Vildagliptin reduces plasma stromal cell-derived factor-1α in patients with type 2 diabetes compared with glimepiride

Kyeong Seon Park1, SooHeon Kwak1, Young Min Cho1, Kyong Soo Park1, Hak C Jang2, Seong Yeon Kim1, Hye Seung Jung1∗

1Department of Internal Medicine, Seoul National University Hospital, Seoul, and 2Department of Internal Medicine, Seoul National University Bundang Hospital, Seongnam, Gyeonggi-do, Korea

ABSTRACT
Aims/Introduction: Dipeptidyl peptidase-4 inhibitors might have pleiotropic protective effects on cardiovascular disease (CVD), in contrast to sulfonylureas. Therefore, we compared various CVD risk factors between vildagliptin and glimepiride.

Materials and Methods: We carried out a randomized, prospective and crossover trial. A total of 16 patients with type 2 diabetes whose glycated hemoglobin was >7% were randomized to add vildagliptin or glimepiride. After 12-week treatment, each drug was replaced with the other for another 12 weeks. Before and after each treatment, glucose homeostasis and CVD risk factors were assessed, and the continuous glucose monitoring system was applied to calculate glycemic variability.

Results: The mean age of the participants was 60 years, 31% were men, body mass index 25.5 kg/m² and HbA1c 8.41%. Both vildagliptin and glimepiride significantly decreased glycated hemoglobin and glycemic variability indices. Despite the improved glucose homeostasis, favorable change of CVD markers was not prominent in both the arms, along with significant weight gain. Only plasma stromal cell-derived factor (SDF)-1α decreased by 30% in the vildagliptin arm. According to regression analyses, the reduction of SDF-1α was independently associated with vildagliptin usage and serum interleukin-6 changes, but white blood cells were not related with the SDF-1α changes.

Conclusion: Compared with glimepiride, vildagliptin arrestingly decreased plasma SDF-1α, and its clinical implications should be further investigated.

INTRODUCTION
The most significant cause of mortality in diabetes mellitus is cardiovascular diseases (CVDs). Not only glycated hemoglobin (HbA1c) representing 3-month mean blood glucose, but also postprandial hyperglycemia and hypoglycemia comprising glycemic variability (GV) have been reported to be independently associated with CVD1,2. Additionally, dyslipidemia, inflammation, and oxidative stress can affect the development and prognosis of CVD3. Therefore, these factors should be taken into account in choosing treatment options for diabetes mellitus.

Dipeptidyl peptidase-4 (DPP-4) inhibitors inhibit degradation of incretin hormones and induce postprandial insulin secretion through augmented incretin effects4. Therefore, they would be preferable to the traditional insulin secretagogue, sulfonylureas, in terms of postprandial hyperglycemia and GV5,6. In addition, DPP-4 inhibitors are suggested to have various pleiotropic protective effects on the cardiovascular system7, whereas some sulfonylureas were reported to increase CVD compared with metformin8. However, the large clinical trials, Trial Evaluating Cardiovascular Outcomes with Sitagliptin (TECOS), Examination of Cardiovascular Outcomes with Alogliptin vs Standard of Care (EXAMINE) and The Saxagliptin Assessment of Vascular Outcomes Recorded in Patients with Diabetes Mellitus (SAVOR)–Thrombolysis in Myocardial Infarction (TIMI) 53, for CVD outcome of DPP-4 inhibitors did not show their superiority compared with conventional antidiabetic agents.

There has been no report from prospective trials comparing CVD outcomes between DPP-4 inhibitors and sulfonylureas; a
head-to-head trial comparing linagliptin and glimepiride (CARdiovascular Outcome Trial of LINAgliptin vs Glimepiride in Type 2 Diabetes [CAROLINA] trial) has been ongoing since 2010, with a total of 6,041 patients. A retrospective analysis using the Korean national health insurance claims database has shown an increased hazard ratio for sulfonylureas plus metformin compared with a DPP-4 inhibitor plus metformin for total CVD. Regarding CVD risk factors, DPP-4 inhibitors have been reported to be favorable to body mass index (BMI), insulin resistance and triglyceride levels compared with sulfonylureas, whereas they were comparable in low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C), arterial stiffness, blood pressure, oxidative stress and high-sensitivity C-reactive protein. Therefore, although DPP-4 inhibitors logically have more favorable influences on CVD than sulfonylureas, clinical evidence is currently lacking. In the present study, we carried out a prospective and crossover study comparing various CVD risk factors between vildagliptin and glimepiride in patients with type 2 diabetes mellitus taking metformin.

