Identification of microRNAs involved in gefitinib resistance of non-small-cell lung cancer through the insulin-like growth factor receptor 1 signaling pathway

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Abstract. Multiple clinical and experimental studies have suggested that epidermal growth factor receptor tyrosine kinase inhibitors (EGFR-TKIs) may be effective at treating advanced non-small cell lung cancer (NSCLC), however, the molecular basis of primary resistance to EGFR-TKIs in NSCLC remains unclear. In the current study, the insulin-like growth factor 1 receptor (IGF-1R) gene in the gefitinib-resistant human lung adenocarcinoma epithelial cell line A549 (A549/GR) was silenced using small interfering RNA (siRNA) in order to determine the role of microRNA (miRNA) in the development of resistance against epidermal growth factor receptor tyrosine kinase inhibitors (EGFR-TKIs) in lung adenocarcinoma. The relative gefitinib-resistant capacity in A549 and A549/GR cells was determined using a cell counting kit 8. A549/GR cells were transfected with chemically synthesized siRNA to silence the IGF-1R gene. A total of 48 h after siRNA transfection, IGF-1R expression in A549/GR cells was evaluated using reverse transcription-quantitative polymerase chain reaction (RT-qPCR) and western blotting. miRNA expression in A549/GR cells and A549/GR cells with silenced IGF-1R was analyzed using a miRNA microarray. The microarray results of 10 miRNAs were then compared with the results of RT-qPCR. The results demonstrated that the gefitinib-resistance capacity of A549/GR cells was six times higher than that of A549 cells. Additionally, RT-qPCR and western blotting demonstrated that the IGF-1R gene in A549/GR cells was successfully silenced by siRNA. The highest silencing rate (72%) of the IGF-1R gene was obtained using siRNA-2. The microarray identified 72 miRNAs with significantly different expression in A549/GR cells with silenced IGF-1R compared with A549/GR cells. Of the 72 differentially expressed miRNAs, 13 miRNAs (including miR-497-3p and miR-1273c) were up-regulated and 59 miRNAs (including miR-361-3p and miR-345-3p) were down-regulated in A549/GR cells with silenced IGF-1R compared with A549/GR cells. The changes in the expression of 10 different miRNAs were confirmed by RT-qPCR. Thus, the present study successfully established an A549/GR cell line with silenced IGF-1R. The results suggest that a number of miRNAs associated with the IGF-1R signaling pathway, including miR-497-3p and miR-144-5p, were involved in the development of resistance against EGFR-TKIs in A549 cells. These miRNAs may provide novel targets to treat lung adenocarcinoma exhibiting resistance against EGFR-TKIs.

Introduction

Lung cancer is the leading cause of cancer-associated mortality worldwide. Non-small-cell lung cancer (NSCLC) is the predominant form of lung cancer accounting for ~85% of all lung cancer cases (1). The primary use of systemic chemotherapy in the treatment of NSCLC is palliative and produces only modest pain relief (2). Multiple clinical and experimental studies have suggested that mutations in the epidermal growth factor receptor (EGFR) are an indicator of diseases, such as NSCLS (3) and colorectal cancer (4). Epidermal growth factor receptor tyrosine kinase inhibitors (EGFR-TKIs) are molecularly targeted drugs, able to target c-Src and Pak1, among others (5), that are effective at treating patients with advanced NSCLC (6). However, the development of resistance to EGFR-TKIs in tumors inhibits their efficacy. A number of potential mechanisms underlying the development of EGFR-TKI resistance in tumors, including mutations in the KRAS proto-oncogene and ALK receptor tyrosine kinase.
gene fusion, have been identified and an increasing number of studies are focusing on the regulation of EGFR bypass signaling pathways (7,8). Receptor tyrosine kinases (RTK) stand out among causal dominant oncogenes in NSCLC, and the ablation of RTK signaling has emerged as a novel tailored therapeutical strategy (9), and inhibition of mitogen-activated protein kinase (MEK) may decrease cell viability of NSCLC (10). The EGFR/AKT signaling pathway and the mTOR signaling pathway have demonstrated important roles in EGFR-TKIs resistance (11). Other receptor tyrosine kinases (RTK) may activate the downstream phosphoinositol-3 kinase (PI3K)/protein kinase B (Akt), Ras-Raf-mitogen-activated protein kinase 1 (MAPK) and signal transducer and activator of transcription (STAT)/STAT signaling pathways when the EGFR signaling pathway is blocked by EGFR-TKIs (12). As one of the most important components in the EGFR bypass signaling pathways, insulin-like growth factor 1 receptor (IGF-1R) is a potential target. IGF-1R is a transmembrane tetraploid that exhibits tyrosine kinase activity. The binding of IGF-1 to its ligands IGF-1 or IGF-2 induces the auto-phosphorylation of intracellular tyrosine kinase, which promotes the proliferation and metastasis of lung cancer cells by activating the PI3K/Akt and Ras-Raf-MAPK pathways (13).

