Effects of foliar and root application of epsom salt on aquaponics beetroot (*Beta vulgaris*) production in confined condition

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Abstract: The study was conducted to evaluate the impact of epsom salt, rich in magnesium and sulfur content beneficial to plant, on beetroot production in aquaponics system. Media based aquaponics systems (nine units) were employed for beetroot and tilapia culture where three treatments T₁, T₂ and T₀ were applied with three replications (R₁, R₂, and R₃) each. Epsom salt was applied in T₁ as foliar spray in the form of 5% solution in each replication, whereas same amount of epsom salt (5 g) was directly applied on vegetable bed in T₂ as root application. But in control (T₀) no epsom salt was used. Beetroot saplings were planted in retrospect at a rate of two saplings per growbed (replication). After thirty days, plants grew enough to support the water clarification in aquaponics, ten tilapia fry were stocked per tank (90 L) in each replication. Fish were fed commercial floating feed containing 30% protein. Although the fish growth parameters in three treatments were statistically similar, the highest mean length gain, weight gain, specific growth rate and fish production were found 5.11±1.64 cm, 30.16±0.11 g, 0.39±0.01 %/day and 26.27±2.47 tons/ha/60 days respectively in T₁ followed by T₂ and T₀. However, response of beetroots to different treatments was statistically different. The highest average leaf area (69.79 ± 0.48 cm²), length of root (31.03±0.43 cm), weight of root (21.17±1.92 g), and production of beetroot 8.67±1.54 tons/ha/90 days were found in T₁ followed by T₂ and T₀. Harvesting time was also reduced in epsom salt based aquaponics system. The overall production of beetroot and tilapia was higher in T₁ (foliar spray) than T₂ (root application) and T₀ (control). Thus, the study signifies the potentials of epsom salt in aquaponics vegetable production. However, further research is needed to verify the dose response to epsom salt in aquaponics system.

Keywords: aquaponics; epsom salt; foliar application; root application; beetroot

1. Introduction

Aquaponics, a perfect engineering of ecology, is the combination of aquaculture (fish or crustaceans) and hydroponic cultivation of plants (soilless production) in a re-circulating system, utilizing the nutrients present in the aquaculture effluents to produce plants with commercial value (Yildiz et al., 2017). Catastrophic climatic change, reduced agricultural land and demand of safe food gradually transfer aquaponics technology to an essential part of life for producing organic food at our dwelling. Beetroot (*Beta vulgaris*) is generally known as garden beet. It is native to Mediterranean region but is widely cultivated in America, Europe and throughout Indian subcontinent. It is being traditionally used in medicine, specifically for the treatment of fertility, cancer, hypertension and urinary tract disorders (Chawla et al., 2016). It makes a wonderful dietary supplement being not only rich in nutrients, minerals, amino acids and vitamins but also has unique phyto-constituents, which have numerous medicinal properties such as anti-oxidant, anti-depressant, anti-microbial, anti-inflammatory, diuretic and expectorant properties. It is one of the natural foods,
which boosts the energy to athletes. It is also used as natural food color in dairy and meat products. Traditionally, beetroot is consumed as food and has high nutritive value. It is now being recognized as a functional food (Ninfáli, 2013). Beetroot contains alkaloids (128.889), steroids (16.4), glycosides (0.652), flavonoids (6.417), terpenoids (115.5), saponins (3.789), and acidity level (5.227) all in mg/100g. The elemental analysis of Beta vulgaris includes the presence of the following minerals: iron, magnesium, copper, sodium, potassium, manganese, calcium and zinc in the ratio 0.76, 18.60, 0.08, 73.60, 31.20, 0.86, 13.80 and 0.29 mg/100 g respectively. Vitamins were also analyzed as vitamin A (2.6 μg/100 g), vitamin K (3.2 μg/100 g), vitamin C, vitamin E, vitamin B1, vitamin B6, vitamin B2, pantothentic acid and cholesterol 4.36, 0.18, 0.35, 0.03, 90.053, 0.034, 0.151 and 0.04 mg/100 g, respectively (Odoh and Okoro, 2013). Because of these potentialities, beetroot was attempted to culture in aquaponics system. However, aquaponics has some limited nutrients supply as it take the traditional soil growbed. Several efforts have been made to supplement aquaponically grown plants with nutrients (Bethe et al., 2017; Zahan et al., 2018). As a continuation of the endeavor, epsom salt (MgSO₄ 7H₂O) has been selected to supply additional minerals in this system. Plants require seventeen essential nutrients for proper development among which magnesium and sulfur are two commonly deficient macronutrients that could be supplemented to plants with epsom salt (hydrated magnesium sulfate) (Kristek et al., 1997; Epstein and Bloom, 2005).

