Dynamic cerebral autoregulation in anterior and posterior cerebral circulation during cold pressor test

Takuro Washio¹,², Hironori Watanabe¹ and Shigehiko Ogoh¹* *

Abstract
We hypothesized that cerebral blood flow (CBF) regulation in the posterior circulation differs from that of the anterior circulation during a cold pressor test (CPT) and is accompanied by elevations in arterial blood pressure (ABP) and sympathetic nervous activity (SNA). To test this, dynamic cerebral autoregulation (dCA) in the middle and posterior cerebral arteries (MCA and PCA) were measured at three different conditions: control, early phase of the CPT, and the late phase of the CPT. The dCA was examined using a thigh cuff occlusion and release technique. The MCA and PCA blood velocities were unchanged at CPT compared with the control conditions despite an elevation in the ABP. The dCA in both the MCA and PCA remained unaltered at CPT. These findings suggest that CPT-induced elevations in the ABP and SNA did not cause changes in the CBF regulation in the posterior circulation compared with the anterior circulation.

Keywords: Posterior cerebral artery, Middle cerebral artery, High blood pressure, Sympathoactivation

Introduction
Interestingly, hypertensive disease-induced primary intracerebral hemorrhage occurs mainly at the small penetrating blood vessels in the posterior cerebral circulation rather than the anterior cerebral circulation [1]. Hypertension is a risk factor for cerebrovascular diseases [2, 3]; therefore, these findings suggest that the posterior cerebral vasculature may not be able to properly regulate an elevation in blood pressure compared with the anterior cerebral vasculature. Recent studies [4–8] have reported that the responses of the posterior cerebral blood flow (CBF) in several physiological conditions (e.g., orthostatic stress, hypoxia, dynamic resistance exercise, heat stress, etc.) are different from those of the anterior CBF. Indeed, it has been reported that the dynamic cerebral autoregulation (dCA) or cerebrovascular carbon dioxide (CO₂) reactivity is lower in the posterior cerebral circulation than that of anterior cerebral circulation [9, 10]. It is well established that these cerebral regulatory mechanisms keep the CBF relatively constant despite changes in cerebral perfusion pressure (CPP), which is critical in preventing cerebral ischemia or hemorrhage [11]. Although it can be expected that this heterogeneous CBF response may be due to different physiological roles, the mechanism of this heterogeneous CBF remains unknown.

On the other hand, some previous studies [12–14] reported that sympathetic blockade (prazosin or trimethaphan) impairs dCA, suggesting that autonomic neural control, i.e., sympathetic nerve activity (SNA), of the cerebral circulation likely plays an important role in the dynamic CBF regulation. The SNA also exerts a greater influence on cerebral circulation in hypertension than in the normotensive condition [15]. Thus, sympathoexcitation accompanied by hypertension may improve dCA and prevent over-perfusion. Importantly, there are anatomical differences between the anterior and posterior cerebral circulations, including regional

*Correspondence: ogoh@toyo.jp
1 Department of Biomedical Engineering, Toyo University, 2100 Kujirai, Kawagoe-shi, Saitama 350-8585, Japan
Full list of author information is available at the end of the article

© The Author(s) 2020. This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.
heterogeneity in the sympathetic innervation of intracranial arterioles [16]. The posterior cerebral circulation may have less sympathetic innervation than the anterior cerebral circulation [16]. Therefore, an effect of sympathoexcitation on dynamic CBF regulation may be less in the posterior cerebral circulation compared with anterior cerebral circulation. Hypertension-induced sympathoexcitation may emphasize the difference between anterior and posterior CBF regulation, and this phenomenon may be associated with hypertensive disease-induced primary intracerebral hemorrhage occurred in mainly at the posterior cerebral circulation. However, an effect of sympathoexcitation on posterior CBF regulation has not been identified.

Against this background, we hypothesized that the dCA or cerebral vasculature tone of the posterior CBF is lower than that of the anterior CBF during an elevation in the arterial blood pressure (ABP) accompanied with sympathoexcitation. To test this hypothesis, the CBF velocity as an index of CBF, cerebral vascular tone, and the dCA in the middle cerebral artery (MCA) and the posterior cerebral artery (PCA) were measured before and during a cold pressor test (CPT). The CPT is widely used as a physiological tool to evoke temporary increases in the ABP with an elevation in the SNA.

Methods
Ethical approval
The protocol was approved by the Institutional Review Board at Toyo University (TU-2017-004) and each subject provided written informed consent prior to participating in the study. The study was performed in accordance with the principles of the Declaration of Helsinki.

