Temperature-dependent competitive outcomes between the fruit flies

*Drosophila santomea* and *D. yakuba*

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Abstract

Changes in temperature associated with climate change can alter species’ distributions, drive adaptive evolution, and, in some cases, cause extinction. Research has tended to focus on the direct effects of temperature, but changes in temperature can also have indirect effects on populations and species. Here we test whether temperature can indirectly affect the fitness of *Drosophila santomea* and *D. yakuba* by altering interspecific competitive outcomes. We show that, when raised in isolation, both *D. santomea* and *D. yakuba* show maximal performance at temperatures near 22°C. However, when raised together, *D. santomea* outcompetes *D. yakuba* at a lower temperature (18°C), while *D. yakuba* outcompetes *D. santomea* at a higher temperature (25°C). We then use a ‘coexistence’ experiment to show that *D. santomea* is rapidly (within 8 generations) extirpated when maintained with *D. yakuba* at 25°C. By contrast, *D. santomea* remains as (or more) abundant than *D. yakuba* over the course of ~10 generations when maintained at 18°C. Our results provide an example of how the thermal environment can indirectly affect interspecific competitive outcomes and suggest that changes in the competitive advantage of species can lead to some species becoming more prone to extinction by competitive exclusion.

Keywords: local adaptation, competition, climate change, temperature, context-dependent
Introduction

Species vary in their physiological tolerance and behavioral preference for different thermal environments (Calosi et al. 2010; Kellermann et al. 2012). Temperature is therefore an important abiotic factor that can drive local adaptation (McNab 1971; Freckleton et al. 2003; Campbell-Staton et al. 2016, 2018; Stager et al. 2016; Delhey 2017, 2019) and shape species’ ranges (Soberón 2007; Calosi et al. 2010; Early and Sax 2011; Kellermann et al. 2012). However, a species’ range and their response to variation in climate is also shaped by biotic interactions. Competition is one outcome of biotic interactions that, like temperature, can drive phenotypic and ecological divergence (i.e. character displacement (Pfennig and Pfennig 2009; Stuart and Losos 2013)), the maintenance of intraspecific variation (Roughgarden 1972; Bolnick 2001; Harris et al. 2008), speciation (Polechová and Barton 2005; Winkelmann et al. 2014), extinction (Park 1954; Davis et al. 1998a; Alexander et al. 2015), and global biogeographic patterns (Pianka 1966; Willig et al. 2003). The majority of biotic interactions, such as competition, occur across a range of abiotic conditions (either temporally or geographically). To understand how biotic interactions, such as interspecific competition, affect a species abundance and evolution, it is important to understand the outcome of those interactions across different abiotic conditions (e.g. thermal environments (Davis et al. 1998a; Alexander et al. 2015)).

The outcome of interspecific competition has been shown to vary with temperature for a number of species inhabiting different environments. For example, experimental work in communities of algae (Goldman and Ryther 1976; Hillebrand 2011), beetles (Park 1954; Wilson et al. 1984), fungi (Carreiro and Koske 1992), alpine plants (Klanderud and Totland 2007; Alexander et al. 2015), and fruit flies (Davis et al. 1998a, 1998b) have all shown how temperature can indirectly affect a species’ relative abundance through competition. However, direct effects of climate and competition have also been reported: experimental communities of tussock tundra plant communities show no noticeable interaction between temperature and competition (Hobbie et al. 1999). Therefore, while the majority of studies point towards temperature as an important abiotic control on the fitness consequences of interspecific competition, more examples are required to understand its generality.

One aspect of temperature-mediated competitive outcomes that remains underexplored is its prevalence in closely related (e.g. sibling) species. Species that have diverged recently and come into secondary contact are likely to share more aspects of their ecology than distantly-
related species, potentially leading to strong competition (Schluter and McPhail 1992; Grant and
Grant 2006). Species pairs that display overlapping or adjacent (parapatric) geographic ranges
therefore provide useful systems in which to test for variable outcomes of interspecific
competition, especially when those ranges occur along an environmental gradient. The sibling
species *Drosophila santomea* and *D. yakuba* provide one such system. *D. santomea* is endemic
to the cool tropical forest of the island of São Tomé in the Gulf of Guinea. In laboratory
experiments adult *D. santomea* show a behavioural preference for moderate temperatures (22°C)
and larval survival and egg hatchability drops at temperatures above 25°C (Matute et al. 2009).
*D. yakuba*, on the other hand, is a generalist species that has a broad distribution across sub
saharan Africa and is regularly found in association with human-modified habitats (Lachaise et
al. 1988; Cooper et al. 2018). In nature, populations of *D. yakuba* therefore experience
environments subject to a much wider range of temperatures than populations of *D. santomea*,
and *D. yakuba* can tolerate an overlapping but broader range of temperatures in the lab (Matute
et al. 2009; Cooper et al. 2018). On the island of São Tomé the distributions of *D. santomea* and
*D. yakuba* are adjacent, with *D. santomea* typically occurring at forested habitats above 800 m
and *D. yakuba* at lower elevation open habitats (Llopart et al. 2005a; Comeault et al. 2016). The
two species form a narrow hybrid zone between 800 and 900m elevation that has been stable for
over 20 years and occurs as lowland agricultural fields give way to upland rainforest habitats
(Lachaise et al. 2000; Llopart et al. 2005b; Matute 2010a; Comeault et al. 2016). The fact that *D.
santomea* and *D. yakuba* show differences in their realized niche in nature, yet display broadly
overlapping thermal tolerances in the lab raises the question of how interspecific competition
may contribute to the maintenance of their distinct ecological niches.

