Expression of the calcium-activated potassium channel in upper and lower segment human myometrium during pregnancy and parturition

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Abstract

Background: Large conductance calcium-activated potassium channel (BKCa) plays an important role in the control of uterine contractility during pregnancy. The change from uterine quiescence to enhanced contractile activity may be associated with the spatial and temporal expression of BKCa within myometrium. The objectives of this study were to examine the expression of BKCa alpha- and beta-subunit in upper segment (US) and lower segment (LS) regions of uterus, and to investigate for the possibly differential expression of these proteins in US and LS myometrium obtained from three functional states: (1) non-pregnant (NP); (2) term pregnant not in labour (TNL) and (3) term pregnant in labour (TL).

Methods: Myometrial biopsies were collected from non-pregnant women at hysterectomy and pregnant women at either elective caesarean section or emergency caesarean section. Protein expression level and cellular localization of BKCa alpha- and beta-subunit in US and LS myometrium were determined by Western blot analysis and immunohistochemistry, respectively.

Results: BKCa alpha- and beta-subunit were predominantly localized to myometrial smooth muscle in both US and LS myometrium obtained from non-pregnant and pregnant patients. The level of BKCa alpha-subunit in US but not in LS was significantly higher in NP myometrium than those measured in myometrium obtained during pregnancy. Lower expression of BKCa alpha-subunit in both US and LS was found in TL than in TNL biopsies. Expression of beta-subunit in both US and LS myometrium was significantly reduced in TL group compared with those measured in TNL group. There was no significant difference in BKCa beta-subunit expression in either US or LS between NP and TNL group.

Conclusion: Our results suggest that expression of BKCa alpha- and beta-subunit in pregnant myometrium is reduced during labour, which is consistent with the myometrial activity at the onset of parturition.
Background

During most of pregnancy, myometrium activity is characterized by poorly coordinated contractions. In late pregnancy, the uterus undergoes preparedness for the stimuli that lead to contractility and labour [1,2]. The mechanisms that initiate labour in women, particularly the molecular processes that convert the myometrium form a state of relative quiescence to the activated and contractile state, are not well understood. An understanding of these processes, at the molecular and cellular level, is essential to developing novel therapeutic strategies for management of associated clinical problems such as preterm labour that accounts for 85% of all perinatal complications and death.

It has been known that uterine myometrial contractility at term is triggered by a number of physiological signals, which orchestrate changes in uterine excitability via ion-channel modulation [3,4]. Potassium channels are central to regulation of cell membrane potential and contractility of smooth muscle [4,5]. Among the diverse families of K+ channels, the large conductance calcium-activated potassium channel (BKCa) is the predominant K+-channel type expressed in human myometrium [6,7]. This channel is activated by membrane depolarization and also by an increase in the intracellular calcium concentration, thereby playing a pivotal role in the modulation of uterine contractility and myometrial calcium homeostasis [3,4,8-10]. Electrophysiological studies have demonstrated changes in BKCa activity during pregnancy. It was reported by Wang et al. [11] that the contribution of BKCa channels to the total outward K+ currents was reduced by about 10% in pregnant myocyte near term compared with non-pregnant myocyte. Khan and colleagues had shown that the sensitivity of Ca2+ and voltage of this channel in human pregnant myometrium was lost at the time of labor [7]. A number of studies suggested altered BKCa expression in myometrium during pregnancy and parturition. Song et al. [12] worked on rat and found that BKCa is decreased by 60% in the myometrium of pregnant rats at labour [7]. Benkusky and co-workers [13] reported that BKCa in mouse myometrium is increased during pregnancy and diminished in post-partum. Studies by Khan’s group demonstrated BKCa expression in lower segment of human term myometrium and found it is decreased in labour onset [14,15].

It has been implicated that there is a functional regionalization in the myometrium during pregnancy and labour. The upper segment (US) region of the uterus expands to accommodate the growing fetus and then at labour contract to cause expulsion of the fetus, while the lower segment (LS) may maintain a relative quiescence to allow passage of the fetus [16]. Current data about BKCa expression in human myometrium during pregnancy are restricted to the LS [14,15,17]. There is no information concerning the expression of BKCa in the different region of uterus during pregnancy and labour that would support its role in the regulation of uterus contractions.

