Effects of taurine on resting-state fMRI activity in spontaneously hypertensive rats

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Abstract

Attention deficit hyperactivity disorder (ADHD) is a global behavior illness among children and adults. To investigate the effects of taurine on resting-state fMRI activity in ADHD, a spontaneously hypertensive rat (SHR) animal model was adopted. Significantly decreased serum C-reactive protein (CRP) was detected in rats of Wistar Kyoto (WKY) high-taurine group and significantly decreased interleukin (IL)-1β and CRP were detected in rats of SHR low-taurine and high-taurine groups. Moreover, significantly higher horizontal locomotion was detected in rats of WKY low-taurine and SHR low-taurine groups than in those of controls. In contrast, significantly lower horizontal locomotion was detected in rats of the SHR high-taurine group than in rats of the SHR control group. Additionally, significantly lower functional connectivity (FC) and mean amplitude of low-frequency fluctuation (mALFF) in the bilateral hippocampus in rats of WKY high-taurine and SHR high-taurine groups was detected. Notably, the mALFF in rats of the SHR low-taurine and high-taurine groups was significantly lower than in those of the SHR control group. These findings suggest that the administration of a high-dose taurine probably improves hyperactive behavior in SHR rats by ameliorating the inflammatory cytokines and modulating brain functional signals in SHR rats.

Introduction

Attention deficit hyperactivity disorder (ADHD) is a very common developmental disorder in both children and adults worldwide, with prevalences of 5–10% [1] and 3–5% [2], respectively. Although the exact etiology of ADHD is still unclear, various causes of the pathogenesis of ADHD have been suggested. Inflammation has been associated with various neuropsychiatric illnesses, including ADHD. Elevated pro-inflammatory cytokines such as interleukin (IL)-1, IL-6, CRP and tumor necrosis factor alpha (TNF-α), are known as common pathogenic parts.
of schizophrenia, ADHD and autism [3]. Children with extremely prematurely birth, recurrent, or persistently inflammation during the first 14 postnatal days are strongly associated with attention deficiency [4]. Accordingly, a systematic review article regarding inflammation among young people with neuropsychiatric diseases reveals that elevated inflammatory markers play critical roles in the development of ADHD [5].

Magnetic resonance imaging (MRI) has been utilized to evaluate the anatomic differences in brain areas between ADHD children and their controls [6–7]. Overall reductions in total brain volume have been reported in ADHD children than those of age- and gender-matched controls [8]. Indeed, evidences consistently show that the average brain volume in ADHD children is significantly smaller than in healthy controls [9]. Brain areas of particular interest in this regard are the prefrontal regions, basal ganglia, corpus callosum, and cerebellum. For instance, differences in distributions of grey and white matter in children with ADHD and healthy controls have been reported [10]. Since specific executive function impairments in particular brain areas of ADHD patients have been associated with the impaired behavioral phenomena [11], functional MRI (fMRI) was employed herein has been used as the method of choice to provide superior spatial resolution in ADHD cases. Based on fMRI measurement, the pathogenesis of ADHD is likely to involve a reduction in volume or function in specific brain areas, resulting in various behavioral problems such as abnormal cognitive processing, attention, motor planning, and speed of processing responses [12].

Taurine, which is known to be the richest amino acid in the central nervous system, performs various functions in the body, including antioxidant, anti-inflammation, osmoregulation, neuromodulation, membrane stabilization, embryogenesis and immune regulation [13–15]. Increasing wariness has recently been paid to the role of taurine in neuro-regulation. Taurine is an endogenous ligand for the glycine receptor in the nucleus accumbens, elevating the dopamine level in this area [16]. Additionally, taurine is reported to induce anti-anxiety effects in an animal model probably owing to its association with GABA and has been used as a natural anxiolytic, or anti-anxiety compound, although some studies reveal mixed results [17–19]. These findings strongly associate taurine with various ADHD-related neurotransmitters. Therefore, we intend to study the effects of taurine on serum inflammatory factors, horizontal locomotion and brain functional activity in an ADHD-like animal model.

