**Bacillus subtilis** biofilm extends *Caenorhabditis elegans* longevity through downregulation of the insulin-like signalling pathway

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Beneficial bacteria have been shown to affect host longevity, but the molecular mechanisms mediating such effects remain largely unclear. Here we show that formation of *Bacillus subtilis* biofilms increases *Caenorhabditis elegans* lifespan. Biofilm-proficient *B. subtilis* colonizes the *C. elegans* gut and extends worm lifespan more than biofilm-deficient isogenic strains. Two molecules produced by *B. subtilis* — the quorum-sensing pentapeptide CSF and nitric oxide (NO) — are sufficient to extend *C. elegans* longevity. When *B. subtilis* is cultured under biofilm-supporting conditions, the synthesis of NO and CSF is increased in comparison with their production under planktonic growth conditions. We further show that the prolongevity effect of *B. subtilis* biofilms depends on the DAF-2/DAF-16/HSF-1 signalling axis and the downregulation of the insulin-like signalling (ILS) pathway.
During the last 30 years, human life expectancy has significantly increased worldwide http://www.who.int/gho/publications/world_health_statistics/2015. Between 1990 and 2012, humans (males and females) born in low- and high-income countries have gained 9.05 and 4.25 years in life expectancy, respectively http://www.who.int/gho/publications/world_health_statistics/2015, which has been attributed to a plethora of genetic and environmental factors1,2. In mammals, the beneficial bacteria residing in the host intestine (that is, gut or commensal flora) play a crucial role in immune system development, tissue morphogenesis and aging3–5. Beneficial effects of the gut flora on host physiology, including slowing of aging, might be improved with the incorporation of probiotic bacteria into the host diet. Probiotics are live organisms that have a beneficial effect on host health when they are administered or present in adequate quantities9,10. However, the mechanisms causing potential prolongevity effects of beneficial bacteria on different hosts remain poorly understood9,10–13.

Microbial biofilms are three-dimensional structured communities of adherent microorganisms encased in a self-produced extracellular matrix, containing networks of channels for nutrient supply and long-distance cell-to-cell communication6,17. Bacteria living in biofilms are physiologically very distinct from their planktonic counterparts, and they function as a cooperative consortium more similar to that of multicellular organisms than a unicellular organism16,18,19. The crucial role of biofilms in the success of different pathogenic bacteria in infecting different hosts is well documented20, but whether biofilms influence the interaction between beneficial bacteria and their host is largely unknown10,16.

*Bacillus subtilis* is a model beneficial bacterium with the ability to display many distinct cell types under developmental control, including the ability to form robust and sophisticated biofilms21–27. Interestingly, a recent study showed that planktonic *B. subtilis* modulates the longevity of the bacteriovorous nematode and model organism *Caenorhabditis elegans* independent of its role as a food source21,28. Worms fed on planktonic *B. subtilis* cells live longer (an approximately 15% increase in lifespan) than worms grown in the presence of other food sources, that is, *E. coli*28,29. Because of the limited understanding of the effect of biofilm formed by beneficial gut bacteria on host health, we determined whether the biofilm of *B. subtilis* might represent a novel anti-aging agent for improving host longevity.

**Results and Discussion**

*C. elegans* longevity is increased by undomesticated *B. subtilis*. The various studies that have reported interactions of *B. subtilis* with *C. elegans* utilized genetically modified laboratory *B. subtilis* (domesticated) strains derived from the wild (undomesticated) Marburg isolate NCIB3610. However, these laboratory strains (that is, strains 168, JH642 and PY79; refs 30–32 are known to harbour several initially inadvertent mutations that affect collective behaviours (that is, social-surface motility and biofilm formation)31,32 that might play an important role during bacteria–host interactions.

To answer this question, we initially compared the lifespan effects produced by the wild strain NCIB3610 (ref. 31) and its isogenic derivate, the domesticated JH642 strain33 (Supplementary Table 1) on *C. elegans*. It has been noted that in contrast to the susceptibility of vegetative cells, *B. subtilis* spores survive the transit through the pharynx and germinate in the worm intestine34,35. Therefore, for all the subsequent experiments, the sporulated form of *B. subtilis* was provided as the worm food source to correlate the effects on worm lifespan with the activity of live bacteria30,21. *C. elegans* fed on domesticated *B. subtilis* cells showed an extended longevity compared with the effect of OP50 *E. coli* cells when used as a food source (Fig. 1a and Supplementary Table 2). The survival of worms fed on JH642 cells increased by an average of 23.56% (average survival, 25.02 days; 95% confidence interval (CI): 23.39–26.65) compared with strain OP50 (average survival, 20.25 days; 95% CI: 18.52–21.97) when used as a food source (Supplementary Table 2). Interestingly, the lifespan effect on *C. elegans* was significantly more pronounced with a statistically right-shifted survival curve (P = 0.00) by log-rank test when the worms were fed on undomesticated *B. subtilis* cells (strain NCIB3610) (Fig. 1a and Supplementary Table 2). In this case, NCIB3610 cells increased the worm longevity by an average of 52.59% compared with the lifespan effect produced by OP50 *E. coli* cells (average value, 30.90 days; 95% CI: 29.14–32.67) (Supplementary Table 2). The lifespan of *C. elegans* also increased by an average of 59.51% (average value, 32.30 days; 95% CI: 30.68–33.90; equality, P = 0.00 by log-rank test) when fed on another undomesticated *B. subtilis* strain (that is, the probiotic *B. subtilis* natto strain RG4365, Supplementary Table 1)22,27. The different effects of these bacterial strains on worm survival cannot be attributed to physiological differences between the domesticated and undomesticated strains because both types of bacterial cells displayed similar efficiencies of spore germination, vegetative growth and spore formation (Supplementary Fig. 1a–c). However, the possibility exists that the observed differential effects on worm survival were due to changes in the mechanical/physical properties of both types of spores. In this case, the worm grinder may not be able to efficiently break the spores of undomesticated cells, leading to higher colonization levels and/or reduced nutritional availability, which might explain the observed longevity effects (Fig. 1a). To test this possibility, we analysed and compared the robustness and resistance of JH642 and NCIB3610 spores. As shown in Supplementary Fig. 1d, spores derived from domesticated and undomesticated cells are indistinguishable one from the other in their resistance to different physico-chemical treatments such as sonication, low and high pH exposure, and lysozyme and protease treatments. Accordingly, several physiological parameters of *C. elegans*, such as its chemotactic behaviour (in response to the different bacterial food sources), growth rate, body size, postembryonic development and egg-laying proficiency were not affected when fed on any of the selected bacterial strains (that is, OP50, JH642 and NCIB3610 cells; Supplementary Fig. 1e–i). Additionally, we used FUdR in the lifespan assays which was previously shown to be a potential co-founder for *C. elegans* lifespan studies36. To test this possibility, we analysed the lifespan effects produced by *B. subtilis* NCIB3610 cells on N2 worms either in the presence or the absence of FUdR, as well as the effect of FUdR in *B. subtilis* biofilm formation proficiency. As shown in Supplementary Fig. 2a, and Supplementary Table 3, the worm lifespan increase produced by *B. subtilis* NCIB3610 cells was essentially the same in the presence (average value, 30.90 days; 95% CI: 29.14–32.67; equality, P = 0.00 by log-rank test) or absence of FUdR (average value, 31.07; 95% CI: 29.45–32.78). Accordingly, the *in vitro* and *in vivo* biofilm formation proficiency of *B. subtilis* NCIB3610 cells was not affected by FUdR (Supplementary Fig. 2b). Overall, these results rule out potential drug–bacterial strain interaction, variations in food preference and/or food availability between the different bacterial food sources as reasons for the observed differences in *C. elegans* lifespan (Fig. 1a and Supplementary Table 2).

