Morphology and taxonomic status of *Aedes aegypti* populations across Senegal

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Abstract

*Aedes aegypti* is the primary vector of dengue, Zika, yellow fever and chikungunya viruses to humans. In Africa, two subspecies, *Ae. aegypti aegypti* (*Aaa*) and *Ae. aegypti formosus* (*Aaf*) have been described. Until very recently, it was considered that the two forms were sympatric in East Africa and that only *Aaf* was present in Central and West Africa. However, recent data suggests that *Aaa* was also common in Senegal without any clear evidence of genetic differences with *Aaf*. This study was carried out in different *Ae. aegypti* populations from Senegal to better clarify their taxonomic status. The larvae, pupae and eggs were collected between July and September 2018 and reared individually to adult stage. For each population, *F*1 progeny from eggs laid by a single female *F*0 were reared as sibling samples. The number of pale scales on the first abdominal tergite (*T*1) and the basal part of the second tergite (*T*2) were counted. Individuals with no pale scale on *T*1 were classified as *Aaf* while those with at least one pale scale on this tergite were classified as *Aaa*. The morphological variations within families of *Aaf* were studied across 4 generations. In total, 2400 individuals constituting 240 families were identified, of which 42.5% were heterogeneous (families with both forms). Multivariate statistical analysis of variance including *T*1 and *T*2 data together showed that populations were significantly different from each other. Statistical analysis of *T*1 alone showed a similarity between populations from the southeast while variations were observed within northwest population. The analysis of family composition across generations showed the presence of *Aaa* and *Aaf* forms in each generation. The classification of *Ae. aegypti* into two subspecies is invalid in Senegal. Populations exhibit morphological polymorphism at the intra-family level that could have biological and epidemiological impacts.

Introduction

Zika (ZIKV), dengue (DENV), yellow fever (YFV) and chikungunya (CHIKV) viruses are transmitted mainly by *Aedes aegypti* worldwide. These arboviruses have experienced a significant geographic expansion, causing epidemics in different countries of Africa, Indian Ocean, Asia, Pacific, Europe and America despite all the considerable efforts made for their control [1,
DENV is the most prevalent arthropod-borne virus in the world. More than half of the world’s population is exposed to dengue fever and the number of infections is estimated at 390 million each year [3]. The four serotypes (DENV1-4) have all been reported in Africa. Recently, epidemics of dengue 1 (DEN1) occurred in Senegal (2017 and 2018) and Burkina Faso (2019) [4]. A re-emergence of CHIKV has been observed [5] especially in Asia and Indian Ocean islands. ZIKV is the most frequently amplified arbovirus in Senegal [2, 6] and recently has risen to considerable notoriety worldwide [7]. Despite the availability of a highly effective vaccine YF outbreaks are still frequent in Africa [8], and has recently occurred in Democratic Republic of Congo and Angola from which imported cases have been reported in China [9, 10]. Forty-seven countries, including 34 in Africa and 13 in Central and South America, are endemic to YF [8, 11]. Without any specific treatment and licensed vaccines (with the exception of YF) against these arboviruses, vector control is the only way of effective prevention and control. However, this vector control requires very precise targeting of the populations actually involved in transmission and therefore better knowledge of their structuration. Ae. aegypti, the most important epidemic vectors of DENV, ZIKV, CHIKV, and YFV, is present in practically all tropical and intertropical areas especially between 35˚ North and 35˚ South latitudes [12]. It is genetically the best characterized species in the genus Aedes [13]. This species presents great morphological and behavioral variability, close proximity to humans and the ability to transmit many pathogens [14, 15]. The first observation of morphological variations in Ae. aegypti was made by Hill in 1921 in Queensland, Australia [16]. This author noted that populations of Ae. aegypti which bred in the bush was darker than that associated with urban environment. This implicit correlation between differences in color and behavior was among the many concerns that prompted Mattingly to reassess the biology and taxonomy of Ae. aegypti [14, 17]. Considering the morphological, ecological and ethological data, he described a pale anthropophilic form which breeds in urban environment and a dark or wild form preferring natural breeding sites and animals for blood meals. Following this correlation between the habitat, the morphology and the behavior of the females, this author conventionally subdivided the species into two subspecies: Ae. aegypti aegypti (Aaa) and Ae. aegypti formosus (Aaf). Aaa was considered as domestic and anthropophilic with at least one pale scale on the first abdominal tergite. As for, Aaf was described as darker and characterized by the total absence of pale scales on this first abdominal tergite. This form Aaf was supposed to be present only in Africa with sylvatic and rather zoophilic behaviors. Referring to Mattingly’s work, McClelland proposed a classification of the different forms based on the coloration encountered, ranging from the black “F” form to the palest form “R” [15]. Applying this classification to a large population of Ae. aegypti from around the world, this author questioned the notion of subspecies as defined by Mattingly [14] and proposed the possibility of an incipient speciation [18]. Until very recently, it was considered that the two forms were sympatric in East Africa without reproductive barrier [19] and only Aaf would be present in Central and West Africa [14, 20]. However, recent data suggests that Aaa was also common in Senegal and presented a northwest–southeast cline with a dominance of Aaa in the northwest and Aaf in the southeast [21, 22]. Unlike morphological and bioecological data, the analysis of the genetic differentiation of Ae. aegypti populations from different localities in Senegal provides no clear evidence of the existence of two genetically distinct groups [21, 23]. In addition, genetic relationships highlighted by several molecular markers such as microsatellites and single nucleotide polymorphisms often do not match with morphological similarities [24]. Overall, these results improve understanding of the diversity of Ae. aegypti in West Africa, but so far, all these studies were done at population level and none has been interested in intra-family morphological variations. However, a study on the polymorphism within 196 families (each coming from a single female) from 18 anthropophilic and non-anthropophilic populations of Ae. aegypti collected from different localities
in South Africa, showed that 60.2% of the families were heterogeneous containing both Aaa and Aaf individuals [25]. These results suggest that the classification of Ae. aegypti into two subspecies is not valid in South Africa. A similar study in West Africa could help to explain the lack of genetic structuring of Ae aegypti subspecies in this area. It is in this context that we conducted this study on intra-family morphological polymorphism in different populations of Ae. aegypti from Senegal to better clarify on their taxonomic status.

