Antidepressant-like effects of translocator protein (18 kDa) ligand ZBD-2 in mouse models of postpartum depression

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Abstract

The 18 kDa translocator protein (TSPO) is primarily localized in the outer mitochondrial membrane of steroid-synthesizing cells in the central and peripheral nervous systems. One of the protein’s main functions is transporting substrate cholesterol into the mitochondria in a prerequisite process for steroid synthesis. Clinical trials have indicated that TSPO ligands might be valuable in treating some neuropathies and psychopathies. However, limited information is known about the role of TSPO in postpartum depression (PPD). The TSPO ligand ZBD-2, a derivative of XBD173, was synthesized in our laboratory. Behavioral tests, enzyme linked immunosorbent assay, and Western blot were employed to evaluate ZBD-2’s efficacy against PPD and to elucidate the potential underlying molecular mechanism. The TSPO levels significantly decreased in the basolateral amygdala of PPD models. After treatment for 2 weeks, ZBD-2 alleviated depression-like behaviors and enhanced the TSPO level in a PPD animal model. The underlying mechanisms of ZBD-2 were related to regulate the hypothalamic-pituitary-adrenal axis, enhance 5-HT and BDNF secretion, and maintain the excitatory and inhibitory synaptic protein expression to normal levels. Our results directly confirm that ZBD-2 exerts a therapeutic effect on PPD, which provides a new target for anti-PPD drug development.

Keywords: Translocator protein 18 kDa, ZBD-2, Postpartum depression, Amygdala

Introduction

Postpartum depression (PPD), a widespread mental disorder, occurs in women soon after giving birth [1]. Data have shown that approximately 40% of new mothers develop moderate to severe depression, and symptoms include sadness and hopelessness [2]. PPD is well known to negatively influence the offspring, which may then acquire deficits in cognitive and social interaction, as well as emotional disorders [3]. However, the underlying etiology remains largely unknown. Peptide and steroid hormones dramatically fluctuate during pregnancy and the postpartum period; these changes may exacerbate symptoms in vulnerable women. In women at risk for depression, many symptoms attack during the period encompassing large variations in estradiol and progesterone [4]. The estrogen and progesterone levels rise steeply in pregnant women and then decrease rapidly after birth. Moreover, immune system, hypothalamic-pituitary-adrenal (HPA) axis hormones, cytokines, and fatty acids are also involved in the occurrence and development of PPD [5]. To date, no effective therapeutic method is available to treat PPD. Mothers and their individual families face a dilemma between the side effects of PPD pharmacotherapy and the adverse effects of untreated depression on their offspring. Therefore, the ideal drugs treating postpartum depression with minimal side effects are required for both mothers and their infants.

Translocator protein (TSPO) is an 18 kDa protein, which is widely distributed in the outer membrane of mitochondria in central and peripheral tissue [6]. This protein was initially identified as a peripheral binding site for diazepam and later functionally and structurally distinguished from the central benzodiazepine receptor [7]. Numerous studies have shown that TSPO plays an important role in cholesterol transport and steroidogenesis.
Meanwhile, steroid hormones modulate TSPO expression and activity in neurons [8]. At sites of injury, inflammation, and neuropathological conditions (stroke, Alzheimer's disease, Parkinson's disease, Huntington's disease, multiple sclerosis, and amyotrophic lateral sclerosis), TSPO expression was robustly enhanced in reactive microglia and astrocytes. Therefore, TSPO ligands are commonly regarded as sensitive biomarkers of brain imaging for neuroinflammation [9]. TSPO ligands have anxiolytic and antidepressant effects without evident side effects of conventional benzodiazepines [10–12]. Our previous work showed that ZBD-2, a TSPO ligand, effectively relieves anxiety [13] and depression [14] in animal models. However, the role of ZBD-2 in PPD is limited.

