**Review**

**Clinical review: Myocardial depression in sepsis and septic shock**

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**Abstract**

Myocardial dysfunction frequently accompanies severe sepsis and septic shock. Whereas myocardial depression was previously considered a preterminal event, it is now clear that cardiac dysfunction as evidenced by biventricular dilatation and reduced ejection fraction is present in most patients with severe sepsis and septic shock. Myocardial depression exists despite a fluid resuscitation-dependent hyperdynamic state that typically persists in septic shock patients until death or recovery. Cardiac function usually recovers within 7–10 days in survivors. Myocardial dysfunction does not appear to be due to myocardial hypoperfusion but due to circulating depressant factors, including the cytokines tumor necrosis factor alpha and IL-1β. At a cellular level, reduced myocardial contractility seems to be induced by both nitric oxide-dependent and nitric oxide-independent mechanisms. The present paper reviews both the clinical manifestations and the molecular/cellular mechanisms of sepsis-induced myocardial depression.

**Keywords** contractility, cytokine, heart, myocardial depression, nitric oxide

Sepsis and septic shock have been recognized as an increasingly serious clinical problem, accounting for substantial morbidity and mortality. The past four decades have seen the age-adjusted mortality of sepsis increase from 0.5 to 7 per 100,000 episodes despite major advances in the understanding of its pathophysiology [1]. The incidence of severe sepsis in the United States today is estimated at 750,000 cases per year, resulting in 215,000 deaths annually [2]. The majority of these sepsis patients die of refractory hypotension and of cardiovascular collapse.

Sepsis has been defined as the systemic inflammatory response to infection [3]. An infectious stimulus (e.g. endotoxin or another microbiologic element) induces the release of local and systemic inflammatory mediators, especially tumor necrosis factor alpha (TNF-α) and IL-1β, from monocytes/macrophages and other cells [4]. These cytokines stimulate polymorphonuclear leukocytes, macrophages and endothelial cells to release a number of downstream inflammatory mediators, including platelet activating factor and nitric oxide (NO), further amplifying the inflammatory response. Several anti-inflammatory mediators are also released as part of this amplification cascade; namely, IL-10, transforming growth factor beta and IL-1 receptor antagonist. The relative contribution of these cytokines will determine the severity of the septic episode. If the inflammatory reaction is particularly intense, homeostasis of the cardiovascular system will be disrupted, leading to septic shock. One of the manifestations of cardiovascular dysfunction in septic shock is myocardial depression.

The present article reviews the clinical manifestations of cardiac dysfunction in sepsis, from the point of view of both the right and left ventricle, as well as cardiovascular prognostic factors in sepsis and septic shock. We will also review the
potential pathophysiologic processes responsible for myocardial depression in sepsis, from the perspective of organ physiology and molecular biology.

Clinical manifestations of cardiovascular dysfunction

Historical perspectives

Our understanding of the cardiovascular manifestations of septic shock has evolved over the years, as new techniques to assess cardiovascular performance have become available. Before the introduction of the pulmonary artery catheter (PAC), two distinct cardiovascular clinical presentations of septic shock were described: a high cardiac output (CO) state, associated with warm, dry skin and a bounding pulse despite hypotension (warm shock); and a low CO state, associated with hypotension, cold, clammy skin and a thready pulse (cold shock) [5]. Clowes et al. [6], in a 1966 study, described these two clinical pictures as different stages of septic shock: patients were believed to initially experience a hyperdynamic phase (warm shock), and then to either recover or progress to hypodynamic shock (cold shock) and death. This view was reinforced by other clinical studies [5,7] that correlated survival with high cardiac index (CI). Only a few studies hinted at the relationship between volume status, the CI and outcome [8,9]. All these studies that supported the concept of terminal cold shock suffered from the fact that they used central venous pressure (CVP) as the best available estimate of left ventricular end-diastolic volume and adequacy of resuscitation. Evidence accumulated over the past 40 years shows that CVP, as a reflection of right ventricular preload, is a poor estimate of left ventricular preload in critically ill patients, and particularly in sepsis [10].

The introduction of the PAC (which could measure pulmonary artery wedge pressure as a more accurate estimate of left ventricular preload) has allowed for better definition of the cardiovascular dysfunction in septic shock and has improved volume resuscitation. Several studies have shown that adequately resuscitated septic shock patients consistently manifest a hyperdynamic circulatory state with high CO and low systemic vascular resistance (SVR) [11,12]. In contrast to previous belief, this hyperdynamic state usually persists until death in nonsurvivors (Fig. 1) [13,14]. Despite the strong evidence characterizing sepsis as a hyperdynamic state, studies that examined myocardial performance still showed left ventricular dysfunction (illustrated by decreased left ventricular stroke work index) in properly resuscitated septic patients [16]. The depression in the Frank–Starling curve demonstrated in these studies, however, could be explained by either a change in myocardial contractility or compliance.

