A Protein Phosphorylation Threshold for Functional Stacking of Plant Photosynthetic Membranes

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Abstract
Phosphorylation of photosystem II (PSII) proteins affects macroscopic structure of thylakoid photosynthetic membranes in chloroplasts of the model plant Arabidopsis. In this study, light-scattering spectroscopy revealed that stacking of thylakoids isolated from wild type Arabidopsis and the mutant lacking STN7 protein kinase was highly influenced by cation (Mg++) concentrations. The stacking of thylakoids from the stn8 and stn7stn8 mutants, deficient in STN8 kinase and consequently in light-dependent phosphorylation of PSII, was increased even in the absence of Mg++. Additional PSII protein phosphorylation in wild type plants exposed to high light enhanced Mg++-dependence of thylakoid stacking. Protein phosphorylation in the plant leaves was analyzed during day, night and prolonged darkness using three independent techniques: immunoblotting with anti-phosphothreonine antibodies; Diamond ProQ phosphoprotein staining; and quantitative mass spectrometry of peptides released from the thylakoid membranes by trypsin. All assays revealed dark/night-induced increase in phosphorylation of the 43 kDa chlorophyll-binding protein CP43, which compensated for decrease in phosphorylation of the other PSII proteins in wild type and stn7, but not in the stn8 and stn7stn8 mutants. Quantitative mass spectrometry determined that every PSII in wild type and stn7 contained on average 2.5 ± 0.1 or 1.4 ± 0.1 phosphoryl groups during day or night, correspondingly, while less than every second PSII had a phosphoryl group in stn8 and stn7stn8. It is postulated that functional cation-dependent stacking of plant thylakoid membranes requires at least one phosphoryl group per PSII, and increased phosphorylation of PSII in plants exposed to high light enhances stacking dynamics of the photosynthetic membranes.

Introduction
Plant chloroplasts contain extremely large and well organized photosynthetic thylakoid membranes with highly stacked membrane layers [1,2] enriched in photosystem II (PSII), which uses light energy to oxidize water and produce oxygen [3]. The flattened sacks of thylakoid membranes form multiple stacks, called grana, which allow fitting of the enormous membrane surface inside a chloroplast. The area-to-volume ratio of plant thylakoids is 70–500 times bigger than that for a sphere-like vesicle, resulting in the increased ability of the chloroplasts to capture light energy [4]. Thus, thylakoids of higher plants are organized in two distinct domains: 80% of the membrane comprises the grana stacks that are connected by non-appressed stroma lamellae membranes [5,6]. The grana stacking of thylakoids is important for several regulatory processes of plant photosynthesis, like balancing of the excitation energy between the thylakoids is important for several regulatory processes of plant stroma lamellae membranes [5,6]. The grana stacking of thylakoids comprises the grana stacks that are connected by non-appressed flattened sacks of thylakoid membranes form multiple stacks, called grana, which allow fitting of the enormous membrane surface inside a chloroplast. The area-to-volume ratio of plant thylakoids is 70–500 times bigger than that for a sphere-like vesicle, resulting in the increased ability of the chloroplasts to capture light energy [4]. Thus, thylakoids of higher plants are organized in two distinct domains: 80% of the membrane comprises the grana stacks that are connected by non-appressed stroma lamellae membranes [5,6]. The grana stacking of thylakoids is important for several regulatory processes of plant photosynthesis, like balancing of the excitation energy between the thylakoids.

We recently demonstrated that phosphorylation of PSII proteins in the model plant Arabidopsis thaliana affects macroscopic structure of thylakoids. The Arabidopsis stn8 and stn7stn8 mutants defective in phosphorylation of PSII have grana stacks that are markedly bigger than in thylakoids of wild type plants [7]. This enhanced grana size, visualized in the leaves of the mutant plants by electron microscopy, obstructs lateral migration of the PSII reaction centre protein D1 and of the processing protease FtsH between the stacked and unstacked membrane domains, and suppresses turnover of damaged D1 in the leaves exposed to high light [7]. Notably, the length of grana stacks in mature chloroplasts of all studied plant species is rather constant, about 400 nm [5], and larger grana diameter could be disadvantageous for lateral protein diffusion processes [8], like it has been demonstrated in Arabidopsis mutants deficient in light-induced phosphorylation of PSII [7]. Formation of thylakoid grana depends on the complex interplay of physicochemical forces of attraction and repulsion [4] The in vitro studies clearly demonstrated that electrostatic forces control the stacking and unstacking of isolated thylakoid membranes [9,10,11,12]. Protein phosphorylation occurs at the outer surface of thylakoid membranes [13,14] and contributes to the total negative charge of the membrane surface [4].

Thylakoid protein phosphorylation is mediated by a redox-sensitive regulatory system reflecting different light and other environmental conditions [14,15,16,17]. The recent years revealed two major protein kinases involved in these phosphorylation events in Arabidopsis. The thylakoid associated Ser-Thr kinase STN7 is essential for phosphorylation of light harvesting complex polypeptides, LHCCI, of the minor light harvesting protein CP29 and of TSP9, a soluble protein involved in regulation of light harvesting [17,18,19,20,21]. The phosphorylation of PSII core proteins is
mediated through an ortholog of STN7 called STN8 kinase [18]. STN8 is involved in phosphorylation of the D1, D2, CP43 and PsbH proteins of PSII, and the calcium-sensing receptor (CaS) protein [22,23]. Phosphorylation of LHCII and PSII core proteins serves important, while different adaptive functions. The first is crucial for the photosynthetic state transitions when reversible phosphorylation of mobile LHCII modulates its association with either PSII or photosystem I, allowing for optimal usage of excitation energy at low light intensities [17,21]. Differential phosphorylation of LHCCI-PSII linker protein CP29 [24,25] and the TPS9 protein, localized at the LHCCI-PSII interface, also regulate state transitions [20,26,27]. Phosphorylation of PSII core proteins is important for sustained photosynthesis in plants exposed to high light and for turnover of the light-damaged D1 core protein of PSII [7,16,28].

Phosphorylation of LHCCI and PSII proteins is regulated according to dark and light conditions. However, LHCCI is quickly phosphorylated under low light and dephosphorylated in darkness, while the PSII core has a more stable phosphorylation during the photoperiod [13]. Moreover, under high light conditions phosphorylation of PSII core proteins is increased while that of LHCCI is drastically reduced [13,15,21,29,30]. The relationship between the dynamic light-dependent phosphorylation of thylakoid proteins on the one hand and stability of thylakoid grana stacks in plant chloroplasts [3,8] on the other hand is not presently resolved. Particularly, there are no quantitative data on the PSII phosphorylation in plant leaves during day/night transitions in relation to thylakoid stacking. Moreover, the threshold of the PSII protein phosphorylation between the stn8 and stn7/stn8 protein kinase mutants with the abnormally grana size of the thylakoids and the wild type and stn7 Arabidopsis plants with the normal membrane organization [7] remains to be determined.

