**Arabidopsis thaliana** SKP1 interacting protein 11 (At2g02870) negatively regulates the release of green leaf volatiles†

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Green leaf volatiles (GLV) are important factors in plant defense, and plant–plant and plant–insect interactions. Kelch-repet containing F-box proteins (KFB) are plant-specific proteins, which recruit substrates for protein degradation. The present study was conducted to evaluate the role of AtSKIP11, an Arabidopsis KFB, in the expression of hydperoxide lyase (HPL) pathway genes and the formation of products by creating transgenic Arabidopsis thaliana plants expressing the AtSKIP11 open reading frame and its antisense sequence. We found that production of C5 volatiles was greatly reduced in these transgenic plants. The production of C6 GLV was reduced by 49% in AtSKIP11 overexpressing plants, whereas the production of these compounds was increased by 12% in the AtSKIP11-antisense suppression plants. Real time quantitative polymerase chain reaction (RT-qPCR) results showed that expression of LOX2, HPL and ADH1 genes was reduced by 1.8, 1.3 and 2.5 fold, respectively, in the AtSKIP11 overexpressing plants, compared with wild-type plants. In the AtSKIP11-antisense plants, expression of LOX2 was reduced 1.2 fold, but HPL and ADH1 genes were up-regulated by 1.2 fold and 1.7 fold, respectively, over the wild-type samples. Our results suggest the involvement of AtSKIP11 in negatively regulating the HPL pathway, especially the expression of ADH1, possibly through degradation of the transcription factors.

**Introduction**

The sessile nature of the plant kingdom does not allow these creatures to move towards a safe or a favourable place. Moreover, these organisms lack the specialized immune system animals use to combat diseases. Alternatively, plants have developed various other mechanisms to combat a variety of threats. One way of doing this is through inducing the production of defense-related proteins and the release of toxic chemicals. These chemical defenses are the products of the secondary metabolism of plants and they obstruct the growth of and potential damage by different pathogens, including viruses, fungi, bacteria and herbivores. Thousands of different kinds of secondary metabolites are synthesized in plants by a large number of inducible pathways. The oxylipin pathway is among the major inducible pathways for the production of defense responses through activation of its genes in response to different biotic and abiotic stresses. These stresses can activate the oxylipin pathway for the release of higher amounts of jasmonates and green leaf volatiles (GLV), which are otherwise released in very low concentrations under normal conditions.

Plant oxylipins constitute a diverse class of bioactive lipids that are produced by the oxidation of α-linolenic acid (ALA, 18:3ω6) and linoleic acid (LA, 18:2ω6) by lipoygenase (LOX) enzymes due to activation of the pathway genes under the stressed conditions. The production of GLV is induced under conditions of stress in order to protect the plants against herbivores and microbial pathogens. The production of these compounds abruptly increases in response to tissue disruption resulting from biotic or abiotic stresses.

The hydroperoxide lyase (HPL) branch of the oxylipin pathway takes its share from the polylsaturated fatty acids (PUFA) hydroperoxide substrates, generated by lipoygenase (LOX) enzymes due to activation of the pathway genes under the stressed conditions. The products of LOX are divided between the two competing branches of the oxylipin pathway in...
a coordinated manner.\textsuperscript{14,15} Allene oxide synthase converts these intermediates to jasmonates and the HPL enzyme releases volatile C9 and C6 aldehydes. These aldehydes are further converted to other aldehydes, alcohols and their acetylated derivatives.\textsuperscript{14} Among the HPL downstream enzymes are included (3Z)-(2E)-hexenal isomerase, responsible for the conversion of Z-3-hexenal to its E-2 isomer, and NAD-dependent alcohol dehydrogenase (ADH), that catalyzes the reduction of aldehydes to their respective alcohols in order to confer higher stability.\textsuperscript{15-17} All these saturated and unsaturated aldehydes and their downstream products synthesized in the plant's vegetative tissue are volatile in nature and are collectively named as GLV. The role of GLV in the plant's response to various stresses, like microbial pathogens and herbivory attack, has been studied since the early 1990s. But they have been getting the special attention of researchers around the world for the last decade and a half because of their emerging role as aerial messengers for plant–plant communication.\textsuperscript{18,19}

