ANTI-INFLAMMATORY ACTIVITY OF MARINE SPONGE CALLYSPONGIA SP. AND ITS ACUTE TOXICITY

ADRYAN FRISTIOHADY¹, WAHYUNI WAHYUNI¹, FADHILYAH MALIK¹, LA ODE MUHAMMAD JULIAN PURNAMA¹, BARU SADARUN², SAHIDIN I*¹

¹Department of Pharmacy, Faculty of Pharmacy, Universitas Halu Oleo, Kendari 93232, Southeast Sulawesi, Indonesia. ²Department of Fisheries, Faculty of Fisheries and Marine Science, Universitas Halu Oleo, Kendari 93232, Southeast Sulawesi, Indonesia.

Email: sahidin02@uho.ac.id

Received: 30 June 2019, Revised and Accepted: 23 October 2019

ABSTRACT

Objective: This study aims to investigate the anti-inflammatory effect of the ethanolic extract of Callyspongia sp. using stabilization of the human red blood cell (HRBC) membrane and its acute toxicity using brine shrimp lethality test (BSLT) method.

Methods: Callyspongia sp. was macerated with 96% ethanol. Extract characterized and screened for the secondary metabolite. Anti-inflammatory activity by stabilization of the HRBC membrane method with a varied dose of 50 ppm; 100 ppm; 200 ppm; 400 ppm; 800 ppm; 1600 ppm; and 3200 ppm. Solutions observed using a photometer to describing stability and ability in preventing membranes hemolytic and statistically analyzed using SPSS. Acute toxicity carried out by the BSLT method and analyzed using Minitab® version 17.2.1.

Results: The phytochemical screening was indicating that Callyspongia sp. contains flavonoid, alkaloid, and terpenoid. The results of the anti-inflammatory activity test showed that the percentage value of stability and hemolysis of extracts with doses of 50, 100, 200, 400, 800, 1600, and 3200 ppm were 55% and 45%, 63% and 37%, 70% and 30%, 74% and 26%, 80% and 20%, 87% and 13%, and 97% and 3%, respectively. It showed that extract of sponge Callyspongia sp. in all varied dose has activity in stabilizing the HRBC membrane thus can be potential as an anti-inflammatory. The results of acute toxicity assay showed that the value of LC₅₀ was 1281.45 μg/ml and categorized as nontoxic to Artemia salina Leach.

Conclusion: Various concentrations of Callyspongia sp. showed effective as an anti-inflammatory in stabilizing HRBC, and categorized as safe.

Keywords: Anti-inflammatory, Callyspongia sp., Red blood cell membranes.

INTRODUCTION

Inflammation is a response of host against to antigen which can cause tissue injury such as infection and burns hence the antigen will not spread. Inflammation is characterized by cardinal signs which are rubor, tumor, calor, dolor, and functiolesia. Inflammation is an important in the healing process however it can disturb activities if suffered for long term. Thus, the anti-inflammation agent is needed to decrease and stop the inflammation process [1].

Callyspongia sp. is one of marine sponges which can be found in Indonesia’s abundant sea. Callyspongia sp. has activity as antibacterial, antifungal, promoting antitumor, antiretroviral, and anti-inflammation [2,3]. Previous studies showed that Callyspongia sp. contains triterpenoid, steroid, alkaloid, and flavonoid [4-6]. Flavonoid and terpenoid contained in Callyspongia sp. potentially have anti-inflammatory activity by its ability to stabilize the membranes. Human red blood cell (HRBC) membranes can be used as a parameter for in vitro anti-inflammatory activity due to its similarity with lysosome membranes, which are responsible for the inflammatory process [1,7].

Callyspongia sp. can be developed as a new alternative in treating inflammation because commercial anti-inflammation drugs such as diclofenac sodium if consumed for long term has side effects. However, each study that involved natural products that have potency as drug or empirical used as drug, need pre-clinical toxicity acute assay to predict the safety following other pharmacological tests [8]. Many previous studies focus in researching marine sponges in each aspect, yet remains have no study reported about anti-inflammatory properties of Callyspongia sp. using stabilization of the HRBC membrane method. Therefore, this study aims to investigate the anti-inflammatory activity of the ethanolic extract of Callyspongia sp. with stabilization of the HRBC membrane method and its acute toxicity acute to Artemia salina Leach larvae.

