Global knockdown of glutamate decarboxylase 67 elicits emotional and auditory abnormalities in mice

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Abstract

Reduced expression of glutamate decarboxylase 67 (GAD67), encoded by the Gad1 gene, is a consistent finding in postmortem brains of patients with several psychiatric disorders, including schizophrenia, bipolar disorder and major depressive disorder. The dysfunction of GAD67 in the brain is implicated in the pathophysiology of these psychiatric disorders; however, the neurobiological consequences of GAD67 dysfunction in mature brains are not fully understood because the homozygous Gad1 knockout is lethal in newborn mice. We hypothesized that the tetracycline-controlled gene expression/suppression system could be applied to develop global GAD67 knockdown mice that would survive into adulthood. In addition, GAD67 knockdown mice would provide new insights into the neurobiological impact of GAD67 dysfunction. Here, we developed Gad1^{TA/STOP-tetO} biallelic knock-in mice using Gad1^{STOP-tetO} and Gad1^{TA} knock-in mice, and compared them with Gad1^{+/+} mice. The expression level of GAD67 protein in brains of Gad1^{TA/STOP-tetO} mice treated with doxycycline (Dox) was decreased by approximately 90%. The GABA content was also decreased in the brains of Dox-treated Gad1^{TA/STOP-tetO} mice. In the open-field test, Dox-treated Gad1^{TA/STOP-tetO} mice exhibited hyper-locomotor activity and decreased duration spent in the center region. In addition, acoustic startle responses were impaired in Dox-treated Gad1^{TA/STOP-tetO} mice. These results suggest that global reduction in GAD67 elicits emotional and auditory abnormalities in mice. These GAD67 knockdown mice will be useful for elucidating the neurobiological mechanisms of emotional abnormalities, such as anxiety symptoms associated with psychiatric disorders.
Introduction

$\gamma$-Aminobutyric acid (GABA), a major inhibitory neurotransmitter, regulates a variety of biological functions. GABA is synthesized from glutamate by glutamate decarboxylase (GAD) existing two isoforms with different molecular weights, 67 kDa (GAD67) and 65 kDa (GAD65), which are independently encoded by the $Gad1$ and $Gad2$ genes, respectively [1,2]. Since GAD67 and GAD65 proteins have different subcellular distributions and a cofactor association [3–6], the physiological roles of GAD67 and GAD65 may be different in the brain.

Decreased expression of the full-length $GAD1$ transcript and GAD67 protein is one of the most consistent findings in the brains of subjects with several psychiatric disorders, including schizophrenia, bipolar disorder and major depressive disorder [7–12]. Alternative splicing of $GAD1$ and the epigenetic state may play roles in brain development and the risk of schizophrenia [12]. Recent studies with whole exome sequencing of schizophrenic patients identified missense mutation mapping at the $GAD1$ gene, which caused a reduction in GAD67 enzymatic activity by ~30% due to impaired homodimerization [13,14]. Therefore, the functional impairment of GAD67 associated with genetic mutations is involved in the neurobiological mechanisms of several psychiatric disorders.

To elucidate the physiological roles of GAD67 in mature brains, a study using transgenic animals with reduced GAD67 expression would be helpful. Because homozygous $Gad1$ knockout ($Gad1^{-/-}$) is lethal in newborn mice [15], conditional $Gad1^{-/-}$ mice generated with a Cre-loxP strategy have often been used for neurobiological studies [16–20]. These studies have provided much information about the function of GAD67 in targeted cells. However,
behavioral and neurochemical consequences of the global dysfunction of GAD67 in mature brains are not fully understood. Several studies have been performed to investigate GAD67 function using mice with GAD67 haplodeficiency; however, physiological changes, such as GABA reduction in the brain and behavioral abnormalities, were mild in those mice [15,21–24].

The tetracycline-controlled gene expression/suppression system allows reduction in a gene of interest by administration of antibiotic tetracyclines and its derivative doxycycline (Dox) [25]. This system requires two distinct transgenic mice: one has a cell type-specific promoter driving a tetracycline-controlled transcriptional activator (tTA)-expressing allele, and the other has a tetracycline operator site (tetO) binding to tTA and driving the expression of the gene of interest. By crossing them, a biallelic knock-in mouse (tTA/tetO) can be obtained with a tTA-mediated gene induction system, which can be turned off by the administration of Dox.

We hypothesized that the tetracycline-controlled gene expression/suppression system could be applied to generate transgenic mice able to postnatally knockdown GAD67. In brief, two knock-in mice were generated: (1) Gad1STOP-tetO knock-in mice, which were generated by inserting the tetO sequence following the STOP sequence upstream of the Gad1 translation initiation site (Fig. 1a), and (2) Gad1tTA knock-in mice [26], which express tTA proteins under the control of an endogenous Gad1 promoter (Fig. 1b). The Gad1tTA/STOP-tetO mice can produce GAD67 protein (Fig. 1c) and survive into adulthood. In addition, the expression of GAD67 protein in the Gad1tTA/STOP-tetO mice is suppressed by treatment with Dox (Fig. 1d). Therefore, Dox-treated Gad1tTA/STOP-tetO mice
can be used as GAD67 knockdown mice for studying the behavioral and neurochemical consequences of the global loss of GAD67 in mature brains. In this study, we successfully developed Gad1STOP-tetO knock-in mice and the subsequent Gad1TA/STOP-tetO mice. Herein, we report the behavioral abnormalities elicited by the global knockdown of GAD67 in mice.

Methods

Ethics

This study was performed in accordance with the Guidelines for Animal Experimentation at Gunma University Graduate School of Medicine and was approved by the Gunma University Ethics Committee (Permit number: 14-006 and 19-009). Every effort was made to minimize the number of animals used and their suffering.

