Speciation and Gene Flow between Snails of Opposite Chirality

Angus Davison1,2,3*, Satoshi Chiba1, Nicholas H. Barton2,4, Bryan Clarke3

1 Graduate School of Life Sciences, Tohoku University, Aramaki-Aza-Aoba, Aoba-ku, Japan, 2 Institute of Evolutionary Biology, School of Biological Sciences, University of Edinburgh, Edinburgh, United Kingdom, 3 Institute of Genetics, School of Biology, University of Nottingham, Nottingham, United Kingdom, 4 Department of Genetics, University of Cambridge, Cambridge, United Kingdom

Left-right asymmetry in snails is intriguing because individuals of opposite chirality are either unable to mate or can only mate with difficulty, so could be reproductively isolated from each other. We have therefore investigated chiral evolution in the Japanese land snail genus Euhadra to understand whether changes in chirality have promoted speciation. In particular, we aimed to understand the effect of the maternal inheritance of chirality on reproductive isolation and gene flow. We found that the mitochondrial DNA phylogeny of Euhadra is consistent with a single, relatively ancient evolution of sinistral species and suggests either recent “single-gene speciation” or gene flow between chiral morphs that are unable to mate. To clarify the conditions under which new chiral morphs might evolve and whether single-gene speciation can occur, we developed a mathematical model that is relevant to any maternal-effect gene. The model shows that reproductive character displacement can promote the evolution of new chiral morphs, tending to counteract the positive frequency-dependent selection that would otherwise drive the more common chiral morph to fixation. This therefore suggests a general mechanism as to how chiral variation arises in snails. In populations that contain both chiral morphs, two different situations are then possible. In the first, gene flow is substantial between morphs even without interchiral mating, because of the maternal inheritance of chirality. In the second, reproductive isolation is possible but unstable, and will also lead to gene flow if intrachiral matings occasionally produce offspring with the opposite chirality. Together, the results imply that speciation by chiral reversal is only meaningful in the context of a complex biogeographical process, and so must usually involve other factors. In order to understand the roles of reproductive character displacement and gene flow in the chiral evolution of Euhadra, it will be necessary to investigate populations in which both chiral morphs coexist.

Citation: Davison A, Chiba S, Barton NH, Clarke B (2005) Speciation and gene flow between snails of opposite chirality. PLoS Biol 3(9): e282.

Introduction

Left-right asymmetry is an integral part of the establishment of a body plan that may ultimately be traced back to a much deeper molecular asymmetry [1,2]. Although substantial progress has been made recently towards understanding the establishment of asymmetry in several organisms [3–5], almost nothing is known about the first stages in any embryo. Snails may be crucial tools for studying left-right asymmetry, because their chirality is determined at a very early cleavage, and because several species have morphological variation, so that the gene(s) involved can be mapped [6]. It is intriguing that molluscan asymmetry is determined by the effects on the developing embryo of a maternal “chirality” gene (or series of closely linked loci). The genotype of the mother determines the phenotype of the offspring [7–11].

Some recent work has focussed on the issue of why snails are almost always invariant in chirality. One approach has been to study the exceptions, species that are chirally dimorphic, but even in these most populations are fixed for a particular type [11–14] (with the exception of Amphidromus [15]). This is because positive frequency-dependent selection tends to drive the chiral majority to fixation; rarer snails of opposite chirality are less likely to find a mate and successfully copulate [13]. In the best-studied case, the Polynesian tree snail Partula suturalis, dimorphic populations occurred in narrow clines between areas of dextral or sinistral snails [12,13]. In “no choice” experiments, interchiral matings occurred only 20% as frequently as intrachiral matings, and fewer young were produced by the mixed pairs [13]. Since dextral populations of P. suturalis tended to coincide geographically with closely related sinistral species, it was suggested that dextral P. suturalis became established because of reproductive character displacement: Dextral individuals were favoured even when rare, because they were less likely to waste time (or gametes) mating with sinistral species [12,13,16].

More general aspects of shell morphology in land snails have been of interest since the observation by Cain that members of widely separate and taxonomically distinct land snail faunas tend to be either high- or low-spired, with relatively few globular forms in between [17]. Following the suggestion of Gittenberger that there is an association between shell shape and the degree of variation in asymmetry [18], Asami et al. went on to report that reciprocal mating between dimorphic low-spired snails is not usually possible, because the genitalia of a sinistral individual cannot engage with those of a dextral snail [14]. In contrast, high-spired
Chirality and Speciation in Snails

Results
Phylogenetic Analysis of Gene Sequences
As expected from earlier studies [30–32], mitochondrial variation was extreme both within and between species. Only the 16S rRNA fragment had a level of variation suitable for resolving both the deeper relationships and those within species (881 bp when aligned, of which 639 bp were used for the phylogeny; for GenBank references, see Table 1). The variation within 12S rRNA was confined to several hyper-variable regions. Cytochrome oxidase c synonymous nucleotide positions were saturated and its amino acids barely variable. There was also not enough variation within the nuclear sequence of ITS2. These latter three gene fragments were therefore not used further (GenBank AY445024–AY445027, AY251858–AY251871, and AY251820–AY251837).

The sinistral species Euhadra quasita (Figure 1A), E. decorata, E. grata, E. scaevola, and E. murayamai (Figure 1B) were confined to a single clade in the 16S rRNA neighbour-joining phylogeny (Figure 3), which also contained the dextral species E. eoa, E. senckenbergiana (E. senck.; see Figure 1C–1F), and E. latispira. Bootstrap support for the branch that defined this group is 76%, so the simplest explanation is that sinistral Euhadra evolved once only. The maximum likelihood phylogeny, using representative sequences from each clade, had the same overall structure, and the same as that discovered by Ueshima and Asami using a different mitochondrial gene fragment [24].

The relationships within the clade that contains the sinistral species were surprisingly complicated (Figures 3 and 4). Six lineages were found: individual E. decorata, E. eoa, E. grata, and E. scaevola lineages, plus two multi-species lineages, one comprising two species, E. senck. and E. latispira, and the other three species, E. quaesita, E. senck., and E. murayamai. The latter lineage group divides into five subgroups (Figure 4), two that have not been discovered before (QUAI and II), and three already reported (QUAI, IV, and V) [33]. The geographic distributions of these subgroups are principally Tohoku and North East Chubu (QUAI and II), Kanto and Miura peninsula (QUAIII), Kanto and Tohoku (QUAIIV), and Kanto and Izu peninsula (QUAV). All the major lineages within the clade that contains the sinistral species were strongly supported by bootstrapping, but their relationship to each other was not resolved (Figures 3 and 4).