The regulation of cellular processes through the degradation of regulatory proteins by the ubiquitin 26S proteasome system (UPS) followed by recognition via specific F-box proteins is a common signaling mechanism. F-box is an emerging family of signaling proteins in eukaryotes and is thought to be one of the largest superfamilies of plant proteins.\textsuperscript{20} The majority of F-box proteins serve as a substrate-recruiting component of SCF (SKP1, Cullin 1, RING-BOX 1 and an F-box protein) complexes. Nearly two decades since the recognition of and giving the name ‘F-box’ to a domain in cyclin-F protein, our understanding of the role of F-box proteins, SCF biology and the molecular mechanisms employed by UPS is steadily developing.\textsuperscript{21} F-box proteins are comprised of a conserved 40–50 amino acids F-box domain on the N-terminal, while the C-terminals contain one or more secondary motifs. These motifs, which include WD40-repeats, leucine rich-repeats (LRR), kelch-repeats and some other motifs, may be the same or variable in a particular protein and are involved in protein–protein interactions.\textsuperscript{22,23} Researchers around the world have identified many F-box proteins since the discovery of cyclin-F, especially in plants. Since hundreds of SCF complexes have been characterized, the number of F-box proteins serving these SCF complexes is also quite large. However, most of these F-box proteins either remain ‘orphans’ or, until now, could not be completely characterized for their function.\textsuperscript{24}

F-box proteins containing the kelch-repeat motifs (KFB) are among the major plant F-box proteins and are considered to be plant specific. The fact that only a couple of KFB have been characterized in non-plant organisms suggests that the KFB subfamily of F-box proteins is exclusively a plant-specific family of genes.\textsuperscript{25,26} This indicates that some very crucial roles have been assigned to these proteins in the plant cell. In recent years, the pace of research involving plant KFB in molecular signaling has advanced at a reasonable rate, but there is still a lot more to be explored. Several KFB have been reported for their signaling roles in plant defense, plant physiology, circadian mechanisms, secondary metabolism, growth and development, but still the majority of these proteins remain ‘orphans’ or under investigation.\textsuperscript{24}

Although several recent studies have highlighted the role of KFB in plant physiology, not even a single KFB has been reported for any role in the HPL branch of the oxylipin pathway.\textsuperscript{27-29} A couple of recent studies identified and characterized a novel KFB gene in Persicaria minor, named PmF-box1. This gene was found to respond to jasmonic acid elicitation and co-up-regulated with PmADH1 and PmLOX1 upon exogenous treatment with jasmonic acid.\textsuperscript{30,33} BLAST searches were run to find a PmF-box1 homolog in Arabidopsis thaliana. The results returned by the BLAST search show that SKP1 interacting protein 11 (SKIP11) possesses the highest (69%) sequence similarity with PmF-box1. AtSKIP11 (At2g02870) is a KFB consisting of 467 amino acid residues with five kelch repeats. Although no specific function has been identified for AtSKIP11 until now, the protein has been found to interact with the Arabidopsis thaliana ASK1 and ASK2 proteins.\textsuperscript{30,31} The present study highlights the role of the AtSKIP11 gene in the HPL branch of the oxylipin pathway for the production of GLV. A full length AtSKIP11 gene sequence was cloned to the pB2GW7 gateway destination vector to form overexpression and antisense suppression constructs. Both the constructs were transferred to Arabidopsis thaliana wild-type (At-WT) plants by Agrobacterium tumefaciens mediated transformation using the floral dip method. Functional analysis was performed on the T2 generation of the transgenic plants, along with the At-WT plants (control plants). Our results have shown that AtSKIP11 overexpressing plants produced a lower amount of GLV as compared to the wild-type Arabidopsis plants. Whereas, the production of GLV was found to be enhanced in the transgenic Arabidopsis plants in which the AtSKIP11 gene transcript was suppressed by the antisense strategy. These results suggest that AtSKIP11 is involved in the biosynthesis of GLV, possibly affecting the transcription factors, and negatively regulates the HPL pathway.

**Experimental**

**Plant material and bacterial cultures**

*Arabidopsis thaliana* ecotype Columbia-0 seeds were obtained from Dr Noor Azmi Shaharudin (Universiti Putra Malaysia). Chemically competent *E. coli* strain TOP10 cells and the GV3101 strain of *Agrobacterium tumefaciens* were obtained from Plant Biotechnology Lab, Universiti Kebangsaan Malaysia.

