HYBRIDIZATION AND MOLECULAR CHARACTERIZATION OF F₁ ALLIUM CEPA × ALLIUM ROYLEI PLANTS

ALICJA CHUDA* AND ADELA ADAMUS

Department of Genetics, Plant Breeding and Seed Science, University of Agriculture in Cracow, 29 Listopada 54, 31-425 Cracow, Poland

Received July 31, 2012; revision accepted September 13, 2012

We made interspecific crosses to facilitate the introgression of desirable traits of Allium roylei into the Allium cepa genome. After hand-pollination, 906 interspecific F₁ Allium cepa × A. roylei plants were obtained by in vitro culture via embryo rescue. Nuclear DNA analysis showed that 97.6% of the regenerants were interspecific F₁ Allium cepa × A. roylei hybrids. Genomic in situ hybridization (GISH) showed that each hybrid had 16 chromosomes, eight of which were identified as A. cepa and eight as A. roylei chromosomes.

Key words: Allium cepa, A. roylei, embryo rescue, genomic in situ hybridization (GISH), interspecific hybrids, molecular markers.

INTRODUCTION

The genus Allium comprises about 750 species including A. cepa, A. fistulosum and A. roylei. Onion (Allium cepa L.) is a valuable vegetable crop preceded on the list of worldwide cultivated vegetable species only by tomato (FAOSTAT, 2011). Onion breeding programs currently focus mainly on improvement of existing cultivars. According to Kik (2002), cultivated and wild Allium species possess many desirable traits (disease and pest resistance, cytoplasmic male sterility, bulb quality) which might be useful for genetic improvement of the bulb onion. Of all the species belonging to Allium, A. cepa, A. vavilovii, A. galanthum, A. roylei, A. fistulosum, A. altaicum, A. pskemense and A. oschaninnii have been recognized as the most important gene pools of onion (Shigyo and Kik, 2008).

According to Peffley and Hou (2000), Kik (2002) and Yamashita et al. (2005), Allium fistulosum possesses many traits agronomically useful for onion. Due to its resistance to onion leaf blight, pink root, anthracnose and onion fly, as well as its earliness, high dry matter content and winter-hardiness, Japanese bunching onion has been used to expand the genetic variation of onion since the last century. Studies of interspecific hybridization between A. cepa and A. fistulosum initiated by Emsweller and Jones (1935) have been continued over the years by Dolezel et al. (1980), van Raamsdonk et al. (1992), van der Valk et al. (1991a,b), Khrustaleva and Kik (1998), Peffley and Hou (2000) and Mangum and Peffley (2005). Unfortunately, subsequent backcrosses of the obtained F₁ A. cepa × A. fistulosum hybrids to A. cepa have proved problematic. The difficulties observed in genome transmission in advanced backcross generations have been explained by the sterility of F₁ A. cepa × A. fistulosum hybrids probably resulting from one of three possible mechanisms: stylar incongruity (van der Valk et al., 1991a), nuclear cytoplasmic incompatibility (Ulloa et al., 1995) or central cell nuclear cytoplasmic incongruity (Mangum and Peffley, 2005).

Due to problems in introgression of Japanese bunching onion traits into onion, so far only four new varieties of A. fistulosum have been developed: Beltsville Bunching, Delta Giant, Top Onion and Wakegi Onion (Kik, 2002).

Allium roylei Stearn is a wild species originating from the Indian subcontinent which harbors disease resistance against downy mildew (Peronospora destructor), leaf blight (Botrytis squamosa) and anthracnose (Colletotrichum gloeosporioides) (van der Meer and de Vries, 1990; de Vries et al., 1992; Galvan et al., 1997; Scholten et al., 2007; Kohli and Gohil, 2009). This species has been used to expand the genetic variation of onion since 1960 (Fritsch and Friesen, 2002). The first crosses between A. roylei and A. cepa were made in 1985 to achieve introgression of downy mildew resistance into onion.
Subsequent studies on reciprocal *Allium cepa* and *A. roylei* hybridization (Kofoet et al., 1990; Kofoet and Zinkernagel, 1990) led to the conclusion that resistance to *Peronospora destructor* is determined by a single dominant gene, *Pd1*. Later, AFLP marker technique combined with SCAR analysis confirmed this (van Heusden, 2000a,b). A long-term investigation on interspecific hybridization and downy mildew resistance led Scholten et al. (2007) to obtain F1BC5-generation plants expected to be resistant to *Peronospora destructor*.

