REVIEW

The impact of nitrogen starvation on the dynamics of lipid and biomass production in Scenedesmus sp.

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ABSTRACT

Microalgal lipid is a major natural feedstock for biodiesel production. However, microalgae-based biofuel technology comes with obstacles to production, such as high investment and operating costs. To overcome these problems, nowadays some strategies have been used during cultivation stage of the microalgae for enhancing biomass and accumulate lipids and carbohydrates which could be used for biofuel production. The most common methods applied to microalgae are classified as nutrient stress and changes in growth conditions that lead to increase the lipid content in the species without decreasing the growth rate of their potential strains or by simultaneously increasing both of these. Scenedesmus sp. are considered to be the most appropriate microalgae to culture commercially due to their high biomass, lipid and carbohydrate yield. The purpose of this review was to describe nutrient stress strategy to develop biofuels as a sustainable alternative to fossil fuels and, in particular, with respect to nitrogen nutrient limitations, the lipid yield and biomass development in Scenedesmus sp. microalgae. The nitrogen starvation/limitation strategy that will increase the general economic feasibility of microalgal lipid production and affect the fatty acid composition was presented.

Keywords: Biofuel, lipid productivity, microalgae, nitrogen starvation, Scenedesmus sp., triacylglycerol (TAG)

1. INTRODUCTION

Biofuels have been considered to be a clean alternative to fossil fuels in recent years because of their sustainability, nontoxicity, biodegradability, and low greenhouse gas emissions [1-3]. Microalgae have a 30% lipid content, which is higher than that in either palm oil or soybean oil, which are currently used in biodiesel production (<5% of dry biomass) [4, 5]. The lipid content in microalgae varies from 20 to 40% on a dry-weight basis; whereas, some microalgae have lipid contents as high as 85% [6, 7]. Microalgae offer many advantages over traditional oil crop biofuels and chemicals of high value because of their properties of high lipid productivity and photosynthetic efficiency, robust environmental adaptation, failure to compete with nutrients or arable lands, rapid fixation of environmental carbon, and ease of cultivation in media similar to wastewater and other media [8]; however, the biggest obstacle to obtaining biofuels from algae is the high cost of biodiesel production from oleaginous microalgae and the need for and cost of pilot-scale studies [9, 10]. Optimizing the conditions under which microalgae strains can be cultivated is crucial to creating enough lipid production to enable biofuel production on an economically significant scale [11]. Stimulating stress responses by limiting nutrients and controlling cultivation is one of the methods by which the lipid content in microalgae can be improved [12-14].

Microalgae are rich in tri- and diglycerides, phospho- and glycolipids, hydrocarbons, and other lipids [6], [15]. Under suitable environmental conditions, microalgae synthesize fatty acids to produce membrane glycolipids, mainly glycolipids and phospholipids; however, several microalgae alter their lipid biosynthetic pathways to produce a large amount of neutral lipids in the form of triglycerides (TAGs), stored mostly in cytosolic lipid bodies under stress conditions [16]. Under these conditions, microalgae cell division will stop, and TAGs will be stored in their cells as a survival approach to withstand these adverse conditions [17]. These TAGs, which are generally found in a cell as storage lipids,
can be converted into biodiesel [18]. Lipids are stored in microalgae cells when accessible nutrients are consumed and depleted from a culture medium and become growth-limiting factors [19]. In addition, the amount and type of lipids are different in different species; therefore, the growth and cultivation conditions of the microalgae species are altered to affect their fatty acids [20]. *Scenedesmus* sp. is considered to be one of the most important microalgae for producing biofuel raw material because of their high biomass, high lipid and carbohydrate content, and ability to grow under various wastewater conditions [21].

Fatty acid ratios and lipid contents in microalgae differ according to environmental and cultural parameters, such as nitrogen and phosphorus concentrations, light intensity, growth phase, light-receiving period, temperature, salinity, and carbon dioxide (CO$_2$) concentration [22, 23]. Among these, nitrogen starvation is the most important technique reported in the literature that increases microalgae biomass and biochemical components, such as lipids and carbohydrates [24]. The purpose of this review was to investigate biomass productivity in the *Scenedesmus* sp., a species with a high lipid content, to enable improvement of the economic feasibility of microalgae-based biofuels and to examine the methods by which nitrogen starvation or limitation increase the lipid yield and carbohydrate production; and also to guide researchers in understanding the reaction of *Scenedesmus* sp. against its cultivation in nitrogen-limited nutrient media.

