Comparison of Two Assays to Determine Anti-Citrullinated Peptide Antibodies in Rheumatoid Arthritis in relation to Other Chronic Inflammatory Rheumatic Diseases: Assaying Anti-Modified Citrullinated Vimentin Antibodies Adds Value to Second-Generation Anti-Citrullinated Cyclic Peptides Testing

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Determination of anti-citrullinated peptide antibodies (ACPA) plays a relevant role in the diagnosis of rheumatoid arthritis (RA). To date, it is still unclear if the use of several tests for these autoantibodies in the same patient offers additional value as compared to performing only one test. Therefore, we evaluated the performance of using two assays for ACPA: second-generation anti-citrullinated cyclic peptides antibodies (anti-CCP2) and anti-mutated citrullinated vimentin (anti-MCV) antibodies for the diagnosis of RA. We compared three groups: RA (n = 142), chronic inflammatory disease (CIRD, n = 86), and clinically healthy subjects (CHS, n = 56) to evaluate sensitivity, specificity, predictive values, and likelihood ratios (LR) of these two assays for the presence of RA. A lower frequency of positivity for anti-CCP2 was found in RA (66.2%) as compared with anti-MCV (81.0%). When comparing RA versus other CIRD, sensitivity increased when both assays were performed. This strategy of testing both assays had high specificity and LR+. We conclude that adding the assay of anti-MCV antibodies to the determination of anti-CCP2 increases the sensitivity for detecting seropositive RA. Therefore, we propose the use of both assays in the initial screening of RA in longitudinal studies, including early onset of undifferentiated arthritis.
1. Introduction

Rheumatoid arthritis (RA) is a chronic inflammatory disorder that involves synovial joints and may develop extra-articular manifestations [1]. Frequently, the diagnosis of RA may pose some difficulties in primary care, particularly during early disease, and this disease may inappropriately be confused with other rheumatic diseases [2]. In this context, a relevant tool to support the diagnosis is the presence of autoantibodies associated with the disease. Although the detection of rheumatoid factor [3] is useful to support the diagnosis and it is detected in 75% of patients with RA, a limitation of this autoantibody is its low specificity, being frequently observed in other rheumatic disorders, chronic infections, and even in healthy elderly people [3]. Different assays are currently used to detect antibodies against cyclic citrullinated antigens as well as noncyclic citrullinated peptides. Therefore, the term anti-citrullinated peptide antibody (ACPA) is commonly used in these days. Assays to identify antibodies against citrullinated cyclic peptides are commonly used as a tool to support the diagnosis of RA, because it has been widely demonstrated that these autoantibodies have higher specificity as compared with the rheumatoid factor (RF). One of the most common assays is the determination of second-generation anti-citrullinated cyclic peptide antibodies (anti-CCP2). Therefore, ACPAs have been included in the most recent classification criteria for RA diagnosis [4]. Nevertheless, around 38% of patients with RA may have negative results for anti-CCP2 [5, 6]. Assays determining antibodies against human mutated vimentin (anti-MCV) have been also proposed recently as a tool for the diagnosis of RA [7, 8]. Nevertheless, still 26% of patients with RA may yield negative results with these assays [7]. To date, there are several studies comparing the performance of different assays of anti-CCP2 versus anti-VCM in the diagnosis of RA [9–11]. These studies support that detection of anti-VCM is as useful as the assays determining anti-CCP2 to distinguish RA from healthy controls [12, 13] and can help in the differential diagnosis of RA from other rheumatic disorders [14–16]. Nevertheless, currently, there are no studies in Mexican patients evaluating if the strategy of performing both tests may increase sensitivity and positive predictive value for the presence of established RA as compared to performing them individually.

Therefore, we evaluated the performance of using two ACPA assays: second-generation anti-citrullinated cyclic peptide antibodies (anti-CCP2) and anti-mutated citrullinated vimentin (anti-MCV) antibodies in established RA, and we correlated the titers observed of these autoantibodies with disease activity.

2. Patients Methods

Design. Cross-sectional study.