Materials and methods

Sampling sites

Samples were collected between July and September 2018 in three climatic areas corresponding to three main rainfall area from south to north (Fig 1).

- Kédougou, PK10, and Tambacounda are located in the south of the country and are undergoing demographic and economic changes. They have a Sudano-Guinean and Sudano-Saharan climate. They are a crossroads of ecosystems characterized by a very diverse flora and fauna which are the result partly of favorable climatic characteristics. It is the rainiest area in the country (between 450 and 1300 mm/year from May to October) with temperatures ranging from 21˚C to 42˚C.

- Dakar and Mbour are located in the west of the country dominated by a wooded savannah. They are among the most urbanized cities in Senegal. This area benefits from a coastal microclimate influenced by the trade winds and the monsoon. The relatively short hot and humid season lasts from July to October with mean temperatures around 27˚C and annual rainfall of 300 to 400 mm/year.

- Barkédji and Louga located in the northwest of the country are characterized by a dry Sahelian climate with a vegetation consisting of a savannah with trees and a long dry season of 9 months or more. The short and unstable rainy season records rainfall between 300 mm and 390 mm in 24 rainy days. Temperatures range between 21˚C to 38˚C and relative humidity between 30 and 75% [26].

All sampling sites in Kédougou, Tambacounda, Dakar, Mbour, Barkédji, and Louga were located in the center of urban areas in the domestic environment while the sampling site at PK10 were within a forest gallery in a sylvatic environment.

Field collection of samples

The samples were collected from various potential breeding sites of Ae. aegypti (Table 1). In the domestic environment, artificial breeding sites (used tires, bricks) were prospected to collect larvae and pupae. These immature stages were also collected in natural breeding sites (tree holes, fruit husks and rock holes) from one single forest gallery located at 10 km from Kédougou city (PK10). In this forest, the choice to study the variations of the two populations (PK10Aaf and PK10Aaa) was motivated by the first notification during our sampling of both forms (Aaf and Aaa) in sympatric in the natural breeding sites in contrast to previous data which only reported the presence of the Aaf form [21]. Eggs were collected with trap consisting of a black painted pot half filled with water in which an oblique piece of wood immersed at 2/3 was used as a laying substrate. These traps were hung in shaded places in the urban environment as well as in the forest. The samples were collected from sites at least 100 m apart.
Ethics statement

No specific permits were required for this study. No specific permissions were required for these activities and the locations investigated are not protected. This study did not involve endangered or protected species. The study protocol was carefully explained to the heads and inhabitants of each household investigated to obtain their informed oral consent.