METHODS

Marine sponge extract
Marine sponge of Callyspongia sp. was determined at Faculty of Fisheries and Aquaculture, Universitas Halu Oleo, Kendari (No. 0097 / UN 29.12.1.LIPP/2018) Marine sponge obtained from Bintang Samudra Edu-Marine Park, Konawe Regency, Southeast Sulawesi with a total of 2.8 kg. Marine sponge then wet sorted and chopped into pieces.

The pieces were macerated with 96% ethanol for 3×24 h. Filtered obtained was concentrated using rotary vacuum evaporator (Rotavapor, Buchi®) at temperature 60°C and water bath (60°C) yielded concentrated extract 31.52 g (1.12%).

Characterization of extract
Characterization of extract conducted was moisture content and ash content.

Moisture content
2 g of extract was put in the oven (105°C) for 3–5 h then, cooled in desiccators for 30 min. Extract then weighed until constant.
Ash content
2 g of extract put in weighed Kruz (A₀), flamed slowly and raised temperature until 600±25°C. The sample then cooled in desiccators and weighed (A₁).

Phytochemical screening
Phytochemical screenings conducted were flavonoid test, alkaloid test, and terpenoid/steroid test.

Flavonoid test
2 ml of extract put in the tube, added with Mg powder and 1 ml HCl. Thereafter, 1 ml amyl alcohol added.

Alkaloid test
1 mg of extract put in tube added 0.5 ml HCl 2%+2–3 drops Dragendorff reagent.

Terpenoid/steroid test
2 ml of extract put in the tube added Liebermann–Burchard reagent.

Ethical approval
This study conducted accordance ethic issued by the Ethical Committee of Health Research, Halu Oleo University No. 916/UN29.20/PPM/2018.

Anti-inflammatory activity by HRBC stabilization method
1 ml of each varied dose of ethanolic extract of Callyspongia sp. (50; 100; 200; 400; 1600; and 3200 ppm) as samples and diclofenac sodium as positive control was added into 1 ml phosphate buffer pH 7.4 (0.15 M), 0.5 ml of red blood cell, and 2 ml of hyposaline solution into tube. Negative control used is 1 ml isosaline was added into 1 ml phosphate buffer pH 7.4 (0.15 M), 0.5 of red blood cell, and 2 ml of hyposaline.
Each solution incubated at 56°C temperature for 30 min. Afterward, solutions centrifuged at 5000 rpm for 10 min. Hemoglobin was measured using photometer 5010 from the supernatant obtained (λ=560 nm).

The percentage of hemolysis of HRBC as calculated as follows [9]:
\[
\% \text{ Hemolysis} = \left( \frac{\text{Optical density of sample}}{\text{Optical density of control}} \right) \times 100
\]

The percentage of stability of HRBC as calculated as follows [9]:
\[
\% \text{ Stability} = 100 - \left( \frac{\text{Optical density of sample}}{\text{Optical density of control}} \right) \times 100
\]

Toxicity acute assay by brine shrimp lethality test method
10 of A. salina larvae in seawater were put in the tube and added 2.5 ml of varied dose sample (25; 50; 100; 2000 ppm). Control used is 2.5 ml Blanco solution added 2.5 ml seawater. Each sample replicated 3 times. Then, we observed for 24 h to count the number of live larvae and dead larvae.

\[
\text{Percentage of lethality calculated as follows } = \left( \frac{\text{Total of death larvae}}{\text{Total larvae}} \right) \times 100\%
\]

If control gives death of larvae, the formula can be corrected using Abbot formula =
\[
\frac{\text{Total of death larvae (Sample –Control)}}{\text{Total larvae}} \times 100\%
\]

Data collected then analyze using probit analysis with Minitab®ver. 17.2.1

Statistical analysis
Data collected were analyzed by SPSS with Kolmogorov-Smirnov test and Levene test, followed by One-Way ANOVA test (confidence interval=95%) and Least Significant Difference test. Probability level p<0.05 indicates significant difference and vice versa.

