Assessing the structure and temporal dynamics of seabird communities: the challenge of capturing marine ecosystem complexity

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Summary

1. Understanding interspecific interactions, and the influences of anthropogenic disturbance and environmental change on communities, are key challenges in ecology. Despite the pressing need to understand these fundamental drivers of community structure and dynamics, only 17% of ecological studies conducted over the past three decades have been at the community level.

2. Here, we assess the trophic structure of the procellariiform community breeding at South Georgia, to identify the factors that determine foraging niches and possible temporal changes. We collected conventional diet data from 13 sympatric species between 1974 and 2002, and quantified intra- and inter-guild, and annual variation in diet between and within foraging habits. In addition, we tested the reliability of stable isotope analysis (SIA) of seabird feathers collected over a 13-year period, in relation to those of their potential prey, as a tool to assess community structure when diets are diverse and there is high spatial heterogeneity in environmental baselines.

3. Our results using conventional diet data identified a four-guild community structure, distinguishing species that mainly feed on crustaceans; large fish and squid; a mixture of crustaceans, small fish and squid; or carrion. In total, Antarctic krill Euphausia superba represented 32%, and 14 other species a further 46% of the combined diet of all 13 predators, underlining the reliance of this community on relatively few types of prey. Annual variation in trophic segregation depended on relative prey availability; however, our data did not provide evidence of changes in guild structure associated with a suggested decline in Antarctic krill abundance over the past 40 years.

4. Reflecting the differences in δ15N of potential prey (crustaceans vs. squid vs. fish and carrion), analysis of δ15N in chick feathers identified a three-guild community structure that was constant over a 13-year period, but lacked the trophic cluster representing giant petrels which was identified using conventional diet data.

5. Our study is the first in recent decades to examine dietary changes in seabird communities over time. Conventional dietary analysis provided better resolution of community structure than SIA. However, δ15N in chick feathers, which reflected trophic (level) specialization, was nevertheless an effective and less time-consuming means of monitoring temporal changes.

Key-words: community, diet, pellets, procellariiform, regurgitations, resource partitioning, seabirds, stable isotopes, stomach contents, trophic guilds

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Introduction

Anthropogenic factors, including climate change and overfishing, are among the dominant forces that impact on the structure and dynamics of marine ecosystems (Halpern et al. 2008; Hoegh-Guldberg & Bruno 2010). Oceanographic changes alter energy flow within food webs by increasing or decreasing the amount of primary and secondary production available to consumers (Beaugrand, Luczak & Edwards 2009; Brown et al. 2010). In addition, fishing redirects energy flow from pathways involving heavily harvested species to those involving species that are exploited little or not at all (Link & Garrison 2002; Smith et al. 2011). Consequently, climate- and fisheries-induced changes in prey availability influence intraspecific competition and resource partitioning of marine predators (Sandvik, Coulson & Saether 2008; Forcada & Trathan 2009; Hill, Phillips & Atkinson 2013). Although our understanding of the impact of anthropogenic exploitation on predator populations has improved (Croxall, Trathan & Murphy 2002; Trathan, Forcada & Murphy 2007; Cury et al. 2011; Lauria et al. 2013), there is a pressing need to develop more integrated evaluations of ecosystem status. However, only 17% of ecological studies in the past three decades have been at the community level (Carmel et al. 2013), suggesting that the characterization of changes in food webs at multiple trophic levels remains challenging.

Extensive research on seabirds has demonstrated not only their vulnerability to environmental perturbations, but also their utility, because they integrate information on multiple taxa, as indicators of changes in the wider ecosystem (Croll et al. 2005; Frederiksen et al. 2006; Piatt, Sydeman & Browman 2007; Einoder 2009; Cury et al. 2011). As environmental variation affects different aspects of their feeding ecology, changes in diet, prey capture rates, chick provisioning and growth, and breeding success may reflect impacts ranging from relatively subtle alterations in behaviour to major repercussions for populations (Votier et al. 2008; Grémillet & Charmanier 2010; Lewison et al. 2012) and can highlight ecosystem-wide events (Miller & Sydeman 2004; Montefeltre et al. 2007; Moreno et al. 2013). Moreover, seabirds have a wide range of ecological roles from secondary to apex consumers, and as scavengers, and whole communities are accessible for sampling during the breeding season; hence, the opportunity exists for developing a reliable, multispecies and multi-trophic level indicator of the ecosystem that can be used in management and conservation (Frederiksen et al. 2006; Piatt, Sydeman & Browman 2007; Grandgeorge et al. 2008; Cury et al. 2011).