Subjects
A total of 11 young men (mean±SD, age 22±1 years, height 175±5 cm, weight 65±8 kg) participated in this study. They did not have any cerebrovascular or cardiovascular disease and were not taking any medications at the time of enrollment. Prior to each experimental session, the participants were required to abstain from caffeine for 12 h and strenuous exercise and alcohol for 24 h. The experiment was performed at least 3 h after a light meal.

Experimental protocol
Following instrumentation, the subjects were asked to rest in the supine position on a bed for at least 15 min before the commencement of the protocol. Each subject had the thigh cuff occlusion-release protocol at three different conditions: control, early phase of the CPT (cuff release at the 30th s of CPT, CPT30), and the late phase of the CPT (cuff release at the 90th s of CPT, CPT90, Fig. 1). Previous studies [17] suggest that the cerebrovascular response may be modified by cold stimulation-induced pain sensation as well as elevations in ABP and SNA during a CPT. In the present study, to identify the cerebrovascular response isolated from pain sensation, the dCA and critical closing pressure (CrCP) were assessed at two different time phases (CPT30 and CPT90). This is because the pain sensation is not changed during a CPT [18], whereas the ABP and SNA increase gradually and reach their peaks at 60–90 s after cold stimulation [19]. Thus, the different measurement parameters between CPT30 and CPT90 may be due to elevations in the ABP and SNA and not pain sensation.

Each subject immersed their left hand into 1–2 °C water starts at the 90th or 30th s after cuff inflation during CPT30 or CPT90, respectively, and continued to

![Fig. 1 Overview of the experimental protocol](image-url)
immersed until 30 s after cuff deflation. All trials were randomized and separated by a minimum of 15 min for the hemodynamics to return to the baseline. The thigh cuff occlusion-release protocol uses acute hypotension to identify the dCA [20]. Each test was performed twice; thus, each subject performed six total trials of the thigh cuff occlusion-release protocol. The thigh cuff occlusion-release technique consists of a resting baseline of 1-min, 2-min inflation of thigh cuffs (> 180 mmHg), and a 30-s deflation period. Throughout the experimental protocol, the subjects were instructed to adjust their respiratory rate (RR) according to the sound of a metronome (15 breaths/min) to maintain the same end-tidal partial pressure (PETCO₂) level [21].

**Experimental measurements**

The heart rate (HR) was measured using a lead II electrocardiogram (bedside monitor, BMS-3400; Nihon Kohden, Japan). The beat-to-beat arterial blood pressure was monitored continuously using finger photoplethysmography (Finapres Medical Systems, Amsterdam, The Netherlands) to determine the systolic blood pressure (SBP), diastolic blood pressure (DBP), and mean arterial pressure (MAP). The stroke volume (SV) was determined from the BP waveform using the Modelflow software program, which incorporates the sex, age, height, and weight of the subject (Beat Scope1.1; Finapres Medical Systems BV). The cardiac output (CO) was calculated by the SV multiplied by the HR. The PETCO₂, minute ventilation (BV), and RR were sampled from a leak-free mask and respired breaths/min) to maintain the same end-tidal partial pressure (RR) according to the sound of a metronome (15 breaths/min) to maintain the same end-tidal partial pressure (PETCO₂) level [21].

**Data analysis**

The beat-to-beat MAP, MCAv, and PCAv were obtained from each waveform. The cerebrovascular conductance index of the MCA (MCA CVCi) or PCA (PCA CVCi) was calculated by dividing the MCAv or PCAv, respectively, by the MAP. The hemodynamic data were averaged using 30 s data points before cuff deflation and 30 s data point before end of cuff deflation at each condition. The dCA was the average of values in two trials at each condition.

As previously reported, the CrCP of the cerebral circulation was estimated as the index of cerebral vascular tone [22]. Pairs of systolic and diastolic values of MCAv or PCAv and ABP were used to determine pressure–flow velocity relationships, i.e., CrCP [23–26]. The ABP axis intercept of the extrapolated regression line between 30 s of consecutive pairs of systolic and diastolic values of ABP (x-axis, mmHg) and MCAv or PCAv (y-axis, cm/s) waveforms determines the CrCP [22, 27].