Here we test how the outcome of competition between *D. santomea* and *D. yakuba* varies
across an ecologically relevant range of thermal environments. First, we study the distribution of
*D. santomea* and *D. yakuba* on the island of São Tomé and find that temperature and seasonality
differ between areas where these two species are found. We then manipulate temperature and the
opportunity for competition in the lab to show that both species display maximal performance at
moderate temperatures, but that at low temperatures *D. santomea* is able to outcompete *D.
yakuba*, while the opposite is true at warmer temperatures. We then use a multigenerational
‘coexistence’ experiment to show that the relative abundances of *D. santomea* and *D. yakuba* are
strongly affected by the thermal environment: at low temperatures, *D. santomea* and *D. yakuba*
coexist, while at higher temperatures *D. yakuba* rapidly drives *D. santomea* to extinction. Our results show that the interaction between temperature and competition helps shape ecological differences between these two species; and, disturbingly, suggest that with warming temperatures, *D. yakuba* will be able to outcompete *D. santomea*, potentially contributing to extinction of this island endemic species.

**Materials and Methods**

*The thermal niche of D. santomea and D. yakuba on São Tomé*

We first qualitatively describe the thermal niche of *D. santomea* and *D. yakuba* found on the island of São Tomé using a previously published dataset of occurrence records for both species (Comeault et al. 2016) and climate data from the Worldclim database (bioclim variables 1-11; Table S1; [http://www.worldclim.org/bioclim](http://www.worldclim.org/bioclim)). We extracted values for bioclim variables 1-11 at a resolution of 2.5 arc-degrees, for each site, using the *raster* R library (Hijmans et al. 2019). Because the transect on São Tomé covers a relatively small geographic area - it spans only four unique sets of bioclim variables - we report the range of (1) mean annual temperatures (bioclim variable 1), (2) isothermality (bioclim variable 3: mean diurnal range of temperatures / annual range of temperatures), and (3) temperature seasonality (bioclim variable 4) for sites where either *D. santomea* or *D. yakuba* is the more abundant species, based on relative abundances reported in Figure S1 of Comeault et al. 2016, rather than summarize climatic variation using a typical decomposition-based method (e.g. principal components analysis).

These three variables were chosen because mean annual temperature was strongly correlated with all ‘thermal’ bioclim variables ($r > 0.9$) except isothermality ($r = 0.11$) and seasonality ($r = 0.36$) and isothermality and seasonality showed more modest correlations with all other thermal bioclim variables (minimum $r = 0.09$, maximum $r = 0.70$; mean $r = 0.26$).

**Details of populations used for experiments**

To measure performance under different temperatures and competitive environments we used individuals from genetically diverse laboratory populations of *D. santomea* and *D. yakuba*. We generated these populations by combining 5 male and 5 female offspring from each of 20 isofemale lines established from inseminated females collected on São Tomé. Females were collected between 1 and 14 February 2015 at the sites “lake7” (for *D. santomea*) and “monte7”
(for D. yakuba) and the synthetic populations were created on 12 and 19 March 2015 after ~2-4
generations in the lab. The two resulting synthetic populations (D. santomea: “san_lake_7S” and
D. yakuba: “yak_monte_7S”) were maintained at large population sizes spread over three to five
175 ml polypropylene bottles (Genesee Scientific, Morrisville, NC) for between 5 and 10
overlapping generations before initiating experiments.

Temperature’s effect on competition between D. santomea and D. yakuba

We tested the relative performance of D. santomea and D. yakuba at each of three
biologically relevant temperatures (18°C, 22°C, and 25°C) when maintained in isolation or
together. To initiate this experiment, we placed 6 one to 9 day old female flies from the stock
populations into individual 30 mL vials containing standard corn-starch medium. Sampling
females from stock populations in this way results in >95% of the females being inseminated and
actively laying viable eggs (see Supporting Information, Appendix I). To quantify performance
in the absence of interspecific competition, 6 females of either D. santomea or D. yakuba were
added to the vials. To quantify the effect of interspecific competition, three females of each
species were added to the same vial. Therefore the total number of laying females remained
constant between the “isolation” and “competition” treatments and adult flies and larvae
experienced intraspecific competition only in the former treatment, while they experienced both
intra and interspecific competition in the latter. We created a total of 30 replicate vials containing
only D. santomea and only D. yakuba, and 30 “competition” replicates which contained both
species. We then randomly assigned 10 replicates of each of the three resulting treatments - D.
santomea in isolation, D. yakuba in isolation, or competition - to each of the three temperature
treatments. Females were then allowed to lay eggs for 7 days and then removed from the vials.
When removing the females we added a dampened (0.5% propionic acid) Kimwipe (Kimberly-
Clark, catalog number: 34155) as a pupation site and to inhibit the growth of fungi. As a measure
of performance across temperatures and in different competitive environments we counted the
total number of each sex of adult fly that ecolsed from each vial over the following 23 days.

