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The mechanistic link between selective vulnerability of the locus coeruleus and neurodegeneration in Alzheimer’s disease

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Abstract
Alzheimer’s disease (AD) is neuropathologically characterized by the intracellular accumulation of hyperphosphorylated tau and the extracellular deposition of amyloid-β plaques, which affect certain brain regions in a progressive manner. The locus coeruleus (LC), a small nucleus in the pons of the brainstem, is widely recognized as one of the earliest sites of neurofibrillary tangle formation in AD. Patients with AD exhibit significant neuronal loss in the LC, resulting in a marked reduction of its size and function. The LC, which vastly innervates several regions of the brain, is the primary source of the neurotransmitter norepinephrine (NE) in the central nervous system. Considering that NE is a major modulator of behavior, contributing to neuroprotection and suppression of neuroinflammation, degeneration of the LC in AD and the ultimate dysregulation of the LC–NE system has detrimental effects in the brain. In this review, we detail the neuroanatomy and function of the LC, its essential role in neuroprotection, and how this is dysregulated in AD. We discuss AD-related neuropathologic changes in the LC and mechanisms by which LC neurons are selectively vulnerable to insult. Further, we elucidate the neurotoxic effects of LC de-innervation both locally and at projection sites, and how this augments disease pathology, progression and severity. We summarize how preservation of the LC–NE system could be used in the treatment of AD and other neurodegenerative diseases affected by LC degeneration.

Keywords Locus coeruleus · Alzheimer’s disease · Tau · Norepinephrine · Selective vulnerability

Abbreviations
Aβ Amyloid-β
AD Alzheimer’s disease
BDNF Brain-derived neurotrophic factor
DHF 7,8-Dihydroxyflavone
DOP-4 N-(2-Chloroethyl)-N-ethyl-bromo-benzylamine
GABA γ-Aminobutyric acid
GSK Glycogen synthase kinase
iNOS Inducible nitric oxide synthase
LC Locus coeruleus
l-DOPS l-3,4-Dihydroxyphenylserine
MRI Magnetic resonance imaging
nbM Nucleus basalis of Meynert
NE Norepinephrine
NFx B Nuclear factor-xB
NFT Neurofibrillary tangle
PiB-PET C-Pittsburgh Compound-B-positron emission tomography
PPARγ Peroxisome proliferator-activated receptor gamma
p-tau Hyperphosphorylated tau
SubC Subcoeruleus

Introduction

The locus coeruleus (LC) is a small brainstem pontine nucleus of neuromelanin-containing neurons, widely recognized as the primary source of the mono-amnergic neurotransmitter norepinephrine (NE). NE plays an important role in modulating many behavioral functions, including attention, mood, motivation, stress and arousal [96, 134]. Furthermore, NE influences blood flow, heart rate, sleep and...
waking patterns [80, 91], and regulates neuroinflammation and neuronal survival [94]. Thus, the noradrenergic system was established as one of the most pivotal neuromodulators in the brain.

Current research supports that dysregulation of the noradrenergic system is a significant player in the development of psychiatric and neurodegenerative disorders, including Alzheimer’s disease (AD), Parkinson’s disease, Lewy body dementia, and frontotemporal lobar degeneration with tau. Analyses of AD brains from progressive stages of the disease have reported significant neuronal and volume loss in the LC [64, 154]. Previous studies have suggested that LC degeneration is age related [50, 85, 93, 157, 164]. However, it is of note that some of these findings were based on a small cohort (n = 5–13) [50, 85] and did not exclude cases with neuropathology elsewhere. Intriguingly, recent unbiased stereological approaches indicated that there are no significant associations between normal aging and changes in the LC [16, 106, 116, 154], suggesting a disease-specific phenomenon.

AD is neuropathologically characterized by extracellular amyloid-β (Aβ) plaque deposits and abnormal accumulation of intracellular hyperphosphorylated tau (p-tau) that form neurofibrillary tangles (NFTs) [103]. In 1991, Braak and Braak defined the six-stage pattern of NFT deposition across the course of AD neuropathologic progression [10]. In 2011, Braak et al. revised the standardized staging scheme to include pretangle stages a–c, which denote the accumulation of subcortical p-tau in the LC that occurs before any cortical tau pathology [12]. This observation situates the LC, together with other interconnected neuromodulatory subcortical structures [143, 153], as one of the first structures to accumulate AD-tau as neuronal inclusions [12], a pathology that ultimately results in neuronal death. The subsequent loss of LC neurons, which occurs with a topographical gradient, is associated with increased Aβ plaque deposition and NFT load in the cortices [8]. Dysfunction of the LC noradrenergic system was also found to be associated with the onset of memory dysfunction and cognitive impairment in AD [55]. Interestingly, LC degeneration correlated better with AD onset and duration than degeneration of the cholinergic nucleus basalis of Meynert (nBM) in the basal forebrain [100, 169], which is also highly vulnerable to tau pathology in AD, indicating that noradrenergic deficits in AD play a major role in disease progression. These findings show that the connection between loss of LC noradrenergic innervation and subsequent onset of neurodegeneration in AD, as well as the detection and treatment of noradrenergic deficits in patients, are important disease aspects that require further research and understanding.

The LC has a unique therapeutic value in AD as it is one of the earliest subcortical regions affected by tau lesions, and prevention of neuropathologic changes in this nucleus could prevent the spread of irreversible changes in the brain [61, 67]. With this in mind, this review will summarize the neuroanatomy of the LC and its function within the noradrenergic system, stating the mechanisms by which NE exerts its responsibility as a key neuromodulator, as well as its role in neuroprotection. We will discuss which AD-related neuropathologic changes are observed in the LC, how the LC–NE system’s dysregulation leads to AD neuropathology, onset and progression, and possible therapeutic interventions to prevent or lessen LC degeneration and NE deficits in AD.

