Essential Role of the Disulfide-bonded Loop of Chromogranin B for Sorting to Secretory Granules Is Revealed by Expression of a Deletion Mutant in the Absence of Endogenous Granin Synthesis

Andreas Krömer, Michael M. Glombik, Wieland B. Huttner, and Hans-Hermann Gerdes
Department of Neurobiology, University of Heidelberg, 69120 Heidelberg, Germany

Abstract. Sorting of regulated secretory proteins in the TGN to immature secretory granules (ISG) is thought to involve at least two steps: their selective aggregation and their interaction with membrane components destined to ISG. Here, we have investigated the sorting of chromogranin B (CgB), a member of the granin family present in the secretory granules of many endocrine cells and neurons. Specifically, we have studied the role of a candidate structural motif implicated in the sorting of CgB, the highly conserved NH2-terminal disulfide-bonded loop. Sorting to ISG of full-length human CgB and a deletion mutant of human CgB (Δcys-hCgB) lacking the 22–amino acid residues comprising the disulfide-bonded loop was compared in the rat neuroendocrine cell line PC12. Upon transfection, i.e., with ongoing synthesis of endogenous granins, the sorting of the deletion mutant was only slightly impaired compared to full-length CgB. To investigate whether this sorting was due to coaggregation of the deletion mutant with endogenous granins, we expressed human CgB using recombinant vaccinia viruses, under conditions in which the synthesis of endogenous granins in the infected PC12 cells was shut off. In these conditions, Δcys-hCgB, in contrast to full-length hCgB, was no longer sorted to ISG, but exited from the TGN in constitutive secretory vesicles. Coexpression of full-length hCgB together with Δcys-hCgB by double infection, using the respective recombinant vaccinia viruses, rescued the sorting of the deletion mutant to ISG. In conclusion, our data show that (a) the disulfide-bonded loop is essential for sorting of CgB to ISG and (b) the lack of this structural motif can be compensated by coexpression of loop-bearing CgB. Furthermore, comparison of the two expression systems, transfection and vaccinia virus–mediated expression, reveals that analyses under conditions in which host cell secretory protein synthesis is blocked greatly facilitate the identification of sequence motifs required for sorting of regulated secretory proteins to secretory granules.

SPECIALIZED eukaryotic cells, such as endocrine cells and peptidergic neurons, contain a regulated secretory pathway in addition to the constitutive pathway present in all cells (Burgess and Kelly, 1987). Proteins specific for either of the two pathways are segregated from each other at the level of the TGN. Morphological and biochemical data indicate that a key feature in this sorting is the formation of aggregates of regulated secretory proteins in the TGN from which constitutively secreted proteins are excluded (Tooze et al., 1993). A paradigm for the aggregation-mediated sorting are chromogranin A (CgA),1 chromogranin B (CgB, secretogranin I), and secretogranin II (SgII), the three classical members of the granin family with widespread occurrence in secretory granules of neuroendocrine cells (Huttner et al., 1991; Rosa and Gerdes, 1994). It was shown that their selective aggregation is triggered by an acidic pH and millimolar concentration of calcium in vitro (Gerdes et al., 1989; Gorr et al., 1989; Yoo and Albanesi, 1990) and in the TGN (Chanat and Huttner, 1991). This low pH/calcium-induced aggregation is thought

1. Abbreviations used in this paper: AT, α1-antitrypsin containing a tyrosine sulfation site; CgA, chromogranin A; CgB, chromogranin B; CV, constitutive secretory vesicles; EGFP, enhanced green fluorescent protein; gpt, guanine phosphoribosyltransferase; hCgB, human CgB; hsPG, heparansulfate proteoglycan; ISG, immature secretory granules; pfu, plaque-forming unit; PI, post infection; POMC, proopiomelanocorticotrophic hormone; rCgB, rat CgB; rSgII, rat SgII; SgII, secretogranin II; S_eff, storage efficiency; vv, vaccinia virus.
to be mediated by structural features common to all granins, in particular, an abundance of acidic residues distributed along the polypeptide chain (Gerdes et al., 1989). Aggregates of regulated secretory proteins exit the TGN in immature secretory granules (ISG), a short lived intermediate in the biogenesis of mature secretory granules (Toozé and Huttner, 1990; Toozé et al., 1991). During maturation of ISG, which includes further condensation of the soluble content and removal of excess membrane, budding of ISG-derived vesicles has been proposed (Toozé et al., 1991). Such vesicles are thought to mediate the constitutive-like secretion of proteinaceous material that remained soluble in the ISG (Arvan and Castle, 1992; Kuliawat and Arvan, 1992).

During exit from the TGN into ISG, regulated secretory proteins are thought to interact with specific components in the membrane of the TGN. Recently it has been proposed that carboxypeptidase E is a membrane-associated binding site, recognizing regulated secretory proteins in the TGN (Cool et al., 1997). The structural features of regulated secretory proteins involved in membrane recognition, often referred to as sorting signals, are poorly understood. To date, no such sorting signals have been unequivocally identified. One dilemma could lie in the fact that it is not single molecules, but rather aggregates, which are sorted to ISG. Therefore, mutated regulated secretory proteins lacking the sorting signal when analyzed by transfection might not be missorted if the mutant protein can coaggregate with endogenous regulated secretory proteins carrying a sorting signal. Coaggregative sorting could explain why mutated regulated secretory proteins were not seen to be affected in their sorting to secretory granules (Burgess et al., 1987; Powell et al., 1988; Chu et al., 1990; Roy et al., 1991; Castle et al., 1992; Chevrier et al., 1993). In the few cases in which effects of mutations have been detected, the analysis was not performed at the level of the TGN, but rather at the level of mature secretory granules (Sevarino et al., 1989; Stoller and Shields, 1989; Cool and Peng Loh, 1994; Cool et al., 1995), and thus it remained open whether sorting to, or storage in, secretory granules was affected. To overcome these two limitations of previous studies we pursued the following strategy. First, we surmised that the identification of sorting signals by a mutagenesis approach would necessitate expression of the mutants under conditions in which there is no synthesis of endogenous regulated secretory proteins, thus precluding coaggregative sorting. Second, to analyze sorting at the level of the TGN, we monitored the exit of proteins from the TGN into ISG using the constitutive-like secretion of carboxypeptidase E (Chanat et al., 1994; Chanat et al., 1993), without affecting its aggregative properties (Chanat et al., 1994), suggesting a role of the disulfide bond in the sorting of CgB to the regulated pathway of secretion. Although DTT-induced missorting was specific for CgB and not observed for SgII, which lacks cysteins, it cannot be entirely excluded that the effect of DTT was not due to its reduction of the disulfide bond of CgB, but reflected an effect on a component necessary for the sorting of CgB and not for SgII. In this report, we analyze the sorting of hCgB and of a deletion mutant of hCgB lacking the highly conserved disulfide-bonded loop at the level of the TGN as well as at the level of the mature secretory granule. Recombinant proteins were expressed in the neuroendocrine cell line PC12, either by transfection, i.e., in the presence of endogenous granin synthesis, or by recombinant vaccinia viruses, i.e., in the absence of endogenous granin synthesis.

**Materials and Methods**

**Materials**

Hydroxyurea was purchased from Boehringer Ingelheim Bioproducts (Heidelberg, Germany), sodium butyrate from Merck (Darmstadt, Germany), radiochemicals from Amersham Intl. (Buckinghamshire, UK), and cell culture reagents from GIBCO BRL (Gaithersburg, MD).

**Cells and Viruses**

RK13 (rabbit kidney cells ATCC CCL 37) and HuL43 Tk- (ATCC CRL 8303) were cultured in Eagle’s minimal essential medium supplemented with 10% fetal calf serum at 37°C in 5% CO₂. PC12 cells (rat pheochromocytoma cells, clone 251; Heumann et al., 1983) were grown in DMEM supplemented with 10% horse serum and 5% fetal calf serum as previously described (Toozé and Huttner, 1990). Vaccinia virus, wild type (strain vv-WR), and a temperature-sensitive mutant, Ts7 (strain Copen-hagen), were kindly provided by H. Stunnenberg (University of Nijmegen, The Netherlands).