In this study we analyze the phosphorylation of PSII proteins in leaves of Arabidopsis thaliana wild type and stn7, stn8, and stn7/stn8 mutants at different photoperiodic time points and during prolonged darkness. For comprehensive analysis of the in vivo protein phosphorylation under various light/dark conditions we use three different complementary techniques: western blotting with two different anti-phosphothreonine antibodies; Diamond ProQ staining of phosphorylated proteins; and quantitative mass spectrometry of non-phosphorylated and phosphorylated peptides released from the thylakoid membranes. Furthermore, we also use light-scattering spectroscopy to monitor changes in thylakoid membrane structure depending on the protein phosphorylation state and varying concentrations of MgCl2, a salt important in maintaining grana structures in isolated thylakoid membranes [9,10]. Our data reveal the state of PSII protein phosphorylation during the photoperiod and demonstrate that phosphorylation of at least one protein per every PSII is required for the natural Mg++-dependent stacking of Arabidopsis thylakoid membranes.

**Results and Discussion**

**Effects of MgCl2 on stacking of isolated thylakoid membranes**

Thylakoid membranes carry net negative charges on their surfaces and electrostatic interactions control the stacking and unstacking of these membranes [4,12]. The extent of the stacking of isolated thylakoids could be directly monitored via light scattering of their suspensions by recording the optical density at 580 nm [9,10]. Furthermore, it had been shown by electron microscopy that chloroplasts isolated in media containing 5 mM MgCl2 retained stacked thylakoids, while chloroplasts isolated in low salt media lost their grana structure and had a reduced absorbance at 580 nm [9,10]. The stn7/stn8 and stn8 mutants of Arabidopsis are deficient in light-induced phosphorylation of PSII and have increased size of stacked thylakoid membranes compared to wild-type and stn7 plants, as has been revealed by transmission electron microscopy [7]. Here, we analyzed the changes in the light-scattering properties of isolated thylakoid membranes by recording the absorbance of their suspensions in the region 400–750 nm. Figure 1A, upper panel, shows the characteristic spectra from wild type, stn7, stn8 and stn7/stn8 membranes resuspended in a buffer containing 5mM MgCl2, to retain the native grana and stroma lamellae structures. It is clearly seen that the scattering properties of the thylakoids are different between wild type and stn7 on one hand and stn7/stn8 on the other hand. The difference is clear due to the higher absorbency of the stn8 and stn7/stn8 membranes in the yellow/orange range of 570–620 nm. Notably, Figure 1A, lower panel, shows the almost identical absorbance spectra of chlorophylls and other pigments after their extraction by acetone from the wild type and all the mutant membranes. This figure also displays the increase in the absorbance of the major chlorophyll peaks after their extraction from the native membranes (compare with Figure 1A, upper panel).

In order to confirm that increase in absorbance at 570–620 nm for the stn8 and stn7/stn8 mutants (Figure 1A) was a consequence of enhanced thylakoid membrane stacking [7,9,10] we performed titration of the membrane suspensions with 0, 5, 10, 20 and 30 mM MgCl2. Figure 1B shows the influence of MgCl2 on the spectral properties of thylakoids from wild type and stn8. The wild type and stn7 membranes followed the same pattern with high increase in absorbance in the yellow/orange spectral range upon increasing concentrations of MgCl2 (Figure 1C). The higher absorbance at increasing concentration of MgCl2 was a consequence of salt-mediated stacking of thylakoid membranes [9]. However, stn8 and stn7/stn8 showed a much higher absorbance of isolated thylakoids already at 0 mM MgCl2 and that absorbance was less affected by MgCl2 (Figure 1C). In order to quantify the increase in absorbance at 580 nm for stn8 and stn7/stn8 versus wild type and stn7 we normalized the signals at 580 nm to those at 678 nm and generated the graph in Figure 1C. This graph shows that wild type and stn7 have an overall lower absorbance value at 580 nm and are more influenced by MgCl2 titration than the stn8 and stn7/stn8 membranes.

As a next step we did analysis of Mg++-dependent folding of thylakoid membranes isolated from wild type plants exposed to three hours of high light treatment. Immunoblotting of the thylakoid proteins from the leaves exposed to darkness, normal light and high light demonstrated the high-light-induced rise in phosphorylation of D1 and D2 proteins of PSII (Figure 2A). The spectroscopic analysis of the isolated thylakoids showed that absorbance of the membranes from dark, normal and high light exposed plants was similar, when the membranes were resuspended in the absence of MgCl2. However, the higher phosphorylation of PSII core proteins from high light exposed plants resulted in the steeper absorbance rise when the membranes were titrated with MgCl2 (Figure 2B). At the higher levels of MgCl2 the dark, normal and high light acclimated membranes had similar absorbance at 580 nm, as a consequence of saturated Mg++-induced stacking. Thus, if the reduced phosphorylation of PSII in the stn8 and stn7/stn8 mutants diminishes Mg++-dependence of thylakoid folding (Figure 1), increased phosphorylation of PSII in the high-light-treated leaves of wild type makes folding of thylakoids more receptive to lower concentrations of Mg++ (Figure 2).

Phosphorylated PSII complexes are facing each other in the compressed thylakoid grana membranes [9,31], so the negatively charged phosphoryl groups repel each other. When Mg++ ions are present in the medium the phosphoryl groups exposed at the membrane surface are neutralized by these cations, which can also
bridge the adjacent membrane layers and thus stabilize the grana structures. In the stn8 and stn7stn8 mutants defective in phosphorylation of PSII the adjacent membranes have a lower negative charge and are more stacked [7], thus the Mg\(^{2+}\) ions induce only minor additional structural changes in the thylakoids. These data demonstrate that a certain stable level of PSII phosphorylation is important for maintenance of optimal cation-dependent stacking of plant thylakoid membranes.

**Photoperiodic changes in thylakoid protein phosphorylation**

To address the question of protein phosphorylation dynamics in thylakoid membranes of *Arabidopsis thaliana* we harvested leaves at different time points of the plant growth photoperiod (16 hours dark and 8 hours light). To preserve the *in vivo* protein phosphorylation we isolated thylakoid membranes in the presence of NaF, an inhibitor of thylakoid protein phosphatases [32]. The phosphorylation state of major thylakoid phosphoproteins was analyzed by immunoblotting with anti-phosphothreonine antibody (Figure S1). In agreement with the earlier studies [13] we observed significant increase in LHCII phosphorylation during the light period and almost complete dephosphorylation during night (Figure 3A). To the contrary, the phosphorylation state of the D1 and D2 proteins of PSII was rather stable during the whole photoperiod, with some increase in phosphorylation during the hours of light exposure (Figure 3A). Strikingly, CP43 phosphor-
ylation was regulated in the opposite way: it was higher during the night and approached its lowest levels during the middle of the light period (Figure 3A). Quantification of phosphorylation levels for CP43, D1, D2 and LHCII proteins (Figure 3A) revealed the most representative time points for night and day at 4 am and 1 pm (time point 13 in the Figure 3A), correspondingly. We also assayed phosphorylation states of the thylakoid proteins in the stn7 and stn8 mutants at these time points representative for night and day (Figure 3B). The light-dependent phosphorylation of LHCII was absent in stn7, while phosphorylation of the D1 and D2 proteins was significantly reduced in stn8, in agreement with the previous studies [18,22]. In respect to the night-dependent rise in phosphorylation of CP43, the stn7 mutant followed the same pattern as in the wild type, while stn8 showed a very low dark-induced CP43 phosphorylation as compared to the wild type and stn7 plants (Figure 3B). A distinct increase in CP43 phosphorylation observed after exposure of the wild type plants to darkness occurred in parallel to the dramatic decrease in phosphorylation of LHCII and some moderate reduction in phosphorylation of the D1 and D2 proteins. Thus, phosphorylation of CP43 during the light to dark transition in Arabidopsis is regulated opposite to that of the other PSII and LHCII proteins.

