Elsholtzia: phytochemistry and biological activities

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Abstract

Plants of the genus Elsholtzia (Lamiaceae) have a long history of medicinal use in folk. The phytochemical investigations revealed the presence of flavonoids, phenylpropanoids, terpenoids, and other compounds. Abundant volatile components are also identified. Pure compounds, volatile constituents and crude extracts from the genus exhibited a wide spectrum of in vitro and in vivo pharmacological activities. The aims of this review hopefully provide comprehensive information on the distribution, phytochemistry, volatile components, and pharmacological research of Elsholtzia for exploring the potential and advance researches.

Review

Background

Elsholtzia is a genus containing at least 33 species in the family Lamiaceae. They have been widely distributed and applied in East Asia, Africa, North America, and European countries for centuries. The genus Elsholtzia plants are mostly aromatic plants, always used as domestic folk medicine, herbal tea, food, spices, beverages, perfumeries, cosmetics, aromatherapies, and the source of honey manufacture.

As folk medicine, the plants in the genus have been used for the treatment of colds, headaches, pharyngitis, fever, diarrhea, digestion disorder, rheumatic arthritis, nephritises, and nyctalopia in China [1-3]. Another important application of the plants is to repair soil that is contaminated by heavy metals. A growing number of research works are focusing on the function of the genus for repairing soil. The most known one is from E. splendens (E. haichowensis), which is a Cu-accumulator plant widely distributed in Cu-mining wastes and Cu-contaminated soil in China [4-7].

Elsholtzia generally exists at hilly grasslands, waste areas, forests, thickets, or valleys in warm area. E. saxatilis, named ‘Yansheng Xiangru’ in Chinese, grows in rocky crevices. And E. saxatilis is different from other plants of Elsholtzia, being distributed in the northeast of Asia, such as northeast of China, Korea, Russia, and Japan, where are cold compared with in South Asia. So far, E. communis and E. argyi have been cultivated in Yunnan province (China) and Vietnam. Except for the two mentioned species, most other plants in the genus are wild [1-3]. The distribution of all 33 species of Elsholtzia is shown in Additional file 1: Table S1.

Distribution

The genus Elsholtzia (Lamiaceae) was widely found in East Asia, Africa, North America, and Europe, especially in China, Korea, Japan, and India. So far, it is reported that at least 33 species plants of the genus are distributed in China [1,2]. And most of Elsholtzia live at an altitude of 1000 to 3000 meters. E. splendens survives at altitude from 200 to 300 meters. E. cephalantha, E. strobilifera and E. eriostachya grow in high altitude of 3000 to 4000 meters.

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Chemical constituents

Previous phytochemical investigations showed that flavonoids are major ingredients in Elsholtzia. They are characterized by the presence of the substitutional groups and modes, as well as their glycosides. Phenylpropanoids, terpenoids, phytosterols, and cyanogenic glycosides are also main chemical constituents in this genus. In this section, we summarize and classify all reported constituents from Elsholtzia. Compounds 1–144 and the corresponding plant sources are list in Additional file 2: Table S2 and the structures 1–132 are showed in Additional file 3: Figure S1.

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C6-C3 constituents

Up to now, there are 68 C6-C3-C6 compounds isolated and reported from *Elsholtzia*, including flavonoids and their glycosides. Its number and content are the most in all secondary metabolites derived from the genus.

Firstly, compounds 1–30 are flavones, in which several hydroxyls, methoxyls and glycosyl groups are linked to the mother nucleus. The oxygenic function groups are most commonly attached to the C-5 and C-7 positions in the flavones. A small amount of 5, 6-dihydroxy-7, 8-dimethoxy flavone (2) from *S. splendens* was obtained by preparative TLC, which is the second report except its first isolation from the roots of *Scutellaria ramosissima* [8]. Luteolin 7-O-[6”-(3’-hydroxy-4”-methoxy cinnamoyl)]-β-D-glucopyranoside (28) and luteolin 7-O-(6’-feruloyl)-β-D-glucopyranoside (29) are very semiable in structure. The difference is that the glucose moiety C-6 position is attached to a 3”'-hydroxy-4”'-methoxy cinnamoyl in 28, while a 6’-feruloyl in 29.

