Phylogenetic lineages in the Botryosphaeriaceae

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Abstract: Botryosphaeria is a species-rich genus with a cosmopolitan distribution, commonly associated with dieback and cankers of woody plants. As many as 18 anamorph genera have been associated with Botryosphaeria, most of which have been reduced to synonymy under Diplodia (conidia mostly ovoid, pigmented, thick-walled), or Fusicoccum (conidia mostly fusoid, hyaline, thin-walled). However, there are numerous conidial anamorphs having morphological characteristics intermediate between Diplodia and Fusicoccum, and there are several records of species outside the Botryosphaeriaceae that have anamorphs apparently typical of Botryosphaeria s.str. Recent studies have also linked Botryosphaeria to species with pigmented, septate ascospores, and Dothiorella anamorphs, or Fusicoccum anamorphs with Dichoherea synamanaphors. The aim of this study was to employ DNA sequence data of the 28S rDNA to resolve apparent lineages within the Botryosphaeriaceae. From these data, 12 clades are recognised. Two of these lineages clustered outside the Botryosphaeriaceae, namely Diplodia-like anamorphs occurring on maize, which are best accommodated in Stenocarpella (Diaphthiales), as well as an unresolved clade including species of Camarosporum/Microbridiophia. We recognise 10 lineages within the Botryosphaeriaceae, including an unresolved clade (Diplodia/Lasiodiplodiella Tiarosporellae), Botryosphaeria s.str. (Fusicoccum anamorphs), Macrohomina, Neoscytalidium gen. nov., Dothiodothy (Dothiorella anamorphs), Neofusicoccum gen. nov. (Botryosphaeria-like teleomorphs, Dichoherea-like synamanaphors), Pseudofusicoccum gen. nov., Saccharata (Fusicoccum- and Diplodia-like synamanaphors), “Botryosphaeria” quercuum (Diplodia-like anamorphs), and Guignardia (Phyllosticta anamorphs). Separate teleomorph and anamorph names are not provided for newly introduced genera, even where both morphs are known. The taxonomy of some clades and isolates (e.g. B. mamane) remains unresolved due to the absence of ex-type cultures.

Taxonomic novelties: Neofusicoccum Crous, Slippers & A.J.L. Phillips gen. nov., Neofusicoccum andinum (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M.J. Wingf. comb. nov., Neofusicoccum arbuti (D.F. Farr & M. Elliott) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum australe (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum eucalypticola (Slippers Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum eucalyptorum (Crous, H. Smith & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum luteum (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum macroclavatum (Burges, Barber & Hardy) Burgess, Barber & Hardy comb. nov., Neofusicoccum mangiferae (Syd. & P. Syd.) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum parvum (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum protearum (Dennman & Crous) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum ribis (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum viticlavatum (Niekerk & Crous) Crous, Slippers & A.J.L. Phillips comb. nov., Neofusicoccum vitulostereum (Niekerk & Crous) Crous, Slippers & A.J.L. Phillips comb. nov., Neoscytalidium Crous & Slippers gen. nov., Neoscytalidium dimidiatum (Penz.) Crous & Slippers comb. nov., Neoscytalidium Crous & Slippers gen. nov., Neoscytalidium dichotomum (Penz.) Crous & Slippers gen. nov., Neoscytalidium salicis (Fries) Crous & Slippers gen. nov., Neoscytalidium novaceps (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M.J. Wingf. comb. nov.

Key words: Ascomycetes, Botryosphaeria, Botryosphaeriaceae, Diplodia, Dothiorella, Fusicoccum, Lasiodiplodia, Macrohomina, Neofusicoccum, Neoscytalidium, Pseudofusicoccum, Scytalidium, Stenocarpella, Tiarosporella, systematics.

INTRODUCTION

The genus Botryosphaeria Ces. & De Not. was introduced in 1863 (Cesati & De Notaris 1863), emended by Saccardo (1877), and is based on the type species Botryosphaeria dothidea (Mouge. : Fr.) Ces. & De Not. (Barr 1972, Slippers et al. 2004). Botryosphaeria is a species-rich genus with a cosmopolitan distribution (Dennman et al. 2000). Species occur on a wide range of monocotyledonous, dicotyledonous and gymnosperm hosts, on woody branches, herbaceous leaves, stems and haulms of grasses, on twigs and in the thalli of lichens (Barr 1987). Taxa range in habit from being saprofic, parasitic and endophytic (Smith et al. 1996, Denman et al. 2000), and can cause die-back and canker diseases of numerous woody hosts (von Arx 1987).

Botryosphaeria has been well circumscribed, and can be defined as forming uni- to multiocular ascomata with multi-layered walls, occurring singly or in clusters, often intermixed with conidiomata, which are pycnidial. Asci are bitunicate, with a thick endotunica, stalked or sessile, clavate, with a well-developed apical chamber, forming in a basal hymenial layer, intermixed among hyaline pseudoparaphyses that are frequently constricted at the septa. Ascospores are hyaline, asperate, fusoid to ellipsoidal or ovoid, bi- to triseriate, mostly without a mucoid sheath or appendages; ascospores turn brown and become septate and even slightly verruculose upon germination (von Arx & Müller 1954, Shoemaker 1964, Eriksson 1981, Sivanesan 1984, Denman et al. 2000, Alves et al. 2004).

Theissen & Sydow (1915) placed Botryosphaeria in the Botryosphaeriaceae, a sub-family of the Pseudosphaeriaceae, which was not assigned to any specific order. The Pseudosphaeriaceae was later placed in the Myriangiales (Theissen 1916), and in 1917 Theissen & Sydow were of the opinion
that the *Pseudosphaeriaceae* should be united with the *Dothideaceae* (Luttrell 1951). The *Dothideales* were characterised by the formation of asci in locules embedded in stromata, and contained the *Dothideaceae*, a family established to accommodate multiciliate forms like *Botryosphaeria*. Petrak (1923) placed *Botryosphaeria* in the sub-family *Pseudosphaeriales*, which was placed in the *Pleosporaceae* (*Sphaeriales*).

Miller (1928) placed *Botryosphaeria* in the *Dothideales* because true perithecial walls were absent. He later recognised three orders, namely the *Sphaeriales* (with perithecia and paraphyses), the *Dothideales* (ascostromatic forms without paraphyses), and the *Pseudosphaeriales* (ascostromatic forms with interthecal threads) and assigned *Botryosphaeria* to the *Pseudosphaeriales*.

Luttrell (1955) identified eight types of centrum development, and highlighted the taxonomic value of sterile, interthecal tissues in the taxonomy of the *Ascomycetes*. He furthermore replaced the name *Pseudosphaeriales* with *Pleosporales*, and assigned *Botryosphaeria* to this order. Luttrell's views were supported by Eriksson (1981) and Barr (1987). The orders proposed by Luttrell and Barr were not accepted by von Arx & Müller (1975) and von Arx (1987), as they comprised a mixture of unrelated genera (von Arx 1987). Von Arx & Müller (1975) only delimited the *Dothideales*, with two sub-orders and 24 families. Their view was that this was a more appropriate means of dealing with the *Dothideales* because true perithecial walls were absent.

