Improving Diagnosis and Clinical Management of Acquired Systemic Autoinflammatory Diseases

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Abstract: Systemic autoinflammatory diseases (SAID) are conditions caused by dysregulation or disturbance of the innate immune system, with neutrophils and macrophages the main effector cells. Although there are now more than 40 distinct, genetically defined SAIDs, the genetic/molecular diagnosis remains unknown for a significant proportion of patients with the disease onset in adulthood. This review focuses on new developments related to acquired/late onset SAID, including phenocopies of monogenic disorders, Schnitzler’s syndrome, Adult onset Still’s disease, VEXAS syndrome, and autoinflammatory complications associated with myelo-dysplastic syndrome.

Keywords: autoinflammation, adult onset Still’s disease, inflammasome, VEXAS, Schnitzler’s syndrome

Introduction

Systemic autoinflammatory diseases (SAID) are defined as conditions primarily caused by dysregulation or disturbance of the innate immune system, with neutrophils and macrophages the main effector cells. Although there are now more than 40 distinct SAIDs for which the genetic basis has been established,¹ the genetic/molecular diagnosis remains elusive for a significant proportion of patients. This is particularly true for patients with adult onset of the disease, in whom the underlying cause is unknown in at least 50% of all cases.²

Inherited or Mendelian forms of SAID are caused by rare pathogenic germline variants arising within relevant genes. These are typically inherited from one or both parents, but in some situations may arise de novo. In all cases, the pathogenic variants are present in most or all tissues and cells of the offspring. Therefore, depending on the penetrance of such variants, most patients are symptomatic from early childhood. In some extreme cases, such as neonatal-onset multisystemic disorders (NOMID), the symptoms might be present from birth. Rarely, disease onset might be in adulthood, particularly in late adulthood. Most patients presenting for the first time in adult life are rarely ever diagnosed with a known monogenic form of SAID.

Recent reports and studies have described late onset SAID due to somatic, post-zygotic mutations. These are either in the known causative genes, or due to pathogenic variants in novel genes such as UBA1,³ as in the case of newly-discovered VEXAS (Vacuoles, E1 ubiquitin, X-linked, autoinflammatory somatic) syndrome. Somatic mutations are those not inherited from a parent, but instead are introduced during the lifetime of the individual as a result of a DNA replication defect or environmental insult (eg, UV radiation). In the case of late onset SAID, such somatic mutations tend to arise within myeloid cell lineage, which is the most relevant for the pathogenies of SAID. It has been reported that the proportions of neutrophils carrying a pathogenic mutation may be as low as 2–5%, and still be enough to cause a full clinical phenotype indistinguishable from a hereditary form of the disease.⁴

Although SAIDs remain relatively rare, their recognition in the pediatric setting has improved. This is particularly the case when making an early diagnosis in more established forms such as hereditary fever syndromes, which has been aided by the development of agreed diagnostic criteria.⁵ At the same time, late onset or acquired forms remain difficult to
| Diseases                        | Key Investigations                                                                 |
|--------------------------------|-------------------------------------------------------------------------------------|
| **Infectious diseases**        |                                                                                     |
| Bacterial                      |                                                                                     |
| Septicemia                     | Blood cultures, PCT                                                                |
| Infectious endocarditis        | Heart ultrasonography                                                              |
| Bilary, colic or urinary infections | Targeted cultures, CT Scan                                                         |
| Tuberculosis                   | IGRAs, PCR, CT Scan                                                                 |
| Brucellosis, yersiniosis …      | Serology, PCR                                                                       |
| **Viral**                      |                                                                                     |
| HIV, viral hepatitis, parvovirus B19, | Serology, PCR                                                                       |
| Herpes viridae, measles, rubella … |                                                                                     |
| **Parasitological**            |                                                                                     |
| Toxoplasmosis, abscessed parasitosis | Serology, PCR                                                                       |
| **Malignant diseases**         |                                                                                     |
| Hematological                  |                                                                                     |
| Hodgkin disease or non-Hodgkin lymphoma | Lymph node biopsy                                                                 |
| Angio-immunoblastic lymphadenopathy | Bone marrow examination                                                            |
| Castelman disease              | CT scanner                                                                          |
| Myeloproliferative disorders   | PET/CT scanner                                                                      |
| Solid cancers                  |                                                                                     |
| Kidney, colon, lung …          | CT scanner, PET/CT scanner, Biopsies                                                 |
| Paraneoplastic syndroms        |                                                                                     |
| **Systemic diseases**          |                                                                                     |
| Autoimmune diseases            | Antinuclear autoantibodies                                                         |
| Systemic lupus erythematosus   | MRI, biopsy                                                                         |
| Polymyositis, dermatomyositis   | RF, ACPA                                                                            |
| Rheumatoid arthritis           | ANCA, arteriography                                                                 |
| Polymyositis nodosa or other vasculitis |                                                                                     |
| **Auto-inflammatory diseases** |                                                                                     |
| Post-streptococcal arthritis   | ASLO                                                                                |
| Reactive arthritis             | HLA-B27, MRI                                                                        |
| Hereditary auto-inflammatory syndromes | Family history                                                                    |
| Familial Mediterranean fever    | MEFV gene analysis                                                                  |
| Mevalonate kinase deficiency   | Urinary mevalonic acid, mevalonate kinase analysis                                   |
| TNF receptor-associated periodic syndrome | TNFRSF1A gene analysis                                                             |
| NLRP3-Associated autoinflammatory disease | NLRP3 gene analysis                                                                |
| Schnitzler syndrome            |                                                                                     |
| VEXAS syndrome                 | UBA1 gene analysis                                                                   |
| **Other Conditions**           |                                                                                     |
| Sarcoidosis                    | ACE, biopsy (granulomatosis)                                                        |
| Neutrophilic dermatosis, Sweet syndrome | Biopsy …                                                                         |
| Drug-related hypersensitivity or other pseudo-lymphoma |                                                                                     |
| Kikuchi-Fujimoto disease       |                                                                                     |

