RESEARCH ARTICLE

Seroprevalence of hepatitis E virus infection in the Americas: Estimates from a systematic review and meta-analysis

Nathalie Verónica Fernández Villalobos1*, Barbora Kessel2, Isti Rodiah2, Jördis Jennifer Ott3,4, Berit Lange2,4, Gérard Krause2,3,4,5

1 Department of Epidemiology, PhD Programme, Helmholtz Centre for Infection Research (HZI), Braunschweig-Hannover, Germany, 2 Department of Epidemiology, Helmholtz Centre for Infection Research (HZI), Braunschweig, Germany, 3 Hannover Medical School, Hannover, Germany, 4 German Centre for Infection Research (DZIF), Braunschweig, Germany, 5 Twincore, Centre for Experimental and Clinical Infection Research, Braunschweig-Hannover, Germany

* Nathalie.Fernandez@helmholtz-hzi.de

Abstract

Background

Hepatitis E virus (HEV) infection is responsible for inflammatory liver disease and can cause severe health problems. Because the seroprevalence of HEV varies within different population groups and between regions of the continent, we conducted a systematic review on the topic in order to provide evidence for targeted prevention strategies.

Methods

We performed a systematic review in PubMed, SCIELO, LILACS, EBSCO, and Cochrane Library and included reports up to 25 May 2021 (PROSPERO registration number: CRD42020173934). We assessed the risk of bias, publication bias, and heterogeneity between studies and conducted a random-effect meta-analysis for proportions using a (binomial-normal) generalized linear mixed model (GLMM) fitted by Maximum Likelihood (ML). We also reported other characteristics like genotype and risk factors.

Results

Of 1212 identified records, 142 fulfilled the inclusion criteria and were included in the qualitative analysis and 132 in the quantitative analysis. Our random-effects GLMM pooled overall estimate for past infection (IgG) was 7.7% (95% CI 6.4%–9.2%) with high heterogeneity (I² = 97%). We found higher seroprevalence in certain population groups, for example in people with pig related exposure for IgG (ranges from 6.2%–28% and pooled estimate of 13.8%, 95% CI: 7.6%–23.6%), or with diagnosed or suspected acute viral hepatitis for IgM (ranges from 0.3%–23.9% and pooled estimate of 5.5%, 95% CI: 2.0%–14.1%). Increasing age, contact with pigs and meat products, and low socioeconomic conditions are the main risk factors for HEV infection. Genotype 1 and 3 were documented across the region.
Conclusion

HEV seroprevalence estimates demonstrated high variability within the Americas. There are population groups with higher seroprevalence and reported risk factors for HEV infection that need to be prioritized for further research. Due to human transmission and zoonotic infections in the region, preventive strategies should include water sanitation, occupational health, and food safety.

Introduction

Hepatitis E virus (HEV) is responsible for a liver disease that affects millions of people worldwide, especially in low- and middle-income countries [1]. In 2005, a disease burden of 20 million HEV infections worldwide and 3.3 million symptomatic cases was estimated [2]. For 2019, the Institute for Health Metrics and Evaluation has also calculated a global HEV prevalence rate of 20.5 (95% uncertainty intervals 16.8–24.7) cases per 100 000 people [3].

Generally, the HEV disease is self-limiting and has mild symptoms; however, in some cases it can result in severe acute hepatitis, extrahepatic disorders, chronic hepatitis leading to cirrhosis, and fulminant hepatitis [4]. Pregnant women have been documented to have an increased risk of fulminant hepatitis [5]. Chronic hepatitis E infection has been reported in immunocompromised people [6], affecting mainly solid organ transplants (SOT) patients with 66% development of chronic hepatitis [7].

HEV genome can be grouped into 8 genotypes (HEV-1 to HEV-8), but only genotype 1–4 can infect humans [8]. Genotype 1 and 2 have been identified mainly in Asia and Africa, and they are related to fecal-oral transmission; while, genotype 3 and 4 are identified as zoonosis and circulate in animals and humans. Genotype 3 has been documented worldwide [8].

Current literature suggests that the main route of transmission of the virus is fecal-oral, mainly through contaminated drinking water or food, and it is commonly found in low- and middle-income countries due to the limited access to drinkable water and poor sanitation infrastructure [6].

Most systematic reviews on HEV focus on Europe [9], the Middle East, and the North African region [10]. Some reviews for HEV seroprevalence included individual countries of the Americas [11, 12], but not the whole continent. A review at global level also included some countries of the Americas [13], but it did not include all available evidence on the situation on the continent. Other reviews have focused on non-endemic countries [14, 15], industrialized countries [16], or on a specific population such as children [17], immunocompromised individuals [18], or pregnant women [19] (Table 2, page 3 in S1 File). Thus, there is a lack of systematically retrieved evidence on the seroprevalence and risk factors of HEV in the majority of the countries in the Americas, and there is not a realistic picture of the HEV seroprevalence in the continent. There is a need to compile evidence that includes different populations, diagnostic methods, and especially considering particular languages and databases of the region.

In this systematic review, we aimed to estimate the HEV seroprevalence of the Americas focusing on the analysis of serological markers, population groups, and regions. We also considered the risk factors, genotypes, and routes of transmission associated with the infection of HEV in the continent. Likewise, the aim was to provide evidence for improving the related burden of this disease and to target prevention strategies for HEV.
Material and methods

Search strategy

We performed a systematic review (registration number in PROSPERO CRD42020173934), following the PRISMA guidelines and checklist (S2 File) [20], in PubMed, SCIELO, LILACS, EBSCO, and Cochrane Library, searching for publications on seroprevalence of Hepatitis E Virus (search terms “hepatitis E” and “seroprevalence,” page 1 in S1 File) without date or language restrictions. The literature search included reports up to 25 May 2021. Further, we applied the snowball method [21] starting with the references of the existing reviews to ensure that we included all available relevant studies. All references were processed in Mendeley Desktop [22].

Inclusion and exclusion criteria

We included reports conducted in the Americas, reporting anti-HEV immunoglobulin G, M, and/or total (IgG, IgM, and/or total anti-HEV), and Ribonucleic acid (RNA) in humans. We defined Americas as the countries included in the Pan American Health Organization (PAHO), and we added one study that was done in Greenland in Inuits; a population that inhabits the Arctic regions of Greenland, Canada, and Alaska [23].

We considered the different serological markers based on the detection of anti-HEV antibodies: i) IgM markers of acute or recent infection, ii) IgG markers of past infection, iii) total antibodies markers of detection of antibodies to HEV, and iv) RNA markers of acute infection.

We included publications using any of the following study designs: observational studies, surveys, cohort studies, cross-sectional studies, case control studies, and outbreak investigation reports. We excluded studies reporting seroprevalence based on fecal samples.

Data extraction

One researcher (NVFV) extracted the following information: first author, journal, year of publication, country, sample size for total anti-HEV, sample size for IgG, sample size for IgM, sample size for RNA, start and end of data collection, assay used for total anti-HEV, assay used for IgG, assay used for IgM, assay used for RNA, seroprevalence values total anti-HEV, seroprevalence values IgG, seroprevalence values IgM, seroprevalence values RNA, unit of measure of effect (outcome = odds ratio (OR) and relative risk (RR)), genotype, population group, age, the proportion of male, risk factors (outcome = list of findings), routes of transmission (outcome = list of findings). Two other researchers (BK & IR) independently crosschecked all extracted data. Any conflicts were resolved in a discussion.

Risk of bias assessment

Two researchers (NVFV & IR) assessed the risk of bias using an adapted version of the ROBINS-I tool [24] for non-randomized studies. Specifically, they analyzed the studies in terms of bias due to confounding (high risk if no age information was provided), selection of participants and follow-up (high risk if different follow-up time for different groups was performed), misclassification of exposure (high if the source was unclear), missing data (high if participants were excluded on large scale for missing data), measurement of outcome (high if the report was not available), or reporting (high if selective subgroup was reported). We measured the risk scales as low, moderate, and high. Any conflicts in the assessment were resolved in a discussion.
Data analysis

Descriptive. We performed a qualitative synthesis with all the included studies and presented their main characteristics as tables and as narrative. Moreover, we organized the findings according to the population groups and regions of the Americas (North America, South America, Central America and the Caribbean). Further, we grouped different population groups as follows: 1) general population, 2) blood donors, 3) ethnic groups, 4) exposed population, 5) occupational group, 6) pig related occupation, 7) viral hepatitis, 8) immunodeficiency, 9) pregnant women, 10) rural population, 11) children, and 12) combined.

For those groups that were not consistent, we applied the following definitions:

- General population: urban, mixed urban, and rural residents, civilian participants, people without exposure, volunteers, and controls.
- Ethnic groups: Indigenous population, Indians population, tribes, and Amazonian and Andean population.
- Exposed population: Cohabitants of the exposed people, inmates, homeless people, alcoholics, sex workers, persons who inject drugs, homosexuals, and travelers.
- Occupational group: veterinary students and workers, healthcare workers, waste pickers, religious groups, workers of natural resources, biologists, hunters, workers in water and sewage companies, soldiers, and people deployed in the military.
- Pig related occupation: Swine related occupations and people who work in pig farms with direct contact with pigs.
- Viral hepatitis: People with acute hepatitis, acute jaundice, positive anti-hepatitis antibodies, suspected cases, sporadic cases, and outbreak cases.
- Immunodeficiency: Hemodialysis patients, people living with human immunodeficiency virus (HIV), transplant or plasma recipients, patients with chronic hepatitis C virus (HCV) or hepatitis B virus (HBV), people with liver failure, cancer, Elevated alanine aminotransferase (ALT), patients with plasmapheresis, patients with hepatosplenic schistosomiasis, patients with *Schistosoma mansoni*, and people subjected to invasive procedures.

From the included studies, we also extracted information related to HEV genotypes found in the region and geographically represented their distribution in the Americas using vector layers based on shapes from Environmental Systems Research Institute (ESRI) [25] and using R [26] version 4.0.2 (using packages "ggplot2" [27], "sf" [28], "dplyr" [29]).