The objectives of the present study were to determine the regional distribution of the BKCa channel in human non-pregnant and term myometrium before and during parturition. Protein levels of BKCa channel were also examined by Western blotting to establish whether the expression of these proteins is changed in pregnancy and labour.

Methods

Tissue Collection

Paired upper and lower uterine segmental myometrial tissues from pregnant and non-pregnant women were collected in Navy General Hospital, the teaching hospital of Second Military Medical University, Beijing, China. Approval of this study was granted by human ethics committee of Navy General Hospital as well as human ethics committee of Second Military Medical University. Written informed consent was obtained from each participant.

Non-pregnant myometrium tissues were obtained from premenopausal, normal, cycling women (mean age 41 ± 4.3 yr, n = 8) undergoing hysterectomy for fibroids. Pregnant myometrial biopsies were collected at cesarean section form the following groups of pregnant women (37–42 wk): term no labour (mean age, 25 ± 2.2 yr, mean gestational age, 39 ± 1.7 weeks, n = 10) and term labour (mean age 26 ± 2.3 yr, mean gestational age, 39 ± 1.6 weeks n = 10). Labour was defined as regular contractions (<5 min apart) plus membrane rupture and cervical dilation (≥3 cm) with no augmentation (oxytocin or PG administration). Indications for cesarean section included breech presentation, placenta previa, previous cesarean section, cephalopelvic disproportion, failure of labour to progress, fetal distress, or maternal request. None of the women included in this study had evidence of underlying disease (e.g. hypertension, diabetes, preeclampsia, intrauterine growth restriction, etc.). LS uterine samples were collected from the upper margin of the LS uterine incision, while US uterine samples were taken from just below fundus. Because there is no LS in non-pregnant uterus, tissues taken from the isthmus of non-pregnant uterus were treated to be equivalent to the LS of pregnant uterus. All non-pregnant tissues were taken from the normal part of uterus without fibrosis contractions. Approval of this study was granted by human ethics committee of Navy General Hospital as well as human ethics committee of Second Military Medical University. Written informed consent was obtained from each participant.

Tissue Collection

Immunohistochemistry

Immunohistochemical investigations were performed with the Histostain-SP kit (Zymed, San Franscisco, CA),
Western blot analysis

Approximately 70 mg of human myometrial tissue was homogenized in ice-cold lysis buffer consisting of 60 mM Tris-HCl, 2% sodium dodecyl sulfate (SDS), 10% sucrose, 2 mM phenylmethylsulfonyl fluoride (Merck, Darmstadt, Germany), 1 mM sodium orthovanadate (Sigma-Aldrich), 10 μg/ml aprotinin (Bayer, Leverkusen, Germany). Lysates were then quickly ultrasonicated in ice bath, boiled 5 min at 95 °C, and stored at -80°C until used. Protein concentrations were measured using a modified Bradford assay and samples were diluted in sample buffer (250 mM Tris-HCl (pH 6.8), containing 4% SDS, 10% glycerol, 2% β-mercaptoethanol, and 0.002% bromphenol blue) and boiled for a further 5 min. Samples were separated on an SDS-8% polyacrylamide gel, and the proteins were electrophoretically transferred to a nitrocellulose filter at 300 mA for 1.5 h in a transfer buffer containing 20 mM Tris, 150 mM glycine, and 20% methanol. The filter was then blocked in TBS containing 0.1% Tween-20 (TBST) and 5% dried milk powder (wt/vol) for 2 h at room temperature. After three washes with TBST, the nitrocellulose filters were incubated with primary antibody for BKCa α- or β-subunit (1:500) at 4°C overnight. After another three washes with TBST, the filters were incubated with a secondary horseradish peroxidase-conjugated IgG (1:1000) for 1 h at room temperature and further washed for 30 min with TBST. Immunoreactive proteins were visualized using the enhanced chemiluminescence Western blotting detection system (Santa Cruz). The light-emitting bands were detected with X-ray film. The resulting band intensities were quantitated using an image scanning densitometer (Furi Technology, Shanghai, China). To control sampling errors, the ratio of band intensities to β-actin was obtained to quantify the relative protein expression level.

