Long Non-coding RNAs in Tuberculosis: From Immunity to Biomarkers

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Tuberculosis (TB) caused by Mycobacterium tuberculosis (Mtb) is the leading lethal infectious disease with 1.3 million deaths in 2020. Despite significant advances have been made in detection techniques and therapeutic approaches for tuberculosis, no suitable diagnostic tools are available for early and precise screening. Many studies have reported that Long non-coding RNAs (lncRNAs) play a regulatory role in gene expression in the host immune response against Mtb. Dysregulation of lncRNAs expression patterns associated with immunoregulatory pathways arose in mycobacterial infection. Meanwhile, host-induced lncRNAs regulate antibacterial processes such as apoptosis and autophagy to limit bacterial proliferation. In this review, we try to summarize the latest reports on how dysregulated expressed lncRNAs influence host immune response in tuberculosis infection. We also discuss their potential clinical prospects for tuberculosis diagnosis and development as molecular biomarkers.

Keywords: tuberculosis, lncRNA, immune response, diagnosis, biomarker

INTRODUCTION

LncRNAs, frequently defined as non-protein-coding transcripts greater than 200 nucleotides in length (McFadden and Hargrove, 2016), were initially thought to be "transcriptional noise" (Ulitsky and Bartel, 2013). Recent studies have unraveled the biogenesis of lncRNAs, which are distinct from that of mRNAs. Those lncRNAs, generally display lower sequence conservation (Pang et al., 2006) and contain fewer exons (Washietl et al., 2014; Guo et al., 2020), and their functions are typically linked to their specific subcellular localizations (Statello et al., 2021). In addition, lncRNAs have been reported to include small open reading frames (SMORFs) encoding short functional peptides (Kim et al., 2014; Huang et al., 2017; Fathizadeh et al., 2020; Zhu et al., 2020). Accumulated evidence demonstrates that they are emerging as essential regulators in gene regulation, such as chromatin modification, mRNA stability, transcriptional, translation and post-translation activation/inhibition (Quinn and Chang, 2016; Yao et al., 2019). Meanwhile, lncRNAs have various regulatory functions in biological processes, including apoptosis (Heydarnezhad Asl et al., 2022).

Tuberculosis, one of the leading lethal infectious diseases worldwide caused by Mtb, has been considered a public health emergency that concerns the world. According to the WHO global TB report, there were 5.8 million newly sick and 1.5 million TB-related deaths in 2021 (WHO, 2021, no date). Approximately one-third of the world’s population has a latent TB infection (LTBI), and about 10% can develop active tuberculosis (ATB) with impairment in the immune system (Pai et al., 2016). In the host-pathogen interactions, intracellular survival and replication of Mtb after macrophage phagocytosis leads to an immune response that converges on granuloma formation...
and/or disease (McCaffrey et al., 2022; Medley et al., 2022). The host cells have adopted a series of clearance mechanisms to facilitate intracellular bacterial killing, such as apoptosis, autophagy, inflammation, and macrophage polarization. Mtb has also evolved with a set of almost perfect immune escape mechanisms to evade the host immune system (Stanley and Cox, 2013). Growing evidence has delineated that the expression of many lncRNAs is involved in TB with a definitive role in orchestrating the biological processes from immune response to host-pathogen interactions. In addition, tissue-specific and condition-specific expression patterns suggest that lncRNAs provide potential biomarkers (Statello et al., 2021). Therefore, in-depth elucidation of the effect and mechanism of lncRNAs may develop clinical prospects for precise TB diagnosis and treatments (Lyu et al., 2021; Wei et al., 2021). In this paper, we review the progress of lncRNA roles in Mtb infection from host immunity to biomarkers and discuss its potential in clinical diagnosis.

REGULATORY MECHANISMS OF LONG NON-CODING RNAs

Depending on specific interactions with DNA, RNA, and proteins, lncRNAs can modulate diverse biological processes through complex and diverse mechanisms, such as execute-as signals, decoys, guides, scaffolds to regulate target genes (Wang and Chang, 2011; Kazemzadeh et al., 2015; Statello et al., 2021).

