Unusual Dynamics of Concentration Fluctuations in Solutions of Weakly Attractive Globular Proteins

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Supporting Information

ABSTRACT: The globular protein γ-crystallin exhibits a complex phase behavior, where liquid–liquid phase separation characterized by a critical volume fraction \( \phi_c = 0.154 \) and a critical temperature \( T_c = 291.8 \) K coexists with dynamical arrest on all length scales at volume fractions around \( \phi \approx 0.3-0.35 \), and an arrest line that extends well into the unstable region below the spinodal. However, although the static properties such as the osmotic compressibility and the static correlation length are in quantitative agreement with predictions for binary liquid mixtures, this is not the case for the dynamics of concentration fluctuations described by the dynamic structure factor \( S(q,t) \). Using a combination of dynamic light scattering and neutron spin echo measurements, we demonstrate that the competition between critical slowing down and dynamical arrest results in a much more complex wave vector dependence of \( S(q,t) \) than previously anticipated.

Over the past years, an increasing effort has been made to describe transitions between liquid- and solid-like states in colloidal dispersions, and experimental, theoretical, and simulation studies of systems with different interparticle interactions have been reported.1–3,5 The majority of studies have focused on two extreme cases, the occurrence of a hard sphere glass transition at very high densities and the formation of colloidal gels as a result of irreversible aggregation due to strong interparticle attractions.6 For colloids with short-range attractions, these two regimes are connected by an arrest line resulting from irreversible aggregation.7 A generic state diagram has been developed that also highlights a possible competition between liquid–liquid phase separation and dynamical arrest,6,14 which can lead to the phenomenon of an arrested spinodal decomposition. This scenario has been supported by a number of observations in systems as diverse as synthetic and food colloids, emulsions, or small globular proteins.14–19 A particularly interesting feature of such systems is that, depending on the actual shape of the interaction potential, the arrest line can intersect the (metastable) binodal close to the critical point, that is, in a region where critical fluctuations are still detectable. This leads to an intriguing competition of two mechanisms that both cause a slowing down of dynamical processes: critical slowing down and dynamical arrest. We expect both to act on different length scales, but there is currently no information available on the resulting relaxation time spectra seen for example in the dynamic structure factor \( S(q,t) \) probed by dynamic light scattering (DLS) and other quasielastic scattering techniques, such as neutron spin echo (NSE).

Globular proteins have already been used as model systems to study critical phenomena in particle systems.20–22 However, the majority of studies have focused on phase diagrams and the behavior of the osmotic compressibility \( \kappa_T \) and the static correlation length \( \xi_\alpha \) and the information on dynamic properties is limited. A notable exception is the work of Fine et al.,22 where DLS has been employed to study the influence of critical phenomena on the collective dynamics of protein solutions. They demonstrated that the decay rate of concentration fluctuations was consistent with the theory for critical dynamics in binary liquids but also noted that an unusually large background contribution had to be assumed to achieve quantitative agreement with theory. However, at that time, the possible existence of an arrest line and its influence on the dynamics had not yet been recognized.

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Here, we now address the possible competition between critical slowing down and dynamical arrest in the vicinity of a critical point for particles with short-range attractive potentials. We combine DLS with NSE, a technique accessing dynamics on a range of length scales down to the order of the protein size, thus allowing to disentangle the effects of critical slowing down and dynamical arrest in concentrated protein solutions. We focus on the globular eye lens protein γB-crystallin, which has a molecular mass of around 21 kDa and a diameter of about 3.6 nm. The static properties of γB-crystallin solutions have been studied intensively in the past, primarily in the context of cataract formation, and are well described by a model of colloidal particles interacting via a short-range attractive potential.17

