**ABCB1** polymorphism as prognostic factor in breast cancer patients treated with docetaxel and doxorubicin neoadjuvant chemotherapy

Hee-Jun Kim,1 Seock-Ah Im,2,3 Bhumsuk Keam,2,3 Hye Seon Ham,2 Kyung Hun Lee,2,3 Tae Yong Kim,2,3 Yu Jung Kim,3,4 Do-Youn Oh,2,3 Jee Hyun Kim,2,3,4 Wonshik Han,3,5 In-Jin Jang,6 Tae-You Kim,2,3 In Ae Park7 and Dong Young Noh1,2,3

1Department of Internal Medicine, Chung-Ang University College of Medicine, Seoul; 2Department of Internal Medicine, Seoul National University College of Medicine, Seoul; 3Cancer Research Institute, Seoul National University, Seoul; 4Department of Internal Medicine, Seoul National University Bundang Hospital, Seongnam; 5Department of Surgery, Seoul National University College of Medicine, Seoul; 6Department of Clinical Pharmacology, Seoul National University College of Medicine, Seoul; 7Department of Pathology, Seoul National University College of Medicine, Seoul, Korea

Key words
ABCB1 Gene, breast cancer, C3435T, neoadjuvant chemotherapy, single nucleotide polymorphism

Expression of the adenosine triphosphate-binding cassette B1 (**ABCB1**) transporter and P-glycoprotein are associated with resistance to anticancer drugs. The purpose of this study was to investigate the role of single nucleotide polymorphism in the **ABCB1** and CYP3A genes in breast cancer patients who were treated with neoadjuvant chemotherapy. Stage II/III breast cancer patients were treated with three cycles of neoadjuvant, after which the patients received curative surgery and adjuvant chemotherapy. The polymorphisms of **ABCB1** and CYP3A were genotyped. The correlation of polymorphism of **ABCB1**, CYP3A, and clinical outcomes was analyzed. Among the 216 patients, **ABCB1** C3435T genotype had a longer overall survival (OS). than CC/CT. Multivariate analyses demonstrated that good PS, invasive ductal carcinoma, non-triple negative phenotype and initial pathological response from neoadjuvant therapy, which has been shown to be associated with a good prognosis.4,5

As neoadjuvant chemotherapy is becoming increasingly more important in clinical practice, a disputed issue is the development of resistance to the cytotoxic agents as well as cross-resistance to substances patients have never been exposed to before. Inter-individual variability of pharmacokinetics of anti-cancer drugs constitutes a major limitation to their clinical use. Since a variety of commonly used chemotherapeutic agents such as docetaxel and doxorubicin have narrow therapeutic indices and significant inter-individual variations in drug disposition, great efforts have been put in determining the mechanism of person-to-person pharmacokinetic diversities.

Single nucleotide polymorphisms (SNPs) in adenosine triphosphate-binding cassette B1 (**ABCB1**) and cytochrome P450 (CYP) genes are among potential candidates that explain the inter-individual variations in drug disposition. SNPs in **ABCB1** gene are associated with phenotypic variation in P-glycoproteins (P-gp), a membrane-bound efflux pump, which removes chemotherapeutic drugs including docetaxel and doxorubicin from the cells. The intestinal P-gp plays a major role in the fecal elimination of drugs by modulating reabsorption of the drug after hepatobiliary secretion.6,7,8 The percentage of tumors expressing P-gp in breast cancer was 41.2% and the expression rate of P-gp increased after treatment which was...
three times more likely to fail in chemotherapy. (7) Recently, SNPs of the \textit{ABCB1} gene have been reported to be associated with taxane clearance and the clinical outcome of patients who were treated with taxane. (8,9) Doxorubicin was also shown to be functionally affected by SNPs of \textit{ABCB1} with regards to the pharmacokinetics. (10,11) It has been shown that \textit{ABCB1} C3435T polymorphism in exon 26 is significantly related to clinical toxicity and high blood levels of docetaxel. (12) \textit{ABCB1} G2677T/A polymorphism in exon 21 are associated with treatment outcomes after paclitaxel monotherapy. (11) \textit{ABCB1} C1236T polymorphism in exon 12 is associated with docetaxel clearance. (8,9)