Traditionally, changes in the diet of seabirds are monitored using stomach contents, pellets or, less commonly, direct observations of prey carried by returning adults, or dropped items collected at breeding colonies. Although these approaches can be biased, the results provide reasonable taxonomic resolution (Karnovsky, Hobson & Iverson 2012) and have been invaluable for examining interspecific dietary segregation (Table 1). However, monitoring the diets of a whole community by such methods is a daunting and time-consuming task. An alternative is to use nitrogen and carbon stable isotope analysis (SIA) of bird tissues, which are less biased but provide coarser taxonomic information and are reliant on a number of assumptions (Layman et al. 2012). It is essential, however, to recognize that marine environments usually show complex spatial and temporal variation in baseline isotope signatures due to oceanographic processes (Graham et al. 2010). In the Southern Ocean, for example, SIA has been used successfully to describe seasonal changes in the isotopic niche space of the seabird community at South Georgia (Bodey et al. 2014). Nevertheless, baseline δ15N and δ13C change with latitude, sea surface temperature, nutrient and Chl-a concentration, which is reflected in consumer tissues (Cherel et al. 2007; Phillips et al. 2009; Stowasser et al. 2012), and may obscure feeding relationships and prevent the estimation of trophic level or specific prey consumption (Ménard et al. 2007; Moreno et al. 2011; Roscales et al. 2011). Therefore, any study of a seabird community should consider the complications associated with high variability in foraging ranges and use of water masses with potentially differing isotopic baselines.

The Southern Ocean has been influenced not only by sealing, whaling and fishing over the last two centuries (Murphy et al. 2007; Trathan & Reid 2009), but also shows some of the strongest signals of global climate warming (Levitus et al. 2000; Gille 2002). Retrospective analyses suggest that abundance of Antarctic krill Euphausia superba in some regions of the Southern Ocean may have declined in the last 40 years as a consequence of reduced sea-ice extent and duration (Atkinson et al. 2004; but see Loeba & Santorab 2015; Steinberg et al. 2015). Thus, establishing feasible methods to describe and monitor the structure and function of Antarctic communities is imperative for a better understanding of ecosystem status, and for developing sustainable management strategies. One of the major breeding sites in the Southern Ocean for seabirds, including many threatened species, is South Georgia (Clarke et al. 2012). During the last four decades, the feeding ecology of most species breeding at this site has been characterized using conventional techniques, but until now, there was no attempt to integrate this wealth of dietary information in a quantitative analysis of variation between and within foraging guilds. Nor has there been a formal test of the reliability or limitations of SIA as a tool for quantifying trophic community structure where there is high spatial and temporal heterogeneity in environmental baselines.

Aiming to better understand resource partitioning within the procellariiform community breeding at Bird Island, South Georgia, we reviewed information from

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conventional dietary assessment of 13 sympatric species between 1974 and 2002 to (i) assess community structure, (ii) examine for the first time, evidence for temporal changes in this structure and (iii) investigate whether the same conclusions would have been drawn if the prey data were resolved only to higher taxonomic levels (i.e. family, rather than genus or species), which would involve much reduced monitoring effort. Furthermore, by comparing conventional dietary information with stable isotope ratios (\(d^{15}N\) and \(d^{13}C\)) in seabird chick feathers collected over a 13-year period in relation to those of their potential prey from different water masses (i.e. with wide intra-specific variability in isotopic signatures), we highlight several important issues that were unresolved in the Antarctic and elsewhere.

**Material and methods**

**STUDY AREA AND SPECIES**

Bird Island (54°00′ S, 38°03′ W) is situated close to the northwest tip of South Georgia, in the maritime subantarctic (Fig. 1). The waters of the South Georgia shelf and slope are characterized by phytoplankton concentrations and rates of primary production that are among the highest in the Southern Ocean (Atkinson et al. 2001), hence the importance of this archipelago for breeding seabirds (Clarke et al. 2012). During several austral summers covering a 13-year period (here and afterwards, the breeding season is given as the year in which the chicks fledged, e.g. austral summer 2001/2002 is denoted 2002, etc.), we collected a random sample of body feathers from chicks of 11 sympatric species of Procellariiform (wandering albatross *Diomedea exulans* – 79

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### Table 1. Sources of diet information used in this study

| Species                  | Type of diet samples | Season       | Month                | Age   | Source                                      |
|--------------------------|----------------------|--------------|----------------------|-------|---------------------------------------------|
| Antarctic prion          | Regurgitations       | 1974         | February–March       | Adults| Prince (1980)                               |
|                          | Regurgitations       | 1986         | February             | Adults| Croxall, Prince & Reid (1997)               |
|                          | Regurgitations       | 1991–1992    | February             | Adults| Liddle (1994)                               |
|                          | Regurgitations       | 1994         | February             | Adults| Reid, Croxall & Edwards (1997a)            |
| Black-browed albatross   | Regurgitations       | 1975–1976    | February–March       | Adults| Prince (1979)                               |
|                          | Regurgitations       | 1986         | February             | Adults| Croxall, Prince & Reid (1997)              |
|                          | Regurgitations       | 1994         | February             | Adults| Croxall, Reid & Prince (1999)              |
|                          | Regurgitations       | 1996–2000    | February–May         | Chicks| Xavier, Croxall & Reid (2003)              |
| Blue petrel              | Regurgitations       | 1974         | December–January     | Adults| Prince (1980)                               |
| Common diving petrel     | Stomach contents     | 1973–1974    | December–March       | Chicks| Payne & Prince (1979)                      |
| Fairy prion              | Stomach contents     | 1987         | November–February    | Adults| Reid et al. (1997b)                        |
| Grey-headed albatross    | Regurgitations       | 1983         | December–February    | Adults| Prince & Copestake (1990)                  |
|                          | Regurgitations       | 1975–1976    | February–March       | Adults| Prince (1980)                               |
|                          | Regurgitations       | 1986         | February             | Adults| Croxall, Prince & Reid (1997)              |
|                          | Regurgitations       | 1994         | February             | Adults| Croxall, Reid & Prince (1999)              |
| Light-mantled sooty      | Regurgitations       | 1977–1978    | November–April       | Adults and chicks| Thomas (1981) |
| albatross                |                      |              |                      |       |                                             |
| Northern giant petrel    | Regurgitations       | 1980–1981    | January               | Chicks| Hunter (1983)                               |
| South Georgia diving     | Stomach contents     | 1972–1973    | December–March       | Chicks| Payne & Prince (1979)                      |
| petrel                   | Stomach contents     | 1986–1987    | December–March       | Adults| Reid et al. (1997b)                        |
| Southern giant petrel    | Regurgitations       | 1980–1981    | January               | Chicks| Hunter (1983)                               |
| Wandering albatross      | Regurgitations       | 1983–1984    | May–September        | Chicks| Croxall, North & Prince (1988)             |
|                          | Pellets              | 1999–2000    | May–August           | Chicks| Rodhouse, Clarke & Murray (1987)           |
|                          |                      |              |                      |       | Xavier, Croxall & Reid (2003)              |
| White-chinned petrel     | Regurgitations       | 1986         | February             | Adults| Croxall, Prince & Reid (1997)              |
| and stomach contents     |                      |              |                      |       |                                             |
| Wilson’s storm petrel    | Regurgitations       | 1996 and 1998| January–March        | Adults| Berrow & Croxall (1999)                    |
|                          | Regurgitations       | 1985         | March                | Adults| Croxall, North & Prince (1988)             |

