Research Article

Osteoprotective Effect of Cordycepin on Estrogen Deficiency-Induced Osteoporosis In Vitro and In Vivo

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The purpose of this study was to verify the effect of cordycepin on ovariectomized osteopenic rats. Fifty Wistar female rats used were divided into 5 groups: (1) sham-operation rats (control), (2) ovariectomized (OVX) rats with osteopenia, and (3) OVX'd rats with osteopenia treated with cordycepin (5mg, 10mg, and 20mg) for 8 weeks. After the rats were treated orally with cordycepin, serum alkaline phosphatase (ALP), tartrate resistant acid phosphatase (TRAP), serum osteocalcin (OC), homocysteine (HCY), C-terminal crosslinked telopeptides of collagen type I (CTX) level, and oxidative stress were examined, respectively. The femoral neck was used for mechanical compression testing. At the same time, we further investigated the effect of cordycepin in vitro assay.

The beneficial effects of cordycepin on improvement of osteoporosis in rats were attributable mainly to decrease ALP activity, TRAP activity, and CTX level. At the same time, cordycepin also increases the OC level in ovariectomized osteopenic rats. The histological examination clearly showed that dietary cordycepin can prevent bone loss caused by estrogen deficiency. These experimental results suggest that complement cordycepin is protective after ovariectomized osteopenic in specific way.

1. Introduction

Osteoporosis is a major concern in public health care and the disease has severe consequences if untreated [1, 2]. It is characterized by low bone mineral density (BMD) and loss of the structural and biomechanical properties that are required to maintain bone homeostasis. Bone is a metabolically active tissue that undergoes remodeling throughout life, with roughly 5% remodeled at any time [3]. Over several weeks, a bone remodeling unit (BMU) will develop that incorporates several cell types, including osteoclasts, osteoblasts, and osteocytes. The loss of sclerostin and alterations in other secreted cytokines and chemotactic factors promote BMU’s formation. Despite current treatment options that include vitamin D, hormone, and bisphosphonates therapy, osteoporosis results in significant morbidity and mortality. Development of novel therapies is vital for therapy against osteolytic bone diseases.

The herbal kingdom is a wide field to search for natural effective osteoporosis protective agent that has no side effects. As potential alternative treatments for osteoporosis, the preventive and therapeutic effects of natural products derived from plants have been reported [4–6]. Cordyceps is a genus of the family Clavicipitaceae that has been used in traditional Oriental medicine for centuries. Recent studies have demonstrated that the bioactive components isolated from this genus have various pharmacological actions [7–9]. Among them, cordycepin, also known as 3-deoxyadenosine, has been shown to possess multiple pharmacological activities such as inhibition of tumour growth, modulation of the immune response, and suppression of reactive oxygen species [10]. In the former paper, we have reported that cordycepin can act as anti-inflammatory agent in magnesium silicate-induced inflammation in osteoporosis [11]. However, the role of cordycepin in estrogen deficiency-induced osteoporosis in ovariectomized rats has not been investigated. The aim of the
The present investigation was to discover cordycepin for effective osteoporosis treatment in vivo and in vitro.

2. Materials and Methods

2.1. Animals. Wistar rats (weighing 225 ± 25 g) were used in the study. This study was performed in accordance with the Guide for the Care and Use of Laboratory Animals. Care was taken to minimize discomfort, distress, and pain to the animals. The study was submitted to, and approved by, the Fourth Military Medical University institutional ethics committee.

2.2. Drugs. Cordycepin with 98% purification was obtained following the extraction and separation using a column chromatographic method [12].

2.3. Experimental Design. Fifty rats were randomly divided into five groups of animals, four ovariectomized (OVX) and another given a sham-operation (control). Then groups 1 (sham) and 2 (OVX) were treated orally with 10 mL of saline; group 3, group 4, and group 5 were treated orally with cordycepin (5 mg, 10 mg, and 20 mg) for 8 weeks, respectively. Cordycepin was dissolved in distilled water and administered orally twice daily using a feeding needle for 21 days.

Cordycepin content was determined using an Osteocalcin EIA kit (Xinyubio-Technology, Inc., China) as described in the manufacturer’s directions. Homocysteine (Hcy) was measured by use of an enzymatic fluorescence polarization immunoassay on an Axsym analyzer (Abbott, Wiesbaden, Germany). C-terminal crosslinked telopeptides of collagen type I (CTX) were quantified by ELISA (Sunbio, Inc., China).