MATERIALS AND METHODS
Study design
We designed a prospective, open-labeled, crossover trial (NCT01812122). Participants were recruited at Seoul National University Hospital from May 2013 through November 2014 by the staff of the diabetes clinic. Enrollment criteria were as follows: patients with type 2 diabetes mellitus aged 20–75 years, receiving metformin monotherapy for >3 months and HbA1c >7%. We excluded patients who had liver function abnormality (threefold higher than normal range), decreased kidney function (estimated glomerular filtration rate [eGFR] <60 mL/min/1.73 m²) and pancreatic diseases. Patients with malignancies, recent history of operation and medical treatment that could affect blood glucose levels were also excluded.

We calculated the sample size according to a hypothesis that the differences of CVD markers between glimepiride and vildagliptin would come from the differences of GV. For a significant difference (a two-sided P-value <0.05) of the mean amplitude of glycemic excursions (MAGE) by 1.1 mmol/L between the agents in a crossover trial, at least 14 participants were required to provide a power of at least 90%, according to previous reports. Including 20% dropout, we collected 18 participants.

Serial numbers of the participants were determined in order of enrollment consecutively, and odd-number patients were to add glimepiride 1 mg twice daily for 12 weeks, and then switch to vildagliptin 50 mg twice daily for 12 weeks. Even-number patients were vice versa: vildagliptin first, and then glimepiride. If hypoglycemia occurs with typical symptoms and self-measured blood glucose <4.4 mmol/L, the dose of glimepiride or vildagliptin was reduced by half. At baseline and at each end of 12-week treatment, anthropometric examinations and laboratory tests were carried out. The continuous glucose monitoring system (CGMS-gold; Medtronic Minimed, Northridge, California, USA) was applied for three consecutive days, too (Figure S1).

Primary end-points were traditional, and novel CVD risk factors (presented in Table 3) and secondary end-points were composite CVD risk scores.

Clinical and laboratory parameters
Medical history, concomitant drugs and anthropometric measures, including blood pressure, heart rate, body weight, height and waist circumferences, were investigated by a trained coordinator at each visit. After 12-h overnight fasting, venous blood was collected and the following were measured: complete blood cell count with differential white blood cell (WBC) types (XE-2100 Hematology Analyzer; Sysmex Corporation, Kobe, Japan) differentiates leukocytes by simultaneously measuring volume, structure and fluorescence), HbA1c (Variant II TURBO HbA1c kit 2.0; BIO-RAD laboratories, Inc., Hercules, California, USA), 1,5-anhydroglucitol (an enzymatic colorimetric assay kit; Kyowa Medex, Tokyo, Japan), insulin (DIAsource INS-IRMA kit; Dia-source Immuno Assays, Ostiguy-Louvain-la-Neuve, Belgium), fasting plasma glucose, aspartate aminotransferase, alanine aminotransferase (Shinyang Diagnostics, Seoul, Korea), creatinine (Jaffe method; Roche Crea, Roche Diagnostic, Basel, Switzerland), LDL-C (RANDOX direct LDL cholesterol kit; Randox Laboratories Ltd, Crumlin, UK), HDL-C (HDL-C plus-Gen.3; Roche Diagnostic), high-sensitivity C-reactive protein (CRP-latex(II)X2; latex-enhanced turbidimetric immunoassay; Denka Seiken Co., Ltd., Tokyo, Japan), lipoprotein(a) (Lp[a]; immunoturbidmetry; Roche Diagnostic), B-type natriuretic peptide (chemiluminescence microparticle immunoassay using Abbott reagent and i2000 Architect analyzer; Abbott, Abbott Park, Illinois, USA), plasmogen activator inhibitor-1 (enzyme-linked immunosorbent assay using Asserachrom PAI-1 kit; STAGO, Paris, France), interleukin (IL)-6 and stromal cell-derived factor-1 alpha (SDF-1α; enzyme-linked immunosorbent assay kits; R&D System, Minneapolis, Minnesota, USA). Albumin and creatinine in the morning spot urine were also measured (immunoturbimetric assay, ALBT2; Roche, Basel, Switzerland; and Jaffe method, CREJ2; Roche, respectively) to calculate the albumin/creatinine ratio. The homeostasis model assessment of insulin resistance and homeostasis model assessment of β-cell function were calculated by an equation using fasting glucose and insulin. The eGFR was calculated by the Modification of Diet in Renal Disease method. Skewed variables were logarithmically converted for statistical analyses.