### 2. Microalgae Culture Conditions

The growth characteristics and composition of microalgae depend largely on four fundamental cultivation conditions—photoautotrophic, heterotrophic, mixotrophic, and photoheterotrophic [25]-[26]. Table 1 provides the characteristics of these different conditions.

| Culture conditions | Energy source | Carbon source |
|--------------------|---------------|---------------|
| Photoautotrophy    | Light         | Inorganic     |
| Heterotrophy       | Organic carbon| Organic       |
| Photoheterotrophy  | Light         | Organic       |
| Mixotrophy         | Light and organic carbon | Inorganic and organic |

Microalgae are divided into two groups—autotrophic and heterotrophic. Autotrophic microalgae use sunlight to convert CO$_2$ into lipids without the need for carbon; however, cultivation requires a large area, and lipid accumulation is slow [28]. On the contrary, heterotrophic microalgae have a high lipid content and rapid growth rate and lipid accumulation; therefore, they have a high capacity for lipid production. Photoautotrophic cultivation, in which microalgae utilize inorganic carbon (such as CO$_2$) and solar energy to generate chemical energy, is the most widespread technique [29]. In heterotrophic cultivation, microalgae can grow in the absence of light by utilizing glucose and other similar organic carbon sources [30]. Although the heterotrophic system provides higher lipid output than the autotrophic system, the need for organic substrates and their cost prevent its industrial-scale production and commercialization [31].

Mixotrophic cultivation combines both autotrophic and heterotrophic systems because both CO$_2$ and organic carbon are simultaneously absorbed [32]. In photoheterotrophic cultivation, microalgae need both organic carbon and light. The difference between the two is that photoheterotrophs use light and mixotrophs use organic carbon as an energy source.

When comparing different microalgae species with different carbon sources, autotrophs generally have satisfactory lipid content, but because their growth rate is low, subsequent lipid production efficiency is also low. The opposite is true for heterotrophs, in which lipid content is low but the growth rate is very high, resulting in higher biomass and lipid production. Mixotrophs utilize both inorganic and organic carbon sources; these conditions result in the highest biomass and lipid yield among all cultivation methods [14].

### 3. Microalgal Lipid Classification and Accumulation Strategies

Microalgae-based biodiesel technologies contain the conditions that determine optimum microalgae growth, define economically feasible and efficient culturing media, harvest and separate microalgae biomass, and produce biodiesel (Fig 1). Biodiesel production from microalgae comprise the following four basic steps: isolating and characterizing the microalgae, producing microalgae biomass, harvesting, and processing [33].

Lipids are not only the energy sources in microalgae cells but also a promising raw material for biodiesel production [37]. When energy input (photosynthesis rate) exceeds energy output (cell growth and division), microalgae synthesize lipids [38]. These microalgae contain refined lipids that can be extracted to create fuel oils [39]. There are more than 300,000 known microalgae species, of which 70% have a high lipid content and a fuel yield of 9500–35000 liters per 1000 m$^2$ [40]. Microalgae have advantages over other microorganisms in biofuel production because of their greater biomass productivity and ability to accumulate high amounts of lipids [42].
The biochemical composition of microalgae comprises four main components—proteins, carbohydrates, nucleic acids in varying ratios among species, and lipids. Lipids are the highest energy source at 37.6 kJ g\(^{-1}\), which can be either polar or nonpolar, followed by proteins at 16.7 kJ g\(^{-1}\) and carbohydrates at 15.7 kJ g\(^{-1}\). Depending on the metabolic rate, the ratio of these lipids changes during different microalgae growth periods [43]. Neutral lipids are composed of glycerol and free fatty acids (FFAs) and because of their low degree of unsaturation, they are more suited for producing biodiesel [44]. Polar lipids, such as phospholipids (PL) and glycolipids (GL), are structural [45] and are important components of the outer membranes of chloroplasts. Nonpolar (neutral) lipids, such as TAGs, diglycerides (DAGs), monoglycerides (MAGs), FFAs, pigments (e.g., chlorophylls), and hydrocarbons, are stored. Algae store lipids differently depending on the species, their growth periods, and environmental conditions. FFA components also vary with different microalgae strains. In most cases, polyunsaturated fatty acids (PUFAs) are located in the structural lipids, while monounsaturated fatty acids (MUFAs) and saturated fatty acids (SFAs) are located in the stored lipids [46]-[48]. Table 2 presents the lipid classifications in microalgae.

