Probiotic viability, viscosity, hardness properties and sensorial quality of synbiotic ice creams produced from goat’s milk

Merve ACU¹, Ozer KINIK², Oktay YERLIKAYA*³

Abstract
This research aimed to examine the probiotic viability of bacteria, rheological and sensorial properties in synbiotic ice creams produced from goat’s milk combined with probiotic culture and prebiotics. Tagatose, Litesse ultra and polydextrose (as prebiotics) were used in ice cream production and mixtures and these mixtures were inoculated with Bifidobacterium bifidum, Lactobacillus paracasei and Bifidobacterium longum combined culture. Frozen raspberry fruits, commercial raspberry and blackberry fruit purees were used as taste flavor enhancers in synbiotic ice cream. Four different ice cream types were produced including control sample. Probiotic bacteria viability, apparent viscosity, hardness and sensory properties were examined during the 120-day storage. It was determined that frozen fruit and fruit purees addition and using prebiotics significantly affected the Lactobacillus paracasei and Bifidobacterium spp, viability and color, appearance, flavor, taste and overall sensory scores of the synbiotic goat’s milk ice creams (P < 0.05). Synbiotic ice cream samples maintained their probiotic properties during storage and were generally well appreciated in terms of sensory properties.

Keywords: goat’s milk; ice cream; prebiotics; probiotic viability; synbiotics.

Practical Application: In this study, ice cream production with functional properties has been developed by using goat’s milk. Within the scope of the study, probiotic cultures that were used less in probiotic products were tried and ice creams were enriched with fruit and fruit purees. As a result of the research, probiotic viability remained at the desired level and a dairy product with the possibility of commercialization was produced.

1 Introduction

Probiotics is defined as “[...] live microorganisms that, when administered in adequate amounts, confer a health benefit on the host [...]” (Hill et al., 2014). Today, numerous dairy products with probiotic additives are produced at the industrial level (Markowiak & Śliżewska, 2017). Lactobacillus acidophilus, Lactobacillus paracasei and Bifidobacterium species isolated from human and animal intestinal tract are the most common bacteria used as probiotics. The use of probiotic preparations including these bacteria in industrial food production systems have become increasingly widespread in accordance with consumer demands (Kechagia et al., 2013).

Dairy products are the most probiotic food carrier (Champagne et al., 2018) with several benefits to consumer health (Sarfraz et al., 2019; Shafi et al., 2019; Vasconcelos et al., 2019) and technological parameters (Guimarães et al., 2020). Non bovine dairy foods present relevance as probiotic carrier (Ranadheera et al., 2018) and in particular ice cream is a probiotic dairy food with several examples (Ayar et al., 2018; Balthazar et al., 2018; Kalicka et al., 2019).

Prebiotics are nutrients that can directly pass to the large intestine without being digested in the small intestine and increase the activity of probiotics, thus, increasing the beneficial effects of probiotics in the bowel system (O’Bryan et al., 2013). Products created by the combined use of probiotics and prebiotics are called synbiotics. With synbiotic application, the life span of probiotic bacteria is prolonged and these bacteria can better colonize in the colon. In vitro studies show that synbiotic administration is more advantageous than the sole uses of prebiotics or probiotics (Pandey et al., 2015; Krumbeck et al., 2016).

Today, interest in probiotics, prebiotics and functional foods is constantly increasing. These subjects have become increasingly more popular in the academic studies and scientific studies on the benefits of probiotics to human health are added to the literature every day (Markowiak & Śliżewska, 2017). Although yogurt and fermented dairy products are the main carriers of probiotic bacteria, products such as milk-based desserts, baby formulas, ice cream, butter, cheese types, capsules and powders dissolving in cold beverages and plant-origin fermented foods have become available in the international market in recent years (Heller, 2001; Shibly & Mishra, 2013).

