The effect of lidocaine on TRPM 2, 6, 7 and 8 channels in liver ischemia / reperfusion model in rats

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Abstract

Aim: The transient receptor potential melastatin (TRPM) channel is subfamilies of the transient receptor potential (TRP) channels, cation permeable channels. Ischemia and reperfusion (I/R) injury is a phenomenon highlighting cellular damage of the liver caused by oxygen. Lidocaine is a local anesthetic that blocks sodium channels and suppresses mitochondrial functions of neutrophils. In this study, we propose to investigate the effects of TRPM2/8 and TRPM6/7 expressions after lidocaine treatment in liver ischemia/reperfusion rat model.

Materials and Methods: The study was carried out on 32 male Wistar rats. The animals were randomly divided into 4 groups including sham, lidocaine, I/R group and I/R-lidocaine group. The portal vein and the hepatic artery branches were clamped for 60 minutes for complete ischemia. TRPM2/8 and TRPM6/7 gene expression levels were assessed by RT-qPCR.

Results: The expression levels of TRPM2 were significantly higher in liver-I/R group compared to sham and lidocaine groups (p<0.05, p<0.000, respectively), TRPM8 genes were significantly higher in liver-I/R group compared to sham and lidocaine groups (p<0.05, p<0.05, respectively). However, the expression levels of TRPM6 gene was significantly higher in liver-I/R group compared to sham, lidocaine, and I/R-lidocaine groups (p<0.05, p<0.01, p<0.05, respectively), TRPM7 genes was significantly higher in liver-I/R group compared to sham, lidocaine and I/R-lidocaine groups (p<0.05, p<0.000, p<0.05, respectively).

Discussion: In conclusion, we firstly showed that an association between the expression level of TRPM2/8, TRPM6/7, and hepatic I/R, I/R-lidocaine groups as well as TRPM2/8 and TRPM6/7 gene expressions are affected by lidocaine in the liver-I/R in a rat model.

Keywords
Sodium channel; Ischemia; Reperfusion; TRPM; Lidocaine
Introduction

The transient receptor potential (TRP) channels are a form of calcium channels and they transport magnesium, calcium, trace metal ions and modulate the driving force for ion entry [1, 2]. TRPs are present in all cellular membranes, with the exception of the nuclear envelope and mitochondria [3]. TRPs are localized in the plasma membrane and almost every cell type expresses them [2]. TRPs multigene superfamily encodes integral membrane proteins and is divided into seven subfamilies: TRPC (canonical), TRPM (melastatin), TRPV (vanilloid), TRPA (ankyrin transmembrane protein), TRPML (mucolipin), TRPN (nom PC-like) and TRPP (polycystin), [4]. The members of the TRPM family (Melastatin) were divided into 4 groups according to the sequence similarity (TRPM1 / 3, TRPM2 / 8, TRPM4 / 5 and TRPM6 / 7) [4]. TRPM2 is most commonly expressed in the brain but is found in various peripheral cell types. TRPM2 is also suggested to play a role in oxidative stress / reactive oxygen species, TNF-α mediated Ca2+ influx and cell death [5]. TRPM6 and 7 show high permeability to Ca2+ and magnesium. In accordance with the role of TRPM6 in Mg+2 homeostasis, the decrease in intracellular Mg+2 concentrations activate the channel [6]. TRPM7 also plays a role in the regulation of neuronal cell death and cell cycle [7]. TRPM8 is activated by cooling (<26˚C) and by menthol. TRPM8 was originally cloned from human prostate but is also expressed in many types of tumor tissues, such as breast, colon, lung, and skin [8]. Intracellular calcium has been described as an important secondary messenger ion, and high calcium concentration leads to compressive compensatory vasodilatation after a vasoconstriction period [9]. All of these causes, including lack of oxygen and nutrients, initiate apoptosis and necrosis [10]. Ischemia and hypoxia cause failed permeability of cell membranes to a high level of free intracellular calcium which causes ischemic liver tissue injury. Liver ischemia-reperfusion (I/R) remains a major problem after partial hepatectomy and transplantation [11, 12].