Lifespan and stress resistance are interrelated1,2. Extended longevity of *C. elegans* correlates with enhanced worm resistance
against different biological, physical and chemical stressors. In this sense, in accordance with the increase in the lifespan effect produced by undomesticated \textit{B. subtilis}, we found a strong positive effect on the thermotolerance of \textit{C. elegans}. As shown in Fig. 1b, \textit{C. elegans} fed on NCIB3610 cells showed higher resistance and survival to heat shock compared with the protective effects produced by domesticated \textit{JH642} (using OP50 cells as a control). When \textit{C. elegans} was shifted from its routine growth temperature of 20 to 34°C, worms fed either on undomesticated or domesticated \textit{B. subtilis} cells lived 278.57% (average survival, 132.16 min) and 179.52% (average survival, 97.58 min) longer, respectively, than did worms fed on OP50 cells (average survival, 34.91 min; Fig. 1b and Supplementary Table 2). Accordingly, we did not observe any significant FUdR interference on the thermotolerance behaviour of \textit{C. elegans} when fed on NCIB3610 cells (Supplementary Fig. 2c and Supplementary Table 3).

To confirm the higher beneficial effects of undomesticated \textit{B. subtilis} cells over domesticated \textit{B. subtilis} cells on worm survival, we measured the lifespan of N2 worms subjected to injuries other than heat shock (that is, osmotic, heavy metal and oxidative stresses). In all the analysed situations, these results indicate that wild (undomesticated) \textit{B. subtilis} strains are significantly more beneficial and efficient than domesticated \textit{B. subtilis} in protecting and prolonging \textit{C. elegans} lifespan (Fig. 1, Table 1A and Supplementary Table 2).
**Table 1 | Summary of all aging experiments shown in this work.**

| C. elegans strain (genetic background) (relevant phenotype) | Bacterial strain (genetic background) (relevant phenotype) | Mean survival days ± s.e.m. | Median survival days (confidence interval with 95%) | P-value | Survival increase (%) |
|----------------------------------------------------------|----------------------------------------------------------|-----------------------------|----------------------------------------------------|---------|------------------------|
| A N2 (wt) | OP50 (wt) | 19.68 ± 0.77 | 18 (17-19) | Control | | |
| | JH642 (wt) | 25.02 ± 0.83 | 26 (24-28) | 0.0004 | 27.13 | |
| | NCIB3610 (wt) | 30.61 ± 0.93 | 32 (29-34) | 0.00 | 55.54 | |
| B N2 | NCIB3610 | 30.61 ± 0.93 | 32 (29-34) | Control | | |
| | RG3603 (ΔbslA) (deficient in biofilm formation) | 22.48 ± 0.87 | 22 (22-24) | 1E-07 | -26.56 | |
| | RG3610 (Δnos) (deficient in NO production) | 25.94 ± 1.02 | 26 (24-29) | 0.0022 | -15.26 | |
| | RG4010 (Δcsf) (deficient in CSF production) | 26.48 ± 0.92 | 27 (24-29) | 0.0085 | -13.49 | |
| | RG3611 (Δnos Δcsf) (deficient in NO and CSF production and biofilm formation) | 20.10 ± 0.79 | 21 (19-23) | 0 | -34.34 | |
| | RG3405 (Δcsf ΔbslA) (deficient in CSF production and biofilm formation) | 20.54 ± 0.85 | 22 (19-25) | 0 | -32.90 | |
| | LC3601 (Δnos Δacsf) (deficient in NO and CSF production) | 23.85 ± 0.92 | 23 (20-25) | 1E-06 | -22.08 | |
| | LC3611 (Δnos ΔbslA Δacsf) (deficient in NO and CSF production and biofilm formation) | 17.43 ± 0.86 | 18 (17-19) | 0 | -43.06 | |
| C | OP50 | 13.87 ± 0.52 | 12 (11-13) | Control | | |
| | CF1038 (daf-16/mu86) | 15.60 ± 0.68 | 12 (11-13) | 0.007 | 12.47 | |
| | JH642 | 17.36 ± 0.74 | 16 (14-17) | 0.00016 | 25.16 | |
| | NCIB3610 | 19.56 ± 0.83 | 20 (18-22) | Control | | |
| D | OP50 | 11.97 ± 0.58 | 10 (9-11) | Control | | |
| | PS3551 (hsf-1/sy441) | 13.85 ± 0.56 | 12 (11-13) | 3.5E-05 | 15.71 | |
| | JH642 | 16.58 ± 0.60 | 16 (14-17) | 0 | 40.35 | |
| | NCIB3610 | 17.68 ± 0.79 | 18 (16-20) | 0.00016 | 25.16 | |
| E | OP50 | 31.82 ± 1.68 | 25 (23-27) | Control | | |
| | CB1370 (daf-2/e1370) | 34.84 ± 1.81 | 28 (24-32) | 0.02 | 9.49 | |

Experimental conditions are described in the ‘Methods’ section. Each data set in the same line (mean survival, median survival, P value and survival increase) was calculated using the log-rank and Kaplan-Meier tests and represents the average value from all the experiments described in this work that involved the same experimental conditions and the same pair of worm-bacterial strains. Shown data are the mean values of the experimental data showed in Figs 1a and b; Figs 2a and b. Median survival (confidence interval with 95%) was calculated using the log-rank and Kaplan-Meier tests and represents the average value from all the experiments described in this work that involved the same experimental conditions and the same pair of worm-bacterial strains. Shown data are the mean values of the experimental data showed in Figs 1a and b; Figs 2a and b.

**C. elegans longevity depends on B. subtilis gut colonization.** To shed light on the reasons for the differences in the magnitude of the lifespan effects produced by domesticated and undomesticated B. subtilis, we analysed the ability of each cell type to grow and colonize the worm intestine. Our initial approach was to measure the intestinal activity of a reporter gene harboured by the bacterium used as the worm food source. Therefore, we fed C. elegans on NCIB3610 and JH642 cells that harboured a reporter of bacterial gut colonization (number of B. subtilis cells/worm). To distinguish between these possibilities, we measured the germination rates, the outgrowth and the β-galactosidase expression driven by the P<sub>srf-lacZ</sub> fusion in domesticated and undomesticated B. subtilis cells inoculated in nematode growth medium (NGM). As shown in Supplementary Fig. 3, the kinetics of spore germination, the vegetative growth rates and the β-galactosidase expression from the reporter P<sub>srf-lacZ</sub> fusion of cells grown in liquid and solid NGM did not significantly differ between the domesticated and undomesticated B. subtilis cells (Supplementary Fig. 3a–c). These results suggest that the different levels of intestinal β-galactosidase activity derived from the two types of B. subtilis cells (Fig. 2a) were due to the different levels of bacterial worm gut colonization.

