Aim of the study: To determine the significance of expression of synaptophysin, chromogranin A, and Ki-67 and their association with clinicopathological parameters, and to find out the possible prognostic factors in gastric neuroendocrine carcinoma (G-NEC).

Material and methods: We investigated the immunohistochemical features and prognosis of 62 G-NECs, and evaluated the association among expressions of synaptophysin, chromogranin A, and Ki-67, clinicopathological variables, and outcome.

Results: Chromogranin A expression was found more commonly in small-cell NECs (9/9, 100%) than in large-cell NECs (27/53, 51%) (p = 0.008). No statistical significance was found in Ki-67 (p = 0.494) or synaptophysin (p > 0.1) expression between NEC cell types. Correlation analyses revealed that Ki-67 expression was significantly associated with mid-third disease of stomach (p = 0.005) and vascular involvement (p = 0.006), and had a trend of significant correlation with tumour relapse (p = 0.078). High expression of chromogranin A was significantly associated with histology of small-cell NECs (p = 0.008) and lesser tumour greatest dimension (p = 0.038). The prognostic significance was determined by means of Kaplan-Meier survival estimates and log-rank tests, and as a result, early TNM staging and postoperative chemotherapy were found to be correlated with longer overall survival (p < 0.05). Univariate analysis revealed associations between poor prognosis in NECs and several factors, including high TNM staging (p = 0.048), vascular involvement (p = 0.023), relapse (p = 0.004), and microscopic/macrosopic residual tumour (R1/2, p < 0.001). In a multivariate analysis, relapse was identified as the sole independent prognostic factor.

Conclusions: No significant correlation between survival and expression of synaptophysin, chromogranin A, or Ki-67 has been determined in G-NECs. Our study indicated that early diagnosis, no-residual-tumour resection, and postoperative chemotherapy were possible prognostic factors.

Key words: neuroendocrine carcinoma, stomach, prognosis, Ki-67, chromogranin A.

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Histological characterisation and prognostic evaluation of 62 gastric neuroendocrine carcinomas

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Introduction

Neuroendocrine carcinomas (NEC) is defined as a high-grade neuroendocrine neoplasm and regarded as a distinct and independent category because of its particularly aggressive biological behaviour and poor prognosis, compared with other neuroendocrine neoplasms such as neuroendocrine tumour/carcinoid [1–5].

Extra-pulmonary NECs are most often found in the gastrointestinal (GI) tract, which accounts for 35–55% of all extra-pulmonary NECs [6]. GI-NECs are mainly located in the oesophagus, stomach, pancreas, and colon. Among which, gastric NEC (G-NEC) is a rare but highly malignant tumour [7, 8], and it has a poorer outcome in comparison to conventional adenocarcinomas [9–11]. However, because of its rarity, limited study had been directed towards its clinicopathological characteristics and prognosis, and the optimal clinical management strategies have not yet been well established.

G-NECs encompass two histopathological entities: small-cell NECs (SC-NECs) and large-cell NECs (LC-NECs). It has been reported that most NECs arising in the glandular mucosa of the gastrointestinal tract are large-cell type, whereas those arising in the squamous mucosa are mostly small-cell type [12], and thus the majority of the NECs are LC-NECs. Both cell types have a high proliferation rate with a Ki-67 labelling index > 20% by definition, but usually much higher (> 75%) [13]. They are also characterised by some specific molecular markers of neuroendocrine differentiation. Numerous studies have supported the validity of the Ki-67 labelling index as a prognostic indicator for GI-NETs, as well as its potential role in planning therapy [14–16]. In addition, synaptophysin (Syn) is usually positive, while chromogranin A (CgA) is less frequently present. Positive CgA staining usually indicates a more mature tumour, and the presence of both Syn and CgA is considered a good prognostic sign [17–19].

In order to identify and better define the useful prognostic markers for G-NECs, in the present study, we examined the morphologic, immunohistochemical, as well as clinicopathological factors in 62 cases of G-NEC that met the definition according to the updated World Health Organization (WHO) classification [20].

