Hypoxia equally reduces the respiratory compensation point and the NIRS-derived [HHb] breakpoint during a ramp-incremental test in young active males

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Abstract
This study investigated the effect of reduced inspired fraction of O$_2$ (FiO$_2$) in the correspondence between the respiratory compensation point (RCP) and the breakpoint in the near-infrared spectroscopy-derived deoxygenated hemoglobin signal ([HHb]$_{bp}$) during a ramp-incremental (RI) test to exhaustion. Eleven young males performed, on two separated occasions, a RI test either in normoxia (NORM, FiO$_2$ = 20.9%) or hypoxia (HYPO, FiO$_2$ = 16%). Oxygen uptake (\(\dot{V}O_2\)), and [HHb] signal from the vastus lateralis muscle were continuously measured. Peak \(\dot{V}O_2\) (2.98 ± 0.36 vs. 3.39 ± 0.26 L min$^{-1}$) and PO (282 ± 29 vs. 310 ± 19 W) were lower in HYPO compared to NORM condition, respectively. The \(\dot{V}O_2\) and PO associated with RCP and [HHb]$_{bp}$ were lower in HYPO (2.35 ± 0.24 and 2.34 ± 0.26 L min$^{-1}$; 198 ± 37 and 197 ± 30 W, respectively) when compared to NORM (2.75 ± 0.26 and 2.75 ± 0.28 L min$^{-1}$; 244 ± 29 and 241 ± 28 W, respectively) (\(p < .05\)). Within the same condition, the \(\dot{V}O_2\) and PO associated with RCP and [HHb]$_{bp}$ were not different (\(p > .05\)). Bland–Altman plots mean average errors between RCP and [HHb]$_{bp}$ were not different from zero in HYPO (0.01 L min$^{-1}$ and 1.1 W) and NORM (0.00 L min$^{-1}$ and 3.6 W) conditions. The intra-individual changes between thresholds associated with \(\dot{V}O_2\) and PO in HYPO from NORM were strongly correlated (\(r = .626\) and 0.752, \(p < .05\)). Therefore, breathing a lower FiO$_2$ during a RI test resulted in proportional reduction in the RCP and the [HHb]$_{bp}$ in terms of \(\dot{V}O_2\) and PO, which further supports the notion that these physiological responses may arise from similar metabolic changes reflecting a common phenomenon.

KEYWORDS
cycling, deoxygenation breakpoint, exercise intensity, RCP, thresholds
INTRODUCTION

During a ramp-incremental (RI) test to exhaustion, the rate of O2 uptake (V̇O2) increases rather linearly from the exercise onset until maximal V̇O2 (V̇O2max) or task failure ensue. At approximately 80% of V̇O2max, two physiological responses become discernible: at the pulmonary level the respiratory compensation point (RCP) (Whipp, Davis, & Wasserman, 1989) is expressed, whereas at the level of the muscle (i.e. the vastus lateralis muscle), a breakpoint in the near-infrared spectroscopy (NIRS)-derived deoxygenated hemoglobin signal ([HHb]bp) is observed (Spencer, Murias, & Paterson, 2012).

At the pulmonary level, the RCP is identified as the onset of a more rapid increase in minute ventilation that is disproportional from the rate of carbon dioxide production (VCO2), which causes the arterial tension of CO2 to fall (Whipp et al., 1989). This hyperventilatory response offers partial compensation to the blood accumulation of hydrogen ions ([H+]i) arising from the accelerated glycolytic rate within the active musculature (Whipp et al., 1989). At the muscle level (within the vastus lateralis of the quadriceps muscles), the [HHb]bp demarcates the beginning of a plateau-like response in the [HHb] signal (Spencer et al., 2012), with the adjustment of the [HHb] being a proxy for local O2 extraction, as it represents the balance between delivery and off-loading of O2 to and out of the capillary microvasculature, respectively. (Grassi & Quaresima, 2016). Although still a subject of debate, our group has suggested that the [HHb]bp during a RI test, and thus provide insights into the relationship between these two physiological indices of exercise intensity (Leo, Sabapathy, Simmonds, & Cross, 2017). Indeed, hypoxia has been shown to limit O2 provision to the peripheral tissues as indicated by a reduction in arterial O2 saturation and capillary partial pressure of O2 (Richardson et al., 2006). These impairments typically result in lower V̇O2max and peak power output (PPO) during incremental exercise (Knight et al., 1992), and a smaller PPO associated with the heavy-to-severe boundary of exercise intensity, as demonstrated by reduced CP in hypoxia (La Monica et al., 2018; Parker Simpson, Jones, Skiba, Vanhatalo, & Wilkerson, 2014; Townsend, Nichols, Skiba, Racinais, & Périard, 2017). Thus, it could be expected that breathing a lower fraction of O2 (FiO2) during a RI test would also affect the RCP and the [HHb]bp by reducing the V̇O2 and PO at which they occur with respect to normoxia. Indeed, previous findings have shown a reduction in the PO associated with the heavy-to-severe boundary of exercise intensity, as demonstrated by reduced CP in hypoxia (Azvedo et al., 2020; Osawa, Kime, Hamaoka, Katsumura, & Yamamoto, 2011). Although suggested by a previous study (Osawa et al., 2011), no investigation has quantified if the changes in the RCP and the [HHb]bp in hypoxia compared to normoxia would be of similar magnitude, and, thus, be linked to one another.

