The diagnostic utility of pleural markers for tuberculosis pleural effusion

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Abstract: Tuberculosis pleural effusion (TPE) is common in clinical practice, and its diagnosis remains a challenge for clinicians. Ziehl-Neelsen staining, PE Mycobacterium tuberculosis culture, and biopsy are the gold standards for TPE diagnosis; however, they are time-consuming, invasive, observer-dependent, and insensitive. PE markers represent a rapid, low-cost, and non-invasive objective diagnostic tool for TPE. In the past decades, several PE biomarkers have been developed, and their diagnostic accuracy has been evaluated in many studies. Here, we reviewed the literature to summarize the diagnostic accuracy of these biomarkers, especially using the evidence from systematic review and meta-analysis. The current research strongly suggests that adenosine deaminase (ADA), interferon-gamma (IFN-γ), and interleukin 27 (IL-27) have extremely higher diagnostic accuracy for TPE, while the diagnostic accuracy of interferon gamma release assays (IGRAs), tumor necrosis factor-α (TNF-α), and interferon-γ-induced protein 10 kDa (IP-10) is moderate. Although some evidence supports C-X-C motif chemokine ligand 9 (CXCL9), CXCL11, CXCL12, sFas ligand, angiotensin-converting enzyme (ACE), calpain-1, spectrin breakdown products (SBDP), matrix metalloproteinase-1 (MMP-1), soluble CD26 (sCD26), soluble interleukin 2 receptor (sIL-2R) as useful diagnostic markers for TPE, more support is needed to validate their diagnostic accuracy. Finally, nucleic acid amplification tests (NAATs) have extremely high diagnostic specificity, but their sensitivity is low. Taken together, ADA is the preferred marker for TPE because of its low cost and suitability for standardization.

Keywords: Tuberculosis pleural effusion (TPE); diagnostic accuracy; sensitivity; specificity; pleural markers

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Introduction

Tuberculosis pleural effusion (TPE) is common in clinical practice, and separating it from other types of pleural effusion (PE), such as malignant pleural effusion (MPE), parapneumonic pleural effusion (PPE), and transudate, is often challenging (1). PE Mycobacterium tuberculosis culture, Ziehl-Neelsen staining, pleural biopsy, either guided by medical thoracoscopy, computed tomography, or ultrasound, are the gold standards for TPE diagnosis. However, these diagnostic tools have some disadvantages. PE Mycobacterium tuberculosis culture is time- and labor-consuming and thus...
does not facilitate timely diagnosis. PE Ziehl-Neelsen staining has high specificity for TPE diagnosis, but its sensitivity is unsatisfactory (2). Pleural biopsy is an invasive tool and its operation-related complications are problematic (3). Further, the accuracy of PE Ziehl-Neelsen staining and biopsy is largely affected by the skill and experience of the operator and observer.

Pleural biomarkers, in contrast to culture, staining, and biopsy, represent an inexpensive, non-invasive, rapid, and objective diagnostic tool for TPE (4). To date, many pleural biomarkers have been developed for TPE diagnosis. Here, we conducted a literature review to summarize the diagnostic accuracy of available pleural biomarkers, with particular attention paid to the results from systematic review and meta-analysis.

**Adenosine deaminase (ADA)**

The first report investigating ADA for TPE diagnosis was published in 1978 by Piras et al. (5). The authors enrolled 54 patients with PE (21 of them are TPE) and found that both the sensitivity and specificity of ADA were 100% at a threshold of 40 IU/L (5). Subsequently, many studies have investigated the diagnostic accuracy of ADA for TPE, but the results were varied.

