Seropositivity and Risk Factors Associated with *Toxoplasma gondii* Infection in Wild Birds from Spain

Oscar Cabezon¹, Ignacio Garcia-Bocanegra², Rafael Molina-Lopez³, Ignasi Marco¹, Juan M. Blanco⁴, Ursula Hofle⁵, Antoni Margalida⁶,⁷, Esther Bach-Raich¹, Laila Darwin⁸,⁹, Israel Echeverria⁹, Elena Obon³, Mauro Hernandez¹⁰, Santiago Lavin¹¹, Jitender P. Dubey¹¹, Sonia Almeria¹²,¹³

¹ Servei d’Ecopatologia de Fauna Salvatge, Departament de Medicina i Cirugia Animals, Facultat de Veterinaria, Universitat Autònoma de Barcelona, Bellaterra, Spain, ² Departamento de Sanidad Animal, Facultad de Veterinaria, Universidad de Córdoba, Córdoba, Spain, ³ Centre de Fauna Salvatge de Torreferrussa, Direcció General del Medi Natural i de la Biodiversitat-Forestal Catalana, Generalitat de Catalunya, Santa Perpètua de la Mogoda, Barcelona, Spain, ⁴ Centro de Estudios de Rápaces Ibéricas, Sevilla de la Jara, Castilla-la-Mancha, Spain, ⁵ Instituto de Investigación en Recursos Cinegéticos (IREC), Centro de Recerca en Sanitat Animal (CRiSA), Universitat Autònoma de Barcelona, Bellaterra, Barcelona, Spain, ⁶ Department de Sanitat i d’Anatomia Animals, Facultat de Veterinària, Universitat Autònoma de Barcelona, Bellaterra, Barcelona, Spain, ⁷ Direcció General del Medio Natural (CRAS, Junta de Castilla y Leon), Departament d’Agricultura, Ramaderia, Pesca, Alimentación i Medi Natural (Torreferrussa, Generalitat de Catalunya), Consejería de Medio Ambiente (Junta de Andalucía), ⁸ SEFAS (Universitat Autònoma de Barcelona) y IREC (Ciudad Real) for donation of wild bird samples. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

*Toxoplasma gondii* is a zoonotic intracellular protozoan parasite of worldwide distribution that infects many species of warm-blooded animals, including birds. To date, there is scant information about the seropositivity of *T. gondii* and the risk factors associated with *T. gondii* infection in wild bird populations. In the present study, *T. gondii* infection was evaluated on sera obtained from 1079 wild birds belonging to 56 species (including *Falconiformes* (n = 610), *Strigiformes* (n = 260), *Ciconiformes* (n = 156), *Gruiformes* (n = 21), and other orders (n = 32), from different areas of Spain. Antibodies to *T. gondii* (modified agglutination test, MAT titre 1:25) were found in 282 (26.1%, IC95%: 23.5–28.7) of the 1079 birds. This study constitute the first extensive survey in wild birds species in Spain and reports for the first time *T. gondii* antibodies in the griffon vulture (*Gyps fulvus*), short-toed snake-eagle (*Circaetus gallicus*), Bonelli’s eagle (*Aquila fasciata*), golden eagle (*Aquila chrysaetos*), bearded vulture (*Gypaetus barbatus*), osprey (*Pandion haliaetus*), Montagu’s harrier (* Circus pygargus*), Western marsh-harrier (*Circus aeruginosus*), peregrine falcon (*Falco peregrinus*), long-eared owl (*Asio otus*), common scops owl (*Otus scops*), Eurasian spoonbill (*Platalea leucorodia*), white stork (*Ciconia ciconia*), grey heron (*Ardea cinerea*), common moorhen (*Gallinula chloropus*); in the International Union for Conservation of Nature (IUCN) “vulnerable” Spanish imperial eagle (*Aquila adalberti*), lesser kestrel (*Falco naumanni*) and great bustard (*Otis tarda*); and in the IUCN “near threatened” red kite (*Milvus milvus*). The highest seropositivity by species was observed in the Eurasian eagle owl (*Bubo bubo*) (68.1%, 98 of 144). The main risk factors associated with *T. gondii* seropositivity in wild birds were age and diet, with the highest exposure in older animals and in carnivorous wild birds. The results showed that *T. gondii* infection is widespread and can be at a high level in many wild birds in Spain, most likely related to their feeding behaviour.

Introduction

*Toxoplasma gondii* is a zoonotic intracellular protozoan parasite of worldwide distribution [1,2]. Wild and domestic felines are the definitive hosts, excreting oocysts in faeces. Humans and virtually all warm-blooded species, including birds, can be intermediate hosts and can become infected by ingestion of food and water contaminated with sporulated *T. gondii* oocysts, by consumption of tissue cysts in infected animal tissues, or congenitally [1,2].

