The CD31 molecule: a possible neuroprotective agent in acute ischemic stroke?

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Abstract

Background: The transmembrane receptor molecule CD31 is known to have immunomodulatory functions, suggesting a possible neuroprotective effect in the context of acute ischemic stroke by restricting an over-activation of secondary immunological processes. This study examines the density of CD31 + cells in mechanically extracted thrombi of stroke patients with the aim to test whether the occurrence of CD31 + cells was associated with a beneficial clinical outcome in those patients.

Methods: Thrombi of 122 consecutive patients with large anterior circulation stroke were collected during intracranial mechanical recanalization. Out of these, 86 immunostained specimens of adequate quality could be analysed. The density of CD31 + cells was quantified and compared with clinical outcome data of the affected patients.

Results: The density of CD31 + cells was positively related to early patient improvement (ΔNIHSS, r = 0.283, p = 0.012) with an even clearer relationship after exclusion of patients who died in the early hospital phase (r = 0.371, p = 0.001). This finding stayed stable also in the multivariate analysis after correction for other outcome-influencing factors (p = 0.049).

Conclusion: This study shows a stable relation between CD31 + cells and early clinical improvement of patients with acute ischemic stroke. This finding is in line with recent reports showing immunomodulatory and potential neuroprotective effects of CD31, suggesting that CD31 may be a promising neuroprotective agent in stroke patients.

Keywords: Stroke, Mechanical thrombectomy, Thrombus, Histology, CD31, Inflammation

Background

Inflammatory processes in the brain tissue after acute ischemic stroke are an important determinant of clinical course and patient outcome. Furthermore, increasing evidence highlights a strong linkage between thrombus formation and inflammatory pathways [1]. These assumptions are widely accepted, but with the results of the big recanalization studies being taken into clinical practice the opportunity is now open to recognizing the magnitude of the effect of these inflammatory processes with their inherent therapeutic possibilities.
One study addressing this question found no clear relation between clinical parameters and CD4+ T cells and CD68+ monocytes [5] in stroke thrombi. Another study found higher CD3+ T cells in clots from strokes of atherothrombotic origin compared to all other causes, without association to outcome parameters [6].

Recent reports suggest immunomodulatory effects of the cell adhesion molecule CD31 (PECAM-1) [7], supported by results that could show protective effects in the context of cardiovascular [8] and atherothrombotic [9] diseases. Here, we examined the presence and density of CD31+ cells in mechanically extracted thrombi of stroke patients. We aimed to test whether the occurrence of CD31+ cells was associated with a beneficial clinical outcome in those patients.

**Methods**

Between 10/2010 and 09/2012, mechanically extracted thrombi of 122 patients with acute anterior circulation stroke were prospectively collected and processed as previously described [3] (all patients with anterior circulation stroke from our last published series [10]).

As primary endpoint, we investigated the relation between the density of CD31+ cells in the thrombus tissue and clinical outcome parameters.

HE-staining and semi-automated quantification of main thrombus components (fibrin/platelet aggregations [F/P], red blood cells [RBC], white blood cells [WBC]) of the paraffin-embedded thrombi were carried out as previously described [3]. CD31 immunostaining was done using an monoclonal mouse antibody, clone JC70A (Dako Deutschland GmbH, Hamburg, Germany), dilution 1:100. Primary antibody staining was carried out overnight at room temperature. Prior to primary antibody staining, incubation with 10% normal goat serum in 1x phosphate buffered saline (PBS) was performed. An affinity purified biotinylated goat anti-mouse IgG antibody (Vector Laboratories, Burlingame, CA) was used as antiserum followed by avidin-peroxidase (Vectastain ABC; Vector). Diaminobenzidine (Invitrogen, Carlsbad, CA) was used as chromogen substrate. Negative control sections without primary antibodies were processed in parallel.

In a first step, all immunostained specimens were independently screened by two experienced investigators and assigned to a 3-point quality scale from 0 = insufficient through 1 = sufficient to 2 = good quality. All insufficient stainings were excluded, resulting in a total of 86 analyzable samples. Total numbers and distribution of CD31+ cells was quite heterogeneous between both different specimens and different regions of the same specimen. Hence, CD31+ cells were independently quantified by two investigators using a semiquantitative 5-point scale (0 = no cells, 1 = sporadic, 2 = few, 3 = some, 4 = many cells). The readers were blinded with regard to clinical and interventional data. In cases of discrepancies, results were reached by consensus.