**Materials and methods**

**Animals and diets**

As a valid and accepted animal model for ADHD, the spontaneously hypertensive rat (SHR) and its control, the Wistar-Kyoto (WKY) rat, were used in this study [20–21]. Three week-old male SHR/NcrCrlj and WKY/NcrCrlj rats (with a body weight of approximately 180g) were purchased from the National Laboratory Animal Center, Taipei, Taiwan. Rats of each the two species were separated into control (Control), low taurine (Low Tau) and high taurine (High Tau) groups (eight rats /group) and housed in an animal room at 22±2˚C with a 12/12 h light-dark cycle. All protocols were followed under the supervision of the Institutional Animal Care and Use Committee at Chung Shan Medical University (IACUC approval number: 1671), following the Guide for the Care and Use of Laboratory Animals that was published by the United States National Institutes of Health. The dosage of taurine used in this study is according to a previous study [19]. All animals were provided with chow diets (Laboratory Rodent Diet 5001, PMI Nutrition International/Purina Mills LLC., USA) and water for one week for acclimation. Subsequently, the rats in the Control, Low Tau and High Tau groups were fed a chow diet, a low taurine diet (22.5 mmol/kg taurine), and a high taurine diet (45 mmol/kg taurine) for four weeks, respectively. Before the animals were sacrificed, horizontal locomotion
and rs-fMIR was performed. Subsequently, the animals were sacrificed at an age of eight weeks by CO$_2$ asphyxiation. Blood was obtained from the heart and stored at -80°C before analysis.

**Measurement of horizontal activity in an open-field assay**

Horizontal activity was detected as described elsewhere [22]. All rats were set in an open-field device (acrylic cylinder; 40 cm diameter × 40 cm height) for two consecutive days prior to the test day to diminish any influences of the treating process. Briefly, each rat was put in the middle of the apparatus under ambient lighting, and then allowed to explore the open field for 90 minutes in the first two days. On day three, the rat was placed in the same open-field device. Horizontal activity was measured for 90 minutes as the total number of interruptions of the beam of a horizontal sensor (SCANET MV-10; Melquest Co., Ltd, Toyama, Japan). All behavioral evaluations were performed between 08:00 and 15:00 h.

**ELISA estimations for IL-1β and CRP**

Serum CRP and IL-1β concentrations were measured in triplicate with a commercially available enzyme-linked immunosorbent assay kit (BD Pharmingen, San Diego, CA, USA) according to the instruction of manufacturer.

**Functional MRI acquisition**

Whole brain images were acquired from all subgroups mentioned above and scanned by 7T MRI (Bruker BioSpin, Ettlingen, Germany). In preparation, each rat was anesthetized with 3.5% isoflurane mixed with 800 ml/min air. Before MRI experiments a short-acting tranquilizer and synthetic medicine, Domitor (0.1 ml/500 g), was injected. The sedation effect of Domitor was about half hour to one hour. It is commonly used in animal surgery and fMRI study to avoid BOLD signal contamination by isoflurane. During experiments, the temperature was maintained at around 37°C using hot pad. All functional images were acquired by gradient echo based echo planar imaging (EPI) with the following parameters: repetition time/echo time (TR/TE) = 2000ms /20 ms, resolution (voxel size) = 3.12 x 3.12 x 1 mm$^3$, slice number = 12, number of repetition = 300, and the scan time = 10 min.

**Functional image pre-processing**

The raw data were first converted into ANALYZE format. The preprocess was then performed, including slice timing, realignment, denoise, detrending and filtering, using Statistical Parametric Mapping (SPM8, Wellcome Department of Cognitive Neurology, London, UK) and Resting-State fMRI Data Analysis Toolkit (REST1.8, Lab of Cognitive Neuroscience and Learning, Beijing Normal University, China) [23]. For motion correction, all EPI images in each run were realigned to first image. To reduce the noise, all data were spatially smoothed by a Gaussian kernel with full width at half maximum (FWHM) of [0.6 mm, 0.6 mm, 2 mm]. To cut down the influences of low-frequency linear and quadratic drifts and physiological signals, detrending and band-pass temporal filtering (0.01–0.12 Hz) were adopted on the time series of each voxel.

**Functional connectivity**

Functional connectivity analysis is a seed-based analysis that can observe the connectivity between brain regions that share functional properties. It indicates selecting regions of interest (ROIs) and correlating the average blood oxygenation level dependent (BOLD) time course of voxels within these ROIs with each other as well as with the time courses of all other voxels.
Using the rat brain in stereotaxic coordinates as a reference, seed region was prescribed for each choosing brain area center, then extract a spherical region with a 0.5mm radius. Spherical seeds were prescribed for each network. We used the posterior cingulate cortex (PCC) as the seed region to identify the default mode network (DMN) [24]. In addition, we used the bilateral amygdala, hippocampus, thalamus and motor as seed points to construct the functional connectivity map. To assess bilateral connectivity, we calculated the correlation between seed point and other sides. A circular ROI was selected on the contralateral side of the seed regions, with the shortest axis of the brain area as the radius; the ROI size in each group was controlled to be the same. We used Functional Magnetic Resonance Imaging of the Brain Software Library (FMRIB Software Library, FSL, Oxford, UK) with FSL-View to make the 3D mask for calculating the correlation between seed points and other sides, and the mean signals in the functional connectivity map. Finally, the average z-scores of the two groups were determined.