Material and methods

Patients and tissue preparation

Sixty-two G-NEC patients were identified among 8052 cases of gastric carcinoma who underwent radical resection of primary tumour (total or...
subtotal gastrectomy) and systematic lymphadenectomy at The First Affiliated Hospital of Fujian Medical University (Fuzhou, Fujian Province, P.R.C.) between January 2007 and February 2013. The patients’ clinicopathological data are shown in detail in Table 1. None of the patients had received chemo- or radiotherapy before tissue collection. The histopathological features of the tumour specimens were classified in accordance with the WHO criteria [20, 21]. The TNM staging was determined according to the 7th Edition of the AJCC guidelines (2010) for NEC. The study protocol was approved by the Human Ethics Review Committee of Fujian Medical University, and signed, informed consent was obtained from each patient. Each specimen was fixed overnight in 10% buffered formalin at room temperature and embedded in paraffin. Serial sections were cut at a thickness of 4 µm and placed onto 3-aminopropyltriethoxysilane-coated glass slides. Some sections were stained with haematoxylin and eosin in a routine manner for histological examination.

Immunohistochemistry for synaptophysin, chromogranin A, and Ki-67

Immunohistochemistry was performed with the indirect enzyme-labelled antibody method, as described previously [22, 23]. The antibodies used in IHC are demonstrated in Table 2. In brief, for detection of Ki-67, synaptophysin, and chromogranin A, paraffin-embedded sections were deparaffinised with toluene and rehydrated in graded alcohols. After being autoclaved for 15 minutes at 120°C in 10 mM citrate buffer (pH 6.0) for antigen retrieval, endogenous peroxidase was inactivated with 0.3% hydrogen peroxide in methanol for 15 minutes. The sections were then pre-incubated with 500 µg/ml normal goat IgG dissolved in 1% BSA in PBS (pH 7.4) for one hour, reacted with primary antibodies for 16 hours, washed with 0.075% Brij 35 in PBS, and then incubated with HRP-conjugated goat anti-rabbit IgG (CgA/Syn) or HRP-conjugated goat anti-mouse IgG (Ki-67) in 1% BSA in PBS for one hour. After washing with 0.075% Brij 35 in PBS, the sites of HRP were visualised with DAB and H2O2. Finally, the nuclei were counterstained with haematoxylin. As a negative control, some sections were reacted with normal rabbit IgG or normal mouse IgG instead of the specific antibodies.

Quantitative evaluation

Staining results were examined by two observers unaware of patients’ clinical information. Another reading by a third observer was needed to reach a consensus when there was a significant discrepancy between initial results. At least five high-power fields and more than 2000 cells were calculated in each case with a light microscope (Zeiss 2021-85; Carl Zeiss Inc, Germany) at 400× magnification. Immunostaining results were evaluated by calculating the percentage of positive staining cells.

Statistical analysis

The SPSS 19.0 statistical software package (SPSS Inc., Chicago, IL, USA) was employed for all analyses. The association between tested markers and different clinicopathological parameters was evaluated using the chi-square test for categorical variables, and the Student’s t-test for continuous variables. A p-value of <0.05 was considered statistically significant.

### Table 1. Clinicopathological parameters

| Parameters                              | No. of cases (%) |
|-----------------------------------------|------------------|
| Median age, years                       | 63.4             |
| Age (y.o.)                              |                  |
| < 63                                    | 30 (48.4)        |
| ≥ 63                                    | 32 (51.6)        |
| Gender                                  |                  |
| Male                                    | 48 (77.4)        |
| Female                                  | 14 (22.6)        |
| Location of disease                     |                  |
| Upper third                             | 31 (50)          |
| Middle third                            | 20 (32.3)        |
| Lower third                             | 11 (17.7)        |
| Greatest dimension                      |                  |
| < 5 cm                                  | 38 (61.3)        |
| ≥ 5 cm                                  | 24 (38.7)        |
| TNM stage                               |                  |
| I                                       | 0                |
| II                                      | 6 (9.7)          |
| III                                     | 48 (77.4)        |
| IV                                      | 8 (12.9)         |
| Histology                               |                  |
| Large cell                              | 53 (85.5)        |
| Small cell                              | 9 (14.5)         |
| Depth of invasion                       |                  |
| T1                                      | 1 (1.6)          |
| T2                                      | 6 (9.7)          |
| T3                                      | 26 (41.9)        |
| T4                                      | 29 (46.8)        |
| LV factor                               |                  |
| Positive                                | 49 (79.0)        |
| Negative                                | 13 (21.0)        |
| V-factor                                |                  |
| Positive                                | 37 (59.7)        |
| Negative                                | 25 (40.3)        |
| LN metastasis                           |                  |
| Positive                                | 49 (79.0)        |
| Negative                                | 13 (21.0)        |
| Distant metastasis                      |                  |
| Positive                                | 24 (38.7)        |
| Negative                                | 38 (61.3)        |
| Postoperative chemotherapy              |                  |
| Yes                                     | 42 (67.7)        |
| No                                      | 20 (32.3)        |
| Relapse                                 |                  |
| Yes                                     | 32 (51.6)        |
| No                                      | 30 (48.4)        |
| Residual tumour classification          |                  |
| R0                                      | 53 (85.5)        |
| R1                                      | 4 (6.4)          |
| R2                                      | 5 (8.1)          |
| Ki-67 expression                        |                  |
| High                                    | 28 (45.2)        |
| Low                                     | 34 (54.8)        |
| Chromogranin A expression               |                  |
| Positive                                | 36 (58.1)        |
| Negative                                | 26 (41.9)        |
| Median follow-up (months)               | 19.3             |

