Texture analysis of cartilage T$_2$ maps: individuals with risk factors for OA have higher and more heterogeneous knee cartilage MR T$_2$ compared to normal controls - data from the osteoarthritis initiative

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Abstract

Introduction: The goals of this study were (i) to compare the prevalence of focal knee abnormalities, the mean cartilage T$_2$ relaxation time, and the spatial distribution of cartilage magnetic resonance (MR) T$_2$ relaxation times between subjects with and without risk factors for Osteoarthritis (OA), (ii) to determine the relationship between MR cartilage T$_2$ parameters, age and cartilage morphology as determined with whole-organ magnetic resonance imaging scores (WORMS) and (iii) to assess the reproducibility of WORMS scoring and T$_2$ relaxation time measurements including the mean and grey level co-occurrence matrix (GLCM) texture parameters.

Methods: Subjects with risk factors for OA (n = 92) and healthy controls (n = 53) were randomly selected from the Osteoarthritis Initiative (OAI) incidence and control cohorts, respectively. The specific inclusion criteria for this study were (1) age range 45-55 years, (2) body mass index (BMI) of 19-27 kg/m$^2$, (3) Western Ontario and McMaster University (WOMAC) pain score of zero and (4) Kellgren Lawrence (KL) score of zero at baseline. 3.0 Tesla MR images of the right knee were analyzed using morphological gradings of cartilage, bone marrow and menisci (WORMS) as well as compartment specific cartilage T$_2$ mean and heterogeneity. Regression models adjusted for age, gender, and BMI were used to determine the difference in cartilage parameters between groups.

Results: While there was no significant difference in the prevalence of knee abnormalities (cartilage lesions, bone marrow lesions, meniscus lesions) between controls and subjects at risk for OA, T$_2$ parameters (mean T$_2$, GLCM contrast, and GLCM variance) were significantly elevated in those at risk for OA. Additionally, a positive significant association between cartilage WORMS score and cartilage T$_2$ parameters was evident.

Conclusions: Overall, this study demonstrated that subjects at risk for OA have both higher and more heterogeneous cartilage T$_2$ values than controls, and that T$_2$ parameters are associated with morphologic degeneration.

Introduction

Osteoarthritis (OA) is a degenerative joint disease that affects more than 27 million people in the US alone [1]. OA is characterized by biochemical and morphologic degradation of joint tissues (in particular, the articular hyaline cartilage). The process of cartilage loss is manifested by biochemical degeneration (proteoglycan loss, increased water content, collagen degradation, and chondrocyte response to tissue damage) as well as morphologic degeneration such as fibrillation and cartilage thinning [2,3]. Biochemical alterations to the articular cartilage often occur prior to morphologic degeneration [4]; thus, evaluating the biochemical composition of cartilage may be valuable for the early detection of OA.
Magnetic resonance (MR) T2 relaxation time is sensitive to biochemical changes that occur during cartilage degeneration, including alterations in hydration, collagen content, and tissue anisotropy [5]. Mean cartilage T2 has been used to distinguish subjects with early OA from healthy subjects [6]. Recent studies have suggested that, in addition to mean T2, the spatial distribution of cartilage T2 values may be important when examining the pathogenesis of OA [7-9]. Early degenerative changes of the cartilage matrix due to disease or injury are reflected by the spatial distribution of T2 values and can be quantified by grey level co-occurrence matrix (GLCM) texture analysis [10]. GLCM entropy of cartilage T2 has been found to be elevated in patients with OA as compared with controls [7,9], demonstrating that not only mean T2 [6] but also the spatial distribution of T2 values is affected by disease.

The Osteoarthritis Initiative (OAI) is a multi-center longitudinal study aimed at assessing biomarkers in OA, including those derived from MR imaging (MRI). The OAI is a cross-sectional and longitudinal dataset that includes both MRI and radiographic images of subjects, scanned annually over 4 years. MR images that can be used to assess joint morphology and cartilage T2 are available. This database provides a means to longitudinally evaluate MRI biomarkers, including T2 relaxation time in the development and progression of OA, thus providing a wealth of information on OA development and progression.

While many previous studies have evaluated subjects with symptomatic and radiographic OA [11-13], the present study evaluates subjects at risk for developing OA (but without radiographic knee degeneration or pain within the week before MRI) as well as normal controls. This patient cohort is unique, facilitating the assessment of early biochemical changes in OA which occur prior to morphologic degeneration detected by radiography. Since early morphologic degeneration in the joint may not be detected by radiography [14,15], this study uses MRI to assess cartilage and meniscus morphology. The MR whole-organ magnetic resonance imaging scores (WORMS) [16] are employed for focal knee evaluation, and MR T2 relaxation time is used for the assessment of cartilage biochemical composition. The goals of this study were (a) to compare the prevalence of focal knee abnormalities, the mean cartilage T2 relaxation time, and the spatial distribution of cartilage MR T2 relaxation times between subjects with risk factors for OA and those without them; (b) to determine the relationship between MR cartilage T2 parameters, age, and cartilage morphology as determined by WORMS; and (c) to assess the reproducibility of WORMS scoring and T2 relaxation time measurements, including the mean and GLCM texture parameters.

