Case Report

Invasive *Aspergillus niger* Is the Sole Etiological Agent for CSOM: A Clinical Case from Nepal

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1.Introduction

Chronic suppurative otitis media (CSOM) is a chronic inflammatory middle ear cleft condition that is aggravated by partial or complete loss of the tympanic membrane and ossicles and often induces permanent sequelae that manifest as deafness and persistent ear discharge [1]. Globally, the burden of CSOM illness accounts for 28000 deaths; 65–330 million with draining ear and 39–200 million people are suffering from significant hearing impairment [2]. In Nepal, it is a major public health burden with a high prevalence rate [3]. If the prevalence of CSOM is greater than 3, it is accounted as a high-risk priority disease in the region. Nepal falls in the high endemic of CSOM with a prevalence of 3.5% [3–5]. We are reporting a case of a CSOM patient who was initially treated with topical antibiotics, whereas the laboratory diagnosis was secured on the evaluation for fungal pathogenesis and revealed invasive *Aspergillus niger* as the sole etiological agent. We, therefore, recommend a strong indicator of clinical suspicion supported by the standard and
relevant microbiological analysis to avoid its misdiagnosis for an effective treatment option.

2. Case Presentation

A 53-year-old man who has a history of CSOM in the right ear was suffering for more than 3 months. He had visited various healthcare facilities and hospitals. He was on an antibacterial drug from the last 10 days but was not benefited. Finally, he visited the department of ENT and Head and Neck Surgery OPD of a tertiary-care hospital and underwent clinical investigation and treatment in March 2016.

3. Investigation and Outcomes

During his visit to the Department of Otorhinolaryngology-Head and Neck Surgery (ENT-HNS), Tribhuvan University and Teaching Hospital (TUTH), a medical team observed his ears with an otoscope and immediately spotted the ear infection with fungus, i.e., otomycosis. Furthermore, the doctor performed microscopic suction clearance of fungal mass. Subsequently, the use of topical antibiotic was discontinued.

For clinical investigation, we aseptically collected four swab samples from the right ear discharge and conducted further downstream assays, primarily microbiological investigations, Gram stain, KOH mount, and bacterial and fungal culture of aural discharge. The patient was advised for follow-up after a week for further treatment.

The first aural swab was processed for the preparation of Gram staining and potassium hydroxide (KOH) mount preparation. The second swab was inoculated into Robertson’s cooked meat (RCM) medium, while the third was aliquoted into the nutrient broth (NB) and then the aliquots were inoculated into chocolate agar (CHOC), 5% sheep blood agar (BAP), and MacConkey medium (MAC), respectively. The fourth swab was utilized for the culture in Sabouraud dextrose agar (SDA), and clonal selection and subculture were conducted in specific media, i.e., malt yeast agar (MYA) and corn meal agar (CMA). All the clinical investigations procedures have been performed in accordance with the guidelines of the Clinical Microbiology Procedures Handbook, Volume 2 by Lynne S. Garcia, and Textbook of Medical Mycology, the Third Edition by Jagdish Chander. All the experiments were performed in triplicate utilizing the relevant positive and negative controls.

The Gram staining of the aural discharge revealed the presence of plenty of pus cells with fungal spores (conidia); however, no bacteria were detected (Figure 1).

The KOH mount exhibited the septe fungal hyphae with dichotomous branching (a typical feature of Aspergillus spp.) (Figure 2). For further microbial characterization and investigation, the aliquot of the culture was incubated for 18–24 hours in all three media plates, i.e., chocolate agar, blood agar, and MacConkey agar. The results indicated that there was no bacterial growth; however, fungal growth was observed in all three media plates (Figure 3).

To rule out bacterial growth, Gram stain was performed with the colonies/growth from all three media plates. For the precise phenotypic characterization of the fungal growth, LPCB preparation was assessed and observed.

After 48 hours of incubation in RCM, the growth was further subcultivated into blood agar with disks of metronidazole plus gentamicin and cultured anaerobically for 48 hours. Pseudomonas aeruginosa (previously isolated in the department) was used as a control strain for aerobic culture and Clostridium sporogenes (previously isolated in the department) was used as a control for anaerobic culture in this study.

