Vitamin-anticancer drug conjugates: a new era for cancer therapy

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ABSTRACT
Background: Following cardiovascular diseases, cancer is the world’s second leading cause of death. Chemotherapy is the conventional gold technique for successful treatment of cancer. There are some drawbacks associated with traditional chemotherapy, namely, low aqueous solubility, limited biological half-life, production of multidrug resistance and non-specificity (lack of targeting ability) or dose-limiting cellular toxicity. To develop a targeted drug delivery for its anticancer effect is still a challenging task.

Methods: We developed literature review methods which included inclusion and exclusion criteria for identifying potentially relevant articles, articles search strategies, abstract review protocols and a comprehensive scoring system for published studies. This study contains a detailed survey of various reported methods such as folic acid-drug conjugates, Cobalamin-Drug Conjugate, Vitamin B12-Conjugated and Paclitaxel-Loaded Micelles etc., all of which were studied for their methods of preparation and possible impact on biological activity.

Results: Due to its specific ability to carry anticancer drugs directly to tumours, vitamin-mediated drug targeting has recently emerged as a novel concept. Solid tumour cancer has an unquenchable appetite for various essential vitamins, resulting in over-expression of the receptors involved in cell internalization of vitamins on the surface of cancer cells. So, the vitamin drug conjugates are specifically important for carrying the anticancer drugs directly to the tumour cells. Biotin, folic acid, vitamin B12 and riboflavin, the vitamin necessary for the division of all cells, especially cancer cells, have recently been examined as targeting agents.

Conclusion: Vitamin-Drug Conjugate methods were found to be the most suitable methods amongst all the other reported methods and they can be applied for current therapy against cancer.

Keywords: Cancer, chemotherapy, anticancer drug and vitamin

INTRODUCTION
Cancer is the unchecked development of cells in the human body and these cells have the capacity to metastasize. If this spread is not managed, cancer will lead to death (Davis, 2019). It is a multacellular and multigenic disease that may occur with a complex etiology from any type of cell and organ (Baskar, Lee, Yeo, & Yeoh, 2012). Cancer has become the world’s second-largest cause of death after cardiovascular disease (Padma, 2015).

Mainstream modalities of treatment include surgery, radiotherapy, immunotherapy, hormonal therapy, laser therapy, and stem cell therapy. Surgery and radiotherapy are the most effec-
tive and common therapies for localized primary tumors and non-metastatic cancers. Cancer drugs (chemotherapy, biological and hormonal therapy) are used to treat metastatic cancer. Drugs spread to all tissues in the body, through the bloodstream (Pérez-Herrero, & Fernández-Medarde, 2015). Thus, for effective cancer treatment, chemotherapy is said to be as the gold standard strategy. Chemotherapy is based on chemotherapeutic drugs (i.e. toxic compounds) that act by inhibiting the cancer cells' rapid proliferation. There are some drawbacks associated with traditional chemotherapy, i.e. chemotherapeutic drugs with cancer cell suppression, which can also prevent the rapid growth needed to maintain normal cells, such as bone marrow, gastrointestinal tract and hair follicle cells, leading to undesirable side effects in cancer treatments (Seifu & Nath, 2019). Some other drawbacks in cancer treatment associated with free chemotherapeutic agents are low aqueous solubility, short biological half-life, production of multidrug resistance and non-specificity (lack of targeting capacity) or dose-limiting cellular toxicity.

