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Editing independent effects of ADARs on the miRNA/siRNA pathways

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Adenosine deaminases acting on RNA (ADARs) are best known for altering the coding sequences of mRNA through RNA editing, as in the GluR-B Q/R site. ADARs have also been shown to affect RNA interference (RNAi) and microRNA processing by deamination of specific adenosines to inosine. Here, we show that ADAR proteins can affect RNA processing independently of their enzymatic activity. We show that ADAR2 can modulate the processing of mir-376a2 independently of catalytic RNA editing activity. In addition, in a Drosophila assay for RNAi deaminase-inactive ADAR1 inhibits RNAi through the siRNA pathway. These results imply that ADAR1 and ADAR2 have biological functions as RNA-binding proteins that extend beyond editing per se and that even genomically encoded ADARs that are catalytically inactive may have such functions.

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Introduction

The most common RNA editing event in human mRNA is deamination of adenosine to inosine. Proteins of the adenosine deaminases acting on RNA (ADAR) family catalyse this editing event. ADAR activity has been implicated in cancer, development, innate immunity and RNA interference (RNAi) (Patterson et al., 1995; George and Samuel, 1999; Beghini et al., 2000; Maas et al., 2001; Scadden and Smith, 2001; Hartwig et al., 2004; Luciano et al., 2004; Scadden, 2005; Scadden and O’Connell, 2005; Yang et al., 2005, 2006; Blow et al., 2006; Takahashi et al., 2006; Ohman, 2007; Paz et al., 2007; Kawahara et al., 2007a, b; Cenci et al., 2008). In humans, the two most studied proteins of the ADAR family are ADAR1 and ADAR2. The ADAR1−/− mice are embryonic lethal with liver disintegration and a deficiency in erythropoiesis (Wang et al., 2000, 2004; Hartner et al., 2004). ADAR2 has been shown to be necessary for the editing of the GluR-B transcript at the critical Q/R site, which limits calcium entry through AMPA receptors. ADAR2−/− mice die around day p20 with seizures (Higuchi et al., 2000).

ADAR1 and ADAR2 have similar domain structures (Figure 1D) but mainly display distinct editing site specificities—although their editing activities do overlap at some sites. There are two major isoforms of ADAR1 because of alternative promoter usage, the longer shuttling p150 isoform and the shorter nuclear p110 isoform. ADAR1 p150 is up-regulated by interferon and can compete with PKR for double-stranded (ds)RNA. At the amino terminus ADAR1 p150 has two Z-DNA-binding domains and a nuclear export signal, which allows it to shuttle in and out of the nucleus, however it is predominantly found in the cytoplasm. Both isoforms have three dsRNA-binding domains, the third one overlaps the catalytic deaminase domain. It is localized to the nucleus by an NLS variant that produces a full-length ADAR1 protein. On the other hand, is most highly expressed in the central nervous system, has two dsRNA-binding domains and a catalytic deaminase domain that is at the carboxy terminus (Strehblow et al., 2002). ADAR1 is widely expressed throughout the body with the exception of skeletal muscle. ADAR2, on the other hand, is most highly expressed in the central nervous system, has two dsRNA-binding domains and a carboxy-terminal deaminase domain. It is localized to the nucleus by an NLS present in the N-terminal region (Desterro et al., 2003; Wong et al., 2003). To what extent each of the different domains of ADAR1 and ADAR2 contributes to their biological activities is an open question. Two other members of the ADAR family in vertebrates are catalytically inactive, ADAR3 that is brain specific and TENR that is expressed in the testis (Melcher et al., 1996; Connolly et al., 2005). Drosophila has a single Adar gene encoding a deaminase with the same domain structure as ADAR2. Adar mutant flies are viable with severe locomotion defects and loss of RNA editing in transcripts expressed in the CNS (Palladino et al., 2000a, b).

Mutations in human ADAR1 are associated with Dyschromatosis Symmetrica Hereditaria (DSH1) (Tojo et al., 2006), an autosomal dominant hyperpigmentation of the hands and feet occurring in Chinese and Japanese families. The majority of DSH1 mutations are associated with single allele truncations of ADAR1 and the dominant phenotype seems to be due to haploinsufficiency for ADAR1. The effect may arise in migrating neural crest cells that develop into melanocytes in the affected skin areas. The point mutant ADAR1 G1007R is an unusual example of a DSH1-associated variant that produces a full-length ADAR1 protein. On the basis of the known crystal structure of the ADAR2 deaminase domain (Macbeth et al., 2005) this mutant should not affect deaminase domain folding and the protein may be capable of RNA binding. However, the mutation introduces an addi-
tional positively charged arginine residue on the RNA-binding face of the deaminase domain very close to the active site and the DSH1 phenotype suggests that the ADAR1 G1007R protein may be catalytically inactive.

dsRNA is required for both RNA editing by ADARs and RNAi. Thus, it is not surprising that there have been several reports of effects of ADARs in RNAi. Most of these studies have focused on the effect of the editing base change on processing of dsRNA or edited microRNA precursors (Scadden and Smith, 2001; Luciano et al., 2004; Scadden, 2005; Scadden and O’Connell, 2005; Yang et al., 2005, 2006; Blow et al., 2006; Ohman, 2007; Kawahara et al., 2007a, b). More recent studies focused on the downstream effects on target selection by edited miRNAs. For example, it was recently shown that an miRNA predicted to target the PRPS1 transcript is edited by ADAR2 and, interestingly, levels of PRPS1 protein are elevated in ADAR2-deficient mice (Kawahara et al., 2007b). This result is the only demonstration so far that miRNA modulation of protein levels is altered by ADARs.

