Blue versus Red Light Can Promote Elongation Growth Independent of Photoperiod: A Study in Four Brassica Microgreens Species

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Abstract. An elongated stem has beneficial effects on microgreen production. Previous studies indicate that under 24-hour light-emitting diode (LED) lighting, monochromatic blue light, compared with red light, can promote plant elongation for some species. The objective of this study was to investigate whether shortened photoperiod can change blue vs. red light effects on elongation growth. The growth and morphology traits of arugula (Brassica erucA, ‘Rocket’), cabbage (Brassica oleracea, unknown variety name), mustard (Brassica juncea, ‘Ruby Streaks’), and kale (Brassica napus, ‘Red Russian’) seedlings were compared during the stage from seeding to cotyledon unfolding under two light quality x two photoperiod treatments: 1) R, monochromatic red light (665 nm) and 2) B, monochromatic blue light (440 nm) using continuous (24-hour light/0-hour dark) or periodic (16-hour light/8-hour dark) LED lighting. A photosynthetic photon flux density of ~100 μmol·m⁻²·s⁻¹ and an air temperature of ~22 °C was used for the preceding treatments. After 7 to 8 days of lighting treatment, regardless of photoperiod, B promoted elongation growth compared with R, as demonstrated by a greater stem extension rate, hypocotyl length, or petiole length in the tested microgreen species, except for mustard. The promotion effects on elongation were greater under 24- vs. 16-hour lighting in many cases. Among the tested species, mustard showed the lowest sensitivity in elongation response to B vs. R, which was independent of photoperiod. This suggests that the blue-light-promoted elongation is not specifically from 24-hour lighting, despite the varying promotion degree under different photoperiods or for different species. The elongation growth promoted by blue LED light under a photoperiod of either 24 hours or 16 hours can potentially benefit indoor production of microgreens.

Microgreens are consumed when cotyledons are fully expanded, retaining their typical color, with or without appearance of the first true leaves (Jones-Baumgardt et al., 2019; Kyriacou et al., 2016). Microgreens can be grown in varying scenarios, including outdoor, greenhouse, and indoor environments (Kyriacou et al., 2016). LED lighting has been increasingly used as a sole light source for indoor production of vegetables such as microgreens (Kozai et al., 2015).

Most microgreens are harvested at 7 to 21 d from seeding with a minimum height of ~5 cm (Kyriacou et al., 2016). Also, commercial microgreen production has been increasingly switching from hand- to machine-harvesting to reduce labor cost. Microgreens with hypocotyls <5 cm are difficult for machine-harvesting, according to the communication with some Canadian growers. It is well known that both red and blue light can mediate stem elongation (Haché-Thériel et al., 2016). Also, monochromatic red and blue LED lights have been successfully used for microgreen cultivation with the advantage of increasing beneficial phytochemicals such as antioxidants (Kopsell and Sams, 2013; Wu et al., 2007). However, limited information is available on the effect of monochromatic red and blue LED lights on stem elongation of microgreens, especially under different photoperiods, because photoperiod can also affect this plant trait (Bergstrand, 2017).

Previous studies indicated that under LED lighting at a photosynthetic photon flux density (PPFD) of 100 μmol·m⁻²·s⁻¹ or 50 μmol·m⁻²·s⁻¹, monochromatic blue vs. red light promoted elongation growth in all the tested bedding plant species, including petunia, calibrachoa, geranium, and marigold (Kong et al., 2018), and some microgreen species such as arugula, cabbage, and kale (Kong et al., 2019). In these studies, a photoperiod of 24 h was used for lighting treatments. Possibly the blue-light-promoted elongation growth is an artifact specifically from 24-h lighting because it is well known that most plants grow naturally under a periodic light/dark environment. However, similar promotion effects by blue vs. red light have also been achieved under a photoperiod of <24 h (i.e., 12–18 h) in other LED studies on seedlings of eggplant (Hirai et al., 2006), cherry tomato (Kim et al., 2014), cucumber (Hernández and Kubota, 2016), marigold (Heo et al., 2002), and sunflower (Schwend et al., 2015) at a PPFD of ~100 μmol·m⁻²·s⁻¹. Thus, under a certain range of light levels (e.g., 100 μmol·m⁻²·s⁻¹), the promoted stem elongation growth by monochromatic blue light, relative to red light, might be a common phenomenon when photoperiod varied between 12 and 24 h. However, this speculation needs confirmation because the preceding studies were performed with different species under different environments. For indoor production, 16-h lighting daily has become popular (Kozai, 2018). When photoperiod is shortened from 24 h to 16 h, it is unknown whether blue vs. red LED lighting at a PPFD of 100 μmol·m⁻²·s⁻¹ could also promote elongation growth for some microgreen species.