Experimental design

We first generated Gad1STOP-tetO knock-in mice and assessed the Gad1 knockout phenotypes (GAD67 deletion, neonatal death and cleft palate) of homozygous Gad1STOP-tetO/STOP-tetO mice to confirm the elimination of Gad1 gene function by inserting the Neo-STOP-tetO cassette. We then crossed heterozygous Gad1TA/+ and Gad1STOP-tetO/+ mice to obtain Gad1TA/STOP-tetO mice and confirmed tTA-mediated GAD67 expression. Afterward, we evaluated whether tTA-mediated GAD67 expression was suppressed by treatment with Dox. It has been
reported that GAD67 haplodeficient mice demonstrate an approximately 40% reduction in GAD67 protein in the brain compared with wild-type mice [27]. Therefore, we judged that GAD67 knockdown mice were successfully developed when the expression level of GAD67 protein in the brain was reduced by more than 40%. These experiments were performed in mice of both sexes.

In the behavioral tests, male mice were only used. We prepared two independent cohorts comparing $Gad1^{+/+}$ mice and $Gad1^{tTA/STOP-tetO}$ mice at the ages of 8 to 10 weeks. One cohort was used for assessing the body weights, motor coordination performance, and GABA and glutamate contents in their brains. The body weights and motor coordination performance of the mice were measured 3 weeks after starting Dox treatment. Immediately after the motor coordination test, the mice were killed by decapitation, and the frontal cortex (FCX), hippocampus (HIP) and cerebellum (CER) were quickly dissected. The collected tissues were immediately frozen in liquid nitrogen and stored at -80°C until use. The frozen tissues were used for measuring GABA and glutamate contents. Another cohort was used for the open-field test and PPI test. Three weeks after treatment with Dox, the open-field test was conducted. After testing, the mice were returned to their home cage, and treatment with Dox was continued. Two days after the open-field test, the acoustic startle responses and prepulse inhibition (PPI) responses were assessed in the mice.

**Animals**
To generate Gad1STOP-tetO knock-in mice, we constructed the Gad1 targeting vector by linking the following elements in tandem: the 4.7-kb 5'-homology arm, 3.4-kb Neo-STOP-tetO cassette [28], 5.9-kb 3'-homology arm, and the MC1 promoter-driven diphtheria toxin A subunit gene (DT) (Fig. 1a). The Neo-STOP-tetO cassette comprised the 1.7-kb PGK-Neo cassette, a 1.3-kb STOP sequence, and a 0.5-kb tetO site. The targeting vector was designed to insert the Neo-STOP-tetO cassette just upstream of the Gad1 translation initiation site. We used B6-derived embryonic cells for homologous recombination. From 179 G418-resistant clones, we obtained 46 recombinant clones. Germline-transmitted offspring were established as Gad1STOP-tetO knock-in mice (Fig. 1a).

The generation of Gad1tTA knock-in mice has already been described [26]. In these mice, the tTA2 cDNA followed by the SV40 polyadenylation signal was inserted into exon 1 of the Gad1 gene in frame with the translation initiation codon, and the tTA protein was expressed under the control of an endogenous Gad1 promoter (Fig. 1b).

Heterozygous mice carrying one STOP-tetO allele (Gad1STOP-tetO/+ mice) were crossed with heterozygous mice carrying one tTA allele (Gad1tTA/+ mice) to obtain four genotypes: Gad1+/+, Gad1tTA/+, Gad1STOP-tetO/+ and Gad1tTA/STOP-tetO mice. To prevent the expression of GAD67 protein, we administered 100 mg of Dox per kg of regular mouse chow CE-2 (CLEA Japan, Inc.).

The animals were housed at 2-3 mice per cage (16.5 × 27 × 12.5 (H) cm) and had free access to food and water. The animal rooms for breeding and experiments were maintained at 22 ± 3°C with a 12-h light-dark cycle.
(lights on at 6:00, lights off at 18:00).

Genotyping

Genotyping of the transgenic mice was performed by PCR using tail genomic DNA with SapphireAmp Fast PCR master mix (Takara Bio Inc., Japan) and the specific primer sets. Primer set 1 determined the existence of the Neo allele (Fig. 1a); the sequences were Neo-F, 5’- CAGCTGTGCTCGACGTTGTC-3’ and Neo-R, 5’-AAGACCGGCTTCATCCGAG-3’. Primer set 2 determined the existence of the Gad1 and tTA-inserted Gad1 alleles (Fig. 1b); the sequences were Gad1-F, 5’- TGGTCTCCCTTCTGTCTCCA-3’, Gad1-R, 5’-TGTAGGCGCAGGTGAG-3’, and tTA-R, 5’-GGGCAAAAGTGAATGGTGCC-3’. After amplification, 5 μL of each reaction mixture and a size marker (Loading Quick 100 bp DNA Ladder, TOYOBO Co. Ltd, Osaka, Japan) were analyzed by 2% agarose gel electrophoresis, and the bands were then visualized by ethidium bromide staining. The lengths of the amplified DNA fragments were 224 bp (Neo allele), 229 bp (Gad1 allele) and 357 bp (tTA-inserted Gad1 allele) (Figs. 1e and 1f).

Palate formation

Mouse neonates were killed by decapitation, and the lower jaw was removed. The cleft palate of the mouse was determined under a stereoscopic microscope.
**Immunoblot analysis**

The mice were killed by decapitation. The brain hemispheres of neonates and the FCX, HIP and CER of adult mice were quickly dissected on an ice-cold stainless plate. The tissues were immediately frozen in liquid nitrogen and stored at -80°C until use. The frozen tissues were homogenized in ice-cold buffered sucrose (0.32 M) solution containing 20 mM Tris-HCl (pH 7.5) and protease inhibitor cocktail (P8340, Sigma-Aldrich, Inc.). The homogenates were centrifuged at 1,000 g for 10 min at 4°C, and the supernatants were collected as the protein samples. The protein concentrations were determined using a TaKaRa BCA Protein Assay Kit (T9300A, Takara Bio Inc., Japan).