There are several key steps in the miRNA pathway, which could be affected by ADARs (for review see Tomari and Zamore, 2005; Valencia-Sanchez et al., 2006). First, primary transcripts (pri-miRNA) are processed in the nucleus by the microprocessor complex, which contains Drosha and its partner DGCR8, to pre-miRNA hairpins (Gregory et al., 2004). Nuclear ADARs could therefore affect this step through steric hindrance of Drosha/DGCR8 binding or by inhibiting Drosha action through the alteration at the edited base in the primary miRNA transcript. As pre-miRNA hairpins can be bound by ADARs it is possible that their export from the nucleus, which is mediated by Exportin 5 (Yi et al., 2003), can be disrupted by binding. The next step in miRNA processing is the Dicer complex-mediated cleavage of the pre-miRNA hairpin into a 21–23 nucleotide RNA duplex in the cytoplasm. Cytoplasmic ADARs could also influence this step through sequestering the pre-miRNA or editing the pre-miRNA. Ultimately, the mature miRNA guide strand guides RISC to miRNAs that have complementarity with the guide strand, resulting in translation inhibition or miRNA degradation. ADAR antagonism of miRNA or siRNA production should result in alteration in the functional activity elicited by miRNAs or siRNAs.

In this work, we sought to determine which properties of ADARs are responsible for influencing miRNA function. Specifically, we have examined the ability of several truncation and point mutations of ADAR1 and ADAR2 to alter the function of mir-376a2 in a cell culture system optimized for analysis of the activity of a single miRNA. Our results indicate that editing can result in retargeting of human mir-376a2 as

Figure 1 Editing levels in the human mir-376 cluster. (A) Diagram of the mir-376 cluster; nomenclature is from miRBASE http://microrna.sanger.ac.uk/sequences. Numbers at bottom indicate intervening lengths of sequences. Bold text regions are the mature microRNAs. Boxes indicate major sites of editing. There is no evidence of editing in mir-654. (B) Editing in mir-376a1 expressed from a complete mir-376 cluster. (C) Editing in mir-376a2. Ratio G/(A + G) indicates the ratio of the guanosine signal to the total (adenosine + guanosine) chromatogram signal. +4 and +44 are positions within mir-376a2 indicated in A. (D) ADAR1 and ADAR2 protein domains showing the positions and features used in the construction of the hADAR expression vectors. hADAR1 p150 contains two Z-DNA-binding domains, three dsRBDs and one deaminase domain. hADAR1 p110 begins at amino-acid position 296 of hADAR1 p150. hADAR2 has three dsRBDs and one deaminase domain. hADAR2 ΔN is a deletion mutant of hADAR2 with amino acids from positions 4–72 removed. A key glutamate residue in the deaminase domain is amino acid 912 for ADAR1 and 319 for ADAR2. Mutation of this residue to alanine renders the ADAR catalytically inactive.
shown earlier for murine mir-376 (Kawahara et al, 2007b). Interestingly, our results also show that even in the absence of editing, ADARs can affect the processing of miRNAs at the Drosha cleavage step, thereby altering the functional activity of miRNAs. This suggests that ADARs may influence a larger set of miRNAs than would be predicted from the numbers that are known to be edited. We have also examined the ability of the human ADARs to alter silencing of the white⁺ eye colour gene in Drosophila by a 300+ plus base-pair hairpin, which is processed into siRNA duplexes (Vagin et al, 2006). We again find that a deaminase-inactive ADAR can antagonize siRNA activity against white⁺. Therefore, RNA binding by itself is an important function of ADAR biology.

Results

Editing of the mir-376 cluster by ADAR isoforms shows asymmetric editing in pri-mir-376a2

The human mir-376 cluster comprises several miRNA hairpins encoding mir-376c, mir-376a2, mir-654, mir-376b and mir-376a1 and others (Figure 1A). It was reported earlier that in human medulla several of the adenosines in this cluster are edited (Kawahara et al, 2007b). To elucidate whether particular domains of human ADAR1 or ADAR2 have prominent roles in the effect of ADARs on miRNA function, we cotransfected HEK 293T cells with constructs that express the mir-376 cluster and with constructs that express individual human ADAR1 or ADAR2 isoforms (Figure 1D summarizes key features of ADAR1 and ADAR2). All mutant proteins were expressed to the same level as the respective wild-type protein, as detected on immunoblots probed with anti-FLAG monoclonal antibody (data not shown). HEK 293T cells do not express detectable levels of the endogenous cluster. RNA editing levels were examined initially, before endeavouring to examine the effects of these ADAR variants on precursor processing and effects on downstream microRNA targets.