**Chemicals and reagents**

All the chemicals and reagents used in this study were of analytical grade or higher. The chemicals used for the molecular work and as the reference standards were obtained from Sigma-Aldrich (USA). For the tissue culture work, chemicals were obtained from Duchefa Biochemical B.V. (Netherlands). The QIAquick Gel Extraction Kit and the DNeasy Plant Mini Kit were from Qiagen (UK). The Gateway Cloning kit, PureLink Quick Plasmid Miniprep Kit, Plant RNA Reagent, Platinum Taq DNA Polymerase, Ambion TURBO DNase and DNA Ladders were obtained from Life Technologies (Carlsbad, CA, USA). KAPA SYBR FAST qPCR Kit, KAPA Taq and KAPA Taq HotStart DNA

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Polymerase were from Kapa Biosystems (Wilmington, Massachusetts). Restriction enzymes and the ProtoScript First Strand cDNA Synthesis Kit were from New England Biolabs (MA, USA).

Construction of AtSKIP11 overexpression and antisense recombinant plasmid

a. Isolation of AtSKIP11 gene and its antisense sequence.
The nucleotide sequence of A. thaliana F-box protein, AtSKIP11 was retrieved from the NCBI database with the GenBank ID At2g02870. About 4–5 healthy leaves were cut from the A. thaliana plants and the total RNA was extracted by PureLink Plant RNA Reagent and treated with a turbo DNA-free kit from Life Technologies according to the manufacturer’s protocol. The cDNA was synthesized by a ProtoScript II First Strand cDNA Synthesis Kit (New England Biolabs), according to the kit manufacturer’s protocol.

For the cloning of the AtSKIP11 gene and its antisense sequence, the full length open reading frame was amplified by a standard PCR reaction, using A. thaliana cDNA as the template. The primers for the isolation of the AtSKIP11 gene with flanking attB sites were designed as guided by the Gateway Cloning system and verified by the OligoAnalyzer 3.1 tool from Integrated DNA Technologies (IDT). Using these primer pairs, fragments of 1555 bps were amplified by PCR for each AtSKIP11 open reading frame and its antisense sequence plus Gateway attB sites (Table 1).

The PCR reaction was run with Platinum Taq DNA Polymerase from Life Technologies using Arabidopsis cDNA as the template. Two separate 50 μL PCR reactions were performed on an Eppendorf Mastercycler machine with the following program: initial denaturation of 1 cycle at 94 °C for 2 min; and 30 cycles of: denaturation at 94 °C for 15 s, primers annealing at 55 °C for 30 s, extension at 68 °C for 1 : 30 min and the final hold at 10 °C. The PCR products were analyzed by 1% (w/v) agarose-EtBr gel electrophoresis and purified by a Purelink PCR purification kit (Life Technologies) according to the manufacturer’s protocol. The quality of the purified PCR product was verified by NanoDrop and then stored at −20 °C until the next step.

b. Cloning of AtSKIP11 gene and AtSKIP11-antisense in the gateway destination vector.
The attB sites containing PCR products were cloned into pENTR221, using ‘BP Clonase II’ enzyme mix (Life Technologies) to yield the entry clones pENTR::AtSKIP11 and pENTR::AtSKIP11-antisense. LR clonase reactions were then carried out to transfer the AtSKIP11 and the AtSKIP11-antisense cDNA fragments from the respective entry clones to the gateway destination vector, pB2GW7. Expression clones, pB2GW7::AtSKIP11 and pB2GW7::AtSKIP11-antisense, were constructed as a result of these reactions. Successful transformants were screened by LB agar-spectinomycin selection and verified by colony PCR using the gene-specific primers given in Table 1. Further confirmation of the integration of the AtSKIP11 gene and the AtSKIP11-antisense sequence in the recombinant plasmids was accomplished by RE digestion analysis and sequencing after extracting the plasmids from the positive clones.

Generation of transgenic Arabidopsis thaliana plants

Agrobacterium tumefaciens strain GV3101 harbouring pBGW7::AtSKIP11 and pBGW7::AtSKIP11-antisense recombinant plasmids was used to transform the A. thaliana (ecotype Columbia) plants by the floral dip method.33 The plants were grown under standard conditions and selected by the herbicide ‘glufosinate’ (basta) due to the presence of the bar gene in the recombinant plasmids. The putative transgenic plants, which survived the herbicide treatment, were confirmed for the integration of the transgenes by PCR, using the genomic DNA as the templates. The PCR was run with Kapa Polymerase Master Mix and the primers listed in Table 2.

