Manganese-12 acetate suppresses the migration, invasion, and epithelial–mesenchymal transition by inhibiting Wnt/β-catenin and PI3K/AKT signaling pathways in breast cancer cells

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Abstract
Background: Breast cancer is the leading cause of cancer-related death in the world, and it is of great value to reveal the molecular mechanisms of breast cancer progression and develop new therapeutic targets.

Methods: Transwell assay is used to analyze the migration and invasion of breast cancer cells. Real-time PCR and western blotting assay are applied to detect the expression levels of epithelial–mesenchymal transition markers and the key members of Wnt/β-catenin and PI3K/AKT signaling pathways.

Results: Manganese-12 acetate (Mn12Ac) significantly inhibited the migration and invasion of MCF7 and MDA-MB-231 breast cancer cells. Western blotting assay further showed that Mn12Ac significantly upregulated E-cadherin, and downregulated N-cadherin and vimentin. We further found that Mn12Ac reduced the mRNA expressions of epithelial–mesenchymal transition-associated transcription factors snail, slug, twist1, and ZEB1 using real-time PCR assay. Importantly, we further found that Mn12Ac significantly reduced the Wnt1 and β-catenin protein expressions, and suppressed the phosphorylation of PI3K and AKT in MCF7 and MDA-MB-231 breast cancer cells. Very interestingly, we also showed that Mn12Ac decreased the mRNA and protein expressions of programmed cell death ligand 1.

Conclusion: Taken together, our results suggested that Mn12Ac inhibited the migration, invasion, and epithelial–mesenchymal transition by regulating Wnt/β-catenin and PI3K/AKT signaling pathways in breast cancer.

Introduction
Breast cancer is one of the most frequent malignancies worldwide, and the second leading cause of cancer-related mortality in the female population.1,2 Despite advances being made in the management of breast cancer over the past several decades, approximately 20–25% of patients with early breast cancer relapse within five years of diagnosis.3 Because metastasis is the leading cause of cancer-related death, revealing the mechanism of breast cancer metastasis is a matter of extreme urgency. Although many efforts have been made to show the mechanisms underlying both the emergence of drug resistance and the metastasis of breast cancer, these issues remain challenges to successful therapy.

Epithelial–mesenchymal transition (EMT) is a significant process, which is characterized as cells losing polarity and cell contact, and is activated in tumor metastasis and resistance to chemoradiotherapy.4,5 Activation of EMT is associated with changed expression of many genes,
including downregulation of epithelial markers, such as occluden-1 and E-cadherin, and upregulation of mesenchymal markers, including N-cadherin and vimentin. Many studies have shown that EMT is a critical process in tumor invasion and metastasis in breast cancer.7–9

Manganese-12 acetate (Mn12Ac) is a magnetically bistable molecule showing a combination of strong magnetic anisotropy and high spin properties.10,11 However, there has been no report about the correlations between Mn12Ac and disease therapy, especially cancer. In the present study, we explored the biological function of Mn12Ac in breast cancer treatment.

**Methods**

**Cell culture**

The human breast cancer cell lines, MCF7 and MDA-MB-231, were cultured in RPMI-1640 medium with 10% fetal bovine serum, 100 U/ml penicillin, and 100 g/ml streptomycin at 37°C in a humidified incubator containing 5% carbon dioxide.

We used 100 nM Mn12Ac (dissolved in H2O) to treat the breast cancer cells for 48 hours, and then detections were carried out.

**Total RNA extraction and real-time PCR assay**

Total RNA was isolated from cancer cells using the RNeasy Mini Kit, as described by the manufacturer (Qiagen, Hilden, Germany), and used for real-time PCR assay.