Therefore, this study explored the changes in the VO2 and PO associated with the RCP and the [HHb]bp in response to a diminished FiO2 (16%) during a RI test to exhaustion. It was hypothesized that a reduction in both the VO2 and PO at the RCP and the [HHb]bp would occur as a consequence of the decreased FiO2, and that these changes would be proportional on an individual basis.
2 METHODS

2.1 Participants

Eleven healthy males were recruited (age: 29 ± 6 years; height: 179 ± 6 cm; weight: 79 ± 9 kg). Participants gave informed written consent to participate in this study after completing the physical activity readiness questionnaire (PAR-Q+) and being cleared for exercise. All participants were free of any medical condition that could have altered their normal cardiovascular responses to exercise. The study was approved by the Conjoint Health Research Ethics Board at the University of Calgary. This study was part of a larger project designed to answer different research questions (Azevedo et al., 2020), whereby, out of 11 participants, 10 were included in our previous publication.

2.2 Experimental design

Participants reported to the laboratory on two separate occasions at similar time of the day (±1 hr). The visits were separated by at least 48 hr, during which participants performed a RI test to exhaustion either in normoxia (NORM, FiO2 = 20.9%) or hypoxia (HYPO, FiO2 = 16%). The FiO2 for hypoxia was chosen with reference of previous investigations demonstrating that this specific FiO2 was effective in altering arterial and muscle O2 saturation during exercise (Amann, Romer, Subudhi, Pegelow, & Dempsey, 2007). Before each RI test, a moderate intensity step transition, consisting of cycling at 20 W for 6 min and at 80 W for another 6 min, was performed to subsequently compute the mean response time (MRT) of \( \dot{V}O_2 \) (Iannetta, Murias, & Keir, 2019) (see data analyses for details). Thereafter, the participants were given 2 min of rest followed by a 4 min baseline cycling at 20 W prior to the RI test. The RI tests to volitional exhaustion consisted of 30 W min−1 continuous increments (i.e. 1 W every 2 s). Each exercise session was preceded by a 5 min wash-in period with the predetermined FiO2 (either hypoxia or normoxia), during which participants rested seated on the bike. All exercise testing was performed on an electromagnetically braked cycle ergometer (Velotron Dynafit Pro; Racer Mate).

2.3 Measurements

2.3.1 Pulmonary O2 uptake

A breath-by-breath metabolic cart system (Quark CPET, Cosmed), which was calibrated before each test as per the manufacturer’s recommendations, was used to measure ventilatory and gas exchange variables. The expiratory and inspiratory air volume rates and the concentrations of inspired and expired O2 and CO2 were measured through a low-resistance flowmeter and high precision gas analyzers, respectively. During all sessions, participants breathed through a mask connected to the flowmeter which was attached to a two-way low-resistance T-valve (Hans Rudolph In.; 2600 series, medium two-way NRBV). To control the FiO2 during HYPO, inspired air was delivered by a gas-mixing device (Altitrainer NP, SMTEC) connected with a tube to the T-valve to provide the desired concentrations of O2. During NORM, the same system was used to deliver room air in order to blind participants from the experimental condition.

2.3.2 NIRS-derived signals

\([\text{HHb}] \) (µM) and tissue O2 saturation index (\( \text{StO2} \) (%)) signals of the VL muscle were measured using a two-channel frequency-domain NIRS device (Oxiplex TS; ISS) at a sampling rate of 2 Hz, as described elsewhere (Iannetta, Qahtani, Millet, et al., 2017). The probe was placed on the belly of VL muscle, midway between the greater trochanter and the proximal border of the patella and secured in place by double-sided tape as well as an elastic strap to prevent movement. Additionally, the NIRS probe was covered with an optically dense, black vinyl sheet to minimize possible intrusion of extraneous light. Before removing the probe, the area was marked to ensure the consistency of the placement for the following visit. The NIRS system was calibrated before each test, as per the manufacturer’s recommendations.

