How Far Have We Come in the Field of Nerve Regeneration After Trigeminal Nerve Injury?

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Abstract Patients suffering from nerve injury with sensory disturbances or orofacial pain have greatly reduced quality of life, and it is a big cost for the society. Abnormal sensations caused by trigeminal nerve injury often become chronic, severely debilitating, and extremely difficult to treat. In general, non-invasive treatment such as drug treatment has been insufficient, and there are currently few available effective treatments. Surgical interventions such as end-to-end connection or nerve grafting have disadvantages such as donor site morbidity or formation of neuroma. There is need for optimizing the technique for nerve repair, especially for the trigeminal nerve system, which has so far not yet been well explored. Recently, tissue engineering using biodegradable synthetic material and cell-based therapies represents a promising approach to nerve repair and it has been reported that mesenchymal stem cell (MSC) has an anti-inflammatory effect and seems to play an important role in nerve healing and regeneration.

Keywords Sensory neurons · Orofacial sensory disturbances · Trigeminal nerve · Inferior alveolar nerve · Lingual nerve · Infraorbital nerve · Hypoesthesia · Nerve regeneration · Mesenchymal stem cells · Adipose derived stem cells · Stem cells therapy

Introduction

Background

Dysfunction of the trigeminal nerve due to trauma, diseases, or unknown causes is for the patients distressing. The sensory disturbances and/or pain are unpleasant conditions. It can involve the function of the mandible, the muscles, the skin of chin and lips, the intra orally mucosa, and the tongue and give rise to several problems such as pain, inability to move the jaw, tongue-lip or cheek biting, inability to maintain food and liquid competence, burning sensation with provocative stimuli, change in speech pattern, and change in taste perception [1]. Studies of patients that are affected by temporomandibular disorders (TMD), which may present a longstanding pain condition where muscles and/or joint are involved, have revealed that psychological factors dominate as a consequence of living with pain [2]. In a study where TMD patients were investigated with a multidisciplinary approach, most of the patients had a long history of pain, significant high levels of catastrophizing, and high occurrence of anxiety and/or depression [3]. Injury to the somatosensory pathways may either increase the nerve transmission like in allodynia and hyperalgesia or decrease the transmission such as in hypoesthesia or anesthesia [4, 5]. An important sequel of nerve injury and other nervous system diseases is neural degeneration.
Abnormal sensation induced by peripheral nerve injury has been considered as a progressive neurodegenerative disease [6].

Traumatic nerve injury of the trigeminal nerve is a major clinical challenge. The frequently most affected trigeminal branch is the inferior alveolar nerve (IAN) followed by the lingual nerve (LN) and finally the infraorbital nerve (ION) [7, 8] (Fig. 1). It has earlier been reported that spontaneous recovery of injured IAN after 6–9 months will leave some degree of long-term permanent disability [9]. Alteration of sensation in the damaged nerve results from either direct or indirect damage due to compression, stretching, or laceration. The degree of alteration depends on the severity of injury [10].

When the nerve is damaged, an inflammation process will start which releases a cascade of prostaglandins to the surrounding tissue which will spread to sensory neurons. The inflammation response will maintain the painful symptoms which in turn can develop to both peripheral and central sensitization [11]. During the inflammation process, interactions of macrophages and monocytes from peripheral blood migrate towards the damage site in the peripheral nerve which is connected via cell bodies in the dorsal spinal cord and the trigeminal ganglion and further to central parts of the brain. They will consequently activate microglia, which are surrounded by satellite glia cells (SGCs), and underlie peripheral sensitization which in turn maintain allodynia and hyperalgesia [12]. This finding has been demonstrated in an animal study where injections of capsaicin to the temporal joint capsule showed that SGCs were activated [13]. Further, activation of sensory neurons in the mandible has shown resultant spreading of neuronal activity not only in the third trigeminal branches but also to the first and second branches (Fig. 1). A cross excitation to extraterritorial sites outside the injured dermatome was the interpretation of the results [14]. Recent evidence in animal studies indicates that deficits of trigeminal nerve system may lead to impairments in learning and memory and neuronal loss in the hippocampus [15].

Orofacial pain induced by trigeminal nerve injury is often a symptom complex rather than a single condition, and it is thought to be caused by multiple factors. However, these factors are poorly understood. A major obstacle in exploring mechanisms and treatments of neuropathic pain is that our conventional understanding of pain physiology and pharmacology has been built primarily on studies of nociceptive pain whereas persistent or neuropathic pain in many aspects differs from, and even is contrary to, nociceptive pain [16]. Clinical research on this problem is difficult as these are not common disorders, and thus, homogenous patient samples for important variables are difficult to obtain. Furthermore, invasive methods often have to be used to address the underlying mechanisms and novel unproven treatments tested.

