Pseudomonas aeruginosa AES-1 Exhibits Increased Virulence Gene Expression during Chronic Infection of Cystic Fibrosis Lung

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Abstract

Pseudomonas aeruginosa, the leading cause of morbidity and mortality in people with cystic fibrosis (CF), adapts for survival in the CF lung through both mutation and gene expression changes. Frequent clonal strains such as the Australian Epidemic Strain-1 (AES-1), have increased ability to establish infection in the CF lung and to superimpose and replace infrequent clonal strains. Little is known about the factors underpinning these properties. Analysis has been hampered by lack of expression array templates containing CF-strain specific genes. We sequenced the genome of an acute infection AES-1 isolate from a CF infant (AES-1R) and constructed a non-redundant micro-array (PANarray) comprising AES-1R and seven other sequenced P. aeruginosa genomes. The unclosed AES-1R genome comprised 6,254Mbp and contained 6957 putative genes, including 338 not found in the other seven genomes. The PANarray contained 12,543 gene probe spots: comprising 12,147 P. aeruginosa gene probes, 326 quality-control probes and 70 probes for non-P. aeruginosa genes, including phage and plant genes. We grew AES-1R and its isogenic pair AES-1M, taken from the same patient 10.5 years later and not eradicated in the intervening period, in our validated artificial sputum medium (ASDMD) and used the PANarray to compare gene expression of both in duplicate. 675 genes were differentially expressed between the isogenic pairs, including upregulation of alginate, biofilm, persistence genes and virulence-related genes such as dihydroorotase, uridylate kinase and cardioplin synthase, in AES-1M. Non-PAO1 genes upregulated in AES-1M included pathogenesis-related (PAGI-5) genes present in strains PACS2 and PA7, and numerous phage genes. Elucidation of these genes’ roles could lead to targeted treatment strategies for chronically infected CF patients.

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Introduction

In cystic fibrosis (CF) patients, chronic Pseudomonas aeruginosa infection and inflammation are associated with biofilm formation, a progressive decline in lung function and premature death [1,2,3]. Early infection of CF infants with P. aeruginosa is usually successfully managed by aggressive antibiotic therapy; however by adolescence most patients have chronic infection. Prominent persisters include the frequent clones Liverpool Epidemic Strain (LES), Manchester strain (c3719), and the Australian Epidemic Strain-1 (AES-1), which infects ca. 40% of adult CF patients in Sydney and Melbourne [4,5]. While frequent clones are often more resistant to antibiotics than infrequent clones, they lack a distinctive antibiotic resistance profile [6], indicating that other factors are involved in their persistence.

P. aeruginosa undergo a number of genetic and expression changes that assist in their ability to survive in the CF lung and to evade detection and clearance by the immune system [7,8]. Acute lung infection in CF patients is associated with expression of virulence determinants that are involved in establishment of infection in animal model systems [9,10,11,12]. However, P. aeruginosa from chronically-infected CF patients usually lack some of these virulence determinants, suggesting that these genes are unnecessary for long-term maintenance of P. aeruginosa infection in vivo [13,14,15].

Genomic studies of two closely related P. aeruginosa strains collected from a CF patient 7.5 years apart [14] showed loss of function mutations in virulence genes required for O-antigen biosynthesis, Type III secretion (T3SS), twitching motility, exotoxin A regulation, multidrug efflux, osmotic balance, phenazine biosynthesis, quorum sensing, and iron acquisition. Chronic infection strains also possess particular characteristics, including large chromosomal inversions (LCI) [16], a specific Type IV pilin allele [17], exhibit enhanced biofilm dispersal properties...
[18] and a progressive loss of T3SS function over time [19]. In vivo conditions can also result in genomic changes to persistent strains and this is probably related to strain hypermutability [20,21].

Persistence may also be related to changes in levels of expression of particular genes. Conversion to the mucoid phenotype, which is dependent on biofilm formation, has been associated with establishment of chronic infection [22]. Chronic growth also appears to activate new virulence strategies, including the catabolism of fatty acids such as phosphotidylcholine and prostaglandin, thus preventing their utilisation by the host [12]. A study of sequential isogenic P. aeruginosa isolates from three adults with chronic infection showed upregulation of anaerobic respiration, microaerobic respiration and the TCA cycle pathways [23], however there have been no studies comparing the gene expression changes between isogenic acute infection and chronic infection isolates. Additionally, the sequencing of the acute AES-1 isolate AES-1R and the use of an array encompassing eight P. aeruginosa genomes has allowed detection of novel genes not present in the PAO1 genome.

This study has used an artificial sputum media closely mimicking CF sputum (ASMDM) [24] and an array based on the genome sequence of AES-1R and other CF isolates (PANarray) to identify the gene expression changes in sequential early and chronic isogenic AES-1 isolates.

