B7 Family Members in Lymphoma: Promising Novel Targets for Tumor Immunotherapy?

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T cells play a vital role in the immune responses against tumors. Costimulatory or coinhibitory molecules regulate T cell activation. Immune checkpoint inhibitors, such as programmed cell death protein 1 (PD-1) and programmed death ligand 1 (PD-L1) have shown remarkable benefits in patients with various tumor, but few patients have displayed significant immune responses against tumors after PD-1/PD-L1 immunotherapy and many have been completely unresponsive. Thus, researchers must explore novel immune checkpoints that trigger durable antitumor responses and improve clinical outcomes. In this regard, other B7 family checkpoint molecules have been identified, namely PD-L2, B7-H2, B7-H3, B7-H4 and B7-H6. The aim of the present article was to address the expression, clinical significance and roles of B7 family molecules in lymphoma, as well as in T and NK cell-mediated tumor immunity. B7 family checkpoints may offer novel and immunotherapeutic strategies for patients with lymphoma.

Keywords: lymphoma, PD-L1, PD-L2, B7-H2, B7-H3, B7-H4, B7-H6

INTRODUCTION

T cells play important roles in antitumor immunity, and their dysfunction results in immune evasion (1). It is unclear how tumor interact with the immune system. Immunotherapeutics that target checkpoints have achieved remarkable clinical responses in tumor treatment. However, many patients remain unresponsive to such therapies, suggesting that there are other mechanisms of T cell exhaustion (2). Thus, researchers must investigate novel co-inhibitory molecules for

Abbreviations: ADC, antibody-drug conjugate; B7-H3×4-BB bispecific antibody targeting on B7-H3 and 4-BB; B7-H6Bi-Ab, bispecific antibody anti-CD3 and anti-B7-H6; BiKE, bispecific killer cell engager; BiTEs, bispecific T cell engagers; CAR, chimeric antigen receptor; cHL, classic Hodgkin lymphoma; CR, complete response; CTLA-4, cytotoxic T lymphocyte antigen 4; DLBCL, diffuse large B cell lymphoma; EBV, Epstein-Barr virus; ENKTL, extranodal NK/T cell lymphoma; FL, follicular lymphoma; GVHD, graft-versus-host disease; HDAC, histone deacetylase; ICOS, inducible T-cell costimulator; JAK, Janus Kinases; LMP1, Latent membrane protein-1; mAb, monoclonal antibody; MCL, mantle cell lymphoma; MM, multiple myeloma; MRD, minimal residual disease; NHL, non Hodgkin lymphoma; NK, natural killer; ORR, overall response rate; PD-1, programmed cell death protein-1; PD-L1, programmed death ligand-1; PD-L2, programmed death ligand-2; PMBCL, primary mediastinal large B-cell lymphoma; scFvs, single chain fragment variables; SCT, stem cell transplantation; STAT, signal transducer and activator of transcription; TAM, tumor associated macrophages; TLT-2, myeloid cell like transcript 2; TNF, tumor necrosis factor.
immunotherapy. Combining new target molecules with present immunotherapies may offer novel strategies and improve clinical responses.

B7 family members have received attention because they are expressed on T cells in cases of immune evasion and tumorigenesis. To date, ten B7 family molecules have been identified: B7-1 (CD80), B7-2 (CD86), B7-H1 (PD-L1, CD274), B7-DC (CD273 or PD-L2), B7-H2 (ICOSLG, CD275), B7-H3 (CD276), B7-H4 (B7S1, B7x or VTCN1), B7-H5 (VISTA, GI24, or PD-1H), B7-H6 (NCR3LG1) and B7-H7 (HHLA2) (3). Using the TCGA and GTEx databases, we investigated the mRNA expression levels of B7 family proteins in lymphoma. As shown in the heatmap showed (Figure 1), all B7 family members reported in the literature other than B7-H5 were more highly expressed in diffuse large B cell lymphoma (DLBCL), suggesting that these molecules may play vital roles in lymphoma immunity, explaining the poor effect of PD-1/PD-L1 therapy.

