Study and use of the probiotic *Lactobacillus reuteri* in pigs: a review

Chengli Hou, Xiangfang Zeng, Fengjuan Yang, Hong Liu and Shiyan Qiao*

**Abstract**

Probiotics are living microorganisms that provide a wide variety of health benefits to the host when ingested in adequate amounts. The bacterial strains most frequently used as probiotic agents are lactic acid bacteria, such as *Lactobacillus reuteri*, which is one of the few endogenous *Lactobacillus* species found in the gastrointestinal tract of vertebrates, including humans, rats, pigs and chickens. *L. reuteri* is one of the most well documented probiotic species and has been widely utilized as a probiotic in humans and animals for many years. Initially, *L. reuteri* was used in humans to reduce the incidence and the severity of diarrhea, prevent colic and necrotic enterocolitis, and maintain a functional mucosal barrier. As interest in alternatives to in-feed antibiotics has grown in recent years, some evidence has emerged that probiotics may promote growth, improve the efficiency of feed utilization, prevent diarrhea, and regulate the immune system in pigs. In this review, the characteristics of *L. reuteri* are described, in order to update the evidence on the efficacy of using *L. reuteri* in pigs.

**Keywords:** Antibiotics, Application, *Lactobacillus reuteri*, Pigs, Probiotics

**Introduction**

Antibiotics are a common additive in livestock feed which have been widely used for growth promotion and prophylaxis purposes in farm animals during the past several decades [1]. However, antibiotic resistance is a looming public health crisis. The use of antibiotics as growth promoters has been forbidden in the European Union, Korea, and Japan. Other countries including the United States and China may ban the feeding of antibiotics within the next few years. As a result, there is increasing interest concerning alternatives to in-feed antibiotics, such as probiotics, prebiotics, plant products and organic acids in the livestock industry [2].

Probiotics are living microorganisms, which, when consumed in adequate amounts, can confer a health benefit to the host [3]. In farm animals, probiotics have been shown to promote growth, improve the efficiency of feed utilization, modulate the gastrointestinal ecosystem, stimulate the immune system and protect the host from gastrointestinal tract (GIT) diseases [4]. Therefore, probiotics provide a potential alternative strategy to in-feed antibiotics [5].

The properties of probiotics are strain-specific, and suitable probiotic strains for pigs are usually selected based on some criteria including pig origin, acid and bile tolerance, their ability to adhere to intestinal cells and to colonise the intestinal tract, production of antimicrobial substances, antibiotic resistance patterns, demonstrable efficacy and safety, and stability to the conditions used in industrial processes [6-8]. The organisms most frequently used as probiotic agents are lactic acid bacteria (LAB) [9], such as *Lactobacillus*, which are a normal inhabitant of the GIT [10]. *Lactobacillus reuteri* is one of the dominant species in the GIT of vertebrates such as humans, rats, pigs and chickens [11]. It is one of the most well documented probiotic species and has been widely utilized as a probiotic in humans and animals [12-15].

In recent years, numerous probiotic strains have been used in pig production. The application of probiotics provide a potential alternative strategy to the use of antibiotics. The aim of this review is to systematically review and update the evidence on the efficacy of using *L. reuteri* in pigs.

* Correspondence: qiaoshy@mafic.ac.cn
State Key Laboratory of Animal Nutrition, China Agricultural University, Beijing 100193, China

© 2015 Hou et al; licensee BioMed Central. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.
**Characteristics of Lactobacillus reuteri**

*L. reuteri* is a heterofermentative bacterium, and is considered to be one of the few true autochthonous *Lactobacillus* species in humans and animals. Many researchers have already selected some specific *L. reuteri* strains isolated from human feces, breast milk, the human vagina, the human oral cavity, guinea pigs, rats, pigs, broilers and sourdough. There is now mounting evidence to show that selected *L. reuteri* strains have probiotic characteristics, and can provide health benefits to their hosts. We have constructed a summary table (Table 1), in order to provide an overview of the reported *L. reuteri* strains used as probiotics.

**Probiotic properties**

Probiotic bacteria encounter various stresses after ingestion by the host, including exposure to a low pH in the stomach and contact with bile in the small intestine. *L. reuteri* I5007, initially known as *Lactobacillus fermentum* I5007, was selected from over 7,000 native Lactobacilli colonies according to criteria including resistance to heat, low pH, copper, and bile salts, as well as storage stability and antagonism to pathogenic agents [16]. Other *L. reuteri* strains also show resistance to low pH and bile salts [12,17-19].