Epsom salt was chosen due to its high solubility in water, vital elements for plant growth, quick nutrient uptake by plants, traditional use as plant fertilizer and ease application as foliar spray. Epsom salt is not true salt, but it is known as a salt due to its chemical structure. It consists of mixture of magnesium and sulfur. In water epsom salt dissociates into magnesium ions, and sulfate ions. These ions are easy to absorb through the plant cell and improves the benefits of epsom salt. Magnesium and sulfur play a vital role in the plant physiology by regulating the enzymatic activity. In practical, magnesium induces chlorophyll synthesis and sulfur activates that chlorophyll for photosynthesis in plants. Besides, these two minerals improves nutrients (mostly nitrogen and phosphorous) absorption capacity in plant, therefore increase plants’ ability to produce flowers and fruits. Consequently, using epsom salt promises a bushier and greener garden (Orlovius, 2001; Merhaut, 2007; Tilley 2016; Elbossaty, 2018)

In order to assure favorable plant growth, nutrients must be available not only in sufficient amounts, but also in appropriate ratios (Maathuis, 2009). Foliar spray is an effective way to avail nutrients for plant growth. As for examples, foliar spray of 30% magnesium (Mg) solution rectified Mg deficiency in tea plants within 14 days of spraying and the growth rate increased by 16 to 13.45% whereas a 20% Mg solution could only correct the insufficiency after a second application (Obatolu, 1999). In another research, foliar spray of 5% epsom solution on sugar beets increased sucrose contents by 0.25 and 0.20% (Kristek et al., 1997). In aquaponics, foliar spray of molasses and compost tea also yield increased production of water spinach (Bethe et al., 2017). Therefore, the present experiment was conducted to assess the effects of epsom salt as foliar spray and root application on beetroot production in aquaponics system. Notably, monosex tilapia (Oreochromis niloticus) was preferably selected to culture in aquaponics system because of hardy and fast growing features of the fish species (Rakocy et al., 2011).

2. Materials and Methods
2.1. Site selection
The experiment was conducted at the Aquaponics Laboratory (Aquaponics Oasis), Department of Aquaculture, Faculty of Fisheries, Bangladesh Agricultural University (BAU), Mymensingh from 17th February to 18th May, 2018.

2.2. Experimental design
The experimental design was comprised of nine sets of aquaponics unit where nine fish holding tanks (diameter 58 cm, height 45 cm) each containing 120 liter of water and nine plastic containers for vegetable growing beds (38.5 cm length, 28.5 cm width, 22 cm height) were installed. There were two treatments (T₁ and T₂) with epsom salt application (foliar spray and direct application on plant grow bed respectively) and a control (T₀) with no epsom application. Each treatment (T) were applied on three separate aquaponics unit as replication (R), therefore denoted as T₁R₁, T₁R₂, T₁R₃, T₂R₁, T₂R₂, T₂R₃, T₀R₁, T₀R₂ and T₀R₃ Which were placed randomly (Figure 1).

2.3. Bed preparation for beetroot cultivation
Plastic containers which were locally available, cheap and good quality used as beetroot culture beds (plant growbeds). Each container (38.5 cm length, 28.5 cm width, 22 cm height) was cut longitudinally 1.27cm inch
below from the upper surface to make it a bed, cleaned with detergent, rinse with clean water for 3 to 4 times and sun dried before use. A hole was made above 0.5 inch from the bottom to facilitate clean water passage from the beetroot bed to the fish tank. Each grow bed was filled with brick lets of 2-5 cm sizes as plant growing substrates. The plastic containers were placed on a wooden structure for convenient management.

2.4. Planting beetroot saplings
For production of beetroot saplings first the seeds were soaked for 24 hours. Then these were replaced to a plastic bucket containing 50% coco dust and 50% vermi-compost and covered with paper. Before that a pore was created into the plastic bucket 0.5 inch above the bottom for emergency water exit. After germination 2 saplings were planted in two opposite corners of each growbed.