Mitochondrial lineages of sinistral E. murayamai from Myojo-san, as well as dextral E. senck. aomoriensis from Iide-san, Tamayama, and Tsugaru were within the E. quaesita, E. senck., and E. murayamai mitochondrial clade (Figure 4; DNA sample sites and distributions are shown in Figure 2, with further details in Figure S1 and Table S1). E. murayamai is found on a single mountain, yet its mitochondrial haplotypes were found in QUAI and II, nested with haplotypes from E. quaesita. Individual haplotypes of dextral E. senck. aomoriensis were found in QUAI, II, and IV (n = 6), three clades of which are predominantly found in Tohoku, but also North East Chubu and Kanto. The other subspecies of E. senck. grouped with E. latispira. Within the QUAIII lineages (Figure 4), snails were therefore either sinistral E. murayamai from Myojo-san (North East Chubu), sinistral E. quaesita from Haguro-san (central Tohoku), or dextral E. senck. aomoriensis from Tsugaru and Tamayama (northern Tohoku).

Analysis of Morphological Characters
The morphological analysis showed that both E. senck. aomoriensis and E. senck. notoensis have genitalia similar to those in E. quaesita and E. murayamai (Table S2). Specifically, ten of 18 E. senck. aomoriensis genital characters were identical to those of E. quaesita and different from those of E. senck. senckenbergiana, whereas only one of 18 was similar to E. senck. senckenbergiana and different from E. quaesita (Table S2). The morphology of the genitalia in the other E. senck. subspecies (E. senck. senckenbergiana, E. senck. minoensis, and E. senck. ibukicola) was similar to that of E. latispira. E. senck. aomoriensis, E. senck. notoensis, E. quaesita, and E. murayamai grouped together in the genital character maximum parsimony tree, distinct from the other E. senck. subspecies as well as from E. latispira (Figure 5A). The analysis of shell characters showed that there is considerable variation between populations (Figure 5B; Table S3). Consequently, different populations of
the same species group closely with other species in the parsimony analysis of shell characters (Figure 5B).

In summary, *E. senck. aomoriensis* and *E. quaesita* specimens grouped together in the mitochondrial DNA phylogeny and have similar genitalia. In contrast, while *E. senck. notoensis* samples have similar genitalia to *E. senck. aomoriensis* and *E. senck. quaesita*, their mitochondrial DNA sequences grouped with the other *E. senck.* subspecies (Figure 5B).

**Analysis and Interpretation of the Model**

Because the chirality gene is maternally expressed, an individual's phenotype may not correspond to its genotype. This means that we needed to follow the proportions of five kinds of snail: Assuming for the moment that the sinistral allele (S) is dominant over the dextral allele (D), sinistral snails may contain SS, SD, or DD genotypes, whereas dextral snails may be SD or DD (Table S4). No dextral snails can have a SS genotype, because all snails with the SS genotype inherit an S allele from their mother, and so are sinistral. The proportions of the five kinds of snail tend to a characteristic equilibrium that can be thought of as analogous to the Hardy-Weinberg proportion (Table S5). Full details of our model are described in the Protocol S1. Here, an overview is given with an emphasis on findings that are directly relevant to *Euhadra*.

Briefly, the model was used to find the equilibrium frequencies of the chirality gene in each chiral morph, depending upon whether there is random or nonrandom mating (i.e., either interchiral mating with no assortment between chiral morphs, $\alpha = 0$, or complete assortment with no interchiral mating, $\alpha = 1$, respectively). The model is powerful because it predicts the frequency of the sinistral offspring from sinistral morphs as a function of the frequency of sinistral morphs in the population (Figure 6). We expected that the model would be useful in understanding the mitochondrial phylogeny because, by extension, if chirality genes are able to flow freely between chiral morphs, then so will any other gene regardless of whether it is physically linked (on the same chromosome) to the chirality gene or not. Throughout the analysis, we assumed that the allele for sinistral coiling is dominant, as in *Partula*, so the interpretation would have to be modified if dextral is dominant in *Euhadra*.

We were able to account for a variety of other factors as
well as those determining whether mating is random or not. They included frequency-dependent selection, in which rare chiral morphs are less likely to find a mate, and selection against hybrid mating, which might occur if snails are less likely to transfer sperm in interchiral mating. We also considered the effects of reproductive character displacement; i.e., a sinistral species would be at a disadvantage if it risked mating with another sympatric sinistral species when hybrid matings between the two species are sterile or when hybrid offspring are less fit. If a new dextral morph arose within a sinistral species, then while it might initially be at a disadvantage (it would have difficulty finding morphs of the same chirality to mate with), the disadvantage could be counterbalanced if the dextral individuals did not waste time mating with the other species. This explanation for the origin of new chiral morphs has been suggested for *Partula* [12], but has not been tested theoretically.

We first investigated the extent of gene flow between different chiral morphs, depending upon their degree of interchiral mating (Section 2 in Protocol S1), using a parameter, $\alpha$, to define the degree of interchiral or assortative mating. Two equilibria are possible in a chirally mixed population. In the first, gene flow between chiral morphs is substantial, sinistral mothers produce a large proportion of dextral offspring (and dextral mothers produce a large proportion of dextrals), and the population rapidly moves to a balanced equilibrium, even if there is absolutely no interchiral mating ($\alpha = 1$). This high degree of gene flow is a direct consequence of the delayed (maternal) expression of chirality. The implication for *Euhadra* is that gene flow between different chiral morphs is inevitable, even if the morphs are themselves unable to mate.

We have illustrated this first equilibrium in Figure 6. In this figure, the axes represent the proportion of sinistrals in the
population (x-axis) and the proportion of sinistral offspring from sinistral mothers (y-axis). These two parameters were used to illustrate the model because they represent data that can be gathered in the field, and because they have predictive value. For example, in Euhadra, interchiral mating is either not possible ( voted = 1), or the model predicts that 80% of the offspring of sinistral snails will be sinistral. In contrast, if sinistral and dextral chiral morphs were able to mate freely ( voted = 0; unlike in Euhadra, but possible in other species), then the model predicts that 65% of the offspring of sinistral snails will be sinistral. Finally, if there is no gene flow, then sinistral snails will not give birth to dextral individuals (and vice versa). This is the second equilibrium (the upper dotted line in Figure 6), which corresponds to single-gene speciation, and is discussed below.