Adhesion of a probiotic strain to the host GIT is important for bacterial colonization, pathogen exclusion,

---

**Table 1 The strains and probiotic characteristics of reported *L. reuteri***

| Strain       | Source          | Characteristics                                                                 | Reference |
|--------------|-----------------|---------------------------------------------------------------------------------|-----------|
| *L. reuteri* I5007 | piglets         | strong adhesion, competitiveness against pathogens; improved pig performance; immune function and antioxidant status; alleviated the weaning stress syndrome; modulated gut microflora | [21,35,46-48,50,52] |
| *L. reuteri* NCIMB 30242 | pig      | improve gastrointestinal health; inhibit sterol absorption; increase mean circulating 25-hydroxyvitamin D | [61-63]   |
| *L. reuteri* ATCC 53608 | pig      | recognize immunoglobulins                                                       | [24,41]   |
| *L. reuteri* BSA 131    | pig      | resistance to pH, oxgall and antibiotics, and antimicrobial activities against enteric pathogenic, improved pig performance | [12]      |
| *L. reuteri* Pg4        | broilers   | tolerate acid and bile salts; inhibit pathogenic bacteria; adhere to intestinal epithelial cells; improved broiler performance | [17]      |
| *L. reuteri* ATCC 55730 | breast milk | colonize the intestinal tract; resistance to tetracycline and lincomycin; maintain intestinal health; prevent diarrhea; modulate the immune system; used in treatment of *Helicobacter pylori* | [13,14,64,65] |
| *L. reuteri* DSM 17938  | ATCC 55730    | a daughter strain derived from ATCC 55730 and has the same properties as ATCC 55730 | [42]      |
| *L. reuteri* ATCC PTA 4659 | breast milk | partly prevent diet-induced obesity possibly via the mechanism of inducing liver expression of Cpt1a | [66]      |
| *L. reuteri* ATCC PTA 6475 | breast milk | protect mice from disease manifestations of enterohemorrhagic *E. coli*         | [67]      |
| *L. reuteri* ATCC PTA 5289 | oral cavity | improve oral malodour; reduce the number of selected periodontal pathogens in the subgingival microbiota | [68,69]   |
| *L. reuteri* DPC16      | human feces   | produce reuterin                                                                | [29]      |
| *L. reuteri* DSM 20016  | human feces   | produce reuterin                                                                | [30]      |
| *L. reuteri* JCM 1112   | human feces   | produce reuterin and cobalamin                                                  | [31]      |
| *L. reuteri* RC-14      | human vagina  | produce hydrogen peroxide; adhere to uroepithelial cells and inhibit uropathogens; modulate immunity | [28,70]   |
| *L. reuteri* GMN-32     | -              | regulate blood glucose levels, protect cardiomyocytes and prevent diabetic cardiomyopathy in diabetes mellitus rats | [71]      |
| *L. reuteri* DSMZ 17648 | -              | reduce the load of *Helicobacter pylori*                                        | [72]      |
| *L. reuteri* GMNL-263   | -              | ameliorate hepatic steatosis observed in high fructose treated rats; protect streptozotocin-induced diabetic rats from hyperglycermia-enhanced renal fibrosis | [73,74]   |
| *L. reuteri* R2         | -              | strong inhibitory activity against the dermatophyte *Trichophyton tonsurans*     | [75]      |
| *L. reuteri* TD1        | rat            | not produce reuterin, exhibit a similar onset of type 1 diabetes                 | [76]      |
| *L. reuteri* 100-23     | rat            | stimulate the development of regulatory T cells; transiently activates intestinal epithelial cells | [36,77]   |
| *L. reuteri* BR11       | guinea pig     | unique antioxidant properties, show promise in the treatment of experimental colitis | [37]      |
| *L. reuteri* CRL1098    | sourdough      | produce vitamin B<sub>12</sub>                                                  | [34]      |
| *L. reuteri* LTH2584    | sourdough      | produce reutericyclin                                                          | [32]      |

Note: *L. reuteri* I5007, initially known as *L. fermentum* I5007; *L. reuteri* BR11, initially known as *L. fermentum* BR11; *L. reuteri* RC-14, initially known as *L. fermentum* RC-14.
and interaction with host cells for the protection of epithelial cells or immune modulation [20]. Several studies have demonstrated that *L. reuteri* have the capacity to colonize, and can adhere to mucin and intestinal epithelial cells [17,21-23]. *L. reuteri* DSM 15007 shows strong adhesion to Caco-2 cells, IEC-6 cells, IPEC-J2 cells, and porcine intestinal mucus [15,21]. The possible mechanism for *L. reuteri* adherence and colonization involved in adhesion, has been linked to mucus-binding protein [24], surface protein [22], D-alanyl-LTA [25], exopolysaccharide [26], glucosyltransferase A and inulosucrase [27].

*L. reuteri* has been reported to produce a variety of antimicrobial substances such as lactic acid, hydrogen peroxide [28], reuterin [29-31], and reutericyclin [32], which have beneficial effects for the host organism. *L. reuteri* strains have been demonstrated to inhibit the *in vitro* growth of many enteric pathogens, including *Escherichia coli*, *Salmonella* Typhimurium, *Staphylococcus epidermidis*, *Staphylococcus aureus*, *Helicobacter pylori*, and rotavirus [12,19,33]. In addition, *L. reuteri* can produce vitamin B12 [31,34], and has the capacity to *de novo* synthesize L-lysine and folic acid based on a computer simulation model [15].

*L. reuteri* exhibited free radical-scavenging capacity *in vitro* [35], and encoded various antioxidant enzymes [15]. Studies in animals and humans have shown that oral administration of *L. reuteri* reduced the incidence and the severity of diarrhea, decreased visceral pain, prevented colic and necrotic enterocolitis, maintained a functional mucosal barrier, and induced colonization and immunomodulation [36-39].