28 was isolated from the whole plants of *E. bodinieri* as a new compound [9]. Secondly, 14 flavonols, 31–44, were isolated from the genus. A number of free hydroxyls are attached to the C-3 positions in 32–34. Compounds 35–44 are 3-O-flavonol glycosides, linked with diverse glycosyl groups, i.e. glucopyranosyl, galactosyl or rhamnopyranosyl. Also, four prenyl-flavonoids (45–48) occur in *E. rugulosa* and *E. stauntonii*, among which 5, 7, 3’, 4’-tetrahydroxy-5’-C-prenylflavone 7-O-β-D-glucoside (46) and muxiangrine III (47) were reported as new compounds [10–13]. 3’-Hydroxy-4’, 5’-dimethoxyfurano- flavone (49) and 3’, 4’, 5’-trimethoxyfurano-flavone (50) are furanoflavone. They possess the characteristic of an unsaturated oxygen-containing furanyl fused into the ring-A’s C-6 and C-7 positions. They are the two unique furanoflavones discovered in the genus so far [14]. Structurally compounds 51–55 are all characterized by a gem-dimethylchromene moiety and one hydroxyl attached to C-3’ and C-4’ of ring-B. Muxiangrine I (51) and muxiangrine II (52) were isolated from *E. stauntonii* [11,12]. Compounds 53–55, named sifanghaoines I–III, were new compounds from *E. blandula* respectively [15,16]. 5’-Dihydroxy-7-acetyl-6,8,3’,3”-tetramethylpyran-(3’,4’)-flavone (53) and 5,5’-dihydroxy-7- (α-methyl) butyroxy-6,8,3’,3”-tetramethylpyran-(3’, 4’)-flavone (54) are structurally similar. The distinguishment is that 53 is linked by an acetate group at C-7 position, while 54 is linked by a (α-methyl) butyroxy moiety. A methylenedioxy is substituted C-6 and C-7 positions in 5,5’-dihydroxy-6,7-methylenedioxy-8,3’,3”-trimethylpyran-(3’,4’)-flavone (55). The same substituent ion also occurs in 47.

Furthermore, eight flavanones (56–63) and one flavanonol (64) are listed in Additional file 2: Table S2. Compounds 58–62 glycosylated at C-7 position, are mainly existed in *E. bodinieri* [17,18]. Eriodictyol 7-O-(6’-feruloyl)-β-D-glucopyranoside (59) and eriodictyol 7-O-[6”-(3’-hydroxy-4”-methoxy cinnamoyl)]-β-D-glucopyranoside (60) were isolated from *E. bodinieri* as new flavanone glycosides [17]. The feruloyl is attached to the glucosyl C-6 position in compound 59. Compound 60 is an isomer of 59, with -OCH3 and -OH groups at the C-3” and C-4” positions in the 3,4-substituted coumaroyl unit. The substituent positions of -OCH3 and -OH groups are opposite of 59. The distinguishment in 59 and 60 is the same as that in 28 and 29. They were all obtained from the plants *E. bodinieri* [17]. It is worth noting that one -O-CH2-O- group is connected with C-6 and C-7 of ring-A to get a furyl-ring in compound 63.

Additionally, iso-formononetin-4’-O-β-D-glucopyranoside (65), amentoflavone (66), (+)-catechin (67), and (–)-epicatechin (68) were also isolated from *Elsholtzia* [11,19–22].

Compounds 69–73 are linear furanocoumarins, in which 70–73 were found in *E. densa* as new furanocoumarins [11,14,23]. 69–72 exhibit a prenyl group or prenyl derivative in the C-5 position and a methoxy group in the C-8 position.

Only three lignanolides were reported in the genus. They are 3-hydroxyarctiin (75) and arctigenin (76) from *E. eriostachya*, together with saussurenoside (77) in *E. ianthina* [24,25].

Terpenoids

Triterpenoids are other major constituents in this genus. The oleane-type triterpenes (82–89) were mainly isolated from the aerial part of *E. bodinieri* [26–29]. The glycosyl is linked to the C-28 (–COO-) of 23-hydroxyechinocystic acid by ester-bond in compounds 87–89. The C-3 position is attached a caffeoyl in compound 87, and linked to an arabinosyl in 88 and 89, respectively. Three ursane-type triterpenes including ursolic acid (90), corosolic acid (91) and 2α,3β,19α-trihydroxyurs-12-en-28-oic acid (92), are obtained from *E. rugulosa*, *E. ciliata* and *E. bodinieri* [17,30–32].