Clinical data were recorded in accordance with the AJCC TNM staging system for carcinoma of the stomach; LV-factor/V-factor – the status of tumour invasion into lymphatic vessels (LV)/vascular invasion (V); LN – lymph node; R0 – no residual tumour/R1 – microscopic residual tumour/R2 – macroscopic residual tumour.
adjuvant chemotherapy and 20 did not. The average number in four cases. Forty-two patients received postoperative chemotherapy. Three patients were suspected, including liver (n = 5) and transverse colon (n = 3). Nonetheless, macroscopic tumour residual was observed only in one patient, invasion into lamina muscularis propria occurred in six patients, invasion into subserosal connective tissue without involving visceral peritoneum or adjacent structures was observed in 26 cases, and invasion into serosa or adjacent structures occurred in 29 patients. There were 24 cases with a greatest dimension of 5 cm or more, while 38 cases had less than 5 cm. As for TNM staging, six cases were stage II, 48 cases stage III, and 8 cases were stage IV. Forty-nine patients were positive with distant metastasis while 13 were absent. Twenty-four patients were found positive with LN metastasis while 13 were absent. Twenty-four patients relapsed after surgery, while no sign of relapse had been found at the end-point of follow-up for the other 30 cases. Fifty-three cases were large-cell NECs and nine were small-cell NECs. Postoperative follow-up data were available for all 62 cases, and the median follow-up duration was 19.3 months, ranging from 1.0 to 54.0 months.

**Results**

**Patients’ clinicopathological data**

As shown in Table 1, the patient population included 48 males and 14 females, and the male-to-female ratio was approximately 3:1. The mean age of the patients was 63.4 years, ranging from 47 to 82 years. The number of upper, middle, and lower third disease were 31, 20, and 11, respectively. Invasion into mucosa/submucosa was observed only in one patient, invasion into lamina muscularis propria occurred in six patients, invasion into subserosal connective tissue without involving visceral peritoneum or adjacent structures was observed in 26 cases, and invasion into serosa or adjacent structures occurred in 29 patients. There were 24 cases with a greatest dimension of 5 cm or more, while 38 cases had less than 5 cm. As for TNM staging, six cases were stage II, 48 cases stage III, and 8 cases were stage IV. Forty-nine patients were present with LN metastasis while 13 were absent. Twenty-four patients were found positive with distant metastasis while 38 negative. All patients underwent surgical resection of the stomach with regional lymphadenectomy, and 53 cases underwent curative resection, i.e. R0 resection, while 8 cases were R1 resection, and the other five were R2 resection. In detail, 11 underwent distal gastrectomy and 51 total gastrectomy. Combined resection of spleen and tail of pancreas was performed in eight patients with advanced tumours in the upper third stomach. Resection was aggressively extended to adjacent organs when tumour invasion was suspected, including liver (n = 5) and transverse colon (n = 3). Nonetheless, macroscopic tumour residual was found in five patients and microscopic tumour residual in four cases. Forty-two patients received postoperative adjuvant chemotherapy and 20 did not. The average number of adjuvant chemotherapy cycle was four. Regimens of adjuvant chemotherapy were as below: fluorouracil, leucovorin plus oxaliplatin (FOLFOX4) for 13 patients, fluorouracil plus oxaliplatin for 12 patients, paclitaxel plus oxaliplatin for four patients, docetaxel plus fluorouracil for four patients, dacarbazine, epirubicin plus fluorouracil for two patients, and other regimens for the other seven patients. Thirty-two patients relapsed after surgery, while no sign of relapse had been found at the end-point of follow-up for the other 30 cases. Fifty-three cases were large-cell NECs and nine were small-cell NECs. Postoperative follow-up data were available for all 62 cases, and the median follow-up duration was 19.3 months, ranging from 1.0 to 54.0 months.