Lidocaine is a local anesthetic that blocks sodium channels, shows anti-inflammatory effects due to its ability to inhibit superoxide formation and leukocyte metabolic activity [13]. Thus, lidocaine is effective for flap recovery after reperfusion injury [14]. The voltage-clamped studies showed that increasing the calcium in the external medium affects sodium and potassium conductance versus voltage curves. Moreover, treatment with a high calcium leads to an increase in the time to peak of the sodium level [15]. Thus, our hypothesis is that the sodium channel blocker (lidocaine) affects calcium level or calcium channels or vice versa.

There is a limited study on the use of sodium channel blockers to prevent ischemia-reperfusion liver injury. However, there are few studies on the relationship between sodium channel blocker administration and TRPM channels [9]. To understand the roles of TRPM calcium channels in liver tissue is important to explain the pathogenesis of ischemia-reperfusion and membrane depolarization. Thus, we propose to investigate the effects of TRPM2/8 and TRPM6/7 expressions after treatment of sodium channel blocker (lidocaine) in a rat model with hepatic ischemia-reperfusion.

Material and Methods

Experimental Design

The study was carried out on 32 male Wistar rats (average weight 225±25g) housed in an environmentally controlled room (240C to 260C temperature) with a 12:12 hour light: dark cycle, and kept on commercial rat chow and tap water ad libitum. The Committee on the Ethics of Animal Experiments of the Mustafa Kemal University has approved the study protocol.

The animals were randomly divided into 4 groups including sham, lidocaine, I/R group and I/R-lidocaine (sodium channel blocker) group.

Sham Group: Rats were pretreated with saline solution and surgical procedures, except for induction of liver ischemia, but including liver resection (n=8).

Sodium channel blocker (Lidocaine) group: The rats were treated with lidocaine as pretreatment (5 mg/kg) [16] and none of the rats was applied ischemia-reperfusion (n=8).

Ischemia-reperfusion group (I/R): Rats were not treated by any substance. For Ischemia atraumatic vascular vein and right hepatic artery were clamping for 60 minutes after the laparotomy. After reperfusion, all group members underwent relaparotomy and the livers were isolated (n=8).

Ischemia-reperfusion/sodium channel blocker (I/R-lidocaine) group: Lidocaine (5 mg/kg) was given orally to the rats 30 minutes before anesthesia as pretreatment. Xylazine/ ketamine (12/80mg/kg) was administered as anesthetic protocol. Ischemia was performed with a clamp on the portal vein and left lateral branches of the hepatic artery for 60 minutes after laparotomy. Relaparotomy was performed in all group members and livers were isolated for 60 minutes after reperfusion (n=8) [6].

Surgical Procedures

The rats were anesthetized by using xylazine/ ketamine (12/80mg/kg) combination and placed in a supine position on a temperature-controlled heating table, maintaining the body temperature in the range of 36.5-37.50C. The rats were allowed breathing spontaneously during surgery. For the preparation of the liver, abdominal skin was sterilized with ethyl alcohol (70%) and shaved. Then, using subcostal incisions and midline laparotomy, the liver was carefully mobilized from all ligamentous attachments. The left lateral branches and portal of the hepatic artery were clamped for 60 minutes in an atraumatic vascular clamp. The abdominal incision lines were closed to the margin and wetted with isotonic saline to accompany those who had removed excess body fluids. The median and left hepatic lobes were taken and then the abdomen was favorably watered with isotonic saline after extraction of the clamp. During I/R periods, the abdomen was covered with a plastic wrap to minimize fluid loss via evaporation. At the end of the first hour of reperfusion, the abdomen was closed by continuous stitches using Vicryl (Ethicon Endo-Surgery, Inc. USA) 4/0 sutures and the animals were returned to their cages. After 60 minutes of reperfusion, animals were anesthetized with an intraperitoneal injection of xylazine/ ketamine (12/80mg/kg) and were sacrificed, then histological samples were taken for RT-PCR and histopathological examination [6, 9].