Following the approach of Cardinaux et al.,17 we first established the state diagram of γB-crystallin as a function of the volume fraction φ shown in Figure 1. The coexistence curve (binodal) for liquid–liquid phase separation was determined using cloud point measurements26 (fuchsia □) and shallow quenches to temperatures T just below the critical temperature Tc followed by macroscopic phase separation (fuchsia △).27 The spinodal was obtained from static light scattering (SLS) measurements (blue ○).28 We find a critical volume fraction φc = 0.154 and Tc = 291.8 K. For quenches below 285 K, the solutions undergo a liquid–amorphous solid transition as a result of a spinodal decomposition that eventually becomes arrested when the dense phase forms a space–spanning network, in agreement with the results reported for lysozyme, another globular protein with short-range attractive interactions.17 The arrest line below the binodal/spinodal (◇) was located by performing temperature quenches deep into the unstable region of the state diagram, followed by centrifugation to separate the solid-like dense from the dilute liquid phase.17 Additionally, DLS measurements reveal a sharp temperature-independent transition from ergodic (green *) to nonergodic (X) behavior of the protein solutions at a volume fraction φarr = 0.34 as shown in Figure S3, allowing us to extend the glass line well beyond the binodal. The glass fluidizes on dilution, showing that this arrest transition is a reversible process and not caused by irreversible aggregation and gelation.

We subsequently used a combination of small-angle X-ray scattering (SAXS) and SLS to investigate the static structure factor S(q) as a measure of structural correlations on different length scales. Here we concentrate on length scales significantly larger than the protein size, that is, on q ≪ 2π/d, where d ≈ 4 nm is the protein diameter. Under these conditions, the total scattering of the proteins S(q,T,φ) can be separated into a noncritical background Scrit(0) (accounting for the athermal excluded volume interactions) and a strongly temperature-dependent critical component Sarr(q,T,φ) that dominates in the vicinity of Tc or the spinodal temperature (Tsp).

The q dependence of Sarr(q) = Scrit(0)/(1 + q2ξs−2) is given by the Ornstein–Zernike (OZ) equation, where Scrit(0) is related to the osmotic compressibility and ξs is the static correlation length as a measure of the characteristic size of density fluctuations caused by critical phenomena, and all relevant structural properties are in quantitative agreement with predictions for critical phenomena in simple binary mixtures (see Supporting Information for details). As both Scrit(0) and ξs diverge at Tc and Tsp and given the shape of the spinodal (Figure 1), we therefore also expect them to exhibit a maximum around φc for a given temperature, which is indeed observed experimentally (Figure 2 a,b). We can quantitatively reproduce the experimentally determined T and φ dependence of S(φ) of γB-crystallin solutions using a model of hard ellipsoidal particles interacting via a square-well potential with range Δ = 0.6 nm (Figure 2a, see Supporting Information for more details).

![Figure 1](4471)

**Figure 1.** State diagram of γB-crystallin in D2O. Shown are the binodal (fuchsia □, cloud point measurements; fuchsia △, shallow quenches) and the spinodal (blue ○) and the corresponding fits (T = Tc[1 − A·(|φ − φc|/φc)1/β]), with β = 0.33; binodal, A = 0.065 and 0.029; spinodal, A = 0.248 and 0.057), as well as the arrest line (dashed line) as a function of φ. The location of the latter was determined by a combination of deep quenches (○) and DLS data (green *, ergodic; X, nonergodic). φc is marked by the dotted line.

![Figure 2](4471)

**Figure 2.** (a) Inverse S(0) from SLS (filled symbols) and SAXS (open symbols) versus φ at different T and the corresponding model fits (full lines). (b) Inverse normalized correlation length versus φ as a function of T. Symbols, inverse static correlation length (from SAXS); dashed lines, inverse dynamic correlation length (actual data points shown in Supporting Information Figure S2). (c) Normalized short-time diffusion coefficient from NSE versus φ as a function of T. The dotted line marks φarr, whereas the dashed line indicates the location of the macroscopic arrest line φarr (cf. Figure 1).
providing further evidence that \( S(0) \) is indeed closely related to the existence of a critical point.