The protein samples were diluted with electrophoresis sample buffer. Proteins (1.5 μg) were separated by 8% SDS-polyacrylamide gels and transferred to a PVDF membrane. Blots were probed with respective antibodies to GAD65/67 (1:1,000, rabbit polyclonal antibody) [29] and GAD67 (1:1,000, mouse monoclonal antibody, Millipore, Code No. MAB5406). Immunoblots were developed using horseradish peroxidase-conjugated secondary antibodies (GE Healthcare) and then detected with chemiluminescence reagents (ECL prime, GE Healthcare) and visualized by the Light Capture AE-9672 (ATTO Co., Ltd.). After the detection of immunoblots, the blotting membranes were washed with PBS several times and reprobed with a mouse monoclonal antibody to β-actin (1:10,000, Medical & Biological Laboratories Co. Ltd., Code No. M177-3). The immunoblots of β-actin...
were developed and visualized by the same protocol described above. The density of the bands was determined using ImageJ software. The band densities of β-actin were used as the loading control. The relative expression level of GAD67 to β-actin was calculated and used for comparisons between the genotypes.

**Double-label immunofluorescence analysis**

Deeply anesthetized mice by continuous inhalation of isoflurane were fixed by perfusion with Mildform 10N (containing 3.7 - 4.3 w/w% formaldehyde; FUJIFILM Wako Pure Chemical Co., Osaka, Japan) through the left ventricle. The brain was removed and postfixed in Mildform 10N overnight at 4°C. The brain hemispheres were cut into 50-μm-thick sagittal sections by a vibrating blade tissue slicer (Neo-LinearSlicer MT, Dosaka EM Co., Ltd., Kyoto, Japan).

Free-floating immunostaining was performed by using a VECTOR M.O.M.® (Mouse on Mouse) Immunodetection Kit (BMK-2202, Vector Laboratories Inc., USA). The sections were incubated overnight at room temperature in the 1st primary antibodies against GAD67 (1:300, mouse monoclonal antibody, MAB5406, Millipore) and parvalbumin (PV) (1:300, guinea pig polyclonal antibody, PV-GP-Af1000, Frontier Institute Co., Ltd., Hokkaido, Japan) with the M.O.M. Blocking reagent after preincubation with 0.3% Triton X-100 in PBS. After rinsing, the sections were incubated in the M.O.M. Biothinylated Anti-Mouse IgG Reagent (1:300) with a secondary antibody (1:300, goat anti-guinea pig IgG conjugated with AlexaFluor488, A-11073, Invitrogen) for 30
min at room temperature. After rinsing, the sections were incubated in a solution containing Streptavidin-DyLight649 (1:50, SA-5649, Vector Lab.) and DAPI (1:500, D523, Dojindo Laboratories, Japan) for 30 min at room temperature. The stained sections were mounted on MAS-coated glass slides (Matsunami Glass Ind., Ltd., Osaka, Japan) with Fluoromount (K024, Diagnostic BioSystems, USA). Fluorescence images were captured with a fluorescence digital microscope (BZ-X800, Keyence, Osaka, Japan).

Three independent mice in the respective groups were assessed. The names of brain regions were referenced to the Allen Mouse Brain Atlas (https://alleninstitute.org/).

Motor coordination test

The performance of motor coordination in mice was tested by a rotarod apparatus (Ugo Basile, Comerio, Italy) according to a previous report [16]. Briefly, each mouse was placed in a separate lane of the apparatus on a rotating cylinder (3 cm diameter) at 20 rounds per minute. The latency until the mouse fell from the cylinder (up to 120 s) was recorded in three consecutive trials with 2 - 3 min intervals, and the median latency was used for the following analysis. If the mouse did not fall within 120 s, the latency to fall was recorded as 120 s.

Open-field test

Each mouse was placed in the center of an open-field apparatus (50 cm × 50 cm × 40 (H) cm) that was
illuminated by light-emitting diodes (30 lux at the center of the field) and allowed to move freely for 5 min. The data were collected and analyzed using ImageJ OF4 (O’Hara & Co., Ltd., Tokyo, Japan), which is modified software that is also based on the public domain ImageJ program. The procedure was performed according to our previous report [30].

Acoustic startle response and PPI test

An acoustic startle reflex measurement system (O’Hara & Co., Ltd., Tokyo, Japan) was used. The startle response was assessed with various stimulus intensities. Five times of 70 to 120 dB (70, 75, 80, 85, 90, 95, 100, 110, and 120 dB) white noise stimuli (40 ms) were presented in quasi-random order and random intertrial intervals (10 - 20 s). In the PPI session, mice experienced five trial types: no stimulus; startle stimulus (120 dB, 40 ms) only; prepulse 70 dB (20 ms, lead time 100 ms) and pulse 120 dB; prepulse 75 dB (20 ms, lead time 100 ms) and pulse 120 dB; and prepulse 80 dB (20 ms, lead time 100 ms) and pulse 120 dB. Each trial was repeated 10 times in quasi-random order and random intertrial intervals (10 - 20 s). PPI was defined as the percent decline of the startle response: 100 - [(startle amplitude after prepulse and pulse)/(startle amplitude after pulse only)] × 100. The procedure was performed according to our previous report [19].

