Structure and Dynamics of Curcumin Encapsulated Lecithin Micelles: A Molecular Dynamics Simulation Study

Lukman Hakim1,2,3, Diah Mardiana1, Urnik Rokhiyah1, Maria Lucia A.D Lestari4, Zubaidah Ningsih1*

1Department of Chemistry, Faculty of Mathematics and Natural Sciences, Universitas Brawijaya, Jl Veteran Malang 65145, Jawa Timur, Indonesia
2Elements Strategy Initiatives for Catalysts and Batteries, Kyoto University, Katsura, Kyoto 615-8520, Japan
3Graduate School of Engineering Science, Osaka University, Toyonaka, Osaka 560-8531, Japan
4Department of Pharmaceutical Science, Faculty of Pharmacy, Universitas Airlangga, Jl Mulyorejo Surabaya 60115, Jawa Timur, Indonesia
*Corresponding author: zubaidah@ub.ac.id

Abstract
Curcumin is a natural product with potential pharmaceutical applications that can be augmented by drug delivery technology such as nano emulsion. Our study focuses on microscopic structural and dynamics response of curcumin encapsulation in micellar system with lecithin as a natural surfactant under variations of composition and temperature using molecular dynamics (MD) simulations. The results highlight the self-assembly of lecithin micelle, with curcumin encapsulated inside, from initial random configurations in the absence of external field. The variation of composition shows that lecithin can aggregate into spherical and rod-like micelle with the second critical micelle concentration lies between 0.17-0.22 mol dm\(^{-3}\). The radial local density centering at the micelle center of mass shows that the effective radius of micelle is indeed defined by the hydrophilic groups of lecithin molecule and the encapsulated curcumin molecules are positioned closer to these hydrophilic groups than the innermost part of the micelle. The spherical micelle is shown to be thermally stable within the temperature range of 277-310 K without a perceivable change in the spherical eccentricity. The dynamics of micelle are enhanced by the temperature, but it is shown to be insensitive to the variation of lecithin-curcumin composition within the studied range. Simulation results are in agreement with the pattern obtained from experimental results based on particle size, polydispersity index, and encapsulation efficiency.

Keywords
Lecithin, Curcumin, Nanoemulsion, Micelle Formation, Molecular Dynamic Simulation.

1. INTRODUCTION
Curcumin is a natural resource and versatile compound that shows anticancer (Tomheh et al., 2019), anti-inflammatory (Jurrenka, 2009), anti-diabetic (Den Hartog et al., 2020), anti-viral, anti-bacterial and anti-fungal activity (Zorofchian M et al., 2014). Nevertheless, low solubility and bioavailability hamper its medical applications. Drug-nanotechnology is considered as a way to overcome the disadvantages and a deep understanding of components interactions in nano emulsion is essential to advance the technology.

Nowadays, the utilization of computer simulation in drug discovery becomes a routine which accelerate the finding of the compound with the highest efficacy with reduced intensive trial and error in laboratory. Computer simulation elaborate drug-target interaction, structure changes, thermodynamics and kinetics aspect which indicate structure stability (Bera and Payghan, 2019; De Vivo et al., 2016; Durrant and McCammon, 2011). In more detail, computer simulation has also been applied in studying the formulation of active compound-loaded nanoemulsion involving liposomal or micellar system (Karjiban et al., 2012; Gupta et al., 2020; Marrink et al., 2000; Moghadasi et al., 2018; Turchi et al., 2019). Nanoemulsion, which consists of active compound, oil-based solvent, emulsifier and water, has widely known to offer higher active compound bioavailability which increases its efficacy in delivering drugs to the targeted cells (Ochoa-Flores et al., 2017; Shelat and Acharya, 2016; Yu and Huang, 2012). Moreover, curcumin nanoemulsion is continuously being developed to improve curcumin solubility and bioavailability.