Most chemotherapeutic agents are eliminated via hepatic metabolism, which is mediated by cytochrome P450 (CYP), mainly CYP 3A4 and CYP3A5. (13) Metabolism of docetaxel consists of CYP3A-mediated oxidation of the tert-butyl propionate side chain, which results in the formation of four metabolites with reduced cytotoxic activity. (14) CYP3A4*1B and CYP3A5*3 are common polymorphisms, which have been shown to have clinical implications in some studies. (14,15) CYP3A4*1B, an A-392G transition in the 5'-flanking region, is a promoter polymorphism. While the polymorphism is commonly found in the African-American population, (14) its minor allele frequency is estimated 0% in Chinese, Taiwanese, Chinese Americans, and Japanese Americans. (16,17) On the other hand, the CYP3A5*3 splice site variant that causes loss of hepatic expression of CYP3A5 is common, but does not account for inter-individual variability of docetaxel disposition in Asian populations. (18,19) P-gp may affect the extent of CYP3A-mediated metabolism of drugs by limiting the intracellular substrate availability. (20) This study hypothesized from the above findings that a genetic variant of \textit{ABCB1} and \textit{CYP3A} contributes to the pharmacokinetic variability of docetaxel and doxorubicin.

This study investigated the distribution of SNPs in \textit{ABCB1} and \textit{CYP3A} genes in breast cancer patients who were treated with neoadjuvant docetaxel and doxorubicin chemotherapy. The correlation between the genetic polymorphism, plasma concentration of chemotherapeutic agents and clinical outcomes was also explored.
**Materials and Methods**

**Patients and study design.** From September 2003 to September 2008, pathologically proven, previously untreated, clinical stage II or III breast cancer patients were enrolled in this prospective study. Patients had to have an Eastern Cooperative Oncology Group (ECOG) performance status (PS) of 0–2, objective measurable disease, and adequate bone marrow, hepatic, cardiac, and renal functions. Initial evaluations included physical examination, mammography, breast ultrasonography, computed tomography (CT) of the chest, bone scan, and breast magnetic resonance imaging (MRI). Initial nodal staging was evaluated by the physical examination and CT.

All patients were treated with three cycles of neoadjuvant chemotherapy which consisted of docetaxel (75 mg/m²) and doxorubicin (60 mg/m²) by intravenous infusion. Granulocyte colony-stimulating factor as primary prophylaxis was injected for 5 days every cycle. After completion of neoadjuvant treatment, the patients were re-evaluated for the response. After the primary surgery, patients were treated with three more cycles of docetaxel and doxorubicin as the adjuvant chemotherapy. Adequate radiation or hormonal therapy was performed as indicated. The radiological response was evaluated using breast MRI for the primary breast lesion and chest CT for the lymph node lesions with RECIST criteria 1.0. Pathologic complete response (pCR) was defined as complete disappearance of invasive carcinoma in both breast and axillary lymph nodes after three cycles of chemotherapy. For estrogen receptor (ER) and progesterone receptor (PR), cases with 10% or more positive staining were grouped as positive. Human epidermal growth factor receptor-2 (HER2) overexpression was demonstrated by immunohistochemistry scoring or fluorescence in situ hybridization (FISH) in paraffin-
sections of their primary carcinomas. Immunohistochemistry scores of only 3+ on a scale of 0 to 3+ or FISH positivity were defined as overexpression of HER2. Triple negative breast cancer was defined as the lack of expression of ER and PR and the absence of HER2 overexpression. Recurrence free survival (RFS) was calculated as the time from the first cycle of chemotherapy to the diagnosis of a recurrent disease in the ipsilateral breast, local, regional, or distant site, and the overall survival (OS) was the time from the first cycle of chemotherapy to death or last follow-up day. Adverse events were assessed in all patients with version 3.0 of the National Cancer Institute's Common Terminology Criteria for Adverse Events. The study protocol was approved by the Institutional Review Board of Seoul National University Hospital (IRB Registration No. H-0510-506-159, H-0610-020-186). This manuscript is a part of a clinical trial (ClinicalTrials.gov Identifier: NCT01396655).

