EFFECTS OF CYCLIC-NUCLEOTIDE DERIVATIVES ON THE GROWTH OF HUMAN COLONIC CARCINOMA XENOGRAFTS AND ON CELL PRODUCTION IN THE RAT COLONIC CRYPT EPITHELIUM

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Summary.—Previous studies have shown that various amine hormones are able to influence the growth rate of human colorectal carcinomas propagated as xenografts in immune-deprived mice, and it is now well known that the effects of many amine and other hormones are mediated by cyclic nucleotides, acting as second messengers within cells. In the present study the influence of various derivatives of cyclic adenosine monophosphate and cyclic guanosine monophosphate on the growth of two different lines of colorectal cancer growing in immune-deprived mice, and on the cell production rate in the colonic crypt epithelium of the rat, was assessed. Growth of each tumour line, as well as crypt-cell production, was suppressed by treatment with N\(^6\)O\(^6\) dibutyryl and N\(^6\) monobutyryl derivatives of cyclic adenosine monophosphate. Dibutyryl cyclic guanosine monophosphate, on the other hand, was found to promote the growth of Tumour HKX4 and to promote crypt cell production, but to have no significant effect on Tumour HXM2.

There is mounting evidence that cell proliferation in the colonic crypt epithelium and in colonic tumours is influenced by amine hormones. Adrenaline and other beta-adrenergic agonists have been shown to inhibit cell division in dimethylhydrazine-induced tumours of rat colon (Tutton & Barkla, 1977a), whereas histamine (acting via histamine-H\(_2\) receptors) and serotonin both promote cell division in these tumours (Tutton & Barkla, 1978a, b; 1980a). By contrast noradrenaline, acting via an \(\alpha\)-adrenergic receptor, promotes cell production in the colonic crypt epithelium (Tutton & Barkla, 1977a; Tutton & Steel, 1979) have shown that the growth rate of human colorectal tumours propagated as xenografts in immune-deprived mice is also influenced by amine hormones. Growth of one tumour line, HKX4, a moderately to well differentiated colonic adenocarcinoma (Nowak et al., 1978) was inhibited by the histamine-H\(_2\)-receptor antagonist, cimetidine, whereas another colonic tumour, HKX7 (a moderately to poorly differentiated colonic adenocarcinoma) was inhibited by an anti-serotoninergic drug, BW501c. The growth of Tumour HKX4 was also inhibited by adrenaline acting through a \(\beta\)-adrenoceptor.

Cyclic nucleotides have been implicated as intracellular mediators of the effects of many hormones, including the biogenic amines adrenaline, noradrenaline, histamine and serotonin. The catecholamines, such as adrenaline and noradrenaline, have been shown to act via receptors which were originally classified into 2 groups, \(\alpha\) and \(\beta\) (Ahlquist, 1948) and which have subsequently been sub-classified into 4 groups, \(\alpha_1\) and \(\alpha_2\) (Berthelson & Pettenger, 1977), and \(\beta_1\) and \(\beta_2\) (Lands et al., 1967). The response to \(\alpha_1\) receptors is mediated via calcium ions, whereas that of alpha2 receptors is mediated by inhibi-

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tion of adenylate cyclases and consequent lowering of intracellular cyclic AMP level (Fain & Garcia-Sainz, 1980). The response to both types of β adrenoceptors is mediated by activation of adenylate cyclase and consequent elevation of cellular cyclic AMP levels (Maguire et al., 1977). The cellular responses to histamine are mediated by receptors which have been termed H1 and H2 (Ash & Schild, 1966; Black et al., 1972) with H1-receptor effects being mediated by calcium ions and H2-receptor effects being mediated by cyclic AMP (Fain & Garcia-Sainz, 1980). The effects of serotonin have been attributed to raised cyclic AMP levels in some tissues (Enjalbert et al., 1978) and to raised cyclic GMP levels in other tissues (Goldberg & Haddock, 1977).

