Genetic Architecture of Winter Hardiness and Frost Tolerance in Triticale

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Abstract

Abiotic stress experienced by autumn-sown crops during winter is of great economic importance as it can have a severe negative impact on yield. In this study, we investigated the genetic architecture of winter hardiness and frost tolerance in triticale. To this end, we used a large mapping population of 647 DH lines phenotyped for both traits in combination with genome-wide marker data. Employing multiple-line cross QTL mapping, we identified nine main effect QTL for winter hardiness and frost tolerance of which six were overlapping between both traits. Three major QTL were identified on chromosomes 5A, 1B and 5R. In addition, an epistasis scan revealed the contribution of epistasis to the genetic architecture of winter hardiness and frost tolerance in triticale. Taken together, our results show that winter hardiness and frost tolerance are complex traits that can be improved by phenotypic selection, but also that genomic approaches hold potential for a knowledge-based improvement of these important traits in elite triticale germplasm.

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Introduction

Triticale (x Triticosecale Wittmack L.; 2n = 6x = 42) combines properties of its two parents, wheat and rye, and is primarily grown in the northern hemisphere. In Central Europe it is commonly planted as winter triticale in autumn, as varieties with a winter growth habit usually have a higher yield potential than varieties planted in spring. A prerequisite for these winter-type varieties is that they possess an adequate tolerance to endure the harsh conditions during winter in temperate zones. The capability of these plants to survive winter can be referred to as winter hardiness, involving among other factors freezing tolerance, desiccation, anoxia and disease resistance [1]. Of these, frost tolerance, i.e. the ability to survive freezing temperatures, has been considered as the major limiting factor in most regions. Owing to the economic importance, these two traits have been investigated for many decades and show complex genetic regulation [1]. Consequently, progress by breeders to improve these traits has been slow, which is further aggravated by the variable occurrence of this type of stress across years.

An additional complication when studying frost tolerance is that the trait can be affected by genes conferring low temperature tolerance as well as developmental genes with pleiotropic effects [2]. As reproductive meristems are more sensitive to low temperatures, small differences in developmental stage can affect the plants’ ability to survive. As a consequence, genes controlling the transition to the reproductive phase by photoperiod (Ppd) or vernalization (Vrn genes) can also have an impact on frost tolerance [2–4]. To date, two major loci affecting frost tolerance have been identified in diploid and polyploid wheat and mapped to chromosome 5. Frost Resistance-1 (Fr-1) maps close to Vernalization-1 (Vrn-1) and has therefore been suggested to present a pleiotropic effect of this major vernalization gene [2,5,6]. Vrn-1 encodes a MADS box transcription factor similar to the Arabidopsis meristem identity gene API and is upregulated during vernalization [7]. The second locus underlying frost tolerance, Fr-2, has been mapped approximately 30 cM proximal to Vrn-1 [8–10] and the molecular characterization has revealed a cluster of multiple Cbf (C-repeat Binding Factor) genes at this locus that are upregulated by low temperatures [11,12]. In wheat, copy number variation at the Fr-2 locus was recently reported to be associated with frost tolerance [13]. For both Fr QTL, homoeologous loci have been reported on the A, B and D genomes of wheat [14–16]. In rye, twelve members of the Cbf family have also been assigned to the long arm of chromosome 5R [17] and some have recently been reported to be significantly associated with frost tolerance [18]. In addition to the two major Fr-1 and Fr-2 loci, the complex genetic architecture underlying this trait suggests the presence of other genes with small effects on frost tolerance [19].

The aim of this study was to dissect the genetic basis underlying winter hardiness and frost tolerance in triticale, identifying targets for knowledge-based improvement of these two agronomically important traits. To this end, we employed multiple-line cross QTL mapping [20] based on a large triticale mapping population with 647 DH lines derived from four families. In particular, the objectives were to (i) investigate the phenotypic inheritance of the
two traits, (ii) detect main effect QTL and (iii) assess the contribution of epistasis.