Despite the fact that curcumin has been developed vastly in nanoemulsion form (Esperón-Rojas et al., 2020; Franklyne et al., 2018; Kamel, 2019; Ningsih et al., 2021; Ochoa-Flores et al., 2017; Páez-Hernández et al., 2019; Shelat and Acharya, 2016; Sholihat et al., 2020; Silva et al., 2018; Teixeira et al., 2016; Yu and Huang, 2012), study of molecules interaction in
curcumin nanoemulsion system is relatively scarce. Simulation studies have been reported on the interaction of curcumin with lipid bilayer (Jalili and Saeedi, 2016; Kopeć et al., 2013) or with inorganic substance (Shi et al., 2019). Although experimental work has been attempted to study the formation of curcumin in soybean oil with Tween-80 surfactant (Jannah, 2020), the molecular-level details on structures and dynamics of the fundamental curcumin encapsulation in lecithin micelle are still not well understood. Molecular dynamics (MD) simulation is a powerful method to elucidate molecular interactions and advance our understanding on the aggregation behaviors at molecular level. The method has been used to study the binding patterns of micelles (Sammalkorpi et al., 2007; Wei et al., 2016), the effects of additive to non-ionic micelles (Piotrovskaya et al., 2006; Vierros and Sammalkorpi, 2018), micelle shape transition (Velinova et al., 2011), as well as surface/interfacial properties (Hakim et al., 2016). MD simulation has been also used to describe the interaction of Tween-80, multiple acid species in soybean oil and water in the absence or presence of curcumin (Moghaddasi et al., 2018), but the work is lacking of systematic approach that start from the fundamental chemical species and incrementally increase the system complexity. On the other hand, it is known that micelle shape in solution can be controlled through variations of temperature, concentration, additives, and ionic strength (Velinova et al., 2011). Here we investigate the curcumin encapsulated lecithin micelle morphological response to the variations of composition and temperature in microscopic details using MD simulation method. In particular, the work signifies the self-assembly process of micelle formation from a random configuration to spherical or rod-like micelles in the absence of an external driving force. These shapes are determined by the concentration of lecithin and less affected by the concentration of curcumin. Simulations also show that the encapsulated curcuminics arrange themselves near the hydrophilic groups of lecithin that forms an interface with the surrounding water molecules.

2. EXPERIMENTAL SECTION

2.1 Method

2.1.1 Simulation Cell and Potential Models

A cubic simulation cell is adopted where lecithin (1-palmitoyl-2-linoleoylphosphatidylcholine) and curcumin (5-hydroxy-1,7-bis(4-hydroxy-3-methoxyphenyl) hepta-1,4,6-trien-3-one) are randomly displaced using PACKMOL (Martinez et al., 2009), followed by insertion of water molecules. In this study, the effects of lecithin-curcumin composition to micelle formation are systematically investigated by preparing several systems whose labels and the corresponding compositions are given in Table 1. The system is labelled as LxKy, where x is the number of lecithin and y is the number of curcumin. The potential models for lecithin and curcumin are built using the Antechamber module in AMBER Tools 18 (Case et al., 2005) and Acrypye according to General Amber Force Field (Wang, 2004). Water molecules are described using the rigid TIP4P/ε water model (Fuentes-Azcuitl and Alejandre, 2014). The model is a re-parameterization from the widely used TIP4P model and has been used to study biomolecular systems (Dominguez, 2016; Onufriev and Izadi, 2018).

2.1.2 Molecular Dynamics Simulations

Molecular dynamics simulations are performed using GROMACS 2018.7 (Abraham et al., 2015). Three dimensional periodic boundary condition and minimum image convention are adopted. The equations of motion are solved using the leapfrog algorithm at a time step of 2 fs. The van der Waals interactions are smoothly truncated at 1.0 nm with a switching function that took effects from 0.8 nm. The long range coulombic interactions are treated using the smooth particle mesh Ewald method with a spline order of 4, a relative tolerance of 10⁻⁵, and the number of reciprocal space mesh was 25 for each of the x, y, and z directions. The temperature is maintained using Nose Hoover thermostat (Hooer, 1985; Nose, 1984) at 277 K, 298 K, and 310 K, with a coupling time of 0.1 ps. The pressure is controlled at 1 bar using the Parrinello Rahman barostat (Parrinello and Rahman, 2005) with a coupling time of 1 ps. Chemical bonds that bind hydrogen atoms are fixed using LINCS algorithm (Hess et al., 1997). The simulation for each system is performed for at least 100 ns.

