Association between Alcoholism and the Genetic Polymorphisms of the GABA<sub>A</sub> Receptor Genes on Chromosome 5q33-34 in Korean Population

INTRODUCTION

Alcoholism is one of the most common psychiatric diseases that have been well defined clinically (1), and has a strong genetic influence. Family, twin and adoption studies have convincingly demonstrated that genes play an important role in the development of alcoholism, accounting for approximately 40-60% of the population variance (2, 3). The alleged candidate genes of alcoholism are the γ-aminobutyric acid (GABA) gene, the serotonin receptor gene, the monoamine oxidase A promoter, the serotonin transporter regulatory gene, the dopamine type 2 receptor gene, the catechol-O-methyltransferase gene, the cannabinoid receptor gene, and the μ-opioid receptor gene (4). Despite the strong evidence that genetic effects contribute to the susceptibility of alcoholism, detecting the specific genes that increase or decrease the risk for alcoholism has proven difficult.

GABA is the major inhibitory neurotransmitter in the human central nervous system and is involved in many of the behavioral effects of alcohol, including motor incoordination, anxiolysis, sedation, withdrawal signs and ethanol preference (5, 6). There are two primary GABA receptor subtypes: the GABA<sub>A</sub> and GABA<sub>B</sub> receptors. The GABA<sub>A</sub> receptors act through intrinsic ion channels and are composed of multiple subunits, designated α, β, γ, δ, and ρ, with several identified genes coding for these subunits (5). The GABA<sub>A</sub> receptor agonists tend to potentiate the behavioral effects of alcohol, whereas the GABA<sub>A</sub> receptor antagonists attenuate these effects. The GABA<sub>A</sub> receptors have also been implicated in ethanol tolerance and dependence (6). However, the precise mechanisms by which the GABA<sub>A</sub> receptors are involved in these actions of ethanol remain unknown. A role for the GABA<sub>A</sub> receptors in the actions of ethanol has not been studied nearly as extensively as that of the GABA<sub>B</sub> receptors. Most of the GABA<sub>A</sub> receptor genes are organized into clusters; the GABA<sub>A</sub> α2, GABA<sub>A</sub> α3, GABA<sub>A</sub> β1 and GABA<sub>A</sub> γ1 gene cluster on chromosome 4p13-p12 (7); the GABA<sub>A</sub> δ2, GABA<sub>A</sub> β3 and GABA<sub>A</sub> γ3 gene cluster on chromosome 15q11-q13 (8); and the GABA<sub>A</sub> α1, GABA<sub>A</sub> α6, GABA<sub>A</sub> β2 and GABA<sub>A</sub> γ2 gene cluster on chromosome 5q33-34 (9).

Several recent genetic studies have addressed the importance of the GABA<sub>A</sub> receptor genes in the development of alcoholism. Animal studies have demonstrated that alterations in the response to drugs and alcohol may be caused by amino acid differences at the GABA<sub>A</sub> α6 and GABA<sub>A</sub> γ2 subunits (9). Several loci related to alcohol withdrawal were also identified on the mouse chromosome 11, which corresponds to the region containing four GABA<sub>A</sub> receptor genes on human chromo-
some 5q33-34 (10). However, human genetic studies have reported conflicting results about the role of the GABAA receptor genes on chromosome 5q33-34 in the development of alcoholism (11-14). Therefore, we explored the association between the genetic polymorphisms of the GABAA receptor genes on chromosome 5q33-34 and alcoholism in Korean population.

**MATERIALS AND METHODS**

**Subjects and clinical assessments**

The study subjects consisted of 162 male alcoholics who were hospitalized in alcoholism treatment institutes (mean age=45.9 yr, SD=7.0 yr) and 172 control subjects (mean age=43.1 yr, SD=11.4 yr). The study was approved by the Institutional Review Board of Gyeongsang National University.

The 334 participants were interviewed by two psychiatrists. Individuals were diagnosed with alcohol dependence using DSM-IV criteria (15), and alcoholic subjects who had major psychiatric histories, including schizophrenia, major depressive disorders, bipolar disorders, anxiety disorder and organic mental disorder, or who had difficulties participating in the objective interview procedures due to cognitive and communication impairments, were excluded from the study.

