Research Article

Indirect Regeneration and Assessment of Genetic Fidelity of Acclimated Plantlets by SCoT, ISSR, and RAPD Markers in Rauwolfia tetraphylla L.: An Endangered Medicinal Plant

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1. Introduction

In India, Ayurveda is a traditional medicinal system which has been practiced for thousands of years and remains as one of the most ancient and yet living traditions across the country, which has a sound philosophical and experimental basis [1, 2]. India is also the largest producer of medicinal herbs and hence appropriately called as the Botanical Garden of the World [3]. Some of these medicinal plants have been featured on Indian postage stamps, where the first set of stamps came out in 1997, showing four different medicinal plants: Ghritkumari (Aloe barbadensis), Sarpagandha (Rauwolfia serpentina), Tulsi (Ocimum sanctum) and Haridra (Curcuma longa) [4] followed by another set of four medicinal plants: Brahmi (Bacopa monnieri), Guggulu (Commiphora wightii), Amla (Emblica officinalis), and Ashwagandha (Withania somnifera) as postal stamps were released [5].

Presently, medicinal plants play a very important role in the modern economy of developing countries. In India, the medicinal plant related trade is estimated to be approximately US $ 1 billion per year [6]. Recent trends have indicated further increase in this trade with the herbal cosmetic and...
pharmaceutical industry playing a major role in fulfilling the demand for herbas worldwide. In addition to the international trade, there is a substantial volume of internal trade for medicinal plants in India, which has created a lot of demand to these medicinal herbs and shrubs [7].

Among the different plant families, Apocynaceae has gained lot of medicinal importance, as most of the plant species of this family produce milky sap which has pharmaceutical applications such as for gastrointestinal ailments, malaria, pain and diabetes [8]. Almost every part (roots, stem bark, leaves and latex) of Apocynaceae family members was used for several treatments and disorders like leprosy, dysentery, worms, ulcers, tumours, skin, and liver diseases [9].

*Rauwolfia* is an important genus in Apocynaceae family which has gained much popularity due to the presence of pharmaceutically significant alkaloids with higher concentrations in roots and stem bark. Approximately 130 species of *Rauwolfia* have been identified and are located mostly in tropical and semitropical regions. Among the different species, *R. serpentina* Benth. ex Kurz is indigenous to India [10, 11] and this species is on the verge of extinction [12] due to its overexploitation by ayurvedic practitioners and pharmaceutical industries [13] and Government of India has also restricted the exploitation of this endangered species [14]. In this scenario the attention of industrialists and medicinal practitioners was diverted to find an alternative plant species as a source of pharmaceutical alkaloids.

Among the other species of *Rauwolfia*, the most important is *Rauwolfia tetraphylla* L., which has gained lot of importance as an alternative source of reserpine and other alkaloids [15]. *R. tetraphylla* is native of tropical America and West Indies [16], but now it is naturalized [17] throughout the tropics which includes the moist deciduous forests of Southeast Asia including Burma, Bangladesh, India, Malaysia, and Sri Lanka [18]. The common names are wild snake root plant, devil pepper, be-still tree, American serpent wood, devil root, milk bush etc. It is being cultivated widely as both an ornamental and a source of pharmaceuticals. This species produces different type of pharmaceutically important alkaloids in their stem bark and roots such as reserpine, deserpidine, ajmalicine, ajmaline, canescine, yohimbine, and recinnamine [19, 20].

Due to presence of various commercial indole alkaloids, this plant species was also exploited during the last three decades and presently it is in endangered status [21]. Propagation of *R. tetraphylla* through vegetative cuttings and seed material is limited (5-10%) due to its poor rooting ability and due to presence of phenolic compounds in seed [22]. In view of the above facts about the mass exploitation of this plant from natural resources and due to its restricted cultivation, there is an urgent need to conserve this medicinally important plant species by proper conservation strategy. Hence, the present study is undertaken with the objective of mass propagation of *R. tetraphylla* through the standardized indirect regeneration protocol by utilizing leaf and stem based callus and to confirm the genetic fidelity of acclimated plants by using SCoT, ISSR, and RAPD primer based PCR analysis.

Even though there are some reports about its multiplication through nodal, shoot tip cultures [23] and *in vitro* flowering [24], but there is scanty information about its regeneration through callus based indirect regeneration protocols. In our previous report, we have used single type of cytokinin (TDZ) in regenerating the low frequency of multiple shoots from leaf and stem based callus [25]. Now, in this study we are reporting a protocol for its conservation through high frequency of plantlet production by using combination of cytokinins amended media.

## 2. Materials and Methods

In this study, most of materials and methods were followed as per the report on indirect regeneration studies in a *Morus* species [26].

### 2.1. Plant Material and Explant Source.

Two-year-old *R. tetraphylla* plants available in the medicinal arboretum, Department of Biotechnology, Kakatiya University, were used as parent plant material in this *in vitro* propagation studies. Fully expanded leaf of about 7 cm in length and 4 cm in breadth, stem (internodal segments), and root parts were collected during the second week of June and were used as explants for the study.