3. RESULTS AND DISCUSSION

3.1 Micelle Shapes

The amphiphilic structure of lecithin is expected to lead the molecules to form a micelle in aqueous environment. MD simulations shows that the dispersed lecithin molecules in water rapidly form a micelle structure whose shape is dependent to the number of lecithin molecules as depicted in Figure 1 and their sizes are given in Table 2. The spherical micelle shape is further quantified in term of eccentricity, e, given by (Palazzesi et al., 2011; Wei et al., 2016).

\[
e = 1 - \frac{3I_{\text{min}}}{I_1 + I_2 + I_3} \tag{1}
\]

with \(I_1, I_2, \) and \(I_3\) are the three principal moments of inertia, and \(I_{\text{min}}\) is the minimum among those three. The equation above shows that the eccentricity value goes to zero when the micelle shape is close to a perfect sphere. The eccentricity values of the lecithin micelle formed under the studied composition are listed in Table 2. Although the parallelism of the eccentricity values with the compositions is not observed, their values that are close to zero shows that lecithin in most of the studied compositions tends to form a spherical micelle. The effective radius of this spherical micelle, \(R_s\), is defined as (Palazzesi et al., 2011; Wei et al., 2016).

\[
R_s = (5/3)^{1/2} <R_g> \tag{2}
\]
Table 1. Variations in The Number of Lecithin and Curcumin Molecules in The Simulated Systems. The Number of Water Molecules is Fixed to 20,189. The Volumetric Concentrations are Calculated from The Equilibrium Volume Given by MD Simulations at 1 bar and 298 K

| System     | Number of Molecules | Concentration (mol dm$^{-3}$) |   |   |
|------------|---------------------|-------------------------------|---|---|
|            | Lecithin | Curcumin | Lecithin | Curcumin |
| L25K5      | 25       | 5       | 0.06     | 0.01     |
| L50K5      | 50       | 5       | 0.12     | 0.01     |
| L75L5      | 75       | 5       | 0.17     | 0.01     |
| L100K5     | 100      | 5       | 0.22     | 0.01     |
| L25K10     | 25       | 10      | 0.06     | 0.03     |
| L25K15     | 25       | 15      | 0.06     | 0.04     |
| L25K20     | 25       | 20      | 0.06     | 0.05     |
| L25K25     | 25       | 25      | 0.06     | 0.06     |

Table 2. Characteristics of Micelle Formed at Various Composition of Lecithin and Curcumin. The Eccentricity $e$, Rod-like Parameter $s$, and Effective Micelle Radius $R_s$ are Sampled in The Window of 50 – 100 ns Simulation Time. The Uncertainty Values Correspond to The Standard Deviation.

| System     | $e$      | $s$      | Micelle Shape | $R_s$ (nm) |
|------------|----------|----------|---------------|------------|
| L25K5      | 0.09 ± 0.03 | -        | spherical     | 2.24 ± 0.02|
| L50K5      | 0.08 ± 0.03 | -        | spherical     | 2.76 ± 0.01|
| L75K5      | 0.12 ± 0.02 | -        | spherical     | 3.15 ± 0.02|
| L100K5     | -        | 0.024 ± 0.013 | rod-like     | -          |
| L25K10     | 0.10 ± 0.03 | -        | spherical     | 2.28 ± 0.02|
| L25K15     | 0.10 ± 0.04 | -        | spherical     | 2.31 ± 0.02|
| L25K20     | 0.13 ± 0.04 | -        | spherical     | 2.36 ± 0.02|
| L25K25     | 0.09 ± 0.03 | -        | spherical     | 2.38 ± 0.02|

where $R_g$ is the radius of gyration and the angled bracket denotes average. The calculated radii are listed in Table 2, which shows that the micelle size notably grows with the number of lecithin molecules. Figure 1, which also shows that lecithin can form a cylindrical micelle, and thus the shape is quantified in term of rod-like factor, $s$, that is defined as \((\text{Vierros and Sammalkorpi, 2018})\).