Gene expression analysis (qRT-PCR)

Total RNA was isolated by using kits (RNAeasy Kit, Qiagen, Germany). cDNA was procured using the reverse transcription process.
assay kit (cDNA RT. Kit, USA). Shortly, 10X RT random primers (2 µl), 10X buffer reverse transcriptase (2 µl), dNTP mix (0.8 µl) (Table 1), AMV RT. (1 µl), RNase and mRNA free water (4.2 µl) were mixed to procure cDNA. The reaction admixture was incubated at 250°C for 10 minutes and 370°C for 120 minutes for reverse transcription and heated at 850°C for 5 minutes to inactive AMV reverse transcriptase. cDNA procured was stored at -20°C until tested. Then, cDNA was denatured at 950°C for 10 minutes, at 950°C for 15 seconds annealed at 600°C for 1 minute (TRPM2/8, TRPM6/7), and extended at 950°C for 3 minutes and at 720°C for 30 seconds. The reaction admixture was exposed to 40 cycles of PCR after an initial 15 seconds denaturation step at 950°C. Expressions of TRPM and p-actin mRNA as the cleaning gene were analyzed by quantitative reverse transcriptase PCR in Rotor-Gene Q (QIAGEN Rotor-Gene Q, Germany). Primary sequence information of the gene regions used in the study is given in Table 1.

Table 1. Primary sequence information of the gene regions.

| Gene      | Primary Sequences Used          | Tm   | Length (bp) |
|-----------|--------------------------------|------|-------------|
| TRPM2     | Left 5'-ATT TTG CTC ATC GCC ATG TT-3' | 55.2 | 20          |
| TRPM2     | Right 5'-CTC GCC CAT GTA CCA GAG ATT TCT GAG GT-3' | 59.4 | 20          |
| TRPM6     | Right 5'-CGT GCC CTG TGT TGC CTG CGT G-3' | 59.4 | 20          |
| TRPM6     | Left 5'-GGA ACA ACT GCC CCT TCC GCG-3' | 59.4 | 20          |
| TRPM7     | Right 5'-ATC CGG GTC TTC TGG CAT CAT TCT-3' | 59.4 | 20          |
| TRPM7     | Left 5'-AGA CCC TTC TGC ATG GCC GC-3' | 57.3 | 20          |
| β-Actin   | Left 5'-CCC GCG ACC ACG ACC TTC T-3' | 58.8 | 19          |
| β-Actin   | Right 5'-GCT CAT CCA TGG TGC ACT-3' | 56.0 | 18          |

Histopathological examination

After euthanasia, rats were necropsied and liver tissues were taken into 10% buffered formalin solution. The samples were then subjected to routine follow-up procedures and embedded in paraffin blocks. The 5 µm sections from the blocks were stained with Hematoxylin-Eosin for necrotic and degenerative changes. Sections examined under light microscopy were evaluated as none (-), mild (+), moderate (+ +) and severe (+ + +).

Data Analysis and Statistical analysis

The data were sorted out by utilizing Rotor-Gene Q Software, and the positive number chambers were corrected to estimate the actual number of copies. Described numbers were used to determine the number of copies in the original sample. As the housekeeping gene for normalization of the expressions, Beta Actin was used (Normalization= CT of Beta Actin/ CT of Gene). The expression levels in lidocaine group, I/R group, and I/R-lidocaine group were compared. The expression levels of TRPM and p-actin mRNA as the cleaning gene were analyzed by quantitative reverse transcriptase PCR in Rotor-Gene Q (QIAGEN Rotor-Gene Q, Germany). Primary sequence information of the gene regions used in the study is given in Table 1.

Data were analyzed using GraphPad Prism 5 program. Data were expressed as mean values ± standard error of the mean (SEM). For normally distributed data, one-way analysis of variance (ANOVA) with the Bonferroni’s multiple comparison post-test was used to test for significant differences. Histopathological data were assessed with the Pearson Chi-Square test which was utilized to compare ratios. P-values < 0.05 were considered as statistically significant.