DNA Level Regulation

A feature of lncRNAs is that they can generate a hybrid structure with DNA to influence gene expression. They mediate DNA methylation and transcriptional inhibition by complementary pairing with unstranded DNA bases. In addition, lncRNAs can inhibit the recruitment of Pol II or alter the binding of transcription factors at the promoter sequences whose function can inhibit the recruitment of Pol II or alter the binding of transcription factors (O'Leary et al., 2015). It as a recruitment platform for gene-silencing machinery (MAT2A) via methylation in response to irradiation, implicating it as a recruitment platform for gene-silencing machinery. Also, RNA–DNA complex formation has been proposed as an example of lncRNA–DNA interplay in mediating gene silencing or activation (Monod et al., 2015; O’Leary et al., 2015). For instance, lncRNA PARTICLE, forming a DNA-lncRNA triplex, repress the tumor suppressor methionine adenosyltransferase (MAT2A) via methylation in response to irradiation, implicating it as a recruitment platform for gene-silencing machinery (O’Leary et al., 2015).

RNA Level Regulation

At the RNA level, lncRNAs often function as lncRNA-miRNA-mRNA competing endogenous RNAs (ceRNAs) that serve as miRNA sponges and inhibit their regulatory effect on the target gene (Figure 1B; Salmena et al., 2011; Wang M. et al., 2019; Wang et al., 2020). For example, a ceRNA network constructed from pulmonary tuberculosis (PTB) patients suggested that lncRNAs regulate mRNAs expression may mediate by acting as sponged miRNAs (Zhang et al., 2020). Furthermore, severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), pandemic H1N1, and H7N9 infection induced upregulation of lncRNA-34087.27, which could serve as ceRNA, competitively binding with miR-302b-3p to stabilize IRF1 mRNA (Yang et al., 2022). Additionally, mRNA splicing is one of the biological functions that can be affected by lncRNAs through forming complementary double strands with transcripts (Figure 1B; De Troyer et al., 2020, p. 3). For instance, lncRNA MALAT1 can modulate the levels of serine/arginine (SR) proteins, thereby regulating the alternative splicing (AS) of pre-mRNA (Tripathi et al., 2010). Moreover, with lncRNA-mRNA dimer formation, lncRNAs can also inhibit RNase degradation and alter the stability and translation of cytoplasmic mRNAs (Figure 1B; Statello et al., 2021).

Protein Level Regulation

Numerous lncRNAs localize on chromatin, interacting with proteins to facilitate or inhibit their binding and activity at targeted DNA regions. On the one hand, it can participate in protein phosphorylation by protein kinase or ubiquitin modification of a protein by ubiquitin-modified enzymes (Statello et al., 2021). For instance, HOTAIR is known to act at the posttranslational level by serving as an assembly scaffold for protein ubiquitination (Figure 1C). In addition, lncRNAs could regulate histone modification and chromatin accessibility, thereby regulating transcription (Figure 1C). A well-described example is the X chromosome dosage compensation process. The X-inactive-specific transcript (Xist) recruits polycomb repressive complex 2 (PRC2) and triggers large numbers of histones methylated as well as a cascade of events that entails chromosome remodeling to achieve stable silencing (Creamer and Lawrence, 2017; Iégú et al., 2017). Similarly, HOTAIR interacts with PRC2 favors epigenetic silencing through Enhance of Zeste2 (EZH2) catalyzed deposition of H3K27me3, facilitating the survival of virulent Mtb (Table 1; Subuddhi et al., 2020). Moreover, lncRNAs can be used as a protein structural component to affect protein spatial conformation.

THE ROLE OF LONG NON-CODING RNAs IN ANTI-TUBERCULOSIS IMMUNITY

Different expression patterns of lncRNAs are emerging as critical regulators to control the function of innate and adaptive immune cell types and initiate effective defense mechanisms in TB (Figure 2 and Table 1; Chen Y. G. et al., 2017; Wei et al., 2021).