Calculation of CVD risk scores
We calculated Z-scores of changes of each CVD marker, and summed them to create a compound CVD risk score. In the case of favorable factors (HDL-C and eGFR), their Z-scores were not added, but subtracted. Markers whose relationships with CVD in diabetes mellitus were not clear (Lp[a] and
SDF-1α) were not included in the calculation of the composite scores.

**Calculation of GV from CGMS data**
The standard deviation (SD), MAGE, continuous overall net glycemic action (CONGA)-6 h, M100 and the area under the curve for blood glucose level ≥180 mg/dL were calculated using the initial 48 h of the CGMS data².

**Statistical analysis**
Data are expressed as mean ± standard deviation or median (range) or n (%) according to the variable's nature. Parametric and non-parametric paired test was carried out to compare the values before and after treatment in each treatment arm. Serial changes of serum SDF-1α were examined by the repeated measures ANOVA. The Student’s t-test, Mann–Whitney test and χ²-tests were used between the treatment arms, to compare continuous and categorical variables, respectively. The relationship of CVD markers and GV indices were identified using partial correlation analysis. Multiple linear regression analysis was carried out to identify variables that best predicted the change of SDF-1α. Statistical analysis was carried out using Spss 20 (IBM Corp., Armonk, New York, USA) and GraphPad Prism 5 (GraphPad Software, La Jolla, California, USA). P < 0.05 was considered statistically significant.

**Ethics**
The study was approved by the institutional review board of Seoul National University Hospital (IRB number H-1212-042-448), and was carried out according to the Declaration of Helsinki. All patients provided written informed consent.

**RESULTS**
A total of 18 patients were enrolled; two dropped out due to follow-up loss, and a total of 16 participants completed the study and were included in the final analysis. The baseline characteristics are shown in Table 1. Men constituted 31% of the sample, the mean age was 60.0 ± 9.6 years, BMI was 25.5 ± 4.1 kg/m², duration of diabetes mellitus was 7.4 ± 5.2 years and the dose of metformin was 1,360 ± 490 mg/day. Although we did not intentionally exclude patients with CVD, there was no history of clinical CVD according to history taking and the medical records.

Because of hypoglycemic episodes, the mean dose of glimepiride became 1.45 ± 0.34 mg/day, and that of vildagliptin 80.4 ± 9.2 mg/day after 12-week treatment. As a result, there was no statistical difference in the occurrence of hypoglycemia between the arms (Table 2). Both agents significantly decreased fasting plasma glucose and HbA1c, and increased 1,5-anhydroglucitol and homeostasis model assessment of β-cell function, but there was no difference between the agents, either. Mean blood glucose (MBG) and GV indices calculated from CGMS data also improved in both arms, except CONGA-6 h; glimepiride did not change CONGA-6 h significantly (Table 2).

Even though the GV indices seemed better after the vildagliptin treatment, there was no statistical significance compared with the glimepiride. Although GV improved by both treatments, duration of hypoglycemia (glucose less than 4.4 mmol/L) increased regardless of the agents, suggesting the improved GV was mainly caused by a reduction of hyperglycemic surges.

