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A Unique Protein Phosphatase with Kelch-Like Domains (PPKL) in *Plasmodium* Modulates Ookinete Differentiation, Motility and Invasion

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Abstract

Protein phosphorylation and dephosphorylation (catalysed by kinases and phosphatases, respectively) are post-translational modifications that play key roles in many eukaryotic signalling pathways, and are often deregulated in a number of pathological conditions in humans. In the malaria parasite *Plasmodium*, functional insights into its kinome have only recently been achieved, with over half being essential for blood stage development and another 14 kinases being essential for sexual development and mosquito transmission. However, functions for any of the plasmodial protein phosphatases are unknown. Here, we use reverse genetics in the rodent malaria model, *Plasmodium berghei*, to examine the role of a unique protein phosphatase containing kelch-like domains (termed PPKL) from a family related to *Arabidopsis* BSU1. Phylogenetic analysis confirmed that the family of BSU1-like proteins including PPKL is encoded in the genomes of land plants, green algae and algae, but not in other eukaryotic lineages. Furthermore, PPKL was observed in a distinct family, separate to the most closely-related phosphatase family, PP1. In our genetic approach, C-terminal GFP fusion with PPKL showed an active protein phosphatase preferentially expressed in female gametocytes and ookinetes. Deletion of the endogenous ppkl gene caused abnormal ookinete development and differentiation, and dissociated apical microtubules from the inner-membrane complex, generating an immotile phenotype and failure to invade the mosquito mid-gut epithelium. These observations were substantiated by changes in localisation of cytoskeletal tubulin and actin, and the micronemal protein CTRP in the knockout mutant as assessed by indirect immunofluorescence. Finally, increased mRNA expression of *dozi*, a RNA helicase vital to zygote development was observed in *ppkl* mutant, with global phosphorylation studies of ookinete differentiation from 1.5–24 h post-fertilisation indicating major changes in the first hours of zygote development. Our work demonstrates a stage-specific essentiality of the unique PPKL enzyme, which modulates parasite differentiation, motility and transmission.

Introduction

Reversible protein phosphorylation is a ubiquitous regulatory process for a variety of eukaryotic and prokaryotic pathways, including cell-cycle regulation, cell to cell signalling, cell proliferation and differentiation [1]. In humans, aberrant regulation of protein phosphorylation has been implicated in cancers [2], and plays a central role in many other pathological diseases [3]. Protein phosphorylation and dephosphorylation is catalysed by protein kinases and phosphatases, respectively [4–7], and nearly two-thirds of the proteins encoded in the human genome are believed to be modified by reversible phosphorylation [8] at over 650,000 phosphosites (PhosphoNET KnowledgeBase (www.phosphonet.ca)), emphasizing the importance of this post-translational modification.

This regulatory mechanism is highly conserved [3], and plays a vital role in development of apicomplexan protozoan parasites of the genus *Plasmodium* [9,10], which are globally responsible for over a million deaths annually through malaria [11]. The *Plasmodium* life-cycle proceeds via a number of distinct developmental stages: asexual exo-erythrocytic proliferation in liver hepatocytes and intra-erythrocytic multiplication in erythrocytes...
Author Summary

Malaria parasites are single-celled organisms, which alternate their life-cycle between vertebrate and mosquito hosts. In the mosquito, the malaria parasite undergoes sexual development, whereby a male and female gamete fuse to form a zygote. This zygote then elongates into an invasive stage, termed an ookinete, which can glide to and penetrate the mosquito’s gut wall in order to form a cyst (called an oocyst). Protein phosphorylation is known to play a vital role during this process; however, the role of Plasmodium kinases (which phosphorylate proteins) during zygote/ookinete maturation is better understood than the completely uncharacterised plasmodial phosphatases (which dephosphorylate proteins). Using a malaria parasite which infects mice, Plasmodium berghei, we show that a unique protein phosphatase containing kelch-like domains (called PPKL) plays a vital role in ookinete maturation and motility. Deleting this gene produces ookinetes whose shape is grossly abnormal, resulting in non-motile parasites that cannot penetrate the lining of the mosquito gut wall. Overall, PPKL is an essential phosphatase that is critical to ookinete development, motility and invasion.

of the vertebrate host, and sexual development in the female Anopheles mosquito [12]. During both asexual and sexual development the parasite utilises a number of signalling pathways, many of which involve reversible protein phosphorylation. Systematic functional analyses of plasmodial protein kinases (PKs) in both the human parasite P. falciparum and the rodent model P. berghei species have revealed that over half of their kinome is essential to asexual blood stage schizogony [9,10]. Furthermore, reverse genetic studies in P. berghei have shown that a further 14 PKs have specific functions during sexual development of the parasite within the mosquito mid-gut lumen and subsequent migration to the salivary glands [10,13–21]. In particular, development of the motile and invasive ookinete within the mosquito mid-gut is known to be highly dependent upon two NIMA-related kinases, NEK2 and NEK4 [14,15]. Of the three invasive stages of malaria parasite development (namely the sporozoite, merozoite and ookinete), the ookinete stage is unique in that it can develop extracellularly and lacks rhoptries, an organelle specifically associated with cell invasion. This is in contrast to the other “zoite” forms (merozoite and sporozoite) that develop intracellularly and contain both rhoptries and micronemes as apical organelles. However, all of these invasive stages comprise a unique cortical structure termed a pellicle, which consists of a parasitic plasma membrane and an underlying double membrane structure termed the inner membrane complex (IMC) [22]. Parasite motility is powered by an actin myosin motor termed the glideosome [23], which resides within the pellicle of invasive stages. At the molecular level, the motility and mid-gut invasion of the ookinete involves the secretion of a number of membrane proteins including the motor complex-associated proteins glideosome-associated protein 45 (GA45) and myosin-A tail domain interacting protein (MTIP) [24], micronemal proteins such as circumsporozoite and trap-related protein (CTRP) [25] and secreted ookinete adhesive protein (SOAP) [26], as well as calcium-dependent protein kinase 3 (CDPK3) [21].

Although the role of plasmodial kinases has been intensively studied, the role of all the complementary protein phosphatases (PPs) during any stage of Plasmodium development is unknown. Biochemical studies have shown that P. falciparum possesses predominantly phosphatase-1-like activities, and chemical inhibition of phosphatase activity by use of calyculin A and okadaic acid significantly reduces asexual blood stage proliferation [27]. In contrast to the human phosphatome (comprising approximately 156 phosphatases [28] [and PhosphoNET KnowledgeBase (www.phosphoenet.ca)], the Plasmodium genome codes for one of the smallest phosphomes of all the eukaryotic phyla known to date, with 27 putative protein phosphatases falling into four major classes: phosphoprotein phosphatases (PPPs), metallo-dependent protein phosphatases (PPMs), protein tyrosine phosphatases (PTPs) and NLI interacting factor-like phosphatases (NIFs) [29].

Like the kinases (although fewer in number), the Plasmodium phosphatome contains members that have no orthologues in mammalian systems [29]. The function of the Shewanella-like PPs (Shelps) and the likely inactive pseudo-phosphatase EF-hand containing phosphatase (EFPP) is completely unknown [29]; however, in P. falciparum, Shelps are postulated to have a role in erythrocyte invasion [30]. Furthermore, a distinctive PP1-related enzyme belonging to the PPP family of phosphatases comprising an N-terminal kelch repeat domain and a C-terminal PP1-like phosphatase domain (named PPKL: Protein Phosphatase with Kelch-Like domains) has been detected in the apicomplexans Cryptosporidium hominis, Toxoplasma gondii and Theileria parva (one gene per genome), as well as in the land plants Arabidopsis thaliana and Oryza sativa (4 and 5 genes respectively) [31,32]. The kelch motif is widespread and involved in many cellular functions, particularly in actin-based cytoskeleton formation and transcriptional regulation [33]. PPKL itself has only been studied in detail in Arabidopsis thaliana (where it is known as BSU1 or bri1 suppressor1) and along with the kinase BIN2, is involved in brassinosteroid hormone signalling and phosphorylation of the transcription factors BZR1 and BES1 [34]. Even though PPKL was first discovered in P. falciparum (initially named PPP3), where RT-PCR analysis showed mRNA expression exclusively in gametocytes and hence suggesting a role during sexual development [35], its function has remained unknown. Transcriptomic and proteomic studies of P. falciparum [36] and P. berghei [37] showed high levels of pppk transcripts in gametocytes and PPKL protein in ookinetes, respectively (PlasmoDB), although discordant results were found in a study of the proteome of sex-specific gametocytes [18].