RNA was extracted from transfected cells, reverse transcribed with random hexamers and cDNA corresponding to individual pri-mir-376 hairpins were amplified using primers located around a 100 nucleotides before and after each hairpin within the long intervening sequences between the hairpins. RNA editing levels at sites within the pri-mir-376 hairpins were measured from mixed sequence chromatograms on the amplified sequence pools for each pri-mir-376 hairpin. We found that sites in pri-mir-376 hairpins were highly edited, as reported earlier for human medulla (Kawahara et al, 2007b).

The different specificities of ADAR1 and ADAR2 account for the different patterns of editing in the pri-mir-376a1 (Figure 1B), -376a2 (Figure 1C) and -376b hairpins (Kawahara et al, 2007b). For instance, ADAR2 edits pri-mir-376a1 at the +4 and +44 site more efficiently than ADAR1 (Figure 1B). In contrast, the two naturally occurring isoforms of ADAR1, p150 and p110, showed similar efficiencies at a range of sites within pri-mir-376a1, with the nuclear ADAR1 p110 editing slightly more efficiently in all cases (Figure 1B). The larger isoform has an extended amino terminus that is due to editing of the Z-DNA-binding domain in human medulla RNA, mir-654 was not edited in HEK 293T cells cotransfected with ADAR.

As anticipated, cotransfection of the ADAR1 1-296 construct that expresses only the first 296 residues, containing the first Z-DNA-binding domain of ADAR or the ADAR1 1-442 construct containing both Z-DNA-binding domains did not increase RNA editing. On the other hand, cotransfection with the ADAR1 443-end construct that expresses a protein lacking both Z DNA-binding domains gave levels of RNA editing similar to intact ADAR1 p110.

To determine whether editing activity is critical for effects of ADARs on microRNA processing and/or function we tested the effects of two ADAR1 mutant proteins. First, a construct with a glutamate to alanine change in an essential catalytic site residue (ADAR1 E912A) was shown to cause only background levels of RNA editing similar to that seen with cotransfection of the control pcDNA3.1 construct. We also cotransfected a construct expressing the naturally occurring G1007R mutation in ADAR1 that has been linked to DSH1 with the construct expressing the mir-376 cluster and found that this protein is incapable of editing any sites in the microRNA precursors. This protein, expressed in and purified from the yeast *Pichia pastoris* binds long dsRNA similarly to wild-type ADAR1 as assayed by filter binding but does not deaminate long dsRNA (Supplementary Figure S1). ADAR2 ΔN is a deletion of ADAR2 amino acids 4–72 that removes the nuclear localization sequence and causes active ADAR2 to accumulate in the cytoplasm (Wong et al, 2003). Surprisingly, the ADAR2 ΔN isoform edited the pri-miRNA, which is normally found only in the nucleus (Figure 1B and C). The observed editing by the cytoplasmic ADAR2 ΔN is due to editing of the pri-miRNA transcripts that escaped to the cytoplasm without being cleaved by the Drosha complex in the nucleus (Supplementary Figure S2).

ADAR2 shows strong inhibition of miRNA activity

As the RNA editing sites in mir-376a2 show the highest levels of preference for editing by ADAR1 or ADAR2 within the entire miR-376 cluster, we generated a construct that expresses pri-mir-376a2 alone. This allows the effects of ADAR2 binding and editing at the +4 site and ADAR1 binding and editing at the +44 site to be studied in the absence of other microRNAs that have strong sequence similarities. We verified that the editing of mir-376a2 alone was the same as editing of mir-376a2 expressed in the intact cluster (Supplementary Figure S3A).

Processing of pre-mir-376a2 by the miRNA processing machinery produces mature 5’ and 3’ microRNAs. To determine how the production and the specific targeting of the mature microRNAs is affected by the presence of ADARs we measured RNA-silencing activities with two sets of luciferase
constructs containing full-length microRNA target sites in the 3’ UTR regions of the luciferase transcripts (Supplementary Figure S3B). The 5’ miR luciferase reporter and the 3’ miR luciferase reporters measure the luciferase-silencing activity of the corresponding wild-type mature unedited microRNAs. The use of target sites with perfect homology extending beyond the seed match may lead to site-directed cleavage of the target transcript as well as translational inhibition. When ADARs edit within miRNAs they have the potential of retargeting miRNAs to another transcript. The RNA-silencing activities of the mature microRNAs retargeted as a result of editing are measured using a second pair of luciferase reporters with single base changes in the target sites (Supplementary Figure S3B).

We found that mir-376a2 expressed and processed in the absence of cotransfected ADAR constructs generated substantial silencing activities corresponding to the mature, unedited 5’ and 3’ miRs, as expected, (Supplementary Figure S3B), reducing luciferase levels 10-fold to approximately 0.1 of the value obtained in the absence of the miRNA. We optimized the level of mir-376a2-expressing construct so that it was sufficient to obtain this maximal level of silencing for both microRNAs.

