Identification of *Candida* spp. in the oral cavity in patients with malignant diseases

Identifikacija vrsta gljivica iz roda *Candida* u usnoj duplji bolesnika sa malignim oboljenjem

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**Background/Aim.** Oral candidiasis frequently causes discomfort in patients treated for malignant diseases, acting as well as a potential source of systemic infection. This disease may present itself through different clinical manifestations of both acute or chronic type. The aim of this study was to identify different *Candida* species from oral cavities of patients suffering from malignant diseases. **Methods.** Thirty patients admitted to the hospital for diagnostics/treatment of malignant diseases were included in this investigation. All subjects had visible changes of oral mucosa in the form of pseudomembranes and inflammation corresponding to oral candidiasis. Control group included 30 non-hospitalized patients diagnosed with candidiasis. *Candida* species was confirmed in all patients by microbiological analysis of tongue swabs. For microbiota identification, three different tests were used: germination test, fungal growth test on corn meal agar, and biochemical identification with commercially available ID 32 C kit (bio-Merieux, Marcy-l’Etoile, France). **Results.** Out of 30 isolates collected from hospitalized patients, 90% was related to *Candida albicans*, 7% was identified as *Candida kefyr*, and 3% as *Candida famata*. In samples collected from non-hospitalized controls, we isolated *Candida albicans* in 90% of the cases, in 7% *Candida kefyr*, while in 3% we identified *Candida glabrata*. **Conclusion.** Based on this investigation, oral candidiasis in patients treated with radiotherapy and chemotherapy is mainly caused by *Candida albicans*. It is to be expected that *Candida albicans* will remain the most significant causative agent of oral candidiasis, although we must bear in mind the possibility of other pathogenic species.

**Key words:** candida; *candida albicans*; mouth; neoplasms; microbiological techniques; candidiasis, oral.

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Introduction

Candida species (Candida spp.) are constitutive members of oral flora. Nearly half of the healthy adult population has these fungi on the mucosal surface without experiencing any symptoms since the immune system controls its excessive growth and development of the disease. However, in cases of some local and systemic predisposing factors, there is a possibility of oral candidiasis development. The local predisposing factors are changes in saliva gland function, use of antibiotics and corticosteroid drugs, carbohydrate-rich diet, changes in oral epithelium, dentures, or excessive tobacco use. Systemic predisposing factors include changes in hormonal status, iron, folic acid and vitamin B12 deficiencies, use of antibiotics, malignant diseases and immune suppression of different origins. Oral candidiasis may be manifested through different clinical signs and symptoms, as acute pseudomembranous, acute atrophic candidiasis, chronic atrophic candidiasis, glossitis and angular cheilitis.

Oral candidiasis is the most common opportunistic infection in patients with malignant diseases. Cytoxic therapy and radiotherapy are both important predisposing factors in its development. Patient's defense mechanisms which have already been weakened by the main disease may be further depleted by cytotoxic and radiation therapy. As a consequence of cytotoxic therapy, oral candidiasis occurs in 30–70% of the patients. Side effects of head and neck radiotherapies, such as dry mouth and thick sticky saliva, are coupled with an increased colonization of oral mucosa with Candida, especially Candida albicans (C. albicans). Such milieu favors the development of oral candidiasis in 17% to 52.5% of the patients. Fungi belonging to Candida spp. are responsible for 75% of all fungal infections in patients with malignancies. According to the literature, C. albicans is isolated in 70–80% of all patients, while Candida glabrata (C. glabrata) and Candida tropicalis (C. tropicalis) appear in 5–8% of the cases. Recent research shows an increase in trend of non-albicans candidal infections (C. glabrata, C. parapsilosis and C. tropicalis), while the share of C. albicans decreased.

Development of oral candidiasis in patients treated with radiotherapy and chemotherapy presents a significant factor for the systemic infection and candidaemia which may act as a direct cause of death. The European Organization for Research and Treatment of Cancer – International Antimicrobial Therapy Group published the data of oral cavity being a direct cause of death. The control group was made by 30 patients from the Clinic of Dental Medicine of the same hospital center. All of them had a fully developed scope of clinical signs and symptoms corresponding to oral candidiasis and did not suffer from any malignancy.

