Vanadium, Ruthenium and Copper Compounds: A New Class of Non-platinum Metallodrugs with Anticancer Activity

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Abstract: Cancer is a group of diseases involving abnormal cell growth. The cells grow uncontrollably with the potential to invade and spread to other parts of the body. This disease is one of the principal death causes in the world, thus becoming a significant topic of scientific research. On the other hand, transition metals play a fundamental role in different living systems. In particular, Metallodrugs represent new and powerful tools for diverse therapeutic applications. To date, various metallodrugs display interesting biological activities for chemotherapy. In this field, cisplatin was the first inorganic compound with high relevance in cancer treatment. This compound was a leader agent in clinical use. Toxicity and resistance problems trigger the development of other platinum drugs with better clinical perspective and also raise the scientific interest for the putative antitumor properties of V, Ru and Cu compounds. Several scientific articles show that complexes of these metals are the new metal-based drugs used in the treatment of several cancers, such as, lung, colon, breast, bladder, etc.

In this review we recapitulate current information and new advances on antitumor in vitro effects of several organic and inorganic compounds derived from copper, ruthenium and vanadium. These metal derived compounds targeting DNA or cell proteins involved in cell signaling pathways related to cancer. The mechanisms of cell death of these metallodrugs have also been comprehensively reviewed.

The knowledge of these mechanisms of death and the relationship between chemical structure and biological activity may be useful for the design of new metal-based drugs with promising pharmacologic applications as anticancer agents.

Keywords: Anticancer agents, vanadium, ruthenium, copper, cell in culture, mechanism of action.

1. INTRODUCTION

Metal ions regulate a vast array of essential cellular processes with high specificity and selectivity. The ability of some metal ions to prevent and cure cancer in humans has since long been known, with early documented cases dating back to the 16th century [1]. In this sense, medicinal chemistry has arisen as a field covering the design and synthesis of novel drugs, based on knowledge related to their function at the molecular level. This new prospect enabled the development of metal coordination compounds (a) acting as useful metallodrugs, which interact with a specific molecular target (pharmacodynamics), and (b) being capable of reaching that target (pharmacokinetics). Medicinal Inorganic Chemistry has arisen as a field covering the design and synthesis of novel drugs, based on the knowledge related to the function of metals at cellular and molecular levels. This new perspective enabled the...
development of metal coordination compounds (a) acting as useful metallodrugs, which interact with a specific molecular target (pharmacodynamics), and (b) increasing through coordination the possibility to reach the cellular and molecular targets (pharmacokinetics).

The extensive range of coordination numbers, physicochemical properties, redox states and geometries, of the metal ions and ligands enable different reactivities that are useful tools for research in this field. Considering these facts, Inorganic Biochemistry presents good opportunities for the design of appropriate therapeutic agents [2,3]. Furthermore, this area has taken advantage of carbohydrate derived ligands to increase the solubility and molecular targeting of metallodrug candidates. As a result of such activity, a significant number of anticancer metal based drugs have emerged in the last years [4-6]. Outstanding such metallodrugs are platinum complexes. It is worthy to mention the success of cisplatin in medicine for the treatment of several kinds of tumors. Its use positioned the metallo-drugs in an interesting place for cancer treatment [3, 7]. Cis-[Pt(NH$_3$)$_2$Cl$_2$] (cisplatin) has been used in a significant number of cancer with high incidence in society but it is usually combined with radiation or other medication. Different metallodrugs that have been used for a cancer treatment have a similar mechanism of action of cisplatin and the design and synthesis of these are based on structural similarities with cisplatin.

On the other hand, serious side effects and resistance limit the therapy with cisplatin [8]. New discoveries in drug design try to reduce toxicity and also open the range of tumors to be treated with some compounds [9,10]. In fact, the disadvantage of cis-Pt has been partially ameliorated by the use of new platinum drugs such as carboplatin and oxaliplatin [11]. Cancer has been and remains to be considered yet as one of the fatal diseases worldwide. Cancer is the general name of any malign neoplasia and involves more than 100 diseases which share an uncontrolled growth of cells that invade organs and tissues placed even at a distance from the original tumor (metastasis) [12]. Cancer cure requires deep knowledge of the processes initiating and developing the chemical and biological phenotypes of the disease. The main features of cancer comprise several biological skills that involve the capacity of evading growth suppressors, resisting cell death, inducing angiogenesis, invasion and metastasis, and evading immune system [13]. The study of these concepts is expected to increase and stimulate the design and synthesis of new metal based drugs with anticancer properties and development of alternative therapies.

Among the ten most active metal derived drugs it is worthy to discuss vanadium compounds [14]. In the case of vanadium, notable is the presence of vanadocene, a promising anticancer agent belonging to the family of metallocenes [15]. However, other forms of vanadium, extending from simple inorganic salts to more defined coordination compounds involving organic and inorganic ligands have also been proposed as anticancer agents [16,17]. Future work will help to decide on their applicability in the fight against cancer, based on the strict chemical criteria governing solubility, bioavailability and molecular specificity at the sub-cellular level. Other interesting metal based drugs with potential anticancer properties are ruthenium derivatives [18,19]. Ruthenium is particularly attractive as the ligand exchange kinetics in its complexes can be similar to those of platinum complexes [20]. Several scientific reports show the interaction of ruthenium compounds with different molecular targets such as DNA, mitochondria, among others [21,22]. Another metal interesting pharmacological properties and different targets for cancer treatment is copper [23]. Copper is an endogenous metal with important functions. It is known that these kinds of components may be less lethal for cells with normal phenotypes than for tumor cells. Nevertheless, copper can also be toxic due to its affinity for binding sites and redox effects. The transformed metabolic processes of tumors provoke differential responses to copper compounds comparing to normal cells [6]. In this present survey, the state of knowledge referred to copper, ruthenium and vanadium derivatives with potential anticancer effects has been reviewed.

2. ANTITUMOR ACTIVITY OF VANADIUM COMPOUNDS

In the last years, the antitumor activity of several vanadium derivatives has been remarked, but the mechanisms of action are not well understood. The principal molecular targets for the anticancer effects of vanadium compounds are the disruption of cellular metabolism through the generation of ROS, the depletion of GSH, the alterations of cellular organelles, the spindle key proteins such as cyclins, cytoskeleton proteins, some signal transduction pathways and caspases, which in turn play a role in cell cycle arrest and programmed cell death.

2.1. Vanadocenes

Within metallocenes field, one of the most important metal based drug are titanocenes and derivatives. The anticancer activity of titanocene dichloride was widespread in the early of 1980. Then, this compound
was studied in phase I clinical trials in 1993 [24] and while phase I clinical trials were not as satisfactory, several phase II clinical trials with patients with breast and a renal tumor have been carried out detecting a very low activity [25]. Nevertheless, in the last ten years, different works of many groups showed that these kinds of metals exhibited anticancer activity toward several types of tumor such as prostate [26], cervix, melanoma, colon and lung, and these results renewed the interest in this field [27-29].

The biological results showed that titanocene Y (bis-[p-methoxybenzyl)cyclopentadienyl] titanium(IV) dichloride) and C (Bis-(N,N-dimethylamino-2(N-methylpyrrolyl) methylcyclopentadienyl] titanium (IV) dichloride) are the most promising metallotherapeutics as anti-tumor agents and in particular show promise for use in combination therapies. The main mechanism of action of these compounds involved to induce apoptosis in a dose-dependent manner, induce DNA damage and a differential damage response and reduced the clonogenic capacity of the cells without affecting the cell cycle progress in prostate tumor cells. Besides, Titanocene Y activated cell signaling affected c-Jun N-terminal kinase and it increased the phosphorylation levels of adenosine monophosphate (AMP)-activated protein kinase α1 (AMPKα1) in small lung cancer cells. On the other hand, Titanocene Y triggered apoptosis in leukemia cells, regardless of the expression of anti-apoptotic Bcl-2 and pro-apoptotic smac. Moreover, the titanium compounds are also effective against leukemia cells that are multidrug resistant due to over-expression of P-gp [30].

Other interesting group of compounds with antitumor properties within metallocenes field are vanadocene derivatives [31]. Vanadocene, is a metallocene, belonging to a group of organometallic derivatives, with a metal ion sandwiched between two cyclopentadienyl rings. The first vanadocene that showed relevant pre-clinic results was the bis(cyclopentadienyl) dichloro-V(IV), vanadocene dichloride, [VCp2Cl2] (1), [32]. This compound exhibits a higher in vitro activity against tumor cell lines upon direct comparison with [TiCp2Cl2] (2) (see Fig. 1). Besides, in vivo studies revealed that vanadocene compounds exhibit significant antitumor properties with vanadocene dichloride being one of the most promising among metallocenes [33]. Recently, speciation studies have shown that [VCp2Cl2] transforms at physiological pH to [VCp2(OH)2] and carbonate, oxalate, lactate, phosphate are able to displace the two OH- ions to yield the adducts with these ligands [VCp2(ox)], [VCp2(CO3)], [VCp2(lactH-1)], and [VCp2(HPO4)] [34]. Several vanadocene derivatives, especially the methyl-and methoxy-substituted vanadocene dichlorides showed cytotoxic effects against T-lymphocytic leukemia cells with MOLT-4 evidencing more activity than their corresponding titanocene analogs. Activation of apoptosis by vanadocene compounds is very similar to the mechanism used by cisplatin, as it triggers DNA injury and involves p53 activation [35]. p53 is a tumor suppressor factor that normally functions in apoptotic mechanisms, cell cycle regulation, and maintenance of genomic stability.

Fig. (1). Schematic structures of vanadocene dichloride (1) and, titanocene dichloride (2).

