Identification of crucial genes associated with rat traumatic spinal cord injury

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Abstract. The aim of the present study was to investigate the key genes associated with traumatic spinal cord injuries (TSCI). The dataset GSE52763 was downloaded from the Gene Expression Omnibus, for which lumbar spinal cord samples were obtained from rats at 1 and 3 weeks following contusive spinal cord injury and 1 week subsequent to a sham laminectomy, and used to identify differentially expressed genes (DEGs). Functional enrichment analysis, co-expression analysis and transcription factor (TF) identification were performed for DEGs common to the 1 and 3 week injury samples. In total, 234 upregulated and 51 downregulated DEGs were common to the 1 and 3 week injury samples. The upregulated DEGs were significantly enriched in Gene Ontology terms concerning immunity (e.g. Itgal and Ccl2) and certain pathways, including natural killer cell mediated cytotoxicity [e.g. Ras-related C3 botulinum toxin substrate 2 (Rac2) and TYRO protein tyrosine kinase binding protein (Tyrobp)]. The downregulated DEGs were highly enriched in female gonad development [e.g. progesterone receptor (Pgr)] and the steroid biosynthesis pathway. A total of 139 genes had co-expression associations and the majority of them were upregulated genes. The upregulated co-expressed genes were predominantly enriched in biological regulation, including TGFβ induced factor homeobox 1 (Tgif1) and Rac2. The downregulated co-expressed genes were enriched in anatomical structure development (e.g. Dnm3). A total of 92 co-expressed genes composed the protein-protein interaction network. Additionally, 9 TFs (e.g. Pgr and Tgif1) were identified from the DEGs. It was hypothesized that the genes including Tgif1, Rac2, Tyrobp, and Pgr may be closely associated with TSCI.

Introduction

Patients with traumatic spinal cord injuries (TSCI) endure low health-associated quality of life and high healthcare costs. They also have a higher mortality rate compared with the general population. In 2012, the estimated incidence of acute spinal cord injury in the United States was 54 cases per 1 million (1). The biological processes of TSCI involve a diverse group of cells and molecules from the nervous, immune and vascular systems. For instance, connexin 43 functions as a mediator of central nervous system inflammation and chronic pain following spinal cord injury (2), altered expression of E2F-associated phosphoprotein regulates reactive astrogliosis and neuronal apoptosis (3) and ginsenoside Rb1 upregulates the expression of Bcl-xL and vascular endothelial growth factor at 7 days after spinal cord injury (4). Investigation of gene changes has contributed to the understanding of the molecular mechanisms of TSCI.

Gene expression profiling by microarray has been used to uncover molecular variations in spinal cord repair and degeneration (5-7). In 2014, using microarray analysis, Shin et al. (8) identified that numerous inflammation-associated genes were upregulated in the lumbar spinal cord at 1 and 3 weeks after traumatic injury, and locomotor function was improved in part by treadmill locomotor training (TMT). However, the molecular mechanisms of TSCI remain to be elucidated and regulatory factors associated with TSCI, including transcription factors (TFs), have not been investigated to the best of the authors’ knowledge.

The present study used the microarray data obtained by Shin et al (8) and screened differentially expressed genes (DEGs) common to the 1 and 3 week injury samples, and then analyzed the functions and interactions of DEGs. Additionally, TFs in DEGs were identified to reveal the regulatory associations of DEGs. These results may provide novel information to aid the understanding of the molecular mechanisms of TSCI.

Materials and methods

Affymetrix microarray data. The raw gene expression profile data GSE52763 (8) were obtained from the public database Gene Expression Omnibus (GEO; http://www.ncbi.nlm.nih.gov/geo/), which is based on the platform of GPL1355 (Rat230_2) Affymetrix Rat Genome 230 2.0 Array. The
dataset contained eight rat lumbar spinal cord samples obtained from rats 1 (n=4) and 3 (n=4) weeks following contusive spinal cord injury at the T9 level (designated as 1 week injury and 3 weeks injury samples), three lumbar spinal cord samples obtained from rats 1 week following sham laminectomy (designated as sham samples), four lumbar spinal cord samples obtained at 3 weeks following contusive spinal cord injury with treadmill training (designated as 3 weeks injury + TMT samples), and three lumbar spinal cord samples from rats which underwent a sham laminectomy followed by 2 weeks of treadmill training (designated as sham + TMT samples). All of the samples were taken from adult (8 weeks) female Sprague-Dawley rats (250-300 g). Only 1 week injury, 3 weeks injury and sham samples were used for analysis in the present study.