The breeding season is given as the year in which the chicks fledged, for example austral summer 1985/1986 is denoted 1986 etc.
individuals in total, black-browed albatross Thalassarche melanophris – 51, grey-headed albatross T. chrysostoma – 58, light-mantled sooty albatross Phoebetria palpebrata – 34, northern giant petrel Macronectes halli – 59, southern giant petrel M. giganteus – 60, white-chinned petrel Procellaria aequinoctialis – 39, blue petrel Halobaena caerulea – 19, Antarctic prion Pachyptila desolata – 19, South Georgia diving petrel Pelecanoides georgicus – 2 and common diving petrel P. urinatrix – 6) to analyse d canoides georgicus – Pachyptila desolata locations reflected a wide spatial range of birds’ foraging distributions at sea (Fig. 1). These sampling fish, squid and carrion sampled in five locations within the isotope ratios of chicks with those of 20 species of crustacean, in the diet recorded at the community level. 

et al. the same procellariiform community at Bird Island (Anderson et al. 2009). Together, these prey species represent 73% of items in the diet recorded at the community level.

**ISOTOPIC ANALYSES**

 Feather samples were washed in chloroform : methanol (2 : 1 v/v) solution, dried, stored in sealed plastic bags and then later ground to a fine powder in a freezer mill operating at liquid nitrogen temperature prior to SIAs. Carbon and nitrogen isotope ratios for feathers were measured by continuous-flow isotope ratio mass spectrometry (CF-IRMS) using both a Carlo Erba (model NA 1500) EA linked to a Finnigan Tracer Mat and a Costech system, assuring good matching of results and allowing any instrument drift to be corrected. Stable isotope ratios were expressed in δ notation as parts per thousand (‰) deviation from the international standards V-Pee Dee Belemnite (carbon) and AIR (nitrogen), according to the following equation:

\[
\delta X = \left( \frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right) \times 1000
\]

where \(X\) is \(\delta^{15}N\) or \(\delta^{13}C\), and \(R\) is the corresponding ratio \(^{15}N/^{14}N\) or \(^{13}C/^{12}C\). Measurement precision of both \(\delta^{15}N\) and \(\delta^{13}C\) was estimated to be \(\pm 0.2 \text{‰}\). All values presented are the mean ± 1 standard deviation unless otherwise stated.

**DIET DATA SOURCES**

 Data on diets from regurgitations, pellets and stomach contents of 13 species (wandering albatross, black-browed albatross, grey-headed albatross, light-mantled sooty albatross, northern giant petrel, southern giant petrel, white-chinned petrel, blue petrel, Antarctic prion, fairy prion Pachyptila turtur, Wilson’s storm petrel Oceanites oceanicus, South Georgia diving petrel and common diving petrel), all collected at Bird Island, were obtained mainly from published sources and are summarized in Table 1. Prey species were summarized into 120 groups based on identification of the lowest taxonomic level: 24 to genus and 96 to species level. Dietary composition was expressed as percentage wet mass of all ingested prey, either measured or reconstructed (see below), excluding prey that were unidentified, or classified as ‘other’. If diet information from the same samples was described in separate papers (fish and cephalopod prey of wandering albatross from 1983 and 1984) or split into tables or results within the same article (diets of white-chinned petrels from 1996 and 1998, light-mantled sooty albatross from 1977 to 1978, diving petrels from 1972 to 1973 and 1973 to 1974, Wilson’s storm petrel from 1985, Antarctic prion from 1974, grey-headed and

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**Fig. 1.** Map of the sampling area. Stations sampled for potential prey within the main foraging distribution of the seabird species from this study are indicated by enlarged circles (Stowasser et al. 2012). *is marking Bird Island, the study area. The island where the procellariiform community monitored in this study breeds.
black-browed albatross from 1975 to 1976, and northern and southern giant petrel from 1980 and 1981), overall composition was calculated accordingly (for references, see Table 1). If data were expressed in terms of numerical abundance, values were converted to mass by multiplying the number of prey items by the estimated mass (diet of diving petrels from 1986 and 1987 – see Table 1). Sufficient diet data were available for wandering albatross, black-browed albatross, grey-headed albatross, white-chinned petrel, South Georgia diving petrel and Antarctic prion, to enable annual comparisons within and between species at three different taxonomic levels (species, family and group – crustaceans, squid, fish and carrion).