*Toxoplasma gondii* infection is prevalent in many avian species including poultry, game and other species in the wild [3,4,5,6,7,8,9]. In addition, clinical cases have been reported and *T. gondii* considered to be the cause of mortality in birds of different species [3,4,5,10,11]. However, there is still only scant information about seroprevalence of *T. gondii* and the risk factors associated to *T. gondii* infection in wild bird populations. Investigation of *T. gondii* seropositivity in birds could be a useful way to assess environmental contamination with oocysts since some avian populations feed directly on the ground and are continuously exposed to oocyst ingestion [11]. Furthermore, the investigation of *T. gondii* infection in avian scavengers might allow the assessment of the possibility of contact with intermediate hosts and the associated risk for public health [12].
In Europe, recent studies of T. gondii seropositivity have shown a high prevalence of T. gondii antibodies in wild birds. Aubert et al [4] reported prevalence levels of 79% of 14 common buzzards (Buteo buteo), 50% of 12 tawny owls (Strix aluco) and 11% of 16 barn owls (Tyto alba) in raptors from France. In Portugal, Lopes et al [6] reported a prevalence of antibodies in 50% of 52 wild birds that included multiple species such as black kite (Milvus migrans), common buzzard, Eurasian sparrowhawk (Accipiter nisus), Northern goshawk (Accipiter gentilis), Eurasian eagle owl (Bubo bubo) and tawny owl. Recently, in Spain, high seroprevalence of T. gondii in common ravens (Corvus corax) (80.5% of 113 sera samples) has been reported [7] and the presence of T. gondii DNA in brain tissues detected in magpies (Pica pica) (http://www.umh.es/_web_rw/ceie/doc/animales/1201_05%20proteccion%20animales%20experimentacion.pdf) of the Ministry of Presidency of Spain.

**Materials and Methods**

**Ethics statement**

Samples were collected in compliance with the Ethical Principles in Animal Research in the wildlife rehabilitation centres. The study of this material (blood serum) did not require the approval of an ethics committee because the collection of sera is considered among routine procedures and the sera was collected and stored before the design of this study. The rehabilitation centres directly depend on the governmental Autonomous Wildlife Services. Thus, protocols, amendments and other resources were done according to the guidelines approved by each Autonomous government following the R.D. 1201/2005 (10th October 2005, BOE 21st October 2005) [13]. The present study analyzed T. gondii seropositivity and provides information on possible associated risk factors, i.e. species, order, dietary habits, migration, sex, age and geographic origin in numerous species of wild birds from Spain, including some endangered species listed in the International Union for Conservation of Nature (IUCN) Red List of Threatened Species [13].

**Samples**

Serum samples from 1079 wild birds of 56 species (Table 1), belonging to the Orders Falconiformes (n = 610), Strigiformes (n = 260), Ciconiiformes (n = 156), Gruiformes (n = 21), and other orders (n = 32), were analyzed. The analyzed species included some listed on the IUCN Red List of Threatened Species [13] as near threatened ([Aegypius monachus, (n = 23), Milvus milvus, (n = 3), Crex crex, (n = 1)], as vulnerable [Aquila adalberti (n = 146); Falco naumanni, (n = 5), Otis tarda, (n = 7)] and; as endangered [Neophron percnopterus, (n = 49)].

Samples were collected from wild birds captured or brought to wildlife rehabilitation centres and other breeding centres between 1996 and 2010. Blood collection was from the jugular vein, the heart, or the brachial vein. Blood was placed in a serum collection tube until clotted and then centrifuged. Sera were stored at −20°C until analysis. The samples were obtained from the main geographic areas from Spain including Northern Spain (n = 332) (Catalonia (228 samples) and Castile and León (4 samples from Valladolid); Central Spain (Castile La Mancha) (n = 563); and Southern Spain (Andalusia) (n = 282). The provinces sampled within Catalonia were Lleida and Barcelona; within Castile-La Mancha the provinces of Toledo and Ciudad Real; and within Andalusia the provinces of Granada, Málaga, Cádiz, Huelva, Seville, Córdoba and Jaén.

Serological test

Sera were examined by the modified agglutination test (MAT) to detect antibodies against T. gondii as described previously [14]. Sera were tested at 1:25, 1:50, 1:100 and 1:500 dilutions. A commercial positive control (Toxotrol-A, Biorneux, France) diluted from 1:25 to 1:3200 (with a minimum titre of 1:200) was included in each test. Negative controls were also included in all tests. Titres of 1:25 or higher were considered positive and those with doubtful results were re-examined. This technique has been previously evaluated in several bird species [3,15].

**Definition of variables and Statistical analysis**

Bird species were classified in regard to feeding behaviour as carnivorous (birds of prey), scavenger (when they feed on dead or decaying matter), granivorous-herbivorous-insectivorous (G/H/I), omnivorous and piscivorous. Whenever possible, sex and age data were collected. Age was classified in two categories (<1 year and ≥1 year). In addition, wild birds were classified as migratory or sedentary (non-migratory), according to their movements [16].