2.5. Application of Epsom salt
Locally available epsom salt was collected and prepared for application in T₁ and T₂. In T₁, 5% epsom salt was applied as foliar spray on beetroot plants of each replication (R). Foliar spray for each plant growbed was prepared by dissolving 5 g of epsom salt in filtered water to make the final volume 100ml and sprayed on plants fortnightly. Whereas, 5g of epsom salt was applied directly on each growbed fortnightly in T₂. T₀ was control, therefore no epsom was supplemented. The concentration of epsom salt was selected according to Kristek et al., 1997.

2.6. Fish tank preparation
Nine fish holding tanks (diameter 58 cm, height 45 cm) with the volume of 90 L of water were used for fish culture. The tanks were prepared by washing, drying, setting up pipes, filling up with water. The tanks were brought from local market. At first tanks were uncovered and then these were washed with disinfectant to remove chemicals if any. An 18 watt submersible pump for watering the vegetables and one air pump with two air stones for oxygen supply were set in each aquaponics unit.

2.7. Stocking and rearing the fish
Monosex tilapia (Oreochromis niloticus) were collected from a renowned hatchery. During transportation oxygenated polythene bags that contained 2/3 oxygen and 1/3 water was used. Acclimatization helps to minimize stress of fingerlings and to adapt with the differences of water temperature, pH, and oxygen between the transported bag and the fish tank water. For acclimatization, the polythene bag containing fish was kept in fish tank for one hour to reduce temperature difference between polythene bag and fish tank. After acclimatization the bag was untied and created current by hand so that the fish can move freely into the tank. The fish which could not swim freely against water current was discarded. Ten fish were stocked in each tank containing 90 L of water. Fish were stocked after thirty days of beetroot plantation to allow beetroot sapling grow enough to support the recirculation process in aquaponics.

2.8. Fish and vegetable sampling and harvesting
Fish and vegetables were sampled fortnightly. During each sampling, 10% fish of each tank (each replication) were caught randomly with scoop net and individual length-weight was measured with an electronic compact balance and wooden measuring scale to ascertain fish growth performance. Number of leaves, leaf area, length and weights of plants and roots were also recorded during each sampling. All data were recorded in notebook. After recording length and weight, fish were released in respective tank. After 90 days of experiment, fish and beetroots were cropped on the day of final harvest.

2.9. Water quality parameters
Physico chemical parameters of tank water were measured to know the suitability of fish culture. Temperature, DO (dissolved oxygen) and pH were measured every 7 days interval. Electric conductivity (EC), carbonate (CO₃), hydrogen carbonate (HCO₃), potassium (K), total nitrogen (N), sulfur (S) and sodium (Na) content of aquaponics water were also measured at monthly interval. The tests were done in the Humboldt Soil Testing Laboratory, Soil Science department, Bangladesh Agricultural University.

2.10. Data processing and analysis
The data were processed for analysis using "Microsoft Excel 2010". The collected data were summarized carefully before final tabulation. Data were analyzed by using one way analysis of variance (ANOVA) and Duncan’s new multiple range test (DMRT). R-Stat software was also employed to visualize the significance of
the treatments. Final results have been depicted in tabular and graphical forms.

3. Results and Discussion

3.1. Plant growth parameters

The growth of beetroot plants was measured at different sampling date throughout the study period and production of beetroot was obtained at final harvest. The highest mean height of the plant was 64.8±2.65 cm in T1 followed by 59.17±4.75 cm and 43.68±1.71 cm in T2 and T0. The epsom salt effects might be the possible reasons for significant differences of plant height among the treatments. Salam et al. (2014) found the highest 85.58±1.53 cm of tomato plant height and Parvin (2016) obtained the highest taro plant height of 75.71±20.15 cm in aquaponics system. The height of beetroot in the present study was less than the other research findings which might be due to species variation. The highest mean leaf number, leaf area and weight of leaf were observed during the final harvest valued as 9.53±0.15, 69.79±0.48 cm² and 41.91±2.41 g respectively in T1 whereas the lowest values were 7.99±0.18, 50.52±0.81 cm² and 31.08±3.11 g respectively that obtained in T0. Notably, there were significant differences in mean leaf number, leaf area and leaf weight of beetroot plants among the treatments (Table 1).