In the blood agar medium, no bacterial growth was observed; however, the light growth of the fungus was spotted which was confirmed through Gram staining and LPCB mount preparation.

After 48 hours of incubation in an SDA medium, the fungal growth (Figure 4) was further inoculated into a corn meal agar (CMA) and cultured for 72 hours. The fungal colonies from SDA and CMA media were proceeded into the lactophenol cotton blue (LPCB) tease mount. The microscopy results from the LPCB mount revealed the presence of multiple conidia, conidiophores, vesicles, phialides, and metulae resembling Aspergillus spp. (Figure 5). Furthermore, we have also conducted the clonal selection and utilized Malt yeast agar (MYA) and CMA-specific culture media for the growth of Aspergillus spp. and successfully isolated and confirmed Aspergillus niger. From all microbial observations and analysis, the finding of the vital characterization of the specific biseriate structure of phialides and the black-coloured conidia phenotypically confirmed Aspergillus niger as the sole etiological agent for this CSOM.

4. Discussion

Otomycosis is a chronic recurring mycosis of the outer ear canal. The incidence rate of otomycosis is high in tropical regions, and the species of Aspergillus are considered the predominant etiological factors which account for 48.9% of the entire otomycosis manifestations [6].

CSOM is one of the chronic types of otitis. It is ubiquitously caused by specific bacteria and fungi. Etiological agents of CSOM of significant aerobic bacterial origins are Pseudomonas aeruginosa, Staphylococcus aureus, Proteus mirabilis, Klebsiella pneumoniae, and Escherichia coli, while anaerobic bacterial origins are Clostridium spp., Peptostreptococcus spp., Fusobacterium spp., and Veillonella spp. Major fungal species involved in CSOM are Aspergillus spp., Candida spp., Rhizopus spp., and Penicillium spp. [7, 8]. Prolonged and irrational use of topical antimicrobial (antibiotics or antibiotics-steroids) ear drops could shift and suppress the commensal and emerge opportunistic bacterial flora [9].

CSOM is a common health burden in low- and middle-income countries where socioeconomic status, overall health, hygiene and sanitation conditions, nutritional status, and accessibility to healthcare services are severely compromised. In addition, frequent upper respiratory tract infections, irrational antibiotic treatment, and nasal diseases
are considered as the vital predisposing factors to CSOM [10, 11]. Without a precise etiological diagnosis of CSOM, the irrational use of antibiotics leads to the emergence of multidrug-resistant microbial strains which gives rise to severe complications that is an ultimate challenge currently faced by low- and middle-income countries. Each year, CSOM accounts for thousands of deaths and directly impacts millions of people who suffer from significant hearing impairment.

Aspergillus species are the most predominant fungal etiology in CSOM. Aspergillus conidia disperse in airborne dust, colonize, and grow rapidly in the ear canal which is supported by nutrients present in cerumen and the slightly acidic environmental condition. Hundreds of Aspergillus conidia can survive within the air; therefore, even if it is inhaled, no disease can be progressed to the immunocompetent individuals. However, the immunocompromised

**Figure 1:** Gram staining revealing plenty of pus cells with fungal spores (Conidia) but lacking bacteria, when observed under a 100X Olympus CX21 compound microscope.

**Figure 2:** KOH mount revealing the septate fungal hyphae with the characteristic dichotomous branching, when observed under a 40X Olympus CX21 compound microscope.

**Figure 3:** Aliquots from the culture sample grown on CHOC, BAP, MAC, and SDA media indicate the growth of Aspergillus spp.

**Figure 4:** Aspergillus spp. observed as dark brown/black colonies grown on the SDA medium.

**Figure 5:** Confirmation of Aspergillus niger with the specific phenotypic characterization of conidiophores, vesicles, metulae, biseriate phialides, and conidia in the LPCB mount, observed under a 100X Olympus CX21 compound microscope.
hosts with diabetes, steroid administration, HIV infection, chemotherapy, and malignancy are susceptible to CSOM with fungal infection [12]. Immunosuppression health condition increases the risk of dissemination of the opportunistic pathogens *Aspergillus* spp. into various body organs either via circulation or by direct tissue invasion [13]. *Aspergillus* species ubiquitously occur in the diverse environment and their conidia mostly disperse in the air. If anyone is exposed to these conidia, the risk of otomycosis/CSOM progression is minimal for immune-competent individuals [14]. *Aspergillus niger* is one of the significant and predominant invasive fungal pathogens responsible for the infection of recurrent drainage of CSOM.