### 2.2. Murashige and Skoog Media Preparation.

MS medium [27] was prepared by using sucrose (3%) and appropriate quantities of vitamins, iron, macronutrients, and micronutrients from stock solutions. Before dispensing the prepared medium to culture tubes, required concentrations and combinations of plant growth regulators were added to it from the hormonal stock solutions. The media was solidified with 0.8% agar and pH was adjusted to 5.6 with 1 or 2 drops of 0.1 N HCl or NaOH solution before autoclaving at 121°C for 15 minutes.

### 2.3. Aseptic Techniques for Sterile Transfer.

All operations including transfer and sterile filtration were made under laminar airflow cabinet. Prior to operation, the laminar airflow bench was swabbed with 70% alcohol to maintain maximum sterility. The laminar airflow was switched on and kept on UV mode for 30 minutes before using it. The scalpel, forceps and spatula were sterilized by flaming with alcohol before using the laminar airflow cabinet.

### 2.4. Explant Surface Sterilization.

Leaf, stem (internodal), and root explants of *R. tetraphylla* were collected from the medicinal arboretum after washing under running tap water for 2 minutes. Then different protocols were followed for the sterilization of leaf, stem and root explants, as leaf explants are more delicate and sensitive in nature to sterilants and due to greater degree of fungal count on stem and root explants. Leaf explants were treated with Tween-20 solution for 1 minute followed by 60% ethanol for 2 minutes and in 0.1% mercuric chloride (HgCl<sub>2</sub>) solution for 2 minutes. Stem and root explants were treated initially with 0.5% bavistin for 10 minutes, followed by Tween-20 solution for 2 minutes and with 0.1% mercuric chloride solution for 4 minutes.
minutes. After every treatment explants were rinsed in sterile distilled water for 2 minutes to remove the adhering chemical sterilants.

2.5. Inoculation and Incubation. Surface sterilized leaf, stem and root explants were blotted dry on sterile tissue paper and inoculated onto MS medium under aseptic conditions of laminar air flow cabinet with the help of sterile forceps and culture containers were immediately closed. During inoculation, the test tubes and flasks containing solid medium were held horizontally; this strongly reduces the contaminations. Culture tubes with inoculated explants were kept in culture room under controlled conditions of 16/8 hours of photoperiod with 3000 lux intensity light at 27°C. For each treatment, 25 replicates were used and each experiment was repeated thrice.

2.6. Callus Induction. For callus induction, fully expanded leaf, stem (inter nodal), and root explants of *R. tetraphylla* (two years old) were inoculated on to MS medium supplemented with 2,4-D or NAA or Kn (1.0-5.0 mg/L). Data on callus induction from three replicates were recorded at the end of 3 weeks of culture. Each replicate consists of 25 cultures.

2.7. Determination of Fresh and Dry Weight of Callus. For determination of fresh and dry weight of callus, 10 identical culture tubes were used. The mean value of each callus was determined. After determining the callus weight, they were dried in an oven at 60°C for 1 hour and average dry weight of the callus was determined. The average fresh and dry weight of the callus was subtracted from the respective callus culture to find out the average gain in fresh and dry weight of the callus. Fresh and dry weights of the callus were noted with a gap of 22 days.

2.8. Shoot Regeneration. In vitro shoot regeneration was carried from the friable callus by using individual and combinations of cytokinins (BAP and TDZ) supplemented MS media. While preparing combinational cytokinin media, BAP of 2 mg/L was kept constant across all the combinations with altering concentrations of TDZ in low amounts (0.1-1.0 mg/L). Data on number shoot bud regeneration from friable callus in three replicates was recorded at the end of 2nd week of culture. Each replicate of regeneration studies consists of 10 cultures.

2.9. Rooting of the Regenerated Shoots. The regenerated shoots were separated from the culture tubes and transferred onto MS medium containing various concentrations (0.5–2.0 mg/L) of individual (IAA or IBA) as well as combination (IAA+IBA) of auxins. Data on root induction from shootlets of three replicates was recorded at the end of 2nd week of culture. Each replicate consists of 10 cultures.

2.10. Hardening and Acclimatization of Rooted Plantlets. Rooted plantlets of *R. tetraphylla* were separated carefully from the culture tubes along with the media and the adhered medium was removed gently by washing under running tap water and then hardened in a plastic pot or plastic cup by using 2:1 ratio of sterile garden soil and sand.

2.11. DNA Isolation and Genetic Fidelity Analysis. Genomic DNA was isolated from acclimated plantlets of *R. tetraphylla* (seven plants were selected randomly, four from leaf callus based and three from stem callus based regenerants) from field along with the mother plant. Fresh leaf tissue (500 mg) of both mother plant and *in vitro* regenerated plants was used as raw material for DNA isolation by following modified cetyl trimethyl ammonium bromide (CTAB) method [28, 29]. The quality parameters of DNA were checked by spectrometric and electrophoretic methods.

