Combined Detection of Preoperative Neutrophil-to-Lymphocyte Ratio and CEA as an Independent Prognostic Factor in Nonmetastatic Patients Undergoing Colorectal Cancer Resection Is Superior to NLR or CEA Alone

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Received 9 December 2016; Accepted 12 February 2017; Published 8 June 2017

Objective. To explore the role of combined detection of carcinoembryonic antigen (CEA) and neutrophil-to-lymphocyte ratio (NLR) in the prognostic assessment of colorectal cancer (CRC).

Methods. We investigated preoperative NLR and CEA in 125 surgical CRC patients, determined the patients’ thresholds by receiver operating characteristic (ROC) curve analysis, and assessed their prognostic values by Kaplan–Meier curve and Cox regression models. In addition, we used nomograms of several risk factors to evaluate the risk in survival and predictive accuracy by using Harrell’s concordance index (c-index).

Results. Resultsofmultivariate analysis showed high NLR, high CEA, and high COCN (combination of CEA and NLR) were significantly correlated with decreased disease-free survival (DFS) [HR: 2.229, 95% CI: 1.012–4.911, and \( P = 0.047 \); HR: 3.652, 95% CI: 1.630–8.179, and \( P = 0.002 \); HR: 3.139, 95% CI: 1.800–5.472, and \( P < 0.001 \)]. But high CEA and COCN remained significant only for decreased overall survival (OS) [HR: 3.713, 95% CI: 1.396–9.873, and \( P = 0.009 \); HR: 3.106, 95% CI: 1.576–6.123, and \( P = 0.001 \)]. High NLR showed higher mortality rates with worse OS (\( P = 0.058 \)), and nomograms containing NLR improved the predictive accuracy. Area under the curve of COCN was higher than that of CEA or NLR.

Conclusion. COCN acts as a better independent prognostic biomarker of CRC than NLR or CEA alone.

1. Introduction

Colorectal cancer (CRC) is one of the most frequently diagnosed tumors and is the fourth leading cause of cancer-related deaths worldwide, with an annual incidence of 148,000 new cases [1]. Overall survival (OS) of CRC patients is poor, and more than one-third of CRC patients die within 5 years [2]. Although the 5-year OS of CRC patients has been improved remarkably in recent years owing to the advances in surgical techniques and other therapies, 40–50% of patients who underwent colorectal resection developed recurrences or died due to metastatic disease [3, 4].

Inflammatory response plays a key role in the survival of cancer patients. Several recent studies have demonstrated that systemic inflammatory response (SIR) markers, including C-reactive protein (CRP) [5], Glasgow prognostic score (GPS) [6], platelet-to-lymphocyte ratio (PLR), and lymphocyte-to-monocyte ratio (LMR) [7], are correlated with poor survival rates of CRC and many other cancers [5, 8]. The neutrophil-to-lymphocyte ratio (NLR) which is considered as one of the SIR markers has been reported to be associated with the prognosis in patients with various types of cancer [9–12]. Few recent studies have reported the role of NLR as a prognostic factor for CRC patients [5, 8]. However, most of these studies focused on the prognostic role of advanced or metastatic CRC [13–17], and few studies have investigated the prognostic value of NLR in resectable stages II and III primary CRC. Specifically, there were no studies till date...
that have explored the prognostic role of combined detection of NLR and CEA in patients undergoing colorectal cancer resection and compared the combination with NLR or CEA alone.

It is increasingly recognized that survival of cancer patients was determined not only by the host SIR but also by tumor characteristics [17]. Carcinoembryonic antigen (CEA) is a widely used tumor-related marker for prognostic prediction of CRC patients [2, 8]. We therefore hypothesize that identifying parameters reflecting both tumor characteristics and host SIR may be a better approach for predicting patient survival, and COCN (combination of CEA and NLR) may be a better biomarker in the prognostic assessment of CRC.

Hence, in the present study we first evaluated the prognostic utility of NLR or CEA alone in patients undergoing surgery for CRC and then explored the prognostic value of COCN, a novel inflammation-based prognostic system with tumor characteristics, in an attempt to provide experimental clues for better prediction of CRC prognosis in patients.