On the other hand, vanadocenes are efficient agents against human testicular cell lines [36]. Vanadocene compounds containing phenanthroline (phen) ligands are promising chemotherapeutic agents, mostly due to their high anticancer activity, solubility and stability properties [37]. In recent years, scientific reports have shown the design and synthesis of new benzyl-substituted vanadocenes, including 3, 4 and 5 which were modeled after bis-[5-(p-methoxybenzyl) cyclopentadienyl] titanium(IV) dichloride (Titanocene Y) and its derivative compound (6) (Fig. 2). The three vanadocene caused slightly better activity than cis-platin in several cancer cell lines [38]. Besides, other novel compounds such as benzyl- substituted vanadocenes with p-methoxybenzylcyclo-pentadienyl, bis-[(3,4-dimethoxybenzyl)cyclo-penta-dienyl] and bis-[(3,4,5-trimethoxybenzyl)cyclo-penta-dienyl] showed high toxicity on pig kidney epithelial cell line [39].

2.2. Vanadium IV and V Complexes

Another interesting class of vanadium compounds with potential application in cancer treatment is formed by some oxidovanadium(IV) complexes. One of the most important and promising example in this area is bis(4,7-dimethyl-1,10-phenanthroline) sulfatooxovanadium (IV) Metvan (Fig. 3). This complex induces cell damage through apoptosis activation in several human cancer cell lines such as multiple myeloma cells, leukemia cells, and solid tumors derived from breast, ovarian, prostate and testis [4,40,41]. One of the main therapeutic advantages of Metvan is the high effectiveness toward ovarian and testicular cancer cell resistant
Fig. (2). Schematic structures of bis(η5-(2-methoxybenzyl) cyclopentadienyl)-vanadium(IV) dichloride (3), bis(η5-(3,4-dimethoxybenzyl) cyclopentadienyl)-vanadium(IV) dichloride (4), bis(η5-(3,4,5-trimethoxybenzyl)cyclopentadienyl)-vanadium (IV) dichloride (5), and bis(η5-(3,4-dimethoxy-benzyl) cyclopentadienyl)-titanium(IV) dichloride (6).

Fig. (3). Schematic structure of bis(4,7-dimethyl-1,10-phenanthroline) sulfato oxovanadium(IV) “Metvan”.

to cisplatin. On the other hand, Metvan shows promising anticancer actions on glioblastoma and breast cancer in invivo models. The wide range of Metvan antitumor activity with auspicious pharmacodynamics features and its low toxicity highlight that this compound has potential to be the first vanadium complex as an alternative to the classical platinum treatment [42]. Flavonoids are a group of natural polyphenols predominantly synthesized by higher plants with numerous pharmacological properties. The main therapeutic actions of flavonoids include antiviral, anticancer, antibacterial and antioxidative activities [43]. On the other hand, metal complexation has an important role in the improvement of the pharmacological activities of metal ions. The anticancer properties of a series of flavonoids (quercetin, hesperidin, morin, silibinin, chrysin) and their complexes with oxidovanadium(IV) were investigated against normal (MC3T3E1) and tumor (UMR106) osteoblasts-like cells [44-46]. Table 1 highlights the effects of flavonoids and VO-flavonoids complexes against normal (MC3T3E1) and tumor (UMR106) osteoblasts-like cells. The comparison was performed at 100 µM concentration. As it can be seen, hesperidin, quercetin and silibinin only altered slightly normal osteoblasts whilst chrysin produced a cytotoxic effect on proliferation of these cell lines and this effect could be probably attributed to structural differences due to the lack of substituents on B ring of the cinnamoyl system. In view of these previous results, VOsil and VOchrys (Fig. 4) have been thoroughly investigated in the human osteosarcoma cells (MG-63). This cell line is an excellent model to study the effects of potential anticancer drugs for osteosarcoma treatment. In human osteosarcoma cells, oxidovanadi um(IV), chrysin and VOchrys impaired cell viability in a concentration-dependent manner. Moreover, VOchrys was the strongest antitumor drug in human osteosarcoma cells having a lower IC50 value (16 µM) than vanadyl cation and chrysin (>100 µM) [47]. In addition, in the same cell line, VOsil, decreased the cell viability in a dose dependent manner with a greater potency than silibinin and vanadyl cation. The complex also displayed a concentration effect both in cyto- and genotoxicity processes. Investigation of the redox status of the cells was determined through the ROS level and GSH/GSSG ratio. These parameters were the most representative for the events involved in the deleterious effects of the complex on human osteosarcoma cells. Moreover, the compound induced cell cycle arrest (G2/M phase) and the activation of caspase 3 that conveyed the cells to apoptosis [48]. From the comparison of the anticancer properties of VOchrys, VOsil and cisplatin in MG-63 cells, it could be concluded that VOchrys was the strongest antiproliferative effects (see Table 2). Table 2 gives IC50 values of VOchrys, VOsil and cisplatin in MG-63 cell line. ROS generation, alterations of the GSH/GSSG ratio, changes in the mitochondria membrane potential (MMP), induction of caspase-3 and DNA damage are the main mechanisms involved in the antitumor activity of VOchrys and VOsil.

All these mechanisms finally trigger the human osteosarcoma cells to apoptosis. Besides, VOchrys and VOsil showed promissory anticancer effects against human colon adenocarcinoma cells. These vanadium compounds caused cytotoxicity and genotoxicity in a concentration dependent manner. The main mechanisms of action of VOchrys involved a decrease in GSH/GSSG ratio and cell cycle deregulation whilst VOsil employed several mechanisms that include a decrease in GSH, programmed cell death activation,
Table 1. Effects of flavonoids and VO-flavonoids complexes against normal (MC3T3E1) and tumor (UMR106) osteoblasts-like cells.

| Flavonoids | MC3T3E1 (%) basal (100 µM) | UMR106 (%) basal (100 µM) | Vanadyl and VO-flavonoid complexes | MC3T3E1 (%) basal (100 µM) | UMR106 (%) basal (100 µM) |
|------------|---------------------------|---------------------------|-----------------------------------|---------------------------|---------------------------|
| Hesperidin  | 90                        | 85                        | VOhesp                            | 20                        | 20                        |
| Quercetin   | 95                        | 78                        | VOquer                            | 60                        | 82                        |
| Silibinin   | 94                        | 60                        | VOsil                             | 46                        | 33                        |
| Chrysin     | 65                        | 40                        | VOchrys                           | 50                        | 42                        |
| Morin       | 70                        | 65                        | VOmor                             | 30                        | 65                        |

Fig. (4). Schematic structures of oxidovanadium(IV) complexes with chrysin (VOchrys) and silibinin (VOsil).

Table 2. IC\(_{50}\) values of VOchrys, VOsil and cisplatin on MG-63 cell line.

| Compounds  | IC\(_{50}\) |
|------------|------------|
| VOchrys    | 16 µM      |
| VOsil      | 74 µM      |
| Cisplatin  | 43 µM      |

The attenuation of NF-kB pathway among others [49]. Another interesting group of vanadium based drugs complexes of oxidovanadium(IV) with ligands that hold multiple donor atoms is able to coordinate with metal centers.

Chelating ligands are relevant in living systems since they can facilitate the uptake and transport of metallo-drugs inside the cells. Oxodiacetate (oda), O(CH\(_2\)COO)\(_2\), is a very versatile ligand that coordinates metal ions by forming chelate rings [50]. V\(^{IV}\)O-complexes of oda, V\(^{IV}\)O(oda), as well as V\(^{IV}\)O (oda)(bipy) and V\(^{IV}\)O(oda)(phen) (Fig. 5), showed significant effects in osteoblast like cells. The three complexes were tested in two cell lines derived from osteoblasts (MC3T3-E1 from mouse calvaria and UMR106 from a rat osteosarcoma cells). V\(^{IV}\)O (oda) inhibited cell viability in both cell lines, but the inhibitory effect was stronger in the normal than in the tumor osteoblasts [51]. The principal mechanisms of action involved in the anticancer activity of V\(^{IV}\)O(oda) are ROS generation, depletion of GSH, disruption of the mitochondria membrane potential (MMP) and activated ERK cascade phosphorylation [51]. Besides, V\(^{IV}\)O(oda)(phen) impaired cell viability in both cell lines (MC3T3-E1 and UMR106), but the cell damage was stronger in the normal than in the tumor cells [52]. On the contrary, V\(^{IV}\)O(oda)(bipy) showed promissory antitumor activity since the cytotoxicity was stronger in the tumor than in the normal cells [53]. Recent reports have shown that the three complexes caused a concentration dependent inhibition of cell viability in human osteosarcoma cells in culture [54]. In this order, V\(^{IV}\)O(oda)(phen) produced strong cytotoxicity affecting several organelles such as lysosomes and mitochondria from 2.5 µM while V\(^{IV}\)O(oda) and V\(^{IV}\)O(oda)(bipy) only caused these effects at higher concentrations (100 µM). Moreover, complex with phen and bipy derivatives triggered apoptosis as a relevant mechanism of cell death. Besides, the nuclease activity of the three compounds revealed that DNA cleavage caused by V\(^{IV}\)O(oda) and V\(^{IV}\)O(oda)(bipy) was similar, whilst V\(^{IV}\)O(oda)(phen) showed a stronger effect. In this way, a very interesting relationship between the bioactivity of the complexes and their struc-
Fig. (5). Schematic structures of oxidovanadium(IV) complexes with oxodiacetate VO(oda), oxodiacetate and 2,2-bipiridine VO(oda)bipy, oxodiacetate and 1.10 phenanthroline VO(oda)phen.