Clinical Setting. Adult consecutive patients with RA seen in an outpatient rheumatology clinic of a secondary-care center in Guadalajara, Mexico (Hospital General Regional I10, Instituto Mexicano del Seguro Social), were invited to participate if they met at least four of the 1987 ACR criteria for RA [17]. They were excluded if they had a history of blood transfusion, chronic infectious diseases, including hepatitis B or C, human immunodeficiency virus, or tuberculosis. Patients with overlapping syndrome, cancer, or other associated autoimmune disorders or pregnant patients were also excluded.

These patients were compared with two distinct non-RA controls selected.

(i) The first comparison group was constituted by patients with other rheumatic inflammatory disorders mainly including systemic lupus erythematosus (SLE, 1982 ACR criteria) [18] or ankylosing spondylitis (AS, 1984 New York modified criteria) [19]. Nevertheless, patients with systemic sclerosis (SSc) and articular manifestations were included if they met the 1980 ACR criteria [20]. All these patients were obtained from the same rheumatology clinic where patients with RA were recruited.

(ii) The second group was constituted by clinically healthy blood donors obtained from the same hospital, without history of blood transfusion or chronic infections.

For these two comparison groups, similar inclusion and exclusion criteria described for patients with RA were applied.

2.1. Clinical Evaluations. A structured assessment for patients with RA was performed including disease characteristics, evaluation of disease activity according to DAS-28 [21], functioning according to the Spanish validated version of HAQ-DI [22], and treatments used.

2.2. ACPA Determinations. A venous blood sample was taken from all included subjects at the same time of the clinical evaluation and the serum was obtained and stored at −20°C until antibodies determination. Anti-CCP2 were determined by ELISA using a commercial kit (Axis-Shield, UK) with a cut-off value for positivity >20 U/mL and anti-MCV were determined by ELISA using also a commercial kit (ORGENTEC, Mainz, Germany) with a cut-off value for positivity >20 U/mL.

3. Statistical Analysis

Qualitative variables were expressed as frequencies and percentages and quantitative variables were expressed as means ± standard deviations. Chi-square tests were used to compare proportions among groups and Student’s t-test was used to compare means between two groups. We selected as “gold standard” the 1987-ACR criteria for diagnosis of established RA. These criteria were used instead of the most recent 2010-ACR criteria because the status of positive ACPA is included within the criteria. The performance of the assays for anti-MCV and anti-CCP2, either individually or tested together, to identify RA was evaluated estimating sensitivity, specificity, and positive and negative predictive values, as well as likelihood ratios. In this study, sensitivity can be defined as the probability of positive anti-CCP2 or anti-MCV in patients with RA. Specificity was defined as the probability of negative results for these autoantibodies in patients or controls without RA. Positive predictive value (PPV+) was
Table 1: General characteristics in patients with rheumatoid arthritis.

| Characteristics                      | N = 142 |
|--------------------------------------|---------|
| Age in years, mean ± SD              | 49 ± 10.69 |
| Women, n (%)                         | 135 (95) |
| Disease duration (years), mean ± DE  | 9 ± 8.07 |
| DAS-28, mean ± SD                    | 4.7 ± 1.5 |
| DAS-28 > 3.2 n (%)                   | 118 (83.1) |
| HAQ-DI, mean ± SD                    | 0.91 ± 0.65 |
| HAQ-DI > 1.25, n (%)                 | 38 (26.6) |
| Treatments                           |         |
| Methotrexate, n (%)                  | 48 (33.8) |
| Chloroquine, n (%)                   | 4 (2.8) |
| Leflunomide, n (%)                   | 17 (12) |
| Azathioprine, n (%)                  | 18 (14) |
| Etanercept, n (%)                    | 8 (5.6) |
| Glucocorticoids, n (%)               | 124 (87.9) |
| Prednisone mg, mean ± SD             | 5.7 ± 1.6 |

SD: standard deviation, mg: milligrams.
DAS-28: disease activity score.
DAS-28: low activity ≤3.2; moderate activity >3.2 ≤5.1; high activity >5.1.
HAQ-DI: Health Assessment Questionnaire-Disability Index: S.

