A case of synchronous multiple myeloma and chronic myeloid leukemia

TO THE EDITOR: Multiple myeloma (MM) is a hematologic malignancy caused by the proliferation of clonal plasma cells in the bone marrow, leading to uncontrolled production of monoclonal immunoglobulin. Chronic myeloid leukemia (CML) is a myeloproliferative neoplasm associated with the presence of the BCR-ABL1 fusion gene. These are uncommon malignancies that account for approximately 1.6% and 0.4% of all newly diagnosed cancers in the United States, respectively [1]. In Korea, these are also rare diseases and the crude incidence rates of those are 2.5/100,000 and 0.8/100,000, respectively [2, 3]. The concurrent diagnosis of MM and CML in one patient is an extremely rare event. In this report, we describe an additional case of synchronous MM and CML.

A 64-year-old man was referred to our hospital for evaluation of a right pleural soft-tissue mass and a sternal osteolytic lesion incidentally detected on chest computed tomography (CT). The patient had no specific symptoms and remarkable findings on physical examinations. A complete blood count showed a white blood cell counts of 13.1×10⁹/L, with a differential count of 65% neutrophils, 15% lymphocytes, 10% monocytes, 5.2% eosinophils, and 4% basophils; hemoglobin levels of 8.3 g/dL; and platelet counts of 234×10⁹/L.

Bone marrow biopsy revealed increased myeloid cell infiltration. Most of the infiltrated cells were positive for CD138 on immunohistochemical staining (Fig. 1). A patchy positive reaction for CD138 was compatible with myeloma involvement, and myeloid cells with varying maturation. A 64-year-old man was referred to our hospital for evaluation of a right pleural soft-tissue mass and a sternal osteolytic lesion incidentally detected on chest computed tomography (CT). The patient had no specific symptoms and remarkable findings on physical examinations. A complete blood count showed a white blood cell counts of 13.1×10⁹/L, with a differential count of 65% neutrophils, 15% lymphocytes, 10% monocytes, 5.2% eosinophils, and 4% basophils; hemoglobin levels of 8.3 g/dL; and platelet counts of 234×10⁹/L.

Blood chemistry showed reversal of the albumin-globulin ratio (total protein 11.8 g/dL, and albumin 2.3 g/dL), renal dysfunction (creatinine clearance 38.4 mL/min), and hypercalcemia (calcium 11.8 mg/dL). A total of 7 g/dL of monoclonal gammopathy (IgA lambda type) was detected with serum immunofixation electrophoresis. The ratio of serum kappa/lambda free light chains was 0.006 and serum β2 microglobulin level was 19.8 mg/L. Radiological investigation revealed multiple osteolytic lesions in the skull and axial skeleton. The needle biopsy of pleural soft-tissue mass showed infiltrations of monotonous round cells with eccentrically located nuclei and different sizes, but no evidence of myeloid cell infiltration. Most of the infiltrated cells were positive for CD138 on immunohistochemical staining (Fig. 1). A patchy positive reaction for CD138 was compatible with myeloma involvement, and myeloid cells were positive for myeloperoxidase. The bone marrow aspirate demonstrated myeloid hyperplasia with increased eosinophils, basophils and plasma cells. Plasma cells showed mature forms with condensed nuclear chromatin, indistinct nucleoli, and abundant basophilic cytoplasm with a perinuclear halo (Fig. 2).

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Bone marrow biopsy revealed increased cellularity (up to 90%), consisting of plasmacytoid round cell infiltrations and myeloid cells with varying maturation. A patchy positive reaction for CD138 was compatible with myeloma involvement, and myeloid cells were positive for myeloperoxidase. The bone marrow aspirate demonstrated myeloid hyperplasia with increased eosinophils, basophils and plasma cells. Plasma cells showed mature forms with condensed nuclear chromatin, indistinct nucleoli, and abundant basophilic cytoplasm with a perinuclear halo (Fig. 2). Chromosome analysis revealed 46,XY, t(2:3)(p15;q26), t(9:22)(q34;q11) in 20 of 21 cells, and fluorescent in situ

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hybridization (FISH) indicated $BCR/ABL$ translocations in 86.5% of cells. Molecular analysis revealed $BCR/ABL$ rearrangement based on results from reverse transcription polymerase chain reactions. Consequently, the patient was diagnosed with stage III MM and low-risk CML in accordance with the International Staging System [4] and Sokal scores [5], respectively. Considering the patient’s old age and toxicity of combination therapy for both CML and MM, the patient was first treated for high-stage MM. Treatment was initiated with a regimen of thalidomide (100 mg daily) and dexamethasone (40 mg on days 1–4 and 15–18) every 4 weeks. However, the response to treatment was not evaluable, since the patient died of pneumonia caused by carbapenem-resistant *Acinetobacter* after 4 weeks.