Similarly, the fluorescence signal originated from the gut of worms fed on undomesticated B. subtilis cells harbouring a reporter gene of GFP expression (bslA-gfp) (Supplementary Table 1) was significantly higher than the originated from worms fed on domesticated B. subtilis cells (Fig. 2b). The observed differences in GFP expression were not due to differences in the growth rate or other physiological parameter between both types of B. subtilis strains because the GFP expression levels displayed by both types of cells were indistinguishable from one another under planktonic growth conditions on NGM agar plates (Supplementary Fig. 3d). Therefore, these results demonstrate the proficiency of B. subtilis, as a commensal organism, to complete an entire cell cycle in...
Figure 2 | Proficiency of \textit{B. subtilis} gut colonization in \textit{C. elegans} intestine. (\textbf{a}) Expression of surfactin operon in worm intestine. Thirty late L4/young-adult-stage worms were allowed to develop on NGM agar plates seeded with OP50 \textit{E. coli}, washed and transferred to fresh NGM plates containing JH642- or NCIB3610-derived isogenic RG4243 (red circles) and RG4220 (blue circles) strains, respectively (Supplementary Table 1) harbouring the \(P_{\text{tapA-sipW-tasA}}\) reporter gene. Determination of \(\beta\)-galactosidase activity was performed in worm extracts. The \(\beta\)-galactosidase activity, expressed as Miller units (MU) over time (h) is shown. ***\(P<0.001\) (analysis of variance (ANOVA) with Bonferroni test). (\textbf{b}) \textit{B. subtilis} biofilm formation in \textit{C. elegans} gut. Thirty late L4/young-adult-stage worms were allowed to develop on NGM agar plates seeded with OP50 \textit{E. coli}, washed and transferred to fresh NGM plates containing NCIB3610- or JH642-derived isogenic (RG4225 (blue) and RG4223 (red) strains, respectively, carrying a \(bslA::gfp\) reporter fusion (Supplementary Table 1)). The worms were taken and imaged by fluorescence microscopy. Fluorescence intensity is indicated as arbitrary units (A.U.) per worm, and error bars show the mean ± (n = 3) ***\(P<0.001\) (ANOVA with Bonferroni test). The inserts shown are epifluorescence micrographs of typically stained N2 \textit{C. elegans} fed on GFP-expressing domesticated and undomesticated \textit{B. subtilis}. The fluorescence images were superimposed to differential interference contrast (DIC) images to depict the localization of the labels within the cells. Scale bar, 20 μm. (\textbf{c}) Worm intestine colonization by \textit{B. subtilis}. Hundred late L4/young-adult-stage worms were allowed to develop on NGM agar plates seeded with OP50 \textit{E. coli}, washed, transferred to fresh NGM plates containing new OP50 cells (green) or spores of the strains JH642 (red) or NCIB3610 (blue). The number of \textit{E. coli} or \textit{B. subtilis} cells (spores and vegetative forms) was measured in the worm gut. Error bars show the mean ± (n = 3). ***\(P<0.001\) (ANOVA with Bonferroni test).

The biofilm is key to colonize \textit{C. elegans} and extend its lifespan. The ability to adhere and form a multicellular structure called a pellicle or biofilm constitutes an attribute of bacteria that is key to their success in colonization and persistence in a particular niche\textsuperscript{16–20}. In studies of host–bacteria interactions, the biofilm formation proficiency of both pathogenic and beneficial bacteria is believed to play a crucial role\textsuperscript{44–52}. Because of the significant difference between domesticated and undomesticated \textit{B. subtilis} strains in their biofilm formation proficiency\textsuperscript{24,31}, we wanted to analyse whether the observed differences in worm longevity and gut colonization produced by \textit{B. subtilis} were due to the different abilities of the bacterial strains to establish a biofilm in the worm intestinal environment. The \(tapA\)-\textit{sipW-tasA} and \textit{epsA-G} operons encode for two key components of the extracellular matrix of the biofilm, the \(tasA\) protein and the EPS exopolysaccharide, respectively\textsuperscript{25,26,53}. In addition, the \(bslA\) gene encodes for an essential hydrophobin responsible for the formation of the hydrophobic surface layer that surrounds and protects the biofilm\textsuperscript{54}. Therefore, to test our hypothesis, we introduced
significance differences between the numbers of B. subtilis spores of the wild-type NCIB3610 (blue) or a mixture of the three mutants deficient in biofilm formation. The worms were processed to measure the formation. Thirty L4 worms were allowed to develop on NGM agar plates seeded with OP50 cells, washed and transferred to fresh NGM plates containing A representative experiment ± were fed either on NCIB3610 (blue) or a mixture of the three B. subtilis mutants deficient in biofilm formation (purple) and worm survival was monitored. (a) Biofilm formation proficiency. Biofilms of undomesticated B. subtilis strains harbouring mutations in genes encoding for essential components of the biofilm matrix (ΔepsG, ΔtasA or ΔbslA) were developed in liquid NGM. (b) Worm longevity is affected when the synthesis of components of the biofilm matrix is reduced. Fifty L4 worms were fed on spores of undomesticated B. subtilis deficient in components of the biofilm matrix (ΔepsG, grey), (ΔtasA, orange) or (ΔbslA, yellow) on NGM agar plates, and the survival was monitored. The figure shows the survival average ± s.e.m. (n = 3). (c) Colonization of the worm gut by B. subtilis strains deficient in biofilm formation. Thirty L4 worms were allowed to develop on NGM agar plates seeded with OP50 cells, washed and transferred to fresh NGM plates containing spores of the wild-type NCIB3610 (blue) or spores of each one of the three mutants deficient in biofilm formation. Worms were processed to measure the number of B. subtilis cells (spores and vegetative forms) in the worm gut. Error bars show the mean ± s.e.m. (n = 3) ***P < 0.001 (analysis of variance (ANOVA) with Bonferroni test). (d) Transcomplementation of a mixture of biofilm-deficient B. subtilis strains to restore C. elegans lifespan. Thirty L4 worms were fed either on NCIB3610 (blue) or a mixture of the three B. subtilis mutants deficient in biofilm formation (purple) and worm survival was monitored. A representative experiment ± s.e.m. is shown. (e) Recolonization of the worm gut by a proportional mixture of B. subtilis strains deficient in biofilm formation. Thirty L4 worms were allowed to develop on NGM agar plates seeded with OP50 cells, washed and transferred to fresh NGM plates containing spores of the wild-type NCIB3610 (blue) or a mixture of the three mutants deficient in biofilm formation. The worms were processed to measure the number of B. subtilis cells in the gut worm. Error bars show the mean ± s.e.m. (n = 3) ***P < 0.001 (ANOVA with Bonferroni test). NS denotes no significance differences between the numbers of B. subtilis spores counted.

mutations in bslA, epsG and tasA genes of the undomesticated strain NCIB3610 to obtain biofilm-defective derivates (Supplementary Table 1). When each of the three undomesticated-derived B. subtilis mutant strains defective in biofilm formation (ΔtasA, ΔepsG and ΔbslA; Fig. 3a) were separately used to feed C. elegans, there was a significant decrease in lifespan (an average decrease of 30%) compared with C. elegans fed on the biofilm-proficient wild-type strain NCIB3610 (Fig. 3b and Supplementary Table 5). For instance, while C. elegans fed on the wild-type NCIB3610 strain showed an increase in longevity of 58.73% (Supplementary Table 5), worms fed on the biofilm-deficient ΔbslA strain (RG3603, Supplementary Table 1) only increased longevity by an average of 19.10% (average value, 23.26 days; 95% CI: 21.16–25.36; equality, P = 0.01 by log-rank test) compared with the lifespan effect produced by OP50 E. coli cells (average value, 19.53 days; 95% CI: 17.70–21.36; equality; Supplementary Table 5).