Two unusual 18,19-secoursane glycopyranosides, bodiniosides A (93) and B (94), were isolated from the whole plant of *E. bodinieri* [17,27]. It was the first report that E-secoursane glycosides occurred in the Lamiaceae family. In addition, 2,3,19-trihydroxy urs-12-en-28-oic acid (92) and hypadicanic acid (95) were also simultaneously obtained from the *E. bodinieri*. Compounds 93–95 could be derived from 92 in the biogenetic relationships [17].

Compounds 98–102, five diterpenoids, were isolated from *E. bodinieri* [21,27,33,34], Ludongnin 5 (98), a tricyclic kaurane diterpenoid, is connected a γ-lactonic at the C6-C19 positions. 98 also has significant and extensive antibacterial effect [34,35]. Sandaracopimar-15-en-8β,12β-diol (99), a tricyclic pimarane diterpenoid, was
isolated from *Elsholtzia* for the first time [21]. An abietane-type diterpenoid, (+)-hinokiol (100), is a minor diterpenoid occurring in plants. Its consuming inhibitive and deactive effects against *Staphylococcus aureus*, *Streptococcus*, *Escherichia coli*, and *Pseudomonas aeruginosa* attract the researchers [33]. It is the infrequent O-H…π stacking that was found in the packing of the crystal structure of 100 except for the existence of hydrogen bonding, when its molecular configuration and conformation were characterized by X-ray diffraction analysis [36]. Two hardwickii acid glycophydrinosides, 6-hydroxy-(-)-hardwickii acid 2′-O-β-D-glucopyranosylbenzyl ester (101) and 6,7-dihydroxy-(−)-hardwickii acid 2′-O-β-D-glucopyranosylbenzyl ester (102) were firstly reported in *E. bodinieri* as two novel clerodane diterpenoids [27].

Three eudesmane-type sesquiterpene glycopyranosides, dictamnoside G (103), β-D-glucopyranosyl-(1→4)-α-L-rhamnopyranosyl-(1→3)-β-D-glycopyranoside (104) and integirioside A (105) were obtained from the root bark of *E. bodinieri* [37].

So far, only one monoterpenoid, 2,6-dimethyl-8-hydroxy-2,6-octadienic acid-8-β-D-glucoside (106), was obtained from *E. bodinieri* as a new one [18].

**Others**

Up to now, only three compounds (107–109) containing nitrogen atoms were reported from the genus, all from *E. rugulosa*. Prunasin (107) and amygdalin (108) are cyanogenic glycosides [30,38,39]. This was the second report of cyanogenic glycosides in Lamiaceae plants. The first case was from an Australian plant *Clerodendrum gravi*.* Since Armeniacae semen* (apricot kernel) containing these compounds, it has been used for cough remedies in Europe and China. The result chemically supported the use of *E. rugulosa* for the treatment of colds and coughs in China. Three maltol glycosides, maltol 3-O-β-D-glucopyranoside (110), maltol 6′-O-β-D-apiofuranosyl-β-D-glucopyranoside (111) and maltol 6′-O-(5-O-p-coumaroyl)-β-D-apiofuranosyl-β-D-glucopyranoside (112) were isolated from *E. rugulosa* [38]. Here, 112 linked with a cinnamoyl can also be classified in the C6-C3 group. Besides, four phytosterols, 113–116, were reported from the genus [11,20,22,25,27,30,32,39,40], and a stilbene’s hydroxylated derivative, trans-3,4,3′,5′-tetrahydroxy-4′-methyl-stilbene 4-O-β-D-xylopyranosyl-(1→6)-β-D-glucopyranoside (130), was isolated from the root bark of *E. bodinieri*, as a new compound [33].

**Volatile chemical constituents**

The plants *Elsholtzia* are aromatic herbs in general, as they possess plentiful volatile oils. The oils have been developed and utilized as medicines, food and the source of honey manufacture [1]. Many phytochemistry and pharmacology scientists are interested in the volatile constituents and its biological activities. The latest paper reported that the volatile constituents exert strong inhibition against central nervous system and take on considerable analgesic effect [41]. It also shows antibacterial effects [42-44].