The expression of B7 family molecules were regulated by various mechanisms and play important roles in lymphoma proliferation, migration, evasion, chemoresistance and immune evasion. Blockade of B7 family molecules released T/NK cells from the inhibitory effects and restores antitumor immunity via promoting T/NK cell activation, proliferation and cytotoxicity, and suppressing inhibitory immune cells and molecules. Immunotherapies targeting B7 family members include monoclonal antibody (mAb), inhibitors, antibody-drug conjugates (ADCs), single chain fragment variables (scFvs), antibody-dependent cell-mediated cytotoxicity (ADCC), bispecific T cell engagers (BiTEs) and chimeric antigen receptor (CAR) T cell therapy (4).

The present review summarizes the research involving B7 family members in lymphoma, namely PD-L1, PD-L2, B7-H2, B7-H3, B7-H4 and B7-H6. The surface expression of these molecules in lymphoma is shown in Figure 3. Further exploration of these molecules is needed to develop effective immunotherapies, either as monotherapy or in combination with other antibodies.

2 PD-L1

The PD-1/PD-L1 axis is a vital checkpoint in tumor progression and immune evasion. The binding of PD-1 to PD-L1 resulted in T cell anergy, exhaustion, apoptosis, and reduced cytotoxicity (5). The drug mechanisms of anti-PD-1/PD-L1 antibodies are similar. They
destroy the immunosuppressive microenvironment and reactivate T cells, allowing them to recognize and kill tumor cells by blocking the binding of PD-L1 on tumor cells to PD-1 on T cells (6).

Anti-PD-1 antibodies have been approved for use in various solid tumors and lymphomas (7). A multicenter, single-arm, phase II trial of sintilimab to treat relapsed or refractory classical Hodgkin’s lymphoma (cHL) was carried out in China and showed that the overall response rate (ORR) was 80.4% (5, 8). Single sintilimab therapy also revealed an anti-tumor effect in extranodal natural killer (NK)/T cell lymphoma (ENKTL) in a phase II trial (5). The ORR was 67.9% and the disease control rate was 85.7% (5). Sintilimab combined with decitabine and the histone deacetylase inhibitor chidamide resulted in partial remission in DLBCL (9). Phase II studies have revealed that the ORR after treatment with either nivolumab or pembrolizumab was 66.3% and 69.0%, respectively, in patients with relapsed or refractory cHL (5, 10, 11). The ORR of pembrolizumab in patients with ENKTL was 78.6% (12). A recent single-sintilimab study reported that geptanolimab showed promising efficacy and manageable toxicity in patients with relapsed/refractory peripheral T cell lymphoma (13). Interestingly, patients with PD-L1 expression > 50% obtained more benefit from geptanolimab treatment, with an ORR of 53.3% and a median progression-free survival of 6.2 months (13). The correlations between PD-L1 expression and response to anti-PD-1 antibodies should to be further investigated in future clinical trials. We have summarized the finished clinical trials in Table 1 and ongoing ones in Table 2.

