Serum Calprotectin, a Marker of Neutrophil Activation, and Other Mediators of Inflammation in Response to Various Types of Extreme Physical Exertion in Healthy Volunteers

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Purpose: While extreme physical exertion is known to induce changes in the status of inflammation comparisons of the responses for various mediators of inflammation after acute bouts of high-intensity exercise have been limited.

Subjects and Methods: We examined the responses in serum levels of novel inflammatory proteins, calprotectin, suPAR, CD163, and pro- and anti-inflammatory cytokines in 12 physically active volunteers (10 men, 2 women, mean age 37±14 years) before and after completing various types of extreme physical exertion (marathon run, half-marathon run or 24-h cross-country skiing). For comparisons, the levels of the biomarkers were also measured at rest in 30 healthy controls (25 men, 5 women, mean age 42 ± 12 years) with low or sedentary activity.

Results: Extreme physical exertion induced significant increases in serum calprotectin (p < 0.0005), suPAR (p < 0.01), CD163 (p < 0.05), IL-6 (p < 0.0005), IL-8 (p < 0.01) and IL-10 (p < 0.0005) (pre- vs 3h-post-exercise). These responses were found to normalize within 48 hours. While the increases in blood leukocytes were of similar magnitude following the different types of exercise, markedly more pronounced responses occurred in serum TNF-α (p < 0.01), IL-8 (p < 0.01) and CD163 (p < 0.05) in those with more intense activity. In 3-h post-exercise samples significant correlations were observed between serum calprotectin and IL-6 (r = 0.720, p < 0.01), IL-10 (r = 0.615, p < 0.05), TNF-α (r = 0.594, p < 0.05), suPAR (r = 0.587, p < 0.05) and blood leukocytes (r = 0.762, p < 0.01).

Conclusion: The present results suggest distinct exercise-intensity dependent changes in mediators of inflammation (including calprotectin, suPAR and CD163) following extreme physical exertion. Our findings indicate that there is a major reversible impact of high-intensity physical exertion on the status of inflammation.

Keywords: acute exercise, cytokine, immune status, leukocyte, oxidative stress

Introduction

Although the health benefits brought about by regular physical activity are proved and wide ranging,¹,² the intensity and types of physical activity associated with either beneficial or adverse effects on the status of inflammation have, however, remained a matter of controversy.³⁻⁷ While regular physical activity with appropriate resting periods is expected to enhance immunity, acute bouts of vigorous activity may also lead to adverse consequences.³⁻⁶,⁸ Several lines of evidence indicate that strenuous exercise is associated with robust inflammatory responses,
mobilization of leukocytes and increases in the levels of inflammatory mediators in circulation.\textsuperscript{4,6,9-15} However, the intensity and duration of exercise needed to elicit clinically notable changes in the mediators of inflammation remain unclear.\textsuperscript{1-6}

Previous studies have indicated that high-intensity physical activity leads to a release of both pro-inflammatory and anti-inflammatory cytokines.\textsuperscript{4,6,12} While recent advances in inflammation research have provided a wide range of new and emerging serum biomarkers of inflammation, only limited information is available on the levels of such markers in response to exercise. Serum calprotectin is secreted from neutrophil granulocytes and may serve as a marker of neutrophil-driven inflammation.\textsuperscript{16-19} Its release has been linked to both anti-infective and anti-inflammatory properties including control of myelopoiesis, chelation of divalent cations, scavenging of reactive oxygen species, chemotaxis and direct antimicrobial action.\textsuperscript{16-19} Soluble urokinase plasminogen activator receptor (suPAR) is expressed on many immunologically active cells, including monocytes, neutrophils and activated T cells.\textsuperscript{20-22} CD163, an endocytic receptor protein for haptoglobin-hemoglobin complexes, is found on macrophages and monocytes: its serum levels increase in conjunction with the involvement of macrophages and free-radical induced tissue damage.\textsuperscript{23,24} Both suPAR and CD163 proteins have emerged as prospective risk factors for adverse clinical outcomes in various inflammatory conditions.\textsuperscript{20-24}