Analysis of \textit{ABCB1} and \textit{CYP3A} genomic polymorphism. Whole blood samples were obtained before the neoadjuvant chemotherapy. Genomic DNA was isolated from mononuclear cells of peripheral blood cells using QiaAmp blood mini kit (Qiagen, Hilden, Germany). \textit{ABCB1} C3435T\(^{(26,27)}\), G2677T\(^{A}\) \(^{(11,27)}\) and \textit{C1236T}\(^{B}\) \(^{(28)}\) polymorphisms were genotyped by polymerase chain reaction (PCR) and restriction fragment length polymorphism (RFLP) assays as previously described.\(^{(29)}\) \textit{CYP3A4*1B} and \textit{CYP3A5*3} polymorphisms were done by the GoldenGate assay (Illumina, San Diego, CA, USA) that consisted of 96 custom selected SNPs within 34 genes. The dissolved biotinylated DNA pellet was extended and ligated. The supernatant was then used in PCR amplification. Double-stranded PCR products were immobilized onto paramagnetic particles, B Reagent (Illumina). The bound PCR products were denatured and the released ssDNAs were tralized with hybridization reagent (Illumina). Hybridization was done and then imaged using a BeadArray Reader (Illumina).

**Pharmacokinetic studies.** Seventy-two out of 216 patients participated in the prospective pharmacokinetic study of docetaxel and doxorubicin. These patients were grouped as the pharmacokinetic (PK) study group. Pharmacokinetic samplings were done at three time points. The plasma concentrations of docetaxel and doxorubicin at pre-concentration (just before infusion), post-concentration (just after infusion), and 24 h-concentration after infusion were determined by liquid chromatography tandem mass (LC/MS/MS) assays. Mass spectrometric analysis was done using an API 3000 MS system (Applied Biosystems, Foster City, CA, USA). All plasma samples were subjected to the sample preparation procedure described previously.\(^{(30)}\) With the above three blood samples, a graph was plotted of the sampling point as the \(x\)-axis against the drug concentration as the \(y\)-axis and was able to draw a diagram corresponding to the area under the plasma concentration (AUC) until 24 h after drug infusion. Based on this diagram, the drug concentration was calculated and was defined as the AUC in this study. The AUC of docetaxel and doxorubicin were dictated as AUC\(_{\text{doc}}\) and AUC\(_{\text{dox}}\), respectively. The differences between the AUCs were examined until the 24 h-concentration after infusion, according to the genotypes of \textit{ABCB1} and \textit{CYP3A5}.

**Statistical analysis.** This is a project of a phase II trial. The primary endpoint in this study was pCR. Secondary end points included rate of breast conserving operation, toxicity, relapse-free survival, overall survival and predictive/prognostic factors. The association between the allelic frequency of \textit{ABCB1} and clinical outcomes was assessed by the \(\chi^2\) test or Fisher’s exact test. The RFS and OS were estimated by the Kaplan–Meier method, and the difference in survival was compared using the log-rank test. Cox regression model was used to compare the clinical outcomes among the

### Table 3. Multivariate analysis for overall survival

| Characteristics                  | Category            | Hazard ratio† | 95% CI          | P-value |
|----------------------------------|---------------------|---------------|-----------------|---------|
| ECOG                             | 0-1                 | 1             |                 |         |
|                                 | 2                   | 9.526         | 2.587–20.493    | 0.003   |
| Histology                        | Invasive ductal carcinoma | 1            |                 |         |
|                                 | Others*             | 3.547         | 1.205–10.444    | 0.022   |
| Triple negative                  | Non-triple          | 1             |                 |         |
| Phenotype                        | Triple negative     | 2.106         | 1.145–3.874     | 0.017   |
| Pathologic                       | pCR                 | 1             |                 |         |
| Response                         | Non-pCR             | 8.204         | 1.810–19.112    | 0.047   |
| ABCB1 C3435T                     | CC + CT             | 1             |                 |         |
|                                 | TT                  | 0.245         | 0.059–1.026     | 0.054   |

†Hazard ratio was calculated by Cox’s proportional hazard model. If the hazard ratio is >1, the hazard ratio can be thought of as the average increased risk of death compared with the reference group (upper line). ‡According to the TNM classification of 2002 AJCC staging. *Among the 10 patients, 7 had invasive lobular carcinomas, 3 medullary carcinomas and 1 tubular carcinoma.