Figures was achieved. V\textsuperscript{IV}O(oda)(phen) showed the most potent anticancer action in human osteosarcoma cells followed by V\textsuperscript{IV}O(oda)(bipy) and subsequently by V\textsuperscript{IV}O(oda)) in agreement to the number of intercalating heterocyclic moieties [54]. Others organically chelated vanadium compounds also showed interesting anticancer activity against lymphoma derived cells. Quinoline and pyridinone molecules are attractive ligands because of their pharmacological activity. The antitumor actions of quinoline complexes on U937 cells showed slight relation with the length and shape of the alkyl chain in the relationship structure-activity analysis [55]. One of these compounds showed stronger antitumor effects than cisplatin. Moreover, this complex activated apoptosis but through a pathway independent of caspase activation [55]. Besides, hydroxiquinoline derived vanadium complexes displayed specifically anticancer activity against cisplatin sensitive/resistant ovarian cells (A2780/A2780cisR). The bioactivity of these vanadium compounds toward the A2780 cell line was dependent on incubation time and concentrations showing IC\textsubscript{50} values in the range of 3-14 \textmu M. In this order, these compounds were significantly more active than cisplatin (22 \textmu M), in the A2780 cells and in the cisplatin-resistant cells A2780cisR (4-8 \textmu M vs. 75.4) [56]. A huge family of vanadyl compounds also includes bound organic ligands based on the salen ligand. A pyridoxal-based vanadium(IV) complex showed selective cytotoxicity for tumor cell lines in contrast with healthy cells. The complex impaired cell viability was stronger in melanoma and lung carcinoma cells than in normal epidermal keratinocytes, lung cells and peripheral blood mononuclear cells [57]. Oxidovanadium (IV) complexes with a Schiff base and thiosemicarbazones as mixed-ligands showed interesting antitumor effects on several colon cancer cell lines (HTC-116, Caco-2, and HT-29) and also with non-cancerous colonic myofibroblasts (CCD18-Co). In general, these compounds exhibited antitumor activity on cancer cells, but did not affect normal cells. Nevertheless, compounds 8-10 (Fig. 6) showed less inhibitory effects on CCD-18Co cells, suggesting a possible cytotoxic selectivity against colon cancer cells and may have a potential in chemotherapy (see Table 3) [58]. Table III shows Antitumor activity of compounds derivatives 8-10 and cisplatin, on different colon adenocarcinoma cell lines after 24, 48, and 72 h of treatment. Data are expressed as IC\textsubscript{50} (\textmu M). IC\textsubscript{50} values are shown as mean ± S.D. obtained from three independent experiments. Others organic vanadium complexes with pharmacological activities are acetylacetonate derivatives.
cancer activity of the complex [60]. On the other hand, several V\(^{V}\)-compounds were reported to exhibit antitumor properties in different experimental models [4,14,16]. The development of V(V)-peroxide species in binary as well as ternary combinations emerged as a promising approach for novel cancer therapy. To this end, different V(V)-peroxide complexes have been designed and synthesized as potential antitumor compounds. The most common form of a bis(peroxido)vanadium complex is \( M[VO(O\text{O}_2)]L \) \( n \cdot xH_2O \) and that of a mono(peroxido)vanadium complex is \( M[VO(O\text{O}_2)L] \) \( n \cdot xH_2O \). In peroxidovanadate complexes, the peroxido ligands are bound to the vanadium center almost exclusively in a side-on fashion in which the geometry around vanadium is octahedral or pentagonal bipyramidal, with varying numbers of water molecules or hydroxo ions. In vitro assays showed that peroxidovanadate complexes displayed inhibitory effects of protein phosphotyrosines suggesting that the effects of protein phosphotyrosines are one of the key mechanisms of action of peroxidovanadate complexes as antitumor agents [61]. V(V)-oxido and V(V)-oxido-peroxide compounds with carboxylic acids \( K_2[VO(O\text{O}_2)(C_2O_4)_{2}] \cdot 1/2H_2O \) and \( K_2[VO_2(C_2O_4)_{2}] \cdot 3H_2O \) containing oxalate ligands showed interesting anticancer activity against human osteosarcoma cells. In this way, the monoperoxide complex showed a very strong antitumor action, whereas the dioxide compound was inactive [62]. Moreover, bis(peroxido)vanadium(V) complexes like \( (\text{Pr4N})[VO(O\text{O}_2)(\text{ox})(\text{phen})] \) \( (\text{Vphen})\text{ox} = \text{oxalate} \) (2-) and \( \text{Pr4N} += \text{tetra(n-propyl)ammonium} \), promote DNA cleavage in human lymphocytes in a concentration-dependent manner [63]. Further screening of probable anticancer activity on a murine leukemia cell line (L1210 cells) showed that they exhibited significant antitumor activity in a concentration and exposure time-dependent manner [63]. Several peroxide species were investigated in order to identify correlations of organic ancillary ligands with their related structures and the relationship to the physiological and antitumor effects. The nature and denticity of employed ligands were variable adopting a pentagonal bipyramidal geometry around the vanadium ion in these compounds. This geometry is typical for a monoperoxido V(V) species with an organic ancillary ligand. In addition, half-maximal inhibitory concentration (IC\(_{50}\)) values in different cancer cell lines could be related to the transition energies (LMCT) of the peroxide group, especially among complexes with analogous ligands. Djordjevic proposed that intramolecular electron transfer between the peroxide group and the metal center would affect the anticancer activity of the peroxidovanadium species and these result were in agreement [29]. It is suggested that ROS generated through intramolecular electron transfer in V(V)-peroxide species, may be one of the mechanisms of action responsible for the observed cytotoxicity. V(V)-peroxide species bearing variable ligands show different antitumor actions and these effects may be ascribed to the distinct electronic contribution of the bound peroxide moiety. Therefore, cytotoxicity and physiological actions of V(V)-peroxide species, correlating structure and activity, can ride

### Table 3. Antitumor activity of compounds derivatives 8–10 and cisplatin, on different colon adenocarcinoma cell lines after 24, 48, and 72 hours treatment. Data are expressed as IC\(_{50}\) (µM). IC\(_{50}\) values are shown as mean ± S.D. from three independent experiments.

| Compounds | HT-29  | Caco-2  |
|-----------|--------|---------|
|           | 24 h   | 48 h    | 72 h   | 24 h   | 48 h    | 72 h    |
| 8         |        |         |        |        |         |         |
| 277.1±10.2| 239.6±7.9| 100.3±5.1| 349.1±19.5| 277.1±16.3| 147.1±10.8|
| 9         |        |         |        |        |         |         |
| 244.2±8.8 | 169.6±15.1| 87.9±0.5| 367.0±21.5| 281.6±14.4| 152.5±12.7|
| 10        |        |         |        |        |         |         |
| 128.6±10.7| 82.4±9.6 | 47.8±5.5| 242.1±25.1| 166.2±15.3| 85.4±14.0|
| Cis-Pt    |        |         |        |        |         |         |
| 84.7±1.9  | 80.6±1.6 | 69.1±3.2| 32.0±1.6  | 22.8±1.5 | 17.9±1.8 |
| HCT-116   |        |         |        |        |         |         |
| 24 h      |        |         |        |        |         |         |
| 203.4±7.7 | 181.3±9.2 | 115.0±6.5| 495.6±23.5| 339.2±16.7| 208.0±11.9|
| 9         |        |         |        |        |         |         |
| 227.4±6.8 | 192.7±11.6| 110.3±10.1| 490.6±27.6| 329.5±15.4| 203.6±12.5|
| 10        |        |         |        |        |         |         |
| 161.3±4.6 | 134.9±3.9 | 89.5±14.5| 382.7±21.9| 246.2±11.4| 152.2±12.0|
| Cis-Pt    |        |         |        |        |         |         |
| 53.8±2.1  | 49.7±1.3 | 41.0±2.7 | 83.0±1.1 | 72.0±1.5 | 64.1±1.6 |
strongly on the nature and electronic properties of the auxiliary ligand, thus guiding the design and synthesis of new peroxidovanadium compounds that may be used as anticancer agents [64,65]. On the other hand, anthraquinone derivatives compounds exhibited specifically higher inhibition on human colorectal adenocarcinoma (HCT-8 cells) than the clinical anticancer drug Fu-5. Along the same lines, the anticancer activity of three different vanadium complexes bearing pyridine Schiff and pyrimidinone-based ligands (Fig. 7) was studied in two different cell lines. The study showed that the V(V)-species resulting from the V(V)-MHCPE are more toxic for tumor cells (HeLa) than for normal cells (3T3-L1 fibroblasts), thereby indicating potential anticancer activity [66,67]. Other relevant Schiff base with pharmacological properties is N,N'-bis(salicylidene)ethylenediamine, more commonly referred to as salen. VO-salen (Fig. 8) inhibited K562 leukemia cell proliferation in the range of 6-32 µM and the complex enhanced taxol anticancer activity. These conclusions support that the combined use of VO-salen and taxol might establish an effective novel approach against leukemia [68]. Semicarbazones are versatile ligands with therapeutic effects. Different semicarbazones and vanadium complexes were tested in three different human tumor cell lines derived from breast, colon and kidney cancer for then activity as potential anticancer drugs, showing selective antitumor activity on kidney derived cells. Results showed that changes in the semicarbazone moiety structure could have a relevant effect on the antitumor activity of the vanadium complexes [69]. Moreover, a complex of vanadium(V) with salicylaldehyde semicarbazone (V(V)-Salsem) (Fig. 9) exhibited anticancer effects on tumor osteoblasts through the production of ROS, the ERK pathway activation and the apoptosis induction [70].

2.3. Polyoxovanadate compounds

Polyoxometalates (POMs) are clusters with transition metal oxygen anion that contain combinations of metal cations (usually the d\textsuperscript{0} species V(V), Mo(VI), and W(VI)) associated by oxide anions. The structural, electronic and physicochemical properties (shape, stability under physiological conditions, polarity, redox potential, etc), make them attractive for different uses in diverse fields. POMs have pharmacological properties as antiviral and anticancer agents [71,72]. Polyoxovanadates are a prominent subclass of POMs that are relatively less investigated in comparison to the Mo and W compounds. These compounds show different topologies and structures such as [V\textsubscript{4}O\textsubscript{12}]\textsuperscript{4-}, [V\textsubscript{5}O\textsubscript{14}]\textsuperscript{6-} and [V\textsubscript{10}O\textsubscript{28}]\textsuperscript{6-} [73]. Other important features of polyoxovanadates are their diverse structural chemistry, combining tetrahedral (VO\textsubscript{4}), square pyramidal (VO\textsubscript{5}) and octahedral (VO\textsubscript{6}) units, a variability of oxidation states and the potential encapsulation of small molecules or ions.