**Notes:** Adapted from Mitrovic S, Fautrel B. New markers for adult-onset Still's disease. Jt Bone Spine. 2018;85(3):285–293. Copyright © 2018 Elsevier Masson SAS. All rights reserved. 
*Abbreviations:* PCT, procalcitonin; CT, computed tomography; IGRAs, interferon gamma release assays; PCR, polymerase chain reaction; PET, positron emission tomography; RF, rheumatoid factor; ACPA, anti-citrullinated antibody; ANCA, anti-neutrophil cytoplasmic antibodies; ASLO, anti-streptolysin O antibody; HLA, human leukocyte antigen; MRI, magnetic resonance imaging; MEFV, Mediterranean fever; TNFRSF1A, tumour necrosis factor receptor superfamily member 1A; ACE, angiotensin converting enzyme.
Table 2 Key Features of Systemic Autoinflammatory Syndromes. Adapted from 3,4,23,24,37,47,118–120,129,130,143,144

|                      | Late-Onset NLRP3-AID | Schnitzler’s Syndrome | AOSD               | VEXAS                     | MDS-SAID                  |
|----------------------|----------------------|-----------------------|--------------------|--------------------------|--------------------------|
| **Average age of onset** | >40 years of age     | Most cases >50 years of age | Possibly two peaks (younger and older adults) | Most cases >60 years of age | Most cases >60 years of age |
| **Gender**           | M=F                  | M=F                   | M<F                | M>>>F                    | M=F                      |
| **Clinical characteristics** |                       |                       |                    |                          |                          |
| **Fever**            | Yes                  | Yes                   | Yes                | Yes                      | Not always               |
| **Weight loss**      | No                   | Possible but not typical | Yes               | Yes                      | Possible                 |
| **Skin**             | NUD                  | NUD                   | Salmon pink rash (typical), urticaria, NUD | Neutrophilic dermatosis (Sweets syndrome), medium vessel vasculitis, Leukocytoclastic vasculitis | Sweets syndrome, PG |
| **Musculoskeletal**  | Arthralgia, arthritis, myalgia | Arthralgia, arthritis, myalgia | Arthralgia, arthritis, myalgia | Arthralgia, arthritis, chondritis, myalgia | Arthralgia, arthritis, myalgia |
| **GI/liver**         | No                   | No                    | Abdominal pain, hepatomegaly | Hepatomegaly, colitis | Rare, colitis |
| **Cardiac**          | No                   | No                    | Pericarditis, myocarditis | Pericarditis | Not usually |
| **Respiratory**      | No                   | No                    | Pleuritis, pulmonary infiltrations | Pulmonary nodules, neutrophilic alveolitis, pleural effusion | Rare |
| **Neurological**     | Headaches, aseptic meningitis, sensorineural hearing loss | No | Rare, aseptic meningitis | Rare, aseptic meningitis, sensorineural hearing loss | Rare |
| **Hematological**    | No                   | MGUS, IgM>>IgG paraprotein | Splenomegaly, lymphadenopathy, cytopenia (usually as a feature of MAS) | Macrocytic anemia, vacuoles in myeloid and erythroid cells (most cases) cytopenia, MDS | Anemia, cytopenia, vacuoles in myeloid and erythroid cells (some cases) |
| **Other frequent features** | Nil                  | Nil                   | Sore throat         | Periorbital edema        | Nil                      |
| **Complications**    | Deafness if untreated | Progression into hematological malignancy (MM, WM, MZL, etc) | MAS; Disseminated intravascular coagulopathy;Thrombotic thrombocytopenic purpura; Diffuse alveolar haemorrhage; Fulminant hepatitis; Pulmonary hypertension | MDS, transfusion dependence; infections, thromboembolic events; MAS rare, progression to AML or CMML rare | Progression into AML and CMML |

(Continued)
diagnose in a timely manner. The focus of this review will, therefore, be on this latter group. We will provide updates on new developments in this field and describe our broad approach to the investigation and management of these patients.

### Phenocopies of Monogenic Disorders

#### Late Onset (Acquired) NLRP3-Associated Autoinflammatory Disease (NLRP3-AID)

Heterozygous gain-of-function (GoF) variants in the NLRP3 gene are known to cause a spectrum of inflammatory conditions previously known as cryopyrin-associated periodic syndrome (CAPS) and now termed NLRP3-AID. The NLRP3 gene codes for NACHT (NAIP, CIITA, HET-E, and TP1), LRR (leucin rich repeat), and PYD (pyrin) domain-containing protein 3 or NLRP3, which forms the sensing part of a larger multimolecular complex known as the NLRP3 inflammasome.\(^\text{145,146}\) The mutated protein reduces the threshold and/or leads to spontaneous NLRP3 inflammasome activation, resulting in release of the active forms of IL-1beta and IL-18. It is inappropriate release of the IL-1beta that produces most of the symptoms associated with NLRP3-AID, since a therapeutic blockade of this cytokine leads to complete symptom control and the disease remission in most patients.\(^\text{147}\)

The range of symptom and disease severity varies significantly across the NLRP3-AID spectrum. The mildest form, also known as familial cold autoinflammatory syndrome (FCAS), is associated with cold induced urticarial-like rash, mild flu-like symptoms, and fever, lasting 1–2 days. Muckle-Wells syndrome (MWS) is of moderate severity, with fever attacks lasting for several days. Before introduction of the modern biological therapies, MWS was also frequently

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**Table 2 (Continued).**

| Pathogenesis/ genetic cause | Late-Onset NLRP3-AID | Schnitzler’s Syndrome | AOSD | VEXAS | MDS-SAID |
|-----------------------------|----------------------|-----------------------|------|-------|---------|
| GOF somatic variants in NLRP3 | MYD88 (p. L265P variant) in selected cases. Significance unclear | Complex, likely polygenic | Somatic mutations in UBA1 | Complex, somatic variants in MDS related genes likely relevant |

