Chemosensor properties of 7-hydroxycoumarin substituted cyclotriphosphazenes∗

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Abstract: The newly synthesized cyclotriphosphazene cored coumarin chemosensors 5, 6, and 7 were successfully characterized by 1H NMR, 31P NMR, and MALDI-TOF mass spectrometry. Additionally, the photophysical and metal sensing properties of the targeted compounds were determined by fluorescence spectroscopy in the presence of various metals (Li+, Na+, K+, Cs+, Mg2+, Ba2+, Cr3+, Mn2+, Fe3+, Co2+, Al3+, Hg+, Cu2+, Zn2+, Ag+, and Cd2+). The fluorescence titration results showed that compounds 5, 6, and 7 could be employed as fluorescent chemosensors for Fe3+ ions with high sensitivity. The complex stoichiometry between final cyclotriphosphazene chemosensors and Fe3+ ions was also determined by Job’s plots.

Key words: Cyclotriphosphazene, coumarin, fluorescent chemosensor, aqueous media

1. Introduction
Fluorescence chemosensor design is the key topic in the determination of ions in biological processes. It is important to examine the fluorescence properties of the compounds to be synthesized as new sensors, their location in living organisms, and their usability for metal ion detection as fluorescent sensors [1–3]. Fe3+, an indispensable element for life, both provides oxygen-forming capacity and serves as a cofactor in many enzymatic reactions in the mitochondrial respiratory chain, and both deficiency and excess amounts of Fe3+ can cause various diseases [4]. Deficiency of Fe3+ ions causes anemia, diabetes, liver damage, Parkinson disease, and cancer. On the other hand, excessive intake of iron in humans is equally harmful and may cause dysfunctions of certain organs, such as the heart, pancreas, and liver [5]. Furthermore, high levels of Fe3+ ions may be toxic, promoting the oxidation of lipids, proteins, and other cellular components. For this reason, the detection of iron ions in biological media is very important [6]. Coumarin-based fluorescent chemosensors show less toxicity and are easily altered. Furthermore, the carbonyl group of coumarin can partake in coordination with metal ions if necessary. This is an ideal model for the design of chemosensors because the response will be fast and efficient when guests, e.g., protons and metal ions, are bound to the host probes. Recently, coumarins being are used as fluorescent chemosensors due to their high optical activities, high light stability, high quantum yield, wide Stokes shift, and low toxicity properties. These compounds are an important part of organic heterocyclic compounds used in sensor technology [4]. Coumarin compounds demonstrate perfect photophysical properties. For this reason, they are used in many applications for their anticoagulant [7], antitumor [8], photosensitizer

∗Dedicated to our supervisor Prof. Dr. Adem Kılıç on his retirement.
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The fluorescence characteristics of these compounds can be varied by substituting different groups at the 7-position of the coumarin skeleton. Because of its donor–acceptor behavior, it has very high potential to be used as a fluorescent chemosensor [16–18].

Phosphazenes are composed of repeating units of [–P=N–] in their structure. They also have a cyclic structure connecting two inorganic or organic side groups (R) to each phosphorus atom. Hexachlorocyclotriphosphazene is one of the most important members of the phosphazene family because different products are obtained as a result of various reactions on the six chlorine groups in the structure. Depending on the attached groups, they are suitable for the synthesis of different compounds for various applications [19,20]. During the past two decades, nucleophilic substitution reactions at phosphorus atoms of hexachlorocyclotriphosphazene have been extensively explored, leading to an enormous variety of materials with interesting properties such as anticancer [21] and antimicrobial agents [22] or fluorescence sensors [23]. Although there are many studies about coumarin, there are few studies about cyclotriphosphazenes bearing coumarin groups. Some of these were synthesis and characterization studies [24], as well as fluorescent chemosensor studies [25,26]. In the current study, new coumarin-based cyclotriphosphazenes (5–7) (Scheme), which are fluorescent chemosensors for Fe$^{3+}$ ions, have been successfully designed and prepared. The photophysical properties and metal-sensing behaviors of the new compounds (5–7) were investigated by UV-Vis electronic absorption and fluorescence spectroscopies.

Scheme. 7-Hydroxycoumarin substituted cyclotriphosphazenes (5–7).
2. Experimental

Materials and methods are given in the Supporting information.

2.1. Syntheses

7-Hydroxy-3-(4-methoxyphenyl)coumarin (1) [27], 7-hydroxy-3-(3,4-dimethoxyphenyl)coumarin (2) [28], and 7-hydroxy-3-(3,4,5-trimethoxyphenyl)coumarin (3) [29] were synthesized according to the procedures given in the literature.