In this study, we assessed the magnitude and patterns of acute changes in serum calprotectin with those of suPAR, CD163 and pro- (IL-6, IL-8, TNF-\textalpha) and anti-inflammatory (IL-10, TGF-\beta) cytokines in a group of 12 healthy volunteers participating in various types of high-intensity exercise. For additional comparisons, these biomarkers were also measured in a group of 30 apparently healthy age- and sex-matched individuals with low or sedentary activity. Our findings support a major reversible impact of high-intensity exercise on the status of inflammation.

**Subjects and Methods**

**Study Subjects**

The sample of individuals attending the different types of strenuous exercise events consisted of 12 healthy physically active volunteers (10 men, two women, mean age 37 ± 14 years), who had no history of diseases or medication at the time of the study. These subjects attended either a marathon run (ie 42.2 km, n = 4) or half-marathon run (ie 21.1 km, n = 7) or in one case, a 24-h skiing event (Table 1). The participants in the skiing event and the full marathon run were healthy endurance trained fitness enthusiasts whereas the half-marathon runners were mainly recreational runners. The details of the exercise protocols and associated medical surveillance have been described previously.\textsuperscript{12,25} The typical weekly physical activity of the participants prior to the experiment had ranged from three to 15 hours of moderate to high intensity aerobic activity per week. Medical examinations were carried out before undertaking the intensive exercise, within two hours after finishing and one week after the trial. All participants were allowed to follow their individual food strategies prior to the experiment and in the restitution period. A two-day rest before the trial was recommended. During the experiment, liberal amounts of various fluids and food (mostly exogenous carbohydrates) were available.

**Table 1 Main Characteristics of Study Subjects Including Physically Active Individuals Participating in Various Types of Extreme Physical Exertion and Individuals with Low/Sedentary Activity**

| Subjects with Extreme Physical Exertion | Exercise Timing Characteristics | Subjects with Low/Sedentary Activity |
|---------------------------------------|---------------------------------|-------------------------------------|
| n = 12                                | Speed (km/h) Mean ± SD | Pace (Min/km) Mean ± SD |
| Age, years, mean ± SD                 | 36.5 ± 13.7                  | 42.2 ± 12.3                         |
| Males, n (%)                          | 10 (83.3%)                   | 25 (83.3%)                          |
| Exercise type, n (%)                  |                               |                                     |
| Marathon run                          | 4 (33.3%)                    | 4:43 ± 0:13                         |
| Half-marathon run                     | 7 (58.3%)                    | 5:37 ± 0:50                         |
| 24-h Cross-country skiing             | 1 (8.3%)                     | 3:32                                |
|                                      | 12.7 ± 0.6                   | 10.9 ± 1.7                          |
|                                      | 17.0                          |                                     |
ad libitum. After the trial, two of the subjects received intravenous hydration (approximately 300 mL/h) for the next 20 hours while being monitored in a hospital setting. Post-exercise symptoms ranged from muscle pain and stiffness to signs of generalized fatigue and nausea. Both the self-concepts expressed by the participants and the medical and psychological post-exercise assessments suggested a rapid recovery in all subjects, which was completed within one week with no apparent signs of residual emotional or physiological exhaustion.

Blood specimens were collected on the day before and at 3 and 48 hours post-exercise by a trained laboratory nurse using previously described standard operating procedures.\cite{12,25} For additional comparisons of the biomarker baseline levels in less physically active individuals, blood samples at rest were also collected from a reference population including 30 healthy but sedentary individuals (25 men, 5 women, mean age 42 ± 12 years), whose physical activity was typically less than 150 minutes of light intensity activity per week over a period of one year prior to sampling. Serum was separated by centrifugation (1500 × g for 10 minutes) and stored frozen at −70 °C prior to the analyses.

