Differing Behavioural Responses of *Bemisia tabaci* MEAM1 and MED to Cabbage Damaged by Conspecifics and Heterospecifics

Hailong Kong¹,², Yang Zeng¹, Wen Xie¹, Shaoli Wang¹, Qingjun Wu¹, Xiaoguo Jiao³, Baoyun Xu¹ & Youjun Zhang¹

The whitefly *Bemisia tabaci* is a serious pest with an extensive host range. Previous research has shown that *B. tabaci* is a species complex with many cryptic species or biotypes and that the two most important species are MEAM1 (Middle East-Minor Asia 1) and MED (Mediterranean genetic group). MEAM1 and MED are known to differ in their preference for cabbage, *Brassica oleracea*, as a host plant, however, the mechanism underlying this preference is unknown. In the current study, a host choice experiment showed that MED prefers to settle and oviposit on undamaged cabbage plants rather than MED-damaged cabbage plants. However, MEAM1 prefers MED-damaged cabbage plants to undamaged plants and does not exhibit a significant preference for undamaged or MEAM1-damaged cabbage plants. On the basis of gas chromatography-mass spectrometry (GC-MS) analysis, the following volatiles were released in larger quantities from Q-damaged cabbage plants than from undamaged plants: 2-ethyl-1-hexanol, benzene methanol, (E)-2-decenol, benzaldehyde, nonanal, acetic acid geraniol ester, 4-hydroxy-4-methyl-2-pentanone, decane, and α-longipinene. Only one volatile, 4-hydroxy-4-methyl-2-pentanone, was released in greater quantities from MEAM1-damaged cabbage plants than from undamaged plants. Our results suggest that differences in herbivore-induced host volatile release may help explain the differences between the preference of *B. tabaci* MEAM1 and MED for cabbage as a host.

Herbivore-induced plant volatiles (HIPVs) are volatile semiochemicals that plants emit either at the site of herbivore damage or at a distance from this site¹. Plants often increase their synthesis and emission of HIPVs when damaged by herbivores². Although HIPVs can benefit the plant by attracting natural enemies of the herbivores and by repelling other herbivores, HIPVs can also harm the plant by attracting conspecific or heterospecific herbivores³. HIPVs are an important aspect of insect-plant interactions, and understanding their effects is important not only from an ecological and evolutionary perspective but also for the development of novel crop-protection strategies. Some volatiles, for example, have been used to mass trap pests or to repel pests from food and oviposition sites⁴,⁵. The use of HIPVs for integrated pest control requires an understanding of their effects on both natural enemies and on conspecific and heterospecific herbivores.

The whitefly *Bemisia tabaci* (Hemiptera: Aleyrodidae), a highly destructive agricultural pest, is a species complex composed of many morphologically indistinguishable biotypes. The two most invasive and destructive species are MEAM1 (Middle East-Minor Asia 1) and MED (Mediterranean genetic group)⁶–⁸. *B. tabaci* is widely distributed in the tropics and subtropics and has become a worldwide pest⁹. In China, *B. tabaci* was first recorded in the late 1940s⁴⁰, but the damage it inflicted on crops was not considered serious until the introduction of MEAM1 in the 1990s⁴¹. In recent years, MED has invaded China and has now displaced MEAM1 in many areas¹²,¹³. Because *B. tabaci* MEAM1 and MED are resistant to insecticides and are not effectively controlled by other traditional methods, alternative methods of control are being investigated⁴¹.
B. tabaci can infest more than 600 species of plants, including cabbage, tomato, cotton, cucumber, sweet pepper, and other economically important species. B. tabaci larvae have very limited mobility, and they therefore rely on maternal choice to determine their host plant. It follows that modification of host plant choice, i.e., of maternal behaviour, could be useful for controlling this pest. Jiao et al. found that the proportion of whiteflies that settled and oviposited on cabbage was significantly higher for MEAM1 than for MED. Thus, the attraction to cabbage was greater for MEAM1 than MED. However, little is known about the mechanism underlying the differences in the selection of cabbage by MEAM1 and MED. We hypothesise that HIPVs may help explain why MEAM1 and MED differ in this preference. To test this hypothesis, we compared the preference of MEAM1 and MED whiteflies for undamaged and damaged cabbage plants and the volatile compounds released by these plants. In this study, the term “damaged plants” refers to plants damaged by conspecifics (or heterospecifics), i.e., by the same (or another) species that was tested for preference.

