Extensive post-transcriptional regulation of microRNAs and its implications for cancer

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MicroRNAs (miRNAs) are short, noncoding RNAs that post-transcriptionally regulate gene expression. While hundreds of mammalian miRNA genes have been identified, little is known about the pathways that regulate the production of active miRNA species. Here we show that a large fraction of miRNA genes are regulated post-transcriptionally. During early mouse development, many miRNA primary transcripts, including the Let-7 family, are present at high levels but are not processed by the enzyme Drosha. An analysis of gene expression in primary tumors indicates that the widespread down-regulation of miRNAs observed in cancer is due to a failure at the Drosha processing step. These data uncover a novel regulatory step in miRNA function and provide a mechanism for miRNA down-regulation in cancer.

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The founding microRNA (miRNA), lin-4, was discovered more than 20 years ago in the nematode Caenorhabditis elegans [Lee et al. 1993]. Elegant studies demonstrated that the gene product of lin-4 was a noncoding RNA that post-transcriptionally regulated several critical genes during development [Wightman et al. 1993]. More recently, a large number of miRNA genes have been identified in plant and animal genomes. To date, >300 miRNAs have been validated in the human genome, and computational predictions suggest that many hundreds more exist [Aravin and Tuschl 2005]. While it is clear that these small, noncoding RNAs have essential functions in mammalian biology, few miRNA genes have been functionally linked to specific cellular pathways. For example, miR-181 modulates hematopoietic differentiation, and miR-1 and miR-133 participate in skeletal and cardiac muscle development. The polycistronic cluster miR-17-92 promotes lymphomagenesis, while Let-7 suppresses proliferation by targeting the oncoprotein Ras (for review, see Wienholds and Plasterk 2005).

For most miRNA genes, however, biological information is limited to expression analyses. In mouse and zebrafish, few miRNAs are expressed in early embryos [Thomson et al. 2004; Wienholds et al. 2005]. During mid- to late embryonic development, large numbers of miRNAs are induced in temporal and spatial patterns [Kloosterman et al. 2006]. This culminates in adult tissues, where a large fraction of the known miRNA genes are expressed. Interestingly, most of these same miRNA genes are down-regulated in cancer, perhaps reflecting a loss of cellular differentiation [Takamizawa et al. 2004; Lu et al. 2005]. The regulatory mechanism behind these expression changes is largely unknown, although the most obvious candidate is transcriptional control. It is difficult, however, to reconcile this model with the widespread, parallel reduction in miRNA expression observed in most types of human cancer.

A second possibility is that miRNA maturation is a regulated event. The biogenesis of miRNAs begins with a primary transcript, termed the pri-miRNA, which is generated by RNA polymerase II [for review, see Kim 2005]. The active species is contained in a stem–loop structure that is liberated by the nuclear ribonuclease III (RNase III) Drosha. This stem–loop, termed the pre-miRNA or precursor, is exported from the nucleus in a Ran/GTP/Exportin 5-mediated event. In the cytoplasm, the precursor is further processed by the RNase III enzyme Dicer to generate the mature miRNA species. In a concerted reaction, this siRNA-like product is loaded into the RNA induced silencing complex (RISC), where it mediates target miRNA translational suppression [Du and Zamore 2005; Gregory et al. 2005; Maniataki and Mourelatos 2005].

In principle, any step during this maturation process could be regulated. The core enzymes are widely expressed, however, and no post-translational regulation of Drosha or Dicer has been reported. Discrepancies between the levels of primary transcript, precursor, and mature miRNA species have been reported; however, no clear model is apparent [Suh et al. 2004; Eis et al. 2005]. Here we report that a large fraction of miRNAs is regulated during the Drosha processing step, and this regulation has a major impact on miRNA expression during embryonic development and in cancer.