Results

TRPM2, TRPM6, TRPM7, and TRPM8 Expressions

The Expression levels of TRPM2, TRPM6, TRPM7, and TRPM8 were shown in Table 2 and Figure 1. The expression levels of TRPM2 were significantly higher in liver-I/R group compared to sham and lidocaine groups (p<0.05, p<0.00, respectively). TRPM6 genes were significantly higher in liver-I/R group compared to sham and lidocaine groups (p<0.05, p<0.05, respectively). However, the expression levels of TRPM6 gene were significantly higher in liver-I/R group compared to sham, lidocaine, and I/R-lidocaine groups (p<0.05, p<0.01, p<0.05, respectively). The normalized CT±SD of TRPM2 values were 0.996±0.00, 0.980±0.01, 1.074±0.00 and 1.019±0.01 for sham, lidocaine, liver-I/R and I/R-lidocaine groups, respectively (Figure 1). The highest expression level of TRPM6 gene was in the liver-I/R group (Normalized CT ± SD: 1.218±0.01) and the values of normalized CT ± SD were 1.148±0.01, 1.127±0.01, and 1.141±0.01 for sham, lidocaine, and I/R-lidocaine groups, respectively (Figure 1). TRPM7 expression was modest in the lidocaine group (Normalized CT±SD: 1.189±0.01) and the values of normalized CT±SD were 1.239±0.01, 1.344±0.01, and 1.237±0.01 for sham, liver-I/R, and I/R-lidocaine groups, respectively (Figure 1). For TRPM8 gene, the values of expressions were 1.156±0.01, 1.152±0.02, 1.249±0.02 and 1.197±0.01 in hepatocytes from the sham, lidocaine, liver-I/R and I/R-lidocaine groups, respectively (Figure 1) (Table 2).

Table 2. Statistical data of gene expression results(Mean±SD)

| Genes | Sham (n=8) | Lid. (n=8) | I/R (n=8) | Lid+I/R (n=8) |
|-------|------------|-----------|-----------|--------------|
| TRPM2 | 0.99±0.00  | 0.98±0.00 | 1.07±0.00 | 1.01±0.00    |
| TRPM6 | 1.14±0.00  | 1.12±0.00 | 1.21±0.00 | 1.14±0.00    |
| TRPM7 | 1.23±0.00  | 1.18±0.00 | 1.34±0.00 | 1.23±0.00    |
| TRPM8 | 1.15±0.00  | 1.15±0.00 | 1.24±0.00 | 1.19±0.00    |

I/R: Ischemia-reperfusion; Lid: Lidocaine; a: compared with Sham group; b: compared with I/R group. *: p<0.05; **: p<0.01; ***: p<0.001

Histopathological examination

After euthanasia, rats were necropsied and liver tissues were taken into 10% buffered formalin solution. The samples were then subjected to routine follow-up procedures and embedded in paraffin blocks. The 5 µm sections from the blocks were stained with Hematoxylin-Eosin for necrotic and degenerative changes. Sections examined under light microscopy were evaluated as none (-), mild (+), moderate (+ +) and severe (+ + +).

Data Analysis and Statistical analysis

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Lidocaine and Sham groups had normal liver histology (Figure 2A). A statistically significant difference was found between ischemia-reperfusion and ischemia-reperfusion-lidocaine groups (p<0.05). Severe levels of necrotic/degenerative changes and intense hemorrhagic areas were observed in the livers of rats in the ischemia-reperfusion group (Figure 2C). Necrotic/ degenerative changes in the liver of rats in the ischemia-reperfusion group with lidocaine were found to be less than in the ischemia-reperfusion group alone (Figure 2D) (Table 3).

Table 3. Statistical data of gene expression results(Mean±SD)

| Genes | Sham (n=8) | Lid. (n=8) | I/R (n=8) | Lid+I/R (n=8) |
|-------|------------|-----------|-----------|--------------|
| Cellularity | 1.47±0.2 | 1.10±0.0 | 2.75±0.2 | 1.60±0.1 |
| Congestion | 1.90±0.3 | 1.71±0.0 | 3.65±0.2 | 3.31±0.1 |
| Polymorph nuclear leukocytes | 1.18±0.1 | 1.14±0.1 | 1.77±0.3 | 1.10±0.3 |
| Apoptosis | 0.28±0.1 | 1.14±0.1 | 0.57±0.2 | 0.34±0.2 |