The variables analyzed included wild bird species, order, feeding behavior, geographic area of sample collection, migratory behavior, sex and age. For all statistical analysis only those species with more than 3 analyzed samples (birds) were included.

Association between explanatory variables and T. gondii seropositivity was tested in three steps. Firstly, a bivariate chi-square test was performed to obtain an indication of the relevance of the explanatory variables to the risk of an animal being seropositive. When observations per category were less than six, Fisher’s exact test was used. Secondly, factors showing a p-value <0.25 were further scrutinized for associations using Spearman’s rank correlation coefficient (r) to avoid collinearity problems. If r was larger than 0.4 the variable more clearly linked to T. gondii infection, based on epidemiological data and previous studies in the literature, was retained. The third step involved a multiple logistic-regression model using a non-automatic backward selection of variables. Biologically plausible confounding factors were tested using Mantel-Haenszel analysis and confounding was considered to be potentially significant if ORs shifted appreciably. Changes in the OR greater than 30% were considered indicative of confounding. The fit of the models was assessed using the Hosmer and Lemeshow goodness-of-fit test [17]. Potential two-way interactions between the variables were tested for significance in the model. The model was re-run until all the remaining variables presented statistically significant values (likelihood-ratio Wald’s test, p < 0.05), and a potential causal relationship with the response variable existed. Statistical analyses were performed using SPSS 15.0 (Statistical Package for Social Sciences (SPSS) Inc., Chicago, IL, USA).

**Results**

The overall seropositivity (MAT≥1:25) against T. gondii was 26.1% (CI95%: 23.5–28.7) (282 positive of 1079 wild birds tested), with titres of 1:25 in 145 samples (51.4%), 1:50 in 112 samples (39.7%), 1:100 in 13 samples (4.6%) and 1:500 in 12 samples (4.3%). The results by analyzed species are shown in Table 1.

In the bivariate analysis, considering those species with more than 3 samples analyzed, statistically significant differences were observed between seropositivity levels and the variables: order, wild bird species, geographic area of sample collection, migratory behaviour, age, year of sampling and feeding behaviour.

Based on their feeding behaviour, carnivorous wild birds showed statistically significant higher seropositivity of T. gondii (36.2% of 588) compared to scavenger (15.6% of 262 samples),
| Species                              | IUCN-Red list   | No. sera | % Positive |
|-------------------------------------|----------------|----------|------------|
| **Falconiformes**                   |                |          |            |
| Griffon Vulture (Gyps fulvus)       | Least Concern  | 175      | 17.71      |
| Spanish Imperial Eagle (Aquila adalberti) | Vulnerable   | 146      | 17.07      |
| Common Buzzard (Buteo buteo)        | Least Concern  | 96       | 51.04      |
| Egyptian Vulture (Neophron percnopterus) | Endangered   | 49       | 0.00       |
| Cinereous Vulture (Aegypius monachus) | Near Threatened | 23      | 26.09      |
| Black Kite (Milvus migrans)         | Least Concern  | 17       | 29.41      |
| Bearded Vulture (Gypaetus barbatus) | Least Concern  | 15       | 42.86      |
| Common Kestrel (Falco tinnunculus)  | Least Concern  | 13       | 30.77      |
| Short-toed Snake-eagle (Circaetus gallicus) | Least Concern | 10     | 50.00      |
| Bonelli’s Eagle (Aquila fasciata)   | Least Concern  | 9        | 11.11      |
| Golden Eagle (Aquila chrysaetos)    | Least Concern  | 8        | 62.50      |
| Osprey (Pandion haliaetus)          | Least Concern  | 7        | 28.57      |
| Montagu’s Harrier ( Circus pygargus) | Least Concern | 7        | 14.29      |
| Eurasian Sparrowhawk (Accipiter nisus) | Least Concern | 7        | 0.00       |
| Western Marsh-harrier ( Circus aeruginosus) | Least Concern | 6       | 50.00      |
| Northern Goshawk (Accipiter gentilis) | Least Concern | 5        | 40.00      |
| Lesser Kestrel ( Falco naumanni)    | Vulnerable     | 5        | 40.00      |
| Peregrine Falcon ( Falco peregrinus) | Least Concern  | 4        | 25.00      |
| European Honey-buzzard (Pernis apivorus) | Least Concern | 3        | 0.00       |
| Red Kite ( Milvus milvus)           | Near Threatened | 3        | 33.33      |
| Northern Harrier (Circus cyaneus)   | Least Concern  | 1        | 0.00       |
| Booted Eagle ( Hieraaetus pennatus) | Least Concern  | 1        | 0.00       |
| **Strigiformes**                    |                |          |            |
| Eurasian Eagle-owl (Bubo bubo)      | Least Concern  | 144      | 68.06      |
| Barn Owl ( Tyto alba)               | Least Concern  | 45       | 13.33      |
| Tawny Owl (Strix aluco)             | Least Concern  | 38       | 13.16      |
| Little Owl (Athene noctua)          | Least Concern  | 19       | 15.79      |
| Long-eared Owl (Asio otus)          | Least Concern  | 9        | 11.11      |
| Common Scops-owl (Otus scops)       | Least Concern  | 4        | 25.00      |
| Short-eared Owl ( Asio flammeus)    | Least Concern  | 1        | 0.00       |
| **Ciconiiformes**                   |                |          |            |
| Eurasian Spoonbill (Platlea leucorodia) | Least Concern | 81      | 6.17       |
| White Stork (Ciconia ciconia)       | Least Concern  | 64       | 14.06      |
| Grey Heron (Ardea cinerea)          | Least Concern  | 5        | 60.00      |
| Purple Heron (Ardea purpurea)       | Least Concern  | 4        | 0.00       |
| Black Stork (Ciconia nigra)         | Least Concern  | 1        | 0.00       |
| Cattle Egret (Bubulcus ibis)        | Least Concern  | 1        | 0.00       |
| **Gruiformes**                      |                |          |            |
| Red-knobbed Coot ( Fulica cristata) | Least Concern  | 11       | 0.00       |
| Great Bustard (Otis tarda)          | Vulnerable     | 7        | 14.29      |
| Corncrake (Crex crex)               | Near Threatened | 1        | 0.00       |
| Common Coot (Fulica atra)           | Least Concern  | 1        | 100.00     |
| Common Moorhen (Gallinula chloropus) | Least Concern | 1       | 100.00     |
| **OTHERS**                          |                |          |            |
| Eurasian Thick-knee (Burhinus oedicnemus) | Least Concern | 6       | 0.00       |
| **Passeriformes**                   |                |          |            |