Publication bias. We used funnel plots from the R package “meta” [30] to visualize the interplay between reported seroprevalence estimates and their precision in order to identify any irregularities, which we then interpreted according to the recommendations by Sterne et al. [31]. We considered the risk of not publishing a result because a particular seroprevalence value has been found to be low in our setting. However, it potentially had a considerable impact on our results due to the number of studies available per country and per population group. Therefore, we visualized the composition of the studies by stacked bar charts (constructed in "ggplot2" [27] package in R) and accounted for it when interpreting the results.

Meta-analysis. We conducted our analysis for each of the considered markers, total anti-HEV, IgG, IgM, RNA, separately. IgM and RNA results were further separated into those obtained from the whole sample and those obtained from previously positive samples (for example IgG or total anti-HEV positive). For RNA in positive samples, we refrained from conducting a meta-analysis due to low sample sizes (only two studies with more than 50 samples).
and varying definition of "previously positive samples" (IgG positive, IgM positive, at least one of the before mentioned, or both at the same time).

In all other cases, using the raw data (the reported number of positive and the reported sample size) for each study, we performed a random-effect meta-analysis for proportions using a (binomial-normal) generalized linear mixed model (GLMM) fitted by Maximum Likelihood (ML). We constructed a 95% confidence interval for the pooled estimate using t-quantiles with k-1 degrees of freedom (k = number of pooled studies). We assessed heterogeneity visually by forest plots, by assessing the percentage of variance over the studies using $I^2$, and by estimating the between study variance $\tau^2$.

We performed subgroup analyses by region, country, and population group. Based on previous knowledge and the observed large heterogeneity between the reported seroprevalences, we expected different seroprevalence levels in some population subgroups. For a formal comparison of two, or more pooled estimates, we used the Q-test for subgroup differences as implemented in the R package “meta” [30] (referred to also as “comparison based on fixed-effects model between subgroups” [32] in literature).

In three instances the reported seroprevalence included in the pooling was based on the National Health and Nutrition Examination Survey (NHANES) dataset, and thus subject to necessary corrections due to the survey design. To be able to include these seroprevalences into the random-effects GLMM framework, we calculated for each of them a fictive number of positive cases and a fictive sample size that would yield exactly the reported seroprevalence, and the reported 95% confidence interval assumed to be constructed on the logit-scale using normal quantiles. The resulting decimal numbers were then rounded when calling the meta-prop function in package “meta” [30].

In each meta-analysis, we included one seroprevalence result per sample. Our choice is detailed in Table 4, pages 9–36 in S1 File; in Fig 3, page 62 in S1 File (for samples analyzed repeatedly by different tests); and in Fig 6, page 65 in S1 File (for studies based on the NHANES dataset).

In case of samples repeatedly analyzed by different tests, we examined the impact of our choice by conducting a sensitivity analysis. We replaced the primarily chosen values by their alternatives one at a time and all at once, and carried out the respective meta-analysis. When pooling within regions involved several seroprevalences reported in one publication, we examined whether allowing for a positive correlation between such results would change our pooled estimates. We did this by endowing the basic random-effects GLMM with an additional random effect (for publication) and fitted the extended model using R package “lme4” [33].

All the analysis were performed using R [26] version 4.0.2 (using package “meta” [30] and “metaphor” [34]). Forest plots were created using R package “forestplot” [35]. The significance level used was 0.05.

Results

We identified 1212 records and retrieved 217 for full-text screening. After verifying the inclusion criteria, we included 142 for our qualitative analysis and 132 for the quantitative analysis (Fig 1). The snowball searching of references in the existing reviews did not reveal any additional relevant publications.

Of the 142 included articles, the majority of studies originated from Brazil (n = 38), United States (USA; n = 33), and Argentina (n = 14). According to region, South America hosted 57% of the included studies (n = 81), North America 37% (n = 52), and Central America and the Caribbean 6% (n = 9). The studies were conducted between 1967 (retrospective) and 2018,
Fig 1. Flow diagram of the selected studies for the HEV seroprevalence.

https://doi.org/10.1371/journal.pone.0269253.g001
and they included different population groups. The sample sizes ranged between 14 and 18,829 participants, with ages between 1–98 years.

Regarding the assays, 20% of the studies performed the analysis with Abbott Laboratories (n = 29), followed by the combination of multiple assays with 15% (n = 21), and by “in house” ELISA (enzyme-linked immunosorbent assay) methods with 14% (n = 20). The characteristics of the included studies are presented in the Table 4, page 9–36 in S1 File.

Risk of bias assessment

We assessed the risk of bias due to a) confounding, b) selection, c) misclassification, d) missing data, and e) measurement of outcome. Confounding was moderate in most studies as adjusted estimates or age information was provided. High risk of bias was found for 18 studies as they did not report any age related information. Selection bias and missing data were mostly moderate, and misclassification was mostly low. The total assessment can be found in the Table 3, page 4–8 in S1 File. Since the majority of studies had a low or moderate risk assessment, we did not exclude any study and included all of them into the qualitative analysis.

HEV seroprevalence

The results of our pooled analyses and the $I^2$ results by population group and by region are presented in the Tables 1 and 2, respectively. Further details can be found in supplementary figures as detailed in the text below in S1 File.

**Total HEV antibodies.** In 21 studies that reported on total anti-HEV antibodies (37 results), we found ranges of seroprevalence from 0%–40.6%. The random effects meta-analysis pooled estimate was 4.3% (95% CI 2.9%–6.4%) with high heterogeneity ($I^2 = 95%$).

The subgroup analysis (Table 1) shows that seroprevalences reported for some of the considered subpopulations tend to be higher. For example, exposed populations [seroprevalence range: 5.9%–13.5% (n = 3), and pooled estimate: 9.6% (95% CI: 3.9%–21.9%)], people with pig related exposure [range: 10.9%–40.6% (n = 3), and pooled estimate: 19.7 (95% CI: 3.8%–60.2%)], or ethnic groups [range: 5.4%–17% (n = 4), and pooled estimate: 8.5% (95% CI: 4.4%–16%)] had higher seroprevalence compared with those reported for pregnant women.

| Population Group                  | Total anti-HEV | IgG          | IgM (total sample) | IgM (+ sample) |
|-----------------------------------|----------------|--------------|-------------------|---------------|
|                                   | Proportion (95% CI) | $I^2$ | Proportion (95% CI) | $I^2$ | Proportion (95% CI) | $I^2$ | Proportion (95% CI) | $I^2$ |
| General population                | 3.1% (1.4%-6.8%)   | 89%          | 7.2% (5.2%-9.9%)  | 93%      | 0.8% (0.3%-2%)  | 84%    | 15.5% (4.2%-43.5%) | 79%    |
| Blood donors                      | 4.2% (1.6%-10.9%)  | 98%          | 6.6% (4.2%-10.3%) | 98%      | 5.7% (2.7%-11.7%)| 94%    | 18.3% (11.6%-28.4%)| 10%   |
| Ethnic groups                     | 8.5% (4.4%-16%)    | 83%          | 5.7% (2.7%-11.7%) | 94%      | 7.1% (3.7%-13.2%)| 68%    | 18.3% (11.6%-28.4%)| 10%   |
| Exposed population                | 9.6% (3.9%-21.9%)  | 60%          | 11.4% (5.4%-22.6%)| 96%      | 13.8% (7.6%-23.6%)| 92%    | -                  | -     |
| Occupational group                | 5.3% (2%-13.7%)    | 92%          | 7.1% (3.7%-13.2%) | 68%      | -                  | -      | -                  | -     |
| Pig related Occupation            | 19.7% (3.8%-60.2%) | 93%          | 13.8% (7.6%-23.6%)| 92%      | -                  | -      | -                  | -     |
| Known viral hepatitis             | -                | -            | 8.2% (4.6%-14.3%) | 97%      | 5.5% (2.0%-14.1%)  | 93%    | 29% (16.2%-46%)    | 32%    |
| Immunodeficiency                  | -                | -            | 10.4% (6.8%-15.4%)| 96%      | 0.9% (0.3%-2.4%)  | 59%    | 29% (16.2%-46%)    | 32%    |
| Pregnant women                    | 0.8% (0.2%-3.1%)   | 19%          | 3.0% (0.7%-11.4%) | 93%      | -                  | -      | -                  | -     |
| Rural population                  | -                | -            | 11.5% (5.4%-22.9%)| 97%      | -                  | -      | -                  | -     |
| Children                          | -                | -            | 1.1% (0.1%-8.1%)  | 88%      | -                  | -      | -                  | -     |

* Number of observations. Importantly, some studies included more than one population group, so this number is different than number of studies.
* Only studies with sample size at least 10 are meta-analyzed.
* Without an outbreak with seroprevalence 100%.

https://doi.org/10.1371/journal.pone.0269253.t001
In many of the subgroups, the heterogeneity of the reported results remained high. See also Fig 4, page 63 in S1 File.

Regarding regions, 11 studies were from South America, nine studies were from North America (only one study outside of USA), and two in Central America and the Caribbean, with seroprevalence ranges from 0%–17%, 0.4%–13.5%, and 2.9%–40.6, respectively. All three regions show similar level of variability in the reported seroprevalences; a formal test does not confirm any significant differences between the pooled estimates (Q-test, p = 0.1678). For more details, see Table 2 and Fig 5, page 64 in S1 File.

### Past hepatitis E exposure (IgG)

Disregarding repeated analyses of the same sample, as described in Methods, we considered 157 seroprevalences reported in 105 publications. The seroprevalence ranges from 0%–71% (median of 8%), and the random-effects GLMM pooled overall estimate was 7.7% (6.4%–9.2%) with high heterogeneity ($I^2 = 97\%$, $\tau^2 = 1.4$).

Fig 2 shows an overview of the pooled results within subgroups, and details of the seroprevalences underlying the pooled estimates are given in the S1 File, pages 66–71.

Compared to general (adult) population (pooled estimate 7.2%, 95% CI: 5.2%–9.9%), children had a lower seroprevalence (1.1%, 95% CI: 0.1%–8.1%, Q test p = 0.0037), and people with a pig related exposure had a higher one (13.8%, 95% CI: 7.6%–23.6%, Q-test p = 0.0183). The heterogeneity in the subgroups remained high.