Results

Homology between AES-1 and other P. aeruginosa genomes

In order to identify all coding sequences (CDS) in the eight P. aeruginosa genomes (AES-1, PAO1, PA7/PSPA7, PA14, PACS2, Pa2192, Pac/PALES and c3719), BLAST analysis was performed on the AES-1R sequence, which produced 7672 clusters, of which 3962 represented CDS common to all strains, while 2710 were specific to AES-1R genes. The region between AES_6966 and AES_7152 are unique to AES-1R (E value less than 10^-4). Two phages (AES_7010, 7011 and 7084), the bacteriophage P2 tail protein and the heme exporter protein were putative or probable phage functions. In general, the genomes of PACS2 (41.8%), PAO1 (41.8%) and c3719 (41.4%), (Pearson’s χ² test: p = 0.0014, p = 0.017 and p = 0.047, respectively), in terms of cluster of orthologous groups (COG) function (not shown), a comparison of AES-1R with PAO1, PA14, Pa2192 and PA7 showed no significant differences in group size (p<0.05) with the exception of inorganic ion transport and metabolism, where AES-1R had a significantly greater number of CDS (308 against 291) compared to PA7 (p = 0.035).

AES-1R had the highest number of unique CDS of the eight genomes (336), though as a percentage of total CDS, AES-1 was at the same level as P. aeruginosa Pa2192 (Table 1). The unique AES-1R genes coded mainly for hypothetical proteins, but also included several characterized proteins such as the SOS-response transcriptional repressor lexA (AES_7031), three Mu-like bacteriophages (AES_7010, 7011 and 7084), the bacteriophage P2 tail protein, a heme exporter protein and polyhydroxyalkanoate synthesis protein pelF (Table S1). Fully 67.4% of CDS in the region between AES_6966 and AES_7152 are unique to AES-1R (E value less than 10^-4 for all homologs), indicating that this 107 CDS region may be part of the AES-1R accessory genome. While most CDS in this region are hypothetical proteins, 16 have putative or probable phage functions.

Genotyping and growth characteristics of CF strains in ASMDM

Pulsed field gel electrophoresis (PFGE) of AES-1R and AES-1M showed no discernible band differences (Fig. 2) therefore they were classed as the same strain. The acute and chronic isolates both had a non-mucoid phenotype on horse blood agar (HBA) and Mueller-Hinton agar (MHA). This is unusual since most chronic isolates are mucoid or revertants from mucoid phenotypes [25,26]. AES-1R and AES-1M may qualify as small colony variants (SCV) due to their small colony size of 2–5 mm after 48 hours, and the upregulation of the polysaccharide genes pelABCDEF due to their small colony size of 2–5 mm after 48 hours, and the upregulation of the polysaccharide genes pelABCDEF in AES-1M may be contributing to this morphotype (see Discussion) [27]. As shown in Fig. 3, there is a difference in the growth patterns of AES-1R and AES-1M. AES-1R grew with a thickened pellicle and projections into the media while AES-1M showed no discernable band differences (Fig. 2) therefore they were classed as the same strain. The acute and chronic isolates both had a non-mucoid phenotype on horse blood agar (HBA) and Mueller-Hinton agar (MHA). This is unusual since most chronic isolates are mucoid or revertants from mucoid phenotypes [25,26]. AES-1R and AES-1M may qualify as small colony variants (SCV) due to their small colony size of 2–5 mm after 48 hours, and the upregulation of the polysaccharide genes pelABCDEF in AES-1M may be contributing to this morphotype (see Discussion) [27]. As shown in Fig. 3, there is a difference in the growth patterns of AES-1R and AES-1M. AES-1R grew with a thickened pellicle and projections into the media while AES-1M had smaller projections at 72h. When incubated for up to 96h, the growth of AES-1M resembled that of AES-1R (not shown), indicating that it is slower growing during chronic infection. The growth characteristics of AES-1M resembled those of P. aeruginosa UCBPP-PA14 in ASMDM [24], which also grew more slowly with smaller anaerobic projections.

| Bacterial strain | Size (Mbp) | Total CDS | Unique CDS | % of genome |
|------------------|------------|-----------|------------|-------------|
| P. aeruginosa AES-1R | 6.254 | 6957 | 338 | 4.9% |
| P. aeruginosa Pa2192 | 6.826 | 5914 | 284 | 4.8% |
| P. aeruginosa PACS2 | 6.492 | 5676 | 44 | 0.8% |
| P. aeruginosa UCBPP-PA14 | 6.537 | 5896 | 183 | 3.1% |
| P. aeruginosa PAO1 | 6.264 | 5568 | 73 | 1.3% |
| P. aeruginosa c3719 | 6.146 | 5221 | 63 | 1.2% |
| P. aeruginosa PaLES | 6.601 | 5925 | 193 | 3.3% |
| P. aeruginosa PSPA7 | 6.439 | 6286 | 54 | 0.9% |

CDS = Coding sequences.
Total adjusted for adjoining duplicates.
All genes were annotated based on BLAST analysis results, with genes showing an E-value of less than 10^-4 designated as unique.