$$ s = \frac{I_3 - I_2}{I_3 - I_1} $$  \hspace{1cm} (3)

Similar to the eccentricity value above, the value of $s$ become close to zero when the micelle shape resembles a cylindrical rod. Table 2 shows that the value of $s$ in L100K5 system suggests that the micelle shape is indeed rod-like, as depicted by Figures 1c and 1d. It is important to note, however, that the cylindrical shape is adopted by the micelle because of the presence of periodic boundary condition that connects both ends of cylinder. In other words, the volumetric concentration of lecithin is a determining factor for the shape transition of curcumin encapsulated lecithin micelle. A second critical concentration of micellization that characterize the transition to rod-like assemblies is then expected between L75K5 and L100K5 that correspond to 0.17-0.22 mol dm$^{-3}$.
The spatial distributions of micelle components are useful to understand the structure of micelle. In spherical micelle, the distribution can be quantified in terms of local radial density $\rho_L$ centered at the micelle’s center of mass under equilibrium conditions. Here we focus on the spatial distribution of hydrophilic groups of lecithin, represented by N and P atoms; hydrophobic tail of lecithin, represented by the 8th carbon atom; and curcumin molecule, represented by its carbonyl group. Figure 3 shows the radial local density as a function of its distance from the center of mass of micelle for L25K5, L25K25, and L75K5 systems. As expected from a surfactant, the hydrophilic groups of lecithin are shown to be located on the outermost part of the micelle, while the hydrophobic carbon tail is secluded in the inner part, near the center of mass. It is worth to note that the distance $r$ of the peaks of hydrophilic groups are comparable to the effective spherical radius computed from the radius gyration shown in Table 2. This further emphasizes that the hydrophilic groups correspond to the surface of spherical micelle and directly interacts with the water phase. Figure 3 also shows that curcumin molecules, as represented by their carbonyl group, arrange themselves close to the hydrophilic group of lecithin. However, the overall curcumin molecules are located inside the micelle and interact with the hydrophobic carbon chain of lecithin since curcumin is not an amphoterics species. As the number of curcumin increases, the peak broadening shows the curcumin molecules begin to distribute closer to the inner part of the micelle as shown by Figure 3b. Similar broadening is also observed when the number of lecithin molecules is increased. Figure 3c shows that spherical micelle of larger size allows broader distributions of atoms inside.

### 3.3 Micelle Dynamics

The mean squared displacement (MSD) of lecithin, curcumin, and water molecules are displayed in Figure 4 for L25K5 system. Compared to the freely moving solvent molecules, the mobility of lecithin micelle is obviously limited. The inset in Figure 4 shows that the displacement of micelle is very limited under the reference of center of mass. Note that the smaller curcumin molecules have the similarly limited displacement with the lecithin molecules as a result of being trapped inside the micelle. The inset also shows that the displacements of lecithin and curcumin molecules are more limited in larger micelle. The MSD profiles for the other compositions are similar in trend.

Table 3 shows the self-diffusion coefficient, $D$, that quantitatively describes the mobility of each molecule species in the system. The diffusion coefficient is calculated from the slope of linear fitting of the mean squared displacement curve at interval 5 to 15 ns. The diffusion of lecithin and curcumin molecules decreases with the increase of lecithin composition that corresponds to the increase of micelle size. However, the variations of diffusion coefficients with the increase of curcumin composition are not discernable within the standard deviation.

