RESEARCH ARTICLE

Renal function is independently associated with circulating betatrophin

Lukas Maurer1,2,3, Franziska Schwarz1,3, Antje Fischer-Rosinsky1,2,3, Nina Schlüeter1,3,4, Sebastian Brachs1,3, Matthias Möhlig1, Andreas Pfeiffer1,4,5, Knut Mai1,2, Joachim Spranger1,2,3,* and Thomas Bobbert1

1 Dept. of Endocrinology, Diabetes and Nutrition, Charité-Universitätsmedizin Berlin, Berlin, Germany, 2 Experimental and Clinical Research Center, Charité-Universitätsmedizin Berlin and Max-Delbrück Zentrum, Berlin-Buch, Germany, 3 Charité Center for Cardiovascular Research (CCR), Charité-Universitätsmedizin Berlin, Berlin, Germany, 4 DZHK (German Centre for Cardiovascular Research), Partner Site Berlin, Berlin, Germany, 5 Dept. of Clinical Nutrition, German Institute of Human Nutrition, Nuthetal, Germany

* joachim.spranger@charite.de

Abstract

Objective

Betatrophin has been identified as a marker linking liver with beta cell function and lipid metabolism in murine models. Until now, the regulation of circulating betatrophin in humans is not entirely clear. We here analyzed the relation of betatrophin levels to phenotypes of the metabolic syndrome and speculated that renal function might influence circulating betatrophin levels and explain age-dependent changes of betatrophin.

Subjects

We analyzed blood samples from 535 individuals participating in the Metabolic Syndrome Berlin Potsdam study.

Results

In a crude analysis we found a positive correlation between betatrophin levels and HbA1c (r = 0.24; p < 0.001), fasting glucose (r = 0.20; p < 0.001) and triglycerides (r = 0.12; p = 0.007). Furthermore betatrophin was positively correlated with age (r = 0.47; p < 0.001), systolic blood pressure (r = 0.17; p < 0.001), intima media thickness (r = 0.26; p < 0.001) and negatively correlated with CKD-EPI eGFR (r = -0.33; p < 0.001) as an estimate of renal function. Notably, eGFR remained highly associated with betatrophin after adjustment for age, waist circumference, gender, HbA1c and lipid parameters in a multivariate linear regression model (β = -0.197, p < 0.001).

Conclusions

Our data suggest that circulating levels of betatrophin depend on age, gender, waist circumference, total/HDL cholesterol ratio and renal function. Especially the association to eGFR highlights the importance for future studies to address renal function as possible influence...
on betatrophin regulation and consider eGFR as potential confounder when analyzing the role of betatrophin in humans.

**Introduction**

A recent report about a newly identified predominantly liver derived circulating factor referred to as betatrophin attracted great attention because of its capability to promote beta cell proliferation and improving glucose tolerance in mice [1]. Betatrophin also called ANGPTL8, RIFL or Lipasin was initially identified in 2012. It is an atypical member of the angiopoietin-like protein family (ANGPTLs) functionally considered as a nutritionally-dependent blood lipid regulator [2–4] inhibiting the lipoprotein lipase and thereby regulating triglyceride (TG) levels [5]. Subsequent studies analyzing the phenotype of betatrophin knock-out animals challenged the perspective of betatrophin having a prominent role in glucose metabolism. While knock-out animals showed clear signs of an altered lipid metabolism with significantly lower TG levels in the fed state no changes in glucose homeostasis were observed [6]. Nevertheless the identification of this liver derived circulating factor potentially involved in the regulation of glucose and lipid metabolism triggered a great research interest in this hormone as potential therapeutic target or new biomarker for the metabolic syndrome, leading to several clinical studies examining the role of betatrophin in humans with multiple pathological conditions. Most studies so far compared betatrophin levels between diabetic and non-diabetic subjects (type 2 diabetes, type 1 diabetes and gestational diabetes), providing quite heterogeneous results. While the majority showed higher serum concentrations for diabetic patients [7–9] others failed to detect any significant differences [10] or even showed decreased betatrophin levels for diabetic compared to non-diabetic individuals [11]. Based on the pre-clinical data indicating that betatrophin is a nutrition dependent lipogenic factor, appearing as a potential mediator of post-prandial fatty acid trafficking and storage to adipose tissue [4], a number of clinical studies investigated the role of betatrophin in lipid metabolism showing positive correlations with triglycerides [9, 12, 13]. Beyond that, the results of studies looking for a putative link to obesity presented a similar heterogeneity as in diabetes, differing for betatrophin levels to be higher [14], lower [15] or equal [16] comparing patients with obesity to lean individuals. Taking into consideration the partial contradictory results on the influence of betatrophin on glucose and lipid metabolism as well as obesity, we analyzed the participants of the Metabolic-Syndrome Berlin-Potsdam follow-up cohort with respect to their betatrophin levels in relation to markers of the metabolic syndrome. We included intima media thickness as a marker for atherosclerosis and CKD-EPI estimated GFR as a marker for renal function in order to analyze two further age dependent phenotypes, given the situation that the majority of clinical studies has pointed towards a strong positive correlation of betatrophin with age [8, 13, 17–19].

