Cyclin K Inhibits HIV-1 Gene Expression and Replication by Interfering with Cyclin-dependent Kinase 9 (CDK9)-Cyclin T1 Interaction in Nef-dependent Manner

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Human immunodeficiency virus-1 (HIV-1) exploits a number of host cellular factors for successful survival and propagation. The viral protein Nef plays an important role in HIV-1 pathogenesis by interacting with various cellular proteins. In the present work, we identified Cyclin K (CycK) as a novel Nef-interacting protein, and for the first time, we showed that CycK inhibits HIV-1 gene expression and replication in a Nef-dependent manner. The positive elongation factor b complex comprising cyclin-dependent kinase 9 (CDK9) and Cyclin T1 is a critical cellular complex required for viral gene expression and replication. Enhanced expression of CycK in the presence of Nef induced CycK-CDK9 binding, which prevented CDK9-Cyclin T1 complex formation and nuclear translocation of CDK9, resulting in inhibition of HIV-1 long terminal repeat-driven gene expression. Furthermore, this effect of CycK was not observed with Nef-deleted virus, indicating the importance of Nef in this phenomenon. Finally, silencing of CycK in HIV-1-infected cells resulted in increased translocation of CDK9 into the nucleus, leading to increased viral gene expression and replication. These data also suggest that endogenous CycK might act as an inhibitory factor for HIV-1 gene expression and replication in T-cells. Thus, our results clearly demonstrate that CycK utilizes HIV-1 Nef protein to displace CycT1 from the positive elongation factor b complex, resulting in inhibition of HIV-1 gene expression and replication.

Human immunodeficiency virus type-1 (HIV-1) perpetuates its survival in the host by utilizing multiple strategies such as irreversible integration of its DNA into the host cell genome to establish the provirus (1), diversification of its genome to escape or tolerate adaptive immune responses (2), and use of the accessory genes (nef, vif, vpu, and vpr) to modulate host cell immune responses further (3–5). These viral accessory genes are frequently seen as dispensable in many in vitro cell culture systems but seem to be indispensable, in the context of natural infections in vivo. Among these accessory genes, pleiotropic functions of Nef have been studied extensively. It is a 27–30-kDa, N-terminally myristoylated protein having a well-characterized CD4 down-regulatory activity (6). Nef does not possess any enzymatic activity, but it modulates diverse signaling pathways and the gene expression of host cell proteins (7, 8). Furthermore, Nef influences the activation state of the host cell by bringing together different host cell proteins, protein kinases, and components of the endocytic machinery and making the cells permissible to the virus (9, 10). All these functions of Nef are manifested by a number of events such as activation of signaling molecules, modulation of apoptosis, activation of transcription factors, mitigation of transcriptional repressors, and an increase in the infectivity of virions (11). Although these functions of Nef have been well studied, controversy exists on its role in viral infectivity and replication as both negative (12, 13) and positive (14, 15) roles have been attributed in the literature. Although a number of reports show that Nef increases viral replication by activating T-cells, the molecular basis of Nef-induced viral gene expression and replication remains to be clearly elucidated. Nef was also shown to physically interact with Tat and augment viral gene expression (16), although availability of Tat is critical for the elongation step of long terminal repeat (LTR)2-driven transcription by RNA polymerase II. Several cellular factors have been identified for their roles in this process, including positive transcription elongation factor b (P-TEFb) complex. Different P-TEFb complexes isolated from mammalian cells contain a common catalytic subunit, cyclin-dependent kinase 9 (CDK9), and one of the regulatory cyclins, CycT1, CycT2a, CycT2b, or CycK (17–19). However, CycT1-containing P-TEFb complex is the key complex responsible for HIV-1 transcription elongation (19). Tat is expressed early in the replicative cycle of HIV and is essential for viral gene expression from the LTR promoter. It recognizes the 5′-bulge in the transactivation response stem loop RNA, which is located at the 5′-end of all viral transcripts. Tat binds to CycT1, and together they form the combinatorial surface that interacts with the transactivation response RNA with high affinity and specificity. The obligate partner of CycT1, CDK9, then hyperphosphorylates the C-terminal domain (CTD) of RNA polymerase II, resulting in increased HIV-1 transcription elongation (20, 21).

HIV-1 gene expression is controlled by several cellular as well as viral proteins, including Nef. Evidences suggest that Nef enhances Tat-mediated viral transcription through a heterogeneous nuclear ribonucleoprotein K-nucleated signaling com-

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2 The abbreviations used are: LTR, long terminal repeat; CDK9, cyclin-dependent kinase 9; HEXIM1, hexamethylene bisacetamide-inducible protein 1; CTD, C-terminal domain; Cyc, Cyclin; P-TEFB, positive elongation factor b; Luc, luciferase; EGFP, enhanced green fluorescent protein.
Cyclin K Inhibits HIV-1 Gene Expression and Replication

In another study, Hsp40 was shown to interact and co-localize with Nef in the nucleus and thereby augment LTR-driven gene expression (23). In contrast, HEXIM1 and Hsp70 negatively regulate HIV-1 gene expression and replication (24, 25). HEXIM1 along with 7SK RNA was shown to be essential for inactivation of the P-TEFb complex (26, 27). Reportedly, the HEXIM1–7SK RNA complex physically competes with Tat for binding to P-TEFb (28). Recently, CycK has been also shown to inhibit viral replication in a T-cell line by inhibiting LTR-mediated transcription (29), although the mechanism of inhibition remains to be elucidated. CycK was first identified as a protein that could restore progression of the cell cycle and was most closely related to human Cyclins C and H (30). The kinase partner of CycK was identified as CDK9 (17), although a recent report also indicates CycK interaction with CDK12 and -13 (31). The complex between CycK and CDK9 was also shown to function as a CTD kinase in vitro (17).