Consequently, it is still difficult to target an anti-cancer drug selectively to cancerous cells and to achieve a breakthrough in cancer research. To achieve this goal, different approaches can be implemented. The most popular is to provide an effective carrier for a medication. Thanks to its ease of use and uncomplicated chemical modification, polymeric drug carriers are the most powerful (Pawar, Badhwar, Kharas, Khandare, & Vavia, 2012; Tripodo, Mandracchia, Collina, Rui, & Rossi, 2014). Such systems usually have the ability to accumulate by means of passive drug targeting in different organs as a result of their removal route or for active organotropism, and a more specific drug targeting can be accomplished by adding a carrier with specific targeting agents, such as vitamins, antibodies, peptides, magnetic particles or hormones (Cavallaro, Maniscalco, Campisi, Schillaci, & Giammona, 2007; Bareford, Avaritt, Ghandehari, Nan, & Swaan, 2013; Tripodo et al., 2014; Gibiansky & Gibiansky, 2014; Qu, Zhou, Chen, Chen, & Shen, 2015). The most commonly used method for drug targeting to specific cells is to attach a drug directly to targeting moiety to form a new chemical entity which is pharmacologically active, i.e. prodrug (Elsadek et al., 2010; Tripodo et al., 2014). In these conditions, the so-called vitamin-mediated drug targeting has recently emerged as a new concept for carrying anticancer drugs particular to tumours (Mahato, Tai, & Cheng, 2011; Bildstein, Dubernet, & Couvreur, 2011; Fortin & Berube, 2013). As it is well known, all living cells require vitamins for their survival, whereas rapidly dividing cells that are found in solid tumour cancer, have an unquenchable appetite for different essential vitamins, resulting in vitamin receptors being over-expressed on the surface of cancer cells. In addition, there has been a claim that the combination of an anticancer drug with a particular vitamin contributes to the development of vitamin-drug conjugates (Figure 1). This vitamin drug conjugate would lead to the targeting of a higher amount of drugs, i.e. a high dose to the targeted cancer cells. Biotin, folic acid, vitamin B12 and riboflavin, which are necessary for the division of all cells, in particular cancer cells, have recently been investigated as targeting agents (Chen et al., 2010; Russell, McTavish, & McEvan, 2011).

**Vitamin-Drug Conjugates**

A drug is linked directly or through a spacer with a targeting moiety to form a pharmacologically active new chemical entity to produce vitamin-drug conjugate.

Advantages: Vitamin drug conjugates will be nontoxic, and they will be specifically internalized into cancerous cells and release anticancer drugs without loss of potency. In addition, they will minimize the systemic toxicity by being stable in blood circulation, and provide a target specific activity by sparing the normal cell that will minimize any side effects (Ojima, Zuniga, Berger, & Seitz, 2012).

**Vitamin-9 (Folic Acid)-Drug Conjugates**

Folic acid (FA) is more closely related to folate receptors (FR). Glycosylphosphatidylinositol (GPI)-linked membrane folate receptor was involved in folate-based chemotherapeutic drug receptor-mediated absorption (Rijnboutt, Jansen, Posthuma, Hynes, & Schornagel, 1996). This GPI-connected membrane protein collects the ligands from the extracellular matrix and transports them through the recycled endosomal pathway to the intracellular space (Figure 2). The folate receptors are also recognized as tumour antigens/biomarkers due to the fact that the folate receptors are commonly absent in normal tissues. This has led to the development of therapeutics and diagnosis approaches for the treatment of cancer (Leamon et al., 2007). Hence molecular payloads with a size range from radiouclides to large constructs of DNA and liposome have been
successfully delivered through the FR pathway within cancer cells. One particularly related approach is to attach powerful chemotherapy drugs to FA to form a small molecule drug conjugate or SMDCs. (Vlahov & Leamon, 2012).

**Folic acid-paclitaxel conjugates**

Yuxuan et al., 2019, engineered the conjugates of folic acid-peptide-paclitaxal (FA-P3/P7-PTX), to resolve drug resistance, achieve targeted delivery to tumours, increase cellular absorption, and allow water-soluble conjugates. The synthesized (FA-P3/P7-PTX) conjugate was a folic acid -lytic peptide conjugate, in which lytic peptides I-3 and I-7 were used as molecular carriers and cell-disrupting peptides. 16-site cysteine-substituted I-3 and I-7 were named as P3 and P7, respectively, and served as a peptide backbone. The conjugates were tested for their efficacy and also showed higher anti-proliferative activity than free paclitaxel in MCF-7/PTX cells. Higher cellular uptake in MCF-7/PTX cells was shown by, FA-P3-PTX than P3-PTX, which was based on folate receptors present. FA-P7-PTX had a much stronger effect on cell toxicity, apoptosis and membrane disturbance behavior in MCC. Similar to FA-P3-PTX, FA-P7-PTX had more ability to suppress tumor growth than PTX. (Wang et al., 2011; Gaspar, Costa, Queiroz, Pichon, & Souse, 2015; Dai et al., 2019).

