Long non-coding RNA NORAD regulates angiogenesis of human umbilical vein endothelial cells via miR-590-3p under hypoxic conditions

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Abstract. Dysregulation of angiogenesis can be caused by hypoxia, which may result in severe diseases of the heart, including coronary artery disease. Hypoxia-inducible factor 1 (HIF-1) modulates angiogenesis via the regulation of several angiogenic factors. However, the underlying mechanism of hypoxia-induced angiogenesis remains unknown. In the present study, it was hypothesized that long non-coding RNA (IncRNA) non-coding RNA activated by DNA damage (NORAD) may serve a role in the process of angiogenesis via the regulation of microRNA(miR)-590-3p under hypoxic conditions. The effect of NORAD and miR-590-3p on cell viability and properties associated with angiogenesis, including cell migration and tube formation in human umbilical vein endothelial cells (HUVECs) under hypoxic conditions, were assessed. Potential downstream angiogenic factors of miR-590-3p were also determined by molecular experiments. It was identified that NORAD expression was upregulated and miR-590-3p expression was downregulated in hypoxia-exposed HUVECs, and also in myocardial infarction (MI) left ventricle tissues in mice. Moreover, down-regulation of NORAD expression resulted in decreased cell viability and angiogenic capacity, but further knocking down miR-590-3p expression reversed these alterations, resulting in increased cell migration and tube formation in HUVECs under hypoxic conditions for 24 h. It was demonstrated that NORAD overexpression also increased cell vitality and tube-formation capacity. Furthermore, NORAD was identified to bind with miR-590-3p directly, and miR-590-3p was shown to target certain proangiogenic agents, such as vascular endothelial growth factor (VEGF)A, fibroblast growth factor (FGF)1 and FGF2 directly. Therefore, the present results suggested that IncRNA NORAD may bind with miR-590-3p to regulate the angiogenic ability of HUVECs via the regulation of several downstream proangiogenic factors under hypoxia. Thus, the IncRNA NORAD/miR-590-3p axis may be a novel regulatory pathway in the angiogenic mechanisms in HUVECs, which highlights a potentially novel perspective for treating ischemia/hypoxia-induced angiogenic diseases.

Introduction

Angiogenesis is a process in which blood vessels are formed from pre-existing vessels, and is vital for vascularization during embryonic development and vessel formation in various organs and tissues (1,2). Vascular endothelial growth factor (VEGF) has been studied as a fundamental growth factor involved in blood vessel formation (3). Moreover, tight regulation of VEGF expression is required during angiogenesis, but this regulation may be altered by certain physiological signals, particularly in hypoxia (3,4). Hypoxia-inducible factor 1 (HIF-1) exerts its effects on every stage of angiogenesis via activation of various angiogenic factors, such as VEGF, or regulation of proangiogenic chemokines (5-7). VEGF also increases the expression of other proangiogenic factors, including fibroblast growth factor (FGF) (5). FGF1 and FGF2 bind to FGF receptors and serve crucial roles in the angiogenic process (8). Hypoxia is primarily caused by ischemia in vivo, which can result in severe diseases, such as coronary artery diseases (9). Coronary artery diseases are one of the leading causes of mortality worldwide (10). Furthermore, ischemic heart disease results in >1 million mortalities worldwide each year, and the morbidity and mortality rates have remained high, particularly in developing countries (10,11). It has been demonstrated that brief continual myocardial ischemia enhances blood vessel formation in the adult heart, resulting in improvement to blood supply and improvement in ischemic areas, which may prevent extended ischemic disorders (12). However, the detailed mechanisms of ischemia/hypoxia-induced angiogenesis are not fully understood.

Long non-coding RNAs (lncRNAs), which are non-coding sequences >200 nucleotides in length, are involved in various physiological and pathological processes, and diseases (13,14). In the heart, a conserved IncRNA mechanism, known as the Mhrt-related circuit, participates in chromatin remodelling and prevent myocardial hypertrophy (15). It has been revealed that cardiac apoptosis-related IncRNA affects abnormal mitochondrial fission and apoptosis, which may improve the
treatment of myocardial infarction (16). Furthermore, IncRNA non-coding RNA activated by DNA damage (NORAD), which is involved in the maintenance of genomic stability, has been identified (17). As its activation is strongly associated with a poor prognosis and survival in breast cancer and pancreatic cancer, and high expression of NORAD is observed in gastric cancer cells and tissues, NORAD is considered to be a potential oncogene (18-20). Additionally, NORAD may serve a role in the development of lung carcinoma by regulating the expression of transforming growth factor-β (TGF-β) and its associated phenotype (21). Furthermore, among the family of microRNAs (miRNAs/miRs), which are small non-coding RNAs ~22 nucleotides in length, hsa-miR-590-5p has been shown to downregulate the TGF-β signalling pathway, resulting in a decreased generation of cardiac cells (22,23). Previous studies have also revealed that hypoxia modulates the activities of abundantly expressed miRNAs by regulating proteins involved in post-transcriptional processes of miRNAs (24).

