Airways are More Reactive to Histamine than to Methacholine in Patients with Mild Airway Hyperresponsiveness, Regardless of Atopy

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Background : The airway muscles from allergen-sensitized animals in vitro show a heightened response to histamine, but not to carbachol. This study investigated whether the airway responsiveness to histamine in vivo is comparable to that of methacholine in human subjects with varying degrees of atopy.

Methods : One-hundred-and-sixty-eight consecutive adult asthma patients or volunteers underwent bronch-oprovocation tests to both histamine and methacholine after determining their blood eosinophil counts, serum total IgE levels and skin test reactivity to 10 common aeroallergens.

Results : The responsiveness to histamine was significantly related to that to methacholine (r=0.609, p<0.001), but many individuals with a negative methacholine test response showed a positive response to histamine. The histamine–bronchial reactivity index (BRindex) was significantly higher than the methacholine–BRindex in subjects with a positive response to none (n=69, p<0.01) or only one (n=42, p<0.001) of histamine and methacholine, while there was no significant difference in the subjects with positive responses to both of them (n=57). The histamine–BRindex was significantly higher than the methacholine–BRindex in the subjects with mild histamine hyperresponsiveness (n=58, 1.28 ±0.01 vs. 1.20±0.02, respectively, p<0.001). Both histamine and methacholine responsiveness was significantly related to the atopy markers. However, the histamine–BRindex/methacholine–BRindex ratio of the atopics was not significantly different from that of the non–atopics.

Conclusions : The airway responsiveness to histamine is comparable to that of methacholine in the subjects with positive responses to both histamine and methacholine, but the airway responsiveness to histamine is greater than that to methacholine in those subjects with mild airway hyperresponsiveness, regardless of atopy.

Key Words : Asthma, Atopy, Histamine, Methacholine

INTRODUCTION

Atopic allergy, including asthma, is characterized not only by the enhanced production of allergen–specific IgE antibodies, but also by increased sensitivity of the target organs). Some people can be sensitized to allergens, but not be allergic, i.e., they lack increased sensitivity of the target organs; however, many individuals with respiratory allergy have an increased airway response specifically to allergens that they are sensitized to or they are nonspecifically sensitized to various stimuli such as histamine and methacholine. Airway responsiveness to histamine or methacholine is widely measured in general practice for
diagnosing asthma\textsuperscript{3}.

Although many patients with asthma have airway hyper-
responsiveness (AHR) to various pharmacological or physical
bronchoconstricting stimuli, it has been recognized that they
often respond to one stimulus, but not to all other stimuli\textsuperscript{3}. Antonnissen et al.\textsuperscript{5, 6} demonstrated a heightened response to
histamine, but not to a cholinergic agonist, in the airway
muscles from allergen–sensitized animals. Histamine is the
primary mediator of the allergic response, and leukotrienes,
another allergic mediator, increase the number of histamine
receptors\textsuperscript{7}. Therefore, atopic individuals might have an
increased specific sensitivity to an allergic mediator. However,
Juniper, et al.\textsuperscript{8} have showed that the histamine–AHR is
comparable to the methacholine–AHR in asthma patients, so
both tests are used interchangeably in general practice to
diagnose asthma. This study investigated whether the airway
responsiveness to histamine is comparable to that of
methacholine in subjects with varying degrees of atopy.

**MATERIALS AND METHODS**

**Subjects**

One-hundred-and-sixty-eight consecutive adult asthma
patients and healthy volunteers whose ages were less than 50
years were recruited from Chonnam National University
Hospital. The enrolled patients were mostly young adult men
who required a medical certificate for asthma in order to be
exempted from obligatory military service, and medical students
and hospital employees, including doctors and nurses, were
also enrolled. The participants were classified as 3 groups,
based on their responses to methacholine and histamine
bronchoprovocation tests. Sixty-nine subjects had negative
responses to both tests (the both negative group); 42 subjects
had a positive response to one test only (the sole positive
group), and the other 57 subjects had positive responses to
both tests (the both positive group). The Institutional Review
Board of Chonnam National University Hospital approved the
study. All the patients were informed of the experimental
procedures and they provided written informed consent for
participation.