GABA and glutamate contents in the brains
The frozen tissues were weighed and then homogenized by BioMasher II (Nippi, Inc., Tokyo, Japan) in 500 μL of 0.1% formic acid in acetonitrile (Wako, Tokyo, Japan) containing an internal standard 2-morpholinoethanesulfonic acid (2-MES; Dojindo, Tokyo, Japan). The standard was spiked at a final concentration of 10 μM. The homogenates were centrifuged at 15,000 g for 15 min at 4°C, and then, the supernatants were collected and filtered through an ISOLUTE PLD+ column (Biotage Japan Ltd., Tokyo, Japan). The 40 μL filtrates were lyophilized and stored at -20°C.

At the time of analysis, the lyophilized samples were dissolved in 1.25 mL of ultrapure water. The prepared sample solutions (3 μL) were then injected on ultra-performance liquid chromatograph coupled to triple-quadrupole mass spectrometer (LC/MS) (LCMS-8050; Shimadzu, Kyoto, Japan). The chromatographic conditions were according to the Shimadzu method package using a pentafluorophenylpropyl (PFPP) column. The MS settings, data acquisition and data analysis were in accordance with the manufacturer’s instructions for analyzing Primary Metabolites version 2.0 (Cat. #: 225-24865A, Shimadzu). The relative values of metabolites from the internal standard 2-MES and the weight of corresponding tissues were calculated and used for the following data analysis.

Standard solutions containing GABA (A2129, Sigma-Aldrich Co. LLC., USA) and L-glutamic acid (G1251, Sigma-Aldrich) at dose ranges of 0.01 - 3 μmol/L and 0.03 - 10 μmol/L, respectively, with internal standard 2-MES (10 μM) were also applied to the LC/MS system. The concentrations of GABA and glutamate in the
sample solutions were determined by the peak heights of the chromatogram. The GABA and glutamate contents per the corresponding tissue weights were calculated.

Statistical analysis

Statistical analyses were conducted using BellCurve for Excel ver. 3.20 (Social Survey Research Information Co., Ltd., Tokyo, Japan). Significant differences among the multiple groups were analyzed by the Bonferroni multiple comparison test after one-way analysis of variance (ANOVA). Significant differences between two groups were analyzed by Student’s t-test. The factorial comparisons in some experiments were performed by two-way ANOVA. Data are expressed as the mean with standard error (SE). Statistical significance was defined as a p value less than 0.05.

Results

Generation of Gad1STOP-tetO knock-in mice

To confirm the elimination of function of the Gad1 gene by inserting the Neo-STOP-tetO cassette, we first generated homozygous Gad1STOP-tetO/STOP-tetO mice by crossing heterozygous Gad1STOP-tetO/+ parents and assessed whether the Gad1STOP-tetO/STOP-tetO mouse showed the Gad1 knockout phenotypes. Gad1STOP-tetO/STOP-tetO mice, Gad1STOP-tetO/+ mice and Gad1+/* mice were born at the expected Mendelian frequency (Fig. 2a). Then, mouse
pups were divided into two groups to determine the survival rates and palate formation. All \( \text{Gad1}^{\text{STOP-tetO/STOP-tetO}} \) mice died within 1 day after birth, but \( \text{Gad1}^{\text{STOP-tetO/+}} \) mice and \( \text{Gad1}^{+/+} \) mice survived (Fig. 2b). All \( \text{Gad1}^{\text{STOP-tetO/+}} \) and \( \text{Gad1}^{+/+} \) mice formed normal palate. However, 57% of \( \text{Gad1}^{\text{STOP-tetO/STOP-tetO}} \) mice exhibited a cleft palate (Fig. 2c). These phenotypes are the same as those of \( \text{Gad1}^{+/+} \) mice previously reported [31]. Western blot analyses demonstrated that the expression of GAD67 protein in the brain was abolished in \( \text{Gad1}^{\text{STOP-tetO/STOP-tetO}} \) mice with or without cleft palate (Figs. 2d and 2e). These observations indicate that the insertion of the Neo-STOP-tetO cassette following the \( \text{Gad1} \) promoter eliminates the function of the \( \text{Gad1} \) gene in mice.

Development of GAD67 knockdown mice

We then crossed heterozygous male \( \text{Gad1}^{\text{tTA/+}} \) mice and female \( \text{Gad1}^{\text{STOP-tetO/+}} \) mice to generate \( \text{Gad1}^{\text{tTA/STOP-tetO}} \) biallelic knock-in mice. Mice with four genotypes (\( \text{Gad1}^{+/+} \), \( \text{Gad1}^{\text{tTA/+}} \), \( \text{Gad1}^{\text{STOP-tetO/+}} \), \( \text{Gad1}^{\text{tTA/STOP-tetO}} \)) were born at the expected Mendelian frequency (Fig. 3 legend). Palate formation in mouse neonates with four genotypes was observed, but none of them exhibited the cleft palate (Fig. 3a). Next, the survival rates of these mice were examined in a small population. All \( \text{Gad1}^{+/+} \) mice (total n = 15) survived until 8 weeks of age (Fig. 3b). Two \( \text{Gad1}^{\text{tTA/+}} \) mice (total n = 15), one \( \text{Gad1}^{\text{STOP-tetO/+}} \) mouse (total n = 17), and eight \( \text{Gad1}^{\text{tTA/STOP-tetO}} \) mice (total n = 20) died within 8 weeks after birth (Fig. 3b). The proportion of genotypes in our breeding colony (total n = 1,319) at the weaning period (P21 - P28) is shown in Fig. 3c. These observations
indicate that the survival rate of $Gad1^{TA/STOP-tetO}$ mice was lower than that of mice with the other genotypes. The protein levels of GAD67 in the FCX ($F(3,8) = 4.355, p = 0.043$, one-way ANOVA) and HIP ($F(3,8) = 10.527, p = 0.004$, one-way ANOVA) were significantly lower in $Gad1^{TA/STOP-tetO}$ mice than $Gad1^{+/+}$ mice at 8 weeks of age (Figs. 3d and 3e). On the other hand, the protein levels of GAD67 in the CER were not significantly different among the genotypes ($F(3,8) = 2.867, p = 0.104$, one-way ANOVA) (Fig. 3f).

