Erythrocyte Alloimmunization in Children with Sickle Cell Disease

Daniela de Oliveira Werneck Rodrigues1,2,4*, Lysla Cardoso Sudário3, Olivia Franco dos Santos1, Leonardo de Angelli Benedito Cardoso1 and Luiz Claudio Ribeiro1

1Universidade Federal de Juiz de Fora, Juiz de Fora, Brazil  
2Fundação Hemominas, Brazil  
3Hospital Maternidade Therezinha de Jesus, Juiz de Fora, Brazil  
4Researcher at NHLBT-NIH, USA

Introduction

Sickle cell disease (SCD) comprises a group of hematological alterations caused by the presence of hemoglobin S (Hb S), either homozygous SS or in combination with other variants of hemoglobins (Hb) - such as HbC and other of hemoglobinopathies like Beta Thalassemia [1,2]. It is estimated that about 3,000 children are born each year with SCD in Brazil [3].

Transfusion of red blood cells (RBC) has a crucial role in the therapeutic arsenal of SCD, resulting in reduced morbidity and mortality [4-9]. One complication of RBC transfusion therapy is alloimmunization, an immune response against blood group antigens triggered by a genetic disparity between donor and recipient [10,11]. According to Schonewille, et al. [7], the risk of alloimmunization depends on the exposure of the recipient to a foreign antigen, the immunogenicity of this antigen and the number and frequency of transfusions. Genetic factors controlling inflammatory responses may activate the innate immune system and convert an inert or even a tolerogenic event into an immunogenic one [9].

Alloimmunization occurs more frequently in SCD when...
compared to other diseases such as cancer and other hemoglobinopathies, with alloimmunization rates between 18-47% for SCD [7,9,10]. The main alloantibodies found are directed against antigens of the Rhesus system, Kell, Duffy, Kidd, Lewis and MNS, and is primarily due to antigenic differences between blood recipients and donors [10,12-17].

The objectives of this study were to determine the prevalence and alloimmunization rate among transfused children with SCD treated at Hemominas Juiz de Fora (JFO), as well as to identify the most common antibodies in alloimmunization and risk factors for alloimmunization in this population.

Materials and Methods

Population

A retrospective cohort study was conducted to evaluate the prevalence of alloimmunization among 120 children born between 1998-2007 diagnosed with SCD through the Programa Estadual de Triagem Neonatal de Minas Gerais (PETN-MG), treated at Hemominas Juiz de Fora (JFO). Standardized case report forms were used to abstract data from medical records. All patients included in the cohort were followed up for at least 7 years.

The researchers decided to include all the children in the study and not to do the sample calculation, considering the reduced number of patients with alloimmunization. The children were stratified into transfused and non-transfused groups. The 72 children transfused were analyzed for this study (Figure 1). Ten children who died (9 from sepsis and 1 from splenic sequestration) were not included.

Variables studied

The data collection period started in August 2013 and was completed in December 2014. Variables abstracted from medical record included: Sex, age, type of hemoglobinopathy, RBC phenotype for ABO, Rh (C, c, E, e, Cw, D), Kell (K, k), Duffy (Fya, Fyb), Kidd (Jka, Jkb), Lewis (Lea, Leb), MNSs, Indirect Antiglobulin Test (IAT - RBC Antibody Screen), date of diagnosis of positive IAT, number of transfusions, location of transfusions, number of transfusions by age, number of previous units transfused to the positive IAT, identified alloantibodies and chronic transfusion treatment.

Children were stratified into patients transfused only at JFO, transfused at JFO and another facility or transfused at another facility only, as well as the type of blood leukoreduced. For the analysis, the number of transfusions was categorized into 2 groups: (1) Up to ten and (2) Above 10 transfusions in the observation period. The outcome variable for all analyses was development of RBC antibody.

Laboratory tests

Testing for abnormal hemoglobin was performed by high performance liquid chromatography (HPLC) and isoelectric focusing electrophoresis (IEF) on blood obtained by heel stick and applied to filter paper.

Confirmatory diagnosis of SCD and their respective classification in SS, SC, S beta-thalassemia (Sβ+, Sβ0) was performed by hemoglobin electrophoresis at alkaline and acid pH at 6 and 12-months-old; determination of A2 Hemoglobin levels by chromatography and Fetal Hemoglobin by radial immunodiffusion.