2.1.1. Synthesis of compound 5

Hexachlorocyclotriphosphazene (4) (0.5 g, 1.43 mmol) was dissolved with tetrahydrofuran in a 100-mL three-necked round-bottomed flask. After that, NaH (0.35 g, 8.58 mmol) dissolved with 5 mL of THF was added to the reaction mixture. Then 7-hydroxy-3-(4-methoxyphenyl)coumarin (1) (2.29 g, 8.58 mmol) dissolved with 10 mL of THF was added drop-wise for about 20 min and the reaction was heated with stirring at 80 °C under an argon atmosphere for 4 days. After this time, the reaction mixture was filtered off with a G4 filter for removing formed NaCl salts. The solvent was completely removed by evaporation. Finally, the reaction mixture was purified by column chromatography using silica gel as column material and an n−hexane-THF (1:5) solvent mixture as an eluent. Yellowish pure product 5 was obtained (0.62 g, 0.35 mmol, 25%). Anal. Calc. for C_{96}H_{66}N_{3}O_{24}P_{3}, MALDI-TOF (m/z) calc. 1738.50, found: 1739.198 [M+H]+, 1762.071 [M+Na]+ (Figure 1a). 31P NMR (proton decoupled) (202 MHz, CDCl_{3}) δP = 7.92 (s, 3P) ppm (Figure 1b). 1H NMR (500 MHz; CDCl_{3}): δH = 7.66 (s, Ar-CH, 6H); 7.53 (d, Ar-CH, 12H, 3J_{H-H} = 8.38); 7.47 (d, Ar-CH, 6H, 3J_{H-H} = 8.41); 7.09 (d, Ar-CH, 6H, 3J_{H-H} = 8.41); 7.01 (s, Ar-CH, 6H); 6.85 (d, Ar-CH, 12H, 3J_{H-H} = 8.38); 3.84 (s, -OCH_{3}, 18H) ppm (Figure 2). FT-IR (cm\(^{-1}\)): 3100–2800 (C-H aromatic/aliphatic); 1728 (C=O); 1610, 1575 (C=C); 1247 (C-O); 1146, 1121 (P=N); 990 (P-O).

2.1.2. Synthesis of compound 6

Hexachlorocyclotriphosphazene (4) (0.5 g, 1.43 mmol) was dissolved with tetrahydrofuran in a 100-mL three-necked round-bottomed flask. After that, NaH (0.35 g, 8.58 mmol) dissolved with 5 mL of THF was added to the reaction mixture. Then 7-hydroxy-3-(3,4-dimethoxyphenyl)coumarin (2) (2.55 g, 8.58 mmol) dissolved with 10 mL of THF was added drop-wise for about 20 min and the reaction was heated with stirring at 80 °C under an argon atmosphere for 4 days. After this time, the reaction mixture was filtered off with a G4 filter for removing formed NaCl salts. The solvent was completely removed by evaporation. Finally, the reaction mixture was purified by column chromatography using silica gel as column material and an n−hexane-THF (1:5) solvent mixture as an eluent. Yellowish pure product 6 was obtained (0.62 g, 0.35 mmol, 25%). Anal. Calc. for C_{102}H_{78}N_{3}O_{30}P_{3}, MALDI-TOF (m/z) calc. 1918.66, found: 1919.099 [M+H]+ (Figure S1). 31P NMR (proton decoupled) (202 MHz, CDCl_{3}) δP = 7.99 (s, 3P) ppm (Figure S2). 1H NMR (500 MHz; CDCl_{3}): ppm: δH = 8.06 (s, Ar-CH, 6H); 7.56 (d, Ar-CH, 6H, 3J_{H-H} = 8.52); 7.28–7.24 (m, Ar-CH, 6H+6H); 6.98 (d, Ar-CH, 6H, 3J_{H-H} = 8.52); 6.79 (d, Ar-CH, 6H, 3J_{H-H} = 8.35); 6.73 (s, Ar-CH, 6H); 3.77 (s, -OCH_{3}, 18H); 3.76 (s, -OCH_{3}, 18H) ppm (Figure S3). FT-IR (cm\(^{-1}\)): 3080–2850 (C-H aromatic/aliphatic); 1724 (C=O); 1610, 1575 (C=C), 1256 (C-O); 1144, 1119 (P=N); 997 (P-O).
2.1.3. Synthesis of compound 7