Expression of the mir-376a2 construct in the absence of ADARs produced much less silencing activity directed against the reporters for the edited microRNAs, reducing the luciferase levels by only approximately two-fold, to approximately 0.5 of the value obtained in the absence of the miRNA (Supplementary Figure S3B). This five-fold discrimination by the mature unedited microRNAs between the unedited and the edited target sites is highly specific, considering that the edited target sites in each case differ by only 1 nucleotide in the 22 base sequences.

Having established conditions to maximize silencing activities of mature miRNAs from the transfected pri-mir-376a2 construct and to determine their target site preferences we cotransfected the mir-376a2 reporter with constructs expressing ADARs and ADAR mutations to assess how ADARs affect the silencing activities of resulting miRNAs. Consistent with their editing of pri-mir-376a2 at position +4, both hADAR1 p110 and hADAR1 p150 increased the silencing activity corresponding to the edited form of the 3’ miR (Figure 2B). This change was largely editing dependent, as the deaminase mutant ADARs (ADAR p150 E912A and ADAR p110 E912A) did not have this effect (Figure 2B); the effect was not dependent on the Z-DNA-binding domains of ADAR1 (ADAR1 443-end). Further, the Z-DNA-binding domains of ADAR1 (ADAR1 1-442) were not sufficient on their own to significantly increase the silencing activity of edited 3’ miRNA.

Notably, it was ADAR2 that produced the most striking inhibitory effects on miRNA function, particularly on the activity of the unedited miRNAs (Figure 2A and C). ADAR2 edits the pri-mir-376a2 at position +4 and thereby increases the silencing activity of edited 5’ miR (Figure 2D), at the expense of unedited 5’ mir activity (Figure 2C). As ADAR2 edits the +4 site in pri-mir-376a2 with 90% efficiency it is surprising that the retargeting effect was not more complete. Silencing of the reporters for edited microRNAs was maximized by cotransfecting equivalent amounts of mir-376a2 constructs bearing A-G point mutations at the RNA editing sites so that the mature miRNAs were ‘pre-edited’ to mimic the inosine-containing product. Cotransfecting a pri-mir-376a2 construct pre-edited at the +4 site gives a silencing activity corresponding to the edited 5’ miRNA that was almost twice as great as seen with ADAR2 (Figure 2D). A possible explanation is that ADAR2 edits pri-mir-376a2 very efficiently but that it also slows Drosha processing of the pri-mir-376a2 so that less mature miRNA is produced.

The hypothesis that ADAR2 inhibits Drosha processing of pri-mir-376a2 is supported, first, by the fact that ADAR2 inhibits the activity of the 3’ miRNA even though it edits the +44 site within this miRNA with only 10% efficiency. Second, the cytoplasmic protein ADAR2 ΔN has no effect either on activity or on retargeting of the 3’ micro RNA (Figure 2A and B). This is consistent with a nuclear inhibition of Drosha being responsible for the ADAR2-mediated inhibition of 3’ miR activity.

ADAR2 ΔN did affect the activity of the 5’ miRNA and increased the activity of the edited form. This appears to be due to base deamination and it does not suggest further cytoplasmic influence of ADAR2 on cytoplasmic processing steps. ADAR2 ΔN, which is cytoplasmic, edits the Drosha product, the pre-miRNA, after it is exported to the cytoplasm (Figure 2 and Supplementary Figure S2).

Drosophila processing of pri-mir-376a2 is inhibited by ADAR2 protein and not by the RNA editing base changes

As mentioned earlier ADAR interactions with miRNAs can occur at all levels of the miRNA processing pathway. We examined the processing of pri-mir-376a2 in transfected cells by northern analysis (Figure 3A and B; Supplementary Figure S4). ADARs were expressed in HEK293T cells in the presence of the mir-376a2 expression construct and the appropriate reporter construct. Only ADAR2 caused a significant reduction in the mature ~22-nucleotide miRNA and in the ~64-nucleotide pre-miRNA, with a reduction of over 65% in both cases. This indicates that ADAR2 activity on the pri-mir-376a2 hairpin disrupts Drosha processing in the nucleus. In further support of this hypothesis, ADAR2 ΔN, a cytoplasmically localized mutant of ADAR2, did not cause a significant reduction in the levels of either the pre-miRNA or the mature miRNA.

By performing in vitro pri-miRNA processing assays with HeLa nuclear extract we found that the addition of purified ADAR2 specifically inhibited Drosha processing of pri-mir-376a2 but not pri-let-7a-1 (Figure 3C), though some pre-mir-376a2 is still produced. To determine whether Drosha cleavage in the HeLa nuclear extract is inhibited by the RNA editing base changes or by binding of ADAR2, in vitro transcribed mutant pri-mir-376a2 transcripts bearing adenosine to guanosine changes at the edited positions were assayed. Specifically, in mir-376a2 a137g, a guanosine replaced adenosine at the +4 position and in mir-376a2 a177g, a guanosine replaced adenosine at the +44 site that is the +6 position of the 3’ miR. Importantly, Drosha cleavage was not affected by either mutation (Figure 3D), clearly showing that the nucleotide changes mediated by editing are not themselves sufficient to inhibit the processing of mir-376a2. The c188t construct that removes the A-C mismatch at the +4 site also had no effect.
Catalytically inactive ADAR2 disrupts pri-mir-376a2 processing

