Correlations between Coronary Plaque Tissue Composition Assessed by Virtual Histology and Blood Levels of Biomarkers for Coronary Artery Disease

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Purpose: We investigated correlations of coronary plaque composition determined by virtual histology (VH) intravascular ultrasound (IVUS) and blood levels of biomarkers that represent the vulnerability of coronary plaques. Materials and Methods: Pre- and postprocedural blood levels of high sensitivity C-reactive protein, soluble CD40 ligand (sCD40L), matrix metalloproteinase-9, and neopterin were measured in 70 patients with stable angina (SA) or unstable angina (UA) who were undergoing percutaneous coronary intervention (PCI) for single lesions. We evaluated the data for correlations between these biomarkers and necrotic core contents in PCI target lesions analyzed by VH. Results: Clinical characteristics, IVUS, VH, and biomarker blood levels were not different between the SA and the UA group except for more frequent previous statin use (52.3% vs. 23.1%, p=0.017) and lower remodeling index in the SA group (0.98±0.09 vs. 1.10±0.070, p<0.001). Among the biomarkers evaluated, only pre-PCI neopterin level showed a weakly significant correlation with the absolute volume of the necrotic core (r=0.320, p=0.008). Pre- and post-PCI blood levels of sCD40L (r=0.220, p=0.072; r=0.231, p=0.062) and post-PCI blood level of neopterin (r=0.238, p=0.051) showed trends toward weakly positive correlations with the absolute volume of necrotic core. Conclusion: We found a weakly positive correlation between the pre-PCI neopterin level and necrotic core volume in the PCI-target lesion. The clinical implications of our findings need to be investigated in further studies.

Key Words: Atherosclerosis, coronary artery disease, inflammation, intravascular ultrasound

INTRODUCTION

The process of coronary plaque destabilization, rupture, and thrombosis is associated with the development of acute coronary syndrome (ACS).¹ Vulnerable atherosclerotic plaques are prone to rupture and are generally characterized by a large lipid core, the presence of inflammatory cells, a paucity of smooth muscle cells
and fibrous tissue, thinning of the fibrous cap, positive remodeling, and intimal neovascularization. Various molecules of inflammation and tissue proteolysis are activated in the formation of vulnerable plaques. C-reactive protein (CRP), soluble CD40 ligand (sCD40L), matrix metalloproteinase (MMP), and neopterin are known to be involved in the process, and their blood levels are elevated in patients with ACS.

Virtual histology (VH) of intravascular ultrasounds (IVUS) was developed to assess the tissue compositions of coronary plaques. VH uses spectral analysis of radiofrequency ultrasound signals and classifies plaques into four major components: fibrous, fibro-fatty, necrotic core, and dense calcium. Previous studies found that culprit lesions in patients with ACS have greater amounts of necrotic core than with target lesions in patients with stable angina (SA), and that necrotic core volume predicts microembolism and no reflow during percutaneous coronary intervention (PCI).

Therefore, the purpose of the present study was to investigate whether the relative content of necrotic core, determined by VH, is correlated with periprocedural blood levels of various biomarkers that represent the vulnerability of coronary plaques in patients undergoing PCI.

**MATERIALS AND METHODS**

**Patients**
A total of 70 consecutive patients with SA or unstable angina (UA), participating in the Real Safety and Efficacy of a 3-month Dual Antiplatelet Therapy Following Zotarolimus-eluting Stents Implantation (RESET) trial who underwent PCI at Severance Cardiovascular Hospital, Yonsei University Health System, Seoul, Korea, were enrolled in this study. Inclusion criteria were age >20 years, the presence of a single significant coronary artery stenosis (>50%) with a clinical presentation of stable or unstable angina, and the presence of a PCI target lesion with a reference diameter of 2.5-4 mm and a lesion length ≤26 mm that could be covered by a single stent. Exclusion criteria were hemodynamic instability, heart failure (left ventricular ejection fraction <40%), history of stroke, peripheral artery disease, significant hepatic (aspartate aminotransferase or alanine aminotransferase ≥3 times the upper limit of normal range) or renal failure (serum creatinine >2.0 mg/dL), previous use of drug-eluting stents, left main disease, bifurcation lesion requiring 2-stent technique, chronic total occlusion, restenotic lesion, all women with childbearing potential or women with pregnancy, known bleeding diathesis or bleeding history within the previous 3 months, and contraindications to antiplatelet agents. We also excluded patients with acute myocardial infarction because in these patients the blood levels of biomarkers might be influenced not only by the vulnerability of coronary plaques but also by the extent of myocardial necrosis. In addition, we excluded patients who took corticosteroids or other anti-inflammatory drugs except low-dose aspirin, and patients with any identifiable condition that was related to infectious, active allergic, connective tissue-related, or other chronic inflammatory diseases febrile conditions, acute infection, or known chronic inflammatory disease. Informed written consent was obtained from all patients, and the study protocol was approved by the institutional review board of Yonsei University Health System in accordance with the Declaration of Helsinki.