The control subjects were recruited from the staff and the men who visited the Health Examination Center at Gyeongsang National University Hospital in Jinju City. All the subjects were asked by a physician to participate in the study, and a written informed consent was obtained from every subject. None of control subjects had a family history of alcoholism and a lifetime history of any other psychiatric, neurological or medical disorders. The control and alcoholic subjects were age-matched.

Patients were interviewed to collect demographic data, including age, gender, occupation, educational level, economic status, marital status and alcohol related questions that inquired about the subjects’ alcohol drinking history and the alcohol-associated problematic history. Several standardized research scales were used for the clinical assessment of alcoholism and a lifetime history of any other psychiatric, neurological or medical disorders. The control and alcoholic subjects were age-matched.

**Genomic DNA extraction**

We collected a 3-4 mL venous blood sample, which were poured into 7.5% potassium-EDTA containing tubes, were extracted immediately or stored at -20°C for later use. Genomic DNA was extracted by using QIAamp DNA blood midi kit according to the manufacturer’s recommendations (QIAamp DNA blood midi kit, QIAGEN Inc., CA, U.S.A.). Briefly, 200 µL of Qiaqen protease and 2.4 mL of buffer AL were added to 2 mL of specimen. Samples were incubated at 70°C for 10 min after which time 2 mL of 100% ethanol was added to each. Each mixture was vortexed and pipetted into a QiAamp Spin column, which was then centrifuged at 1,850 × g for 3 min at room temperature. Samples were washed by the adding 2 mL of AW1 to each spin column, followed by centrifugation at 4,500 × g for 5 min. Washing was repeated with 2 mL of AW2, followed by centrifugation at 4,500 × g for 5 min. Spin columns were then placed in clean centrifuge tubes, and a 300 µL volume of AE was added to elute the DNA. Elution was accomplished with a final centrifugation step at 4,500 × g for 5 min. DNA yield is determined by measuring the concentration of DNA in the eluate by absorbance at 260 nm and 280 nm, an eluate containing 50-100 ng DNA/µL. Pure DNA has an A260/A280 ratio of 1.6-1.9.

**Determination of GABAA a1 receptor gene polymorphism**

PCR was carried out with the primers, forward (5'-GCT ATG GAT TGG TTT ATT GCC GTG TG-3) and reverse (5'-ATA ATA TTG ATG TAC TAG AGG GAC-3'). A 50 µL total volume of the reaction mixture contained the following: 100 ng genomic DNA, 30 pmole of each primer, 1.25 mM dNTP, 2.5 mM MgCl2, and 1 unit of Taq polymerase. After an initial 5 min denaturation at 95°C, the reaction mix was processed by use of a Perkin Elmer thermocycler according to the following program; 40 cycles of 20 sec at 95°C, 30 sec at 55°C, 40 sec at 72°C, with final extension for 10 min at 72°C. The PCR product (15 µL) was then electrophoresed in 2% agarose gel and visualized with ethidium bromide for viewing. Two units of Taq polymerase was subjected to 2% agarose gel electrophoresis and stained with ethidium bromide and ultraviolet light. If the nucleotide 15 of the last intron of the GABAA a1 is a G, the restriction enzyme cuts the 165 bp fragments into two fragments of 141 and 24 bp. If the nucleotide 15 is an A, the PCR fragment remains uncut.

**Determination of the GABAA a6 receptor gene polymorphism**

Forward exon primer, 5'-GGA GGC ACC AGT AAA ATA GAC CAG-3' and reverse intronic primer, 5'-AAT ACT GAA CAA TGG AAG ACA AAA G-3', were used to amplify a specific region of 423 bp fragment. The reaction mixture contained PCR buffer, 2.5mM MgCl2, 1 unit of Taq DNA polymerase, 1.25 mM dNTP, 30 pmole of each primer and 100 ng of genomic DNA. After an initial 5 min denaturation at 95°C, there followed 40 cycles of PCR (denaturation, 1 min 30 sec at 95°C; annealing, 30 sec at 56°C; polymerization, 40 sec at 72°C) with final extension for 10 min at 72°C on a Perkin Elmer GeneAmp 9600 thermocycler. The PCR product was subjected to 2% agarose gel electrophoresis and stained with ethidium bromide for viewing. Two units of Nhe I (Roche, Mannhein, Germany) were digested with 22.3 µL of the PCR products at 37°C for 4 hr. Electrophoresis of the
digest was carried out with 3% agarose gel, and the bands were visualized by ethidium bromide staining. If the nucleotide 1,519 in the non-coding region of the the GABA\(_{\alpha}\) receptor gene is a T, the restriction enzyme cuts the 423 bp fragments into two fragments of 257 and 166 bp. If the nucleotide 1,519 is a C, the PCR fragment remains uncut.