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Overall changes in expression in AES-1 over time

In all, 675 genes were differentially-expressed between the frequent clone isolates AES-1R and AES-1M at p < 0.05 (Table S2) including 365 upregulated and 310 downregulated in the chronic AES-1M. Five hundred and twenty five of these genes were differentially expressed $\geq 2$fold, while 74 (11.2%) of all differentially expressed genes did not have homologues in the PAO1 genome sequence (Table S3). These genes would not have been detectable on the Affymetrix PAO1 array. They included 10 phage genes and one integrase gene, eight of which were upregulated (average upregulation 3.7fold), two upregulated pathogenesis-related proteins from $P$. aeruginosa gene island-5 (PAGI-5): PaerPA_01000873 (3.8fold) and PSPA7_4490 (5.8fold), the enzyme uridylate kinase (6.4fold), and the downregulated proteins polyketide synthase ($\geq 4.3$fold) and luminal binding protein ($\geq 3.4$fold).

Gene expression differences between acute and chronic infection AES-1 isolates

Against expectations that chronic infection leads to a downregulation of virulence-related genes, a number of genes with virulence functions were upregulated in chronic AES-1M compared to its acute infection counterpart (Table 2). These included alcohol dehydrogenase adaA (5.5fold), the T3SS regulator pscD (3.2fold), uroporphyrinogen decarboxylase hemE (16-fold), peptidyl-prolyl cis-trans isomerase ppcA (2.5-fold), alginate-associated genes algD,E,F,8 and amnZ (1.9 to 10.7fold), dihydroorotase pyrC (12.1fold), uridylate kinase pyrH (6.4fold), the extracellular polysaccharide genes pelC (5.6fold) and pelE (3.7fold) part of the pelABCDE operon, phospholipase N (pelN) (3.2fold) and cardiolipin synthase cls (7.0fold). Genes with putative or probable virulence roles upregulated in chronic AES-1M included a probable haloacid dehalogenase (21.6fold), threonine dehydratase isocA (9.7fold) xanthine phosphoribosyltransferase xpt (16.7fold) and polyhydroxyalkanoate synthesis protein phaF (6.0fold) (Table S2). The downregulation of homogenitase 1,2-dioxygenase hmgA (-7.5fold) is also virulence related since its downregulation de-represses pyomelanin production, which enhances persistence.

Gene expression by quantitative PCR

The average quantitative PCR ratios of the selected virulence-related (Table 2) and other genes correlated well with their microarray expression ratios (correlation coefficient: R$^2 = 0.8053$ – Fig. 4). All genes showed either up- or downregulation consistent with the microarray results, despite eight of these being quantified using different RNA samples for array and qPCR.

Discussion

The sequencing of the genome of the Australian epidemic strain AES-1 (isolate R) has provided the first opportunity to examine the similarities and differences between this frequent clone widespread in eastern Australia and other frequent clones such as PaLES and c3719. In terms of overall size, AES-1R (6.254 Mbp) falls between c3719 (6.146 Mbp) and PaLES (6.601 Mbp), and close to that of PAO1 (6.264 Mbp). As the c3719 and PaLES genomes are closed and the AES-1R genome is not, a direct comparison of CDS is not possible, however in terms of genes of known function there are significant differences between AES-1R and c3719. In subcellular localisation (Fig. 1) AES-1R has a significantly smaller proportion of cytoplasmic protein content (39.7%) compared to PACS2 (41.8%), PAO1 (41.8%) and c3719 (41.4%) (Pearson’s $\chi^2$ test: $p = 0.0014$, $p = 0.017$ and $p = 0.047$, respectively).
Australia), and digested using restriction enzyme SpeI to generate a small number (ca. 15–40) of large DNA fragments. Band patterns were analysed using cluster analysis software (GelComparITM, Applied Maths, Belgium) and established criteria [55] (same genotype if less than two bands difference). The AES-1R and AES-1M genomes fall within these criteria.