In this study, we have used P. berghei to elucidate the function of the PPKL enzyme during the Plasmodium life-cycle. We show by using reverse genetics that Plasmodium PPKL has an essential function during ookinete differentiation, affecting apical end integrity, collar and pellicle morphology, and microtubule linkage to the IMC that are crucial for parasite shape, motility and invasion.

Results

PPKL is a unique Ser/Thr phosphatase with N-terminal kelch domains and is limited to Alveolata and Viridiplantae

To identify PPKL-like phosphatases in Plasmodium, we used BSU1 from Arabidopsis thaliana to seed an iterative profile-based similarity search [38]. Phylogenetic analysis based on a common phosphoesterase domain (Pfam: PF00149) revealed a robust family of BSU1-like proteins encoded in the genomes of land plants, green algae and alveolates (Figure 1A, B), but not other eukaryotic lineages, in agreement with previous studies [31,32]. All members of the BSU1-like family including the single example contained in each Plasmodium spp share a distinctive conserved protein

PPKL Regulates Ookinete Morphology and Invasion
architecture, with N-terminal kelch repeats followed by the phosphatase domain.

*P. berghei* PPKL contains five complete and one truncated kelch repeat (Figure 1A), a configuration which is conserved in most identified orthologues. Phylogenetic trees and identity matrices of individual kelch domains showed that the fourth domain has the highest identity between species, while the sixth was the least similar (Figure S1). The phosphatase domain of *P. berghei* PPKL shows the greatest similarity to type 1 and 2A protein phosphatases. All the important signature motifs of the serine/threonine phosphatase (STP) family are present (Figure S2); they form the active site, and are known to have a role in metal ion binding (GDxHG, GDxVDRG, and GNHE) [39]. The binding site for the inhibitor microcystin is conserved, but some of the residues important for docking the PP1 inhibitor Inhibitor-2 and the PPP inhibitor okadaic acid are not conserved, making it difficult to predict *in silico* the inhibitor specificity of this enzyme. Additionally, we identified 4 conserved insertions (Figure S2I-IV) in the catalytic domain, which are only present in *Apicomplexa*, suggesting a specific role for these sequences in this group.

PPKL has protein phosphatase enzyme activity, and is highly expressed in female gametocytes and ookinetes.

Transcriptomic and proteomic studies in *Plasmodium* [36,37] have suggested that *ppkl* transcripts and PPKL protein are present in gametocytes and ookinetes only, respectively, although PPKL is not present in the proteome of sex-specific gametocytes [18]. To confirm this, we analysed *ppkl* mRNA expression by qRT-PCR and PPKL protein expression and localisation by generation of a C-terminal green fluorescent protein (GFP) fusion protein of endogenous *ppkl* (PBANKA_132950) using a single crossover recombination strategy (Figure S3A-E).

*ppkl* mRNA is expressed in asexual blood stages, gametocytes and ookinetes, with the highest expression in schizonts (Figure 2A) relative to *hsp70* and arginyl-tRNA synthetase genes used as controls in this assay. The intensity of PPKL-GFP fluorescence in sexual stages was highest in the nuclear and cytoplasmic compartments of female gametocytes and zygotes, and in the apical cytoplasm of ookinetes. The protein was also detected in oocysts and sporozoites, but was not observed in activated microgametocytes or microgametes (Figure 2B). These data suggest that in sexual stages PPKL is female-specific and has a role during sexual development.

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**Figure 1. Phylogenetic analysis of Plasmodium PPKL.**

A. Schematic representations of the predicted protein architectures (Pfam domains) of the PPKL orthologues in *A. thaliana* and *P. falciparum*. B. Bayesian phylogeny of kelch-like phosphatases. Tree shown is the consensus of trees from four independent inferences based on a trimmed alignment of the phosphatase domain. Numbers beside nodes represent support from Bayesian posterior probabilities.

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We also investigated whether PPKL is an active phosphatase in *P. berghei*. Using 3-O-methylfluorescein phosphate (MFP) as a substrate, we found that PPKL-GFP immunoprecipitated with GFP-TRAP beads from parasite lysates had phosphatase activity proportional to the amount of lysate in the assay; whereas the WT parasite lysate controls produced much lower fluorescence levels (Figure 2C). The residual fluorescence observed in the WT lysate compared to the corresponding control without MFP substrate might be due to endogenous phosphatase activity non-specifically pulled-down by the GFP-TRAP beads. Our results therefore indicate that PPKL has protein phosphatase activity, correlating with its orthologue in *Arabidopsis*, BSU1 [34].

An absence of PPKL results in abnormal ookinete morphology and is contributed by the female gamete

To examine the function of PPKL during the *Plasmodium* life-cycle, we used a double homologous recombination strategy to replace the endogenous *ppkl* gene with a *dhfr*/*ts* selectable marker from *Toxoplasma gondii* (Figure S4A–E). Analysis of two *ppkl* deletion mutant clones from two independent transfections, named *ppkl*− c13 and *ppkl*− c19 identified no phenotypic changes during asexual proliferation in terms of parasite growth, erythrocyte invasion or morphology and no effect on sexual stage cell development (gametocytogenesis), as assessed on blood smears (data not shown). Gametogenesis in the activated microgametocyte was also comparable to wild-type controls (Figure 3A). However, analysis of *in vitro* cultures for 18–24 h to monitor differentiation into ookinete stages [40] showed that cultures of *ppkl*− mutants were dominated by grossly abnormal retort forms [41] (Figure 3B, C, S5C). As a result of this observation, we performed a time-course analysis of ookinete differentiation over 6, 9, 12, 15, 18 and 24 h. Development of the zygote through stages I–III of ookinete maturity (0–9 h post-fertilisation) [41] was indistinguishable in the *ppkl*− mutants compared to wild-type. Wild-type controls showed normal progression and maturation to stage IV (at 12 h), with 69% of all macrogamete-derived parasites progressing to stage VI 24 h post-fertilisation (Figure 3D). In contrast, the majority of *ppkl*− mutants did not progress from stage III to stage IV, but produced a high proportion of abnormal retort forms 24 h post fertilisation (35% of the total population) with only 4% progressing to stage IV–VI. This suggests that PPKL is essential for maturation, differentiation and morphological development of ookinetes from stage III to stage IV.

We next examined whether the defect was sex-specific by performing genetic crosses between *ppkl*− parasites and lines deficient in either male (*map2*−) or female (*nek4*−) gametes [14,40]. Cross-fertilization with *nek4*− parasites did not rescue the phenotype, whereas crossing with *map2*− gametocytes resulted in 26% of all macrogamete-derived parasites progressing to stage VI 24 h post-fertilisation (Figure 4A). This suggests that PPKL is essential for maturation, differentiation and morphological development of ookinetes from stage III to stage IV.

PPKL is essential for gliding motility and mosquito invasion

To substantiate the *in vitro* findings, parasites in mice infected with either wild-type or *ppkl*− gametocytes were fed to mosquitoes to analyse oocyst development. Wild-type oocysts developed normally; whereas no oocysts were found in the guts of mosquitoes
fed on ppkl− parasites when analysed 14 and 21 days after feeding (Figure 4B). This result confirms that PPKL is vital to ookinete development and that oocyst formation is completely blocked in the ppkl− parasites.

Due to the ablation of oocyst development in our in vivo study (Figure 4B), as well as defects in the morphology and maturation of ppkl− mutants, we analysed gliding motility of wild-type and ppkl− parasites by embedding cultured ookinetes in dilute Matrigel and using time-lapse video microscopy to quantify gliding movement [20]. Using this method, we found that wild-type parasites followed a characteristic helical gliding motion with an average speed of 4.74 μm/min (Figure 4C and Video S1), in close agreement with previous studies [20]. Strikingly, gliding motility in ppkl− retorts was significantly reduced compared to wild-type parasites (0.13 μm/min; \( p < 0.001 \)), although occasional limited forward motion and "flexing" at the apical end was observed (Figure 4C and Video S2).