Ice cream, which is used as a probiotic carrier, is a dairy product obtained by processing milk and dairy products, sweeteners, stabilizers, emulsifiers, color and aroma substances by churning to have air spaces in the mixture (Marshall & Arbuckle, 1996). There are many studies on synbiotic ice creams and their properties, where probiotics and prebiotics are used separately or in combination. Ice cream could present interesting properties, such as better melting features and desirable texture, if prepared...
with goat milk. When use of goat’s milk in ice cream production, the color of ice cream becomes whiter. It also becomes rich in terms of total amount of dry matter (Homayouni et al., 2008, 2012; Ranadheera et al., 2019). Goat milk is also important for human health since it has high digestibility, low allergen effect properties, and contains functional compounds and proteins. In addition, the ice cream produced using goat milk has been shown to have better quality criteria and sensory properties (Paz et al., 2014; Zenebe et al., 2014; Lad et al., 2017). The aim of this research was to determine the viability of probiotic cultures, viscosity, hardness properties and sensorial quality of synbiotic goat’s ice cream having improved functional properties.

2 Materials and methods

2.1 Materials

The raw goat’s milk and skimmed milk powder were obtained from local dairy plants located in Bornova-Izmir while powdered sahlep (produced from the wild orchid plant) were provided from a local food market. Tagatose, Litesse® Ultra™ and polydextrose were provided from commercial manufacturers. Frozen raspberry fruit (Superfresh, Turkey), raspberry and blackberry purees (Aromsa, Turkey) were melted at room temperature and made ready for use. Probiotic lactic cultures (Lactobacillus paracasei subsp. paracasei Lafti L-26, Bifidobacterium longum + Bifidobacterium bifidum Lafti B-94) were obtained from DSM Food Specialties USA, Inc.

2.2 Methods

Functional ice cream production

Two replicate productions were carried out in the study with two duplicates. Sample codes and ice cream production stages of synbiotic ice cream samples are given in Figure 1.

The mixture consisted of skimmilk powder, tagatose (5%), Litesse ultra (0.67%), polydextrose (1%) and sahlep. These ingredients were added to the raw milk and a subsequent pasteurization procedure was carried out at 90-95 °C for 5-10 minutes. Then the mixtures were cooled to 37-40 °C and probiotic starter culture was inoculated in accordance with the commercial company’s recommendations (10⁸-10⁹ Log cfu/g). The inoculated mixtures were left to incubate at 37 °C. The incubation was carried out until the pH values reached 4.8-4.9 and ice cream samples stored at +4 °C for 24 hours for ageing. The mixtures were divided into four groups and three of the groups were fortified with 10% frozen fruit or fruit purees. No fruit puree was added to the control group. Freezing procedure was carried out in a batch type ice cream machine and then the samples were packaged. The samples were analyzed for probiotic viability, viscosity, hardness values and sensory properties on the 1st, 15th, 30th, 60th, 90th, and 120th days of the storage. All of the samples were produced in the pilot dairy plant located in Ege University, Izmir. For the production of raspberry puree, the raspberries were thawed, and then grounded and stirred on the stove.

Probiotic bacteria counts in synbiotic goat’s ice creams

MRS-Vancomycin media for L. paracasei subsp. paracasei; MRS-NNLP (nalidixic acid, neomycin sulfate, lithium chloride, paromomycin sulfate) media for Bifidobacterium spp. was used. Petri plates were incubated at 37 °C for 72 h using Anaerocult® A (Merck, Darmstadt, Germany) in anaerobic jars (Merck, Darmstadt, Germany). 0.03 g nalidixic acid; 0.2 g neomycin sulfate, 6 g lithium chloride and 0.2 g paromomycin sulfate were added to the 1000 mL MRS Agar media by using 0.22 µm diameter microfilter. 0.05% L-cysteine was added to medium. At the end of the incubation, dark white, high centered colonies with diameter of 1-1.5 mm were evaluated (Dave & Shah 1996; Tharmaraj & Shah, 2003; Donkor et al., 2006).

Figure 1. Flow diagram of synbiotic ice cream with fruit puree.
Viscosity and textural analyses in synbiotic goat’s ice creams

Viscosity analysis were performed by using Brookfield viscometer DV-II model at 60 rpm for mixtures, and at 120 rpm for ice cream samples at 8-10 °C using 63=LV3 spindle. The measurements were saved and the results were obtained using Rheocalc application (Brookfield Engineering Laboratories Inc., ABD) software.