The efficacy and safety of anti-PD-L1 antibodies in lymphoma patients have also been assessed in clinical trials, involving patients with lymphoma. At present, the following anti-PD-L1 antibodies are used in clinical practice: avelumab, durvalumab and atezolizumab. The finished clinical trials are summarized in Table 1 and ongoing ones in Table 2. Avelumab is a fully human IgG1 mAb that selectively blocks PD-L1 and enhances anti-tumor T-cell activity (7). A phase I study of avelumab demonstrated that ORR and complete response (CR) were 54.8% and 63%, respectively, in patients with relaps/refractory cHL who had suffered progression following stem cell transplantation (SCT) or SCT-ineligible (7). A phase II trial demonstrated that the CR of avelumab was 24% and that the ORR was 38% in patients with relapsed or refractory ENKTL (14). The response to avelumab was strongly correlated with PD-L1 expression in tumor tissues (14). A phase Ib/II, multicenter, open-label study of ibrutinib plus durvalumab in relapsed/refractory follicular lymphoma (FL) or DLBCL showed ORR values of 25% among all patients, 26% among patients with FL, and 13% among patients with germinal center B-cell DLBCL (15). A multicenter open-label, phase I-II trial of patients with solid tumors or lymphomas observed that atezolizumab was well tolerated with generally comparable exposure across populations (16). A phase Ib study involving patients with PD-L1 + large B-cell lymphoma demonstrated that the ORR of CD19-PD-1/ CD28-CAR-T-cell therapy was 58.8%, and that the CR was 41.2%. No severe cytokine release syndrome or neurologic toxicity was reported in that study (17).

### Table 1: The finished clinical trials targeting on B7 family molecules in lymphoma.

| Targets | Drug | Trial ID | Phase | N | Diagnosis | Response | Ref |
|---------|------|---------|-------|---|-----------|----------|----|
| PD-L1   | Avelumab | NCT03439501 | II | 21 | ENKTL | CR 24%, ORR 38% | (14) |
| PD-L1   | Durvalumab | NCT02401048 | Ib/II | 61 | r/r DLBCL, r/r FL | ORR 25% | (15) |
| PD-L1   | Atezolizumab | NCT02541604 | III | 90 | r/r solid tumors and lymphoma | ORR 5% | (16) |
| PD-1    | CD19-PD-1/CD28-CAR T cell | NCT03258047 | Ib | 17 | PD-L1+ LBL | CR 41.2%, ORR 58.8% | (17) |
| PD-1    | Nivolumab | NCT01592370 | I | 23 | r/r HL | ORR 87%, CR 17%, PFS rate 86% | (18) |
| PD-1    | Nivolumab | NCT02181736 | II | 80 | chL | ORR 66.3%, CR 8.8%, PR 57.5%, PFS 10 m | (19) |
| PD-1    | Nivolumab | NCT02029847 | III | 144 | DLBCL, FL | CR 61%, PR 14%, SD 3% | (19) |
| PD-L1   | Pembrolizumab | NCT01963692 | Ib | 31 | r/r chL | ORR 65%, CR 16%, | (20) |
| PD-L1   | Pembrolizumab | NCT01953692 | Ib | 21 | r/r PMBL | ORR 48%, CR 33% | (21) |
| PD-1    | Pembrolizumab | NCT02576990 | II | 53 | r/r PMBL | ORR 45%, CR 13% | (21) |
| PD-1    | Pembrolizumab | NCT02332980 | II | 9 | r/r DLBCL | ORR 44% | (22) |
| PD-1    | Pembrolizumab | NCT02453954 | II | 210 | r/r chL | ORR 71.9%, CR 27.6%, PR 44.3%, PFS 13.7 m, DOR 16.6 m, 3 years OS 86.4% | (23) |
| PD-1    | Pembrolizumab | NCT02541565 | I | 33 | DLBCL, FL | ORR 90%, CR 77%, PFS 83% | (24) |
| PD-1    | Pembrolizumab | NCT03150329 | I | 30 | DLBCL, PMBL, FL, chL | ORR 30%, CR 30%, DOR 6 m, PFS 59% | (25) |
| PD-1    | Pembrolizumab+ Vorinostat | NCT03155042 | II | 75 | chL | CR 28%, PR 48% | (26) |
| PD-1    | Nivolumab | NCT03209973 | II | 70 | r/r chL | PR 87.6%, CR 62.9%, ORR 87.1%, CR 62.9%, 9 m PFS=74.5% | (27) |
| PD-1    | Nivolumab Brentuximab | NCT02581631 | I, II | 30 | PMBL | ORR 73%, CR 37% | (28) |
| PD-1    | Ipilimumab | NCT01822509 | I | 28 | hematologic cancer | ORR 32%, PFS 1 year | (29) |
| PD-1    | Nivolumab | NCT03502629 | II | 102 | r/r PTCL | ORR 40.4%, CR 14.6%, PR 25.8%, DOR 11.4 m | (13) |

