Phenotypic clustering of dilated cardiomyopathy patients highlights important pathophysiological differences

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Aims
The dilated cardiomyopathy (DCM) phenotype is the result of combined genetic and acquired triggers. Until now, clinical decision-making in DCM has mainly been based on ejection fraction (EF) and NYHA classification, not considering the DCM heterogeneity. The present study aimed to identify patient subgroups by phenotypic clustering integrating aetiologies, comorbidities, and cardiac function along cardiac transcript levels, to unveil pathophysiological differences between DCM subgroups.

Methods and results
We included 795 consecutive DCM patients from the Maastricht Cardiomyopathy Registry who underwent in-depth phenotyping, comprising extensive clinical data on aetiology and comorbidities, imaging and endomyocardial biopsies. Four mutually exclusive and clinically distinct phenogroups (PG) were identified based upon unsupervised hierarchical clustering of principal components: [PG1] mild systolic dysfunction, [PG2] auto-immune, [PG3] genetic and arrhythmias, and [PG4] severe systolic dysfunction. RNA-sequencing of cardiac samples (n = 91) revealed a distinct underlying molecular profile per PG: pro-inflammatory (PG2, auto-immune), pro-fibrotic (PG3; arrhythmia), and metabolic (PG4, low EF) gene expression. Furthermore, event-free survival differed among the four phenogroups, also when corrected for well-known clinical predictors. Decision tree modelling identified four clinical parameters (auto-immune disease, EF, atrial fibrillation, and kidney function) by which every DCM patient from two independent DCM cohorts could be placed in one of the four phenogroups with corresponding outcome (n = 789; Spain, n = 352 and Italy, n = 437), showing a feasible applicability of the phenogrouping.

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Conclusion

The present study identified four different DCM phenogroups associated with significant differences in clinical presentation, underlying molecular profiles and outcome, paving the way for a more personalized treatment approach.

Graphical Abstract

## Introduction

Non-ischaemic, non-valvular, dilated cardiomyopathy (DCM) represents a heterogeneous group of patients, as it results from a variety of genetic and acquired triggers.\(^1\) The heterogeneity makes it difficult to classify DCM with great precision to guide clinical decision-making. Until now, clinical decision-making in DCM has mainly been based on ejection fraction (EF) and NYHA classification,\(^3\) which does not recapitulate the complexity of the interactions with comorbidities and underlying aetiologies in the development and progression of DCM.\(^5\) Therapeutic modalities and outcome may differ depending on the underlying aetiologies.\(^7\) Overall, only 40–50% of DCM patients show left ventricular reverse remodelling (LVRR) after 12–24 months of optimal heart failure therapy.\(^9\) Classification which integrates the aetiological heterogeneity with cardiac function and comorbidities, may better reflect the biological differences between patients, and may allow guidance for a more targeted therapy.\(^10\) In an ideal situation, patients with corresponding pathophysiology would be classified together, creating more homogeneous patient populations.

Phenomapping based upon unsupervised clustering of clinical data may help to create homogeneous DCM subgroups, called phenogroups.\(^11\) Machine learning aids in detecting patterns between variables which explain the heterogeneity in a dataset.\(^12\) The
methodology has previously helped to create clinically valid phenogroups in heart failure with preserved and reduced EF.\textsuperscript{11,13} So far, studies applying machine learning to create phenogroups in patients with DCM are missing.

The goal of this study is (i) to identify patterns in clinical data and cluster patients based on phenotypic similarities; (ii) to study their underlying molecular profiles and possible pathomechanisms by performing RNA-sequencing on cardiac biopsies of patients from each phenogroup; and (iii) to create a clinical classifier based on the generated patients clusters and test its applicability in external DCM cohorts (Figure 1).

Methods

Study population and clinical information

The DCM diagnosis was defined according to the World Health Organization criteria and the latest ESC proposal.\textsuperscript{14,15} Enrolled patients presented with a left ventricular (LV) EF $\leq$50% at baseline echocardiographic evaluation in the absence of any of the following conditions: obstruction $\geq$50% of a major coronary artery branch [at coronary angiography (CAG)], pericardial diseases, congenital heart diseases, cor pulmonale, and active myocarditis. Patients, when not contraindicated, received guideline-directed medical therapy titrated to the maximal tolerated dose as well as device therapy (ICD and CRT-D implantation) according to the latest ESC guidelines.\textsuperscript{13} Three European DCM cohorts were analysed. The study was performed according to the declaration of Helsinki and approved by all the local Institutional Review Boards. All patients gave written informed consent.

Index cohort

The index cohort was derived from the prospective Maastricht Cardiomyopathy Registry of the Maastricht University Medical Center (MUMC, Maastricht, The Netherlands) between 2012 and 2019. As part of the diagnostic protocol, all patients received genetic counselling and DNA testing using a 47 DCM-associated gene panel (Supplementary material online, Table S1), and underwent a physical examination, blood sampling, 12-lead electrocardiogram, 24-h Holter monitoring, a complete echocardiographic and Doppler evaluation, and CAG at baseline. See the Supplementary material online, Methods for a detailed overview of the inclusion criteria, definitions of non-genetic disease modifiers and performed genetic analysis. Endomyocardial biopsies (EMB) and cardiovascular magnetic resonance imaging were performed if patient consented and was able to undergo these procedures. Table 1 demonstrates the phenotypic domains covering all of the collected clinical variables.

