Interference of \textit{Mycoplasma} spp. or \textit{Ureaplasma} spp. in Ovine Semen Quality

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Abstract The presence of mycoplasma in ovine semen was associated in 9.09\% (3/33) to the microscopic and macroscopic alterations of this fluid. \textit{Mycoplasma} spp. was isolated in 36.36\% (12/33) from semen samples while \textit{Ureaplasma} spp. was isolated in 12.12\% (4/33). The mollicute infection rates in studied semen samples indicated that the diagnosis of these bacteria in ovine must be a routine procedure for the quality of this biological product.

Keywords Mycoplasma, Ureaplasma, Ovine Semen

1. Introduction

The reproductive disorders in animals interfere the economic investments in cattle breeding. The related diseases may be caused by handling deficiencies, environmental and genetic influences and infectious agents. The urogenital infections may also be caused by some Mollicutes and \textit{Mycoplasma} spp. and \textit{Ureaplasma diversum} are mostly detected or isolated and associated with the reproductive disorders in bovines\cite{1,2}.

In 1955 Albertsen, in Denmark raised the possibility that the antibiotic-resistant microorganisms were responsible for the persistent low fertility bulls\cite{25}. The first mycoplasma was isolated in 1898 by Nocard and Roux in France in a clinical material obtained from a bovine during an outbreak of contagious bovine pleuropneumonia.

A bovine \textit{Ureaplasma} was isolated firstly in 1978 and by the time many bovine reproductive disorders were controversially associated because this mollicute was also isolated from healthy animals. This bacteria was studied through experimental and natural infections in bovines causing abortion, infertility and vulvovaginitis syndrome\cite{3,4,5,6}. Ball and Mc Caughey\cite{7} mentioned that the presence of mycoplasmas in vulvar or cervix side may be normal but their isolation from cervix or uterus was considered a pathological condition.

The association of Mollicutes with reproductive disorders in caprines and ovines was mentioned less frequently \cite{8-10}. \textit{Mycoplasma} serotype 11-strain 2D, is a not yet classified strain and was related with cases of vulvovaginitis and reproductive disorders in Australia, USA, India, England, France and Nigeria\cite{10}. Other species were isolated from reproductive tract of ovine and caprine with circumstantial evidences of pathogenesis. In this context are included \textit{M. mycoides} subsp. \textit{mycoides}, \textit{M. bovigenitalium}, \textit{M. mycoides} subps. \textit{capri}, \textit{M. arginini}, \textit{M. alkalescens} e \textit{Acholeplasma} spp.\cite{11,12}. Fe et al.,\cite{13} obtained three positive PCRs and cultures for \textit{Mycoplasma} spp. in 146 samples of caprine semen. Using serological and PCR methodology the isolates were identified \textit{M. alkalescens}.

Livingston et al.,\cite{14} and Ball et al.,\cite{7} inoculated in caprines and ovines field isolates of \textit{ureaplasma} recovered from other diseased animals and related them to the serotype IX as causative of infertility and abortion in sheep.

Actually seven species of \textit{ureaplasma} are recognized. They are human, bovine, canine, feline and avian origin. It was described other five species, but they also did not received yet a nomenclature\cite{15}. These isolates were included in those recovered from ovine or caprine and nine described serotypes. Their DNA and some polypeptides pointed a similarity with \textit{U. diversum}\cite{8,16,17}. The pathogenicity of these microorganisms in reproductive disorders in caprines and ovine are unknown.

The adherence of mentioned Mollicutes to the bovine spermatozoa is the initial step in this host-parasite relationship. Nicholas et al. (1999) described decreased motility of spermatozoa in semen of bulls belonging to herds infected with \textit{Mycoplasma ovine/caprine} serogroup 1. These animals presented low fertility rates, and degeneration of the tail in cases of orchitis. AK et al (1995) inoculated experimentally \textit{M. agalactiae} and observed a decrease of volume, activity, motility and semen.
Some species of *Mycoplasma* and *Ureaplasma diversum* were associated with seminal vasculitis, balanoposthitis, epididymitis,
[27] other functional disorders and morphology of spermatozoa
[28,29] including a reduced motility.
[30,31,32,33,34,35,36]

The virulence mechanisms by which organisms Family
Mycoplasmataceae cause problems for cells are related to
interaction with the immunological system, changes in
humoral and cellular immune response and the production
and induction of cytotoxic components such as ammonia
and peroxidase.