The second equilibrium is a special case because it is the only equilibrium in which there is no gene flow between the two morphs, so that they are reproductively isolated. It can only occur if the different morphs are completely unable to mate ( voted = 1). However, this equilibrium is unstable and will return to the first equilibrium (free gene flow between chiral morphs) if any sinistral snails have recessive dextral alleles (or alternatively, dextral snails have dominant sinistral alleles), or if there is a very low frequency of interchiral mating (e.g., voted ≈ 0.99). The only requirement, therefore, to enable gene flow between different chiral morphs in this second equilibrium is that intrachiral matings occasionally produce offspring of the opposite asymmetry to the mother, or else that interchiral mating occasionally occurs. Intrachiral matings could produce offspring of the opposite asymmetry by either de novo gene mutation or accidents of development, which can happen when embryos develop in extreme environments. At present we do not know the frequency of interchiral mating in species such as Euhadra. Although no interchiral matings have been observed, they may be possible (i.e., voted ≤ 1).

We next considered the effects of a lower frequency or fertility of mating in the rarer chiral morph, giving the more-common morph an advantage (Figure 7; section 3.1 in Protocol S1). To simplify the equations, we also assumed that differences between selection on “males” and “females” are not of significant effect, which is reasonable since most snails are hermaphrodites. We imagine that fertility is reduced in interchiral mating because sperm is less likely to be transferred. Perhaps more importantly, an individual of the rare chirality might produce fewer offspring, because it will waste time searching for a suitable mate.

Although the model and its equations became more complicated when the above selective factors were included (section 3.1 in Protocol S1), the results were remarkably similar to the first set of circumstances (described above). With random mating (no barriers to interchiral mating, voted = 0), the population moves close to an equilibrium level of gene flow within two generations, even when selection is close to its maximum value. With complete assortment (no interchiral mating, voted = 1), the population moves to equilibrium rather more slowly, since gene flow between morphs is somewhat restricted, but eventually reaches a similar position (Figure 6).

We used the same axes as in Figure 6 to illustrate the equilibria that are reached when selection is included (Figure 7). As the description of the outcome is somewhat more complicated, we used the figure to illustrate the extreme condition only, where no interchiral mating is possible ( voted = 1). The graph shows eight trajectories that start close to the boundary separating two alternative outcomes. As there was no interchiral mating, if the initial condition is two separate subpopulations (i.e., sinistral snails contain only sinistral alleles and vice versa), then the two types stay distinct: the population moves along the top horizontal (dotted) line, with competition between two reproductively isolated types. As before, this situation is equivalent to single-gene speciation. However, if there are any chiral morphs that contain the opposite allele, then the population will move towards an apparent equilibrium in which there is extensive gene flow between the morphs (dotted line running from bottom left to top right in Figure 7), irrespective of the starting frequency of chiral phenotypes and genotypes. Once near this quasi-equilibrium, frequency-dependent selection moves the population towards fixation of one or other morph. Under these conditions, the opportunity for single-gene speciation must be very limited, and even more so if interchiral mating is possible ( voted < 1). Moreover, if single-gene speciation does occur, then as before, it is unstable because of the introduction of new chiral alleles or accidental reversals of chirality.

Another consideration was that selection might act against hybrids between different chiral morphs (section 3.1i in Protocol S1), because genetic differences leading to postmating isolation might be held in disequilibrium with the chirality locus itself. While the chirality gene and genes that determine whether hybrids are less fit (or not) are not likely to be physically linked (on the same chromosome), they may be associated because of coincident selection against the physical act of interchiral mating and because the offspring from interchiral matings tend to be less fit. While this seems unlikely because gene flow should be extensive between different chiral morphs (recombination will break down the disequilibrium), selection against hybrids could occur if the different chiral morphs originally evolved in distinct populations and then came together into a single population. In the most extreme circumstance, all heterozygotes would die, interchiral matings would not produce viable offspring, and the system would be stable.

We also considered a more realistic model, in which heterozygotes have their fertility or survival reduced by a fixed proportion (section 3.1i in Protocol S1). As in the former models, except when there is an almost complete lack of interchiral mating ( voted > 0.99), the model shows that there should be high gene flow between morphs, so that heterozygotes are abundant, and selection causes fixation of the more-common allele relatively quickly. When assortment is almost complete ( voted ≈ 1), there is a narrow range of parameters with two alternative ways for the population to reach fixation: either rapidly, with high gene flow between morphs, or much more slowly, with little gene flow. Nonetheless, the outcome in all circumstances (except when all heterozygotes die) is the same: extensive gene flow between chiral morphs.