microRNAs (miRNAs) make up only 1% of the human genome, however they regulate the expression of >30% of all genes (14). The abnormal expression of multiple miRNAs has been identified in the development, angiogenesis, metastasis and drug resistance of lung cancer. The expression of miRNAs associated with the IGF-1R bypass signaling pathways are altered following the inactivation of IGF-1R, which contributes to the proliferation of drug-resistant NSCLC cells (15). In the present study, the expression of IGF-1R in the gefitinib-resistant human lung adenocarcinoma epithelial cell line A549/GR was silenced using small interfering (si) RNA. A number of differentially expressed miRNAs were identified based on the results of a miRNA microarray. It was then determined which of these miRNAs may serve a role in the development of resistance to EGFR-TKIs in patients with NSCLC. The aim of the current study was to identify strategies to inhibit the development of primary resistance to EGFR-TKIs.

Materials and methods

Establishment of the gefitinib-resistant A549/GR cell line. Cells from the human lung carcinoma epithelial cell line A549 (CCL-185; ATCC, Manassas, VA, USA) were used. A569 cells were purchased from American Type Culture Collection (ATCC, Manassas, USA). Gefitinib-resistant A549 cells (A549/GR cells) were established by stepwise selection following exposure to increasing doses of gefitinib (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). Briefly, A549 cells were gradually exposed to gefitinib of increasing concentrations of 5, 10, 20 and 30 µM, at room temperature. After 10 months, A549 cells grew stably at 37°C in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum, 100 U/ml penicillin and 100 g/ml streptomycin (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) containing 30 µM gefitinib. The stability of the gefitinib-resistant cells was confirmed following culture in gefitinib-free medium for 6 months at room temperature.

Evaluation of A549 and A549/GR cell proliferation using the cell counting kit-8 (CCK-8). A549 and A549/GR cells in the logarithmic growth phase were seeded onto a 96-well culture plate (2x10^3-3x10^3 cells/well), the DMEM was purchased from Gibco; Thermo Fisher Scientific, Inc. Different concentrations of gefitinib (2.5, 5, 10, 20, 30, 40, 60, 80 and 120 µM) were added to the DMEM culture medium (Gibco, MA, USA). For the negative control and blank groups, the same amount of culture medium and PBS, respectively, were added instead of gefitinib. For each group, the experiment was conducted in three wells. The cells were cultured in an incubator containing 5% CO2 at 37°C for 72 h. The medium was then replaced with 10 µl CCK-8 kit (catalogue no. 96992; Sigma-Aldrich; Merck KGaA) and 90 µl medium, and the cells were then cultured at room temperature for 3 h. The optical density (OD) value at 450 nm was determined to evaluate the vital rate (VR) of cells according to the following formula: VR(%)=(OD of the gefitinib group-OD of the blank group)/(OD of the negative control group-OD the blank group) x100%. The half maximal inhibitory concentration (IC50) of gefitinib was calculated on the basis of the logarithm of the concentration of gefitinib and the linear regression of VR. The resistance index (RI) of gefitinib was calculated using the following formula: RI=IC50 (A549/GR)/IC50 (A549). The experiments were repeated 3 times with duplicate samples per group.