2. Materials and Methods

2.1. Patient Selection. One hundred and sixty patients were included in this study with pathologically confirmed stages II and III CRC without distant metastasis or local recurrence who received surgical resection in Changhi Hospital (Shanghai, China) between March 2013 and October 2014. All patients received curative resection and the surgical procedures were performed by the same surgical team of the same department in all patients. The inclusion criteria included patients who were presented with clinical signs of systemic inflammation or infection, hematological diseases, evidence of hyperpyrexia, enterobrosis, onset of intestinal obstruction during hospitalization, or a history of other malignancies. All the procedures were performed in accordance with the ethical standards of the institutional and/or national research committee and with the Helsinki declaration and were approved by the Ethics Committee of Changhi Hospital with the permit number of 2012B0076, and informed consent was obtained from all included patients.

2.2. Clinical Data and Laboratory Methods. Neutrophil count, lymphocyte count, red blood cell (RBC), platelet count, fibrinogen (Fib), CEA, and carbohydrate antigen 199 (CA199) were measured within 3 days prior to the surgery as part of the routine preoperative workup in these patients. Full blood count (FBC) was analyzed with a Sysmex XN-9000 automated hematology analyzer (Sysmex Corporation, Kobe, Tokyo, Japan). Fib was measured with a Sysmex CS-5100 automatic coagulation analyzer (Sysmex UK Ltd., Milton Keynes, UK). CEA and CA199 were conducted by Roche Elecsys 2010 Chemistry Analyzer (Basel, Switzerland). NLR or PLR was calculated by dividing the absolute number of neutrophils or platelets by the absolute number of lymphocytes, respectively.

The clinical data were collected via the hospital information system, which included age, gender, smoking, cancer site, tumor stage, histological class, differentiation, tumor size, and family history. Tumor staging was performed according to the tumor-node-metastasis (TNM) classification of the American Joint Committee on Cancer (AJCC, 7th edition).

2.3. Survival and Follow-Up. All 160 patients were put on a regular follow-up program on the outpatient basis for every 3–6 months for the first 2 years and every 6 months for the next 3 years, which included physical examination, CEA and CA199 test, chest X-ray, abdominal ultrasonography, or abdominal CT and colonoscopy every 1–3 years. Of them, 35 patients were excluded from the study due to loss of follow-up for various reasons.

Recurrence was detected by a combination of imaging studies and tumor markers, such as CEA and CA199, and finally was confirmed by pathological examination [2]. OS was calculated from the date of diagnosis to the date of death or last follow-up visit in survivors (23) [18]. Disease-free survival (DFS) was measured from the date of surgery to the date of disease recurrence or the date of last follow-up visit in patients without recurrences [18].

2.4. Statistical Analysis. The cut-off values of NLR, PLR, Fib, CA199, CEA, and RBC were determined by receiver operating characteristic (ROC) curve analysis. According to each cut-off value, patients were divided into two groups. Furthermore, according to COCN score as shown in Table 1, patients were assigned into three groups. Differences in clinicopathological characteristics as grouped by NLR and CEA were compared using the Pearson $\chi^2$ test or Fisher’s exact test. Survival analysis was performed using the Kaplan–Meier method and Cox proportional hazard model. The variates with significant differences were identified from the univariate analysis and were selected for forward multivariate Cox regression survival analysis. Differences were estimated by log-rank test. Statistical significance was set at a level of $P < 0.05$. All analyses were performed using SPSS software 21.0 (SPSS Inc., Chicago, USA), and nomogram was explored by R version 3.0.0 software (Institute for Statistics and Mathematics, Vienna, Austria). The optimal thresholds were selected using R package MAXSTAT [19].

| Scoring system | Score |
|----------------|-------|
| NLR ≥ 2.43     | 1     |
| NLR < 2.43     | 0     |
| CEA ≥ 5 ng/mL  | 1     |
| CEA < 5 ng/mL  | 0     |
| Combination of CEA and NLR (COCN) | |
| NLR ≥ 2.43 and CEA ≥ 5 ng/mL | 2   |
| NLR ≥ 2.43 or CEA ≥ 5 ng/mL | 1   |
| Neither NLR ≥ 2.43 nor CEA ≥ 5 ng/mL | 0   |