2.4. Mechanical Testing. The mechanical strength of the femoral neck was measured by applying a vertical load to the femoral head using a Shimadzu EZ-1 pressure system. The fracture load was recorded at the peak force as Newton (N) at the point that the femoral neck fractured [13].

2.5. Histomorphometry of Osteoblast Surface. The tibia and the lumbar vertebrae were decalcified in formic acid, embedded in paraffin, and longitudinally sectioned. Histomorphometric analyses were made by tracing the section image onto a digitizing platen with the aid of a camera lucida attachment on the microscope and Osteomasure bone analysis software. To reveal osteoclasts, sections were stained for immunoreactivity to cathepsin K, an osteoclast marker [14]. Osteoblast perimeter was determined by scoring osteoblasts in direct contact with cancellous bone surfaces.

2.6. Plasma Enzyme Measurements. ALP and TRAP activity were determined by nitrophenol based method as described by Bessy et al. [15] and Godkar [16], respectively.

2.7. Plasma Proteins Measurements. Serum osteocalcin (OC) content was determined using an Osteocalcin EIA kit (Biomedical Technology, Inc., China) as described in the manufacturer’s directions. Homocysteine (Hcy) was measured by use of an enzymatic fluorescence polarization immunoassay on an Axsym analyzer (Abbott, Wiesbaden, Germany). C-terminal crosslinked telopeptides of collagen type I (CTX) were quantified by ELISA (Sunbio, Inc., China).

2.8. In Vitro Assay and Alkaline Phosphatase (ALP) and Tartrate Resistant Acid Phosphatase (TRAP). The murine mesenchymal stem cell line was purchased from the Beijing Lihaoy Co., Ltd., China, and grown in a DMEM medium supplemented with 10% fetal bovine serum (FBS), penicillin (100 U/mL), and streptomycin (100 μg/mL). All cultured cells were incubated in a humidified atmosphere at 37°C and at 5% CO₂. The study used cells with passages 5–10 (after purchase) for all experiments in cell lines. Cells (3 × 10⁵ cells/well) were incubated in a 96-well plate overnight and cotreated with different concentrations of cordycepin in the medium for 48 h. ALP activity was measured in total cell lysates after homogenization in a buffer containing 1 mmol/L Tris–HCl (pH 8.8), 0.5% Triton X-100, 10 mmol/L Mg₂⁺, and 5 mmol/L p-nitrophenolphosphate as substrates. The absorbance was read at 405 nm. The differentiated osteoblast cells from monocytes were measured by a TRAP activity assay and staining using the Acid-Phosphatase Kit (Shanghai Jinma Biological Technology, Inc., China).

2.9. Oxidative Stress Assay. In serum, glutathione peroxidase (GPx) activity, glutathione reductase (GR) activity, catalase (CAT) activity, Na⁺K⁺ATPase activity, and glutathione S transferase (GST) activity were quantified by ELISA (Sunbio, Inc., China).

2.10. Statistical Analysis. Data were expressed as the mean ± S.E.M. and the results analyzed by ANOVA followed by Dunnett’s t-test. A P value of <0.05 was considered significant.
3. Results and Discussion

Ovariectomized (OVX) animal models, in a variety of species, have been used to evaluate the mechanism of or to assess the effect of drugs on osteoporosis. Mechanical strength of bones is the most important parameter related to fracture risk. Therefore, this study first investigated the effect of cordycepin on mechanical strength in OVX osteopenic rats. The average maximum fracture loading to the femoral necks was lower in the OVX group compared with the sham group (Figure 1). Mechanical strength was significantly increased by treatment with cordycepin. It indicates that cordycepin had the positive effect on ovariectomized osteopenic rats.

Based on the results of mechanical strength, the trabecular number and thickness were studied. In the current study, we found that treatment of osteopenic OVX rats with cordycepin significantly increased maximal load compared to OVX animals. At the end of the 8-week treatment period, osteoblast surface in the lumbar vertebrae was not affected by OVX or any treatment (Figure 2(a)). A different cellular response was observed in the proximal tibial metaphysis. Cordycepin treatment caused a 3-fold increase in osteoblast surface compared with that in OVX rats (P < 0.05) (Figure 2(b)). It indicates that cordycepin administration improved bone strength mainly by increasing trabecular thickness. It agrees with the report of Ulrich [17].

Estrogen deficiency induces increased body weight in ovariectomized rats. The body weight gain pattern is shown in Figure 3. By the end of the fifth week, the ovariectomized rats gained significant weight compared to all other groups. From 5 weeks after the treatment was initiated, cordycepin-20 significantly increased body weights compared to sham group (Figure 3). The increase in rats' body weight in the cordycepin groups in the present study could be due to increased food intake as a result of lower leptin secretion though the impact is less severe compared to OVX rats [18].