**ORR**: overall response rate; **CR**: complete response; **DOR**: Duration of overall response; **PFS**: progression-free survival; **SD**: stable disease.
Currently, clinical trials targeting B7 family members are ongoing in patients with lymphoma. Abnormality in chromosome 9p24.1, which encodes PD-L1 and PD-L2 protein and Janus kinase 2, is the main cause of PD-L1 and PD-L2 overexpression (34). Chromosomal rearrangement of PD-L2 is associated with abnormal overexpression in malignant cells of mycosis fungoides (35). BCL6 is a key negative regulator of PD-L1 and PD-L2 in germinal center B cells. Moreover, microRNA-BHRF1-2-5p plays a regulatory role in LMP1 driven PD-L1 and PD-L2 amplification (39). So further identification of microRNAs that target immune checkpoints allow RNA-based therapy. The regulatory mechanisms of PD-L1 and PD-L2 expression and their function are summarized in Figure 2.

### 3 PD-L2

Programmed death ligand-2 (PD-L2), is a PD-1 receptor. It is mainly expressed in dendritic cells, macrophages, mast cells and B cells, as well as in hematological malignancies, including multiple myeloma (MM), acute leukemia and chronic lymphocytic leukemia (33). However, it has little or no significant effect on prognosis in these diseases (33).

It is reported that PD-L2 was expressed on the surface of malignant cells in 65-100% patients with cHL and 54% of patients with nodular lymphocyte predominant Hodgkin’s lymphoma. Abnormality in chromosome 9p24.1, which encodes PD-L1 and PD-L2 protein and Janus kinase 2, is the main cause of PD-L1 and PD-L2 overexpression (34). Chromosomal rearrangement of PD-L2 is associated with abnormal overexpression in malignant cells of mycosis fungoides (35). BCL6 is a key negative regulator of PD-L1 and PD-L2 in germinal center B cells. It directly binds to the promoter region of PD-L1 and intron 2 of PD-L2 to inhibit its transcription and maintain the size of follicular T cells during the development of germinal center (36). The IL-27/STAT3 signaling pathway induces PD-L1 and PD-L2 expression in in infiltrating macrophages of lymphoma (37). PD-L1 and PD-L2 are highly expressed in Epstein-Barr virus (EBV)-positive lymphomas, including DLBCL, extranodal NK/T-cell lymphoma, aggressive NK cell leukemia and T-cell lymphoproliferative diseases (38). Latent membrane protein-1 (LMP1) induced the expression of PD-L1 and PD-L2. Cristino et al. reported that when LMP1 was activated, PD-L1 and PD-L2 expression was significantly increased during the transformation of B cells from the late germinal center to early and late activated B cells. Moreover, microRNA-BHRF1-2-5p plays a regulatory role in LMP1 driven PD-L1 and PD-L2 amplification (39). So further identification of microRNAs that target immune checkpoints allow RNA-based therapy. The regulatory mechanisms of PD-L1 and PD-L2 expression and their function are summarized in Figure 2.

One recent study reported that high levels of plasma-soluble PD-L1 and signal transducer and activator of transcription (STAT) 3 were related to worse progression-free survival and overall survival in patients with DLBCL (30). Another study showed that vincristine induced PD-L1 expression via p-STAT3 and augmented the efficacy of PD-L1 blockade therapy by activating effector T cells and increasing the antitumor immune response in DLBCL (31). The expression levels of PD-L1 on monocytes are increased in patients with NK/T-cell lymphoma and constitute a novel predictor of prognosis (32). More clinical trials involving anti-PD-1/PD-L1 antibodies are currently ongoing in patients with lymphoma.