For testing the genetic homogeneity of regenerated plantlets with that of mother plant, three types of primers (RAPD, ISSR, and SCoT) were used. A total of 30 primers were used, *i.e.*, SCoT1 (10 primers), ISSR (10 primers), and RAPD (10 primers) for genetic fidelity studies. PCR amplifications were performed using a total volume of 50 μL containing 100 ng DNA, 1X PCR master mix (GCC biotech), and 10 pmole primer. The amplification reaction was carried out in a thermal cycler (Eppendorf, Germany) for 30 cycles with an initial denaturation at 94°C for 5 minutes, followed by denaturation at 94°C for 30 seconds, and annealing was carried at 50°C for 45 seconds for SCoT and ISSR and at 37°C for 45 seconds for RAPD primer based analysis, followed by extension at 72°C for 2 minutes and final extension at 72°C for 5 minutes. After completion of cycles, amplified products were allowed to cool down at a holding temperature of 4°C. PCR products were then separated on 1% agarose gel electrophoresis using TAE (Tris acetic acid EDTA) buffer. The size of amplicons was estimated using 1kb DNA ladder (Thermo Scientific). All amplification reactions with RAPD, ISSR, and SCoT primers were repeated at least three times to check the reproducibility. The gels were photographed using gel documentation system and only clear and scorable DNA bands were considered for analysis.

2.12. Statistical Analysis. The data obtained in this research study was statistically analyzed using one-way ANOVA in SPSS Version 17 (SPSS Inc., Chicago, USA) and means were compared using Tukey’s tests at the 5% level of significance. All means are presented with ± Standard Error (SE).

3. Results and Discussion

3.1. Callus Induction. In this study leaf, stem, and root explants of *R. tetraphylla* cultured on MS basal medium did not produce any callus, but when cultured on MS medium supplemented with various concentrations of plant growth regulators (2,4-D, NAA, and Kn), copious amounts of friable callus were produced, after 4 weeks of culture (Figures 1(a)–1(d)). The produced callus is of different types, such as friable white, friable pale brown, and compact brown callus. At lower concentrations (1-2 mg/L) of 2,4-D, NAA, and Kn, all the explants have produced friable white callus (Figures 1(a) and 1(b)) whereas friable green callus (Figure 1(c)) to
Friable brown callus (Figure 1(d)) was produced from the optimum (3 mg/L) and higher concentrations (4-5 mg/L) of plant growth regulators (2,4-D, NAA, and Kn), respectively.

On 2,4-D supplemented MS media, maximum amount of mean fresh and dry weight (mg) of callus (378.7±0.67, 19.2±0.15), (241.5±0.34, 10.3±0.08), and (323.8±0.36, 15.5±0.12) was observed in leaf, stem, and root explants cultured on 5, 4, and 3 mg/L of concentrations, respectively, after 3 weeks of culture (Table 1). On NAA supplemented media, maximum amount of mean fresh and dry weight (mg) of callus (313.5±0.23, 19.5±0.27), (412.8±0.49, 25.1±0.15), and (277.4±0.14, 16.1±0.03) was observed in leaf, stem, and root explants cultured on 2, 6, and 2 mg/L concentrations, respectively, after 3 weeks of culture (Table 2). On Kn supplemented media, maximum amount of mean fresh and dry weight (mg) of callus (349.5±0.46, 22.8±0.19) and (343.2±0.64, 14.4±0.26) was observed in leaf and stem explants cultured on 5 mg/L concentration of Kn, after 3 weeks of culture (Table 3). Root explants have not responded on Kn supplemented media even after 4 weeks of culture.

Though the rate of callus growth was negligible in the first two weeks, the entire explant was covered with healthy callus after 3 weeks. Major portion of callus that developed from leaf, stem, and root was friable creamy white in color after 3 weeks of culture but became brown when cultured for more than six weeks on the same medium, due to the release of phenolic compounds, which is commonly observed in the callus cultures of shrub and tree species [30]. As the primarily induced callus of leaf and stem explants were friable and creamy white in color, in order to make them friable, green, and organogenic, the primary callus of leaf and stem explants of *R. tetraphylla* were subcultured for 3 passages of time (2 months) on the same combination of plant growth regulators.