**Histological findings and immunohistochemical staining for synaptophysin, chromogranin A, and Ki-67**

The tumors were sub-classified into large-cell and small-cell NECs, as defined by the WHO classification and previous reports [4, 24]. LC-NECs manifest as tumour cells exhibiting round to markedly irregular nuclei with coarse chromatin and prominent nucleoli and moderate amounts of cytoplasm (Fig. 1A). Peripheral palisading and/or rosette formation were present in the majority of LC-NECs, which also exhibited hyperchromatic nuclei with finely granular chromatin and eosiophilic granular cytoplasm. SC-NECs, however, showed a markedly higher nuclear/cytoplasmic ratio, exhibiting hyperchromatic nuclei with finely granular chromatin and scant cytoplasm (Fig. 1B). Other factors like presence of necrosis, mitotic counts (count per 10 HPF using a Zeiss 2021-85 microscope), lymphatic and vascular involvement, association with non-NEC components, and the statuses of the surrounding mucosa, including the presence of chronic gastritis and intestinal metaplasia, were also evaluated.

Immunohistochemically, Ki-67 was localised predominantly in the nuclei, while both chromogranin A and synaptophysin in the cytoplasm (Fig. 1G-H). Here we used 56% as the cut-off point to determine high and low Ki-67 expression. The number of patients positive for synaptophysin staining was 61 (98.4%), whereas the numbers for positive chromogranin A staining and high Ki-67 expression was 26 (41.9%) and 28 (45.2%), respectively. For different histologies, 25 of 53 LC-NECs were found to have high expression of Ki-67 while in SC-NECs the number was three (p = 0.494). As for chromogranin A expression, 27 of 53 large-cell and all nine small-cell NECs were positive.

### Table 2. List of antibodies used in immunohistochemistry

| Antibody                          | Working dilution/concentration | Manufacturer                               |
|-----------------------------------|--------------------------------|--------------------------------------------|
| Monoclonal, mouse anti-human Ki-67 (Clone: MIB-1) | 1 : 400                          | DakoCytomation, Glostrup, Denmark           |
| Monoclonal, rabbit anti-human Chromogranin A (Clone: SP12) | 1 : 100                          | Thermo Fisher Scientific, Fremont, CA, USA  |
| Monoclonal, rabbit anti-human Synaptophysin (Clone: EP158) | 1 : 200                          | Epitomics, Inc. Burlington, CA, USA         |
| Normal goat IgG                  | 1 : 20                          | Sigma Chemical Co., MO, USA                |
| HRP-conjugated goat anti-rabbit/mouse IgG | 1 : 200                          | DakoCytomation, Glostrup, Denmark           |
Fig. 1. Histologic and immunohistochemical features of LC-NEC and SC-NEC. A) HE staining for large-cell NEC with a trabecular growth pattern. Of note, the tumour cells have round and hyperchromatic nuclei and eosinophilic and granular cytoplasm (white arrowheads), showing nuclear moulding and suggesting neuroendocrine differentiation; B) HE staining for small-cell NEC, showing a markedly higher nuclear/cytoplasmic ratio and hyperchromatic nuclei with finely granular chromatin and scant cytoplasm; C) LC-NEC, positive staining for Ki-67; D) SC-NEC, positive staining for Ki-67; E) LC-NEC, positive staining for chromogranin A; F) SC-NEC, positive staining for chromogranin A; G) LC-NEC, positive staining for synaptophysin; H) SC-NEC, positive staining for synaptophysin. Scale bar represents 20 µm
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No statistical significance was found for synaptophysin expression within two NECs ($p > 0.1$).