In this study we sought to obtain and characterize interspecific F1 *Allium cepa* × *A. roylei* hybrids. To our knowledge this is the first report of the use of embryo rescue technique in vitro to produce interspecific F1 *Allium cepa* × *A. roylei* hybrids. To identify the obtained hybrids we generated a system of three novel independent molecular markers. We also developed a genomic in situ hybridization (GISH) protocol in order to determine the cytological characteristics of the generated F1 hybrids.

**MATERIALS AND METHODS**

PRODUCTION OF F1 *A. CEPA* × *A. ROYLEI* PLANTS VIA EMBRYO RESCUE

In 2004–2007 we crossed six different onion breeding lines (A1–A6) and two cultivars (Bila, Kutnowska) with two *Allium roylei* ecotypes (333, 334). The onion breeding materials came from three Polish seed companies: PlantCo Zielonki, Spojnia Nochowo and KHNO Polan. *A. roylei* plants were kindly provided by Dr. T. Kotlinska of the Plant Genetic Resources Laboratory, Research Institute of Vegetable Crops (RIVC), Skierkiewice, Poland.

*Allium cepa* and *A. roylei* plants were grown in the greenhouse from February to June or in the field from April to August. The interspecific crosses were performed in a screen-covered cage in the greenhouse from June to August. Flowering plants from maternal stocks were subsequently emasculated (Fig. 1a) and manually pollinated with *A. roylei* pollen (Fig. 1b). At 7–14 days after pollination, ovaries (Fig. 1c) were harvested and sterilized in 70% ethanol for 1 min followed by 20% chloramine B for 20 min. The material was then washed three times in sterile water. Ovules were aseptically excised from ovaries and placed on two different induction media (Fig. 1d). Medium A was based on B5 basal medium (Gamborg et al., 1968) with 20 g/l sucrose and 7.0 g/l agar, pH 6.0; medium B was based on BDS basal medium (Dustan and Short, 1977) with 20 g/l sucrose and 7.0 g/l agar, pH 6.0. The media were autoclaved at 121°C for 20 min. Twenty ovules were cultured on each 60 × 10 mm Petri dish. The dishes were sealed with parafilm and kept at 25±2°C in the dark. After ca. one month, at the stage of leaf and root formation, developing embryos (Fig. 1e) were transferred to MS medium (Murashige and Skoog, 1962) with 30 g/l sucrose and 7.0 g/l agar, pH 6.0, and kept in glass tubes in a growth chamber at 25±2°C under a 16 h photoperiod and light intensity 55 μms-1m-2 (Fig. 1f). The developed plantlets were placed in soil and acclimatized to ex vitro conditions in a chamber with regulated temperature and humidity. After acclimatization, F1 *A. cepa* × *A. roylei* plants (Fig. 1g) were cultivated in the greenhouse.

The total effectiveness of F1 *A. cepa* × *A. roylei* embryo development is expressed as the number of developed embryos per number of ovules plated on media, and the total effectiveness of F1 *A. cepa* × *A. roylei* plants through in vitro culture is expressed as the number of acclimatized plants per number of ovules cultured.

**MOLECULAR MARKERS**

The hybrid nature of all of the obtained F1 *Allium cepa* × *A. roylei* regenerants was verified with three different molecular markers (Tab. 1). SIR, ACS and A markers were developed on the basis of three *A. cepa* sequences available in the NCBI (2005). SIR-F/SIR-R primers were based on the *A. cepa* sulfite reductase gene. ACS-F/ACS-R primers were designed on the basis of the *A. cepa* anthocyanidin synthase gene. A-F/A-R primers synthesized accord-
ing to van Heusden et al. (2000a,b) were based on the A. cepa alliinase gene.

