Occurrence and sources of selected phenolic endocrine disruptors in Ria de Aveiro, Portugal

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Received: 19 May 2009 / Accepted: 21 November 2009 / Published online: 17 December 2009
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Abstract

Ria de Aveiro (Portugal) is a shallow coastal lagoon of high economic and ecological importance. Hardly any data on its chemical pollution by polar organic pollutants are available in literature. This study focused on the presence and sources of a series of phenolic endocrine-disrupting compounds (EDCs) in this area, including parabens, alkylphenolic compounds and bisphenol-A (BPA). A number of possible sources of pollution are present in the area, including the large harbours present in the lagoon, the city of Aveiro and the rivers discharging into the area. A recently constructed submarine wastewater outfall, located a few kilometres from the lagoon inlet has also been suggested as a possible source of pollution to Ria de Aveiro in several publications. The aim of the current field study was to investigate the occurrence and main sources of phenolic endocrine disruptors in Ria de Aveiro.

Materials and methods

An extensive sampling campaign was performed, with surface water and wastewater grab samples taken at over 50 locations, in duplicate on different days. Samples were treated using solid phase extraction and analysed by liquid chromatography tandem mass spectrometry.

Results and discussion

Concentrations in lagoon water were generally low: not exceeding 20 ng/L for most analytes. Levels in river water exceeded those in the lagoon by a factor 3 to 500 (o-phenylphenol (PhP) and nonylphenoxy ethoxy acetic acids (A9PEC), respectively), with concentrations up to 700 ng/L for BPA and 7,300 ng/L for A9PEC. Samples from the harbours showed EDC levels similar to those in the rest of the lagoon, but in the city of Aveiro, elevated concentrations were observed for alkylphenol ethoxylates (A9PEO), A9PEC, PhP and BPA. Wastewater effluents showed low levels for parabens, whilst alkylphenolic compounds reached several micrograms per litre. The effluents are discharged into the ocean via a submarine outfall, but as marine water near the outfall showed slightly elevated concentrations only for A9PEO, it does not seem to be a significant source of these EDCs for the area.

Conclusions

All the studied phenolic EDCs were detected in the study area, with high levels found in some of the rivers discharging into the lagoon, and generally low concentrations in the lagoon itself. The main sources for all investigated EDCs were the rivers Caster and Antuã which discharge into the lagoon. The city of Aveiro was identified as a secondary source. As the tidal water exchange volume is much larger than the freshwater input from the rivers, concentrations of phenolic EDCs remained low in the lagoon.

Keywords Endocrine disruptors · Ria de Aveiro · Fate study · Alkylphenols · Parabens
1 Background, aim and scope

Endocrine-disrupting effects in organisms have been shown in a number of field studies over the past 15 years (Jobling et al. 1998; Matthiessen et al. 2002; Vethaak et al. 2005), including several investigations along the Portuguese coast (Barroso et al. 2000; Sousa et al. 2005, 2007; Galante-Oliveira et al. 2006). In several of these studies, a relationship between the presence of organotin contaminants and endocrine-disrupting effects could be established. Other endocrine-disrupting compounds (EDCs) have so far received less attention in Portugal, and hardly any published data can be found on the environmental presence of these substances in Portuguese waters (Azevedo et al. 2001; Céspedes et al. 2004; Quirós et al. 2005). In the present study, the environmental occurrence of a combination of some well-known and some newly emerged phenolic EDCs was studied in the Ria de Aveiro lagoon in Portugal. The studied substances are all high production volume chemicals and reach the environment mainly via wastewater. Analytical methods for aqueous samples using solid phase extraction (SPE) and liquid chromatography tandem mass spectrometry (LC-MS/MS) were optimised to be able to determine all these EDCs within one method based on work of Benijts et al. (2004).

The current study includes the non-ionic surfactants nonylphenol ethoxylates (A₉PEOₙ) and their main metabolites nonylphenoxy ethoxy acetic acids (A₉PEₐC) and nonylphenol (NP), as well as octylphenol (OP), which is a metabolite of the octylphenol ethoxylate surfactants (Ahel et al. 1994; Jonkers et al. 2001). The study also includes the EDC bisphenol-A (BPA) that is mainly used in the production of polycarbonate plastics and epoxy resins. A large number of studies exist on the environmental behaviour of A₉PEOₙ and their metabolites as well as on BPA, and several reviews on this subject are available (Thiele et al. 1997; Staples et al. 1998; Fromme et al. 2002; Ying et al. 2002; Knepper and Berna 2003; Montgomery-Brown and Reinhard 2003; Kang et al. 2006; Giger et al. 2009).