### Table 2 | The ongoing clinical trials targeting on B7 family molecules.

| Targets     | Drug                      | Disease          | Phase | Status                  | Trial ID               |
|-------------|---------------------------|------------------|-------|-------------------------|------------------------|
| PD-L1       | Durvalumab                | NKTCL            | II    | Not yet recruiting      | NCT03357224            |
| PD-L1       | Durvalumab                | PCNSL            | I     | Not yet recruiting      | NCT03817666            |
| PD-L1       | Acabrutinib               | PCNSL            | I     | Not yet recruiting      | NCT0460653             |
| PD-L1       | Atezolizumab              | PCNSL            | I     | Not yet recruiting      | NCT04462328            |
| PD-L1       | Atezolizumab              | DLBCL            | Recruiting |                           | NCT03850028           |
| PD-L1       | Rituximab                 | r/r DLBCL        | II    | Active, not recruiting  | NCT03422523            |
| PD-L1       | Oxaliplatin               | CTCL, SS         | II    | Active, not recruiting  | NCT03357224            |
| PD-L1       | Atezolizumab              | r/r ENKTL        | II    | Active, not recruiting  | NCT03439601            |
| PD-L1       | Avelumab                  | Advanced HL      | II    | Recruiting              | NCT03817666            |
| PD-L1       | Avelumab                  | PTCL             | II    | Active, not recruiting  | NCT04185028            |
| B7-H3       | B7-H3 CAR T               | DIPG, DMG, r/r CNS tumors | I     | Recruiting              | NCT04077866            |
| B7-H3       | B7-H3 CAR T               | r/r Glioblastoma | II    | Recruiting              | NCT04483778            |
| B7-H3       | B7-H3 CAR T               | r/r solid tumors | I     | Recruiting              | NCT04432649            |
| B7-H3       | 4SCAR-276                 | solid tumors     | II    | Recruiting              | NCT04470068            |
| B7-H3       | B7-H3 CAR T, Fludarabine, Cyclophosphamide | Epithelial Ovarian Cancer | Not recruiting | NCT02923180          |
| B7-H3       | MG0218                    | Prostate Cancer  | II    | Active, not recruiting  | NCT02923180            |
| B7-H3       | Enoblituzumab, Retifanlimab, Tebotelimab | Head and neck caner | Not recruiting | NCT04834825          |
| B7-H3       | MGC018                    | Advanced solid tumors | I     | Recruiting              | NCT03729596            |
| B7-H3       | MGD009                    | Advanced solid tumors | I     | Active, not recruiting  | NCT02923180            |
| B7-H4       | FPA150                    | Advanced solid tumors | I     | Active, not recruiting  | NCT03406940            |
| B7-H6       | BI 765049                 | Advanced solid tumors | I     | Not recruiting          | NCT04752215            |
| B7-H6       | BI 754091                 | Advanced solid tumors | I     | Not recruiting          | NCT04752215            |

CNS, central nervous system; CTCL, cutaneous T-cell lymphomas; DIPG, diffuse intrinsic pontine glioma; DMG, diffuse midline glioma; DLBCL, diffuse large B-cell lymphoma; ENKTL, extranodal natural killer/T-cell lymphomas; HL, Hodgkin Lymphomas; PCNSL, primary central nervous system lymphoma; PTCL, Peripheral T-Cell Lymphomas; r/r, relapsed or refractory; SCNSL, secondary central nervous system lymphoma; SS, Sezary syndrome.
and multivariate analysis has shown that the differential expression of PD-1 and PD-L2 genes in Th-1/Th-2 status guarantees the prognosis of primary central nervous system lymphoma (42). PD-L2 RNA in situ hybridization was a sensitive, specific and practical marker to identify primary mediastinal large B-cell lymphoma (PMBCL) (43).