Although Nef was earlier reported as a negative factor for viral replication (32), recent evidence has convincingly demonstrated Nef as an enhancer of viral replication (16, 22, 33–36). Nef performs most of its functions by interacting with cellular proteins. The majority of the functional studies of Nef to date have been performed with HIV-1 subtype B Nef protein, and there have been few studies using subtype C Nef protein. In the present study, we attempted to identify novel Nef-interacting host cell proteins using subtype C Nef as a bait in a yeast two-hybrid system. Our results show that both HIV-1 subtype C and B Nef interact with human Cyclin K in vitro and in vivo. Furthermore, CycK overexpression led to reduction in LTR-mediated gene expression and virus replication in the presence of Nef. Finally, CycK along with Nef disrupted CDK9-CycT1 interaction and thereby inhibited nuclear translocation of CDK9, consequently inhibiting viral gene expression and replication.

**EXPERIMENTAL PROCEDURES**

*Plasmids, Cell Lines, and Antibodies—The nef gene from HIV-1 subtype C Indian isolate IN301904 (37) was cloned into pAS2-1 (Clontech) and pcDNA3.1 vector (Invitrogen), respectively, as described earlier (23). The sequence of cloned nef was confirmed by DNA sequencing using an ABI 310 Genetic Analyzer (Applied Biosystems). Immunoblotting was used to confirm the expression of Nef from both vectors. The pAS2-1 Nef plasmid expressing subtype C Nef and Gal4 DNA binding domain as a fusion protein was used as the bait in yeast two-hybrid screening of a cDNA library of human leukocytes in the pACT2 vector obtained from Clontech. The NL4-3 molecular clone (pNL4-3) was obtained from the National Institutes of Health AIDS Reagent Program (38). The nef-deleted NL4-3 molecular clone (pNL4-3ΔNef) and glutathione S-transferase (GST)-Nef (B) plasmids were obtained from Dr. John C. Gautelli (39) and Dr. K. Saksela, respectively. The vectors expressing HA-Nef and HA-Nef mutants used in this study were obtained from Dr. W. C. Greene (40). The subtype C Nef-expressing GST-Nef plasmid was constructed by cloning subtype C nef gene into BamHI and EcoRI sites of pGEX-5X.1 (GE Healthcare). The sequence of cloned nef was confirmed by DNA sequencing as above and also by Western blotting with Nef- and GST-specific antibodies. EGFP-Nef plasmid was constructed by cloning the complete GFP-Nef fusion gene sequence from pEGFPN2-Nef in EcoRI and NotI sites of pcDNA6HisC (Invitrogen). Cyclin K (CCNK) gene encoding the 354-amino acid isoform (30) was cloned in-frame into BamHI and EcoRI sites of pcDNA6HisC to obtain pcDNA6HisC-Cyclin K (pc-CycK). The cloning of pc-CycK was confirmed by restriction digestion and by DNA sequencing in an ABI 310 Genetic Analyzer (Applied Biosystems). The expression was confirmed by Western blotting using anti-Xpress epitope and CycK antibodies. HEK-293T cells (human embryonic kidney cell line) and Jurkat cells (CD4+ human T cell line) were obtained from the National Centre for Cell Science Cell Repository in India. CEM-GFP, a CD4+ human T cell line, was obtained from the National Institutes of Health AIDS repository. Jurkat-1G5 cells (CD4+ human T cell line) having a copy of integrated LTR-Luc in the genome were obtained from the National Institutes of Health AIDS Reagent Program. Antibodies specific for Cyclin K, Cyclin T1, CDK9, GAPDH, and HA tag were obtained from Santa Cruz Biotechnology. Anti-Nef monoclonal antibody was obtained from Fitzgerald, anti-Nef polyclonal serum was obtained from Dr. Shahid Jameel, International Centre for Genetic Engineering and Biotechnology, New Delhi, India (41), and anti-p24 gag polyclonal serum was obtained from the National Institutes of Health AIDS repository.

*Yeast Two-hybrid Screening—A human leukocyte cDNA library in pACT2 vector (Clontech) was screened for Nef-interacting proteins by co-transformation with the pAS2-1 Nef plasmid into yeast strain AH109 (Clontech) as described previously (23). Positive clones were selected based on growth in medium lacking adenine, histidine, tryptophan, and leucine and also by expression of β-galactosidase. Co-transformants in AH109 yeast strain were screened three times for growth on these selection plates and also for β-galactosidase activity to exclude false positive clones. A liquid β-galactosidase assay was finally performed to confirm the interaction using the yeast β-galactosidase assay kit (Pierce) according to the manufacturer’s protocol. The interacting protein in the positive clone was identified by rescuing the gene fragment using PCR amplification with pACT2-specific primers followed by DNA sequencing and BLAST analysis.

*HIV-1 Infection and Virus Quantitation—*CEM-GFP cells (2 × 106) were infected with HIV-1NL44.3 virus at a multiplicity of infection of 0.5 in the presence of Polybrene (1 μg/ml) as described earlier (23). 24 h postinfection, cells were transfected with 200 nM siRNA, and then 48 h later, cells were harvested and lysed for preparation of whole cell lysate. The culture supernatants from infected and molecular clone-transfected cells were used to determine virus production by p24 gag antigen capture ELISA (Perkin Elmer Life Sciences).