Despite the significantly improved HbA1c and GV, favorable change of CVD risk factors was not prominent in both the arms (Table 3). When we analyzed the changes of traditional CVD risk factors, bodyweights significantly increased in both the arms. Among novel biomarkers recently observed to be related with CVD17–22, resting heart rates increased in the glimepiride arm and Lp(a) increased in the vildagliptin arm, but the final measures were not different between the arms. The most remarkable finding was a reduction of SDF-1α from 188.1 ± 31.2 to 133.8 ± 30.8 pmol/L, by 30% in the vildagliptin arm, causing a significant difference between the two agents (P = 0.005). The composite risk scores calculated excluding Lp(a) and SDF-1α were comparable between the arms, too.

When we showed the SDF-1α levels separately according to the treatment order, a significant decrease in SDF-1α was found only after vildagliptin, and then a subsequent switch to glimepiride recovered the levels (Figure 1). Therefore, lowering of SDF-1α was a specific and reversible effect by vildagliptin. In partial correlation analyses using all the variables (presented in Tables 2 and 3), a change of SDF-1α by either agent was associated with changes of LDL-C (r = 0.361, P = 0.046), log(IL-6) (r = 0.366, P = 0.043) and log(CONGA-6) (r = 0.302, P = 0.098) after adjustment by the treatment of vildagliptin. Because SDF-1α is a regulator of immune cells and platelets23, changes in total WBC counts, differential composition and

### Table 1 | Baseline characteristic of the participants

| Variables | Values |
|-----------|--------|
| Age (years) | 60.0 ± 9.6 |
| Men (%) | 31 |
| BMI (kg/m²) | 25.5 ± 4.1 |
| Diabetes duration (years) | 7.4 ± 5.2 |
| Hypertension (%) | 60 |
| History of CVD (%) | 0 |
| Diabetic retinopathy (%) | 13 |
| Urine albumin/creatinine (mg/g) | 58.3 ± 112.4 |
| eGFR (mL/min/1.73 m²) | 91.0 ± 21.4 |
| AST (IU/L) | 27 ± 17 |
| ALT (IU/L) | 30 ± 22 |
| Metformin dose (mg/day) | 1,360 ± 490 |
| ACE inhibitors and ARB use (%) | 53 |
| Statin use (%) | 67 |

Data are presented as mean value ± SD or number (%); n = 16. ACE, angiotensin-converting enzyme; ALT, alanine transaminase; ARB, angiotensin II receptor blocker; AST, aspartate transaminase; BMI, body mass index; CVD, cardiovascular disease; eGFR, estimated glomerular filtration rate.
Table 2 | Changes in glycemic control by glimepiride and vildagliptin

|                      | Baseline       | Glimepiride   | Vildagliptin   | p†   |
|----------------------|----------------|---------------|---------------|------|
| Dose (mg/day)        | NA             | 1.45 ± 0.34   | 804 ± 9.2     | NA   |
| Symptomatic hypoglycemia, n (12 weeks) | NA             | 0.75 ± 1.24   | 0.44 ± 1.26   | 0.381 |
| SMBG at hypoglycemic episodes (mmol/L) | NA             | 43 ± 0.7      | 38 ± 0.1      | 0.464 |
| FPG (mmol/L)         | 96 ± 1.4       | 82 ± 1.8†     | 77 ± 2.4†     | 0.496 |
| HbA1c, % (mmol/mol)  | 8.4 ± 0.6 (68.0 ± 56) | 6.7 ± 0.4† (50.0 ± 25) | 6.6 ± 0.9† (49.0 ± 56) | 0.799 |
| 1,5-AG (mg/L)        | 34.8 ± 23.8    | 74.0 ± 56.6§  | 85.4 ± 42.2§  | 0.569 |
| HOMA-B               | 25.8 ± 14.4    | 53.0 ± 30.5§  | 615 ± 39.6§   | 0.499 |
| CGMS data            |                |               |               |      |
| Log(MBG) (mmol/L)    | 1.03 ± 0.11    | 0.93 ± 0.09‡  | 0.91 ± 0.12‡  | 0.547 |
| Log(MAGE) (mmol/L)   | 0.75 ± 0.17    | 0.64 ± 0.18‡  | 0.62 ± 0.15‡  | 0.768 |
| Log(SD) (mmol/L)     | 0.38 ± 0.15    | 0.29 ± 0.15‡  | 0.25 ± 0.14‡  | 0.456 |
| CONGA-6 (mmol/L)     | 67.8 ± 31.5    | 54.3 ± 23.3   | 46.9 ± 23.2‡  | 0.373 |
| Log(M100)            | 1.41 ± 0.44    | 0.99 ± 0.46§  | 0.87 ± 0.51§  | 0.493 |
| AUC180 (mmol/L·min)  | 6,103 ± 6,206  | 1,913 ± 2,566§ | 2,137 ± 4,129§ | 0.855 |
| Duration of glucose <4.4 mmol/L (min) | 8,14 ± 16,72 | 6,875 ± 184,81§ | 8,566 ± 190,56§ | 0.752 |