The development of portable radionuclide cineangiography and its application to critically ill patients has further improved our understanding of cardiovascular dysfunction in septic shock, by allowing differentiation between impaired contractility and impaired compliance.

Figure 1

The mean (± SEM) cardiac index plotted against time for all patients, survivors, and nonsurvivors. The hatched areas show the normal range. All groups maintained an elevated cardiac index throughout the study period. The difference between the survivors and nonsurvivors was not statistically significant. Reproduced with permission from [16].

Left ventricular function

Parker et al. [16] showed, using radionuclide cineangiography, that survivors of septic shock demonstrated a decreased left ventricular ejection fraction (LVEF) and an acutely dilated left ventricle, as evidenced by an increased left ventricular end-diastolic volume index (LVEDVI) (Fig. 2). These parameters returned to normal over 7–10 days in survivors. Nonsurvivors maintained normal LVEF and LVEDVI throughout the course of their illness until death. All patients in this study [16] had normal or elevated CI and low SVR, as measured by the PAC.

In 1988, Ogibene et al. compared left ventricular performance curves (plotting left ventricular stroke work index versus LVEDVI) of septic and nonseptic critically ill patients (Fig. 3). They showed a flattening of the curve in septic shock patients, with significantly smaller left ventricular stroke work index increments in response to similar LVEDVI increments when compared with nonseptic critically ill controls [17]. Subsequent studies have confirmed the presence of significant left ventricular systolic dysfunction in septic patients [18,19].
Left ventricular diastolic function in septic shock is not as clearly defined. The dilatation of the left ventricle [16] and the lack of discordance between pulmonary artery wedge pressure (PAWP) and left ventricular end-diastolic volume [17] both argue against significant diastolic dysfunction in sepsis. More recent studies using echocardiography, however, have demonstrated slower left ventricular filling [20] and aberrant left ventricular relaxation [21,22] in septic patients, suggesting that impaired compliance may significantly contribute to myocardial depression in sepsis.

**Right ventricular function**

Low peripheral vascular resistance in sepsis leads to decreased left ventricular afterload. However, the right ventricular afterload is frequently elevated due to increased pulmonary vascular resistance from acute lung injury [23]. These different physiologic conditions mean that the right ventricle cannot be expected to behave like the left ventricle in septic patients. Multiple studies have therefore specifically examined right ventricular function in sepsis.

A number of studies have documented right ventricular systolic dysfunction in volume-resuscitated septic patients, as evidenced by decreased right ventricular ejection fraction (RVEF) and right ventricular dilation [24–27]. Kimchi et al. [24] and Parker et al. [26] also showed that right ventricular dysfunction occurred independently of pulmonary vascular resistance and pulmonary artery pressure, suggesting that increased right ventricular afterload could not be the dominant cause of right ventricular depression in septic shock. Parker et al. [26] further demonstrated a close temporal parallel between right ventricular and left ventricular dysfunction in sepsis. In their study, survivors experienced significant right ventricular dilation and decreased RVEF and right ventricular stroke work index, all of which returned to normal within 7–14 days (Fig. 4). Nonsurvivors had moderate right ventricular dilation and a marginally decreased RVEF, neither of which improved throughout their illness.

There is also evidence of right ventricular diastolic dysfunction in septic patients. Kimchi et al. [24] noticed a lack of correlation between right atrial pressure and right ventricular
end-diastolic volume, suggesting altered right ventricular compliance. Schneider et al. [25] similarly identified a sub-group of patients who failed to exhibit an increased right ventricular end-diastolic volume index in response to volume loading, despite a rise in CVP. However, the relative contribu-
tion of systolic and diastolic dysfunction to right ventricular depression in sepsis remains largely unknown.

**Cardiovascular prognostic factors in septic shock**

Early studies of cardiovascular dysfunction in septic shock suggested that a low or decreasing CI invariably carried a poor prognosis [5–8]. As previously discussed, these studies relied on CVP measurements to assess volume status. It is now known that CVP is a poor reflection of left ventricular preload in critical illness and cannot accurately determine adequacy of resuscitation. Introduction of the PAC showed that adequately volume-resuscitated septic shock patients (as measured by PAWP) predictably exhibited a high CI and low SVR, including nonsurvivors [11,12]. CI is therefore not a reliable predictor of mortality in sepsis.

Recognition of the significant post-resuscitation peripheral vasodilation (low SVR) in septic shock led to the theory that peripheral vascular failure could be a major determinant of mortality in septic shock. Baumgartner et al. [28] noticed that patients with an extremely high CI (>7.0 l/min/m²) and low SVR had a uniformly poor outcome. Groeneveld et al. [29] retrospectively examined data from septic shock patients. They found that, for equivalent CI, nonsurvivors had a lower SVR than survivors. They concluded that peripheral vascular resistance was closely linked to outcome in septic shock.