**Material and methods**

For this study blood samples of 535 individuals of the Metabolic Syndrome Berlin Potsdam (MesyBePo) follow-up study were analyzed. Details of baseline phenotyping were described previously [20]. The study protocol was approved by the ethics committees of the Landesarztekammer Brandenburg and the Freie Universität Berlin. All study samples were taken at 8:00 a.m. after an overnight fast. An oral glucose tolerance test (OGTT) was performed in all individuals, and fasting blood samples were taken before OGTT. Patient classification with respect to
impaired glucose tolerance (IGT), impaired fasting glucose (IFG) and T2DM was made according to either prior diagnosis of T2DM with accompanying antidiabetic medication or based on the results of fasting glucose and 2h glucose levels after GTT according to the official WHO diagnostic criteria [21]. EDTA and citrate plasma as well as serum was taken from participants. Plasma was placed on ice and spun down within 30 min at 4°C. After centrifugation, the plasma was immediately transferred to a −20°C freezer. Within the next 24 h, all samples were transferred on dry ice to a −80°C freezer, where aliquots were stored until assay. Serum was left at room temperature for 30 min to allow clotting, thereafter, the sample was centrifuged. The serum was then transferred to a −20°C freezer and within 24 h to a −80°C freezer, comparable to all plasma samples. Betatrophin levels were measured by validated enzyme-linked immunosorbent assay (ELISA) kit provided by EIAAB (Catalogue number E1164H; Wuhan, China). The internal quality control revealed an intra-assay coefficient of variability of 5.2% and an inter-assay CoV of 13.6% for the measured full-length betatrophin levels in plasma samples. Carotid arterial IMT was measured at the posterior wall of the common carotid artery at three different positions using a high resolution ultrasound (Kretz AG, Germany). Mean and maximum values for IMT, which included possible plaques, were calculated. CKD-EPI estimated glomerular filtration rate was calculated according to the established mathematical formula from the Chronic Kidney Disease Epidemiology Collaboration [22]. The study was carried out in accordance with the recommendations of the Declaration of Helsinki, and the experimental protocol of the study was approved by the institutional ethical committee. All subjects gave written informed consent. Statistical calculations were performed using SPSS 22.0 software (IBM Statistics, Germany). All values are presented as mean and SEM, if not otherwise mentioned. Logarithmic transformation of betatrophin levels was performed to achieve normal distribution tested by visual inspection and Shapiro Wilk test. The statistical tests in general were performed with ln(betatrophin) values. To provide a better comparability and understanding non-transformed values were depicted when absolute plasma concentration were presented. One-way ANOVA was performed followed by post-hoc Bonferroni test. A value of p < 0.05 was considered as statistically significant. Pearson Correlation coefficients were analyzed with respect to log Betatrophin. Multivariate linear regression models were calculated to identify independent relations between Betatrophin and ageing, renal function and multiple markers of the metabolic syndrome. Three models were calculated: crude without adjustment, model 1 was adjusted for age and sex only and model 2 was additionally adjusted for waist circumference, systolic blood pressure, HbA1c, total cholesterol/HDL cholesterol ratio, triglycerides and smoking (fully adjusted model).

Results
Betatrophin levels were analyzed in participants of the MesyBePo study follow-up cohort (Metabolic Syndrome Berlin-Potsdam) that included healthy control subjects as well as patients with impaired glucose metabolism including overt type 2 diabetes. Baseline Characteristics of the study cohort are summarized in Table 1.

