The Potential of Curcumin-Capped Nanoparticle Synthesis in Cancer Therapy: A Green Synthesis Approach

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Abstract: Cancer nanotherapeutics is an important field of research which utilizes nanomaterials as an approach to cancer therapy. Nano-mediated therapeutic delivery systems overcome the adverse side effects of traditional cancer treatment methods. Nanoparticles (NPs) are considered excellent tumor-targeting vehicles due to their compact and variable size, large surface area, ability to load several genes and drugs, and mediation of increased therapeutic payload uptake. Despite the rapid development of nanotechnology, there is growing concern regarding the possible long-term side effects of NPs on the environment and human health. Green chemistry using plant materials, such as curcumin, is a sustainable alternative to conventional reduction methods and confers dual reducing and capping properties. Curcumin is a bioactive compound isolated from the rhizome of the Curcuma longa plant, which exhibits various medicinal properties. Curcumin-capped NPs exhibit increased solubility, bioavailability, therapeutic indices, and antitumor properties. This review highlights the potential and antitumor properties of economical, simple, and eco-friendly curcumin-synthesized and capped NPs for the localized delivery of therapeutic genes and drugs to the cancer tumor microenvironment with fewer adverse side effects.

Keywords: curcumin; green chemistry; cancer; nanoparticles; delivery systems

1. Introduction

Cancer remains a leading cause of death worldwide with a poor prognosis. It is one of the deadliest diseases in human history with an inferior prognosis [1]. Cancer transforms healthy cells through inherited and acquired genetic mutations conferring growth and survival advantages and ultimately generates malignant neoplasms that invade adjacent tissues and spread to distant organs [2]. In 2020, 19.3 million novel cases and 10.0 million deaths were reported as a result of cancer, according to the World Health Organization (WHO), making the disease the second most common cause of death worldwide. The WHO further predicted that the number of cancer patients will increase to 29.4 million annually by 2040 [3]. The high global incidence and mortality rate of this non-communicable disease costs the economy billions of dollars [4]. Cancer displays great complexity at the cellular, epigenetic, and genetic levels, resulting in a reduction in long-term efficiency and the development of multiple drug resistances, non-specificity, dosage limitations, and adverse side effects. Conventional cancer treatment is also limited by the tumors’ pathological characteristics and the abnormal architecture of blood vessels [5].

Nanotherapeutics can enhance therapeutic indices, targeting, biodistribution, oral bioavailability, and aqueous solubility, making them promising candidates to challenge conventional cancer treatments [6,7]. Nanoparticles (NPs) have many uses in nutraceuticals, pollution management, and gene or drug delivery systems [8]. Various chemical, physical, and biological methods have been used to synthesize NPs. Although physical and chemical means of NP synthesis yield well-defined NPs, hazardous and expensive reducing reagents are frequently utilized, restricting upscaling and possibly damaging the environment [9,10].
Biological or green synthesis of NPs has gained much attention, as it employs plant extracts to synthesize NPs in an ecologically sound, environmentally friendly, and non-toxic manner [11]. In addition, using biological resources for NP manufacturing does not require complicated processes. Globally, a host of botanical moieties have been successfully used to synthesize different NPs [12]. Many of these NPs contain biologically active compounds that display medicinal properties [13], with curcumin being one such compound.

Curcumin is the principal curcuminoid obtained from the rhizomes of *Curcuma longa*. This bioactive component has many biological and pharmacological properties, including antioxidant, hypotensive, anti-inflammatory, anticoagulant, antiulcer, antimicrobial, antivenom, anti-fibrotic, antimutagenic, antidiabetic, anticarcinogenic, and most importantly anticancer activities [14]. Although the health benefits of curcumin have been well established, the full potential of curcumin in biomedical applications has yet to be fully exploited due to its crystalline structure, low solubility in water, and bioavailability (~1%) [15]. Reducing the size of the curcumin improves its solubility and, consequently, its bioavailability, with current studies aiming to increase the bioavailability of curcumin [16] specifically. This has led to the discovery of hybrid NPs, which comprise organic or inorganic NPs enveloped by curcumin [17].

Multidrug-loaded nanocarriers or curcumin NPs are a potential strategy for fighting cancer. Incorporating co-delivery systems as a feasible treatment method has projected synergistic benefits and limited undesirable effects [18]. Curcumin-capped NPs can introduce a synergistic effect by reducing the amount of the main therapeutic component needed. This can improve the therapeutic activity while reducing toxicity. Compared with NPs only and free curcumin, curcumin-capped NPs have shown a high level of cytotoxicity in some malignant cells [13]. Curcumin-containing nanocomplexes’ strong anticancer characteristics and their positive in vitro and in vivo results have prompted their in vivo investigation [19]. This review discusses curcumin’s antitumor properties and the potential of curcumin as a reducing agent in the synthesis of NPs for cancer therapy. Although anticancer studies have been conducted using curcumin, its use as a reducing or capping agent for synthesizing organic and inorganic NPs (primarily metal NPs) to produce synergistic NPs has yet to be fully explored in cancer therapy and is the main focus of this review.

2. Cancer Epidemiology

Cancer is a significant cause of death worldwide, with a global incidence and mortality rate of 19.3 million and 9.6 million per year, respectively (Figure 1). Lung, colorectum, liver, stomach, breast, cervical, and bladder cancer account for more than half of the annual deaths (Figure 2) [20]. By 2017, the global cancer mortality rate had almost doubled that seen in 1990 [21]. Cancer is a multi-factorial disease induced by multistage carcinogenesis involving genetic, cellular, and epigenetic abnormalities that transform healthy cells into malignant ones [6]. The evolution of cells to their cancerous state is induced by mutational damage to cancer susceptibility genes such as the tumor suppressor, DNA repair, and proto-oncogenes [22].