Parker et al. [13] reviewed hemodynamic data from septic shock patients on presentation and at 24 hours to identify prognostic value. They found that, on presentation, only a heart rate <106 beats/min suggested a favorable outcome. At 24 hours, a heart rate <95 beats/min, a SVR index >1529 dynes s/cm²/m², a decrease in heart rate >18 beats/min and a decrease in CI >0.5 l/min/m² all predicted survival. In a subsequent study [14], the same authors confirmed previous findings of decreased LVEF and increased LVEDVI in survivors of septic shock but not in non-
survivors. However, although nonsurvivors as a whole did not exhibit left ventricular dilation, they could be divided into two groups: the first with decreasing LVEDVI and decreasing CI on serial measurements, and the other group with progressively increasing LVEDVI and maintained CI.

There are three hemodynamic patterns of death in septic shock [13]. Early deaths are due either to distributive shock (low SVR and refractory hypotension despite preserved CI) or to a cardiogenic form of septic shock (decreased CI). Late deaths are due to multisystem organ failure. Correlating this with Parker et al.’s [14] findings, it is possible that nonsur-
vivors who are unable to dilate their left ventricle (decreasing LVEDVI and CI) succumb to the cardiogenic form of septic shock. Those who have increasing LVEDVI and preserved CI die of the classic distributive shock.

The value of right ventricular performance parameters in predicting outcome is less clear. Multiple studies [24–27] have shown that, in sepsis, the right ventricle behaves similar to the left ventricle, exhibiting acute dilation and decreased RVEF. Persistence of right ventricular systolic dysfunction is associated with poor outcome [26,27]. Opinions diverge, however, on whether initial right ventricular dilation and decreased RVEF portend poor prognosis. Vincent et al. [27] suggested that survival correlates with higher initial RVEF, whereas Parker et al. [28] observed lower RVEF in survivors of septic shock than in nonsurvivors. The question requires further study.

Patients with sepsis and septic shock can show resistance to the vasopressor and inotropic effects of catecholamines. Nonsurvivors of septic shock have been shown to have an attenuated inotropic response to a dobutamine stress test compared with survivors [30]. Conversely, increased SVI, increased mixed venous oxygen saturation, ventricular dilation and drop in diastolic blood pressure in response to a dobuta-
mine stress test all predict survival in septic patients [31].

**Etiology of myocardial depression in sepsis and septic shock**

**Myocardial hypoperfusion**

The possibility of myocardial dysfunction in sepsis was originally proposed and described in the 1960s. Its etiology, however, remained a mystery. For many years, the leading theory was that sepsis was associated with a globally
decreased myocardial perfusion, leading to ischemic injury and myocardial depression. Two studies disproved that view. Cunnion et al. [32] performed serial measurements of coronary blood flow and metabolism using thermodilution coronary sinus catheters in septic patients (Fig. 5). They found normal or elevated coronary blood flow in septic patients compared with normal controls with comparable heart rates, and found no difference in blood flow between patients who developed myocardial dysfunction and those who did not. There was no net myocardial lactate production. Dhainaut et al. [33], using the same technique, confirmed these findings.

Furthermore, studies on animal models of sepsis [34] demonstrated that myocardial oxygen metabolism and high-energy phosphates were preserved in septic shock, neither of which is compatible with myocardial ischemia. However, Turner et al. [35] recently measured increased troponin I levels in patients with septic shock, demonstrating some degree of myocardial cell injury in the course of septic shock. It remains unclear from their study whether direct cardiac injury plays a role in sepsis-induced myocardial dysfunction or is the result of other factors, including a myocardial depressant substance (MDS) or exogenous catecholamine administration.

**Circulatory depressant substances**

Wiggers’ landmark report [36] in 1947 postulating the presence of a circulating myocardial depressant factor in hemorrhagic shock provided the basis for the accepted current theory of myocardial dysfunction in septic shock. The presence of a myocardial depressant factor in sepsis was later confirmed experimentally by Lefer [37] in the late 60s. Clinical studies performed at the same time associated death from septic shock with a hypodynamic circulatory state marked by a decreased CO [6–8]. Although these earlier studies depended on the measurement of CVP to assess preload, a factor now recognized as unreliable in the critically ill, Lefer’s early contributions furthered research into MDS in sepsis [37].

The first study to actually show a link between septic shock-associated myocardial depression in humans with the myocyte depressant effects of a patient’s own septic serum was carried out by Parrillo et al. in 1985 [38]. Serum from patients with septic shock generated a significant concentration-dependent depression of in vitro myocyte contractility (Fig. 6). The investigators were also able to demonstrate a strong correlation between the timing and degree of septic shock-associated decrease in LVEF in vivo and cardiac myocyte depression in vitro induced by exposure to serum from the same patients.