The proper clinical diagnosis of CSOM is carried out by the standard bacteriological and mycological assessments. Microbial phenotypic studies specifically focusing on fungal culture, wet mount, and staining are able to distinguish the specific etiological agent of CSOM in resource-constraint laboratory settings. Therefore, the clinical investigation and diagnosis of the etiological agent of CSOM can be achieved by the observation/demonstration of microbial evidence by the variety of standard methods, primarily Gram staining, KOH mount preparation, and bacterial and fungal culture in selective and differential media.

In this case study, we have clinically investigated the aural discharge from the right ear of a 53-year-old man suffering from CSOM. The patient probably got infected being immunocompromised due to his diabetic condition. We have systematically investigated this case study with the standardized microbial assessments which is shown in the flowchart for this study (Figure 6). The findings revealed no bacterial growth was able to be cultured from the collected samples of the patient; however, the fungus was isolated and identified.

Clonal selection and the specific culture in MYA and CMA generated black colonies of *Aspergillus* spp. which was further phenotypically investigated. For further characterization of *Aspergillus* spp., the revelation of the specific phenotypic characterization of conidiophores, vesicles, metulae, biseriate phialides, and conidia in microscopy phenotypically confirmed *Aspergillus niger* as the sole etiological agent of this case of CSOM.

The current recommended treatment for otomycosis due to *Aspergillus* spp. is clotrimazole (1% w/v) ear drops, and they are given for 3-4 weeks. It is considered as the effective antifungal therapeutic option as it stops fungal growth in the ear blocking them from the formation of their protective covering [15, 16].

For the therapy of this clinical case, the patient was also treated with the recommended medication of Candid Ear Drop with lidocaine (2% w/v) plus clotrimazole (1% w/v) for 4 weeks. When we followed up after 4 weeks of treatment and made the patient undergo overall ear check-up investigations, his ear and health conditions were improved remarkably. For further clinical investigation, swab samples were also aseptically collected from both ears and microbial assessments were performed; however, no microbial growth was isolated and detected.

5. Conclusions

CSOM caused by both bacterial and fungal origins in laboratory diagnosis has frequently been reported; however,
CSOM associated with the clinically relevant fungal flora alone is a unique case.

In this clinical case report, we have clinically and microbiologically investigated the aural discharge from a CSOM patient, isolated and phenotypically distinguished invasive *Aspergillus niger* as the sole etiological pathogen associated with the CSOM presented.

To sum up, irrational and prolonged use of antimicrobials ear drops may lead to the emergence of the microbial superinfection-CSOM with opportunistic virulent pathogens. Therefore, a definite and prompt investigation for fungal etiology with the standardized laboratory assessments is recommended for each CSOM case which subsequently guides for the proper and timely management and treatment of CSOM and prevents CSOM-associated severe complications.

**Abbreviations**

BAP: 5% sheep blood agar plate  
CMA: Corn meal agar  
CHOC: Chocolate agar  
CSOM: Chronic suppurative otitis media  
ENT-HNS: Otorhinolaryngology-head and neck surgery  
HIV: Human immunodeficiency virus  
KOH: Potassium hydroxide  
LPCB: Lactophenol cotton blue  
MAC: MacConkey agar  
MYA: Malt yeast agar  
NB: Nutrient broth  
OPD: Outpatient department  
RCM: Robertson’s cooked meat  
SDA: Sabouraud dextrose agar  
TUTH: Tribhuvan University Teaching Hospital.

**Data Availability**

No data were used to support this study.

**Conflicts of Interest**

The authors declare no conflicts of interest.

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**References**

[1] S. N. Merchant, M. J. McKenna, and J. J. Rosowski, “Current status and future challenges of tympanoplasty,” *European Archives of Oto-Rhino-Laryngology*, vol. 255, no. 5, pp. 221–228, 1998.

