An Optimized *Ustilago* Chassis for Itaconic Acid Production at Theoretical Maximal Yield

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Abstract: *Ustilago maydis*, member of the Ustilaginaceae family, is a promising host for the production of several metabolites including itaconic acid. This dicarboxylate has great potential as a bio-based building block in the polymer industry, and is of special interest for pharmaceutical applications. Several itaconate overproducing *Ustilago* strains have been generated by metabolic and morphology engineering. This yielded stabilized unicellular morphology through *fuz7* deletion, reduction of by-product formation through deletion of genes responsible for itaconate oxidation and (glyco)lipid production, and the overexpression of the regulator of the itaconate cluster *ria1* and the mitochondrial tricarboxylate transporter encoded by *mttA* from *Aspergillus terreus*. In this study, itaconate production was further optimized by consolidating these different optimizations into one strain. The combined modifications resulted in itaconic acid production at theoretical maximal yield, which was achieved under biotechnologically relevant fed-batch fermentations with continuous feed.

Keywords: Itaconic acid; *U. maydis*; Metabolic engineering; Fungi; Yeast

1. Introduction

Itaconic acid and its derivatives are found in many application fields, such as the production of paper, paints, and fibers, or in waste water treatment [1] [2] [3] [4] providing a stable market for this bio-based chemical. In 2004, the organic acid was classified as one of the top 12 value-added platform chemicals derived from biomass [5]. There is also a strong interest in this molecule in the medical and pharmaceutical sectors, both as an antibacterial compound [6] and as an immunoregulator for the treatment of autoimmune diseases [7] and viral infections including SARS-COV2 [8]. However, its relatively high production cost compared to fossil counterparts like acrylic acid prevents an even more expanded usage range [9] [10]. Given the already high yield, titer, and rate of the industrially established itaconic acid production process with *Aspergillus terreus* [11], a qualitative breakthrough in other dimensions of the process window is needed. Non-conventional itaconate producing yeasts like *Ustilago*, *Candida*, or *Pseudozyma* [12] [13] [14] may offer such a breakthrough because of their unicellular morphology as well as their lower sensitivity to medium impurities. It also enables easier handling and scale-up using cheaper raw substrates or even waste streams [10] [15] [16]. However, compared to the filamentous ascomycete *A. terreus*, the product to substrate yield has to be improved to make *U. maydis* competitive, and this yield also needs to be achievable under industrially relevant conditions [17].

In nature, *U. maydis* is known for its pathogenicity towards maize (*Zea mays*) causing corn smut disease [18] [19]. To penetrate and invade maize tissue, *U. maydis* switches from a yeast-like, non-pathogenic to a filamentous, pathogenic cell form [20]. Although predominantly growing in its...
yeast form in fermentation processes, \textit{U. maydis} can switch to filamentous growth under stress conditions \cite{21} \cite{22} \cite{23}. Filamentous growth causes issues such as high viscosity, reduced oxygen supply, and cell adherence to reactor walls \cite{9}. This can be avoided through the deletion of \textit{fuz7}, the product of which plays an essential role in regulating pathogenicity and the switch to filamentous growth \cite{21}. This deletion stabilizes the yeast-like morphology of \textit{U. maydis} and \textit{U. cynodontis} without impacting the fitness of the cells under biotechnologically relevant stresses \cite{24} \cite{25}.

\textit{U. maydis} produces itaconate as one product from a potpourri of metabolites including organic acids such as malate, succinate, and (S)-2-hydroxyparacarboxylic acid, polyols such as erythritol and mannitol, and different lipidic products including glycolipids and triglycerides \cite{26} \cite{27} \cite{28} \cite{29} \cite{30}. It can also metabolize a range of renewable carbon sources, which besides sugar also include glycerol \cite{32}, galacturonic acid \cite{33}, cellulose \cite{34}, xylan \cite{35}, and pectin \cite{36}. Although these features make \textit{U. maydis} an attractive candidate for industrial applications \cite{37} \cite{9} \cite{38}, it also poses a drawback because often multiple products are produced simultaneously. This hinders handling and downstream processing, and it also reduces yield on substrate by diverting carbon flux away from the main product \cite{39}. If itaconic acid is to become a bulk chemical, yield is the most relevant production parameter because substrate cost is the decisive price-determining factor. For itaconate production from glucose, the reported maximal theoretical yield is 0.72 mol\textit{ITA} mol\textit{glu}^{-1}, which equals 1 mol\text{ITA} mol\text{glu}^{-1}. The yield achieved in practice is affected by different factors such as cell density, the metabolic pathway leading up to itaconate (i.e., anaplerosis), bottlenecks in the itaconate biosynthesis pathway itself, side product formation, redox cofactor balancing, and cell maintenance demand \cite{40} \cite{41}.

In previous studies several knockouts, promotor replacements and overexpression of genes were implemented to increase itaconate production, reduce by-product formation, and stabilize the unicellular morphology \cite{42} \cite{43} \cite{10} \cite{24} \cite{39}. Those metabolic engineering approaches resulted in several itaconate hyper-producing \textit{Ustilago} strains with individual modifications. In this study, these modifications are consolidated into one strain, based on the previously engineered \textit{U. maydis} ITA chassis \cite{39}. The resulting strain K14 produces itaconate from glucose at maximum theoretical yield. The catalytic vigor of the strain was demonstrated in fed-batch cultures.

2. Materials and Methods

3.1. Media and culture conditions

All strains used in this work are listed in Table 1. \textit{U. maydis} strains were grown in YEPS medium containing 10 g L^{-1} yeast extract, 10 g L^{-1} peptone, and 10 g L^{-1} sucrose. As screening medium for production experiments, \textit{U. maydis} was cultivated in modified Tabuchi medium (MTM) according to Geiser \textit{et al.} \cite{28}. Besides varying glucose and buffer (2-(N-morpholino)ethanesulfonic acid (MES; pH adjusted to 6.5 with NaOH) or CaCO\textsubscript{3}) concentrations, this medium contained 0.8 g L^{-1} NH\textsubscript{4}Cl, 0.2 g L^{-1} MgSO\textsubscript{4}·7H\textsubscript{2}O, 0.01 g L^{-1} FeSO\textsubscript{4}·7H\textsubscript{2}O, 0.5 g L^{-1} KH\textsubscript{2}PO\textsubscript{4}, 1 mL L^{-1} vitamin solution, and 1 mL L^{-1} trace element solution. The vitamin solution contained (per liter) 0.05 g D-biotin, 1 g D-calcium pantothenate, 1 g nicotinic acid, 25 g myo-inositol, 1 g thiamine hydrochloride, 1 g pyridoxol hydrochloride, and 0.2 g para-aminobenzoic acid. The trace element solution contained (per liter) 1.5 g EDTA, 0.45 g ZnSO\textsubscript{4}·7H\textsubscript{2}O, 0.10 g MnCl\textsubscript{2}·4H\textsubscript{2}O, 0.03 g CoCl\textsubscript{2}·6H\textsubscript{2}O, 0.03 g CuSO\textsubscript{4}·5H\textsubscript{2}O, 0.04 g Na\textsubscript{2}MoO\textsubscript{4}·2H\textsubscript{2}O, 0.45 g CaCl\textsubscript{2}·2H\textsubscript{2}O, 0.3 g FeSO\textsubscript{4}·7H\textsubscript{2}O, 0.10 g H\textsubscript{3}BO\textsubscript{3}, and 0.01 g KI. When using CaCO\textsubscript{3}, medium components were added relative to the total volume of solids plus liquid, leading to a higher aqueous concentration of soluble components. Shaking cultures of \textit{U. maydis} were performed in 24-well System Duetz\textsuperscript{®} plates with a filling volume of 1.5 mL (shaking diameter = 50 mm, n = 300 rpm, T = 30 °C and Φ = 80%) \cite{44} or in 500 mL shaking flasks with a filling volume of 50 mL (shaking diameter = 25 mm, n = 200 rpm, T = 30 °C and Φ = 80%). When using System Duetz\textsuperscript{®}, cultures were inoculated in parallel into multiple plates in order to ensure continuous oxygenation by taking a complete plate as sacrificial sample for each sample point.
### Table 1. *U. maydis* MB215 strains used in this study.