2.3.3 Pulse O2 saturation

The pulse oxygen saturation (\( \text{SpO2} \)) was measured via a pulse oximetry pod (ADInstruments) from the forefinger of the right hand at a 1 Hz sampling rate and at wavelengths of 660 and 910 nm for red and infrared lights, respectively. The oximeter was connected to the acquisition apparatus (Power Lab, ADInstruments) linked to a computer software (LabChart 8, ADInstruments).

2.3.4 Blood lactate concentration

Within 1 min after the RI task failure, capillary blood lactate concentrations ([Lac−]) were measured with a portable lactate analyzer (Lactate Scout, SensLab GmbH). A lancet was used to perform a pinprick after wiping the finger with an alcohol swab, and a 2 µl capillary sample of whole blood was collected and immediately analyzed.
2.4  |  Data analyses

2.4.1  |  $\dot{V}O_2$

$\dot{V}O_2$ data were individually analyzed as previously described (Lamarr, Whipp, Ward, & Wasserman, 2017). Briefly, aberrant data points that were three standard deviations (SD) from the local mean were removed. Data were then linearly interpolated to $1 \text{s}$ intervals. $\dot{V}O_{2\text{max}}$ was defined as the highest $\dot{V}O_2$ computed from a $20-\text{s}$ rolling average. Gas exchange threshold (GET) and RCP were identified by three independent investigators by examining raw respiratory data. Briefly, GET corresponded to the breakpoint in the $\dot{V}O_2$-to-PCO$_2$ relationship (i.e. V-slope method) concomitant with an increase in the ventilatory equivalent of O$_2$ and a leveling off of end-tidal pressure of CO$_2$ (PCO$_2$) (Beaver, Wasserman, & Whipp, 1986). The RCP corresponded to the second disproportional increase (i.e. second breakpoint) in the $\dot{V}O_2/\dot{V}O_2$ relation, where end-tidal PCO$_2$ began to fall after a period of isocapnic buffering (Whipp et al., 1989). In case of disagreement of more than $100 \text{ ml min}^{-1}$, investigators would reevaluate together the profiles until consensus was reached.

To account for the circulatory transit time delay of deoxygenated hemoglobin from the active musculature to reach the lungs and the kinetics of $\dot{V}O_2$, the MRT was calculated on an individual basis, as previously described (Iannetta, Murias, et al., 2019), using a customized function of a computer software (Origin, Origin Lab). A linear regression of the $\dot{V}O_2$ versus PO relationship was fitted from the onset of the systemic rise in $\dot{V}O_2$ until the previously established GET. The steady-state $\dot{V}O_2$ from the moderate step transition (i.e. $80 \text{ W}$ for $6 \text{ min}$) was superimposed on the $\dot{V}O_2$ versus PO relationship. Thereafter, the MRT corresponded to the difference between (a) the PO equivalent to the abscissa of the intersection between the $\dot{V}O_2$ and the linear fit versus PO, and (b) the steady-state $\dot{V}O_2$ corresponding to $80 \text{ W}$ (i.e. measured during the step transition) (Iannetta, Murias, et al., 2019).

2.4.2  |  NIRS-derived signals

The [HHb]-time relationship for VL was modeled with the following piecewise equation that included two linear segments (e.g. “double linear”), as previously described (Spencer et al., 2012):