Less well-known EDCs included in this study are the parabens and ortho-phenylphenol. The biocide ortho-phenylphenol (PhP) is used in diverse products such as glues, concrete additives, leather and as active ingredient in disinfectants. Parabens are a group of chemicals formed by para-hydroxybenzoic acid esters, with alkyl substituents ranging from methyl to butyl or benzyl groups. They are extensively applied as preservatives (usually as mixtures) in a large number of products including cosmetics and toiletries such as shampoos, skin care products and toothpastes (Rastogi et al. 1995; Routledge et al. 1998; Madsen et al. 2001). The parabens most commonly present in consumer products are methylparaben (MeP) and propylparaben (PrP). Both the parabens and ortho-phenylphenol are readily biodegradable under aerobic conditions; however, their high consumption amounts and continuous introduction into the environment may lead to a “pseudo-persistent” situation (Madsen et al. 2001).

The objective of the current field study was to investigate the occurrence and main sources of the series of endocrine disruptors mentioned above in Ria de Aveiro in Portugal. With respect to endocrine disruption, this estuarine system is a particularly interesting study area as several studies have shown endocrine-disrupting effects to occur in the area, including high levels of female gastropod molluscs masculinisation and sterility induced by organotin (OT) compounds (Barroso et al. 2000, 2005; Galante-Oliveira et al. 2009a, b; Sousa et al. 2005). Apart from data on OTs and heavy metals, hardly any information is available in the literature on the presence of EDCs or other organic microcontaminants in Ria de Aveiro.

Another interesting feature of Ria de Aveiro is the submarine wastewater outfall which was constructed several years ago and discharges all the treated wastewater of the area into the Atlantic Ocean, a few kilometres from the inlet of the estuary. Some literature suggests that a considerable part of this effluent flows back into the lagoon with tidal currents and could therefore be a relevant source of contaminants to the lagoon area (Figueiredo da Silva et al. 2002a). In the current study, the wastewater outfall was considered as one possible main source of EDCs for Ria de Aveiro. Other possible sources include the large harbours present in the lagoon, the city of Aveiro and the rivers discharging into the area.

2 Materials and methods

2.1 Reagents and standards

The standards of methyl-, ethyl-, propyl- and butylparaben as well as 4-nonylphenol (mixture of isomers) and 2-phenylphenol were obtained from Acros Organics (New Jersey, USA). Benzylparaben, 4-tert-octylphenol and bisphenol-A were purchased from Aldrich (Gillingham, UK). The internal standards d₁₆-BPA and nNPE₂C were from Cambridge Isotope Laboratories, and ¹³C₆-NP and ¹³C₆-A₉PEO₁₅ were a kind gift from P. Lee Ferguson (SUNY Stony Brook, New York, USA). Solvents (HPLC-grade) were purchased from Scharlau (Sentmenat, Spain), and ammonium acetate was from Merck (Darmstadt, Germany).

2.2 Study area

Ria de Aveiro (Fig. 1) is a shallow estuarine system in the northwest of Portugal formed by a complex system of
channels and wide intertidal areas (mudflats and salt marshes). The lagoon has a maximum width and length of 10 and 45 km, respectively, and is connected to the sea by an artificial inlet at the west side. The average depth of the lagoon is about 1 m, with a maximum of around 20 m in the channel near the inlet, and the water column can be considered as vertically homogeneous (Dias and Lopes 2006). The tidal regime is semi-diurnal with a tidal range of around 2 m at the inlet. Tide generates strong currents in deep and narrow channels, but not in the intertidal areas (Dias et al. 1999, 2000, 2001). The maximum tidal flow is about 4,700 m$^3$/s. This tidal contribution is much higher than the inputs from the main rivers Vouga and Antuã which, on average, contribute about 50 and 5 m$^3$/s, respectively (Lopes et al. 2001). Minor rivers flowing into the lagoon are the Rio Caster and Rio Boco.

