Antiproliferative activity of marine stingray *Dasyatis sephen* venom on human cervical carcinoma cell line

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Abstract

**Background:** Venoms comprise mixtures of numerous bioactive compounds that have a wide range of pharmacologic actions. Toxins from venomous animals have attracted the attention of researchers because of their affinity for primary sites responsible for lethality and their efficacy at extremely low concentrations. The venoms of marine stingrays have not been extensively studied and limited data is available on them. The present study aims to evaluate the antiproliferative and biochemical properties of the venom obtained from a species of marine stingray (*Dasyatis sephen*) on human cervical cancer cell line HeLa.

**Methods:** The antiproliferative effect of *D. sephen* venom was determined by MTT assay, and the oxidative stress was determined by lipid peroxidation method along with assessment of changes in the enzymatic and non-enzymatic antioxidant status. We observed intracellular reactive oxygen species (ROS) levels by DCFH-DA method, mitochondrial membrane potential alterations by rhodamine 123 staining and apoptotic morphological changes by acridine orange/ethidium bromide dual staining method.

**Results:** *D. sephen* venom enhances lipid peroxidative markers such as thiobarbituric acid reactive substance, conjugated diene, and lipid hydroperoxide in HeLa cell lines. Stingray venom enhances the ROS levels, which is evidenced by the increased 2–7-diacetyl dichlorofluorescein fluorescence. Further, *D. sephen* venom treatment altered the mitochondrial membrane potential in HeLa cells. Additionally, we observed increased apoptotic morphological changes in *D. sephen* venom-treated groups.

**Conclusions:** *Dasyatis sephen* venom exhibits potent antiproliferative effect on HeLa cell line and upon further purification it could be a promising antiproliferative agent.

**Keywords:** Marine organisms, Membrane potential, Oxidative stress, ROS, Stingray, Venom

Background

Marine organisms comprise approximately one-half of the total global biodiversity; therefore, they offer an important source for novel compounds that has been classified as the largest reservoir of natural molecules to be evaluated for drug activity [1]. A different type of environment exists in the ocean, where organisms live in competitive and aggressive surroundings that differ in many aspects from the terrestrial environment. This competitive environment demands the production of quite specific and potent active molecules by marine organisms [2]. These organisms have been continuously screened for pharmacologically active substances and till date over 6500 marine natural products have been isolated [3].

Cancer is characterized by the uncontrolled growth and spreading of abnormal cells. In normal cell function and tissue homeostasis, proliferation and apoptosis are balanced. Cancer cells display abnormal *in vivo* proliferation that is not balanced by compensatory apoptosis [4]. Apoptosis, defined as a controlled form of cell death, might...
represent a pivotal point in cancer treatment development [5]. For the past few decades, venom components have become the focus of researchers and have been extensively studied for their various anticaner properties with effective inhibition of proliferation. The anticaner potential of venom on adenocarcinoma cells was first reported for Naja snake venoms by Calmette et al. [6]. In addition, bee venom has also been found to have antiproliferative activity in vitro and ability to reduce tumor growth in vivo [7]. Scorpion venom toxins demonstrate antiproliferative activity against human glioma and leukemic cells as well [8, 9]. Marine environment is highly diverse and several compounds from marine origin have been approved for clinical use, including vidarabine (recurrent epithelial keratitis and superficial keratitis), cytarabine (cancer), ziconotide (chronic pain in cancer or AIDS), trabectedin (soft tissue sarcoma) and halaven (metstatic breast cancer) [10, 11]. But, concerning fish venoms, limited data are available about their bioactive potential.

Marine stingrays are venomous fish of the Elasmobranch family and are mainly found in temperate and tropical areas of the world. Stingrays are considered to be one of the most significant venomous fish in the world [12]. They have serrated spines at the base of the tail that penetrate the body of the victim. The epidermal covering of the spine releases venom into the sting site causing severe pain and tissue necrosis [13]. Stingray venom exhibits neurotoxic, cardiotoxic, fibrinogenolytic and anticoagulant activities and are composed of proteins, serotoxin, vasoconstrictor peptides and several other unidentified components [14–18]. In this study, the mechanism by which the venom of the marine stingray D. sephen inhibits cell proliferation and the associated apoptotic pathways are examined in human cervical cancer cells (HeLa).