All subjects were informed about the study and gave their written informed consent for participation. The protocol was approved by the local ethical committee (Northern Ostrobothnia Hospital District Institutional Review Board, Oulu, Finland). The study was conducted according to the provisions of the Declaration of Helsinki.

**Laboratory Analyses**

Serum was separated by centrifugation (1500 × g for 10 minutes) and stored at −70 °C prior to the measurement of the various biomarkers. Serum calprotectin concentrations were measured using BÜHLMANN MRP8/14 ELISA kit according to the instructions of the manufacturer (BÜHLMANN Laboratories AG, Schönenbuch, Switzerland). Serum suPAR levels were measured using the suPARnostic enzyme-linked immunosorbent assay (ELISA) kit according to the instructions of the manufacturer (Virogates, Birkerød, Denmark). The measurements of CD163 were carried out using Quantikine human CD163 ELISA assay (R&D Systems, Abingdon Science Park, UK). The concentrations of interleukins (IL-6, IL-8, IL-10, TNF-α and TGF-β) in serum were determined using Quantikine high sensitivity ELISA kits (R&D Systems, Abingdon, Science Park, UK). Routine blood chemistry analyses were carried out using standard clinical chemical methods on an Abbott Architect c8000 automated clinical chemistry analyser (Abbott Diagnostics, Abbott Laboratories, Abbott Park, IL, USA) and blood cell counts were performed using a Sysmex XE-5000 automated Hematology Analyser. All measurements were carried out in SFS-EN ISO 15189:2013 accredited laboratory.

**Statistical Methods**

The values are reported as mean ± standard deviation (SD) or medians and interquartile ranges, as indicated. The levels of different mediators of inflammation were compared between the study groups using the Mann–Whitney U-test. The comparisons between the different time points related to physical exercise were made using paired samples t-test. Both absolute changes and relative changes from baseline levels were calculated for each parameter. Relative changes were based on the pre-exercise values of the subjects and are expressed as a percentage (% change=100*absolute change/pre-exercise value). Comparisons of the changes were done using the Mann–Whitney U-test. Spearman’s rank correlation coefficient (r_s) was used for evaluating the correlations between the study variables. The correlations between postrace values were calculated after an adjustment for baseline variability using percent change (pre–post) for each outcome variable. A p-value of < 0.05 was considered statistically significant. Statistical analyses were carried out using IBM SPSS Statistics 24.0 (Armonk, NY: IBM Corp.).

**Results**

The details of these exercise protocols and associated medical surveillance have been described previously.\cite{12,25} Table 1 summarizes the main characteristics of the study subjects and timing details for each type of long-term endurance exercise events. The efforts by all participants represented exercise stress of over 90 minutes at >70% of VO₂ max and the self-reported ratings of perceived exertion (RPE) ranged from hard to maximal exertion (Borg scale ≥15 on a 6 to 20 rating scale).

The medians and interquartile ranges of the main biomarkers of inflammation before and after the acute bouts of exercise are summarized in Figure 1. When compared to pre-exercise baseline levels, significant 3-h post-exercise increases occurred in serum calprotectin (p < 0.0005), suPAR (p < 0.01), CD163 (p < 0.05), IL-6 (p < 0.0005), IL-8 (p < 0.01) and IL-10 (p < 0.01). All parameters returned to baseline levels within 48 hours (Figure 1, Table 2). Although the values of IL-6, suPAR and TGF-β tended to
be lower in the physically active group at baseline than those in the blood samples taken at rest from a reference group of the individuals with low or sedentary activity (Figure 1, Table 2), the differences did not reach statistical significance.

