Original article

Pericardial and myocardial involvement after SARS-CoV-2 infection: a cross-sectional descriptive study in healthcare workers

Rocio Eiros, Manuel Barreiro-Pérez, Ana Martín-García, Julia Almeida, Eduardo Villacorta, Alba Pérez-Pons, Soraya Merchán, Alba Torres-Valle, Clara Sánchez-Pablo, David González-Calle, Oihane Pérez-Escurza, Inés Toranzo, Elena Díaz-Peláez, Blanca Fuentes-Herrero, Laura Macías-Alvarez, Guillermo Oliva-Ariza, Quentin Lecrevisse, Rafael Flucta, José L. Bravo-Grande, Alberto Orfao and Pedro L. Sánchez

* Servicio de Cardiología, Hospital Universitario de Salamanca, Salamanca, Spain
** Centro de Investigación Biomédica en Red Enfermedades Cardiovasculares (CIBERCV), Spain
* Instituto de Investigación Biomédica de Salamanca (IRBASAL), Salamanca, Spain
* Facultad de Medicina, Universidad de Salamanca, Salamanca, Spain
* Centro de Investigación del Cáncer, Universidad de Salamanca-CSIC, Salamanca, Spain
* Servicio de Citometría, Nucleus - Universidad de Salamanca, Salamanca, Spain
* Centro de Investigación Biomédica en Red Cáncer (CIBERONC), Madrid, Spain
* Servicio de Prevención de Riesgos Laborales, Hospital Universitario de Salamanca, Salamanca, Spain

Article history:
Received 28 May 2021
Accepted 22 October 2021
Available online 5 November 2021

Keywords:
SARS-CoV-2
COVID-19
Healthcare worker
Cardiac injury
Pericarditis
Myocarditis
Cardiac magnetic resonance
Immunе response
Immune cells
Serology

A B S T R A C T

Introduction and objectives: The cardiac sequelae of SARS-CoV-2 infection are still poorly documented. We conducted a cross-sectional study in healthcare workers to report evidence of pericardial and myocardial involvement after SARS-CoV-2 infection.

Methods: We studied 139 healthcare workers with confirmed past SARS-CoV-2 infection. Participants underwent clinical assessment, electrocardiography, and laboratory tests, including immune cell profiling and cardiac magnetic resonance (CMR). Clinically suspected pericarditis was diagnosed when classic criteria were present and clinically suspected myocarditis was based on the combination of at least 2 CMR criteria.

Results: Median age was 52 (41-57) years, 71.9% were women, and 16.5% were previously hospitalized for COVID-19 pneumonia. On examination (10.4 [9.3-11.0] weeks after infection-like symptoms), participants showed hemodynamic stability. Chest pain, dyspnea or palpitations were present in 41.7% participants, electrocardiographic abnormalities in 49.6%, NT-proBNP elevation in 7.9%, troponin in 0.7%, and CMR abnormalities in 60.4%. A total of 30.9% participants met criteria for either pericarditis and/or myocarditis: isolated pericarditis was diagnosed in 5.8%, myopericarditis in 7.9%, and isolated myocarditis in 17.3%. Most participants (73.2%) showed altered immune cell counts in blood, particularly decreased eosinophil (27.3%; P < .001) and increased cytotoxic T cell numbers (17.3%; P < .001). Clinically suspected pericarditis was associated (P < .005) with particularly elevated cytotoxic T cells and decreased eosinophil counts, while participants diagnosed with clinically suspected myopericarditis or myocarditis had lower (P < .05) neutrophil counts, natural killer-cells, and plasma cells.

Conclusions: Pericardial and myocardial involvement with clinical stability are frequent after SARS-CoV-2 infection and are associated with specific immune cell profiles.

© 2021 Sociedad Española de Cardiología. Published by Elsevier España, S.L.U. All rights reserved.

https://doi.org/10.1016/j.rec.2021.11.001
1885-5857 © 2021 Sociedad Española de Cardiología. Published by Elsevier España, S.L.U. All rights reserved.
Afección pericárdica y miocárdica tras infección por SARS-CoV-2: estudio descriptivo transversal en trabajadores sanitarios

RESUMEN

Introducción y objetivos: Las secuelas cardíacas tras la infección por SARS-CoV-2 todavía están poco documentadas. Se realizó un estudio transversal en trabajadores sanitarios para estudiar la prevalencia de afección pericárdica y miocárdica tras la infección por SARS-CoV-2.

Métodos: Se estudió a 139 trabajadores sanitarios con infección previa confirmada por SARS-CoV-2. Los participantes se sometieron a una evaluación clínica, electrocardiograma, laboratorio, incluido el perfil de células inmunitarias, y resonancia magnética cardíaca (RMC). El diagnóstico clínico de pericarditis se realizó ante la presencia de los criterios clásicos y el diagnóstico clínico de miocarditis ante la presencia de al menos 2 criterios de RMC.

Resultados: La mediana de edad fue de 52 (41–57) años, el 71,9% eran mujeres, y el 16,5% había sido hospitalizado previamente por neumonía por COVID-19. En la evaluación (10,4 [9,3–11,0] semanas después de los síntomas de infección), todos los participantes presentaban estabilidad hemodinámica. El 41,7% presentaba dolor torácico, disnea o palpitaciones; el 49,6%, alteraciones electrocardiográficas; el 7,9%, elevación de NT-proBNP; el 0,7%, elevación de troponina; y el 60,4%, alteraciones en la RMC. Un total de 30,9% de participantes cumplieron los criterios clínicos establecidos de pericarditis o miocarditis: pericarditis aislada en el 5,8%, miopericarditis en el 7,9% y miocarditis aislada en el 17,3%. La mayoría de los participantes (73,2%) mostraron recuentos de células inmunitarias alteradas en sangre; en particular disminución de eosinófilos (27,3%; p < 0,001) y aumento del número de células T citotóxicas (17,3%; p < 0,001). La sospecha clínica de pericarditis se asoció (p < 0,005) particularmente con un elevado número de células T citotóxicas y recuento de eosinófilos disminuidos; mientras que los participantes con sospecha clínica de miopericarditis o miocarditis tenían recuentos de neutrófilos, células natural killer y células plasmáticas más bajos (p < 0,05).

Conclusiones: La afección pericárdica y miocárdica con estabilidad hemodinámica es frecuente después de la infección por SARS-CoV-2 y se asocia con perfiles de células inmunitarias específicas.

© 2021 Sociedad Española de Cardiología. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.