*Transient Transfection and Luciferase Assay—*HEK-293T cells were transfected with HIV-1 LTR-reporter vector (pLTR-Luc) along with other expression vectors using the calcium phosphate precipitation method and were harvested 36 h post-transfection for luciferase assay. Jurkat-1G5 cells were transfected with pNL4-3 and pc-CycK using Lipofectamine 2000 (Invitrogen) according to the manufacturer’s protocol, and cells
were harvested 48 h post-transfection for luciferase assay. The cells were then lysed in cell lysis reagent (Promega), and luciferase assays were performed using Lucite substrate (Perkin-Elmer Life Sciences). Normalization of transfection efficiency was done using enhanced green fluorescent protein reporter (pEGFP-N1) co-transfection and quantitation as described earlier (42). For RNAi experiments, Jurkat-1G5 cells were transfected with 100 nM siRNAs along with expression vectors using Lipofectamine 2000 (Invitrogen) according to the manufacturer’s protocol. The culture supernatants were also collected at the time of luciferase assay from HIV-1 NL4-3–molecular clone–transfected cells to determine virus production.

**GST Pulldown, Immunoprecipitation, and Immunoblotting—Escherichia coli BL21(DE3) cells expressing either GST or GST-Nef were induced with isopropyl β-d-thiogalactoside followed by purification of proteins using glutathione-Sepharose beads (Amersham Biosciences). Jurkat cells were lysed in lysis buffer (50 mM Tris-HCl, pH 7.4, 5 mM EDTA, 0.12 M NaCl, 0.5% Nonidet P-40, 0.5 mM NaF, 1 mM dithiothreitol, 0.5 mM phenylmethylsulfonyl fluoride) with a protease inhibitor mixture (Roche Applied Bioscience) on ice for 45 min. The clarified lysates were incubated with either GST or GST-Nef protein immobilized on glutathione-Sepharose beads at 4 °C and subjected to five washes with lysis buffer. The complexes were resuspended in Laemml sample buffer, boiled, and resolved by 12% SDS-PAGE. Proteins were transferred onto polyvinylidene difluoride (PVDF) membrane, and the membrane was probed with anti-CycK antibody.

Infected and uninfected CEM-GFP cells and co-transfected HEK-293T cells were lysed in lysis buffer as mentioned above. The clarified lysate was incubated with polyclonal antibody, and the antigen–antibody complex was pulled down by an equal mixture of Protein A and G beads followed by resolution by 12% SDS-PAGE. The proteins were transferred onto PVDF membrane, and then the membrane was probed with specific antibodies mentioned above. The blots were developed using the ECL Plus system (Amersham Biosciences). Furthermore, equal amounts of protein were taken from cell lysates and were resolved by SDS-PAGE followed by immunoblotting for CycK and other proteins.

**Immunofluorescence Microscopy—**HEK-293T cells grown on coverslips were co-transfected with expression vectors using Lipofectamine (Invitrogen). Cells were harvested 24 h post-transfection and stained with CycK and Nef antibodies after fixing with 2% paraformaldehyde. The secondary antibodies used were Cy3-conjugated, Cy5-conjugated, and fluorescein isothiocyanate-conjugated specific antibodies (Chemicon). After washing, cells were mounted in mounting medium containing antifade reagent (Santa Cruz Biotechnology) on the slide, and the samples were analyzed with a confocal microscope (Zeiss LSM 510).

**Reverse Transcription–PCR—**RNA was prepared from HIV-1 NL4-3–infected and uninfected CEM-GFP cells or from Jurkat-1G5 cells using TRIzol reagent (Invitrogen). The cDNA was made using Moloney murine leukemia virus reverse transcriptase (Invitrogen) followed by amplification by PCR for CCNK, β-ACTIN, and HIV-p24 gene with Taq polymerase (Bangalore Genei) using standard conditions and gene-specific oligonucleotide primers. The primers used were as follows: CCNK: forward primer, 5′-TGAAAAGACCAACTCAAGGGT-3′; reverse primer, 5′-AACAAACGGCTCCCCCTCCTC-3′; β-ACTIN: forward primer, 5′-ATCTACCTGGTGCACACCATGGG-CAGT-3′; reverse primer, 5′-GGAGGTAGCAGGTTGCGTT-TACGAGATC3′; and HIV-p24: forward primer, 5′-ATACCTATCCCCAGTAGAGAATT-3′; reverse primer, 5′-TTTGGTCCTGTCTTATGTCCAGAATGC-3′.

**Preparation of Nuclear and Cytoplasmic Extracts—**HIV-1-infected CELM-GFP and transfected HEK-293T cells were harvested and washed with ice-cold phosphate-buffered saline (PBS). Nuclear and cytoplasmic extracts were prepared using ProteoJET reagent (Fermentas) according to the manufacturer’s protocol. Equal amounts of nuclear and cytoplasmic extracts were used for immunoblotting.

**Statistical Analysis—**Each individual experiment was repeated at least three times. The error bars in the figures represent the mean ± S.D. of three independent experiments. Statistical analysis of the experimental data was performed using Student’s t test with the levels of significance defined as p < 0.05 (*) and p < 0.01 (**).

**RESULTS**

HIV-1 Nef protein interacts with a number of cellular proteins, most of which perform critical functions in signaling pathways. Although the role of Nef in HIV-1 pathogenesis has been well studied, the molecular mechanism of how Nef utilizes these interacting cellular factors to modulate viral replication and gene expression is still a conundrum. The functions attributed to Nef are believed to be conserved across various HIV-1 subtypes, and to date, the majority of the functional studies have been performed with subtype B Nef protein. As subtype C has become the most prevalent subtype in the world at present, we wanted to identify novel cellular factors, if any, that may interact with subtype C Nef protein. The subtype C Nef from Indian isolate IN301904 (37) used in the yeast two-hybrid screening was shown to have 84% similarity with subtype B NL4-3 Nef (23).