Sample processing in the laboratory

Larvae and eggs were maintained under standard insectarium conditions [27] (temperature of 27±1˚C, relative humidity of 80% and a photoperiod of 12:12h) until pupal stage. These pupae, as well as those collected directly in the field, were individually placed in test tubes.
After their emergence, adult mosquitoes (F₀) were identified morphologically according to the descriptions of Mattingly and Huang [14, 28] and then grouped by sex. Subsequently, they were gently cold-anesthetized and their wings were spread using needles to check the presence or not of pale scales on the first abdominal tergite under binocular dissecting microscope. After identification, male and female individuals of the same form were pooled according to their origin for mating. The Aaa forms were chosen in Dakar, Mbour, Barkédji and Louga for the F₀ parents while Aaf was chosen in Tambacounda and Kédougou and both forms in PK10 forest (Table 1).

**F₀ families egg batches production**

For each population, 30 fully engorged Aaa or Aaf females were selected for individual egg batches production. Each female was placed in a cup covered with a mosquito net and a cotton wool soaked in water were deposited at the bottom to collect the eggs. These females were subsequently fed with 10% sucrose and maintained under standard insectarium conditions as describe previously.

**Morphology of F₁ progeny**

Each egg batch (family) was reared separately into adults, and 10 F₁ females identified. For that, the wings of the mosquitoes were cut off at their base and theirs bodies fixed using a needle horizontally crossing the thorax. The number of pale scales on the first abdominal tergite (T₁) and the basal part of the second tergite (T₂) were counted under a binocular dissecting microscope (Motic ST-36C-6LED) at 40 times magnification.

To follow the morphological variations within families across 4 generations, egg batches were produced from 5 pairs of Aaf for each generation. The different egg batches were separately reared into adults and 10 females identified, as described previously, per egg batch.

**Data analysis**

Mosquito specimens without any pale scale on the T₁ tergite were classified Aaf while those with one or more pale scales on this T₁ tergite were classified Aaa. Thus, families in which all individuals had the same forms (Aaa or Aaf) were considered as homogeneous while families which presented both forms (Aaa and Aaf) were considered as heterogeneous. The mean numbers of pale scales on both tergites (T₁ + T₂) of the different populations were compared by multivariate analysis of variances using the Wilk’Lambda test. The mean numbers of pale scales on each tergite (T₁ and T₂) were compared using the Waller-Duncan t-test [29]. The relative abundance of the two forms across the generations was compared with the $\chi^2$ test.

| Locality         | Latitude N       | Longitude W     | Breeding sites | Stages collected | Morphology of F₀ |
|------------------|------------------|-----------------|----------------|------------------|------------------|
| Kédougou         | 12°33′45.3″      | 12°10′31.9″     | Used tires, bricks | Larvae + pupa    | Aaf              |
| PK10 forest      | 12°36′43″        | 12°14′46.80″    | TH, RH, FH, Ovitraps | Larvae + pupa    | Aaa, Aaf         |
| Tambacounda      | 14°5′52.73″      | 14°5′58.98″     | Used tires      | Larvae + pupa    | Aaf              |
| Louga            | 15°37′16.5″      | 16°14′7.6″      | Ovitraps        | Eggs             | Aaa              |
| Barkédji         | 15°16′37.4″      | 15°51′46.8″     | Ovitraps        | Eggs             | Aaa              |
| Mbour            | 14°25′7.6″       | 16°5′23.1″      | Used tires, poultry waterers | Larvae + pupa    | Aaa              |
| Dakar            | 14°40′22.5″      | 17°28′36.9″     | Used tires, flower pots | Larvae + pupa    | Aaa              |

TH = Tree holes, RH = Rock holes, FH = Fruit husks.

https://doi.org/10.1371/journal.pone.0242576.t001

Table 1. Breeding sites and collection stages of *Aedes aegypti* populations by locality, July-September 2018, Senegal.
Statistical analyses were performed using the R software version 2.15.1 [30] and results were considered significant when \( P < 0.05 \).

**Results**

A total of 2400 female progenies belonging to 240 families were identified and the number of pale scales on their \( T_1 \) and \( T_2 \) counted. Out of the 240 families, 42.5% (102/240) and 15% (36/240) were respectively \( Aaa \) and \( Aaf \) homogeneous families. The remaining families (42.5% of the 240 families investigated) were heterogeneous, containing both the \( Aaa \) and \( Aaf \) forms (Table 2). For each population studied, part of the \( F_1 \) offspring were morphologically different from their \( F_0 \) parents. Populations from the southeast (Kédougou, PK10 and Tambacounda) presented higher heterogeneity rates (Fig 2) compared to those from the northwest (Dakar, Mbour, Louga and Barkédji) (\( P < 0.05 \)).