| Strain designation | Resistance | Reference |
|--------------------|------------|-----------|
| *U. maydis* MB215  |            | [45]      |
| *U. maydis* MB215 ∆*cyp3* ∆*Pao::Poff* |            |           |
| *U. maydis* MB215 ∆*umag*-05079 *Pao::mttA* | hyg§, cbx§ | [46]      |
| *U. maydis* MB215 ∆*cyp3* ∆*fuz7* ∆*Pao::Poff* *Pao::mttA* | hyg§, cbx§ | [10]      |
| *U. maydis* MB215 ∆*cyp3* ∆*MEL* ∆*UA* ∆*gat* ∆*Pao::Poff* (∆*tGA* chass) |            | [39]      |
| *U. maydis* MB215 ∆*cyp3* ∆*MEL* ∆*UA* ∆*gat* ∆*Pao::Poff* ∆*fuz7* |            | this study |
| *U. maydis* MB215 ∆*cyp3* ∆*MEL* ∆*UA* ∆*gat* ∆*Pao::Poff* ∆*fuz7* *Pao::mttA* | cbx§ | this study |
| *U. maydis* MB215 ∆*cyp3* ∆*MEL* ∆*UA* ∆*gat* ∆*Pao::Poff* ∆*fuz7* *Pao::mttA* *K3* | cbx§ | this study |
| *U. maydis* MB215 ∆*cyp3* ∆*MEL* ∆*UA* ∆*gat* ∆*Pao::Poff* ∆*fuz7* *Pao::mttA* *K9* | cbx§ | this study |
| *U. maydis* MB215 ∆*cyp3* ∆*MEL* ∆*UA* ∆*gat* ∆*Pao::Poff* ∆*fuz7* *Pao::mttA* *K10* | cbx§ | this study |
| *U. maydis* MB215 ∆*cyp3* ∆*MEL* ∆*UA* ∆*gat* ∆*Pao::Poff* ∆*fuz7* *Pao::mttA* *K14* (∆*K14* strain) | cbx§ | this study |

Pulsed fed-batch fermentations were performed in New Brunswick BioFlo® 115 bioreactors (Eppendorf, Germany) as described in [10]. Fed-batch fermentations with continuous feed were performed in a 2.0 L DASGIP® Bioblock bioreactor (Eppendorf, Germany) with a starting volume of 1.0 L. The medium contained 120 g L⁻¹ glucose, 0.8 g L⁻¹ NH₄Cl, 0.2 g L⁻¹ MgSO₄7H₂O, 0.01 g L⁻¹ FeSO₄7H₂O, 0.5 g L⁻¹ KH₂PO₄, 1 mL L⁻¹ vitamin solution, 1 mL L⁻¹ trace element solution, and 1 g L⁻¹ yeast extract. When the glucose concentration reached approximately 50 g L⁻¹, a constant feed of a 50% glucose solution was started. Feeding rates of 0.75 and 2.8 g h⁻¹ were estimated from the glucose consumption rates of previous pulsed fed-batch fermentations under similar conditions. During cultivation, the pH was controlled by automatic addition of 5 M NaOH and 1 M HCl. 0.5 mL Antifoam 204 (Sigma Life Science, USA) was added manually every 24 h to avoid foam formation. The DO was controlled at 30% by using a cascade mode including stirring 800-1200 rpm, air flow 1.2 L min⁻¹ and the addition of pure oxygen. The CO₂ formation was determined with the DASGIP GA4 module, employing IR sensors (BlueSens). The cultivations were performed at 30 °C. Bioreactors were inoculated to an OD₆₀₀ of 0.75 from a 48 h preculture in 50 mL MTM.

#### 3.1. Analytical methods

When using CaCO₃ as buffer, 1 mL culture broth was taken for OD₆₀₀ and HPLC analysis. The CaCO₃ was dissolved 1:1 with 4 M HCl prior to further measurements as described in Zambanini *et al.* [47]. Cell densities were measured by determining the absorption at 600 nm with an Ultrospec 10 Cell Density Meter (Amersham Biosciences, UK).

For high performance liquid chromatography (HPLC) analysis, all samples were filtered with Rotilab® (CA, 0.2 µm, Ø 15 mm) or Acrodisc® (GHP, 0.2 µm, Ø 13 mm) syringe filters and diluted 1:5 or 1:10 with 5 mM H₂SO₄ or deionized water. Products in the supernatant were analyzed using a DIONEX UltiMate 3000 HPLC System (Thermo Scientific, Germany) or a Agilent-1260 Infinity HPLC system (Agilent, Waldbronn, Germany) with an ISERA Metab-AAC column 300 x 7.8 mm column (ISERA, Germany). As solvent, 5 mM H₂SO₄ with a constant flow rate of 0.6 mL min⁻¹ and a temperature of 40 °C was used. When using the DIONEX UltiMate 3000 HPLC System, detection was carried out by a DIONEX UltiMate-3000 Variable Wavelength Detector set to 210 nm and a SHODEX RI-101 detector (Showa Denko Europe GmbH, Germany). When using the Agilent 1260
Infinity HPLC system, detection was done by a diode array detector (DAD) at 210 nm and a refractive index (RI) detector. Analytes were identified via retention time and UV/RI ratio compared to corresponding standards.

All values are the arithmetic mean of at least two biological replicates. For \( n = 2 \), error bars indicate the deviation from the mean and for \( n > 2 \) error bars indicate the standard error of the mean. Statistical significance was evaluated by t test (two-tailed distribution, heteroscedastic, \( p \leq 0.05 \)).

3.1. Plasmid cloning and strain engineering

Plasmids were constructed by Gibson assembly [48] using the NEBuilder® HiFi DNA Assembly Cloning Kit (New England Biolabs (NEB), Ipswich, MA, USA). Primers were ordered as DNA oligonucleotides from Eurofins Genomics (Ebersberg, Germany).  As polymerase, Q5® High-Fidelity DNA Polymerase (NEB) was used. Detailed information about utilized primers and plasmids are listed in Table 2 and supplemental Table S1. Competent E. coli DH5α were used for standard cloning and plasmid maintenance according to Sambrook et al. [49]. Plasmids were confirmed by PCR or sequencing. Generation of protoplasts and transformation of \( U. maydis \) were performed according to Brachmann et al. [50]. Genomic DNA of \( U. maydis \) was isolated according to Hoffman et al. [51]. For the deletion of \( fuz7 \), homologous recombination with 1000 bp flanking regions including FRT-site and a hygromycin resistance cassette were used [52]. Successful integration and deletion was verified by PCR and sequencing. For the overexpression of \( mttA \), the plasmid \( P_{opt}-\text{Cbx-AT_mttA} \) was used [46].