Mycoplasma spp and *Ureaplasma diversum* has a
worldwide distribution and can be spread through
international trade of animals, semen and industrial
products for transferring embryos.
[30,37,38]

Table 1. Estimated risk regarding to association between *Mycoplasma*
spp isolation and poor semen quality of ovine

| Seminal alterations | Positive (%) | ODa | CI95% |
|---------------------|--------------|-----|-------|
| ↓Vigor              | 3 (0.09)     | 2.00| 0.33 – 11.97 |
| ↓Motility           | 7 (21.21)    | 0.86| 0.20 – 3.66 |
| ↓Sperm concentration| 6 (18.18)    | 2.50| 0.57 – 10.94 |
| Presence of clumps  | 3 (0.09)     | 2.00| 0.33 – 11.97 |
| Polymorphonuclear   | 4 (12.12)    | 4.75| 0.72 – 31.38 |

*aOD=Odds Ratio, CI=Confidence Interval and ↓=Low*

Table 2. Estimated risk regarding to association between *Ureaplasma* 
spp isolation and poor semen quality of ovine

| Seminal alterations | Positive (%) | ODa | CI95% |
|---------------------|--------------|-----|-------|
| ↓Vigor              | 0            | 0.50| 0.02 – 10.58 |
| ↓Motility           | 2 (6.06)     | 0.61| 0.07 – 4.99 |
| ↓Sperm concentration| 2 (6.06)     | 1.90| 0.23 – 15.59 |
| Presence of clumps  | 2 (6.06)     | 6.25| 0.67 – 57.93 |
| Polymorphonuclear   | 1 (3.03)     | 1.60| 0.14 – 18.73 |

*aOD=Odds Ratio, CI=Confidence Interval and ↓=Low*

Table 3. Estimated risk regarding to association between *Mycoplasma* 
spp and *Ureaplasma diversum* spp isolation and poor semen quality of ovine

| Seminal alterations | Positive (%) | ODa | CI95% |
|---------------------|--------------|-----|-------|
| ↓Vigor              | 0            | 0.54| 0.02 – 11.80 |
| ↓Motility           | 2 (6.06)     | 1.33| 0.11 – 16.40 |
| ↓Sperm concentration| 1 (3.03)     | 0.86| 0.07 – 10.66 |
| Presence of clumps  | 2 (6.06)     | 13.0| 0.94 – 178.88 |
| Polymorphonuclear   | 1 (3.03)     | 2.50| 0.19 – 33.19 |

*OD=Odds Ratio, CI=Confidence Interval and ↓=Low*

4. Discussion

Mycoplasma isolation from ovine semen was not
reported yet in Brazil. Although it was studied 33 samples the obtained frequencies of 36.36% for Mycoplasma spp. and 12.12% for Ureaplasma spp. are the first national reference and points that the percentage of this infection is probably higher.

Isolation of mycoplasmas in ovine is not usually performed in other countries. Kappor et al.,[11], in India, Mycoplasma spp. and Acholeplasma spp. were isolated in 17.6% from 68 semen samples. Trichard et al.,[20], in South Africa found mycoplasmas in 83% of samples from ovine with ulcerative balanoposthitis and vulvovaginitis and 36% in healthy animals. Nicholas et al.,[10], in Great Britain reported the isolation of Mycoplasma serogroup 11 in vaginal mucus from 23 female infertile sheep. It was showed the interference of these microorganisms in ovine breeding.