As primary outcome parameter, the National Institute of Health Stroke Scale (NIHSS) was determined on admission and at discharge by the respective responsible neurologist, the modified Rankin Scale (mRS) up to 90 days post stroke was assessed by personal clinical examination, by phone interview, or by evaluating the medical reports of the rehabilitation centers.

Basic clinical and interventional data including age and sex, blood leukocyte count at admission and at day 1, occlusion site, time from symptom onset to reperfusion, number of retraction maneuvers, application of intra venous alteplase, stroke etiology according to the Trial of Org 10172 in Acute Stroke Treatment (TOAST) [11] classification, and recanalization success according to the thrombolysis in cerebral infarction (TICI) score [12] were assessed, as summarized in Table 1.

The two-sided Spearman’s rank correlation coefficient was used to assess correlations between the amount of CD31+ cells and primary clinical variables and outcome variables (NIHSS at admission and at discharge, ΔNIHSS between admission and discharge, mRS until 90 days). Differences in CD31+ densities were assessed by the nonparametric Kruskal–Wallis test, and boxplot diagrams were generated. For multivariate analysis, a logistic regression

| Table 1 Clinical patient characteristics |
|-----------------------------------------|
| Characteristic                  | n  |
| Age (years): median, range          | 72, 21–92 |
| Sex                                   |     |
| Female                                | 40 (47%) |
| Occlusion site                        |     |
| MCA                                   | 50 (58%) |
| ICA including carotid-T               | 22 (26%) |
| Combined ICA and MCA/ACA              | 13 (15%) |
| ACA                                   | 1 (1%)  |
| Baseline NIHSS (range, median)       | 15, 2–27 |
| IV rtPA                               | 57 (66%) |
| Time symptom onset to reperfusion (min): median, range | 20 (0–495) |
| mRS (90 days)                         | 41 (47.7%) |
| 0–2                                   | 17 (39.1%) |
| > 2                                   | 24 (60.9%) |
| Stroke etiology (TOAST)              |     |
| 1 = arterioembolic                    | 16 (18.8%) |
| 2 = cardioembolic                     | 40 (47.1%) |
| 4 = other determined cause            | 9 (10.6%)  |
| 5 = cryptogenic                      | 20 (23.5%) |

Basic demographic and clinical patient characteristics (MCA middle cerebral artery, ICA internal carotid artery, ACA anterior cerebral artery, IV rtPA intravenous recombinant tissue plasminogen activator)
model was used to calculate the effect of several factors (age, recanalization success, NIHSS at admission, and time from symptom onset to reperfusion) on a dichotomized outcome parameter. Dichotomization was performed according to previous reports [13] using the following scheme: either a difference between baseline NIHSS and NIHSS at discharge ≥8 or NIHSS at discharge ≤1 as cutoff between major or minor neurological improvement. SPSS Statistics version 23.0 (SPSS Inc., IBM, Ehningen, Germany) was used for all statistical analyses.

Results

Demographic and clinical patient data
Most extracted thrombi (58%) were from isolated occlusions of the medial cerebral artery (MCA); the dominant stroke etiology was cardioembolic (47.1%); and most patients were heavily affected with a median NIHSS at admission of 15. Two-thirds of the patients received intravenous thrombolysis. Baseline clinical and interventional data of all 86 patients with specimens of sufficient staining quality are summarized in Table 1.

Correlation analyses
Quantitative analyses of main thrombus components showed median proportions of RBCs of 47% (3–96%), F/P of 44% (2–89%), and WBCs of 6% (1–25%).

The initial correlation analysis (Spearman-Rho, two-sided) showed no association between the amount of CD31+ cells and other histopathological parameters like RBC, F/P and WBC count inside the clot or leukocyte count in the peripheral blood. Additional group comparisons showed no significant differences between the CD31+ cell density groups concerning the amount of other clot components (RBC: $p = 0.373$, F/P: $p = 0.276$, WBC: $p = 0.833$).

There were also no correlations with the initial clinical appearance (NIHSS pre-treatment) or the clinical status at discharge (NIHSS post). However, a positive correlation with NIHSS improvement (ΔNIHSS) was apparent ($r = 0.283$, $p = 0.012$). To exclude possible coincidental or interfering effects, an additional multivariate logistic regression model was used that included age, recanalization success, received thrombolytic therapy, NIHSS at admission, and time from symptom onset to reperfusion as possible relevant outcome-influencing factors. Dichotomization of NIHSS improvement was done as described earlier.