$$f = \begin{cases} g(x), & x < TD, \\ h(x), & x \geq TD, \end{cases}$$

where $f$ is the “double-linear” function, $x$ is time and $y$ is [HHb], $TD$ (i.e. time delay) is the time coordinate corresponding to the interception of the two regression lines (i.e. $[\text{HHb}]_{b_p}$), $l_1$ and $l_2$ are the intercepts of the first and second linear function, respectively and $s_1$ and $s_2$ are the slopes (i.e. SL1 and SL2, respectively). Model parameter estimates were determined by linear least-square regression analysis in which the best fit was defined by minimization of the residual sum of squares (RSS) and highest coefficient of determination ($R^2$). The “double-linear” fit was performed plotting the [HHb] data against PO with the fit starting at the onset of the systematic increase in the [HHb] signal until the last data point corresponding to the end of the RI test. Aberrant data that were $3 \pm SD$ from the local mean were removed. The piecewise equation was selected to fit this specific range of data where the equation parameters (i.e. $l_1$, $s_1$, $s_2$, and $TD$) were not previously fixed and were automatically selected by the mathematical software. Thereafter, the $\dot{V}O_2$ associated with the $[\text{HHb}]_{b_p}$ was identified after having “left-shifted” the $\dot{V}O_2$ data (Fontana et al., 2015). Finally, the end-exercise SpO$_2$ and StO$_2$ (i.e. last $10\%$ of the RI test) were utilized to compare the effectiveness of distinct FiO$_2$ in altering the O$_2$ saturation at the arterial and muscle level, respectively. Figure 1 shows a representative participant in whom the RCP and the $[\text{HHb}]_{b_p}$ were identified.

2.5  |  Statistical analyses

Descriptive data are presented as mean ± standard deviation (SD). Data normal distribution was tested by Shapiro–Wilk test and Q-Q plots. End-exercise PPO, $\dot{V}O_{2\text{max}}$, SpO$_2$, StO$_2$, and [Lac$^-$] were compared using paired samples $t$ tests. The $\dot{V}O_2$ and the PO associated with the RCP and the $[\text{HHb}]_{b_p}$ variables (i.e. absolute and relative values) and the $[\text{HHb}]$ NIRS-derived slopes before and after the $[\text{HHb}]_{b_p}$ (i.e. SL1 and SL2, respectively) were compared utilizing a two-way repeated measures ANOVA. When $F$ values were significant, Tukey post hoc were used to determine the loci of significant differences. The effect size was computed as partial eta squared ($\eta^2_p$) for ANOVA comparisons [i.e. assuming the small (<0.02), medium (0.02–0.26)]
NORM

VO₂ (L⋅min⁻¹)
0.00 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 50 100 150 200

Ve (L⋅min⁻¹)
0.00 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 50 100 150 200

PetCO₂ (mmHg)
0.0 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 10 20 30 40 50 60

[HHb] (µmol L⁻¹)
0.0 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 10 20 30 40 50 60 70

Time (s)
0 100 200 300 400 500 600 700

HYPO

VO₂ (L⋅min⁻¹)
0.00 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 50 100 150 200

Ve (L⋅min⁻¹)
0.00 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 50 100 150 200

PetCO₂ (mmHg)
0.0 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 10 20 30 40 50 60

[HHb] (µmol L⁻¹)
0.0 0.5 1.0 1.5 2.0 2.5 3.0 3.5 4.0 4.5
0 10 20 30 40 50 60 70

Time (s)
0 100 200 300 400 500 600 700

NORM

i₁ = 34.0
s₁ = 0.05
TD = 454
s₂ = -0.001
R² = 0.93

454 s
(227 W)

HYPO

i₁ = 33.8
s₁ = 0.04
TD = 382
s₂ = 0.01
R² = 0.95

382 s
(191 W)
and large (>0.26) effect sizes (Bakeman, 2005). For two group comparisons as Cohen D (d) [i.e. ranked as trivial (0–0.19), small (0.20–0.49), medium (0.50–0.79), and large (0.80 and greater) effect sizes] (Cumming, 2014). Bland–Altman plot analyses were used to compare the VO₂ and PO average errors and limits of agreement (LoA) between the RCP and [HHb]bp at each FiO₂ condition (Bland & Altman, 1986; Inglis et al., 2019). In addition, the association between values of VO₂ and PO at the RCP and the [HHb]bp were tested by linear regression and Pearson’s product moment correlation. Finally, the VO₂ and the PO associated with the RCP and the [HHb]bp in HYPO condition were compared based on the delta change from NORM condition with a Pearson’s product moment correlation. The significance level was set at p < .05. All statistical analyses were performed using a statistical software package (Statistica, version 10.0).

3 | RESULTS

3.1 | Ramp-incremental physiological responses and [HHb] signal adjustment in NORM and HYPO conditions