Ria de Aveiro has three main branches extending from the sea inlet: the Mira, São Jacinto and Ilhavo channels. Seawater intrudes far into the branches, and only deep inside the branches, water salinity decreases below seawater levels in the summer. The whole estuary is heavily influenced by human activities, including tourism (a number of small tourist harbours are present), fishery and aquaculture, transport (large chemical and commercial harbours near the city of Aveiro), agriculture and industry (Jonkers et al. 2009).

The area surrounding the lagoon has a population of approximately 700,000 inhabitants (INE 2008), and a number of industries are present, including a pulp mill, steel and agrochemical industries. The wastewater treatment facilities in the estuary were built relatively recently, and until 2000, the effluents were discharged directly into the lagoon. Presently, all legal sources of effluent discharge are directed towards the ocean via a submarine outfall, which discharges domestic (30%) and industrial (70%) effluents after secondary treatment into the coastal area at 3.3 km offshore, 5 km north of the lagoon inlet (Figueiredo da Silva et al. 2002a). The prevailing inshore water current along the Aveiro coast is to the south. In a study on the Ria de Aveiro wastewater outfall using aerial observations and in situ water sampling, Figueiredo da Silva et al. (2002a, b) observed no offshore transport of the wastewater plume, but most commonly transport in the direction of the lagoon inlet channel. This suggests that impacts of pollutants discharged can be anticipated in nearshore waters and potentially within the estuarine system (Figueiredo da Silva et al. 2002a).

2.3 Sampling strategy

The goal of the sampling campaign was to determine the levels of EDC contamination for the whole Ria de Aveiro lagoon, to trace the main sources of phenolic EDCs in the area and to identify possible contamination hot spots. To this end, an extensive sampling campaign was performed in August 2006 in which water grab samples were taken at over 50 locations throughout the study area. One week later, sampling was repeated at the same locations to get an idea of the short-term variability of the system. Locations were selected near possible sources as well as locations where no sources were anticipated and “background levels”
of EDCs were expected. The following possible sources were considered:

- The submarine wastewater outfall: Marine water samples were taken near the outfall as well as near the inlet of the lagoon where part of the wastewater possibly reenters the lagoon.
- The city of Aveiro: Samples were taken in five of the city’s channels and the main entrance canal into the city.
- The large harbours in the vicinity of Aveiro: Two fishing ports, the commercial and the chemical ports were sampled.
- The rivers discharging into the lagoon: The 6 sources of freshwater into the lagoon were sampled, from north to south: Rio Caster, Rio Antuã, Rio Águeda (which flows into Rio Vouga), Rio Vouga, Rio Boco and the far end of the Mira channel.

Diffuse sources of EDCs from (illegally) discharged wastewater and tourism were anticipated as well, but no specific sampling was possible to trace these sources.

In addition, duplicate 24-h wastewater effluent samples were taken from the three wastewater treatment plants (WWTPs) in the study area: WWTP Cacia (ETAR Norte), WWTP Gafanha (ETAR Sul) and wastewater from a large pulp mill. This treated wastewater is not discharged into the estuary, but is led to the submarine wastewater outfall in the ocean. Duplicate samples of the wastewater reaching the outfall were taken as well.

Samples were taken from bridges, piers or from quays using a homemade bottle sampling device on a rope. All samples were taken at low tide. The marine water samples near the submarine wastewater outfall were taken by boat using a Van Dorn sampler and were collected just once.

2.4 Sample cleanup

A solid phase extraction method was optimised based on the method of Benijts et al. (2004). After filtration of the water using glass microfiber (GF/C) filters, the samples were frozen at −20°C until treatment. Before extraction, a mix of internal standards was added, including $^{13}$C$_6$-NP, $^{13}$C$_6$-NPEO, d$_{16}$-BPA and nNPE$_{2C}$ (an isomer having a linear alkyl chain and therefore a longer retention time). Extraction sample volumes were 150 mL for wastewater effluent and 250 mL for surface waters, with 5% methanol added to each sample. The SPE cartridges (Oasis HLB 200 mg, Waters) were conditioned with hexane, methyl tert-butyl ether (MTBE)/2-propanol (1:1), methanol and nanopure water. Then, the filtered samples (neutral pH) were passed through. After loading the cartridges with the sample, 4 mL of nanopure water was passed through to rinse the cartridge from salts and reduce possible matrix effects during analysis. Then, the cartridges were dried under vacuum. The cartridges were eluted with 3 mL MTBE/2-propanol (1:1), followed by 3 mL methanol. Finally, the extract was evaporated under a gentle nitrogen stream and transferred to 250 μL methanol/H$_2$O (1:1).