The Waller-Duncan t-test on \( T_1 \) showed a similarity between \( Ae. \) aegypti populations from Kédougou, PK10 and Tambacounda (Table 2) with an average number of 1 to 2 pale scales on \( T_1 \) (\( p > 0.05 \)). However, variations were noted within \( Ae. \) aegypti populations from northwest sites with on average of pale scales ranging from 31 to 40. The population from Barkédji showed more pale scales on \( T_1 \) (\( p < 0.0001 \)) whereas that from Dakar presented fewer pale scales on \( T_1 \) than the others (\( p < 0.0001 \)). On the other hand, the populations from Mbour and Louga were comparable (\( p > 0.05 \)). All of these populations had more pale scales on \( T_1 \) than those from the southeast (\( p < 0.0001 \)).

The same analysis made on \( T_2 \) showed significant variations between populations from the southeast and those from the northwest with the exception of Dakar and PK10Aaf which were comparable (Table 2). A similarity was noted between populations from Barkédji and Louga (\( p > 0.05 \)) and between those from Kédougou and Tambacounda (\( p > 0.05 \)).

When both tergites means (\( T_1 + T_2 \)) were compared together by multivariate analysis, significant variations were observed across the 8 populations studied (\( p < 0.0001 \)).

The analysis of the composition of the offspring of \( Aaf \) families across 4 generations showed that the two forms (\( Aaa \) and \( Aaf \)) were present in each generation (Fig 3). The relative abundance of the two forms was statistically different from one generation to another (\( p < 0.05 \)).

### Table 2. Comparison of the average numbers of pale scales on \( T_1 \) and \( T_2 \) tergites and classification of \( F_1 \) families in 8 \( Ae. \) aegypti populations, July-September 2018, Senegal.

| Locality       | Nb  | min-max | Mean  | sd   | p-value | min-max | Mean  | sd   | p-value | Aaa | Aaf | Aaf-Aaa |
|----------------|-----|---------|-------|------|---------|---------|-------|------|---------|-----|-----|---------|
| Barkédji       | 300 | 0–122   | 40.09 | 22.04| a       | 0–68   | 17.62 | 12.76| a       | 21  | 0   | 9       |
| Dakar          | 300 | 0–83    | 25.77 | 16.31| b       | 0–76   | 8.13  | 10.13| b       | 26  | 0   | 4       |
| Louga          | 300 | 0–124   | 31.76 | 18.18| c       | 0–42   | 16.23 | 10.43| a       | 25  | 0   | 5       |
| Mbour          | 300 | 0–87    | 31.89 | 14.45| c       | 0–04   | 12.00 | 8.49 | c       | 27  | 0   | 3       |
| PK10Aa         | 300 | 0–35    | 2.36  | 5.03 | d       | 0–17   | 1.56  | 2.91 | df      | 1   | 11  | 18      |
| Kédougou       | 300 | 0–23    | 2.66  | 4.72 | d       | 0–23   | 0.92  | 2.56 | d       | 2   | 9   | 19      |
| PK10Af         | 300 | 0–32    | 1.79  | 4.37 | d       | 0–16   | 6.89  | 2.77 | eb      | 0   | 5   | 25      |
| Tambacounda    | 300 | 0–34    | 2.39  | 6.04 | d       | 0–23   | 3.35  | 5.18 | f       | 0   | 11  | 19      |
| **Total**      | 2400|         |       |      |         |         |       |      |         | 102 | 36  | 102     |

Identical letters indicate populations with a comparable average number of scales. Nb, number of individuals examined; sd, standard deviation; min, minimum number of scales; max, maximum number of scales; \( Aaa \), homogeneous family \( Ae. \) aegypti \( aegypti \); \( Aaf \), homogeneous family \( Ae. \) aegypti \( formosus \); \( Aaf + Aaa \), heterogeneous family.