Hexachlorocyclotriphosphazene (4) (0.5 g, 1.43 mmol) was dissolved with tetrahydrofuran in a 100-mL three-necked round-bottomed flask. After that, NaH (0.35 g, 8.58 mmol) dissolved with 5 mL of THF was added to the reaction mixture. Then 7-hydroxy-3-(3,4,5-trimethoxyphenyl)coumarin (3) (2.81 g, 8.58 mmol) dissolved with 10 mL of THF was added drop-wise for about 20 min and the reaction was heated with stirring at 80 °C under an argon atmosphere for 4 days. After this time, the reaction mixture was filtered off with a G4 filter for removing formed NaCl salts. The solvent was completely removed by evaporation. Finally, the reaction mixture was purified by column chromatography using silica gel as column material and an n-hexane-THF (1:5) solvent mixture as an eluent. Then yellowish pure product 7 was obtained (0.63 g, 0.30 mmol, 21%). Anal. Calc. for C\textsubscript{108}H\textsubscript{90}N\textsubscript{3}O\textsubscript{36}P\textsubscript{3}, MALDI-TOF (m/z) calc. 2097.45, found: 2097.05 [M]+, 2117.05 [M+Na]+ (Figure S4).

\(^{31}\)P NMR (proton decoupled) (202 MHz, CDCl\textsubscript{3}) \(\delta\textsubscript{P} = 7.82\) (s, 3P) ppm (Figure S5).

\(^{1}\)H NMR (500 MHz; CDCl\textsubscript{3}): ppm: \(\delta\textsubscript{H} = 7.78\) (s, Ar-CH, 6H); 7.58 (d, Ar-CH, 6H, \(^{3}\)J\textsubscript{H−H} = 8.51); 7.17 (d, Ar-CH, 6H, \(^{3}\)J\textsubscript{H−H} = 8.51); 6.97 (s, Ar-CH, 6H); 6.88 (s, Ar-CH, 12H); 3.89 (s, -OCH\textsubscript{3}, 18H); 3.84 (s, -OCH\textsubscript{3}, 36H) ppm (Figure S6).

FT-IR (cm\textsuperscript{-1}): 3090-2845 (C-H aromatic/aliphatic); 1724 (C=O); 1610, 1514 (C=C); 1256 (C-O); 1119, 1122 (P=N); 981 (P-O).

3. Results and discussion

3.1. Syntheses and characterizations

7-Hydroxycoumarin derivatives substituted cyclotriphosphazenes (5-7) were synthesized and their fluorescence chemosensor properties were also reported. 7-Hydroxy-3-(4-methoxyphenyl)coumarin (1), 7-hydroxy-3-(3,4-
dimethoxyphenyl)coumarin (2), and 7-hydroxy-3-(3,4,5-trimethoxyphenyl)coumarin (3) were synthesized according to the procedures given in the literature [27–29]. Compounds 1–3 and hexachlorocyclotriphosphazene (4) were separately reacted in a 6:1 molar ratio in the presence of NaH as a base in THF under an argon atmosphere at room temperature (Scheme). All the products (5–7) were purified by column chromatography and characterized by MALDI-TOF MS, FT-IR, $^1$H NMR, $^{31}$P NMR, and fluorescence spectroscopy. The MS, $^1$H NMR, and $^{31}$P NMR results for each new compound were compatible with the data given in Section 2. All of the proton decoupled $^{31}$P NMR spectra of compounds 5–7 showed a sharp single peak due to the equivalent chemical environment of all the phosphorus nuclei, suggesting an A$_3$ spin system. The $^{31}$P NMR spectrum of compound 5 is depicted as an example in Figure 1b.

The $^1$H NMR data also confirmed the structures of compounds 5–7. The aromatic protons for all compounds were observed between 8.06 and 6.73 ppm and some of them were distinguishable from each other. The NMR signals of the -OCH$_3$ groups were observed as a singlet or two singlets between 3.76 and 3.89 ppm. The $^1$H NMR spectrum of compound 5 is given as an example in Figure 2. FT-IR frequencies of various diagnostic bands for compounds 5–7 are given in Section 2. These spectra showed characteristic stretching bands for aromatic/aliphatic -CH vibrations between 3100 and 2845 cm$^{-1}$. The vibrations bands assigned to the stretching of the P=N- bands were observed at frequencies in the range of 1100–1150 cm$^{-1}$.

