Research Article

Upregulation of Serum miR-629 Predicts Poor Prognosis for Non-Small-Cell Lung Cancer

Fayong Liu, Tianshui Li, Ping Hu, and Li Dai

Department of Respiratory and Critical Care Medicine, Beijing Jishuitan Hospital, Beijing 100000, China

Correspondence should be addressed to Fayong Liu; fyliudoctor@163.com

Received 23 June 2020; Revised 19 January 2021; Accepted 5 February 2021; Published 3 March 2021

1. Introduction

Lung cancer is one the most common cancers and by far the leading cause of cancer-associated mortality in the world. Non-small-cell lung cancer (NSCLC) accounts for approximately 85% of lung cancer [1]. The incidence and mortality rates of NSCLC are still increasing annually in both developed and developing countries [2]. Genetic susceptibility, poor diet, occupational exposures, and cigarette smoking are the major risk factors for NSCLC. Currently, the therapeutic methodologies for NSCLC include surgery, immunotherapy, radiotherapy, and targeted therapy [3]. Although great progress has been made in improving the therapeutic strategies, the prognosis of NSCLC remains poor and the five-year overall survival rate is only 15% [4, 5]. Therefore, developing highly efficient biomarkers for prognosis prediction of NSCLC is critical for improving the clinical outcome of NSCLC.

MicroRNAs (miRNAs) are a class of highly conserved small noncoding RNAs that play a role in posttranscriptional regulation of gene expression [6, 7]. miRNAs have been identified to be involved in diverse biological processes such as cellular differentiation, proliferation, apoptosis, and development processes [8]. In addition, aberrant expression of miRNAs has been observed in different types of human cancer including NSCLC [9]. For instance, overexpression of miR-154 significantly repressed the migration and invasion abilities of NSCLC cells in vitro, suggesting that miR-154 may be a potential anticancer therapeutic target for NSCLC [10]. Similarly, miR-101 is reported to act as a tumor-suppressive miRNA in NSCLC [11].

Previous studies have demonstrated that miR-629 was highly expressed in various cancers, such as lung adenocarcinoma, gastric cancer, and pancreatic cancer [12–15]. However, the clinical significance of serum miR-629 in NSCLC is still unclear. Therefore, the aim of the present study was to determine the expression level of serum miR-629 in NSCLC and explore its potential prognostic value.

2. Materials and Methods

2.1. Subject Recruitment and Sample Collection. This study received ethical approval from the Ethics Committee of
Beijing Jishuitan Hospital, and the protocol was in accordance with the Declaration of Helsinki. Written informed consent was obtained from each participant. A total of 166 patients with NSCLC, 70 patients with nonmalignant lung diseases, and 100 healthy volunteers were enrolled in our study. All the NSCLC cases were pathologically confirmed. The detailed information of the participants is shown in Table 1. The patients with nonmalignant lung diseases and healthy controls were matched with NSCLC cases for age and gender. The serum was separated from the blood by centrifugation at 1200 g for 10 min at room temperature. All extracted serum samples were stored at −80 °C until further analysis.

2.2. RNA Extraction and qRT-PCR. An miRNeasy Serum-/Plasma kit (Qiagen, Hilden, Germany) was used to extract the total RNA from serum samples. The RNA quality and concentration were assessed using a NanoDrop™ 1000 Spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA). The OD260/OD280 ratios were 1.8-2.2, and the OD260/OD230 ratios were 2.0-2.2. Complementary DNA was synthesized by reverse transcription reaction using the TaqMan MicroRNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA, USA). Quantitative real-time PCR for the relative quantification of serum miR-629 was performed using a miScript SYBR Green PCR kit (Qiagen) on a 7500 Real-Time PCR System (Applied Biosystems). The PCR conditions were 95°C for 5 min, followed by 40 cycles at 94°C for 15 sec and 55°C for 30 sec. Spiked-in Cel-miR-39 was used as a normalizer for serum miR-629 quantification. The relative level of serum miR-629 was calculated through the 2−ΔΔCt method.

2.3. Enzyme-Linked Immunosorbent Assay. The levels of CYFRA21-1 and carcinoembryonic antigen (CEA) in NSCLC patients and healthy subjects were determined with the CYFRA21-1 ELISA Kit (MyBioSource, San Diego, CA, USA) and Human Carcinoembryonic Antigen ELISA Kit (MyBioSource), respectively.