Materials and Methods

Plant material, field trials and phenotypic data

The plant material and field trials underlying this study have been described previously [21,22]. In brief, the mapping population consisted of 647 doubled haploid (DH) [23,24] triticale (x Triticosecale Wittmack L.) lines derived from four families, DH06 (131), DH07 (120), EAW74 (200), and EAW78 (196) that have been described by Alheit et al. [25,26]. The DH lines were grown in partially replicated designs [27] including common checks with 960 plots per location and analyzed by alpha analysis of variance. In 2012 two locations (Eckartsweier and Hohebuch) showed severe frost without snow coverage. In Eckartsweier the average temperature in February was −2.0°C and the minimum temperature was −17.8°C while in Hohebuch the average temperature was −2.2°C and the minimum temperature −17.7°C. At both locations winter hardiness was scored as the appearance of the plants at the end of winter (beginning of April) on a scale between 1 (no damage) to 9 (maximum damage, i.e., no plant survived). In addition, the direct effect of frost was scored at one location (Eckartsweier) after the frost had occurred (i.e. frost damage at the end of February), with a score between 1 (no damage) to 9 (maximum damage). For all lines the Best Linear Unbiased Estimates (BLUEs) were determined. All mixed model calculations were performed using the software ASReml 3.0 [28].

Multiple-line cross QTL mapping

The DH lines were genotyped with DArT markers and QTL mapping was done based on the integrated consensus linkage map described by Alheit et al. [25]. For QTL mapping, an additive genetic model was chosen and a joint analysis was performed with a model assuming specific QTL effects for every family [20,29] as described in detail by Steinhoff et al. [30]. In brief, the multiple-line cross QTL mapping model was:

\[ Y = JM + X_cB_c + \sum_{i=1}^{p} X_qB_q + \varepsilon \]

where \( Y \) was a \( N \times 1 \) column vector of the BLUE values of phenotypic data of \( N \) progenies coming from \( P \) families. \( J \) was a \( N \times P \) matrix whose elements were 1 or 0 according to whether or not individual \( i \) belonged to family \( p \) and \( M \) was a \( P \times 1 \) vector of family specific means. \( X_c (X_q) \) was a \( N \times P \) matrix containing the expected number \( ( \text{from } 0 \text{ to } 2) \) of allele \( k \) for each individual in family \( p \) at QTL \( q \) (cofactor \( c \)), and \( B_{cq} \) was a \( P \times 1 \) vector of the expected allele substitution effects of QTL \( q \) (cofactor \( c \)) in family \( p \). \( \varepsilon \) was the vector of the residuals.

Cofactor selection was performed using PROC GLMSELEQT in the statistical software SAS [31]. The presence of a putative QTL in an interval was tested using a likelihood-ratio test with the statistical software R [32]. LOD-thresholds of 4.7 for winter hardiness and 4.66 for frost tolerance were used corresponding to an experiment-wise type I error of \( P<0.01 \), based on 2000 permutations [33]. The proportion of genotypic variance explained by the detected QTL was estimated as \( R^2 = q^2 / \sigma^2 \) [34]. The support interval of a QTL was defined as a LOD fall-off of 1.0 expressed as position on the chromosome in centimorgans (cM) [35] and cofactors were excluded within a distance to the marker interval under consideration smaller than 10 cM. QTL were declared as overlapping between winter hardiness and frost tolerance if they fell within an arbitrarily defined 10 cM interval surrounding the QTL. Fivefold cross-validation was done as described previously [36,37].

The epistasis scan for pairwise interactions was done with the model described above which was extended by the term \( X_qX_q \) for the second locus and the interaction term \( X_qX_q \) between the two loci \( q, q' \). We used an \( \alpha \)-level of 0.05 and followed the suggestion of Holland et al. [30] dividing the \( \alpha \)-level by the number of possible independent pairwise interactions between chromosome regions, assuming two separate regions per chromosome \((P<5.3e-5)\). The circular plots illustrating the epistatic interactions were created with Circos [39].

Results

In the mapping population with 647 DH lines, we observed significant \( (P<0.01) \) genotypic variances \( (\sigma^2_G) \) for both winter hardiness and frost tolerance (Table 1). For winter hardiness, which was evaluated at two locations, the genotype by environment interaction variance \( (\sigma^2_{G 	imes E}) \) could be estimated. This was also found to be significant and the ratio of genotypic variance to genotype by environment interaction variance was 9:1. For both traits the observed phenotypic values covered the full range from highly susceptible to fully tolerant (Table 1, Figure 1). The heritability was high with 0.37 for winter hardiness and the repeatability was 0.91 for frost tolerance, while the the phenotypic correlation between both traits was 0.88 \((P<0.01)\). The phenotypic values of the parents differed to varying degrees ranging from 2.9 to 5.9 for winter hardiness and from 3.9 to 6.6 for frost tolerance (Figure 1). The trait distributions approximately followed a normal distribution and in each family DH lines transgressed the respective parents (Figure 1). Taken together, these results illustrate that the mapping population is well suited to study the genetics underlying winter hardiness and frost tolerance in triticale.