Data are presented as mean ± standard deviation. †Student’s t-test or Mann–Whitney test between glimepiride and vildagliptin. ‡p < 0.05 vs baseline by paired t-test or Wilcoxon matched-pairs signed rank test. §p < 0.01 vs baseline by paired t-test or Wilcoxon matched-pairs signed rank test. 1,5-AG, 1,5-anhydroglucitol; AUC180, area under the curve for glucose above 180 mg/dL; CONGA-6, continuous overlapping net glycemic action calculated with 6-h time intervals; FPG, fasting plasma glucose; HOMA-B, homeostasis model assessment for β-cell function; M100, weighted average of glucose values; MAGE, mean amplitude glycemic excursion; MBG, mean blood glucose; NA, not applicable; SD, standard deviation; SMBG, self-measured blood glucose.

Table 3 | Changes in cardiovascular risk factors

|                      | Baseline       | Glimepiride   | Vildagliptin   | p†   |
|----------------------|----------------|---------------|---------------|------|
| Traditional risk factors |                |               |               |      |
| Weight (kg)          | 65.8 ± 12.3    | 68.1 ± 12.7§  | 67.6 ± 12.8§  | 0.907 |
| Waist (cm)           | 90.6 ± 8.7     | 92.0 ± 9.6‡   | 91.8 ± 9.4    | 0.946 |
| SBP (mmHg)           | 125 ± 14       | 129 ± 18      | 126 ± 14      | 0.687 |
| DBP (mmHg)           | 80 ± 12        | 82 ± 11       | 79 ± 10       | 0.481 |
| LDL-C (mmol/L)       | 2.36 ± 0.53    | 2.41 ± 0.38   | 2.38 ± 0.59   | 0.844 |
| HDL-C (mmol/L)       | 1.37 ± 0.35    | 1.43 ± 0.42   | 1.35 ± 0.30   | 0.551 |
| Log(HOMA-IR)         | 0.44 ± 0.22    | 0.58 ± 0.33   | 0.43 ± 0.33   | 0.215 |
| Novel biomarkers     |                |               |               |      |
| Pulse pressure (mmHg)| 45 ± 8         | 47 ± 14       | 47 ± 11       | 0.945 |
| Heart rate (b.p.m.)  | 78 ± 9         | 82 ± 8§       | 80 ± 11       | 0.495 |
| eGFR (mL/min/1.73 m²)| 91.0 ± 21.4    | 89.8 ± 21.7   | 88.0 ± 18.9   | 0.796 |
| Log(Lp[a]) (µmol/L)  | −0.27 ± 0.46   | −0.21 ± 0.36  | −0.13 ± 0.32‡ | 0.514 |
| Log(IL-6) (µmol/L)   | −3.01 ± 1.48   | −3.31 ± 1.46  | −3.41 ± 1.48  | 0.842 |
| Log(hsCRP) (µmol/L)  | −0.42 ± 0.72   | −0.61 ± 0.51  | −0.42 ± 0.56  | 0.323 |
| Log(PAI-1) (µmol/L)  | 2.56 ± 0.35    | 2.63 ± 0.20   | 2.63 ± 0.14   | 0.981 |
| Log(BNP) (µmol/L)    | 1.44 ± 1.18    | 1.47 ± 0.88   | 1.33 ± 1.18   | 0.703 |
| SDF-1α (µmol/L)      | 188.1 ± 31.2   | 180.8 ± 47.7  | 133.8 ± 40.8§ | 0.005 |
| Risk scores          |                |               |               |      |
| Traditional factors  |                |               |               |      |
| Novel markers        | 0.46 ± 2.34    | 0.46 ± 2.78   | 0.317         |
| Entire markers       | 0.50 ± 4.58    | 0.50 ± 2.80   | 0.464         |