Contamination of semen and preputial mucus with U. diversum was firstly mentioned in 1969 in England, when ten preputial cultures and 84% in 32 samples of fresh semen were positive for the agent.[40] ONOVIRAN et al., (1975), in Canada observed 132(35%) positive cultures for U. diversum in preputial mucus, 140(24%) in fresh semen samples and 42(14%) in processed semen. In 1978, Czechoslovakia, this ureaplasma was recovered from 202 samples of bull semen in insemination centers.[41]

The highest rate of positive cultures for Mycoplasma (71%) was obtained in cows with low fertility compared with 24% of cows culled for other reasons.[42]

FISH et al. (1985), Canada, reported that 28% of bulls used for artificial insemination had semen contaminated with species of Mycoplasma and M. bovis was not recovered. GARCIA et al. (1986), in the same country, studied 2950 samples of semen and M. bovis was. Ball et al. (1987)[43] isolated Mycoplasma spp in 46% of 332 samples of fresh semen of bulls In 32% of these positive samples, were also infected with U.diversum.

LE GRAND et al. (1995)[44], in France, isolated U. diversum in 74% (37/50) of semen samples. Serogroups B and C were predominant in males.

M. bovis was the first bovine mycoplasma identified in Brazil by ROSSINI, (1978)[45] in calves with pneumonia in a property located in State of São Paulo. Subsequently, LIBERAL et al. (1982)[46] reported the isolation of Mycoplasma spp. in cases of bovine pneumonia in the State of Rio de Janeiro.

The mycoplasma transmission through the infected semen justify in part the diversity of findings. Lingwood et al.,[21] and Quinn et al.,[22] showed the adherence of U. diversum to bovine spermatozoa trough sulfoglicolipids. Eaglesomme and Garcia[22] described the interference of M. bovis in the fertilization processes. The presented data and the diversity of the host-parasite relationship in mycoplasmolgy strongly suggest a better control of these bacteria in ovine breeding or rising.

The motility and other parameters of ovine spermatozoa analysed in present study were influenced by mycoplasma infection. It was possible to estimate a risk of 4.75 higher to found these bacteria in polymorphonuclear-PMN cells in infected ovine semen (Table. 1). When the semen is infected with Ureaplasma spp the estimative risk to produce semen clots was 6.25 higher if compared with mycoplasma free semen. It was detected an OR>1 for a lactescent appearance, spermatozoa density and PMN (Table 2). The same OR and semen parameters was obtained for the co-infection of Mycoplasma spp and Ureaplasma spp. In this study the estimated risk was 1.33, 13, 2.5 times higher for alterations of spermatozoa motility, clots and PMN in semen respectively(Table. 3).

Cardoso et al.,[24], studied bovine semen infected with Mycoplasma spp and obtained a risk of 1.02 higher for the lactescent semen and 2.7 higher for spermatozoa density. In presence of U. diversum the risk was 1.32 higher for clumping. In the present study it was detected a higher risk in ovine semen quality when infected with mycoplasma than described for bovines. Nicholas et al.,[10] mentioned a decreasing motility of bovine spermatozoa and low fertility in reproductive cattle infected with Mycoplasma serogroup 11. Animals with orchitis presented a degeneration of spermatoza tails. Ak et al.,[12], experimentally inoculated M. agalactiae in bovine semen and mentioned a decreasing volume, motility and density of spermatoza and an increase of abnormal spermatozoa.

In 1977 Jurmanova and Sterbova associated a low sperm motility(60%) and the semen contamination by mycoplasma and ureaplasma adhered on these cells . It was observed by a reduced motility (60%) and[26]. There are infectious agents more commonly associated with animal reproductive disorders : Brucella spp, Campylobacter spp, Letospira spp.[28][35] However, many other are less mentioned, as Mollicutes , listed in schedule B of the OIE, and therefore are extremely important and should be studied.[39]

Eaglesomme (1980) studies found that M. bovis interfered extensively in various stages of fertilization in vitro, first causing the decrease in the percentage of sperm capacitation, fusion with the oocyte and interfering with the organization of the pronucleus after chromatin decondensation, without, however, cause changes in their viability and motility.

5. Conclusions

It was possible to found estimate risk of 4.75 higher in polymorphonuclear-PMN cells in infected mycoplasma ovine semen and 6.25 higher estimate risk semen clots in semen infected with Ureaplasma spp. In this study the estimated risk was 1.33, 13, 2.5 times higher for alterations of spermatozoa motility, clots and PMN in infected semen respectively. The monitoring of mycoplasma infection in ovine production must be included in routine analysis.

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