RESULTS AND DISCUSSION

Characterization of extract
Based on the study conducted, found that moisture content and ash content of marine sponge Callyspongia sp. were 4.5% and 17% consecutively. Information about the extract characterization is presented in Table 1.

Moisture content obtained from this study was 4.5% compared with literature; the ideal moisture content of extract is must <10%. Moisture content must comply with specified standards due to the effect of the durability of extract and avoid activity from microbes [10,11].

Table 1: Characterization of marine sponge Callyspongia sp.

| S. No. | Moisture content (%) | Ash content (%) |
|--------|----------------------|-----------------|
| 1.     | 4.5                  | 17              |

Table 2: Phytochemical screening of marine sponge Callyspongia sp.

| Phytochemical screening | Reagent                     | Reference                               | Result                             | Conclusion |
|------------------------|-----------------------------|-----------------------------------------|------------------------------------|------------|
| Alkaloid               | HCl 2%+Dragendorff          | Formed red brick, red, or orange color deposit [13] | Formed red color deposit           | Positive   |
| Flavonoid              | Mg+concentrated HCl+amyl alcohol | Discoloration into red, orange, or yellow [13] | Discoloration into red             | Positive   |
| Terpenoid/Steroid      | Liebermann–Burchard         | Discoloration into green or blue for steroid; and red or violet for terpenoid [14] | Discoloration into red             | Positive   |

Ethical approval
This study conducted accordance ethic issued by the Ethical Committee of Health Research, Halu Oleo University No. 916/UN29.20/PPM/2018.
Ash content obtained from this study was 17% and if compared with literature is must be <7%. Higher content of ash of extract describes extract contains a higher level of minerals and metals contamination [10,12].

Phytochemical screening
The information about the secondary metabolite of marine sponge Callyspongia sp. is presented in Table 2. It concluded that Callyspongia sp. contains flavonoid, alkaloid, and terpenoid. The study was in line with the previous study showed that flavonoid, alkaloid, and terpenoid were secondary metabolites that provide anti-inflammatory properties with their own mechanisms [15].

Anti-inflammatory activity
Anti-inflammatory activity ethanolic extract of Callyspongia sp. conducted to investigate the ability of extract in stabilizing red blood cell membranes. Stabilization of red blood cell membrane method used due to similarity with the lysosome membrane which is involved in inflammation response [1]. Stabilization of lysosome membrane has a role in inhibiting inflammation response due to inflammation mediators released by lysosome that can affect tissue damage and induce inflammation response [16].

The ability of the ethanolic extract of Callyspongia sp. in preventing red blood cell hemolysis in the stabilization of red blood cell membrane method is observed from the values of measured absorbance. The value absorbance describes the lysis of the red blood cell membrane. The greater the absorbance value, the greater the lysis occurs and vice versa. The study was done in triplication, with data obtained were presented in mean±standard deviation (SD).

Ethanolic extract of Callyspongia sp. at varied dose 50 ppm; 100 ppm; 200 ppm; 400 ppm; 800 ppm; 1600 ppm; and 3200 ppm has activity in stabilizing red blood cell (Figs. 1 and 2). This is proven by the stability percentage value of extract compared with positive control was almost similar to control with value 55–97% and 60–92%, respectively. Besides that, the hemolysis percentage value of extract compared with control was almost similar to value 45–3% and 40–8%, respectively. The maximum dose of 3200 ppm gave a higher effect in stabilizing and preventing hemolysis of the red blood cell membrane with value 97% and 3% consecutively. The minimum dose 50 ppm gave minimum effect in stabilizing and preventing hemolysis of red blood cell membrane with value 55% and 92% consecutively. The study concluded that extract and positive control with the same dose showed no significant difference (p>0.05) in stabilizing the membranes at 50 ppm; 100 ppm; 200 ppm; 400 ppm; 800 ppm; 1600 ppm; and 3200 ppm compared to positive control (diclofenac sodium) extract at 50 ppm; 100 ppm; 200 ppm; 400 ppm; 800 ppm; 1600 ppm; and 3200 ppm has activity in stabilizing red blood cell (Figs. 1 and 2). This is proven by the stability percentage value of extract compared with positive control was almost similar to control with value 55–97% and 60–92%, respectively. Besides that, the hemolysis percentage value of extract compared with control was almost similar to value 45–3% and 40–8%, respectively.