We have calculated dCA in both MCA and PCA at each condition according as previous study [20]. The pre-release values of CBF (proportional to velocity) and MAP were defined by their means during the 4 s before thigh cuff release. The relative changes to prerelease value in the MAP, MCAv or PCAv, and MCA CVCi or PCA CVCi during thigh cuff release value were calculated (value/baseline value; normalized units relative to control pre-release values). A slope of the relationship between relative changes to prerelease in the MAP and MCA CVCi or PCA CVCi from 1.0 to 3.5 s after the cuff release was computed as the rate of regulation (RoR), which was used as an index of dCA [20]:

\[ \text{RoR} \ (\text{s}^{-1}) = (\Delta \text{MCA CVCi or } \Delta \text{PCA CVCi} / \Delta T) \Delta \text{MAP}, \]

where \( (\Delta \text{MCA CVCi or } \Delta \text{PCA CVCi} / \Delta T) \) is the slope of the linear regression between MCA CVCi or PCA CVCi and time (\( T \)), and \( \Delta \text{MAP} \), the magnitude of the step, was calculated by subtracting control MAP from the averaged MAP during the interval from 1.0 to 3.5 s [20].

**Statistical analysis**

All data are expressed as mean±SD. One-way analysis of variance (ANOVA) with repeated measures was used to compare for hemodynamics each condition (control, CPT30, and CPT90). Two-way ANOVA with repeated measures (3 conditions; control, CPT30, and CPT90) was performed with Bonferroni’s post hoc test (SPSS 24, IBM, Tokyo, Japan) where appropriate. Effect sizes for ANOVA are reported as partial eta squared (\( \eta_p^2 \)). \( P \) values of <0.05 were regarded as statistically significant.

**Results**

Any differences in hemodynamics during the resting baseline between the different conditions were not observed (\( P>0.238 \)), suggesting that the 15-min interval between the test conditions was enough for the recovery of hemodynamics to the baseline value.

There was no significant difference in the subjective pain sensation between the CPT30 (6.94±2.24) and CPT90 (7.45±2.13) (\( P=0.347 \)). At the CPT30, DBP, SV, and CO were unchanged (\( P>0.234 \)), but the HR, SBP, and MAP increased compared with the control.
conditions ($P<0.025$) (Table 1). In addition, at CPT90, SBP, and DBP, the MAP increased further from the CPT30 ($P<0.008$), but the HR did not change ($P=1.00$). Because the subjects maintained their voluntary RR during the experiment ($P=0.883$), $V_E$ and $P_{ET}CO_2$ were unchanged compared with the control throughout the CPT ($P>0.065$).

Both the MCAv and PCAv were unchanged at the CPT30 and CPT90 compared with control condition (MCA; $P=0.846$, PCA; $P=0.958$, Table 1). The CVCi was decreased in both the MCA and PCA (MCA; $P=0.053$, PCA; $P=0.022$, Table 1), suggesting that CPT causes cerebral vasoconstriction. However, there was no significant difference in changes in the CVCi between MCA and PCA ($P=0.310$, $\eta_p^2=0.111$; Fig. 2). MCA CrCP and PCA CrCP were lower at the CPT90 and CPT30 ($P=0.012$ and $P=0.019$, respectively) compared with the control (Table 1). Although the change in the MCA CrCP and PCA CrCP reduced during CPT, the change in the MCA CrCP from the control conditions was not different from that of the PCA CrCP ($P=0.075$, $\eta_p^2=0.229$; Fig. 2).

The release of the thigh cuffs elicited an acute decrease in the ABP at all the test conditions (Fig. 3). Changes in the MAP at both CPT30 and CPT90 were similar to the control conditions ($P=0.601$). The nadir of the MCAv and PCAv responses to cuff release was not different among the three conditions ($P=0.579$, $\eta_p^2=0.053$; Fig. 3), and there was no difference in the RoR as an index of dCA between conditions or arteries ($P=0.558$, $\eta_p^2=0.057$; Fig. 4).

**Discussion**

The present study examines whether CBF regulation in the posterior cerebral circulation is different from the anterior cerebral circulation during a CPT. Both the MCAv and PCAv were unchanged at the CPT30 and CPT90 from the baseline, despite an elevation in the ABP. In addition, the dCA in both MCA and PCA remained unaltered at CPT30 and CPT90. Interestingly, the MCA CrCP and PCA CrCP, an index of cerebral vascular tone, decreased during the CPT. However, there was no difference in the CrCP between the MCA and PCA. In contrast to our hypothesis, these findings indicate that CPT did not cause any difference in CBF regulation between the posterior and anterior cerebral circulations. These findings indicate that both the anterior and posterior cerebral vasculature were well-regulated (i.e., no over-perfusion occurs) via the dCA despite CPT-induced high blood pressure accompanied with a high SNA.

