Impacts of detritivore diversity loss on instream decomposition are greatest in the tropics

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The relationship between detritivore diversity and decomposition can provide information on how biogeochemical cycles are affected by ongoing rates of extinction, but such evidence has come mostly from local studies and microcosm experiments. We conducted a globally distributed experiment (38 streams across 23 countries in 6 continents) using standardised methods to test the hypothesis that detritivore diversity enhances litter decomposition in streams, to establish the role of other characteristics of detritivore assemblages (abundance, biomass and body size), and to determine how patterns vary across realms, biomes and climates. We observed a positive relationship between diversity and decomposition, strongest in tropical areas, and a key role of abundance and biomass at higher latitudes. Our results suggest that litter decomposition might be altered by detritivore extinctions, particularly in tropical areas, where detritivore diversity is already relatively low and some environmental stressors particularly prevalent.
A key question in contemporary ecology is whether changes in biodiversity lead to alterations in the functioning of ecosystems and associated biogeochemical cycles. Interest in this topic emerged in the 1990s, motivated in part by the remarkable increase in global biodiversity loss, and led to hundreds of experiments that manipulated biodiversity at different levels (species, genes or functional traits) in different groups of terrestrial and aquatic organisms, to examine possible effects on ecosystem processes. While this large body of primary research and subsequent syntheses have demonstrated a strong, positive role of diversity of primary producers on biomass production, the patterns for decomposition have proven to be weaker and less consistent. This contrast may occur because decomposition can be simultaneously affected by the diversities of plant litter, microbial decomposers and animal consumers, with consequently more complex relationships.

Plant litter decomposition is a key process in the biosphere, as 90% of the annual plant production escapes herbivory and eventually becomes litter, which is ultimately decomposed or sequestered in terrestrial or aquatic ecosystems. Streams play a particularly relevant role in receiving and processing litter from their catchments, contributing significantly to global carbon and nutrient fluxes. Litter enters streams mainly in the form of leaves, and it is decomposed by microorganisms (mostly aquatic hyphomycetes) and specialised invertebrates (litter-consuming detritivores) that can obtain carbon and nutrients from the litter and associated fungi. Multiple studies have manipulated detritivore diversity and assessed its effect on decomposition locally in streams or in laboratory microcosms, with inconsistent results. These inconsistencies have been attributed to the existence of different species interactions driving either positive or negative effects, which can compensate for each other and sometimes result in overall neutral effects. However, there has been no global assessment of the relationship between detritivore diversity and decomposition in streams, which would help account for local and regional environmental contingencies in the diversity–decomposition relationship. A meta-analysis of terrestrial and aquatic studies revealed strong effects of detritivore diversity on decomposition, but there was no separate assessment of instream decomposition. Several stream studies have suggested a direct link between faster decomposition and greater detritivore diversity in temperate streams, but did not explore the relationship explicitly. A large-scale study demonstrated that decomposition in streams was enhanced when detritivore assemblages were more complex (large- and medium-sized organisms as opposed to medium-sized only), although it did not examine detritivore diversity.

Here, we describe results from a global-scale decomposition experiment conducted by partners of the GLOBE collaborative research network in 38 streams distributed across 23 countries in all inhabited continents. We use a standardized design and methodology to examine global-scale ecological questions, which reduces the number of confounding factors that need to be statistically controlled for in a meta-analysis. Our main working hypothesis is that detritivore diversity has a major positive effect on decomposition, although we also expect an influence of other detritivore assemblage characteristics such as abundance, biomass, and body size. Moreover, we predict that biotic drivers of decomposition vary across sites at different latitudes, possibly because of the varying interplay between positive and negative species interactions.

Multiple detritivore species were found in the majority of the streams, and the number of species was highest in the neotropical and afrotropical realms, temperate and boreal areas; and that abundance and diversity were highest in the Neotropical, Afrotropical and Indomalayan realms, tropical wet forests and savannas and xeric shrublands, and equatorial climates. Biome and body size were highest in the GLOBE-Net project, but the highest growing season occurred in the tropical and boreal areas, and the highest growing season was in the temperate and boreal areas.