The amygdala is involved in modulation of stress and emotional disorders. In the amygdala, the basolateral amygdala (BLA) is a critical component that receives most of the cortical and subcortical inputs. BLA contains two major types of glutamatergic principal neurons and GABAergic interneurons. In the present study, the effects of ZBD-2 on PPD were determined in the BLA of animal model. These positive results suggested that ZBD-2 reduced anxiety-like and depression-like behaviors may be through regulating the HPA axis, enhancing 5-HT secretion, and maintaining the excitatory and inhibitory synaptic protein expression to normal levels in the BLA.

Results
ZBD-2-mediated relief of anxiety- and depression-like behaviors in PPD models
In the OF test, the traveled distance and time in the central area were markedly reduced in the PPD model mice relative to those of the control mice (distance traveled: F(6, 35) = 151.27, P < 0.001, LSD test; time in the central area: F(6, 35) = 51.07, P < 0.001, Dunnett T3 test, Fig. 1a and b). Meanwhile, the PPD models showed decreased numbers of entries and time spent in open arms in the EPM test (percent time spent in open arms: F(6,35) = 18.47, P < 0.001, LSD test; percent number of entries in open arms: F(6,35) = 19.09, P < 0.001, LSD test, Fig. 1c and d). These data indicate that the PPD models exhibited anxiety-like behaviors. Next, we detected the depression-like behaviors through sucrose preference, TST, and FST tests. The sucrose consumption ratio was significantly decreased (F(6,35) = 18.27, P < 0.001, LSD test, Fig. 1e), and the immobility times were markedly enhanced (TST immobility time: F(6,35) = 26.22, P < 0.001, LSD test; FST immobility time: F(6,35) = 19.61, P < 0.001, LSD test, Fig. 1f and g) in the PPD animal models than those of the control mice. These results suggest that the PPD animal models suffered from co-morbid depression and anxiety. ZBD-2 significantly relieved anxiety-like behaviors, as shown by the increased distance traveled and time in the central area during the OF test (Fig. 1a, b), as well as the increased open-arm entries and time spent in open arms in the EPM test (Fig. 1c, d). ZBD-2 treatment also ameliorated depression-like behaviors, as shown by the increased sucrose intake and reduced immobility times (Fig. 1e, f and g). The effects of ZBD-2 (1.5 mg/kg) were comparable with those of fluoxetine (a commonly used antidepressant in clinical) (P<0.05, Fig. 1b-g). The effects of ZBD-2 were blocked by PK11195 (a selective antagonist of TSPO), which indicated that ZBD-2 takes effect through activating TSPO.

Effects of ZBD-2 on HPA axis hormones in PPD models
We detected the levels of HPA axis hormones because they are involved in the occurrence and development of PPD [15]. The dose (1.5 mg/kg) of ZBD-2 was employed in the following experiments on the basis of above behavioral tests. The levels of CRH, ACTH, and CORT were obviously increased (CRH: F(4,25) = 40.58, P < 0.001, LSD test; ACTH: F(4,25) = 38.28, P < 0.001, LSD test; CORT: F(4,25) = 17.41, P < 0.001, LSD test, Fig. 2a, b and c), and level of 5-HT was markedly decreased in the sera of PPD models as compared to those of control mice (F(4,25) = 7.35, P < 0.001, LSD test, Fig. 2d). ZBD-2 significantly reverted the CRH, ACTH, CORT, and 5-HT concentrations to basal levels (Fig. 2). However, PK11195 treatment abolished the effects of ZBD-2 (P < 0.001, Fig. 2). There was no difference between the effects of fluoxetine and ZBD-2 (P>0.05, Fig. 2).