**Antibodies**

The hybridoma supernatant monoclonal antibody 67-C7-2 was used for hCgB detection (Rosa et al., 1992). A polyclonal antibody was used for detection of rat CgB (rCgB; Rosa et al., 1992). For rat SgII (rSgII) detection, a rabbit antiserum against the NH₂-terminal peptide (-QRNQLLKQEPDL-RLENV-) of rSgII was raised. Anti-myc ascites was a gift from E. Karsenti (European Molecular Biology Laboratory [EMBL], Heidelberg, Germany). A polyclonal anti-human α₁-antitrypsin antibody was kindly provided by J.L. Brown (Denver, CO) and bought as IgG fraction from Sigma Chemical Co. (St. Louis, MO).

**Expression Vector Cloning**

wt-hCgB. The cDNA of hCgB was released from vector pDS6/SgI (Benedum et al., 1987) by EcoRI/BamHI digestion, blunt-ended, and subcloned into the Smal site of pGEM4 (Promega, Heidelberg, Germany). hCgB was subcloned into p160/10 (gift from U. Ruehler, Medizinische Hochschule, Hannover, Germany) using HindIII and EcoRI as restriction sites. It was exized from p160/10 with HindIII and Notf and subcloned into pCDMS (Invitrogen, Carlsbad, CA).

Δψp-hCgB. The deletion mutant of hCgB was constructed using an oligonucleotide-directed mutagenesis system (Amersham Intl., version 2.1). In short, the hCgB cDNA was subcloned from pGEM4 into M13mp19 using HindIII and EcoRI as restriction sites. The single-stranded antisense oligonucleotide-directed mutagenesis system (Amersham Intl., version 2.1). In short, the hCgB cDNA was subcloned from pGEM4 into M13mp19 using HindIII and EcoRI as restriction sites. The single-stranded antisense oligonucleotide was used as a template for mutagenesis. The mutagenic primer, TTCAGGACCTTGGCGCGAGGATCACATTC, was used to obtain the desired deletion. After mutagenesis, double-stranded phase DNA was isolated and screened by digestion with HaeI. Three out of five analyzed phase plaque DNAs showed the expected restriction fragment pattern. The DNA sequence of the deletion mutation from one phase plaque was confirmed by sequencing. DNA from this phase was digested with Sall and Xhol and the restriction fragment containing the deleted region was used to replace the corresponding wild-type restriction fragment in pCDMS/wt-hCgB.
AT. By HindIII/XbaI digestion of pEC/EAlPITS (Leitinger et al., 1994) the α,-antitrypsin cDNA containing the tyrosine sulfation site (AT) of rat cholecystokinin precursor was released and subcloned into pRC/CMV.

For the construction of wt-hCGB and Δεs-hCGB containing a my epitope, a Clal and a KpnI site (underlined) were introduced into the COOH terminus of hCGB by PCR using a T7-primer and as a reverse primer oligonucleotide 5′CCGATGCATCATGGTGACCCCTTTTGGCCTAGATTTCGEM4/hCGB served as a template. The amplified fragment was subcloned into pSP73 (Promega) using XhoI and Clal sites. The resulting vector was digested with Clal and KpnI. Next, two complementary oligonucleotides, 5′CCATGGAACAAAAACTCATCTCAGAAGGAGATTGGAAGAGATATAT and 5′CGATATCTCCTCCAGA-TCTCTTCTGAGATGAGTTTTTGTTCCATGGGTAC encoding for the COOH terminus of hCGB was obtained from the pEMBL-I3-gpt plasmid (kindly provided by K. Krömer et al., 1988). Therefore, a gpt gene was introduced into the PstI site of the plasmid transfer vector with a 7.5K vaccinia promoter.

Recombinant virus was prepared using Hu143 TK− cells infected at 33°C for 2 h with Ts7 virus and transfected with a mixture of wt vaccinia virus DNA and recombinant plasmid essentially as described (Hänggi et al., 1986). The transfected cells were incubated for 2 d at the restrictive temperature (39.5°C) for Ts7 virus. Recombinant viruses were then further selected and plaque-purified on Hu143 TK− cells in the presence of 100 µg/ml 5-bromodeoxyuridine. Plaque-pure recombinants were tested for expression of hCGB on RK13 cells by immunofluorescence analysis (Stratagene, La Jolla, CA). The XhoI–EcoRV restriction fragment was subcloned into pBluescript KS (Stratagene, La Jolla, CA). The XhoI–EcoRI restriction fragment of the fragment was subcloned into pSP73 (Promega) using XhoI and ClaI sites.

Viral Amplifications and Titrations

Preparation of Recombinant Viruses

vwt-hCGB. A plasmid transfer vector with a 7.5K vaccinia promoter flanked by viral thymidine kinase sequences (Chocran et al., 1985), modified by Bujard and co-workers (von Brunn et al., 1988) was linearized with BamHI and Clal and blunted-end. The cDNA of hCGB (2.5 kb) was released from vector pDS6/SgI (Benedum et al., 1987) by EcoRI/BamHI digestion, blunt-ended, and subcloned into the plasmid transfer vector under control of the 7.5K promoter.

Recombinant virus was prepared using Hu143 TK− cells infected at 33°C for 2 h with Ts7 virus and transfected with a mixture of wt vaccinia virus DNA and recombinant plasmid essentially as described (Hänggi et al., 1986). The transfected cells were incubated for 2 d at the restrictive temperature (39.5°C) for Ts7 virus. Recombinant viruses were then further selected and plaque-purified on Hu143 TK− cells in the presence of 100 µg/ml 5-bromodeoxyuridine. Plaque-pure recombinants were tested for expression of hCGB on RK13 cells by immunofluorescence analysis with anti-hCGB antibody and metabolic labeling (see below).

vΔes-hCGB, vAT, vwt-hCGB-EGFP. A dominant selection system for recombinant viruses based on the xanthine-guanine phosphoribosyl-transferase (gpt) gene of Esherichia coli was used (Falkner and Moss, 1988). Therefore, a gpt gene was introduced into the PsI site of the plasmid transfer vector (constructed by von Brunn et al., 1988). The transfer vector was linearized with PstI and blunt-ended. A DNA fragment encoding the gpt gene was obtained from the pEMBL-I3-gpt plasmid (kindly provided by H. Stunnenberg) after Clal/EcoRI digestion. The fragment was blunt-ended and ligated into the prepared transfer vector, such that the 7.5-K promoter and the gpt promoter were oriented in opposite directions. In addition, a SalI site in the NH2-terminal region of the gpt gene was removed by partial digestion, treatment of the recessive ends by Klenow polymerase, and religation. The resulting plasmid transfer vector, termed pAB, was used to generate three recombinant viruses. The vector was linearized with XhoI and SalI and blunt-ended. Δεs-hCGB was released from pCDM8/Δεs-hCGB by SalI/EcoRI digestion, α,-antitrypsin containing the tyrosine sulfation site of rat cholecystokinin precursor was released from pECE/AIPITS (Leitinger et al., 1994) by HindIII/XbaI digestion and hCGB-EGFP (Kaether et al., 1997) was released from pSP73 by restriction with HindIII/EcoRI. Each fragment was blunt-ended and subcloned into pAB. Generation of recombinant virus and gpt’ selection was carried out as described (Falkner and Moss, 1988) except that recombinant viruses were generated in HuTK− cells and gpt’ selection was carried out with RK13 cells.

Viral Amplifications and Titrations

Vaccinia virus-infected RK13 cells were scraped into the medium 2.5 d post infection (PI), pelleted for 1 h at 31,000 g, resuspended in 10 mM Tris, pH 9.0, and lysed by Dounce homogenization. After low-speed centrifugation to remove nuclei, the virus was sedimented through a 36% sucrose cushion (10 mM Tris, pH 9.0) in an SW28 rotor (Beckman Instruments, Inc., Fullerton, CA) at 15,000 rpm for 1 h. The virus pellet was resuspended in PBS, briefly sonicated, and stored in aliquots at −20°C. Titre estimations of viral stocks were performed on RK13 cells by infecting confluent dishes with dilution series in complete culture medium.