Root length and weight were also measured on the final harvesting date. The significantly highest length and weight of roots were 31.03±0.43 cm and 21.17±1.92 g obtained in T1 followed by T2 and T0 whereas the lowest were found 19.48±1.63 cm and 14.24±1.01 g in T0 (Table 1). Remarkably, the differences among the mean values of the parameters were significant therefore suggesting positive effects of epsom salt on the plants wellbeing.

The highest and lowest weight of beetroot plants were 160.11±1.51 g in T1 and 129.06±3.05 g in T0 respectively (Table 1). Salam et al. (2014) found that the plant weight of tomato in aquaponics system was 109.59±116.72 g, even as Afrin (2018) found the highest weight of cauliflower plant 170.43 g in aquaponics system which has similarity to the present findings.

3.2. Beetroot production

The highest and lowest individual mean weights of harvested beetroot were 95.28±6.40 g and 73.87±2.71 g in T1 and T0 respectively (Table 1). This might be due to the epsom salt effects on beetroot production as Elamin and Wilcox (1985) also concluded that tomato production was increased by 27.9% with Mg application.

Similarly, the significantly highest beetroot production was obtained in T1 figured as 8.67±1.43 tons/ha/90 days. Whereas T2 produced 7.48±2.71 tons/ha/90 days which was also significantly higher than T0 (6.61±1.82 tons/ha/90 days) (Figure 2). As because all other growth factors were maintained similar in all the treatments over the study period, it could be concluded that addition of epsom salt raised the beetroot production and improved plants’ wellbeing. And foliar spray of epsom is much effective than direct application on growbeds. Similarly, Bethe et al. (2017) also concluded that foliar spray with compost tea and molasses had positive impacts on aquaponically grown water spinach (Ipomoea aquatica) than no spray in control. Although not certain, but foliar spray might effects more positively as it is applied on plant leaves and stems directly whereas in growbed application roots are responsible for mineral transportation to other parts of plants. Therefore, foliar spray in T1 might have better effects on beetroots than direct growbed application of epsom salt in T2. However, Anna and Maria (2013) carried out a field experiment with beetroot employing conventional, integrated and organic methods and found production of 16.14, 29.42 and 28.80 tons/ha/90 days in conventional, integrated and organic methods respectively. The production of beetroot in the present experiment was lower than the above findings due to high temperature (air temperature ranged 27-32°C over the period). Beetroots require cool temperature and optimum growing temperature is 12-19°C (Odoh, 2013).

3.3. Proximate composition of beetroot

The proximate compositions of beetroots were nearly similar in in T1 and T2 which were comparatively better than T0 except the carbohydrate content. The highest average value of lipid was 1.25±0.01 in T1 and lowest was 1.1±0.1 in T0. The highest mean protein and moisture contents were 0.97±0.01% and 89.46±0.58% in T2 respectively. The highest mean ash content was 0.94±0.01 in T3 and the lowest was 0.93±0.29 in T0. The highest mean fiber was 6.2±0.1 in both T1 and T2 and the lowest was 5.93±0.29 in T0. The highest mean carbohydrate was 21±0.72 obtained in T3 and the lowest was 0.95±0.67 in T2 (Table 2). Odoh et al. (2013) conducted a study on Quantitative phytochemical (proximate/nutritive) composition analysis of beetroot (Beta vulgaris) and stated that it contains 1.35, 0.3, 2.56, 87.4 and 1.4 % of protein, fats and oils, fiber, moisture, and ash value respectively. The lipid, fiber and moisture contents were slightly higher and the percentage of protein was less...
in the present study than the above findings. However, the significantly highest nutritive value was found in T₁ and T₂ than T₀ that implies epsom salt gave the better supplementation of nutrients in T₁ and T₂.

The proximate composition was analyzed on fresh weight basis. Proximate composition of beetroot under different treatments is given in Table 2.