**Figure 2.** High light induced increase in PSII phosphorylation and its effect on MgCl2–dependence of thylakoid membrane light scattering. A, thylakoid membrane proteins from wild type plants adapted to dark, or exposed to normal or high light for 3 hours were separated on SDS-PAGE and immunoblotted with anti-phosphothreonine antibody from Zymed laboratories. The samples were loaded at three different chlorophyll concentrations, as indicated. Positions of the phosphorylated thylakoid proteins and of the molecular mass markers are indicated. B, titration of the light-scattering changes in thylakoid membranes from the leaves exposed to normal or high light, as indicated, using 0, 5, 10, 15, 20 and 30 mM MgCl2 and normalization of absorbance at 580 nm to the 678 nm signals. Error bars represent S.D. of at least three independent experiments. doi:10.1371/journal.pone.0010963.g002

**Figure 3.** Thylakoid protein phosphorylation during the photoperiod. A, relative changes in the PSII and LHCII protein phosphorylation determined by immunoblotting analyses of SDS-PAGE separated thylakoid proteins with anti-phosphothreonine antibody. Thylakoids were prepared from leaves of the wild type plants harvested at the indicated time points during night or day (4, 7, 10, 13, 16 are the hours of the dark phase and 10, 13, 16 are the hours of the light phase of the photoperiod). Error bars represent S.D. of at least three independent experiments. B, immunoblot of thylakoid proteins from the stn mutants and wild type, as indicated, with anti-phosphothreonine antibody from Zymed Laboratories. Positions of the phosphorylated thylakoid proteins and of the molecular mass markers are indicated. Time points correspond to the highest and lowest phosphorylation of CP43 in the wild type plant leaves: after 12 hours of darkness (time point 4) and after 5 hours of light (time point 13). doi:10.1371/journal.pone.0010963.g003

**Phosphorylation dynamics of PSII proteins in darkness**

To make a detailed examination of PSII phosphorylation state in darkness, with particular attention to the unexpected increase in CP43 phosphorylation level, we studied protein phosphorylation during prolonged dark incubation of plants using several complementary assays. Thylakoid membranes were isolated from leaves of Arabidopsis plants exposed to 4 hours of normal growth light, the time point that we used as a control, or from the plants which were kept in darkness for 4, 16, 20 and 24 hours. We analyzed phosphorylation levels of thylakoid proteins using two complementary anti-phosphothreonine antibodies from Zymed Laboratories and from New England Biolabs (Cell Signaling), which differ from each other in sensitivity against major thylakoid phosphoproteins [20,29,33]. Phosphorylation of LHCII and of two PSII core proteins, D1 and D2, decreased and stayed at different steady state levels during prolonged incubation in darkness (Figure S2 and Figure 4A,B). These changes in phosphorylation patterns were detected with both Zymed (Figure...
To investigate if the observed changes in protein phosphorylation patterns depend on photoperiod or only the light/dark transitions we exposed Arabidopsis plants to 3 hours of light (normal photoperiod), transferred plants to darkness for 4 hours, and then exposed them to light for 1 or 2 hours. The thylakoid membranes were isolated from the leaves harvested at these time points and analyzed by western blotting using anti-phosphothreonine antibody (Figure S2C). The quantitative analysis of phosphorylation changes for CP43, D1, D2 and LHCII proteins under these conditions is shown in Figure 4C. LHCII from the light-exposed plants had a high phosphorylation state, but was rapidly dephosphorylated upon the transition to darkness and phosphorylated again when the plants were re-exposed to light (Figure 4C).

The changes in phosphorylation of the PSII core proteins were more complex. Phosphorylation of D1 and D2 decreased, while phosphorylation of CP43 increased after the transition to darkness for 4 hours. However, none of these three PSII proteins changed its phosphorylation state significantly during the following exposure of the plants to light for 2 hours (Figure 4C).

To characterize the changes in endogenous phosphorylation of the PSII proteins by an independent analytical technique we also used Pro-Q Diamond phosphoprotein gel stain in combination with Sypro Ruby total protein gel staining. The combination of these two staining techniques provides a measure of the relative phosphorylation level of the protein in each band on the gel. The Pro-Q staining showed a clear difference in the directions of phosphorylation change for CP43 compared to D1, D2 and LHCII in the plants transferred to darkness (Figure 5A). The Sypro Ruby total protein stain demonstrated that the amount and expression of all the analyzed proteins was stable during the dark incubation (Figure 5B). The phospho-stain data and the total protein-staining allowed calculation of a relative phosphorylation ratio for the CP43, D1, D2 and LHCII proteins (Figure 5C). CP43 phosphorylation was up-regulated during the dark-transition, while phosphorylation of D1, D2 and, most obviously, LHCII was down regulated in the dark. These data confirmed the results obtained with anti-phosphothreonine antibodies and reinforced an important finding, namely that CP43 phosphorylation in darkness is regulated differently from that of the D1 and D2 core proteins of PSII or LHCII polypeptides.

### Protein phosphorylation in the kinase mutants during prolonged darkness

In order to investigate how the PSII phosphorylation dynamics is affected during prolonged incubation in darkness the absence of either STN7 or STN8 kinase, or both of them, we used the stn7, stn8 and stn7stn8 mutants. To this end we did analyses using both western blotting and ProQ phospho-stain. First, the thylakoid samples isolated from mutants and wild type plants exposed to growth light for 4 hours or darkness for 16 and 24 hours were immunoblotted using anti-phosphothreonine antibody. Wild type, stn7 and stn8 plants had a high phosphorylation state, but was rapidly dephosphorylated upon the transition to darkness and phosphorylated again when the plants were re-exposed to light (Figure 4C). LHCII from the light-exposed plants had a high phosphorylation state, but was rapidly dephosphorylated upon the transition to darkness and phosphorylated again when the plants were re-exposed to light (Figure 4C). LHCII was down regulated in the dark. These data confirmed the results obtained with anti-phosphothreonine antibodies and reinforced an important finding, namely that CP43 phosphorylation in darkness is regulated differently from that of the D1 and D2 core proteins of PSII or LHCII polypeptides.
requirement of both STN7 and STN8 for quantitative phosphorylation of the thylakoid proteins [7,17].