**Table 1 Results of the best additive models explaining variation in total and detritivore-mediated litter decomposition based on detritivore diversity, abundance, biomass, mean body size, latitude, and interactions between detritivore variables and latitude.**

| Effect                       | edf | F   | p    |
|------------------------------|-----|-----|------|
| Total decomposition          |     |     |      |
| Diversity                    | 4.00| 6.94| <0.001|
| Abundance                    | 3.14| 6.34| <0.001|
| Biomass                      | 1.00| 2.00| 0.159 |
| Mean body size               | 1.86| 2.10| 0.102 |
| Latitude                     | 1.00| 3.01| 0.085 |
| Diversity × latitude         | 14.56| 6.17| <0.001|
| Abundance × latitude         | 1.00| 8.67| 0.004 |
| Biomass × latitude           | 7.91| 4.20| <0.001|
| Detritivore-mediated decomposi |     |     |      |
| Diversity                    | 4.00| 0.53| 0.716 |
| Abundance                    | 1.05| 0.01| 0.912 |
| Biomass                      | 1.00| 0.04| 0.843 |
| Mean body size               | 1.08| 1.00| 0.843 |
| Latitude                     | 1.71| 0.27| 0.763 |
| Diversity × latitude         | 14.14| 4.74| <0.001|
| Abundance × latitude         | 8.76| 3.30| <0.001|
| Biomass × latitude           | 7.99| 4.36| <0.001|

All predictors were fitted as tensor product interaction smooths. We show effective degrees of freedom (edf) and values of F and p for each factor. Models explained 69% and 78% of variation in the data, respectively.
Fig. 1 Generalised additive models exploring the influence of detritivore diversity, abundance and biomass on decomposition in different latitudinal zones (tropical: ≤23°; temperate: 24–60°; and boreal: >60°). Variation in total and detritivore-mediated decomposition (measured as the proportion of litter mass loss per degree day, dd; mean ± SE) with (a) detritivore diversity (number of families per litterbag), (b) log-transformed abundance (number of individuals per litterbag) and (c) log-transformed biomass (mg per litterbag), in different latitudinal zones. Lines represent the smoothers and shading the 95% confidence intervals from generalised additive models for significant relationships (p-value < 0.05); whole-model results are given in Supplementary Table 3.
Fig. 2 Global distribution of study sites in different biogeographic realms (Pa, Palearctic; Na, Nearctic; Au, Australasian; Nt, Neotropical; At, Afrotropical; Im, Indomalayan); \( n = 38 \). Box plots show the median, interquartile range and minimum-maximum range of litter-consuming detritivore diversity (number of families per litterbag), abundance (number of individuals per litterbag), biomass (mg per litterbag) and mean body size (mm) in each realm (ordered from highest to lowest diversity); different letters indicate significant differences. The NMDS ordination of litter-consuming detritivores with realms is represented by polygons of different colours as in maps and box plots. Significant differences in assemblage structure were: Pa vs. Na, At, Au, Im; Na vs. Nt, Au; Nt vs. Au.
Fig. 3 Global distribution of study sites in different biomes (Tu, tundra; TeBF, temperate broadleaf forest; TeCF, temperate coniferous forest; MeF, Mediterranean forest; XeS, xeric shrubland; TrWF, tropical wet forest; TrS, tropical savanna); n = 38. Box plots show the median, interquartile range and minimum-maximum range of litter-consuming detritivore diversity (number of families per litterbag), abundance (number of individuals per litterbag), biomass (mg per litterbag) and mean body size (mm) in each biome (ordered from highest to lowest diversity); different letters indicate significant differences. The NMDS ordination of litter-consuming detritivores with biomes is represented by polygons of different colours as in maps and box plots. Significant differences in assemblage structure were: TrWF vs. TeBF, TeCF, MeF.
equatorial climates. Assemblage composition mostly differed between the Palearctic/Nearctic (with many families of Laurasian origin) and other realms (families of Gondwanan distribution); between tropical wet forests and several other biomes; and between equatorial and other climates.