Infections

PC12 cells were plated onto laminin-coated dishes (for metabolic labeling) or on poly(1-L)-lysine-coated coverslips (for immunofluorescence) 5 d before infection. Single infections were done at 100 plaque-forming units per cell (pfu/cell) in complete growth medium containing 5 mM hydroxyurea. For double infections, vwt-hCGB-EGFP was used at 400 pfu/cell to adjust the expression level to that of Δεs-hCGB, which was used at 100 pfu/cell. Hydroxyurea was present in the inoculum and in all subsequent culture media. After 1 h at 37°C and 10% CO2 with intermittent agitation, the inoculum was aspirated, the cells were washed twice with complete growth medium, and then propagated in complete growth medium. Standard infection times before analysis were 3 h. Deviations from these standard conditions are indicated in the figure legends.

Transfections

Standard protocol: PC12 cells were trypsinized, washed with PBS, and re-suspended in PBS at a density of 1.5–2.5 × 10^5 cells/ml. Electroporation was done with 50 µg DNA in a total volume of 0.8 ml cell suspension using a BioRad Gene Pulser at 960 µF and 250 V (Hercules, CA). Transfected cells were diluted in 15 ml growth medium and plated onto cell culture dishes. Growth medium was changed the next day and, except for the stimulated release experiments, replaced by growth medium containing 10 mM sodium butyrate 17.5 h before pulse labeling.

Metabolic Labeling

To label PC12 cells (3.5-cm dishes) with [35S]methionine, cells were depleted of endogenous methionine by incubation for 30 min at 37°C in methionine-free medium (DME lacking methionine) with 1% dialyzed horse serum and 0.5% dialyzed fetal calf serum, and, thereafter, pulse labeled for 5 min with [35S]methionine. At the end of the pulse, dishes were transferred to 4°C, the medium was aspirated, and the cells were washed twice with 1 ml ice-cold PBS supplemented with 5 mM methionine and 5 mM cysteine. Thereafter, cells were scraped off in 0.5 ml of isotonic buffer (20 mM Tris/HCl, pH 7.4, 0.3 M sucrose, 10 µg/ml leupeptin, 10 µg/ml aprotonin, 0.25 mM PMSF), and divided into two aliquots. One aliquot was analyzed directly on SDS-PAGE (referred to as total homogenate) to estimate the amount of isolated cells by scanning Coomassie-stained cellular protein bands. From the other aliquot, a heat stable fraction highly enriched for the granins was prepared, as described previously (Benedum et al., 1987) followed by quantitation of labeled rSgII from the fluorogram.

To label the granins present in the TGN, PC12 cells were preincubated for 30 min with sulfate-free DME, supplemented with 10% of the normal concentration of methionine and cysteine. Thereafter, they were pulse labeled for 5 min with sulfate-free DME containing 3–4 µCi/ml carrier-free [35S]sulfate. To chase the sulfate label, the normal growth medium was supplemented with twice the normal concentration of sulfate. The total amount of [35S]sulfate-labeled proteins always increased slightly over time due to a lag time of chase with [35S]sulfate, as observed previously (Baeuerle and Huttner, 1987).

Depolarization-induced Release

3 or 3 h after transfection, PC12 cells were labeled with [35S]sulfate for 10 or 5 min, respectively. After 90 min of chase, cells were washed twice with either 5K/Ca medium (127 mM NaCl, 5 mM KCl, 2.2 mM CaCl2, 0.33 mM Na2HPO4, 0.44 mM KH2PO4, 2.2 mM NaHCO3, 5.6 mM glucose, and 10 mM Hepes-NaOH; pH 7.4; for 5 mM KCl control and 55 mM KCl release) or 5K/Mg medium (5K/Ca medium in which 2.2 mM CaCl2 was replaced by 10 mM MgCl2; for 55 mM KCl plus 10 mM MgCl2 control). Thereafter, cells were incubated for 10 min in 5K/Ca medium (for 5 mM KCl control), 55K/Mg medium (5K/Mg medium in which 127 mM NaCl and 5 mM KCl were replaced by 77 mM NaCl and 55 mM KCl, for 55 mM KCl plus 10 mM MgCl2 control) or 55K/Ca medium (5K/Mg medium in which 127 mM NaCl and 5 mM KCl were replaced by 77 mM NaCl and 55 mM KCl for 55 mM KCl release). At the end of the incubation, dishes were placed on ice, medium was removed, and cells and media were analyzed, either after immunoprecipitation or directly by SDS-PAGE and phosphoimaging.
Subcellular Fractionation

The exit of the granins from the TGN into constitutive secretory vesicles (CV) and immature secretory granules was followed by sequential velocity and equilbrium sucrose gradients according to standard procedures (Tooze and Huttner, 1990; Tooze and Huttner, 1992), except for the following changes: velocity and equilibrium gradients were fractionated into 1 ml aliquots, fractions 2-5 from the top of the velocity gradients were pooled and analyzed by equilibrium gradient centrifugation on linear 0.6-1.6 M sucrose gradients.

Immunoprecipitation

Myc-tagged proteins. After the indicated chase times, the medium was collected and cells were washed twice with ice-cold PBS. Cells were lysed for 10 min in 0.8 ml (6-cm dishes), or 1 ml (10-cm dishes) myc lysis buffer (10 mM Tris-HCl, pH 7.5, 100 mM NaCl, 1 mM EDTA, 1% Triton X-100, 10 μg/ml leupeptin, 0.25 mM PMSF). The lystate was cleared by sequential centrifugation for 10 min at 800 g and 30 min at 100,000 g diluted with 1 vol cell dilution buffer (10 mM Tris-HCl, pH 8.0, 100 mM NaCl, 1 mM EDTA, 10 μg/ml leupeptin, 0.25 mM PMSF). Chaste or release media were cleared as done for cell lysates and diluted with 1 vol medium dilution buffer (10 mM Tris-HCl, pH 8.0, 50 mM NaCl, 2 mM EDTA, 1% Triton X-100, 10 μg/ml leupeptin, 0.25 mM PMSF). Samples were incubated for 4 h after which 40 μl of a 50% (wt/vol) suspension of protein A-Sepharose CL-4B (Pharmacia Diagnostics AB, Uppsala, Sweden), swollen and washed three times in PBS and a 1:1 mixture of cell lysis and cell dilution buffer, was added. After a 2-h incubation, the immunocomplexes were washed four times with myc wash buffer (10 mM Tris-HCl, pH 8.0, 100 mM NaCl, 1 mM EDTA, 0.2% Triton X-100). The final pellet was resuspended in Laemmli sample buffer (Laemmli, 1970), boiled, and subjected to SDS-PAGE. In some experiments, an aliquot of the supernatants before dilution with cell or medium dilution buffer was mixed with Laemmli sample buffer, and directly subjected to SDS-PAGE.

AT was immunoprecipitated like myc-tagged proteins with the following changes: lysis buffer: 10 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.5% deoxycholic acid, 10 μg/ml leupeptin, 0.25 mM PMSF; cell dilution buffer: 50 mM Tris-HCl, pH 8.0, 150 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.5% deoxycholic acid, 0.5% SDS, 1 mg/ml BSA, 0.5% low-fat milk powder, 10 μg/ml leupeptin, 0.25 mM PMSF; Medium dilution buffer: cell dilution buffer with 2% Triton X-100 and 1% deoxycholic acid. The antibody incubation was done overnight. Immunocomplexes were washed twice with wash buffer A (10 mM Tris-HCl, pH 8.0, 150 mM NaCl, 1 mM EDTA, 0.2% Triton X-100) and B (10 mM Tris-HCl, pH 8.0, 500 mM NaCl, 1 mM EDTA, 0.2% Triton X-100) and once with C (10 mM Tris-HCl, pH 8.0, 0.2% Triton X-100). For immunoprecipitation, gradient fractions were diluted with 4 vol gradient dilution buffer M (12.5 mM Tris-HCl, pH 8.0, 125 mM NaCl, 1.25 mM EDTA, 0.63% Triton X-100) before addition of antibody.

Electrophoresis and Immunoblotting

SDS-PAGE was performed and gels were processed as described (Lee and Huttner, 1983). For the analysis of gradient fractions, equal aliquots were acetone precipitated before analysis. Western blots were probed with a monoclonal anti-hCGB (67-CT-2) followed by a goat anti-mouse antibody (Dianova, Hamburg, Germany) and 125I-protein A.