![Figure 2](image_url). The $^1$H NMR spectrum of compound 5 in CDCl$_3$ solution.

### 3.2. Electronic absorption and fluorescence behavior

Absorption and emission properties of coumarin-based cyclotriphosphazene compounds were investigated in different solvents at room temperature (Figures S7–S9). The absorption spectra of the studied compounds (5–7) did not show any changes in the different studied solvents. The absorption bands observed at around 340 nm were attributed to the substituted coumarin groups because cyclotriphosphazenes are optically inert.
in the UV-Vis region [30]. In addition, the ground state absorption spectra of the compounds in THF/H₂O (20:1) were measured at different concentrations. From these spectra, the molar extinction coefficients of the compounds (5–7) were calculated according to the Beer–Lambert law (Figures S10–S12). The solvent effects on the fluorescence emission behavior of the studied compounds (5–7) were determined in different organic solvents as well as THF/H₂O (20:1) solution (Figures S13–S15). All studied coumarin substituted cyclotriphosphazenes (5–7) demonstrated similar fluorescence behavior (Figures 3, S16, and S17) and the maxima of the fluorescence emission peaks in THF/H₂O (20:1) solution are listed in the Table. The best fluorescence emission intensity with a large Stokes shift (158 nm) was observed for compound 7 in THF/H₂O (20:1) at 502 nm (Figure 3).

### Figure 3
Excitation and emission spectra of compound 5 in THF/H₂O (20:1) solution. Excitation wavelength: 340 nm, C = 6 µM.

### Table
Absorption, excitation, and emission spectral data for coumarin substituted cyclotriphosphazenes (5–7) THF/H₂O (20:1) solution.

| Comp. | λ_{max} (nm) | log ε | λ_{Ex} (nm) | λ_{Em} (nm) | Δ_{Stokes} (nm) | τ_F (ns) |
|-------|--------------|-------|-------------|-------------|----------------|----------|
| 5     | 340          | 5.00  | 340         | 465         | 125            | 0.874 (20%), 3.876 (80%) (for 5)<br>0.672 (21%), 3.839 (79%) (for 5+Fe) |
| 6     | 351          | 4.88  | 352         | 498         | 146            | 0.956 (26%), 3.732 (73%) (for 6)<br>1.035 (28%), 3.710 (72%) (for 6+Fe) |
| 7     | 341          | 4.86  | 344         | 502         | 158            | 0.743 (85%), 2.865 (15%) (for 7)<br>0.745 (87%), 3.175 (13%) (for 7+Fe) |

The fluorescence lifetimes (τ_F) of coumarin substituted cyclotriphosphazenes (5–7) were also measured using the time-correlated single-photon counting (TCSPC) technique (Figures 4, S18, and S19). The lifetimes were found to be biexponential and the τ_F values are given in the Table. Changes in the fluorescence lifetime values indicated the binding of Fe^{3+} ions to the studied compounds (5–7).

### 3.3. Chemosensor properties
The cation binding affinity of the newly synthesized coumarin substituted cyclotriphosphazenes (5–7) toward environmentally active metal ions was investigated using fluorescence spectroscopy (Figures 5, S20, and S21). Stock solutions of corresponding metal chlorides (nitrate derivative for Ag^{+} ion) were prepared in water. Figure 5 shows that the addition of Li^{+}, Na^{+}, K^{+}, Cs^{+}, Mg^{2+}, Ba^{2+}, Ca^{2+}, Cr^{3+}, Mn^{2+}, Co^{2+}, Cu^{2+}, Zn^{2+},
Al\(^{3+}\), Ag\(^{+}\), Cd\(^{2+}\), and Hg\(^{2+}\) caused virtually no change to the fluorescence spectra of the studied compounds at room temperature in THF/H\(_2\)O (20:1) solutions. Unlike the rest of the metal ions, Fe\(^{3+}\) addition caused a dramatic decrease in the emission intensity of these compounds (5–7). The novel coumarin substituted cyclotriphosphazenes (5–7) showed ‘turn-off’ chemosensor properties towards the Fe\(^{3+}\) ions in the solution.

The continuous variation method (Job’s plot) was used to determine the binding stoichiometry between the cyclotriphosphazene compounds (5–7) and Fe\(^{3+}\) cation in THF/H\(_2\)O (20:1) solution. The resulting Job’s plot with a maximum mole fraction for the Fe\(^{3+}\) cation was observed as 0.3, which indicates 2:1 (ligand:metal) binding stoichiometry for the complex formed between the chemosensor compound and Fe\(^{3+}\) cation (Figures 6, S22B, and S23B).

