Molecular typing of *Staphylococcus aureus* isolate responsible for staphylococcal poisoning incident in home-made food

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Abstract

In October 2012, two persons fell ill with symptoms consistent with staphylococcal food poisoning after eating home-canned tuna fish and tomatoes. Laboratory investigation detected the enterotoxins in the home-canned tuna and molecular analysis of the isolated *Staphylococcus aureus* confirmed it carried toxin genes. Qualitative enzyme-linked immunosorbent assay and enzyme linked fluorescent assay methods and quantitative assay identified the enterotoxins in the food leftovers, specifically staphylococcal enterotoxins type A (SEA) and D (SED), respectively 0.49 and 2.04 ng/g. The laboratory results are discussed considering the relation to the fish in oil, survival and heat resistance of *S. aureus*, and presumptive microbial contamination due to improper handling during home-canning procedures. This is the first reported cluster of foodborne illnesses due to staphylococcal enterotoxins in tuna in Italy. In this study, we reported cases described and analysed for their *spa*-type. Showing a high heterogeneity of isolates, *spa*-type r13252 is correlated in a node of the minimum spanning tree and it has never been reported as responsible for foodborne outbreak. This case underlines the importance of risk communication and dissemination of home-canning guidelines to reduce the incidence of foodborne outbreaks caused by homemade conserves.

Introduction

Staphylococcal food poisoning (SFP), a frequent following of foodborne diseases worldwide, occurs following the ingestion of staphylococcal enterotoxins (SEs) produced by enterotoxigenic strains of coagulase-positive staphylococci, particularly *Staphylococcus aureus* (Bianchi et al., 2014). In 2012, SEs were responsible for 346 foodborne outbreaks (FBOs) in the European Union, representing 6.4% of all outbreaks reported in the EU. These figures are consistent with those of the previous year, 2011, when 345 outbreaks were reported. Foodborne outbreaks due to SEs are frequently reported by mixed foods (31.4%), followed by cheese (20.0%), broiler meat and products thereof (8.6%), pig meat and products thereof (5.7%); while, to our knowledge, fish and fishery products have not been reported as source of SFP (EFSA, 2013, 2014). The incubation period varies depending on the amount of toxins ingested and individual susceptibility. The duration of illness is short and often self-limited. SFP results from the ingestion of improperly prepared or stored foods in which toxins produced by *S. aureus* (>5 log CFU/g) are present. In order to type *S. aureus* responsible for FBO, different techniques are established; the most widely used method for this type of epidemiological investigations is *spa*-typing, based on the determination of the polymorphic X region of the gene encoding staphylococcal protein A (*spa*). In this study, we report the results of the investigation of a foodborne outbreak caused by SEs in contaminated home-canned tuna fish occurred in October 2012 in Catanzaro and we compare the strain isolated in 32 strains responsible for other outbreaks published, with the aim of determining the relationship between *S. aureus* isolated strains in FBO cases.

Materials and Methods

Outbreak investigation and sampling

In the middle of October 2012, four members of a family (parents and two daughters aged 11 and 16 years) ate home-canned tuna fish preserved in olive oil, which had been received as a gift from a friend. About 3 hours later, the two girls developed nausea, abdominal cramps, vomiting and diarrhea. They were taken to the emergency room of a local hospital for diagnosis and treatment where they were subsequently admitted to the Pediatrics Department for observation. The hospital did not collect stool or vomit samples; however, it notified the surveillance system for foodborne outbreaks which, jointly with the local veterinary and public health services, collected samples of the leftovers.