Immunofluorescence

Single and double immunofluorescence was performed following standard procedures (Rosa et al., 1989). The coverslips were mounted on glass slides with 90% glycerol containing 1 mg/ml p-phenylendiamine. Fluorescence was viewed using a conventional Zeiss Axioskop microscope (objective 63× and 40×; Oberkochen, Germany) or a confocal microscope (Leica TCS 4D, Heerbrugg, Switzerland) with ~800 nm depth for the analyzed sections.

Quantitations

Unless indicated, quantitation was done using a FUJI X-BAS 1000 phosphoimager (Tokyo, Japan). Fluorograms were quantitated by densitometric scanning using the Pharmacia-LKB ultrascan XL.

For determination of the infection rate, cells were fixed after different time periods of infection followed by immunofluorescence with anti-hCGB. For each time point, the percentage of immunopositive cells was calculated by analyzing ~2,000 cells (Zeiss microscope, objective 40×).

Results

Transfected Wild Type and Δcys-hCGB Are Stored in Secretory Granules

To test whether the disulfide-bonded loop of CGB is an essential structure for its sorting to secretory granules, a deletion mutant of hCGB (Δcys-hCGB) was constructed (Fig. 1). This mutant lacks amino acids 16–37 comprising the disulfide-bonded loop of hCGB. Transfection experiments were carried out with wild-type hCGB (wt-hCGB) and Δcys-hCGB containing a myc tag at their COOH termini. Trafficking of the recombinant proteins was analyzed by biochemical and morphological methods. We (a) determined secretion kinetics and intracellular storage of recombinant proteins by pulse-chase analysis, (b) characterized the type of vesicle to which recombinant proteins were sorted upon exit from the TGN by sucrose gradient centrifugation, (c) tested the storage of recombinant proteins in secretory granules by depolarization-induced release, and (d) analyzed their subcellular distribution by immunofluorescence analysis.

First, we analyzed whether wt-hCGB and Δcys-hCGB were sorted to the regulated pathway of secretion during ongoing synthesis of endogenous regulated secretory proteins. Both proteins were transiently expressed in the neuroendocrine cell line PC12. Expression of transfected proteins was monitored by [35S]sulfate-labeling, a post-translational modification of the TGN (Baueuerle and Huttner, 1987), followed by immunoprecipitation with an antibody against the myc tag. At the end of the 5-min pulse, wt-hCGB and Δcys-hCGB were detected as single bands in cell lysates indicating the full-length products (Fig. 2 A, bracket). In addition, after a 90-min chase, two fragments of hCGB of ~68 kD were observed in cell lysates and media (Fig. 2 A, diamond), which could also be immunoprecipitated with an antibody against hCGB (data not shown). As a marker for constitutive secretion, recombinant human α1-antitrypsin containing a tyrosine sulfation site at the COOH terminus (AT; Leitinger et al., 1994) was expressed in PC12 cells. AT was chosen because its isoelectric point is similar to that of hCGB (Owen and Carrell, 1992; Gerdes and Huttner, 1995), but, in contrast to the latter protein, does not undergo calcium- and low pH-induced aggregation in vitro (Krömer, A., and H.-H. Gerdes, unpublished observation). By immunoprecipitation of [35S]sulfate-labeled AT from cell lysates and media a major band of ~62 kD (Fig. 2 A, asterisk) was detected.
At all time points, the total amount of $^{35}$S sulfate-labeled less efficiently in PC12 cells, suggesting that granins of species other than rat are stored to 40.8% reduction in storage efficiency of constitutive marker AT to 9.3%. In the presence of rSgII in transfected cells (Seff: 93.4%) this value was lower than that determined for endogenous rSgII in transfected cells (Seff: 40.8%).

Rapid secretion of all three proteins occurred during the first 60 min and approached a plateau after 90 min of chase (see Fig. 2). Note the slightly slower mobility of cys-hCgB as compared with wt-hCgB. (C) Secretion of wt-hCgB, cys-hCgB, and AT after different chase times. (C) Storage efficiencies were determined after 90 min of chase (four independent experiments). Error bars show standard deviation. In B and C, the labeled protein in the medium (B) and in the cell (C) is expressed as percent of total (sum of cell plus medium).

A minor band of lower molecular weight observed in cell lysates at the end of the 5-min pulse (Fig. 2 A, arrowhead) probably reflected sulfation of incompletely sialylated AT, because this form disappeared already after short chase times (data not shown). Secretion kinetics of transfected wt-hCgB, Δcys-hCgB, and of AT were determined after 5-min sulfate pulse-labeled following by chase times for up to 3 h (Fig. 2 B). At all time points, the total amount of $^{35}$S sulfate-labeled proteins had increased slightly (see Materials and Methods) indicating that degradation of $^{35}$S sulfate-labeled proteins did not take place during the chase periods. Rapid secretion of all three proteins occurred during the first 60 min and approached a plateau after 90 min of chase. The relative amounts that were released into the medium differed for each protein. We determined the storage efficiency (Seff) for each protein by quantitation of the relative amount stored in the cell after 90 min of chase (Fig. 2 C). Wt-hCgB was stored with a Seff of 70.9% ± 6.0 SD. This value was lower than that determined for endogenous rSgII in transfected cells (Seff: 93.4% ± 2.7 SD), suggesting that granins of species other than rat are stored less efficiently in PC12 cells. Δcys-hCgB lacking the disulfide-bonded loop was stored to 40.8% ± 6.8 SD and the constitutive marker AT to 9.3% ± 1.2 SD. Although the reduction in storage efficiency of Δcys-hCgB as compared to wt-hCgB points to a role of the disulfide-bonded loop in sorting of hCgB, this structure is apparently not essential for its sorting after transfection.

The subcellular compartment to which wt-hCgB and Δcys-hCgB were sorted was characterized by three different approaches. First, we determined the type of vesicle into which wt-hCgB and Δcys-hCgB were sorted upon exit from the TGN. PC12 cells were transfected, pulse labeled for 5 min with $^{35}$S sulfate, and chased for 12 min, after which the transfected protein had completely left the TGN (data not shown). Thereafter, postnuclear supernatants were prepared and subjected to an established subcellular fractionation protocol, which consists of a sequential velocity and equilibrium sucrose gradient centrifugation (Tooze and Huttner, 1990). The distribution across the equilibrium gradients of the heparansulfate proteoglycan (hsPG), a marker for CV, SgII, a marker for ISG and transfected proteins is shown in Fig. 3. Both wt-hCgB (Fig. 3, left) and Δcys-hCgB (Fig. 3, right) were found in fractions also containing rSgII indicating sorting to ISG. A minor amount of Δcys-hCgB (~30%) comigrated with hsPG (Fig. 3, right), suggesting that it exited from the TGN in CV. The observation that nearly 100% of transfected wt-hCgB and ~70% of Δcys-hCgB exited into ISG within 12 min but remained in the cell to only ~70% and ~40% after 90 min of chase (see Fig. 2 C), respectively, suggests basal release of mature secretory granules or constitutive-like secretion from ISG of the transfected proteins (see Discussion). In contrast to the gradient profiles obtained after 12 min of chase, complete codistribution of wt-hCgB and Δcys-hCgB with rSgII was observed for equilibrium gradients prepared after 90 min of chase at a density characteristic of mature secretory granules (data not shown).

Second, we tested whether secretion of wt-hCgB and Δcys-hCgB could be induced by depolarization of transfected PC12 cells. After transfection with either wt-hCgB or Δcys-hCgB, PC12 cells were pulse labeled for 10 min with $^{35}$S sulfate and chased for 90 min. Depolarization of

Figure 2. Storage efficiencies of wt-hCgB, Δcys-hCgB, and AT transiently transfected in PC12 cells. PC12 cells transfected with wt-hCgB, Δcys-hCgB, or AT were pulse labeled for 5 min with $^{35}$S sulfate and chased for the indicated times. Equal aliquots of cells and media collected at the end of the chase were analyzed by immunoprecipitation and SDS-PAGE followed by phosphoimaging. (A) Autoradiograms of cell lysates (lanes C) and media (lanes M) at the end of the pulse (0 min of chase) and after 90 min of chase. (Bracket: hCgB; diamond: hCgB fragments; asterisk: AT). Note the slightly slower mobility of Δcys-hCgB as compared to wt-hCgB. (B) Secretion of wt-hCgB, Δcys-hCgB, and AT after different chase times. (C) Storage efficiencies were determined after 90 min of chase (four independent experiments). Error bars show standard deviation. In B and C, the labeled protein in the medium (B) and in the cell (C) is expressed as percent of total (sum of cell plus medium).