Textural properties (hardness) of synbiotic goat’s ice cream samples were determined with Brookfield CT-3 model texture analyzer. Texture readings were conducted between -6 °C to -2 °C and texture application software (Brookfield Engineering Laboratories Inc., ABD) was utilized in the calculation of the results. TA 15/1000 conical probe was used for the analysis. Test speed was 2 mm/s, distance was 15 mm, trigger load was 6.8 g, the length was 40 mm and the diameter were 60 mm.

Sensory analysis

Scoring test (Uysal et al., 2004) was utilized for the sensory evaluation of plain and fruit puree flavored synbiotic ice cream samples. The evaluation of sensory properties was conducted according to the sensory evaluation principles set forth in TS 4265 and each sample was evaluated accordingly. Sensory analyses of the synbiotic ice cream samples were carried out by trained and semi-trained panelists from Ege University, Faculty of Agriculture, Department of Dairy Technology. Accordingly, for scoring, evaluation card by Bodyfelt et al. (1988) was used with modifications. Ice cream samples were evaluated with the highest 5 points [very good (5), good (4) less defective (3) defective (2)] in terms of color-appearance, structure-consistency, odor and taste criteria. Sensory analyses were conducted on 1ª, 15ª, 30ª, 60ª, 90ª and 120ª days of the storage for each sample.

Statistical analyses

SPSS software package v15.00 (SPSS Inc. Chicago, Illinois) was used for the statistical analyses in the study. The significant differences were analyzed according to the Duncan multiple comparison test (P < 0.05). This analysis was made to assess the effects of prebiotics and different types of fruit purees on probiotic viability, rheology and sensorial properties of the samples.

3 Results and discussion

3.1 Probiotic viability in synbiotic goat’s ice creams

It is undeniable that the viability of probiotic cultures during the shelf life decreased in the low storage temperature conditions and high acidity increases. Fluctuations in storage temperatures in ice cream technology can cause crystallization and therefore viability may decrease by cell fractionation (Ranadheera et al., 2012). In Table 1, 2, provides the viable count of probiotic cultures in synbiotic ice cream samples during the storage. It was found that B sample on day 1 of the storage period had the highest L. paracasei subsp. paracasei count (9.32 log cfu/g) and A sample day 1 of the storage period had the lowest count (7.70 log cfu/g). Furthermore, it was shown that B sample on day 1 of the storage period had the highest Bifidobacterium spp. count (9.15 log cfu/g), and A sample on day 1 of the storage period had the lowest count (7.69 log cfu/g).

It was detected that fruit purees and prebiotics had significant effect on L. paracasei subsp. paracasei and Bifidobacterium spp. counts (P < 0.05). While Bifidobacterium spp. count did not change significantly (P > 0.05); it was seen that L. paracasei subsp. paracasei count changed only 1ª day of storage significantly. Tokuc et al. (2008) stated that Lactobacillus spp. remained stable as 7 log during 6 months storage period in probiotic ice cream production. Ranadheera et al. (2012) indicated that levels of Lactobacillus acidophilus La-5 and B. animalis subsp. lactis BB-12 were protected 56.14% and 66.46%, respectively in probiotic ice cream samples produced with goat milk. In compatible with the researchers’ findings during 120 days of storage period probiotic bacteria count of synbiotic ice cream samples did not fall below 7.0; therefore, it could be said that ice cream samples maintained its probiotic characteristics. Probiotic foods are

Table 1. L. paracasei subsp. paracasei counts of synbiotic ice creams produced from goat’s milk (log cfu/g).

| Samples | 1ª day     | 15ª day    | 30ª day    | 60ª day    | 90ª day    | 120ª day   |
|---------|------------|------------|------------|------------|------------|------------|
| C       | 8.13 ± 0.16 | 8.12 ± 0.05 | 8.02 ± 0.18 | 8.63 ± 0.01 | 8.44 ± 0.08 | 8.19 ± 0.09 |
| F       | 8.37 ± 0.39 | 8.02 ± 0.10 | 7.88 ± 0.19 | 8.47 ± 0.10 | 8.31 ± 0.07 | 8.03 ± 0.14 |
| A       | 7.70 ± 0.12 | 8.16 ± 0.14 | 8.06 ± 0.38 | 8.38 ± 0.01 | 8.23 ± 0.14 | 7.99 ± 0.16 |
| B       | 9.32 ± 0.02 | 8.34 ± 0.13 | 8.06 ± 0.20 | 8.49 ± 0.02 | 8.43 ± 0.02 | 8.17 ± 0.14 |

*; Values with the different letters in the same column differ significantly (P < 0.05); **; Values with the different letters in the same row differ significantly (P < 0.05); C: Sample produced from freezing the fermented mixture no adding fruit puree, F: Sample produced from addition of frozen raspberries to the fermented mixture, A: Sample produced from addition of commercial raspberry puree to the fermented mixture, B: Sample produced from addition of commercial blackberry puree to the fermented mixture.

