ULTRASTRUCTURAL CHARACTERISTICS OF SPERMIOGENESIS IN RHYNCHOPHORUS FERRUGINEUS (COLEOPTERA: CURCULIONIDAE)

ABDULLAH M. ALZAHRANI1,*, SALAHELDIN A. ABDELSALAM1,2, OMAR M. ELMENSHAWY1,3 AND ASHRAF M. ABDEL-MONEIM1,4

1Department of Biological Sciences, Faculty of Science, King Faisal University, Hofouf, Saudi Arabia
2Zoology and Entomology Department, Faculty of Science, Assiut University, Egypt
3Department of Zoology, Faculty of Science, Al-Azhar University, Egypt
4Department of Zoology, Faculty of Science, University of Alexandria, Egypt

*Corresponding author; E-mail: aalzahra@kfup.edu.sa

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ABSTRACT

The red palm weevil (RPW), Rhynchophorus ferrugineus Olivier (Coleoptera: Curculionidae), is a pest that is rapidly spreading across the globe. Here, the ultrastructure of R. ferrugineus spermatogenesis and sperm are described. The histology of the testis, sperm ultrastructure, and spermiogenesis were investigated using light and transmission electron microscopy. The differentiation of spermatids was observed to occur within spermiogenetic cysts. Inside each cyst, the spermatids were at the same stage of maturation. During early stages, mitochondria aggregated, fused, and elongated beside the growing flagellar axoneme, while the proacrosome transformed into a triple-layered acrosome, with a perforatorium, acrosomal vesicle, and extra-acrosomal layer. The centriolar adjunct was present in early spermatids but was absent from later spermatid stages and sperm. The sperm’s tail displayed a typical axoneme with a 9 + 9 + 2 microtubule arrangement, 2 mitochondrial derivatives of unequal size, and 2 accessory bodies. A small number of sperms exhibited twin or multiple tails due to membrane fusion. Our results support systematic relationships within the family Curculionidae.

Key Words: axoneme, acrosome, spermatogenesis, microtubules, nebenkern, pterygote spermatozoan, ultrastructure, red palm weevil

RESUMEN

El picudo rojo (RPW), Rhynchophorus ferrugineus Olivier (Coleoptera: Curculionidae), es una plaga que se está extendiendo rápidamente en todo el mundo. Se describe la ultraestructura de la espermatogénesis y los espermatocistos de R. ferrugineus. Se investigó la histología de los testículos, ultraestructura de espermatocistos y espermatogénesis, con microscopía óptica y de transmisión electrónica. Se observó que la diferenciación de las espermatidas ocurre dentro de los quistes spermiogenéticos. Dentro de cada quiste, las espermatidas se encontraban en la misma fase de maduración. Durante las primeras etapas, los mitocondrias se agregaron, juntaron, y alargaron al lado del axonema flagelar creciente, mientras que el proacrosoma se transformó en un acrosoma de tres capas, con un perforatorium, vesícula acrosomal y la capa extra-acrosomal. El complemento centriolar estuvo presente en las primeras espermatidas, pero estuvo ausente en las etapas posteriores de las espermatóndidas y espermatocistos. La cola del espermatozoiode pareció como un axonema típico con un 9 + 9 + 2 disposición de microtúbulos, 2 derivados mitocondriales de tamaño desigual y 2 cuerpos auxiliares. Un pequeño número de espermatocistos exhibió colas dobles o múltiples debido a la fusión de la membrana. Nuestros resultados apoyan las relaciones sistemáticas dentro de la familia Curculionidae.

Palabras Clave: axonema, acrosoma, espermatogénesis, microtúbulos, nebenkern, espermatocistos, espermatocistos pterygotes, ultraestructura, picudo rojo
The red palm weevil (RPW), *Rhynchophorus ferrugineus* Olivier (Coleoptera: Curculionidae), is one of the most destructive pests of coconuts and date palm trees in the world (Faleiro et al. 2003). Its larvae feed on the interior tissues of the palm trunk, destroying the tree. Currently, *R. ferrugineus* is widely distributed in Oceania, the Near, Middle, and Far East (Li et al. 2009), and Europe and America (Dembilio & Jacas 2012).

*Rhynchophorus ferrugineus* spermatogenesis was briefly described by Bartlett (1983). The spermatozoa of most Coleoptera, especially the Curculionidae, display the classic structure, which is characterized by a head region formed by a 3-layered acrosome and nucleus. Two enormous accessory bodies and 2 large and crystallized mitochondrial derivatives support the tail region. A 9 + 9 + 2 axoneme pattern is the most common pattern within the Curculionidae (Gassner et al. 1975; Burrini et al. 1988; Name et al. 2007; Werner & Simmons 2011). Only the Rynchiitidae exhibit an axoneme pattern of 9 + 9 + 0 (Burrini et al. 1988). The number of curculionid species is overwhelming. Therefore, it is difficult to construct systematic lines based on phylogenetic data. However, accumulation of spermatological information on Curculionid species can aid in understanding spermatogenesis in these destructive pests and help to resolve taxonomic and phylogenetic discrepancies among them.