Anitha and Kumari (2006a) [31] were first to report callus induction from leaf explants of *R. tetraphylla* on MS + 2,4-D (9 μM/L) and the fresh and dry weights of callus were reported as 0.3354 and 0.0826 grams, respectively. Callus induction was also reported from petiole, cotyledonary leaf, and hypocotyl explants of *R. tetraphylla* cultured on MS medium supplemented with 4.52μM of 2,4-D [32]. In our
Table 1: Effect of 2, 4-dichlorophenoxy acetic acid (2, 4-D), on callus induction from leaf, stem (internode), and root explants of *Rauwolfia tetraphylla* L.

| Explant        | MS + 2,4-D (mg/L) | Morphogenetic Response | Callus Mean fresh weight (mg) | Callus Mean dry weight (mg) |
|----------------|------------------|------------------------|-------------------------------|-----------------------------|
| Leaf           |                  |                        |                               |                             |
| 1.0            | Friable white callus | 234.4 ± 0.12^a         | 08.2 ± 0.44^a                 |
| 2.0            | Friable white callus | 255.9 ± 0.23^b         | 09.6 ± 0.24^c                 |
| 3.0            | Friable white callus | 287.8 ± 0.91^c         | 11.4 ± 0.49^d                 |
| 4.0            | Friable white callus | 330.6 ± 0.76^e         | 16.7 ± 0.68^f                 |
| 5.0            | Friable white callus | 378.7 ± 0.67^f         | 19.2 ± 0.15^g                 |
| Stem (Internode)| 1.0                | Friable white callus | 92.6 ± 0.19^h                 |
|                | 2.0                | Friable white callus | 123.5 ± 0.28^i                |
|                | 3.0                | Friable white callus | 185.6 ± 0.36^j                |
|                | 4.0                | Friable white callus | 241.5 ± 0.34^k                |
|                | 5.0                | Compact brown callus  | 232.2 ± 0.24^l                |
| Root           | 1.0                | Friable white callus | 123.6 ± 0.34^m                |
|                | 2.0                | Friable white callus | 256.7 ± 0.12^n                |
|                | 3.0                | Friable pale brown callus with roots | 303.6 ± 0.55^o |
|                | 4.0                | Friable pale brown callus with roots | 291.5 ± 0.18^p |
|                | 5.0                | Pale brown callus    | 242.4 ± 0.23^q                |
|                |                    |                        |                               |                             |
|                |                    |                        | 253.3 ± 0.14^r                |
|                |                    |                        | 255.4 ± 0.40^s                |
|                |                    |                        | 303.6 ± 0.47^t                |
|                |                    |                        | 343.1 ± 0.21^u                |
|                |                    |                        | 412.8 ± 0.49^v                |
|                |                    |                        | 256.5 ± 0.38^w                |
|                |                    |                        | 277.4 ± 0.14^x                |
|                |                    |                        | 262.1 ± 0.21^y                |
|                |                    |                        | 258.4 ± 0.74^z                |
|                |                    |                        | 243.8 ± 0.36^aa               |

Data represents average of three replicates. Each replicate consists of 25 cultures. Data scored at the end of 3 weeks of culture. Mean ± standard error. Mean followed by the same superscript in a column is not significantly different at P=0.05.

Table 2: Effect of naphthalene acetic acid (NAA), on callus induction from leaf, stem (internode), and root explants of *Rauwolfia tetraphylla* L.

| Explant        | MS + NAA (mg/L) | Morphogenetic Response | Callus Mean fresh weight (mg) | Callus Mean dry weight (mg) |
|----------------|-----------------|------------------------|-------------------------------|-----------------------------|
| Leaf           |                  |                        |                               |                             |
| 1.0            | Friable white callus | 311.7 ± 0.31^c         | 19.1 ± 0.23^c                 |
| 2.0            | Friable white callus | 313.5 ± 0.28^d         | 19.5 ± 0.67^b                 |
| 3.0            | Friable pale brown callus with roots | 303.6 ± 0.55^e         | 19.0 ± 0.54^f                 |
| 4.0            | Friable pale brown callus with roots | 291.5 ± 0.18^g         | 17.2 ± 0.33^h                 |
| 5.0            | Pale brown callus    | 242.4 ± 0.23^i         | 15.6 ± 0.80^j                 |
| Stem (Internode)| 1.0                | Friable white callus | 153.3 ± 0.14^k                |
|                | 2.0                | Friable white callus | 255.4 ± 0.40^l                |
|                | 3.0                | Friable white callus | 303.6 ± 0.47^m                |
|                | 4.0                | Pale brown callus    | 343.1 ± 0.21^n                |
|                | 5.0                | Pale brown callus    | 412.8 ± 0.49^o                |
| Root           | 1.0                | White friable callus  | 256.3 ± 0.38^p                |
|                | 2.0                | White friable callus  | 277.4 ± 0.14^q                |
|                | 3.0                | Pale brown callus    | 262.1 ± 0.21^r                |
|                | 4.0                | Brown callus         | 258.4 ± 0.74^s                |
|                | 5.0                | Dark brown callus    | 243.8 ± 0.36^t                |

Data represents average of three replicates. Each replicate consists of 25 cultures. Data scored at the end of 3 weeks of culture. Mean ± standard error. Mean followed by the same superscript in a column is not significantly different at P=0.05.