A total of 572 volatile constituents were identified from the 21 species of *Elsholtzia* by hydro-distillation and gas chromatographic-mass spectrometry (GC-MS). Among them, α-pinene, β-pinene, acetophenone, caryophyllene oxide, carvacrol, benzaldehyde, β-caryophyllene, 1,8-cineole, α-phellandrene, and α-terpineol widely exist. Especially, α-pinene and β-pinene are most significant two, which were detected and identified in 15 species in the genus [45-66]. Acetophenone, caryophyllene oxide, carvacrol, benzaldehyde, β-caryophyllene, α-phellandrene, and α-terpineol are also major examples. It is a remarkable matter that the plant source, growing environment, harvesting time, extraction methods, and analysis methods of the study plants play important impact on the sorts and contents of some volatile components [45,67-70]. For instance, the content of α-pinene in *E. blanda* was up to 4.84% in Yunnan province, and decreased to 1.43% in Sichuan province, China [46,47]. A paper illustrated that the content and sort of volatile components from *E. stauntonii* were obviously contrasted with different extraction methods, respectively [67,69]. The researches focusing on the volatile components from *E. splendens*, *E. bodinieri*, *E. stauntonii*, and *E. ciliate* are much more than on other species [45,51,52,55,56,61-63,67-78]. The identified volatile components from the genus are shown in Additional file 4: Table S3, associated with the corresponding plant sources.

**Pharmacological activities**

Pharmacological investigations on the extracts and pure compounds from *Elsholtzia* cover the activities of anti-viral, antibacterial, anti-inflammatory, anti-oxidant, and myocardial ischemia protection, as well as other activities. Researchers are increasingly concerning on the pharmacological activities of the genus.

**Antiviral activity**

Apigenin (12), apiin (15), luteolin (16), galuteolin (20), luteolin 3′-glucuronyl, methyl ester (30), and the ethyl acetate extract of *E. rugulosa* were reported to exhibit remarkable inhibition against the neuraminidases (Nas) from three typical influenza viruses A/PR/8/34 (H1N1), A/Jinan/15/90 (H3N2) and B/Jiangsu/10/2003. They inhibited influenza NAs at the different half maximal inhibitory concentration (IC50) values ranging from 7.81 μg/mL to 28.49 μg/mL. Especially, 12 and 16 exhibit significant effect against H3N2, with its IC50 values of
1.43 and 2.06 μg/mL, respectively. And the antiviral ability of 12 is 3 times higher than positive control, ribavirin. 16 has a similar capacity of antiviral activity with 12 [79]. Many Elsholtzia species, such as E. bodinieri and E. bland a contain rich luteolin (16) and its derivatives (16–23) [80-82]. Its high content, such as up to 16.0 mg/g in the leaves of E. bland a, provide a convenient for the development of antiviral activity [80].

Essential oil from E. densa showed a significant inhibitory properties against Asia influenza virus A and Orphan virus in vitro. And it can postpone the symptom appearance by 72–96 h after being infected by virus in vivo. Also, it showed inhibition against H3N2 subtype of influenza A virus, and exhibited remarkable therapeutic effect on mouse pulmonic induced by influenza virus when the mouse was administered with the essential oil (100 mg/kg) [41].

Antibacterial activity
Luteolin (16), quercetin (33) and ludongnin 5 (98) were isolated from the roots of E. bodinieri [9,21,26,34]. An antimicrobial assays indicated that these compounds had the inhibitory and bactericidal activities against S. aureus, Bacillus subtilis and E. coli in varying degrees. The minimal inhibitory concentrations (MIC) of 98 are 5, 10 and 80 μg/mL, respectively. The MIC values of 16 against E. coli and S. aureus 50 and 40 μg/mL and of 33 against S. aureus and B. subtilis with its MIC values of 60 and 90 μg/mL, respectively [34].

The ethanol extracts of E. bland a and E. rugulosa exhibited remarkable inhibitory activity against methicillin-resistant S. aureus, with its MIC values of 1.32 and 1.43 mg/mL, respectively [83].