The use of siRNA to silence IGF-1R. To design the IGF-1R siRNA targets using short hairpin, the IGF-1R nucleotide sequence was obtained from NCBI GenBank databases (https://www.ncbi.nlm.nih.gov/gene/?term=NM_000875;Accession number: NM_000875). The online siRNA design tool (http://www.mwg-biotech.com) was used to design three specific siRNAs: forward 5′-TTTCAAGGAGGCTTCTCTC-3′ and reverse 5′-GAGAGAGCCTCTGTGAAA-3′ (siRNA-1); forward 5′-TAGGACTGGATTATTCCT-3′ and reverse 5′-GGAGAATATCCGGTC-3′ (siRNA-2) and forward 5′-TGATACGATGAGG-3′ and reverse 5′-CCATCAACA ATGAGTACAA-3′ (siRNA-3), to target the IGF-1R gene in the current study. siRNA sequences, positive and negative controls were synthesized by Guangzhou RuiBo Biological Technology Co., Ltd (Guangzhou, China). siRNAs were stored at -20°C, at a concentration of 20 pmol/µl. A549/GR cells were divided into six groups: i) siRNA-1 transfection group (cells transfected with siRNA-1); ii) siRNA-2 transfection group (cells transfected with siRNA-2); iii) siRNA-3 transfection group (cells transfected with siRNA-3); iv) blank group, (untransfected A549/GR cells); v) positive control group (A549/GR cells transfected with GAPDH) and vi) negative control group (A549/GR cells transfected with unrelated siRNA). A549/GR cells (0.5x10^4 per well) were seeded onto a 12-well culture plate. The transfection solution was prepared by mixing 4 µl siRNA-1, siRNA-2 and siRNA-3, respectively, 2 µl lipofectamine reagent (Gibco; Thermo Fisher Scientific, Inc.) and 200 µl DMEM without serum (Gibco; Thermo Fisher Scientific, Inc.). After standing at room temperature for 20 min, the transfection solution was added to each well. Cells were cultured with the transfection solution in an incubator containing 5% CO2 at 37°C for 5-6 h. The supernatant was then replaced with culture medium and cells were cultured at 37°C for 48 h.
Evaluation of IGF-1R mRNA using reverse transcription-quantitative polymerase chain reaction (RT-qPCR).

Total RNA was isolated from the cells of each group using TRIzol (Thermo Fisher Scientific, Inc., Waltham, MA, USA) and the quality and concentration of total RNA was assessed on a UV spectrophotometer (Ultraspex 3300; Biochrom, Ltd., Cambridge, UK). Complementary DNA (cDNA) was obtained using a reverse transcription kit (DTR056A; Takara Bio, Inc., Otsu, Japan). The primers used in qPCR were as follows: IGF-1R, forward 5'-AACAGCCCCACAGGTATG-3' and reverse 5'-GCTGCTTGGCAGCTTGAG-3'; GAPDH, forward 5'-GGACCTGACCTGCGCTTAG-3' and reverse 5'-GAGGAGTGGTGTGCTGGTT-3'. The PrimeScript RT Reagent kit (catalogue no. RR391A, Takara Bio, Inc.) was used for qPCR and SYBR Green I (catalogue no. RR086A; Takara Bio, Inc.) was used as the reporter fluorophore, which was conducted in a 20 µl reaction system with 10 µl 2x Mix buffer, forward and reverse primers (0.4 µl each), 1 µl cDNA, and 15.4 µl double-distilled water. The reaction was performed as follows: 94°C for 5 min, 30 cycles of 94°C for 1 min, 45°C for 1 min and 72°C for 40 sec. The reaction was performed in a Bio-Rad CFX qPCR machine (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The 2-ΔΔCq method was applied for quantification of relative expression of target genes (16).

Evaluation of IGF-1R protein expression using western blotting. The expression of IGF-1R was evaluated using western blot analysis. Cells from each group (2 ml) were seeded into a 6-well plate (2x10⁶ cells/well). Following 24 h incubation at 37°C, cells were collected and centrifuged at 12,000 g x g for 10 min at 4°C. The protein was extracted by using cold RIPA buffer (Gibco; Thermo Fisher Scientific, Inc.) and centrifuged at 12,000 g x g for 15 min at 4°C. Subsequently, IGF-1R protein was quantified using the Bradford protein quantification method, and by measuring absorbance at 595 nm using a microplate reader (Bio-Rad® Benchmark microplate reader, Bio-Rad Laboratories, Inc.). Proteins (50 µg/well) were separated by 12.5% SDS-PAGE and transferred onto a nitrocellulose membrane. Following blocking in 5% non-fat milk at room temperature overnight, bands were visualized using enhanced chemiluminescence (ECL; catalogue no. ab6721; Abcam) at room temperature for 1 h. The membranes were incubated horseradish peroxidase (HRP)-conjugated anti-rabbit secondary antibodies (1:5,000; catalogue no. ab8245; Abcam, Cambridge, MA, USA) at room temperature overnight. Anti-GAPDH (catalogue no. ab1418; Abcam) was used as an internal control. After washing with PBS twice for 10 min, the membranes were incubated horseradish peroxidase (HRP)-conjugated anti-rabbit secondary antibodies (1:5,000; catalogue no. ab6271; Abcam) at room temperature for 1 h and visualized using enhanced chemiluminescence (ECL; catalogue no. WP20005; Invitrogen; Thermo Fisher Scientific, Inc.) according to manufacturers' protocol. Band intensity was determined using chemiluminescent (ECL) reagents (Pierce; Thermo Fisher Scientific, Inc.) and analyzed on Image J software version 14.1 (National Institutes of Health, Bethesda, MD, USA).