Total genomic DNA was isolated from young leaves of A. cepa, A. roylei and all the F₁ regenerants with the DNeasy Plant Maxi Kit (Qiagen, Germany). PCR was performed in a 20 μl reaction containing 0.05 U Taq DNA polymerase (recombinant, Fermentas), 10 × Taq Buffer with KCl (Fermentas), 2 mM MgCl₂, 0.25 mM dNTP Mix (Fermentas), 0.5 μM of each primer and 10 ng plant DNA. DNA was amplified in a Master Gradient (Eppendorf) thermal cycler programmed as follows: 94°C/2 min; 35 cycles of [94°C/30 s, 58°C/60 s, 68°C/120 s] for SIR-F/SIR-R primers, [94°C/30 s, 65°C/30 s, 68°C/90 s] for ACS-F/ACS-R primers or [94°C/30 s, 51°C/60 s, 68°C/120 s] for A-F/A-R primers; and 68°C/5 min. Products of amplification with SIR-F/SIR-R and A-F/A-R primers were resolved for 2 h in 1% agarose gel in 1 × TBE buffer. PCR products obtained after amplification with ACS-F/ACS-R primers were resolved for 4 h in 6% polyacrylamide gel in 1 × TBE buffer. After electrophoresis the gels were stained with ethidium bromide, visualized under UV transillumination and photographed.

CHROMOSOME PREPARATION AND GENOMIC IN SITU HYBRIDIZATION (GISH)

Twenty-four F₁ Allium cepa × A. roylei hybrids were analyzed in terms of genomic in situ hybridization (GISH). Chromosome composition was examined in hybrids originating from three different cross combinations: F₁ A. cepa cv. Kutnowska × A. roylei ecotype 333, F₁ A. cepa cv. Bila × A. roylei ecotype 333, and F₁ A. cepa A5 breeding line × A. roylei ecotype 333.

Young roots of the hybrids were treated for 8 h in 0.03% aqueous solution of 8-hydroxyquinoline at 15°C and fixed in a 3:1 solution of methanol and glacial acetic acid. The root tips were rinsed three times in sterile water and incubated in 20 μl aqueous enzyme mixture containing 4% Onozuka cellulase (Serva Electrophoresis, Heidelberg, Germany) and 2% Pectolyase Y-23 (Sigma Aldrich), pH 4.8, for 1 h at 37°C (Pląder et al., 1998). The macerated root tips were spread by dissecting and squashing them on grease-free slides and cooled to -20°C in a drop of fixative solution. Under a phase-contrast microscope, slides with a full set of chromosomes in well-spread metaphases (2n=16) were selected for GISH experiments.

In this experiment we used total genomic DNA extracted from young A. roylei (ecotype 333) and A. cepa (cv. Kutnowska) leaves as probe and blocking DNA, respectively. DNA isolation was done with the DNeasy Plant Maxi Kit (Qiagen). To prepare the probe, Allium roylei DNA was autoclaved for 4 min, which generated 100–1500 bp fragments, and labeled with digoxigenin-11-dUTP using a Nick Translation Kit (Roche Diagnostics, Germany). To prepare the block, isolated onion DNA was replicated with the REPLI-g Midi Kit (Qiagen) to increase the DNA concentration and autoclaved for 12 min, yielding 100–500 bp fragments.