Table 2: Concordance between the results of assays for anti-CCP2 and anti-MCV in rheumatoid arthritis.

|                  | Anti-CCP2 |
|------------------|-----------|
|                  | Positive  | Negative |
| Anti-CCP2        | n = 115   | n = 48    |
| Positive         | 88 (61.97%) | 27 (19.01%) |
| Negative         | 6 (4.22%)  | 21 (14.78%) |

Total patients with RA assessed = 142, and values in parenthesis represent the percentage of the total 142 patients. Kappa = 0.42.

An evaluation of utility values for the strategies of testing each assay, anti-CCP2 or anti-MCV alone, or testing both assays in established RA compared with clinically healthy blood donors is shown in Table 3. The highest sensitivity was observed when both autoantibodies tests were performed (85%) followed by testing anti-MCV alone (81%), whereas the lowest sensitivity was observed when only anti-CCP2 test was performed. On the other side, specificity and PPV(+) were similar with the three strategies, and the NPV(−) increased substantially, if both assays were negative.

The utility values for the strategies of performing only anti-CCP2 or anti-MCV or both of these assays in established RA compared with other rheumatic inflammatory diseases are shown in Table 4. The highest sensitivity was again observed when both assays were performed (85%) and the lowest sensitivity was attained when using only anti-CCP2 (66%). The highest specificity was observed when only anti-MCV was performed (96%). PPV(+) values were higher with the anti-MCV assay alone (97%), whereas the highest NPV(−) was observed when both assays were negative (79%).

5. Discussion

In our study, we observed that the assay for anti-MCV antibodies showed more sensitivity and specificity than the assay for anti-CCP2 antibodies to distinguish established RA patients from other systemic inflammatory rheumatic diseases. Using the strategy of performing both assays, we obtained an increase in sensitivity in comparison with using either assay individually. In our study, the Kappa between both assays indicates that determination of both tests should be complementary and consequently increases the utility of both tests in the clinical armamentarium without decreasing specificity.

Previous studies have reported, for anti-CCP2, specificities greater than 90% [23–25], similar to our findings where we found a specificity of 92% for CIRD and 94% for CHS, this assay being very useful to exclude people who do not have RA.

Nevertheless, in terms of a screening test, a higher sensitivity is extremely relevant; therefore, strategies to increase the values of sensitivity are required to establish an earlier diagnosis and opportune reference to the rheumatologist. To this regard, in the present study, the utilization of an assay for anti-CCP2 exclusively had only 66% of sensitivity,
Table 3: Utility values of anti-CCP2, anti-MCV, or any of these assays in rheumatoid arthritis in comparison with clinically healthy subjects (CHS).

| Utility values of the assays for anti-CCP2 and anti-MCV results | Anti-CCP2 | Anti-MCV | Anti-CCP2 or anti-MCV |
|---------------------------------------------------------------|----------|---------|-----------------------|
| Sensitivity % (95% CI)                                       | 66 (58–74) | 81 (73–87) | 85 (78–91)           |
| Specificity % (95% CI)                                       | 94 (84–99) | 94 (84–99) | 94 (84–99)           |
| Positive predictive value % (95% CI)                        | 97 (91–99) | 97 (93–99) | 97 (93–99)           |
| Negative predictive value % (95% CI)                        | 51 (41–61) | 65 (53–75) | 70 (58–81)           |
| LR+                                                           | 11.69 (3.87–35.32) | 14.31 (4.75–43.07) | 15.05 (5–45.28) |
| LR−                                                           | 0.36 (0.28–0.46) | 0.20 (0.14–0.28) | 0.16 (0.11–0.23) |
| Prevalence                                                    | 73 (66–79) | 73 (66–79) | 73 (66–79)           |

LR+: positive likelihood ratio; LR−: negative likelihood ratio.