Oncogene mutations occur in the alleles of genes responsible for cellular growth, development, and maintenance. Mutations in the Ras family mediate cellular communication pathways, growth, and death. The *HER2* oncogenes, which mediate the growth and spread of cancer, lead to breast, ovarian, and cervical cancer [23]. Tumor suppressor genes regulate cell growth by slowing cell division, repairing DNA irregularities, and inducing apoptosis or programmed cell death. More than 50% of cancers are caused by mutations within the *p53* tumor suppressor gene [24], while germline mutations induce breast, ovarian, prostate, and pancreatic cancer in the *BRCA1* or *BRCA2* genes [25]. DNA repair genes that correct genome errors act as tumor suppressor genes. These mutations may be inherited or acquired [26].
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**Figure 1.** Mortality and incidence rates of cancer cases at ages <70 years in 2020. Adapted from [20].

The first step in cancer development involves initiating or altering the cancer-susceptible genes. Once a cell is transformed, it becomes susceptible to the effects of promoters, which bind to receptors on the cell’s surface. This inhibits the apoptotic pathways, leading to uncontrolled cell proliferation [27]. As cancer progresses, the cancerous cells are transported throughout the body via the lymphatic system or bloodstream. Once these cells reach their destination, they proliferate and develop into new tumors through metastasis [28]. Figure 3 illustrates the progression of a cancer cell.

**Figure 2.** Global mortality numbers for different cancers in 2020. Adapted from [20].
where they produce tumors [8] and induce the formation of new blood vessels through the secretion of hormones. The sprouting of the new capillaries invades the surrounding tissues via angiogenesis to allow for a continuous blood supply to the tumor microenvironment [29].

The age-standardized incidence and death rates were 1.5 times higher in males than females, with individuals over 50 and residing in developing countries displaying a higher cancer incidence [21]. The increase in cancer morbidity and mortality is attributed to the population’s growth, distribution, and dominant age as well as the distribution of cancer risk factors often associated with socioeconomic development [5]. Several risk factors influence an individual’s susceptibility to cancer, including lifestyle, age, genetics, physical inactivity, weak immune systems, infections, environment, sex, dietary choices, carcinogens, and physical agents [30]. Oral cancers are induced by alcohol (7–19%), smoking or chewing tobacco (25%), and micronutrient deficiency (10–15%). In South Asian countries such as Sri Lanka, Vietnam, Indonesia, India, and Malaysia, the chewing of betel nut accounts for 50–70% of oral cancers [31]. Higher skin cancer and melanoma fatalities were observed in regions with high levels of UV exposure due to damage to the ozone layer [32].

3. The Promise of Cancer Nanotherapeutics

3.1. The Limitations of Conventional Cancer Therapy

Despite growing knowledge of cancer epigenetics, genetics, biology, and etiology, cancer treatment strategies remain suboptimal, primarily due to their off-target side effects, multiple drug resistances, and physiological barriers, limiting their optimal dosages and efficiency [5]. Furthermore, the pathological properties of tumors and their aberrant blood vessel architecture decrease traditional cancer therapy’s effectiveness [8]. Chemotherapy, the preferred choice, fails to distinguish tumor cells from non-metastatic, healthy cells and targets all rapidly dividing cells, including cells of the lymph, bone marrow, gastrointestinal tract, and hair follicles [33]. This often leads to nausea, fatigue, loss of appetite, hair loss, nephrotoxicity or kidney damage, neuropathy or peripheral neuropathy, anemia, neutropenia, and thrombocytopenia [34]. Surgical removal of tumors is often challenging, leading to further complications, such as metastasis and recurrence. The organ may also be
inoperable due to the size and distribution of the tumor [35]. Hence, the intervention by nanomedicine may be beneficial in overcoming many of these challenges.

3.2. Overcoming Cancer Therapy Limitations with Nanomedicine

Nanomedicine is a rapidly evolving field of biotechnology. The core concept of nanotechnology dates to 1959, when Feynman demonstrated the ability of NPs to aid in the detection and pharmaceutical treatment of various human ailments [36]. The scientific understanding of the principles governing the interaction of matter with biological systems at the nanoscopic scale has advanced dramatically during the last three decades [37]. Nanomedicine utilizes nanomaterials (1–100 nm) for various biomedical applications, including tissue engineering, therapy, imaging, and diagnostics [31]. The therapeutic effect of nanomaterials is strongly influenced by their surface properties (hydrophilic-to-hydrophobic ratio and charge), physical characteristics (shape and size), and functionalization [38]. Figure 4 provides a general illustration of the favorable physicochemical properties of NPs. Carbon nanostructures, inorganic NPs (e.g., gold, silver, and selenium), lipids (e.g., liposomes), dendrimers, mesoporous silica, magnetic NPs, and polymeric NPs are among the NPs used in nanomedicine [4], with inorganic metal NPs yet to be fully explored [39].

![Figure 4. Summary of the tunable physicochemical properties of nanoparticles.](image)

NPs protect the therapeutic agent from opsonization and premature phagocytosis. They are absorbed at an enhanced rate by epithelial diffusion [40]. They can alter the therapeutic compound’s distribution profile and pharmacokinetics within the tumor microenvironment (TME), promoting intracellular efflux in the cells [41]. Furthermore, NPs can act as passive or active targeting agents to deliver therapeutic agents to the TME and elevate intracellular anticancer activity. Passive targeting enhances the anticancer agents’ permeability and retention effects due to the NPs’ favorable size and shape [42]. Active targeting involves the conjugation of ligands (e.g., folate) on the NP surface that can bind to receptors overexpressed on the surface of cancer cells [43,44]. This is easily facilitated
by the flexible surface chemistry of the NP, which allows for the conjugation of various targeting ligands.

