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Transition from Dendritic to Cell-like Crystalline Structures in Drying Droplets of Fetal Bovine Serum under the Influence of Temperature

Marina Efstratiou, John R. E. Christy,* Daniel Bonn, and Khellil Sefiane

ABSTRACT: The desiccation of biofluid droplets leads to the formation of complex deposits which are morphologically affected by the environmental conditions, such as temperature. In this work, we examine the effect of substrate temperatures between 20 and 40 °C on the desiccation deposits of fetal bovine serum (FBS) droplets. The final dried deposits consist of different zones: a peripheral protein ring, a zone of protein structures, a protein gel, and a central crystalline zone. We focus on the crystalline zone showing that its morphological and topographical characteristics vary with substrate temperature. The area of the crystalline zone is found to shrink with increasing substrate temperature. Additionally, the morphology of the crystalline structures changes from dendritic at 20 °C to cell-like for substrate temperatures between 25 and 40 °C. Calculation of the thermal and solutal Benard–Marangoni numbers shows that while thermal effects are negligible when drying takes place at 20 °C, for higher substrate temperatures (25–40 °C), both thermal and solutal convective effects manifest within the drying drops. Thermal effects dominate earlier in the evaporation process leading, we believe, to the development of instabilities and, in turn, to the formation of convective cells in the drying drops. Solutal effects, on the other hand, are dominant toward the end of drying, maintaining circulation within the cells and leading to crystallization of salts in the formed cells. The cell-like structures are considered to form because of the interplay between thermal and solutal convection during drying. Dendritic growth is associated with a thicker fluid layer in the crystalline zone compared to cell-like growth with thinner layers. For cell-like structures, we show that the number of cells increases and the area occupied by each cell decreases with temperature. The average distance between cells decreases linearly with substrate temperature.

INTRODUCTION

Deegan’s paper in 1997, attributing the “coffee ring” effect to a capillary outward flow in droplets of suspensions or soluble solids,1 triggered wide interest in studying sessile drop evaporation of both pure and complex fluids as well as investigation of the patterns arising from drop dryout.12−13 For colloidal suspensions, because of continuity, the outward flow carries the particles to the contact line, causing the formation of a ring deposit. During the evaporation of colloidal drops, multiple hydrodynamic and physicochemical processes take place, which result in the formation of complex patterns.14 These processes include the spreading and adhesion of the drops to the substrate, contact line pinning, induced flows and component redistribution, crystallization, stress accumulation, and release.15,16 As a result of these processes, various morphologies have been reported within the final desiccation patterns, including rings, cracks, and crystalline regions.11,17−20

Biological fluids are considered to be complex colloidal systems, mainly composed of proteins, electrolytes, and water.14,18 During the desiccation of colloidal droplets, water evaporation causes the generation of surface tension gradients, which arise due to concentration gradients within the drying drops. In the case of a heated substrate, significant temperature gradients also arise because of the temperature difference between the warmer base of the drops, which is close to the substrate temperature, and the cooler apex of the drops, close to the ambient temperature. The development of concentration or temperature gradients causes convection and mixing within the droplet, leading to redistribution of the components. Because of the higher evaporation rate at the periphery of the drying drops, the solute concentration in that region increases, leading to supersaturation and preferential protein precipitation near the contact line, and hence, to the formation of an outer ring, mostly composed of proteins. Low molecular weight components, on the other hand, are observed to precipitate in the central region of the droplet. The evolution of the pattern over time and the final pattern formation, depend on the fluid composition.21

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Understanding of the mechanisms involved during desiccation of biological fluid droplets, as well as of the emerging patterns, is of utmost importance for their application to medical diagnostics and forensic analysis. Studies have reported that changes in the composition of biological fluids, caused by disease, alter the final desiccation patterns. A multitude of studies have compared the final patterns arising from droplet evaporation of physiological and pathological biological fluids such as blood and blood serum. Additionally, Sobac and Brutin have described the different stages of blood droplet desiccation.

