelihood of GBS disease in newborn babies has been observed in several seroepidemiological studies. The goal of these studies was to identify a maternal serological correlate of protection against GBS disease in the infant to help facilitate vaccine development.27-29 Baker and colleagues27 did a Bayesian evaluation of absolute disease risk that provided predictions of serotype III early onset disease risk corresponding to a 100% reduction relative to the risk in the general population of colonised women for serotype III-specific IgG concentrations of more than 0·5 μg/mL. For serotypes Ia and V, higher serotype-specific concentrations (>1 to 2 μg/mL) predicted reductions in early onset disease risk of approximately 50–60%. Results reported by Fabbri and colleagues28 for serotypes Ia and III in European infants were generally consistent with these findings, predicting risk reductions of approximately 75–80% for antibody concentrations of more than 1 to 1·5 μg/mL. Dangor and colleagues30 noted a 90% risk reduction (in early onset disease and late-onset disease combined) in South African infants when using antibody concentration thresholds of 3–6 μg/mL, depending on serotype. Thus, a sufficient concentration of maternal capsular-specific antibody at delivery might provide protection to infants against invasive GBS disease.

Results from these seroepidemiological studies are limited by geographic variability, small samples sizes, and use of different assays, which make direct comparison of proposed protective antibody titres difficult. However, all the studies showed an association between greater maternal antibody concentration and decreased infant disease risk, suggesting that some sufficient concentration of maternal capsular-specific antibody at delivery might provide protection to infants against invasive GBS disease. This association was also shown in a preclinical model of GBS6 before the initiation of this study.20

We selected a threshold IgG concentration of at least 1 μg/mL at 1 month as an early assessment of the immune response after vaccination with GBS6 on the basis of the findings of Baker and colleagues.27 Our results show a robust response to the vaccine. Because there is no definitive protective threshold confirmed for any serotype, and until other seroepidemiological studies have been completed, it is unclear at this time whether IgG concentrations of at least 1 μg/mL are a conservative estimation of a protective threshold. Further studies might also elucidate whether one or multiple
antibody protective thresholds (depending on serotype) apply to multivalent GBS vaccines. A limitation of this study is that it was done in the USA among quite a homogeneous non-pregnant population, which might reduce the generalisability of our results; however, clinical assessment of GBS6 is already ongoing in another geographic region in a study of pregnant women (NCT03765073). Data from this study will provide the opportunity to assess GBS6 in diverse populations.

This first-in-human trial showed that GBS6 was safe, tolerable, and elicited an immune response among healthy, non-pregnant adults at all doses and formulations tested. The findings from this trial support further evaluation of GBS6 in pregnant women and the amount of passive antibody transfer in newborn babies.

Contributors
JA, KJC, WJW, SM, and DAS designed the overall study. WCG, KUJ, and ASA provided feedback on the study design. ILS, PCG, DP, and ASA did the immunological analyses. NS, SLB, and JP collected data as study investigators. YP developed the statistical design and oversaw the data analysis. JA drafted the original manuscript. All authors reviewed and edited multiple versions of the manuscript and approved the final version.

Declaration of interests
JA, KJC, ILS, PCG, WJW, WCG, KUJ, YP, SM, DP, and ASA are current employees of Pfizer and hold stock options. KUJ and ASA have US patent 9226525 issued. ILS has patent WO2016178123A1 issued. NS, SLB, and JP received compensation from Pfizer in their role as study investigators as part of the submitted work.

Data sharing
Upon request, and subject to certain criteria, conditions, and exceptions (see https://www.pfizer.com/science/clinical-trials/trial-data-and-results for more information), Pfizer will provide access to individual de-identified participant data from Pfizer-sponsored global interventional clinical studies conducted for medicines, vaccines, and medical devices: (1) for indications that have been approved in the USA, the EU, or both; or (2) in programmes that have been terminated (ie, development for all indications has been discontinued). Pfizer will also consider requests for the protocol, data dictionary, and statistical analysis plan. Data may be requested from Pfizer trials 24 months after study completion.

Declarations of interest
Participants as part of the submitted work.

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