Real-time PCR was used to detect the mRNA expressions of E-cadherin, N-cadherin, vimentin, snail, slug, twist1, ZEB1, and programmed cell death ligand 1 (PD-L1). The PCR reactions were carried out in a total volume of 20 μL, including 10 μL of 2XPower SYBR Green PCR Master Mix (Applied Biosystems, Warrington, UK), 2 μL of cDNA (5 ng/μL) and 1 μL of primer mix (10 μM each). PCR amplification and detection were carried out in a LightCycler 480 II (Roche Applied Science, Penzberg, Germany) as follows: an initial denaturation at 95°C for 10 minutes; 40 cycles of 95°C for 15 seconds and 60°C for 1 minute. The relative gene expression was calculated using the comparative CT method. The gene expression of the target gene was normalized to an endogenous reference (glyceraldehyde 3-phosphate dehydrogenase [GAPDH]), and relative to the calibrator were given by the formula 2−ΔΔCT. ΔCT was calculated by subtracting the average GAPDH CT from the average CT of the gene of interest. The ratio defines the level of relative expression of the target gene to that of GAPDH. The primers were as follows: E-cadherin forward primer, 5′-GAACGCATTGCCCACATA CAC-3′; E-cadherin reverse primer, 5′-GATTCCGGGCTT GTTGTCAT-3′; N-cadherin forward primer, 5′-GGGTTCA TTAGCAAGGGGAATCG-3′; N-cadherin reverse primer, 5′-GGGTTCTCCGTTCACACTGAGGAGG-3′; vimentin forward primer, 5′-TGAGTACCGGAGACAGGT CGAG-3′; vimentin reverse primer, 5′-TGTCGTAG GGTCTGCTGGAA-3′; slug forward primer, 5′-GCGAT GCTTATTGCTTAC-3′; slug reverse primer, 5′-AGGCATT TGAGGAAAGGTATTG-3′; twist1 forward primer, 5′-AGAAGTCTGGGGCTGTGGCGG-3′; twist1 reverse primer, 5′-GAGGCGAGGGTGGAAGTATC-3′; ZEB1 forward primer, 5′-CTACTCAACTACGGGTCAAGCC-3′; ZEB1 reverse primer, 5′-TGGCCCCGTGTAAGATCAG AG-3′; PD-L1 forward primer, 5′-GAACACTCTGGCC CACTCCT-3′; PD-L1 reverse primer, 5′-CACATCCCATCAT TCCCTTCC-3′; GAPDH forward primer, 5′-AAATCC CATCACCATCTTGAG-3′; GAPDH reverse primer, 5′- GAGTCCCTCCAGATACAAAGTTGG-3′.

**Western blotting assay**

Cells were lysed in the lysis buffer (20 mM Tris, 2 mM ethylenediaminetetraacetic acid, 50 mM 2-mercaptoethanol, 10% glycerol, pH 7.4). The homogenates were placed on ice for 30 minutes and centrifuged at 12 000 g for 15 minutes at 4°C. Afterward, the protein concentrations of the lysates were determined using a Protein Assay Kit (Bio-Rad, Richmond, CA, USA). Equal amounts of total proteins were loaded onto a 10% gradient polyacrylamide gel, electrophoretically transferred to polyvinylidene difluoride membrane, and then blocked with 10% non-fat milk for two hours at room temperature. The membranes were incubated with specific primary antibodies overnight at 4°C and probed with corresponding secondary antibodies for one hour at room temperature. The protein bands were visualized using ECL Blotting Detection Reagents (Applygen, Beijing, China). The primary antibodies were as follows: E-cadherin (1:1000; Santa Cruz Biotechnology, CA, USA), N-cadherin (1:1000; Santa Cruz Biotechnology) Vimentin (1:1000; Santa Cruz Biotechnology), Wnt1 (1:1000; Santa Cruz Biotechnology), β-catenin (1:1000; Santa Cruz Biotechnology), p-Pi3K (1:1000; Santa Cruz Biotechnology), Pi3K (1:1000; Santa Cruz Biotechnology), p-AKT (1:1000; Santa Cruz Biotechnology), AKT (1:1000; Santa Cruz Biotechnology), PD-L1 (1:1000; Cell Signaling Technology, Danvers, MA, USA), and β-actin (1:5000; Santa Cruz Biotechnology).

**Cell proliferation assay**

The tumor cell proliferation was analyzed by a Cell Counting Kit-8 (Dojindo Laboratories, Kumamoto, Japan) based
on the manufacturer’s instructions. A total of 48 hours after Mn12Ac treatment, cells were seeded into 96-well plates at a density of $8 \times 10^3$ cells per well with 100 μL medium, and continued to incubate at 37°C. At the indicated time-points, 10 μL Cell Counting Kit-8 solutions were added to each well. After incubation at 37°C for one hour, the absorbance was measured with a plate reader at 450 nm. The growth curves were shown to reveal the growth rates.

**Transwell assay**

For the transwell invasion assay, 60 μL of matrigel was diluted with precooled serum-free medium in 1:4 ratios, and was added to the bottom of the transwell chamber and incubated for one hour at 37°C. Then, $2 \times 10^5$ cells were seeded into the upper chambers (24-well insert; pore size 8 μm), while medium supplemented with 10% FBS (600 μL) was placed in the lower chamber. After incubation at 37°C for 24 hours, cells on the top side of the inserts were removed gently with a cotton swab. The inserts were then fixed by 4% methanol for 15 minutes and stained with 0.1% crystal violet for 15 minutes. The average migratory cells were counted by randomly choosing five fields each time under the microscope. The experiments were repeated three times.

