Simultaneous Silencing of Two Arginine Decarboxylase Genes Alters Development in Arabidopsis

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Polyamines (PAs) are small aliphatic polycations that are found ubiquitously in all organisms. In plants, PAs are involved in diverse biological processes such as growth, development, and stress responses. In *Arabidopsis thaliana*, the arginine decarboxylase enzymes (ADC1 and 2) catalyze the first step of PA biosynthesis. For a better understanding of PA biological functions, mutants in PA biosynthesis have been generated; however, the double *adc1adc2* mutant is not viable in *A. thaliana*. In this study, we generated non-lethal *A. thaliana* lines through an artificial microRNA that simultaneously silenced the two ADC genes (amiR:ADC). The generated transgenic lines (amiR:ADC-L1 and -L2) showed reduced *AtADC1* and *AtADC2* transcript levels. For further analyses the amiR:ADC-L2 line was selected. We found that the amiR:ADC-L2 line showed a significant decrease of their PA levels. The co-silencing revealed a stunted growth in *A. thaliana* seedlings, plantlets and delay in its flowering rate; these phenotypes were reverted with PA treatment. In addition, amiR:ADC-L2 plants displayed two seed phenotypes, such as yellow and brownish seeds. The yellow mutant seeds were smaller than *adc1*, *adc2* mutants and wild type seeds; however, the brownish were the smallest seeds with arrested embryos at the torpedo stage. These data reinforce the importance of PA homeostasis in the plant development processes.

Keywords: artificial microRNA, *Arabidopsis thaliana*, arginine decarboxylase, polyamines, development, seed

INTRODUCTION

In plants, the growth and development processes are tightly regulated through a great variety of molecules with specific functions in a dynamic scenario. Within these molecules, the polyamines (PAs) are recognized as critical regulators in both important processes. Additionally, the PAs also modulate stress responses in plants (Wimalasekera et al., 2011; Gupta et al., 2013; Minocha et al., 2014).

The PAs are small, positively charged, organic molecules; and the mainly four ubiquitous PAs in plants are: diamine putrescine (Put), triamine spermidine (Spd), tetraamines spermine (Spm), and thermospermine (tSpm). Put is synthesized either directly from ornithine by ornithine decarboxylase (ODC) or from arginine by three sequential reactions catalyzed by arginine decarboxylase (ADC), agmatine iminohydrolase (AIH), and N-carbamoylputrescine amidohydrolase (NCPAH). Particularly, in *Arabidopsis thaliana*, the gene encoding ODC is...
absent (Hanfrey et al., 2001; Jiménez-Bremont et al., 2004) so it seems that the ADC pathway is the main route for PA biosynthesis in this plant. Put, is converted into Spd and Spm by the consecutive action of the spermidine synthase (SPDS) and spermine synthase (SPMS), respectively. Both reactions, catalyze the transfer of an aminopropyl group provided by decarboxylated S-adenosylmethionine (dcSAM), which is formed by S-adenosylmethionine decarboxylase (SAMDC) using S-adenosylmethionine (SAM) as a substrate (Kusano et al., 2008). An additional aminopropyl transferase named ACAULIS5 (ACL5), which is present only in plants and some prokaryotic organisms can synthesize the tetraamine tSpm (Minguet et al., 2008).

To elucidate the function of the PAs in diverse biological processes their levels have been manipulated; this can be achieved by biochemical or genetic strategies. In the first one, the use of inhibitors to critical enzymes in PA biosynthesis or application of exogenous PAs has been widely used. However, the use of inhibitors has certain limitations because a lot of them are non-specific being responsible for side effect responses. The second strategy has been generated by both loss-function mutants and gain-function transgenic plants of key enzymes of PA biosynthesis and other enzymes of PA metabolism (Mattoo et al., 2010; Hussain et al., 2011; Tiburcio et al., 2014).

*A. thaliana* has two *ADC* genes: *AtADC1* (AT2G16500) and *AtADC2* (AT4G34710), both encode functional enzymes that catalyze the first rate limiting step in the PA biosynthesis so its manipulation is critical to obtain knowledge of PAs function during plant development. In the latest 90’s, Soyka and Heyer (1999), reported that *A. thaliana* loss-function mutant of *AtADC2* exhibits no obvious phenotype under normal growth conditions but it was impaired to increase the ADC activity by osmotic stress in comparison to the wild type. Moreover, Urano et al. (2004) reported another *A. thaliana* mutant plant of *AtADC2* gene, which was more sensitive to salt stress than wild type plants. Finally, Urano et al. (2005) performed a double mutant of *AtADC1* and *AtADC2*, and reported that both genes are essential for seed development since the double mutant dies during the embryo stage. In all the previous report, it has been inferred a certain degree of functional redundancy between the two isoforms because mostly no obvious phenotype during normal conditions of the single mutants was observed, and since the PA levels did not suffer a drastic change (Soyka and Heyer, 1999; Urano et al., 2004). Nevertheless, the scenario is more complicated because it seems that *A. thaliana* ADC genes are differentially regulated, since *AtADC1* is constitutively expressed, and *AtADC2* is responsive to abiotic stresses, such as salinity, drought, wounding, ABA, MeJA, K⁺ deficiency, and bacterial pathogens (Takahashi and Tong, 2015).