The expression of PD-L2 is correlated with favorable prognosis in patients with DLBCL. Moreover, the high expression levels of PD-L2 are related to low expression of PD-1 and upregulation of CD80 in CD4/CD8 T cells. In addition, fluorescent in situ hybridization has shown that changes in the PD-L2 gene were related to the survival rate and gene expression profile of patients (44). However, other studies have reported that PD-L2 expression is associated with poor disease-free survival and overall survival in DLBCL (45). DLBCL with Janus kinases (JAK) 2/PD-L2 amplification shows PMBCL like replication number changes and poor prognosis (46). PD-L2 replication or amplification has been found in the malignant B cells of 64% of patients with T-cell/histiocyte rich large B-cell lymphoma (47). High expression levels of PD-L2 are associated with poor prognosis in FL, while low expression levels are positively correlated with 24-month disease-free survival (48). PD-L2 must be further explored in the future, especially in clinical trials.

4 B7-H2

B7-H2 binds to inducible T-cell costimulator (ICOS) and augments Th1 and Th2 function by inducing effector cytokine secretion (49). Few studies have evaluated B7-H2 in hematological tumors, although one found that it is highly expressed in FL B cells that it induced the generation of ICOS+ regulatory T cells, inhibiting the function of conventional T cells (50). In one murine lymphoma model, miR21 is a serum oncogenic biomarker. miR21 indicated that the sensitivity of B cell lymphoma sensitivity to ABT-199 through the ICOS and ICOS ligand signal involved interaction between Treg cells and endothelial cells (51).

5 B7-H3

B7-H3 is extensively expressed in various tumors, tumor-infiltrating dendritic cells, and macrophages (52). The exact receptor of B7-H3 remains unclear. Previous studies have reported that myeloid cell-like transcript 2 (TLT-2) binds to B7-H3. However, others found that B7-H3 and TLT2 did not bind to each other (53, 54). A circulating soluble isoform of B7-H3 also exists in serum and other body fluids (55).
Most studies have demonstrated that B7-H3 inhibited T cell function and promoted tumor progression, and one reported that B7-H3 is overexpressed in patients with mantle cell lymphoma (MCL) and cell lines, and that miR-506 negatively regulates the expression of B7-H3, inhibiting cell growth, invasion and migration in MCL. These effects were reversed by the restoration of B7-H3 expression (56). B7-H3 silencing by RNAi suppressed tumor progression and augmented chemosensitivity to chemotherapeutic drugs in U937 cells and MCL cells (57, 58). B7-H3 was correlated with progression-free survival and overall survival time of patients with MM (59, 60). LncRNA NEAT1 sponged miR-214 to induce M2 macrophage polarization by regulating B7-H3, and promoted MM progression through JAK2/STAT3 signaling pathway (59). B7-H3 promoted MM cell survival and growth via ROS/Src/c-Cbl signaling pathway (60).

The B7-H3 checkpoint may serve as a promising and novel target for immunotherapy against tumors. B7-H3 inhibition resulted in reduced growth of multiple tumors and enhanced antitumor immunity via NK and CD8+ T cells (52). Combining blockades of B7-H3 and PD-1 led to further augmented therapeutic effects on late-stage tumors (52). B7-H3-targeted CAR-T cells showed significant antitumor activity against hematologic malignancies and solid tumors (61). B7-H3 was highly and homogeneously expressed in extranodal nasal NK/T cell lymphoma cell lines. A new anti-B7-H3/CD3 BiTE antibody and B7-H3-directed CAR-T cells have been constructed. They effectively target and kill NKTCL cells and inhibited the growth of tumors (62). A B7-H3-redirected CAR based on scFvs from mAb 376.96 demonstrated strong cytotoxicity and cytokine production against target anaplastic large cell lymphoma cells in vitro and promptly eradicated tumor cells in mouse xenografts. In addition, B7-H3 CAR-T cells show growth capacity and a memory phenotype after stimulation using repeated antigen (63). A bispecific antibody targeting B7-H3 and 4-1BB (B7-H3×4-1BB) has been developed. B7-H3×4-1BB showed antitumor activity in mice and promoted CD8 T cell proliferation and cytokine secretion. B7-H3×4-1BB combined with PD-1 blockade synergistically suppressed tumor growth and increased terminally differentiated CD8 T cells (64). B7-H3 CAR-T cells effectively suppressed tumor growth, both in vitro and in vivo. B7-H3 CAR and B7-H3/CD16 bispecific killer cell engager (BiKE) have also been generated. B7-H3/CD16 BiKE has been shown to trigger NK cell activity via CD16 signaling, enhanced NK cell activation and improved antitumor efficacy in vitro and in vivo (65). The regulatory mechanisms and therapies targeting B7-H3 are summarized in Figure 3. The ongoing clinical trials are summarized in Table 2.