**Folic acid-DAVLBH conjugates (Vintafolide)**

Vintafolide (formerly EC145), the most active FA-SMDC, developed by Leamon et al., 2014, was a water-soluble conjugate that selectively delivers the medication desacetyl vinblastine monohydrazine (DAVLBH) to tumours over-expressed to FR-alpha (Leamon et al., 2014). Preclinical studies have shown a higher affinity of vintafolide to bind to FRa, and therefore have a very strong and selective action against FRa-positive xenografts as compared to untargeted DAVLBH (Vlahov et al., 2009).

Vintafolide consists of 4 part modules: 1) a folic acid moiety targeting FRa, 2) a hydrophilic peptide spacer, 3) an autoimmolative disulfide linker, and 4) a DAVLBH microtubule-stabilizing compound (Figure 3).

Leamon et al., 2014, conducted a study to assess the impact of changing three of the vintafolide’s constituent elements out of the four. It was noted that by changing the spacer composition, given the spacer remained hydrophilic, the vintafolide’s potency was minimally affected. By comparison, the bioreleasable linker such as glutathione (GluSH) in the endosomal setting is cleaved by the intracellular thiols which is of critical importance for the conjugate action and has been the most successful approach to activated drug release within the cell (Figure 4). As an example, self-immolative disulfide and acyl hydrazone were exercised both in vitro and in vivo activity, while vintafolide analogs showed more stable amide linkers and thioether linkers did not show (Leamon et al., 2014). In addition, it was shown that by replacing DAVLBH with other clinically approved vinca alkaloid drugs (such as vincristine, vinflunine, vinorelbine and vindesine) while retaining the cleavable disulfide linker, vintafolide was the only variant showing biological activity in vitro and in vivo. It was later hypothesized that other absence of activity in other vinca alkaloid was found due to alteration in the chemical structures, following disulfide reduction and linker release.
EC145 was the first clinically tested FA-SMDC. Vintafolide showed reliability, both as a single agent and in conjunction with doxorubicin, in two phase II tests (ovarian and non-small cell lung cancers) and in a controlled open-label Phase II test (platinum-resistant ovarian cancer). It entered the clinical trials phase-III for advanced platinum-resistant ovarian cancer in 2010 (Leamon et al., 2014). This FA-SMDC failed to meet the previously stated progression-free survival requirements and the trial was prematurely terminated due to this outcome. Despite expectations of doing otherwise, the Phase III trials were unable to demonstrate the dominance of the targeted FR therapy over conventional chemotherapy. Future studies and experiments therefore need to be performed for a good range of qualifying patients who may adequately benefit from anti-FR therapy. (Leamon et al., 2014 Sausville et al., 2007; Reddy et al., 2007; Naumann et al., 2013; Naumann & Coleman, 2011; Vlahov et al., 2009; Fernandez, Javaid, & Chudasama, 2018).

**Folate-taxoid conjugates**

Seitz, 2015, developed a folate-taxoid conjugate of the next decade that is highly potent for use against drug-sensitive and drug-resistant cell lines of cancer. This folate-taxoid conjugate (Figure 5) contains both a highly potent taxoid, i.e an analog of the chemotherapeutic drug Taxol, and also folic acid moiety. This SMDC has a hydrophilic PEGylated dipeptide spacer and an autoimmolative disulfide linker, similar to vintafolide (Seitz, Vineberg, Herlihy, Park, & Melief, 2015).