Enumeration of coagulase-positive staphylococci, phenotypic characterisation of *Staphylococcus aureus* and detection of staphylococcal enterotoxins in food samples

A food sample was sent to the Institute for Experimental Veterinary Medicine of Southern Italy, Catanzaro. Laboratory analysis of the home-canned tuna included isolation and identification of coagulase-positive staphylococci strains, screening for SEs by means of VIDAS SET2 (BioMérieux, Craponne, France) and for *Clostridium botulinum* and its neurotoxins (CDC, 1998). The VIDAS SET2 is an automatic qualitative test performed on VIDAS instruments and is based on an enzyme linked fluorescent assay (ELFA) technique for the detection of classical SEs [*staphylococcal enterotoxins type A* to type E (*SEA to SEE*)] in food. A food sample was sent to the Italian National Reference Laboratory for coagulase-positive staphylococci (CPS), including *S. aureus* (ITNRL-CPS) in Turin, where confirmatory analyses of SEs were performed according...
to the European Screening Method of the European Reference Laboratory (EU-RL) for CPS-ver. 5 (Ostyn et al., 2010b). The EU-RL for CPS quantified the SEs (types SEA to SEE) using a quantitative enzyme-linked immunosorbent assay (ELISA) (Ostyn et al., 2010a). Coagulase-positive staphylococci were counted in sample using the standard method EN ISO 6888 part 2 (ISO, 1999). Coagulase-positive staphylococci isolate were confirmed by real time using the TaqMan® Staphylococcus aureus detection kit (Applied Biosystems, Foster City, CA, USA) and biotyped as described by Devriese (1984).

Polymerase chain reactions and detection of virulence genes
Polymerase chain reaction assays were carried out on the isolated strains to detect the genes encoding SEs (Ostyn et al., 2010a), methicillin resistance (mecA), and Panton-Valentine Leukocidin (pel) (Lamand et al., 2012). In addition the polymorphic X region of the protein A, gene spa was sequenced to assign the spa-type (Harmsen et al., 2003).

Cluster analysis of isolates spa-type involved in foodborne poisoning
Since 2009 in Europe spa-types of isolates involved in cases of SFP were reported. Thirty-two isolates were analyzed and used for cluster analyses of spa sequences, including the isolates induced the poisoning described in this work and 15 strains related to an additional case (SFP ID 31) (Table 1). The sequences were compared and aligned using an algorithm-based model for pairwise alignment of repeats (default cost matrix in Bionumerics), used to make a minimum spanning tree (MST) setting the bin grouping distance at 1.00%.

Results
Investigation and epidemiological information
The two girls developed symptoms about 3 hours after eating the meal. Food history interviews with the family yielded that during the 8 hours preceding symptoms onset the girls had eaten only one meal (consisting of tomatoes and the home-canned tuna fish). Unopened jars were not present when local authorities went to the cooker’s house; they had already been eaten or distributed as gifts. Given the rapid onset of symptoms and considering the characteristics of the food preparation process, the home-canned tuna was suspected as the intoxication cause, although nobody else was reported to be ill, probably due to a non-homogeneous meat contamination among the jars.

Scombroid poisoning was initially considered a likely cause of the outbreak because this type of poisoning with a short incubation period is most frequently associated with tuna and similar fish. The scombroid poisoning, or more accurately defined as histamine food poisoning, can occur after the consumption of spoiled tuna fish where bacteria metabolise histidine into histamine when fish is not kept at refrigerator temperatures (Feng et al., 2016). The symptoms of histamine food poisoning include tongue dysesthesias, face and torso flushing, and a throbbing headache; it often progresses to diarrhea, nausea and vomiting (Wilson et al., 2012; McCarthy et al., 2015). In this case, no allergic reactions were noted, and the gastrointestinal complaints developed about 3 hours after the meal. As the course of symptoms of histamine food poisoning is rapid, within minutes to 30 min, it did not seem a likely cause and histamine levels were not measured.

In 2011, the majority of strong evidence out-
# Table 1. Report of *Staphylococcus aureus* isolated.