Next, the expression levels of GAD67 protein in the brains of $Gad1^{TA/STOP-tetO}$ mice were examined in the absence and presence of Dox treatment. We noticed that some $Gad1^{TA/STOP-tetO}$ mice died during the 3 weeks after starting the Dox treatment. Approximately 44% of $Gad1^{TA/STOP-tetO}$ mice survived just after 3 weeks of Dox treatment (Fig. 4a). The expression levels of GAD67 protein in the FCX ($F(3,8) = 20.563, p < 0.001$, one-way ANOVA), HIP ($F(3,8) = 189.298, p < 0.001$, one-way ANOVA) and CER ($F(3,8) = 22.760, p < 0.001$, one-way ANOVA) were significantly decreased by treatment with Dox in $Gad1^{TA/STOP-tetO}$ mice compared with $Gad1^{+/+}$ mice (Figs. 4b – 4d). Importantly, the expression level of GAD67 protein in the CER of $Gad1^{TA/STOP-tetO}$ mice was markedly decreased in the presence of Dox compared with in the absence of Dox (Fig. 4d). By immunofluorescence, GAD67 immunoreactivity in $Gad1^{+/+}$ mice was detected widely in the brain, particularly at high levels in the olfactory bulb, globus pallidum, olfactory tubercle, substantia nigra, superior and inferior colliculi, and deep cerebellar nuclei (Fig. 4e, upper panel). In brains of $Gad1^{TA/STOP-tetO}$ mice, the overall immunoreactivity was reduced moderately without Dox treatment and severely with the treatment. Dox treatment to $Gad1^{+/+}$ mice did not affect GAD67 immunoreactivity (data not shown). PV is expressed in a major subclass of
GAD67-positive inhibitory neurons [32]. No discernible changes in PV immunoreactivity were found between
Gad1<sup>+/+</sup> mice and Gad1<sup>TA/STOP-tetO</sup> mice with or without Dox treatment (Fig. 4e, lower panel). We assessed 3
independent mice in the respective groups and observed similar findings. These results suggest that Dox treatment
globally suppresses the expression of GAD67 in the brains of Gad1<sup>TA/STOP-tetO</sup> mice.

Behavioral abnormalities in GAD67 knockdown mice

We compared the behavioral phenotypes of Gad1<sup>TA/STOP-tetO</sup> mice with those of Gad1<sup>+/+</sup> mice in the presence
of Dox treatment. To avoid the effects of sex differences, male mice were only used in the following experiments.

We first investigated the body weights and the performance of motor coordination in Dox-treated
Gad1<sup>TA/STOP-tetO</sup> and Gad1<sup>+/+</sup> mice. No difference was observed in the body weights (t(17) = 1.066, p = 0.301,
Student’s t-test, Fig. 5a) or the latency to fall from the cylinder in the rotarod test (t(17) = 0.772, p = 0.451,
Student’s t-test, Fig. 5b) between the two genotypes.

We next conducted the open-field test, which is a well-accepted behavioral test to evaluate the anxiety-like
state of rodents [33]. The total distance, total duration of movement, moving speed, distance per movement, and
duration per movement were significantly increased in Dox-treated Gad1<sup>TA/STOP-tetO</sup> mice compared with
Dox-treated Gad1<sup>+/+</sup> mice (Table 1). In contrast, the total number of movement episodes was significantly
decreased in Dox-treated Gad1<sup>TA/STOP-tetO</sup> mice compared with Dox-treated Gad1<sup>+/+</sup> mice (Table 1). These
observations indicate that $Gad1^{\text{TA/STOP-tetO}}$ mice walk longer distances with less frequency. In addition, Dox-treated $Gad1^{\text{TA/STOP-tetO}}$ mice walked a long time in the wall side and a short time in the center region compared with Dox-treated $Gad1^{+/+}$ mice (Table 1 and Fig. 5c), indicating that $Gad1^{\text{TA/STOP-tetO}}$ mice exhibited anxiety-like behavior in the open-field test.

We further assessed acoustic startle responses and PPI responses in Dox-treated $Gad1^{\text{TA/STOP-tetO}}$ and $Gad1^{+/+}$ mice. The PPI response provides an operational index of sensorimotor gating, and an impaired PPI response is observed in subjects with schizophrenia [34]. The amplitude of acoustic startle responses was significantly affected by the effect of genotype × sound level interaction ($F(8,168) = 2.745, p = 0.007$, two-way ANOVA). The simple main effect of genotypes was statistically significant at the sound levels of 110 dB ($F(1, 92) = 6.345, p = 0.014$) and 120 dB ($F(1, 92) = 13.550, p < 0.001$) (Fig. 5d). The PPI responses were significantly affected by the effect of prepulse intensity ($F(2,42) = 6.713, p = 0.003$, two-way ANOVA) but not by the effect of genotype × prepulse intensity interaction ($F(2,42) = 0.577, p = 0.566$, two-way ANOVA) or genotype ($F(1,21) = 0.514, p = 0.481$, two-way ANOVA) (Fig. 5e).

**Brain GABA and glutamate content in GAD67 knockdown mice**

The GABA content was significantly lower in Dox-treated $Gad1^{\text{TA/STOP-tetO}}$ mice than in $Gad1^{+/+}$ mice in the FCX, HIP and CER (Table 2). On the other hand, the glutamate content in the respective brain regions was
comparable between these genotypes (Table 2).