In a crude analysis (Table 2) with respect to different markers of the metabolic syndrome, we found a strong positive correlation of serum betatrophin concentrations with age (r = 0.47; p < 0.001) as it has been described previously [8, 13, 17–19]. Furthermore serum levels of betatrophin were positively correlated to markers of glucose metabolism HbA1c (r = 0.24; p < 0.001) as well as fasting glucose levels (r = 0.20; p < 0.001) and serum glucose concentration 2h after performing a standard oral glucose tolerance test (r = 0.15; p < 0.001). In accordance calculated indices for insulin sensitivity HOMA-IR (r = 0.19; p < 0.001) and QUICKI (r = -0.14, p = 0.001) showed similar associations. Comparing the different subgroups of our cohort with
In our study we found no correlations with parameters of cholesterol metabolism, total cholesterol (r = 0.01; p = 0.83), LDL cholesterol (r = -0.06; p = 0.191) or HDL cholesterol (r = -0.02; p = 0.636) but a positive correlation with triglyceride levels (r = 0.12; p = 0.007). With respect to obesity we found a positive correlation of betatrophin with waist circumference (r = 0.23; p < 0.001) and with participants body mass index (r = 0.12; p = 0.008). Interestingly we found a distinct negative correlation of betatrophin with CKD-EPI eGFR (r = -0.33; p < 0.001). The majority of participants from the MeSyBePo cohort showed a normal to moderately decreased renal function (GFR category 1 | > 90ml/min: 27.8%, GFR category 2 | 60–89 ml/min: 63.1%, GFR category 3 | 30–59 ml/min: 6.0%). In correspondence to the mentioned association the
comparison of betatrophin levels with respect to the usual GFR categories revealed significant differences between each standard GFR category with the highest betatrophin level in the bottom GFR group: Betatrophin—GFR category 1: 469.3 pg/ml (S.E.M. 16.3), GFR category 2: 550.4 pg/ml (S.E.M. 11.3), GFR category 3: 799.0 pg/ml (S.E.M. 79.5), p < 0.001. Since no patients with an eGFR < 30 ml/min participated in our study, we are unfortunately not able to address the role of betatrophin in patients with a severely impaired renal function. Additionally intima media thickness as marker for arteriosclerosis showed a positive correlation with betatrophin (r = 0.26; p < 0.001) as well systolic blood pressure (r = 0.17; p < 0.001). The positive correlation of betatrophin (β = 0.261, p < 0.001) with the intima media thickness was not found when adjusting for age, sex and cardiovascular risk factors.

Notably the negative correlation for renal function remained associated with betatrophin (β = -0.197, p < 0.001) after adjustment for age, gender, abdominal obesity and standard parameters of lipid and glucose metabolism in a multivariate linear regression model (Table 3).

### Conclusion

The initial hope to use betatrophin as drug therapy target for diabetic patients following the discovery that it was potentially able to promote beta-cell proliferation in mice[1] has not been fulfilled so far. On one hand subsequent preclinical studies examining the effects of deletion as well as overexpression of betatrophin showed no evidence for a prominent isolated role of betatrophin in controlling beta-cell proliferation[23, 24]. On the other hand the clinical evaluation of betatrophin in humans yielded a quite heterogeneous pattern with partially contradictory results [11, 18, 25, 26]. The recent retraction of the initial paper describing betatrophin as potential beta cell mitogen [27] and the collected data from clinical studies largely invalidate the perspective of a clinically relevant player in beta cell proliferation. Nevertheless the so far accumulating data on betatrophin suggest a more intermingled role in metabolic function.

### Table 2. Correlation coefficients and corresponding p-values.

| Variable                      | R     | p-value |
|-------------------------------|-------|---------|
| Age                           | 0.469 | < 0.001 |
| Intima Media Thickness        | 0.261 | < 0.001 |
| CKD-EPI eGFR                  | -0.329| < 0.001 |
| Creatinine                    | 0.277 | < 0.001 |
| Systolic Blood Pressure       | 0.171 | < 0.001 |
| Diastolic Blood Pressure      | 0.014 | 0.758   |
| BMI                           | 0.115 | 0.008   |
| Waist Circumference           | 0.227 | < 0.001 |
| Total Cholesterol             | -0.009| 0.830   |
| LDL Cholesterol               | -0.057| 0.191   |
| HDL Cholesterol               | -0.021| 0.636   |
| Triglycerides                 | 0.116 | 0.007   |
| HbA1c                         | 0.235 | < 0.001 |
| Fasting Glucose               | 0.200 | < 0.001 |
| Glucose 2h after OGTT         | 0.150 | 0.001   |
| ASAT                          | 0.064 | 0.141   |
| ALAT                          | 0.055 | 0.205   |
| Smoking                       | 0.073 | 0.093   |