Results

The preferences of B. tabaci B and Q adults for settling and ovipositing on undamaged cabbage plants vs. cabbage plants damaged by conspecifics and heterospecifics. The proportion of B whiteflies that settled on cabbage plants did not significantly differ between MEAM1-damaged and undamaged plants (Fig. 1A) (t = 1.51, df = 16, P > 0.05); there was a tendency, however, for more MEAM1 whiteflies to settle on MEAM1-damaged plants than on undamaged plants. In contrast, a significantly smaller proportion of MED whiteflies settled on MED-damaged than on undamaged cabbage plants (Fig. 1B) (t = -15.77, df = 16, P < 0.001). For MEAM1 whiteflies, the proportion of eggs deposited did not significantly differ between MEAM1-damaged and undamaged cabbage plants (Fig. 1C) (t = 1.27, df = 16, P > 0.05), but there was a tendency for larger numbers on the damaged plants. MED whiteflies, in contrast, deposited significantly fewer eggs on MED-damaged than on undamaged cabbage plants (Fig. 1D) (t = -13.18, df = 16, P < 0.001).

In addition, a significantly higher proportion of MEAM1 whiteflies settled on MED-damaged than on undamaged cabbage plants (Fig. 1E) (t = 8.57, df = 16, P < 0.001), and MEAM1 whiteflies deposited significantly more eggs on MED-damaged than on undamaged cabbage plants (Fig. 1F) (t = -15.64, df = 16, P < 0.001).

Volatile released by undamaged, MEAM1-damaged, and MED-damaged cabbage plants. According to GC-MS chromatograms, the volatiles released by MED-damaged and undamaged cabbage plants differed quantitatively (Fig. 2; Table 1). MED-damaged cabbage plants emitted significantly more of the following volatiles than undamaged plants: 2-ethyl-1-hexanol, benzenemethanol, (E)-2-decenol, benzaldehyde, nonanal, acetic acid geraniol ester, 4-hydroxy-4-methyl-2-pentanone, decane, and α-longipinene. However, one volatile, 4-hydroxy-4-methyl-2-pentanone, was released in greater quantities from MEAM1-damaged cabbage plants than from undamaged plants.

Discussion

We have demonstrated that B. tabaci MED prefers undamaged cabbage plants to those damaged by conspecifics. Although the settling and oviposition behaviour of B. tabaci MEAM1 did not significantly differ between MEAM1-damaged and undamaged cabbage plants, MEAM1 whiteflies tended to prefer MEAM1-damaged cabbage plants to undamaged cabbage plants. Furthermore, MEAM1 whiteflies significantly preferred MED-damaged cabbage plants to undamaged cabbage plants. These results may help explain the previous finding that a cabbage is a preferred host for MEAM1 but not for MED; once infested by MED but not MEAM1 whiteflies, a cabbage plant will repel many conspecifics.

According to optimal oviposition theory, herbivores preferentially oviposit on plants that provide optimal conditions for offspring development. Jiao et al. found that female fecundity, female longevity, and nymph survival were greater on cabbage for MEAM1 than for MED whiteflies. The present results are therefore consistent with optimal oviposition theory. This theory is also supported by previous results on the preference of B. tabaci and its fitness on the pepper. Jiao et al. found that the effects of the pepper on MEAM1 and MED fitness (adult longevity, fecundity, and nymph survivorship) matched those on settling and oviposition.