Thus, B7-H3 inhibitors may be an effective and safe therapeutic agent against tumors as monotherapy and in combination with other therapeutic agents. Further studies need to be carried out in preclinical and clinical studies.

**FIGURE 3** The regulatory mechanisms and therapies targeting B7-H3.
6 B7-H4

B7-H4 negatively modulates T cell immunity and promotes tumor progression (66). The receptor for B7-H4 has not been identified (67). It has been reported that B7-H4 is induced by IL-6, IL-10 and tumor associated macrophages (TAM) and that it protects NHL cells from T cell-mediated killing by secreting IL-6 and IL-10 (68). In another study, B7-H4 augmented the differentiation of mouse leukemia-initiating cells by deleting the phosphatase and tensin homolog in the Akt/RCOR2/RUNX1 signaling pathway (69). In cancer cells, B7-H4 is upregulated by hypoxia via hypoxia-inducible factor-1α and promotes tumor cell growth (70).

It has been demonstrated that B7-H4 is overexpressed in EBV-positive DLBCL and that it inhibits apoptosis via ERK1/2 and Akt signaling pathways (71). Moreover, B7-H4 appears to play a critical role in prognosis while PD-L1 expression weakened (72). One investigation found that B7-H4 engagement in EBV-positive lymphomas inhibited tumor cell proliferation and regulated cell cycle arrest at the G0-G1 phase via down-regulation of the Akt signaling pathway (73). Thus, B7-H4 presents as a new potential target for EBV-positive lymphoma immunotherapy. In another study, B7-H4 overexpression in myeloid cells from human cancers was related to CD8+ T cell dysfunction (74). The combination of B7-H4 and PD-1 blockade demonstrated synergic effects and enhanced anti-tumor immune responses (74). Therefore, targeting the B7-H4 co-inhibitory pathway may augment the therapeutic effect of current anti-PD-1 therapy to treat cancers.

7 B7-H6

B7-H6, which is selectively expressed on the tumor cell surface, is a ligand for NKp30, which may be a promising target for novel cancer immunotherapy strategies and has been investigated in CAR-T therapy and novel immunoligands (77). NKp30 induces efficient NK cell-mediated antitumor immune responses.
triggered by B7-H6 (78). In a recent study, incorporation of affinity-matured B7-H6 into NKp30 therapy enhanced NK cell-mediated tumor cell killing and significantly increased proinflammatory cytokine release of bispecific immunoligands (79).