CEL files were downloaded and the gene expression data of all samples were preprocessed through background correction, quantile normalization and probe summarization using the Robust Microarray Analysis algorithm of the affy package of Bioconductor (http://www.bioconductor.org/packages/release/bioc/html/) (9).

**DEGs screening.** The linear models for the microarray data package of Bioconductor (10) was used to identify DEGs between 1 week and 3 weeks injury samples and sham samples. The P-value for each gene was calculated by t-test and only the genes with P-value <0.05 and fold change ≥1.5 were selected as DEGs. Subsequently, the DEGs common to the 1 and 3 week injury samples were screened for subsequent analyses.

**Enrichment analysis.** Gene Ontology (GO) functional and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses for DEGs were conducted using the Database for Annotation, Visualization and Integrated Discovery (http://david.abcc.ncifcrf.gov) database, which provides a set of functional annotation tools to aid investigators in comprehending the biological importance underlying numerous genes (11). P<0.05 and gene count ≥2 were set as the cut-off criteria.

**Co-expression analysis.** The Pearson correlation coefficient was calculated to analyze the co-expression associations between DEGs (11). The co-expression pairs with a Pearson correlation coefficient >0.9 were screened out, and the co-expression network was visualized using Cytoscape [http://cytoscape.org; (12)].

Subsequently, GO functional enrichment analysis in biological process was performed using the plug-in Bingo (13) in Cytoscape. P<0.05 was set as the cut-off criterion. Additionally, the Search Tool for the Retrieval of Interacting Genes/Proteins (http://string-db.org) database was used to analyze the protein-protein interactions (PPIs) of co-expressed genes, and the PPI network was visualized by Cytoscape.

**Identification of TFs from DEGs.** TFs in the DEGs common to the 1 and 3 week injury samples were identified using the Animal Transcription Factor Database [AnimalTFDB; http://www.bioguo.org/AnimalTFDB/species_index.php; (14)].

![Figure 1. The boxplots for microarray data prior and subsequent to normalization. The abscissa displays the samples and the ordinate represents gene expression. ‘1 week sham’ represents lumbar spinal cord samples obtained from rats 1 week following a sham laminectomy, three repeats; ‘1 week injury’ and ‘3 week injury’ represents lumbar spinal cord samples obtained from rats 1 and 3 weeks following contusive spinal cord injury at the T9 level (four repeats for each treatment).](image-url)

**Results**

**Identification of DEGs.** Based on the normalization of the microarray data, the boxplot of preprocessed data displayed that medians of each sample data were almost on a line, indicating that the data after preprocessing met the standard for further analyses (Fig. 1).

In total, 322 upregulated and 78 downregulated DEGs were screened between 1 week injury and sham samples, in addition to 354 upregulated and 285 downregulated ones between 3 week injury and sham samples. Among them, 234 upregulated and 51 downregulated DEGs were common to the 1 and 3 week injury samples. The hierarchical cluster analysis of the data demonstrated that the DEGs can be used to
GO functional and KEGG pathway enrichment analyses. To identify the functions of DEGs common to the 1 and 3 week injury samples, GO functional and KEGG pathway enrichment analyses were performed. According to GO functional enrichment analysis, the upregulated DEGs were significantly enriched in several GO terms concerning immunity, including immune response [e.g. integrin subunit α L (Itgal), similar to guanylate binding protein family, member 6 and C-C motif chemokine ligand 2 (Ccl2)], defence response (e.g. TNF α induced protein 8 like 2, apolipoprotein B mRNA editing enzyme catalytic subunit 1 and Ccl2) and cell activation (e.g. exonuclease 1, Intercellular Adhesion Molecule 1 and pleckstrin; Table I). The downregulated DEGs were highly enriched in female gonad development [e.g. Pgr, vascular endothelial growth factor A (Vegfa) and BCL2 like 1], neuron projection [e.g. ATPase plasma membrane Ca2+ transporting 2, dynamin 3 (Dnm3) and glutamate metabotropic receptor 7 (Grm7)] and gated channel activity [e.g. γ-aminobutyric acid type A receptor α3 subunit (Gabra3), Grm7 and calcium voltage-gated channel auxiliary subunit β4; Table II].

Meanwhile, a set of upregulated DEGs were markedly enriched in certain pathways, including natural killer cell mediated cytotoxicity [e.g. Ras-related C3 botulinum toxin substrate 2 (Rac2) and TYRO protein tyrosine kinase binding protein (Tyrobp)] and the B cell receptor signaling pathway [e.g. fc fragment of IgG receptor IIb (Fcgr2B) and Rac2]. Several downregulated DEGs were significantly enriched in the pathways of steroid biosynthesis (cytochrome P450 family 51, transmembrane 7 superfamily member 2 and lanosterol synthase) and neuroactive ligand-receptor interaction (e.g. Gabra3 and Grm7; Table III).

