ABSTRACT

Brown adipose tissue (BAT) is a specialized tissue critical for non-shivering thermogenesis producing heat through mitochondrial uncoupling; whereas white adipose tissue (WAT) is responsible of energy storage in the form of triglycerides. Another type of fat has been described, the beige adipose tissue; this tissue emerges in existing WAT depots but with thermogenic ability, a phenomenon known as browning. Several peripheral signals relaying information about energy status act in the brain, particularly the hypothalamus, to regulate thermogenesis in BAT and browning of WAT. Different hypothalamic areas have the capacity to regulate the thermogenic process in brown and beige adipocytes through the sympathetic nervous system (SNS). This review discusses important concepts and discoveries about the central control of thermogenesis as a trip that starts in the hypothalamus, and taking the sympathetic roads to reach brown and beige fat to modulate thermogenic functions.

1. Introduction

Brown adipose tissue (BAT), and more recently beige adipose tissue, are responsible for heat production through non-shivering thermogenesis (NST). Initially, BAT was thought to exist only in small or hibernating mammals and newborn humans. However, in 2009 functional BAT was identified in adult humans through positron-emission and computed tomographic (PET-CT) scans [1–5]. Since then interest by the scientific community in studying the molecular mechanisms involved in the regulation of NST have expanded greatly. Another type of thermogenic fat was described, “brown-like” adipocytes present in WAT depots named the beige or brite (“brown in white”) adipose tissue, that emerges in located depots of white adipose tissue (WAT) under some stimuli and which possesses thermogenic properties [1,2]. Thermogenesis in BAT and browning of WAT are activated with cold exposure and sympathetic stimulation [3–5]. Several central mechanisms regulating thermogenesis in brown and beige adipocytes have been described, which are the focus of this review. For this purpose, we will start a trip at the control center, namely the hypothalamus, the most studied region of the central nervous system (CNS) with respect to the control of energy balance. From the hypothalamus, we will follow the road through the autonomous nervous system (ANS), via the sympathetic fibers that connect and regulate the different fat pads, where the thermogenic pathway ends.

2. The hypothalamus: the beginning of the travel

The control of energy homeostasis is orchestrated by the CNS with the hypothalamus playing a major role. Indeed, most of the peripheral cues, such as sensory and nutritional signals and humoral factors arrive at the hypothalamus to provide information about the nutritional status of the body. The information is integrated to emit a response through regulation of food intake, energy expenditure (EE) and nutrient partitioning [6–14]. The hypothalamus is composed of several neural populations distributed in different hypothalamic nuclei (Figs. 1 and 2) that are interconnected to each other and to other brain areas. Furthermore, peripheral energy balance is controlled by the hypothalamus through the ANS. Thermogenesis in brown and beige adipose tissues can be induced by activation of the sympathetic nervous system (SNS) [7,8,11,15–17] (Figs. 1 and 2). The control of ANS by the hypothalamus is complex, involving several neuronal populations and signaling pathways in various nuclei.
Several peripheral signals act on the hypothalamus to inform about the nutritional status of the body. Therefore, leptin, glucagon like peptide (GLP-1), bone morphogenetic protein 8B (BMP8B), estradiol (E2) or thyroid hormones (THs) act in the ventromedial nucleus of the hypothalamus (VMH), decreasing AMP-activated protein kinase (AMPK) which activates the sympathetic nervous system (SNS) signaling to brown and beige adipose tissues. Events such as decreasing the endoplasmic reticulum (ER) stress in the VMH or increasing orexins (OX) in the lateral hypothalamic area (LHA) also activate thermogenesis through sympathetic activation. rRPa: rostral raphe nucleus, IO: inferior olive.

Cytokines PGE2
COLD

Fig. 1. Brain adipose axes. Several peripheral signals act on the hypothalamus to inform about the nutritional status of the body. Therefore, leptin, glucagon like peptide (GLP-1), bone morphogenetic protein 8B (BMP8B), estradiol (E2) or thyroid hormones (THs) act in the ventromedial nucleus of the hypothalamus (VMH), decreasing AMP-activated protein kinase (AMPK) which activates the sympathetic nervous system (SNS) signaling to brown and beige adipose tissues. Events such as decreasing the endoplasmic reticulum (ER) stress in the VMH or increasing orexins (OX) in the lateral hypothalamic area (LHA) also activate thermogenesis through sympathetic activation. rRPa: rostral raphe nucleus, IO: inferior olive.

Fig. 2. Hypothalamic circuits regulating thermogenesis of BAT and browning of WAT. The preoptic area (POA) receives peripheral information about external temperature, being activated by cold and prostaglandin E2 (PGE2), and mediate the febrile response. The POA projects to other hypothalamic nuclei such as the ventromedial nucleus of hypothalamus (VMH) leading the activation of BAT sympathetic traffic through projections to rostral raphe nucleus (rRPa). Several peripheral signals converge in the VMH to regulate thermogenesis: activation of estrogen receptor α (ERα), glucagon like peptide 1 receptor (GLP1-R), thyroid hormone receptor (TR), or bone morphogenetic protein 8 receptor (BMP8R) inhibit AMPK in the VMH stimulating thermogenesis in in brown adipose tissue (BAT) and white adipose tissue (WAT), as well as lipotoxicity-induced ER stress. The dorsomedial nucleus of the hypothalamus (DMH) is also involved in the thermogenesis regulation, since it is inhibited under normothermic environment by a GABAergic tone which is disinhibited by POA stimulation inducing thermogenesis. The arcuate nucleus of hypothalamus (ARC) contain neurons that express orexigenic factors such as agouti-related protein (AgRP) along with neuropeptide Y (NPY) and anorexogenic neurons expressing proopiomelanocortin (POMC). Leptin induces POMC expression increasing the release of POMC products in the second order neurons. Leptin action in the ARC also activate RIP-Cre expressing neurons, which through disinhibition of GABAergic population act on paraventricular nucleus of the hypothalamus (PVH) to induce thermogenesis. Conversely, AgRP and NPY in the ARC act on the PVH decreasing the sympathetic activation of BAT. In the lateral nucleus of the hypothalamus (LHA), neurons expressing orexins (OX) promote BAT thermogenesis, a process that can be activated by low AMPK activity in the VMH. LepR: Leptin receptor; GABA-R: GABA receptor; MPO: medial POA; MnPO: median part of the POA; MC4R: melanocortin 4 receptor; β3-AR: beta 3 adrenoreceptor.
2.1. The preoptic area