doi:10.1371/journal.pone.0173197.t002
potentially involving the cross talk of lipid and glucose regulation [28]. In our analysis we found betatrophin to be positively correlated to HbA1c, fasting blood glucose and triglyceride levels as well as BMI and waist circumference. This is in line with the majority of the so far published clinical studies. One important aspect addressing the divergent results for betatrophin in humans might be the use of different ELISA kits for detecting circulating betatrophin levels. In this analysis we used the commercially available ELISA kit provided by EIAAB (Catalogue number E1164H; Wuhan, China) which is based on an antibody recognizing the N-terminus and is the most commonly used detection method so far [8, 13, 17, 19, 25, 29, 30]. A recent report compared this ELISA kit to a second commonly used kit from Phoenix Pharmaceuticals (Catalogue number EK-051-55; Burlingame, CA, USA) that is based on an antibody against the C-terminus [31]. While both assays adequately recognized human recombinant betatrophin the results indicate that betatrophin undergoes proteolytic regulation releasing the C-terminal fragments. As consequence the different ELISAs measure either full-length betatrophin only or full-length protein and C-terminal fragment. It is currently not known if either full-length betatrophin or the C-terminal fragment is responsible for the biological effect nor what further processes are involved in betatrophin degradation and clearance. Nevertheless this discrepancy may in part explain the largely diverse results of the clinical studies conducted so far. Numerous different ELISA kits with variable technical properties for detecting the different protein subsets have been used for the analysis of the published clinical data. This aspect points out particularly that great caution is advised for the interpretation and comparison of betatrophin levels and the proteins potential relevance in metabolic regulation. We recently published a study on a potential role of betatrophin in the postprandial switch from lipid to glucose metabolism showing that the impaired betatrophin response in obese subjects is restored after weight loss to a degree comparable to lean individuals [28]. For this trial we used the same ELISA kit for detecting betatrophin further prompting the notion of the full length protein might be the active form of betatrophin. Given betatrophin’s putative dual role in lipid and glucose metabolism as well as its correlation with ageing and in some studies towards an atherogenic cholesterol profile [25, 30], it is tempting to investigate a possible influence on markers of arteriosclerosis. In our study we see no correlation of betatrophin with total, LDL or HDL cholesterol but a positive correlation with systolic blood pressure and intima media thickness. These associations were only detectable in the crude analysis and disappeared when adjusting for age and gender making an independent betatrophin effect less likely. Interestingly, even

| Parameter                  | Correlation | Standardized Beta | Correlation x Standardized Beta x 100 | p-Value       |
|----------------------------|-------------|------------------|---------------------------------------|---------------|
| Age                        | 0.301       | 0.322            | 9.70                                  | < 0.001       |
| CKD-EPI eGFR               | -0.194      | -0.197           | 3.81                                  | < 0.001       |
| Gender                     | -0.157      | -0.150           | 2.34                                  | < 0.001       |
| Waist Circumference        | 0.123       | 0.121            | 1.49                                  | 0.006         |
| Total/HDL Cholesterol Ratio| -0.095      | -0.096           | 0.91                                  | 0.033         |
| HbA1c                      | 0.071       | 0.066            | 0.47                                  | 0.112         |
| Triglycerides              | 0.055       | 0.056            | 0.31                                  | 0.217         |
| Systolic Blood Pressure    | -0.028      | -0.027           | 0.07                                  | 0.533         |
| Smoking                    | 0.001       | 0.001            | 0.00                                  | 0.986         |
| Total                      |             |                  | 19.11                                 | < 0.001       |

The multiplicative term (Correlation x Standardized β x100) explains the variation of betatrophin explained by the respective parameter in percent.