2.4. Statistical Analysis. The differences in the serum miR-629 level between/among different groups were determined by the Mann-Whitney U-test or Kruskal-Wallis test. Receiver operating characteristic (ROC) curves and the area under the ROC curve (AUC) were generated to assess the diagnostic accuracy of serum miR-629, CYFRA 21-1, and CEA. The overall survival and disease-free survival were evaluated by Kaplan-Meier analysis and log-rank test. Statistical analysis was performed using GraphPad Prism 7.0 software (GraphPad Prism Software Inc., La Jolla, CA, USA). A P value less than 0.05 was considered to be statistically significant.

3. Results

3.1. Serum miR-629 Levels Were Significantly Upregulated in NSCLC. The expression level of miR-629 was examined in 166 NSCLC patients, 70 patients with nonmalignant lung diseases, and 100 healthy controls with quantitative RT-PCR. The intra-assay and interassay coefficients of variation (CV) for serum miR-629 were as follows: NSCLC group (intra-assay CV = 2.73%, interassay CV = 3.67%), nonmalignant lung disease group (intra-assay CV = 1.41%, interassay CV = 3.15%), and healthy control group (intra-assay CV = 1.56%, interassay CV = 4.22%). Our results showed that the expression

| Parameters         | NSCLC     | NMLDs     | Healthy controls |
|--------------------|-----------|-----------|------------------|
| Age                | 62.35 ± 8.52 | 63.17 ± 9.05 | 61.45 ± 7.56     |
| Gender             |           |           |                  |
| Male               | 119       | 48        | 64               |
| Female             | 47        | 22        | 36               |
| Smoker             |           |           |                  |
| Yes                | 93        | 38        | 56               |
| No                 | 73        | 32        | 44               |
| Primary location   |           |           |                  |
| Left lung          | 79        | 33        | —                |
| Right lung         | 87        | 37        | —                |
| Histological type  |           |           |                  |
| AC                 | 96        | —         | —                |
| SCC                | 70        | —         | —                |
| Differentiation    |           |           |                  |
| Well/moderate      | 92        | —         | —                |
| Poor               | 74        | —         | —                |
| Lymph node metastasis |       |           |                  |
| Negative           | 66        | —         | —                |
| Positive           | 100       | —         | —                |
| Stage              |           |           |                  |
| I-II               | 88        | —         | —                |
| III-IV             | 78        | —         | —                |

Table 1: The clinical information of the study cohort.
level of serum miR-629 was significantly higher in patients with NSCLC than in patients with nonmalignant lung diseases and healthy controls \((P < 0.001)\). No significant difference was found between patients with nonmalignant lung diseases and healthy controls \((P = 0.921)\) (Figure 1).

### 3.2. The Diagnostic Potential of Serum miR-629 in NSCLC

The diagnostic value of serum miR-629 for NSCLC was performed with ROC curve analysis. The results showed that the AUC value of serum miR-629 for discriminating NSCLC patients from healthy controls was 0.835 (Figure 2(a)), which was higher than that of CYFRA 21-1 \((AUC = 0.672)\) (Figure 2(b)) and CEA \((AUC = 0.766)\) (Figure 2(c)).

3.3. The Association between Serum miR-629 Expression and Clinicopathological Features of NSCLC

As shown in Figure 3, the NSCLC patients with poorly differentiated grade \((P < 0.001)\), positive lymph node metastasis \((P < 0.001)\), or advanced clinical stages \((P < 0.001)\) had significantly higher serum miR-629 levels than those with well/moderate differentiated grade, negative lymph node metastasis, and early clinical stages. In addition, no significant difference was found between serum miR-629 and other analyzed clinico-pathological parameters including age \((P = 0.242)\), gender \((P = 0.166)\), primary location \((P = 0.855)\), smoking history \((P = 0.397)\), and histological type \((P = 0.617)\).

### 3.4. The Association between Serum miR-629 Expression and Therapeutic Response as well as Survival

A total of 135 NSCLC patients in this cohort received surgical treatment. We then compared the level of serum miR-629 in pretreated samples and posttreated samples. Our results showed that the level of serum miR-629 was markedly decreased in NSCLC patients following surgical treatment \((P < 0.001)\) (Figure 4(a)). According to the median expression level of serum miR-629, the NSCLC patients were divided into a high-expression group \((n = 83)\) and a low-expression group \((n = 83)\). Kaplan-Meier analysis demonstrated that patients in the high serum miR-629 group had a significantly shorter overall survival than those in the low serum miR-629 group \((P = 0.002)\) (Figure 4(b)). In addition, patients with a higher level of serum miR-629 had worse DFS compared with those with a lower level of serum miR-629 \((P = 0.011)\) (Figure 4(c)).