The PCR Master Mix was prepared as given in the manufacturer’s manual. The following PCR program was employed: 1 cycle of initial denaturation at 95 °C for 3 min; and 30 cycles of: denaturation at 98 °C for 30 s, primers annealing at 62 °C for 30 s, primers extension at 72 °C for 1 : 30 min and 1 cycle of final extension at 72 °C for 3 min. The products of PCR were analyzed by running 1% agarose-EtBr gel electrophoresis and comparing the bands with a 1 kb DNA ladder to confirm the integration of the DNA insert.

Molecular analysis of transgenic A. thaliana

Molecular analysis of transgenic A. thaliana plants was initiated by growing the third generation. Three lines were selected from each type of transgenic plant for the analysis of metabolic products and the gene expression of the HPL pathway. To ensure the growth of only the desired transgenic plants and to avoid the growth of any non-transgenic contaminants, the young seedlings were again subjected to the Basta selection. Confirmed transgenic plants were grown in the growth chamber until six weeks of age under standard conditions before sampling. Two hours prior to sampling, the plants were treated with 250 μM methyl jasmonate aqueous solution to activate the plant defence related gene expression and the production of plant defence compounds. After two hours the samples were

Table 1  Primer sequence used to prepare the cDNA insert in the preparation of construct pBGW7::AtSKIP11 and pBGW7::AtSKIP11-antisense

| Primer                  | Sequence (5′-3′)                                                                 | Amplicon size |
|------------------------|---------------------------------------------------------------------------------|---------------|
| At-SKIP11-attB1 (F)    | GGGGACAAGTTTTGATACAAAAAAGCAGGCTGGTTTGGGATTTGGAATTGGATGTATCTGA                  | 1555 bps      |
| At-SKIP11-attB2 (R)    | GGGGACAAGTTTTGATACAAAAAAGCAGGCTGGTTTGGGATTTGGAATTGGATGTATCTGA                  | 1555 bps      |
| At-SKIP11-antisense    | GGGGACAAGTTTTGATACAAAAAAGCAGGCTGGTTTGGGATTTGGAATTGGATGTATCTGA                  | 1555 bps      |
| At-SKIP11-antitB2      | GGGGACAAGTTTTGATACAAAAAAGCAGGCTGGTTTGGGATTTGGAATTGGATGTATCTGA                  | 1555 bps      |
collected for real-time quantitative PCR (RT-qPCR) and solid phase micro-extraction GC-MS (SPME-GC-MS) analysis.

**RT-qPCR analysis**

Traces of any methyl jasmonate, sprayed two hours before, were adsorbed from the leaf surface using a clean tissue paper. The upper parts of 4–5 healthy leaves (≈ 300 µg) were cut with a clean pair of scissors and the plant material was quickly immediately frozen in liquid nitrogen. Total RNA was then transferred to labelled 1.5 mL RNase free microfuge tubes and adsorbed from the leaf surface using a clean tissue paper. The traces of any methyl jasmonate, sprayed two hours before, were collected for the sample. Total RNA was extracted by using the Purelink Plant RNA Reagent (Life Technologies, USA) according to the manufacturer's instructions. After removing any contaminating DNA, first strand cDNA synthesis was accomplished by a ProtoScript II First Strand cDNA Synthesis Kit (New England Biolabs) using 500 ng of purified total RNA as the template.

Analysis for the expression of Arabidopsis Actin2 (Act2), LOX2, HPL and ADH1 genes was performed by RT-qPCR, using a KAPA Fast qPCR Master Mix (2×), 0.125 µM of each primer and nuclease-free water to a total of 15 µL. The PCR program adjusted for the amplification of target fragments, using a Bio-Rad IQ5 optical detection system, consists of: one cycle at 95 °C for 3 min; and 40 cycles of: 95 °C for 10 s, 60 °C for 30 s and 72 °C for 30 s. The amplification curve program was followed by a melting curve program that consisted of 61 cycles, increasing the temperature in each cycle by 0.5 °C and dwelling for 6 s from 65 °C to 95 °C. The PCR amplification of the reference and the target genes from transgenic A. thaliana cDNA templates, as well as cDNA templates from the wild-type plants, was performed in separate reactions. The cDNA from the wild-type plants was used as a control, to calculate the fold change in gene expression levels by determining the threshold cycle (Ct) values of the target and the calibrator genes from the control and the test samples using the Livak method. The PCR products were also analysed on 1.2% (w/v) agarose gel electrophoresis to identify the amplification of any non-specific sequences.