The determination of the RBC phenotype and screening for RBC antibodies relied on the collection of 5 ml of whole blood, as defined in the Sickle Cell Disease Guideline - Basic Flue treatment - available in http://www.saude.gov.br/bvs. All samples with positive IAT - antibody screening had antibody identification with erythrocyte antigens panel. The samples were tested for the direct and reverse blood typing using the tube technique [18]. The Direct and Indirect Anti-globin Test, erythrocyte panel, autoantibody screen and phenotyping of other blood systems were performed by gel centrifugation technique using specific gel-test card (DiaMed® Latin America SA, Brazil).

Erythrocyte phenotyping for the Rh (D, C, E, e), Kell (K, k), Duffy (Fya, Fyb) and Kidd (Jka, Jkb) systems antigens was performed by column-gel-test (Diamed AG, Morat, Switzerland) with polyclonal or monoclonal antibodies. The research and identification of irregular antibodies was performed using a gel-test hemaglutination method using phenotyped red cells (DiaMed® AG, Cressier sur Morat®, Switzerland). Whenever necessary, direct polymorphic or mono specific human antiglobulin tests, antibody adsorption and elution tests, and indirect human antiglobulin tests with selected phenotype red blood cells were also performed.

The IAT - antibody screening is annual and recommended three months after each transfusion. In the ideal situation, RBC antibody tests after transfusion should be performed twice, the first time shortly after transfusion, to detect boosting of existing antibodies or fast appearing new antibodies (i.e., anti-Jka and anti-E), and a second time after a longer period of time, to detect slower developed antibody specificities. Our study design does not allow us to be more specific on the time points.

Statistical analysis

The statistical analysis of the association was performed using the Mann-Whitney U test for non-parametric data, Student’s T-Test for parametric data and Chi-square test for proportions using SPSS Inc., Chicago14® for all analyses. The statistical analysis was univariate due to the reduced number of cases with positive irregular antibody. A p value of < 0.05 was considered statistically significant.
Ethical considerations

This study project was submitted to the Plataforma Brasil and approved by the Research Ethics Committee of Fundação Hemominas (No. 716.359). It is also in accordance with the provisions of Resolution No. 466/12 of the National Health Council, Declaration of Helsinki (2008) and the Code of Medical Ethics 1988 (Article 122-130).

Results

Of the 120 children born between the years 1998-2007 with SCD diagnosis through the PETN-MG and treated at Hemominas 110 were evaluated for this study and 72 were transfused (Figure 1). Among these 110 children studied, 52.7% were male.

The average age at the end of follow-up was 11.2 ± 2.84 years-old (range 6-16). The majority of the patients were SS and the distribution of the genotypes is shown in the graph below (Figure 2).

There were 10 children (9.1%) with anti-erythrocyte antibodies, 8 children had received transfusion and 2 had naturally alloimmunized without previous history of transfusion or pregnancy. As for children with positive IAT - antibody screening, 6 were male, 9 children had SS and 1 a child was of the SC genotype.

The analyzed group was composed of 72 transfused children, 55.6% of which were male. The average age at the end of the extraction of the data was 11.5 years (range 7-16 and standard deviation 2.74). Regarding the diagnosis, 76.4% had SS, 15.3% SC, 5.6% Sβ+ and 2.8% Sβ0.
As for the blood classification in the ABO/Rh system between children with SS, there was a higher prevalence of the O/Positive (45.5%), followed by group A/Positive (23.6%). Similar distribution was shown in the transfused group. In SCD SC type and S/b group, O/Positive was also predominant (54.5% and 75%, respectively).

The study of irregular antibodies was positive in 8 transfused individuals with prevalence of 11.1%. All alloimmunized children had SS, three belonging to blood group O/Positive, two AB/Positive, two B/Positive and two A/Positive. The mean age at the time of positive IAT - antibody screening was 5.25-years-old (range 1-8 and standard deviation 2.37).

Among the 48 children who received between 1 to 10 transfusions, 3 were RBC antibody positive (6.3%), and among the 24 who received more than 10 transfusions, 5 had RBC antibodies (20.8%). The relative risk (RR = 0.300 – 95% Confidence Interval: 0.078-1.151) indicates that the risk of having positive IAT - antibody screening among subjects who received up to 10 transfusions is 70% lower than those who received higher amounts, though this difference was not statistically significant (p = 0.063).