**Methods**

**Chemicals**

Bovine serum albumin, thiobarbituric acid (TBA), phenazine methosulphate (PMS), nitroblue tetrazolium (NBT), 5,5-dithiobis(2-nitrobenzoic acid) (DTNB), 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT), 2−7-diacetyl dichlorofluorescein (DCFH-DH), rhodamine 123 (Rh-123), acridine orange, ethidium bromide, heat inactivated fetal calf serum (FCS), minimum essential medium (MEM), McCoY’s modified medium, glutamine, penicillin-streptomycin, EDTA and trypsin were purchased from Sigma Chemicals Co. (USA).

**Venom extraction**

Stingray specimens were collected from Parangipettai coast (11°30’1.19”N 79°46’20.50”E, Tamil Nadu, India) by local fishermen. The spines that are usually discarded by fishermen due to the lack of commercial value were removed from the base of the tail and transported to the laboratory in an icebox. The tissue covering the stingers was scratched and homogenized in PBS buffer, pH 7.4 and centrifuged at 5000 × g for 10 min. The supernatant (venom) was stored at −20 °C until use [19]. Protein concentration of the supernatant was estimated by standard method using BSA as protein standard. Details about the D. sephen and stinger images are provided in Additional file 1.

**Cell lines and culture conditions**

The present work was carried out on human cervical cancer cell line (HeLa). The cell line was obtained from the National Centre for Cell Science (NCCS), Pune, India. HeLa cells were grown as monolayer in MEM medium supplemented with 10 % FCS, 2mM glutamine, and 100 U/mL penicillin-streptomycin at 37 °C in 5 % CO₂ atmosphere. Stocks were maintained in 25 cm² tissue culture flasks.

**Study group and dose fixation study**

Cells were treated with different concentrations of stingray venom (2, 4, 8, 12, 16, 20 or 24 μg/mL) and the cytotoxicity was observed by MTT assay. PBS used as a sham control. The results assessed by MTT assay were employed in further experiments in which five groups were defined based on the venom administration doses:

- **Group I:** untreated HeLa cells (control)
- **Group II:** HeLa cells + D. sephen venom (4 μg/mL)
- **Group III:** HeLa cells + D. sephen venom (8 μg/mL)
- **Group IV:** HeLa cells + D. sephen venom (12 μg/mL)
- **Group V:** HeLa cells + D. sephen venom (16 μg/mL)

**Drug sensitivity assay**

The growth inhibitory activity of cells was determined by the MTT assay based on the detection of mitochondrial dehydrogenase activity in living cells [20]. Each culture well was incubated for 24 h with different doses of D. sephen venom. Ten microliters of MTT solution (5 mg/mL in PBS) was added and incubated for 4 h to allow color development. An equal volume of DMSO was then added to stop the reaction and to solubilize the blue crystals formed. The absorbance was taken at the wavelength of 570 nm.

**Measurement of intracellular ROS in cells by spectrofluorimetric and fluorescence microscopic methods**

Intracellular ROS was measured by using a nonfluorescent probe, DCFH-DA, which penetrates into the intracellular
matrix of cells to be oxidized by ROS to fluorescent dichlorofluorescein (DCF) [21].Cells were incubated for 24 h with different concentrations of D. sephen venom. Fluorescent dye DCFH-DA was then added to the cells and incubated for 30 min. The cells were washed with PBS to remove the excess dye before fluorescent measurements that were carried out with excitation and emission filters set at 485 ± 10 and 530 ± 12.5 nm, respectively (Shimadzu RF-5301 PC Spectrofluorometer, Japan). Fluorescence microscopic images were taken using blue filter (450–490 nm) (Nikon, Eclipse TS100, Japan).

**Alterations in mitochondrial membrane potential**

After cell incubation with D. sephen venom for 24 h, fluorescent dye Rh-123 (10 μg/mL) was added to the cells. The cells were then incubated for 30 min, washed with PBS and analyzed in fluorescence microscope using blue filter [21]. Polarized mitochondria were marked by orange-red fluorescence, and depolarized mitochondria were marked by green fluorescence.

**Apoptotic morphological changes by acridine orange-ethidium bromide dual staining method**

Staining of DNA with acridine orange (AO) and ethidium bromide (EBr) allowed visualization of the condensed chromatin of dead apoptotic cells [21]. Stained cells were viewed under a fluorescence microscope. The number of cells showing features of apoptosis was counted as a function of the total number of cells present in the field.

**Estimation of lipid peroxidation markers**

The cells were harvested by trypsinization. The pellet obtained was suspended in PBS and taken for the measurement of lipid peroxidative markers such as thiobarbituric acid reactive substances (TBARS), conjugated dines (CD), and lipid hydro peroxide (LHP), according to the procedures described elsewhere [22–24].