**Safety and stability aspects**

*L. reuteri* has the most extensive safety assessment record of any probiotic strain. A number of studies conducted both *in vivo* and *in vitro* indicate that *L. reuteri* is safe for human consumption, even in large amounts [38,40]. However, as is the case for all other species of LAB, plasmids can be found in some strains of *L. reuteri* [15,41,42], and some of these plasmids have been shown to encode for antibiotic resistance genes [42]. According to the European Food Safety Authority, probiotics should not contain known antibiotic resistance traits. *L. reuteri* ATCC 55730 is a commercially available probiotic strain which has been found to carry potentially transferable resistance traits for tetracycline and lincomycin. Therefore, it has been replaced by *L. reuteri* DSM 17938, a strain where the two resistance plasmids have been removed without losing any probiotic characteristics [42].

Probiotic strains must be able to resist any adverse conditions encountered during industrial production in order to survive [43]. *L. reuteri* is sensitive to heat, and therefore, freeze-drying is commonly used for maintaining the stability of *L. reuteri*. Subjecting *L. reuteri* to a higher fermentation temperature (47°C) or a neutral pH (pH 6.7) has been shown to increase the survival of *L. reuteri* during subsequent freeze-drying [44].

**Strains**

Not all *L. reuteri* strains are the same or provide a beneficial response, and the evolution of *L. reuteri* with vertebrates resulted in the emergence of host specialization [45]. Probiotic strains need to be carefully chosen, and evaluated for their safety and effectiveness using *in vitro* assays, animal models, and clinical trials. There are numerous strains of *L. reuteri*, which have some minor differences that make them unique (Table 1).

*L. reuteri* DSM 17938, *L. reuteri* NCIMB 30242, *L. reuteri* ATCC PTA 6475 which are of human origin are the most commonly used in dietary supplements and have been researched the most. In pigs, *L. reuteri* I5007, was isolated from the colonic mucosa of healthy weaning piglets, and has been demonstrated in several studies to have probiotic properties [15,21,35,46-49].

**Applications of probiotic *L. reuteri* for pigs**

In pigs, the administration of *L. reuteri* has been shown to have beneficial effects on performance, prevention of diarrhea, stress relief, altered gut microbiota, and immunomodulation. The applications of *L. reuteri* for pigs are listed in Table 2. Noticeably, *L. reuteri* is mainly used in neonatal piglets and during the post-weaning period.

**Improved performance**

In the pig industry, the use of probiotics improves intestinal health which can improve pig performance. Supplementation of *L. reuteri* has resulted in improved growth and feed efficiency in neonatal and growing pigs. Liu et al. [46] reported that *L. reuteri* I5007 (6 × 10⁹ CFU/d) supplementation increased average daily gain (ADG) in formula-fed piglets. Wang et al. [47] found that administration of *L. reuteri* I5007 significantly increased weight gain and feed conversion compared with weaned pigs fed without *L. reuteri* I5007. Also, Wang et al. [50] reported that weaned piglets supplemented with *L. reuteri* had faster growth and higher feed intakes than unsupplemented piglets. However, feed conversion was unaffected by *L. reuteri* supplementation. In addition, Wang et al. [48] showed that dietary supplementation with *L. reuteri* or aureomycin significantly improved the performance of weaning piglets, and there was no difference between the two feed additives.

Other studies using *L. reuteri* BSA131 tended to show improved ADG and feed conversion in weaned pigs [12]. Wang et al. [51] also reported that supplementation with *L. reuteri* X-1 increased ADG and feed conversion. Yu et al. [52] determined the influence of different levels of *L. reuteri* I5007 on performance, nutrient digestibility and immunity of weaned pigs. The results demonstrated...
that the ideal supplemental concentration of *L. reuteri* was $5.8 \times 10^7$ CFU/g feed.

**Prevention of diarrhea**

Diarrhea is one of the most frequent causes of heavy economic losses in swine operations [53]. The effect of *L. reuteri* against diarrhea in pigs was confirmed in several reports [16,46,47,54,55]. Diarrhea incidence was lower in piglets fed *L. reuteri* 15007 compared with a control [46]. Enterotoxigenic *E. coli* (ETEC) are a major cause of diarrhea in neonatal and weaned pigs [55]. Huang *et al.* [16] showed that a native Lactobacilli complex preparation (including *L. gasseri*, *L. reuteri*, *L. acidophilus* and *L. fermentum*) could effectively prevent weaning piglet diarrhea when administered before challenge with an *E. coli* solution (serovars K99, K88 and 987P at a ratio of 1:1:1). Wang *et al.* [47] reported that 12, 24, and 48 h after challenge, pigs challenged with *E. coli* had mild diarrhea and mild fecal scores. Supplementation of *L. reuteri* 15007 did not alleviate these effects. Only on day 10, did feeding *L. reuteri* 15007 decrease the occurrence of diarrhea. Chen *et al.* [54] demonstrated that reuteran produced by *L. reuteri* may prevent piglet diarrhea by reducing adhesion of ETEC K88.