In our earlier studies, we have used single cytokinin (TDZ) for regeneration of shoot buds from leaf and stem based callus and obtained maximum frequency of shoot regeneration (12.0±0.28) from stem based callus at 2.27 µM/l concentration of TDZ [25]. Now through this research paper we are reporting a standardized protocol for shoot regeneration from leaf, stem and root explants of *Rauwolfia tetraphylla* L.
Table 3: Effect of Kinetin (Kn) on callus induction from leaf, stem (internode), and root explants of *Rauwolfia tetraphylla* L.

| Explant          | MS + Kn (mg/L) | Morphogenetic Response | Callus | Mean fresh weight (mg) | Mean dry weight (mg) |
|------------------|----------------|------------------------|--------|------------------------|---------------------|
|                  |                |                        |        |                        |                     |
| Leaf             | 1.0            | No response            |        |                        |                     |
|                  | 2.0            | Friable white callus   | 201.4  | 11.1 ± 0.1 f           |                     |
|                  | 3.0            | Friable green callus   | 226.3  | 13.5 ± 0.22 d          |                     |
|                  | 4.0            | Pale brown callus      | 334.6  | 18.5 ± 0.68 b          |                     |
|                  | 5.0            | Pale brown callus      | 349.5  | 22.8 ± 0.89 a          |                     |
| Stem (Internode) | 1.0            | Friable white callus   | 218.6  | 3.6 ± 0.19 i           |                     |
|                  | 2.0            | Friable White callus   | 233.3  | 4.2 ± 0.17 h           |                     |
|                  | 3.0            | Friable White callus   | 264.9  | 7.8 ± 0.23 g           |                     |
|                  | 4.0            | Pale brown callus      | 309.4  | 11.5 ± 0.61 e          |                     |
|                  | 5.0            | Pale brown callus      | 343.2  | 14.4 ± 0.26 c          |                     |
| Root             | 1.0            | Single shoot bud       |        |                        |                     |
|                  | 2.0            | Two shoot buds         |        |                        |                     |
|                  | 3.0            | Multiple shoot buds    |        |                        |                     |
|                  | 4.0            | Multiple shoot buds    |        |                        |                     |
|                  | 5.0            | Multiple shoot buds    |        |                        |                     |

Data represents average of three replicates. Each replicate consists of 25 cultures. Data scored at the end of 3 weeks of culture. Mean ± standard error. Mean followed by the same superscript in a column is not significantly different at *P* = 0.05.

![Figure 2: Effect of TDZ (Thidiazuron) on mean number of shoot buds induced in leaf and stem calluses of *R. tetraphylla*. Bars represent mean ± SE and bars denoted by the same letter are not significantly different by Tukey’s test used at *P* = 5% probability.](image2)

![Figure 3: Effect of BAP (6-benzyl aminopurine) on number of shoot buds induced in leaf callus and stem callus of *R. tetraphylla*. Bars represent mean ± SE and bars denoted by the same letter are not significantly different by Tukey’s test used at *P* = 5% probability.](image3)

with much higher frequency of shoot regeneration from the friable callus of leaf and stem explants on MS media supplemented with individual (Figures 2 and 3) and combination of cytokins (TDZ and BAP) (Figure 4).

In this study, three times subcultured callus of leaf and stem explants of *R. tetraphylla* has shown good response of shoot regeneration with maximum number of shoot buds on combinational media (TDZ+BAP) rather than individually supplemented media (TDZ or BAP) (Figure 4). On MS media amended with individual TDZ plant growth regulators, regeneration of shoot buds with a frequency of 3.2 ± 0.1 and 12.2 ± 0.28 was obtained at 0.25 mg/l and 0.50 mg/l concentration from leaf and stem based callus, respectively (Figure 5(a)). On BAP amended media, regeneration of shoot buds with a frequency of 10.0 ± 0.42 and 9.0 ± 0.34 was obtained at 4.0 mg/l concentration from leaf and stem based callus, respectively. Overall, the maximum number of shoot buds with high frequency of regeneration (25 ± 0.25 and 20 ± 0.76) was obtained from leaf and stem calluses, respectively, when cultured on MS media supplemented with TDZ (0.25 mg/l) + BAP (2 mg/l), after 4 weeks of culture (Table 4 and Figure 6(a)).

Thidiazuron (TDZ) (N-phenyl-N’-1,2,3-thiadiazol-5-ylurea) is a thidiazole-substituted phenylurea, which was reported to possess more cytokinin activity as compared to phenyl urea derivatives and other active adenine types of cytokins [34]. Thidiazuron was also reported to possess actions of other plant growth regulators such as auxins, gibberellins, and ethylene [35]. Thidiazuron (TDZ) was used
by several researchers in regeneration studies of medicinal plants with higher frequency of shoot let production from friable callus [36, 37]. In our earlier studies we have induced low frequency of multiple shoots from stem callus of *R. tetraphylla* on TDZ amended media [25].