Figure 3. Transfected wt-hCgB and Δcys-hCgB are sorted to immature secretory granules upon exit from the TGN. PC12 cells transfected with wt-hCgB (left) or Δcys-hCgB (right) were pulse labeled with $^{35}$S sulfate for 5 min and chased for 12 min. Postnuclear supernatants were prepared and subjected to sequential velocity and equilibrium sucrose gradient centrifugation. Equal aliquots of equilibrium gradient fractions were analyzed, either after immunoprecipitation (hCgB) or directly (hsPG, rSgII) by SDS-PAGE and phosphoimaging. The graphs show the quantitation of $^{35}$S sulfate-labeled hCgB, hsPG, and rSgII across the equilibrium sucrose gradient (fraction 1, top). The arrows indicate the peak of constitutive secretory vesicles (CV) and immature secretory granules (ISG).
and chased for 90 min. Cells were then incubated for 10 min, as indicated. Equal aliquots of cells and media were analyzed after immunoprecipitation by SDS-PAGE and phosphoimaging. The graph shows the quantitation of [35S]sulfate-labeled hCgB secreted during the 10-min incubation. Each value is expressed as percentage of total (sum of cells plus medium). Error bars show standard deviation of four independent experiments.

Third, the subcellular distribution of transfected proteins was analyzed by confocal double immunofluorescence. After incubation of transfected PC12 cells with cycloheximide for 90 min, both wt-hCgB (Fig. 5 B) and Δcys-hCgB (Fig. 5 D) were detected as a punctate pattern at the periphery of the cell. Punctate structures immunoreactive for wt-hCgB and Δcys-hCgB were also immunoreactive for endogenous rCgB (compare arrowheads in Fig. 5, A vs. B and C vs. D), suggesting the presence of wt-hCgB and Δcys-hCgB in secretory granules. On the other hand, the marker protein for the constitutive pathway, AT, which could be detected as a perinuclear staining without cycloheximide treatment (Fig. 5 E), was barely detectable in cells after 90 min of cycloheximide chase (Fig. 5 F), consistent with its efficient secretion during pulse-chase labeling (Fig. 2 B and C). In summary, we conclude that both wt-hCgB and Δcys-hCgB expressed by transfection are sorted to ISG after exit from the TGN, and are stored in secretory granules.

**Vaccinia Virus Infection of PC12 Cells Blocks Endogenous Protein Synthesis**

We next analyzed sorting of the same recombinant proteins expressed in the absence of endogenous granins. Because all known cell lines bearing the regulated pathway of protein secretion express granins, we set up the vaccinia expression system for PC12 cells. Infection with vaccinia virus has been shown to inhibit host cell protein synthesis and to induce the synthesis of virus-encoded proteins (Moss, 1968).

The viral system was established by infection of PC12 cells with a recombinant virus vv:wt-hCgB which contains wt-hCgB cDNA under the control of the viral 7.5-K promoter. This promoter is active at early and late stages of viral infection. Most importantly, viral infections were performed in the presence of hydroxyurea, a drug known to inhibit the expression of late viral genes (Pogo and Dales, 1971). Thereby viral assembly, in particular, two steps of viral membrane recruitment, at the intermediate compartment (Sodeik et al., 1993) and at the TGN (Schmelz et al., 1994), are blocked, thus preventing disturbance of the secretory pathway.

Infection of PC12 cells with a recombinant virus led to a striking decrease in newly synthesized host cell proteins as determined by pulse labeling of PC12 cells with [35S]me-
thionine. This effect was visible by comparison of total homogenates prepared from either mock-infected or infected PC12 cells (Fig. 6 A, compare total, lanes m and vv). In particular, the cessation of protein synthesis of the two major secretory proteins of PC12 cells, rSgII and rCgB, could be demonstrated after preparation of heat stable fractions (Fig. 6 A, HS). Whereas in heat stable fractions of mock-infected cells rSgII (arrowhead) and rCgB (bracket) were the most prominent bands, in infected cells their expression was reduced to a low level. Residual expression of endogenous granins was due to noninfected cells (see below). At the same time hCgB was expressed under viral control to approximately the same level as either of the two endogenous granins, rSgII and rCgB, and was already visible in total homogenates (Fig. 6 A, total and HS, lanes vv, arrows). The expression level was unchanged from 3 to 8 h PI (data not shown). The identity of hCgB was confirmed by Western blotting of total homogenates with a species-specific antibody that recognizes human, but not rat, CgB. A signal of the expected molecular weight was visible in total homogenates (Fig. 6 A, HS), compare total, lanes m and vv, arrow).

We quantitatively correlated cessation of host cell protein synthesis with the onset of hCgB expression. Expression of hCgB at the cellular level was detected by indirect immunofluorescence with the antibody specific for the human protein. As a marker protein for regulated secretory protein synthesis of the host cell, we followed expression of rSgII. Synthesis of rSgII was monitored by pulse labeling of PC12 cells with [35S]methionine after various times of infection with 0, 20, and 100 pfu per cell. The time courses of the synthesis of rSgII and of the percentage of noninfected cells are shown in Fig. 6 C. For all conditions applied, the block of synthesis of rSgII upon infection was paralleled by the decrease of noninfected cells. By trypan blue staining, after 8 h of infection with 100 pfu/cell, <1% positive cells were detected, indicating that the decrease in rSgII synthesis was not due to cell death (data not shown). The expression of hCgB and shut-off of host cell protein synthesis, therefore, are concurrent events and the remaining expression of rSgII seen at 8 h PI is due to noninfected cells. This implies that the shut-off of host cell protein synthesis starts at the latest with the onset of early viral expression. Furthermore, the decrease in methionine-labeled or sulfate-labeled endogenous rSgII was compared after 3 h PI of either vv:wt-hCgB or vv:Δcys-hCgB at 100 pfu/cell (not shown). Because the same amount in decrease of labeled rSgII was detected for both recombinant viruses, we conclude that the shut-off of endogenous granin synthesis of vv:Δcys-hCgB was as complete as for vv:wt-hCgB.

To obtain direct evidence at the single cell level for a complete block of host cell protein synthesis in infected cells, we used confocal double immunofluorescence. Two constraints of our system have to be considered in this context. First, shut-off at the cellular level cannot be monitored by a lack of endogenous granins because PC12 cells store them in secretory granules located in the cell periphery. Second, immunoreactivity for the endogenous granins (in contrast to that for exogenous hCgB) was, for unknown reasons, not detectable at the level of the PC12 cell Golgi, not even after application of various transport blocks (data not shown). We, therefore, analyzed whether vaccinia virus–expressed hCgB colocalized with endoge-

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Figure 6. Vaccinia virus infection of PC12 cells blocks endogenous protein synthesis. In A and B, PC12 cells were either mock-infected (m) or infected with vv:wt-hCgB (vv) for 8 h with 20 pfu/cell. (A) Cells were pulse labeled for 5 min with [35S]methionine. Equal aliquots of cell homogenates (total) and of heat stable fractions prepared therefrom (HS) were analyzed by SDS-PAGE and fluorography. Full-length hCgB, arrows; rCgB, bracket; rSgII, arrowhead. (B) Immunoblot of cell homogenates (total) for hCgB (arrow). (C) PC12 cells were either mock-infected (0 pfu/cell) or infected with vv:wt-hCgB at 20 or 100 pfu/cell. After the indicated times periods of infection (hrs p. i.), cells were either fixed and analyzed for infection by immunofluorescence for hCgB, or analyzed for the synthesis of rSgII by metabolic labeling with [35S]methionine. The percentage of noninfected cells, defined by the lack of hCgB immunofluorescence (broken lines with diamonds), or the extent of rSgII synthesis, expressed as percentage of that obtained before infection (0 hrs p. i.; continuous lines, filled squares) are shown. Data are from single experiments (0 pfu, 0, 2, 10 hrs p. i.; 20 pfu, all time points; 100 pfu, 0, 6, 8 hrs p. i.), or are the mean of duplicate experiments (0 pfu/cell, 4, 5, 6, 8 hrs p. i.), or of triplicate experiments (100 pfu, 2, 3, 4 hrs p. i.). Error bars indicate SEM or the variation of individual values from the mean.