Table 2. Bifidobacterium spp. counts of synbiotic ice creams produced from goat’s milk (log cfu/g).

| Samples | 1ª day     | 15ª day    | 30ª day    | 60ª day    | 90ª day    | 120ª day   |
|---------|------------|------------|------------|------------|------------|------------|
| C       | 8.03 ± 0.41 | 8.13 ± 0.02 | 7.91 ± 0.23 | 8.55 ± 0.05 | 8.41 ± 0.14 | 8.15 ± 0.02 |
| F       | 8.53 ± 0.31 | 8.04 ± 0.10 | 7.91 ± 0.15 | 8.34 ± 0.23 | 8.34 ± 0.05 | 8.07 ± 0.04 |
| A       | 7.69 ± 0.22 | 7.93 ± 0.16 | 7.75 ± 0.60 | 8.27 ± 0.00 | 8.31 ± 0.15 | 7.73 ± 0.24 |
| B       | 9.15 ± 0.07 | 8.19 ± 0.17 | 8.12 ± 0.11 | 8.41 ± 0.03 | 8.40 ± 0.04 | 8.07 ± 0.12 |

*; Values with the different letters in the same row differ significantly (P < 0.05); C: Sample produced from freezing the fermented mixture no adding fruit puree, F: Sample produced from addition of frozen raspberries to the fermented mixture, A: Sample produced from addition of commercial raspberry puree to the fermented mixture, B: Sample produced from addition of commercial blackberry puree to the fermented mixture.
required to carry a specific live microorganism at a level of at least 10^6-10^7 cfu/g during the storage period.

### 3.2 Viscosity and hardness properties in synbiotic goat’s ice creams

Viscosity and hardness values of synbiotic goat’s ice cream samples are presented in Table 3. As is seen in the results, viscosity values varied between 613.75-979.25 cP. The highest viscosity during storage period was determined in F sample at the 30th day (979.25 cP), and the lowest values was found in C sample at the 1st day (613.75 cP). The most challenging measurements were performed in raspberry puree ice creams. B sample could not be measured at 120 rpm at the 60th day. This may be due to the different viscosity of prebiotic supplement and different fruit purees. Generally speaking, viscosity values increased at the beginning of the storage and showed a decline at the further days. In general, it is normal for control sample to have lower values. Many ingredients adding the food composition can increase the viscosity values of the products. It could be said that addition of fruit purees and prebiotics increased the viscosity. Although fat content and viscosity are directly proportional, this relation was not as proportional in ice cream samples as is in the mixtures. However, this can be associated with the seed particles in the ice cream samples.

In all ice cream samples changes caused by storage and ingredients were significant (P < 0.05) whereas prebiotics and fruit puree used did not cause a significant difference between the samples only at the 30th day of the storage (P > 0.05). Guven & Karaca (2002), in their study on fruit ice cream containing sugar and strawberry at different ratios and vanilla yogurt ice creams, reported that viscosity increased parallel to the increase in sugar and fruit content. Muse & Hartel (2004), in their study on ice cream production by using different emulsifiers and sweeteners, found the viscosity values between 621-935 cP. Akin (2005), reported the viscosity of probiotic yogurt ice cream samples between 842-1312 cP. Kesenkas et al. (2013) found the lowest viscosity during storage in ice cream samples produced from cow’s milk while the highest viscosity was found in ice cream samples produced from the mixture prepared with the addition of kefir at 50% to the mixture produced with soy milk. Carbohydrate based ice cream mixtures containing fat substitutes which exhibiting a viscous behavior due to their water absorption capacity increase the viscosity of the system (Cottrell et al., 1979; Schmidt et al., 1993).

