Comparison of laparotomic omentopexy vs. laparoscopic abomasopexy treatments of left displaced abomasum in dairy cows under field conditions: biochemical analysis

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Abstract

The objective of this retrospective study was to compare the effects of the two methods (laparoscopic or laparotomic) of LDA (left displaced abomasum) correction under field conditions by means of survival rate and biochemical profile evaluation. Holstein cows from one farm with LDA over a 20-month period were included in the study. Cows underwent laparoscopic abomasopexy (Janowitz’s method, LS) or a laparotomic right flank omentopexy (Dirksen’s method, LT). Blood samples for biochemical analysis were taken just prior to surgery (D1) and on days 10 (D10) and 30 (D30) following surgery. Blood profiles of healthy cows identified from the farm’s routine transition cow monitoring program were used as control (C). Aiming to minimize disruptive effects of quite a long period of data collection, the cows’ biochemical profile was evaluated in three orthogonal comparisons: LS cows vs. control group, LT cows vs. control group, and LS cows vs. LT cows. The rate of animal survival was similar for both methods (loss of about 11% until 30 days after treatment). Most of the blood parameters from LS and LT groups were comparable in all three sampling terms indicating continual organism recovery. At D10 the difference of higher cholesterol concentration and lower haptoglobin concentration were favourable for the LS group (P < 0.05). Total protein, calcium, magnesium and albumin showed more positive dynamics in the LS group too. This proves previous laboratory and clinical indices of expediency of LDA laparoscopy treatment under field conditions.

Abomasal displacement, LDA, cattle, laparotomy, laparoscopy, recovery, surgery

Left displaced abomasum (LDA) is considered as an important production disease of high producing dairy cows primarily related to the periparturient period, a challenging time when cows undergo changes in their metabolism and are more susceptible to diseases (Markusfeld 1986; Pistkova et al. 2019). Reported prevalence of LDA is approximately 2% on well-managed farms (Sickingner 2017). Left displaced abomasum disrupts the passage of digesta with continuing excretion of hydrochloric acid. This scenario has a negative impact on acid-base and ion balance, dehydration, and the total nutritional balance of cow. Affected animals can suffer from hypokalaemia, hyponatraemia and hypochloraemia together with metabolic alkalosis. Dehydration and haemoconcentration are identified. At the same time catabolic processes occur manifested by high blood urea concentration as a final product of protein catabolism, as well as by hypoproteinaemia and hypoalbuminaemia which sometimes reflect protein synthesis depression too (Delgado-Lecaroz et al. 2000; Constable et al. 2013). Indices of liver dysfunction as well as of negative energy balance (NEB) are documented in affected cows (Whitaker 2004). An increase of haptoglobin in cows with LDA indicates inflammation processes (Simões et al. 2017). Contemporarily, NEB, namely higher beta
hydroxybutyrate (BHB) concentration, is considered as an important predisposing factor of LDA (LeBlanc et al. 2005). Another predisposing factor is hypocalcaemia (Neves et al. 2018).

Surgery is a well-documented mode of LDA correction (Niehaus 2016). Right flank laparotomic omentopexy (Dirksen 1967) is a commonly used method. A disadvantage of the method is substantial invasion into the organism. Laparoscopic methods are generally less invasive and less painful (Newman 2009). Among them, ventral abomasopexy according to Janowitz (1998) is the most common approach of laparoscopic LDA correction under field conditions.

Limited numbers of studies compared the effectiveness of laparoscopic ventral abomasopexy with laparotomic right flank omentopexy according to Dirksen. Wittek et al. (2009) mentioned improved recovery of rumen and abomasum motility and quicker return of milk production, although these results were only numerically but not significantly different. In a study focused on indicators of peritoneal inflammation, laparoscopy was evaluated as a favourable method (Wittek et al. 2012). In a university clinical study, Seeger et al. (2006) found significant differences in normalization of bilirubin concentration and glutamate dehydrogenase (GDH) activity which were more favourable for animals after laparoscopic surgery. In contrast, Roy et al. (2008) did not find any significant differences in all parameters monitored in their field study.

