Abcb1 in Pigs: Molecular cloning, tissues distribution, functional analysis, and its effect on pharmacokinetics of enrofloxacin

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P-glycoprotein (P-gp) is one of the best-known ATP-dependent efflux transporters, contributing to differences in pharmacokinetics and drug-drug interactions. Until now, studies on pig P-gp have been scarce. In our studies, the full-length porcine P-gp cDNA was cloned and expressed in a Madin-Darby Canine Kidney (MDCK) cell line. P-gp expression was then determined in tissues and its role in the pharmacokinetics of oral enrofloxacin in pigs was studied. The coding region of pig Abcb1 gene was 3,861 bp, encoding 1,286 amino acid residues (Mw = 141,966). Phylogenetic analysis indicated a close evolutionary relationship between porcine P-gp and those of cow and sheep. Pig P-gp was successfully stably overexpressed in MDCK cells and had efflux activity for rhodamine 123, a substrate of P-gp. Tissue distribution analysis indicated that P-gp was highly expressed in brain capillaries, small intestine, and liver. In MDCK-pAbcb1 cells, enrofloxacin was transported by P-gp with net efflux ratio of 2.48 and the efflux function was blocked by P-gp inhibitor verapamil. High expression of P-gp in the small intestine could modify the pharmacokinetics of orally administrated enrofloxacin by increasing the Cmax, AUC and Ka, which was demonstrated using verapamil, an inhibitor of P-gp.

P-glycoprotein (P-gp, encoded by the Abcb1 gene) is an important ATP-dependent efflux transporter, which was initially identified in Chinese hamster ovary cells in 1976. P-gp has been documented to be highly expressed in the intestine, liver, and kidney of humans and rodents and as such has essential roles in bioavailability and drug-drug interaction of substrate xenobiotics. P-gp has been shown to recognize structurally and pharmacologically diverse compounds, including drugs widely used in veterinary medicine (e.g. ivermectin, macrolides, and fluoroquinolones); thus, the relevance of P-gp in veterinary medicine and drug development is significant. However, little information is available for veterinarians on oral absorption, elimination and drug-drug interactions related to P-gp.

The complete cDNA of P-gp from human, mouse, rat, ovine, dog, feline and the Salmon louse have been cloned. Pig is an important model species for veterinary medicine, and it is commonly used in toxicological and pharmacological studies. Until now, information about porcine P-gp is lacking, and expression and function of P-gp in pharmacokinetically important tissue is scarce. Here, full-length cDNA for porcine P-gp was cloned and a Madin-Darby Canine Kidney (MDCK) cell line that stably expressing pig P-gp was established. Its expression in tissue, as well as its participation in enrofloxacin pharmacokinetics, was assessed in pigs.

Results

Sequence analysis of porcine P-gp. Based on sequence homology, three gene fragments of porcine Abcb1(971, 2480 and 712 bp) (Fig. 1A) were amplified (Fig. 1B) using primers depicted in Table 1. Fusion PCR was then performed to obtain full-length cDNA (Fig. 1C). The Abcb1 gene was confirmed by DNA sequencing and BLASTN analysis at NCBI. The cDNA was 4,060 bp, consisting of a 5'-untranslated region of 66 bp, an uninterrupted open reading frame of 3,861 bp, and a 3'-untranslated region of 133 bp. The sequence was submitted to GenBank and assigned an accession number (GenBank ID: KP233220). The nucleotide sequence shared 89, 88, 89 and 82% identities with that of cow (XM590317.7), sheep (NM001009790.1), human (NM000927.4) and...
mouse (NM011075.2), respectively. The deduced amino acid sequence of porcine P-gp was 1,286 amino acid residues in length and the estimated molecular weight was 141.966 kDa and the theoretical isoelectric point was

Table 1. Primers used in this study.