**Discussion**

Our study demonstrates a positive influence of detritivore diversity on decomposition, supporting previous suggestions that latitudinal gradients in detritivore diversity and instream decomposition are linked\(^24,25\) and agreeing with results of a
meta-analysis of controlled experiments performed in terrestrial and aquatic ecosystems. Our result also agrees with results of a meta-analysis of controlled experiments performed in terrestrial and aquatic ecosystems. These variables have previously been found to be important predictors of decomposition in some tropical streams, but here their importance was lower in the tropics than elsewhere. In temperate areas, both relationships were non-linear and complex (with decomposition first decreasing and then increasing with higher abundance or biomass), which impedes predictions about how decomposition might be altered by changes in these variables. Moreover, responses of abundance and biomass to environmental stressors are not as straightforward as diversity loss, because lost species can be replaced by more tolerant ones that thrive under stressful conditions and can cause an overall increase in numbers. However, this variation could be due to taxonomic differences rather than to size. Our results suggest that species replacements under environmental stress could result in an overall increase in biomass, but this possibility needs confirmation.

The distribution of most detritivore families corresponded to broad realms (Fig. 5), with 26 families showing a Laurasian distribution (i.e. being present in the Palearctic and/or Nearctic realms) and 14 families a Gondwanan distribution (Neotropical, Afrotropical, Australasian, and/or Indomalayan realms). Although we did not perform phylogenetic analyses, this dichotomy, together with the observation that diversity and abundance of detritivores were higher in the Palearctic and Nearctic (and their predominant biomes and climates), suggests that patterns of variation in diversity and abundance were at least partly determined by biogeography. Our findings contrast with those for angiosperms, current distributions of which do not correspond to tectonic history, possibly because of the existence of high transoceanic dispersal; however, they support patterns for organisms with lower dispersal, such as liverworts and conifers, which show clear Laurasian–Gondwanan disjunctions.

The strong influence of biogeography on detritivore diversity and abundance, and the fact that these two variables are key...
drivers of decomposition, suggest that the split of Pangea in the Late Jurassic (≈200 Ma ago) had a crucial legacy effect on the current functioning of stream ecosystems and the influence of ongoing environmental change. The lower detritivore diversity of tropical streams and the higher susceptibility of their fauna to extinction make these streams more vulnerable to reductions in decomposition rates that are associated with impaired ecosystem functioning. This observation, together with the over-exploitation of natural resources that severely affects tropical stream ecosystems, indicates that tropical detritivore species should be of high conservation concern globally.

Methods

Study sites. We conducted our study in 38 headwater streams located in different regions in 23 countries (Figs. 2–4). A random distribution of sites was unfeasible, so some regions were underrepresented (mostly Africa and northern Asia), which is usually the case for globally distributed experiments. Streams were similar in size (mean ± SE: wetted channel width, 3.9 ± 0.1 m; water depth, 28.7 ± 0.4 cm; 1st–3rd order) and physical habitat (alternating riffles and pools). Most had rocky substrate and were shaded by a dense riparian vegetation (64 ± 1%) representative of the region. They were located in 6 realms, 7 biomes, and 10 Köppen climate classes. In each stream we selected a ca. 100-m-long reach with 5 consecutive pool habitats in which to conduct the experiment. Further information on site physicochemical characteristics is given in Supplementary Table 4.

Field and laboratory work. At each site, we incubated 6 different 3-species litter mixtures, which included 9 species in total (Supplementary Table 5). The species and mixtures were chosen to represent different levels of functional diversity for a companion study, but here our interest was to use a variety of mixtures and thus increase the generality of our results (as opposed to working with a single or a few species). The 9 species were collected at different locations around the world and distributed among partners; we considered the possible home-field-advantage effect of using litter from different origins negligible based on available literature.