Effects of ZBD-2 on TSPO, 5-HT receptors, and the neural plasticity protein BDNF in PPD mice
To clarify the molecular mechanism of ZBD-2 on PPD, the levels of TSPO, 5-HT receptor subtype 1A (5-HT1A) and BDNF were measured by Western blot. The results showed that the levels of TSPO, 5-HT1A, and BDNF were significantly reduced in the BLA of PPD models. (TSPO: F(4,25) = 25.52, P < 0.001, LSD test; 5-HT1A: F(4,25) = 8.90, P < 0.001, LSD test; BDNF: F(4,25) = 56.47, P < 0.001, LSD test, Fig. 3a, b, c and d). Treatment with ZBD-2 (1.5 mg/kg) upregulated the levels of TSPO, 5-HT1A and BDNF in PPD models (Fig. 3a, b, c and d). The effects of ZBD-2 were comparable with the effects of fluoxetine (P<0.05, Fig. 3), and the effects of ZBD-2 on TSPO, 5-HT1A, and BDNF were blocked by PK11195 (P < 0.001, Fig. 3b; P < 0.05, Fig. 3c, d).

Effects of ZBD-2 on the excitatory glutamic receptors in the BLA of PPD mice
PPD is closely related to neurotransmitter disorder in CNS [5]. AMPA and NMDA receptors are two major glutamate receptors in CNS [16]. Therefore, the levels of AMPA and NMDA receptors were detected in the BLA. The levels of GluA1, phosphorylation of GluA1 at ser845 site (p-GluA1-Ser845), and GluN2B were enhanced in the BLA of PPD models (GluA1: F(4,25) = 3.285, P =
Fig. 1 (See legend on next page.)
0.072, LSD test; ρ-GluA1-Ser845: F(4,25) = 3.687, P = 0.061, Dunnett T3 test; GluN2B: F(4,25) = 4.431, P = 0.077, LSD test, Fig. 4b, c, and f), which were reversed by ZBD-2 administration (Fig. 4b, c, and f). In addition, treatment with PK11195 abolished the effects of ZBD-2 (P < 0.05, Fig. 4b, c). However, the level of GluN2A was not influenced in the BLA of PPD model with or without ZBD-2 and PK11195 treatment (Fig. 4e). The effects of ZBD-2 were comparable with those of fluoxetine (P > 0.05, Fig. 4).

**Effects of ZBD-2 on GABA receptors in PPD mice**

GABA is an important inhibitory neurotransmitter in CNS, and imbalance of excitatory and inhibitory transmission contributes to mental disorder [17]. PPD decreased the levels of GABA_A-α2 and GABA_A-γ2 in the BLA (GABA_A-α2: F(4,25) = 21.016, P = 0.002, Dunnett T3 test; GABA_A-γ2: F(4,25) = 3.821, P = 0.015, LSD test, Fig. 5a, b, and c), whereas ZBD-2 treatment upregulated their levels (Fig. 5a, b, and c). The effects of ZBD-2 were similar with those of fluoxetine (P > 0.05), and PK11195 administration abolished the effects of ZBD-2 on GABA_A-α2 and GABA_A-γ2 levels (P < 0.001, Fig. 5b; P < 0.05, Fig. 5c).

**Discussion**

In the present study, ZBD-2 treatment significantly ameliorated anxiety- and depression-like behaviors in the PPD mice. The underlying mechanisms of ZBD-2 included regulating HPA axis hormones, the levels of 5-HT, BDNF, AMPA, NMDA, and GABA_A receptors in the BLA of PPD animals.

**ZBD-2-mediated relief of anxiety- and depression-like behaviors in PPD models**

PPD is the major mood disorder in the postpartum period; up to 50% of women experiences various degrees of PPD [18]. However, the pathogenesis of PPD is highly complex and has not been completely clarified. In present study, a PPD model was established by hormone-simulated pregnancy (HSP) to evaluate the effects of ZBD-2 on PPD.
The "ovarian steroid withdrawal" hypothesis is based on the fact that significant variation in the levels of estrogen, progesterone, monoamine neurotransmitters and receptors, metabolic products, and other biological factors within the HPA axis cause the occurrence of PPD [15]. HSP-treated rats mimicked the high estrogen and progesterone levels in late pregnancy that rapidly declined after delivery. The decline in circulating ovarian steroids caused postpartum symptoms [19, 20]. In this study, PPD mice exhibited anxiety- and depression-like behaviors in EPM, OF, FST, and TST tests, indicating that HSP was suitable to establish PPD animal model.