Methods

This research included thirty patients hospitalized at the Clinic of the Internal Medicine and Clinic of Radiology and Oncology, Clinical Hospital Center Rijeka. Table 1 presents demographic and clinical characteristics of hospitalized patients. At the time of sampling, 19 of them were subjected to cytostatic therapy, while 11 of them went through radiotherapy. All subjects had visible changes in the oral mucosa with pseudomembranes whose clinical appearance corresponded to oral candidiasis. All subjects were microbiologically tested in order to confirm the diagnosis. Patients did not use any antifungal drugs for at least one month before sampling procedures.

The control group was made by 30 patients from the Clinic of Dental Medicine of the same hospital center. All of them had a fully developed scope of clinical signs and symptoms corresponding to oral candidiasis and did not suffer from any malignancy.

Both groups were introduced to the purpose of the research upon the inclusion and by signing the Informed consent agreement to participate. The research had been approved by the Ethics Committee of the Clinical Hospital Center Rijeka.

Cultivation and identification of Candida

Subjects were taken the oral mucosa swabs from the tongue region with a sterile cotton swab (Copan, Zagreb, Croatia). Immediately upon collection, the material was struck out on solid Sabouraud dextrose agar plates (PanreacQuimica, Cultimed, Spain) and incubated at 37°C for 72 h. Distinctive colonies sized 2–3 mm, with smooth and shiny surface and clean margins, white to cream in color, and with typical yeast smell, were transferred and multiplied on new solid medium plates. Positive strains were identified by standard mycological methods, by germ tube production, chlamydospore development in the microculture in cornmeal Tween 80 (Difco, Detroit, USA) and API ID 32 Candida identification kit (bio-Merieux, Marcy-l’Etoile, France). The germ-tube test involved the induction of hyphal outgrowths from yeast cultured in rabbit serum for 3 h at 37°C. Microscopic slides were examined under light microscope. This test was used for C. albicans identification. Chlamydospore production was also associated with C. albicans. C. albicans produced thick-walled, dormant growth forms induced in vitro by culture agar supplemented with Tween 80. The inoculated area was covered with the cover slip and the agar incubated at 22°C for 72 h. Under such conditions, cornmeal agar also included a characteristic filamentous growth which could aid in the identification of C. albicans. The API ID 32 C system consists of a single use disposable plastic strip with 32 wells containing substrates for 29 assimilation tests (carbohydrates, organic acids, and amino acids), one susceptibility test (cytoheximide), one colorimetric test (esculin), and one negative control. The yeast identification procedures were conducted in accordance with the manufacturer's instructions. One day-cultures and sterile distilled water were used to prepare the suspensions with final turbidity equivalent to McFarland #2. Five drops of this suspension were then dispensed to ampoules of C medium provided by the manufacturer and homogenized to prepare an even dispersion of inoculum. The inoculum suspensions were used to inoculate the wells. The systems were incubated at 30°C for 48 h. The results were visually examined and transformed into numerical bio-codes. At the end, the isolates were identified by ID 32 Analytical Profile Index.
Table 1

| Hospitalized patients | Gender | Age (years) | Type of malignancy | Type of therapy |
|-----------------------|--------|-------------|-------------------|----------------|
|                       |        |             |                   | surgery | chemotherapy | radiotherapy |
| 1                     | m      | 73          | Non-Hodgkin lymphoma | –       | +            | –            |
| 2                     | f      | 57          | Leukemia          | –       | +            | –            |
| 3                     | m      | 71          | Colon cancer      | +       | +            | –            |
| 4                     | f      | 51          | Ovarian cancer    | +       | +            | –            |
| 5                     | m      | 73          | Oral cancer       | +       | –            | +            |
| 6                     | f      | 55          | Breast cancer     | +       | –            | +            |
| 7                     | m      | 51          | Colon cancer      | +       | +            | –            |
| 8                     | m      | 71          | Prostate cancer   | +       | +            | –            |
| 9                     | f      | 69          | Thyroid cancer    | –       | –            | +            |
| 10                    | f      | 31          | Leukemia          | –       | +            | –            |
| 11                    | m      | 52          | Colon cancer      | +       | +            | –            |
| 12                    | m      | 64          | Oral cancer       | +       | –            | +            |
| 13                    | f      | 51          | Breast cancer     | +       | –            | +            |
| 14                    | f      | 49          | Breast cancer     | +       | –            | +            |
| 15                    | f      | 66          | Thyroid cancer    | +       | –            | +            |
| 16                    | f      | 65          | Uterine cancer    | +       | +            | –            |
| 17                    | f      | 71          | Oral cancer       | +       | –            | +            |
| 18                    | m      | 66          | Prostate cancer   | +       | +            | –            |
| 19                    | f      | 78          | Breast cancer     | +       | –            | +            |
| 20                    | f      | 59          | Leukemia          | –       | +            | –            |
| 21                    | f      | 64          | Breast cancer     | +       | –            | +            |
| 22                    | f      | 69          | Colon cancer      | +       | +            | –            |
| 23                    | f      | 49          | Non-Hodgkin lymphoma | –       | +            | –            |
| 24                    | m      | 70          | Oral cancer       | +       | –            | +            |
| 25                    | f      | 72          | Breast cancer     | +       | +            | –            |
| 26                    | m      | 58          | Gastric cancer    | +       | +            | –            |
| 27                    | f      | 65          | Leukemia          | –       | +            | –            |
| 28                    | f      | 61          | Breast cancer     | +       | –            | +            |
| 29                    | f      | 54          | Colon cancer      | +       | +            | –            |
| 30                    | m      | 64          | Prostate cancer   | +       | +            | –            |