Analysis of co-expressed genes. Gene co-expression analysis is a powerful method to predict the function of genes and/or to identify genes that are functionally associated with query genes. Based on the cut-off criterion of Pearson correlation coefficient >0.9, 1894 co-expression pairs in 139 DEGs were obtained (Fig. 3). Notably, the majority of co-expressed genes were upregulated DEGs.

According to GO functional enrichment analysis, the upregulated co-expressed genes [e.g. Rac2, fc fragment of IgG receptor Ia and including TGFB induced factor homeobox 1 (Tgif1)] were significantly enriched in a series of GO terms, including biological regulation and response to stimulus. The downregulated co-expressed genes were markedly enriched in certain GO terms, including anatomical structure development (e.g. Dnm3 and Vegfa) and intracellular signal transduction (e.g. mitogen-activated protein kinase kinase kinase kinase 5 and Grm7) (Table IV).

The PPI network was composed of 92 co-expressed genes (83 upregulated and 9 downregulated) and 351 interactions. The connectivity degree of six genes was more than 20 and they were Tyrobp (degree=35), CD68 molecule (Cd68; degree=34), Rac2 (degree=29), integrin subunit β2; (degree=28), CD53 molecule (degree=25), C-type lectin domain family 4 member A (degree=22). Tyrobp interacted with multiple genes, including Cd68 and Rac2 (Fig. 4).

Analysis of TFs. Based on the AnimalTFDB database, a total of 9 TFs were identified from the DEGs common to the 1

Figure 2. The cluster heat maps for the differentially expressed genes between the sham samples and the 1 and 3 week injury samples. Each row represents a single gene and each column represents a spinal cord sample. The gradual color change from red to green represents the changing process from upregulation to downregulation. ‘1 week sham’ represents lumbar spinal cord samples obtained from rats 1 week following a sham laminectomy, three repeats; ‘1 week injury’ and ‘3 week injury’ represents lumbar spinal cord samples obtained from rats 1 and 3 weeks following contusive spinal cord injury at the T9 level (four repeats for each treatment).

accurately distinguish 1 and 3 week injury samples from sham samples (Fig. 2).
Table I. Top five enriched GO terms with the highest P-value in biological process, cellular component and molecular function for the up-regulated differentially expressed genes common to the 1 week and 3 weeks injury samples.