Data are presented as mean ± standard deviation. †Student’s t-test between glimepiride and vildagliptin. ‡p < 0.05 vs baseline by paired t-test. §p < 0.01 vs baseline by paired t-test. †These variables were excluded in the calculation of risk scores. BNP, B-type natriuretic peptide; DBP, diastolic blood pressure; HDL-C, high-density lipoprotein cholesterol; HOMA-IR, homeostasis model assessment for insulin resistance; hsCRP, high-sensitive C-reactive protein; IL-6, interleukin-6; LDL-C, low-density lipoprotein cholesterol; Lp(a), lipoprotein(a); PAI-1, plasminogen activator inhibitor-1; SBP, systolic blood pressure; SDF-1α, stromal cell-derived factor-1α.
platelet counts were analyzed with regard to SDF-1α, but there was no significant result. In the multiple linear regression analyses with several independent variables (Table 4), vildagliptin use was the most powerful determinant of SDF-1α change, and the change of IL-6 was also a significant factor.

Next, because GV has been suggested to increase oxidative stress and CVD risk, we examined the relationships of GV indices with CVD markers. Changes of the five indices – MAGE, SD, CONGA-6, M100 and area under the curve for blood glucose level ≥180 mg/L – by either agent were positively correlated with not only each other, but also change of MBG (P < 0.05; data not shown). The changes of GV indices also showed a positive correlation with changes of fasting plasma glucose, and a negative correlation with changes of 1,5-anhydroglucitol (P < 0.05, data not shown), as expected. Among the CVD risk factors, changes of HbA1c, plasminogen activator inhibitor-1, heart rates and the composite risk scores were positively correlated with the changes of MBG, but change of Lp(a) was negatively correlated with it (Table S1). Among the GV indices, M100 and area under the curve for blood glucose level ≥180 mg/L showed similar patterns with MBG, and adjustment with the change of MBG left nothing significant. In the case of MAGE, the association between changes of MAGE and of LDL-C was left significant after adjustment with the change of MBG. A change of SD was positively associated with novel CVD risk factors, changes of HbA1c, plasminogen activator inhibitor-1, heart rates and the composite risk scores were positively correlated with the changes of MBG, but change of Lp(a) was negatively correlated with it (Table S1). Among the GV indices, M100 and area under the curve for blood glucose level ≥180 mg/L showed similar patterns with MBG, and adjustment with the change of MBG left nothing significant. In the case of MAGE, the association between changes of MAGE and of LDL-C was left significant after adjustment with the change of MBG. A change of SD was positively associated with novel score independently from the change of MBG. A negative correlation between SD change and eGFR change was presumed to result from the increase of weight and body surface area; further adjustment with weight change removed the statistical significance.

**DISCUSSION**

In the present study, we could observe that there was a significant difference in the plasma levels of SDF-1α between glimepiride and vildagliptin, even though their favorable effects on glycemia were comparable. Except for SDF-1α levels, other
clinical and laboratory factors related with CVD were not significantly different between the agents. SDF-1α/C-X-C motif chemokine 12 is a highly-conserved chemokine, and the biological effects are mediated by the chemokine receptor, C-X-C chemokine receptor type 4. SDF-1α is a major regulator of stem/progenitor cell trafficking in the bone marrow and tissues, suggesting its role in tissue regeneration, although it is also a potent platelet agonist highly expressed in atherosclerotic plaques, suggesting its contribution to atherogenesis. SDF-1 governs the homing of endothelial progenitor cells from bone marrow to areas of vascular injury for angiogenesis and repair. There is also an association among CXCL12 genetic variation, circulating SDF-1 levels and circulating endothelial progenitor cells. Therefore, an understanding of SDF-1α–CXCR4 signaling and associated biological functions with respect to CVD seems complicated now. Recently, in the 3,359 Framingham Heart Study participants, high plasma SDF-1α was reported to be associated with older age, lower levels of HDL-C, cigarette smoking and lower CD34+ cell frequency. Cox regression (median 9.3 years) showed that high plasma SDF-1α was associated with heart failure and all-cause mortality risk, but not with new-onset CVD and myocardial infarction. As the SDF-1α was negatively correlated with circulating CD34+ frequency in the study, we could infer that constitutively high plasma SDF-1α might reflect the reactive response to low circulating CD34+ cells, and the impaired regenerative capacity might induce heart failure rather than new-onset coronary heart disease. Emerging data show that diabetes is associated with impaired bone marrow structure and function, attenuating vascular regenerative cells and contributing to vascular disease. Delayed stem cell mobilization and/or impaired differentiation towards the endothelial phenotype in type 2 diabetes mellitus might increase plasma SDF-1α; however, it has not been established.