VO₂max was lower in HYPO (2.98 ± 0.36 L min⁻¹) compared to NORM (3.39 ± 0.26 L min⁻¹) (p < .001, d = −1.32). Similarly, PPO was lower in HYPO (282 ± 29 W) than in NORM (310 ± 19 W) (p = .002, d = −1.12). [Lac−] was not different between conditions (HYPO, 9.6 ± 2.6 mM L⁻¹; NORM: 10.0 ± 2.3 mM L⁻¹; p = .522, d = −0.16). SpO₂ and StO₂ in HYPO (81% ± 4% and 58% ± 11%, respectively) were lower than in NORM (90% ± 5% vs. 65% ± 10%, respectively) (p = .001 and 0.007, d = −1.85 and −0.63) at the last 10% of the RI test. The slope of the [HHb] NIRS signal before the [HHb]bp (i.e. SL1: HYPO, 0.039 ± 0.022 µM s⁻¹; NORM, 0.044 ± 0.024 µM s⁻¹) was greater than after the [HHb]bp (i.e. SL2: HYPO, 0.019 ± 0.022 µM s⁻¹; NORM, 0.019 ± 0.020 µM s⁻¹) (p = .003, η² = 0.583) but there was no difference between conditions (p = .562, η² = 0.034). The “double-linear” fitting parameters for [HHb]bp identification showed high values for goodness-of-fit within the data set, in accordance with a previous study (Spencer et al., 2012), as given by the RSS and R² for HYPO (RSS = 709 ± 269; R² = 0.92 ± 0.06) and NORM (RSS = 658 ± 247; R² = 0.93 ± 0.03) conditions.

3.2 | The VO₂ and PO associated with the RCP and the [HHb]bp variables between the different FiO₂ conditions

The RCP and the [HHb]bp associated with absolute and relative VO₂ and PO values are shown in Table 1. The absolute values for VO₂ associated with the RCP and the [HHb]bp were lower in HYPO compared to NORM condition (p < .001, η² = 0.785) but there was no difference between variables within the same condition (p = .964, η² < 0.001). The %VO₂max associated with the RCP and the [HHb]bp were neither different between conditions (p = .235, η² = 0.069) nor variables within the same condition (p = .924, η² < 0.001). The absolute values for the PO associated with the RCP and the [HHb]bp were lower in HYPO when compared to NORM (p < .001, η² = 0.826), but there was no difference between the RCP and the [HHb]bp within the same condition (p = .848, η² = 0.001). The %PPO associated with RCP and [HHb]bp in HYPO were lower compared to NORM (p < .001, η² = 0.575) but there was no difference between the RCP and the [HHb]bp within the same condition (p = .756, η² = 0.004). The MRT obtained for HYPO (37 ± 10 s) and NORM (39 ± 13 s) conditions were not different (p = .363, d = −.329).

3.3 | Correspondence between the RCP and the [HHb]bp variables within the same FiO₂ condition

The comparison between the VO₂ and the PO associated with the RCP and the [HHb]bp within the HYPO and NORM conditions are shown in the Bland–Altman plots (Figure 2). The VO₂ mean average error for HYPO (0.01 L min⁻¹, LoA, lower:

| TABLE 1 | Respiratory compensation point (RCP) and deoxyhemoglobin breakpoint ([HHb]bp) variables in each FiO₂ condition |
|---|---|
| **NORM** | **HYPO** |
| **RCP** | **[HHb]bp** | **RCP** | **[HHb]bp** |
| VO₂ (L·min⁻¹) | 2.75 ± 0.26* (2.24–3.26) | 2.75 ± 0.28* (2.19–3.31) | 2.35 ± 0.24 (1.89–2.81) | 2.34 ± 0.26 (1.82–2.86) |
| VO₂ (%) | 81 ± 5 | 81 ± 6 | 79 ± 6 | 78 ± 5 |
| PO (W) | 244 ± 29* (188–300) | 241 ± 28* (186–294) | 198 ± 37 (126–270) | 197 ± 30 (138–255) |
| PO (%) | 79 ± 6* | 78 ± 7* | 69 ± 7 | 69 ± 5 |

Note: Data are presented as mean ± SD and 95% confidence intervals in brackets. Abbreviations: %, percentage of peak power output and VO₂max; [HHb]bp, deoxygenated hemoglobin breakpoint; HYPO, hypoxia condition (FiO₂ = 16.0%); NORM, normoxia condition (FiO₂ = 20.9%); VO₂, oxygen uptake; PO, power output; RCP, respiratory compensation point. *Statistically different from HYPO (p < .05).
FIGURE 2 Bland–Altman plots comparison between the \( \dot{V}O_2 \) and the PO associated with the respiratory compensation point (RCP) and the deoxyhemoglobin breakpoint ([HHb]bp) for each FiO2 condition. Black lines indicate mean difference error between the oxygen uptake (\( \dot{V}O_2 \)) associated with RCP and [HHb]bp; Dashed black lines indicate lower and upper limits of agreement (±2 SD). (a) \( \dot{V}O_2 \) associated with RCP and [HHb]bp in normoxia condition (FiO2 = 20.9%); (b) Power output associated with RCP and [HHb]bp in normoxia condition; (c) \( \dot{V}O_2 \) associated with RCP and [HHb]bp in hypoxia condition (FiO2 = 16.0%); (d) PO associated with RCP and [HHb]bp in hypoxia condition.