**Estimation of antioxidant enzyme activity**

The supernatant obtained after centrifuging the trypsinized cells was used for the measurement of activities of antioxidant enzymes, superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx), according to the procedures described elsewhere [25–27].

**Estimation of reduced glutathione levels**

The levels of reduced glutathione (GSH) were determined in the supernatant obtained after centrifuging the trypsinized cells according to the procedures described elsewhere [28].

**Statistical analysis**

Statistical analysis was performed using one way analysis of variance (ANOVA) followed by Duncan’s multiple range test (DMRT) by using Statistical Package of Social Science (SPSS) version 12.0 for Windows. The values were expressed as mean ± SD for six samples in each group. In addition, p value of < 0.05 was considered as statistically significant.

**Results**

**Effect of D. sephen venom on cell proliferation**

Effect of D. sephen venom on cell proliferation was determined by MTT assay. The proliferation of HeLa cells was significantly inhibited by D. sephen venom. Figure 1 shows the changes in the percentage of cell death in control and venom-treated cells. The treatment with 2 μg/mL of D. sephen venom did not show significant (p < 0.05) proliferation inhibition. The treatment with 4, 8, 12 and 16 μg/mL of D. sephen venom significantly inhibited in HeLa cells. Concentrations of 16 and 20 μg/mL displayed almost the same inhibitory effect (80 and 81 % inhibition) on HeLa cells. Hence, for further tests D. sephen venom concentrations of 4, 8, 12 and 16 μg/mL were used on HeLa cells.

**D. sephen venom generates intracellular ROS**

Levels of ROS in control and D. sephen venom-treated cells are depicted in Fig. 2 – A (i) and B (i). Venom treatment significantly increased ROS level in HeLa cells. Venom concentrations of 4, 8, 12, and 16 μg/mL increased ROS levels to 12, 30, 63 and 75 %, respectively, when compared to control group. Among all tested doses, 16 μg/mL showed maximum generation of ROS in HeLa cells.

**D. sephen venom modulates mitochondrial membrane potential**

Changes in mitochondrial membrane potential in control and D. sephen venom-treated cells are depicted in Fig. 2 – A (ii) and B (ii). D. sephen venom treatment significantly increased mitochondrial depolarization in HeLa cells. After 24-hour incubation, venom treated cells showed significantly decreased mitochondrial membrane potential to 57, 35, 18, and 10 % at 4, 8, 12, and 16 μg/mL concentrations of D. sephen venom in HeLa cells, respectively, when compared to control group. Among all the doses tested, 16 μg/mL of D. sephen venom showed the highest level of mitochondrial depolarization in HeLa cells. Polarized mitochondria were marked by orange-red fluorescence and depolarized mitochondria were marked by green fluorescence.

**Effect of D. sephen venom on apoptotic morphological changes**

Figure 2 – A (iii) and B (iii) show the effect of D. sephen venom on apoptotic morphological changes. Figure 2 – A (iii) shows the fluorescence microscopic observation of the total number of cells present in the field.
of the untreated and treated cancer cells. Untreated cancer cells appeared in green (AO stained) whereas venom treated cells appeared in red/orange (EtBr stained) that revealed the presence of apoptotic cells. Figure 2 – B (iii) shows the percentage of apoptosis. Venom concentrations of 4, 8, 12, and 16 μg/mL increased apoptotic cells levels to 30, 50, 74 and 78 %, respectively, when compared to the control group. Among all the doses tested, 16 μg/mL of D. sephen venom showed increased apoptotic levels in HeLa cells.

Changes in the levels of lipid peroxidative markers and the activities of enzymatic antioxidants

We observed the levels of lipid peroxidative markers, such as TBARS, CD and LHP in control and venom-treated cells (Fig. 3). D. sephen venom treatment increased the levels of lipid peroxidation in Hela cells. Among all the concentrations (4, 8, 12, and 16 μg/mL) tested, 16 μg/mL significantly increased levels of TBARS, CD and LHP in HeLa cells.