### Treatment

- **Steroid responsive**
  - Partial

- **Colchicine**
  - No

- **Anti-IL-1 blockade**
  - Yes

- **Anti-IL-6**
  - No

- **Anti-TNF**
  - No

- **JAKi**
  - No

- **Allogeneic HSCT**
  - No

- **Other**
  - Nil

**Abbreviations:** AOSD, adult onset Still’s disease; NUD, neutrophilic urticarial dermatosis; MDS, myelodysplastic syndrome; SAID, systemic autoinflammatory disease; PG, polyarteritis nodosa; AML, acute myeloid leukemia; CMML, chronic myelomonocytic leukemia; MM, multiple myeloma; MZL, marginal zone lymphoma; WM, Waldenstrom macroglobulinemia; MGUS, monoclonal gammopathy of undetermined significance; MAS, macrophage activation syndrome.
associated with chronic systemic inflammation resulting in complications such as amyloidosis, sensorineural hearing loss and aseptic meningitis. As the name suggests, NOMID presents in the neonatal period and it is the most severe form of NLRP3-AID. Without treatment, NOMID causes almost continuous fevers, with chronic systemic inflammation leading to the complications seen in MWS in addition to visual loss, epiphyseal enlargements, and cognitive impairment.

The urticarial-like rash which is seen in all forms of NLRP3-AID to a different degree is non-itchy. The lesional biopsy typically shows neutrophilic predominant infiltration, hence the name of this entity being the neutrophilic urticarial dermatosis, which also confirms the central role neutrophils play in the pathogenesis of this condition.  

Description of late onset NLRP3-AID follows a previous discovery that some cases of NOMID result from somatic mosaicism with de novo rare variants arising within the NLRP3 gene post-zygotically. Although these patients presented with classical symptoms in the neonatal period, their genetic diagnosis was initially missed by standard Sanger sequencing method. The importance of these discoveries was two-fold. They showed that complete clinical phenotypes can be replicated even when only a relatively small proportion of cells carries a pathogenic mutation, and that next generation sequencing methods are needed to diagnose such cases. Subsequently, late onset NLRP3-AID were described. Here, patients who were previously fit and well and with no relevant family history developed symptoms consistent with FCAS or MWS for the first time in their 4th, 5th and later decades of life. These reports showed not only that a small proportion of cells is sufficient to carry a pathogenic mutation, but also that these cells need to be relevant for the disease pathogenesis, which in these cases were cells of the myeloid lineage and in particular neutrophils. This concept was recapitulated recently in a mouse model, where animals were engineered so that only neutrophils carried a pathogenic mutation in NLRP3. This was again sufficient to reproduce a complete NLRP3-AID phenotype. In the largest case series of late onset-NLRP3-AID (8 patients) the symptom onset occurred between 31–71 years. The clonal expansion of those myeloid cells carrying the pathogenic NLRP3 allele was also noted. In one patient there was an increase in the mutated NLRP3 gene from 5.1% to 45% over 12 years, which interestingly was associated with a greater requirement for anti-IL-1 inhibition for symptom control. An increase like this suggests a mechanism of either increased production, through non-malignant clonal expansion of hematopoietic stem cells with the NLRP3 mutation, or a selective survival advantage in the myeloid cells with the NLRP3 mutation. Neither possibility has yet been determined.

**Acquired NLRC4-AID**

NLRC4 (NOD-like receptor family CARD-containing 4 protein) is an intracellular receptor whose activation, like NLRP3, leads to the production of an inflammasome and resulting pyroptosis, through caspase-1 mediated cleavage of gasdermin D. Activation of NLRC4, typically by intracellular Gram-negative bacteria, causes it to bind to ASC leading to caspase-1 activation and subsequent cleavage and release of the pro-inflammatory cytokines IL-18 >> IL-1β. Germline GoF variants of the NLRC4 gene have been discovered in patients presenting with autoinflammatory phenotypes similar to FCAS, with characteristic rashes and features, as well as infantile enterocolitis and recurrent macrophage activation syndrome (MAS). The excess of IL-18 rather than IL-1β is thought to be more important for the pathogenesis of NLRC4-AID, and in particular MAS. Selective targeting of IL-18 in this setting has been shown to have therapeutic benefit.

Up until very recently NLRC4 somatic mosaicism had only been identified in two neonates, both born prematurely and presenting with florid systemic autoinflammation. One of these patients (mutation p.Ser171Phe) presented with necrotising enterocolitis and MAS, passing away at 2 months, whilst the other (mutation p.T177A) was given a diagnosis of NOMID and responded well to IL-1 blockade.

As of 2022 there have been two reported cases of somatic NLRC4 mutations identified in older adults with no personal or familial history of systemic inflammation. In one case a 69-year-old Chinese woman presented with a history of intermittent prolonged fevers and a recurrent rash. Results showed elevated inflammatory markers, elevated lactate dehydrogenase, and changes consistent with cutaneous vasculitis on skin biopsy. A good clinical response was achieved with Tocilizumab. Genetic analysis revealed a somatic missense variant (p.His443Gln) in the NLRC4 gene with a mosaicism ratio of 2.61%, with the mutation found in both myeloid and lymphoid cells suggesting a late onset mutation occurring in a hematopoietic stem cell progenitor. In another case, a 57 year old Spanish woman was worked up for a
10 year history of periodic symptoms; beginning with 2–3 day episodes of temperatures and features of systemic inflammation occurring every 6–8 weeks and slowly progressing in severity. An excellent clinical and biochemical response, including the resolution of mesenteric lymphadenopathy, was eventually achieved with anakinra 200 mg/day. Canakinumab was later shown to be equally effective. Genetic analysis suggested a likely pathogenic post-zygotic variant (p.Ser171Phe) with a mutant allele fraction of ~3%.

A consideration of both acquired NLRP3-AIDs and acquired NLRC4-AIDs is how a relatively small proportion of peripheral blood cells carrying the pathogenic NLRP3 variants can produce identical phenotypes to their germline counterparts. One possibility is that the activated inflammasome complexes of pathogenic cells retain their enzymatic activity after release via pyroptosis and are taken up by surrounding cells, propagating a “prionoid” inflammatory cascade. In support of this, soluble ASC protein specks can be detected in the serum of patients with NLRP3-AIDs and the levels can change in line with disease activity. Another possibility is the presence of more pathogenic NLRP3 or NLRC4 variants in somatic cases as a result of somatic mutations rarely found in inherited disease, something seen in the cases of NLRP3 somatic mosaicism. Though these mutations arise at the same gene hotspots, their much greater prevalence in somatic disease suggests when they do occur as germline mutations the resulting severity is not compatible with a viable embryo.