The effects of amine hormones on xenograft growth are not readily explained in terms of cyclic nucleotide actions, for two reasons. First, in Tumour Line HXK4, adrenaline (acting through a β-adrenoceptor and hence presumably cAMP-linked) and histamine (acting through a histamine-H2 receptor, also presumably cAMP-linked) have opposing actions on tumour growth rate. Secondly, growth appears to be promoted in one tumour line (HXK4) by a cAMP-linked agent (histamine via H2 receptors) and in the other line (HXK7) by an agent (serotonin) which is possibly a cGMP-linked agent. In an attempt to resolve these apparent inconsistencies, the effect of various derivatives of cAMP and cGMP on the growth of a histamine-dependent line and a serotonin-dependent line of colonic tumours, and on cell production in the colonic crypt epithelium, was examined.

**MATERIALS AND METHODS**

*Estimation of the mitotic rate in the colonic crypt epithelium.*—Male Sprague–Dawley rats were fed Clark No. 2 pellets, tap water *ad libitum*, and housed at 21–24°C with artificial light from 07:00 to 21:00 and darkness from 21:00 to 07:00. Animals were given vinblastine sulphate (Velbe, Elly Lilly and Co., Indianapolis, U.S.A.) at a dose of 4 mg/kg by i.p. injection at 12:00 and were killed by decapitation between 12:45 and 16:00. This dose of vinblastine was found, in preliminary experiments, to be the lowest that provides reliable metaphase arrest in the colonic crypts. Counts of metaphase and non-metaphase cells in histological sections in crypts of Lieberkuhn of the descending colon were made as previously described (Tutton & Barkla, 1976). All metaphase indices were then corrected for the differences in size between metaphase and non-metaphase nuclei and for the geometric artefact described by Tannock (1967). Details of these correction factors are described elsewhere.

Graphs of corrected metaphase index vs duration of vinblastine treatment were then constructed for each experimental group of tissues with mitoses blocked for 0:75–4 h. The least-squares estimate of the regression coefficient for each of the graphs was then calculated; this value represents the rate at which cells enter metaphase and has the units of mitoses/cell/h. The statistical significance of apparent differences between regression coefficients for different experimental groups of tissue was then estimated by analysis of variance (Bliss, 1967).

Initially, cell proliferation was studied in the colonic crypts of 14 normal rats.

Additional groups of 6 rats were treated with N6O2 dibutyryl, N6 monobutyryl, O2 monobutyryl and 8-bromo cyclic AMP or N6O2 dibutyryl cyclic GMP, each at a dose of 2 mg/kg injected i.p. All cyclic-nucleotide derivatives were purchased from the Sigma Chemical Company, St Louis, U.S.A. A further group of 6 rats was treated with sodium butyrate at a dose of 7 mg/kg. This dose was used since it represents the amount of butyrate moiety that was contained in the highest dose of dibutyryl cAMP.

**Xenograft technique.**—Male and female CBA/lac mice were bred in our laboratory and immune-deprived by a technique similar to that reported by Steel et al. (1978). Mice aged 16–20 days were thymectomized under ketamine anaesthesia (Ketlar, 0-15 mg/kg i.p.). After an interval of 18–21 days the mice were injected with cytosine arabinoside (Cytosar, the Upjohn Company) as a dose of 200 mg/kg i.p. and then subjected, 48 h later, to 8-5 gray of whole-body irradiation from an X-ray source at 180 keV. Pre-treatment with Cytosar obviated the need for marrow reconstitution that was previously necessary.
Mann-Whitney, time relative linearity volume logarithm meter of betweenthe assessment using rank of thecontrol observations were calculated from histamine-dependent, in the sense that its growth in immune-deprived mice was inhibited by administration of the histamine-H₂-receptor antagonist Cimetidine (Totton & Steel, 1979). Tumour Line HXK4 was established by the authors from a surgically resected specimen of descending colon tumour. This tumour was histologically poorly differentiated and assessed as being serotonin-dependent, in the sense that its growth was inhibited by the antiserotoninergic drug BW501c (Totton & Barkla, unpublished).