Discussion

We first generated $Gad1^{STOP-tetO}$ knock-in mice. All homozygous $Gad1^{STOP-tetO/STOP-tetO}$ mice died on the day of birth, and 57% of $Gad1^{STOP-tetO/STOP-tetO}$ mice exhibited a cleft palate. The expression of GAD67 protein was lacking in the brains of $Gad1^{STOP-tetO/STOP-tetO}$ mice with or without the cleft palate. These phenotypes in $Gad1^{STOP-tetO/STOP-tetO}$ mice are consistent with those in $Gad1^{+/}$ mice [15,31]. Therefore, the function of the $Gad1$ gene was eliminated by the insertion of the Neo-STOP-tetO cassette in the 5'-untranslated region of the $Gad1$ gene in mice. It has been reported that neonatal death in $Gad1^{+/}$ mice is caused by respiratory failure rather than impairment of suckling [15,35]. Therefore, neonatal death in $Gad1^{STOP-tetO/STOP-tetO}$ mice may also be caused by respiratory failure.

We next developed $Gad1^{TA/STOP-tetO}$ biallelic knock-in mice by crossing $Gad1^{TA+/}$ and $Gad1^{STOP-tetO/+}$ parents. Approximately 40% of $Gad1^{TA/STOP-tetO}$ mice died on the day of birth, and the number of $Gad1^{TA/STOP-tetO}$ mice at P21-P28 in our breeding colony was smaller than the numbers of mice with the other genotypes. None of the $Gad1^{TA/STOP-tetO}$ mice demonstrated a cleft palate. Unexpectedly, some adult $Gad1^{TA/STOP-tetO}$ mice died by treatment with Dox. Therefore, GAD67 is important for survival not only in the neonatal period but also in adulthood. However, the cause of death in Dox-treated $Gad1^{TA/STOP-tetO}$ mice is currently unknown. To resolve this
question, pathological examination is required in a future study.

Adult mice with Gad1 haplodeficiency demonstrated an approximately 40% reduction in GAD67 protein levels in the whole brain compared with Gad1<sup>+/+</sup> mice [27]. Consistently, we observed that heterozygous Gad1<sup>TATA+</sup> and heterozygous Gad1<sup>STOP-tetO+</sup> knock-in mice exhibited a 30 - 50% reduction in GAD67 protein levels in the FCX, HIP and CER compared with Gad1<sup>+/+</sup> mice. In the absence of Dox treatment, the expression level of GAD67 protein in Gad1<sup>TATA/STOP-tetO</sup> mice relative to Gad1<sup>+/+</sup> mice was dependent on the brain regions. In the immunoblotting analysis, the expression of GAD67 protein in the CER was comparable between Gad1<sup>TATA/STOP-tetO</sup> mice and Gad1<sup>+/+</sup> mice. However, the expression of GAD67 protein in the FCX and HIP was significantly lower in Gad1<sup>TATA/STOP-tetO</sup> mice than Gad1<sup>+/+</sup> mice. Importantly, in the presence of Dox treatment, GAD67 expression was reduced by approximately 90% in the brains of Gad1<sup>TATA/STOP-tetO</sup> mice, compared with Dox-treated Gad1<sup>+/+</sup> mice. The brain-wide reduction of GAD67 expression in Dox-treated Gad1<sup>TATA/STOP-tetO</sup> mice was also observed in the immunofluorescence analysis. These findings suggest that GAD67 expression is suppressed by treatment with Dox in the brains of Gad1<sup>TATA/STOP-tetO</sup> mice.

In adult mice with Gad1 haplodeficiency, the GABA content in the brain was reduced by 7 - 20% from those in wild-type control mice [15,32]. In this study, we found that the GABA contents in the FCX, HIP and CER of Dox-treated Gad1<sup>TATA/STOP-tetO</sup> mice were reduced by 27.0% - 56.8% from those in Dox-treated Gad1<sup>+/+</sup> mice. Therefore, the GABA reduction in the brains of Dox-treated Gad1<sup>TATA/STOP-tetO</sup> mice was larger than that in Gad1
haplodeficient mice. Because approximately half of the brain GABA is produced by GAD65 in adulthood [36],
the remaining GABA in the brain of Dox-treated \textit{Gad1}^{TA/STOP-tetO} mice is mainly synthesized by GAD65. The
brain glutamate contents in Dox-treated \textit{Gad1}^{TA/STOP-tetO} mice were comparable to those in Dox-treated \textit{Gad1}^{+/-}
mice. Therefore, the glutamatergic system may be normal in Dox-treated \textit{Gad1}^{TA/STOP-tetO} mice.

GAD67 haplodeficient mice demonstrated several abnormal behaviors, such as hyper-locomotor activity,
reduced interactions with an unfamiliar mouse, and aggressive behavior. However, the emotional behaviors in
\textit{Gad1} haplodeficient mice were normal in the open-field test, the light-dark avoidance test and the elevated
plus-maze test [21,23]. In the current open-field test, Dox-treated \textit{Gad1}^{TA/STOP-tetO} mice walked longer distances
than Dox-treated \textit{Gad1}^{+/-} mice. In addition, Dox-treated \textit{Gad1}^{TA/STOP-tetO} mice preferentially walked for more time
along the walls and for less time in the center region. These observations indicate that Dox-treated \textit{Gad1}^{TA/STOP-tetO}
mice exhibited behavioral abnormalities, including the hyper-locomotor activity and anxiety-like behavior, in the
open-field test. Since \textit{Gad1}^{TA/STOP-tetO} mice exhibited normal body weight and motor coordination in the presence
of Dox treatment, the changes in exploratory behavior are unlikely to be associated with physical dysfunction. It
is well accepted that inhibition of GABAergic tone elicits anxiety-like behavior in the open-field test [33].
Therefore, the reduction in brain GABA in Dox-treated \textit{Gad1}^{TA/STOP-tetO} mice may cause the induction of
anxiety-like behavior. The mice lacking GAD67 in protein phosphatase 1 regulatory subunit 2
(Ppp1r2)-expressing cells, in which Cre recombinase expression is largely confined to GABA interneurons of the
cerebral cortex and the hippocampus, demonstrated the hyper-locomotor activity and anxiety-like behavior in the
open-field test [37]. In addition, we recently reported that mice lacking GAD67 in somatostatin-expressing GABA interneurons demonstrated anxiety-like behavior in the open-field test without affecting locomotor activity [20]. Therefore, the anxiety-like behavior in Dox-treated Gad1\textsuperscript{T/STOP-tetO} mice may be due to GAD67 knockdown from somatostatin-expressing GABA interneurons in cortical and hippocampal areas. In addition, the hyper-locomotor activity in Dox-treated Gad1\textsuperscript{T/STOP-tetO} mice may be associated with GAD67 knockdown from the other subtypes of GABA interneurons.