Microarray analysis of miRNA gene expression. A total of 2,565 miRNA probes were included on the RiBo Array™ miRNA Human Array (Shanghai Biotechnology Corporation, Shanghai, China). The probes were designed using the miRBase database (Sanger miRBase Release 21.0; http://www.mirbase.org). Following hybridization, the laser scanner GenePix 4000B (Molecular Devices, LLC, Sunnyvale, CA, USA) was used to obtain hybridization images. The results were analyzed using the Array-Pro 3.0 software (Axon Instruments, Inc., Union City, CA, USA). The Locally-Weighted Regression (LOWESS) method (17) was used to normalize the microarray results and calculate the fold changes and P-values for the paired t test. Fold changes of >1.5 or < -1.5 were determined to be statistically significant (P<0.05). Significant differences in the expression of miRNAs between the A549/GR-siRNA2 and the negative control group were identified using heat map clustering and scatter plots. The scatter plot was used to assess variation in miRNA expression between the siRNA2 and negative control group. The raw data from the microarray images were then extracted using Feature Extraction software version 11.0.1.1 (Agilent Technologies, Santa Clara, CA, USA), and the raw data was normalized using Locally-Weighted Regression (LOWESS; Silicon Genetics, Inc., Redwood, CA, USA). The values obtained for the X and Y axes in the scatter plot were the normalized signal values determined for the samples (log2 scaled) or the average normalized signal values obtained for the groups of samples (log2 scaled). Heat map clustering was performed using Cluster 3.0 software (http://rana.iblb.gov/EisenSoftware.htm; Lawrence Berkeley National Laboratory, Berkeley, CA, USA). The colors in the map present the relative values of all tiles within two groups. Red represents up-regulated and green represents down-regulated miRNAs. The brightness of the color indicates the degree of up- or down-regulation.

Confirmation of microarray results: miRNA levels measured by RT-qPCR. A total of 10 miRNAs that exhibited the most significant differences in expression between the A549/GR cells and A549/GR cells with silenced IGF-1R, were selected for RT-qPCR. Total RNA was isolated from cells of each group using TRIzol reagent according to the manufacturer protocol (Thermo Fisher Scientific, Inc.) and the quality and concentration of total RNA was assessed on a UV spectrophotometer (Ultraspex 3300; Biochrom, Ltd., Cambridge, UK). cDNA was obtained using the One Step Prime Script™ RT-PCR kit (catalogue no. RR064A; Takara Bio, Inc.). The cDNA samples were then used for RT-qPCR measurement with miRNA-specific primers. SYBR Green I was used as the reporter fluorophore (Takara Bio, Inc.). The primers used in this study were as follows: i) miR-144-5p Forward 5'-CTGCAAGGATATCATCATC-3'; Reverse 5'-GTGAGGCTTCCGAG-3'; ii) miR-550a-5p Forward 5'-CTGACAGTGCTGAGGGAG-3'; Reverse 5'-GTGCAGGTCTCCGAGGT-3'; iii) miR-361-3p Forward 5'-GTGCAGGTCTCCGAGGT-3'; Reverse 5'-CTGCACTCCCCCACCCTG-3'; iv) miR-345-3p Forward 5'-CTGCACGCGCCTAGAG-3'; Reverse 5'-CTGCACGGTCCGAGGT-3'; v) miR-4450 Forward 5'-CTGCACTGGAGATTTTG-3'; Reverse 5'-GTGACAGGGTCCGAGGT-3'; vi) miR-135a-3p Forward 5'-CTGCACTGGAGATTTTG-3'; Reverse 5'-GTGACAGGGTCCGAGGT-3'; vii) miR-135b-3p Forward 5'-CTGCACTGGAGATTTTG-3'; Reverse 5'-GTGACAGGGTCCGAGGT-3'; viii) miR-137-3p Forward 5'-CTGCACTGGAGATTTTG-3'; Reverse 5'-GTGACAGGGTCCGAGGT-3';
CCK8 demonstrated that the viability of A549/GR and A549 cells decreased as concentrations of gefitinib increased in the culture medium. However, the IC<sub>50</sub> value of gefitinib for A549/GR cells was 48.05±4.33 µmol/l, which is significantly greater than that of A549 cells (8.02±0.63 µmol/l; P<0.01; Fig. 1), this suggested that A549/GR exhibits significantly increased resistance to gefitinib than A549 cells.