### Table 2. Lipid classifications in microalgae [49]

| Neutral Lipids | Glycolipids | Phospholipids |
|----------------|------------|---------------|
| Triglycerides  | Phosphatidylcholine | Polymerized triglyceride |
| Wax esters     | Phosphatidylethanolamine | Monogalactosyldiglyceride |
| Hydrocarbons   | Phosphatidylserine | Digalactosyldiglyceride |
| Free fatty acids | Phosphatidylglycerol |                      |
| Sterols        | Phosphatidylinositol |                      |

Algae growth has five different phases [50] as follows: 1) lag or acclimation, 2) logarithmic, 3) decreasing, 4) stationary, and 5) death. To obtain the highest biomass yield, the growth rate in the media should always be within the logarithmic growth phase. The highest lipid synthesis takes place during the transition between the lag and the stationary growth phases (i.e., stages 2 and 3) [51]. Microalgae harvested during the stationary phase have less polar lipids and more neutral lipids (e.g., TAGs) than those harvested during the logarithmic growth phase, so harvesting at the stationary phase is more suitable for biodiesel production [47]. The most well-known lipids are TAGs formed by one glycerol and three fatty acids, and PLs (and GLs), formed by two fatty acids and one phosphate (and carbohydrate). The natural lipids produced by microalgae are TAGs, which makes them the main focal point because of having the necessary molecular structure for biodiesel production [52]. TAG molecules function as electron pools when the electron supply of energy and carbon storage compounds and photosynthesis cannot meet the demand of growing cells [47]. The common methyl esters used in biodiesels are methyl palmitate (C16:0), methyl stearate (C18:0), methyl oleate (C18:1), methyl linoleate (C18:2), and methyl linolenate (C18:3). In other words, microalgae lipids are
generally composed of neutral lipids with a lower amount of unsaturation [43], which is the natural fatty acid profile of microalgae [53]. The optimum fatty acid ratio from microalgae for producing biodiesel is 5:4:1 for C16:1, C18:1, and C18:2, respectively, which gives biodiesel its distinguishing properties, such as high cetane number (CN), low iodine value (IV), and low cold filter plugging point (CFPP) [11].

The reaction of fatty acids in 1 mole of triglycerides and 3 moles of alcohol during biodiesel production yields fatty acid methyl esters (FAME) and glycerol (side product) (Fig 2). The glycerol can be separated from the biodiesel using phase separation techniques or “transesterification,” in which glycerol is displaced by methanol in the presence of a catalyst [6].

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\begin{align*}
\text{CH}_2-\text{OCOR}_1 & \quad \text{catalyst} \quad \text{CH}_2-\text{OH} & \quad R_1-\text{COOCH}_3 \\
\text{CH} - \text{OCOR}_2 + 3\text{HOCH}_3 & \quad \text{CH} - \text{OH} + R_2-\text{COOCH}_3 \\
\text{CH}_2-\text{OCOR}_3 & \quad \text{CH}_2-\text{OH} + R_3-\text{COOCH}_3
\end{align*}
\]

Fig 2. Algae biodiesel production using transesterification [54]

Table 3. Parameters that affect lipid production in microalgae [37]

| Physicochemical Properties | Species-Dependent Factors | Culture System Improvement |
|---------------------------|---------------------------|---------------------------|
| **Light** | *Lipid productivity | *Photosynthetic efficiency |
| - Intensity | -Biomass productivity | -Light penetration |
| - Spectral quality | -Lipid contents | -Light distribution |
| **Temperature** | *Fatty acid profiles | *Mass transfer rate |
| - Reactive oxygen species level | -Optimal fatty acid ratio | -Aeration rate |
| - Fatty acid composition | *Harvesting method | *Nutrient control |
| **Carbon dioxide (CO2) and pH** | | |
| - CO2 concentration | -Cell size | -Semipermeable membrane |
| - Bicarbonate ion (HCO3-) concentration | -Cell density | |
| **Nutrient starvation** | | |
| - Nitrogen starvation | -Surface property | |

(under specific conditions after inducing stresses and depending on the species, the lipid content in the microalgae can change to between 1 and 90% of their dry weight [6]. Table 3 lists the parameters to which changes can induce lipid accumulation.)
The cost of the microalgal-based biodiesel is comparable to that of petroleum diesel, and this has driven many researchers to study how to increase lipid production in microalgae to further decrease the cost of producing biodiesel [63-66]. Put simply, by following new methods by which high biomass yields, the desired lipid content, and the desired lipid composition can be generated, the overall efficiency of lipid production and other valuable chemicals produced by microalgae can increase [16, 43].