![Fig. (8). Schematic representation of VO-salen.](image)

![Fig. (9). Schematic structure of vanadium(V) complex with salicylaldehyde semicarbazone (V(V)-Salsem).](image)

Polyoxovanadates form diverse anionic species in solution that can coexist simultaneously in equilibrium, like mononuclear [VO\textsubscript{4}]\textsuperscript{3-}, dinuclear [V\textsubscript{2}O\textsubscript{7}]\textsuperscript{4-}, tetranuclear [V\textsubscript{4}O\textsubscript{12}]\textsuperscript{4-} and decanuclear [V\textsubscript{10}O\textsubscript{28}]\textsuperscript{6-}. The pH of solutions is a relevant property to study the speciation of oxovanadates. In this sense, a decrease in pH increased the generation of the protonated forms. The pharmacological effects of polyoxovanadates include antiviral, antibacterial and antitumor activities. Scientific reports showed the antitumor activity of octacosaoidodecavane-dcavane-date(V)Na\textsubscript{5}Co(H\textsubscript{2}O)\textsubscript{3}V\textsubscript{10}O\textsubscript{28}18 H\textsubscript{2}O (CoV10) against liver and ovary cancer cell lines [74]. Moreover, octacosaoidodecavanate(V) diamonium derivatives, (H\textsubscript{2}men\textsubscript{2})V\textsubscript{10}O\textsubscript{28}·6H\textsubscript{2}O and (H\textsubscript{2}en)\textsubscript{3}V\textsubscript{10}O\textsubscript{28}·2H\textsubscript{2}O, decreased the cell viability showing a high anticancer activity in lung adenocarcinoma and leukemia cells [75]. The structures of these compounds are very similar (skeletal structure of octacosaoidodecavanate(V) anion and cation), however, the main

![Fig. (7). Schematic illustration of plausible forms of complex V(V)MHCPE.](image)
difference is the four methyl (CH3) substituents of cation in the former species. Therefore, the compound (H2tmen)3V10O28·6H2O has higher anticancer activity since its amphiphilic structure facilitates its permeation through the monolayer cell membrane. In the last year, several mixed-valence V(IV)/V(V) octacosaoxidodecavanadates were interestecompounds due to their structural, spectroscopic, nuclease activity and DNA-protecting actions. One of the more relevant compounds is (Me4N)6[V15O35Cl] that have important chemical interaction properties with alkylating agents [76]. Different DNA interaction studies showed the chemoprotective action of this compound toward alkylating agents such as diethylsulphate (DES) and dimethylsulphate(DMS) using pUC19 plasmid DNA. The main mechanism of action elucidated for this chemoprotective activity exposed the progressive generation of V(IV), V(V) and mixed-valence aggregates, suggesting mechanistically preferential transfer of the alkyl group to the vanadate instead of DNA [76]. One of the most important biomedical polyoxovanadates are the decavanadate derivatives [V10O28]6− that play a key role in many biological processes [77]. These compounds may act as effective ion pump inhibitors and have the potential therapeutic applications in the treatment of different diseases such as cancer, diabetes, ischemic heart disease, ulcers, etc. [78]. Recently scientific reports showed the antidiabetic actions of metforminium decavanadate (H2Metf)3[V10O28]8H2O in Wistar rats as biological model. This compound has therapeutic properties as a hypoglycemic, lipid-lowering and metabolic regulator [79]. Considering other promising biological roles of these kinds of compounds, several groups reported novel synthesis of different polyoxovanadates based on decavanate structure [80,81].

3. ANTITUMOR ACTIVITY OF RUTHENIUM COMPOUNDS

Historically, cisplatin discovery opened the perspective to use drugs with a metal core for cancer treatment, despite its good effects over tumor cells, it also showed a narrow therapeutic window, with severe and undesirable side effects on patients [1]. This safety issue with the cisplatin raised the number of investigations related to platinum and non-platinum metallodrugs in order to find substances capable of killing cancer cell but with better toxicological profile. Within the transition metal group, the platinum like elements have played an important role in the research and development of new active pharmaceutical ingredients (API). Among them, ruthenium (Ru) showed attractive chemical properties like its range of oxidation states (8, 6, 4, 3, 2, 0 and -2), atomic radius and the variable kind of ligands that can form complexes or bonds with Ru. Especially for the medicinal chemistry, the oxidation states 3 and 2 have a particular importance due to their stability under physiological pH and the equilibrium between those species make them suitable for potential pharmaceutical uses [82]. The ruthenium compounds with biological interest can be classified into the following main categories: coordination and organometallic ruthenium derivatives.

3.1. Coordination Compounds

As models of coordination compounds, the well-known and vastly explored NAMI-A, KP1019, KP418 and NKP-1339 were found. The main feature in these complexes is related to their six-coordinated sphere of coordination, achieving an octahedral geometry, where ligands containing soft nitrogen and sulphur atoms (e.g. imidazole, indazole and DMSO) are found in the axial positions and are the responsible of the steric and electronic properties of the complex [83,84]. Four axillary positions, occupied with ligands can be easily replaced or exchanged (e.g. chloride), depending on the environmental conditions (see Fig. 10). The interaction of these complexes with blood components in vitro and in vivo, proved strong relation between the ability of the complexes to bound to serum albumin and transferrin with their pharmacokinetic profile and the final therapeutic effect. Such proteins have a main role in transportation and storage of ruthenium complexes [85]. In the case of trans-tetrachloridobis(1H-indazole) ruthenate(III)], Jarosz et al. showed that the complex can bind to transferrin in apo and holo forms, out of the Fe binding site and is still recognizable by the transferrin receptor in the cell [86]. Some studies about ruthenium complexes reviewed by Pessoa and Tomaz demonstrated that the presence of transferrin-complex adducts enhanced the anticancer activity of the complexes [87]. This highlights the importance of this protein in the mode of action. The Ru-transferrin adduct reaches the tumor cell, almost in a targeted way, since it is known that cancer cells have a higher need of iron and it has been proved that this causes an overexpression of transferrin receptor on the cell surface [88-90]. This approach led to the development of this kind of complexes (protein adducts) as a “Trojan horse” (Fig. 11). The activation by reduction of Ru (III) to Ru (II) is the currently accepted hypothesis by which the complexes of Ru (III) act as a pro-drug (see Fig. 12). It is believed that the complexes once released from the transport proteins in zones with low pressure of oxygen (acidic
environment) are reduced to Ru(II) complexes which are more reactive and ready to interact with proteins (imidazole nitrogen from histidine side chains) or nucleobases (nitrogen 7 in the purine bases) into the cell [82,91]. The mechanism of action for these complexes actually goes beyond the DNA cross-linking and adducts formation previously investigated [92]. There is some evidence that this kind of compounds also cause a disruption in the mitochondrial and cellular redox balance by producing reactive oxygen species (Fenton-like reaction) and GSH depletion. Different carried out experiments suggested that the addition of ROS scavengers reduce the cytotoxicity of these complexes [93-96]. However, the most accepted mechanisms of action based on KP1019 and NKP-1339, are redox balance disruption, cell cycle arrest on G2/M, DNA synthesis blockage and finally apoptosis induced mitochondrial pathway [91]. As previously exposed, the ability to exchange ligands in the coordination sphere for the Ru complexes, and the possibility to induce modifications in their structure are the key factors to achieve a rational design of drugs and the establishment of structure activity relationship [97]. Since the mechanism of action of Ru (III) drugs is conservative in many compounds, the keystone in the investigation is more related to the characteristics of the ancillary ligands, aiming to improve pharmacokinetic behavior [98]. Another subfamily of the Ru coordination compounds is that composed of complexes containing at least one polypyridyl ligand (e.g. bipyridine or phenanthroline). The accepted mechanism of action of these drugs is the binding and cleaving of DNA inducing apoptosis in the cancer cells [99]. At normal physiological conditions, at least one of the polypyridyl ligands will remain inertly bound to the Ru center. This could be extended to the case in which every ligand in the complex is inert to hydrolysis reactions and therefore the coordination sphere of the metal remains saturated. In this last situation, the mechanism of the cytotoxicity should be different from that known for other metallopharmaceuticals (i.e. the activation by hydrolysis step is not viable).

3.2. Ru-Arene Complexes (Organometallic)

A second class of Ru related compounds, which have gained an important place in the research field of medicinal chemistry are the Ru-Arene complexes. The main concern about Ru (II) complexes is related to the stability and the ligand exchange rate. Nevertheless, Sadler and coworkers found out that arene moiety acts as a stabilizer of the Ru atom and increases the lipophilicity, which can increase the uptake by the cell enhancing the cytotoxic effect [100]. The Ru(II)-arene complexes have the general formula \([\eta^6 \text{- arene} \text{Ru}(X)(Y)(Z)]\) where X (e.g. Chloride) is a leaving group, and YZ are chelating ligands [101]. In general, the cy-

![Fig. (11). Schematic representation of the overexpression of transferrin receptor on tumor cells and the possible delivery mechanism and recognition mediated by transferrin-Ru adduct.](image)
totoxic effect increases in function of the size (lipophilic character) of the arene ligand, probably related to the capability to intercalate with the DNA. In this order, the bicyclic arenes showed higher rate of DNA damage compared with the single cycle arenes, and this effect is linked with the ability to form arene base π-π stacking (see Fig. 13) [102-104]. The ligands occupying the position (Y)(Z), have an important role in the antitumor activity since it was demonstrated that monodentate ligands have lower activity compared with chelated ligands in the inclusion of ethylenediamine, which led to an increase in the biological activity even on cisplatin resistant cell lines [105,106]. Studies varying the chelating ligand with N-N, N-O and O-O groups revealed that the anticancer activity of complexes with N-N moiety was stronger than N-O and O-O [107, 108]. These complexes should be in an active form, capable of interacting with the molecular target (mainly attributed to DNA). The most described activation mechanism is the aquation, where the leaving group (chloride or another halide) is substituted by a water molecule. \[(\eta^6 - \text{arene})\text{Ru}^2\text{Cl}_2(Y)(Z) \rightarrow [(\eta^6 \text{arene})\text{Ru}(H_2O)(Y)(Z)].\] In the bloodstream the complex with halide is mostly kept in an inactive form due to high chloride extra cellular concentration (c.a. 100 mM). However, when the compound enters to the cell, the Cl⁻ concentration is about 4 mM which facilitates the replacement of the leaving group by the water molecule [104,105,109]. The rational design of complexes allows to set up the rate and extend of hydrolysis based on the properties of the arene, leaving the group and the chelate ligand. This is a useful tool to control the selectivity and half-life of the complex [110].

