Expanding structural diversity in a library of disulfide macrocycles through in-situ imide hydrolysis

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We describe here an unorthodox approach to dynamic covalent chemistry in which the initially-unexpected in-situ hydrolysis of a bis-imide is employed to control the composition of a library of structurally diverse macrocycles. A single building block is used to generate a library of numerous disulfide-based architectures in a one-pot single-step process. The dual-stimuli method is based on simultaneous changes in pH and DMSO concentration to expand the structural diversity of the macrocyclic products. Mechanistic details of this complex process are investigated by the kinetics analysis. We delivered a facile strategy for the synthesis of water-soluble, multicomponent and dynamic macrocycles equipped with number of different functional groups, thus giving a prospect of their application in guest-driven phase transfer.

Dynamic combinatorial chemistry (DCC) is of continuing interest among various fields of modern chemistry1–3. Many reversible linkages are employed in DCC, such as imines, acyl hydrazones, disulfides, boronic esters, or coordination bonds4–6. Dynamic disulfides are of particular interest due to their useful applications as chemical receptors7,8, nanocapsules for molecular transport9–11, or in metallousramolecular architectures12. Recently, there has been a growing interest in the search for new methods and conditions for the oxidation of thiols to disulfides in DCC using metalloid catalysis13,14, solvent effects15, iodine16 or mechanochemical stimuli17–19. To date, numerous water-soluble disulfide systems of distinct topologies such as macrocycles, cages, catenanes or knots have been described, most of which are based on organic cysteine-functionalized components20–29. Macrocyclic structures, especially those based on modified naphthalenediimides (NDIs) have been heavily explored in the last decade as they present interesting properties and functions such as molecular sensors, host–guest complexes, molecular devices, catalysis through π-anion interactions and non-covalent binding with DNA for bio-applications30,31. Several different strategies of controlled, stimuli-based, modification of the dynamic library composition through physical and chemical factors have been described for these systems32. However, it remains a challenge to externally control DCL compositions and product constitutions.

Here we present an unorthodox approach to disulfide DCC by employing in-situ imide hydrolysis which leads in our system to expanded library diversity and larger macrocycles. We describe a robust method for the use of a mixture of water and DMSO as an excellent environment for the oxidation and exchange of thiols and we show the control of the library composition through appropriate pH and DMSO concentration. Previously, we described new building blocks based on 3,3′,4,4′-Biphenyltetracarboxylic dianhydride (BPDA)33, similar to the NDI-based components, but with an extra degree of geometrical flexibility. One of these was the bis-imide equipped with two cysteine moieties A (Fig. 1). During the standard DCL procedure (water, pH 8.0) with A, instead of the expected macroyclic oligomer library we noticed products resulting from the hydrolysis of A.

Literature evidence on this process encouraged us to investigate the potential of this phenomenon in concert with disulfide DCC34,35. In basic aqueous solution, imide hydrolysis may take place, and results in opening of the five-membered ring, thereby providing additional features in conformational flexibility, polarity (new carboxylic groups) and intramolecular non-covalent interactions (NH hydrogen bond donor) during disulfide formation.

Results and discussion

The work began with the synthesis of dithiol component A starting from the BPDA and S-Trityl-L-cysteine. We employed our earlier reported microwave method33 to obtain an intermediate STr-protected diimide product. In the second step the trityl groups were removed by the standard Et3SiH/TFA/DCM reaction. Natural L-cysteine provides several important functions for further self-assembly, such as the carboxyl groups ensuring

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high polarity and water solubility, a stable amino-acid chiral centre, and -SH groups for oxidation located on a
conformationally labile arm.

Additionally, the component A contains a single bond between two phenyl rings, which provides additional
rotational flexibility to the entire system, a significant difference compared with rigid NDIs.