**Study design**

On the first day of the study, baseline spirometry, allergy
skin–prick tests and laboratory tests for the complete blood
counts and measuring the total serum IgE were performed after
taking a history and conducting a physical examination. The
airway responsiveness to methacholine or histamine (selected
randomly) was then measured. The test was performed again
the next day, at the same time of day, with the other agent.

**Studies for atopy markers\textsuperscript{6, 9}\textsuperscript{10}**

The total IgE levels were determined using nephelometry
(normal < 100 IU/mL: Behring Diagnostics GmbH, Germany).
Allergy skin–prick tests were carried out using ten common
derivative extracts (Allergopharma, Reinbek, Germany): *Derma-
tophagoides farinae, D. pteronyssinus, cockroach (Periplaneta
americana), cat, dog, Aspergillus, hazel, birch, timothy, and
ragweed: histamine (1 mg/mL) and saline (0.9%) solutions were
used as the positive and negative controls, respectively. The
longest and perpendicular diameters of each wheal were
measured using vernier calipers (Absolute Digimatic: Mitutoyo,
Japan) 15 minutes after a prick with a 26-gauge needle, and
the arithmetic mean of the recorded measurements was used
as the representative value. Reactivity was graded according to
the ratio of the size of the allergen–induced wheal to the size of
the wheal elicited by the histamine solution (the A/H ratio),
and this was categorized as follows: 1+: 25–49%, 2+: 50–99%,
3+: 100–199%, 4+: ≥200%. The sums of the grades for the
ten allergens and for the two house dust mite extracts were
defined as the atopy score and the house dust mite score,
respectively. The skin test reactivity of ≥3+ was regarded as
indicative of a clinically significant positive response\textsuperscript{10},
and the subjects having at least one significant positive response were
deemed as atopic.

**Airway responsiveness**

The bronchial challenge tests were followed a standardized
tidal breathing method\textsuperscript{11}. The forced expiratory volume in one
second (FEV\textsubscript{1}) was measured in triplicate before the test and in
duplicate (at 30 and 90 sec) after each inhalation period with
using a spirometer (Spiro Analyzer ST-250, Fukuda Sangyo,
Tokyo, Japan). The selected predictive equation for FEV\textsubscript{1} was
that recommended by the Intermountain Thoracic Society\textsuperscript{12}:
Isotonic saline, followed by a methacholine (Sigma–Aldrich, St.
Louis, MO) solution or a histamine (Sigma–Aldrich) solution, was
aerosolized at room temperature in a DeVilbiss 646 nebulizer
(DeVilbiss, Somerset, PA: output 0.13 mL/min). In view of the
chemical stability of methacholine and histamine solutions, they
were prepared fresh on the morning of each challenge day.
The dilution increments used in the first year of this study were
0.075, 0.15, 0.31, 0.62, 1.25, 2.5, 5.0, 10 and 25 mg/mL. Since
many of the subjects could not tolerate the highest histamine
dose, this highest dose was then reduced from 25 to 16 mg/mL
in the second year. The aerosols were inhaled by tidal breathing
over a 2 minute period at 5-min intervals, through the mouth
with the nose clipped. The challenge test was discontinued if
the FEV\textsubscript{1} dropped by 20% or more from the post–saline FEV\textsubscript{1},
or if the maximum concentration of agonist was administered.
The provocation concentration of methacholine or histamine
resulting in a 20% fall in the FEV\textsubscript{1} (PC\textsubsymbol{20}) was calculated by
Table 1. Clinical characteristics of the subjects grouped according to their responsiveness to histamine and methacholine