Multiple-line cross QTL mapping revealed 9 QTL for winter hardiness as well as for frost tolerance (Table 2, 3, Figure 2) of which 6 were overlapping between both traits. The detected QTL explained 63.0 and 59.8 percent of the genotypic variance of winter hardiness and frost tolerance, respectively. The proportion of genotypic variance explained by single QTL ranged from 2.0 to 24.1 percent for winter hardiness and from 1.9 to 16.3 percent for frost tolerance (Table 3). For winter hardiness as well as for frost tolerance three major QTL explaining more than 10 percent of genotypic variance were detected on chromosomes 5A, 1B and 5R. Interestingly, the major QTL on chromosome 1B appeared to segregate in all four families while family EAW74 did not segregate for the QTL on 5R but was the only family segregating for the QTL on chromosome 5A. The single family analysis revealed one more major QTL for frost tolerance located on chromosome 6A, segregating in family DH07.

We used fivefold cross-validation to obtain less biased estimates of the proportion of genotypic variance \( (\rho_G) \) explained by the QTL (Table 2). The cross-validated \( \rho_G \) was 31.1 and 30.0 percent for winter hardiness and frost tolerance, respectively. The QTL frequency distributions revealed that some QTL including the three major QTL were detected in a high number of runs further substantiating their importance (Figure 2).

The full 2-dimensional epistasis scan revealed seven epistatic QTL for winter hardiness and six for frost tolerance (Figure 3). Interestingly, the epistatic QTL for frost tolerance between chromosomes 5A and 1R involves the same chromosomal region on chromosome 5A (at \( \approx 53 \) cM) that was identified as major QTL. The contribution of these epistatic QTL to the genotypic variance was small and ranged between 0.1 and 3.5 percent for
winter hardiness and between 0.9 and 4.1 percent for frost tolerance.

Discussion

Frost tolerance is one of the most important abiotic stresses. In both hemispheres it is a factor limiting the geographic distribution of crops but can also have severe effects on crop production potentially resulting in complete losses. The higher yield makes winter varieties attractive but requires breeding for increased winter hardiness and frost tolerance to ensure survival of the plants during winter. Whereas winter hardiness and frost tolerance have been studied in wheat and rye, little is known about the inheritance of both traits in triticale. The aim of this study therefore was to investigate the genetic architecture of winter hardiness and frost tolerance in triticale.

Phenotypic evaluation

Triticale (AABBRR) contains both wheat (AABB) and rye (RR) genomes, with rye being an extremely frost tolerant species among small grain cereals [40,41]. Although triticale is less frost tolerant than rye, we observed significant genotypic variances for winter hardiness and frost tolerance in the mapping population. The estimated genotypic values covered the full range from highly tolerant to fully susceptible illustrating the potential to improve these traits by breeding. The trait distributions highlight the quantitative nature of the traits, suggesting a contribution of many small-effect QTL but not excluding the presence of few major effect QTL. We observed substantial differences between the parents for each family. The transgressive segregation, however, suggests that the less tolerant parent carried favourable alleles not present in the better parent. Such positive alleles can be combined in the progeny resulting in lines that are superior to the parental lines, illustrating the potential of phenotypic selection to improve these traits in triticale. Consequently, crosses including the most frost tolerant lines identified here appears as a promising strategy to further increase the level of frost tolerance in elite triticale germplasm. The high correlation between winter hardiness and frost tolerance confirms frost tolerance as a major factor contributing to the plants’ survival during winter.

A major limitation, however, for the conventional improvement of winter hardiness and even more so of frost tolerance is the wide variation in occurrence of these stresses across years. Semi-controlled tests have recently been shown to correlate well with results from field trials in durum wheat and thus offer a possibility to screen for frost tolerance irrespective of the snow coverage on the field [42]. Another option is the identification of QTL for marker-assisted selection [43]. The phenotypic data from the field trials in the year 2012 with its exceptionally strong frost in the absence of snow offers an excellent basis for the detection of QTL underlying winter hardiness and frost tolerance in triticale.

