Liver Fluke Induces Cholangiocarcinoma

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Opisthorchiasis and Clonorchiasis: Major Regional Public Health Problems

Liver fluke infection caused by *Opisthorchis viverrini*, *O. felineus*, and *Clonorchis sinensis* is a major public health problem in East Asia and Eastern Europe. Currently, more than 600 million people are at risk of infection with these trematodes [1]. *O. viverrini* is endemic in Southeast Asian countries, including Thailand, Lao People’s Democratic Republic, Vietnam, and Cambodia [2], and *C. sinensis* infection is common in rural areas of Korea and China. Opisthorchiasis has been extensively studied in Thailand, where an estimated 6 million people are infected with the liver fluke (calculated from overall 9.4% prevalence within the population in 2001) [3].

Infection with these food-borne parasites is prevalent in areas where uncooked cyprinoid fish is a staple of the diet. Due to poor sanitation practices and inadequate sewerage infrastructure, people infected with *O. viverrini* and *C. sinensis* pass parasite eggs in their faeces into natural water reservoirs, where the parasite eggs are eaten by intermediate host snails, for example, aquatic snails of the genus *Bithynia*, the first intermediate host of *O. viverrini*. After hatching, free swimming parasites, called cercariae, are released from the infected snails. Cercariae then locate their next intermediate host, cyprinoid fishes, encyst in the fins, skin, and muscles of the fish, and become metacercariae. The metacercariae are infective to humans and other fish-eating mammals upon ingestion of raw or undercooked fish in dishes such as *koi-pla* (Figure 1), and in turn the parasite’s life cycle is completed (Figure 2).

Most people with opisthorchiasis or clonorchiasis have no symptoms. Only 5%–10% of infected people, in general those with heavy fluke infections, have non-specific symptoms such as right upper quadrant abdominal pain, flatulence, and fatigue [4,5]. Enlargement of the gall bladder can be detected by ultrasonography, and is reversed after elimination of flukes by praziquantel [6]. Nonetheless, heavy, long-standing infection is associated with a number of hepatobiliary diseases, including cholangitis, obstructive jaundice, hepatomegaly, fibrosis of the periporal system, cholecystitis, and cholelithiasis [7–10]. Moreover, both experimental and epidemiologic evidence strongly implicates liver fluke infection in the aetiology of one of the liver cancer subtypes—cholangiocarcinoma (CCA), or cancer of the bile ducts [2,11].

The pathology of clonorchiasis was recently reviewed in detail by Rim [12]. Unlike *O. viverrini*, *C. sinensis* is not considered a Group I carcinogen (known to be carcinogenic in humans) [2], despite its widespread prevalence [13].

Funding: We gratefully acknowledge support from the United States National Institute of Allergy and Infectious Diseases, grant number 1U01 AI065871, and the Sandler Family Supporting Foundation. Neither the National Institute of Allergy and Infectious Diseases nor the Sandler Foundation was involved in the preparation of this article or in the decision to publish.

Competing Interests: PJB is the Principal Investigator of the National Institute of Allergy and Infectious Diseases grant number 1U01 AI065871, which deals with liver fluke–induced cholangiocarcinoma.

Citation: Sripa B, Kaewkes S, Sithithaworn P, Mairiang E, Laha T, et al. (2007) Liver fluke induces cholangiocarcinoma. PLoS Med 4(7): e201. doi:10.1371/journal.pmed.0040201

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Abbreviations: CCA, cholangiocarcinoma; ES, excretory/secretory; HCC, hepatocellular carcinoma; NO, nitric oxide

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The Neglected Diseases section focuses attention either on a specific disease or describes a novel strategy for approaching neglected health issues in general.
this review we therefore focus on opisthorchiasis, where the link between fluke infection and CCA is more robust than for clonorchiasis. Further, the article reviews the molecular pathogenesis of opisthorchiasis and associated cholangiocarcinogenesis, particularly nitrative and oxidative DNA damage and clinical manifestations of CCA that were either not addressed or available in our previous review [14].

Liver Fluke Infection and Cholangiocarcinoma

Second to tobacco use, infections are the most important preventable source of human malignancies [15–17] (Table 1). The association between the occurrence of CCA and the presence of liver flukes has been known for about 50 years [18].