Table 1. Seroprevalence of antibodies against *Toxoplasma gondii* (*T. gondii*) in wild birds from Spain using the modified agglutination test (MAT $\geq 1:25$).
omnivorous (13.2% of 68), G/H/I (11.8% of 34), and piscivorous (10.3% of 97) (P<0.001) species.

The order Strigiformes showed statistically higher seropositivity (44.0% of 259) compared to all the other orders (P<0.01). In addition, the seropositivity in the order Falconiformes (23.8% of 602) was significantly higher than that in the order Ciconiiformes (11% of 154) (P<0.05). Seropositivity in Gruiformes order was 5.6% (1 of 18), and in other orders was 12.5% (2 of 16).

The highest seropositivity by species was observed in the Eurasian eagle owl 68.1% of 144 samples. A higher than 50% seropositivity was observed in: golden eagle (Aquila chrysaetos) (62.5% of 8), grey heron (Ardea cinerea) (60% of 5) and, in common buzzard (Buteo buteo) (51% of 96). Interestingly, no antibodies were observed in the endangered Egyptian vulture (Neophron percnopterus) (0 of 49 samples). Statistically significant differences were observed between B. bubo and B. buteo compared to A. adalberti, Ciconia ciconia, Fulica cristata, G. fulvus, N. percnopterus, Platalea leucorodia, S. aluco and T. alba (P<0.05).

The positivity of infection was significantly higher (P<0.001) in samples collected from wild birds from Northern (mostly Catalonia) and Central Spain (27.2 and 30.2%, respectively) compared to Southern Spain (18.3%), (P<0.001).

Sedentary wild birds had significantly higher seropositivity (31.1% of 795 samples) compared to migratory birds (12.0% of 249) (P<0.001).

Significant differences were observed between samples from birds older than 1 year showed statistically significant higher seropositivity (38.6% of 158) than birds younger than 1 year (9.5% of 148). Significant differences were also observed also among years of sampling (P<0.001). Samples collected up to the year 2000 showed high seropositivity but the number of analyzed samples was low (mean of the 1996–2000 period: 40.0% of 55 samples), (Figure 1). The period of highest seropositivity was observed between 2001 and 2005 (mean of the 2001–2005 period, 54.6% of 132 samples), then seropositivity decreasde from 2006 onward maintaining a similar level up to 2010 (mean of the period 2006–2010, 21.2% of 641 samples) (Figure 1). Seropositivity in the latter period was significantly lower compared to the other two periods (P<0.01).