The preoptic area (POA) is the main brain nucleus that senses temperature. It acts as a thermostat that receives the peripheral and central thermal signals through temperature-sensitive neurons to regulate body temperature according to the external environment [18–22]. The POA receives input from thermosensitive areas elsewhere in the body. Thus, detection of cold signals by the median POA (mPOA) triggers hypothalamic mechanisms to induce thermogenesis in BAT [23]. More specifically, POA connects directly with the ventromedial nucleus of the hypothalamus (VMH) to activate BAT thermogenesis in response to cold, as demonstrated by the fact that destruction of the VMH abolishes the ability of external cold to stimulate BAT thermogenesis [24,25].

The POA also controls the febrile response, a physiological mechanism to induce a hyperthermic environment needed to defend against pathogens [26]. During infections, prostaglandins (PG) are released in the vasculature and peripheral tissues and upon arrival to the POA trigger activation of the BAT thermogenic program [27–29]. Specifically, POA contains PG receptors subtype EP3, which are connected to the dorsomedial nucleus of the hypothalamus (DMH), as well as the rostral raphe pallidus (rRPA) in the brainstem that control thermogenesis in BAT to induce fever through a mechanism that involve cAMP [27–30]. A population of the POA neurons expressing EP3 subtype of PGE receptor is mainly GABAergic and projects to DMH and the rRPa antagonizing some inhibitory fibers to induce fever [30]. Furthermore, neurons of the mPOA which are enriched in melanocortin 4 receptors (MC4R) and connected to the DMH can also activate thermogenesis in brown and beige adipocytes [31], although the exact role of the POA in regulating browning remains unknown.

2.2. The arcuate nucleus of the hypothalamus

The arcuate nucleus of the hypothalamus (ARC) is located just above the median eminence (ME) where the blood brain barrier (BBB) is permeable allowing access of peripheral signals to the brain [7,8,11,15–17]. Thus, the ARC is considered as one of the gates for the brain to sense circulating factors. The ARC is heterogeneous, containing several populations of neurons, but the most studied are those expressing the orexigenic neuropeptides, such as agouti-related peptide (AgRP) and neuropeptide Y (NPY), or anorexigenic neuropeptide precursors, such as pro-opiomelanocortin (POMC) and cocaine- and amphetamine-regulated transcript (CART) [7,8,11,15–17]. In addition to the control of feeding behavior, the ARC also regulates energy expenditure [14,32,33]. For example, leptin action in the ARC is required for the induction of action potential and firing of the sympathetic nerves subserving BAT [34,35]. Interestingly, activation of ARC neurons expressing NPY was found to decrease BAT thermogenesis [36,37]. A recent study describes that this observation is sexually dimorphic, with female mice lacking corticotropin-releasing hormone receptor 1 (CRFR-1) in AgRP neurons showing decreased thermogenesis in brown and beige adipocytes in response to cold [38]. On the other hand, POMC neurons have been related to increased thermogenesis through activation of SNS to BAT [39–41]. As a consequence, mice lacking genes downstream to POMC neurons, such as the Mc4r gene display suppressed leptin-dependent augmentation of thermogenesis in BAT and WAT [42]. Conversely, absence of protein tyrosine phosphatases 1B (PTP1B) and tyrosine-protein phosphatase non-receptor type 2 (TCPPTP), both of which suppress signaling of leptin and insulin receptors, promote WAT browning and energy expenditure in response to insulin and leptin, preventing the development of diet-induced obesity [43]. The capacity of leptin to induce BAT thermogenesis by through the ARC may also involve another population of neurons; the GABAergic RIP-Cre (Cre-mediated expression of rat insulin II promoter)- expressing neurons through their projections to other hypothalamic sites, such as the paraventricular nucleus (PVH) [44–46]. Overall, these findings suggest that both orexigenic and anorexigenic neural populations of the ARC can regulate thermogenesis.

The molecular mechanism mediating the effect of the ARC on thermogenesis remains unclear. There is evidence pointing to endoplasmic reticulum (ER) stress in the ARC as an important mechanism involved in the regulation of energy expenditure. Indeed, increased ER stress in POMC neurons reduced thermogenesis in BAT [47–49], while reduction of ER stress, or O-linked-β-N-acetylglucosamine transferase (OGT) expression are associated with increased energy expenditure and thermogenesis in BAT and WAT [50,51].

2.3. The dorsomedial nucleus of the hypothalamus

It is widely reported that the DMH is involved in the central control of thermogenesis in brown and beige adipose tissues through sympathetic transmission. The DMH is involved in the febrile response, during which some DMH glutamatergic neurons activate sympathetic fibers to BAT through rRPas. This process involves two groups of neurons. Initially, the GABAergic neurons within the DMH inhibit the glutamatergic neurons. However, under prostaglandin (PG)E2 activation of the POA, stimulation of a second group of GABAergic neurons in the POA inhibit the DMH GABAergic neurons [11,30,52–56].

