Regulation of Na\(^+\) and K\(^+\) homeostasis in plants: towards improved salt stress tolerance in crop plants

Diego M. Almeida, M. Margarida Oliveira and Nelson J. M. Saibo

1Genomics of Plant Stress Unit, Instituto de Tecnologia Química e Biológica António Xavier, Universidade Nova de Lisboa and Instituto de Biologia Experimental e Tecnológica, Oeiras, Portugal.

Abstract

Soil salinity is a major abiotic stress that results in considerable crop yield losses worldwide. However, some plant genotypes show a high tolerance to soil salinity, as they manage to maintain a high K\(^+\)/Na\(^+\) ratio in the cytosol, in contrast to salt stress susceptible genotypes. Although, different plant genotypes show different salt tolerance mechanisms, they all rely on the regulation and function of K\(^+\) and Na\(^+\) transporters and H\(^+\) pumps, which generate the driving force for K\(^+\) and Na\(^+\) transport. In this review we will introduce salt stress responses in plants and summarize the current knowledge about the most important ion transporters that facilitate intra- and intercellular K\(^+\) and Na\(^+\) homeostasis in these organisms. We will describe and discuss the regulation and function of the H\(^+\)-ATPases, H\(^+\)-PPases, SOS1, HKTs, and NHXs, including the specific tissues where they work and their response to salt stress.

Keywords: Salinity, sodium, potassium, proton pumps, ion transporters.

Received: April 13, 2016; Accepted: August 18, 2016.

Salt stress effects on plant growth and yield

Soil salinity is a major environmental constraint to crop production, affecting millions of hectares of land throughout the world and costing billions of dollars every year (Munns, 2005; Munns and Tester, 2008; Shabala and Cuin, 2008). High salinity affects over 6% of the world’s total land area. Most of this affected land has arisen from natural causes, such as rainfall, windblown salt from ocean, tsunamis, and rock weathering. Apart from natural causes, soil salinization is commonly associated to land clearing by removal of deep root vegetation, thus accumulating more water and consequently raising the levels of salty groundwater, or irrigation practices, such as the use of water with high salt concentration. Currently it is estimated that 20% of the total irrigated land is salt-affected. Given that irrigated land produces at least twice as much as rain-fed land and is responsible for one third of the world’s food production, it raises awareness for salinity as a serious problem for crop productivity (Munns, 2005; Munns and Tester, 2008).

High soil salinity is a condition characterized by a high concentration of soluble salts, in which NaCl is the most soluble and widespread salt. Soils are classified as saline when the electrical conductivity (EC) is 4 dS/m (= 40 mM NaCl) or higher. At this soil salt concentration, growth and yield of most crops are significantly reduced. Rice, as well as most crop plants, is a glycophyte and therefore it can only tolerate relatively low concentrations of salt. Among cereal crops, rice is the most salt sensitive one, showing salt stress symptoms and reduced yield even when the EC is lower than 4.0 dS/m (Munns and Tester, 2008). The salinity threshold for rice is 3.0 dS/m with a 12% reduction in yield per dS/m beyond this threshold (Gao et al., 2007). However, some degree of genotype diversity for salt stress tolerance is available in rice germplasm. Among 180,000 rice genotypes screened by the International Rice Research Institute (IRRI, 2013), 17% showed acceptable tolerance at an EC of 10 dS/m at seedling stage (Gregorio et al., 2002).

High salinity affects plants in two distinct phases. The first phase is the osmotic effect, which is independent of the accumulation of salt in the shoot. Salts dissolved in the soil solution reduce the soil water potential. This makes the water uptake from roots thermodynamically hampered and induces water deficit (Pardo, 2010; Roy et al., 2014). A water deficit signal is rapidly transmitted (within minutes) from roots to shoots and will cause intracellular turgor reduction and decreased cell expansion (Munns, 2005; Munns and Tester, 2008). This signal also promotes the biosynthesis of abscisic acid (ABA), which leads to a lower stomatal conductance (Munns, 2005; Munns and Tester, 2008; Roy et al., 2014). The lower stomatal conductance causes a lower carbon assimilation, biomass production and decreased yield. The second phase of salinity is ionic specific; this is due to the accumulation to toxic concentrations of sodium.
(Na⁺) and/or chloride (Cl⁻) ions, especially in the older leaves, inducing tissue necrosis and early leaf senescence (Roy et al., 2014). For most plant species Na⁺ appears to reach a toxic concentration earlier than Cl⁻ (Tester and Davenport, 2003). For rice Na⁺ has been shown to be the primary toxic ion (Chi Lin and Huei Kao, 2001; Tsai et al., 2004). Both osmotic and ionic effects disturb aerobial metabolism and induce the accumulation of reactive oxygen species (ROS) beyond the plant’s capacity for cellular oxidant detoxification, which in turn negatively affects cellular structures and metabolism (Chaves and Oliveira, 2004; Chaves et al., 2009).

A deleterious effect imposed by salt stress, during the second phase, is ion imbalance (Munns and Tester, 2008). Potassium (K⁺) is an essential macronutrient that plays important functions related to enzyme activation, osmotic adjustment and turgor generation, regulation of membrane potential, and cytoplasmic pH homeostasis (PPI, 1998; Barragan et al., 2012). Due to similarity in physicochemical properties between Na⁺ and K⁺ (i.e., ionic radius and ion hydration energy), the former competes with K⁺ for major binding sites in key metabolic processes in the cytoplasm, such as enzymatic reactions, protein synthesis and ribosome functions (Marschner, 1995; PPI, 1998). Na⁺ inhibits the enzyme activity of many enzymes that require K⁺ for functioning (Duggleby and Dennis, 1973). With over 50 different cytoplasmic enzymes being activated by K⁺, disruption of the K⁺ homeostasis leads to severe metabolism impairment, both in root and leaf tissues (Marschner, 1995; PPI, 1998). It has been suggested that plant survival under salt stress requires a high cytosolic K⁺/Na⁺ ratio in the cytoplasm. The restriction of Na⁺ accumulation in shoots under salt stress has been correlated with salt stress tolerance in rice (Lutts et al., 1996) and maize (Zea mays L.) (Tester and Davenport, 2003).

Sodium uptake from soil, sensing and signaling mechanisms

The very low membrane potential across the plasma membrane of root cells (more negative inside) promotes the passive transport of Na⁺ into the cells, and especially so when the sodium concentration increases in the soil solution. In contrast, Na⁺ efflux (i.e., removal from the cell) is not passive and requires energy expenditure (Maathuis et al., 2014). The passive Na⁺ uptake into root cells at high soil salinity is mainly mediated by a family of Non-Selective Cation Channels (NSCCs family), for which the molecular identity remains largely unknown (Blumwald et al., 2000; Kronzucker and Britto, 2011) (Figure 1). In addition to the Na⁺ flow across cellular membranes to enter into root cells (symplast flow), it has been reported that, at least in some species, interruptions in the endodermis (passage cells) allow the movement of water and solutes (i.e., Na⁺) through the cell wall and intercellular spaces. This type of transport into the xylem stream, without crossing the plasma membrane, is referred as “apoplast flow” (Yeo et al., 1987; Kronzucker and Britto, 2011) (Figure 1). Casparian strips and suberine layers in the root endoderm and exodermal layers provide some barrier to apoplast flow (Yeo et al., 1987). In many plant species, such as rice, the apoplast flow is considered to be the major port of Na⁺ entry (~50% of total Na⁺ uptake) (Yeo et al., 1987), especially at high salinity levels, and is responsible for a significant amount of Na⁺ transported to the shoot (Yeo et al., 1987; Kronzucker and Britto, 2011). Na⁺ ions taken up by the roots are then transported to shoots via xylem vessels by bulk flow (Figure 1). This is driven by the tension in the xylem, which causes the continuous movement of water from the root through the plant to the surrounding atmosphere during transpiration (Nobel, 2009).

Sodium has also a strong inhibitory effect on K⁺ uptake by cells, probably by inhibiting K⁺ transporters, such as AKT1 (hyperpolarization-activated inward-rectifying K⁺ channel), a major player in K⁺ acquisition by plants (Hirsch et al., 1998; Fuchs et al., 2005), and HAK5 (carrier-type HUP/HAK/KT transport) (Nieves-Cordones et al., 2010), both present in the plasma membrane of root cells. Additionally, membrane depolarization caused by large cytosolic Na⁺ influx results in increased K⁺ efflux, possible through depolarization-activated outward-rectifying K⁺ channels (e.g., GORK) (Adams and Shin, 2014) and also NSCCs (Sun et al., 2009).

Very little is known about how Na⁺ is sensed in most cellular systems. In theory, Na⁺ can be sensed either outside or inside the cell, or both. Extracellular Na⁺ may be sensed by a membrane receptor, whereas intracellular Na⁺ may be sensed either by membrane proteins or by any of the Na⁺ sensitive enzymes in the cytoplasm (Conde et al., 2011). The plasma membrane Na⁺/H⁺ antiporter SOS1 (SALT OVERLY SENSITIVE 1) has been described as a possible Na⁺ sensor (Shi et al., 2000). Its transport activity is essential for Na⁺ efflux from cells (Quintero et al., 2002), but its unusually long cytoplasmatic tail is thought to be involved in Na⁺ sensing (Shi et al., 2000) (Figure 2). However, this mechanism is not fully clear.

In plant cells, Ca²⁺ acts as a second messenger connecting a wide range of extracellular stimuli with various intracellular responses (Conde et al., 2011). Salt stress originates a fast and transient increase in free cytosolic Ca²⁺, likely released from the vacuole (Pottosin et al., 2009), which is decoded by Ca²⁺ sensors, such as calmodulin (CaM), calcineurin B-like proteins (CBLs) and CBL-interacting protein kinases (CIPKs). When acting as a CBL-CIPK complex, these Ca²⁺ sensors are often designed as calcium-dependent protein kinases (CDPKs) (Yang and Poo-vaiah, 2003; Conde et al., 2011). Cytosolic Ca²⁺ sensors in turn trigger many signal transduction pathways involved in the regulation of ion channels activity (e.g., NSCCs are strongly blocked by external Ca²⁺), as well as enzymatic activity and gene transcription, ending up in ion homeostasis (Pardo and Quintero, 2002; Yamaguchi et al., 2005; Marti-
Mechanisms of salt tolerance in plants

Salt stress frequently affects plant habitats and many species evolved different mechanisms to cope with it. The mechanisms for salt tolerance can be classified into three main categories. The first one is osmotic stress tolerance, which is regulated by long distance signals that reduce shoot growth (Roy et al., 2014) and involves the biosynthesis and accumulation of compatible solutes to maintain water uptake (Peleg et al., 2011). The second mechanism is ion exclusion, in which Na⁺ transporters reduce the accumulation of toxic Na⁺ within roots and leaves. This system operates by controlling the Na⁺ loading into the xylem and Na⁺ retrieval from the xylem, before reaching the photosynthetic tissues in the shoot (Figure 1). Finally, the third mechanism is tissue tolerance, in which high salt concentration is found in leaves, but Na⁺ is compartmentalized at the cellular and intracellular level (especially in the vacuole) reducing the deleterious effect of Na⁺ in the cytosol and driving water uptake to cells (Figure 1) (Munns and Tester, 2008). In most cases, the plant salt stress tolerance relies on the three mechanisms together, rather than on only one mechanism in particular (Munns and Tester, 2008; Roy et al., 2014; Pires et al., 2015).

Sodium transporters and plant salt stress tolerance

The study of salt stress tolerance in plants usually focuses on the control of Na⁺ movement, namely on: Na⁺ exclusion from roots, Na⁺ long distance transport, and Na⁺...
compartmentalization at both cellular and tissue level (Munns, 2005; Conde et al., 2011; Roy et al., 2014). These processes are mediated by membrane transporters, reason why the manipulation of their activity has an enormous potential to improve plant performance under high salinity (Brini and Khaled, 2012). Here, we focus on the specific membrane transporters involved in the above outlined tolerance processes. In contrast to animal cells, higher plants do not have Na+-ATPases or K+/Na+-ATPases and, rely on H+-ATPases and H+-pyrophosphatases (PPases) to create the proton-motive force necessary to drive Na⁺ transport across membranes (Conde et al., 2011). The plasma membrane localized SOS1 (Martínez-Atienza et al., 2007; Ji et al., 2013) and the vacuole membrane (tonoplast) localized NHX1 (Jiang et al., 2010; Fukuda et al., 2011) are two Na⁺/H⁺ antiporters involved in Na⁺ exclusion back to the soil and in Na⁺ compartmentalization in the vacuole, respectively. In addition, members of the HKT1 family of HKTs (High Affinity Potassium Transporters) are involved in the control of Na⁺ long distance transport by reabsorption of Na⁺ from the xylem sap into the root cells, preventing the large accumulation of Na⁺ in the above-ground tissues (Rus et al., 2004) (Figure 1). It is noteworthy that the HKT1 Na⁺ exclusion mechanism from the transpiration stream has been frequently indicated as a strong trait in salt tolerance of different cereals, such as rice (Ren et al., 2005) and durum wheat (Triticum turgidum L. subsp. durum) (James et al., 2006).

In the following sections, the role that different Na⁺ transporters and H⁺-pumps play in plant salt stress response is discussed.

**H⁺-Pumps and the plant response to salt stress**

Proton gradients are crucial for the transport of ions and solutes across the different plant cell membranes. Three primary proton transport proteins are found in plant cells: (1) plasma membrane (PM) and (2) vacuolar H⁺-ATPases, which couple ATP hydrolysis with proton transport, and (3) PM and vacuolar H⁺-PPase, which couple pyrophosphate hydrolysis with proton transport (Gaxiola et al., 2007; Fuglsang et al., 2010). The H⁺-Pumps generate an electrochemical potential gradient across membranes,
which is the motive force for a large set of secondary transports.