|                        | Both Negative\(^a\) (n=69) | Sole Positive\(^a\) (n=42) | Both Positive\(^a\) (n=57) |
|------------------------|----------------------------|-----------------------------|-----------------------------|
| Age (years)            | 26.9±0.6                   | 26.2±1.0                    | 24.8±0.8                    |
| Gender (M/F)           | 48/21                      | 28/14                       | 36/21                       |
| Height (cm)            | 169.9±0.9                  | 169.8±1.4                   | 169.3±1.1                   |
| Blood eosinophils > 400/μL\(^b\) | 6 (8.7%)                  | 3 (7.1%)                    | 9 (15.8%)                   |
| Serum IgE > 100 IU/mL\(^c\)* | 32 (46.4%)                | 17 (40.5%)                  | 42 (73.7%)                  |
| Atopy\(^d\)**          | 32 (47.1%)                 | 23 (54.8%)                  | 44 (78.6%)                  |
| House dust mites SPT\(^†\) (+)** | 30 (44.1%)               | 20 (47.6%)                  | 43 (76.8%)                  |
| FEV\(_1\) (% predicted) | 92.2±2.0                   | 88.5±1.9                    | 87.5±1.5                    |
| Histamine PC\(_{20}\)\(^***\) (Geometric mean, mg/mL) | >16 | 5.36 | 1.54 |
|                        | > 16 mg/mL                 | 69 (100%)                   | 0                           |
|                        | 2~16 mg/mL                 | 0                           | 32 (76.2%)                  |
|                        | 0.2~2 mg/mL                | 6 (14.3%)                   | 29 (50.9%)                  |
|                        | < 0.2 mg/mL                | 1 (2.4%)                    | 2 (3.5%)                    |

\(^a\)negative or positive airway hyperresponsiveness [PC\(_{20}\) [provocation concentration resulting in a 20% fall in forced expiratory volume in one second (FEV\(_1\))] <16 mg/mL] to histamine and methacholine; \(^b\) Skin prick test; \(^c\) p <0.01 and \(^d\) p <0.001.

RESULTS

The clinical characteristics of the subjects are presented in Table 1. The age, gender, height and baseline FEV\(_1\) values did not differ significantly among the groups. However, the histamine PC\(_{20}\) and atopy markers, except the blood eosinophil counts, i.e., the serum total IgE levels, the atopy score and house dust mite score, were significantly different among the groups. The sole positive group had a mild AHR to histamine (PC\(_{20}\): 2~16 mg/mL) in 83.3% of the subjects as compared to the both positive group that had a mild AHR to histamine in 45.6% of the subjects (p<0.001).

The BRindex and maximal % fall in FEV\(_1\) in response to histamine were significantly related to the responses to methacholine (BRindex: r=0.609, maximal FEV\(_1\) fall: r=0.439; all p<0.001) (Figure 1, Table 2). However, many values were distributed in the upper areas of the lines of identity: in other words, the histamine–BRindex and FEV\(_1\) fall were higher in 113 (67.3%) and 109 (64.9%) subjects, respectively, and lower only in 53 (31.5%) and 50 (29.8%) subjects, respectively, than the methacholine–BRindex and FEV\(_1\) fall, respectively. The BRindex and FEV\(_1\) fall of the response to histamine were significantly higher than those to methacholine (1.24±0.01 vs. 1.18±0.01 and 20.6±0.9 vs. 15.5±0.8%, respectively, all p<0.001).

Thirty-eight individuals with a negative methacholine test response showed a positive response to histamine, whereas only four persons with a negative histamine test response showed a positive response to methacholine. The BRindex of the response to histamine was significantly higher than that to methacholine in the both negative group (1.12±0.01 vs. 1.09±0.01, p<0.01) and the sole positive group (1.29±0.02 vs. 1.14±0.01, p<0.001), while there was no significant difference in the both positive group (1.37±0.02 vs. 1.36±0.02, p>0.05) (Figure 2). Similarly, the maximal % fall in the FEV\(_1\) for the response to histamine was significantly higher than that to methacholine in the both negative group (1.24±0.01 vs. 1.18±0.01, p<0.01) and the sole positive group (1.29±0.02 vs. 1.14±0.01, p<0.001), while there was no significant difference in the both positive group (1.37±0.02 vs. 1.36±0.02, p>0.05) (Figure 2).
Both histamine and methacholine responsiveness were significantly related to the blood eosinophil counts, the serum total IgE levels, the atopy score and the skin responsiveness to the house dust mite extracts (Table 2). However, the histamine/methacholine-BRindex ratio was not significantly related to the atopy markers. Since the airway responsiveness to histamine was significantly higher than to methacholine in the both negative group and the sole positive group, but not in the both positive group, the measurements of the atopic markers were compared between the histamine-sensitive and methacholine-sensitive groups as classified according to the histamine-BRindex/methacholine-BRindex ratio in the both negative group and the sole positive group (Table 3). There was no significant difference between the groups. In addition, the histamine-BRindex/methacholine-BRindex ratio in the atopics (n=55) was not significantly different from that in the non-atopics (n=45) whose atopy scores were zero (1.07±0.02 in the atopics vs. 1.08±0.02 in the non-atopics, p=0.622). The subjects with mild histamine AHR also showed a similar result [1.07±0.02 in the atopics (n=32) vs. 1.09±0.03 in the non-atopics (n=20), p=0.451].