The Role of Long Non-coding RNAs in Macrophages

Accumulating lines of evidence have revealed that lncRNAs play an important role in regulating the innate immune response of
FIGURE 1 | Schematic overview of lncRNAs’ molecular mechanisms. (A) lncRNAs regulate downstream gene transcription by binding to transcription factors, repression of RNA Pol II, complementary pairing with unstranded DNA bases, and DNA methylation. (B) lncRNAs serve as miRNA sponges to competitively bind with miRNA. lncRNAs regulate mRNA transcription by modulating mRNA stability and pre-mRNA splicing. lncRNAs can inhibit RNase degradation and promote mRNA stability. (C) lncRNAs can serve as a scaffold for protein phosphorylation and ubiquitination and regulate chromatin modification by employing histone-modifying complex.

host macrophages, which are the sentinels to phagocytose and eliminate Mtb (Figure 2A,B). For one thing, lncRNAs participate in macrophage-mediated immune-inflammatory responses that an inducible program of inflammatory gene expression is central to anti-microbial defenses. For example, lincRNA-Cox2 mediates the activation/repression of immune genes that may activate NF-κB and STAT3 to regulate inflammatory responses for resistance of Mtb infection (Figure 2A; Carpenter et al., 2013; Li D. et al., 2020). In addition, lncRNA-PACER (also known as lncRNA-Cox2) was induced in Mtb infected macrophages, acting as a positive regulator of its proximal pro-inflammatory gene PtgS-2 (also known as Cox2). Via mechanisms involving the sequestration of repressive NF-κB subunit p50 away from PtgS-2 promoter (Krawczyk and Emerson, 2014; Tamguez et al., 2021). LncRNA-Cox2 can promote the activation of macrophages toward the pro-inflammatory M1 phenotype known to be efficient in killing Mtb (Figure 2A; Ye et al., 2018; Tamguez et al., 2021). Recently, some studies have found that in Mtb infected macrophages, the down-regulated lncRNA GAS5 might facilitate the cell vitality and the inflammatory response by sponging miR-18a-5p (Figure 2A; Li et al., 2021), and NEAT1 participates in inflammatory response through targeted regulation of miR-377-3p (Figure 2A; Sun et al., 2021).

For another, previous studies indicated that massive lncRNAs could regulate Mtb-induced apoptosis and autophagy of macrophages, playing a vital role in the pathogenesis of TB (Behar et al., 2011; Kim et al., 2019). For example, the Bacillus Calmette-Guerin (BCG)-infected macrophages induced apoptosis with upregulated lincRNA-Cox2 expression. Knockdown of Cox2 aggravated reactive oxygen species (ROS) accumulation and initiated apoptosis by activating the PERK-eIF-2α-CHOP signaling pathway (Figure 2B; Xu et al., 2021). A similar mechanism has been described for lncRNA-EPS, modulating apoptosis and autophagy by activating the JNK/MAPK signaling pathway in macrophages (Figure 2B; Ke et al., 2020). Additionally, down-regulation of lncRNA-MEG3 in infected macrophages induced autophagy and enhanced eradication of intracellular Mycobacterium Bovis BCG (Pawar et al., 2016). Recent studies have revealed that several lncRNAs act as ceRNAs to regulate apoptosis and autophagy. For example, lncRNA PCED1B-AS1 decreased in ATB patients compared to healthy individuals. The suppression of PCED1B-AS1 significantly attenuated apoptosis and enhanced autophagy in macrophages by sponging the miR-155 and inhibiting its targets FOXO3/Rheb (Figure 2B; Li et al., 2019). Moreover, lncRNA MIAT modulated macrophage...
TABLE 1 | The regulatory role of lncRNAs in anti-TB immunity.