Dothideomycetes, which separated the examined *Botryosphaeria* spp. into two groups, correlating to those species with *Diplodia*-like anamorphs and those with *Fusicoccum*-like anamorphs (Jacobs & Rehner 1998, Denman et al. 2000). Later studies including additional species and a larger suite of DNA-based markers supported this view (Zhou & Stanoz 2001, Alves et al. 2004, Slippers et al. 2004). However, this apparently clear sub-division is questioned by *Saccharata proteae* Denman & Crous (as *Botrysphaeria proteae* (Wakef.) Denman & Crous with *Fusicoccum* and *Diplodia* synanamorphs), which is morphologically and phylogenetically distinct from representatives of the *Diplodia-* and *Fusicoccum*-like groups (Crous et al. 2004). Some authors have continued to use *Lasiodiplodia* Ellis & Everh. as a genus distinct from *Diplodia* Fr., because of its distinct phylogenetic (usually ITS or EF-1α) and morphological (striae conidia and paraphyses) characteristics (Pavlic et al. 2004). Recently, the name *Dothiorella* Sacc. has also been re-introduced as a distinct *Botryosphaeria* anamorph (conidia brown, septate while still attached to the conidiogenous cells) (Philips et al. 2005a) and *Dichomera* Cooke has been linked to *Botryosphaeria* species with *Fusicoccum* anamorphs (Barber et al. 2005). Many of the other 18 coelomycete genera linked to *Botryosphaeria* remain untested in terms of phylogenetic association to the above groups.

The representation and phylogenetic understanding of major groups within *Botryosphaeria* remains poor. Previous analyses based on DNA sequence comparisons have included limited numbers of species, not representing the full anamorph diversity associated with *Botryosphaeria*. The value of the intron-dominated sequences of the ITS, β-tubulin and EF-1α loci (on which most previous studies were based) to infer phylogenetic relationships across the diversity of the genus, is also unclear. The more conserved mtSSU data have, for example, suggested that *B. dothidea* and *B. corticis* (Demaree & Wilcox) Arx & E. Müll. are unrelated to *Fusicoccum s.str.* (Zhou & Stanoz 2001) even though they are typically assigned to this genus.

*Botryosphaeria* as a single genus is clearly unaligned with evolutionary radiations in the group, as exemplified by the morphologically and phylogenetically distinct anamorph genera linked to it. A preferable approach would be natural unit classification, also referred to as the “genus for genus concept” (Seifert et al. 2000). Here, morphologically distinct anamorph genera are linked to unique teleomorphs on a one for one basis, correlating with phylogenetic DNA data. This is an approach that has been applied to genera such as *Calonectria* De Not. (Crous 2002), *Cryphonectria* (Sacc.) Sacc. & D. Sacc. (Gryzenhout et al. 2006 – this volume), *Ophiostoma* Syd. & P. Syd. (Zipfel et al. 2006 – this volume) and *Botryosphaeria* (Rossman & Samuels 2005). The primary aim of the present study is to delineate the phylogenetic lineages of the *Botryosphaeriaceae*, and to discuss the morphological differences and generic concepts that can be ascribed to them. For this purpose, we have chosen comparisons of sequences for the 28S rDNA gene (LSU) because of its favourable size (approx. 900–1000 bp. used), and its relatively conserved (medium–high level) nature, suitable to consider taxonomic sub-divisions at the generic level.

**MATERIALS AND METHODS**

**Isolates**

Single-conidial or ascospore isolates were made from ascomata or pycnidia on dead or dying twigs of various hosts as explained in Slippers et al. (2004). Other isolates of representative *Botryosphaeria* spp. were obtained from the Centraalbureau voor Schimmelcultures (CBS), Utrecht, the Netherlands and the Culture Collection of the Tree Protection Co-operative Programme (CMW), FABI, University of Pretoria, South Africa (Table 1).
Cultural characteristics were determined on plates containing 2 % malt extract agar (MEA), 2 % potato-dextrose agar (PDA), and oatmeal agar (OA) (Gams et al. 1998).

**DNA phylogeny**

The isolation protocol of Lee & Taylor (1990) was used to extract genomic DNA from fungal mycelia grown on MEA. The primers ITS1 (White et al. 1990) and LR5 (Vilgalys & Hester 1990) were used to amplify part of the nuclear rRNA operon using the PCR conditions recommended by the authors and spanning the 3' end of the 18S rRNA gene, the internal spacers, the 5.8S rRNA gene and a part of the 5' end of the 28S rRNA gene. PCR products were separated by electrophoresis at 80 V for 1 h in a 0.8 % (w/v) agarose gel in 0.5 × TAE running buffer (0.4 M Tris, 0.05 M NaAc, and 0.01 M EDTA, pH 7.85) and visualised under UV light using a GeneGenius Gel Documentation and Analysis System (Syngene, Cambridge, U.K.) following ethidium bromide staining. The amplification products were purified using a GFX PCR DNA and Gel Band Purification Kit (Amersham Pharmacia Biotech Europe GmbH, Germany). The purified products were sequenced in both directions using an ABI PRISM Big Dye Terminator v. 3.1 Cycle Sequencing Ready Reaction Kit (PE Biosystems, Foster City, CA) containing AmpliTaq DNA Polymerase as recommended by the manufacturer. The primers LROR (Rehner & Samuels 1994), LR3R (http://wwwbiology.duke.edu/fungi/mycolab/primers.htm), LR16 (Moncalvo et al. 1993), and LR5 (Vilgalys & Hester 1990) were used to ensure good quality sequences over the entire length of the amplicon. The resulting fragments were analysed on an ABI Prism 3100 DNA Sequencer (Perkin-Elmer, Norwalk, CN).

DNA sequences were assembled and added to the outgroups and additional GenBank sequences using Sequence Alignment Editor v. 2.0a11 (Rambaut 2002), and manual adjustments for improvement were made by eye where necessary. The phylogenetic analyses of sequence data were done in PAUP (Phylogenetic Analysis Using Parsimony) version 4.0b10 (Swofford 2002) and consisted of neighbour-joining analysis with the uncorrected ("p"), the Kimura 2-parameter and the HKY85 substitution model in PAUP. Alignment gaps were treated as missing data and all characters were unordered and of equal weight. Any ties were broken randomly when encountered. For parsimony analysis, alignment gaps were treated as both missing and as a fifth character state and all characters were unordered and of equal weight. Maximum parsimony analysis was performed using the heuristic search option with simple taxa additions and tree bisection and reconstruction (TBR) as the branch-swapping algorithm. Branches of zero length were collapsed and all multiple, equally parsimonious trees were saved. The robustness of the trees obtained was evaluated by 1000 bootstrap replications (Hillis & Bull 1993). Tree length (TL), consistency index (CI), retention index (RI) and rescaled consistency index (RC) were calculated and the resulting trees were printed with TreeView v. 1.6.6 (Page 1996).

Bayesian analysis was conducted on the same aligned LSU dataset as the distance analysis. First MrModeltest v. 2.2 (Nylander 2004) was used to determine the best nucleotide substitution model. Phylogenetic analyses were performed with MrBayes v. 3 (Ronquist & Huelsenbeck 2003) applying a general time-reversible (GTR) substitution model with gamma (G) and proportion of invariable site (I) parameters to accommodate variable rates across sites. The Markov Chain Monte Carlo (MCMC) analysis of 4 chains started from random tree topology and lasted 10 000 000 generations. Trees were saved each 100 000 generations, resulting in 1000 saved trees. Burn-in was set at 500 000 generations after which the likelihood values were stationary, leaving 950 trees from which the consensus trees and posterior probabilities were calculated. PAUP 4.0b10 was used to reconstruct the consensus tree, and maximum posterior probabilities were assigned to branches after a 50 % majority rule consensus tree was constructed from the 950 sampled trees.