The final multivariate model showed that the main risk factors associated with T. gondii seropositivity in wild birds were age and feeding behaviour. Older wild birds had a 6.8 times higher risk of

| Table 1. Cont. |
|----------------|
| Species | IUCN-Red list | No. sera | % Positive |
| Falconiformes* | | | |
| Black-billed Magpie (Pica pica) | Least Concern | 4 | 0.00 |
| Eurasian Linnet (Carduelis cannabina) | Least Concern | 3 | 0.00 |
| European Goldfinch (Carduelis carduelis) | Least Concern | 1 | 100.00 |
| Eurasian Jackdaw (Corvus monedula) | Least Concern | 1 | 0.00 |
| Northern House-martin (Delichon urbicum) | Least Concern | 1 | 0.00 |
| European Serin (Serinus serinus) | Least Concern | 1 | 0.00 |
| Anseriformes | | | |
| Mallard (Anas platyrhynchos) | Least Concern | 6 | 33.33 |
| Northern Pintail (Anas acuta) | Least Concern | 1 | 100.00 |
| Red-crested Pochard (Netta rufina) | Least Concern | 1 | 0.00 |
| Apodiformes, Columbiformes, Galliformes, Podicipediformes and Pelecaniformes | | | |
| Common Pheasant (Phasianus colchicus, Galliformes) | Least Concern | 2 | 0.00 |
| Eurasian Collared-dove (Streptopelia. decaocto, Columbiformes) | Least Concern | 1 | 0.00 |
| Great Cormorant (Phalacrocorax carbo, Pelecaniformes) | Least Concern | 1 | 0.00 |
| Little Grebe (Tachybaptus ruficollis, Podicipediformes) | Least Concern | 1 | 0.00 |
| Common Swift (Apus apus, Apodiformes) | Least Concern | 1 | 0.00 |
| Eurasian Hoopoe (Upupa epops, Apodiformes) | Least Concern | 1 | 0.00 |

*Different letters among orders indicate statistically significant differences.

**Only those species with more than 3 analyzed samples (birds) were included in the statistical analysis.

doi:10.1371/journal.pone.0029549.g001

Figure 1. Seropositivity of Toxoplasma gondii (95% confidence limits) in wild birds from Spain among periods (years) of sampling.

doi:10.1371/journal.pone.0029549.g001
being seropositive compared to younger birds (>1 year compared to <1 year birds (OR = 6.8; 95% CI = 2.71–17.29) and carnivorous wild birds had a 5.4 times higher risk compared to piscivorous wild birds (taken as reference) (OR = 5.36; 95% CI = 2.70–10.6) (Table 2).

Discussion

The present study included a wide range of wild bird species and a high number of analyzed samples supplementing the existing data on seropositivity of T. gondii in wild birds worldwide, and adds numerous new species of wild birds to the possible intermediate host list for T. gondii infection. To our knowledge, this is the first report of T. gondii antibodies in griffon vulture, short-toed snake-eagle, Bonelli’s eagle, golden eagle, bearded vulture, osprey, Montagu’s harrier, Western marsh-harrier, peregrine falcon, long-eared owl, common scops owl, Eurasian spoonbill, white stork, grey heron, common moorhen, the IUCN “vulnerable” Spanish imperial eagle, lesser kestrel and great bustard; and the IUCN “near threatened” red kite and it is the first survey of these wild bird species in Spain. Detection and/or isolation of the parasite from these species would be necessary to corroborate that these species are intermediate hosts of T. gondii.

The results showed that T. gondii infection is widespread in many wild birds in Spain, with high variation among different species, orders, geographical regions and feeding behaviour. The main risk factors associated with T. gondii seropositivity in wild birds were age and feeding behaviour, with higher exposure observed in older animals and in species with a meat-based diet. The observed age effect is in accordance with previous studies of T. gondii seroprevalence in other animal species such as pigs, Iberian lynx (Lynx pardinus), Spanish ibex (Capra pyrenaica) and cats in Spain [18,19,20,21] as well as in numerous studies of other species worldwide [2], and would be an indication of cumulative likelihood for exposure to T. gondii with age [2,22] and to lifelong persistence of antibodies. The higher seropositivity observed in older individuals could also be an indication of horizontal transmission as the main route of transmission of T. gondii in these species.

Carnivorous species showed a significantly higher seropositivity of T. gondii. In this respect, the order Strigiformes (in which virtually all the species are carnivorous) showed the highest seropositivity, followed by the order Falconiformes, in which the majority of species are carnivorous and/or scavenger species. The order Gruiformes, with mostly G/H/I species, showed the lowest seropositivity. This agrees with the results obtained in mammals, in which several studies have observed that carnivorous species have higher seroprevalence of T. gondii antibodies compared to omnivorous, herbivorous or insectivorous species [23,24,25,26]. A possible explanation could be the cumulative ingestion of infected prey, as suggested by similar results from comparison of seroprevalence levels in carnivorous mammals such as cats [21], omnivorous such as wild boars [27] and herbivorous such as red deer and other wild ruminants [28] in Spain. The main mode of infection of wild birds, as of other intermediate hosts, with T. gondii is through ingestion of food and water contaminated with sporulated oocysts (mainly in ground feeding birds) and by consumption of cysts contained in tissues of infected animals (mainly in carnivorous, scavenger birds) [2]. The present results suggest that the main mode of transmission of T. gondii in the analyzed wild birds was consumption of tissue cysts.