In vitro research was conducted to compare the behavior of the free taxoid and taxoid conjugate in FRα-positive and FRα-negative cells. For both cell lines, i.e., free (taxoid) SB-T-1214 was highly potent as expected. Moreover, taxoid conjugate demonstrated significant cytotoxicity to the FRα-positive cell lines. In addition, taxoid conjugate also showed more than a 1000-fold decline in toxicity to normal cells compared to the free drug (Fernandez et al., 2018).

Such FA–SMDCs are from a vast field of conjugates using a disulfide linker for cytotoxic drug release. It is particularly relevant to note that folate is conjugated to many other medications, such as camptothecins (Henne, Kularatne, Hakenjos, Carron, & Henne, 2013), tubulysins (Reddy et al., 2009; Bartouskova, Melichar, & Mohelnikova, 2015), mitomycins (Leamon et al., 2007), and maytansinoids, all of which were prepared and evaluated by disulfide linker.

**5-Fluorouracil loaded PLGA-1, 3-diaminopropane-folic acid nanoparticles**

Because of a low FA conjugation ratio, (Wang et al., 2015), developed a PLGA (poly (lactic co-glycolic acid))-based drug delivery carrier with a targeting mood such as folic acid (FA) that has poor targeting performance. A crosslinker 1, 3-diaminopropane was used in this work to produce a FA-conjugated PLGA device and achieved a high 46.7% (mol / mol) conjugation level. The prepared biomaterial based on PLGA was then used for encapsulation into nanoparticles of 5-fluorouracil (5-FU). The IC50 of 5-FU loaded PLGA-1, 3-diaminopropanefolic acid nanoparticles on HT-29 cancer cells was observed to be 5.69 mg/mL in vitro experiments and is smaller than that of 5-FU loaded PLGA nanoparticles and 5-FU with only 14.17 and 22.9 mg/mL, IC₅₀ respectively. The images of fluorescent microscopy showed that targeting nanoparticles have more affinity with cancer cells, and nanoparticles with FA are taken up in greater amount than pure drugs and untreated nanoparticles, by the cancer cells with HT-29. The 5-FU primed PLGA-1, 3-diaminopropane-folic acid nanoparticles are therefore one of the highly efficient methods for the targeted delivery of the drug to the cancer cells (Wang, Li, Chen, Gao, Zeng, & Kong, 2015).

**Folic acid- trimethyl chitosan- paclitaxel conjugates**

He & Yin, 2017, developed a trimethyl chitosan (TMC-PTX) conjugated paclitaxel (PTX) modified with folic acid (FA), (FA-TMC-PTX) for oral and intravenous delivery of PTX. Modification of FA has been carried out to alleviate conjugate protein adsorption (Pawar et al., 2012; Henne et al., 2013, He & Yin, 2017).

**Folic acid coupled to the pegylated-liposomes of 5-fluorouracil**

Pegylated-liposomes coupled with folic acid were produced by Gupta et al., 2007, and they increased the in vitro absorption by up to 11 times compared to non-coupled pegylated liposomes. The tumor-inhibitory effect of FA–SL (stabilized liposome) was significantly greater than that of free 5-FU and SL. Therefore, this device can be successfully used to target 5-FU to the tumour cells. (Gupta, Jain, Jain, & Jain, 2007).

**Abrabinogalactan-folic acid- drug conjugate**

The FA-AGGFLG-MTX conjugate, illustrated by Pinhasi et al., 2009, shows the distribution of a cytotoxic cargo to FRα-overexpressing cells. They also showed, a target-activated release
mechanism by adding an endosomally cleavable peptide (GFLG) into the conjugate. Compared to normal cells, this conjugate showed a 6.3-fold increase in cytotoxic activity of cancer cells. This result creates a novel FA bound polymer nano-conjugate to transmit methotrexate to over-expressed cancer cells in FR. (Pinhasi et al., 2009).