The limit of detection (LOD) (for compounds 5–7 with Fe\(^{3+}\) ions were determined from fluorescence titration experiments using fluorescence spectroscopy (Figures 7, S22A, and S23A). This value was calculated using the \(3\sigma/k\) equation (\(\sigma = \) standard deviation of the blank signal, \(k = \) slope of calibration curve) [31]. The LOD values were found as 4.32 \(\mu\)M, 2.19 \(\mu\)M, and 1.13 \(\mu\)M for compounds 5, 6, and 7, respectively, which allows detection in micromolar concentration ranges. These results showed that the novel coumarin substituted cyclotriphosphazene compounds (5–7) have high sensitivity towards Fe\(^{3+}\) ions.
Figure 6. Job's plot of $\text{5-Fe}^{3+}$ complex in THF/H$_2$O (20:1) solution. Excitation wavelength = 340 nm.

Figure 7. A) The fluorescence titration of compound 5 by addition of Fe$^{3+}$ cation [0–75 μM in THF/H$_2$O (20:1)] for determination of LOD. Excitation wavelength = 340 nm.

4. Conclusions

In this study, three novel 7-hydroxycoumarin substituted cyclotriphosphazene derivatives (5–7) were synthesized. All the synthesized novel compounds (5–7) were characterized by mass spectrometry, FT-IR, and $^1$H and $^{31}$P NMR spectroscopies. The electronic absorption and fluorescence behaviors of the synthesized cyclotriphosphazene derivatives were also studied by UV-Vis and fluorescence spectroscopy in THF/H$_2$O (20:1) solution. The novel studied cyclotriphosphazenes (5–7) were observed with absorption peaks at approximately 340 nm due to the optic properties of substituted coumarin. The fluorescence behaviors of the synthesized compounds were explored and intense emission bands were observed between 465 and 502 nm in THF/H$_2$O (20:1) solution. A significant decrease in the fluorescence emission intensities of the compounds (5–7) by the addition of the Fe$^{3+}$ cation solution was observed and this was evidence of the ‘turn-off’ chemosensor properties of these compounds against the Fe$^{3+}$ ions in the solution. These compounds could easily be used as chemosensors for Fe$^{3+}$ ions in solution because they showed low detection limits for Fe$^{3+}$ ions.
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References

1. Jonaghani MZ, Zali-Boeini H, Moradi H. A coumarin based highly sensitive fluorescent chemosensor for selective detection of zinc ion. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy 2019; 207: 16-22. doi: 10.1016/j.saa.2018.08.061

2. Liu F, Fan C, Pu S. A new “turn-on” fluorescent chemosensor for Zn2+ based on a diarylethene derivative and its practical applications. Journal of Photochemistry and Photobiology A: Chemistry 2019; 371: 248-254. doi: 10.1016/j.jphotochem.2018.11.020

3. Jang M, Kang S, Han MS. A simple turn-on fluorescent chemosensor for CO2 based on aggregation-induced emission: application as a CO2 absorbent screening method. Dyes and Pigments 2019; 162: 978-983. doi:10.1016/j.dyepig.2018.11.031

4. Yao J, Dou W, Qin W, Liu W. A new coumarin-based chemosensor for Fe3+ in water. Inorganic Chemistry Communications 2009; 12 (2): 116-118. doi: 10.1016/j.inoche.2008.11.012

5. Şenkuytu E, Yenilmez Çiftçi G. Structural and chemosensor properties of FDA and FDP derivatives of fluorenyledene bridged cyclotetraphosphazenes. Polyhedron 2016; 115: 247-256. doi: 10.1016/j.poly.2016.04.046

6. Qin JC, Yang ZY, Wang GQ, Li CR. FRET-based rhodamine–coumarin conjugate as a Fe3+ selective ratiometric fluorescent sensor in aqueous media. Tetrahedron Letters 2015; 56 (35): 5024-5029. doi: 10.1016/j.tetlet.2015.07.023

7. Salvador JP, Tassies D, Reverter JC, Marco MP. Enzyme-linked immunosorbent assays for therapeutic drug monitoring coumarin oral anticoagulants in plasma. Analytica Chimica Acta 2018; 1028: 59-65. doi:10.1016/j.aca.2018.04.042

8. Arcau J, Andermark V, Aguiló E, Gandioso A, Moro A et al. Luminescent alkynyl-gold(i) coumarin derivatives and their biological activity. Dalton Transactions 2014; 43 (11): 4426-4436. doi: 10.1039/C3DT52594E

9. Zou Q, Fang Y, Zhao Y, Zhao H, Wang Y et al. Synthesis and in vitro photocytotoxicity of coumarin derivatives for one- and two-photon excited photodynamic therapy. Journal of Medicinal Chemistry 2013; 56 (13): 5288-5294. doi: 10.1021/jm400025g

10. Nofal ZM, El-Zahar MI, Abd El-Karim SS. Novel coumarin derivatives with expected biological activity. Molecules 2000; 5 (2): 99-113.