3.3. RAPTA Compounds

This class of compounds have a stabilization mechanism mediated by arene moiety, the family of \([(\eta^6-\text{arene})(\text{PTA})\text{X}_2],\) PTA = 1,3,5-triaza-7-phosphaadamantane.

Chemically these compounds are configured as follows: three Ru coordination positions are occupied by the arene ligand, a position is filled by the PTA (phosphine) ligand which gives it a hydrophilic (water soluble) character and two positions with labile ligands (mainly chloride) which are available for aquation activation (see Fig. 14) [111]. These compounds have a low cytotoxic effect in vitro, but in further investigation it was found that they have a similar property to inhibit the metastasis process similar to the NAMI-A [112]. RAPTA-C suffers a rapid aquation driving to very high activation [109]. Modification of the core using bidentate ligands instead of Cl⁻ leads to an increase in the stability, solubility and aquation rate (carbo-RAPTA and oxalo-RAPTA). In relation to the molecular target of these complexes, it is very interesting that RAPTA-C shows a selective pH dependent damage of DNA on acidic environment (pH< 7). In this

Fig. (12). Scheme of the potential mechanisms of action and transport.

Fig. (13). Chemical structures of arene moiety sorted in decreasing order of anticancer activity.

Fig. (14). Representation of the structure of RAPTA.
order, scientific reports showed that DNA damage in tumor cells (acidic pH) is higher than in healthy cells (neutral pH). This effect can be attributed to the PTA ligand protonation at low pH, which is a more active form of the RAPTA-C compound [109]. The RAPTA complexes do not show selective in vitro binding to DNA, proteins and RNA [89]. But, this complex inhibited cell growth and division through G2/M arrest and apoptosis induction [113]. In Fig. (15), the first members of the RAPTA series of compounds are displayed. Dyson and collaborators made modifications in the arene ring and the PTA moiety to evaluate the structure activity relationship, these changes increased the stability and led to control the aquation process and enhance the cytotoxic and antimetastatic activity [111]. Nevertheless, Ru-Arene complexes overshadowed the investigations related to these complexes due to their better cytotoxic profile compared with the RAPTA family. Undoubtedly, the ruthenium complexes and compounds have proven be potential tools for the development of future therapeutics for cancer treatment. A development in Ru based drugs is an open door to a rational design where the flexibility to change the ligands led to find new cellular targets, enhancing the selectivity of the compounds. A great number of Ru compounds have been synthesized and their cytotoxic effects on several tumor cell lines have been demonstrated, despite the heterogeneity in their chemical structure. However, it should be noted that the effect of the lipophilic ligands, which in the most of the cases induced a strong anticancer effect on solid tumors, even with IC50 lower than those obtained with cisplatin. (see Table 4). Nowadays the action mechanism of Ru metallodrugs is quite unclear, but scientific studies pointed out some targets recognized for the different compounds (see Fig. 16) [83]. Table 4 shows the structure-activity on several tumor cell lines of different ruthenium complexes. [114-127].

3.4. Ru Complexes: Role in Photodynamic Therapy (PDT).

Among the practical applications of Ru complexes as alternative to the cancer treatment, beyond chemotherapy, photoactivated chemotherapy (PACT) and photodynamic therapy (PDT) have gained a notorious relevance in the last decade. According to SCOPUS® database in 2006, two documents related to Ru and photodynamic therapy were published, whilst in 2015, thirty-three scientific articles were published.

The photodynamic therapy (PDT) is the approach which mostly studied the cases of anticancer drugs activation through light which can be split into two categories: photoactivated chemotherapy (PACT) and photodynamic therapy (PDT).

The photodynamic therapy requires the presence of three agents: a photosensitizer (PS), light and molecular oxygen (O2). The first one is a molecule with the ability to be excited by light in the range of 650 to 800 nm. The activation yields an electron displacement to higher energy levels with the subsequent energy release as fluorescence emission, phosphorescence or a non-radioactive decay, called intersystem crossing. [128-130]

The excited PS can be present as singlet state and/or triplet state, and produce two types of reactions. The type I: derived from an excited singlet state involves the oxidation or the reduction of the substrates or even the photosensitizer itself to produce superoxide, hydroxyl and peroxides radicals which can start the cascade of cytotoxic effects leading to the cellular death. The type II: involves the presence of the excited triplet state, produced by the intersystem crossing, and the PS in this state can react directly with molecular oxygen. This reaction gives two forms of oxygen singlet (which are highly reactive oxygen species) that can directly attack biomolecules as amino acid residues, unsaturated lipids and nucleobases. Most studies have pointed out that the cell death is mainly mediated by the last one mechanism [131].

The classic model of photosensitizers can be found in the tetrapyrrrole ring (porphyrins, chlorins) which have been extensively studied in photochemistry. In this way, porphyrin derivatives were approved for human use in PDT for the treatment of several types of
Fig. (16). Summary of the putative action mechanism of Ru compounds.

Table 4. Structure, cell line, IC50 values of different ruthenium complexes.

| Structure | Cell line             | IC50 (µM)          | Reference |
|-----------|-----------------------|--------------------|-----------|
|           | A2780                 | 0.54 ± 0.1         | [114]     |
|           | MCF7                  | 5.4 ± 2.0          |           |
|           | HeLa                  | 3.6 ± 1.1          |           |
|           | J774                  | 1.9 ± 0.1          |           |
|           | 518A2                 | 94 (24h)           | [115]     |
|           | 518A2                 | 16 ± 3.4 (48h)     |           |
|           | HL-60                 | 11 ± 2.4 (24h)     |           |
|           | HL-60                 | 9 ± 1.4 (48h)      |           |
|           | HL-60                 | 9 ± 2.4 (24h)      |           |
|           | Kb-V1/Vbl             | 9 ± 1.4 (48h)      |           |
|           | Kb-V1/Vbl             | 6 ± 1.4 (48h)      |           |
|           | MCF-7/Topo            | 50 (24h)           |           |
|           | MCF-7/Topo            | 1.5 ± 0.3 (48h)    |           |
|           | 518A2                 | 45 ± 1.5 (24h)     | [115]     |
|           | 518A2                 | 13 ± 4.2 (48h)     |           |
|           | HL-60                 | 6 ± 4.2 (24h)      |           |
|           | HL-60                 | 3 ± 3.3 (48h)      |           |
|           | HL-60                 | 46 ± 5.6 (24h)     |           |
|           | Kb-V1/Vbl             | 20 ± 4.8 (48h)     |           |
|           | Kb-V1/Vbl             | 48 ± 5.8 (24h)     |           |
|           | MCF-7/Topo            | 11 ± 3.8 (48h)     |           |
|           | MCF-7/Topo            | 8 ± 3.8 (48h)      |           |
|           | 518A2                 | 31 ± 2.3 (24h)     | [115]     |
|           | 518A2                 | 13 ± 4.2 (48h)     |           |
|           | HL-60                 | 2 ± 1.1 (24h)      |           |
|           | HL-60                 | 0.8 ± 0.25 (48h)   |           |
|           | HL-60                 | 50 (24h)           |           |
|           | Kb-V1/Vbl             | 3 ± 0.7 (48h)      |           |
|           | Kb-V1/Vbl             | 50 (24h)           |           |
|           | MCF-7/Topo            | 0.2 ± 0.14 (48h)   |           |
|           | MCF-7/Topo            | 8 ± 0.14 (48h)     |           |
|           | 518A2                 | 60 ± 1.7 (48h)     | [115]     |
|           | HL-60                 | 95 ± 5 (48h)       |           |
|           | Kb-V1/Vbl             | 59 ± 3.1 (24h)     |           |
|           | Kb-V1/Vbl             | 3 ± 4.7 (48h)      |           |
|           | MCF-7/Topo            | 49 ± 3.9 (24h)     |           |
|           | MCF-7/Topo            | 29 ± 3.4 (48h)     |           |
(Table 4) contd….

| Structure | Cell line | IC$_{50}$ (µM) | Reference |
|-----------|-----------|----------------|-----------|
| ![Structure Image] | 3a | 518A2 | 3 ± 0.3 (24h) | [115] |
| | | 518A2 | 2.2 ± 0.2 (48h) | |
| | | HL-60 | 1.3 ± 0.02 (24h) | |
| | | HL-60 | 1 ± 0.2 (48h) | |
| | | Kb-V1/Vbl | 20 ± 4.6 (24h) | |
| | | Kb-V1/Vbl | 7 ± 1.5 (48h) | |
| | | MCF-7/Topo | 75 ± 13.7 (24h) | |
| | | MCF-7/Topo | 7 ± 2.7 (48h) | |
| ![Structure Image] | SiHa | 37.1 | [116] |
| ![Structure Image] | SiHa | 16.91 | [116] |
| ![Structure Image] | SiHa | 11.11 | [116] |
| ![Structure Image] | HeLa | 0.230 ± 0.053 | [117] |
| | HEP-2 | 0.226 ± 0.002 | |
| | 7T | 0.330 ± 0.037 | |
| | HCT 116 | 0.249 ± 0.010 | |
| | H460 | 0.300 ± 0.033 | |
| | MDA-MB-485 | 0.297 ± 0.015 | |
| ![Structure Image] | HeLa | 0.103 ± 0.006 | [117] |
| | HEP-2 | 0.108 ± 0.015 | |
| | 7T | 0.109 ± 0.006 | |
| | HCT 116 | 0.106 ± 0.009 | |
| | H460 | 0.117 ± 0.004 | |
| | MDA-MB-485 | 0.123 ± 0.018 | |
| ![Structure Image] | HeLa | 0.203 ± 0.025 | [117] |
| | HEP-2 | 0.180 ± 0.015 | |
| | 7T | 0.212 ± 0.020 | |
| | HCT 116 | 0.180 ± 0.001 | |
| | H460 | 0.208 ± 0.018 | |
| | MDA-MB-485 | 0.329 ± 0.174 | |
| ![Structure Image] | HeLa | 7.42 ± 0.01 | [118] |
| | A2780 | 9.65 ± 0.01 | |
| | A2780cisR | 12.95 ± 0.04 | |
| | HFL-1 | 18.48 ± 0.02 | |
### Table 4 contd.:

| Structure | Cell line | $IC_{50}$ ($\mu$M) | Reference |
|-----------|-----------|---------------------|-----------|
| ![Structure 1](image1.png) | HeLa, A2780, A2780cisR, HFL-1 | $2.55 \pm 0.01$ | [118] |
| ![Structure 2](image2.png) | HeLa, A2780, A2780cisR, HFL-1 | $11.42 \pm 0.02$ | [118] |
| ![Structure 3](image3.png) | HeLa, A2780, A2780cisR, HFL-1 | $3.65 \pm 0.03$ | [118] |
| ![Structure 4](image4.png) | HeLa, A2780, A2780cisR, HFL-1 | $7.98 \pm 0.01$ | [118] |
| ![Structure 5](image5.png) | HeLa, A2780, A2780cisR, HFL-1 | $0.84 \pm 0.25$ | [118] |
(Table 4) contd….