**Cobalamin-Drug-Conjugate**

The invention provides a cobalamin-drug conjugate in which cobalamin is indirectly covalently bound to an anti-tumour drug through a cleavable linker and one or more optional spacers, and is suitable for the treatment of tumour-related diseases. Cobalamin is covalently bound by the 5'-OH of the cobalamin ribose ring to a cleavable linker or the first spacer. The drug is connected to the cleavable linker’s Second Spacer through an additional or current functional group on the surface. This conjugate then forms a transcobalamin complex (any of its isoforms). This complex binds to the receptor present on the cell membrane and is the result of cellular internalization, and the intracellular enzymes then separate the conjugates and activate the drug. Based on the conjugate form, the specific class or type of intracellular enzyme influences the cleavage. As there is a high demand of rising cells for cobalamine, tumour cells have more appetite than the normal cells. Compared to a free drug, the technology conjugate has an increased potency and reduced systemic toxicity. (Jung & Keller, 2012).

**VB12-Conjugated and PTX-Loaded Micelles (VB12-Serinc-PBLG (Poly (Gamma-Benzyl-L-Glutamate))-PTX)**

Guo et al., 2018, introduced sericin micelles to reverse drug resistance in their previous research, but these sericin micelles could not selectively bind to the cells of gastric cancer (GC). They developed sericin micelles conjugated with vitamin B12 (VB12) for the targeted treatment of gastric cancer. Researchers studied their physicochemical properties, the function of antitumours, and the cellular uptake. It was shown that VB12-sericin-PBLG-PTX micelles have an acceptable particle size, have strong dispersion, and are bio-safe. Through following transcobalamin-II (CD320)-receptor-mediated endocytosis, the micelles were internalized cellularly with gastric targeting and improved cellular uptake capacities, they reversed drug resistance, and modified the mitochondrial transmembrane / apoptosis pathway (Guo et al., 2018).

**Vitamin B12-Metal Conjugates for the Therapeutic and Diagnostic Application**

Vitamin B12 (cyanocobalamin) is an essential nutrient characterized by very poor bioavailability. Due to the rapid rate of proliferation of cancerous cells, they have a greater demand of nutrients including a greater vitamin B12 uptake compared to normal cells. Such a cyanocobalamin appetite can be used for site-specific delivery of the medication to tumour cells by conjugating the vitamin B12 (carrier) anticancer drug for therapeutic use or by conjugating a fluorophore or radionuclide diagnostic agent with vitamin B12 for diagnostic use. Vitamins such as B12 are transformed through biologically active cofactors, i.e. methylcobalamin (which is used to manufacture methionine) and adenosylcobalamin (which is used as a coenzyme to generate energy through the synthesis of carboxylic acids), and it is noteworthy that the quickly dividing tumour cell needs higher amounts of methionine and energy for replication, allowing vitamin B12 to be more aimed at cancer.

**Therapeutic vitamin B12- metal conjugates**

An impressive number of studies on platinum-based and other metal-based anticancer drugs were generated from the marketing and clinical establishment of anticancer drug cisplatin, but the main drawback associated with such treatment is low water solubility, low bioavailability, non-specific tumour cell binding, and higher toxicity. There has lately been a greater interest in the development of vitamin-metallodrug conjugates, but none of them have yet reached the clinical trial stage and have thus far only explored their therapeutic efficacy.

**Platinum:** Florea & Büsßelburg, 2011, developed a strategy to address the issue of non-specific anticancer medication binding, such as cisplatin, and its severe side effects. Conjugating cisplatin to cyanocobalamin as an anticancer medication has been speculated as a possible way of improving tumour-specific attachment with a better clinical output. The metal containing scaffolds are directly linked by the nitrogen atom to the cyanogroup present on vitamin B12, producing a heterodinuclear derivative (B12-Coll-III-M), in which the present vitamin serves as a ligand. For vitamin B12 to be transformed into its cofactor (methylcobalamin or adenosylcobalamin) requires first of all the reduction of Co (III) to Co (II) by removing the main cyanogroup. Thus, when mixing a metal product with the moiety B12-Coll-III-CN, the release of the cyanometal species (CN-M) will occur directly within the cells, so this vitamin B12-metal conjugate would be considered a prodrug.