Because of the safety issues and difficulties involved in the acquisition of physiological fluids, many researchers have focused on the investigation of similar but simpler model systems, such as inorganic or organic colloids and colloidal solutions with salt admixtures. Various structures are observed when different types of ions are added to bovine serum albumin (BSA) solutions due to the interactions taking place within the solutions, leading to droplet spreading, gelation, crystallization, and crack formation. Pathak et al. have investigated how the addition of different salts (KCl and MgCl2) in aqueous BSA droplets affects the final desiccation patterns. Tarasevich and Pravoslavnova have developed a model for the evaporation of biological fluid drops, taking into account both diffusion and evaporative capillary flows.

The desiccation of droplets consisting of biological fluids and the morphology of the final desiccation deposits depend on multiple factors including the type and concentration of macromolecules and electrolytes within the solution, the substrate properties such as wettability, and the ambient conditions under which evaporation takes place. Carreón et al. studied the effects of substrate temperature on the desiccation patterns of protein mixtures and protein—salt mixtures to find the most suitable temperature for efficient diagnosis.

The aforementioned studies offer a basis for the understanding of the impact of substrate temperature on the drying of protein—protein and protein—salt mixtures. Nevertheless, blood serum is a very complex mixture which consists of various types of macromolecules, electrolytes, antibodies, antigens, and hormones. Because of the complexity of biological fluids such as blood and serum, further research is required to gain a better understanding of the mechanisms involved in the desiccation of such systems. Additionally, the pH of the mixtures, which has a significant effect on protein conformation and aggregation by altering the electrostatic repulsive and attractive van der Waals forces, was not controlled in these studies. By affecting the interactions between the components, the pH can significantly influence the magnitude of the intermolecular forces and can therefore alter the final desiccation patterns of the dried drops.

In this work, we use fetal bovine serum (FBS) droplets to examine the addition of different salts (KCl and MgCl2) in aqueous BSA droplets and to study the effect of substrate temperature on the desiccation patterns arising from droplet evaporation of biological fluids such as blood and blood serum.

### Table 1. Type and Concentration of Proteins in the FBS Used in the Experiments

| Type of Protein | Typical Concentration Range | Concentration in Our Sample |
|-----------------|-----------------------------|-----------------------------|
| BSA             | 17–35 mg/mL                 | 23 mg/mL                    |
| α-globulin      | 7–20 mg/mL                  | 16 mg/mL                    |
| β-globulin      | 3–9 mg/mL                   | 3.6 mg/mL                   |
| γ-globulin      | 10–200 μg/mL                | 23.73 μg/mL                 |
| Hemoglobin      | 0.01–0.30 mg/mL             | 0.1401 mg/mL                |

Glass microscope slides (MS/1 Scientific Glass Laboratories Ltd., UK) were placed in an ultrasonic bath with deionized water for 15 min and then rinsed with ethanol (ethanol, 99%+, Absolute, Fisher Scientific, UK) and dried by using an air gun. Sessile drops of FBS (1.2 μL) were gently placed on the glass slides and left to dry at different substrate temperatures. The initial contact angle between the liquid FBS droplets and the substrates was 35 ± 5°. The substrate temperature was controlled with the use of a heater mat (SRFRA-4/10-230V, Omega Engineering Ltd., UK), connected to a PID controller that enabled the temperature adjustments, placed underneath the glass surfaces, and a thermocouple mounted on the slide surface (Figure S1 in the Supporting Information). Both the ambient temperature and the relative humidity (RH) levels were monitored during the experimental procedure via the use of a temperature—humidity meter (HH311, Omega Engineering Ltd., UK). RH levels were 45 ± 5% during the experiments, and the ambient temperature was 20 ± 1 °C.

The temporal evolution of droplet volume, diameter, and contact angle were monitored with the use of a goniometer (DSA-305 Drop Shape Analyzer, KRÜSS, Germany) which enabled the investigation of the side-profile evolution of the drops. Images were captured up to completion of the desiccation process under different magnifications (2×, 5×, 10×) for the investigation of the final deposition patterns. Top view images of the droplets were also captured during desiccation using an optical microscope (Euromex, Netherlands) connected to a CMOS camera (MAKO G-507 Allied Vision Technologies, UK) under 2× magnification. All the deposits were also examined 24 h after the completion of the experiments.