nous granins at the level of mature secretory granules, as has been shown to be the case for hCgB expressed by transfection (Fig. 5). Lack of colocalization would imply that virally expressed hCgB and endogenous granins were not packaged into the same secretory granule and, thus, were not present in the TGN at the same time. Infected
PC12 cells were chased for 90 min with cycloheximide before fixation. In vv:wt-hCgB–infected cells (Fig. 7A), as was the case in noninfected cells (not shown), endogenous rCgB and rSgII were largely colocalized, with most of the immunostaining being observed at the cell periphery. This result is consistent with both proteins being present in the TGN simultaneously and being copackaged into secretory granules. In contrast, vaccinia virus–expressed wt-hCgB chased to mature secretory granules (see also Figs. 9 and 10) did not colocalize with endogenous rCgB (Fig. 7B) nor endogenous rSgII (Fig. 7C), but was observed as distinct punctate structures distributed largely at the cell periphery and to some extent scattered in the cytoplasm. A quantitative analysis of >100 cells revealed that <5% of punctate signals were positive for both virally expressed hCgB and endogenous granins. Bearing in mind that wt-hCgB in transfected cells colocalized with endogenous granins (Fig. 5), we conclude that the synthesis of endogenous granins is shut off under vaccinia virus infection and that the secretory granules containing endogenous granins were made before shut-off, whereas secretory granules containing hCgB were made after shut-off.

**ER to trans-Golgi Transport Rate Is Unaffected after Prolonged Shut-off**

To analyze the transport of hCgB in the early secretory pathway of infected cells, we monitored the conversion of newly synthesized hCgB to the mature form, i.e., its arrival in the trans-Golgi, by pulse-chase labeling. After a 3-min pulse with [35S]methionine, the ER form of hCgB was detected as a doublet (Fig. 8 top, arrowhead). With increasing chase time, this form shifted to a higher molecular mass form of ~125 kD (Fig. 8 top, arrow), which comigrated with [35S]sulfate-labeled hCgB (data not shown), a marker for the mature form in the TGN (Baeuerle and Huttner, 1987). The calculated t1/2 of transport from ER to Golgi was ~7.5 min (Fig. 8, bottom). In uninfected PC12 cells, transport of rCgB to the trans-Golgi occurred with a t1/2 of ~7 min (Chanat et al., 1993). Our results indicate that host cell protein synthesis shut-off has virtually no effect on ER to Golgi transport of hCgB. Furthermore, the...
MgCl₂ in the absence of calcium, respectively). These data showed that hCgB is stored in a compartment which can undergo maturation as has been shown for ISG in uninfected cells (Tooze et al., 1991), equilibrium gradient analysis was performed after a 90-min chase period. As shown in Fig. 10, under these conditions, the peak of wt-hCgB was shifted by one fraction towards higher density as compared to 12 min of chase (Fig. 10, compare fractions 8 and 9, left, to fractions 9 and 10, right). Notably, after 90 min of chase wt-hCgB was detected in the same gradient fractions as rSgII synthesized in noninfected cells. This suggests that maturation of ISG in infected cells occurred within a similar time window as in noninfected cells. In contrast Δcys-hCgB colocalizing with the peak of hsPG after 12 min of chase was barely detectable on equilibrium gradients after a 90-min chase (not shown) suggesting its constitutive secretion into the medium (see below). We conclude from the gradient analyses that after vaccinia virus expression (a) the majority of wt-hCgB is sorted from the TGN into ISG, which undergo maturation during a 90-min chase and (b) Δcys-hCgB exits from the TGN exclusively into CV.

**Vaccinia-expressed wt-hCgB but Not Δcys-hCgB Is Stored in Secretory Granules**

The differential exit of wt-hCgB and Δcys-hCgB from the TGN indicated sorting of wt-hCgB but not of the mutant to the regulated pathway of secretion. To determine the storage efficiencies of vaccinia-expressed proteins, PC12 cells were infected with either vv:wt-hCgB, vv:Δcys-hCgB, or vv:AT and analyzed by a 5-min pulse with [³⁵S]sulfate followed by a 90-min chase. The recovery of [³⁵S]sulfate-labeled hCgB and AT in cells and media after 90 min of chase was slightly higher than that obtained at the end of the pulse (see Materials and Methods) indicating that no degradation of sulfated proteins occurred under infection conditions. Virally expressed wt-hCgB was stored with an efficiency of 32.2% ± 1.0 SD which is ~4.5 times higher than that of Δcys-hCgB (S_{cys-hCgB} 7.0% ± 2.7 SD; Fig. 11). Notably, after vaccinia expression Δcys-hCgB was secreted like the constitutive marker protein AT (S_{cys-hCgB} 8.3% ± 0.8 SD). This result was different from that obtained after transfection. Together with the data obtained by gradient centrifugation it indicates secretion of virally expressed Δcys-hCgB via CV during 90 min of chase.

The finding that on the one hand ~70% wt-hCgB exited...
the TGN in ISG (Fig. 10, left) and on the other hand the storage efficiency determined after 90 min was $\sim 32\%$ (Fig. 11) suggests that constitutive-like secretion from ISG or an increased basal release of secretory granules occurred. A similar observation was made in the transfection studies (see Discussion).

**Immunofluorescence Analysis of Vaccinia-mediated Expression of wt-hCgB and Δcys-hCgB**

Storage of wt-hCgB but not Δcys-hCgB could also be demonstrated by immunofluorescence analysis of infected cells. 3 h PI, cells were immediately fixed or chased for 90 min in the presence of cycloheximide and subsequently fixed. Immunofluorescence analysis for hCgB is shown in Fig. 12. Without cycloheximide treatment, both wt-hCgB (Fig. 12 A, arrows) and Δcys-hCgB (Fig. 12 C, arrows) were detected as a perinuclear staining, which colocalized with TGN38 (data not shown), a marker protein of the TGN. After cycloheximide treatment, the signal in vv:Δcys-hCgB infected cells was almost completely lost (Fig. 12 D), reflecting its efficient secretion as observed by metabolic pulse-chase labeling (Fig. 11). In contrast, virally expressed wt-hCgB showed a peripheral distribution after a 90-min cycloheximide treatment (i.e., 4.5 h PI), characteristic of secretory granules (Fig. 12 B, arrowheads), that is consistent with its storage observed by pulse-chase analysis. Peripheral staining was already visible, although less pronounced, before incubation with cycloheximide (i.e., 3 h PI) in vv:wt-hCgB (Fig. 12 A, arrowheads) but not in vv:Δcys-hCgB (Fig. 12 C) infected cells. Incubation of vv:wt-hCgB infected cells with cycloheximide for up to 3 h did not significantly decrease the hCgB signal when compared to that obtained after 90 min, suggesting that wt-hCgB is neither secreted nor degraded once it has reached the mature secretory granule (data not shown).

**Sorting of Vaccinia Virus–expressed Δcys-hCgB Is Rescued by Coinfection with a Virus Expressing Full-Length hCgB**

Comparison of the sorting behavior of Δcys-hCgB after
transfection or infection suggests that, in the case of transfection, the lack of the disulfide-bonded loop was compensated by coaggregation of Δcys-hCgB with loop-containing full-length hCgB. If so, double infection of vv:Δcys-hCgB with a recombinant virus expressing full-length hCgB should mimic the transfection conditions and rescue sorting of the mutant to secretory granules. We investigated this issue using [35S]sulfate pulse-chase in combination with subcellular fractionation. This approach has the advantage of providing a direct analysis of protein sorting from the TGN to ISG. In contrast, the three other approaches used to analyze protein sorting to the regulated pathway of secretion (determination of storage efficiency, stimulation of secretion, immunofluorescence) are indirect in that conclusions about sorting at the level of the TGN can only be made if the protein under study that is sorted to ISG is also retained in secretory granules during their maturation.