Unexpectedly, the posterior CBF response to the CPT was not different from that of the anterior CBF (Table 1 and Fig. 2). In addition, both MCAv and PCAv were unchanged during the CPT (Table 1 and Fig. 2), suggesting that the posterior and anterior cerebral circulations were well-regulated despite an elevation in the ABP. Generally, blood flow is determined by the balance between perfusion pressure and vasomotion. However, the cerebral circulation has a dCA, a specific physiological mechanism that maintains adequate cerebral perfusion against a variation in the ABP. Indeed, impairments in the dCA are associated with cerebrovascular diseases [28, 29]. Therefore, the present study identified the dCA and CrCP as indices of cerebral vascular tone during the CPT to address the mechanisms of CBF response to the CPT. As with the CBF response, the response of the dCA and CrCP to the CPT in the posterior cerebral circulation was not different from that of the anterior cerebral circulation. Based on this, the lack of a difference in CBF between the posterior and anterior cerebral circulation was likely due to the response of the dCA and CrCP to the CPT.

The CrCP is a well-established index of cerebral vascular tone, influenced by changes in metabolism (e.g., $CO_2$ and neural activities), transmural pressure (e.g., intracranial pressure), and the SNA [13, 22, 30]. Moreover, previous studies have suggested that an increase in the CrCP might protect the blood–brain barrier.

**Table 1 Hemodynamic parameters at control, CPT30, and CPT90**

|                | Control   | CPT30     | CPT90     |
|----------------|-----------|-----------|-----------|
| HR (bpm)       | 70.2 ± 17.7| 81.5 ± 17.1*| 81.0 ± 15.5*|
| SBP (mmHg)     | 129.1 ± 11.2| 139.9 ± 11.1*| 152.1 ± 15.8*†|
| DBP (mmHg)     | 74.0 ± 8.3  | 78.6 ± 8.3  | 87.8 ± 8.4*†|
| MAP (mmHg)     | 92.4 ± 8.4  | 99.0 ± 8.2*| 109.2 ± 10.2*†|
| SV (mL)        | 90.8 ± 16.9| 94.0 ± 12.4| 920 ± 11.6|
| CO (L/min)     | 63.3 ± 18   | 76.8 ± 18   | 74 ± 1.7   |
| RR (breath/min)| 148.0 ± 0.7 | 148.0 ± 0.7 | 148.0 ± 0.6|
| $V_E$ (L/min)  | 83.1 ± 16   | 87.1 ± 14   | 99 ± 2.5   |
| $P_{ET}CO_2$ (mmHg) | 37.5 ± 2.7 | 37.2 ± 1.9  | 364 ± 2.5  |
| MCAv (cm/s)    | 58.8 ± 6.1  | 60.4 ± 10.3 | 598 ± 12.7 |
| MCA CVCi (cm/s/ mmHg) | 0.64 ± 0.05 | 0.61 ± 0.09 | 0.55 ± 0.11 |
| MCA CrCP (mmHg) | 16.7 ± 13.5 | 10.9 ± 13.2 | 60 ± 16.6* |
| PCAv (cm/s)    | 33.3 ± 7.3  | 33.4 ± 6.7  | 33.1 ± 8.2 |
| PCA CVCi (cm/s/mmHg) | 0.36 ± 0.07 | 0.34 ± 0.07 | 0.30 ± 0.07* |
| PCA CrCP (mmHg) | 13.6 ± 15.9 | 4.6 ± 14.4* | 4.1 ± 19.9 |