[2] World Health Organization, *Chronic Suppurative Otitis Media: Burden of Illness and Management Options*, World Health Organization, Geneva, Switzerland, 2004.

[3] P. Adhikari, B. Sinha, N. Pokharel, B. Kharal, R. Aryal, and J. Ma, “Prevalence of chronic suppurative otitis media in school children of Kathmandu district,” *Journal of Institute of Medicine Nepal*, vol. 29, no. 3, pp. 10–12, 2007.

[4] M. M. Rovers, H. Straatman, G. A. Zielhuis, K. Ingels, and G.-J. Van Der Wilt, “Seasonal variation in the prevalence of persistent otitis media with effusion in one-year-old infants,” *Paediatric & Perinatal Epidemiology*, vol. 14, no. 3, pp. 268–274, 2000.

[5] N. Nwakoye, L. Egwari, A. Coker, O. Olubi, E. Ugoji, and S. Nwachukwu, “Predisposing and bacteriological features of otitis media,” *African Journal of Microbiology Research*, vol. 6, no. 3, pp. 520–525, 2012.

[6] S. Tasić-Otašević, M. Golubović, S. Denić et al., “Species distribution patterns and epidemiological characteristics of otomycosis in Southeastern Serbia,” *Journal of Medical Mycology*, vol. 30, no. 3, Article ID 101011, 2020.

[7] S. Saranya, G. Vazhavandal, B. Vallab Ganesh, M. Ismail, A. Uma, and T. Subramaniam, “Bacteriological and mycological profile of chronic suppurative otitis media in a tertiary teaching hospital, Trichy, Tamilnadu,” *International Journal of Pharmaceutical Science Invention*, vol. 4, no. 1, pp. 13–19, 2015.

[8] R. Prakash, D. Juyal, V. Negi et al., “Microbiology of chronic suppurative otitis media in a tertiary care setup of Uttarakhand state, India,” *North American Journal of Medical Sciences*, vol. 5, no. 4, p. 282, 2013.

[9] D. Juyal, V. Negi, M. Sharma, S. Adekhandi, R. Prakash, and N. Sharma, “Significance of fungal flora in chronic suppurative otitis media,” *Annals of Tropical Medicine and Public Health*, vol. 7, no. 2, p. 120, 2014.

[10] S. K. Thakur, S. K. Singh, R. Acharya, A. Anwar, and N. Ghimire, “Sociodemographic profile and the associated factors of chronic otitis media in rural areas of eastern Nepal,” *International Journal of Otorhinolaryngology and Head and Neck Surgery*, vol. 3, no. 2, pp. 222–227, 2017.

[11] L. Hunt, W. Mulwafu, V. Knott et al., “Prevalence of paediatric chronic suppurative otitis media and hearing impairment in rural Malawi: a cross-sectional survey,” *PLoS One*, vol. 12, no. 12, Article ID e0188950, 2017.

[12] B. Viswanatha and K. Naseeruddin, “Fungal infections of the ear in immunocompromised host: a review,” *Mediterranean journal of hematology and infectious diseases*, vol. 3, no. 1, Article ID e2011003, 2011.

[13] J. Mohanty, S. Mohanty, R. Sahoo, S. Ghosh, N. Chayani, and B. Mallick, “Clinico-microbial profile of otomycosis in Berhampur,” *India Journal of Otolology*, vol. 5, no. 2, pp. 81–83, 1999.

[14] P. S. Morris, A. J. Leach, S. Halpin et al., “An overview of acute otitis media in Australian Aboriginal children living in remote communities,” *Vaccine*, vol. 25, no. 13, pp. 2389–2393, 2007.

[15] J. C. Stern, M. K. Shah, and F. E. Lucente, “In vitro effectiveness of 13 agents in otomycosis and review of the literature,” *The Laryngoscope*, vol. 98, no. 11, pp. 1173–1177, 1988.

[16] A. Jackman, R. Ward, M. April, and J. Bent, “Topical antibiotic induced otomycosis,” *International Journal of Pediatric Otorhinolaryngology*, vol. 69, no. 6, pp. 857–860, 2005.