### 4. Discussion

In this study, we have shown that serum miR-629 was significantly upregulated in patients with NSCLC. In addition, serum miR-629 well differentiated patients with NSCLC from healthy controls. Moreover, serum miR-629 expression was notably decreased after treatment. NSCLC patients with higher serum miR-629 expression had worse clinical characteristics and prognosis. These findings might suggest that
serum miR-629 might serve as a novel potential prognostic biomarker for NSCLC.

Consistent with our findings, a positive correlation was found between the level of miR-629 and tumor invasion in lung adenocarcinoma (LUAD) specimens. In addition, over-expression of miR-629 enhanced the invasive capacity of lung cancer cells, indicating that miR-629 plays a tumor-promoting role in LUAD [12]. Similarly, miR-629-3p was also found to be upregulated in LUAD tissues and cell lines. Enforced expression of miR-629-3p promoted cell proliferation via downregulating the expression of SFTPC [13].

Aberrant expression of miR-629 has also been reported in other types of cancer. For instance, Li et al. demonstrated that the expression of miR-629 was upregulated in gastric cancer. Ectopic expression of miR-629 suppressed gastric cancer cell proliferation and promoted apoptosis by targeting FOXO3 [14] Similarly, Yan et al. showed that miR-629 expression is markedly upregulated in pancreatic cancer tissues. miR-629 overexpression enhanced cell proliferation and metastasis of pancreatic cancer cells in vitro and in vivo [15]. Shi et al. reported that miR-629 was significantly increased in the tissues and serum samples from pancreatic cancer patients. In addition, serum miR-629 was demonstrated to be a promising biomarker for the diagnosis and prognosis of pancreatic cancer [16]. The expression of miR-629 was also strongly associated with tipifarnib resistance in breast cancer cell lines [17]. Tao et al. reported that miR-629 was significantly upregulated in hepatocellular carcinoma (HCC) tissues compared with adjacent normal tissues. In addition, an in vitro assay showed that overexpression of miR-629 promoted the proliferation, migration, and invasiveness of HCC cells, while knockdown of miR-629 led to the opposite effects [18]. In colorectal cancer, miR-629 was found to be significantly upregulated in colorectal cancer tissues and cell lines, and upregulation of miR-629 enhanced cell proliferation and migration as well as suppressed cell apoptosis by directly downregulating CXXC4 [19].

One of the potential limitations of our study was the relatively small sample size. Further studies with larger sample sizes are urgently needed to validate our findings. In addition, as we have discussed above, serum miR-629 might be also upregulated in other types of cancer or other human diseases. Therefore, combining the serum miR-629 and currently known biomarkers for NSCLC as well as clinical parameters might contribute to improving the diagnosis and prognosis prediction of NSCLC. Moreover, the molecular mechanisms accounting for the tumor-promoting role in NSCLC need further investigation.

In conclusion, to the best of our knowledge, this is the first study to report that serum miR-629 is upregulated in

---

**Figure 3:** The association between serum miR-629 level and clinicopathological parameters of NSCLC.
NSCLC. An increased level of serum miR-629 is associated with a poor clinical outcome of NSCLC, indicating that serum miR-629 might serve as a promising and novel prognosis biomarker for NSCLC.

Data Availability
The data that support the findings of this study are available from the corresponding author upon reasonable request.

Conflicts of Interest
The authors declare that they have no conflicts of interest.

Authors’ Contributions
Dr. Liu FY, Dr. Li TS, Dr. Hu P, and Dr. Dai L designed the study, conducted the experiments, analyzed the data, and wrote the manuscript. All authors have read and approved the final version of the manuscript.