2.5 Analysis of phenolic EDCs

For analysis, a HP1100 liquid chromatography system (Hewlett-Packard, Waldbronn, Germany) coupled to an API4000 triple quadrupole mass spectrometer (Applied Biosystems, Foster City, USA) were used. Chromatographic separation was performed with a Lichrospher 100 RP18ec column (Macherey-Nagel, Dueren, Germany), with nanopure water containing 4 mM ammonium acetate and methanol as mobile phases. An electrospray interface was used in positive ionisation mode for A$_9$PEO$_n$ and negative mode for all other analytes. Detection was done in multiple-reaction monitoring (MRM) mode, with one mass transition for each analyte (see Table 1). LC-MS/MS parameters can be found in the Electronic supplementary material. Quantification was based on external calibration with areas relative to the internal standard areas (at least eight calibration standard solutions, $r^2$ was always above 0.97). The internal standard used for the parabens, phenylphenol and bisphenol-A was d$_{16}$-bisphenol-A, whilst for the alkylphenols, A$_9$PEC, A$_9$PEO$_{1,2}$ and A$_9$PEO$_{3-10}$, the internal standards $^{13}$C$_6$-NP, n-A$_9$PE$_{2C}$, $^{13}$C$_6$-A$_9$PEO$_2$ and $^{13}$C$_6$-A$_9$PEO$_3$ were used, respectively. The concentrations determined were corrected for average blank value and relative recovery.

2.6 Quality control

Along with each extraction round of 12 samples, a blank and a spiked sample (at levels from 150 to 1,000 ng/L) were processed. From these samples, relative recoveries were calculated. Limits of quantification were calculated as 10 times the standard deviation of the blank values, or for analytes where no significant blank signals were present, as signal-to-noise ratios of 10.

3 Results and discussion

3.1 Quality control

Eight field samples (from sea, river, lagoon and wastewater) were spiked to check the recovery of the extraction and possible interferences or matrix effects in the LC-MS/MS analysis. Recoveries were close to 100% (see Table 1).
some analytes, the average recoveries were above 100%, which indicates that matrix effects either cause an analyte signal enhancement (slightly) higher than for the internal standard or an internal standard signal suppression stronger than for the analyte. Environmental samples were corrected for recovery percentage.

Blank levels were relatively high for NP, leading to a higher limit of quantification (LOQ) for this analyte, as shown in Table 1. Blank levels (those of NP) were always below those of the environmental samples. The average of the blank levels was subtracted from the samples.

3.2 Salinity

At most of the locations sampled, salinities around 38 psu were measured. Clearly, the influence of ocean water is much larger than that of freshwater entering the study area. Only deep into the Mira Channel (southwestern channel) and in the rivers were lower salinities measured. In almost all cases, the highest concentrations of phenolic EDCs were found at salinity 0 psu. However, with the current data set, it is not meaningful to evaluate the correlations between salinity and EDC concentrations in detail as the data are strongly clustered around salinity 38 and 0 psu.

3.3 Occurrence of phenolic EDCs

When comparing the phenolic EDC concentrations of the two samples from each location (taken 1 week apart), an overall median concentration difference over all analytes of a factor of 1.9 was found. This indicates that concentrations were reasonably constant during the sampling period and that the grab samples give a representative indication of contaminant levels in the study area.

The dissolved concentrations of phenolic EDCs are presented in Table 2. Median and average concentrations for the different types of water sample are given, as well as minimum and maximum concentrations found. In Fig. 1, the spatial distribution of the concentrations for \( \text{A9PEO1}_{10} \) and BPA are shown.

The wastewater effluents were dominated by the alkylphenolic compounds, with the oxidised metabolites (\( \text{A9PEO1}_{10} \)) present at highest concentrations. Median concentrations were 3,260, 990 and 1,500 ng/L for total \( \text{A9PEO1}_{10} \) and NP, respectively. Effluent concentrations of BPA and PhP were generally above 100 ng/L, whilst OP and total paraben concentrations were lower, with medians of 17 and 27 ng/L, respectively. The paraben species with highest concentrations were MeP and PrP, which reflects the composition of paraben mixtures in common consumer products (Rastogi et al. 1995). Clearly, the secondary treatment in the WWTPs does not fully remove the phenolic EDCs present in the wastewater.