Table 2. Plasmids used in this study.

| plasmid                  | description                                                                 | reference                                                                 |
|-------------------------|----------------------------------------------------------------------------|----------------------------------------------------------------------------|
| pJET1.2/blunt           | Ori ColE1; AmpR                                                             | Thermo Scientific, Germany                                                 |
| pFLPexpC                | Pcrg1 promoter; synthetic FLP recombinase gene; cbxR; ARS; AmpR             | Prof. M. Feldbrügge, Heinrich-Heine University Düsseldorf, Germany         |
| pUMa1523                | FRTm1-HygR-FRTm1 cassette; GentR                                           | Dr. K. Schipper, Heinrich-Heine University Düsseldorf, Germany            |
| pJET1.2-\textit{fuz7} 5'-UTR flank - FRTm1-HygR-FRTm1-\textit{fuz7} 3'-UTR flank | pJET1.2 with 5'- and 3'-UTR flank of UMAG_01514 as deletion construct; HygR; FRT m1 recombination sites | this study                                                                |
| \( P_{opt}-\text{Cbx-AT_mttA} \) | constitutive \( P_{opt} \) promoter, dicodon-optimized version of \textit{A. terreus} ATEG_09970 (\textit{mttA}), cbxR, ampR | [46]                                                                      |

Quantitative PCR was applied to determine the copy number of \( mttA \) integrated into the \( U. maydis \) genome using Luna® Universal qPCR Master Mix (NEB, Germany). Primers were designed using “GenScript Real-Time PCR (TaqMan) Primer Design” tool (Gen Script, New Jersey, USA). Primer sequences are given in Table 3. As reference genes, UMAG_02592 and UMAG_03726 were amplified with the primer pairs JB-126/JB-127 and JB-128/JB-129. Primers JB-132/JB-133 specifically bound within the \( mttA \) sequence (supplemental Table S1). Amplification curves were taken by Bio-Rad CFX ConnectTM Real-Time PCR Detection system and data were analyzed by using Bio-Rad CFX ManagerTM 3.1 software (Bio-Rad Laboratories, Hercules, USA) using the \( \Delta Ct \) method according to Pfaffl [53].
3. Results and Discussion

3.1. Prevention of filamentous growth by fuz7 deletion and its influence on itaconate production

In previous work, itaconate production with Ustilago has been significantly improved. The characterization and upregulation of the itaconate gene cluster [42] [54] [55] as well as the engineering of the mitochondrial carrier for cis-aconitate [46] has laid the foundation for this improvement. Those achievements were combined with further modifications including the deletion of genes responsible for itaconate oxidation (cyp3) and (glyco)lipid production (MEL, UA, dgat), resulting in the U. maydis MB215 ITA chassis (∆cyp3 ∆MEL ∆UA ∆dgat Δpet::Petef) with reduced by-product formation [39]. Filamentous growth was observed for this strain, similar to other engineered U. maydis variants [10]. The morphology switch is likely induced by the additional stress imposed by the metabolic engineering itself, and the resulting high itaconate titers and associated low pH. This drawback can be overcome by the deletion of fuz7 (UMAG_01514) [25] [24]. The influence of this knockout in the U. maydis MB215 ITA chassis was assessed in System Duetz® cultivations in MTM containing 50 g L⁻¹ glucose and 100 mM MES (Figure 1). Under these conditions, the ∆fuz7 strain produced 1.3-fold more itaconate than the reference ITA chassis strain (Figure 1 A, Table 3). Full consumption of glucose by both strains resulted in an equivalently improved yield of the ∆fuz7 strain to 0.45 ± 0.01 g ita g glu⁻¹. The overall production rate was increased by 12% and the maximal rate even by 26% (Table 3).

These results clearly illustrate the benefit of preventing filamentous growth. In these deeply engineered strains of U. maydis, cells start to adhere to the walls of the culture plates during the production phase when low pH stress and increasing product concentrations occur. This results in inhomogeneous oxygen and nutrient supply and exclusion from the buffer effect in the medium for the affected cells. This cell accumulation on the walls was reflected in decreasing optical density values after 72 h (Figure 1 B) and a decreasing itaconate productivity (Figure 1 A). This is in contrast to the fuz7 mutant, which showed a constant itaconate production rate in the production phase until glucose was depleted.
Table 3. ITA production parameters of engineered strains of the *U. maydis* MR215 ITA chassis (Δ_cyp3 Δ_MEL Δ_AUA Δ_gsat Δ_PraC::P_adh). *z*-values indicate the standard error of the mean (*n* = 3). Symbols refer to Figures 1, 2, and 3.

| Conditions | Symbol | Strain modification | ITA titer<sub>max</sub> (g L<sup>-1</sup>) | q<sub>m</sub> (g L<sup>-1</sup> h<sup>-1</sup>) | q<sub>P_max</sub> (g L<sup>-1</sup> h<sup>-1</sup>) | y<sub>wt</sub> (%) |
|------------|--------|---------------------|---------------------------------|-------------------------------|--------------------------------|-----------------|
| 100 mM MES, 50 g L<sup>-1</sup> glucose | ▲      | control             | 19.4 ± 0.3                      | 0.25 ± 0.01                   | 0.43 ± 0.00       | 0.41 ± 0.01     |
|           | ▶      | Δ_fuz7              | 24.4 ± 0.5                      | 0.28 ± 0.01                   | 0.44 ± 0.01       | 0.45 ± 0.01     |
|           | □      | P<sub>oc</sub>mttA_K3 | 23.5 ± 0.6                      | 0.20 ± 0.00                   | 0.46 ± 0.02       | 0.53 ± 0.01     |
|           | ▪      | P<sub>oc</sub>mttA_K8 | 29.1 ± 0.1                      | 0.35 ± 0.01                   | 0.49 ± 0.03       | 0.54 ± 0.01     |
|           | ■      | P<sub>oc</sub>mttA_K9 | 28.9 ± 0.3                      | 0.24 ± 0.00                   | 0.44 ± 0.01       | 0.57 ± 0.01     |
|           | ▬      | P<sub>oc</sub>mttA_K10 | 32.3 ± 0.8                      | 0.37 ± 0.00                   | 0.54 ± 0.01       | 0.60 ± 0.02     |
|           | ▣      | P<sub>oc</sub>mttA_K14 | 29.9 ± 0.7                      | 0.25 ± 0.00                   | 0.44 ± 0.01       | 0.64 ± 0.03     |
| 66 g L<sup>-1</sup> CaCO<sub>3</sub>, 100 g L<sup>-1</sup> glucose | □      | Δ_fuz7              | 48.8 ± 1.3                      | 0.42 ± 0.01                   | 0.70 ± 0.07       | 0.47 ± 0.01     |
|           | ▣      | P<sub>oc</sub>mttA_K3 | 33.8 ± 0.5                      | 0.29 ± 0.00                   | 0.43 ± 0.06       | 0.58 ± 0.04     |
|           | ▤      | P<sub>oc</sub>mttA_K8 | 56.5 ± 1.7                      | 0.48 ± 0.01                   | 0.74 ± 0.09       | 0.51 ± 0.02     |
|           | ▤      | P<sub>oc</sub>mttA_K9 | 48.1 ± 2.9                      | 0.41 ± 0.02                   | 0.52 ± 0.02       | 0.56 ± 0.03     |
|           | ▤      | P<sub>oc</sub>mttA_K10 | 49.0 ± 3.8                      | 0.42 ± 0.03                   | 0.64 ± 0.07       | 0.44 ± 0.03     |
|           | ▤      | P<sub>oc</sub>mttA_K14 | 54.4 ± 0.2                      | 0.46 ± 0.00                   | 0.82 ± 0.01       | 0.57 ± 0.00     |

a. Overall itaconate production rate ([glucose] > 5.5 g L<sup>-1</sup>).  
b. Maximum itaconate production rate.  
c. Yield itaconate per consumed glucose.