Our results reveal a complex regulatory process for CP43 phosphorylation, which includes both dark- and photoperiod-dependence. We determine that STN8 is at least partially responsible for the dark-induced phosphorylation of CP43, since its remaining phosphorylation in stn8 membranes is low during either light to dark transitions or photoperiod. Besides the redox regulation [29,34,35], thylakoid protein phosphorylation is also controlled at the substrate level by conformational changes of membrane proteins in response to light [36]. Furthermore, it has been shown that CP43 phosphorylation depends on the light-induced structural changes in this protein [37]. This work demonstrated that illumination of PSII cores affected the conformation of CP43 and expose of its N-terminus, containing the phosphorylation site, to the enzymes such as kinases and proteases [37]. We present evidence that phosphorylation of CP43 is decreased during the day. Accordingly, the light-induced conformational change in CP43 may be beneficial to the phosphatase accountable for its dephosphorylation. The enzymatic process responsible for dephosphorylation of CP43 has been characterized [32], while the corresponding protein phosphatase is still unknown and it is clearly distinct from the PPH1 phosphatase, which does not dephosphorylate the PSII core proteins [58].

Quantitative analysis of PSII protein phosphorylation

To quantify phosphorylation changes during day and night in the wild type, stn7, stn8 and stn7stn8 plants we used high performance liquid chromatography coupled with mass spectrometry. Thylakoid membranes isolated from wild type and mutant plants were subjected to proteolytic shaving by trypsin to remove the surface exposed phosphorylated peptides for vectorial proteomics [13,14,39]. This approach depends on complete proteolytic cleavage of the peptides from the membrane proteins, particularly when used quantitatively. We assayed proteolytic cleavage of the thylakoid proteins using western blotting. Figure 7A shows the results of analyses of thylakoid proteins with antibodies against CP43, D2, D1 C-terminus, D1 DE-loop and anti-phosphothreonine antibodies before and after trypsin treatment. These data demonstrate that the proteolytic cleavage of the membrane proteins was very efficient and no intact protein signal was detected after 3 hours of trypsin digestion.

HPLC in conjunction with Electrospray Ionization Mass Spectrometry (ESI-MS) allows resolution and detection of the major phosphorylated peptides and their non-phosphorylated counterparts for determination of the phosphorylation stoichiometry for PSII proteins [13]. In the present work we improved this earlier described method. To obtain accurate data on quantitative phosphorylation of PSII proteins we introduced a normalization procedure that takes into account the differences in ionization and in signal intensities for phosphorylated and corresponding non-phosphorylated peptides [40]. We determined the flyability ratio [40] for each particular peptide/phosphopeptide pair and used these ratios to correct the signal intensities of the phosphorylated peptides for absolute phosphorylation stoichiometry measurements. This approach is built on the fact that the decrease in the amount of phosphorylated peptide during enzymatic dephosphorylation is equal to increase in the amount of its non-phosphorylated cognate peptide. To generate samples with varying phosphorylation stoichiometry the thylakoid peptide mixtures were treated with alkaline phosphatase under controlled conditions: the aliquots were taken at 0, 10, 20 and 40 min of the phosphatase treatment. Figure 7B shows the ion intensity signals of phospho (678.42+), de-phospho (638.42+) and de-phospho (638.42+) forms of the CP43 peptide (Ac-TLFGNTLALAGR) at different time points and decrease in amount of phosphorylation upon the phosphatase treatment. The similar measurements were done for peptide/phosphopeptide pairs from the D1, D2 and PsbH proteins and their flyability ratios were calculated using 0, 10, 20, 40 min time points (Table 1). These ratios were used to correct the signal intensities of the phosphorylated peptides in measurements of the protein phosphorylation states by calculation of peptide/phosphopeptide ratios for each particular PSII protein (see below and Table 2).

We performed LC-MS analyses of the peptide mixtures from thylakoids isolated from leaves harvested during the day or night. The typical chromatogram is shown in Figure 7C with indication of elution times for the phosphorylated and non-phosphorylated peptides from the PSII core proteins. Phosphorylation stoichiometry of these proteins under day or night conditions was determined in the wild type, stn7, stn8 and stn7stn8 plants. The results are presented in Table 2. Phosphorylation state of the PSII core proteins in leaves of the wild type plants harvested during day or night was rather stable. In agreement with the earlier study [13] at least one third of CP43, D1, D2 or PsbH was phosphorylated during the dark or light phases of the photoperiod. Only appearance of the doubly phosphorylated form of PsbH [13,22] was strictly light-dependent (Table 2). Account of the differences in ionization and in signal intensities for phosphorylated and corresponding non-phosphorylated peptides and use of the

Figure 5. Phosphorylation of thylakoid proteins during prolonged darkness assayed with ProQ phosphoprotein stain. A, ProQ phospho-stain analysis of SDS-PAGE separated thylakoid proteins from wild type plants exposed to normal light for 4 hours (Light) and then transferred to darkness for 4, 16, 20 and 24 hours, as indicated. B, Sypro total protein stain on the same gel as in A. Positions of the phosphorylated thylakoid proteins and of the molecular mass markers are indicated. C, relative quantification of the protein phosphorylation of elution times for the phosphorylated and non-phosphorylated peptides from the PSII core proteins. Phosphorylation stoichiometry the thylakoid peptide mixtures were treated with alkaline phosphatase under controlled conditions: the aliquots were taken at 0, 10, 20, 40 min of the phosphatase treatment. Figure 7A shows the results of analyses of thylakoid proteins with antibodies against CP43, D2, D1 C-terminus, D1 DE-loop and anti-phosphothreonine antibodies before and after trypsin treatment. These data demonstrate that the proteolytic cleavage of the membrane proteins was very efficient and no intact protein signal was detected after 3 hours of trypsin digestion.
Flyability constants (Table 1) allowed for detection of increase in phosphorylation of the CP43 protein during night (Table 2). This quantitative MS analysis supported our immunoblotting and staining results: the extent of CP43 phosphorylation in the wild type increased from 34% during the day to 49% during the night, which corresponded to a relative 44% increase in the phosphorylation upon day to night transition.

Phosphorylation of the PSII core proteins during day or night in leaves of stn7 was very similar to that in wild type (Table 2), showing that the light-regulated kinase STN7 [17,18,35] is not directly involved in control of PSII phosphorylation during dark or light phases of the photoperiod. On the contrary, phosphorylation of the CP43, D1 and D2 core proteins in stn8 was 3 to 4 times lower than in wild type or stn7 plants and no increase in phosphorylation of the CP43 protein during night was found (Table 2). No significant phosphorylation of the PSII core proteins was detected in leaves of the stn7stn8 double mutant plants. Using the MS approach we found doubly phosphorylated PsbH corresponding to about 50% of total PsbH in light-exposed leaves of wild type and stn7, but it was totally absent in stn8 plants. On average at least one third of each of the PSII core protein was always phosphorylated during photoperiod in wild type and stn7, while in stn8 plants this phosphorylation was three times lower (Table 2).