https://doi.org/10.1371/journal.pone.0242576.t002
Discussion

This study conducted on 2400 females belonging to 240 families, showed significant intra-family morphological variations in different populations of *Ae. aegypti* from Senegal. Our data showed that a part of the F₁ progeny was morphologically different from their F₀ parents whatever the population studied. This intra-family heterogeneity in the offspring noted during this study suggest that the classification of *Ae. aegypti* into two subspecies by Mattingly [14] based on the presence of pale scales on the first abdominal tergite (T₁) should be considered as invalid in Senegal. The presence of both forms (Aaf and Aaa) across the country (at family level) is discordant with an earlier study which showed a southeast/northwest cline in the distribution of these two forms at population level with the exclusive presence of Aaf in the southeast, Aaa in the northwest and both forms in the center of the country [21]. The non-detection
of Aaa in the southeast and Aaf in the northeast, in this previous study, could be explained by their low intra-family proportions in the populations studied and the possible influence of some factors (temperature, relative humidity, rainfall, etc.) favoring a high mortality of the less represented forms. The impact of these factors could explain the southeast/northwest cline of Aaa and Aaf as previously observed [21]. Comparative studies on the survival of both forms from the southeast, center and northeast of Senegal are necessary to assess the possible impact of these factors. Our results are similar to those obtained during a study of the polymorphism

Fig 3. Relative abundance of Aaa and Aaf forms across four generations (F2 to F5) of Aaf parents. Different letters indicate a significant difference from one generation to another.

https://doi.org/10.1371/journal.pone.0242576.g003
of *Ae. aegypti* populations in South Africa [25]. Indeed, the author by rearing separately the progenies of several females of *Aaf* or *Aaa* has observed the presence of both forms in several families. As in South Africa, the proportion of homogeneous *Aaf* families was lower than that of the homogeneous *Aaa* and the heterogeneous families. However, the proportion of homogeneous *Aaf* families was more important in Senegal. These intra-family morphological variations could explain the significant variations observed within and between different *Ae. aegypti* populations worldwide. These variations also explain the presence in sympathy of both forms, formerly considered as 2 different subspecies, in sub-Saharan Africa from natural and artificial breeding sites [14, 21, 31–33]. It is interesting to note that our results are in perfect agreement with all the genetic studies which did not show any clear differentiation between individuals belonging to the two forms collected in several localities of Senegal [21–23, 34, 35]. The intra-family morphological variations of *Ae. aegypti* populations in other parts of Africa should be systematically reviewed to determine their taxonomic status. The heterogeneity rates in the progeny were higher in the populations from the southeast (Kédougou, PK10 and Tambacounda) compared to those from the northwest (Dakar, Mbour, Louga and Barkédji). That could be explained by an increase of *Aaa* specimens in this area in correlation with the beginning of an adaptation to the domestic environment of these so-called wild populations. Consistent with these data, a recent study showed the existence of a highly anthropophilic neo-population of *Ae. aegypti* in this area (unpublished data). Based on the average number of pale scales on *T*₁ tergite, our results showed a similarity in populations from the southeast (Kédougou, PK10 and Tambacounda). This could be explained by a conservation of sylvatic characters, in particular the dark coloring of tergites in these populations evolving under the same ecological conditions and dominated by the *Aaf* form [21, 23]. Indeed, an impact of climatic factors including temperature and relative humidity on the geographic distribution of the two forms has been noted by some authors [32, 36]. In agreement with other studies, our results showed variations in coloration on the basal part of *T*₂ ranging from complete absence to well-marked bands [15, 25]. These variations less reflected the geographic distribution of the two forms in Senegal compared to *T*₁.

The presence of both forms across the four generations of *Aaf* parents seems to confirm that the coloring of scales on *T*₁ is a polymorphic morphological character within families. *Ae. aegypti* should be viewed as a highly polymorphic rather than a polytypic species.

Other studies reported chromosomal inversions in *Aaf* from Senegal [37] and elsewhere with genetic introgression between *Aaa* and *Aaf* [38] suggesting a chromosomal polymorphism in *Ae. aegypti* populations. These chromosomal inversions have been directly associated with behaviors such as feeding behavior [39, 40], oviposition site preferences [41, 42], insecticide resistance [43] and immune response to parasites [44, 45] in malaria mosquito vectors. Moreover, mutant markers for abdominal coloring have been reported on *Ae. aegypti* chromosomes [13, 46].

Many studies showed that *Aaa* populations have higher vector competence compared to *Aaf* [21, 47]. This difference in vector competence could be linked to differences in competence of individuals of both forms. Thus, to test this hypothesis, it would be interesting to compare the intra-family variations in vector competence of individuals belonging to the *Aaf* and *Aaa* forms.

**Conclusion**

This study revealed a morphological polymorphism at intra-family level in different populations of *Ae. aegypti* from Senegal. The presence of pale scales on *T*₁, as a classification criterion for the two forms should be considered as invalid in Senegal. However, this study reveals two...
distinct groups; a group located in the southeast of the country with an average of 1 to 2 pale scales on T₁ and another group in the northwest with an average of 31 to 40 pale scales on same tergite. Additional detailed chromosome and/or genomic studies could give more explanation to these intra-family and inter-population morphological variations which could have an impact on biological parameters and the transmission of pathogens by Ae. aegypti in Senegal.