In 2003, two studies systematically reviewed the diagnostic accuracy of ADA (6,7) and pooled the results using meta-analysis. They reported that both the sensitivity and specificity of ADA were higher than 0.90. Notably, the method used in these two meta-analyses are based on summary receiver operating characteristic (sROC), which is not recommended in meta-analysis of diagnostic test accuracy study (8,9). In 2008, a meta-analysis (10) including 63 studies with 2,796 TPE and 5,297 non-TPE cases reported that the sensitivity of specificity of ADA for TPE were 0.90 and 0.92, respectively, with the area under sROC curve (AUC) being 0.96. An subsequent meta-analysis published in 2019 validated the meta-analysis published in 2008 (11). Another meta-analysis published in 2014 pooled the results of the studies published between 1990 and 2014 (12). A total of 12 studies were included, and the authors found that the sensitivity, specificity, and AUC of ADA were 0.86, 0.88, and 0.93, respectively (12). The low sensitivity and specificity in this meta-analysis may be due to the fact that the literature searching strategy in this meta-analysis had some limitations. In fact, some studies included in both a later (11) and previous (10) study were missed by this meta-analysis. The researcher included 174 studies comprising 10696 TPEs 16313 non-TPEs (11) and found that the pooled sensitivity and specificity and AUC were 0.90 and 0.92, respectively. Notably, a large portion of studies set the threshold between 30 and 50 IU/L (7,10,11).

Some meta-analyses have been performed to investigate the diagnostic accuracy of ADA in Indian (13), Spanish (14), and Brazilian (15) studies. All these studies reported both a sensitivity and specificity of ADA at around 0.90, indicating that the country source of PE has limited effects on the diagnostic accuracy of ADA.

**Interferon-gamma (IFN-γ)**

The first study dealing with the diagnostic accuracy of IFN-γ for TPE was published in 1988 (16). The authors found that pleural INF-γ level was significantly higher than MPE, PPE, and transudate (16). Notably, sera INF-γ was dramatically lower than that in PE, and there were no statistical differences between TPE, MPE, PPE and transudate, indicating that pleural INF-γ is produced locally in pleural compartment and does not arise due to the passive diffusion from sera (17). Subsequently, some studies investigated the diagnostic accuracy of pleural INF-γ and yielded similar results (17-19).

The first meta-analysis investigating the diagnostic accuracy of IFN-γ was published in 2003 (6). Because this meta-analysis used sROC to calculate sensitivity and specificity, other metrics such as positive and negative likelihood ratios (PLR and NLR) and diagnostic odds ratio (DOR) were not reported. In 2007, a comprehensive meta-analysis included 22 studies with 782 TPE and 1,319 non-TPE patients (20). The pooled sensitivity and specificity of IFN-γ were 0.89 and 0.97, respectively. Also, the AUC was 0.98, indicating that IFN-γ has extremely higher diagnostic accuracy for TPE.

During the past decade, many studies have been performed to investigate the diagnostic accuracy of IFN-γ TPE (17-19), but no updated meta-analysis has been published so far. Therefore, an updated meta-analysis is...
needed to provide new evidence.

**Interleukin 27 (IL-27)**

IL-27 is a member of the IL-12 family consisting of p28 and Epstein-Barr virus-induced gene 3 subunits (21). In 2012, Yang et al. were first to report that PF IL-27 level in TPE patients was significantly higher than that in MPE, PPE, and transudate (22). In their study, PE IL-27 had an extremely high diagnostic accuracy for TPE, with an AUC of 0.994 (22). At a threshold of 1,007 ng/L, the sensitivity and specificity were 0.927 and 0.991, respectively (22). In addition, sera IL-27 was not increased in TPE patients, indicating that high PE IL-27, like IFN-γ, is produced locally, rather than being a product of passive diffusion from sera to the pleural compartment (22). Using flow cytometry, the researchers found that pleural CD4+ T cells, CD8+ T cells, macrophages, monocytes, NKT cells, B cells, and mesothelial cells are major sources of IL-27 (22).

The high diagnostic accuracy of IL-27 has been validated by further studies (17,23,24). As of now, three meta-analyses have been performed to investigate the diagnostic accuracy of PE IL-27 for TPE (25-27). All these studies have demonstrated that PE IL-27 had an AUC more than 0.95, and both sensitivity and specificity were higher than 0.90, also indicating that pleural IL-27 has extremely high diagnostic accuracy for TPE. In another meta-analysis, Liu et al. investigated the ability of pleural IL-27 for differentiating TPE and MPE (28). They found that the overall diagnostic accuracy of pleural IL-27 was high, with both a sensitivity and specificity of 0.97 (28).