INTRODUCTION

The novel severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2) is currently causing a sustained COVID-19 pandemic, with the risk of causing long-term cardiac sequelae in the infected population.¹ The fear of SARS-CoV-2 causing greater myocardial damage than other conventional viruses is based on its mechanism of infecting human cells by binding to the transmembrane angiotensin converting-enzyme 2—which is mainly expressed by cells in alveoli and myocardial tissue—the rise in troponin levels observed in COVID-19 patients hospitalized with pneumonia and its association with increased mortality, and the probably reduced innate antiviral defenses against a novel virus.²

Pericarditis and myocarditis after conventional viral infections both stem from an inadequate or excessive immune response driven by T and B cell-mediated mechanisms.²³⁻¹⁰ If there is an inadequate response, continued viral replication in the perimyocardium protracts inflammation by attracting killer T cells and the concomitant production of chemokines and cytokines. In contrast, molecular mimicry can result in the production of autoantibodies against cardiac proteins, leading to a cardio-specific autoimmune response that causes sustained inflammation, effusion, or cardiac remodeling. However, the specific immune profiles that occur after SARS-CoV-2 infection, particularly in patients showing cardiac sequelae remain unknown.³⁻⁷

The present study was designed to search for evidence of pericardial and myocardial involvement after past SARS-CoV-2 infection comprehensively studied by clinical assessment, laboratory tests, electrocardiography and cardiac magnetic resonance (CMR) imaging. Additionally, participants underwent an in-depth characterization of the immune cell compartments in blood and the virus-specific humoral immune response. As healthcare workers have been the group most affected by SARS-CoV-2 in Spain but have also been subject to more testing than the rest of the population, we decided to conduct the study in this singular cohort.

METHODS

Study design and healthcare worker participants

This cross-sectional study consecutively recruited 142 healthcare workers with laboratory confirmed SARS-CoV-2 infection in the University Hospital of Salamanca, and who volunteered for the study. Among them, 106 healthcare workers tested positive for SARS-CoV-2 by reverse-transcriptase polymerase chain reaction (RT-PCR) from nasopharyngeal swab between March 13 and April 25, 2020; and 36 healthcare workers were diagnosed after testing positive for anti-SARS-CoV-2-immunoglobulin G (IgG) antibodies by ELISA between April 10 and May 22, 2020, as part of a major hospital campaign. The purpose of this second group was to also provide data from participants with past SARS-CoV-2 infection, who are more likely to have mild symptoms of viral infection, and because population-based SARS-CoV-2 seroprevalence studies are becoming more established.⁶⁻⁷ Study enrolment began on May 25 and finished on June 12, 2020.
Institutional approval (2020/05/490) for the study was provided by the University Hospital of Salamanca Ethics Committee, and all participants provided written informed consent. The study is registered with ClinicalTrials.gov NCT04413071. The responsibility for the study design, data collection and data interpretation lay solely with the study investigators. An internal adjudication monitoring board reviewed all cardiac study findings and adjudicated study outcomes. The authors had full access to all the data and elaborated all materials to submit for publication.

Investigation process and procedures

All participants underwent clinical evaluation, electrocardiography, laboratory tests and CMR imaging at the same visit. After obtaining written informed consent, trained interviewers used a structured questionnaire to collect baseline data in face-to-face interviews. A cardiologist took a complete medical history, performed a physical examination and reviewed the completeness of the questionnaire in a separate room, where an electrocardiogram was performed, and blood samples were drawn immediately before the CMR. Electrocardiograms were interpreted in consensus by 2 experienced readers, who were blinded to participant identification, clinical history, symptoms, physical examination, and other findings.

CMR was performed using a clinical 1.5 whole-body magnetic resonance scanner in the cardiac imaging laboratory of the University Hospital of Salamanca. The imaging acquisition protocol is described in detail in Methods of the supplementary data. CMR images were globally and regionally analyzed using dedicated software, in consensus by 2 experienced readers, who were blinded in a similar manner to the electrocardiogram protocol. T2 and T1-based markers of myocardial inflammation were analyzed in each of the 16 segments of the 17-segment model of the American Heart Association (the true apex was excluded), where only positive segment concordances from the different T2 and T1-based markers were considered. Because myocarditis was diagnosed according to these T2 and T1-based CMR markers and an adequate selection of normal reference values is fundamental, we used a population-based control CMR imaging group from 50 sex- and aged-matched individuals without cardiac disease. The prevalences of cardiovascular risk factors (hypertension, diabetes mellitus, dyslipemia, current smoking) in the control cohort were similar to those in the study population.

Immunophenotypic analysis of (> 250) immune cell populations was performed in peripheral blood samples collected in K3-ethylendiaminetracetate-acetic acid (EDTA, 10 mL/sample, and stained with the EuroFlow lymphocyte screening tube and the cluster of differentiation 4 T cell (TCD4), natural killer (NK)/TCD8, beta-lactoglobulin hydrolyses (BigHy) and monocyte-derived dendritic cell (MoDC) immune monitoring tubes by flow cytometry (FACSCANTO II and LSR-Fortessa, respectively; Becton/Dickinson Biosciences, United States) using a dual-platform assay previously described in detail. Reference values for the individual immune cell subsets investigated in blood by flow cytometry were defined based on a population-based control group of 463 age-matched adults (median age 52 [IQR 47–61] years) studied prior to the SARS-CoV-2 pandemic. Anti-SARS-CoV-2-IgM (AnshLabs, Webster, United States), IgG and IgA (Mikrogen Diagnostik, Neuried, Germany) antibody levels were measured in parallel in plasma from the same blood samples using commercially available in vitro diagnostic medical device approved (semi quantitative) ELISA kits, strictly as instructed by the manufacturers.

Study outcomes and definitions

Study outcome measures were the prevalence of clinical pericarditis and of myocarditis, and the characterization of the immune cell compartments in blood, and the virus-specific humoral immune response. Clinically suspected pericarditis was diagnosed if at least 2 of the following criteria were present, following current guidelines: pericarditic chest pain, pericardial rub on auscultation, widespread ST-elevation or PR segment depression on electrocardiogram, and evidence of pericardial effusion on CMR. Elevation of inflammation markers, C-reactive protein, and evidence of pericardial inflammation on CMR were used as additional supporting findings. The diagnosis of clinically suspected myocarditis was based on CMR criteria11; we considered as main CMR criteria positive edema-sensitive T2-based markers (T2-weighted images or T2-mapping) or positive T1-based tissue characterization markers (abnormal T1-relaxation time or extra-cellular volume or late gadolinium enhancement), and as supportive CMR criteria either pericardial effusion, or evidence of pericardial inflammation on CMR, or systolic left ventricular wall motion abnormalities. Considering that participants were being examined after the acute phase of SARS-CoV-2 infection, in this study clinically suspected myocarditis was defined as the presence of a combination of at least 2 T2 or T1-based CMR main criteria or the presence of combination of only 1 T2 or T1-based main criterion with 1 additional CMR supportive criterion.

As we were aware that pericarditis and myocarditis occur together in clinical practice, we therefore defined as clinically suspected myopericarditis those cases of pericarditis with associated myocarditis on CMR but without left ventricular wall motion abnormalities, and as clinically suspected perimyocarditis those cases where left ventricular wall motion abnormalities were present.