To assess whether the ablation of oocyst development in ppkl− mutant parasites was also due to a defect in invasion of the mid-gut epithelium, we bypassed the gut barrier by injecting ppkl− parasites directly into the haemocoel of A. stephensi mosquitoes and analyzed salivary gland invasion 20 days post-injection [42]. Using this method, we found that ppkl− parasites were able to form viable sporozoites, which could migrate to the salivary gland and actively invade (Table 1). Onward transmission experiments via a mosquito biting resulted in infection in mice with both wild-type and ppkl− lines. Subsequent analysis of parasites recovered from these lines and cultured in ookinete medium for 24 h confirmed the knockout phenotype in the ppkl− lines. This strongly suggests that sporozoites of ppkl− mutants are able to migrate to and invade the salivary glands, as well as undergo exo- and intra-erythrocytic proliferation, but the abnormal retorts of ppkl− are not able to penetrate the epithelial lining of the mosquito mid-gut.

**ppkl− parasites show defects in ookinete apical end integrity**

To examine whether the morphology of the ppkl− mutant retorts could be linked to their inability to glide and invade, we performed ultrastructure analyses using transmission electron microscopy (TEM).
Significant structural differences were detected between wild-type and ppkl2 mutant parasites at the apical end of the ookinete. Wild-type ookinetes had a very uniform appearance with a conical shaped apical end (Figures 5Ai), with unique and complex structures [43] consisting of an enclosing plasmalemma, beneath which is a conical electron dense collar with a central aperture and in firm contact with the IMC (Figure 5Aiii, S5A). Beneath the collar, but in close contact with it, is a second somewhat smaller and less electron dense layer termed the apical ring (Figure 5Aiii, S5A), which is subtended by the sub-pellicular microtubules. The apical end also has a large number of micronemes with fine ducts running through the aperture in the collar to the apical plasmalemma (Figure 5Aiii, S5A). In contrast, ppkl2 retort forms showed marked variation in shape with clear constriction and elongation of the apical end (Figure 5Aii, 5Avii and S5B). Detailed examination of the apical end showed structural differences from the wild-type, namely: the electron dense collar’s reduced length (Figure 5Aiv, S5B); the apical ring appeared reduced in size compared to the wild-type, resembling a series of small vesicles and some loss of connection to the collar (Figure 5Aiv); the microtubules run longitudinally from the ring, but while they were closely associated with the IMC in the wild-type parasite (Figure 5Av), they were more disorganised in the mutant and the electron dense links between the microtubules and the IMC were not found (Figure 5Avi); in severe cases, collapse of the apex resulted in groups of tubules within a neck-like region not shown in the dot plot. Bar = arithmetic mean; n = 17 for wild-type and 28 for ppkl2 lines.

**Table 1.** Sporozoite infectivity in ookinetes directly injected into the haemocoel.

| Experiment | Parasite line | Number of salivary gland sporozoites/mosquito | Biteback (%) | First day of detectable blood stage infection |
|------------|---------------|---------------------------------------------|--------------|----------------------------------------------|
| 1          | Wild type     | 1840                                        | 2/2 (14/23)  | 4 dpi                                        |
|            | ppkl2         | 147                                         | 1/2 (11/8)   | 6 dpi                                        |
| 2          | Wild type     | 29’321                                      | 2/2 (31/23)  | 4 dpi                                        |
|            | ppkl2         | 20’500                                      | 2/2 (27/23)  | 4 dpi                                        |

Y = number of mosquitoes fed.
dpi = days post-infection.

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associated with the IMC (Figure 5Avii, viii). Due to the collapse of the apex, fewer micronemes were located in the apical region, but those present appeared normal with ducts running to the apex (Figure 5Aiv, S5B). The other cytoplasmic and nuclear features were similar in both wild-type and mutant parasites (Figure 5Ai, ii). In summary, there is a loss of apical integrity due to defects in the collar and the attachment between the microtubules and IMC possibly resulting in an immotile parasite.

Indirect immunofluorescence analysis using antibodies for the cytoskeletal proteins tubulin and actin showed mislocalisation with intense atypical tubulin staining at the apical end of the \( ppkl^- \) mutant as well as loss of the normal apical staining for actin. While the motor complex proteins GAP45 and MTIP showed no obvious abnormal pattern in \( ppkl^- \) lines, micronemal CTRP showed less distinct apical staining and is more distributed throughout the body of the retort. This may be due to the structural and cytoskeletal changes at the apical end. However, no change in the normally diffuse intracellular distribution of the microneme-associated protein SOAP was observed (Figure 5B).

Expression of genes essential to zygote development is differentially regulated in PPKL mutants

The protein kinase NEK4 and the DDX6-class RNA helicase DOZI are essential to female/zygote development [14,18,44].
ascertain whether PPKL is regulated by either of these, we analysed mRNA expression of the phosphatase in mutants of NEK4 and DOZI by qRT-PCR. Expression of ppkl compared to wild-type was only significantly altered in both non-activated and activated gametocytes of the nek4<sup>−/−</sup> mutant (p<0.001 for both). In ppkl<sup>−/−</sup> mutants, transcript levels of nek4 compared to wild-type controls were indistinguishable in both total asexual blood stages and non-activated and activated gametocytes (Figure 6A). However, dozi was significantly up-regulated compared to wild-type (p<0.01 for all stages analysed), suggesting that PPKL could be involved in the function of this RNA helicase.

**PPKL is hyper-phosphorylated in activated gametocytes compared to schizonts**

As studies in *Arabidopsis thaliana* have shown that the activity of BSU1 is regulated by phosphorylation [45], we investigated whether PPKL was phosphorylated in *P. berghei* using PPKL-GFP parasites metabolically labelled for 30 min with 32P-orthophosphate, lysed and immunoprecipitated with GFP-TRAP to assess PPKL phosphorylation *in vivo*. Activated gametocytes showed a higher level of phosphorylation compared to schizonts (7.6 times) as assessed by measuring the ratio of the intensity of the PPKL phosphorylation signal on the autoradiograph to the intensity of the PPKL protein band on the Western blot (Figure 6C, arrow).

**Global phosphorylation is altered in ppkl<sup>−/−</sup> mutants**

As PPKL is essential for ookinete maturation, we assessed the impact of ppkl deletion on protein phosphorylation during ookinete development. This was achieved by analysing global phosphorylation profiles of wild-type and ppkl<sup>−/−</sup> parasites 1.5, 6 and 24 h post gametocyte activation (pga) using a metabolic labelling technique [46]. Three representative fractions (9–11), in which significant differences in phosphorylation were observed are shown in Figure 6C. Equal protein loading was assessed by Coomassie blue staining (data not shown). While the majority of the phosphorylated proteins remained unchanged in wild-type versus ppkl<sup>−/−</sup> lysates, as early as 1.5 h pga (Figure 6C, arrows), and also later in zygote development (6 h) (Figure 6C, arrowhead) and 24 h pga (Figure 6C, asterisk), we observed several specific changes, including both increases and decreases in phosphorylation levels.

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**Figure 6. Differential transcript and phosphorylation levels in ppkl<sup>−/−</sup> and mutants of genes essential for zygote development.**

A. Relative expression of ppkl, nek4 and dozi in ppkl<sup>−/−</sup>, nek4<sup>−/−</sup> and dozi<sup>−/−</sup> mutant parasites compared to wild-type controls (Pfaffl method). Error bar = ±SEM, n = 3 from three independent experiments. ***p<0.001; **p<0.01. ASB = asexual blood stages; NAG = non-activated gametocytes; AG = activated gametocytes. B. Upper panel: autoradiograph showing phosphorylation in lysates of schizonts and activated gametocytes from WT-GFP and PPKL-GFP parasite lines. Lower panel: corresponding Western blot using anti-GFP antibody. Sch = schizonts; AG = activated gametocytes. C. Autoradiograph from three fractions showing alterations in global phosphorylation 1.5, 6 and 24 h post-activation of wild-type and ppkl<sup>−/−</sup> gametocytes. Differential phosphorylation in ppkl<sup>−/−</sup> compared to wild-type 1.5 h after gametocyte activation is represented by arrows, 6 h after gametocyte activation by an arrowhead and 24 h after gametocyte activation by an asterisk. Representative radiographs of three independent experiments are shown.

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These data suggest that although PPKL is involved in the regulation of the phosphorylation status of specific proteins throughout ookinete development, the largest impact of PPKL on protein phosphorylation occurs early (1.5 h) after gametocyte activation.