While we counted the total number of flies, we focus analyses below on male counts because
male D. santomea and D. yakuba are easily and unambiguously identified based on
pigmentation: D. santomea males are a solid yellow and D. yakuba males have black
pigmentation on the last three tergites of their abdomen (females show some variation in color).
Despite potential for misclassification of females, we found no difference in sex ratio across treatments or temperatures (binomial GLM; Figure S1) and summarize results from analyzing all offspring in the Results and Figure S2. We stopped counting emerging flies after 23 days because this duration spanned peak eclosion at all temperatures (Figure S3).

To test whether temperature affected the outcome of competition between \textit{D. santomea} and \textit{D. yakuba} we first modeled the mean (per-female) number of male flies that eclosed from an experimental replicate as a function of temperature, competition treatment (three levels: \textit{D. santomea}, \textit{D. yakuba}, or interspecific competition), and the interaction between temperature and competition. This model was fit using the \textit{glm} function in R (Core Team 2017) assuming Poisson distributed error. To determine if there was a significant interaction between temperature and competition treatment, we conducted a likelihood ratio test (LRT) that compared the model described above to one that lacked the interaction term using the \textit{anova} function in R. Because this analysis identified a significant interaction between temperature and competition (see Results) on the mean number of male offspring produced, we also fit independent generalized linear models, splitting the data by temperature treatment. We then used Tukey’s post-hoc tests, as implemented with the \textit{glht} function in the \textit{multcomp} R package (Hothorn et al. 2008), to test for significant pairwise differences in performance between competition treatments, for each temperature treatment. Finally, for replicates that contained both \textit{D. santomea} and \textit{D. yakuba} (i.e. the “competition” treatment) we tested for differences in performance between these species, at different temperatures, by conducting dependent-samples sign-tests (these data are naturally paired by replicate) as implemented by the \textit{SIGN.test} function in the \textit{bsda} R package (Arnholt and Evans 2017). Because \textit{D. santomea} on the island of São Tomé are found in cooler environments than \textit{D. yakuba}, we predicted that \textit{D. santomea} would outcompete \textit{D. yakuba} at 18°C and \textit{D. yakuba} would outcompete \textit{D. santomea} at 22°C and 25°C.

\textit{Competition during experimental sympatry at different temperatures}

We found a significant interaction between temperature and competition treatments in the single-generation experiment described above. Building on this result, we next tested whether competition between \textit{D. santomea} and \textit{D. yakuba} at different temperatures could lead to one species competitively displacing the other. To test this, we created two-species experimental ‘communities’ in mesh cages (24.5 cm x 24.5 cm x 24.5 cm; \textit{www.bugdorm.com}) and
maintained them at either 18°C or 25°C (four replicates at each temperature). Each cage was prepared by adding ~1 inch of dampened coconut fiber as a substrate to help maintain a relative humidity between ~40 and 80%. We then added two 175 ml polypropylene bottles (Genesee Scientific, Morrisville, NC) containing ~30ml of standard cornmeal medium and a damped kimwipe to each cage along with 24 males and 24 females of both *D. santomea* and *D. yakuba* (96 flies founded each experimental community). The flies used to initiate this experiment were collected 14 hours after eclosion and were 3 days old when introduced to the cages. Every two weeks, we added two fresh bottles containing cornmeal medium and bottles were removed after they were in the cages for at least four weeks (see Table S2 for details). We then randomly sampled flies from each experimental cage at 34 days, 57 days, 90 days, 113 days, and 148 days after initiating the experiment. 148 days is ~10 generations at 18°C and ~12 generations at 25°C for both of these species. To test for consistent changes in the relative abundance of the two species within each cage we conducted Mann-Kendall Trend Tests on the proportion of *D. santomea* within each cage. We expect a significant (and consistent) change in the proportion of *D. santomea* within each cage if it was either at a competitive advantage (increase) or disadvantage (decrease) over *D. yakuba*.

**Figure 1.** Mean annual temperature (A) and temperature seasonality (B) and mean annual precipitation (C) on the island of São Tomé. Black line outlines the coast of São Tomé, white space is the Atlantic Ocean. Filled black points denote sites where *D. yakuba* is the more abundant species and open black circles represent sites where *D. santomea* was more abundant. SD = standard deviation.

**Results**
We found that along the altitudinal transect on São Tomé, D. santomea is more abundant than D. yakuba at cooler sites, where mean annual temperatures range from 17.8 to 20.6ºC and seasonality (in units of standard deviations in temperature) ranges from 8.24 to 8.29 (Figure 1). By contrast, D. yakuba is more abundant at sites experiencing higher mean annual temperatures (20.6 to 25.5ºC) and stronger seasonality (8.27 to 8.35; Figure 1). Isothermality – measured as the mean diurnal range over the annual temperature range – was effectively invariable across the sampled sites (range = 0.70 to 0.71). These qualitative patterns are consistent with previously published results that suggest that D. santomea has a behavioral preference for cooler temperatures than D. yakuba (Matute et al. 2009).