Results and Discussion

A striking event during mouse development is the massive induction of Let-7 family miRNAs at ~10.5 d of gestation. This is exemplified by Let-7g. This miRNA is housed in the second intron of the WD-40 repeat gene AK037620 [Fig. 1A]. Processing of this primary transcript by Drosha yields a 79-nt stem–loop precursor. Further processing by Dicer leads to the 21-nt mature species. We analyzed the expression of all three molecular forms by Northern blotting [Fig. 1B]. As previously reported, mature Let-7g is undetectable in embryonic stem (ES) cells and P19 embryonal teratocarcinoma cells [Thomson et al. 2004]. Expression is detectable at 10.5 d of gestation and is high at 14.5 d. The precursor is detectable slightly earlier than the mature miRNA, suggesting a delay at the Dicer processing step. Similar results have
Regulation of microRNA processing

Figure 1. Expression of let-7g during mouse development. (A) The genomic organization of Let-7g is shown. RT-PCR primer sites are indicated by arrows. Primers Ex1 and Ex2 were used for PCR of the primary transcript. Primers In1 and In2 were used for PCR of the unspliced primary transcript. (B) Northern blot analysis of the molecular species of Let-7g. 18S rRNA and U6 snRNA were used for loading controls for pri-miRNA and pre-miRNA/mature, respectively. (C) Nonquantitative RT-PCR of the unspliced primary transcript. Control reactions without the reverse transcription step are shown. (D) Nonquantitative RT-PCR of the unspliced primary transcript and mature species. The U6-snRNA reference is also shown. The right lanes are control reactions without reverse transcription step. (E) Real-time RT-PCR analysis of the let-7g primary transcript and mature species. The U6-snRNA reference is also shown. The right lanes are control reactions without reverse transcription step. Expression is shown as relative values on a logarithmic scale.

been observed in mouse and Drosophila development (Hutvagner et al. 2001; Schulman et al. 2005). Surprisingly, the primary transcript is highly expressed throughout development, with little difference in expression between ES cells and 14.5-d embryos. This discrepancy is not resolved by the amount of unspliced pri-miRNA, since this is also essentially constant (Fig. 1C). We next quantitated the levels of primary and mature species by real-time RT-PCR. We adapted a published procedure to measure the amount of both species from a single RT reaction (see Supplemental Figs. 1, 2 for validation, Fig. 1D illustrates a nonquantitative example; Shi and Chiang 2005). U6 snRNA was used for normalization for mature and primary species. This RNA exhibited more consistent expression during embryonic development than more typical reference RNAs such as β-2 microglobulin, glyceraldehyde-3-phosphate dehydrogenase, and β-glucuronidase. Using this procedure, we determined that the amount of mature Let-7g increases 4200-fold between ES cells and 14.5-d embryos. As observed by Northern blot analysis, expression levels of the primary transcript do not match the mature miRNA but are relatively constant (Fig. 1E, note log scale).

There are several possible explanations for the different expression levels of the pri-miRNA and the mature miRNA. There could be rapid turnover of the mature species in ES cells compared to late embryos. This is unlikely, since mature miRNAs from other gene families are present in ES cells (e.g., mir-290–295), and published data suggest that all miRNAs are loaded into RISC.

complexes nondiscriminately [Liu et al. 2004]. A second possibility is that the precursor for Let-7g is retained in the nucleus or otherwise made inaccessible to Dicer. In that circumstance, we would expect to see an accumulation of precursor. Our data disagree with this, since the precursor is essentially undetectable in ES cells. A third possibility, most congruous with our data, is that processing of Let-7g is blocked at the Drosha step. Release of the block would enable production of mature Let-7g. It is worth emphasizing that this is not simply a delay in processing during development, but a complete block. ES cells are grown continuously in culture, express high levels of Let-7g primary transcript, and never process a significant quantity to the precursor or mature species.

We next investigated processing of other miRNA genes that are expressed during development (Fig. 2). Let-7 family miRNAs are located at eight genomic loci (Griffiths-Jones et al. 2006). Of these we were able to detect primary transcripts for four loci. Notable is the Let-7i-2/miR-98 cluster, which is located within an intron of the widely expressed ubiquitin ligase gene HUWE-1. All four primary transcripts were expressed at similar levels throughout development. Similar to Let-7g, all mature miRNAs within these genes were elevated several thousandfold between ES cells and 14.5 d of gestation. The correlation between primary transcript expression and mature miRNA expression was close to zero, with the exception of Let-7i (see Table 1). For this miRNA, the primary transcript is induced during development, although there is still a large difference in processing efficiency (300-fold) (Table 1). Two of the four Let-7 clusters we analyzed were intronic, and two were exonic. Thus there is no correlation between regulated processing by Drosha and intron/exon location.