**Discussion**

Reversible phosphorylation is a major regulator for many cellular processes. The kinases are well recognised as important signalling molecules and drug targets in various diseases, but phosphatases have been neglected and their roles remained elusive until recently [32,47]. Although recent systematic functional analyses of the *Plasmodium* kinome have identified a number of potential drug targets [9,10], studies on phosphatases in *Plasmodium* have been mostly limited to biochemical studies. Recently however, the total protein phosphatome of *P. falciparum* was published and showed that the genome of this malaria parasite codes for one of the smallest known eukaryotic phosphatomes, comprising 27 phosphatase sequences [28]. We have shown that the unique phosphatase PPKL is encoded by a single-copy gene belonging to a robust family that is only detected in *Viridiplantae* and *Alveolata*, confirming previous studies [31,32]. Analysis of wild-type mRNA transcription and protein expression confirmed the findings of previous global studies [36,37], suggesting that PPKL is expressed during sexual development. Furthermore, we have also shown that PPKL is an active phosphatase, which in gametocyte/gamete stages is female-specific and in the ookinete is found preferentially at the apical end. However, the localisation is not uniquely nuclear but is also cytosolic, contrasting the exclusively nuclear localisation of the *Arabidopsis* BSU1 phosphatase [34].

Our functional studies have shown that the only point of essentiality for PPKL in the malaria parasite is ookinete differentiation, particularly in the latter stages when the machinery for motility and invasion is formed. Zygote development is normal in *ppkl* mutants, but ookinete differentiation is grossly impaired, producing a parasite population dominated by retorts with abnormal morphology and varying degrees of structural deformities. By using a motility assay we show here that the gliding motility of *ppkl* abnormal retorts is virtually abolished. The data suggest that apical structure and organelle distribution are determined at the early female/zygote stage. We also show that this defect is inherited through the female line. Previous studies have shown that many defects at the zygote/ookinete stage are carried through maternal inheritance of a number of ookinete specific molecules [42]. In addition, the haemocoeal infection experiment clearly suggests that this impairment is restricted to the ookinete stage since by-passing the mid-gut barrier by direct injection of mutant retorts into the haemocoeal allowed oocyst formation and the production of invasive sporozoites that could cause blood stage infection after a mosquito bite-back. However, the abnormality persisted at the subsequent ookinete stage in these *ppkl* parasites.

The ookinete is morphologically and biochemically distinct from sexual stage gametocytes and zygotes, as well as the later oocyst and sporozoite stages. It is also the only motile and invasive stage that is formed extracellularly, does not require invasion of a new host cell as it moves between the mosquito gut cells in traversing to the basal surface and also develops extracellularly into the next stage. Ultrastructure analyses using TEM revealed severe structural abnormalities in the apical end of the mutant retort forms, particularly that of microtubule organisation and association with the IMC, as well as a reduction in the length of the apical ring and distinct shortening of the apical collar. The apical complex of the ookinete is unique amongst the invasive *Plasmodium* stages. In particular, the distinctive ookinete morphology is maintained through an array of microtubules, the subpellicular network [22], and a lattice of intermediate filaments.

These are enclosed by the pellicle consisting of the plasmalemma and two underlying closely associated unit membranes formed from flaten vacuoles and referred to as the IMC. The structural abnormalities seen in the microtubular association with the IMC, as well as the mislocalisation of cytoskeletal proteins in *ppkl* lines could explain the distinctive morphology of the mutant. It would also explain why no *ppkl*–associated defects are observed in the rest of the life-cycle since the unique apical complex of the ookinete is absent in both merozoites and sporozoites.

Immunofluorescence studies with antibodies to the microneme marker CTRP suggested that in *ppkl* lines CTRP is distributed throughout the parasite body and the apical end localisation as seen in the wild-type parasite is not observed. It is important to note however, that even though CTRP and SOAP are both microneme-associated proteins, the localisation we see in our wild-type parasites is consistent with previous studies [26,48], with CTRP showing intense apical staining and SOAP localisation more diffuse throughout the cell body. The altered distribution of CTRP in *ppkl* lines could more likely be due to the structural abnormality at the apical end, since we still observe a high concentration of apical micronemes in the mutants. Although this may explain their inability to move and invade the gut wall, it has been shown previously in misfit disruption mutants [49] that even in the absence of micronemes, parasites could still form oocysts. Nevertheless, this particular defect in MISFIT is quite different to *ppkl* because it shows paternal inheritance and a range of distinct molecular defects. The location of markers of the motor complex such as GAP45 and MTIP did not show any marked difference between wild-type and *ppkl* parasites. However, the mislocalisation of the cytoskeletal protein tubulin and of actin in *ppkl* parasites is consistent with the structural defects identified in the ultrastructure studies. In mammalian systems protein phosphatases are important in microtubule formation and are regulated during the cell cycle [50]; it is therefore possible that PPKL has a direct role in microtubule distribution, consistent with the aberrant morphology of the *ppkl* retorts.

Cellular processes are regulated through complex signalling networks, although very little is known in *Plasmodium*. Studies of kinases have demonstrated the importance and specificity of protein phosphorylation at every stage of the *Plasmodium* life-cycle. For example, a number of kinases are implicated in male gamete (CDPK4, SRPK, MAP2), female gamete (NEK2, NEK4), zygote (NEK2, NEK4), and ookinete (PK7, GAK) development [10,51]. We have shown that *ppkl* transcripts are down-regulated in nek4 mutants. Furthermore, we have also shown that PPKL is phosphorylated in both schizonts and activated gametocytes, confirming phosphoproteomic studies indicating that PPKL is itself phosphorylated [32] and suggesting that its activity or interactions with other proteins may also be regulated by phosphorylation. As a precedent, phosphorylation of the *Arabidopsis* PPKL orthologue, BSU1, by the constitutive differential growth 1 (CDG1) kinase results in its activation and the subsequent dephosphorylation of BIN2 [45], suggesting a fine balance between phosphorylation/dephosphorylation pathways in signalling in plants [53]. Whether such a mechanism operates in *Plasmodium* is to be elucidated in future studies.

The global protein phosphorylation profile during ookinete development of the *ppkl* mutant is evidently different from that of wild-type parasites. Most changes were seen within 1.5 h of gametocyte activation and zygote formation, suggesting that the
function of PPKL is crucial at the early zygote stage even though the phenotype is observed at the oocyte stage. For example, the molecular blueprint for apical end formation may be established following fertilization in the early zygote stage. One intriguing observation is that the qRT-PCR analyses revealed that transcripts of dozi (which is involved in mRNA processing) [44] are significantly up-regulated in ppklI lines. This might suggest that in the absence of the phosphatase the modulation of DOZI is perturbed and this could directly affect the process of translational repression for many proteins in early zygote development. Previous studies have identified IMC-associated proteins that are vital for maintenance of oocyte morphology and virulence, namely IMC1b and IMC1h (or Alv3) [54–56]. Disruption and deletion mutants of the nucleotide cyclase guanylyl cyclase vital for maintenance of ookinete morphology and virulence, previous studies have identified IMC-associated proteins that are repression for many proteins in early zygote development.

In summary, this is the first functional analysis, to our knowledge, of a protein phosphatase in Plasmodium and demonstrates that like the kinases, a phosphatase is also involved in a regulatory pathway in a stage-specific and essential manner.

Materials and Methods

Ethics statement

All animal work has passed an ethical review process and was approved by the United Kingdom Home Office. Work was carried out in accordance with the United Kingdom ‘Animals (Scientific Procedures) Act 1986’ and in compliance with ‘European Directive 86/609/EEC’ for the protection of animals used for experimental purposes. The permit number for the project licence is 40/3344.

Animals

Tuck-Ordinary (TO) (Harlan) outbred mice were used for all experiments except for mosquito “bite-back” infections of mice, where C57/Bl6 mice were used.