NLR = neutrophil-to-lymphocyte ratio; CEA = carcinoembryonic antigen.
3. Results

3.1. ROC Curves of CEA and NLR for Both DFS and OS. Using ROC analysis, the cut-off value of NLR was calculated based on which patients were assigned to a high NLR (≥2.43) group and low NLR (<2.43) group. Area under the curve (AUC) was 0.605 (95% CI: 0.487 to 0.723) for DFS and 0.619 (95% CI: 0.487 to 0.751) for OS (Figures 1(a) and 1(b)). A cut-off value of 2.43 was chosen as an optimal NLR value for evaluating DFS and OS. This value showed a 0.688 sensitivity and 0.606 specificity for OS and 0.630 sensitivity and 0.622 specificity for DFS. Similarly, based on ROC analysis, a cut-off value of 5 was used for CEA (ng/mL) in our study. Similarly, ROC analysis was applied to PLR, Fib, CA199, and RBC, which showed cut-off values of 113.5, 3.30 (g/mL), 7.42 (U/mL), and 4.43 (×10^12/mL), respectively.

3.2. Correlations of NLR and CEA with Clinicopathological Factors. Correlations of NLR and CEA with various clinicopathological factors including Fib, CA199, PLR, and other clinicopathological characteristics were analyzed. As shown in Table 2, high NLR was significantly correlated with cancer
Table 2: Characteristics of the 125 patients grouped by NLR and CEA.

| Variables          | Sum (%) | NLR ≥2.43 | P value | CEA (ng/mL) ≤5 | P value |
|--------------------|---------|-----------|---------|----------------|---------|
|                    |         | <2.43     |         |                |         |
| Age (year)         |         |           |         |                |         |
| <65                | 70 (56) | 32        | 38      | 0.587          | 23      | 47      | 0.265 |
| ≥65                | 55 (44) | 22        | 33      |                | 24      | 31      |        |
| Gender             |         |           |         |                |         |
| Female             | 52 (0.42) | 24   | 28      | 0.588          | 22      | 30      | 0.454 |
| Male               | 73 (0.58) | 30   | 43      |                | 25      | 48      |        |
| Smoking            |         |           |         |                |         |
| Yes                | 116 (0.92) | 50  | 66      | 1.000          | 45      | 71      | 0.481 |
| No                 | 9 (0.08)  | 4    | 5       |                | 2       | 7       |        |
| Family history     |         |           |         |                |         |
| No                 | 118 (0.944) | 51  | 67      | 1.000          | 45      | 73      | 0.710 |
| Yes                | 7 (0.056)  | 3    | 4       |                | 2       | 5       |        |
| Cancer site        |         |           |         |                |         |
| Rectum             | 88 (0.7)  | 31   | 57      | 0.006          | 32      | 56      | 0.689 |
| Colon              | 37 (0.3)  | 23   | 14      |                | 15      | 22      |        |
| Histologic class   |         |           |         |                |         |
| AC                 | 120 (0.96) | 50  | 70      | 0.165          | 45      | 75      | 1.000 |
| NAC                | 5 (0.04)   | 4    | 1       |                | 2       | 3       |        |
| Differentiation    |         |           |         |                |         |
| Well/moderate      | 4 (0.03)   | 4    | 0       | 0.033          | 1       | 3       | 0.663 |
| Poor               | 121 (0.97) | 50   | 71      | 0.149          | 46      | 75      | 0.272 |
| Tumor stage        |         |           |         |                |         |
| II                 | 56 (0.452) | 20   | 36      |                | 18      | 38      | 0.165 |
| III                | 68 (0.548) | 34   | 35      | 0.289          | 38      | 71      |        |
| Tumor size (cm)    |         |           |         |                |         |
| <5                 | 116 (0.92) | 45   | 64      | 0.289          | 38      | 71      | 0.165 |
| ≥5                 | 9 (0.08)   | 9    | 7       |                | 9       | 7       |        |
| Fib (g/mL)         |         |           |         |                |         |
| <3.30              | 47 (0.38)  | 19   | 29      | 0.580          | 11      | 37      | 0.008 |
| ≥3.30              | 76 (0.62)  | 35   | 42      |                | 36      | 41      |        |
| RBC (<10^{12}/mL)  |         |           |         |                |         |
| <4.43              | 77 (0.62)  | 38   | 39      | 0.095          | 28      | 49      | 0.850 |
| ≥4.43              | 48 (0.38)  | 16   | 32      |                | 19      | 29      |        |
| PLR                |         |           |         |                |         |
| <113.5             | 48 (0.38)  | 4    | 44      | <0.001         | 18      | 30      | 1.000 |
| ≥113.5             | 77 (0.62)  | 50   | 27      |                | 29      | 48      |        |
| NLR                |         |           |         |                |         |
| <2.43              | 71 (0.57)  | —    | —       |                | 24      | 47      | 0.354 |
| ≥2.43              | 54 (0.43)  | —    | —       |                | 23      | 31      |        |
| CEA (ng/mL)        |         |           |         |                |         |
| <5                 | 78 (0.72)  | 31   | 47      | 0.206          | —       | —       |        |
| ≥5                 | 47 (0.38)  | 23   | 24      |                | —       | —       |        |
| CA199 (U/mL)       |         |           |         |                |         |
| <7.42              | 50 (0.40)  | 18   | 32      | 0.202          | 13      | 37      | 0.038 |
| ≥7.42              | 75 (0.60)  | 36   | 39      |                | 34      | 41      |        |