The hybridization mixture (30 μl per slide), denatured before hybridization for 10 min at 80°C, contained 50% deionized formamide, 10% sodium dextran sulfate, 20 × SSC, 50 ng/μl probe DNA, 3.5 μg/μl blocking DNA and 5.5 ng/μl DNA from herring sperm (Sigma Aldrich). Slides with metaphase spreads were denatured in 70% formamide for 1.5 min at 80°C, washed in an ethanol series (70%, 90%, and 100% for 5 min each) and air-dried. Hybridization was performed overnight in a humid chamber at 37°C. Steps of posthybridization washes and detection were performed according to Dong et al. (2000) with some minor modifications. Digoxigenin-labeled probes were detected with 1% fluorescein isothiocyanate (FITC)-conjugated antidigoxigenin antibody (Roche Diagnostics). Chromosomes were counterstained with 4′,6-diamidino-2-phenylindole (DAPI) in ProLong Gold antifade reagent (Invitrogen). Slides were examined with an Axio Imager M1 microscope equipped with separate excitation filter sets for DAPI and FITC (Carl Zeiss MicroImaging). Images were captured digitally with an AxioCam and Axio Vision Release 4.6.3. (Carl Zeiss MicroImaging). We used Adobe Photoshop ver. 9.0.2 (San Jose, California, U.S.A.) to adjust the images and optimize the contrast and background of the presented figures.

---

TABLE 1. Primer sequences based on three onion cDNA clones

| cDNA clone | Marker | Primers | Primer sequence (5’-3’)
|------------|--------|---------|------------------|
| AY753557   | SIR    | SIR-F   | AGTCACTGTGCCAACCTGACAACCTCAG |
|            |        | SIR-R   | TAACCCAAATGTCGACCCAAACGAGG  |
| AY585678   | ACS    | ACS-F   | AAGCCTGATAATTGACCCACACCAGCTCC |
|            |        | ACS-R   | ATGCCAGATGTTGCGGGGGGCATTGT   |
| L48614     | A      | A-F     | TGGGTACTCCATAAGCCAGTG       |
|            |        | A-R     | TGCAACCTTCGGAGAACAG          |
RESULTS

PRODUCTION OF F1 ALLIUM CEPA × A. ROYLEI PLANTS

During the four years of the studies we pollinated 370 A. cepa inflorescences with A. roylei pollen (Tab. 2) and plated 35,880 ovules on regeneration media, from which 1768 (4.9%) embryos started to develop. Analysis of F1 embryo development revealed that cv. Kutnowska was the most embryogenic (7.1%), and the least embryogenic (0.5%) was the A4 breeding line. The mean effectiveness of F1 plant regeneration in vitro was 55.0%. After acclimatization to ex vitro conditions, 906 (93.1%) F1 Allium cepa × A. roylei plants were produced via embryo rescue (Tab. 2).

CONFIRMATION OF HYBRIDITY

PCR analysis with SIR-F/SIR-R, ACS-F/ACS-R and A-F/A-R primers was performed for 906 F1 Allium cepa × A. roylei regenerants.

Following PCR with SIR-F/SIR-R primers, the presence of a monomorphic 400 bp product was revealed for A. cepa, A. roylei and all examined F1 plants (Fig. 2). For paternal plants, the SIR marker produced an additional amplicon of 1.2 kb. The presence of this product in the profiles confirmed that the examined regenerants were interspecific Allium cepa × A. roylei hybrids.

For the A marker, amplification resulted in three different fragments (Fig. 3). Two of them (350 bp, 750 bp) were characteristic of the maternal plants, while the third amplicon (800 bp) was characteristic of A. roylei. This product, present in electrophoretic profiles of the regenerants, confirmed their hybridity.

Use of ACS-F/ACS-R primers yielded a fragment of 760 bp, observed for both A. roylei ecotypes (Fig. 4). Additionally, the ACS marker generated five different amplicons (770–780 bp) characteristic of five maternal genotypes (A3, A4, A5, A6 lines, cv. Kutnowska). The profiles of F1 Allium cepa × A. roylei hybrids had two products characteristic of the maternal and paternal plants.

Molecular analysis carried out with SIR, A and ACS markers revealed amplicons specific to both A. cepa and A. roylei plants in 97.6% of the examined regenerants. These results confirmed that 884 of 906 plants were true F1 interspecific Allium cepa × A. roylei hybrids.