**Amplitude of low frequency fluctuations**

The amplitude of low frequency fluctuations (ALFF) calculates low-frequencies spectral power (0.01–0.12 Hz) of BOLD signal voxel-by-voxel [25]. The ALFF measures the correlation of low frequency fluctuations of BOLD signal at each brain region, which is used to proxy the absolute intensity of spontaneous brain activity [26]. The ALFF have numerous similarities with fluctuations in neural metabolic, hemodynamic, and neurophysiological parameters [27]. Therefore, the ALFF during resting state is considered to be physiologically meaningful and reflective of spontaneous neural activity [28]. For calculating mALFF, the time series was first converted to the frequency domain using a Fast Fourier Transform for a given voxel. The square root of the power spectrum was measured, averaged and normalized across a predefined frequency interval, which was termed the ALFF at the given voxel [28]. To reduce the global effects of variability across rats, the ALFF of each voxel was divided by the global mean ALFF value for each subject, resulting in a relative ALFF, mALFF. We then performed two-sample t-tests with FDR correction to assess the difference in mALFF between the SHR and WKY rats.

**Statistical analysis**

All data are presented as mean ± standard error. Two-way analysis of variance (ANOVA) followed by Dunnett’s test was performed to evaluate the horizontal activity data. The comparisons among groups were performed using GraphPad Prism 5 software (GraphPad Software, Inc., La Jolla, CA, USA) by one-way analysis of variance (One-way ANOVA) followed by Tukey multiple comparisons test. A value of P < 0.05 was considered statistically significant. Additionally, to investigate the associations between fMRI indices (FC and mALFF) and behavior/biochemical measures (horizontal activity, IL-1β and CRP), the scatter plots were performed.

**Results**

**Effects of taurine on horizontal locomotion in WKY and SHR rats**

The effect of taurine on locomotion of SHR and WKY rats was examined by detecting their horizontal locomotion. Rats of the SHR control group exhibited significantly higher horizontal locomotion than those of the WKY control group (Fig 1). Significantly higher horizontal locomotion was detected in rats of WKY low-taurine than in those of the WKY control group (Fig 1). Similar result was observed that SHR rats that were fed with low-dose taurine exhibited significantly higher horizontal locomotion than in those of the SHR control group (Fig 1).
Conversely, the SHR rats that were fed with a high dose of taurine exhibited significantly less horizontal locomotion than in those of the SHR control group (Fig 1).

Effects of taurine on serum IL-1β and CRP levels in WKY and SHR rats

Elevation of inflammatory factors induces the development of neurological and mental pathologies, including ADHD. This study further investigates the effects of taurine on inflammatory factors, including IL-1β and CRP, in both SHR and WKY rats. Significantly higher level of serum IL-1β was detected in rats of the SHR control group than in those of the WKY control group (Fig 2A). No significant difference between serum IL-1β levels was detected among WKY rats that were fed with low or high dose of taurine and those of the control group (Fig 2A). A significantly lower serum IL-1β levels were detected in SHR rats that were fed with low or high dose of taurine that in those of the SHR control group (Fig 2A). Significantly higher level of serum CRP was detected in rats of the SHR control group than in those of the WKY control group (Fig 2B). Significantly lower serum CRP level was detected in rats of the WKY high-taurine group than in those of the WKY control group (Fig 2B). Notably, the serum CRP level in rats of SHR low and high taurine groups were significantly lower than in those of the SHR control group (Fig 2B).

Effects of taurine on functional connectivity

To investigate the effects of taurine on functional signals in various brain regions of SHR and WKY rats, the signal of functional connectivity (FC) was detected using resting-state
Effects of taurine in ADHD animal model

**Fig 2.** Serum (A) IL-1β and (B) CRP levels in WKY and SHR rats. Both WKY and SHR rats were divided into three subgroups, including control group (Control), low dose taurine group (Low Tau), and high dose taurine group (High Tau). The data are expressed as the mean ± standard error. The symbols, * (WKY Control vs. SHR Control), # (SHR Low Tau or SHR High Tau vs. SHR Control) and & (WKY High Tau vs. WKY Control), indicate statistically significant (P<0.05).

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functional magnetic resonance imaging (rs-fMRI). The FC maps are presented as individual animals, which were composed of six individual representative rat brain maps, respectively. The color bar represents z-scores. In Fig 3, the FC of the bilateral hippocampus in rats of the SHR control group was significantly higher than in those of WKY control group. The FC of the bilateral hippocampus in rats of the WKY high-taurine group was significantly lower than in those of the WKY control group (Fig 3B). However, no significant variation of the FC in the bilateral hippocampus was detected in rats of the SHR low-taurine and high-taurine groups than in those of the SHR control group (Fig 3B). No significant variation in the FC of the other brain regions was detected among the groups (data not shown).