By region, 63 studies (97 results) were from South America, 36 (52 results) from North America (only one study outside of USA), and two in Central America and the Caribbean. Brazil (33 studies), the USA (20 studies), Argentina (13 studies), and Mexico (10 studies) were the countries with more number of published studies.

The seroprevalence by region ranged from 0%–66.3%, 0%–43.6%, and 0%–71% in South America, North America, and Central America and the Caribbean respectively. The pooled estimates (see Fig 2) were not statistically significantly different (Q-test, p = 0.4797). When we compared the regions within subgroups with more evidence (for example in blood donors, general population, viral hepatitis, and immunodeficiency), the collected data suggested a regional difference (between North and South America) only in the latter case. Details can be found in S1 File, pages 66–69.

Fig 2 shows also the pooled seroprevalences by country. They were mostly comparable with perhaps the exception of Peru, Bolivia, and Colombia (Fig 14, page 73 in S1 File). The heterogeneity within countries remained high (Fig 13, page 72 in S1 File).

### Acute hepatitis E (IgM)

Due to some studies that reported IgM results based on positive samples, we divided this group into two: those that use a total sample and those that used only samples with positive results.

---

Table 2. Pooled analysis of HEV seroprevalence by region.

| Region                             | Total anti-HEV | IgG | IgM (total sample) | IgM (+ sample) |
|------------------------------------|----------------|-----|--------------------|---------------|
|                                    | Proportion (95% CI)  | $I^2$ | $k$ | Proportion (95% CI)  | $I^2$ | $k$ | Proportion (95% CI)  | $I^2$ | $k$ | Proportion (95% CI)  |
| North America                      | 3.4% (1.5%-7.2%) | 98% | 11 | 9% (6.6%-12.2%) | 97% | 52 | 0.8% (0.4%-1.5%) | 77% | 13 | 2.5% (0.1%-48.1%) |
| South America                      | 4.1% (2.5%-6.5%) | 87% | 21 | 7.2% (5.7%-9.1%) | 96% | 97 | 1.5% (0.6%-3.8%) | 79% | 9 | 13.8% (7.7%-23.6%) |
| Central America and the Caribbean  | 10.7% (2.4%-36.4%) | 93% | 5 | 6.5% (1.9%-20.3%) | 96% | 8 | - | - | - | - |

* Number of observations. Importantly, some studies included more than one population group, so this number is different than number of studies.

† Only studies with a sample size of at least 10 were meta-analyzed.

https://doi.org/10.1371/journal.pone.0269253.t002
The total sample for IgM analysis. Without including the viral hepatitis group, 23 IgM seroprevalences were reported by 21 studies (Fig 15, page 74 in S1 File). The observed seroprevalences ranged from 0% to 10% (median 1%). Values above 5% were reported for pediatric patients in Mexico [36] (6.1%, n = 99), ethnic groups in Argentina [37] (8.7%, n = 126), and female blood donors in Brazil [38] (10%, n = 20). A pooled seroprevalence was 0.9% (95% CI: 0.6%–1.5%, $I^2 = 83\%$, tau$^2 = 0.97$).

In 12 studies reporting IgM results from sera of patients with confirmed or suspected acute viral hepatitis (viral hepatitis group), the seroprevalence ranged from 0% to 100%, with a median of 8%. Seroprevalences higher than 10% were reported from Venezuela [39], Chile [40], Colombia [41], and Cuba [42–44], the latter country accounted also for the observed prevalence of 100% [44] (Fig 16, page 75 in S1 File). Without this apparent HEV outbreak, the pooled seroprevalence was 5.5% (95% CI: 2.0%–14.1%, $I^2 = 93\%$, tau$^2 = 2.1$), significantly (Q-test, p = 0.006) higher than in the other subgroups (pooled together).

Comparing South and North America, we did not find a significant difference between the pooled estimates (Q-test, p = 0.2211), the former being 1.5% (95% CI: 0.6%–3.8%), the latter
0.8% (95%CI: 0.4%–1.5%) (Fig 17, page 75 in S1 File). Central America and the Caribbean was not considered in the comparison due to the low number of reported results (n = 1).

IgM among positive samples. In 26 studies that presented 33 results regarding IgM among IgG or total anti HEV positive cases, we found ranges of seroprevalence from 0%–100%, with median 16%. Considering only results based on a sample size of at least 10, the range was 0%–50%.

The pooled seroprevalence was 11.4% (95% CI: 6.2%–19.9%, I² = 55%, τ² = 1.8), split into subgroups we got 29% (95% CI: 16.2%–46%, I² = 79%, τ² = 1.5) for the viral hepatitis group, and 15.5% for blood donors (95% CI: 4.2%–43.5%, I² = 32%, τ² = 0) (Fig 18, page 76 in S1 File).

By region, 19 studies were performed in South America (Brazil = seven, and Argentina = seven), six in North America (USA = five), and one in Central America and the Caribbean (Fig 19, page 77 in S1 File). The data did not confirm differences between South and North America (p = 0.218 for the subgroup comparison), with a pooled estimate of 13.8% (95% CI: 7.7%–23.6) in the south and 2.5% (95% CI: 0.1%–48.1%) in the north.

Acute and chronic hepatitis E (RNA). As before, we divided the RNA group into those that use a total sample and in those that used only positive samples.

The total sample for RNA analysis. In 23 studies that presented RNA results from the total sample (24 results reported altogether), we found ranges of seroprevalence from 0%–54.5% with median of 0. All but one study reported RNA-seroprevalence under 10.4%. The one outstanding study [36] reported a seroprevalence of 54.5% among pediatric patients in Mexico; in contrast, IgG (IgM) seroprevalence in this group was 3% (6%).

From the other studies, nonzero seroprevalences were reported for cirrhosis patients in Argentina [45] (21%), renal transplant recipients in Brazil (10.4% [46] and 3.1% [47]), and patients with acute viral hepatitis in Argentina [48] (6.3%) and Cuba [49] (8.5%). For more details, see Fig 20, page 78 in S1 File.

RNA among positive samples. In 23 studies that reported information related to RNA positive results among IgG and/or IgM positive samples (26 results reported altogether), we found ranges of seroprevalence from 0%–100%, with median of 0%.

In eight cases, the underlying sample size was less than 10 and in two cases as low as one sample. Leaving these cases out, the proportion of RNA positive results ranged from 0%–60% (median of 0%). Non-zero proportions were observed especially for people with acute viral hepatitis (Fig 21, page 79 in S1 File).

Risk factors and route of transmission

Of the 142 included studies, 96 mentioned risk factors related to the HEV seropositivity (Table 5, page 37 in S1 File). Of those, 41 studies reported a measure of association such as Odds Ratios (OR, n = 39) or Prevalence Ratios (PR, n = 2).

Increasing age. In the general population, increasing age is associated with HEV infection (OR 1.06 (1.05–1.06)) [50], increasing from 30 years [51], increasing age (OR 3.50 (1.39–8.87)) with high prevalence in older than 46 years old group [52], and among those aged between 41–60 years (OR 3.2 (1.09–9.7)) [53]. In children and young adults, seroprevalence was higher in people between 26 to 29 years of age compared to children below 5 years (OR 15.50 (2.17–110.90)) [54].

Higher age is also a risk factor among blood donors, with a 4-fold increase odds of having HEV at an advanced age (45–59 years) than at younger age (OR 3.96 (1.54–10.22)) [55], and with a 3-fold increase odds for age ≥ 50 against <50 (OR 3.33 (1.11–9.95)) [56].
In the exposed population, being older than 30 (OR 3.61 (1.31–9.94)) [57] or older than 45 years old (OR 3.2 (1.1–10.7)) [58] was associated with HEV.

In the occupational group, US service members older than 35 years had higher odds to be HEV positive (OR 2.9 (0.9–8.8)) [59], and waste pickers older than 40 years were more likely to be HEV positive (OR 5.2 (1.5–17.5)) [60]. HEV seropositivity was associated with increasing age in waste pickers by OR 6.52 (1.95–21.78) [61] and in Mennonites by OR 1.05 (1.00–1.09) [62] in comparison with the control group.

Higher age also increases odds for HEV positivity in people with acute hepatitis (OR 2.04 (1.09–3.79) for those ≥ 60 years against <60 [63], and OR 1.5 (1.03–1.07) per-year increase) [64], and in those with immune deficiency (adjusted OR, 3.34 (1.54–7.24) for age ≥ 60 against <60 [65], and adjusted OR 1.05 (1.02–1.09) per 1 year increase) [66].

In rural population, increasing age, for each 1-year increase, was associated with HEV seropositivity (OR 1.03 (1.01–1.05) [67], OR 1.04 (1.04–1.05) [68], OR 1.05 (1.04–1.07) [69]), especially in the 21–30 [67], and in the 41–50 age groups [70].

Sex. In the general population, it was found that females had a higher prevalence than males (OR 1.2 (1.06–1.38)) [50], and those young females (6–39 years) were found to have higher HEV seropositivity compared to males (OR 1.44 (1.03–2.02)) [71]. In contrast, one study found that males had a higher anti-HEV IgG prevalence rate than females [72].

In the immunosuppressed group, being male was a risk factor for HEV (adjusted OR 2.07 (1.17–3.66)) [66], as well as for the viral hepatitis group (OR 2.10 (1.39–3.19)) [64].

Ethnicity. For the general population, three studies referred to this factor. Two studies in USA found that non-Hispanic-Asian ethnicity was associated with HEV seropositivity when comparing with other groups (OR 1.69 (1.12–2.56)) [50]. In addition, non-Hispanic black people had lower rates of HEV infection than white participants (OR 0.60 (95% CI 0.46–0.79)) [71]. Moreover, place of birth was associated as an HEV factor [73, 74]. One study found that people born outside the United States were more likely to be HEV positive (OR 1.78 (1.23–2.58)) [71].