Application cohorts

The cohort from Italy was derived from the prospective Heart Muscle Disease Registry of Trieste Registry.\textsuperscript{56} Patients were enrolled between 2006 and 2019. All patients underwent physical examination, blood sampling, 12-lead electrocardiogram, 24-h ECG Holter monitoring, a complete echocardiographic and Doppler evaluation at baseline and during structured, systematic follow-up.\textsuperscript{57} Genetic testing was performed using a 23 DCM-associated gene panel. In order to exclude ischaemic heart disease, a CAG was systematically performed in patients $\geq$35 years, with cardiovascular risk factors and/or without familial history for DCM.

The cohort from Madrid included DCM patients followed in the Hospital Puerta de Hierros Inherited Cardiac Diseases Unit from 2004–19.\textsuperscript{57} All patients underwent a physical examination, blood sampling, genetic testing, a 12-lead electrocardiogram, echocardiogram, and systematic follow-up.

Follow-up

The median follow-up time of the Maastricht cohort was 53 months (interquartile range 28–98 months), 85 months (interquartile range 34–145 months) for Madrid and 40 months (interquartile range 13–77 months) for Trieste. Information about the occurrence of adverse events at follow-up was retrieved from the hospital medical records, the Personal Records Database, and/or telephone contact with the patient or their general practitioners in the three cohorts. We collected information regarding the following composite outcome measures: (i) death due to cardiovascular (CV) disease (heart failure or thrombo-embolic events), (ii) heart transplantation (Htx), (iii) LV assist device (LVAD) implantation, and (iv) life-threatening arrhythmias (LTA) defined as non-fatal ventricular fibrillation (with or without ICD shock), and/or sustained ventricular tachycardia treated with appropriate ICD shock. The combined endpoint is defined as the first occurrence of at least one of the above-mentioned adverse events: CV-related death, Htx, LVAD implantation, or an LTA.

Data processing

Figure 1 shows the study aims and design including all data processing steps, performed statistical analysis and application. The detailed description of these steps and used methods can be found in the Supplementary material online, Methods. Table 1 includes all 47 clinical parameters which were measured in the DCM patients, see Supplementary material online, Table S2 for an overview of the variable selection process. All 47 variables were used for data imputation using proximity from Random Forest for mixed-type data. Cardiovascular magnetic resonance parameters were imputed based on their equivalent echocardiographic parameters (Supplementary material online, Figure S1). The correlation matrix among variables is comparable between the imputed and non-imputed data, depicting high similarity (Supplementary material online, Figure S2). Fourteen variables with more than 25% missing data in the unimputed raw data were excluded from further analysis (Supplementary material online, Figure S3). Only late gadolinium enhancement (LGE) was kept in the analysis despite 29.8% of missing data, due to the established clinical importance of this variable.\textsuperscript{18} Five variables were excluded due to redundancy (Supplementary material online, Methods, Table S2). This left 28 variables for the clustering analysis.

Phenotypic clustering

We used optCluster (v1.3.0) to determine the stability and validity of various clustering methods to help determine the optimal clustering method (Supplementary material online, Methods). Phenotypic clusters were defined based on 28 clinical parameters without high multicollinearity using unsupervised hierarchical clustering of principal components (HCPC) approach, in which principle components of the clinical data were obtained with Factor Analysis for Mixed Data (FAMD), ensuring the balance of the influence between continuous and categorical variables. The optimal number of clusters was determined based on the gain in within-inertia (inside group variance) and using the Nbclust package (Supplementary material online, Figure S4). The over- or underrepresentation of character variables in each cluster were analysed by v-test based on the hypergeometric distribution. The clustering was performed with FactoMiner package v1.41 in R. The patient groups identified with the unsupervised clustering method were used as input for the decision tree modelling to identify the key parameters able to distinguish the created...
groups. Meaning that the decision tree modelling creates a classifier which can be applied on external cohorts, which is based on the unsupervised clustering. For this, conditional interference tree methods were used, a recursive binary partitioning with less overfitting and variable selection problems using the party package v1.3-3 in R. A detailed description of the used methods can be found in the Supplementary material online.

Methods. The Jaccard similarity index was calculated using the clusteval package to estimate the robustness between the unsupervised and supervised clustering.
Figure 2 Four mutually exclusive phenogroups as determined by hierarchical clustering of principal component using phenotypical information as input. The most distinct clinical characteristics are listed per phenogroup. Variables with an asterisk are key parameters to distinguish the phenogroups, as selected by supervised decision tree modelling (A). Characteristic plots of the four proposed phenogroups including their most representative clinical variables. The over- or underrepresentation of a variable within a cluster was analysed by v-test within the hierarchical clustering of principal component function, based on the hypergeometric distribution. A positive value indicates overrepresentation of this variable in the applicable phenogroup, a negative value indicates underrepresentation of the corresponding variable (B).
Table 2  Clinical characteristics stratified by phenogroup