The observed decrease in the worm lifespan was not associated with a diminished ability of the biofilm-mutant strain
The biofilm and nitric oxide produce synergetic anti-aging effects. The beneficial effect of domesticated *B. subtilis* over OP50 on *C. elegans* longevity has been recently attributed to the bacterium’s nitric oxide (NO) production proficiency. NO-mediated signalling plays key roles in all living organisms, and for an unknown reason, *C. elegans* is unable to produce its own NO but is able to incorporate the NO produced by *B. subtilis*. Most organisms produce NO through aerobic conversion of l-arginine to l-citrulline in a reaction catalysed by the enzyme NO synthetase encoded by the nos gene. *E. coli* strains, several of which are routinely used to feed the worms (that is, OP50, HB101; refs 14,15,29), are not proficient in aerobic NO production because they lack a functional copy of nos. However, *E. coli* can produce NO under anaerobic/microaerophilic conditions by a series of biochemical reactions associated with the anaerobic respiratory chain of the bacterium (Supplementary Table 6 and ref. 56). Under this scenario, *E. coli* might find permissive conditions for NO production in the oxygen-depleted environment of the worm intestine; therefore, the positive effect of *B. subtilis* on worm longevity could be due to at least one as-yet unknown additional factor apart from the intestinal NO provision. First, we were interested in determining whether the biofilm proficiency-dependent lifespan effect of *B. subtilis* on *C. elegans* was due to NO production. As shown in Fig. 4a, the mutations affecting biofilm formation (*ΔbslA* mutant) or NO synthesis (*Δnos* mutant) (Supplementary Table 1) produced a negative effect on *C. elegans* lifespan compared with the lifespan effect produced by the wild-type NCIB3610 strain. Interestingly, the negative effect on worm longevity was more pronounced when *C. elegans* was fed *B. subtilis* cells deficient in biofilm formation (but proficient in NO production; average value, 22.03 days; 95% CI: 20.13–23.94; equality, *P* = 0.04 by log-rank test) compared with *B. subtilis* cells deficient in NO production (but proficient in biofilm formation; average value, 25.55 days; 95% CI: 23.31–27.79; equality, *P* = 2.5 × 10^-7 by log-rank test; Supplementary Table 7). The average worm lifespan increases of 29.17 and 11.38% were observed when the worms were fed on *B. subtilis* cells defective in NO or biofilm production, respectively, in comparison with the 53.13% lifespan increase (related to OP50 cells) obtained when the worms were fed on wild-type *B. subtilis* (average value, 30.29 days; 95% CI: 28.26–32.33; equality, *P* = 0.00 by log-rank; Fig. 4a and Supplementary Table 7). Furthermore, the double mutant of *B. subtilis* deficient in biofilm formation and NO synthesis (*Δnos-ΔbslA*; Supplementary Table 1) produced the most negative impact on worm survival (average value, 20.06 days; 95% CI: 18.52–21.62; equality, *P* = 0.67 by log-rank test) with a final increase in worm survival of only 1.42% compared with OP50 cells (Fig. 4a and Supplementary Table 7). In addition, spor e robustness and worm chemotactic behaviour toward *Δnos* (60.0 ± 2.0% of *Δnos* spore survival after 30 s of treatment at pH 2.0 and chemotaxis index of 0.95 ± 0.2, respectively) and *Δnos-ΔbslA* (59.4 ± 1.9% of *Δnos-ΔbslA* spore survival after 30 s of treatment at pH 2.0 and chemotaxis index of 0.91 ± 0.3, respectively) cells were not significantly different compared with the NCIB3610 strain. The different effects on *C. elegans* lifespan correlated with the capacity of each *B. subtilis* strain to form a biofilm (Fig. 4b), produce NO (Fig. 4c) and colonize the worm intestine (Fig. 4d). Thus, the positive lifespan effects of NO production and biofilm formation proficiency are different and synergistic.

If NO production and biofilm formation are distinct but complementary processes improving the longevity of *C. elegans*, then we hypothesize that a mixture of mutant *B. subtilis* cells, that is, cells deficient in biofilm formation (*ΔbslA*) and cells deficient in NO synthesis (*Δnos*), would complement each other and restore the full lifespan effect of wild-type NCIB3610 cells on *C. elegans*. As predicted by this hypothesis, a mixture of *ΔbslA* and *Δnos* cells (strains RG3610 plus RG3603, light blue curve in Fig. 4a) restored the worm lifespan to levels comparable to the effect of wild-type NCIB3610 cells (average value, 29.51 days; 95% CI: 27.77–31.75; equality, *P* = 0.0141 by log-rank test) compared with OP50 cells (final increase in worm survival, 49.19%; Fig. 4a and Supplementary Table 7).

Overall, these results demonstrate that biofilm formation proficiency constitutes a new attribute, different from NO production, for the anti-aging effect of *B. subtilis* on *C. elegans* (Table 1B). The discovery that NO production is not the only factor that explains how *B. subtilis* extends the longevity of *C. elegans* opens the possibility of the existence of other factors that positively contributes to increase worm longevity.

**B. subtilis** quorum sensing extends worm longevity. Intra- and interspecific quorum sensing (QS) constitutes a widely used...
strategy that bacteria use in nature to communicate with each other and with cells of different kingdoms. Recent studies have shown the potential effect of bacterial QS on C. elegans physiology. C. elegans recognized Vibrio cholerae QS signalling molecules through the AWC

Figure 4 | Biofilm formation and nitric oxide production are required to extend C. elegans lifespan. (a) Effects upon longevity of worms fed on undomesticated B. subtilis defective in biofilm production and/or NO synthesis. Fifty L4 worms were fed either on spores of undomesticated NCIB3610 (blue) or RG3610 (Δnos, defective in NO production, orange) or RG3603 (ΔbslA, defective in biofilm synthesis, yellow) or RG3611 (ΔnosΔbslA, defective both in NO production and biofilm synthesis, grey) or a mixture of RG3610 and RG3603, light blue) B. subtilis on NGM agar plates, and survival was monitored. A representative experiment ± s.e.m. is shown. (b) Biofilm formation proficiencies of undomesticated B. subtilis strains harbouring mutations in genes essential for biofilm formation and/or NO production. Biofilms were developed in liquid NGM. (c) NO quantification of undomesticated B. subtilis strains harbouring mutations in genes essential for biofilm formation and/or NO production. The nitrite and nitrate concentrations are expressed as μM. Error bars show the mean ± s.e.m. (n = 3). **P < 0.001 and *P < 0.05 (analysis of variance (ANOVA) with Bonferroni test). (d) Colonization of the worm gut by B. subtilis strains deficient in biofilm formation and/or NO production. L4 worms were allowed to develop on NGM agar plates seeded with OP50 cells, washed and transferred to fresh NGM plates containing spores of the wild-type NCIB3610 (blue) or RG3610 (Δnos, defective in NO production, orange) or RG3603 (ΔbslA, defective in biofilm synthesis, yellow) or RG3611 (ΔnosΔbslA, defective both in NO production and biofilm synthesis, grey) B. subtilis strains, respectively. The worms were taken and processed to measure the number of B. subtilis cells in the worm gut, expressed as CFUs per worm. Error bars show the mean ± s.e.m. (n = 3). **P < 0.001, *P < 0.01 and +P < 0.05 (ANOVA with Bonferroni test). Images shown on the bottom are epifluorescence micrographs of typically stained N2 C. elegans fed epsA:γfp-expressing B. subtilis strains (RG3620, wild-type; RG3621, Δnos; RG3622, ΔbslA; and RG3623, ΔnosΔbslA). The fluorescence images were superimposed to differential interference contrast (DIC) images to depict the localization of the labels within the cells. Scale bar, 20 μm.

epithelial cells. Because an orthologue of octn2 is present in C. elegans, we were interested in determining whether the CSF molecule affects the lifespan of C. elegans. As shown in Fig. 5, C. elegans fed on NCIB3610-isogenic B. subtilis strain deficient in CSF synthesis (Δcsf mutant, strain RG4010; Supplementary Table 1) showed a decreased lifespan compared with worms fed on wild-type NCIB3610 cells. In this case, worms fed on Δcsf cells showed an increase in survival of 29.89% (average value, 26.16 days; 95% CI: 24.41–27.91 days; equality, P = 0.04 by log-rank test), compared with OP50 cells (average value, 20.14 days; 95% CI: 18.53–21.75; Fig. 5a and Supplementary Table 8).
26.16 days (95% CI: 24.41–27.91 days; equality, \( P = 0.003 \) by log-rank test) and 29.68 days (95% CI: 27.59–31.77 days; equality, \( P = 0.004 \) by log-rank test) were obtained with wild-type \( B. \ subtilis \) cells (deficient in biofilm formation and NO synthesis) and \( \Delta \text{cfs} \) cells (deficient in CSF synthesis) to feed the worm (average worm survival obtained with each strain individually used as food: 20.06 days and 26.16 days, respectively; Supplementary Tables 7,8) complemented the deficiency of each mutant strain and re-established the lifespan of \( C. \ elegans \) to levels comparable to the lifespan effect produced by wild-type \( B. \ subtilis \) cells. Average worm survival values of 30.74 days (95% CI: 28.9–32.58 days; equality, \( P = 0.001 \) by log-rank test) and 26.16 days (95% CI: 24.41–27.91 days; equality, \( P = 0.04 \) by log-rank test) were obtained with wild-type cells and mixture of \( \Delta \text{bslA} \Delta \text{nos} \) plus \( \Delta \text{cfs} \) cells, respectively (Fig. 5a and Supplementary Table 8). Accordingly, the use of FUdR did not interfere with the effects produced by the \( \Delta \text{bslA} \), \( \Delta \text{nos} \) and \( \Delta \text{cfs} \) \( B. \ subtilis \) mutant strains on \( C. \ elegans \) survival (Supplementary Table 9).