Available Treatments

Treatment of abnormal sensation of sensory nerves such as pain is usually, as a first choice, pharmacological. Acute pain can be treated successfully with paracetamol, non-steroidal anti-inflammatory drugs (NSAID), and/or morphine. When the postoperative pain develops into persistent neuropathic pain, medication with antiepileptic drugs such as gabapentin or carbamazepine is used with different outcomes [12]. The side effects of this type of medication can be a problem for the patient, which include drowsiness, dry mouth, and negative mood changes that affect their quality of life.

Searching for new targets in the pharmacological field to treat neuropathic pain is important. Transient receptor potential (TRP) channels are present in sensory neurons and are involved in the development of pain. Neuropathic injury in humans has been shown to increase the expression of TRPA1 [17], and studies on tooth injury have specific shown to increase the expression of the TRPA1 channels [18]. TRP channel antagonists could be promising as novel analgesic agents [19]. There are several antagonists that have been shown to block the TRPA1-induced neuropeptide release in dental pulp. Recently, we have shown that a novel TRPA1 antagonist inhibits TRPA1 agonist-stimulated release of neuropeptides from dental pulp biopsies in an in vivo model [20].
Serotonin (5-HT) is a neuromodulator and plays an important role as a mediator of pain both centrally and peripherally [21]. It has been suggested that 5-HT$_3$ receptors are activated in humans with muscle pain [22]. Recently, it was shown from our group that 5-HT$_3$ receptors were highly expressed in masseter muscles of women with TMD compared to controls [23]. It was concluded that in myofacial TMD up-regulated 5-HT$_3$ receptors could serve as a biomarker. In the search for 5-HT$_3$ receptor antagonists as a target for blocking orofacial pain, only one study was found in animals. Painful Injection of formalin to the masseter muscle in rats attenuated nociceptive behavior by both local and systemic administration of a 5-HT$_3$ receptor antagonist [24].

Non-invasive treatment modalities such as local anesthesia have been used as neural therapy which approach for long-term relief of pain after nerve injury. Neural therapy should be repeated several times with increasing time intervals. The effect of Local anesthetic has been speculated to provide protection against sprouting in sympathetic nerves [25] and give a pro-inflammatory effect [26]. Recently, a review by Weinschenk raised the question whether or not local anesthetics can interrupt the liberation of pro-inflammatory substances at the terminal plate in neurogenic inflammation [27].

Another non-invasive type of treatment is low-level laser therapy (LLLT), which has shown good results in subjects with nerve injuries that are identified immediately [28]. For longstanding injuries, there has been registered some improvement with LLLT [29, 30]. In a study, LLLT showed some efficacy for long term of sensory disturbances following third molar surgery [31]. For TMD or temporomandibular joint derangement (TMJD), two systematic reviews have concluded that LLLT is probably more effective for the treatment of TMJD and less effective for TMD [32, 33].

Treatments available for nerve injury have shown some functional recovery in humans, i.e., more sensation and/or less pain, but evidence lacks for nerve regeneration. Surgical procedures for reconstruction of peripheral nerves are available such as microsurgical approaches with direct end-to-end connection. The gold standard for nerve grafting is autologous nerve, sural nerve, or auricular nerve to be used for the reconstruction. The critical size of the gap depends on the number of sensory nerves or nerve fibers that can be bridged. A large number of animal studies have been carried out, mostly in rats and mice. The critical size of the gap between the axons is 10 mm. The choice of analyzing methods for measurement of nerve regeneration employs anatomic and histological methods but a functional evaluation is necessary. There is still need for optimizing the NGC, especially for the trigeminal nerve, which has so far not yet been well explored. Nanomaterials mimic the properties for natural tissues and may resolve the numerous problems associated with today’s limitations.

Use of mesenchymal stem cells as a new treatment is of interest due to the core properties of these cells. It has been reported that MSCs have an anti-inflammatory effect and they seem to play an important role in nerve healing and regeneration [43]. In animal studies, promising results have been presented where neuropathic trigeminal pain has been reduced following MSC treatment [44+]. Recently, a preliminary report described the outcomes following injection of autologous stem
cells into the pain fields in female patients with different diagnosis of neuropathic pain including trigeminal neuralgia, PDAP, and BMS. It was found that the pain intensity scores and use of anti-neuropathic medication were strongly reduced for 6 months after administration of the cells [45–46]. It has also been shown that multiple or high doses prolong the therapeutic effect much longer than for a single dose [46–47].