Shortened photoperiod is known to reduce elongation for some species (Bergstrand, 2017; Schüssler and Bergstrad, 2012). Also, a recent study on petunia indicated that the stem elongation was not promoted by blue vs. red light until the exposure duration increased up to 5 d, and the blue light promotion was proportional to the lighting duration time (Fukuda et al., 2016). It is possible that shortening photoperiod within a certain range may reduce, rather than eliminate, promotion effects of blue vs. red light on plant elongation at least in some species. However, in that study, the petunia plants developed expanded true leaves, so both photosynthesis and photomorphogenesis were involved in blue vs. red light effects on plant elongation under different lighting duration. For microgreens without appearance of true leaves, photomorphogenesis is the main contributor to plant elongation. Therefore, it needs confirmation in the microgreens that shortened photoperiod from 24 h to 16 h can reduce blue light promotion effect on elongation to some degree.

Arugula, cabbage, kale, and mustard are popular species used for microgreen production. In a previous study on these microgreens with unfolded true leaves under 24-h lighting, different species varied in their elongation-promotion response to blue vs. red light at a PPFD of 100 μmol·m⁻²·s⁻¹, showing a lower sensitivity in mustard than other species (Kong et al., 2019). However, it needs confirmation that microgreens without appearance of true leaves (i.e., from seeding to cotyledon unfolding) also differ in the blue light response among species. In addition to promoted elongation, some other typical shade-avoidance responses such as reduced side branch number and cotyledon size and increased biomass allocation to main stem also occurred under blue vs. red light, which
varied with different species (Kong et al., 2018, 2019). It was concluded that blue-light-promoted elongation is a shade-avoidance response with varying sensitivity among species (Kong et al., 2018, 2019). Unfortunately, the conclusion was drawn from the studies on some bedding plants and microgreens under 24-h lighting, although the shade-avoidance responses to blue light have been also reported for Arabidopsis under a photoperiod of 24 h (de Wit et al., 2016; Keller et al., 2011; Pedmale et al., 2016). For microgreens, it is still unclear whether the species difference in blue light’s effect on elongation as a shade-avoidance response could also be found under noncontinuous (e.g., 16-h) lighting.

On the basis of the preceding information, the following three hypotheses were proposed for arugula, cabbage, kale, and mustard seedling growth from seeding to cotyledon unfolding. Under LED lighting at a PPFD of 100 μmol·m⁻²·s⁻¹ with a photoperiod of 24 h or 16 h, 1) shortened photoperiod (16 h) cannot eliminate the blue light promotion effect on plant elongation relative to red light, 2) the elongation promoted by blue light is greater under 24-h than 16-h lighting at least for some species, and 3) species differ in elongation response to blue light and the interspecies difference is unaffected by photoperiod. The objective of this study was to explore the mode of blue light action on plant elongation in four microgreen species by testing the foregoing hypotheses.

Materials and Methods

Plant materials and growing conditions. The experiment was conducted on four microgreen species with three replicates over time (Table 1) at the University of Guelph, Guelph, ON, Canada during the summer of 2018. Seeds were sown (one seed per cell) in 128-cell (8 × 16 cell) trays containing Sunshine Mix #5 substrate (Sun Gro Horticulture, MA). Each treatment had three trays in total. Each tray contained four species, and each species occupied two rows of cells in one tray. The sowed trays were placed inside a walk-in growth chamber to start the light treatment. The irrigation strategy and nutrient solution used for these plants were the same as those in the literature (Kong et al., 2019). The temperature and relative humidity were set at ±22 °C and 70%, respectively.