To confirm that inhibition of Drosha cleavage is due to ADAR2 binding rather than because of the base change at the edited site an active site-mutant ADAR2 E319A was co-expressed. ADAR2 E319A retained the ability to reduce the silencing activity of unedited 5' miR but, as expected, it was unable to retarget the silencing activity (Figure 4A). Furthermore, ADAR2 E319A caused inhibition of the processing of pri-miRNA-376a2, as detected on small RNA northern blot with RNA from cotransfected cells (Figure 4B). ADAR2 E319A disruption of Drosha processing verifies the conclusion that ADAR2 interference with miRNA processing is independent of editing. ADAR2 ΔN E319A, a deaminase domain mutant localized to the cytoplasm, did not inhibit 3' miR activity and only weakly affected 5' miR activity (Figure 4A). This shows further that it is nuclear Drosha cleavage that is affected and not cytoplasmic Dicer activity.

Human ADAR1 protein expressed in Drosophila inhibits RNAi through a mechanism that is partially independent of RNA editing activity

We generated a series of Drosophila strains that express different human ADAR proteins using the GAL4/UAS binary

Figure 2  Effects of ADARs on functional activities of mature miRNAs processed from pri-mir-376a2. (A) Inhibition of 3' mature miRNA activity. A value more than 1 means that the miRNA is less active when the ADAR is present than when the pri-mir-376a2 is expressed alone. (B) Retargeting of 3' mature miRNA activity. A value less than 1 means that the miRNA is more active in the presence of the ADAR than when the pri-mir-376a2 is expressed alone. Pre-edited mir-376a2 (a177g) indicates a construct expressing pri-mir-376a2 with the + 44 site (position 177) mutated from an adenosine to a guanosine. (C) Inhibition of 5' mature miRNA activity. (D) Retargeting of 5' mature miRNA activity. The activity for pre-edited mir-376a2 (a137g) is from a construct expressing pri-mir-376a2 with the + 4 site (position 137) mutated from an adenosine to a guanosine.
expression system. Interactions between expressed human ADARs and the RNAi system were examined by crossing these UAS-ADAR-bearing strains to a fly strain in which silencing of the wild-type white gene in the eye by a cytoplasmic white RNA hairpin construct is also directed by the GAL4/UAS binary expression system. Silencing of the white gene in the eye is directed by expressing a UAS-white hairpin construct under the control of the eye-specific GMR-GAL4 driver (Kalidas and Smith, 2002). Silencing of white results in flies with white eyes and this has been used successfully in a screen to find genes that antagonize the siRNA pathway (Lee and Carthew, 2003). The
white hairpin primary transcript contains an intron and has been designed to be efficiently processed, polyadenylated and exported to the cytoplasm. Silencing is mediated by the Dicer processing of the resulting cytoplasmic RNA hairpin, which is over 300-nucleotides long (Vagin et al., 2006). After Dicer cleavage, the white hairpin-derived siRNA targets the white⁺ mRNA for silencing by RISC.

When strains containing UAS-ADAR are crossed with the silencing reporter strain (homozygous w⁺; GMR>wIR), ADAR is co-expressed with the white hairpin in the eye. The interaction of ADAR with RNAi is observed as an alteration in the silenced white eye colour. The strongest effect of a human ADAR on white hairpin-mediated silencing was caused by ADAR1 p150, the shuttling isoform of ADAR1 that accumulates in cytoplasm (Figure 5C). This ADAR1 isoform antagonized RNAi strongly to restore the eye colour to red by allowing expression of the intact wild-type white⁺ gene in the strain background. This effect on the eye colour was not variegated but was absolutely consistent across the eyes and between flies.

Figure 4 Deaminase-inactive ADAR2 fails to retarget miRNAs but still inhibits their processing. (A) Top panel: effects of normal versus deaminase mutant ADAR2 on production of mature unedited and edited miRNA activities. Inactive ADAR2 E319A retained the inhibitory effect of ADAR2 on the unedited 5’ miRNA activity but was unable to retarget the activity by editing. Bottom panel: effects of cytoplasmic ADAR2 ΔN and ADAR2 ΔN E319A on the activity of mir-376a2. ADAR2 ΔN E319A did not affect the 5’ reporter, indicating a loss in retargeting. Neither hADAR2 ΔN nor hADAR2 ΔN deaminase mutant affected the 3’ mature miRNA activity. (B) Effect of ADAR2 E319A on processing of mir-376a2 in cotransfected HEK 293T cells. Small RNA northern blot of RNA from cotransfected cells probed with antisense probes to the 3’ miRNA. Blots were also probed with a 5S rRNA-specific probe as a loading control (data not shown). Deaminase mutant ADAR2 E319A expression reduced both pre-miRNA and mature miRNA levels. Percentage of pre-miRNA signal relative to pri-mir-376a2 expressed alone is noted below the blot. (C) Effects of main isoforms of ADAR1 and ADAR2 on functional activities of 5’ and 3’ miRNAs expressed from a construct bearing mir-376a1 alone. As in the case of mir-376a2, ADAR2 was able to inhibit the activity of mir-376a1 independently of deaminase activity. (D) The ADAR2 E319A deaminase domain mutant reduced the level of pre-mir-376a1 in cotransfected HEK 293T cells. Small RNA northern blot of RNA from transfected cells probed with antisense probe to the 3’ miRNA. The percentage of pre-mir-376a1 signal relative to pri-mir-376a1 expressed alone is listed below the blot.
human ADAR that is presumably because of the endogenous fly protein (Supplementary Figure S5A). Of the human proteins only the cytoplasmic ADAR1 p150 showed a substantially increased level of editing. This suggests that cytoplasmic localization is required for efficient editing and for full inhibition of siRNA activity in this assay.