**Significant Trends or Developments**

To our knowledge, published data on MSC treatment for patient with neuropathic pain are very sparse. In one case study, Ichim et al. (2010) reported a positive result for suppressing neuropathic pain from expanded umbilical cord by intrathecal injection of MSCs [47]. More recently, in a study involving ten patients, Vickers and colleagues (2014) demonstrated a significant effect of MSC treatment on neuropathic trigeminal pain [45–47]. They reported that approximately 56% of patients (5 of 9), who suffered from chronic pain for 4 months to more than 6 years, showed a reduction of pain intensity scores. Moreover, during the investigation, all patients were given anti-neuropathic medication, amitriptyline, and gabapentin. The change in daily dosage requirements of medication showed a near significant reduction in gabapentin and minor reduction in amitriptyline, which indicated a possible biological priority of stem cells in recovery myelinated fibers over unmyelinated fibers. Interestingly, the same group investigators also found that one of the most responders was an 80-year-old patient. They concluded that MSCs can produce many factors to achieve the therapeutic effect and the secretion profile of the stem cells remains unaffected by age, which is consistent with other observations [48–50]. To a significant extent, the study clarified a positive outcome from neuropathic pain patients in response to a single dose of MSCs, suggesting that a possible enhanced therapeutic effect could be achieved with multiple dosage strategies.

**Conclusion**

Patients suffering from nerve injury-induced sensory disturbances and/or pain have greatly reduced quality of life, and it is a big cost for the society. The vast majority of the work on sensory disturbances/pain mechanisms has been carried out in spinal nerve systems. These studies have provided great insight into mechanisms regarding pain of the spinal area. However, it is clear that the pathophysiology of the trigeminal nerve is in many ways different to that found in spinal nerves. Treatments that are available today are not enough to cure the patient, recover the nerve sensibility, and/or reduce pain. Stem cells therapy could be a future solution to solve the situation for the patients.

**Compliance with Ethical Standards**

**Conflict of Interest** Arezo Tardast, Tie-Jun Shi, and Annika Rosén declare that they have no conflict of interest.

**Human and Animal Rights and Informed Consent** This article does not contain any studies with human or animal subjects performed by any of the authors.

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**References**

Papers of particular interest, published recently, have been highlighted as:

- Of importance
- Of major importance

1. Boffano P, Rocca F, Gallesio C. Lingual nerve deficit following mandibular third molar removal: review of the literature and medical considerations. Oral Surg Oral Med Oral Pathol Oral Radiol. 2012;113(3):e10–8.

2. Slade GD, Ohrbach R, Greenspan JD, Fillingim RB, Bair E, Sanders AE, Dubner R, Diatchenko L, Meloto CB, Smith S, Maixner W. Painful temporomandibular disorder: decade of discovery from OPPERA studies. J Dent Res. 2016;95:1084–92.

3. Berge T, Schjødt B, Bell RF, Johansson A, Paulsberg A-G, Geitung JT, Rosén A. Assessment of patients with severe temporomandibular disorder in Norway—a multidisciplinary approach. Nordic theme 2016. Nor Tannlegeforen Tid. 2016;126:114–21.

4. Hansson P, Backonja M, Bouhassira D. Usefulness and limitations of quantitative sensory testing: clinical and research application in neuropathic pain states. Pain. 2007;129(3):256–69.

5. Svensson P et al. Guidelines and recommendations for assessment of somatosensory function in oro-facial pain conditions—a taskforce report. J Oral Rehabil. 2011;38(5):366–94.

6. Zimmermann M. Pathobiology of neuropathic pain. Eur J Pharmacol. 2001;429(1–3):23–37.

7. Hillerup S. Iatrogenic injury to the inferior alveolar nerve: etiology, signs and symptoms, and observations on recovery. Int J Oral Maxillofac Surg. 2008;37(9):704–9.

8. Schultz-Mosgau S, Krems H, Ott R, Neukam FW. A prospective electromyographic computer-aided thermal sensitivity assessment of nerve lesions after sagittal split osteotomy. J Oral Maxillofac Surg. 2001;59(2):128–38.

9. Robinson PP. Observations on the recovery of sensation following inferior alveolar nerve injuries. Br J Oral Maxillofac Surg. 1988;26(3):177–89.

