Whole-Genome Sequences of Five *Geobacillus stearothermophilus* Strains Isolated from Processing Lines of Powdered Infant Formula

Tae Jin Cho, Dongmin Yang, Byeonghyeok Park, In-Geol Choi, Min Suk Rhee

*Department of Biotechnology, College of Life Sciences and Biotechnology, Korea University, Seoul, South Korea*

**ABSTRACT** *Geobacillus stearothermophilus* is the thermophile present in processing lines of powdered infant formula (PIF). We report the whole-genome sequences of *G. stearothermophilus* strains isolated from work-in-process products (sterilized and concentrated milk) of manufacturing plants. Understanding the genomic basis governing the metabolism of *G. stearothermophilus* can contribute to the safety management of PIF during its manufacture.

*Geobacillus stearothermophilus* is a Gram-positive spore-forming thermophile ubiquitously present in powdered infant formula (PIF) processing plants (1, 2). The general processing steps of PIF consist of the blending of raw ingredients, heat sterilization, condensation, and spray drying (3). Since the high heat resistance of *G. stearothermophilus* spores means the bacterium can endure the heat treatment processes during the manufacture of PIF, growth of *G. stearothermophilus* in poststerilization steps has been regarded to be the major cause of the production of spoilage metabolites (4, 5). To understand the metabolic pathways and major determinants for metabolism which result in the deterioration of PIF products, functional genes governing key metabolisms should be identified by genomic analysis.

We sequenced five strains of *G. stearothermophilus* obtained from the work-in-process products (WIP) of PIF processing lines through the following isolation method: WIP samples (25 ml of concentrated or pasteurized milk) (Table 1) were homogenized with 225 ml of sterile 0.85% (wt/vol) saline using a stomacher (Circulator 400, Seward) at 230 rpm for 2 min. One milliliter of homogenized sample was added in 0.85% saline for serial 10-fold dilutions, and 100 μl of diluted sample was spread plated on plate count agar (PCA; Difco, Sparks, MD, USA), followed by incubation at 55°C for 48 h. Single colonies on PCA were substreaked onto tryptic soy agar (TSA; Difco, Becton, Dickinson), and the TSA was incubated, followed by the propagation of a single colony on those plates in 5 ml tryptic soy broth (TSB; Difco) at 55°C for 48 h. Bacterial suspensions were mixed with a 50% glycerol solution to prepare stock cultures and stored at −80°C.

Each stock culture (50 μl) was incubated in 4.95 ml TSB at 55°C for 24 h. The enriched culture was streaked onto TSA and propagated at 55°C for 24 h. Then, a single colony from a subcultured TSA plate was inoculated in 5 ml TSB for incubation at 55°C, and the enriched culture with a cell density of ca. 9-log CFU/ml (incubation time was set by the preliminary tests) was used for the extraction of genomic DNA (gDNA). The DNeasy blood and tissue kit (Qiagen, Valencia, CA) was utilized to extract gDNA from a 1-ml aliquot of bacterial culture after the pretreatment specific for Gram-positive bacteria according to the manufacturer’s instructions.

A library for whole-genome sequencing was prepared with a Nextera DNA library prep kit (Illumina, Inc., San Diego, CA, USA). Sequencing was carried out with 600 cycles of reads by the Illumina MiSeq platform. An average 2,251,205 paired-end reads with a read size of 301 × 2 bp were obtained (Table 1). Trimming of the adapter sequences and
low-quality bases (<Q20) from raw reads was performed using the Trim Galore! software (version 0.4.4; https://www.bioinformatics.babraham.ac.uk/projects/trim_galore/). An average 2,236,883 reads per strain could pass the quality control and were assembled using the SPAdes genome assembler (version 3.12.0) (6). The assemblies were annotated preliminarily using the Prokaryotic Contigs Annotation Pipeline Server (P-CAPS, version 0.1) with default parameters (7). The final annotation was generated with the NCBI Prokaryotic Genome Annotation Pipeline (PGAP, version 4.6) with default parameters (8).

A summary of the characteristics and genome features of the five strains of *G. stearothermophilus* is presented in Table 1. The general statistics for the genomes were as follows: sequencing reads in total (paired ends), 1,487,742 to 2,651,209 sequences (seqs); total read length, 815 to 1,507 Mbp; total length of assembled sequence, 2,667,663 to 2,894,956 bp; number of contigs, 150 to 330; overall GC content, 52.13 to 52.94%; and coverage depth, 470.42- to 875.59-fold.

The whole-genome sequence information reported here is expected to broaden our knowledge regarding the genetic and functional characteristics of *G. stearothermophilus* with regard to the production of metabolites during the manufacture of PIF.

**Data availability.** All genome sequences and raw reads reported here were deposited in GenBank under the accession, BioSample, and BioProject numbers listed in Table 1. The versions described in this paper are the first versions.

**ACKNOWLEDGMENTS**

This work was supported by the Korea Institute of Planning and Evaluation for Technology in Food, Agriculture, Forestry (IPET) through the High Value-added Food Technology Development Program, funded by the Ministry of Agriculture, Food and Rural Affairs (MAFRA) (grant 314046-3). This research was also supported by a Korea University grant.

We thank the School of Life Sciences and Biotechnology of Korea University for BK 21 PLUS and the Institute of Biomedical Science and Food Safety, CJ-Korea University Food Safety Hall for providing the equipment and facilities.

We declare no competing interests.

**REFERENCES**

1. Yuan DD, Liu GC, Ren DY, Zhang D, Zhao L, Kan CP, Yang YZ, Ma W, Li Y, Zhang LB. 2012. A survey on occurrence of thermophilic bacilli in commercial milk powders in China. Food Control 25:752–757. https://doi.org/10.1016/j.foodcont.2011.12.020.

2. McHugh AJ, Feehily C, Hill C, Cotter PD. 2017. Detection and enumeration of spore-forming bacteria in powdered dairy products. Front Microbiol 8:109. https://doi.org/10.3389/fmicb.2017.00109.

3. Jiang YJ, Guo M. 2014. Processing technology for infant formula, p
211–229. In Guo M (ed), Human milk biochemistry and infant formula manufacturing technology, 1st ed. Woodhead Publishing, Sawston, United Kingdom.

4. Cho TJ, Kim HW, Kim NH, Park SM, Kwon JI, Kim YJ, Lee KW, Rhee MS. 2018. New insights into the thermophilic spore-formers in powdered infant formula: implications of changes in microbial composition during manufacture. Food Control 92:464–470. https://doi.org/10.1016/j.foodcont.2018.05.036.

5. Burgess SA, Lindsay D, Flint SH. 2010. Thermophilic bacilli and their importance in dairy processing. Int J Food Microbiol 144:215–225. https://doi.org/10.1016/j.ijfoodmicro.2010.09.027.

6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.

7. Park B, Baek M-J, Min B, Choi I-G. 2017. Prokaryotic Contig Annotation Pipeline Server: Web application for a prokaryotic genome annotation pipeline based on the shiny app package. J Comput Biol 24:917–922. https://doi.org/10.1089/cmb.2017.0066.

8. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.