| ID   | SFP ID | Origin                  | Detail                                      | spa-type | Phenotypic characteristics | Genotypic characteristics | Geographical ID | Suspected food spreading agent | Causative agent | People involved | References               |
|------|--------|-------------------------|---------------------------------------------|----------|---------------------------|--------------------------|----------------|-------------------------------|-----------------|-----------------|---------------------------|
| 1    | 1      | Human                   | Case, stool                                 | t127     | SEA+                      | sea, seh, seh, seh       | Freiburg (Germany) | Ice-cream                   | Ice-cream        | 20 patients    | (Fetsch et al., 2014) |
| 2    | 2      | Human                   | Case, stool                                 | t127     | SEA+                      | sea, seh, seh, seh       | Turin (Italy)     | Food handler                 | Seafood salad    | 9 patients     | (Gallina et al., 2013)  |
| 3    | 3      | Human                   | Nasal swab                                  | t160     | SED+                      | sea, seh                | Baden-Wurttemberg (Southwest Germany) | Food handler | Food handler                 | 12 patients     | (Johler et al., 2013)  |
| 4    | 4      | Human                   | Nasal mucosa of food handler #5             | t701     | /                         | sea, sei                | Children and staff members of a Swiss boarding school (Switzerland) | Seafood salad | 14 patients     | (Johler et al., 2015)  |
| 5    | 5      | Human                   | Nasal swab of caterer’s nose #1             | t018     | SEA+                      | sea, seh, seh            | Austria           | Raw milk                     | 80 patients      | 40 patients     | (Schmid et al., 2007)  |
| 6    | 6      | Human                   | Nasal swab of caterer’s nose #2             | t018     | SEA+                      | sea, seh, seh            | Italy             | Milk                         | 80 patients      | This work       |                           |
| 7    | 7      | Human                   | Nasal swab                                  | t018     | SEA+                      | sea, seh, seh            | Italy             | Home-cooked tuna fish        | 2 patients       | This work       |                           |
| 8    | 8      | Human                   | Nasal swab of caterer’s nose #3             | t018     | SEA+                      | sea, seh, seh            | Italy             | Home-cooked tuna fish        | 2 patients       | This work       |                           |
breaks in the EU were caused by foodstuffs of animal origin. Fish and fish products were implicated in 10.1% of all outbreaks. Of the 25 SEs outbreaks, 18 were general outbreaks and seven were household outbreaks (EFSA 2013). Generally, 31.4% of foodborne outbreaks are due to food prepared in domestic kitchens and 28.6% in a commercial kitchen or other licensed facilities (EFSA, 2014). As in previous years, in 2012 foodstuffs of animal origin reach the majority of the strong evidence outbreaks, and the most common single foodstuff category reported as food vehicle was eggs and egg products, responsible for 168 outbreaks (22.0%) (EFSA, 2014). Fish and fish products were responsible for the 9.2% of all cases. The type of outbreak was provided for 34 cases: 26 (76.5%) were general outbreaks and eight (23.5%) were household outbreaks. The most commonly reported settings were restaurant, café, pub, bar, hotel (11 outbreaks), followed by household/domestic kitchen in nine outbreaks (EFSA, 2014). Household outbreaks were also frequent (52.0%) in Piedmont Region (north-western Italy) (Abelli et al., 2013). The reported cases analysed for their *spa*-type showed a high heterogeneity of isolates; *spa* type t13252 resulted correlated in a node of the MST and was never reported as responsible of foodborne outbreak. The most common reported *spa* types (in three FBO cases) are t018 and t084 (Fetsch et al., 2014; Johler et al., 2013, 2015; Schmid et al., 2007; Wattinger et al., 2012); t018 strain was reported as responsible for the production of SEA in SFP ID, 8 strains isolated in SFP ID 27 resulted negative, and strains responsible for SFP ID 11 reported by Wattinger et al. (2012) lack the genotypic and phenotypic characteristics of the isolate. In this reported case, the SEs concentrations were comparable to previously reported outbreaks: 0.625 ng/g of SEA in canned mushrooms (Levine et al., 1996) and 0.45 ng/g of SEE in cheese (Ostyn et al., 2010a). In this case, an average meal might be around 100 g, and 50 g of this may be represented by home-canned tuna fish. Considering the level of combined revealed toxins (2.5 ng/g), the intake may be 125 ng comparable to estimated human dose (around 144 ng) of SE from a large outbreak of SFP involving chocolate milk in a school district (Evenson et al., 1988).

**Home-canning procedure**

The tuna was boiled in salted water for about one hour and then cooled in an uncovered container at room temperature for 12 hours. The meat chunks were added and compacted in nine glass jars (about 400 mL volume) by hand and then coated with corn oil and olive oil. The jars were then closed and placed in boiling water for 30 minutes. The family received it one week before the meal and stored it at room temperature. The jar was opened just before eating and tuna was put in a dish plate.