Statistical analysis

Descriptive statistics were used to summarized the data. Results are presented as the proportion (%) of valid cases for categorical variables and as the median [IQR] for continuous variables. Differences between groups were analyzed by the Fisher exact test for categorical variables and by the nonparametric Mann-Whitney or Kruskal-Wallis tests for continuous data. Comparisons between immune cell counts in the blood of patients and controls were adjusted for age and sex (covariates) using the 1-way ANCOVA (analysis of covariance) univariate general linear modeling test (SPSS statistical software package v25.0, IBM, Armonk, United States). We compared characteristics of participants and examinations, all tables, according to the final clinical diagnosis (nonpericardial and myocardial manifestations vs pericarditis vs myopericarditis vs myocarditis). For 2-dimensional visualization of flow cytometry data, multivariate canonical analysis with multidimensional reduction of data via linear discriminant analysis, and the t-distributed stochastic neighbor embedding machine-learning algorithm visualization tools were used (Infinicyt software, Cytognos, Universidad de Salamanca).13

RESULTS

Study population

Figure 1 depicts the flowchart for participant selection among healthcare workers. Of the 142 recruited healthcare workers who signed informed consent, 1 participant did not complete the CMR due to claustrophobia. Two additional participants were excluded due to a history of severe hypertrophic myocardiopathy in 1 case,
and inherited immune deficiency in the other. Thus, a total of 139 participants completed the clinical assessment, electrocardiography, laboratory tests and CMR. Of these, 103 (74.1%) had been diagnosed by RT-PCR and 36 (25.9%) by serology. No participants were clinically diagnosed with post-COVID-19 cardiac involvement at the time of their index presentation, nor were those who were previously hospitalized.

All participant characteristics are shown in table 1. By professional categories, 49 (35.3%) were nurses, 35 (25.2%) physicians, and the remaining 55 (39.6%) included different profiles such as auxiliary nurses and other hospital staff. A total of 67 (48.2%) healthcare workers were infected while directly attending COVID-19 hospitalization wards.

Among the overall study population, 106 (76.3%) had at least 1 comorbidity and 8 (5.8%) healthcare workers had a history of cardiovascular disease: 1 with chronic ischemia with stent revascularization, 3 with paroxysmal atrial fibrillation, 2 with intranodal supraventricular tachycardias treated with ablation, and 2 with an episode of acute pericarditis several years previously.

Most (137 [98.6%]) healthcare workers experienced a viral prodrome during SARS-CoV-2 infection and cardiac symptoms with shortness of breath, chest pain, palpitations or dizziness were reported by 86 (61.9%) participants. A total of 27 (19.4%) healthcare workers were previously diagnosed with COVID-19 pneumonia and 23 (16.5%) required hospitalization (none of these were diagnosed with pericarditis or myocarditis during this hospitalization index).

Chronic drug therapy and treatment during SARS-CoV-2 infection are shown in table 1 of the supplementary data. Overall, the drug therapy aimed at ameliorating the disease was heterogeneous: hydroxychloroquine was given in 33 (23.7%) participants, lopinavir-ritonavir in 17 (12.2%), oral glucocorticoids in 9 (6.5%), high-dose intravenous bolus of methylprednisolone in 15 (10.8%), and interleukin inhibitors in 18 (12.9%).

Symptoms, electrocardiographic, biochemical, and cardiac magnetic resonance profiling

The study examinations (table 2) were performed 10.4 (9.3–11.0) weeks after the start of symptoms of infection. All participants had vital and exploratory signs of hemodynamic stability on examination. A total of 91 (65.5%) healthcare workers still had symptoms, which were cardiac-related in 58 (41.7%).

Of the 139 electrocardiograms, electrocardiographic abnormalities were reported in 69 (49.6%) cases (table 2 of the supplementary data). A total of 33 (23.2%) electrocardiograms met the criteria for pericarditis-like changes (figure 1 of the supplementary data). As the Occupational Health Service of the hospital provide healthcare workers a baseline medical evaluation before starting their jobs, we were able to recover and review 53 (76.8%) prior baseline electrocardiograms from the 69 patients with electrocardiographic changes at the study examination. Of these 53 electrocardiographic comparisons, 67.9% electrocardiographic changes at the study examination were not present previously and consisted mainly of pericardial like changes and ST-segment depression or T-wave inversion (table 3 of the supplementary data).

Cardiac-specific and inflammatory biomarkers were within the normal range in most participants (table 2). CMR abnormalities were observed in 84 (60.4%) participants (table 3 and tables 4 and 5 of the supplementary data, figure 2). Two (1.4%) participants showed increased myocardial T2-relaxation time, 5 (3.6%) edema on T2-weighted images, 40 (28.8%) increased native myocardial T1-relaxation time, 27 (19.4%) increased T1-extracellular volume, 10 (7.2%) T1-late gadolinium enhancement, 42 (30.2%) pericardial effusion, 1 (0.7%) a pericardial thickness of 3 mm and 7 (5.0%) systolic left ventricular wall motion abnormalities, global or regional.

Clinically suspected pericarditis and myocarditis prevalence

A total of 43 (30.9%) participants fulfilled the criteria for either clinically suspected pericarditis or myocarditis. Clinically suspected isolated pericarditis was diagnosed in 8 (5.8%) participants, isolated myocarditis in 24 (17.3%), and myopericarditis in 11 (7.9%). These were no cases of perimyocarditis. Descriptions of criteria combinations are provided in figure 3 and baseline and examination characteristics for each diagnostic group are detailed in table 1 and table 2.
Table 1
Baseline characteristics of the study cohort

|                      | All participants (n = 139) | Presence of pericardial and myocardial manifestations | P       |
|----------------------|----------------------------|------------------------------------------------------|---------|
|                      | No (n = 96)                | Pericarditis (n = 8)                               | Myopericarditis (n = 11) | Myocarditis (n = 24) |
| Age, y               | 52 [41-57]                 | 52 [39-58]                                          | 50 [34-56]                  | 53 [49-58]          | .704                |
| Female sex           | 100 (71.9)                 | 64 (66.7)                                           | 7 (87.5)                      | 8 (72.7)             | 21 (87.5)           | .164                |
| Coexisting conditions|                           |                                                     |                                  |                      |                     |
| Obesitya             | 17 (12.2)                  | 14 (14.6)                                           | 1 (12.5)                        | 0                    | 2 (8.3)              | .620                |
| Hypertension         | 17 (12.2)                  | 12 (12.5)                                           | 1 (12.5)                       | 1 (9.1)              | 3 (12.5)             | 1.000               |
| Diabetes             | 2 (1.4)                    | 2 (2.1)                                             | 0                                | 0                    | 0                    | 1.000               |
| Dyslipidemia         | 27 (19.4)                  | 20 (20.8)                                           | 1 (12.5)                       | 2 (18.2)             | 4 (16.7)             | .977                |
| Current smoking      | 6 (4.3)                    | 5 (5.2)                                             | 0                                | 1 (9.1)              | 0                    | .534                |
| Past smoking         | 70 (50.4)                  | 52 (54.2)                                           | 2 (25.0)                       | 4 (36.4)             | 12 (50.0)            | .333                |
| Alcohol useb         | 23 (16.5)                  | 12 (12.5)                                           | 2 (25.0)                       | 2 (18.2)             | 7 (29.2)             | .140                |
| Cardiovascular disease| 8 (5.8)                  | 5 (5.2)                                             | 0                                | 2 (18.2)             | 1 (4.2)              | .297                |
| Pulmonary diseasc    | 8 (5.8)                    | 6 (6.3)                                             | 0                                | 0                    | 2 (8.3)              | .898                |
| Sleep apnea-hypopnea syndrome | 8 (5.8) | 6 (6.3)                                             | 1 (12.5)                       | 1 (9.1)              | 0                    | .326                |
| Chronic kidney disease | 5 (3.6)                   | 2 (2.1)                                             | 0                                | 0                    | 3 (12.5)             | .118                |
| Cancer               | 4 (2.9)                    | 3 (3.1)                                             | 0                                | 0                    | 1 (4.2)              | 1.000               |
| At least 1 of the above | 106 (76.3)               | 75 (78.1)                                           | 5 (62.5)                       | 7 (63.6)             | 19 (79.2)            | .493                |