SA was defined as no change in the frequency, duration, or intensity of symptoms within 6 weeks before the intervention. UA was defined as chest pain occurring at rest and lasting longer than 20 min, new-onset chest pain of at least Canadian Cardiovascular Society class III severity, or previously diagnosed angina that has become distinctly more frequent, longer in duration, or lower in threshold.

**Percutaneous coronary intervention**
All patients were placed on chronic aspirin (100-325 mg/day) and clopidogrel (75 mg/day) therapy for ≥5 days or received aspirin (250 mg) and clopidogrel (300-600 mg) loading at least 12 hours before PCI. Unfractionated heparin was administered according to the local standards of care (target activated clotting time >250 sec) at a dosage consonant with label instructions. In all patients, PCI target lesions were predilated prior to stenting and treated with drug-eluting stents such as a zotarolimus-eluting stent (Endeavor Sprint® or Endeavor Resolute®, Medtronic Vascular Inc., Santa Rosa, CA, USA), sirolimus-eluting stent (Cypher®, Cordis, Johnson & Johnson, Miami Lakes, FL, USA) or everolimus-eluting stent (Xience V®, Abbott Vascular, Santa Clara, CA, USA). After implantation of the stent, aspirin (100 mg/day) was administered indefinitely, and clopidogrel (75 mg/day) was given for 3 or 12 months according to the RESET study protocol.

**Intravascular ultrasound and virtual histology**
IVUS investigations were performed in all PCI target le-
Fisher’s exact tests were used. Continuous data were ex-
guished chi-square tests. When the expected cell number was <5,
variables were expressed as percentages and compared us-
spatial software (SPSS Inc., Chicago, IL, USA). Categorical
All analyses were performed using SPSS version 18.0 statis-
technique.

which use a quantitative sandwich enzyme immunoassay
immunoassays (R&D Systems, Minneapolis, MN, USA),
sCD40L and neopterin were determined using Quantikine
(Seoul Clinical Laboratories, Seoul, Korea). Serum hsCRP
neopterin, were measured at an external clinical laboratory
for subsequent analysis. Investigational biomarkers, includ-
both plasma and serum were aliquoted and stored at -80°C
PCI. The blood samples were centrifuged immediately, and
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heparin and PCI in the catheterization room and 24 h after
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for subsequent analysis. Investigational biomarkers, including
high sensitivity CRP (hsCRP), MMP-9, sCD40L, and
neopterin, were measured at an external clinical laboratory
(Seoul Clinical Laboratories, Seoul, Korea). Serum hsCRP
was measured using a latex-enhanced turbidimetric immu-
nounoassay. The serum levels of MMP-9 and plasma levels of
sCD40L and neopterin were determined using Quantikine®
imunoassays (R&D Systems, Minneapolis, MN, USA),
which use a quantitative sandwich enzyme immunoassay
 technique.

All analyses were performed using SPSS version 18.0 statis-
tical software (SPSS Inc., Chicago, IL, USA). Categorical
variables were expressed as percentages and compared us-
ing chi-square tests. When the expected cell number was <5,
Fisher’s exact tests were used. Continuous data were ex-
pressed as mean±SD. Continuous variables were analyzed
using unpaired t-tests and the Mann-Whitney U-test, as ap-
propriate. Categorical data and proportions were analyzed
using the χ²-test or Fisher’s exact test, as required. Pearson’s
two-way test was used to assess the relationships between
two quantitative variables with normal distributions. For all
tests, two-tailed p values <0.05 were considered signif-

tant

RESULTS

Baseline clinical characteristics
Among the 70 enrolled patients (age 60.4±8.9 years, male
57.1%), there were 26 UA and 44 SA patients. There were
no significant differences between the two patient groups in
baseline characteristics such as age, gender, diabetes melli-
tus, hypertension, hypercholesterolemia, current smoking,
renal failure, previous stroke, and multivessel disease (Table 1).
However, the SA group had a higher frequency of
previous statin use (52.3% vs. 23.1%, p=0.017). Conse-
sequently, total cholesterol (170.4±37.7 vs. 194.6±38.2 mg/
dl, p=0.013) and low density lipoprotein cholesterol levels
(95.5±34.1 vs. 114.9±37.4 mg/dL, p=0.032) were lower in
the SA group than in the UA group.