| Fragments of Abcb1 | Sequences (5′-3′) | Length (bp) |
|--------------------|------------------|-------------|
| F1                 | F: GTCTGCCCTCTCTCTCTCCAAAAATC <br> R: GCACCAATAAGAATGTTGGCTGTAATAGC |
|                    | 971              |
| F2                 | F: GAAGAAGCCAAAGGATTGGAATAAAGAAA <br> R: CACTGGACGCTTCAGTTTTTTAATCTCCC |
|                    | 2,480            |
| F3                 | F: GTGCTAATGGAGCCAGGGAGATTGA <br> R: GTGGAATGATCTTCAATGGTAGGAGG |
|                    | 712              |

| Genes | Sequences (5′-3′) | Length (bp) |
|-------|------------------|-------------|
| Abcb1 | F: AGTCTAATAAGAAGAGGAT <br> R: GCCATTCTGTTAATTCA |
|       | 145              |
| Gapdh | F: GAAGGTCGGAGTGAAAGGAT <br> R: CATTGCTAGATCATCTGGAACA |
|       | 149              |

| Genes | Sequences (5′-3′) | Length (bp) |
|-------|------------------|-------------|
| Abcb1 | F: GCACCATGCTGCTGATCTTGAAGAAGGCG <br> R: GCAACTAGCTAGCTAGCTGAGAGG |
|       | 3,861            |
8.99. The protein sequence of porcine P-gp shared 89, 88 and 87% identity to that of human (NP_000918.2), cow (XP_590317.6) and sheep (NP_001009790.1), respectively (Fig. 2A). Unique to the sequences of P-gp in ruminants (sheep and cow) is a 4-amino acids deletion at position 22–25 of the amino acid sequence.

A phylogenetic tree was constructed using the neighbor-joining (NJ) method with a 1,000 bootstrap test based on the multiple alignments, which indicated that the selected 14 protein sequences of P-gp were clustered into two groups: avian and mammal. The porcine P-gp was located in vertebrates group and was closely related to cow and sheep, suggesting a close evolutionary relationship between them (Fig. 2B).

**Protein structural model of porcine P-gp.** Porcine P-gp was predicted to possess 12 transmembrane helices with extracellular N- and C-termini (Fig. 3A) and comprised four structural domains: two cytoplasmic domains containing the nucleotide-binding domains (NBD) and another two hydrophobic transmembrane domains (TMD), which was similar to human P-gp (see Fig. 3). The potential putative N-glycosylation sites of
Porcine and human P-gp was marked in red (Fig. 3), indicating seven in the porcine P-gp, whereas there were ten in human P-gp. The tertiary structure of porcine P-gp contained 42 α-helices and 21 β-strands (Fig. 4A). Fourteen binding site residues were found to be located in TM5 (Tyr-308), TM6 (Phe-337, Leu-340, Ile-341, Phe-344), TM7 (Gln-726, Phe-729, Ser-730, Phe-733), TM11 (Tyr-954) and TM12 (Phe-979, Ser-980, Val-983, Ala-986) in the porcine P-gp (Fig. 4B,C).

**Establishment and characterization of an MDCK cell line stably-transfected with porcine Abcb1.** The pig Abcb1 gene was amplified using primers shown in Table 1 with XhoI and XbaI sites, respectively, and a recombinant pcDNA3.1-pAbcb1 plasmid was constructed. MDCK cells were transfected with pcDNA3.1-pAbcb1. A single cell colony resistant to G418 was selected as a stable transfectant (MDCK-pAbcb1). The presence of Abcb1 was confirmed with qRT-PCR and western blot (Fig. 5A,B). The accumulation of Rho123 (substrate of human P-gp) was measured to investigate whether MDCK-pAbcb1 cell line has enhanced porcine P-gp protein function. Figure 5C,D show that MDCK-pAbcb1 cells accumulated less Rho123 than MDCK cells (control), indicating a significantly enhanced extruding capacity of Rho123 in MDCK-pAbcb1 cells (p < 0.01). Furthermore, verapamil (100 μM), an inhibitor of human P-gp, significantly (p < 0.01) increased Rho123 accumulation in MDCK-pAbcb1 cells, demonstrating that such an accumulation could be reversed by verapamil.