IGF-1R expression is successfully silenced by siRNAs. To examine the effect of siRNAs on the expression of IGF-1R, the expression of IGF-1R mRNA and protein was assessed using RT-qPCR and western blotting. The expression of IGF-1R mRNA was inhibited by siRNA-1, siRNA-2, and siRNA-3 (Fig. 2). A total of 48 h after transfection with siRNA, the relative Cq value of IGF-1R for the siRNA-1, siRNA-2, siRNA-3 groups were 0.55±0.05, 0.28±0.02, and 0.62±0.03, respectively. The expression of IGF-1R in all three siRNA groups was significantly lower than in the blank control group (P<0.05; Fig. 2A). These results demonstrate that siRNAs successfully inhibit the expression of IGF-1R mRNA. The siRNA silencing efficiency was calculated according to the following formula: siRNA silencing efficiency=(1-IGF-1R expression in the siRNA group/IGF -1R expression in the blank group) x100%. siRNA-2 exhibited the highest efficiency (72%) of silencing (Fig. 2B). Western blot analysis demonstrated that in cells transfected with all three siRNAs, there was a significant decrease in IGF-1R expression (P<0.01) and that siRNA-2 exhibited the highest inhibition efficiency (Fig. 3). Therefore, siRNA-2 was used in subsequent experiments to silence IGF-1R and evaluate alterations in miRNA expression.

Differentially expressed miRNAs in A549/GR cells with silenced IGF-1R compared with A549/GR cells. Based on the miRNA microarray, 72 differentially expressed miRNAs were selected for further analysis. The primers for qPCR were designed as follows: Forward 5'-CTG TAT GGC GAC AAA ACG-3', Reverse 5'-GTG CAG GGT CCG AGG T-3'; x) miR-497-3p Forward 5'-CTG CGA GGT GA CCG AGG T-3', Reverse 5'-CTG CGA GGT GA CCG AGG T-3'. The Takara Kit (Takara Bio, Inc.) was used for qPCR, which was conducted in a 20 µl reaction system with 10 µl 2x Mix buffer, forward and reverse primers (0.4 µl each), 1 µl cDNA, and 15.4 µl double distilled water. The PCR parameters were as follows: 95˚C for 1 min, followed by 40 cycles of 95˚C for 10 sec and extension at 64˚C for 25 sec. The 2<sup>ΔΔCq</sup> method was used to determine the relative expression of miRNAs (16).

Statistical analyses. Statistical analyses were conducted using SPSS software 19.0 (IBM Corp., Armonk, NY, USA). Data were expressed as mean ± standard deviation. Comparisons between different groups were conducted using the t-test for two independent samples and P<0.05 was considered to indicate a statistically significant difference.

Results

A549/GR cells exhibit gefitinib resistance. The results of CCK8 demonstrated that the viability of A549/GR and A549 cells decreased as concentrations of gefitinib increased in the culture medium. However, the IC<sub>50</sub> value of gefitinib for A549/GR cells was 48.05±4.33 µmol/l, which is significantly higher than that of A549 cells (8.02±0.63 µmol/l; P<0.01; Fig. 1), this suggested that A549/GR exhibits significantly increased resistance to gefitinib than A549 cells.