Because most microalgae accumulate a considerable amount of lipids under stress, lipid production can be highly efficient under specific conditions leading stresses. On the other hand, under these stress conditions in which lipids and TAGs are biosynthesized, cell growth and biomass yield are jeopardized, which also leads to protein decomposition and can affect the overall lipid yield differently depending on the species [36].

Different *Scenedesmus* species show differences in the number of cell spikes and the texture of their cell walls. Morphologic changes in *Scenedesmus* species result from changes in nutrient concentrations, pH, and/or allelochemicals [71, 72].

**Nutrient stress**

Nitrogen, iron, phosphorus, magnesium, sulfur, and silicon are important for photosynthesis, cell division, respiration, intercellular transport, and protein synthesis in microalgae [73]. In particular, nitrogen and phosphorus are the two basic macronutrients for microalgae growth. Nitrogen is required for the synthesis of growth metabolites and protein, which are limiting factors on the overall growth rate, and is part of the chemical structure of nucleic acids, proteins, and other biomolecules [74]. Theoretically, these elements in the microalgae growth media should be within a stoichiometric ratio of C168H218O43N27P for optimum growth (C for carbon, H for hydrogen, O for oxygen, N for nitrogen, P for phosphorus). When the nitrogen: phosphorus ratio is 5:1, the environment is classified as nitrogen limiting, and when the ratio is 30:1, the environment is phosphorus limiting [43].

**General properties of Scenedesmus sp.**

The success of mass cultivation of microalgae, especially considering a low-value product, such as biodiesel, depends on selecting the correct species [55]. Even though there are many ways suggested in the literature by which to improve lipid production in microalgae, if the appropriate species is not used, several limitations can arise [37]. *Nannochloropsis* sp., *Chlorella* sp., *Chlamydomonas* sp., *Scenedesmus* sp., *Dunaliella* sp., *Isochrysis* sp., and *Botryococcus braunii* are multifunctional microalgae that accumulate lipids between 10 and 75% of their dry weights [21]. Among these, *Scenedesmus* sp. are considered to be the most appropriate microalgae for producing biodiesel because of their high biomass, lipid and carbohydrate yield and ability to grow in various wastewater environments, and short doubling time [21, 67, 68].

Rapid growth rate increases the biomass yield of microalgae, and stress conditions increase their lipid contents [75, 76]. Microalgae have a tendency to accumulate polysaccharides and/or neutral lipids under stress. This defense mechanism is exploited in the production of various metabolites; such as neutral lipids, carotenoids, and polysaccharides [77]. Fig 4 illustrates the increase in the lipid content in microalgae under different stress conditions.

While a nutrient efficiency ceases cell growth, it helps in channeling metabolic fluxes towards fatty acid biosynthesis. It is resuluted in accumulation of storage lipid in the form of triacylglycerols (TAGs). Under the nutrient starvation-driven lipid accumulation concept, the activation of diacylglycerol acyltransferase enzyme is thought to convert acyl-CoA to triglyceride (TAG) that results in lipid accumulation [78].

A large amount of lipids can be accumulated in microalgae under nitrogen-limiting conditions [79]; however, the challenge is to find the balance between increased lipid production and biomass yield. Nitrogen has a huge impact on both processes and is directly proportional to biomass yield and negatively proportional to lipid content [80]. As a result, both high biomass yield and high lipid content cannot be attained at concurrently [81]. Stresses adversely affect microalgal growth and, in the end, reduce output of

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**Fig 3. Scenedesmus sp.** (a) Freshwater green microalgae Scenedesmus sp. [69], (b) Scanning electron micrographs of *S. quadricauda* [70].

**Fig 4.** Illustrates the increase in the lipid content in microalgae under different stress conditions.
the desired product; however, doubling the biomass yield can reduce biodiesel prices by 41–42% [82]. Under nutrient-limiting conditions, microalgae relocate their carbon resources into energy-rich lipid and starch compounds [79]. The proportion of polar and nonpolar lipids can be controlled by changing the ratio of nitrogen, phosphorus, and inorganic carbon in the growth media [83].