Regarding the type of transfused blood component, 59.3% of children received RBC, 11% received phenotyped leukoreduced RBC and 29.7% received both RBC and phenotyped leukoreduced RBC. The analysis of the positive IAT - antibody screening, type of RBC infused and the local of transfusion showed that despite the use of phenotyped Trocar para and leukoreduced RBC there was development of alloantibodies in this study (p = 0.00).

Children with SCD were studied in age ranges with the extraction of the number of transfusions in each interval. In the group with antibodies, the total number of transfusions in the 0-4 years-old subgroup was 5 RBC units (range 0-22 and standard deviation 7.8). The children between 5 and 10 years received an average of 11.6 units (range 1-35 and standard deviation of 12.7). From the age of 11, children with SCD had an average of 2.6 units transfused RBC (minimum of 0, maximum of 17 and standard deviation of 5.9). Statistical analysis was performed for comparison with non-transfused group alloimmunized, using the Mann-Whitney test and there was no statistical significance (p = 0.420, p = 0.064 and p = 0.825 for each age interval).

As for the amount of transfusions, the positive IAT - antibody screening group has average 19.25, median 11.5, with minimum number of two and maximum of 67 units. Negative IAT - antibody screening group has average 11.8, median 5.0, minimum one unit, and maximum 67 units.

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### Table 1: Demographic data of Alloimmunized vs. Not-alloimmunized.

| Demographic data                          | Alloimmunized (n = 8) | Not-Alloimmunized (n = 64) | P value |
|------------------------------------------|-----------------------|----------------------------|---------|
| Age (years old)                          | 12.25 (± 2.5)         | 11.48 (± 2.8)              | 0.460*  |
| Gender                                   |                       |                            |         |
| Male                                     | 5 (62.5%)             | 35 (54.7%)                 | 0.675** |
| Female                                   | 3 (37.5%)             | 29 (47.3%)                 |         |
| Diagnose                                 |                       |                            | 0.095** |
| SCA                                      | 8 (100%)              | 47 (73.4%)                 |         |
| SCD                                      | 0                     | 17 (26.6%)                 |         |
| Type of Blood Component                  |                       |                            | 0.464** |
| RBC (NON Phenotyped leukoreduced)        | 4 (50%)               | 38 (59.3%)                 |         |
| Phenotyped leukoreduced RBC              | 2 (25%)               | 7 (11%)                    |         |
| RBC and Phenotyped leukoreduced RBC      | 2 (25%)               | 19 (29.7%)                 |         |
| Total number of transfusions             |                       |                            |         |
| Total                                    | 19.25 (± 21.9)        | 11.8 (± 20.2)              | 0.085** |
| Total number of transfusions within Age groups |     |                            |         |
| 0-4 years-old                            | 5.0 (± 7.8)           | 2.2 (± 4.0)                | 0.420** |
| 5-10 years-old                           | 11.6 (± 12.7)         | 6.3 (± 10.8)               | 0.064*  |
| Over 10-years-old                        | 2.6 (± 5.9)           | 3.3 (± 13.2)               | 0.825** |
| Total number of transfusions             |                       |                            | 0.063** |
| ≤ 10 transfusions                        | 3 (6.3%)              | 45 (93.8%)                 |         |
| > 10 transfusions                        | 5 (20.8%)             | 19 (79.2%)                 |         |
| RBC units transfused previous positive IAT - Antibody screening | 8.0 (± 12.9) | -                         |         |
| Chronic Transfusion Therapy              | 2 (22.2%)             | 7 (77.8%)                  | 0.257*  |

* Student’s T-test, ** Mann-Whitney’s U test, *** Pearson’s Chi Square Test
Immunocompetent subjects (exposed to RBC transfusion, simple or chronic) described that 50 to 70% of patients with SCD could never develop alloantibodies despite frequent blood transfusions or chronic transfusion treatment [12].

The average age of identification of irregular antibody was 5.25-years-old. Verduzco and Nathan reported that blood transfusion exposure at an early age could have likely protective role against alloimmunization [22].

The analysis of our data on the type of transfused blood components showed that 25% of the subjects received phenotyped and leukoreduced RBC. In other health institutions, 50% of children received transfusion of RBC concentrate on a sporadic basis. These data are very important due to possible correlation between the administration of phenotypically identical blood components and lower alloimmunization rates in SCD populations with multiple transfusions that were performed at Fundação Hemominas [22,23].