**Neuroanatomy and function of the locus coeruleus**

The LC is a “tube-like” collection of noradrenergic neurons located in the dorsolateral pontine tegmentum, beginning rostrally at the inferior colliculus level and continuing caudally to the lateral face of the fourth ventricle [50, 68]. The LC forms part of the isodendritic core, a group of interconnected and phylogenetically conserved subcortical nuclei [125, 153]. This group also includes the dorsal raphe nucleus, substantia nigra in the midbrain, and nbM in the forebrain [125, 153]. The isodendritic core network plays an important role in neuromodulation by regulating behavior and homeostasis through aminergic and cholinergic projections to the cortices [153].

Unbiased stereological estimates of the total LC cell population showed an average of 98,000 neurons, constituting an overall volume of approximately 13 mm³ [154]. LC neurons are identifiable by their neuromelanin pigment, as well as immunolabeling with dopamine β-hydroxylase, the enzyme that converts dopamine to NE [138, 146] or tyrosine hydroxylase, the enzyme that catalyzes the rate-limiting step in NE biosynthesis [121] (Fig. 1). Using such methods, several studies have identified distinct cellular heterogeneity within the LC, characterizing two classes of medium-sized neurons; the large multipolar cells (~35 μm) and smaller fusiform cells (~20 μm) [138, 145]. Their distinct cellular morphologies could indicate differences in their characteristics and function. Though both cell types are dispersed throughout the LC, the small fusiform cells dominate the densely packed rostral portion, indicating a cytoarchitectonic bias [145]. In addition, recent studies have indicated phenotypic variability among subsets of LC neurons [17, 124, 165]. Hippocampal and prefrontal-innervating LC neurons differ in their physiological response to the α₂-adrenoreceptor agonist clonidine, suggesting a functional heterogeneity within the LC neuron population [165].

The structure and organization of neurons within the LC and adjacent subcoeruleus (SubC) are essential for its function as the primary source of NE. Retrograde labeling techniques using radioactive or fluorescent tracers have
established that each single LC axon innervates extensively into several brain regions, in particular the forebrain, cerebellum, brainstem and spinal cord [122] (Fig. 2), with 90% of these LC efferent projections remaining ipsilateral [95, 128]. Several groups have identified that topographic organization of the LC neurons are dependent on their output target, with rostrally located cells projecting to the forebrain region, innervating the hippocampus and septum; whereas, cells in the middle and caudal portions of the LC project to the cerebellum, basal ganglia and spinal cord, and regulate autonomic function [86, 95, 122, 136] (Fig. 2).

LC neuronal projections to the cortex occur in three ways: (1) monosynaptically, whereby the axons innervate the region directly [38, 112]; (2) through the thalamus, using this structure as a 'hub' [47, 76]; or (3) via the cholinergic basal forebrain nuclei, including the nbM [53]. At the projection target area, NE is released by two different types of LC axonal terminals; typical synaptic transmission and varicosities [41, 94, 134]. Axonal varicosities may permit the extra-synaptic release of NE commonly referred to as volume transmission, which refers to the diffuse release of neurotransmitters [41, 94, 134]. This method of release is an important and essential characteristic of neuromodulatory neurons as it influences the excitability and synaptic plasticity of a vast range of proximal cells [41, 94, 134].

Once released, NE exerts its neuromodulatory effects through binding to G-coupled α- and β-adrenoreceptors (Fig. 1), which are present on neurons, glia and immune cells, and can be detected in cerebral microvessels [124, 127, 147]. One of the most important receptors in mediating the downstream effects of NE is the α2-adrenoreceptor [57, 127] that functions as both inhibitory autoreceptor on noradrenergic neurons and modulator of noradrenergic innervation on postsynaptic cells [127]. α2-Adrenoreceptors are also highly expressed by LC neurons, contributing to the neuromodulatory effects of NE [138]. The α2-adrenoreceptor subtype, α2A-adrenoreceptor, is found abundantly in the cortex, hippocampus, hypothalamus, amygdala, brainstem, and spinal cord; regions that are highly innervated by LC-projecting axon terminals [78, 87, 127]. The abundance of the α2A-adrenoreceptor subtype in such regions suggests that it may play an important role in mitigating NE signaling [78, 87, 127]. On binding to adrenoreceptors, NE initiates a vast array of downstream pathways, leading to further neurotransmitter release, regulation of inflammatory processes and growth factor expression, all of which are important in modulating its many roles in the brain [94, 127].

It is widely recognized that NE plays a critical role in regulating neuroinflammation. For example, LC axonal terminals were found in close contact to astrocytes and microglia [23, 83], suggesting a modulatory relationship. In early in vitro studies, NE was shown to inhibit the induction of MHC class II antigen expression on astrocytes by IFN-γ, indicating that NE downregulates the immune response in the brain [45]. Since then, several studies have shown that NE plays a fundamental role in reducing a number of inflammatory genes, including NOS2, IL-1β, ICAM-1, adhesion molecules, and TNF-α, in cells such as microglia, astrocytes and endothelial cells [27, 40, 99].

Findings indicate that much of the neuromodulatory effects of NE occur through downstream cAMP pathways. Through the induction of this pathway, NE was shown to increase the expression of peroxisome proliferator-activated receptor gamma (PPARγ) in astrocytes and neurons, and research suggests that this upregulation partly mediates the anti-inflammatory effects of NE [74]. This is supported by the findings which suggest that treatment with PPARγ agonists restores the depleted levels of inflammatory markers that are observed in LC-lesioned animal models, as well as inhibiting the activation of proinflammatory molecules such as nuclear factor-κB (NFκB) [59] (Table 1). Another mechanism by which NE exhibits its neuromodulatory effects
includes increased IκBα expression, a protein involved in inhibiting the NFκB pathway whose action is highly immunosuppressive [48]. This is supported by findings that animals with LC degeneration have reduced basal levels of inhibitory IκB proteins [59].

In addition to releasing neurotransmitters, the LC also co-transmits a variety of neuropeptides, most namely neuropeptide Y and galanin [79], which are expressed in 20% and 80% of neurons, respectively [62]. Studies have suggested that the co-expression of these neuropeptides helps regulate central adrenergic transmission mediated through α2-adrenoreceptors [163]. Furthermore, these cells express essential neurotrophins, including brain-derived neurotrophic factor (BDNF), which dictates neuronal survival and differentiation [18, 25], as well as receptors for neuropeptide Y, galanin, somatostatin and hypocretin/orexin [41].