Statistical analysis

Protein levels of BKCa α- and β-subunit were determined by densitometric analysis (Furi Technology, Shanghai, China). Peak count values were expressed as densitometric units. The data are presented as mean ± SEM. All data were tested for homogeneity of variance by Bartlett's test. The results indicated that the data were normally distributed. Individual comparisons were made by Student's t-test for paired data. One-way ANOVA with Student-Newman-Keuls was used for multiple comparisons. P-value of <0.05 was considered to be significant.

Results

Expression of the α- and β-subunit of BKCa in non-pregnant and pregnant myometrium

Positive immunoreactivity for BKCa α- and β-subunit was identified in both US and LS myometrium from non-pregnant and pregnant women. Immunohistochemistry revealed that BKCa α- and β-subunit were predominantly localized to smooth muscle cells of myometrium. Smooth muscle cells lining blood vessel were also positively stained for these proteins (Fig 1 and 2). Immunoreactivity was abolished when the antibody was preabsorbed with excess peptide, thereby confirming the specificity of antibody (Fig 1E and 1F).

Western blot analysis, using α- and β-subunit-specific antibodies, detected bands of 110 and 35 kDa, respectively (Fig 3A). To given an overall expression profile in the US and LS, the expression values from all the pregnant patients were combined. When the overall expression level of each protein was compared in the pregnant upper and lower myometrium samples, there were no significant differences in either α- or β-subunit levels between US and LS myometrium (Fig 3B and 3C).

Pregnancy and labour associated changes in the expression of BKCa α-subunit

Within US myometrium, the expression level of α-subunit protein was significantly down-regulated in pregnant samples compared to that of the non-pregnant samples (NP versus all other groups, P < 0.01, Fig 4A). It was further decreased in TL samples compared to TNL samples (P < 0.05, Fig 4A).

Within LS, however, no significant differences in the expression of α-subunit protein were observed among NP
Immunolocalization of BKCa channel α- and β-subunit in non-pregnant myometrium. Positive staining for the α-subunit of BKCa (arrow) in (A) US myometrium and (B) LS myometrium. Positive staining for the β-subunit (arrow) in (C) US myometrium and (D) LS myometrium. (E-H) Negative controls. The primary antibody was substituted with (E) normal rabbit serum or (G) PBS. Sections were stained with (F) α-subunit preabsorption antibody or (H) β-subunit preabsorption antibody. Original magnification ×400.
Immunolocalization of BK$_{Ca}$ channel $\alpha$- and $\beta$-subunit in myometrium from pregnant women at term in labour or not in labour. A-D shows representative sections for positive staining for $\alpha$-subunit in (A) US myometrium not in labour, (B) LS myometrium not in labour, (C) US myometrium in labour and (D) LS myometrium in labour. E-H shows representative sections for positive staining for $\beta$-subunit in (E) US myometrium not in labour, (F) LS myometrium not in labour, (G) US myometrium in labour and (H) LS myometrium in labour. Arrow: positive staining. Original magnification $\times$400.
and TNL groups. This protein was significantly down-regulated in TL samples compared to that in TNL samples ($P < 0.01$, Fig. 4B). Within LS, β-subunit protein level was significantly lower in TL group than that in TNL or NP group ($P < 0.01$, Fig. 5B). No marked changes in either US or LS were observed in pregnancy (NP versus TNL).

**Discussion**
The present study demonstrated, for the first time, the expression of α- and β-subunit of BKCa in paired US and LS tissues from non-pregnant and pregnant women. BKCa α- and β-subunit were mainly localized to myometrial smooth muscle cells in the US and LS. The levels of these proteins were significantly down-regulated in both US and LS at the time of labour.

Studies by Khan’s group showed that the expression of both BKCa α- and β-subunit in LS region of uterus was significantly reduced during labour [14,15]. Curley et al. [17] also found decreased mRNA level of the α-subunit of BKCa in LS with labour onset. In consistence with the above studies, the present study also showed the labour associated decrease in the expression of α- and β-subunit proteins within LS myometrium. In addition, we also investigated the localization and expression pattern of BKCa within US during pregnancy and labour. Our data suggest that α- and β-subunit of BKCa in US region is significantly down-regulated at the onset of labour.