**In vitro cytotoxicity experiments on human ovarian adenocarcinoma A2780 cells and human breast adenocarcinoma MCF7 cells have shown that the isolated Pt (II)-cyanoderivative has a comparable anti-cancer action to cisplatin, thereby indicating that conjugate (B12-Coll-III-CN-PtII) can be identified as a prodrug because it has the ability to release Pt II, a cytotoxic agent, directly into the body. Unfortunately, this hypothesis was not consistent with evidence that the initial conjugate was less cytotoxic than free cisplatin. (Ruiz-Sanchez, Konig, Ferrari & Alberto, 2011; Tran, Sturup, Lambert, Gammelgaard, & Furger, 2016; Pettenuzzo, Pigot, & Ronconi, 2017).**

**Diagnostic vitamin B12-metal conjugates**

Recent developments for diagnostic applications of vitamin B12-metal conjugates using radiodiagnostic or fluorescent samples are discussed below. A variety of biomolecules are used as a carrier for radionuclides, some of which have been approved by Food and Drug Administration, such as peptides and monoclonal antibodies. Among all of these, vitamins were researched as a target-specific delivery carrier in which vitamin B12 is the least studied.

**Gadolinium:** -Gadolinium (III) is a paramagnetic material that is used for magnetic resonance imaging (MRI) and may also be cytotoxic. Siega et al., 2009, formed two metal chelating ligands of vitamin B12, either diethylenetriamine-N, N,
N,N',N'',N'''-pentaacetic acid (DTPA) or triethyleneetetramine-N, N',N''-N,N',N'',N'''-hexaacetic acid (TTHA), and the corresponding gadolinium(III) combines Gd-1 and Gd-2 (Figure 6).

On human immortalized myelogenous leukemia K562 cells, in vitro cytotoxicity was performed; in which it was found that the Gd-1 conjugate emitted the Gd<sup>3+</sup> molecule, resulting in reduced cell viability. On the other hand, when the same test was performed of more stable Gd-2 ion, it did not show Gd<sup>3+</sup> cell internalization, and also no significant influence was observed on cell viability (Siega et al., 2009; Pettenuzzo et al., 2017).

**Vitamin D3-Drug Conjugates**

Patil, Gawali, Patil, & Basu, (2014), developed nanoparticles of vitamin D3 to deliver some of the commonly used and clinically licensed anticancer drugs such as doxorubicin (a DNA-damaging agent), paclitaxel (a microtubule-stabilizing agent) and PI103 (a phosphatidylinositol-3-kinase inhibitor). Next, they developed vitamin D3-drug conjugates (Figure 7-compound no 3, 4 and 5), then the biocompatible vitamin D3 nanoparticles were synthesized from these conjugates using a process of hydration-extrusion solvent evaporation-lipid film (Figure 8).

Such vitamin D3-nanoparticles were examined in the drug release model. A gradual and continuous release of medications over a period of time and an accelerated release was found at pH 5.5 relative to pH 7.4. Connections in vitamin-drug conjugates, i.e. vitamin-D3-PI103 conjugates, contain phenolic-ester connections, vitamin D3-paclitaxel conjugates contain ester connections, and amide-connected vitamin-D3-doxorubicin conjugates were more labile at pH 5.5 relative to pH 7.4, resulting in increased drug release at acid pH. In addition, the cellular absorption process of vitamin D3-NPs was studied in HeLa cells by treating them with VitD3-Dox-NPs. It was observed that internalization in HeLa cells was through a low pH lysosomal compartment, whereas free drug doxorubicin was internalized through the diffusion path. A similar endocytosis was expected for VitD3-paclitaxel-NPs and vitD3-PI103-NPs (Patil et al., 2014; Petros & DeSimone, 2010; Rajendran, Knolker & Simons, 2010; Iversen, Skotland, & Sandvig, 2011).