For the transwell migration assay, the remaining protocol was the same as the transwell invasion assay, except for the pre-coat of the matrigel in the inserts.

**Statistical analysis**

All data were analyzed by Student’s t-test and one-way analysis of variance using SPSS (SPSS, Chicago, IL, USA) and GraphPad Prism 5.0 (GraphPad, La Jolla, CA, USA). $P < 0.05$ was considered as a significant difference. *$P < 0.05$, **$P < 0.01$, ***$P < 0.001$ and ****$P < 0.0001$.

**Results**

**Mn12Ac significantly inhibited the EMT of MCF7 and MDA-MB-231 breast cancer cells**

EMT is activated and plays important roles in tumor metastasis; therefore, we explored whether Mn12Ac affected the EMT process of breast cancer cells. The results showed that Mn12Ac significantly upregulated the epithelial marker E-cadherin, and downregulated the mesenchymal markers N-cadherin and vimentin in MCF7 and MDA-MB-231 cells by real-time PCR assay (Fig 2a–c). We further found that in protein level, Mn12Ac also significantly upregulated E-cadherin, and reduced the expressions of N-cadherin and Vimentin by western blotting assay (Fig 2d).

**Mn12Ac significantly inhibited the EMT-associated transcription factors in MCF7 and MDA-MB-231 breast cancer cells**

Snail, slug, twist1, and ZEB1 are the EMT-associated transcription factors in cancer. Therefore, we want to know whether Mn12Ac regulated the EMT by affecting these transcription factors. Using real-time PCR, we revealed that Mn12Ac significantly decreased the mRNA expressions of snail, slug, twist1, and ZEB1 in MCF7 and MDA-MB-231 cancer cells (Fig 3a–d). These results suggested that Mn12Ac inhibited the EMT of breast cancer cells by downregulating EMT-inducing transcription factors.

**Mn12Ac significantly inhibited the Wnt/β-catenin and PI3K/AKT signaling pathways**

Wnt/β-catenin and PI3K/AKT signaling pathways play important roles in the regulation of EMT, and we predicted that Mn12Ac might inhibit the EMT of breast cancer cells through regulating these two pathways. Using western blotting assay, we found that Mn12Ac significantly reduced the expressions of Wnt1 and β-catenin (Fig 4a). Mn12Ac could also inhibit the phosphorylation of PI3K and AKT in MCF7 and MDA-MB-231 breast cancer cells (Fig 4b).

**Mn12Ac decreased the expression of PD-L1 both in mRNA and protein levels**

PD-1/PD-L1-targeted immunotherapy has emerged as a promising therapeutic strategy for breast cancer, and PD-L1 expression was associated with EMT transition in breast cancer. Therefore, we explored whether Mn12Ac affected the PD-L1 in breast cancer. Very interestingly, the results showed that Mn12Ac could reduce the mRNA and protein expression levels of PD-L1 in MCF7 and MDA-MB-231 breast cancer cells.
breast cancer cells by real-time PCR and western blotting assay (Fig 4c,d).

**Discussion**

Breast cancer is the most common malignancy and the leading cause of cancer-related death among women worldwide.\(^1\) Women aged from 20 to 59 years are most likely to be afflicted by breast cancer, and the age of onset has reduced in recent years and has been a public health problem.\(^2\) Cancer cell migration and invasion is a necessary step for metastatic dissemination that leads to rapid relapse and a poor prognosis in breast cancer patients. Development of a novel and effective treatment method to inhibit tumor metastasis is urgently required.

Wnt/β-catenin and PI3K/AKT signaling pathways play important roles in regulating the EMT process in the carcinogenesis. In the present study, we found that Mn12Ac could reduce the protein expressions of Wnt1 and β-catenin, and inhibit the phosphorylation of PI3K and AKT in MCF7 and MDA-MB-231 breast cancer cells. Lu et al. reported that L1, L5, and VLDL could promote breast cancer progression and metastasis through Akt-induced EMT.\(^3\) Leptin was reported to promote the breast cancer cells EMT by activating the PI3K/AKT signaling pathway.\(^4\) miR-564 could directly target the AKT2 and inhibit the PI3K/AKT pathway, and further inhibit the EMT process, proliferation, and invasion of breast cancer.\(^5\) Xie et al. found that FYN was transcriptionally regulated by FOXO1, and mediated FGF2-induced EMT through both the PI3K/AKT pathway.
AKT and ERK signaling pathways. Harper et al. found that Her2+ disseminated cancer cells could activate the Wnt-dependent EMT-like dissemination program, but without complete loss of the epithelial phenotype, and this effect could be reversed by Her2 or Wnt inhibition. Overexpression of DACT2 induced breast cancer cell apoptosis,

Figure 2 Manganese-12 acetate (Mn12Ac) inhibited the epithelial-mesenchymal transition. (a–c). mRNA expressions of epithelial-mesenchymal transition markers using Real-time PCR assay. (d). Protein expressions of epithelial-mesenchymal transition markers using Western blotting assay. MCF7, MDA-MB-231. GAPDH, glyceraldehyde 3-phosphate dehydrogenase; NC, negative control.