### Table 4. Toxicities in total patients (n = 216)

| Grade | Neutropenia | Thrombocytopenia | Febrile neutropenia | Nausea | Vomiting | Stomatitis | Diarrhea | Neuropathy |
|-------|-------------|------------------|---------------------|--------|----------|------------|----------|------------|
| 1 (%) | 16 (7.4)    | 11 (5.1)         | 16 (7.4)            | 28 (13.0) | 20 (9.3) | 16 (7.4) | 14 (6.5) | 22 (10.1) |
| 2 (%) | 13 (6.0)    | 5 (2.3)          | 9 (4.2)             | 13 (6.0) | 15 (6.9) | 6 (2.8)   | 9 (4.2)  | 7 (3.2)    |
| 3 (%) | 14 (6.4)    | 9 (4.2)          | 3 (1.4)             | 3 (1.4) | 12 (5.6) | 3 (1.4)   | 12 (5.6) | 7 (3.2)    |
| 4 (%) | 7 (3.2)     | –                | –                   | –       | –        | –          | –        | –          |

Toxicity was evaluated by CTCAE ver. 3. CI, confidence interval; HR, hazard ratio; IDC, invasive ductal carcinoma; OS, overall survival.
Table 5. Toxicities according to the polymorphism of ABCB1 gene

| SNPs | Neutropenia† | Diarrhea | Stomatitis† | Nausea† | Febrile neutropenia† | RR | (95% CI) | P  | RR | (95% CI) | P  | RR | (95% CI) | P  |
|------|-------------|----------|------------|---------|---------------------|----|----------|-----|----|----------|-----|----|----------|-----|
| C3435T | CT vs. TT | 7.0 | 1 (2.3-11.6) | 0.037 | 3.2 | 1 (0.7-17.7) | 0.123 | 1.6 | 6.5 | 3.6 (0.5-21.0) | 0.121 | 1.6 | 4.4 | 2.1 (1.3-17.4) | 0.114 |
| GG vs. Non-GG | TT vs. CT | 9.7 | 1 (2.1-16.9) | 0.201 | 1.3 | 1 (0.5-4.1) | 0.209 | 2.8 | 1.8 | 0.4 (4.1-14.5) | 0.121 | 1.6 | 4.4 | 2.1 (1.3-17.4) | 0.114 |
| C1236T | CC vs. TT | 11.2 | 1 (2.1-16.9) | 0.201 | 1.3 | 1 (0.5-4.1) | 0.209 | 2.8 | 1.8 | 0.4 (4.1-14.5) | 0.121 | 1.6 | 4.4 | 2.1 (1.3-17.4) | 0.114 |
| Non-GG vs. GG | C2 vs. TT | 11.2 | 1 (2.1-16.9) | 0.201 | 1.3 | 1 (0.5-4.1) | 0.209 | 2.8 | 1.8 | 0.4 (4.1-14.5) | 0.121 | 1.6 | 4.4 | 2.1 (1.3-17.4) | 0.114 |

†-Grade 3 Toxicities.
‡RR, relative risk; 95% CI, 95% confidence interval.

Results

Patients and treatment outcomes. A total of 216 breast cancer patients were enrolled. The median follow-up duration was 85.4 months (range 26.2-117.8 months) and 75 patients (34.7%) experienced relapse and 43 patients (19.9%) died. Baseline characteristics of the study population are presented in Table 1. The median age of the study population was 44 years (range 25-69 years). ABCB1 C3435T and ABCB1 C1236T were divided into two groups as the CC/CT vs. TT genotype and the ABCB1 G2677T/A was grouped into the GG vs. non-GG group.

Pharmacokinetic analysis was done in 72 patients. Baseline characteristics of these patients did not differ greatly from those of the total patient group.

Association between genotypes and treatment outcomes. Frequency of ABCB1 3435TT genotype was 14.4% in total 216 patients and 9.7% in the PK group of 72 patients. In PK group, the AA genotype of CYP3A4 was 100% and the AA genotype of CYP3A5 had the lowest percentage, 4.2%. The proportion of each genotype was maintained similarly in 72 PK group patients. Each observed genotype distribution of ABCB1 genes conformed to Hardy–Weinberg equilibrium (each P > 0.050). Linkage disequilibrium for all pairs of exons 26, 21, and 12 were observed by use of the PL-EM algorithm (P < 0.001).