**Tumor necrosis factor-α (TNF-α)**

TNF-α is a well-known inflammatory factor involved in infectious disease. Early research indicated that protein-peptidoglycan complex and lipoarabinomannan have the ability to upregulate the expression of TNF-α in pleural fluid mononuclear cells (29). A later study reported that pleural TNF-α level was higher in TPE than MPE and transudates (30). This result indicates that pleural TNF-α is a potential diagnostic marker for TPE.

This first study investigating the diagnostic accuracy of TNF-α for TPE was published in 1996 (31). Subsequently, several studies have investigated the accuracy of TNF-α for TPE (32-34). In 2015, the first meta-analysis on the diagnostic accuracy of TNF-α for TPE was published (35). The authors included 7 studies and found that the sensitivity and specificity of TNF-α were 0.89 and 0.82, respectively. In another meta-analysis published in 2016, the authors included 12 studies and reported a sensitivity and specificity for TNF-α of 0.85 and 0.80, respectively (36). However, it should be noted that some studies also found that pleural TNF-α in TPE is not significantly higher than non-TPE (37,38). Therefore, more evidence is needed to evaluate the diagnostic accuracy of TNF-α.

**Nucleic acid amplification tests (NAATs)**

NAATs, which are based on polymerase chain reaction (PCR), have been widely used in tuberculosis diagnosis. Many types of specimens can be used in NAATs, including cerebrospinal fluid, tissue, urine, and PE. For TPE, both tissue and PE can be used for NAATs. Here, we focused on PE because of its invasiveness. NAATs are categorized as commercial or in-house tests. In 2004, a meta-analysis investigated the diagnostic accuracy of both the commercial and in-house NAATs for TPE, and found that the sensitivities were 0.62 and 0.71, and the specificities were 0.98 and 0.93, respectively (39). This result indicates that NAAT is useful for TPE in confirming, but not ruling out, a diagnosis.

Xpert® MTB/RIF, a commercial NAAT, has been widely used in clinical practice. Many meta-analyses have been performed to evaluate the diagnostic accuracy of Xpert® MTB/RIF for TPE (40-45). All of them found the specificity of Xpert® MTB/RIF to be around 0.99, but with a sensitivity only around 0.3 and 0.5. Notably, in two studies, the authors investigated the diagnostic accuracy of Xpert in pleural fluid versus composite reference standard (CRS), and both of them found a sensitivity around 0.20 (42,43).

**Interferon gamma release assays (IGRAs)**

Lymphocytes from tuberculosis patients have previously been exposed to *Mycobacterium tuberculosis*, and there
are many memory lymphocytes in circulation or in the pleural compartment. These memory lymphocytes, either in circulating or pleural effusion, are hypersensitive to mycobacterial antigen stimulation (e.g., ESAT-6, CFP-10). When stimulated with mycobacterial antigens, lymphocytes of TB infected patients release more interferon-γ than the lymphocytes of uninfected patients (46). Therefore, interferon-γ released by lymphocytes is an indicator of tuberculosis infection. Two types of commercially available kits have been developed, namely Quanti-FERON-TB Gold and T-SPOT-TB. The former one uses enzyme-linked immunosorbent assay (ELISA) to detect interferon-γ while the latter one uses enzyme-linked immunosorbent spot (ELISpot).

IGRA was first developed for latent TB diagnosis, but some studies have investigated the diagnostic accuracy of IGRA for TPE. The first meta-analysis of IGRA was published in 2008 (47). The author included 8 cohorts and found that the sensitivity, specificity, and AUC of pleural IGRA were 0.75, 0.82, and 0.88, respectively. The accuracy of blood IGRA is similar to that of pleural IGRA (47). In 2015, the meta-analysis was updated (48). The author included 16 cohorts with 516 TPE and 416 non-TPE patients and found that the sensitivity and specificity of pleural IGRA were 0.75 and 0.79, respectively. Interestingly, another three meta-analyses published in 2015 found that the diagnostic accuracy of IGRA was high (49-51), with a sensitivity between 0.82 and 0.94, and a specificity between 0.80 and 0.90. The AUC of the T-SPOT-TB test was higher than that of ELISA (0.98 vs. 0.84, respectively) (49). A meta-analysis only analyzed the diagnostic accuracy of T-SPOT TB and found that the sensitivity and specificity were 0.94 and 0.80, respectively (50). Notably, all three meta-analyses mentioned (49-51) included a large sample size study with extremely high diagnostic accuracy (52). This may explain the different diagnostic accuracy between these meta-analyses.