**Expression of P-gp in pigs.** Porcine P-gp mRNA and protein were analyzed by qRT-PCR and Western blot, respectively. As shown in Fig. 6A, Abcb1 mRNA was detected in all tested tissues. In the CNS, the brain capillaries had the most Abcb1 mRNA and the cerebellum had the least. Abcb1 mRNA levels in the cerebral cortex, cerebellum, midbrain, hypothalamus and hippocampus was very low. In the peripheral tissues, Abcb1 were highly expressed in the jejunum, ileum, colon and liver. Abcb1 mRNA transcription in the ileum was greater than in the cecum and kidney (p = 0.005, p = 0.001) and significantly higher than that in the duodenum and rectum.

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**Figure 3.** Secondary structure prediction of porcine P-gp (A) and human P-gp (B). Potential N-glycosylation sites of P-gp are colored with red (indicating position 87, 104, 297, 495, 705, 810 and 1035 in porcine P-gp sequence those differ from human P-gp).
Abcb1 mRNA in the jejunum, colon and liver was significantly higher than that in the kidney ($p = 0.046$, $p = 0.018$, $p = 0.030$, respectively).

P-gp protein (shown in Fig. 6C) was ~170 kDa as detected in different pig tissues, indicating that antibody Mdr-1 directed against the human isoform could also recognize porcine P-gp. Protein expression in tissues
appears in Fig. 6B. The most P-gp protein was in the brain capillaries and the least was observed in the hypothalamus. The P-gp protein level was correlated to its mRNA level ($r = 0.863, p < 0.05$).

**The effect of P-gp on in vitro transport and in vivo pharmacokinetics of enrofloxacin.**

Enrofloxacin was measured using HPLC, and the lowest limit of detection (LOD) of enrofloxacin was 0.02 $\mu$g/mL, based on a signal-to-noise ratio > 3. The lowest limit of quantification (LOQ) was 0.05 $\mu$g/mL with a signal-to-noise ratio > 10. The mean percent recovery of enrofloxacin exceeded 81%, and intra-assay and inter-assay reproducibility had a relative standard deviation (RSD) < 11%. Assay linearity was good over 0.05–10 $\mu$g/mL with $r^2 = 0.9999$.

MDCK and MDCK-pAbcb1 cells were grown to confluent monolayers on porous membrane filters, and vectorial transport of the enrofloxacin (12 $\mu$M) across the cells was determined. In the MDCK parental cell line, apically and basolaterally directed translocation of enrofloxacin were similar. However, in the pAbcb1 expressing MDCK cells, $P_{\text{app}}$(BL-AP) was significantly higher than $P_{\text{app}}$(AP-BL) ($P < 0.01$). The P-gp-mediated transport was completely inhibited by its inhibitor verapamil, resulting in a similar efflux ratio, compared to that of the MDCK parental cell line (Table 2). These results showed highly efficient transport of enrofloxacin by porcine P-gp.

To further confirm whether P-gp affects the pharmacokinetics of enrofloxacin in pigs, verapamil (an inhibitor of P-gp) was administrated prior to enrofloxacin, and the plasma concentration-time curve and pharmacokinetics of enrofloxacin are summarized in Fig. 7 and Table 3, respectively. Compared with the control group, $C_{\text{max}}$, AUC and $K_a$ of enrofloxacin were increased, whereas $T_{\text{peak}}$ and $T_{1/2\text{ka}}$ of enrofloxacin were shortened in animals when co-administered with verapamil. In summary, enrofloxacin pharmacokinetics was altered by P-gp inhibition, indicating that porcine P-gp can influence enrofloxacin behavior in vivo.
Oral drug administration is convenient intake route for humans, as well as for animals. The effects of P-gp on drug pharmacokinetics are important for the evaluation of the capacity of oral drug intake in pigs. The localization of P-gp in different tissues in pigs was studied in our previous work15; however, how P-gp functions in pigs is unclear. To address this deficit, we cloned full-length cDNA of porcine P-gp, and characterized the transport activity of P-gp by exogenously expressing the *Abcb1* gene in MDCK cells and measuring the porcine P-gp at the protein and transcriptional levels. Then, we assessed its role in the pharmacokinetics of oral enrofloxacin in pigs. This represents the first cloning of full-length porcine cDNA of P-gp.