In agreement with the above results, the highest seropositivity was observed in the Eurasian eagle owl (68.1% of 144), a carnivorous species recently included in the list of birds with T. gondii antibodies by Lopes et al [6]. High seropositivity was also observed in other carnivorous species such as the common buzzard, which has been previously reported to have high seroprevalence in France (79% of 14 samples) [4], and in the golden eagle. The main diet of these species of carnivorous wild birds includes rodents and other small mammals, which are well known reservoirs of T. gondii infection for many animal species, especially pigs [25,29,30]. On the other hand, high seropositivity was also observed in some omnivorous species, such as the grey heron (60% of 5) and in some species of duck [Anas acuta (100% of 1) and A. platyrhynchos (33.3% of 6)], although the number of analyzed samples was relatively low. The grey heron mainly feeds in shallow water catching fish, frogs, and insects, but will also eat small mammals, reptiles and other birds [31] and, like other Ciconiformes, has been moving into urban environments where they also make use of food discarded by humans. The fact that grey heron samples were mainly from Catalonia (4 of 5 samples, with all the positive samples from this location), which seems to be one of the most prevalent areas of T. gondii infection in Spain, with higher prevalence of T. gondii compared to other locations for species such as wild rabbits [32], roe deer [33] and domestic pigs [18], and where higher detection of the parasite has also been observed in tissues from wild birds [9], could also be an explanation for the increased seropositivity observed in this species. Moderate precipitation and humidity in those areas might favour oocyst survival and sporulation in the environment facilitating T. gondii spread and maintenance [1]. The ducks analyzed in the present study were from different geographical regions, indicating that seroprevalence was not due to location. High seroprevalence in ducks has been recently reported by Alvarado-Esquivel et al [8] (2 positive samples, in 1 A. platyrhynchos and 1 A. durazi, of 4 ducks analyzed).

Table 2. Logistic regression analysis of potential risk factors associated with T. gondii seropositivity in wild birds in Spain.

| Variable | Category | β  | P-value | OR   | 95% CI        |
|----------|----------|----|---------|------|---------------|
| Food     | Carnivorous | 1.68 | <0.001  | 5.36 | (2.69–10.68)  |
|          | Granivorous/Herbivorous/Insectivorous | -0.97 | 0.221  | 0.38 | (0.08–1.78)   |
|          | Omnivorous | -18.86 | 0.999  | 0.00 |                  |
|          | Scavenger  | 0.71  | 0.231  | 2.04 | (0.63–6.53)   |
|          | Piscivorous |      |        |      |               |
| Age      | >1 year old | 1.92 | <0.001  | 6.85 | (2.71–17.29)  |
|          | <1 year old |      |        |      |               |

*Reference category.

doi:10.1371/journal.pone.0029549.t002
Similarly, it was also unexpected to observe high seropositivity of *Toxoplasma gondii* in bearded vultures (*G. barbatus*) (42.9% of 15) because this species feeds primarily upon livestock bones. A possible explanation is that during the chick-rearing period, small and medium-sized mammals and other birds are also included in the chick’s diet, and even in the parents diet [34]. Furthermore, most bearded vultures were also obtained in the northern area. Further studies including more samples of this species will need to be performed to confirm this observation.

To the contrary, the Egyptian vulture, a small vulture inhabiting open landscapes in arid and rugged regions in the Mediterranean, Africa and Asia, did not show antibodies to *T. gondii*. The diet of Egyptian vultures is based on carcasses of small and medium-sized wild and domestic animals and, in a lesser degree, livestock, usually ingesting small pieces of skin or muscle [35] which could be the reason for the observed lack of contact with *T. gondii* in the analyzed samples.

Interestingly, some piscivorous birds showed contact with the parasite, including some species such as the Eurasian spoonbill (*P. leucorodia*) analyzed in high numbers, (6.2% of 81 samples). This result could be an indication of water contamination by oocysts.

Until recently, waterborne transmission of *T. gondii* was considered uncommon. However, a large human outbreak linked to contamination of a municipal water reservoir in Canada by wild felids and the widespread infection of marine mammals in the USA provided reasons to question this view [36]. Antibodies have been observed in Mediterranean wild dolphins [37] and a case of systemic toxoplasmosis in a pregnant dolphin from the Spanish Mediterranean coast has been reported [38].