**Plasma membrane H\(^+\)-ATPase**

The PM H\(^+\)-ATPase belongs to a class known as P-type ATPases (P-ATPases), and is encoded by a large gene family (Gaxiola et al., 2007; Fuglsang et al., 2010). The pump is formed by a single subunit protein, which contains 10 trans-membrane helices and a large cytoplasmatic domain (Fuglsang et al., 2010). Arabidopsis and rice genomes encode 11 and 10 P-ATPases, respectively (Axelsen and Palmgren, 2001; Arango et al., 2003).

The proton motive force created by P-ATPases is largely responsible for a negative potential across the plasma membrane, which is essential for root nutrient uptake, stomatal aperture, phloem loading, and cell growth (Blumwald et al., 2000; Gaxiola et al., 2007; Mansour, 2014). Besides regulation of many physiological processes, the P-ATPases have a critical role in plant adaptation to salt stress conditions. Higher P-ATPases activity under salt stress conditions repolarizes the NaCl-induced depolarization of PM. This response has been strongly associated with salt stress tolerance (Mansour, 2014). The maintenance of the PM potential under salt stress through P-ATPases activity has a great effect on reduction of Na\(^+\) influx via depolarization-activated NSCCs and K\(^+\) efflux via KORs and NSCCs, which help to restore higher K\(^+\)/Na\(^+\) levels (Sun et al., 2009). The higher P-ATPases activity under salt stress also energizes the active transport that exclude Na\(^+\) from root cells, a process dependent of the SOS1 Na\(^+\)/H\(^+\) antiporter (Gaxiola et al., 2007). Furthermore, it was reported that higher activation of P-ATPases is often found in halophytes and salt tolerant genotypes, which may correlate with salt stress tolerance (Mansour, 2014). For instance, in rice callus lines, a higher activation of P-ATPases occurred in salt-tolerant lines as compared to less tolerant ones (Pons et al., 2011).

The salt-dependent activation of PM H\(^+\) pump is associated with increased levels of gene expression as well as post-translational modifications of the enzyme present in a preexisting pool (Gaxiola et al., 2007; Mansour, 2014). However, it is likely that regulation of the pump activity occurs mostly at post-translational level (Gaxiola et al., 2007; Fuglsang et al., 2010). The pump activity can be modulated by phosphorylation/dephosphorylation of the penultimate amino acid residue of the cytoplasmatic C-terminus domain, a threonine residue. The phosphorylated threonine residue promotes binding of the activating 14-3-3 protein (Fuglsang et al., 2010).

Stomatal aperture involves regulation of osmotic pressure within the guard cells, a process powered by P-ATPases activity and responsive to a wide variety of external signals (Gaxiola et al., 2007). Blue light perception in guard cells is mediated by phototropins, which initiate a signal transduction signal pathway that involves an upstream protein phosphatase I and a downstream protein kinase that phosphorylates the penultimate C-terminus amino acid residue of the P-ATPase (Takemiya et al., 2006; Gaxiola et al., 2007). Under drought and salt stress conditions, stomatal closure is induced by ABA through a mechanism that involves production of hydrogen peroxide (H\(_2\)O\(_2\)) and dephosphorylation of the P-ATPases (McAinsh et al., 1996; Zhang et al., 2001; Gaxiola et al., 2007).

**Vacuolar H\(^+\)-ATPase**

Among the three proton-pumps found in plant cells, the vacuolar H\(^+\)-ATPase (V-ATPase) is the most complicated one (Gaxiola et al., 2007). The V-ATPase was first found associated with endomembrane system where it acidifies and generates a proton force motive within diverse cell compartments (e.g., vacuole, endoplasmic reticulum and trans-Golgi network) (Ratajczak, 2000). However, V-ATPases have also been associated with cell plasma membrane (Hanitzsch et al., 2007). The ability of the V-ATPase to maintain the cytosolic pH homeostasis and to acidify the endomembrane compartments is crucial during essential processes, such as cell growth and elongation (Hanitzsch et al., 2007).

Vacuolar H\(^+\)-ATPases are multisubunit enzymes composed of two subcomplexes (V\(_1\) and V\(_0\)): the peripheral V\(_1\) complex consists of eight subunits (A, B, C, D, E, F, G and H) responsible for ATP hydrolyses, and the V\(_0\) membrane-integral complex consists of up to six subunits (a, c, c', c", d and e) responsible for proton translocation (Gaxiola et al., 2007) (Figure 3). In plants, the subunit c' is not found and many of the V-ATPase subunits are encoded by gene families. In Arabidopsis and rice, the 13 subunits which compose the vacuolar H\(^+\)-ATPases (A, B, C, D, E, F, G, H, a, c, c', c", d and e) are encoded by a total of 27 genes and 22 genes respectively (known as VHA genes). If all possible isoform combinations are used, we will have hundreds of different V-ATPase complexes (Sze et al., 2002; Hanitzsch et al., 2007).

By convention, the subunits of V\(_1\) and V\(_0\) complexes are distinguished with capital and case letters, respectively. The V\(_1\) complex consists of: (1) a globular hexameric head with three alternating copies of subunits A and B forming a ring, (2) a central rotational stalk composed of single copies of subunits D and F, and (3) a outer stalk made of subunits C, E, G, and H. Subunits A and B mediate the hydrolysis of ATP at three reaction sites associated with subunit A. Both the central rotational stalk and fixed outer stalk connect the V\(_1\) complex to the membrane inserted V\(_0\) complex. The proton transporting V\(_0\) complex consists of six or more c subunits, also forming a ring structure. In addition, each V\(_0\) complex contains one copy of subunits a, d and e (Beyenbach and Wieczorek, 2006; Hanitzsch et al., 2007) (Figure 3). It has been reported that structural changes of the V-ATPase complex or presence/absence of individual protein isoforms could be correlated with differences in V-ATPase localization and activity between plant organs and/or tissues (Gaxiola et al., 2007).
The plant vacuole plays a very important role in the maintenance of cellular metabolism due to its role in long term storage of toxic ions, long or short term storage of mineral and/or organic acids and in pH and Ca2+ cytoplasmatic homeostasis. Furthermore, the V-ATPase is the most abundant H⁺-pump in the tonoplast and it has been shown that its activity is modulated to cope with environmental and metabolic changes (Ratajczak, 2000). For instance, under salt stress, a general increase of V-ATPase activity has been reported in many plant species (Matsutomo and Chung, 1988; Silva and Gerós, 2009). The V-ATPase provides the driving force necessary for Na⁺ vacuole compartmentalization, a process related on the NHX1 antiporter activity (Jiang et al., 2010; Bassil and Blumwald, 2014).

The ability to respond to high salinity via changes in the expression of the V-ATPase subunits encoding genes might be a prerequisite and a characteristic of salt stress tolerance in plants. It has been reported that the transcript levels of some subunits are up-regulated in response to high salinity (Narasimhan et al., 1991; Kirsch et al., 1996; Silva and Gerós, 2009). However, the expression of V-ATPase genes does not always involve a fixed stoichiometry of mRNAs for the different subunits (Silva and Gerós, 2009). Other factors may also account for the regulation of V-ATPase activity. For instance, the VHA-A subunit from barley (*Hordeum vulgare L.*) was shown to interact to 14-3-3 proteins, well known activators of PM ATPases, in a phosphorylation-dependent way. That interaction was suggested to activate V-ATPase activity (Klychnikov et al., 2007).

### V-ATPase

**Figure 3** - Structural model of the plant V-ATPase adapted from Gaxiola et al. (2007). The peripheral V₁ complex (blue) and the membrane integral V₀ complex (orange) are linked through a peripheral stalk formed by subunits a, C, E, G and H. Hydrolysis of ATP is coupled with H⁺ transport to the vacuole.

**Figure 3**

Plasma membrane and vacuolar H⁺-PPase

H⁺-pyrophosphatases (H⁺-PPase) are highly hydrophobic single subunit proteins that generate proton gradient across the vacuole, Golgi and plasma membrane using the energy of hydrolysis of pyrophosphate (PPI) molecules (Gaxiola et al., 2007). Plants have two phylogenetically distinct types of H⁺-PPases: type I and type II. Type I H⁺-PPases depend on cytosolic K⁺ for their activity and are moderately sensitive to inhibition by Ca²⁺, and type II H⁺-PPases are K⁺ insensitive but extremely Ca²⁺ sensitive.

The Arabidopsis genome encodes two H⁺-PPases: a type I H⁺-PPase (AVP1) and a type II H⁺-PPase (AVP2) (Drozdzowicz et al., 2000). The rice genome also encodes two H⁺-PPases: OVP1 and OVP2 (Sakakibara et al., 1996). However, more isoforms have been proposed (Choura and Rebai, 2005). Phylogenetic analysis of V-PPase sequences showed that rice H⁺-PPases are likely to be type I H⁺-PPases (Drozdzowicz et al., 2000). Type I H⁺-PPases are mainly suggested to acidify the vacuole (Gaxiola et al., 2007). However these H⁺-pumps were also found in the plasma membrane (Ratajczak et al., 1999; Alexandersson et al., 2004). Arabidopsis type II H⁺-PPase, AVP2, has been shown to localize exclusively to Golgi apparatus (Mitsuda et al., 2001).

The expression levels of the H⁺-PPases are strictly regulated at transcriptional level in response to various environmental conditions or developmental stages. It has been shown that the pollen-specific cis-acting region of the AVP1 gene is involved in the regulation of the gene expression during pollen development. AtCAM15, AtCAMTA 1 (calmoduline-binding transcription factors) (Mitsuda et al., 2003), AtVOZ1, and AtVOZ2 (Arabidopsis thaliana Vascular plant One Zinc finger protein) (Mitsuda et al., 2004) were identified as binding to the cis-acting region of the AVP1 gene (Silva and Gerós, 2009; Fuglsang et al., 2010). Salt stress was reported to increase H⁺-PPase activity (Maeshima, 2000). However, a comprehensive mechanism of H⁺-PPase gene expression and post-translational regulation is still needed. It is likely that the protein C-terminus plays an essential role in supporting the physiological function of H⁺-PPase activity (Fuglsang et al., 2010).

Given the importance of the pH homeostasis in the cytosol for cell metabolism, it is likely that the activity of all three H⁺-pumps (P-ATPase, V-ATPase and H⁺-PPase) is regulated by common regulatory mechanisms. 14-3-3 proteins, which are known to regulate many membrane localized proteins, particularly cell ion pumps (Bunney et al., 2002), may be involved in such mechanisms.

**SOS1 and the plant response to salt stress**

Comparisons of unidirectional Na⁺ fluxes and rates of net accumulation of Na⁺ in root indicate that 70-99% of the Na⁺ transported into the root is extruded back to the apoplastic (Munns, 2005; Tester and Davenport, 2003). For rice, that value is indicated as 96% (Munns, 2005), meaning that over time Na⁺ will accumulate in roots and being transferred via the transpiration stream to the shoot, later accumulating there. Since it is important to maintain low cytoplasmatic Na⁺ concentrations for growth and survival under saline conditions, plants have developed a direct...
mechanism to extrude Na\(^+\) from cells across the plasma membrane to the soil or apoplastic. Small differences in Na\(^+\) exclusion capacity create major changes in Na\(^+\) net accumulation (Tester and Davenport, 2003; Munns, 2005; Brini and Khaled, 2012). However, the role of cellular Na\(^+\) efflux is not intuitive in multicellular plants, as Na\(^+\) transport out of one cell would negatively impact the surrounding neighbor cells. So, the role of Na\(^+\) efflux has to be considered in specific tissues and in the context of the whole plant (Zhu, 2003). Sodium efflux is catalyzed by the plasma membrane Na\(^+\)/H\(^+\) antiporter encoded by SOS1 (Salt Overly Sensitive1 = AtNHX7) gene, identified in several plants including Arabidopsis (Wu et al., 1996), rice (Martinez-Atienza et al., 2007), wheat (Xu et al., 2008), and tomato (Xu et al., 2008). SOS1 uses the proton gradient established by P-ATPase and/or plasma membrane H\(^+\)-PPase to exchange Na\(^+\) for H\(^+\) across the membrane (Shi and Zhu, 2002; Qiu et al., 2004; Ji et al., 2013). Activity of the Arabidopsis SOS1 promoter is detected ubiquitously in virtually all tissues, but it appears to be more active in: (1) root epidermal cells (particularly at the root tip), suggesting that meristem requires special protection, since the root tip cells have very small vacuoles and thus are incapable of vacuolar Na\(^+\) compartmentalization, and (2) root parenchyma cells lining the vasculature (Shi and Zhu, 2002; Kronzucker and Britto, 2011). The SOS1 gene expression pattern, together with the results of ion analysis in sos1 mutant plants, suggest that SOS1 has several roles: (1) Na\(^+\) efflux from roots; (2) slowing down Na\(^+\) accumulation in the cytoplasm in order to gain time for Na\(^+\) storage in the cytoplasm (Zhu, 2003; Conde et al., 2011). SOS1 may mediate active loading of Na\(^+\) to the xylem under mild salinity (25 mM NaCl). However, at high salinity (100 mM NaCl), expression of SOS1 is induced and SOS1 may function in Na\(^+\) retrieval from the xylem (Shi et al., 2002). Such a role for SOS1 in long-distance transport is important for the coordination between transpiration Na\(^+\) flow and Na\(^+\) vacuolar sequestration in leaves. However, a thermodynamic analysis by Munns and Tester (2008) indicated that the Na\(^+\) removal from the xylem is unlikely to be mediated by a Na\(^+\)/H\(^+\) antiporter such as SOS1, because its operation “in reverse” under high Na\(^+\) conditions is thermodynamically unfavorable. Instead, class I HKTs have been shown to be involved in xylem unloading of Na\(^+\) (Ren et al., 2005; James et al., 2006; Davenport et al., 2007). Thus, the role of SOS1 in long-distance Na\(^+\) transport remains unclear. Nevertheless, many reports suggest that SOS1 plays a critical role in Na\(^+\) exclusion, thus maintaining cellular ion homeostasis and allowing plants to survive and grow under salt stress conditions (Shi et al., 2003; Cuin et al., 2011) (Table 1).