In this study, Dox-treated Gad1\textsuperscript{T/STOP-tetO} mice showed a normal response to PPI. Therefore, global knockdown of GAD67 does not affect the PPI response. However, we previously reported that mice with conditional Gad1 heterozygous knockout predominantly in parvalbumin-positive cells demonstrated an impaired response to PPI [19]. It is possible that a mild reduction in GAD67 in parvalbumin-positive cells might be required for impairing the PPI response. Interestingly, the startle responses elicited by large acoustic stimuli were impaired in Dox-treated Gad1\textsuperscript{T/STOP-tetO} mice. GABAergic interneurons expressing GAD67 proteins constitute auditory neural networks and contribute to auditory function [38–40]. Therefore, it is possible that global knockdown of GAD67 in the brain induces the impairment of auditory function.

The GAD1 gene and GAD67 protein have often been targeted in human studies to elucidate an association with the pathophysiology of psychiatric disorders [41–43]. Reduced full-length GAD1 transcript and GAD67 protein is a consistent finding in the postmortem brains of patients with several psychiatric disorders including
schizophrenia, bipolar disorder and major depressive disorder [7–12]. Because GAD67 reduction was predominantly observed in parvalbumin-positive GABAergic interneurons in the postmortem brains of schizophrenic patients [44–46], mice with conditional knockout of GAD67 in parvalbumin-positive cells have been used as an animal model of schizophrenia [18,19,47,48]. On the other hand, the specific subsets of GABAergic interneurons reducing GAD67 expression have not yet been identified in other psychiatric disorders, which might indicate that GAD67 expression is globally reduced in the brain of subjects with the other psychiatric disorders. In this study, global knockdown of GAD67 elicited anxiety-like behavior in mice. Therefore, we suggest the possibility that the global reduction in GADI transcript and GAD67 protein in the brain might be related to the occurrence of anxiety symptoms frequently comorbid in several psychiatric disorders.

We did not examine whether administration of anxiolytic, antidepressant or antipsychotic drugs improves anxiety-like behavior in Dox-treated Gad1tTA/STOP-tetO mice. Because of the high lethality rate, we could not immediately prepare a sufficient number of animals for conducting a pharmacological study. Therefore, we will determine the utility of Dox-treated Gad1tTA/STOP-tetO mice as a tool for screening potential medications for anxiety symptoms in a future study.

In summary, Gad1tTA/STOP-tetO biallelic knock-in mice showed GAD67-knockdown phenotypes when treated with Dox. We suggest that the global reduction in GAD67 elicits emotional and auditory abnormalities in mice. The use of GAD67 knockdown mice will provide new insights into the neurobiological impact of GAD67.
dysfunction and elucidate the neurobiological mechanisms of emotional abnormalities associated with psychiatric disorders.

Abbreviations

GAD67: glutamate decarboxylase 67; GAD65: glutamate decarboxylase 65; Dox: doxycycline; GABA: \( \gamma \)-aminobutyric acid; tTA: tetracycline-controlled transcriptional activator; tetO: tetracycline operator site; FCX: frontal cortex; HIP: hippocampus; CER: cerebellum; CP: cleft palate; PV: parvalbumin; PPI: prepulse inhibition; ANOVA: analysis of variance; SE: standard error

Declarations

Ethics approval and consent to participate

This study was performed in accordance with the Guidelines for Animal Experimentation at Gunma University Graduate School of Medicine and was approved by the Gunma University Ethics Committee (Permit number: 14-006 and 19-009). Every effort was made to minimize the number of animals used and their suffering.
Consent for publication

Not applicable.

Availability of data and materials

All data are available within the manuscript.

Competing interests

The authors declare that they have no conflicts of interest with the contents of this manuscript.

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**Authors’ contributions**

S.M. and Y.Y. conceptualization; S.M. data curation; S.M. formal analysis; Y.Y. and K.S. funding acquisition; S.M., H.O. and T.H. investigation; S.M., T.K., K.F. and Y.Y. methodology; S.M. project administration; J.N., M.T., S.I., M.W., K.T., M.A. and K.S. resources; Y.Y. supervision; S.M. and Y.Y. writing-original draft; all authors writing-review&editing.