| Structure | Cell line | IC₅₀ (µM) | Reference |
|-----------|-----------|-----------|-----------|
| ![Structure 1](image1) | HeLa A2780 A2780cisR HFL-1 | 19.40 ± 0.03 6.13 ± 0.25 6.19 ± 0.01 49.40 ± 0.31 | [118] |
| ![Structure 2](image2) | HeLa A2780 A2780cisR HFL-1 | 6.87 ± 0.23 5.25 ± 0.03 4.24 ± 0.01 7.30 ± 0.03 | [118] |
| ![Structure 3](image3) | HeLa A2780 A2780cisR HFL-1 | 7.95 ± 0.03 6.97 ± 0.33 6.80 ± 0.05 6.47 ± 0.23 | [118] |
| ![Structure 4](image4) | HeLa A2780 A2780cisR HFL-1 | 2.26 ± 0.03 5.12 ± 0.03 1.33 ± 0.02 13.88 ± 0.03 | [118] |
| ![Structure 5](image5) | HeLa A2780 A2780cisR HFL-1 | 5.66 ± 0.01 9.47 ± 0.03 3.14 ± 0.01 4.27 ± 0.03 | [118] |
| Structure | Cell line | IC\textsubscript{50} (µM) | Reference |
|-----------|-----------|-------------------------|-----------|
|           | HeLa      | 7.88 ± 0.01             | [118]     |
|           | A2780     | 4.54 ± 0.01             |           |
|           | A2780cisR | 2.25 ± 0.03             |           |
|           | HFL-1     | 3.84 ± 0.01             |           |
|           | HeLa      | 11.15 ± 0.01            | [118]     |
|           | A2780     | 6.13 ± 0.02             |           |
|           | A2780cisR | 2.14 ± 0.04             |           |
|           | HFL-1     | 6.32 ± 0.02             |           |
|           | HeLa      | 2.69 ± 0.01             | [118]     |
|           | A2780     | 2.26 ± 0.03             |           |
|           | A2780cisR | 2.14 ± 0.04             |           |
|           | HFL-1     | 36.61 ± 0.48            |           |
|           | HeLa      | 6.81 ± 0.03             | [118]     |
|           | A2780     | 7.70 ± 0.12             |           |
|           | A2780cisR | 2.77 ± 0.01             |           |
|           | HFL-1     | 13.69 ± 0.01            |           |
|           | HeLa      | 1.72 ± 0.01             | [118]     |
|           | A2780     | 3.75 ± 0.02             |           |
|           | A2780cisR | 3.76 ± 0.03             |           |
|           | HFL-1     | 2.81 ± 0.03             |           |
(Table 4) contd….

| Structure | Cell line | IC$_{50}$ (µM) | Reference |
|-----------|-----------|----------------|-----------|
| ![Structure](image1) | A2780 A2780cisR HEK-293 | 113 ± 6 / 112 ± 16 / 232 ± 32 | [119] |
| ![Structure](image2) | A2780 A2780cisR HEK-293 | 195 ± 4 / 103 ± 10 / 247 ± 18 | [119] |
| ![Structure](image3) | A2780 A2780cisR HEK-293 | 132 ± 18 / 96 ± 6 / 90 ± 5 | [119] |
| ![Structure](image4) | A2780 A2780cisR HEK-293 | 19 ± 3 / 25 ± 1 / 20 ± 1 | [119] |
| ![Structure](image5) | A2780 A2780cisR HEK-293 | 69 ± 12 / 90 ± 7 / 61 ± 2 | [119] |
| Structure | Cell line | IC<sub>50</sub> (µM) | Reference |
|-----------|-----------|----------------------|-----------|
| ![Structure 1](image1) | SiHa | 37.10 (1) 16.91 (2) 11.11 (3) | [120] |
| ![Structure 2](image2) | CT26 A549 | 15.9 ± 2.4 21.0 ± 1.7 | [121] |
| ![Structure 3](image3) | CT26 A549 | 5.3 ± 1.1 3.8 ± 0.4 | [121] |
| ![Structure 4](image4) | CT26 A549 | 11.3 ± 2.0 13.4 ± 3.0 | [121] |
| ![Structure 5](image5) | CT26 A549 | 5.5 ± 0.90 6.5 ± 0.5 | [121] |
(Table 4) contd….

| Structure | Cell line         | $IC_{50}$ (µM) | Reference |
|-----------|-------------------|----------------|-----------|
| ![Structure 1](image1.png) | A549, MDA-MB-231, HeLa, MRC5 | 6.9 ± 2.9 | [122] |
| ![Structure 2](image2.png) | A549, MDA-MB-231, HeLa, MRC5 | > 100 | [122] |
| ![Structure 3](image3.png) | A549, MDA-MB-231, HeLa, MRC5 | 24.5 ± 0.1 | [122] |
| ![Structure 4](image4.png) | HeLa, Hep-G2, BEL-7402, A549, A549/CDDP, LO-2 | > 100, > 100 | [123] |
| ![Structure 5](image5.png) | HeLa, Hep-G2, BEL-7402, A549, A549/CDDP, LO-2 | 51.4, 69.2, 45.3, 79.2, > 100 | [123] |
Table 4 contd....

| Structure | Cell line | IC<sub>50</sub> (µM) | Reference |
|-----------|-----------|----------------------|-----------|
| ![Structure 1](image1.png) | HeLa, HeLa, Hep-G2, Hep-G2, BEL-7402, A549, A549/CDDP, LO-2 | 3.3, 5.6, 7.4, 3.7, 4.1, 10.2 | [123] |
| ![Structure 2](image2.png) | HeLa, MCF-7 | 21.0 ± 0.6 (<X=O>), 18.0 ± 0.5 (<X=S>), 6 ± 1 (<X=O>), 2 ± 1 (<X=S>) | [124] |
| ![Structure 3](image3.png) | Hep-G2, MCF-7, MO59J, GM07492A | 7.6 ± 0.9, 47 ± 6, 8 ± 2, 2 ± 1 | [125] |
| ![Structure 4](image4.png) | Hep-G2, MCF-7, MO59J, GM07492A | 5 ± 3, 9 ± 3, 6.5 ± 0.2, 6 ± 2 | [125] |
| ![Structure 5](image5.png) | SW480, LS174T | >200 (1), >200 (2), 41.4 ± 2.6 (1), >500 (2) | [126] |
| ![Structure 6](image6.png) | BEL-7404, A549, MGC80-3, HeLa, Hep-G2, BEL-7402, HL-7702 | 7.4 ± 0.4, 10.8 ± 0.2, 10.4 ± 0.1, 9.0 ± 0.6, 16.2 ± 0.7, 14.7 ± 0.3, 69.2 ± 0.3 | [127] |
cancer, macular degeneration, etc. In the Fig. (17), the structure of porfirmer sodium (Photofrin®) is presented which is a Haematoporphyrin derivative, used in the treatment of bladder cancer and non-small cell lung carcinoma [132].

Later, Schmitt compared the effect of the arene ligand derivation in monopyrlylporphyrin (mpp) and tetrapyrlylporphyrin (tpp) compounds, and also studied the effect of the isomer used (3-pp and 4-pp) (Fig. 19), in Me300 cell line. The cytotoxicity at 72h, without light stimulation revealed that monosubstituted porphyrins exerted a weak effect on cellular proliferation, while the tetrastitituted analogues demonstrated an IC_{50} value in the 10-20 µM range. This cytotoxic effect can be related to DNA synthesis inhibition. In relation to the photosensitizer behavior the isomer in the position 3 showed a better performance, since it needed low light radiation (0.5J/cm²) and low concentrations (5µM) to decrease the cell viability significantly. In conclusion they propose a molecule tetracoordinated with Ru-arene in the position 3 of the tetrapyrlylporphyrin moiety, as was proposed very interesting candidate for PDT[134].

Davia reported the synthesis of a compound porphyrin was functionalized in three positions with vainillin derivative, and the fourth position was coordinated with Ru-polypyridyl complexes. The results showed a high DNA binding rate for the compound and cell death at 10µM and activation through 60W tungsten bulb but importantly highlighted the lack of cytotoxic effects without light activation [135] (see Fig. 20).

A Ru porphyrin (RuP) compound increased the cytotoxicity of AY27 cell line (rat bladder cancer) at 20 µM through necrosis pathway. However, there is not a big difference between dark toxicity and photoactivated toxicity with 15.6J/cm² (17% and 14% cell viability compared to basal, respectively). Despite the generation of singlet oxygen the compound does not have good spectral properties to act as photosensitizer. Nevertheless, RuP has lower IC_{50} compared with cisplatin [136] (Fig. 21).

Pernot et al. evaluated the anticancer in vivo effects of two arene ruthenium porphyrin compounds (Rut1 and Rut4, see Fig. 22) on a mice model with ectopic human oral carcinoma xenograft. The results depicted that the presence of the Ru groups enhanced the photodynamic efficiency and Rut4 had a better performance. Associated with biodistribution high concentrations of Ru were found in the liver, kidney, spleen and plasma maybe due to the reticuloendothelial system elimination. Varying concentrations 1.45 and 1.50% of the administered dose were found in the tumor during 24 - 96h post injection. The subcellular distribution of the complex was found in the cytoplasm and not in the nucleus as expected for the Ru compounds [137].