This Drosha regulatory step has not been observed in other organisms. C. elegans let-7, for example, is induced at the fourth larval stage and continues into the adult. Expression is controlled by defined enhancer elements in the promoter of the primary transcript [Johnson et al. 2003]. Expression of the primary transcript is closely correlated with mature let-7 [Bracht et al. 2004].

The developmental function of Let-7 family members in mammals is not known. There have been reports of decreased Let-7 expression in primary tumors (Takamizawa et al. 2004; Lu et al. 2005). This may have functional consequences. The oncogene Ras is a validated target of Let-7 [Johnson et al. 2005]. Thus, decreased Let-7 expression would lead to increased Ras expression, which would promote survival of tumor cells. Our data raise the possibility that Let-7 down-regulation in tumors is post-transcriptional (see below).

Several other families of miRNAs are up-regulated during mouse development. We were able to detect pri-
mary transcripts for nine of these miRNAs. Most cases exhibited regulated Drosha processing (Fig. 2; Table 1). Exceptions were miR-21 and the miR-143/miR-145 cluster. Both primary transcripts matched expression of the mature species (correlation = 1.0 and 0.99, respectively). Another interesting example is the miR-15/miR-16 cis-tron, which is located at two genomic loci. Each primary transcript displays distinct expression kinetics. The Chromosome 14 copy is located within an uncharacterized gene that is induced during development. The Chromosome 3 copy, in contrast, is located within the condensin subunit SMC4L1, which is constitutively expressed. Thus, each pri-miRNA contributes to the mature species in a complex manner, with a post-transcriptionally regulated component (Chromosome 3) and a transcriptionally regulated component (Chromosome 14).

It should be noted that coordinate expression of the primary transcript and the mature miRNA does not prove that Drosha processing is not regulated. Such miRNAs [Table 1, bottom panel] may be regulated at transcription and Drosha processing. It remains a possibility that all miRNAs are regulated at the Drosha step, except for miRNAs that have demonstrated expression in the early mouse embryo. Interestingly, this has implications for experiments that use ectopic expression of miRNAs in ES cells. We predict that this would fail to produce the mature species, unless cis-regulatory elements were removed from the expression cassette.

Our data suggest that the differentiation events that occur during embryonic development activate Drosha processing of specific miRNAs. To directly test this we used the teratocarcinoma cell line P19, which can be differentiated in culture into multiple cell types (Rudnicki et al. 1990). miRNA microarray expression analysis of untreated and differentiated P19 cells mirrors the expression changes observed during mouse development (data not shown). Embryonic miRNAs, such as the miR-290–295 cluster, are down-regulated during differentiation. In contrast, miRNAs that are up-regulated during mouse development, including Let-7 family members, are increased upon differentiation. As in embryonic development, the increases in Let-7 miRNAs are not coupled to transcription of the pri-miRNA (see Fig. 3).

| miRNA gene | pri-miRNA/mature correlation | Δ processing efficiency |
|------------|------------------------------|------------------------|
| let-7g     | +0.29                        | 5400                   |
| let-7f-2/miR-98 | -0.12                   | 2700                   |
| let-7i     | +1.00                        | 300                    |
| let-7d/let-7a-1/let-7f-1 | -0.21              | 1400                   |
| miR-140    | -0.20                        | 290                    |
| miR-23b/miR-24-2/miR-27b | -0.29          | 110                    |
| miR-30/b/miR-30d | +0.89                | 2.5                    |
| miR-103-1  | -0.38                        | 18                     |
| miR-103-2  | -0.87                        | 120                    |
| miR-15b/miR-16-2 | -0.68              | 19                     |
| miR-15a/miR-16-1 | +0.98               | 0.23                   |
| miR-21     | +0.99                        | 1.4                    |
| miR-143/miR-145 | +0.99           | 9.3                    |

Pearson correlations were calculated for each mature miRNA/pri-miRNA pair and are shown in the middle column. The change in processing efficiency is defined as the ratio of mature to pri-miRNA in a 14.5-d embryo divided by the ratio of mature to pri-miRNA in ES cells, and is shown in the right column. The table is divided into miRNAs regulated by Drosha [top] and miRNAs potentially unregulated by Drosha [bottom].