Fibr = fibrinogen; RBC = red blood cell; PLR = platelet-to-lymphocyte ratio; NLR = neutrophil-to-lymphocyte ratio; CEA = carcinoembryonic antigen; CA199 = carbohydrate antigen 199; COGN = combination of CEA and NLR; AC = adenocarcinoma; NAC = nonadenocarcinoma.
Table 3: Univariate Cox regression survival analysis for all CRC patients undergoing surgery (n = 125).

| Variables                        | OS         |              |              | DFS         |              |              |
|----------------------------------|------------|--------------|--------------|-------------|--------------|--------------|
|                                  | HR         | 95% CI       | P value      | HR          | 95% CI       | P value      |
| Age (≥65 versus <65)             | 1.469      | 0.596–3.622  | 0.404        | 1.405       | 0.660–2.991  | 0.378        |
| Gender (male versus female)      | 1.952      | 0.703–5.424  | 0.191        | 1.692       | 0.740–3.865  | 0.212        |
| Smoking                          | 2.745      | 0.798–9.451  | 0.109        | 1.953       | 0.587–6.493  | 0.275        |
| Family history                   | 1.618      | 0.372–7.040  | 0.521        | 1.755       | 0.527–5.844  | 0.360        |
| Cancer site                      | 1.264      | 0.477–3.345  | 0.637        | 1.370       | 0.612–3.064  | 0.444        |
| Histologic class                 | 1.492      | 0.199–11.204 | 0.698        | 1.059       | 0.143–7.813  | 0.956        |
| Differentiation                  | 21.018     | 0–3707867689 | 0.621        | 0.697       | 0.094–5.146  | 0.723        |
| Tumor stage                      | 4.491      | 1.308–15.418 | 0.017        | 3.049       | 1.231–7.557  | 0.016        |
| Tumor size (≥5 cm versus <5 cm)  | 1.923      | 0.634–5.827  | 0.248        | 2.069       | 0.833–5.141  | 0.117        |
| Fib (≥3.30 g/mL versus <3.30 g/mL) | 1.092    | 0.429–2.777  | 0.853        | 1.263       | 0.567–2.812  | 0.568        |
| RBC (≥4.43 versus <4.43 ×10^12/mL) | 1.460 | 0.593–3.596  | 0.410        | 1.538       | 0.723–3.275  | 0.264        |
| PLR (≥113.5 versus <113.5)       | 2.656      | 0.881–8.010  | 0.083        | 2.048       | 0.865–4.846  | 0.103        |
| NLR (≥2.43 versus <2.43)         | 3.193      | 1.213–8.405  | 0.019        | 2.582       | 1.182–5.644  | 0.017        |
| CEA (≥5 ng/mL versus <5 ng/mL)   | 4.351      | 1.645–11.509 | 0.003        | 4.023       | 1.803–8.974  | 0.001        |
| CA199 (≥0.4 ng/mL versus <0.4 ng/mL) | 4.491   | 0.593–3.596  | 0.410        | 1.538       | 0.723–3.275  | 0.264        |

AC = adenocarcinoma; NAC = nonadenocarcinoma; Fib = fibrinogen; RBC = red blood cell; PLR = platelet-to-lymphocyte ratio; NLR = neutrophil-to-lymphocyte ratio; CEA = carcinoembryonic antigen; CA199 = carbohydrate antigen 199; COCN = combination of CEA and NLR; HR = hazard ratio; OS = overall survival; DFS = disease-free survival.