**Determination of the GABA\(_{\alpha}\) \(\beta2\) receptor gene polymorphism**

Forward exon primer, 5'-AAG CAC AAT GCT AGC CTA TGG TGC-3' and reverse intronic primer, 5'-GTT CAC ATA ATA AAG CCA ATA GAC GAT-3', were used to amplify a specific region of 253 bp fragment. The reaction mixture contained PCR buffer, 1.5 mM M\(g\)Cl\(_2\), 1 unit of Taq DNA polymerase, 1.25 mM dNTP, 30 pmole of each primer and 100 ng of genomic DNA. After an initial 5 min denaturation at 95\(^\circ\)C, the reaction mix was processed by use of a Perkin Elmer thermocycler according to the following program; 40 cycles of 30 sec at 95\(^\circ\)C, 30 sec at 56\(^\circ\)C, 40 sec at 72\(^\circ\)C, with final extension for 10 min at 72\(^\circ\)C. The PCR product was subjected to 2% agarose gel electrophoresis and stained with ethidium bromide for viewing. Two units of Nhe I (Roche, Mannhein, Germany) were digested with 22.3 \(\mu\)L of the PCR products at 37\(^\circ\)C for 4 hr. Electrophoresis of the digest was carried out with 3% agarose gel, and the bands were visualized by ethidium bromide staining. If the nucleotide 1,412 in the last exon before the non-coding region of the GABA \(_{\alpha}\) \(\beta2\) receptor gene is a C, Ban I digests the 253 bp fragments into two fragments of 229 and 24 bp. If the nucleotide 1,412 is a T, the PCR fragment remains uncut.

**Determination of the GABA\(_{\alpha}\) \(\gamma2\) receptor gene polymorphism**

Forward exon primer, 5'-AAA GAT AAA AAG AAG AAA AAC CCT-3' and reverse intronic primer, 5'-CAC AGA AAA TAG AAA CAG ACT TGA-3', were used to amplify a specific region of 224 bp fragment. The reaction mixture contained PCR buffer, 1.5 mM M\(g\)Cl\(_2\), 1 unit of Taq DNA polymerase, 1.25 mM dNTP, 30 pmole of each primer and 100 ng of genomic DNA. After an initial 5 min denaturation at 95\(^\circ\)C, the reaction mix was processed by use of a Perkin Elmer thermocycler according to the following program; 40 cycles of 30 sec at 95\(^\circ\)C, 30 sec at 56\(^\circ\)C, 40 sec at 72\(^\circ\)C, with final extension for 10 min at 72\(^\circ\)C. The PCR product was subjected to 2% agarose gel electrophoresis and stained with ethidium bromide for viewing. Two units of Nhe I (Roche, Mannhein, Germany) were digested with 22.3 \(\mu\)L of the PCR products at 37\(^\circ\)C for 4 hr. Electrophoresis of the digest was carried out with 3% agarose gel, and the bands were visualized by ethidium bromide staining. If the nucleotide 1,142 in the last exon before the non-coding region of the GABA\(_{\alpha}\) \(\gamma2\) receptor gene is a C, Bam I digests the 224 bp fragments into two fragments of 229 and 24 bp. If the nucleotide 1,142 is a T, the PCR fragment remains uncut.