AD-related neuropathologic changes in the locus coeruleus

Hyperphosphorylated tau accumulation in the locus coeruleus

During the development of AD, NFT pathology progresses in the cortex in six stages: in stages I/II, NFTs are present in the transentorhinal and entorhinal cortex; stages III/IV, NFTs appear in the limbic system and temporal cortices; and finally stages V/VI, where NFTs are present throughout the association and primary cortices [10]. However, a recent revision of the original staging system by Braak and colleagues in 2011 has incorporated the LC as the earliest site of p-tau accumulation [12]. According to the updated staging, ‘pretangle stage a/b’ refers to p-tau accumulation confined only to the pontine tegmentum, in or in close proximity to the LC [12]. In ‘pretangle stage a’, the LC cell processes are positive for AT8 immunoreactivity; whereas in ‘pretangle stage b’, p-tau has extended further down the axon with spiked protrusions along the neuron outer somatic rim [12]. Further research indicated that at Braak stage 0, 8% of LC neurons are p-tau-positive, which doubled by Braak
stage I [32]. By Braak stage VI, 100% of LC neurons had tau pathology [5] (Fig. 3).

Recent studies have shown that p-tau begins to accumulate in the LC early in life, in some cases as young as 10 years of age, with up to 90% of individuals having some tau pathology in the LC by the age of thirty [11] (Fig. 3). Similar findings were reported by other groups who observed tau lesions, including NFTs and neuropil threads, in 72% of

| Table 1 | Animal model studies used in locus coeruleus research |
|-----------------|-----------------|-----------------|-----------------|-----------------|
| References in order of appearance | Model(s) | Species | LC Lesion type | Section(s) |
| Heneka et al. 2003 [59] | Control | Rat | DSP-4 | Neuroanatomy and function of the locus coeruleus |
| Heneka et al. 2006 [61] | APP23 | Mouse | DSP-4 | Locus coeruleus atrophy in Alzheimer’s disease |
| Kalinin et al. 2007 [67] | APP V717F | Mouse | DSP-4 | Locus coeruleus atrophy in Alzheimer’s disease |
| Kelly et al. 2019 [72] | Tg344-19 | Rat | DBH-sap | Locus coeruleus atrophy in Alzheimer’s disease |
| Hammerschmidt et al. 2013 [56] | APP/PS1 | Mouse | DBH−/− | Locus coeruleus atrophy in Alzheimer’s disease; l-DOPS (droxidopa) |
| Chalermpalanupap et al. 2018 [19] | Tau P301S | Mouse | DSP-4 | Locus coeruleus atrophy in Alzheimer’s disease |
| Heneka et al. 2002 [58] | Control | Rat | DSP-4 | Locus coeruleus atrophy in Alzheimer’s disease |
| Heneka et al. 2010 [60] | APP V717I & APP/PS1 | Mouse | DSP-4 | Locus coeruleus atrophy in Alzheimer’s disease |
| Kang et al. 2020 [69] | Tau P301S & MAPT/AEP−/− | Mouse | DBH−/− | Neuronal vulnerability in the locus coeruleus |
| Castren et al. 1995 [18] | Control | Rat | None | The role of locus coeruleus in neuroprotection |
| Fawcett et al. 1998 [39] | DBH–BDNF | Mouse | None | The role of locus coeruleus in neuroprotection |
| Mastunaga et al. 2004 [97] | Control | Rat | None | The role of locus coeruleus in neuroprotection |
| Nakai et al. 2006 [114] | Control | Rat | None | The role of locus coeruleus in neuroprotection |
| Ádori et al. 2015 [1] | SSTR2−/− | Mouse | None | The role of somatostatin in neuroprotection |
| O’Meara et al. 2000 [115] | GAL−/− | Mouse | None | Neuropeptides expressed by the locus coeruleus exhibit neuroprotective effects |
| Elliott-Hunt et al. 2004 [33] | GAL−/− & GAL overexpression | Mouse | None | Neuropeptides expressed by the locus coeruleus exhibit neuroprotective effects |
| Kalinin et al. 2012 [68] | 5xFAD | Mouse | None | l-DOPS (droxidopa) |
| Devi et al. 2012 [28]; Zhang et al. 2014 [171] | 5xFAD | Mouse | None | 7,8-Dihydroxyflavone |
| Takeda et al. 1984 [150] | Control | Mouse | None | Vindeburanol |
| Labatut et al. 1988 [77] | Control | Rat | None | Vindeburanol |
| Braun et al. 2014 [13] | 5xFAD | Mouse | None | Vindeburanol |
| Wang et al. 2010 [166]; Sun et al. 2012 [144] | 3xTgAD | Mouse | None | Allopregnanolone |
| Torres-Sanchez et al. 2018 [158] | Control | Rat | None | Brain stimulation technologies |
| Follesa et al. 2007 [42] | Control | Rat | None | Brain stimulation technologies |
| Rorabaugh et al. 2017 [129] | TgF344-AD | Rat | None | Brain stimulation technologies |

LC locus coeruleus, DSP-4 N-(2-chloroethyl)-N-ethyl-bromo-benzylamine, APP amyloid precursor protein, PS1 presenilin-1, DBH dopamine β-hydroxylase, BDNF brain-derived neurotrophic factor, SSTR2 Somatostatin Receptor 2 gene, GAL galanin gene, MAPT tau gene, AEP Asparagine endopeptidase
individuals aged 31–40 years, and 94% of individuals aged 41–50 years [123]. In terms of the onset of AD, it is suggested that NFT formation in the LC occurs 10 years prior to any cognitive changes [20, 55, 153] and at least 25 years before significant neuronal loss [16].