In Alaska, USA, native Americans were less likely to be seropositive to HEV than non-native people [75]. Moreover, Talarmin, A et al. also documented that seroprevalence rates differed significantly between ethnic groups, with higher adjusted-per-age rates among Brazilians (OR 4.8 (1.9–11.9), Chinese and Hmongs (OR 4.0 (1.5–10.9), and Haitians (OR 3.3 (1.1–9.9) when comparing against Creole [76].

Among viral hepatitis patients, being born outside the United States or Canada (OR 2.09 (1.03–4.25)), or being Asian and less educated people living in Canada or in USA (OR 2.92 (1.42–6.01)) were also documented as a risk factor for HEV seropositivity [64].

Immunosuppressed system. Comorbidities such as cirrhosis (OR 8.95 (4.0–15.99)) [45], other liver diseases (adjusted OR 7.78 (3.43–17.64)) [66], or having low CD4 counts (OR 0.96 (0.94–0.99) for a CD4 increase of 10 counts) [77] increased the odds of having HEV. Moreover, one study demonstrated an association between the risk of post-transplant HEV infection and graft rejection (OR, 14.2 (1.26–160.0)) [78].

Occupation. People occupationally exposed to pigs have more risk to be HEV positive than those without exposure (PR 2.42 (1.66–3.53)) [79]. Moreover, swine veterinarians were 1.51 times more likely to be anti-HEV positive than normal blood donors when tested with swine HEV antigen (CI 95% 1.03–2.20) [80]. Having HEV antibodies is associated with increasing years of occupation in a slaughterhouse (OR 8.81 (1.28–60.34), for employment = 20 years) [81]. Especially raising pigs, within waste pickers, was associated with being HEV positive (OR 12.01 (1.48–97.26)) [61].

Food consumption practices. Consumption of meat or self-grown products is a potential risk factor for HEV in the general population. People that ingested self-grown food had higher
odds to have positive HEV than those who did not (OR 1.87 (1.41–2.48)) [71]. Consuming liver or other organ meats more than once per month (OR, 1.38 (1.01–1.88)) [82], or consuming meat more than 10 times per month increased the odds of being HEV positive [59]. Consumption of undercooked meat is related to the increased odds of being HEV positive in undergraduate and veterinary students (OR 12.9 (1.71–97.19)) [83].

In the immunocompromised group, fish consumption was a risk factor (OR 9.33 (2.07–42.2)) [84]. Moreover, a positive association between alcohol consumption and HEV seropositivity in immunocompromised patients (OR 3.43 (1.39–8.49)) [45] was reported.

**Rurality and low education.** Other risk factors for infection in the general population were living in rural communities (OR 1.63 (1.16–2.30)) [54], and having a low educational level (OR 2.08 (1.04–4.17) [54]; PR 1.76 (1.02–3.04) [85]). Dwelling in a rural settlement for more than 5 years was associated with HEV seropositivity (OR 3.4 (1.2–9.6)) [86].

For patients with viral hepatitis additional contact with pigs (OR 1.99) [63], having low education level, living in a rural area, or having poor sanitation practices were potential risks for HEV infection [41, 87, 88].

**Sanitation conditions.** In the general population, five studies found water and sanitation as a potential factors associated with HEV infection. Having a source of tap water lower the odds of having HEV (OR 0.78 (95% CI, 0.63–0.97)), while having any pet in the household (OR 1.19 (1.01–1.40), including dogs and cats), or having a dog in the household (OR 1.22 (1.04–1.43)) increases the odds [82]. Another study found higher HEV prevalence in people that have their water supply outside rather than inside their home (OR 3.15 (1.38–7.17)) [89].

In the rural population, consumption of untreated water (OR 1.92 (1.06–3.46)) and availability of water at home (OR 1.87 (1.07–3.27)) were factors associated to HEV [68]. For the immunocompromised group, piped water availability (OR 0.08 (0.01–0.66)) [90] decreases the odds of being HEV seropositive.

**Inner-country residence and travelling.** In the USA, blood donors from the Midwest were more likely to be antibody positive compared to the rest of the country (OR 2.23, (1.92–2.88)) [91]. In Brazil, the highest prevalence was observed in the Central (OR 6.93, (1.15–41.61)) and South-Central (6.28 (1.22–32.22)) among the demographic zones in the city of Sao Paulo (with Northeast as the reference) [55].

Fearon, MA et al. reported that Canadian blood donors with a history of living outside Canada (OR 2.9, (1.56–5.32)) or contact with farm animals (OR 1.5 (1.01–2.28)) were associated with HEV seropositivity [92]. Moreover, seropositive blood donors were more likely to have travelled to endemic regions than seronegative ones (OR 3.4, (1.0–14.5)) [93]. Another study also found that national trips within Mexico were associated with HEV exposure (OR 5.38 (1.02–28.16)) [94].

**Pregnancy.** Three studies have reported that HEV was associated with pregnancy (OR 3.5 (1.1–10.5)) [95], adjusting also for age and place of residence, or with having more than three pregnancies (OR 1.69 (1.04–2.75)) [96]. Moreover, Tissera, G et al. found an association between low age of a pregnancy (≤25 years old) and HEV seroprevalence (OR 3.2 (1.2–8.9)) [95].

**Transmission route.** The main route of transmission mentioned in the studies was fecal-oral; [69, 97] one study found blood transfusion history as a risk factor [68].

**Genotyping**

Twenty studies reported genotypes (Table 6, page 59 in S1 File). The most frequently studied genotypes in the Americas were 1 (Argentina, Venezuela, USA, Mexico, and Cuba) and 3
Fig 3. HEV genotype distribution in the Americas according to the included studies.

https://doi.org/10.1371/journal.pone.0269253.g003

(Arkentina, Brazil, Canada, Colombia, Venezuela, Bolivia, Mexico, USA, Cuba, and Uruguay) (Fig 3). Genotype 3 is more prevalent in this region and specifically the subtype 3a.

**Discussion**

Our systematic review and meta-analysis summarized the seroprevalence of the hepatitis E virus and its variation according to different serological markers, population groups, and regions. Anti-HEV IgG, which indicates past HEV infection, was the most studied antibody with an overall seroprevalence of 7.7% (95% CI 6.4%–9.2%). Nevertheless, this estimate is influenced by the composition of studied populations as reported in the literature, since the seroprevalence seems to differ in certain subpopulations. For example, our analysis found a higher anti-HEV IgG seroprevalence in people exposed to pigs (13.8% (7.6%–23.6%)), exposed population (11.4% (5.4%–22.6%)), and rural population (11.5% (5.4%–22.9%)). Risk factors for HEV prevalence that we identified from published data were age (highest in people over 30 years), contact with pigs and meat products, and some socioeconomic factors such as low sanitation access and low education level. Finally, we found predominance of genotype 1 and 3 in the American continent.
We found high variability in the seroprevalence estimates within regions of the Americas, countries, and within the considered population subgroups. A detailed inspection of the IgG seroprevalences reported from Brazil (Fig 13, page 72 in S1 File) suggests that this heterogeneity may be partly due to the different assays used. For example, the seroprevalences obtained using a test from Wantai tend to be higher than those obtained by a test from Abbott Laboratories. Pooling only results obtained by one test would yield in Brazil a seroprevalence of 15% (95% CI: 11%– 21%) in the former and of 4% (95% CI: 2%– 7%) in the latter case. This difference seems to be in line with the recently found low specificity (75%) for the Wantai test [98]. However, in our analysis, we do not correct for test characteristics, since the seroprevalence studies often do not report the numbers relevant for their investigations. Nevertheless, accounting for the test performance would be necessary to obtain more reliable seroprevalence estimates. Other studies also identified the employed assays as an important source of variability, together with the variance between regional areas, and the study design [99].

Our pooled seroprevalence estimate and the observed heterogeneity may be compared to findings from other regions. Data from Europe indicates that the anti-HEV IgG seroprevalence rate ranges from 0.6% to 52.5% with high heterogeneity [99], with an overall anti-HEV IgG seroprevalence of 19% (95% CI 14%–26%) [16]. As in our review, the European HEV seroprevalence is higher in individuals exposed to swine/wild animals and increases with age [99]. In the Middle East and in the Eastern Mediterranean region, the anti-HEV antibody IgG seroprevalence was 12.17% (95% CI 11.79%–12.57%), and 11.81% (95% CI 11.43%–12.21%), respectively [100]. In terms of general population, global estimates for anti-HEV IgG found a seroprevalence of 12.47% (95% CI 10.42% - 14.67%; I² = 100%) [13].

Considering differences between countries and regions of the Americas, the pooled seroprevalences were comparable, with perhaps the exception of Bolivia, Peru and Colombia showing a higher anti-HEV IgG seroprevalence (Fig 2). However, the overall number of studies available for these three countries was rather low and consequently not all subpopulations were represented in the results. Thus, the apparent higher seroprevalence does not have to indicate a higher background seroprevalence in these countries. A similar limitation applies to the formally observed difference between North and South America within the immunodeficiency subgroup. The difference may reflect the particular choices for the studied populations within each region, since the immunodeficiency subgroup has been defined rather broadly. Other studies have found differences in the estimated human HEV seroprevalence in industrialized countries [16] and between European countries [99].

In terms of the risk factors reported by the studies, we identified increasing age as being associated with higher prevalence through all the population groups, especially in those over 30 years. This suggests a lifelong cumulative exposure to the virus [15, 101, 102]. Ditah et al. had suggested that there is a higher probability of exposure to HEV in those people that live longer, and for that reason the prevalence increased with age [73]. In terms of aging, there are multiple factors that affect disease incidence and mortality such as young age at exposure, the accumulation over time of exposure, and the effect of aging on immune function, genomic instability, and other aging processes [103]. In chronic diseases mainly, the exposition to a certain risk factor occurs through adult life, childhood or prenatally, and its biological effects will be cumulative and possibly advanced at older ages [104].

Higher anti-HEV seropositivity is related to the occupational population with contact with pig or with pig products [13, 102, 105]. This is relevant not only in low- and middle-income countries, but also in high-income countries because it confirms its relationship with zoonotic transmission and contaminated food. It can be prevented with the implementation of medical and veterinary health strategies, surveillance and disease control activities, and safe food handling practices and procedures. People can also avoid the consumption of raw or undercooked...
products. Likewise, it will be important to perform studies in populations exposed to pigs and by assessing other potential risks and their contribution to HEV seroprevalence in those slaughterhouses and meat companies.

