Original Research Article

In vitro Antimicrobial Activity of Bangladeshi Artocarpus lakoocha Roxb Fruits

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A B S T R A C T

Microorganisms play a crucial role in living organisms. Some microorganisms are beneficial for us some are pathogenic. The aim of the study is to evaluate the activity of the fruits of Artocarpus lakoocha Roxb. against certain pathogenic microbial strains. Petroleum ether, chloroform and methanol extract of these fruits were used by disk diffusion method on Escherichia coli, Bacillus subtilis, Staphylococcus aureus, Salmonella typhi, Bacillus cereus, Salmonella paratyphi and two fungal strains. All the fruit extracts showed the antimicrobial activity against all the microorganisms. Among the three extracts, methanolic fruit extracts exhibited significant activity against the microorganisms and fruit extracts of petroleum ether showed the least activity against the certain microorganisms. The fruit extracts showed antibacterial activity with a zone of inhibition of 4 to 20 mm. The methanolic fruit extract of Artocarpus lakoocha Roxb showed antibacterial activity with a zone of inhibition of 17 to 20 mm. The study recommended that the fruits of Artocarpus lakoocha Roxb. exhibited significant antimicrobial activity.

Keywords
Artocarpus lakoocha, Antibacterial activity, Antifungal activity, Zone of inhibition

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Introduction

For a long period of time, medicinal plants are used as a rich source in traditional and sanctioned medicine (Prashant et al., 2008). As medicinal plants have a potential source of potent medicinal value, convenience to users, economic viability and low toxicity so nowadays these plants are increasing attraction for producing drugs (Calixto, 2000). It is proved that after using long term plant-derived drug do not have any side effects whereas synthetic drugs have diverse side effects (Afolayan, 2003). Due to the continuous exposure of drug-resistant organisms the efficacy of current antimicrobial agents has been reduced. Although plant-based antimicrobials have enormous therapeutic potential and usefulness in the treatment of infectious diseases these antimicrobials represent a vast unexploited source of medicine (Aliero et al., 2006, Khare, 2007).

Artocarpus lakoocha (Moraceae) is a widely used medicinal plant found in Bangladesh,
India, Thailand and Southeast Asia. All parts of this medicinal plant have diverse medicinal value (Charoenlarp et al., 2007). Generally, the fruits are eaten fresh. The fruits comprise vitamin C and β carotene. Also, the fruits of *Artocarpus lakoocha* have antioxidative value. For the maintenance of normal health and protection from the cardiovascular disease, the antioxidative properties of fruits play an important role. The oxidative properties also combat with cancer (Jahan et al., 2011, Pandey et al., 2009). The edible fruit pulp is used as a restorative for the liver. Pickles and chutney are also made by raw fruits. The brown powder called Puag-Haad has been used as a traditional anthelmintic drug for treatment of tapeworm infestation in Thailand (Salguero, 2003; Puntumchai et al., 2004). The tree bark is used to treat skin lesion and the powder form of bark is applied to cure wound (Tomar et al., 2015). The seed and bark also reduce the stomach and liver diseases. Although the ripe fruits are sweet and used as a liver tonic the unripe fruit is sour and also cause loss of appetite, blood infelicity (Piyush et al., 2014). β-amyrin acetate and lupeol acetate are present in fruits having a potential anti hyperglycemic and hypolipidemic effects that could be used as a lead compound for the production of effective in diabetes and atherosclerosis medicine (Perry, 1980). The roots of this plant showed antibacterial and cytotoxic activity (Likhitwitayuwid et al., 2005). The heartwood of *A. lakoocha* has exhibited moderate activity against herpes simplex virus and HIV (Piyush et al., 2014). The fruit pericarp of *A. lakoocha* also has antibacterial, antioxidant, anthelmintic and insecticidal activity (Sein et al., 2009). There are no reports on the antimicrobial activity of *A. lakoocha* fruits that are grown in Bangladesh.

The main objective of this study is to determine the antimicrobial potential of fruit extracts of *A. lakoocha* that are grown in Chittagong, Bangladesh using disk diffusion assay for bacteria and fungi.

**Materials and Methods**

**Collection of plant materials**

The fresh fruits of *Artocarpus lakoocha* Roxb were collected from University of Chittagong during April 2018. Collected fruits were washed thoroughly in running tap water than in distilled water. After that wiped with a paper towel and weighed whole after removing the seeds. The edible portions were cut into small pieces and shade dried at room temperature (25°C±1) for one week in the open air. After dry the fruits were pulverized into powder form with a grinder. Then the powder was stored in airtight closed bottles and kept in a refrigerator at 4°C before used for analysis.

**Microorganisms**

Reference bacteria and fungi strains were executed from Biochemistry and Molecular Biology Department, University of Chittagong, and they included *Escherichia coli*, *Bacillus subtilis*, *Staphylococcus aureus*, *Salmonella typhi*, *Bacillus cereus*, *Salmonella paratyphi* and *Candida albicans*, *Candida krusei*.