Effects of taurine on mean amplitude of low-frequency fluctuations

The signal of mean amplitude of low-frequency fluctuation (mALFF) was detected to clarify the effects of taurine in various regions of rat brain. The mALFF maps are presented as individual animals, which were composed of six individual representative rat brain maps, respectively. The color bar represents z-scores. In Fig 4, the mALFF of the bilateral hippocampus in rats of the SHR control group was significantly higher than that in rats of the WKY control group. The mALFF of the bilateral hippocampus in rats of the WKY high-taurine group was significantly lower than in those of the WKY control group (Fig 4B). After treatment with a low or high dose of taurine, the mALFF of the bilateral hippocampus in rats of the SHR low-taurine and high-taurine groups was significantly lower than in those of the SHR control group (Fig 4B). No significant variation in mALFF in the other brain regions was detected among groups (data not shown).

Correlative analysis of fMRI indices biochemical measures

To investigate the correlation between fMRI indices, including FC and mALFF, and biochemical measures, including horizontal activity, IL-1β and CRP, the scatter plots were performed. Accordingly, positive correlation between FC and IL-1β (r = 0.85), FC and CRP (r = 0.90), mALFF and IL-1β (r = 0.98), and mALFF and CRP (r = 0.99) were detected (Fig 5). However, no significant correlation was found between FC and horizontal activity or mALFF and horizontal activity (data not shown).

Discussion

Accumulating evidence suggests a strongly pathophysiological relationship between inflammation and attention-deficit hyperactivity disorder (ADHD) [5]. Elevated levels of pro-inflammatory cytokines such as interleukin (IL)-1 and C-reactive protein (CRP) are known to be diagnostic markers and crucial pathogenic factor of ADHD [3, 29–30]. When cell damage occurs, CRP binds to phosphocholine on cell membranes and promotes the binding of complements [31]. Evidence reveals that IL-1 has both neuromodulatory and neurodevelopmental functions that involve turnover neurotransmitters in various brain regions [32]. Interleukin-1 (IL-1α and IL-1β), IL-1Ra, and IL-1 receptors are found to be expressed in the brain [33]. Especially in relation to ADHD, IL-1β induces changes in dopamine (DA) and norepinephrine (NE) in the prefrontal cortex. Various studies have also shown that the systemic administration of IL-1β enhances DA and NE utilization in the prefrontal cortex (PFC) in both mice and rats [32, 34]. In this work, serum CRP and IL-1β levels were significantly lower in SHR rats that were administered a low dose or high dose of taurine. These results suggest that taurine can ameliorate inflammatory factors, resulting in ameliorating ADHD-like behaviors, such as hyperactivity, in SHR rats.
Fig 3. The functional connectivity of bilateral hippocampus in WKY and SHR rats. The (A) representative images of functional connectivity (FC) in bilateral hippocampus of (a-c) WKY and (d-f) SHR rats. Both WKY and SHR rats were divided into three subgroups, including (a, d) control group (Control), (b, e) low dose taurine group (Low Tau), and (c, f) high dose taurine group (High Tau). (B) Bar graphs showing the functional connectivity (FC) values for each group. The asterisk (*) indicates a statistically significant difference (p < 0.05) compared to the control group, and the ampersand (&) indicates a significant difference (p < 0.05) between the low and high dose taurine groups.
Functional magnetic resonance imaging (fMRI) is a specialized MRI method that is used to measure hemodynamic responses; it provides images of the brain microvasculature in which the image contrast reflects blood oxygen level [35]. The signals of blood-oxygen-level-dependent (BOLD) dominate the mechanism that gives rise to functional connectivity in the resting human brain [36]. Low-frequency temporal components (<0.1Hz) in resting-state BOLD fMRI reflect spontaneous fluctuations in brain physiology and metabolism, which regards as imaging markers of brain function [37]. Several indices of spontaneous low-frequency fluctuations of BOLD fMRI, including the amplitude of low-frequency fluctuations (ALFF) [38], regional homogeneity (ReHo) and functional connectivity (FC) [39], have been introduced to evaluate resting brain function in patients with various psychiatric disorders, such as ADHD [40]. Evidence suggests that increased hippocampus volume may reflect neural activities such as temporal processing and delay aversion within the hippocampus in ADHD [8]. However, these findings are not very stable among samples from different ADHD patients and the contradictory findings may be related to the different locations of alterations in the complex circuits that are responsible for the various symptoms of ADHD [41–42]. A previous study reported that ADHD patients exhibited higher ALFF values in the left superior frontal gyrus and sensorimotor cortex (SMC) and suggested irregular frontal activities in the resting state that are associated with the underlying physiopathology of ADHD [43]. Other evidence indicates that ADHD patients exhibit impaired executive function and abnormal fMRI indices, such as increased ALFF in both left and right globus pallidus and right dorsal superior frontal gyrus, and increased FC in the frontostriatal circuit, relative to healthy controls [44]. Notably, the SHR rats that were fed with taurine exhibited significantly lower mALFF values than those in SHR control group in the hippocampus, as well as less horizontal locomotion and lower levels of inflammatory proteins. These findings suggest that taurine mitigates mALFF in SHR rats and probably result in reduced ADHD-like behavior in SHR rats. However, further investigations are required to elucidate the precise associations between fMRI indices and ADHD symptoms.