Experimental design and treatments. The experiment was conducted as a 2 × 2 factorial (photoperiod × light quality × plant species) in a split-split-plot design with three replicates over time (Table 1). The photoperiod, light quality, and plant species treatments were allocated to the main plots, subplots, and sub-subplots, respectively. For photoperiod treatments, continuous (24-h light/0-h dark) or periodic (16-h light/8-h dark) lighting were used, and for 16-h photoperiod, the lights were turned on between 9:00 a.m and 1:00 p.m. Light quality treatments included 1) R, monochromatic red light with a peak wavelength at 665 nm; and 2) B, monochromatic blue light with a peak wavelength at 440 nm. LED lighting system (Pro Series 325) was provided by LumiGrow, Inc. (Emeryville, CA).

For each replicate, two photoperiod treatments were randomly allocated to the two ends in the growth chamber, which were far away from each other to avoid neighboring effects. Also, within each photoperiod treatment plot, the two light quality treatments were randomly allocated to the two compartments. The locations of the four treatments of photoperiod × light quality were switched by changing the timer setting and light spectral output of the LEDs in each compartment for each of the three replicates (Table 1). For each replicate, a PPFD of around 100 μmol·m⁻²·s⁻¹ at the plant canopy level were achieved in each compartment by adjusting the light intensity output of the LEDs. Compartments were separated by curtains to prevent neighboring effects. Light spectra and intensities were set up and verified using a USB2000 + ultraviolet/VIS spectrometer (Ocean Optics, Inc., Dunedin, FL). The light levels and other environmental data of different light treatments are presented in Table 2.

Growth and morphology measurements. When more than 50% of seeds germinated under all the treatments, the germination percentages were investigated. At the beginning of cotyledon unfolding, for each species, 16 plants in the middle cells were sampled from each treatment for each replicate to measure initial main stem (MS) length. At the end of the light treatments, the final MS length was measured on the same plants as those for initial measurement. On the basis of the MS measurements, stem extension rate (SER; cm·d⁻¹) was calculated using Eq. [1],

\[
SER = \frac{L_{Sf} - L_{Si}}{3}
\]

where \(L_{Sf}\) and \(L_{Si}\) are the final and initial MS lengths, respectively. The denominator, 3, represents the number of days between the initial and final measurements for each plant species.

At 7 d (mustard and kale) and 8 d (arugula and cabbage) after the start of light treatments in each of the three replicates of the experiment, six plants from each species were randomly selected for observation of stem and cotyledon morphology under each light treatment combination (two light quality treatments × two photoperiod treatments). For the morphology observations, the sampled plants were cut at the root-shoot junction, and the cotyledons, with petals, were cut from the stems. Then the stems were straightened, and the cotyledons were fully unfolded using sticky, clear, plastic tape and laid on an white sheet of A4 paper with the upper cotyledon surface outward. The papers with stems and cotyledons and the standard scale were then scanned using a CanonScan LiDE 25 scanner (Canon Canada Inc., Brampton, ON, Canada). After scanning, another 10 plants from each species for each replicate were randomly selected for the measurements of biomass accumulation and partitioning. The sampled plants were cut at the root-shoot junction. After weighing the total aerial fresh weight (FW), the aerial parts were separated into cotyledons with petals, and stems. The separated aerial plant parts were put into two separate paper bags and dried in an oven at 65 °C to determine dry weight (DW) of each component. The biomass partitioning trait, stem/aerial DW, was then calculated using Eq. [2].

\[
\text{Stem/aerial DW (\%)} = \left(\frac{\text{DW of stem (g)}}{\text{DW of aerial part (g)}}\right) \times 100
\]

After harvesting was completed for all the seedlings, the scanned images were processed.
using ImageJ 1.42 software (National Institute of Health). Hypocotyl length, diameter, and color; cotyledon maximum blade length, maximum blade width, area, and color; and petiole length were determined from the scanned images. For hypocotyl and cotyledon color measurements, the detailed process could be found in the literature (Kong et al., 2019). Plant size and color measurements were calibrated by scanning a ruler and Munsell color chips (GretagMacbeth LLC, New Windsor, NY), respectively.