The expression and regulation of B7-H6 have been explored. In one study, B7-H6 in its soluble and in soluble forms, was induced at the surface of proinflammatory monocytes and neutrophils by ligands of toll-like receptors and proinflammatory cytokines including interleukin-1β and tumor necrosis factor (TNF) α (80). In another investigation, metalloproteases induced B7-H6 release from the tumor cell surface and treatment with metalloprotease inhibitors resulted in both increased surface levels of B7-H6 and augmented NK cell-mediated tumor cell lysis (81). Promoter analyses demonstrated that the proto-oncogene Myc induced B7-H6 expression in tumor cells (78). In one study, suppression of c-Myc or N-Myc markedly reduced the expression levels of B7-H6, and both mRNA and surface protein expression of B7-H6 was reduced by histone deacetylase (HDAC) inhibitors and small interfering RNA-mediated knockdown of HDAC 2 or 3 (82). In another study, B7-H6 downregulation was related to reduced B7-H6 reporter activity and histone acetylation at the B7-H6 promoter (82). Treatment with cisplatin and 5-fluorouracil chemotherapy, radiotherapy, non-lethal heat shock, and TNF-α therapy-induced B7-H6 expression in tumors and enhanced tumor sensitivity to NK cell cytotoxicity (83).

In primary lymphoma tissues, B7-H6 mRNA levels are increased and related to HDAC3 expression (82). HDAC inhibitors reduces B7-H6 expression and NKp30-dependent efficient functions of NK cells. The mRNA levels of c-Myc are significantly correlated with B7-H6 expression, and inhibition of c-Myc damaged NKp30-mediated degranulation of NK cells (78). In B cell NHL, B7-H6 knockdown suppressed tumor progression and enhanced chemosensitivity. Downstream target investigation has indicated that STAT3 pathway is involved in B7-H6 knockdown-mediated antitumor immunity (84). B7-H6 is overexpressed in DLBCL, T-lymphoblastic lymphoma and lymph node reactive hyperplasia tissues promoting cell growth, migration, and invasion through the Ras/MEK/ERK signaling pathway (85).

The combination of recombinant immunoligands ULBP2:7D8 and B7-H6:7D8 increases NK cell-mediated ADCC in lymphoma (86). Moreover, bispecific antibody anti-CD3 and anti-B7-H6 (B7-H6Bi-Ab) armed T cells showed significant cytotoxicity induction in B7-H6 positive hematological tumor cells via the production of granzyme B and perforin (1). In addition, B7-H6Bi-Ab armed T cells secreted more T cell-derived cytokines and expressed much higher level of the activation marker CD69 (1). Another study demonstrated that B7H6-specific BiTEs directed T cells to mediate cytolysis and IFN-γ production against tumors (87). In vivo, B7-H6-specific BiTE significantly increases the survival of lymphoma-bearing mice via perforin and IFN-γ secretion. Moreover, BiTE protein reduces tumor burden in melanoma and ovarian cancer-bearing mice. Therefore, combining therapeutic antibodies may provide a promising insight to further enhance the efficacy of antibody therapy. This strategy

![FIGURE 5](image-url)
may be especially encouraging for eradicating minimal residual disease cells after transplantation. The regulatory mechanisms and therapies targeting B7-H36 are summarized in Figure 5. The ongoing clinical trials are summarized in Table 2. Taken together, B7-H6 may be a promising immunotherapy target for hematological and solid tumors. Further explorations of B7-H6 targeted immunotherapy should be carried out in the preclinical and clinical studies.

CONCLUSION

In summary, B7 family members may provide novel strategies to inhibit or kill tumors by triggering antitumor immune responses. Blocking the PD-L1/PD-1 pathway has generated therapeutic success in human tumors. However, PD-L1/PD-1 have a low response rate. Therefore, researchers must combine novel checkpoint inhibitors with PD-L1/PD-1 inhibitors, or used them in monotherapy. The B7 family member pathways represent novel immunosuppressive mechanisms in tumor immunity, as well as a potential target for immunotherapy. Additional studies, involving immunoregulatory mechanisms and clinical trials are needed for further exploration. In the future, immunotherapy based on combined B7 family members may represent a promising strategy for treating hematologic malignancies.

AUTHOR CONTRIBUTIONS

WZ and YQ designed the study and wrote the manuscript. YF and XX participated in data acquisition and statistics analysis. LW and ZC edited and revised the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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