### Materials and methods

#### Subjects

A subset of subjects from the incidence (n = 92) and control (n = 53) cohorts of the OAI [17] was selected on the basis of the inclusion criteria of this study. The incidence cohort did not have symptomatic knee OA - criteria were no frequent knee symptoms in the past 12 months, defined as “pain, aching, or stiffness in or around the knee on most days” for at least 1 month during the past 12 months, and no radiographic tibiofemoral knee OA, defined as definite tibiofemoral osteophytes (Osteoarthritis Research Society International atlas grades 1 to 3, equivalent to Kellgren-Lawrence (KL) grade of at least 2 on fixed flexion radiographs in either knee at baseline)' [17] - but did have risk factors for OA, including being overweight (defined using gender- and age-specific cut-points for weight: males of greater than 92.9 kg and females of greater than 77.1 kg from the age of 45 to 69 years) or having knee injury (defined as a history of knee injury causing difficulty walking for at least 1 week), knee surgery (defined as a history of knee surgery, including meniscal and ligamentous repairs and unilateral total knee replacement for OA), family history of total knee replacement (defined as a total knee replacement for OA in a biological parent or sibling), or Heberden nodes (defined as self-report of bony enlargement of 1+ distal interphalangeal joint in both hands) [17]. Subjects from the control cohort had no knee symptoms or risk factors for OA.

The exclusion criteria for the study included rheumatoid arthritis, bilateral total knee joint replacement, and a positive pregnancy test. The specific inclusion criteria for this study were (a) age range of 45 to 55 years, (b) body mass index (BMI) of 19 to 27 kg/m², (c) Western Ontario and McMaster Universities Osteoarthritis Index (WOMAC) pain score of 0, and (d) KL score of 0 at baseline. These parameters were chosen in order to examine a middle-aged, non-obese, and asymptomatic population without radiographic evidence of OA. The following OAI datasets were assessed in this study: baseline clinical dataset 0.2.2 and baseline imaging datasets 0.E.1 and 0.C.2. The institutional review boards at all units participating in the OAI, including the clinical centers and the OAI Coordinating Center at University of California San Francisco, have reviewed and approved the protocol and consent forms for the OAI study. All OAI study participants signed informed consent forms for participation in the study.

#### Knee radiographs

Bilateral standing posterior-anterior fixed flexion knee radiographs were acquired at baseline. Knees were positioned in a plexiglass frame (SynaFlexer; CCBR-Synarc, Newark, CA, USA) with 20° to 30° flexion and 10°
internal rotation of the feet. In an additional reading performed for the present study, knee radiographs were graded by two radiologists (LN and WV) in consensus by using the KL scoring system [18]. The KL score included only the tibiofemoral joint and not the patellofemoral joint since the OAI used the posterior-anterior “fixed flexion” knee radiograph protocol, which is a primary protocol for tibiofemoral joint radiography.

**Magnetic resonance imaging**

MR images were obtained with four identical 3.0 Tesla scanners (Siemens Magnetom Trio, Erlangen, Germany) and quadrature transmit-receive coils (USA Instruments, Aurora, OH, USA) in Columbus, OH; Baltimore, MD; Pittsburgh, PA; and Pawtucket, RI. The following sequences were acquired and used for image analysis: sagittal two-dimensional (2D) intermediate-weighted (IW) fast spin-echo (FSE) sequence (resolution = 0.357 × 0.511 × 3.0 mm) and a coronal 2D IW FSE sequence (resolution = 0.365 × 0.456 × 3.0 mm). A sagittal 2D multi-slice multi-echo (MSME) sequence (TE1-TE7 = 10, 20, 30, 40, 50, 60, 70 ms, resolution = 0.313 × 0.446 × 3.0 mm, and 0.5 mm gap) was used for T2 measurements [19].

**Image analysis**

All images were analyzed with a Sun Workstation (Sun Microsystems, now part of Oracle Corporation, Redwood Shores, CA, USA). Knee articular cartilage was segmented manually in five compartments (patella, medial femur, medial tibia, lateral femur, and lateral tibia) as previously reported [20,21]. An IDL (Interactive Data Language, Research Systems, Boulder, CO, USA) software routine was implemented to manually segment the cartilage from the T2 maps by one operator (HA). Segmentation was performed on a slice-by-slice basis (spanning all slices), and each region of interest encompassed the entirety of the cartilage tissue. To exclude potential chemical shift artifacts or fluid from the region of interest, the user simultaneously examined the T2 map and the first echo of the MSME sequence (in neighboring slice number, and zoom.

T2 maps were computed on the basis of Equation 1 from the MSME images on a pixel-by-pixel basis by using six echoes (TE = 20 to 70 ms) and three parameter fittings accounting for noise [22,23].

\[
S(TE)^2 = S_0^2 e^{-\frac{2\times TE}{T_2}} + B^2
\]

In Equation 1, S is the signal intensity at a given echo time (TE), \(S_0\) is the signal intensity at TE = 0 ms, and B is the estimated noise at a given TE. To reduce potential errors resulting from stimulated echoes in a multi-echo Carr-Purcell-Meiboom-Gill sequence [24,25], the first echo (TE = 10 ms) was not included in the \(T_2\) fitting procedure. A noise-corrected algorithm was implemented based on results from a recent study demonstrating increased accuracy and precision of \(T_2\) relaxation time when using a noise-corrected algorithm as compared with the traditional uncorrected exponential fit [22,23].