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| Number | miRNA name               | Fold change | P-value   |
|--------|--------------------------|-------------|-----------|
| 1      | hsa-miR-19a-5p           | 5.0666666667 | 0.001502792 |
| 2      | hsa-miR-4477a            | 2.419354839 | 0.006793443 |
| 3      | hsa-miR-497-3p           | 2.0560632911 | 0.006056884 |
| 4      | hsa-miR-76412            | 1.920393783 | 0.006714308 |
| 5      | hsa-miR-4285             | 1.907407407 | 0.025745132 |
| 6      | hsa-miR-4804-3p          | 1.650793651 | 0.01466040 |
| 7      | hsa-miR-487b-3p          | 1.621626122 | 0.017437008 |
| 8      | hsa-miR-1973             | 1.595256312 | 0.00733254 |
| 9      | hsa-miR-1273c            | 1.58258483  | 0.002789136 |
| 10     | hsa-miR-337-3p           | 1.571428571 | 0.025546335 |
| 11     | hsa-miR-4467             | 1.553892216 | 0.028963891 |
| 12     | hsa-miR-544b             | 1.532608696 | 0.03936994 |
| 13     | hsa-miR-3679-3p          | 1.505596721 | 0.01655788 |
| 14     | hsa-miR-144-5p           | -0.47360917 | 0.046285311 |
| 15     | hsa-miR-6789-5p          | -0.966396292 | 0.000284606 |
| 16     | hsa-miR-6885-5p          | -2.902923977 | 0.043100598 |
| 17     | hsa-miR-550a-5p          | -2.372917298 | 0.004168534 |
| 18     | hsa-miR-4706             | -2.249421742 | 0.012286458 |
| 19     | hsa-miR-1238-3p          | -2.171066526 | 0.019221921 |
| 20     | hsa-miR-3131             | -2.112351179 | 0.040649198 |
| 21     | hsa-miR-762              | -0.094218057 | 0.006013498 |
| 22     | hsa-miR-524-3p           | -0.075471698 | 0.040855564 |
| 23     | hsa-miR-1233-5p          | -0.041464467 | 0.028882678 |
| 24     | hsa-miR-6856-5p          | -0.034500649 | 0.033718572 |
| 25     | hsa-miR-877-3p           | -0.028571429 | 0.032463565 |
| 26     | hsa-miR-4722-5p          | -1.928336079 | 0.021888284 |
| 27     | hsa-miR-6851-5p          | -1.925488809 | 0.015705194 |
| 28     | hsa-miR-3197             | -1.923085887 | 0.017365446 |
| 29     | hsa-miR-361-3p           | -1.891474966 | 0.01367353 |
| 30     | hsa-miR-6835-5p          | -1.884668508 | 0.041693405 |
| 31     | hsa-miR-6835-3p          | -1.882758621 | 0.007597698 |
| 32     | hsa-miR-6746-5p          | -1.862366121 | 0.020666861 |
| 33     | hsa-miR-8066             | -1.848739496 | 0.004502012 |
| 34     | hsa-miR-4758-5p          | -1.81223111  | 0.010309085 |
| 35     | hsa-miR-598-5p           | -1.8058977035 | 0.010516834 |
| 36     | hsa-miR-1193             | -1.808841099 | 0.00103303 |
| 37     | hsa-miR-345-3p           | -1.8        | 0.04155795 |
| 38     | hsa-miR-6724-5p          | -1.770495483 | 0.006848465 |
| 39     | hsa-miR-6750-5p          | -1.769516792 | 0.006843986 |
| 40     | hsa-miR-4313             | -1.741487144 | 0.036712854 |
| 41     | hsa-miR-573              | -1.715715973 | 0.010358878 |
| 42     | hsa-miR-6824-5p          | -1.731818182 | 0.022809783 |
| 43     | hsa-miR-4450             | -1.727125068 | 0.046979788 |
| 44     | hsa-miR-6889-5p          | -1.725773196 | 0.016773382 |
| 45     | hsa-miR-4665-5p          | -1.722864198 | 0.030246907 |
| 46     | hsa-miR-3683             | -1.721925134 | 0.034157758 |
| 47     | hsa-miR-555              | -1.711229947 | 0.025664868 |
| 48     | hsa-miR-4783-3p          | -1.706832298 | 0.040236296 |
| 49     | hsa-miR-4741             | -1.67372657 | 0.006643116 |
| 50     | hsa-miR-6800-3p          | -1.673211781 | 0.016012437 |
| 51     | hsa-miR-6786-5p          | -1.670338552 | 0.038049277 |

were identified in A549/GR cells that had undergone IGF-1R silencing compared with control A549/GR cells (P<0.05). Of the 72 differentially expressed miRNAs, 13 were up-regulated and 59 were down-regulated (Table I). The levels of expression of 15 miRNAs in A549/GR cells with IGF-1R silencing were two times higher or lower than in A549/GR cells. These differentially expressed miRNAs may serve important roles in the development of geritinib-resistance in A549/GR cells. Heat map clustering was performed based on ‘All Targets value-miRNA’ using the siRNA-2 group and negative control group; the clustering results revealed distinct miRNA expression profiles in the two groups (Fig. 4). A scatter plot was constructed indicating the overlapping miRNA expression profiles of the siRNA-2 and negative control groups. The values obtained for the X and Y axes in the scatter-plot are the normalized signal values of the samples (log2 scaled) or the averaged normalized signal values of groups of samples (log2 scaled: Fig. 5). That there were differences in the expression profiles of 10 miRNAs between the siRNA-2 and negative control in A549 cells, indicating that these 10 miRNAs expression profiles were changed after transfection with IGF-1R siRNA-2 in A569 cells. The scatter plot was used to assess variation in miRNA expression between the A549 cells transfected with siRNA-2 and the negative control group. The axes of the scatter plot are the normalized signal values of the samples (the ratio scale), red color represents upregulated miRNAs, green color represents downregulated miRNAs, gray color represents equally-expressed miRNAs.
miRNA expression confirmed by RT-qPCR. The miRNA microarray results were further evaluated using RT-qPCR to evaluate the expression of 10 miRNAs, including miR-144-5p, miR-550a-5p, miR-361-3p, miR-345-3p, miR-4450, miR-135a-3p, miR-124-3p, hsa-miR-337-3p, hsa-miR-1273c and hsa-miR-497-3p, which exhibited the strongest hybridization signal. It was demonstrated that the results of RT-qPCR were consistent with those of the microarray, suggesting that the results of the miRNA microarray are reliable (Fig. 6).