A major objective of this study was to utilize the AES-1R sequence and the PANArray to elucidate the differences and similarities in gene expression between the acute and chronic isolates of AES-1. The finding that certain virulence-related genes were upregulated in chronic infection isolate AES-1M has relevance in areas including biofilm development, transmissibility, antibiotic resistance, pyomelanin-related persistence and lung surfactant secretion (Table 2). The upregulation of adhA is required for P. aeruginosa biofilm development [32] and biofilms are important in virulence and persistence. We previously reported that AES-1 strains produce significantly larger biofilms than non-epidemic strains [33] thus the upregulated expression of biofilm-related genes in AES-1M would enhance its ability to persist. T3SS genes are generally switched off or downregulated at chronic infection and several including pscV, pscC and pscI were significantly downregulated in AES-1M; however pscD, a regulatory T3SS gene [34] was significantly upregulated in AES-1M. As a functioning regulator, pscD may trigger the T3SS and enable AES-1M to infect other CF patients from the environment or transmit to other patients via aerosols from an infected patient. ppxA is one of four periplasmic isomerases in E. coli, inactivation of which leads to a decreased growth rate and increased susceptibility to certain antibiotics [35]. It has been shown in E. coli that hemE is the first enzyme transcribed in heme synthesis, and heme synthesis gene expression has been linked to regulation of alginate secretion [36]. While AES-1M appeared non-mucoid on HBA and MHA (see Results), there was significant upregulation of most alginate genes in ASMMD, suggesting that this mucus-like medium triggers their enhanced expression. Alginate is considered a chronic infection virulence factor [12] and is involved in biofilm production and resistance to leukocyte killing [37]. The exception was a significant downregulation of algC in AES-1M. algC is transcribed separately from the other alginate genes, which are in a single operon under the control of algD [38] and has another function as a requirement for lipopolysaccharide (LPS) biosynthesis [39]. Thus downregulation of algC would suggest defects in production of A band LPS in these isolates.
Dihydroorotase (pyrC-upregulated 12.1fold) catalyzes the third of six enzymatic steps in the biosynthesis of uracil monophosphate (UMP) from glutamine and aspartate precursors while uridylate kinase (pyrH upregulated 6.4fold) catalyses the conversion of UMP to UDP. Inhibition of the uracil biosynthetic pathway has been demonstrated to repress biofilms and all three QS pathways (Rhl, Las and Pqs) in P. aeruginosa [40] while Staphylococcus aureus pyrC mutants showed a fivefold reduction in virulence in a BALB/c mouse competitive systemic infection model and threefold in a non-competitive systemic infection model [41]. Of the genes with probable virulence roles upregulated in AES-1M, pel genes are involved in P. aeruginosa biofilm formation [42] and enhance a strain’s capacity to persist. Expression of pel genes by P. aeruginosa has also been shown to competitively disrupt S. aureus biofilms [43]. The phospholipase C precursor plcN is important in extracellular virulence as it degrades phosphatidylcholine, a constituent of lung surfactant [44]. Cardiolipin synthase (cls) plays an important role in membrane fluidity and P. putida cls mutants have shown increased sensitivity to antibiotics [45].

Other genes upregulated in AES-1M included the putative chemotactic transducer pctA (AES_5777 8.2fold) and a P. aeruginosa pathogenicity island-1 (PAPI-1) gene (PA14-59980 5.6fold) also found in UCBPP-PA14. PctA is essential for taxis towards sugars, organic acids and L-amino-acids and in the acquisition of nitrogen

Table 2. Known virulence-related genes differentially expressed in P. aeruginosa AES-1M compared to AES-1R (p<0.05).

| Gene name | FC  | Description | Role in P. aeruginosa |
|-----------|-----|-------------|-----------------------|
| hemE      | 16.1| Uroporphyrinogen decarboxylase | Regulation of alginate secretion |
| pyrC      | 12.1| Dihydroorotase | Biofilm development, QS and virulence |
| pyrH      | 6.4 | Uridylate kinase | Biofilm development, QS and virulence |
| adhA      | 5.5 | Alcohol dehydrogenase | Required for biofilm development |
| peB, peE  | 5.6, 3.7 | Extracellular polysaccharide | Competitive disruption of S. aureus biofilms |
| cls       | 7.0 | Cardiolipin synthase | Increases membrane lipids and AB resistance |
| pscD      | 3.2 | T3SS export protein | Regulator of T3SS - virulence |
| plcN      | 3.2 | Phospholipase C precursor | Degrades phosphatidylcholine in lung surfactant |
| algD,E,F,R,arnZ | 1.9-10.7 | Alginate biosynthesis | Biofilm development |
| ppiA      | 2.5 | Peptidyl-prolyl cis-trans isomerase | ΔppiA-decreased growth- susceptible to AB* |
| hmgA      | -7.2 | Homogentisate 1-2-dioxygenase | Loss induces pyomelanin - greater persistence |
| algC      | -9.3 | Phosphomannomutase | |

*FC = Fold change - indicates up or downregulated in AES-1M relative to AES-1R.
*AB = Antibiotic.

Figure 4. Differential expression of virulence-related genes by microarray and qPCR. Thirteen mainly virulence-related genes significantly differentially expressed by microarray were also quantified for expression by qRT-PCR. Five of these (adhA, algC, AES_6147, AES_2005 and pyrC) were quantified for qPCR using the same RNA sample as for microarray and eight (pelB, cls, algD, ppiA, pscD, hemE, pvcC and hmgA) were quantified using a different RNA sample to that for microarray, and gave an overall correlation coefficient R² = 0.8053.
The presence of PAPI-1 pathogenicity island genes in AES1 and their upregulation is of great interest due to the role of this island in UCBPP-PA14 virulence [47] and its mode of spread to CF strains [48].