doi:10.1371/journal.pone.0173197.t003
after fully adjusting for cardiovascular risk factors age, gender, waist circumference, total/HDL cholesterol ratio and renal function remain significantly correlated with betatrophin. Similar to the above mentioned results correlations of betatrophin and age can be found in recent clinical studies to be either positive [18, 19] or negative [32]. With respect to sex specific differences betatrophin levels have been described to be significantly higher in males [16] as well as in females [11]. Besides differences in detection methods other factors might be responsible for the discrepancies. The results come from different study populations differing largely in age, ethnicity, basic disease state and medication. In our cohort adjusting for the intake of antihypertensive, lipid lowering or antidiabetic medication has no substantial impact on the described associations while only antidiabetic drug treatment exerts independent influence in the fully adjusted model (std. β = 0.143, p = 0.001). Given the low prevalence of antidiabetic drug treatment (2.8% of all participants) it remains elusive if a distinct pharmacological influence on betatrophin need to be taken into account in general. Notably, data on a possible influence of renal function on circulating betatrophin levels are sparse, taking into consideration that study populations were varying largely by age and are affected by different manifestations of the metabolic syndrome that predispose for an impaired renal function. Some smaller studies identified weak positive correlations with serum creatinine levels not reaching statistical significance [8, 13, 32]. One recent Japanese study in diabetic patients found a significant negative correlation of betatrophin and creatinine clearance as well as eGFR (calculation adjusted for Japanese population) that seemed mainly driven by a common association of the two factors with age [19]. In slight contrast a recent German study, analyzing betatrophin levels in diabetic patients undergoing hemodialysis compared to diabetic individuals with sustained renal function, found a significant positive correlation of betatrophin and eGFR [33]. In this study patients on hemodialysis had significantly lower betatrophin levels compared with subjects having an eGFR > 50ml/min/1.73m². A finding that may argue against a possible renal elimination of betatrophin, that has been the initial hypothesis of the authors. Furthermore a third recent study from Taiwan investigated the correlation of full-length betatrophin with urinary albumin excretion and eGFR as markers of diabetic nephropathy in patients suffering from type 2 diabetes. They found serum betatrophin levels to be higher in patients with macroalbuminuria compared to normoalbuminuria with an independent inverse association to eGFR and speculate betatrophin to be a novel endocrine regulator involved in the progression of diabetic nephropathy [34]. Unfortunately our data acquisition did not include albumin excretion in 24h urine nor urine creatinine levels. Solemnly albumin concentration in the spot urine was measured and didn’t reveal a significant correlation with betatrophin. Given the low prevalence of elevated urinary albumin excretion in our cohort (albumin concentration < 20 mg/l: 86.0%; albumin concentration 20–200 mg/l: 13.3%; albumin concentration > 200 mg/l: 0.7%), we cannot properly address the aspect of an association between betatrophin and urinary albumin excretion or albumin/creatinine ratio as additional markers for kidney function. Nevertheless in the study by Chen et. al. elevated betatrophin levels in type 2 diabetes have been found to be associated with impaired renal function and also with urinary albumin excretion. The same detection method for full-length betatrophin has been applied there and the findings have been interpreted as potential link of betatrophin function with respect to lipid metabolism and the progression of diabetic nephropathy. In our own cohort consisting of 535 patients from the MesyBePo follow-up study with a normal to mildly impaired renal function (average eGFR of 81.2 ml/min at an average of 55.6 years; 6% of participants diagnosed with diabetes) betatrophin showed a similarly strong independent correlation of circulating hormone levels and renal function. Interestingly for other members of the angiopoietin-like protein family such as ANGPTL3, on which betatrophin has a regulatory effect by promoting ANGPTL3 cleavage, a higher serum concentration in patients with kidney disease has been described [35]. The same phenomenon of elevated blood concentrations in
patients with impaired renal function has been documented for a number of other hepatokines and adipokines e.g. FGF21 \cite{36}, chemerin \cite{37} and leptin \cite{38}. Taken together circulating betatrophin levels are independently associated with renal function. Our study further extends the data on betatrophin’s potential role in diabetic nephropathy in terms of linking betatrophin to renal function even in absence of a decisive renal pathology. At this point it remains an open question if betatrophin exerts an effect on kidney function or the progression of a pre-existing nephropathy or if wise versa betatrophin turnover depends on renal function. Further experiments are needed to elucidate physiological significance of this finding and future studies on betatrophin should take a possible influence of renal function into special account.