The presence of NFTs in the LC can have a multitude of effects regarding differential gene expression. NFT accumulation in the LC is associated with colocalized expression of tau kinases, including MAPK/ERK, SAPK-JNK, p38, glycogen synthase kinase (GSK) and GSK-3β, with several kinases showing immunoreactivity restricted to neurons bearing p-tau [2]. In addition, NFTs in the LC have increased colocalization with neuroketal adducts (oxidation product of docosahexenoic acid) [2], which was shown to rapidly induce protein cross-linking and aggregation, and are markers of age-related neurodegeneration [29]. Further, NFT accumulation in the LC was shown to correlate with decreased expression of superoxide dismutase 1, suggesting a reduced response to oxidative stress [2]. LC neurons containing truncated tau were shown to have decreased levels of voltage-dependent anion channels, suggesting a decrease in the number of functional mitochondria [2]. Whole-transcriptome analysis of the LC at Braak stage IV identified upregulated expression of proteins associated with heat shock protein-binding and ATP metabolism, and downregulation of DNA-binding and members of the small nucleolar RNAs family protein expression [2]. This suggests that abnormal tau accumulation triggers multiple dynamic changes in LC neurons, perhaps as compensatory mechanisms or to induce cell death. Similarly, active caspase-6 antibody positivity, a marker of apoptosis, was shown to increase in NFT-bearing LC neurons with advancing Braak stage [155]. In contrast, macroautophagy, identified by the autophagosome marker LC3, was found to be decreased during AD progression [155]. As macroautophagy is responsible for the degradation of misfolded proteins and was shown to reduce p-tau accumulation in affected neurons [66], dysregulation of this system could contribute to the formation of NFTs in the LC, ultimately leading to activation of caspase-6 and subsequent neuronal death [155]. Furthermore, neurons associated with
aggregan-containing perineuronal nets are spared from the formation of NFTs [14]. Perineuronal nets are assemblies of extracellular matrix proteins that are implicated in neuroprotection and neural activity regulation [168], thus suggesting a protective mechanism against the development of AD pathology [14]. Interestingly, the LC, which is highly affected by tau pathology, is devoid of perineuronal nets, and its neurons are not contacted by aggregan-immunoreactive axonal coats, indicating a heightened vulnerability to tau-associated neuronal death [104].

**Amyloid-β in the locus coeruleus**

Despite the early implications of p-tau and the LC, Aβ plaques do not occur in this region until Thal amyloid phase 5 [152], where 62% of cases were found to have Aβ deposits (Fig. 3). However, studies by Muresan and Muresan indicated that LC-derived cell lines are prone to intracellular Aβ accumulation in both the neurites [109] and at the axonal terminals [110]. Interestingly, intracellular Aβ accumulation in these cells did not induce apoptosis or necrosis [109]. With projection sites from LC neurons innervating the entire brain, the group hypothesized that early intracellular accumulation of Aβ at axonal terminals could serve as seeds for further Aβ propagation and aggregation [110]. Furthermore, recent findings have indicated that Aβ oligomers may be involved in the abnormal phosphorylation of tau [170]. Aβ oligomers were shown to bind to an allosteric site on α2A-adrenoreceptor, which are present on LC neurons, inducing a redirection of NE-induced signaling to GSK-3β [170]. GSK-3β induces tau phosphorylation, and therefore, activation of this pathway by Aβ oligomers may lead to tau hyperphosphorylation and the development of tau-related pathology in AD [170]. This could implicate Aβ oligomers in the susceptibility of noradrenergic neurons of the LC to NFT formation, a hypothesis that needs to be explored in the future.

**Neuronal cell loss**

The AD brain was shown to have marked reductions in the size and volume of the LC (Fig. 3). Early studies reported that the rostrocaudal extent of the nucleus shrinks by an average of 6 mm in these patients [49]. More recently, using unbiased stereological analysis, reports have shown that as the Braak stage increases by one unit, the LC volume decreases by 8.4% [154]. Furthermore, 3T magnetic resonance imaging (MRI) assessment of LC signal intensity, which is determined by the intrinsic neuromelanin-driven contrast and, thus, may correlate with LC neuron population [7], showed a marked decrease in the LC signal in AD dementia patients. This finding was regardless of clinical presentation and independent of the [11C]-Pittsburgh Compound-B-positron emission tomography (PiB-PET) Aβ load, which correlated with memory performance in typical AD [117]. Although it did not reach significance, AD dementia patients with an atypical (non-annestic) clinical presentation were observed to have a lower LC signal than typical AD dementia patients [117]. It will be important for future studies to further investigate whether potential differences exist in atypical AD dementia patients; especially given they are more commonly younger at symptom onset [111]. A recent study using MRI to examine brainstem volumetric changes indicated that participants with neuropsychologically confirmed mild cognitive impairment or clinically diagnosed AD dementia had smaller LC volumes as compared to cognitively normal individuals [31].

Reduction in LC volume can be reflected by a vast loss of noradrenergic neurons or neuronal shrinkage. Studies have shown that LC neuronal loss can average 63% in AD [49] and occurs midway through disease progression [154]. Increased neuronal loss is also associated with earlier age of onset [169], increased disease duration, and earlier age of death [8]. In terms of clinical presentation, 30% of LC neurons are lost between progression from cognitively unimpaired to amnestic mild cognitive impairment, with an additional 25% reduction of LC neurons during progression to mild/moderate AD [71]. An unbiased stereological study using neuropsychological and neuropathologic measures to determine AD stage reported that LC neuronal loss is detectable at preclinical stages, and that progressive loss of LC neurons parallels the progression of the disease to severe dementia [3]. Interestingly, Hoogendijk and colleagues reported that AD dementia patients exhibit an 82% and 39% decrease in the number of large multipolar and small fusiform neurons, respectively [64]. This manifested in a reduction in the large to small pigmented neuron ratio, suggesting that decreased LC volume and neuronal count in AD is not entirely due to cell death, but also cell shrinkage and phenotype loss [64]. Similarly, it was reported that LC neurons display characteristic morphological changes in the early stages of AD, such as contracted dendrites and swollen cell bodies, which could also augment noradrenergic dysfunction [153].