For all analytes, the highest concentrations in surface water were found in the river samples of Rio Caster, followed by Rio Antuã. Again, alkylphenolic compounds showed the highest levels, with maximum concentrations of 8,200, 1,540 and 233 ng/L for total \( \text{A9PEO1}_{10} \), \( \text{A9PEO1}_{10} \), and NP, respectively. For \( \text{A9PEO1}_{10} \), the concentration ranges in the Rio Caster are similar to those found in the wastewater effluents. The average concentration ratio of \( \text{A9PEO1}_{10} \) to \( \text{A9PEO1}_{10} \) of around 8 shows that in these rivers, the oxidative hydrolytic degradation route prevails over the non-oxidative route for \( \text{A9PEO1}_{10} \) (Jonkers et al. 2001). BPA levels were high in Rio Caster as well, with a maximum of 683 ng/L. The other rivers (Rio Vouga, Águeda and Boco) showed considerably lower levels, which were only slightly higher than in the saline waters.

Samples from the Aveiro city channels showed concentrations higher than in the lagoon for alkylphenolic compounds and BPA. Maximum levels in the channels for total \( \text{A9PEO1}_{10} \), \( \text{A9PEO1}_{10} \) and NP were 251, 266, 74 and 18 ng/L, respectively. For both the city and river samples, the \( \text{A9PEO1}_{10} \) oligomers present at highest abundance were of medium chain length (\( \text{A9PEO} \)). This indicates that inputs of \( \text{A9PEO1}_{10} \) occurred recently as in surface water, these surfactants are easily biodegraded into the metabolites \( \text{A9PEO1}_{2} \) or \( \text{A9PEC} \) (Jonkers et al. 2001).

Lagoon waters showed generally low levels of phenolic EDCs, with concentrations up to several tens of nanograms per litre. Samples taken in the harbours showed concentrations similar to those found elsewhere in the lagoon for all phenolic EDCs.

Levels in the coastal zone and lagoon were comparable for all compounds. For \( \text{A9PEO1}_{10} \) only, concentrations were slightly elevated near the marine outfall compared to the other coastal locations.

3.4 Sources of EDCs

From a comparison of EDC concentrations in different sample types, the main sources of phenolic EDCs to the Ria de Aveiro can be derived.

For the alkylphenolic compounds, concentrations were considerably higher in the rivers (especially Rio Caster and Rio Antuã) and city channels than in the lagoon or sea samples. Therefore, these are considered as the main sources of \( \text{A9PEO} \) and \( \text{A9PEC} \) to the lagoon. It is difficult to compare the contribution of the river and city sources quantitatively as the city channels are flushed with relatively clean seawater during each high tide and are therefore likely to vary strongly (with the current low tide samples representing a maximum concentration), whilst the concentrations in the river are not influenced by tidal movement.
The strong concentration differences for the different rivers (averages of 17 and 817 ng/L for A9PEO1-10 and 17 and 6530 ng/L for A9PE1-4C in the Rio Vouga and Rio Caster, respectively) can be partially explained by the differences in population densities in the river catchments, which is around 110 inhabitants per square kilometre for the Rio Vouga and 470 inhabitants per square kilometre for the Rio Caster and Antuã (Figueiredo da Silva et al. 2002b). An additional explanation for the concentration differences is the stronger dilution capacity in the Rio Vouga as the water flows of the rivers are 50 m³/s for Rio Vouga, 5 m³/s for Rio Antuã and <1 m³/s for Rio Caster and Rio Boco (yearly averages; Lopes et al. 2005). In the winter season, both river water flows and the fraction of river water in the lagoon increase. Differences in water flows as large as approximately 3 m³/s in September compared to approximately 150 m³/s in May have been reported for the Rio Vouga (Figueiredo da Silva et al. 2002b). Salinity at the far end of the São Jacinto channels decreases to around 15 psu in the northern half of the channel (Lopes et al. 2005). Therefore, in the winter season, concentrations of EDCs originating from the rivers may be higher at the far ends of the lagoon channels. However, the higher river water flows do not necessarily mean higher loads of EDCs (as the river water may be diluted), and therefore, seasonal fluctuations may play only a minor role.