It has been shown that IncRNAs serve an important role in the regulation of angiogenesis to further affect cancer development or diseases (25). However, the function and mechanisms of the majority of IncRNAs have not been previously determined. It was hypothesized that NORAD may regulate endothelial cell properties via miR-590-3p, and modulated angiogenesis.

Cell culture and grouping treatment. HuVecs were purchased from the Shanghai Zhong Qiao Xin Zhou Biotecnology Company and cultured in M199 complete medium (Gibco; Thermo Fisher Scientific, Inc.) supplemented with 20% FBS (Sigma‑Aldrich; Merck KGaA) at 37˚C with 5% CO₂. HuVecs or transfected HuVecs were plated in 96-well plates with 5 replicates per condition. Following incubation at 37°C with 5% CO₂ for 24 h, cells underwent normoxia or hypoxia treatment and were cultured for 12 or 24 h, respectively. Subsequently, 10 µl CCK-8 reagent (Sigma-Aldrich; Merck KGaA) was added and incubated at 37°C for 1 h according to the manufacturer's instructions. Absorbance was measured at 450 nm using a microplate reader (BioTek Instruments, Inc.).

Transwell migration assay. Transwell chambers with 8.0 µm pore polycarbonate membrane inserts (Corning, Inc.) were placed in 24-well-plates. A total of 4x10³ normal or transfected HuVECs were suspended in serum-free M199 medium and seeded into the upper chamber, and 800 µl medium supplemented with 20% FBS was added to the lower chamber. Cells were cultured under normoxic or hypoxic conditions at 37°C for 24 h and washed twice with PBS. Cells that had migrated through the filter were fixed with 4% paraformaldehyde (Sigma-Aldrich Chemical Reagent Co., Ltd.) for 20 min and stained using 0.1% crystal violet (Amresco, LLC) for 5 min at room temperature. The stained cells were visualized using an inverted phase contrast microscope (magnification, x200) (Olympus Corporation), and the number of cells in 5 randomly chosen fields was counted.

Cell Counting Kit-8 (CCK-8) assay. A total of 3x10³ cells/well were transfected HuVECs were plated in 96-well plates with 5 replicates per condition. Following incubation at 37°C with 5% CO₂ for 24 h, cells underwent normoxia or hypoxia treatment and were cultured for 12 or 24 h, respectively. Subsequently, 10 µl CCK-8 reagent (Sigma-Aldrich; Merck KGaA) was added and incubated at 37°C for 1 h according to the manufacturer's instructions. Absorbance was measured at 450 nm using a microplate reader (BioTek Instruments, Inc.).

Materials and methods

Cell culture and grouping treatment. HuVECs were purchased from the Shanghai Zhong Qiao Xin Zhou Biotecnology Company and cultured in M199 complete medium (Gibco; Thermo Fisher Scientific, Inc.) supplemented with 20% FBS (Sigma-Aldrich; Merck KGaA) at 37°C with 5% CO₂. When cells reached ~70% confluency within 3 passages, cells were divided into two groups (normoxia and hypoxia) for hypoxia treatment. In the present study, the effects of NORAD and miR-590-3p on angiogenesis in human umbilical vein endothelial cells (HUVECs), and whether miR-590-3p affected NORAD-mediated regulation of endothelial cell activities were investigated.