| Clinical Characteristic | Group 1—mild systolic dysfunction (n = 331) | Group 2—auto-immune (n = 83) | Group 3—cardiac arrhythmias (n = 165) | Group 4—severe systolic dysfunction (n = 216) |
|-------------------------|---------------------------------------------|-------------------------------|--------------------------------------|-----------------------------------------------|
| Age, year               | 50.5 ± 13.1                                 | 52.5 ± 13.8                   | 57.9 ± 11.5                          | 56.3 ± 11.4                                   |
| Female, n (%)           | 150 (45)                                    | 49 (59)                       | 20 (12)                              | 77 (36)                                       |
| Disease modifiers, n (%)|                                            |                               |                                      |                                               |
| Genetic mutation        | 45 (14)                                     | 9 (11)                        | 94 (57)                              | 27 (13)                                       |
| TTNtv                   | 14 (4)                                      | 6 (7)                         | 43 (26)                              | 9 (4)                                         |
| LMNA                    | 0 (0)                                       | 1 (1)                         | 20 (12)                              | 1 (0)                                         |
| Other                   | 30 (9)                                      | 1 (1)                         | 24 (15)                              | 17 (8)                                        |
| Multiple                | 1 (0)                                       | 1 (1)                         | 7 (4)                                | 0 (0)                                         |
| Familial disease        | 59 (18)                                     | 15 (20)                       | 78 (41)                              | 48 (23)                                       |
| Toxic trigger           | 31 (9)                                      | 7 (8)                         | 9 (5)                                | 36 (17)                                       |
| Auto-immune disease     | 7 (2)                                       | 57 (69)                       | 2 (1)                                | 3 (1)                                         |
| Cardiac inflammation    | 50 (15)                                     | 16 (19)                       | 20 (12)                              | 44 (20)                                       |
| Significant viral load  | 31 (9)                                      | 11 (13)                       | 15 (9)                               | 25 (12)                                       |
| Electrical trigger      | 137 (41)                                    | 12 (14)                       | 73 (44)                              | 99 (46)                                       |
| NYHA functional class   | 33 (10)                                     | 26 (31)                       | 49 (30)                              | 114 (53)                                      |
| Hypertension, n (%)     | 117 (35)                                    | 36 (43)                       | 72 (44)                              | 72 (33)                                       |
| Body mass index, kg/m²  | 26.8 ± 5.2                                   | 24.4 ± 5.1                    | 28.3 ± 4.4                           | 26.4 ± 4.7                                    |
| Diabetes mellitus, n (%)| 22 (7)                                      | 14 (17)                       | 21 (13)                              | 29 (13)                                       |
| Heart rate, b.p.m.      | 72.3 ± 14.6                                  | 80.1 ± 18.1                   | 73.3 ± 16.6                          | 77.7 ± 17.2                                   |
| Systolic blood pressure, mm Hg | 134.6 ± 19.5 | 136.9 ± 26.3 | 135.8 ± 19.6 | 128 ± 19.6 |
| Diastolic blood pressure, mm Hg | 78.6 ± 12 | 79.9 ± 13.7 | 80.3 ± 12.6 | 78.4 ± 12.5 |
| Echocardiography        |                                            |                               |                                      |                                               |
| LV ejection fraction, % | 37.6 ± 8.6                                   | 33.4 ± 10.2                   | 32.7 ± 9.7                           | 21.8 ± 7.2                                    |
| LV EDD index, mm/m²     | 29 ± 3.7                                     | 30.9 ± 5.3                    | 28.9 ± 3.8                           | 34.7 ± 5.2                                    |
| LV ESD, mm              | 45 ± 7.3                                     | 46 ± 9.5                      | 49 ± 8.9                             | 60 ± 8.8                                      |
| LA volume index, mL/m²  | 39.2 ± 13.8                                  | 43.6 ± 16.8                   | 53.2 ± 16.5                          | 50.3 ± 19.1                                   |
| LV mass index, g/m²     | 95.4 ± 22.9                                  | 108.1 ± 32                    | 107 ± 25                             | 140.2 ± 36                                    |
| E/A ratio               | 1.1 ± 0.5                                    | 1.1 ± 0.5                     | 1.4 ± 0.9                            | 1.2 ± 0.8                                     |
| E/e’ ratio              | 7.5 ± 2.7                                    | 11.6 ± 5.2                    | 10.8 ± 5.1                           | 13.1 ± 4.3                                    |
| Posterior WT, mm        | 8.6 ± 1.3                                    | 9.1 ± 1.8                     | 9.1 ± 1.6                            | 9.4 ± 1.6                                     |
| Intraventricular ST, mm | 8.7 ± 1.5                                    | 9.1 ± 2.2                     | 9.6 ± 2                              | 9.2 ± 1.7                                     |
| Laboratory data         |                                            |                               |                                      |                                               |
| NT-proBNP, pmol/L       | 16 [7–42]                                    | 102 [39–520]                  | 94 [39–230]                          | 60 [19–229]                                   |
| Hs-Troponin T, ng/L     | 8 [4–12]                                     | 19 [9–48]                     | 18 [10–32]                           | 17 [10–36]                                    |
| Creatinine, µmol/L      | 83 [73–94]                                   | 113 [87–149]                  | 103 [89–120]                         | 97 [84–113]                                   |
| sIL-2R elevation, n (%) | 39 (19)                                      | 38 (60)                       | 17 (20)                              | 40 (29)                                       |
| Neopterin elevation, n (%) | 46 (22)                                      | 38 (62)                       | 29 (35)                              | 52 (39)                                       |
| Cardiac auto-antibodies, n (%) | 8 (12)                                      | 4 (4)                         |                                      | 5 (4)                                         |
| C-reactive protein, mg/L| 2 [1–5]                                      | 5 [2–11]                      | 2 [1–5]                              | 3 [1–11]                                      |
| ASAT, U/L               | 23 [19–28]                                   | 24 [19–28]                    | 28 [22–36]                           | 26 [21–33]                                    |
| ALAT, U/L               | 24 [19–34]                                   | 20 [15–27]                    | 32 [24–42]                           | 27 [19–40]                                    |
| ECG/Holter, n (%)       |                                            |                               |                                      |                                               |
| Atrial fibrillation     | 43 (13)                                      | 17 (20)                       | 100 (61)                             | 31 (14)                                       |
| Non-sustained VT        | 49 (15)                                      | 16 (19)                       | 103 (62)                             | 60 (28)                                       |
| Left bundle branch block| 94 (28)                                      | 8 (10)                        | 40 (24)                              | 83 (38)                                       |
| Atrioventricular block  | 26 (8)                                       | 7 (8)                         | 42 (25)                              | 21 (10)                                       |
| Out of hospital cardiac arrest | 12 (4)                                      | 2 (2)                         | 24 (15)                              | 13 (6)                                        |
| Cardiovascular MR (n = 558) |                                        |                               |                                      |                                               |
| LV mass index, g/m²     | 62.7 ± 15.4                                  | 66.8 ± 24.3                   | 66.4 ± 14.8                          | 94.2 ± 27.4                                   |
| LV EDV index, mL/m²     | 110.1 ± 24.6                                 | 113.5 ± 31                    | 115.1 ± 28.8                         | 178.1 ± 52.5                                  |