Acknowledgments
The authors would like to thank Mamoudou Ba, Oumar Ba, Moussa Ba and Ibrahima Niang for their technical assistance and collaboration.

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References
1. Weaver SC, Barrett AD. Transmission cycles, host range, evolution and emergence of arboviral disease. Nat Rev Microbiol. 2004; 2: 789–801. https://doi.org/10.1038/nrmicro1006 PMID: 15378043
2. Diallo D, Dia I, Diagne CT, Gaye A, Diallo M. Emergences of Chikungunya and Zika in Africa. Chikungunya and Zika Viruses. Elsevier; 2018. pp. 87–133.
3. Bhatt S, Gething PW, Brady OJ, Messina JP, Farlow AW, Moyes CL, et al. The global distribution and burden of dengue. Nature. 2013; 496: 504–507. https://doi.org/10.1038/nature12060 PMID: 23563266
4. WHO. WHO AFRO Outbreaks and Other Emergencies, Week 44: 28 October—03 November 2017 (Data as reported by 17:00; 03 November 2017)—Senegal. In: ReliefWeb [Internet]. 2017 [cited 26 Mar 2020]. Available: https://reliefweb.int/report/senegal/who-afro-outbreaks-and-other-emergencies-week-44-28-october-03-november-2017-data
5. Vasconcelos PFC, Powers AM, Hills S. The Emergence of Chikungunya and Zika Viruses in the Americas. Chikungunya and Zika Viruses. Elsevier; 2018. pp. 215–235. https://doi.org/10.1016/B978-0-12-811865-8.00007-6
6. Diallo D, Sall AA, Diagne CT, Faye O, Faye O, Ba Y, et al. Zika virus emergence in mosquitoes in southeastern Senegal, 2011. PLoS One. 2014; 9: e109442. https://doi.org/10.1371/journal.pone.0109442 PMID: 25310162
7. Diallo D, Diallo M. Why is Zika virus so rarely detected during outbreaks and how can detection be improved? BMC Res Notes. 2017; 10: 524. https://doi.org/10.1186/s13104-017-2854-8 PMID: 29084593

8. Diallo D, Sall AA, Diagne CT, Faye O, Hanley KA, Buenermann M, et al. Patterns of a sylvatic yellow fever virus amplification in southeastern Senegal, 2010. Am J Trop Med Hyg. 2014; 90: 1003–13. https://doi.org/10.4269/ajtmh.13-0404 PMID: 24615140

9. Ahmed QA, Memish ZA. Yellow fever from Angola and Congo: a storm gathers. Trop Doct. 2017; 47: 92–96. https://doi.org/10.1177/0049475517699726 PMID: 28424031

10. Wilder-Smith A, Leong WY. Importation of yellow fever into China: assessing travel patterns. J Travel Med. 2017; 24. https://doi.org/10.1093/jtm/tax008 PMID: 28426111

11. WHO. Yellow fever. 2019 [cited 25 Mar 2020]. Available: https://www.who.int/news-room/fact-sheets/detail/yellow-fever

12. Kraemer MU, Sinka ME, Duda KA, Mylne AQ, Shearer FM, Barker CM, et al. The global distribution of the arbovirus vectors Aedes aegypti and Aedes albopictus. Jit M, editor. eLife. 2015; 4: e08347. https://doi.org/10.7554/eLife.08347 PMID: 26126267

13. Severson DW, Mori A, Zhang Y, Christensen BM. Linkage Map for Aedes aegypti Using Restriction Fragment Length Polymorphisms. J Hered. 1993; 84: 241–247. https://doi.org/10.1093/oxfordjournals.jhered.a111333 PMID: 8101854

14. Mattingly PF. Genetic Aspects of the Aedes aegypti Problem: I.—Taxonomy and Bionomics. Ann Trop Med Parasitol. 1957; 51: 392–408. https://doi.org/10.1080/00034983.1957.11685991

15. McClelland GAH. A preliminary study of the genetics of abdominal colour variations in Aedes aegypti (L.) (Diptera, Culicidae). Ann Trop Med Parasitol. 1960; 54: 305–320. https://doi.org/10.1080/00034983.1960.11685991

16. Hill GF. Notes on Some Unusual Breeding-Places of Stegomyia Fasciata, F. ABR, in Australia. Ann Trop Med Parasitol. 1921; 15: 91–92. https://doi.org/10.1080/00034983.1921.11684253

17. Mattingly PF. Genetical Aspects of the Aedes aegypti Problem: II. Disease Relationships, Genetics and Control. Ann Trop Med Parasitol. 1958; 52: 5–17. PMID: 13521698

18. McClelland GAH. A worldwide survey of variation in scale pattern of the abdominal tergum of Aedes aegypti (L.) (Diptera: Culicidae). Trans R Entomol Soc Lond. 1974; 126: 239–259.