_Tumour measurement._—Starting on the 24th to 28th day after implantation, tumours were measured every 1 or 2 days. The greatest and least superficial diameters were measured, using vernier calipers, and the xenograft volume calculated by the formula (mean diameter)^3 \times \pi / 6. The volume of each tumour after t days of assessment was divided by the volume of the same tumour at the start of assessment (V₀) to obtain the relative tumour volume (Vₜ/V₀). The mean of the logarithm of this quotient was then plotted as a function of time for each treatment group of mice. The relative tumour volume was calculated because inter-xenograft variations in this parameter arise only during treatment; the logarithm of the quotient was plotted against time because of the previously observed linearity of log volume vs time for xenografts (Lamerton & Steel, 1975). Tumour volumedoubling times for control tumours were calculated from the linear regression of log relative tumour volume vs time. The statistical significance of any apparent difference between the relative volumes of various groups of xenografts at a particular time after the start of treatment was assessed using the Mann–Whitney, non-parametric test for ranked observations (Sokal & Rohlf, 1969). The control group of tumours for Line HXK4 contained 16 xenografts and the control group of tumours for Line HXM2 contained 17 xenografts. Each experimental group consisted of 10–16 xenografts. Mice were treated with N⁰O²'- dibutyryl, N⁶ monoobutyryl, O²' monobutyryl or 8-bromo cyclic AMP at doses of either 2-0 or 20 mg/kg, N⁰O²' dibutyryl cyclic GMP at doses of either 0-2 or 2 mg/kg, or a combination of dibutyryl cyclic AMP (20 mg/kg) and a phosphodiesterase inhibitor, papaverine HCl at a dose of 50 mg/kg (Amer & Keighbaum, 1975). An additional group of mice bearing Tumour HXK4 was treated with sodium butyrate at a dose of 7 mg/kg. All mice were treated twice daily.

**RESULTS**

The influence of various cyclic nucleotide derivatives and of sodium butyrate on the colonic crypt-cell production rate is shown in the Table.

| Treatment          | Dose (mg/kg) | Mitotic rate— | P (vs control) |
|--------------------|--------------|---------------|----------------|
| Nil                |              |               |                |
| Sodium butyrate    | 7-0          | 0-040 ± 0-004 | NS             |
| Dibutryl cGMP      | 2-0          | 0-035 ± 0-006 | NS             |
|                    | 0-2          | 0-063 ± 0-012 | < 0-001        |
| Dibutryl cAMP      | 20           | 0-008 ± 0-004 | < 0-01         |
|                    | 0-2          | 0-008 ± 0-004 | < 0-01         |
| N⁶-monobutyryl cAMP| 2-0          | 0-012 ± 0-002 | < 0-05         |
| O²'-monobutyryl cAMP| 2-0        | 0-019 ± 0-003 | NS             |
| 8-Bromo cAMP       | 2-0          | 0-036 ± 0-007 | NS             |

The control group of tumours for Line HXK4 contained 16 xenografts and the control group of tumours for Line HXM2 contained 17 xenografts. Each experimental group consisted of 10–16 xenografts. Mice were treated with N⁰O²'- dibutyryl, N⁶ monoobutyryl, O²' monobutyryl or 8-bromo cyclic AMP at doses of either 2-0 or 20 mg/kg, N⁰O²' dibutyryl cyclic GMP at doses of either 0-2 or 2 mg/kg, or a combination of dibutyryl cyclic AMP (20 mg/kg) and a phosphodiesterase inhibitor, papaverine HCl at a dose of 50 mg/kg (Amer & Keighbaum, 1975). An additional group of mice bearing Tumour HXK4 was treated with sodium butyrate at a dose of 7 mg/kg. All mice were treated twice daily.

_Growth curves for Tumour HXK4 are illustrated in Figs 1, 2 and 3, and those for HXM2 in Figs 4 and 5. Tumour HXK4 had a mean doubling time of 8-1 days and HXM2 had a mean doubling time of 7-3 days. In each case db cAMP was seen to inhibit xenograft growth, though in the case of HXK4 using a dose of 20 mg/kg, and HXM2 using a dose of 2 mg/kg, this response was short-lived. Of the other cAMP derivatives tested, N⁶ monobutyryl cAMP was of similar effectiveness to db_
cAMP, whereas other derivatives were marginally effective or ineffective. In HXK4 tumours treated with db cAMP and the phosphodiesterase inhibitor, papaverine HCl, growth remained inhibited for 5 days and, in HXM2 tumours treated with db cAMP alone at a dose of 20 mg/kg, growth remained inhibited for 6 days, and the mean tumour volume after 8 days of treatment was similar to that at the beginning of the experiment. Papaverine alone also inhibited the growth of Tumour HXK4, but was less effective than db cAMP alone. Treatment with db cGMP at a dose of 2 mg/kg had a short-term stimulatory effect on Tumour HXK4 and no statistically significant effect on tumour HXM2.
DISCUSSION