It was reported that thidiazuron in combination with other plant growth regulators induced callus [38], somatic embryos [39], and shoot regeneration from friable callus [40]. Hence, in this study we have used TDZ in combination with BAP for the indirection regeneration of *R. tetraphylla*. In our study, combination of TDZ (0.25mg/L) + BAP (2mg/L) has resulted in inducing maximum number of shoot buds (25±0.25 and 20±0.76) from leaf and stem calluses, respectively, which is much higher compared to our earlier reports. GDZ+BAP combinations have been reported for successful multiple and rapid shoot regeneration of various species, e.g., *Ancistrocladus heyneanus* [41], *Luffa cylindrica* L. [42], *Rauwolfia tetraphylla* L. [25], *Oryza sativa* [43], *Mirabilis jalapa* L. [44], *Achyranthes aspera* [40], and *Morus* Sps. cv. PPR-1 [26].

As the regenerated shoots are minute in the form of micro shoots, they were transferred onto MS basal media (Figures 5(b) and 6(b)) and cultured for 2 weeks for elongation of micro shoots into shoot lets (Figures 5(c), 5(d), 5(e), and 6(c)).

### 3.3. Rooting of In Vitro Regenerated Shoots

The regenerated shoots of *R. tetraphylla* L. were rooted successfully with maximum rooting frequency of 98.0±1.2 on combinational media of auxins with IAA (1.0 mg/L) and IBA (1.0 mg/L), after 2 weeks of culture, and well developed plantlet with extensive rooting was observed after 4 weeks of culture (Figures 5(f), 6(d), and 6(e)). On individual auxins supplemented media, rooting was induced with a frequency of 72.4±5.6 and 90.6±6.2 from IAA (0.5 mg/L) and IBA (1.0 mg/L), respectively (Table 5 and Figure 7). Rapid growth of plant lets

### Table 4: Effect of TDZ and BAP on shoot bud differentiation from 3 times subcultured leaf, stem, and root callus of *Rauwolfia tetraphylla* L.

| Three times sub-cultured callus | TDZ (mg/L) | BAP (mg/L) | No. of shoot buds |
|---------------------------------|------------|------------|-------------------|
| Leaf                            |            |            |                   |
| 0.10                            | 2.00       | 4 ± 0.12³  |
| 0.25                            | 2.00       | 25 ± 0.25³ |
| 0.50                            | 2.00       | 10 ± 0.17³ |
| 0.75                            | 2.00       | 8 ± 0.11³  |
| 1.00                            | 2.00       | 2 ± 0.15³  |
| Stem                            |            |            |                   |
| 0.10                            | 2.00       | 4 ± 0.43³  |
| 0.25                            | 2.00       | 20 ± 0.76³ |
| 0.50                            | 2.00       | 14 ± 0.13³ |
| 0.75                            | 2.00       | 4 ± 0.28³  |
| 1.00                            | 2.00       | 2 ± 0.26³  |

Data represents average of three replicates. Each replicate consists of 10 cultures. Data scored at the end of 2 weeks of culture. Mean ± standard error. Mean followed by the same superscript in a column is not significantly different at *P*=0.05.

### Table 5: Effect of auxins (IAA and IBA) on root induction from *in vitro* shoot lets of *Rauwolfia tetraphylla* L.

| IAA (mg/L) | IBA (mg/L) | Rooting (%) | No of roots/shoot |
|------------|------------|-------------|-------------------|
| 0.5        | -          | 72.4 ± 5.6³ | 3.54 ± 0.62³     |
| 1.0        | -          | 52.1 ± 6.4³ | 3.12 ± 0.24³     |
| 1.5        | -          | 48.3 ± 3.8³ | 2.18 ± 0.28³     |
| 2.0        | -          | 32.7 ± 2.2³ | 2.08 ± 0.13³     |
| -          | 0.5        | 82.2 ± 5.7³ | 5.16 ± 0.44³     |
| -          | 1.0        | 90.6 ± 6.2³ | 7.40 ± 0.72³     |
| -          | 1.5        | 60.3 ± 3.4³ | 3.22 ± 0.84³     |
| -          | 2.0        | 44.8 ± 2.8³ | 4.86 ± 0.32³     |
| 0.5        | 0.5        | 84.6 ± 4.6³ | 6.04 ± 0.54³     |
| 0.5        | 1.0        | 92.4 ± 5.8³ | 6.30 ± 0.28³     |
| 1.0        | 0.5        | 69.8 ± 4.2³ | 1.44 ± 0.12³     |
| 1.0        | 1.0        | 98.0 ± 1.2³ | 8.32 ± 0.84³     |