Amongst genes downregulated in AES-1M, hmgA and thick stand out. Studies have shown loss of hmgA expression induces pyomelanin production, which in turn leads to increased persistence [49] and we have detected downregulation of HmgA at the protein level in AES-1M. Thioredoxin has been shown to be induced in P. aeruginosa PAO1 biofilms under strict anaerobic conditions [50], yet in our studies P. aeruginosa grown in ASMDM was unlikely to have experienced strictly anaerobic conditions as both surface and subsurface growth has previously been shown to have access to oxygen [24]. Thus AES-1R may be able to cope with hydrogen peroxide-mediated oxidative stress by induction of the thioredoxin pathway, while chronic AES-1M has adapted by not inducing the host cell-mediated response. Iron uptake is critical in the iron-starved ASMDM and CF sputum [24], thus the synthesis of the siderophores such as pyoverdine and pyochelin would be expected to be high in both acute and chronic isolates. Interestingly, in the pyoverdine biosynthesis operon pcvABCD, pcvB was significantly upregulated (3.3fold) while pcvC was significantly downregulated (~7.5fold). A recent study suggests pcvABCD is required not for pyoverdine production but for dihydroxycurcumarin expression [51]. pcvAB makes an intermediate compound which is oxidized to the catechol form by pcvCD. It is possible that the final product is not required by AES-1M, leading to either mutation or downregulation of pcvC.

Hoboth et al. [23] compared an early mutator with an end-stage non-mutator phenotype from the same patient, seeing a downregulation of virulence-related genes including T3SS, chemotaxis, QS and flagellin genes. While that study did not use isogenic strains, there are some findings in common with our study. Several chemotaxis transducers (waspD, AES_2270 and AES_4899), and T3SS genes (pcrV, pscC and pscF) were downregulated in AES-1M, but most genes in these groups were not differentially expressed. Mutation in any of the chemotaxis genes (waspABCDEF) leads to loss of motility and auto-aggregation in P. aeruginosa [52]. Therefore the downregulation of waspD suggests decreased motility and swarming, possibly leading to less expansion and spread of biofilm microcolonies compared to AES-1R. Our phenotypic data (not shown) demonstrate that AES-1M does swarm less than AES-1R, though not significantly (Pearson’s χ² test: p = 0.062) however it swims and twitches significantly more (p = 0.29 and p = 0.015, respectively). With respect to expression of metabolic genes, only one TCA cycle metabolism gene (isocitrate dehydrogenase-idh) downregulated in the end-stage mutator [23], was also downregulated in AES-1M, with the remainder not differentially expressed at p<0.05. Possible reasons for the different metabolic profiles include growth media and conditions, and AES-1-strain specific characteristics. Our study used ASMDM, which resembles the composition of lung sputum, and selected the entire biofilm (anaerobic and aerobic growth) for analysis, compared to microaerobic growth in Laría-Bartani broth used in the Hoboth study.

Conclusions

The sequencing of the AES-1 isolate AES-1R and the use of a broad-capture array has for the first time enabled detection of the expression of AES-1 genes not found in the reference strain PAO1. Furthermore, the use of ASMDM has provided a profile of differential expression of genes under sputum-like conditions. The upregulation of certain virulence-related genes, including biofilm-enhancing, competitive inhibition and persistence genes seen in chronic AES-1M grown in ASMDM offers some explanation for the spread and serious health outcomes associated clinically with AES-1, including person-to-person transmission, more exacerbations and more hospitalisations. In vivo analysis of knockout mutants of differentially expressed genes in persister strains will aid in identifying the factors leading to persistence.

Materials and Methods

Strains used in this study

Ethics approval for this study was given by the University of Sydney Human Research Ethics Committee (Protocol Number: X07-0029, Reference Number 6999). The Australian Epidemic Strain-1 (AES-1, previously known as m16, C3789 or PI), is one of two dominant eastern Australian mainland clonal complexes. AES-1R was isolated from a child aged 14 months at the time as the deaths of five CF-infants infected with AES-1 [53]. AES-1M was isolated from the same patient at 11 years 9 months. AES-1 was not eradicated in the patient in the intervening period (D. Armstrong pers. comm.). Written informed consent was obtained by the Royal Children’s Hospital Melbourne, and The Southern Health Service, Melbourne.

Genotyping of AES-1R and AES-1M

AES-1R and AES-1M were genotyped using pulse field gel electrophoresis (PFGE) [54]. Briefly, bacterial cells embedded in agarose plugs were lysed using EC lysis buffer (Sigma-Aldrich Australia), and digested using restriction enzyme SpeI to generate a small number (ca. 15–40) of large DNA fragments. After electrophoresis of plugs containing digested DNA on 1.2% w/v agar, band pattern analyses were performed using cluster analysis software (GeCompAryTM, Applied Maths, Belgium) and the criteria developed by Tenover et al [55] (different genotype if more than a two band difference).