The aim of the study was to verify by assessment of biochemical profile whether laparoscopic abomasopexy and laparotomic omentopexy procedures for LDA correction had similar recovery success for cows under field conditions.

**Materials and Methods**

The study was performed as a retrospective data analysis of herd and veterinary records from a farm of 1000 high-producing Holstein cows with an average of 11,500 kg milk production. Processed data come from the period of 20 months during which both methods of LDA surgery – laparotomic omentopexy (LT; Dirksen 1967) and ventral laparoscopic abomasopexy (LS; Janowitz 1998) – were performed.

**Animals**

Cows indicated for LDA surgery with no other intercurrent disease were randomly assigned to the LT or LS group. Nine cows ranging from first to fourth lactation were subjected to LT. The LDA was diagnosed on day 1 to day 22 after parturition. The LS method was used in 44 cows ranging from first to sixth lactations. The LDA was diagnosed on day 4 to day 48 after parturition. Blood samples for biochemical analyses were taken on the day of surgery (D1) and day 10 (D10) and 30 (D30) after surgery to monitor recovery. A group of 26 cows that were closest to the age and days in milk (DIM) of the experimental cows and assessed as not having LDA or any other major health concern, were selected as controls (C) and their data from the herd’s transition cow monitoring program were used.

**Biochemical analysis**

Biochemical parameters of harvested serum were used for this study. Fluid colorimetry (analyser Erba Lachema XL200) with the use of commercial kits (Erba Lachema, s.r.o., Brno, Czech Republic) were used for analysis non-esterified fatty acids (NEFA), triacylglycerols (TAG), cholesterol (CHOL), total protein (TP), albumin (ALB), urea (UREA), gamma-glutamyl transferase (GGT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), phosphorus (P), calcium (Ca) and magnesium (Mg). Electrolytes, sodium (Na), potassium (K) and chloride (Cl), were analysed by ionic selective electrodes on the same analyser. Haptoglobin (HAPT) was analysed by commercial kit (Haptoglobin Bovine, Tridelta Development, Ireland), beta-hydroxybutyrate (BHB) was analysed by commercial kit (RANDOX NEFA, RANDOX RANBUT, Laboratories Ltd., UK) and glucose (GLU) was analysed by commercial kit (GLUL 500 S, PLIVA – Lachema diagnostika s.r.o., Brno, Czech Republic).

Data by Pechova et al. (2009) were used as reference limits for individual biochemical parameters.

**Statistical analysis**

For basic surgical outcome analysis chi-squared test was used with a $P$ value < 0.05.

Because of the relatively long period of data collecting (20 months), biochemical parameters were evaluated under three partial comparisons, always comparing animals of the same period of sample collection: laparoscopy operated cows vs. control group (LS × C), laparotomy operated cows vs. control group (LT × C) and laparoscopy operated cows vs. laparotomy operated cows.
operated cows vs. laparotomy operated cows (LS × LT). Thirty-seven animals were included in LS × C analysis (LS – 12, C – 25). Thirteen animals were included in LT × C analysis (LT – 5, C – 8). Seventeen animals were included in the LS × LT analysis (LS – 11, LT – 6).

Data underwent exploratory data analysis focusing on detecting outliers and verifying the normality of distribution of all dependent variables. The data that were not normally distributed (NEFA, BHB, TAG, CHOL, UREA, GGT, AST, ALP, K, P, HAPT) were transformed by natural logarithms. Data were evaluated by mixed linear model with repeated measures procedure MIXED (version 9.3; SAS Institute Inc., Cary, NC). Parameters were estimated by the REML method. The model was structured to determine the effect of period and the combined effect of group (LS, LT, C) and day after surgery. Due to repeated measures within each cow, random (co)variances between days after surgery were summarized by residual R matrix. As alternatives, the compound symmetry, heterogeneous compound-symmetry, unstructured, autoregressive of order 1 and the Toeplitz covariance structures were compared. The compound-symmetry (GLU, NEFA, TAG, CHOL, TP, ALB, UREA, GGT, Cl, P, Ca) and heterogeneous compound-symmetry (Na, K, Mg, BHBlog, ASTlog, ALPlog, HAPTlog) structures were found to be the most appropriate in accordance with Akaike’s Information Criterion and the Schwartz Bayesian Criterion (Littell et al. 2000). The least square means were calculated, and multiple comparisons were made in simple-effect slices, with $P$ values adjusted using Tukey’s procedure. The least squares method (LSM) of transformed variables and their confident limits (α = 0.05) were retransformed to the original scale.