In this experiment, beetroots’ performance was considerably better in T₁ and T₂ rather T₀. All these positive responses might be attributed to the presence of the two very crucial elements for plants’ growth namely sulfur and magnesium in the supplied epsom salt (hydrated magnesium sulfate). It is well known from the literature that, magnesium helps chlorophyll formation in plants therefore aid in photosynthesis and ease nutrient uptake by plants. Although, magnesium is mostly available in the traditional soil, aquaponics plants (grown without soil) could easily suffer from magnesium deficiency and subsequent leaf curing as well as stunted growth. Similarly, sulfur regulates inner working of plants and is not as available in aquaponics substrates as in traditional soil (Kristek et al., 1997). Again, the experimental plant “beetroot” is one of the high mineral demanding plants therefore field trials (1998-2000) implied that foliar application of epsom salt with the addition of manganese and boron increased yield of sugar beets by 3–5% (Orlovius, 2001). So, it is justified that magnesium and sulfur are to be supplied in aquaponically grown beetroot plants to activate plant proteins and enzymes for plants’ wellbeing. Besides these, epsom salt has additional benefits on water clarification in aquaponics. Epsom salt has been reported to increase nitrate uptake of plants from fish waste water that are produced by nitrifying bacteria residing the plant root system in aquaponics (Nell et al., 2006).

### 3.4. Fish growth performance and production

During foliar application of Epsom salt the vegetable beds were kept covered with polyethylene sheet and during root (growbed) application water pumps were kept stopped for 15-20 minutes to avoid epsom salt get mixed with fish tank water. Therefore, epsom salt had negligible or no effect on fish wellbeing reared in tanks. Tilapia (Oreochromis niloticus) was cultured for 60 days. The initial mean length of tilapia stocked in different treatments was 15.13±1.84 cm and final mean length were 21.30 ± 1.60, 19.17 ±0.44, and 18.58 ±0.33 cm in T₁, T₂ and T₀, respectively. The initial mean weight was 55.12±7.02 g that were increased up to 85.52 ±0.78, 81.70 ± 0.30, and 78.05 ±0.15 g in T₁, T₂ and T₀, respectively (Figure 3). Similarly, Bethe et al., (2017) recorded the initial mean length 9.06±1.22 cm and mean weight 17.27±6.50 g thereby after 90 days of rearing they obtained final mean length and weight of tilapia 20.83 ±3.04 cm and 187.65 ±81.93 g respectively in aquaponics system. However, Fish culture in small tank with high stocking density might be the possible reason for lower final mean length and weight in aquaponics system. The highest mean length gain of tilapia was 5.11±1.64 cm in T₁ followed by 4.76±0.81 and 3.64±1.97 cm in T₂ and T₀, respectively. Whereas, the highest mean weight gain of tilapia was 30.16±0.11 g in T₁ followed by 26.51±0.17 and 23.47±0.20 g in T₂, and T₀, respectively (Table 3). Bethe et al. (2017) in their 90 days of trial recorded the highest mean length gain of aquaponically grown tilapia was 11.72± 2.49 cm and weight gain of 170.38±78.7 g which is higher than the present study. Zahan et al. (2018) found the mean weight gain of tilapia 295.03±12.04 g after 109 days of rearing which is also higher than the present study. This might be due to short culture period, high stocking density and inferior fish seed quality in the study.

The specific growth rate (SGR) of tilapia in T₁, T₂ and T₀ were 0.39 ± 0.01, 0.37 ± 0.01 and 0.34 ± 0.01 %/day, respectively (Table 3). In contrast, Roy et al. (2013) found that the SGR for Tilapia were 0.82, 0.79, and 0.42 %/day in three ponds which are higher than the present study. The lower SGR values in the current study might be attributed to the high stocking densities in small container, short rearing period and poor quality fish fry. The feed conversion ratios (FCR) were 3.52, 2.87 and 3.68 found in T₁, T₂ and T₀ respectively (Table 3). Rakocy et al. (1992) and Bethe et al. (2017) obtained FCR 2.05 and 2.33 of tilapia feeding with 30% protein containing diet. The quality of feed and associated feed loss might be responsible for comparatively higher FCR values in the present study.

At the end of the experiment total fish production was 26.27, 20.08 and 19.55 ton/ha/60 days in T₁, T₂ and T₀, respectively (Table 3). The present findings are lower, might be due to the aforementioned limitations of the study, than the findings of Afrin (2018) who recorded the total production of tilapia 183.1, 247.09 and 189.60 ton/ha/72 days respectively in aquaponics system. Bethe et al. (2017) also obtained 134.30 ton/ha/180 days production of tilapia from aquaponics.