### 3.4 Thermal Stability

To further examine the effects of temperature to the micelle structure and dynamics, we choose L25K5 system, which produce the smallest effective radius as the subject, and apply three different simulation temperatures: 277 K, 298 K and 310 K. Within the investigated temperature range, increasing the temperature does not result in a spike in the potential energy of the interaction between the molecules in the system. The change of eccentricity values of micelle with the change in temperature are also not noticeable within the standard deviation as shown by Table 4. However, the thermal expansion on micelle size can be seen from the increase of its effective spherical radius.
Figure 3. The Local Radial Density $\rho_l$ Profile of (a) L25K5, (b) L25K25 and (c) L75K5 System at 298 K. The distance of origin ($r=0$) corresponds to the center of mass of the micelle. Each curve is normalized by the number density of the corresponding molecule species.

Table 3. Diffusion Coefficients of Lecithin, Curcumin, and Water Molecules at 298 K

| Sample  | Lecithin  | Curcumin | Water     |
|---------|-----------|----------|-----------|
| L25K5   | 0.10 ± 0.00 | 0.10 ± 0.02 | 4.80 ± 0.02 |
| L50K5   | 0.08 ± 0.03 | 0.11 ± 0.03 | 4.52 ± 0.00 |
| L75K5   | 0.05 ± 0.02 | 0.08 ± 0.02 | 4.23 ± 0.02 |
| L100K5  | 0.06 ± 0.01 | 0.05 ± 0.01 | 4.00 ± 0.00 |
| L25K10  | 0.08 ± 0.05 | 0.08 ± 0.06 | 4.780 ± 0.01 |
| L25K15  | 0.21 ± 0.02 | 0.23 ± 0.02 | 4.77 ± 0.00 |
| L25K20  | 0.10 ± 0.05 | 0.12 ± 0.05 | 4.78 ± 0.01 |
| L25K25  | 0.07 ± 0.04 | 0.07 ± 0.04 | 4.70 ± 0.01 |

Figure 4. Mean Squared-displacement (MSD) of Lecithin, Curcumin and Water Molecules in L25K5 System at 298 K. The Enlarged Plot is Given as an Inset. In Addition, MSD of Each Molecule Species in L75K5 System, that Gives Larger Micelle, at 298 K is Also Plotted as Dashed Lines in The Inset.

radius, and the diffusion of each species increases with the increase of temperature. With relatively small changes in all of the scrutinized parameters, it signifies that nanoemulsion in L25K5 system is thermally stable within the studied different temperatures.

3.5 Relevance with Experimental Observations

In comparison with the previously reported experimental works on the formation of nanoemulsion in lecithin and curcumin with the addition of Tween-80 (Jannah, 2020), the results from MD simulations show some similar patterns. Although the predicted micelle size from all-atom simulations is generally smaller than the experimental results in the laboratory, the observed trend is mostly the same. Experimental works showed that the nanoemulsion size increases with the addition of lecithin, but relatively constant with the addition of curcumin. The experimental results on the efficiency of encapsulation also shows that the curcumin can be encapsulated inside the micelle under the corresponding concentration given in Table 1.

Experiment showed that polydispersity index increases when lecithin molecules number is increasing. At concentrations that correspond to L25K5 up to L75K5, the observed polydispersity index indicates uniformity of the particle size. In contrast, at a concentration that correspond to L100K5, we observed polydispersity index that implies non-uniform particle size, which might also portray a non-similar particle shape (Jannah, 2020). This observation is in line with simulation results where we could identify a shape transition from a spherical to a rod-like structure. It is possible that at higher lecithin concentration, micelle structure is diversified in rod-like as well as spherical structure. On the other hand, the above phenomenon is not observed in the addition of curcumin molecule where the size and shape of the micelles are relative constant. Poly-
disperity index does not change drastically which indicates that the particles produced in this formulation are relatively homogeneous in size and it is possible that the shape of the particles is more uniform in spherical structure. These patterns are reproduced by the MD simulations.