Contamination during post-cooking and handling, especially putting the meat chunks in jars, was suspected as the possible source of this food poisoning incident. Inadequate final heat treatments in home-canning may allow *S. aureus* cells to survive and produce SEs in food products, including those preserved in oil. SEs are able to retain some biological activity after heat treatment at 121°C for 28 minutes. The decimal reduction (D-value) for *S. aureus* at 60°C has been reported as 4.8-6.6 minutes in broth as compared with 20.5 minutes when encapsulated in fish and oil (Paulin et al., 2012). Additional evidences for contamination during food handling in the post-cooking phase as the most likely cause are finding of preformed SEA and SEE and of enterotoxinogenic strain carrying the relative encoding genes (*sea* and *sed*) in the home-canned tuna.

### Conclusions

The lack of information regarding hand wounds among the family of patients and microbiological contamination of utensils used for cooking and serving is the limit of the study. Likewise, a food-handler was the most likely common source of an SFP gastroenteritis occurring in 2011 in Turin, Italy, following a catered dinner party at a private home (Galliina et al., 2013). This investigation was unable to discover the exact source of the *S. aureus* or the enterotoxins because it was not possible to take nasal or skin swab samples from any of the family members or the utensils used for cooking and serving. Furthermore, spoilage of homemade conserves can result from contamination by food handlers themselves and a lack of knowledge of proper and good practices. The high incidence of domestic outbreaks underlines the importance of risk communication and dissemination of home-canning guidelines to raise awareness for and to reduce the risk of foodborne diseases and outbreaks. Consumers become ill from food consumed at home, which is an important place of consumption for tracing the source of foodborne pathogens, the origin and the typing of responsible isolates. However, apart from identifying the location and the level of the contamination of homemade foods, the exact point for

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### Table 2. Analysis performed and results in food and strain typing.

| SE presence (ELISA qualitative) | SE analysis | Isolated strain analysis | Genotypic ses mPCR |
|---------------------------------|-------------|--------------------------|-------------------|
| Positive                        | SEA         | NHS type I               | *sea* detected    |
|                                 | SEB         | New *spa*-type t13252    | *seb* not detected|
|                                 | SEC         | *sec* not detected       |                   |
|                                 | SED         | *sed* detected           |                   |

|                  | Biotyping | Phenotypic Spa-typing |                      |
|------------------|-----------|-----------------------|----------------------|
| 2.04 ng/mL in 25 g/mL | Biotyping | Phenotypic Spa-typing |                      |
| <LoQ ng/mL in 25 g/mL | Biotyping | Phenotypic Spa-typing |                      |
| <LoD ng/mL in 25 g/mL | Biotyping | Phenotypic Spa-typing |                      |
| 11.03 ng/mL in 25 g/mL | Biotyping | Phenotypic Spa-typing |                      |

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**SE**, staphylococcal enterotoxins; ELISA, enzyme-linked immunosorbent assay; *spa*, gene encoding staphylococcal protein A; *sea*, staphylococcal enterotoxin A; *sec*, staphylococcal enterotoxin B; *sed*, staphylococcal enterotoxin C; *sei*, staphylococcal enterotoxin D; *sel*, limit of quantification; *LoQ*, limit of detection; *NHS*, not host specificity; *see*, staphylococcal enterotoxin type E gene; *sea*, staphylococcal enterotoxin type B gene; *sed*, staphylococcal enterotoxin type D gene; *sei*, staphylococcal enterotoxin type C gene; *sel*, staphylococcal enterotoxin type E gene; *see*, staphylococcal enterotoxin type G gene; *sea*, staphylococcal enterotoxin type H gene; *sei*, staphylococcal enterotoxin type I gene; *sel*, staphylococcal enterotoxin-like type J gene; *sea*, staphylococcal enterotoxin type F gene; *see*, staphylococcal enterotoxin type R gene.

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addressing the guidelines also for small productions should be pinpoint to better identify the causes of illness and to allocate resources in a risk-based manner.

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