However, the differences in fish growth parameters among all the three treatments were non-significant. Perhaps, it happens due to similar plant growbed quality, equal water recirculation efficacy of all aquaponics units, same species of plants and fish in all the treatments, similar fish rearing strategies, prohibition of epsom leakage into fish tanks etc. Notably, fish mortality rate was zero in all the treatments and fish growth was satisfactory in terms of length and weight, hence signifying the feasibility of aquaponics system in fish rearing.
3.5. Water quality parameters

pH indicates the acidity or alkalinity of water. Water becomes acidic at a pH less than 7 and alkaline at a pH more than 7. Fish may die, if the pH of water reaches to 11 or more and drops to 4 or below. At the beginning of the experiment, all the fish tanks were supplied with clean tap water having pH 7.21 (±0.01). However, over the period pH of fish tank water varied slightly in all the treatments and finally the values were 6.49 (± 0.03), 6.48 (± 0.01) and 6.53 (± 0.05) in T1, T2 and T3 respectively. The range of pH from 6.5 to 9 is the desirable range for tilapia culture. Sallenave (2016) reported that at pH 7.0 or below, most ammonia (>96%) remain in non-toxic form. All these references verify the suitable pH ranges of the fish holding tank in the experiment.

EC (electric conductivity) is a measure of total nutrients contained in water. Initially the EC value of fish tank water was 747.33 (±0.01) that increased over the period in all the treatments. On the final sampling, the highest EC value was 1521.0 (±59.40) µs/cm found in T1 and the lowest value of EC 1302.33 (±187.79) µs/cm was found in T0. Bablee et al., 2019 showed the EC ranged between 747-1388.5±26.27 µs /cm in aquaponics system. The observed EC implies usual range for aquaponics system.

The mean values of carbonate were 4.0 (±1.14) and 32.67 (±2.67) mg/l observed in T2 and T0 respectively at the end of the experiment. Whereas carbonate was absent in T1, Boyd (1990) mentioned that acceptable carbonate range in pond aquaculture is 0-20 mg/l. So it could be concluded that CO2 range was within the suitable range for fish culture in the experiment.

Initially, the hydrogen carbonate value started from 286.73 (±0.01) mg/l in all the treatments and decreased with the passing of time. This decrease was probably due to the accelerating plants and fish growth in the system irrespective to treatments. However, during the final sampling the highest hydrogen carbonate value was 167.00 (±9.24) mg/l found in T0 and the lowest value was 126.50 (±13.17) mg/l in T1. Therefore, beetroot plants in T1 were more active in ionic utilization than other treatments. Boyd (1990) mentioned that acceptable hydrogen carbonate range in pond aquaculture is 50-300 (mg/l). So, it can be concluded that hydrogen carbonates in the experimental fish tanks were within acceptable range over the study period.

During final sampling of fish tank water, the highest total N value 8.86 (±2.91) mg/l was found in T0 and the lowest in T1 counted as 6.5 (±0.80) mg/l, whereas, the highest value of total-N was 16.35±0.64 mg/l estimated by Salam et al. (2014) in aquaponics system. The amount of total-N in individual tank water was lower than the above findings. This was because the absorption rate of nitrogen by beetroot plants from fish waste water in the form of nitrates was enhanced with magnesium and sulfur supplementation through epsom salt (Nell et al., 2006). However the observed values of total N in fish tanks comply the suitable range for fish culture (Boyd, 1990).

However, other nutrients such as phosphorus (P), sulfur (S), sodium (Na) and potassium (K) contents in fish tanks’ water were within acceptable ranges for aquaculture (Boyd, 1990).

Water temperature is a very crucial parameter for maintaining normal physiological process in fish body and could strongly affect aquaculture production (León et al., 2006). Throughout the study, water temperature varied among different treatments likewise 22.9 to 27.4, 24.2 to 28.2 and 25.4 to 28.5°C in T1, T2 and T3 respectively (Figure 5). According to Losordo et al. (1998) a wide range of 25 to 32°C temperature gave acceptable growth rates for tilapia and nitrification can also be accomplished between 15 to 25°C while undisturbed to 30°C. Therefore, fish in tanks and nitrifying bacteria residing the root system of beetroot plants were quite comfortable for performing their activities (Tyson et al., 2004).