Combining UAS-Dicer-2 with UAS-ADAR1 p150 in crosses to the homozygous w⁺; GMR> wIR strain showed that increasing DICER-2 expression above normal levels increases the background level of RNAi in the absence of ADAR1p150 in the progeny (Figure 5, compare panels E and A) and also partly but not completely suppresses the antagonism of RNAi by ADAR1 (Figure 5, compare panels C and F). This suggests that the antagonism of RNAi by ADAR1p150 is partly attributable to competition with DICER-2 for long dsRNA, although ADAR1p150 does also bind the shorter dsRNA products of DICER cleavage (Yang et al, 2005) and changing dosages of R2D2 and ARGONAUTE-2 might have similar effects.

The visual assay is very consistent, especially because groups of ADAR-expressing flies can be compared with sibling progeny of the same age, which have white silencing but lack ADAR. This gives sensitivity to the assay and testing further ADAR1 derivatives shows that all the domains of ADAR1, including the Z-DNA-binding domains, have perceptible effects on RNAi in this assay. The truncation mutant, ADAR1 1-442, containing two Z-DNA-binding domains showed an effect in this assay and removing the Z-DNA-binding domains in the ADAR1 443-end construct weakened the inhibition (data not shown).

When human ADAR1p150EA, the deaminase-inactive mutant, was expressed in this system it was found to still antagonize the RNAi activity of the white hairpin (Figure 5G and H). Again the effect was very consistent between flies. The red eye colour was 40% as strong as ADAR1 p150, based on quantization of red pigment in ethanol-extracted heads and in digital images of eyes (Supplementary Figure S5B). This result suggests that, as in the case of miRNA processing, a significant part of the antagonism is independent of deaminase activity. Very surprisingly, expressing the ADAR2 ΔN protein that lacks the nuclear localization signal increased the antagonism by ADAR2 only very slightly (not shown). This suggests that the specificity of the different ADARs is also important for this effect.

Discussion

Earlier work on the interaction between ADARs and the miRNA/siRNA pathways in mammals has focused on the effects of the base changes introduced by RNA editing. This editing was shown to affect microRNA processing or produce retargeting (Scadden and Smith, 2001; Luciano et al, 2004; Scadden, 2005; Yang et al, 2005; Blow et al, 2006; Landgraf et al, 2007; Kawahara et al, 2007a, b). Our data show, for the first time, that catalytically inactive ADAR2 can inhibit the processing and decrease the downstream function of miRNA, independent of base deamination. In addition, we show that deaminase-inactive ADAR1 p150 inhibits siRNA activity in a Drosophila test system. ADAR1 p150 also inhibits siRNA in ADAR1⁻/⁻ MEFs (Yang et al, 2005). As ADARs may bind to a wider range of pre-miRNAs than they successfully edit, the influence of ADARs on miRNA function may also

Neither nuclear ADAR1 p110 nor ADAR2 had any effect on white hairpin-mediated silencing (Figure 5B and D). The human ADARs are expressed in addition to the normal level of endogenous Drosophila ADAR in the flies shown here. Drosophila ADAR, which is also nuclear, behaved like ADAR2 in this assay: neither increasing the expression with a UAS-ADAR construct nor deleting the single endogenous Drosophila ADAR gene on the X chromosome affected silencing (data not shown). When editing of the white hairpin over a stretch of several hundred base pairs was analysed in these flies there was some editing even in the absence of any
extend beyond the subset of miRNA, which are edited. Modulation of miRNA biogenesis by binding Lin-28 protein (Newman et al., 2008; Piskounova et al., 2008) or hnrNP A1 (Guil and Caceres, 2007; Michlewski et al., 2008) to miRNA precursors has also been reported recently. Therefore, regulation of pri-miRNA processing by accessory RNA-binding proteins may be more common than appreciated earlier.

There is specificity in how different ADARs affect different RNAi phenomena as only ADAR2 robustly affects the processing of pri-mir-376a2. Despite overexpressing other ADAR proteins with dsRNA-binding domains, they did not radically inhibit processing of this pri-miRNA by Drosha. What is really surprising is that ADAR1 binds to and edits the +4 position in mir-376a2 to between 40 and 50% yet its affect on Drosha processing is not as strong as ADAR2 (Figure 3). The +4 editing site is a mere three base pairs from the Drosha cleavage site.