Transfection. Specific short hairpin (sh)RNA against IncRNA NORAD (shNORAD) and control shRNA (shNC1) were obtained from Wanleibio Co., Ltd. miR-590-3p mimics (5'-UAA UUUUAUGUAUGCUAGU-3'), miR-590-3p inhibitor (5'-ACUAGCUAUCAUAUAAUA-3') and scrambled control (mimics-NC and inhibitor-NC) were purchased from JTS Scientific Co., Ltd. The overall length of NORAD was 5,378 bp, and the overexpression plasmid pcdNA3.1+ (Clontech Laboratories, Inc.) containing the first 2,000 bp of sequence NORAD (NORAD1,200 OE) was established. This sequence was used to ensure the potential binding sequence with miR-590-3p was included, and to avoid the binding site of shNORAD. HUVECs were transfected with 100 pmol of shNORAD (shNC1), miR-590-3p mimics (mimics-NC), NORAD1,200 OE plasmid (empty plasmid/vector) or shNORAD combined with miR-590-3p inhibitor (inhibitor-NC) using Lipofectamine® 2000 (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer protocol. Then, 48 h after transfection, cells were underwent hypoxia treatment as described above.
Reverse transcription-quantitative (RT-q) PCR. Total RNA was extracted from HUVECs and tissues using TRIpure reagent (BioTeke Corporation), and RT into cDNA using Super M-MLV reverse transcriptase (BioTeke Corporation), dNTPs, RNase inhibitor (BioTeke Corporation), 5X reaction buffer, and primers (RT primer for miRNA, Oligo(dT)15 for other genes) at 42°C for 30 min and 70°C for 10 min (miRNA) or 42°C for 50 min and 80°C for 10 min (other genes). Subsequently, qPCR was performed using diluted 2X Power Taq PCR MasterMix (BioTeke Corporation) with SYBR-Green (Sigma-Aldrich; Merck KGaA). The thermocycling conditions for miRNA were: 94°C for 4 min, 40 cycles of 94°C for 15 sec, 60°C for 20 sec and 72°C for 15 sec; for other genes they were: 94°C for 5 min, 40 cycles of 94°C for 15 sec, 60°C for 20 sec and 72°C for 30 sec. The expression of hsa-miR-590-3p was normalized to 5S, and the rest of the genes was normalized to β-actin. Primers were purchased from GenScript and the sequences were: hsa-miR-590-3p/mmu-miR-590-3p forward, 5'-Taa TTT TATGTATAAGCTAGT-3'; hsa-miR-590-3p/mmu-miR-590-3p reverse, 5'-GCAGGGTCCAGGATTCT-3'; S5 forward, 5'-GAT CTCGGAGACGTAAACGAG-3'; S5 reverse, 5'-TGGTGCAAGG GCAGGAGGTAT-3'; mmu-5S forward, 5'-CTAAAGATTTCC GTGGAGAG-3'; mmu-5S reverse, 5'-TGGTGCAAGGGTCC AGGAGGT-3'; IncRNA MIRAD forward, 5'-GGGAATCTG CTTAAGACT-3'; IncRNA MIRAD reverse, 5'-CCACACACCC AATTGATATG-3'; mmu-IncRNA NORDAD forward, 5'-GAT TGGCAGCCAGGGTAT-3'; mmu-IncRNA NORDAD reverse, 5'-CTGAAACAAACGGAGCA-3'; HIF-1α forward, 5'-GAA ACCTCTTGATGCTTGTTG-3'; HIF-1α reverse, 5'-CAACT GAGTTAATCCCATG-3'; VEGFA forward, 5'-TCACCAAGG CGACACATAG-3'; VEGFA reverse, 5'-GGGACACAAACGT ACACCGT-3'; FGFl forward, 5'-GAGCGACAGCACATCA G-3'; FGFl reverse, 5'-TCTCTTCACGCTTCCAGC-3'; FGFl forward, 5'-AGAACGAACGCCCCTCAT-3'; FGFl reverse, 5'-AAAGAAXACACTCATCCGTA-3'; β-actin forward, 5'-GGCCACCCAGCAATGAAA-3'; β-actin reverse, 5'-TACGACATTGTCGCTTG-3'; mmu-β-actin forward, 5'-TCG TGCCCATCTACGAGGTAT-3'; and mmu-β-actin reverse, 5'-TTTGATGTCACGCACTTTCC-3'. Fold changes of gene expressions were calculated using the 2^ΔΔCt method (27). Western blotting. Cell Lysis Buffer for Western or IP (Beyotime Institute of Biotechnology) was used to extract the total protein. Protein concentration was determined using a bicinchoninic acid protein assay kit (Beyotime Institute of Biotechnology). A protein of 20–40 μg was loaded on an 8–12% SDS gel and resolved using SDS-PAGE. The resolved proteins were transferred to PVDF membranes (EMD Millipore) and were blocked in 5% non-fat milk (Yili Group) at room temperature for 1 h. Membranes were incubated with primary antibodies against HIF-1α (AF1009, Affinity Biosciences) and β-actin (sc-47778; Santa Cruz Biotechnology, Inc.) both at 1:1,000 at 4°C overnight. Incubation of membranes with horseradish peroxidase-conjugated goat anti-rabbit (A0208, 1:5,000; Beyotime) or anti-mouse (A0216, 1:5,000; Beyotime) secondary antibody was performed at 37°C for 45 min. An ECL reagent (Beyotime Institute of Biotechnology) was used to visualize the signals, and densitometry analysis was performed using Gel-Pro-Analyzer software (version 4; Media Cybernetics, Inc.).