The histamine-BRindex/methacholine-BRindex ratio was significant higher in the subjects with histamine-AHR or methacholine-AHR than in those subjects without it, regardless of atopy (Figure 3). Among the subjects without AHR, the ratio in the atopics was not higher than that in the others: the subjects with AHR also showed a similar result. Many of the subjects complained of side effects following high doses of inhaled histamine.

Table 2. Relationship among the variables

|                  | Histamine BRindex | Histamine FEV1 fall* | Methacholine BRindex | Methacholine FEV1 fall* | H/M BRindex |
|------------------|-------------------|----------------------|----------------------|------------------------|-------------|
| Methacholine BRindex | 0.609***           | 0.434***             | -                    | 0.935***               | -           |
| Methacholine FEV1 fall* | 0.589***           | 0.439***             | 0.282**              | 0.308**                | -0.122      |
| Blood eosinophils | 0.177              | 0.132                | 0.304***             | 0.253***               | -0.053      |
| Serum total IgE  | 0.284***           | 0.245***             | 0.331***             | 0.306***               | -0.108      |
| Atopy score      | 0.257***           | 0.214***             | 0.368***             | 0.394***               | -0.106      |
| House dust mites score | 0.304***           | 0.253***             | 0.331***             | 0.308***               | -0.053      |

BRindex: bronchial reactivity index; *: maximal % fall in forced expiratory volume in one second (FEV1) after histamine or methacholine bronchoprovocation; H/M: histamine/methacholine, p<0.05, **p<0.01, and ***p<0.001.

Figure 1. Relationships of the bronchial reactivity index (BRindex), which was calculated using a formula: log10 (10 + the maximal % fall in FEV1/log10 (the dose in mg/dL of the stimulus required to produce it)), and maximal % fall in the forced expiratory volume in one second (FEV1) after a bronchial challenge between histamine and methacholine. The dashed line is the line of identity.
Figure 2. Comparisons of the bronchial reactivity index (BRindex) and maximal % fall in the forced expiratory volume in one second (FEV₁) after a bronchial challenge between histamine and methacholine in the subjects with negative responses to both, a positive response to one, or positive responses to both histamine and methacholine tests.

Figure 3. Histamine (H)/methacholine (M)- bronchial reactivity index (BRindex) ratios in the study groups.
with mild histamine AHR suggests that further work is necessary for correctly interpreting the histamine-AHR test results of these subjects in both positive test responses or either one positive test response to histamine and methacholine.

Table 3. Clinical characteristics of the histamine-sensitive or methacholine-sensitive subjects as classified according to the histamine-BRindex/methacholine BRindex ratio > or < 1 for the subjects with both negative test responses or either one positive test response to histamine and methacholine

|                        | Histamine Sensitive | Methacholine Sensitive |
|------------------------|---------------------|------------------------|
| Age (years)            | 26.5±0.6            | 26.9±1.2               |
| Gender (M/F)           | 57/23               | 17/12                  |
| Height (cm)            | 170.5±0.9           | 168.2±1.5              |
| Blood eosinophils > 400/µL | 4 (5.0%)         | 5 (17.2%)              |
| Serum IgE > 100 IU/mL  | 34 (42.5%)          | 15 (51.7%)             |
| Atopy                  | 38 (48.1%)          | 17 (58.6%)             |
| House dust mites SPT† (+) | 34 (43.0%)      | 16 (55.2%)             |
| FEV1 (% predicted)     | 91.5±1.4            | 88.8±4.0               |

BRindex, bronchial reactivity index; FEV1, forced expiratory volume in one second
† Skin prick test.