Litter mixtures were enclosed within paired coarse-mesh (5 mm) and fine-mesh (0.4 mm) litterbags containing the same amount and type of litter. The two types of litterbag respectively quantified total and microbial decomposition, and allowed the calculation of detritivore-mediated decomposition (see below). There were 60 litterbags per stream (n = 5 per litter mixture and mesh size), each containing 3 g of senescent litter (1 g per species), which had been collected freshly fallen from the forest floor, air-dried and distributed among research partners. Litterbags were deployed in each stream (one litterbag per litter mixture type and mesh size in a different stream pool, with all 5 pools consecutive) in 2017–2019 at the local time of the year with the greatest litter input and were retrieved after 23–46 d, depending on water temperature in each stream, thereby halting the decomposition process at a comparable stage (mean ± SD: 32 ± 17% litter mass loss on average for all the litter mixtures, 41 ± 18% for the fastest decomposing mixture; mean values for each biome are given in Supplementary Fig. 1). Litterbags were transported to the laboratory on ice enclosed individually in zip-lock bags and rinsed with filtered stream water to remove attached sediment and invertebrates. Litter was oven-dried (70 °C, 72 h) and a subsample weighed, incinerated (500 °C, 4 h) and re-weighed to calculate the final ash-free dry mass (AFDM). Invertebrates were sorted, and litter-consuming detritivores were counted and identified under a binocular microscope to the highest taxonomic level possible (mostly species or genus, and family in some cases), using available literature and local expert knowledge.

Calculation of variables. We quantified litter decomposition in each litterbag as the proportion of litter mass loss (LML) per degree day (dd), to account for differences in temperature across sites. LML = \( \frac{\text{final AFDM} - \text{initial AFDM}}{\text{initial AFDM}} \times \text{LML}_{\text{calc}} \), where initial AFDM was previously corrected by leaching, drying and ash content, which were estimated in the laboratory. We calculated detritivore-mediated decomposition as the difference in LML between paired coarse-mesh and fine-mesh litterbags. Total and detritivore-mediated decomposition were strongly correlated (\( r^2 = 0.90, p < 0.001 \)), but we used both as response variables in the analyses because the former is more relevant at the ecosystem level and the latter reflects patterns mediated solely by detritivores.

We quantified detritivore diversity in each coarse-mesh litterbag as taxon and family richness; as they were strongly correlated (\( r^2 = 0.90, p < 0.0001 \)), we used family richness for analyses to avoid taxonomic inconsistencies among sites. We quantified abundance as the number of individuals per litterbag. We estimated total biomass based on mean body size by using published equations for each family, and mean body size based on abundance and the mean of a body size category (2.5–5.0, 5.0–10.0, 10–20, 20–40 and 40–80 mm) that was assigned to each family using available literature.

Data analyses. We examined the influence of detritivore diversity, abundance, biomass, mean body size, latitude and the interactions between detritivore variables and latitude on decomposition, using generalised additive models (GAMs, gam function, ‘mgcv’ package v. 1.8.31) and a model selection (dredge function,
Each predictor, and no pattern was observed. Variation in assemblage composition model estimates. As interactions of detritivore variables with latitude were significant, we explored the relationships for tropical (≥23° of latitude), temperate (24°–60°) and boreal regions (≥60°) through a model that was similar to the one described above, but with latitude as a categorical rather than continuous predictor. This was done to facilitate the representation and interpretation of complex nonlinear relationships between two continuous predictors.