AES-1R genome sequencing and construction of the PanArray

The AES-1R genome was sequenced using a 454 Genome Sequencer GS20 (Roche Diagnostics, Basel, Switzerland). This produced 598131 reads totalling 58 Mbp, providing approximately nine times coverage of the genome. De novo assembly using the Newbler assembler with default parameters, produced 1968 contigs totalling 6.28 Mbp which were ordered and oriented to the P. aeruginosa PAO1 genome [56]. Putative genes were predicted using GeneMarkS 4.6b [57]. The whole genome shotgun sequence of the AES-1R genome (ID: 64619) is available at http://www.ncbi.nlm.nih.gov/nuccore/AFNF00000000. To examine gene homology amongst the eight P. aeruginosa genomes (AES-1R, PAO1, PA7/PSPA7, PA14, PACS2, Pa_2192, Pae/PLES and c3719), all gene protein sequences were clustered into orthologous groups using OrthoMCL version 1.4 [58]. Genes were assigned an origin based on the highest identity by BLAST analysis as described on the web based annotation system for prokaryotes of the Victorian Bioinformatics Consortium at Monash University (WASABI) http://vbc.med.monash.edu.au/wasabi/. The AES-1R genes, plus all those from the other seven P. aeruginosa genomes were collated for input into the CombiMatrix® array design software (CombiMatrix Corp. WA. USA). The resultant non-redundant array contained 12,543 gene probe spots; comprising 12,147 P. aeruginosa gene probes (including 1,996 gene probes spotted twice), 326 quality-control probes and 70 probes for non-P. aeruginosa genes, including phage and plant genes. Lists of genes unique to each PanArray genome (Table 1) were annotated based on BLAST analysis results, with genes showing an E-value of less than 10⁻⁵ designated as unique.
Media preparation and growth conditions
ASMDM was prepared as described previously [24] and triphenyl tetrazolium chloride to a final concentration of 0.025% w/v was added to distinguish respiring cells. Ten milliliters of ASMDM in 20 ml glass screw-cap bottles was inoculated with 50 μl of the diluted culture just under the surface of the media. The cultures were incubated with a loose lid to permit gas exchange at 37°C, and checked for growth every 24 h. ASMDM-grown cells were harvested at 72 h. An unoinoculated control was included to detect media contamination.

Cell preparation and RNA extraction from ASMDM-grown P. aeruginosa
Bacteria and associated biofilm material were removed from the thick surface growth and the deep anaerobic projections at 72h (Fig. 3) for RNA extraction. Cells were washed to increase cell yield and remove debris. Briefly, the bacterial culture (ca. 10 ml) was transferred to a 50 ml Falcon tube and washed 5 times in an equal volume of ice-cold 1×PBS or until the pellet was cleared of non-cellular debris; by pelleting (5 min/5000 g, 4°C), and re-suspension in fresh ice-cold 1×PBS. All steps were carried out on ice or at 4°C to avoid cell and RNA degradation. Cells were extracted for RNA as previously described [33].

cDNA synthesis and hybridization
cDNA was synthesized as previously described [59]. cDNA was fragmented using DNaseI and quality-checked using a Bioanalyser (Agilent, Germany). Chosen samples were KreaTech labelled (Agilent) and then hybridized to the Combimatrix® PANarray at the Australian Genome Research Facility (AGRF Ltd, Melbourne) using Cy5 dye. Hybridization conditions were as previously described [33,59].

Replicates for microarray analysis
AES-1R, AES-1M were arrayed as biological duplicates (same isolate, different culture, different RNA extraction, and different microarray) giving a total of four data sets, to assess biological variability at the level of culture. Substitution of different biological (culture) replicates was conducted for AES-1R to test for variability, and these had little or no effect on the prediction of differentially expressed genes.

Data analysis
The raw data were re-scaled to account for differences in individual hybridization intensities. Background corrected data were imported into Partek® software, version 6.4 2010 (Partek Inc.), and quantile normalisation (Bolstad et al, 2003) was carried out on the four AES-1 data sets. Data for each pair of biological duplicates were averaged and the average of the acute and chronic isolates was used to determine differential-expression. Genes that were differentially expressed were determined by an ANOVA model with a cut-off value of p = 0.05. All microarray data is MIAME compliant and both the raw and normalized data have been deposited in the MIAME compliant database Gene Expression Omnibus (GEO) http://www.ncbi.nlm.nih.gov/projects/geo under platform accession number GPL13924/Series GSE28152.

Gene expression by quantitative PCR
Thirteen genes mainly virulence-related genes that showed significant differential expression by microarray (adkA, algC, AES_6147, AES_2005, pyrC, pelB, cls, algD, ppiA, pscD, hmeE, pocG and hmgF) were also checked for expression by quantitative SYBR-green-PCR (qPCR) using a Rotor-Gene6000 Real-Time amplification system (Qiagen), and performed on cDNA synthesised from the microarray RNA or synthesised from RNA extracted from later equivalent experiments. Genes were chosen based on high differential expression and/or association with virulence (Tables 2 and 3). Oligonucleotide primers were designed using Primer Express (Applied Biosystems), cDNA was synthesised as described (Invitrogen), by reverse transcription (RT) using 50U SuperScriptII RT (Invitrogen) and 1 μg total RNA.