The transcript level of SOS1 is upregulated by high salinity (Shi et al., 2000). Analysis of the 2 Kb upstream of the SOS1, CIPK24/SOS2 and CBL4/SOS3 transcription initiation sites revealed that the promoter of these genes contains several binding elements for transcription activation of the bZIP, NAC, WRKY, and TCP classes (Ji et al., 2013). However, transcription factors (TFs) mediating promoter activity of SOS genes have not yet been identified. Up-regulation of SOSI transcript levels under high salinity is suggested to be regulated at the post-transcriptional level, as SOS1 promoter activity is not up-regulated by salt stress, but the SOS1 gene expression driven by the constitutive Cauliflower mosaic virus 35S promoter is (Shi et al., 2003). This may indicate that the SOS1 transcript is unstable in the absence of salt stress and that the salt stress causes a post-transcriptional stabilization of the transcript (Shi et al., 2003). More recently, it was suggested that the Na\(^+\) stress induced SOS1 mRNA stability is mediated by ROS (Chung et al., 2008). In addition, regulation of SOSI transcript levels by high salinity is partly under the control of SOS2 and SOS3 (Shi et al., 2000). CIPK24/SOS2 is a protein kinase and CBL4/SOS3 is a calcium sensor that, together with SOS1, are the three key components comprising the Salt Overly Sensitive (SOS) signaling pathway identified in Arabidopsis (Wu et al., 1996) and rice (Martinez-Atienza et al., 2007). At the cellular level, the SOS signaling pathway has been proposed to mediate cellular signaling under salt stress to maintain the ion homeostasis (Ji et al., 2013).

Activation of the Na\(^+\)/H\(^+\) antiporter activity of SOS1 by salt stress is controlled by SOS3 and SOS2 (Zhu, 2003; Ji et al., 2013). In response to an external stimulus, such as high Na\(^+\) concentration, transient increases in cytoplasmatic Ca\(^{2+}\) occur and that is decoded by the calcineurin B and neuronal Ca\(^{2+}\) sensor-like protein SOS3. Activation of SOS3 requires N-myristoylation and Ca\(^{2+}\) bound on EF-hand Ca\(^{2+}\) binding sites. Activated SOS3 physically interacts with the auto-inhibitory domain of SOS2, a member of the SnRK (sucrose non-fermenting-related serine/threonine kinase) family, which activates the kinase and facilitates the localization of the SOS2-SOS3 complex to the plasma membrane. The SOS2-SOS3 complex associates with the Na\(^+\)/H\(^+\) antiporter SOS1, phosphorylating its C-terminal auto-inhibitory domain, which becomes activated and thus pumps Na\(^+\) out of the cell (Pardo, 2010; Brini and Khaled, 2012; Hasegawa, 2013; Ji et al., 2013) (Figure 2).

The SOS pathway is not limited to the three main proteins, as it interacts with other stress related proteins. A SOS3 homolog SOS3-LIKE Calcium Binding Protein8 (SCABP8/CBL10) interacts with SOS2 to form an alternative protein kinase complex that regulates SOS1 activity in the plasma membrane in response to salt stress, mainly in shoots, while SOS3 functions primarily in the root (Quan et al., 2007) (Figure 2). SOS2 phosphorylates CBL10 in a Ca\(^{2+}\) independent manner upon salt stress, and this phosphorylation stabilizes the SOS2-CBL10 complex association with the plasma membrane and increases SOS1 antiporter activity (Kim et al., 2007; Quan et al., 2007; Hasegawa, 2013). Abscisic acid insensitive 2 (ABI2) inter-
| Transporter | Species | Transport selectivity | Tissue localization | Subcellular localization | Function in planta | Refs. |
|-------------|---------|----------------------|---------------------|-------------------------|--------------------|-------|
| AtNHX1      | Arabidopsis | Na⁺/K⁺ | Roots: Vascular tissues Shoots: Floral and vascular tissues, guard cells, trichome | Tonoplast | K⁺ homeostasis and pH regulation | Rodriguez-Rosales et al., 2009; Bassil et al., 2011b; Yokoi et al., 2002 |
| AtNHX2      | Arabidopsis | Na⁺/K⁺ | Roots Shoots: High in guard cells | Tonoplast | K⁺ homeostasis and pH regulation | |
| AtNHX3      | Arabidopsis | Na⁺/K⁺ | Mainly in roots | Tonoplast | N/A | |
| AtNHX4      | Arabidopsis | Na⁺/K⁺ | Shoots: Mainly in mature pollen and seeds | Tonoplast | N/A | |
| AtNHX5      | Arabidopsis | Na⁺/K⁺ | Roots Shoots: High in guard cells | TGN, PVC | pH homeostasis in TGN, PVC | |
| AtNHX6      | Arabidopsis | Na⁺/K⁺ | Roots Shoots: High in guard cells | TGN, PVC | pH homeostasis in TGN, PVC | |
| AtNHX7/SOS1 | Arabidopsis | Na⁺ | Roots: Epidermal cells (particularly root tip), parenhyma cells lining the vasculature Shoots | PM | Na⁺ efflux | Martínez-Atienza et al., 2007; Shi and Zhu, 2002; Kronzucker and Britto, 2011 |
| AtNHX8      | Arabidopsis | N/A | N/A | PM | N/A | N/A |
| OsNHX1      | Rice | Na⁺/K⁺ | Roots: Stela, emerging parts of lateral roots. Shoots: Basal part of seedling shoot, vascular bundle, flag leaf sheaths, panicles, guard cells, trichome | Tonoplast | N/A | Fukada et al., 2004; Fukada et al., 2011; Bassil et al., 2012 |
| OsNHX2      | Rice | Na⁺/K⁺ | Shoots: Flag leaf sheaths, panicles | Tonoplast | N/A | |
| OsNHX3      | Rice | Na⁺/K⁺ | Shoots: Flag leaf sheaths, panicles | Tonoplast | N/A | |
| OsNHX4      | Rice | N/A | N/A | Tonoplast | N/A | |
| OsNHX5      | Rice | Na⁺/K⁺ | Roots: Stela, emerging parts of lateral roots, root tip. Shoots: Basal part of seedling shoot, vascular bundle, flag leaf sheaths, panicles, pollen grain | TGN, PVC | N/A | |
| OsNHX7/OsSOS1 | Rice | Na⁺ | Roots and shoots | PM | Na⁺ efflux | Martínez-Atienza et al., 2007 |
acts with SOS2 to prevent the SOS3 binding to SOS2 and kinase activation. Such ABI2-SOS2 interaction may represent an integrating node between salt stress and ABA signaling (Ohta et al., 2003; Hasegawa, 2013).

The SOS pathway may also regulate the Na⁺ vacuolar compartmentalization. Interaction of SOS2-CBL10 may result in localization of the kinase complex at the vacuolar membrane where it is possibly involved in the regulation of Na⁺/H⁺ exchange at the tonoplast, presumably by regulation of NHX antiporter(s) activity (Qu et al., 2004; Kim et al., 2007). However, no NHX antiporter has already been shown to be directly regulated by SOS2 and/or by the SOS2-complex. In addition, SOS2 has been suggested to regulate the V-ATPase activity. SOS2 was found to interact with the B1 and B2 subunits of the V-ATPase in the absence of CBL proteins, and tonoplast vesicles from the Arabidopsis sos2-2 mutant showed reduced ATPase and H⁺-translocation activities (Batelli et al., 2007).

Potassium homeostasis has also shown to be modulated by the SOS signaling pathway. The protein CBL10 has been indicated to directly interact with AKT1 channel and negatively regulate its activity in roots (Ren et al., 2013). It is well known that plant salt stress tolerance is closely related to maintenance of high K⁺/Na⁺ cytosolic ratio under stress (Tester and Davenport, 2003). The possibility that CBL10 functions as an interconnecting regulator of SOS1 and AKT1 may indicate that CBL10 plays a crucial role in ion homeostasis (K⁺/Na⁺) under salt stress by regulating both K⁺ and Na⁺ uptake/exclusion (Ren et al., 2013).

HKTs and the plant responses to salt stress

Another important determinant of salt stress tolerance in plants is the activity of the HKT (high affinity potassium transporter) proteins (Munns and Tester, 2008; Roy et al., 2014). The HKT family is quite diverse, and this diversity reflects its large number of functions (Munns and Tester, 2008; Almeida et al., 2013; Roy et al., 2014). The HKT family is divided into two distinct classes according to their transport characteristics. The main distinguishing feature is the amino acid sequence that constitutes the first pore domain (PD) (Platten et al., 2006). Members of class I transporters (HKT1) have a serine (S), forming an S-G-G-G motif, where most of the members of class II (HKT2) have a G in the position occupied by the S in class I transporters, forming a G-G-G-G motif (Maser et al., 2002). The presence of either S or G at this position is critical for the cation specificity of transporter. The presence of an S (HKT1) is characterized by a preference for Na⁺ conductance over other cations, whereas the presence of a G (HKT2) is characterized by transport of Na⁺ and/or K⁺ depending on the external concentrations of these two ions (Platten et al., 2006; Kronzucker and Britto, 2011). However, there are notable exceptions, in particular HKT2;1 from cereals, in which the G has reverted to S (Kronzucker and Britto, 2011), but it has been clearly shown to be involved in mediating Na⁺ and K⁺ entry into roots (Munns and Tester, 2008; Kronzucker and Britto, 2011). The main role of HKT1 is believed to be Na⁺ retrieval from the transpiration stream avoiding the over accumulation of Na⁺ in the photosynthetic tissues.

HKT1 family

The best characterized member of HKTs class I is AtHKT1;1 from Arabidopsis. Disruption of AtHKT1;1, the only member of HKT family in Arabidopsis, caused a higher accumulation of Na⁺ in the shoots but reduced concentration in roots, with little effect on the net Na⁺ uptake (Rus et al., 2004; Pardo, 2010; Kronzucker and Britto, 2011) AtHKT1;1 is preferentially expressed in the plasma membrane of xylem parenchyma cells and phloem cells of both roots and leaves, where it is suggested to regulate the Na⁺ distribution between roots and shoots (Sunarpi et al., 2005; Moller et al., 2009; Pardo, 2010; Kronzucker and Britto, 2011) (Figure 1 and Table 2). Two complementary functions for AtHKT1;1 have been proposed. In the phloem recirculation model, AtHKT1;1 loads Na⁺ into shoot phloem cells to be transferred to roots via the downward stream, preventing Na⁺ overaccumulation in the shoot. However, the overall Na⁺ retranslocation potential via phloem should not exceed 10% of the total Na⁺ loaded in the shoot xylem transpiration stream (Berthomieu et al., 2003). Another function of AtHKT1;1 is to unload Na⁺ from the xylem transpiration stream, thereby restricting the amount of Na⁺ reaching the photosynthetic tissues and supporting salt stress tolerance.

Analysis of several QTLs of salt tolerance in rice (Ren et al., 2005; Kronzucker and Britto, 2011) and wheat (James et al., 2006; Byrt et al., 2007; Kronzucker and Britto, 2011) has provided further evidence for the importance of HKT class I genes in controlling Na⁺ accumulation in leaves upon salt stress. In rice, QTL analyses showed that higher shoot K⁺ content of the salt-tolerance indica genotype, Nona Bokra, cosegregated with an allelic variant of SKCI (Shoot K⁺ Content 1) with higher activity as compared to that of the salt-sensitive japonica genotype, Koshihikari (Ren et al., 2005). SCK1, now referred to as OsHKT1;5 (OsHKT8) is a plasma membrane, K⁺ independent, and Na⁺ selective transporter that is preferentially expressed in the parenchyma cells surrounding xylem vessels (Ren et al., 2005; Pardo, 2010; Almeida et al., 2013) (Table 2). The Nona Bokra OsHKT1;5 has four amino acids different from the Koshihikari protein, and this difference has been associated with greater Na⁺ transporter activity and increased ability for maintenance of K⁺/Na⁺ homeostasis under salt stress (Ren et al., 2005). Rice contains four more HKT1 members in the genome, OsHKT1;1, OsHKT1;2, OsHKT1;3, OsHKT1;4 (Ren et al., 2005; Huang et al., 2006; Wu et al., 2009; Cotsaftis et al., 2012; Almeida et al., 2013) (See Table 2 for further information). OsHKT1;4 gene expression is up-regulated in the leaf sheaths under salt stress (Cotsaftis et al., 2012) and encodes three different splicing forms, identified in both rice genotypes Pokkali
(salt-tolerant) and Nipponbare (salt-susceptible). All OsHKT1;4 splicing forms are translated into protein, nevertheless only the longer splicing form seems to be translated into a functional protein (Cotsaftis et al., 2012). Interestingly, Pokkali is able to maintain a much higher ratio of functional OsHKT1;4 transcripts in younger leaf sheaths as compared to Nipponbare. In addition, transcript levels of the functional transcripts were inversely correlated with the individual leaf blade Na⁺ concentration in both genotypes (Cotsaftis et al., 2012). At this point it seems that the longer OsHKT1;4 splicing form is the key transporter controlling the sheath-to-blade transfer of Na⁺ in rice shoots (Cotsaftis et al., 2012).