Discussion

IGF-1R and EGFR share two downstream signaling pathways, the Ras-Raf-MAPK and PI3K-Akt signaling pathways, which are involved in cell proliferation and apoptosis, as well as angiogenesis (18). In the present study, A549 cells were selected to identify why gefitinib exhibits poor efficacy in lung cancer cells. A549 cells are Ras-mutated, PI3K-mutated and EGFR wild-type and exhibit resistance to first-generation EGFR TKIs, such as gefitinib (19). Previous studies have demonstrated that EGFR-TKIs are effective against lung cancer in vitro and in vivo (20,21). Inhibition of EGFR by TKIs, including gefitinib and erlotinib, has provided hope for patients with NSCLC. However, a number of studies have reported that one of the EGFR TKIs is a more effective treatment for patients with NSCLC that exhibit EGFR mutations than in patients with wild-type EGFR, as patients with EGFR mutations are typically highly sensitive to EGFR TKIs (22,23). Due to acquired resistance of EGFR TKIs via EGFR-mutant NSCLC that arise through various molecular mechanisms including EGFR secondary mutations, MET proto-oncogene amplification and hepatocyte growth factor (HGF) overexpression, the anti-tumor effect of EGFR TKIs may also be weakened (24,25). Hence, the present study used the EGFR wild-type lung cancer cell line A569 as the cell model, to avoid the EGFR tyrosine kinase secondary mutation in the process of stepwise gefitinib selection.

A previous study by the current authors identified the increased expression of IGF-1R mRNA in A549/GR cells, which is associated with gefitinib-resistance (26). The results of the current study suggest that IGF-1R activation contributes to the development of secondary resistance to gefitinib. Activated IGF-1R bypasses the EGFR pathway to directly activate the downstream Ras-Raf-MAPK and PI3K-Akt signaling pathways, which promote the proliferation and metastasis of tumor cells and secondary resistance to gefitinib (27).

Aberrant expression of miRNA has been identified in many tumors (28,29). The complementary binding between miRNAs and their target mRNAs induces the formation of RNA-induced silencing complexes (RISCs), which degrade mRNAs or inhibit the translation of mRNAs (30). During cancer development and metastasis, miRNAs acts as a proto-oncogene and a tumor suppressor gene. One miRNA may target multiple mRNAs and several miRNAs may coordinate to regulate the expression of a single mRNA (31). In addition, the transcription of miRNA is regulated by transcription factors, which form a complicated regulatory network (32). The characterization of miRNAs involved in the IGF-1R pathway may facilitate the identification of novel targets to treat different types of cancer that are resistant to gefitinib. As an effective tool for gene knockout, siRNA interference has been widely used in biological research. Following siRNA transfection into host cells, the siRNA duplex melts and integrates into the RISC. The reverse strand of siRNA guides the complementary binding between RISC and mRNA, leading to the efficient and specific degradation of intracellular mRNA and gene silencing (33). Silenced expression of a gene may inactivate the downstream signaling pathway, which facilitates the identification and characterization of the regulatory miRNAs involved in the signaling pathway (34). To efficiently silence a specific gene, a number of siRNAs must be designed and their silencing efficiency compared under the same conditions (35). In the present study, three siRNAs were designed on the basis of the nucleotide sequence of IGF-1R gene and a pair of unrelated nucleotides was used as a negative control.

It has been demonstrated that exogenous double-stranded RNA >50 base pairs long may activate the interferon pathway, thus inducing apoptosis (36). However, the transfection of double-stranded RNA of 19-23 nucleotides long can efficiently...
and rapidly silence target genes without affecting the interferon pathway. Therefore, in the current study, siRNAs 19 nucleotides long were used to avoid interference with the interferon pathway. In addition, numerous studies have demonstrated that siRNAs containing 30-50% guanine and cytosine (GC) are more efficient at gene silencing than siRNAs containing...
is a reliable tool to use to identify differentially expressed the microarray results, suggesting that the miRNA microarray measured using RT-qPCR and were consistent with those of hsa-miR-497-3p, hsa-miR-1273c and hsa-miR-337-3 were miR-361-3p, miR-345-3p, miR-4450, miR-135a-3p, miR-124-3p, hsa-miR-337-3p, hsa-miR-1273c and hsa-miR-497-3p. It was demonstrated that the results of RT-qPCR results were consistent with the microarray results. RT-qPCR, reverse transcription-quantitative polymerase chain reaction; miRNA, microRNA.