Quantitative data from Table 2 and account of the equivalent molar amounts of the CP43, D1, D2 and PsbH proteins in the PSII core allowed calculations of average number of phosphoryl groups per PSII in leaves of the wild type, stn7 and stn8 plants during day or night. In the wild type Arabidopsis every PSII contained 1.38±0.11 or 2.44±0.13 phosphoryl groups during night or day, respectively. In the stn7 mutant these numbers were very similar: 1.35±0.13 or 2.51±0.08, correspondingly. However, in the stn8 mutant only 0.49±0.06 phosphoryl groups per PSII were present during night and 0.46±0.07 groups during the day. Phosphorylation of PSII during either day or night in the stn7stn8 plants was much lower (Table 2).

Phosphorylation of PSII and thylakoid stacking

It had been suggested that the physiological significance of PSII core protein phosphorylation is in regulation of the repair cycle of...
photodamaged PSII: phosphorylation facilitates migration of damaged PSII centers from grana to stroma thylakoid regions, were the damaged D1 protein is dephosphorylated, degraded and exchanged to a newly synthesized copy [28,41]. The thylakoid membranes contain big protein complexes and the high molecular ratio of proteins to lipids makes the thylakoids crowded. In the grana region, where PSII complexes are densely packed, movement of protein complexes may be considerably restricted [8]. This implies that the thylakoid membranes, especially in the grana stacks should be highly dynamic to allow for PSII migration [31]. Importantly, PSII-LHCII supercomplexes occupy about 50% of the grana space [31], so phosphorylation of PSII infers a massive negative change of the membrane surface and, consequently, repulsion of the adjacent grana stacks, as is schematically outlined in Figure 8. Quantitative mass spectrometry determined that every PSII in the wild type and stn7, stn8, stn7stn8 plants from normal light (4 hours of light) and dark (12 hours of darkness).

Figure 7. Quantification of PSII core protein phosphorylation using LC-MS. A, immunoblotting analyses of thylakoid membrane proteins from wild type plants with antibodies against CP43, D2, D1 and phosphothreonine before (−) and after (+) trypsin treatment, as indicated. B, LC-MS extracted ion chromatograms of the phosphorylated and non-phosphorylated N-terminal peptide from the CP43 protein after incubation of the peptide mixture with alkaline phosphatase for different time, as indicated. The ratios of phosphorylated to non-phosphorylated peptide intensities at each time point were used for calculations of CP43 phosphorylation level. C, LC-MS profile of total ion chromatogram with indication of the elution times for peptide/phosphopeptide pairs from PSII core proteins. Experimental procedures as in A, B and C were done for wild type and stn7, stn8, stn7stn8 plants from normal light (4 hours of light) and dark (12 hours of darkness).

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PSII are retarded in cation-dependent stacking (Figure 1) and the lateral migration of absence of the electrostatic repulsion. These thylakoids lost the second PSII has a phosphoryl group and the adjacent grana deficient in light-induced phosphorylation of PSII less than every simple explanation of all experimental data. In the mutant plants phosphoryl group per PSII. The model in Figure 8 provides a possible link between the high-light-induced increase in PSII phosphorylation and facilitation in migration of damaged PSII centers from grana to stroma thylakoid regions during the PSII repair cycle [28,41]. Notably, the recent study demonstrated more prominent photoinhibition of PSII in stacked than in unstacked thylakoids isolated from spinach, suggesting that unstacking of thylakoids has a crucial role in facilitating degradation of the photodamaged D1 under light stress [42].

Conclusion

Using quantitative measurements of PSII core protein phosphorylation we determined the threshold of the phosphorylation required for the naturally selected optimal stacking of thylakoid membranes in Arabidopsis. The lowest phosphorylation state of the PSII core proteins D1, D2, CP43 and PsbH on average does not decrease below 30% in the wild type plants. Phosphorylation of these proteins in stn7 is similar, while the absence of the LHCII phosphorylation in this mutant does not affect the thylakoid membrane structure in the leaves [7] or cation-dependent stacking of isolated thylakoids. Thus, LHCII phosphorylation does not influence macroscopic structure of thylakoids in Arabidopsis. Decrease in phosphorylation state of the PSII proteins to an average 12% in stn7 mutant plants increases size of thylakoid grana in the leaves [7] and diminishes cation-dependent folding of isolated thylakoids. The similar effects are caused by almost

### Table 1. Flyability ratios for PSII peptide ions and their phosphorylated cognates.

| Protein Peptide/phosphopeptide sequence | z | Flyability ratio |
|----------------------------------------|---|-----------------|
| CP43 Ac-TLFONGTLAAGR/Ac-TLFONGTLAAGR | 2+ | 1.23 ± 0.14 |
| PsbH ATQTVEDSSR/AIQTVEDSSR            | 2+ | 0.94 ± 0.17 |
| PsbH ATQTVEDSSR/AIQTVEDSSR            | 2+ | 0.89 ± 0.13 |
| PsbH ATQTVEDSSR/AIQTVEDSSR            | 2+ | 1.14 ± 0.21 |
| D1 Ac-TAILER/AtqTAILER                | 1+ | 1.17 ± 0.11 |
| D2 Ac-TIALGK/AtqTIALGK                | 1+ | 1.29 ± 0.10 |

The thylakoid peptide mixtures were treated with alkaline phosphatase for 0, 10, 20 and 40 min to obtain samples with varying degrees of phosphorylation. The signal intensity of each peptide ion was obtained using LC-MS and flyability ratio for each peptide/phosphopeptide pair was determined as (IpA/IpB), where IpA and IpB are the ion intensities of the phosphorylated and non-phosphorylated peptides at 0 min time point, and IpA and IpB are the corresponding ion intensities at the 10, 20 or 40 min time points. An average flyability ratio for each peptide/phosphopeptide pair was calculated from the data corresponding to all time points of alkaline phosphatase treatment. (IpB), were IpA and IA are the ion intensities of the phosphorylated and non-phosphorylated peptides at 0 min time point, and IpB and IB are the corresponding ion intensities at the 10, 20 or 40 min time points. An average flyability ratio for each peptide/phosphopeptide pair was determined as (IpA/IpB), were IpA and IA are the ion intensities of the phosphorylated and non-phosphorylated peptides at 0 min time point, and IpB and IB are the corresponding ion intensities at the 10, 20 or 40 min time points. An average flyability ratio for each peptide/phosphopeptide pair was calculated from the data corresponding to all time points of alkaline phosphatase treatment.

### Table 2. In vivo phosphorylation stoichiometry (%) for the PSII core proteins from the leaves of wild type, stn7, stn8 and stn7stn8 mutant plants harvested during day or night.

| Protein | % of phosphorylation | % of phosphorylation |
|---------|----------------------|----------------------|
| P-CP43 | 49±12 | 34±13 | 16±8 | 13±7 |
| P-D1   | 27±12 | 36±15 | 12±9 | 14±8 |
| P-D2   | 33±11 | 42±12 | 10±6 | 10±4 |
| P-PsbH | 29±9  | 24±12 | 11±8 | 9±8  |
| PP-PsbH| N.D.  | 54±14 | N.D. | N.D. |
| P-CP43 | 44±13 | 36±9  | N.D. | N.D. |
| P-D1   | 29±12 | 35±8  | N.D. | N.D. |
| P-D2   | 33±13 | 44±7  | <3*  | <4*  |
| P-PsbH | 29±15 | 22±8  | <4*  | <4*  |
| PP-PsbH| N.D.  | 57±9  | N.D. | N.D. |

N.D. - not determined.