In wheat, QTL analyses using durum wheat (Triticum turgidum L. subsp. durum) breeding Line 149 led to the identification of two loci, Nax1, and Nax2, which decreased Na⁺ accumulation in the leaf blade (James et al., 2006; Byrt et al., 2007; Kronzucker and Britto, 2011). In addition, bread wheat (Triticum aestivum), which is an allohexaploid (2n = 6s = 42, genome AABBDD), was found to be more salt tolerant than the allotetraploid pasta wheat (AABB genomes). It was shown that the D genome carries a locus (Kna1) responsible for maintenance of high K⁺/Na⁺ ratio during salt stress justifying the salt tolerance of bread wheat (Dubcovsky et al., 1996; Byrt et al., 2007; Kronzucker and Britto, 2011). The process controlled by the Nax2 and Kna1 loci reduces net root xylem loading of Na⁺, while the Nax1 locus reduces Na⁺ accumulation in the leaf blade by restricting Na⁺ loading into root xylem and partitioning Na⁺ into the leaf sheath (James et al., 2006; Hasegawa, 2013).

Table 2 - List of HKT transporters including information about class, species, transport selectivity, tissue localization, subcellular localization and plant function for each HKT transporter described in this review. No information available (N/A), Plasma membrane (PM).

| Transporter | Species | Transport selectivity | Tissue localization | Subcellular localization | Function in planta | Refs. |
|-------------|---------|-----------------------|--------------------|-------------------------|--------------------|-------|
| AtHTK1;1    | Arabidopsis | Na⁺      | Roots: Xylem parenchyma, phloem Shoots: Phloem | PM                    | Unload Na⁺ from the xylem transpiration stream | Berthomieu et al., 2003; Møller et al., 2009; Sunarpi et al., 2005 |
| OsHTK1;1    | Rice     | Na⁺      | Roots: Similar as OsHTK2;1 Leaves: Bulliform cells and vascular tissues | PM                    | N/A                 | Jabnoune et al., 2009 |
| OsHTK1;2    | Rice     | N/A      | N/A | PM                    | N/A                 | Wu et al., 2009 |
| OsHTK1;3    | Rice     | Na⁺      | Roots: Cortex and vascular tissues in the stele. Leaves: Bulliform cells and vascular tissues, mesophyll cells. | PM                    | N/A                 | Jabnoune et al., 2009 |
| OsHTK1;4    | Rice     | N/A      | Leaves sheaths | PM | Control of sheath to blade transfer of Na⁺ | Cotsaftis et al., 2012 |
| OsHTK1;5    | Rice     | Na⁺      | Roots and Shoots: Xylem parenchyma | PM | Unload Na⁺ from the xylem transpiration stream | Ren et al., 2005 |
| TaHKT1;4    | Wheat    | Na⁺      | Roots and Leaves | PM | Unload Na⁺ from the xylem transpiration stream | Huang et al., 2006 |
| TaHKT1;5    | Wheat    | Na⁺      | Roots | PM | Unload Na⁺ from the xylem transpiration stream | Byrt et al., 2007; Munns et al., 2012 |
| OsHKT2;1    | Rice     | Na⁺/K⁺   | Roots: Epidermis, exodermis, cortex differentiated into aerenchyma, stele (mainly phloem). Leaves: Bulliform cells, xylem, phloem, mesophyll cells. | PM | Nutritional Na⁺ uptake from the external medium. | Jabnoune et al., 2009; Horie et al., 2007 |
| OsHKT2;2    | Rice     | Na⁺/K⁺   | Roots | PM | Na⁺ and K⁺ uptake under K⁺ starvation conditions. | Yao et al., 2010; Oomen et al., 2012 |
| TaHKT2;1    | Wheat    | Na⁺/K⁺   | Roots: Cortical and stele Leaves: Vascular tissue of mesophyll. | PM | Na⁺ uptake from external medium. | Schachtman and Schroeder, 1994 |
| HvHKT2;1    | Barley   | Na⁺/K⁺   | Roots: Cortex. Leaves: Blade and sheath. | PM | K⁺ uptake at very low K⁺ concentrations (possible) and uptake of Na⁺ in the roots (possible). | Mian et al., 2011; Haro et al., 2005 |
Using high-resolution mapping, Nax1 and Nax2 were identified as members of the HKT1;4 gene family and Kna1 as member of the HKT1;5 gene family (Table 2). Because both Nax genes are originated from a wheat relative, Triticum monococcum, that was crossed with a durum wheat, they were named TmHKT1;4-A2 and TmHKT1;5-A, respectively. The Nax2 region of the breeding Line 149 was found to correspond to the Kna1 region of the breed of wheat and Kna1 was named TaHKT1;5-D (Almeida et al., 2013). Nax1 and Nax2 genes do not exist in modern bread or durum wheat genotypes, and introgression of Nax1 or Nax2 into bread wheat led to reduced leaf blade Na⁺ accumulation and increased leaf blade Na⁺ exclusion relative to the parent respectively. The combination of Nax1 and Nax2 further decreased Na⁺ accumulation in the leaf blade (James et al., 2011), showing that these genes clearly have similar functions as AtHKT1;1 in Arabidopsis and OsHKT1;5 and OsHKT1;4 in rice (Ren et al., 2005; James et al., 2006; Almeida et al., 2013) (Table 2). Moreover, field trials with durum wheat, carrying the Nax2 gene, growing under high saline soils showed a 25% increase in grain yield and reduced Na⁺ accumulation in flag leaf as compared to a near isogenic line without the Nax2 locus (Munns et al., 2012). Altogether, these results indicate that HKT1 mediated Na⁺ exclusion from shoot is an effective mechanism for enhancing salt stress tolerance in crop plants.

Regarding HKT1 transcriptional regulation, some transcriptional regulatory elements have been identified in the AtHKT1;1 promoter. The tandem repeat regions (R1 and R2) found in the distal AtHKT1 promoter region located about 3.9 kb upstream of the translational start codon were responsible for expression of AtHKT1 in roots (Rus et al., 2004; Baek et al., 2011). The repeat sequence R2 which is closest to ATG acts as an enhancer element of AtHKT1 expression. Its inactivation caused reduced AtHKT1 expression in root and higher Na⁺ accumulation in shoot (Rus et al., 2004; Baek et al., 2011). The AtHKT1 promoter contains a highly methylated GC region (250 bp) at 2.6 kb upstream of the translational start codon. Interestingly, methylation in the leaf is higher than in roots, which suggests that higher methylation in this promoter region is required to maintain AtHKT1 expression at low levels and perhaps in a correct pattern of expression in the different tissues (Baek et al., 2011). Furthermore, this region contains a putative small RNA target site, which was suggested to be involved in methylation guided by small RNAs (Baek et al., 2011).

HKT2 family

HKT2 class 2 proteins are generally found in monocotyledonous species, and no HKT class 2 homologs have been identified in dicotyledonous species (Platten et al., 2006; Adams and Shin, 2014). Four HKT class 2 members have been characterized in detail: OsHKT2;1 and OsHKT2;2 in rice, TaHKT2;1 in wheat, and HvHKT2;1 in barley (Hordeum vulgare L.) (Table 2). These transporters have common properties thought to be shared by all HKT2 class 2 members, such as a role in Na⁺ uptake from external medium under K⁺ limiting conditions (Almeida et al., 2013).

The two characterized rice members of this HKT family, OsHKT2;1 and OsHKT2;2, have been reported to mediate Na⁺ uptake from soil under K⁺ limiting conditions (Table 2). OsHKT2;1 gene expression is induced by K⁺ deficiency (Horie et al., 2001; Yao et al., 2010). OsHKT2;1 is an atypical HKT class 2 member, which has an S residue in the first PD and mediates high-affinity Na⁺ uptake. However, OsHKT2;1 can also mediate K⁺ transport depending on the external concentration of both K⁺ and Na⁺ (Jabnoune et al., 2009; Yao et al., 2010; Almeida et al., 2013). OsHKT2;1 is known to be highly involved in “nutritional” absorption of Na⁺ and its relevance in Na⁺ uptake during salt stress may be limited since it has a micromolar affinity for Na⁺ and its activity is rapidly downregulated at high Na⁺ concentration. Interestingly, RNA levels of at least three other OsHKTs genes have been shown to be inhibited by an external Na⁺ concentration as low as 30 mM (Horie et al., 2001). On the other hand, OsHKT2;2 has only been found in the salt-tolerant Nona Bokra and Pokkali genotypes, being absent in the rice salt-sensitive Nipponbare genotype, which suggests that the presence of OsHKT2;2 is an evolutionary advantage for salt-tolerant genotypes (Horie et al., 2001; Almeida et al., 2013). OsHKT2;2 is expressed in roots among other tissues and transporting both K⁺ and Na⁺, but under salinity only Na⁺ is transported (Kader et al., 2006; Oomen et al., 2012) (Table 2). Other OsHKT class 2 members have also been identified (OsHKT2;2/1, OsHKT2;3, OsHKT2;4), however these will be not described. For further information, see Almeida et al. (2013) and Kronzucker and Britto (2011).

In wheat, TaHKT2;1 seems to have a function in root Na⁺ influx similar to rice OsHKT2;1 (Horie et al., 2009). TaHKT2;1 is expressed in the root cortex and is induced by K⁺ deficiency (Schachtman and Schroeder, 1994). In planta, TaHKT2;1 has been suggested to have a role in Na⁺ transport with a possible role in root Na⁺ uptake, though TaHKT2;1 was also reported to transport K⁺ (Almeida et al., 2013) (Table 2).

In barley, a relative salt-tolerant species, HvHKT2;1 is preferentially expressed in root cortex and to a much lower level in leaf blade and sheaths, and it is induced by K⁺ deficiency in roots and shoots and by high Na⁺ concentration in shoots. HvHKT2;1 mediates both K⁺ and Na⁺ transport (Haro et al., 2005; Mian et al., 2011; Almeida et al., 2013) (Table 2). Transgenic barley lines over-expressing HvHKT2;1 result in higher Na⁺ concentration in xylem, enhanced translocation of Na⁺ to shoots and Na⁺ accumulation in the leaves higher than in the non-transformed plants, supporting the hypothesis that this transporter is able to mediate root Na⁺ uptake (Mian et al., 2011). Moreover, transgenic plants showed a significant increase in shoot K⁺
content in plants growing in limiting K\(^+\) conditions, suggesting that \(HvHKT2;1\) may also play a role in K\(^+\) absorption or re-absorption at very low K\(^+\) concentrations (Mian et al., 2011).

**NHX and the plant response to salt stress**

At the cellular level, high amounts of Na\(^+\) can be tolerated by intracellular partitioning so that the concentration in the cytoplasm is kept as low as 10-30 mM (Munns and Tester, 2008). This strategy can be used by plants for the alleviation of excessive cytosolic Na\(^+\) by sequestrating Na\(^+\) into the vacuole, which typically makes up to 80-90\% of the cell volume. Other organelles, such as endosomal compartments, plastids and mitochondria, may also accumulate Na\(^+\) and thus contribute to the overall subcellular Na\(^+\) sequestration (Zhu, 2003). The vacuolar sequestration of Na\(^+\) that occurs in all tissues is not only important for Na\(^+\) detoxification in the cytosol, but it is also a critical mechanism of osmotic adjustment to maintain water uptake from saline solutions (Zhu, 2003; Munns and Tester, 2008; Bassil et al., 2012).

An increased vacuolar Na\(^+\) concentration requires a coordinated increase in the osmotic pressure of the other subcellular components, including the cytosol, to maintain the osmotic pressure and thereby the volume. This can be achieved by an increase in the K\(^+\) concentration to a sub-toxic level, as well as by the synthesis and accumulation of compatible solutes (e.g., proline, sucrose, glycine betaine, etc.). Nevertheless, the latter represents a major drawback due to the high energetic cost associated with solute synthesis (Munns and Tester, 2008; Maathuis et al., 2014).

The tonoplast controls the movement of inorganic and organic solutes to and from the cytoplasm through a wide range of pumps, carriers and ion channels (Conde et al., 2011). Cation/H\(^+\) antiporters mediate the transport of Na\(^+\) into the vacuole, driven by the electrochemical gradient of protons generated by the V-ATPase and V-PPase enzymes (Jiang et al., 2010; Bassil and Blumwald, 2014). This Na\(^+\)/H\(^+\) exchange is mediated by members of a family of transporters referred to as Na\(^+\)/H\(^+\) antiporters (NHXs) in plants or Na\(^+\)/H\(^+\) exchange (NHEs) in animals (Jiang et al., 2010; Bassil and Blumwald, 2014). In addition, plant NHX antiporters mediate both Na\(^+\)/H\(^+\) and K\(^+\)/H\(^+\) exchange, therefore affecting both salinity tolerance and K\(^+\) nutrition (Venema et al., 2002; Leidi et al., 2010).