Continued
### RNA-sequencing

RNA was isolated from cardiac biopsies from DCM patients and checked for quality and integrity. The mRNA-sequencing library was generated using TruSeq mRNA sample preparation kit (Illumina) and sequenced on the NextSeq 500 (Illumina). Molecular pathway analysis was performed using gene enrichment in Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways. The RNA-sequencing steps and bioinformatical analysis are described in more detail in the Supplementary material online, Methods.

### Statistical analysis

The link between clinical variables and the created clusters was analysed by ANOVA for numeric and χ²-test for categorical variables. The over- or underrepresentation of a variable within a cluster was analysed by v-test within the HCPC function, based on the hypergeometric distribution. A P-value < 0.05 was considered as statistically significant.

For outcome analyses, we used unadjusted and multivariable-adjusted Cox proportional hazards models to determine the independent association between patient groups and outcome, after testing the proportional hazards assumption. Primary outcome was the combined endpoint of CV death, Hb, LVAD implantation, and/or an LTA. Covariates included in the adjusted analysis included variables known to be predictive of outcomes in DCM such as NT-proBNP, EF, creatinine, heart rate, NYHA class, hypertension, LGE, atrial fibrillation (AF), age, and gender. Model discrimination (c-statistics comparison using DeLong test), calibration (likelihood ratio test), and reclassification ([continuous] net reclassification index (NRI), and integrated discrimination index (IDI)) were evaluated for the clinical predictors with and without phenogrouping. Kaplan–Meier survival curves were estimated, and differences between groups were assessed by the log-rank test, using time at diagnosis as time zero. Calculations were done using SPSS version 23.0 (SPSS Inc., Chicago, IL, USA).

### Results

#### Comparison of clinical characteristics among phenogroups

Figure 1 summarizes the data processing of the clinical variables, selecting 28 variables as input for the cluster analysis (Table 1, Supplementary material online, Table S2). Four was the minimal number of clusters which could accurately reflect the phenotypic variation in the index cohort (Supplementary material online, Figure S4). Twenty-seven of the 28 clinical input variables had a contribution in defining the patient clusters. Only the cardiac viral load in EMBs did not contribute (Figure 2, Supplementary material online, Table S2). This led to the identification of four phenogroups with significant differences in their clinical characteristics (Table 2, Figure 2).

Phenogroup 1 (PG1; n = 331; 42%) had a moderate reduction in left ventricular ejection fraction (LVEF; 43% ± 9%) and overall smallest cardiac dimensions, lower NT-proBNP [16 (7–42) pmol/L] and NYHA class I and II (90%) compared with the other patients. Phenogroup 4 (PG4; n = 216; 27%) appears to be the opposite pole of severity, consisting of patients with a low LVEF (23% ± 8%), and the largest cardiac dimensions. More than half of the patients in this subgroup had NYHA III or IV (53%).

Patients in phenogroup 2 (PG2) and 3 (PG3) more reflect the aetiological diversity, irrespective of cardiac function and clinical presentation. Patients in PG2 (n = 83; 10%) consisted primarily of patients with an auto-immune disease (69%), females (59%), and chronic kidney disease [creatinine: 113 (87–149) μmol/L]. In contrast, PG3 (n = 165; 21%) consisted mainly of males (88%) with a high prevalence of cardiac arrhythmias [mainly AF (61%) and NSVT (62%)], pathogenic gene variants (57%), and LGE at MRI (59%). Two-third of the gene variants were pathogenic TTNtv and LMNA variants (Table 2).