The aim of the current study is to provide a detailed understanding of *R. ferrugineus* spermiogenesis through elucidation of the sperm ultrastructure using light and electron microscopy techniques and to compare it to published data on other related species.

**MATERIALS AND METHODS**

**Insect Culture**

The insects used in this study were obtained from infested palm trees in the Alahsa area of eastern Saudi Arabia. The insects were cultured in a rearing room at 25 ± 2 °C, 70 ± 5% RH and 12:12 h L:D. Sugarcane was used to feed the larvae, while the adults were reared on sugarcane pieces and cotton wicks saturated with a 10% sugar solution for feeding and egg laying.

**Light Microscopy**

Testes were dissected in Ringer’s physiological solution and fixed in a 10% formalin solution overnight. The specimens were washed, dehydrated, and cleared in xylene before being embedded in paraffin wax. Multiple serial sections, with a thickness of 5 μm, were obtained and stained with hematoxylin and eosin. The sections were observed with a Nikon 80i light microscope (Nikon, Tokyo, Japan).

Free spermatozoa were obtained by dissecting testes in phosphate buffer saline (pH 7.2) and fixed in 4% paraformaldehyde. Sperm smears were stained with Harris’s Haematoxylin and mounted in a drop of DPX.

**Transmission Electron Microscopy**

For electron microscopic examination, small pieces of testicular tissues were fixed at 4 °C in a 3% glutaraldehyde solution buffered with 0.1 M sodium cacodylate at pH 7.2, then post-fixed in 1% osmium tetroxide in the same buffer for 2 h, dehydrated in a graded series of ethanol, transferred to propylene oxide, and embedded in an Epon-Araldite mixture. Ultra-thin sections were obtained using diamond knives with a Leica EM UC6 ultramicrotome (Leica Co., Vienna, Austria). The samples were subsequently stained with 2% uranyl acetate and lead citrate prior to examination on a Jeol JEM 1011 transmission electron microscope (Jeol Ltd., Tokyo, Japan) at 80 kV.

**Image Analysis**

Quantitative analysis of sperm dimensions were detected using the freeware ImageJ v1.33 downloaded from the NIH website (http://rsb.info.nih.gov/ij).

The figures in this articled are also shown in the supplementary material for this article in Florida Entomologist 96(4) (December 2013) online at http://purl.fcla.edu/fcla/entomologist/browse.

**RESULTS**

*Rhynchophorus ferrugineus* males had 2 testes, which were subdivided into tubular testicular follicles, and each follicle was enclosed by a layer of epithelial cells (Fig. 1). The follicles were filled with cysts at various stages of development. The outer rim of the testis was occupied by zones containing spermatogonia, while zones containing spermatocytes and spermatids were situated internally. Numerous spermatozoa were observed in the lumen of deferent duct using light microscopy. The sperm cells of *R. ferrugineus* appeared linear and very thin with an overall length of 87.23 ± 14.12 μm (mean ± standard deviation, n = 9) (Fig. 2). The headpiece measured approximately 10.12 ± 1.19 μm in length (n = 14), comprising about 10% of the total sperm length.

Inside spermiogenetic cysts, all spermatids were in the same stage of maturation. During the early spermatid phase, the nucleus was similar to that of somatic cells, displaying dispersed chromatin with some electron-dense areas. The early spermatids contained many mitochondria, expanded Golgi complexes and some smooth
endoplasmic reticulum cisternae. The mitochondria were aggregated into 2 parts (Fig. 3), which were fused together, giving rise to 2 conspicuous nebenkerns in a juxtanuclear position (Fig. 4). An acrosome vesicle, derived from the Golgi complex, was observed near the nuclear envelope (Fig. 5). In this phase, an electron-dense material, the centriole adjunct, began to accumulate around the centriole in the region of flagellum implantation (Fig. 6). During the intermediary stages of spermatic maturation, there was a gradual condensation of nuclear chromatin, causing an increase in electron density. The structural reorganization of chromatin during nuclear condensation does not follow the same course in all insect species, and chromatin condensation was not uniform in *R. ferrugineus*. During some stages, the region near the nuclear envelope contained homogeneously condensed chromatin, while the central region was distinguished by the presence of chromatin fibers (Fig. 7). Thus, nuclear chromatin condensed gradually, and large blocks were associated with the nuclear envelope (Fig. 8). In mature spermatids, we observed a compact nucleus surrounded by a layer of manchette microtubules, and there were small spots of less electron-dense areas within the chromatin mass (Fig. 9). Simultaneous with chromatin condensation and nuclear and cellular elongation, the acrosomic complex covered the anterior of the nucleus, and development of the flagellum was initiated, with the centriolar adjunct disappearing in late spermatid stages (Fig. 10). The axoneme elongated from a centriole (Fig. 11), and 2 mitochondrial derivatives developed from the nebenkern toward the posterior end and exhibited cristae that were arranged perpendicular to the axoneme (Fig. 12). During the formation of the mitochondrial derivatives, these structures were separated from each other, and there were microtubules present in the surrounding cytoplasm (Fig. 13). These 2 structures showed different sizes, and the larger one exhibited a dense crystallloid in the mitochondrial matrix in the region opposite the axoneme (Fig. 14). The mitochondrial derivatives in the tail region were also associated with a well-developed agranular endoplasmic reticulum (ER). Laterally, on both sides of the axoneme, 2 electron-dense arches were observed. Some testicular cysts contained bi- or multi-flagellate spermatids with 2 or more axonemes. In the bi- or tri-flagellate varieties, 2 to 3 nebenkern derivatives were noted (Fig. 15).
The spermatozoon of *R. ferrugineus* had a homogenously dense nucleus (Fig. 16). The diameter of the nucleus was reduced progressively from 0.81 to 0.32 μm before its apex was reached. When examined via electron microscopy, the coneshaped acrosome was very short (about 1.24 μm in length) and triple layered; the perforatorium-like inner core was surrounded by electron-dense acrosomal contents, which was itself covered by an extra-acrosomal layer (Fig. 17). The transition between the basal part of the perforatorium and the anterior part of the nucleus was visible (approximately 25 nm). In cross-sections, the acrosome displayed profiles that varied from circular to oval (data not shown). The flagellar region included an axoneme, a pair of mitochondrial derivatives, a pair of triangular accessory bodies, and a puff-like corpuscle with electron-dense areas (Fig. 18). In addition, the mitochondria derivatives were about 0.33 μm at their widest diameter. In the axoneme, we observed a 9 (outer singlets) + 9 (intermediate doublets) + 2 (central singlets) pattern of microtubules. Twin or multiple sperm tails were also observed (Fig. 19). In these cases, each axoneme was part of a distinct flagellum.