**Statistical Analysis**

Comparative analyses of diet among years, and between different techniques (SIA and stomach content assessment), were carried out with the PRIMER software package (Plymouth Routines In Multivariate Ecological Research, version 6; Clarke and Gorley, 2006). A variety of resemblance metrics are available, which offer different advantages and disadvantages depending on the context (Somerfield, Clarke & Olsgard 2002; Clarke, Somerfield & Chapman 2006). We calculated Bray–Curtis similarity indices, which are unaffected if taxa are absent for both samples that are being compared. This is because species can be absent for many different reasons, and it is counter-intuitive to infer that two samples are similar because neither contains particular species (Clarke, Somerfield & Chapman 2006). Diet composition and trophic niche segregation were quantified using hierarchical agglomerate clustering and non-metric multidimensional scaling (using in both cases, the Bray–Curtis similarity index), followed by analysis of similarities (ANOSIM). The key output of the pairwise tests carried out by ANOSIM is an R value that gives an absolute measure of group separation on a scale of 0 (indistinguishable) to 1 (all similarities within groups are less than any similarity between groups). R > 0.5 indicates that the groups are well separated, R > 0.5 reflects an overlap but a clear difference, and R < 0.25 means that the groups are barely separable (Plymouth Routines in Multivariate Ecological Research, version 6; Clarke and Gorley, 2006). We also conducted Bray–Curtis analyses of SI ratios, to enable a comparison between these and the conventional diet data without any potential confounding effect of a difference in methodology.

The proportions of the variance in δ¹³N and δ¹⁵N explained by prey group (crustaceans, squid, fish and carrion) and prey species (all individual species in each group) were investigated using a combination of random effects models and variance components analysis. The random effects model was fitted using the package nlme in the programme R. In the global model, the response variable was δ¹⁵N or δ¹³C, and the random effects were species nested within prey group, fitted with normal errors and an identity link. Model selection was performed using backward-stepwise removal of each of the random effects, with the significance of the consequent increase in residual variance tested using ANOVA. Variance components, expressed as proportions of the total variance, were calculated from the selected model using the package ape.

**Results**

Results of the comparison in conventional diet of 13 species at taxonomic species level are shown in Figs 2a and 3. This generated four significantly different trophic guilds (R = 0.78, P < 0.001): (i) Antarctic prion, fairy prion, blue petrel, common diving petrel, South Georgia diving petrel and Wilson’s storm petrel; (ii) black-browed albatross, grey-headed albatross, light-mantled sooty albatross and white-chinned petrel; (iii) northern giant petrel and southern giant petrel; and (iv) wandering albatross.

For the six seabird species for which there were 3 or more years of detailed conventional diet data, the species were grouped into the same three significantly different trophic guilds when prey taxonomic resolution was at species (R = 0.84, P < 0.001) and family level (R = 0.88, P < 0.001), regardless of year. These guilds comprised the following: (i) wandering albatross; (ii) Antarctic prion and blue petrel; and (iii) black-browed albatross, grey-headed albatross and white-chinned petrel (Fig. 4). In contrast, the species were no longer grouped into the same three trophic guilds when diet data were aggregated at group level. Although ANOSIM analysis showed a pattern of strong niche segregation, diet similarity values were sensitive to the taxonomic level of diet categorization (see similarity values in Fig. 4). For example, in the analysis at prey species and family level for black-browed and grey-headed albatrosses, there was a greater similarity in diet between the two species in the same year (i.e. 1994 and 2000), than within each species in different years. However, overlap between the diet of grey-headed and black-browed albatrosses in 2000 and white-chinned petrel and Antarctic prion in 1986, and overlap between the diet of wandering albatrosses in 1983, 1984, 1999 and 2000 and grey-headed and black-browed albatrosses in 1994, appeared as an artefact of analysis of diet at the coarsest level (by group).

There was no significant temporal trend in δ¹⁵N in chick feathers of the 11 species that were sampled in multiple years. In cluster analyses, these species were grouped into three significantly different trophic guilds (R = 0.82, P < 0.001): (i) Antarctic prion, blue petrel, common diving petrel and South Georgia diving petrel; (ii) black-browed albatross, grey-headed albatross, light-mantled sooty albatross and white-chinned petrel, northern giant petrel and southern giant petrel; and (iii) wandering albatross (Fig. 2b). The relative variability between years and among species is clearly illustrated in a standard δ¹⁵N–δ¹³C biplot (Fig. 5). Note that there were no significant differences in δ¹³C between the main groups of potential prey (Fig. 6), and hence the relationships between δ¹³C and distribution will be explored in more detail in another paper.