The porcine P-gp shared high similarities with P-gps from cow, sheep, human and mouse, suggesting a similar physiological role in these animals. Drug-binding pockets are predicted to reside in transmembrane segments, which formed a translocation path for drug substrates as they exit the membranes16. The nucleotide binding domain may bind ATP or its analogs and can hydrolyze ATP17–20. There is “cross-talk” between nucleotide binding and transmembrane domains as evidenced by cross-linking studies21, and transmembrane domains of porcine P-gp were generated here with Protter protein display software. The prediction showed that porcine P-gp contains twelve transmembrane domains which was similar to human P-gp, but glycosylation site prediction revealed seven potential putative N-linked glycosylation sites on porcine P-gp, fewer than that in human P-gp. Some studies have shown that N-glycosylation can participate in many biological processes such as regulation of intracellular targeting, protein folding and maintenance of protein stability22,23. However, other studies have reported that N-linked glycosylation is not essential for protein expression, plasma membrane localization or overall function.24,25. Further studies are necessary to clarify whether glycosylation affects the function of porcine P-gp.

To better understand how porcine P-gp transports various hydrophobic compounds, we predicted residues that participate in drug binding. Our results indicated that amino acids participating in substrate binding were located at TMD1 sites 5 and 6 and TMD2 sites 7, 11 and 12 of porcine P-gp, whereas previous studies have shown that human P-gp substrate binding sites were located in TMD1 sites 5 and 6 and TMD2 sites 11 and 1226. This may

**Table 3.** Oral enrofloxacin in healthy pigs with/without verapamil (mean ± SEM, n = 4). *P < 0.05, **P < 0.01 significant difference for enrofloxacin in presence/absence of verapamil in healthy pigs. Ke, elimination rate constant; Ka, absorption rate constant; T1/2ka, the absorption half-life; T1/2ke, the elimination half-life; Tpeak, the time to reach peak concentration; Cmax, the peak concentration; AUC, the area under the plasma concentration-time curve; CL/F, Clearance/F, where F is the fraction of dose absorbed; V/F, volume of distribution/F, where F is the fraction of dose absorbed.

**Figure 7.** Mean plasma enrofloxacin in piglets after enrofloxacin administrated alone (10 mg/kg, po) and with verapamil (10 mg/kg, po). Each point represents the mean ± SEM of four piglets.
influence substrate types. Site-directed mutagenesis studies of Abcb1 can be performed to prove whether these residues in porcine P-gp participate in ligand binding. Knowing substrate binding sites may enable development of drugs that bypass recognition of P-gp.

Human colorectal adenocarcinoma cell line (Caco-2) has been approved by the US Food and Drug Administration (FDA) for in vitro transport studies of P-gp27,28. However, pharmacokinetics might be different because of the specific functions of P-gp across different species29. An alternative approach was to establish a porcine P-gp expression system. The MDCK cell line is an approved model for overexpressing Abcb1 originating from other species due to its negligible expression of endogenous transporters30,31. MDCK cell line overexpression porcine P-gp could be used to screen substrate drugs and potential inhibitors or inducers.

The efflux efficiency of P-gp substrate is determined by the P-gp number or density on the cell membrane32. Differences expression pattern of P-gp in tissues may influence therapeutic outcomes by affecting the accumulation of drugs in specific tissues. Transcription of Abcb1 had been measured using RT-PCR in the liver and brain capillary endothelial cells (pBCECs)33 and in the kidneys34. We measured Abcb1 mRNA level in the CNS, intestine, liver and kidneys. The data demonstrated that its expression in intestine was not uniform. In the peripheral tissues, the greatest expression was in the ileum. The porcine liver also expressed relatively high Abcb1 mRNA, which was consistent with previous data from human and canines35,36. Renal Abcb1 mRNA is lower in the pig, compared to that in humans35,36. P-gp in intestinal epithelial cells may prevent pharmacotherapeutic agents from entering the systemic circulation and therefore change their pharmacokinetics35. Hepatic metabolism is significant, and high expression level of P-gp is vital for biliary drug, toxicant, and metabolite excretion35. More Abcb1 mRNA in the intestine and liver suggests that porcine P-gp may limit absorption and facilitate secretion of substrates into the intestines. In addition, we measured P-gp protein expression level in 60-day old piglets and found that this was coincident with mRNA levels, suggesting a tissue-specific post-transcriptional mode of regulation. Hence, RNA analysis of pig tissue samples should be done in conjunction with protein analysis, because RNA and protein may differ among certain organs.