In Thailand, the first evidence for such an association came from hospital case series. CCA was observed in a high percentage of liver cancers from northeast Thailand, where the prevalence of *O. viverrini* infection is higher than elsewhere in this country [19]. More formal correlation studies showed that the incidence of CCA in the five major regions of Thailand varied by at least 12-fold and had a strong positive correlation with the prevalence of *O. viverrini* infection, as measured by anti-*O. viverrini* antibody titres in the general population (Figure 3) [20,21]. Similar correlations have been observed in populations in Laos [22,23]. Hepatocellular carcinoma (HCC) shows no such relationship.

Several similar surveys of Thai villagers showed a strong association between the intensity of *O. viverrini* infection, parasite-specific antibody response, and abnormalities of the biliary tract, including suspected CCA [8,24]. A linear trend of the frequency of suspected CCA and *O. viverrini* faecal egg count was observed, where an odds ratio of 14.1 was found in a group with elevated egg counts [25]. Moreover, elevated anti-*O. viverrini* antibody titres increase the risk of appearance of CCA by up to 27-fold [26].

The geographical pattern of liver fluke infection in Thailand is not uniform, with the greatest prevalence...
occurring in the north (19.3%) and northeast (15.7%), compared with the central (3.8%) and southern regions (0%) [3]. Khon Kaen province in northeast Thailand has the highest incidence of this type of cancer in the world [27,28].

In Thailand, despite widespread implementation of chemotherapy with praziquantel in the past, the prevalence of *O. viverrini* in some endemic areas approaches 70%. Moreover, in Thailand, liver cancer is the most prevalent of the fatal neoplasias, and rates of CCA in regions where the parasite is endemic are unprecedented. By way of comparison, CCA is responsible for about 19% of liver cancers in the United States, compared with 71% in Khon Kaen, Thailand, representing the highest incidence of CCA in the world in this region [28].

**Clinical Manifestations**

Most primary malignant liver cancers are of two main histologic types distinguished by their cellular origin. HCC derives from hepatocytes, the main cells in the liver, and is the most common form of liver cancer throughout the world. There are several important risk factors that have been demonstrated with varied mechanisms of action; 75%–80% of cases are attributable to hepatitis B and C infection [29,30]. CCA is derived from cholangiocytes, which form the epithelial lining of both intrahepatic and extrahepatic bile ducts, except for those of the gallbladder. This form of liver cancer has a lower operable rate than HCC and most cases are currently untreatable except for complete liver transplantation [31]. Compared to HCC, CCA is a considerably less common form of liver cancer, except in regions within East Asia where infection with *O. viverrini*, and to a lesser extent, *C. sinensis*, is widespread [32].

CCA represents a formidable diagnostic and treatment challenge [33]. The wide variation in CCA biology and the rarity of the condition (which represents only 3% incidence of all gastrointestinal cancers) have significantly increased the difficulty of any studies involving this cancer, because of broad variations in presentation and low study participant numbers [34]. The disease’s rarity, combined with its lack of early symptoms and a range of possible alternative diagnoses, contributes to the challenge of identifying and investigating this deadly cancer [35]. Practically all cases are derived from glandular tissue, making them adenocarcinomas. Details of the gross morphology, histology, mode of spreading, and clinical manifestations are outlined in Figure 4 [36,37].

**Pathogenesis of Liver Fluke-Induced CCA**

**Mechanical injury and fluke metabolic products.** Mechanical injury from the activities of feeding and migrating flukes contributes to biliary damage in the human host. Both oral and ventral suckers of the fluke hook onto the biliary epithelium, resulting in tissue damage even early in infection [38]. As the parasite matures, the lesion becomes more pronounced and ulcerates. Fluke eggs become entrapped in the periductal tissue through the ulcer and induce granulomatous inflammation around the eggs. The granulomata are readily visualized in experimental hamster infections and occasionally in human cases with bile duct obstruction.

The liver fluke secretes or excretes metabolic products, some of which are highly immunogenic, from the tegument and excretory openings into the bile or culture medium in vitro [39,40]. Apart from inducing host immune responses, the metabolic products themselves may be toxic to or interact with the biliary epithelium [41]. Indeed, mouse fibroblasts co-cultured with *O. viverrini* adult worms (but physically separated in Transwell plates) underwent more than 4-fold greater proliferation than cells in media alone [42].