\[ \dot{V}O_2 \] and PO associated with the RCP and [HHb]bp in HYPO condition were equally decreased in HYPO compared to NORM, as demonstrated by a significant correlation for each variable (Figure 3a and c). Moreover, the mean average error bias for \( \dot{V}O_2 \) (0.00 L min\(^{-1}\); LoA: lower, −0.39 L min\(^{-1}\); upper, 0.41 L min\(^{-1}\); \( p = .877 \)) (Figure 3b) and PO (−2 W, LoA, lower: −32 W; upper, 27 W; \( p = .572 \)) (Figure 3d) associated with the RCP and the [HHb]bp in HYPO compared to NORM condition was not statistically different from zero.

4 | DISCUSSION

This study investigated whether the strong relationship between the RCP and the [HHb]bp that is typically observed in normoxia during a RI test (Fontana et al., 2015; Iannetta, Qahtani, Maturana, & Murias, 2017; Iannetta, Qahtani, Millet, et al., 2017; Inglis et al., 2019; Keir et al., 2015; Murias, Keir, Spencer, & Paterson, 2013) was affected by breathing hypoxic air (FiO2 16%). The main and novel findings were that: (a) the RCP and [HHb]bp were equally reduced during hypoxia in terms of both \( \dot{V}O_2 \) and PO; (b) these changes were highly correlated, showing small and nonsignificant biases within each individual in each experimental condition (i.e. NORM and HYPO). Taken together, these results indicate that breathing a lower FiO2 during a RI test results in a proportional reduction in both the \( \dot{V}O_2 \) and PO associated with the RCP and the [HHb]bp. These data further support the notion that these indices of intensity reflect a similar physiological response that is linked to metabolic changes within the active muscles.

In line with previous studies (Knight et al., 1992; Osawa et al., 2011; Richardson et al., 2006), the detrimental effects of hypoxia in the present study are evidenced by the reduced SpO2 and StO2, and lower \( \dot{V}O_2 \)max and PPO throughout and at the end of RI test, respectively, compared to NORM. Most importantly, the earlier development of metabolic perturbations associated with exercise in hypoxia is evident in the attenuated \( \dot{V}O_2 \) and PO at which RCP and [HHb]bp occurred. Acute changes in FiO2 have detrimental effects on exercise performance and maximal aerobic capacity due to reduced \( O_2 \) delivery and diffusive driving pressure of \( O_2 \) at the muscular level (Knight et al., 1992; Osawa et al., 2011; Richardson et al., 2006). Thus, in order to maintain the adequate balance...
between O₂ delivery and O₂ utilization, several physiological adjustments need to occur (Calbet, Rådegran, Boushel, & Saltin, 2009). For example, for a given absolute submaximal exercise intensity, there is a greater cardiac output and increased local vasodilatory responses when compared to normoxia (Calbet et al., 2009). In fact, it has been suggested that the hypoxia-induced hyperemia response is proportional to the hypoxia-induced fall in arterial O₂ content (Casey & Joyner, 2011). However, as the exercise intensity increases, these adjustments cannot counteract the reduced driving pressure of O₂ at the capillary-to-muscle interface, limiting mitochondrial ATP resynthesis rate (Richardson, Leigh, Wagner, & Noyszewski, 1999). Thus, during a RI test to exhaustion in hypoxic conditions, there is an earlier reliance on anaerobic resources to sustain the ATP requirement when compared to normoxia (Connett, Honig, Gayeski, & Brooks, 1990; Linnarsson, Karlsson, Fagraeus, & Saltin, 1974). This is also due to increased circulation of catecholamines (i.e. epinephrine and noradrenaline) and an earlier recruitment of type II fibers which increase the glycolytic contribution to the ATP resynthesis in hypoxia compared to normoxia (Moritani, Sherman, Shibata, Matsumoto, & Shinohara, 1992; Osawa et al., 2011). This is evidenced by previous studies showing greater lactate efflux from the muscle to blood and greater depletion of intramuscular phosphagen resources at a given submaximal absolute work rate in hypoxia compared to normoxia (Hogan, Richardson, & Haseler, 1999; Linnarsson et al., 1974). Additionally, the present results and previous findings (Azevedo et al., 2020; Osawa et al., 2011) have consistently shown that the occurrence of [HHb]bp is associated with lower absolute PO but the rate of change on [HHb] signal (i.e. SL1 and SL2) is not different either before or after the [HHb]bp (Azevedo et al., 2020). Thus, based on those abovementioned mechanisms, it would be plausible to suggest that breathing hypoxic air promotes a “left-shift” in the physiological responses throughout a RI test when compared to normoxia.