In terms of δ¹³N of prey (Fig. 6), removal of the random effects of both the prey group and species terms resulted in significant increases in the residual deviance (prey group; LR = 37.38, d.f. = 1, P < 0.0001; species; LR = 110.9, d.f. = 2, P < 0.0001). Both terms were therefore retained in the selected model. Variance components analysis of this model revealed that prey group (group 1 = crustaceans; group 2 = carrion, squid and fish) explained 82% of the variation in δ¹³N, species explained...
8%, and residual variance was 11%. The variability was over 10 times greater across the prey groups than within individual prey species sampled in areas of different productivity.

In terms of $\delta^{13}$C (Fig. 6), the removal of the prey group term resulted in a significant increase in deviance ($LR = 28.3, \text{d.f.} = 1, P < 0.0001$), but the removal of species did not ($LR = 0.0, \text{d.f.} = 1, P > 0.9$). Variance components were therefore calculated from the model that included prey group but omitted species. Prey group explained only 1.6% of the variance, and the remainder (98.4%) was residual.

**Discussion**

Our study highlighted that although the procellariiform seabird community from Bird Island includes small-to-large species from different trophic levels and with disparate foraging strategies, only 15 prey species comprised three-quarters of their diet. The cluster analyses of conventional diet indicated that despite differences in the degree of niche segregation between years depending on availability of prey, there was no evidence of a consistent change in the trophic guild structure related to the suggested decline of krill in the last 40 years (Atkinson et al. 2004). Similarly, analysis of SI data from chick feathers did not indicate substantial changes within the 13-year study period and provided a similar, if somewhat less resolved indication of trophic guild structure to the conventional diet analysis (missing a cluster representing the carrion-feeding giant petrels). Our detailed picture of a diverse Antarctic seabird community demonstrates that analysis of diet composition at higher taxonomic levels can provide reliable insights into community dynamics. In addition, so long as the potentially confounding influence of a complex underlying marine isoscape can be overcome, SIA is an effective and less time-consuming means of assessing temporal changes in community trophic structure.
The analysis of conventional diet data collected between 1974 and 2002 for the seabird community breeding at Bird Island highlighted a four guild structure defined by species feeding mainly on crustaceans (Antarctic prion, fairy prion, blue petrel, common diving petrel, South Georgia diving petrel, Wilson’s storm petrel), large fish and squid (wandering albatross), a mixture of crustaceans, small fish and squid (black-browed albatross, grey-headed albatross, light-mantled sooty albatross and white-chinned petrel) and carrion (northern and southern giant petrel; Figs 2a and 3). Although most previous community studies using conventional methods (Table 2) have not quantified fully the variation between and within foraging guilds, the descriptions of diet during the breeding period facilitate a comparison of patterns of feeding segregation in relation to the intrinsic characteristics of different oceanic environments. Temperate and polar communities foraging in productive ecosystems (i.e. frontal zones, coastal upwelling, highly productive shelves) typically include a wide diversity of feeding strategies including surface-seizing, filtering, plunge and pursuit diving. Such communities, including that at South Georgia described in this study, include specialist planktivorous and piscivorous species, together with squid consumers, apex predator–scavengers and generalists that eat squid, fish and crustaceans in various proportions (Hobson, Piatt & Pitocchelli 1994; Ridoux 1994; Sydeman et al. 1997; this study). In contrast, tropical seabird communities feeding on marine environments that show limited seasonality and low productivity mainly consist of surface predators foraging in multispecies flocks that prey largely upon flying fish (family Exocoetidae) and squid (Diamond 1983; Harrison, Hida & Seki 1983).

The comprehensive analysis reported here highlights the key food resources for the seabird community at South Georgia, which is in a region clearly affected by rapid and ongoing environmental change. The ecological significance of Antarctic krill for top predators in the Scotia Sea, including several of the albatrosses and petrels at South Georgia, has been pointed out previously (Croxall & Prince 1987; Croxall, Prince & Reid 1997; Murphy et al. 2007; Stowasser et al. 2012). Accordingly, when we considered all years and species, 32% of the community diet during the breeding period consisted of krill, supporting its...
Fig. 4. Dendrogram (left) and multidimensional scaling (right) of conventional diet data from six procellariiform species sampled in different years at Bird Island, South Georgia (WA, wandering albatross; BBA, black-browed albatross; GHA, grey-headed albatross; WCP, white-chinned petrel; AP, Antarctic prion; SGDP, South Georgia diving petrel) from several years (Table 1) based on contribution by mass at three different taxonomic levels: (a) species, (b) family and (c) group. Dashed lines in dendrograms at 23%, 35% and 70% diet similarity indicate the three trophic guilds significant at $P < 0.05$ and defined by ANOSIM analysis.

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prominent role in the transfer of energy from primary producers to seabirds. However, when considering all species except scavengers (northern and southern giant petrels), our analysis also highlighted that only 15 of the total of 120 prey species that were included (i.e. 12.5%) explained 75% of the differences between trophic guilds and comprised 78% of their diets. Thus, the structure of this community depended largely on the relative importance of a few key species of fish, crustaceans and squid, besides Antarctic krill (Table 3). This raises the issue that long-term fluctuations in the availability of a small minority of prey, not just krill, will greatly increase interspecific competition.