The anxiolytic and antidepressant effects of ZBD-2 in the PPD model mice were blocked by PK11195, a potent TSPO antagonist, suggesting the effects of ZBD-2 are through activating TSPO. Previous studies have shown that TSPO ligands can increase the concentrations of pregnenolone, progesterone, and 3α,5α-tetrahydroprogesterone (3α,5α-THP) in brain. XBD173, a TSPO ligand, has been shown to potentiate GABA-mediated neurotransmission and is a promising therapeutic drug for mental disorder [8]. Fluoxetine, a first-line antidepressant drug, was used as positive control in present study. It is a selective 5-HT reuptake inhibitor and is a modulator of

Fig. 3 ZBD-2 increased the TSPO, 5-HT1A, and BDNF levels in PPD mice. a The representative Western blot analyses for TSPO, 5-HT1A, and BDNF were shown in the BLA. b, c, d ZBD-2 increased the levels of TSPO, 5-HT1A, and BDNF in the BLA of PPD mice. Similarly, PK11195 abolished the effects of ZBD-2, and the effects of ZBD-2 were comparable to those of fluoxetine. n = 5 in each group. *p < 0.05, **p < 0.01 compared with the control; †p < 0.05, ‡p < 0.01 compared with the PPD model; §p < 0.05, §§p < 0.01 compared with the ZBD-2 group. PK: PK11195

Fig. 4 ZBD-2-induced reversal of glutamate receptor expression in PPD mice. a-d The representative Western blot analyses for GluA1, p-GluA1-Ser845, GluN2A, and GluN2B are presented in the BLA. b, c, f ZBD-2 administration reversed the increased levels of GluA1 (b), p-GluA1-Ser845 (c), and GluN2B (f) expression in the PPD model mice. The effects of ZBD-2 were blocked by PK11195. e ZBD-2 did not affect the levels of GluN2A-containing NMDA receptors in the BLA of PPD mice. There were no differences between ZBD-2 and fluoxetine treatment groups. n = 5 in each group. *p < 0.05 compared with the control; †p < 0.05 compared with the PPD model; §p < 0.05 compared with the ZBD-2 group. PK: PK11195
neurosteroidogenesis in brain [21]. It has been found that fluoxetine also alleviates many premenstrual dyshoric disorder symptoms, which is strongly associated with ovarian hormones including progesterone [22]. Fluoxetine increases the 3α,5α-THP levels in the brains of depressed patients [23], and 3α,5α-THP is a potent positive allosteric modulator of the GABA<sub>α</sub> receptor [24]. Accordingly, fluoxetine exerts its antidepressant effect partially through elevating neurosteroid levels and regulating GABAergic neurotransmission in brain.

Mechanism of ZBD-2 on PPD
The HPA axis is essential in restoring normal homeostatic function following psychological stress [25]. It also plays a key role in the postpartum period [26, 27]. The activity of HPA axis is altered during pregnancy and postpartum stress [28]. HPA axis hormones are mainly composed of CRH, ACTH, and CORT, which are dysregulated in PPD patients [29]. The declining levels of monoamines, especially 5-HT, lead to functional and structural neuronal weakness, which cannot adapt to the stimulation of stress. Meanwhile, cognitive function and emotional regulation disorders are accompanied by neuroendocrine immune dysfunction, eventually leading to depression [29]. ZBD-2 significantly recovered CRH, ACTH, CORT, and 5-HT concentrations to basal levels in the PPD models; thus, ZBD-2 is involved in regulating the HPA axis hormones and 5-HT.