Table 4: Utility values of anti-CCP2, anti-MCV, or any of these assays in rheumatoid arthritis in comparison with other chronic inflammatory rheumatic diseases (CIRD).

| Utility values of the assays for anti-CCP2 and anti-MCV results | Anti-CCP2 | Anti-MCV | Anti-CCP2 or Anti-MCV |
|---------------------------------------------------------------|----------|---------|-----------------------|
| Sensitivity % (95% CI)                                       | 66 (58–74) | 81 (73–87) | 85 (78–90)           |
| Specificity % (95% CI)                                       | 92 (84–97) | 96 (90–99) | 92 (84–97)           |
| Positive predictive value % (95% CI)                        | 93 (86–97) | 97 (93–99) | 94 (89–98)           |
| Negative predictive value % (95% CI)                        | 62 (53–71) | 75 (66–83) | 79 (70–86)           |
| LR+                                                           | 8.13 (3.96–16.7) | 23.22 (7.62–70.77) | 10.47 (5.13–21.36) |
| LR−                                                           | 0.37 (0.29–0.47) | 0.20 (0.14–0.28) | 0.16 (0.11–0.24) |
| Prevalence                                                    | 62 (67–89) | 62 (68–89) | 62 (56–68)           |

LR+: positive likelihood ratio; LR−: negative likelihood ratio.

whereas when both assays, anti-CCP2 and anti-MVC, were done in the same patients, the sensitivity increased to 85%, with an improvement in the utility of these assays as a tool for clinicians. Regarding specificity of anti-CCP2, some studies have shown a wide variability ranging from 40% to 83% [26, 27], the frequency of negatives being a limitation to establish the diagnosis in RA. Genetic factors may contribute to these differences in sensitivity, characteristic of the study population, including variables such as disease duration or severity of the disease, and characteristics of assays used to detect these autoantibodies [28], although, in our study, anti-MCV antibodies were more sensitive than anti-CCP2 antibodies for RA and these findings have been reported by others [29].

To this regard, around 1 of 5 patients with established RA had a negative anti-MCV test result. Therefore, the question arises if the utility value of the test could be increased by using both assays. We observed that using both assays in the same patients the sensitivity increases to 85% with an LR+ of 10.47 in comparison to other CIRD, constituting an excellent support in the clinical armamentarium for RA.

Several factors could contribute to explaining why we observed that the anti-VCM assay was more sensitive than the anti-CCP2 assay. One of them is that vimentin contains 43 arginine residues. Each arginine residue can potentially be citrullinated by peptidylarginine deiminase (PAD) resulting in a variety of citrullinated epitopes. In contrast, in the anti-CCP2 test only a few epitopes are presented [30–32].

Some authors reported recently that combining determinations of anti-MCV, anti-CCP2, and RF increases the sensitivity [15]. Nicase-Roland et al. [29] described, in a cohort of patients with early RA and undifferentiated arthritis, an increase in sensitivity when two tests are associated. Therefore, these data support our findings implying gains in clinical utility when two assays for ACPA are applied in the same patient. Our study, however, revealed that still 6% of controls without any rheumatic disorders had positive anti-CCP2 or anti-MCV antibodies; these data are relevant because the presence of a positive antibody without clinical manifestations is insufficient to support the presence of disease, although we ignore it if these patients would have an increase in risk for a CIRD in the future. Cohort studies will help to identify the evolution of these patients with positive anti-MCV.

Some limitations of the study due to its cross-sectional nature are that we were unable to identify if controls without rheumatic disorders who depicted positivity to one or both autoantibodies will have progression to a CIRD in the future; nevertheless, this hypothesis should be tested in cohort models, increasing the number of patients. On the other side, we did not apply these tests to specific subgroups of patients, such as RA with extra-articular manifestations, undifferentiated arthritis, or early RA, where the performance of these diagnostic tests may have substantial variations to those observed in defined RA. Another limitation was that we did not include an assay for testing anti-CCP3. Anti-CCP3 assays rely upon additional epitopes not present in the anti-CCP2 antigen sequence [33, 34]. Szekanecz et al. evaluated the sensitivity of cyclic citrullinated antibodies second-generation (anti-CCP2) and third generation (anti-CCP3 and anti-CCP3.1); the diagnostic sensitivity of anti-CCP2 was 74.8%, anti-CCP3 was 78.8%, and anti-CCP3.1 was 83.0%; the specificity of anti-CCP2 was 95.7%, anti-CCP3 was 97.8%.
96.6%, and anti-CCP3. 98.3% [35]. However, Shidara et al. show no evident increase in utility values when comparing anti-CCP3 and anti-CCP2 assays; the sensitivity of anti-CCP2 was 88.7% and specificity of anti-CCP2 was 89.5%, whereas; the sensitivity of anti-CCP3 was 91.5% and specificity was 87.7% [36]. An assay for anti-CCP3 may provide an increase in sensitivity as compared to that observed with the assay for anti-CCP2 used in this study.