\(T_2\) quantification was performed with an in-house program created with Matlab (MathWorks, Natick, MA, USA).

**Texture analysis**

Texture analysis was performed on a slice-by-slice basis on the cartilage \(T_2\) maps. This method is based on the GLCM as described by Haralick and colleagues [10]. The GLCM determines the frequency that neighboring grey-level values occur in an image. GLCM texture parameters, including contrast, variance, and entropy, were calculated in each cartilage region. The equations for contrast, variance, and entropy are shown below (Equations 2-4), respectively.

\[
\text{Entropy} = \sum_{i=1}^{N} \sum_{j=1}^{N} P(i,j) (-\ln P(i,j))
\]

\[
\text{Variance} = \sum_{i,j=0}^{N-1} P_{ij}(i - \mu_{ij})^2
\]

\[
\text{Contrast} = \sum_{i=1}^{N} \sum_{j=1}^{N} P(i,j)(i - j)^2
\]

Where

\[\mu_{ij} = \frac{\#(i,P_{ij})}{\#(P_{ij})}\]

P represents the probability of the co-occurrence of pixel values i and j in an image. N represents the total number of pixel value co-occurrences in the image. A pixel offset of one pixel was chosen based on the fact that approximately three to four pixels span the cartilage thickness. Analysis was performed by averaging the GLCM parameters across four orientations: 0° (corresponding to the anterior-posterior axis), 45°, 90° (corresponding to the superior-inferior axis), and 135°.

**WORMS scoring**

MR images of the right knee were reviewed on picture archiving communication system workstations (Agfa, Ridgefield Park, NJ, USA). A board-certified radiologist (WV) with 7 years of experience and a fourth-year
radiology resident (LN) with 3 years of experience read the images independently and graded meniscus, cartilage, and bone marrow lesions. Cartilage and bone marrow lesions were assessed in five compartments (patella, medial femur, medial tibia, lateral femur, and lateral tibia) by using a modified semi-quantitative WORMS [16,26,27], and the highest grade of lesion was recorded for each region. In case of disagreement, a consensus reading was performed with a musculoskeletal radiologist with 22 years of experience (TML). For calibration purposes, the first 20 cases were read simultaneously by the three readers in consensus. Compared with the original WORMS grading system, only six compartments were analyzed as relatively mild lesions were expected. This could have potentially affected the number of grade 4 or grade 6 cartilage lesions as well as grade 3 bone marrow lesions, all of which, however, are rare. Cartilage signal and morphology were scored with an 8-point scale: 0 = normal thickness and signal, 1 = normal thickness but increased signal on T2-weighted images, 2.0 = partial-thickness focal defect of less than 1 cm in greatest width, 2.5 = full-thickness focal defect of less than 1 cm in greatest width, 3 = multiple areas of partial-thickness (grade 2.0) defects intermixed with areas of normal thickness or a grade 2.0 defect of wider than 1 cm but less than 75% of the region, 4 = diffuse (at least 75% of the region) partial-thickness loss, 5 = multiple areas of full-thickness loss (grade 2.5) or a grade 2.5 lesion of wider than 1 cm but less than 75% of the region, and 6 = diffuse (at least 75% of the region) full-thickness loss. Meniscal morphology was assessed in six regions by using a modified WORMS: the medial and lateral sides of the anterior, body, and posterior regions; an additional grade was added to the meniscal classification ‘intrasubstance degeneration’ to better assess early degenerative disease. The grading scale ranged from 1 to 4: 0 = normal, 1 = intrasubstance abnormalities, 2 = non-displaced tear, 3 = displaced or complex tear, and 4 = complete destruction. Subarticular bone marrow abnormalities were defined as poorly marginated areas of increased signal intensity in the normal subchondral and epiphyseal bone marrow on T2-weighted FSE fast-suppressed MR images. A 4-point grading scale was employed to assess the size of the bone marrow abnormalities: 0 = none, 1 = minimal (less than 25% of region), 2 = moderate (25% to 50% of region), and 3 = severe (greater than 50% of region) [20].

Reproducibility

The reproducibility of WORMS scoring for meniscus, cartilage, and bone marrow tissues was investigated in 15 subjects and read out twice by two radiologists independently. An intraclass correlation coefficient (ICC) was calculated to determine the intra- and inter-reader reproducibility errors [28]. The ICC is mathematically equal to the weighted kappa using quadratic weights [29,30]. The reproducibility of mean T2 and texture analysis was determined by segmenting the cartilage in five subjects, three times by one operator (HA). The reproducibility error was calculated as the root mean square (RMS) coefficient of variation (CV) of the repeated measurements as described by Glüer and colleagues [31].