There are NPY-expressing neurons in the DMH that are able to regulate thermogenesis through sympathetic activation to BAT and WAT. In fact, disruption of NPY in the DMH increases BAT thermogenesis and browning of WAT [31,57]. The existing evidence suggests that hypothalamic NPY plays an important role in the control of thermogenesis and promote energy storage in fat [58]. However, under some conditions, such as high energy demand, diet-induced obesity or upon cold exposure, NPY in the DMH increases the thermogenesis process, a phenomenon different from what happens in the ARC where NPY expression is reduced after cold exposure, and where NPY reduces BAT activity [57,59]. Taken together, NPY in the DMH could play a role in thermogenic regulation as part of a basal inhibitory tone that can be disinhibited under some stimuli, for example the induction of the febrile response from the POA. However, such possibility remains to be tested.

Finally, there are some neurons expressing leptin receptor (Lepr) in the DMH, which contribute to sympathetic activation to some fat depots in obese mice [60,61]. Accordingly, lack of Lepr in the DMH was found to decrease BAT thermogenesis promoting weight gain [60], suggesting that leptin in the DMH plays a key role in the sympathetically-mediated activation of BAT thermogenesis.

2.4. The ventromedial nucleus of the hypothalamus

The importance of the VMH in the regulation of thermogenesis is well established. The main role of this nucleus consists in the integration of various peripheral signals to coordinate the thermogenic response, particularly the sympathetic tone to BAT and WAT. This is supported by the anatomical link between the VMH and the brown and white fat pads [62,63] through the rRPas and inferior olive (IO) in the brainstem [32,64–69]. Thereby, inhibitory factors, such as GABA agonists in the VMH impair the PGE2-induced thermogenesis [27], while action of excitatory neuropeptides such as glutamate, noradrenaline, serotonin and tryptophan in the VMH activate BAT thermogenesis [70–76]. A novel paradigm postulates that the VMH-dependent control of thermogenesis is mediated by circadian rhythms, since mice lacking the clock-control gen Bmal1 in the VMH show impaired thermogenesis during the night phase [77].

As mentioned above, several peripheral signals arrive at the VMH to activate thermogenesis in BAT and browning of WAT. Many of these signals such as thyroid hormones (THs) [66,78,79], bone morphogenetic protein 9B (BMP9B) [80,81], leptin [82], estradiol (E2) [67], glucagon-like-peptide-1 (GLP1) analogues [83], and drugs such as nicotine [84] have been demonstrated to act through a common
mechanism, involving inhibition of AMP-activated protein kinase (AMPK) which leads to the activation of sympathetic fibers to BAT and WAT [79,81,83]. More recent data demonstrate that BMP8β acts by decreasing AMPK in VMH neurons connected to the LHA through glutamatergic fibers that stimulate the orexin (OX) system which in turn activate the SNS promoting thermogenesis in BAT and WAT [81]. Conversely, adiponectin (ADP) and resistin (RSTN) caused a decrease in BAT thermogenesis by activation of hypothalamic AMPK [85–91], although the role of the VMH in these actions remains unproven. Additional peripheral signals that act on the hypothalamus eliciting sympathetic firing to BAT and activating thermogenesis include amylin and uroguanylin (UNG), with UNG also inducing browning of WAT [92–94]. However, it remains unknown whether the mechanism mediating the actions of these two hormones involves a decrease of AMPK activity in the VMH. The current evidence points to the energy sensor AMPK in the VMH as a canonical regulator of the brain-adipose axes through the SNS [16]. This makes hypothalamic AMPK a potential therapeutic target in the fight against obesity [16].

Hypothalamic complex lipids, such as ceramides, have been involved in the regulation of thermogenesis. Low levels of ceramides are implicated in cellular functions such as growth, differentiation, adhesion and apoptosis [95–97]. High levels of hypothalamic ceramides have been shown to be lipotoxic, by promoting ER stress in the VMH and reducing the BAT sympathetic tone which impairs the thermogenic process [98]. Overexpression of the chaperone glucose related protein 78 (GRP78) in the VMH, decreases ER stress thereby promoting BAT thermogenesis and browning of WAT causing weight loss and improvement in the metabolic phenotype of diet-induced obese rats in manner independent of feeding [99,100]. These data suggest that modulation of hypothalamic ER stress could be a promising therapy to treat obesity and associated metabolic alterations.

2.5. The lateral hypothalamic area

The LHA has long been implicated in the control of thermogenesis in BAT and more recently in browning of WAT. Most of the evidences point to orexins A and B (OX-A and -B), that are orexigenic neuropeptides expressed predominantly in the LHA, as well as thermogenic inducers in BAT and WAT [81,101–106]. Interestingly, OXs are also required to correct development, differentiation and function of brown adipocytes [107]. In fact OXs act on both components of energy expenditure, thermogenesis and locomotor activity [104,107–110].

As discussed in the previous section, the LHA seems to receive projections from the VMH and overexpressing OXs activate sympathetic fibers innervating BAT and WAT [81,108–115]. It should be noted however, that OX null mice display normal temperature although they are unable to induce thermogenesis in response to stress induced by handling, suggesting that OX plays an important role in stress-dependent thermogenesis [116]. The involvement of OX neurons in BMP8β-induced thermogenesis has been aforementioned [81]. The possible induction of OX in the LHA to activate thermogenesis by other peripheral signals that trigger AMPK-VMH remains to be determined.