**Taxonomy**

Morphological descriptions were made for isolates sporulating on 2 % water agar (WA) with sterilised pine needles as substrate, at 25 °C under near-UV light, to induce sporulation. Structures were mounted in lactic acid, and 30 measurements at × 1000 magnification were made of each structure where possible. The 95 % confidence levels were determined, and the extremes of spore measurements given in parentheses. All cultures used in this study are maintained in the CBS culture collection.

**RESULTS**

**DNA phylogeny**

For the LSU gene, approximately 1000 bases were determined for the isolates listed in Table 1. Additional sequences, some of which were shorter, were also obtained from GenBank and added to the alignment. The manually adjusted alignment contained 115 taxa (including the two outgroups) and 576 characters including alignment gaps in TreeBASE (S1505, M2707). Of the 576 characters used in the phylogenetic analysis, 190 were parsimony-informative, 19 were variable and parsimony-uninformative, and 367 were constant when gaps were treated as missing characters. When gaps were treated as a new character state, seven more parsimony informative characters were added. Neighbour-joining analysis using the three substitution models on the sequence data yielded trees with similar topology and bootstrap values.

Twelve clades could be identified in the distance tree obtained using the HKY85 substitution model (Fig 1). These are discussed in the Taxonomy and Discussion sections. Parsimony analysis with gaps treated as missing characters yielded 79604 equally parsimonious trees (TL = 582 steps; CI = 0.509; RI = 0.905; RC = 0.460). Treating gaps as new states resulted in 79775 equally parsimonious trees (TL = 603 steps; CI = 0.524; RI = 0.910; RC = 0.477).
strict consensus trees calculated from the equally parsimonious trees were identical to each other and are shown in TreeBASE. Between the neighbour-joining and parsimony analyses, the same clades were supported with two exceptions. The first exception is *Botryosphaeria mamane* D.E. Gardner (CBS 117444), which resides in Clade 3 (Fig. 1), but is basal to Clades 1 to 6 in the strict consensus trees. Also, Clade 7 groups with Clade 10 in the strict consensus trees, but this is not supported with a bootstrap analysis (data not shown).

Bayesian analysis resulted in a tree with largely the same topology and clades (Fig. 2). Differences observed were related to the position of *B. mamane*, which clustered close to Clade 3 in the distance analysis, but clustered in Clade 4 in the Bayesian analysis. A further difference was that in the Bayesian analysis Clades 8–9 clustered basal to the *Botryosphaeriaceae*.

**Taxonomy**

A total of 113 isolates representing most of the morphological variation presently recognised in the *Botryosphaeriaceae* were subjected to DNA sequence analysis. These analyses revealed 11 clades in the family. These phylogenetic clades can also be correlated with distinct morphological features.
Clade 1 includes species with Diplodia, Sphaeropsis Sacc. and Lasiodiplodia anamorphs clustering together. Although sequences of gene regions such as ITS, EF-1α and β-tubulin, support the synonymy of Sphaeropsis under Diplodia, in various cases they separate Lasiodiplodia from the Diplodia clade (Pavic et al. 2004, Phillips et al. 2005a). This is in contrast to the LSU dataset (Fig. 1) in which species having Diplodia anamorphs could not be separated from those in Lasiodiplodia. Inclusion of additional strains resulted in a basal polytomy with low bootstrap support. Furthermore, uncertainty remains as to which teleomorph name is best suited for this clade, as the form genus Diplodia is known to be polyphyletic (Sutton 1980, Sivanesan 1984). The clustering of three species of Tiarospora Höhn. in this clade was also unexpected.

Clade 2 is represented by the type species of the genus Botryosphaeria, namely B. dothidea, and its anamorph Fusicoccum aesculi Corda. The genus Macrohomorphosphaeria N.E. Stevens & Baechler, represented by a strain identified as M. coronillae (Desm.) Petr. (type species) (CBS 769.71), clusters in this clade, as does the genus Dichomera Cooke, represented by a strain identified as D. saubinetti (Mont.) Cooke (type species) (CBS 990.70).

Clade 3 is represented by isolates of Macrohomophora phaseolina (Tassi) Goë. This fungus is the coelomycete synanamorph of “Rhizoctonia” batalicola (Tauben.) E.J. Butler, which is shown to be a member of the Botryosphaeriaceae. Conidia also have apical mucous appendages early in their development, which has in the past led to confusion, and the allocation of this species to the genus Tiarospora (von Arx 1981). With age, conidia lose their apical appendages, and become brown and slightly roughened, appearing more Diplodia-like in morphology.

Clade 4 represents Fusicoccum dimidiatum (Penz.) D.F. Farr. This species, which has a large number of synonyms (Farr et al. 2005), is unusual in having a Fusicoccum-like coelomycete anamorph (with mucoid apical appendages). It also has a powdery Scytalidium-like hyphomycete synanamorph that is lacking in other species in the Botryosphaeriaceae. Scytalidium, typified by S. lignicola Pesante (CBS 233.57) clusters outside of the Botryosphaeriaceae. However, DNA sequence data derived from the Scytalidium species present in the CBS collection lead us to conclude that this genus is also polyphyletic (results not given).

Botryosphaeria mammame has a Fusicoccum anamorph and clusters most closely with species in either Clade 3 or Clade 4 in the various analyses. The Fusicoccum anamorph is morphologically most similar to species residing in Clade 2 (Fusicoccum s.str.). It does not have the apical appendages or discoloration found in species residing in Clade 3, nor does it have a Scytalidium-like synanamorph occurring in species residing in Clade 4. Consequently, its taxonomic position remains unresolved.

Clade 5 represents Botryosphaeria-like teleomorphs with pigmented, septate ascospores for which the genus Dothidotthia Höhn. is available (Barr 1987, 1989). Anamorphs of this genus reside in Dothiorella Sacc. (Phillips et al. 2005a).

Clade 6 represents Botryosphaeria-like species with Fusicoccum-like anamorphs and Dichomera-like synanamorphs, for which the name Neofusicoccum gen. nov. is introduced. The Dichomera-like synanamorphs in this clade are characterised by globose to pyriform conidia. The older, brown conidia in Clade 2 (Botryosphaeria s.str.) are obovoid, ellipsoid or fusiform, never globose or subglobose (Phillips et al. 2005b, Barber et al. 2005).

Clade 7 represents isolates of “Fusicoccum” stromaticum Mohali, Slippers & M.J. Wingf. (Mohali et al. 2006). This taxon is distinguished from Fusicoccum aesculi and other Fusicoccum-like genera by having conidia that are enclosed in a persistent mucous sheath, for which the genus Pseudofusicoccum is proposed.
Clade 8 is represented by a single species, *Botryosphaeria quercuum* (Schwein.) Sacc., for which the genus *Melanops* Nitschke ex Fuckel is available. *Saccharata* Denman & Crous (anamorph *Fusicoccum*-like; synanamorph *Diplodia*-like) (Clade 9) is morphologically distinguished from *Botryosphaeria* s. str. by having unilocular ascomata that develop under a clypeus.

Clade 10 includes species of *Guignardia* Viala & Ravaz with *Phyllosticta* Pers. anamorphs (Van der Aa & Vaney 2002). Clade 11 contains several distinct genera, namely *Camarosporium* Schulzer [type = *C. quaternatum* (Hazsl.) Sacc.], *Phyllosticta* *flevolandica* Aa, and other morphologically distinct taxa. *Diplodia* macropora Earle and *Diplodia* *maydis* (Berk.) Sacc. (Clade 12) are shown to be distinct from the *Botryosphaeriaceae*. *Stenocarpella* Syd. & P. Syd. is appropriate for them, as they cluster apart from the *Botryosphaeriaceae*. Surprisingly, they cluster in the *Diaporthales* although no teleomorph connections are currently known for species of *Stenocarpella*.