Correlation of Ki-67 and chromogranin A expression with clinicopathological parameters

To determine the correlation of Ki-67 and chromogranin A expression and clinicopathological parameters, and to determine its prognostic impact, chi-square together with univariate, multivariate, and Kaplan-Meier survival analyses were performed. As shown in Table 3, high Ki-67 expression was significantly associated with the mid-third disease ($p = 0.005$), vascular involvement ($p = 0.006$), and had a trend of significant correlation with tumour relapse ($p = 0.078$). High expression of chromogranin A was significantly associated with small-cell histology ($p = 0.008$) and lesser greatest dimension of tumour ($\leq 5$ cm, $p = 0.038$). However, Kaplan-Meier survival analyses revealed no difference of disease-free survival or OS in patients with different levels of Ki-67 or chromogranin A expression ($p > 0.05$). Univariate analysis demonstrated associations between poor prognosis and several factors, including high TNM staging ($p = 0.048$), vascular involvement ($p = 0.023$), relapse ($p = 0.004$), and microscopic/macroscopic residual tumour (R1/2, $p < 0.001$). Using a Cox proportional

Table 3. Association of Ki-67 and chromogranin A with clinicopathological parameters in G-NEC patients

| Variables                  | Ki-67       | CgA        |
|---------------------------|-------------|------------|
| Agea (y.o.)               |             |            |
| $\geq 63$                  | 32          | 32         | 0.459  | 0.829 |
| $< 63$                    | 30          | 30         | 0.420  | 0.917 |
| Gender                    |             |            |
| Male                      | 48          | 48         | 0.005  | 0.876 |
| Female                    | 14          | 14         | 0.0005 |            |
| Location                  |             |            |
| Upper                     | 31          | 31         | 0.005  |            |
| Middle                    | 20          | 20         | 0.0005 |            |
| Lower                     | 11          | 11         | 0.0005 |            |
| TNM stageb                |             |            |
| I/II                      | 6           | 6          | 0.681  | 1.000 |
| III/IV                    | 56          | 56         | 0.681  | 1.000 |
| Histologyb                |             |            |
| Small-cell                | 9           | 9          | 0.494  | 0.008 |
| Large-cell                | 53          | 53         | 0.494  |            |
| Depth of invasionb        |             |            |
| T1-2                      | 7           | 7          | 0.442  | 0.119 |
| T3-4                      | 55          | 55         | 0.442  |            |
| LV-factor                 |             |            |
| Yes                       | 49          | 49         | 0.585  | 0.775 |
| No                        | 13          | 13         | 0.585  |            |
| V-factor                  |             |            |
| Yes                       | 37          | 37         | 0.006  | 0.426 |
| No                        | 25          | 25         | 0.006  |            |
| LN metastasis             |             |            |
| Yes                       | 49          | 49         | 0.585  | 0.775 |
| No                        | 13          | 13         | 0.585  |            |
| Distant metastasis        |             |            |
| Yes                       | 24          | 24         | 0.660  | 0.574 |
| No                        | 38          | 38         | 0.660  |            |
| Greatest dimension        |             |            |
| $\geq 5$ cm               | 24          | 24         | 0.257  | 0.038 |
| $< 5$ cm                  | 38          | 38         | 0.257  |            |
| Relapse                   |             |            |
| Yes                       | 32          | 32         | 0.078  | 0.829 |
| No                        | 30          | 30         | 0.078  |            |
| Ki-67                     |             |            |
| High                      | –           | –          | –      | 0.243 |
| Low                       | –           | –          | –      |            |

LV-factor – lymphatic vessel involvement; V-factor – vascular involvement; LN metastasis – lymph node metastasis

$^a$Mean age

$^b$Fisher exact tests (2-sided); $\chi^2$ test for all the other analyses.
Univariate and multivariate analyses of factors associated with OS.

| Variables                        | Hazard Ratio (95% CI) | P-value |
|----------------------------------|----------------------|---------|
| **Univariate analysis**          |                      |         |
| Age (y.o.) (≥ 63 vs. < 63)       | 1.519 (0.735–3.140)  | 0.260   |
| Gender (male vs. female)         | 1.688 (0.645–4.419)  | 0.286   |
| Location (upper/middle/lower)    | 1.139 (0.730–1.778)  | 0.566   |
| Greatest dimension (cm)          | 0.594 (0.290–1.219)  | 0.156   |
| TNM stage (I/II vs. III/IV)      | 7.587 (1.019–56.464) | 0.048   |
| Histology (small cell vs. large cell) | 0.700 (0.243–2.020) | 0.510   |
| Depth of invasion (T1-2 vs. T3-4) | 1.028 (0.357–2.957) | 0.959   |
| LV-factor (yes vs. no)           | 1.894 (0.754–4.757)  | 0.174   |
| V-factor (yes vs. no)            | 2.508 (1.137–5.533)  | 0.023   |
| LN metastasis (yes vs. no)       | 1.894 (0.754–4.757)  | 0.174   |
| Distant metastasis (yes vs. no)  | 1.654 (0.807–3.390)  | 0.169   |
| Relapse (yes vs. no)             | 77.455 (4.036–1486.380) | 0.004 |
| Residual tumour classification (R0 vs. R1/2) | 5.911 (2.544–13.735) | 0.000 |
| Ki-67 expression (high vs. low)  | 0.635 (0.297–1.359)  | 0.242   |
| Chromogranin A (positive vs. negative) | 0.898 (0.435–1.851) | 0.770   |