SARS-CoV-2 infection diagnosis

RT-PCR 103 (74.1) 67 (69.8) 7 (87.5) 10 (90.9) 19 (79.2) 0.395

Anti-SARS-CoV-2-IgG antibodies 36 (25.9) 29 (30.2) 1 (12.5) 1 (9.1) 5 (20.8) 0.000

Symptoms during SARS-CoV-2 infection

General

Fatigue 117 (84.2) 79 (82.3) 6 (75.0) 11 (100) 21 (87.5) .404
Fever 94 (67.6) 65 (67.7) 7 (87.5) 9 (81.8) 13 (54.2) .269
Cough 91 (65.5) 64 (66.7) 6 (75.0) 7 (63.6) 14 (58.3) .849
Headache 90 (64.7) 60 (62.5) 4 (50.0) 9 (81.8) 17 (70.8) .499
Myalgia 83 (59.7) 55 (57.3) 6 (75.0) 7 (63.6) 15 (62.5) .810
Anosmia 73 (52.5) 48 (50.0) 6 (75.0) 6 (54.5) 13 (54.2) .617
Ageusia 66 (47.5) 40 (41.7) 5 (62.5) 5 (45.5) 16 (66.7) .133
Abdominal pain or diarrhea 64 (46.0) 44 (45.8) 1 (12.5) 7 (63.6) 12 (50.0) .161
Chills 60 (43.2) 40 (41.7) 3 (37.5) 6 (54.5) 11 (45.8) .846
Score throat 55 (39.6) 37 (38.5) 3 (37.5) 4 (36.4) 11 (45.8) .940
Nausea or vomiting 30 (21.6) 17 (17.7) 2 (25.0) 5 (45.5) 6 (25.0) .154
Clumsiness 20 (14.4) 15 (15.6) 1 (12.5) 2 (18.2) 2 (8.3) .856
Memory loss 19 (13.7) 13 (13.5) 0 (0.0) 2 (18.2) 4 (16.7) .733
Skin lesions 5 (3.6) 4 (4.2) 0 (0.0) 0 (0.0) 1 (4.2) 1.000

Cardiac

Shortness of breath 68 (48.9) 42 (43.8) 5 (62.5) 9 (81.8) 12 (50.0) .090
Palpitations 44 (31.7) 25 (26.0) 5 (62.5) 7 (63.6) 7 (29.2) .017
 Chest pain 40 (28.8) 24 (25.0) 3 (37.5) 4 (36.4) 9 (37.5) .455
Dizziness 4 (2.9) 0 (0.0) 1 (12.5) 2 (18.2) 1 (4.2) .003

At least 1 cardiac symptom 86 (61.9) 54 (56.3) 6 (75.0) 10 (90.9) 16 (66.7) .114

IgG, immunoglobulin G; RT-PCR, reverse-transcriptase-polymerase-chain-reaction.

Data are expressed as No. (%) or median [interquartile range].

a Obesity was considered if body-mass index was 30 or more.
b Alcohol use was considered as an average of at least 1 drink a day.
c All participants with previous pulmonary disease reported asthma. P value for comparison among the 4 participants groups.

A higher percentage of participants with clinically suspected pericarditis, myopericarditis or myocarditis had cardiac symptoms during SARS-CoV-2 infection (32 [74.4%] vs 54 [56.3%]; P = .058) and on study examination (26 [60.5%] vs 32 [33.3%]; P = .005) than participants without these manifestations. Chronic drug therapy with statins was more frequent in participants without pericardial or myocardial manifestations than in those with clinically suspected pericarditis, myopericarditis or myocarditis (16 [16.7%] vs 1 [2.3%]; P = .022).

Participants with infection confirmed through anti-SARS-CoV-2-IgG detection

Among the 36 participants diagnosed with past infection through anti-SARS-CoV-2-IgG detection (data for this group, compared with RT-PCR participants, are shown in table 6 of the supplementary data), 34 (94.4%) reported at least 1 of the 18 collected symptoms of COVID-19 infection (ie, fever, persistent cough and/or anosmia, whose presence, according
to current guidelines would require isolation and testing was present in 27 (75.0%) of these 36 participants; 28 (77.8%) had previously tested negative by RT-PCR after developing SARS-CoV-2 symptoms and 8 (22.2%) were never RT-PCR tested. A lower percentage of participants diagnosed through positive serology still had symptoms on examination compared with RT-PCR participants (18 [50.0%] vs 73 [70.9%]; \(P = .027\)); nonetheless, the prevalence of clinically suspected pericarditis, myopericarditis or myocarditis was high in both groups (figure 4).