3D topographical studies were also conducted on the desiccated deposits. A confocal laser scanning microscope (VK-X1000, Keyence, Netherlands) was utilized to obtain 3D topographs, in combination with the Keyence MultiFile Analyzer software for height measurements in different zones of the dried deposits. Topographical data were acquired under 20× magnification. Because of the view of the entire desiccation pattern was challenging under such high magnification, multiple regions of each drop were examined separately, and then stitched together in Keyence MultiFile Analyzer, to provide the topography of the final desiccation pattern. The experimental procedure was similar as described above by using the goniometer. Topographical investigation of the deposits allowed measurements of the average thickness (height) and radius of each region of the desiccation patterns. Based on the measured radius, the area of each deposit was estimated. This allowed the calculation of the final deposit volume and the volume of each zone within the dried deposits, based on measurements of thickness and radius.

Each set of experiments was repeated at least three times for each substrate temperature. The final deposits showed good reproducibility regardless of the final morphology for each of the temperatures in all of these methods.

### EXPERIMENTAL SECTION

Fetal bovine serum (FBS South American A3160802, Thermofisher Scientific, UK) was received frozen in dry ice and used after defrosting and gentle mixing at room temperature. It should be noted that no vortex mixers or sonicators were used to avoid protein denaturation. FBS is a complex solution consisting of multiple proteins (Table 1), ions, and hormones. The ionic strength and pH of the solution are 0.14 M and 7.4, respectively.

**Table 1. Type and Concentration of Proteins in the FBS Used in the Experiments**

| Type of Protein | Typical Concentration Range | Concentration in Our Sample |
|-----------------|-----------------------------|-----------------------------|
| BSA             | 17–35 mg/mL                 | 23 mg/mL                    |
| α-globulin      | 7–20 mg/mL                  | 16 mg/mL                    |
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| γ-globulin      | 10–200 μg/mL                | 23.73 μg/mL                 |
| Hemoglobin      | 0.01–0.30 mg/mL             | 0.1401 mg/mL                |
RESULTS AND DISCUSSION

Four desiccation stages were identified during drying for all the examined substrate temperatures. These are pregelation, gelation, crystallization and crack formation. During pregelation, the drying is dominated by water evaporation at the contact line where the evaporation rate is higher. Because of the higher evaporation rate at the periphery, a radially outward capillary flow develops, carrying fluid toward the contact line. This leads to the aggregation and precipitation of proteins at the contact line and induces pinning of the drop to the substrate. Protein adsorption follows a complex series of adsorption−displacement steps in which proteins of lower molecular weight adsorb to the substrate first and are then displaced by proteins of higher molecular weights. This phenomenon is called the “Vroman effect”. Because of pinning, evaporation proceeds in a constant contact radius (CCR) mode, during which the contact angle decreases over time. The droplet volume decreases linearly over time, indicating a constant evaporation rate, until the onset of gelation (continuous lines), after which a small decrease of the evaporation rate is observed, as shown in Figure 1. The dashed lines in Figure 1 show the extrapolation of the initial evaporation kinetics according to the evaporation rate at the early stage of pregelation. The average times for the onset of gelation are ~250, ~163, ~129, ~66, and ~38 s for droplets evaporating on glass slides with temperature 20, 25, 30, 35, and 40 °C, respectively. It should be noted that the volume data points appear to asymptote to a constant value during crystallization and cracking. This is due to limitations of the experimental apparatus to make accurate measurements when the remaining volume is very small. The final volume of the deposits is in the order of tens of nanoliters.

Effect of Temperature on the Morphology of the Final Deposits. It was shown in a previous work that for aqueous drops of 7% BSA−0.9% NaCl solution, four distinct zones are observed in the desiccation deposits after evaporation. Moving from the edge of the drop toward the center, these zones are (1) a zone of homogeneous protein (peripheral ring), (2) a zone of protein structures, (3) the protein gel, and (4) the crystalline zone within the protein gel. The proteins exist at different conditions within each zone, forming materials of different properties: a glassy (high volume fraction) protein ring on the periphery and a protein gel (lower volume fraction) in the interior of the ring. In the central area of the desiccated drops, the volume fraction of proteins is lower, but the ionic strength increases. The same zones were observed in our experiments, with the zones of protein structures and gel being more evident between 25 and 40 °C. The different zones for the desiccated deposits are shown in Figure 2.