In this study, we performed a simultaneous silencing of the two *ADC* paralogous genes in *A. thaliana* targeting a consensus region between both genes by an artificial microRNA (amiR:ADC). We explored whether the silencing led to a decrease in PA levels and the impact in *A. thaliana* development. Our observations suggest that the silencing strategy was successful due to significant lower expression levels of *AtADC1* and *AtADC2*, and consequently a drastic decrease in PA levels. Furthermore, we compared the amiR:ADC line with parental (Ws) and single *adc1* and *adc2* mutants. Excitingly, amiR:ADC line showed several development defects, such as stunted growth, including a delay in flowering rate, and morphology alteration of seed coat and viability. Our observations suggest that ADC paralogs in *A. thaliana* play a crucial role in plant growth and development during life span.

**MATERIALS AND METHODS**

**Plant Material and Growth Conditions**

Seeds of *A. thaliana* ecotype Wassilewskija (Ws), and the T-DNA insertional mutants for *AtADC1* (*adc1*) and *AtADC2* (*adc2*) were used (Urano et al., 2005). First, the seeds were surface-sterilized for 7 min with 20% (v/v) chlorine solution and then washed four times with sterile distilled water. Next, the seeds were germinated and grown on Murashige and Skoog (MS) 0.5X plates, pH 5.7, containing 0.5% (w/v) sucrose, and 1.2% (w/v) agar (Murashige and Skoog, 1962). Seeds were stratified for 2 days at 4°C in the dark, and then the plates were incubated at 22 ± 2°C in a growth chamber with a 16 h light (120 µmol m⁻² s⁻¹) and 8 h darkness.

**AmiRNA Design, Vector Construction, and Plant Transformation**

The full-length nucleotide sequences of *AtADC1* and *AtADC2* transcripts were used to generate the artificial microRNA in the Web MicroRNA Designer (http://wmd3.weigelworld.org). The amiR:ADC is targeted of a consensus region of 21 nucleotides between the *AtADC* genes (*AtADC1*:1418–1438 and *AtADC2*:1535–1555 nt, respectively). The amiR:ADC was introduced into the *A. thaliana* miR319a precursor sequence by an overlapping PCR, as previously described by Schwab et al. (2006) and Ossowski et al. (2008). For this purpose, the pRS300 vector was used as a template and specific oligonucleotides were employed (Supplemental Table 1). The amiR:ADC precursor was cloned into pCR8/GW/TOPO entry vector (Invitrogen, Carlsbad, CA, USA) and sub-cloned into the pMDC32 binary vector (Curtis et al., 1994). The cloning was performed by site-specific recombination using Gateway LR Clonase II Enzyme Mix (Invitrogen). Finally, pMDC32-amiR:ADC construction was transformed into *Agrobacterium tumefaciens* GV2260 strain to generate *A. thaliana* transgenic lines by “floral dip” method (Zhang et al., 2006). Transformed plants were selected on MS 0.5X medium supplemented with 50 µg/mL hygromycin B. For this study, T4 generation homozygous plants were analyzed.

**RNA Isolation and qRT-PCR Analysis**

The RNA isolation from Ws, amiR:ADC-L1, and -L2 15-day-old seedlings was carried out using Concert reagent (Invitrogen). RNA was purified according to manufacturer’s instructions. The genomic DNA was removed through DNAsel Turbo (Ambion, Austin, TX, USA) treatment. Finally, RNA concentration was quantified using an Epoch 2 microplate reader (Biotek Instruments, Winskosi, USA).

The synthesis and quantification of cDNA were performed through iScript™ kit, One-Step RT-PCR kit, and the SYBR
Green fluorophore, respectively (Applied Biosystems, USA). The qRT-PCR was performed in a 10 μL reaction mixture containing 50 ng of total RNA as template, 5 μL Power SYBR Green RT-Mix (2X), 200 nM oligonucleotides, and 0.08 μL RT enzyme mix (125X) using the direction system StepOne Real-Time PCR with the StepOne V2.1 software (Applied Biosystems, USA) as described previously (Rodríguez-Hernández et al., 2014; Ortega-Amaro et al., 2015). The gene expression was analyzed with the following conditions: 30 min at 48°C (cDNA synthesis), 10 min at 95°C (activation of AmpliTaq Gold DNA) following by 40 cycles of PCR for 15 s at 95°C (denaturation) and 1 min at 60°C (annealing/extension). Melting curves were performed with cycles of 15 s at 95°C (denaturation), 15 s at 60°C (alignment), and 15 s to 95°C (denaturation), incrementing 0.3°C per cycle. The A. thaliana ubiquitin 5 (UBQ5) gene was used as expression control. The relative gene expression levels of AtADC1 and AtADC2 genes were presented as 2^−ΔCt, where Δ Ct = CtAtADC1 or AtADC2−CtUBQ5 (Livak and Schmittgen, 2001). The oligonucleotides that we used are described in Supplemental Table 1. For each sample, three biological replicates (n = 3) were analyzed with their respective technical replicates. Experiments were repeated at least twice with similar results.

**Free Polyamines Extraction and Quantification**

The A. thaliana free polyamines (Put, Spd, and Spm + tSpm) were determined as dansyl-derivatives by reversed phase HPLC (high performance liquid chromatography) approach. PAs extraction was done with 0.2 g plant material homogenized in 0.8 mL 5% (v/v) perchloric acid and incubated overnight at 4°C. After centrifugation, 0.2 mL of the supernatants was dansylated in a mixture containing 0.1 mL of saturated Na2CO3, 0.2 mL dansylchloride (5 mg/mL acetone), and 1 mM 1,7-diaminoheptane as internal standard. The mixture was incubated overnight in darkness at room temperature. Reaction was stopped by adding 0.1 mL proline (100 mg/mL), and dansylated PAs were extracted with 0.4 mL toluene. Finally, the organic phase was vacuum-evaporated and dansylated PAs were dissolved in 0.05 mL acetonitrile.