Multiple drug resistance (MDR) occurs when cancer cells build resistance to numerous chemotherapeutic agents, leading to drug inactivation and efflux from the malignant cell. MDR poses a severe hurdle to treatment [45]. The compact size of the NPs enables the therapeutic to be administered at safe doses, increasing their antitumor effect and overcoming tumor drug resistance [46]. This further improves the therapeutic indices and pharmacokinetics of the delivered biomolecule.

NPs are amenable to steric stabilization using polyethylene glycol (PEG), which affords further stability to the NP by reducing surface-surface interactions and aggregation [47]. The PEG coating prevents the opsonization of the nanocomplexes. It allows the nanocomplex to avoid phagocytosis and clearance by the reticuloendothelial system (RES) and mononuclear phagocyte system, which are effective defense systems in the body that remove foreign material from the blood [48–50]. This increases the circulation time of the nanocomplex and allows the therapeutic to accumulate within the TME [51]. Figure 5 provides a simple illustration of the above process.

![Prevention of nanoparticle opsonization using long-chained polymers such as polyethylene glycol.](image)

**Figure 5.** Prevention of nanoparticle opsonization using long-chained polymers such as polyethylene glycol.

4. Green Synthesis of NPs

Various chemical, physical, and biological methods have been used to synthesize NPs, all with some advantages and disadvantages (Table 1). Although physical and chemical means of NP synthesis yield well-defined NPs, hazardous and expensive reducing reagents are frequently utilized, restricting upscaling and possibly damaging the environment [9,10]. This has created a niche for developing novel methods of NP synthesis which produce NPs with desired shapes, high thermal stability, and which use few or no toxic compounds [52,53]. This has prompted the start of the green nanotechnology revolution, where eco-friendly bioactive agents are employed in NP synthesis.

**Table 1.** Summary of nanoparticle synthesis methods and their advantages and disadvantages.

| Synthesis  | Advantages                             | Disadvantages                          | Types                  |
|------------|----------------------------------------|----------------------------------------|------------------------|
| Physical   | - Simple procedure                     | - High energy consumption              | Radiation              |
|            | - Produced in large quantities         | - Expensive                            | Sonication             |
|            | - Controlled particle interspacing      | - Specialized equipment required        | Laser ablation         |
|            |                                        |                                        | Membrane filtration    |
|            |                                        |                                        | Ion exchange            |
Table 1. Cont.

| Synthesis         | Advantages                                      | Disadvantages                                      | Types                     |
|-------------------|------------------------------------------------|---------------------------------------------------|---------------------------|
| Chemical          | - Size, shape, and morphology controlled        | - Toxic chemicals                                  | Reduction                 |
|                   | - Simple procedure                              | - Long reaction time                                | Oxidative process         |
|                   | - Narrow size distribution                      | - Limited by external factors (pH and temperature) | Photochemical             |
|                   |                                                  |                                                   | Electrochemical destruction|
|                   |                                                  |                                                   | Condensation              |
|                   |                                                  |                                                   | Sol-gel method            |
| Biological        | - Eco-friendly                                  | - Degree of reducibility                           | Plants                    |
|                   | - Use of non-toxic chemicals                    | - Reducing extract needs to be elucidated          | Bacteria                  |
|                   | - Inexpensive                                   |                                                   | Fungi                     |
|                   | - Less energy required                          | - Limited knowledge of controlled shape and size   | Viruses                   |

Peralta-Videa et al. (2016) stated that although plant-based green synthesis is comparable to conventional physical and chemical methods, they do not provide sufficient information on the ion’s reduction mechanisms, yield, or the stability of the NPs [54]. The mass production of NPs requires highly reactive substances and energy-consuming procedures, which are not considered environmentally friendly. The growing need to overcome these problems has prompted researchers to experiment with more economical and environmentally friendly methods. The green synthesis approach to creating NPs has gained tremendous strides as an alternative to physical and chemical syntheses [55].

Green synthesis can be defined as the use of various biological or bioactive agents such as plant extracts, microorganisms, fungi, and even biowastes to effectively synthesize metallic NPs in an eco-friendly and bio-reductive manner (Figure 6) [56,57]. Green synthesis vaunts a mechanism of reducing NPs while utilizing relatively low energy and maintaining a cost-effective method [58,59]. This approach, which was initially brought about due to the urgent need for “sustainable development”, requires three fundamental aspects for its success: a non-toxic reducing agent, a solvent of an environmentally friendly nature (for example, ethanol, water, and their combinations), and finally a stabilizer [60,61]. Furthermore, green synthesis offers a biocompatible, environmentally friendly, and non-toxic approach, as the byproducts and capping agents are natural to the environment, which is highly beneficial in the medical sector [56]. These NPs were reported to have a larger surface area with reduced aggregation than those produced with toxic chemical-reducing agents such as formaldehyde, hydrazine, sodium borohydride, aniline, polyvinylpyrrolidone, and sodium dodecyl sulfate [62–64]. In addition, these NPs showed improved photocatalytic and antioxidant capacities [54].