**Statistical analysis**

For the case-control genetic comparisons, differences in genotype and allele frequencies between groups were evaluated by using the chi-square test or Fisher's exact test, where appropriate. Univariate comparisons of clinical data between groups were conducted with independent t-tests or ANOVA were used. The Duncan test was used for the post-hoc analysis. Since we compare genotypic and allelic distributions of four independent polymorphic markers between healthy comparison subjects and alcoholism, a Bonferroni correction was applied to correct for multiple testing level of significant was set to alpha<0.05/4=0.0125). The Hardy-Weinberg equilibrium was tested for by the chi-square test. All significance levels were set at p<0.05 and the statistical analyses were run on SPSS software, version 11.0 (SPSS, Chicago, IL, U.S.A.).

**RESULTS**

Genotype and allele frequencies of the GABA\(_{\alpha}\) receptor gene polymorphisms

The genotype frequency of the polymorphisms did not deviate from that expected according to Hardy-Weinberg equilibrium (GABA\(_{\alpha}\) \(g1\), \(p=0.893\); GABA\(_{\alpha}\) \(g6\), \(p=0.892\); GABA\(_{\alpha}\) \(\beta2\), \(p=0.650\); GABA\(_{\alpha}\) \(\gamma2\), \(p=0.918\)). When compared with the controls, an association between polymorphisms of the GABA\(_{\alpha}\) \(g1\) and \(g6\) receptor gene was with alcohol dependence. Allelic distribution of polymorphisms of the GABA\(_{\alpha}\) \(g1\) and \(g6\) receptor gene was associated with alcohol dependence. These associations remained statistically significant even after Bonferroni correction. However, there were no significant associations between alcohol dependence and the genetic polymorphisms of the GABA\(_{\alpha}\) \(\beta2\) and GABA\(_{\alpha}\) \(\gamma2\) receptor genes (Table 1).

**Association of the clinical characteristics with genetic polymorphisms of the GABA\(_{\alpha}\) receptor genes**

Genetic polymorphisms of the GABA\(_{\alpha}\) \(g1\) receptor gene were associated with the mean onset age of alcoholism and the alcohol withdrawal symptoms and mean score of the ADS. The GG genotype of the GABA\(_{\alpha}\) \(g1\) receptor gene was associated with the early onset of alcoholism and the alcohol withdrawal symptoms. The ADS mean score was significantly higher for alcoholics with the GG genotype than for the alcoholics with the AA and AG genotypes of the GABA\(_{\alpha}\) \(g1\) receptor gene. However, the GABA\(_{\alpha}\) \(g1\) receptor gene polymorphisms were not associated with the scores of the BDI and OCDS. There were no associations found between the other
Table 1. Genotype and allele frequencies of the GABA<sub>a</sub> receptor gene polymorphisms on chromosome 5q33-34 in alcoholic and control subjects

| Genotype | Allele |
|----------|--------|
| GABA<sub>a</sub> 1 | AA | AG | GG | p | A | G | p |
| **receptor gene** | | | | | | | |
| Alcoholics | 28 | 70 | 64 | 0.002 | 126 | 198 | 0.001 |
| (17.3) | (43.2) | (39.5) | (38.9) | (61.1) |
| Controls | 42 | 92 | 38 | 176 | 168 |
| (24.4) | (53.5) | (22.1) | (51.2) | (48.8) |
| GABA<sub>a</sub> 2 | CC | CT | TT | p | C | T | p |
| **receptor gene** | | | | | | | |
| Alcoholics | 54 | 72 | 36 | 0.001 | 180 | 144 | 0.000 |
| (33.3) | (44.5) | (22.2) | (55.6) | (44.4) |
| Controls | 82 | 74 | 16 | 238 | 106 |
| (47.7) | (43.0) | (9.3) | (69.2) | (30.8) |
| GABA<sub>a</sub> 6 | CC | CT | TT | p | C | T | p |
| **receptor gene** | | | | | | | |
| Alcoholics | 160 | 12 | 0 | 0.250 | 306 | 18 | 0.197 |
| (88.9) | (7.0) | (0) | (94.4) | (5.6) |
| Controls | 144 | 18 | 0 | 332 | 12 |
| (93.0) | (7.0) | (0) | (96.5) | (3.5) |
| GABA<sub>b</sub> 2 | AA | AC | CC | p | A | C | p |
| **receptor gene** | | | | | | | |
| Alcoholics | 36 | 76 | 50 | 0.551 | 148 | 176 | 0.264 |
| (22.2) | (46.9) | (30.9) | (45.7) | (54.3) |
| Controls | 46 | 80 | 46 | 172 | 172 |
| (26.7) | (46.4) | (26.7) | (50.0) | (50.0) |

*Values are presented as number (proportion in the sample in %).