In the multivariate analysis, the effect of CD31+ cells on NIHSS improvement was, although shortly not significant, still visible after correcting for the factors mentioned above ($p = 0.057$), as well as the effects of age ($p = 0.051$) and time to reperfusion ($p = 0.002$). Successful recanalization showed no effect on early patient improvement in the multivariate analysis, probably due to few cases with unsuccessful recanalization (TICI 0–2a, $p = 0.999$), as well as NIHSS at admission ($p = 0.569$) and received thrombolytic therapy ($p = 0.497$).

In a second step, assuming that this possible positive effect may not manifest itself completely if the patient suffers from a major bleeding or malignant infarction, we performed an additional subgroup analysis with the purpose of further specifying the group of patients who may have the largest benefit of the described effect. Therefore, we excluded all patients who died in the early stroke phase up to discharge ($n = 7$), resulting in a patient subgroup of 79 patients, according to the scheme shown in Fig. 1.

As expected, the observed relation between the NIHSS improvement and the CD31+ count was even more evident in the subgroup correlation ($r = 0.371$, $p = 0.001$) and multivariate analysis ($p = 0.049$). The results of the correlation analysis of CD31+ cells and outcome parameters are summarized in Table 2.

Figure 2 shows representative clot samples with low and high densities of CD31+ cells as well as the boxplot analysis illustrating relationship and variance of the amount of CD31+ cells with regard to early clinical patient improvement (ΔNIHSS):

Discussion

To our concern this study contains by far the largest number of patients with an immunohistochemical analysis of CD31+ cells in mechanically retrieved clots. We can show a clear relationship of the amount of CD31+ cells in extracted stroke thrombi with early clinical improvement. First, it seems surprising that one certain cell type inside the vessel-occluding thrombus material should have a measurable impact on patient outcome parameters. On the other hand, several published data as well as pathophysiological considerations support the plausibility of our results.

The CD31 molecule is a receptor protein expressed by multiple cell types involved in coagulatory and immunological processes (endothelial cells, platelets, and all types of leukocytes). Recent reports indicate a broad spectrum of different functions of this molecule [7], possibly affecting secondary processes in the course of acute stroke. Most importantly, CD31 seems to be able to modulate focal and excessive neuroinflammatory processes in the early post-stroke phase without affecting patients’ immunocompetence in general. This can be achieved by two main modes of action: Adhesional functions of CD31 might be required to re-establish endothelial integrity, which is compromised, for example, by hypoxic stress [7, 14]. In this context, Graesser et al. [15] showed that CD31 can protect against excessive neuroinflammation, presumably by limiting the migration of
immunocompetent cells in the extravasal compartment. Additionally, and presumably more important, is the signaling function of CD31 leading to raised activation thresholds of lymphocytes (T and B cells) and subsequent decreased production of inflammatory mediators [16]. This downregulation of inflammatory pathways within ischemic brain lesions may protect stressed neurons against overactivated microglia [17] (by reducing their activation level). These well-known processes can lead to secondary infarct growth in the first few days after the initial stroke. Therefore, as CD31 very probably can inhibit these detrimental secondary effects in the early post-stroke phase, the observed positive effect on early patient improvement seems plausible from a pathophysiological point of view. Of course do these assumptions apply, if the amount of CD31 inside the thrombus material reflects its concentration in the peripheral blood which is supposable but has to be confirmed in further studies.

However, the observed early beneficial effects do not carry over to a better outcome after 3- months. From our initial patient cohort 3-month mRS values were available in only about half of these cases (34 patients), so perhaps the effect is simply masked due to the low patient number. Additionally, although the mRS value represents the standard outcome parameter in most stroke studies, it is known that this parameter is of limited accuracy compared with the NIHSS score, especially in older patients [18], probably also contributing to the loss of an observable long-term-effect.

A possible neuroprotective effect of CD31 in acute stroke is of special interest, as two studies showed the dependence of the immunmodulating effect of CD31 on the molecular integrity of the extracellular domains [19]. If this integrity is lost by enzymatic shedding, it seems to be possible to re-activate the immunmodulating signaling pathway by administering a CD31 derived peptide, which binds at the extracellular domain six of the truncated molecule [20]. This in turn implicates a possible neuroprotective treatment option in acute stroke patients, especially targeting the acute hyperinflammatory reaction in post-ischemic brain parenchyma in the early post-stroke phase. To follow-up, we plan to analyze CD31 counts (soluble parts and cell-bound molecules) in the peripheral blood of acute stroke patients prospectively. In addition, the neuroprotective effect of the CD31 based peptide will be evaluated in a mouse model of acute stroke.