Gel and Central Crystalline Zone. Increasing the substrate temperature causes the central crystalline zone in the desiccation patterns to shrink. This is shown both on the microscopic images acquired after evaporation and on the 3D topographical data acquired via confocal laser scanning microscopy (Figure S2). Image analysis showed that variation of the ratio of the crystalline area (A_c) to the total area of the desiccation deposit (A_total) with temperature can be approximated by a second-order
polynomial curve (Figure 3). The reason for this is not clear yet. Because of the shrinking of the crystalline zone, the width of the gel between the glassy protein ring and the crystalline zone increases with temperature. The increase in the width of the protein gel is accompanied by a decreasing height with increasing temperature. The gel is more profound at the highest examined temperatures of 35 and 40 °C (Figure 4 and Figure S3). The glassy peripheral protein ring appears to be the thickest feature of the desiccation patterns for the range of temperatures examined in this work. Topographical investigation has shown that the height of the deposit decreases from the peripheral protein ring (red) toward the region of protein aggregates and the gel (yellow and green) and the central crystalline zone which appears to be the thinnest feature as shown in Figure 4.

The temperature increase appears to have an effect on the morphology of the final crystalline structures forming in the central region of the dried deposit, as shown in Figure 5. The morphology of the crystals is probably related to the thickness of the crystalline zone, as will be discussed further later. Although for droplets evaporating at 20 °C the final crystalline deposit consists of dendritic structures, at higher temperatures, the central region is composed of cell-like structures, which become finer with increasing temperature from 25 to 40 °C. The time for the onset of crystal nucleation and the duration of crystallization differ for each of the examined temperatures. Dendrite nucleation is initiated at approximately 60% of the droplet lifetime, whereas cell-like structures start to form at ~78% of the lifetime for droplets drying at 25 °C (Figure S4). For cell-like structures, the time at which crystallization commences, as well as its duration, decreases with increasing temperature because the evaporation rate varies with temperature.

In an attempt to probe the effect of temperature on the formation of crystalline structures, we proceed to the investigation of the central crystalline zone. Investigation of the desiccation deposits via confocal laser scanning microscopy enables the determination of the average height (thickness) and area of each zone within the deposits. Focusing on the central crystalline zone, we measure the final average thickness of this region with substrate temperature (Figure 6). The thickness of the central crystalline zone varies significantly in the case of dendrites, formed on substrates at 20 °C, compared to the case of cell-like structures, formed on substrates of higher temperatures (25–40 °C). The average final thickness in the case of dendritic formation is approximately 9.5 μm, whereas for cell-like structures it varies between ~4.9 and ~6.2 μm, depending on the substrate temperature. The final thickness decreases from approximately 6.2 μm at 25 °C to ~5.4 and ~4.9 μm at 30 and 35 °C, respectively, and it increases from 35 to 40 °C (~6.2 μm at 40 °C). This indicates that the thickness of the central crystalline zone is significantly thicker when drying takes place on a substrate with temperature of 20 °C compared to higher temperatures, i.e., 25–40 °C. This finding suggests that the lower evaporation rate at lower substrate temperatures leads to the formation of a thicker film in the central region of the dried deposit, manifesting dendritic structures. The thinner film forming in the central region at higher temperatures, on the other hand, consists of cell-like crystalline structures. Therefore, the type of crystalline structures occurring could be related to thickness of the central region.

Hydrodynamic and Intermolecular Forces Acting within the Droplets at the Onset of Evaporation. In an attempt to understand the phenomena governing the pattern formation, we have performed an approximate analysis of the hydrodynamic (capillary, drag) and intermolecular forces acting in the droplets at the onset of drying (see the Supporting Information). To do this, we considered the spherical-like geometry of the protein macromolecules in fetal bovine serum.41–43 The approach for the calculation of hydrodynamic and intermolecular forces is similar to that employed in other studies,42, 44, and 45, treating proteins as spherical macromolecules. The equations used for the estimation of electrostatic and van der Waals forces on macromolecules may also be found in other works.46 and 47. This analysis suggests that the hydrodynamic fluid forces dominate initially, while macromolecules are in solution, but that once deposition starts, at the molecular level intermolecular attractive protein–protein and protein–substrate forces dominate over electrostatic forces, which may partially explain why the salts tend to diffuse out from the protein deposited at the contact line. Capillary forces acting on protein macromolecules (O(10−9 N)) are found to be orders of magnitude higher compared to drag forces or intermolecular forces. The attractive protein interactions between proteins of the same type are in the order of O(10−14 N) at 25 °C. The order of magnitude of these forces varies from O(10−14 N) to O(10−15 N), depending on the type of biomolecule, when the substrate temperature is 40 °C. Protein–substrate interactions may increase from O(10−15 N) to O(10−14 N) or drop from O(10−14 N) to O(10−15 N) for different types of proteins, when the substrate temperature changes from 25 to 40 °C.