Temperature’s effect on competition between D. santomea and D. yakuba

When raised in isolation, performance, as measured as the number of emergent male offspring, varied with temperature for both D. santomea and D. yakuba (GLMs: D. santomea: $X^2 = 14.26; P = 0.0008$; D. yakuba: $X^2 = 8.54; P = 0.014$). Drosophila santomea produced significantly more male offspring when maintained at 22ºC (mean = 8.1; S.D. = 0.5) compared to when they were maintained at 18ºC (mean = 4.8; S.D. = 0.3) (Tukey’s HSD: $Z = 2.5; P = 0.03$) or 25ºC (mean = 4.5; S.D. = 0.3) (Tukey’s HSD: $Z = 3.6; P = 0.001$). Drosophila yakuba also produced significantly more male offspring when maintained at 22ºC (mean = 8.0; S.D. = 0.5) compared to 25ºC (mean = 5.1; S.D. = 0.3) (Tukey’s HSD: $Z = 2.9; P = 0.012$), but there was not a significant difference (Tukey’s HSD: $P > 0.1$) in performance between either 22ºC and 18ºC (mean = 5.9; S.D. = 0.4) or 25ºC and 18ºC. These results suggest that both D. santomea and D. yakuba perform best at temperatures in the low 20’s (ºC), consistent with previous results (Matute et al. 2009).
Figure 2. The effect of temperature and competition on performance. A) The mean number of offspring per female when *D. santomea* (*san*) and *D. yakuba* (*yak*) were raised with the opportunity for intraspecific competition only (“isolation”) or with both intra and interspecific competition (“competition”). Groups that showed a significant difference in the mean number of offspring produced are indicated by horizontal bars (Tukey’s pairwise contracts; generalized linear models run separately for each temperature). When maintained at 18°C and in competition with *D. santomea, D. yakuba* showed a significant reduction in fitness (leftmost panel of A), while the opposite is true at 25°C (rightmost panels). At 22°C, there is a large variance in the number of offspring that emerged and we did not detect a difference between *D. santomea* and *D. yakuba*. B) Competition data presented in A), but as paired data. Lines connect the number of male *D. santomea* and *D. yakuba* for each of 10 replicates conducted at each temperature.
In addition to temperature, the interaction between temperature and competitive environment had a strong effect on performance (LRT: $X^2 = -28.69; P < 1.0 \times 10^{-4}$; Figure 2A). When maintained together (i.e., with the opportunity for interspecific competition) at 18°C, *D. santomea* had higher performance than *D. yakuba* (Tukey’s HSD: $Z = 3.09; P = 0.01$) and *D. yakuba* performed worse than when maintained in isolation (Tukey’s HSD: $Z = 2.65; P = 0.04$) (Figure 2A). When maintained together at 25°C, *D. yakuba* outperformed *D. santomea* (Tukey’s HSD: $Z = 3.55; P = 0.002$), but there was not a significant difference in *D. santomea*’s performance when maintained in isolation or together with *D. yakuba* at 25°C (Tukey’s HSD: $Z = 1.42; P = 0.48$). At 22°C, there was no significant effect of competitive environment on per-female performance in either *D. santomea* or *D. yakuba* (Tukey’s HSD: all $P > 0.05$). We report results from parallel analyses including both female and male offspring in Appendix II and Figure S1 (Supporting Information). The only difference we observed when analyzing total offspring (i.e. male and female offspring both included) was that *D. yakuba* that experience interspecific competition at 22°C produce fewer offspring than when maintained in the absence of interspecific competition at 22°C (Tukey’s HSD: $Z = -2.724; P = 0.0325$; Figure S2).

When treating performance of *D. santomea* and *D. yakuba* within each replicate of the interspecific competition experiment as paired, *D. santomea* tended to produce more offspring than *D. yakuba* at the low temperature (18°C), resulting in a marginally significant effect ($S = 8, P = 0.055$; Figure 2B, left panel), *D. yakuba* produced more offspring than *D. santomea* at the high temperature (22°C; $S = 0, P = 0.001$; Figure 2B, right panel), and there was no consistent difference in the number of offspring produced by either species at 22°C ($S = 9, P = 0.828$; Figure 2B, middle panel).

**Competition during experimental sympatry at different temperatures**

Consistent with the results we observed over the course of a single generation, when maintained together at 25°C the proportion of *D. santomea* monotonically decreased over the course of the experiment ($S = -9, -9, and -12; P = 0.036, 0.036, and 0.013$, respectively, for the three cages that maintained viable populations over the entire course of the experiment). Under our experimental conditions, *D. santomea* went extinct in three of the four cages maintained at 25°C after 57 days (4-5 generations; in the fourth cage only one of 67 sampled individuals was *D. santomea* (Figure 3 & S4)). By contrast, when maintained together at 18°C, the proportion of
D. santomea in each cage did not monotonically change over the course of the experiment (S = 7, 3, 5, and -7; P = 0.13, 0.35, 0.23, and 0.87, respectively), and D. santomea was frequently more abundant than D. yakuba in cages maintained at 18°C (Figure 3). This trend of D. santomea being more abundant than D. yakuba in cages maintained at 18°C was not statistically significant (generalized linear mixed model assuming Poisson distributed error with sample date and cage as random effects: fixed effect of species: Z = -1.314; P = 0.19).

Figure 3. The multi-generation effect of temperature on the coexistence of D. santomea and D. yakuba in experimental sympatry. When D. santomea and D. yakuba are maintained together in cages at 18°C (blue lines) the proportion of D. santomea within the cages tends to be near or above 0.5. By contrast, when maintained at 25°C (red lines), D. santomea is competitively excluded from the cages within ~4-7 generations. The approximate number of generations is indicated, for both temperature treatments, by shaded rectangles (blue = generation times when maintained at 18°C; red = generation times when maintained at 25°C).