| Category | Term | Count | P-value | Genes |
|----------|------|-------|---------|-------|
| BP       | GO:0006955-immune response | 37    | 3.71E-22 | Tnfaip8L2, Itgal, Loc685067, Ccl2, Apobec1, C3, Endou, Tlr2, Rsad2, Tlr7, C1Qc, Btk, Gch1, Klh16, Fcgr1A, Cfh, Fcrl1G, Inpp5D, Fcgr3A, Exol1, Ptpn6, Icam1, Ptprc, Tnfsf4, Rl1-1C12, Myo1F, Vav1, Psmb8, Psmb9, C1Qa, C1Qb, Cd86, Cybb, Fcgr2B, Cd300A, Cxcl16, Gbp2 |
|          | GO:0006952-defense response  | 29    | 8.07E-15 | Tnfaip8L2, Apobec1, Ccl2, C3, Tlr2, Rsad2, Itgb2, Tlr7, C1Qc, Btk, Gch1, Casp4, Hmox1, Cybb, Fcgr1A, Cfh, Pycard, Fcrl1G, Ptpn6, Ptprc, Tnfsf4, Pdpn, Hck, Myo1F, C1Qa, C1Qb, Cd86, Fcgr2B, Cxcl16 |
|          | GO:0002252-immune effector process | 18    | 3.17E-14 | Exol1, Ptprc, Ptpn6, Icam1, Apobec1, C3, Myo1F, Rsad2, Tlr7, C1Qc, Btk, C1Qa, C1Qb, Fcgr2B, Fcrl1A, Cfh, Fcrl1G, Inpp5D |
|          | GO:0001775-cell activation | 21    | 4.15E-12 | Exol1, Icam1, Ptprc, Itgal, Tnfsf4, Plek, Aif1, Tlr2, Myo1F, Itgb2, Tlr7, Vav1, Igam, Timp1, Btk, Cd48, Cd86, Fcgr2B, Fcrl1G, Fcgr3A, Blnk |
|          | GO:0004532-leukocyte activation | 19    | 3.69E-11 | Exol1, Icam1, Ptprc, Itgal, Tnfsf4, Aif1, Tlr2, Myo1F, Itgb2, Tlr7, Vav1, Igam, Btk, Cd48, Cd86, Fcgr2B, Fcrl1G, Fcgr3A, Blnk |
| CC       | GO:0009897-external side of plasma membrane | 13    | 5.95E-07 | Cd48, Icam1, Itgal, Ptprc, Cd86, Emr1, Fcgr2B, Fcgr1A, Tlr2, Fcrl1G, Cd22, Clec7A, Itgam |
|          | GO:0009986-cell surface | 16    | 1.03E-05 | Icam1, Ptprc, Itgal, Tnfsf4, Tlr2, Itgb2, Cd53, Igam, Cd48, Cd86, Fcgr2B, Emr1, Fcgr1A, Cd22, Fcrl1G, Clec7A |
|          | GO:0044459-plasma membrane part | 33    | 7.58E-05 | Itgal, Gna15, Aif1, Tlr2, Rsad2, Itgb2, Abca1, Itgam, Cd48, P2Ry6, Laptm5, Fcgr1A, Hmox1, Cd22, Fcrl1G, Gpmb, Ptpn6, Icam1, Ptprc, Plek, Rl1-1C12, Pdpn, Anxa1, Axl, Stom, Cd86, Cybb, Gng2, Emr1, Fcgr2B, Clec7A, Cpf, Gbp2 |
|          | GO:0005886-plasma membrane | 47    | 2.18E-04 | Itgal, Gna15, Aif1, Ifitm3, Tlr2, Rsad2, Itgb2, Abca1, Itgam, Scl7A7, Cd48, P2Ry6, Laptm5, Mall, Fcgr1A, Hmox1, Cfh, Fcrl1G, Cd22, Pik3Ap1, Inpp5D, Fcgr3A, Gpmb, Csf1r1, Blnk, Ptprc, Ptpn6, Icam1, F10, Plek, Rl1-1C12, Pdpn, Anxa1, Axl, Clic1, Stom, Cd86, Cybb, Gng2, Gpr34, Fcgr2B, Emr1, Cd300A, Clec7A, Pcyox1, Cpf, Gbp2 |
|          | GO:0005576-extracellular region | 30    | 3.59E-04 | Fmod, Ccl2, Spock3, C3, Tnc, C1Qc, Timp1, Lgals3Bp, Glipr1, Fcrl1, Cfh, Pycard, Ptn, Casp1, Tfpi2, Matn2, Icam1, Ctsz, F10, Tnfsf4, Lgals3, Plek, Anxa1, C1Qa, C1Qb, Grn, Cxcl16, Cpf, Pcyox1, Pros1 |
| MF       | GO:0030246-carbohydrate binding | 19    | 8.31E-08 | Ptprc, Ccl2, Lgals3, Spock3, Endou, Hexb, Tlr2, Itgam, Tnfaip6, Pygl, Fcn1, Cfh, Grf6in, Clec4A1, Ptn, Clec4A3, Clec7A, Gpmb, Cd302 |
|          | GO:0030247-polysaccharide binding | 11    | 1.06E-06 | Tnfaip6, Ptprc, Ccl2, Spock3, Endou, Cfh, Tlr2, Ptn, Clec7A, Gpmb, Itgam |
**Table I. Continued.**

| Category | Term | Count | P-value |
|----------|------|-------|---------|
| GO:0001871 | pattern binding | 11 | 1.06E-06 |
| GO:0019865 | immunoglobulin binding | 5 | 8.51E-06 |
| | | | |
| GO:0005539 | glycosaminoglycan binding | 9 | 2.63E-05 |
| | | | |
| TNF | α | Integrin subunit L; binding protein family, member 6; Ccl2, C-C motif chemokine ligand 2; proteasome subunit 8; C1Qa, complement C1q A chain; C1Qb, complement C1q B chain; Cybb, cytochrome B-245 b chain | 2001 |
| | | | |
| Fcgr2B | apolipoprotein B MRNA editing enzyme catalytic subunit 1; | 21 |
| | | | |
| Fcgr1A | fc fragment of IgG receptor Ia; | 207 |
| | | | |
| Fcgr3A | Fc fragment of IgE receptor Ig; | 217 |
| | | | |
| Lgals3 | galectin 3; | 239 |
| | | | |
| Tnfaip6 | TNF α induced protein 6; | 215 |
| | | | |
| Ptprc | galectin‑related inter‑fiber protein; | 261 |
| | | | |
| Ccl2 | C-C motif chemokine ligand 2; | 118 |
| | | | |
| Spock3 | EGF-Like Module Receptor 1; | 250 |
| | | | |
| Cfh | complement factor H; | 221 |
| | | | |
| Tlr2 | toll like receptor 2; | 210 |
| | | | |
| Ptn | exonuclease 1; | 228 |
| | | | |
| Gpnmb | protein tyrosine phosphatase, receptor type C; | 239 |
| | | | |
| Itgam | integrin subunit α T; | 207 |
| | | | |
| Timp1 | | 299 |
| | | | |
| Icam1 | Intercellular Adhesion Molecule 1; | 118 |
| | | | |
| Vav1 | vav guanine nucleotide exchange factor 1; | 207 |
| | | | |
| Itgb2 | integrin subunit b2; | 185 |
| | | | |
| Rsd2 | radical s-adenosyl methionine domain containing 2; | 7B, SRY-box 18, activating TF 3 (Af3), MAF BZIP TF B (Mafb), Tgif1, Fli-1 proto-oncogene, ETS TF (Fli1) and T-box 4 (Tbx4). Among them, Af3, Mafb, Tbx4, Tgif1 and Fli1 were all upregulated in 1 and 3 week injury samples, compared with sham samples, while the others were downregulated (Fig. 5).