Supporting Information
Table S1 Genes unique* to P. aeruginosa AES-1R based on BLAST analysis of the AES-1R genome. (DOC)
Table S2 Genes differentially expressed between P. aeruginosa AES-1R and AES-1M grown in ASMDM (p<0.05). (DOC)
Table S3 Genes without homologues in PAO1* that were differentially expressed between P. aeruginosa AES-1R and AES-1M grown in ASMDM (p<0.05). (DOC)

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Author Contributions
Conceived and designed the experiments: JM SN BR TS TT CW LT PB DP. Performed the experiments: JM SN TS TT DP SC. Analyzed the data: JM SN TS TT BR. Contributed reagents/materials/analysis tools: JM TS TT DP BR. Wrote the paper: JM SN BR. Editing of manuscript: JM SN BR TT TS CW PB LT PB DP.

References
1. Koch C, Hoaby N (1993) Pathogenesis of cystic fibrosis. Lancet 341: 1065-1069.
2. Kosorok MB, Zeng L, West SE, Rock MJ, Spijelaard ME, et al. (2001) Acceleration of lung disease in children with cystic fibrosis after Pseudomonas aeruginosa acquisition. Pediatr Pulmonol 32: 277–287.
3. Al-Aloul M, Crawley J, Winstanley C, Hart CA, Ledson MJ, et al. (2004) Increased morbidity associated with chronic infection by an epidemic Pseudomonas aeruginosa strain in CF patients. Thorax 59: 334–336.
4. Griffiths AL, Jamsen K, Carlin JB, Grimwood K, Carzino R, et al. (2005) Effects of segregation on an epidemic Pseudomonas aeruginosa strain in a cystic fibrosis clinic. Am J Respir Crit Care Med 171: 1020–1025.
5. Gibson RL, Burns JL, Ramsey BW (2003) Pathophysiology and management of pulmonary infections in cystic fibrosis. Am J Respir Crit Care Med 168: 910–951.
6. Sadikot RT, Blackwell TS, Christman JW, Prince AS (2005) Pathogen-host interactions in Pseudomonas aeruginosa pneumonia. Am J Respir Crit Care Med 171: 1209–1223.
7. Mayer JM, Neely A, Stinzzi A, George C, Holder IA (1996) Pyoverdin is essential for virulence of Pseudomonas aeruginosa. Infect Immun 64: 518–523.
8. Ostroff RM, Wretlid B, Vasil ML (1989) Mutations in the hemolytic-phospholipase C operon result in decreased virulence of Pseudomonas aeruginosa.
PAO1 grown under phosphate-limiting conditions. Infect Immun 57: 1369–1373.
11. Wilderman PJ, Vasil AI, Johnson Z, Vasil MI (2001) Genetic and biochemical analyses of a eukaryote-like phosphodiesterase D of Pseudomonas aeruginosa suggest horizontal acquisition and a role for persistence in a chronic pulmonary infection model. Mol Microbiol 39: 291–303.
12. Lindsey TL, Hagedun JS, Sokol PA, Silo-Suh LA (2008) Virulence determinants from a cystic fibrosis isolate of Pseudomonas aeruginosa include inositol lyase. J Med Microbiol 57: 1381–1387.
13. Dacheux D, Attre J, Toussaint B (2001) Expression of ExsA in trans confers type III secretion system-dependent cytotoxicity on nontypeable Pseudomonas aeruginosa cystic fibrosis isolates. Infect Immun 69: 530–542.
14. Smith EE, Buckley DG, Wu Z, Naughton S, Turnbull L, Tingpej P, et al. (2010) Gene dynamics of Adaptive Microevolution of Hypermutable Pseudomonas aeruginosa during Chronic Pulmonary Infection in Patients with Cystic Fibrosis. Journal of Infectious Diseases 200: 118–130.
15. Fung C, Naughton S, Burns JL, Speert DP, Turnbull L, Tingpej P, et al. (2010) Gene expression of Pseudomonas aeruginosa in a mucin-containing synthetic growth medium mimicking cystic fibrosis lung sputum. J Med Microbiol 59: 1089–1100.
16. Ciuffo O, Lee B, Johannesson M, Hermansen NO, Meyer P, et al. (2008) Investigation of the algT operon sequence in mucoid and non-mucoid Pseudomonas aeruginosa isolates from patients with cystic fibrosis. Microbiology 154: 3264–3274.
17. Jain M, Bar-Meir M, McColley S, Cullina J, Potter E, et al. (2008) Evolution of Pseudomonas aeruginosa type III secretion in cystic fibrosis: a paradigm of chronic infection. Transl Resp Med 125: 257–264.
18. Mena A, Smith EE, Burns JL, Speert DP, Moskowitz SM, et al. (2008) Genetic adaptation of Pseudomonas aeruginosa to the airways of cystic fibrosis patients is catalyzed by hypermutation. J Bacteriol 190: 7910–7917.
19. Oliver A, Canton R, Campo P, Baquerizo F, Blazquez J (2008) High frequency of hypermutable Pseudomonas aeruginosa in cystic fibrosis lung infection. Science 320: 1251–1254.