Figure 2. miRNA expression during mouse development. The expression levels of primary transcript and mature species from 12 representative miRNAs are shown. Red bars represent mature miRNA, and blue bars represent primary transcript. All analyses are performed with the following RNA samples, from left to right: P19 teratocarcinoma cell line; embryonic stem cell line; 10.5-d gestation mouse embryo; 14.5-d gestation mouse embryo. Reactions were performed in triplicate and normalized to U6 cycle threshold values. Expression is shown as relative values on a logarithmic scale. The bold font indicates the mature miRNA gene that was used for RT-PCR analysis. In all cases, these are unique miRNA sequences in the mouse genome.

Figure 3. miRNA expression during P19 cell differentiation. P19 teratocarcinoma cells were differentiated by forming embryoid bodies for 4 d in the presence of all-trans retinoic acid [RA], followed by plating without RA. Arrows indicate RA treatment and plating time. Primary transcript and mature miRNA species were quantitated by real-time RT-PCR. Reactions were performed in triplicate and normalized to U6 cycle threshold values. Expression is shown as relative values on a linear scale. Red bars indicate mature miRNA, and blue bars indicate pri-miRNA.
One experimental observation of cancer cells is the widespread alterations in miRNA expression. While some miRNAs are elevated, for example, the oncogenic cluster miR-17–92, most miRNAs have significantly reduced expression [He et al. 2005; Lu et al. 2005]. Our data raise the possibility that these reductions are a consequence of the Drosha processing block. To address this, we analyzed existing expression data, comparing mature miRNA levels with the primary transcript. The most complete data set, from the Broad Institute, has Affymetrix mRNA expression data and Luminex miRNA data from a wide range of primary tumors and normal tissues [Ramaswamy et al. 2001; Lu et al. 2005]. We combined the data set in the following manner: 68 tumors and 21 normal tissue samples had data for miRNA and mRNA expression. Of 217 miRNAs, 22 could be mapped to a primary transcript that was present on the Affymetrix microarray. When this restricted miRNA data set was hierarchically clustered, the normal samples were located in a single node with some tumor samples, while most tumors clustered in a separate node [Fig. 4A]. The tumor samples had overall reduction in miRNA expression levels. This was qualitatively similar to the entire miRNA expression map, as previously published [Lu et al. 2005].

We then created an expression map of primary transcript expression levels [Fig. 4A]. The samples and genes are arranged in the same order as for the miRNA expression map. The high and low clusters of expression are completely lost in the primary transcript expression map. We performed correlation analysis between each mature/pri-miRNA expression pair, for each sample. The data are represented in the histogram in Figure 4B. There is no correlation between pri-miRNA and mature expression in the tumor samples, while the normal tissue samples had positive correlation (KS test, p < 0.01). This demonstrates that the miRNA alterations that occur in tumors, for this limited set of genes, are not due to misregulated transcription.

Our data suggest a multistep model for the control of miRNA expression. Transcription of the pri-miRNA can be regulated, as has been demonstrated for tissue-specific miRNAs [Fazi et al. 2005; O’Donnell et al. 2005; Zhao et al. 2005; Chen et al. 2006]. Processing at the Dicer step can be delayed or inhibited [Hutvagner et al. 2001; Schuman et al. 2005; Obernosterer et al. 2006]. We demonstrate that further suppression of miRNA production is achieved at the Drosha step. This may be necessary for early development, where inappropriate expression of even small amounts of Let-7 may promote differentiation, with disastrous consequences. This model also allows miRNAs to be located in essential housekeeping genes that are ubiquitously expressed.

The biochemical nature of Drosha regulation is unknown. We measured the expression levels of Drosha and its binding partner DGCR8, during the P19 differentiation time course [Supplemental Fig. 3]. No induction occurred concurrent with the production of mature miRNAs. It is possible these proteins are regulated by post-translational modification. Alternatively, additional regulatory binding proteins may be required for specific miRNA processing. Recently, adenosine deaminase (ADAR) editing of specific pri-miRNAs has been reported [Yang et al. 2006]. This editing event leads to decreased processing of the miRNA by Drosha and increased turnover by the Tudor-SN nuclease. We tested whether this mechanism is responsible for the Drosha block during embryogenesis. We failed to find evidence of A-I editing of Let-7 pri-miRNAs in ES cells [data not shown]. Our data do not, however, discount the possibil-