Angiographic characteristics and procedural data
Angiographic characteristics and procedural data did not
differ significantly between the UA and SA patient groups
and are summarized in Table 2. The most common target
 lesion location was the left anterior descending artery. The
majority of the lesions were type B2 or C lesions according
to the American College of Cardiology/American Heart As-

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IVUS and virtual histology data
Preprocedural IVUS and VH data are presented in Table 3.
There were no significant differences in vessel, lumen, and
P&M area at the target lesion cross section with the mini-
imum lumen area or in the vessel, lumen, and P&M volume
of the target lesion between the SA and UA patient groups.
VH also demonstrated no significant intergroup differences
in plaque composition at the target lesion cross section with
minimum lumen area or in the tissue component of the whole
target lesion. However, patients with UA showed a trend to-
ward greater necrotic volume (9.4±3.8% vs. 7.5±3.7%,

Laboratory assessment
Blood samples were obtained before bolus administration of
heparin and PCI in the catheterization room and 24 h after
PCI. The blood samples were centrifuged immediately, and
both plasma and serum were aliquoted and stored at -80°C
for subsequent analysis. Investigational biomarkers, including
high sensitivity CRP (hsCRP), MMP-9, sCD40L, and
neopterin, were measured at an external clinical laboratory
(Seoul Clinical Laboratories, Seoul, Korea). Serum hsCRP
was measured using a latex-enhanced turbidimetric immu-
nounoassay. The serum levels of MMP-9 and plasma levels of
sCD40L and neopterin were determined using Quantikine®
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 technique.
in pre- and postprocedural blood levels of these biomarkers. However, there was a significant intergroup difference in blood sCD40L levels observed before and after the procedure. The UA patient group showed a significantly greater increase in the sCD40L level than the SA group (2.9 ± 0.8 vs. -0.1 ± 0.3 ng/mL, p = 0.043). Table 5 shows the correlations between the necrotic core content in the target lesion and the biomarker blood levels. Among the biomarkers evaluated, 7.6%, p = 0.110) than SA patients. The remodeling index was significantly greater for the UA group than the SA group (1.10 ± 0.070 vs. 0.98 ± 0.09, p < 0.001).

### Blood levels of biomarkers

Pre- and postprocedural blood levels of hsCRP, sCD40L, MMP-9, and neopterin are shown in Table 4. Between the SA and UA patient groups, there were no significant differences in pre- and postprocedural blood levels of these biomarkers. However, there was a significant intergroup difference in blood sCD40L levels observed before and after the procedure. The UA patient group showed a significantly greater increase in the sCD40L level than the SA group (2.9 ± 0.8 vs. -0.1 ± 0.3 ng/mL, p = 0.043). Table 5 shows the correlations between the necrotic core content in the target lesion and the biomarker blood levels. Among the biomarkers evaluated,
DISCUSSION

The major findings of the present study are as follows: 1) The relative content of necrotic core tended to be higher and the relative content of fibro-fatty tissue tended to be lower in patients with unstable angina than in patients with stable angina. 2) Pre- and post-PCI blood levels of biomarkers such as hsCRP, sCD40L, MMP-9, and neopterin showed a mildly significant correlation with the absolute volume of the necrotic core in the target lesion (r=0.320, p=0.008). Pre- and post-PCI blood levels of sCD40L (r=0.220, p=0.072; r=0.231, p=0.062) and post-PCI blood level of neopterin (r=0.238, p=0.051) also showed trends toward positive correlation with the absolute volume of necrotic core in the target lesion. However, other biomarker levels were not correlated with relative or absolute content of necrotic core.