I/R: Ischemia-reperfusion; Lid: Lidocaine; a: compared with Sham group; b: compared with I/R group. *: p<0.05; **: p<0.01; ***: p<0.001
The effect of lidocaine on TRPM 2, 6, 7 and 8 channels

Figure 1. The expression levels of TRPM2, TRPM6, TRPM7 and TRPM8 genes

Figure 2. A: Sham group. Normal histological appearance. B: Lidocaine group. Normal histological appearance. C: Ischemia-Reperfusion group. Severe levels of necrotic/ degenerative changes in hepatocytes (*) and hemorrhagic areas (arrowheads). D: Ischemia-Reperfusion + Lidocaine group. Moderate necrotic / degenerative changes in hepatocytes (*).
In conclusion, in this study, we evaluated expression changes of TRPM2/8 and TRPM6/7 genes in hepatocytes of rats which were exposed to I/R after administration of lidocaine. The increase of calcium in the ECM affects sodium and potassium conductance versus voltage curves. The added high calcium leads to an increase in the time to peak of sodium level at medium [15]. However, all calcium channels including TRPs, ryanodine, and calcium pumps should be investigated. We think that administration of lidocaine may protect liver from ischemia-reperfusion injury by reducing increased TRPM2/8 and TRPM6/7 genes expression in Liver I/R injury and liver transplantation.

Scientific Responsibility Statement
The authors declare that they are responsible for the article's scientific content including study design, data collection, analysis and interpretation, writing, some of the main line, or all of the preparation and scientific review of the contents and approval of the final version of the article.

Animal and human rights statement
All procedures performed in this study were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. No animal or human studies were carried out by the authors for this article.

Conflict of interest
None of the authors received any type of financial support that could be considered potential conflict of interest regarding the manuscript or its submission.