Other Examples

Other acquired SAIDs, in which phenocopies due to somatic mosaicism have been found to occur, include tumour necrosis factor receptor-associated periodic syndrome (TRAPS) and Blau syndrome (BS).

TRAPS, which classically presents in early childhood with periodic fevers, a rash and arthralgia, occurs due to mutations in the TNFRSF1A gene, encoding the TNFR1 (tumor necrosis factor receptor 1) receptor. Patients who carry what is thought to be low penetrance variants (R92Q and P46L) tend to present later, sometimes for the first time in adulthood, and with a phenotype that is overall milder compared to classical TRAPS. Acquired mutations have recently been identified in two patients presenting with symptoms consistent with systemic autoinflammation. One was a 41-year-old gentleman with a deletion mutation in TNFRSF1A (p.Ser86_Glu93del) and 7.6% whole blood mutant allele frequency. The other was a 60-year-old gentleman in whom the de novo missense mutation (p.Phe89Leu) was detected in the patients NK cells, neutrophils, and B cells, though not in monocytes and T cells.

BS is a granulomatous autoinflammatory syndrome occurring due to mutations in the NOD2 gene, typically presenting in early childhood with cutaneous sarcoidosis, granulomatous arthritis, and uveitis. A 22 year old male presenting in such a way was found to have a known pathogenic NOD2 mutation (p.Arg334Gln) with 0.9–12.9% somatic NOD2 mosaicism. Interestingly his two daughters presented with the condition in their first year of life, strongly suggesting gonadal tissue mosaicism, engendering “intrafamilial recurrence” or development of a germline mutation in these affected children.

Schnitzler’s Syndrome

Schnitzler’s syndrome (SchS) is a rare, late onset, acquired autoinflammatory disorder with prominent cutaneous features. Unlike VEXAS syndrome and late onset NLRP3-AID, the genetic basis of it is undeciphered and, hence, diagnostic criteria (Strasbourg criteria) have been devised to make a diagnosis. These include two required features, firstly, a monoclonal protein which is usually IgM and secondly the presence of a chronic urticarial looking rash which is neutrophil rich on biopsy. Other manifestations include recurrent fever, bone pain, lymphadenopathy, peripheral neuropathy, headache, arthralgia, fatigue, weight loss, and raised inflammatory markers.

Patients with SchS have been found to have elevated levels of ASC, IL-6, and IL-18, however the underlying molecular mechanism is unclear. Due to its clinical similarity with NLRP3-AID, it was suggested that SchS in some cases might also be caused by low level somatic mutations in NLRP3. However, two studies which included 32 patients in total failed to find any evidence for this. More recently another study which included a further 40 SchS cases also did not find any patients with rare variants in NLRP3. Interestingly, the same study also showed that SchS patients do not have any pathogenic variants in UBA1 either, confirming that SchS has a unique pathology. Further clues to possible aetiology of SchS stems from the observation that a proportion of patients with SchS develop Waldenstrom’s
macroglobulinaemia (WM) suggesting that both disorders may have a similar cause.24 Approximately 90% of patients with WM have been found to have the p.L205P variant in MYD88.27 MYD88 is a cytoplasmic adaptor protein which mediates interaction of toll like receptors and IL-1 receptor families. This is crucial for propagating downstream signalling of NFKB and MAPK pathways. The L265P may have different consequences dependent on the cell affected. In lymphoid cells it is thought to drive lymphoid malignancy via toll like receptor signalling pathways.28 Conversely, in myeloid cells the same mutation is thought to drive downstream signalling from the IL-1 receptor.29 The finding that in one study 9/30 patients were found to have this variant supports this hypothesis, simultaneously it does not offer a universal mechanism for all cases of SchS.25 It is therefore likely that there are other factors at play. The clonal lymphoid expansion seen in SchS may have a role in some cases. Clonal haematopoiesis of undetermined potential (CHIP) is characterised by the acquisition of somatic mutations in stem cells and is associated with an increased level of the pro-inflammatory cytokines IL-1 and IL-6.30 Additionally, CHIP related somatic mutations in TET2 and U2AF1 stimulate the generation of reactive oxygen species and this in turn is thought to trigger the NLRP3 inflammasome.30–32 This at best may account for a small proportion of cases as only 1/30 patients were found to have a CHIP associated mutation in one study.25 Finally, the IgM paraprotein may have a direct pathogenic effect, the mechanism for this is unclear but the finding that the disease seems to remit when underlying hematological malignancy is treated and the paraprotein disappears supports this hypothesis.33,34 Ultimately there continues to be gaps in our understanding of the pathophysiology of SchS which is a promising area for future study.

SchS is dramatically responsive to IL-1 antagonism, in one study the initiation of anakinra led to a significant reduction in CRP and a meaningful reduction in quality-of-life measures; 95% reported the disappearance of all symptoms accompanied by normalization of plasma CRP concentration.17 Similarly, canakinumab has also proved to be efficacious.35 IL-6 blockade may offer an alternate treatment option, as illustrated in an increasing number of case series and reports.36,37 Notably, it may be an option in patients who fail IL-1 blockade. More recently, Huang et al.38 described a patient with SchS who received Ibrutinib (there was no lymphoplasmacytic lymphoma on bone marrow examination) with the intent of treating his autoinflammatory symptoms, and there was a dramatic symptom improvement in parallel with a reduction in paraprotein levels. In other cases, treating the underlying malignancy proves to be effective.33,34

**Adult Onset Still’s Disease**

Adult onset Still’s disease (AOSD) is an autoinflammatory condition characterised by prolonged intermittent fevers, arthralgia, and evanescent rash in the absence of infection, malignancy, or rheumatological disease.39 AOSD and systemic juvenile idiopathic arthritis (sJIA) have different diagnostic criteria but are increasingly understood to be the same disease, with commonality in clinical features, immunological profiles, and responses to treatment.39