*The PsbH (AIQTVEDSSR) and D2 (Ac-tIALGK) phosphopeptide signals were detected in the samples from stn7stn8 only in one or two out of three different experiments. The phosphorylation state of each protein was determined from signal intensities of peptide and phosphopeptide ion pairs corrected using corresponding flyability ratios from the Table 1. Data are the average from three experiments in each condition and three LC-MS runs for every experiment.

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complete absence of PSII core protein phosphorylation in the stn7stn8 mutant of *Arabidopsis*.

The data indicate that the PSII core protein phosphorylation in plant thylakoids affects both macroscopic structure and dynamic properties of the photosynthetic membranes. We present evidence that CP43 phosphorylation is up-regulated during night, which partially compensates for the decrease in phosphorylation of other PSII core proteins during the dark phase of the photoperiod and could be important for the stable structure of plant thylakoids. The phosphorylation of CP43 in darkness is regulated differently from that of the D1 and D2 core proteins of PSII or LHCII polypeptides. PSII phosphorylation controls membrane stacking and lateral movements of the membrane components, which consequently may regulate processes such as PSII repair, plastoquinone diffusion and membrane biogenesis. These dynamic properties of thylakoid membranes allow plants to fine-tune photosynthesis, regulate the mechanisms of photoprotection and in general the adaptation to different environments. Elucidating the mechanisms involved in regulation of thylakoid structural flexibility we came closer to understanding of the functional properties of extremely large and organizationally complex photosynthetic membranes in the chloroplasts of higher plants.

**Materials and Methods**

**Plant Material**

*Arabidopsis thaliana* wild type (ecotype Columbia) plants, *stn7* (SALK 073254) [18], *stn8* (SALK 060869) [22], and double mutant *stn7stn8* [7,18] in Columbia background used in this study were grown hydroponically [43] at 23°C, 65–70% relative humidity, photosynthetic flux of 120 μmol photons m⁻² s⁻¹ and photoperiod of 8 h light and 16 h dark. In the case of high light experiments photosynthetic flux of 900 μmol photons m⁻² s⁻¹ has been used.

**Isolation and characterization of thylakoids**

Four-week-old plants were used for the preparation of chloroplasts and thylakoids. The thylakoid membranes were isolated from 4g of *Arabidopsis* leaves harvested at time points indicated. The leaves were homogenized in 20 ml of ice-cold 25 mM Tricine (pH 7.8), 330 mM sorbitol, 1mM EDTA, 10mM KCl, 0.15% bovine serum albumin, 4 mM sodium ascorbate, 7mM L-cysteine in a metal blender for four periods of 1 sec at high speed. The homogenate was immediately filtered through four layers of nylon mesh (20 μm pore size), and the filtrate was centrifuged for 3 min at 1000 g. The pellet was resuspended in the same buffer to wash the thylakoids and centrifuged for 5 min at 1000 g. The chloroplast pellet was resuspended in 10 mM Tricine, 5 mM MgCl₂, and 10 mM NaF and allowed to stand for 5 min in the dark on ice in order to lyse the chloroplasts. Following lysis, the thylakoids were pelleted by centrifugation for 5 min at 6000 g. To wash the thylakoids the pellet was resuspended in 100 mM sorbitol, 25 mM Tricine (pH 7.8), 5 mM MgCl₂, 10 mM KCl, and 10 mM NaF and centrifuged for 5 min at 6000 g. The pellet was resuspended in a small volume of the same buffer. NaF was used as a phosphatase inhibitor in the buffers when phosphorylation was quantified. All procedures were made under weak green light at 4°C, and the sample was kept on ice throughout the whole process.

**Immunoblotting**

Thylakoid membrane proteins were separated by SDS-PAGE (6% acrylamid stacking gel+14% separation gel+6M urea) and the proteins were transferred to a PVDF membrane (Immobilon, Millipore). For the anti-phosphothreonine antibodies, purchased from Zymed Laboratories Inc. or New England Biolabs (Cell Signalling), the membranes were blocked with 5% bovine serum albumin. For specific antibodies against the DE-loop in D1 protein [44], D1 C-terminus, residues 230–245 in the D2 protein and against CP43 protein [45], the blocking was done with 10% skimmed milk. The membranes were then incubated with horseradish peroxidase-conjugated secondary antibody and analyzed using ECL detection kit (GE Healthcare) and chemiluminescence imaging (LAS-1000) procedure. Immunoresponse linearity was determined by a series of sample dilutions in the range of 0.05–2.0 μg chlorophyll. Quantification of the immunoblots was done using Fujifilm LAS-1000 software.

**ProQ Diamond and Sypro staining**

Proteins from thylakoids containing equal amounts of chlorophyll were separated by SDS-PAGE and stained with ProQ® Diamond phosphoprotein gel stain and SYPRO® Ruby protein-staining kit (Synchromics). The thylakoids were isolated from 4g of leaves harvested at time points indicated. The leaves were homogenized in 20 ml of ice-cold 25 mM Tricine (pH 7.8), 330 mM sorbitol, 1mM EDTA, 10mM KCl, 0.15% bovine serum albumin, 4 mM sodium ascorbate, 7mM L-cysteine in a metal blender for four periods of 1 sec at high speed. The homogenate was immediately filtered through four layers of nylon mesh (20 μm pore size), and the filtrate was centrifuged for 3 min at 1000 g. The pellet was resuspended in the same buffer to wash the thylakoids and centrifuged for 5 min at 1000 g. The chloroplast pellet was resuspended in 10 mM Tricine, 5 mM MgCl₂, and 10 mM NaF and allowed to stand for 5 min in the dark on ice in order to lyse the chloroplasts. Following lysis, the thylakoids were pelleted by centrifugation for 5 min at 6000 g. To wash the thylakoids the pellet was resuspended in 100 mM sorbitol, 25 mM Tricine (pH 7.8), 5 mM MgCl₂, 10 mM KCl, and 10 mM NaF and centrifuged for 5 min at 6000 g. The pellet was resuspended in a small volume of the same buffer. NaF was used as a phosphatase inhibitor in the buffers when phosphorylation was quantified. All procedures were made under weak green light at 4°C, and the sample was kept on ice throughout the whole process.

**Thylakoid Phosphorylation**

A model for phosphorylation- and cation-dependent stacking of plant photosynthetic membranes. Deficiency in light-induced phosphorylation of PSII in the stn8 and stn7stn8 mutant plants cause enlargement of grana and loss of the cation-dependent stacking of thylakoids (left). Presence of at least one phosphoryl group per PSII in the wild type thylakoids causes electrostatic repulsion of the adjacent grana membranes (center) and makes their stacking highly cation-dependent (right).