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**Fig. 2.** Consensus phylogram of 950 trees resulting from a Bayesian analysis of 115 LSU sequences. Bayesian posterior probabilities are given at the nodes. Clades are numbered from 1–12 next to the brackets, following the number-to-clade assignment presented in Fig. 1. The tree was rooted to *Gaeumannomyces graminis* var. *avenae* (AF362556) and *Magnaporthe grisea* (AF362554) (*Diaporthales*, *Magnaporthaceae*).
DISCUSSION

Based on the LSU phylogeny obtained in the present study (Fig. 1), 11 clades could be recognised for species that have been applied to or that have anamorphs allied to the Botryosphaeriaceae. An additional 12th Clade included two species that have been treated in Diplodia and that reside in the Diaporthales. These 12 clades are discussed individually as follows:

Clade 1: Diplodia/Lasiodiplodia (several teleomorph genera available)

Species in Clade 1 are poorly resolved in both distance and Bayesian analyses. This is due to the few informative sites in this clade (19) in the section of LSU selected for this study. A larger segment of the LSU or additional gene regions will be required to resolve the phylogenetic relationships of species residing in this clade.

In the past, anamorphs of Botryosphaeria have been described in up to 18 different genera (Denman et al. 2000), many of which were not clearly defined and contain dark conidia typical or similar to those typical of Diplodia. Sutton (1980) reduced Macrophoma (Sacc.) Berl. & Voglino to synonymy with Sphaeropsis. Pennycook & Samuels (1985) reduced Macrohomospora to synonymy with Fusicoccum. Crous & Palm (1999) showed that Botryodiapodia (Sacc.) Sacc. was a nomen dubium, and reduced Dothiorella to synonymy with Diplodia. Denman et al. (2000) also regarded Sphaeropsis and Lasiodiplodia as synonyms of Diplodia. Phillips et al. (2005a) again separated Dothiorella from Diplodia, and also provided evidence to show that the teleomorphs were different.

Recent treatments of Botryosphaeria anamorphs have revealed that they cluster in two clades, namely Diplodia (dark, mostly >10 µm broad, thick-walled conidia), and Fusicoccum (hyaline, mostly <10 µm broad, thin-walled conidia) (Jacobs & Rehner 1998; Denman et al. 2000, Zhou & Stanosz 2001, Alves et al. 2004). With age, however, conidia in species of Fusicoccum become dark, and frequently also septate. This complicates identification, especially where this is attempted based on structures occurring on natural substrates and in the absence of fresh cultures.

Pavlic et al. (2004) described a new species of Lasiodiplodia based on its characteristic conidial ornamentation typical of the genus (von Arx 1987). Xiao & Rogers (2004) described a new species of Diplodia, and placed it in Sphaeropsis. The correct generic names to be used for the dark-spored anamorphs of Botryosphaeria thus remains uncertain.

The genus Sphaeropsis is based on S. visci (Fr.) Sacc., while the genus Lasiodiplodia is based on L. theobromae (Pat.) Griff. & Maubl. In the present study we included cultures of both species. Based on the LSU phylogeny, it is clear that strains of Sphaeropsis visci reside in this clade (CBS 186.97, 100163). Furthermore, our data also reveal that the strains deposited in CBS under the name L. theobromae represent several distinct species with this typical conidial ornamentation; the LSU phylogenetic data could not resolve the Lasiodiplodia Diplodia clade. Lasiodiplodia gonubiensis and three new species of Lasiodiplodia recently described (Burgess et al., unpubl. data), were interspersed among species of Diplodia. The rather atypical Diplodia species recently described by Van Niekerk et al. (2004) as D. porosum Van Niekerk & Crous, also clustered in this clade. Botryosopheria subglobosa (C. Booth) Arx & E. Müll. is another interesting example, as it has an anamorph described as Sphaeropsis subglobosa C. Booth. Although the latter species is illustrated to have what appears to be a germ slit in its asceptate conidia
from designating a teleomorph name for this clade, in anticipation of the additional research that is needed to elucidate the status of these older names. A further possibility is that the taxa in Clade 1 still represent more than one genus.

Clade 2: *Botryosphaeria* (anamorph *Fusicoccum*)
Barber et al. (2005) have recently shown that species of *Fusicoccum* can have *Dichomera* Cooke synanamorphs. Furthermore, two species of *Dichomera*, *D. versiformis* Z.Q. Yuan, Wardlaw & C. Mohammed and *D. eucalypti* (G. Winter) B. Sutton also cluster in Clade 6 with the majority of the “*Fusicoccum*” species. Phillips et al. (2005b) also reported that conidia of *Fusicoccum aesculi* from olives can become pigmented, ovoid, ellipsoid or fusiform, 1–2-septate, similar to those observed by Barber et al. (2005) for *F. aesculi*. Phillips et al. (2005b) also revealed *Fusicoccum dalmaticum* (Thüm.) Vanev is the same as *Camarosporium dalmaticum* (Thüm.) Zachos & Tzav.-Klon., both being later synonyms of *F. aesculi*. Sutton (1980) stated that *Camarosporium* Schulzer was the pycnidial analogue of *Dichomera*, which had more stromatic conidiomata. Given our current knowledge of the phylogenetic value of conidiomatal structure in the *Dothideomycetes*, it seems redundant to separate these anamorph genera based on this character alone (also see illustrations in Sutton 1980). The generic name *Camarosporium* (1870) is older, and thus has preference over *Dichomera* (1876). When a strain identified as *D. saubinetii* (Mont.) Cooke (CBS 990.70; sterile, morphology unconfirmed) was subjected to sequence analysis, it clustered in Clade 2, while strains identified as *C. quaternatum* (CBS 134.97, 483.95; fertile, matching the original description) clustered outside of the *Botryosphaeriaceae*. In *Fusicoccum* s.str. (*F. aesculi*) conidia are fusiform to ellipsoid, and with age become septate and brown, to some extent appearing *Dichomera*-like. Whether strains matching *D. saubinetii* in morphology will cluster in Clade 2 remains to be determined.

Sutton (1980) stated that *Macrophomopsis coronillae* was closely related to *F. aesculi*, but distinguishable by its anellidic conidiogenous cells. Crous & Palm (1999) showed that percurrent proliferation occurred in the type of *F. aesculi*, and concluded that Pennycook & Samuels (1985) were probably correct to reduce *M. coronillae* to synonymy with *F. aesculi*. As shown in the current study, an isolate of *M. coronillae* (CBS 769.71) was indistinguishable from *F. aesculi* based on sequence data. However, this isolate produced pycnidial paraphyses in culture, which had not been observed on the type specimen of *F. aesculi* (Sutton 1980). Phillips (2000) used this feature to distinguish *F. populi* A.J.L. Phillips from *F. aesculi*. DNA sequence comparisons revealed, that this feature is uninformative at the species level (*F. aesculi* = *F. populi*), and that not all strains of *F. aesculi* form paraphyses. Furthermore, some pycnidia in the culture CBS 769.71 produced numerous paraphyses, while they were almost completely absent in others. A note by H.A. van der Aa in the CBS database mentions the fact that this...
The asexual, sterile basidiomycete genus *Rhizoctonia* DC. has been linked to a number of teleomorphs in different families, namely the *Platygloeaceae*, *Exidiaceae*, *Tulasnellaceae*, *Ceratobasidiaceae*, etc. (Stalpers & Andersen 1996). The reason these species are treated in the form-genus *Rhizoctonia*, is that they are mostly sterile, and share the same vegetative features typical of *Rhizoctonia* spp. Species have thus been distinguished based on features such as sclerotium production, mycelial colour, wide hyphae, all with two or several nuclei, length of cells, shape and size of monilioid cells, and sclerotial size (Parmeter & Whitney 1970).