#### RNA-sequencing of cardiac biopsies to unravel underlying pathophysiology

To gain better insight into the cardiac pathophysiological differences between the phenogroups, genome-wide transcriptome analysis (RNA-sequencing of EMB) was performed in a sub-cohort of patients with available spare biopsies for RNA-sequencing (n = 91, distribution phenogroups 1–4 = 21/9/35/26). All phenogroups had a distinct transcriptomic signature separating them from the other groups (Figure 3A). PG4 (low EF) had large differences in gene expression (Figure 3A), again reflected in the transcriptomic signature of the created clusters (Figure 3B).

**Figure 1**: Summary of the data processing of the clinical variables, selecting 28 variables as input for the cluster analysis (Table 1, Supplementary material online, Table S2). Four was the minimal number of clusters which could accurately reflect the phenotypic variation in the index cohort (Supplementary material online, Figure S4). Twenty-seven of the 28 clinical input variables had a contribution in defining the patient clusters. Only the cardiac viral load in EMBs did not contribute (Figure 2, Supplementary material online, Table S2). This led to the identification of four phenogroups with significant differences in their clinical characteristics (Table 2, Figure 2).

- **Phenogroup 1 (PG1)**: n = 331; 42%. Moderate reduction in LVEF (43% ± 9%) and overall smallest cardiac dimensions. Lower NT-proBNP [16 (7–42) pmol/L] and NYHA class I and II (90%) compared with other patients. PG4 appears to be the opposite pole of severity, consisting of patients with a low LVEF (23% ± 8%), and the largest cardiac dimensions. More than half of the patients in this subgroup had NYHA III or IV (53%).
- **Phenogroup 2 (PG2) and 3 (PG3)**: Reflect aetiological diversity, irrespective of cardiac function and clinical presentation. PG2 consisted primarily of patients with an auto-immune disease (69%), females (59%), and chronic kidney disease (creatinine: 113 (87–149) μmol/L). PG3 consisted mainly of males (88%) with a high prevalence of cardiac arrhythmias (mainly AF and NSVT), pathogenic gene variants (57%), and LGE at MRI (59%). Two-third of the gene variants were pathogenic TTNtv and LMNA variants (Table 2).

**Figure 2**: The link between clinical variables and the created clusters was analysed by ANOVA for numeric and χ²-test for categorical variables. The over- or underrepresentation of a variable within a cluster was analysed by v-test within the HCPC function, based on the hypergeometric distribution. A P-value < 0.05 was considered as statistically significant.

**Figure 3A**: Large differences in gene expression (Figure 3A) again reflected in the transcriptomic signature of the created clusters (Figure 3B).

**Table 2** Continued

| Clinical Characteristic | Group 1—mild systolic dysfunction (n = 331) | Group 2—auto-immune (n = 83) | Group 3—cardiac arrhythmias (n = 165) | Group 4—severe systolic dysfunction (n = 216) |
|------------------------|------------------------------------------|-----------------------------|--------------------------------------|------------------------------------------|
| LV ESV index, mL/m²    | 63.4 ± 21                                | 73.8 ± 31.5                 | 75.1 ± 27.4                          | 138.7 ± 51.6                            |
| LV stroke volume index, mL/m² | 46.8 ± 11                               | 40.1 ± 11                   | 40.3 ± 12.5                          | 39 ± 12.8                               |
| LV ejection fraction, % | 42.9 ± 8.8                               | 37.4 ± 12.5                 | 36.1 ± 11.1                          | 22.8 ± 8.2                              |
| Late gadolinium enhancement, n (%) | 33 (52)                                 | 68 (59)                     | 42 (29)                              |                                           |
| Endomyocardial biopsy  |                                         |                             |                                      |                                          |
| Collagen volume fraction, %/area | 4.7 [2–7]                              | 6 [3–10]                    | 6 [3–11]                             | 6.2 [3–10]                              |

Categorical variables are presented as counts and percentages; continuous variables are presented as mean ± SD; and right-skewed variables are presented as median (25th–75th percentile).

ALAT, alanine transaminase; ASAT, aspartate transaminase; EDD, end-diastolic diameter; EDV, end-diastolic volume; ESD, end-systolic diameter; ESV, end-systolic volume.; LA, left atrial; LMNA, lamin A/C; LV, left ventricular; MR, magnetic resonance; NYHA, New York Heart Association; sIL-2R, soluble interleukin-2 receptor; ST, septum thickness; TTNtv, truncating titin variants; VT, ventricular tachycardia; WT, wall thickness.
expression, as reflected by the number of differentially expressed genes and strong separation (*Figure 3B*). Subsequent gene set enrichment analysis with KEGG terms revealed unique expression of molecular pathways per phenogroup (*Figure 3C, Table 3*). Overall, cardiac metabolism was the strongest differentially expressed genes in the comparison between two corresponding PG (FDR < 0.01 + fold change >1.5) (*B*). Significantly enriched Kyoto Encyclopedia of Genes and Genomes pathways (*P*-value < 0.05) in the comparison between the PG (Table 4) (*C*).