If we analyze the hydrodynamic fluid forces in the central region following deposition of proteins at the outer edge, these will be influenced by the temperature difference between the heated substrate and the liquid vapor surface at which evaporation takes place. We suspect that the pattern formation in the central crystalline region might be related to a thin film phenomenon. After gelation, the central region of the drying drop does not have a hemispherical shape anymore, but it resembles a thin film instead due to water evaporation. When the substrate is close to room temperature, no significant temperature gradients are expected within the film, and we do not anticipate any significant thermally driven surface temperature gradients. With a lack of driving force we expect the salt
concentration in the central region to increase until the level of supersaturation results in nucleation of a crystal, with subsequent crystallization extending from the one nucleation site.

In the case of the heated substrate, we expect significant temperature gradients to develop initially between the base of the drop and the apex of the drop. The formation of cell-like structures for temperatures between 25 and 40 °C may be attributed to thermal Bénard–Marangoni convection, leading to the development of instabilities and resulting in the formation of polygonal (hexagonal-like) convective cells within the drying liquid film. To probe whether the cell-like formation could be

Figure 4. Left: top view images of the deposits illustrating the horizontal line along which the height profile is shown. Right: height profile of the deposits occurring after droplet drying at (A) 20, (B) 25, (C) 30, (D) 35, and (E) 40 °C. The gray arrows indicate the increase in the width of the gel region for droplets evaporating at higher temperatures (35 and 40 °C).
due to the development of instabilities, we proceed to investigate the convective effects for the substrate temperatures examined in this study.

Bénard–Marangoni Convection. For small droplets deposited on substrates with \( h \ll R \), the gravitational effects may be neglected because of the small Bond number \( \left( Bo = \frac{gR^2}{\eta} \sim 0.3 \right) \). In this case, convection is driven by surface tension effects (Bénard–Marangoni) rather than buoyancy effects (Rayleigh–Bénard). Bénard–Marangoni convection develops due to a surface tension gradient on the free surface.

This surface tension gradient may result from a temperature gradient (thermal Bénard–Marangoni) and/or a concentration gradient (solutal Bénard–Marangoni). The effect of thermal and solutal Bénard–Marangoni instabilities may be evaluated by calculation of the dimensionless thermal \((Ma_T)\) and solutal \((Ma_S)\) Bénard–Marangoni numbers, respectively.

The thermal Bénard–Marangoni number is defined as

\[
Ma_T = \frac{\eta \alpha}{\Delta T h} \frac{\partial \gamma}{\partial T}
\]

where \( \frac{\partial \gamma}{\partial T} \) is the change of the surface tension with temperature, \( \Delta T \) is the temperature difference at the air/liquid interface, \( h \) is the film thickness (height), \( \eta \) is the dynamic viscosity of the fluid, and \( \alpha \) is the thermal diffusivity. Thermal Bénard–Marangoni instabilities develop because of a temperature gradient on the free surface, giving rise to a surface tension gradient.

The solutal Bénard–Marangoni number, on the other hand, is given by

\[
Ma_S = \frac{\eta D}{\alpha C} \frac{\partial \gamma}{\partial C}
\]

where \( \frac{\partial \gamma}{\partial C} \) is the change of the surface tension with concentration, \( \Delta C \) is the concentration difference at the air/liquid interface, and \( D \) is the diffusion coefficient. Solutal Bénard–Marangoni instabilities may develop from a concentration gradient on the free surface. The solvent evaporation from the free surface can lead to the development of a concentration gradient across the air/liquid interface, between the base and the apex.