Figure 3 Manganese-12 acetate (Mn12Ac) reduced the epithelial-mesenchymal transition associated transcription factors. MCF7, MDA-MB-231. GAPDH, glyceraldehyde 3-phosphate dehydrogenase; NC, negative control.
and further inhibited breast tumor cell proliferation, migration, and EMT, by suppressing the Wnt/β-catennin signaling pathway.\textsuperscript{26} The present results further suggested that Mn12Ac might develop as a therapeutic drug in breast cancer treatment.

Mechanically, in our results, the phosphorylation of PI3K and AKT was inhibited by Mn12Ac. Previous studies reported that BRD4, FTO, and MACC1 could positively regulate the phosphorylation of PI3K and AKT, and miR-200a and miR-200b could negatively regulate the levels of p-PI3K and p-AKT.\textsuperscript{27–30} Whether Mn12Ac inhibited the phosphorylation of PI3K and AKT by regulating the above genes or other downstream genes should be the subject of future studies.

Programmed death-1 (PD-1)/programmed cell death ligand 1 (PD-L1) were identified as negative immunoregulatory molecules that promoted immune evasion of tumor cells.\textsuperscript{31} The interaction of PD-1 and PD-L1 inhibited the function of T cells while increasing the function of immunosuppressive regulatory T cells. Very interestingly, the present results found that Mn12Ac could significantly reduce the expression of PD-L1 both in mRNA and protein expression levels in breast cancer cells.

Taken together, our results showed that Mn12Ac could significantly inhibit the migration, invasion, and EMT of breast cancer cells by suppressing the Wnt/β-catennin and PI3K/AKT signaling pathways, and Mn12Ac could also reduce the expression of PD-L1. However, the direct target of Mn12Ac is still unknown. In future studies, we will further explore the mechanism underlying the suppression of Mn12Ac on the migration, invasion, and EMT of breast cancer cells, and further identify the target of Mn12Ac.

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Disclosure

No authors report any conflict of interest.