## Table 2: The application of probiotic *L. reuteri* in pigs

| Strain | Dose | Animal | Significant results | Reference |
|--------|------|--------|----------------------|-----------|
| *L. reuteri* 15007 | $6 \times 10^7$ CFU/d | newborn piglets | increased average daily gain; reduced diarrhea incidence; affected the colonic microbial communities, in particular, reduced numbers of *Clostridium* sp; reduced mRNA expression of IL-1β in the ileum | [46] |
| *L. reuteri* 15007 | $2 \times 10^8$ CFU/d | weaned pigs | increased weight gain and feed conversion; decreased the occurrence of diarrhea; enhanced T-cell differentiation and induced cytokine expression in the ileum | [47] |
| *L. reuteri* 15007 | $2 \times 10^9$ CFU/d | weaned pigs | had faster growth and higher feed intakes; improved the anti-oxidative defence system and alleviated damage caused by diquat | [50] |
| *L. reuteri* 15007 | $5.8 \times 10^7$ CFU/g | weaned pigs | increased weight gain, feed intake and apparent crude protein digestibility; increased serum specific anti-OVA IgG level | [52] |
| Lactobacilli complex | $10^7$ CFU/g | weaned pigs | increased weight gain and feed intake compared with carbadox; prevented diarrhea; decreased *E. coli* and anaerobe counts, increased Lactobacilli and anaerobe counts in digesta and mucosa | [16] |
| *L. reuteri* BSA131 | $2 \times 10^6, 2 \times 10^8$ CFU/g | weaned pigs | improved weight gain and feed conversion; decreased the number of enterobacteria in the feces | [12] |
| *L. reuteri* X-1 | $10^8$ CFU/g | weaned pigs | improved weight gain and feed conversion; decreased serum IgG and IgM concentrations, increased serum DAO and D-lactate concentrations | [51] |
| *L. reuteri* 15007 | $1.02 \times 10^8$ CFU/g | growing pigs | increased total antioxidant capacity | [35] |
| *L. reuteri* 15007 | $10^8$ CFU/g | weaned pigs | increased weight gain, neither body weights nor weight gains differed between the *L. reuteri* and aureomycin groups; alleviated weaning stress syndrome | [48] |
| *L. reuteri* and *L. plantarum* complex | $10^6$ CFU/g | weaned pigs | increased apparent total tract digestibility of nitrogen, gross energy, and fecal Lactobacillus concentration; decreased fecal gas emission, diarrhea score, and *E. coli* concentration | [78] |
| *L. reuteri* 357 and *L. plantarum* 4.1 | $10^{10}$ CFU/d | sows and piglets | were found in the faeces; decreased the population of Enterobacteriaceae; decreased β-glucuronidase activity of all pigs | [79] |

Note: *L. reuteri* 15007 was initially known as *L. fermentum* 15007.

Lactobacilli complex including *L. gasseri*, *L. reuteri*, *L. acidophilus* and *L. fermentum* (renamed *L. reuteri* 15007).

**Alleivate stress**

Pigs in industrial farming systems are frequently exposed to oxidative stress, which results in decreased performance and reduced immune function. *L. reuteri* has been shown to be effective in scavenging free radicals *in vitro*, and could be used to alleviate oxidative stress [35,37]. Wang *et al.* [35] reported that supplementation of *L. reuteri* 15007 improved the antioxidant status of growing-finishing pigs (from 50 to 90 kg) as evidenced by increased levels of antioxidant enzymes such as superoxide dismutase and glutathione peroxidase, and decreased levels of malondialdehyde. Wang *et al.* [50] determined the anti-oxidative effect of *L. reuteri* 15007 in weaning piglets using an oxidative stress model induced by diquat. Their results showed that diquat-injection decreased the performance of weaning pigs and increased plasma levels of cortisol, adrenaline, corticosteroid, and malondialdehyde. *L. reuteri* supplementation alleviated oxidative stress and enhanced the performance of weaning pigs.

Weaning is one of the most stressful periods that results in gastrointestinal, immunological, and behavioral changes [56]. Wang *et al.* [48] demonstrated that *L. reuteri* 15007...
alleviated weaning stress syndrome by enhancing the levels of proteins involved in energy metabolism, lipid metabolism, cell structure and mobility, protein synthesis, and immune response, thereby facilitating cellular proliferation and depressing apoptosis.

Modulation of gut microbiota

*L. reuteri* in neonatal piglets can be used to support the development of a stable microbiota, to stimulate the immune system and to prevent diarrheal diseases. During the weaning and post-weaning periods, *L. reuteri* is used in pigs to modulate the gastrointestinal microbiota as it aims to prevent post-weaning diarrhea and stimulate growth. Liu et al. [46] reported that *L. reuteri* I5007 plays a positive role in gut development in neonatal piglets by modulating the microbial composition and intestinal development. Denaturing gradient gel electrophoresis (DGGE) revealed that *L. reuteri* I5007 affected the colonic microbial communities on day 14 and, in particular, reduced numbers of *Clostridium* spp. In weaning pigs, administration of *L. reuteri* BSA131 decreased the number of enterobacteria in the feces [12]. Huang et al. [16] showed that a Lactobacilli compound (including *L. gasseri*, *L. reuteri*, *L. acidophilus* and *L. fermentum*) significantly decreased *E. coli* and aerobe counts, and increased Lactobacilli and anaerobe counts in the digesta and mucosa of most sections of the GIT compared with a control group.