AOSD typically presents in younger people, with onset occurring in a bimodal pattern, at ages 15–25 and 35–45, though some studies suggesting a further peak between ages 60–65.40–44 Epidemiological data on AOSD is somewhat limited, with an annual incidence between 0.24–0.62/100,000 and point prevalence of 2.7–6.9/100,000, with women predominantly affected more often than men.43–46 A retrospective study performed in the USA between 2008–2013 gave an inpatient mortality of 2.6%.47 The most common clinical features of AOSD are an intermittent fever of ≥39°C lasting ≥1 week, arthralgia lasting ≥2 weeks, and a skin rash that is classically a salmon-pink, non-pruritic, maculopapular eruption occurring during the fever. Myalgias, pharyngitis, lymphadenopathy, and splenomegaly also characteristically feature.39,48

AOSD was traditionally felt to occur as one of three broad patterns; monocytic systemic, polycyclic systemic, and chronic articular. More recently, a dichotomous classification system, with characteristic clinical and immunological profiles, has been proposed, whereby AOSD can be either systemic (high fever, rash, multi-organ involvement, and predominant IL-18/1β cytokine signature) or articular (prominent joint disease and a predominant IL-6 cytokine signature).49–51 Further work has been done to highlight the clinical heterogeneity of AOSD, with a study showing they may fall into four distinct phenotypes, each characterised by a single prominent clinical feature (high CRP, high ferritin, high systemic score, low CRP/ferritin), raising the possibility of risk stratification and targeted therapies in future.52
When comparing presentation by age, those that develop AOSD aged 65 or above similarly present with fever, arthralgia, and skin rashes, though are less likely to develop pharyngitis and more likely to develop pleuritis. Those with an elderly onset are 3–5-times more likely to develop life-threatening complications such as disseminated intravascular coagulopathy (DIC) and MAS, highlighting the need for prompt recognition and management in this population.

Biochemical findings include raised CRP/ESR, a neutrophilic leukocytosis, anemia, and thrombocytopenia. Hyperferritinemia is common in AOSD but in isolation is poorly predictive of the disease. However, ferritin levels have a role in monitoring disease activity in AOSD as they are a reliable marker of disease activity and severity, whilst a profound increase suggests hemophagocytosis and MAS. AOSD has a characteristic pattern on FDG PET/CT, showing increased uptake in the bone marrow and spleen with multiple reactive lymph nodes in a symmetrical distribution in the axilla and neck. This characteristic pattern combined with the intensity of uptake has been shown to effectively differentiate AOSD from other rheumatological diseases, though the radiation burden and costs likely preclude it from becoming a key tool of the diagnostic process.

The most widely used and validated criteria for diagnosing AOSD are Yamaguchi’s criteria, with a sensitivity of 96.2% and specificity of 92.1%. This classification includes the common clinical features stated above, though requires the exclusion of infections, malignancies, and rheumatic diseases (Table 1), which can significantly delay diagnosis. In 2002, Fautrel et al developed a new classification including glycosylated ferritin fraction ≤20% and removing any exclusion criteria, producing a classification with a sensitivity of 80.6% and an improved specificity of 98.5%. The Fautrel classification has since been validated in a separate study, giving a sensitivity of 96.3%, specificity of 98.9%, and positive and negative predictive values of 94.5% and 99.3%, respectively, providing an effective tool for earlier diagnosis and treatment.

Genetics
Unlike monogenic systemic autoinflammatory syndromes (mSAIDs), AOSD has a presumed polygenic basis. These underlying genetic factors, through unclear immunological pathogenesis, engender a susceptibility to the disease which is then likely triggered by a combination of infective and environmental factors. There is a degree of genetic overlap between AOSD and other SAIDs, with one case series of 162 AOSD/sJIA patients showing 51 (31.4%) carried at least one genetic variant associated with the periodic fever syndromes Familial Mediterranean Fever (FMF), CAPS, TRAPS, or BS. Where these genetic changes indicate shared pathogenic mechanisms of the innate immune system, unique to AOSD is the association of HLA subtypes with the disease, suggesting involvement of the adaptive immune system. HLA-DRB1*11 (OR 2.3), HLA-DQB1*06:02 (OR 2.70), HLA-DRB1*15:01 (OR 2.44), and HLA-DQA1*01:02 (OR 1.97) have all demonstrated association with the disease against healthy controls. Furthermore, studies have shown associations with mild, self-limiting disease (HLA-Bw35 and HLA-DRB1*14), chronic disease (HLA-DRB1*2501 and HLA-B1*1501), and those with chronic and systemic AOSD (HLA-DQB1*0602).

Pathogenesis
The pathogenesis of AOSD is felt to be driven in large part by a pro-inflammatory cytokine environment with IL-1, IL-6, IL-18, and TNF-α driving an inflammatory cascade. Underpinning this process are new mechanisms gradually being elucidated.

IL-1 is the most potent human endogenous pyrogen and has two forms, IL-1α and IL-1β, both of which stimulate the IL-1R1 receptor and are countered by the endogenous competitive inhibitor IL-1 receptor antagonist (IL-1ra). IL-1β, IL-18, which is part of the IL-1 family of cytokines, and IL-6 are all pro-inflammatory cytokines elevated in the context of AOSD. IL-6 and IL-18 profiles have even been suggested to indicate clinical subsets, with raised IL-6 increasing the likelihood of articular AOSD (prominent joint disease) and raised IL-18 suggesting an increased risk of developing MAS. Excellent clinical responses in AOSD to anti-IL-1 therapy and anti-IL-6 therapy further underscore the role of these proinflammatory cytokines in active disease.

The interplay between Toll-like receptors (TLRs) and neutrophils, through their various mediators, is emerging as a key component in the development of the sterile inflammation found in AOSD/sJIA. TLRs are pattern recognition receptors that detect pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs), leading to the activation of the innate immune system. This activation results in the production of pro-inflammatory cytokines, chemokines, and other inflammatory mediators.