The surface tension gradients arising at the air/liquid interface from thermal or solutal effects give rise to the development of internal flows in the liquid film. The strength of these flows and the velocities developing in the fluid are related to the magnitude of Bénard–Marangoni numbers; hence, we estimate the maximum values that these numbers can take in our system.
for the temperatures studied. For the estimation of the thermal \( (\text{Ma}_T) \) Bénard–Marangoni numbers we assume the temperature difference at the free surface with the base having the temperature of the heated substrate, whereas the apex is at the ambient temperature (20 °C). The critical thermal Marangoni number \( \text{Ma}_T \) for the formation of convective thermal Bénard–Marangoni instabilities has been estimated to be ~80 for water films.

The values of parameters for the calculations of \( \text{Ma}_S \) and \( \text{Ma}_T \) are given in the Supporting Information (Table S5). Because of the preferential deposition of proteins at the contact line, the volume fraction of proteins is higher at the droplet edge, whereas the preferential deposition of proteins at the contact line, the MaT number

\[ \text{Ma}_T \]

is higher than \( \text{Ma}_S \) at the base. Thus, the temperature gradient leads to thermal Bénard–Marangoni convection, creating toroidal vortices within each cell.

Although thermal Bénard–Marangoni convection appears to be responsible for the setup of the convective cells early in the evaporation process defining the number and the shape of the cells, it is possible that as drying proceeds, the maintenance of circulation in the cells is enhanced by solutal Bénard–Marangoni convection. At the latter stages of evaporation, convective cells are fully formed in the drying film. We believe that the toroidal vortices formed within each cell by thermal Bénard–Marangoni effects result in more concentrated solution flowing upward to the air/liquid interface of each cell. As drying proceeds, the fluid is carried upward at the edge of the cell to the air/liquid interface, reaching supersaturation at the surface and leading to crystal nucleation in the center of the base of each cell. Once nucleation commences, the fluid moving upward the edge of the cell is saturated rather than supersaturated. This internal circulation, initiated by thermal effects, is sustained by solutal effects, since surface tensions are higher for both lower temperatures and higher salt concentrations, maintaining the toroidal flow patterns within each cell. For the calculations of the solutal (\( \text{Ma}_S \)) Bénard–Marangoni numbers, we consider that

\[ \Delta C = C_{\text{sat}} - C_{\text{sat},r} \]

where \( C_{\text{sat}} \) is the supersaturation concentration (at the apex) and \( C_{\text{sat},r} \) is the saturation concentration (at the base). The estimated solutal Bénard–Marangoni numbers at the end of gelation just prior to the onset of crystallization are approximately 6.2 \( \times 10^4 \), 6.8 \( \times 10^4 \), 7.1 \( \times 10^4 \), and 39.9 \( \times 10^4 \) for 25, 30, 35, and 40 °C, respectively. In all cases, we believe the existence of the cells to be related to surface tension effects, with thermal Bénard–Marangoni convection being responsible for the formation of solutal Bénard–Marangoni convection being responsible for the maintenance of cells during crystallization.

Figure 8 shows a schematic diagram of the convective cells formed in the thin films, giving rise to the cell-like structures. The area and distribution of cell-like structures were investigated for temperatures between 25 and 40 °C (Figure 9) via ImageJ (v1.53c) software. The number and area of the crystalline structures vary with temperature. It has been found that elevated temperatures give rise to a higher number of cell structures in the crystalline area. Structures with areas of 0.005 mm\(^2\) have been observed in the samples for all the examined temperatures. Nevertheless, at 25 and 30 °C, a number of larger structures exist in the central region with a maximum area of 0.025 and 0.020 mm\(^2\), respectively, leading to the conclusion that the base area of each cell decreases as the substrate temperature increases. This would be in agreement with stronger Bénard–Marangoni convection caused by the higher temperature gradient between the apex and the base of the drying drops, leading to a higher number of smaller convective cells. As a result of the cell formation, the salt diffusion within the

Figure 7. Estimation of thermal (\( \text{Ma}_T \)) Bénard–Marangoni numbers at the end of gelation for different substrate temperatures.