4. CONCLUSIONS

The structures and dynamics of lecithin micelle encapsulating curcumin have been elucidated using MD simulations and the trends are in agreement with the reported experimental works. Within lecithin concentration of 0.06 – 0.17 mol dm$^{-3}$, lecithin forms a spherical micelle whose size is proportional to the number of lecithin molecules. At higher concentration 0.22 mol dm$^{-3}$, lecithin forms a cylindrical micelle. By fixing the lecithin concentration at 0.06 mol dm$^{-3}$, the addition of curcumin up to 0.06 mol dm$^{-3}$ does not alter the micelle shape, but a slight increase in micelle radius is observed as a result of curcumin encapsulation. The local radial density profile shows that the spherical radius of micelle correspond to the hydrophobic groups of lecithin. These groups arrange themselves on the outermost part of the micelle and directly interact with water molecules. The encapsulated curcumin molecules are located close to the hydrophilic groups, instead of being in the innermost part of the micelle where the hydrophobic carbon chains of lecithin are positioned. The self-diffusion coefficients further show that the mobility of curcumin follows the mobility of lecithin. The nanoemulsion is shown to be thermally stable within 277–310 K.

5. ACKNOWLEDGEMENT

This study is supported by Hibah Peneliti Pemula 2020 Universitas Brawijaya Indonesia.

REFERENCES

Abraham, M. J., T. Murtola, R. Schulz, S. Páll, J. C. Smith, B. Hess, and E. Lindahl (2015). GROMACS: High performance molecular simulations through multi-level parallelism from laptops to supercomputers. *SoftwareX, 1*: 19–25.

Bera, I. and P. V. Payghan (2019). Use of molecular dynamics simulations in structure-based drug discovery. *Current Pharmaceutical Design, 25*(11): 3339–3349.

Case, D. A., T. E. Cheatham III, T. Darden, H. Gohlke, R. Luo, K. M. Merz Jr, A. Onufriev, C. Simmerling, B. Wang, and R. J. Woods (2005). The Amber biomolecular simulation programs. *Journal of Computational Chemistry, 26*(16): 1668–1688.

De Vivo, M., M. Masetti, G. Bottegoni, and A. Cavalli (2016). Role of molecular dynamics and related methods in drug discovery. *Journal of Medicinal Chemistry, 59*(9): 4035–4061.

Den Hartogh, D. J., A. Gabriel, and E. Tsiani (2020). Antidiabetic properties of curcumin II: evidence from in vivo studies. *Nutrients, 12*(1): 58.

Dominguez, H. (2016). Molecular dynamics simulations to study the solvent influence on protein structure. *Chemical Physics Letters, 651*: 92–96.

Durrant, J. D. and J. A. McCarmon (2011). Molecular dynamics simulations and drug discovery. *BMC Biology, 9*(1): 1–9.

Esperón-Rojas, A. A., R. Baeza-Jiménez, D. Santos-Luna, L. d. C. Velasco-Rodríguez, L. R. Ochoa-Rodríguez, and H. S. García (2020). Bioavailability of curcumin in nanoemulsions stabilized with mono-and diacylglycerols structured with conjugated linoleic acid and n-3 fatty acids. *Bio-catalysis and Agricultural Biotechnology, 26*: 101638.

Franklyne, J. S., A. Nadarajaj, A. Ebazner, N. Tiwari, A. Mukherjee, and N. Chandrasekaran (2018). Preparation and characterization of edible oil nanoemulsions for enhanced stability and oral delivery of curcumin. *International Journal of Applied Pharmaceutics, 10*: 139–46.

Fuentes-Azcáiz, R. and J. Alejandro (2014). Non-polarizable force field of water based on the dielectric constant: TIP4P/ε. *The Journal of Physical Chemistry B, 118*(3): 1263–1272.

Gupta, K. M., S. Das, P. S. Chow, and C. Macbeath (2020). Encapsulation of ferulic acid in lipid nanoparticles as antioxidant for skin: Mechanistic understanding through experiment and molecular simulation. *ACS Applied Nano Materials, 3*(6): 5351–5361.

Hakim, L., W. D. Saputri, and S. M. Ulfa (2016). Study of curcumin behavior programs. *Journal of Computational Chemistry, 26*(16): 1668–1688.

Hess, B., H. Bekker, H. Berendsen, and J. Fraaije (1997). LINCS: A Linear Constraint Solver for Molecular Simulations. *Journal of Computational Chemistry, 18*: 1463–1472.