Detection of main effect QTL

We identified a total of nine QTL for winter hardiness and frost tolerance of which six were detected for both traits. The high number of overlapping QTL detected for winter hardiness and frost tolerance further substantiates frost tolerance as a major contributor to winter survival. Among the identified QTL, three explained more than 10 percent of the genotypic variance for both traits and can be considered as major QTL (Table 3). One of these

| Table 1. Summary statistics for winter hardiness and frost tolerance. |
|----------------------|----------------------|
|                      | Winter hardiness     | Frost tolerance   |
| Min                  | 0.9                  | 0.4               |
| Mean                 | 3.9                  | 4.8               |
| Max                  | 8.1                  | 8.9               |
| $\sigma^2_G$         | 3.00**               | 2.36**            |
| $\sigma^2_{G \times E}$ | 0.33**         | n.a.              |
| $\sigma^2_e$         | 0.81                 | 0.34              |
| $h^2$                | 0.87                 | 0.91              |

Genotypic variance ($\sigma^2_G$), genotype by location interaction variance ($\sigma^2_{G \times E}$), error variance ($\sigma^2_e$), and heritability ($h^2$). ** significant at the 0.01 probability level. doi:10.1371/journal.pone.0099848.t001

Figure 1. Histograms of the phenotypic values. Shown for (a) winter hardiness and (b) frost tolerance for the entire population (All) and for each of the four families (DH06, DH07, EAW74, EAW78). The arrowheads indicate the phenotypic values of the respective parents. doi:10.1371/journal.pone.0099848.g001
Table 2. Results of QTL mapping and fivefold cross-validation.

| QTL | Winter hardness | Frost tolerance |
|-----|----------------|-----------------|
| QTLDS | 9 | 9 |
| pG-DS | 63.0 | 59.8 |
| QTLES | 8.5 | 8.8 |
| pG-ES | 52.5 | 52.0 |
| pG-TS | 31.3 | 30.0 |
| Relative bias | 40.4 | 42.3 |

Number of detected QTL (QTLDS), proportion of genotypic variance (%) explained by the detected QTL across all families in the data set (pG-DS), number of QTL (QTLES) and proportion of genotypic variance averaged over estimation sets (pG-ES) and averaged over test sets (pG-TS), and relative bias (%) in the estimation of pG.

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Table 3. QTL detected for winter hardiness and frost tolerance.

| Chromosome | Position in cM [support interval] | pGDH06 | pGDH07 | pG EAW74 | pG EAW78 |
|------------|----------------------------------|--------|--------|----------|----------|
| Winter hardiness | 2A | 62.1 [58.0–63.8] | 2.2 | 0.6 | 3.6 | 0.3 | 0.2 |
| | 5A | 52.7 [51.6–52.9] | 14.1 | 1.2 | 0.2 | 40.2 | 0.0 |
| | 7A | 63.6 [63.5–63.7] | 2.3 | 0.0 | 0.5 | 3.1 | 3.3 |
| | 1B | 54.1 [53.2–54.3] | 12.5 | 14.4 | 17.4 | 7.6 | 18.8 |
| | 2B | 107.4 [107.0–108.1] | 2.9 | 4.5 | 3.4 | 3.1 | 4.1 |
| | 5B | 39.9 [39.3–40.7] | 2.8 | 2.1 | 2.2 | 5.3 | 1.3 |
| | 6B | 56.8 [55.3–58.7] | 2.0 | 1.5 | 6.8 | 0.7 | 0.2 |
| | 4R | 35.7 [35.3–36.1] | 2.6 | 0.0 | 0.2 | 4.8 | 2.3 |
| | 5R | 58.9 [58.6–58.9] | 24.1 | 44.8 | 20.4 | 3.4 | 29.8 |
| | A genome | 18.6 | 1.8 | 4.3 | 43.6 | 3.5 |
| | B genome | 20.2 | 22.5 | 29.8 | 16.7 | 24.4 |
| | R genome | 26.7 | 44.8 | 20.6 | 8.2 | 32.1 |