Plant PAs were analyzed by HPLC using a 4.6 × 150 mm C18 reverse phase column, in which the flow was 1.5 mL/min, and the elution gradient was prepared with eluent A (water) and eluent B (acetonitrile). Then, it was equilibrated with 70% B and 30% A before injecting 0.01 mL samples. This was followed by a linear gradient ending with 100% B during 9 min. The final step was held for 4 min before regenerating the column. Detection was done with a fluorometer using excitation and emission wavelengths of 415 and 510 nm, respectively, according to Flores and Galston (1982). A relative calibration procedure was used to determine the PAs in the samples, using 1,7-diaminoheptane as the internal standard and PA concentrations ranging from 0.3 to 1.5 nmol (Sigma-Aldrich Mexico). The results were expressed as nmols per gram of fresh weight (nmol/gFW).

**Measurement of Fresh Weight, Primary Root Length, Lateral Root Number, and Seed Weight**

The A. thaliana seeds from Ws, adc1, adc2, and amiR:ADC-L2 lines were grown on 0.5X MS plates for 7 and 14 days. Fresh weight (mg) of seedlings was obtained on an analytical scale and the values obtained represent the means from seven groups comprised of four seedlings per line (n = 7). Subsequently, the estimation of primary root length (cm) was determined by measuring the length of 28 roots of each line (n = 28). The image analysis software IMAGE J (http://rsb.info.nih.gov/ij) was used to evaluate primary root length. The lateral root number was measured using a stereomicroscope Motic SMZ-143 (n = 28). Seed weight was calculated from three replicates for each line (Ws, adc1, adc2, and amiR:ADC-L2), where 200 seeds represent each replicate (n = 3). In particular for amiR:ADC-L2 line seeds, 200 yellow and 200 brownish seeds were estimated. Each seed lot (time post-harvest) was measured on an analytical scale, and weights are expressed in mg. All measurements were repeated at least twice with similar results.

**Measurement of Flowering Time**

Ten days-old Ws, adc1, adc2, and amiR:ADC-L2 seedlings grown on 0.5X MS plates were transferred to soil pots into a growth chamber under 16 h of 120 μmol m⁻² s⁻¹ light and 8 h of darkness. The flowering time was estimated by recording the number of days after sowing in which the inflorescence reached 1 cm in length. The flowering time data was represented as the percentage of plants with floral tissues (n = 10).

**Exogenous Polyamine Application**

The Ws and amiR:ADC-L2 stratified seeds (2 days at 4°C) were germinated and grown on 0.5X MS plates into a growth chamber for 7 days. After, the seedlings were transferred to divided plates, containing both 0.5X MS medium and 0.5X MS supplemented with 10 μM each PA (Put, Spd, and Spm). The plates were incubated into a grown chamber for 9 days. Subsequently, the mean of fresh weight and primary root length per plant were calculated (n = 15). In the case of flowering rate recovering, amiR:ADC-L2 14-day-old seedling grown on 0.5X MS plates were transferred to soil pots and their aerial tissues were sprayed with a solution containing 10 μM Put for 15 days.

**Microscopic Analyses**

The immature and mature siliques from Ws, adc1, adc2, and amiR:ADC-L2 were dissected from the plant, and the valves were gently removed from siliques using forceps and scalp. The septum and seeds were observed under a stereomicroscope (Motic SMZ-143). The magnification for the images captured was 15x to mature siliques, and 40x for immature siliques. Ten immature and mature siliques of each line were analyzed, and a representative image is shown in each column. For scanning electron microscopy (SEM) analysis, dry seeds were glued onto pure carbon containing polymer films and they were examined using a high-resolution scanning electron microscope (FEI ESEM-QUANTA 200; Low Vacuum/Water).
operating at 25 kV and 90–100 Pa. Micrographs were digitalized using the basic photo program from the OS X system (Apple, CA, USA). Ten seed and embryos of each line were analyzed, and a representative image is shown in each column.

**Statistical Analyses**

One-Way Analysis of Variance (ANOVA) and Tukey's post-test analyses were performed to assess statistical significance. The GraphPad Prism version 5.0b (GraphPad Software, San Diego, California, USA) was used. Data represent the mean ± SEM. Differences at $P < 0.05$ were considered significant.

**RESULTS**

**Silencing of *A. thaliana* ADC1 and ADC2 Genes by Artificial microRNA**

In order to simultaneously silence both ADC genes from the *A. thaliana*, we generated an artificial microRNA (amiR:ADC) using the Web MicroRNA Designer (WMD; Schwab et al., 2006; Ossowski et al., 2008). The designed amiR:ADC is targeted to consensus region of 21 nucleotides between the AtADC genes (AtADC1:1418–1438 bp and AtADC2:1535–1555 bp; Figure 1A). As is shown in the alignment, exist one mismatch at position 17 for AtADC2 target, and other mismatch for both mRNAs at 1 and 21 positions (Figure 1B). These mismatches should reduce potential transitive effects due to priming and extension by RNA dependent RNA polymerase (Ossowski et al., 2008). Besides, there is no mismatch at the cleavage site (position 10–11). We analyzed the expression levels of each AtADC1 and AtADC2 genes by qRT-PCR approach in the generated *A. thaliana* transgenic lines. In this sense, a strong reduction of both AtADC transcript levels was observed in the amiR:ADC-L1 and -L2 transgenic lines (8 to 9.6 fold for AtADC1 and 6 fold for AtADC2) compared to the expression of wild type (Ws) seeds (Figure 1C). Our data show that the expression of designed amiR:ADC reduce the transcript levels from both *A. thaliana* AtADC1 and AtADC2 genes. For the following experiments amiR:ADC-L2 line was used.