m – male; f – female.

Statistical analysis

Statistical analysis was performed using the Statistica 12.7 software (StatSoft, Inc., Tulsa, OK, USA).

The Kolmogorov-Smirnov normality test was applied to our data. The Student-\( t \)-test was used to analyze the age difference between groups while the \( \chi^2 \) test was used to compare the genders. Fisher’s exact test was used to analyze oral candida distribution in hospitalized and non-hospitalized group of patients.

Statistically significant difference was set to \( p < 0.05 \).

Results

The hospitalized group of patients suffering from oral candidiasis \((n = 30)\) included 17 women and 13 men. Chemotherapeutic protocol for treatment of leukemia was assigned for 4 patients, 2 for treatment of lymphoma, 3 for breast cancer, 6 for malignancies of digestive organs, 3 for prostate cancer and 1 for ovarian cancer. A total of 4 subjects was treated with irradiation therapy for breast cancer, 4 for oral cancer, 2 for thyroid cancer, and 1 for uterine cancer. All patients had pronounced inflammatory changes on the oral mucosa with pseudomembranes. Microbiological analysis proved oral candidiasis in all patients. Average patient age was \( 61.23 \pm 9.07 \) years (Table 2).

The control group of patients with oral candidiasis \((n = 30)\) included 11 males and 19 females. Denture stomatitis was diagnosed in 14 patients, 13 had acute atrophic candidiasis, and 3 patients developed acute pseudomembranous candidiasis. Clinical diagnosis of oral candidiasis was confirmed in all patients through microbiological analysis. Average patient age in this group was similar to the study group (Table 2). The distribution of different species in both groups is presented in table 3. Of 30 isolates, collected from 30 patients, \( C. \) albicans was detected in 27 (90%), two (7%) isolates...
collect from non-hospitalized patients, the main isolate was Candida famata (C. famata).
When analyzing 30 isolates were identified as Candida kefyr (C. kefyr), and one (3%) as Candida famata (C. famata). When analyzing 30 isolates collected from non-hospitalized patients, the main isolate was C. albicans in 27 (90%) cases, in two (7%) samples C. kefyr, and in one (3%) C. glabrata.

Discussion

Fungal infections pose a significant source of complications in patients suffering from malignancies. During the last 50 years their numbers increased, they frequently develop in early stages of the disease, and are caused by the species which had not been previously considered as pathogenic. Cytotoxic and radiation therapies are important predisposing factors in the development of oral candidiasis. While the organism had already been weakened by the principal disease, its defense mechanisms are further depleted by those treatment modalities. In addition, a consequence of these therapies is frequently oral mucositis, which is mainly caused by their direct cytotoxic effect on mucosal cells as well as a negative effect of long-standing inflammation due to inadequate immune reaction to fungal infection. It is estimated that 30–70% of the patients on cytotoxic therapy develop oral candidiasis. Moreover, negative side-effects of radiation therapy in the head and neck regions, such as dry mouth and thick sticky saliva, are coupled with an increase in colonization with Candida spp, especially C. albicans. Such a milieu favors the development of oral candidiasis in 17% to 52.5% of the patients. In all our test subjects, clinical examination and microbiological analysis proved oral candidiasis.