To investigate whether loop-containing full-length hCgB would rescue the sorting of Δcys-hCgB upon double infection, a recombinant virus, vv:wt-hCgB-EGFP, expressing wt-hCgB tagged with enhanced green fluorescent protein (EGFP; Kaether et al., 1997) was generated. The EGFP tag, which does not interfere with the sorting of wt-hCgB to secretory granules (Kaether et al., 1997), makes it possible to distinguish wt-hCgB from Δcys-hCgB on SDS-PAGE due to a shift in molecular weight. PC12 cells were double-infected with vv:Δcys-hCgB and vv:wt-hCgB-EGFP and subjected to sucrose gradient analysis after sulfate pulse-chase as described (Fig. 10, left, middle) to analyze for coaggregative sorting during exit from the TGN. In parallel the same analysis was performed with PC12

**Figure 12.** Immunofluorescence of PC12 cells expressing wt-hCgB or Δcys-hCgB by infection with recombinant viruses. PC12 cells were infected with vv:wt-hCgB (A and B) or vv:Δcys-hCgB (C and D) for 3 h, and either fixed (A and C) or incubated for an additional 90 min with cycloheximide, and fixed thereafter (B and D). The micrographs show single immunofluorescence for hCgB. Note that wt-hCgB but not Δcys-hCgB was detected in the cell after incubation with cycloheximide. Arrows, perinuclear staining; arrowheads, peripheral staining; all images are of same magnification. Bar (B), 10 μm.

**Figure 13.** Vaccinia coexpression of full-length hCgB rescues sorting of Δcys-hCgB to ISG. PC12 cells were either single-infected with vv:Δcys-hCgB (top, bottom) or double-infected with vv:Δcys-hCgB and vv:wt-hCgB-EGFP (middle, bottom), pulsed for 5 min with [35S]sulfate and chased for 12 min. Postnuclear supernatants were prepared and fractionated by sequential, velocity, and equilibrium sucrose gradient centrifugation. Equal aliquots of the equilibrium gradient (fraction 1, top) were analyzed by SDS-PAGE and phosphoimaging. Top: single infection; □, position of Δcys-hCgB and fragments (curly bracket, see also Fig. 10, middle). Middle: double infection; ●, position of wt-hCgB-EGFP; ○, position of Δcys-hCgB and fragments (curly bracket, see also Fig. 10, middle). Top, middle: endogenous hsPG and rSgII of noninfected cells are indicated by open double arrowheads and open arrows, respectively. Bottom: quantitation of virally expressed recombinant proteins across the equilibrium gradients shown in top and middle panels. The position of CV and ISG is indicated.
Our results obtained by expression of wt-hCgB-EGFP are largely found in fractions 8 and 9, the position of ISG, and to a minor extent in fraction 6, the position of CV (Fig. 15 middle, bottom, △). In double-infected cells, at least 50% of Δcys-hCgB colocalized with wt-hCgB-EGFP at the position of ISG (Fig. 13 middle, bottom, ○), whereas in single infections with vVΔcys-hCgB, the deletion mutant was almost exclusively found at the position of CV (Fig. 13, top, bottom, □; also Fig. 10, middle). In conclusion, coexpression of Δcys-hCgB with wt-hCgB-EGFP rescues sorting of the mutant to ISG in the vaccinia system.

**Discussion**

**The Disulfide-bonded Loop Is Essential for Sorting of CgB during Vaccinia Expression**

In our study we provide biochemical and morphological evidence that the 22 amino acids of hCgB comprising the disulfide-bonded loop are essential for sorting of CgB to secretory granules and that they affect sorting at the level of the TGN. This result was accomplished using a vaccinia virus-based expression system which inhibits host cell protein synthesis. We have shown by different methods that under these conditions wt-hCgB but not Δcys-hCgB is sorted to secretory granules. First, sucrose gradient analysis showed a differential exit of wt-hCgB and Δcys-hCgB from the TGN. Whereas wt-hCgB was sorted mainly into ISG, which were shown to undergo maturation, Δcys-hCgB was found exclusively in CV (Fig. 10). The reduction in sorting efficiency of wt-hCgB in the vaccinia system (∼70% versus ∼100% after transfection) is most likely due to the shut-off of host cell protein synthesis, which could make factors involved in sorting rate-limiting. Consistent with this explanation, we observed that the sorting efficiency of virally expressed wt-hCgB decreased with increasing infection times (data not shown). Second, after pulse-chase labeling, Δcys-hCgB was secreted like the constitutive marker protein AT whereas wt-hCgB was stored ∼4.5-fold more (Fig. 11). Third, immunofluorescence analysis after 90 min of cycloheximide treatment revealed a pattern for wt-hCgB, which was reminiscent of granule staining. For the deletion mutant, no signal was detected after 90 min of cycloheximide treatment reflecting its efficient secretion (Fig. 12). Furthermore, we have demonstrated that sorting of Δcys-hCgB to ISG can be rescued by coexpression with full-length CgB, which resembles the presence of endogenous granins, as in the case of transfection.

From the finding that the disulfide-bonded loop is necessary for the sorting of hCgB to secretory granules, we conclude that the disulfide-bonded loop comprises either a part or the entire sorting signal. This explanation is consistent with the data obtained by Chanat et al. (1993), who showed that reductive cleavage of the disulfide bond of CgB by DTT caused its missorting to the constitutive secretory pathway. However, the previous DTT data did not strictly exclude an effect of DTT on a component of the secretory machinery for CgB, rather than CgB itself. Our results obtained by expression of Δcys-hCgB clearly indicate that missorting is due to the mutation in CgB.

**Sorting of Transfected Δcys-hCgB by Coaggregation**

In contrast to the data obtained with the vaccinia expression system, Δcys-hCgB was efficiently sorted to the regulated pathway after transfection. This was determined by pulse-chase labeling (Fig. 2), sucrose gradient centrifugation (Fig. 3), depolarization-induced calcium-dependent release (Fig. 4), and confocal double immunofluorescence analysis (Fig. 5). How can one reconcile the opposing results for Δcys-hCgB obtained by the two different expression systems? The major difference between the two systems is the ongoing synthesis of endogenous proteins after transfection, in contrast to a shut-off of host cell protein synthesis during viral infection, resulting in the presence or absence of endogenous granins in the TGN (Fig. 14). Ongoing protein synthesis during transfection would allow the formation of aggregates between transfected and endogenous regulated secretory proteins enabling the former to enter the regulated secretory pathway by coaggregation. The fact that approximately one-third of Δcys-hCgB was missorted to CV after transfection may be explained by the formation of homotypic aggregates of the mutant. Because, under transfection, Δcys-hCgB is expressed approximately two to three times higher than endogenous granins (not shown), the probability of homotypic Δcys-hCgB aggregates is increased. Indeed the absence and presence of rescuing molecules (full-length CgB) as the principal difference between the two expression systems is clearly shown by coexpression in the vaccinia system of Δcys-hCgB with full-length CgB, which rescues sorting of the mutant to secretory granules (Fig. 13). This implies that sorting of the deletion mutant is accomplished by coaggregation with loop-bearing CgB, i.e., endogenous rCgB in the case of transfection and recombinant wt-hCgB in the vaccinia system (Fig. 14).

This explanation is supported by the demonstration of heterophilic interactions between regulated secretory proteins in vitro (Gorr et al., 1989; Palmer and Christie, 1992; Colomer et al., 1996). In particular, it was shown that CgB interacts with CgA at low pH (Yoo, 1996). Because the aggregative properties of CgB are maintained after reduction of the disulfide bond (Chanat et al., 1994) and deletion of the disulfide-bonded loop (A. Krömer and H.-H. Gerdes, unpublished observations), Δcys-hCgB, like wt-hCgB, can be expected to interact with other regulated secretory proteins. Taken together, our data suggest that Δcys-hCgB lacking the disulfide-bonded loop, when expressed by transfection, is sorted to the regulated pathway of secretion by coaggregation with endogenous regulated secretory protein, the latter bearing the sorting signal.