When feeding resources become less abundant, diet overlap may decrease and niche width increase because of the greater reliance on a wide range of suboptimal prey types (MacArthur & Pianka 1966; Krebs & Davies 1981). One compensatory response to low availability of a key prey is to switch to alternatives, which has been investigated at South Georgia only for white-chinned petrels, grey-headed and black-browed albatrosses. Krill abundance was high throughout 1996 (c. 26.7 g m\(^{-2}\)) but apparently low in early 1998 (c. 5 g m\(^{-2}\)), yet the diet of white-chinned petrels was similar between years, and krill was always the most important prey item followed by fish and squid (Berrow & Croxall 1999). In contrast, Croxall, Reid & Prince (1999) demonstrated that a fourfold difference in krill biomass between 1986 (c. 30 g m\(^{-2}\)) and 1997 (c. 7 g m\(^{-2}\)) around northwest South Georgia (Brierley, Watkins & Murray 1997) caused a reduction of 88–90% in the consumption of krill, and a compensatory increase of fish in the diet of both black-browed and grey-headed albatrosses (the latter also showed an increase in diet diversity). Although dietary overlap indices between the 2 years were very similar for the two albatrosses, the overlaps between albatrosses and penguins were greatly reduced in 1997.

An analysis of krill density in the Southern Ocean from 1926 to 2003 suggested a major decline since the 1970s as a consequence of the reduction in sea-ice extent and duration (Atkinson et al. 2004; but see Loeba & Santorab 2015; Steinberg et al. 2015). Effects of a possible krill shortage on the guild structure of the procellariiform

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Fig. 5. Mean \(\delta^{15}N\) and \(\delta^{13}C\) (±SD) in feathers of chicks of 11 procellariiform species (WA, wandering albatross; BBA, black-browed albatross; GHA, grey-headed albatross; LMSA, light-mantled sooty albatross; NGP, northern giant petrel; SGP, southern giant petrel; WCP, white-chinned petrel; BP, blue petrel; AP, Antarctic prion; SGDP, South Georgia diving petrel; CDP, common diving petrel) sampled at Bird Island, South Georgia, during several years covering a 13-year period.

Fig. 6. Mean \(\delta^{15}N\) and \(\delta^{13}C\) (±SD) in 20 species of crustacean, fish, squid and carrion (for seals Arctocephalus gazella and Macaroni penguin Eudyptes chrysolophus) sampled in four locations within the birds’ foraging distribution at sea that differed greatly in temperature, productivity and nutrients (Stowasser et al. 2012).
community remain untested. Despite the limitations associated with pooling of somewhat disparate data sets in this study, the analysis of conventional data for six species from three trophic guilds in multiple years has provided the first insights into possible temporal changes in the fundamental structure of the community at South Georgia. Although, as described above, variation between years in niche segregation may be influenced by abundance or availability of key prey, including krill, the temporal perspective provided here shows that, overall, the similarities remained much greater within than between the three trophic guilds, and thus, the main structure remains consistent through time (Fig. 4a).

The painstaking work involved in sorting and identifying diet samples to species is such that community-level analyses are labour-intensive, time-consuming and therefore expensive. One potential means of overcoming this problem is to exploit the redundancy in community data by only analysing the samples to higher taxonomic levels, such as family. For the marine macro- and meio-benthos, aggregations of species data to higher taxonomic levels have been used to assess how much information is lost compared with the full species-level analysis (Olsgard, Sommerfeld & Carr 1998; Olsgard & Sommerfeld 2000). However, to our knowledge, there are no equivalent studies for seabird communities. The MDS and pairwise comparisons in our analyses confirm that the six species were grouped according to diet composition into the same three trophic guilds regardless of whether the analysis was carried out at species or family level (Fig. 4a, b). Therefore, our results demonstrate that a less intensive monitoring programme that involves prey identification only to a coarse taxonomic level can nevertheless provide reliable insights into the structure of seabird communities.

Although general community structure could be determined from aggregated data, it remained sensitive to taxonomic resolution. In particular, the similarity within and between species in different trophic guilds depended on the level to which prey were identified (Fig. 4). To illustrate, unusual oceanographic conditions in 2000 resulted in a much greater consumption of crustaceans by both grey-headed and black-browed albatrosses, and reduced reliance on what would otherwise have been their main prey, cephalopods and fish, respectively (Xavier, Croxall & Reid 2003). This switch was reflected in the analysis carried out at species and family level, which grouped together the diet of grey-headed and black-browed albatross in 2000 (Fig. 4a, b). However, some counter-intuitive results arose as artefacts of analyses at coarser taxonomic levels (Fig. 4c).