Based on our literature review, we conclude that more than ethnicity and country of birth or place of residence, the main risk factors are related to the socio-economic, education, and sanitary conditions that some populations have, especially those rural and indigenous communities where the fecal-oral transmission is more likely. Those areas that have compromised economic conditions have higher odds of exposure to animals and to contaminated surfaces such as soil or water [13]. Other authors have reported that there is evidence that premature deaths, having communicable diseases, or being affected by some pathogens is attributable to socioeconomic inequality [106–108]. In China for example, a higher prevalence of HEV in ethnic groups has been reported, especially in areas with mixed farming of domestic animals, or overpopulated and less developed areas [109, 110].

We documented that both genotype 1 and 3 are present in the Americas, which is consistent with the findings that fecal-oral transmission among humans as well as infections of zoo-notic origin play a role in the Americas. This also indicates that transmission routes in the Americas are apparently very similar to high-income countries of other continents where the genotype 3 predominates [9, 111, 112]. This supports the evidence that HEV strains circulating among swine and humans are closely genetically related, supporting as well the transmission of HEV through consumption of raw or undercooked meat products [4]. Genotype 3 is higher documented in the region and specifically the subtype 3a. One European study has mentioned that subtype 3a has a worldwide presence and can be found in samples from humans, pigs, and wild animals, and especially in American samples it could be an indigenous subtype [113].

Regarding other subtypes, 1a is more prevalent in Asia, and 1d is found in Africa [4]. Subtypes 3a and 3b circulate in the United States and Japan, and 3f and 3i circulate in Europe [4]. Although there are different HEV-3 subtypes, there is no clear evidence of the clinical significance of infection based on the subtype. For example, European studies have found that the risk of HEV-3-infected patients being hospitalized varied with the subtype and that patients infected with subtype 3c were at lower risk of hospitalization than those infected with subtypes 3f or 3e [114, 115].

Besides the seroprevalence reported from the general population, specific patient groups were studied for acute and recent HEV infections. The IgM seroprevalence was higher (Q-test, p = 0.006) in patients with confirmed or suspected viral hepatitis (pooled seroprevalence 5.5%, 95% CI: 2.0%–14.3%) when compared to the other subgroups (pooled seroprevalence 0.9%, 95% CI: 0.6%–1.5%). Similarly, in two studies an RNA positivity of 6.3% [48] and of 8.5% [49] was reported in acute viral hepatitis patients. This suggests that in suspected cases of acute hepatitis it would be important to look also for hepatitis E virus, as this agent can be the cause of the disease.

To our knowledge, this is the first systematic review that focused on the Americas and the seroprevalence of HEV. The strengths are the broad search of data and the inclusion of studies without restrictions, for example by language, as well as the critical and systematic assessment of available data. Our focus was on HEV in different subgroups and the analysis according to different serological markers such as total antibodies, IgG, IgM, or RNA. We also performed a robust statistical analysis considering multiple options in terms of proportions by region, population groups, and countries.

Aiming at including all relevant evidence available, the main limitation of our approach results from the heterogeneity of studies in terms of methods and effect size. As shown in the heterogeneity and in the publication bias analysis (Figs 1 and 2, pages 60–61 in S1 File), we found that there are important differences between the studies not only in the sample size but
also in the methods and analysis, and in the small study effect. We studied the reasons behind the high heterogeneity between the studies. Nevertheless, we could not explain it, but we suspect that it could be because: 1) difference between the assays and their specificity and sensitivity, 2) difference between the years of the study, 3) difference between countries and within the regions and cities from the countries. Therefore, due to this heterogeneity, we consider our approach more explorative, and we are aware that our findings are to some extent influenced by bias and error.

Our results give us some ideas to develop. We can highlight gaps in evidence in the area of acute hepatitis and pig related exposure. It is important to further investigate the causality of HEV in acute hepatitis cases and outbreaks and in those people that work with pigs or meat products, and to assess attributable fractions of HEV. We recommend including testing for HEV as a standardized practice in suspected hepatitis cases and outbreaks when financially possible. We also consider measuring the real risk of infection that people who work with pigs or meat products have.

In conclusion, the seroprevalence of HEV varies between certain population groups, serological markers, and within regions and countries of the Americas. The main risk factors involved in the infection are increasing age, contact with pig or pig products, and poor socioeconomic conditions. This is in line with the findings that genotype 1 and 3 are the two most prevalent genotypes in the Americas and further supports the transmission of HEV as zoonotic and as human fecal-oral origin. Our results underline that water sanitation, occupational health, and food safety must be priority components for the prevention of HEV infections.

Supporting information
S1 File. Supplementary material. (PDF)
S2 File. PRISMA checklist. (PDF)

Author Contributions
Conceptualization: Nathalie Verónica Fernández Villalobos, Barbora Kessel, Jórdis Jennifer Ott, Berit Lange.
Data curation: Nathalie Verónica Fernández Villalobos, Barbora Kessel, Isti Rodiah.
Formal analysis: Nathalie Verónica Fernández Villalobos, Barbora Kessel.
Investigation: Nathalie Verónica Fernández Villalobos.
Methodology: Nathalie Verónica Fernández Villalobos, Barbora Kessel.
Supervision: Jórdis Jennifer Ott, Berit Lange, Gérard Krause.
Validation: Barbora Kessel, Isti Rodiah.
Visualization: Nathalie Verónica Fernández Villalobos, Barbora Kessel.
Writing – original draft: Nathalie Verónica Fernández Villalobos.
Writing – review & editing: Nathalie Verónica Fernández Villalobos, Barbora Kessel, Isti Rodiah, Jórdis Jennifer Ott, Berit Lange, Gérard Krause.
References

1. Blasco-Perrin H, Abravanel F, Blasco-Baque V, Péron JM. Hepatitis E, the neglected one. Liver Int. 2016; 36: 130–134. https://doi.org/10.1111/liv.13014 PMID: 26725910

2. Rein DB, Stevens GA, Theaker J, Wittenborn JS, Wiersma ST. The global burden of hepatitis E virus genotypes 1 and 2 in 2005. Hepatology. 2012; 55: 988–997. https://doi.org/10.1002/hep.25505 PMID: 22121109

3. Institute of Health Metrics and Evaluation. Acute hepatitis E—Level 4 cause. 2019 [cited 1 Jul 2021] pp. 4–5. http://www.healthdata.org/results/gbd_summaries/2019/acute-hepatitis-e-level-4-cause

4. Kamar N, Dalton HR, Abravanel F, Izopet J. Hepatitis E virus infection. Clin Microbiol Rev. 2014; 27: 116–138. https://doi.org/10.1128/CMR.00057-13 PMID: 24396139

5. Kumar A, Beniwal M, Kar P, Sharma JB, Murthy NS. Hepatitis E in pregnancy. Int J Gynecol Obstet. 2004/05/18. 2004; 85: 240–244. https://doi.org/10.1016/j.ijgo.2003.11.018 PMID: 15145258

6. World Health Organization. Hepatitis E. 2021 [cited 8 Feb 2021]. https://www.who.int/news-room/fact-sheets/detail/hepatitis-e

7. Kamar N, Garrouste C, Haagsma EB, Garrigue V, Pischke S, Chauvet C, et al. Factors associated with chronic hepatitis in patients with hepatitis E virus infection who have received solid organ transplants. Gastroenterology. 2011/03/01. 2011; 140: 1481–1489. https://doi.org/10.1053/j.gastro.2011.02.050 PMID: 21354150

8. Bruni R, Villano U, Equestre M, Chionne P, Madonna E, Trandeva-Bankova D, et al. Hepatitis E virus genotypes and subgenotypes causing acute hepatitis, Bulgaria, 2013–2015. PLoS One. 2018/06/08. 2018; 13: e0198045. https://doi.org/10.1371/journal.pone.0198045 PMID: 29879148

9. Aspinall EJ, Couturier E, Faber M, Said B, Ijaz S, Tavoschi L, et al. Hepatitis E virus infection in Europe: Surveillance and descriptive epidemiology of confirmed cases, 2005 to 2015. Eurosurveillance. 2017; 22. https://doi.org/10.2807/1560-7917.ES.2017.22.26.30561 PMID: 28681720

10. Yazbek S, Kreidieh K, Ramia S. Hepatitis E virus in the countries of the Middle East and North Africa region: an awareness of an infectious threat to blood safety. Infection. 2016; 44: 11–22. https://doi.org/10.1007/s15010-015-0807-5 PMID: 26112744

11. Tengan FM, Figueiredo GM, Nunes AKS, Manchiero C, Dantas BP, Magri MC, et al. Seroprevalence of hepatitis E in adults in Brazil: A systematic review and meta-analysis. Infect Dis Poverty. 2019/01/17. 2019; 8: 3. https://doi.org/10.1186/s40249-018-0514-4 PMID: 30646964

12. Carrilho FJ, Mendes Clemente C, Da Silva LC. Epidemiology of hepatitis A and E virus infection in Brazil. Gastroenterol Hepatol. 2005/03/18. 2005; 28: 118–125. https://doi.org/10.1157/13072011 PMID: 15771857

13. Li P, Liu J, Li Y, Su J, Ma Z, Bramer WM, et al. The global epidemiology of hepatitis E virus infection: A systematic review and meta-analysis. Liver Int. 2020/04/14. 2020; 40: 1516–1528. https://doi.org/10.1111/liv.14468 PMID: 32281720

14. Wilhelm B, Waddell L, Greig J, Young I. Systematic review and meta-analysis of the seroprevalence of hepatitis E virus in the general population across non-endemic countries. PLoS One. 2019/06/08. 2019; 14: e0216826. https://doi.org/10.1371/journal.pone.0216826 PMID: 31173594