**The amiR:ADC Seedlings Show Reduced Polyamine Levels**

Since AtADC1 and AtADC2 are rate-limiting enzymes for PA biosynthesis, we explored whether the silencing led to a decrease in PA levels. We quantified the total free PAs content (Put, Spd, and Spm + tSpm) in the Ws, adc1 and adc2 single mutants, and amiR:ADC-L2 12-day-old seedlings using HPLC technique. Our analyses reveal that the amiR:ADC-L2 line contain the lowest PA levels in contrast to all lines evaluated; even, the Put levels were lower than in the adc2 mutant, which it has been known to show a reduction of their Put levels (Urano et al., 2004). In particular, the Put, Spd, and Spm + tSpm levels were reduced 85, 88, and 89%, respectively, compared to the parental Ws (Figure 1D). These results indicate that the simultaneous silencing of *A. thaliana* AtADC genes impact severely in their PAs content.

**The amiR:ADC Seedlings Display a Stunted Growth Phenotype**

In order to evaluate the physiological roles of diminished PA level in amiR:ADC-L2 line, we have analyzed the growth rate, such as fresh weight, primary root length, and lateral root number of the Ws, adc1, adc2, and amiR:ADC-L2 seedlings grown on MS medium. Seven days-old amiR:ADC-L2 seedlings showed 25% fresh weight reduction in contrast to the wild type Ws (2.9 ± 0.2 and 3.9 ± 0.2 mg, respectively; Figures 2A,B); however, significant differences were not found in the fresh weight of adc1 and adc2 lines respect to Ws (Figure 2B). Likewise, the amiR:ADC-L2 seedlings disclosed impaired development, which is reflected in a 30% reduction of primary root length compared to parental Ws (0.9 ± 0.03 and 1.3 ± 0.04 cm, respectively). In contrast, there was no difference in the main root length of adc1 and adc2 lines (Figure 2C). Additionally, we observed a significant augment of the lateral roots number in amiR:ADC-L2 (1.5 ± 0.2) in comparison to Ws (0.6 ± 0.1), the adc1, adc2 lines (1 ± 0.2 and 0.9 ± 0.1, respectively) showed increment of lateral root number compared to the parental line (Figure 2D).

When the plantlets were 14 days old, we observed similar behavior in the physiological parameters that were measured (Figure 2E). The amiR:ADC-L2 plantlets reduced their fresh weight in 31% respect to the Ws plantlets (11 ± 0.5 and 16 ± 0.7 mg, respectively). Also, a significant difference was found in the fresh weight of adc1 (12 ± 0.9 mg) in comparison to Ws (Figure 2F). The primary root length was diminished 38% in the amiR:ADC-L2 line compared to the Ws (2.1 ± 0.1 and 3.4 ± 0.1, respectively; Figure 2G). Noticeably, the lateral roots number was higher in the amiR:ADC-L2 than Ws, adc1, and adc2 lines (Figure 2H). Additionally, we evaluated the independent transgenic line amiR:ADC-L1, which showed a slightly reduction of the primary root length in 7 days and it increased the number of lateral roots in 7 and 14 days. However, this line did not present the pronounced phenotype that was observed in the amiR:ADC-L2 line (Supplemental Figure S1). These data suggest that the PA homeostasis is necessary for normal early development in *A. thaliana*.

**PA Additions Rescued the Stunted Growth of amiR:ADC Seedlings**

In addition, we were interested in exploring if the stunted growth was reverted by the exogenous addition of PAs. For this purpose, amiR:ADC-L2 7-day-old seedlings were grown in the right side of split Petri dishes in presence of 10 μM Put, Spd, or Spm (Figure 3A). In the left side of the split dish, the Ws seedlings without PA treatment were grown as control. Both physiological parameters: fresh weight and primary root length (Figures 3B,C, respectively) increased in the silenced plants after Put, Spd or Spm treatments, being able to reach the parental levels. However, discrete differences were observed between PAs, as the Spm generated the lower recovery in the primary root length in comparison to Put and Spd treatments.
FIGURE 1 | The artificial microRNA amirR:ADC is effective to silence both ADC genes and decreased the polyamine levels of *A. thaliana*. (A) Schematic representation of *AtADC1* and *AtADC2* transcripts. Target regions of the artificial microRNA are indicated in red. (B) Multiple alignments of *AtADC1* and *AtADC2* target region with the predicted mature amiR:ADC sequence (5′-3′). (C) Transcript levels of *AtADC1* and *AtADC2* genes estimated in Ws, amirR:ADC-L1, and -L2 lines by qRT-PCR approach. The *A. thaliana* ubiquitin 5 (*UBQ5*) gene was used as expression control and the relative gene expression levels of *AtADC1* and *AtADC2* genes were presented as $2^{-\Delta\Delta Ct}$, where $\Delta Ct = Ct_{AtADC1} - Ct_{UBQ5}$. (D) Quantification of free polyamine levels [putrescine (Put), spermidine (Spd), and spermine + thermospermine (Spm + tSpm)] in Ws, *adc1*, *adc2*, and amirR:ADC-L2 12-day-old seedlings using HPLC technique. Error bars denote SE (n = 3) and different letters are used to indicate means that differ significantly according to the One-way ANOVA analysis and Tukey’s multiple comparison test ($P < 0.05$).
FIGURE 2 | Stunted growth of the amiR:ADC-L2 seedlings. (A,E) Phenotype of 7- and 14-day-old Ws, adc1, adc2, and amiR:ADC-L2 seedlings grown on MS plates, respectively. The scale bar corresponds to 1 cm. Measure of fresh weight (B,F), primary root length (C,G), and number of lateral roots (LR) (D,H). To fresh weight the values represent means of seven groups of four seedlings (n = 7). To primary root length and number of lateral roots (LR) error bars represent means SE (n = 28) and different letters are used to indicate means that differ significantly according to the One-way ANOVA analysis and Tukey’s multiple comparison test (P < 0.05).
Co-Silencing of ADC Genes Alters Flowering Rate in A. thaliana