Statistical analysis

Statistical analysis was performed with STATA 11 software (StataCorp LP, College Station, TX, USA). Descriptive statistics (i.e. mean age, gender, and BMI) were calculated for each group, and differences between groups were assessed by using regression models and a Pearson chi-square test. The association between age and mean T2 was assessed by using regression models and partial correlations adjusting for group, gender, and BMI.

The primary compartmental predictors of this study were the medial femur, the medial tibia, and the average of all compartments. The medial femur and medial tibia were chosen based on the following rationale: the medial side of the knee is a concentrated region of weight-bearing [32], the medial side of the knee has a higher incidence of OA than the lateral side [33], and meniscal and cartilage lesions are more prevalent on the medial side of the joint [33]. The remaining compartments, including the lateral femur, lateral tibia, and patella, were examined in an exploratory manner. Additionally, three GLCM texture parameters were analyzed (GLCM contrast, GLCM variance, and GLCM entropy) and were regarded as representative parameters from each of the three texture groups (contrast, statistics, and order, respectively). These texture parameters were selected based on of results from previous studies demonstrating their elevation in subjects with OA [7-9].

Two separate analyses were performed to assess the prevalence of morphologic knee abnormalities in each group: the first analysis defined the prevalence of cartilage (and meniscus) lesions as present for any compartment that had WORMS of greater than 0. The second analysis defined prevalence as present for any compartment that had WORMS of at least 2. The rationale for these chosen cutoff points was to assess subjects with any features of cartilage degeneration (WORMS of greater than 0) and subjects with mild degeneration (WORMS of at least 2). The prevalence of subjects with severe degeneration (WORMS of greater than 4) [34] was scarce (5 subjects overall); thus, this study did not focus on these subjects. The differences in the prevalence of morphologic knee abnormalities between groups were assessed by using logistic regression models (independent variable: group; dependent variable:
WORMS prevalence). The prevalence of bone marrow lesions was defined as a BML score greater than 0, and a logistic regression model (described above) was performed to assess the differences in the prevalence of bone marrow lesions between groups.

The differences in T2 parameters between control group (CG) and incidence group (IG) were assessed by using regression models (independent variables: group; dependent variable: T2 parameters). To compare the differences in T2 parameters between groups, the following equation was implemented: \( \frac{(T2_{\text{parameter}}_{\text{IG}} - T2_{\text{parameter}}_{\text{CG}})}{(\text{the average standard deviation (SD) of both groups})} \).

The relationship between the prevalence of morphologic abnormalities and cartilage T2 was investigated by using regression models and partial correlations (independent variables: T2 parameter and group; dependent variable: WORMS max score\(^2\)). The WORMS max score is defined as the maximum of the WORMS in all compartments per patient. All models were adjusted for age, gender, and BMI.

**Results**

The control (n = 53) and incidence (n = 92) groups had no significant differences (\( P > 0.05 \)) in age or BMI (age-control = 50.30 ± 3.03 years, age-\( \text{incidence} = 50.65 ± 2.89 \) years, \( P = 0.49 \); BMI-control = 23.90 ± 2.23 kg/m\(^2\); BMI-\( \text{incidence} = 23.78 ± 2.25 \) kg/m\(^2\); \( P = 0.78 \)). The incidence group consisted of 50 (54.34%) females, whereas the control group consisted of 36 (67.92%) (\( P > 0.05 \)) (Table 1). The incidence group had the following distribution of risk factors: 44 had a previous injury, 19 had previous knee surgery, 19 had a family history of knee replacement, and 17 had Heberden nodes.

The reproducibility results for the WORMS grading are listed in Table 2. The intra-observer reproducibility in all tissues (meniscus, cartilage, and bone marrow) was at least 96%, whereas the inter-observer reproducibility was at least 97%. The reproducibility results for the mean cartilage T2 and the GLCM texture analysis are listed in Table 3. The mean T2 values had RMS CVs ranging from 0.85% in the lateral femur to 2% in the medial tibia. GLCM entropy exhibited the lowest CVs (<1%), whereas GLCM contrast had CVs of less than 5% in all compartments except for the lateral tibia (8.67%) and medial tibia (11.41%). The CVs for GLCM variance were less than 5%, except for the medial tibia, which had a CV of 7.91%.

A significant association (\( r = 0.19, P = 0.04 \)) was evident between subject age and mean T2 (adjusting for group, gender, and BMI) in all compartments combined for both the incidence and control groups. The GLCM texture parameters and WORMS max scores were not significantly related to age (\( P > 0.05 \)) in both groups.

The prevalence of focal knee abnormalities (cartilage lesions, bone marrow lesions, and meniscus lesions) was not significantly (\( P > 0.05 \)) different between the incidence and control groups (Tables 4, 5, 6). No significant differences between groups were observed when evaluating the overall prevalence of knee abnormalities (Table 4) or the prevalence of knee abnormalities by compartment (Tables 5 and 6). The patella had the highest prevalence of cartilage defects (WORMS of greater than 0: 54.5% in the incidence cohort and 57.7% in the control cohort), followed by the lateral tibia (WORMS of greater than 0: 19.3% in the incidence cohort and 22.2% in the control cohort). Also, meniscal tears were most abundant in the medial posterior compartment (WORMS of greater than 0: 43.1% in the incidence group and 33.3% in the control group), followed by the medial body (WORMS of greater than 0: 12.5% in the incidence group and 20.0% in the control group).