11. Vargas-Soto FA, Céspedes-Acuña CL, Aqueveque-Muñoz PM, Alarcón-Enos JE. Toxicity of coumarins synthesized by Pechmann-Duisberg condensation against Drosophila melanogaster larvae and antibacterial effects. Food and Chemical Toxicology 2017; 109: 1118-1124. doi: 10.1016/j.fct.2017.05.051

12. Reddy DS, Kongot M, Netalkar SP, Kurjogi MM, Kumar R et al. Synthesis and evaluation of novel coumarin-oxime ethers as potential anti-tubercular agents: their DNA cleavage ability and BSA interaction study. European Journal of Medicinal Chemistry 2018; 150: 864-875. doi: 10.1016/j.ejmech.2018.03.042

13. Lingaraju GS, Balaji KS, Jayarama S, Anil SM, Kiran KR et al. Synthesis of new coumarin tethered isoxazolines as potential anticancer agents. Bioorganic & Medicinal Chemistry Letters 2018; 28 (23): 3606-3612. doi: 10.1016/j.bmcl.2018.10.046

14. Hamdy AM, Khaddour Z, Al-Masoudi NA, Rahman Q, Hering-Junghans C et al. Synthesis of arylated coumarins by Suzuki–Miyaura cross-coupling. Reactions and anti-HIV activity. Bioorganic & Medicinal Chemistry 2016; 24 (21): 5115-5126. doi: 10.1016/j.bmc.2016.08.029
15. Kumar R, Saha A, Saha D. A new antifungal coumarin from Clausena excavata. Fitoterapia 2012; 83 (1): 230-233. doi: 10.1016/j.fitote.2011.11.003
16. Jones G, Jackson WR, Choi CY, Bergmark WR. Solvent effects on emission yield and lifetime for coumarin laser dyes. Requirements for a rotary decay mechanism. Journal of Physical Chemistry 1985; 89 (2): 294-300. doi: 10.1021/j100248a024
17. Jones G, Jimenez JAC. Azole-linked coumarin dyes as fluorescence probes of domain-forming polymers. Journal of Photochemistry and Photobiology B: Biology 2001; 65 (1): 5-12. doi: 10.1016/S1011-1344(01)00236-6
18. El-Kenawy M, Retig W. Multiple emission in coumarins with heterocyclic substituents. Physical Chemistry Chemical Physics 2003; 5 (23): 5221-5228. doi: 10.1039/B308712C
19. Tataroğlu A, Özen F, Koran K, Dere A, Gorgülü AO et al. Structural, electrical and photoresponse properties of Si-based diode with organic interfacial layer containing novel cyclotriphosphazene compound. Silicon 2018; 10 (3): 683-691. doi: 10.1007/s12633-016-9513-x
20. Tanrıverdi Eçik E, Şenkuytu E, İbişoğlu H, Zorlu Y, Yenilmez Çiftçi G. Synthesis and fluorescence properties of cyclotriphosphazenes containing thiazole or thiazole rings. Polyhedron 2017; 135: 296-302. doi: 10.1016/j.poly.2017.07.017
21. Şenkuytu E, Yıldırım T, Ölçer Z, Uludağ Y, Yenilmez Çiftçi G. DNA interaction analysis of fluorenylidene double bridged cyclotriphosphazene derivatives. Inorganica Chimica Acta 2018; 477: 219-226. doi: 10.1016/j.ica.2018.02.035
22. Ün İ, İbişoğlu H, Kilç A, Ün ŞŞ, Yüksel F. Nucleophilic substitution reactions of adamantane derivatives with cyclotriphosphazenes. Inorganica Chimica Acta 2012; 387: 226-233. doi: 10.1016/j.ica.2012.01.021
23. Yenilmez Çiftçi G, Şenkuytu E, Durmuş M, Yüksel F, Kilç A. Fluorenylidene bridged cyclotriphosphazenes: ‘turn-off’ fluorescence probe for Cu2+ and Fe3+ ions. Dalton Transactions 2013; 42 (41): 14916-14926. doi: 10.1039/C3DT51426A
24. Yenilmez Çiftçi G, Eçik ET, Bulut M, Yüksel F, Kilç A et al. Synthesis and characterization of dicoumarol substituted cyclotriphosphazenes. Inorganica Chimica Acta 2013; 398: 106-112. doi: 10.1016/j.ica.2012.01.019
25. Yenilmez Çiftçi G, Şenkuytu E, Bulut M, Durmuş M. Novel coumarin substituted water soluble cyclotriphosphazenes as ‘turn-off’ type fluorescence chemosensors for detection of Fe3+ ions in aqueous media. Journal of Fluorescence 2015; 25 (6): 1819-1830. doi: 10.1007/s10895-015-1672-4