Dissolved oxygen (DO) concentration of culture water can lead to stressful condition for fish if not present adequately. DO of 5 ppm in water is the best condition for fish growth and it is hampered if kept below 5 ppm for long time (Boyd, 1990). With the use of aerator in fish tanks, the dissolved oxygen (DO) concentration was remained well above 5 ppm in all the treatments whereas the highest mean DO of 8.2 ppm was recorded in T2. So, the DO contents of tank water were within the acceptable limit for fish (Figure 5).

The toxic elements ( mostly ammonia and nitrite) in fish tank water, produced from fish excreta and feed wastes, were subjected to nitrification by autotrophic nitrifying bacteria (primarily Nitrosomonas and Nitrobacter) harboring the plant root system in aquaponics while passing the tank water through aquaponics growbeds (Tyson et al., 2004; Haug and McCarty, 1972). The final result is the transformation of toxic ammonia and nitrates into plant usable nitrates. This process keeps the tank water livable for fish and supply nutrients to the plants grown in soilless media (aquaponics). The maximum ammonia was recorded in T1 followed by T2 and T0 on 16 May, 2018 (Figure 4). The maximum nitrite and nitrate were found in T1 0.73 ± 0.15 and 39.56 ppm respectively. The acceptable range of ammonia, nitrite and nitrate in aquaponics system are <1 ppm, <1 ppm, 5–150 ppm (Sallenave, 2016). So, it can be concluded that the ammonia, nitrite and nitrate contents of tank water were tolerable for fish.
Table 1. Growth parameters of beetroot plants after final harvesting.

| Treatments | Length of roots (cm) | Wt. roots (g) | Wt. of leaves (g) | Wt. of beetroot (g) | Length of plant (cm) | Weight of plant (g) | Production (tons/ha/90days) |
|------------|----------------------|---------------|------------------|--------------------|----------------------|---------------------|-----------------------------|
| T1         | 31.03 (±0.43)        | 21.17         | 41.91            | 95.28              | 64.81                | 160.11              | 8.67                        |
|            | (±1.92)              | (±2.41)       | (±6.40)          | (±6.45)            | (±1.51)              | (±1.43)             |                             |
| T2         | 26.05 (±1.58)        | 18.73         | 39.43            | 80.58              | 59.17                | 136.58              | 7.48                        |
|            | (±1.91)              | (±2.25)       | (±3.09)          | (±4.75)            | (±4.68)              | (±2.71)             |                             |
| T0         | 19.48 (±1.63c)       | 14.24         | 31.08            | 73.87              | 43.68                | 129.06              | 6.61                        |
|            | (±1.01c)             | (±3.11b)      | (±2.71b)         | (±1.71b)           | (±3.05b)             | (±1.82b)            |                             |
| P-Value    | 0.001393             | 0.0008763     | 0.02396          | 0.01271            | 0.00678              | 0.00103             | .012                        |
| Level of Significant | ** ** *** *** ** ** ** ** ** | | | | | | |
| CV (%)     | 5.475                | 4.16          | 3.934            | 5.74               | 4.18                 | 2.546               | 1.65                        |

In a column, figures with same letters or without letter do not differ significantly whereas figures with dissimilar letter differ significantly as per Duncan’s New Multiple Range Test (DMRT).

* = Significant at 5% level of probability
** = Significant at 1% level of probability
*** = Significant at 0.1% level of probability

Table 2. Proximate composition of beetroot.

| Treatment | Protein (%) | Lipid (%) | Fiber (%) | Ash (%) | Moisture (%) | Carbohydrate (%) |
|-----------|-------------|-----------|-----------|---------|--------------|------------------|
| T1        | 0.94 (±0.015) | 1.25 (±0.015) | 6.2 (±0.1) | 0.92 (±0.015) | 89.7 (±0.59) | 1.67 (±0.84) |
|           | (±0.015)    | (±0.015)   | (±0.1)    | (±0.1)  | (±0.015)    | (±0.84)          |
| T2        | 0.97 (±0.013) | 1.23 (±0.015) | 6.2 (±0.1) | 0.94 (±0.01) | 89.46 (±0.58) | 0.95 (±0.67) |
|           | (±0.013)    | (±0.015)   | (±0.1)    | (±0.1)  | (±0.015)    | (±0.67)          |
| T0        | 0.91 (±0.01) | 1.1 (±0.01) | 5.93 (±0.015) | 0.93 (±0.029) | 89.00 (±0.89) | 2.1 (±0.72) |
|          | (±0.01)     | (±0.01)    | (±0.015)  | (±0.029) | (±0.89)     | (±0.72)          |
| P value   | 0.0017       | 0.028       | 0.011      | 0.29     | .00025       | .000013           |
| Level of significance | ** ** *** *** ** ** ** ** ** | | | | | |
| CV (%)    | 0.85         | 5.06        | 1.04       | 12.57    | 0.06         | 3.49              |

In a column, figures with same letters or without letter do not differ significantly whereas figures with dissimilar letter differ significantly as per Duncan’s New Multiple Range Test (DMRT).