**Results**

In the LS group 5 cows died within 30 days after surgery, 5 animals were culled before finishing the 305-d lactation and 37 cows finished the 305-d lactation. One LT cow died, and 8 cows finished the 305-d lactation. There was no significant difference in these losses in terms of surgical procedure.

Results of serum biochemical analyses of cows that underwent laparoscopy (LS group) and results of a related control group are shown in Table 1. Results of serum biochemical analyses of cows that underwent laparotomy (LT group) compared to the control group are shown in Table 2. Comparisons of serum parameter values between LS and LT groups are shown in Table 3. Parameters without any significance (effect of group or time course) in a particular analysis are not shown.

**Discussion**

The rate of basic surgical intervention outcome was similar for both of the methods verified. The loss of about 11% is also comparable to published papers (Seeger et al. 2006; Roy et al. 2008).

The biochemical profile indicates metabolic changes typical of LDA but also a metabolic overload typical of the transition period: On D1, evidence of NEB in conjunction with lipomobilization was obvious both for LT and LS cows with NEFA and BHB concentrations elevated above limits (> 0.35 mmol/l and > 1.0 mmol/l, respectively) and low CHOL concentration (limits 2.6–5.2 mmol/l). Cows with BHB concentrations higher than 1.0 mmol/l are at higher risk of health issues or decreased milk productivity (Whitaker 2004). Evaluation of LS and C cows shows also a significant increase of TAG in diseased cows (limits 0.17–0.51 mmol/l). This one-off finding is probably related to higher NEFA values of diseased animals from this selected dataset (LSM = 1.15 mmol/l), as TAG are synthesized in the liver from circulated NEFA (Illek et al. 2011).

These biochemical changes are considered not only as a consequence of LDA but also as a risk factor for LDA in high producing cows after calving (Van Winden and Kuiper 2003; LeBlanc et al. 2005). Concentrations of NEFA, BHB and CHOL indicated that C cows also underwent NEB, although not as severe as LDA cows. For this reason, on D30 (i.e., one month from the operation plus an additional month from calving) blood serum values returned to normal ones not only at LDA groups but at C groups as well. However, the mean values of BHB on D30 are generally close to the upper limit, which reflects ongoing NEB in many cows at this period. Because these cows were 30–70 DIM, we consider this as a normal situation (Whitaker 2004; Pechova et al. 2009).
The lipomobilization effect and starvation in dairy cows are associated with a higher load and decrease of synthetic activity of the liver parenchyma (Staufenbiel et al. 2007). This was reflected by high AST activity or by low values of TP, ALB and UREA. Comparison of LS and LT groups (Table 3) on D10 showed that there is a certain delay of negative effect of LDA on UREA concentrations in both groups and also on the ALB value (this for the LT group only).

Hypocalcaemia as an important predisposing factor is often diagnosed in cows with LDA (Neves et al. 2018). Our dataset indicates that both cows with LDA and cows in the control group suffered from subclinical hypocalcaemia (Ca limits: 1.6–2.25 mmol/l). After surgical correction Ca concentrations returned to normal values. Lower than normal concentrations were also observed for other ions, namely Na, Cl, K and Mg while comparable values were found in the LDA groups and the control group. Occasional significant changes in time (Mg increase or Na decrease) were not related to LDA. Hypokalaemia was found only in the LT group in the LT × C analysis. Electrolytes Na, K, and Cl are considered as indicators of disease severity. Higher concentrations of Na and K are presented in animals with doubtful or poor prognosis. Hypokalaemia is considered as a consequence of retaining K in the digestive system (Rohn et al. 2004).