To confirm that CSF production constitutes a novel pro-longevity factor for \( C. \ elegans \), we supplemented the worm diet with pure CSF (100 nM) in Petro dishes containing OP50 (Fig. 5b) or LC3611 (\( \Delta \text{bslA} \Delta \text{nos} \Delta \text{cfs} \); Fig. 5c) bacterial cells as a worm food source. Interestingly, the added CSF increased the lifespan of \( C. \ elegans \) fed on OP50 cells, with average survival values of 18.68 days (95% CI: 17.44–19.92 days) and 23.98 days (95% CI: 22.14–25.85 days; equality, \( P = 0.0001 \) by log-rank test) in the absence or presence of CSF supplementation, respectively (Fig. 5b and Supplementary Table 8). Similarly, the addition of CSF to \( C. \ elegans \) fed on the \( B. \ subtilis \) mutant strain LC3611 (\( \Delta \text{bslA} \Delta \text{nos} \Delta \text{cfs} \)) increased the worm lifespan to levels even higher than those obtained when \( \Delta \text{bslA} \Delta \text{nos} \) cells (strain RG3611, proficient in CSF production) were used as a food source (Fig. 5c). Average worm survival values of 16.06 days (95% CI: 13.93–18.19 days), 17.52 days (95% CI: 15.55–19.49 days; equality, \( P = 0.15 \) by log-rank test) and 20.02 days (95% CI: 18.48–21.56 days; equality, \( P = 0.55 \) by log-rank test) were obtained with the \( \Delta \text{bslA} \Delta \text{nos} \Delta \text{cfs} \), \( \Delta \text{bslA} \Delta \text{nos} \) and \( \Delta \text{bslA} \Delta \text{nos} \Delta \text{cfs} \) (plus CSF) cells, respectively (Fig. 5c and Supplementary Table 8). In sum, these results confirm the QS molecule CSF as the third factor that contributes to the lifespan extension effect of \( B. \ subtilis \) on \( C. \ elegans \) (Table 1B).

In addition, these results strongly suggested that the positive lifespan effect of \( B. \ subtilis \) on \( C. \ elegans \) is primarily, or exclusively, dependent on the three properties studied here (that is, NO, CSF and biofilm production), which produced positive synergic effects on worm survival.

Using Cox proportional hazard model (Supplementary Table 10), we confirmed that the biofilm formation proficiency (absence or presence of \( \text{bslA} \) activity) constitutes the more important \( B. \ subtilis \) property responsible for extending worm longevity. Worms fed on biofilm-deficient \( B. \ subtilis \) have a risk

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**Figure 5** | The quorum-sensing factor of \( B. \ subtilis \) (CSF) extends \( C. \ elegans \) longevity. (a) Synergic effects of biofilm formation proficiency (\( \Delta \text{bslA} \)), NO production (\( \Delta \text{nos} \)) and CSF synthesis (\( \Delta \text{cfs} \)) on \( C. \ elegans \) longevity. Late L4/young-adult-stage N2 worms were fed either on spores of undermithicic NCIB3610 (blue) or RG4010 (\( \Delta \text{cfs} \), defective in CSF production, orange) or LC3601 (\( \Delta \text{nos}\Delta \text{cfs} \), defective in NO and CSF production, yellow) or LC3611 (\( \Delta \text{nos}\Delta \text{bslA}\Delta \text{cfs} \), defective in NO production, biofilm synthesis and CSF production, light blue) or a mixture of RG3611and RG4010 (red) \( B. \ subtilis \) on NGM agar plates, and survival was monitored for signs of life. The data shown are the average \( \pm \) s.e.m. (\( n = 3 \)).

(b) Exogenous CSF peptide increased the longevity of worms fed OP50 \( E. \ coli \). Late L4/young-adult-stage N2 worms were fed either on OP50 \( E. \ coli \) (green) or OP50 \( E. \ coli \) plus 100 nM CSF peptide (red) on NGM agar plates, and the survival was monitored for signs of life. The data shown are the average \( \pm \) s.e.m. (\( n = 3 \)), and the survival increase is expressed as a percentage of the total number of worms fed OP50 \( E. \ coli \).

(c) Exogenous CSF peptide restored the longevity of worms fed \( B. \ subtilis \) deficient in QS. Late L4/young-adult-stage N2 worms were fed either on spores RG3611 (\( \Delta \text{nos}\Delta \text{bslA} \), defective both in NO production and biofilm synthesis, red) or LC3611 (\( \Delta \text{nos}\Delta \text{bslA}\Delta \text{cfs} \), defective in NO production, biofilm synthesis and CSF production, light blue) or LC3611 plus 100 nM CSF peptide (green) \( B. \ subtilis \) on NGM agar plates, and survival was monitored for signs of life. A representative experiment \( \pm \) s.e.m., and the survival increase as a percentage of the total number of worms fed on LC3611 cells is shown.
The cell-to-cell communication and division of labour that occurs inside the biofilm produce significant differences in gene expression compared with the gene expression pattern of cells grown under planktonic conditions\(^{16,19,23,58}\). In this sense, the levels of NO and CSF produced in \( \textit{B. subtilis} \) cultures have been measured only under planktonic growth conditions\(^{82}\). To determine whether NO and/or CSF production is affected in \( \textit{B. subtilis} \) cells that develop as a biofilm, we grew NCIB3610 cells under planktonic and biofilm supporting conditions and measured the production levels of the two longevity molecules. As shown, the production levels of NO (Table 2) and CSF (Table 2) were significantly enhanced when \( \textit{B. subtilis} \) was grown as a biofilm compared with the levels of NO and CSF produced under planktonic conditions. These results (Table 2 and Figs 2 and 3) reinforce the importance of the biofilm formation proficiency in the longevity of \( \textit{C. elegans} \).

The results shown in Table 2 also suggest that \( \textit{B. subtilis} \) viability inside the worm gut would be an important prerequisite for the prolongevity effect. Only alive (and not dead) vegetative \( \textit{B. subtilis} \) cells, originated from worm gut germinated spores, are able to form a biofilm and enhance production of the anti-aging molecules NO and CSF. If this interpretation is correct, there should be a difference in the magnitude of the prolongevity effect between worms fed on alive or dead \( \textit{B. subtilis} \) cells. To test this hypothesis, we fed \( \textit{C. elegans} \) on dead and alive \( \textit{B. subtilis} \) spores to measure the magnitude of the produced lifespan. Interestingly, \( \textit{B. subtilis} \) must be alive in order to produce its beneficial prolongevity effect (Supplementary Fig. 6 and Supplementary Table 11). Dead \( \textit{B. subtilis} \) used to feed \( \textit{C. elegans} \) produced a lower (\(-14.32.0\%\) lifespan increase (average survival, 25.79 days; 95% CI: 24.22–27.36 days; equality, \( P = 0.000002 \) by log-rank test) than alive \( \textit{B. subtilis} \) (average survival, 30.10 days; 95% CI: 25.35–29.29 days) (Supplementary Fig. 6 and Supplementary Table 11).