Data are presented as means ± SD.  
HR heart rate, SBP systolic blood pressure, DBP diastolic blood pressure, MAP mean arterial pressure, SV stroke volume, CO cardiac output, RR respiratory rate, $V_E$ ventilation, $P_{ET}CO_2$ end-tidal partial pressure of carbon dioxide, MCAv middle cerebral artery mean blood velocity, MCA CVCi middle cerebral artery cerebral vascular conductance index, MCA CrCP middle cerebral artery critical closing pressure, PCAv posterior cerebral artery mean blood velocity, PCA CVCi posterior cerebral artery cerebral vascular conductance index, PCA CrCP posterior cerebral artery critical closing pressure.  
* $P<0.05$ different from control, †$P<0.05$ different from CPT30.
from the over-perfusion induced by exercise-induced hypertension [23, 24]. It has been well-established that there is regional heterogeneity in the sympathetic innervation of the intracranial artery [15]. Therefore, we expected that the CPT will lead to differences in the CrCP and different CBF between the MCA and PCA because increases in the SNA and BP enhance the CrCP [13, 22]. Unexpectedly, the MCA CrCP or PCA CrCP decreased rather than increase at the CPT30 ($P = 0.019$, Fig. 2) and CPT90 ($P = 0.012$, Fig. 2) compared with the control conditions despite an elevation in the ABP. In addition, there was no difference in the change in the CrCP and CBF from the control conditions between the MCA and PCA ($P = 0.075$; Fig. 2). Therefore, the decrease in the CrCP is not consistent with the concept that the cerebral vascular response protects the blood–brain barrier from over-perfusion. Although the CBF response to CPT may be associated with no difference in the response of the CrCP to the CPT between the anterior and posterior cerebral circulation, it is unclear how CPT-induced decreases in the CrCP led to no changes in the CBF with an elevation in the ABP. The CrCP is an index of changes in smaller arterioles, which are more responsive to regional brain metabolic demands [30, 31]. Thus, the reduction in the CrCP may be associated with neural activity-induced metabolic changes at the regional level. Neuroimaging studies have shown that the neural activity of the brainstem is increased at the initial phase of a CPT, whereas high-order regions are activated at the later phase [32, 33]. Therefore, CPT-induced decreases in the CrCP may be affected by neural activation in the brain area supported by the MCA and PCA. However, the CVCi decreased in both the MCA and PCA, suggesting that CPT causes cerebral vasoconstriction. The physiological mechanism responsible for the decrease in CrCP during CPT despite an increase in vascular resistance in the cerebral artery remains unclear.

In the present study, the RoR as an index of the dCA in the MCA and PCA was unchanged during CPT. In contrast to these results, Hilz et al. [34] reported that CPT impaired the dCA in the MCA. One possible reason for this inconsistent result is that the foot was stimulated rather than the hand for the CPT. Although there was no difference in the pressor response during CPT between the hand and foot, the endocrine response and subjective pain sensation during cold stimulation of the foot were larger compared with that of the hand [35]. Because it has been reported that pain intensity differences could modify the CBF response [36], different pain sensation
due to different stimulation methodology may account for the inconsistent result.

In contrast to our hypothesis, the RoR as an index of the dCA in the PCA was unchanged at CPT30 and CPT90, but was not different from that of the MCA ($P=0.558$; Fig. 3). It has been reported that the dCA is lower in the posterior cerebral circulation than in the anterior cerebral circulation under normal physiological conditions [37, 38], but these findings are inconsistent [39, 40]. Alternatively, few studies have compared the anterior and posterior dCA during physiological stimulation. Interestingly, though there is no difference in the dCA between the anterior and posterior cerebral circulation under the supine position, orthostatic stress evokes regional differences in the dCA [10]. This heterogeneous dCA may be partially related to regional heterogeneity in the sympathetic innervation of intracranial arterioles. Indeed, the posterior cerebral circulation may have a less sympathetic innervation than the anterior cerebral circulation [16]. This finding indicates that if the SNA partly contributes to the cerebral vasculature, CPT-induced sympathoexcitation influences are smaller in the posterior circulation than in the anterior circulation and, consequently, cause less cerebral vasoconstriction and less augmented dCA in the posterior circulation. This concept may support the finding that hypertensive disease-induced primary intracerebral hemorrhage occurs mainly in the posterior cerebral circulation rather than the anterior cerebral circulation [1] because the vascular response has a weaker capacity to protect the blood–brain barrier against cerebral over-perfusion during an acute elevation in the ABP. However, in the present study, both the MCAv and PCAv were unchanged during CPT. Moreover, in contrast to our hypothesis, the posterior cerebral circulation was well-regulated despite an elevation in the ABP and SNA. An intact dCA during CPT in the posterior cerebral circulation may be associated with an adequate posterior CBF that is preserved during acute elevations in the ABP and SNA. In addition, these results suggest that the effects of different SNA between the anterior and posterior cerebral circulation during CPT on the dCA may be minimal.