Discussion

Our results show how the thermal environment modifies competitive outcomes between the co-occurring sibling species D. yakuba and D. santomea. We find that in the absence of interspecific competition, both species display the highest performance at a moderate temperature of ~22°C (Figure 2A). However, the competitive dominance hierarchy displayed by these two species depends on the thermal environment under which competition takes place: at higher temperatures D. yakuba outcompetes D. santomea, while at lower temperatures D.
santomea outcompetes D. yakuba (Figure 2A). The direction of competitive advantage therefore depends on the thermal environment under which competition takes place and is consistent with the thermal niche these two species inhabit in nature (Figure 1). These results suggest that the temperature-dependent outcome of competition between these two species is contributing to the maintenance of their narrow band of sympatry at mid elevations on the island of São Tomé.

**Competition’s role in moderating local adaptation to different thermal environments**

We did not observe a symmetrical effect of competition at the low and high temperatures that we tested, as D. santomea did not competitively exclude D. yakuba within ~10 generations of being maintained together at 18ºC (Figure 3). While D. santomea’s competitive advantage at lower temperatures meant that it tended to be the more abundant species in experimental enclosures maintained at 18ºC, this effect was minor (see blue lines in Figure 3 and Figure S4). Given D. yakuba is able to maintain high performance at relatively low temperatures, even in the presence of D. santomea, what stops D. yakuba from moving to higher elevation habitats on São Tomé? One variable that our experiments do not account for is the behavioral preferences of D. santomea and D. yakuba for different environments. Laboratory experiments have shown that D. santomea displays a behavioral preference for cooler environments than D. yakuba (Matute et al. 2009). D. yakuba is also a broadly distributed generalist species found across sub saharan Africa (Lachaise et al. 1988; Yassin et al. 2016), while D. santomea has primarily been found in association with figs and is endemic to São Tomé (Lachaise et al. 1988, 2000). It is therefore likely that multiple ecological factors, including temperature, humidity, habitat type, and diet, affect the realized distributions of D. santomea and D. yakuba on São Tomé. Future studies testing competition and performance on different diets and at temperatures below 18ºC (the lowest we tested) are needed to more fully understand the factors defining the ranges of D. santomea and D. yakuba. Our results show that differences in competitive ability across thermal environments is one factor that is likely to affect the realized niches of these two species.

**Context dependent responses to climate change**

Climate change exposes species to warmer mean annual temperatures and their demographic and evolutionary responses to warmer temperatures will depend on both direct and indirect effects of the thermal environment. For example, species may show direct responses to
different thermal environments through the evolution of novel or different physiological traits
(Eliason et al. 2011; Cooper et al. 2012; Campbell-Staton et al. 2020). In the context of climate
change, a species will only be able to persist and maintain demographically viable populations if
the benefit of direct evolutionary responses to warmer (or more variable) temperatures is not
outweighed by negative changes in biotic interactions. Previous studies have, for example,
shown how changes in the thermal environment and/or the community of interspecific
competitors can lead to lower fitness or rapid extirpation (Davis et al. 1998a; Alexander et al.
2015). Competitive exclusion has therefore been discussed as an important outcome of climate
change in a number of species (Finstad et al. 2011; Bulgarella et al. 2014), including even
between anatomically modern humans and Neanderthals (Banks et al. 2008).

One of the predictions generated from our results with respect to competition between D.
santomea and D. yakuba is that under warmer mean daily temperatures, the highland forest
endemic species, D. santomea, may be challenged by competitive exclusion by D. yakuba,
ultimately leading to extinction. There are three caveats to consider when interpreting this
prediction. First, the multigenerational experiment we conducted that showed competitive
exclusion of D. santomea by D. yakuba at 25ºC (Figure 3) did not include appropriate
experimental controls, where the two species were maintained in isolation. We therefore assume
that D. santomea is capable of maintaining viable populations at 25ºC under the same
experimental conditions in the absence of competition. Two lines of evidence support this
assumption. First, when we raised D. santomea in isolation at 25ºC, over a single generation,
each female produced an average of 5.6 male and 6.5 female offspring (Figure 2), suggesting
they can maintain positive population growth at this temperature. Second, previous work on
thermal performance traits in D. yakuba and D. santomea show that males and females of both
species remain fertile and reproductively active at 24ºC (Matute et al. 2009). We therefore
interpret the multigenerational experiment summarized in Figure 3 as providing evidence for
competitive exclusion of D. santomea at 25ºC. Under this interpretation, this result highlights the
importance of considering both direct and indirect effects when estimating the impacts that
climate change will have on biodiversity.

Second, we did not account for frequency- or density-dependent processes that may alter
competitive advantages over the course of the multigenerational experiment. For example, if D.
santomea exhibited strong competitive dominance over D. yakuba at 18ºC one prediction is that
they would consistently be the more abundant species across the course of the experiment. While there was a weak trend in our data to suggest *D. santomea* tended to be more abundant than *D. yakuba* at 18°C, this trend was not statistically significant (GLMM: $P = 0.19$; Figure S4). Our method of estimating relative abundances within experimental cages (non-standardized sampling effort across cages and sampling dates) also did not allow us to test density-dependent effects. Future future work is needed to understand the density-dependence of competitive outcomes between *D. santomea* and *D. yakuba* across different thermal environments.