Accordingly, dead pathogenic bacteria (that is, killed \( \textit{P. aeruginosa} \)) used to feed \( \textit{C. elegans} \) increase worm survival in comparison with live pathogenic bacteria used as a food source. In addition, \( \textit{E. coli} \) strains, including the ‘non-virulent’ OP50 strain, are mildly pathogenic to aging \( \textit{C. elegans} \)\(^{14,15,29}\). We found that dead OP50 cells effectively prolonged \( \textit{C. elegans} \) survival (21% lifespan increase), with average survival of 24.73 days (95% CI: 23.23–26.23 days; equality, \( P = 2.4 \times 10^{-7} \) by log-rank test) in comparison with live OP50 cells (average survival, 20.51 days; 95% CI: 19.35–21.67 days) used as food sources. Overall, these results confirmed that \( \textit{B. subtilis} \) constitutes a beneficial flora for \( \textit{C. elegans} \) and showed that the prolongevity effect of this bacterium depends on its viability inside the worm gut.

\( \textit{B. subtilis} \) downregulates the insulin-like signalling pathway. The lifespan of \( \textit{C. elegans} \) is subject to regulation by conserved signalling pathways and transcription factors that sense stress, environmental cues and nutrient availability\(^{29}\). Dietary restriction (DR) and the insulin-like signalling (ILS) pathway are central for the regulation of longevity in different animal models, including \( \textit{C. elegans} \)\(^{1,2,65–68}\). These longevity regulatory pathways

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**Table 2 | The \( \textit{B. subtilis} \) biofilm enhances the production of the anti-aging molecules NO and CSF.**

| Media | Growth condition | NO production \((\text{NO}_2 + \text{NO}_3) \mu\text{mol per 1.0} \times 10^8 \text{ cells}\) | \( \text{csf–} \text{lacZ} \) activity (Miller Units per 1.0 \times 10^8 \text{ cells}) |
|-------|-----------------|--------------------------------|---------------------------------|
| M3gg  | Planktonic      | 55.60 ± 0.82                  | 160.45 ± 8.3                   |
| M3gg  | Biofilm         | 257.60 ± 25.20                | 540.50 ± 21.1                  |
| NGM   | Planktonic      | 180.00 ± 19.10                | 225.83 ± 15.2                  |
| NGM   | Biofilm         | 1,160.00 ± 45.20              | 1,740.15 ± 75.1                |

Biofilms of the wild-type undomesticated strain NCIB3610 were developed in M3gg and NGM as indicated in the ‘Methods’ section. After incubation for 48 h at 25 °C, the developed biofilms were disrupted as described previously\(^{23,24}\) and assayed for their NO-derivate species (nitrites and nitrates) content\(^{28}\) and CSF production levels, which are measured as levels of β-galactosidase activity driven by the promoter of the \( \text{csf} \) gene\(^{42}\). The average values of at least three independent experiments ± s.e.m. are presented.

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**Figure 6 | The prolongevity effects of \( \textit{B. subtilis} \) require the activity of the anti-aging factors DAF-16 and HSF-1.** Undomesticated and domesticated \( \textit{B. subtilis} \) cells (NCIB3610 and JH642, respectively) partially extend the lifespan of \( \text{daf-16} \) and \( \text{hsf-1} \) worms compared with their lifespan effect produced on \( \textit{N2} \) worms. Late \( \text{L4/young-adult-stage} \) \( \text{daf-16} \) (a) or \( \text{hsf-1} \) (b) worms were fed either on spores of undomesticated NCIB3610 (blue) or domesticated JH642 (red) \( \textit{B. subtilis} \) on \( \textit{NGM} \) agar plates, and the survival was monitored for signs of life. Each graph is the average ± s.e.m. \((n = 3)\), and the survival increase is expressed as a percentage of the total number of worms fed on OP50 \( \textit{E. coli} \) (green).

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The \( \textit{B. subtilis} \) biofilm enhances the production of NO and CSF. The result of death 2.45-fold higher than worms fed on biofilm-proficient cells (Supplementary Table 10). Interestingly, this statistical analysis showed that the effects of NO and CSF on lifespan were not additive and independent one from the other but showed the existence of a genetic dependence between \( \text{nos} \) and \( \text{csf} \). Up to now, we do not know the molecular reason for this bacterial gene interaction in the worm gut environment but it shows up the complexity of the mechanism responsible for the extended host longevity. We are currently investigating this interesting finding.
(DR and ILS) converge on the positive and negative regulation of the transcription factors DAF-16 and HSF-1, respectively.

To determine how B. subtilis affects the signalling pathways involved in the regulation of C. elegans longevity, we fed daf-16 (mu86) and hsf-1 (sy441) mutant worms (strains CF1038 and PS3551, respectively, Supplementary Table 1) with wild-type B. subtilis and isogenic mutants showing a decreased effect on wild-type N2 worm lifespan (ΔbslA, Δnos and Δcsf cells). Interestingly, the lifespan extension effect of wild-type B. subtilis on daf-16 and hsf-1 worms was approximately half of the lifespan extension effect on wild-type N2 worms (Fig. 6a,b and Supplementary Table 12). While the average lifespan extension effect of NCIB3610 and JH642 cells (compared with OP50 cells) on wild-type (N2) worms was 55.54 and 27.13%, respectively (Table 1A), in daf-16 worms, these percentages were 23.79 and 12.47%, respectively, compared with the use of wild-type NCIB3610 (Table 1C,D).

Similarly, the lifespan extension effect on hsf-1 worms fed on NCIB3610 or JH642 cells (Fig. 6b) was partial in comparison with the lifespan extension effect on wild-type worms fed the same type of B. subtilis cells. In this case, the average lifespan extension effect produced by NCIB3610 and JH642 cells (compared with OP50 cells) on hsf-1 worms was 35.76% and 15.71%, respectively (Fig. 6b and Supplementary Table 12). Because HSF-1 and DAF-16 are active in daf-16 and hsf-1 worms, respectively, our results suggest that the prolongevity effect of B. subtilis on C. elegans depends on the activity of DAF-16 and HSF-1 (Table 1C,D).

How does each B. subtilis prolongevity property depend on DAF-16 and/or HSF-1 activities? To answer this key question, we measured the survival of N2, daf-16 and hsf-1 worms fed on NCIB3610-isogenic mutants defective in biofilm (ΔbslA), NO (Δnos) and CSF (Δcsf) production (Fig. 7). The abolishment of each of the B. subtilis prolongevity properties was responsible for partial lifespan decreases of 26.35, 12.52 and 10.96% for wild-type N2 worms fed on ΔbslA-, Δnos- and Δcsf- B. subtilis cells, respectively, compared with the use of wild-type NCIB3610 cells (Fig. 7a and Supplementary Table 13). Accordingly, the survival of daf-16 and hsf-1 worms fed on biofilm-deficient (ΔbslA) B. subtilis cells showed a lifespan decrease of 15.04% (average value, 14.91 days; 95% CI: 14.04–15.78 days) and 15.16% (average value, 14.72 days; 95% CI: 13.65–15.79 days), respectively, compared with the average lifespan obtained with wild-type NCIB3610 cells used to fed daf-16 (average value, 17.55 days; 95% CI: 16.48–18.62 days) and hsf-1 worms (average value, 17.35 days; 95% CI: 16.18–18.52 days) (Fig. 7b,c and Supplementary Table 13). Similarly, the average worm survival percentage decreased by 9.69% (average value, 15.85 days; 95% CI: 14.67–17.03 days) and 6.57% (average value, 16.21 days; 95% CI: 14.98–17.44 days) in daf-16 and hsf-1 worms, respectively, when fed on B. subtilis cells defective in NO synthesis (Δnos) in comparison with feeding daf-16 and hsf-1 worms wild-type NCIB3610 cells (Fig. 7b,c and Supplementary Table 13). Finally, survival decreased by 6.72% (average value, 16.37 days; 95% CI: 15.20–17.54 days) and 4.61% (average value, 16.55 days; 95% CI: 15.35–17.75 days) in daf-16 and hsf-1- worms, respectively, when fed on Δcsf B. subtilis cells compared with feeding wild-type NCIB3610 cells to daf-16 and hsf-1 worms (Fig. 7b,c and Supplementary Table 13). In addition, we confirmed that the lifespan effects produced by B. subtilis NCIB3610 and its isogenic derivatives (Δnos, ΔbslA and Δcsf) on wild-type daf-16 and hsf-1 worms were not affected by the inclusion of FUdR in the assays (Supplementary Table 9). In sum, these results indicated that each of the three anti-aging properties of B. subtilis require the activity of HSF-1 and DAF-16 (ref. 69; Fig. 7 and Supplementary Tables 12,13) and reinforced the former conclusion suggesting that the biofilm formation proficiency of B. subtilis is the most important property causing the beneficial effect on C. elegans longevity.