Figure 5. Scatter plot. The scatter plot was used to assess variation in miRNA expression between the two groups. The values obtained for the X and Y axes in the scatter plot are the normalized signal values determined for the samples (log2 scaled) or the average normalized signal values obtained for the groups of samples (log2 scaled). miRNA, microRNA; siRNA, small interfering RNA; NC, negative control.

miRNAs. The expression of miR-497-3p was significantly increased in A549/GR cells with silenced IGF-1R. miR-497-3p, a member of the miR-15/16/195/424/497 family, is located on the short arm of chromosome 17 (17p13.1). As with other members of the miR-15/16/195/424/497 family, miR-497-3p is involved in mammalian meiosis by regulating the expression of cell cycle-related genes (39). It has been demonstrated that miR-497 transfection leads to G1/G0 cell cycle arrest in host cells (39). Zhao et al (40) identified the down-regulation of miR-497 in NSCLC tissue, as well as A549, SPC-A1, H1299 and H460 cells. Another study demonstrated that miR-497 promotes the apoptosis of NSCLC cells through the hepatoma-derived growth factor (HDGF) (40). Han et al (41) reported that miR-497 and miR-34a inhibited the proliferation and metastasis of lung cancer cells by binding to the 3'-untranslated region (UTR) of the cyclin E1 gene and inhibiting its expression, the synergistic actions of miR-497 and miR-34a partly correlated with cyclin E1 levels, cyclin E1 is downregulated by both miR-497 and miR-34a, which synergistically retard the growth of human lung cancer cells. In the present study, the expression of miR-497-3p in A549/GR cells with silenced IGF-1R was significantly different than that in A549 cells, suggesting that miR-497-3p is involved in the development of gefitinib resistance in NSCLC cells through the IGF-1R bypass pathway.

Of the 71 differentially expressed miRNAs, miR-144-5p exhibited the most significant difference in expression between A549/GR cells and A549/GR cells with silenced IGF-1R. However, the role of miR-144-5p in the development and progression of tumors is controversial. Studies have suggested that miR-144-5p acts as a tumor suppressor gene during the development of osteosarcoma. Ye et al (42) reported that miR-144-5p promotes the progress of colon cancer. Zha et al (43) determined that the expression of miR-144-5p in NSCLC tissues was significantly lower than in adjacent tissues (P<0.01) and that the expression of miR-144-5p in A549 cells was significantly lower than in normal bronchial epithelial cells (P<0.01). Increased expression of the apoptotic marker apoptosis related protein 3 (ARP3) and cytochrome C has been detected in A549 cells overexpressing miR-144-5p (44).
In addition, it has been demonstrated that miR-144-5p inhibits the growth and promotes the apoptosis of A549 cells (45). Therefore, it was speculated that miR-144-5p serves a tumor suppressor role in NSCLC and that zinc finger proteins may be involved in the underlying mechanism. Xiang et al (46) demonstrated that miR-144-5p inhibits the proliferation and invasion of A549 and H460 cells through the TP53-inducible glycolysis and apoptosis regulator. Furthermore, Han et al (47) demonstrated that the down-regulation of miR-223 may activate the IGF-1R/phosphatidylinositol 3-kinase (PI3K)/Akt pathway in lung cancer PC9/ER and PC9/CD133+ cells, suggesting that miR-233 is a potential therapeutic target for overcoming resistance to EGFR-TKIs.

In the present study, the results from the microarray and RT-qPCR identified significant differences in the expression of miR-497-3p and miR-144-5p in A549/GR cells with silenced IGF-1R compared with A549/GR cells. This suggests that miR-497-3p and miR-144-5p stimulate the development of gefitinib-resistance in A549 cells through the IGF-1R bypass pathway. However, the underlying molecular mechanisms have not yet been reported and further studies are therefore required.

In conclusion, the current study identified 72 differentially expressed miRNAs using a miRNA microarray in A549/GR cells in which the IGF-1R gene was silenced using siRNA. The results of the current study provide a basis for investigating the roles of miRNAs in the development of resistance to gefitinib in lung adenocarcinoma. Given that miRNAs involved in the IGF-1R bypass pathway serve important roles in the proliferation of drug-resistant NSCLC cells, the identification of these miRNAs may establish novel targets to treat gefitinib-resistant NSCLC.

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