| LncRNAs          | Expression lever | Targets                             | Sample         | Mtb strains   | Effect                                                                                           | References        |
|------------------|------------------|-------------------------------------|----------------|---------------|-------------------------------------------------------------------------------------------------|-------------------|
| NEAT1            | Up               | miR-377-3p                          | Macrophages    | H37Ra         | Promote the expression of IL-6 and inhibit apoptosis                                             | Sun et al., 2021  |
| HOTAIR           | Down             | EZH2                                | Macrophages    | H37Rv         | Promote the SATB1 and DUSP4 transcript and inhibit the production of ROS                         | Subuddhi et al., 2020 |
| Cox2             | Up               | NF-κB and Stat3                     | Macrophages    | H37Ra         | Activate NF-κB and Stat3 to regulate inflammatory responses                                        | Li D. et al., 2020 |
| Cox2             | Up               | PERK/eIF2α/CHOP                      | Monocyte-Derived Macrophages | BCG | Inhibit apoptosis and promote autophagy                                                             | Xu et al., 2021    |
| PACER            | Up               | Ptgs-2                              | Monocyte-Derived Macrophages | HN878 (clinical hypervirulent strain) | Promote ptgs2 transcription.                                                                        | Tamgue et al., 2021 |
| GAS5             | Down             | miR-18a-5p                          | Macrophages    | H37Rv         | Promote cell viability and inflammatory response                                                  | Li et al., 2021    |
| MEG3             | Down             | Not known                           | Macrophages    | M. bovis BCG  | Promote autophagy                                                                                  | Pawar et al., 2016 |
| EPS              | Down             | JNK/MAPK                            | Macrophages    | BCG           | Attenuate apoptosis and promote autophagy                                                          | Ke et al., 2020    |
| PCED1B-AS1       | Down             | miR-155                             | Macrophages    | /             | Attenuate apoptosis and promote autophagy                                                          | Li et al., 2019    |
| MIAT             | Up               | miR-665                             | Macrophages    | BCG           | Attenuate autophagy and promote apoptosis                                                          | Jiang et al., 2021 |
| CD244            | Up               | EZH2                                | CD8(+) T cells | /             | Inhibit IFN-γ/ TNF-α expression and promote Mtb proliferation                                      | Wang et al., 2015  |
| Lnc AC145676.2.1-6 | Down             | miR-29a                             | Whole blood    | /             | Interference with cytokine-cytokine receptor interactions and TLR signaling path ways.           | Bai et al., 2019  |
| Lnc-TGS1-1       | Down             | MIR-143                             | Whole blood    | /             | Thrombocytopenia and interference with the TLR signaling                                           | Bai et al., 2019  |

apoptosis and autophagy upon BCG infection through the miR-665/ULK1 crosstalk (Figure 2A; Jiang et al., 2021).

The Role of Long Non-coding RNAs in T Cells

Once the innate immune system is breached, the "human guardians," including T and B cells, immediately enter a fighting state and initiate adaptive immunity. LncRNAs also regulate T cell-mediated immune regulation in TB (Figure 2C). CD4 + T cells play a dominant role in the host immune response of TB (Jasenosky et al., 2015). The study analyzed the lncRNA profile in CD4 + T cells and revealed that compared with healthy controls, lncRNAs showed abnormal expression in ATB and LTBI (Yi et al., 2014). In addition, significantly enriched signaling pathways based on deregulated mRNAs were cytokine-cytokine receptor interaction, mitogen-activated protein kinase (MAPK), and TLR signaling pathway (Yi et al., 2014).