Generation of transgenic parasites

The targeting vector for ppkl was constructed using the PBS-DHFR plasmid, which contains polylinker sites flanking a Toxoplasma gondii dhfr/ts expression cassette conveying resistance to pyrimethamine, as described previously [10]. PCR primers P0011P (5’-CCCGGGCCGATGTTTATTTGTTT-GGC-3’) and P0012P (5’-GGGGAAGCTTCAACATCCGTG- TTTCTTTAAATGATCC-3’) were used to generate a 788 bp fragment of 5’ upstream sequence of ppkl from genomic DNA, which was inserted into ApaI and HindIII restriction sites upstream of the dhfr/ts cassette of PBS-DHFR. A 694 bp fragment generated with primers P0013P (5’-CCGGGAATTCGCCCATCAGCCCACC- CAAGAGTCACAAGCG-3’) and P0014P (5’-GGGGTTCTAGA- CGGGGAATTGATGAAATCCGC-3’) from the 3′ flanking region of ppkl was then inserted downstream of the dhfr/ts cassette using EcoRI and XhoI restriction sites. The linear targeting sequence was released using AapI/XhoI. For GFP-tagging by direct Nucleic Acid Labelling and Detection kit (GE Healthcare). Parasites were also visualized on a Zeiss AxioImager M2 (Carl Zeiss, Inc) microscope fitted with an AxioCam ICc1 digital camera (Carl Zeiss, Inc) and analysed by Western blot to confirm GFP expression.

Phenotypic analysis

Infections for phenotype screens were initiated by intraperitoneal injection of infected blood containing 5×10⁶ parasites into mice pre-treated with 0.2 ml of 6 mg/ml phenylhydrazine in PBS injected intraperitoneally to induce reticulocytosis 3 days prior to infection. Asexual stages and gametocyte production were monitored on Giemsa-stained blood films. Exflagellation was examined on day 4–5 post infection. 10 ml of gametocyte-infected blood were obtained from the tail with a
heparinized pipette tip and mixed immediately with 40 μl of ookinete culture medium (RPMI1640 containing 25 mM HEPES, 20% fetal bovine serum, 10 mM sodium bicarbonate, 50 μM xanthurenic acid at pH 7.6). The mixture was placed under a Vaseline-coated cover slip and 15 min later exflagellation centres were counted by phase contrast microscopy in 12–15 fields of view using a 63× objective and 10× ocular lens. Ookinete formation was assessed the next day. 10 μl of infected tail blood were obtained as above, mixed immediately with 40 μl ookinete culture medium, and incubated for 2 h at 20°C to allow completion of gametogenesis and fertilization. Each culture was then diluted with 0.45 ml of ookinete medium and incubated at 20°C for a further 21–24 h to allow ookinete differentiation. Cultures were pelleted for 2 min at 5000 rpm and then incubated with 50 μl of ookinete medium containing Hoechst 33342 DNA dye to a final concentration of 5 μg/ml and a Cy3-conjugated mouse monoclonal antibody 13.1 [16] recognizing the P28 protein on the surface of ookinetes and any undifferentiated macrogametes or zygotes. P28-positive cells were counted with a Zeiss AxiosImager M2 microscope (Carl Zeiss, Inc) fitted with an AxioCam ICc1 digital camera. Ookinete conversion was expressed as the percentage of P28 positive parasites that had differentiated into ookinetes [40]. For mosquito transmission experiments 20–50 Anopheles stephensi SD500 mosquitoes were allowed to feed for 20 min on anaesthetized infected mice whose asexual parasitaemia had reached ~5–7% and were carrying comparable numbers of gametocytes as determined on Giemsa stained blood films. Day 14 post feeding approximately 20 mosquitoes were dissected and oocysts on their mid-guts counted. Oocyst formation was examined by Hoechst 33342 staining for 10–15 min and guts were washed and mounted under Vaseline-rimmed cover slips. Images were recorded using a 63× oil immersion objective on a Zeiss AxiosImager M2 microscope fitted with an AxioCam ICc1 digital camera. Day 21 post feeding another 20 mosquitoes were dissected and their guts and salivary glands crushed separately in a loosely fitting homogenizer to release sporozoites, which were then quantified using a haemocytometer. Due to day-to-day variations in transmission levels, all data were normalized to a matching number of wild type controls analyzed on the same day.

Bioinformatic analysis

Using Arabidopsis thaliana BSU1 sequence (RefSeq: NP_171844) as a seed, the iterative strategy enacted by the ‘jackhammer’ method of HMMER3 [38] was used to search the predicted proteomes of 46 diverse eukaryotes [60] with the addition of the P28-positive cells were counted with a Zeiss AxiosImager M2 microscope (Carl Zeiss, Inc) fitted with an AxioCam ICc1 digital camera. Ookinete conversion was expressed as the percentage of P28 positive parasites that had differentiated into ookinetes [40]. For mosquito transmission experiments 20–50 Anopheles stephensi SD500 mosquitoes were allowed to feed for 20 min on anaesthetized infected mice whose asexual parasitaemia had reached ~5–7% and were carrying comparable numbers of gametocytes as determined on Giemsa stained blood films. Day 14 post feeding approximately 20 mosquitoes were dissected and oocysts on their mid-guts counted. Oocyst formation was examined by Hoechst 33342 staining for 10–15 min and guts were washed and mounted under Vaseline-rimmed cover slips. Images were recorded using a 63× oil immersion objective on a Zeiss AxiosImager M2 microscope fitted with an AxioCam ICc1 digital camera. Day 21 post feeding another 20 mosquitoes were dissected and their guts and salivary glands crushed separately in a loosely fitting homogenizer to release sporozoites, which were then quantified using a haemocytometer. Due to day-to-day variations in transmission levels, all data were normalized to a matching number of wild type controls analyzed on the same day.

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BLAST searches of eukaryotic genomes. The sequences were aligned using ClustalW2 and optimised using CLC Genomics Workbench (CLC bio, Cambridge, MA). After identifying the kelch-domains the kelch-domain coding sequences were realigned using the same program and neighbour-joining bootstrap trees were generated. The phylogenetic trees were drawn using Fig Tree v1.3.1.

Purification of schizonts, gametocytes and ookinetes

Purification of gametocytes was achieved using a protocol modified from [64]. Mice were treated by intra-peritoneal injection of 0.2 ml of phenylhydrazine (6 mg/ml) (Sigma) in PBS to encourage reticulocyte formation four days prior to infection with parasites. Day four post infection (p.i.) mice were treated with sulfadiazine (Sigma) at 20 mg/L in their drinking water for two days to eliminate asexual blood stage parasites. On day six p.i. mice were bled by cardiac puncture into heparin and gametocytes separated from uninfected erythrocytes on a 48% NycoDenz gradient (27.6% w/v NycoDenz in 5 mM Tris-HCl, pH 7.20, 3 mM KCl, 0.3 mM EDTA) in coelenterazine loading buffer (CLB), containing PBS, 20 mM HEPES, 20 mM Glucose, 4 mM sodium bicarbonate, 1 mM EGTA, 0.1% w/v bovine serum albumin, pH 7.25. Gametocytes were harvested from the interface and washed twice in RPMI 1640 ready for activation of gamete formation. Blood cells from day 5 p.i. mice were placed in culture for 24 h at 37°C for schizonts (with rotation at 100 rpm) and 20°C for ookinete production as described above. Schizonts and ookinetes were purified on a 60% and 63% NycoDenz gradient, respectively and harvested from the interface and washed.

Quantitative RT-PCR

Schizonts and gametocytes were purified as described above and frozen in Trizol (Sigma) prior to RNA extraction. Asexual blood parasites were extracted as for gametocytes above but on day four p.i. with very low gametocytaemia and no sulfadiazine treatment. RNA was isolated according to manufacturer’s instructions. Isolated RNA was treated with DNase I (Promega) and used in reverse transcription reactions (SuperScript III Reverse Transcription kit, Invitrogen) from 500 ng of total RNA.