In addition, oral administration of *L. reuteri* 15007 not only increased the concentration of butyrate and other branched chain fatty acids but also decreased *Clostridium* strains accompanied by a lowered pH in the colonic digesta [46]. This indicates that administration of *L. reuteri* modulates gut microbiota, and thereby affected the microbial metabolites.

Immunomodulation

Probiotics such as *L. reuteri* may stimulate or suppress innate immune responses via several mechanisms including modulation of pro-inflammatory cytokines. *L. reuteri* strains can be divided into two subsets, immunosuppressive (ATCC PTA 6475 and ATCC PTA 5289) and immunostimulatory strains (ATCC 55730 and CF48-3A), and each subset has potential therapeutic value [57]. The effects of *L. reuteri* on immunomodulation were documented in pigs. Wang et al. [47] reported that oral administration of *L. reuteri* I5007 could enhance T-cell differentiation and induce ileal cytokine expression, which suggests that this probiotic strain could modulate immune function in weaned piglets. Yu et al. [52] showed that *L. reuteri* I5007 supplementation increased serum specific anti-OVA IgG levels. In neonatal piglets, *L. reuteri* has been found to decrease the mRNA expression of IL-1β in the ileum [46]. Azevedo et al. [58] found that *L. reuteri* combined with *L. acidophilus* could help to maintain

---

**Figure 1** Mechanisms of *L. reuteri* modulating in the gut.

1. *L. reuteri* can produce a variety of antimicrobial substances (AMS) such as lactic acid, and reuterin [28-30].
2. *L. reuteri* has the capacity to colonize, and can adhere to mucin and intestinal epithelial cells [17,21,22].
3. *L. reuteri* has been shown to stimulate or suppress innate immune responses by affected the production of cytokines in macrophages (M), monocytes, and dendritic cells (DCs). The modulation of dendritic cells by *L. reuteri* has been shown to be mediated through dendritic cell-specific intercellular adhesion molecule-3-grabbing non-integrin (DC-SIGN) and promote development of regulatory T cells producing high amounts of interleukin-10 (IL-10) and transforming growth factor-β (TGF-β) [59,60].
4. *L. reuteri* has been reported affected the colonic microbial communities and short chain fatty acid (SCFA) concentration [46]. Please see text for details and references.
immunological homeostasis in neonatal gnotobiotic pigs infected with human rotavirus by regulating TGF-β production.

Conclusions and perspectives

In conclusion, *L. reuteri* is a probiotic bacteria that is one of the few true autochthonous *Lactobacillus* species. Numerous studies have demonstrated that they can positively improve performance, prevent diarrhea, alleviate stress, alter gastrointestinal microbiota, regulate the immune system, and thereby improve pig performance and health. The beneficial effects of *L. reuteri* in pigs have been related to different modes of action. The improvements in pig performance of supplemental *L. reuteri* are mostly due to the fact that *L. reuteri* has the ability to elongate the GIT, produce antimicrobial substances and stimulate the intestinal immune system (Figure 1), thereby promoting nutrient metabolism and improve health. However, a clear mode of action has yet to be described. It appears from the data presented that the beneficial effects of *L. reuteri* are strain specific. It will be important to select more powerful or targeted strains. Unfortunately, the viability of *L. reuteri* is a key criteria for developing *L. reuteri* products. To expand the probiotic *L. reuteri* application in pigs, care must be taken during processing techniques such as microencapsulation to maintain bacterial stability.

Pig husbandry has entered an era when the use of antibiotics is increasingly unwelcome. Probiotics, which are a potential alternative to in feed antibiotics, can expect a promising future. Besides selection of excellent strains and improved processing techniques, more research, especially in the form of well-designed animal trials, is needed to evaluate the efficacy of *L. reuteri*. More studies are also needed to explore the mechanisms of action of *L. reuteri* in pigs. An important fact is that *L. reuteri* added to pig diets may potentially help improve performance. With evolving knowledge, effective use of *L. reuteri* will be possible in the future.

Competing interests

The authors declare that they have no competing interests.

Authors’ contributions

CH and SQ carried out the literature study and drafted the manuscript. XZ, FY and HL critically evaluated the manuscript. All authors read and approved the final manuscript.

Acknowledgments

This research was financially supported by the National Natural Science Foundation of China (Grant Number 31420103908).

Received: 26 October 2014 Accepted: 26 March 2015
Published online: 09 April 2015