The presence of antibodies belonging to Rh system was observed in 9.8%, and two (22.2%) patients on chronic transfusion regimen were alloimmunized. Chou, et al. [20] identified the same phenotypes of the Rh system in patients with SCD undergoing hypertransfusion regime with SCD and sporadic transfusions (12%), suggesting that the implementation of extended genotyping research could avoid isoimmunization with reduction of genetic differences between receiver and donor. The Fundação Hemominas is gradually implementing extended genotyping to all donors and the Rh system for patients with SCD in order to improve the compatibility between transfusions and reduce the risk of developing alloantibodies.

Eight patients were observed in chronic transfusion treatment, receiving 416 transfusions (52 units per patient) which give an alloimmunization rate of 0.41/100. Recent studies have found alloimmunization rate of 0.55/10024; 0.30/10020 and 0.11/10023. The complexity of alloimmunization becomes increasingly a challenge in transfusion therapy in SCD, since beyond the phenotypic differences between donor and recipient there is likely a relationship between the increased formations of antibodies with the increasing number of exposures [24]. Our results did not show any significance between alloimmunization and the amount of transfusions. Therefore, this data should be analyzed with caution, since the number of alloimmunized is quite low (8 individuals) and the p value was only slightly higher than 5% (0.085).

This study, as well as Fasano, et al. did not show statistical significance related to the number of transfused units, presence of alloantibodies and positive IAT-antibody screening. It was not possible to confirm the risk of alloimmunization in patients undergoing chronic transfusion therapy, despite increased exposure to erythrocyte antigens [5], perhaps because of the small...
population enrolled in this study. There is an ongoing multicenter study searching to establish the association between the number of transfused units and the risk of developing alloimmunization (REDS-III).

Nickel, et al. showed that alloimmunization was more significant in older children with an average age of 13 years and with more exposure to transfusion, without association with sex, age of onset of chronic transfusion treatment and transfusion outside the study institution. The data found in our study were different regarding the average age of alloimmunization (5.25-years-old) and consistent in relation to other variables. Research in Immunohematology area and expanded genotyping may help to prevent and reduce the incidence of alloimmunization in this population [25-31].

Red blood cell transfusion was common in Brazilian patients with SCD, with use driven by CTT. Kelly, et al. described in the Recipient Epidemiology and Donor Evaluation (REDS)-II Brazil cohort, a prevalence of erythrocyte antibody formation in 26.2% of children, consistent with the data found in this study and in the literature [32].

Conclusions

Alloimmunization is frequent in patients with SCD, attended at JFO, particularly in multiple transfusions. The immunologic sensitization in patients in the chronic transfusion group occurred on dates prior to the beginning of chronic transfusion treatment. Transfusion therapy in the presence of pro-inflammatory events, particularly in SCD, can generate immune response and trigger alloimmunization. There is also a need for genetically similar blood donors, and the miscigenation of the Brazilian population may contribute in the development of alloimmunization.

The prevalence of alloimmunization in children in this study was 11.1% and 50% of the expressed antibodies belonged to the Rh system, as already presented in the literature. It is important to note that even with the offer of phenotyped and leukoreduced RBC and patients without any transfusion therapy exposure the alloimmunization phenomenon could be detected.

The uses of blood products not phenotyped/leukoreduced and cumulative exposure during emergency treatment are likely contributors to the awareness of children with SCD. Alloimmunization has become frequent with the wide use of chronic transfusion treatment, new studies and methods, such as the implementation of genotyping erythrocyte are mandatory to reduce the identification of gaps gene variants of the Rh system.

The development of alloantibodies is considered a challenge in the treatment of SCD, with restrictive implications for transfusion therapy, considering the early diagnosis, the implementation of transcranial doppler, access to treatment and increased survival in SCD, especially among children with formal indication for chronic transfusion treatment because Stroke risk.

The prescription of phenotyped leukoreduced and genotyped RDC throughout the health team involved in the treatment and follow-up of children with sickle cell disease should be a strategy to try to reduce the development of alloimmunization related to transfusion therapy and the immune complications determined by the presence of alloantibodies.

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Conflicts of Interest

The authors declare no conflicts of interest.

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