Having verified that the static properties of the protein solutions are quantitatively captured by a simple coarse-grained model of short-range attractive particles, we next proceed to a characterization of their dynamic properties. Here, we use DLS, which provides a measure of the dynamics of concentration fluctuations on length scales of order \( 2\pi / q \). We concentrate on the analysis of the initial decay of the measured correlation or intermediate scattering function (ISF) \( f(q,t) \), assuming an exponential decay \( f(q,t) = \exp(-\Gamma(q)t) \), where \( \Gamma(q) \) describes the temporal decay of the ISF at the given \( q \) value. For fluid-like protein solutions, where \( dq \ll 1 \) and particle motion is diffusive, we thus obtain the collective diffusion coefficient \( D_c = \Gamma(q)/q^2 \). This allows us to define a dynamic correlation length \( \xi_d \) or apparent hydrodynamic radius \( R_{\text{app}} \) (where \( \xi_d \equiv R_{\text{app}} \)) through the Stokes–Einstein relation \( D_c = k_B T/[6\pi\eta(T)R_{\text{app}}] \), where \( k_B \) is the Boltzmann constant and \( \eta(T) \) the T dependent solvent viscosity. In order to directly compare \( R_{\text{app}} \) with the corresponding static properties \( S(0) \) and \( \xi_d \) (Figure 2a,b and Supporting Information Figure S2), we use a normalized quantity \( D_c/D_0 \) where \( D_0 = k_B T/[6\pi\eta(T)R_{\text{app}}] \) is the free diffusion coefficient of a noninteracting \( \beta \)-crystallin with \( R_{\text{app}} = 2.27 \) nm. Note that \( D_c/D_0 = R_{\text{app}}/R_{\text{app}} \) retains only the T dependence of the apparent hydrodynamic radius.

For simple binary liquids in the vicinity of the critical point or the spinodal line, \( \xi_d \) mirrors the behavior of \( \xi_s \) and is completely dominated by critical concentration fluctuations. This is obviously not the case for our protein solutions. Here, concentration fluctuations at small \( q \) are completely dominated by critical concentration fluctuations at small \( q \), the increase of the dynamic correlation length \( \xi_d \) at large \( q \) is due to the formation and decay of a transient network driven by temporary bonds induced by the short-range attractions. This second decay is most pronounced close to the glass line but far away from the spinodal, that is, for large \( \phi \) and high \( T \) (first two data sets in Figure 3c). However, simultaneously approaching both the spinodal and the glass line results in the previously described logarithmic decay over a range of at least 2 orders of magnitude (last data set in Figure 3c).

These findings are reminiscent of the dynamics near higher-order glass-transition singularities. There, the predicted and observed logarithmic time dependence of the correlation functions is attributed to the interplay between two different arrest scenarios for \( \phi \approx 0.6 \): caging of particles by their nearest neighbors (repulsion-driven glass) and temporary bonding between the particles (attraction-driven glass). However, despite the evident similarities, there are two major differences between this "multiple glassy state" system and our concentrated \( \beta \)-crystallin solutions: (1) We find dynamical arrest at much smaller \( \phi \) (around 0.3–0.35, depending on \( T \)), where caging due to the hard core repulsion is negligible. (2) The vicinity of the spinodal leads to large-scale concentration fluctuations, strongly affecting the observed collective dynamics. It is thus rather the combination of critical fluctuations at quite large length scales and a true macroscopic arrest that leads to the logarithmic decay of the DLS correlation functions, once critical slowing down has pushed the former process close to the time scale of the attraction-driven bonding processes.

These two effects can be further disentangled with a systematic investigation of the short-time dynamics of \( \gamma_B \)-crystallin solutions on length scales comparable to the nearest-neighbor distance of the proteins. NSE experiments performed at the corresponding \( q = q_s = 0.2 \) Å\(^{-1}\) give us access to the local protein short-time diffusion, unaffected by the critical behavior that appears in the DLS data. The effective short-time diffusion coefficients \( D_s(q) \) can be extracted from single-exponential decay fits to the initial short-time decay of these ISFs (see Figure S4 in Supporting Information).

Figure 3c shows that the normalized \( D_s(q = q_s)/D_0 \) (probing the local dynamics over length scales of a protein) is entirely unaffected by large-scale fluctuations around \( \phi_c \). It also shows...
that while $D_s(q = q^*)/D_0$ decreases by 2 orders of magnitude upon approaching the arrest line, dynamics on these short length and time scales do not completely arrest, similar to previous observations in hard sphere glasses or dense particle gels,\textsuperscript{29,30} quite in contrast to arrest on macroscopic length scales as probed by DLS (Figure 1 and Supporting Information Figure S3). Moreover, $D_s(q = q^*)/D_0$ barely changes with temperature, in agreement with the nearly vertical continuation of the glass line outside of the 2-phase region of the state diagram (Figure 1).