**Identification of Cyclin K as Subtype C Nef-interacting Protein—**To identify cellular factors interacting with subtype C Nef, we performed a yeast-two hybrid screening of a human leukocyte cDNA library using full-length subtype C Nef as bait. Positive clones were identified after several rounds of screening and culture on X-gal-containing dropout plates (Fig. 1A). The gene expressed in the positive clone was amplified by PCR using pACT2-specific primers followed by DNA sequencing. Sequence analysis and database comparison revealed that the clone expressed a gene encoding for Cyclin K (GenBank™ Gene ID: 8812). To validate this interaction, a liquid β-galactosidase assay was performed, and the clone expressing Cyclin K and Nef showed β-galactosidase activity comparable with the positive control (Fig. 1B). To further confirm our finding, a GST–Nef (subtype C) pulldown assay was performed using Jurkat cell lysate as described under “Experimental Procedures.” The result of the pulldown clearly indicates that Cyclin K (CycK) interacts with subtype C Nef but not with GST (Fig. 1C). To see whether these proteins interact in vivo, immunofluorescence staining was performed in subtype C...
Cyclin K interacts with HIV-1 subtype C Nef in the yeast two-hybrid assay. A human leukocyte cDNA library was screened using subtype C Nef as bait in the yeast two-hybrid system. One of the clones co-expressing Cyclin K and Nef was found to be positive after repeated screening and was grown on a quadruple knock-out plate with appropriate controls. B, β-galactosidase activity of Cyclin K-Nef yeast co-transformant in liquid β-galactosidase activity assay as described in the text. C, GST-Nef (subtype C) pulls down Cyclin K from Jurkat lysate. GST and GST-Nef (subtype C) were incubated with Jurkat cell lysate followed by SDS-PAGE and immunoblotting with Cyclin K antibody. D, co-localization of Cyclin K and subtype C Nef in co-transfected HEK-293T cells. HEK-293T cells were grown on coverslips and transfected with pcDNA-NefC and pc-CycK. After 24 h, transfected cells were stained with Nef and Cyclin K antibodies as described in the text. The immunofluorescence was visualized by confocal microscopy. E, GST-Nef (subtype B) also pulls down Cyclin K from Jurkat lysate. GST and GST-Nef (subtype B) were incubated with Jurkat cell lysate followed by SDS-PAGE and immunoblotting with Cyclin K antibody. F, co-localization of Cyclin K and subtype B Nef in co-transfected HEK-293T cells. HEK-293T cells were grown on coverslips and transfected with pNL4-3 and pc-CycK. The immunofluorescence staining and visualization were performed as described above. G, Nef and Cyclin K interact in HIV-1-infected CEM-GFP cells. CEM-GFP cells were infected with HIV-1NL4-3 virus as described in the text. The cells were lysed on day 5 postinfection. The lysates of uninfected (U) and infected (I) CEM-GFP cells were co-immunoprecipitated (IP) with either Nef or Cyclin K antibody followed by immunoblotting with Cyclin K or Nef antibody, respectively, as presented in the figure. NEG, negative; POS, positive. The red lines in panels D and F indicate 10 μm scale.

Nef-transfected 293T cells. Interestingly, these proteins co-localized primarily in the cytoplasm with some localization in the nucleus (Fig. 1D).

Cyclin K Also Interacts with Subtype B Nef Both in Vitro and in Vivo—To test whether Cyclin K also interacts with subtype B Nef, a GST pulldown assay was performed as described above. Jurkat cell lysate was incubated with bead-bound GST and GST-Nef (subtype B) followed by immunoblotting with Cyclin K antibody, which showed that Cyclin K specifically interacts with GST-Nef but not with GST alone (Fig. 1E). Immunofluorescence staining was also performed in pNL4-3- and pc-CycK-co-transfected HEK-293T cells to confirm in vivo interaction of Cyclin K with Subtype B Nef. Both proteins were found to be co-localized extensively in the cytoplasm as shown in Fig. 1F. We further validated this interaction in vivo by co-immunoprecipitation from uninfected and HIV-1NL4-3-infected CEM-GFP cells. Uninfected and HIV-1-infected cell lysates were immunoprecipitated with either Nef or Cyclin K antibody followed by immunoblotting with Cyclin K or Nef antibody. The analyses of immunoprecipitates clearly suggest that Cyclin K and Nef specifically interact in HIV-1-infected cells (Fig. 1G). The uninfected cell lysate served as a control in co-immunoprecipitation experiments. These results clearly suggest that Nef interacts with Cyclin K both in vitro and in vivo.