Hoover, W. G. (1985). Canonical dynamics: Equilibrium phase-space Distributions. *Physical Review A, 31*(3): 1695–1697.

Jalili, S. and M. Saeedi (2016). Study of curcumin behavior
in two different lipid bilayer models of liposomal curcumin using molecular dynamics simulation. *Journal of Bioenergetics and Biomembranes*, **34**(8): 185–198.

Jannah, L. M. I. E. N. Z., and D. S. R. Mahdin. (2020). Emulsion Formulation of Curcumin in Soybean Oil with a Combination Surfactant of Tween-80 and Lecithin Using Wet Ball Milling Method. *AIP Conference Proceedings*, **2167**(1): 1–6.

Jurrenka, J. S. (2009). Anti-inflammatory properties of curcumin, a major constituent of Curcuma longa: a review of preclinical and clinical research. *Alternative Medicine Review*, **14**(2): 141–153.

Kamel, F. M. L. D., and A. E. (2019). Curcumin-loaded nanosctructured lipid carriers prepared using PecofTM and olive oil in photodynamic therapy: development and application in breast cancer cell line. *International Journal of Nanomedicine*, **14**: 5073–5085.

Karjiban, R. A., M. Basri, M. B. A. Rahman, and A. B. Salleh (2012). Structural properties of nonionic Tween80 micelle in water elucidated by molecular dynamics simulation. *APCBE Proceedings*, **3**: 287–297.

Kopeć, W., J. Telenius, and H. Khandelia (2013). Molecular dynamics simulations of the interactions of medicinal plant extracts and drugs with lipid bilayer membranes. *The FEBS Journal*, **280**(12): 2785–2805.

Marrink, S., D. Tieleman, and A. Mark (2000). Molecular dynamics simulation of the kinetics of spontaneous micelle formation. *The Journal of Physical Chemistry B*, **104**(31): 12165–12173.

Martinez, L., R. Andrade, E. Birgin, and J. Martinez (2009). Software news and update packmol: a package for building initial configurations for molecular dynamics simulations. *Journal of Computational Chemistry*, **30**(13): 2157–2164.

Moghaddasi, F., M. R. Housaindokht, M. Darroudi, M. R. Bozorgmehr, and A. Sadeghi (2018). Soybean oil-based nanoemulsion systems in absence and presence of curcumin: Molecular dynamics simulation approach. *Journal of Molecular Liquids*, **264**: 242–252.

Ningsih, Z., M. L. A. Lestari, and S. A. R. Maharin (2021). Preparation and Characterization of Curcumin Nanoemulsion in Olive Oil-Tween 80 System using Wet Ball Milling Method. *ICS Physical Chemistry*, **1**(1): 16–16.

Nosé, S. (1984). A unified formulation of the constant temperature molecular dynamics methods. *The Journal of Chemical Physics*, **81**(1): 511–519.

Ochoa-Flores, A., J. A. Hernández-Becerra, A. Cavazos-Garduño, I. Soto-Rodríguez, M. Guadalupe Sanchez-Otero, E. J Vernon-Carter, and H. S García (2017). Enhanced bioavailability of curcumin nanoemulsions stabilized with phosphatidylcholine modified with medium chain fatty acids. *Current Drug Delivery*, **14**(3): 377–385.

Onufriev, A. V. and S. Izadi (2018). Water models for biomolecular simulations. *Wiley Interdisciplinary Reviews: Computational Molecular Science*, **8**(2): e1347.

Páez-Hernández, G., P. Mondragón-Cortez, and H. Espinosa-Andrews (2019). Developing curcumin nanoemulsions by high-intensity methods: Impact of ultrasonication and microfluidization parameters. *LWT Journal*, **111**: 291–300.

Palazzesi, F., M. Calvaresi, and F. Zerbetto (2011). A molecular dynamics investigation of structure and dynamics of SDS and SDBS micelles. *Soft Matter*, **7**(19): 9148–9156.