Comparisons of the relative increases in the mediators of inflammation in relation to the type of the exercise (based on the percentage change from the pre-race values) are summarized in Table 3. While the increases in blood leukocytes following the different types of exercise were similar, markedly more pronounced responses in those with more intense activity were found to occur in serum TNF-α (p < 0.01), IL-8 (p < 0.01) and CD163 (p < 0.05) (Table 3). In addition, striking increases in serum calprotectin, IL-6 and TGF-β were observed in the individual who completed the most extreme and prolonged exercise (24-h skiing) (Table 3).

The changes in serum calprotectin (3-h post-exercise and adjusted for baseline levels), correlated with those of IL-6 (rₚ = 0.720, p < 0.01), IL-10 (rₚ = 0.615, p < 0.05), TNF-α (rₚ = 0.594, p < 0.05), suPAR (rₚ = 0.587, p < 0.05) and blood leukocytes (rₚ = 0.762, p < 0.01). Comparisons of the relative changes in the mediators of inflammation and biochemical indices of rhabdomyolysis or other possible adverse health effects of extreme exercise indicated significant correlations between the change in TNF-α and creatine kinase (CK) (rₛ = 0.667, p < 0.05), aspartate aminotransferase (AST) (rₛ = 0.733, p < 0.05) and C-reactive protein (CRP) (rₛ = 0.670, p < 0.05). The changes in IL-8 also correlated with those of TNF-α (rₛ = 0.883, p < 0.001), CK (rₛ = 0.633, p < 0.05), AST (rₛ = 0.733, p < 0.05) and CD163 (rₛ = 0.700, p < 0.05).

**Discussion**

Our study comparing serum calprotectin, a marker of neutrophil activation, with other biomarkers of inflammation after extreme exercise demonstrates distinct characteristics in the patterns of exercise-induced immune responses. The data also indicates differences in the magnitude of the responses in proportion to the intensity and duration of the exercise.
Although the beneficial health effects of physical activity in proper amounts have been well established, extreme physical exertion may also trigger adverse consequences on immune function.1–7,26–28 The present data indicates that serum calprotectin, IL-6 and IL-10 respond to an acute bout of physical exercise in a highly sensitive manner. The findings of increased IL-6 and IL-10 in both marathon and half-marathon finishers is in accordance with recent findings by Nielsen and coworkers from Norway.6 Post-exercise increases in serum calprotectin also coincide with increased leukocyte counts suggesting that calprotectin could also serve as a biomarker for myeloid reactions when analyzing the body’s immune response to exercise. Calprotectin release may indicate exercise-induced activation of neutrophil-driven inflammation, which have previously been linked with anti-inflammatory processes and scavenging of tissue-damaging reactive oxygen species.16,18 A switch of the immune system towards either pro- or anti-inflammatory consequences in association with extreme physical exertion may be influenced by multiple possible precipitating factors, such as changes in the extracellular pH during high-intensity exercise17 or the status of regular exercise training.29 Calprotectin could play a pivotal regulatory role in controlling the inflammatory cascades and leukocyte trafficking to yield more coordinated immune reactions.17–19,29 Thus, it could also contribute to the elicitation of an appropriate inflammatory response and prevention of tissue damage.18,19,29

Previous studies comparing multiple serum cytokines in response to acute bouts of exercise have yielded variable findings.3,4,6,8,12 While some studies have found increases in IL-6, IL-8 and TNF-α levels, other studies have shown relatively stable levels of these biomarkers. The intensity, type, duration and familiarity of the exercise may all play a role in explaining such divergent findings.4,28 Studies on the relationships between physical activity and the status of inflammation have indicated that even rather short sessions of regular exercise can reduce inflammation through downregulation of TNF-α production via beta-adrenergic activation.30 While IL-6 levels after acute exercise appear to increase more than other