Up to this point, we assumed a single population, whereas the more-common circumstance may be that most populations are fixed in their chirality, with narrow clines between
| Genus          | Species     | Chirality | Site, Prefecture, (Site Number) or [Reference Source] | GenBank Accession Number |
|---------------|-------------|-----------|------------------------------------------------------|--------------------------|
| Euhadra       | amaliae     | Dextral   | Nara, Nara (27)                                      | AF098712                 |
|               | awensis     | Dextral   | Anan, Tokushima (2)                                  | AY445016                 |
|               | brandtii    | Dextral   | Shiribeshi, Hokkaido (29)                            | AY251875                 |
|               | callizona   | Dextral   | Heta, Shizuoka (9)                                   | AY251873                 |
|               | congenita   | Dextral   | Hachiman, Kobe, Hyogo (7)                             | AY251874                 |
|               | decorata    | Sinistral | Asamushi, Aomori (3)                                 | AY251907                 |
|               |             |           | Myojo-san, Iwate (31)                                | AY445011                 |
|               |             |           | Myojo-san, Akita (23)                                | AY445023                 |
|               | dixoni      | Dextral   | Daisei, Totori (6)                                   | AF098711                 |
|               | eoa         | Dextral   | Syuzenji, Shizuoka (30)                              | AY251903                 |
|               |             |           | Yugasahima, Shizuoka (40)                            | AY251903                 |
|               |             |           | Irohama, Fukui (15)                                  | AY445019                 |
|               |             |           | Myojo-san, Niigata (24)                              | AY251895                 |
|               |             |           | Myojo-san, Niigata (24)                              | AY251897                 |
|               |             |           | Myojo-san, Niigata (24)                              | AY251896 (2)             |
|               |             |           | Myojo-san, Niigata (24)                              | AY251894                 |
|               |             |           | Myojo-san, Niigata (24)                              | AY251895                 |
|               |             |           | Izu, Shizuoka (16)                                   | AY251878                 |
|               |             |           | Kanto, Izu, Miura [52]                               | AF104039–AF104057        |
|               |             |           | Sendai, Miyagi (28)                                  | AY251897                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251879                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251880                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251881                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251882                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251883                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251884                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251885                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251888                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251889                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251890                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY251891                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY445008                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY445022 (2)             |
|               |             |           | Sendai, Miyagi (28)                                  | AF213716–AF213730, AF354233–AF354257 |
|               |             |           | Sendai, Miyagi (28)                                  | AF098710                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY445020                 |
|               |             |           | Sendai, Miyagi (28)                                  | AY445021                 |
|               |             |           | Tenryu, Shizuoka (33)                                | AY251899                 |
|               |             |           | Kawachi, Shiga (18)                                  | AY445009                 |
|               |             |           | Kawachi, Shiga (18)                                  | AY445010                 |
|               |             |           | Kanto [33]                                           | AF213716–AF213730, AF354233–AF354257 |
|               |             |           | Tsugaru, Aomori (35)                                 | AY251892                 |
|               |             |           | Iide-san, Yamagata (12)                              | AY251886 (3)             |
|               |             |           | Iide-san, Yamagata (12)                              | AY251886 (1)             |
|               |             |           | Tamayama, Iwate (31)                                 | AY445004                 |
|               |             |           | Mt. Fujiwara, Mie (22)                               | AY445014 (3)             |
|               |             |           | Kanazawa, Kanazawa (17)                              | AY444998                 |
|               |             |           | Toyama, Toyama (34)                                  | AY445000 (2)             |
|               |             |           | Ichihashi, Gifu (11)                                 | AY445001                 |
|               |             |           | Azuchi-jo, Shiga (4)                                 | AY445002                 |
|               |             |           | Myojo-san, Niigata (24)                              | AY444999                 |
them, as in *Partula* [12]. The model suggests that regardless of whether mating is random or there is no interchiral mating at all, the position of the cline will move in favour of the dominant allele (section 4.i in Protocol S1) because the dominant allele tends to increase the frequency of its corresponding phenotype following interchiral matings. This conclusion was reached independently by Johnson et al. [25] and Mallet (“dominance drive”) [34]. It is interesting that the shape of the cline will be almost identical regardless of the degree of assortative mating, and in all cases long tails of introgression are expected on either side of the centre of the cline.

A final use of the model was to try to understand the conditions under which new chiral morphs can become established in the first place (section 5 in Protocol S1). If the

---

**Table 1. Continued**

| Genus       | Species | Chirality | Site, Prefecture, (Site Number) or [Reference Source] | GenBank Accession Number |
|-------------|---------|-----------|-----------------------------------------------------|--------------------------|
| *Imajyo*    | subnimbosa | Dextral | Imajyo, Fukui (13) | AY445014 |
| *Imajyo*    | subnimbosa | Sinistral | Yashima, Kagawa (38) | AF0988710 |
| *Imajyo*    | subnimbosa | Sinistral | Nakano, Ehime (26) | AY251908 |
| *Imajyo*    | subnimbosa | Sinistral | South Daito Islands, Okinawa | AF0988713 |

*aFor key to site numbers, see Figure S1 and Table S1.*

DOI: 10.1371/journal.pbio.0030282.t001

---

16S rRNA rate-corrected neighbour-joining phylogeny, rooted using *Nesiohelix bipyramidalis*, showing the relationship between mitochondrial DNA lineages from dextral and sinistral *Euhadra*. The most parsimonious explanation is a single, relatively ancient evolution of sinistral snails. Lineages within the box are shown in detail in Figure 4. Bootstrap support of more than 70% is shown below the node. Shape parameter = 0.30.

DOI: 10.1371/journal.pbio.0030282.g003

---

16S rRNA rate-corrected neighbour-joining phylogeny (subtree from Figure 3) showing the relationship between *E. quaesita*, *E. senck. aomoriensis*, and *E. murayamai* mitochondrial lineages. All unlabelled tips are *E. quaesita*. Both *E. senck. aomoriensis* and *E. murayamai* are polyphyletic. The polyphyly of *E. senck. aomoriensis* can be explained by either single-gene speciation or mitochondrial introgression. Bootstrap support shown for important nodes only.

DOI: 10.1371/journal.pbio.0030282.g005
population were initially entirely sinistral, then positive frequency-dependent selection would act to prevent the establishment of dextral morphs. This is because new chiral morphs would not be able to find a mating partner. However, if a proportion of all matings would otherwise be with another sinistral species, and these matings did not give viable offspring (or the hybrids were less fit), then the mechanical isolation that leads to lower fitness in attempted interspecific interchiral matings would give the new chiral morphs an advantage, because of reproductive character displacement. The model shows that the dextral morphs would be fixed and begin to spread out from the region in which it originated.

If the new morph went on to establish a population beyond the region of overlap with the other chiral morph, a cline might develop, as presumably happened in *P. suturalis* [13]. Moreover, if the allele for the new morph were dominant, then it would tend to move forward due to “dominance drive” [25,34], although movement might be impeded by local barriers. Since the dominance relations in *Euhadra* are unknown, it is difficult to make predictions that are more specific, but in *P. suturalis* the dextral gene is recessive, so dominance drive could have acted against the establishment of a new dextral morph.

An effect of population density could aid the establishment of new morphs. In *Heliconius* butterflies, the interaction between frequency and density is often overlooked as a factor that could assist the establishment of new wing-pattern morphs [29]. At low density, purifying selection against new morphs is strong because predators remain naive, but when the population density is high, selection is weak over a broad range of intermediate frequencies [29]. In *Euhadra*, the parallel situation is that new morphs are less likely to find a...
mate at low density than at high density. New chiral morphs are therefore more likely to become established in populations at high density, because mates of the same chirality are more easily found.

**Single-Gene Speciation in Sympatry or Mitochondrial DNA Introgression?**

The phylogeny did not resolve the relationship between the six major groups within the sinistral clade, as it contains two exclusively dextral groups, three sinistral groups, and one mostly sinistral group that also includes dextral *E. senck. aomoriensis* (see Figures 3 and 4). Recently, Ueshima and Asami found a similar phylogeny using a different set of mitochondrial gene fragments [24]. One interpretation of these phylogenies is that the polyphyly of *E. senck. aomoriensis* within the sinistral mitochondrial DNA *E. quaesita* clade is suggestive of recent gene flow between sinistral *E. quaesita* and dextral *E. senck. aomoriensis*. An alternative, as concluded by Ueshima and Asami, is that the polyphyletic *E. senck. aomoriensis* lineages are in fact a different (i.e., new) species that has been derived from *E. quaesita* by single-gene speciation [24].