Table 3: Percentage of lethality Artemia salina larvae. *Values are presented in mean±standard deviation, n=3 with various doses 25 ppm, 50 ppm, 500 ppm, 1000 ppm, and 2000 ppm

| Varied dose (µg/ml) | Larvae death | Average | Lethality (%) | LC50 (µg/ml) |
|---------------------|--------------|---------|---------------|--------------|
| 25 ppm              | 0            | 0       | 0             | 1281.45      |
| 50 ppm              | 0            | 0       | 0             | 0            |
| 500 ppm             | 0            | 0       | 2             | 0            |
| 1000 ppm            | 2            | 3       | 1             | 2            |
| 2000 ppm            | 9            | 10      | 10            | 9.6          |
| Control             | 0            | 0       | 0             | 0            |
in converting arachidonic acid into prostaglandin as an inflammatory mediator [1].

**Acute toxicity**
The acute toxicity test was analyzed by probit analysis for observing the correlation of concentration and the mortality rate of larvae. The lethal concentration of the ethanolic extract of *Callyspongia* sp. was more than equal 500 ppm. Dose 25 ppm and 50 ppm showed no lethality to *A. salina* larvae similar to control. Information is presented in Table 3 and Fig. 3.

The LC₅₀ of the ethanolic extract of *Callyspongia* sp. was 1281.45 µg/ml (Table 3). The LC₅₀ of *A. salina* is declared as toxic if the value of LC₅₀ ≥1000 µg/ml, and vice versa declared as nontoxic (safe) if the LC₅₀ ≤1000 µg/ml [21]. Based on the value of LC₅₀ of the extract concluded that extract *Callyspongia* sp. is regarded as safe or nontoxic.

**CONCLUSION**
Secondary metabolite found in *Callyspongia* sp. is flavonoid, alkaloid, and terpenoid/steroid. Ethanolic extract of *Callyspongia* sp. has anti-inflammatory by stabilizing membrane at dose 50 ppm; 100 ppm; 200 ppm; 400 ppm; 800 ppm; 1600 ppm; and 3200 ppm. Moreover, *Callyspongia* sp. is nontoxic that the value of LC₅₀ is 1281.45 µg/ml.

**ACKNOWLEDGEMENT**
The authors thank the Ministry of Research, Technology, and Higher Education of the Republic of Indonesia, for a research grant scheme (Penelitian Dasar Unggulan Perguruan Tinggi 2019) for financial support.

**AUTHORS’ CONTRIBUTIONS**
Study concepts: IS, AF, and BS. Study design: IS, AF, and WW. Data acquisition: AF, WW, FM. Quality control of data and algorithms: AF, WW, FM, and LOMJP. Data analysis and interpretation: IS, AF, WW, FM, and BS. Manuscript preparation: IS, AF, LOMJP, and BS. Manuscript editing: IS, AF, and LOMJP. All authors reviewed the manuscript.