### Table 2 Mean Absolute and Relative Changes (%) in Mediators of Inflammation as Compared to Baseline (Pre-Exercise) Levels of the Physically Active Individuals

| Physically Active Subjects | Pre-Exercise | 3-h Post | 48-h Post | Subjects with Low/Sedentary Activity |
|----------------------------|--------------|----------|-----------|------------------------------------|
| Mean ± SD Exercise-Induced Change from Baseline, Units (%) | Mean ± 5 SD Exercise-Induced Change from Baseline, Units (%) | Mean ± SD Exercise-Induced Change from Baseline, Units (%) | Difference from Baseline *, Units (%) |
| Calprotectin | 1.8 ± 2.0 μg/mL | +5.8 (+320%) *** | −0.3 (−19%) | −0.4 (−23%) |
| suPAR | 1.8 ± 0.3 ng/mL | +0.6 (+33%) ** | −0.04 (−2%) | +0.8 (+43%) |
| CD163 | 528 ± 208 ng/mL | +142 (+27%) | +0.7 (+32%) | +30 (+6%) |
| IL-6 | 2.2 ± 1.8 pg/mL | +143 (+648%) *** | +0.5 (+22%) | +0.7 (+32%) |
| IL-8 | 12.0 ± 4.2 pg/mL | +12.1 (+101%) ** | −0.7 (−5%) | +2.3 (+19%) |
| IL-10 | 0.6 ± 0.1 pg/mL | +0.8 (+139%) *** | −0.06 (−11%) | −0.02 (−4%) |
| TNF-α | 0.08 ± 0.2 pg/mL | +0.3 (+45%) | +0.2 (+23%) | +0.04 (+5%) |
| TGF-β | 516 ± 284 pg/mL | +68 (−13%) | +85 (+16%) | +135 (+26%) |
| Leukocytes | 5.9 ± 2.210⁷/l | +8.2 (−139%) | +0.2 (+3%) | −0.1 (−2%) |

Notes: Mean relative change significantly different from baseline (pre-exercise) values. (Mann–Whitney U-test), **p < 0.01, ***p < 0.001; *Mean difference compared to pre-exercise levels of the physically active individuals.

Abbreviations: suPAR, soluble urokinase plasminogen activator receptor; a biomarker of immune activation; CD163, a biomarker of monocyte-macrophage activation; IL, interleukin; TNF-α, tumor necrosis factor-alpha; TGF-β, transforming growth factor-beta.

### Table 3 Relative Changes (% from Pre-Exercise Levels) in Mediators of Inflammation After Various Types of Exertion

| Marathon Run | Half-Marathon Run | 24-h Skiing |
|--------------|------------------|------------|
| Calprotectin | +342             | +304       | +1427     |
| suPAR        | +53              | +20        | +51       |
| CD163        | +71 *            | +9         | +4        |
| IL-6         | +687             | +561       | +2178     |
| IL-8         | +252 **          | +18        | +251      |
| IL-10        | +228             | +95        | +40       |
| TNF-α        | +160 **          | −28        | +140      |
| TGF-β        | −24              | −3         | +721      |
| Leukocytes   | +128             | +148       | +132      |

Notes: Relative change significantly different between marathon run and half-marathon run (Mann–Whitney U-test), *p < 0.05, **p < 0.01.

Abbreviations: suPAR, soluble urokinase plasminogen activator receptor; a biomarker of immune activation; CD163, a biomarker of monocyte-macrophage activation; IL, interleukin; TNF-α, tumor necrosis factor-alpha; TGF-β, transforming growth factor-beta.
moderate weekly physical activity is generally associated with lower levels of IL-6, IL-8, TNF-α and calprotectin following the most intense activity. The relatively paracrine changes observed here in pro- and anti-inflammatory cytokines, IL-6, IL-8, TNF-α and anti-inflammatory cytokines, IL-10, TGF-β pathways following strenuous physical activity is also in accordance with the view that a coordinated balance between pro- and anti-inflammatory cascades is needed to counteract exercise-induced inflammatory threat and tissue damage. Both IL-6 and IL-10 can stimulate the Th2 pathway, activate anti-inflammatory cascades and inhibit TNF-α.