Subsequent work focused on identifying the presence of a MDS. In a study of 34 patients with septic shock [39], researchers noted high levels of myocardial depressant activity that correlated with higher peak serum lactate, with increased ventricular filling pressures and with increased LVEDVI. In addition, a trend towards higher mortality was present in patients with high levels of myocardial depressant activity compared with patients with lower or absent activity (36% versus 10%) [39]. These studies would argue in favor of a circulating substance rather than hypoperfusion as the causative factor in septic shock-associated myocardial depression.

**Myocardial depression in sepsis: cytokines**

Although the existence of MDS was demonstrated by the previous studies [36–39], the identity of the molecules remained in question. Potential circulating inflammatory mediators that could cause septic myocardial depression include the prostaglandin group, leukotrienes, platelet activating factor, histamine and endorphins. However, the substance was found to be heat labile, soluble in water, and its activity in filtration studies was present in the >10 kDa fraction [39]. Although full molecular characterization was not possible with the available data, the characteristics were most consistent with either a polypeptide or protein. The list of potential
cytokine mediators of myocardial depression is exhaustive; however, TNF-α and IL-1β play a central role and deserve further consideration.

TNF-α shares a similar biochemical profile with MDS, making it a plausible mediator of the myocardial effects of sepsis and septic shock [38,40,41]. Experimentally, increased levels of TNF-α produce fever, lactic acidosis, disseminated intravascular coagulation, acute lung injury and death. The cardiovascular effects are similar to clinical septic shock; namely, hypotension, increased CO and low SVR [42,43]. Human volunteers given TNF-α infusions demonstrate similar responses [44,45].

A number of studies have shown that, when TNF-α is administered to human and animal myocardial tissue in vitro or ex vivo, the result is a concentration-dependent depression of contractility [46–49]. Furthermore, removal of TNF-α from the serum of patients with septic shock partially eliminates its myocardial depressant effect [47]. Although larger phase III clinical trials have shown no overall survival benefit when anti-TNF-α monoclonal antibody has been administered to patients with septic shock, left ventricular function did improve in this patient group [50].

IL-1β induces similar pathophysiologic responses to TNF-α. IL-1β is increased in both human and animal models of sepsis and septic shock [51]. When exposed to IL-1β, in vitro as well as ex vivo myocardial contractility is depressed [49,52,53]. Immunoabsorption of IL-1β partially neutralizes cardiac myocyte depressant activity of human septic serum [47]. Administration of IL-1β antagonist attenuates the hemodynamic and metabolic manifestations of septic shock [54,55]. As with TNF-α, overall mortality has not been improved in randomized trials with IL-1β [54,55].

A number of studies have postulated that cytokine synergy plays a key role in septic myocardial depression. When considering IL-1β and TNF-α independently, substantially higher concentrations of each cytokine are required to depress contractility in rat cardiac myocytes than when these two cytokines are combined [47]. These findings have also been validated in ex vivo studies of isolated human atrial trabeculae [56]. The combination of TNF-α and IL-1β may cause myocyte depression at concentrations 50–100 times lower than would be required if applied individually. Such concentrations are well within those found in blood during human septic shock. Available data supports a causative role for TNF-α and IL-1β acting synergistically in septic myocardial depression.

**Cellular mechanisms of septic myocardial depression**

Depression of myocardial contractility by TNF-α, IL-1β and septic serum in vitro occurs in two distinct time frames. The early phase of cardiac myocyte depression occurs within minutes of exposure either to TNF-α, to IL-1β, to TNF-α and IL-1β given together or to septic serum [47,48]. In vivo studies have shown that treatment with anti-TNF-α and IL-1β monoclonal antibodies in the acute phase of sepsis renders the myocardium refractory to the depressant effects of these cytokines [50].

Figure 6

The effect of serum from septic shock patients and control groups on the extent of myocardial cell shortening of spontaneously beating rat heart cells in vitro. Septic shock patients during the acute phase demonstrated a statistically significant lower extent of shortening (P<0.001) compared with any other group. Reproduced with permission from [38].
canine studies also demonstrate the ability of TNF-α to induce rapid myocardial depression [43,57]. Furthermore, there is a convincing relationship between the degree of in vitro early cardiac myocyte depression produced by human septic serum and the decrease in LVEF from those same patients during acute septic shock [38]. Other studies also document a delayed depressant effect of TNF-α, IL-1β and supernatants of activated macrophages on in vitro myocardial tissue [52,53,57,58]. This effect begins hours after exposure and may persist for days. This late phase of myocardial depression appears to occur by a distinctly different biochemical pathway than the early depression, and may involve de novo protein synthesis.