Biogeochemical markers such as ${\delta^{15}}N$ reflect trophic level and have provided substantial insights into feeding ecology in previous studies of seabirds (Phillips et al. 2007; Moreno et al. 2010, 2013; Votier et al. 2010). However, in marine ecosystems, ${\delta^{15}}N$ not only reflects trophic interactions but also correlates with nutrient availability and primary productivity (Graham et al. 2010; Stowasser et al. 2012). As a consequence of the simultaneous influence of diet and geographic variation in ${\delta^{15}}N$ baselines, a difference between ${\delta^{15}}N$ of consumers could indicate mainly a change in the inorganic nitrogen source utilized by primary producers, a different trophic position or a combination thereof. Information on isotopic ratios of potential prey from different foraging areas is critical for distinguishing the relative importance of prey vs. habitat specialization (Bugoni, McGill & Furness 2010; Moreno et al. 2012). As a consequence of the simultaneous influence of diet and geographic variation in ${\delta^{15}}N$ baselines, a difference between ${\delta^{15}}N$ of consumers could indicate mainly a change in the inorganic nitrogen source utilized by primary producers, a different trophic position or a combination thereof. Information on isotopic ratios of potential prey from different foraging areas is critical for distinguishing the relative importance of prey vs. habitat specialization (Bugoni, McGill & Furness 2010; Moreno et al. 2012).

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Table 3. Percentages of wet mass for the most important 15 prey species in the diet of the procellariiform community breeding at Bird Island from 1974 to 2002 (Table 1)

|                       | Wandering albatross | Black-browed albatross | Grey-headed albatross | Light-mantled sooty albatross | Antarctic prion | Fairy Prion | Blue petrel | Common diving petrel | South Georgia diving petrel | White-chinned petrel | Wilson’s storm petrel | Northern giant petrel | Southern giant petrel |
|-----------------------|---------------------|------------------------|-----------------------|--------------------------------|----------------|-------------|-------------|----------------------|--------------------------|---------------------|----------------------|----------------------|---------------------|
| Squid                 | 22.0                | 7.2                    | 12.9                  | 0.9                            | 0              | 0           | 0           | 0                    | 0                        | 2.5                 | 0                     | 5.9                  | 10.0                |
| Koudakovia longimana  | 2.6                 | 2.0                    | 2.3                   | 0.6                            | 0              | 0           | 0           | 0                    | 0                        | 0                   | 0                     | 0                    | 0                   |
| Moroteuthis knipovitchi | 0.6               | 12.8                   | 27.5                  | 0.0                            | 0              | 0           | 0           | 0                    | 0                        | 2.8                 | 0                     | 0.1                  | 0.2                 |
| Martiana hyadesi      | 0.4                 | 2.0                    | 5.8                   | 0.0                            | 0              | 0           | 0.4         | 0                    | 0                        | 0                   | 0                     | 0                    | 0                   |
| Todarodes sagittatus  | 0.8                 | 3.5                    | 4.9                   | 0.0                            | 0              | 0           | 0           | 0                    | 0                        | 1.9                 | 0                     | 0.1                  | 0.2                 |
| Galiotus glaucalis    | 28.0                | 0                      | 0                     | 0.0                            | 0              | 0           | 0           | 0                    | 0                        | 0                   | 0                     | 1.6                  | 10.0                |
| Dijsostichus eleginoides | 1.0              | 13.5                   | 5.7                   | 0.0                            | 0              | 0           | 0           | 0                    | 3.1                      | 0                   | 0                     | 0                    | 0                   |
| Champsocephalus gunnari | 13.8             | 8.3                    | 3.4                   | 0.0                            | 0              | 0           | 0           | 0                    | 0                        | 0                   | 0                     | 0                    | 0                   |
| Pseudochaenichthys georgius | 0.0             | 0.1                    | 8.3                   | 0.0                            | 0              | 0           | 0           | 0                    | 0                        | 0                   | 0                     | 0                    | 0                   |
| Geotria australis     | 0.0                 | 1.5                    | 0.8                   | 2.4                            | 0              | 0           | 0           | 0                    | 8.0                      | 0                   | 0                     | 0                    | 0                   |
| Gymnoscopelus nicholsi | 0.0              | 6.3                    | 6.3                   | 0.0                            | 0              | 0           | 0           | 0                    | 0                        | 0                   | 0                     | 0                    | 0                   |
| Magnisudis primorsa  | 0.0                 | 37.4                   | 18.5                  | 38.4                           | 41.7           | 80.1        | 74.6        | 13.4                  | 59.4                      | 47.4                | 36.1                   | 18.1                  | 16.0                |
| Euphausia superba     | 0.0                 | 0                      | 0                     | 0.0                            | 12.3           | 0.9         | 0.6         | 34.1                  | 19.6                      | 0                   | 0                     | 0                    | 0                   |
| Calamoides acutus     | 0.0                 | 0                      | 0                     | 0.0                            | 21.6           | 3.2         | 0.7         | 25.3                  | 4.3                       | 0                   | 0                     | 0                    | 0                   |
| Rhincalanus gigas     | 0.0                 | 0                      | 0                     | 0.0                            | 4.8            | 15.8        | 0           | 4.8                   | 0.2                      | 0.7                 | 31.2                   | 0                    | 0                   |
| Themisto gaudichaudii | 26.4                | 27.4                   | 53.4                  | 0.9                            | 0              | 0           | 0.4         | 0                    | 7.2                       | 0                   | 0                     | 6.0                  | 14.0                |
| Fish                  | 42.7                | 29.7                   | 24.6                  | 2.4                            | 0              | 0           | 0           | 0                    | 0                        | 11.1                | 0                     | 1.6                  | 10.0                |
| Crustaceans           | 0.0                 | 37.4                   | 18.5                  | 38.4                           | 80.5           | 100.0       | 75.9        | 77.5                  | 83.5                      | 48.1                | 67.3                   | 18.1                  | 16.0                |
| Combined prey         | 69.1                | 94.5                   | 96.4                  | 41.7                           | 80.5           | 100.0       | 76.4        | 77.5                  | 83.5                      | 66.4                | 67.3                   | 25.7                  | 18.4                |