2.6. The paraventricular nucleus of the hypothalamus

The PVH regulates feeding behavior and energy homeostasis, and is widely interconnected with other hypothalamic and extra-hypothalamic areas. However, the exact role of the PVH in the control of thermogenic processes is controversial due to conflicting findings. For instance, direct stimulation of the PVH was reported to stimulate [71], inhibit [46] or have no effect [117] on BAT thermogenesis.

Some evidence suggests that the PVH activates thermogenesis during the febrile process by inducing thermogenesis in BAT and browning of WAT through sympathetic activation, while lesions in the PVH reduce fever [116,118–120]. Furthermore there is evidence for anatomical connection between the PVH and the BAT [4,63,114]. The link between the PVH and BAT is further supported by the fact that administration of various excitatory substances in the PVH, such as corticotropin -releasing hormone (CRH), hydroxybutyrate, glutamate, noradrenaline, serotonin, tryptophan, cholecystokinin (CCK), brain-derived neurotrophic factor (BDNF), histamine, UCN, CART, PGE2, leptin or melanocortins leads to the activation of BAT thermogenesis [44,71,74,121–131]. It is worth mentioning that genetic ablation of single-minded homolog 1 (SIM1; a factor necessary for the correct development of the PVH) reduces thermogenesis in BAT leading to obesity, suggesting that overall the PVH induces BAT thermogenesis [132]. There is also evidence suggesting that the PVH induces browning of white fat. For instance, CART administration into the PVH induces uncoupling 1, 2 and 3 (UCP1, UCP2 and UCP3) expression in brown and beige adipocytes [126]. Moreover, genetic ablation of liver X receptors (LXRs) leads to browning of WAT through activation of thermoprogenic releasing hormone (TRH) in the PVH [133]. Together these results demonstrate that the PVH regulates thermogenesis in both BAT and WAT.

Since stimulation or disinhibition of PVH neurons impairs thermogenesis in BAT and reverses the effects induced by cold or by N-methyl-D-aspartate receptor (NMDA; a specific agonist at the NMDA receptor mimicking the action of glutamate) administration into the rRPa led to the suggestion of the existence of a group of neurons in the PVH that project to the brainstem inhibiting the BAT sympathetic activation [46,134]. This is supported by the demonstration that NPY administration into the PVH reduces BAT thermogenesis [37,135]. On the other hand, the melanocortin system in the PVH regulates feeding behavior but appears not involved in the regulation of thermogenesis [136]. Finally, an interesting recent report described a negative link between oxidative stress in the PVH and thermogenesis, since inhibition of NADPH-oxidase, a reactive oxygen species (ROS) inducer, selectively in the PVH of obese mice increased BAT thermogenesis and browning of WAT through sympathetic activation protecting against diet-induced obesity [137].

Taken together, it appears that PVH negatively regulates BAT thermogenesis under basal conditions and some stimuli that activate the inhibitory presynaptic GABAergic neurons hinder the inhibition of thermogenesis by the PVH. However, further investigations are needed to establish the exact role of PVH in thermogenic regulation.

3. Next stop: the brainstem

As mentioned above, the different hypothalamic areas which receive several inputs including hormones and nutritional signals integrate the information that is transmitted through efferent fibers to many other CNS areas particularly the brainstem [11,30,109,110,138]. Among the brainstem nuclei, rRPa neurons receive tonic inhibitory inputs most notably from warm-sensitive GABAergic POA neurons, and disinhibition of rRPa neurons by various thermogenic signals increases BAT sympathetic activity [30]. The rRPa sympathetic premotor neurons integrate several signals from peripheral thermoreceptors, from the hypothalamus and other brain areas implicated in the regulation of the body temperature to control sympathetic output that activates BAT. Thus several neuronal networks converge in the rRPa which controls the sympathetic nerves subserving BAT and WAT [11,22]. Viral tracing evidence demonstrates that sympathetic nerves that innervate subcutaneous WAT originate from rRPa [139]. Thus the role of the rRPa in the control of BAT and WAT sympathetic tone is well established.

Substantial evidence points to the importance of the brainstem nuclei such as the rRPa in mediating the sympathetically-mediated effects of the hypothalamic nuclei on BAT thermogenesis and WAT browning. The VMH is anatomically linked to rRPa as well as the IO [17,64–69]. It has been also demonstrated that a morphological connection between neurons expressing OX in the LHA with the rRPa neurons exists [111,114]. Moreover, activation of the PVH neurons attenuates the increase in BAT activity evoked by NMDA injections into
the rPAs, suggesting the existence of inhibitory projections from the PVH to rPAs decreasing BAT sympathetic nerve activity [46]. Although the cellular mechanisms mediating these actions are not fully understood, it has been reported that sympathetic premotor neurons expressing vesicular glutamate transporter type 3 (vGlut3) in the rPAs activate BAT [11,22,140]. Neurons in the rPAs also express serotonin (5-HT) that plays a role in BAT activation [64]. Thus, cold activates 5-HT neurons in the rPAs [141], and accordingly pharmacological activation of 5-HT receptors in the rPAs affects sympathetic tone to BAT in response to cold [142,143]. Furthermore, inhibition of 5-HT biosynthesis inhibits thermogenesis [144].

Other brainstem areas have been implicated in the control of BAT sympathetic tone. For instance, catecholaminergic neurons in the ventrolateral medulla, also located in the brainstem, inhibit rPAs BAT premotor neurons through activation of α2 adrenergic receptors probably regulating a hypothermic process [134,145]. The lateral parabrachial nucleus, periaqueductal gray, and locus coeruleus have also been linked to the control of thermogenesis [146–150], their exact role remains unclear.