**Dual drug loaded vitamin D3 nanoparticles**

One of cancer chemotherapy’s most challenging problems is combating drug resistance. Drug cocktails can overcome drug resistance. Double vitamin D3 nanoparticles (i.e. a fair mixture of PI103 and doxorubicin or paclitaxel or cisplatin) were developed by Palvia et al. 2014, (Table 1). Nanoparticles dual drug release was observed to be higher at pH 5.5 relative to pH 7.4 at a slow rate over 72 hours in a prolonged fashion, and at 37°C and 40°C for more than 15 days. Cell death by dual drug loaded nanoparticles in human hepatocellular...
carcinoma (Hep3B cell) was found to be increased at 24 hours compared to monotherapy (Persidis, 1999; Garraway & Janne, 2012; Parhi, Mohanty, & Sahoo, 2012; Palvai, Nagraj, Mapara, Chowdhury, & Basu, 2014).

Vitamin-E-Drug-Conjugates

Vitamin-E analogue-neomycin conjugate for RNA-I drug
Iwata et al., 2015, synthesized a vitamin E-conjugated neomycin derivative to administer RNAi (RNA interference) medication to the cell of liver cancer and tested it for its biological and physicochemical properties. SiRNA delivery studies were not entirely successful, but some of the neomycin derivatives showed significant RNAi activity in liver cancer cells. (Iwata et al., 2015).

Biotin-Drug-Conjugates

Biotin-taxoid conjugates
Chen et al., 2010, developed a biotin (vitamin or vitamin B7)-taxoid conjugate for a tumour-targeted drug delivery system. These conjugates included biotin as a mood-targeted tumour, a mechanism-based self-immolative linker, and a cytotoxic agent (SB-T-1214) (Figure 9). They developed a tumour-specific mechanism for the delivery of drugs based on endocytosis mediated by vitamin receptors. Reports of L1210FR leukemia cells that over-express biotin receptors were performed. The result was an excellent site-specific delivery of the conjugates, reducing unwanted toxicity to healthy cells (Chen et al., 2010; Tripodo, Mandracchia, Collina, Rui, & Rossi, 2014).

Biotin-doxorubicin conjugates
Ibsen et al., 2010, developed and synthesized the biotin-doxorubicin conjugate in which the amine component of doxorubicin was derivatized with a photocell biotinylated spacer (Figure 10) (Tripodo et al., 2014).

Ibsen et al., 2010, demonstrated that the doxorubicin (i.e. active drug) is only released after the conjugates are internalized into the cancerous cells and then further activated via exposure to UV at 350 nm of the photocleavable group in the conjugate, leading to a decrease in the cytotoxic effects on normal cells.

The cell proliferation assay results clearly evidenced a lower cytotoxicity of conjugate than that of free doxorubicin (Sutherland & Griffin, 1981; Ibsen et al., 2010).

The brief summary of all the reported methods and their advantages over conventional chemotherapy is shown in Table 2.
Figure 10. Chemical structure of biotin-doxorubicin conjugate.

Table 2. Vitamin-drug conjugates and their advantages over chemotherapeutic drugs.