As mentioned, host longevity is mainly regulated by the signalling pathways of DR and the nutrient-sensing pathway ILS that converge in the regulation of DAF-16 and HSF-1 activities. Interestingly, ILS could mediate part of the DR prolongevity effect because DR downregulates the activity of ILS1,2. Therefore, we tested the putative participation of the ILS pathway in the prolongevity effect of B. subtilis on C. elegans. DAF-2 is the C. elegans homologue of the insulin-like receptor

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**Figure 7 | Dissecting the contribution of each B. subtilis anti-aging property on worm survival pathways.** Late L4/young-adult-stage N2 (a), daf-16 (b) or hsf-1 (c) worms were fed either on spores of RG3610 (Δnos, defective in NO production, red) or RG3603 (ΔbslA, defective in biofilm synthesis, yellow) or RG4010 (Δcsf, defective in CSF production, green) B. subtilis on NGM agar plates, and the survival was monitored for signs of life. Undomesticated NCIB3610 B. subtilis (blue) was used as a control. A representative experiment ± s.e.m. (n = 3) is shown, and the survival increase is expressed as a percentage of the total number of worms fed on NCIB3610 cells (blue).
(IGFR), a transmembrane receptor that negatively regulates DAF-16 and HSF-1 (ref. 69). Loss-of-function mutations affecting DAF-2 have been shown to increase longevity and resistance to heat stress68. Interestingly, most of the positive lifespan effects of *B. subtilis* on wild-type *C. elegans* disappeared when this bacterium was used to feed *daf-2* (e1370) worms. Average values of 31.82 days (95% CI: 28.84–35.10 days) and 34.84 days (95% CI: 31.26–38.39 days) were obtained when *daf-2* worms were fed on OP50 and NCIB3610 cells, respectively (Fig. 8a and Supplementary Table 14). These lifespan values represent a 9.49% increase in the longevity of NCIB3610-fed *daf-2* worms (Fig. 8a) versus a 55.54% average increase in the longevity of NCIB3610-fed N2-worms compared with OP50 cells used to feed *daf-2* and N2 worms, respectively (Table 1A,E). These results strongly indicated that the anti-aging effect of *B. subtilis* on *C. elegans* is mainly transduced through the ILS pathway.

Aging is a multifactorial and poorly understood process characterized by progressive impairment of the host response to stresses and general cellular deterioration of key metabolic pathways1,2. Intensive work in different animal models (that is, yeasts, worms, flies, mice and monkeys) has identified a number of factors that promote longevity. Restricting food intake (DR), decreasing insulin/IGF-1 signalling (IIL), slowing mitochondrial respiration, reducing germline function or lowering temperature can all extend lifespan1,2. In addition, probiotics are associated with a broad spectrum of positive effects on host health, including positive effects on host longevity9–15,21,22.

To our knowledge, the present study is the first to formulate and address the question of how the bacterial biofilm might affect the prolongevity effect of beneficial gut bacteria on the host. Our evidence directly implicates the biofilm formation proficiency as the primary cause of the increased lifespan and stress
resistance (that is, healthy longevity) of *C. elegans* when fed *B. subtilis* (Figs 1–3 and Table 1).

In the case of the host (*C. elegans*), the benefits of the biofilm are evident: lifespan extension, higher stress resistance and probably competitive exclusion of pathogens2,21,22,42. In addition to our results, it has been reported that *B. subtilis* strains proficient in biosurfactant production (that is, lycengycin) are able to protect *C. elegans* from infection by bacterial pathogens22. In addition, the ability to establish a biofilm in connection with the host cells would allow for a stronger and closer interaction between the bacterium and the host13–14. The finding that exogenously added CSF (Fig. 5) and NO28 extend the lifespan of *C. elegans* fed *E. coli* cells supports the notion of the underlaying phenomenon of worm–bacteria QS. This dual microbial–worm interaction would allow the bacterium to colonize and establish a multicellular biofilm in the friendly environment of the worm gut mucosa, a similar scenario to the formation of stable biofilms during the beneficial bacteria–plant interaction of some Bacilli (that is, the notion of the underlying phenomenon of worm–bacteria QS). In addition, the bacterial properties and behaviour in a multicellular biofilm differ from the behaviour and properties of bacteria grown as individuals under planktonic conditions16–18. Specifically, our results demonstrated the importance of the biofilm for the success of *B. subtilis* to healthy *C. elegans* gut colonization (Figs 2 and 3), increase the production of the anti-aging molecules NO/CSF (Table 2), and increase worm longevity (Figs 4 and 5). Future studies would address how the production of the anti-aging molecules NO and CSF is enhanced in *B. subtilis* cells grown as a biofilm (Table 2) and whether this higher NO and CSF production might be influenced by signals originating from the host (Supplementary Table 10).

The present work also indicated that the prolongevity effect of *B. subtilis* on *C. elegans* requires the activity of FOXO/DAF-16 and IHSF-1 (Figs 6 and 7 and Table 1). At the top of the regulatory pathways that modulate the activity of HSF-1 and DAF-16 in *C. elegans* are the ILS pathway (homologous to the insulin/IGF-1 pathway present in humans) and the process of DR1,2,29. We showed that the prolongevity effect of *B. subtilis* on *C. elegans* primarily depends on the functionality (downregulation) of the ILS pathway (Fig. 8a). This result points to the ILS pathway as the target of the *B. subtilis* biofilm to increase host longevity. Interestingly, it has been observed that healthy human centenarians likely have IGF-1 receptor genetic variants associated with reduced function, an intriguing observation that correlates with our findings. As ILS pathway is part of the nutrient signalling pathway, it is also (negatively) controlled by DR a scenario that open the possibility that ultimately DR might sense and control the ILS-dependent prolongevity effect of *B. subtilis* (Fig. 8b). Finally, driven by the Microbiome Project3,8,10,7,24, literature about the microbiota and its effects on human health and longevity has grown exponentially in the last decade. This scenario and the present results open the possibility to test if *B. subtilis* might produce beneficial effects on human longevity7,24–26.

**Lifespan analysis.** Lifespans were monitored at 20°C as described previously28. Briefly, to obtain a synchronized population, embryos were isolated by treating the culture containing the worm strains with alkaline hypochlorite to kill vegetative cells. In all the cases, late L4/young adult stage worms were used at t=0 for lifespan analysis, and 16 µM 5-fluoro-2'-deoxyuridine (FUDR, Sigma Co.) was used to inhibit progeny growth. During the quantification of the number of dead/live worms, the investigator was blinded to the group allocation. All the experiments were repeated at least three times. Worms were considered dead when they ceased pharyngeal pumping and did not respond to prodding with a platinum wire. Worms with internal hatching were removed from the plates and excluded from lifespan calculations. Given that some temperature-sensitive mutations in some genes lead to the formation of dauer larvae, the lifespan experiments involving *daf-2*(e1770) were performed shifting the temperature from 15 to 20°C, the non-permissive temperature, at late L4/young adult stage. For lifespan analysis with no FUDR, a synchronous population was obtained by sodium hypochlorite treatment of gravid hermaphrodites to obtain eggs that were then raised on standard NGM plates. Late L4/young adult stage worm were transferred to fresh plates at the complete absence of FUDR every 2 days, while any of them were fertile. Survival was scored as described above.