The global mean T2, GLCM contrast, and GLCM variance (medial femur, medial tibia, and average of all compartments) were significantly (\( P < 0.05 \)) elevated in

**Table 1** Subject characteristics

| Characteristic               | Incidence cohort | Control cohort | \( P \) value |
|-----------------------------|-----------------|----------------|--------------|
| Number of subjects          | 92              | 53             |              |
| Age in years, mean ± SD     | 50.65 ± 2.89    | 50.30 ± 3.03   | 0.49\(^a\)   |
| Body mass index in kg/m\(^2\), mean ± SD | 23.78 ± 2.25 | 23.90 ± 2.23 | 0.78\(^a\) |
| Number of females           | 50              | 36             | 0.10\(^b\)   |
| WOMAC pain score            | 0               | 0              |              |
| Kellgren-Lawrence score     | 0               | 0              |              |

\(^a\)Regression model; \(^b\)Pearson chi-square test. SD, standard deviation; WOMAC, Western Ontario and McMaster Universities Osteoarthritis Index.

**Table 2** Interclass correlation coefficient [28] of the whole-organ magnetic resonance imaging scores (WORMS) in the meniscus, cartilage, and bone marrow lesions

| Tissue                      | Reader 1 | Reader 2 | ICC  |
|-----------------------------|----------|----------|------|
| Meniscus                    | 0.98     | 0.96     | 0.97 |
| Cartilage                   | 0.98     | 0.96     | 0.98 |
| Bone marrow lesions         | 0.98     | 0.97     | 0.98 |

The reproducibility of WORMS scores was investigated in 15 subjects and read out twice by two readers independently. ICC, interclass correlation coefficient.
the incidence group compared with the control group. Mean T2 in the medial femur was greater in the incidence cohort (37.68 ± 2.28 ms) than in the control cohort (36.85 ± 2.16 ms). GLCM entropy (medial femur, medial tibia, and average of all compartments) was elevated in the incidence group but the differences were not significant (P > 0.05). Table 7 summarizes the average values of T2 parameters in the incidence and control groups. An additional exploratory analysis of remaining compartments demonstrated similar results, but they were not significant. Subjects at risk for OA had elevated mean T2, GLCM contrast, and GLCM variance in the lateral femur (P > 0.05), the lateral tibia (P > 0.05), and the patella (P > 0.05). The incidence and control groups had a 0.21 SD difference in mean T2, a 0.28 SD difference in entropy, a 0.31 SD difference in variance, and a 0.14 SD difference in entropy (average of all compartments). Figure 1 illustrates two representative T2 maps from a control and a subject at risk for OA, respectively. While both subjects do not have cartilage abnormalities (WORMS = 0), the subject from the incidence cohort has greater mean T2, GLCM contrast, GLCM variance, and GLCM entropy of cartilage T2.

Subjects with cartilage abnormalities (cartilage WORMS of greater than 0; n = 92) had significantly (P < 0.05) elevated cartilage T2 parameters (mean T2, GLCM variance, GLCM contrast, and GLCM entropy) than subjects without abnormalities (WORMS = 0; n = 41) in the average of all compartments, in the medial femur, and the patella. The remaining compartments did not demonstrate a significant relationship (P > 0.05). This analysis pooled the incidence and control cohorts and accounted for group in the regression model. Similar trends were observed when subdividing the analysis by group. Note that eight subjects from the control group and four subjects from the incidence group did not have whole-organ magnetic resonance imaging score (WORMS) readings available. Various subjects had lesions in multiple compartments.

A positive relationship between cartilage WORMS max score and T2 parameters (mean cartilage T2, partial correlation adjusting for age, gender, and BMI) r = 0.31, P = 0.0007, GLCM variance (r = 0.18, P = 0.04), GLCM contrast (r = 0.17, P = 0.03), and GLCM entropy (r = 0.31, P = 0.09) was demonstrated in the medial femur and across both the control and incidence groups. The remaining compartments demonstrated similar trends but the correlations were not significant (P > 0.05).

Table 3 Reproducibility (coefficient of variation) [31] of T2 measurements in five subjects segmented three times each by one operator

| Parameter        | Femur  | Tibia  | Femur  | Tibia  | Patella |
|------------------|--------|--------|--------|--------|---------|
| Mean T2          | 0.99   | 2.08   | 0.85   | 1.51   | 0.86    |
| Contrast         | 4.60   | 11.41  | 3.61   | 8.67   | 4.07    |
| Entropy          | 0.47   | 0.75   | 0.05   | 0.03   | 0.55    |
| Variance         | 2.40   | 7.91   | 2.12   | 4.31   | 3.34    |

The texture parameters analyzed were at 0 degrees and at 1 pixel offset.