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Legends for Figures

Fig. 1. Doxycycline (Dox)-regulated Gad1 suppression system. (a) Generation of Gad1<sup>STOP</sup>-tetO knock-in mice. Schematic diagram depicting Gad1 genomic DNA (Gad1<sup>+</sup>), targeting vector, and Gad1 genomic DNA inserted into the Neo-STOP-tetO cassette (Gad1<sup>STOP</sup>-tetO). Arrows indicate the PCR primers (Primer set 1). (b) Schematic representation of Gad1 genomic DNA (Gad1<sup>+</sup>) and DNA containing the tetracycline-controlled transactivator (tTA) gene (Gad1<sup>tTA</sup>). Arrows indicate the PCR primers (Primer set 2). (c, d) Schematic diagram of the Dox-regulated Gad1 suppression system in Gad1<sup>tTA/STOP-tetO</sup> mice. Before Dox treatment, tTA binds to the tetracycline operator site (tetO) and promotes Gad1 transcription and GAD67 production (c). Dox treatment interferes with tTA binding to tetO and suppresses Gad1 transcription (d). Representative results of PCR genotyping for Gad1<sup>tTA/STOP-tetO</sup> mice (e), Gad1<sup>STOP-tetO/STOP-tetO</sup> mice (f) and their littermates. M; 100-bp size marker, +/-; Gad1<sup>+/+</sup>, tTA/tet; Gad1<sup>tTA/STOP-tetO</sup>, tTA/+; Gad1<sup>tTA/+</sup>, tet/+; Gad1<sup>STOP-tetO/+</sup>, tet/tet; Gad1<sup>STOP-tetO/STOP-tetO</sup>.

Fig. 2. Characterization of homozygous Gad1<sup>STOP-tetO/STOP-tetO</sup> mice. (a) The number of mouse pups born from heterozygous Gad1<sup>STOP-tetO/+</sup> parents. +/-; Gad1<sup>+/+</sup>, tet/+; Gad1<sup>STOP-tetO/+</sup>, tet/tet; Gad1<sup>STOP-tetO/STOP-tetO</sup>. (b) The survival rates of mouse pups born from heterozygous Gad1<sup>STOP-tetO/+</sup> parents. (c) Microscopic images of the palate of mice. The arrow indicates a cleft palate (CP). (d) Representative western blot analysis of the expression of GAD65/67 and β-actin proteins in the brains of Gad1<sup>+/+</sup>, Gad1<sup>STOP-tetO/+</sup> and Gad1<sup>STOP-tetO/STOP-tetO</sup> mice. (e) Western
blot analysis of the expression of GAD65/67 and β-actin proteins in the brains of 3 Gad1STOP-tetO/STOP-tetO mice with and without CP. The Gad1+/+ mouse protein sample was used as a positive control for GAD67 immunoblotting.

**Fig. 3.** Characterization of Gad1TA/STOP-tetO mice in the absence of Dox treatment. (a) Microscopic images of the palates of mice. (b) The survival rates of Gad1+/+ (+/+), Gad1TA/STOP-tetO (tTA/tet), Gad1TA/+ (tTA/+) and Gad1STOP-tetO/+ (tet/+) mice. The number of mice at postnatal day 0 was 15 (+/+), 20 (tTA/tet), 15 (tTA+) and 17 (tet/+). (c) The number of mice with four genotypes at postnatal days 21 - 28. (d - f) Representative western blot analysis of the expression of GAD65/67 and β-actin proteins in the frontal cortex (d), the hippocampus (e) and the cerebellum (f) of Gad1+/+, Gad1TA/STOP-tetO, Gad1TA/+ and Gad1STOP-tetO/+ mice. β-Actin was used as a loading control. Three independent mice of the respective genotypes were examined. The band densities of GAD67 and β-actin were quantified by ImageJ software. GAD67 protein levels were normalized to β-actin protein expression, and the % change was calculated relative to Gad1+/+ mice. The means with SE are demonstrated as columns. *p < 0.05 vs. the value of Gad1+/+ mice (Bonferroni test).

**Fig. 4.** Characterization of Gad1TA/STOP-tetO mice in the presence of Dox treatment. (a) The survival rates of mice with Gad1+/+ (+/+), and Gad1TA/STOP-tetO (tTA/tet) mice during Dox treatment (weeks 0, 1, 2, and 3). (b - d) Representative western blot analysis of the expression of GAD65/67 and β-actin proteins in the frontal cortex (b),
the hippocampus (c) and the cerebellum (d) of Gad1+/+ and Gad1\textsuperscript{TA/STOP-tetO} mice treated and not treated with Dox.

β-Actin was used as a loading control. Three independent mice in the respective groups were examined. The band densities of GAD67 and β-actin were analyzed by ImageJ software. GAD67 protein levels were normalized to β-actin protein expression, and the % change was calculated relative to Gad1+/+ mice. The means with SE are demonstrated as columns. *p < 0.05 vs. the value of nontreated Gad1+/+ mice (Bonferroni test). #p < 0.05 vs. the value of nontreated Gad1\textsuperscript{TA/STOP-tetO} mice (Bonferroni test). (e) Representative immunoreactivities of GAD67 and parvalbumin (PV) proteins in the brain sections of nontreated Gad1+/+ mice, nontreated Gad1\textsuperscript{TA/STOP-tetO} mice and Dox-treated Gad1\textsuperscript{TA/STOP-tetO} mice. The white bars in the images indicate 2 mm length.

**Fig. 5.** Behavioral consequences of GAD67 knockdown in mice. Body weights (a) and latency to fall from a cylinder in the rotarod test (b) for Dox-treated Gad1+/+ and Gad1\textsuperscript{TA/STOP-tetO} mice. (c) Examples of the path traveled in the open-field test by two Gad1+/+ and two Gad1\textsuperscript{TA/STOP-tetO} mice treated with Dox. (d) Acoustic startle responses of Dox-treated Gad1+/+ and Gad1\textsuperscript{TA/STOP-tetO} mice. Startle amplitudes by the sounds indicate the startle responses (A.U.). *p < 0.05 and ***p < 0.001 between genotypes (simple main effect of two-way ANOVA). (e) PPI responses of Dox-treated Gad1+/+ and Gad1\textsuperscript{TA/STOP-tetO} mice. PPI was defined as the percent decline in the startle response (% Inhibition).