3.2. Overexpression of *mttA* from *A. terreus* and its impact on itaconate production

The transport of *cis*-aconitate from the mitochondria to the cytosol is the rate-limiting step in the itaconate production pathway of *U. maydis* [54] [42]. In the ITA chassis this bottleneck is addressed through the overexpression of *mtt1* caused by the promoter exchange of the itaconate cluster regulator encoded by *ria1* [39] [54]. However, the *mttA* transporter from *A. terreus* was shown to cause a higher metabolic flux towards itaconate than its *U. maydis* counterpart *Mtt1* [10] [46]. Overexpression of both *mtt1* and *mttA* further enhances itaconate production. Therefore, a *P<sub>oc</sub>mttA* construct was targeted to the *ip*-locus on the genome of the novel Δ_fuz7 strain.

Targeted integration of CbxR constructs into the *ip*-locus is not perfect. Often, multicopy integration and/or ectopic insertion into random genomic sites occurs. Therefore, several clones were picked to identify the best itaconate producer. Insertion of at least one *mttA* copy was verified by PCR for five clones resulting in the strains listed in Table 3. The strains significantly differed from each other regarding itaconate production, glucose consumption, and growth (Figure 2).
Figure 2. System Duetz® cultivation of *U. maydis* MB215 strains expressing *mttA* in MTM with 50 g L\(^{-1}\) glucose and 100 mM MES, incubated in 24-well plates with a filling volume of 1.5 ml (shaking diameter = 50 mm, n = 300 rpm, T = 30 °C and Φ = 80%). Itaconate production (continuous lines) and glucose consumption (dotted lines) (A) are shown, as well as OD\(_{600}\) (continuous lines) and pH (dotted lines) (B) of *U. maydis* MB215 ∆cyp3 ∆MEL ∆UA ∆dgat ∆P\(_{nia::PETef}\) ∆fuz7 (red) and five *Petef\(_{mttA}\)* transformants named K3 (grey), K8 (green), K9 (orange), K10 (pink), and K14 (black). Error bars indicate the standard error of the mean (n = 3).

With the exception of K3, all *mttA* transformants outperformed the reference strain with 18-32% improvements in titer (Figure 2 A, Table 3). Clones K3, K9, and K14 did not consume all glucose during 120 h of cultivation (Figure 2 A), which further boosted the yield. All *mttA* transformants achieved a higher yield than the reference (Table 3), with K14 reaching 0.64 ± 0.03 g ITA g\(_{Glucose-1}\)\(^{-1}\), which is 89% of the theoretical maximum. Strains K8 and K10 had the highest rates, which were 25% and 32% higher, respectively, compared to the reference. Overexpression of *mttA* also had a strong impact on the growth (Figure 2 B). While *U. maydis* MB215 ∆cyp3 ∆MEL ∆UA ∆dgat ∆P\(_{nia::PETef}\) ∆fuz7 reached OD\(_{600}\) values above 30, all *mttA* transformants remained well below this value. A similar effect was observed in other *mttA* overexpressing strains, where growth and glucose consumption were also strongly decreased [46]. The constitutive *PETef* promoter causes expression of *mttA* during the growth phase [56]. It is assumed that this forces cis-aconitate export from the mitochondria to the cytosol, leading to the observed growth defects.

The trends regarding OD\(_{600}\), yield, and rate were similar under screening conditions with a higher substrate concentration and CaCO\(_3\) as buffer (Figure 3, Table 3). Much higher titers of up to 56.5 ± 1.7 g L\(^{-1}\) were reached compared to the MES-buffered cultivation due to the higher buffer capacity of CaCO\(_3\), higher substrate concentration, and in situ precipitation of calcium itaconate, which alleviates product inhibition.

Figure 3. System Duetz® cultivation of six *U. maydis* MB215 mutants in MTM with 100 g L\(^{-1}\) glucose and 66 g L\(^{-1}\) CaCO\(_3\), incubated in 24-well plates with a filling volume of 1.5 ml (shaking diameter = 50 mm, n = 300 rpm, T = 30 °C and Φ = 80%). Concentrations of itaconate (continuous lines) and glucose (dotted lines) (A) are shown, as well as OD\(_{600}\) (B) of *U. maydis* MB215 ∆cyp3 ∆MEL ∆UA ∆dgat ∆P\(_{nia::PETef}\) ∆fuz7 (red) and five *Petef\(_{mttA}\)* transformants *U. maydis* MB215 ∆cyp3 ∆MEL ∆UA ∆dgat ∆P\(_{nia::PETef}\) ∆fuz7 *Petef\(_{mttA}\)*, K3 (grey), K8 (green), K9 (orange), K10 (pink), and K14 (black). Error bars indicate the standard error of the mean (n = 3).
3.3. Correlation between copy number of \(P_{\text{etef}}mttA\) and impact on itaconate production

In order to test whether these differences in production parameters and growth are a result of differences in the copy number of \(P_{\text{etef}}mttA\), the copy number was determined by quantitative PCR. Primer efficiencies and Ct values of \(mttA\) and two reference genes, UMAG_02595 and UMAG_03726, were 2.014 for JB-126/JB-127 \((R^2 = 0.999)\), 1.994 for JB-128/JB-129 \((R^2 = 0.999)\), and 1.976 for JB-132/JB-133 \((R^2 = 1.0)\). Ratios between \(mttA\) and each reference gene were calculated independently according to Pfaffl et al\(^{195}\) and the resulting mean of both ratios was rounded to an integer value. As positive control, \(U.\ maydis\ MB215\ \Delta\text{UMAG}_05079\ \Delta fuz7\ \Delta P_{\text{etef}}mttA\) was used. This strain was previously proven to be a single-copy \(mttA\) transformant by Southern blot [46], which was confirmed by the qPCR method. For the \(mttA\) transformants, copy numbers between 1 and 4 were determined (Table 4).