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Table 3. Preprocedural Intravascular Ultrasound and Virtual Histology Data for the Target Lesions

|                      | Stable angina (n=44) | Unstable angina (n=26) | p value |
|----------------------|----------------------|------------------------|---------|
| Target lesion CSA with minimum lumen area |                      |                        |         |
| EEM CSA, mm$^2$      | 14.0±4.0             | 14.3±3.9               | 0.754   |
| Lumen CSA, mm$^2$    | 2.5±0.6              | 2.7±0.9                | 0.323   |
| P&M CSA, mm$^2$      | 11.5±3.9             | 11.7±3.8               | 0.891   |
| Plaque burden, %     | 81.3±5.5             | 80.4±6.5               | 0.557   |
| Fibrous CSA, %       | 67.2±13.2            | 65.2±14.5              | 0.555   |
| Fibrofatty CSA, %    | 10.2±5.5             | 8.3±4.2                | 0.126   |
| NC CSA, %            | 14.3±9.8             | 15.4±8.5               | 0.631   |
| DC CSA, %            | 8.3±7.3              | 11.1±10.4              | 0.186   |
| Remodeling index     | 0.98±0.09            | 1.10±0.07              | <0.001  |
| Whole target lesion volume |                      |                        |         |
| EEM volume, mm$^3$   | 257.8±92.3           | 249.1±88.4             | 0.701   |
| Lumen volume, mm$^3$ | 98.9±36.3            | 95.7±36.1              | 0.723   |
| P&M volume, mm$^3$   | 158.8±65.5           | 153.3±63.9             | 0.735   |
| Plaque burden, %     | 61.3±7.9             | 60.9±8.7               | 0.853   |
| Fibrous volume, %    | 65.3±10.8            | 62.4±12.2              | 0.313   |
| Fibrofatty volume, % | 9.4±3.8              | 7.5±3.7                | 0.052   |
| NC volume, %         | 15.1±8.1             | 18.3±7.6               | 0.110   |
| DC volume, %         | 9.4±6.5              | 11.4±7.2               | 0.226   |

CSA, cross sectional area; EEM, external elastic membrane; P&M, plaque and media; NC, necrotic core; DC, dense calcium.

Table 4. Pre- and Postprocedural Blood Levels of Biomarkers

| Biomarker              | Stable angina (n=44) | Unstable angina (n=26) | p value |
|------------------------|----------------------|------------------------|---------|
| hsCRP (mg/L)           |                      |                        |         |
| Pre-PCI                | 0.7±0.7              | 1.2±1.3                | 0.148   |
| Post-PCI at 24 hrs     | 2.0±2.2              | 2.7±2.1                | 0.203   |
| Δ                      | 1.2±1.9              | 1.5±1.7                | 0.490   |
| Soluble CD40 ligand (ng/mL) |                  |                        |         |
| Pre-PCI                | 2.6±2.3              | 2.0±2.6                | 0.376   |
| Post-PCI at 24 hrs     | 2.6±3.0              | 3.4±3.1                | 0.299   |
| Δ                      | -0.1±0.3             | 2.9±0.8                | 0.043   |
| MMP-9 (μg/L)           |                      |                        |         |
| Pre-PCI                | 303.1±112.3          | 359.3±190.9            | 0.189   |
| Post-PCI at 24 hrs     | 826.9±258.7          | 799.0±282.8            | 0.691   |
| Δ                      | 505.5±256.4          | 451.0±214.7            | 0.390   |
| Neopterin (nmol/L)     |                      |                        |         |
| Pre-PCI                | 5.1±2.7              | 5.5±3.4                | 0.543   |
| Post-PCI at 24 hrs     | 6.5±3.1              | 6.4±3.3                | 0.580   |
| Δ                      | 1.4±1.5              | 0.8±0.9                | 0.087   |

hsCRP, high sensitivity C-reactive protein; PCI, percutaneous coronary intervention; MMP, matrix metalloproteinase.
Inflammation plays a key role in atherosclerosis and plaque vulnerability. CRP is a liver-derived acute phase protein involved in innate immune response. CRP has been found to induce the expression of adhesion molecules, such as Vascular Cell Adhesion Molecule-1, Inter-Cellular Adhesion Molecule-1 and E-selectin, as well as monocyte chemoattractant factor-1. CRP also increases tissue factor expression in monocyte-macrophages; promotes monocyte chemotaxis and adhesion to endothelial cells; stimulates the release of reactive oxygen species and MMP-1; and pro-