From the different biological agents used for green NP synthesis, microorganisms have shown some promising results. Since many inorganic metal salt ions can be toxic to microorganisms, they manufacture extra or intracellular enzymes that convert hazardous ions into harmless NPs [65]. A change in the redox state within the cell caused by foreign ions enables microbial reduced NPs to be applied in bioremediation [66]. Microorganisms can produce NPs within their cell walls. For example, the Klebsiella aerogenes bacterium was used to synthesize cadmium NPs [67], and Escherichia coli and Deinococcus radiodurans synthesized gold NPs [68,69]. Fungi are also attractive for large-scale production since they have a high tolerance for toxic elements and produce vast amounts of extracellular enzymes, as evidenced by silver and gold NPs extracted from Aspergillus oryzae and Verticillium [70,71]. However, compared with microbe-based synthesis, green synthesis of metallic NPs using plant-based extracts is a relatively more straightforward process which eliminates the constant maintenance of cultures [72].
Nanomaterials derived from plant-based syntheses have been employed to eradicate biofilms from clinically relevant surfaces, for targeted drug delivery, as 3D culture models, and in cancer therapy. In addition, the raw resources used in this green synthesis are renewable [73]. Plant extracts have been noted to produce NPs of varied sizes and shapes that range from rod-like to spherical, cubic, and triangular [74]. This is due to the phytochemical make-up of the plant utilized, which promotes natural stability in NP creation while doubling as a reducing agent [75]. Plant-based green synthesis is an extensive process involving the use of the whole plant biomass or extracts from different parts of the plant (stems, leaves, flowers, roots, seeds, bark, etc.) to synthesize different types of NPs [54]. The use of whole plants to synthesize metallic NPs is an intrinsic process with NPs deposited within the plant tissues, while plant extract-mediated NP synthesis occurs extracellularly. The composition of the plants affects their bioreductive ability, which in turn determines the morphology, composition, and dimensions of the nanoparticles formed [56]. Complementary to the effects of the phytochemicals present, biomolecules (carbohydrates, co-enzymes, and proteins) found in the plant extract also portray an exemplary reduction of metal salts into the desired metallic NPs. Functional compounds obtained from plant extracts, such as carboxylic acids, alkene, alkane, amine, and methylene, serve as promising reducing agents for synthesizing metallic NPs [72]. The biological activity of the synthesized NPs is primarily determined and fine-tuned by the biomaterial utilized for the stability and reduction of the metal ions [76]. The secondary metabolites of plants, such as tannins, flavones, and polyphenols, possess antioxidant, antimicrobial, and anticarcinogenic properties [77].

Apiin, isolated from Lawsonia inermis, reduced silver and gold metal salts via electrostatic interaction between the extract’s carbonyl groups and metal ions [78]. Phyllanthin extracted from Phyllanthus amarus was used to synthesize silver and gold NPs by exchanging the metal ions and the plant extract’s methoxide group [79]. Researchers discovered that compounds isolated from propolis extract could produce a broader range of NP sizes. These findings suggest that many chemicals with various reducing properties can complicate the synthesis process, impacting its simplicity and NP size distribution [80]. Spherical copper, gold, silver, selenium, and platinum of <100 nm in size were produced by extracts from either leaves, flowers, stems, fruit, or seeds from various plants [81–89]. However,
triangular and hexagonal shapes were also reported for green synthesized gold, silver, and silver-selenium bimetallic NPs [86,88,90], while titanium dioxide NPs produced unusual tetragonal shapes [91,92].

However, there are still barriers to be overcome in relation to more conventional methods for synthesizing NPs [9]. The ability to control the shape and size of biologically synthesized NPs has been a significant impediment, since the size and shape of the NP are predetermined by the different phytochemical compositions of the plant extract [81]. Similar plants grown in other geographical areas or harvested at different times can produce varied bioactive constituents. As a result, the shape and size of the NPs produced are altered [84]. This in turn can cause a decline in their market value, as the morphology of commercially manufactured NPs is uniform. Moreover, plant extracts contain many active phytochemicals which require isolation and purification [73]. However, the benefits of green synthesis outweigh the disadvantages, providing a niche for scientists to improve their means of plant-mediated synthesis.

5. The Properties of Curcumin

*Curcuma longa*, also known as turmeric, is an ancient perennial plant native to India that belongs to the Zingiberaceae family. Curcuma has evolved via continuous crossbreeding and selection. Over 100 Curcuma species have been identified to date [93]. The perennial herb grows in subtropical and tropical regions worldwide and is widely cultivated in Asian countries, such as India, Taiwan, Japan, Vietnam, Indonesia, Thailand, Burma, Bangladesh, and China [94]. The *C. longa* rhizome has an oblong pyriform shape with short branches. Turmeric is considered a significant plant in Ayurvedic history, having been used to treat a broad range of ailments in Indian Ayurvedic medicine since 1900 BC, including wounds, aches and pains, sprains, gastrointestinal system disorders, and liver disorders [95]. The biological constituent has been extensively studied for its bioactivity [96]. Curcumin exhibits antimicrobial, antioxidant, anticancer, anti-inflammatory, hyperlipidemic, hepatoprotective, and neuroprotective activities [95].

Turmeric can be broken down into three curcuminoids (Figure 7): bisdemethoxycurcumin, demethoxycurcumin, and diferuloylmethane [96]. The latter, a polyphenol, is a primary constituent of turmeric which accounts for turmeric’s vibrant yellow color and is commonly referred to as curcumin. The additional components of turmeric include proteins, sugars, resins, and volatile oils (zingiberone, atlantone, and turmerone) [94]. Curcumin (C_{21}H_{20}O_{6}) comprises two polyphenolic rings connected by a C7-linker containing an unsaturated β-diketone motif [97]. The bioactive component is insoluble in water under neutral and acidic pH levels. Still, it is soluble in acetone, dimethyl sulfoxide (DMSO), and ethanol, with a melting temperature and molecular weight of 183 °C and 368.37 g·mol^{-1}, respectively [95].

Curcumin contains two tautomeric forms: keto and enol, with the former being energetically stable in both the solid and liquid phases. However, the bioactive components take the form of a bis-keto (1,7-bis(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione) under acidic and neutral conditions [96]. It was reported that curcumin had enhanced antioxidant properties compared with its curcuminoid counterparts. This was attributed to the transition metal chelation attaching the o-methoxy and diketone phenols [93]. Curcumin blocks the NFkB and hemeoxygenase-1 pathways, which are responsible for the structural moieties of the α, β-unsaturated diketone that serves as an acceptor in the Michael reaction. Various studies have elaborated on curcumin’s biological and pharmaceutical properties [94,98–100].
The FtsZ protein is responsible for an essential stage of cell division [94,101]. Reports conclude that curcumin’s hydroxyl and methoxy groups are directly linked to antimicrobial activity. The oxygen molecules of these functional groups linked to curcumin’s phenolic rings catalyze the FtsZ GTPase protein, inducing premature cell death. Huang et al. demonstrated the antibacterial properties of curcumin-capped silver micelles [103]. The antiviral effects of curcumin micelles on hepatitis C virus (HCV) attachment were also demonstrated, where HCV cells treated with the nanocomplex had a lower viral load [104]. Similar results were observed in cells infected with the respiratory syncytial virus (RSV) when treated with curcumin-capped silver NPs [105].