In general, CD8 + T cells were previously thought to be less critical than CD4 + T cells in the immune response to TB. It has been newly stated its non-redundant role with a specific CD8 + T cell response (Lin and Flynn, 2015). A recent study confirmed that lncRNA-CD244 epigenetically repressed the IFN-γ and TNF-α expression in CD8 + T cells (Figure 2C; Wang et al., 2015). Adoptive transfer of CD244–depressed CD8 + T cells to Mtb-infected mice reduced infection and pathology compared to mice transplanted with wild-type CD8 + cells (Wang et al., 2015). Moreover, Heme Oxygenase 1 (HMOX1) was increased after Mtb infection and can distinguish LTBI from ATB in the previous research (Costa et al., 2016). Also, 328 differentially expressed lncRNAs were found in CD8 + T cells’ response to ATB. Among them, lncRNA XLOC_014219 was upregulated, while its nearby protein-coding gene HMOX1 was significantly decreased. It is essential to uncover why HMOX1 is downregulated and whether lncRNA XLOC_014219 relates to it, which is ultimately involved in the dysfunction of CD8 + T cells (Fu et al., 2017a).

The Role of Long Non-coding RNAs in B Cells

It has come to light that lncRNAs can influence antibodies produced by B cells and impact B cell biology by regulating survival signals during activation (Zeni and Mraz, 2021). For instance, a study reported that 844 lncRNAs differentially expressed in B cell samples. Additionally, SOCS3 is an essential
negative regulator of cytokine response to Mtb infection, and its upstream lncRNA XLOC_012582 highly increased in B cells of ATB. Whether upregulation of XLOC_012582 leads to overexpression of SOCS3 and ultimately participates in the progression of TB needs in-depth investigation (Fu et al., 2017b). Findings provided new insight into the pathogenesis of TB. However, discoveries related to B cells are like a tip of an iceberg.

In summary, the immune protection mechanism of TB is complex and comprehensive. Although there have been
preliminary studies on the role of lncRNAs in different host cells to Mtb, their specific functions in immunity are mainly unexplored.

**LONG NON-CODING RNAs AS DIAGNOSIS BIOMARKERS IN TUBERCULOSIS**

So far, most clinical diagnosis methods have inherent limitations (Pai et al., 2016). Methods like smear microscopy and mycobacterium culture have insufficient sensitivity and timeliness and have a poor detection rate of smear-negative PTB (Walzl et al., 2011; Fang et al., 2021; Mirzaei et al., 2021), while interferon-gamma release assays (IGRA) unable to discriminate between LTBI and ATB (Walzl et al., 2018). The recent recommendations of the WHO include non-pathogen-based detection to improve the identification of clinically diagnosed TB with rapid and universal methods (Martinez and Andrews, 2019). Therefore, biomarkers of the host immune responses might provide critical insights to solve this problem. Some ncRNAs are considered to be biomarkers (Beermann et al., 2016), such as miR-889 targets that can be manipulated for antimycobacterial therapeutic purposes and candidate biomarkers for LTBI (Chen et al., 2020). Relative to investigated in high detail miRNAs, the roles of lncRNAs remain largely elusive (Lee, 2012). Nevertheless, a rising number of dysregulation processes indicate that lncRNAs are highly promising as biomarkers of TB (Table 2).

### Biomarkers in Peripheral Blood Mononuclear Cells

Accumulating lines of studies have revealed the abnormally expressed lncRNAs in peripheral blood mononuclear cells (PBMCs) of TB patients (Table 2). A study reported that two significantly aberrantly expressed lncRNA (MIR3945HG V1 and MIR3945HG V2) in PBMCs samples from active PTB patients have the potential to be novel diagnostic biomarkers (Yang et al., 2016). Furthermore, the expression of NEAT1 (both NEAT1_1 and NEAT1_2) in TB patients was higher than healthy control, declined gradually with treatment, and was restored to the normal level. This dynamic change could reflect the efficacy of anti-TB therapy. Therefore, NEAT1 may serve as a potential indicator for patient prognosis of TB (Huang et al., 2018). Additionally, the study suggested that downregulated PCED1B-AS1 in PBMCs and THP-1 show promise as a new early diagnostic biomarker for ATB (Li et al., 2019). Nevertheless, little information has been done on the underlying mechanisms of lncRNAs above. To enhance the PTB identification, lncRNA n344917 was confirmed down-regulated in PBMC of PTB. Hence, a web-based prediction model combining the molecular biomarker n344917, laboratory, and EHR variables was constructed and could serve as a user-friendly, accurate platform to improve the clinical diagnosis of PTB (Meng et al., 2021).