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gel stain according to manufacturer’s instructions (Molecular Probes, Eugene, OR, USA). The gels were fixed in 50% methanol with 10% acetic acid for 30 min, the solution was changed and the fixation continued overnight, followed by washing 3×10 min in MilliQ water. Immediately prior to use the ProQ Diamond stain was equilibrated to room temperature and vigorously mixed. Gels were stained for 90 min in darkness, followed by destaining three times for 30 min with 4% acetonitrile in 50 mM sodium acetate pH 4.0, and finally washed twice in MilliQ water prior to scanning on a FLA-5100 imaging system at 532 nm. The gels were then immediately put in Syprio Ruby total protein stain and incubated over night followed by two washes in 10% methanol, 7% acetic acid and finally rinsed twice with MilliQ water. Total protein was detected on FLA-5100 imaging system at 473 nm.

Characterization of protein phosphorylation by mass spectrometry

The thylakoids isolated from the wild type or vtnb, vtn7, vtn7stn7 mutant plants were resuspended in 25 mM NH4HCO3, 10 mM NaF to a final concentration of 2.5 mg of chlorophyll/ml and incubated for 3 h at 22°C with a sequencing grade-modified trypsin from Promega (Madison, WI, USA) at 5 µg of enzyme/mg of chlorophyll [22]. The peptides cleaved by trypsin were separated from the thylakoid membranes by centrifugation at 100 000 g for 30 min. The peptides were chromatographically separated from the thylakoid membranes by centrifugation at 100 000 g for 30 min. The peptides were performed using collision induced dissociation of peptide ions.

Controlled protein dephosphorylation

Controlled dephosphorylation of the peptides derived from thylakoids by trypsin treatment, as described above, was performed using alkaline phosphatase at a concentration of 100 millunits/µl (Phosphatase alkaline, Sigma) in a buffer containing 25 mM NH4HCO3 (pH 8.0). The aliquots were taken at 0, 10, 20 and 40 min time points and dephosphorylation reactions were stopped by 1% formic acid (pH 4.0).

Spectroscopic measurements

The spectroscopic measurements were performed using a PerkinElmer Lambda 25 spectrophotometer. The data were sampled at 2-nm intervals in the 400–750 nm range. Thylakoid membranes were resuspended at 0.0125µg chlorophyll/µl in a buffer containing 100 mM sorbitol, 25 mM Tricine (pH 7.8), 5 mM MgCl2, 10 mM KCl, and 10 mM NaF; which was used to preserve the native conformation of the membranes. For titration studies with different concentrations of MgCl2 the membranes were resuspended in the same buffer without MgCl2 or with MgCl2 at the corresponding concentration and incubated on ice for 5 min with occasional careful mixing of the solution. For pigment analysis the same membranes were dissolved in 90% acetone at 0.0125µg chlorophyll/µl.

Supporting Information

Figure S1 Thylakoid protein phosphorylation during the photoperiod. Immunoblot of SDS-PAGE separated thylakoid proteins with anti-phosphothreonine antibody from Zymed Laboratories. Positions of the phosphorylated thylakoid proteins and of the molecular mass markers are indicated. Thylakoids were prepared from leaves of the wild type plants harvested at the indicated time points during night or day (4, 7, 19 are the hours of the dark phase and 10, 13, 16 are the hours of the light phase of the photoperiod).

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Figure S2 Phosphorylation of thylakoid proteins during prolonged darkness or during dark/light transitions not related to photoperiod. Thylakoid membrane proteins from wild type plants were separated on SDS-PAGE and immunoblotted with anti-phosphothreonine antibody. A and B, thylakoids were isolated from leaves exposed for 4 hours to normal light of 120 µmol photons m-2 s-1 (Light) or for 16, 20 and 24 hours of darkness, as indicated. A, immunoblot with anti-phosphothreonine antibody from Zymed Laboratories. B, immunoblot with anti-phospho-threonine antibody from Cell Signaling. Lower panels in A and B demonstrate immunoblots of the same membranes with antibodies against the CP43 and D1 proteins, correspondingly. C, thylakoids were isolated from the wild type plants exposed to normal light for 3 hours (Light), then incubated for 4 hours in darkness and re-exposed to normal light for 1 or 2 hours, as indicated. Thylakoid membrane proteins were separated on SDS-PAGE and immunoblotted with anti-phosphothreonine antibody from Zymed Laboratories.

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Author Contributions

Conceived and designed the experiments: AVV. Performed the experiments: RF PG. Analyzed the data: RF PG. Wrote the paper: RF AVV.

References

1. Mustardly L, Garab G (2005) Granum revisited. A three-dimensional model - where things fall into place. Trends Plant Sci 10: 117–122.
2. Mullineaux CW (2005) Function and evolution of grana. Trends Plant Sci 10: 521–525.
3. Barber J (2006) Photosystem II: an enzyme of global significance. Biochem Soc Trans 34: 619–631.
4. Chow WS, Kim EH, Horton P, Anderson JM (2005) Granal stacking of thylakoid membranes in higher plant chloroplasts: the physicochemical forces at work and the functional consequences that ensue. Photochem Photobiol 81: 1081–1090.
5. Albertsson PA, Andreassen E (2004) The constant proportion of grana and stroma lamellae in plant chloroplasts. Physiol Plant 113: 334–342.
6. Dekker JP, Boekema EJ (2005) Supramolecular organization of thylakoid membrane proteins in green plants. Biochim Biophys Acta 1706: 12–39.
7. Fristedt R, Willig A, Granath P, Grevecoeur M, Rochaix JD, et al. (2009) Phosphorylation of Photosystem II Controls Functional Macroscopic Folding of Photosynthetic Membranes in Arabidopsis. Plant Cell 121: 3950–3964.
8. Kirchhoff H (2008) Molecular crowding and order in photosynthetic membranes. Trends Plant Sci 13: 201–207.
9. Itozawa S, Good NE (1966) Effect of Salts and Electron Transport on the Conformation of Isolated Chloroplasts. I. Light-Scattering and Volume Changes. Plant Physiol 41: 535–543.
10. Itozawa S, Good NE (1966) Effect of Salts and Electron Transport on the Conformation of Isolated Chloroplasts. II. Electron Microscopy. Plant Physiol 41: 544–552.
11. Barber J (1982) Influence of surface charges on thylakoid structure and function. Annu Rev Plant Physiol 33: 261–295.
12. Chow WS, Thorne SW, Duniec JT, Sculley MJ, Boardman NK (1980) The stacking of chloroplast thylakoids. Effects of cation screening and binding, studied by the digitonin method. Arch Biochem Biophys 201: 347–355.

13. Vener AV, Harms A, Sussman MR, Vierstra RD (2001) Mass spectrometric resolution of reversible protein phosphorylation in photosynthetic membranes of Arabidopsis thaliana. J Biol Chem 276: 6959–6966.

14. Vener AV (2007) Environmentally modulated phosphorylation and dynamics of proteins in photosynthetic membranes. Biochim Biophys Acta 1767: 449–457.

15. Vener AV, Ohad I, Andersson B (1998) Protein phosphorylation and redox sensing in chloroplast thylakoids. Cur Opin Plant Biol 1: 217–223.