As in the original evolution of sinistral *Euhadra* that was discussed earlier, initially rare dextral morphs of *E. quaesita* could have been favoured in their spread by reproductive character displacement from another species. The most widespread sinistral species with which *E. quaesita* might have hybridized is *E. decorata*, which is found in the northern part of Tohoku. This explanation is reasonable because *E. quaesita* has a restricted distribution in northern Tohoku, whereas *E. senck. aomoriensis* and *E. decorata* are quite widespread there (see Figure 2). Thus, dextral *E. senck. aomoriensis* may have evolved from sinistral *E. quaesita*, because otherwise *E. quaesita* would tend to hybridize with sympatric, sinistral *E. decorata*. If so, this process could have been quite recent and in situ, because northern Tohoku is considered to have been unvegetated during the last glaciation (Figure 8).

However, it is presently not possible to resolve whether hybridization or single-gene speciation best explains the mitochondrial DNA phylogeny, especially since the dominance of the chirality gene is unknown, and the morphological analysis complicates the interpretation (see Figure 5). We therefore used our model to compare the two competing explanations. We found that when there is even a slight degree of interchiral mating (incomplete assortment; \( \alpha < 1 \)), there is a single feasible equilibrium that is always stable. This equilibrium, which is analogous to a Hardy-Weinberg equilibrium while accounting for the maternal inheritance of the chirality gene, is one of substantial gene flow between the chiral morphs (see Figure 6).

When there is no interchiral mating (complete assortment; \( \alpha = 1 \)), two equilibria become possible. As before, in the first equilibrium there is substantial gene flow between morphs. This is interesting because it means that even when different chiral morphs are unable to mate (as in *Euhadra*), gene flow will still be substantial because of the delayed (maternal) inheritance. This observation was first pointed out by Johnson et al. [25]. In the second equilibrium, there is...
complete reproductive isolation so it amounts to single-gene speciation, but is only possible when sinistral snails are entirely sinistral homozygotes, and dextral snails are entirely dextral homozygotes. The latter equilibrium is unstable to the introduction of sinistral snails carrying dextral alleles or vice versa, and to a low frequency in interchiral mating. The empirical studies that have been carried out in Partula and also Achatinella tend to support the predictions from the model of high gene flow between morphs [38–40]. While assortative mating should be much greater in Euhadra because interchiral mating is not possible ($\alpha \approx 1$) [14], the prediction is that gene flow still should be substantial.

Another factor that could reduce gene flow between chiral morphs is selection. First, the more-common morph could be at an advantage because it spends less time searching for its own morph to mate with, and may well have more offspring as a consequence. Second, snails are probably less likely to transfer sperm successfully in interchiral matings. There is direct evidence for reduced fertility in Partula, since fewer young are produced by mixed pairs [13]. When we incorporated this mode of selection into the model, with random mating (inter- and intrachiral mating equally likely; $\alpha = 0$) the population moves rapidly to equilibrium gene flow. Even with a complete lack of interchiral mating ($\alpha = 1$), there is still substantial gene flow. The main difference is that the population moves towards equilibrium rather more slowly (see Figure 7).

Although there are no data for Euhadra, it is possible that a selection operates against hybrids between chiral morphs because genetic differences leading to postmating isolation are held in association (disequilibrium) with the chiral gene itself, preventing introgression. Evidence from the model suggests that such disequilibrium is unlikely to build up in sympathy. However, if two different morphs met after a period in allopatry, then the population could fall into an equilibrium in which gene flow is reduced, and so maintain the initial association. Even in this circumstance, the model shows that for all but very strong assortment ($\alpha > 0.99$), there still should be high gene flow between morphs, so that heterozygotes would be abundant and selection would cause fixation of the more-common viability allele relatively quickly.

The results from the model have important implications for the interpretation of the mitochondrial DNA phylogeny, and can help to make suggestions for future research. Reproductive character displacement can explain how new chiral morphs arise and become the majority. However, even with reproductive character displacement there will still be a significant flow of the chirality gene (and other genes) between morphs. In these circumstances, speciation by chiral reversal is meaningful only in the context of a complex biogeographical process, which must usually involve other factors. Allopatry, perhaps combined with ecological selection, will be required for genetic differences and reproductive isolation to build up between chiral morphs. As this must have happened in Euhadra and several other species, it will be interesting to disentangle the relative contributions of each towards reproductive isolation.

The immediate question is whether sinistral E. quaesita reverted back to a dextral species, producing E. senck. aomoriensis, or whether there has been gene flow between E. quaesita and E. senck. aomoriensis. To resolve this issue, a priority should be to investigate the phylogeny of nuclear gene sequences and compare them with the mitochondrial DNA phylogeny. If dextral E. senck. aomoriensis arose by single-gene speciation, then most nuclear loci should place these specimens within the E. quaesita clade. Alternatively, if the polyphyletic position of dextral E. senck. aomoriensis in the mitochondrial DNA phylogeny is due to hybridization and introgression, then the genomes of the two populations should be a mosaic in their extent of differentiation [41–43].

However, even with a complete lack of interchiral mating, introgression and fixation of lineages will occur rapidly, so that introgression may be difficult to distinguish from single-gene speciation.

Another priority should be to establish the dominance relations of the chirality genes in Euhadra. This is because a prediction of the model and previous simulations [25,26,34] is that morphs are more likely to be established by single-gene speciation if the mutation involved is dominant. One means to establish dominance would be to attempt breeding experiments in the laboratory. However, there are no known dimorphic species, and interchiral mating has not been observed, so this may not be possible. A solution would be to discover a rare coiling variant, as has been done in Bradybaena similaris [24]. The final priority is to finely map the distributions of the different morphs, especially sinistral E. quaesita, sinistral E. decorata, and dextral E. senck. aomoriensis, to test whether the patterns are consistent with reproductive
character displacement as an explanation. Individuals from zones of contact could be used in captive breeding experiments to determine the dominance relations, the proportions of each chirality allele, and the extent of gene flow (if any).