4. The sympathetic highway

The ANS is a major and powerful regulatory system that ensures homeostasis of the body’s functions. The metabolic organs are innervated and tightly regulated by two antagonistic branches of the ANS, the SNS and the parasympathetic nervous system (PSNS) [4,151–153]. Generally, the SNS has been associated with emergency and threat situations and thus referred to as the "fight or flight" response. The PSNS compensates for the SNS to restore the body to a state of calm and thus thought to mediate the “rest and digest” response.

Both BAT and WAT are innervated by the SNS with a wide presence of nerve terminals and postsynaptic β3-adrenergoreceptors (β3-AR) on adipocytes [3,154]. Induction of thermogenesis in adipocytes is triggered by the arrival of the sympathetic signal, consisting in the release of noradrenaline (NA) which binds to β3-AR activating lipolysis in WAT and the thermogenesis in brown and beige adipocytes. Stimulation with NA, which binds to β3-AR activating lipolysis in brown adipocytes [180], β3-AR is coupled to a stimulatory G-protein that triggers activation of adenylate cyclase (AC) which in turn catalyzes the conversion of ATP into cyclic AMP (cAMP) which leads the activation of protein kinase A (PKA). Several downstream pathways that increase thermogenesis are activated PKA in adipocytes. First, PKA activates p38 mitogen-activated protein kinase (MAPK) and extracellular signal regulated kinases (ERK)1/2, which facilitate UCP1 transcription. Moreover, PKA induces lipolysis, activating several lipases, such as adipose tissue triglyceride lipase (ATGL), hormone-sensitive lipase (HSL) and monoacylglycerol lipase (MGL), which hydrolyze triacylglycerol, diacylglycerol and monoacylglycerol, respectively, into free fatty acids (FFAs) and glycerol. Then, carnitine palmitoyltransferase 1a (CPT1a) introduces FFAs into the mitochondria, where they are oxidized through β-oxidation leading to reduction of NAD+ and FAD to NADH and FADH2, which serve as fuel of the electron transport chain. Oxidation drive protons out of the mitochondrial matrix from where they are re-introduced by UCP1, dissipating energy as heat [7,170,179]. This heat-producing process in brown or beige adipocytes, named thermogenesis, is the final destination of our trip.

6. The main reason for this travel: the impact of thermogenesis on obesity

Clearly, the autonomic control of thermogenic processes is critical for maintaining energy homeostasis and dysregulation in this regulatory mechanism is involved in metabolic disease and obesity. Thus, it may possible to use therapeutic strategies to reverse obesity-associated defects in autonomic control of thermogenesis. Indeed, evidence gathered in recent years suggests that activation of thermogenesis could become a promising target to curb obesity due its capacity to burn fat, especially after the recognition of the importance of BAT and browning in adult humans [180–184]. It is estimated that cold-induced thermogenesis in BAT in lean healthy volunteers burn as much as 25–400 kcal/day [185]. It should be noted that activation of thermogenesis not only induces weight loss, but also has a positive impact on other alterations in the metabolic syndrome such as insulin and leptin resistance, hepatic steatosis, hyperlipidemia, hyperglycemia, hypercholesterolemia and hypertriglycerideremia, likely due to the ability of the thermogenic adipocytes to uptake lipids and glucose from the circulation [15,161,186–191]. Studies targeting central pathways such as hypothalamic AMPK [66,67,80,81,83,84] or ER stress [48,98,99], have demonstrated the beneficial and specific effects of activating BAT thermogenesis and browning of WAT. Currently, some agents that have demonstrated action on hypothalamic targets such as AMPK including metformin, nicotine or liraglutide are used in the clinic [16].

7. Conclusions

The hypothalamus plays a major role in the regulation of systemic energy balance by regulating feeding behavior and energy expenditure. Our knowledge of the neuronal and molecular mechanisms underlying the control of thermogenesis has been expanded in recent years by the identification of many neuronal and peripheral signals that interact together to form a network that ensure tight regulation of energy
The new knowledge about the processes that regulate thermogenesis, especially the findings that brown fat exist in humans, has generated great enthusiasm and hope to identify new pharmacological targets against obesity and associated metabolic alterations. The potential and serious side effects of activating a major and powerful system, such as the ANS must be taken also in consideration. In line with this, it is important to highlight that most of the studies focused on the central control of thermogenesis do not assess the possible effects of these signals on cardiovascular system, which is highly influenced by the ANS [158]. For this reason, strategies aiming to target neuronal circuits controlling the thermogenic activity of BAT and beige fat must have a high specificity in order to avoid undesired deleterious effects. This highlights the need to understand and define the different routes that travel from the CNS to peripheral organs of the body, before the way through the thermogenic pathways can be exploited safely.

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References

[1] N.Z. Jespersen, T.J. Larsen, L. Pejja, S. Daugaard, P. Homoe, A. Laft, J. de Jong, N. Mathur, B. Cannon, J. Nedergaard, B.K. Pedersen, K. Moller, C. Scheele, A classical brown adipose tissue mRNA signature partly overlaps with brite in the supraclavicular region of adult humans, Cell Metab. 17 (2013) 798–805.
[2] J. Wu, P. Bostrom, L.M. Sparks, L. Ye, J.H. Choi, A.H. Giang, M. Khandekar, K.A. Virtanen, P. Nautila, G. Schaar, K. Huang, H. Tu, W.D. van Marken Lichtenbelt, J. Hoeks, S. Enerback, P. Schrauwen, B.M. Spiegelman, Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human, Cell 150 (2012) 366–376.
[3] A.M. Cypess, L.S. Weiner, C. Roberts-Toler, E. Franquet Elia, S.H. Kessler,
C. Contreras et al.  
Redox Biology 12 (2017) 854–863

[10] C. Magnan, B.E. Levin, S. Luquet, Brain lipid sensing and the neural control of energy balance. Mol. Cell Endocrinol. 438 (2016) 107–115.