Table 4 Prevalence of focal knee abnormalities in the incidence and control groups

| Lesion           | Control groupa (n = 53) | Incidence groupa (n = 92) | Odds ratiob | 95% confidence interval |
|------------------|-------------------------|---------------------------|--------------|-------------------------|
| Cartilage        |                         |                           |              |                         |
| WORMS >0         | 33 (73.3%)              | 59 (67.0%)                | 0.83         | 0.36 - 1.88             |
| WORMS ≥2         | 19 (42.2%)              | 44 (50.0%)                | 1.42         | 0.66 - 3.05             |
| Bone marrow lesions | 19 (42.2%)         | 29 (32.9%)                | 0.64         | 0.29 - 1.40             |
| Meniscus         |                         |                           |              |                         |
| WORMS >0         | 18 (40.0%)              | 42 (47.7%)                | 1.52         | 0.71 - 3.30             |
| WORMS ≥2         | 9 (15.5%)               | 22 (25.0%)                | 2.16         | 0.73 - 6.35             |

*Eight subjects from the control group and four subjects from the incidence group did not have whole-organ magnetic resonance imaging score (WORMS) readings available. All P values were greater than 0.05 using a logistic regression both when unadjusted and when adjusted for age, body mass index, and gender.

Table 5 Prevalence of cartilage abnormalities (cartilage WORMS >0 and WORMS ≥2) in the incidence and control groups by compartment

| Lesion           | Control groupa, b (n = 53) | Incidence groupa, b (n = 92) | Odds ratiob | 95% confidence interval |
|------------------|---------------------------|-----------------------------|--------------|-------------------------|
| Prevalence of WORMS >0 |                     |                             |              |                         |
| Groupa, b Femur | 11 (12.5%)               | 4 (4.5%)                    | 1.52         | 0.71 - 3.30             |
| Tibia           | 8 (9.0%)                 | 17 (19.3%)                  | 1.52         | 0.71 - 3.30             |
| Patella         | 6 (7.2%)                 | 10 (11.3%)                  | 1.52         | 0.71 - 3.30             |
| Prevalence of WORMS ≥2 |                   |                             |              |                         |
| Groupa, b Femur | 10 (11.1%)               | 3 (3.4%)                    | 1.52         | 0.71 - 3.30             |
| Tibia           | 6 (6.8%)                 | 13 (14.6%)                  | 1.52         | 0.71 - 3.30             |
| Patella         | 5 (5.5%)                 | 11 (12.1%)                  | 1.52         | 0.71 - 3.30             |

*All P values were greater than 0.05 using a logistic regression both when unadjusted and when adjusted for age, body mass index, and gender.
This study evaluated the differences in knee morphology and biochemical composition in the incidence and control groups of the OAI. While there was no significant difference in the prevalence of knee abnormalities (cartilage lesions, bone marrow lesions, and meniscus lesions) between the incidence and control groups, T2 parameters (mean T2, GLCM contrast, and GLCM variance) were significantly elevated in the incidence group. These results demonstrate that subjects at risk for OA may experience early breakdown of the cartilage extracellular matrix (ECM), such as changes to the collagen structure and increased mobility of water, prior to cartilage degeneration. It is interesting that both subject groups had neither pain (WOMAC pain = 0) nor radiographic evidence (KL score of 0 in the tibiofemoral joint) of OA at baseline yet had varying biochemical compositions. These results suggest that T2 mapping may be useful in detecting early arthritic biochemical cartilage changes that precede morphologic degeneration in OA.

While radiography did not demonstrate joint space narrowing or osteophytes in either subject group, MRI detected cartilage and meniscus defects in both groups. The patellar cartilage had the highest prevalence of abnormalities compared with the other compartments, and this corroborates previous studies in athletes [35], candidates for cartilage repair surgery [36], and controls and subjects who developed frequent knee symptoms over 15 months [34]. The posterior horn of the medial meniscus had the highest prevalence of meniscus degeneration, and this has been previously reported [37-39]. Previous studies have demonstrated discordant findings between radiographic and arthroscopic joint damage: subjects with normal radiographic KL scores often demonstrated advanced OA when arthroscopy was used [14]. Thus, soft tissue degeneration in the knee may not

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### Table 6 Prevalence of meniscus abnormalities (meniscus WORMS >0 and WORMS ≥2) in the incidence and control groups by compartment

| Group | Compartments | Incidence group (n = 92) | Control group (n = 53) |
|-------|--------------|--------------------------|------------------------|
|       | Anterior     | Medial                   | Lateral                |
|       | Body         | Posterior                | Body                   |
|       | Posterior    |                          | Posterior              |

| Group | Compartments | Incidence group (n = 92) | Control group (n = 53) |
|-------|--------------|--------------------------|------------------------|
|       | Anterior     | Medial                   | Lateral                |
|       | Body         | Posterior                | Body                   |
|       | Posterior    |                          | Posterior              |

*aAll *P* values were greater than 0.05 using a logistic regression both when unadjusted and when adjusted for age, body mass index, and gender. *b*Eight subjects from the control group and four subjects from the incidence group did not have whole-organ magnetic resonance imaging score (WORMS) readings available. Various subjects had lesions in multiple compartments.