**Table 5. Correlations of Necrotic Core Contents Determined by Virtual Histology with Periprocedural Blood Levels of Biomarkers**

| Necrotic core content | Biomarkers             | r     | p value |
|-----------------------|------------------------|-------|---------|
| Relative content of necrotic core at the cross section with minimum lumen area | hsCRP (pre-PCI) | 0.088 | 0.474   |
|                       | hsCRP (post-PCI 24 hrs) | 0.081 | 0.513   |
|                       | hsCRP (Δ)               | 0.067 | 0.590   |
|                       | sCD40L (pre-PCI)        | 0.170 | 0.165   |
|                       | sCD40L (post-PCI 24 hrs) | 0.172 | 0.168   |
|                       | sCD40L (Δ)              | -0.016 | 0.894   |
|                       | MMP-9 (pre-PCI)         | 0.173 | 0.165   |
|                       | MMP-9 (post-PCI 24 hrs) | 0.077 | 0.546   |
|                       | MMP-9 (Δ)               | 0.033 | 0.794   |
|                       | Neopterin (pre-PCI)     | 0.097 | 0.433   |
|                       | Neopterin (post-PCI 24 hrs) | 0.032 | 0.797   |
|                       | Neopterin (Δ)           | -0.140 | 0.255   |
| Relative content of necrotic core in the target lesion volume | hsCRP (pre-PCI) | 0.109 | 0.376   |
|                       | hsCRP (post-PCI 24 hrs) | 0.125 | 0.314   |
|                       | hsCRP (Δ)               | 0.093 | 0.453   |
|                       | sCD40L (pre-PCI)        | 0.179 | 0.166   |
|                       | sCD40L (post-PCI 24 hrs) | 0.234 | 0.058   |
|                       | sCD40L (Δ)              | -0.043 | 0.726   |
|                       | MMP-9 (pre-PCI)         | 0.161 | 0.197   |
|                       | MMP-9 (post-PCI 24 hrs) | 0.112 | 0.382   |
|                       | MMP-9 (Δ)               | 0.024 | 0.853   |
|                       | Neopterin (pre-PCI)     | 0.155 | 0.207   |
|                       | Neopterin (post-PCI 24 hrs) | 0.071 | 0.567   |
|                       | Neopterin (Δ)           | -0.178 | 0.147   |
| Absolute necrotic core volume in the target lesion | hsCRP (pre-PCI) | 0.086 | 0.486   |
|                       | hsCRP (post-PCI 24 hrs) | 0.152 | 0.221   |
|                       | hsCRP (Δ)               | 0.149 | 0.224   |
|                       | sCD40L (pre-PCI)        | 0.220 | 0.072   |
|                       | sCD40L (post-PCI 24 hrs) | 0.231 | 0.062   |
|                       | sCD40L (Δ)              | -0.060 | 0.629   |
|                       | MMP-9 (pre-PCI)         | 0.155 | 0.213   |
|                       | MMP-9 (post-PCI 24 hrs) | 0.025 | 0.844   |
|                       | MMP-9 (Δ)               | -0.022 | 0.861   |
|                       | Neopterin (pre-PCI)     | 0.320 | 0.008   |
|                       | Neopterin (post-PCI 24 hrs) | 0.238 | 0.051   |
|                       | Neopterin (Δ)           | -0.153 | 0.214   |

hsCRP, high sensitivity C-reactive protein; PCI, percutaneous coronary intervention; sCD40L, soluble CD40 ligand; MMP, matrix metalloproteinase.