The observed inhibitory effect of cAMP derivatives on the growth of both lines of human colonic tumour resembles the effects of this agent on the mitotic rate in rat primary colonic tumours (Tutton & Barkla, 1980c) and the stimulating effect of db cGMP on Tumour Line HXM2 also resembles the effect of this agent on rat tumours (Tutton & Barkla, 1980b). Since the growth of Tumour Line HXK4 is inhibited by cAMP derivatives and promoted by cGMP derivatives, it would appear unlikely that the previously reported growth promoting effect of histamine on this tumour is mediated by cAMP; in fact these results raise the possibility that the effects of histamine-H₂-receptor agonism is mediated by cGMP.

The Yin Yang hypothesis (Goldberg et al., 1974), which proposes that various biological processes, including cell division, are controlled by the molar ratio of cyclic GMP to cyclic AMP, is not, however, uniformly supported by data from our laboratory. We have reported previously that in the jejunal crypt epithelium significant inhibition of cell proliferation by db cAMP was not obtained with doses between 0.2 and 20 mg/kg (Tutton & Barkla, 1980c). It must be admitted that in these experiments direct inhibition by cAMP may have been opposed by indirect stimulation by increased circulating levels of thyroid hormones, which have previously been shown to promote jejunal crypt-cell proliferation (Carriere, 1976; Tutton, 1965). Secretion of glucocorticoids may also be increased by treatment with cAMP derivatives (Perlmutter et al., 1971) but this would not appear to explain the present results, since glucocorticoids themselves appear not to influence colonic crypt-cell proliferation, and to promote rather than inhibit cell proliferation in colonic tumours (Tutton & Barkla, submitted for publication). Clearly, the suggested role of cyclic nucleotides as intracellular mediators for the amine hormones which appear to influence intestinal epithelial-cell proliferation needs to be confirmed using radioimmunoassay and possibly quantitative immunocytochemistry of cyclic nucleotides in amine-treated and control tissues.

If cyclic GMP is a growth stimulant in colonic tumours and a mitogen in both colonic tumours and in the colonic crypt epithelium (Tutton & Barkla, 1980b), has the metabolism of biogenic amines and cyclic nucleotides any relevance to the basic biology of neoplastic change? At present the only optimistic lead to this question lies in the reported differences between particulate (membrane-bound) and soluble (cytoplasmic) forms of guanylate cyclase in various tissues (Kimura & Murad, 1975). Exposure of the colonic epithelium to the carcinogen N-methyl-nitro-nitrosoguanidine results in a shift from particulate to soluble guanylate cyclase (de Rubertis & Craven, 1977). Stimulation of soluble guanylate cyclase by biogenic amines requires the cells to have either an amine-uptake process or an amine-regulated calcium-gating uptake (Fain & Garcia-Sainz, 1980). Evidence for such an amine-uptake process in neo-
plastic but not in normal colonic epithelial cells has been reported; toxic analogues of serotonin, (5,6- and 5,7-dihydroxytryptamine) cause rapid cytoplasmic changes in DMH-induced colonic tumour cells, but not in jejunal crypt cells (Tutton & Barkla, 1977b; 1979). The biological significance of amine uptake by tumour cells remains to be assessed.

Whatever differences exist between the mechanism controlling the synthesis and degradation of cyclic nucleotides in normal and malignant tissues, the fact remains that cellular levels of cGMP are higher, and levels of cAMP are lower in colonic tumours than in colonic mucosa (de Rubertis et al., 1976). In addition, plasma and urinary levels of cyclic GMP are raised in patients with disseminated bowel cancer (Chawla et al., 1979) and pharmacokinetic studies have shown that this increase in plasma and urinary levels of cGMP is due to a substantial increase in its production (Murray et al., 1979). These cyclic-nucleotide assay results, seen in the light of our cell-kinetic results, suggest that cyclic nucleotides may have an exceedingly important role in controlling tumour growth.

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