Gene expression was quantified by SYBR green PCR using Fast mastermix on an ABI 7500 QPCR System (Applied Biosystems). Primers were designed using the PerlPrimer software program [65] to be 18–22 bp in length, with 30–60% GC content, to amplify a 50–150 bp long and when possible, to bind within 600 bp of the 3’ end of the genes of interest. Primer efficiencies were all between 90–110%, with qRT-PCR resulting in no detectable primer dimers, as determined by dissociation curves. cDNA was diluted 1:20 with DEPC-treated water before use. Reactions consisted of 3.6 μl of diluted cDNA, 5 μl SYBR green fast mastermix (Applied Biosystems), 0.2 μl each of forward and reverse primer and 1 μl of DEPC water. Cycling conditions were: 95°C for 20 s followed by 40 cycles of 95°C, 3 s, and 60°C, 30 s, followed by dissociation curve. Three biological replicates, with three technical replicates from each biological replicate were performed for each assayed gene. Wild-type gene expression was determined using the comparative cycle threshold method [66], whereas relative quantification in mutant lines was determined using the Pfaffl method [67]. Both methods used hsp70 (PBANKA_068190) (forward, 5’-GTATTTATTAATGAACC-CACC-CTC-3’; reverse, 5’-GAAAACATCATAATGACC-CTTG-3’) and arginy1-RNA synthetase (PBANKA_143420) (forward, 5’-TTGATTGGAGATGGATTCG-3’; reverse, 5’-ATCCGCTTTGGCCTTCCAG-3’) as reference genes. ppk1 primers were: forward, 5’- TTCTAAGTACCTTGCAACAA-
Isolation of PPKL-GFP protein and phosphatase assay

Blood aliquots from mice infected with PPKL-GFP and WT parasites were incubated in ookinete medium for 30 min at 20°C and processed as described previously [46]. The resulting parasite pellets were incubated for 30 min at 4°C in lysis buffer (10 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.5 mM EDTA, 0.5% NP-40) supplemented with protease inhibitors (Roche), then the lysates were centrifuged at 20,000 g for 5 min and the supernatants were subjected to immunoprecipitation using GFP-TRAP beads (ChromoTek) according to manufacturer’s instructions. Protein phosphatase activity of the immunoprecipitate was assessed using the SensoLyte MFP Protein Phosphatase Assay Kit (AnaSpec) according to manufacturer’s instructions. Briefly, the GFP-TRAP beads were resuspended and diluted in phosphatase assay buffer (100 mM Tris-HCl pH 7.5, 4 mM DTT, 0.2 mM EDTA, 0.5 mM MnCl₂, 0.4 mg/ml BSA), incubated for 30 min at 37°C in the presence or absence of MFP fluorogenic phosphate substrate, and centrifuged for 2 min at 2700 g. Supernatants were subjected to immunoprecipitation using GFP-TRAP beads (ChromoTek), the immunoprecipitated proteins were then resuspended in Laemmli sample buffer and separated by SDS-PAGE. ³²P-labelled proteins were visualized using a phosphorimager (Molecular Dynamics) and GFP-tagged proteins analysed by Western blot as described above. The relative PPKL-GFP phosphorylation levels in activated gametocytes with respect to schizonts were obtained by taking the normalized ratio between the intensity of the phosphorylation signal from the phosphoimager and the intensity of the GFP immunoreactive signal from the corresponding Western Blot by using the ImageJ software (National Institute of Health).

PPKL in vivo phosphorylation

Blood aliquots from infected mice were incubated overnight, from which schizonts were purified as described previously [46]. Gametocytes were purified and activated for 25 min at 20°C in ookinete medium as described above. Schizonts and activated gametocytes were then washed in phosphate-free Krebs buffer (118 mM NaCl, 4.7 mM KCl, 4.2 mM NaHCO₃, 1.2 mM MgSO₄, 11.7 mM glucose, 10 mM HEPES, 1.3 mM CaCl₂, pH 7.4) and metabolically labelled with 3–5 MBq ³²P-orthophosphate in phosphate-free Krebs buffer for 30 min at 20°C or 37°C for activated gametocytes and schizonts, respectively. After two washes in phosphate-free Krebs buffer, the labelled parasites were lysed for 30 min at 4°C in lysis buffer (10 mM Tris-HCl pH 7.5, 150 mM NaCl, 0.5 mM EDTA, 0.5% NP-40) supplemented with protease and phosphatase inhibitors (Roche), the resulting lysate was centrifuged at 20,000 g for 5 min and the supernatant collected. PPKL-GFP protein was isolated using GFP-TRAP beads (ChromoTek), the immunoprecipitated proteins were then resuspended in Laemmli sample buffer and separated by SDS-PAGE. ³²P-labelled proteins were visualized using a phosphorimager (Molecular Dynamics) and GFP-tagged proteins analysed by Western blot as described above. The relative PPKL-GFP phosphorylation levels in activated gametocytes with respect to schizonts were obtained by taking the normalized ratio between the intensity of the phosphorylation signal from the phosphoimager and the intensity of the GFP immunoreactive signal from the corresponding Western Blot by using the ImageJ software (National Institute of Health).

Metabolic labelling for global phosphorylation profile

Gametocytes from wild type, pbd⁻¹ and nek4⁻ mutant parasites were purified as described above from the blood of infected mice. Purified gametocytes were placed for 60 min, 5.5 h and 23.5 h in ookinete medium at 20°C to activate both male and female gametocytes to form gametes. For metabolic labelling, the parasites were washed once with 1 ml of phosphate-free Kreb’s buffer and resuspended in 500 µl of the same buffer, 20–25 µl ³²P-orthophosphate (7–9.25 MBq) was added to the suspension and parasites incubated at 20°C for 30 min. The labelled parasites were then lysed in lysis buffer (50 mM Tris, 0.5 mM EDTA, 5% β-glycerophosphate, pH 7.6, supplemented with protease/phosphatase inhibitors (Roche) and 1% NP-40). Following incubation on ice for 10 min, the samples were centrifuged for 3 min at 20000 g and the supernatants were collected for further fractionation. Fractionation was carried out on an AKTA chromatographer (Amersham Pharmacia Biotech) using Resource Q (Amersham Pharmacia Biotech) anion-exchange column (matrix volume 1 ml). The proteins were eluted using a linear gradient of 0–1.0 M NaCl.

Western blotting

Western blot analysis was performed on cell lysates prepared by re-suspending parasite pellets in a 1:1 ratio of PBS containing Protease inhibitor (Roche) and Laemmli sample buffer, boiling and separating on a 4%–15% SDS-polyacrylamide gel (BioRad). Samples were subsequently transferred to nitrocellulose membranes (Amersham Biosciences) and immunoblotting performed using the Western Breeze Chemiluminescent Anti-Rabbit kit (Invitrogen) and anti-GFP polyclonal antibody (Invitrogen) at a concentration of 1:1250, according to the manufacturer’s instructions.

Electron microscopy

Samples cultured in ookinete medium as described above were fixed in 4% glutaraldehyde in 0.1 M phosphate buffer and processed for routine electron microscopy as previously described [68]. Briefly, samples were post fixed in osmium tetroxide, treated en bloc with uranyl acetate, dehydrated and embedded in Spurr’s epoxy resin. Thin sections were stained with uranyl acetate, dehydrated and embedded in Spurr’s epoxy resin. Thin sections were stained with uranyl acetate, dehydrated and embedded in Spurr’s epoxy resin. Thin sections were stained with uranyl acetate and osmium tetroxide. The presence of PPKL-GFP in the bead pellets was assessed by Western blot.

Immunofluorescence assay

IFAs were performed on cell lysates from ookinete cultures produced as previously described [46]. Briefly, infected blood from mice with 7–10% gametocytaemia was incubated for 24 h in ookinete medium at 20°C, then ookinete smear slides were prepared and air dried. For various antibodies different procedures were followed. For α-tubulin, slides were fixed in 4% paraformaldehyde in MTBS buffer [46] and the mouse monoclonal α-tubulin (Sigma) was used as the primary antibody (1:1000 dilution). For actin staining the procedure of [21] was followed. Briefly, ookinete were freshly fixed in 4% paraformaldehyde in MTBS with 0.2% Triton, followed by methanol fixation for 5 min and stained with monoclonal Dictyostelium actin-anti-actin antibody (1:1000 dilution). For GAP45 and MTIP, cells were fixed in 4% paraformaldehyde in MTBS buffer and stained with antibodies against GAP45 (1:250) and MTIP (1:250) [69]. For CTRP, air-dried slides were fixed for 5 min in 1% formaldehyde and mouse monoclonal anti-CTRP antibody (1:1000) was used [25]; for anti-SOAP antibodies, slides were fixed in 4% paraformaldehyde and a 1:100 dilution of antibody was used [26]. For mouse monoclonal antibodies 568 AlexaFluor-labelled anti-mouse (Invitrogen) (1:1000) was used as a secondary antibody. For GAP45 and MTIP antibodies, AlexaFluor 466 labelled anti-rabbit (Invitrogen) (1:1000) was used as a secondary antibody.
in running buffer (10 mM Tris, 5 mM EDTA and 20 mM β-glycerolphosphate, pH 7.4). Fractions (1 ml) were collected and analysed further by resolution on SDS-PAGE gels. 32P-labelled proteins were visualised by autoradiography.