16. Aro E-M, Ohad I (2003) Redox regulation of thylakoid protein phosphorylation. Antioxid Redox Signal 5: 53–67.

17. Ročaix JD (2007) Role of thylakoid protein kinases in photosynthetic acclimation. FEBS Lett 581: 2768–2773.

18. Bellaflore S, Barneche F, Pehier G, Ročaix JD (2005) State transitions and light adaptation require chloroplast thylakoid protein kinase STN7. Nature 433: 492–495.

19. Deppe N, Bellaflore S, Ročaix JD (2003) Role of chloroplast protein kinase Stn7 in LHCCI phosphorylation and state transition in Chlamydomonas. Science 299: 1572–1575.

20. Friseth R, Carlberg I, Zogadio A, Pippol M, Nurmi M, et al. (2009) Intrinsically unstructured phosphoprotein TSP9 regulates light harvesting in Arabidopsis thaliana. Biochemistry 48: 499–509.

21. Tikkanen M, Pippol M, Suorsa M, Sirpiä S, Mulo P, et al. (2006) State transitions revisited - a buffering system for dynamic low light acclimation of Arabidopsis. Plant Mol Biol 62: 779–793.

22. Vainonen JP, Hannon M, Vener AV (2005) STN8 protein kinase in Arabidopsis thaliana is specific in phosphorylation of photosystem II core proteins. J Biol Chem 280: 35679–35686.

23. Vainonen JP, Sakuragi Y, Stael S, Tikkanen M, Allahverdijaev Y, et al. (2008) Light regulation of CaS, a novel phosphoprotein in the thylakoid membrane of Arabidopsis thaliana. FEBS J 275: 1767–1777.

24. Kargul J, Turrina MV, Nield J, Benson S, Vener AV, et al. (2005) Light-harvesting complex II protein CP29 binds to photosystem I of Chlamydomonas reinhardtii under State 2 conditions. FEBS J 272: 4797–4806.

25. Turquina MV, Kargul J, Tosi M, Niel J, Komolinos S, et al. (2006) Environmentally modulated phosphoproteome of photosynthetic membranes in the green alga Chlamydomonas reinhardtii. Mol Cell Proteomics 5: 1412–1425.

26. Hannon M, Dupuis T, Stromquist R, Andersson B, Vener AV, et al. (2007) The mobile thylakoid phosphoprotein TSP9 interacts with the light-harvesting complex II and the peripheries of both photosystems. J Biol Chem 282: 16214–16222.

27. Kargul J, Barber J (2008) Photosynthetic acclimation: structural reorganisation of light harvesting antenna role of redox-dependent phosphorylation of major and minor chlorophyll a/b binding proteins. FEBS J 275: 1056–1068.

28. Tikkanen M, Nurmi M, Kangasjarvi S, Ročaix EM (2008) Core protein phosphorylation facilitates the repair of photodamaged photosystem II at high light. Biochim Biophys Acta 1777: 1432–1437.

29. Rintamäki E, Salonen M, Suoranta UM, Carlberg I, Andersson B, et al. (1997) Phosphorylation of light-harvesting complex II and photosystem II core proteins shows different irradiance-dependent regulation in vivo. Application of phosphothreonine antibodies to analysis of thylakoid phosphoproteins. J Biol Chem 272: 30476–30482.

30. Barna-Gonzalez E, Barbato R, Aro E-M (1999) Role of phosphorylation in repair cycle and oligomeric structure of photosystem two. Planta 208: 196–204.

31. Kihchhoff H, Lehnert S, Buchel C, Chl I, Nield J (2000) Probing the organization of photosystem II in photosynthetic membranes by atomic force microscopy. Biochemistry 47: 431–440.

32. Vener AV, Ročka A, Fulgoji H, Andersson B, Herrmann RG (1999) A cyclophilin-regulated PP2A-like protein phosphatase in thylakoid membranes of plant chloroplasts. Biochemistry 38: 14953–14963.

33. Aro EM, Ročka A, Vener AV (2004) Determination of phosphoproteins in higher plant thylakoids. Methods Mol Biol 274: 271–286.

34. Vener AV, Van Kan PM, Rich PR, Ohad I, Andersson B (1997) Phosphoprotein at the quinol oxidation site of reduced cytochrome b mediates signal transduction between light and protein phosphorylation: Thylakoid protein kinase deactivation by a single-turnover flash. Proc Natl Acad Sci U S A 94: 1383–1390.

35. Lemelle S, Willig A, Deppe-Fargeix N, Delessert C, Bassi R, et al. (2009) Analysis of the chloroplast protein kinase Sn7 during state transitions. PLoS Biol 7: e45.

36. Zer H, Vink M, Keven N, Dilsh-Harwis HG, Paulsen H, et al. (1999) Regulation of thylakoid protein phosphorylation at the substrate level: reversible light-induced conformational changes expose the phosphorylation site of the light-harvesting complex II. Proc Natl Acad Sci U S A 96: 8277–8282.

37. Vink M, Zer H, Herrmann RG, Andersson B, Ohad I (2000) Regulation of Photosystem II core protein phosphorylation at the substrate level. Light induces exposure of the CP43 chlorophyll a protein complex to thylakoid protein kinase(s). Photosynthesis Research 64: 209–219.

38. Shapiguzov A, Ingelsson B, Samuel I, Andre C, Kesler F, et al. (2010) The PPH1 phosphatase is specifically involved in LHCCI dephosphorylation and state transitions in Arabidopsis. Proc Natl Acad Sci U S A 107: 4762–4767.

39. Vener AV, Strallos P (2005) Vectorial proteomics. IUBMB Life 57: 433–440.

40. Steen H, Jebanathirajah JA, Springer M, Kirschner MW (2005) Stable isotope-free and absolute quantitation of protein phosphorylation stoichiometry by MS. Proc Natl Acad Sci U S A 102: 3948–3953.

41. Aro E-M, Virgin I, Andersson B (1993) Photoinhibition of Photosystem II. Inactivation, protein damage and turnover. Biochim Biophys Acta 1143: 113–134.

42. Khatoon M, Inagawa K, Pospisil P, Yamashita A, Yoshioka M, et al. (2009) Quality control of photosystem II: Thylakoid unstacking is necessary to avoid further damage to the D1 protein and to facilitate D1 degradation under light stress in spinach thylakoids. J Biol Chem 284: 25343–25352.

43. Norén H, Svensson P, Andersson B (2004) A convenient and versatile hydroponic cultivation system for Arabidopsis thaliana. Physiologia Plantarum 121: 343–348.

44. Spetea C, Hundal T, Lohmann F, Andersson B (1999) GTP bound to plastidial thioplyl synthase: photosynthetic electron transport by a single-turnover flash. Proc Natl Acad Sci U S A 96: 6547–6552.

45. Koivuniemi A, Aro E-M, Andersson B (1995) Degradation of the D1- and D2-proteins of photosystem II in higher plants is regulated by reversible phosphorylation. Biochemistry 54: 16022–16029.