Of note, the P-gp expression pattern in tissues affects pharmacokinetics of enrofloxacin, a fluoroquinolone (FQ) widely used in veterinary medicine either orally or parenterally. Sørgel’s group30 and others41–43 reported that the concentrations of ciprofloxacin and danofloxacin-mesylate in the gut lumen are higher than those in plasma. Other studies proved that ciprofloxacin and danofloxacin-mesylate were substrates of multiple human ABC transporters44–46, indicating that intestinal efflux is the underlying mechanism for danofloxacin-mesylate and ciprofloxacin secretion into the lumen. In this study, the net efflux ratio of enrofloxacin across MDCK-pAbcb1 cells was more than 2 and basolateral-to-apical transport of enrofloxacin could be counteracted by the P-gp inhibitor verapamil. Although the concentration of enrofloxacin was not measured in the luminal compartment, its concentration in plasma was increased when animals received verapamil. These results suggest that enrofloxacin might be a substrate of porcine P-gp, similar to the data obtained in chickens47. Initially, we treated piglets with the P-gp inhibitor PSC-833, but some animals began vomiting. Thus, PSC-833 was replaced with verapamil, which could inhibit P-gp function and be monitored in an MDCK-pAbcb1 cell line.

Our data also suggested that co-administration of P-gp inhibitors may be an alternative strategy to improve oral bioavailability and therapeutic efficacy of enrofloxacin when used to treat systemic bacterial infection in the swine industry. Three generations of P-gp inhibitors have been identified but none of them show improved therapeutic efficacy due to their broad activity and toxicity48. Berberine can significantly inhibit P-gp (data not shown) so it may be a potential P-gp inhibitor as it is not absorbed after oral administration and only inhibits P-gp in the gastrointestinal tract.

In conclusion, the complete cDNA of porcine P-gp was cloned for the first time. The protein structure was modeled and analyzed. The phylogenetic analysis on P-gp proteins from different species was performed, and its protein expression and mRNA levels in different tissues were studied. Also, the effect of P-gp on enrofloxacin transmembrane transport and in vivo pharmacokinetics was evaluated. More studies are required to fully elucidate the complex epigenetic regulation of porcine P-gp during development, tissue-specific expression patterns, and the contribution of epigenetics to species variability in drug disposition and therapeutic response.

Materials and Methods

Cells. MDCK (Madin-Darby canine kidney) and IPEC-J2 cells were obtained from Shanghai Institute of Cell Biology, the Chinese Academy of Sciences (Shanghai, China) and grown in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum (FBS, HyClone Laboratories Inc.). Cells were maintained at 37 °C in a humidified atmosphere of 5% CO2 and sub-cultured once 80% confluent.

Animals. Sixty-day-old healthy crossbred pigs (large white x Landrace x Duroc, 20 ± 2 kg) were bought from Jiangsu Agricultural Academy and reared under standard conditions of light and temperature. Feed and water were provided ad libitum during the study. All pigs were fed according to the breeding standards of the Chinese Local Pigs and National Research Council (NRC). Animal use and handling protocols were approved by the regional Animal Ethics Committee and Nanjing Agricultural University. The protocol of the study was conducted in accordance with guidelines of the regional Animal Ethics Committee and Nanjing Agricultural University.