In summary, the static properties of $\gamma_B$-crystallin solutions are well described by a coarse-grained model of ellipsoids with a weak square-well attraction. The $T$ and $\phi$ dependence of the low-$q$ limit of the static structure factor is quantitatively reproduced by a combination of a critical component given by a simple OZ equation, where the parameters follow universal scaling laws for critical phenomena in binary mixtures, and an additional $T$-independent background contribution that corresponds to the athermal limit due to hard sphere interactions. However, the corresponding dynamic structure factor is much more complex. We observe a rich $T$ and $\phi$ dependence caused by competing contributions from critical slowing down and dynamical arrest. This results in strongly differing decay scenarios for the ISF at different wave vectors, which can be clearly seen when using a combination of DLS and NSE.

There remain, however, a number of intriguing findings that warrant future investigations. The comparison between the static and dynamic properties (Figure 2) clearly indicates that although the decay of large-scale concentration fluctuations is strongly influenced by critical slowing down and qualitatively follows the static correlation length, this decay is superimposed on a large noncritical background contribution. It is important to understand the origin of this contribution, possibly linked to the presence of an arrest line at quite low values of $\phi$. However, we currently lack models that are capable of quantitatively reproducing our data. The relatively low values of $\phi_c$ and $\phi_{arr}$ and the evidence for the formation of transient network structures seen in DLS furthermore indicate that we may need to consider the influence of patchiness, both on the state diagram as well as on the dynamic properties.\textsuperscript{31} We believe that these findings are not only relevant for our fundamental understanding of dynamic processes in attractive colloids but may also have implications for the properties of the eye lens. A recent study has demonstrated the existence of a hard sphere-like glass transition in $\alpha$-crystallin solutions, another major eye lens protein, and has pointed out the importance of various arrest scenarios in dense eye lens protein solutions for cataract and presbyopia.\textsuperscript{31} Our study has now demonstrated that for $\gamma_B$-crystallin solutions such liquid—solid transitions can occur already at much lower volume fractions and lead to dramatic changes in the decay of concentration fluctuations responsible for light scattering.

**EXPERIMENTAL METHODS**

Bovine $\gamma_B$-crystallin was purified from calf lenses, obtained fresh as a byproduct from a local slaughterhouse, following the method described by Thurston\textsuperscript{25} (using Superdex 200 prep grade instead of Sephacryl S-200 high resolution for the size-exclusion chromatography). Concentration, determined by UV absorption spectroscopy,\textsuperscript{27} was converted to volume fraction $\phi$ using $\phi = c v$, where $v = 0.7$ mL/g is the protein voluminosity.\textsuperscript{21}

LS experiments and transmission measurements were performed on either a home-built multianalyte instrument\textsuperscript{32} or a commercial goniometer system (3D LS Spectrometer, LS Instruments AG), both implementing the so-called 3D cross-correlation technique to suppress contributions from multiple scattering, essential for measurements in the vicinity of a critical point or spinodal line.\textsuperscript{33} The $q$ range covered by LS is $0.001–0.003$ Å$^{-1}$, where the scattering vector $q = [4\pi n(\sin (\theta/2))]$ depends on the scattering angle $\theta$, the refractive index $n$ of the solution, and the wavelength $\lambda$.

SAXS measurements were performed on a pinhole camera (Ganesha 300 XL, SAXSLAB) covering a $q$ range from 0.003 to 2.5 Å$^{-1}$. Dividing the concentration-normalized scattering intensity $I(q)/c$ of a concentrated sample by that of a dilute sample yields the structure factor $S(q)$.

NSE experiments were carried out on IN15 at ILL probing a $q$ range from 0.01 to 0.23 Å$^{-1}$ and a Fourier time range between 0.03 and 600 ns. In this Letter, for the analysis of the NSE data, we focus on measurements performed at $q = q^* = 0.2$ Å$^{-1}$, where $q^*$ closely corresponds to the nearest-neighbor peak in the structure factor of $\gamma_B$-crystallin solutions.

**COMPUTATIONAL METHODS**

The theoretical isothermal compressibility plots in Figure 2a have been obtained for a monodisperse system of hard ellipsoids (HE, semiaxes $a = 2.75$ nm, $b = 1.625$ nm, and $c = 1.375$ nm), complemented with a square-well (SW) potential, using successive umbrella sampling (SUS) Monte Carlo (MC) simulations.\textsuperscript{34}

**ASSOCIATED CONTENT**

Supporting Information
The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacslett.5b02092.

Details of sample preparation and the experimental and computational methods. (PDF)

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Notes
The authors declare no competing financial interest.

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