An interesting result was observed in this study. The horizontal locomotion of SHR rats that were fed with a low dose of taurine was significantly higher than that of rats of the SHR control group. However, the SHR rats that were fed with a high dose of taurine exhibited significantly less horizontal locomotion, owing to the effect of the dose of taurine. Accordingly, a recent study found that glycine receptors were activated by administrating a low-dose taurine (0.5 mM), whereas a higher dose of taurine (3 mM) induced both glycine and gamma-aminobutyric acid A (GABAA) receptors in preoptic hypothalamic area (PHA) neurons [45]. Another study reported that taurine supplementation increases locomotor activity and anxiety whereas taurine injection suppressed locomotor activity and anxiety [18]. It indicates that taurine could induce opposite effects through different ways of administration. However, investigations are needed to clarify whether high-dose taurine supplementation is similar to the effect of lower-dosage taurine injection. Although further works are required to elucidate the influences of different dosages of taurine as well as its administrative methods, these findings may provide a possible explanation of why taurine has ameliorate effects on ADHD-like behaviors in SHR rats.

Although the cause of ADHD remains unclear, impaired prefrontal cortex (PFC) circuits plays crucial roles in the pathogenesis of ADHD [46]. Pharmacotherapy to treat ADHD is based on evidence that norepinephrine and dopamine are extensively involved in various
Fig 4. The mALFF of bilateral hippocampus in WKY and SHR rats. The (A) representative images of mALFF in hippocampus of (a-c) WKY and (d-f) SHR rats. Both WKY and SHR rats were divided into three subgroups, including (a, d) control group (Control), (b, e) low taurine group (Low Tau), and (c, f) high taurine group (High Tau). (B) The mean amplitude of low-frequency fluctuations (mALFF) of hippocampus was measured and compared between WKY and SHR rats. The mALFF was significantly higher in SHR rats compared to WKY rats in both hippocampus regions (R and L), and the effect was more pronounced in the high taurine group.
functional brain regions such as motor control, working memory, attention, and executive function [47]. Therefore, a reduced GABA concentration was detected in cases of ADHD and associated with its pathogenesis [48]. Taurine is the second most abundant neurotransmitter in the central nervous system, which plays important roles in brain neuronal proliferation, stem cell proliferation, and differentiation [49]. Clinically, taurine can be directly used in the treatment of brain development because it has no toxic effects on humans [49]. Also, taurine can interact with dopamine in the striatum [50] and induces the activation of extrasynaptic GABA A receptors in the thalamus [51], taurine can elevate dopamine levels in the nucleus accumbens [16, 52]. The above findings suggest diverse roles of taurine in modulating ADHD-related neurotransmitters. Although further investigations are needed to confirm the precise

![Graphs showing correlations between fMRI indices and inflammatory-cytokine levels in SHR rats.](https://doi.org/10.1371/journal.pone.0181122.g005)
effect of taurine on neurotransmitters in cases of ADHD, and the multiple modulatory effects of taurine, especially in activating the GABA A receptor and elevating dopamine levels, this study may provide a rational explanation of taurine’s amelioration of ADHD-like symptoms in SHR rats by modulating neurotransmitters, such as GABA, dopamine and glutamate or their receptors, in the brain.

**Conclusion**

This study is the first to reveals the beneficial effects of taurine on an ADHD animal model. The treatment of SHR rats with a high dose of taurine significantly reduces the levels of pro-inflammatory cytokines and horizontal locomotor activity. Treatment with a low or high dose of taurine caused the FC of the bilateral hippocampus in SHR rats to return to the levels in rats of the WKY control group. Taurine significantly reduces the mALFF of bilateral hippocampus in SHR rats. Although no significant correlation was found between FC and horizontal activity or mALFF and horizontal activity, this study still disclosers the ameliorative effect of high-dose taurine on hyperactivity in SHR rats and suggests that taurine may be an alternative treatment for ADHD. However, further animal and clinical studies are required to determine the efficacy, safety, optimal dosage and precise mechanism of taurine for clinical purposes.