**Extract preparation**

10 g of air-dried powder was added to 100 ml of petroleum ether, chloroform and methanol (Sigma Chemicals Co., St. Louis, MO, USA) separately in a conical flask, plugged with cotton wool and then kept for 7 days at room temperature with occasional stirring. The supernatant was collected by filtering with Whatman no1 filter paper and the solvent was slowly evaporated under reduced pressure below 50°C through an evaporator (RE200; Bibby Starling, Staffordshire, England). The
concentrated extract was collected in a Petri dish and allowed to air-dry for the complete evaporation of solvents in the absence of sunlight and stored at 4°C in airtight bottles.

**Reference antibiotics**

Kanamycin and Nystatin were used as positive control for bacteria and fungi respectively.

**Study of antibacterial activity**

**Media preparation**

Standard nutrient agar (Difco) media was maintained for the growth (37 °C and pH) of the bacterial strains. The bacteria were sub-cultured overnight in nutrient agar broth and it was adjusted to get turbidity comparable to 0.5 McFarland standard when requisite. For the maintenance of cultures test tube slants of nutrient agar medium were prepared. With the help of sterilized needles, small amount of the collected microorganisms were transferred to the test tubes. Under laboratory condition, the inoculated slants were inoculated at room temperature.

**Disk diffusion assay**

The antibacterial assay was performed using the paper disc diffusion method described by Navarro et al., 1998; Swain et al., 2008. Distinct concentrations of the fruit extracts (0.075, 0.10, 0.20, 0.40, 0.60, and 1.00mg/μl) were prepared with petroleum ether, chloroform and methanol. Nutrient agar was inoculated with a microbial cell suspension (200 μl in 20 ml of medium) and poured into sterile Petri dishes and rotated clockwise and counterclockwise for a few times to be seeded uniformly. Dried and sterilized filter paper discs 6 mm in diameter were treated with 20 μl of each extract concentration and using micropipette and placed on the inoculated agar surface. Standard 6 mm antibiotic discs Kanamycin containing (30 μg/disc Oxoid, Hampshire, England) was used as positive control was placed on the agar surface with sterile forceps. Negative controls were made using paper discs treated with 20 μl of the solvents. After pre-incubation for 2 h in a low temperature (4-6°C) and allowed to diffuse the test materials (antimicrobial) from the disc to the surrounding medium by this time. Then the plates were incubated overnight at 37°C for 24 h to allow the maximum growth of the organisms.

**Study of antifungal activity**

**Fungal strains**

The in vitro antifungal activity of the *A. lakoocha* fruits were studied against two human pathogenic fungal strains *Candida albicans* (*C. albicans*) and *Candida krusei* (*C. krusei*).

**Determination of antifungal activity**

Antifungal effect of *A. lakoocha* fruits was determined by poisoned food technique (Grover et al., 1962, Mishra et al., 1992, Nene et al., 2002). Potato dextrose agar was used as a culture medium. For this 10% sample solution was taken with a sterilized pipette in a sterilized petri dish and then 15 ml of medium was poured into the petri dish to mix well and allowed to solidify. Inoculation was done at the centre of each plate with 5 mm of mycelium block for each fungus. The mycelium block was prepared with the help of cork-borer from the growing area of a five-day-old culture of the test fungi on potato dextrose agar. To get greater contact of the mycelium with the culture medium the blocks were placed at the centre of each petri dish in an inverted position. After 5 days of incubation, the diameters of fungal colonies were measured. The experiment was repeated
three times. Nystatin 30 μg/disc was used as positive control. Negative controls were made using paper discs treated with 20 μl of the solvents.

**Results and Discussion**

The present study demonstrated that all the extracts of *Artocarpus lakoocha* fruits showed antibacterial activity (Table 1) and antifungal activity (Table 2) against reference microorganisms. Petroleum ether and chloroform exhibited antibacterial activity at the concentration of 0.20, 0.40, 0.60, and 1.00mg/μl. But methanol extract showed antibacterial activity at the concentration of 0.10, 0.20, 0.40, 0.60, and 1.00mg/μl.

Distinct solvents reveal different activity against microorganisms. In our experiment, three different solvents were used which have different polarities. Among the three solvents, petroleum ether is non-polar, chloroform is more polar than petroleum ether and methanol is most polar than previous two solvents.

Petroleum ether extract of fruits of *Artocarpus lakoocha* Roxb showed the lowest activity compared to chloroform and methanol extracts. Among the three extracts, methanolic extract of the fruit exhibited strongest antimicrobial activity against the reference microorganisms which certainly point out that methanolic extract contains a higher concentration of active antimicrobial agents. Also, this could be due to the polarity nature to the active microbial compounds including alkaloids, carbohydrates, glycosides, saponin is present in these fruits.