The high concentrations of A9PE and A9PEC in wastewater did not lead to significantly increased concentrations in seawater near the submarine outfall for most of the analysed oligomers. Only for A9PEO3-7 were slightly increased concentrations found near the outfall. However, since the seawater samples were taken at ~1-m depth, it cannot be excluded that a deeper lying current with more polluted water exists.

For NP, the median concentrations in all surface water types were similar, which illustrates the ubiquitous nature

| Analyte          | MW   | MRM transition | Retention time (min) | Relative recovery (%) (n=8) |
|------------------|------|----------------|----------------------|-----------------------------|
| Methylparaben    | 152.2| 151.1→92.0     | 9.3                  | 111                         |
| Ethylparaben     | 166.2| 164.9→92.0     | 11.6                 | 107                         |
| Propylparaben    | 180.2| 178.9→92.0     | 13.8                 | 113                         |
| Butylparaben     | 194.2| 193.1→92.0     | 16.3                 | 121                         |
| Benzylparaben    | 228.3| 227.2→92.0     | 16.4                 | 118                         |
| Bisphenol A      | 228.3| 227.2→211.8    | 14.1                 | 100                         |
| Phenylphenol     | 170.2| 168.9→114.9    | 15.7                 | 91                          |
| Octylphenol      | 206.3| 205.2→132.8    | 24.2                 | 124                         |
| Nonylphenol      | 220.4| 219.1→133.0    | 26.3                 | 121                         |
| A9PE1 4C         | 278.3| 277.2→218.9    | 17.2                 | 123                         |
|                  | 322.3| 321.2→218.9    | 18.3                 | 123                         |
|                  | 366.3| 365.2→218.9    | 18.7                 | 123                         |
|                  | 410.3| 409.2→218.9    | 18.9                 | 123                         |
|                  | 244.3| 241.0→223.2    | 14.1                 | 123                         |
|                  | 322.3| 321.2→218.9    | 20.7                 | 123                         |
|                  | 226.4| 225.3→139.1    | 26.3                 | 123                         |
|                  | 264.4| 282.2→127.1    | 6.9                  | 95                          |
|                  | 308.5| 326.2→183.2    | 7.1                  | 96                          |
|                  | 352.5| 370.3→227.1    | 7.2                  | 117                         |
|                  | 396.5| 414.4→271.1    | 7.2                  | 117                         |
|                  | 440.5| 458.4→315.2    | 7.2                  | 117                         |
|                  | 484.5| 502.4→359.2    | 7.2                  | 117                         |
|                  | 528.6| 546.5→291.2    | 7.2                  | 117                         |
|                  | 572.6| 590.5→291.2    | 7.2                  | 117                         |
|                  | 616.6| 634.6→291.2    | 7.2                  | 117                         |
|                  | 660.6| 678.6→291.2    | 7.2                  | 117                         |
|                  | 314.5| 332.4→189.2    | 7.1                  | 117                         |
|                  | 358.5| 376.3→233.2    | 7.2                  | 117                         |
| Parent ions are ammonium adducts for A9PEO1 10 and deprotonated molecules for all other analytes