**Author Contributions**

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**References**

1. Polanczyk G, deLima MS, Horta BL, Biederman J, Rohde LA. The worldwide prevalence of ADHD: A systematic review and meta regression analysis. Am J Psychiatry. 2007; 164: 942–948. https://doi.org/10.1176/appi.ajp.2007.164.6.942 PMID: 17541055

2. Kessler RC, Adler LA, Barkley R, Biederman J, Conners CK, Demler O, et al. The prevalence and correlates of adult ADHD in the United States: Results from the National Comorbidity Survey Replication. Am J Psychiatry. 2006; 163(4): 716–723. https://doi.org/10.1176/appi.ajp.163.4.716 PMID: 16585449

3. Zubarev OE, Klimenko VM. Elevation of proinflammatory cytokines level at early age as the risk factor of neurological and mental pathology development. Ross Fiziol Zh Im I M Sechenova. 2011; 97: 1048–1059. PMID: 22292271

4. O’Shea TM, Joseph RM, Kuban KC, Allred EN, Ware J, Coster T, et al. Elevated blood levels of inflammation-related proteins are associated with an attention problem at age 24 mo in extremely preterm infants. Pediatr Res. 2014; 75: 781–787. https://doi.org/10.1038/pr.2014.41 PMID: 24614800

5. Mitchell RH, Goldstein BI. Inflammation in children and adolescents with neuropsychiatric disorders: a systematic review. J Am Acad Child Adolesc Psychiatry. 2014; 53: 274–296. https://doi.org/10.1016/j.jaac.2013.11.013 PMID: 24563356

6. Hynd GW, Semrud-Clikeman M, Lorys AR, Novey ES, Eliopoulos D. Brain morphology in developmental dyslexia and attention deficit disorder/hyperactivity. Arch Neurol. 1990; 47: 919–926. PMID: 2375699
7. Plessen KJ, Bansal R, Zhu H, Whitteman R, Amat J, Quackenbush GA, et al. Hippocampus and amygdala morphology in attention-deficit/hyperactivity disorder. Arch Gen Psychiatry. 2006; 63: 795–807. https://doi.org/10.1001/archpsyc.63.7.795 PMID: 16818869

8. Frodl T, Skozauskaus N. Meta-analysis of structural MRI studies in children and adults with attention deficit hyperactivity disorder indicates treatment effects. Acta Psychiatr Scand. 2012; 125: 114–126. https://doi.org/10.1111/j.1600-0447.2011.01786.x PMID: 22118249

9. Durston S, Hulshoff Pol HE, Casey BJ, Giedd JN, Buitelaar JK, van Engeland H. Anatomical MRI of the developing human brain: What have we learned? J Am Acad Child Adolesc Psychiatry. 2001; 40: 1012–1020. https://doi.org/10.1097/00004583-200109000-00009 PMID: 11556624

10. Krain AL, Castellanos FX. Brain development and ADHD. Clin Psychol Rev. 2006; 26: 433–444. https://doi.org/10.1016/j.cpr.2006.01.005 PMID: 16480802

11. Barkley RA. Behavioral inhibition, sustained attention, and executive functions: constructing a unifying theory of ADHD. Psychol Bull. 1997; 121: 65–94. PMID: 9000892

12. Cortese S. The neurobiology and genetics of attention-deficit/hyperactivity disorder (ADHD): what every clinician should know. Eur J Paediatr Neurol. 2012; 16: 422–433. https://doi.org/10.1016/j.ejpn.2012.01.009 PMID: 11843263

13. Lima L, Obregon F, Cubillos S, Fazzino F, Jaimes I. Taurine as a micronutrient in development and regeneration of the central nervous system. Nutr Neurosci. 2001; 4: 439–443. PMID: 11843263

14. Bidri M, Chaoy P. Taurine: a particular amino acid with multiple functions. Ann Pharm Fr. 2003; 61: 385–91. PMID: 14639190

15. Schuller-Levis GB, Park E. Taurine and its chloramine: modulators of immunity. Neurochem Res. 2004; 29: 117–126. PMID: 14992270

16. Ericson M, Molander A, Stomberg R, Söderpalm B. Taurine elevates dopamine levels in the rat nucleus accumbens; antagonism by strychnine. Eur J Neurosci. 2006; 23: 3225–3229. https://doi.org/10.1111/j.1460-9568.2006.04868.x PMID: 16820013

17. Zhang CG, Kim SJ. Taurine induces anti-anxiety by activating strychnine-sensitive glycine receptor in vivo. Ann Nutr Metab. 2007; 51: 379–386. https://doi.org/10.1159/000107687 PMID: 17728537