TSPO is involved in neurodegenerative diseases and psychiatric disorders [30]. Moreover, TSPO level is enhanced in multiple sclerosis, amyotrophic lateral sclerosis, Parkinson’s disease, Huntington’s disease, AD, and stroke [31]. However, reduced TSPO expression has been observed in patients with co-morbidities of anxiety and depression or bipolar disorder [32] and has been associated with distress and aggression [33]. Our data showed that the TSPO levels were downregulated in the BLA of PPD models and restored to basal levels after ZBD-2 treatment. Therefore, TSPO downregulation is a possible response to PPD. We also found that the effect of ZBD-2 on PPD was comparable to that of fluoxetine and suggested the high complexity of PPD pathogenesis. Thus, multiple target therapy is needed beside 5-HT reuptake inhibitors.

BDNF, a neurotrophin related to the modulation of synaptic plasticity and long-term potentiation in brain [34], is essential to placental development during pregnancy and involved in major depression [35]. BDNF level decrease induced by stress results in aberrant neurogenesis and subsequent depression [36, 37], whereas its increase follows anti-depressant treatment [38]. BDNF is also reduced during pregnancy and in the postpartum period when concomitant with depressive symptoms [39]. Diminished BDNF levels are believed to be a potential pathological mechanism underlying the impaired neurogenesis in depression [40]. We found that administering ZBD-2 reversed the downregulation of BDNF expression in the BLA of PPD models. This result suggested that the antidepressant effect of ZBD-2 is closely related to the correction of abnormal BDNF levels in the BLA.

ZBD-2-mediated amelioration of the imbalance in GABAergic and glutamatergic transmission
The balance between excitatory and inhibitory neurotransmitters is the basis of normal neurological CNS function. Glutamate is a major excitatory neurotransmitter and GABA is the most important inhibitory neurotransmitter in brain [41]. The large increase in progesterone-derived neurosteroids during pregnancy and their precipitous decrease at parturition may have considerable effects on GABA<sub>α</sub>Rs during pregnancy and postpartum [42]. Glutamate levels are sensitive to ovarian hormone fluctuations, pregnancy and the postpartum period [43]. The dysfunction of GABAergic and glutamatergic transmission is connected with PPD [41]. In the BLA of PPD mice, the levels of excitatory glutamate receptors including GluA1, P-GluA1-Ser845, and GluN2B increased, whereas those of inhibitory GABA<sub>α</sub>-a2 and GABA<sub>α</sub>-γ2 receptor decreased. Meanwhile, ZBD-2 significantly reversed these alterations but did not affect the levels of GluN2A-containing NMDA receptors in the BLA of PPD mice. The difference between GluN2A and GluN2B levels suggests that GluN2A and GluN2B play different roles in the development of PPD.

**Fig. 5** ZBD-2-induced reversal of the decreased expression of GABA<sub>α</sub>-2 and GABA<sub>α</sub>-γ2 in the PPD mice. **a** The representative Western blot analyses of GABA<sub>α</sub>-2 and GABA<sub>α</sub>-γ2 were performed. The effects of ZBD-2 were abolished by PK11195. There were no differences between ZBD-2 and fluoxetine treatment groups. **b** The representative Western blot analyses of GABA<sub>α</sub>-2 and GABA<sub>α</sub>-γ2 were performed. The difference between ZBD-2 and fluoxetine treatment groups. **c** The representative Western blot analyses of GABA<sub>α</sub>-2 and GABA<sub>α</sub>-γ2 were performed. The difference between ZBD-2 and fluoxetine treatment groups.
Synaptic and extrasynaptic NMDA receptors couple different intracellular signaling pathways [45]. Thus, the effects of ZBD-2 against PPD may be due to modulate the balance between excitatory and inhibitory transmission in the BLA.

In conclusion, we investigated the effects and potential molecular mechanisms of ZBD-2 on PPD treatment. The mechanisms of ZBD-2 are related to regulate the HPA axis, enhance 5-HT and BDNF secretion, and maintain the excitatory and inhibitory synaptic protein expressions to normal levels. These results provide important evidence that TSPO level in the BLA is involved in PPD development and ZBD-2 is an effective antidepressant drug against PPD.