The behavioural response of herbivores to host plants damaged by conspecifics varies among herbivores and plants. De Moraes et al., for example, found that females of the moth Petasites paradoxus and Adenostyles alliariae when they were damaged by conspecifics than when they were undamaged. B. tabaci MED’s preference for undamaged cabbage plants over MED-damaged cabbage plants in the current study may be explained by significant differences in the volatile compounds released by the two types of cabbage plants, i.e., perhaps MED is attracted to volatiles from undamaged cabbage plants but repelled by volatiles from MED-damaged plants, because natural enemies may be attracted to the volatiles released from herbivore-damaged plants. Li et al. found that root JA and shoot JA-induced Chinese broccoli volatiles have high efficiencies in attracting the parasitoid Encarsia formosa Gahan and the predator Serangium japonicum, natural enemies of B. tabaci. Therefore, MED may have evaded to avoid such damaged plants to reduce its risk of attack from natural enemies. This is evidently not the case, however, for MEAM1. Although there are no significant differences in the behaviour of MEAM1 on damaged and undamaged plants, MEAM1 whiteflies tend to prefer MEAM1-damaged cabbage plants to undamaged cabbage plants. We also found that MEAM1 whiteflies significantly prefer MED-damaged cabbage plants to undamaged cabbage plants. This means that MEAM1 adults may be using the volatiles emitted by the damaged plants as cue for finding new sources of food, but, in doing so, their risk of exposure to natural enemies increases. This preference is likely to represent a trade-off between the detection of a more apparent host and suboptimal nutritional food quality and the risk of predation or parasitisation. Paradoxically, a preference for odours from a suboptimal host, risk of predation or parasitisation but detectable and damaged hosts might be a behavioural strategy to cope...
with uncertainties of host-plant location. Carroll et al.\textsuperscript{22} found that fall armyworm (\textit{Spodoptera frugiperda}) larvae preferred odours from damaged over undamaged maize seedlings. As noted earlier, MEAM1 and MED are morphologically identical and are substantially similar genetically, but are reproductively isolated\textsuperscript{7,23}. Given this reproductive isolation, it seems possible that the effects of MEAM1 and MED damage on volatiles emitted by a host plant and their behavioural responses to the same host plant might be different.

Volatiles often play a key role in the location of hosts by herbivores\textsuperscript{19}. It follows that the effect of herbivore injury on the behavior of later-arriving conspecifics may result from herbivore-induced changes in the volatiles released by a plant. In the current study, we found that the quantities of nine volatiles released by MED-infested cabbage plants increased significantly. Previous studies have documented that some plant volatiles repel or attract whiteflies. Wang et al.\textsuperscript{24} found that \textit{Chaitophorus salmigra} exhibited taxis in response to benzaldehyde. Cao et al.\textsuperscript{25} showed that myrcene attracted \textit{B. tabaci} MEAM1. High levels of p-cymene and curcumene were reported to be strongly repellent.

**Figure 1.** The preferences of \textit{B. tabaci} MEAM1 and MED adults for undamaged cabbage plants vs. cabbage plants damaged by conspecifics in two-choice bioassays. (A) The proportions of MEAM1 whiteflies that settled on MEAM1-damaged vs. undamaged cabbage plants. (B) The proportions of MED whiteflies that settled on MED-damaged vs. undamaged cabbage plants. (C) The proportions of eggs deposited by MEAM1 whiteflies on MEAM1-damaged vs. undamaged cabbage plants. (D) The proportions of eggs deposited by MED whiteflies on MED-damaged vs. undamaged cabbage plants. Values are means ± SE. (E) The proportions of MEAM1 whiteflies that settled on MED-damaged vs. undamaged cabbage plants. (F) The proportions of eggs deposited by MEAM1 whiteflies on MED-damaged vs. undamaged cabbage plants. Values are means ± SE.
Figure 2. Total ion chromatograms of volatile compounds released by undamaged cabbage plants and cabbage plants damaged by the MEAM1 or MED species of *B. tabaci*. Those compounds that differed significantly in quantity among the three types of cabbage plants were: 1 = (E)-2-decenol; 2 = 4-hydroxy-4-methyl-2-pentanone; 3 = benzaldehyde; 4 = decane; 5 = acetic acid geraniol ester; 6 = 2-ethyl-1-hexanol; 7 = α-longipinene; 8 = benzenemethanol; and 9 = nonanal.