26. Şenkuytu E, Eker Y, Yenilmez Çiftçi G. 4-Hydroxycoumarin functionalized cyclotriphosphazenes: synthesis, characterization and fluorescence properties. Inorganica Chimica Acta 2017; 459: 45-50. doi: 10.1016/j.ica.2017.01.023
27. Pişkin M, Durmuş M, Bulut M. Highly soluble 7-oxy-3-(4-methoxyphenyl)coumarin bearing zinc phthalocyanines: synthesis and investigation of photophysical and photochemical properties. Journal of Photochemistry and Photobiology A: Chemistry 2011; 223 (1): 37-49. doi: 10.1016/j.jphotochem.2011.07.014
28. Bondarenko SP, Frasinyuk MS, Khilya VP. Aminomethylation of 3-ary1-7-hydroxycoumarins. Chemistry of Heterocyclic Compounds 2010; 46 (5): 529-535. doi: 10.1007/s10593-010-0541-y
29. Kaya EN, Yüksel F, Özpinar GA, Bulut M, Durmuş M. 7-Oxy-3-(3,4,5-trimethoxyphenyl)coumarin substituted phthalonitrile derivatives as fluorescent sensors for detection of Fe3+ ions: experimental and theoretical study. Sensors and Actuators B: Chemical 2014; 194: 377-388. doi: 10.1016/j.snb.2013.12.044
30. Alidağı HA, Tümay SO, Şenocak A, Yeşilot S. Pyrene functionalized cyclotriphosphazene-based dyes: synthesis, intramolecular excimer formation, and fluorescence receptor for the detection of nitro-aromatic compounds. Dyes and Pigments 2018; 153: 172-181. doi: 10.1016/j.dyepig.2018.02.012
31. Parker CA, Rees WT. Correction of fluorescence spectra and measurement of fluorescence quantum efficiency. Analyst 1960; 85 (1013): 587-600. doi: 10.1039/AN9608500587
Supporting information

1. Experimental

1.1. Materials and methods

Hexachlorocyclotriphosphazene (98%), 3,4-methoxyphenylacetic acid (97%), 3,4-dimethoxyphenylacetic acid (97%), 3,4,5-trimethoxyphenylacetic acid (97%), and 2,4-dihydroxybenzaldehyde (98%) were obtained from Aldrich. Tetrahydrofuran (THF) (99.0%), dichloromethane (99.0%), and n-hexane (95.0%) were obtained from Merck. Sodium hydride (NaH) at 60% dispersion in mineral oil (Merck) was purified prior to use by washing with dry n-hexane followed by decantation. Silica gel 60 (230–400 mesh) for column chromatography was obtained from Merck. CDCl$_3$ for NMR spectroscopy was obtained from Goss Scientific. Positive ion and linear mode MALDI-MS of compounds was performed in dihydroxybenzoic acid as a MALDI matrix using a nitrogen laser accumulating 50 laser shots using a Bruker Microflex LT MALDI-TOF mass spectrometer. All reactions were monitored using thin-layer chromatography (TLC) on Merck silica gel plates (Merck, Kieselgel 60, 0.25 mm in thickness) with an F254 indicator. Column chromatography was performed on silica gel (Merck, Kieselgel 60, 230–400 mesh; for 3 g of crude mixture, 100 g of silica gel was used in a column of 3 cm in diameter and 60 cm in length). All reactions were carried out under an argon atmosphere. $^1$H and $^{31}$P NMR spectra were recorded in CDCl$_3$ solutions on a Varian INOVA 500 MHz spectrometer. Absorption spectra in the UV-Vis region were recorded with a Shimadzu 2101 UV-Vis spectrophotometer. Fluorescence excitation and emission spectra were recorded on a Varian Eclipse spectrofluorometer using 1-cm path length cuvettes at room temperature. The fluorescence lifetimes were obtained using a Horiba-Jobin-Yvon-SPEX Fluorolog 3-2iHR instrument with Fluoro Hub-B Single Photon Counting Controller at an excitation wavelength of 470 nm. Signal acquisition was performed using a TCSPC module (NanoLED-390 emitting 390 nm).