We were able to find a visible change from flower initiation in the amiR:ADC-L2 transgenic line. We found that silenced plants showed a delay in flowering time (up to 9 days) relative to Ws under long day conditions (Figures 4A,B). In addition, the adc2 mutant line displayed a slight delay of 2 days in the inflorescence emergence in comparison to Ws (Figure 4B). Previously, we observed that the stunted growth in 16-day-old amiR:ADC-L2 seedlings was reverted with the applying of Put (Figure 3). Consequently, we analyzed whether the flowering time could be recovered after Put treatment (Figure 4). In this sense, 14-day-old amiR:ADC-L2 seedlings that were sprinkled with 10 µM Put for 15 days showed an reduce of 5 days in their flowering time in comparison to non-treated plants (Figures 4A,B).

Afterwards, we investigated the phenotypic characteristics of 45- and 55-day-old amiR:ADC-L2 plants growth under a long day condition (Figure 5). At 45 days, the amiR:ADC-L2 plants remained with the slow growth pattern (Figure 5A). Another feature observed in the amiR:ADC-L2 mutant line was the serrate rosette leaves (Figure 5A). Furthermore, we examined the number of siliques in 55 day-old-plants. Our data shows that the amiR:ADC-L2 had a low number of siliques at this stage compared to the wild type Ws (Figure 5B).

The Seed Development is Affected by Co-Silencing of ADC Genes

We found a striking phenotype in amiR:ADC-L2 seeds, suggesting that the seed development is also altered in the silenced plants. As we can notice in the Figure 6A, stereomicroscope analyses showed that amiR:ADC-L2 immature and mature siliques had some abortive seeds. Because it was clear that the seeds were altered in amiR:ADC siliques, we were interested in analyzing the number of seeds and weight. Our data showed approximately 20% reduction of seed number in amiR:ADC-L2 siliques compared to parental Ws (40 ± 2 and 48 ± 2 seed number per siliques, respectively; Figures 6A,B); similarly, the adc2 single mutant produced less seeds per silique (42 ± 2; Figure 6B). Stereomicroscope analyses reveal that both amiR:ADC-L2 immature and mature siliques contain a 80% of normal seeds (large and yellow) and a 20% of abortive seeds (shrunken and brownish; Figures 6A,C). The phenotype of abortive seeds in mature siliques from amiR:ADC-L2 was reverted by exogenously addition of Put (Supplemental Figure S2). Additionally, we analyzed the weight of 200 seeds of

FIGURE 3 Polyamine application alleviates the stunted phenotype of amiR:ADC-L2 seedlings. (A) Ws, adc1, adc2, and amiR:ADC-L2 16-day-old seedlings were grown on split plates containing both MS medium (left side) and MS supplemented with 10 µM Put, Spd, or Spm (right side). The scale bar corresponds to 1 cm. Measure of fresh weight (B), and primary root length (C) of the plantlets. The mean of fresh weight is presented as mg per plant. Error bars denote SE (n = 15) and different letters are used to indicate means that differ significantly according to the One-way ANOVA analysis and Tukey's multiple comparison test (P < 0.05).
FIGURE 4 | The amiR:ADC-L2 plants are altered in their flowering rate. (A) Phenotype of 35-day-old Ws, adc1, adc2, and amiR:ADC-L2 plants with or without putrescine (Put) treatment grown on soil pots under long day conditions. The plants were sprinkled with 10 µM Put for 15 days. The scale bar corresponds to 2.5 cm. (B) Percentage of flowering rate of Ws, adc1, adc2, and amiR:ADC-L2 plants under long day conditions (n = 10). The percentage of flowering rate recovering is showed with red rhombus in (B).

each line, including the 2 types of seeds of the amiR:ADC-L2. The weight of Ws and adc1 mutant seeds did not present significant differences between them. In contrast, the weight of adc2 mutant and amiR:ADC-L2 yellow seeds had decreased 16 and 21%, respectively, compared to the Ws (Figure 6D). In addition, the reduction in the brownish seeds, which had severe defects in cell morphology, recorded a weight of 0.5 ± 0.03 mg per 200 seeds in comparison to 3.5 ± 0.07 mg in Ws (Figure 6D).