Cyclin K Inhibits HIV-1 LTR-driven Gene Expression in Presence of Nef—Cyclin K was first identified as a novel C type cyclin that is associated with an in vitro kinase activity toward CTD of RNA polymerase II (30). As this activity is important for HIV-1 gene expression, we wanted to examine the effect of Cyclin K on HIV-1 LTR-driven viral gene expression. First, HEK-293T cells were co-transfected with the pLTR-Luc construct along with Tat and Cyclin K expression vectors. As shown in Fig. 2A, overexpression of Cyclin K in the presence of Tat alone did not cause any significant change in LTR-driven reporter activity. However, in the presence of Nef, Cyclin K overexpression led to inhibition of both Tat- and Nef-induced LTR-driven gene expression in 293T cells in dose-dependent manner (Fig. 2B). This result clearly shows that Cyclin K inhibits HIV-1 LTR-driven gene expression in the presence of Nef. To explore this finding further, we then used Jurkat-1G5 cells, a human CD4+ T-cell line having a copy of “integrated” LTR-Luc in the genome (43). Jurkat-1G5 cells were transfected with increasing amounts of Cyclin K along with an HIV-1 molecular clone (pNL4-3) followed by a luciferase activity assay 48 h post-transfection. There was a significant reduction in luciferase activity upon Cyclin K overexpression in a dose-dependent manner (Fig. 2C). Concurrently, Western blotting indicated that p24 expression in these cells was also reduced in a dose-dependent manner with increasing expression of Cyclin K (data not shown). To further elucidate the effect of Cyclin K on viral gene expression and replication, a p24 antigen capture ELISA of culture supernatant of these cells was also performed. Virus production in Cyclin K-overexpressing cells analyzed by p24 antigen release in the culture supernatant was significantly reduced in a dose-dependent manner as compared with pNL4-3 only-transfected cells (Fig. 2D, gray bars). Moreover, when Jurkat-1G5 cells were...
transfected with a Nef-deleted molecular clone (pNL4-3/H9004Nef), CycK-mediated reduction in virus production was not observed at all (Fig. 2D, black bars). These data clearly suggest that CycK acts as a cellular inhibitory factor against HIV-1 in a Nef-dependent manner.

**Endogenous CycK Reduces Viral Gene Expression and Replication in T-cells**—To identify the role of endogenous CycK in HIV-1 gene expression and replication, Jurkat-1G5 cells were co-transfected with pNL4-3 and specific siRNA to silence CycK expression as described under “Experimental Procedures.” After 72 h of transfection, cells were harvested, and culture supernatants were collected for estimation of virus production by a p24 antigen capture ELISA. The harvested cells were used for cell lysate preparation. The efficacy of siRNA was checked by immunoblotting, and the cell lysates were also used for a luciferase activity assay. As compared with control siRNA-transfected Jurkat-1G5 cells, transfection of CycK siRNA showed significant silencing of CycK expression (Fig. 3A, inset). The luciferase activity in CycK siRNA-transfected cells was significantly increased as compared with control siRNA-transfected cells (Fig. 3B). These results clearly indicate that CycK down-regulation leads to an increase in HIV-1 gene expression and virus production.

The expression level of p24 was higher in CycK siRNA-transfected cells as compared with control (Fig. 3A, inset). Furthermore, a similar result was obtained when virus production from siRNA-transfected cells was determined by a p24 antigen capture ELISA. The amount of virus in the culture supernatant of CycK siRNA-transfected cells was significantly increased as compared with control siRNA-transfected cells (Fig. 3B). These results clearly indicate that CycK down-regulation leads to an increase in HIV-1 gene expression and virus production.

To assess the role of CycK in a more physiological scenario, we then performed the above experiment in an HIV-1-infected CEM-GFP reporter T-cell line. The CEM-GFP cells were first transfected with siRNA, and after 24 h, they were infected with HIV-1NL4-3 at 0.5 multiplicity of infection. On day 3 postinfection, cells were harvested to monitor progression of infection by flow cytometry. The cells transfected with CycK siRNA showed significantly increased as compared with control siRNA-transfected cells (Fig. 3B). These results clearly indicate that CycK down-regulation leads to an increase in HIV-1 gene expression and virus production.

**FIGURE 2.** CycK inhibits HIV-1 LTR-driven gene expression and replication in Nef-dependent manner. A, CycK does not modulate LTR-driven luciferase expression in the presence of Tat alone. HEK-293T cells were transfected with HIV-1 LTR-Luc reporter vector along with pc-CycK in increasing amounts (0.3–1.2 µg) and with pcDNA-Tat (0.25 µg) followed by a luciferase assay 36 h post-transfection. The total amount of DNA transfected was equal in all lanes. B, CycK down-regulates Tat-induced LTR-driven luciferase expression in the presence of Nef. HEK-293T cells were transfected with HIV-1 LTR-Luc reporter vector along with pc-CycK in increasing amounts (0.3–1.2 µg) and with pcDNA-Nef (0.5 µg) and pcDNA-Tat (0.25 µg). 36 h post-transfection, cells were lysed in Promega cell lysis reagent, and a luciferase assay was performed. C, CycK inhibits integrated LTR-driven gene expression in T-cells. Jurkat-1G5 cells were transfected with pNL4-3 (1 µg) and increasing amounts of pc-CycK (0.25–0.5 µg). 36 h post-transfection, cells were lysed in Promega cell lysis reagent followed by a luciferase assay. D, Nef is necessary for CycK-mediated inhibition of HIV-1 replication. Jurkat-1G5 cells were transfected with either pNL4-3 (gray bars) or pNL4-3Nef (black bars) and with increasing amounts of pc-CycK (0.25–0.5 µg) as presented in the figure. 48 h post-transfection, the culture supernatant was collected, and the amount of virus present in the culture supernatant was estimated by a p24 gag antigen capture ELISA.
pared with the control siRNA-transfected cells (Fig. 3C). A similar pattern was obtained by immunoblotting for p24 (Fig. 3C, inset), thereby indicating that CycK acts as an inhibitory factor in HIV-1-infected cells.