**Culturing bacteria from worms.** The N2 *C. elegans* eggs were isolated using a solution of 10% commercial bleach and 1 N NaOH followed by four washes with M9 buffer (22 mM KH2PO4, 42 mM Na2HPO4, 85 mM NaCl and 1 M MgSO4). Approximately 500 eggs were transferred to a 60 mm plate with NGM agar and incubated overnight at 20°C with agitation to allow L1 larvae to emerge. Three or more 5-day-old worms were transferred to each well-plate containing 200 mM NaCl (for osmotic stress), 50 µM cadmium (for metal stress), 25 mM H2O2 (for oxidative stress) and 1% Triton X-100. The worms were treated with 25 mM levamisole to induce temporal paralysis, superficially sterilized with 3% commercial bleach for 15 min and washed three times with M9 buffer. After the worms were surface sterilized, worms devoid of outside bacteria were disrupted using a pellet pestle (Sigma Co.), centrifuged and resuspended in 500 µL M9 buffer. Finally, 50 µL of each cell suspension was used to prepare serial dilutions of the bacteria before counting. To this end, 100 µL of the appropriate serial dilutions was spread with a Drigalski scraper on LB Petri dishes. The number of colony-forming units (CFUs) was determined after 24 h of incubation at 37°C. To determine the number of spores, samples were heat-treated at 80°C for 20 min before spreading. This treatment kills the vegetative cells, while the spore cells remain alive33. The number of CFUs obtained before and after the heat treatment represents an estimation of the number of vegetative and spore cells, respectively, inside the worm.

**β-Galactosidase activity assay.** β-Galactosidase activity was assayed in *B. subtilis* cells harbouring lacZ fusions, and the specific activity was expressed in Miller Units (MU)34. To determine gene-derived β-galactosidase activity, 1-day-old L4 worms were separately fed on NGM plates with a homogenate of each *B. subtilis* strain. At different incubation times, 50 worms were transferred to Eppendorf tubes containing M9 buffer, and 1% Triton X-100. The worms were treated with 25 mM levamisole to induce temporal paralysis, superficially sterilized with 3% commercial bleach for 15 min and washed three times with M9 buffer. After the worms were surface sterilized, worms devoid of outside bacteria were disrupted using a pellet pestle (Sigma Co.), centrifuged and resuspended in 500 µL M9 buffer. Finally, 50 µL of each cell suspension was used to prepare serial dilutions of the bacteria before counting. The number of colony-forming units (CFUs) was determined after 24 h of incubation at 37°C. To determine the number of spores, samples were heat-treated at 80°C for 20 min before spreading. This treatment kills the vegetative cells, while the spore cells remain alive35. The number of CFUs obtained before and after the heat treatment represents an estimation of the number of vegetative and spore cells, respectively, inside the worm.

**Methods.**

**Strains and growth media.** The *B. subtilis* strains used in this study were the domesticated H642 (ref. 33), the undomesticated RG4365 (ref. 27) and NCIB3610 (ref. 23), and their isogenic derivatives (Supplementary Table 1). The OP50 *E. coli* strain was obtained from the Caenorhabditis Genetic Center (CGC). As indicated, bacteria were grown in Luria-Bertani (LB) broth and Schaeffer’s sporulation medium (SM)35. Sporulation was induced by the starvation method: *B. subtilis* strains were grown in SM medium at 37°C for 48 h (ref. 30). After this incubation period, the culture was heat-treated for 20 min at 80°C to kill vegetative cells30. To obtain pure spores, the heat-treated culture was treated three times with lysozyme (25 µg mL−1, Sigma Co.), washed each time with cold water and centrifuged until 100% of the culture consisted of phase-bright spores. When indicated, the pure spore solution was autoclaved twice to kill spores. Wild-type N2 (Bristol) and mutant strains (obtained from CGC) worms were maintained at 20°C on Nematode Growth Medium agar (NGM)28 plates seeded with OP50 *E. coli* or *B. subtilis* cells with or without supplementation with ampicillin (100 µg mL−1).

To obtain pure *B. subtilis* cells harboured the *parA*::lacZ reporter fusion. At different times, approximately 100 worms were taken from the NGM plates containing each
**Microscopic visualization of bacterial cells colonizing the nematode intestinal tract.** Day 5 adult worms fed B. subtilis strains carrying an integrated epsA::gfp or bilaA::gfp reporter fusion were pipetted out from plates using M9 buffer and 0.25 mM sodium azide (Sigma Co.) and covered with a glass coverslip. Bacterial colonization of the nematode digestive tract was observed using an Olympus FV1000 laser confocal scanning microscope. Images were analysed with Olympus software and Fiji software.

**Bacterial pulse-chase experiment.** Day 5 adult worms fed different B. subtilis strains (JH642 or NCIB3610) were washed and transferred to NGM plates seeded with OP50 on C176. E. coli or NCIB3610/JH642 strains (JH642 or NCIB3610) were washed and transferred to NGM plates containing lawns of OP50 on C176. E. coli or NCIB3610/JH642 or B. subtilis cells were anaesthetized with sodium azide, straightened and photographed. Their length in pixels was then compared with a 1 mm scale bar.

**Chemoatxos assay.** Batches of 20 age-synchronized worms (late L4/young adult stage) were transferred to a plate containing lawns of OP50 E. coli or JH642 or NCIB3610 B. subtilis. Unstaged eggs were placed at 20°C and allowed to hatch for 4 h. Larvae that hatched during this period were placed singly on fresh plates and monitored every 5 h until they reached late L4/young adult stage. To measure the worm size, ~20 randomly picked worms fed OP50 E. coli or NCIB3610/JH642 B. subtilis cells were anaesthetized with sodium azide, straightened and photographed. Their length in pixels was then compared with a 1 mm scale bar.

**Self-brood size and egg production rate.** These experiments were performed essentially as described5,6. The eggs from strain N2 nematodes were isolated, treated with hypochlorite and incubated for 20 h at 20°C in S-buffer (0.5 M KH₂PO₄, 0.5 M K₂HPO₄ and 0.1 M NaCl). A synchronized population of L1 arrested worms was then placed on NGM agar plates seeded with OP50 E. coli or NCIB3610/JH642 B. subtilis cells. Five late L4/young adult stage animals were picked manually and transferred to a new plate. The worms were transferred twice a day to prevent overcrowding until egg laying ceased. The progeny was counted 3 days after removal of the parents. The experiment was performed in triplicate.

**Cultural media and antibiotics.** Bacterial cultures were grown on Luria-Bertani agar (LB) plates. Bacterial colonies were transferred to LB broth and cultured with or without shaking, respectively. In MSgg31 or biofilm-supporting conditions (with or without shaking, respectively) in MSgg31 or B. subtilis 168 TOP10 cells were transformed by electroporation. Transformation efficiency was determined by plating a serial dilution of the transformation mix on LB agar plates and counting the number of colonies. The transformation efficiency was generally 10,000–100,000 transformants per microgram of plasmid DNA. Appropriate aliquots of each cell-free supernatant were assayed for the presence of NO-derived metabolites (NO₂⁻ and NO₃⁻) as reported previously29. For determining the β-galactosidase activity driven from the csf-promoter, the cells were grown as a biofilm or planktonically, concentrated by centrifugation and treated as reported previously30. The production levels of NO and CSF are expressed as μmol and MU, respectively, per 1 × 10⁶ CFUs counted from the cultures developed under planktonic and biofilm-supporting conditions.