In summary, the present study extends the range of species of wild birds that have antibodies to *T. gondii* and indicates widespread exposure to *T. gondii* among many wild bird species in Spain, with highly variable positivity levels among species and orders. The main risk factors associated with *T. gondii* seropositivity in wild birds were age and feeding behaviour, with higher exposure in older animals and in carnivorous wild birds. The results indicate that *T. gondii* infection can be high in some species, mainly related to their feeding behaviour, and that these species may have an important role in the epidemiology of *T. gondii* infection. The observed contact with the parasite in some vulnerable or near threatened wild bird species suggests that the analysis of this parasite should be taken into account in relevant wild bird conservation programs.

### Acknowledgments

The authors would like to thank Paul Cairns for assistance with the English translation. The study was part of an agreement with Direcció General del Medi Natural [Junta de Castilla y León], Departament d’Agricultura, Pesca, Alimentació i Medi Natural [Tarragona], Generalitat de Catalunya, Consejeria de Medio Ambiente [Junta de Andalucía], SEFaS (Universitat Autònoma de Barcelona) and IREC (Ciudad Real) for donation of wild bird samples. We thank all the staff that participated in the sample collection and in the care of the animals.

### Author Contributions

Conceived and designed the experiments: OC SA IGB. Performed the experiments: OC LD IE. Analyzed the data: IGB. Contributed reagents/materials/analysis tools: RMI JM UB AM EBR EO MH SL JPD. Wrote the paper: OC SA IGB AM.