**Statistical analysis**

Statistical significance was assessed by using GraphPad Prism 5. Data is presented as mean ± SD and the comparison among the three groups was assessed by using one-way ANOVA followed by Dunnett’s multiple comparison test. Differences under P < 0.05 were considered statistically significant.

**Extraction of GLV for SPME-GC-MS analysis**

Pre-labelled 20 mL glass vials from Agilent Technologies (USA) were used to collect the samples for this purpose. In each vial, 4 glass beads were added to disrupt the plant tissue for the release of GLV. About 1–2 g of healthy leaves were cut from each pot of plants with a razor blade and quickly transferred to the respective vials. The leaves were further cut into small pieces by the same blade during shifting of plant material to the vial, so as to facilitate the leaves’ disruption process. The vials were sealed with parafilm and the crimp-top caps were tightened before weighing the vials.

The GLV emitted from the Arabidopsis leaf samples were analysed by extracting these volatiles through the headspace solid-phase micro extraction (HS-SPME) technique and GC-MS, following a recently published protocol from Matsui et al. (2012) with some modifications. The tissues were completely disrupted by vigorously vortexing the vials for 5–10 min. Then the vials were incubated at 25 °C for 10 min followed by the addition of 1 mL of saturated solution of calcium chloride to deactivate any enzyme activity.

| Table 3 Reference and gene-specific primers for RT-qPCR |
| --- |
| **Target gene/primers** | **Sequence (5’–3’)** | **Amplicon size** |
| Act2-forward | CTTGCACCAAGAGCATGAA | 68 |
| Act2-reverse | CCGATCCAGCACCTGTACTTTTCTT | |
| Lox2-forward | CCGTCGACGAGCAAGTTCTGTTCA | 130 |
| Lox2-reverse | CGGGGAACCTCTCTGTTCA | |
| Hpl-forward | CGCAGCTGGGAGAAAAGA | 88 |
| Hpl-reverse | CCGCTGTAACCAAAAGAAGGTCA | |
| Adh-forward | GGATGGTAGCTACGCTTTGGTGT | 130 |
| Adh-reverse | GGATAGGGAAGCTGTGGGCTT | |
SPME-GC-MS analysis
GLV formed by completely disrupted plant material were analyzed by exposing an SPME fiber (50/30 mm DVB/Carboxen/PDMS, Sigma-Aldrich, USA) in the headspace of the vials at 40 °C for 30 min. Subsequently, the fiber was inserted into the insertion port of a GC (Agilent 7890A) containing a 0.25 mm by 30 m BP20 (WAX) column [SGE Analytical Science, Australia] for thermal desorption of the volatiles. The GC was directly coupled to the MS of an Agilent 5975C inert MSD with a triple-axis detector. To completely remove any compound absorbed on the fiber matrix, the fiber was held in the injection port for 10 min. The temperature of the injector was 200 °C and the interface was at 230 °C. The column temperature program followed was: 40 °C for 1 min, rising at a rate of 15 °C per min to 230 °C and then kept there for 1 min. Helium was used as the carrier gas and delivered at a flow rate of 1 mL per min. The analysis was performed by splitless injection with a sampling time of 1 min. The MSD Chemstation was used to find all the peaks in the raw GC chromatogram. A library search was carried out for all the peaks using NIST/EPA/NIH version 2.0, and the results were combined in a single peak table. Among all the GLV, Z-3-hexen-1-ol can serve as the best compound for the analysis of the HPL pathway because it is the first C6 alcohol formed in the pathway. A standard curve was constructed for Z-3-hexen-1-ol consisting of four concentration points at 5, 50, 500 and 1500 nanomolar (nM) concentrations.

Results and discussion
The physiological role of AtSKIP11 in the Arabidopsis HPL pathway was determined by generating transgenic A. thaliana plants, integrating the overexpression and the antisense suppression constructs of the AtSKIP11 gene. Three of the transgenic lines from each of the AtSKIP11 overexpressing plants and the AtSKIP11-antisense plants were selected for further analysis by RT-qPCR and SPME-GC-MS. Wild-type plants were also grown along with the transgenic plants. Seeds from each of the above-mentioned lines were planted on soil and the plants grown for six weeks under standard conditions. The plants were treated with methyl-jasmonate, to activate the Arabidopsis defence-related pathways, including the oxylipin pathway genes, two hours prior to collecting the samples for analysis. The leaf samples were analyzed to measure any changes in the levels of oxylipin gene expression and the GLV formed in transgenic plants against the control plants.