Animal experiment. A total of 20 male C57 mice (8 weeks old, 22±2 g; Beijing HFK Bioscience Co., Ltd.) were used to establish a myocardial infarction (MI) model via left anterior descending (LAD) ligation surgery. Prior to surgery, all mice were maintained in a 12 h light/dark cycle with constant temperature of 22±1°C and 45–55% humidity, with free access to food and water. After week, mice were immobilized and anesthetized. Then, endotracheal intubation and ventilation were performed following incision of the neck trachea, followed by left thoracotomy between the third and fourth intercostal space. Blood flow in the anterior descending branch was blocked using a sterile 8/0 suture ligation. Mice in the control group (n=6) underwent the same operation, but without ligation. After 24 h following suture, the left ventricle tissues were removed for analysis. Each group contained six mice. Animal experiments were performed in accordance with the principles described in the Guide for The Care and Use of Laboratory Animals (28), and were approved by The Ethics Committee of The First Hospital of China Medical University. Statistical analysis. Data were presented as the mean ± SD of 3 repeats for the in vitro experiments, and for the animal experiments there were 6 animals per group. GraphPad Prism version 8 (GraphPad Prism Software, Inc.) was used to perform the statistical analysis. Unpaired t-tests were used to compare two independent groups, and differences between ≥3 groups were compared using the one-way ANOVA combined with Tukey’s multiple comparisons post hoc test. P<0.05 was considered to indicate a statistically significant difference.

Results. HUVEC tube-formation ability is increased under hypoxic conditions. To determine the effects of non-coding RNAs on hypoxia exposed HUVECs, experiments were performed to assess the activities of HUVECs under hypoxic conditions. mRNA expression levels of HIF-1α and its downstream target VEGFA were determined 24 h after hypoxia culturing using RT-qPCR, and it was identified that both had increased expression levels following hypoxia treatment (Fig. 1A). Western blotting results also demonstrated increased protein expression of HIF-1α under hypoxic conditions (Fig. 1B). Thus, these results suggested that hypoxia treatment was effective. Furthermore, the CCK-8 assay results indicated that, compared with cells in the normoxic condition, cell viability was decreased.
of HUVECs was increased after 24 h of hypoxia exposure (Fig. 1C). Moreover, Transwell assays were performed to assess the migratory ability of cells following hypoxia for 24 h, and it was identified that hypoxia significantly increased migration (Fig. 1D). Furthermore, results from the tube formation assay in the hypoxia-exposed HUVECs demonstrated that tube count and length were increased compared with normoxic cells (Fig. 1E). Therefore, the present results suggested that short-term hypoxia exposure caused increased tube-formation in HUVECs.

IncRNA NORAD expression is increased and miR-590-3p expression is decreased in hypoxic HUVECs in vitro and MI left ventricular tissues in vivo. The relative mRNA expression levels of IncRNA NORAD and miR-590-3p in HUVECs under hypoxia were assessed using RT-qPCR. It was demonstrated that NORAD expression was increased and miR-590-3p expression was decreased in HUVECs exposed to hypoxia for 24 h (Fig. 2A). In vivo, mice MI models were also used to assess the expression levels of these 2 factors in the left ventricle following MI modelling for 24 h, and similar results were observed as those identified in vitro (Fig 2B).

Thus, it was identified that hypoxia increased expression of IncRNA NORAD, and decreased expression of miR-590-3p, in HUVECs in vitro and in left ventricular tissues in vivo.