In conclusion, using both assays, anti-CCP2 and anti-MCV, increases the sensitivity for the presence of RA as compared to performing only one assay; therefore, this strategy should be included in the clinical armamentarium to improve the value of these assays as screening tests.

**Ethical Approval**

The Institutional Research Committee of the Hospital approved Project number R-2009-1301-57. All the included patients and controls signed a voluntary informed consent. This protocol followed the guidelines of the Helsinki declaration.

**Conflict of Interests**

All the authors declare that there is no conflict of interests to disclose.

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**References**

[1] J. A. Rindfleisch and D. Muller, “Diagnosis and management of rheumatoid arthritis,” The American Family Physician, vol. 72, no. 6, pp. 1037–1047, 2005.

[2] J. I. Gamez-Nava, L. Gonzalez-Lopez, P. Davis, and M. E. Suarez-Almazor, “Referral and diagnosis of common rheumatic diseases by primary care physicians,” British Journal of Rheumatology, vol. 37, no. 11, pp. 1215–1219, 1998.

[3] M. A. M. van Boekel, E. R. Vossenaar, F. H. J. van den Hoogen, and W. J. van Venrooij, “Autoantibody systems in rheumatoid arthritis: specificity, sensitivity and diagnostic value,” Arthritis Research, vol. 4, no. 2, pp. 87–93, 2002.

[4] D. Aletaha, T. Neogi, and A. J. Silman, “2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League against Rheumatism collaborative initiative,” Annals of the Rheumatic Diseases, vol. 69, pp. 1580–1588, 2010.

[5] P. Taylor, J. Gartemann, J. Hsieh, and J. Creeden, “A systematic review of serum biomarkers anti-cyclic citrullinated peptide and rheumatoid factor as tests for rheumatoid arthritis,” Autoimmune Diseases, vol. 1, no. 1, Article ID 815038, 2011.

[6] E. Zinzstaras, A. A. Papathanasiou, D. C. Ziogas, and M. Vougalierlis, “The reporting quality of studies investigating the diagnostic accuracy of anti-CCP antibody in rheumatoid arthritis and its impact on diagnostic estimates,” BMC Musculoskeletal Disorders, vol. 13, article 113, 2012.

[7] H. Poulos and P. J. Charles, “Antibodies to citrullinated vimentin are a specific and sensitive marker for the diagnosis of rheumatoid arthritis,” Clinical Reviews in Allergy and Immunology, vol. 34, no. 1, pp. 4–10, 2008.

[8] E. Wagner, M. Skoumal, P. M. Bayer, and K. Klaushofer, “Antibody against mutated citrullinated vimentin: a new sensitive marker in the diagnosis of rheumatoid arthritis,” Rheumatology International, vol. 29, no. 11, pp. 1315–1321, 2009.

[9] C. Dejaco, W. Klotz, H. Larcher, C. Duftner, M. Schirmer, and M. Herold, “Diagnostic value of antibodies against a modified citrullinated vimentin in rheumatoid arthritis,” Arthritis Research and Therapy, vol. 8, no. 4, article R119, 2006.

[10] K. Raza, L. Mathsson, C. D. Buckley, A. Filer, and J. Rönnelid, “Anti-modified citrullinated vimentin (MCV) antibodies in patients with very early synovitis,” Annals of the Rheumatic Diseases, vol. 69, no. 3, pp. 627–628, 2010.

[11] J. Ursum, M. M. J. Nielen, D. van Schaardenburg et al., “Antibodies to mutated citrullinated vimentin and disease activity score in early arthritis: a cohort study,” Arthritis Research and Therapy, vol. 10, no. 1, article R12, 2008.