### References

1. Dubey JP, Beattie CP (1988) Toxoplasmosis of Animals and Man. In: CRC Press, Boca Raton, FL, pp. 1–220.
2. Dubey JP (2009) Toxoplasmosis in animals and humans. Second Edition. CRC Press, Boca Raton, FL. 313 p.
3. Dubey JP (2002) *A review of Toxoplasmosis in wild birds*. Vet Parasitol 106: 172–173.
4. Aubert D, Terrier ME, Dumetre A, Barrat J, Villena I (2008) *Prevalence of Toxoplasma gondii in raptors from France*. J Wildl Dis 44: 172–173.
5. Dubey JP, Felix TA, Kwok OC (2010) *Serological and parasitological results from Cryptocephalus rufus from Kenya*. J Wildl Dis 46: 937–939.
6. Lopa AP, Sargo R, Rodrigues M, Cardoso I (2010) *High seroprevalence of antibodies to Toxoplasma gondii in wild birds from Portugal*. Parasitol Res 108: 1163–1169.
7. Molina Lépez R, Cabezon A, Palomín O, Darwich L, Oboñ E, et al. (2011) *High seroprevalence of Toxoplasma gondii and Neospora caninum in Common raven (Corvus corax) in the Northeast of Spain*. Res Vet Sci. In press;doi:10.1016/j.rvsc.2011.05.011.
8. Alcarado-Espavilh C, Rajendreren C, Ferreira I, Kwok O, Choudhury S, et al. (2011) *Prevalence of Toxoplasma gondii infection in wild birds in Durango, Mexico*. J Parasitol (In press). doi:10.1645/GE-2044.1.
9. Darwich L, Cabezon O, Echeverria I, Pabón M, Marco I, et al. (2011) *Presence of Toxoplasma gondii and Neospora caninum DNA in brain of wild birds*. Vet Parasitol (in press). doi:10.1016/j.vetpar.2011.07.024.
10. Tenter AM, Heckerth AR, Weiss LM (2000) *Toxoplasma gondii: From animals to humans*. Int J Parasitol 30: 1217–1258.
11. Gondim LS, Abe-Sandes K, Uze D, Silva MS, Santos SL, et al. (2010) *Toxoplasma gondii and Neospora caninum in sparrows (Passer domesticus) in the Northeast of Brazil*. Vet Parasitol 168: 121–124.
12. Dubey JP, Jones JL (2008) *Toxoplasma gondii infection in humans and animals in the United States*. Int J Parasitol 38: 1257–1275.
13. IUCN (2010) *IUCN Red List of Threatened Species*. Version 2010.1 website: Available: http://www.iucnredlist.org (accessed 2011, December 1st).
14. Dubey JP, Desmonts G (1987) *Serological responses of equids fed Toxoplasma gondii oocysts*. Equine Vet J 19: 337–339.
15. Godoi FS, Nishi SM, Pena HF, Gennari SM (2010) *Toxoplasma gondii: diagnosis of experimental and natural infection in pigeons (Columba livia) by serological, biological and molecular techniques*. Rev Bras Parasitol Vet 19: 238–243.
16. De Juana E, Varela JM (2001) *Guía de las aves de España*. Peninsula, Baleares y Canarias. Lynx Edicions, Barcelona. 225 p.
17. Houmer DW, Lemeshow S (1989) *Applied logistic regression*. Johns Wiley & Sons, New York, USA.
18. García-Bocanegra I, Simon-Griffith M, Dubey JP, Casal J, Martin GE, et al. (2010a) *Seroprevalence and risk factors associated with Toxoplasma gondii in domestic pigs from Spain*. Parasitol Int 59: 421–426.
19. García-Bocanegra I, Dubey JP, Martinez F, Vargas A, Cabezón O, et al. (2010b) *Factors affecting seroprevalence of Toxoplasma gondii in the endangered Iberian lynx (Lynx pardinus)*. Vet Parasitol 167: 36–42.
20. García-Bocanegra I, Cabezón O, Pabón M, Gómez-Guillamón F, Arenas A, et al. (2011) *Prevalence of Toxoplasma gondii and Neospora caninum antibodies in Spanish-Iberian vultures (Cathartes ayresii ibериaca)*. J Vet Med (in press);doi:10.1016/j.jvm.2011.09.011.
21. Gauss CB, Almería S, Ortúñor A, García F, Dubey JP (2003) *Seroprevalence of Toxoplasma gondii antibodies in domestic cats from Barcelona, Spain*. J Parasitol 89: 1067–1068.
22. Roelke ME, Johnson WE, Milland J, Palomares F, Revilla E, et al. (2008) *Exposure to disease agents in the endangered Iberian lynx (Lynx pardinus)*. Eur J Wildl Res 54: 171–178.
23. Kappurund G (1978) *Survey of toxoplasmosis in wild and domestic animals from Norway and Sweden*. J Wild Dis 14: 157–162.
24. Smith DD, Fenkel JK (1995) *Prevalence of antibodies to Toxoplasma gondii in wild mammals of Missouri and East Central Kansas: biologic and ecologic considerations of transmission*. J Wild Dis 31: 15–21.
25. Hejvicek K, Litesek J, Necalova I (1997) *Toxoplasmosis in wild mammals from the Czech Republic*. J Wild Dis 33: 400–405.
26. Zhang SY, Wei MX, Zhou ZY, Yu JY, Shi XQ (2000) *Prevalence of antibodies to Toxoplasma gondii in the sera of rare wildlife in the Shanghai Zoological Garden, people’s Republic of China*. Parasitol Int 49: 171–174.
27. Gauss CB, Dubey JP, Vidal D, Ruiz F, Vicente J, et al. (2005) *Seroprevalence of Toxoplasma gondii in wild pigs (Sus scrofa) from Spain*. Parasitology 131: 151–156.
28. Gauss CB, Dubey JP, Vidal D, Cabezón O, Ruiz-Fons F, et al. (2006) *Prevalence of Toxoplasma gondii antibodies in red deer (Cervus elaphus) and other wild ruminants from Spain*. Vet Parasitol 136: 193–200.
29. Wengel RM, Dubey JP, Siegel AM, Kitron UD, Mannelli A, et al. (1995) *Risk factors for transmission of Toxoplasma gondii on swine farms in Illinois*. J Parasitol 81: 736–741.
30. Daherza HA, Miller MA, Gardner IA, Packham AE, Avril EI, et al. (2000) *Risk factors for Toxoplasma gondii infection in wild rodents from central coastal California and a review of T. gondii prevalence in rodents*. J Parasitol 94: 675–683.
31. Pistorius PA (2008) Grey Heron (Ardea cinerea) predation on the Aldabra White-throated Rail (Dryolimnas cuvieri aldabranus). Wilson J Ornithol 120: 631–632.
32. Almería S, Calvete C, Pagés A, Gauss C, Dubey JP (2004) Factors affecting the seroprevalence of Toxoplasma gondii infection in wild rabbits (Oryctolagus cuniculus) from Spain. Vet Parasitol 123: 265–270.
33. Gamarra JA, Cabezon O, Pabón M, Arnal MC, Laco DF, et al. (2008) Prevalence of antibodies against Toxoplasma gondii in roe deer from Spain. Vet Parasitol 153: 152–156.
34. Margalida A, Bertran J, Heredia R (2009) Diet and food preferences of the endangered Bearded vulture Gypaetus barbatus: a basis for their conservation. Ibis 151: 235–243.
35. Margalida A, Benitez JR, Sanchez-Zapata JA, Avila E, Arenas R, et al. (2011) Long-term relationship between diet breadth and breeding success in a declining population of Egyptian Vultures Neophron percnopterus. Ibis (in press). doi: 10.1111/j.1474-919X.2011.01189.x.
36. Jones JL, Dubey JP (2010) Waterborne toxoplasmosis—recent developments. Exp Parasitol 124: 10–25.
37. Cabezon O, Resendes AR, Domingo M, Raga JA, Agusti C, et al. (2004) Seroprevalence of Toxoplasma gondii antibodies in wild dolphins from the Spanish Mediterranean coast. J Parasitol 90: 643–644.
38. Resendes AR, Almería S, Dubey JP, Obón E, Juan-Salles C, et al. (2002) Disseminated toxoplasmosis in a Mediterranean pregnant Risso’s dolphin (Grampus griseus) with transplacental fetal infection. J Parasitol 88: 1029–1032.