Incretin-based antidiabetic agents, DPP-4 inhibitors’ effects on SDF-1α have been examined, because SDF-1α is one of the substrates of DPP-4. DPP-4 specifically cleaves dipeptides from substrates containing a penultimate proline or alanine residue at the NH2-terminus. SDF-1α and B-type natriuretic peptide are regarded as important substrates of DPP-4, inactivated by DPP-4. As a result, they are supposed to mediate favorable effects of DPP-4 inhibitors on CVD. DPP-4 inhibition around acute ischemic injury, such as hind limb ischemia and cardiac ischemia/reperfusion in animal models, has consistently enhanced recruitment of SDF-1α and endothelial progenitor cells in the damaged tissue, promoting tissue regeneration. Therefore, administration of DPP-4 inhibitors in the case of chronic subclinical ischemia, such as diabetes, would also be expected to increase plasma SDF-1α levels. However, in our participants, SDF-1α levels were rather decreased by vildagliptin (Table 3). In the case of non-diabetic HIV-positive patients, sitagliptin treatment up to 24 weeks also decreased serum SDF-1α levels compared with a placebo. More recently, Aso et al. also found that sitagliptin to type 2 diabetes mellitus lowered plasma SDF-1α levels compared with glimepiride. Long-term administration of DPP-4 inhibitors might induce such contradictory reduction, because active SDF-1α levels acutely increased by 4-day treatment of linagliptin. However, SDF-1α reduction could result from the assay method of the SDF-1α levels. Most researchers including the present authors used an enzyme-linked immunosorbent assay kit for total SDF-1α from the same company, including both active and inactive forms. It is possible that DPP-4 inhibitors increased active SDF-1α causing a reactive reduction in total form, which is not clear now.

Anyway, several clinical studies in type 2 diabetes mellitus consistently showed that DPP-4 inhibitors increased circulating stem/progenitor cell numbers, although causal relationships with SDF-1α change were controversial. We could not check stem cell frequency, having only total WBC, lymphocytes and monocytes as available data. Vildagliptin decreased neutrophil frequency and increased monocyte frequency without effects on total WBC counts (data not shown), but any changes of these were not related to SDF-1α change. We can speculate that decreased circulating stem cells in type 2 diabetes mellitus might induce plasma SDF-1α, and administration of DPP-4 inhibitors decreased it by stem cell mobilization. There is a report that different DPP-4 inhibitors had different effects on plasma SDF-1α levels in type 2 diabetes mellitus. If some DPP-4 inhibitors differentially increase the risk of heart failure, it would be related to the different effects on SDF-1α involving tissue protection and angiogenesis, and progenitor cell recruitment might mediate the mechanisms.

According to multiple regression analyses, plasma SDF-1α was associated with serum IL-6 independently from the use of vildagliptin (Table 4). IL-6 is one of the inflammatory markers associated with CVD risk and mortality, and it has been recently suggested as the key causal cytokine compared with CRP and fibrinogen in the pathogenesis of CVD, by large-scale human genetic and biomarker data. Specific interactions between IL-6 and SDF-1α have not been established in this setting, while there were some reports on the interrelationships between IL-6 and SDF-1α.