References

1. Thacker PA. Alternatives to antibiotics as growth promoters for use in swine production: a review. J Anim Sci Biotechnol. 2013;4:35.
2. Wang Z, Zeng X, Mo Y, Smith K, Guo Y, Lin J. Identification and characterization of a bile salt hydrolase from *Lactobacillus salivarius* for development of novel alternatives to antibiotic growth promoters. Appl Environ Microbiol. 2012;78:8795–802.
3. Guamer F, Schaafsma GJ. Probiotics. Int J Food Microbiol. 1998;39:237–8.
4. Gaggia F, Mattarelli P, Biavati B. Probiotics and prebiotics in animal feeding for safe food production. Int J Food Microbiol. 2010;141 Suppl 1:515–28.
5. Cho HJ, Zhao PY, Kim IH. Probiotics as a dietary additive for pigs: a review. J Anim Vet Adv. 2011;10:217–34.
6. Kalilasapaty K, Chin J. Survival and therapeutic potential of probiotic organisms with reference to *Lactobacillus acidophilus* and *Bifidobacterium spp.* Immunol Cell Biol. 2000;78:80–8.
7. Tuomola E, Crittenden R, Payne M, Isolauri E, Salminen S. Quality assurance criteria for probiotic bacteria. Ann Clin Nutr. 2001;3:393S–8.
8. Shoikyazdan P, Sioe CC, Kalavathy R, Liang JB, Altheen NB, Jahorni MF, et al. Probiotic potential of *Lactobacillus* strains with antimicrobial activity against some human pathogenic strains. Biomed Res Int. 2014;2014:927268.
9. Land MH, Rooster-Stevens K, Woods CR, Cannon ML, Crotta J, Shetty AK. *Lactoba- cilus* sepsis associated with probiotic therapy. Pediatrics. 2005;115:178–81.
10. Tannock GW, Tilsala-Timisjarvi A, Rottmans S, Ng J, Munro K, Aaltonesa T. Identification of *Lactobacillus* isolates from the gastrointestinal tract, silage, and yoghurt by 16S-23S rRNA gene intergenic spacer region sequence comparisons. Appl Environ Microbiol. 1999;65:464–7.
11. Of Pl, Benson AK, Peterson DA, Patil PB, Moriyama EN, Roos S, et al. Diversification of the gut symbiont *Lactobacillus reuteri* as a result of host-driven evolution. ISME J. 2010;4:377–87.
12. Chang YH, Kim JK, Kim HJ, Kim YB, Park YH. Selection of a potential probiotic *Lactobacillus* strain and subsequent in vivo studies. Antonie Van Leeuwenhoek. 2001;80:193–9.
13. Cocconullo P, Strisciuglio C, Martinelli M, Miele E, Greco L, Taiiano A. *Lactobacillus reuteri* (DSM 17938) in infants with functional chronic constipation: a double-blind, randomized, placebo-controlled study. Pediatr. 2010;157:598–602.
14. Francavilla R, Lionetti E, Castellanza S, Ciruzzi F, Indrio F, Macciale A, et al. Randomised clinical trial: *Lactobacillus reuteri* DSM 17938 vs. placebo in children with acute diarrhea-a double-blind study. Aliment Pharmacol Ther. 2012;36:363–9.
15. Hou C, Wang Q, Zeng X, Yang F, Zhang J, Liu H, et al. Complete genome sequence of *Lactobacillus reuteri* IS007, a probiotic strain isolated from healthy piglet. J Biotechnol. 2014;179:63–4.
16. Huang CH, Qiao SY, Li DF, Piao XS, Ren JP. Effects of *Lactobacillus* on the performance, diarrhea incidence, VFA concentration and gastrointestinal microbial flora of weaning pigs. Asian-Aust J Anim Sci. 2004;17:401–9.
17. Yu B, Liu JR, Chou MY, Hsu YR, Chiou PWS. The effects of probiotic *Lactobacillus reuteri* Pg4 strain on intestinal characteristics and performance in broilers. Asian-Aust J Anim Sci. 2007;20:1243–51.
18. Whitehead K, Versavitc J, Roos S, Britton RA. Genomic and genetic characterization of the bile stress response of probiotic *Lactobacillus* species. ATCC 55730. Appl Environ Microbiol. 2008;74:1812–9.
19. Seo BJ, Mun MR, Rejish Kumar J, Kim CI, Lee J, Chang YH, et al. Bile tolerant *Lactobacillus reuteri* isolated from pig feces inhibits enteric bacterial pathogens and porcine rotavirus. Vet Res Commun. 2010;34:323–33.
20. Lebeer S, Vanderleyden J, De Keersmaecker SC. Genes and molecules of lactobacilli supporting probiotic action. Microbiol Mol Biol Rev. 2012;76:728–64.
21. Li XX, Yue LY, Guan XF, Qiao SY. The adhesion of putative probiotic lactobacilli to cultured epithelial cells and porcine intestinal mucus. J Appl Microbiol. 2008;104:1082–91.
22. Wang B, Wei H, Yuan J, Li Q, Li Y, Li N, et al. Identification of a surface protein from *Lactobacillus reuteri* JCM1081 that adheres to pig gastric mucin and human enterocyte-like HT-29 cells. Curr Microbiol. 2008;57:33–8.
23. Miyoshi Y, Okada S, Uchimura T, Satoh E. A mucus adhesion promoting protein, MapA, mediates the adhesion of *Lactobacillus reuteri* to Caco-2 human intestinal epithelial cells. Biosci Biotechnol Biochem. 2006;70:1622–8.
24. Mackenzie DA, Jeffers F, Parker ML, Vibert-Vallet A, Bongaerts RJ, Roos S, et al. Stain-specific diversity of mucus-binding proteins in the adhesion and aggregation properties of *Lactobacillus reuteri*. Microbiology. 2010;156:3638–78.
25. Walker JB, Doih MK, Rockell C, Hermann C, Pfitznermaier M, et al. D-alanyl ester depletion of teichoic acids in *Lactobacillus reuteri* 100–23 results in impaired colonization of the mouse gastrointestinal tract. Environ Microbiol. 2007;9:1750–60.
26. Wang Y, Gänzle MG, Schwab C. Escherichia coli binds to porcine enterocytes. Appl Environ Microbiol. 2010;76:4863–6.
27. Walter J, Schwab C, Loach DM, Ganzle M, Tannock GW. Gluconolactone reuteri Lactobacillus reuteri TMW1.106 contribute to cell aggregation, in vitro biofilm formation, and colonization of the mouse gastrointestinal tract. Microbiology. 2008;154:72–80.