In the present study, we investigated the correlations among SIR, clinicopathological characteristics, and survival in patients with primary CRC undergoing surgical resection. Our results demonstrated that high NLR was correlated with cancer site, tumor differentiation, and PLR, while high CEA was correlated with Fib and CA199 level. In addition, CEA and NLR were both independent prognostic factors.
Table 4: Multivariate Cox regression survival analysis for all patients undergoing surgery (n = 125).

| Variables | OS          |              |              | DFS         |              |              |
|-----------|-------------|--------------|--------------|-------------|--------------|--------------|
|           | HR          | 95% CI       | P value      | HR          | 95% CI       | P value      |
| NLR (≥ 2.43 versus <2.43) | 2.571 | 0.968–6.832 | 0.058 | 2.229 | 1.012–4.911 | 0.047 |
| CEA (≥5 ng/mL versus <5 ng/mL) | 3.713 | 1.396–9.873 | 0.009 | 3.652 | 1.630–8.179 | 0.002 |
| COCN (0, 1, 2) | 3.106 | 1.576–6.123 | 0.001 | 3.139 | 1.800–5.472 | <0.001 |
| Tumor stage | 3.384 | 0.972–11.778 | 0.055 | 2.449 | 0.978–6.131 | 0.056 |

NLR = neutrophil-to-lymphocyte ratio; CEA = carcinoembryonic antigen; COCN = combination of CEA and NLR; HR = hazard ratio; OS = overall survival; DFS = disease-free survival.

Figure 2: Kaplan–Meier survival curves for overall survival (OS) and disease-free survival (DFS) in 125 patients undergoing primary colorectal cancer resection according to their NLR and CEA levels. (a) OS according to NLR. (b) DFS according to NLR. (c) OS according to CEA levels. (d) DFS according to CEA levels. (e) OS according to COCN. (f) DFS according to COCN. Patients with neither NLR ≥ 2.43 nor CEA ≥ 5 were assigned as COCN = 0, with NLR ≥ 2.43 or CEA ≥ 5 as COCN = 1 and with NLR ≥ 2.43 and CEA ≥ 5 as COCN = 2.

associated with DFS in patients with primary CRC. CEA was an independent positive prognostic marker for OS in CRC, while high NLR showed a significant trend with lower OS rates (P = 0.058). Moreover, the current study for the first time demonstrated that COCN was more effective candidate prognostic biomarker in patients undergoing surgical resection of CRC than NLR or CEA alone.

Based on the results of ROC analysis, we chose a cut-off value of 2.43 for NLR because it yielded the highest sensitivity and specificity. Cut-off value of our study was approximately
Figure 3: Nomograms that predict the probability of NLR in OS (a) and DFS (b).
equal to 2.5 as per Shibutani et al. [20] study and 2.4 as per Neofytou et al. [21] study in CRC patients, and a little lower than 2.57 was reported in Yao et al. [22] study of breast cancer. Therefore, a cut-off value of 2.43 for NLR was considered as an acceptable value in several types of cancer.

Our study demonstrated that elevated NLR was an independent prognostic factor for poor survival of CRC patients. This finding was consistent with previous reports regarding CRC. For instance, a study by Shin et al. [23] demonstrated that preoperative NLR could predict survival rate in resectable patients with stage T1-2N0 CRC. Kubo et al. [24] also demonstrated that the pre- and postoperative NLR were both considered to be good predictor for the long-time survival in CRC patients. These studies together with ours suggest that high NLR values have a prognostic significance in CRC patients.

Although the reason for association between high NLR and poor prognosis is very complex and remains to be elucidated, there are several potential mechanisms. Patients with elevated NLR showed a relative lymphopenia and a high-circulating neutrophil level [25]. Neutrophils are known to play a key role in all stages of tumor progression by (1) producing a number of ligands and secretion of matrix metalloproteinases (MMPs), thus inducing tumor cell proliferation and invasion [26]; (2) releasing proangiogenic chemokines and other cytokines to promote tumor vascular formation [27, 28]; and (3) interacting with T cell, thus affecting tumor cell proliferation, angiogenesis, and metastasis [29]. Taken together, neutrophils may act as a tumor-promoting factor in various stages of cancer [30]. In contrast, lymphocytes could inhibit the proliferation and metastasis of tumor cells by participating in cytotoxic cell death and inducing the secretion of cytokines against tumor formation [20]. Given the tumor-promoting role of neutrophils and the antitumor effect of lymphocytes, elevation of NLR may reflect the increased protumor activity of neutrophils or the reduced antitumor immune responses by lymphocytes, consequently resulting in poorer survival of patients with elevated NLR. This might explain as to why patients with elevated NLR had a significantly poorer prognosis in the the present study.