Statistical analysis. Data were subjected to analysis of variance using a Data Processing System Software (DPS, version 7.05; Refine Information Tech. Co., Hangzhou, China) and were presented as means ± SE (standard error). Separation of means was performed using Duncan’s new multiple range test at the \( P \leq 0.05 \) level. The CV under light quality treatments for each affected growth and morphological trait was calculated to compare variation magnitude of response to B vs. R light between the two photoperiod treatments and among the four plant species.

Results

There was no difference in germination under B and R light (data not shown). During the experimental period, plants under B vs. R light showed a greater SER regardless of photoperiod for cabbage, kale, and arugula (Fig. 1A). At harvest, both FW and DW of aerial parts were similar for the plants from B and R light (data not shown). However, regardless of photoperiod, B vs. R light increased the ratio of stem/aerial DW for cabbage, kale, and arugula (Fig. 1B). The plants harvested from B vs. R appeared to be taller regardless of photoperiod for cabbage, kale, and arugula (Fig. 1C–F). There was an interaction effect between photoperiod and light quality on SER. Also, species interacted with light quality, affecting all the plant size traits. However, there were no interaction effects on the plant size traits for photoperiod, species, and light quality.

| Photoperiod | Light quality | PPFD \(^{a}\) (\(\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}\)) | DLI (mol\cdot m\(^{-2}\)\cdot d\(^{-1}\)) | PPS | Air temp. (\(^{\circ}\text{C}\)) | RH (%) |
|-------------|---------------|-----------------|-----------------|-----|-----------------|-------|
| 24 h        | Red light     | 98.4 ± 0.7\(^{b}\) | 8.5 ± 0.1       | 0.89 ± 0.00 | 21.9 ± 0.1 | 68.7 ± 0.4 |
|             | Blue light    | 102.6 ± 0.4     | 8.9 ± 0.0       | 0.46 ± 0.00 | 21.8 ± 0.1 | 68.4 ± 0.5 |
| 16 h        | Red light     | 97.1 ± 1.0      | 5.6 ± 0.1       | 0.89 ± 0.00 | 21.8 ± 0.1 | 68.6 ± 0.3 |
|             | Blue light    | 102.6 ± 1.6     | 5.9 ± 0.1       | 0.47 ± 0.01 | 21.8 ± 0.1 | 68.3 ± 0.4 |

\(^{a}\)PPFD = photosynthetic photon flux density. The wavelength range 400 to 700 nm was used for calculating the PPFD. DLI = daily light integral; PPS = phytochrome photostationary state, which is the estimated phytochrome photoequilibrium according to the method by Sager et al. (1988). RH = relative humidity. 

Data are means ± se (n = 3).
Generally there was no difference in cotyledon color between plants under B and R light (data not shown). For stem color, B vs. R did not change hue angle which was around 77° (yellow-green), 354° (purple), 21° (red), and 86° (yellow-green) for arugula, cabbage, kale, and mustard, respectively (data not shown) but reduced saturation and increased brightness in stem color for arugula and kale (Fig. 3). This suggested that the stems under B vs. R showed less greenness and redness for arugula and kale, respectively. For two traits, there were no interaction effects for photoperiod and light quality or for photoperiod, light quality, and species, but species showed interaction with light quality.