The generation of NO may be central to both early and late depression of myocytes in in vitro and in vivo models of septic shock. The role of NO in cardiac contractility is still under study. NO is produced by the conversion of L-arginine to L-citrulline by the nitric oxide synthase, which exists in inducible nitric oxide synthase (iNOS) and constitutive nitric oxide synthase (cNOS) forms. Studies support the role of NO generated by cNOS in the physiologic regulation of cardiac contractility [59–61]. In vitro studies of cardiac myocytes superperfused with either NO, nitroprusside (NO donor) or the NO donor SIN-1 demonstrate reductions in myocardial contractility [62]. In human studies, nitroprusside infusion into coronary arteries depresses intraventricular pressure while improving diastolic relaxation and distensibility [63].

In sepsis and septic shock, the pathophysiologic production of NO may contribute to cardiovascular dysfunction. The initial process is thought to occur from sequential NO and cGMP generation via cNOS activation rather than by de novo synthesis of iNOS. This is consistent with the short time frame to the onset of myocardial depression [48,56,64]. These studies cannot, however, account for the late-onset of cytokine-driven myocardial depression discussed earlier. Further studies have shown, in in vitro myocardial preparations, that the generation of iNOS, NO and cGMP may be responsible for late onset myocardial depression [48,53,65,66].

Studies thus suggest that NO-mediated depression of myocardial contractility appears in two separate time frames by two distinct pathways. Prior to Kinugawa et al’s study [67], it might have been thought these mechanisms were mutually exclusive. Kinugawa et al. demonstrated, using an avian cardiac myocyte model, a biphasic response to IL-6 administration. IL-6 given in high concentration produced both early (<30 min) and late (24 hours) cardiac myocyte depression, associated with a decrease in intracellular calcium. Early depression could be blocked by pretreatment with a calcium chelator, implicating that the calcium/calmodulin-dependent cNOS is involved. However, late depression could not be blocked by the chelator, supporting the role for the calcium/calmodulin-independent iNOS.

The study of Kinugawa et al. clearly suggests that cytokines (or sepsis) can stimulate the heart to sequentially produce NO by both cNOS and iNOS. The data would suggest that TNF-α, IL-1β and possibly other cytokines involved in septic shock could mediate their cardiodepressant effects by at least two distinct mechanisms. Early cardiodepressant activity may involve both a NO-dependent but β-adrenoreceptor-independent mechanism and a NO-independent defect of β-adrenoreceptor signal transduction [68]. Late depression involves an iNOS-dependent defect of β1-adrenorenergic signal transduction with further addition of an undefined defect of myocardial contractility. While NO generation may contribute to the cardiodepressant effects seen in sepsis, its precise role and the role of other mechanisms underlying septic myocardial dysfunction continue to be defined.

**Conclusion**

Myocardial dysfunction is an important variable in sepsis and septic shock. The impairment in cardiac function results from a combination of systolic and diastolic dysfunction. The presence of a circulating MDS, which probably represents different cytokines, including TNF-α and IL-1β, acting synergistically is clear. The myocardial depressant effect of these cytokines is at least partly mediated through mechanisms of NO generation. While the current standard of therapy in patients with septic shock is still directed towards re-establishing organ and tissue perfusion and oxygen delivery by fluid resuscitation and inotropic support, ongoing research aimed at understanding the cellular mechanisms of cardiac dysfunction may lead to exciting new therapeutic options.

**Competing interests**

None declared.

**References**

1. Center for Diseases Control and Prevention: National Center for Health Statistics: mortality patterns — United States, 1990. *Monthly Vital Stat Rep* 1993, 41:5.
2. Angus DC, Linde-Zwirble WT, Lidicker J, Clermont G, Carcillo J, Pinsky MR: Epidemiology of severe sepsis in the United States: analysis of incidence, outcome and associated costs of care. *Crit Care Med* 2001, 29:1303-1310.
3. Bone RC, Balk RA, Cerra FB, Dellinger RP, Fein AM, Knaus WA, Schein RM, Sibbald WJ: Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. *Crit Care Med* 1992, 101:1644-1655.
4. Van der Poll T, Van Deventer JH: Cytokines and anticytokines in the pathogenesis of sepsis. *Infect Dis Clin North Am* 1999, 13:413-426.
5. MacLean LD, Mulligan WG, McLean APH, Duff JH: Patterns of septic shock in man: a detailed study of 56 patients. *Ann Surg* 1967, 166:543-562.
6. Clowes GHA, Vucinic M, Weidner MG, Circulatory and metabolic alterations associated with survival or death in peritonitis. *Ann Surg* 1966, 163:844-866.
7. Nishijima H, Weil MH, Shubin H, Cavanilles J: Hemodynamic and metabolic studies on shock associated with gram-negative bacteremia. *Medicine (Baltimore)* 1973, 52:287-294.
8. Weil MH, Nishijima H. Cardiac output in bacterial shock. *Am J Med* 1978, 64:920-922.
9. Blain CM, Anderson TO, Pietras RJ, Gunnar RM: Immediate hemodynamic effects of gram-negative vs gram-positive bacteremia in man. *Arch Intern Med* 1970, 126:260-265.
10. Packman ML, Rackow EC: Optimum left heart filling pressure during fluid resuscitation of patients with hypovolemic and septic shock. Crit Care Med 1983, 11:165-169.