Most superficial fungal infections of the oropharyngeal region and digestive system are caused by C. albicans. In immunologically challenged patients, this fungus may cause an invasive infection through damage and ulcerations of the mucosal surfaces. Disseminated type of the infection may appear in cases of neutropenia, hematologic malignancies, and in patients on high dose-regimens of antimicrobial and cytotoxic therapy. In this research, C. albicans was the most common cause of the infection, and these results correspond to the majority of other investigations. Nicolatou-Galitis et al. investigated patients in irradiation therapy; a total of 61 patients participated, and pseudomembranous candidiasis developed in 31 patients. The most commonly isolated fungus was C. albicans (84%), followed by C. tropicalis (9%), C. glabrata (3.4%), Candida krusei (1.2%) and Candida holmii (1.2%). Similar results were obtained by Swoboda-Kopec et al. The main causative agent was C. albicans, while of non-albicans Candida significant role in infection was played by C. glabrata, C. krusei, C. tropicalis, C. parapsilosis and C. kefyr. The cases where C. albicans is isolated as the main causative agent range from 28.1% to 94%.

These results are in line with our results since in our patients 90% of the infections were caused by C. albicans.

To a lesser extent (6%) we were able to isolate C. kefyr in our patients. It is well known that C. kefyr may cause superficial infections. Besides, some cases of fungaemia caused by this fungus have been described in hospitalized patients both from departments of surgery and oncology. C. famata is a saprophyte which rarely causes infections in humans, however, there are case reports of fungaemia and peritonitis in hospitalized patients. In addition, it may cause urogenital infections and deep fungal infections of heart or lung tissue with the subsequent development of sepsis. In our investigation, one of the subjects was positive on Candida glabrata infection.

C. glabrata had been considered as a harmless member of the oral flora; however, it has recently been identified as a significant causative agent of infections in immunologically compromised patients. In patients submitted to irradiation therapy, C. glabrata is regarded as an important cause of oropharyngeal candidiasis. The probability of C. glabrata-caused infection of the oral mucosa is significantly increased if the previous fungal infection was treated with fluconazole or ketoconazole. In hospitalized patients who participated in this research, no C. glabrata isolate has been detected, but we have isolated this fungus from the one of the control patients’ oral mucosa.

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Table 2

Clinical characteristics of hospitalized and non-hospitalized group of patients

| Parameter                           | Hospitalized | Non-hospitalized | p-value |
|-------------------------------------|--------------|-----------------|---------|
| Total number of patients            | 30           | 30              |         |
| Gender (m/f), n                     | 13/17        | 11/19           | 0.60*   |
| Age (years), \( \bar{x} \pm SD \)   | 61.2 ± 9.0   | 58.7 ± 11.6     | 0.05**  |

m – male; f – female; \( \bar{x} \) – mean value; SD – standard deviation.

*Student-t test; **Fisher’s exact test.

Table 3

Oral candida distribution in hospitalized and non-hospitalized group of patients

| Candida spp.   | Hospitalized patients (n = 30) | Non-hospitalized patients (n = 30) | p-value |
|----------------|---------------------------------|------------------------------------|---------|
| Candida albicans | 27 (90)                         | 27 (90)                            |         |
| Candida kefyr*  | 2 (7)                           | 2 (7)                              |         |
| Candida famata* | 1 (3)                           | 1 (3)                              | 1**     |
| Candida glabrata* | 0 (0)                         | 0 (0)                              |         |

*Categories pooled for data analysis because of the small number in cells; **Fisher’s exact test.
Conclusion
This study confirmed that oral candidiasis in patients with malignant diseases treated with irradiation and chemotherapy is mostly caused by *C. albicans*. It is to be expected that *C. albicans* will remain the most significant causative agent of oral candidiasis, although other species should also be taken into account (such as *C. kefyr* and *C. famata*).