GABA<sub>a</sub> receptor gene polymorphisms and the clinical characteristics of the alcoholics (Table 2).

**DISCUSSION**

Alcoholism is a complex, multifaceted disorder that has long been recognized to run in families. There is substantial evidence from twin research and adoption studies that a major genetic component operates in the development of alcoholism. In this study, we investigated the association between alcoholism and the genetic polymorphisms of the GABA<sub>a</sub> receptor genes, and the association between the genetic polymorphisms and the clinical characteristics of alcoholics. We found that genetic polymorphisms of the GABA<sub>a</sub> a1 and a6 receptor genes were associated with alcoholism, but the GABA<sub>a</sub> b2 and y2 receptor genes were not. A number of groups have investigated the role of the GABA<sub>a</sub> receptor genes polymorphisms that are located on chromosome 5, and these studies have reported mixed results. In research on several populations, revealed a possible association between alcoholism and the genetic polymorphisms of the GABA<sub>a</sub> a1 or a6 receptor genes (11, 12). Although there was some evidence for association, previous studies also showed no association of the GABA<sub>a</sub> a6, b2 and y2 receptor genes polymorphisms with alcoholism or familial alcoholism (13, 14). These controversial results from other studies might be due to either ethnic

Table 2. Clinical characteristics and genetic polymorphisms of the GABA<sub>a</sub> receptor genes on chromosome 5q33-34

| Genotype | GABA<sub>a</sub> a1 receptor gene |
|----------|---------------------------------|
| Genotype | AA | AG | GG | p |
| Onset<sup>1</sup>, mean (SD) (yr) | 30.46 | 29.61 | 27.03 | 0.006<sup>*</sup> |
| (4.01) | (6.03) | (5.19) |
| Withdrawal<sup>1</sup>, mean (SD) (yr) | 41.50 | 39.00 | 35.97 | 0.001<sup>†</sup> |
| (1.69) | (5.80) | (5.44) |
| ADS, mean (SD) | 38.50 | 40.83 | 47.91 | 0.001<sup>†</sup> |
| (8.98) | (5.80) | (10.21) |
| BDI, mean (SD) | 36.38 | 38.13 | 40.47 | 0.272 |
| (12.89) | (11.38) | (11.45) |
| OCDS, mean (SD) | 17.85 | 16.48 | 19.25 | 0.073 |
| (6.69) | (6.32) | (7.11) |

| Genotype | GABA<sub>a</sub> a6 receptor gene |
|----------|---------------------------------|
| Genotype | CC | CT | TT | p |
| Onset<sup>1</sup>, mean (SD) (yr) | 27.84 | 29.58 | 28.24 | 0.214 |
| (5.42) | (7.79) | (5.67) |
| Withdrawal<sup>1</sup>, mean (SD) (yr) | 37.76 | 38.16 | 37.00 | 0.700 |
| (6.21) | (4.95) | (6.07) |
| ADS, mean (SD) | 42.63 | 43.17 | 44.22 | 0.722 |
| (9.42) | (8.13) | (10.74) |
| BDI, mean (SD) | 38.38 | 38.63 | 40.06 | 0.899 |
| (11.99) | (11.15) | (12.67) |
| OCDS, mean (SD) | 18.00 | 17.63 | 18.25 | 0.951 |
| (7.17) | (6.47) | (7.01) |

| Genotype | GABA<sub>a</sub> y2 receptor gene |
|----------|---------------------------------|
| Genotype | AA | AC | CC | p |
| Onset<sup>1</sup>, mean (SD) (yr) | 28.86 | 27.44 | 0 | 0.309 |
| (5.72) | (3.85) | (0) |
| Withdrawal<sup>1</sup>, mean (SD) (yr) | 37.83 | 37.29 | 0 | 0.738 |
| (6.38) | (5.32) | (0) |
| ADS, mean (SD) | 43.11 | 44.11 | 0 | 0.664 |
| (8.70) | (12.52) | (0) |
| BDI, mean (SD) | 38.84 | 38.67 | 0 | 0.954 |
| (11.85) | (10.85) | (0) |
| OCDS, mean (SD) | 17.99 | 17.11 | 0 | 0.610 |
| (6.97) | (5.41) | (0) |