*Macrophomina phaseolina* is the type species of the genus *Macrophomina* Petr., and is also the name given to the coelomycete synanamorph of *Rhizoctonia bataticola* (Taubenh.) E.J. Butler. The latter fungus is a root inhabitant, and has been implicated in numerous root rot diseases of a wide range of crops, commonly causing charcoal rot and ashy stem blight (Holliday & Punithalingam 1998). The name of the synanamorph, and its taxonomic placement, has been the topic of much controversy. In their IMI description sheet of the fungus, Holliday & Punithalingam (1988) mention that the conidiophores are hyaline phialides, short obpyriform to cylindrical, 5–13 × 4–6 µm. Conidia are described as being hyaline, ellipsoid to obvoid, 14–30 × 5–10 µm. Von Arx (1981) introduced the name *Tiarosporella phaseolina* (Tassi) van der Aa for this fungus, and also reduced the genus *Macrophomina* to synonymy under *Tiarosporella* Höhn. As no explanation was given, this treatment has largely been ignored by the plant pathological and mycological community, though von Arx (1987) retained it. The genus *Tiarosporella* [based on *T. paludosa* (Sacc. & Fiori ex P. Syd.) Höhn.] is characterised by having conidia formed from smooth, hyaline conidiogenous cells that lack periclinal thickenings and percurrent proliferations, and hyaline, subcylindrical to fusiform conidia that have irregular apical mucoid appendages (Nag Raj 1993). H.A. van der Aa (pers. comm.) informed the first author that the decision was based upon the fact that he had succeeded to induce sporulation of *M. phaseolina* in culture, and observed that the conidia have apical mucoid appendages, which prompted von Arx to place this fungus in *Tiarosporella*. In the present study we have been able to induce numerous strains of *M. phaseolina* to sporulate on sterile pine needles, and found that conidia form apical mucoid appendages (Figs 5–6) as previously suggested by van der Aa. As this is inconsistent with the description provided by Holliday & Punithalingam (1988), we provide an emended description below:

*Macrophomina phaseolina* (Tassi) Goid., Annali Sper. agr. N.S. 1: 457. 1947. Figs 5–6.

*Basionym:* *Macrophoma phaseolina* Tassi, Bull. Lab. Ort bot. Siena IV: 9. 1901.

= *Tiarosporella phaseoli* (Maubl.) Aa, Verh. Kon. Ned. Akad. Wetensch., Sectie 2, 68: 4. 1977.

Additional synonyms listed by Holliday & Punithalingam (1988).

Sclerotia occurring in host tissue or in soil, black, smooth, hard, 100–1000 µm diam. *Conidiomata* pycnidial, dark brown to black, solitary or gregarious, up to 200 µm diam, opening by a central ostiole; wall multilayered, cells dark brown, thick-walled. *Conidiogenous cells* lining the inner surface of the conidioma, hyaline, short obpyriform to subcylindrical, proliferating several times percurrently near the apex, 6–12 × 4–6 µm; young conidiogenous cells covered by a mucous layer that extends over the apex of the developing conidiom. *Conidia* ellipsoid to obvoid, (16–)20–24(–32) × (6–)7–9(–11) µm; immature conidia hyaline, enclosed in a mucous sheath, that upon dehiscence encloses the top half of the conidiom, transformed into two lateral tentaculiform, apical mucoid appendages (type C, Nag Raj 1993); mature conidia becoming medium to dark brown, with a granular outer layer that in some cases appears pitted, without any mucoid appendages; conidial hilum frequently with a marginal frill.

**Notes:** Although *Macrophomina phaseolina* can have conidia with apical mucoid appendages as found in *Tiarosporella* (Sutton & Marasas 1976), it is distinguished by having percurrently proliferating conidiogenous cells [not seen in any species of *Tiarosporella sensu* Nag...
Rag (1993), nor in those investigated here, and conidia that become dark brown at maturity. Based on these differences (and in the absence of authentic cultures of T. paludosa), the genus Macrophomina and the name M. phaseolina is retained. The three species of Tiarosporella that were available for this study clustered in Clade 1 (Figs 7–8), suggesting that the latter clade is still unresolved.

Clade 4: Neoscytalidium dimidiatum (teleomorph unknown)
Several distinct Fusicoccum-like fungi with conidia that become septate with a darker central cell have been treated under the epithet “mangiferae”. Some isolates, however, formed a Scytalidium-like synanamorph, while others did not.

Dothiorella mangiferae Syd. & P. Syd. was originally described from mango (Sydow et al. 1916). Nattrass (1933) described a similar fungus from pome and stone fruit trees, but noticed a pigmented conidial state, which led him to describe Hendersonula toruloidea Nattrass. Sutton & Dyko (1989) revised the genus Hendersonula Speg. and synonymised both D. mangiferae and H. toruloidea under the redescribed Nattrassia mangiferae (Syd. & P. Syd.) B. Sutton & Dyko. Furthermore, the mycelial synanamorph was described as Scytalidium dimidiatum (Penz.) B. Sutton & Dyko. Farr et al. (2005) made the point that the oldest name for the fungus was Torula dimidiata Penz., and hence introduced the combination Fusicoccum dimidiatum (Penz.) D.F. Farr, stating that the type species of the genera Nattrassia and Scytalidium were synonyms of F. dimidiatum. As seen in the present study, this fungus, with its powdery disarticulating aerial mycelium, is a genus in its own right within the Botryosphaeriaceae. Furthermore, the ex-type strain of Scytalidium, S. lignicola Pesante, (CBS 233.57) clusters outside the Botryosphaeriaceae (results not given), and hence Scytalidium is unavailable for this fungus.

Fusicoccum mangiferae (Syd. & P. Syd.) Johnson, Slippers & M.J. Wingf. (≡ D. mangiferae, ≡ N. mangiferae) is a distinct taxon (see Clade 6, Neofusicoccum) that should not be confused with F. dimidiatum. When Sutton & Dyko (1989) and Johnson (1992) re-examined the type of D. mangiferae, they did not observe the Scytalidium-like anamorph on the type specimen, in accordance with Sydow et al. (1916). Slippers et al. (2005) studied isolates identified as D. mangiferae (= N. mangiferae) from mango in Australia, and found them to belong to Fusicoccum, for which they introduced the name F. mangiferae (now Neofusicoccum, Clade 6). They also did not observe the Scytalidium-like synanamorph as described by Sutton & Dyko (1989). The synonymy of H. toruloidea (which has a Scytalidium-like synanamorph) with F. mangiferae (which does not appear to have a Scytalidium-like synanamorph), is thus rejected here. A new genus is proposed to accommodate this fungus.

Neoscytalidium Crous & Slippers, gen. nov. MycoBank MB500868.
Genus anamorphosis hyphomycetum. Arthroconidia catenata in mycelio aero, pulvulenta, disarticulanta, cylindrica-truncata, oblongo-obtusa vel doliformia, fusca, crassitunicata, 0–2-septata.