Total RNA extraction and cDNA synthesis. All pigs were anesthetized and killed by decapitation. Samples of liver, kidney, intestines and brain tissues were taken immediately (within 5–10 min after death). Brain capillaries were isolated from ~10 g of fresh cerebral cortices according to published methods with minor modification49. In brief, meninges and choroid plexus from the brains were removed and cerebral matter was homogenized in 20 volumes of cold Ringer’s solution containing 10 mM HEPES (Santa Cruz, Heidelberg, Germany) at pH 7.4 in a glass homogenizer. Ten upwards and downwards strokes were applied during the homogenization. The homogenates were collected and filtered initially through a 150 μm nylon mesh, the filtrate was then re-filtered.
through a 60 μm nylon mesh. The brain capillaries are trapped on the 60 μm nylon meshes and were collected in the tube. All samples were frozen in liquid nitrogen and then stored at −80 °C until RNA and protein extraction.

Total RNA was extracted from tissues using TRIzol (TaKaRa, Japan) following the manufacturer’s instructions. Quality and concentration were measured using a photometer (Eppendorf Biophotometer, Germany). RNA integrity was assessed with RNA electrophoresis. cDNA templates were synthesized from 1 μg of total RNA using a HiScript 1st strand cDNA synthesis kit (Vazyme, Nanjing, China).

Cloning and sequencing of full-length cDNA of porcine Abcb1. PCR primers (Table 1) toward three overlapping cDNA fragments of porcine Abcb1 (F1, F2, F3, shown in Fig. 1A) were designed based on sequence alignments of conserved regions from various species (Human, NM_000927.4; sheep, NM_001009790.1; dog, NM_001003215.1; mouse, NM_011075.2). Three overlapping fragments of porcine Abcb1 were amplified by touchdown PCR using PrimeSTAR GXL DNA Polymerase (TaKaRa, Japan) under these conditions: 95 °C for 5 min, 36 cycles of 95 °C for 30 s, 65–55 °C for 30 s (initial annealing temperature of 65 °C was reduced by 2 °C after every six cycles to 55 °C for the final six cycles), 72 °C for 1 min, and an extra extension of 15 min at 72 °C. PCR products were verified using published methods. After the three products were confirmed to be part of the Abcb1 gene by sequencing, full-length cDNA was obtained from the three fragments using fusion PCR as follows: 95 °C for 5 min, 30 cycles of 95 °C for 30 s, 62.5 °C for 30 s, 72 °C for 1 min, and 72 °C for 15 min. Final PCR products were confirmed by sequencing and the sequence was submitted to GenBank to obtain an accession number.

Bioinformatics analysis of porcine Abcb1 and its protein structural model. cDNA sequence analysis was conducted using NCBI BLAST (http://www.ncbi.nlm.nih.gov/blast). Calculated molecular weights and predicted isoelectric points were obtained with the ExPasy online server (http://www.expasy.org/tools/). Multiple alignments of P-gp protein sequences were generated using the Bio Edit program. A phylogenetic analysis was used to determine relatedness of porcine P-gp to other mammalian P-gp sequences, and alignment data were imported into MEGA version 4.1. The phylogenetic tree was constructed using the neighbor-joining method with Poisson correction. Bootstrap analysis was performed using 1,000 replicates.

The secondary structure of porcine and human P-gp was predicted and compared with Proter1.0 servers (http://wlab.ethz.ch/protter/start/). Secondary and predicted isoelectric points were obtained with the ExPasy online server (http://www.expasy.org/tools/). A phylogenetic analysis was conducted using NCBI BLAST. Calculated molecular weights and predicted isoelectric points were obtained with the ExPasy online server (http://www.expasy.org/tools/). Multiple alignments of P-gp protein sequences were generated using the BioEdit program. A phylogenetic analysis was used to determine relatedness of porcine P-gp to other mammalian P-gp sequences, and alignment data were imported into MEGA version 4.1. The phylogenetic tree was constructed using the neighbor-joining method with Poisson correction. Bootstrap analysis was performed using 1,000 replicates.