18. El Idrissi A, Boukarou L, Heany W, Malliaros G, Sangdee C. Effects of taurine on anxiety-like and locomotor behavior of mice. Adv Exp Med Biol. 2009; 643: 207–215. PMID: 19239151

19. Toyoda A, Iio W. Antidepressant-like effect of chronic taurine administration and its hippocampal signal transduction in rats. Adv Exp Med Biol. 2013; 775: 29–43. https://doi.org/10.1007/978-1-4614-6130-2_3 PMID: 23392922

20. Ueno KI, Togashi H, Mori K, Matsumoto M, Ohashi S. Behavioral and pharmacological relevance of stroke-prone spontaneously hypertensive rats as an animal model of a developmental disorder. Behav Pharmacol. 2002; 13: 1–13. PMID: 11990715

21. Sagvolden T, Russell VA, Aase H, Johansen EB, Farshbaf M. Rodent models of attention-deficit/hyperactivity disorder. Biol Psychiatry. 2005; 57: 1239–1247. https://doi.org/10.1016/j.biopsych.2005.02.002 PMID: 15949994

22. Umehara M, Molander A, Stomberg R, Söderpalm B. Taurine elevates dopamine levels in the rat nucleus accumbens; antagonism by strychnine. Eur J Neurosci. 2006; 23: 3225–3229. https://doi.org/10.1111/j.1460-9568.2006.04868.x PMID: 16820013

23. Song XW, Dong ZY, Long XY, Li SF, Zuo XN. REST: a toolkit for resting-state functional magnetic resonance imaging data processing. PLoS One. 2011; 6: e25031. https://doi.org/10.1371/journal.pone.0025031 PMID: 21949842

24. Wu CW, Gu H, Lu H, Stein EA, Chen JH. Frequency specificity of functional connectivity in brain networks. Neuroimage. 2008; 42: 1047–1055. https://doi.org/10.1016/j.neuroimage.2008.05.035 PMID: 18632288

25. Zou QH, Zhu CZ, Yang Y, Zuo XN, Long XY. An improved approach to detection of amplitude of low-frequency fluctuation (ALFF) for resting-state fMRI: fractional ALFF. J Neurosci Methods 2008; 172: 137–141. https://doi.org/10.1016/j.jneumeth.2008.04.012 PMID: 18501669

26. Zhang YF, He Y, Zhu CZ, Cao QJ, Sui MQ, Liang M, et al. Altered baseline brain activity in children with ADHD revealed by resting-state functional MRI. Brain Dev 2007; 29: 83–91. https://doi.org/10.1016/j.braindev.2006.07.002 PMID: 18919409

27. Zhang X, Zhu X, Wang X, Zhu X, Zhong M. First-episode medication-naïve major depressive disorder is associated with altered resting brain activity in the affective network. PLoS One. 2014; 9: e85241. https://doi.org/10.1371/journal.pone.0085241 PMID: 24416367

28. Yue Y, Jia X, Hou Z, Zang Y, Yuan Y. Frequency-dependent amplitude alterations of resting-state spontaneous fluctuations in late-onset depression. Biomed Res Int 2015; 2015: 505479. https://doi.org/10.1155/2015/505479 PMID: 25705666
29. Lovell B, Moss M, Wetherell M. The psychosocial, endocrine and immune consequences of caring for a child with autism or ADHD. Psychoneuroendocrinology 2012; 37: 534–542. https://doi.org/10.1016/j.psyneuen.2011.08.003 PMID: 21889267

30. Holtmann M, Poustka L, Zepf FD, Manaschewski T, Priller J. Severe affective and behavioral dysregulation in youths is associated with a proinflammatory state. Psychother 2013; 41: 393–399.

31. Hennig EM. C-reactive protein and biological defense. Tidsskr Nor Laegeforen 1995; 115: 1632–1635. PMID: 7778080

32. Lacosta S, Merali Z, Anisman H. Influence of interleukin-1 beta on exploratory behaviors, plasma ACTH, corticosterone, and central biogenic amines in mice. Psychopharmacology (Berl). 1998; 137: 351–361.