28. Martinez RC, Seney SL, Summers KL, Nomizo A, De Marinis EC, Reid G. Effect of Lactobacillus rhamnosus GR-1 and Lactobacillus reuteri RC-14 on the ability of Candida albicans to infect cells and induce inflammation. Microbiol. Mol. Biol. Rev. 2009;73:487–95.

29. Bian L. In vitro antimicrobial and safety study of Lactobacillus reuteri DPC16 for probiotic concept. Master thesis: Massey University. 2008.

30. Armin HM, Hashem AM, Ashour MS, Hatti-Kaul R. 1,2 Propanediol utilization by Lactobacillus reuteri DSM 20016, role in bioconversion of glycerol to 1,3 propanediol, 3-hydropyrorinonealdehyde and 3-hydroxyproprionic acid. J Genet Eng Biotechnol. 2013;11:53–9.

31. Morita H, Toh H, Fukuda S, Horikawa H, Oshima K, Suzuki T, et al. Comparative genome analysis of Lactobacillus reuteri and Lactobacillus fermentum reveal a genomic island for reuterin and cobalamin production. DNA Res. 2008;15:151–61.

32. Gärde MG, Hölzl J, Walter J, Jung G, Hammes WP. Characterization of reuterinycilin produced by Lactobacillus reuteri LTH2584. Appl Environ Microbiol. 2000;66:4325–33.

33. Mukai T, Asaka T, Sato E, Mori K, Matsumoto M, Ohori H. Inhibition of binding of Helicobacter pylori to the glycolipid receptors by probiotic Lactobacillus reuteri: FEMS Immunol Med Microbiol. 2002;32:105–10.

34. Taranto MP, Vera JL, Hugenholtz J, De Valdez GF, Senna F. Lactobacillus reuteri CR-1098 produces cobalamin. J Bacteriol. 2003;185:5643–7.

35. Wang AN, Yi XW, Yu HF, Dong B, Qiao SY. Free radical scavenging activity of Lactobacillus reuteri in vitro and its antioxidative effect on growing-finishing pigs. J Appl Microbiol. 2009;107:1140–8.

36. Hoffmann M, Rath E, Holzwimmer G, Quintanilla-Martinez L, Loach DM, Tannock G, et al. Lactobacillus reuteri 100–23 transiently activates intestinal epithelial cells of mice that have a complex microbiota during early stages of colonization. J Nutr. 2008;138:1684–91.

37. Atkins HL, Geier MS, Prisciandaro LD, Pattanaik AK, Forder RE, Turner MS, et al. Effects of a Lactobacillus reuteri BR11 mutant deficient in the cystine-transport system in a rat model of inflammatory bowel disease. Dig Dis Sci. 2012;57:713–9.

38. Ulbratska M, Szajewska H. The efficacy of Lactobacillus reuteri DSM 17938 in infants and children: a review of the current evidence. Eur J Pediatr. 2014;173:1327–37.

39. Dickved J, Scheiber O, Willing B, Petersson J, Rang S, Phillipson M, et al. Lactobacillus reuteri maintains a functional mucosal barrier during DSS treatment despite mucus layer dysfunction. PLoS One. 2012;7, e46399.

40. Lee DY, Seo YS, Rayamajhi N, Kang ML, Lee SI, Yoo HS. Isolation, development and alters the intestinal microbiota in formula-fed piglets. J Anim Sci Biotechnol. 2013;4:19.

41. Jones ML, Martoni CJ, Ganevsky JG, Gulemankhel J, Ghali P, Prakash S. Improvement of gastrointestinal health status in subjects consuming Lactobacillus reuteri NCIMB 30242 capsules: a post-hoc analysis of a randomized controlled trial. Expert Opin Biol Ther. 2013;13:1643–51.

42. Jones ML, Martoni CJ, Prakash S. Cholesterol lowering and inhibition of sterol absorption by Lactobacillus reuteri NCIMB 30242, a randomized controlled trial. J Clin Lipidol. 2012;6:1234–41.

43. Jones ML, Martoni CJ, Prakash S. Oral supplementation with probiotic L. reuteri NCIMB 30242 increases mean circulating 25-hydroxyvitamin D: a post hoc analysis of a randomized controlled trial. J Clin Endocrinol Metab. 2013;98:2944–51.

44. Dommels YE, Kemperman RA, Zebbreges YE, Draisma RB, Jol A, Wouters DA, et al. Survival of Lactobacillus reuteri DSM 17938 and Lactobacillus rhamnosus GG in the human gastrointestinal tract with daily consumption of a low-fat probiotic spread. Appl Environ Microbiol. 2009;75:198–204.

45. Liu Y, Fatheree NY, Mangalat N, Rhoads JM. Human-derived probiotic Lactobacillus reuteri strains differentially reduce intestinal inflammation. Am J Physiol Gastrointest Liver Physiol. 2010;299:G1087–96.