Besides, some Elsholtzia essential oils also showed antimicrobial activity against bacteria, i.e. E. coli, Shigell a flexneri, S. epidermidis, beta Streptococcus, Bacterium paratyphosum B, B. typhi murium, B. dysenteriae, B. diphtheriae, B. meningitidis purulentae, B. proteus, Allthrax bacillus, and Neisseria intracellularis [41].

Essential oils from E. splendens are inhibitory against S. aureus, P. acnes and S. epidermidis. Its MIC against P. acnes was 0.31 μL/mL. As we known, P. acnes and S. epidermidis are involved in the formation acne, thus the inhibition against the two bacteria supports the considerable potential of the E. splendens essential oil for the treatment of acne [42]. It was also reported that volatile components from E. ciliata and E. rugulosa inhibit common bacteria, such as S. aureus, P. aeruginosa, B. enteritidis, B. subtilis, Proteus vulgaris, Shigella dysenteriae, and E. coli [43,44].

Anti-inflammatory
The 75% ethanol extract of the aerial part of E. splendens can significantly inhibit acute inflammation (mouse ear edema by croton oil-inducing) and subchronic inflammation (ear edema by phorbol ester inducing). E. splendens significantly inhibited PGE2 production by pre-induced cyclooxygenase-2 of lipopolysaccharide-treated RAW 264.7 cells. It was thus believed that inhibition against cyclooxygenase-2 is probably one of the function mechanisms [84].

On chemical view, luteolin (16), a widely contained flavone with many hydroxyl substitutions, is a bioactive constituent in Elsholtzia plants for anti-inflammatory activity. It can inhibit the production of NO and generation of other inflammatory cytokine, such as TNF-α, IL-1β, IL-6, NF-κB, etc. [41]. Therefore, rich production of hydroxylated flavone and/or its derivatives is one of reasons to some extend to explain the anti-inflammatory action for Elsholtzia plants.

Anti-oxidant activity
The extracts of the aerial parts of E. rugulosa and E. bodinieri displayed significant anti-oxidant activity in a radical-scavenged assay, which perhaps can elucidate why E. rugulosa has an anti-aging effect [85,86]. The flower of E. rugulosa is rich in flavonoids with its content up to 0.2352 mg/mL, and the flavonoids significantly scavenge the OH− and O2− ions, with its scavenging rate 30.8 and 40.5%, respectively [87]. The extract and extract-loaded nanoparticles of the flower of E. splendens showed a concentration-dependent manner in DPPH radical scavenging assay. E. splendens was also found to activate the antioxidant defense system against 7,12-dimethyl-benz(a)anthracene (DMBA)-induced oxidative stress and reduce several biomarkers of oxidative stress such as thio-barbituric acid reactive substance, protein carbonyls, serum 8-hydroxy-20-deoxyguanosine, and ovary CHO-K1 cells aging [88-92]. The maximum LPO inhibition ratio of the flavonoid extracts of E. bland a was 70.8% with the IC50 0.23 mg/L. And the inhibition of LPO was induced by OH free radical [93]. The genus Elsholtzia owns rich polyphenols, thus having radical scavenging effects [94,95].