In the study, hardness values varied between 521.75-2127.25 g. The highest texture value during storage period was determined in C sample at the 90th day, while the lowest value was found in B sample at the 30th day. In a general evaluation texture values, there were irregular fluctuations. Texture readings were performed with multi parallels and calculations were done based on the closest values.

Changes only in C samples in all ice cream groups were significant (P < 0.05), while the effect of prebiotics and fruit purees were statistically significant only at the 90th day of the storage (P < 0.05). El-Nagar et al. (2002), in their study on the effect of inulin in yogurt ice creams, reported that inulin addition increased the hardness values of ice cream samples. Some researchers have also reported that low fat ice cream samples decreased the gumminess values during storage. Akalin et al. (2008) in their study on fat reduced and low fat ice creams, reported that fat ratio had a significant effect on the hardness values of ice cream samples (P < 0.05). Kesenkas et al. (2013) have reported that the texture values of the ice creams varied between 1237.7 and 4270.5. Lower results obtained in our study can be associated with the soft structure of ice cream samples.

### 3.3 Sensory properties in synbiotic goat’s ice creams

Compared to classic ice cream products, flavor profiles of probiotic ice cream can substantially vary. Inulin and oligofructose improve the sensory properties including smoother feeling in the mouth, prolonged flavor with lower aftertaste and mild

---

**Table 3. Viscosity and hardness values of synbiotic ice creams produced from goat’s milk.**

| Days | C                  | F                  | A                  | B                  |
|------|--------------------|--------------------|--------------------|--------------------|
|      | Viscosity (cP)     |                    |                    |                    |
| 1    | 613.75 ± 24.25<sup>ax</sup> | 857.50 ± 55.00<sup>bh</sup> | 777.75 ± 35.75<sup>bh</sup> | 725.50 ± 20.50<sup>ay</sup> |
| 15   | 690.75 ± 36.25<sup>ab</sup> | 969.75 ± 12.75<sup>c</sup> | 687.75 ± 30.25<sup>ah</sup> | 971.00 ± 13.00<sup>b</sup> |
| 30   | 907.25 ± 7.25<sup>c</sup> | 979.25 ± 1.75<sup>c</sup> | 973.50 ± 8.00<sup>c</sup> | 924.25 ± 50.25<sup>c</sup> |
| 60   | 812.25 ± 14.25<sup>c</sup> | 844.00 ± 8.50<sup>fs</sup> | 784.00 ± 2.50<sup>b</sup> | ND                 |
| 90   | 626.50 ± 6.50<sup>ab</sup> | 663.25 ± 26.25<sup>a</sup> | 775.50 ± 14.50<sup>c</sup> | 779.25 ± 22.25<sup>y</sup> |
| 120  | 654.25 ± 8.25<sup>ab</sup> | 635.25 ± 30.75<sup>a</sup> | 768.25 ± 22.25<sup>b</sup> | 928.25 ± 18.25<sup>x</sup> |
| Hardness (g) | 1510.00 ± 236.25<sup>ab</sup> | 1709.00 ± 560.70 | 667.50 ± 104.25 | 656.25 ± 95.75 |
| 15   | 766.25 ± 82.25<sup>a</sup> | 1022.62 ± 257.12 | 1123.12 ± 336.12 | 927.00 ± 357.75 |
| 30   | 1210.00 ± 229.50<sup>ab</sup> | 1156.75 ± 271.00 | 1192.12 ± 180.12 | 521.75 ± 111.50 |
| 60   | 1998.47 ± 450.87<sup>c</sup> | 1359.37 ± 238.62 | 1468.12 ± 282.87 | 246.37 ± 198.37 |
| 90   | 2127.25 ± 67.00<sup>c</sup> | 1065.75 ± 210.25<sup>h</sup> | 993.37 ± 337.12<sup>h</sup> | 765.87 ± 203.62<sup>c</sup> |
| 120  | 1019.25 ± 72.50<sup>a</sup> | 885.90 ± 96.25 | 1587.37 ± 345.87 | 871.50 ± 141.50 |