19. Moore DF. Hybridization and mating behavior in Aedes aegypti (Diptera: Culicidae). J Med Entomol. 1979; 16: 223–228. https://doi.org/10.1093/jmedent/16.3.223 PMID: 537005

20. Kröpelin S, Verschuren D, Lezine A-M, Eggermont H, Cocquyt C, Francus P, et al. Climate-Driven Ecosystem Succession in the Sahara: The Past 6000 Years. Science. 2008; 320: 765–768. https://doi.org/10.1126/science.1154913 PMID: 18467583

21. Sylla M, Bosio C, Urdaneta-Marquez L, Ndiaye M, Black WC. Gene Flow, Subspecies Composition, and Dengue Virus-2 Susceptibility among Aedes aegypti Collections in Senegal. Harris E, editor. PLoS Negl Trop Dis. 2009; 3: e408. https://doi.org/10.1371/journal.pntd.0000408 PMID: 19365540

22. Paupy C, Brengues C, Ndiath O, Toty C, Hervé J-P, Simard F. Morphologic and genetic variability within Aedes aegypti in Niakhar, Senegal. Infect Genet Evol. 2010; 10: 473–480. https://doi.org/10.1016/j.meegid.2010.03.001 PMID: 20223297

23. Huber K, Dia I, Mathiot C, Sall AA, Diallo M. Aedes aegypti in Senegal: Genetic Diversity and Genetic Structure of Domestic and Sylvatic Populations. Am J Trop Med Hyg. 2008; 79: 1003–13. PMID: 18689628

24. Powell JR, Tabachnick WJ. History of domestication and spread of Aedes aegypti—A Review. Mem Inst Oswaldo Cruz. 2013; 108: 11–17. https://doi.org/10.1590/0074-0276130395 PMID: 24473798

25. Jupp PG, Kemp A, Franos C. The potential for dengue in South Africa: morphology and the taxonomic status of Aedes aegypti populations. Mosq Syst 1991; 23: 182–190.

26. ANSD. Conseil Régional de Kédougou / Tambacounda / Dakar/Lougà/Thiès.). 2016 [cited 4 Mar 2020]. Available: http://www.ansd.sn/index.php

27. Gerberg EJ, Barnard DR, Ward RA. Manual for mosquito rearing and experimental techniques. American Mosquito Control Association, Inc. Lake Charles; 1994. Available: https://www.cabdirect.org/cabdirect/abstract/19952010555

28. Huang Y-M. The subgenus Stegomyia of Aedes in the Afrotropical Region with keys to the species (Diptera: Culicidae). Zootaxa. 2004; 700: 1–120. https://doi.org/10.1164/zootaxa.700.1.1

29. Waller RA, Duncan DB. A Bayes Rule for the Symmetric Multiple Comparisons Problem. J Am Stat Assoc. 1969; 64: 1484–1503. https://doi.org/10.1080/01621459.1969.10501073
30. R core Team. R: A language and environment for statistical computing. R foundation for statistical computing Vienna, Austria; 2016.

31. Futami K, Ishisita H, Higa Y, Lutiali PA, Sonye GO, Mwatele C, et al. Geographical Distribution of Aedes aegypti and Aedes aegypti formosus (Diptera: Culicidae) in Kenya and Environmental Factors Related to Their Relative Abundance. Wilkerson R, editor. J Med Entomol. 2019; 57: 772–779. https://doi.org/10.1093/jme/tjz233 PMID: 31815285

32. Higa Y, Abilio AP, Futami K, Lázaro MAF, Minakawa N, Gudo ES. Abundant Aedes (Stegomyia) aegypti mosquitos in the 2014 dengue outbreak area of Mozambique. Trop Med Health. 2015; 43: 107–109. https://doi.org/10.2149/tmh.2014-29 PMID: 26060423

33. Tabachnick WJ, Munstermann LE, Powell JR. Genetic distinctness of sympatric forms of Aedes aegypti in East Africa. Evolution. 1979; 33: 287–295. https://doi.org/10.1111/j.1558-5646.1979.tb04682.x PMID: 28568173