Analysis of clinical outcomes of the total patients. The overall clinical response rate (RR) was 76.8%. Twenty patients (9.3%) achieved clinical complete response (cCR) and 18 (8.3%) achieved pCR. Among the PK group, RR was 82.0% with 4.2% of pCR. ABCB1 C3435T and ABCB1 C1236T did not predict the responders. As shown in Figure 1, ABCB1 3435TT genotype had a longer OS than the CT/TT genotype with statistical significance (P = 0.024). A similar trend was observed for the RFS, that is, ABCB1 3435TT genotype tended to have a longer RFS although it was not statistically significant (P = 0.234). ABCB1 G2677T/A and C1236T were analyzed, and there were no significant differences in the RFS and OS (Fig. S1).

With univariate analysis of the OS, good PS (ECOG 0-1), invasive ductal carcinoma, initial operable stage, ER-positivity, non-triple negative phenotype, breast conserving surgery and the TT genotype of ABCB1 C3435T were associated with a lower risk of death (Table 2). Multivariate analyses of the OS demonstrated that poor PS (HR 9.526, 95% CI 2.587-20.493; P = 0.003), triple negative phenotype (HR 2.106, 95% CI 1.145-3.874; P = 0.017), other histologic type (HR 3.547, 95% CI 1.205-10.444; P = 0.022), initial locally advanced clinical stage (HR 2.406, CI 1.270-4.555; P = 0.003) and non-pathologic complete remission (HR 8.204, 95% CI 1.810-22.112; P = 0.047) were significantly
associated with the OS (Table 3). The \(ABCB1\) 3435TT genotype was also associated with a lower risk of death with marginal significance not shown to be an independent prognostic factor (HR 0.245, 95% CI = 0.059–1.026; \(P = 0.054\)). The result was similar when breast cancer staging was adjusted instead of operable staging (HR 0.238; 95% CI, 0.057–0.999; \(P = 0.050\)).

Toxicities. Toxicities according to the NCI-CTC version 3.0 are summarized in Table 4. One hundred seventeen patients (54.2%) of the studied patients suffered one or more adverse event of any grade. Overall, Grade 3–4 toxicities were observed in 34 patients (15.7%). Grade 3–4 neutropenia developed in 21 patients (9.6%) with 9 (4.2%) febrile episodes, as grade 3–4 diarrhea did in 12 patients (5.6%). No treatment-related mortality was encountered in the study. TT genotype of \(ABCB1\) C3435T was associated with higher frequency of grade 3–4 hematologic and non-hematologic toxicities including neutropenia (\(P = 0.037\)) and diarrhea (\(P = 0.017\)) compared with CC/CT genotype (Table 5).

Pharmacokinetics and \(ABCB1\) and \(CYP3A5\) genotypes. The \(ABCB1\) 3435TT genotype showed a higher AUC\(_\text{doc}\) than the CC/CT genotype with statistical significance (\(P = 0.031\)) (Fig. 2). However, the \(ABCB1\) G2677T/A or C1236T genotypes showed no association with AUC\(_\text{doc}\) or AUC\(_\text{dox}\). Among the genotypes of \(CYP3A5\), AA (⁄)*1/*1)/AG/AA/*3/+ genotypes had a higher AUC of docetaxel than the GG (⁄)*3/*3) genotype with statistical significance (\(P = 0.024\)). The \(ABCB1\) 3435TT genotype was correlated with neutropenia (\(P = 0.039\)), febrile neutropenia (\(P = 0.218\)), and diarrhea (\(P = 0.057\)). \(CYP3A5\) polymorphism did not affect survival and toxicities.

Discussion

In recent years, various genetic variants in \(ABCB1\) have been described to affect the transporter expression and function.\(^{31}\) Docetaxel and doxorubicin are subjected to transport by the \(ABCB1\) transporter. \(ABCB1\) 3435C (glutamate) to T (glutamate) substitution does not result in an amino acid sequence change nor is this gene located in a regulatory region.\(^{30}\) This synonymous \(ABCB1\) C3435T polymorphism has been associated with mRNA instability and a lower expression of P-gp in the duodenum.\(^{27}\) P-gp expression and \(ABCB1\) C3435T polymorphism have been identified as an independent factor for RFS and RR for chemotherapy in breast cancer.\(^{11,33}\) Hoffmeyer et al. reported that the TT genotype of C3435T was correlated with reduced expression of P-gp and, therefore, reduced cellular elimination and maintained higher plasma concentrations of chemotherapeutic drugs.\(^{27,32}\) From these results, we hypothesize that patients who have the \(ABCB1\) 3435TT genotype might show a better treatment response to docetaxel and doxorubicin chemotherapy and get survival benefit compared to \(ABCB1\) 3435 CC/CT.