### Table 7 Average values of T2 parameters in the incidence and control groups

| Parameter | Compartment | Incidence group (n = 92) | Control group (n = 53) | *P* value unadjusted | *P* value adjustedb | Coefficientc | 95% confidence interval |
|-----------|-------------|--------------------------|------------------------|----------------------|---------------------|--------------|------------------------|
| T2 Mean, ms | All         | 32.65 ± 1.55            | 32.07 ± 1.38           | 0.056                | 0.018               | -0.72        | -1.31 -0.12            |
|           | MF          | 37.68 ± 2.28            | 36.85 ± 2.16           | 0.042                | 0.003               | -1.18        | -1.94 -0.42            |
|           | MT          | 30.27 ± 1.88            | 29.51 ± 1.77           | 0.029                | 0.036               | -0.72        | -1.39 -0.04            |
| GLCM Contrast | All        | 248.67 ± 38.39          | 227.41 ± 35.00         | 0.005                | 0.003               | -22.81       | -37.48 -8.13           |
|           | MF          | 364.17 ± 68.98          | 333.37 ± 66.58         | 0.013                | 0.009               | -32.97       | -57.72 -8.21           |
|           | MT          | 257.30 ± 63.19          | 226.96 ± 47.83         | 0.004                | 0.013               | -25.40       | -45.49 -5.48           |
| Entropy   | All         | 6.34 ± 0.16             | 6.30 ± 0.19            | 0.142                | 0.286               | 0.03        | -0.03 -0.10 0.03       |
|           | MF          | 6.96 ± 0.17             | 6.90 ± 0.17            | 0.055                | 0.062               | 0.01        | -0.06 -0.12 0.03       |
|           | MT          | 6.17 ± 0.28             | 6.09 ± 0.34            | 0.050                | 0.127               | 0.07        | -0.07 -0.15 0.02       |
| Variance  | All         | 187.77 ± 26.69          | 171.74 ± 24.57         | 0.003                | 0.002               | -16.60       | -27.09 -6.27           |
|           | MF          | 255.64 ± 42.38          | 233.10 ± 38.60         | 0.003                | 0.001               | -24.86       | -39.82 -9.91           |
|           | MT          | 183.65 ± 37.88          | 162.66 ± 32.01         | 0.001                | 0.005               | -17.89       | -30.31 -5.47           |

*a*All average of all compartments: medial femur (MF), medial tibia (MT), lateral femur, lateral tibia, and patella. *b*P value adjusted for age, gender, and body mass index (BMI). *c*The coefficient is the difference in the T2 parameter between the incidence and control groups, adjusted for age, gender, and BMI. A coefficient below 0 indicates that the control group has a lower estimated T2 parameter in comparison with the incidence group. GLCM, grey level co-occurrence matrix.
closely correspond with joint space narrowing [14,15], and radiography may not be optimal for assessing early-stage arthritic joint degeneration.

Interestingly, the prevalence of cartilage and meniscus morphologic abnormalities was similar between controls and subjects at risk for OA. One might expect that subjects at risk for OA may have an increased number of morphologic abnormalities; however, this was not the case in this study. Similar results were reported in a study by Crema and colleagues [39], who demonstrated that the prevalence of meniscal abnormalities was similar between patients with OA (frequent knee symptoms and KL score of 2 to 3) and controls. In addition, Javaid and colleagues [34] reported that the prevalence of cartilage lesions (any feature damage, whole knee) was similar between OA subjects (KL score of 0 at baseline) who developed frequent knee symptoms over 15 months (80.6%) and controls (67.2%); however, severe cartilage lesions were significantly more prevalent in subjects with OA (22.2% in subjects with OA and 8.6% in controls). The results of these studies suggest that control subjects have a similar prevalence of morphologic abnormalities as those at risk for OA and those with mild/moderate OA; thus, the use of morphologic grading to discriminate between subjects with early OA and controls may be challenging.

While the prevalence of morphologic abnormalities was similar between groups, the mean $T_2$ significantly differed, indicating that subjects at risk for OA have altered cartilage biochemistry. Cartilage $T_2$ relaxation time is sensitive to the mobility of water in cartilage tissue [40], water content [41], and collagen fiber orientation [42]; changes to these elements of the ECM characterize the initial stages of early OA, eventually leading to gross joint degeneration as detected by morphologic MRI. The elevation of cartilage $T_2$ suggests that early cartilage biochemical changes may be of primary interest when assessing subjects at risk for OA.

While elevated mean $T_2$ values are associated with OA, the heterogeneous nature of cartilage tissue is also an important consideration when quantifying cartilage tissue integrity. Nissi and colleagues [43] reported that healthy bovine cartilage samples showed a laminar appearance while spontaneously degenerated bovine cartilage tissue did not, demonstrating changes in the distribution of cartilage ECM components with degeneration. In addition, previous studies have shown varying $T_2$ relaxation times from the cartilage-bone interface to the joint surface [24,40,44-47] and varying spatial patterns of $T_2$ values in osteoarthritic cartilage [48]. Therefore, quantifying only mean values of cartilage $T_2$ may mask important information regarding the
spatial changes occurring in the ECM during degeneration.