**Discussion**

In the present study, 234 upregulated and 51 downregulated DEGs were common to the 1 week and 3 week injury samples, compared with the sham samples. Among them, 139 genes had co-expression associations and the majority of them were upregulated genes. The upregulated co-expressed genes were predominantly enriched in several GO terms of biological regulation, including Tgif1 and Rac2.

Tgif1 was identified as a TF in the present study. It belongs to the three-amino acid loop extension superclass of atypical homeodomains (15). Studies (16-18) have showed that Tgif1 exerts crucial functions in the nervous system. Additionally, a previous study (19) identified TGF1 as a novel regulator of macrophage activation in immune response. In the present study, Tgif1 had a co-expression associations with Rac2 and Tyrobp, the two of which had a higher degree in the PPI network. Rac2 encodes a member of the Ras superfamily of small guanosine triphosphate (GTP)-metabolizing proteins, and it modulates diverse processes, including secretion, cell polarization and phagocytosis (20). In the present study, Rac2 was identified to be significantly enriched in several pathways, including natural killer cell-mediated cytotoxicity. Natural killer cells participate in immune processes after spinal cord injury (21) and there is evidence that suppression of Rac activity in the injured spinal cord enhances cell survival (22). It has been demonstrated that the expression of Rac2 is activated in inflammatory responses (23). Furthermore, Ras GTPases exert critical functions in multiple procedures during axonogenesis in injured spinal cords (24). Tyrobp is a transmembrane signaling polypeptide which has an immunoreceptor tyrosine-based activation motif and it serves a role in signal transduction, brain myelination, and inflammation (25,26).

In the current study, Tyrobp was enriched in natural killer cell mediated cytotoxicity, interacted with Rac2 in the PPI network and co-expressed with Tgif1. Together, Tgif1, Rac2 and Tyrobp may play pivotal roles in TSCI.

Among the downregulated genes, Pgr, identified as a TF, was highly enriched in female gonadal development and the ovulation cycle process. In axonal regeneration, gonadal steroids function as promoting factors (27). Estrogens have direct neuroprotective effects, including modification of humoral immune responses, and gestagens can prevent neuronal death and promote the growth of nervous cells and the formation of new synapses (28). A previous study (29) confirmed that progesterone provides neuroprotection to the injured central and peripheral nervous system in the injured spinal cord. Therefore, Pgr may serve a key role in the regulation of nervous regeneration in spinal cord injuries.

In conclusion, 234 upregulated and 51 downregulated DEGs were differentially expressed in 1 and 3 week injury samples. Among them, the upregulated genes Rac2 and...
Table II. Top five enriched GO terms with the highest P-value in biological process, cellular component and molecular function for the downregulated differentially expressed genes common to the 1 week injury and 3 week injury samples.