The 15 prey species are further aggregated into four major groups: squid, fish, crustaceans and all prey combined (bottom of table).
et al. 2011), particularly in regions where there are strong isotopic gradients. However, as these factors are usually to some extent conflated, especially in southern marine ecosystems, inferring seabird community structure from $\delta^{15}N$ remains a challenge. Similarly, although some recent studies have demonstrated the potential of SIA for assessing the structure of large seabird communities at a scale of 1000–2000 km (Hobson, Piatt & Pitocchelli 1994; Forero et al. 2004; Bugoni, McGill & Furness 2010), these did not assess the influence of spatial variability on isotopic signatures of the various prey species.

The need to return regularly to provision chicks constrains the foraging range of breeding seabirds and therefore provides the opportunity for separating the effect of prey specialization from that of geographic variation in $\delta^{15}N$. The most detailed analysis to date of the food web within the foraging areas of the albatrosses and petrels included in our study (Stowasser et al. 2012) revealed a clear spatial variation in $\delta^{15}N$ of particulate organic matter and several organisms, highlighting the wide intraspecific variability in isotopic signatures. A more targeted analysis restricted to potential prey of the procellariiform community indicated no significant differences between the $\delta^{15}N$ of squid, fish and carrion, but a clear distinction between the $\delta^{15}N$ of crustaceans and other types of prey (Fig. 6).

Although only one species of squid was included in this analysis and there was no detectable difference in isotope ratios between squid, fish and carrion, a more complete study of $\delta^{15}N$ in muscle of a wider range of squid species (Anderson et al. 2009) indicates that some have much higher $\delta^{15}N$ than the species reported here (i.e. 10.51–11.36‰). Reflecting the differences in $\delta^{15}N$ of potential prey (crustaceans vs. squid vs. fish and carrion), we found a clear correspondence between the four-guild community structure obtained using conventional dietary data (Fig. 2a) and that using $\delta^{15}N$ of chick feathers (Fig. 2b).

The latter also discriminated species that feed mainly on crustaceans (Antarctic prion, blue petrel, common diving petrel, South Georgia diving petrel), large fish and squid (wandering albatross) and a mixture of crustaceans, small fish and squid (black-browed albatross, grey-headed albatross, light-mantled sooty albatross, white-chinned petrel). However, given the similarity in isotope ratios of fish and carrion (Fig. 6), analysis of $\delta^{15}N$ of feathers failed to discriminate the scavenging giant petrels from black-browed, grey-headed and light-mantled sooty albatrosses, and white-chinned petrel. There were no data available for a direct assessment of temporal variation in the $\delta^{15}N$ baseline across the very large foraging ranges of the procellariiform species included here. However, stable isotope ratios in chick feathers sampled from multiple species from 2001 to 2013 indicated that differences in $\delta^{15}N$ between years were much less than those between trophic levels (Figs 5 and 6); hence, annual variation in baselines will have minimal impact on the isotopic assessment of trophic relationships. Although conventional dietary analysis provided better resolution of the community structure, our study also demonstrates that despite the potentially confounding influence of natural biogeochemical gradients in baseline stable isotope signature, $\delta^{15}N$ in chick feathers is determined largely by trophic (level) specialization and therefore can also be used to monitor changes in the structure of the community.

Previous isotopic studies have highlighted that patchy knowledge of spatial heterogeneity in stable isotope signatures means that values in predator tissues require careful interpretations (Cherel & Hobson 2007; Phillips et al. 2009; Weiss et al. 2009; Moreno et al. 2011). The detailed picture of the seabird community and associated food web in the Antarctic ecosystem provided here indicates that to obtain a reliable estimate of trophic level or given the extensive overlap in isotope ratios, the proportion of a specific fish, squid or crustacean in the diet using mixing models may be impossible when diets are diverse, and seabirds feed in more than one water mass. As shown here, however, it is possible to use this pragmatic approach to reconstruct overall community trophic structure if the diets consist of components that are isotopically distinct at a coarse taxonomic level (crustaceans vs. fish and carrion vs. squid), providing an effective means for assessing long-term changes in community interactions.

Our review highlighted that in the past four decades, barely 20 published studies have attempted to describe seabird communities, only seven of which considered more than 10 species and none monitored temporal changes (Table 2). By comparing conventional diet with isotopic data from predators, our analyses both explored the limitations and demonstrated the potential of combining multiple lines of evidence. The scarcity of such studies reflects a profound gap in knowledge of the basic mechanisms driving seabird community structure, and highlights the necessity of further research.

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Data accessibility

Data from this study are archived at the British Antarctic Survey Data Holdings, http://doi.org/6m5

References

Ainley, D.G., Fraser, W.R., Smith, W.O., Hopkins, T.L. & Torres, J.J. (1991) The structure of upper level pelagic food webs in the Antarctic: effect of phytoplankton prey distribution. Journal of Marine Systems, 2, 111–122.