**CONFLICTS OF INTEREST**
The authors declare there are no conflicts of interest.

**REFERENCES**
1. Wiranto E, Mahamad AW, Puji A. Aktivitas antiinflamasi secara in vitro ekstrak teripang butuh keling (*Holothuria leucospilota* Brandt) dari pulau lemukutan. J Kimia Khatulistiwa 2016;5:52-7.
2. Warad VB, Habbu P, Shastri R. Antimicrobial activity of *Callyspongia diffusa* (Marine Sponge) associated endophytic bacteria 1 strains. Int J Pharm Pharm Sci 2017;9:90-6.
3. Ibrahim HA, El-Naggar HA, El-Damhougy KA, Bashar MA, Senna FM. *Callyspongia crassa* and *C. siphonella* (Portiera, Callyspongidae) as a potential source for medical bioactive substances, aqaba gulf, red sea, Egypt. J Phytopharmacol 2017;7:8-7.
4. Cowan MM. Plant products as antimicrobial agents. Clin Microbiol Rev 1999;12:564-82.
5. Suriani S, Hanapi U, Ahyar A. Isolasi, karakterisasi, dan uji bioaktivitas metabolit sekunder dari spons *Callyspongia* sp. Marina Chim Acta 2012;12:2-3.
6. Meenupriya J, Thangaraj M. Isolation and molecular characterization of bioactive secondary metabolites from *Callyspongia* spp. Associated fungi. Asian Pac J Trop Med 2016;3:738-40.
7. Ko HH, Hung CF, Wang JP, Lin CN. Antiinflammatory triterpenoids and steroids from *Ganoderma lucidum* and G. Tsugae. Phytochemistry 2008;69:234-9.
8. Frengki, Roslizawaty, Desi P. Uji toksisitas ekstrak etanol sarang semut lokal aceh (*Myrmecocystus sp.*) dengan metode BSLT terhadap larva *Udang Artemia salina* Leach. J Med Vet 2014;8:60-2.
9. Chippada SC, Sharan SV, Srinivasa RB, Meena V. In vitro antiinflammatory activity of methanolic extract of *Centella asiatica* by hrbc membrane stabilisation. Rasayan J Chem 2011;4:457-60.
10. Badan Pengawas Obat dan Makanan. Farmakope Herbal. 1* ed. Jakarta: Menteri Kesehatan RI; 2009.
11. Sastrawan, IN, Meiske S, Vanda K. Skrining fitokimia dan uji aktivitas antioksidan ekstrak biji adas (*Foeniculum vulgare*) menggunakan metode DPPH. J Ilmiah Sains 2013;13:110-5.
12. Isawwati A, Sukmayati A, Lucie W. Karakterisasi ekstrak etanol biji klabet (*Trigonella foenum-graecum* L.) sebagai tanaman obat pelancar ASI. Bul Penelitian Kesehatan 2013;41:103-10.
13. Nurgrahani R. Andayani Y, Hakim A. Skrining fitokimia dari ekstrak buah buncis (*Phaseolus vulgaris* L.) dalam Sediaan Serbuk. JPPIP 2016;2:98.
14. Illing I, Erfiana WS. Uji fitokimia ekstrak buah dengan. J Dinamika 2017;8:77-8.
15. Mohammed MS, Osman WJ, Garelnabi EA, Osman Z, Osman B, Khalid HS, et al. Secondary metabolites as anti-inflammatory agents. Int J Phytopharmacol 2014;3:275-85.
16. Iswariya S, Uma TS. Evaluation of in vitro anti-inflammatory and antimicrobial activity of aqueous and methanolic seed extracts of *Citrus lanatus*. Int J Pharm Pharm Sci 2017;9:29-33.
17. Saleem TK, Azeem AK, Dilip C, Sankar C, Prasanth NV, Duraisami R, et al. Anti-inflammatory activity of the leaf extracts of *Gendarussa vulgaris* Nees. Asian Pac J Trop Biomed 2011;1:147-9.
18. Oyedapo OO, Akinpelu BA, Adeyinka MO, Sipeolu FO. Red blood cell membrane stabilizing potentials of extracts of bioactive secondary metabolites from *Callyspongia crassa* (Porifera, Callyspongiidae) as a potential source for medical bioactive substances, aqaba gulf, red sea, Egypt. J Basic Appl Zool 2017;78:9.
19. Manivannana R, Sukumar D. The RBC membrane stabilisation in an in vitro method by the drug isolated from *Leucas aspera*. Int J Appl Sci Eng 2007;5:133-8.
20. Sadarun B, Hajrul M, Wahyuni W, Sahidin S. Toksisitas akut senyawa metabolit sekunder dari spons *Callyspongia crassa* sp. Pharmauho 2017;3:6-9.
21. Meyer BN, Ferrigni NR, Putnam JE, Jacobsen LB, Nichols DE, McLaughlin JL, et al. Brine shrimp: A convenient general bioassay for active plant constituents. Planta Med 1982;45:31-4.