<sup>*</sup> group difference, AA-AG (p=0.641), AA-GG (p=0.000); <sup>†</sup> group difference, AA-AG (p=0.300), AA-GG (p=0.000); 1, group difference, AA-AG (p=1.000), AA-GG (p=0.002), AG-GG (p=0.026); 2, group difference, AA-AG (p=0.452), AA-GG (p=0.004), AG-GG (p=0.013); 3, Onset age of alcoholism; 4, Onset age of the alcohol withdrawal symptoms; ADS, alcohol dependence scale; BDI, Beck depression inventory; OCDS, Obsessive-Compulsive Drinking Scale.
differences in the allele frequencies or the difference of the phenotype definition. Because Koreans are a unitary race that is genetically homogeneous (19), the subjects in this study have the merit of minimizing the stratification bias that can originate from association studies when the subjects of these studies consist of diverse races. Therefore, our results support the evidence that genetic polymorphisms of the GABA\(\alpha_1\) and \(\alpha_6\) receptor genes may have crucial roles for the development of alcoholism in Koreans.

The GG genotype of the GABA\(\alpha_1\) receptor gene was significantly associated with the early onset of alcoholism and the alcohol withdrawal symptoms. Furthermore, the GG genotype was associated with a high mean ADS score, which suggests a tendency toward severe alcoholism. Typology studies have suggested a way of reducing the etiological heterogeneity. The best known of these typologies is the Type 1/Type 2 distinction developed by Cloninger from the studies that were done on the adopted sons of Swedish alcoholics (2). Type 1 alcoholics are characterized by the late onset of problem drinking, few childhood risk factors, a prominence of guilt and anxiety related to drinking, relatively mild dependence, few alcohol related problems and little psychopathology. In contrast, type 2 alcoholics are characterized by the early onset of alcoholism, many childhood risk factors, significant psychopathology, a strong family history of alcohol abuse and the absence of guilt and fear concerning drinking (20). An adoption study based on Cloninger’s typology reported that the lifetime risk of severe alcoholism was increased six-fold for the adopted sons having the type 2 genetic background compared with type 1 subjects, regardless of their postnatal environment (21). In contrast, neither the genetic nor environmental risk factors for type 1 alcoholism were sufficient enough to cause alcoholism. These finding indicate that the age of onset and the antisocial behavior represent two characteristics that are susceptible to the genetic components of alcoholism. Our results support possibility that the GG genotype of the GABA\(\alpha_1\) receptor gene may be associated with Cloninger’s type 2 alcoholics. However, further evaluations for the antisocial behavior and the family history of alcoholics will be required to elucidate a positive association between Cloninger’s type 2 alcoholics and the genetic polymorphisms of the GABA\(\alpha_1\) receptor gene.

A possible limitation in case-control association studies is the risk of spurious associations as a result of population admixture. However, this objection is less likely in the present study, as our study group was composed of subjects of Korean descent for generations and because the Italian population is relatively homogeneous. In addition, the size of our sample is relatively small and replication of our results on a larger and independent sample is required. Moreover, another limitation was that this study lacked a structured diagnostic interview. Finally, only male patients were genotyped. Therefore, the generalization of the results to all alcoholics is limited.

In conclusion, there was a significant association between the genetic polymorphisms of the GABA\(\alpha_1\) and \(\alpha_6\) receptor genes and alcoholism. The GG genotype of the GABA\(\alpha_1\) receptor gene was associated with not only the onset age of alcoholism and the alcohol withdrawal symptom, but also the ADS score in Korean population. These results indicate that genetic polymorphisms of the GABA\(\alpha_1\) and \(\alpha_6\) receptor genes may have a crucial role in the development of alcoholism. In particular, the GG genotype of the GABA\(\alpha_1\) receptor gene may play an important role in the development of early onset drinking and the more severe alcoholism in Korean population.

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