**Soluble Fas (sFas) ligand**

In 2001, Wu et al. were first to describe significantly higher PE sFas ligand in TPE than that of MPE and transudate (53). This result was reproduced by several subsequent studies (54-56). However, the results were not reproduced by Mitani et al., who found that patients with MPE had significantly higher sFas ligand than TPE (57). None of these studies evaluated the diagnostic accuracy of sFas ligand using metrics such as sensitivity, specificity, and AUC.

In 2010, Wu et al. evaluated the diagnostic accuracy of sFas ligand in pleural effusion (58). A total of 23 TPE and 56 non-TPE patients were enrolled, and the diagnostic performance of sFas ligand was evaluated by ROC curve. They found that, at a cutoff value of 39.85 pg/mL, the sensitivity and specificity of sFas ligand for TPE were 0.96 and 0.80, respectively. The AUC of sFas ligand was 0.88, which is not significantly inferior to that of ADA (0.91) and INF-γ (0.91). Subsequently, two studies (59,60) validated the findings of Wu et al. Both studies found that, at a cut-off value around 40 pg/mL, both the sensitivity and specificity were around 0.90, and the AUC of sFas ligand was comparable to that of ADA, IFN-γ, and interferon-γ-induced protein 10 kDa (IP-10) (60).

**C-X-C motif chemokine receptor 3 (CXCR3) ligands**

CXCR3 ligands are typical IFN-γ-induced proteins, and include three chemokines: CXCL9, CXCL10, and CXCL11. CXCL10, also known as IP-10, has been extensively investigated as a diagnostic marker for TPE. In 2005, Okamoto (61) was first to report that IP-10 has high diagnostic accuracy for TPE (AUC =0.93). This result was validated by subsequent studies (19,60,62-66). In 2017, a meta-analysis investigated the diagnostic accuracy of CXCL10 for TPE (67). The authors included 14 included studies with 715 TPEs and 667 non-TPEs and found that the sensitivity and specificity of CXCL10 were 0.84 and 0.90, respectively. The AUC under sROC of CXCL10 was 0.94.

Thus far, only one study has investigated the diagnostic accuracy of CXCL9 and CXCL11 for TPE (68). The researchers enrolled 336 subjects with PE (106 TPE and 230 non-TPE) and simultaneously evaluated the diagnostic accuracy of ADA, IFN-γ, CXCL9, and CXCL11. They found that these four markers had comparable diagnostic accuracy, with AUCs all higher than 0.95 (68).

The diagnostic accuracy of CXCL12 was assessed in 2012; the researchers included 60 PE patients, 15 of whom were TPE (69). At a threshold of 4,600 pg/mL,
the diagnostic sensitivity and specificity of CXCL12 were 0.60 and 0.93 respectively. Pleural CXCL12 concentration was approximately two times higher than that in sera, indicating that CXCL12 is produced locally in the pleural compartment. However, in another study, which set MPE as the control, the diagnostic accuracy of CXCL12 was fair, with an AUC of 0.69 (70), indicating that more evidence is needed to validate the diagnostic accuracy of CXCL12.

### Pleural fibrosis biomarkers

Pleural fibrosis is a common complication of TPE and occurs in approximately half the patients with TPE (71). Therefore, fibrosis-related markers are considered to be a useful diagnostic tool for TPE. In a recently published study, the researchers enrolled 47 TPE, 28 MPE, and 10 transudate patients (72). They reported that angiotensin-converting enzyme (ACE), calpain-1, spectrin breakdown products (SBDP), and matrix metalloproteinase-1 (MMP-1) were significantly higher in TPE. The AUCs of ACE, calpain, SBDP, and MMP-1 was 0.80, 0.72, 0.83, and 0.79, respectively (72). Given that this study did not include PPE, further studies are needed to validate the findings (73). Notably, in an early study, the authors reported that rheumatoid arthritis (RA) patients have higher ACE than TPE (74), indicating that RA-induced PE should be considered for patients with higher ACE.