Conidia occurring in arthric chains in aerial mycelium, powdery to the touch, disarticulating, cylindrical–truncate, oblong–obtuse to doliform, dark brown, thick-walled, 0–2-septate.

Type species: Neoscytalidium dimidiatum (Penz.) Crous & Slippers, comb. nov.

Neoscytalidium dimidiatum (Penz.) Crous & Slippers, comb. nov. MycoBank MB500869. Fig. 9. Basionym: Torula dimidiata Penz., Michelia 2: 466. 1882 (basionym; hyphomycete synanamorph).

≡ Scytalidium dimidiatum (Penz.) B. Sutton & Dyko, Mycol. Res. 93: 484. 1989.
≡ Fusicoccum dimidiatum (Penz.) D.F. Farr, Mycologia 97: 740. 2005.
≡ Hendersonula toruloidea Nattrass, Trans. Brit. Mycol. Soc. 18: 197. 1933 (coelomycete synanamorph).

Clade 5: Dothidotthia (anamorph Dothiorella)
Concepts defining morphological features of Botryosphaeria (ascomata, asci and hamathecium) have developed slowly (Denman et al. 2000). This has resulted in confusion between Botryosphaeria and superficially similar genera such as Physalospora Niesl. and Guignardia (von Arx & Müller 1954, Hanlin 1990), Auerswaldiella Theiss. & Syd., Discococha Höhn., Dothidotthia Höhn., Neoedightonia C. Booth, Homostegia Fuckel and Otthia Nitschke. Subsequent
Fig. 6. *Macrophomina phaseolina* (CPC 11052). A. Conidia and sclerotia formed on pine needles. B. Conidiogenous cells. C–H. Hyaline conidia with apical, mucoid appendages. I–M. Brown, mature conidia with verrucose walls. Scale bars = 8 µm.

Fig. 7. *Tiarosorella graminis* var. *karoo* (CBS 118718). A. Pycnidium on a pine needle. B–C. Conidiogenous cells. D–G. Conidia (arrows denote apical, mucoid appendages). H. Conidia. Scale bars: A = 200 µm, B–D = 7 µm.
Fig. 8. Tiarosporella tritici (CBS 118719). A. Pycnidia on a pine needle. B–D. Conidiogenous cells. E–F. Conidia (arrows denote apical, mucoid appendages). Scale bars: A = 300 µm, B–F = 10 µm.

Fig. 9. Neoscytalidium dimidiatum (CBS 312.90). A–B. Neoscytalidium conidia. C. Conidiogenous cells of coelomycete synanamorph. D–F. Conidia of coelomycete synanamorph (arrows denote sheath). Scale bars = 4 µm.

Fig. 10. Dothiorella pyrenophora (K 54912). A–B. Pycnidia on stems. B. Spermatia and spermatogenous cells. D–H. Conidia. Scale bars = 10 µm.
to the review of Denman et al. (2000), two of these genera, namely *Otthia* and *Dothidotidia*, have been variously treated, and they are discussed here.

The genus *Otthia* was described as having short-stalked, cylindrical, bitunicate asci containing hyaline ascospores that become brown and 1-septate at maturity (Dennis 1981, Sivanesan 1984). Booth (1958) designated *Otthia spiraeae* (Fuckel)Fuckel as lectotype of the genus, citing *Diplodia sarmentorum* (Fr.)Fr. as anamorph. Von Arx (1974) listed *Otthia* as the teleomorph of *Aplosporella* Speg., which is indistinguishable from *Sphaeropsis* Sacc., and thus similar to *Diplodia sarmentorum*. Denman et al. (2000) were thus of the opinion that *Otthia* should be reduced to synonymy with *Botryosphaeria*, but stated that further morphological and DNA sequence comparisons were first required. Van Niekerk et al. (2004) showed that the Wollenweber isolate of *Diplodia sarmentorum* (CBS 120.43) is identical to the Booth isolate of *Otthia spiraeae* (IMI 63581b). Recently, Phillips et al. (2005a) re-examined the lectotype of *O. spiraeae* (K 104853), and found that it represents a fungus distinct from that collected and treated by Booth (1958) as "*O. spiraeae*". These authors also showed that the anamorph-teleomorph connection reported by Booth (1958), was in fact incorrect. Consequently, the new species, *Botryosphaeria sarmentorum* A.J.L. Phillips, Alves & Luque (anamorph: *Dothiorella sarmentorum* (Fr.) A.J.L. Phillips, Alves & Luque) was introduced for the fungus treated by Booth (1958) and Wollenweber (1941). The incorrect link between *Otthia* and "*Diplodia* sarmentorum" as reported by Booth (1958) was thus resolved. *Otthia* was retained as a distinct, but poorly known genus, characterised by cylindrical asci, brown, 1-septate ascospores that are obliquely uniseriate, and thin, sparingly septate pseudoparaphyses. Due to the lack of cultures, the correct placement of *Otthia* remains unknown.

In the phylogenetic analysis of DNA sequence data of the ITS region and EF-1α gene, Phillips et al. (2005a) demonstrated that the "*Botryosphaeria*" species with pigmented, septate ascospores and *Diplodia*-like anamorphs formed a separate clade. *Dothiorella* had been reduced to synonymy under *Diplodia* by Crous & Palm (1999), who did not have access to cultures and DNA sequence data and thus used a wider morphological concept for *Diplodia*. Phillips et al. (2005a) re-examined the type of *Dothiorella pyrenophora* Sacc. (K 54912) (Fig. 10), and stated that it differs from *Diplodia* by having conidia that are brown and 1-septate early in their development, while they are still attached to the conidiogenous cells. In *Diplodia* (*D. mutila*), conidial darkening and septation takes place after discharge. We have re-examined the types of both genera in the present study, and concur with Phillips et al. (2005a) that the genera have distinct conidial characteristics. Teleomorphs of *Dothiorella* have pigmented, septate ascospores, for which the genus *Dothidotidia* is available. The latter genus had been recognised as a member of the *Botryosphaeriaceae* by Barr (1987, 1989), and is known to have *Diplodia*-like anamorphs, which are now accommodated in *Dothiorella*. The new taxa described by Phillips et al. (2005a) must thus be placed in *Dothidotidia*. These taxa will be treated elsewhere (A.J.L. Phillips, in prep.).

**Clade 6: Neofusicoccum (telemorph Botryosphaeria-like)**

An interesting issue to resolve is the morphological distinction between the two larger *Fusicoccum* clades, namely *Fusicoccum s. str.* (based on *F. aesculi*, and linked to the name *Botryosphaeria*), and the larger *Fusicoccum*-like clade (linked to *Botryosphaeria*-like teleomorphs), which includes most of the species that are currently known from DNA sequence data. Although the teleomorphs are similar in both clades, their anamorphs provide some clues for a possible separation. There is little to choose in their *Fusicoccum* anamorphs, but as seen in *Sacchararia*, the distinguishing feature is to be found in their synanamorphs. In *Fusicoccum s. str.* (*F. aesculi*) conidia are fusiform to ellipsoid, and with age turn septate and brown. In the *Fusicoccum*-like clade, two distinct conidial types are seen, namely the first with a *Fusicoccum*-like morphology, which can turn brown and septate with age (as seen in Clade 2). The second conidial form has globose to pyriform conidia, which are brown, slightly verruculose, and muriformly septate. It is debatable if *Fusicoccum s. str.* (Clade 2) forms a distinct synanamorph. The synanamorph observed in the *Fusicoccum*-like clade (Clade 6) cannot be accommodated in *Camarosporium* (the stromal analogue of *Dichomera*), as the type species of *Camarosporium*, *C. quaternatum* (CBS 134.97, 483.95) clusters outside the *Botryosphaeriaceae*. As far as we could establish, however, no genus is presently available for this clade, and thus a new one is proposed below. We introduce a single generic name, namely for the anamorph (which occurs with a *Dichomera*-like synanamorph), which is the more informative morphological state:

*Neofusicoccum* Crous, Slippers & A.J.L. Phillips, gen. nov. MycoBank MB500870.