[11] R. Lage, J. Forno, R. Nogueiras, C. Dieguez, Hypothalamic AMPK: a canonical regulator of whole-body energy balance. 
Nat. Rev. Endocrinol. 12 (2016) 421–432.

[12] J.A. Boulant, Role of the preoptic-anterior hypothalamus in thermoregulation and fever. Clin. Infect. Dis. 31 (Suppl. 5) (2000) S157–S161.

[13] C. Fuller, B.A. Horwitz, J.M. Horowitz, Shivering and nonshivering thermogenic responses of cold-exposed rats to hypothalamic warming. Am. J. Physiol. 228 (1975) 1519–1524.

[14] J.D. Guieu, J.D. Hardy, Effects of heating and cooling of the spinal cord on preoptic unit activity. J. Appl. Physiol. 29 (1970) 675–683.

[15] I. Imai-Matsumura, K. Matsumura, T. Nakayama, Involvement of ventromedial hypothalamic brown adipose tissue induced by preoptic cooling in rats. Jpn. J. Physiol. 49 (1999) 939–946.

[16] K. Nakamura, Central circuits for body temperature regulation and fever. Am. J. Physiol. Regul. Integr. Comp. Physiol. 301 (2011) R2107–R2128.

[17] K. Nakamura, S.F. Morrison, Preoptic mechanism for cold-defensive responses to skin cooling. J. Physiol. 566 (2008) 2611–2620.

[18] E. Preston, J. Trianda, Neurons containing mRNA encoding glucagon-like peptide 1 (GLP-1), and GLP-2 in the rat hypothalamus. Brain Res. 928 (2002) 113–120.

[19] S. Amir, A. Schiavetto, Injection of prostaglandin E2 into the anterior hypothalamus promotes development of brown adipocytes and thermogenesis in mice. Am. J. Physiol. Regul. Integr. Comp. Physiol. 296 (2009) R831–R843.

[20] X. Zhang, G. Zhang, H. Zhang, M. Karin, H. Bai, D. Cai, Hypothalamic iKBkappa/ER stress and ER stress link overnutrition to energy imbalance and obesity. Cell 135 (2008) 677–687.

[21] M. Schneeburger, M.O. Dietrich, S. Imberon, C. Castano, A. Garcia, Y. Esteban, A. Gonzalez-Franquesa, I.C. Rodriguez, A. Bortolozzi, P.M. Garcia-Roves, R. Gomis, R. Nogueiras, T.L. Horvath, A. Zorzano, M. Claret, Minofin in POMC neurons connects stress with leptin resistance and energy imbalance. Cell 155 (2013) 172–187.

[22] B. Cao, W. Fan, S.F. Morrison, Neurons in the paraventricular nucleus of the hypothalamus promote thermogenesis in response to cold exposure. J. Physiol. 595 (2008) 860–873.

[23] A.A. Steinier, J. Antunes-Rodrigues, L.G. Branco, Role of preoptic sympathetic neurons in hypothalamic heat-stress responses to cold. Neuroscience 39 (1990) 138–142.

[24] F. Yoon, Q. Fong, C. Suda, P.M. Fuller, M.J. Krashes, L. Vong, R.S. Dyer, P.O. Olson, B.B. Lowell, GABAergic RP-Cre neurons in the arcuate nucleus selectively regulate energy expenditure. Cell 151 (2012) 645–657.

[25] J.A. Boulant, Role of the preoptic-anterior hypothalamus in thermoregulation and fever. Clin. Infect. Dis. 31 (Suppl. 5) (2000) S157–S161.

[26] C. Fuller, B.A. Horwitz, J.M. Horowitz, Shivering and nonshivering thermogenic responses of cold-exposed rats to hypothalamic warming. Am. J. Physiol. 228 (1975) 1519–1524.

[27] J.D. Guieu, J.D. Hardy, Effects of heating and cooling of the spinal cord on preoptic unit activity. J. Appl. Physiol. 29 (1970) 675–683.

[28] K. Nakamura, Central circuits for body temperature regulation and fever. Am. J. Physiol. Regul. Integr. Comp. Physiol. 301 (2011) R2107–R2128.

[29] C. Fuller, B.A. Horwitz, J.M. Horowitz, Shivering and nonshivering thermogenic responses of cold-exposed rats to hypothalamic warming. Am. J. Physiol. 228 (1975) 1519–1524.

[30] J.D. Guieu, J.D. Hardy, Effects of heating and cooling of the spinal cord on preoptic unit activity. J. Appl. Physiol. 29 (1970) 675–683.

[31] K. Nakamura, Central circuits for body temperature regulation and fever. Am. J. Physiol. Regul. Integr. Comp. Physiol. 301 (2011) R2107–R2128.

[32] E. Preston, J. Trianda, Neurons containing mRNA encoding glucagon-like peptide 1 (GLP-1), and GLP-2 in the rat hypothalamus. Brain Res. 928 (2002) 113–120.