The wide range of studies related to ruthenium chemistry found that some complexes have photo-
Gasser and co-workers [138] reported a pool of six complexes of Ru(II), 2,2'-bipyridine and dipyrido[3,2-a:2',3'-c]phenazine (dppz). Substituents as NH₂, OMe, OAc, OH, CH₂OH and CH₂Cl were added on dppz moiety (Fig. 23). The complexes showed a oxygen singlet quantum yield varying in the range of 50 - 94% and showed good stability in human plasma after 48h incubation. Cytotoxicity assays were carried out showing low effect on Hela (cervix cancer) and MRC-5 (normal phenotype) cell lines in dark (without light stimulation), however, when the cells were light-radiated (9.27 J/cm² at 420 nm), the IC₅₀ value ranged between 2.0 and 20.5 µM, for the complexes 1, 2, 4 and 6. The complexes 1 and 2 induced a DNA photocleavage effect at 10, 30, and 50 µM concentration. This effect can be correlated with high binding affinity of the complexes by the DNA.

The design and synthesis of three highly charged ruthenium(II) polypyridyl complexes were reported by Chao, Gasser and co-workers. The complexes demonstrated high phototoxic effect on 2D and 3D Hela cells models. Complexes Ru1, Ru2 and Ru3 (Fig. 24) had low dark toxicity with IC₅₀ valu ranged between 300 - 400 µM in the monolayer and greater than 500 µM in the 3D model. After ethe photoactivation the IC₅₀ values were 1.1, 2.2 and 2.9 µM in the monolayer system, and in the case of the spheroids for one photon PDT, the IC₅₀ values were 10.1, 15.4, 21.2 µM and 1.9, 5.4 and 7.2 µM for two photon PDT. Besides, the results showed that the complexes were up taken via endocytosis and suffer lysosomal accumulation [139].

Glazer et al. reported the synthesis of two ruthenium complexes with bathophenanthroline and batho
Fig. (22). Rut1 and Rut4 structures.

Fig. (23). Complexes of Ru(II) and 2,2'-bipyridine and dipyrido[3,2-a:2',3'-c]phenazine (dppz).

phenanthroline disulfonate (Fig. 25) and the effects of these complexes on jurkat, HL60 and A549 cell lines. The compound with bathophenantroline induced dark cytotoxicity (IC$_{50}$ ranged 0.62 to 3.15 $\mu$M) for three cell lines and even lower after irradiation with visible light. This complex was located at mitochondrial level, causing an impairment in the mitochondrial function leading to death mediated by necrosis.

Fig. (24). Ru1, Ru2 and Ru3: two photon activated photosensitizer.

The sulfonated analogue had a very low dark toxicity (IC$_{50}$ greater than 300 $\mu$M) while the photoactivation reduced the IC$_{50}$ value to a range of 3.3 to 17.3 $\mu$M. The main mechanism of action observed after light stimulation was apoptosis. In addition, both complexes exerted photocleavage of the DNA [140].

On the other hand, Fig. (26) shows the summary of the main mechanism of action involved in the antitumor properties of Ru compounds using in the PDT therapies.

Fig. (25). Ru-bathophenanthroline and Ru-bathophenanthroline disulfonate complexes.

4. COPPER COMPLEXES

Copper is an essential element associated with biological pathways since it acts as cofactor in most aerobic organisms [141]. Copper-based complexes have been studied because it has been shown that endogenous metals are less toxic for normal cells than for cancer cells. Moreover, normal and tumor cells respond differently to copper and this is the major development of copper complexes having antiproliferative characteristics.

Even though copper level in the human organism is strictly controlled, high serum and tissue levels of Cu were found in many types of human cancers including
breast [142], prostate [143], colon [144], lung [145] and brain cancer [146]. The reasons for this phenomenon are connected to the important function that copper plays in angiogenesis [147], which is essential for growth, invasion and metastasis [148]. Therefore, it has been claimed that copper chelators may have high therapeutic value acting as antiangiogenic molecules [149].

Several antitumor studies have postulated copper complexes as anticancer or anticarcinogenic agents in *in vitro* and *in vivo* systems [150-152]. In particular, it has been demonstrated that copper thiosemicarbazones have antitumor activity [153] and developments in this area are still in progress [154,155]. In fact, novel copper thiosemicarbazone complexes showed cell growth inhibition, apoptosis induction and cell cycle arrest on cisplatin-resistant neuroblastoma cells [155].

It has been shown that a dimeric copper complex of the unsubstituted pyridoxal thiosemicarbazone induces apoptosis and arrest the cell cycle significantly on two human lymphocytes (CEM and U937 cell lines), but it is inactive on K562 human lymphoblasts *in vitro* and on TLX5 lymphoma *in vivo*. However, copper complexes with N-substituted pyridoxal thiosemicarbazones did not produce inhibition on cell proliferation on the aforementioned cell lines [156].

Isolavones are a type of naturally occurring isoflavonoids which act as antioxidants because of their ability to trap singlet oxygen showing antitumor activity [157,158]. Copper(II) complexes showed moderate cytotoxicity against five human cancer cell lines, with IC$_{50}$ values in the 10–50 µM range [159] pointing out a benefit of the complexation of these ligands, since the complexes showed a stronger inhibitory effects than free isolavones or metal ions. Moreover, complexes provoked a significant G(2)/M phase arrest, which developed early apoptosis.

Hesperetin and apigenin are biologically active flavonoids which have been investigated as antitumor drugs against breast cancer and hepatoma cell lines [160]. Copper complexes with hesperetin and apigenin showed growth inhibition of SGC-7901 and HepG2 cell lines in comparison to the free ligands [161].

In a recent study, it was demonstrated that a Cu(II) complex with quercetin can cleavage plasmid DNA generating single and double DNA strand breaks, intercalate the DNA strand and increase the level of reactive oxygen species which produce oxidative DNA damage. Moreover, induction of apoptosis has been observed along with a decrease in the levels of survivin protein expression and increase in caspase-3 on A549 cells [162].

This study previously reported the biological actions of a copper(II) complex with santonic acid. The compound produced a cytotoxic effect on four cell lines (MC3T3-E1, UMR-106, Caco-2, TC7) displaying alteration in the nuclei and cytoplasm of treated cells [163].

Moreover, the lowering blood pressure drug valsartan coordinated with copper(II) through its carboxylate group giving rise to a binuclear compound, which showed antiproliferative activity toward two selected cell lines (normal MC3T3-E1 and tumor UMR-106) in the micromolar range, whereas valsartan itself had no effect up to 100 mM [164] (see Fig. 27 for a schematic representation of the complex).
Copper complexes have been demonstrated to show potential as cytotoxic drugs in 
*in vitro* assays, however, there is little information on the cellular mechanisms of 
action or the molecular targets implicated (see Fig. 28 for a schematic summary).

Previously, it has been shown that copper binds to DNA and interacts with DNA in a non-covalent way, 
through intercalation, electrostatic forces, and groove binding [6].

An example of the interaction of planar polycyclic containing copper compounds with DNA was shown 
for Casiopeinas™. These mixed-chelate complexes carrying aromatic substituted diimine (phen or bipy and 
substituted analogs) showed antineoplastic properties (Fig. 29) [165,166].

The authors described that Casiopeinas stacked between DNA bases produced a DNA breakup detected 
in either pUC19 plasmid or in HeLa cells [167,168]. Moreover, it has been demonstrated that only planar 
and unsubstituted ligands could intercalate into DNA.

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**Fig. (27).** Schematic representation of structure of copper(II) complex with valsartan.

**Fig. (28).** Schematic representation of molecular targets involved in antitumor activity of copper complexes.

**Fig. (29).** Schematic structures of CASIOPEINAS™ compounds.
[169-171] whereas nonplanar aromatic heterocyclic ligands did not show intercalation properties [172,173]. That is why physicochemical properties, such as the planarity play important roles in the mechanism of interaction with DNA [6]. On the other hand, Cu(II) complexes have been demonstrated to produce DNA oxidative cleavage due to their redox properties. Many copper complexes generated plasmid DNA cleavage through an oxidative pathway [170,174,175]. This phenomenon was observed through a Fenton-type reaction, where hydroxyl radicals [176] or different metal-based radicals were formed, producing oxidation and the breakup of DNA [177]. After DNA injury, many copper complexes trigger apoptotic cell death. Synthesis of proapoptotic or antiapoptotic proteins is deregulated as it was described for Cu(N9-ABS)(phen)$_2${[9]} (H$_2$N9-ABS = N-(9H-purin-6-yl)benzenesulfonamide) which induced a rise in p53 protein and a reduction of Bcl-2 expression in Caco-2 cells [178]. On the other hand, recent research has demonstrated that the inhibition of topoisomerases is another mechanism of action of copper(II) compounds. These enzymes regulate the overwinding or underwinding of DNA generating a transient DNA break which allows the DNA to be untangled [179].

Topoisomerase inhibitors have been postulated as important drugs for cancer chemotherapy treatments. A copper(II) complex of 2-furaldehyde oxime was found to injure topoisomerase activity by blocking the phosphorylation activation [180]. More recently, a copper salicylaldoxime, which effectively inhibited the L1210 leukemia cells, restrained topoisomerase dimmer formation with consequent induction of single-strand breaks in the DNA. A series of novel copper(II) complexes with quinolinone Schiff bases induced cytotoxicity and inhibition of DNA synthesis in HepG2 cells in a concentration and time-dependent fashion due to their topoisomerase inhibitory potential [181].

Copper complexes containing the planar heterocyclic phenanthroline showed that they can bind selectively to DNA by intercalation and electrostatic forces, and inhibit topoisomerase I, leading to apoptotic cell death in nasopharyngeal cancer cell line HK1 but not on normal NP69 cells [182].