### Biomarkers in Plasma or Serum

Differentially expressed lncRNAs in the plasma or serum of TB patients have also been explored as potential diagnostic biomarkers (Table 2). For instance, expression levels of ENST0000354432 and ENST0000427151 were suggested to act as biomarkers for the early detection of TB (He et al., 2017). LOC152742 in plasma had higher specificity in ATB and gradually downregulated in the treatment. Hence it could serve as a novel biomarker for the diagnosis and therapy of ATB (Wang L. et al., 2019). Also, plasma lncRNAs might act as potential biomarkers to evaluate TB cure in an efficient and precise manner. lncRNAs uc.48 + and NR_105053 may serve as biomarkers to distinguish between untreated TB patients and cured TB subjects (Li Z.-B. et al., 2020). Recently, studies reported some lncRNA sets with high diagnostic sensitivity and specificity. For example, ceRNA analysis of four differentially expressed lncRNAs (NR_038221, NR_003142, ENST0000570366, and ENST0000422183) was demonstrated to discriminate PTB from healthy individuals. The results showed that NR_038221 was the most significantly associated with TB (Chen Z.-L. et al., 2017). Their previous study had verified hsa-miR-378a-3p as a potential biomarker for PTB, which was associated with NR_038221, indicating that NR_038221 and hsa-miR-378a-3p might play a similar function during the biological process of PTB (Zhang et al., 2013; Chen Z.-L. et al., 2017). At present, the development of high-throughput experimental technologies has led to a rapid expansion of lncRNA research. For example, an integrated analysis of the GEO dataset and the NONCODE database identified four significantly downregulated lncRNAs (NON-HSAT101518.2, NON-HSAT067134.2, NON-HSAT148822.1, and NON-HSAT078957.2) in serum exosomes of ATB patients. ROC curve analysis suggests that these four lncRNAs can discriminate ATB from healthy individuals with high specificity and sensitivity (Fang et al., 2021).

### Biomarkers in Bacteria-Negative Tuberculosis

Accurate diagnosis of complete inactivation of TB lesions remains a challenge concerning sputum-negative TB, one of the significant factors for the development and spread of ATB (Horsburgh, 2004). Clinically diagnosed PTB patients without microbiological evidence of Mtb often lead to misdiagnosis or delayed diagnosis. Therefore, A study validated the lncRNAs and corresponding predictive models to effectively diagnose these patients, finding differentially expressed lncRNAs (ENST0000497872, n333737, n335265, NR_038221, indicating that NR_038221 and hsa-miR-378a-3p might play a similar function during the biological process of PTB (Zhang et al., 2013; Chen Z.-L. et al., 2017). At present, the development of high-throughput experimental technologies has led to a rapid expansion of lncRNA research. For example, an integrated analysis of the GEO dataset and the NONCODE database identified four significantly downregulated lncRNAs (NON-HSAT101518.2, NON-HSAT067134.2, NON-HSAT148822.1, and NON-HSAT078957.2) in serum exosomes of ATB patients. ROC curve analysis suggests that these four lncRNAs can discriminate ATB from healthy individuals with high specificity and sensitivity (Fang et al., 2021).
### TABLE 2 | Overview of the candidate IncRNA biomarkers in TB.