* = Significant at 5% level of probability
** = Significant at 1% level of probability
*** = Significant at 0.1% level of probability
NS = non significant

Table 3. Growth performances of tilapia in aquaponics system of the experiment.

| Treatments | Mean length gain (cm) | Mean weight gain (g) | Percent length gain (%) | Percent weight gain (%) | Survival (%) | Specific growth rate (% per day) | FCR | Production (ton/ha/60 days) |
|------------|----------------------|----------------------|-------------------------|-------------------------|--------------|----------------------------------|-----|--------------------------|
| T1         | 5.11 (±1.64)         | 30.16 (±0.11)        | 36.62 (±5.00)           | 53.15 (±0.27)           | 76.66        | 0.39 (±0.01)                     | 3.52 | 26.27                    |
|            | (±1.64)              | (±0.11)              | (±5.00)                 | (±0.27)                 | (±0.19)      | (±0.01)                         | (±0.1) | (±2.47)               |
| T2         | 4.76 (±0.81)         | 26.51 (±0.17)        | 31.33 (±7.31)           | 46.11 (±4.29)           | 73.33        | 0.37 (±0.01)                     | 2.98 | 20.08                    |
|            | (±0.81)              | (±0.17)              | (±7.31)                 | (±0.29)                 | (±0.31)      | (±0.01)                         | (±0.1) | (±1.87)               |
| T0         | 3.90 (±1.97)         | 23.47 (±4.53)        | 28.91 (±0.21)           | 43.86 (±0.51)           | 66.66        | 0.34 (±0.01)                     | 3.68 | 19.55                    |
|            | (±1.97)              | (±4.53)              | (±0.21)                 | (±0.51)                 | (±0.01)      | (±0.01)                         | (±0.01) | (±1.56)               |
| P-Value    | 0.57                 | 0.17                 | 0.32                    | 0.33                    | 0.28         | 0.12                             | 0.63 | 0.65                     |
| Level of Significant | NS | NS | NS | NS | NS | NS | NS | NS |
| CV (%)     | 8.80                 | 6.60                 | 4.71                    | 9.14                    | 6.22         | 4.78                             | 0.78 | 5.94                    |

In a column, figures with same letters or without letter do not differ significantly whereas figures with dissimilar letter differ significantly as per Duncan’s New Multiple Range Test (DMRT).

NS = non significant
Figure 1. Experimental design.
Figure 2. Production of beetroot (ton ±SD) over the study period. Vertical bar of each treatment represents standard deviation.

Figure 3. Weight of tilapia in different treatments on different sampling dates.

Figure 4. Ammonia and nitrite contents of fish tank water during the study period. Vertical bar of each treatment represents standard deviation (SD).
Figure 5. Water temperature and dissolved oxygen (DO) contents of fish tank water. Vertical bar of each treatment represents standard deviation (SD).

4. Conclusions
Magnesium and sulfur are the two very crucial minerals for plant growth. Although they are quite available in the agricultural lands, aquaponically grown vegetables might require to be supplemented with these nutrients as they lack soil contact. This experiment has used epsom salt as the primary supplier of magnesium and sulfur to beetroot in aquaponics system with a view to summarizing its effect on plants’ growth without hampering the fish well being. Growth and production of beetroot was better in T₁ (epsom salt as foliar spray) than T₂ (epsom salt as root application) and T₀ (control), which might favor to conclude that addition of epsom salt supplied maximum minerals and nutrients to the plants. In the present experiment, beetroot was appeared in epsom salt based aquaponics system 15 days earlier than T₀. Therefore, it may be stated that epsom salt has a significant role in decreasing the harvesting period and increasing production of beetroot in aquaponics. However, in depth research is needed to verify the effects and dose response to epsom salt in aquaponics system supplemented as plant growth promoter.

Conflict of interest
None to declare.

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