Previous studies had also demonstrated that *Artocarpus lakoocha* Roxbis very rich in proteins and steroids (Dubey et al., 2015). The fruit extracts showed antibacterial activity with a zone of inhibition of 4 to 20 mm. The methanolic fruit extract of *Artocarpus lakoocha* Roxb. paraded a broad-spectrum antibacterial activity with a zone of inhibition of 17 to 20 mm. Different reports showed that methanolic extracts exhibited broader spectrum antimicrobial activity (Hossain et al., 2014, Jeong et al., 2009, Patrick et al., 2011). Our result indicated the similar result. On the other hand, chloroform extract exhibited a broad-spectrum antibacterial activity with a zone of inhibition of 17 to 20 mm against some bacterial species and petroleum ether extract showed comparatively narrow spectrum antibacterial activity with a zone of inhibition of 11 to 15 mm against reference bacterial species. The activity of the standard drug, Kanamycin was found higher than these fruit extract concentrations showing 24-30 mm in diameter against all the tested bacterial strains.

Moreover, the extracts of the fruit were active against the tested fungal species. Methanolic fruit extract had a very promising inhibitory effect on fungal strains compared to the reference antifungal drug nystatin same as antibacterial activity.

Comparisons of the three extracts it is clear that the methanolic fruit extract has significant activity, chloroform extract has moderate activity and petroleum ether extract has the least activity against *Candida albicans* and *Candida kruisei*. From our experiment, it is clear that more polar solvents have significant antimicrobial activity than non-polar solvents which have similarities with other reports (Das et al., 2012; Mwambete et al., 2009). It was reported that steroids are a major component that acts as an antifungal secondary metabolite (Onwuliri et al., 2005). As the fruits have steroids (Dubey et al., 2015) so these findings suggest that the antifungal effect of *Artocarpus lakoocha* extract is probably due to the individual or interactive effect of the secondary metabolites present in the extract.
**Table 1** Antibacterial activity of fruits of *Artocarpus lakoocha* Roxb

| Microorganisms          | Different extracts of fruits of *Artocarpus lakoocha* Roxb (Zone of inhibition) | Petroleum ether(mg/µl) | Chloroform(mg/µl) | Methanol(mg/µl) |
|-------------------------|---------------------------------------------------------------------------------|------------------------|-------------------|-----------------|
|                         |                                                                                 | 0.075 0.10 0.20 0.40 0.60 1.00 | 0.075 0.10 0.20 0.40 0.60 1.00 | 0.075 0.10 0.20 0.40 0.60 1.00 |
| *Escherichia coli*      |                                                                                 | - - + + ++ ++ - - + + ++ ++ +++ - + ++ +++ +++ +++ |
| *Bacillus subtilis,*    |                                                                                 | - - + + ++ - - - - + + ++ ++ +++ - + ++ ++ +++ +++ |
| *Staphylococcus aureus,*|                                                                                 | - - + + + - - - - + + ++ ++ +++ - + ++ +++ +++ +++ |
| *Salmonella typhi,*     |                                                                                 | - - + + ++ ++ - - + + ++ +++ - + + ++ ++ +++ +++ |
| *Bacillus cereus,*      |                                                                                 | - - + + ++ - - - - + + ++ ++ +++ - + ++ +++ +++ +++ |
| *Salmonella paratyphi*  |                                                                                 | - - + + ++ - - + + ++ +++ - + + ++ ++ +++ +++ |
| Positive control        |                                                                                 |                       | +++              | +++             |
| *Kanamycin*             |                                                                                 |                       | +++              | +++             |

Experiments were done in triplicate. Diameter of zone of inhibition: - < 4; + = 5-10; ++ = 11-15; +++ > 16.

**Table 2** Antifungal activity of fruits of *Artocarpus lakoocha* Roxb

| Microorganisms          | Different extracts of fruits of *Artocarpus lakoocha* Roxb | Petroleum ether(mg/µl) | Chloroform(mg/µl) | Methanol(mg/µl) |
|-------------------------|-------------------------------------------------------------|------------------------|-------------------|-----------------|
|                         |                                                              | 0.075 0.10 0.20 0.40 0.60 1.00 | 0.075 0.10 0.20 0.40 0.60 1.00 | 0.075 0.10 0.20 0.40 0.60 1.00 |
| *Candida albicans*      |                                                              | - - - - + + - - - - + + ++ - - + ++ ++ ++ +++ |
| *Candida krusei*        |                                                              | - - - - + + - - - - + ++ +++ - - + ++ +++ +++ |
| Positive control        |                                                              |                       | +++              | +++             |

Nystatin (30µg/disk) ++++++++  ++++++++  ++++++++  

Experiments were done in triplicate. Diameter of zone of inhibition: - < 4; + = 5-10; ++ = 11-15; +++ > 16.
In conclusion, this study represents the preliminary report on the antimicrobial activity of the fruit extracts of Artocarpus lakoocha Roxb. against both the bacterial and fungal strains. The results suggest that Artocarpus lakoocha fruits are very promising as an antimicrobial agent. Further studies are recommended that will involve various parts of the plant from distinct areas and select different fractions of crude extracts and also purify the most active antimicrobial components.

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