33. Licinio J, Wong ML. The role of inflammatory mediators in the biology of major depression: central nervous system cytokines modulate the biological substrate of depressive symptoms, regulate stress-responsive systems, and contribute to neurotoxicity and neuroprotection. Mol Psychiatry. 1999; 4: 317–327. PMID: 9728078

34. Kamikawa H, Hori T, Nakane H, Aou S, Tashiro N. IL-1beta increases norepinephrine level in rat frontal cortex: Involvement of prostanooids, NO, and glutamate. Am J Physiol. 1998; 275: R803–R810. PMID: 9728078

35. Ogawa S, Lee TM, Kay AR, Tank DW. Brain magnetic resonance imaging with contrast dependent on blood oxygenation. Proc Natl Acad Sci USA. 1990; 87: 9868–9872. PMID: 2124706

36. Biswal BB, Van Kylen J, Hyde JS. Simultaneous assessment of flow and BOLD signals in resting-state functional connectivity maps. NMR Biomed. 1997; 10: 165–170. PMID: 9430343

37. Fox MD, Raichle ME. Spontaneous fluctuations in brain activity observed with functional magnetic resonance imaging. Nat Rev Neurosci. 2007; 8: 700–711. https://doi.org/10.1038/nrn2201 PMID: 17704812

38. Yang H, Long XY, Yang Y, Yan H, Zhu CZ. Amplitude of low frequency fluctuation within visual areas revealed by resting-state functional MRI. Neuroimage 2007; 36: 144–152. https://doi.org/10.1016/j.neuroimage.2007.01.054 PMID: 17434757

39. Wu T, Zang Y, Wang L, Long X, Hallett M. Aging influence on functional connectivity of the motor network in the resting state. Neurosci Lett 2007; 422: 164–168. https://doi.org/10.1016/j.neulet.2007.05.011 PMID: 17611031

40. Mitterschiffthaler MT, Ettinger U, Mehta MA, Mataix-Cols D, Williams SC. Applications of functional magnetic resonance imaging in psychiatry. J Magn Reson Imaging 2006; 23: 851–861. https://doi.org/10.1002/jmri.20590 PMID: 16652410

41. Plessen KJ, Bansal R, Zhu H, Whiteman R, Amat J. Hippocampus and Amygdala Morphology in Attention-Deficit/Hyperactivity Disorder. Arch Gen Psychiatry 2006; 63: 795–807. https://doi.org/10.1001/archpsyc.63.7.795 PMID: 16818869

42. Perlov E, Philipsen A, Tebartz van Elst L, Ebert D, Henning J. Hippocampus and amygdala morphology in adults with attention-deficit hyperactivity disorder. J Psychiatry Neurosci 2008; 33: 509–515. PMID: 18982173

43. Yang H, Wu QZ, Guo LT, Li QQ, Long XY. Abnormal spontaneous brain activity in medication-naïve ADHD children: a resting state fMRI study. Neurosci Lett 2011; 502: 89–93. https://doi.org/10.1016/j.neulet.2011.07.028 PMID: 21810451

44. Li F, He N, Li Y, Chen L, Huang X. Intrinsic brain abnormalities in attention deficit hyperactivity disorder: a resting-state functional MR imaging study. Radiology. 2014; 272: 514–523. https://doi.org/10.1148/radiol.14131622 PMID: 24785156

45. Bhattachar J, Park SJ, Chun SW, Cho DH, Han SK. Activation of synaptic and extrasynaptic glycine receptors by taurine in preoptic hypothalamic neurons. Neurosci Lett. 2015; 608: 51–56. https://doi.org/10.1016/j.neulet.2015.10.012 PMID: 26453764

46. Arnsten AF. The emerging neurobiology of attention deficit hyperactivity disorder: The key role of the prefrontal association cortex. J Pediatr. 2009; 154: I–S43. https://doi.org/10.1016/j.jpeds.2009.01.018 PMID: 20596295

47. Thapar A, Cooper M. Attention deficit hyperactivity disorder. Lancet. 2016; 387: 1240–1250. https://doi.org/10.1016/S0140-6736(15)00238-X PMID: 26836541

48. Edden RA, Crocetti D, Zhu H, Gilbert DL, Mostofsky SH. Reduced GABA concentration in attention-deficit/hyperactivity disorder. Arch Gen Psychiatry. 2012; 69: 750–753. https://doi.org/10.1001/archgenpsychiatry.2011.2280 PMID: 22752239

49. Li XW, Gao HY, Liu J. The role of taurine in improving neural stem cells proliferation and differentiation. Nutr Neurosci. https://doi.org/10.1080/1028415X.2016.1152004
50. Kontro P. Interactions of taurine and dopamine in the striatum. Adv Exp Med Biol. 1987; 217: 347–355. PMID: 3434428

51. Jia F, Yue M, Chandra D, Keramidas A, Goldstein PA, Homanics GE. Taurine is a potent activator of extrasynaptic GABA(A) receptors in the thalamus. J Neurosci. 2008; 28: 106–115. https://doi.org/10.1523/JNEUROSCI.3996-07.2008 PMID: 18171928

52. Garcia de Yebenes Prous J, Carlsson A, Mena Gomez MA. The effect of taurine on motor behaviour, body temperature and monoamine metabolism in rat brain. Arch Pharmacol. 1978; 304: 95–99.