Establishment of an MDCK cell line stably-transfected with porcine Abcb1. Primers used to amplify pig Abcb1 complementary DNAs (cDNAs) were designed based on our previously submitted sequence (GenBank ID: KP233220) containing an Xhol site within the sense primer and an Xba I site within the antisense primer (See Table 1 for primers). PCR amplifications were performed as previously described in the Methods. Pig Abcb1 full cDNA was cloned into the plasmid pcDNA 3.1 and transformed into Escherichia coli DH5α cells (Vazyme, China). Plasmid was purified using an Omega Endo-Free Plasmid Mini Kit and was sequenced using vector primers to confirm inserted genes (Invitrogen, Shanghai, China).

To generate cell lines stably expressing porcine P-gp, an MDCK cell line was transfected with pcDNA3.1-pAbcb1 plasmid using LipofectAMINE 2000 reagent (Invitrogen) according to the manufacturer’s instructions. Stable transfectants were selected with G418 (600 μg/mL, Gibco). When single-cell stable colonies resistant to G418 were observed, cells were sub-cultured again and distinct colonies were isolated using cloning cylinders. Cells were grown in the presence of G418 to maintain expression of transfected genes. MDCK-pAbcb1 cells were propagated and sub-cultured in the same medium with the addition of G418 (300 μg/mL). MDCK, IPEC-J2 and three colonies of MDCK-pAbcb1 were used to measure exogenous expression of porcine P-gp. qRT-PCR and western blot were used according to published methods, with different primers (Table 1) and antibodies (Mdr-1, Santa Cruz, Heidelberg, Germany), dilution 1:200.

Rhodamine 123 accumulation assay. MDCK and MDCK-pAbcb1 cells were seeded in six-well plates, and cells were washed with PBS and rhodamine 123 (Rho123; 5 μM) (Sigma-Aldrich, Castle Hill, Australia) was added in the presence and absence of verapamil (100 μM) to identify P-gp mediated Rho123 accumulation. To study the time-course of Rho123 accumulation, cells were harvested at 15, 30, 60 and 120 min after the addition of Rho123 and accumulation was measured using a FACS Calibur (BD Biosciences, Bedford, MA) with CellQuest Pro software. Data were collected for a minimum of 10,000 gated events per sample as geometric mean fluorescent intensity for all samples.

Tissues distribution of P-gp (Abcb1) in pigs. Pigs (N = 6) were used to measure constitutive P-gp (Abcb1) expression in different tissues with real time quantitative PCR and western blot. Abcb1 mRNA was measured with qRT-PCR and gene-specific primers for PCR were designed according to the cloned sequence of porcine Abcb1 (KP233220 in GenBank). GAPDH was as internal control for each sample. Primer pairs for genes appear in Table 1. qRT-PCR was performed on a CFX96 Real-Time PCR Detection System (Bio-Rad, USA), and SYBR Green was the dye (Toyobo, Japan). mRNA was measured using 12.5 μL SYBR Green real-time PCR Master Mix in a 25 μL volume with 2 μL cDNA. Each run consisted of an initial 1 min activation cycle at 95 °C, followed by 40 cycles of denaturation for 20 s at 95 °C, annealing for 30 s at 60 °C, and elongation for 31 s at 72 °C. Uniform amplification of products was analyzed according to melting curves of the amplified products. Abcb1 mRNA relative expression in different tissues was measured using the 2−ΔΔCt method.

P-gp relative expression was measured with standard western blot with minor modifications. Tissue protein was extracted using a membrane and cytosol protein extraction kit (Beyotime, Haimen, China). Briefly, equal amounts of membrane proteins (5 μg/lane) were fractionated with SDS-PAGE and then transferred onto PVDF membranes (Bio-RAD, USA). Membranes were blocked with 5% bovine serum albumin and incubated with primary antibody at the appropriate dilution (Mdr-1, Santa Cruz, Germany, 1:200; β-actin, TransGen, China,
composition was 0.1 M phosphoric acid (adjusted pH to 3.5 with triethylamine)/acetonitrile (84:16, v/v) and assessed using published methods.