Bone histomorphometry was also performed to determine the effects of cordycepin treatment on cancellous bone mass and levels of bone formation and resorption. A sample photomicrograph is presented in Figure 4. It is quite clear that trabecular bone loss is much higher in the vertebrae of rat with OVX (Figure 4(b)), whereas the vertebrae of cordycepin-fed OVX rat (Figure 4(a)) appear to be near normal (Figure 4(c)).

The current studies demonstrated that systemic treatment with cordycepin has a strong bone anabolic effect in OVX rats. The mechanism of it also needs to be studied in this paper.

ALP is a noncollagenous protein secreted by osteoblast, which is essential for bone mineralization [19]. Increased ALP level in serum has been observed in conditions such as rapid bone loss [20] and fracture risk [21, 22]. TRAP is secreted by osteoclasts during bone resorption, and serum TRAP activity correlates with resorptive activity in disorders of bone

![Figure 2: Effects of cordycepin on bone ObPm/BPm. Data are expressed as the mean ± SEM (n = 10 per group). *P < 0.05, significant difference from vehicle-treated OVX rats.](image1)

![Figure 3: Effects of cordycepin on body weight changes for the study (● sham group, ■ OVX group, □ cordycepin-5 group, ▲ cordycepin-20 group, and ○ cordycepin-10 group). Body weights were higher in the ovariectomized (OVX) animals than in sham-operated ones. Cordycepin-20 had similar body weights with the OVX animals in the former 5 weeks. From 5 weeks after the treatment was initiated, cordycepin-20 significantly increased body weights compared to sham group.](image2)

![Figure 4: (a) Bone histomorphometry showing trabecular bone loss in the vertebrae of OVX rat and (b) near normal appearance in the cordycepin-fed rat.](image3)
metabolism. In the present study significant increase in ALP and TRAP levels was observed in OVX control (Table 1). On the contrary, cordycepin significantly decreased ALP and TRAP levels ($P < 0.01$). Based on the above results, we further investigated the effect of cordycepin in vitro assay. Treatment of 50$\mu$g/mL of cordycepin showed significantly decreased ALP activity (Figure 5(a)) and TRAP activity (Figure 5(b)). It suggested that the potency of cordycepin is due to decrease ALP activity, TRAP activity in OVX rats.

Osteocalcin (OC), homocysteine (HCY), and collagen type I (CTX) are known as serum markers reflecting osteoblast activities including bone formation and turnover [23–25]. The effects of treatment with cordycepin on OC, HCY, and CTX level were shown in Table 2. Treatment of 50$\mu$g/mL of cordycepin increased OC level ($P < 0.01$). It also significantly decreased CTX level ($P < 0.01$). However,
and Na of oxidative stress. Activities of various antioxidant enzymes, that is, GPx, GR, CAT, and GST, as well as Na+K+ATPase, have been performed to estimate the amount of endogenous antioxidant enzymes. Therefore, the measurement of endogenous antioxidants, that is, GPx, GR, CAT, and GST, as well as Na+K+ATPase, has been used to estimate the amount of oxidative stress. Activities of various antioxidant enzymes and Na+K+ATPase of different groups have been listed in Table 3. The activity of endogenous antioxidant enzymes was decreased significantly (P < 0.01) in the OVX group, as compared to the sham group, whereas in the cordycepin group, cordycepin treatment showed a significant (P < 0.05–0.01) restoration in the level of various enzyme as compared with OVX group.

In conclusion, our findings first showed that oral administration of cordycepin can counteract the bone loss in an experimental model of established osteoporosis. These findings suggested that the mechanism of cordycepin is due to decrease ALP activity and TRAP activity both in vitro and in vivo. At the same time, oral administration of cordycepin can increase the OC level and decrease CTX and CTX level as well as restoring the oxidative stress in OVX animals. This suggests that cordycepin may be a good natural herbal medicine candidate for the treatment of osteoporosis.