RT-qPCR analysis
Total RNA was extracted from 100 mg of the leaf powder by PureLink Plant RNA Reagent (Life Technologies), according to the manufacturer’s protocol. cDNA was synthesized with a ProtoScript II First Strand cDNA Synthesis Kit (New England Biolabs), according to the manufacturer’s protocol, using 500 ng of purified total RNA as the template. The cDNA product was diluted with nuclease-free water to a final concentration of 2 ng μL⁻¹, based on the concentration of the RNA.

The cDNAs of the AtSKIP11 overexpressing and the AtSKIP11-antisense sequence expressing transgenic lines were used as templates to determine any change in the gene expression levels by RT-qPCR, using a pair of gene-specific primers for the target genes and the calibrator gene (reference gene). The relative expression of the genes of the HPL pathway was calculated by averaging the threshold cycle (CT) values of each line and the fold changes in the expression levels of the target genes were calculated by applying the 2⁻DDCT method (Appendix A, ESI†). Changes in the gene expression levels for the transgenic plant cDNAs were plotted on a graph to highlight the differences (Fig. 1).

Reduction in the expression of LOX2, HPL and ADH1 was shown by the AtSKIP11 overexpressing plant cDNA templates. The highest change was observed in the expression of ADH1, in which more than 2-fold down-regulation was recorded as compared with the control samples. On the other hand, the plants expressing the AtSKIP11-antisense sequence gave a different pattern of expression. In these cDNA samples, LOX2 was down-regulated by about 1.2-fold, while the HPL expression was increased by the same value and the ADH1 was up-regulated by more than 1.7-fold as compared to control plants. The data assessed by using one-way ANOVA followed by Dunnett’s multiple comparison Test shows that the difference in expression levels of ADH1 is statistically significant with a p-value of 0.0013.

SPME-GC-MS analysis of GLV
Products of the HPL pathway were analyzed by GC-MS after extracting the volatiles, produced from the transgenic A. thaliana plants integrating the AtSKIP11 gene and the AtSKIP11-antisense sequence by HS-SPME. The GC-MS chromatograms yielded well-resolved peaks for different volatiles with similar patterns (Appendix B, ESI†).

Amounts of different GLV produced by each line of transgenic A. thaliana plants and the wild-type A. thaliana plants were
calculated by averaging the mass abundance of each compound per gram of fresh leaf material from the respective chromatograms and the RSD was determined (Appendix B, ESI†). Average mass abundance values of each volatile compound for the transgenic and control plants are computed in Table 4 along with their retention times.

The amounts of different GLV per gram of fresh leaf material and their RSD values were plotted on a graph, to compare the levels of volatiles produced in the transgenic plants and the wild-type plants (Fig. 2 and 3). The synthesis of all target GLV compounds was found to be greatly reduced in the AtSKIP11 overexpressing plants as compared to the control plants, with the highest reduction observed in the formation of two C5 GLV, (E)-2-pentenal and 1-penten-3-ol (Fig. 2). The AtSKIP11-antisense sequence expressing plants also produced lower amounts of some of the volatile compounds, except (Z)-3-hexen-1-ol and 1-hexanol that were either produced in higher concentrations in the transgenic plants or remained almost the same in both types of plants, respectively (Fig. 3). The amounts of (Z)-3-hexen-1-ol released by the AtSKIP11 overexpressing plants was about 55% less, but that released by the AtSKIP11-antisense sequence expressing plants was about 20% higher than that produced by the control plants. Concentrations of (Z)-3-hexen-1-ol, the key GLV in this study, released by the AtSKIP11 overexpressing and the AtSKIP11-antisense expressing transgenic plants were calculated to be 89 nM and 313 nM per g of fresh leaf, respectively. Shiojiri et al. (2012) reported that the maximum concentration of (Z)-3-hexenal, (E)-2-hexenal, and (Z)-3-hexenol released by completely damaged tissue was calculated to be 496 nM GLV per g of the fresh weight. The reduction in the levels of C5 volatiles to a very significant degree show that the gene may be affecting LOX1 more than LOX2 in the Arabidopsis. The genes involved in the biosynthesis of C5 volatiles have not been identified until now and, moreover, synthesis of C5 volatiles is found to be independent of HPL. Shen et al. (2014) have elucidated the role of TomloxC, a tomato 13-LOX in the biosynthesis of C5 volatiles. Arabidopsis LOX1 catalyses the oxygenation at carbon number 9 of the hydrocarbon backbone in LA and ALA to yield (9S,10E,12Z)-9-hydroperoxyoctadeca-10,12-dienoic acid (9-HPOD) and (9S,10E,12Z,15Z)-9-hydroperoxy-10,12,15-octadecatrienoic acid (9-HPOT), respectively. The AtSKIP11 protein has five kelch-repeats and interacts with the substrate proteins possibly through the blades of its β-propeller, thus forming a SCFAt2g02870 (SCFAtSKIP11) complex.