**Telemorph: Botryosphaeria-like**

**Synanamorph: Dichomera-like**

Genus anamorphosis coelomyceticum. Fusicoceco similis sed synanamorpha Dichomerae similis et conidios bruneus, globosis vel pyriformibus, distinguendum.

Resembling species of *Fusicoccum*, but distinct in forming a *Dichomera*-like synanamorph with brown, globose to pyriform conidia.

**Type species:** *Neofusicoccum parvum* (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips, comb. nov.

Most species that have thus far been described in *Fusicoccum* appear to reside in this clade, as can be seen in the present, as well as other recent studies. To facilitate clarity, new combinations are proposed for those known to us from culture:

**Neofusicoccum andinum** (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M.J. Wingf. comb. nov. MycoBank MB500871.

*Basionym:* *Fusicoccum andinum* Mohali, Slippers & M.J. Wingf., Mycol. Res. 10: 408. 2006.
**Neofusicoccum arbuti** (D.F. Farr & M. Elliott) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500872.

_Basionym:_ Fusicoccum arbuti D.F. Farr & M. Elliott, Mycologia 97: 731. 2005.

**Neofusicoccum australi** (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500873.

_Basionym:_ Fusicoccum australi Slippers, Crous & M.J. Wingf., Mycologia 96: 1035. 2004.

_Teleomorph:_ "Botryosphaeria" australis Slippers, Crous & M.J. Wingf., Mycologia 96: 1035. 2004.

**Neofusicoccum eucalypticola** (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500874.

_Basionym:_ Fusicoccum eucalypticola Slippers, Crous & M.J. Wingf., Stud. Mycol. 50: 351. 2005.

_Teleomorph:_ "Botryosphaeria" eucalypticola Slippers, Crous & M.J. Wingf., Stud. Mycol. 50: 351. 2005.

**Neofusicoccum eucalyptorum** (Crous, H. Sm. ter & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500875.

_Basionym:_ Fusicoccum eucalyptorum Crous, H. Sm. ter & M.J. Wingf., Mycologia 93: 280. 2001.

_Teleomorph:_ "Botryosphaeria" eucalyptorum Crous, H. Sm. ter & M.J. Wingf., Mycologia 93: 280. 1998.

**Neofusicoccum luteum** (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500876.

_Basionym:_ Fusicoccum luteum Pennycook & Samuels, Mycotaxon 24: 456. 1985.

_Teleomorph:_ "Botryosphaeria" lutea Phillips, Sydowia 54: 59. 2002.

**Neofusicoccum macroclavatum** (Burgess, Barber & Hardy) Burgess, Barber & Hardy, **comb. nov.** MycoBank MB500877.

_Basionym:_ Fusicoccum macroclavatum Burgess, Barber & Hardy, Austral. Pl. Pathol. 34: 562. 2005.

_Note:_ For further information about this recently published species, see Burgess et al. (2005).

**Neofusicoccum mangiferae** (Syd. & P. Syd.) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500878.

_Basionym:_ Dothiorella mangiferae Syd. & P. Syd., Ann. Mycol. 14: 192. 1916.

≡ Natrassia mangiferae (Syd. & P. Syd.) B. Sutton & Dyko, Mycol. Res. 93: 484. 1989.

≡ Fusicoccum mangiferae (Syd. & P. Syd.) Johnson, Slippers & M.J. Wingf., Mycologia 97: 106. 2005.

_Teleomorph:_ "Botryosphaeria" sp.

**Neofusicoccum parvum** (Pennycook & Samuels) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500879.

_Basionym:_ Fusicoccum parvum Pennycook & Samuels, Mycotaxon 24: 455. 1985.

_Teleomorph:_ "Botryosphaeria" parva Pennycook & Samuels, Mycotaxon 24: 455. 1985.

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**Fig. 11. Pseudofusicoccum stromaticum** (CBS 117448). A. Pycnidia on pine needles. B–C. Conidiogenous cells. D–H. Conidia (arrows denote mucoid sheaths). Scale bars = A = 90 µm, B = 6 µm.
Neofusicoccum protearum (Denman & Crous) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500880.

_Basionym:_ Fusicoccum protearum Denman & Crous, Mycologia 95: 301. 2003.

_Teleomorph:_ “Botryosphaeria” protearum Denman & Crous, Mycologia 95: 301. 2003.

Neofusicoccum ribis (Slippers, Crous & M.J. Wingf.) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500881.

_Basionym:_ Fusicoccum ribis Slippers, Crous & M.J. Wingf., Mycologia 96: 96. 2004.

_Teleomorph:_ “Botryosphaeria” ribis Grossenb. & Duggar, Tech. Bull. N.Y. Agric. Exp. St. 18: 128. 1911.

Neofusicoccum viticlavatum (Niekerk & Crous) Crous, Slippers & A.J.L. Phillips, **comb. nov.** MycoBank MB500882.

_Basionym:_ Fusicoccum viticlavatum Niekerk & Crous, Mycologia 96: 792. 2004.

Clade 7: “Fusicoccum” stromaticum (teleomorph unknown)

“Fusicoccum” stromaticum Mohali, Slippers & M. J. Wingf. (Mohali _et al._ 2006) was described for a new _Fusicoccum_-like species occurring on _Eucalyptus_ and _Acacia_ spp. in Venezuela. The taxon was distinguished from other species of _Fusicoccum_ based on its unusually large conidiomata, the ability to grow at 35 °C, and thick-walled conidia. Strains of this species have conidia that are encased in a persistent mucous sheath, which is absent in other species of _Fusicoccum_, and is the character that distinguishes it as genus from _Fusicoccum_ s.str.

_Pseudofusicoccum_ Mohali, Slippers & M.J. Wingf. **gen. nov.** MycoBank MB500884.

Genus anamorphosis coelomyceticum. Fusicocco simile, sed conidiis strato mucido persistente circumdatis distinguendum.

Resembling species of _Fusicoccum_, but distinct in having conidia encased in a persistent mucous sheath.

_Type species:_ Pseudofusicoccum stromaticum (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M. J. Wingf., **comb. nov.**

_Pseudofusicoccum stromaticum_ (Mohali, Slippers & M.J. Wingf.) Mohali, Slippers & M.J. Wingf., **comb. nov.** MycoBank MB500885. Fig. 11.

_Basionym:_ Fusicoccum stromaticum Mohali, Slippers & M.J. Wingf., Mycol. Res. 110: 408. 2006.

Clade 8: “Botryosphaeria” quercuum (anamorph Diplodia-like)

Von Arx & Müller (1954) placed _B. melanops_ (Tul. & C. Tul.) G. Winter under _B. quercuum_ (Schwein.) Sacc., which is a complex in need of revision, including _Melanops tulasnei_ (Tul. & C. Tul.) Fuckel, the oldest generic name available for this clade. To resolve the status of _B. quercuum_, however, authentic cultures of all these names need to be studied, and linked to existing names. The strain in Clade 8 closely matches the morphology of the type specimen (Phillips _et al._, unpubl. data). No name is, however, proposed for this genus pending the outcome of studies based on authentic isolates.