46. Jones SE, Versalovic J. Probiotic Lactobacillus reuteri biofilms produce antimicrobial and anti-inflammatory factors. BMC Microbiol. 2009;9:35.

47. Azevedo MS, Zhang W, Wen K, Gonzalez AM, Saff LJ, Yousef AE, et al. Lactobacillus acidophilus and Lactobacillus reuteri modulate cytokine responses in gnotobiotic pigs infected with human rotavirus. Benef Microbes. 2012;33:3–42.

48. Walter J, Britton RA, Roos S. Host-microbial symbiosis in the vertebrate gastrointestinal tract and the Lactobacillus reuteri paradigm. Proc Nat Acad Sci U S A. 2011;108 Suppl 1:4645–52.

49. Liu Y, Fatheree NY, Mangalat N, Rhoads JM. Human-derived probiotic Lactobacillus reuteri strains differentially reduce intestinal inflammation. Am J Physiol Gastrointest Liver Physiol. 2010;299:G1087–96.

50. Wang AN, Cai CJ, Zeng XF, Zhang FR, Zhang GL, Thacker PA, et al. Dietary supplementation with Lactobacillus reuteri DSM 5007 improves the anti-oxidative activity of weaning piglets challenged with diquat. J Appl Microbiol. 2013;114:1582–91.

51. Wang SP, Yang LY, Tang XS, Cai LC, Liu G, Kong XF, et al. Dietary supplementation with high-dose Baccillus subtilis or Lactobacillus reuteri modulates cellular and humoral immunology and improves performance in weaned pigs. J Food Agric Environ. 2011;9:181–7.

52. Yu H, Wang A, Li X, Qiao S. Effect of viable Lactobacillus reuteri on the growth performance, nutrient digestibility and immunity of weaned pigs. J Anim Feed Sci. 2008;17:61–9.

53. Fairbrother JM, Nadeau E, Gyles CL. Escherichia coli in postweaning diarrhea in pigs: an update on bacterial types, pathogenesis, and prevention strategies. Anim Health Res Rev. 2005;6:17–39.

54. Chen XY, Woodward A, Zijlstra RT, Ganzle MG. Exopolysaccharides synthesised by Lactobacillus reuteri protect against enterotoxigenic Escherichia coli in piglets. Appl Environ Microbiol. 2014;80:7572–60.

55. Francis DH. Enterotoxigenic Escherichia coli infection in pigs and its diagnosis. J Swine Health Prod. 2002;10:171–5.

56. Campbell JM, Greamesh JD, Polo J. The biological stress of early weaned pigs. J Anim Sci Biotechnol. 2013;4:19.
70. Kohler GA, Assefa S, Reid G. Probiotic interference of Lactobacillus rhamnosus GR-1 and Lactobacillus reuteri RC-14 with the opportunistic fungal pathogen Candida albicans. Infect Dis Obstet Gynecol. 2012;2012:636474.

71. Lin CH, Lin CC, Shibu MA, Liu CS, Kuo CH, Tsai FJ, et al. Oral Lactobacillus reuteri GMN-32 treatment reduces blood glucose concentrations and promotes cardiac function in rats with streptozotocin-induced diabetes mellitus. Br J Nutr. 2014;111:598–605.

72. Mehling H, Busjahn A. Non-viable Lactobacillus reuteri DSMZ 17648 (Pylopass) as a new approach to Helicobacter pylori control in humans. Nutrients. 2013;5:3062–73.

73. Hsieh FC, Lee CL, Chai CY, Chen WT, Lu YC, Wu CS. Oral administration of Lactobacillus reuteri GWNL-263 improves insulin resistance and ameliorates hepatic steatosis in high fructose-fed rats. Nutr Metab (Lond). 2013;10:35.

74. Lu YC, Yin LT, Chang WT, Huang JS. Effect of Lactobacillus reuteri GWNL-263 treatment on renal fibrosis in diabetic rats. J Biosci Bioeng. 2010;110:709–15.

75. Guo J, Mauch A, Galle S, Murphy P, Arendt EK, Coffey A. Inhibition of growth of Trichophyton tonsurans by Lactobacillus reuteri. J Appl Microbiol. 2011;111:474–83.

76. Valladares R, Sankar D, Li N, Williams E, Lai KK, Abdelgeliel AS, et al. Lactobacillus johnsonii N6.2 mitigates the development of type 1 diabetes in BB-DP rats. PLoS One. 2010;5, e10507.

77. Livingston M, Loach D, Wilson M, Tannock GW, Baird M. Gut commensal Lactobacillus reuteri 100–23 stimulates an immunoregulatory response. Immunol Cell Biol. 2010;88:99–102.

78. Zhao PY, Kim IH. Effect of direct-fed microbial on growth performance, nutrient digestibility, fecal noxious gas emission, fecal microbial flora and diarrhea score in weanling pigs. Anim Feed Sci Technol. 2015;200:86–92.

79. De Angelis M, Siragusa S, Caputo L, Ragni A, Burzigotti R, Gobbetti M. Survival and persistence of Lactobacillus plantarum 4.1 and Lactobacillus reuteri 357 in the gastrointestinal tract of pigs. Vet Microbiol. 2007;123:133–44.