Gene microarrays were recently used to explore transcriptional changes induced in these murine fibroblasts that were co-cultured with *O. viverrini*, and the fluke molecules induced over-expression of several mRNAs encoding growth-promoting proteins such as transforming growth factor [43]. Moreover, we observed that human CCA cell lines underwent excessive proliferation upon stimulation with *O. viverrini* soluble excretory/secretory (ES) products (B. Sripa et al., unpublished data). These studies clearly indicate that metabolic products of *Opisthorchis* contain mitogen-like activity and can induce cell proliferation. This is in agreement with earlier findings of hyperplasia of biliary epithelial cells in opisthorchiasis [38,44].

It is intriguing that *O. viverrini* would secrete a protein that promotes cell proliferation and eventually cancer, since the
benefit to the parasite is unclear. One possibility might be that over-production of biliary cells may provide an abundant food source for the flukes, which graze on epithelial cells and/or their secreted mucins, though it has been proposed that *C. sinensis* ingests host blood through the damaged mucosa [12].

**Immunopathology.** It has long been suspected that host immune responses and immunopathologic processes mediate hepatobiliary damage in opisthorchiasis [38,45,46]. Sripa and Kaewkes [40] showed that inflammation around infected hamster bile ducts was a consequence of the host’s cellular response to *Opisthorchis* antigens. Marked infiltration of inflammatory cells at the periportal areas of infected hamster liver was associated with the presence of parasite antigens in the bile duct epithelium as detected by immunohistochemistry. Small bile ducts, the first-order ducts (where flukes do not reside because the diameter of the ducts is too small), were also positive for *O. viverrini* antigens and were markedly inflamed. *Opisthorchis* antigens were also observed in macrophages, epithelioid cells, and giant cells of the granuloma [40].

Biliary cell damage induced by *O. viverrini* likely also stems from the actions of oxygen radicals such as nitric oxide (NO) released from effector cells activated by inflammatory cytokines [47,48]. These radicals can induce oxidative DNA damage to the infected biliary epithelium. Moreover, excess NO and other reactive oxygen intermediates produced by inflammatory cells during infection might exert direct cytotoxic and mutagenic effects and cause increased cell proliferation [49–51]. Increased formation of 8-nitroguanine (8-NO2-G) and 8-oxo-7,8-dihydro-2’-deoxyguanosine (8-oxodG) is associated with various pathological conditions [52,53], and 8-oxodG is considered to be mutagenic. Recently, we reported diffuse nitrative and oxidative DNA damage (8-nitroguanine and 8-oxodG) in biliary epithelium of hamsters infected with *O. viverrini* [48], and these DNA lesions still remained in the epithelium at least 180 days post-infection. Moreover, repeated infections with liver fluke result in enhanced biliary DNA damage [54]. This may be explained by the fact that repeated infection increased inducible nitric oxide synthase expression in the bile duct epithelium. The DNA damage in infected biliary cells is proven to be a result of the inflammatory response caused by *O. viverrini* because 8-nitroguanine and 8-oxodG disappeared after praziquantel treatment [55]. Usually genotoxic events with DNA damage lead to either DNA-mismatch repair mechanisms or, if the damage is beyond repair, to cell death through apoptosis. NO not only induces DNA damage but has been reported to mediate DNA repair inhibition [56,57]. Moreover, NO has also been demonstrated to inhibit

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**Figure 3.** Incidence of CCA and *O. viverrini* in Thailand from 1990–2001

Increasing intensity of red represents increasing prevalence of *O. viverrini*, while increasing number of dots represents increasing cancer rates. In general, higher *O. viverrini* prevalence correlates with a higher CCA burden, although sporadic anthelmintic therapy has influenced this relationship. It should be noted that even one spot represents significant cancer rates anywhere else in the world. *Truncated age-standardised incidence from 35–64 years. **Age-standardised incidence of CCA throughout registered regions. Adapted from [20].

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**TABLE 2.**

| CCA incidence * | Khon Kaen Province Average 24.5% |
|-----------------|----------------------------------|
| (per 100,000 person yrs) |                                |
| Khon Kaen Avg 118.8 |
| <109           | Below 4%                        |
| 110-169        | 5-10.9%                         |
| 170-229        | 11-17.9%                        |
| 230-279        | 18-24.9%                        |
| >280           | 25-33.9%                        |
|                 | Above 34%                       |
apoptosis downstream of cytochrome c [58]. All of these manifestations facilitate carcinogenesis.