**Association of phenogroups with outcome**

We tested whether the four different phenogroups were associated with the combined outcome measure as defined by CV death, Htx, LVAD implantation or the occurrence of an LTA. PG1 (mrEF) has the lowest, and PG3 (arrhythmia) the highest risk for an adverse event (1.1 vs. 6.5 events per 100 patient-years; *P* < 0.001; *Figure 4, Table 4*). Both PG2 (auto-immune) and PG4 (low EF) shared a high-risk profile with 4.2 events per 100 patient-years. PG3 (arrhythmia) remained a high-risk subgroup, even after correction of well-established prognostic factors (Table 4). The addition of phenogroup information improved the outcome prediction beyond the clinical predictors (*c*-statistic 0.63–0.734,
The arrow indicates the direction of the pathway (up- or down-regulation) in the cardiac RNA-sequencing data from the underlined phenogroup compared with the phenogroup in direct comparison.

KEGG, Kyoto Encyclopaedia of Genes and Genomes.

\*Number of significant differentially expressed genes between datasets which are represented in the specific pathway.

\*A P-value < 0.05 indicates a significant enrichment of the corresponding pathway in the comparison.

***Supervised decision tree modelling to enhance clinical utility***

We applied supervised decision tree modelling with the phenogroups from the clustering as input, to create a classifier consisting

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**Table 3** Molecular Kyoto Encyclopaedia of Genes and Genomes pathway analysis in RNA-seq data from endomyocardial biopsies of DCM patients stratified per phenogroup

| Phenogroup 2 vs. Phenogroup 1 | Number of genes | P-value | Top enriched KEGG pathway (functional annotation chart) | Number of genes | P-value |
|-------------------------------|----------------|---------|-------------------------------------------------------|----------------|---------|
| NF-kappa B signalling pathway | 12             | 7.5E-6  | NF-kappa B signalling pathway | 12             | 7.5E-6  |
| RIG-I-like receptor signalling pathway | 7             | 7.0E-3  | Metabolic pathways | 201             | 4.1E-4  |
| Glutathione metabolism | 4             | 3.4E-2  | Focal adhesion | 7.9E-3 | 4.3E-3 |

**Phenogroup 3 vs. Phenogroup 1**

| KEGG pathway | Number of genes | P-value | KEGG pathway | Number of genes | P-value |
|--------------|----------------|---------|--------------|----------------|---------|
| ECM-receptor interaction | 7         | 7.9E-2 | Focal adhesion | 12         | 4.3E-3 |
| Focal adhesion | 12     | 4.3E-3 | TNF-signalling pathway | 17         | 5.0E-5 |

**Phenogroup 4 vs. Phenogroup 1**

| KEGG pathway | Number of genes | P-value | KEGG pathway | Number of genes | P-value |
|--------------|----------------|---------|--------------|----------------|---------|
| DNA replication | 19       | 1.3E-7 | Metabolic pathways | 201         | 4.1E-4 |
| Dilated cardiomyopathy | 16        | 1.4E-3 | Focal adhesion | 7.9E-3 | 4.3E-3 |
| Fatty acid metabolism | 7       | 1.8E-1 | Focal adhesion | 7.9E-3 | 4.3E-3 |

**Phenogroup 2 vs. phenogroup 3**

| KEGG pathway | Number of genes | P-value | KEGG pathway | Number of genes | P-value |
|--------------|----------------|---------|--------------|----------------|---------|
| RIG-I-like receptor signalling pathway | 5         | 2.1E-2 | Metabolic pathways | 17         | 5.0E-7 |
| T-cell receptor signalling | 8       | 3.9E-2 | DNA replication | 12         | 4.3E-3 |
| B-cell receptor signalling | 5       | 7.0E-2 | DNA replication | 12         | 4.3E-3 |

**Phenogroup 4 vs. phenogroup 2**

| KEGG pathway | Number of genes | P-value | KEGG pathway | Number of genes | P-value |
|--------------|----------------|---------|--------------|----------------|---------|
| cGMP-PKG signalling pathway | 29       | 5.3E-3 | Metabolic pathways | 188        | 5.2E-3 |
| DNA replication | 10       | 3.7E-2 | Dilated cardiomyopathy | 15        | 6.7E-3 |
| Fatty acid metabolism | 7       | 1.8E-1 | Focal adhesion | 7.9E-3 | 4.3E-3 |

**Phenogroup 4 vs. phenogroup 3**

| KEGG pathway | Number of genes | P-value | KEGG pathway | Number of genes | P-value |
|--------------|----------------|---------|--------------|----------------|---------|
| DNA replication | 16       | 9.2E-5 | Metabolic pathways | 200        | 5.3E-2 |
| PI3K-Akt signalling pathway | 11       | 4.6E-2 | DNA replication | 12         | 4.3E-3 |
| Focal adhesion | 15       | 6.5E-2 | DNA replication | 12         | 4.3E-3 |

\( P = 0.006; \) likelihood ratio test \( P < 0.001; \) IDI 6%; \( P < 0.001; \) NRI 20%; \( P = 0.026; \) Supplementary material online, Table S3. Overall, the prognosis of PG2, 3 and 4 were comparable (Figure 4), reflecting the influence of aetiological fraction (Group 2: auto-immune disease Group 3: genetics) on prognosis irrespective of cardiac function (Group 4). Patients classified in PG3 (arrhythmia) had specifically an increased risk for LTAs compared with the other phenogroups (4.1 events per 100 patient-years; Table 4).