Third, we did not test whether local adaptation to the experimental environment (biotic and abiotic) altered the competitive interaction between *D. santomea* and *D. yakuba*. Studies of local adaptation in these and other species of Drosophilid flies have shown that they can show evolutionary responses to selection in as few as 10 generations (Matute 2010a, 2010b; Bergland et al. 2014; Tobler et al. 2014; Comeault et al. 2016; Behrman et al. 2018). Local adaptation in *D. santomea*, *D. yakuba*, or both species therefore could have altered the nature of competition or population growth rates over the course of our experiment. Future experiments that either control for evolution or explicitly measure evolution’s effects on species interactions under different environmental conditions have the potential to greatly increase our understanding of species’ responses to environmental change (e.g. (Germain et al. 2020)).

In addition to competitive interactions, temperature is likely to affect, and modify, other types of biotic interactions. For example, the fitness cost of parasites can vary with temperature in a similar way to competition (Lashomb et al. 1987; Vale et al. 2008) and, in some cases, mutualists can become parasites when temperature increases (Baker et al. 2018). In other cases, symbionts can alter the thermal tolerance of their host and act to facilitate colonization of new niches (Russell and Moran 2006). Together with these examples, the results we present here illustrate how predicting a species’ response to environmental change (such as global warming) requires an understanding of both the direct and indirect effects imposed by that change.

*A role for biotic interactions in range-shifts associated with climate change*

An often discussed (and observed) response that species have to climate change is a poleward or upslope shift in their range (Parmesan 2006; Colwell et al. 2008; Lenoir et al. 2008; Schuetz et al. 2019). Range-shifts can be driven by a species tracking favorable abiotic conditions, such as temperature, but the rate and extent of range-shifts is likely to vary among
species, resulting in “community reorganization” (Van der Putten 2012). Climate associated range-shifts are therefore likely to affect interspecific interactions in at least two ways. First, they can change the identity of the interacting members of a community and, in the case of interspecific competition, alter competitive dominance hierarchies (Alexander et al. 2015). Second, they can change the environmental context of the interaction - such as when tracking one environmental variable results in a change in a second - and alter the outcome of specific interactions (Bronstein 1994; Davis et al. 1998a; Tylianakis et al. 2008; Chamberlain et al. 2014; Harrower and Gilbert 2018). The results we have presented here provide an example of the latter and point to the importance of quantifying interspecific interactions under different environments in order to better predict responses to climate change. This may be particularly important for tropical endemic species threatened by the invasion of sibling species that can displace them through competitive exclusion.

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References

Alexander, J. M., J. M. Diez, and J. M. Levine. 2015. Novel competitors shape species’ responses to climate change. Nature 525:515–518.

Arnholt, A. T., and B. Evans. 2017. BSDA: Basic Statistics and Data Analysis.

Baker, D. M., C. J. Freeman, J. C. Y. Wong, M. L. Fogel, and N. Knowlton. 2018. Climate change promotes parasitism in a coral symbiosis. The ISME Journal 12:921–930.

Banks, W. E., F. d’Errico, A. T. Peterson, M. Kageyama, A. Sima, and M.-F. Sánchez-Goñi. 2008. Neanderthal Extinction by Competitive Exclusion. (H. Harpending, ed.) PLoS ONE 3:e3972.

Behrman, E. L., V. M. Howick, M. Kapun, F. Staubach, A. O. Bergland, D. A. Petrov, B. P. Lazzaro, et al. 2018. Rapid seasonal evolution in innate immunity of wild Drosophila melanogaster. Proceedings. Biological Sciences 285.

Bergland, A. O., E. L. Behrman, K. R. O’Brien, P. S. Schmidt, and D. A. Petrov. 2014. Genomic Evidence of Rapid and Stable Adaptive Oscillations over Seasonal Time Scales in Drosophila. PLoS Genetics 10.

Bolnick, D. I. 2001. Intraspecific competition favours niche width expansion in Drosophila melanogaster. Nature 410:463–466.

Bronstein, J. L. 1994. Conditional outcomes in mutualistic interactions. Trends in Ecology & Evolution 9:214–217.

Bulgarella, M., S. A. Trewick, N. A. Minards, M. J. Jacobson, and M. Morgan-Richards. 2014. Shifting ranges of two tree weta species (Hemideina spp.): competitive exclusion and changing climate. Journal of Biogeography 41:524–535.

Calosi, P., D. T. Bilton, J. I. Spicer, S. C. Votier, and A. Atfield. 2010. What determines a species’ geographical range? Thermal biology and latitudinal range size relationships in European diving beetles (Coleoptera: Dytiscidae). Journal of Animal Ecology 79:194–204.

Campbell-Staton, S. C., A. Bare, J. B. Losos, S. V. Edwards, and Z. A. Cheviron. 2018. Physiological and regulatory underpinnings of geographic variation in reptilian cold tolerance across a latitudinal cline. Molecular Ecology 27:2243–2255.

Campbell-Staton, S. C., S. V. Edwards, and J. B. Losos. 2016. Climate-mediated adaptation after mainland colonization of an ancestrally subtropical island lizard, Anolis carolinensis. Journal of Evolutionary Biology 29:2168–2180.

Campbell-Staton, S. C., K. M. Winchell, N. C. Rochette, J. Fredette, I. Maayan, R. M. Schweizer, and J. Catchen. 2020. Parallel selection on thermal physiology facilitates repeated adaptation of city lizards to urban heat islands. Nature Ecology & Evolution 1–7.