This study was designed to identify the clinical impact of \(ABCB1\) SNPs in Korean patients with breast cancer who were treated with docetaxel and doxorubicin, a regimen widely accepted as neoadjuvant chemotherapeutic agents.\(^{21}\) In accordance with previous studies, this study showed that the TT genotype of \(ABCB1\) 3435 was associated with a longer OS and higher plasma concentration of docetaxel than CT/TT.\(^{33,34}\) It also suggests that a higher plasma level of docetaxel was maintained for a longer time in the TT genotype of \(ABCB1\) translating into more intense exposure to docetaxel. As hematopoietic stem cells and intestinal epithelial cells are reported to express P-gp,\(^{35}\) higher drug concentrations in homozygote 3435TT leads to more frequent grade 3–4 neutropenia, febrile neutropenia, and diarrhea. Since the number of patients in the pharmacokinetic group was small, the OS tended to be longer in the \(ABCB1\) 3435TT genotype without statistical significance. There was a trend toward a higher percentage of grade 3–4 neutropenia in the \(ABCB1\) 3435TT subjects in the PK group (28.6% versus about 3.1% for the CC/CT subjects). It provides valuable information to clinicians that the \(ABCB1\) homozygote carriers have increased efficacy and increased toxicity simultaneously. There were no significant differences in the OS at the analyzed data of G2677T/A and C1236T. To our knowledge, this is the first study demonstrating that \(ABCB1\) C3435T polymorphism is significantly associated with plasma docetaxel concentration and OS after neoadjuvant docetaxel/doxorubicin combination chemotherapy in Asian patients with stage II or III breast cancer.

Since liver metabolism of docetaxel to inactive hydroxylated metabolites is mediated by \(CYP3A4\) and \(CYP3A5\),\(^{12,36}\) polymorphisms of such enzymes could affect the survival outcome of the treatment in breast cancer patients. Intuitively, it is reasonable to infer that the side effects of docetaxel treatment could be related to those metabolic enzymes. The most common variant, \(CYP3A4^*1B\) allele founded in the African-American population, was not detected in the present study as expected based on previously published data.\(^{37}\) In contrast to \(CYP3A4^*1B\), the frequency of \(CYP3A5^*3\) allele was 65–85% in Asia\(^{18}\) and 76.8% in our study. The \(CYP3A5\) is linked to a common transition in intron 3 of the \(CYP3A5\) gene (\(CYP3A5^*3\)) which introduces a frameshift during translation and results in a truncated, nonfunctional protein.\(^{38}\) At this point, homozygote (GG genotype,\(^*3/\*3\)) for \(CYP3A5^*3\) allele would be expected to be also strongly associated with low \(CYP3A5\)
protein content and AUC in a homozygote for the CYP3A5*3 allele which would be higher than CYP3A5*1. However, our findings show somewhat conflicting results: the homozygote of CYP3A5*3 tend to have a lower docetaxel concentration and a higher risk of grade 3 diarrhea than the CYP3A5*1/*1 genotype (P = 0.072). Concordantly, Goh et al. reported the same result that docetaxel clearance in patients with at least one CYP3A5*1 allele was significantly lower than CYP3A5*3 homozygotes. What causes this difference in predictions is as follows: the AA genotype (CYP3A5*1/*1) of CYP3A5 was only in three patients (4.2%) and was correlated with the TT genotype of C3435T with statistical significance. So a high docetaxel AUC revealed in the ABCB1 C3435T could be associated with the AA genotype of CYP3A5. Although CYP3A5 can account for >50% of the total CYP3A, we speculate that docetaxel metabolism was affected by the presence of the CYP3A5*3 allele to a small degree.

One of the merits of this study is the homogeneity of the population who received the same neoadjuvant chemotherapy followed by surgery for stage II/III breast cancer. Second, the study was designed prospectively; pharmacogenomics samples and plasma drug concentration samples were collected at specific time points according to the written prospective protocol in the chemotherapy-naive patients. The clinical data were gained through the same therapeutic scheme. With all of these advantages, the major limitation of this study is the relatively small sample size of the pharmacokinetic study population.