**Discussion**

Blue vs. red light can also promote plant elongation under noncontinuous lighting. In a previous study on microgreens, under 24-h lighting at a PPFD of 50 or 100 μmol·m⁻²·s⁻¹, B vs. R light promoted elongation growth for cabbage, kale, and arugula (Kong et al., 2019). In the present study, although the LED light, environmental conditions, and growth period were different from those in the previous study, B vs. R light under either 24- or 16-h lighting at a PPFD of 100 μmol·m⁻²·s⁻¹ promoted elongation growth by increasing stem extension rate, hypocotyl length, or petiole length for all the tested microgreen species except mustard. It confirmed the first hypothesis that shortened photoperiod (16 h) cannot eliminate the blue light promotion effect on plant elongation relative to red light. In the present study, under 16-h or 24-h lighting, blue-light-promoted microgreen elongation met the demand of machine-harvesting (i.e., around 5 cm hypocotyl length). Also, under 16-h lighting, B vs. R light promoted elongation without compromising yield compared with 24-h lighting. Crop yield is important for microgreen growers because it directly affects economic return (Jones-Baumgardt et al., 2019). It implies that if blue light is used for microgreen production, shortening photoperiod from 24 h to 16 h may potentially reduce lighting cost and increase production efficiency.

The promoted elongation by B vs. R light was considered as one of the shade-avoidance responses mediated by blue light associated with weak phytochrome activity under certain light levels in previous studies (Kong et al., 2018, 2019). This conclusion was also supported by the results in the present study. B vs. R light has a much lower phytochrome photo-stationary state (PPS) value, 0.46 vs. 0.89 (Table 2), and there is a general consensus that a PPS >0.6 can induce active-phytochrome response (Stutte, 2009). Monochromatic blue light seems to act like far-red light with regard to phytochrome. In addition to promoting elongation growth, B vs. R light also increased biomass partition to stem, reduced cotyledon size, or hypocotyl coloring showing typical shade-avoidance responses, which varied with different species. The reduced cotyledon size, or hypocotyl coloring under B vs. R light may negatively affect the appearance quality of microgreens to some degree because microgreens with large leaf size and deep stem color are normally more attractive to most consumers. In fact, the reduction of stem color in the present study was difficult to perceive by human vision possibly due to similar hue angle. Nevertheless, to reach a balance on the consideration of hypocotyl length and appearance quality (e.g., leaf size and stem Fig. 2. Plant size traits under different light treatments in four microgreen species. Data are means ± se (n = 3). B = monochromatic blue light; R = monochromatic red light. At the bottom of each panel, 24 h and 16 h indicate that the photoperiod is 24 and 16 h, respectively. For a given plant trait, symbols for light quality (Q), photoperiod (P), plant species (S), or the interaction of light quality and photoperiod (Q·P), light quality and plant species (Q·S), photoperiod and plant species (P·S), or light quality, photoperiod, and plant species (Q·P·S) are located closely above the frames followed by ns, *, **, or *** denote that treatment effects are not significant or significant at P ≤ 0.05, 0.01, or 0.001, respectively. Within the same species, data bearing the same letter are not significantly different at P ≤ 0.05, according to Duncan’s new multiple range test.
color) of microgreens, after ≈7-d blue light treatment, exposing these plants to red light for some days may be an option because these microgreen species can be harvested without or with unfolded first true leaves (i.e., 7–8 d or 11–14 d after seeding) (Jones-Baumgardt et al., 2019). This needs further study to confirm the feasibility.

It is worthwhile to note that in the previous study on the same microgreen species, B vs. R light changed cotyledon color (Kong et al., 2019). However, this change did not occur in the present study. The difference might be due to earlier plant harvest (7–8 d vs. 11–14 d after seeding) in the present vs. previous study. The lighting treatment period for the present study was from seeding to cotyledon unfolding, so the unfolded cotyledons during de-etiolation experienced a very short period of lighting to elicit color changes. During the de-etiolation stage, cotyledon color changing from yellow to a genotype-inherent color such as green or red involves initiated chloroplast development or increased anthocyanin biosynthesis (Kong and Zheng, 2019). The difference in pigment formation under B vs. R light might not be obvious until the lighting treatments reached a certain time length. For example, seedlings of nonheading Chinese cabbage increased chlorophyll content after 28 to 30 d of B vs. R LED light treatment (Fan et al., 2013; Li et al., 2012).