Carreiro, M. M., and R. E. Koske. 1992. The effect of temperature and substratum on competition among three species of forest litter microfungi. Mycological Research 96:19–24.
Chamberlain, S. A., J. L. Bronstein, and J. A. Rudgers. 2014. How context dependent are species interactions? Ecology Letters 17:881–890.

Colwell, R. K., G. Brehm, C. L. Cardelús, A. C. Gilman, and J. T. Longino. 2008. Global Warming, Elevational Range Shifts, and Lowland Biotic Attrition in the Wet Tropics. Science 322:258–261.

Comeault, A. A., A. Venkat, and D. R. Matute. 2016. Correlated evolution of male and female reproductive traits drive a cascading effect of reinforcement in Drosophila yakuba. Proceedings of the Royal Society B: Biological Sciences 283.

Cooper, B. S., L. A. Hammad, N. P. Fisher, J. A. Karty, and K. L. Montooth. 2012. In a Variable Thermal Environment Selection Favors Greater Plasticity of Cell Membranes in Drosophila melanogaster. Evolution 66:1976–1984.

Cooper, B. S., A. Sedghifar, W. T. Nash, A. A. Comeault, and D. R. Matute. 2018. A Maladaptive Combination of Traits Contributes to the Maintenance of a Drosophila Hybrid Zone. Current Biology 28:2940-2947.e6.

Core Team, R. 2017. R: A language and environment for statistical computing (Vol. 1). URL http://www.R-project.org/, Vienna, Austria.

Davis, A. J., L. S. Jenkinson, J. H. Lawton, B. Shorrocks, and S. Wood. 1998a. Making mistakes when predicting shifts in species range in response to global warming. Nature 391:783–786.

Davis, A. J., J. H. Lawton, B. Shorrocks, and L. S. Jenkinson. 1998b. Individualistic species responses invalidate simple physiological models of community dynamics under global environmental change. Journal of Animal Ecology 67:600–612.

Delhey, K. 2017. Gloger's rule. Current Biology 27:R689–R691.

———. 2019. A review of Gloger’s rule, an ecogeographical rule of colour: definitions, interpretations and evidence. Biological Reviews 94:1294–1316.

Early, R., and D. F. Sax. 2011. Analysis of climate paths reveals potential limitations on species range shifts. Ecology Letters 14:1125–1133.

Eliason, E. J., T. D. Clark, M. J. Hague, L. M. Hanson, Z. S. Gallagher, K. M. Jeffries, M. K. Gale, et al. 2011. Differences in Thermal Tolerance Among Sockeye Salmon Populations. Science 332:109–112.

Finstad, A. G., T. Forseth, B. Jonsson, E. Bellier, T. Hesthagen, A. J. Jensen, D. O. Hessen, et al. 2011. Competitive exclusion along climate gradients: energy efficiency influences the distribution of two salmonid fishes. Global Change Biology 17:1703–1711.

Freckleton, R. P., P. H. Harvey, and M. Pagel. 2003. Bergmann’s Rule and Body Size in Mammals. The American Naturalist 161:821–825.
Germain, R. M., D. Srivastava, and A. L. Angert. 2020. Evolution of an inferior competitor increases resistance to biological invasion. Nature Ecology & Evolution 4:419–425.

Goldman, J. C., and J. H. Ryther. 1976. Temperature-influenced species competition in mass cultures of marine phytoplankton. Biotechnology and Bioengineering 18:1125–1144.

Grant, P. R., and B. R. Grant. 2006. Evolution of character displacement in Darwin’s finches. Science 313:224–246.

Harris, W. E., A. J. McKane, and J. B. Wolf. 2008. The Maintenance of Heritable Variation Through Social Competition. Evolution 62:337–347.

Harrower, J., and G. S. Gilbert. 2018. Context-dependent mutualisms in the Joshua tree–yucca moth system shift along a climate gradient. Ecosphere 9:e02439.

Hijmans, R. J., J. van Etten, M. Sumner, J. Cheng, A. Bevan, R. Bivand, L. Busetto, et al. 2019. raster: Geographic Data Analysis and Modeling.

Hillebrand, H. 2011. Temperature mediates competitive exclusion and diversity in benthic microalgae under different N:P stoichiometry. Ecological Research 26:533–539.

Kellermann, V., J. Overgaard, A. A. Hoffmann, C. Flojgaard, J.-C. Svenning, and V. Loeschcke. 2012. Upper thermal limits of Drosophila are linked to species distributions and strongly constrained phylogenetically. Proceedings of the National Academy of Sciences of the United States of America 109:16228–16233.

Lachaise, D., M.-L. Cariou, J. R. David, F. Lemeunier, L. Tsacas, and M. Ashburner. 1988. Historical Biogeography of the Drosophila melanogaster Species Subgroup. Pages 159–225 in M. K. Hecht, B. Wallace, and G. T. Prance, eds. Evolutionary Biology. Springer US, Boston, MA.

Lachaise, D., M. Harry, M. Solignac, F. Lemeunier, V. Bénassi, and M. L. Cariou. 2000. Evolutionary novelties in islands: Drosophila santomea, a new melanogaster sister species from São Tomé. Proceedings of the Royal Society of London B 267:1487–1495.