IncRNA NORAD knockdown inhibits tube-formation ability of HUVECs under hypoxia. To investigate how IncRNA NORAD affected the HUVECs under hypoxia, cells were transfected with shNORAD and shNC1. RT-qPCR was performed to detect the relative expression of NORAD in transfected HUVECs, and it was revealed that shNORAD effectively knocked down the expression of IncRNA NORAD in HUVECs (Fig. 3A). Moreover, hypoxic treatment for 24 h in shNORAD transfected HUVECs also demonstrated that NORAD expression was significantly knocked down. It was also identified that NORAD downregulation caused an increase in miR-590-3p expression under hypoxia (Fig. 3B). Cell viability of transfected HUVECs was assessed using a CCK-8 assay. In contrast to hypoxia-exposed shNC1 cells, knockdown of NORAD resulted in decreased viability of shNORAD transfected HUVECs (Fig. 3C). Furthermore, the Transwell migration assays showed that HUVECs transfected with shNORAD exhibited a lower migratory
capacity compared with cells transfected with shNC1 under hypoxia (Fig. 3D). NORAD knockdown also decreased tube formation ability of HUVECs exposed to hypoxia compared with the NC cells (Fig. 3E). Therefore, the present results suggested that lncRNA NORAD knockdown inhibited the angiogenesis-associated abilities of HUVECs under hypoxia, indicating decreased blood vessel formation.

**lncRNA NORAD overexpression promotes tube-formation in HUVECs under hypoxia.** Cells were transfected with the NORAD_{1,2000} OE plasmid to study the effects of NORAD overexpression on HUVECs under hypoxia. Under normoxic conditions, the transfection of NORAD_{1,2000} OE plasmid in HUVECs was highly efficient (Fig. 4A). Moreover, the relative expression of NORAD was significantly upregulated, and the expression of miR-590-3p was decreased in the NORAD_{1,2000} OE transfected cells under hypoxia (Fig. 4B). Cell viability, migratory capacity and tube formation, including tube count and length, were assessed in NORAD overexpressed cells under hypoxia, and were identified to be increased (Fig. 4C-E). Collectively, the present results indicated that NORAD overexpression in HUVECs reversed the reduced tube-formation ability caused by NORAD knockdown under hypoxia.

**Overexpression of miR-590-3p suppresses tube formation in HUVECs under hypoxia.** To determine the role of miR-590-3p in the vasculogenic ability of HUVECs under hypoxia, cells were transfected with miR-590-3p mimics and mimics-NC. The RT-qPCR results indicated that miR-590-3p was efficiently overexpressed when cells were transfected with miR-590-3p mimics, under both normoxic and hypoxic conditions (Fig. 5A). Furthermore, overexpression of miR-590-3p resulted in similar effects on cell viability, cell migration and tube formation ability as knockdown of NORAD under hypoxia (Fig. 5B-D). In the dual luciferase assay, 293T cells co-transfected with NORAD-wild-type and miR-590-3p mimics exhibited significantly decreased luciferase activity compared with cells transfected with mutant NORAD and miR-590-3p mimics (Fig. 5E), which suggested that NORAD could bind with miR-590-3p directly. Thus, it was hypothesized that miR-590-3p also affected HUVEC tube-forming ability under hypoxia, which may be associated with NORAD targeting.

**lncRNA NORAD affects angiogenic characteristics of HUVECs under hypoxia via miR-590-3p.** To investigate whether the effects of NORAD on HUVEC tube-formation ability were regulated by miR-590-3p under hypoxia, HUVECs were co-transfected with shNORAD and miR-590-3p inhibitor. The transfection efficiency of the miR-590-3p inhibitor under normoxic conditions was assessed using RT-qPCR, which identified that the expression of miR-590-3p was significantly downregulated (Fig. 6A). Subsequently, the relative expression levels of lncRNA NORAD and miR-590-3p were assessed using RT-qPCR following co-transfection. Cells co-transfected with shNORAD and miR-590-3p inhibitor exhibited decreased miR-590-3p expression levels, but increased NORAD expression, compared with cells transfected with shNORAD and inhibitor-NC (Fig. 6B). Therefore, it was
speculated that miR-590-3p negatively regulated NORAD, and similar results were observed under hypoxia (Fig. 6C). Cell viability in the transfected hypoxia-exposed HUVECs, in which NORAD and miR-590-3p expression levels were downregulated, was significantly increased compared with cells transfected with shNORAD and inhibitor-NC (Fig. 6D). Furthermore, downregulation of NORAD and miR-590-3p in the HUVECs also increased migration and tube formation under hypoxia compared with the control cells (Fig. 6E and F). Thus, downregulation of both NORAD and miR-590-3p in HUVECs reversed the decrease in angiogenic activity caused by knockdown of NORAD alone, which resulted in enhanced angiogenic properties under hypoxic conditions. Collectively, the present results suggested that miR-590-3p lay downstream of the regulatory effects of IncRNA NORAD on angiogenic activity in hypoxia-exposed HUVECs.