\[ \text{Ma}_T \]

\[ \text{Ma}_T \]

\[ \text{Ma}_T \]

\[ \text{Ma}_T \]

\[ \text{Ma}_T \]

\[ \text{Ma}_T \]

\[ \text{Ma}_T \]

\[ \text{Ma}_T \]
between cells (substrate temperatures. A schematic of the average distance consistent with a more intense thermal convection at higher decrease in the size and average distance between cells is inversely proportional to the substrate temperature. This substrate temperature, suggesting that the average cell distance average repetition distance between cells (substrate temperature, which grow and propagate within the central crystalline zone with no spatial restrictions. As a result, crystallization extends from the formed nucleation site, covering the crystalline zone. However, in the case of cell-like structures forming at higher substrate temperatures, when the thermal Bénard–Marangoni convection is stronger, the number of convective cells forming in the central crystalline zone increases. In this case, fluid is trapped in each convective cell and each cell does not share fluid with neighboring cells. At the latter evaporation stages, when solutal Bénard–Marangoni convection dominates, each convective cell may act as an individual nucleation site. This would mean that the number of crystal nuclei is related to the number of convective cells forming.

For droplets evaporating at 25 and 30 °C, crystals nucleate as circular individual spots in the central region of the drying drop, merging a few seconds after nucleation and propagating toward the periphery of the drop (Figures S9 and S10). For the highest examined temperatures of 35 and 40 °C, on the other hand, an arc-like crystal growth characterizes the onset of crystallization (Figures S11 and S12).

**CONCLUSIONS**

The drying of biological fluid droplets gives rise to distinct desiccation patterns. The conditions under which drying takes place, such as temperature and relative humidity, affect the drying process and can lead to distinct variations in the dried deposits. Therefore, it is important to understand how these conditions affect the evaporative dynamics and the final deposit formation. In this work, we focus on the effect of substrate temperature on the crystalline pattern formation in dried deposits of FBS droplets. We examine substrate temperatures between 20 and 40 °C, as these are the most relevant for diagnostic applications. We estimate the forces acting at the final deposit finding that hydrodynamic fluid forces dominate.

Examining the desiccation deposits and focusing on the central crystalline zone, we find that the ratio of the crystalline
area over the entire deposit area ($A_c/A_{total}$) decreases with substrate temperature. A transition in the morphology of the crystals is observed, from dendrites at 20 °C to cell-like structures for substrate temperatures between 25 and 40 °C. We believe that this transition occurs because of the interplay between thermal and solutal Bénard–Marangoni convection in the drying droplets, leading to the development of convective instabilities. In an attempt to explain the change in the morphology of the crystalline structures, we calculate the thermal and solutal Bénard–Marangoni numbers at the end of the gelation stage. Thermal effects are negligible throughout drying in the case of dendritic crystalline structures forming on substrates of 20 °C. However, in the case of cell-like structures, both thermal and solutal convective effects act within the drying drops. Thermal convective effects dominate during the early drying stages, leading to the development of instabilities and the

**Figure 9.** Average number and area distribution of cell-like structures observed in the crystalline area for droplets evaporating at (A) 25, (B) 30, (C) 35, and (D) 40 °C. (E) Average distance between cells ($\lambda$) with substrate temperature based on hexagonal geometry.
formation of convective cells. As evaporation proceeds, the solutal effects dominate, maintaining the circulation and leading to crystallization within the formed cells. These findings support our hypothesis that the formation of cell-like structures results from the interplay between thermal and solutal effects in the drying droplets. In the case of dendrites, the crystalline zone is found to be significantly thicker compared to the case of cell-like structures. For cell-like structures, image analysis has shown that the number of cells increases with increasing substrate temperature, whereas the area occupied by each cell-like structure decreases. The average distance between cells (λ) has been calculated and was found to be inversely proportional to the substrate temperature. Temperature also affects the nucleation and growing manner of the crystals from one or two circular nuclei at 20 °C, to multiple circular nuclei that grow into crystals and merge at 25 and 30 °C, to an arc-like growth at higher temperatures. The findings of this work could contribute to the understanding of the impact of environmental conditions on the desiccation patterns of biological fluid droplets as a tool for disease diagnosis. Investigation of FBS deposits at a larger range of temperatures may be required for a better understanding of the mechanisms affecting the final desiccation patterns; however, this was beyond the scope of this study.

■ ASSOCIATED CONTENT

Supporting Information
The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.langmuir.2c00019.
Additional experimental information, calculations and values of parameters used for calculations, and figures (PDF)

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Notes
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■ ABBREVIATIONS

BSA, bovine serum albumin; CCR, constant contact radius; FBS, fetal bovine serum

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