There are three degrees of microalgae sensitivity to the manipulation of nutrient concentration—starvation, limitation, and depletion. Khozin-Goldberg and Cohen [84] presented three possible reasons for lipid accumulation under nitrogen starved conditions: (1) decreased in cellular content of thylakoid membrane, (2) activation of acyl hydrolase and (3) stimulation of phospholipids hydrolysis [78, 85, 86]. Conventional process biochemistry approaches involving nutrient limitation/deprivation or physiological stress conditions might be useful for enhancing lipid content on dry weight basis. However, the increment in lipid content happens at the cost of biomass. So, it fails to enhance lipid productivity significantly. Therefore, the proportion of lipid productivity versus biomass formation remains same [78]. In Fig 5, the effects of physico-chemical parameters on lipid accumulations was given in a microalgal cell factory. As a nitrogen stress condition, nitrate starvation was defined in the algal cell. Other physico-chemical stress conditions also affect lipid accumulation in the algal cell but the probable mechanism for lipid accumulation is not known. Therefore, the knowledge of the biochemical mechanisms and molecular insights for lipid accumulation influenced by the environmental conditions might be improved by metabolic and molecular engineering.

During starvation, the microalgae are first grown in a nutrient-rich environment and then transferred into a nutrient-depleted environment. This nutrient deficiency results in the generation of high-energy compounds and their accumulation. In nutrient-limited media, one nutrient that limits maximum biomass production and induces a physiological reaction is limited while the others are abundant. The fundamental idea behind this condition is called “the law of the minimum”, which assumes that there is usually one nutrient lacking in a media and that limits biomass growth while all other nutrients are in excess. This method is sequentially conducted in a culture medium. First, the cells are cultivated in a nutrient-rich environment and cell density and growth rate increase until the nutrients are depleted. Then, with changes in some metabolic processes, energy-rich compounds increase and growth rate and photosynthesis decrease [43].

**Nitrogen starvation**

Nitrogen is not only a significant element found in the structure of proteins and genetic material, but also one of the most prevalent elements in the entire structures, after carbon, hydrogen and oxygen; therefore, cells require nitrogen to grow and multiply [87]. Nitrogen deficiency in the microalgae growth media is applied as either nitrogen depletion or limitation (Fig 6). In nitrogen depletion, microalgae grow in a media without a nitrogen source; in nitrogen limitation, there is a specific nitrogen source but it is limited compared to that of other nutrients [88]. When nitrogen is limited more than other nutrients, photosynthesis will continue, but the chemicals produced in microalgae will have less nitrogen and be more energy rich (e.g., will contain more lipids and carbohydrates). Moreover, when the nitrogen in the cell is depleted, microalgae begin to decompose nitrogen-containing nonlipid cellular chemicals and free the nitrogen [89]. Nitrogen deficiency is the cheapest and easiest method by which to enhance lipid production; therefore, utilization of this method is widespread and is trustworthy and effective for many species [83].
Nitrogen is quantitatively important in growth media and its limitation, depending on the species, inhibits microalgal growth [90]. Declining cell multiplication changes the route through which lipids are synthesized from membrane lipids to neutral lipids [91].

Nitrogen limitation or starvation causes an increase in lipid and carbohydrate content while decreasing growth rate, protein synthesis, photosynthesis, and cell size [92]. Some examples for nitrogen starvation effect on lipid production of *Scenedesmus* sp. were shown in Table 4. Schnurr et al. [93] have researched the possibility of increasing the neutral lipid content in a nutrient-deficient algae biofilm using *S. obliquus* and *Nitzschia palea*. Although the neutral lipid content in suspensions of the same species was doubled, there was no concentration increase observed in their biofilms.

Many researchers have reported that nitrogen starvation decreases photosynthesis and protein synthesis but increases lipid and carbohydrate synthesis [92]. In addition, environmental conditions, especially nitrogen-rich nutrients, play a significant role in the amount and quality of the lipids produced in the microalgae [47, 93].

Lipid production in microalgae increases with nitrogen deficiency [95]-[96], and changes in the fatty acid chain length and unsaturation results in more suitable TAG production for biodiesel. Many microalgae species can similarly adapt their metabolism under nitrogen deficiency [47, 97]. During the stationary growth phase, nutrient starvation is the
most well-known method by which to improve lipid content. In addition, temperature, salinity, light, and changes during the growth phase also impact microalgae metabolism [98]. Breuer et al. [99] have found that the highest lipid content, independent of light intensity, was observed in S. obliquus and changed from 18 to 40% of dry weight with changes in pH and temperature. This lipid accumulation stores the energy to be allocated for growth to prevent a decline in growth rate and, subsequently, in biomass and lipid production [100]. Toledo-Cervantes et al. [101] have evaluated CO₂, aeration, and light intensity impacts on the growth of and lipid production in S. obstusiusculus under nitrogen-limiting conditions. Without any nutrient limitations, in 20% nonpolar algae-containing culture with 5% CO₂ and 134 µmole m⁻² s⁻¹ light intensity, biomass productivity was 500 g m⁻³ d⁻¹ and maximum biomass concentration was 6000 g m⁻³. Under nitrogen-limiting conditions with 5% CO₂, lipid accumulation was 55.7% of the dry weight.