Curcumin is a classical phenolic antioxidant which traps free radicals such as reactive nitrogen and oxygen species by enhancing the production of free radical scavenging enzymes [106–111]. Rajasekar (2015) demonstrated the antioxidant properties of curcumin nanocrystals in Wistar rats [108]. Nanocurcumin structures were observed to counteract the toxic side effects of aluminum phosphide by enhancing the stabilization of oxidative stress and scavenging free radicals [15].
Curcumin is a pleiotropic bioactive component which interacts with many inflammatory targets, including interleukins (IL) 1, 2, 5, 6, 8, and 12, macrophage inflammatory proteins (MIP), TNF-α (tumor necrosis factor-α), and the monocyte chemoattractant protein (MCP) [93,109,112,113]. The bioactive component hinders essential inflammatory response mediators, proinflammatory leukotrienes, and prostaglandin synthesis by downregulating the activity of lipoxygenase (LOX), cyclooxygenase-2 (COX-2), nitric oxide synthase (iNOS), and phospholipase A2 (PLA2) [101,110]. Thus, curcumin is effective in reducing post-surgical inflammation. The inflammatory response of curcumin can be attributed to its association with the arachidonic acid pathway for eicosanoid biosynthesis through the downregulation of COX-2 and LOX [95]. This produces many lipids, including prostaglandins, leukotrienes, prostacyclins, and thromboxanes. Nahar et al. (2015) demonstrated the ability of curcumin-capped lipid NPs to hinder the activation of NF-κB in murine macrophages through the downregulation of lipopolysaccharide-induced proinflammatory mediators, including IL-6, NO, and PGE2 [104,114]. Similarly, the obstruction of NF-κB activation by curcumin-loaded PLGA NPs downregulated iNOS and COX-2 expression [103,115].

5.2. The Anticancer Properties of Curcumin

The anticancer properties of curcumin in humans have gained significant interest in the past few decades. Kuttan et al. (1987) initially reported the anticancer properties of curcumin in clinical trials on patients with external cancer lesions. The study revealed that curcumin effectively relieved the symptoms of pain and itching while decreasing the lesion size [113]. Curcumin has anticancer activities that alter various cell growth cycle stages [114]. It acts as a blocking agent, hindering the initial stages of cancer by suppressing the proliferation of malignant cells during carcinogenesis. The bioactive component acts on transcription factors, oncogenes, and signaling proteins, which facilitate cancer cells’ growth and metastasis at different carcinogenesis stages [105] (Figure 8). Curcumin can also suppress matrix metalloproteinases’ activity, thus averting cancer metastasis. This results from curcumin suppressing the Bcl-xL, nuclear factor-kappa (NF-κB), BCL-2, cyclin D1, and c-MYC genes involved in tumor proliferation, growth, and apoptosis [115]. Furthermore, curcumin downregulates mitogen and epidermal growth factor receptor-activated protein kinases in lung and pancreatic cancer cells. It also displayed anti-amyloid activity, which reduced β-secretase and acetylcholinesterase activity as well as amyloid-β-protein aggregation and inflammation [116].

Curcumin is a potential anticancer agent against several cancers, including thyroid, prostate, lung, liver, myeloma, melanoma, colorectal, breast, and cervical cancer. Curcumin NPs showed anticancer properties in skin, lung, and liver cancer cells [117]. PLGA-curcumin NPs enhanced apoptosis and lysosomal activity and deregulated nuclear β-catenin and androgen receptor (AR) activity in prostate cancer cells [118]. An in vitro study reported the ability of curcumin to inhibit the metastasis of the papillary thyroid tumor cells by regulating the expression pattern of E-cadherins and metalloprotease-9 to enhance mesenchymal-epithelial transition [94]. An increase in HIF-1α, nuclear p65, and NF-κB expression was also noted, which altered the carcinogenic activity in breast cancer cells [119,120]. In vitro studies revealed enhanced the cytotoxicity and anti-invasive, anti-migratory, and apoptosis properties induced by curcumin-loaded NPs in metastatic pancreatic cancer [121]. The ability of curcumin nano-formulations to overcome the limitations of conventional treatment in colorectal cancer has been described in the literature [19].
6. Nanocurcumin Synthesis

Curcumin controls various signaling molecules based on the cell and target background, thus allowing it to act on multiple targets in cellular pathways [122]. However, the bioactive component has low bioavailability due to its limited solubility in water and its crystalline form [105]. To circumvent these constraints, researchers have attempted to increase curcumin’s biological and pharmacological potency by reducing its size [108]. This led to the discovery of nanocurcumin, which enhances the biological activity of curcumin, increasing its bioavailability, solubility, long-time circulation, and retention in the body [123].

The chemical and physical properties are essential in altering curcumin into its nanoform. The hydrophobicity, particle size, surface area, and charge are important physicochemical properties that make nanocurcumin a more effective anticancer agent than its native form [124]. The size of nanocurcumin being 1–100 nm is considered an ideal choice to use as a therapeutic agent because it has a larger surface area for better contact with the solvent. This enhances its solubility properties. Nanocurcumin structures can enter organs that are inaccessible to native curcumin [125]. Furthermore, nanocurcumin may have a higher intracellular absorption capacity, allowing the bioactive component to target foreign entities [126]. Zou et al. (2015) noted the high systemic bioavailability in the plasma and tissues of nanostructured curcumin compared with free curcumin [127]. Moreover, nanocurcumin increases the in vivo bioavailability and distribution, increasing the biological half-life 60-fold compared with treatment with native curcumin [93]. The loading and entrapment efficiency of nanodrugs depend highly on the preparation method and type of carrier system used to produce nanodrugs [128].