**Downstream targets of the IncRNA NORAD/miR-590-3p axis in HUVECs under hypoxia.** In total, 3 possible regulated genes were proposed, VEGFA, FGF1 and FGF2, all of which are closely associated with angiogenesis and cell proliferation (5,8). After 24 h of hypoxia treatment, it was demonstrated that the mRNA expression levels of all 3 genes were significantly decreased in NORAD knockdown HUVECs, but significantly increased in cells where NORAD and miR-590-3p were both downregulated (Fig. 7A). Moreover, ELISAs were performed to assess the changes in the levels of all 3 proteins, and it was identified that when NORAD was downregulated, the concentrations of VEGFA, FGF1 and FGF2 were decreased under hypoxia. However, when miR-590-3p was knocked down, the expression levels of these genes were increased significantly under the same condition (Fig. 7B). Binding of miR-590-3p to these genes was determined using a dual
luciferase assay, which identified that cells co-transfected with the 3'UTR of Vegfa, FGF1 or FGF2, and miR-590-3p mimics, exhibited lower luciferase activity compared with cells transfected with their mutated 3'UTR and miR-590-3p mimics (Fig. 7c). Therefore, the present results suggested that a lncRNA norad/miR-590-3p axis regulated the activity of the angiogenic indicators Vegfa, FGF1 and FGF2, and that these genes were direct downstream targets of miR-590-3p.

Discussion

The aim of the present study was to determine whether the lncRNA NORAD served a role in angiogenesis induced by hypoxia, and the involvement of miR-590-3p in this process. lncRNA NORAD is activated by DNA damage and maintains genomic stability via isolating Pumilio proteins, which bind with mRNAs to suppress their stability and translation activity (17). Thus, NORAD is crucial for cell division and growth during cell development (29). In the present study, in short term hypoxia (24 h), downregulation of NORAD in the HUVECs resulted in decreased angiogenic properties, including cell viability, cell migration and tube formation. Moreover, NORAD overexpression resulted in opposite effects, suggesting that NORAD regulates the angiogenic properties of HUVECs under hypoxic conditions. It was identified that the overexpression of miR-590-3p caused a similar decrease in tube formation in hypoxia-exposed HUVECs. The present results observed in cells overexpressing miR-590-3p were consistent with the effects of NORAD downregulation, and when NORAD was downregulated, miR-590-3p expression was increased both in vitro and in vivo. Several miRNAs, including miR-20, miR-130 family and miR-199, are involved in angiogenesis under hypoxic conditions by modulating the expression or activity of proteins, including HIF-1 and...
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Based on the present results, it was speculated that miR-590-3p may also be considered an additional miRNA involved in angiogenesis under hypoxic conditions.

The present results suggested that NORAD directly binds to miR-590-3p, and miR-590-3p directly targeted the 3’UTR of VEGFA, FGF1 and FGF2. In addition, knockdown of both NORAD and miR-590-3p could alleviate and reverse the decreased angiogenic activities and expression levels of proangiogenic indicators caused by knockdown of NORAD alone, leading to increased cell viability, migration, tube formation and expression of associated proangiogenic factors. Collectively, the present results indicated the presence of a regulatory pathway in which IncRNA NORAD binds to miR-590-3p to regulate angiogenesis in HUVECs, by affecting the expression levels of downstream proangiogenic proteins, such as VEGFA, FGF1 and FGF2, under hypoxic conditions. It is possible that other positive factors involved in the regulation of angiogenesis also serve a role in the NORAD/miR-590-3p

Figure 5. Overexpression of miR-590-3p suppresses tube formation in HUVECs under hypoxia. (A) HUVECs were transfected with miR-590-3p mimics and mimics-NC. Relative expression of miR-590-3p following transfection was assessed prior and subsequent to hypoxia treatment. ***P<0.001 vs. respective NC controls. (B) Cell viability, (C) cell migration (magnification, x200) and (D) tube formation (magnification 100x) were examined in cells transfected with miR-590-3p mimics under hypoxia. *P<0.05, **P<0.01 and ***P<0.001 vs. Hypoxia + mimics-nc group. (E) Relative luciferase activity was measured 48 h after co-transfection of miR-590-3p mimics with norad-wt or norad-mut in 293T cells. The binding sequences of the wt and mut NORAD are shown. **P<0.01 vs. NORAD-mut + miR-590-3p mimics group. IncRNA, long non-coding RNA; IncRNA NORAD, IncRNA non-coding RNA activated by DNA damage; miR, microRNA; NC, negative control; mut, mutant; wt, wild-type; OD, optical density.