Myocardial ischemia protection
The total flavones from E. bland a (TFEB) could improve the recovery of myocardial function, and keep heart from ischemic damage due to coronary occlusion in Beagle dogs. The effect was achieved by the inhibition of serum creatine kinase-MB (CK-MB) and malondialdehyde (MDA), together with by the lowering of mean arterial pressure (MAP), coronary vascular resistance (CVR), etc. [96]. The TFEB not only could reduce infarct size during acute myocardial infarction (AMI) by inhibiting myocardial apoptosis through modulation of Bcl-2 family [97], but also might decrease the myocardial ischemia and ‘Xiongbi Symtom’ [98]. Luteolin 7-O-β-D-
exhibited certain effect on depressing blood-lipid by re-
cells (CMECs) against amyloid-
12 effects against the Alzheimer’s disease (AD) in cell mod-
es. Additionally, Acanthopanax senticosus ese Traditional Compound Medicines consisting of [105]. The extracts of luteolin (35) -induced toxicity. Endothelial cells of cerebral capil-
16 lowering low-density lipoprotein (LDL)-cholesterol E
12 other activities. It is notable that apigenin (12) and luteolin (16) from E. rugulosa displayed protecting effects against the Alzheimer’s disease (AD) in cell models. 12 protects rat cerebral microvascular endothelial cells (CMECs) against amyloid-β peptide 25–35 (Aβ25–
35) -induced toxicity. Endothelial cells of cerebral capil-
aries forming the blood–brain barrier play an important role in the pathogenesis and therapy of AD. Aβ 25–35 showed toxicity on CMECs, and broke the barrier in-
tegrity and function [100]. Copper can trigger the neuro-
toxicity in amyloid precursor protein Swedish (APPsw) overexpressing cells, which exacerbated the amyloid-β (Aβ) neurotoxicity and can be taken as a model of AD. Luteolin (16) treatment exerted neuroprotection through mechanisms that decrease amyloid-β precursor protein (AβPP) expression, lower Aβ secretion, regulate the redox imbalance, preserve mitochondrial function, and depress caspase family-related apoptosis [101]. E. splendens exhibited significant analgesic activity against mouse acetic acid-induced writhing. And the inhibition is up to 50% at 400 mg/kg [84].
A study indicated that E. splendens could obviously re-
lieve symptoms of premenstrual syndrome. And the scores of depression and anxiety and the premenstrual instability decreased significantly [102]. The extracts of E. splendens had the potential of inducing structural ab-
erration of chromosome. The ethanolic extracts of E. splendens have been found for reducing blood lipid by lowing low-density lipoprotein (LDL)-cholesterol [103,104]. And ’Ciwujia Xiangru Decoction’, one of Chin-
ese Traditional Compound Medicines consisting of Acanthopanax senticosus and E. splendens, has been investigated the mentioned effect and used in clinic [105]. The extracts of E. splendens and E. stauntonii had fungitoxic activity, and may be potential fumigants for integrated pest management programs of stored-grain insect [106,107]. Additionally, E. bodinieri extracts exhibited certain effect on depressing blood-lipid by re-
ducing the level of total cholesterol of rats [108].

Conclusion remarks
The review paper summarized a total of 144 compounds and abundant volatile components that were reported from the genus Elsholtzia, with 117 references cited. We noted that Elsholtzia has an extensive distribution, di-
verse biological and pharmacological activities of pure
compounds, extracts and volatile components described. Previous phytochemical researches on the genus revealed the extensive presence of flavone, coumarin, terpenoid, and other compound types, together with prolific essential oils. The pharmacological activities of volatile constituents mainly were regarded on antiox-
dant, antiviral and antibacterial activities.

From the review, it can be seen that phytochemical investigations mainly focus on 10 Elsholtzia species, E. blanda, E. bodinieri, E. ciliata, E. cristata, E. densa, E. eriostachya, E. ianthina, E. rugulosa, E. splendens, and E. stauntonii. And the volatile constituents’ analyses pri-
marily concentrated on 20 species. However, related chemical and biological toward other Elsholtzia species, including E. kachinensis, E. capituligera, E. cephalantha, E. cypriani, E. eriocalyx, E. flava, E. glabra, E. hetero-
phylla, E. hunannensis, E. kachinensis, E. luteola, E. ochroleuca, E. oldhamii, E. penduliflora, E. pilosa, E. pyg-
maea, E. saxatilis, E. souliei, E. stachyodes, and E. winiti-
ana, are still blank. So, plenty of further studies are necessary in order to illustrate the chemo-diversity and to make full use of the biological significance of the compounds and extracts of Elsholtzia, especially the antiviral and anti-inflammatory activities. The authors wish the review can provide a valuable data for explora-
tions and advanced researches of Elsholtzia species.

Additional files

Additional file 1: Table S1. The list of Elsholtzia species [1,2].
Additional file 2: Table S2. The name, plant source of compounds 1–
144 from Elsholtzia [9-33,35,38-42,106,109-114].
Additional file 3: Figure S1. The structures of compounds 1–132 from
Elsholtzia.
Additional file 4: Table S3. The volatile chemical components from
Elsholtzia [81,43,44,47-49,51-79,106,114-118].

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
GZ, LZ, WX, and SG have been involved in preparing the manuscript. LW, JR
and CR participated in the discussion of views in the paper. All authors have
read and approved the final manuscript.

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