[33] S. Amir, A. Schiavetto, Injection of prostaglandin E2 into the anterior hypothalamus promotes development of brown adipocytes and thermogenesis in mice. Am. J. Physiol. Regul. Integr. Comp. Physiol. 296 (2009) R831–R843.

[34] X. Zhang, G. Zhang, H. Zhang, M. Karin, H. Bai, D. Cai, Hypothalamic iKBkappa/ER stress and ER stress link overnutrition to energy imbalance and obesity. Cell 135 (2013) 677–687.

[35] M. Schneeburger, M.O. Dietrich, S. Imberon, C. Castano, A. Garcia, Y. Esteban, A. Gonzalez-Franquesa, I.C. Rodriguez, A. Bortolozzi, P.M. Garcia-Roves, R. Gomis, R. Nogueiras, T.L. Horvath, A. Zorzano, M. Claret, Minofin in POMC neurons connects stress with leptin resistance and energy imbalance. Cell 155 (2013) 172–187.

[36] B.H. Ruan, M.O. Dietrich, Z.W. Liu, M.R. Zimmer, M.D. Li, J.P. Singh, K. Zhang, R. Yin, J. Wu, T.L. Horvath, X. Yang, O-GlcNAc transferase enables AgRP neurons to suppress bwarming of white fat. Cell 159 (2014) 306–317.

[37] T. Lu, T. Hovatta, S.F. Morrison, Glutamate receptors in the raphe pallidus mediate brown adipose tissue thermogenesis evoked by activation of dorsomedial hypothalamic neurons. Neuropharmacology 51 (2006) 426–437.

[38] A.A. Steinier, J. Antunes-Rodrigues, L.G. Branco, Role of preoptic sympathetic neurons in hypothalamic heat-stress responses to cold. Neuroscience 39 (1990) 138–142.

[39] F. Yoon, Q. Fong, C. Suda, P.M. Fuller, M.J. Krashes, L. Vong, R.S. Dyer, P.O. Olson, B.B. Lowell, GABAergic RP-Cre neurons in the arcuate nucleus selectively regulate energy expenditure. Cell 151 (2012) 645–657.

[40] S. Amir, A. Schiavetto, Injection of prostaglandin E2 into the anterior hypothalamus promotes development of brown adipocytes and thermogenesis in mice. Am. J. Physiol. Regul. Integr. Comp. Physiol. 296 (2009) R831–R843.

[41] X. Zhang, G. Zhang, H. Zhang, M. Karin, H. Bai, D. Cai, Hypothalamic iKBkappa/ER stress and ER stress link overnutrition to energy imbalance and obesity. Cell 155 (2013) 677–687.
adipose tissue in spite of systemic leptin resistance, J. Neurosci. 31 (2011) 12189–12197.

[62] M. Bamshad, V.T. Aoki, M.G. Adikson, W.S. Warren, T.J. Bartness, Central nervous system origins of the sympathetic nervous system outflow to white adipose tissue. Am. J. Physiol. 275 (1998) R291–R299.

[63] M. Bamshad, C.K. Song, T.J. Bartness, CNS origins of the sympathetic nervous system outflow to brown adipose tissue. Am. J. Physiol. 276 (1999) R1569–R1578.

[64] G. Cano, A.M. Passerin, G. Schilts, J.P. Card, O. Sano, S. Tovar, D. Fernandez-Nievas, R. Lage, P.B. Martinez de Morentin, I. Gonzalez-Garcia, L. Martins, R. Lage, D. Fernandez-Nievas, N. Martinez-Sanchez, J.M. Moreno-Navarrete, C. Contreras, E. Rial-Pensado, M. Tanida, N. Yamamoto, T. Shibamoto, K. Rahmouni, Involvement of hypothalamic AMPK and fatty acid metabolism mediate thyroid regulation of energy balance, Nat. Med. 16 (2010) 1001–1008.

[65] P.B. Martinez de Morentin, I. Gonzalez-Garcia, L. Martins, R. Lage, D. Fernandez-Mallo, N. Martinez-Sanchez, F. Ruiz-Pino, J. Liu, D.A. Morgan, L. Pinilla, R. Gallego, A.K. Saha, A. Kalibek, E. Filers, P.H. Bichhop, C. Dieguez, R. Nogueiras, K. Rahmouni, M. Tena-Sempere, M. Lopez, Estradiol regulates hypothalamic and peripheral lipid metabolism in a nutritional-depen-
dent fashion, Endocrinology 149 (2008) 455–454.

[66] B.J. Oldfield, M.G. Piggott, P.J. Williams, I. Gonzalez-Garcia, R.R. Sessle, T. Kawai, R. Gallego, C. Fernández-Santos, Z. Zhang, H. Zheng, Orexin inputs to caudal raphe neurons involved in thermal, cardiovascular, and gastrointestinal thermogenesis, J. Neurosci. 31 (2011) 15944–15955.

[67] D. Sellayah, C.J. Madden, D. Tapoun, An orexinergic projection from paraf-inal hypothalamic to raphe pallidus increases rat brown adipose tissue thermogenesis, J. Neurosci. 31 (2011) 15944–15955.

[68] C.J. Madden, D. Tapoun, An orexinergic projection from parafinal hypothalamic to raphe pallidus increases rat brown adipose tissue thermogenesis, J. Neurosci. 31 (2011) 15944–15955.
[172] Y. Jeanson, A. Carriere, L. Castella, A. New Role, for browning as a redox and stress Adaptive mechanism? Front. Endocrinol. 6 (2015) 158.

[173] R.S. Ahima, Y. Qi, N.S. Singhal, M.B. Jackson, P.E. Scherer, Brain adipocytokine action and metabolic regulation, Diabetes 55 (Suppl. 2) (2006) S145-S154.