Proteasome inhibition is another mechanism by which cells regulate many cellular functions, such as proliferation, apoptosis, angiogenesis, and metastasis formation [183]. An excellent anticancer strategy is targeting the ubiquitin–proteasome pathway, since cancer cells are more vulnerable to proteasome inhibition than normal cells [184], and currently, development of proteasome inhibitors is under exhaustive investigation. Several copper compounds have been investigated to selectively inhibit the proteasome in vitro and in vivo in human leukemia cells, however proteasome is not inhibited in non-transformed human natural killer cells under the same treatment [185]. Moreover, it has been established that certain types of organic ligands could bind to copper found inside the tumor cells and form proteasome inhibitors that lead to apoptosis [1]. Copper(II) complexes with asymmetric ligands containing the methylypyridin-amino-methylphenol moiety have been shown to inhibit the proteasome in vitro and in vivo [186]. These compounds were studied in prostate cancer cells resulting in an induction of apoptosis associated with a significant proteasome inhibition. It has been suggested that these copper complexes can cross over the cell membrane transporting the copper ion inside the cell [187]. It has been observed that ternary complexes binding with 1,10-phenanthroline reduce proteasome activity before induction of apoptosis in MDA-MB-231 human breast cancer cells, as well as in PC-3 human prostate cancer cells [188]. The hypothesis is based on the fact that phenanthroline transports copper into the cancer cell, leading to direct proteasome inhibition or oxidation of the proteasome.

5. NEW PERSPECTIVES: DRUG DELIVERY SYSTEMS FOR ADMINISTERING ANTICANCER RU, V, CU BASED DRUGS

Certain pharmacological issues of drugs such as bioavailability, site specific targeting, degradation and solubility may be improved by the application of different drug delivery systems. Nanotechnology is expected to address some of the most important problems of conventional anticancer chemotherapy. Nanodrugs are promissory in cancer chemotherapy because of their selective access to cancer cells and tissues [189]. Low efficiency of transport processes of active species to biological targets has also improved because nanotechnologies have been designed and developed. They act as vectors for metallodrug or simply as protectors of active species of the complexes for amplifying their activities and reducing their degradation [190, 191].

Numerous molecules and macromolecules have been employed as drug delivery systems for metallo-
drugs such as cyclodextrins, liposomes, lipid nanocapsules, proteins, carbon nanotubes, polymeric nanoparticles and ceramic materials [192].

Actually, copper complexes are being stabilized into nano-formulations for improving their biological activity. Copper complexes containing inorganic ligands (Cu(NH₄)₂Cl₄ and CuCl₂) were loaded on a functionalized titania to obtain drug delivery systems. They were tested as toxic agents on different cancer cell lines, as well as on cisplatin as a reference material. It was found that independently of the copper complex and also the cell line used, low concentrations of each copper compound were sufficient to kill almost 100% of cancer cells. Both copper complexes alone as well as those loaded on TiO₂ had higher toxic effect than cisplatin [193].

Moreover, it has been shown that multivalent copper(II)-conjugated phosphorus dendrimers and their mononuclear copper(II) complexes exerted potential antiproliferative properties against HCT116, MCF-7, OVCAR8 and U87 cell lines [194]. A direct association was found between the number of terminal moieties or the amount of copper complexed to the dendrimer and the growth inhibitory effect. These results demonstrated that the number of terminal moieties and Cu atoms increased cytotoxicity. Therefore, burdening copper complexes onto nanostructures improved the anticancer properties. However, there have only been a few reports on the nano-functionalization of anticancer copper complexes so far, which strongly promoted the investigation of different copper complexes as anticancer agents in nano-formulations.

On the other hand, molecular vectors were designed to transport ruthenium complexes to cells with an additional protection against physiological degradation during long circulation times. Since despite their high effectiveness, they are poorly stable in physiological conditions and are present short half-life times in aqueous media [192]. Three amphiphilic nucleoside-based ruthenium complexes, ToThyRu, HoThyRu and DoHu-Ru were combined with a new organometallic ruthenium complex, named AziRu and stabilized in POPC phospholipid formulations, which were stable for months and showed high in vitro antiproliferative activity [195]. Another efficient drug delivery strategy, warranting high ruthenium content within the formulation, presented long half-life in physiological media and enhanced cell uptake. Amphiphilic ruthenium anionic complexes, based on the formation of stable nanoparticles with the cationic lipid 1,2-dioleyl-3-trimethylammoniumpropane chloride (DOTAP) showed IC₅₀ values in the low µM range against MCF-7 and WiDr cells, thus, proving to be 10-20-fold more active than AziRu with a marked tendency to accumulate within, or in proximity of the nuclei (see Fig. 30 for a schematic representation) [196]. Moreover, Valente and co-workers [197] reported the synthesis of a D-glucose end-capped polylactide ruthenium cyclopenta-

Fig. (30). Schematic graphic of accumulation within nuclei of amphiphilic ruthenium anionic complexes, based on the formation of stable nanoparticles with the cationic lipid 1,2-dioleyl-3-trimethylammoniumpropane chloride (DOTAP).
Table 5. The most prominent V, Cu and Ru compounds as antitumor agents.

| Name                                      | Structure                              | Biological data                                                                                                                                                                                                                   | References |
|-------------------------------------------|----------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------|
| **Metal complexes under clinical investigation** |                                        |                                                                                                                                                                                                                                | [98]       |
| NAMI-A                                    | ![NAMI-A](image)                        | In a phase I study it was concluded that NAMI-A can be safely administered as a three hour i.v. infusion at a dose of 300 mg/m2/day for five days, every three weeks. In this study, one patient (total: 24 patients) with non-small cell lung cancer showed stable disease. |            |
| KP1019                                    | ![KP1019](image)                       | KP1019 showed promising results in a phase I clinical study. Five of six evaluable patients with solid tumours experienced stable disease. Only mild toxicities were observed and further investigation in phase II was suggested. In contrast to NAMI-A, KP-1019 was also active at the primary tumour site. | [93, 95]   |
| Casiopeína IIgly                          | ![Casiopeína IIgly](image)              | CasII-gly prevents malignant cells to continue with their life cycle, by inhibiting estrogen-mediated G1/S cell cycle progression. The compound inhibited uncontrolled cell migration of cancer cells. Currently, CasII-gly is in the phase I clinical trials. | [165, 166] |
| **Metal complexes under preclinical investigation** |                                        |                                                                                                                                                                                                                                |            |
| [VCp₂Cl₂]                                 | ![VCp₂Cl₂](image)                      | This metallodrug exhibiting a higher in vitro activity on several such us leukemias P388 and L1210, colon 38 and Lewis lung carcinomas, B16 melanoma, among others. In addition, in vivo studies revealed that vanadocene compounds show significant antitumor properties in MOLT-4 leukemia and human peripheral blood mononuclear cells | [33]       |
| Metvan                                    | ![Metvan](image)                       | METVAN induces apoptosis in human leukaemia cells, multiple myeloma cells and solid tumour cells derived from breast cancer, glioblastoma, ovarian, prostate and testicular cancer patients | [40]       |
dienyl complex (RuPMC), which displayed IC50 values in the micromolar range against human MCF7 and MDA-MB-231 breast and A2780 ovarian adenocarcinoma. Cellular distribution studies also showed that RuPMC was predominantly found in the nucleus and in the membrane. Encapsulation and delivery of cytotoxic ruthenium complexes within nanostructures have shown an advance in the enhance permeability and retention effect, along with improvements in the cytotoxic effects, decrease in the harmfulness and evasion of complex structure degradation. However, further studies still have to be performed for developing novel mechanisms of drug delivery structures.

Different drug delivery systems were described for vanadium compounds. Vanadocene dichloride could bind to transferrin and enhanced its anticancer activity in proliferating malignant cells that over-express transferrin receptors, indicating that transferrin plays an important role in the transport and targeted delivery of vanadocene dichloride into cancer cells [198]. An alternative delivery system for a complex of vanadium (IV) with aspirin (VOAspi) was developed using poly(beta-propiolactone) (PbetaPL) films by one of. UMR106 osteosarcoma cells were used as a model to evaluate the anticarcinogenic effects of the VOAspi released from the PbetaPL film. VOAspi-PbetaPL film inhibited cell proliferation in a dose-response manner and induced formation of approximately half of the thiobarbituric acid reactive substances, and an index of lipid peroxidation compared to that with free VOAspi in solution [199].

CONCLUSION

The tremendous success of cisplatin, carboplatin and oxaliplatin in cancer chemotherapy provoked a great impulse of research in Inorganic Medicinal Chemistry. Scientific efforts have been specially devoted to solve problems such as side effects, and resistance of platinum therapy. Intensive research has been done to design and develop novel non platinum-based metal anti-cancer drugs (Table 5). Different promising results have been reported in preclinical research for vanadium, ruthenium and copper complexes and several of these metal based compounds have been already investigated in clinical trials. In the preclinical stage, the main mechanisms of action involved with the antitumor properties of V, Ru and Cu compounds are: ROS generation, GSH/GSSG imbalance, cell cycle arrest, DNA fragmentation and apoptosis. Nevertheless, several compounds have attracted attention based on modes of action different from that of cisplatin (covalent binding to the DNA) whilst some metallocompounds interact with other specific molecular targets different from DNA (such as tyrosinkinases, proteasome, p53, etc). This research revealed new leading compounds for cancer therapy with well defined structure-activity relationships. In this order, the search of novel cellular targets including new inorganic and organic metal complexes is of great significance. Clinical research trials focused on copper and ruthenium compounds showed that the copper compound Cas-GlyII was entered in phase I due to the interesting antitumor profile found in pre-clinical studies. Besides, the ruthenium agent NAMI-A acts by its anti-metastasis properties rather than by conventional cell growth inhibitory effects. In addition, KP1019 in contrast to NAMI-A activated the primary tumour site.

Currently, both Ruthenium complexes have completed phase II of clinical trials. Actually, clinical studies on vanadium, ruthenium and copper based drugs are ongoing and they will bring new candidates for modern chemotherapy in the next few years.

Besides, this research in this field tried to overcome the problems existing between the therapeutic action and the adverse side effects of V, Ru and Cu compounds. The balance towards the beneficial side is the crucial issue for the future use of these compounds as alternative therapies against cancer.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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