| Analysis | Candidates of IncRNA biomarkers | Sample | Effect | References |
|----------|---------------------------------|--------|--------|------------|
| PBMCs    | Upregulated (MIR3945HG V1 and MIR3945HG V2) in PTB patients | Active PTB patients and healthy donors vaccinated with BCG | Promising candidate diagnostic markers for TB | Yang et al., 2016 |
| PBMCs    | NEAT1 (both NEAT1_1 and NEAT1_2) declined gradually with treatment | TB patients and healthy group | Potential indicator for patient prognosis of TB | Huang et al., 2018 |
| PBMCs    | Downregulated (PCED1B-AS1) in ATB patients | ATB patients and healthy individuals | May represent a novel early diagnostic marker of ATB | Li et al., 2019 |
| PBMCs    | Downregulated (n344917) in clinically diagnosed PTB | clinically diagnosed PTB, PTB with an etiological evidence and non-TB disease controls | Potential molecular biomarker for the clinically diagnosed PTB | Meng et al., 2021 |
| Plasma   | Upregulated (ENST00000354432, ENST00000427151) in TB patients | TB patients, community acquired pneumonia and healthy individuals | Potential molecular biomarkers for the rapid diagnosis of TB | He et al., 2017 |
| Sputum and plasma | Upregulated LOC152742 in ATB patients | ATB patients, obselete TB patients, individuals affected with BCG, and normal individuals, | Potential biomarker for diagnosis and therapy of ATB | Wang L. et al., 2019 |
| Plasma   | Differently expressed (IncRNAs uc.48 + and NR_105053) between the untreated and the cured TB | Untreated TB and cured TB subjects | Potential biomarkers to distinguish between untreated and cured TB, provide an experimental basis to evaluate the effect of TB treatment | Li Z.-B. et al., 2020 |
| Plasma   | Upregulated (NR_038221, NR_003142, ENST00000570366), downregulated (ENST00000422183) | ATB patients and healthy control | Potential biomarkers for early diagnosis of TB | Chen Z.-L. et al., 2017 |
| Serum exosomes | Downregulated (NON-HSAT101518.2, NON-HSAT067134.2, NON-HSAT148822.1, NON-HSAT078967.2) in ATB patients | ATB patients and healthy individuals | Discriminate ATB from healthy individuals | Fang et al., 2021 |
| PBMC     | Differently expressed (ENST00000497872, n333737, and n335265) in PTB and healthy control | Clinically diagnosed PTB, microbiologically confirmed PTB cases, non-TB disease controls, healthy subjects | Facilitate the early identification of PTB cases among suspected patients with negative Mtb microbiological evidence | Hu et al., 2020 |
| Lung tissue | Upregulated (ENST00000429730.1 and MSTRG.93125.4) | Clinically diagnosed PTB, microbiologically confirmed PTB cases, non-TB disease controls, healthy subjects | Potential indicators of metabolic activity in TB lesions for sputum-negative tuberculosis | Wang et al., 2021 |
| Whole blood | AC079767.4 | Clinically diagnosed PTB, microbiologically confirmed PTB cases, non-TB disease controls, healthy subjects | AC079767.4 polymorphisms may potentially act as biomarkers for TB diagnostic and even as therapeutic targets | Zhao et al., 2017 |
| Whole blood | Downregulated (AC145676.2.1-6 and TGS1-1) in TB patients | Clinically diagnosed PTB, microbiologically confirmed PTB cases, non-TB disease controls, healthy subjects | Facilitate the early identification of PTB cases among suspected patients with negative Mtb microbiological evidence | Bai et al., 2019 |
| Whole blood | RiPK2 | Clinically diagnosed PTB, microbiologically confirmed PTB cases, non-TB disease controls, healthy subjects | TGS1-1 and its variants 4737420 may be predictive indicators of anti-TB drug-induced adverse drug reactions | Song et al., 2019a |
| Peripheral blood | HNF1B-3:1 | Clinically diagnosed PTB, microbiologically confirmed PTB cases, non-TB disease controls, healthy subjects | Facilitate the early identification of PTB cases among suspected patients with negative Mtb microbiological evidence | Bai et al., 2019 |
| Whole blood | RP11-37B2.1 | TB patients and healthy individuals | Might serve as a hazard for TB in the Chinese Han population | Song et al., 2019b |
| Whole blood | CASC8 | Potential biomarker for the progression of clinical TB | Potential biomarker for the progression of clinical TB | Liu et al., 2020 |
Long Non-coding RNA Polymorphism and Tuberculosis Susceptibility