11. Winslow EJ, Loeb HS, Rahimtoola SH, Kamath S, Gunnar RM: Hemodynamic studies and results of therapy in 50 patients with bacteremic shock. Am J Med 1973, 54:421-432.

12. Krauss MM, Perel A, Eimerl D, Cotev S: Cardiopulmonary effects of volume loading in patients with septic shock. Ann Surg 1977, 185:429-434.

13. Parker SM, Shellhammer JH, Natanson C, Alling DW, Parrillo JE: Serial cardiovascular variables in survivors and non-survivors of human septic shock: heart rate as an early predictor of prognosis. Crit Care Med 1987, 15:923-929.

14. Winslow EJ, Loeb HS, Rahimtoola SH, Kamath S, Gunnar RM: Hemodynamic studies and results of therapy in 50 patients with bacteremic shock. Am J Med 1973, 54:421-432.

15. Krauss MM, Perel A, Eimerl D, Cotev S: Cardiopulmonary effects of volume loading in patients with septic shock. Ann Surg 1977, 185:429-434.

16. Winslow EJ, Loeb HS, Rahimtoola SH, Kamath S, Gunnar RM: Hemodynamic studies and results of therapy in 50 patients with bacteremic shock. Am J Med 1973, 54:421-432.

17. Ognibene FP, Parker MM, Natanson C, Shelhamer JH, Parrillo JE: Depressed left ventricular performance. Response to volume infusion in patients with sepsis and septic shock. Chest 1988, 93:903-910.

18. Ellrodt AG, Riedinger MS, Kimchi A, Berman DS, Maddahi J, Swan HJC, Murata GH: Left ventricular performance in septic shock: reversible segmental and global abnormalities. Am Heart J 1985, 110:402-409.

19. Raper RF, Sibbald WJ, Driedger AA, Gerow K: Relative myocardial depression in normotensive sepsis. J Crit Care 1989, 4:9-18.

20. Jafri SM, Lavine S, Field BE, Thill-Baharozian MC, Carlson RW: Left ventricular diastolic function in sepsis. Crit Care Med 1991, 18:709-714.

21. Munt B, Jurj S, Strick W, sclercolling L, Pfeft. Right ventricular systolic stiffness in septic shock. J Appl Physiol 1990, 69:1329-1335.

22. Poelert A, De Clerck C, Vogelaers D, Collard VN, Visser CA: Left ventricular diastolic filling in septic shock. Circ Shock 1991, 38:27-34.

23. Bibb WA, Paterson N, Mallory DE, Anderson RA, Lobd TR, Duff JH: Pulmonary hypertension in sepsis: measurement by the pulmonary artery diastolic-pulmonary wedge pressure gradient and the influence of passive and active factors. Chest 1990, 97:833-839.

24. Kimchi A, Ellrodt GA, Berman DS, Murata GH, Riedinger MS, Swan HJC, Murata GH: Right ventricular performance in septic shock: a combined radiouclide and hemodynamic study. J Am Coll Cardiol 1984, 4:945-951.

25. Schneider AJ, Teuini AF, Natanson C, Ognibene FP, Shellhammer JH, Parrillo JE: Responses of left ventricular function in survivors and non-survivors of septic shock. J Crit Care 1989, 4:19-25.

26. Wiegand RD, Vito L, Dennis RC, Hechtman HB: Myocardial Intensive Care Med 1997, 23(5):560.

27. Parker MM, McCarthy KE, Ognibene FP, Parrillo JE: Right ventricular dysfunction and dilatation, similar to left ventricular changes, characterise the cardiac depression of septic shock in humans. Chest 1990, 97:126-131.

28. Vincent JL, Reuse C, Frank N, Contrempe B, Kahn RJ: Right ventricular dysfunction in septic shock: assessment by measurements of right ventricular ejection fraction using the echodensitometrie technique. Acta Anaesthesiol Scand 1989, 33:34-38.

29. Bussmann J, Vaney C, Perrot C: An extreme form of hyperdynamic syndrome in septic shock. Intensive Care Med 1984, 10:245-249.

30. Groeneweld ABJ, Nauta JJ, Thijssen L: Peripheral vascular resistance in septic shock: its relation to outcome. Intensive Care Med 1988, 14:141-147.

31. Schupp E, Kumar A, Bunnell E, Uretz E, Calvin J, Ali A, Parrillo JE: The cardiovascular response to incremental doses of dobutamine in septic shock: prediction of survival [abstract]. Crit Care Med 1994, 22:A109.