GENOMIC CONSTITUTION

To determine their chromosome composition, we used genomic in situ hybridization (GISH) to analyze 24 interspecific F1 Allium cepa × A. roylei hybrids. Both parental species have 16 chromosomes (2n). GISH revealed that all the F1 hybrids also had 16 chromosomes (2n). No differences in chromosome morphology, deletions or additional chromosomes were observed in the spread metaphases of any of the examined plants (Fig. 5a–c). We employed total genomic DNA from A. roylei as probe together with A. cepa as blocking DNA to determine the parental origin of the chromosomes of F1 A. cepa cv. Kutnowska × A. roylei ecotype 333, F1 A. cepa cv. Bila × A. roylei ecotype 333, and F1 A. cepa A5 breeding line × A. roylei ecotype 333 hybrids (Fig. 5a–c). In GISH profiles of every analyzed hybrid plant, eight chromosomes hybridized with A. roylei probe DNA and showed green fluorescence; the other eight chromosomes displayed blue fluorescence, confirming their maternal origin. No recombinant chromosomes were observed in the examined metaphase spreads (Fig. 5a–c).

| Maternal form (Allium cepa) | No. of A. cepa inflorescences | No. of ovules plated on media (A+B) | Embryo development | Plant regeneration | Plant acclimatization |
|---------------------------|-------------------------------|------------------------------------|-------------------|-------------------|----------------------|
| A1                        | 28                            | 3455                               | 130               | 87                | 83                   |
| A2                        | 10                            | 1166                               | 15                | 7                 | 6                    |
| A3                        | 43                            | 3923                               | 176               | 97                | 95                   |
| A4                        | 18                            | 1306                               | 7                 | 1                 | 1                    |
| A5                        | 81                            | 8175                               | 387               | 236               | 226                  |
| A6                        | 88                            | 7703                               | 424               | 209               | 197                  |
| ‘Bila’                    | 52                            | 5623                               | 309               | 151               | 137                  |
| ‘Kutnowska’               | 50                            | 4529                               | 320               | 185               | 161                  |
| Total                     | 370                           | 35880                              | 1768              | 973               | 906                  |

Mean (%) 4.9 55.0 93.1
DISCUSSION

Embryo rescue methods have enabled successful intergeneric and interspecific hybridization of many vegetables (Lycopersicon, Solanum), cereals (Oryza, Triticum) and fruits (Actinidia, Fragaria), helping to expand genetic variability within many plant genera (Sharma et al., 1996). This technique has also helped increase the pool of genetic resources within the genus Allium. Crosses between onion and numerous distant Allium species followed by embryo development in vitro have produced a considerable number of interspecific hybrids (Gonzalez and Ford-Lloyd, 1987; Keller et al., 1996). In vitro culture has also been applied to produce interspecific hybrids within edible Alliaceae. After crossing Allium cepa and Allium fistulosum plants and culturing hybrid embryos, Dolezel et al. (1980) obtained both reciprocal interspecific A. cepa × A. fistulosum hybrids. Nascent embryo rescue yielded interspecific hybrids between onion and fertile garlic (Ohsumi et al., 1993). Yanagino et al. (2003) used ovary culture to produce an interspecific A. ampeloprasum × A. sativum hybrid, and Peterka et al. (1997) combined ovary and ovule culture to hybridize onion with leek. Unfortunately, embryo rescue technique proved ineffective in an attempt to obtain interspecific A. cepa × A. schoenoprasum hybrids (Gonzalez and Ford-Lloyd, 1987).

In this study we obtained a large population of F1 Allium cepa × A. roylei hybrids through embryo culture. To our knowledge this is the first report of production of such hybrids in culture in vitro. Others have succeeded in hybridizing A. cepa with A. roylei, but van der Meer and de Vries (1990) and Scholten et al. (2007) obtained only individual hybrid plants from seeds harvested after hand-pollination of both parental species. We achieved 2.5% overall efficiency of production of interspecific F1 Allium cepa × A. roylei hybrids through embryo rescue. This compares with 1.6% mean efficiency of production of F1 Allium cepa × A. roylei hybrids from seeds harvested after crossing A. cepa and A. roylei plants (data not shown). These results are in accord with those of Dolezel et al. (1980) and Gonzalez and Ford-Lloyd (1987), who used embryo rescue technique to increase the effectiveness of reciprocal Allium cepa × A. fistulosum hybridiza-
tion versus normal seed sowing by 40% and 70%, respectively. In vitro culture evidently can facilitate hybrid embryo development and overcome genetic incompatibility between taxonomically distant parental species and the post-fertilization barriers that hinder seed maturation in planta.