Clade 9: Saccharata (anamorph Fusicoccum-like)

Wakefield (1922) described an ascomycete associated with leaf spots and stem cankers of _Protea_ and _Leucospermum_ species as _Phyllachora proteae_ Wakef. This latter fungus is characterised by having unilocular ascomata that develop under a very small epidermal clypeus, cylindrical asci, pseudoparaphyses, and hyaline, aseptate, ellipsoidal ascospores. Doidge (1942) found the ascomatal walls are continuous with, and similar in structure to the clypeus, and stated that the fungus should be allocated elsewhere, possibly in _Botryosphaeria_. Denman _et al._ (1999) recollected this species, placed it in _Botryosphaeria_, and also established a cultural link with its anamorph, “Fusicoccum” proteae Denman & Crous, which also

Fig. 12. Camarosporium quaternatum (CBS 134.97). A. Pycnidia in agar. B–C Conidiogenous cells. D–E. Conidia. Scale bars: A = 100 µm, B = 6 µm.
forms a Diplodia-like synanamorph in culture. On the basis of ITS DNA sequence comparisons, Denman et al. (2000) later showed that the fungus clustered outside the Botryosphaeria clades that accommodated Fusicoccum and Diplodia anamorphs. Given its unilocular ascomata, the presence of a clypeus, and its unusual Fusicoccum- and Diplodia-like synanamorphs, Crous et al. (2004) established a new genus, Saccharata Denman & Crous to accommodate S. proteae (Wakef.) Denman & Crous. The LSU phylogenies in this study support Saccharata as a distinct genus that is basal to, but probably outside the Botryosphaeriaceae.

Clade 10: Guignardia (anamorph Phyllosticta)
The genus Phyllosticta was revised by Van der Aa & Vanev (2002), who treated 2936 taxa, accepting 143 species in Phyllosticta, many of which have teleomorphs in Guignardia (Botryosphaeriaceae), and Leptodothiorella spermatial states. As seen with “Phyllosticta” flevolandica Aa, it is to be expected that more “Phyllosticta” taxa will be allocated elsewhere once subjected to DNA analysis (Crous et al. in prep.).

Clade 11: Camarosporium and relatives
Several morphologically discordant taxa group in this clade, including C. quaternatum, the type species of Camarosporium (Fig. 12), but also an unidentified Dothiorella-like strain (1-septate, brown conidia), and another seen as an atypical Phyllosticta, namely P. flevolandica (hyaline, 0–1-septate conidia) (Van der Aa 1973). Other taxa include Karstenula Spec. (anamorph Microdiplodia Tassi), Letendrea Sacc. (Tubeufiaceae?), and Byssothecium Fuckel (anamorph Chaetophoma-like). It appears that the Dothiorella-like strain (CPC 12268) is, in fact, a species of Microdiplodia. Sutton (1977) reported that the genus was introduced for stem- or branch-inhabiting species with small, brown, 1-septate conidia. This is consistent with the results of this study where the strain examined was isolated from branches of Sophora chrysophylla collected in Hawaii. As we have shown here that the genus Microdiplodia Tassi could be used for these small-spored Dothiorella-like species, the genus needs to be lectotypified according to Tassi’s original concept (see Sutton 1977).

Clade 12: Stenocarpella (teleomorph unknown)
The genus Stenocarpella Syd. & P. Syd. is based on S. macrospora (Earle) B. Sutton. The genus contains two species that cause Diplodia ear rot of maize, namely S. macrospora and S. maydis (Berk.) B. Sutton. These taxa were formerly treated in Diplodia by Sutton (1964), and Macrodiplodia Sacc. by Petrak & Sydow (1927). Sutton (1964, 1980) was of the opinion that these species should be accommodated in a genus other than Diplodia, and because the status of Macrodiplodia was unknown, he placed them in Stenocarpella (Sutton 1977). This treatment has not been widely accepted, and plant pathologists refer to “Diplodia ear rot”, and continue to use the Diplodia names.

Fig. 13. Stenocarpella maydis on Zea mays. A. Pycnidium on upper leaf surface. B–C. Conidial cirri. D. Conidiogenous cells giving rise to conidia. E–H. Conidia. Scale bars = 10 µm.
Because cultures were not available for *S. maydis* and *S. macrospora*, these two species were recollected as part of the present study, and subjected to DNA sequence comparisons. Interestingly, they clustered in the *Diaporthales*, clearly supporting the decision by Sutton (1964, 1980) to move them to their own genus, *Stenocarpella*. Species of *Diplodia* are quite variable in morphology, and hence it is difficult to see immediately which morphological features separate *Stenocarpella* from *Diplodia*. Species of *Stenocarpella* tend to have unicellular, thick-walled pycnidia with walls of brown *textura angularis*. Under moist conditions, they exude a long cirrhus of brown conidia via a central ostiule (reminiscent of *Phaeophilespora* Rangel). Conidiogenous cells are phialidic, thin-walled, subcylindrical to narrowly obclavate (Fig. 13), thus different from typical fusoid to ellipsoid, thick-walled conidia of *Diplodia*. To facilitate future research with these pathogens, epitype specimens and cultures are designated below:

**Stenocarpella maydis** (Berk.) B. Sutton, in Sutton, *The coelomycetes, fungi imperfecti with pycnidia, acervuli and stromata* : 432. 1980.

*Basionym:* *Sphaeria maydis* Berk., Hooker’s J. Bot. London 6: 15. 1847.

Additional synonyms listed in Sutton (1980).

Types: **U.S.A.** Salem, Bethlehem, *Zea mays*, Schweinitz, 1832, IMI 96546 *isotype. South Africa*, KwaZulu-Natal, Simlantengtsha, *Bt Zea mays* hybrid from 2003-04 season, J. Rheeder, *epitype designated here*. CBS 117558, preserved as freeze-dried inactive strain (= MRC 8613): ibid. CBS 117557 = MRC 8612; Hlabisa, commercial hybrid PAN-6043, MRC 8614 = CBS 117559.

**Stenocarpella macrospora** (Earle) B. Sutton, Mycol. Pap. 141: 202. 1977. Fig. 13.

*Basionym:* *Diplodia macrospora* Earle, Bull. Torrey Bot. Cl. 24: 29. 1897.

Additional synonyms in Sutton (1980).

Types: **U.S.A.** Alabama, Auburn, *Lee Co.*, F.S. Earle, 1896, IMI 12790, *isotype. South Africa*, KwaZulu-Natal, Hlabisa, rain damaged *Bt Zea mays* hybrid, 2003-04 season, J. Rheeder, *epitype designated here*. CBS 117560, preserved as freeze-dried inactive strain (= MRC 8815).

This study provides a framework to align the taxonomy of the *Botryosphaeriaceae* with the phylogenetic lineages within the group. It also highlights the previously unrealised morphological and evolutionary complexity of the group. Specific studies are now needed to clarify some remaining, and arising, taxonomic and phylogenetic questions within this family. To resolve remaining taxonomic uncertainties, epitypes of key species, representing the oldest names in the respective groups, will need to be collected, studied and designated. To resolve the phylogenetic uncertainties (e.g. Clade 1 and *B. mambane*) sequences for additional gene regions (to add more informative sites, and from unlinked loci) will have to be added. Specific studies focussing on the phylogenies within the clades, and expanding on the current set of available cultures (e.g. Clades 1, 10 and 11), will add valuable information on the evolution within these groups, and also help identify definitive morphological characters.

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