| Category | Term                                      | Count | P-value      | Genes                        |
|----------|-------------------------------------------|-------|--------------|------------------------------|
| BP       | GO:0008585-female gonad development       | 5     | 7.32E-05     | Pgr, Vegfa, Bcl2L1, Pcyt1B, Vgf |
|          | GO:0046545-development of primary female sexual characteristics | 5     | 9.52E-05     | Pgr, Vegfa, Bcl2L1, Pcyt1B, Vgf |
|          | GO:0022602-ovulation cycle process        | 5     | 1.05E-04     | Pgr, Vegfa, Bcl2L1, Pcyt1B, Vgf |
|          | GO:0046660-female sex differentiation     | 5     | 1.28E-04     | Pgr, Vegfa, Bcl2L1, Pcyt1B, Vgf |
|          | GO:0042698-ovulation cycle                | 5     | 1.46E-04     | Pgr, Vegfa, Bcl2L1, Pcyt1B, Vgf |
| CC       | GO:0043005-neuron projection              | 8     | 6.07E-04     | Pgr, Atp2B2, Dnm3, Grm7, Dicer1, Slc18A3, Vegfa, Pex5L |
|          | GO:0045202-synapse                       | 7     | 1.68E-03     | Dnm3, Clstn2, Gabra4, Gabra3, Grm7, Rps6Kb1, Rph3A |
|          | GO:0044456-synapse part                   | 6     | 1.80E-03     | Dnm3, Clstn2, Gabra4, Gabra3, Grm7, Rph3A |
|          | GO:0030425-dendrite                      | 5     | 8.17E-03     | Atp2B2, Dnm3, Grm7, Dicer1, Pex5L |
|          | GO:0005886-plasma membrane                | 16    | 9.36E-03     | Cadm3, Clstn2, Gabra4, Gabra3, Trhr, Rps6Kb1, Bcl2L1, Cacnb4, Rph3A, Slc9A3R2, P2Rx5, Atp2B2, Mastl, Grm7, Rasgpl1, Slc6A5 |
| MF       | GO:0022836-gated channel activity         | 5     | 1.32E-02     | P2Rx5, Gabra4, Gabra3, Grm7, Cacnb4 |
|          | GO:0005230-extracellular ligand-gated ion channel activity | 3     | 2.14E-02     | P2Rx5, Gabra4, Gabra3 |
|          | GO:0005216-ion channel activity           | 5     | 2.74E-02     | P2Rx5, Gabra4, Gabra3, Grm7, Cacnb4 |
|          | GO:0022838-substrate specific channel activity | 5     | 3.00E-02     | P2Rx5, Gabra4, Gabra3, Grm7, Cacnb4 |
|          | GO:0046983-protein dimerization activity  | 6     | 3.33E-02     | Zbtb7B, Cadm3, Grm7, Trhr, Vegfa, Bcl2L1 |

GO, Gene Ontology; BP, biological process; CC, cellular component; MF, molecular function; Pgr, progesterone receptor; Vegfa, vascular endothelial growth factor A; Bcl2L1, BCL2 like 1; Pcyt1B, phosphate cytidylyltransferase 1, choline, β; Vegf, VGF nerve growth factor inducible; Atp2B2, ATPase plasma membrane Ca²⁺ transporting 2; Dnm3, dynamin 3; Grm7, glutamate metabotropic receptor 7; Dicer1, Dicer 1; Ribonulease III; Slc18A3, solute carrier family 18 member A3; Pex5L, peroxisomal biogenesis factor 5 like; Clstn2, calsyntenin 2; Gabra4, γ-aminobutyric acid type A receptor α4 subunit; Gabra3, γ-aminobutyric acid type A receptor α3 subunit; Rps6Kb1, ribosomal protein S6 kinase B1; Rph3A, rabphilin 3A; Cadm3, cell adhesion molecule 3; Trhr, thyrotropin releasing hormone receptor; Cacnb4, calcium voltage-gated channel auxiliary subunit β 4; Slc9A3R2, SLC9A3 regulator 2; P2Rx5, purinergic receptor P2X 5; Zbtb7b, zinc finger and BTB domain containing 7B.
Table III. Top five enriched pathways for the upregulated differentially expressed genes and two enriched pathways for the downregulated differentially expressed genes common to the 1 week injury and 3 week injury samples.

| Category | Term | Count | P-value | Genes |
|----------|------|-------|---------|-------|
| Upregulated | rno04650: Natural killer cell mediated cytotoxicity | 11 | 9.78E-07 | Cd48, Itgal, Ptpn6, Icam1, Fcgr2B, Rac2, Fcer1G, Itgb2, Fcgr3A, Vav1, Tyrobp |
| | rno04662: B cell receptor signaling pathway | 9 | 7.93E-06 | Ptpn6, Fcgr2B, Rac2, Cd22, Pik3Ap1, Inpp5D, Vav1, Btk, Blnk |
| | rno04666: Fcγ R-mediated phagocytosis | 8 | 2.01E-04 | Arpc1B, Fcgr1A, Rac2, Hck, Fcgr1A, Inpp5D, Vav1 |
| | rno05322: Systemic lupus erythematosus | 8 | 2.32E-04 | C1Qa, C1Qb, Cd86, Fcgr2B, C3, Fcgr1A, Fcgr3A, C1Qc |
| | rno04610: Complement and coagulation cascades | 7 | 3.92E-04 | C1Qa, C1Qb, F10, C3, Cfh, Pros1, C1Qc |
| Downregulated | rno00100: Steroid biosynthesis | 3 | 1.94E-03 | Cyp51, Tm7sf2, Lss |
| | rno04080: Neuroactive ligand-receptor interaction | 5 | 1.77E-02 | P2rx5, Gabra4, Gabra3, Grm7, Trhr |