| \(U.\ maydis\ strain\) | \(C_t\) value UMAG_02595 | \(C_t\) value UMAG_03726 | \(C_t\) value \(mttA\) | Ratio \(mttA\) to UMAG_02595 | Ratio \(mttA\) to UMAG_03726 | Rounded mean |
|------------------------|-------------------------|-------------------------|-------------------------|-----------------------------|-----------------------------|--------------|
| wildtype               | 27.6 ± 0.10             | 27.9 ± 0.29             | 35.9 ± 1.20             | 0.0                         | 0.0                         | 0            |
| \(\Delta\text{UMAG}_05079\ \Delta P_{\text{etef}}mttA\) | 25.7 ± 0.14             | 25.8 ± 0.24             | 25.7 ± 0.32             | 1.0                         | 1.0                         | 1            |
| \(P_{\text{etef}}mttA\_K3\) | 28.3 ± 0.12             | 28.4 ± 0.31             | 26.7 ± 0.36             | 3.2                         | 3.1                         | 3            |
| \(P_{\text{etef}}mttA\_K8\) | 27.8 ± 0.10             | 27.9 ± 0.11             | 28.5 ± 0.21             | 0.7                         | 0.6                         | 1            |
| \(P_{\text{etef}}mttA\_K9\) | 28.2 ± 0.27             | 28.5 ± 0.16             | 26.4 ± 0.18             | 3.7                         | 3.9                         | 4            |
| \(P_{\text{etef}}mttA\_K10\) | 26.8 ± 0.17             | 26.6 ± 0.35             | 26.6 ± 0.10             | 1.2                         | 0.9                         | 1            |
| \(P_{\text{etef}}mttA\_K14\) | 26.6 ± 0.03             | 27.0 ± 0.13             | 25.2 ± 0.13             | 2.8                         | 3.3                         | 3            |

Cells of strain K8 with a single \(mttA\) copy reached the highest \(OD_{600}\) and itaconate production rate (Figure 4). The transformants with higher copy numbers showed lower \(OD_{600}\) and production rates, but significantly higher yields. Transformant K3 represents an outlier, which is most apparent in the direct comparison to K14 having the same copy number of 3. Possibly, one or more copies of the \(mttA\) construct were inserted into a different locus, which may lead to different expression levels or to defects in growth due to gene disruption. Overall, transformant K14 showed the best balance of high yield with minimal reduction in growth and production rates. Therefore, this strain was selected for further characterization.

**Figure 4.** Production parameters of \(U.\ maydis\ MB215\ \Delta cyp3\ \Delta\text{AMEL}\ \Delta\text{UA}\ \Delta\text{dgat}\ \Delta P_{\text{etef}}\ \Delta fuz7\ \Delta P_{\text{etef}}mttA\) transformants with different \(mttA\) copy numbers incubated in MTM with 50 g L\(^{-1}\) glucose and 100 mM MES (A) and with 100 g L\(^{-1}\) glucose and 66 g L\(^{-1}\) CaCO\(_3\) (B). Error bars indicate the standard error of the mean \((n = 3)\).
3.4. Evaluation of itaconate production with the novel engineered strain K14 in a bioreactor

The final consolidated \textit{U. maydis} MB215 \textit{Δcyt3 ΔMEL ΔUA Δgat ΔPut::P\textit{def} Δfuz7 P\textit{cyp3} \textit{ΔmitA} K14, henceforth named strain K14 for ease of reference, was deeply engineered to reduce by-product formation, to stabilize the yeast-like morphology, and to alleviate bottlenecks in the itaconate production pathway. This engineering strongly impacted growth, and thus might also affect its catalytic vigor under stress. To assess the performance of the novel engineered strain K14 under more industrially relevant conditions, fed-batch bioreactor experiments were performed.

Previously, in situ precipitation of calcium itaconate was successfully used to achieve very high titters with \textit{U. maydis} MB215 \textit{Δcyt3 ΔPut::P\textit{def} Δfuz7 P\textit{cyp3} \textit{ΔmitA}} [10]. Under similar conditions with 200 g L\textsuperscript{-1} glucose and 4 g L\textsuperscript{-1} NH\textsubscript{4}Cl in the presence of CaCO\textsubscript{3}, strain K14 achieved very similar yield, titer and rate of itaconate production (supplemental Table S2). A titer of 205.6 ± 1.1 g L\textsuperscript{-1} itaconate was achieved within 481 h with an overall productivity of 0.43 ± 0.00 g L\textsuperscript{-1} h\textsuperscript{-1} and a yield of 0.32 ± 0.00 g \textit{g\textsuperscript{TA}} \textit{g\textsuperscript{glu}}\textsuperscript{-1} (Figure 5). Although a very high titer was reached under these conditions due to the alleviation of product inhibition, the yield is relatively low compared to the shaken cultures. After the fermentations, extensive clumping of solids attached to reactor walls and components was discovered, indicating that the shown itaconate production from broth samples during cultivation may be an underestimation. Although the in situ precipitation of itaconate is a very promising strategy, it clearly requires extensive optimization of solids feeding and reactor mixing in order to realize the full potential of the engineered strain.

The intermittent stress imposed by the pulsed feeding strategy likely also negatively affected production. This is especially apparent in NaOH-titrated pulsed fed-batch fermentations, where only 60 g L\textsuperscript{-1} itaconate was produced with a yield of 0.42 g \textit{g\textsuperscript{TA}} \textit{g\textsuperscript{glu}}\textsuperscript{-1} (supplemental Figure S1). Although this is a considerable improvement compared to a similar fermentation with \textit{U. maydis} MB215 \textit{Δcyt3 ΔPut::P\textit{def} Δfuz7 P\textit{cyp3} \textit{ΔmitA}} [10] (supplemental Table S2), the achieved yield is still far lower from that obtained in shake flasks. This indicates that the strain modifications, especially the overexpression of \textit{mitA}, reduced the tolerance of the engineered strains to osmotic stress. Consequently, the novel engineered strain was cultivated in fed-batch fermentations with continuous feed to achieve a lower baseline glucose concentration. High-density fermentations with 4 g L\textsuperscript{-1} NH\textsubscript{4}Cl as well as low-density fermentations with 0.8 g L\textsuperscript{-1} NH\textsubscript{4}Cl were performed. A starting glucose concentration of 120 g L\textsuperscript{-1} was allowed to drop to approximately 50 g L\textsuperscript{-1} during growth, at which point a constant feed of 0.75 or 2.8 g h\textsuperscript{-1} was started, for the low and high-density fermentation, respectively.

![Figure 5](https://example.com/figure5.png)

**Figure 5.** High-density pulsed fed-batch fermentation of \textit{U. maydis} strain K14. (A) Concentration of glucose (+) and \textit{OD\textsuperscript{600}} values (▲) and (B) concentration of itaconate (■) during fermentation in a bioreactor containing batch medium (200 g L\textsuperscript{-1} glucose, 4 g L\textsuperscript{-1} NH\textsubscript{4}Cl, CaCO\textsubscript{3} as buffering agent, 30 °C, 1000 rpm, top pitched blade impeller, bottom Rushton impeller). Arrows indicate the addition of 80 g glucose + 50 g CaCO\textsubscript{3} (black arrow), 80 g glucose (grey arrow), or 50 g CaCO\textsubscript{3} (light grey arrow). Error bars indicate the deviation from the mean (n = 2).
Figure 6. High-density fed-batch fermentation with continuous feed of *U. maydis* strain K14. (A) concentration of glucose, itaconate and ammonium, and OD<sub>600</sub> values and (B) filling volume, CO<sub>2</sub> production and the added volumes of 5 M NaOH and 50% glucose during fermentation in a bioreactor containing batch medium with 120 g L<sup>-1</sup> glucose and 4 g L<sup>-1</sup> NH<sub>4</sub>Cl. The pH was kept at 6.5 by automatic titration with NaOH. Cultures were fed with an additional 130 g glucose at a rate of 2.8 g h<sup>-1</sup> during the indicated time interval. Error bars indicate the deviation from the mean (n = 2).