Injections of ookinetes into the mosquito haemocoel

Ookinete were cultured using standard methods and harvested by pelleting. A. stephensi mosquitoes were anaesthetised with CO2 and approximately 500 oocinete in 69 μl of complete oocinete medium were injected into the thorax using a microinjector (Drummond, Nanoject II). Bitebacks and dissections for quantification of salivary gland sporozoites were performed 20 days after injection.

Statistical analyses

All statistical analyses were performed using GraphPad Prism (GraphPad Software). For oocinete motility analysis, non-parametric t-tests were used. For relative quantification of qRT-PCR reactions, a pair-wise fixed reallocation randomisation test [67] was used.

Accession numbers

Sequences are derived from Uniprot (http://www.uniprot.org/), P. berghei (Q4Z2N2), P. yoelii (Q7RA97), P. chabaudi (Q4XN17), P. falciparum (Q84K15), P. knowlesi (B3L999), P. vivax (A5K396), T. gondii (B6KK99), T. parva (Q1N2G8), C. maris (B6A994), T. thermophila (Q22BC4), C. reinhardtii (A8HNE3), A. thaliana (Q8L7U5), P. marinus (C5K554), B. bovis T2Bo (A7AT39), C. variabilis (E1ZE2), V. carteri f. nagaresis (D8UHM6), S. moellendorffii (D0RUC3), P. tetraurelia strain d12 (A0CLB3), M. pennula CCMP1545 (C1MQB3), O. lucimarinus CCE9901 (A4S3K6), N. canum (F0VC41), I. multifiliis (G0QV54).

Supporting Information

Figure S1 Bioinformatic analysis of individual PPKL kelch domains in Plasmodium. The P. berghei Kelch repeat protein phosphatase contains five full (A–E, kelch 1–5) and one truncated (F, kelch 6) kelch repeats. The trees were constructed using Kelch phosphatase sequences identified in BLAST searches of eukaryotic genomes. The sequences were aligned using ClustalW2 and optimised using CLC Genomics Workbench. After identifying the 6 kelch-domains the kelch-domain coding sequences were realigned using the same program and neighbour-joining bootstrap trees were generated. The phylogenetic trees were drawn using Fig Tree v1.3.1. (TIF)

Figure S2 Sequence alignment of the protein phosphatase domains of kelch phosphatases from Alveolata and Virdiplanetae. The invariant residues conserved in individual phosphatases are highlighted in grey and metal ion binding sites are marked with solid circle. The amino acids with role in inhibitor binding are highlighted: Inhibitor-2 binding amino acids (1–7, blue squares); microcystin binding amino acids (1–4, orange squares); okadaic acid and microcystin binding amino acids (1–5, green squares); Inhibitor-2, microcystin, okadaic acid binding amino acid (1–2, blue boxes). Conserved insertions identified in Alveolates are marked with black bars (I–IV) above the sequences. A. thaliana and P. berghei type 1 protein phosphatases (PP1) were included in the alignment as markers for the PP1 catalytic subunit. (TIF)

Figure S3 gfp tagging of the endogenous ppkl locus. A. Schematic representation of the gene targeting strategy used for gene tagging the endogenous locus with gfp via single homologous recombination. Primers 1+2 used for diagnostic PCR are indicated, as well as the EcoRI site used for Southern blotting. Probe location used for detection by Southern blotting is indicated. B. Diagnostic PCR confirming successful integration of the tagging sequence. C. Southern blot analysis of EcoRI digested ppkl genomic DNA using the 3′ UTR of the targeting construct as a probe. Band sizes for PPKL-GFP (tag) and wild-type (wt) are indicated. D. Pulse-field gel electrophoresis blot hybridized with Pb 3′ UTR which detects the endogenous chromosome 7 locus and hdp6 of the C-fusion gfp sequence integrated on chromosome 13–14. E. Western blot analysis using an anti-GFP antibody against control wild-type-GFP (wt) and transgenic (tag) oocinete (ook) non-activated gametocytes (NAG) and activated gametocytes (AG) showing bands of expected sizes of 29 kDa for wild-type-GFP and 128 kDa for PPKL-GFP. (TIF)

Figure S4 Deletion of the ppkl gene. A. Schematic representation of the gene targeting strategy used for gene disruption via double homologous recombination. Primers 1–4 used for diagnostic PCR are indicated, as well as the EcoRI digestion site used for Southern blotting. Probe location used for detection by Southern blotting is indicated. B. Diagnostic PCR confirming successful integration of the disruption sequence of ppkl in mutant clone 3 (cl3), clone 9 (cl9) and a third clone in the P. berghei ANKA 2.34 line constitutively expressing GFP (clone 6–cl6 GFP). Primers 1+2 were used to verify successful integration at the correct locus. Primers 3+4 were used to confirm loss of the endogenous gene. C. Southern blot analysis of EcoRI digested clone 3 and 9 genomic DNA using the 3′ UTR of the targeting construct as a probe. Band sizes for ppkl+ clone 3 (cl3), clone 9 (cl9) and wild-type (wt) are indicated. D. Pulse-field gel electrophoresis blot hybridized with Pb 3′ UTR which detects the endogenous chromosome 7 locus and disrupted locus on chromosome 13 in both clones. E. Bar graph showing relative expression of endogenous ppkl in ppkl− mutants using qRT-PCR compared to wild-type. Error bars = ±SEM, n = 3 from three separate experiments in both clone 3 and clone 9. (TIF)

Figure S5 Montage of wild-type and ppkl− parasites. A. Montage of longitudinal sections through the apex of six different wild-type oocinete illustrating the similarity in appearance of the apical structures. Bar = 100 nm. B. Montage of longitudinal sections of the anterior of ppkl− mutants showing the variability of appearances ranging from relatively normal to severe collapse. In all cases the electron dense collar appeared to be reduced in variety of different ppkl− morphologies after in vitro culture in oocinete medium for 24 h. Bar = 1 μm. (TIF)

Video S1 Wild-type oocinete motility. (MOV)

Video S2 ppkl− retort motility. (MOV)

Acknowledgments

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Author Contributions
Conceived and designed the experiments: DSG BP AAH RT. Performed the experiments: DSG BP BW PLC CR DB EMP US LS RT. Analyzed the data: DSG BP BW PLC CR EMP JLG ABT RES AAH RT. Contributed reagents/materials/analysis tools: DSG RT. Wrote the paper: DSG BP AAH RT.