Nanocurcumin has been synthesized in various ways, including spray drying, emulsion polymerization, microemulsion, antisolvent precipitation, ultra-sonication, ionic gelation, single emulsion, solvent evaporation, wet milling, solid dispersion, thin film hydration, the Fessi method, and the coacervation technique (Figure 9). Each technique has its advantages and disadvantages, which have reviewed by many researchers [94,129,130].
its advantages and disadvantages, which have reviewed by many researchers [94,129,130]. The antisolvent precipitation and ionic gelation methods are regarded as the most efficient of the curcumin nano-formulation techniques [93].

These different techniques determine the shape and size of the curcumin NPs. Mukerjee and Vishwanatha (2009) reported the synthesis of 30–50-nm nanosphere curcumin structures using the polymerization emulsion technique [131]. Other studies reported the synthesis of crystal structured curcumin (150–200 nm) by the single emulsion–solvent evaporation method [132], small clusters (50 nm) using the inclusion complexation method [133], nanospheres (132 nm) using the redox-free radical polymerization technique [134], and nanocrystal curcumin NPs (30–40 nm) using the nanoprecipitation method [135]. Hence, it is evident that each technique can produce nanocurcumin of a defined size or shape. Based on these studies, researchers looking to synthesize curcumin NPs for biomedical applications would need to examine the size limitations of the target tissue or organ and the optimal shape of the NP for cellular uptake before opting for a particular synthesis method.

Nanocurcumin particles are not tissue-specific and act on healthy tissues surrounding the TME and cancer cells. Therefore, future studies must develop nano-delivery systems targeted at specific tissues [93]. Overall, the therapeutic effect of nanocurcumin remains at the concept level. Several questions and challenges prevail before nanocurcumin can be recommended as a promising candidate for therapeutic applications [136].

7. Curcumin-Capped NPs in Cancer Therapy

Curcumin can effectively reduce metal salts and cap the metal NPs. Curcumin-capped metal NPs have exhibited potent cytotoxicity in cancer cells [13]. Encapsulating therapeutic agents within NPs can enhance their pharmacokinetics and provide targeted delivery and controlled release [17]. Curcumin-capped NPs have a relatively larger surface area to interact with the solvent than naked curcumin. This property improves their aqueous solubility, leading to better bioavailability of the bound or encapsulated therapeutic. This enhances the therapeutic’s response to a specific molecular target and improves its pharmacological

Figure 9. A summary of the methods used in nanocurcumin synthesis.

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activity [137] by promoting controlled drug release [138]. Although nanocurcumin has a higher water solubility than curcumin-capped NPs, reducing and capping metal NPs with curcumin prevents their aggregation, making them highly stable in a solution [139–143]. This good stability and the ability of curcumin-capped NPs to be easily dissolved in an aqueous solution enhances their cellular internalization [8].

To date, solid lipid, polymeric, magnetic, and gold-based NPs have all been employed to enhance curcumin’s therapeutic application (Table 2).

Table 2. Curcumin-conjugated nanoparticles and their anticancer activities.

| Curcumin-Conjugate Nanoparticle (Source) | Morphology | Cancer Model | Anticancer Activity | Ref |
|----------------------------------------|------------|--------------|---------------------|-----|
| Solid lipid (Stearic acid and lecithin) | Spherical 20–80 | Breast cancer | Increased Bax/Bcl-2 ratios. Enhanced biocompatibility. | 140 |
| Micelles (d-hydroxyethyl starch) | Spherical 34.9 | Lung and colorectal cancer | Improved bioavailability, blood circulation, solubility, and stability of the nanocomplex. Enhanced antiproliferation and apoptosis. | 129 |
| Liposomes (L-α-phosphatidyl choline and cholesterol) | Irregular spherical 10–50 | Melanoma and Lung cancer | Greater encapsulation activity. Enhanced antiproliferation and apoptosis. | 137 |
| Polymeric (Poloxamer 188) | Spherical 248.4 | Colon and ovarian carcinoma | Enhanced uptake and cell specificity. Increased cytotoxicity. Improved blood circulation. | 143 |
| Silver nitrate | Spherical 15.5 | Breast cancer | Enhanced cellular uptake. Enhanced antiproliferation and apoptosis. | 141 |
| Gold (Chloroauric acid) | Spherical 26–28.2 | Breast, colon, prostate, and renal carcinoma | Improved blood circulation, solubility, and stability of the nanocomplex. Enhanced antiproliferation and apoptosis. | 140,142,144 |
| Albumin (Bovine serum albumin) | Spherical 112–198 | Breast cancer | Enhanced antiproliferation and apoptosis. | 145 |
| Graphene oxide and quantum dots (Graphite powder) | Crystal-like 15.62 | Breast cancer | Enhanced cellular uptake. Increased cytotoxicity. | 146 |

The antioxidant property of curcumin facilitates the reduction of metal salts by transferring electrons from curcumin to metal ions. The functional carbonyl and hydroxyl groups of the free curcumin further stabilize the NPs [126–129]. Solid lipid NPs are colloidal particles of natural or synthetic lipids. They are stable, easily scalable, and display enhanced biocompatibility, further improving solubility [140]. Solid lipid NPs conjugated with curcumin enhanced the therapeutic agent’s solubility, cellular uptake, dispersibility, and stability [111]. Wang et al. (2013) demonstrated that curcumin had increased inhibition (from 19.5% to 69.3%) and enhanced apoptosis in lung cancer cells both in vitro and in vivo [140].