### Table 1A. Biochemical profile of cows operated using laparoscopy vs. control.

| Indicator (mmol/l) | Day | Laparoscopy | Control |
|-------------------|-----|-------------|---------|
|                   |     | Mean | SE  | Mean | SE  |
| NEFA (mmol/l)     | 1   | 1.15 |      | 0.74 | 1.77 |
|                   | 30  | 0.12 | 1.85 | 0.18 | 0.36 |
| BHB (mmol/l)      | 1   | 2.14 | 1.39 | 3.28 | 0.62 |
|                   | 30  | 0.63 | 1.15 | 0.84 | 0.69 |
| TAG (mmol/l)      | 1   | 0.17 | 0.47 | 0.18 | 0.69 |
|                   | 30  | 0.11 | 0.10 | 0.13 | 0.13 |
| CHOL (mmol/l)     | 1   | 1.63 | 1.26 | 2.10 | 2.50 |
|                   | 30  | 3.55 | 2.73 | 4.62 | 4.14 |
| AST (μkat/l)      | 1   | 2.72 | 2.13 | 3.46 | 1.45 |
|                   | 30  | 1.05 | 0.87 | 1.26 | 1.15 |
| HAPT (g/l)        | 1   | 1.20 | 0.88 | 1.62 | 0.55 |
|                   | 30  | 0.49 | 0.29 | 0.81 | 0.57 |

^- mean values with the same superscript within the row differ significantly (P < 0.05)

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Table 1B. Biochemical profile of cows operated using laparoscopy vs. control.

| Indicator (mmol/l) | Day | Laparoscopy | Control |
|-------------------|-----|-------------|---------|
|                   |     | Mean | SE  | Mean | SE  |
| Na                | 1   | 127.53 | 1.85 | 125.02 | 1.37 |
|                   | 30  | 119.82 | 2.95 | 121.18 | 2.19 |
| Ca                | 1   | 2.05 | 0.07 | 2.25 | 0.05 |
|                   | 30  | 2.20 | 0.08 | 2.39 | 0.0 |
A common finding in LDA groups was a highly significant increase of HAPT on D1 in comparison to the control group and its decrease in time. Haptoglobin is one of the acute phase proteins considered as an important indicator of inflammation in the organism (Simões et al. 2017).

Direct comparison of animals treated with the LS method as opposed to LT revealed that most of the blood indices in the LS and LT groups were comparable at all three sampling terms, but on D10 the differences of higher CHOL and lower HAPT concentrations were favourable for the LS group. There was also a quicker normalization of other parameters in the LS group. The total protein, Ca, Mg and ALB even showed a significant decrease on D10 in the LT group (but not in the LS group). These findings agree with Wittek et al. (2009), who documented a more rapid normalization of abomasum and rumen motility after laparoscopy or with the Wittek et al. (2012), where the same team - based on indicators of peritoneal inflammation - evaluated laparoscopy as probably less stressful and more favourable method. Also, Seeger et al. (2006) found significant differences in normalization of bilirubin level and activity of GDH favourable for animals with laparoscopic surgery. Nevertheless, all these studies were conducted under clinical or even