Table 2: Effects of cordycepin on plasma proteins.

| Groups     | Serum OC (ng/mL) | Serum HCY (μmol/L) | Serum CTX (ng/mL) |
|------------|------------------|--------------------|-------------------|
| Sham      | 81.0 ± 5.0       | 7.7 ± 1.1          | 75.5 ± 4.2        |
| OVX       | 61.4 ± 5.1       | 9.2 ± 2.0          | 100.3 ± 5.0       |
| Cordycepin-20 | 84.1 ± 5.1**  | 8.0 ± 1.0          | 79.0 ± 8.1**      |
| Cordycepin-10 | 73.6 ± 4.0*     | 9.2 ± 3.3          | 85.1 ± 9.0        |
| Cordycepin-5  | 70.6 ± 3.2      | 8.0 ± 2.1          | 90.3 ± 4.4        |

Values are mean ± SEM. n = 10. *P < 0.05 versus OVX control; **P < 0.01 versus OVX control.

Table 3: Effect of cordycepin on the activity of various enzymes.

| Different groups | GPx     | GR             | GST      | CAT     | Na'+K' ATPase |
|------------------|---------|----------------|----------|---------|--------------|
| Sham             | 15.98 ± 1.23*** | 35.55 ± 2.51*** | 17.00 ± 1.22** | 7.11 ± 0.33* | 4.52 ± 0.32** |
| OVX              | 8.01 ± 0.42   | 20.88 ± 2.11   | 10.07 ± 1.11  | 4.22 ± 0.13 | 2.00 ± 0.13  |
| Cordycepin-20    | 13.16 ± 1.32** | 29.01 ± 2.21** | 13.66 ± 0.90* | 5.78 ± 0.20* | 4.11 ± 0.22* |
| Cordycepin-10    | 9.10 ± 1.02   | 26.00 ± 2.22   | 13.00 ± 0.91  | 4.70 ± 0.21 | 3.51 ± 0.23  |
| Cordycepin-5     | 8.16 ± 1.31   | 21.21 ± 2.20   | 10.10 ± 0.92  | 4.28 ± 0.33* | 2.10 ± 0.11  |

Values are shown as means ± SEM. *P < 0.05 versus OVX group, **P < 0.01 versus OVX group, and ***P < 0.001 versus OVX group.

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References

[1] O. Johnell and J. Kanis, “Epidemiology of osteoporotic fractures,” Osteoporosis International, vol. 16, no. 2, supplement, pp. S3–S7, 2005.
[2] S. Das and J. C. Crockett, “Osteoporosis—a current view of pharmacological prevention and treatment,” Drug Design, Development and Therapy, vol. 7, pp. 435–448, 2013.
[3] T. J. Martin, N. A. Sims, and K. W. Ng, “Regulatory pathways revealing new approaches to the development of anabolic drugs for osteoporosis,” Osteoporosis International, vol. 19, no. 8, pp. 1125–1138, 2008.
[4] S.-J. Suh, W.-S. Yun, K.-S. Kim et al., “Stimulative effects of Ulmus davidiana Planch (Ulmaceae) on osteoblastic MC3T3-E1 cells,” Journal of Ethnopharmacology, vol. 109, no. 3, pp. 480–485, 2007.
[5] K. W. Kim, S. J. Suh, T. K. Lee et al., “Effect of safflower seeds supplementation on stimulation of the proliferation, differentiation and mineralization of osteoblastic MC3T3-E1 cells,” Journal of Ethnopharmacology, vol. 115, no. 1, pp. 42–49, 2008.
[6] W. Caichompoop, Q.-Y. Zhang, T.-T. Hou, H.-J. Gao, L.-P. Qin, and X.-J. Zhou, “Optimization of extraction and purification of active fractions from Schisandra chinensis (Turcz.) and its osteoblastic proliferation stimulating activity,” Phytotherapy Research, vol. 23, no. 2, pp. 289–292, 2009.
[7] K. Yue, M. Ye, Z. Zhou, W. Sun, and X. Lin, “The genus Cordyceps: a chemical and pharmacological review,” Journal of Pharmacy and Pharmacology, vol. 65, no. 4, pp. 474–493, 2013.