Characterisation of the host immune responses—including the cytokine phenotypes—to *O. viverrini* and their roles in immune/inflammatory cell-induced tissue damage, as well as their relationship with host susceptibility, will likely yield a more complete understanding of the immunopathologic process involved in liver fluke–induced CCA.

**Multi-factorial aetiology of CCA.** Liver fluke–induced CCA is therefore generally accepted to be the result of chronic inflammation [53,58,59]. Several mechanisms by which *O. viverrini* infection may enhance cholangiocarcinogenesis have been proposed above, and are summarised in Figure 5. Indeed, it is almost certain that a combination of those pathologies described above (mechanical damage, parasite secretions, and immunopathology) culminate in CCA after chronic infection with *O. viverrini*. The primary pathologic change, i.e., epithelial desquamation, may be due to mechanical irritation caused by the liver fluke and/or its metabolic products. However, immunopathologic processes may contribute to the long-standing hepatobiliary damage. During liver fluke infection, inflammation, periductal fibrosis, and proliferative responses, including epithelial hyperplasia, goblet cell metaplasia, and adenomatous hyperplasia, may represent predisposing lesions that enhance susceptibility of DNA to carcinogens [11,60]. Several N-nitroso compounds and their precursors occur at low levels in fermented food such as preserved mud fish paste, *pla-ra*, a condiment that is a ubiquitous component of the cuisine of northeastern Thailand and Laos.

Indeed, it has been hypothesised that N-nitroso compounds (e.g., nitrosamine) are a primary carcinogen leading to CCA in humans in this region [61,62]. We recently reported dissimilar gene expression profiles in CCA tissues from patients from sites endemic for *O. viverrini* in Thailand versus non-endemic areas in Japan [63]. Of particular interest was the selective up-regulation in Thai patients with CCA of genes involved in xenobiotic metabolism, implying that these genes may be involved in the detoxification of possible carcinogens such as nitrosamines [63]. CCA can be reliably induced in hamsters experimentally infected with *O. viverrini* and exposed to sub-carcinogenic doses of nitrosamine, but not in liver fluke–infected hamsters not exposed to nitrosamine [11].

Apart from exogenous carcinogens, however, endogenous nitrosation caused by liver fluke infection has also been investigated in animals and humans. Experimental *Opisthorchis* infection in hamsters can induce NO synthase expression by immune effector cells in inflamed areas surrounding the bile ducts and increased endogenous nitrosation of thiazolidine-4-carboxylic acid (thioproline) [64]. Several human studies suggest that infected people have a higher endogenous nitrosation potential than uninfected people [65,66]. Restricted diet studies clearly demonstrated an increase in endogenous generation of NO and N-nitroso compounds as indicated by increased levels of plasma, urinary nitrate and nitrite, and by nitrosation of proline and thioproline among infected men compared to members of an uninfected control group [25]. Moreover, the difference was specifically abolished by praziquantel treatment and by co-administration of ascorbic acid with proline. Enhanced immune responses to *O. viverrini* are also associated with an increase of endogenous nitrosation in infected individuals [67]. Both exogenous and in situ nitrosamine formation may lead to DNA alkylation and deamination in predisposed and inflamed tissues [49–51].

Apoptosis is rarely observed in either *O. viverrini*–infected hamster liver or in biliary cells that are co-cultured with

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**Figure 4.** Characteristics of CCA

Morphologically this cancer falls into three subtypes: mass forming, periductal infiltrating, and intraductal. This classification is independent of the location of the affected bile duct, which may be intrahepatic (within the liver) or extrahapecatic (outside the liver, excluding the gallbladder). The line between these subtypes tends to blur as the malignancy matures. Tumour dimensions, mode of spreading, and clinical manifestations are noted. Adapted from [36,37] by Giovanni Maki.