The biocompatibility and compact size of polymeric NPs enable them to circulate within the blood system for an extended period. Chaurasia et al. (2016) observed the ability of a Eudragit R E100 cationic copolymer to enhance the uptake, binding, and cytotoxicity of curcumin-conjugated polymeric NPs in colon-26 cells [142].
The chemical properties of magnetic NPs, formed from a metallic oxide core, are easily manipulated regarding shape and size. Furthermore, these low-cost magnetic NPs display unique physical properties, promoting their biocompatibility in the human body [38]. PEGylated magnetic NPs conjugated with curcumin displayed enhanced biocompatible and antitumor responses [147]. The sustainable delivery of curcumin-loaded, thiolated, starch-capped iron-oxide NPs to lymphocyte cells, inducing cytotoxicity in several cancer cell lines, was demonstrated. Curcumin-capped Fe$_3$O$_4$-magnetic NPs illustrated enhanced uptake and targeted drug delivery to tumor cells [148].

Gold is a valuable metal, prized for its economic value and aesthetic appeal. Gold NPs serve as potential therapeutic gene delivery vehicles due to their favorable properties and ease of manufacturing [149]. Furthermore, due to their tunable stability, resilience, biodegradability, low cytotoxicity, biocompatibility, therapeutic gene protection from systemic degradation, and synthetic surface amenability, gold NPs have been used in various biomedical applications [46,150,151]. Targeted delivery of therapeutic agents and enhanced apoptosis in colon tumor cells was demonstrated using curcumin-reduced and capped chitosan-gold NPs [152]. Curcumin-capped gold NPs also effectively induced apoptosis in prostate and renal cancer cells [127,153]. Furthermore, green synthesized curcumin-capped gold NPs were found to improve antiproliferative and apoptotic activities in breast (MCF-7) and colon (HCT-116) cancer cells [126]. Gold NPs capped with curcumin and folic acid were observed to decrease tumor proliferation in breast cancer cells of Balb/c mice by 51% [154].

8. Clinical Trials Involving Curcumin

Numerous clinical trials have explained curcumin’s pharmacokinetic profile, safety, and effectiveness in different diseases. Clinical trials showed positive results where curcumin arrested or even eliminated cancer cell growth [6,137,139]. A search for clinical trials using curcumin or curcumin-reduced and capped NPs revealed that no curcumin-capped NPs are currently being screened. Most trials involved the delivery of curcumin alone or in combination with another therapeutic. Only one study utilized albumin NPs in combination with curcumin for pancreatic cancer therapy. The pilot phase I trial study demonstrated the ability of the nanotherapeutic agent to slow down cancer growth by halting tumor cell division. Table 3 provides a summary of the clinical trials involving curcumin [155]. Trials that have been withdrawn or terminated are not reflected.

Table 3. Selected completed or ongoing clinical trials from 2004 to date utilizing curcumin in cancer therapy (adapted from [155]).

| Cancer | Study Title | Therapeutic | Start and End Dates |
|--------|-------------|-------------|---------------------|
| **Breast** | “Window Trial” on Curcumin for Invasive Breast Cancer Primary Tumors | * Curcumin | January 2020–December 2022 |
| | Curcumin in Reducing Joint Pain in Breast Cancer Survivors with Aromatase inhibitor-induced Joint Disease # Curcumin for the Prevention of Radiation-induced Dermatitis in Breast Cancer Patients # Pilot Study of Curcumin for Women with Obesity and High Risk for Breast Cancer # Phase II Study of Curcumin vs Placebo for Chemotherapy-Treated Breast Cancer Patients Undergoing Radiotherapy # Prophylactic Topical Agents in Reducing Radiation-Induced Dermatitis in patients With Non-inflammatory Breast Cancer # Curcumin in Combination with Chemotherapy in Advanced Breast Cancer # Disposition of Dietary Polyphenols and Methylxanthines in Mammary Tissues from Breast Cancer Patients | ** Curcumin Nanoemulsion | March 2019–July 2022 |
| | Curcumin c3 | * Curcumin | January 2008–April 2011 |
| | | * Curcumin | June 2013–September 2016 |
| | | * Curcumin | May 2015–July 2018 |
| | | * Curcumin | October 2015–September 2016 |
| | | ** Curcumin, Paclitaxel | March 2017–June 2019 |
| | | ** Curcumin Polyphenol | June 2017–December 2019 |
| Cancer     | Study Title                                                                 | Therapeutic          | Start and End Dates                        |
|------------|----------------------------------------------------------------------------|----------------------|--------------------------------------------|
| Colon      | Study Investigating the Ability of Plant Exosomes to Deliver Curcumin to Normal and Colon Cancer Tissue | * Curcumin           | January 2011–December 2022                 |
|            | # Curcumin Biomarkers                                                       | * Curcumin c3        | November 2010–January 2013                 |
|            | # Combining Curcumin with FOLFOX Chemotherapy in Patients with inoperable Colorectal Cancer | ** Curcumin, Chemotherapy | February 2012–May 2017                     |
|            | # Effect of Curcumin on Dose Limiting Toxicity and Pharmacokinetics of Irinotecan in Patients with Solid Tumors | ** Curcumin, Irinotecan | June 2013–October 2016                     |
|            | # Avastin/FOLFIRI in Combination with Curcumin in Colorectal Cancer Patients with Unresectable Metastasis | ** Curcumin, Avastin/FOLFIRI | August 2015–2019                           |
|            | Curcumin in Advanced Cervical Cancer ** Curcumin, Pembrolizumab Radiation, Vitamin D Aspirin, Lansoprazole Cyclophosphamide | ** Curcumin, Piperine | July 2017–June 2021                        |
| Cervical   | # Trial on Safety and Pharmacokinetics of Intravaginal Curcumin             | * Curcumin           | January 2010–2012                          |
|            | # Study of Pembrolizumab, Radiation and Immune Modulatory Cocktail in Cervical/Uterine Cancer | ** Curcumin, Pembrolizumab Radiation, Vitamin D Aspirin, Lansoprazole Cyclophosphamide | July 2017–June 2021 |
| Prostate   | Adjuvant Curcumin to Assess Recurrence-Free Survival in Patients Who Have Had a Radical Prostatectomy | * Curcumin           | May 2014–June 2023                         |
|            | Trial of Curcumin to Prevent Progression of Low-risk Prostate Cancer Under Active Surveillance | * Curcumin           | March 2016–November 2026                  |
|            | Curcumin and Piperine in Patients on Surveillance for Monoclonal Gammopathy, Smoldering Myeloma or Prostate Cancer | ** Curcumin, Piperine | December 2021–May 2023                    |
|            | # Comparison of Duration of Treatment Interruption with or Without Curcumin During the off-Treatment Periods in Patients with Prostate Cancer Undergoing Intermittent Androgen Deprivation Therapy | * Curcumin           | August 2007–2015                           |
|            | # Radiosensitizing and Radioprotective Effects of Curcumin in Prostate Cancer | * Curcumin           | March 2011–October 2019                    |
|            | # Multicentre International Study for the Prevention with Ialuril® of Radio-induced Cystitis (MISTIC) | ** Curcumin, Radiotherapy | April 2017–May 2019                        |
|            | # Correlative Analysis of the Genomics of Vitamin D and Omega-3 Fatty Acid Intake in Prostate Cancer | ** Curcumin, Vitamin D, Omega-3 | September 2017–December 2019               |
| Lung       | Phase II Trial to Modulate Intermediate Endpoint Biomarkers in Former and Current Smokers | ** Curcumin, Lovaza | June 2019–October 2023                     |
|            | The Thoracic Peri-Operative Integrative Surgical Care Evaluation Trial-Stage II | ** Curcumin, Vitamin D3 Coriolus Versicolor Provitalix Green Tea Extract | April 2022–May 2025                        |
| Head and Neck | # Curcumin Biomarker Trial in Head and Neck Cancer                       | * Curcumin c3        | June 2010–January 2016                     |
|            | # Curcumin Bioavailability in Glioblastoma Patients                      | * Curcumin           | October 2012–May 2013                      |
|            | # The Effect of Curcumin on Treatment of Cancer Anorexia-Cachexia Syndrome in Patients with Stage III-IV of Head and Neck Cancer | * Curcumin           | February 2020–March 2021                  |
| Leukaemia  | Safety and Efficacy of Curcumin in Children with Acute Lymphoblastic Leukemia | * Curcumin           | August 2021–September 2022                |
| Oral       | # Oral Curcumin for Radiation Dermatitis                                   | * Curcumin           | February 2011–January 2015                |
Table 3. Cont.