The results of this study demonstrated that subjects at risk for OA have localized variations in their cartilage composition, as evidenced by their elevated GLCM contrast, GLCM entropy, and GLCM variance. Specifically, GLCM contrast is a measure of the differences in neighboring pixel values; high contrast signifies that many pixels with different values are neighboring. GLCM entropy is a measure of disorder in an image; high entropy signifies that the probability of pixel co-occurrence is uniform throughout an image. GLCM variance is a measure of the distribution of pixels about the mean; high variance signifies a high dispersion of co-occurrences of relaxation times. Previous studies have demonstrated differences in the spatial distribution of cartilage relaxation times in subjects with OA and those without OA. For example, Carballido-Gamio and colleagues [9] demonstrated elevated GLCM contrast and GLCM entropy of T1 relaxation time in rotating frame (T1\textsubscript{r}) and T2 in subjects with mild OA as compared with controls; Li and colleagues [8] demonstrated elevated GLCM contrast and entropy of patellar cartilage T1\textsubscript{r} in patients with OA compared with controls; and Blumenkrantz and colleagues [7] demonstrated elevated GLCM entropy of cartilage T2 in patients with OA as compared with controls. Additionally, Burstein and colleagues [49] illustrated a loss of normal spatial dependency of cartilage T2 relaxation times in a patient with anterior knee pain and chronic chondral injury. The authors suggested that areas of high T2-weighted signal (frequently associated with cartilage injury) are often adjacent to areas with low T2. Such degenerative changes in cartilage tissue due to disease or injury are reflected by the spatial distribution of T2 values and can be quantified by GLCM texture analysis.
This study demonstrated that cartilage abnormalities were associated with elevated and more heterogeneous cartilage $T_2$ values, corroborating previous research: Blumenkrantz and colleagues [50] found an association between cartilage $T_2$ and cartilage thickness, Mosher and colleagues [51] reported changes in cartilage $T_2$ and cartilage thickness after running, Stahl and colleagues [52] reported associations between cartilage $T_2$ and cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness, and Stehling and colleagues [20] demonstrated a relationship between patellar cartilage volume and thickness. These results highlight the complex interrelationship between biochemical cartilage changes and consequent morphologic cartilage loss and suggest that biochemical cartilage composition as measured by $T_2$ may be associated with cartilage loss.

Several limitations are pertinent to this study: it may have been useful to subdivide the cartilage into weight-bearing and non-weight-bearing regions. To minimize errors due to multiple comparisons, this type of segmentation was not performed. Furthermore, other technologies such as dGEMRIC (delayed gadolinium-enhanced MRI of cartilage) or $T_{1g}$ may have been useful in investigating the ECM during OA progression; however, this study did not employ these methods, as the required MRI sequences were not acquired in the OAI protocol.

Because the feasibility of GLCM texture analysis by using the OAI dataset was demonstrated by Carballido-Gamio and colleagues [53], their study provided the foundation for the present study. The present study evaluated a larger subject cohort (145 versus 13 subjects), examined distinct subject groups (we examined subjects at risk for OA and healthy controls while Carballido-Gamio and colleagues [53] examined subjects with symptomatic and radiographic OA), and assessed joint morphology in addition to cartilage $T_2$. Thus, the present study is unique in assessing the spatial distribution of cartilage $T_2$ values by using GLCM texture analysis in a large cohort at risk for OA.

Conclusions

This study demonstrated that subjects at risk for OA have both higher and more heterogeneous $T_2$ values than controls and that subjects with cartilage abnormalities have elevated cartilage $T_2$ parameters compared with subjects without abnormalities. While joint morphology was similar in both groups, cartilage $T_2$ parameters showed significant differences, suggesting that $T_2$ relaxation time may be a valuable early marker for OA.

Abbreviations

2D, two-dimensional; BMI, body mass index; CG, control group; CV, coefficient of variation; ECM, extracellular matrix; FSE, fast spin-echo; GLCM, grey level co-occurrence matrix; ICC, intraclass correlation coefficient; IG, incidence group; IW, intermediate-weighted; KL, Kellgren-Lawrence; MR, magnetic resonance; MRI, magnetic resonance imaging; MSME, multi-slice multi-echo; OA, osteoarthritis; OAI, Osteoarthritis Initiative; RMS, root mean square; SD, standard deviation; $T_{1g}$, $T_2$ relaxation time in rotating frame; TE, echo time; WOMAC, Western Ontario and McMaster Universities Osteoarthritis Index; WORMS, whole-organ magnetic resonance imaging score; WORMS max score: the maximum of the whole-organ magnetic resonance imaging scores in all compartments per patient.

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Authors’ contributions

GBJ assisted with the study design, performed $T_2$ assessment and statistical analysis, and drafted the manuscript. TB assisted in designing the study, supervised the cartilage segmentation, and helped interpret the data and perform the analysis. JC-G developed the software for $T_2$ mapping quantification and texture analysis. LN performed WORMS grading and cartilage segmentations. WJ performed WORMS grading. HA performed cartilage segmentation. JAL participated in the study design and patient selection. CEM advised with and helped perform the statistical analysis. SM participated in the conceptual design of the study, data interpretation, and analysis. TML participated in the design of the study, interpretation of data, performing WORMS scoring, and manuscript revision. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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