### Altered immune cell and humoral profiles in blood

Most study participants (101 [73.2%]) displayed altered cell counts in blood for at least 1 major immune cell population, as
| Table 3 | Cardiac magnetic resonance measures |
|---|---|
| | Population-based controls (n = 50) | All participants (N = 139) | | Presence of pericardial and myocardial manifestations | |
| | | | | No (n = 96) | Pericarditis (n = 8) | Myopericarditis (n = 11) | Myocarditis (n = 24) | |
| **Age, y** | 51 [42-56] | 51 [42-58] | .335 | 52 [39-58] | 51 [34-56] | 50 [44-61] | 53 [49-58] | .704 |
| **Female sex** | 35 (70.0) | 100 (71.9) | .856 | 64 (66.7) | 49 [34-61] | 51 (54-61) | 53 (49-58) | .164 |
| **Height, cm** | 166 [160-172] | 165 [160-173] | .844 | 166 [161-174] | 164 [162-169] | 167 [162-169] | 162 [158-170] | .300 |
| **Weight, kg** | 65 [59-75] | 69 [58-80] | .506 | 72 [60-83] | 58 [54-75] | 60 [56-71] | 59 [51-74] | .003 |
| **Body surface area, m²** | 1.7 [1.6-1.9] | 1.8 [1.6-1.9] | .555 | 1.8 [1.7-2.0] | 1.6 [1.6-1.8] | 1.6 [1.6-1.8] | 1.6 [1.6-1.8] | .044 |
| **Haematocrit, %** | 42 [40-43] | 39 [37-41] | < .001 | 40 [38-42] | 39 [36-41] | 41 [36-41] | 37 [35-39] | .002 |
| **Left ventricle** | | | | | | | | |
| End-diastolic indexed volume, mL/m² | 74 [58-82] | 72 [67-83] | .483 | 72 [65-84] | 73 [60-89] | 77 [71-96] | 72 [69-79] | .322 |
| End-systolic indexed volume, mL/m² | 23 [20-31] | 26 [23-31] | .079 | 27 [22-31] | 27 [20-34] | 26 [25-40] | 26 [23-30] | .802 |
| Indexed left ventricle mass, g/m² | 48 [43-60] | 51 [46-60] | .071 | 53 [46-62] | 49 [41-58] | 50 [43-59] | 50 [47-55] | .488 |
| Left ventricular ejection fraction, % | 66 [61-70] | 64 [60-67] | .019 | 63 [59-67] | 64 [60-67] | 65 [61-66] | 65 [61-67] | .782 |
| Wall regional motion abnormalities | 0 | 7 [5.0] | .193 | 4 [4.2] | 0 | 0 | 3 [12.5] | .291 |
| Global myocardial T₂-relaxation time, msec | 52 [48-54] | 50 [48-51] | < .001 | 50 [48-51] | 49 [45-51] | 50 [48-53] | 50 [50-52] | .086 |
| Global native myocardial T₁-relaxation time, msec | 1024 [991-1040] | 1021 [1005-1040] | .615 | 1020 [1002-1039] | 1028 [997-1046] | 1027 [1024-1045] | 1030 [1008-1052] | .411 |
| Global T₁-extracellular volume, % | 25 [22-30] | 26 [24-28] | .355 | 25 [23-27] | 27 [26-27] | 28 [25-30] | 28 [26-30] | < .001 |
| Left ventricle global longitudinal strain | −23 [−26 to −21] | −22 [−24 to −20] | .004 | −21 [−24 to −20] | −24 [−25 to −18] | −22 [−24 to −21] | −22 [−25 to −20] | < .001 |
| Left ventricle global circumferential strain | −33 [−36 to −28] | −32 [−32 to −26] | < .001 | −28 [−32 to −25] | −29 [−31 to −24] | −31 [−32 to −25] | −29 [−32 to −26] | .952 |
| Left ventricle global radial strain | 73 [60-88] | 59 [45-70] | < .001 | 58 [44-68] | 60 [43-70] | 60 [51-71] | 60 [49-76] | .676 |
| Right ventricle | | | | | | | | |
| End-diastolic indexed volume, mL/m² | 71 [61-80] | 74 [67-85] | .094 | 74 [66-84] | 78 [62-92] | 78 [69-90] | 76 [69-80] | .449 |
| End-systolic indexed volume, mL/m² | 26 [18-30] | 27 [22-34] | .032 | 27 [22-33] | 30 [18-40] | 27 [23-36] | 29 [22-34] | .909 |
| Right ventricular ejection fraction, % | 66 [60-69] | 63 [59-68] | .095 | 63 [58-67] | 61 [56-72] | 66 [60-69] | 63 [60-66] | .839 |
| Free wall right ventricle global longitudinal strain | −28 [−32 to −24] | −25 [−27 to −21] | < .001 | −25 [−27 to −21] | −25 [−25 to −19] | −25 [−26 to −20] | −26 [−28 to −22] | .214 |
| Atrium | | | | | | | | |
| Left atrium indexed area, cm²/m² | 11 [10-13] | 11 [10-13] | .947 | 11 [10-12] | 13 [11-13] | 13 [12-14] | 12 [10-14] | .067 |
| Left atrium emptying fraction, % | 58 [53-66] | 60 [54-66] | .540 | 61 [56-66] | 60 [37-68] | 58 [52-69] | 60 [54-66] | .859 |
| Left atrium global longitudinal strain | 41 [34-50] | 41 [35-52] | .522 | 41 [35-53] | 43 [28-55] | 40 [33-56] | 43 [34-51] | .983 |
| Right atrium indexed area, cm²/m² | 10 [9-12] | 10 [9-12] | .407 | 10 [9-11] | 11 [9-14] | 11 [10-13] | 10 [9-12] | .087 |
| **Main CMR Lake-Louise criteria for myocarditis** | | | | | | | | |
| Increase in myocardial T₂-relaxation time | 0 | 2 [1.4] | .540 | 0 | 0 | 0 | 2 [8.3] | .094 |
| T₂-weighted hyperintensity | 0 | 5 [3.6] | .211 | 0 | 0 | 1 [9.1] | 4 [16.7] | .002 |
| Increase in native myocardial T₁-relaxation time | 0 | 40 [28.8] | < .001 | 19 [19.8] | 0 | 5 [45.5] | 16 [66.7] | < .001 |
| Increase in T₁-extracellular volume | 0 | 27 [19.4] | < .001 | 8 [8.3] | 0 | 6 [54.5] | 13 [54.2] | < .001 |
| T₁-late gadolinium enhancement | 0 | 10 [7.2] | .042 | 2 [2.1] | 0 | 4 [36.4] | 4 [16.7] | < .001 |
| At least 1 main criterion | 0 | 63 [45.3] | < .001 | 28 [29.2] | 0 | 11 [100] | 24 [100] | < .001 |
| **Supportive CMR Lake-Louise criteria for myocarditis** | | | | | | | | |
| Pericardial effusion | 0 | 42 [30.2] | < .001 | 11 [11.5] | 7 [8.75] | 11 [100] | 13 [54.2] | < .001 |

R. Eiros et al. / Rev Esp Cardiol. 2022;75(9):734-746
The data are expressed as No. (%) or median [interquartile range].

a Pericardial effusion greater than 3 mm.

b Pericardial thickness of 3 mm on T2-weighted spin-echo CMR.

c P value for comparisons between population-based controls vs all participants.

d Right value for comparison among the 4 participants groups.

DISCUSSION

This study examined the prevalence of clinically suspected pericarditis and myocarditis in a cohort of SARS-CoV-2–positive healthcare workers. In one of the largest cohort of participants with CMR imaging assessment reported so far, we demonstrate that pericardial and myocardial involvement is prevalent after SARS-CoV-2 infection in association with an altered immune response.