Cyclin K Inhibits HIV-1 Gene Expression and Replication

FIGURE 3. Cyclin K silencing increases LTR-mediated gene expression and virus production in T-cells. A, inhibition of Cyclin K expression by siRNA leads to an increase in LTR-driven luciferase activity in Jurkat-1G5 cells. Jurkat-1G5 cells were transfected with 100 nM control (siCtrl) siRNA or CycK-specific siRNA (siCycK), and 24 h post-transfection, cells were transfected with pNL4-3 vector (1 μg). 48 h post-transfection, cells were harvested, and a luciferase assay was performed. The inset shows the efficiency of siRNA-mediated Cyclin K silencing by immunoblotting. B, inhibition of Cyclin K expression by siRNA leads to an increase in virus production from Jurkat-1G5 cells. The virus released in the culture supernatant of cells in the experiment described in A was estimated by a p24 gag antigen capture ELISA. C, Cyclin K silencing increases virus production in HIV-1NL4-3-infected CEM-GFP cells. CEM-GFP cells were first transfected with 200 nM control or CycK-specific siRNA. 24 h post-transfection, cells were infected with HIV-1NL4-3 at 0.5 multiplicity of infection, and on day 3, the culture supernatant was used for a p24 antigen capture ELISA. The inset shows the efficacy of siRNA-mediated silencing by immunoblotting.

FIGURE 4. Cyclin box 1 domain of Cyclin K is necessary for its inhibitory activity on LTR-driven gene expression and HIV-1 virus production. A, schematic representation of Cyclin K mutants generated and used in the present study. B, cyclin box 1 domain is required for the inhibition of HIV-1 LTR-driven luciferase activity. HEK-293T cells were transfected with LTR-Luc reporter and pNL4-3 molecular clone along with various CycK mutant expression vectors as indicated in the figure. 24 h post-transfection, cells were harvested followed by a luciferase assay of the lysate. C, cyclin box 1 domain is required for the inhibition of HIV-1 virus production. HEK-293T cells were transfected with LTR-Luc reporter and pNL4-3 molecular clone along with various CycK mutant expression vectors as indicated in the figure. 24 h post-transfection, cell supernatants were used to analyze the virus production by a p24 antigen capture ELISA. D, expression of different Cyclin K mutant proteins in transfected HEK-293T cells. HEK-293T cell lysates were examined for the expression of various CycK mutants using Xpress epitope antibody and GAPDH by Western blot analysis in the experiments in B and C.

Cyclin Box 1 Is Required for CycK-mediated Reduction of LTR-driven Gene Expression—Human CycK is a 354-amino acid protein; however, a less abundant isoform of CycK consisting of 580 amino acids also exists. The overall structure of CycK
is similar to other cyclin structures. It consists of 10 helical
domains, which are arranged in two cyclin boxes along with two
short helices (HNa and HNb) preceding the N-terminal cyclin
box. The five helices comprising the N- and C-terminal cyclin
domains are designated as H1–H5 and H1’–H5’, respectively.
The N-terminal domain (amino acids 1–151) of CycK is
responsible for binding to CDK9; however, the two N-terminal
helices HNa and HNb are not essential (44). The region from
amino acids 152 to 267 forms cyclin box 2, and the region com-
prising amino acids 268–354 can be considered as the C termi-
nus of the protein (Fig. 4A). Different deletion mutants of
Cyclin K, as shown in Fig. 4A, were cloned in pcDNA6HisC.

To investigate the effect of different CycK mutants on LTR-
mediated gene expression, HEK-293T cells were transfected
with pLTR-Luc reporter plasmid along with pNL4-3 and wild
type or different CycK mutants followed by a luciferase activity
assay. Upon wild type CycK overexpression (Fig. 4B, third bar),
the luciferase activity was reduced by ~5-fold as compared with
that of vector control (second bar). A similar pattern was
observed when CycK(1–267) (fourth bar) and CycK(1–151)
(sixth bar) mutants were used in the experiment (Fig. 4B). How-
ever, no significant change in luciferase activity was observed
when cyclin box 1 deletion mutants like CycK(152–354),
CycK(152–267), and CycK(268–354) were co-transfected with
LTR-Luc and pNL4-3 vectors (Fig. 4B), indicating that the
N-terminal end of Cyclin K was important for CycK activity,
whereas the C-terminal end was not required for this activity.

To find the region of Cyclin K important for interaction with Nef,
immunoprecipitation of wild type or CycK mutants and
Nef-transfected HEK-293T cell lysate was performed with anti-
Nef antibody followed by immunoblotting of the complex with
anti-Xpress epitope antibody. Our results clearly show that Nef
specifically interacts with only wild type CycK and the two
mutants having the region comprising amino acids 1–151 (Fig.
5A). These data suggest that domain 1–151 is responsible for
the Nef binding property of CycK, which correlates well with
the inhibitory activity of CycK delineated in the earlier figure
(Fig. 4C). It has been reported that Nef has some signature
motifs responsible for binding to various cellular proteins (45).
To explore the motif of Nef important for binding to CycK, we
used a set of Nef point mutants (40). HEK-293T cells were co-
transfected with HA-Nef mutant expression vectors along with
CycK-WT expression vector. The cell lysates were then sub-
jected to immunoprecipitation with CycK antibody followed by
immunoblotting with Nef antibody. The co-immunoprecipita-
tion analysis indicates that myristoylation of Nef is required for
CycK interaction as complete loss of interaction was observed
when the glycine at the second position, which is critical for
myristoylation, was replaced with alanine in Nef (Nef-G2A
mutant) (Fig. 5B).
Cyclin K Inhibits HIV-1 Gene Expression and Replication