Ide et al. [6] reviewed 12 cases in which MM and CML coexisted. In four of the cases, patients were diagnosed first with MM followed by CML, while in four other cases, MM developed after CML. The interval between the two types of cancer ranged from 14 months to 113 months. In the remaining four cases, MM and CML were diagnosed simultaneously.

Although there are no established treatment regimens for simultaneously occurring MM and CML, there are two cases in which MM with CML was treated with a combination of bortezomib, dexamethasone, or lenalidomide, and imatinib or dasatinib [7, 8]. The combination treatment targeting both MM and CML resulted in successful outcomes in these cases. Our patient received only anti-MM therapy owing to old age and overall poor health status. Although data from a small study suggest that thalidomide treatment along with imatinib is efficacious for the treatment of CML [9], our patient did not respond to thalidomide treatment.
We presented our experience with a patient who was diagnosed with MM and CML simultaneously. Evaluation of other cases is required to shed light on clinical characteristics of the disease states, as well as to explore potential evaluable treatments.

TO THE EDITOR: Primary cardiac amyloidosis accompanying heart failure, angina, and/or arrhythmia is very serious and has a poor prognosis [1]. Sequential heart and autologous stem cell transplantation has resulted in some promising outcomes in a few series [2-4]. We present a case of primary amyloidosis with cardiac involvement that was successfully managed with these combined approaches.

Case
A 62-year-old woman was referred to our clinic with 3 months of dyspnea on exertion; she was categorized in New York Heart Association class III, and had abnormal echocardiographic findings. She had no other medical history of note. On initial physical examination, her vital signs were as follows: blood pressure, 88/45 mmHg; pulse rate, 79 beats/min; and body temperature, 36.6°C. Neck vein engorgement and pretrivial pitting edema were noted. Heart and lung sounds on auscultation were normal. Initial laboratory tests were as follows: white blood cell count, 5,600/μL; hemoglobin, 12.4 g/dL; platelet count, 150,000/μL; protein, 6.0 g/dL; albumin, 3.6 g/dL; blood urea nitrogen, 21 mg/dL; serum creatinine, 1.33 mg/dL; aspartate aminotransferase, 19 IU/L; alanine aminotransferase, 20 IU/L; alkaline phosphatase, 128 IU/L; troponin I, 0.041 ng/mL; and brain natriuretic peptide (BNP), 629 pg/mL. There were no abnormal findings on urinalysis.

A chest radiograph revealed cardiomegaly with a cardiothoracic ratio of 0.7 and increased interstitial markings suggesting pulmonary edema. Both costophrenic angles were blunted with bilateral pleural effusion. Electrocardiography displayed low voltage in leads I, II, and III and T-wave inversion in leads V5 and V6. Transthoracic echocardiography revealed thickened ventricle walls with minimal pericardial effusion and impaired diastolic function. Left ventricle (LV) filling pressure was high, with an E/E’ of 37. No regional wall motion abnormality was observed and the LV ejection fraction was 59%. Cardiac magnetic resonance imaging indicated diffuse transmural or subendocardial enhancement at both ventricular walls on a delayed enhancement image, which were consistent with cardiac amyloidosis (Fig. 1A). Endomyocardial biopsy with a femoral venous approach was performed to confirm this diagnosis. On pathologic examination, amyloid deposits were confirmed by Congo-red staining. Immunohistochemical staining results were as follows: prealbumin (+); kappa chain (+); lambda chain (-); and amyloid A (-) (Fig. 1B, C). Although paraproteinemia or Bence-Jones proteinuria were not evident by electrophoresis and immunofixation, the patient’s serum free light-chain ratio was increased to 114 (kappa, 2,040.0 mg/L; lambda, 17.9 mg/L). A bone mar-

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Sequential heart and autologous stem cell transplantation for light-chain cardiac amyloidosis

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