Wider Significance

The interpretation of the model could impact upon the understanding of the evolution of any gene with an indirect genetic effect, especially maternal-effect genes [44]. Maternal-effect genes are important in development because maternally produced molecules (e.g., mRNAs) in the egg affect early developmental processes such as axis formation, and often have later pleiotropic effects. Maternal-effect genes have been implicated in causing infertility [45], with a few also directly implicated in causing hybrid sterility [46], although generally it has been realized that their contribution to postzygotic isolation might be limited [47]. Finally, sex determination in many species involves interactions between maternal and zygote genes, so it has been proposed that conflicting selective pressures between maternally and zygotically expressed sex-determining loci can in turn have a role in shaping the evolution of sex determining systems [48]. However, the detailed discussion of the relevance of our model to these and other issues is beyond the scope of this paper.

Conclusions

The concept of single-gene speciation in snails has attracted attention because it is an exception to the accepted view that at least two pairs of interacting genes are required for reproductive isolation [21,22,24]. We have shown that reproductive isolation between chiral morphs is possible, but unstable, and will lead almost inevitably to gene flow. While chiral morphs may be a step towards distinct species, complete reproductive isolation must require other factors, such as divergent selection for habitat use or geographic separation of populations. Assuming that the differences in genitalia are not due to dominance or epistasis, chiral morphs can separate of populations. Assuming that the differences in genitalia are not due to dominance or epistasis, chiral morphs can separate of populations. Assuming that the differences in genitalia are not due to dominance or epistasis, chiral morphs can separate of populations. Assuming that the differences in genitalia are not due to dominance or epistasis, chiral morphs can separate of populations. Assuming that the differences in genitalia are not due to dominance or epistasis, chiral morphs can separate of populations. Assuming that the differences in genitalia are not due to dominance or epistasis, chiral morphs can separate (5).

Materials and Methods

Samples. The 22 species of Euhadra (Bradybaenidae) are distributed throughout Japan and the neighbouring Korean island of Jeju [50]. We sampled all five sinistral species for morphological and molecular analysis, in addition to 14 of the 17 dextral species (Figure S1: Tables 1 and S1). The remaining dextral species, E. sadonis, E. nuchiosa, and E. sigens have restricted distributions. The distributions of four species are shown in Figure 2, using data compiled from [50,51] and the Web site http://www.biodie.go.jp/site_/maps/site_map.html (in Japanese). We were also able to use an extensive set of 16S rRNA sequences from E. peliomphala [52] and E. quaesita [33]. For the outgroup, Nesohelix bryanioidalis was used because a prior analysis has suggested that it is suitable [31].

To enable a clear comparison between our results and those of Ueshima and Asami [24], we refer to subspecies where necessary, especially with regard to E. senck. Note that Ueshima and Asami refer to E. senck as “E. omoerisensis” and E. murayamae as “E. quaesita murayamae” [24].

Morphological analysis. Morphological characters were measured in subset of the species, primarily those from the “sinistral” clade, and then used to make a maximum parsimony tree. In the first analysis of genital shape and structure, the characters were classified as follows: (1) wings on the basal part of dart: 0, absent; 1, small; and 2, large (ordered). (2) Wings on the middle part of dart: 0, absent; 1, small; and 2, large (ordered). (3) Curve of wings on the front part: 0, absent; 1, present. (4) Shape of wings: 0, absent; 1, straight; and 2, curved (ordered). (5) Maximum number of wings: 0, absent; 1, two; and 2, three (ordered). (6) Shape of cross-section of dart: 0, round or oval; 2, triangular; and 1, crescent (ordered). (7) Flatness of the front section of the dart: 0, very flat (rounded); 1, flat; and 2, curved (ordered). (8) Flatness of the middle section of dart: 0, round; 1, flat; and 2, very flat (ordered). (9) Flatness of the basal section of dart: 0, round; and 1, flat. (10) Shape of the front part of dart-sac: 0, round; 1, oval; and 2, long- oval (ordered). (11) Shape of the mid-dorsal part of dart-sac: 0, curved; and 1, straight. (12) Shape of the dart-sac near atrium: 0, round; and 1, pointed. (13) Length of the middle part relative to the front part of dart-sac: 0, short; and 1, long. (14) Size of the sheath at the basal part of dart-sac: 0, small; and 1, large. (15) Relative size of sub-dart sac: 0, large; 1, medium; and 2, small (ordered). (16) Shape of sub-dart sac: 0, round; and 1, flat. (17) Number of nuculus glands: 0, > 10; 1, from 5 to 10; and 2, < 5 (ordered). (18) Region between sub-dart sac and dart sac: 0, not contracted; and 1, contracted.

DNA extraction and PCR amplification. Genomic DNA was isolated using previously described methods [53]. Primers for PCR amplification of an approximately 900-bp fragment of 16S rDNA are described in [51]. Variation in the mitochondrial 12S rRNA and cytochrome oxidase subunit 1 genes, and nuclear ITS2 region was also investigated, using primers described in [31,55,54]. All PCR reactions used Takara Taq (Takara Biomedicals, Tokyo, Japan) and buffers, with annealing temperatures of 50 °C. Cycle sequencing was carried out with both forward and reverse primers, using about 80–100 pmol of PCR product in the reaction and the BigDye Terminator v3.0 Cycle Sequencing Ready Reaction Kit (Applied Biosystems, Foster City, California, United States). DNA sequences were electrophoresed on a 310 Genetic Analyzer (Applied Biosystems).

Phylogenetic analyses. Sequences were aligned using the ClustalX software, and then checked manually. All insertion and deletion sites (indels), as well as several difficult-to-align regions, were removed before phylogenetic analysis. Phylogenetic relationships were analyzed using two methods: neighbour-joining and maximum likelihood, both with PAUP*4.0b8 [55]. Multiple hits were corrected using the general time reversible (GTR) model. The tree matrix, base frequencies, and shape parameter (a) of the gamma distribution (based on 16 rate categories) were estimated using likelihood, by iteration from an initial neighbour-joining tree. Parameters estimated from the initial tree were used to make a new neighbour-joining tree. The parameters were then re-estimated at every step of the process was repeated until there was no further improvement in likelihood. Bootstrap values were then calculated using 1,000 replicates. Maximum likelihood methods used a heuristic procedure with tree-bisection-recombination, and 100 bootstrap replicates.

Assumptions. In the species that have been investigated, asymmetry (or chirality) is under control of one or a few linked loci, where the phenotype of the offspring is controlled by the maternal genotype (maternal inheritance). Dominance is variable, even within genera: dextral is dominant in Lymnaea pyrga, L. stagnalis, B. similaris, and P. natalensis [5,7,8,10,11,24] and is dominant in D. Pulex, Lymnaea, and Lyncinaea [9,25]. Reversed asymmetry can sometimes be agenetic, when development is disrupted due to environmental extremes, usually heat or cold shock. We assumed that asymmetry is also
maternally inherited in Euhadra. In Partula, mating is possible between different morphs (although it occurs at reduced frequency) because they are high-spired and mate by shell-mounting [19]. There is no such detailed information for Euhadra, but evidence suggests that interchiral mating is difficult and perhaps impossible, because they mirror “face-to-face” [14].