15. Wilhelm B, Waddell L, Greig J, Young I. A systematic review and meta-analysis of predictors of human hepatitis E virus exposure in non-endemic countries. Zoonoses Public Health. 2020/03/21. 2020; 67: 391–406. https://doi.org/10.1111/zph.12696 PMID: 32196945

16. Capai L, Falchi A, Charrel R. Meta-analysis of human IgG anti-HEV seroprevalence in industrialized countries and a review of literature. Viruses. 2019/01/24. 2019; 11. https://doi.org/10.3390/v11010084 PMID: 30695517

17. Verghese VP, Robinson JL. A Systematic Review of Hepatitis E Virus Infection in Children. Clin Infect Dis. 2014/05/23. 2014; 59: 689–697. https://doi.org/10.1093/cid/ciu371 PMID: 24846637

18. Tavakoli A, Alaviam SM, Moghoofe M, Mostafaei S, Abbasi S, Farahmand M. Seroepidemiology of hepatitis E virus infection in patients undergoing maintenance hemodialysis: Systematic review and meta-analysis. Ther Apher Dial. 2020/04/30. 2021; 25: 4–15. https://doi.org/10.1111/1744-9987.13507 PMID: 32348032

19. Bigna JJ, Modyinji AF, Nansseu JR, Amouougou MA, Nola M, Kenmoe S, et al. Burden of hepatitis E virus infection in pregnancy and maternalfetal outcomes: A systematic review and meta-analysis. BMC Pregnancy Childbirth. 2020/07/30. 2020; 20: 426. https://doi.org/10.1186/s12884-020-03116-2 PMID: 32723309

20. Moher D, Liberati A, Tetzlaff J, Altman DG. Preferred Reporting Items for Systematic Reviews and Meta-Analyses: The PRISMA Statement. PLoS Med. 2009; 6: e1000097. https://doi.org/10.1371/journal.pmed.1000097 PMID: 19621072
21. Wohlin C. Guidelines for snowballing in systematic literature studies and a replication in software engineering—EASE ’14. New York, New York, USA: ACM Press; 2014. pp. 1–10.

22. Elston DM. Mendele y. J Am Acad Dermatol. 2019; 81: 1071. https://doi.org/10.1016/j.jaad.2019.06.1291 PMID: 31279032

23. Langer BCA, Frösner GG, Von Brunn A. Epidemiological study of viral hepatitis types A, B, C, D and E among Inuits in West Greenland. J Viral Hepat. 1997/10/06. 1997; 4: 339–349. https://doi.org/10.1046/j.1365-2893.1997.00063.x PMID: 9310933

24. Sterne JAC, Hernán MA, Reeves BC, Savović J, Berkman ND, Viswanathan M, et al. ROBINS-I: a tool for assessing risk of bias in non-randomized studies of interventions. BMJ. 2016; 355.

25. Porto Tapiquén CE. Orogeñesis Soluciones Geográficas: “América.” Porlamar, Venezuela; 2015. http://tapiquen-sig.jimdo.com

26. Andy Bunn MK. A language and environment for statistical computing. R Foundation for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing; 2017. pp. 11–18. http://www.r-project.org

27. Valero-Mora PM. ggplot2: Elegant Graphics for Data Analysis. Journal of Statistical Software. Springer-Verlag New York; 2010. https://doi.org/10.18637/jss.v035.b01

28. Pebesma E. Simple Features for R: Standardized Support for Spatial Vector Data. R J. 2015; 67. https://doi.org/10.32614/RJ-2018-009

29. Balduzzi S, Rücker G, Schwarzer G. How to perform a meta-analysis with R: A practical tutorial. Evid Based Ment Health. 2019; 22: 153–160. https://doi.org/10.1136/ebmental-2019-300117 PMID: 31563865

30. Remondegui C, Ceballos S, Arce LP, Pintado E, Vidaurre R, Nitschko H, et al. Serologic evidence of the circulation of the hepatitis E virus and the prevalence of antibodies against hepatitis A in an indigenous population in northern Argentina. Rev Argent Microbiol. 2021/03/03. 2021; 53: 314–324. https://doi.org/10.1016/j.ram.2020.10.006 PMID: 33648797

31. Blatt S, Botelho TK, Dalmacco E. Sorología para el virus de la Hepatite E en gestantes: clinicamente importante o desnecesario? Rev bras anal clin. 2007; 39: 269–273. Available: https://pesquisa.bvsalud.org/porta l/resource/es/lil-490977

32. García CG, Sánchez D, Villalba MCM, Pujol FH, de los Ángeles Rodríguez Lay L, Pinto B, et al. Molecular characterization of hepatitis E virus in patients with acute hepatitis in Venezuela. J Med Virol. 2012/08/16. 2012; 84: 1025–1029. https://doi.org/10.1002/jmv.22377 PMID: 22985719

33. Hurtado H, Muñoz G, Braham B J. Anti-VHEIgM en casos de infección por el virus hepatitis E. Rev Med Chil. 2005/08/30. 2005; 133: 645–647. https://doi.org/10.4067/s0034-988720050000600004 PMID: 16075127

34. Peñáez-Carvajal D, Martínez-Vargas D, Escalante-Mora M, Palacios-Vivero M, Contreras-Gómez Lady, Coinfection of hepatitis E virus and other hepatitis virus in Colombia and its genotypic characterization. Biomedica. 2015/01/01. 2016; 36: 69–78.

35. Lay LDLAR, Quintana A, Villalba MCM, Lemos G, Corredor MB, Moreno AG, et al. Dual infection with hepatitis A and E viruses in outbreaks and in sporadic clinical cases: Cuba 1998–2003. J Med Virol. 2008/03/25. 2008; 80: 798–802. https://doi.org/10.1002/jmv.21147 PMID: 18360913
43. Lemos G, Jameel S, Panda S, Rivera L, Rodríguez L, Gavilondo J V. Hepatitis E virus in Cuba. J Clin Virol. 2000/02/19. 2000; 16: 71–75. https://doi.org/10.1016/s1386-6532(99)00062-1 PMID: 10680744

44. Villalba MCM, Lay LDLAR, Chandra V, Corredor MB, Frometa SS, Moreno AG, et al. Hepatitis E virus genotype 1, Cuba. Emerg Infect Dis. 2008/08/06. 2008; 14: 1320–1322. https://doi.org/10.3201/eid1408.080049 PMID: 18680671

45. Fantilli AC, Trinks J, Marciano S, Zárate F, Balderramo DC, Wassaf MGM, et al. Unexpected high seroprevalence of hepatitis E virus in patients with alcohol-related cirrhosis. PLoS One. 2019/10/28. 2019; 14: e0224404. https://doi.org/10.1371/journal.pone.0224404 PMID: 31648288

46. Hering T, Passos AM, Perez RM, Bilar J, Fragano D, Granato C, et al. Past and current hepatitis E virus infection in renal transplant patients. J Med Virol. 2014/03/13. 2014; 86: 948–953. https://doi.org/10.1002/jmv.23915 PMID: 24619955

47. Passos AM, Heringer TP, Medina-Pestana JO, Ferraz MLG, Granato CFH. First report and molecular characterization of hepatitis E virus infection in renal transplant recipients in Brazil. J Med Virol. 2013/02/06. 2013; 85: 615–619. https://doi.org/10.1002/jmv.23494 PMID: 23382036

48. Munné MS, Altabert NR, Otegui M LO, Vladimirskey SN, Moreiro R, Espul MP, et al. Updating the knowledge of hepatitis E: New variants and higher prevalence of anti-HEV in Argentina. Ann Hepatol. 2014/08/26. 2014; 13: 496–502. https://doi.org/10.1016/s1665-2681(19)31248-7 PMID: 25152981

49. Montalvo-Villalba MC, Castellano-Girones Y, Bello-Corredor M, de los Á Rodríguez-Lay L. Detección del genotipo 1d del virus de la hepatitis E en pacientes con sospecha de hepatitis viral aguda, Cuba 2013. Rev bioméd. 2016; 77: 25–73.

50. Cangin C, Focht B, Harris R, Strunk JA. Hepatitis E seroprevalence in the United States: Results for immunoglobulins IGG and IGM. J Med Virol. 2018/09/01. 2019; 91: 124–131. https://doi.org/10.1002/jmv.25299 PMID: 30168589

51. Bartoloni A, Bartalesi F, Roselli M, Mantella A, Arce CC, Paradisi F, et al. Prevalence of antibodies against hepatitis A and E viruses among rural populations of the Chaco region, southern-eastern Bolivia. Rev Panam Salud Publica. 2019/10/28. 2019; 40: 251–254. https://doi.org/10.1128/jcm.35.5.1244-1247.1997 PMID: 10427876

52. Passos-Castilho AM, De Sena A, Granato CFH. High prevalence of hepatitis E virus antibodies in a population of recyclable waste pickers in Brazil. J Med Virol. 2014/09/28. 2014; 86: 948–953. https://doi.org/10.1002/jmv.23915 PMID: 24619955

53. Passos-Castilho AM, de Sena A, Granato CFH. High prevalence of hepatitis E antibodies in a population of recyclable waste pickers in Brazil. J Med Virol. 2015/07/28. 2016; 88: 361–364. https://doi.org/10.1002/jmv.24336 PMID: 26211918

54. Villalba MCM, Guan M, Pérez A, Corredor MB, Frometa SS, Moreno AG, et al. Seroprevalence of antibodies to hepatitis E virus in two large communities in Havana, Cuba. Trans R Soc Trop Med Hyg. 2010/08/28. 2010; 104: 772–776. https://doi.org/10.1016/j.trstmh.2010.08.006 PMID: 20870260

55. Alvarez-Muñoz MT, Torres J, Damasio L, Gómez A, Tapia-Conyer R, Muñoz O. Seroepidemiology of hepatitis E virus infection in Mexican subjects 1 to 29 years of age. Arch Med Res. 1999/07/31. 1999; 30: 241–254. https://doi.org/10.1002/(SICI)1098-1161(199909)30:3251-254.10427876