Limitations
Some potential limitations of the present study should be considered. First, the TCD-determined MCAv and PCAv can be used as indices of the anterior and posterior CBF, with the assumption of a constant diameter of the insonated artery. In this regard, several studies have reported no change in the MCA diameter in response to physiological stimulations such as orthostatic stress [41, 42]. In contrast, a recent report demonstrated that dynamic handgrip exercise response caused significant
changes in the MCA diameter [43]. However, if sympathetic activation during CPT elicits a decrease in the MCA or PCA diameter, measuring changes in the MCAv or PCAv would overestimate the anterior or posterior CBF because a reduction in diameter would increase blood velocity. Thus, because there is no change in the MCAv and PCAv during CPT, it can be inferred that changes in the MCAv and PCAv likely reflect changes in blood flow. Second, a few previous studies have suggested that the dCA is relatively more effective at protecting the brain against transient hypertension than hypotension [44, 45]. Therefore, our findings may not reflect the RoR using transient hypertensive stimuli. Third, we did not examine the effect of CPT without cuff inflation on hemodynamics response to reduce the burden of subjects in the present study. However, there was no significant difference in MAP, MCAv, and PCAv between the resting baseline and cuff inflation (MAP; \(P = 0.179\), MCA; \(P = 0.114\), PCA; \(P = 0.201\)). Therefore, the effect of cuff inflation on hemodynamics response to CPT may be minimal. Finally, the participants in the present study were young healthy subjects. Therefore, the results may differ in older aged individuals with a greater risk of hypertension or stroke. Indeed, a previous study reported that the CBF response to cold stimulation is different between old and young individuals [46]. Thus, a dedicated study on the effect of aging in CBF regulation during CPT in humans is required to address this issue.

**Conclusion**

Contrary to our hypothesis, there was no difference in CBF and its regulation between the anterior and posterior cerebral circulation during an elevation in ABP and SNA. This finding suggests that CPT-induced acute elevations in blood pressure and SNA did not affect the regulation of posterior cerebral vasculature, and the posterior CBF and the anterior cerebral circulation was well-regulated. However, the effects of a chronic elevation in ABP (hypertension) on the posterior cerebral circulation may be different from that of an acute elevation in ABP.

**Abbreviations**

ABP: arterial blood pressure; CBF: cerebral blood flow; CO2: carbon dioxide; CO: cardiac output; CPP: cerebral perfusion pressure; CPT: cold pressor test; CrCP: critical closing pressure; CVC: cerebrovascular conductance index; DBP: diastolic blood pressure; dCA: dynamic cerebral autoregulation; HR: heart rate; MAP: mean arterial pressure; MCA: middle cerebral artery; PCA: posterior cerebral artery; \(P_{a,CO2}\): end-tidal partial pressure of carbon dioxide; RoR: rate of regulation; RR: respiratory rate; SBP: systolic blood pressure; SNA: sympathetic nervous activity; SV: stroke volume; VAS: visual analog scale; VE: minute ventilation.

**Acknowledgements**

We appreciate the commitment of all participants of this study and their staff.

**Authors’ contributions**

TW, SO conception and design of research. TW, SO performed experiments. TW analyzed data. TW, SO interpreted results of experiments. TW prepared figures. TW, HW, SO drafted manuscript. All authors edited and revised manuscript. All authors read and approved the final manuscript.

**Funding**

This study was supported by “Grant-in-Aid for Scientific Research from the Japanese Ministry of Education, Culture, Sports, Science and Technology” (No. 18J20884 to T.W.; No. 18H03156 to S.O.) and “the INOUE ENRYO Memorial Grant, TOYO University.”

**Availability of data and materials**

All relevant data are within the paper.

**Ethical approval and consent to participate**

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. All the participants signed an informed consent form agreeing to submit to the procedures involved in the study.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

**Author details**

1 Department of Biomedical Engineering, Toyo University, 2100 Kujirai, Kawagoe-shi, Saitama 350-8585, Japan. 2 Research Fellow of Japan Society for the Promotion of Science, Tokyo, Japan.

**Received:** 21 October 2019 **Accepted:** 22 January 2020 **Published online:** 29 January 2020

**References**

1. Dunatov S, Antonicic I, Bralic M (2011) Hemodynamic changes in the posterior cerebral circulation triggered by insufficient sympathetic innervation—cause of primary intracerebral hemorrhage? Med Hypotheses 76:668–669

2. Tuirn TC, Okamura T, Afzal AR, Rumana N, Watanabe M, Higashiyama A, Nakao Y, Nakai M, Takegami M, Nishimura K et al (2016) Hypertension and lifetime risk of stroke. J Hypertens 34:116–122

3. Takahashi I, Geyer SM, Nishi N, Ohshita T, Takahashi T, Akahoshi M, Fujiwara S, Kodama K, Matsumoto M (2011) Lifetime risk of stroke and impact of hypertension: estimates from the adult health study in Hiroshima and Nagasaki. Hypertens Res 34:649–654

4. Ogoh S, Sato K, Hirasawa A, Sadamato T (2019) The effect of muscle metaboreflex on the distribution of blood flow in cerebral arteries during isometric exercise. J Physiol Sci 69:375–385