In the high density fermentation, 74.9 ± 1.25 g L<sup>-1</sup> itaconate was produced by *U. maydis* strain K14 within 140 h with an overall productivity of 0.53 ± 0.01 g L<sup>-1</sup> h<sup>-1</sup> and a yield of 0.54 ± 0.02 g<sub>ita</sub> g<sub>glu</sub><sup>-1</sup> (Figure 6). The low density fermentation resulted in a similar itaconate titer of 75.7 ± 1.3 g L<sup>-1</sup>. The five-fold reduction in NH<sub>4</sub>Cl as growth limiting nutrient only resulted in an approximately two-fold reduction of the maximum OD<sub>600</sub> value as well as of the overall production rate (supplemental table S2). A similar trend was observed previously for both itaconate and malate producing strains [47] [42]. The lower substrate requirement for biomass formation enabled a higher yield of 0.66 ± 0.02 g<sub>ita</sub> g<sub>glu</sub><sup>-1</sup> (Figure 7). Compared to the low density pulsed fed batch (supplemental Figure S1), the fed batch with continuous feed increased the titer by 27%, the overall productivity by 14% and the yield by 57%. These results clearly illustrate the benefit of the lower glucose concentration, probably due to a reduced osmotic stress and the absence of osmo shocks caused by glucose pulses. A similar trend was observed for an engineered *U. cynodontis* strain, where a constant glucose feed controlled by an in-line glucose sensor significantly increased the production parameters [25].

The yield of 0.66 ± 0.02 g<sub>ita</sub> g<sub>glu</sub><sup>-1</sup> is the highest yield ever reported for *U. maydis*, and it is also higher than most reported yields achieved with *A. terreus* [41]. In fact, the low density fermentation achieved the theoretical maximal yield during the production phase. When disregarding the glucose consumed during the first 24 h in the growth phase, the yield was 0.72 ± 0.02 g<sub>ita</sub> g<sub>glu</sub><sup>-1</sup>, or 1.00 ± 0.03 mol<sub>ita</sub> mol<sub>glu</sub><sup>-1</sup>. 

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Figure 7. Low-density fed-batch fermentation with continuous feed of *U. maydis* strain K14. (A) concentration of glucose, itaconate and ammonium, and OD<sub>600</sub> values and (B) filling volume, CO<sub>2</sub> production and the added volumes of 5 M NaOH and 50% glucose during fermentation in a bioreactor containing batch medium with 120 g L<sup>-1</sup> glucose, 0.8 g L<sup>-1</sup> NH<sub>4</sub>Cl. The pH was kept at 6.5 by automatic titration with NaOH. Cultures were fed with an additional 130 g glucose at a rate of 0.75 g h<sup>-1</sup> during the indicated time interval. Error bars indicate the deviation from the mean (n = 2).

3. Conclusions

This study explored the limits of microbial itaconic acid production with *U. maydis* by combinatorial metabolic and morphological engineering. These modifications, especially the overexpression of *mttA*, had a major effect on growth of the final strain *U. maydis* K14. This reduction in growth did affect its performance in fed-batch reactors, but this effect could be avoided by reducing osmotic stress with a continuous glucose feed. Under these conditions, itaconate was produced from glucose at 100% of the theoretical maximum yield during the production phase in a low density fermentation. Looking forward, osmotolerance of *U. maydis* may be enhanced by laboratory evolution. Further, production of itaconic acid at low pH values is also paramount. Although *U. maydis* grows relatively poorly at low pH, the shaken cultures clearly indicate that the K14 strain is able to produce itaconic acid at pH levels below 4. A pH shift between growth and production may therefore enable low-pH production. Since substrate cost is usually the main price determining factor for commodity products, the high yield achieved in this work will significantly contribute to the establishment of an *Ustilago*-based industrial itaconate production process, further enabled by the facile, yeast-like growth of this strain.
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Conflicts of Interest: N. Wierckx, L. M. Blank and H. Hosseinpour Tehrani are inventors of patent applications on morphological engineering and organic acid production with Ustilago and related fungi. The other authors declare no conflict of interest.

References

1. Okabe, M.; Lies, D.; Kanamasa, S.; Park, E.Y. Biotechnological production of itaconic acid and its biosynthesis in Aspergillus terreus. Applied Microbiology and Biotechnology 2009, 84, 597-606, doi:10.1007/s00253-009-2132-3.

2. Willke, T.; Vorlop, K.D. Biotechnological production of itaconic acid. Appl Microbiol Biotechnol 2001, 56, 289-295, doi:10.1007/s002530100685.

3. Klement, T.; Milker, S.; Jäger, G.; Grande, P.M.; Domínguez de María, P.; Büchs, J. Biomass pretreatment affects Ustilago maydis in producing itaconic acid. Microbial Cell Factories 2012, 11, 43-43, doi:10.1186/1475-2859-11-43.

4. Steiger, M.G.; Wierckx, N.; Blank, L.M.; Mattanovich, D.; Sauer, M. Itaconic acid—an emerging building block. Industrial biotechnology, products and processes. Weinheim, Germany: Wiley-VCH Verlag GmbH & Co. KGaA 2017.

5. Werpy, T.; Petersen, G. Top Value Added Chemicals from Biomass: Volume I -- Results of Screening for Potential Candidates from Sugars and Synthesis Gas; DOE/GO-102004-1992; TRN: US200427%%671 United States 10.2172/15008859 TRN: US200427%%671 NREL English; ; National Renewable Energy Lab., Golden, CO (US): 2004; p Medium: ED; Size: 76 pp. pages.

6. Michelucci, A.; Cordes, T.; Ghelfi, J.; Pailot, A.; Reiling, N.; Goldmann, O.; Binz, T.; Wegner, A.; Tallam, A.; Rausell, A., et al. Immune-responsive gene 1 protein links metabolism to immunity by catalyzing itaconic acid production. Proceedings of the National Academy of Sciences 2013, 110, 7820-7825, doi:10.1073/pnas.1218599110.

7. Mills, E.L.; Ryan, D.G.; Prag, H.A.; Dikovskaya, D.; Menon, D.; Zaslona, Z.; Jedrychowski, M.P.; Costa, A.S.H.; Higgins, M.; Hams, E., et al. Itaconate is an anti-inflammatory metabolite that activates Nrf2 via alkylation of KEAP1. Nature 2018, 556, 113-117, doi:10.1038/nature25986.

8. Olagnier, D.; Farahani, E.; Thesis, J.; Blay-Cadanet, J.; Herengt, A.; Idorn, M.; Haid, A.; Hernæz, B.; Knudsen, A.; Iversen, M.B., et al. SARS-CoV2-mediated suppression of NRF2-signaling reveals potent antiviral and anti-inflammatory activity of 4-octyl-itaconate and dimethyl fumarate. Nature Communications 2020, 11, 4938, doi:10.1038/s41467-020-18764-3.
9. Klement, T.; Bächs, J. Itaconic acid – A biotechnological process in change. *Bioresource Technology* **2013**, *135*, 422-431, doi:10.1016/j.biortech.2012.11.141.