**Nef Induces Cyclin K Interaction with CDK9**—The CDK9-CycT1 complex (P-TEFb complex) plays a central role in HIV-1 LTR-driven gene expression (46). Both CycK and CycT1 are known to bind CDK9 through their N-terminal cyclin box of which 53 of 80 residues are identical (17, 44). Based on this report, we initiated studies to elucidate the mechanism of CycK-mediated reduction in viral gene expression. To test whether the presence of CycK alters the physical association between CycT1 and CDK9, we performed a co-immunoprecipitation experiment by binding to CDK9 and sequestering it in the cytoplasm. We hypothesized that inhibition of CDK9-CycT1 complex formation by CycK could be responsible for restricting CDK9 from translocating into the nucleus to form active P-TEFb complex and thus causing reduction in viral gene expression. We therefore analyzed whether ectopic expression of CycK inhibits CDK9 nuclear translocation. We performed Western analysis of nuclear and cytoplasmic extracts of HEK-293T cells transfected with Nef and different amounts of CycK. It was clearly evident that the presence of Nef and CycK significantly reduced CDK9 translocation into the nucleus as analyzed by immunoblotting (Fig. 7A). This experiment also showed that with increasing CycK expression there was a substantial decrease in CDK9 levels in the nucleus (Fig. 7A). To further substantiate this finding, we transfected HEK-293T cells with EGFP-Nef and CycK expression vectors to perform an immunofluorescence study. In EGFP-Nef-transfected cells, CDK9 was evidently co-localized in the cytoplasm with CycK and EGFP-Nef (Fig. 7B). When EGFP alone was used as a control, CDK9 was visible mostly in the nucleus. Thus, the CycK-Nef complex may act as an inhibitory complex, which may restrict viral replication by binding to CDK9 and sequestering it in the cytoplasm.

To further confirm the above mentioned finding, we directly examined the localization of CDK9 in pNL4-3- and pNL4-3ΔNef-transfected cells in the presence of CycK by immunofluorescence. Cells were counted, and the percentage of cells with CDK9 localized in the cytoplasm and nucleus was calculated for three independent experiments. The results presented in Fig. 7C clearly show that CDK9 was localized more in the nucleus of pNL4-3ΔNef-transfected cells as compared with pNL4-3-transfected cells. Representative immunofluorescence data are shown in Fig. 7D. Put together, all these results clearly indicate
that CycK inhibits nuclear translocation of CDK9 in the presence of Nef.

Endogenous Cyclin K Restricts CDK9 Nuclear Translocation in HIV-1-infected Cells—For further confirmation of the above findings, we then investigated the localization of CDK9 in HIV-1-infected T-cells in which Cyclin K expression was silenced. We made nuclear and cytoplasmic extracts of CycK-silenced and HIV-1-infected CEM-GFP cells and performed immunoblotting. The infected CEM-GFP cells that were transfected with CycK and CDK9 as described in the text. Cells were then mounted onto a glass slide and visualized by a confocal microscope. CDK9 translocation into the nucleus is increased in the cells transfected with the Nef-deleted molecular clone. HEK-293T cells were grown on a coverslip and were transfected with pNL4-3 or pNL4-3ΔNef along with pc-CycK. Immunofluorescence staining was performed with CDK9 and CycK antibodies. Cells stained for CDK9 in the nucleus or cytoplasm were counted and plotted as a bar diagram with data from three independent experiments. Black and gray bars indicate CDK9-stained cells transfected with pNL4-3 and pNL4-3ΔNef, respectively. D, representative immunofluorescence images of cells transfected with pNL4-3 and pNL4-3ΔNef along with pc-CycK, respectively. BF, bright field; +ve, positive.

DISCUSSION

The dependence of HIV-1 on host cellular factors is well established. The multifunctional protein Nef is capable of imparting extensive aid for HIV-1 pathogenesis by making a niche in the form of an interactome. The Nef interactome consists of multiple branches leading to critical cellular functions like receptor down-modulation, cell signaling, cell migration, apoptosis, and transcription regulation. To date, Nef has been shown to interact with several important cellular factors, and new proteins are being added frequently, unraveling its novel functions. In the present study, we show for the first time that
HIV-1 Nef interacts with Cyclin K, which was identified by yeast two-hybrid screening and was further confirmed by immunoprecipitation and co-localization studies. This interaction is not subtype-specific as both subtypes B and C Nef bind to CycK. Our data seem to corroborate previous reports (48), suggesting that despite significant sequence differences in the Nef gene from various subtypes functional properties remain relatively well conserved.

CycK was first identified as a protein that could restore progression through the cell cycle and was most closely related to human cyclins C and H (30). The kinase partner of CycK was identified as CDK9 and was reported to function as a CTD kinase in vitro (17). Recently Cyclin K was also shown to interact with CDK12 and -13 (31), but its functional implication remains to be clearly elucidated. CycT1 on the other hand is an obligate cyclin partner of CDK9, and this complex (P-TEFb) plays a key role in HIV-1 gene expression (19). As both CycK and CycT1 bind to CDK9, it was reasonable to speculate a possible role of Nef-CycK interaction in HIV-1 LTR-driven gene expression. Our results clearly show that CycK reduces LTR-driven gene expression in a dose-dependent manner. Moreover, CycK was able to reduce LTR activity only in the presence of Nef, which suggests an apparent role of Nef-CycK interaction in this phenomenon. Furthermore, CycK ectopic expression in Jurkat-1G5 cells was able to reduce LTR-mediated gene expression and thus virus production. Additionally, when endogenous CycK was knocked down using specific siRNA, the viral gene expression was enhanced, and virus particle release was also augmented in Jurkat-1G5 cells. Hence, our findings convincingly indicate that CycK might possess an HIV restricting property and that it is not a cell type-dependent phenomenon.