In relation to the NORM condition, the present results are in line with previous studies which have shown a correspondence between the RCP and the [HHb]bp both in terms of VO₂ and PO in normoxia condition (Fontana et al., 2015; Iannetta, Qahtani, Millet, et al., 2017; Keir et al., 2015). Murias et al. (2013) were the first to demonstrate that the RCP and the [HHb]bp occurred at the same VO₂ throughout a RI test, which was later confirmed by several other studies in large and heterogeneous samples of individuals and in a test–retest conditions (Fontana et al., 2015; Iannetta, Qahtani, Millet, et al., 2017; Inglis et al., 2019; Keir et al., 2015). In direct connection with the goal of this investigation, the present data further confirm the correspondence between the RCP and [HHb]bp when exercising under distinct FiO₂ conditions. Based on the Bland–Altman plots analysis, the bias obtained for the VO₂ associated with the RCP and [HHb]bp was not different from zero neither in NORM (0.00 L min⁻¹) nor in HYPO conditions (0.01 L min⁻¹) (Figure 2). These biases are similar to those...
previously reported in other studies (Fontana et al., 2015; Iannetta, Qahtani, Maturana, et al., 2017). Similarly, the PO associated with the RCP and [HHb]bp displayed biases that were small in NORM (i.e. 3.6 W) and HYPO (i.e. 1.1 W), and similar to previously reported findings (Inglis et al., 2019). In addition to this, the V̇O₂ and the PO change in HYPO from NORM condition for RCP and [HHb]bp were significantly correlated (Figure 3a and c), showing a bias that was not different from zero (Figure 3b and d). This is an important finding, as it indicates that both, the V̇O₂ and the PO associated with the RCP and the [HHb]bp, were negatively affected by the same degree in hypoxia.

The fact that the RCP and [HHb]bp continued to occur at the same metabolic rate, independently of the FiO₂, provides further support to the idea that these events might be triggered by common physiological responses occurring at the systemic and muscular level (Fontana et al., 2015; Keir et al., 2015). In other words, both the RCP and the [HHb]bp are linked to a metabolic rate corresponding to the transition from the heavy to the severe exercise intensity domain (Keir et al., 2015), which elicits important metabolic changes as a consequence of metabolites accumulation within the muscle and bloodstream (Whipp et al., 1989). For example, up to a critical metabolic rate, increases in ventilation are sufficient to constraint the increases in arterial [H⁺] linked to CO₂ production (Wasserman, Beaver, Sun, & Stringer, 2011). However, the increase in rate of [H⁺] accumulation, when progressively increasing the intensities above the critical metabolic rate, exceeds the capacity of this buffering system, which results in a more rapid accumulation of [H⁺], triggering a reflex increase in ventilation (i.e. the RCP) as a result of its uncoupling in relation to VCO₂. Even though the exact mechanisms underpinning the hyperventilatory response throughout a RI test, and more specifically above the RCP, are still under debate (Nicolò, Marcora, & Sacchetti, 2020), there are several relevant systemic and local muscular changes that may play a direct effect (Wasserman et al., 2011; Whipp et al., 1989). Furthermore, the transition into the severe intensity domain is characterized not only by an increase in [H⁺], with a subsequent reduction in pH, but also by a decrease in the partial pressure of O₂ within the active muscles, which are known to favor local vasodilatation (Casey & Joyner, 2011) and, consequently, redistribution of blood flow to the active tissues. Accordingly, it has been suggested that increased blood flow distribution to the vastus lateralis muscle might be the main mechanism responsible for the observed plateau in the [Hb] signal during ramp-incremental exercise (Azevedo et al., 2020; Iannetta, Okushima, et al., 2018; Inglis et al., 2017; Muriás, Spencer, et al., 2013), as the improved local blood flow would reduce the need to further increase O₂ extraction to support the continued increase in muscle V̇O₂. This is in line with findings from animal models, whereby the greater activation of type II fibers when surpassing the heavy-to-severe boundary of exercise intensity triggers disproportionate increases in blood flow in these fibers (Copp, Hirai, Musch, & Poole, 2010). However, although in humans the superficial portions of the vastus lateralis (which are the target of the NIRS light) are characterized by a greater proportion of type II muscle fibers (Johnson, Polgar, Weightman, & Appleton, 1973), this interpretation must be taken with caution because, since the human skeletal muscle is characterized by a mosaic of different fiber types, which may present disparate vascular dynamics and not necessary demonstrate such abrupt changes. In this regard, the leveling off of the [HHb] signal in the vastus lateralis muscle has been demonstrated not to be depth dependent (Iannetta, Okushima, et al., 2018) or affected by differential muscle recruitment patterns (Okushima et al., 2020) and exercise modes (Iannetta, Passfield, Qahtani, MacInnis, & Murias, 2019), which would suggest that this phenomenon may not be ascribed to a unique fiber type population.