5. Ogoh S, Sato K, Okazaki K, Miyamoto T, Hirasawa A, Sadamato T, Shibasaki M (2015) Blood flow in internal carotid and vertebral arteries during graded lower body negative pressure in humans. Exp Physiol 100:259–266

6. Sato K, Ogoh S, Hirasawa A, Oue A, Sadamato T (2011) The distribution of blood flow in the carotid and vertebral arteries during dynamic exercise in humans. J Physiol 589:2847–2856

7. Ogoh S, Sato K, Nakahara H, Okazaki K, Subudhi AW, Miyamoto T (2013) Effect of acute hypoxia on blood flow in vertebral and internal carotid arteries. Exp Physiol 98:692–698

8. Ogoh S, Sato K, Okazaki K, Miyamoto T, Hirasawa A, Morimoto K, Shibasaki M (2013) Blood flow distribution during heat stress: cerebral and systemic blood flow. J Cereb Blood Flow Metab 33:1915–1920

9. Sato K, Sadamato T, Hirasawa A, Oue A, Subudhi AW, Miyazawa T, Ogoh S (2012) Differential blood flow responses to CO2 in human internal and external carotid and vertebral arteries. J Physiol 590:3277–3290
10. Sato K, Fisher JP, Seifert T, Overgaard M, Secher NH, Ogoh S (2012) Blood flow in internal carotid and vertebral arteries during orthostatic stress. Exp Physiol 97:1272–1280

11. Diedler J, Sykora M, Rupp A, Poli S, Karpel-Massler G, Sakowitz O, Steiner T (2009) Impaired cerebral vasomotor activity in spontaneous intracerebral hemorrhage. Stroke 40:815–819

12. Ogoh S, Brothers RM, Eubank WL, Raven PB (2008) Autonomic neural control of the cerebral vasculature: acute hypotension. Stroke 39:1979–1987

13. Purkayastha S, Saxena A, Eubank WL, Hoxha B, Raven PB (2013) alpha1-Adrenergic receptor control of the cerebral vasculature in humans at rest and during exercise. Exp Physiol 98:451–461

14. Zhang R, Zuckerman JH, Iwasaki K, Wilson TE, Crandall CG, Levine BD (2002) Autonomic neural control of dynamic cerebral autoregulation in humans. Circulation 106:1814–1820

15. Heistad DD, Marcus ML, Gross PM (1978) Effects of sympathetic nerves on cerebral vessels in dog, cat, and monkey. Am J Physiol 235:H544–552

16. Edvinsson L, Owman C, Sjoberg NO (1976) Autonomic nerves, mast cells, and amine receptors in human brain vessels. A histochemical and pharmacological study. Brain Res. 115:377–393

17. Perry BG, Bear TL, Lucas SJ, Mundel T (2016) Mild dehydration modifies the cerebrovascular response to the cold pressor test. Exp Physiol 101:135–142

18. Kriegel KC, Seals DR, Callister R (1992) Sympathetic nervous system activity during skin cooling in humans: relationship to stimulus intensity and pain sensation. J Physiol 454:359–371

19. Yamamoto K, Iwase S, Mano T (1992) Responses of muscle sympathetic nerve activity and cardiac output to the cold pressor test. Jpn J Physiol 42:239–252

20. Aaslid R, Lindegaard KF, Sorteberg W, Nornes H (1989) Cerebral autoregulation dynamics in humans. Stroke 20:45–52

21. Ogoh S, Sato K, Akimoto T, Oue A, Hirasawa A, Sadamoto T (1985) Critical closing pressure of the cerebral circulation during dynamic exercise in humans. J Appl Physiol 1980(108):1701–1705

22. Panerai RB (2003) The critical closing pressure of the cerebral circulation. Med Eng Phys 25:621–632

23. Ogoh S, Fisher JP, Young CN, Fadel PJ (2011) Impact of age on critical closing pressure of the cerebral circulation during dynamic exercise in humans. Exp Physiol 96:417–425

24. Ogoh S, Brothers RM, Jeschke M, Secher NH, Raven PB (2010) Estimation of cerebral vascular tone during exercise; evaluation of age closing pressure in humans. Exp Physiol 95:678–685

25. Czosnyka M, Smielewski P, Piechnik S, Al-Rawi PG, Kirkpatrick PJ, Matta BF, Pickard JD (1999) Critical closing pressure in cerebral vasoregulation in humans. J Neurol Neurosurg Psychiatry 66:606–611