[12] X. Liu, R. Jia, J. Zhao, and Z. Li, “The role of anti-mutated citrullinated vimentin antibodies in the diagnosis of early rheumatoid arthritis,” Journal of Rheumatology, vol. 36, no. 6, pp. 1136–1142, 2009.

[13] H. Bang, K. Egerer, A. Gauliard et al., “Mutation and citrullination modifies vimentin to a novel autoantigen for rheumatoid arthritis,” Arthritis and Rheumatism, vol. 56, no. 8, pp. 2530–2511, 2007.

[14] E. Besada, C. Nikolaisen, and H. Nossent, “Diagnostic value of antibodies against mutated citrullinated vimentin for rheumatoid arthritis,” Clinical and Experimental Rheumatology, vol. 29, no. 1, pp. 85–88, 2011.

[15] T. Zhu and L. Feng, “Comparison of anti-mutated citrullinated vimentin, anti-cyclic citrullinated peptides, anti-glucose-6-phosphate isomerase and anti-keratin antibodies and rheumatoid factor in the diagnosis of rheumatoid arthritis in Chinese patients,” International Journal of Rheumatic Diseases, vol. 16, no. 2, pp. 157–161, 2013.

[16] E. Bartoloni, A. Alunno, O. Bistoni et al., “Diagnostic value of anti-mutated citrullinated vimentin in comparison to anti-cyclic citrullinated peptide and anti-viral citrullinated peptide 2 antibodies in rheumatoid arthritis: an Italian multicentric study and review of the literature,” Autoimmunity Reviews, vol. 11, no. 11, pp. 815–820, 2012.

[17] F. C. Arnett, S. M. Edworthy, D. A. Bloch et al., “The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis,” Arthritis and Rheumatism, vol. 31, no. 3, pp. 315–324, 1988.

[18] A. Doria, P. Vesco, F. Zulian, and P. F. Gambari, “The 1982 ARA/ACR criteria for the classification of systemic lupus erythematosus in pediatric and adult patients,” Clinical and Experimental Rheumatology, vol. 12, no. 6, pp. 689–690, 1994.

[19] S. van der Linden, H. A. Valkenburg, and A. Cats, “Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria,” Arthritis and Rheumatism, vol. 27, no. 4, pp. 361–368, 1984.

[20] A. T. Masi, G. P. Rodnan, and T. A. Medsger, “Preliminary criteria for the classification of systemic sclerosis (scleroderma). Subcommittee for scleroderma criteria of the American Rheumatism Association Diagnostic and Therapeutic Criteria Committee,” Arthritis and Rheumatism, vol. 23, no. 5, pp. 581–590, 1980.
[21] M. L. L. Prevoo, M. A. Van ’T Hof, H. H. Kuper, M. A. van Leeuwen, L. B. A. van de Putte, and P. L. C. M. van Riel, "Modified disease activity scores that include twenty-eight-joint counts: development and validation in a prospective longitudinal study of patients with rheumatoid arthritis," *Arthritis and Rheumatism*, vol. 38, no. 1, pp. 44–48, 1995.

[22] M. Cardiel, M. Abello-Banfi, R. Ruiz-Mercado, and D. Alarcón-Segovia, "How to measure health status in rheumatoid arthritis in non-English speaking patients: validation of a Spanish version of the Health Assessment Questionnaire Disability Index (Spanish HAQ-DI)," *Clinical and Experimental Rheumatology*, vol. 11, no. 2, pp. 117–121, 1993.

[23] A. M. El-Barbary, E. M. Kassem, M. A. S. El-Sergany, S. A.-M. Essa, and M. A. Eltomey, "Association of anti-modified citrullinated vimentin with subclinical atherosclerosis in early rheumatoid arthritis compared with anti-cyclic citrullinated peptide," *Journal of Rheumatology*, vol. 38, no. 5, pp. 828–834, 2011.