DISCUSSION

Although spermiogenesis and spermatozoan morphology are similar among most insect species, many species also exhibit certain distinctive characteristics. In this report, the ultrastructural aspects of spermiogenesis in *R. ferrugineus* are described. The spermatozoa of *R. ferrugineus* largely follow the generalized model of pterygote spermatozoa proposed by Baccetti (1972). The arrangement of the mitochondrial cristae and the quantity and distribution pattern of electron-dense material within the mitochondrial derivatives are characteristic of each species (Baccetti 1972; Rosati et al. 1976). In *R. ferrugineus*, the nuclei of early spermatids were observed to be displaced to the cell periphery, while the nebenkern concentrated at the opposite pole to form 2 clusters. In contrast, only one nebenkern cluster is found in the tenebrionid beetle, *Palembus der-
The gradual condensation of the nuclear material is accompanied by a change in the shape of the nucleus from spherical to elongated. During the elongation of the nucleus, the chromatin is reorganized, resulting in a long, highly compact nucleus. In some instances, the nucleus contains small spots of less electron-dense areas within the chromatin mass, rather than a homogeneous texture. The nuclear material does not undergo uniform morphological changes during spermiogenesis, as either the central zone or the periphery can be the first to condense (Chevailler 1970). Similar structural changes during nuclear development have been described in other beetles (Báo et al. 1993a; Báo 1996; Name et al. 2007).

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9 + 9 + 0 axoneme pattern and a limited degree of asymmetry in the tail organelles (Burrini et al. 1988). The flagellum of *R. ferrugineus* exhibits 2 equal-sized accessory bodies and one puff-like corpuscle, which are common characteristics in most coleopterans, except in the flagellum of *Coelomera lanio* Dalman and *Cerotoma arcuata* (Olivier) (Coleoptera: Chrysomelidae), which present a single accessory body (Báo 1996, 1998). The puff-like corpuscles, with a flocculent aspect, exhibit different shapes and sizes. Similar structures have been observed in other Chrysomelidae (Baccetti & Daccordi 1988) and Curculionoidea (Burrini et al. 1988; Name et al. 2007). The puff-like corpuscles seem to be characteristic components of the coleopteran flagellum, and it is likely that these corpuscles help to maintain the equilibrium of the flagellar structure during motility (Name et al. 2007).

In the present study, bi- and multi-flagellate spermatids and spermatozoa were observed at a low rate. This phenomenon is common in insects (Wolf 1997), although it is not observed in the Curculionidae *S. zeamais* and *S. oryzae* (Name et
al. 2007). One explanation for this phenomenon, introduced by Wolf (1997), is that there may be a secondary fusion of cells, which could occur during gonial mitosis, where a complex system of cytoplasmic bridges connects the germ cells throughout spermatogenesis. The cytoplasmic bridges widen occasionally, and cells fuse with one another. Another possibility is that supernumerary chromosomes (B chromosomes) may be present, which would increase the incidence of cell fusion during spermatogenesis (Suja et al. 1987, 1989). To our knowledge, only 5 weevil species have been reported to exhibit B chromosomes (Ennis 1972; Smith & Brower 1974; Dey 1989; Holecová et al. 2005). Determining the specific reason for the existence of bi- and multi-flagellate sperm in R. ferrugineus will require further investigation.

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