Our research demonstrated NLR and preoperative CEA as independent prognostic factors in predicting the survival rates, and our results were similar to the previous reports [31–33]. Toiyama et al. [31] suggested that elevated CEA was a predictor of poor OS in rectal cancer patients who were treated with preoperative radiotherapy and chemotherapy. Thirunavukarasu et al. [33] confirmed that serum CEA was an independent prognostic marker in CRC patients with a mean follow-up period of 27 months. Graham et al. [32] considered that testing the CEA level of CRC patients was the most cost-effective for predicting the postoperative recurrences. Because of the prognostic usefulness of CEA, the American Society of Clinical Oncology (ASCO) and the European Society for Medical Oncology (ESMO) recommended CEA level to be considered as a golden follow-up standard after CRC therapy [34, 35].

We did not find significant correlations of the PLR and clinical characteristics with DFS or OS. This is not consistent with the result reported by Kwon et al. [36], who found that high PLR was independently associated with poorer OS and DFS. There are several reasons associated with differences in the results. One reason might be due to the different cut-off values used in this study. Presently, there is no consensus on cut-off values of PLR and the optimal cut-off value for PLR was determined either by ROC analysis as was the case with our study or according to the median of PLR. Another possible reason was due to tumor specificity and underlying genetic and biological differences between distinct patient cohorts. Finally, analyzers from different manufacturers could also cause differences in the PLR levels [37]. All these reasons put together may lead to the differences in the results as observed in these studies. Therefore, it is difficult to make a fair comparison between these studies and further studies are required to validate these possible reasons.

A more recent study reported that tumor progression was significantly correlated with tumor characteristics and host SIR [38]. Therefore, identifying parameters that reflect both tumor characteristics and host SIR provides a better prognostic value. Meanwhile, high CEA levels were reported to be related to the invasion and metastasis of tumor cells [34] and elevated NLR was considered to be a main factor of host SIR to tumors [39]. Our results further showed that CEA and NLR were both independent prognostic markers in CRC patients. We therefore hypothesized that combined detection of CEA and NLR may have a more important prognostic value in CRC. As shown in Table 4, COCN was an independent prognostic biomarker. Our study result was similar to the study results of He et al. [40], who supported that NLR in combination with CEA may provide a useful prognostic value in metastatic CRC patients. Further to this, our study for the first time indicated useful prognostic value of COCN in the primary resected CRC patients (Table 4), but it was also considered to be a better predictor of postoperative prognosis than NLR or CEA alone.

Our study demonstrated that COCN was correlated with survival of patients with resected primary CRC and supported the categorization of CRC patients into groups as favorable or poor prognosis based on the combined detection of inflammation-based and tumor-related factors. Highlights of the present study included the use of a uniform approach for patient assessment. On the other hand, COCN is easy to measure because of its low cost and convenience. Additionally, compared with NLR and CEA alone, COCN not only reflects the inflammatory and immune status of the patient but also represents the tumor characteristics and therefore is considered as a good prognostic marker in primary CRC patients.

Our study has few limitations. Firstly, selection bias could not be completely excluded due to the single-institutional and retrospective nature of the study. Secondly, the number of patients was small and the follow-up duration was relatively short. Finally, standardization of all clinical assays was another important problem. Although NLR and CEA are easy to measure, their utility might be affected by several factors and, therefore, more standard criteria need to be considered. Despite these limitations, still our study suggests that COCN was proved to be a better independent prognostic biomarker of CRC than NLR or CEA alone. Future studies are needed...
to prospectively validate the prognostic usefulness of COCN in CRC patients and assess the predictive nature of COCN to guide CRC therapy as well.

In summary, our research showed COCN to be an independent prognostic biomarker for CRC. Owing to the convenience, low cost, and high prognostic value, COCN may serve as a good biomarker in optimizing patient selection for further treatment, predicting OS and DFS, and decreasing the morbidty in patients undergoing curative resection for primary CRC.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Authors’ Contributions

Xiaoli Zhan and Xiaobo Sun contributed to this paper equally.

Acknowledgments

The project was supported by the National Natural Science Foundation of China (no. 81501401).

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