| Frost tolerance | 1A | 80.1 [80.0–80.4] | 2.6 | 0.0 | 8.3 | 1.4 | 1.5 |
| | 5A | 52.5 [51.6–52.9] | 12.4 | 2.1 | 0.5 | 40.2 | 0.3 |
| | 6A | 14.5 [14.0–14.6] | 2.2 | 0.0 | 13.1 | 0.0 | 0.2 |
| | 7A | 71.6 [66.8–79.3] | 3.4 | 0.4 | 4.1 | 3.8 | 1.9 |
| | 1B | 54.1 [53.2–54.3] | 16.3 | 19.0 | 15.4 | 5.1 | 25.3 |
| | 2B | 79.3 [78.5–79.3] | 2.9 | 5.6 | 3.4 | 4.3 | 4.1 |
| | 6B | 57.8 [55.3–58.7] | 1.9 | 1.6 | 2.5 | 3.1 | 0.6 |
| | 4R | 25.3 [25.1–25.3] | 3.0 | 0.8 | 4.5 | 6.9 | 0.0 |
| | 5R | 55.6 [55.4–56.2] | 14.7 | 31.7 | 9.0 | 1.5 | 18.2 |
| | A genome | 20.6 | 2.5 | 26.0 | 45.4 | 3.9 |
| | B genome | 21.1 | 26.2 | 21.3 | 12.5 | 30.0 |
| | R genome | 17.7 | 32.5 | 13.5 | 8.4 | 18.2 |

Chromosome, position with support interval with a LOD fall off of 1.0 and proportion of genotypic variance explained by the QTL (pG in %) in the entire population and in each of the families, and the summary for the three genomes.

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major QTL identified on chromosome 1B requires further research. Our findings illustrate that all genomes contribute to winter hardiness and frost tolerance in triticale. Interestingly, more of the detected QTL are derived from the wheat A and B genomes and not from the rye genome (Table 3), despite rye being the more frost tolerant species [41]. While primary triticale has been obtained by crosses between durum wheat with low winter hardiness and rye, secondary triticale has resulted from backcrosses with wheat, potentially explaining the source of the winter hardiness and frost tolerance QTL on the A and B genomes. In addition, phenotypic evaluation of octoploid and hexaploid triticale has revealed that their cold hardness levels were always more similar to their wheat parents suggesting a suppression of the excellent rye cold hardness in triticale, either by suppression of the responsible rye genes or by a dilution of their effects due to the polyploid nature of triticale [40]. Alternatively, important frost tolerance QTL alleles on the rye genome may be fixed in all six parents and may thus not be segregating in the mapping population. It must be noted that only segregating QTL will contribute to the phenotypic variation and can be detected,
Contribution of epistasis

Epistasis refers to interactions between the alleles at two or more genetic loci in the genome [46] and has recently been shown to contribute to the genetic architecture of complex traits in different crops including maize, wheat and rapeseed [47–51]. Interactions among key factors underlying frost tolerance are present on the molecular level [1] suggesting that epistasis could contribute to the genetics of this trait. One significant epistatic interaction has been reported for the candidate gene association mapping study on frost tolerance in rye [18] and a number of epistatic interactions for the genome-wide association mapping approach in wheat [19]. Here, we identified seven epistatic QTL for winter hardness and six for frost tolerance of which one might be identical in both traits (Chr 6A - 4R; Figure 3). Interestingly, one epistatic interaction involved the major QTL identified on chromosome 5A suggesting that this locus, which might be Vrn-1, is also involved in epistatic interactions. Our results thus corroborate previous findings from wheat and rye and show the contribution of epistasis to the genetic architecture of winter hardness and frost tolerance in triticale.

Conclusions

In this study, we employed multiple-line cross QTL mapping based on a large DH mapping population to dissect the genetic architecture of winter hardness and frost tolerance in triticale. We identified main effect QTL on all three genomes and three major effect QTL, some of which potentially correspond to known frost tolerance loci. In addition, our results reveal the contribution of epistasis to both traits. In summary, winter hardness and frost tolerance in triticale are complex traits that can be improved by phenotypic selection but also hold the potential for a knowledge-based selection of superior lines.

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Author Contributions

Conceived and designed the experiments: TW HPM VH. Performed the experiments: HPM EAW VH. Analyzed the data: WL GL MG TW. Contributed to the writing of the manuscript: TW MRT.

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