In the first experiment a simple aqueous solution of A at pH 8.5 in unsealed vial was set up (see ESI S2 for
details). After several minutes we observed complete decay of A and the formation of a single new product.
This was isolated and based on NMR and MS analysis was shown to be the unsymmetrical product of double
hydrolysis of A where the newly formed two amide groups are placed para and meta to the inter-biphenyl bond
A-2Hyd (Fig. 2). Analysis of the 1H and COSY (see Fig. S8 in ESI) spectra in DMSO clearly shows the forma-
tion of a pair of amide N–H signals (δ8.83 and δ8.75 ppm) coupled to a pair of methine C-H cysteine signals.

By chemical intuition, one would expect a symmetrical product with a double meta or para configuration.
Presumably, due to electronic and entropic factors, the system favors two-step hydrolysis path, which leads
to an unsymmetrical product (for more details see Fig.S34). To the best of our knowledge, such directional
hydrolysis has not been recorded previously for BPDA derivatives. After hydrolysis, the A-2Hyd should display
extra structural flexibility. To check how this new feature of A-2Hyd influences the formation of disulfides, we
set a new solution of A at pH 8.5 for oxidation with air exposure (Fig. 3a). The post-reaction mixture contained
mostly one product (90%), which was identified by 1H NMR spectroscopy (Fig. 4c) and ESI–MS as the mono-
protonated cation \([\text{M + H}]^+\) remarkable cyclic \(m\text{-, }\rho\text{-dimer with two disulfide bonds (A-2Hyd}) formed from
two molecules of A-2Hyd. The second component AS (10%) is the monomeric A-2Hyd derivative cyclised by a
single intramolecular disulfide bond.

To accelerate the oxidation, we used the known DMSO effect15. We started with A dissolved in pure DMSO
(Fig. 4a) with expectation of rapid and full conversion36,37. After 3 days we observed via LC–MS the selective
formation of only one product, which was identified as dimeric macrocycle A2 with two disulfide bonds (Fig. 3b).
This compound was isolated and analyzed by NMR (Fig. 4b). The 1H spectrum of the dimer A2 shows the upfield
shifts of aromatic signals characteristic of tight aromatic macrocycles, and downfield shifts of aliphatic C-H and
CH2 signals. The absence of any amide signals indicates that no hydrolysis took place. A comparison of both A2
and A2-Hyd dimers shows that A has a structural tendency to form dimeric systems.

Here we also checked whether hydrolysis is possible after thiol oxidation. The isolated A2 was dissolved in
water (5 mM, pH 8.5, r.t.) and monitored by HPLC. After 6 h the complete hydrolysis of A2 to A2-4Hyd occurred

Figure 1. Schematic representation of the DCLs formed from component A under three different conditions,
in DMSO, in water at pH 8.5, and in 5% DMSO/aqueous AcONH4 buffer. The inset contains a diagram of the
irreversible imide hydrolysis. The –COOH group resulting from the imide hydrolysis is marked as a red ellipse.
DMSO and the pH in the range from 4.0 to 8.5 in 0.1 M AcONH₄ buffer adjusted to the desired pH with AcOH abundance after approx. 2 h. Simultaneously, A take place either in meta or para position. However, these isomers are indistinguishable by LC–MS, so we were unable to determine exactly which isomers are observed in the generated DCLs.

We showed that the library composition can be controlled by pH. At slightly acidic pH (4.0), an unhydrolyzed dimer A₃ dominates; a pH close to neutral (6.5–7.25) promotes the formation of the tetramer A₄-Hyd. Due to similarity in the polarity of the library components, we were unable to isolate preparative amounts of separate products despite numerous attempts. Therefore we decided to analyze the intact DCL. The library was isolated by evaporation to dryness and analyzed by NMR (Fig. 4d). The ¹H spectrum of the library contains significantly more signals (in comparison to A and A₃) indicating a loss of symmetry of the trimeric and tetrameric products. In the COSY spectrum, tiny N–H signals coupled with the C-H signals (similar to A-Hyd) are observed. This is additional proof for the presence of amides in the structures of some mono-hydrolyzed macrocycles in DCL.