Finally, the scanning electron microscopy (SEM) approach was used to analyze the external morphology alterations in amiR:ADC-L2 seeds. The micrographs revealed that the cells surface from Ws, adc1, adc2, and amiR:ADC-L2 (yellow) seeds showed similar polygonal structures and central elevation, in contrast to brownish seeds of amiR:ADC-L2 that showed an irregular shape (Figure 7A). In particular, the brownish seeds exhibited a reduction of 28 and 40% in length and width seed, respectively, compared to Ws (Figures 7B,C). We therefore examined the embryo development of amiR:ADC-L2 seeds and found that the brown seeds had a late arrest in development of embryos corresponding at torpedo stage (Figure 7A).

DISCUSSION

Plant growth and development processes involve environmental and endogenous signals that contribute to determine the anatomy, physiology and molecular features of the plant. Within the endogenous signals, the polyamines (PAs) have been recognized in higher plants as key players in these biological processes (Mattoo et al., 2010; Takahashi and Kakehi, 2010; Tiburcio et al., 2014). Moreover, the literature indicates that PAs are implicated in a variety of cellular processes, including control cell division, root formation, flowering, retardation of senescence, and response to stress (Watson et al., 1998; Pandey...
et al., 2000; Alcázar et al., 2005; Jiménez-Bremont et al., 2007; Ortega-Amaro et al., 2012).

We generated a non-lethal A. thaliana line with very low levels of major polyamines by using an artificial microRNA amiR:ADC. An effective tool in plants to silence genes in a simultaneously manner, when the single or multiple mutants have a lethal phenotype, is made through an artificial microRNA because it provides the opportunity to analyze the mutant lines during life span. In this sense, the artificial miRNA design targets a consensus region in both ADC genes, and the fact that the two ADC genes in A. thaliana are attractive targets of genetic manipulation is because this plant lacks of the ODC gene (Hanfrey et al., 2001), thus biosynthesis de novo of PA is only through the ADC route. Plants with lowest levels of PAs are desired in order to provide further evidence of the importance of PA homeostasis in plant growth and development processes.

Important biochemical and genetic efforts have been performed in order to manipulate the ADC expression or activity that have negative impact in PAs levels and their association with defects in development. For example, in the latest 80’s the DL-alpha difluoromethylarginine (DMFA), an irreversible inhibitor of ADC activity, was commonly used in plants (Slocum and...
FIGURE 6 | The seed development is affected in amiR:ADC-L2 plants. (A) Mature and immature siliques (left and right panel, respectively) from 55-day-old Ws, adc1, adc2, and amiR:ADC-L2 plants grown under long day conditions. The white arrows indicate the abortive seeds distribution in amiR:ADC-L2 siliques. In the left and right panels the scale bars correspond to 1500 and 500 µm, respectively. (B) Seed number per mature siliques from 55-day-old plants. (C) Percentage of both yellow (Y) and brownish (B) seeds per siliques in the amiR:ADC-L2. The measures were carried out using 10 siliques per plant of 10 plants (n = 10). (D) Weight of 200 seeds from mature siliques of Ws, adc1, adc2, and amiR:ADC-L2. To amiR:ADC-L2 lines and their normal (Y) and abortive (B) seeds were separated and analyzed individually (n = 3). Error bars denote SE and different letters are used to indicate significant differences among lines according to the One-way ANOVA analysis and Tukey's multiple comparison test (P < 0.05).
Albino mutants (Urano et al., 2004). An alternative strategy is the use of RNA interference (RNAi) and microRNA (miRNA) methods. The miRNA approach has been demonstrated to be more specific than the RNAi approach and to have a lower toxicity. Therefore, the miRNA approach was used to downregulate the expression of the AtADC genes. The AtADC miRNA approach was achieved by introducing an AtADC-specific miRNA (amiR:ADC) into Arabidopsis plants through Agrobacterium-mediated transformation. The resulting plants exhibited a range of phenotypic changes, including stunted growth, delayed flowering, and altered morphology. The phenotypic changes observed are consistent with the role of PAs in plant development. These results suggest that the AtADC genes play a crucial role in Arabidopsis development and provide a new approach for manipulating PA metabolism in Arabidopsis.
Besides, we detected an altered morphology of the seeds reflected by lower weight and fewer seed number per siliquae in bothadc2 and amir:ADC Arabidopsis A. thaliana plants. In particular, the amir:ADC siliquae contain 20% of abortive seeds, in which the embryo was arrested in torpedo stage. As expected, this phenotype was reverted by exogenously addition of Put. In this sense, Urano et al. (2005) reported that the A. thaliana siliquae from heterozygous double mutant of ADC genes contain 25% of abortive seeds; moreover, all the homozygous ADC mutated seeds are unable to germinate since their embryos are arrested at the heart to torpedo transition stage. Also, they described the tissue-specific expression of the promoter of AtADC1 and AtADC2 genes and they found that AtADC2 promoter was detected in siliquae and through the early developmental stages of embryo, endosperm, and still active during embryo maturation; remarkably, the AtADC1 promoter was not detected in these tissues. So it seems that AtADC2 is more functional during embryo development in contrast to AtADC1. Nevertheless, the amir:ADC-L2 line showed a more drastic negative phenotype in the seeds in comparison with the single mutantadc2. On the other hand, it has recently been reported in wheat that exogenous application of Spd and Spm increased the grain filling rate and weight (Liu et al., 2013). The importance of PAs during embryo maturation is also recognized by the facts that mainly accumulate during early stages of seed development (Urano et al., 2005).