| Analyte          | MW   | MRM transition | Retention time (min) | Relative recovery (%) (n=8) |
|------------------|------|----------------|----------------------|-----------------------------|
| Methylparaben    | 152.2| 151.1→92.0     | 9.3                  | 111                         |
| Ethylparaben     | 166.2| 164.9→92.0     | 11.6                 | 107                         |
| Propylparaben    | 180.2| 178.9→92.0     | 13.8                 | 113                         |
| Butylparaben     | 194.2| 193.1→92.0     | 16.3                 | 121                         |
| Benzylparaben    | 228.3| 227.2→92.0     | 16.4                 | 118                         |
| Bisphenol A      | 228.3| 227.2→211.8    | 14.1                 | 100                         |
| Phenylphenol     | 170.2| 168.9→114.9    | 15.7                 | 91                          |
| Octylphenol      | 206.3| 205.2→132.8    | 24.2                 | 124                         |
| Nonylphenol      | 220.4| 219.1→133.0    | 26.3                 | 121                         |
| A9PE1 4C         | 278.3| 277.2→218.9    | 17.2                 | 123                         |
|                  | 322.3| 321.2→218.9    | 18.3                 | 123                         |
|                  | 366.3| 365.2→218.9    | 18.7                 | 123                         |
|                  | 410.3| 409.2→218.9    | 18.9                 | 123                         |
| Sample type          | Number of samples | \( \text{AOPE}_{1} \) | \( \text{AOPE}_{2} \) | \( \text{AOPE}_{2.7} \) | \( \text{AOPE}_{6.10} \) | \( \text{APE}_{1} \) | \( \text{APE}_{2} \) | \( \text{APE}_{2.7} \) | \( \text{APE}_{6.10} \) | \( \text{NP} \) |
|---------------------|-------------------|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|-----------------------|
| Lagoon              | 28                | <7                    | <7                    | <7                    | <7                    | 0.7-3.6               | 0.8                    | 0.8                    | 0.9-9.36              | 0.9-9.9               |
| Harbour             | 10                | <7                    | <7                    | <7                    | <7                    | 0.7-1.2               | 0.2                    | 0.4                    | 0.9-2.1              | 0.9-9.9               |
| Sea                 | 12                | <7                    | <7                    | <7                    | <7                    | 0.7-0.8               | 0.7                    | 0.2                    | 0.9-4.4              | 0.9-9.9               |
| Sea near outfall    | 20                | <7                    | <7                    | <7                    | <7                    | 0.7-5.6               | 1.7                    | 2.1                    | 3.1-4.9              | 1.6-4.2              |
| City                | 14                | <7-3.5                | 7.4                   | 10                   | 0.7-6.3               | 15                   | 17                    | 0.9-3.66              | 18                    | 0.9-9.3              |
| Rivers Águeda and Vogãa | 12          | <7-19                 | 2.1                   | 3.8                  | 0.7-145               | 1.6                   | 14                    | 0.9-1510             | 2.3                   | 134                  |
| Rivers Castor and Antaial WWTP effluent | 8 | 20-851               | 372                   | 455                  | 116-311              | 195                   | 206                   | 122-617              | 304                   | 316                  |

Median, average, minimum and maximum concentrations are given. The number of samples taken per sample type is also shown.
of this substance. Maximum NP concentrations were found in the Rio Caster and Rio Antuã, and therefore, these two rivers are considered as the main source of NP to the lagoon. However, even in those rivers, the Environmental Quality Standard (EQS=0.3 µg/L) set by the European Water Framework Directive was not exceeded (European Parliament and Council 2007). Concentrations of OP were extremely low in all surface waters, and therefore, no source could be indicated for this substance.

For BPA and PhP, the main sources again seem to be the Caster and Antuã river and, to a lower extent, the city of Aveiro. Although BPA and PhP were present in the wastewater, this did not lead to significantly elevated concentrations near the submarine outlet.

For MeP, concentrations are similar in all sample types, and no obvious sources can be distinguished. For the other parabens, concentrations in almost all surface water samples are very low, which is not surprising as these compounds are readily biodegradable (Madsen et al. 2001). The only elevated concentrations found are in the Rio Caster (up to 64 ng/L for PrP), which suggests a continuous discharge of untreated (or ineffectively treated) wastewater into this river.

The harbours in the lagoon, which have received relatively much attention in the past due to the presence of alkyltin EDCs (used as anti-fouling agents), do not seem to be significant sources of the EDCs investigated in this study.

As concentrations throughout the whole lagoon area remained low, diffuse sources from illegal discharges seem to be not very relevant. Only the elevated concentrations of some EDCs (alkylphenolic compounds and BPA) in the channels of Aveiro suggest some minor diffuse sources in the city. All of the city’s sewage system is connected to the submarine wastewater outfall.

3.5 Comparison to literature

For the Ria de Aveiro, not many data on micropollutant concentrations are available in literature. In a study on nutrient inputs into the lagoon, the rivers Antuã and Caster were found to contain higher concentrations of ammonium, nitrate and total phosphorus than the Vouga River, which is in agreement with our results for EDC concentrations (Figueiredo da Silva et al. 2002b). In that study, a considerable increase in nutrient loads into the lagoon from the ocean was expected, originating from the submarine outlet when it would come into use (in 2005). This expectation is not sustained by our current findings of low EDC concentrations near the outlet.