[24] A. Al-Shukaili, S. Al-Ghafri, S. Al-Marhoobi, and J. Alkaabi, "Evaluation of anti-mutated citrullinated vimentin antibodies, anti-cyclic citrullinated peptide antibodies and rheumatoid factor in omani patients with rheumatoid arthritis," *International Journal of Rheumatology*, vol. 2012, Article ID 285854, 5 pages, 2012.

[25] C. H. C. Maraina, A. K. Nurdayana, D. Rusni, and Y. Azwany, "Diagnostic value of anti-modified citrullinated vimentin in rheumatoid arthritis," *International Journal of Rheumatic Diseases*, vol. 13, no. 4, pp. 335–339, 2010.

[26] S. Rantapää-Dahlqvist, B. A. W. De Jong, E. Berglin et al., "Antibodies against cyclic citrullinated peptide and iga rheumatoid factor predict the development of rheumatoid arthritis," *Arthritis and Rheumatism*, vol. 48, no. 10, pp. 2741–2749, 2003.

[27] L. Mathsson, M. Mullazehi, M. C. Wick et al., "Antibodies against citrullinated vimentin in rheumatoid arthritis: higher sensitivity and extended prognostic value concerning future radiographic progression as compared with antibodies against cyclic citrullinated peptides," *Arthritis and Rheumatism*, vol. 58, no. 1, pp. 36–45, 2008.

[28] E. L. Gomez, S. C. Gun, S. D. Somananth, K. Chinna, and A. K. Radhakrishnan, "Ethnic differences in the prognostic utility of rheumatoid factor isotypes and anticyclic citrullinated peptides in rheumatoid arthritis patients: a cross-sectional study," *Modern Rheumatology*, vol. 23, no. 4, pp. 716–721, 2013.

[29] P. Nicaise-Roland, L. Nogueira, C. Demattei et al., "Autoantibodies to citrullinated fibrinogen compared with anti-MCV and anti-CCP2 antibodies in diagnosing rheumatoid arthritis at an early stage: data from the French ESPOIR cohort," *Annals of the Rheumatic Diseases*, vol. 72, no. 3, pp. 357–362, 2013.

[30] E. R. Vossenaar, N. Després, E. Lapointe et al., "Rheumatoid arthritis specific anti-Sa antibodies target citrullinated vimentin," *Arthritis Research & Therapy*, vol. 6, no. 2, pp. R142–150, 2004.

[31] G. A. Schellekens, B. A. W. de Jong, F. H. J. van den Hoogen, L. B. A. van de Putte, and W. J. van Venrooij, "Citrulline is an essential constituent of antigenic determinants recognized by rheumatoid arthritis-specific autoantibodies," *Journal of Clinical Investigation*, vol. 101, no. 1, pp. 273–281, 1998.

[32] K. N. Verpoort, K. Cheung, A. Ioan-Facsinay et al., "Fine specificity of the anti-citrullinated protein antibody response is influenced by the shared epitope alleles," *Arthritis and Rheumatism*, vol. 56, no. 12, pp. 3949–3952, 2007.

[33] O. Vittecoq, B. Incaurregarat, F. Jouen-Beades et al., “Autoantibodies recognizing citrullinated rat filaggrin in an ELISA using citrullinated and non-citrullinated recombinant proteins as antigens are highly diagnostic for rheumatoid arthritis," *Clinical and Experimental Immunology*, vol. 135, no. 1, pp. 173–180, 2004.

[34] A. Saraux, J. M. Berthelot, V. Devauchelle et al., "Value of antibodies to citrulline-containing peptides for diagnosing early rheumatoid arthritis," *Journal of Rheumatology*, vol. 30, no. 12, pp. 2535–2539, 2003.

[35] Z. Szekanecz, Z. Szabó, M. Zeher et al., "Superior performance of the CCP3.1 test compared to CCP2 and MCV in the rheumatoid factor-negative RA population," *Immunologic Research*, vol. 56, no. 2-3, pp. 439–443, 2013.

[36] K. Shidara, E. Inoue, E. Tanaka et al., “Comparison of the second and third generation anti-cyclic citrullinated peptide antibody assays in the diagnosis of Japanese patients with rheumatoid arthritis," *Rheumatology International*, vol. 31, no. 5, pp. 617–622, 2011.