In addition to double mutantadc1/adc2, the double spds1/spds2 mutant is also lethal at embryo stages (Imai et al., 2004). Thus, it is clear that is not possible to generate a polyamine-auxotrophic plant with traditional approaches, so it seems that the ADC silencing via an artificial miRNA strategy can provide us with valuable information regarding to the minimum levels that are required for optimum plant development, and also for a more detailed description of the growth defects associated with this genetic background. In this study, the phenotypic characterization of the simultaneous silencing of two ADC genes in A. thaliana reinforces the role of PAs during growth and development. This genetic background will be a very useful tool for further molecular studies in order to enlighten and to give more information related to the specific functions of PAs during homeostasis in plant development processes, and in stress responses.

**AUTHOR CONTRIBUTIONS**

DS, AC, AR, IM, and JJ designed and carried out the experiments, analyzed the results, and wrote the manuscript. KU and KS contributed scientific advice, and wrote the manuscript. All authors have read and approved the final manuscript.

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**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: http://journal.frontiersin.org/article/10.3389/fpls.2016.00300

**REFERENCES**

Alcázar, R., García-Martínez, J. L., Cuevas, J. C., Tiburcio, A. F., and Altabella, T. (2005). Overexpression of ADC2 in Arabidopsis induces dwarfism and late-flowering through GA deficiency. Plant J. 43, 425–436. doi: 10.1111/j.1365-313X.2005.02465.x

Curts, I. S., Power, J. B., Blackhall, N. W., de Laat, A. M. M., and Davey, M. R. (1994). Genotype-independent transformation of lettuce using Agrobacterium tumefaciens. J. Exp. Bot. 45, 1441–1449. doi: 10.1093/jxb/45.10.1441

Flores, H. E., and Galston, A. W. (1982). Analysis of polyamines in higher plants by high performance liquid chromatography. Plant Physiol. 69, 701–706. doi: 10.1104/pp.69.3.701

Gupta, K., Dey, A., and Gupta, B. (2013). Plant polyamines in abiotic stress responses. Acta Physiol. Plant. 35, 2015–2036. doi: 10.1007/s11738-013-1239-4

Hanfrey, C., Sommer, S., Mayer, M. J., Burtin, D., and Michael, A. J. (2001). Arabidopsis polyamine biosynthesis: absence of ornithine decarboxylase and the mechanism of arginine decarboxylase activity. Plant J. 27, 551–560. doi: 10.1046/j.1365-313X.2001.01100.x

Hanzawa, Y. (2000). ACAULIS, an Arabidopsis gene required for stem elongation, encodes a spermine synthase. EMBO J. 19, 4248–4256. doi: 10.1093/emboj/19.16.4248

Hussain, S. S., Ali, M., Ahmad, M., and Siddique, K. H. M. (2011). Polyamines: natural and engineered abiotic and biotic stress tolerance in plants. Biotechnol. Adv. 29, 300–311. doi: 10.1016/j.biotechadv.2011.01.003

Imai, A., Matsuyama, T., Hanzawa, Y., Akiyama, T., Tamaoki, M., Saji, H., et al. (2004). Spermidine synthase genes are essential for survival of Arabidopsis. Plant Physiol. 135, 1565–1573. doi: 10.1104/pp.104.041699

Jiménez-Bremont, J. F., Camacho-Villasana, Y. M., Cabrera-Ponce, J. L., de la Rosa, A. P. B., and Ochoa-Alejo, N. (2004). Sequence comparison of plant ornithine decarboxylases reveals high homology and lack of introns. Biol. Plant. 48, 193–198. doi: 10.1023/B:BIOP.0000033444.17603.f2

Jiménez-Bremont, J. F., Ruiz, O. A., and Rodríguez-Kessler, M. (2007). Modulation of spermidine and spermine levels in maize seedlings subjected to long-term salt stress. Plant Physiol. Biochem. 45, 812–821. doi: 10.1016/j.phyto.2007.08.001

Kaur-Sawhney, R., Tiburcio, A. F., and Galston, A. W. (1988). Spermidine and flower-bud differentiation in thin-layer explants of tobacco. Planta 173, 282–284. doi: 10.1007/BF00403022

Kusano, T., Berberich, T., Tateda, C., and Takahashi, Y. (2008). Polyamines: essential factors for growth and survival. Planta 228, 367–381. doi: 10.1007/s00425-008-0772-7

Liu, Y., Gu, D., Wu, W., Wen, X., and Liao, Y. (2013). The Relationship between polyamines and hormones in the regulation of wheat grain filling. PLoS ONE 8:e78196. doi: 10.1371/journal.pone.0078196

Livak, K. J., and Schmittgen, T. D. (2001). Analysis of relative gene expression using real-time quantitative PCR and the 2−ΔΔCT method. Methods 25, 402–408. doi: 10.1006/meth.2001.1262

Mattoo, A. K., Minocha, S. C., Minocha, R., and Handa, A. K. (2010). Polyamines and cellular metabolism in plants: transgenic approaches reveal different
responses to diamine putrescine versus higher polyamines spermidine and spermine. Amino Acids 38, 405–413. doi: 10.1007/s00726-009-0399-4

Minguez, E. G., Vera-Sirera, F., Marina, A., Carbonell, J., and Blázquez, M. A. (2008). Evolutionary diversification in polyamine biosynthesis. Mol. Biol. Evol. 25, 2119–2128. doi: 10.1093/molbev/msn161

Minocha, R., Majumdar, R., and Minocha, S. C. (2014). Polyamines and abiotic stress in plants: a complex relationship. Front. Plant Sci. 5:175. doi: 10.3389/fpls.2014.00175