did not differ between the SA group and the UA group. However, the increase in the blood level of sCD40L after PCI was significantly higher in UA patients than in SA patients. 3) Among the various biomarkers evaluated, only pre-PCI blood level of neopterin showed a weakly significant correlation with the absolute volume of the necrotic core in the target lesion. Pre- and post-PCI blood levels of sCD40L and post-PCI blood level of neopterin also showed trends toward a positive correlation with the absolute volume of necrotic core in the target lesion.
motes oxidized low-density lipoprotein uptake, which results in increased foam cell formation. recently, several studies discovered CRP in coronary and carotid atherosclerosis. However, a question of whether CRP directly contributes to atherogenesis is still controversial. Activated platelets express CD40 ligand, which is then cleaved and released as sCD40L. The sCD40L binds to circulating monocytes through both its receptor, CD40, and through the monocyte/macrophage integrin Mac-1, and this binding promotes monocyte adhesion to the vascular endothelium. The sCD40L also binds to CD40 on endothelial cell surfaces, which activates the expression of adhesion molecules on endothelial cell surfaces. MMPs are a family of more than 20 zinc-containing endoproteinas that contribute to collagen degradation and weakening of the fibrous cap. Increased expressions of MMP-1, -3 and -9 have been found at the rupture prone shoulder regions of atherosclerotic plaques. Neopterin is also produced by activated macrophages and is thought to be a marker of immune activation and macrophage activity. Neopterin can act as a pro-oxidant, enhancing oxidant production and promoting cell death or atherosclerotic plaque instability. Neopterin can act as a pro-oxidant, enhancing oxidant production and promoting cell death or atherosclerotic plaque instability. Avanzas, et al. 30 reported that serum neopterin is an independent predictor of major adverse coronary events in patients with chronic stable angina pectoris, and Zouridakis, et al. 31 also demonstrated that increased serum neopterin levels were associated with the rapid progression of coronary artery disease. The blood levels of these biomarkers are reported to be increased in patients with ACS. However, only a few in vivo studies have investigated the relationship between estimates of tissue composition determined by VH and periprocedural blood levels of biomarkers. Kubo, et al. 34 demonstrated a weak but positive correlation between hsCRP and the amount of necrotic core in the target lesion, and Otake, et al. 35 also found a positive correlation between serum hsCRP and the necrotic core ratio and an inverse relationship between serum adiponectin and the necrotic core ratio in both culprit and non-culprit lesions in patients with ACS. Park, et al. 36 measured plasma levels of MMP-2 and -9, tissue inhibitor of MMP-1, adiponectin, and macrophage migration inhibitory factor in patients with coronary artery disease and found that only the MMP-9 level showed a weak correlation with the total number of ruptured plaques in both target and non-target vessels. In the present study, we found only a weak correlation between pre-PCI blood level of neopterin and necrotic volume in the PCI target lesion. We also observed that the increase in the blood level of sCD40L after coronary stenting was significantly higher in UA patients than in SA patients. Previous studies have shown that blood levels of inflammatory biomarker rise after PCI possibly due to vascular and myocardial injury related to the procedure. Kozinski, et al. 38 also reported that the rise in the serum levels of hsCRP and serum amyloid A after coronary intervention was greater for UA compared with SA. Although the clinical implication of elevated inflammatory biomarkers after coronary stenting is not fully clarified, a correlation between the levels of CRP after coronary stenting and the cardiovascular event rate during follow-up has been suggested. 39,40

There are several potential explanations for weak or poor correlations between the biomarker levels and the relative content of necrotic core by VH in our study. First, our study sample may have been too small to detect any significant relationships between tissue composition analyzed by VH and blood biomarker levels. Second, the blood levels of biomarkers generally represent systemic inflammation and may not reflect local inflammation status or tissue characteristics of PCI target lesions. For instance, we cannot rule out the possibility that the blood level of biomarkers may have been more affected by non-PCI target lesion with mild-to-moderate plaque burden and/or non-coronary atherosclerotic lesions than by the PCI target lesions. Furthermore, non-vascular origins of inflammation may have been responsible for the status of systemic inflammation, although we excluded patients with known infectious or inflammatory disease. Third, potential impact of previous use of statin on the results of the present study cannot be ruled out. Statins are known to reduce blood levels of inflammatory biomarkers and to exert effects on vascular remodeling and plaque composition. However, whether statins had different degrees of influence on blood levels of biomarkers and tissue composition of target lesions in our study cannot be clarified within the limits of the present study. Fourth, tissue characterization by VH may not be sufficient. VH validation studies on human coronary arteries ex vivo have focused solely on the detection of particular tissue types in limited regions of interest. However, VH has never been validated for determination of the absolute or relative areas of any particular tissue component in human coronary arteries. Recently, Thim, et al. 41 reported that the necrotic cores of minipig coronary artery plaques assessed by VH did not correlate with pathologic findings, especially when calcifications in the necrotic core were not present or not detected. Therefore, VH may require more pathologic vali-
dation to support its reliability.

In conclusion, we found a weak correlation between the pre-PCI neopterin level and necrotic core volume in the PCI-target lesion. The clinical implications of our findings need to be investigated in further studies.

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