We decided to carry out a study in healthcare workers as this sector has been disproportionally infected in Spain, which provided us with the opportunity to study the prevalence of clinically suspected pericarditis and myocarditis in SARS-CoV-2-infected cases that were confirmed by positive RT-PCR or positive serology. In addition, because the proportion of female healthcare workers is high in Spain, our study does not underrepresent women who constituted more than two thirds of recruited participants. Unlike other observational studies suggesting that myocarditis may be slightly more prevalent in men than in women,14 men in our study had a lower prevalence of pericardial and myocardial involvement than women (7 [17.9%] vs 36 [36.0%]; P = .043). Recent findings suggest that the immune landscape in COVID-19 differs considerably between the sexes,15 with female patients having a more robust activation of T cells than men. Considering that pericarditis and myocarditis both stem from a T and B cell–mediated mechanism, our observation is of interest and provides new evidence indicating sex differences in the clinical outcomes of COVID-19 disease that should be corroborated in future studies.
Previous studies have reported the prevalence of myocardial injury after COVID-19. An initial retrospective observation in 26 recovered patients with COVID-19 pneumonia showed cardiac complaints during hospitalization revealed the presence of myocardial edema in 54% of the patients and late gadolinium enhancement in 31%. Observations similar to a prospective study in 100 patients recovered from COVID-19 pneumonia were the presence of myocardial edema in 60% and late gadolinium enhancement in 32%. Our observations, performed mostly in nonhospitalized participants (83.5%) and also including participants diagnosed through serology (25.9%), showed lower percentages of CMR myocardial injury (specially myocardial edema) than the aforementioned studies and are more in agreement with the rates of myocardial injury observed in influenza—in which elevated cardiac enzymes, electrocardiographic, echocardiographic and histologic findings have been reported in approximately one third of cases. Two recent pathological studies in postmortem patients with COVID-19 pneumoniae have shown lymphocytic myocarditis in 14.3% and 26.7% of the cases, which is more in line with our finding (17.3% for isolated myocarditis and 25.2% for any...
**Table 4**
Distribution of subsets of myeloid and lymphoid immune cells in blood

|                          | Healthy donors (n=463) | All participants (N=139) |   | Pericardial and myocardial manifestations |   |
|--------------------------|------------------------|--------------------------|---|------------------------------------------|---|
|                          |                        |                          |   | Presence of pericardial and myocardial  |   |
|                          |                        |                          |   | manifestations                            |   |
|                          |                        |                          |   | Healthy donors vs all participants |   |
|                          |                        |                          |   | Left P value                               |   |
|                          |                        |                          |   | Right P value                              |   |
|                          |                        |                          |   | Comparisons between healthy donors vs    |   |
|                          |                        |                          |   | all participants                          |   |
|                          |                        |                          |   | Comparisons among the 4 participants      |   |
|                          |                        |                          |   | groups                                    |   |
|                          |                        |                          |   | Comparisons between healthy donors vs    |   |
|                          |                        |                          |   | all participants are adjusted for age and  |   |
|                          |                        |                          |   | sex.                                      |   |
| Neutrophils               | 3723 [2969-4613]       | 3430 [2633-4225]         | .010 | 3468 [2647-4223]                          | .491 |
|                          | 18 (12.9)/5 (3.6)      | < .001/.001              | 10 (10.4)/4 (4.2) | 1 (12.5)/0                             | 7 (29.2)/1 (4.2) | .051/.844 |
| Eosinophils               | 157 [101-249]          | 74 [47-149]              | < .001 | 82 [50-130]                          | 48 [31-189]                          | 59 [46-69]                          | 102 [39-191]                          | .463 |
|                          | 38 (27.3)/1 (0.7)      | < .001/.231              | 24 (25.0)/1 (1.0) | 4 (50.0)/0                             | 3 (27.3)/0                             | 7 (29.2)/0                             | .499/.929 |
| Basophils                 | 38 [21-52]             | 47 [35-62]              | .007 | 49 [37-62]                          | 34 [23-55]                          | 36 [29-56]                          | 51 [38-65]                          | .293 |
|                          | 4 (2.9)/5 (3.6)        | < .001/.001              | 2 (2.1)/3 (3.2) | 1 (12.5)/1                             | 1 (9.1)/1                             | 0 (1/4.2)                             | .169/.719 |
| Monocytes                 | 317 [245-433]          | 405 [328-523]            | < .001 | 418 [345-533]                          | 369 [307-424]                          | 404 [347-476]                          | 358 [282-574]                          | .724 |
|                          | 3 (2.2)/8 (5.8)        | < .012/.001              | 3 (3.1)/5 (5.2) | 0 (1/12.5)                             | 0 (8)/0                                | 0 (8/2)                               | .712/.639 |
| Dendritic cells           | 29 [20-35]             | 28 [20-35]              | .561 | 28 [20-35]                          | 28 [20-33]                          | 29 [18-42]                          | 23 [19-35]                          | .920 |
|                          | 14 (10.2)/5 (3.6)      | < .001/.001              | 11 (1.7)/5 (5.3) | 0 (8)/0                                | 1 (9.1)/0                             | 2 (8)/0                                | .755/.508 |
| Lymphocytes               | 1675 [1332-2223]       | 2221 [1717-2563]         | < .001 | 2294 [1854-2760]                          | 1990 [1555-2422]                          | 1660 [1379-2089]                          | 1998 [1619-2439]                          | .055 |
|                          | 3 (2)/8 (6)            | < .012/.001              | 1 (1.0)/6 (3.3) | 1 (12.5)/1                             | 1 (9.1)/0                             | 0 (1/4.2)                             | .054/.683 |
| NK cells                  | 260 [162-372]          | 213 [137-321]            | .427 | 217 [143-337]                          | 233 [142-281]                          | 120 [76-321]                          | 156 [137-248]                          | .240 |
|                          | 9 (6.5)/6 (4.3)        | < .001/.001              | 5 (5.2)/5 (5.2) | 0 (8)/0                                | 3 (27.3)/0                             | 1 (4/21)                              | .003/.791 |
| B cells                   | 154 [108-228]          | 208 [158-297]            | < .001 | 234 [175-327]                          | 233 [119-443]                          | 174 [153-188]                          | 181 [136-230]                          | .020 |
|                          | 2 (1.4)/16 (11.5)      | < .053/.001              | 1 (1.0)/10 (12.5) | 1 (12.5)/2                             | 0 (0)/0                                | 0 (2)/8                                | .525/.362 |
| Plasma cells              | 2 [0.8-3]              | 0.8 [0.3-2]              | < .001/0.9 | 0.9 [0.5-2]                          | 1 [0.2-1]                              | 0.4 [0.3-0.8]                          | 1 [0.3-2]                              | .362 |
|                          | 11 (7.9)/1 (0.7)       | < .001/.231              | 7 (7.3)/1 (1.0) | 0 (8)/0                                | 2 (18.2)/0                             | 2 (8)/0                                | .506/.929 |

NK, natural killer.

Values are expressed as the median [interquartile range] of cells/μL of blood or No. (%) of cases lower than 5th percentile (\(5^{\text{th}}\)p) and higher than the 95th percentile (\(95^{\text{th}}\)p) for each cell subset in age-matched healthy donors. Left P value for comparisons between healthy donors vs all participants. Right P value for comparison among the 4 participants groups. Comparisons between healthy donors vs all participants are adjusted for age and sex.
degree of myocardial involvement on CMR. In addition, pericardial injury after COVID-19 is prevalent; a recent study in 54 consecutive student athletes with positive RT-PCR or serology to SARS-CoV-2 has shown pericardial inflammation with associated pericardial effusion in 39.5% of cases, which is similar to the 30.2% pericardial effusion observed in our study. Of note, athletes had increased native T1-relaxation time values in 19.0% individuals but native T2 findings were normal in all participants.