It is widely recognized that a vulnerability gradient of the LC across the rostrocaudal axis exists in AD and differs between neurodegenerative diseases. The rostral portion is affected to a greater extent in AD [32], with an 83% loss of length, as compared to the middle and caudal parts (23% and 15%, respectively) [64]. The degree of rostral degeneration was shown to correlate with aggressive behavior in patients with dementia [98]. With rostral neurons projecting to the forebrain and cortical structures, degeneration of this portion could explain behavioral changes observed in AD dementia patients.
To reduce the consequences of LC pathology, many compensatory mechanisms are implemented by the remaining noradrenergic neurons in order to maintain its essential role within the brain. Evidence from postmortem AD and Lewy body disease brain studies with significant LC neuronal loss indicated that the noradrenergic system undergoes major changes that are consistent with compensation [149]. This includes increased tyrosine hydroxylase mRNA expression and NE synthesis, reduced NE uptake at synaptic terminals within the surviving neurons, as well as dendritic sprouting into the peri-LC dendritic zone and axonal sprouting in projection areas [148, 149]. Upregulation of neurotransmitter biosynthesis could explain the increase in NE levels in the cerebrospinal fluid of AD dementia patients with severe LC degeneration, which is also associated with poor cognitive function [35, 63, 153].

**Locus coeruleus atrophy in Alzheimer’s disease**

LC neuronal dysfunction in neurodegenerative diseases can have a multitude of effects, both locally and at the projection areas. On a local basis, many cellular changes may augment the dysregulation of the noradrenergic system. Unbiased stereological methods have indicated that LC neurons positive for p-tau have significant reductions in synaptic connectivity, as shown by a loss of approximately 50% of synaptophysin-immunoreactive perineuronal dots [2]. Furthermore, microarray analysis using mRNA from LC neurons displayed a significant reduction in expression of genes associated with neuritic and structural plasticity, as well as mitochondrial respiration and redox gene expression, in patients with amnestic mild cognitive impairment and mild/moderate AD as compared to controls [71]. The downregulation of such genes was found to be associated with cognitive deficits [71]. These findings suggest that LC neurons have a lessened functional capacity, especially in terms of receiving activating inputs, which can ultimately lead to decreased noradrenergic signaling.

Loss of NE concentration throughout the brain was reported by many groups. In AD, NE concentrations in the mid-temporal cortices are decreased by 31% [98] and up to 50% only two years after dementia onset [49]. In addition, AD dementia patients who died before the age of 79 and those with greater loss of LC neurons were found to have a NE deficit in the cingulate cortex [8, 131].

LC and NE network dysfunction can also contribute to the early behavioral alterations seen in preclinical AD, such as sleep disturbances and mood changes. During the awake state, LC neurons are highly active, and consistent fluctuations are observed throughout the day for certain waking behaviors [4]. The LC firing frequency then decreases during slow-wave and non-REM sleep, and is completely diminished by the REM stage and throughout paradoxical sleep [4, 156]. Clinical studies have reported a high frequency of AD dementia patients exhibiting sleep disturbances [159], which is associated with disease severity [102, 130] that could be in part due to loss of LC neurons. In addition to sleep deficits, it is widely recognized that noradrenergic dysfunction can lead to the development of mood disorders [126]. NE upregulation was shown to have antidepressant effects in patients with major depression, and with depression being an early symptom of AD dementia, it is possible that early noradrenergic de-innervation could augment this behavioral phenotype [44, 126, 156, 174].

Diminished signaling to projection areas can result in varied phenotypes. In AD, although α2-adrenoreceptor expression is increased in the hippocampus at Braak stage I, its expression is reduced at Braak stage IV [2]. Tyrosine hydroxylase expression also declines with age and disease progression, especially within the hippocampus and amygdala at Braak stages III and IV [2]. A study using APP/PS1 mice crossbred with dopamine β-hydroxylase (–/–) mice, which resulted in a mouse with reduced NE production, showed that depletion of NE impaired long-term synaptic plasticity in the hippocampus, leading to spatial memory deficits [56]. NE deficiency also modified the expression of plasticity-related synaptosomal subunit proteins, such as CAMKII and NMDAR [56]. Similarly, inducing LC degeneration using N-(2-chloroethyl)-N-ethyl-bromo-benzylamine (DSP-4) in an APP transgenic mouse reduced cerebral glucose metabolism, acetylcholinesterase activity and neuronal integrity, leading to increased memory deficits [61]. LC degeneration also led to increased levels of inducible nitric oxide synthase (iNOS) in the projection areas, resulting in augmented neuronal loss in the frontal cortex and hippocampus [61]. Furthermore, lesioning of the LC in a P301S mouse model of tauopathy resulted in spatial and associative memory deficits, as well as exacerbated hippocampal tau burden and increased lethality, suggesting a direct role of LC degeneration in potentiating cognitive deficits and inducing AD-related pathology and mortality [19].

Research has shown, both in human [8, 92, 157] and animal studies [61, 67], that loss of LC neurons leads to increased Aβ deposition. In human studies, neuronal loss across the rostrocaudal axis of the LC was shown to topographically correspond to the distribution of Aβ plaques within the cerebral cortex of AD brains [92]. Post-mortem analysis have shown that loss of neurons from the rostral, middle and caudal portions of the LC correlates with increased Aβ plaque burden in the frontal cortex, temporal cortex and occipital cortex, respectively [92].

In animal models, DSP-4-treated mice display a five-fold increase in Aβ plaque burden and increased average plaque size [67]. Furthermore, LC-lesioned animals...
exhibit increased levels of APP C-terminal cleavage fragments, as well as decreased expression and activity of the Aβ-degrading enzyme neprilysin, both of which correlated with augmented Aβ pathology [67]. Similarly, other studies using LC-lesioned rats revealed 30% and 20% increases in the deposition of Aβ in the cortex and hippocampus, respectively [72]. Interestingly, mice with diminished abilities to produce NE do not exhibit exacerbated Aβ deposition, suggesting that LC degeneration itself causes the increase in Aβ levels, and it is probable that such mechanisms are independent of NE signaling, specifically [56].