1.2. Chemosensor studies

Spectroscopic changes upon addition of metal salts to the ligand were recorded using a fluorescence spectrophotometer. All fluorescence emission spectral studies were performed in tetrahydrofuran/water (20:1) solutions (C = 6 $\mu$M) of the coumarin substituted cyclotriphosphazene compounds (5–7) at room temperature. The water solutions (C = 0.1 M) of the corresponding metal chlorides (nitrate derivative for Ag ion) were used as the source of metal ions. The UV-Vis spectra were routinely acquired at 25 °C in a 1-cm path length quartz cuvette with 2 mL of cyclotriphosphazene compounds (5–7) by the addition 0.1 mL of different metal solutions (Li$^+$, Na$^+$, K$^+$, Cs$^+$, Mg$^{2+}$, Ba$^{2+}$, Ca$^{2+}$, Cr$^{3+}$, Mn$^{2+}$, Fe$^{3+}$, Co$^{2+}$, Cu$^{2+}$, Zn$^{2+}$, Al$^{3+}$, Ag$^+$, Cd$^{2+}$, and Hg$^{2+}$). Titration experiments of cyclotriphosphazene compounds (5–7) in the presence of Fe$^{3+}$ were carried out with freshly prepared stock solutions of cyclotriphosphazene compounds (5–7) in tetrahydrofuran/water (20:1) and Fe$^{3+}$ solution in water using a fluorescence spectrophotometer.
Figure S1. Mass spectrum of compound 6.

Figure S2. $^{31}$P NMR spectrum of compound 6 in CDCl$_3$. 
Figure S3. $^1$H NMR spectrum of compound 6 in CDCl$_3$.

Figure S4. Mass spectrum of compound 7.
Figure S5. $^{31}$P NMR spectrum of compound 7 in CDCl$_3$.

Figure S6. $^1$H NMR spectrum of compound 7 in CDCl$_3$. 
**Figure S7.** UV-Vis absorption spectra of compound 5 in different solutions.

**Figure S8.** UV-Vis absorption spectra of compound 6 in different solutions.
**Figure S9.** UV-Vis absorption spectra of compound 7 in different solutions.

**Figure S10.** Molar extinction coefficient of compound 5 in 20-1 THF-water solution.
Figure S11. Molar extinction coefficient of compound 6 in 20-1 THF-water solution.

Figure S12. Molar extinction coefficient of compound 7 in 20-1 THF-water solution.
Figure S13. Fluorescence emission spectra of compound 5 in different solutions.

Figure S14. Fluorescence emission spectra of compound 6 in different solutions.
Figure S15. Fluorescence emission spectra of compound 7 in different solutions.

Figure S16. Excitation (dashed lines) and emission spectra (solid lines) of compound 6 in THF-H₂O (20/1).
Figure S17. Excitation (dashed lines) and emission spectra (solid lines) of compound 7 in THF-H$_2$O (20/1).

Figure S18. Fluorescence decay profiles of compound 6 in the absence and presence of Fe$^{3+}$ ion using laser excitation source of 390 nm.
Figure S19. Fluorescence decay profiles of compound 7 in the absence and presence of Fe$^{3+}$ ion using laser excitation source of 390 nm.
Figure S20. A) Fluorescence emission spectra and B) bar graph of compound 6 (6 µM) with the addition of metal ions.
Figure S21. A) Fluorescence emission spectra and B) bar graph of compound 7 (6 µM) with the addition of metal ions.
Figure S22. A) Job’s graph and B) the fluorescence titration of compound 6 by addition of Fe^{3+} cation [0–50 µM in THF/water (20:1)] for determination of LOD (excitation wavelength = 340 nm).
Figure S23. A) Job’s graph and B) the fluorescence titration of compound 7 by addition of Fe^{3+} cation [0–50 µM in THF/water (20:1)] for determination of LOD (excitation wavelength = 340 nm).