Mathematical model. We developed a mathematical model to understand gene flow between chiral morphs, as well as the route by which new chiral morphs can become established. The methods are described in detail in Protocol S1.

Supporting Information

Protocol S1. The Mathematical Model

Found at DOI: 10.1371/journal.pbio.0030282.sd001 (89 KB DOC).

Figure S1. Sample Sites Used in This Study

The sample sites used in this study, highlighting in particular the sites where E. quaesita, E. murrayamai and E. senchengeriana were collected (underlined if used for DNA methods), m, E. murrayamai; q, E. quaesita; sa, E. sench. amornensis; si, E. sench. ibukicolae; sm, E. sench. minorensis; sn, E. sench. notoniensis; ss E. sench. sench. 
Found at DOI: 10.1371/journal.pbio.0030282.sg001 (661 KB TIF).

Table S1. Species Collected at Each Site

Found at DOI: 10.1371/journal.pbio.0030282.st001 (24 KB DOC).

Table S2. Genital Characters

See Materials and Methods for definitions.
Found at DOI: 10.1371/journal.pbio.0030282.st002 (24 KB DOC).

Table S3. Shell Characters

See Materials and Methods for definitions.

References

1. Brown NA, Wolpert L (1990) The development of handedness in left/right asymmetry. Development 109: 1–9.
2. McManus C (2002) Right hand left hand. The origins of asymmetry in brains, bodies, atoms and cultures. London: Weidenfeld and Nicolson. 412 p.
3. Nonaka S, Shiratori H, Saijoh Y, Hamada H (2002) Determination of left-right patterning of the mouse embryo by artificial nodal flow. Nature 418: 969–972.
4. Bergmann DC, Lee M, Robertson B, Tsou MFR, Rose LS, et al. (2003) Embryonic handedness choice in C. elegans involves the G alpha protein GPA-16. Development 130: 5731–5740.
5. Shibazaki Y, Shimizu M, Kuroda R (2004) Body handedness is directed by genetically determined cytoskeletal dynamics in the early embryo. Curr Biol 14: 1462–1467.
6. Hosoi Y, Harada Y, Kuroda R (2005) Construction of a backcross progeny collection of dextral and sinistral individuals of a freshwater gastropod, Lymnaea stagnalis. Dev Genes Evol 213: 193–198.
7. Boycott AE, Diver C (1925) On the inheritance of sinistrality in the gastropod E. quaesita where D describes the strength of assortative mating, β describes the mating advantage of dextral over sinistral snails (averaged over the sexes), and δ describes the difference in mating advantage between the reciprocal crosses. Necessarily, α < 1–δ/β2.

Found at DOI: 10.1371/journal.pbio.0030282.st003 (27 KB DOC).

Table S4. The Five Classes of Snail and Their Frequencies

The formulas used in calculating frequencies were $Q + P = 1$, and $d + s = 1$.
Found at DOI: 10.1371/journal.pbio.0030282.st004 (33 KB DOC).

Table S5. The Contribution of Each of the Four Phenotypic Mating Combinations to the Next Generation

Note that α describes the strength of assortative mating, β describes the mating advantage of dextral over sinistral snails (averaged over the sexes), and δ describes the difference in mating advantage between the reciprocal crosses. Necessarily, α < 1–δ/β2.
Found at DOI: 10.1371/journal.pbio.0030282.st005 (25 KB DOC).

Acknowledgments

We are grateful to Yuji Watanabe and Morito Hayashi for providing some of the 16S rRNA sequences. In addition, Jonathan Hobley, Osamu Takahashi, Jun Yokoyama, Norio Wakayama, Osamu Miura, and Lazaro Echenique Diaz helped collect snails. We thank Sara Goodacre, John Brookfield, Chris Wade, Adele Grindon, and Kester Jarvis for helpful comments; and Menno Schilthuizen, James Mallet, and three further anonymous referees for improvements to the manuscript. AD was supported by a Royal Society “2+2” fellowship. NB was supported by a Natural Environment Research Council postdoctoral fellowship.

Competing interests. The authors have declared that no competing interests exist.

Author contributions. AD, SC, NHB, and BC conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, and wrote the paper.

Found at DOI: 10.1371/journal.pbio.0030282.st002 (24 KB DOC).