Parrinello, M. and A. Rahman (2005). Polymorphic transitions in single crystals: A new molecular dynamics method. Polymorphic transitions in single crystals: A new molecular dynamics method. *Journal of Applied Physics*, **52**: 7182–7190.

Piotrovskaya, E., A. Vanin, and N. Smirnova (2006). Molecular dynamics simulation of micellar aggregates in aqueous solution of hexadecyl trimethylammonium chloride with different additives. *Molecular Physics*, **104**(22): 3645–3651.

Sammalkorpi, M., M. Karttunen, and M. Haataja (2007). Structural properties of ionic detergent aggregates: a large-scale molecular dynamics study of sodium dodecyl sulfate. *The Journal of Physical Chemistry B*, **111**(40): 11722–11733.

Shelat, D. Y. and S. R. Acharya (2016). CUR-CA-THIONE: A novel curcumin conection with enhanced water solubility and brain bio-availability. *International Journal Pharmacy and Pharmaceutical Sciences*, **8**: 265–270.

Shi, X., Y. Wang, H. Sun, Y. Chen, X. Zhang, J. Xu, and G. Zhai (2019). Heparin-reduced graphene oxide nanocomposites for curcumin delivery: in vitro, in vivo and molecular dynamics simulation study. *Biomaterials Science*, **7**(3): 1011–1027.

Sholihat, S. I., E. Indahyanti, M. L. A. Lestari, and Z. Ningsih (2020). Preparation of Curcumin Nanoemulsion in Soybean Oil–Tween 80 System by Wet Ball Milling Method. In *IOP Conference Series: Materials Science and Engineering*, volume 833. IOP Publishing, page 012044.

Silva, H. D., J. Poejo, A. C. Pinheiro, F. Donsi, A. T. Serra, C. M. Duarte, G. Ferrari, M. A. Cerqueira, and A. A. Vicente (2018). Evaluating the behaviour of curcumin nanoemulsions and multilayer nanoemulsions during dynamic in vitro digestion. *Journal of Functional Foods*, **48**: 605–613.

Teixeira, C., L. Mendonça, M. Bergamaschi, R. Queiroz, G. Souza, L. Antunes, and L. Freitas (2016). Microparticles containing curcumin solid dispersion: stability, bioavailability and anti-inflammatory activity. *AAPS PharmSciTech*, **17**(2): 252–261.

Tomheh, M. A., R. Hadianamrei, and X. Zhao (2019). A review of curcumin and its derivatives as anticancer agents. *International Journal of Molecular Sciences*, **20**(5): 1033.

Turchi, M., Q. Cai, and G. Lian (2019). In Silico Prediction Method. *Journal of Computational Chemistry*, **40**(3): 1011–1027.

Velino, M., D. Sengupta, A. V. Tadjer, and S.-J. Marrink (2011). Sphere-to-rod transitions of nonionic surfactant micelles in aqueous solution modeled by molecular dynamics simulations. *Langmuir*, **35**(33): 10855–10865.

Vierros, S. and M. Sammalkorpi (2018). Effects of 1-hexanol on C12 E10 micelles: a molecular simulations and light scattering study. *Physical Chemistry Chemical Physics*, **20**(9):
6287–6298
Wang, J. K. P. C., Wolf RM; Caldwell (2004). Development and testing of a general amber force field. *Journal of Computational Chemistry, 25;* 1157–1174
Wei, Y., H. Wang, G. Liu, Z. Wang, and S. Yuan (2016). A molecular dynamics study on two promising green surfactant micelles of choline dodecyl sulfate and laurate. *RSC Advances, 6*(87); 84090–84097
Yu, H. and Q. Huang (2012). Improving the oral bioavailability of curcumin using novel organogel-based nanoemulsions. *Journal of Agricultural and Food Chemistry, 60*(21); 5373–5379
Zorofchian M, S., Abdul Kadir, P. Hassandarvish, H. Tajik, S. Abubakar, and K. Zandi (2014). A review on antibacterial, antiviral, and antifungal activity of curcumin. *BioMed Research International, 2014*