**Blue light promotion effects on elongation are more obvious under 24-h than non-24-h lighting.** There were interaction effects between photoperiod and light quality on the elongation growth traits in the present study. For the tested species, B vs. R light effects on elongation were greater under continuous than noncontinuous lighting in many cases (Fig. 3). For example, B vs. R light increased stem extension rate and hypocotyl length for mustard, and petiole length for mustard and kale under 24-h lighting rather than 16-h lighting. Under 24 vs. 16-h lighting, the B light promotion effects were greater for stem extension rate and hypocotyl length in kale and for petiole length in arugula. This confirmed our second hypothesis that the elongation promoted by blue light is more obvious under 24- than 16-h lighting at least for some species. It appears that continuous lighting might provide a better platform to present and even amplify B vs. R light effects on plant elongation at least for some species. When considering all the light-quality-affected plant traits together, in most cases, B vs. R light effects were greater under 24-h than 16-h lighting for the tested species except cabbage (Fig. 4). For cabbage, 24-h lighting showed the smallest difference from 16-h lighting in terms of B vs. R light promoted elongation growth, and this species even showed contrasting results from other species on cotyledon size (e.g., maximum blade length) (Fig. 2). Considering this point, when blue LED light is used for increasing plant elongation of cabbage microgreens, 16-h rather than 24-h lighting may be adopted in production. Although the involved mechanism for the species difference is not clear and needs further study, it suggests that species differences in blue light effects cannot be ruled out.

**Species differences in blue light effects on elongation are independent of photoperiod.** In the previous study on microgreens with unfolded true leaves, B vs. R light effects on the plant growth and morphology varied with species (Kong et al., 2019). Similar results were found in the present study despite shorter growth period (from seeding to cotyledon unfolding). This was supported by interaction
effects between plant species and light quality on all the plant traits affected by light quality. Generally, mustard showed the lowest sensitivity in elongation response to B vs. R light under either 24-h or 16-h photoperiod among all the tested species (Fig. 4). It appeared that for red-leaved mustard, elongation promoted by blue light was less obvious, compared with other green-leaved species. A similar different elongation response to B vs. R light was achieved in the seedlings of different cabbage cultivars with different leaf color: ‘Kinsushin’ (green leaf), and ‘Red Rookie’ (red leaf) (Mizuno et al., 2011). Possibly, the different leaf color can partly help explain the species’ difference in elongation response. For red-leaved mustard rather than other green-leaved species, the increased red pigments in the cotyledons might increase the reflection and then reduce the transmission of red light to its main photoreceptors, phytochromes. Previous studies have indicated that the blue light promoted elongation is related to weak phytochrome activity compared with red light (Kong et al., 2018) and stem elongation signal comes from phytochromes in cotyledons (Black and Shuttleworth, 1974; Kim et al., 2016). Under lower light levels (i.e., 100 μmol·m⁻²·s⁻¹), the reduced red light signaling in red-leaved mustard rather than other green-leaved species might trigger similarly weak phytochrome activity as blue light.

In the present study, although there were species variations in responses to B vs. R light, the interaction between plant species and light quality appeared to be independent of photoperiod, which confirmed our third hypothesis that species differ in elongation response to blue light, and the difference is unaffected by photoperiod. For example, under either 24-h or 16-h lighting, arugula and mustard showed the highest and lowest sensitivity, respectively, in response to B vs. R light among the four species (Fig. 4). This was also supported by the result that there were no interaction effects of species × photoperiod × light quality, despite significant interaction effects of species × light quality on all the plant traits affected by B vs. R light. Previous studies indicate that blue-light-mediated elongation response is a shade-avoidance response with varying sensitivity among species (Kong et al., 2018, 2019). Possibly, the species difference in elongation promoted by B vs. R light is due to inherent genetic differences in response to shade (Gommers et al., 2013), which may not be removed by change in some environmental factors, such as photoperiod.

In summary, at a PPFD of ≈100 μmol·m⁻²·s⁻¹, monochromatic blue vs. red LED light can also promote elongation growth under 16-h lighting for the tested microgreen species except mustard. The promotion effects by blue light are more obvious under 24-h than 16-h lighting in most cases. However, species vary in elongation response to blue light regardless of photoperiod. It suggests that blue-light-promoted elongation growth is not an artifact specifically from 24-h lighting, implying a potential use of monochromatic blue LED light to promote hypocotyl elongation of some microgreen species.

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