However, the distribution of genotype in this study was well matched to what has been reported previously in Koreans, and in a line with those with Asian population. In a sense, we believe the small sample size of this study did not led to selection bias, and thus does not undermine the significance of the findings.

In conclusion, this study shows that the genetic polymorphism of ABCB1 C3435T is associated with higher docetaxel concentration, a longer OS and more frequent toxicities. Our results suggest the future of guided drug selection by predicting drug toxicities according to the patient’s genotype. Further prospective trials with large scale as well as in vitro and in vivo functional studies are warranted.

Acknowledgments

This research was supported by a grant from Basic Science Research Program (grant number : 2010-0022299) through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology and a grant of the Korea Health Technology R&D Project through the Korea Health Industry Development Institute (KHIDI), funded by the Ministry of Health & Welfare, Republic of Korea (grant number : HI14C1277).

Disclosure Statement

The authors have no conflicts of interest.

References

1. Fisher B, Brown A, Mamounas E et al. Effect of preoperative chemotherapy on local-regional disease in women with operable breast cancer: findings from National Surgical Adjuvant Breast and Bowl Protocol B-18. J Clin Oncol 1997; 15: 2483-93.
2. Rastogi P, Anderson SJ, Bear HD et al. Preoperative chemotherapy: updates of National Surgical Adjuvant Breast and Bowl Protocol B-18 and B-27. J Clin Oncol 2008; 26: 778-85.
3. Hontobaygi GN, Spanswick W, Montague ED, Buzdar AU, Yap HY, Blumenschein GR. Treatment of locoregionally advanced breast cancer with surgery, radiotherapy, and combination chemotherapy+maintenance. Int J Radiat Oncol Biol Phys 1983; 9: 643–50.
4. Kuerer HM, Sahn AA, Hunt KK et al. Incidence and impact of documented eradication of breast cancer axillary lymph node metastases before surgery in patients treated with neoadjuvant chemotherapy. Ann Surg 1999; 230:72–8.
5. Fisher B, Mamounas EP. Preoperative chemotherapy: a model for studying the biology and therapy of primary breast cancer. J Clin Oncol 1995; 13: 537–46.
6. van Zuylen L, Verweij J, Nooter K, Brouwer G, Spijkerboomen A. Role of intestinal P-glycoprotein in the plasma and fecal disposition of docetaxel in humans. Clin Cancer Res 2000; 6: 2598–603.
7. Trock BJ, Leonesa F, Clarke R. Multidrug resistance in breast cancer: a meta-analysis of MDR1/gp170 expression and its possible functional significance. J Natl Cancer Inst 1997; 89: 917–31.
8. Bosch TM, Huitema AD, Doodeman VD et al. Pharmacogenetic screening of CYP3A and ABCB1 in relation to population pharmacokinetics of docetaxel. Clin Cancer Res 2006; 12: 5786–93.
9. Green H, Soderkvist P, Hovarth G, Peterson C. mdr1 single nucleotide polymorphisms in ovarian cancer tissue: G2677T/A correlates with response to paclitaxel chemotherapy. Clin Cancer Res 2006; 12: 854–9.
10. Kafka A, Sauer G, Jaeger C et al. Polymorphism C3435T of the MDR-1 gene predicts response to preoperative chemotherapy in locally advanced breast cancer. Int J Oncol 2003; 22: 1117–21.
11. Tatsuta M, Minami Y, Fujita H et al. Expression of P-glycoprotein in human placenta: relation to genetic polymorphism of the multidrug resistance (MDR-1) gene. J Pharmacol Exp Ther 2001; 297: 1137–43.
12. Tran A, Jullien V, Alexandre J et al. Pharmacokinetics and toxicity of docetaxel: role of CYP3A, MDR1, and GST polymorphisms. Clin Pharmacol Ther 2002; 71: 505–9.
13. Shou M, Martinet M, Korzekwa KR, Krausz KW, Gonzalez FJ, Gelboin HV. Role of human cytochrome P450 3A4 and 3A5 in the metabolism of taxotere and its derivatives: enzyme specificity, interindividual distribution and metabolic contribution in human liver. Pharmacogenetics 1998; 8: 391–401.
14. Ball SE, Scatina J, Kao J et al. Population distribution and effects on drug metabolism of a genetic variant in the 5’ promoter region of CYP3A4. Clin Pharmacol Ther 1999; 66: 288–94.
15. Rebbeck TR, Jaffe JM, Walker AH, Wein AJ, Malkowicz SB. Modification of clinical presentation of prostate tumors by a novel genetic variant in CYP3A4. J Natl Cancer Inst 1998; 90: 1225–5.
16. Hsieh RP, Lin YY, Cheng CL et al. Novel mutations of CYP3A4 in Chinese. Drug Metab Dispos 2001; 29: 268–73.
17. Walker AH, Jaffe JM, Gunasegaran S et al. Characterization of an allelic variant in the nifedipine-specific element of CYP3A4: ethnic distribution and implications for prostate cancer risk. Mutations in brief no. 191. Online. Hum Mutat 1998; 12: 289.
18. Kuehl P, Zhang J, Lin Y et al. Sequence diversity in CYP3A promoters and characterization of the genetic basis of polymorphic CYP3A5 expression. Nat Genet 2001; 27: 383–91.
19. Goh BC, Lee SC, Wang LZ et al. Explaining interindividual variability of docetaxel pharmacokinetics and pharmacodynamics in Asians through phenotyping and genotyping strategies. J Clin Oncol 2002; 20: 3683–90.
20. Lan LB, Dalton J, Schuetz EG. Mdr1 limits CYP3A metabolism in vivo. Mol Pharmacol 2000; 58: 863–9.
21. Han S, Kim SB, Kang SS et al. A phase II study of neoadjuvant docetaxel plus doxorubicin (KBCS-01) in stage II, III breast cancer. Breast Cancer Res Treat 2006; 98: 57–61.
22. Keam B, Im SA, Kim HJ et al. Prognostic impact of clinicopathologic parameters in stage II/III breast cancer treated with neoadjuvant docetaxel and doxorubicin chemotherapy: paradoxical features of the triple negative breast cancer. BMC Cancer 2007; 7: 203.
23. Recht A, Edge SB, Solin LJ et al. Postmastectomy radiotherapy: clinical practice guidelines of the American Society of Clinical Oncology. J Clin Oncol 2001; 19: 1539–69.
24. Kong M, Hong SE. Which patients might benefit from postmastectomy radiotherapy in breast cancer patients with T1–2 tumor and 1–3 axillary lymph nodes metastasis? Cancer Res Treat 2013; 45: 103–11.
25. Therasse P, Arbuck SG, Eisenhauer EA et al. New guidelines to evaluate the response of cancer therapy in solid tumors. J Natl Cancer Inst 2000; 92: 205–16.
27 Hoffmeyer S, Burk O, von Richter O et al. Functional polymorphisms of the human multidrug-resistance gene: multiple sequence variations and correlation of one allele with P-glycoprotein expression and activity in vivo. Proc Natl Acad Sci USA 2000; 97: 3473–8.