### Soluble CD26 (sCD26)

CD26, also named dipeptidyl peptidase 4 (DPP4), is preferentially expressed on the T helper. The expression of CD26 in T helper is correlated with interferon-γ and thus represents a potential biomarker for TPE (75). There are two types of CD26, a membrane-bound form and a soluble form. In 2001, a study with a small sample size investigated the diagnostic accuracy of sCD26 for TPE (76). The authors set MPE, PPE, and heart failure as controls, and found that the sensitivity and specificity of sCD26 were 0.46 and 0.95, respectively (76). Subsequently, three studies have investigated the diagnostic accuracy of sCD26 for TPE (19,77,78). The sensitivity in these studies ranged from 0.46 to 0.87 while the specificity ranged from 0.82 to 0.95. None of these studies reported the AUC of sCD26.

### Soluble interleukin 2 receptor (sIL-2R)

Interleukin-2 (IL-2) is a cytokine that can promote the proliferation of active T-cells. Its receptor, which is expressed on the surface of activated T cells, can be released into circulation, which is termed sIL-2R (79). In the early 1990s, a study revealed that patients with pulmonary tuberculosis had higher sera sIL-2R compared with sarcoidosis and healthy volunteers, and anti-tuberculosis agents could decrease the sera sIL-2R level (80). Subsequent studies indicated that patients with TPE had significantly higher pleural sIL-2R than MPE, PPE, and transudate patients, and that pleural sIL-2R is positively correlated with ADA (81). These results suggest that pleural sIL-2R is a potential diagnostic marker for TPE.

In 1994, two studies investigated the diagnostic accuracy of sIL-2R TPE (82,83), and both revealed that pleural sIL-2R is a useful diagnostic marker for TPE. Subsequently, several studies have investigated the diagnostic accuracy of sIL-2R for TPE, but the results have been varied (84-88). The sensitivity of sIL-2R ranged from 0.74 to 0.91, and the specificity ranged from 0.31 to 1.00. Notably, a study reported that the specificity of sIL-2R is only 0.31 (85), which is obviously different from previous studies. The reason explaining difference between these studies needs to be identified in future studies.

### Conclusions

Table 1 summarizes the evidence from meta-analyses. Currently, the diagnostic accuracy of ADA, IFN-γ, IL-27, IGRA, TNF-α, NAAT, and IP-10 has been evaluated by meta-analysis. Among the available markers, ADA, IFN-γ, and IL-27 are the most promising. Considering that ADA has advantages of low cost and suitability for standardization, we recommend ADA for TPE diagnosis. The diagnostic accuracy of IGRA, TNF-α, and IP-10 seems inferior to that of ADA, IFN-γ, and IL-27. NAATs have extremely high diagnostic specificity, but their sensitivity is low.

Table 2 summarizes the evidence from single studies. Although some preliminary research attests to CXCL9, CXCL11, CXCL12, MMP-1, ACE, calpain-1, SBDP, sFas ligand, sCD26, and sIL-2R being useful diagnostic markers for TPE, the results are varied. Therefore, more evidence is
Table 1 Pleural markers for tuberculosis pleural effusion: evidence from meta-analyses