Acknowledgments

The authors like to thank Nadine Huckauf and Candy Kalischke for excellent technical assistance.

Author Contributions

Conceptualization: LM TB JS.
Data curation: FS AF NS SB.
Formal analysis: LM TB.
Funding acquisition: JS.
Investigation: TB KM.
Methodology: KM AP MM.
Project administration: JS TB.
Resources: JS AP.
Supervision: TB JS KM AP MM.
Validation: FS AF NS SB.
Visualization: LM.
Writing – original draft: LM TB JS.
Writing – review & editing: LM TB JS.

References

1. Yi P, Park JS, Melton DA. Betatrophin: a hormone that controls pancreatic beta cell proliferation. Cell. 2013; 153(4):747–58. PubMed Central PMCID: PMC3756510. doi: 10.1016/j.cell.2013.04.008 PMID: 23623304
2. Quagliarini F, Wang Y, Kozlitina J, Grishin NV, Hyde R, Boerwinkle E, et al. Atypical angiopoietin-like protein that regulates ANGPTL3. Proceedings of the National Academy of Sciences of the United States of America. 2012; 109(48):19751–6. PubMed Central PMCID: PMC3511699. doi: 10.1073/pnas.1217552109 PMID: 23150577
3. Ren G, Kim JY, Smas CM. Identification of RIFL, a novel adipocyte-enriched insulin target gene with a role in lipid metabolism. American journal of physiology Endocrinology and metabolism. 2012; 303(3): E334–51. PubMed Central PMCID: PMC3423120. doi: 10.1152/ajpendo.00084.2012 PMID: 22569073
4. Zhang R. Lipasin, a novel nutritionally-regulated liver-enriched factor that regulates serum triglyceride levels. Biochemical and biophysical research communications. 2012; 424(4):786–92. doi: 10.1016/j.bbrc.2012.07.038 PMID: 22809513
5. Fu Z, Yao F, Abou-Samra AB, Zhang R. Lipasin, thermoregulated in brown fat, is a novel but atypical member of the angioptelin-like protein family. Biochemical and biophysical research communications. 2013; 430(3):1126–31. doi: 10.1016/j.bbrc.2012.12.025 PMID: 23261442

6. Wang Y, Quagliarini F, Gusarova V, Gromada J, Valenzuela DM, Cohen JC, et al. Mice lacking ANGPTL8 (Betatrophin) manifest disrupted triglyceride metabolism without impaired glucose homeostasis. Proc Natl Acad Sci U S A. 2013; 110(40):16109–14. PubMed Central PMCID: PMCPMC3791734. doi: 10.1073/pnas.1315292110 PMID: 24043787

7. Fu Z, Berhane F, Fite A, Seyoum B, Abou-Samra AB, Zhang R. Elevated circulating lipasin/betatrophin in human type 2 diabetes and obesity. Sci Rep. 2014; 4:5013. doi: 10.1038/srep05013 PMID: 24852694

8. Espes D, Lau J, Carlsson PO. Increased circulating levels of betatrophin in individuals with long-standing type 1 diabetes. Diabetologia. 2014; 57(1):50–3. PubMed Central PMCID: PMCPMC3855541. doi: 10.1007/s00125-013-3071-1 PMID: 24078058

9. Trebolic TK, Klimek P, Thomas A, Fenzl A, Leitner K, Springer S, et al. Circulating Betatrophin Is Strongly Increased in Pregnancy and Gestational Diabetes Mellitus. PLoS One. 2015; 10(9):e0136701. PubMed Central PMCID: PMCPMC4556632. doi: 10.1371/journal.pone.0136701 PMID: 26325425

10. Guo K, Lu J, Yu H, Zhao F, Pan P, Zhang L, et al. Serum betatrophin concentrations are significantly increased in overweight but not in obese or type 2 diabetic individuals. Obesity (Silver Spring). 2015; 23(4):793–7.