Data represents average of three replicates. Each replicate consists of 10 cultures. Data scored at the end of 2 weeks of culture. Mean ± standard error. Mean followed by the same superscript in a column is not significantly different at *P*=0.05.
Figure 5: In vitro plant regeneration from leaf callus (3 times subcultured) in R. tetraphylla. (a) Shoot bud induction (represented with arrows) in leaf callus cultured on MS + TDZ (0.5 mg/L). (b) Transfer of individual micro shoots on MS basal media. (c) Single shoot elongation on MS basal medium. (d) Increase in shoot length up to 1 cm on MS basal medium after 1 week of culture. (e) Increase in shoot length up to 2 cm on MS basal medium after 2 weeks of culture. (f) Root induction in single shoot cultured on MS + IAA (1mg/L) + IBA (1 mg/L). (g) Complete plantlet transferred to plastic pot containing sterile soil for hardening.
Figure 6: In vitro plant regeneration from stem callus (3 times sub cultured) in R. tetraphylla. (a) Shoot bud induction from stem callus cultured on MS + TDZ (0.25 mg/L) + BAP (2 mg/L). (b) Multiple shoot buds transferred onto MS basal media. (c) Elongation of multiple shoot buds into small shoot lets on MS basal media. (d) Root induction in single shoot cultured on MS + IAA (1 mg/L) + IBA (1 mg/L). (e) Root induction in single shoot cultured on MS + IAA (1 mg/L) + IBA (1 mg/L). (f) Complete plantlet transferred to plastic cup containing sterile soil for hardening. (g) Field transfer (2 months old). (h) Acclimatized plant (24 months old).

was observed once the primary roots were initiated from the ends of shoot lets.

3.4. Plant Acclimatization to Ex Vitro Conditions. In vitro raised plantlets of R. tetraphylla were separated carefully from the media and hardened in a plastic pot (Figures 5(g) and 6(f)) using 2:1 ratio of sterile garden soil and sand. Initially the hardened plantlets were kept in hardened pots and cups for 4-6 weeks under controlled conditions by irrigating with broth solution of half strength MS salts and then transferred and acclimatized to field conditions (Figures 6(g) and 6(h)) with 86% survival. With garden soil and sand, similar types of results were obtained in acclimatizing the in vitro raised Morus Sps [45, 46].
### Table 6: List of primers, their sequence, number and size of DNA bands generated by SCoT.

| # | Primer code | Primer Sequence | Number of Bands | Band Length (in bp) |
|---|-------------|-----------------|-----------------|---------------------|
| 1 | S1          | CAACAATGGCTACCACCA | 2               | 900-1200            |
| 2 | S2          | CAACAATGGCTACCACC | 2               | 1000-1500           |
| 3 | S3          | CAACAATGGCTACCACCG | 3               | 1000-1400           |
| 4 | S4          | CAACAATGGCTACACCT | 4               | 500-2000            |
| 5 | S5          | CAACAATGGCTACCAGGA | 4               | 500-2000            |
| 6 | S6          | CAACAATGGCTACCACGC | 4               | 500-2000            |
| 7 | S7          | CAACAATGGCTACCACGG | 3               | 500-2000            |
| 8 | S8          | ACGACATGGCGACCACCA | 3               | 400-2100            |
| 9 | S9          | ACCATGGCTACCACGAC | 2               | 500-1200            |
| 10 | S10       | CCATGGCTACCACCCGAG | 3               | 400-1200            |

### Table 7: List of primers, their sequence, number and size of DNA bands generated by ISSR.

| # | Primer code | Primer Sequence | Number of Bands | Band Length (in bp) |
|---|-------------|-----------------|-----------------|---------------------|
| 1 | R1          | AGAGAGAGAGAGAGAGG | 5               | 500-2000            |
| 2 | R2          | AGAGAGAGAGAGAGAYT | 5               | 750-2500            |
| 3 | R3          | AGAGAGAGAGAGAGATC | 4               | 700-1700            |
| 4 | R4          | CTCTCTCTCTCTCTCTRA  | 4               | 500-2000            |
| 5 | R5          | GAGAGAGAGAGAGATGAT | 6               | 400-2000            |
| 6 | R6          | GAGAGAGAGAGAGAGAC | 4               | 500-2000            |
| 7 | R7          | ACACACACACACACACT | 3               | 750-1500            |
| 8 | R8          | CTCTCTCTCTCTCTCTCT | 3               | 600-1100            |
| 9 | R9          | GAGAGAGAGAGAGAYTG  | 6               | 300-2000            |
| 10 | R10        | GAGAGAGAGAGAGAGAYG | 6               | 600-2500            |

3.5. Assessment of Genetic Stability Using SCoT, ISSR, and RAPD Markers. Genetic fidelity of micropropagated and acclimated plantlets of *R. tetraphylla* was assessed by using three generations of markers, i.e., RAPD, ISSR, and SCoT. For better analysis of genetic homogeneity and for discarding the possibility of somaclonal variants, it was always recommended to use more than one marker while carrying out the genetic stability studies [47].