**Diversity of plant NHX antiporters**

Plant NHX proteins belong to a large superfamily of monovalent cation/proton antiporters (CPAs) made up of two subgroups, CPA1 and CPA2. The CPA2 family includes members of the less known Cation/H\(^+\) Exchangers (CHXs) and K\(^+\) efflux antiporters (KEA). The CPA1 family includes members of the NHX-type, which are ubiquitous in all eukaryotic organisms (Rodriguez-Rosales et al., 2009; Bassil et al., 2012). In Arabidopsis, NHX-type antiporter family members comprise eight members that are divided into two distinct classes; two divergent members located at the plasma membrane (SOS1/AtNHX7 and AtNHX8), and six intracellular members located either at the tonoplast (AtNHX1-AtNHX4) or the endosomal membrane (Golgi, trans-Golgi network and prevacuolar compartments) (Rodriguez-Rosales et al., 2009; Bassil et al., 2012; Reguera et al., 2015). In rice, six NHX-type antiporter family members were identified as belonging to two distinct classes with different cellular localizations: one in the plasma membrane (SOS1) (Martinez-Atienza et al., 2007), and five intracellular members that are either in the tonoplast, OsNHX1 to OsNHX4, or in the prevacuolar compartment OsNHX5 (Fukuda et al., 2011) (Figures 1 and 2, Table 1). In Arabidopsis, the most abundant members of NHX-types are AtNHX1 and AtNHX2, accounting for a significant amount of the K\(^+\)/Na\(^+\)/H\(^+\) antiport activity in tonoplast vesicles (Barragan et al., 2012). Detailed information regarding AtNHXs and OsNHXs tissue localization is described in Table 1.

**NHX gene expression under stress conditions**

In Arabidopsis seedlings, \(AtNHX1\) and 2 were shown to be induced by salt stress (NaCl), hyperosmotic stress (mannitol) and ABA treatment, whilst \(AtNHX5\) was only induced by salt stress (NaCl) (Yokoi et al., 2002). In rice seedlings, salt stress (NaCl), hyperosmotic stress (mannitol) and ABA treatment increased the transcript levels of \(OsNHX1\), 2, 3 and 5 (Fukuda et al., 2011). These reports show that NHX genes are components of the plant salt stress response. Interestingly, treatment with a high KCl concentration induced the expression of \(OsNHX1\) and 2 (Fukuda et al., 2011), and \(AtNHX1\) (Yokoi et al., 2002). \(AtNHX1\) and 2 were induced by ABA but not by NaCl in the ABA-deficient \(aba2-1\) mutant, showing that NaCl induction of these members depends on ABA signaling (Yokoi et al., 2002). \(AtNHX1\) and 2 promoter sequences do not have ABA-responsive elements (ABRE). Nevertheless, the promoter of each gene contains MYC/MYB cis-regulatory elements, suggesting that \(AtNHX1\) and 2 are outputs of the ABA-dependent pathway regulated by these transcription factors (Yokoi et al., 2002). On the other hand, the \(OsNHX1\) promoter (up to 1.8 kb upstream of the translational start codon) shows several ABA-responsive elements (ABRE), as well as drought responsive elements MYC/MYB cis-regulatory elements (Almeida et al., unpublished results), indicating that \(OsNHX1\), similar to \(AtNHX1\), is also transcriptionally regulated by an ABA-dependent pathway. Interestingly, the SOS pathway also appears to regulate the activity of vacuolar Na\(^+\)/H\(^+\) antiporters (Yokoi et al., 2002; Qiu et al., 2004). The activity of AtNHX1 is possibly regulated through interaction with the protein kinase \(SOS2\) (Qiu et al., 2004) (Figure 2). It was reported that the vacuolar Na\(^+\)/H\(^+\) antiporter activity of Arabidopsis membrane vesicles was significantly reduced in vesicles obtained from \(sos2\) null mutants, as compared to
wild type controls, and could be stimulated *in vitro* by the addition of activated SOS2 protein (Qiu et al., 2004). The activity was further inhibited by AtNHX1 antibodies. However, phosphorylation of AtNHX1 by SOS2 was not shown. It was further shown that SOS2 also interacts with several V-ATPase subunits (Figure 2), and that vesicles isolated from sos2 null mutants show a considerable lower V-ATPase acidification (Batelli et al., 2007). Thus, comparisons of antiporter activity in vesicles from sos2 mutant and wild type is complex, as the proton force motive that drives ion transport is not similar in both cases (Batelli et al., 2007).

### NHX regulation and structural organization

**Transcriptional regulation**

Although multiple functional studies on NHX-type proteins, especially NHX1, have been carried out, the details of how NHX1 is transcriptionally regulated remain poorly explored. Adler et al. (2010) reported that NHX1 from the relatively salt-tolerant crop, sugar beet (*Beta vulgaris* L.) is regulated under salt stress by one or more MYB transcription factors, which could not be identified yet. Despite the importance of rice, only one study reported the identification of a TF interacting with the OsNHX1 promoter. Using a chromatin immunoprecipitation assay, an OsbZIP71 TF was identified by Liu et al. (2014) as directly binding to the OsNHX1 promoter. It was shown that OsbZIP71 gene expression was strongly induced by drought, polyethylene glycol (PEG), and ABA treatments, but repressed by salt treatment. Transgenic rice lines overexpressing OsbZIP71 (p35S::OsbZIP71) showed improved tolerance to drought, salt and PEG-induced drought stresses, suggesting that OsbZIP71 plays an important role in ABA-mediated drought and salt tolerance in rice (Liu et al., 2014). However, the authors did not show whether the identified TF is relevant for OsNHX1 activation under stress.

**Post-translational modifications**

The cation selectivity of Arabidopsis NHX1 appears to be regulated by its C-terminal tail through the interaction with the vacuolar lumen-localized calmodulin-like protein 15 (AtCaM15), in a Ca\(^{2+}\) and pH-dependent manner (Yamaguchi et al., 2003). Under control physiological conditions, when the vacuole pH is acidic (pH 5.5) and the Ca\(^{2+}\) concentration is high, AtCaM15 is bound to the AtNHX1 C-terminal tail, resulting in a higher K\(^{+}/H^+\) exchange activity over Na\(^+\)/H\(^+\) activity (Yamaguchi et al., 2003). On the other hand, salt stress often causes alkalization of the vacuole, which reduces AtCaM15 binding to AtNHX1. This leads to an increased Na\(^+\)/H\(^+\) exchange activity over K\(^{+}/H^+\) activity and subsequent enhanced vacuolar Na\(^+\) sequestration (Yamaguchi et al., 2003; Rodriguez-Rosales et al., 2009). In addition, phosphoproteomic studies in Arabidopsis and rice suggested that NHX antiporters are regulated by phosphorylation (Whiteman et al., 2008a,b). In rice, the vacuolar OsNHX3 was reported to be phosphorylated at residue S471 located in the C-terminus. The same residue is conserved among the three other rice vacuolar NHXs members (OsNHX1, 2 and 4) (Bassil et al., 2012). Sequence comparison analyses between rice NHXs and Arabidopsis NHX3, 5 and 6 revealed that the Arabidopsis NHXs members contained the same S residue at a similar position (Bassil et al., 2012). So far, the biological role of such post-translational modifications has not yet been functionally characterized. Negrão et al. (2013) identified a significant nonsynonymous mutation at OsNHX1, serine 477 to asparagine (S477N), present in the rice salt-susceptible genotypes IR 29 and IR 64, but also in the salt-tolerant genotype FL 478 (a recombinant inbred line derived from an IR 29 x Pokkali cross). The loss of an S residue can imply the loss of a putative phosphorylation site, and S477 sits in a cluster of S residues with high phosphorylation probability and is itself a potential phosphorylation target (Negrão et al., 2013). This residue is located in the C-terminus of the OsNHX1 protein, and it was suggested by the authors that the nonsynonymous mutation may affect the phosphorylation of the OsNHX1 C-terminal possibly by SOS2, which in turn results in lower activation of OsNHX1 exchanger activity. However, further studies are needed to test this hypothesis.

**Topology**

The crystallographic structure of NHXs antiporters is not yet available. Epitope tagging and protease protection assays applied to full length expressed AtNHX1 in a yeast heterologous system unveiled that AtNHX1 has nine transmembrane domains, with an additional three “buried” domains that do not entirely span the membrane (Yamaguchi et al., 2003). This study also showed that the hydrophilic C-terminus is oriented to the vacuolar lumen, a feature that is strikingly different from the proposed cytosolic C-terminus orientation of animal NHEs (Yamaguchi et al., 2003; Bassil et al., 2012). However, another study performed by Sato and Sakaguchi (2005), using only protein fragments, showed that AtNHX1 contains eleven transmembrane domains and a cytosolic C-terminus, resembling an overall membrane topology of the human NHE. The C-terminus of NHX members is highly divergent, even among closely related members within the same species. Because the C-terminus regulates the antiporter activity, it has been suggested that divergent C-terminal sequences may constitute a novel way to differentially regulate individual members (Bassil et al., 2012).

**Function of the NHX antiporters**

**Salt tolerance**

Na\(^+\)/H\(^+\) exchange at the tonoplast is generally accepted to play a major role in plant salt stress tolerance. Several reports indicate that NHX overexpression (in ho-
mologous or heterologous systems) confer salt stress tolerance in a wide range of plant species. Constitutive overexpression of AtNHX1 appears to increase salt stress tolerance significantly in yeast (Aharon et al., 2003). Arabidopsis (Apse et al., 1999), tomato (Zhang and Blumwald, 2001) and cotton (He et al., 2005). Constitutive overexpression of various cereal NHX homologs has been also reported to improve the salt stress tolerance of Arabidopsis (Brini et al., 2007), rice (Fukuda et al., 2004; Zhao et al., 2006) and wheat (Xue et al., 2004). These results show the fundamental role of these proteins in plant salt stress tolerance and explain why they have been a major focus for genetic engineering (Jiang et al., 2010; Bassil et al., 2012). However, increased salt stress tolerance was not always associated with an increased vacuolar Na+ accumulation (Fukuda et al., 2004; Rodriguez-Rosas et al., 2008, 2009; Jiang et al., 2010). Most of the characterized NHX members can transport both K+ and Na+, and may have similar Km for these substrates (Jiang et al., 2010). This means that, unless the cytoplasmatic Na+ concentration is significantly higher than that of K+(difficult to occur even under salt stress conditions), NHX exchangers mainly mediate K+ exchange rather than Na+/H+ exchange (Jiang et al., 2010; Maathuis et al., 2014). Indeed, nhx1/nhx2 double null mutants in Arabidopsis resulted in impaired vacuolar K+ accumulation, enhanced vacuolar Na+ uptake, and a salt (NaCl) insensitive phenotype, compared to wild-type (Barragan et al., 2012). Thus, it is likely that the contribution of NHX-like protein to plant salt stress tolerance is the maintenance of K+ homeostasis rather than sequestration of Na+ into the vacuole (Maathuis et al., 2014). Nevertheless, it is still not clear what are the primary ions being transported by NHX-like proteins in planta. Given that NHX1 cation selectivity is regulated by interacting partners (AtNHX1 is regulated by CaM15), it is difficult to interpret the ion content results from plants overexpressing AtNHX1, and possibly other NHXs members, due to a possible shortage of interacting partners (Rodriguez-Rosas et al., 2009).

K+ homeostasis

Besides their key role in salt stress tolerance, at control growth conditions vacuolar NHX proteins have a key role in mediating K+/H+ exchange for turgor regulation and pH control. Potassium is an essential plant nutrient and the most abundant cation in plants, comprising up to 10% of plant dry matter. K+ is an important cofactor in many biosynthetic processes, and in the vacuole it plays key roles in cell volume regulation (Barragan et al., 2012; Andrés et al., 2014).

During grape berry (Vitis vinifera L.) development, high VvNHX1 transcript levels during the véraison and post-véraison stages would indicate that the increase in vacuolar K+ accumulation, mediated by VvNHX1 is needed for vacuolar expansion. This process is coupled with a rapid accumulation of sugars that drives water uptake to the berry and the concomitant berry size increase, typical of the post-véraison growth stage (Hanana et al., 2007).

Genetic studies in Arabidopsis firmly demonstrate the importance of NHXs in the regulation of K+ and pH homeostasis (Rodriguez-Rosas et al., 2008; Leidi et al., 2010; Bassil et al., 2011a; Barragan et al., 2012). The Arabidopsis AtNHX1 single knockout mutant displayed an altered phenotype under control growth conditions, including smaller cells, smaller leaves, and other developmental irregularities associated with altered K+ homeostasis, which was correlated with lower K+/H+ and Na+/H+ antiport activity (Apse et al., 2003; Sottosanto et al., 2004). AtNHX2 knockout did not display any obvious growth phenotype, but mutants lacking both AtNHX1 and 2 displayed a significant reduction in cell expansion in all tissues, especially in rapidly elongating organs such as flowers filaments and hypocotyls of etiolated seedlings, as compared to Atnhx1 mutant or wild-type plants (Bassil et al., 2011a). These plants displayed poor seed set because their filaments did not elongate enough to position the anther close to the stigma. Though these plants had non-dehiscent anthers, flowers could be artificially pollinated (Bassil et al., 2011a). In root and leaf cells of the double mutant, the vacuolar K+ content was about one-third of that from wild-type cells. The double mutant was also highly sensitive to the addition of external K+ (nhx1nhx2 mutant has higher K+ cytosolic content), which may indicate that these vacuolar NHX antiporters are the main mediators of cytosolic K+ uptake into the vacuole; it also suggests that variations of K+ supply, which would otherwise result in a fluctuation of cytosolic K+ content, is essentially buffered by vacuolar K+/H+ exchange, likely promoted by the activity of vacuolar NHX proteins (Bassil et al., 2011a; Bassil and Blumwald, 2014). Impaired osmoregulation in the nhx1nhx2 mutant leads to lower leaf water content, lower cell turgor and consequent defective stomatal movement. Altogether, it results in a poor plant water status maintenance (Barragan et al., 2012). Stomatal movements rely on guard cell turgor and require massive bidirectional K+ fluxes across the guard cells plasma and tonoplast membranes. The double mutant displayed markedly reduced tonoplast vesicles, K+/H+ activity, and disruption in K+ accumulation in guard cells, which in turn may affect the guard cells osmoregulation capacity and stomatal movement (Andrés et al., 2014). In addition, the nhx1nhx2 mutant exhibited more acidic vacuoles and the disappearance of the highly dynamic remodeling of vacuolar structure associated with stomatal movements (Andrés et al., 2014). Altogether, these data suggest that NHX1 and NHX2 are the main transporters mediating K+ uptake to the vacuole.

pH homeostasis

Cellular pH homeostasis is one of the most important factors for cellular function. In plants cells, cytoplasmatic pH is regulated by the primary action of H+-pumps and metabolic process producing H+ or OH-. Cation/H+
antiporters constitute proton leak pathways allowing rapid cytoplasmatic pH adjustments (Rodriguez-Rosales et al., 2009). Involvement of plant NHX antiporters in vacuolar pH regulation is best illustrated by studies of Japanese morning glory (*Ipomoea nil* or *Pharbitis nil*) flower petal coloration. During flower development, *Ipomea* petals begin to accumulate anthocyanins in vacuole, which are red at low pH but turn blue as pH increases. During the color transition, petal vacuolar pH shifts from ca. 6.5 to 7.5 and is accompanied by increased V-ATPase, V-PPase and InNHX1 expression and activity (Yamaguchi et al., 2001; Yoshida et al., 2009). The purple (*pr*) mutation of *Ipomea nil*, which abolishes the activity of *InNHX1*, partially hampers vacuole alkalinization and prevents the full color shift from red to blue in opening flowers (Yamaguchi et al., 2001). The partial pH change and color in *pr* mutant has been suggested to rely on the activity of another abundantly expressed vacuolar NHX-type member, *InNHX2* (Ohnishi et al., 2005).