Alternatively, it has been proposed that even though the RCP and the [HHb]bp might share somewhat similar underpinning mechanisms, they manifest an order of occurrence (Boone, Barstow, et al., 2016; Boone, Vandekerckhove, Coomans, Prieur, & Bourgois, 2016). Accordingly, as intensity increases there is an increasing recruitment of type II fibers leading to the development of metabolic acidosis that triggers the hyperventilatory response (i.e. RCP) and, subsequently, the occurrence of the [HHb]bp as a consequence of the attainment of a supposedly maximal O₂ extraction (Boone, Barstow, et al., 2016; Boone, Vandekerckhove, et al., 2016). It should be noted, however, that one of these studies has actually shown that the RCP and the [HHb]bp occurred at the same metabolic rate (Boone, Barstow, et al., 2016). Furthermore, it has been recently discussed that discrepancies in views as to whether or not the RCP and the [HHb]bp are expressed simultaneously or in sequential order might be explained by the approach used to account for the MRT, as detailed elsewhere (Boone, Caen, Vermeire, Bourgois, & Bourgois, 2019; Keir et al., 2019).

4.1 | Experimental considerations

It should be noted that the goal of this study was to determine whether changes (or the lack thereof) in the V̇O₂ and PO associated with the RCP and the [HHb]bp when breathing hypoxic air were proportional. In this context, it could be argued that including a hypoxia condition would have provided additional insights. In fact, this study originally included such condition and, when the RCP and the [HHb]bp in hypoxia were analyzed (data presented elsewhere (Azevedo et al., 2020)), they occurred at the same %VO₂max and %PPO compared to normoxia condition. However, as stated in our previous manuscript (Azevedo et al., 2020) and also reported
by other studies (Amann et al., 2007; Oussaidene et al., 2013; Rausch, Whipp, Wasserman, & Huszczuk, 1991; Ulrich et al., 2017), measurements of \( \dot{V}O_2 \) using current systems are often not valid during hyperoxic conditions, (even after attempting corrections as proposed elsewhere (Lang, Herold, Kraft, Harth, & Preisser, 2018)). Therefore, the inclusion of the hyperoxia condition is not appropriate since the \( \dot{V}O_2 \) data are not physiologically justifiable.

In conclusion, the present study demonstrated that the RCP and the [HHb]bp during a RI test occur at the same metabolic rate and PO, even under distinct FiO\(_2\) conditions that alter the absolute values at which these intensity indices occur during the test. Furthermore, and most importantly, the magnitudes of the changes in both the RCP and the [HHb]bp from normoxic to hypoxic conditions were proportional on an individual basis. Taken together, data from the present study reinforce the idea that the RCP and the [HHb]bp parameters represent a similar physiological response and that the mechanisms underpinning their occurrence are likely linked to metabolic changes that take place when surpassing the heavy-to-severe boundary of exercise intensity.

5 | PERSPECTIVES AND SIGNIFICANCE

This study is the first to evaluate the correspondence between the RCP and the [HHb]bp during a RI test under distinct FiO\(_2\) conditions (i.e. normoxia and hypoxia). It was found that the \( \dot{V}O_2 \) and PO associated with the RCP and the [HHb]bp were not different within conditions but both reduced under hypoxia condition. Most importantly, the changes in hypoxia, in terms of the \( \dot{V}O_2 \) and PO, negatively affected the RCP and the [HHb]bp in a similar magnitude compared to normoxia. Thus, the present data reinforce the idea that both markers of intensity represent a similar physiological response that delimitates the metabolic rate associated with the transition from the heavy to the severe intensity domain. These findings have important implications to further our understanding on the physiological underpinnings of these markers.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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