References
1. Graves JD, Krebs EG (1999) Protein phosphorylation and signal transduction. Pharmacol Ther 62: 111–121.
2. Elmore J, Hunter T (2001) Oncogenic kinase signalling. Nature 411: 355–365.
3. Tan CS, Bodenmiller B, Pascaud E, Jovancevic M, Hengartner MO, et al. (2009) Comparative analysis reveals conserved protein phosphorylation networks implicated in multiple diseases. Sci Signal 2: ra39.
4. Barford D, Das AK, Egloff MP (1998) The structure and mechanism of protein kinases: insights into catalysis and regulation. Annu Rev Biophys Biomol Struct 27: 133–164.
5. Gallogo M, Virshup DM (2005) Protein serine/threonine phosphatases: life, death, and sleeping. Curr Opin Cell Biol 17: 197–202.
6. Hanks SK (2003) Genomic analysis of the eukaryotic protein kinase superfamily: a perspective. Genome Biol 4: 111.
7. Hanks SK, Quinn AM (1991) Protein kinase catalytic domain sequence database: identification of conserved features of primary structure and classification of family members. Methods Enzymol 200: 38–62.
8. Kostich M, English J, Madison V, Gheyas F, Wang L, et al. (2002) Human members of the eukaryotic protein kinase family. Genome Biol 3: RESEARCH0043.
9. Solyakov I, Hallberg J, Alam MM, Semblat JP, Dorin-Semblat D, et al. (2011) Global kinase and phospho-proteomic analyses of the human malaria parasite Plasmodium falciparum. Nat Commun 2: 365.
10. Tewari R, Strachisl U, Baten B, Bilde N, Herrve J, et al. (2010) The systematic functional analysis of Plasmodium protein kinases identifies essential regulators of mosquito transmission. Cell Host Microbe 8: 377–387.
11. Murray CJL, Rosenfeld LC, Lim SS, Andrews KG, Foreman KJ, et al. (2012) Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980–2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet 380: 2095–2127.
12. Bannister LH, Sherman IW (2009) Plasmodium. Encyclopedia of Life Sciences (ELS). Chichester: John Wiley & Sons, Ltd.
13. Rangarajan R, Bel AK, Jethwahey D, Malonado P, Dorin D, et al. (2005) A mitogen-activated protein kinase regulates male gametogenesis and transmission of the malaria parasite Plasmodium berghei. EMBO Rep 6: 46–49.
14. Reiniger I, Billker O, Trevari R, Mukhopadhyay A, Fennell C, et al. (2005) A NIMA-related protein kinase is essential for completion of the sexual cycle of malaria parasites. J Biol Chem 280: 31957–31964.
15. Murray CJL, Rosenfeld LC, Lim SS, Andrews KG, Foreman KJ, et al. (2012) Global malaria mortality between 1980 and 2010: a systematic analysis. The Lancet 380: 437–443.
16. Janse CJ, Mons B, Rouwenhorst RJ, Van der Klooster PF, Overdulve JP, et al. (2001) Sexual development of Plasmodium by translational repression. Science 291: 1480–1483.
17. Tewari R, Dorin D, Moon R, Doerig C, Billker O (2005) An atypical mitogen-activated protein kinase controls cytokinesis and flagellar motility during male gamete formation in a malaria parasite. Mol Microbiol 58: 1253–1263.
18. Billker O, Dechamps S, Tewari R, Wenig G, Franke-Fayard B, et al. (2004) Calcium and a calcium-dependent protein kinase regulate gamete formation and mosquito transmission in a malaria parasite. Cell 117: 503–514.
19. Khan SM, Franke-Fayard B, Mir GR, Lasonder E, Janse CJ, et al. (2005) Proteome analysis of separated male and female gametocytes reveals novel sex-specific Plasmodium biology. Cell 121: 675–687.
20. McRoberts L, Taylor CJ, Deng W, Freedman QL, Cummings RM, et al. (2008) Gametogenesis in malaria parasites is mediated by the cGMP-dependent protein kinase. PLoS Biol 6: e139.
21. Moon RW, Taylor CJ, Bex C, Schepers R, Goulding D, et al. (2009) A cyclic GMP-signalling module that regulates gliding motility in a malaria parasite. Plasmod Pathog 5: e1000399.
22. Siden-Kiamos I, Ecker A, Wilkes JM, Doerig C (2008) The protein-phosphatome of the human malaria parasite Plasmodium falciparum contains a large N-terminal extension and five unique inserts in the catalytic domain. Mol Biochem Parasitol 159: 257–255.
23. Sevi R, Tewari R, Ning J, Blagborough AM, Garbom S, et al. (2008) The conserved protein serine/threonine phosphatase 1. Nature 376: 743–753.
24. Liu J, Baker DA (1998) A putative protein serine/threonine phosphatase from Plasmodium falciparum modulates the response to brassinosteroids in Arabidopsis. Genes Dev 12: 448–460.
25. Le Roch KG, Zhou Y, Blair PL, Giragace M, Moch JK, et al. (2003) Discovery of gene function by expression profiling of the malaria parasite life cycle. Science 301: 1530–1538.
26. Hall N, Karras M, Rainé JD, Carlton JM, Koos TW, et al. (2005) A comprehensive survey of the Plasmodium life cycle by genomic, transcriptomic, and proteomic analyses. Science 307: 82–86.
27. Eddy SR (2009) A new generation of homology search tools based on probabilistic inference. Genome Inform 23: 205–211.
28. Goldberg J, Huang HB, Kwon YG, Greengard P, Nairn AC, et al. (1995) Three-dimensional structure of the catalytic subunit of protein serine/threonine phosphatase-1. Nature 376: 743–753.
29. Liu Y, Tewari R, Ning J, Blagborough AM, Garbom S, et al. (2008) The conserved screen identifies six proteins important for malaria development in the mosquito. Mol Microbiol 70: 209–220.
30. Canning LE, Sinden RE (1973) The organization of the oocyte and observations on nuclear division in oocytes of Plasmodium berghei. Parasitology 67: 29–40.
31. Mair GR, Brako JA, Garver LS, Wiegant JC, Hall N, et al. (2006) Regulation of sexual development of Plasmodium by translational repression. Science 313: 667–669.
32. Kim TW, Guan S, Burlingame AL, Wang ZY (2011) The CDG1 kinase mediates brassinosteroid signal transduction from BR1 receptor kinase to BSU1 phosphatase and GSK3-like kinase BIN2, Mol Cell 45: 561–571.
33. Gutteny DS, Ferguson DJ, Poilles AM, Xu Z, Strachil U, et al. (2012) Putative Homologue of CDC20/CDH1 in the Malaria Parasite Is Essential for Male Gamete Development. PLoS Pathog 8: e1002554.
34. Virshup DM, Shenodkar S (2009) From promiscuity to precision: protein phosphatases get a makeover. Mol Cell Biol 31: 537–543.
35. Nacer A, Underhill A, Har M (2008) The microneme proteins CTRP and SOAP are not essential for Plasmodium berghei oocyste to oocyte transformation in vitro in a cell free system. Mol Biol Cell 19: 337–345.
36. Ozalp O, Belkahla Y, Chory J (2006) Brassinosteroid signalling: a paradigm for steroid hormone signaling from the cell surface. Science 314: 1410–1411.
54. Tremp AZ, Dessens JT (2011) Malaria IMC1 membrane skeleton proteins operate autonomously and participate in motility independently of cell shape. J Biol Chem 286: 5383–5391.

55. Tremp AZ, Khater EI, Dessens JT (2008) IMC1b is a putative membrane skeleton protein involved in cell shape, mechanical strength, motility, and infectivity of malaria ookinetes. J Biol Chem 283: 27604–27611.

56. Khater EI, Sinden RE, Dessens JT (2004) A malaria membrane skeletal protein is essential for normal morphogenesis, motility, and infectivity of sporozoites. J Cell Biol 167: 425–433.

57. Hirai M, Arai M, Kawai S, Matsuoka H (2006) PbGCbeta is essential for Plasmodium ookinete motility to invade midgut cell and for successful completion of parasite life cycle in mosquitoes. J Biochem 140: 747–757.

58. Kadota K, Ishino T, Matsuyama T, Chinzei Y, Yuda M (2004) Essential role of membrane-attack protein in malarial transmission to mosquito host. Proc Natl Acad Sci U S A 101: 16310–16315.

59. Janse CJ, Franke-Fayard B, Mair GR, Ramesar J, Thiel C, et al. (2006) High efficiency transfection of Plasmodium berghei facilitates novel selection procedures. Mol Biochem Parasitol 145: 60–70.

60. Wickstead B, Gull K, Richards TA (2010) Patterns of kinesin evolution reveal a complex ancestral eukaryote with a multifunctional cytoskeleton. BMC Evol Biol 10: 110.

61. Wickstead B, Gull K (2007) Dyneins across eukaryotes: a comparative genomic analysis. Traffic 8: 1708–1721.

62. Katoh K, Misasa K, Kuma K, Miyata T (2002) MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. Nucleic Acids Res 30: 3059–3066.

63. Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19: 1572–1574.

64. Beetsma AL, van de Wiel TJ, Sauerwein RW, Eling WM (1998) Plasmodium berghei ANKA: purification of large numbers of infectious gametocytes. Exp Parasitol 88: 69–72.

65. Marshall OJ (2004) PerlPrimer: cross-platform, graphical primer design for standard, bisulphite and real-time PCR. Bioinformatics 20: 2471–2472.

66. Schmittinger TD, Livak KJ (2008) Analyzing real-time PCR data by the comparative C(T) method. Nat Protoc 3: 1101–1108.

67. Pfaffl MW (2001) A new mathematical model for relative quantification in real-time RT-PCR. Nucleic Acids Res 29: e45.

68. Ferguson DJ, Henriquez FL, Kirisits MJ, Muench SP, Prigge ST, et al. (2005) Maternal inheritance and stage-specific variation of the apicoplast in Toxoplasma gondii during development in the intermediate and definitive host. Eukaryot Cell 4: 814–826.

69. Green JL, Rees-Channer RR, Howell SA, Martin SR, Knuepfer E, et al. (2008) The motor complex of Plasmodium falciparum: phosphorylation by a calcium-dependent protein kinase. J Biol Chem 283: 30980–30989.