VEGF (30).
axis under hypoxia, but this was not investigated in the present study. TGF-β has been shown to indirectly regulate angiogenesis and also be involved in a miR-590 regulated pathway in cardiac diseases and NORAD-associated functions in lung cancer (21-23,31,32). Thus, TGF-β may also affect this NORAD/miR-590-3p regulated hypoxia-induced angiogenesis, but further studies are required to assess this.

The mechanisms regulating angiogenesis are complicated and involve not only the regulation of angiogenic indicators and chemokines, but also the participation of endothelial cells, macrophages or pericytes during various functions of organs under different conditions (5,12,33). Hypoxia-induced diseases are primarily divided into two types: Hypoxia-related ischemic diseases; and angiogenesis-caused cancer types (5). Hypoxia/HIF-1-dependent injuries or angiogenesis are highly associated with ischemic diseases (5). A previous study showed that ischemia/hypoxia-induced angiogenesis briefly improves the blood supply to an ischemic heart, and that this phenomenon is associated with innate immune system activation and macrophage participation in the growing vessels, rather than...
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However, the detailed mechanisms of this effect have not been determined. Furthermore, a study investigating the angiogenic response under hypoxia in vivo showed that hypoxia-induced angiogenesis of endothelial cells was modulated by a complex network of interactors, and included both the regulation of angiogenic factors such as VEGF, and the regulation of the expression of VEGFRs and VEGF inhibitors (33). In the present study, the regulatory mechanisms of hypoxia-induced angiogenesis in endothelial cells via the norad/mir-590-3p axis in vitro were assessed, thus identifying a novel pathway of regulation of angiogenesis. However, in vivo studies are required to confirm the relevance of this axis.

The present study identified the need to use different strategies for treating hypoxia-induced diseases based on the specifics of each case. Therapies targeting a single angiogenic factor, such as VEGF, have been shown to be insufficient, which highlights the requirement for the use of a combination of other drugs to maintain or regulate angiogenesis (1,5,33). Therefore, the hubs of hypoxia-induced angiogenesis should be investigated further. The expression levels of several miRNAs have been shown to be age-associated and cell type-specific (30,34), which needs to be considered clinically during angiogenic therapy. While there are few previous studies on IncRNAs in hypoxia-induced angiogenesis, the present results suggested that IncRNA NORAD could be involved in angiogenic activity of endothelial cells under hypoxic conditions, which may be a potential target for the treatment of associated diseases, such as ischemic heart disease.

In conclusion, the present results indicated that IncRNA NORAD was upregulated, and miR-590-3p was downregulated, in hypoxic HuVECs in vitro and MI left ventricular tissues in vivo. Furthermore, downregulation of IncRNA NORAD in HuVECs decreased cell viability and angiogenic activity, and overexpression of miR-590-3p resulted in similar effects. It was identified that knockdown of both NORAD and miR-590-3p reversed this decrease, and resulted in enhanced angiogenic activity. NORAD was also identified to target miR-590-3p and negatively regulate its expression. It was demonstrated that miR-590-3p further targeted the angiogenic factors VEGF, FGF1 and FGF2. Therefore, the present results suggested that an IncRNA NORAD/miR-590-3p axis may be involved in the regulation of angiogenesis in HuVECs under hypoxia, which highlights potential targets for treating hypoxia-induced angiogenic diseases.

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Availability of data and materials
The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Authors' contributions
XZ designed the study, performed the experiments, analysed and interpreted the data, and drafted the manuscript. XWe
and XWa performed the experiments and collected the data. GQ designed and supervised the study, interpreted the data and critically revised the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Animal experiments were performed in accordance with the principles described in the Guide for The Care and Use of Laboratory Animals, and were approved by The Ethics Committee of The First Hospital of China Medical University.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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