With the noradrenergic system highly innervating the cerebrovasculature, it is no surprise that LC degeneration can have a major effect on its integrity. Loss of the neural plexus originating from the LC results in increased immunologic infiltration of the brain parenchyma microvasculature and thickening of capillary walls, subsequently leading to the development of microangiopathy and alterations to normal blood–brain barrier functions, a common phenotype of AD [137]. In animal models, lesioning of the LC resulted in vast Aβ deposition within the prefrontal cortex microvessels, indicating the neuropathologic onset of cerebral amyloid angiopathy [72]. In addition, LC-lesioned rats exhibited increased albumin staining, which suggests decreased blood–brain barrier integrity and increased microvasculature permeability and leakage [72]. Moreover, loss of LC–NE signaling also resulted in significant compensatory remodeling of the arterioles in these animals, as displayed by a 45% increase in wall-to-lumen ratio [72]. Together, these findings suggest that LC degeneration plays a key role in the dysregulation of the cerebrovascular system, as seen in AD.

Many studies have suggested that the dysfunction of the noradrenergic system potentiates the neurotoxic proinflammatory condition seen in AD. The presence of NFTs in the LC is associated with the upregulation of inflammatory markers, as well as increased microglia infiltration into the LC [2]. LC-lesioned P301S mice exhibit increased activation of microglia and astrocytes in the hippocampus, which may contribute to hippocampal degeneration in this model [19]. LC cell loss also resulted in amplified microglial and astroglial activation in the frontal cortex and hippocampus in mutant APP mouse models [67] and, thus, the dysregulation of the LC–NE system may play a major role in disease-related inflammation.

Studies have indicated that LC degeneration exacerbates the proinflammatory response to Aβ deposition in the cortex sooner, more robustly, and with longer duration than in control animals [58]. Intriguingly, Aβ injection induced expression of iNOS in microglia in control animals, as compared to LC-lesioned animals whose iNOS expression was localized to neurons, which is in parallel to AD brains [58]. The proinflammatory condition was attenuated by injection of either NE or a β-adrenergic receptor agonist, suggesting that both NE depletion and LC degeneration enhance neuroinflammation and neuronal cell death in AD [58]. Similarly, research shows that NE protects cortical neurons from cytotoxic microglia by decreasing NOS2 production, leading to reduced cell death [88]. Further studies by the same group revealed that LC lesioning leads to increased glial inflammation and Aβ deposition in LC projection areas, including the hippocampus and frontal cortex; whereas, non-projection areas such as the paraventricular thalamus remain unaffected [61]. Findings suggest that increased Aβ deposition following LC degeneration may be in part due to impaired recruitment of microglia to Aβ plaque sites, as well as diminished capabilities of the glial cells to phagocytose Aβ [60]. These findings suggest LC degeneration contributes substantially to the development of AD pathology and disease onset.

**Neuronal vulnerability in the locus coeruleus**

The extent to which the LC is affected in AD suggests that its neurons are especially vulnerable to neuropathologic changes [167]. One hypothesis that could confer this susceptibility is the cytoarchitecture of these cells, as LC neurons have long and thin axons that have poor or incomplete myelination [153]. Such a characteristic forces the cell to increase its energy output to perform efficient action potentials that lead to increased cellular oxidative stress, and gaps in the myelin sheath allow greater exposure to environmental toxins [153].

Research has shown that the LC innervates the vast majority of the brain microvasculature, with the estimation that, on average, each LC neuron is responsible for innervating 20 m of capillaries, a coverage that surpasses any other neuron [118]. The exposure of LC neurons to the circulatory system of the brain suggests that they can readily take up environmental toxins from the blood, and the extent to which this occurs can be increased when blood–brain barrier integrity is lost, such as in AD [73, 135]. Furthermore, the LC is in close proximity to the fourth ventricle, which enhances its exposure to the toxins present in the cerebrospinal fluid [107]. Research has shown that the LC is sensitive to the presence of environmental toxins, and increased levels of heavy metals, such as mercury, bismuth and silver, were reported in the LC neurons of AD brains [41, 119]. Studies suggest that accumulation of heavy metals may lead to the development of AD pathology within the LC [141]. Interestingly, AD brains have approximately the same proportion of LC neurons that contain heavy metals (19%) as those that contain p-tau (21%) [119], suggesting that insult to the LC via the accumulation of environmental toxins plays an important role in its degeneration and dysfunction.

Research has suggested that molecular mechanisms may contribute to the development of early tau pathology in...
selectively vulnerable LC neurons. 3,4-Dihydroxyphenylglycolaldehyde, a metabolic product of NE that is produced exclusively in noradrenergic neurons, was shown to activate asparagine endopeptidase cleavage of tau into aggregation-and propagation-prone forms [69]. 3,4-Dihydroxyphenylglycolaldehyde-induced tau aggregation resulted in LC neurotoxicity, propagation of tau pathology to interconnected brain regions and cognitive impairment in an AD mouse model [69]. Furthermore, activation of asparagine endopeptidase and subsequent tau cleavage required the presence of NE [69], suggesting that production and ultimately the metabolism of NE by noradrenergic neurons could potentiate the selective vulnerability of the LC to neuropathologic changes.

Another factor that may exacerbate the vulnerability of LC neurons is their high bioenergetic need [167]. To conserve their essential physiological function within the brain, LC neurons sustain their spiking rate by exhibiting autonomous activity in the absence of glutamate and γ-aminobutyric acid (GABA) inputs [133]. This is achieved through activity-dependent Ca\(^{2+}\) entry, which upon activation, leads to the induction of mitochondrial oxidative stress and results in increased LC neuronal vulnerability [133]. Epidemiological studies have also identified a link between NE availability and an increased risk of developing AD dementia. Polymorphisms in the dopamine β-hydroxylase gene, which influences NE availability in the brain, cause a significant decrease in NE concentration and individuals carrying such polymorphism exhibit a twofold increased risk of developing AD dementia [24].

**The role of locus coeruleus in neuroprotection**

NE was shown to have an essential role in neuroprotection by modulating inflammation, stress, and Aβ-mediated neurotoxicity [26, 82, 88, 162]. To exert its neuroprotective effect, NE binds to β₁- and β₂-adrenergic receptors to activate downstream signaling pathways involving cAMP response element-binding protein, subsequently leading to the expression of endogenous nerve growth factor (NGF) and BDNF [26]. Interestingly, NGF was found to inhibit tau accumulation and phosphorylation in rat hippocampal neurons by activating cAMP-dependent pathways [172]. Significant loss of NE levels following LC degeneration can, therefore, exacerbate many mechanisms involved in inducing neuronal death and subsequent brain atrophy.