We were excited to get a deeper understanding of the system kinetics and mechanistic features. We employed HPLC to monitor the DCL equilibration over time (1 ml volume, pH 6.5, 5% DMSO, 5 mM of A). The DCL was analyzed every 30 min for 24 h at room temperature. The first injection (t₀) has been done immediately after A was dissolved in the reaction buffer. The distribution curves of individual species during the equilibration were plotted, based on the integrations of the relative peak areas (RPA) from the collected chromatograms (Fig. 5).

It revealed that the system reaches equilibrium just after approx. 10 h and does not change any further up to 24 h. Several preliminary conclusions concerning the kinetics and mechanism of this complex process can be drawn. Firstly, substrate A (brown plot) irreversibly hydrolyzes to A-Hyd (reaction I, grey) and reach 10% abundance after approx. 2 h. Simultaneously, A dimerizes into a linear AA intermediate (II, black) with a single S–S bond and reaches maximum abundance of about 20% after approx. 1.5 h. Then AA undergoes intramolecular cyclization to macrocyclic A₂ (IV, green). It seems that the synthesis of the larger macrocycles A₃ (gold) and A₄ (yellow) depends on the presence of AA and A₃ (VI-IX) because they appear in the mixture only after 2 h. No

(Fig. 1, top). Placing the sample of A₂-4Hyd in pure DMSO (5 mM, r.t.) did not lead to A₃ dimide again, confirming the irreversible nature of the hydrolysis process under such conditions.

Based on the results so far, we decided to use a combination of pH control and DMSO in an attempt to expand to DCLs with larger macrocycles (Fig. 3c–f). To find the right conditions, we screened both concentration of DMSO and the pH in the range from 4.0 to 8.5 in 0.1 M AcONH₄ buffer adjusted to the desired pH with AcOH and ammonia (see Figs. S28–30).

It became clear that pH had the greatest influence on the DCL product distribution. In this system, the most interesting results occur for pH 6.5 and 7.25, which fits with the usual optimal thiol oxidation conditions (Fig. 3e–f)³⁸. pH 8.5 is usually applied to fully deprotonate the COOH groups and thus achieve water solubility and ammonia (see Figs. S28–30).

Our combined method with the appropriate pH and the DMSO as oxidation accelerator resulted in a multi-component library containing trimeric and tetrameric macrocycles. Based on LC–MS analysis (Fig. 3g), we found that in most cases (pH 5.0–7.25) a dynamic combinatorial library of six products was formed, which were identified as dimers, trimers and tetramers (as monoprotonated species [M + H]+). These are three pairs of dimeric (A₂ and A₂-Hyd), trimeric (A₃ and A₃-Hyd) and tetrameric (A₄ and A₄-Hyd) macrocycles, in which the first member is completely unhydrolyzed, while the second has one hydrolyzed imide group (based on MS). In the case of mono-hydrolyzed products, it should be noted that the ring-opening during the hydrolysis can take place either in meta or para position. However, these isomers are indistinguishable by LC–MS, so we were unable to determine exactly which isomers are observed in the generated DCLs.
traces of the expected AAA and AAAA linear intermediates were found in the obtained HPLC–MS data. However, intermediates AAA and AAAA seem to be an obvious step in chain elongation and ring closure processes. Therefore, it should be assumed that these individuals are formed in a mixture from the smaller molecules (A and AA) and are immediately consumed in the cyclization reaction to the macrocyclic products (A₂ and A₃).

Similar results were observed for mono-hydrolyzed macrocycles, which seem to be strongly dependent from the A-Hyd concentration. The A-Hyd reacts with available A and forms AA-Hyd intermediate (III), which is rapidly intermolecularly cyclized into the major mono-hydrolyzed A₂-Hyd macrocycle (V, red). The synthesis of the...