56. Passos-Castilho AM, Reinaldo MR, de Sena A, Granato CFH. High prevalence of hepatitis E virus antibodies in Sao Paulo, Southeastern Brazil: analysis of a group of blood donors representative of the general population. Brazilian J Infect Dis. 2017; 21: 535–539. https://doi.org/10.1016/j.bjid.2017.05.004 PMID: 28606414

57. Passos-Castilho AM, de Sena A, Geraldo A, Spada C, Granato CFH. High prevalence of hepatitis E virus antibodies among blood donors in Southern Brazil. J Med Virol. 2015/07/28. 2016; 88: 361–364. https://doi.org/10.1002/jmv.24336 PMID: 26211918

58. Mahajan R, Collier MG, Kamili S, Drobeniuc J, Garfein RS, et al. Hepatitis E virus among persons who inject drugs, San Diego, California, USA, 2009–2010. Emerg Infect Dis. 2013/09/21. 2013; 19: 1664–1666. https://doi.org/10.3201/eid1910.130630 PMID: 24047923

59. Thomas DL, Yarbough PO, Vlahov D, Tsarev SA, Nelson KE, Saah AJ, et al. Seroreactivity to hepatitis E virus in areas where the disease is not endemic. J Clin Microbiol. 1997/05/31. 1997; 35: 1244–1247. https://doi.org/10.1128/jcm.35.5.1244-1247.1997 PMID: 9114415

60. Martins RMB, Freitas NR, Kozlowski A, Reis NRS, Lopes CLR, Teles SA, et al. Seroprevalence of hepatitis E antibodies in a population of recyclable waste pickers in Brazil. J Clin Virol. 2014/02/01. 2014; 59: 188–191. https://doi.org/10.1016/j.jcv.2014.01.002 PMID: 24480723

61. Alvarado-Esquivel C, Gutierrez-Martinez VD, Ramírez-Valles EG, Sifuentes-Alvarez A. Hepatitis E virus infection and waste pickers: A case-control seroprevalence study. J Med Virol. 2021; 93: 3779–3785. https://doi.org/10.1002/jmv.26688 PMID: 33230851
62. Alvarado-Esquivel C, Sanchez-Anguiano LF, Hernandez-Tinoco J. Seroepidemiology of Hepatitis E Virus Infection in Merchmates in Mexico. J Clin Med Res. 2014/12/02. 2015; 7: 103–108. https://doi.org/10.14740/jocmr1993w PMID: 25436027

63. Bricks G, Senise JF, Pott H, Junior, Grandi G, Passarini A, Caldeira DB, et al. Seroprevalence of hepatitis E virus in chronic hepatitis C in Brazil. Brazilian J Infect Dis. 2018; 22: 85–91. https://doi.org/10.1016/bjid.2018.02.001 PMID: 29494796

64. McGivern DR, Lin HHS, Wang J, Benzine T, Janssen HLA, Khalili M, et al. Prevalence and impact of hepatitis E virus infection among persons with chronic hepatitis B living in the US and Canada. Open Forum Infect Dis. 2019/05/30. 2019; 6: ofz175. https://doi.org/10.1093/ofid/ofz175 PMID: 3139669

65. Unzueta A, Valdez R, Chang YHH, Desmarteur YM, Heilman RL, Scott RL, et al. Hepatitis E virus serum antibodies and RNA prevalence in patients evaluated for heart and kidney transplantation. Ann Hepatol. 2015/12/03. 2016; 15: 33–40. https://doi.org/10.5604/16652681.1184202 PMID: 26626638

66. Altvig M, Shire NJ, Barrett A, Rouster SD, Sherman KE, Shata MT. Hepatitis E virus antibodies in patients with chronic liver disease. Emerg Infect Dis. 2009/02/26. 2009; 15: 479–481. https://doi.org/10.3201/eid1503.080740 PMID: 19239770

67. Vitral CL, da Silva-Nunes M, Pinto MA, de Oliveira JM, Gaspar AMC, Pereira RCC, et al. Hepatitis A and E seroprevalence and associated risk factors: A community-based cross-sectional survey in rural Amazonia. BMC Infect Dis. 2014/08/26. 2014; 14: 458. https://doi.org/10.1186/1471-2334-14-458 PMID: 25149658

68. Alvarado-Esquivel C, Sanchez-Anguiano LF, Hernandez-Tinoco J. Seroepidemiology of hepatitis E virus infection in general population in rural Durango, Mexico. Hepat Mon. 2014/07/01. 2014; 14:e16876. https://doi.org/10.5812/hepatmon.16876 PMID: 24976837

69. Campolmi I, Spinicci M, Mayaregua DR, Barahona HG, Mantella A, Lara Y, et al. Seroprevalence of hepatitis A, hepatitis E virus, and helicobacter pylori in rural communities of the Bolivian Chaco, 2013. Am J Trop Med Hyg. 2018/03/28. 2018; 98: 1275–1280. https://doi.org/10.4269/ajtmh.17-0747 PMID: 29582736

70. Dell’Amico MC, Cavallo A, Gonzales JL, Bonelli SI, Valda Y, Pieri A, et al. Hepatitis E virus genotype 3 in humans and swine, Bolivia. Emerg Infect Dis. 2011/08/02. 2011; 17: 1488–1490. https://doi.org/10.3201/eid1708.100769 PMID: 22801633

71. Diehl TM, Adams DJ, Nyland CM. Ingesting self-grown produce and seropositivity for hepatitis E in the United States. Gastroenterol Res Pract. 2018/08/18. 2018; 2018: 7980413. https://doi.org/10.1155/2018/7980413. PMID: 30116267

72. Moraes Dos Santos DC, Souto FJD, Lopes Dos Santos DR, Vitral CL, Gaspar AMC. Seroepidemiological markers of enterically transmitted viral hepatitis A and E in individuals living in a community located in the north area of Rio de Janeiro, RJ, Brazil. Mem Inst Oswaldo Cruz. 2002/09/10. 2002; 97: 637–640. https://doi.org/10.1590/s0074-02762002000500007 PMID: 12219125

73. Ditah I, Ditah F, Devaki P, Ditah C, Kamath PS, Charlton M. Current epidemiology of hepatitis E virus infection in the United States: Low seroprevalence in the National Health and Nutrition Evaluation Survey. Hepatology. 2014/05/16. 2014; 60: 815–822. https://doi.org/10.1002/hep.27219 PMID: 24824965

74. Campbell C. Seroprevalence and Characteristics of Hepatitis E in the United States. ProQuest Dissertations and Theses. 2019. http://ezproxy.unal.edu.co/dissertations-theses/servlet/characteristics-hepatitis-e-united/docview/2320963166/servlet-accountid=137090

75. Miernyk KM, Bruden D, Parkinson AJ, Hurlburt D, Klejka J, Berner J, et al. Human Seroprevalence to 11 Zoonotic Pathogens in the U.S. Arctic, Alaska. Vector-Borne Zoonotic Dis. 2019/02/26. 2019; 19: 563–575. https://doi.org/10.1089/vbz.2018.2390 PMID: 30789314

76. Talarmin A, Kazanjii M, Cardoso T, Poulikken JF, Sankale-Suzannon J, Sarthou JL. Prevalence of antibodies to hepatitis A, C, and E viruses in different ethnic groups in French Guiana. J Med Virol. 1997/08/01. 1997; 52: 430–435. PMID: 9260693

77. Debes JD, Pas SD, Groothuismink ZMA, van der Ende ME, de Man RA, Boonstra A. A mutation in the progesterone receptor predisposes to HEV infection in HIV-positive patients. Liver Int. 2017/12/30. 2018; 38: 792–796. https://doi.org/10.1111/liv.13678 PMID: 29285885

78. Sue PK, Pisanic N, Heaney CD, Forman M, Valsamakis A, Jackson AM, et al. Hepatitis E virus infection among solid organ transplant recipients at a North American transplant center. Open Forum Infect Dis. 2016/03/26. 2016; 3: ofw006. https://doi.org/10.1093/ofid/ofw006 PMID: 27047170

79. Gutiérrez-Vergara C, Rodrízquez B, Parra-Suescun J, Correa-Londoño G, López-López L, López-Herrera A, et al. Determinación de anticuerpos totales (IgG/IgM) y específicos (IgM) para el virus de la hepatitis E y detección molecular del virus en heces de humanos con o sin exposición ocupacional a porcinos en 10 municipios de Antioquia. Iatreia. 2015; 28: 248–258. https://doi.org/10.17533/udea.iatreia.v28n3a03
80. Meng XJ, Wiseman B, Elvinger F, Guenette DK, Toth TE, Engle RE, et al. Prevalence of antibodies to hepatitis E virus in veterinarians working with swine and in normal blood donors in the United States and other countries. J Clin Microbiol. 2002/01/05. 2002; 40: 117–122. https://doi.org/10.1128/JCM.40.1.117-122.2002 PMID: 11773103

81. Huerta Lorenzo B, Tarradas-Iglesias C, González M, Peralta B, Mateu de Antonio EM, Astorga Márquez RJ, et al. Seroprevalencia y factores asociados a la infección por el virus de la hepatitis E (Genotipo 3) en perros del sur de España. An la Real Acad Ciencias Vet Andalucía. 2011; 24: 135–144. Available: http://repebis.upb.edu.pe/articulos/dia/v52n4/a4.pdf

82. Kuniholm MH, Purell RH, McQuillan GM, Engle RE, Wasley A, Nelson KE. Epidemiology of hepatitis E virus in the United States: Results from the third national health and nutrition examination survey, 1988–1994. J Infect Dis. 2009/05/29. 2009; 200: 48–56. https://doi.org/10.1086/599319 PMID: 19473098

83. Cossaboom CM, Heffron CL, Cao D, Yugo DM, Houk-Miles AE, Lindsay DS, et al. Risk factors and sources of foodborne hepatitis E virus infection in the United States. J Med Virol. 2016/02/19. 2016; 88: 1641–1645. https://doi.org/10.1002/jmv.24497 PMID: 26889628

84. Pisano MB, Balderramo D, Wassaf MM, Lotto M, Carlino Y, Ré VE, et al. Hepatitis E virus infection in patients on dialysis and in solid organ transplant recipients in Argentina: exploring associated risk factors. Arch Virol. 2016/11/26. 2017; 162: 787–792. https://doi.org/10.1007/s00705-016-3171-6 PMID: 27885564