11. Gomez-Ambrosi J, Pascual E, Catalan V, Rodriguez A, Ramirez B, Silva C, et al. Circulating betatrophin concentrations are decreased in human obesity and type 2 diabetes. J Clin Endocrinol Metab. 2014; 99(10):E2004–9. doi: 10.1210/jc.2014-1568 PMID: 25059091

12. Gao T, Jin K, Chen P, Jin H, Yang L, Xie X, et al. Circulating Betatrophin Correlates with Triglycerides and Postprandial Glucose among Different Glucose Tolerance Statuses—A Case-Control Study. PLoS One. 2015; 10(8):e0133640. PubMed Central PMCID: PMCPMC4527834. doi: 10.1371/journal.pone.0133640 PMID: 26247824

13. Yamada H, Saito T, Aoki A, Asano T, Yoshida M, Ikoma A, et al. Circulating betatrophin is elevated in patients with type 1 and type 2 diabetes. Endocr J. 2015; 62(5):417–21. doi: 10.1507/endocrj.EJ14-0525 PMID: 25753914

14. Crujeiras AB, Zuulet MA, Abete I, Amil M, Carreira MC, Martinez JA, et al. Interplay of atherogenic factors, protein intake and betatrophin levels in obese-metabolic syndrome patients treated with hypocaloric diets. Int J Obes (Lond). 2015.

15. Barja-Fernandez S, Folgueira C, Secoane LM, Casanueva FF, Dieguez C, Castelao C, et al. Circulating Betatrophin Levels Are Increased in Anorexia and Decreased in Morbidly Obese Women. J Clin Endocrinol Metab. 2015; 100(9):E2004–9. doi: 10.1210/JC.2015-1595 PMID: 26171798

16. Wu S, Gao H, Ma Y, Fu L, Zhang C, Luo X. Characterisation of betatrophin concentrations in childhood and adolescent obesity and insulin resistance. Pediatr Diabetes. 2014.

17. Abu-Farha M, Abubaker J, Al-Khairi I, Cherian P, Noronha F, Hu FB, et al. Higher plasma betatrophin/ANGPTL8 level in Type 2 Diabetes subjects does not correlate with blood glucose or insulin resistance. Sci Rep. 2015; 5:10949. PubMed Central PMCID: PMCPMC4650613. doi: 10.1038/srep10949 PMID: 26077345

18. Hu H, Sun W, Yu S, Hong X, Qian W, Tang B, et al. Increased circulating levels of betatrophin in newly diagnosed type 2 diabetic patients. Diabetes Care. 2014; 37(10):2718–22. doi: 10.2337/dc14-0602 PMID: 25024395

19. Tokumoto S, Hamamoto Y, Fujimoto K, Yamaguchi E, Okamura E, Honjo S, et al. Correlation of circulating betatrophin concentrations with insulin secretion capacity, evaluated by glucagon stimulation tests. Diabet Med. 2015; 32(5):653–6. doi: 10.1111/dme.12696 PMID: 25655786

20. Bobbert T, Schwarz F, Fischer-Rosinsky A, Pfeiffer AF, Mohlig M, Mai K, et al. Fibroblast growth factor 21 predicts the metabolic syndrome and type 2 diabetes in Caucasians. Diabetes Care. 2013; 36(1):145–9. PubMed Central PMCID: PMCPMC3526237. doi: 10.2337/dc12-0703 PMID: 22933429

21. Organization WH. Report of a WHO/IDF Consultation: Definition and diagnosis of diabetes mellitus and intermediate hyperglycemia. 2005.

22. Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF 3rd, Feldman HI, et al. A new equation to estimate glomerular filtration rate. Ann Intern Med. 2009; 150(9):604–12. PubMed Central PMCID: PMCPMC2763564. PMID: 19414839

23. Gusarova V, Alexa CA, Na E, Stevis PE, Xin Y, Bonner-Weir S, et al. ANGPTL8/betatrophin does not control pancreatic beta cell expansion. Cell. 2014; 158(3):691–6. PubMed Central PMCID: PMCPMC4243040. doi: 10.1016/j.cell.2014.09.027 PMID: 25417115
24. Jiao Y, Le Lay J, Yu M, Najj A, Kaestner KH. Elevated mouse hepatic betatrophin expression does not increase human beta-cell replication in the transplant setting. Diabetes. 2014; 63(4):1283–8. PubMed Central PMCID: PMCPMC3964501. doi: 10.2337/db13-1435 PMID: 24353178

25. Chen X, Lu P, He W, Zhang J, Liu L, Yang Y, et al. Circulating betatrophin levels are increased in patients with type 2 diabetes and associated with insulin resistance. J Clin Endocrinol Metab. 2015; 100(1):E96–100. doi: 10.1210/jc.2014-2300 PMID: 25303484