| Cancer | Study Title | Therapeutic | Start and End Dates |
|--------|-------------|-------------|---------------------|
| Pancreatic | Gemcitabine Hydrochloride, Paclitaxel Albumin-Stabilized Nanoparticle Formulation, Metformin Hydrochloride, and a Standardized Dietary Supplement in Treating Patients with Pancreatic Cancer That Cannot Be Removed by Surgery | ** Curcumin, Gemcitabine, Metformin | January 2016–December 2022 |
| # Gemcitabine With Curcumin for Pancreatic Cancer | | # Completed trials. |
| # Trial of Curcumin in Advanced Pancreatic Cancer | # Completed trials. |

* Single therapy. ** Dual therapy. # Completed trials.

9. Conclusions and Future Perspectives

Green synthesized NPs have revolutionized nanotechnology. The NPs work synergistically with the conjugated plant extract, enhancing anticancer activity and biocompatibility. Using green nanotechnology to treat cancer provides the exciting prospect of inducing apoptosis while causing minimal damage to healthy cells surrounding the TME. Due to the toxicity challenges associated with chemical synthesis, the possibility of novel green synthesis methods emerging to overcome this problem is imminent. More research is needed to fully understand the various mechanisms involved in reducing organic and inorganic salts to produce NPs by the various green synthesis techniques. Green nanotechnology still has many unknowns to unravel and much to accomplish. Plants especially contain bioactive molecules that can act as reducing agents in NP synthesis. One such compound is curcumin.

Curcumin, due to its range of biomedical properties, including anticancer activity, has attracted the attention of researchers. Curcumin’s limited solubility in water, crystalline form, and low bioavailability have led to the formulation of nanocurcumin. Nanocurcumin enhances the cellular uptake, antitumor properties, tissue specificity, and pharmacokinetics of the conjugated therapeutic agent. The synthesis of inorganic and metal NPs by curcumin reduction especially has yet to be taken advantage of. This reduction process leads to the curcumin capping of the NPs, imparting synergistic activity with the therapeutic cargo. Optimizing synthesis protocols for the further application of curcumin capped-NPs is necessary to ensure these NPs are inexpensive, non-toxic, and can be formulated on a large scale for commercial use. Great potential lies in the curcumin reduction of metal salts to produce metal NPs such as gold and silver, which are commonly chemically synthesized, and improve their potential as delivery vehicles in cancer therapy. Comparative studies of chemical versus green synthesis need to be undertaken before definitive conclusions can be made. The crucial properties such as stability, toxicity, size, and shape must be optimized. The challenges of controlling the size and shape using curcumin or other green synthesis methods must be overcome.

Most of the research to date has been carried out in preclinical models at the proof-of-concept stage. Overall, there is still a significant dearth of knowledge about the full impact of curcumin and its capped NPs and their long-term risks in humans. Hence, before introducing curcumin-synthesized nano-formulations to the pharmaceutical sector, more in vitro and in vivo studies are required before clinical trials can be undertaken.

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