References
[1] C. Aggarwal and H. Borghaei, “Treatment paradigms for advanced non-small cell lung cancer at academic medical centers: involvement in clinical trial endpoint design,” The Oncologist, vol. 22, no. 6, pp. 700–708, 2017.
[2] L. A. Torre, F. Bray, R. L. Siegel, J. Ferlay, J. Lortet-Tieulent, and A. Jemal, “Global cancer statistics, 2012,” CA: A Cancer Journal for Clinicians, vol. 65, no. 2, pp. 87–108, 2015.
[3] R. Garcia-Campelo, R. Bernabe, M. Cobo et al., “SEOM clinical guidelines for the treatment of non-small cell lung cancer (NSCLC) 2015,” Clinical & Translational Oncology, vol. 17, no. 12, pp. 1020–1029, 2015.
[4] M. Dietel, L. Bubendorf, A. M. Dingemans et al., “Diagnostic procedures for non-small-cell lung cancer (NSCLC): recommendations of the European Expert Group,” Thorax, vol. 71, no. 2, pp. 177–184, 2016.
[5] N. Ramnath, T. J. Dilling, L. J. Harris et al., “Treatment of stage III non-small cell lung cancer,” Chest, vol. 143, no. 5, pp. e314S–e340S, 2013.
[6] D. P. Bartel, “MicroRNAs: genomics, biogenesis, mechanism, and function,” Cell, vol. 116, no. 2, pp. 281–297, 2004.
[7] X. Zhao and L. Cui, “A robust six-miRNA prognostic signature for head and neck squamous cell carcinoma,” Journal of Cellular Physiology, vol. 235, no. 11, pp. 8799–8811, 2020.
[8] G. A. Calin and C. M. Croce, “MicroRNA signatures in human cancers,” Nature Reviews. Cancer, vol. 6, no. 11, pp. 857–866, 2006.
[9] V. Del Vescovo and M. A. Denti, “MicroRNA and lung cancer,” Advances in Experimental Medicine and Biology, vol. 889, pp. 153–177, 2015.
[10] X. Lin, Z. Yang, P. Zhang, Y. Liu, and G. Shao, “miR-154 inhibits migration and invasion of human non-small cell lung
cancer by targeting ZEB2,” *Oncology Letters*, vol. 12, no. 1, pp. 301–306, 2016.

[11] J. G. Zhang, J. F. Guo, D. L. Liu, Q. Liu, and J. J. Wang, “MicroRNA-101 exerts tumor-suppressive functions in non-small cell lung cancer through directly targeting enhancer of zeste homolog 2,” *Journal of Thoracic Oncology*, vol. 6, no. 4, pp. 671–678, 2011.

[12] Y. Li, H. Zhang, L. Fan et al., “miR-629-5p promotes the invasion of lung adenocarcinoma via increasing both tumor cell invasion and endothelial cell permeability,” *Oncogene*, vol. 39, no. 17, pp. 3473–3488, 2020.

[13] B. Li, Y. Q. Meng, Z. Li et al., “miR-629-3p-induced downregulation of SFTPC promotes cell proliferation and predicts poor survival in lung adenocarcinoma,” *Artif Cells Nanomed Biotechnol*, vol. 47, no. 1, pp. 3286–3296, 2019.

[14] M. Li, Y. Wang, X. Liu, Z. Zhang, L. Wang, and Y. Li, “miR-629 targets FOXO3 to promote cell apoptosis in gastric cancer,” *Experimental and Therapeutic Medicine*, vol. 19, no. 1, pp. 294–300, 2020.

[15] H. Yan, Q. Li, J. Wu et al., “miR-629 promotes human pancreatic cancer progression by targeting FOXO3,” *Cell Death & Disease*, vol. 8, no. 10, article e3154, 2017.

[16] W. Shi, Y. Lu, R. Gong, J. J. Sun, and G. Liu, “Serum miR-629 is a novel molecular marker for diagnosis and the prognosis of pancreatic cancer,” *European Review for Medical and Pharmacological Sciences*, vol. 22, no. 16, pp. 5187–5193, 2018.

[17] K. Uhr, W. J. Prager-van der Smissen, A. A. Heine et al., “MicroRNAs as possible indicators of drug sensitivity in breast cancer cell lines,” *PLoS One*, vol. 14, no. 5, article e0216400, 2019.

[18] X. Tao, X. Yang, K. Wu et al., “miR-629-5p promotes growth and metastasis of hepatocellular carcinoma by activating β-catenin,” *Experimental Cell Research*, vol. 380, no. 2, pp. 124–130, 2019.

[19] J. Lu, S. Lu, J. Li, Q. Yu, L. Liu, and Q. Li, “miR-629-5p promotes colorectal cancer progression through targeting CXXC finger protein 4,” *Bioscience Reports*, vol. 38, no. 4, article BSR20180613, 2018.