Activities of enzymatic antioxidants such as SOD, CAT, and GPx are depicted in Fig. 4. D. sephen venom (4, 8, 12, and 16 μg/mL) treatment significantly decreased the activities of SOD, CAT, and GPx in HeLa cells. Among all tested doses, 16 μg/mL of D. sephen venom significantly decreased enzymatic activities when compared with other doses in HeLa cells. In this study, the effect of D. sephen venom on glutathione levels in venom-treated cancer cells was examined. Levels of GSH in control and venom-treated cells are showed in Fig. 5. The treatment of D. sephen venom (4, 8, 12, and 16 μg/mL) decreased GSH levels in HeLa cells. Regarding all the doses tested, 16 μg/mL of D. sephen venom significantly decreased GSH levels in HeLa cells.

Discussion

Venoms from many marine sources have demonstrated to be highly effective in low concentrations. Marine fish venoms have not been fully explored and their biological potentials have not been completely characterized. In the present study, we evaluated in vitro antiproliferative potential of stingray venom extracts on HeLa cell line. D. sephen venom venom significantly decreased the viability of cancer cells. There was a significant increase in cytotoxicity (26, 42, 68 and 80 %) of HeLa cells with increasing concentrations of this venom at 4, 8, 12, and 16 μg/mL, respectively. Our results indicate that concentrations of the compounds play a vital role in cytotoxicity. It is very likely that D. sephen venom may disrupt mitochondrial dehydrogenase activity of cancer cells at higher concentrations [29]. Mitogenic and cytotoxic effects of venom from marine sources such as S. verrucosa and H. rubripinnis on regular and tumor cell lines have been reported earlier [30]. However, the molecular role of venom in disrupting intracellular signaling pathways leading to apoptosis (programmed cell death) of cancer cells have not yet been elucidated [31]. Fleury et al. [32] stated that an increase in the production of ROS associated with disturbance in the oxidative status results in the impairment of mitochondrial function that triggers the apoptosis pathway. Hence, in this study, assessment of ROS species, mitochondrial function and antioxidant enzyme activity determination was carried out.

ROS is known to be generated intracellularly though three pathways: as the byproduct of normal aerobic...
Fig. 2 (See legend on next page.)
metabolism; as second messengers in various signal transduction pathways; and in response to environmental stress. Depending upon the concentration, ROS is known to elicit a wide spectrum of biological responses ranging from mitogenic to proliferative effects at low concentrations, and macromolecular damage leading to cell death at high concentrations [33]. Previous reports have suggested that Okinawa habu apoxin protein-1 (OHAP-1) from Okinawa habu (T. flavovirudis) venom induced apoptosis in malignant glioma cell lines by promoting the generation of intracellular ROS [34]. Similarly, in our study D. sephen venom caused a rapid increase of intracellular ROS levels (63 and 76 % at 12 and 16 μg/mL concentrations, respectively) in HeLa cells after 24 h of incubation.

ROS are known to be generated from the reaction of leaked electrons with oxygen under various systems including lipid peroxidation. Lipid peroxidation is a multistep process in which the initially formed lipid radicals are converted to TBRS via the unstable intermediate products CD and LPH [35]. We observed the levels of lipid peroxidative products, such as TBARS, CD and LHP in control and D. sephen venom-treated cells. D. sephen venom treatment increased the levels of lipid peroxidative products in HeLa cells. Among all the concentrations tested (4, 8, 12, and 16 μg/mL), 16 μg/mL of venom showed significantly increased levels of TBARS, CD and LHP in HeLa cells.

Mitochondrion is one of the most important organelles that regulate cell death and mark apoptosis [36]. Functional alterations of mitochondria have been shown to play an important role in cell apoptosis [37]. The mitochondria of normal cells pump H⁺ from initial ground substance to the outside of the endomembrane creating a transmembrane potential. In the present study, mitochondria membrane potential was calculated in order to examine whether apoptosis is accompanied by the loss of mitochondrial transmembrane potential. We observed D. sephen venom-treated cells showed significantly decreased mitochondrial membrane potential by 57, 35, 18, and 10 % at concentrations of 4, 8, 12, and 16 μg/mL, respectively, in HeLa cells when compared to the control group. Rh-123, a mitochondria-specific membrane permeable dye, was accumulated in the mitochondria of control cells whereas venom-treated cells exhibited decreased uptake of it with increasing concentrations. Our results indicate that the mitochondrial damage occurred during D. sephen venom treatment, which suggests that the alteration of mitochondrial membrane potential (MMP) may have a role in venom-induced cell death.
HeLa cell death induced by venom might be due to mitochondrial toxicity, as the MMP collapsed before apoptosis. This observation corroborates a previous study in which treatment with the *O. doriae* venom induced reactive nitrogen intermediates, caspase 3 and depolarization in mitochondria [38].