The difference in the amount of DNA between *Allium cepa* (33.5 pg DNA/2C) and *A. roylei* (28.5 pg DNA/2C) is only 5 pg (Khrustaleva and Kik, 1998). Due to the similarity of nuclear DNA content of the parental species, the hybridity of the obtained F₁ *A. cepa × A. roylei* plants could not be examined by flow cytometry. That is why we verified the hybrid character of the obtained F₁ regenerants by nuclear DNA analysis. We know of only a few reports on practical application of molecular markers to confirm the hybrid nature of plants obtained from crosses within the genus *Allium* (Ohsumi et al., 1993; Bark et al., 1994; Yanagino et al., 2003), so it seemed worthwhile to develop molecular markers to differentiate *A. cepa* and *A. roylei* accessions, which would be useful tools for confirming the hybrid nature of plants obtained from crossing the two species. In this study we developed three novel, independent molecular markers which confirmed almost all (97.6%) of the regenerants as true interspecific hybrids. As expected, each marker disclosed polymorphic fragments from both parental species in the molecular profiles of F₁ interspecific hybrids. These molecular studies developed three nuclear DNA markers useful in assessing the level of DNA polymorphism between *A. cepa* and *A. roylei* species. The high percentage of F₁ plant population hybridity demonstrates that embryo rescue technique is an effective method for producing F₁ *Allium cepa × A. roylei* hybrid plants.

In this work, genomic in situ hybridization (GISH) clearly distinguished *A. cepa* and *A. roylei* chromosomes in the F₁ hybrids. The overriding aim of this project is to introgress the downy mildew resistance locus from *A. roylei* into the bulb onion genome. This method should prove useful for genomic constitution analysis of subsequent backcross (*A. cepa × A. roylei) × Allium cepa* generations. Previous studies on introgression of downy mildew resistance into onion based on AFLP markers and GISH analysis revealed that the *A. roylei* fragment harboring the downy mildew resistance probably is at the distal end of the long arm of chromosome 3 of the analyzed backcross progenies (van Heusden et al., 2000b; Scholten et al., 2007).

The next step of our investigation will entail advanced backcrosses of the obtained F₁ *Allium cepa × A. roylei* hybrids with the bulb onion in order to eliminate undesirable *A. roylei* traits. Preliminary studies in this area suggest that due to the genetic distance between *A. cepa* and *A. roylei* species, the fertility of F₁ hybrid plants might be impaired, probably as a result of lack of homology between the parental genomes. Currently our research focuses on explaining the barriers that disable successful (*Allium cepa × A. roylei) × Allium cepa* backcrosses.

**ACKNOWLEDGMENTS**

We thank Dr. Ewa Grzebelus for help and advice with genomic in situ hybridization, and Dr. Dariusz Grzebelus for assistance with molecular markers. Financial support from the Polish Ministry of Agriculture and Rural Development (grant no. G6513) and the Polish Ministry of Science and Higher Education (grant no. N301 041 32/1508) is gratefully acknowledged.

**REFERENCES**

Bark OH, Havey MJ, and Corgan JN. 1994. Restriction fragment length polymorphism (RFLP) analysis of progeny from an *Allium fistulosum × A. cepa* hybrid. *Journal of the American Society for Horticultural Science* 119 (5): 1046–1049.

Dolezel J, Novak FJ, and Luzný J. 1980. Embryo development and in vitro culture of *Allium cepa* and its interspecific hybrids. *Zeitschrift für Pflanzenzüchtigung* 85: 177–184.

Dong F, Song JS, Naess K, Helgeson JP, Gebhardt G, and Jiang J. 2000. Development and applications of a set of chromosome specific cytogenetic DNA markers in potato. *Theoretical and Applied Genetics* 101: 1001–1007.