| Markers or tools, (reference) | Year | N   | TPE/non-TPE | Method | Sensitivity | Specificity | AUC  | PB |
|-------------------------------|------|------|-------------|--------|-------------|------------|------|----|
| ADA, (6)                      | 2003 | 31   | 1,621/3,117 | sROC   | 0.93        | 0.93       | –    | –  |
| ADA, (7)                      | 2003 | 40   | –           | sROC   | 0.92        | 0.92       | –    | –  |
| ADA, (10)                     | 2008 | 63   | 2,796/5,297 | REM    | 0.92        | 0.90       | 0.96 | Yes|
| ADA, (15)                     | 2008 | 9    | 857/817     | REM    | 0.92        | 0.88       | 0.97 | –  |
| ADA, (12)                     | 2014 | 12   | 865/1,379   | REM    | 0.86        | 0.88       | 0.93 | –  |
| ADA, (13)                     | 2016 | 40   | 2,058/1,466 | BVM    | 0.94        | 0.89       | 0.97 | No |
| ADA, (11)                     | 2019 | 174  | 10,696/16,313 | BVM  | 0.92        | 0.90       | –    | Yes|
| ADA, (14)                     | 2019 | 16   | 1,172/2,975 | BVM    | 0.93        | 0.92       | 0.97 | No |
| IFN-γ, (6)                    | 2003 | 13   | 419/770     | sROC   | 0.96        | 0.96       | –    | –  |
| IFN-γ, (20)                   | 2007 | 22   | 782/1,319   | REM    | 0.89        | 0.97       | 0.98 | Yes|
| IL-27, (25)                   | 2018 | 7    | 323/834     | REM    | 0.94        | 0.92       | 0.98 | Yes|
| IL-27, (26)                   | 2017 | 8    | 380/756     | BVM    | 0.93        | 0.95       | 0.95 | No |
| IL-27, (27)                   | 2017 | 9    | 425/807     | REM    | 0.92        | 0.90       | 0.97 | No |
| IL-27, (28)                   | 2018 | 7    | 285/265     | REM    | 0.93        | 0.97       | 0.99 | No |
| NAAT, commercial, (39)        | 2004 | 14   | 127/1,384   | REM    | 0.62        | 0.98       | –    | No |
| NAAT, in-house, (39)          | 2004 | 26   | 528/939     | REM    | 0.71        | 0.93       | –    | Yes|
| NAAT, Xpert® MTB/RIF, (42)    | 2014 | 14   | 92/749      | BVM    | 0.46        | 0.99       | –    | –  |
| NAAT, Xpert® MTB/RIF, (40)    | 2014 | 9    | 79/572      | BVM    | 0.34        | 0.98       | –    | –  |
| NAAT, Xpert® MTB/RIF, (44)    | 2015 | 13   | –           | BVM    | 0.37        | 0.98       | –    | –  |
| NAAT, Xpert® MTB/RIF, (43)    | 2016 | 21   | 760/1,407   | BVM    | 0.51        | 0.99       | 0.84 | No |
| NAAT, Xpert® MTB/RIF, (41)    | 2018 | 23   | 1,194/1,452 | BVM    | 0.30        | 0.99       | 0.86 | No |
| NAAT, Xpert® MTB/RIF, (45)    | 2018 | 27   | 607/3,309   | BVM    | 0.51        | 0.99       | –    | –  |
| IGRA, (47)                    | 2011 | 8    | 213/153     | REM    | 0.75        | 0.82       | 0.88 | No |
| IGRA,(48)                     | 2015 | 16   | 516/416     | BVM    | 0.75        | 0.79       | –    | Yes|
| IGRA, (49)                    | 2015 | 17   | 806/842     | REM    | 0.82        | 0.87       | 0.91 | Yes|
| IGRA, (50)                    | 2015 | 9    | –           | REM    | 0.94        | 0.80       | 0.97 | No |
| IGRA, (51)                    | 2015 | 9    | 549/309     | REM    | 0.93        | 0.90       | 0.96 | Yes|
| IP-10, (67)                   | 2017 | 14   | 715/667     | REM    | 0.84        | 0.90       | 0.94 | No |
| TNF-α, (35)                   | 2015 | 7    | 159/338     | REM    | 0.89        | 0.82       | 0.86 | No |
| TNF-α, (36)                   | 2016 | 12   | 399/623     | REM    | 0.85        | 0.80       | 0.89 | No |

–, not reported. N, number of included studies; TPE, tuberculosis pleural effusion; AUC, area under curve; PB, publication bias; ADA, adenosine deaminase; sROC, summary receiver operating characteristic; REM, random-effects model; BVM, bivariable model; INF-γ, interferon-gamma; IL-27, interleukin 27; NAAT, nucleic acid amplification tests; IGRA, interferon gamma release assay; IP-10, interferon-γ-induced protein 10 kDa; TNF-α, tumor necrosis factor-α.
needed to validate their diagnostic performance.