Apoptosis plays an important role in determining cellular cytotoxicity following drug treatment [39]. It is known to be associated with a characteristic set of morphological features, including membrane blabbing, chromosomal condensation, nuclear fragmentation, cell shrinkage and formation of apoptotic bodies. In the present study, we observed that *D. sephen* venom pretreatment significantly increased morphological changes associated in HeLa cells leading to 74 to 78% of apoptosis at concentrations of 12 to 16 μg/mL. Fluorescence microscopic observation of the cells showed a typical apoptotic morphology – cell pyknosis, chromosome condensation and nuclear fragmentation in *D. sephen* venom-treated cells when stained with EtBr/AO.

Acridine orange (AO) is a cationic dye known to penetrate the intact membrane of living cells and stain DNA. On the other hand, EtBr is not incorporated by live cells. Hence, control cells can be observed as green under blue emission. On the contrary, cells undergoing apoptosis are not able to exclude EtBr and then exhibit more intense red color [40]. Increased ROS levels and loss of mitochondrial membrane potential might be the reason for increased apoptotic morphological changes such as cell pyknosis and chromosome condensation observed in the venom-treated cells.
In our body, antioxidant enzymatic defense is a very important tool to neutralize oxygen free radical-mediated tissue injury [41]. Free radical scavenging enzymes such as CAT, SOD, and GPx are the first line of cellular defense against oxidative injury, decomposing \( \text{O}_2^\cdot \) and \( \text{H}_2\text{O}_2 \) before their interaction to form the more reactive hydroxyl radical. Antioxidant enzymes can antagonize initiation and promotion phases of carcinogenesis and their levels are reduced in many malignancies [42]. Cell malignancy or transformation is often accompanied by decrease in activity of antioxidant enzymes like CAT, SOD, and GPx, which increases the cell sensitivity to prooxidant compounds [43]. *D. sephen* venom at concentrations of 4, 8, 12, and 16 \( \mu \text{g/mL} \) significantly decreased the activities of SOD, CAT, and GPx in HeLa cells. Among all the doses tested, 16 \( \mu \text{g/mL} \) expressly decreased enzymatic activities when compared with other doses in HeLa cells. It has been shown in the literature that anticancer agents deplete intracellular antioxidants and because of that, there is increased accumulation of free radicals inside the cells which modulate the opening of the mitochondrial permeability transition pore resulting in apoptosis [21, 35]. The present finding indicates that the *D. sephen* venom has caused cell death by damaging mitochondria, increasing intracellular ROS formation and depletion of cellular antioxidant enzyme in a dose-dependent manner in HeLa cells. This corroborates a previous study that showed the cytotoxic potential of *Conus vexillum* venom, which induced oxidative stress [44].

GSH plays a key role in protecting cells from electrophilic compounds and free radicals generated during cellular metabolism. Depletion of GSH can lead to tumor cell death in vitro, especially in melanocytic cells that generate high levels of oxiradicals [45, 46]. In this study, the effect of *D. sephen* venom on reduced glutathione levels in HeLa cancer cells was examined. The treatment with venom (4, 8, 12, and 16 \( \mu \text{g/mL} \)) decreased GSH levels in HeLa cells. Among all the doses tested, 16 \( \mu \text{g/mL} \) significantly decreased GSH levels in HeLa cells. The resistance in most cases is associated with higher GSH levels within these cancer cells. Thus, approaches to cancer treatment could potentially benefit from a selective GSH depleting strategy [47]. Therefore, our results indicate that the prominent decrease of GSH levels in cancer cells treated with *D. sephen* venom, and further purification of its active constituents could lead to a novel drug candidate.

**Conclusions**

The present findings suggest that *D. sephen* venom initiates cancer cell death by decreasing cell proliferation, antioxidant status and mitochondrial membrane potential; and by increasing intracellular ROS, lipid peroxidation and apoptosis in human cervical (HeLa) cancer cells. The current results clearly demonstrate the involvement of an oxidative mechanism for the antiproliferative effect of *D. sephen* venom on HeLa cells. Thus, marine stingray venoms could be used as a source of antiproliferative agents after further purification in the future.
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Erratum: Antiproliferative activity of marine stingray *Dasyatis sephen* venom on human cervical carcinoma cell line

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Figure 2A inadvertently contained a mistake. The second panel of line (ii) is the same of the third one, of column "D. sephen venom (8 µg/mL)"

The correct version of the figure is:

![Correct Figure 2A](image)

The correction does not affect the discussion or conclusions of the original article.