| Sr. No. | Method                                      | Advantages over chemotherapeutic drugs                                                                 |
|--------|---------------------------------------------|-------------------------------------------------------------------------------------------------------|
| 1.     | Folic acid-Paclitaxel Conjugates            | Conjugate showed higher anti-proliferative activity, Higher cellular uptake than free paclitaxel in MCF-7/PTX cells. FA-P7-PTX had a much stronger effect on cell toxicity, apoptosis and membrane disturbance behavior in MCC. FA-P3-PTX, FA-P7-PTX conjugates had more ability to suppress tumour growth than PTX. (Dai et al., 2019). |
| 2.     | Folic acid-DAVLBH (desacetyl vinblastine monohydrazine) conjugate / Vintafolide. | Vintafolide showed higher affinity to bind FRa (folate receptor-alpha), and therefore has a very strong and selective action against FRa-positive xenografts as compared to untargeted DAVLBH. (Vlahov et al., 2009). |
| 3.     | Folate-Taxoid Conjugates.                   | Taxoid conjugate demonstrated significant cytotoxicity to the FRa-positive cell lines. In addition, taxoid conjugate also showed more than 1000-fold decline in toxicity to normal cells compared to the free drug (Fernandez et al., 2018). |
| 4.     | 5-Fluorouracil Loaded PLGA-1, 3-Diaminopropane-Folic Acid Nanoparticles | Targeting nanoparticles have more affinity with cancer cells and nanoparticles with FA are taken up in greater amount than pure drugs and untreated nanoparticles, by the cancer cells with HT-29. (Wang et al., 2015). |
| 5.     | Folic Acid-Trimethyl Chitosan-Paclitaxel Conjugates | Folic acid modification in TMC-PTX alleviated the protein adsorption of conjugate. FA-TMC-PTX conjugate showed enhanced antitumor activity. (He & Yin, 2017; Pawar et al., 2012; Henne et al., 2013). |
| 6.     | Folic Acid Coupled to the Pegylated-Liposomes of 5-Fluorouracil | Pegylated-liposomes coupled with folic acid showed increased in-vitro absorption by up to 11 times compared to non-coupled pegylated liposomes. The tumour-inhibitory effect of FA–SL (stabilized liposome) was significantly greater than that of free 5-FU and SL. (Gupta et al., 2007). |
| 7.     | Abrabinogalactan-Folic Acid-Drug Conjugate. | This conjugate showed a 6.3-fold increase in cytotoxic activity of cancer cells compared to the normal cells (Pinhassi et al., 2009). |
| 8.     | Cobalamin-Drug Conjugate.                  | As there is a high demand of rising cells for cobalamine, tumor cells have more appetite than the normal cells. Compared to a free drug, the technology conjugate has an increased potency and reduced systemic toxicity. (Jung & Keller, 2012). |
| 9.     | VB12-Conjugated And PTX-Loaded Micelles (VB12-Sericin-PBLG (Poly (Gamma-Benzyl-L-Glutamate))-PTX) | Sericin micelles could not selectively bind to the cells of gastric cancer, the vitamin B12conjugated micelles were internalized celluarly with gastric targeting and improved cellular uptake capacities, reversed drug resistance, and modified the mitochondrial transmembrane / apoptosis pathway. (Guo et al., 2018). |
CONCLUSION

Such vitamin-drug conjugates identify and manipulate the inherent morphological and physiological distinctions between normal cells/tissues and cancerous ones. For example, cancer cells that are rapidly growing overexpress cancer-specific receptors to increase nutrient and vitamin intakes. These receptors can be used as targets for the targeted delivery of cytotoxic drugs to cancer cells via receptor-mediated endocytosis (RME). Our review focuses on the conventional treatment of cancer and its advantages and disadvantages. It was noted that there are many novel approaches involved in overcoming these disadvantages. Out of several formulation therapies involved, methods such as conjugation with metal ions or conjugation with several vitamins etc. The Vitamin Drug conjugates were found to be a most effective therapy. Progress in Vitamin-drug conjugates, including some of the recent vitamin-drug conjugates for target specific delivery via vitamin-receptor mediated endocytosis mechanism, reduces the undesirable toxicity to healthy cells. Some of the vitamin-metalldrug conjugates are also given, which are used for therapeutic and diagnostic applications in cancer treatment. Out of all the methods reviewed, the vitamin B12 conjugate method was found to be more specific, and also had a more positive impact on biological activity.

From the literature survey, the conjugation of vitamins with an anticancer agent is of great interest and is becoming a highly innovative approach to improving efficacy and reducing toxicity to healthy cells.

Although very few vitamin-drug conjugates have thus far been developed, this field is in constant growth, as there is a wider need for a targeted drug delivery for cancer therapy.

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