10. Hosseinpour Tehrani, H.; Becker, J.; Bator, I.; Saur, K.; Meyer, S.; Rodrigues Lóia, A.C.; Blank, L.M.; Wierckx, N. Integrated strain- and process design enable production of 220 g L–1 itaconic acid with *Ustilago maydis*. *Biotechnology for Biofuels* **2019**, *12*, 263, doi:10.1186/s13068-019-1605-6.

11. Hevekerl, A.; Kuenz, A.; Vorlop, K.-D. Influence of the pH on the itaconic acid production with *Aspergillus terreus*. *Applied microbiology and biotechnology* **2014**, *98*, 10005-10012.

12. Tabuchi, T.; Sugisawa, T.; Ishidori, T.; Nakahara, T.; Sugiyama, J. Itaconic Acid Fermentation by a Yeast Belonging to the Genus *Candida*. *Agricultural and Biological Chemistry* **1981**, *45*, 475-479, doi:10.1080/00021369.1981.10864534.

13. Levinson, W.E.; Kurtzman, C.P.; Kuo, T.M. Production of itaconic acid by *Pseudozyma antarctica* NRRL Y-7808 under nitrogen-limited growth conditions. *Enzyme and Microbial Technology* **2006**, *39*, 824-827, doi:https://doi.org/10.1016/j.enzmictec.2006.01.005.

14. Krull, S.; Lünsmann, M.; Prüße, U.; Kuenz, A. *Ustilago Rabenhorstiana*—An Alternative Natural Itaconic Acid Producer. *Fermentation* **2020**, *6*, 4.

15. Wierckx, N.; Agrimi, G.; Lübeck, P.S.; Steiger, M.G.; Mira, N.P.; Punt, P.J. Metabolic specialization in itaconic acid production: a tale of two fungi. *Current Opinion in Biotechnology* **2020**, *62*, 153-159, doi:https://doi.org/10.1016/j.copbio.2019.09.014.

16. Regestein, L.; Klement, T.; Grande, P.; Kreyenschulte, D.; Heyman, B.; Maßmann, T.; Eggert, A.; Sengpiel, R.; Wang, Y.; Wierckx, N., et al. From beech wood to itaconic acid: case study on biorefinery process integration. *Biotechnology for Biofuels* **2018**, *11*, 279, doi:10.1186/s13068-018-1273-y.

17. Straathof, A.J.J.; Wahl, S.A.; Benjamin, K.R.; Takors, R.; Wierckx, N.; Noorman, H.J. Grand Research Challenges for Sustainable Industrial Biotechnology. *Trends in Biotechnology* **2019**, *37*, 1042-1050, doi:https://doi.org/10.1016/j.tibtech.2019.04.002.

18. Kämper, J.; Kahmann, R.; Bölk, M.; Ma, L.-J.; Brefort, T.; Saville, B.J.; Banuett, F.; Kronstad, J.W.; Gold, S.E.; Müller, O., et al. Insights from the genome of the biotrophic fungal plant pathogen *Ustilago maydis*. *Nature* **2006**, *444*, 97-101, doi:10.1038/nature5248.

19. Kahmann, R.; Steinberg, G.; Basse, C.; Feldbrügge, M.; Kämper, J. *Ustilago maydis*, the Causative Agent of Corn Smut Disease. 2000; 10.1007/978-94-015-9546-9_12pp. 347-371.

20. Christensen, J.J. Corn smut caused by *Ustilago maydis*. *Monographs. American Phytopathology Society* **1963**, *2*.

21. Klose, J.; De Sá, M.M.; Kronstad, J.W. Lipid-induced filamentous growth in *Ustilago maydis*. *Molecular microbiology* **2004**, *52*, 823-835, doi:10.1111/j.1365-2958.2004.04019.x.

22. Lovely, C.B.; Aulakh, K.B.; Perlin, M.H. Role of Hsl7 in morphology and pathogenicity and its interaction with other signaling components in the plant pathogen *Ustilago maydis*. *Eukaryotic cell* **2011**, *10*, 869-883, doi:10.1128/ec.00237-10.

23. Lovely, C.B.; Perlin, M.H. Cla4, but not Rac1, regulates the filamentous response of *Ustilago maydis* to low ammonium conditions. *Commun Integr Biol* **2011**, *4*, 670-673, doi:10.4161/cib.17063.

24. Hosseinpour Tehrani, H.; Tharmasothirajan, A.; Track, E.; Blank, L.M.; Wierckx, N. Engineering the morphology and metabolism of pH tolerant *Ustilago cynodontis* for efficient itaconic acid production. *Metabolic Engineering* **2019**, *54*, 293-300, doi:https://doi.org/10.1016/j.ymben.2019.05.004.

25. Hosseinpour Tehrani, H.; Saur, K.; Tharmasothirajan, A.; Blank, L.M.; Wierckx, N. Process engineering of pH tolerant *Ustilago cynodontis* for efficient itaconic acid production. *Microbial Cell Factories* **2019**, *18*, 213, doi:10.1186/s12934-019-1266-y.
26. Guevarra, E.D.; Tabuchi, T. Accumulation of Itaconic, 2-Hydroxyparaconic, Itatartaric, and Malic Acids by Strains of the Genus Ustilago. *Agricultural and Biological Chemistry* 1990, 54, 2353-2358, doi:10.1080/00021369.1990.10870333.

27. Feldbrügge, M.; Kellner, R.; Schipper, K. The biotechnological use and potential of plant pathogenic smut fungi. *Applied Microbiology and Biotechnology* 2013, 97, 3253-3265, doi:10.1007/s00253-013-4777-1.

28. Geiser, E.; Wiebach, V.; Wierckx, N.; Blank, L.M. Prospecting the biodiversity of the fungal family Ustilaginaceae for the production of value-added chemicals. *Fungal Biology and Biotechnology* 2014, 1, 2, doi:10.1186/s40694-014-0002-y.

29. Bölker, M.; Basse, C.W.; Schirawski, J. *Ustilago maydis* secondary metabolism-from genomics to biochemistry. *Fungal genetics and biology* : FG & B 2008, 45 Suppl 1, S88-93, doi:10.1016/j.fgb.2008.05.007.

30. Aguilar, L.R.; Pardo, J.P.; Lomeli, M.M.; Bocardo, O.I.L.; Juárez Oropeza, M.A.; Guerra Sánchez, G. Lipid droplets accumulation and other biochemical changes induced in the fungal pathogen *Ustilago maydis* under nitrogen-starvation. *Archives of microbiology* 2017, 199, 1195-1209, doi:10.1007/s00203-017-1388-8.

31. Moon, H.-J.; Jeya, M.; Kim, I.-W.; Lee, J.-K. Biotechnological production of erythritol and its applications. *Applied microbiology and biotechnology* 2010, 86, 1017-1025.

32. Zambanini, T.; Tehrani, H.H.; Geiser, E.; Sonntag, C.K.; Buescher, J.M.; Meurer, G.; Wierckx, N.; Blank, L.M. Metabolic engineering of *Ustilago trichophora* TZ1 for improved malic acid production. *Metabolic engineering communications* 2017, 4, 12-21.

33. Müller, M.J.; Stachurski, S.; Stoffels, P.; Schipper, K.; Feldbrügge, M.; Büchs, J. Online evaluation of the metabolic activity of *Ustilago maydis* on (poly) galacturonic acid. *Journal of Biological Engineering* 2018, 12, 1-17.