| Compounds                  | Retention time | Undamaged plants | MEAM1-damaged plants | MED-damaged plants |
|----------------------------|----------------|------------------|----------------------|-------------------|
| **alcohols**               |                |                  |                      |                   |
| 2-ethyl-1-hexanol          | 12.29          | 41.70 ± 11.28a   | 49.46 ± 8.47a        | 181.04 ± 31.56b   |
| benzenemethanol            | 13.79          | 10.36 ± 1.52a    | 8.52 ± 1.75a         | 41.11 ± 9.44b     |
| **aldehydes**              |                |                  |                      |                   |
| (E)-2-decenol              | 6.54           | 6.38 ± 1.80a     | 11.27 ± 3.22a        | 44.69 ± 12.67b    |
| benzaldehyde               | 9.08           | 58.03 ± 13.94a   | 111.31 ± 25.49a      | 193.67 ± 32.43b   |
| nonanal                    | 13.94          | 38.44 ± 9.94a    | 30.71 ± 7.65a        | 152.27 ± 37.87b   |
| **esters**                 |                |                  |                      |                   |
| acetic acid geraniol ester | 11.37          | 94.69 ± 18.00a   | 80.73 ± 16.66a       | 448.25 ± 131.32b  |
| **ketones**                |                |                  |                      |                   |
| 4-hydroxy-4-methyl-2-pentanone | 8.87    | 50.02 ± 13.56a   | 96.60 ± 13.84b       | 165.87 ± 24.79b   |
| **terpenoids**             |                |                  |                      |                   |
| α-longipinene              | 13.59          | 556.96 ± 131.49a | 1008.27 ± 143.93a    | 2360.18 ± 402.57b |
| **others**                 |                |                  |                      |                   |
| decane                     | 10.93          | 17.48 ± 3.93a    | 17.23 ± 4.21a        | 71.69 ± 11.76b    |

Table 1. Peak areas of volatiles released from *B. tabaci* MEAM1-damaged, *B. tabaci* MED-damaged, and undamaged cabbage plants. Values are means ± SE. Means within a row followed by different letters are significantly different at *P* < 0.05.
to *B. tabaci* MED. In the present study, more benzaldehyde and α-longipinene were released from MED-damaged than undamaged cabbage plants. The different behavioural responses of MEAM1 and MED to undamaged and damaged cabbage plants may be attributed to qualitative differences in the volatile compounds emitted by plants. However, the effects on *B. tabaci* behaviour of the nine volatiles detected in this study requires further investigation.

Nonetheless, the levels of all volatiles detected (except for 4-hydroxy-4-methyl-2-pentanone) did not differ significantly between MEAM1-damaged and undamaged cabbage plants. The lack of a significant change in volatiles has also been reported in other studies involving phloem-feeding insects and host plants. For example, Schwartzberg *et al.* showed that the pea aphid, *Acyrthosiphon pisum*, did not induce a detectable change in plant volatiles released from broad bean (*Vicia faba*). Turlings *et al.* documented that even a heavy infestation of the aphid *Rhopalosiphum maidis* failed to induce measurable emissions of volatiles in maize. The effect of 4-hydroxy-4-methyl-2-pentanone on *B. tabaci* behaviour requires further investigation.

Our results show that *B. tabaci* MEAM1 and MED differ in their responses to undamaged cabbage plants and cabbage plants damaged by conspecifics or heterospecifics. The results also show that these differences correspond to differences in the quantity of volatiles released by the damaged plants. To our knowledge, these results provide the first evidence of a possible direct effect of herbivore-induced plant volatiles (HIPVs) on the differing preference of *B. tabaci* MEAM1 and MED for cabbage. Further experiments are needed to determine which volatiles have the most adverse (or beneficial) effect on host-plant selection by whiteflies. The volatiles from such studies may be able to be used as repellents or attractant traps for whiteflies in the field.

**Methods**

**Insects and plants.** The MEAM1 population of *B. tabaci* used in this study was originally collected from cabbage (*Brassica oleracea* cv. Jingfeng1) in 2004 in Beijing, China. The MED population of *B. tabaci* used in this study was originally collected from poinsettia (*Euphorbia pulcherrima* Wild. ex Klotz.) in 2009 in Beijing. Subsequently, both populations have been maintained on tomato (*L. esculentum* var. Zhongza 9) in separate insect-proof screened cages in a greenhouse at 23 ± 2 °C and with natural lighting. The purity of each population was monitored every two generations by determining the DNA sequence of the haplotypes following amplification by mtCOI primers.