References

1. Ferlay J, Steliarova-Foucher E, Lortet-Tieulent J \textit{et al.} Cancer incidence and mortality patterns in Europe: Estimates for 40 countries in 2012. \textit{Eur J Cancer} 2013; 49: 1374–403.
2. Malvezzi M, Bertuccio P, Rosso T \textit{et al.} European cancer mortality predictions for the year 2015: Does lung cancer have the highest death rate in EU women? \textit{Ann Oncol} 2015; 26: 779–86.
3. DeSantis CE, Lin CC, Mariotto AB \textit{et al.} Cancer treatment and survivorship statistics, 2014. \textit{CA Cancer J Clin} 2014; 64: 252–71.
4. Smith BN, Bhowmick NA. Role of EMT in metastasis and therapy resistance. \textit{J Clin Med} 2016; 5 (2): 17.
5. Lamouille S, Xu J, Derynck R. Molecular mechanisms of epithelial-mesenchymal transition. \textit{Nat Rev Mol Cell Biol} 2014; 15: 178–96.
6 Seton-Rogers S. Epithelial-mesenchymal transition: Untangling EMT’s functions. Nat Rev Cancer 2016; 16: 1.
7 Fici P, Gallerani G, Morel AP et al. Splicing factor ratio as an index of epithelial-mesenchymal transition and tumor aggressiveness in breast cancer. Oncotarget 2017; 18: 2423–2436.
8 Shao P, Liu Q, Maina PK et al. Histone demethylase PHF8 promotes epithelial to mesenchymal transition and breast tumorigenesis. Nucleic Acids Res 2017; 45: 1687–1702.
9 Zhan Y, Li X, Liang X et al. MicroRNA-182 drives colonization and macroscopic metastasis via targeting its suppressor SNAI1 in breast cancer. Oncotarget 2017; 8: 4629–4641.
10 Rakvin B, Zilic D, North JM, Dalal NS. Probing magnetic fields on crystals of the nanomagnet Mn12-acetate by electron paramagnetic resonance. J Magn Reson 2003; 165: 260–4.
11 Morello A, Bakharev ON, Brom HB, Sessoli R, de Jongh LJ. Nuclear spin dynamics in the quantum regime of a single-molecule magnet. Phys Rev Lett 2004; 93: 197202.
12 Amoroso MR, Matassa DS, Agliarulo I et al. TRAP1 downregulation in human ovarian cancer enhances invasion and epithelial-mesenchymal transition. Cell Death Dis 2016; 7: e2522.
13 Liang W, Lai Y, Zhu M, Huang S, Feng W, Gu X. Combretastatin A4 regulates proliferation, migration, invasion, and apoptosis of thyroid cancer cells via PI3K/Akt signaling pathway. Med Sci Monit 2016; 22: 4911–7.
14 Hammers H, Fu C, Gerber S et al. Epithelial-mesenchymal transition: A mechanism of resistance to VEGF pathway inhibition in genitourinary cancers. J Clin Oncol 2012; 30: 390.
15 Liu MH, WJ FU, Cui YH, Guo QN, Zhou Y. Downregulation of Semaphorin-3F is associated with poor prognostic significance in osteosarcoma patients. Am J Cancer Res 2016; 6: 2252–62.
16 Liu B, Pan CF, He ZC et al. Long noncoding RNA-LET suppresses tumor growth and EMT in lung adenocarcinoma. Biomed Res Int 2016; 2016: 4693471.
17 Zhu FQ, Chen MJ, Zhu M et al. Curcumin suppresses epithelial-mesenchymal transition of renal tubular epithelial cells through the inhibition of Akt/mTOR pathway. Biol Pharm Bull 2017; 40: 17–24.
18 Alsuliman A, Colak D, Al-Harazi O et al. Bidirectional crosstalk between PD-L1 expression and epithelial to mesenchymal transition: Significance in claudin-low breast cancer cells. Mol Cancer 2015; 14: 149.
19 Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. Int J Cancer 2010; 127: 2893–917.
20 Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. CA Cancer J Clin 2014; 64: 9–29.
21 LU CW, Lo YH, Chen CH et al. VLDL and LDL, but not HDL, promote breast cancer cell proliferation, metastasis and angiogenesis. Cancer Lett 2016; 388: 130–8.
22 Wei L, Li K, Pang X et al. Leptin promotes epithelial-mesenchymal transition of breast cancer via the upregulation of pyruvate kinase M2. J Exp Clin Cancer Res 2016; 35: 166.
23 Mutlu M, Saatci O, Ansari SA et al. miR-564 acts as a dual inhibitor of PI3K and MAPK signaling networks and inhibits proliferation and invasion in breast cancer. Sci Rep 2016; 6: 32541.
24 Xie YG, Yu Y, Hou LK, Wang X, Zhang B, Cao XC. FYN promotes breast cancer progression through epithelial-mesenchymal transition. Oncol Rep 2016; 36: 1000–6.
25 Harper KL, Sosa MS, Ennentberg D et al. Mechanism of early dissemination and metastasis in Her2+ mammary cancer. Nature 2016; 540: 588–92.
26 Xiang T, Fan Y, Li C et al. DACT2 silencing by promoter CpG methylation disrupts its regulation of epithelial-to-mesenchymal transition and cytoskeleton reorganization in breast cancer cells. Oncotarget 2016 2016; 7: 70924–70935.
27 Hao J, Yang Z, Wang L et al. Downregulation of BRD4 inhibits gallbladder cancer proliferation and metastasis and induces apoptosis via PI3K/AKT pathway. Int J Oncol 2017; 51: 823–31.
28 Liu Y, Wang R, Zhang L, Li J, Lou K, Shi B. The lipid metabolism gene FTO influences breast cancer cell energy metabolism via the PI3K/AKT signaling pathway. Oncol Lett 2017; 13: 4685–90.
29 Wu Q, Lu RL, Li JX, Rong LJ. MiR-200a and miR-200b target PTEN to regulate the endometrial cancer cell growth in vitro. Asian Pac J Trop Med 2017; 10: 498–502.
30 Zhou W, Liu L, Xue Y et al. Combination of endothelial-monocyte-activating polypeptide-II with Temozolomide suppress malignant biological behaviors of human Glioblastoma stem cells via miR-590-3p/MACC1 inhibiting PI3K/AKT/mTOR signal pathway. Front Mol Neurosci 2017; 10: 68.
31 Grinberg-Bleyer Y, Ghosh S. A novel link between inflammation and cancer. Cancer Cell 2016; 30: 829–30.