[174] A. Rodriguez, S. Esquerrro, L. Menendez-Gimenez, S. Becerril, G. Fruhbeck, Revisiting the adipocyte: a model for integration of cytokine signaling in the regulation of energy metabolism, Am. J. Physiol. Endocrinol. Metab. 309 (2015) E691–E714.

[175] P. Villarnoya, A. Vidal-Puig, Beyond the sympathetic tone: the new brown fat activators, Cell Metab. 17 (2013) 638–643.

[176] P. Lee, C.D. Werner, E. Kebebew, F.S. Celi, Functional thermogenic beige adipogenesis is inducible in human neck fat, Int. J. Obes. 38 (2014) 170–176.

[177] M.E. Lidell, M.J. Betz, O. Dalhqvist Leinhard, M. Heglind, L. Elander, M. Slawik, T. Mussack, D. Nilsson, T. Romu, P. Nuutila, K.A. Virtanen, F. Beuschlein, A. Persson, M. Borga, S. Enerback, Evidence for two types of brown adipose tissue in humans, Nat. Med. 19 (2013) 631–634.

[178] L.Z. Sharp, K. Shinoda, H. Ohno, D.W. Scheel, E. Tomoda, L. Ruiz, H. Hu, L. Wang, Z. Pavlova, V. Gilsanz, S. Kajimura, Human BAT possesses molecular signatures that resemble beige/brite cells, PloS One 7 (2012) e49452.

[179] A.J. Whittle, M. López, A. Vidal-Puig, Using brown adipose tissue to treat obesity - the central issue, Trends Mol. Med. 17 (2011) 405–411.

[180] W.D. van Marken Lichtenbelt, J.W. Vanhommerig, N.M. Smulders, J.M. Drossaerts, G.J. Kemerink, N.D. Bouvy, P. Schrauwen, G.J. Teule, Cold-activated brown adipose tissue in healthy men, N. Engl. J. Med. 360 (2009) 1500–1508.

[181] M.C. Zingaretti, F. Crosta, A. Vitali, M. Guerrieri, A. Frontini, B. Cannon, J. Nedergaard, S. Cinti, The presence of UCP1 demonstrates that metabolically active adipose tissue in the neck of adult humans truly represents brown adipose tissue, FASEB J. 23 (2009) 3113–3120.

[182] J. Nedergaard, T. Bengtsson, B. Cannon, Unexpected evidence for active brown adipose tissue in adult humans, Am. J. Physiol. Endocrinol. Metab. 293 (2007) E444-E452.

[183] A.M. Cypess, S. Lehman, G. Williams, I. Tal, D. Rodman, A.B. Goldfine, P.C. Kuo, E.L. Palmer, Y.H. Tseng, A. Doria, G.M. Kolodny, C.R. Kahn, Identification and importance of brown adipose tissue in adult humans, N. Engl. J. Med. 360 (2009) 1509–1517.

[184] K.A. Virtanen, M.E. Lidell, J. Orava, M. Heglind, R. Westergren, T. Niemi, M. Taittonen, J. Laine, N.J. Savisto, S. Enerback, P. Nuutila, Functional brown adipose tissue in healthy adults, N. Engl. J. Med. 360 (2009) 1518–1525.

[185] A.M. Cypess, C.R. Haft, M.R. Laughlin, H.H. Hu, Brown fat in humans: consensus points and experimental guidelines, Cell Metab. 20 (2014) 408–415.

[186] A. Bartelt, O.T. Bruns, R. Reimer, H. Hohenberg, H. Ittrich, K. Peldschus, M.G. Kaul, U.I. Tromsdorf, H. Weller, C. Waurisch, A. Eychmuller, P.L. Gordts, F. Rusinger, K. Bruegelmans, B. Freund, P. Nielsen, M. Merkel, J. Heeren, Brown adipose tissue activity controls triglyceride clearance, Nat. Med. 17 (2011) 200–205.

[187] S.C. Gunawardana, D.W. Piston, Reversal of type 1 diabetes in mice by brown adipose tissue transplant, Diabetes 61 (2012) 674–682.

[188] C.W. Meyer, M. Willenauer, M. Jastroch, B.C. Rouze, T. Fromme, R. Oelkrug, G. Heldmaier, M. Klingenpor, Adaptive thermogenesis and thermal conductance in wild-type and UCP1-KO mice, Am. J. Physiol. Regul. Integr. Comp. Physiol. 299 (2010) R1396–R1406.

[189] K.Y. Chen, R.J. Brychta, J.D. Linderman, S. Smith, A. Courville, W. Dieckmann, P. Herscovich, C.M. Millo, A. Remaley, P. Lee, F.S. Celi, Brown fat activation mediates cold-induced thermogenesis in adult mice in response to a mild decrease in ambient temperature, J. Clin. Endocrinol. Metab. 98 (2013) E1218–E1223.

[190] M. Chondronikola, E. Volpi, E. Borsheim, C. Porter, P. Annamalai, S. Enerback, M.E. Lidell, M.K. Saraf, S.M. Labbe, N.M. Hurren, C. Ylanti, T. Chao, C.R. Andersen, F. Cesana, H. Hawkins, L.S. Sidossis, Brown adipose tissue improves whole-body glucose homeostasis and insulin sensitivity in humans, Diabetes 63 (2014) 4089–4099.

[191] T. Yoneshiro, Recruited brown adipose tissue as an antiobesity agent in humans, 123, 2013, pp. 3404–3408.