Current data outlining pregnancy associated expression of BKCa protein in LS region of human uterus are not fully consistent. Zhou et al. [18] reported that BKCa α-subunit protein in LS region remained virtually unchanged in pregnant human myometrium, compared to non-pregnant myometrium, whereas β-subunit protein was lower in pregnant tissues than in non-pregnant tissues. Matharoo-Ball et al. [15] showed that BKCa α-subunit was increased whereas β-subunit was decreased during pregnancy compared to the non-pregnant state. However, no studies to date appear to have examined BKCa in US myometrium of pregnant and non-pregnant women. In the present study, within LS region, no significant pregnancy associated changes in the expression of either α- or β-subunit were observed. Within US, α-subunit expression was decreased in pregnancy. In animal studies, a decrease in BKCa α-subunit protein levels close to or at term in rat myometrium [12] and an increase in the expression of BKCa α and β1 subunit throughout gestation in mouse myometrium [19] have been reported.

A lot of studies indicate differential expression of a variety of proteins including connexin-43, G proteins and prostaglandin receptors between US and LS myometrium with labour [20-24], which support the idea that increased contractility of the fundus compared to the lower segment during labour. However, similar expression levels of a few contraction-associated proteins (CAPs) in US and LS with
Figure 4
**Semiquantitation of Western blot signals of BKCa α-subunit in US and LS myometrium.** Myometrial tissues were obtained from non-pregnant women (n = 8) and pregnant women at term before the onset of labour (n = 10) or during active labour (n = 10). (A) Levels of α-subunit in US samples. (B) α-subunit expression in LS samples. Representative protein bands were presented on the top of the histogram. Data were expressed as mean ± SEM. *P < 0.05, **P < 0.01 with NP; #P < 0.05, ###P < 0.01 with TNL.

Figure 5
**Semiquantitation of Western blot signals of BKCa β-subunit in US and LS myometrium.** Myometrial biopsies were obtained from non-pregnant women (n = 8) and pregnant women at term not in labour (n = 10) or during active labour (n = 10). (A) BKCa β-subunit expression in US, (B) BKCa β-subunit level in LS. Representative protein bands were presented on the top of the histogram. Data were expressed as mean ± SEM. **P < 0.01 with NP; #P < 0.05, ###P < 0.01 with TNL.
labour have also been reported [21,25,26]. For instance, Havelock et al. [26] showed that S100A9 mRNA was up-regulated in both US and LS during labour. Our results indicate that the similar expression pattern of BKCa in US and LS is occurred with the onset of labour. Together, it suggests that the contractile activity of US or LS region is not likely to be determined by a single specific protein but rather by a combination of all CAPs.

The smooth muscle BKCa channel is formed by tetrameric assembly of an α subunit and an accessory β-subunit [27]. The α-subunit forms the functional BKCa channel [28,29]. The presence of the β-subunit confers BKCa with higher Ca2+ and voltage sensitivity [10,30]. Thus, reduced expression of BKCa α- and β-subunit would permit an increase intracellular Ca2+ levels without permit an increase opposing K+ conductance, thereby enhancing contractility of smooth muscle. Because the BKCa channel is particular abundant in uterine myocytes, a reduction of this channel in protein level would translate into a considerable shift toward electrical excitability in the human uterus and likely promote contractile activity.

Conclusion
Our results indicate that BKCa α- and β-subunit are predominately localized to myometrial cells in US and LS region of pregnant and non-pregnant human uterus. It has been implicated that, at the time of labour, the uterus differentiates into the highly contractile activity. Because BKCa channel play a pivotal role in the modulation of uterine excitability and contractility, the down-regulation of BKCa channel expression in myometrium following labour may contribute, in part, to the enhanced contractility during parturition.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
LG and BC carried out all experimental work. LZ recruited patients, organized the collection of tissues. XN conceived of the study, and participated in its design and coordination. All authors read and approved the final manuscript.

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