| Indicator | Day | Laparotomy | Control |
|-----------|-----|------------|---------|
| NEFA (mmol/l) | 1 | 0.67<sup>1</sup> | 1.63 | 0.21<sup>a</sup> |
| | 30 | 0.22 | 0.31 | 0.17 | 0.09 |
| BHB (mmol/l) | 1 | 1.70<sup>1</sup> | 0.72 | 0.38 | 1.35 |
| | 30 | 0.49<sup>1</sup> | 0.23 | 0.74 | 0.03 |
| CHOL (mmol/l) | 1 | 1.51<sup>a</sup> | 0.74 | 0.38 | 1.35 |
| | 30 | 3.98<sup>1</sup> | 5.80 | 4.65<sup>1</sup> | 3.56 |
| UREA (mmol/l) | 1 | 2.77 | 5.02<sup>a</sup> | 4.05 | 6.23 |
| | 30 | 3.09<sup>1</sup> | 4.31 | 4.05 | 6.23 |
| GGT (μkat/l) | 1 | 0.27<sup>1</sup> | 0.28 | 0.21 | 0.09 |
| | 30 | 0.39<sup>1</sup> | 0.32 | 0.25 | 0.18 |
| HAPT (g/l) | 1 | 1.15<sup>A</sup> | 0.65 | 0.32<sup>a</sup> | 0.23 |
| | 30 | 0.37<sup>1</sup> | 0.75 | 0.30 | 0.18 |
### Table 3A. Biochemical profile of cows operated using laparoscopy vs. cows operated using laparotomy.

| Indicator | Day | Laparoscopy | Laparotomy |
|-----------|-----|-------------|------------|
|           | Mean | SE          | Mean | SE          |
| NEFA (mmol/l) | 1    | 0.8        |        | 1.32       | 0.66 | 1.25 |
|           | 10   | 0.24       |        | 0.72       | 0.25 | 0.72 |
|           | 30   | 0.12       |        | 0.56       | 0.25 | 0.56 |
| BHB (mmol/l)  | 1    | 2.02       |        | 2.20       | 0.09 | 0.09 |
|           | 10   | 0.67       |        | 0.65       | 0.09 | 0.09 |
|           | 30   | 0.16       |        | 0.15       | 0.09 | 0.09 |
| CHOL (mmol/l) | 1    | 63.80      |        | 68.00      | 4.71 |
|           | 10   | 74.84      |        | 73.88      | 4.75 |
|           | 30   | 80.35      |        | 76.90      | 4.97 |
| UREA (mmol/l) | 1    | 2.02       |        | 2.20       | 0.09 | 0.09 |
|           | 10   | 0.67       |        | 0.65       | 0.09 | 0.09 |
|           | 30   | 0.16       |        | 0.15       | 0.09 | 0.09 |

**Note:**
- Mean values with different superscripts within the row differ significantly ($P < 0.05$).
- Mean values with different superscripts within the column differ significantly ($P < 0.01$).

### Table 3B. Biochemical profile of cows operated using laparoscopy vs. cows operated using laparotomy.

| Indicator | Day | Laparoscopy | Laparotomy |
|-----------|-----|-------------|------------|
|           | Mean | Lower | Upper | Mean | Lower | Upper |
| TP (g/l)   | 1    | 0.8    | 1.32  | 0.66 | 0.34  | 1.25  |
|           | 10   | 0.12   | 0.25  | 0.16 | 0.16  | 0.25  |
|           | 30   | 0.16   | 0.25  | 0.15 | 0.16  | 0.25  |
| ALB (g/l)  | 1    | 2.02   | 2.20  | 0.09 | 0.09  | 0.09  |
|           | 10   | 0.67   | 0.65  | 0.09 | 0.09  | 0.09  |
|           | 30   | 0.16   | 0.15  | 0.09 | 0.09  | 0.09  |

**Note:**
- Mean values with different superscripts within the row differ significantly ($P < 0.05$).
- Mean values with different superscripts within the column differ significantly ($P < 0.01$).

**Abbreviations:**
- NEFA - non-esterified fatty acids; BHB - beta-hydroxybutyrate; CHOL - cholesterol; UREA - urea; GGT - gamma-glutamyl transferase; AST - aspartate aminotransferase; ALP - alkaline phosphatase; HAPT - haptoglobin
laboratory conditions. Contrary to them, a study by Roy et al. (2008) under field conditions did not find any significant differences in any of the tested parameters.

In conclusion, we showed that although the changes in the biochemical profile related to cows’ recovery from LDA were predominant, their dynamics were more favourable for laparoscopic correction of LDA (Janowitz’s method) compared to laparotomy (Dirksen’s method). It confirms previous laboratory and clinical indices of expediency of LDA laparoscopic treatment under field conditions.

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