Several studies have shown that many of the neuroprotective mechanisms of NE are implemented by BDNF, which plays a major role in influencing axonal branching, outgrowth and synaptic plasticity [65, 84]. Accumulating evidence suggests that the Trk family of tyrosine protein kinase receptors, namely TrkB, is activated by BDNF to mediate its biological function [6]. This is supported by findings which show that the neuroprotective effects of NE are inhibited by the Trk receptor antagonist K252a [82].

The involvement of the noradrenergic system in the production of BDNF could follow the neurotrophic factor hypothesis, which states that such factors are produced and released by neurons to modulate differentiation and survival of the neurons that they innervate [39]. In animal models, LC neurons were shown to produce a large amount of BDNF [18], which is transported from the cell soma into the noradrenergic projection terminals that widely innervate many regions of the brain [39]. Furthermore, the upregulation of BDNF in noradrenergic neurons leads to increased activation of TrkB throughout the innervated neocortex, which results in long-term cortical morphologic alterations [39]. Interestingly, exogenous BDNF infusion induced a substantial increase in noradrenergic axon density within the frontal cortex of aged rats, which is not recapitulated in young or middle-aged animals, suggesting that BDNF is essential for maintaining noradrenergic cells within the aged brain [97]. Similarly, exogenous BDNF increased multipletreshold antidromic response in rat LC neurons, indicating that BDNF stimulates remodeling of presynaptic axon terminals, leading to the functional changes seen within the projection areas of the aging rat brain [114].

Despite their high expression in adult rodent LC neurons, it was reported that BDNF and TrkB expression is very low in adult human LC neurons [151]. However, NE-induced expression of BDNF and TrkB, or lack of, may still be of importance in other brain regions innervated by the noradrenergic system. In terms of neurodegenerative disease, mounting evidence has indicated that the expression of BDNF and TrkB is downregulated in the AD brain [46], especially in hippocampal and neocortical areas that are highly innervated by noradrenergic LC neurons [140]. Likewise, TrkB expression is markedly downregulated in the cholinergic neurons of the nbM throughout AD progression, indicating a fundamental function in disease development [52]. Intriguingly, BDNF immunoreactivity was found in Aβ plaques and NFT-bearing neurons [108] and stimulation of the BDNF/TrkB pathway in mouse cells caused significant dephosphorylation of tau [34], suggesting a neuroprotective role in AD.

**Norepinephrine protects against Aβ-induced neurotoxicity**

Recent studies support a significant role of NE in preventing Aβ-induced neurotoxicity. NE was shown to protect human neurotoblastoma cultures from Aβ₁₋₄₂ and Aβ₂₅₋₃₅ toxicity by preventing increases in intracellular reactive oxygen species, mitochondrial membrane depolarization and
Somatostatin expression was found to be greatly reduced in the temporal cortex of AD brains in association with aberrant clustering and bulging of tyrosine hydroxylase-positive afferents [1]. Decreasing somatostatin receptor 2 mRNA expression in the LC was observed from Braak stages III/IV and throughout the progression of the disease [1]. Following the generation of animal models, somatostatin receptor knockout mice were found to exhibit selective loss of noradrenergic innervation on a global scale [1]. As expression levels of other major LC transcripts, such as tyrosine hydroxylase, dopamine β-hydroxylase and galanin, remain unchanged, these findings suggest that somatostatin receptor loss plays a major role in LC neuronal vulnerability and contributes to dysfunction of the noradrenergic system as a whole.

Neuropeptides expressed by the locus coeruleus exhibit neuroprotective effects

Galanin is one of the most abundant neuropeptides in the brain, is highly expressed by LC neurons, and was shown to have implications in the development of neurodegenerative diseases. In AD, studies have observed that galanin expression is preserved within the remaining neurons of the LC, suggesting a neuroprotective effect [101]. Furthermore, galanin-containing fibers are found to be in close proximity to cholinergic basal forebrain neurons, and this innervation is enriched in AD brains, which may suggest neurodegenerative compensation [9]. In animal models, loss of galanin function resulted in vast cholinergic degeneration in the medial septum and basal forebrain and cognitive deficits [115]. Galanin overexpression and activation of its receptor using a high-affinity agonist reduced chemically-induced cell death in the hippocampus of transgenic animals [33]. This provides evidence suggestive of a major role of galanin in the neuroprotective effects of the LC.

Neuromelanin: toxic or protective?

Neuromelanin, a pigment commonly used to identify noradrenergic neurons, slowly accumulates in the LC with age, followed by a significant decrease within individuals aged 60 and older [139]. Intriguingly, research has shown that neuromelanin may play a neuroprotective and/or a neurotoxic role in AD progression [175]. In this ill-fated rescue, the protective mechanism set in place is hypothesized to initiate a cascade of events that ultimately leads to a toxic state that overwhelms the neuron. In a neuroprotective sense, neuromelanin chelates heavy metals such as mercury, cadmium, lead and iron, and diminishes their toxic nature [175]. However, the accumulation of heavy metals in
neuromelanin-containing organelles can result in a neurodegenerative response by which the neurons die and release the toxins into the extracellular space, activating microglia and inducing neurotoxic inflammation [175].

Possible therapy implication

To compensate for the significant loss of NE following LC de-innervation, several studies have demonstrated that certain therapies could partially reverse the dysregulation of the noradrenergic system, thus minimizing the neurodegenerative process [120, 161] (Table 2). Such treatments could be a promising therapy for not only AD, but also other neurodegenerative diseases where loss of NE and LC atrophy is observed, such as Parkinson’s disease and Lewy body dementia.

1-DOPS (droxidopa)

l-3,4-Dihydroxyphenylserine (l-DOPS), a synthetic catecholamine that is a precursor of NE, can be used to directly increase levels of NE [41]. l-DOPS can be administered orally and can readily permeate the blood–brain barrier, where it is converted by carboxylation to NE [54] (Fig. 1) and was shown to effectively treat dopamine β-hydroxylase deficiency and neurogenic orthostatic hypotension [41].