Dustan DI, and Short KC. 1977. Improved growth of tissue cultures of the onion, *Allium cepa*. *Physiologia Plantarum* 41: 70–72.

Emsweller SL, and Jones HA. 1935. An interspecific hybrid in *Allium. Hilgardia* 9: 265–273.

FAOSTAT. http://faostat.fao.org/. Accessed 02 August 2011.

Fritsch RM, and Friesen N. 2002. Evolution, domestication and taxonomy. In: Rabinowitch HD and Currah L [eds], *Allium Crop Science: Recent Advances*, 5–38. CABI Publishing Oxon UK.

Galvan GA, Wietsma WA, Putrasemedja S, Permadji AH, and Kir C. 1997. Screening for resistance to anthracnose (*Colletotrichum gloeosporioides* Penz.) in *Allium cepa* and its relatives. *Euphytica* 95(2): 173–178.

Gambaro OL, Miller RA, and Ojima O. 1968. Nutrient requirements of suspension cultures of soybean root cell. *Experimental Cell Research* 50: 151–158.

Gonzalez LG, and Ford-lloyd BV. 1987. Facilitation of wide-crossing through embryo rescue and pollen storage in interspecific hybridization of cultivated *Allium* species. *Plant Breeding* 98: 318–322.

Keller ERJ, Schubert I, Fuchs J, and Meister A. 1996. Interspecific crosses of onion with distant *Allium* species and characterization of the presumed hybrids.
by means of flow cytometry, karyotype analysis and genomic in situ hybridization. *Theoretical and Applied Genetics* 92: 417–424.

**Khristaleva LI** and **Kik C.** 1998. Cyto-genetical studies in the bridge cross *Allium cepa* × (*A. fistulosum* × *A. roylei*). *Theoretical and Applied Genetics* 96: 8–14.

**Kik C.** 2002. Exploitation of wild relatives for the breeding of cultivated *Allium* species. In: Rabinowitch HD and Currah L [eds], *Allium Crop Science: Recent Advances*, 81–100. CABI Publishing Oxon UK.

**Kohli B.** and **Gohil RN.** 2009. Need to conserve *Allium roylei* Steinr: a potential gene reservoir. *Genetic Resources and Crop Evolution* 56: 891–893.

**Kofoet A.** and **Zinkernagel V.** 1990. Resistance to downy mildew (*Peronospora destructor* (Berk.) Cresp.) in *Allium* species. *Journal of Plant Diseases and Protection* 97(1): 13–23.

**Kofoet A.** **Kik C.** **Wietsma WA.** and **de Vries JN.** 1990. Inheritance of resistance to downy mildew (*Peronospora destructor* [Berk.] Cresp.) from *Allium roylei* Steinr in the backcross *Allium cepa* L. × (*A. roylei* × *A. cepa*). *Plant Breeding* 105: 144–149.

**Manjun PD.** and **Peffley EB.** 2005. Central cell nuclear-cytoplasmic incongruity: a mechanism for segregation distortion in advanced backcross and selfed generations of [(*Allium cepa* L. × *A. fistulosum* L.) × *A. cepa*] interspecific hybrid derivatives. *Cytogenetic and Genome Research* 109: 400–407.

**Murashige T.** and **Skroog F.** 1962. A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum* 15: 473–497.

**NCBI.** http://www.ncbi.nlm.nih.gov/. Accessed 17 January 2005.

**Ohsumi C.** **Kojima A.** **Hinata K.** **Etoh T.** and **Hayashi T.** 1993. Interspecific hybrid between *Allium cepa* and *Allium sativum*. *Theoretical and Applied Genetics* 85: 969–975.

**Peffley EB.** and **Hou A.** 2000. Bulb-type onion introgressants possessing *Allium fistulosum* L. genes recovered from interspecific hybrid backcrosses between *A. cepa* L. and *A. fistulosum* L. *Theoretical and Applied Genetics* 100: 528–534.