26. Gokulakrishnan K, Manokaran K, Pandey GK, Amutha A, Ranjani H, Anjana RM, et al. Relationship of betatrophin with youth onset type 2 diabetes among Asian Indians. Diabetes Res Clin Pract. 2015; 109(1):71–6. doi: 10.1016/j.diabres.2015.04.028 PMID: 25981223

27. Yi P, Park J-S, Melton DA. Retraction Notice to: Betatrophin: A Hormone that Controls Pancreatic β-Cell Proliferation. Cell. 2017; 168(1–2):326. doi: 10.1016/j.cell.2016.12.017 PMID: 28038792

28. Maurer L, Brachs S, Decker AM, Brachs M, Leupelt V, Jumpertz von Schwartzenberg R, et al. Weight Loss Partially Restores Glucose-Driven Betatrophin Response in Humans. J Clin Endocrinol Metab. 2016; 101(11):4014–20. doi: 10.1210/jc.2016-1788 PMID: 27459526

29. Espes D, Martinell M, Carlsson PO. Increased circulating betatrophin concentrations in patients with type 2 diabetes. Int J Endocrinol. 2014; 2014:323407. PubMed Central PMCID: PMCPMC4055101. doi: 10.1155/2014/323407 PMID: 24963292

30. Fenzl A, Itariu BK, Kosi L, Fritzer-Szekeres M, Kautzky-Willer A, Stulnig TM, et al. Circulating betatrophin correlates with atherogenic lipid profiles but not with glucose and insulin levels in insulin-resistant individuals. Diabetologia. 2014; 57(6):1204–8. doi: 10.1007/s00125-014-3208-x PMID: 24623100

31. Fu Z, Abou-Samra AB, Zhang R. An explanation for recent discrepancies in levels of human circulating betatrophin. Diabetologia. 2014; 57(10):2232–4. doi: 10.1007/s00125-014-3346-1 PMID: 25099942

32. Ebert T, Kralisch S, Wurst U, Lossner U, Kratzsch J, Blüher M, et al. Betatrophin levels are increased in women with gestational diabetes mellitus compared to healthy pregnant controls. Eur J Endocrinol. 2015; 173(1):1–7. doi: 10.1530/EJE-14-0815 PMID: 25808282

33. Ebert T, Kralisch S, Hoffmann A, Bachmann A, Lossner U, Kratzsch J, et al. Circulating angiopoietin-like protein 8 is independently associated with fasting plasma glucose and type 2 diabetes mellitus. J Clin Endocrinol Metab. 2014; 99(12):E2510–7. doi: 10.1210/jc.2013-4349 PMID: 25325797

34. Chen CC, Susanto H, Chuang WH, Liu TY, Wang CH. Higher serum betatrophin level in type 2 diabetes subjects is associated with urinary albumin excretion and renal function. Cardiovasc Diabetol. 2016; 15:3. PubMed Central PMCID: PMCPMC4704426. doi: 10.1186/s12933-015-0326-9 PMID: 26739836

35. Mahmood D, Makoveichuk E, Nilsson S, Olivecrona G, Stegmayr B. Response of angiopoietin-like proteins 3 and 4 to hemodialysis. Int J Artif Organs. 2014; 37(1):13–20. doi: 10.5301/ijao.5000252 PMID: 24634330

36. Stein S, Bachmann A, Lossner U, Kratzsch J, Blüher M, Stumvoll M, et al. Serum levels of the adipokine FGF21 depend on renal function. Diabetes Care. 2009; 32(1):126–8. PubMed Central PMCID: PMCPMC2066845. doi: 10.2337/dc08-1054 PMID: 18840768

37. Pfau D, Bachmann A, Lossner U, Kratzsch J, Blüher M, Stumvoll M, et al. Serum levels of the adipokine chemerin in relation to renal function. Diabetes Care. 2010; 33(1):171–3. PubMed Central PMCID: PMCPMC2797967. doi: 10.2337/dc09-1351 PMID: 19837789

38. Merabet E, Dagogo-Jack S, Coyne DW, Klein S, Santiago JV, Hmiel SP, et al. Increased plasma leptin concentration in end-stage renal disease. J Clin Endocrinol Metab. 1997; 82(3):847–50. doi: 10.1210/jcem.82.3.84717 PMID: 9062494