Figure 4. miRNA expression in primary tumors. (A) Normalized miRNA expression data from Lu et al. [2005] were clustered hierarchically in both dimensions and are displayed as an expression map. Yellow indicates increased expression, and blue indicates decreased expression, relative to the median. Primary transcript expression data from Ramaswamy et al. [2001] were mapped in the same order as the mature miRNA expression map. Mature miRNA/primary transcript gene pairs are indicated on the right. Tumor samples are indicated in red, and normal tissues are indicated in black. (B) Pearson correlation values for each mature/pri-miRNA value pair were calculated. Values for normal samples (dashed line) and tumor samples (solid line) are plotted as a histogram.
ity of other structural changes in the pri-miRNA that block processing. An alternative mechanism is the sequetration of regulated pri-miRNAs away from the processing apparatus. The exact nature of the regulatory mechanism awaits further investigation.

Materials and methods

Cell lines

The feederless ES cell line E14Tg2A4 was grown on gelatin-coated plates in Glasgow MEM [Sigma] supplemented with 2 mM glutamine, 1 mM pyruvate, 1x nonessential amino acids, 15% fetal bovine serum [HyClone], 0.1 mM β-mercaptoethanol, and 10⁵ units/mL leukocyte inhibitory factor [Chemicon]. The P19 teratocarcina cell line was grown in DMEM [Sigma] supplemented with 7.5% calf serum (Hyclone) and 2.5% fetal bovine serum. P19 cells were differentiated as follows: Cells were seeded on nonadhesive plates in α-MEM [Sigma] supplemented with 5% fetal bovine serum and 500 nM all-trans retinoic acid (Sigma). After 4 d, the aggregated cells were plated on standard tissue culture dishes in DMEM supplemented with 7.5% calf serum and 2.5% fetal bovine serum, and grown for the indicated number of days.

RNA isolation

Dissected whole embryos from pregnant CD1 mice were flash-frozen in liquid nitrogen and pulverized into powder. Total RNA was extracted from pooled embryos of two to eight embryos or from cell lines with Trizol Reagent [Invitrogen] according to the manufacturer's instructions.

Northern blot analysis of mRNA

Five micrograms of total RNA was resolved on a 1% agarose formaldehyde gel and transferred overnight to a positively charged nylon membrane by capillary transfer with 10× SSC. The membrane was washed once for 15 min at 42°C, followed by three washes in 0.1× SSC, 0.1% SDS. PhosphorImaging using the Opticon 2 (MJ Research) was used to detect the grey-level signal. The membrane was washed once for 15 min at 42°C in 2× SSC, 0.1% SDS and then twice for 15 min at 65°C in 0.2× SSC, 0.1% SDS. PhosphorImaging using the Storm system (Molecular Dynamics) revealed the Let-7g primary transcript in Church and Gilbert high-stringency buffer. PCR primers specific for the mouse Let-7g primary transcript [let-7g-probe-F, 5′-TTTAGTCCATTGGCCAGAC-3′; let-7g-probe-R, 5′-CATCACCACA CACCCCTGATCT-3′] were used to make the probe. The membrane was washed once for 15 min at 42°C in 2× SSC, 0.1% SDS and then twice for 15 min at 65°C in 0.2× SSC, 0.1% SDS. Hybridization was performed in Church and Gilbert high-stringency buffer at 65°C, followed by three washes in 0.1× SSC at 65°C.

Quantitative RT-PCR

Total RNA (20 µg) was DNase I [Promega] treated according to the manufacturer's instructions, phenol:chlorofrom extracted, ethanol precipitated, and dissolved in DEPC-treated H₂O. RNA [10 µg] was polyadenylated using Poly[A] polymerase [Ambion] according to the manufacturer's instructions, phenol:chlorofrom extracted, ethanol-precipitated, and dissolved in DEPC-treated H₂O. A modified cDNA was made as follows: 10 µg of polyadenylated RNA was reverse-transcribed using Superscript II reverse transcriptase [Invitrogen] with 2.5 µg of random hexamers and 500 ng of oligo(dT) primer adapter primer (5′-GGCGAGCACAG AATTAATACGACTCACTATAGGTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT
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