In the east of the lagoon, an area called Laranjo Bay is known to be polluted (a.o. by heavy metals) due to industrial activities (Lopes et al. 2001, 2005). The phenolic EDCs did not show elevated concentrations in this area, which can be explained by the fact that for several of the large industries in the area, the wastewater is no longer discharged into the lagoon but into the ocean via the submarine outfall.

In a study by Quiros et al. (2005), estrogenic activity and the occurrence of several EDCs in several Portuguese rivers were assessed using a recombinant yeast assay and LC-MS analysis. In the Ria de Aveiro, samples were taken in the Vouga River near the point where it enters the lagoon. Whilst in that study the Ria de Aveiro system was one of the sites showing low estrogenic activity compared to samples of other Portuguese sites, the average concentrations of the measured EDCs (A9PEO1,2, NP, OP and BPA) were a factor 10 higher than in the current samples from the Vouga River or the lagoon. At the time of sampling (2001–2002), the regional sewage collection system (connected to the marine outlet) was not yet fully operational (Figueiredo da Silva et al. 2002b). Since that time the number of inhabitants connected to the system has almost tripled (Figueiredo da Silva et al. 2002b), which may explain the considerably lower river concentrations found in the present study.

Compared to data on the occurrence of phenolic EDCs in coastal areas of other South European countries, the concentrations from the current study are relatively low (Kveštak and Ahel 1994; Petrovic et al. 2002; Pojana et al. 2007; Ardisoglu and Voutsa 2008). Petrovic et al. (2002) found aqueous concentrations of A9PEO along the Spanish coast above 1 µg/L on several locations which were impacted by wastewater discharges, with a maximum of 11 µg/L. A study in a Croatian lagoon showed maximum concentrations of 1.2 and 1.3 µg/L for NP and A9PEO2, respectively, which is more than an order of magnitude higher than in the current study (Kveštak and Ahel 1994). Lower levels similar to those in the present study were found recently in the lagoon of Venice (Italy) by Pojana et al. (2007), with concentrations in the low nanogram per litre range for BPA, NP and A9PEO1.

An explanation for the relatively low concentrations found in the current study may be the effective dilution of the wastewater discharged by the submarine outfall and the reduction in emissions after the recent EU restrictions in use of A9PEO (Cox and Drys 2003).

Concentrations in the present study found for alkylphenolic compounds and BPA in the Rio Caster are comparable to levels generally found in Northern European rivers (Heemken et al. 2001; Jonkers et al. 2003; Voutsa et al. 2006) and lower than some heavily polluted rivers such as near Barcelona (Spain, Céspedes et al. 2005), Porto and Lisbon (Portugal; Quiros et al. 2005).

The few literature data available on the environmental occurrence of parabens show low concentrations similar to
those of the present study. In river water from Belgium, maximum paraben concentrations of 0.085 µg/L were found (Benijts et al. 2004). In Canadian municipal wastewater, individual paraben concentrations were observed up to 2.4 and 0.04 µg/L for influent and effluent, respectively, with methyl- and propylparaben present at the highest levels (Lee et al. 2005). A study on a Spanish wastewater treatment plant showed maximum paraben concentrations of 2.9 µg/L in influent and 0.06 µg/L in effluent (Canosa et al. 2006).

4 Conclusions

All phenolic EDCs investigated were detected in water from the Ria de Aveiro lagoon system, with high levels found in some of the rivers discharging into the lagoon and generally low concentrations in the lagoon itself. The main EDC sources found were the rivers Caster and Antuã, whilst the city of Aveiro was found to be a minor source (for A9PEO, A9PEC and BPA). In contrast to suggestions in some recent scientific publications, the submarine wastewater outfall does not seem to be an important source of phenolic EDCs to the estuarine system as near the outfall, slightly elevated levels were found only for A9PEO.

Acknowledgements Isabel Quintaneiro and Margarida Esteves from the SIMRIA wastewater authority are kindly acknowledged for providing wastewater samples. We thank René Schönberger and Josef Asmin for their assistance in the laboratory and Marc Suter for scientific advice. This work was financed by the Swiss National Science Foundation (National Research Program NRP50, Project PHENCON).

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