**Vesicular trafficking**

Eukaryotic cells synthesize, modify, and deliver molecular cargo in and between distinct cellular components through a complex and coordinated system of intracellular trafficking of cargo via vesicles. Vesicular trafficking depends on numerous molecular players and biochemical and biophysical factors. Among the principal factors, the vesicular luminal pH must be maintained within a narrow range that is unique to each specific intracellular compartment of the endomembrane system (Bassil et al., 2012). In vivo pH measurements of intracellular compartments along the secretory pathway revealed a general gradual pH acidification with maturity, ranging from pH ~ 7.1 in the endoplasmatic reticulum to pH ~ 5.5 in the vacuole (Martinieri et al., 2013). pH is critical not only for the compartmentalization of specific biochemical reactions but also for maintaining vesicular identity (through receptor association), sorting of newly synthesized or modified cargo, endocytosis, coat protein formation, energizing secondary transport system, as well as the degradation of molecules. The establishment of vesicular acidification is achieved by the action of V-ATPase and V-PPase. If vesicular pH was solely regulated by V-ATPase, the vesicular luminal pH could reach a pH below 3. Therefore, vesicular pH homeostasis is regulated by the active H⁺ transport mediated by H⁺-pumps and by luminal H⁺ leaks, thus establishing the optimal endosomal pH (Bassil et al., 2012). In yeast, the function of *S. cerevisiae NHX1* seems to be related to its involvement in protein sorting through endosomal pH regulation (Brett et al., 2005; Rodriguez-Rosales et al., 2009). Disruption of *ScNHX1* blocked the trafficking out from the Golgi/Pre-vacuolar compartment (Bowers et al., 2000). The mutant also had an acidic cytoplasm and vacuole, as well as growth sensitivity to acidic media. In addition, protein processing and mis-sorting also occurred, because ~35% of the newly synthesized soluble vacuolar protein carboxypeptidase Y (CPY) was secreted to the apoplast (Bowers et al., 2000; Bassil et al., 2012).

In plants, the most direct evidence demonstrating a requirement for endosomal NHX antiporters in vesicular trafficking was generated using null mutants lacking both endosomal *AtNHX5* and *6* (Bassil et al., 2011b; Reguera et al., 2015). *AtNHX5* and 6 reside in the Golgi, *trans*-Golgi network, and prevacuolar compartments (Reguera et al., 2015). Plants lacking both *AtNHX5* and 6 displayed severely reduced growth (mainly due to reduced cell expansion), with smaller and fewer cells, and increased sensitivity to salt stress. In addition, trafficking of CPY was mis-sorted to the apoplast in the *nhx5nhx6* mutant, in a similar phenotype reported for *ScNHX1* disruption (Bowers et al., 2000; Bassil et al., 2011b). In a more recent report Reguera et al. (2015) fully characterized the *nhx5nhx6* mutant. The work showed that *AtNHX5* and *AtNHX6* are crucial for the maintenance of endomembrane luminal pH and supports the concept that proper vacuolar trafficking requires endomembrane pH homeostasis. In addition, transcriptional profile analyses of the Arabidopsis *nhx1* mutant revealed changes in the expression of a significant number of genes encoding proteins associated with intravesicular trafficking, trafficking to the nucleus, and Golgi processing (Sottosanto et al., 2004). This indicates that, similar to what was reported for the yeast ortholog *Nhx1p* (Ali et al., 2004), *AtNHX1* also plays an important role in protein trafficking and targeting, probably via regulation of the intravesicular pH (Sottosanto et al., 2004). Hamaji et al. (2009) reported that NHX1 is present in vesicles in the cytoplasm of salt-treated cells, suggesting that at least under salt stress, NHX1 may have a function in vesicular trafficking. Collectively, these reports indicate that endosomal NHX-type antiporters are critical regulators of endosomal trafficking likely by controlling the endosomal pH.

**Conclusion**

Salt stress is a major constrain for agriculture worldwide. However, the development of salt-tolerant crops has been far too slow. There are many reasons delaying this progress, but the fact that salt-tolerance relies on the combined regulation of hundreds of genes, might be the main one. Plants have evolved remarkable mechanisms to regulate K⁺ and Na⁺ tissue and cellular homeostasis under salt stress. Many of these mechanisms relay on H⁺, K⁺ and Na⁺ transporters. Over the last years, several molecular studies have established a strong involvement of SOS1, HKTs, and NHXs transporters in K⁺ and/or Na⁺ homeostasis and salt tolerance. Manipulation of some of these genes in model and crop plants yielded promising results in controlled conditions, but application in real agricultural conditions has been limited so far. The development of salt tolerant crops is unlikely to be successful until the development of new technical solutions allowing the fine-tuning regulation of multiple genes, preferentially in a tissue-specific manner. In addition, some of the ion transporters are also involved in
key cellular processes, and overexpression of those genes can introduce excessive perturbations of related cellular and physiological processes, limiting the improvement of the salt stress response. Therefore, in order to provide critical insights for the development of salt-tolerant crop plants, future research efforts should be directed towards a better understanding of the molecular mechanisms (e.g. epigenetic modifications, transcription factors, post-translation modifications) underlying the regulation of those transporters.

Acknowledgments

DMA was funded by Fundação para a Ciência e Tecnologia (FCT) through the fellowship SFRH/BD/65229/2009. NS was supported by Programa Ciência 2007 and FCT Investigator, funded by POPH (QREN). This work was also funded by Research unit GREEN-it “Bioresources for Sustainability” (UID/Multi/04551/2013) and by FCT project PTDC/BIA_BCM/099836/2008.

References

Adams E and Shin R (2014) Transport, signaling, and homeostasis of potassium and sodium in plants. J Integr Plant Biol 56:231-249.

Adler G, Blumwald E and Bar-Zvi D (2010) The sugar beet gene encoding the sodium/proton exchanger 1 (BvNHX1) is regulated by a MYB transcription factor. Planta 232:187-195.

Aharon GS, Apse MP, Duan S, Hua X and Blumwald E (2003) Characterization of a family of vacuolar Na+/H+ antiporters in Arabidopsis thaliana. Plant Soil 253:245-256.

Alexandersson E, Saalbach G, Larsson C and Kjellbom P (2004) Arabidopsis plasma membrane proteomics identifies components of transport, signal transduction and membrane trafficking. Plant Cell Physiol 45:1543-1556.

Ali R, Brett CL, Mukherjee S and Rao R (2004) Inhibition of sodium/proton exchange by a Rab-GTPase-activating protein regulates endosomal trafficking in yeast. J Biol Chem 279:4498-4506.

Almeida P, Katschig D and de Boer AH (2013) HKT transporters - State of the art. Int J Mol Sci 14:20359-20385.

Andrés Z, Perez-Hormaeche J, Leidi EO, Schlucking K, Steinhorst L, McLachlan DH, Schumacher K, Hetherington AM, Kudla J, Cubero B and Pardo JM (2014) Control of vacuolar dynamics and regulation of stomatal aperture by tonoplast potassium uptake. Proc Natl Acad Sci U S A 111:E1806-E1814.

Apse MP, Sottosanto JB and Blumwald E (2003) Vacuolar cation/H+ exchange, ion homeostasis, and leaf development are altered in a T-DNA insertion mutant of AtHKT1, the Arabidopsis vacuolar Na+/H+ antiporter. Plant J 36:229-239.

Apse MP, Aharon GS, Snedden WA and Blumwald E (1999) Salt tolerance conferred by overexpression of a vacuolar Na+/H+ antiport in Arabidopsis. Science 285:1256-1258.

Arango M, Gevaudant F, Ouattoule M and Boutilier M (2003) The plasma membrane proton pump ATPase: The significance of gene subfamilies. Planta 216:355-365.

Axelsen KB and Palmgren MG (2001) Inventory of the superfamily of P-type ion pumps in Arabidopsis. Plant Physiol 126:696-706.

Bacet D, Jiang J, Chung JS, Wang B, Chen J, Xin Z and Shi H (2011) Regulated AtHKT1 gene expression by a distal enhancer element and DNA methylation in the promoter plays an important role in salt tolerance. Plant Cell Physiol 52:149-161.

Barragan V, Leidi EO, Andres Z, Rubio L, De Luca A, Fernandez JA, Cubero B and Pardo JM (2012) Ion exchangers NHX1 and NHX2 mediate active potassium uptake into vacuoles to regulate cell turgor and stomatal function in Arabidopsis. Plant Cell 24:1127-1142.

Bassil E and Blumwald E (2014) The ins and outs of intracellular ion homeostasis: NHX-type cation/H+ transporters. Curr Opin Plant Biol 22:1-6.

Bassil E, Tajima H, Liang YC, Ohto MA, Ushijima K, Nakano R, Esumi T, Coku A, Belmonte M and Blumwald E (2011a) The Arabidopsis Na+/H+ antiporters NHX1 and NHX2 control vacuolar pH and K+ homeostasis to regulate growth, flower development, and reproduction. Plant Cell 23:3482-3497.

Bassil E, Ohto MA, Esumi T, Tajima H, Zhu Z, Cagnac O, Belmonte M, Peleg Z, Yamaguchi T and Blumwald E (2011b) The Arabidopsis intracellular Na+/H+ antiporters NHX5 and NHX6 are endosome associated and necessary for plant growth and development. Plant Cell 23:224-239.

Bassil E, Coku A and Blumwald E (2012) Cellular ion homeostasis: Emerging roles of intracellular NHX Na+/H+ antiporters in plant growth and development. J Exp Bot 63:5727-5740.

Batelli G, Verslues PE, Agius F, Qiu Q, Fujii H, Pan S, Schumaker KS, Grillo S and Zhu JK (2007) SOS2 promotes salt tolerance in part by interacting with the vacuolar H+-ATPase and upregulating its transport activity. Mol Cell Biol 27:7781-7790.

Berthomieu P, Conejero G, Nublat A, Brackenbury WJ, Lambert C, Savio C, Uozumi N, Oik S, Yamada K, Cellier F, et al. (2003) Functional analysis of AtHKT1 in Arabidopsis shows that Na+ recirculation by the phloem is crucial for salt tolerance. EMBO J 22:2004-2014.

Beyenbach KW and Wieczorek H (2006) The V-type H+ ATPase: Molecular structure and function, physiological roles and regulation. J Exp Biol 209:577-589.

Blumwald E, Aharon GS and Apse MP (2000) Sodium transport in plant cells. Biochim Biophys Acta 1465:140-151.

Bowers K, Levi BP, Patel FI and Stevens TH (2000) The sodium/proton exchanger NhX1 is required for endosomal protein trafficking in the yeast Saccharomyces cerevisiae. Mol Biol Cell 11:4277-4294.

Brett CL, Tukaye DN, Mukherjee S and Rao R (2005) The yeast endosomal Na+K+/H+ exchanger NhX1 regulates cellular pH to control vesicle trafficking. Mol Biol Cell 16:1396-1405.

Brini F and Khalek M (2012) Ion transporters and abiotic stress tolerance in plants. ISRN Mol Biol 2012:927436.

Brini F, Hanin M, Mezghani I, Berkowitz GA and Masmoudi K (2007) Overexpression of wheat Na+/H+ antiporter TNHX1 and H+-pyrophosphatase TVP1 improve salt- and drought-stress tolerance in Arabidopsis thaliana plants. J Exp Bot 58:301-308.
Bunney TD, van den Wijngaard PW and de Boer AH (2002) 14-3-3 protein regulation of proton pumps and ion channels. Plant Mol Biol 50:1041-1051.

Byrt CS, Platten JD, Spielmeyer W, James RA, Lagudah ES, Dennis ES, Tester M and Munns R (2007) HKT1;5-like cation transporters linked to Na⁺/Cl⁻ exclusion loci in wheat, Nax2 and Kna1. Plant Physiol 143:1918-1928.

Chaves MM and Oliveira MM (2004) Mechanisms underlying plant resilience to water deficits: Prospects for water-saving agriculture. J Exp Bot 55:2365-2384.