Partula suturalis. Dev Genes Evol 213: 193–198.
8. Sturtevant AH (1925) Inheritance of direction of coiling in Lymnaea. Science 58: 269–270.
9. Degener E (1925) Der erbgang der inversion bei Lucinaria triplicata MTG (Gast Pulum). Mitt Humb Zool Mus Inst 51: 3–61.
10. Murray J, Clarke B (1976) Supergenes in polymorphic land snails 2. Partula suturalis. Heredity 37: 271–282.
11. Freeman G, Lundelius JW (1982) The developmental genetics of dextrality and sinistrality in the gastropod Lymnaea peregra. Proc R Soc Lond B Biol Sci 95: 207–213.
12. Sturtevant AH (1925) Inheritance of direction of coiling in Lymnaea. Science 58: 269–270.
13. Johnson MS (1982) Polymorphism for direction of coil in Partula. Biol J Linn Soc 14: 146–151.
14. Freeman G, Lundelius JW (1982) The developmental genetics of dextrality and sinistrality in the gastropod Lymnaea peregra. Roux Arch Dev Biol 191: 69–83.
15. Clarke B, Murray J (1969) Ecological genetics and speciation in land snails of the genus Partula. Biol J Linn Soc 1: 31–42.
16. Freeman G (1982) Polymorphism for direction of coil in Partula suturalis—Behavioral isolation and positive frequency-dependent selection. Heredity 49: 145–151.
17. Asami T, Cowie RH, Ohbayashi K (1998) Evolution of mirror images by sexually asymmetric mating behavior in hermaphroditic snails. Am Nat 152: 225–236.
18. Gittenberger E (1988) Symaptic speciation in snail—A largely neglected model. Evolution 42: 826–828.
19. Lipton CS, Murray J (1979) Courtship of lands snails of the genus Partula. Malacologia 19: 129–146.
20. Asami T (1993) Genetic variation and evolution of coiling chirality in snails. Mol Ecol 2: 96–99.
21. Dobzhansky T (1937) Genetics and the origin of species. New York: Columbia University Press. 364 p.
22. Muller HJ (1942) Isolating mechanisms, Evolution and temperature. Biol Symp 6: 71–125.
23. Orr HA (1991) Is single-gene speciation possible? Evolution 45: 764–769.
24. Ueshima R, Asami T (2003) Single-gene speciation by left-right reversal—A land-snail species of polythetic origin results from chirality constraints on mating. Nature 425: 679–679.
25. Johnson MS, Clarke B, Murray J (1990) The coil polymorphism in Partula suturalis does not favor sympatric speciation. Evolution 44: 459–464.
26. van Ratenburg FHD, Gittenberger E (1996) Ease of fixation of a change in coiling: Computer experiments on chirality in snails. Heredity 76: 278–286.
27. Stone J, Bjorklund M (2002) Delayed prezygotic isolating mechanisms: Evolution with a twist. Proc R Soc Lond B Biol Sci 269: 861–865.
28. Mallet J, Singer MC (1987) Individual selection, kin selection, and the shifting balance in the evolution of warning colors—The evidence from butterflies. Biol J Linn Soc Lond 32: 337–350.
29. Mallet J, Joron M (1999) Evolution of diversity in warning color and mimicry. Polymorphisms, shifting balance, and speciation. Annu Rev Ecol Syst 30: 201–233.
30. Thomaz D, Guiller A, Clarke B (1996) Extreme divergence of mitochondrial DNA within species of pulmonate land snails. Proc R Soc Lond B Biol Sci 263: 363–368.
31. Chiba S (1999) Accelerated evolution of land snails Mandarina in the oceanic Bonin Islands: Evidence from mitochondrial DNA sequences. Evolution 55: 460–471.
32. Davison A (2002) Land snails as a model to understand the role of history and selection in the origins of biodiversity. Popul Ecol 44: 129–136.
33. Watanabe Y, Chiba S (2001) High within-population mitochondrial DNA variation due to microvariance and population mixing in the land snail Euhadra quaesita (Pulmonata: Bradybaenidae). Mol Ecol 10: 2635–2645.
34. Mallet J (1986) Hybrid zones of Heliconius butterflies in Panama and the stability and movement of warning color clines. Heredity 56: 191–202.
35. Peake J (1975) Species isolation in sympatric populations of the genus Diplomatomis (Gastropoda, Prosobranchia, Cyclophoridae, Diplomatini). J Zool Lond 177: 593–615.
36. Uit de Weerd DR, Groenenberg DSJ, Schilthuizen M, Gittenberger E (2005) Reproductive character displacement by inverision of coiling in clausiliid snails (Gastropoda, Pulmonata). Biol J Linn Soc Lond. In press.
37. Schilthuizen M, Davison A (2005) The convoluted evolution of snail chirality. Naturwissenschaften. In press.
38. Johnson MS, Murray J, Clarke B (1987) Independence of genetic subdivision and variation for coil in Partula suturalis. Heredity 58: 307–313.
39. Goodacre SL (2002) Population structure, history and gene flow in a group of closely related land snails: Genetic variation in Partula from the Society Islands of the Pacific. Mol Ecol 11: 365–375.
40. Hoolland BS, Hadfield MG (2002) Islands within an island: Phylogeography and conservation genetics of the endangered Hawaiian tree snail Aschatinella mustelina. Mol Ecol 11: 365–375.
41. Abernethy K (1994) The establishment of a hybrid zone between red and sika deer (Genus Cervus). Mol Ecol 3: 551–562.
42. Goodman SJ, Barton NH, Swanson G, Abernethy K, Pemberton JM (1999) Introgression through rare hybridization: A genetic study of a hybrid zone between red and sika deer (genus Cervus) in Argyll, Scotland. Genetics 152: 355–371.
43. Wu CI (2001) The genic view of the process of speciation. J Evol Biol 14: 851–865.
44. Wolf JB, Brodie ED, Cheverud JM, Moore AJ, Wade MJ (1998) Evolutionary consequences of indirect genetic effects. Trends Ecol Evol 13: 64–69.
45. Boswell RE, Prout ME, Steichen JC (1991) Mutations in a newly identified Drosophila melanogaster gene, mago nashi, disrupt germ cell formation and result in the formation of mirror image symmetrical double abdomen embryos. Development 113: 373–384.
46. Beeman RW, Friesen KS (1999) Properties and natural occurrence of maternal-effect selfish genes (“Medea” factors) in the red flour beetle Tribolium castaneum. Heredity 82: 293–299.
47. Hurst GDD, Schilthuizen M (1998) Selfish genetic elements and speciation. Heredity 80: 2–8.
48. Werren JH, Hatcher MJ, Godfray HCJ (2002) Maternal-offspring conflict leads to the evolution of dominant zygotic sex determination. Heredity 88: 102–111.
49. Mayr E (1963). Mayr E (1963) Animal species and evolution: Harvard University Press.
50. Azuma M (1995) [Coloured illustrations of the land snails of Japan]. Osaka: Hoikusya. 359 p.
51. Anonymous (2002) The national survey on the natural environment: Report on the distributional survey of Japanese animals (land and fresh water mollusca). Tokyo: Biodiversity Center of Japan, Ministry of the Environment. 1317 p.
52. Hayashi M, Chiha S (2000) Intraspecific diversity of mitochondrial DNA in the land snail Euhaedra peliomphala (Bradybaenidae). Biol J Linn Soc Lond 70: 391–401.
53. Teshima H, Davison A, Kuwahara Y, Yokoyama J, Chiha S, et al. (2003) The evolution of extreme shell shape variation in the land snail Aiolohelix editha: a phylogeny and hybrid zone analysis. Mol Ecol 12: 1869–1878.
54. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R (1994) DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Mol Mar Biol Biotech 3: 294–299.
55. Swofford DL (1999) PAUP* 4.b10. Sunderland, MA: Sinauer Associates.
56. Kaizuka S (1977) Geology and geomorphology of the Bonin Islands. Bulletin of Ogasawara Research 1: 29–34.
57. Yoshikawa T, Kaizuka S, Sakaguchi Y, Sugimura S, Ohta Y (1973) Geomorphology of Japan. Tokyo: University of Tokyo Press. 415 p.
58. Kaizuka S, Chinzei K (1995) Nature of Japan: Mountains in Japan. Tokyo: Iwanami-Shoten. 270 p.