**Limitations**

Although our study contains the largest published series of immunohistochemically analyzed thrombi to date, the loss of nearly one-third of the initially included patients might implicate a potential selection bias. As the major cause for exclusion of the patients was low tissue quality (mostly due to technical staining difficulties, variable preservation times after retrieval, and in some cases not

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**Table 2** Correlation analysis

|          | CD31   | NIHSS pre | NIHSS post | ΔNIHSS | mRS 90 days |
|----------|--------|-----------|------------|--------|-------------|
| Correlation coefficient (r) | Significance (p) | n         |            |        |             |
| CD31     | 1.00   | 0.035     | −0.151     | 0.371  | −0.095      |
|          | —      | 0.767     | 0.201      | 0.001  | 0.593       |
|          | 79     | 75        | 72         | 34     |             |

Correlation analysis of clinical outcome parameters and the amount of CD31⁺ cells. (Bold type of the ΔNIHSS values indicate significance)
enough thrombus material), a systematic bias concerning the target endpoint seems extremely unlikely.

For quantification of the amount of CD31+ cells, we decided to use a semiquantitative 5-point-grading scale rather than manual or automated cell counting. This approach may be questioned as unusual, but, in our experience, exact quantification of immunostainings of thrombus specimens by automated or manual cell counting is a complicated and error-prone process: First, stroke thrombi are of very different sizes, leading to problems in including all thrombus fragments in the counted fields without including overlapping regions and doubled countings. Secondly, the immunostainings are often inhomogeneous in the staining intensity, between different specimens as well as on one slide. In our opinion, a grading system is most suitable to mitigate these kind of difficulties and to avoid a "pseudo-accuracy" that is not covered by the underlying data quality. Nevertheless, as complete standardization of immunohistochemical methods is not feasible, some misassignments to the specified groups are possible due to staining inhomogenities.

**Conclusion**

This study shows a stable relation between CD31+ cells and early clinical improvement of patients with acute ischemic stroke. This finding is in line with recent reports showing immunomodulatory and potential neuroprotective effects of CD31, suggesting that CD31 may be a promising neuroprotective agent in stroke patients.

**Abbreviations**

ACA: Anterior cerebral artery; F/P: Fibrin/platelet aggregations; ICA: Internal carotid artery; IV rPA: Intravenous recombinant tissue Plasminogen Activator; MCA: Middle cerebral artery; mRS: Modified rankin scale; NIHSS: National Institute of Health Stroke Scale; RBC: Red blood cells; TICI: Thrombolysis in Cerebral Infarction; TOAST: Trial of Org 10172 in Acute Stroke Treatment; WBC: White Blood Cells

![Fig. 2](image_url) Relation between CD31+ cells and early patient improvement. Example of an immunostained specimen with almost no stained nucleated cells (a) and an example with many CD31+ stained cells (b). In this example, there is almost no "staining-negative" cell visible. c Boxplot analysis with regression line in red, showing the relationship between amount of CD31+ cells (0 = no cells, 1 = sporadic, 2 = few, 3 = some, 4 = many) and early patient improvement (represented by ΔNIHSS)
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Availability of data and materials
The dataset used and analysed during the current study is available from the corresponding author on reasonable request.

Author's contributions
TBB and HP: conception and design of the study. TBB analyzed and interpreted the clinical and histological patient data. SS and LS performed the histological examination of the specimen and analyzed the data. JK performed the statistical analyses. TBB wrote the manuscript. JK, LS, JK, CZ and HP were major contributors in writing the manuscript. All authors read, revised critically and approved the final manuscript.

Competing interests
The authors declare that they have no competing interests.

Consent for publication
Not applicable.

Ethics approval and informed consent
This study was approved by the local ethics committee at the Klinikum rechts der Isar of the Technical University of Munich, Germany, in accordance with the ethical standards of the 1964 Declaration of Helsinki and its later amendments. Written and informed consent of patients has been obtained.