Importantly, clinical assessment of our participants with suspected pericarditis and myocarditis showed clinical stability without any participants showing severe pericardial effusion, heart failure, or left ventricular dysfunction (only 3 participants with myocarditis showed wall motion abnormalities). However, follow-up studies are necessary to determine the outcome of cardiac sequelae observed even in asymptomatic and pauci-symptomatic participants after SARS-CoV-2 infection. The participants diagnosed with past infection through serology who were more likely to be asymptomatic or mildly symptomatic, and who might better represent the cases detected in population-wide seroprevalence studies, also showed a high
prevalence of pericardial and myocardial manifestations although the prevalence was lower than in RT-PCR positive participants (7 [19.4%] vs 36 [35.0%]; P = .097).

At present, there is much interest in the long-term sequelae of COVID-19. It is intriguing that clinically suspected pericardial and myocardial manifestations were observed long after SARS-CoV-2 infection (more than 10 weeks after initial viral prodrome at infection) as well as in some currently asymptomatic participants (9 cases; 1 out of every 5 clinically suspected cases of pericarditis, myopericarditis or myocarditis diagnoses). These long-term manifestations may be due to an inadequate innate and adaptive immune response with very limited data on the longer-term immunological consequences of past SARS-CoV-2 infection, and no study has specifically focused on the settings of pericarditis and clinically suspected myocarditis. In this study, in-depth investigation of the distribution of major and minor populations of immune cells in blood showed a high frequency of overall altered immune profiles.

Several of the immune cell alterations identified mimic abnormalities reported during active infection for the general population with COVID-19, including decreased eosinophil and NK cell counts. The overall pattern in this study emerges as a unique SARS-CoV-2-associated immune profile. For example, while decreased eosinophil counts in blood have been reported among participants infected with influenza, no association has yet been reported with increased counts of cytotoxic (CD4 CD8 T) or plasmablasts in blood, which have been identified among HIV-infected participants in the absence of esoinopenia.

More detailed analysis of the altered immune profiles among the different groups of participants showed that those with clinically suspected myopericarditis or myocarditis had closer to normal lymphocyte counts, but reduced numbers in blood of circulating eosinophils and NK cells. Such a unique profile mimics what has been described recently during the acute phase of SARS-CoV-2 infection, suggesting an ongoing cytotoxic response with increased tissue migration or death by apoptosis of specific subsets of cytotoxic cells. These findings suggest that a less pronounced (potentially insufficient) or a delayed humoral response may occur in these participants, which may lead to decreased neutralization, opsonization and/or clearance of the virus locally at the perimyocardium; local viral persistence would favor an increased tissue-homing (or early death) of eosinophils, immunomodulatory and intermediate monocytes, in addition to cytotoxic (effector) cells. Thus, similar to influenza, although SARS-CoV-2 pneumoniae is the most widely recognized complication, the coronavirus could trigger pericarditis or myocarditis as part of the host immune response rather than viral-mediated myocarditis per se. In this sense, cases of cardiac involvement are beginning to be described after the second dose of COVID-19 vaccine.

CONCLUSIONS

This study shows that clinically suspected pericarditis and myocarditis are frequent in healthcare workers after SARS-CoV-2 infection, as well as in some currently asymptomatic individuals; in addition, we provide evidence for an altered immune cell distribution in blood which affects cells involved in both the innate (eg, eosinophils, monocytes and NK cells) and the adaptive cellular (eg, cytotoxic T cells) and humoral (eg, B cells and plasma cells) immune responses. Pericardial and myocardial involvement after SARS-CoV-2 infection might then indirectly result from this host immune response. These observations may indicate that cardiac sequelae might occur late after SARS-CoV-2 infection.

FUNDING

This study was supported by CIBERCV (CB16/11/00374), CIBERONC (CB16/12/00400) and the COV20/00386 grant from the Instituto de Salud Carlos III and FEDER, Ministerio de Ciencia e Innovación, Madrid, Spain, and by GRS COVID 26/A/20 from the Gerencia Regional de Salud, Junta de Castilla y León, Spain.

AUTHORS’ CONTRIBUTIONS

R. Eiros and M. Barreiro-Pérez contributed equally to this work and share first authorship.

A. Orfao and P.L. Sánchez contributed equally to this work and share last authorship.

R. Eiros, M. Barreiro-Pérez, J. Almeida, J. L. Bravo-Grande, A. Orfao, P. L. Sánchez conceived and designed the study; R. Eiros, M. Barreiro-Pérez, A. Orfao, P. L. Sánchez contributed to the literature search; R. Eiros, S. Merchán, I. Toranzo contributed with clinical evaluation of the participants; S. Merchán, C. Sánchez-Pablo, I. Toranzo, E. Díaz-Peláez, L. Macías-Alvarez contributed with clinical data collection; M. Barreiro-Pérez, A. Martín-García contributed to data analysis (CMR); E. Villacorta, D. González-Calle contributed to data analysis (ECG); A. Pérez-Pons, A. Torres-Valle, O. Pérez-Escurza, B. Fuentes-Herrero performed immunophenotypic and serology experiments and collected data; J. Almeida, B. Fuentes-Herrero, Gu. Oliva-Arizá, Q. Lecrevissé, R. Fluxa, A. Orfao contributed to data analysis and interpretation (immunophenotypic and serology); R. Eiros, M. Barreiro-Pérez, A. Martín-García, E. Villacorta, A. Orfao, P. L. Sánchez contributed to clinical data analysis and interpretation; M. Barreiro-Pérez, J. Almeida, J. L. Bravo-Grande, A. Orfao, P. L. Sánchez contributed to writing of the report.

CONFLICTS OF INTEREST

A. Orfao and J. Almeida report being one of the inventors on the EuroFlow-owned European patent 119646NL00 registered on November 2019 (“Means and methods for multiparameter flow cytometry based leukocyte subsetting”) and A. Orfao and J. Almeida are also authors of the PCT patent WO 2010/140885A1 (“Methods, reagents and kits for flow cytometric immunophenotyping”). The Infinicyt software is based on intellectual property of the University of Salamanca in Spain. All above mentioned intellectual property and related patents are licensed to Cytognos (Salamanca, ES) and Becton/Dickinson Biosciences (San José, California), and these companies pay royalties to the EuroFlow Consortium. These royalties are exclusively used for continuation of the EuroFlow collaboration and sustainability of

Limitations

The major limitation of this study is that clinically suspected myocarditis was not confirmed via endomyocardial biopsy. CMR T1 and T2 measures, although significant, were small between the participants with SARS-CoV-2 infection and the control group. The study analysis was limited to healthcare workers and therefore has limited external generalizability to other nonhealthcare settings. However, the strength of this study is the addition of nonhospitalized participants, as well as the inclusion of participants diagnosed with past SARS-CoV-2 infection through serology, who also had a high prevalence of pericardial and myocardial involvement.
the EuroFlow Consortium. There are no other conflicts of interest related to this study or for the remaining authors.