<sup>1</sup> Viscosity values could not be measured at 120 rpm; ND: not determined at 120 rpm; <sup>a,b,c</sup>; Values with the different letters in the same row differ significantly (P < 0.05); <sup>A,B,C</sup>; Values with the different letters in the same column differ significantly (P < 0.05); C: Sample produced from freezing the fermented mixture no adding fruit puree; F: Sample produced from addition of frozen raspberries to the fermented mixture, A: Sample produced from addition of commercial raspberry puree to the fermented mixture, B: Sample produced from addition of commercial blackberry puree to the fermented mixture.
sweetness. These properties were associated with high sensory scores in some studies (Di Criscio et al., 2010). Sensory properties of a product are the most important features that determine the appreciation of consumers. In this manner, the sensory evaluation of color and appearance, structure and consistency, smell and taste properties of ice cream samples by the panelists were given in Table 4. Data showed that the addition of berry like fruit purees and probiotics in ice cream had effect on the sensory properties of ice cream. The overall acceptability of samples in storage period were in the range of 3.81-4.87. Texture and taste evaluations yielded good results and no off-flavor was reported. These results were consistent with those reported by Di Criscio et al. (2010) who observed a synbiotic effect of prebiotics that helps in the survival of probiotic bacteria. Include the need of performing additional sensory studies as projective methods (Pinto et al., 2018), innovative methods based in consumer perception (Torres et al., 2017) and sensory properties (Mituniewicz-Małek et al., 2019)

### 4 Conclusions

As a result, it was determined that enrichment with fruit puree had a significant effect on *Lactobacillus paracasei* subsp. *paracasei* and *Bifidobacterium* spp. viability and color, appearance, flavor, taste and overall sensory scores of the ice cream samples. It was determined that the samples maintained their probiotic properties during storage and generally received good sensory scores. The effects of reducing the fat ratio and using sweeteners, in addition to natural sugars, on glycemic index can be investigated in the future studies.

### Acknowledgements

The authors thank to Scientific Research Fund Council of Ege University (2012-ZRF-004) for the financial support.

### References

Akalin, A. S., Karagozlu, C., & Unal, G. (2008). Rheological properties of reduced-fat and low-fat ice cream containing whey protein isolate and inulin. *European Journal of Food Research and Technology*, 227(3), 889-895. http://dx.doi.org/10.1007/s00217-007-0800-z.

Akin, M. S. (2005). Effects of inulin and different sugar levels on viability of probiotic bacteria and the physical and sensory characteristics of probiotic fermented ice cream. *Milchwissenschaft. Milk Science International*, 60(3), 297-300.

Ayar, A., Siçramaz, H., Öztürk, S., & Öztürk Yilmaz, S. (2018). Probiotics properties of ice cream products with dietary fibres from by-products of the industry. *International Journal of Dairy Technology*, 71(1), 174-182. http://dx.doi.org/10.1111/1471-0307.12387.

Balthazar, C. F., Silva, H. L. A., Esmerino, E. A., Rocha, R. S., Moraes, J., Carmo, M. A. V., Azevedo, L., Camps, I., K.D. Abud, Y., SantAnna,
C., Franco, R. M., Freitas, M. Q., Silva, M. C., Raices, R. S. L., Escher, G. B., Granato, D., Senaka Ranadheera, C., Nazarro, F., & Cruz, A. G. (2018). The addition of inulin and Lactobacillus casei 01 in sheep milk ice cream. *Food Chemistry*, 246, 464-472. http://dx.doi.org/10.1016/j.foodchem.2017.12.002. PMID:29291874.

Bodyfelt, F. W., Tobias, J., & Trout, G. M. (1988). *The sensory evaluation of dairy products* (398 p.). Westport: AVI Publ.

Champagne, C. P., Gomes da Cruz, A., & Daga, M. (2018). Strategies to improve the functionality of probiotics in supplements and foods. *Current Opinion in Food Science*, 22, 160-166. http://dx.doi.org/10.1016/j.cofs.2018.04.008.

Cottrell, J. I. L., Pass, G., & Phillips, G. O. (1979). Assessement of polysaccharides as ice cream stabilizers. *Journal of Food Science and Agriculture*, 30(11), 1085-1089. http://dx.doi.org/10.1002/jsfa.274031111.