Chaves MM, Flexas J and Pinheiro C (2009) Photosynthesis under drought and salt stress: Regulation mechanisms from whole plant to cell. Ann Bot 103:551-560.

Choura M and Rebai A (2005) Identification and characterization of new members of vacuolar H⁺-Pyrophosphatase family from Oryza sativa genome. Russ J Plant Physiol 52:821-825.

Chung JS, Zhu JK, Bressan RA, Hasegawa PM and Shi H (2008) Reactive oxygen species mediate Na⁺-induced SOS1 mRNA stability in Arabidopsis. Plant J 53:554-565.

Conde A, Chaves MM and Gerós H (2011) Membrane transport, sensing and signaling in plant adaptation to environmental stress. Plant Cell Physiol 52:1583-1602.

Cotsaftis O, Plett D, Shirley N, Tester M and Hrmova M (2012) A two-staged model of Na⁺ exclusion in rice explained by 3D modeling of HKT transporters and alternative splicing. PLoS ONE 7:e39865.

Cuin TA, Bose J, Stefano G, Jha D, Tester M, Mancuso S and Shibata S (2011) Assessing the role of root plasma membrane and tonoplast Na⁺/H⁺ exchangers in salinity tolerance in wheat: In planta quantification methods. Plant Cell Environ 34:947-961.

Davenport RJ, Munoz-Mayor A, Jha D, Essah PA, Rus A and Tester M (2007) The Na⁺ transporter AtHKT1;1 controls retrieval of Na⁺ from the xylem in Arabidopsis. Plant Cell Environ 30:497-507.

Drozdowicz YM, Kissinger JC and Rea PA (2000) AVP2, a sequence-divergent, K⁺-insensitive H⁺-translocating inorganic pyrophosphatase from Arabidopsis. Plant Physiol 123:353-362.

Dubcovsky J, Maria GS, Epstein E, Luo MC and Dvorak J (1996) Mapping of the K⁺/Na⁺ discrimination locus Kna1 in wheat. Theor Appl Genet 92:448-454.

Duggleby RG and Dennis DT (1973) Pyruvate kinase, a possible regulatory enzyme in higher plants. Plant Physiol 52:312-317.

Fuchs I, Stölzl S, Ivashikina N and Hedrich R (2005) Rice K⁺ uptake channel OsAKT1 is sensitive to salt stress. Planta 221:212-221.

Fuglsang AT, Paez-Valencia J and Gaxiola RA (2010) Plant proton pumps: Regulatory circuits involving H⁺-ATPase and H⁺-PPase. In: Geisler M and Venema K (eds) Transporters and Pumps in Plant Signaling. Springer, Berlin pp 39-64.

Fukuda A, Nakamura A, Hara N, Toki S and Tanaka Y (2011) Molecular and functional analyses of rice NUX-type Na⁺/H⁺ antiporter genes. Planta 233:175-188.

Gao JP, Chao DY and LinHX (2007) Understanding abiotic stress tolerance mechanisms: Recent studies on stress response in rice. J Integr Plant Biol 49:742-750.

Gaxiola RA, Palmgren MG and Schumacher K (2007) Plant proton pumps. FEBS Lett 581:2204-2214.

Gregorio GB, Senadhira D, Mendoza RD, Manigbas NL, Roxas JP and Guerta CQ (2002) Progress in breeding for salinity tolerance and associated abiotic stresses in rice. Field Crops Res 76:91-101.

Hamaji, Nagira M, Yoshida K, Ohnishi M, Oda Y, Uemura T, Goh T, Sato MH, Morita MT, Tasaka M, et al. (2009) Dynamic aspects of ion accumulation by vesicle traffic under salt stress in Arabidopsis. Plant Cell Physiol 50:2023-2033.

Hanana M, Cagnac O, Yamaguchi T, Hamdi S, Ghorbel A and Blumwald E (2007) A grape berry (Vitis vinifera L.) cation/proton antiporter is associated with berry ripening. Plant Cell Physiol 48:804-811.

Hanitzsch M, Schnitzer D, Seidel T, Goldack D and Dietz KJ (2007) Transcript level regulation of the vacuolar H⁺-ATPase subunit isoforms VHA-a, VHA-E and VHA-G in Arabidopsis thaliana. Mol Membr Biol 24:507-518.

Haro R, Bañuelos MA, Senn ME, Barrero-Gil J and Rodríguez-Nava A (2005) HKT1 mediates sodium uniport in roots. Pitfalls in the expression of HKT1 in yeast. Plant Physiol. 139:1495-1506.

Hasegawa PM (2013) Sodium Na⁺ homeostasis and salt tolerance of plants. Environ Exp Bot 92:19-31.

He C, Yan J, Shen G, Fu L, Holaday AS, Auld D, Blumwald E and Zhang H (2005) Expression of an Arabidopsis vacuolar sodium/proton antiporter gene in cotton improves photosynthetic performance under salt conditions and increases fiber yield in the field. Plant Cell Physiol 46:1848-1854.

Hirsch RE, Lewis BD, Spalding EP and Sussman MR (1998) A role for the AKT1 potassium channel in plant nutrition. Science 280:918-921.

Horie T, Hauser F and Schroeder JJ (2009) HKT transporter-mediated salinity resistance mechanisms in Arabidopsis and monocot crop plants. Trends Plant Sci 14:660-668.

Horie T, Yoshida K, Nakayama H, Yamada K, Oiki S and Shinmyo A (2001) Two types of HKT transporters with different properties of Na⁺ and K⁺ transport in Oryza sativa. Plant J 27:129-138.

Horie T, Costa A, Kim TH, Han MJ, Horie R, Leung HY, Miyao A, Hirochika H, An G and Schroeder JJ (2007) Rice OsHKT2;1 transporter mediates large Na⁺ influx component into K⁺-starved roots for growth. EMBO J 20:3003-3014.

Huang S, Spielmeyer W, Lagudah ES, James RA, Platten JD, Dennis ES and Munns R (2006) A sodium transporter (HKT7) is a candidate for Nax1, a gene for salt tolerance in durum wheat. Plant Physiol. 142:1718-1727.

IRRI (2013) Rice Almanac. 4th edition. GRiSP (Global Rice Science Partnership) International Rice Research Institute, Los Baños, pp 298.

Jabnoune M, Espeut S, Mieulet D, Fitzames C, Verdeil JL, Concejero G, Rodriguez-Nava A, Sentenac H, Guiderdoni E, Abdellay C, et al. (2009) Diversity in expression patterns and functional properties in the rice HKT transporter family. Plant Physiol 150:1955-1971.
James RA, Davenport RJ and Munns R (2006) Physiological characterization of two genes for Na+ exclusion in durum wheat, Na+x1 and Na+x2. Plant Physiol 142:1537-1547.

James RA, Blake C, Byrt CS and Munns R (2011) Major genes for Na+ exclusion, Na+ x1 and Na+ x2 (wheat HKT1;4 and HKT1;5), decrease Na+ accumulation in bread wheat leaves under saline and waterlogged conditions. J Exp Bot 62:2939-2947.

Ji H, Pardo JM, Batelli G, Van Oosten MJ, Bressan RA and Li X (2013) The Salt Overly Sensitive (SOS) pathway: Established and emerging roles. Mol Plant 6:275-286.

Jiang X, Leidi EO and Pardo JM (2010) How dovacuolar NHX exchangers function in plant salt tolerance? Plant Signal Behav 55:792-795.

Kader MA, Seidel T, Golddack D and Lindberg S (2006) Expressions of OsHKT1, OsHKT2, and OsVHA are differentially regulated under NaCl stress in salt-sensitive and salt-tolerant rice (Oryza sativa L.) cultivars. J Exp Bot 57:4257-4268.

Kim BG, Waadt R, Cheong YH, Pandey GK, Dominguez-Solis JR, Schultheis S, Lee SC, Kudla J and Luan S (2007) The calcium sensor CBL10 mediates salt tolerance by regulating ion homeostasis in Arabidopsis. Plant J 52:473-484.

Kirsch M, An Z, Viereck R, Low R and Rausch T (1996) Salt stress induces an increased expression of V-type H+-ATPase in mature sugar beet leaves. Plant Mol Biol 32:543-547.

Klyuchnikov OI, Li KW, Lill H and de Boer AH (2007) The V-ATPase from etiolated barley (Hordeum vulgare L.) shoots is activated by blue light and interacts with 14-3-3 proteins. J Exp Bot 58:1013-123.

Kronzucker HJ and Britto DT (2011) Sodium transport in plants: A critical review. New Phytol 189:54-81.

Leidi EO, Barragan V, Rubio L, El-Hamdaoui A, Ruiz MT, Cubero B, Fernandez JA, Bressan RA, Hasegawa PM, Quintero FJ, et al. (2010) The AtNHX1 exchanger mediates potassium compartmentation in vacuoles of transgenic tomato. Plant J 61:495-506.

Liu C, Mao B, Ou S, Wang W, Liu L, Wu Y, Chu C and Wang X (2014) OshZIP71, a bZIP transcription factor, confers salinity and drought tolerance in rice. Plant Mol Biol 84:19-36.

Mansour MM (2014) The plasma membrane transport systems of Oryza sativa L. cultivars differing in salinity resistance Plant Growth Regulation 78:527-528.

Maeshima M (2000) Vacuolar H+-pyrophosphatase. Biochim Biophys Acta 1465:37-51.

Marschner H (1995) Mineral nutrition of higher plants. Ann Bot 78:527-528.

Martínez-Atienza J, Jiang X, García-deblas B, Mendoza I, Zhu JK, Pardo JM and Quintero FJ (2007) Conservation of the salt overly sensitive pathway in rice. Plant Physiol 143:1001-1012.

Martineiro A, Bassil E, Jublance E, Alcon C, Reguera M, Sentenac H, Blumwald E and Paris N (2013) In vivo intracellular pH measurements in tobacco and Arabidopsis reveal an unexpected pH gradient in the endomembrane system. Plant Cell 25:4028-4043.

Maser P, Hosoo Y, Goshima S, Horie T, Eckelman B, Yamada K, Yoshida K, Bakker EP, Shimmyo A, Oki S, et al. (2002) Glycine residues in potassium channel-like selectivity filters determine potassium selectivity in four-loop-per-subunit HKT transporters from plants. Proc Natl Acad Sci USA 99:6428-6433.

Matsumoto H and Chung GC (1988) Increase in proton-transport activity of tonoplast vesicles as an adaptive response of barley roots to NaCl stress. Plant Cell Physiol 29:1133-1140.

McAinsh MR, Clayton H, Mansfield TA and Hetherington AM (1996) Changes in stomatal behavior and guard cell cytoplasmic free calcium in response to oxidative stress. Plant Physiol 111:1031-1042.

Mian A, Oomen RJ, Isayenkov S, Sentenac H, Maathuis FJ and Very AA (2011) Over-expression of an Na+-and K+-permeable HKT transporter in barley improves salt tolerance. Plant J 68:468-479.

Mitsuda N, Enami K, Nakata M, Takeyasu K and Sato MH (2001) Novel type Arabidopsis thaliana H+-PPase is localized to the Golgi apparatus. FEBS Lett 488:29-33.

Mitsuda N, Isono T and Sato MH (2003) Arabidopsis CAMTA family proteins enhance V-PPase expression in pollen. Plant Cell Physiol 44:975-981.

Mitsuda N, Hisabori T, Takeyasu K and Sato MH (2004) VOZ; isolation and characterization of novel vascular plant transcription factors with a one-zinc finger from Arabidopsis thaliana. Plant Cell Physiol 45:845-854.

Møller IS, Gilliham M, Jha D, Mayo GM, Roy SJ, Coates JC, Haseloff J and Tester M (2009) Shoot Na+ exclusion and increased salinity tolerance engineered by cell type-specific alteration of Na+ transport in Arabidopsis. Plant Cell 7:2163-2178.

Munns R (2005) Genes and salt tolerance: Bringing them together. New Phytol 167:645-663.

Munns R and Tester M (2008) Mechanisms of salinity tolerance. Annu Rev Plant Biol 59:651-681.

Munns R, James RA, Xu B, Athman A, Conn SJ, Jordans C, Byrt CS, Hare RA, Tyerman SD, Tester M, et al. (2012) Wheat grain yield on saline soils is improved by an ancestral Na+ transporter gene. Nat Biotechnol 30:360-364.

Narasimhan ML, Binzel ML, Perez-Prat E, Chen Z, Nelson DE, Singh NK, Bressan RA and Hasegawa PM (1991) NaCl regulation of tonoplast ATPase 70 kilodalton subunit mRNA in tobacco cells. Plant Physiol 97:562-568.

Negrão S, Almedanim MC, Pires IS, Abreu IA, Maroco J, Courtois B, Gregorio GB, McNally KL and Oliveira MM (2013) New allelic variants found in key rice salt-tolerance genes: An association study. Plant Biotechnol J 11:87-100.

Neves-Cordones M, Alemán F, Martínez V and Rubio F (2010) The Arabidopsis thaliana HAK5 K+ transporter is required for plant growth and K+ acquisition from low K+ solutions under saline conditions. Mol Plant 3:326-333.

Nobel PS (2009) Physicochemical and Environmental Plant Physiology. 4th edition. Academic Press, Amsterdam, pp 589.

Ohnishi M, Fukada-Tanaka S, Hoshino A, Takada J, Inagaki Y and Iida S (2005) Characterization of a novel Na+/-H+ antiporter gene InNHX2 and comparison of InNHX2 with InNHX1, which is responsible for blue flower coloration by increasing the vacuolar pH in the Japanese morning glory. Plant Cell Physiol 46:259-267.
Ohta M, Guo Y, Halfter U and Zhu JK (2003) A novel domain in the protein kinase SOS2 mediates interaction with the protein phosphatase 2C AB12. Proc Natl Acad Sci U S A 100:11771-11776.