20. Govan JR, Deretic V (1996) Microbial pathogenesis in cystic fibrosis mucoid Pseudomonas aeruginosa and Burkholderia cepacia. Microbiol Rev 60: 539–574.
21. Hoberoth C, Hoffmann R, Echterm A, Henke C, Schmidott S, et al. (2009) Dynamics of Adaptive Microevolution of Hypermutable Pseudomonas aeruginosa during Chronic Pulmonary Infection in Patients with Cystic Fibrosis. Journal of Infectious Diseases 200: 118–130.
22. Fung C, Naughton S, Turnbull L, Tingpej P, Rose B, et al. (2010) Gene expression of Pseudomonas aeruginosa in a mucin-containing synthetic growth medium mimicking cystic fibrosis lung sputum. J Med Microbiol 59: 1089–1100.
23. Ciuffo O, Lee B, Johannesson M, Hermansen NO, Meyer P, et al. (2008) Investigation of the algT operon sequence in mucoid and non-mucoid Pseudomonas aeruginosa isolates from patients with cystic fibrosis. Microbiology 154: 3264–3274.
24. Jain M, Bar-Meir M, McColley S, Cullina J, Potter E, et al. (2008) Evolution of Pseudomonas aeruginosa type III secretion in cystic fibrosis: a paradigm of chronic infection. Transl Resp Med 125: 257–264.
25. Mena A, Smith EE, Burns JL, Speert DP, Moskowitz SM, et al. (2008) Genetic adaptation of Pseudomonas aeruginosa to the airways of cystic fibrosis patients is catalyzed by hypermutation. J Bacteriol 190: 7910–7917.
26. Oliver A, Canton R, Campo P, Baquerizo F, Blazquez J (2008) High frequency of hypermutable Pseudomonas aeruginosa in cystic fibrosis lung infection. Science 320: 1251–1254.
27. Govan JR, Deretic V (1996) Microbial pathogenesis in cystic fibrosis mucoid Pseudomonas aeruginosa and Burkholderia cepacia. Microbiol Rev 60: 539–574.
28. Hoberoth C, Hoffmann R, Echterm A, Henke C, Schmidott S, et al. (2009) Dynamics of Adaptive Microevolution of Hypermutable Pseudomonas aeruginosa during Chronic Pulmonary Infection in Patients with Cystic Fibrosis. Journal of Infectious Diseases 200: 118–130.
29. Fung C, Naughton S, Turnbull L, Tingpej P, Rose B, et al. (2010) Gene expression of Pseudomonas aeruginosa in a mucin-containing synthetic growth medium mimicking cystic fibrosis lung sputum. J Med Microbiol 59: 1089–1100.
30. Ciuffo O, Lee B, Johannesson M, Hermansen NO, Meyer P, et al. (2008) Investigation of the algT operon sequence in mucoid and non-mucoid Pseudomonas aeruginosa isolates from patients with cystic fibrosis. Microbiology 154: 3264–3274.
31. Jain M, Bar-Meir M, McColley S, Cullina J, Potter E, et al. (2008) Evolution of Pseudomonas aeruginosa type III secretion in cystic fibrosis: a paradigm of chronic infection. Transl Resp Med 125: 257–264.
32. Mena A, Smith EE, Burns JL, Speert DP, Moskowitz SM, et al. (2008) Genetic adaptation of Pseudomonas aeruginosa to the airways of cystic fibrosis patients is catalyzed by hypermutation. J Bacteriol 190: 7910–7917.
33. Oliver A, Canton R, Campo P, Baquerizo F, Blazquez J (2008) High frequency of hypermutable Pseudomonas aeruginosa in cystic fibrosis lung infection. Science 320: 1251–1254.
34. Govan JR, Deretic V (1996) Microbial pathogenesis in cystic fibrosis mucoid Pseudomonas aeruginosa and Burkholderia cepacia. Microbiol Rev 60: 539–574.
35. Hoberoth C, Hoffmann R, Echterm A, Henke C, Schmidott S, et al. (2009) Dynamics of Adaptive Microevolution of Hypermutable Pseudomonas aeruginosa during Chronic Pulmonary Infection in Patients with Cystic Fibrosis. Journal of Infectious Diseases 200: 118–130.
36. Fung C, Naughton S, Turnbull L, Tingpej P, Rose B, et al. (2010) Gene expression of Pseudomonas aeruginosa in a mucin-containing synthetic growth medium mimicking cystic fibrosis lung sputum. J Med Microbiol 59: 1089–1100.
37. Ciuffo O, Lee B, Johannesson M, Hermansen NO, Meyer P, et al. (2008) Investigation of the algT operon sequence in mucoid and non-mucoid Pseudomonas aeruginosa isolates from patients with cystic fibrosis. Microbiology 154: 3264–3274.
38. Jain M, Bar-Meir M, McColley S, Cullina J, Potter E, et al. (2008) Evolution of Pseudomonas aeruginosa type III secretion in cystic fibrosis: a paradigm of chronic infection. Transl Resp Med 125: 257–264.
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