Cd48, CD48 molecule; Itgal, Integrin subunit αL; Icam1, Intercellular Adhesion Molecule 1; Fcgr2B, Fc fragment of IgG receptor IIb; Rac2, Ras-related C3 botulinum toxin substrate 2; Fcer1G, Fc fragment of IgE receptor Iγ; Itgb2, integrin subunit β2; Fcgr3A, Fc fragment of IgG receptor IIIa; Vav1, vav guanine nucleotide exchange factor 1; Tyrobp, TYRO protein tyrosine kinase binding protein; Ptpn6, protein tyrosine phosphatase, non-receptor type 6; Cd22, CD22 molecule; Pik3Ap1, phosphoinositide-3-kinase adaptor protein 1; Inpp5D, inositol polyphosphate-5-phosphatase D; Btk, braton tyrosine kinase; Blnk, B-cell linker; Arpc1B, actin related protein 2/3 complex subunit 1B; Ptprc, protein tyrosine phosphatase, receptor type C; Hck, HCK proto-oncogene, src family tyrosine kinase; Fcgr1A, Fc fragment of IgG Receptor Iα; Inpp5D, inositol polyphosphate-5-phosphatase D; C1Qa, complement C1q A chain; C1Qb, complement C1q B chain; Cd86, CD86 molecule; C3, complement C3; C1Qc, complement C1q C chain; F10, coagulation factor X; Cfh, complement factor H; Pros1, Protein S (α); Cyp51, cytochrome P450 family 51; Tm7sf2, transmembrane 7 superfamily member 2; Lss, lanosterol synthase; P2rx5, purinergic receptor P2X 5; Gabra4, gamma-aminobutyric acid type A receptor α4 subunit; Gabra3, gamma-aminobutyric acid type A receptor α3 subunit; Grm7, glutamate metabotropic receptor 7; Trhr, thyrotropin releasing hormone receptor.
Table IV. Top five enriched GO terms with the highest P-value in biological process for the co-expressed up- and downregulated genes.

| Category | Term | Count | P-value | Genes |
|----------|------|-------|---------|-------|
| Co-expressed upregulated genes | GO:0065007-biological regulation | 44 | 2.39E-02 | S100A4, Tspo, Speck3, C3, Aif1, Rbm3, Unc93B1, Rsad2, Itgb2, Cd53, C1Qc, Tiprl, Timp1, Cd48, P2Ry6, Rac2, Fcgr1A, Hmox1, Pycard, Cfh, Ptn, Fcerg1G, Inpp5D, Csf1R, Ptprc, Ptd, Pldp, Bcl2A1, Anx1, Ptk, I6R, Anxa3, Psm8b, Arhgap25, Psmb9, C1Qa, C1Qb, Arp1B, Grn, Cxcl16, Tgif1, Cn, Smarca1 |
| GO:0050789-regulation of biological process | | 43 | 1.12E-02 | S100A4, Tspo, Speck3, C3, Aif1, Rbm3, Unc93B1, Rsad2, Itgb2, Cd53, C1Qc, Tiprl, Timp1, Cd48, P2Ry6, Rac2, Fcgr1A, Hmox1, Pycard, Cfh, Ptn, Fcerg1G, Inpp5D, Csf1R, Ptprc, Ptd, Pldp, Bcl2A1, Anx1, Ptk, I6R, Anxa3, Psm8b, Arhgap25, Psmb9, C1Qa, C1Qb, Arp1B, Grn, Cxcl16, Tgif1, Smarca1 |
| GO:0050896-response to stimulus | | 41 | 8.03E-04 | Itgal, Tspo, Rtp4, Ifitm1, C3, Aif1, Rbm3, Ifitm3, Endou, Rsad2, Itgb2, Cd53, C1Qc, P2Ry6, Rac2, Alox5Ap, Fcgr1A, Hmox1, Pycard, Cfh, Ptn, Fcerg1G, Inpp5D, Csf1R, Ptprc, Ptd, Pldp, Bcl2A1, Anx1, Ptk, I6R, Anxa3, Psm8b, Arhgap25, Psmb9, C1Qa, C1Qb, Arp1B, Grn, Cxcl16, Tgif1, Cn, Smarca1 |
| GO:0048856-anatomical structure development | | 8 | 4.26E-02 | Pcsk2, Lingol1, Atp2B2, Dnm3, Vegfa, Rp6Kb1, Fgf1, Scl9a3R2 |
| GO:007399-nervous system development | | 6 | 4.26E-02 | Pcsk2, Lingol1, Atp2B2, Dnm3, Vegfa, Fgf1 |
| GO:0035556-intracellular signal transduction | | 5 | 4.26E-02 | Lingol1, Map4K5, Gm7, Rp6Kb1, Fgf1 |
| GO:002376-immune system process | | 28 | 6.07E-15 | Itgal, Aif1, C3, Endou, Unc93B1, Rsad2, Itgb2, C1Qc, Itgam, Cd48, Fcgr1A, Cfh, Fcerg1G, Inpp5D, Grf3A, Blnk, Ptprc, Ptd, Pldp, Anxa1, I6R, Anxa3, Psm8b, Psmb9, C1Qa, C1Qb, Cxcl16, Grn, Tgif1, Smarca1 |
| GO:0002376-immune system process | | 28 | 6.07E-15 | Itgal, Aif1, C3, Endou, Unc93B1, Rsad2, Itgb2, C1Qc, Itgam, Cd48, Fcgr1A, Cfh, Fcerg1G, Inpp5D, Grf3A, Blnk, Ptprc, Ptd, Pldp, Anxa1, I6R, Anxa3, Psm8b, Psmb9, C1Qa, C1Qb, Cxcl16, Grn, Tgif1, Smarca1 |
| GO:0002376-immune system process | | 28 | 6.07E-15 | Itgal, Aif1, C3, Endou, Unc93B1, Rsad2, Itgb2, C1Qc, Itgam, Cd48, Fcgr1A, Cfh, Fcerg1G, Inpp5D, Grf3A, Blnk, Ptprc, Ptd, Pldp, Anxa1, I6R, Anxa3, Psm8b, Psmb9, C1Qa, C1Qb, Cxcl16, Grn, Tgif1, Smarca1 |
| GO:0035556-intracellular signal transduction | | 5 | 4.26E-02 | Lingol1, Map4K5, Gm7, Rp6Kb1, Fgf1 |
| GO:0007243-intracellular protein kinase cascade | | 4 | 3.54E-02 | Lingol1, Map4K5, Rp6Kb1, Fgf1 |
| GO:0007243-intracellular protein kinase cascade | | 4 | 3.54E-02 | Lingol1, Map4K5, Rp6Kb1, Fgf1 |