52.5% of the patients showed LVRR after 12 months of guideline-directed medical therapy (Supplementary material online, Table S4). Patients in PG4 (low EF) had the highest rate of LVRR (61%) compared with PG1-3 (48%, 49%, and 50%, respectively). Although patients in PG2-4 had similar or a higher rate of LVRR compared with PG1 (mrEF), they still had a higher event rate, which could be partly explained by differences in disease mechanism.

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**Figure 4** Event-free survival stratified by phenogroup. Kaplan–Meier curves for the combined outcome of life-threatening arrhythmias, cardiovascular death, heart transplantation, or left ventricular assist device implantation stratified by phenogroup.
of only a few clinical variables which can still place a patient in their corresponding phenogroup. A low number of variables in such classifier will increase the likelihood that it can be applied in other DCM patient cohorts, instead of the extensive dataset which was needed to identify the phenogroups. The presence of an auto-immune disease, LVEF, AF, and creatinine were selected as the most important variables characteristic for these phenogroups with a combined accuracy of 71% (Figure 5, Supplementary material online, Figure S5, Table S5). However, placing patients in PG2 (auto-immune) had an accuracy of 95%, mostly by the presence of an auto-immune disease (Supplementary material online, Table S5). The decision tree has the most difficulty in recognizing patients from PG3 (arrhythmia), mainly due to the placement is completely independent of LVEF (nodes 5 and 9 in Supplementary material online, Figure S5; accuracy of 80% Supplementary material online, Table S5).

### Discussion

We identified four distinct phenogroups with unique clinical characteristics, reflecting the integration of cardiac function, comorbidities, and underlying aetiology. Each phenogroup had a distinct cardiac transcriptomic profile, which could pave the way to a more targeted and personalized treatment approach (Take home figure).

### Identified phenogroups in a heterogeneous dilated cardiomyopathy dataset

The four mutually exclusive phenogroups integrated a broad range of clinical variables: (i) younger patients with mild systolic dysfunction, small cardiac dimensions, and relatively few symptoms; (ii) young females with a clinical history of auto-immune disease with cardiac and renal involvement; (iii) males with a genetic mutation and prevalent ventricular and supraventricular arrhythmias; and (iv) patients with severe systolic dysfunction, large cardiac dimensions, moderate diastolic dysfunction, and pronounced symptoms. All phenogroups had a distinct cardiac transcriptomic signature, characterizing differences in underlying molecular pathology. This confirms that the generated phenogroups are more homogeneous patient subgroups who could share a common pathophysiology compared with patients in other phenogroups. Supervised decision tree modelling identified four parameters that are sufficient to adequately cluster patients in these subgroups: EF (PG1 and 4), auto-immune disease and creatinine (PG2), and AF (PG3). We could successfully apply this decision tree on an external application cohort placing every patient in one of the four phenogroups.

### Clinical validity of created phenogroups

The present study puts forward an approach of DCM patient classification based on the integration of extensive, but well-accessible...
A recent post-hoc analysis of the POSEIDON-DCM trial revealed a genetic aetiology in a large percentage of non-responders. This illustrates the heterogeneity of DCM and the difficulty to recognize all clinical variables influencing the clinical course of a patient. The variety of underlying disease mechanisms in DCM could be an important reason for the variation in treatment efficacy in clinical trials. The identification of valid DCM patient subgroups with distinct underlying molecular profiles may therefore help to better guide future DCM clinical trials.

Figure 5 A simplified adaptation of the decision tree model shows the clinical parameters which are the core of the phenogroups. The black box indicates that the model cannot accurately place the patient in one of the four phenogroups (see Supplementary material online, Figure S5 for exact accuracy).

Figure 6 Event-free survival stratified by phenogroup as determined by the supervised decision tree model. Kaplan–Meier curves for the combined outcome of life-threatening arrhythmias, cardiovascular death, heart transplantation, or left ventricular assist device implantation stratified by phenogroup in the application cohort (Madrid and Trieste).

diagnostic tools. A recent post-hoc analysis of the POSEIDON-DCM trial revealed a genetic aetiology in a large percentage of non-responders. This illustrates the heterogeneity of DCM and the difficulty to recognize all clinical variables influencing the clinical course of a patient. The variety of underlying disease mechanisms in DCM could be an important reason for the variation in treatment efficacy in clinical trials. The identification of valid DCM patient subgroups with distinct underlying molecular profiles may therefore help to better guide future DCM clinical trials.