34. Brown JE, Evans BR, Zheng W, Obas V, Barrera-Martinez L, Egizi A, et al. Human impacts have shaped historical and recent evolution in Aedes aegypti, the dengue and yellow fever mosquito: evolutionary genetic history of Aedes aegypti. Evolution. 2014; 68: 514–525. https://doi.org/10.1111/evo.12281 PMID: 26060423

35. Wallis GP, Tabachnick WJ, Powell JR. Macrogeographic genetic variation in a human commensal: Aedes aegypti, the yellow fever mosquito. Genet Res. 1983; 41: 241–258. https://doi.org/10.1017/s0016672300002131 PMID: 6884773

36. Machado-Allison CE, Craig GB. Geographic Variation in Resistance to Desiccation in Aedes aegypti and Aedes atropalpus (Diptera: Culicidae). Ann Entomol Soc Am. 1972; 65: 542–547. https://doi.org/10.1093/aes/65.3.542

37. Bernhardt SA, Blair C, Sylla M, Bosio C, Black IV WC. Evidence of multiple chromosomal inversions in Aedes aegypti formosus from Senegal. Insect Mol Biol. 2009; 18: 557–569. https://doi.org/10.1111/j.1365-2583.2009.00895.x PMID: 19754736

38. Redmond SN, Sharma A, Sharakhov I, Tu Z, Sharakhova M, Neafsey DE. Linked-read sequencing identifies abundant microinversions and introgression in the arboviral vector Aedes aegypti. BMC Biol. 2020; 18: 1–14. https://doi.org/10.1186/s12915-019-0728-3 PMID: 31898513

39. Coluzzi M, Sabatini A, Petrarca V, Di Deco MA. Chromosomal differentiation and adaptation to human environments in the Anopheles gambiae complex. Trans R Soc Trop Med Hyg. 1979; 73: 483–497. https://doi.org/10.1016/0035-9203(79)90036-1 PMID: 394408

40. Main BJ, Lee Y, Ferguson HM, Kihonda A, Govella NJ, Kreppel KS, et al. The genetic basis of host preference and indoor resting behavior in the major African malaria vector, Anopheles arabiensis. bioRxiv. 2016; 044701.

41. Manoukis NC, Powell JR, Touré MB, Sacko A, Edillo FE, Coulibaly MB, et al. A test of the chromosomal theory of ecotypic speciation in Anopheles gambiae. Proc Natl Acad Sci. 2008; 105: 2940–2945. https://doi.org/10.1073/pnas.0709806105 PMID: 18287019

42. Sanford MR, Ramsay S, Cornel AJ, Marsden CD, Norris LC, Patchoke S, et al. A preliminary investigation of the relationship between water quality and Anopheles gambiae larval habitats in western Cameroon. Malar J. 2013; 12: 225. https://doi.org/10.1186/1475-2875-12-225 PMID: 23819866

43. Brooke BD, Hunt RH, Coetzee M. Resistance to dieldrin+fipronil assorts with chromosomal inversion 2La in the malaria vector Anopheles gambiae. Med Vet Entomol. 2000; 14: 190–194. https://doi.org/10.1046/j.1365-2915.2000.00222.x PMID: 10872863

44. Petrarca V, Beier JC. Intraspecific chromosomal polymorphism in the Anopheles gambiae complex as a factor affecting malaria transmission in the Kismu area of Kenya. Am J Trop Med Hyg. 1992; 46: 229–237. https://doi.org/10.4269/ajtmh.1992.46.229 PMID: 1539757

45. Riehle MM, Bukhari T, Gnerne A, Guebego WM, Coulibaly B, Fofana A, et al. The Anophels gambiae 2La chromosome inversion is associated with susceptibility to Plasmodium falciparum in Africa. Elife. 2017; 6: e25813. https://doi.org/10.7554/eLife.25813 PMID: 28643631

46. Munstermann LE, Craig GB. Genetics of Aedes aegypti. J Hered. 1979; 70: 291–296. https://doi.org/10.1093/oxfordjournals.jhered.a109261

47. Diallo M, Ba Y, Faye O, Soumare ML, Dia I, Sall AA. Vector competence of Aedes aegypti populations from Senegal for sylvatic and epidemic dengue 2 virus isolated in West Africa. Trans R Soc Trop Med Hyg. 2008; 102: 493–498. https://doi.org/10.1016/j.trstmh.2008.02.010 PMID: 18378270