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References
1. De Meyer SF, Denomme F, Langhauser F, Geuss E, Fluri F, Kleinschmit C. Thromboinflammation in stroke brain damage. Stroke. 2016;47(4):1165–72.
2. Chamorro A, Meisel A, Planas AM, Urra X, van de Beek D, Veltkamp R. The immunology of acute stroke. Nat Rev Neurol. 2012;8(7):401–10.
3. Boech-Behrens T, Schubert M, Forschler A, Prothmann S, Kreiser K, Zimmer C, Riegger J, Bauer J, Neff F, Kehl V, et al. The impact of histological clot composition in embolic stroke. Clin Neuroradiol. 2016;26(2):189–97.
4. Huber T, Kleine JF, Kaesmacher J, Bette S, Poppert H, Zimmer C, Boech-Behrens T. Blood leukocytes as prognostic parameter in stroke thrombectomy. Cerebrovasc Dis. 2016;42(1–2):32–40.
5. Schuhmann MK, Gurrenbein L, Kleinschmit C, Kraft P. Immunohistochemical analysis of cerebral thrombi retrieved by mechanical thrombectomy from patients with acute ischemic stroke: Int J Mol Sci. 2016;17(3):298.
6. Dargazani C, Rigau V, Eker O, Riquelle Bareiro C, Machi P, Gasco G, Arquzian C, Aylinmaci Y, Moursand I, Corlode A, et al. High CD3+ cells in intracranial thrombi represent a biomarker of atherothrombotic stroke. PLoS One. 2016;11(5):e0154945.
7. Marelli-Berg FM, Clement M, Mauro C, Caligiuri G. An immunologist’s guide to CD31 function in T-cells. J Cell Sci. 2013;126(Pt 11):2343–52.
8. Flégé D, Severino A, Troita F, Previtero M, Ucci S, Zara C, Pedicino D, Massaro G, Biasucci LM, Liuzzo G, et al. Altered CD31 expression and activity in helper T cells of acute coronary syndrome patients. Basic Res Cardiol. 2014;109(6):448.
9. Caligiuri G, Rossignol P, Julia P, Groyer E, Mouradian D, Urban D, Misra N, Ollivier V, Sapoval M, Bouteuyrie P, et al. Reduced immunoregulatory CD31 + T cells in patients with atherosclerotic abdominal aortic aneurysm. Arterioscler Thromb Vasc Biol. 2006;26(3):618–23.
10. Boech-Behrens T, Kleine JF, Zimmer C, Neff F, Scheipl F, Pelsiek J, Schirmer L, Nguyen K, Carats D, Poppert H. Thrombus Histology Studies Cardioembolic Cause in Cryptogenic Stroke. Stroke. 2016;47(7):1864–71.
11. Adams Jr HP, Bendixen BH, Kappelle LJ, Biller J, Love BB, Gordon DI, Marsh 3rd EE. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. TOAST. Trial of Org 10172 in Acute Stroke Treatment. Stroke. 1993;24(1):35–41.
12. Hijishita RA, Furlan AJ, Roberts H, Connors B, Barr J, Dillon W, Warach S, Broderick J, Tilly B, et al. Trial design and reporting standards for intra-arterial cerebral thrombolysis for acute ischemic stroke. Stroke. 2003;34(8):109–17.
13. Kerr DM, Fulton RL, Lees KR, Collaborators V. Seven-day NIHSS is a sensitive outcome measure for exploratory clinical trials in acute stroke: evidence from the Virtual International Stroke Trials Archive. Stroke. 2012;43(5):1401–3.
14. Biswas P, Canosa S, Schoenfeld D, Schoenfeld J, Li P, Cheas LC, Zhang J, Cordova A, Sumpio B, Madri JA. PEACAM-1 affects GSK-3beta-mediated beta-catenin phosphorylation and degradation. Ann J Pathol. 2006;169(1):314–24.
15. Graesser D, Soloviej A, Bruckner M, Osterweil E, Juedes A, Davis S, Ruddle NH, Engelhardt B, Madri JA. Altered vascular permeability and early onset of experimental autoimmune encephalomyelitis in PEACAM-1-deficient mice. J Clin Invest. 2002;109(3):383–92.
16. Newton-Nash DK, Newman PJ. A new role for platelet-endothelial cell adhesion molecule-1 (CD31): inhibition of TCR-mediated signal transduction. J Immunol. 1999;163(2):682–8.
17. Brown GC, Neher J. Microglial phagocytosis of live neurons. Nat Rev Neurosci. 2014;15(4):209–16.
18. Kleine JF, Boech-Behrens T, Prothmann S, Zimmer C, Liebig T. Discrepancy between early neurological course and mid-term outcome in older stroke patients after mechanical thrombectomy. J Neurointerv Surg. 2016;8(7):671–6.
19. Fornasa G, Groyer E, Clement M, Dimitrov J, Compain C, Gaston AT, Varthaman A, Khallou-Laschet J, Newman DK, Graff-Dubois S, et al. TCR stimulation drives cleavage and shedding of the ITIM receptor CD31. J Immunol. 2010;184(10):5485–92.
20. Fornasa G, Clement M, Groyer E, Gaston AT, Khallou-Laschet J, Morvan M, Guerd K, Kaven SV, Tedgui A, Michel JB, et al. A CD31-derived peptide prevents angiotensin II-induced atherosclerosis progression and aneurysm formation. Cardiovasc Res. 2012;94(1):30–7.