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**Footnote**

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**Table 2** Pleural markers for tuberculosis pleural effusion: evidence from the single studies

| Biomarker, (reference) | Year | Country | TPE/non-TPE | Threshold | Sensitivity | Specificity | AUC |
|------------------------|------|---------|-------------|-----------|------------|------------|-----|
| CXCL9, (68)            | 2017 | Korea   | 106/230     | 1,522.1 pg/mL | 0.97       | 0.90       | 0.98|
| CXCL11, (68)           | 2017 | Korea   | 106/230     | 151.5 pg/mL | 0.92       | 0.91       | 0.95|
| CXCL12, (69)           | 2012 | Japan   | 15/45       | 4,600 pg/mL | 0.84       | 0.60       | 0.93|
| CXCL12, (70)           | 2015 | China   | 44/39       | 3,710 pg/mL | 0.46       | 0.82       | 0.69|
| sFas ligand, (58)      | 2010 | China   | 23/56       | 39.85 pg/mL  | 0.96       | 0.80       | 0.88|
| sFas ligand, (59)      | 2019 | Poland  | 60/162      | 41.9 pg/mL  | 0.91       | 0.88       | 0.93|
| sFas ligand, (60)      | 2014 | Poland  | 44/159      | 45.0 pg/mL  | 0.95       | 0.90       | 0.95|
| ACE, (72)              | 2018 | Korea   | 47/38       | 47.16 ng/mL | 0.55       | 0.95       | 0.80|
| Calpain-1(72)          | 2018 | Korea   | 47.3/38     | 787 ng/mL   | 0.96       | 0.47       | 0.72|
| MMP-1, (72)            | 2018 | Korea   | 47/38       | 7.229 pg/mL | 0.74       | 0.66       | 0.79|
| SBDP, (72)             | 2018 | Korea   | 47/38       | 2.745 ng/mL | 0.87       | 0.79       | 0.83|
| sCD26, (76)            | 2001 | Japan   | 46/46       | 544.5 ng/mL | 0.46       | 0.95       | –   |
| sCD26, (77)            | 2015 | Spain   | 30/129      | 470 ng/mL   | 0.87       | 0.82       | –   |
| sCD26, (19)            | 2012 | China   | 78/44       | 75 ng/mL    | 0.89       | 0.82       | –   |
| sCD26, (78)            | 2009 | Turkey  | 18/69       | 27 IU/L     | 0.68       | 0.90       | –   |
| sIL-2R, (83)           | 1994 | China   | 42/69       | 4,291.4 U/mL| 0.81       | 1.00       | –   |
| sIL-2R, (82)           | 1994 | China   | 27/66       | 5,000 U/mL  | 0.74       | 0.94       | –   |
| sIL-2R, (84)           | 2000 | Spain   | 23/109      | 4,700 U/mL  | 0.91       | 0.95       | 0.96|
| sIL-2R, (85)           | 2002 | Greece  | 11/39       | 2,980 U/mL  | 0.91       | 0.31       | –   |
| sIL-2R, (86)           | 2003 | Japan   | 10/36       | 4,000 U/mL  | 0.90       | 0.97       | –   |
| sIL-2R, (87)           | 2004 | Japan   | 20/35       | –           | 0.90       | 0.97       | 0.99|
| sIL-2R, (88)           | 2015 | Turkey  | 52/68       | 4.8 ng/mL   | 0.83       | 0.71       | –   |

--, not reported. TPE, tuberculosis pleural effusion; AUC, area under curve; CXCL, C-X-C motif chemokine ligand; sFas, soluble Fas; ACE, angiotensin-converting enzyme; MMP-1, matrix metalloproteinase-1; SBDP, spectrin breakdown products; sCD26, soluble CD26; sIL-2R, soluble interleukin 2 receptor.
other conflicts of interest to declare.

**Ethical Statement:** The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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