**Table 4** Retention times and the abundance of different volatile compounds in the control plants and the transgenic plants overexpressing the AtSKIP11 gene and AtSKIP11-antisense sequence, obtained by averaging the data from GC-MS chromatograms

| GLV            | RT (min) | At-WT plants       | AtSKIP11 overexpress | AtSKIP11 antisense |
|----------------|---------|--------------------|----------------------|-------------------|
| Hexanal        | 4.80    | 47 282 321         | 18 185 543           | 31 974 550        |
| (E)-2-pentenal | 6.27    | 87 611 380         | 29 430 781           | 62 141 721        |
| 1-Penten-3-ol  | 7.60    | 313 233 340        | 47 704 923           | 102 645 089       |
| 1-Pentanol     | 10.36   | 14 968 059         | 7 383 709            | 9 437 418         |
| 1-Hexanol      | 13.42   | 64 605 639         | 28 187 494           | 61 574 701        |
| (Z)-3-Hexen-1-ol | 14.27 | 274 028 143        | 124 994 804          | 337 579 587       |

The overall picture obtained from the results of these transgenic plants has highlighted the involvement of AtSKIP11 in the HPL branch of the oxylipin pathway. The protein is suggested to up-regulate a certain portion of the pathway through degradation of some transcription factors or other regulatory
proteins. Overall, these results suggest that AtSKIP11 is involved at some stage, most probably downstream of the HPL enzyme, of the HPL biosynthetic pathway in a ‘negatively regulating’ manner. This means that higher the levels of AtSKIP11, the lower will be the production of GLV and vice versa. Recently, a few KFBs have been reported to participate in plant physiological phenomena by degrading the inhibitors of the pathway. In a recent report, a novel KFB encoding gene from rice, an inhibitor for brown furrows1 (IBF1) was cloned to show that IBF1 is involved, as a negative regulator in the biosynthesis of flavonoids and to reduce the accumulation of brown pigments in rice hulls.43 Similarly, a KFB encoding gene in rice, LARGER PANICLE (LP) was found to be involved in the regulation of panicle architecture. This study reported that LP is an endoplasmic reticulum localized protein and possibly modulates the cytokinin levels by interacting with rice SKP1-like protein. Moreover, the mutant plants were found to have increased panicle size, better plant architecture and improved grain yield per plant.44 The role of the KMD family of proteins was recently reported in the regulation of Arabidopsis cytokinin signalling. In the SCF\textsuperscript{KMD} complex, KMD proteins were suggested to physically interact with some key transcription factors in the cytokinin signalling. KMD loss of function mutants were more sensitive to cytokinin, while transgenic lines overexpressing KMD were insensitive to cytokinin, showing that KMD proteins negatively regulate the cytokinin signalling in Arabidopsis.45 Recently, a couple of similar studies have highlighted the role of a family of four similar KFBs in Arabidopsis in the regulation of the phenylpropanoid biosynthetic pathway. Zhang et al. (2013 and 2014) have shown that these proteins regulate the biosynthesis of phenylpropanoids by interaction with and proteolytic turn-over of four phenylalanine ammonia-lyase (PAL) isozymes that catalyse the first and the committed step in phenylpropanoid biosynthesis.45,46