Figure 3. (a–f) HPLC chromatograms (254 nm) of 3 days old A DCLs in various conditions of DMSO concentration and pH. The library material distribution is represented as percentages next to the corresponding peaks (relative peak area). The peak colour indicates each macrocycle. See Figs. S17–S24 for LC–MS details. (g) HR-MS analysis of macrocycles as ions [M + H]⁺ (or [M + NH₄]⁺ for A₃, A₃-Hyd, see Figs. S18–27).
Figure 4. Comparison of $^1$H NMR spectra (300 MHz, 298 K, DMSO-d$_6$) of (a) A, (b) A$_2$, (c) A$_2$-4Hyd (contaminated with AS) and (d) DCL mixture from Fig. 3f. Ties show coupled N–H signals with C–H signals (based on COSY, see Figs. S32–33).

Figure 5. Kinetic plots of A DCL equilibration at pH 6.5 (substrate and products), and scheme of crucial reactions in the equilibrating mixture.
**Figure 6.** (a) synthesis scheme of the dimeric macrocycle $B_2$ and comparison of $^1$H NMR spectra (300 MHz, 298 K, DMSO-$d_6$) of (b) B and (c) $B_2$.

$A_3$-Hyd (blue) and $A_4$-Hyd (violet) may go through various pathways (X-XIII). It could also be observed that the DMSO accelerated oxidation and imides hydrolysis run in parallel with a slight vantage of first one during the first two hours. It confirms our main hypothesis that both reactions occur orthogonally during the entire process of DCL equilibration. The final ratio of the hydrolyzed to non-hydrolyzed products is approx. 3:6 (see Fig. 3f). The observed unusual imide hydrolysis in slightly acidic conditions may be powered by the presence of thiols. There are examples in the literature of the thiol-catalyzed hydrolysis of acyl groups. We observed that the sample of unhydrolyzed $A_3$ placed in pH 6.5 buffer has not changed over time (3 days). The addition of NaOH to pH 8.5 resulted in a rapid hydrolysis into the $A_2$-4Hyd (as shown in Fig. 1). These results indicate that the hydrolysis in DCLs at pH 4–6.5 is dependent on the presence of unoxidized thiols. The latter significantly slow down the hydrolysis process or stop it completely when the oxidation is complete.

Thus, we have shown that the use of a cysteine-modified phthalimide analogue gives unusual properties and allows the formation of multi-component DCLs. In order to demonstrate that the dynamics of the described system are strongly dependent on the hydrolysis of imides, and are not a property of the biphenyl structure, we performed a control experiment. We synthesized component B based on modified biphenyl-4,4′-dicarboxylic acid with two cysteine moieties (Fig. 6).

This compound is linear and has no imide groups but is otherwise analogous to component A. We repeated all the experiments that were described earlier with A. Under all pH and DMSO conditions only the cyclic dimer $B_2$ was observed as the result of oxidation. Those experiments unequivocally confirmed the correlation between imide presence and component A activity.

**Conclusions**

We have successfully demonstrated that the use of hydrolysable imides in dynamic disulfide chemistry can effectively act to enhance disulfide exchange factor for building multi-component DCLs. Additionally, we have shown that this effect could be externally modulated by pH and DMSO concentration. Until now, DMSO has been considered only as an oxidation accelerant, and we have shown that by changing the DMSO concentration, the library composition could be controlled. The presented methodology can be applied in the generation of new dynamic molecular transporters or receptors (e.g. cages or macrocycles) in which the water solubility could be modulated through a controlled process of imides hydrolysis. Finally, we delivered new insights into the kinetics and mechanistic properties of such type of complex disulfide system.

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Author contributions
M.K. performed all experiments (synthesis, LC–MS, HPLC, NMR analysis) and co-wrote the paper. A.R.S. designed the project, interpreted results and co-wrote the paper.

Competing interests
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