Oomen RJ, Benito B, Sentenc H, Rodriguez-Navarro A, Talón M, Véry AA and Domingo C (2012) HKT2;2/1, a K⁺-permeable transporter identified in a salt-tolerant rice cultivar through surveys of natural genomic polymorphism. Plant J 71:750-762.

Pardo JM (2010) Biotechnology of water and salinity stress tolerance. Curr Opin Biotechnol 21:185-196.

Pardo JM and Quintero FJ (2002) Plants and sodium ions: Keeping company with the enemy. Genome Biol 3:1017.1-1017.4.

Peleg Z, Apse MP and Blumwald E (2011) Engineering salinity and water-stress tolerance in crop plants: Getting closer to the field. Adv Bot Res 57:405-443.

Pires IS, Negrão S, Oliveira MM and Purugganan MD (2015) Comprehensive phenotypic analysis of rice (Oryza sativa) response to salinity stress. Physiol Plant 155:43-54.

Platten JD, Cotsaftis O, Berthomieu P, Bohnert H, Davenport RJ, Pires IS, Negrão S, Oliveira MM and Purugganan MD (2015) Nomenclature for HKT transporters, key determinants of plant salinity tolerance. Trends Plant Sci 11:372-374.

Pons R, Corneo MJ and Sanz A (2011) Differential salinity-induced variations in the activity of H⁺-pumps and Na+/H⁺ antiporters that are involved in cytoplasm ion homeostasis as a function of genotype and tolerance level in rice cell lines. Plant Physiol Biochem 49:1399-1409.

Pottosin I, Wherrett T and Shabala S (2009) SV channels dominate the vacuolar Ca²⁺ release during intracellular signaling. FEBS Lett 583:921-926.

PP1 - Potash and Phosphate Institute (1998) Potassium for Agriculture. Better Crops with Plant Food. Vol 32, N. 3. Potash and Phosphate Institute, Atlanta.

Qiu QS, Guo Y, Quintero FJ, Pardo JM, Schumaker KS and Zhu JK (2004) Regulation of vacuolar Na⁺/H⁺ exchange in Arabidopsis thaliana by the Salt-Overly-Sensitive (SOS) pathway. J Biol Chem 279:207-215.

Quan R, Lin H, Mendoza I, Zhang Y, Cao W, Yang Y, Shang M, Chen S, Pardo JM and Guo Y (2007) SCABP8/CBL10, a putative calcium sensor, interacts with the protein kinase SOS2 to protect Arabidopsis shoots from salt stress. Plant Cell 19:1415-1431.

Quintero FJ, Ohta M, Shi H, Zhu JK and Pardo JM (2002) Reconstitution in yeast of the Arabidopsis SOS signaling pathway for Na⁺ homeostasis. Proc Natl Acad Sci U S A 99:9061-9066.

Ratajczak R (2000) Structure, function and regulation of the plant vacuolar H⁺-translocating ATPase. Biochem Biophys Acta 1465:17-36.

Ratajczak R, Hinz G and Robinson DG (1999) Localization of pyrophosphatase in membranes of cauliflower in florence cells. Planta 208:205-211.

Reguera M, Bassil E, Tajima H, Wimmer M, Chanoca A, Otegui MS, Paris N and Blumwald E (2015) pH Regulation by NHX-type antiporters is required for receptor-mediated protein trafficking to the vacuole in Arabidopsis. Plant Cell 27:1200-1217.

Ren XL, Qi GN, Feng HQ, Zhao S, Zhao SS, Wang Y and Wu WH (2013) Calcineurin B-like protein CBL10 directly interacts with AKT1 and modulates K⁺ homeostasis in Arabidopsis. Plant J 74:258-266.

Ren ZH, Gao JP, Li LG, Cai XL, Huang W, Chao DY, Zhu MZ, Wang ZY, Luan S and Lin HX (2005) A rice quantitative trait locus for salt tolerance encodes a sodium transporter. Nat Genet 37:1141-1146.

Rodriguez-Rosales MP, Jiang X, Galvez FJ, Aranda MN, Cubero B and Venema K (2008) Overexpression of the tomato K⁺/H⁺ antiporter LeNHX2 confers salt tolerance by improving potassium compartmentalization. New Phytol 179:366-377.

Rodriguez-Rosales MP, Galvez FJ, Huertas R, Aranda MN, Baghous M, Cagnac O and Venema K (2009) Plant NHX cation/proton antiporters. Plant Signal Behav 4:265-276.

Roy SJ, Negrão S and Tester M (2014) Salt resistant crop plants. Curr Opin Biotechnol 26:115-124.

Rus A, Lee BH, Munoz-Mayor A, Sharkhua A, Miura K, Zhu JK, Bressan RA and Hasegawa PM (2004) AtHKT1 facilitates Na⁺ homeostasis and K⁺ nutrition in plants. Plant Physiol 136:2500-2511.

Sakakibara Y, Kobayashi H and Kasamo K (1996) Isolation and characterization of cDNAs encoding vacuolar H⁺-pyrophosphatase isoforms from rice (Oryza sativa L.). Plant Mol Biol 31:1029-1038.

Sato Y and Sakaguchi M (2005) Topogenic properties of transmembrane segments of Arabidopsis thaliana NHX1 reveal a common topology model of the Na⁺/H⁺ exchanger family. J Biochem 138:425-431.

Schachtman DP and Schroeder JI (1994) Structure and transport mechanism of a high-affinity potassium uptake transporter from higher plants. Nature 370:655-658.

Shabala S and Cuin TA (2008) Potassium transport and plant salt tolerance. Physiol Plant 133:651-669.

Shi H and Zhu JK (2002) Regulation of expression of the vacuolar Na⁺/H⁺ antiporter gene AtNHX1 by salt stress and abscisic acid. Plant Mol Biol 50:543-550.

Shi H, Ishitani M, Kim C and Zhu JK (2000) The Arabidopsis thaliana salt tolerance gene SOS1 encodes a putative Na⁺/H⁺ antiporter. Proc Natl Acad Sci U S A 97:6896-6901.

Shi H, Quintero FJ, Pardo JM and Zhu JK (2002) The putative plasma membrane Na⁺/H⁺ antiporter SOS1 controls long-distance Na⁺ transport in plants. Plant Cell 14:465-477.

Shi H, Lee BH, Wu SJ and Zhu JK (2003) Overexpression of a plasma membrane Na⁺/H⁺ antiporter SOS1 under controls long-distance Na⁺ transport in plants. Plant Cell 136:2500-2511.

Silva P and Gerós H (2009) Regulation by salt of vacuolar H⁺-ATPase and H⁺-pyrophosphatase activities and Na⁺/H⁺ exchange. Plant Signal Behav 4:718-726.

Sottosanto JB, Gelli A and Blumwald E (2004) DNA array analyses of Arabidopsis thaliana lacking a vacuolar Na⁺/H⁺ antiporter: Impact of AtNHX1 on gene expression. Plant J 40:752-771.

Sun J, Dai S, Wang R, Chen S, Li N, Zhou X, Lu C, Shen X, Zheng X, Hu Z, et al. (2009) Calcium mediates root K⁺/Na⁺ homeostasis in poplar species differing in salt tolerance. Tree Physiol 29:1175-1186.

Sunarpi HT, Motoda J, Kubo M, Yang H, Yoda K, Horie R, Chan WY, Leung HY, Hattori K, et al. (2005) Enhanced salt tolerance mediated by AtHKT1 transporter-induced Na unload-
ing from xylem vessels to xylem parenchyma cells. Plant J. 44:928-938.

Sze H, Schumacher K, Muller ML, Padmanaban S and Taiz L (2002) A simple nomenclature for a complex proton pump: \textit{VHA} genes encode the vacuolar H\(^+\)-ATPase. Trends Plant Sci 7:157-161.

Takemiya A, Kinoshita T, Asanuma M and Shimazaki K (2006) Protein phosphatase 1 positively regulates stomatal opening in response to blue light in \textit{Vicia faba}. Proc Natl Acad Sci U S A 103:13549-13554.

Tester M and Davenport R (2003) Na\(^+\) tolerance and Na\(^+\) transport in higher plants. Ann Bot 91:503-527.

Tsai Y-C, Hong C-Y, Liu L-F and Kao CH (2004) Relative importance of Na\(^+\) and Cl\(^-\) in NaCl-induced antioxidant systems in roots of rice seedlings. Physiol Plant 122:86-94.

Venema K, Quintero FJ, Pardo JM and Donaire JP (2002) The Arabidopsis Na\(^+\)/H\(^+\) exchanger AtNHX1 catalyzes low affinity Na\(^+\) and K\(^+\) transport in reconstituted liposomes. J Biol Chem 277:2413-2418.

Whiteman SA, Nuhse TS, Ashford DA, Sanders D and Maathuis FJ (2008a) A proteomic and phosphoproteomic analysis of \textit{Oryza sativa} plasma membrane and vacuolar membrane. Plant J 56:146-156.

Whiteman SA, Serazetdinova L, Jones AM, Sanders D, Rathjen J, Peck SC and Maathuis FJ (2008b) Identification of novel proteins and phosphorylation sites in a tonoplast enriched membrane fraction of \textit{Arabidopsis thaliana}. Proteomics 8:3536-3547.

Wu SJ, Ding L and Zhu JK (1996) SOS1, a genetic locus essential for salt tolerance and potassium acquisition. Plant Cell 8:617-627.

Wu Y, Hu Y and Xu G (2009) Interactive effects of potassium and sodium on root growth and expression of K\(^+\)/Na\(^+\) transporter genes in rice. Plant Growth Regulation 57:271-280.

Xu H, Jiang X, Zhan K, Cheng X, Chen X, Pardo JM and Cui D (2008) Functional characterization of a wheat plasma membrane Na\(^+\)/H\(^+\) antiporter in yeast. Arch Biochem Biophys 473:8-15.

Xue Z-Y, Zhi D-Y, Xue G-P, Zhang H, Zhao Y-X and Xia G-M (2004) Enhanced salt tolerance of transgenic wheat (\textit{Triticum aestivum} L.) expressing a vacuolar Na\(^+\)/H\(^+\) antiporter gene with improved grain yields in saline soils in the field and a reduced level of leaf Na\(^+\). Plant Sci 167:849-859.

Yamaguchi T, Apsel MP, Shi H and Blumwald E (2003) Topological analysis of a plant vacuolar Na\(^+\)/H\(^+\) antiporter reveals a luminal C-terminus that regulates antiporter cation selectivity. Proc Natl Acad Sci U S A 100:12510-12515.

Yamaguchi T, Aharon GS, Sottosanto JB and Blumwald E (2005) Vacuolar Na\(^+\)/H\(^+\) antiporter cation selectivity is regulated by calmodulin from within the vacuole in a Ca\(^{2+}\) - and pH-dependent manner. Proc Natl Acad Sci U S A 102:16107-16112.

Yamaguchi T, Fukuda-Tanaka S, Inagaki Y, Saito N, Yonekura-Sakakibara K, Tanaka Y, Kusumi T and Iida S (2001) Genes encoding the vacuolar Na\(^+\)/H\(^+\) exchanger and flower coloration. Plant Cell Physiol 42:451-461.

Yang T and Poooviah BW (2003) Calcium/calmodulin-mediated signal network in plants. Trends Plant Sci 8:505-512.

Yao X, Horie T, Xue S, Leung HY, Katsuhara M, Brodsky DE, Wu Y and Schroeder JI (2010) Differential sodium and potassium transport selectivities of the rice OsHKT2;1 and OsHKT2;2 transporters in plant cells. Plant Physiol 152:341-355.

Yeo AR, Yeo ME and Flowers TJ (1987) The contribution of an apoplastic pathway to sodium uptake by rice roots in saline conditions. J Exp Bot 38:1141-1153.

Yokoi S, Quintero FJ, Cubero B, Ruiz MT, Bressan RA, Hasegawa PM and Pardo JM (2002) Differential expression and function of \textit{Arabidopsis thaliana} NHX Na\(^+\)/H\(^+\) antiporters in the salt stress response. Plant J 30:529-539.

Yoshida K, Miki N, Memonoki K, Kawachi M, Katou K, Okazaki Y, Uozumi N, Maeshima M and Kondo T (2009) Synchrony between flower opening and petal-color change from red to blue in morning glory, \textit{Ipomoea tricolor} cv. Heavenly Blue. Proc Jpn Acad Ser B Phys Biol Sci 85:187-197.

Zhang H-X and Blumwald E (2001) Transgenic salt-tolerant tomato plants accumulate salt in foliage but not in fruit. Nat Biotechnol 19:765-768.

Zhang X, Zhang L, Dong F, Gao J, Galbraith DW and Song CP (2001) Hydrogen peroxide is involved in abscisic acid-induced stomatal closure in \textit{Vicia faba}. Plant Physiol 126:1438-1448.

Zhao F-Y, Zhang X-J, Li P-H, Zhao Y-X and Zhang H (2006) Co-expression of the \textit{Suaeda salsa} \textit{SnNHX1} and Arabidopsis \textit{AVP1} confer greater salt tolerance to transgenic rice than the single \textit{SnNHX1}. Mol Breed 17:341-353.

Zhu JK (2003) Regulation of ion homeostasis under salt stress. Curr Opin Plant Biol 6:441-445.

License information: This is an open-access article distributed under the terms of the Creative Commons Attribution License (type CC-BY), which permits unrestricted use, distribution and reproduction in any medium, provided the original article is properly cited.