32. Cunin RE, Schaar GL, Parker MM, Natanson C, Parrillo JE: The coronary circulation in human septic shock. Circulation 1986, 73:837-844.

33. Dhainaut JJ, Huygebaert MF, Monsallier JJ, Leferve G, Dall’Avana S, Picciotto J, Brunet F, Villenfant D, Carli A, Raichvar D: Coronary hemodynamics and myocardial metabolism of lactate, free fatty acids, glucose and ketones in patients with septic shock. Circulation 1987, 75:533-541.

34. Solomon MA, Correa R, Alexander HR, Koeve LA, Cobb JP, Kim DII, Roberts WC, Quezado ZMN, Schott TD, Cunin RE, Hoffman WD, Bacher J, Tatis I, Danner RL, Banks SM, Ferrans VJ, Babalan RS, Natanson C: Myocardial energy metabolism and morphology in a canine model of sepsis. Am J Physiol 1994, 266:H757-H768.

35. Escher A, Tsamitros M, Bellomo R: Myocardial cell injury in septic shock. Crit Care Med 1999, 27:1775-1780.

36. Wiggers CJ: Myocardial depression in shock. A survey of cardiodynamics studies. Am J Heart 1947, 33:633-650.

37. Lefler AM: Mechanisms of cardiodepression in endotoxin shock. Circ Shock 1978, 1:197-208.

38. Parrillo JE, Burch C, Shellhammer JH, Natanson C, Scuette W: A circulating myocardial depressant substance in humans with septic shock. Septic shock patients with a reduced ejection fraction have a circulating factor that depresses in vitro cardiac cell performance. J Clin Invest 1985, 76:1539-1553.

39. Reilly JM, Cunin RE, Burch-Whitman C, Parker MM, Shellhammer JH, Parrillo JE: A circulating myocardial depressant substance is associated with cardiac dysfunction and peripheral hypoperfusion (lactic acidemia) in patients with septic shock. Chest 1989, 95:1072-1080.

40. Cunin RE, Parrillo JE: Myocardial dysfunction in sepsis. Recent Insights. Chest 1989, 95:941-945.

41. Schepker P, Vey E, Turcatti G, Wiegfield P, Dayer JM: Tumor necrosis factor inhibitor: purification, NH2-terminal amino acid sequence and evidence for anti-inflammatory and immunomodulatory activities. Eur J Immunol 1990, 20:1167-1174.

42. Eichacker PQ, Hoffman WD, Farese A, Banks SM, Kuo GC, MacVitie TJ, Natanson C: TNF but not IL-1 in dogs causes lethal lung injury and multiple organ dysfunction similar to human sepsis. J Appl Physiol 1991, 71:1979-1989.

43. Eichenholz PW, Eichacker PQ, Hoffman WD, Banks SM, Parrillo JE, Danner RL, Natanson C: Tumor necrosis factor challenges in canines: patterns of cardiovascular dysfunction. Am J Physiol 1992, 263:H668-H675.

44. van der Poll T, van Deventer SJ, Hack CE, Woblink GJ, Aarden LA, Butler HR, ten Cate H: Effects on leukocytes following injection of tumor necrosis factor into healthy humans. Blood 1992, 79:693-698.

45. van der Poll T, Romijn JA, Endert E, Born JJ, Butler HR, Sauerwein HP: Tumor necrosis factor mimics the metabolic response to acute infection in healthy humans. Am J Physiol 1991, 261:C447-C465.

46. Gu M, Bose R, Bose D, Yang J, Li X, Light RB, Mink S: Tumor necrosis factor-alpha but not septic plasma depresses cardiac myofilament contraction. Can J Anesth 1998, 45:280-286.

47. Kumar A, Thota V, Dee L, Olson J, Uretz E, Parrillo JE: Tumor necrosis factor-alpha and interleukin-1-beta are responsible for depression of in vitro myocardial cell contractility induced by serum from humans with septic shock. J Exp Med 1996, 183:949-958.

48. Finkels OS, Oddis CV, Jacobs TD, Watkins SC, Hattler BG, Simmons RL: Negative inotropic effects of cytokines mediated by nitric oxide. Science 1992, 257:387-389.

49. Weisneder D, Berenter-Hahn J, Low-Friedrich I: Effects of cytokines on the contractility of cultured cardiac myocytes. Int J Immunopharmacol 1993, 15:581-587.

50. Vincent JL, Bakker J, Marecax G, Schandene L, Kahn RJ, Dupont E: Administration of anti-TNF antibody improves left ventricular function in septic shock patients: results of a pilot study. Chest 1992, 101:810-815.
51. Hesse DG, Tracey KJ, Fong Y, Manogue KR, Palladino MA, Cerami A, Shines GT, Lowry SF: Cytokine appearance in human endotoxemia and primate bacteremia. Surg Gynecol Obstet 1988, 166:147-153.