Abundant evidence from the investigations suggests that host genetic factors contribute to determine susceptibility to TB disease (Møller and Hoal, 2010). Recent advances in lncRNA research have preliminarily explored the relationship between lncRNA single nucleotide polymorphisms (SNPs), TB susceptibility, and clinical manifestations in TB patients of the western Chinese Han population (Table 2). For example, a study uncovered that the lncRNA AC079767.4 might be involved in the progression of TB infection. In addition, the C allele of SNP rs12477677 in AC079767.4 was associated with reduced susceptibility to PTB (Zhao et al., 2017). Moreover, the SNPs (rs12477677 and rs1055229) may influence clinical TB disease. Conceivably, AC079767.4 polymorphisms may serve as novel biomarkers for TB diagnostic and even as therapeutic targets (Zhao et al., 2017). Similarly, the potential TB-associated promoting effects were identified for the decreased expression levels of Inc-AC145676.2-1-6 and Inc-TGS-1 (Bai et al., 2019). Recently, the SNPs of rs39509 G allele in PIPK2 Near gene-3 region, lncRNA - HNF1B - 3:1, lncRNA - RP11-37 b2. 1, lncRNA - CASC8 has also been proved to be associated with TB susceptibility (Song et al., 2019a,b; Liu et al., 2020; Wu et al., 2020). These lncRNA polymorphisms are promising molecular biomarkers for clinical TB infection and/or efficacy evaluation. However, there are some questions need to be answered, such as are there sex and ethnic differences in differentially expressed lncRNAs?

DISCUSSION

Overall, the advancement of sequencing technology has facilitated the discovery of lncRNAs with unknown functions. These lncRNAs can thwart disease or lead to disease progression. Importantly, lncRNAs have presented a diverse perspective on the regulation of TB, especially complicated immune regulation for multiple biomolecular interactions. By high-throughput sequencing and deepening validation, lncRNAs have been found as potential biomarkers for diagnosis, monitoring progression, and clinical efficacy of TB. However, the complexity of the lncRNAs themselves and the lack of accurate databases for the lncRNAs discovered are restricting research to the single analysis of differences in gene expression. We are still far from completely understanding how lncRNAs influence complex physiopathological processes of TB, and several aspects of lncRNA remain enigmatic. For instance, whether lncRNA regulates Mtb infection as an infection phenomenon or a specific bacterial infection phenomenon under certain conditions. In addition, there is a lack of using lncRNAs as biomarkers to indicate progression from latent infections to clinical disease. Despite the questions, lncRNAs are highly ideal biomarkers. Further validation studies on different ethnic populations and function experiments in a large-scale cohort help to confirm the roles of the lncRNAs, and may also help identify biomarkers for latent infections. Most of the research on ncRNAs mainly focuses on miRNA, but understanding the lncRNA function can only be achieved from more studies on a case-by-case basis.

Several new biotechnologies such as Xpert MTB/RIF Ultra have been developed in this context. It has increased sensitivity for diagnosis but is sophisticated. More accurate, rapid, and cost-effective tests are needed to improve TB detection. Various platforms have explored other possible biomarkers, such as host marker signatures, including host gene expression, protein, and metabolites. Some have combined omics techniques which have made some progress in distinguishing LTBI from ATB to explore progression to TB. However, such measurement of expression of markers requires substantial laboratory infrastructure and is time-consuming. Additionally, the most work done so far has been small case-control studies, and the diagnostic potential of these markers still has to be confirmed. The continued study of TB-associated lncRNAs will reveal more unanticipated biomarkers for predicting progression, response to treatment, and relapse. Moreover, the personalized lncRNA-targeted drugs in host-directed therapy (HDTS) are under development.

AUTHOR CONTRIBUTIONS

XZ collected the related manuscript and drafted the manuscript. XZ, CC, and YX discussed and developed its conceptual framework. CC and YX proofread the manuscript. YX and XZ revised the review. All authors contributed to the article and approved the submitted version.

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