**Multivariate analysis**

| Variables                        | Hazard Ratio (95% CI) | P-value |
|----------------------------------|----------------------|---------|
| Relapse (yes vs. no)             | 77.455 (4.036–1486.380) | 0.004 |

Discussion

In the present study, we investigated the expressions of Ki-67, chromogranin A, and synaptophysin in G-NECs immunohistochemically, analysed the associations between the tested markers and clinicopathological parameters, and tried to determine the prognostic factors that would probably affect the survival.

First of all, we found that high expression of Ki-67 was significantly associated with middle third disease of the stomach (p = 0.005) and vascular infiltration (p = 0.006), and had a trend of significant correlation with tumour relapse (p = 0.078). Ki-67 was reported to be overexpressed at both mRNA and protein levels in many cancerous tissues [25–28], and correlated with invasiveness, increased proliferation, and poor outcome. Sorbye et al. [29] reported that patients with a Ki-67 less than 55% were less responsive to platinum-based chemotherapy, but had a longer survival. Boo et al. [13] demonstrated that a high Ki-67 PI (> 60%) was correlated with tumour recurrence and worse histological differentiation. Boo et al. [13] used 60% without any specific explanation, while Sorbye et al. [29] used 55% as the cutoff point determined by receiver operating characteristic (ROC) analysis, which sounded more reasonable. In the present study, we used 56% as the cut-off point, and found that the high Ki-67 PI was closely related to middle third disease of the stomach (p = 0.005) and vascular infiltration (p = 0.006), and had a trend of significant correlation with tumour relapse (p = 0.078), which was consistent with the findings of Rindi et al. [30]. To our knowledge, our data demonstrated for the first time a direct association of expression of Ki-67 with vascular invasion and location of disease of G-NECs. Although in our study neither univariate (p = 0.242) nor multivariate analysis revealed high Ki-67 PI as a poor prognostic factor in G-NECs, the result still suggested that high proliferation index contributed to worse prognosis because both angiogenesis and relapse were confirmatively affecting the outcome of G-NEC patients.

Chromogranin A (CgA) is localised in neurosecretory granules, and its expression thus reflects the degree of neuroendocrine differentiation. According to Bakkeldu’s findings [31], signet ring cells in gastric carcinomas were derived from neuroendocrine cells. Fujiyoshi et al. [17] demonstrated that a significant positive correlation existed between CgA positivity and favourable prognosis, with a tendency for greater positivity to be associated with better OS. In the current study, our finding indicated that high expression of chromogranin A was significantly associated with various clinicopathological parameters including small-cell histology of NECs (p = 0.008) and lesser tumour greatest dimension (p = 0.038), which were consistent with the findings of Pape et al. [18] and Welin et al. [19], while we failed to confirm the significant correlation with better prognosis (p = 0.770). Further study with a large scale of specimens would be needed.

Most significantly, we found that high TNM staging (p = 0.048), vascular involvement (p = 0.023), relapse (p = 0.004), and microscopic/macrosopic residual tumour (R1/2, p < 0.001) were closely associated with poor out-

Survival and its relation with clinicopathological factors

As shown in Figure 2B–F, median OS was 28 months. For histology of large- and small-cell NECs, their median survival was 29 and 28 months, respectively (p = 0.0384). Furthermore, for stage III the median survival was 26 months, and 11 months for stage IV (p = 0.0033). Owing to the limited number of stage II cases, its median survival could not be determined. In addition, for 42 patients given postoperative chemotherapy their median survival was 29 months, while it was 22 months for 20 patients without postoperative chemotherapy (p = 0.0303). For patients with different level of Ki-67 expression, the median survival was 26 and 30 months, respectively (p = 0.2287).