The mobile phase solution was used to concentrate the samples for 5 times. Twenty microliters of the mixture were injected into the HPLC column (Kromasil C18 columns, 5 mm particle size, 250 mm × 4.6 mm). The mobile phase composition was 0.1 M phosphoric acid (adjusted pH to 3.5 with triethylamine)/acetonitrile (84:16, v/v) and the flow rate of mobile phase was set to 0.85 mL/min. UV absorbance was measured at 278 nm. Assay validation including recovery rate, inter- and intra-assay precision, accuracy and assay linearity of enrofloxacin were assessed using published methods.

HPLC analysis of enrofloxacin. HPLC analysis of enrofloxacin was performed as published with minor modifications. Concentrations of enrofloxacin were measured using an Agilent 1200 HPLC system. In brief, plasma was thawed at room temperature and centrifuged at 2,000 × g for 5 min. The supernatant (0.5 mL) was mixed with acetonitrile and separated into organic and water phases by centrifugation. The organic phase was evaporated to dryness under a nitrogen stream and residue was resuspended with mobile phase solution. The samples from the transport assays were directly evaporated to dryness under a nitrogen stream and 40 μL of the mobile phase solution was used to concentrate the samples for 5 times. Twenty microliters of the mixture were injected into the HPLC column (Kromasil C18 columns, 5 mm particle size, 250 × 4.6 mm). The mobile phase composition was 0.1 M phosphoric acid (adjusted pH to 3.5 with triethylamine)/acetonitrile (84:16, v/v) and the flow rate of mobile phase was set to 0.85 mL/min. UV absorbance was measured at 278 nm. Assay validation including recovery rate, inter- and intra-assay precision, accuracy and assay linearity of enrofloxacin were assessed using published methods.

Papp Calculation and Pharmacokinetic analysis. Apparent permeability coefficients (Papp) were calculated using the following equation: Papp = (dQ/dt)/(A × C0), where A is the area of filter membrane, C0 is the initial concentration of the test drug, dQ is the amount of transported drug, and dt is time elapsed. The efflux ratio (ER) was calculated from (Papp B→A)/(Papp A→B). Where Papp B→A and Papp A→B are BL to AP and AP to BL apparent permeability coefficients, respectively. The net efflux ratio was calculated using the following equation: Net efflux ratio = efflux ratio in MDCK-pAbcb1/efflux ratio in MDCK.

Pharmacokinetics were calculated for individual data using sp3p97 practical pharmacokinetic software (Version 97, Chinese Pharmacologic Association, Beijing, China). The best fit compartment model was assessed according to Akaike’s information criterion.

Statistical analysis. All data were analyzed for statistical significance using SPSS Statistics 17.0 (version 17.0, SPSS Inc., Chicago, IL, USA). The differences between means were considered significant at p < 0.05 and very significant at p < 0.01. Data were shown as means ± SEM.

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Acknowledgements
The authors are grateful for the manuscript correction from Prof. Shile Huang from Louisiana State University Health Sciences Center in USA and Prof. Xiang Mao from Chinese Academy of Agricultural Sciences. The study was supported in part by Qinlan Project of Jiangsu Province, National Natural Science Foundation of China (No. 31572567), the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD) and its effect on pharmacokinetics of enrofloxacin.

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Conceived and designed the study: T.G. and L.W. Performed the experiments: T.G., J.H. and L.D. Contributed material/analysis tools: D.G., L.G., F.H., H.Z. and Z.A.B. Analyzed the data: T.G., L.D. and L.W. Statistical analyses: T.G. and L.D. Writing of manuscript: T.G. and L.W. Preparation of tables and figures: T.G. and L.D. All authors reviewed the manuscript.

Additional Information
Supplementary information accompanies this paper at http://www.nature.com/srep

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Guo, T. et al. Abcb1 in Pigs: Molecular cloning, tissues distribution, functional analysis, and its effect on pharmacokinetics of enrofloxacin. Sci. Rep. 6, 32244; doi: 10.1038/srep32244 (2016).

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