Dave, R. I., & Shah, N. P. (1996). Evaluation of media for selective enumeration of Streptococcus thermophilus, Lactobacillus delbrueckii ssp. bulgaricus, *Lactobacillus acidophilus*, and *Bifidobacteria*. *Journal of Dairy Science*, 79(9), 1529-1536. http://dx.doi.org/10.3168/jds.S0022-0302(96)76513-X. PMID:8899517.

Di Criscio, T., Fratianni, A., Mignogna, R., Cinquanta, L., Coppola, R., Sorrentino, E., & Panfilì, G. (2010). Production of functional probiotic, prebiotic, and sybiotic ice creams. *Journal of Dairy Science*, 93(10), 4555-4564. http://dx.doi.org/10.3168/jds.2010-3355. PMID:20854989.

Donkor, D. N., Henriksson, A., Vasiljevic, T., & Shah, N. P. (2017). Impact of probiotics on human health – a review. *International Journal of Microbiology and Applied Science*, 6(5), 1781-1792. https://doi.org/10.20546/ijmas.2017.605.194.

Esmerino, E. A., Silva, M. C., Sant'Ana, A. S., Duarte, M. C. K., & Baptista, C. F. (2014). Biological quality and stability of yoghurt with added inulin. *Current Opinion in Food Science*, 87(1), 1-10. http://dx.doi.org/10.1016/j.cofo.2019.12.002.

Gyftopoulou, K., Skarmoutsou, N., & Fakiri, E. M. (2013). Health benefit of probiotics – a review. *ISRN Nutrition*, 2013, 481651. http://dx.doi.org/10.5402/2013/481651. PMID:24959545.

Hutkins, R. W. (2016). Prebiotics and synbioticsI: dietary strategies for improving gut health. *Current Opinion in Gastroenterology*, 32(2), 110-119. http://dx.doi.org/10.1097/MOG.0000000000000249. PMID:26825589.

Lad, S. S., Aparnathi, K. D., Mehta, B., & Velpula, S. (2017). Goat milk in human nutrition and health – a review. *International Journal of Microbiology and Applied Science*, 6(5), 1781-1792. https://doi.org/10.20546/ijmas.2017.605.194.

Markowiak, P., & Sliżewska, K. (2017). Effects of probiotics, prebiotics, and synbiotics on human health. *Nutrients*, 9(9), 1021. http://dx.doi.org/10.3390/nu9091021. PMID:28914794.

Marshall, R. T., & Arbuckle, W. S. (1996). *Ice cream* (5th ed.). New York: Chapman and Hall. http://dx.doi.org/10.1007/978-1-4613-0477-7.

Munirwicz-Malek, A., Ziebińska, D., & Ziarno, M. (2019). Probiotic monocultures in fermented goat milk beverages – sensory quality of final product. *International Journal of Dairy Technology*, 72(2), 240-247. http://dx.doi.org/10.1111/1471-0307.12576.

Muse, M. R., & Hartel, R. W. (2004). Ice cream structural elements that affect melting rate and hardness. *Journal of Dairy Science*, 87(1), 1-10. http://dx.doi.org/10.3168/jds.S0022-0302(04)73135-5. PMID:14765804.

O’Bryan, C. A., Pak, D., Crandall, P. G., Lee, S. O., & Ricke, S. C. (2013). The Role of prebiotics and probiotics in human health. *Journal of Probiotics and Health*, 1(2), 1000108. http://dx.doi.org/10.4172/2329-8901.1000108.

Pandey, K. R., Naik, S. R., & Vakil, B. V. (2015). Probiotics, prebiotics and synbiotics – a review. *Journal of Food Science and Technology*, 52(12), 7577-7587. http://dx.doi.org/10.1007/s13197-015-1921-1. PMID:26604335.

Paz, N. F., Oliveira, E. G., Kairuz, M. S. N., & Ramón, A. N. (2014). Characterization of goat milk and potentially synbiotic non-fat yogurt. *Food Science and Technology (Campinas)*, 34(3), 629-635. http://dx.doi.org/10.1590/1678-457x.2014.66. PMID:24912386.
Ranadheera, C. S., Evans, C. A., Baines, S. K., Balthazar, C. E., Cruz, A. G., Esmerino, E. A., Freitas, M. Q., Pimentel, T. C., Wittwer, A. E., Naumovski, N., Graça, J. S., Sant’Ana, A. S., Ajlouni, S., & Vasiljevic, T. (2019). Probiotics in goat milk products: delivery capacity and ability to improve sensory attributes. Comprehensive Reviews in Food Science and Food Safety, 18(4), 867-882. http://dx.doi.org/10.1111/1541-4337.12447.