In a 5xFAD transgenic mouse model, co-treatment of l-DOPS and the NE reuptake inhibitor atomoxetine was found to increase NE levels in the brain, as well as improve cognitive functions, reduce neuronal cell death, decrease astrocyte activation and increase BDNF and NGF levels in the cortex and hippocampus [68]. In addition, l-DOPS and atomoxetine co-treatment upregulated the expression of neprilysin, leading to an increase of Aβ degradation and, thus, a reduction in Aβ plaque burden [68]. l-DOPS treatment of NE-depleted APP/PS1 mice also partially rescued spatial memory impairment [56], as well as restored microglial migration and Aβ phagocytosis [60]. Together, these findings suggest that l-DOPS is an encouraging therapeutic method in the treatment of AD.

Norepinephrine transporter inhibitors

The NE transporter, located in the plasma membrane of noradrenergic neurons, facilitates the removal of excess NE from the synaptic cleft and is, thus, the primary mechanism of noradrenergic signaling inactivation [173]. Targeting the NE transporter, which in turn inhibits the reuptake of NE and increases synaptic NE concentration, was shown to alleviate the symptoms of neuropsychiatric disorders such as depression and attention deficit hyperactivity disorder [173]. Therefore, NE transporter inhibitors such as atomoxetine are routinely used as treatment strategies in these diseases. An ongoing clinical trial (ClinicalTrials.gov Identifier: NCT01522404) is evaluating the efficacy of atomoxetine in participants diagnosed with mild cognitive impairment, to reverse the loss of NE in the brain and slow neurodegeneration. As atomoxetine is already an FDA-approved drug that is safe in the elderly, this is a promising therapeutic measure for treating LC–NE dysregulation in AD.

7,8-Dihydroxyflavone

BDNF plays an important role in exhibiting the neuroprotective actions of NE. Therefore, increasing brain levels of BDNF could act as a therapeutic mechanism in AD. When given orally, BDNF is degraded by digestive enzymes [30]. When given systemically, BDNF administration is hindered by its inability to penetrate the blood–brain barrier and its

| References in order of appearance | Therapeutic method | Target | Mechanism of action |
|----------------------------------|--------------------|--------|---------------------|
| Feinstein et al. 2016 [41]; Hammerschmidt et al. 2013 [56]; Heneka et al. 2010 [60]; Kalinin et al. 2012 [68] | l-DOPS (droxidopa) | DOPA decarboxylase | Increase levels of NE |
| Zhou 2004 [173]; ClinicalTrials.gov, NCT01522404 | Atomoxetine | NE transporter | Increase levels of NE at synapse |
| Devi et al. 2012 [28]; Du and Hill, 2015 [30]; Liu et al. 2010 [81]; Zhang et al. 2014 [171] | 7,8-Dihydroxyflavone | TrkB receptor | Increase levels of BDNF |
| Braun et al. 2014 [13]; Ginovart et al. 1996 [51]; Labatut et al. 1988 [77]; Takeda et al. 1984 [150]; Zyzek et al. 1990 [176] | Vindeburnol | Unknown | Increase NE turnover |
| Charalampopoulos et al. 2005 [22]; Singh et al. 2012 [142]; Sun et al. 2012 [144]; Wang et al. 2010 [166] | Allopregnanolone | GABAA receptor | Increase levels of NE |
| Torres-Sanchez et al. 2018 [158] | Deep brain stimulation | Multiple brain regions | Increase NE release |
| Chang et al. 2018 [21]; Follesa et al. 2007 [42] | Vagus nerve stimulation | Vagus nerve | Increase levels of NE and BDNF |
| Rorabaugh et al. 2017 [129] | Chemogenetics | LC neurons | Rescue impaired reversal learning |

LC locus coeruleus, NE norepinephrine, BDNF brain-derived neurotrophic factor, GABA γ-aminobutyric acid
Brain stimulation technologies

Pilot studies have suggested that brain stimulation technologies, which are used to modulate cognitive function in neuropsychiatric disorders, could help ameliorate cognitive deficits in AD.

In animal studies, deep brain stimulation was reported to increase NE release in the area of stimulation [158]. Deep brain stimulation of the ventromedial prefrontal cortex was shown to increase tyrosine hydroxylase expression in the LC, as well as increase LC neuron activity [158]. Additionally, vagus nerve stimulation was shown to lessen cognitive deficits in AD dementia patients [21]. In a rat model, acute vagus nerve stimulation was shown to increase NE concentration, as well as increase BDNF expression in the hippocampus and cerebral cortex [42]. As it projects to the LC, it is possible that stimulation of the vagus nerve could directly activate the LC, leading to increased NE at projection sites. In addition, stimulation of the LC using chemogenetic approaches could also be an important therapeutic strategy in AD. In a TgF344-AD rat model, impaired reversal learning was rescued by chemogenetic LC activation [129]. Thus, brain stimulation technologies could be used as essential therapeutic strategies in alleviating the cognitive effects of LC–NE deinnervation in AD.

Conclusion

In conclusion, research has shown that LC degeneration and subsequent loss of the noradrenergic system play a major role in AD pathogenesis. Lack of noradrenergic innervation leads to a multitude of effects, including neurotoxic inflammation, increased neuropathologic burden and vast neuronal death, especially in the cortical projection areas of the LC. This de-innervation can enhance the progression of cognitive deficits and memory impairment in AD. LC signal intensity, as assessed by MRI, could be used as an early biomarker for AD. Furthermore, the implementation of therapeutics such as 1-DOPS and brain stimulation technologies could alleviate the neuropathologic processes and symptoms of this disease. Since the LC–NE system is a key player in modulating neuroprotection and regulating neuroinflammation, and its degeneration plays an essential role in the development of AD, a better understanding of the mechanistic link between LC loss and onset of AD could be vital for the diagnosis and therapeutic efforts against disease progression.

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Compliance with ethical standards

Conflict of interest Ms. Matchett, Dr. Grinberg, and Dr. Theofilas report no disclosures. Dr. Murray served as a paid consultant for AVID Radiopharmaceuticals.

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