Murashige, T., and Skoog, F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol. Plantarum 15, 473–497. doi: 10.1111/j.1399-3054.1962.tb08052.x

Naka, Y., Watanabe, K., Sagar, G. H. M., Niitsu, M., Arumugam Pillaí, M., Minocha, R., Majumdar, R., and Minocha, S. C. (2014). Polyamines and Murashige, T., and Skoog, F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol. Plantarum 15, 473–497. doi: 10.1111/j.1399-3054.1962.tb08052.x

Ortega-Amaro, M. A., Rodríguez-Hernández, A. A., Rodríguez-Kessler, M., Herrández-Lucero, E., Rosales-Mendoza, S., Ibáñez-Salazar, A., et al. (2015). Overexpression of AtGRDP2, a novel glycine-rich domain protein, accelerates plant growth and improves stress tolerance. Front. Plant Sci. 5:782. doi: 10.3389/fpls.2014.00782

Ortega-Amaro, M. A., Rodríguez-Kessler, M., Becerra-Flora, A., and Jiménez-Bremont, J. F. (2012). Modulation of Arabidopsis CYCB1 expression patterns by polyamines and salt stress. Acta Physiol. Plant. 34, 461–469. doi: 10.1007/s11738-011-0842-5

Ossowski, S., Schwab, R., and Weigel, D. (2008). Gene silencing in plants using artificial microRNAs and other small RNAs. Plant J. 53, 674–690. doi: 10.1111/j.1365-313X.2007.03328.x

Pandey, S., Ranade, S. A., Nagar, P. K., and Kumar, N. (2000). Role of polyamines and ethylene as modulators of plant senescence. J. Biosci. 25, 291–299. doi: 10.1007/BF02703938

Rodríguez-Hernández, A. A., Ortega-Amaro, M. A., Delgado-Sánchez, P., Salinas, J., and Jiménez-Bremont, J. F. (2014). ATGRDP1 Gene encoding a glycine-rich domain protein is involved in germination and responds to ABA signalling. Plant Mol. Biol. Report. 32, 1187–1202. doi: 10.1007/s11105-014-0714-4

Schwab, R., Ossowski, S., Riester, M., Warthmann, N., and Weigel, D. (2006). Highly specific gene silencing by artificial microRNAs in Arabidopsis[WI][OA]. Plant Cell 18, 1121–1133. doi: 10.1105/tpc.105.039834

Scolum, R. D., and Galston, A. W. (1985). Changes in polyamine biosynthesis associated with postfertilization growth and development in tobacco ovary tissues. Plant Physiol. 79, 336–343.

Soyka, S., and Heyer, A. G. (1999). Arabidopsis knockout mutation of ADC2 gene reveals inducibility by osmotic stress. FEBS Lett. 458, 219–223. doi: 10.1016/S0014-5793(99)01125-4

Takahashi, T., and Kakehi, J.-I. (2010). Polyamines: ubiquitous polycations with unique roles in growth and stress responses. Annu. Bot. 105, 1–6. doi: 10.1093/aob/mcp259

Takahashi, T., and Tong, W. (2013). “Regulation and diversity of polyamine biosynthesis in plants,” in Polyamines, eds T. Kusano and H. Suzuki (Tokyo: Springer), 27–44.

Tiburcio, A. F., Altabella, T., Bitrián, M., and Alcázar, R. (2014). The roles of polyamines during the lifespan of plants: from development to stress. Planta 240, 1–18. doi: 10.1007/s00424-014-2055-9

Urano, K., Hobo, T., and Shinozaki, K. (2005). Arabidopsis ADC genes involved in polyamine biosynthesis are essential for seed development. FEBS Lett. 579, 1557–1564. doi: 10.1016/j.febslet.2005.01.048

Urano, K., Yoshiba, Y., Nanjo, T., Igarashi, Y., Seki, M., Sekiguchi, F., et al. (2003). Characterization of Arabidopsis genes involved in biosynthesis of polyamines in abiotic stress responses and developmental stages. Plant Cell Environ. 26, 1917–1926. doi: 10.1046/j.1365-3040.2003.01108.x

Urano, K., Yoshiba, Y., Nanjo, T., Ito, T., Yamaguchi-Shinozaki, K., and Shinozaki, K. (2004). Arabidopsis stress-inducible gene for arginine decarboxylase AtADC2 is required for accumulation of putrescine in salt tolerance. Biochem. Biophys. Res. Commun. 313, 369–375. doi: 10.1016/j.bbrc.2003.11.119

Watson, M. B., Emory, K. K., Piatak, R. M., and Malmberg, R. L. (1998). Arginine decarboxylase (polyamine synthesis) mutants of Arabidopsis thaliana exhibit altered root growth. Plant J. 13, 231–239. doi: 10.1046/j.1365-313X.1998.0027.x

Wimalasekera, R., Tebartz, F., and Scherer, G. F. E. (2011). Polyamines, polyamine oxidases and nitric oxide in development, abiotic and biotic stresses. Plant Sci. 181, 593–603. doi: 10.1016/j.plantsci.2011.04.002

Zhang, X., Henriques, R., Lin, S.-S., Niu, Q.-W., and Chua, N.-H. (2006). Agrobacterium-mediated transformation of Arabidopsis thaliana using the floral dip method. Nat. Protoc. 1, 641–646. doi: 10.1038/nprot.2006.97

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