Conflict of Interests

The authors declare that there is no conflict of interests.
[8] R. R. M. Paterson, “Cordyceps—a traditional Chinese medicine and another fungal therapeutic biofactory?” *Phytochemistry*, vol. 69, no. 7, pp. 1469–1495, 2008.
[9] X. Zhou, Z. Gong, Y. Su, J. Lin, and K. Tang, “Cordyceps fungi: natural products, pharmacological functions and developmental products,” *Journal of Pharmacy and Pharmacology*, vol. 61, no. 3, pp. 279–291, 2009.
[10] S. Takahashi, M. Tamai, S. Nakajima et al., “Blockade of adipocyte differentiation by cordycepin,” *British Journal of Pharmacology*, vol. 167, no. 3, pp. 561–575, 2012.
[11] D.-W. Zhang, Z.-L. Wang, W. Qi, W. Lei, and G.-Y. Zhao, “Cordycepin (3′-deoxyadenosine) down-regulates the proinflammatory cytokines in inflammation-induced osteoporosis model,” *Inflammation*, vol. 37, no. 4, pp. 1044–1049, 2014.
[12] H. Ni, X.-H. Zhou, H.-H. Li, and W.-F. Huang, “Column chromatographic extraction and preparation of cordycepin from *Cordyceps militaris* waster medium,” *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, vol. 877, no. 22, pp. 2135–2141, 2009.
[13] C. M. Bagi, P. Ammann, R. Rizzoli, and S. C. Miller, “Effect of estrogen deficiency on cancellous and cortical bone structure and strength of the femoral neck in rats,” *Calcified Tissue International*, vol. 61, no. 4, pp. 336–344, 1997.
[14] L. Xia, J. Kilb, H. Wex et al., “Localization of rat cathepsin K in osteoclasts and resorption pits: inhibition of bone resorption and cathepsin K-activity by peptidyl vinyl sulfones,” *Biological Chemistry*, vol. 380, no. 6, pp. 679–687, 1999.
[15] O. A. Bessy, O. H. Lowry, and M. J. Brock, “A method for the rapid determination of alkaline phosphatase with five cubic millimetres of serum,” *The Journal of Biological Chemistry*, vol. 164, pp. 321–329, 1946.
[16] P. Godkar, “Enzymes,” in *Textbook of Medical Laboratory Techniques*, pp. 149–167, Bhalani Publishing House, Mumbai, India, 1994.
[17] D. Ulrich, B. van Rietbergen, A. Laib, and P. Ruegsegger, “The ability of three-dimensional structural indices to reflect mechanical aspects of trabecular bone,” *Bone*, vol. 25, no. 1, pp. 55–60, 1999.
[18] R. Torto, S. Boghossian, M. G. Dube, P. S. Kalra, and S. P. Kalra, “Central leptin gene therapy blocks ovariectomy-induced adiposity,” *Obesity*, vol. 14, no. 8, pp. 1312–1319, 2006.
[19] L. M. Havill, L. G. Hale, D. E. Newman, S. M. Witte, and M. C. Mahaney, “Bone ALP and OC reference standards in adult baboons (*Papio hamadryas*) by sex and age,” *Journal of Medical Primatology*, vol. 35, no. 2, pp. 97–105, 2006.
[20] P. D. Ross and W. Knowlton, “Rapid bone loss is associated with increased levels of biochemical markers,” *Journal of Bone and Mineral Research*, vol. 13, no. 2, pp. 297–302, 1998.
[21] G. Patrick, E. Sornay-Rendu, B. Claustrat, and P. D. Delmas, “Biochemical markers of bone turnover, endogenous hormones and the risk of fractures in postmenopausal women: the OFELY study,” *Journal of Bone and Mineral Research*, vol. 15, no. 8, pp. 1526–1536, 2000.
[22] P. D. Ross, B. C. Kress, R. E. Parson, R. D. Wansich, K. A. Armour, and I. A. Mizrahi, “Serum bone alkaline phosphatase and calcaneus bone density predict fractures: a prospective study,” *Osteoporosis International*, vol. 11, no. 1, pp. 76–82, 2000.
[23] D. Polak-Jonkisz and D. Zwolinska, “Osteocalcin as a biochemical marker of bone turnover,” *Nephrology*, vol. 4, no. 5–6, pp. 339–346, 1998.
[24] J.-M. Koh, Y.-S. Lee, Y. S. Kim et al., “Homocysteine enhances bone resorption by stimulation of osteoclast formation and activity through increased intracellular ROS generation,” *Journal of Bone and Mineral Research*, vol. 21, no. 7, pp. 1003–1011, 2006.
[25] M. Herrmann, N. Umaniskaya, B. Wildemann et al., “Stimulation of osteoblast activity by homocysteine,” *Journal of Cellular and Molecular Medicine*, vol. 12, no. 4, pp. 1205–1210, 2008.
[26] J. T. Coyle and P. O. Puttfarcken, “Oxidative stress, glutamate, and neurodegenerative disorders,” *Science*, vol. 262, no. 5134, pp. 689–695, 1993.
[27] J. S. Bains and C. A. Shaw, “Neurodegenerative disorders in humans: the role of glutathione in oxidative stress-mediated neuronal death,” *Brain Research Reviews*, vol. 25, no. 3, pp. 335–358, 1997.