Ranadheera, C. S., Naumovski, N., & Ajlouni, S. (2018). Non-bovine milk products as emerging probiotic carriers: Recent developments and innovations. Current Opinion in Food Science, 22, 109-114. http://dx.doi.org/10.1016/j.cofs.2018.02.010.

Sarfraz, F., Farooq, U., Shafi, A., Hayat, Z., Akram, K., & Rehman, H. U. (2019). Hypolipidaemic effects of synbiotic yoghurt in rabbits. International Journal of Dairy Technology, 72(4), 1-6. http://dx.doi.org/10.1111/1471-0307.12618.

Schmidt, K., Lundy, A., Reynolds, J., & Yee, L. N. (1993). Carbohydrate or protein based fat mimicker effect on ice milk properties. Journal of Food Science, 58(4), 761-779. http://dx.doi.org/10.1111/j.1365-2621.1993.tb09353.x.

Shafi, A., Naem Raja, H., Farooq, U., Akram, K., Hayat, Z., Naz, A., & Nadeem, H. R. (2019). Antimicrobial and antidiabetic potential of synbiotic fermented milk: a functional dairy product. International Journal of Dairy Technology, 72(1), 15-22. http://dx.doi.org/10.1111/1471-0307.12555.

Shiby, V. K., & Mishra, H. N. (2013). Fermented milks and milks products as functional foods-a review. Critical Reviews in Food Science and Nutrition, 53(5), 482-496. http://dx.doi.org/10.1080/10408398.2010.547398. PMid:23391015.

Tharmaraj, N., & Shah, N. P. (2003). Selective Enumeration of Lactobacillus delbrueckii ssp. bulgaricus, Streptococcus thermophilus, Lactobacillus acidophilus, Bifidobacteria, Lactobacillus casei, Lactobacillus rhamnosus and Propionibacteria. Journal of Dairy Science, 86(7), 2288-2296. http://dx.doi.org/10.3168/jds.S0022-0302(03)73821-1. PMid:12906045.

Tokuc, K., Demirci, M., Bilgin, B., & Arici, M. (2008, May 21-23). Baby origin Lactobacillus spp. produce probiotic ice cream using and identify some other properties with probiotic bacteria viability during storage. In Proceedings of the 10th Food Congress. Erzurum, Turkey: Food Technology Association.

Torres, F. R., Esmerino, E. A., Carr, B. T., Ferrão, L. L., Granato, D., Pimentel, T. C., Bolini, H. M. A., Freitas, M. Q., & Cruz, A. G. (2017). Rapid consumer-based sensory characterization of requijao cremoso, a spreadable processed cheese: performance of new statistical approaches to evaluate check-all-that-apply data. Journal of Dairy Science, 100(8), 6100-6110. http://dx.doi.org/10.3168/jds.2016-12516. PMid:28571992.

Uysal, H., Kinik, O., & Kavas, G. (2004). Sensory testing techniques applied in milk and milk products (No. 560, 101 p.). Izmir: Faculty of Agriculture Publications, Ege University.

Vasconcelos, F. M., Silva, H. L. A., Poso, S. M., Barroso, M. V., Lanzetti, M., Rocha, R. S., Graça, J. S., Esmerino, E. A., Freitas, M. Q., Silva, M. C., Raices, R. S. L., Granato, D., Pimentel, T. C., Sant’Ana, A. S., Cruz, A. G., & Valença, S. S. (2019). Probiotic prato cheese attenuates cigarette smoke-induced injuries in mice. Food Research International, 123, 697-703. http://dx.doi.org/10.1016/j.foodres.2019.06.001. PMid:31285019.

Zenebe, T., Ahmed, N., Kabeta, T., & Kebede, G. (2014). Review on medicinal and nutritional values of goat milk. Academic Journal of Nutrition, 3(3), 30-39. http://dx.doi.org/10.5829/idosi.ajn.2014.3.3.93210.