**Constitutive-like Secretion in PC12 Cells**

Gradient analysis of transfected PC12 cells showed that wt-hCgB and Δcys-hCgB pulse labeled for 5 min with [35S]sulfate exited completely within 12 min from the TGN into post-TGN vesicles. At the end of a 12-min chase, transfected wt-hCgB was detected exclusively in ISG, whereas Δcys-hCgB was found in ISG and to a minor extent in CV (Fig. 3), indicating that ∼30% of the mutant had entered the constitutive pathway of secretion upon exit from the TGN. These data are consistent with a role of the disulfide-bonded loop in sorting to ISG. We assume
that the proportion of Δcys-hCgB entering CV did not coaggregate with endogenous granins but formed homotypic aggregates (Fig. 14). After 90 min of chase, a time point at which ISG have largely been converted to mature secretory granules (t1/2 ~45 min; Tooze et al., 1991), 70.9 and 40.8% of transfected wt-hCgB and Δcys-hCgB, respectively, were stored in secretory granules (Fig. 2 C). Comparison of the storage efficiencies determined after 90 min of chase with the proportion sorted to ISG after 12 min suggests that a similar proportion of both proteins was secreted in an unstimulated manner after entering the ISG. Two mechanisms, basal release of ISG (Matsuuchi and Kelly, 1991) and constitutive-like secretion (Kuliyawat and Arvan, 1992), have been described which could account for this effect. For PC12 cells it has been shown, following rSgII as a marker protein, that basal release occurred at a very low level if at all (Gerdes et al., 1989; Tooze and Huttner, 1990). Similar results were obtained in this study for transfected PC12 cells (Seff of rSgII: 93.4% ± 2.7 SD). Therefore, it is unlikely to be basal release but rather constitutive-like secretion that accounts for unstimulated secretion of wt-hCgB and Δcys-hCgB between 12 and 90 min of chase. This explanation is consistent with data obtained from pancreatic islet cells, in which constitutive-like secretion was restricted to the maturation period of ISG (Kuliyawat and Arvan, 1992). During vaccinia expression, unstimulated secretion of wt-hCgB (sorting into ISG ~65% versus S_{eff} ~52%) from ISG was observed to a similar extent as in the case of transfection (sorting into ISG ~100% versus S_{eff} ~71%). This suggests that in both systems, transfection and infection, the difference between sorting and storage of wt-hCgB is caused by constitutive-like secretion.

**Sorting during Host Cell Synthesis Shut-off**

The observation that virally expressed wt-hCgB is sorted and packaged into secretory granules under host cell protein synthesis shut-off allows the following conclusions: First, sorting of wt-hCgB to secretory granules is independent of other regulated secretory proteins, i.e., it carries all signals necessary for sorting to the regulated pathway of secretion. Second, involvement of chaperones with a short half-life in the secretory pathway assisting transport or aggregation of wt-hCgB is unlikely. Third, recruitment of other components necessary for granule formation, e.g., membrane proteins and cytosolic factors is, at least for the infection time analyzed, mostly independent of ongoing protein synthesis. However, inhibition of host cell protein synthesis was not analyzed under the conditions used. Because the degree of inhibition varies significantly with viral multiplicity (Moss, 1968) and with cell
type (Bablanian et al., 1978), it remains unclear whether processing and sorting was determined in the presence or absence of host cell protein synthesis.

Secretory granule biogenesis under protein synthesis arrest was subject of a study by Moore and co-workers (Brion et al., 1992). Using cycloheximide to block protein synthesis and analyze glycosaminoglycan chain trafficking, Brion et al. found that formation of secretory vesicles was severely impaired. In contrast to a total block of protein synthesis after cycloheximide application, vaccinia infection inhibits only host cell protein synthesis. Under the latter condition, we have shown that expression of a single regulated secretory protein is sufficient for the formation of secretory granules.

**Analysis of Sorting Signals in Other Regulated Secretory Proteins**

Prosomatostatin is a regulated secretory protein whose sorting to secretory granules has been studied intensively. Its sorting information was shown to reside in the first 82 (Stoller and Shields, 1989) or 78 amino acids (Sevarino et al., 1989) of prosomatostatin. A more detailed mutational analysis of prosomatostatin did not reveal a unique amino acid sequence sufficient for sorting (Sevarino and Stork, 1991). Taking into account that the proregion of somatostatin encodes for more than half of the prohormone, the defined region might exhibit aggregative properties rather than functioning as a sorting signal. Furthermore, by using stimulated secretion to monitor sorting to the regulated pathway of protein secretion, it is not clear to which extent sorting in the TGN or sorting by retention in mature secretory granules had occurred.

Proopiomelanocorticotrophic hormone (POMC) is another model protein for studying sorting to the regulated pathway. When mutants of this hormone with a 25– or 26–amino acid deletion at the NH2 terminus of the proregion were analyzed in Neuro 2A cells, opposing results for the existence of an NH2-terminal–sorting motif were reported (Roy et al., 1991; Chevrier et al., 1993; Cool and Peng Loh, 1994; Cool et al., 1995). Using a retroviral expression system (which does not shut off host cell protein synthesis) Roy et al. showed that the NH2-terminal 26 amino acids are not necessary for sorting. In addition, they provided evidence that sorting information is present in multiple domains of POMC and speculated that cooperation between these domains is needed for sorting of the precursor to secretory granules (Chevrier et al., 1993). In contrast, Cool et al. reported that transfection of a deletion mutant of POMC lacking amino acids 2–26 or 8–20 resulted in its constitutive secretion (Cool and Peng Loh, 1994; Cool et al., 1995). Furthermore, the authors present data that in AtT20 cells the NH2-terminal 26 amino acids of POMC are sufficient to sort the constitutive reporter protein chloramphenicol acetyltransferase to secretory granules (Tam et al., 1993). However, it remains unclear to what degree homo- or heterophilic interactions of the transfected proteins with endogenously regulated secretory proteins were involved in the sorting process. It is possible that such interactions vary considerably with the expression level of the transfected proteins and might account for the opposing results. In addition, as for prosomatostatin, sorting was studied at the level of mature secretory granules, and not in the TGN. Nevertheless, it is interesting that the proposed sorting motif of POMC like the loop of CgB is stabilized by a disulfide bond. This raises the question of whether an exposed loop of amino acids is a general feature of the signal for granular sorting. From theoretical calculations based on the primary sequence analysis of regulated proteins it was suggested that the sorting signal may contain a degenerated amphipathic helix (Kizer and Tropsha, 1991). Consistent with the latter suggestion, the disulfide-bonded loop in hCgB forms an amphipathic helix, as determined by secondary structure prediction.

In summary, our data show that the vaccinia-based expression is a powerful tool for the identification of sorting signals of regulated secretory proteins. Most importantly, it allows us to distinguish sorting signals from aggregative properties. Using this system it should be possible to identify two classes of regulated secretory proteins: (a) proteins containing all the structural information necessary for sorting to secretory granules and which, therefore, are properly sorted when they are present in the TGN as the only cargo protein and (b) proteins lacking a specific sorting signal, but which are competent for aggregation, and whose sorting therefore depends on the former class of regulated secretory proteins. A paradigm for the former class is CgB analyzed in this study. Recently, Natori and Huttner (1996) provided in vivo evidence for a helper function of CgB in sorting of proteins to secretory granules. In AtT20 cells, CgB was shown to promote sorting of the 23-K fragment of POMC, which was further processed in ISG to ACTH. Other candidates that may rely on helper proteins for sorting may be thyrotrpin hormone and luteinizing hormone whose aggregative properties have been analyzed in vitro by Cohn and co-workers (Gorr et al., 1989), and Rindler and co-workers (Colomer et al., 1996), respectively.

We have demonstrated here using the vaccinia expression system that the NH2-terminal disulfide-bonded loop of CgB is necessary for its sorting to secretory granules. Likewise, this system can be used in future studies to analyze sorting of regulated secretory proteins for which transfection studies failed in the identification of a sorting signal. In particular, it will be interesting to identify the nature of sorting signals in regulated secretory proteins that do not contain cysteins, e.g., SgII. It may turn out that exposed loops of amino acids stabilized by other means than disulfide-bonds serve as signal in these proteins.

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