Cabbage (*B. oleracea* var. Jingfeng 1) was grown from seed in a greenhouse at 23 ± 2 °C and with natural lighting. Two weeks after seeds were sown, seedlings were transplanted to 5-cm-diameter plastic pots (one seedling per pot). When they had six main leaves, plants of similar size were selected for each experiment.

**MEAM1 and MED preferences for undamaged cabbage plants vs. cabbage plants damaged by conspecifics or heterospecifics.** Bioassays were used to assess the preferences of MEAM1 and MED whiteflies for damaged or undamaged cabbage plants. Damaged plants were obtained by attaching three clip-cages per plant with 100 MEAM1 or MED adults per clip-cage. Undamaged cabbage plants were treated in the same manner, but whiteflies were not added to the clip-cages. The bioassays were conducted as described by Omondi *et al.* and used cabbage plants of approximately the same size. In brief, two cabbage plants were placed in diagonal corners of a 60 × 40 × 80-cm screen cage (one plant per corner, two per cage); one plant was undamaged, and the other had been damaged by MEAM1, for assays with MEAM1, or by MED, for assays with either MED or MEAM1. The clip cages and whiteflies within them were not removed after the plants were placed in the cages. Approximately 300 assayed MEAM1 or MED adult whiteflies were collected between 7:00 and 8:00 hours and were released near the centre-bottom of the cage. This was accomplished by placing the aspirator sampling bottle containing whiteflies in a clear plastic tumbler in the centre of the cage. Whiteflies moved to and flew out of the open top of the sampling bottle. The number of whiteflies on each plant was determined after 48 h. All of the leaves from each plant were then removed, and the whitefly eggs on each leaf were counted with the aid of a stereomicroscope (Leica, M205C). There were nine replicates for each screen cage. The cages were arranged in blocks in a greenhouse with natural lighting and at ambient temperature (23 ± 2 °C).

**Plant volatile collection and analysis.** Cabbage plants were undamaged or were damaged by MEAM1 or MED as described in the previous section. For collection of volatiles emitted, the plants were kept in a greenhouse at 23 ± 2 °C and with 50–70% RH and natural lighting. The plants were transferred individually to large desiccators (40 cm i.d., 60 cm high); an inlet at the top of each desiccator was used to supply pressurised air (100 ml/min) that had been filtered by passage through a jar containing distilled water. Volatiles were collected for 8h (from 8:00 to 16:00) in a glass tube filled with 150 mg of Tenax resin that had been placed in the outlet of each desiccator. Three plants (one from each treatment) were sampled simultaneously, and each treatment was represented by five replicate plants. The sampling tubes were stored at −20 °C until analysis.

Volatiles were eluted off the Tenax with 2 ml of pentane: diethyl ether (4: 1) and were subjected to gas chromatography-mass spectrometry (GC-MS-2010plus) using a VF-5MS column (0.25 mm × 30 mm, J and W Scientific, Folsom, CA). The temperature programme was as follows: from 40°C to 95°C at 3°C/min, then to 165°C at 2°C/min, and finally to 250°C at 15°C/min. The volatiles were detected by the MS operated at 70 eV in EI mode. Mass spectra were acquired in full-scan mode (33–400 AMU, 0.4 scan/sec). Compounds were identified by their mass spectra using NIST 2005 (National Institute of Standards and Technology, USA, http://www.nist.gov) and Wiley 9th edition spectral libraries. Relative quantification was based on the peak area of each component of the volatiles.

**Data analysis.** The independent samples *t*-test was used to compare whitefly settling and oviposition preference and the relative abundance of individual volatile compounds emitted from undamaged and from MEAM1- or MED-damaged cabbage plants. Statistical analyses were performed with SPSS (version 13.0; SPSS Inc., Chicago, IL, USA).
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Author Contributions
H.L.K. and Y.J.Z. designed the experiment. H.L.K. and Y.Z. performed the experiment. W.X., S.L.W., Q.J.W., Y.J.Z. and B.Y.X. contributed reagents/materials. H.L.K. and Y.Z. performed the experiment. W.X., S.L.W., Q.J.W., Y.J.Z. and B.Y.X. contributed reagents/materials. H.L.K., X.G.J. and Y.J.Z. wrote the paper.

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Additional Information
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