PG1 (mrEF) and PG4 (low EF) represent two opposites of DCM severity, which is mainly reflected in their LVEF. PG4 (low EF) had a distinct transcriptomic profile, reflecting a cardiac metabolic shift away from fatty acid metabolism towards glycolytic substrate usage, which is characteristic for an increasing heart failure severity as the heart has to metabolically adapt to the increased cardiac energy demand. Furthermore, purine and pyrimidine metabolism was increased, which was in line with the increased DNA replication pathways, which is also a hallmark of heart failure. PG2 (auto-immune) and PG3 (arrhythmia) are distinct disease subgroups independent of LVEF. Cardiac involvement in patients with an auto-immune disease (PG2) was associated with a high-risk profile (PG2). As one would expect, pro-inflammatory pathways predominated in the autoimmune PG2, and this PG could therefore benefit from additional
immunomodulatory therapy as suggested in the position statement of the ESC.\textsuperscript{22}

Patients in PG3 are characterized by cardiac arrhythmias, pathogenic gene variants, and increased cardiac fibrosis. Patients in PG3 had the highest risk profile, with a five times higher risk for LTAs compared with DCM patients in PG1 even after adjustment for well-known clinical predictors such as LVEF and LGE. The close interaction between arrhythmias, fibrosis, and gene variants created difficulties for the decision tree to pick one variable that could place patients in PG3. Expanding genotype-phenotype knowledge (i.e. unveiling the association and disease expression of specific gene variants with AF) could be valuable input to finetune PG3 and enhance the decision tree accuracy. Finding the gene variant which causes the electrical disturbance may help to provide more targeted therapy in otherwise heart failure (HF)-therapy resistant patients in the future. This is exemplified in DCM patients with a pathogenic lamin A/C (\textit{LMNA}) variant, in which hyperactivation of the p38\textsubscript{a} signalling pathway plays a central pathophysiological role.\textsuperscript{23} ARRY-797, an oral, selective p38 inhibitor is currently under investigation in DCM patients with an \textit{LMNA} variant in a randomized, double-blind, placebo-controlled phase 3 trial (NCT03439514). It is not unexpected that this PG had the highest prevalence of \textit{LMNA} and \textit{TTN} variants along with NSVTs, both known risk factors for LTA.\textsuperscript{5,24} This could indicate that patients in this specific phenogroup could benefit the most from prophylactic ICD therapy before advanced LV dysfunction as is currently only implemented in the guidelines for \textit{LMNA}, shedding new light on the outcome of the DANISH trial.\textsuperscript{4,25}

**Future directions of dilated cardiomyopathy patient care**

The main aim of this study was to find patterns in clinical data which represent patient groups who share a similar pathophysiology. This finding precedes potential therapeutic implications which deserve further investigation: (i) response to currently used HF medication per PG and (ii) targeted therapy aimed at the underlying

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**Take home figure** This study identified four phenotypic clusters based on patterns in clinical data which had a unique underlying molecular profile identified by RNA-sequencing on cardiac biopsies of patients from each phenogroup. A classifier derived from the initial clustering could be successfully applied on an external cohort placing every DCM patient in a specific phenogroup.

| Phenogroup 1 | Phenogroup 2 | Phenogroup 3 | Phenogroup 4 |
|-------------|-------------|-------------|-------------|
| Mild systolic dysfunction | Auto-immune | Cardiac arrhythmias | Severe systolic dysfunction |

**EXTERNAL APPLICATION OF CLUSTERING**

- Auto-immune disease
- Left ventricular ejection fraction
- Atrial fibrillation
- Creatinine

**APPLICATION COHORT (n=789)**

**PG1 – Mild systolic dysfunction**

**PG2 – Auto-immune**

**PG3 – Cardiac arrhythmias**

**PG4 – Severe systolic dysfunction**

**INDEX COHORT (n=795)**

**DILATED CARDIOMYOPATHY**

**RNA-SEQUENCING**

- Pro-inflammatory
- Pro-fibrotic
- Cardiac metabolism
- Beta oxidation

**NFkB**
Pathomechanism of the specific PG. In-depth analysis of transcriptomic data per phenogroup and additional functional studies are next steps to elucidate specific molecular targets for treatment.

Although we focused on clinical utility, the effect on clinical decision-making remains elusive. We acknowledge that the current approach is a major departure from the classical studies, but it is a first attempt to a more personalized approach of the DCM patient in everyday clinical practice. Application of patient clustering as a tool to guide clinical decision-making should be further explored in prospective studies. Besides a refined DCM clustering and ensuing targeted therapies, an additional goal would be the development of a comprehensive DCM risk score for sudden cardiac death and LTA, as already available for hypertrophic cardiomyopathy.\(^{26}\)

Limitations and perspectives
In this study, we chose for an unsupervised approach with hierarchical k-means clustering of principal components to look in an unbiased way for underlying clinical patterns. The HCPC method allows us to use principal components instead of individual variables and maximize robustness by an additional k-means consolidation step, which can be used for FAMD. This is a different strategy from model-based clustering methods, which are semi-supervised and more appropriate to improve prognostic assessment. The analysis of the registries is of a retrospective nature, although patients are included prospectively. The results and clinical implications of this study need therefore to be validated in a prospective manner, not only for predicting outcome, but also for its more value in predicting the response to existing or novel HF therapies.

The definition of systemic immune-mediated diseases (based on the recent position statement of the ESC\(^{22}\)) includes a broad spectrum of immune-mediated diseases with diverse implication in cardiac disease development. Larger registries studying this specific immune-mediated subset of DCM patients are needed to be able to look at the specific immune-mediated diseases and their impact on long-term outcome.

Conclusions
The present study identified four different DCM phenogroups associated with significant differences in clinical presentation, underlying molecular profiles and outcome, paving the way for a more personalized treatment approach.

Data availability
The data that support the findings of this study are available from the corresponding author on reasonable request.

Supplementary material
Supplementary material is available at European Heart Journal online.

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