Endogenous CycK seems to substantially reduce HIV-1 gene expression and virus production as was evident in RNAi experiments performed with HIV-1NL4-3-infected CEM-GFP cells. In a report in 2008, Urano et al. (29) demonstrated that CycK overexpression leads to reduction in HIV-1 production from T-cell lines; however, the mechanism behind CycK-mediated

**FIGURE 8.** Cyclin K silencing leads to increased CDK9 nuclear translocation and virus production in HIV-1-infected CEM-GFP T-cells. A, Cyclin K down-regulation leads to increased CDK9 translocation into the nucleus of HIV-1-infected T cells. CEM-GFP cells were transfected with 200 nM control (sc(Cl) and CycK-specific siRNA (siCycK). 24 h post-transfection, cells were infected with HIV-1NL4-3 at 0.5 multiplicity of infection; on day 3, cells were harvested; and nuclear and cytoplasmic fractions were prepared as mentioned under “Experimental Procedures.” Equal amounts of extracts were used for immunoblotting with CDK9, CycT1, CycK, and Nef antibodies. GAPDH and Lamin A/C were used as loading controls. B, Cyclin K down-regulation leads to increased virus production from HIV-1-infected T-cells. The amount of virus present in the culture supernatant of the cells described in A was estimated by a p24 antigen capture ELISA. Nuc. Ext, nuclear extract; Cyt. Ext, cytoplasmic extract; scr, scrambled.

**FIGURE 9.** Proposed mechanistic model for Cyclin K-mediated inhibition of HIV-1 gene expression and replication. The results obtained in the present study indicate that in the presence of the Nef-Cyclin K complex CDK9-CycT1 interaction is abrogated, causing a reduction in active P-TEFb complex in the nucleus and resulting in inhibition of HIV-1 LTR-driven gene expression and replication. However, in the absence of the Nef-Cyclin K complex, CDK9 is free to bind to CycT1 and translocate into the nucleus, consequently leading to enhanced viral gene expression and replication.
reduction of HIV-1 gene expression remained unexplored. Our co-immunoprecipitation experiment along with immunofluorescence studies confirmed that CycK not only binds to CDK9 in the presence of Nef but also reduces binding of CycT1 to CDK9 and thereby restricts CDK9 translocation into the nucleus. It is worth mentioning here that our mutational studies indicate CycK binding to only myristoylated Nef. Thus, it might be hypothesized that membrane-associated Nef-CycK complex only interacts with CDK9 and thereby restricts it from translocating into the nucleus. Moreover, immunofluorescence studies with wild type and a nef-deleted HIV-1 molecular clone demonstrated that Nef is essential for CycK-mediated restriction of CDK9 nuclear translocation. Furthermore, immunoblotting analyses of nuclear and cytoplasmic extracts of CycK-specific siRNA-treated HIV-1-infected CEM-GFP cells showed increased translocation of CDK9 into the nucleus and increased virus production.

Although CDK9-CycK forms an active kinase complex that possesses CTD phosphorylating activity in vitro (17), its role in HIV-1 transcription has not been studied. CycK and CycT1 have 53 identical residues in the cyclin homology box domain, which is responsible for binding to CDK9 (44); hence, the possibility of competition between CycK and CycT1 for binding to CDK9 cannot be ruled out. The Tat-interacting region, also known as the Tat-responsive motif, located outside the cyclin homology box of CycT1, is not present in CycK (49, 50), and thus a possible role of the CDK9-CycK complex in modulating HIV-1 transcription can be ruled out. To function as a part of an active P-TEFb complex for HIV-1 gene transcription, CDK9 needs to be translocated into the nucleus, and it shuttles between the nucleus and cytoplasm under the influence of CycT1 levels in the cell (47). However, if CDK9 is restricted in the cytoplasm, the amount of active P-TEFb complex in the nucleus will be reduced, leading to inhibition of transcription. The present work in the context of Nef-CycK interaction has shown convincingly that CDK9 binding to CycK increases in the presence of Nef, thereby reducing its binding to the obligate partner CycT1. Hence, it may be hypothesized that binding to Nef is responsible for the increased affinity of CycK toward CDK9.

Thus, the present study provides insight into the molecular mechanism of CycK-mediated reduction in HIV-1 gene expression and replication. In summary, we conclude that Nef interacts with CycK, and the resulting complex leads to reduction in HIV-1 LTR-driven gene expression by interfering with CDK9 binding to CycT1 and its translocation into the nucleus. Our data also indicate that CycK function is not cell type-specific, and endogenous CycK is capable of reducing viral gene expression. Finally, our results also suggest that Nef-CycK interaction leads to reduction in the formation of active P-TEFb complex, consequently inhibiting HIV-1 LTR-driven viral gene expression and replication. Based upon our findings, we propose a model depicting how Nef-Cyclin K interaction leads to restriction of CDK9 in the cytoplasm, thereby reducing the formation of active CDK9-CycT1 complex and consequently reducing HIV-1 gene expression and virus production (Fig. 9). It is interesting to note that a recent report finds that CycK along with CDK9 binds to checkpoint proteins and thus plays an important role in replication-induced stress (51). Finally, identification of CycK as a novel Nef-interacting host factor and the consequences of this interaction provide another example for utilization of a viral protein by the host in combating the pathogen.

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Cyclin K Inhibits HIV-1 Gene Expression and Replication