DISCUSSION

In this study, the histamine-AHR was significantly related to, but significantly greater than the methacholine-AHR in the both negative group and the sole positive group, whereas it did not differ in the both positive group. Juniper et al.7 investigated patients with asthma and found that the histamine-AHR was comparable to the methacholine-AHR, which is consistent with our results of the both positive group. However, the greater AHR to histamine versus methacholine found in the subjects with mild histamine AHR suggests that further work is necessary for correctly interpreting the histamine-AHR test results of these groups. Woolcock et al.14 also reported greater sensitivity to histamine versus methacholine in COPD patients, whereas there was no difference in asthma patients. Cockcroft15 suggested a higher sensitivity to histamine versus methacholine even in asthma patients. It seems unlikely that the differences in sensitivity in our study resulted from any difference in the chemical stability of the histamine and methacholine solutions because the solutions were prepared fresh on the morning of each challenge day.

In addition, this study showed that AHR was related to atopy. It has been shown that the methacholine-AHR is significantly related to eosinophilia, the total IgE level and the skin test reactivity to allergens in asthma patients, which is consistent with our results. Then, because histamine is the typical mediator of the atopic allergic reaction and the number of histamine receptors is increased by leukotrienes, it can be speculated that atopic individuals may be specifically sensitive to histamine. Not only the animal studies by Antonissen et al.4, 5 but also the in vitro studies using human bronchial tissues have shown that exposure to allergen or an allergic mediator increases the response to histamine, but not the response to cholinergic agents. Moreover, Spector and Farr21 reported that patients with asthma were more sensitive to histamine than to methacholine, and atopic asthmatics could not tolerate high concentrations of histamine. Cockcroft et al.22 found that the allergen-induced increase in reactivity to histamine tended to be greater than that to methacholine, and it tended to persist longer.

However, for the patients with positive responses to both histamine and methacholine in this study, their histamine-AHR was not significantly different from their methacholine-AHR. Nonspecific AHR in asthma occurs via a variety of mechanisms, including genetic linkage, airway inflammation, smooth muscle alteration and so on. Therefore, even if the histamine-specific hypersensitivity in atopics would contribute to the overall AHR, it may be trivial in full-blown asthma, resulting in minimal difference between the histamine-AHR and the methacholine-AHR. Alternatively, due to the airway obstruction in asthma, the inhaled histamine may not readily reach its receptors, which are primarily localized in small airways, and this is unlike the methacholine receptors that are predominantly localized in the central airways, and so this obscures the atopy-related histamine-specific AHR.

In the subjects with mild histamine AHR in this study, atopy was not associated with an increased reactivity to histamine over methacholine. This discrepancy between our study and the previous studies4, 5, 19-22 may be related to the difference in disease activity between them. Most of the subjects in this study had no symptoms of atopic diseases on the study day, and only about 1/10 of the subjects had blood eosinophilia. The presence of AHR may suggest that the underlying airway disease is somewhat active, for which the leukotrienes that increase histamine receptor numbers are working without regard to atopy. Moreover, chronic exposure to histamine in active allergic diseases may decrease the response to histamine through the mechanism of tachyphylaxis25. Longitudinal studies are needed to examine whether histamine reactivity is predominantly increased during exacerbation of atopic diseases.

Inhaled histamine more frequently produces side effects such as throat irritation, flushing and headache than does inhaled methacholine. Many of our subjects complained of side effects when given high doses of inhaled histamine. Unfortunately, we did not record their side effects in detail and so we could not determine whether the atopic individuals had more side effects than the non-atopic individuals. Further, a positive bronchial challenge test does not always mean that a patient has asthma because AHR has been described in patients with allergic rhinitis and also in those patients with airflow limitation caused by conditions other than asthma. Therefore, further investigations involving taking a through history of allergy symptoms and...
conducting peak expiratory flow monitoring for a considerable period of time are required.

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