Analysis with SCoT primers (10 primers) has produced a total of 30 monomorphic bands and 3 as the average number of bands per primer with band size (in bp) ranging from 400 to 2000 (Table 6). Among the 10 SCoT primers, three primers, namely, S4, S5, and S6, have produced the highest number of bands (4 bands) and three primers, namely, S1, S2, and S9, have produced the least number of bands (2 bands) which were monomorphic to the mother plant. Amplification with 10 ISSR primers has yielded 45 bands in total with 4 as the average number of bands per primer and band size (in bp) ranging from 300 to 2500 (Table 7). ISSR primer, R10, has given good amplification and produced 6 DNA bands which are clear, distinct, and scorable, in the range (in bp) of 600-2500. Similarly, with 10 RAPD primers a total of 42 bands with
Table 8: List of primers, their sequence, number and size of DNA bands generated by RAPD.

| #  | Primer code | Primer Sequence | Number of Bands | Band Length (in bp) |
|----|-------------|-----------------|-----------------|---------------------|
| 1  | OPA1        | CAGGGCCCTTC     | 3               | 300-1500            |
| 2  | OPA2        | TGCGGAGCTG      | 5               | 300-1000            |
| 3  | OPA3        | AGTCAGCCAC      | 6               | 300-2200            |
| 4  | OPA4        | AATCGGGGCTG     | 3               | 400-1200            |
| 5  | OPA5        | AGGGGTCTTG      | 4               | 500-1600            |
| 6  | OPA6        | TGGGGCCTAA      | 6               | 400-2000            |
| 7  | OPA7        | GGATGACCTC      | 5               | 300-1800            |
| 8  | OPA8        | TGGGGCCTCAA     | 4               | 400-1900            |
| 9  | OPA9        | CCACGACCTT      | 3               | 600-1100            |
| 10 | OPA10       | GACTGCACAC      | 3               | 600-1200            |

Figure 8: SCoT profiles of mother plant (Lane 2) and tissue culture raised plantlets (Lanes 3-9) of R. tetraphylla using S2-SCoT primer. Lanes 1 and 10: DNA ladder of 1Kbp size. Lane 2: DNA banding pattern of mother plant. Lanes 3-6: DNA banding pattern of acclimated plants which were raised from leaf explants. Lanes 7-9: DNA banding pattern of acclimated plants which were raised from stem explants.

Table 8: List of primers, their sequence, number and size of DNA bands generated by RAPD.

4 as the average number of bands per primer and band size (in bp) ranging from 300 to 2000 (Table 8) were obtained. RAPD primer, OPA3, has given good amplification and produced 6 DNA bands which are clear, distinct, and scorable, in the range (in bp) of 300-2200.

Even though we have carried amplification and produced monomorphic bands with RAPD primers, we cannot ensure the reproducibility of results with RAPD primers, hence in our study along with RAPD primers we have also used other advanced primers, i.e., ISSR and SCoT, which are reliable and reproducible type in assessing the genetic stability of regenerants compared to RAPD [48]. All amplification reactions with RAPD, ISSR, and SCoT primers were repeated three times to check the reproducibility and results were, i.e., reproducible type with all the markers (R10-ISSR, S2-SCoT, and OPA3-RAPD).

Among the various markers, nowadays SCoT markers are gaining much attention due to their superiority and better resolvability over other markers [49, 50]. SCoT markers can be used effectively for both genetic diversity and similarity studies [51, 52]. SCoT markers were already reported for assessing the genetic stability of several plant species such as Cleome gynandra [53], Pittosporum eriocarpum [48], Albizia julibrissin [54], and Simmondsia chinensis [55]. Similarly, ISSR primer based assessment of regenerants was reported in Tylophora indica [56], Simmondsia chinensis [57], Rauwolfia tetraphylla L. [58], Spilanthes calva [59], Cucumis melo L. [60], Cornus alba [61], and Zea mays [62].

In our study, DNA bands produced by amplification with RAPD, ISSR, and SCoT markers were monomorphic across the micropropagated plantlets and the mother plant (Figures 8, 9, and 10), which confirms the true-to-type and genetically stable nature of acclimated plantlets of R. tetraphylla. RAPD and ISSR primers were also used by earlier researchers for confirming the genetic homogeneity of regenerants in R. tetraphylla [25, 63, 64]. But, this is the first report of using SCoT primers for the assessment of genetic stability of regenerants in R. tetraphylla.
4. Conclusion

An efficient and repeatable protocol was developed for conservation of economically important plant species, *i.e.*, *R. tetraphylla* L., by high frequency of indirect shoot regeneration from leaf and stem based callus. Further, the genetic homogeneity of *in vitro* regenerated plantlets was confirmed by using SCoT, ISSR, and RAPD markers. This standardized protocol could be utilized for the mass propagation of true-to-type genotype of this endangered medicinal plant.

**Data Availability**

The data used to support the findings of this study are included within the article and can be freely available to authors with proper citation in their research work.

**Conflicts of Interest**

The authors declare that there are no conflicts of interest in this research study.

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