85. de Almeida e Araújo DC, de Oliveira JM, Haddad SK, da Roza DL, de O Bottino F, Faria SBSC, et al. Hepatitis E virus infection in pregnant women, Argentina. BMC Infect Dis. 2020/05/26. 2020; 20: 368. https://doi.org/10.1186/s12879-020-05087-3 PMID: 32448132

86. De Freitas NR, Teles SA, Caetano KAA, De Matos MA, Carneiro MADS, Gardinali NR, et al. Hepatitis E virus infection in patients with acute non-A, non-B, non-C hepatitis in Central Brazil. Mem Inst Oswaldo Cruz. 2016/10/21. 2016; 111: 692–696. https://doi.org/10.1590/0074-02760160256 PMID: 27759769

87. Munne MS, Altabert NR, Vladimirsky SN, Moreiro R, Mares LOO, Soto SS, et al. Identifications of poly-phyletic variants in acute hepatitis suggest an underdiagnosed circulation of hepatitis E virus in Argentina. J Clin Virol. 2011/07/29. 2011; 52: 138–141. https://doi.org/10.1016/j.jcv.2011.06.015 PMID: 21795113

88. Perez OM, Morales W, Paniagua M, Strannegard O. Prevalence of antibodies to hepatitis A, B, C, and E viruses in a healthy population in Leon, Nicaragua. Am J Trop Med Hyg. 1996/07/01. 1996; 55: 17–21. https://doi.org/10.4269/ajtmh.1996.55.1.tm0550010017

89. Bezerra LA, de Oliveira-Filho EF, Silva JVJ, Santos Morais VM, Gonçalves JP, da Silva DM, et al. Risk analysis and seroprevalence of HEV in people living with HIV/AIDS in Brazil. Acta Trop. 2018/10/08. 2019; 189: 65–68. https://doi.org/10.1016/j.actatropica.2018.09.026 PMID: 30292751

90. Stramer SL, Moritz ED, Foster GA, Ong E, Linnen JM, Hogema BM, et al. Hepatitis E virus: Seroprevalence and frequency of viral RNA detection among US blood donors. Transfusion. 2015/10/06. 2016; 56: 481–488. https://doi.org/10.1111/trf.13355 PMID: 26434952

91. Fearon MA, O’Brien SF, Delage G, Scalia V, Bernier F, Bingham M, et al. Hepatitis E virus in a Canadian blood donor population. Transfusion. 2017/04/11. 2017; 57: 1420–1425. https://doi.org/10.1111/trf.14089 PMID: 28394029

92. Mast EE, Ken Kuramoto I, Favorov MO, Schoening VR, Burkholder BT, Shapiro CN, et al. Prevalence of and risk factors for antibody to hepatitis E virus seroreactivity among blood donors in Northern California. J Infect Dis. 1997/07/01. 1997; 176: 34–40. https://doi.org/10.1086/514037 PMID: 9207347

93. Alvarado-Esquivel C, Gutierrez-Martinez VD, Ramirez-Valles EG, Sifuentes-Alvarez A. Hepatitis E Virus Infection and Butchers: A Case-Control Seroprevalence Study. Gastroenterol Res. 2021/05/20. 2021; 14: 96–103. https://doi.org/10.14740/gr1198 PMID: 34007351

94. Tissera G, Lardizabal MC, Torres SB, Fantilli AC, Martinez Wassaf MG, Venezuela F, et al. Hepatitis e virus infection in pregnant women. BMC Infect Dis. 2020/05/26. 2020; 20: 368. https://doi.org/10.1186/s12879-020-05087-3 PMID: 32448132

95. Hardtke S, Rocco R, Ogata J, Braga S, Barbosa M, Wranke A, et al. Risk factors and seroprevalence of hepatitis E evaluated in frozen-serum samples (2002–2003) of pregnant women compared with female blood donors in a Southern region of Brazil. J Med Virol. 2018/08/01. 2018; 90: 1856–1862. https://doi.org/10.1002/jmv.25274 PMID: 30063252
97. Pang L, Alencar FEC, Cerutti C, Milhous WK, Andreade AL, Oliveira R, et al. Short report: Hepatitis E infection in the Brazilian Amazon. Am J Trop Med Hyg. 1995/04/01. 1995; 52: 347–348. https://doi.org/10.4269/ajtmh.1995.52.347 PMID: 7741175

98. Zhang Q, Zong X, Li D, Lin J, Li L. Performance evaluation of different commercial serological kits for diagnosis of acute hepatitis e viral infection. Polish J Microbiol. 2020; 69: 217–222. https://doi.org/10.33073/pjm-2020-025 PMID: 32548990

99. Hart J, Otto B, Madden RG, Webb G, Woolson KL, Kristen L, et al. Hepatitis E seroprevalence in Europe: A meta-analysis. Viruses. 2016; 8: 211. https://doi.org/10.3390/v8080211 PMID: 27509518

100. Karbalai Niy MH, Rezaee-Zavareh MS, Ranaei A, Alavian SM. Hepatitis E virus seroprevalence rate among Eastern Mediterranean and middle eastern countries; A systematic review and pooled analysis. Microb Pathog. 2017; 110: 252–256. https://doi.org/10.1016/j.micpath.2017.06.045 PMID: 28688980

101. Mansuy JM, Bendali R, Legrand-Abravanel F, Sauné K, Miédouge M, Ellis V, et al. Hepatitis E virus antibodies in blood donors, France. Emerg Infect Dis. 2011/12/17. 2011; 17: 2309–2312. https://doi.org/10.3201/eid1712.110371 PMID: 22172156

102. Mansuy JM, Sauné K, Rech H, Abravanel F, Mengelle C, L’Homme S, et al. Seroprevalence in blood donors reveals widespread, multi-source exposure to hepatitis E virus, southern France, october 2011. Eurosurveillance. 2015/05/21. 2015; 20: 27–34. https://doi.org/10.2807/1560-7917.ES2015.20.19.21127 PMID: 25990359

103. Izumi S, Sakata R, Yamada M, Cologne J. Interaction between a single exposure and age in cohort-based hazard rate models impacted the statistical distribution of age at onset. J Clin Epidemiol. 2016; 71: 43–50. https://doi.org/10.1016/j.jclinepi.2015.10.004 PMID: 26475570

104. Kaplan GA, Haan MN, Wallace RB. Understanding changing risk factor associations with increasing age in adults. Annu Rev Public Health. 1999; 20: 89–108. https://doi.org/10.1146/annurev.publhealth.20.1.89 PMID: 10352851

105. Teixeira J, Mesquita JR, Pereira SS, Oliveira RMS, Abreu-Silva J, Rodrigues A, et al. Prevalence of hepatitis E virus antibodies in workers occupationally exposed to swine in Portugal. Med Microbiol Immunol. 2016/10/23. 2017; 206: 77–81. https://doi.org/10.1007/s00430-016-0484-8 PMID: 27770276

106. Dowd JB, Zajacova A, Aiello A. Early origins of health disparities: Burden of infection, health, and socioeconomic status in U.S. children. Soc Sci Med. 2009; 68: 699–707. https://doi.org/10.1016/j.socscimed.2008.12.010 PMID: 19152993

107. Greene SK, Levin-Recto A, Hadler JL, Fine AD. Disparities in Reportable Communicable Disease Incidence by Census Tract-Level Poverty, New York City, 2006–2013. Am J Public Health. 2015; 105: e27–e34. https://doi.org/10.2105/AJPH.2015.302741 PMID: 26180961

108. Lewer D, Jayatunga W, Aldridge RW, Edge C, Marmot M, Story A, et al. Premature mortality attributable to socioeconomic inequality in England between 2003 and 2018: an observational study. Lancet Public Health. 2020; 5: e33–e41. https://doi.org/10.1016/S2468-2667(19)30219-1 PMID: 31813773

109. Feng Y, Feng YM, Wang S, Xu F, Zhang X, Zhang C, et al. High seroprevalence of hepatitis E virus in the ethnic minority populations in Yunnan, China. Khudyakov YE, editor. PLoS One. 2018; 13: e0197577. https://doi.org/10.1371/journal.pone.0197577 PMID: 29787589

110. Li H, Zhang Y, Ma Z, Liu Z, Ikrabam A, Liu L, et al. HEV prevalence and potential risk factors in a large multi-ethnic youth cohort in China. Virol J. 2021; 18: 3. https://doi.org/10.1186/s12985-020-01470-3 PMID: 33407606

111. Nicot F, Dimiglio C, Miguere M, Jeanne N, Latour J, Abravanel F, et al. Classification of the Zoonotic Hepatitis E Virus Genotype 3 Into Distinct Subgenotypes. Front Microbiol. 2021; 11. https://doi.org/10.3389/fmicb.2020.634430 PMID: 33584599

112. Lin S, Zang Y-J. Advances in Hepatitis E Virus Biology and Pathogenesis. Viruses. 2021; 13: 267. https://doi.org/10.3390/v13020267 PMID: 33572257

113. Vina-Rodriguez A, Schlosser J, Becher D, Kaden V, Groschup MH, Eiden M. Hepatitis E virus genotype 3 diversity: Phylogenetic analysis and presence of subtype 3b in wild boar in Europe. Viruses. 2015; 7: 2704–2726. https://doi.org/10.3390/v7052704 PMID: 26008708

114. Subissi L, Peeters M, Lamoral S, Klamar S, Suin V, Van Gucht S. Subtype-specific differences in the risk of hospitalisation among patients infected with hepatitis E virus genotype 3 in Belgium, 2010–2018. Epidemiol Infect. 2019; 147: e224. https://doi.org/10.1017/S0950268819001122 PMID: 31364564

115. Abravanel F, Dimiglio C, Castanier M, Péron JM, Kamar N, Lhomme S, et al. Does HEV-3 subtype play a role in the severity of acute hepatitis E? Liver Int. 2020; 40: 333–337. https://doi.org/10.1111/liv.14329 PMID: 31837187