The specificity of ADAR effects is also clear in relation to Dicer processing. It was recently shown that pre-mir-151 could be edited by ADAR1 p110 and ADAR p150 in vitro, and that this editing led to disruption of Dicer processing (Kawahara et al., 2007a). However, cytoplasmic ADAR2 AN did not result in disruption of Dicer processing of the pre-mir-376a2 in this study. Neither was there degradation of miRNA duplexes by Tudor-SN but instead there was retargeting of 5’ miRNA activity by ADAR2 AN because of editing. In addition, ADAR1 p150 did not have a major impact on mir-376a2 processing. Therefore, not all editing events in pre-miRNA in the cytoplasm necessarily disrupt Dicer processing just as not all editing events in pri-miRNA in the nucleus affect Drosha cleavage.

We found that single nucleotide changes in miRNA target sites impair the ability of a particular miRNA to target the mRNA. Several publications have documented the decrease in the ability of miRNA to silence an mRNA when there are nucleotide changes in the miRNA responsive site (reviewed in Tomari and Zamore, 2005). It seems that siRNA activity is even more sensitive to nucleotide changes. Therefore, editing may have a more dramatic effect on silencing of a transcript if it occurs within a miRNA responsive site in the 3’UTRs of specific miRNAs rather than if the miRNA itself is edited. This may be important in some species like *Caenorhabditis elegans* where there is significant editing of 3’UTRs.

We have shown here that one DSH1-associated variant of ADAR1 (Miyamura et al., 2003; Zhang et al., 2004, 2008; Liu et al., 2006) produces an ADAR1 protein that binds dsRNA with a similar Kd to wild type but is catalytically inactive (Supplementary Figure S1). Members of the family having this variant show the same dermatological symptoms of DSH1, which are associated with protein truncations of ADAR1 and that probably reflect decreased RNA editing activity in other families. However, affected members of the Japanese family having the ADAR1 G1009R allele also show additional symptoms including progressive neuromotor defects and calcium accumulation in basal ganglia. On the basis of our demonstration of efficient RNA binding by this protein we suggest that these additional symptoms arise because ADAR1 G1009R partially antagonizes normal RNA editing. Inhibition of mir-376 cluster processing in the brain may be relevant but the mutant protein would also probably bind RNA editing target sites such as the Q/R editing site in the *GluR-B* transcript and reduce editing by ADAR2 and ADAR1 at this site. This would lead to calcium-permeable AMPA receptors and possibly contribute to the brain calcium accumulations and to the slowly progressing neurological symptoms in this family.

In conclusion, we report a number of different and very selective effects of ADARs on RNAi processes that appear to depend significantly on the RNA-binding activities of ADARs. It has been shown earlier that ADARs can bind to many transcripts without editing them (Klaue et al., 2003). A biological function can now be ascribed to this binding activity. Binding of ADAR2 to pri-mir-376a2 is sufficient to inhibit processing by Drosha, an inactive form of ADAR1 p150 is sufficient to inhibit RNAi in a *Drosophila* assay, and disease symptoms in a Japanese family are probably explained by production of an ADAR1 protein that binds RNA but lacks catalytic activity. This implies a broader functional role for ADARs as RNA-binding proteins in different cellular compartments interacting with a range of dsRNA-based phenomena.

**Materials and methods**

**Expression plasmids**

miRNA-expressing plasmids contained mir-376 cluster genomic DNA were amplified with PCR primers and cloned using BamHI and *HindIII* restriction enzyme sites in the primers into the pcDNA3.1 vector. All constructs had 76–100 nucleotides of genomic sequence on both the 5’ and 3’ sides of the encoded mirs depicted in Figure 1A (see Supplementary Table 1 for sequences of primers). Site-directed mutagenesis was performed to introduce the a137g, a177g and c188t mutations into pri-mir-376a2.

Human ADAR-expressing plasmids, containing ADAR coding sequences with N-terminal Flag and C-terminal His epitope tags, were constructed by Gateway Cloning (Invitrogen) with pc3D as the destination vector. pc3D was derived by blunt-end ligation of the gateway cloning cassette-B from Invitrogen into pcDNA3.1. Therefore, the vector for all expression constructs was pcDNA3.1.

**miRNA reporter plasmids**

Target sites for each miRNA were subcloned into the 3’ UTR of *B. subtilis* luciferase in psichek 2.0 (Promega) using the Xhol and Noti restriction enzyme sites. As a transfection control the vector also contains humanized Firefly luciferase under its own promoter. This is also a control for any general effects on translation. (see Supplementary Figure 3 for sequences of target sites).

**Editing assays**

RNA was purified from transiently transfected cell cultures and subjected to reverse transcription with random hexamers (Invitrogen) and MMLV-RnaseH (Invitrogen). Editing was determined by direct sequencing of PCR product pools with primers located within the *Renilla* luciferase in psicheck 2.0 (Promega) using the Xhol and Noti restriction enzyme sites. As a transfection control the vector also contains humanized Firefly luciferase under its own promoter. This is also a control for any general effects on translation. (see Supplementary Figure 3 for sequences of target sites).