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**Table 1.** Characteristics of CCA subtypes

| CCA Subtype       | Dimensions                                                                 | Location (Intra or Extra-hepatic) | Pathology                                                                 | Method of Spread                                                                 | Symptoms of Bile Duct Obstruction? |
|-------------------|----------------------------------------------------------------------------|-----------------------------------|--------------------------------------------------------------------------|--------------------------------------------------------------------------------|-----------------------------------|
| Mass forming      | Central mass; depends on location (HI up to 15 cm; EH 1–2 cm)              | Intra-hepatic                      | • Gray white mass                                                        | • Grows outward into lumen                                                        | Symptoms occasionally occur        |
| Periductal-       | 0.5–6 cm long (up to 1 cm in the case of EH tumors)                        | Extra-hepatic                      | • Concentric thickening of bile duct wall                                 | • Invades bile duct wall                                                          | Viscous mucus produced by the tumor can impede bile flow and produce intermittent obstructive symptoms |
| growing           |                                                                            |                                    | • Later stages appear branch-like                                         | • Spreads along axis of bile ducts                                                |                                    |
|                   |                                                                            |                                    | • Usually highly differentiated                                           |                                                                                   |                                    |
| Intraductal       | Usually small and flat; later stages may fill bile duct lumen              |                                    | • Tumors within lumen                                                     | • Spreads superficially along mucosal surface                                      | Narrowing of bile ducts eventually leads to symptoms |
|                   |                                                                            |                                    | • Frond-like foldings                                                     | • Sloughing of tumor cells can initiate secondary tumors                          |                                    |
|                   |                                                                            |                                    |                                                                         | • Invasive intraductal CCA can also occur                                         |                                    |
flukes in vitro (B. Sripa et al., unpublished data). This may be explained by the mechanisms described above and/or the effect of bile acids [58]. Elevation of bile acids has also been reported in opisthorchiasis [68]. Bile acids are potent tumour promoters and have also been shown to block protein degradation of myeloid cell leukemia protein 1 (MCL-1), a potent anti-apoptotic protein of the BCL-2 family, via activation of EGFR signalling [69]. Anti-apoptosis has been described in infection-associated cancers such as those caused by *Helicobacter pylori* [70,71]. In the case of *O. viverrini* infection, DNA damage is caused in biliary epithelial cells while apoptotic mechanisms are dysregulated, resulting in genetic alterations which may become fixed, leading to malignant transformation.

**Implications for Improving Public Health**

From a public health perspective, thorough cooking of the fish hosts efficiently blocks infection with these parasites. In this regard, liver fluke–associated CCA is preventable by changes in eating habits. Unfortunately, age-old culinary preferences for uncooked dishes such as *koi-pla* (Figure 1) do not readily allow for this possibility. Moreover, fish farming of grass carp and other susceptible species in ponds that are routinely contaminated by untreated sewage has resulted in the establishment of infection in fish populations at large, which, along with the involvement of animal reservoir hosts, makes control of liver fluke infection even more challenging [13,72]. Nonetheless, given this extraordinary linkage between a metazoan parasite and a tumour, characterisation of the nature and action of carcinogens of *O. viverrini* or *C. sinensis* may provide fundamental insights into carcinogenesis at large.

In the short term, enhanced knowledge of pathogenesis, particularly inflammation and its associated DNA damage—both nitrative and oxidative—may assist in disease control, provided the at-risk populations can be educated of the...

**Figure 5. Proposed Mechanisms of *Opisthorchis*-Derived CCA Initiation**

The three proposed pathways linking the parasite to CCA initiation are mechanical damage (in yellow), molecular products (in blue), and immunopathology (in green). Combined, these mechanisms result in several common elements (purple) that all lead to DNA damage. The inhibition of a normal DNA damage response is the final oncogenic factor proposed to dramatically increase the likelihood of a malignant transformation. This invariably leads to progression of CCA. (Adapted from [10].) The photographs underneath the schematic show two livers with advanced CCA.
elevated risk of CCA from repeated infections. Also, dietary changes that include natural products rich in anti-oxidants and increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from *Opisthorchis viverrini* infection in hamsters.

### Supporting Information

**Alternate Language Summary S1.** A summary of the article translated into Thai by BS

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### Acknowledgments

The authors appreciate the generous in-kind support for the "Pathogenesis of liver fluke-induced cancer in Thailand" project provided by our institutions: Khon Kaen University, Tulane University, Queensland Institute of Medical Research, and George Washington University.

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