Using the results of present study in the context of these published results, a mechanism for the function of AtSKIP11 is proposed here. AtSKIP11 interacts with SKP1 through its N-terminal F-box domain and interacts with the target proteins, like LOX2, HPL, ADH1 or the regulatory elements of these genes, through its C-terminal domain. The C-terminal of AtSKIP11 consists of a five kelch-repeats containing domain, which is thought to form a bladed β-propeller. Each blade of the β-propeller consists of a single kelch-repeat that is folded into four sheets and may be capable of independently interacting with other different proteins.25 Therefore, it is suggested that multiple interacting partners may exist for the SCF\textsuperscript{AtSKIP11} complex. The results of the present study show that some proteins, including the enzymes of the oxylipin pathway do interact with the SCF\textsuperscript{AtSKIP11} complex. Degradation of these proteins, that may be enzymes themselves or regulatory proteins, such as transcription factors, reduce the formation and release of certain GLV, as shown by the transgenic plants overexpressing the AtSKIP11 gene. The levels of various GLV were found to be reduced in these plants and the expression of the HPL pathway genes was also found to be decreased as compared with the control plants (Fig. 1–3). On the other hand, in the transgenic plants expressing the antisense suppression construct, levels of some GLV were found to be enhanced and also the expression of the LOX2 and HPL genes was slightly up-regulated, while the expression of ADH1 was significantly up-regulated (Fig. 1–3). That means that the suppression of the translation of the AtSKIP11 gene transcript leads to formation of a reduced amount of SCF\textsuperscript{AtSKIP11} complex. This is in accordance with the results of some recent studies, in which certain KFB overexpressing plants resulted in the down-regulation of the metabolic pathways under study while the mutant plants came up with up-regulation of the pathways.44 As a result, the degradation of the protein targets for the SCF\textsuperscript{AtSKIP11} complex will be decreased. The reduced or the retarded protein degradation, of either the enzymes or the transcription factors, will allow the production of more GLV and the up-regulation of certain enzymes of the pathway. According to our proposed mechanism, the impaired or lower expression of AtSKIP11 leads to decreased production of SCF\textsuperscript{AtSKIP11} complex and thus less degradation of GLV-producing proteins.

It is obvious from the literature review of the GLV biosynthetic pathway that the pathway is quite complex. Release of PUFAs by the degradation of membrane lipids is still not completely understood. LA and ALA, released by phospholipases, serve as substrates for the oxylipin pathway, but there may exist some other PUFA candidates as well. Although LOX2 contributes more towards the generation of oxygenated intermediates, the role of the regulatory components switching between the two types of LOX may also contribute significantly. The most important step in the pathway is the distribution of oxygenated substrates between the HPL and the AOS branches of the pathway. The importance of jasmonic acid is not only to rescue the plant during any type of stress, but it is involved in almost every aspect of plant growth and development.9 While only trace amounts of GLV are released by plants under normal physiological conditions, they are formed very rapidly under stressful conditions. Numerous studies have suggested that the two competing branches of the oxylipin pathway crosstalk during metabolism and the stress response.14 The role of the regulatory proteins at this stage of the pathway is of great importance, in which SCF complexes may be very crucial. The degradation of certain proteins will allow the pathway to switch between the two branches of the pathway. Among the downstream genes, ADH1 appears to be more responsive towards the KFB of the present study. Either ADH1 itself or the transcription factors that enhance its expression seem to be degraded by over-expressed AtSKIP11 (Appendix C, ESI†).

**Conclusions and future prospects**

The results of the analysis of transgenic plants manipulating the AtSKIP11 gene, which is endogenous to Arabidopsis and its antisense suppression sequence, suggest a role for the KFB AtSKIP11 in the Arabidopsis HPL branch of the oxylipin pathway. The release of a higher concentration of GLV under stressed conditions led us to conclude that gene expression for the GLV biosynthetic pathway is induced by signals from infection or stress. Therefore, the nature of stress signals and the mechanisms by which these signals give rise to the generation of the
defence response remain elusive. Although many studies have shown encouraging results, most are limited to laboratory experiments that were performed using synthetic GLV. It is the need of the hour to devise new strategies and to design novel substrates for identifying the ‘missing link’ between the perception of a stress signal and the generation of a defence response. The proposed mechanism from our results suggests that the reduced expression of AtSKIP11 leads to decreased production of the SCF<sup>AtSKIP11</sup> complex and thus less degradation of GLV-producing proteins and vice versa. So, AtSKIP11 may be one of the ‘missing links’ in the Arabidopsis thaliana HPL pathway and may serve as a molecular switch for controlling the production of GLV.

**Conflicts of interest**

The authors declare no conflicts of interest.

**Abbreviations**

ALA  α-Linolenic acid.

DES Divinyl ether synthase.

DGL DONGLE.

GLV Green leaf volatiles.

HIPVs Herbivore-induced plant volatiles.

13-HOT 13-Hydroxytridecanoic acid.

13-(9Z,11E,13S)-13-Hydroperoxyoctadeca-9,11-dienoic acid.

HPOT trienoic acid.

LA Linoleic acid.

PUFAs Polyunsaturated fatty acids.

KFBs Kelch-repeat containing F-box proteins.

MeJA Methyl jasmonate.

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