The role of TLRs in AOSD is well-established. TLRs are expressed on various immune cells, including macrophages, dendritic cells, and neutrophils. These receptors are activated by ligands, which are either derived from the pathogen or from the host cell itself. In AOSD, TLRs can be activated by various agents, such as bacterial lipopolysaccharides, nucleic acids, and DAMPs, leading to the production of pro-inflammatory cytokines, chemokines, and other inflammatory mediators.

TLR activation in AOSD can lead to the production of IL-18, which is a critical mediator in the sterile inflammation seen in AOSD. IL-18 is a pro-inflammatory cytokine that plays a key role in the development of MAS. It is produced by activated macrophages and dendritic cells and acts to recruit and activate neutrophils, further exacerbating the inflammatory response.

In addition to the role of TLRs in AOSD, other pro-inflammatory mechanisms are also involved in the pathogenesis of the disease. For example, the role of cytokines, such as IL-1, IL-6, and TNF-α, in AOSD has been well-documented. These cytokines are produced by various immune cells and play a crucial role in the development and maintenance of inflammation in AOSD.

One of the most important pro-inflammatory cytokines in AOSD is IL-18. IL-18 is a member of the IL-1 family of cytokines and is known to play a critical role in the development of MAS. It is produced by activated macrophages and dendritic cells and acts to recruit and activate neutrophils, further exacerbating the inflammatory response.

The pathogenesis of AOSD is felt to be driven in large part by a pro-inflammatory cytokine environment with IL-1, IL-6, IL-18, and TNF-α driving an inflammatory cascade. Underpinning this process are new mechanisms gradually being elucidated.
receptors (PRRs) involved in the detection of PAMPS and DAMPS. TLR signalling leads to activation of the NLRP3 inflammasome with downstream promotion of pyroptosis and release of the key pro-inflammatory cytokines IL-1β and IL-18.74,75

Neutrophils play a crucial role in this activation, as suggested by the high degree of neutrophilic leucocytosis (>80% of patients) during an acute flare, in part through the production of neutrophil extracellular traps (NETs), which contain the proteins S100A8, S100A9, and S100A12.76–78 These S100 proteins are DAMPs and ligands for TLR4, inducing activation and further release of chemokines and cytokines to increase neutrophil recruitment and pyroptosis, precipitating a positive feedback loop.79

S100A12 and the S100A8/S100A9 dimer (calprotectin) are reliably raised in sJIA a and AOSD, offering a biomarker for diagnosis and disease activity which has shown superiority to CRP in differentiating sJIA from other SAIDs.72,80–86 Of further interest is the increased type I interferon (IFN) response found in AOSD as neutrophils exposed to IFN-α are primed to form NETs.73 This recent insight may inform future avenues of research and therapy, particularly as IFNs signal via the Janus kinase (JAK)-signal transducer and JAK inhibitors have shown promise in treating the condition.73,87,88

**MAS in Context of AOSD**

In MAS, also known as secondary hemophagocytic lymphohistiocytosis (sHLH), a pro-inflammatory environment driven by autoinflammation, infection (eg, previous EBV or CMV), malignancy, or autoimmune disease reduces the ability of natural killer (NK) and cytotoxic CD8+ T cells to lyse antigen presenting cells (APC).89,90 The resulting increased contact time between lytic cells and APCs leads to a pro-inflammatory cascade, involving interferon-gamma (IFN-γ), tumor necrosis factor (TNF), IL-1, IL-6, and IL-18, which in turn leads to macrophage activation and hemophagocytosis. IFN-γ is particularly felt to play a role, supported by its role in macrophage activation, characteristic type 1 polarised T-cell phenotypes seen in the condition, as well as evidence of increased neopterin, an inflammatory marker secreted by IFN-γ activated macrophages and dendritic cells (DCs), in patients with MAS as compared to control groups.91,92 The role of IFN-γ has been further underscored by the reported success of the anti-IFN-γ antibody Emapalumab in primary HLH, as well as the ongoing trials into its efficacy, specifically in MAS.93,94 In the context of AOSD and systemic juvenile idiopathic arthritis (a pediatric equivalent of AOSD), MAS occurs in 7–14% of children with sJIA95,96 and 10–15% of patients with AOSD.97,98 In another 30–40% of patients MAS might be present in a subclinical form.99 Elevated IL-18 levels have been shown to be predictive of developing MAS in both sJIA and AOSD50,100–102

Macrophage dysfunction leading to MAS has also been suggested in sJIA lung disease (sJIA-LD), with one study showing 64% (23/36) of patients had pulmonary alveolar proteinosis and/or endogenous lipoid pneumonia (PAP/ELP) as their predominant lung pathology.103 Interestingly, those with sJIA-LD had significantly higher levels of IL-18, were more likely to develop MAS, and were more likely to have an adverse reaction to biologic therapy, compared to sJIA without lung disease.104 One observational study showed that those with lung disease in AOSD had an increased mortality compared to those without, suggesting these patients may require a more tailored therapeutic approach to mediate these risks.105

It should be noted that excessive inflammatory cytokine production, with a failure of negative feedback mechanisms, leading to hyperinflammation and organ dysfunction, are the hallmark features of a cytokine storm106 MAS is the archetypal cytokine storm syndrome (CSS), though there are other distinct CSS (eg, driven by sepsis, COVID-19) that have their own unique pathogenesis, immunological profiles, and show marked heterogeneity on attempts to apply pre-existing classification systems, like those used for MAS.106,107

MAS classically presents with a triad of fever, hyperrferritinemia, and pancytopenia, with an accompanying clinical picture of multi-organ dysfunction.89,90 Any patient presenting this way, with no clear evidence of underlying infection, malignancy, or autoimmunity should be referred for specialist consideration of autoinflammatory disease, including AOSD.