**In vitro RNA binding and editing**

In vitro RNA-binding assay was performed as described by Ohman (Ohman et al., 2000). In vitro RNA editing assays was performed as described by O’Connell (O’Connell and Keller, 1994).

**Cell culture**

HEK 293T cells were cultured in DMEM supplemented with 10% FCS.
Transient transfections

Transient transfections were performed in 24-well plates seeded 1 day before transfection with 0.14 × 10⁵ cells, with 1.005 μg DNA and 1.01 μL Lipofectamine 2000. The amounts of miRNA-expressing plasmid, ADAR-expressing plasmid and luciferase reporter target construct plasmid were 500, 500 and 5 ng, respectively. Cells were collected on the third day after the transfection.

Luciferase assays

The miRNA target plasmid, derived from pscheck 2.0, contains humanized Firefly and Renilla luciferase. The miRNA responsive site is in the 3'UTR of Renilla luciferase. Assays for Firefly and Renilla luciferase were performed with the Dual-Glo luciferase kit from Promega. Luciferase was measured on a Perkin-Elmer VictorII. To ascertain the activity of mir-376a2, pri-mir-376a2 was co-expressed with the luciferase reporters so that the amount of DNA was constant. The resulting luciferase level was compared to that obtained in a cotransfection of pcDNA3.1 and the luciferase reporter without the pri-mir-376a2 expression vector. Fold Luc activity is the activity remaining in the presence of the miRNA.

Fold Luc = (luciferase activity with the pri-mir-376a2 construct)/(luciferase activity without the pri-mir-376a2 construct).

Assays on ADARs were performed by co-expressing the ADAR, pri-mir-376a2 and the appropriate luciferase reporter. The resulting luciferase level was compared to co-expression of ADAR, pcDNA3.1 and the appropriate reporter without pri-mir-376a2. For the ADAR constructs.

Fold Luc = (luciferase level with pri-mir-376a2 and ADAR construct)/(luciferase level without pri-mir-376a2 and with the relevant ADAR construct).

The activity was then normalized to the ratio obtained with pcDNA3.1 and the pri-miRNA expression construct alone. Fold Luc over pri-mir-376a2 alone (labelled on the x-axis in Figure 2) is the fold change in luciferase activity compared with expression of the pri-miRNA alone. This indicates how the activity of the microRNAs to direct silencing of luciferase activity from the different reporters has been affected by the presence of ADAR proteins. A value greater than 1 indicates inhibition of the miRNA activity, a value less than 1 indicates increased miRNA activity.

Small RNA northerns

RNA samples were electrophoresed on a denaturing 12% PAGE 8 M Small RNA northerns to direct silencing of luciferase activity from the different reporters.

Competent for Drosha cleavage. After incubation at 37 °C, reactions were stopped and electrophoresed on a denaturing PAGE gel.

For Drosha cleavage assays in the presence of hADAR2, the recombinant hADAR2 had been expressed and purified from P. pastoris (Ring et al., 2004).

Drosophila transgenic lines

ADAR coding sequences were cloned in the pUAST plasmid bearing GAL4-binding sites. Constructs were injected in Drosophila eggs and transgenic lines were obtained and balanced by standard methods. Female flies bearing UAS-ADAR constructs on chromosomes II or III were crossed to males homozygous for both the GMR-GAL4 and UAS-whiteR constructs (w/+; GMR-GAL4, UAS-wR). Levels of red pigment in the eyes of female progeny flies in the RNAi assay were quantitated by ethanol extraction of batches of 50 heads followed by optical absorption measurements (Pal-Bhadra et al., 2004). The maximal level of red pigment in the presence of UAS-ADAR1 p150 is 12% of the wild-type level.

The images in Figure 5 were made with an imaging system comprising a Nikon AZI100 macroscope with 2 × plan fluor objective at 3 × zoom setting (Nikon UK Ltd, Kingston-on-Thames, UK), a Qimaging MicroPublisher 5 cooled colour camera (Qimaging, Burnaby, BC, Canada). Image capture and control of the filterwheel and image analysis were performed using IPLab Spectrum (Scanalytics Corp, Fairfax, VA). White balance for the illumination level used was set on a white background, and all images were captured in a single session using identical capture settings.

The data for Supplementary Figure 5B were imaged on a Leica MZFL II/II stereo-microscope with 0.5 ×, 0.63 ×, 1 ×, 2 × objectives. Images were captured with a Hamamatsu Orca AG CCD camera (Hamamatsu Photonics (UK) Ltd, Welwyn Gardens City, UK) and CRI liquid crystal RGB filter (Cambridge Research & Instrumentation, Woburn, MA). Image capture and analysis were performed with in-house scripts written for IPLab Spectrum (Scanalytics Corp). The pixel intensity was measured on both eyes from 10 different flies from each genotype.

Supplementary data

Supplementary data are available at The EMBO Journal Online (http://www.embojournal.org).

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Conflict of interest

The authors declare that they have no conflict of interest.
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