References
1. Du JD, Zheng X, Chen YL, Huang ZQ, Cai SW, Jiao HB, et al. Elevated Transient Receptor Potential Melastatin 8 (TRPM8) Expression Is Correlated with Poor Prognosis in Pancreatic Cancer. Med Sci Monit. 2018;24:3720-25.
2. Nilius B, Owsianik G. The transient receptor potential family of ion channels. Genome Biol. 2011;12(3):218.
3. Cuajungco MP, Grimm C, Oshima K, D’Hoedt D, Nilius B, Mensenkamp AR, et al. PACSINs bind to the TRPV4 cation channel. PACSIN 3 modulates the subcellular localization of TRPV4. J Biol Chem. 2006;281(27):28753-62.
4. Pan Z, Yang H, Reinach PS. Transient receptor potential (TRP) gene superfamily of cation channels. Hum Genomics. 2011;5(2):108-16.
5. Wang H, Song T, Wang W, Zhang Z. TRPM2 participates the transformation of acute pain to chronic pain during injury-induced neuropathic pain. Synapse. 2019;73(10):e22117. DOI: 10.1002/syn.22117.
6. Dohukovic R, Gagbekon B, Yumukas O, Bazeyik I, Goke H, Demir T. Expressions of TRPM6 and TRPM7 and histopathological evaluation of tissues in ischemia reperfusion performed rats. Ren Fail. 2014;36(6):932-6.
7. Gwanyanya A, Amuzescu B, Zakharov SI, Macianskienė R, Sipido KR, Bolotina VM, et al. Magnesium-inhibited, TRPM6/7-like channel in cardiac myocytes: permeation of divalent cations and pH-mediated regulation. J Physiol. 2004;559(Pt 3):761-76.
8. Okamoto Y, Okubo T, Ikebe T, Yamazaki J. Blockade of TRPM8 activity reduces the invasion potential of oral squamous carcinoma cell lines. Int J Oncol. 2012;40(5):1431-40.
9. Bilecik T, Karateke F, Elkan H, Goke H. The effects of TRPM2, TRPM6, TRPM7 and TRPM8 gene expression in hepatic ischemia reperfusion injury. Eur Rev Med Pharmacol Sci. 2019;23(7):3088-95.
10. Chang WJ, Chehab M, Kink S, Toledo-Pereyra LH. Intracellular calcium signaling pathways during liver ischemia and reperfusion. J Invest Surg. 2010;23(4):228-36.
11. Wu HH, Huang CC, Chang CP, Lin MT, Niu KC, Tian YF. Heat Shock Protein 70 (Hsp70) Reduces Hepatic Inflammatory and Oxidative Damage in a Rat Model of Liver Ischemia/Reperfusion Injury with Hyperbaric Oxygen Preconditioning. Med Sci Monit. 2018;24:8096-104.
12. Zhou H, Yu Y, Zhang J, Zhang Y, Luan Q, Wang G. Protective Effects the Akt Activator SC79 in Hepatic Ischemia-Reperfusion Injury. Med Sci Monit. 2018;24:4386-94.
13. Zhu J, Liu J, Shen G, Zhong T, Yu X. Comparison of Efficacy Outcomes of Lidocaine Spray. Topical Lidocaine Injection, and Lidocaine General Anesthesia in Nasal Bone Fractures Surgeries: A Randomized, Controlled Trial. Med Sci Monit. 2018;24:4386-94.
14. Cassuto J, Sinclair R, Bendorovic M. Anti-inflammatory properties of local anesthetics and their present and potential clinical implications. Acta Anaesthesiol Scand. 2006;50(3):265-82.
15. Blautstein MP, Goldman DE. Competitive action of calcium and procaine on lobster axon. A study of the mechanism of action of certain local anesthetics. J Gen Physiol. 1966;49(5):1043-63.
16. Lu HR, Yang P, Remeyzen S, Saels A, Dai DZ, De Clerck F. Ischemia/reperfusion-
induced arrhythmias in anaesthetized rats: a role of Na+ and Ca2+ influx. Eur J Pharmacol. 1999;365(2-3):233-9.
17. Dokuyucu R, Demir T, Yumrutas O, Erbagci AB, Orkmez M, Bahar AY, et al. The role of hepcidin and its related genes (BMP6, GDF-15, and HJV) in rats exposed to ischemia and reperfusion. Turk J Med Sci. 2014;44(4):576-81.
18. Bhasale G, Sharpe JA, Sundier SY, Duchen MR. Calcium signaling as a mediator of cell energy demand and a trigger to cell death. Ann N Y Acad Sci. 2015;1350:107-16.
19. Lei B, Cottrell JE, Kass IS. Neuroprotective effect of low-dose lidocaine in a rat model of transient focal cerebral ischemia. Anesthesiology. 2001;95(2):445-51.
20. Ebel D, Lifert P, Frassdorf J, Preckel B, Mullenheim J, Thamer V, et al. Lidocaine reduces ischemic but not reperfusion injury in isolated rat heart. Br J Anaesth. 2001;86(6):846-52.
21. Faouzi M, Penner R. Trpm2. Handb Exp Pharmacol. 2014;222:403-26.
22. Dusmez D, Cengiz B, Yumrutas O, Demir T, Oztuzcu S, Demiryurek S, et al. Effect of verapamil and lidocaine on TRPM and NaV1.9 gene expressions in renal ischemia-reperfusion. Transplant Proc. 2014;46(1):33-9.
23. Fontfria E, Murdock PR, Casdin FS, Benham CD, Keasel RE, McNulty S. Tissue distribution profiles of the human TRPM cation channel family. Journal of receptor and signal transduction research. 2006;26(3):159-78.
24. Zhang Y, Zhou L, Zhang X, Bai J, Shi M, Zhao G. Ginsenoside-Rd attenuates TRPM7 and ASIC1a but promotes ASIC2a expression in rats after focal cerebral ischemia. Neuronal Sci. 2012;33(5):1125-31.
25. Sun HS, Jackson MF, Martin LJ, Jansen K, Teves L, Cui H, et al. Suppression of hippocampal TRPM7 protein prevents delayed neuronal death in brain ischemia. Nat Neurosci. 2009;12(10):1300-7.
26. Madrid R, Donovan-Rodriguez T, Meseguer V, Acosta MC, Belmonte C, Viana F. Contribution of TRPM8 channels to cold transduction in primary sensory neurons and peripheral nerve terminals. J Neurosci. 2006;26(48):12512-25.

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