As shown in Table 4, univariate analysis revealed that TNM staging (I/II vs. III/IV; p = 0.048), angioinvasion (V-factor, yes vs. no; p = 0.023), tumour relapse (yes vs. no; p = 0.004), and residual tumour classification (R0 vs. R1/2; p < 0.001) were significantly correlated with survival (tumour depth, lymph node metastasis, and residual tumour classification/curability were classified according to the seventh edition of AJCC for G-NECs). Multivariate analysis indicated that tumour relapse (hazard ratio, 77.455; 95% confidence interval, 4.036–1486.380; p = 0.004) was the sole independent prognostic factor for survival.
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Fig. 2. Kaplan-Meier cumulative survival curves and log-rank test. A) Chart illustration of number of cases of immunohistochemical staining for synaptophysin, chromogranin A, and Ki-67; B) Overall survival; C) large-cell NECs versus small-cell NECs ($p = 0.4987$); D) Survival according to AJCC stages, 7th Edition ($p = 0.0033$); E) Presence and absence of postoperative adjuvant chemotherapy ($p = 0.0303$); F) high Ki-67 expression vs. low Ki-67 expression ($p = 0.2287$)

In general, TNM staging, vascular involvement, tumour relapse and residual tumour classification were considered the most important factors affecting prognosis in GI-NECs [11, 30], as was also confirmed in the present study. In patients with R1/2 resection, their hazard ratio of tumour recurrence was 4.911-fold higher than those with R0 resection (Table 4). Kubota et al. [32] reported maximum tumour diameter, depth of tumour invasion, LN metastasis, lymphatic invasion, p-Stage, and curability had significant correlations with survival. Recently, Ishida et al. [11] confirmed curative surgery as the only independent prognostic factor related to survival. Although our findings demonstrated that surgical curability is one of the prognostic factors of vital importance, which would inevitably affect the probability of relapse and long-term outcome, it is not confirmed as an independent one. How-
ever, our data suggested relapse as the only independent prognostic factor.

When further investigating the factors resulting in unfavourable outcome, we inferred that poor prognosis seemed to result from potent microvessel invasiveness and subsequent frequent further metastasis to lymph nodes or other organs even in the early stages of the disease [3], and a significant correlation between liver metastasis and vascular invasion has been well demonstrated in gastric cancers [33–35]. Chiba et al. [36] revealed in their retrospective review of 119 cases that G-NECs seemed significantly to have more frequent incidences of lymphatic invasion (88.9% vs. 56.6%), vascular invasion (75.6% vs. 31.6%), and LN metastases (82.1% vs. 58.8%) compared to those in ordinary gastric cancer. In our study, lymphatic invasion and LN metastasis were present in 49 cases to those in ordinary gastric cancer. In our study, lymphatic invasion and LN metastasis were present in 49 cases compared with 71 cases; however, our data suggested relapse as the only independent prognostic factor for survival (Table 4). Based on these facts, we suggest that every effort should be made to achieve an R0 resection for every surgical candidate, so as to obtain a better outcome and postpone the event of tumour relapse.

Our findings also suggested that there was obvious discrepancy with the OS between patients receiving postoperative chemotherapy or not. As shown in Figure 2E, patients who underwent postoperative chemotherapy had a significantly better outcome with a median overall survival of 29 months; much longer than those who did not, which was only 22 months. In our patient cohort, the common regimens used were fluorouracil, leucovorin plus oxaliplatin (FOLFOX4), fluorouracil plus oxaliplatin, paclitaxel plus oxaliplatin, and docetaxel plus fluorouracil. In addition, Okita et al. [37] reported that chemotherapy with cisplatin plus irinotecan was also effective for gastric poorly-differentiated NECs, whose response rate was 75%, median progression-free survival time was seven months, and median survival was 22.6 months.

Some limitations of this research should be noted. First of all, the number of the dataset was small, so the statistical power would be somewhat limited. Further study with a larger sample size would be necessary to validate the present results. Secondly, some of the follow-up time of the cases is somewhat short. Although other regimens like radiotherapy would be used after progression had been proven, the contribution of other therapies to overall survival had not been taken into account in the survival analysis because surgery plus chemotherapy was considered the key therapy for patients at this stage, and thus would probably lead to some bias.

In summary, G-NEC patients have a poor prognosis, and surgical candidates should be considered for no-residual resection and postoperative chemotherapy without delay. In addition, TNM staging, vascular involvement, and relapse were the most important negative prognostic factors for survival in this retrospective study. In contrast, despite the significant morphologic diversity of G-NECs, their histologic subclassification was not of prognostic significance.

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