10. Burnett MG, Zager EL. Pathophysiology of peripheral nerve injury: a brief review. Neurosurg Focus. 2004;16(5):E1.

11. Austin PJ, Kim CF, Perera CJ, Moalem-Taylor G. Regulatory T cells attenuate neuropathic pain following peripheral nerve injury and experimental autoimmune neuritis. Pain. 2012;153:1916–31.
12. Ren K, Dubner R. Interactions between the immune and nervous systems in pain. Nat Med. 2010;16:1267–76.

13. Durham PL, Garrett FG. Development of functional units within trigeminal ganglia correlates with increased expression of proteins involved in neuron-glia interactions. Neuron Glia Biol. 2010;6(3):171–81.

14. Thalakoti S, PatilVV, Damodaram S, Vause CV, Langford LE, Freeman SE, Durham PL. Neuron-glia signaling in trigeminal ganglion: implications for migraine pathology. Headache. 2007;47(7):1008–23.

15. He Y, Zhu J, Huang F, Qin L, Fan W, He H. Age-dependent loss of cholinergic neurons in learning and memory-related brain regions and impaired learning in SAMP8 mice with trigeminal nerve damage. Neural Regen Res. 2014;9(2):1985–94.

16. McKay Hart A, Brannstrom T, Wiberg M, Terenghi G. Primary sensory neurons and satellite cells after peripheral axotomy in the adult rat: timescourse of cell death and elimination. Exp Brain Res. 2002;142(3):308–18.

17. Anand U, Otto WR, Facer P, Zebda N, Selmer I, Gunthorpe MJ. TRPA1 receptor localization in the human peripheral nervous system and functional studies in cultured human and rat sensory neurons. Neurosci Lett. 2008;438:221–7.

18. Haas ET, Rowland K, Gautam M. Tooth injury increases expression of the cold sensitive TRP channel TRPA1 in trigeminal neurons. Arch Oral Biol. 2011;56:1604–9.

19. Brederdon JD, Kym PR, Szallasi A. Targeting TRP channels for pain relief. Eur J Pharmacol. 2013;716:61–76.

20. Nyman E, Franzen B, Nolting A, Klement G, Liu G,Nilsson M, Rosén A, Björck C, Weigelt D, Wollberg P, Karila P, Raboisson P. In vitro pharmacological characterization of a novel TRPA1 antagonist and proof of mechanism in a human dental pulp model. J Pain Res. 2013;6:59–70.

21. Zeitz KP, Guy N, Malmberg AB, Dirjalal S, Martin WJ, Sun L, Bonhaus DW, Stucky CL, Julius D, Basbaum AI. The 5-HT3 sub-type of serotonin receptor contributes to nociceptive processing via a novel subset of myelinated and unmyelinated nociceptors. J Neurosci. 2002;22(3):1010–9.

22. Sung D, Dong X, Embarg M, Kumar U, Cairns BE. Serotonin (5-HT) excites rat mastocytous muscle afferent fibers through activation of peripheral 5-HT3 receptors. Pain. 2008;134(1–2):41–50.

23. Christidis N, Kang I, Cairns BE, Dong X, Rosén A, Kopp S, Embarg M. Expression of 5-HT3 receptors and TTX resistant sodium channels (Na(V)1.8) on muscle nerve fibers in pain-free humans and patients with chronic myofascial temporomandibular disorders. J Headache Pain. 2014;15:63. doi:10.1186/1129-2377-15-63.

24. Okamoto K, Imbe H, Tashiro A, Kumaibe S, Senba E. Blockade of peripheral 5HT3 receptor attenuates the formalin-induced nociceptive behavior in persistent temporomandibular joint inflammation of rat. Neurosci Lett. 2004;367(2):259–63.

25. Takatori M, Kuroda Y, Hirose M. Local anesthetics suppress nerve growth factor mediated neurite outgrowth by inhibition of tyrosine kinase activity of TrkA. Anesth Analg. 2006;102(2):462–7.

26. Cassuto J, Sinclair R, Bonderovic M. Anti-inflammatory properties of local anesthetics and their present and potential clinical implications. Acta Anaesthesiol Scand. 2006;50(3):265–82.

27. Weinschenk S. Neural therapy—a review of the therapeutic use of local anesthetics. Acupuncture and Related Therapies. 2012;1:5–9.

28. Miloro M, Repasky M. Low-level laser effect on neurosensory recovery after sagittal ramus osteotomy. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2000;90(1):12–8.

29. Khullar SM, Brodin P, Barkvoll P, Haanes HR. Preliminary study of low-level laser for treatment of long-standing sensory aberrations in the inferior alveolar nerve. J oral and Maxillofac Surg. 1996;54(1):2–7.