34. Schlembach, I.; Tehrani, H.; Blank, L.M.; Büchs, J.; Wierckx, N.; Regestein, L.; Rosenbaum, M. Consolidated bioprocessing of cellulose to itaconic acid by a co-culture of *Trichoderma reesei* and *Ustilago maydis*. 2020.

35. Geiser, E.; Reindl, M.; Blank, L.M.; Feldbrügge, M.; Wierckx, N.; Schipper, K. Activating intrinsic carbohydrate-active enzymes of the smut fungus *Ustilago maydis* for the degradation of plant cell wall components. *Applied and environmental microbiology* 2016, 82, 5174-5185.

36. Stoffels, P.; Müller, M.J.; Stachurski, S.; Terfrüchte, M.; Schröder, S.; Ihling, N.; Wierckx, N.; Feldbrügge, M.; Schipper, K.; Büchs, J. Complementing the intrinsic repertoire of *Ustilago maydis* for degradation of the pectin backbone polygalacturonic acid. *Journal of biotechnology* 2020, 307, 148-163.

37. Maassen, N.; Panakova, M.; Wierckx, N.; Geiser, E.; Zimmermann, M.; Bölker, M.; Klinner, U.; Blank, L.M. Influence of carbon and nitrogen concentration on itaconic acid production by the smut fungus *Ustilago maydis*. *Engineering in Life Sciences* 2014, 14, 129-134.

38. Olicón-Hernández, D.R.; Araiza-Villanueva, M.G.; Pardo, J.P.; Aranda, E.; Guerra-Sánchez, G. New Insights of *Ustilago maydis* as Yeast Model for Genetic and Biotechnological Research: A Review. *Current Microbiology* 2019, 1-10.

39. Becker, J.; Hosseinpour Tehrani, H.; Gauert, M.; Mampel, J.; Blank, L.M.; Wierckx, N. An *Ustilago maydis* chassis for itaconic acid production without by-products. *Microbial Biotechnology* 2020, 13, 350-362, doi:10.1111/1751-7915.13525.

40. Karaffa, L.; Kubicek, C.P. Citric acid and itaconic acid accumulation: variations of the same story? *Applied Microbiology and Biotechnology* 2019, 103, 2889-2902, doi:10.1007/s00253-018-09607-9.

41. Kuenz, A.; Krull, S. Biotechnological production of itaconic acid—things you have to know. *Applied Microbiology and Biotechnology* 2018, 102, 3901-3914, doi:10.1007/s00253-018-8895-7.
42. Geiser, E.; Przybilla, S.K.; Engel, M.; Kleineberg, W.; Büttner, L.; Sarikaya, E.; Hartog, T.d.; Klankermayer, J.; Leitner, W.; Bölker, M., et al. Genetic and biochemical insights into the itaconate pathway of *Ustilago maydis* enable enhanced production. *Metabolic Engineering* **2016**, *38*, 427-435, doi:https://doi.org/10.1016/j.mynb.2016.10.006.

43. Zambanini, T.; Hosseinpour Tehrani, H.; Geiser, E.; Merker, D.; Schleese, S.; Krabbe, J.; Buescher, J.M.; Meurer, G.; Wierckx, N.; Blank, L.M. Efficient itaconic acid production from glycerol with *Ustilago vetiveriae* TZ1. *Biotechnology for Biofuels* **2017**, *10*, 131, doi:10.1186/s13068-017-0809-x.

44. Duetz, W.A.; Rüedi, L.; Herrmann, R.; O’Connor, K.; Büchs, J.; Witholt, B. Methods for Intense Aeration, Growth, Storage, and Replication of Bacterial Strains in Microtiter Plates. *Applied and Environmental Microbiology* **2000**, *66*, 2641-2646, doi:10.1128/aem.66.6.2641-2646.2000.

45. Hewald, S.; Josephs, K.; Bölker, M. Genetic analysis of biosurfactant production in *Ustilago maydis*. *Applied and environmental microbiology* **2005**, *71*, 3033-3040.

46. Hosseinpour Tehrani, H.; Geiser, E.; Engel, M.; Hartmann, S.K.; Hossain, A.H.; Punt, P.J.; Blank, L.M.; Wierckx, N. The interplay between transport and metabolism in fungal itaconic acid production. *Fungal genetics and biology : FG & B* **2019**, *125*, 45-52, doi:10.1016/j.fgb.2019.01.011.

47. Zambanini, T.; Sarikaya, E.; Kleineberg, W.; Buescher, J.M.; Meurer, G.; Wierckx, N.; Blank, L.M. Efficient malic acid production from glycerol with *Ustilago trichophora* TZ1. *Biotechnology for Biofuels* **2016**, *9*, 67, doi:10.1186/s13068-016-0483-4.

48. Gibson, D.G.; Young, L.; Chuang, R.Y.; Venter, J.C.; Hutchison, C.A., 3rd; Smith, H.O. Enzymatic assembly of DNA molecules up to several hundred kilobases. *Nature methods* **2009**, *6*, 343-345, doi:10.1038/nmeth.1318.

49. Sambrook, J.; Russell, D.W. Preparation and Transformation of Competent *E. coli* Using Calcium Chloride. *CSH protocols* **2006**, doi:10.1101/pdb.prot3932.

50. Brachmann, A.; König, J.; Julius, C.; Feldbrügge, M. A reverse genetic approach for generating gene replacement mutants in *Ustilago maydis*. *Molecular Genetics and Genomics* **2004**, *272*, 216-226, doi:10.1007/s00438-004-1047-z.

51. Hoffman, C.S.; Winston, F. A ten-minute DNA preparation from yeast efficiently releases autonomous plasmids for transformation of *Escherichia coli*. *Gene* **1987**, *57*, 267-272, doi:https://doi.org/10.1016/0378-1119(87)90131-4.

52. Khrunyk, Y.; Münch, K.; Schipper, K.; Lupas, A.N.; Kahmann, R. The use of FLP-mediated recombination for the functional analysis of an effector gene family in the biotrophic smut fungus *Ustilago maydis*. *New Phytologist* **2010**, *187*, 957-968, doi:10.1111/j.1469-8137.2010.03413.x.

53. Pfaffl, M.W. A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Res* **2001**, *29*, e45, doi:10.1093/nar/29.9.e45.

54. Geiser, E.; Przybilla, S.K.; Friedrich, A.; Buckel, W.; Wierckx, N.; Blank, L.M.; Bölker, M. *Ustilago maydis* produces itaconic acid via the unusual intermediate trans-aconitate. *Microbial Biotechnology* **2016**, *9*, 116-126, doi:10.1111/1751-7915.12329.

55. Geiser, E.; Hosseinpour Tehrani, H.; Meyer, S.; Blank, L.M.; Wierckx, N. Evolutionary freedom in the regulation of the conserved itaconate cluster by Ria1 in related Ustilaginaceae. *Fungal Biology and Biotechnology* **2018**, *5*, 14, doi:10.1186/s40694-018-0058-1.

56. Zambanini, T.; Hartmann, S.K.; Schmitz, L.M.; Büttner, L.; Hosseinpour Tehrani, H.; Geiser, E.; Beudels, M.; Venc, D.; Wandrey, G.; Büchs, J., et al. Promoters from the itaconate cluster of *Ustilago maydis* are induced by nitrogen depletion. *Fungal Biology and Biotechnology* **2017**, *4*, 11, doi:10.1186/s40694-017-0040-3.