**Treatment**

First line therapy of AOSD involves corticosteroids (0.5–1 mg/kg/day), with studies suggesting higher initial doses (eg, ≥40 mg or ≥0.8 mg/kg/day) lead to quicker resolution with fewer relapses.108,109 The response to corticosteroids should be seen within a few days.48,109 DMARDs such as methotrexate are then utilised as steroid sparing agents and even in
low doses (7.5–17.5 mg/week) can effectively induce remission and allow the cessation of corticosteroids. Despite this, a significant number (~22%) fail on this combination of corticosteroids and DMARDs, a condition termed refractory AOSD. This cohort requires biologics, typically in the form of either IL-1 antagonists, IL-6 antagonists, or TNF-inhibitors. However, in patients with severe and rapidly progressing disease and those at risk of developing MAS, use of biologics, particularly anti-IL-1 therapy, should be considered early and before conventional synthetic DMARD’s.

The IL-1 inhibitor anakinra, the IL-1β inhibitor canakinumab, and the IL-6 inhibitor Tocilizumab have all been shown to cause rapid and sustained improvement in clinical symptoms and inflammatory markers associated with AOSD. Anakinra should be first line in so-called systemic AOSD.

In an observational study of anakinra assessing 140 patients over 12 months, there was a significant reduction in clinical and biochemical features and steroid sparing effect, with 69.2% of patients remaining on the therapy at 12 months and 28.1% stopping due to clinical remission. In an observational study of 50 patients receiving canakinumab for refractory disease, there was complete response in 78% of patients within 3 months (median) and a partial response in 20%, with around 51% benefitting from a sustained corticosteroid wean. After 27 months (median), 44% of patients were still on canakinumab treatment whilst 18% achieved clinical remission. A Phase II, randomised, double-blind placebo-controlled trial for canakinumab showed significantly superior per-protocol response rates in those receiving canakinumab, as measured by the American College of Rheumatology criteria (ACR30, 50 and 70) after 12 weeks.

It has been suggested that TNF inhibitors and Tocilizumab should be prioritised in refractory AOSD with pronounced joint disease, whereas Anakinra should be first line in so-called systemic AOSD. Other therapies that are beginning to show promise include JAK inhibitors and the IL-18 binding protein tadekinig alfa. In an observational study of 14 patients the JAK1/3 inhibitor tofacitinib was shown to induce complete remission in 50%, partial remission in 43%, and reduce the average dose of daily prednisolone from 37.3 mg to 5 mg/day at 12 months. A Phase II safety and efficacy trial of tadekinig alfa in AOSD found that both 80 mg and 160 mg, three times a week, had a favourable safety profile and produced a biochemical and/or clinical response in 50% of participants, irrespective of dose. These results highlight the way in which elucidating the pathogenesis of these rare syndromes informs the development of highly targeted, effective treatments.

**VEXAS**

VEXAS syndrome (vacuoles, E1 enzyme, autoinflammatory, somatic) was originally described in 2020, the disorder represents the prototype of the newly-proposed group of “hemato-inflammatory” diseases. It is caused by somatic mutations in UB1 which encodes the E1 ubiquitin ligase, an enzyme that plays a key role in post-translational ubiquitination of cellular proteins. The ubiquitin system regulates innate immune-signaling pathways and plays a crucial role in adaptive immune responses and tolerance. UB1 lies on the X-chromosome, hence the disease almost exclusively effects males. In the original report all 25 men had missense mutations in codon 41 affecting the methionine-41 codon (p.Met41) leading to a loss of the cytoplasmic isoform UB1b, and the subsequent gain of a new isoform, UB1c. This catalytically impaired isoform is thought to drive inflammation. More recently, mutations effecting regions outside of codon 41 have been described. Affecting patients have highly activated inflammatory signatures in pathways associated with myeloid related inflammation such as TNF, IL-6, and IFN-γ. Indeed, animal models replicating loss of UB1 replicate upregulation of multiple cytokines, as seen in VEXAS syndrome.

VEXAS is severe and progressive, initially thought to be rare and the rapid rate of increasingly published reports suggest that the true prevalence may be under-appreciated. Clinically the disease presents with the cardinal features of fever, fatigue, and raised inflammatory markers combined with both rheumatological and hematologic features. There is systemic inflammation of multiple organ systems leading to various manifestations including Sweet’s syndrome, relapsing polychondritis, inflammatory arthritis, polyarteritis nodosa, pulmonary infiltrates, and giant cell arteritis. Hematologic features include macrocytic anemia, thrombocytopenia, thromboembolic disease, and progressive bone marrow failure, which rarely evolves to hematologic malignancy.

Our understanding of the phenotype is constantly evolving with hemophagocytic lymphohistiocytosis, myositis, and Behcet’s disease being described in association with the disease. The combination of relapsing polychondritis, a mean corpuscular volume of >100 fl or platelet count <200x10^9/l in a male patient is highly predictive of a patient with VEXAS.
Vacuoles are a characteristic feature of VEXAS. Vacuoles are rare in marrow myeloid and erythroid precursor cells, when found there is a limited range of differentials including copper deficiency/zinc toxicity and myeloid neoplasms. Consequently, sequencing of UBA1 must be considered when cytoplasmic vacuoles are seen on hematopathological examination of a specimen. Distinguishing features include the finding that the cytoplasmic vacuoles in VEXAS patients ubiquitination are predominantly localized in promyelocytes, myelocytes, erythroid precursors, and blasts in bone marrow aspirates.

There is currently no approved cure for VEXAS. High dose steroid therapy is effective but a reduction in dose invariably leads to disease relapse. Various drugs have been used to treat patients including methotrexate, calcineurin inhibitors, TNF alpha blockade, IL-1 blockade, IL-6 blockade, as well as JAK inhibitors. Bourbon et al compared the effectiveness of different therapeutic strategies by evaluating the time to change between drugs in a case series, hypomethylating agent and signaling inhibitors achieved notable results; the median time to next treatment was 12.7 months for cyclosporine, 21.9 months for azacytidine. Moreover, a marked regression of cutaneous lesions was observed after the introduction of ruxolitinib in one patient. Azacytidine holds potential promise. It is currently used to manage high risk myelodysplastic syndromes and may target myeloid cells harboring UBA1 mutations. Notably, Azacytidine has been proposed as a treatment for autoimmune disorders associated with MDS, based on data reporting control of the inflammatory manifestations in two-thirds of the patients. Allogenic hemopoetic stem cell transplantation is more likely to offer a curative treatment, recently a case series of six patients who received a transplant were described. At the time of publication five of the six patients were in complete remission, one unfortunately died post-transplant. The lack of effective safe treatments requires further evaluation.