We explored differences in detritivore variables across realms, biomes and climates with linear mixed-effects models (lme function, ‘lme’ package v. 3.1.151) where realm, biome or climate were fixed factors and litter mixture type was a random factor, followed by pairwise comparisons using adjusted P-values (glht and mcp functions, ‘multcomp’ package v. 1.4.13)77. The variance was allowed to differ among realms and biomes using the VarIdent structure. Normalised residuals of the final model were inspected with plots of residuals vs. each predictor, and no pattern was observed. Variation in assembly composition was explored with non-metric multidimensional scaling (NMDS, monoMDS function, ‘vegan’ package v. 2.5-6)78 calculated on Hellinger transformed abundance data and permutational analysis of variance (PERMANOVA) based on a Bray–Curtis dissimilarity matrix. We compared realms, biomes and climates (adonis function, ‘vegan’ package), followed by pairwise comparisons (pairwise. adonis function), and determined which were the most representative families in each assemblage (simper function). All analyses were run on R v. 4.0.2.

Reporting summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability
Data supporting the findings of this study are available at https://doi.org/10.6084/m9.gshare.14245538.v1.

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References
1. van der Plas, F. Biodiversity and ecosystem functioning in naturally assembled communities. Biol. Rev. 94, 1220–1245 (2019).
2. Ibel, F. et al. Quantifying effects of biodiversity on ecosystem functioning across times and places. Ecol. Lett. 21, 763–778 (2018).
3. Naemy, S., Thompson, L. J., Lawler, S. P., Lawton, J. H. & Wood, L. Mechanisms behind positive diversity effects on ecosystem functioning: testing global hypotheses in ecology and environmental science. Front. Ecol. Environ. 11, 147–155 (2013).
4. Woodward, G. et al. Continental-scale effects of nutrient pollution on stream ecosystem functioning. Science 336, 1438–1440 (2012).
5. Jonsson, M. & Malmqvist, B. Ecosystem process rate increases with animal species richness: evidence from leaf-eating, aquatic insects. Oikos 89, 519–523 (2000).
6. Boyero, L. et al. Global patterns of stream detritivore distribution: implications for biodiversity loss in changing climates. Glob. Ecol. Biogeogr. 21, 134–141 (2012).
7. Boyero, L. et al. Global distribution of a key trophic guild contrasts with common latitudinal diversity patterns. Ecology 92, 1839–1848 (2011).
8. Handa, I. T. et al. Consequences of biodiversity loss for litter decomposition across biomes. Nature 509, 218–221 (2014).
9. Borre, E. T. et al. Finding generality in ecology: a model for globally distributed experiments. Methods Ecol. Evol. 5, 65–73 (2014).
10. Fraser, L. H. et al. Coordinated distributed experiments: an emerging tool for testing global hypotheses in ecology and environmental science. Front. Ecol. Environ. 11, 147–155 (2013).
11. Cornejo, A. et al. A common fungicide impairs stream ecosystem functioning through effects on aquatic hyphomycetes and detritivorous caddisflies. J. Ecol. Manag. 263, 110425 (2020).
12. Zuberb, J. P. et al. Long-term effects of fungicides on leaf-associated microorganisms and shredder populations—an artificial stream study. Environ. Toxicol. Chem. 36, 2178–2189 (2017).
13. Rasmussen, J. J. et al. Effects of a triazole fungicide and a pyrethroid insecticide on the decomposition of leaves in the presence or absence of macroinvertebrate shredders. Aquat. Toxicol. 118-119, 54–61 (2012).
14. Dillon, M. E., Wang, G. & Huey, R. B. Global metabolic impacts of recent climate warming. Nature 467, 704–706 (2010).
15. Dai, A. Drought under global warming: a review. Clim. Change 2, 45–65 (2011).
16. Tonin, A. M., Hepp, L. U., Restello, R. M. & Gønczálvus, J. E. Understanding of colonization and breakdown of leaves by invertebrates in a tropical stream is enhanced by using biomass as well as count data. Hydrobiologia 740, 79–88 (2014).
17. Pérez, J., Basaguren, A., Descals, E., Larrañaga, A. & Pozo, J. Leaf-litter processing in headwater streams of northern Iberian Peninsula: moderate levels of eutrophication do not explain breakdown rates. Hydrobiologia 718, 306–317 (2013).
18. Friberg, N. et al. Bio-monitoring of human impacts in freshwater ecosystems: the good, the bad and the ugly. Adv. Ecol. Res. 44, 211–278 (2011).
Pennington, R. T., Cronk, Q. C. B. & Richardson, J. A. Introduction and
synthetic plant phylogeny and the origin of major biomes. Philos. Trans. R.
Soc. Lond. B 359, 1455–1464 (2004).