Long-term strenuous exercise was also found here to be associated with significant increases in certain serum proteins, which have previously been established as indicators of harmful responses and disease risks in various inflammatory conditions. suPAR is elevated in patients with severe systemic inflammation and the elevated levels of this biomarker show an association with poorer outcomes in such conditions. While IL-6 has been usually regarded as a pro-inflammatory cytokine, it may also stimulate the production of anti-inflammatory cytokines. IL-6 secretion also appears to be associated with the regulation of the acute phase response, hematopoiesis and tissue regeneration. The relatively parallel changes observed here in pro- (IL-6, IL-8, TNF-α) and anti-inflammatory (IL-10, TGF-β) pathways following strenuous physical activity is also in accordance with the view that a coordinated balance between pro- and anti-inflammatory cascades is needed to counteract exercise-induced inflammatory threat and tissue damage. Both IL-6 and IL-10 can stimulate the Th2 pathway, activate anti-inflammatory cascades and inhibit TNF-α.

In light of exercise-induced changes in CD163, it is of interest to note that CD163 has been shown to play a role in innate immune defense by sequestering hemoglobin-bound iron and scavenging oxidative stress-induced by-products of hemoglobin.

Further studies appear warranted to examine whether the use of current biomarkers could be of value in the follow-up of immune function in physically active individuals engaged in frequent episodes of strenuous exercise. It may be assumed that skewed balances in the ratios of pro- and anti-inflammatory cytokines correlate with unfavorable over-activation of the sympathetic nervous system, production of reactive oxygen species and oxidative stress. The extent of pro-inflammatory changes may also be important in determining the status of individual susceptibility to coincidentally occurring pathogens. In the present study, the post-exercise changes in serum creatine kinase (CK), a laboratory indicator of muscle damage, were found to correlate with increases in IL-8 and TNF-α. Given the key role of TNF-α in low-grade inflammation in various pathological conditions, future studies are warranted to examine whether the generation of a pro-inflammatory milieu and excess TNF-α production could also play a pathogenic role in the development of adverse health effects of high-intensity physical exercise, including exertional rhabdomyolysis and cardiac or renal injuries. Extreme endurance exercise can also impair gastrointestinal integrity and function, which is associated with both local and systemic inflammation, endotoxemia and increased levels of proinflammatory cytokines.

Additional research is also needed to address the questions of how and to what extent physical activity may lead to a reduction of inflammation and oxidative stress as recently proposed by several lines of evidence. At an individual level, adherence to a healthy diet may also be a significant contributor to the status of inflammation and oxidative stress. In the current study, data on the dietary status of the participants were not collected so its possible influence could not be evaluated. It should further be noted that the inflammatory responses related to physical activity and associated changes in biomarkers may occur in an age-dependent manner. Since exercise training has been suggested as a tool to ameliorate aging-related immunosenescence, future studies are warranted to examine the potential role of physical exercise.
as part of therapeutic and preventive strategies in various age-categorized populations.

Conclusions
Taken together, the present data demonstrates significant changes in mediators of inflammation as a result of extreme physical exercise; these changes appear to become more pronounced in proportion to the intensity of the exercise. These findings help to provide a more comprehensive view on the relationships between extreme physical exertion and the status of inflammation. The observed changes may be associated with physiological inflammatory responses to exercise and early acute phase responses to muscle damage and oxidative stress. Our findings on alterations in the balance of pro- and anti-inflammatory mediators also support the view of a transient open window of altered immunity after extreme physical exertion, which could lead to increased susceptibility to infectious agents after completion of strenuous exercise.

Abbreviations
AST, aspartate aminotransferase; CD163, a biomarker of monocyte-macrophage activation; CK, creatine kinase; CRP, C-reactive protein; IL, interleukin; suPAR, soluble urokinase plasminogen activator receptor, a biomarker of immune activation; TGF-β, transforming growth factor-beta; TNF-α, tumor necrosis factor-alpha.

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Author Contributions
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