**GO, Gene Ontology; BP, biological process; Itgb2, integrin subunit β2; Cd53, CD53 Molecule; Rac2, Rac-related C3 botulinum toxin substrate 2; Fcgr1A, fragment of IgG receptor Ia; Hmox1, heme oxygenase 1; C1Qb, complement C1q B chain; Arp1B, actin related protein 2/3 complex subunit IB; Grn, granulin; Cxcl16, C-X-C motif chemokine ligand 16; Tgif1, TGFβ induced factor homeobox 1; P2Ry6, pyrimidinergic receptor P2Y6; Arhgap25, r GTPase activating protein 25; Psmb9, proteasome subunit 9; C1Qa, complement C1q A chain; Smarca1, WI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily A, member 1; Itgam, integrin subunit α M; Tiprl, TOR signaling pathway regulator; Timp1, TIMP metalloproteinase inhibitor 1; Cd48, CD48 molecule; Pycard, PYD and CARD domain containing; Cfh, complement factor H; Cyb, cytochrome B-245 β chain; Ptd, pleckstrin; Pldp, podoplanin; Anxa1, annexin A1; I6R, interleukin 6 receptor; Anxa3, annexin A3; Tyrobp, TYRO protein tyrosine kinase binding protein; Igal, Integrin subunit α I; Blnk, B-cell linker; Ptprc, protein tyrosine phosphatase, receptor type C; Bcl2L1, BCL2 like 1; Pcsk2, proprotein convertase subtilisin/kexin type 2; Lingol1, leucine rich repeat and Ig domain containing 1; Atp2b2, ATPase plasma membrane Ca2+ transporting 2; Dnm3, dynamin 3; Vegfa, vascular endothelial growth factor A; Rp6Kb1, ribosomal protein S6 kinase B1; Fgf1, fibroblast growth factor 1; Scl9a3R2, SLC9A3 regulator 2; Map4K5, mitogen-activated protein kinase kinase kinase kinase 5; Gm7, glutamate metabotropic receptor 7.
Tyrobp, which are associated with natural killer cell-mediated cytotoxicity, may have crucial functions in TSCI. Tgif1 and Pgr may exert a regulatory function in TSCI. These observations require experimental validation, however they are expected to aid the elucidation of the molecular mechanisms in TSCI.

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