28 Tang K, Ngoi SM, Gwee PC et al. Distinct haplotype profiles and strong linkage disequilibrium at the MDR1 multidrug transporter gene locus in three ethnic Asian populations. Pharmacogenetics 2002; 12: 437–50.

29 Yamaguchi H, Hishinuma T, Endo N et al. Genetic variation in ABCB1 influences paclitaxel pharmacokinetics in Japanese patients with ovarian cancer. Int J Gynecol Cancer 2006; 16: 979–85.

30 Wang LZ, Goh BC, Grigg ME, Lee SC, Khoo YM, Lee HS. A rapid and sensitive liquid chromatography/tandem mass spectrometry method for determination of docetaxel in human plasma. Rapid Commun Mass Spectrom 2003; 17: 1548–52.

31 Sparreboom A, Danesi R, Ando Y, Chan J, Figg WD. Pharmacogenomics of ABC transporters and its role in cancer chemotherapy. Drug Resist Updat 2003; 6: 71–84.

32 Kim RB, Leake BF, Choo EF et al. Identification of functionally variant MDR1 alleles among European Americans and African Americans. Clin Pharmacol Ther 2001; 70: 189–99.

Supporting Information

Additional supporting information may be found in the online version of this article:

Fig. S1. Kaplan–Meier survival analysis according to G2677TA and C1236T polymorphisms of the ABCB1 gene.