**Peterka H.** **Budin H.** and **Schrader O.** 1997. Interspecific hybrids between onion (*Allium cepa* L.) with S-cytoplasm and leek (*Allium ampeloprasum* L.). *Theoretical and Applied Genetics* 94: 383–389.

**Plader W.** **Hoshi Y.** and **Malepszy S.** 1998. Sequential fluorescent staining with CMA and DAPI for somatic chromosome identification in cucumber (*Cucumis sativus* L.). *Journal of Applied Genetics* 39(3): 249–258.

**Scholten OE.** **van Heusden AW.** **Khristaleva LI.** **Burger-Meljer K.** **Mank RA.** **Antonise RGC.** **Harrewijn JL.** **van Haecke W.** **Oost EH.** **Peters RJ.** and **Kik C.** 2007. The long and winding road leading to the successful introgression of downy mildew resistance into onion. *Euphytica* 156: 345–353.

**Sharma DR.** **Kaur R.** and **Kumar K.** 1996. Embryo rescue in plants – a review. *Euphytica* 89: 325–337.

**Shigyo M.** and **Kik C.** 2008. Onion. In: Prohens J. Nuez F [eds], *Vegetables II: Fabaceae, Liliaceae, Solanaceae and Umbelliferae*, 121–159. Springer New York.

**Ulloa GM.** **Corgan JN.** and **Dunford M.** 1995. Evidence for nuclear-cytoplasmic incompatibility between *Allium fistulosum* and *A. cepa*. *Theoretical and Applied Genetics* 90: 746–754.

**van Heusden AW.** **van Ooijen JW.** **Vrielink-van Ginkel R.** **Verbeek WHJ.** **Wietsma WA.** and **Kik C.** 2000a. A genetic map of an interspecific cross in *Allium* based on amplified fragment length polymorphism (AFLP™) markers. *Theoretical and Applied Genetics* 100: 118–126.

**van Heusden AW.** **Shigyo M.** **Tashiro Y.** **Vrielink-van Ginkel R.** and **Kik C.** 2000b. AFLP linkage group assignment to the chromosomes of *Allium cepa* L. via monosomic addition lines. *Theoretical and Applied Genetics* 100: 480–486.

**van der Meer QP.** and **de Vries JN.** 1990. An interspecific cross between *Allium roylei* Steinr and *Allium cepa* L., and its backcross to *A. cepa*. *Euphytica* 47: 29–31.

**van Raamsdonk LWD.** **Wietsma WA.** and **de Vries JN.** 1992. Crossing experiments in *Allium* L. section *Cepa*. *Botanical Journal of the Linnean Society* 109: 293–303.

**van der Valk P.** **de Vries SE.** **Everink JT.** **Vorstappen F.** and **de Vries JN.** 1991a. Pre- and post-fertilization barriers to backcrossing the interspecific hybrid between *Allium fistulosum* L. and *A. cepa* L. with *A. cepa*. *Euphytica* 53: 201–209.

**van der Valk P.** **Kik C.** **Vorstappen F.** **Everink JT.** and **de Vries JN.** 1991b. Independent segregation of two isozyme markers and inter-plant differences in nuclear DNA content in the interspecific backcross (*Allium fistulosum* L. × *A. cepa* L.) × *A. cepa* L. *Euphytica* 55: 151–156.

**de Vries JN.** **Wietsma WA.** and **de Vries JN.** 1992. Introgression of leaf blight resistance from *Allium roylei* Steinr into onion (*A. cepa* L.). *Euphytica* 62: 127–133.

**Yamashita K.** **Takatori Y.** and **Tashiro Y.** 2005. Chromosomal location of a pollen fertility-restoring gene, Rf, for CMS in Japanese bunching onion (*Allium fistulosum* L.) possessing the cytoplasm of *A. galanthum* Kar. et Kir. revealed by genomic in situ hybridization. *Theoretical and Applied Genetics* 111: 15–22.

**Yanagino T.** **Sugawara E.** **Watanabe M.** and **Takahata Y.** 2003. Production and characterization of an interspecific hybrid between leek and garlic. *Theoretical and Applied Genetics* 107: 1–5.