**Table 4. Effect of nitrogen limitation/deprivation on lipid production of Scenedesmus sp.**

| Strain of Scenedesmus sp. | Lipid productivity | References |
|--------------------------|--------------------|------------|
| S. obliquus              | 0.18 g m⁻² d⁻¹ lipid productivity | [93] |
| S. obliquus              | 322 mg L⁻¹ d⁻¹ lipid productivity | [99] |
| S. obstusiusculus        | For the inlet CO₂ concentrations of 0.04% and 5% CO₂, the maximum lipid productivities were 51 and 200 g m⁻³ d⁻¹ respectively. | [101] |
| S. obliquus CNW-N        | 45.48 mg L⁻¹ d⁻¹ lipid productivity | [103] |
| S. acutus                | The amount of lipid in the 50% nitrogen-limited media was 19.48% higher than that in the control group | [104] |
| S. dimorphus             | 0.17 g L⁻¹ d⁻¹ lipid productivity | [106] |
| S. abundans              | 5.999 g L⁻¹ d⁻¹ lipid productivity | [107] |
| S. quadricauda           | 25.13 mg L⁻¹ d⁻¹ lipid productivity | [108] |
| S. obtusiusculus         | 0.34 g L⁻¹ d⁻¹ lipid productivity | [109] |
| Scenedesmus sp. LX1     | 204 mg L⁻¹ d⁻¹ lipid productivity | [85] |

Although these nutrient-starvation conditions are generally considered for systems with long turnover periods, in recent studies, there have also been satisfactory results for systems with short-turnover periods [102]. The lipid content of Scenedesmus sp. changes with three different phases. The biomass on day 2 during the nitrogen-consuming logarithmic phase is dark green with 20% lipid content. During the early stress phase (5–7 d), the biomass is greenish yellow with ~35% lipid content. During the late stress period (10–14 d), the biomass is brownish with ~45% lipid content. Ho et al. [103] have applied nitrogen starvation to Scenedesmus obliquus and observed that the highest lipid content was 22.4% on day 5. Agirman and Cetin [104] have investigated the development of Scenedesmus acutus and changes in its protein and lipid contents under nitrogen stress. According to their results, at <50% nitrogen limitation, the lipid content was 19.48% higher than that of the control group. In addition, an inverse correlation was observed among cell development, lipid content, and nitrogen concentration.

Nitrogen starvation triggers several reactions in microalgae from the decomposition of nitrogen-containing compounds, such as proteins, chlorophyll, and DNA, to the accumulation of energy-rich compounds, such as carbohydrates [72]. The critical point here is that nitrogen-rich chlorophyll supports biomass growth and cellular development. Microalgae synthesize chlorophyll in huge amounts when nitrogen is plentiful and within their reach; however, when the opposite is true, they begin to use chlorophyll to obtain free nitrogen [101]. Because chlorophyll is green in microalgae, its decomposition results in microalgae becoming first greenish yellow and then brown when nearly all chlorophyll has been decomposed [32]. Wu et al. [105] have reported that in Scenedesmus sp., the concentration of nitrogen and phosphorus is highest during the exponential growth phase and lower during the stationary phase. Similarly, Wang et al. [106] have set up a
photobioreactor that is open to the atmosphere, and under nitrogen starvation, observed that over a 10-d period, the protein content in \textit{Scenedesmus sp.} decreased from 33 to 10% and the total lipid content increased from 8 to 37%.

4. CONCLUSIONS

Microalgae are promising candidates for replacing fossil fuels because of their rapid growth, uniquely highly efficient solar energy conversion, sustainability and renewability, high lipid content, and high potential for biofuel production. Microalgae use has been increasing as feedstock for biofuel in response to the energy crisis. In particular, the energy stores in microalgae are the raw materials for the biodiesel used in the transportation sector, and enhancement of microalgae’s lipid content and its subsequent production have gained considerable attention among researchers. This review has illustrated that limiting nitrogen in the culture of high lipid-accumulating \textit{Scenedesmus sp.} is a feasible strategy by which to increase lipid production.

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