REFERENCES

1. Garcia-Cuesta C, Sarrin-Pérez MG, Bañón JV. Current treatment of oral candidiasis: A literature review. J Clin Exp Dent 2014; 6(5): 1575–82.
2. Canekurci M, Bokor-Bratil M. Candida albicans infection in patients with oral squamous cell carcinoma. Vojnosanit Pregl 2010; 67(9): 760–70. (Serbian)
3. Dineshshankar J, Sivakumar M, Karthikeyan M, Udayakumar P, Shanmugam KT, Kesavan G. Immunology of oral candidiasis. J Pharm Bioall Sci 2014; 6(Suppl 1): 9–12.
4. Byadurahally Raju S, Rajappa S. Prevalence of oral candidiasis. Oral Health 2012; 2(3): 71–4.
5. Krishnan P. Fungal infections of the oral mucosa. Indian Dent Res 2012; 23(S): 650–9.
6. Kurnatowski P, Moqbil S, Kuczynszyck D. Signs, symptoms and the prevalence of fungi detected from the oral cavity and pharynx of radiotherapy subjects with head and neck tumors, and their susceptibility to chemotherapeutics. Ann Parastoi 2014; 60(3): 207–13.
7. Singh A, Verma R, Maruti A, Agrawal A. Oral candidiasis: An overview. J Oral Maxillofac Pathol 2014; 18(Suppl 1): S81–5.
8. Barker BJ, Barker GJ. Oral complications and management of cancer chemotherapy. Northwest Dent 1990; 60(3): 207–8.
9. Wong HM. Oral Complications and Management Strategies for Patients Undergoing Cancer Therapy. Sci World J 2014; 2014: 14.
10. Paranyzhitusov E, Frantczekszki F, Fevleri A, Armaganidis A, Dimopoulos G. Invasive fungal infections in the ICU: How to approach, how to treat. Molecules 2014; 19(1): 1085–119.
11. Farah CS, Lynch N, McCallough MJ. Oral fungal infections: an update for the general practitioner. Aust Dent J 2010; 55(Suppl 1): 48–54.
12. Raman R, Groniadecki S, Pinces DH, Salkin HF, Chatsovedi V. Efficacy of API 20C and ID 32C systems for identification of common and rare clinical yeast isolates. J Clin Microbiol 1998; 36(11): 3396–11.
13. Donnelly P, Bellm LA, Epstein JB, Keshan G. Antimetabolites and carcinogens in the oral cavity. J Pharm Bioall Sci 2014; 6(Suppl 1): 9.
14. Richardson EJ, Rajappa S. Oral mucositis complicating chemotherapy and/or radiotherapy: Options for prevention and treatment. CA Cancer J Clin 2001; 51(5): 290–315.
15. Nicolaou-Galitis O, Darchas J, Markoulafto P, Saripoulou-Lountou A, Kyriacou K, Kaldis G, et al. Oral pseudomembranous candidiasis, herpes simplex virus-1 infection, and oral mucositis in head and neck cancer patients receiving radiotherapy and granulocyte-macrophage colony-stimulating factor (GM-CSF) mouthwash. J Oral Pathol Med 2001; 30(8): 471–80.
16. Hofer E, Jensen SB, Pedersen AM, Bardow A, Nauntofte B. Oral Microflora in Patients with Salivary Gland Hypofunction. Oral Biosci Med 2004; 1(2): 93–108.
17. Martinec EM, Bagan JV, Jimney Y, Snily C. Evaluation of dental health and the need for dental treatment prior to radiotherapy in eighty-three patients with head and neck cancer. Oral Biosci Med 2004; 1(3): 181–5.
18. Xu L, Zhang H, Liu J, Chen X. Investigation of the oral infections and manifestations seen in patients with advanced cancer. Pak J Med Sci 2013; 29(5): 1112–5.
19. Bulicco L, Paz M, Ramadán S, Ramos J, Patróna C, Sortino M, et al. Oral infections caused by yeasts in patients with head and neck cancer undergoing radiotherapy. Identification of the yeasts and evaluation of their antifungal susceptibility. J Mycol Med 2012; 22(4): 346–53.
20. Arendrup MC. Candida and candidaemia. Susceptibility and epidemiology. Dan Med J 2013; 60(11): B4698.
21. Vásárhelyi-Kapcs L, Karszteki D, Jankovics M, Keszegh M, Laszlo M. Epidemiology and susceptibility to antifungal agents of fungi isolated from clinical specimens from patients hospitalized in the Department of General and Liver Surgery of the Medical University of Warsaw. Transplant Proc 2003; 35(6): 2298–303.
22. Patel A, Gruber P. Severe infections in neutropenic patients. Curr Opin Crit Care 2015; 21(6): 586–92.
23. Dufresne SF, Marr KA, Sydnor E, Staab JF, Karp JE, Lu K, et al. The combination of oral amphotericin B with azoles prevents severe infections in neutropenic patients. Br J Haematol 2001; 112(1): 175–80.
24. Richards M. Preventing mucositis. Radiol Endod 2004; 97(1): 47–52.

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