There were several unexpected findings for the CVD markers in the present study. Another study comparing high-dose gli-mepiride (6 mg/day) and vildagliptin (100 mg/day) reported a better profile of weight and insulin resistance in the vildagliptin arm. Unlike that study, we observed significant weight gain in both the arms similarly (Table 3). Although we did not quantitate food intake and physical activity in the present study, we presume that the weight gain was caused mainly from the improvement in hyperglycemia, because we did not reinforce concurrent lifestyle modification during the study, which can induce weight gain. The reason why the participants in the vildagliptin arm also gained weight seems to come from the crossover design in part; increased weight by glimepiride would not be easily lost by a switch to vildagliptin. In addition, there was no washout period in the study design, and the dose of glimepiride was much smaller than previous studies, which could
induce no difference with vildagliptin in weight gain. This unexpected weight gain in the vildagliptin arm could have interfered with improvements in other CVD markers.

In addition, according to the literature, an increase of heart rate by sulfonylurea (Table 3) is not a usual finding, but enhanced sympathetic activity by glibenclamide has been described. The small sample size could have also influenced this unexpected finding.

Another unexpected finding was the increase of LP(a) after vildagliptin treatment (Table 3). LP(a) changes were also negatively correlated with MBG changes (Table S1). LP(a) is an LDL-like particle consisting of an apolipoprotein. A moiety linked to one molecule of apolipoprotein B(100), and there has been highly suggestive evidence for a potentially causal role of LP(a) in affecting CVD risk in general populations. However, plasma LP(a) levels were observed to be inversely associated with type 2 diabetes mellitus, prediabetes and insulin resistance in several recent studies, and epidemiological studies of LP(a) and CVD risk in diabetes mellitus generated inconsistent results. LP(a) might differentially affect CVD risk between patients with diabetes mellitus and the general population.

Finally, contrary to previous reports between DPP-4 inhibitors and sulfonylureas, there was no significant difference in the GV indices between glimepiride and vildagliptin (Table 2). Indeed, a study examining 5-day effects of glimepiride and vildagliptin in well-controlled patients (HbA1c 7.6%) also failed to show statistically significant differences in MAGE and SD between them. Glucose fluctuation is composed of both hyperglycemia and hypoglycemia. We tried to prevent recurrent hypoglycemia by dose reduction in patients who had complained of it, as should be done in real-world practice. As a result, the dose of glimepiride was lower than that usually used in clinical studies, and there was no significant difference in the degree and frequency of hypoglycemia between glimepiride and vildagliptin (Table 2). We speculate that the attenuated risk of hypoglycemia in the glimepiride arm could have improved GV. Among the GV indices, changes in MAGE and SD were associated with changes in LDL-C and compound risk score by novel biomarkers, respectively (Table S1). MAGE has also been shown to be positively associated with oxidized LDL-C in adolescents. Therefore, among the indices examined in the present study, these two most popular GV indices seemed to be able to provide additional information about MBG with respect to CVD risk. However, because of the limitation of the small sample size of the present study, further investigation would be required for this issue.

There were several limitations to the present study; as for the study design, the participants were not blinded, not really randomized and did not undergo a washout period, although the duration of each treatment seemed long enough to counteract most effects of previous treatment. The small sample size seemed to contribute to the failure in obtaining statistically significant differences in most CVD markers between the agents. Another weak point is that there was no mechanistic study of the SDF-1α change, leaving the clinical implications unclear.

In conclusion, in poorly-controlled patients with type 2 diabetes mellitus without established CVD, vildagliptin decreased SDF-1α, which has been reported to be positively associated with heart failure and mortality, whereas glimepiride did not. This change would be especially meaningful, because other CVD markers were not so significantly different in the present small study. Plasma SDF-1α change was not related with glycemic control including GV, but serum IL-6 change was closely and independently associated.

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DISCLOSURE
The authors declare no conflict of interest.

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SUPPORTING INFORMATION
Additional Supporting Information may be found in the online version of this article:

Table S1 | Correlation analyses between glycemic variability indices and cardiovascular disease risk factors.
Figure S1 | Diagram of the study design. CGMS, continuous glucose monitoring system; CVD, cardiovascular disease; HbA1c, glycated hemoglobin; T2DM, type 2 diabetes mellitus.