Myelodysplastic Syndromes and Systemic Inflammation

Prior to the discovery of VEXAS, a number of studies had already established a close link between myelodysplastic disorders (MDS) and systemic inflammation. MDS is a pre-malignant bone marrow condition characterised by persistent peripheral blood cytopaenias. These result from ineffective stem cell maturation due to acquired genetic abnormalities, leading to clonal hematopoiesis. The pathogenesis of MDS involves progressive accumulation of somatic mutations, affecting various cellular processes such as transcription, epigenetic regulation, cohesin complex, RNA splicing, translation, and telomerase function. A proportion of patients (up to 33%) progress to develop acute myeloid leukemia.

The link between MDS and systemic inflammation is bi-directional. For example, evolving evidence suggests that NLRP3 inflammasome activation is a core feature of MDS. A recent mechanistic study also identified that chronic IL-1β exposure can trigger selective expansion of CEBPα-deficient multipotent hematopoietic progenitors, indicating a role for inflammation. Conversely, given the effector role of myeloid cells in SAID pathogenesis, somatic mutations in genes such as TET2 which initiate MDS development and lead to NLRP3 inflammasome activation might also be responsible for evolution of inflammatory complications seen in this condition. Both autoimmune and autoinflammatory complications occur in MDS. The latter include, among others, inflammatory seronegative arthritis, polymyalgia rheumatica, giant-cell arteritis and various neutrophilic dermatosis such as Sweet’s syndrome and pyoderma gangrenosum. A recent study demonstrated that autoinflammatory complications might be more frequent than autoimmune conditions in MDS and that the former were probably under-recognised. Many patients demonstrated an undifferentiated autoinflammatory state, defined by the presence of inflammatory symptoms such as fever, non-specific myalgia, and arthralgia, and associated with elevated CRP in the absence of overt infection. Such patients were found to be younger (on average 3 years) and also had the type of MDS which had higher risk of progression to AML.

The treatment of autoinflammatory complications associated with MDS is challenging for several reasons. Firstly, patients often present with an indistinct clinical picture which makes diagnosis difficult and means it can be problematic to follow a specific treatment guideline. There is an overall lack of defined treatment targets, and the use of immunosuppressive treatments is further complicated by the pre-existing risk of infection and cytopenia.

Typically, systemic corticosteroids have been the mainstay of the early treatment approach. In many cases, steroid treatment will lead to initial disease remission. However, refractory cases with steroid dependence are common. There is no unified approach to treatment beyond steroids, but this might involve a trial of traditional DMARDs or, increasingly,
biological and synthetic DMARDs. There is emerging evidence that treatments such as azacytidine, a hypomethylating agent used in the treatment of high risk MDS, might be helpful in these cases. Ultimately, some patients require allogenic stem cell transplantation, which has been effective for treating both MDS and inflammatory complications.

**Conclusion**

The recognition, diagnosis, and treatment of patients with late onset or acquired SAID remains challenging and requires a high degree of clinical suspicion. In part this is due to their rarity, but also results from overlapping clinical features and pathogenesis (Table 2). Furthermore, they share many of the non-specific clinical manifestations seen in very common conditions such as infection, autoimmune disorders, and malignancies, which must be excluded first. In malignancy, the situation is further complicated by the fact that some acquired SAID’s such as VEXAS and SchS often develop on a background of pre-malignant disease states. Therefore, the boundary between the autoinflammation seen in acquired SAID and that resulting from malignancy, is increasingly blurred.

There are few specific diagnostic tests beyond genetic investigation, so patients with suspected late-onset SAID’s require detailed diagnostic work up which is predominantly focused on the exclusion of overlapping conditions. The value of PET-CT in investigating patients with pyrexia of unknown origin has largely been the exclusion of malignancy, since findings in SAID patients are largely non-specific. A retrospective study demonstrated that bone scans can show a distinct pattern of inflammation in SchS and that the scintigraphic score correlates with disease activity and treatment response. Patients with SchS often complain of non-specific, deep bone pain, and bone imaging might prove useful in the titration of anti-IL-1 therapy, beyond just measuring CRP.

Routine genetic investigations form an important part of the diagnostic work up. Even if the onset of symptoms is in adulthood, it is still important to screen for the known monogenic forms of SAID, since several conditions such as FMF, NLRP12-AID, and haploinsufficiency A20 can all present for the first time in later life. However, in the investigation of late-onset or acquired SAID, additional testing might be necessary to identify the relevant genetic changes. For example, sequencing analysis pipelines, predominantly focused on detection of germline variants in hereditary diseases, should be updated to include somatic-aware variant calling, such as MuTECT2. Furthermore, sequencing should be performed using modern, massive parallel techniques so that sufficient coverage is provided in order to detect low-level mutation down to 2–5%. Lastly, it might occasionally be necessary to perform cell separation and sequence DNA from each relevant cell type.

Whilst for some conditions, such as SchS, treatment options are well established, for others such as VEXAS the optimum medical management is poorly understood. There are a significant proportion of patients with undifferentiated SAID in whom the precise cause of disease is unknown, but who nevertheless require treatment. In such cases, empirical trials of medications typically used to treat SAID, such as colchicine and anakinra, might prove effective and safe in the long-term. In addition, their use may improve our understanding of the possible pathological mechanisms driving such conditions. In patients who fail colchicine and anti-IL-1 therapy, JAKi are increasingly used in a similar, empirical fashion.

Recent discovery of VEXAS syndrome provides a paradigm shift in thinking around the role of somatic mutations in the disease pathogenesis other than malignancy. The role of such acquired process in the development of many more common disorders is just starting to be fully appreciated.

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