52. Gulick T, Chung MK, Piejer SJ, Lange LG, Schreiner GF: IL-1 and TNF inhibit cardiac myocyte adrenergic responsiveness. Proc Natl Acad Sci 1989, 86:6753-6757.

53. Hosenpud JD, Campbell SM, Mendelson DJ: Interleukin-1 induced myocardial depression in an isolated beating heart preparation. J Heart Transplant 1989, 8:460-464.

54. Fisher CJ, Dhainaut JF, Opal SM, Pribble JP, Balk RA, Slotman GJ, Iberti TJ, Rackow EC, Shapiro MJ, Greenman RL, Reines HD, Shelly MP, Thompson BW, LaBrecque JF, Catalano MA, Knaus WA, Saddoff JC: Recombinant human IL-1 receptor antagonist in the treatment of patients with the sepsis syndrome: results from a randomized, double blind placebo controlled trial. JAMA 1994, 271:1836-1843.

55. Fisher CJ, Jr., Slotner GJ, Opal SM, Pribble JP, Bone RC, Emmanuel G, Ng D, Bloedow DC, Catalano MA: Initial evaluation of human recombinant IL-1 receptor antagonist in the treatment of the sepsis syndrome: a randomized, open-label, placebo-controlled multicenter trial. Crit Care Med 1994, 22:12-21.

56. Cain BS, Meldrum DR, Dinarello CA, Meng X, Joo KS, Banerjee A, Harken AH: Tumor necrosis factor-α and interleukin-1 β synergistically depress human myocardial function. Crit Care Med 1999, 27:1309-1318.

57. Walley KR, Hebert PC, Wakai Y, Wilcox PG, Road JD, Cooper DJ: Decrease in left ventricular contractility after tumor necrosis factor-α in infusion in dogs. J Appl Physiol 1994, 76:1060-1067.

58. DeMeules JE, Pigula FA, Mueller M, Raymond SJ, Gamelli RL: Tumor necrosis factor and cardiac function. J Trauma 1992, 32:686-692.

59. Hare JM, Keaney JF, Balligand JL, Loscalzo J, Smith TW, Colucci WS: Role of nitric oxide in parasympathetic modulation of beta-adrenergic myocardial contractility in normal dogs. J Clin Invest 1995, 95:360-366.

60. Sawyer DB, Colucci WS: Nitric oxide in the failing myocardium. Cardiol Clin North Am 1998, 16:657-664.

61. Balligand JL, Kobzik L, Han X, Kaye DM, Belhassen L, O’Hara DS, Kelly RA, Smith TW, Michel T: NO-dependent parasympathetic signaling is due to the activation of constitutive endothelial (type III) nitric oxide synthase in cardiac myocytes. J Biol Chem 1995, 270:14582-14586.

62. Brady AJ, Warren JB, Poole-Wilson PA, Williams TJ, Harding SE: Nitric oxide attenuates cardiac myocyte contraction. Am J Physiol 1993, 265:H176-H182.

63. Paulus WJ, Vantrimpont PJ, Shah AM: Acute effects of nitric oxide on left ventricular relaxation and diastolic distensibility in humans. Assessment by biconorony sodium nitroprusside infusion. Circulation 1994, 89:2070-2078.

64. Kumar A, Brer R, Wang P, Doe L, Skorupa G, Khadour F, Schulz R, Parrillo JE: The role of nitric oxide and cyclic GMP in human septic serum-induced depression of cardiac myocyte contractility. Am J Physiol 1999, 276:R265-R276.

65. Rozanski GJ, Witt RC: IL-1 inhibits beta-adrenergic control of the cardiac calcium current: role of L-arginine/nitric oxide pathway. Am J Physiol 1994, 267:H1753-H1758.

66. Smith JA, Radomski MW, Schulz R, Moncada S, Lewis MJ: Porcine ventricular endocardial cells in culture express the inducible form of nitric oxide synthase. Br J Pharmacol 1993, 108:1107-1110.

67. Kinugawa K, Takahaashi T, Kohmoto O, Yao A, Aoyagi T, Monomura S, Hirata Y, Serizawa T: Nitric oxide-mediated effects of IL-6 on [Ca2+]i, and cell contraction in cultured chick ventricular myocytes. Circ Res 1994, 75:285-295.

68. Kumar A, Krieger A, Symmans S, Kumar A, Parrillo JE: Myocardial dysfunction in septic shock: Part II. Role of cytokines and nitric oxide. J Cardiothorac Vasc Anesth 2001, 15:485-511.

69. Parillo JE, Parker MM, Natanson C, Suffredini AF, Danner RL, Cunnion RE, Ognibene FP: Septic shock in humans. Advances in the understanding of pathogenesis, cardiovascular dysfunction and therapy. Ann Intern Med 1990, 113:227-242.