Proches, S. Latitudinal and longitudinal barriers in global biogeography. Biol.
Let. 2, 69–72 (2006).

Vanderpoorten, A., Gradvert, S. R., Carine, M. A. & Devos, N. The ghosts of
Gondwana and Laurasia in modern liverwort distributions. Biol. Rev. 85,
477–497 (2010).

Young, R. G., Matthaei, C. D. & Townsend, C. R. Organic matter breakdown and
ecosystem metabolism: functional indicators for assessing river ecosystem
health. J. North Am. Benthol. Soc. 27, 605–625 (2008).

Gessner, M. O. & Chauvet, E. A case for using litter breakdown to assess
functional stream integrity. Ecol. Appl. 12, 498–510 (2002).

Ramírez A., Pringle C. M., Wantzen K. M. in Tropical Ecology (ed. Dudgeon, D.)
(Academic Press, 2008).

Tiegs, S. D., Akinwole, P. O. & Gessner, M. O. Litter decomposition across
continental scales: plant phylogeny and the origin of major biomes.
Behav. Ecol. Sociobiol. 49, 575–584 (2002).

Weya, J. M., Rumbiak, N. S., Hariyanto, S., Irawan, B. & Soegianto, A. Length-
mass regressions of freshwater gastropods in Nevada Spring ecosystems.
Malacologia 49, 133–143 (2003).

Benke, A. C., Huryn, A. D., Smock, L. A. & Wallace, J. B. Length-mass
relationships for freshwater macroinvertebrates in North America with
particular reference to the southeastern United States. J. North Am. Benthol.
Soc. 18, 308–343 (1999).

Miyasaka, H. et al. Relationships between length and weight of freshwater
macroinvertebrates in Japan. Limnology 9, 75–80 (2008).

Costa, L. C., Küfer, W. P. J., Casotti, C. G. & Moretti, M. S. Size-mass
relationships in Trichodactylus flavivialis (Decapoda: Brachyura:
Trichodactylidae), a macroconsumer in coastal streams of the Atlantic Forest,
southeastern Brazil. J. Crust. Biol. 38, 539–546 (2018).

Wood, S. N. Stable and efficient multiple smoothing parameter estimation for
generalized additive models. J. Am. Stat. Assoc. 99, 673–686 (2004).

Wood S. N. Generalized Additive Models: An Introduction with R 2nd edn
(Chapman and Hall/CRC, 2017).

Wagenmakers, E. J. & Farrell, S. AIC model selection using Akaike weights.
Psychon. Bull. Rev. 11, 192–196 (2004).

Zuur, A. F., Ieno, E. N., Walker, N., Saveliev, A. A. & Smith G. M. Mixed
Effects Models and Extensions in Ecology With R (Springer, 2009).

Ieno, E. N. & Zuur, A. F. Beginner’s Guide to Data Exploration and
Visualisation with R (2015).

Pinheiro, J. C., Bates, D. M., DebRoy, S., Sarkar, D. & Team R. C. nlme:
Linear and Nonlinear Mixed Effects Models. R Package Version 3.1-151.
https://CRAN.R-project.org/package=nlme (2020).

Hothorn, T., Bretz, F. & Westfall, P. Simultaneous inference in general
parametric models. Biom. J. 50, 346–363 (2008).

Oksanen, J. et al. vegan: Community Ecology Package. R Package Version 2.5-6.
https://CRAN.R-project.org/package=vegan (2019).

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Competing interests
The authors declare no competing interests.

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