30. Leung YY, Fung PP, Cheung LK. Treatment modalities of neurosensory deficit after lower third molar surgery: a systematic review. J Oral Maxillofac Surg. 2012;70(4):768–78.

31. Ozen T, Orhan K, Gorur I, Ozturk A. Efficacy of low level laser therapy on neurosensory recovery after injury to the inferior alveolar nerve. Head Face Med. 2006;2:3.

32. Melis M, Di Giosia M, Zawawi KH. Low level laser therapy for the treatment of temporomandibular disorders: a systematic review of the literature. Crainio. 2012;30(4):304–12.

33. Petrucci A, Sgolastra F, Gatto R, Matti A, Monaco A. Effectiveness of low-level laser therapy in temporomandibular disorders: a systematic review and meta-analysis. J Orofac Pain. 2011;25(4):298–307.

34. Bagheri SC, Meyer RA. Management of mandibular nerve injuries from dental implants. Atlas Oral Maxillofac Surg Clin N Am. 2011;19:47–61.

35. Siemionow M, Brzezicki G. Chapter 8: current techniques and concepts in peripheral nerve repair. Int Rev Neuroiolo. 2009;87:141–72.

36. Moore AM, MacEwan M, Santos A, KB, et al. Acetaldehyde nerve allografts in peripheral nerve regeneration: a comparative study. Muscle Nerve. 2011;44(2):221–34.

37. Pindrik J, Belzberg AJ. Peripheral nerve surgery; primer for the imagers. Neuroimagi Clin N Am. 2014;24(1):193–210.

38. Kitada M. Mesenchymal cell populations: development of the induction systems for Schwann cells and neuronal cells and finding the unique stem cell population. Anat Sci Int. 2012;87(1):24–44.

39. di Summa PG et al. Adipose-derived stem cells enhance peripheral nerve regeneration. J Plast Reconstr Aesthet Surg. 2010;63(9):1544–52.

40. Matsumoto T, Okabe T, Ikawa T, Iida T, Yasuda H, Nakanura H, Wakitani S. Articular cartilage repair with autologous bone marrow mesenchymal cells. J Cell Physiol. 2010;225(2):291–5.

41. Im GI. Adipose stem cells and skeletal repair. Histol Histopathol. 2013;28(5):557–64.

42. Nectow AR, Marra KG, Kaplan DL. Biomaterials for the development of peripheral nerve guidance conduits. Tissue Eng Part B Rev. 2012;18(1):40–50.

43. Dado-Nachum M et al. Differentiated mesenchymal stem cells for sciatic nerve injury. Stem Cell Res. 2011;7(3):664–71.

44. Guo W et al. Bone marrow stromal cells produce long-term pain relief in rat models of persistent pain. Stem Cells. 2011;29(8):1294–303. This manuscript represents important preclinical evidence of using stem cells in treatment of long-duration persistent pain associated with TMJD.

45. Vickers ER et al. A preliminary report on stem cell therapy for neuropathic pain in humans. J Pain Res. 2014;7:255–63. This manuscript represents the clinical trial of stem cell administration for neuropathic pain in humans for the first time.

46. Franchi S et al. Intravenous neural stem cells abolish nociceptive hypersensitivity and trigger nerve regeneration in experimental neuropathy. Pain. 2012;153(4):850–61. This manuscript represents a pioneer preclinical study of the use of stem cells in treatment of neuropathic pain.

47. Ichim TE, Solano F, Lara F, Paris E, Ugalde F, Rodriguez JP, Minev B, Bogin V, Ramos F, Woods EM, Murphy MP, Patel AN, Hamran RJ, Riordan NH. Feasibility of combination allogenic stem cell therapy for spinal cord injury: a case report. Int Arch Med. 2010;3:30.

48. Blaber SP, Webster RA, Hill CJ, Breen EJ, Kuah D, Vesey G, Herbert BR. Analysis of in vitro secretion profiles from adipose-derived cell populations. J Transl Med. 2012;10:17.

49. Rehman J, Traktuev D, Li J, Merfeld-Clauss S, Temm-Grove CJ, Bovenkerk JE, Pell CL, Johnstone BH, Considine RV, March KL. Secretion of angiogenic and antiapoptotic factors by human adipose stromal cells. Circulation. 2004;109(10):1292–8.

50. De Francesco F, Tirino V, Desiderio V, Ferraro G, D’Andrea F, Giuliani M, Libondi G, Pirozzi G, De Rosa A, Papaccio G. Human CD34/CD90 ASCs are capable of growing as sphere clusters, producing high levels of VEGF and forming capillaries. PLoS One. 2009;4(8):e6537.