26. Richards HK, Czosnyka M, Pickard JD (1999) Assessment of critical closing pressure in the cerebral circulation as a measure of cerebrovascular tone. Acta Neurochir. 141:1221–1227

27. Burton AC (1951) On the physical equilibrum of small blood vessels. Am J Physiol 146:319–329

28. Aries MJ, Ething JW, De Keyser J, Kremer BP, Vroomen PC (2010) Cerebral autoregulation in stroke: a review of transcranial Doppler studies. Stroke 41:2697–2704

29. Chi NF, Hu HH, Wang CY, Chan L, Peng CK, Novak V, Hu CJ (2018) Dynamic cerebral autoregulation is an independent functional outcome predictor of mild acute ischemic stroke. Stroke 49:2605–2611

30. Panerai RB, Eyre M, Potter JF (2012) Multivariate modeling of cognitive-motor stimulation on neurovascular coupling: transcranial Doppler used to characterize myogenic and metabolic influences. Am J Physiol Regul Integr Comp Physiol 303:R395–407

31. Wei EP, Kontos HA, Patterson JL, Jr (1980) Dependence of pial arteriolar response to hypercapnia on vessel size. Am J Physiol 238:697–703

32. Petrovic P, Petersson KM, Hansson P, Ingvar M (2004) Brainstem involvement in the initial response to pain. Neuroimage 22:995–1005

33. Price DD (2009) Psychological and neural mechanisms of the affective dimension of pain. Science 288:1769–1772

34. Hilz MJ, Wang R, Marthol H, Liu M, Tilmann A, Riss S, Hauck P, Hosi KM, Wasmiejer G, Stempner B, Kohrmann M (2016) Partial pharmacologic blockade shows sympathetic connection between blood pressure and cerebral blood flow velocity fluctuations. J Neurol Sci 365:181–187

35. Larra MF, Schilling TM, Rohrig P, Schachinger H (2015) Enhanced stress response by a bilateral feet compared to a unilateral hand Cold Pressor Test. Stress 18:589–596

36. Montoro CJ, Duscheck S, de Guevara CA, Reyes Del Paso GA (2016) Patterns of cerebral blood flow modulation during painful stimulation in fibromyalgia: a transcranial Doppler Sonography Study. Pain Med 17:2256–2267

37. Haubrich C, Wendt A, Diehl RR, Klotzsch C (2004) Dynamic autoregulation testing in the posterior cerebral artery. Stroke 35:848–852

38. Sorond FA, Khvayi R, Serrador JM, Lipsitz LA (2005) Regional cerebral autoregulation during orthostatic stress: age-related differences. J Gerontol A Biol Sci Med Sci 60:1484–1487

39. Nakagawa K, Serrador JM, Larose SL, Mosleh F, Lipsitz LA, Sorond FA (2009) Autoregulation in the posterior circulation is altered by the metabolic state of the visual cortex. Stroke 40:2062–2067

40. Reinhard M, Schork J, Allignol A, Weiller C, Kaube H (2012) Cerebellar and cerebral autoregulation in migraine. Stroke 43:987–993

41. Giller CA, Bowman G, Dyer H, Mootz L, Krippner W (1993) Cerebral arterial diameters during changes in blood pressure and carbon dioxide during craniotomy. Neurosurgery. 32:737–741

42. Serrador JM, Picot PA, Rutt BK, Shoemaker JK, Bondar RL (2000) MRI measures of middle cerebral artery diameter in conscious humans during simulated orthostasis. Stroke 31:1672–1678

43. Verbree J, Bronzwaer A, van Buchem MA, Daemen M, van Lieshout JJ, van Osch M (2009) Autoregulation in the posterior circulation is altered by the metabolic state of the visual cortex. Stroke 40:2062–2067

44. Brassard P, Ferland-Dutil H, Smid JT, Paquette M, Le Blanc O, Malenfant S, Ainslie PN (2017) Evidence for hysteresis in the cerebral pressure-flow relationship in healthy men. Am J Physiol Heart Circ Physiol 312:H701–H704

45. Tseng YC, Willie CK, Atkinson G, Lucas SJ, Wong A, Ainslie PN (2010) Cerebrovascular regulation during transient hypotension and hypertension in humans. Hypertension 56:268–273

46. Fluck D, Ainslie PN, Bain AR, Wildfong KW, Morris LE, Fisher JP (1985) Extra- and intracranial blood flow regulation during the cold pressor test: influence of age. J Appl Physiol 1987(123):1071–1080

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.