Lashomb, J., D. Krainacker, R. K. Jansson, Y. S. Ng, and R. Chianese. 1987. Parasitism of Leptinotarsa decemlineata (Say) eggs by Edovum putteri Grissell (Hymenoptera: Eulophidae): Effects of host age, parasitoid age, and temperature. The Canadian Entomologist 119:75–82.

Lenoir, J., J. C. Gégout, P. A. Marquet, P. de Ruffray, and H. Brisse. 2008. A Significant Upward Shift in Plant Species Optimum Elevation During the 20th Century. Science 320:1768–1771.
Llopart, A., D. Lachaise, and J. A. Coyne. 2005a. An anomalous hybrid zone in Drosophila. Evolution 59:2602–2607.

———. 2005b. Multilocus analysis of introgression between two sympatric sister species of Drosophila: Drosophila yakuba and D. santomea. Genetics 171:197–210.

Matute, D. R. 2010a. Reinforcement of gametic isolation in Drosophila. PLoS Biology 8:e1000341.

Matute, D. R. 2010b. Reinforcement can overcome gene flow during speciation in Drosophila. Current Biology 20:2229–2233.

Matute, D. R., C. J. Novak, and J. a. Coyne. 2009. Temperature-based extrinsic reproductive isolation in two species of Drosophila. Evolution 63:595–612.

McNab, B. K. 1971. On the Ecological Significance of Bergmann’s Rule. Ecology 52:845–854.

Park, T. 1954. Experimental Studies of Interspecies Competition II. Temperature, Humidity, and Competition in Two Species of Tribolium. Physiological Zoology 27:177–238.

Parmesan, C. 2006. Ecological and Evolutionary Responses to Recent Climate Change. Annual Review of Ecology, Evolution, and Systematics 37:637–669.

Pfennig, K. S., and D. W. Pfennig. 2009. Character Displacement: Ecological and Reproductive Responses to a Common Evolutionary Problem. The Quarterly Review of Biology 84:253–276.

Pianka, E. R. 1966. Latitudinal Gradients in Species Diversity: A Review of Concepts. The American Naturalist 100:33–46.

Polechová, J., and N. H. Barton. 2005. Speciation Through Competition: A Critical Review. Evolution 59:1194–1210.

Roughgarden, J. 1972. Evolution of Niche Width. The American Naturalist 106:683–718.

Russell, J. A., and N. A. Moran. 2006. Costs and benefits of symbiont infection in aphids: variation among symbionts and across temperatures. Proceedings of the Royal Society B: Biological Sciences 273:603–610.

Schluter, D., and J. D. McPhail. 1992. Ecological Character Displacement and Speciation in Sticklebacks. The American Naturalist 140:85–108.

Schuetz, J. G., K. E. Mills, A. J. Allyn, K. Stamieszkin, A. L. Bris, and A. J. Pershing. 2019. Complex patterns of temperature sensitivity, not ecological traits, dictate diverse species responses to climate change. Ecology 42:111–124.

Soberón, J. 2007. Grinnellian and Eltonian niches and geographic distributions of species. Ecology Letters 10:1115–1123.
Stager, M., H. S. Pollock, P. M. Benham, N. D. Sly, J. D. Brawn, and Z. A. Cheviron. 2016. Disentangling environmental drivers of metabolic flexibility in birds: the importance of temperature extremes versus temperature variability. Ecography 39:787–795.

Stuart, Y. E., and J. B. Losos. 2013. Ecological character displacement: glass half full or half empty? Trends in Ecology & Evolution 28:402–408.

Tobler, R., S. U. Franssen, R. Kohler, P. Orozco-terWengel, V. Nolte, J. Hermisson, and C. Schlötterer. 2014. Massive Habitat-Specific Genomic Response in D. melanogaster Populations during Experimental Evolution in Hot and Cold Environments. Molecular Biology and Evolution 31:364–375.

Tylianakis, J. M., R. K. Didham, J. Bascompte, and D. A. Wardle. 2008. Global change and species interactions in terrestrial ecosystems. Ecology Letters 11:1351–1363.

Vale, P. F., M. Stjernman, and T. J. Little. 2008. Temperature-dependent costs of parasitism and maintenance of polymorphism under genotype-by-environment interactions. Journal of Evolutionary Biology 21:1418–1427.

Van der Putten, W. H. 2012. Climate Change, Aboveground-Belowground Interactions, and Species’ Range Shifts. Annual Review of Ecology, Evolution, and Systematics 43:365–383.

Willig, M. R., D. M. Kaufman, and R. D. Stevens. 2003. Latitudinal Gradients of Biodiversity: Pattern, Process, Scale, and Synthesis. Annual Review of Ecology, Evolution, and Systematics 34:273–309.

Wilson, D. S., W. G. Knollenberg, and J. Fudge. 1984. Species packing and temperature dependent competition among burying beetles (Silphidae, Nicrophorus). Ecological Entomology 9:205–216.

Winkelmann, K., M. J. Genner, T. Takahashi, and L. Rüber. 2014. Competition-driven speciation in cichlid fish. Nature Communications 5:1–8.

Yassin, A., V. Debat, H. Bastide, N. Gidaszewski, J. R. David, and J. E. Pool. 2016. Recurrent specialization on a toxic fruit in an island Drosophila population. Proceedings of the National Academy of Sciences 113:4771–4776.