WHAT IS KNOWN ABOUT THE TOPIC?
- There is increasing evidence of cardiac sequelae after SARS-CoV-2 infection.
- Although pericarditis and myocarditis are the 2 most frequent cardiac manifestations observed after a viral infection, peer-reviewed literature is limited to isolated case reports of pericarditis and small observations of myocarditis mainly in hospitalized RT-PCR patients with COVID-19 pneumonia, which are insufficient to generalize conclusions about the true prevalence of pericardial and myocardial involvement after SARS-CoV-2 infection.
- No study has investigated the immunological consequences of SARS-CoV-2 infection in the settings of pericarditis and myocarditis.

WHAT DOES THIS STUDY ADD?
- The prevalence of pericarditis and myocarditis, with clinical stability, after SARS-CoV-2 infection is high; up to 31% of cases and also in some presently asymptomatic participants (1 out of every 5 cases diagnosed), in association with notably altered immune cell profiles in blood.
- Female participants were more likely to have pericarditis or myocarditis.
- Clinical pericarditis and myocarditis are associated with specific immune cell profiles, paving the way for a better understanding of the immune mechanisms involved. In this regard, pericardial and myocardial involvement is beginning to be described after COVID-19 vaccination.

APPENDIX. SUPPLEMENTARY DATA
Supplementary data associated with this article can be found in the online version available at https://doi.org/10.1016/j.rec.2021.11.001

REFERENCES
1. Boukhris M, Hillani A, Moroni F, et al. Cardiovascular Implications of the COVID-19 Pandemic: A Global Perspective. Can J Cardiol. 2020;36:1068–1080.
2. Blanco-Melo D, Nilsson-Payant BE, Liu WC, et al. Imbalanced Host Response to SARS-CoV-2 Drives Development of COVID-19. Cell. 2020;181:1036–1045.e9.
3. Adler Y, Charron P, Inazio M, et al. 2015 ESC Guidelines for the diagnosis and management of pericardial diseases: The Task Force for the Diagnosis and Management of Pericardial Diseases of the European Society of Cardiology (ESC) Endorsed by: The European Association for Cardio-Thoracic Surgery (EACTS). Eur Heart J. 2015;36:2921–2964.
4. Sagar S, Liu PP, Cooper LT. Myocarditis Lancet. 2012;379:738–747.
5. Vabret N, Britton GJ, Gruber C, et al. Immunology of COVID-19: Current State of the Science. Immunity. 2020;52:910–941.
6. Stringhini S, Winiak A, Piussatti G, et al. Seroprevalence of anti-SARS-CoV-2 IgG antibodies in Geneva, Switzerland (SEROCoV-POP): a population-based study Lancet. 2020;396:313–319.
7. Pollao M, Perez-Gomez B, Pastor-Barrusso R, et al. Prevalence of SARS-CoV-2 in Spain (ENE-COVID): a nationwide, population-based seroepidemiological study. Lancet. 2020;396:535–544.
8. Barreiro-Pérez M, Tundidor-Sanz E, Martín-García A, et al. First Magnetic Resonance Managed by a Cardiology Department in the Spanish Public Healthcare System. Experience and Difficulties of an Innovative Model. Rev Esp Cardiol. 2018;71:365–372.
9. Melero-Alegria JF, Casco M, Romero A, et al. SALMANTICOR study. Rationale and design of a population-based study to identify structural heart disease abnormalities; a spatial and machine learning analysis. BMJ Open. 2019;9:e024605.
10. Botafogo V, Pérez-Andres M, Jara-Acevedo M, et al. Age Distribution of Multiple Functionally Relevant Subsets of CD4+ T Cells in Human Blood Using a Standardized and Validated 14-Color EuroFlow Immunometrically Monitoring Tube. Front Immunol. 2020;11:166.
11. Ferreira VM, Schulz-Menger J, Holmvang G, et al. Cardiovascular Magnetic Resonance in Nonischemic Myocardial Inflammation: Expert Recommendations. J Am Coll Cardiol. 2018;72:3158–3176.
12. Imazio M, Trincherio R. Myopericarditis: Etiology, management, and prognosis. Int J Cardiol. 2008;127:17–26.
13. Flores-Montero J, Grigore G, Fluxá R, et al. EuroFlow Lymphoid Screening Tube (LST) data base for automated identification of blood lymphocyte subsets. J Immunol Methods. 2019;475:112662.
14. Shah Z, Mohammed M, Vuddanda V, Ansari MW, Masoomi R, Gupta K. National Trends, Gender Management, and Outcomes of Patients Hospitalized for Myocarditis. Am J Cardiol. 2019;124:131–136.
15. Takahashi T, Ellingston MK, Wong P, et al. Sex differences in immune responses that underlie COVID-19 disease outcomes. Nature. 2020;588:315–320.
16. Huang L, Zhao P, Tang D, et al. Cardiac involvement in recovered COVID-19 patients identified by magnetic resonance imaging. JACC: Cardiovasc Imaging. 2020;13:2330–2339.
17. Punthammi VO, Carej ML, Wieters I, et al. Outcomes of Cardiovascular Magnetic Resonance Imaging in Patients Recently Recovered From Coronavirus Disease 2019 (COVID-19). JAMA Cardiol. 2020;5:1265–1273.
18. Kotecha T, Knight DS, Razvi Y, et al. Patterns of myocardial injury in recovered troponin-positive COVID-19 patients assessed by cardiovascular magnetic resonance. Eur Heart J. 2021;42:1866–1878.
19. Sellers SA, Hagan RS, Hayden FC, Fischer WA. The hidden burden of influenza: A review of the extra-pulmonary complications of influenza infection. Influenza Other Respir Viruses. 2017;11:372–393.
20. Rasso C, Leone O, Rizzo S, et al. Pathological features of COVID-19-associated myocardial injury: a multicentre cardiovascular pathology study. Eur Heart J. 2020;41:3827–3835.
21. Bois MC, Boire NA, Layman AJ, et al. COVID-19-associated Non-Occlusive Fibrin Microthrombi in the Heart. Circulation. 2021;143:230–243.
22. Brito D, Meester S, Yanamala N, et al. High Prevalence of Pericardial Involvement in College Student Athletes Recovering From COVID-19. JACC Cardiovasc Imaging. 2021;14:541–555.
23. Qin C, Zhou L, Hu Z, et al. Dysregulation of immune response in patients with COVID-19 in Wuhan. China Clin Infect Dis. 2020;71:762–768.
24. Flick H, Drescher M, Prattes J, et al. Predictors of H1N1 influenza in the emergency department: proposition for a modified H1N1 case definition. Clin Microbiol Infect. 2014;20:0105–8.
25. Chou A, Serpa JA. Eosinophils in patients infected with human immunodeficiency virus. Curr HIV/AIDS Rep. 2015;12:313–316.
26. Gopal R, Marinelli MA, Alcorn JP. Immune Mechanisms in Cardiovascular Diseases Associated With Viral Infection. Front Immunol. 2020;11:570681.
27. Bautista García J, Peña Ortega P, Bonilla Fernández JA, Cárdenes León A, Ramírez Burgos L, Caballero Dorta E. Acute myocarditis after administration of the BNT162b2 vaccine against COVID-19. Rev Esp Cardiol. 2021;74:812–814.