**Methods**

**Materials**

ZBD-2 was prepared at our laboratory as previously described [13]. Anti-β-actin antibody was purchased from Sigma (St. Louis, MO). Anti-GluN2A, anti-GluN2B, anti-GluA1, anti-γ-GluA1-ser845, anti-5-HT1A and anti-BDNF antibodies were purchased from Abcam (Cambridge, UK). Anti-TSPO, anti-GABA A-α2 and anti-GABA A-γ2 antibodies were purchased from Chemicon (Temecula, CA, USA). All secondary antibodies conjugated with horseradish peroxidase (HRP) were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). The CRH (Corticotropin-releasing hormone), ACTH (Adreno-cortico-tropic hormone), CORT (Corticosterone) and 5-HT (5-hydroxytryptamine) ELISA kits were purchased from (Cusabio, Wuhan, China). All of the chemicals and reagents used were standard biochemical quality and commercially available.

**Animals**

Adult (8-week old) female C57BL/6 mice, weighing 18–22 g, were used in this experiment. The samples of BLA were dissected immediately after behavioral tests.

The holding room was maintained at room temperature at 22–25 °C with humidity (50%–60%) and 12-h light/day cycle. All experimental procedures were approved by the Animal Ethics Committee of the Fourth Military Medical University.

**Surgical procedures**

At beginning of the experiment, adult mice were bilaterally ovariectomized (OVX). The surgery was performed using aseptic techniques while under 4% chloral anesthesia. One dorsal lateral incision of waist was made for each side, the ovaries were isolated and sterile suture was tied tightly around the ovaries, and the ovaries were removed. The muscle layer and the cutaneous incision were sutured separately [46]. The mice were allowed to recover for at least 7 days prior to next procedure.

**Hormone-simulated pregnancy**

After one-week recovery following the OVX, mice were administered hormones (estradiol and progesterone dissolved in 0.1 ml sesame oil) for 23 days to establish hormone-simulated pregnancy (HSP) as shown in Fig. 6 [46]. The “ovarian steroid withdrawal” hypothesis is based on the onset of PPD when the levels of estrogen and progesterone are rapidly decreased after delivery. The sham group was subcutaneously injected with the same volume sesame oil (vehicle) after OVX.

**Drug treatment**

The animals received administration of vehicle or ZBD-2 (0.75 mg/kg, 1.5 mg/kg, 3.0 mg/kg, p.o.), PK11195 (3.0 mg/kg, i.p.) and Fluoxetine (3.0 mg/kg, p.o.) once daily for 2 weeks (from Day 18 to Day 31). Behavioral tests were performed 1 h after the last administration. The samples of BLA were dissected immediately after behavioral tests.

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**Fig. 6** Timeline of the experiment and HSP was employed to establish PPD mouse model.
Behavioral tests
Before the behavioral tests, mice were pretreated with mild stroking two times per day for 7 consecutive days to eliminate their nervousness. On the day of behavioral tests, the mice were moved to behavioral testing room at least 1 h for habituation.

Open field test (OFT)
The OFT was evaluated by analyzing the spontaneous activity of mice in open space [47]. Mice were gently placed in the center of an open acrylic box (30 cm × 30 cm × 30 cm) with clear plexiglas walls and white floor, and allowed mice to freely explore for 10 min. The trial for each subject was recorded in a 15 min session by a camera and the data were analyzed with a video-tracking system. The central square area of the box (one fourth of the whole area) was defined as the center zone.

Elevated plus maze (EPM)
The EPM apparatus is consisted of two open arms (without walls) and two enclosed arms (with walls). Each arm was 25 cm long and 8 cm wide, and the wall for the closed arm was 12 cm high. The maze is raised to a height of 50 cm above the ground. The mice were placed in the central square with their heads facing open arms and allowed mice to explore freely for 5 min. The numbers of arm entries and total time spent in open and enclosed arms were recorded by video for 5 min, and analyzed by video tracking system [14].

Sucrose preference test (SPT)
SPT was performed as previously described [48]. All mice were housed individually in a cage and deprived of water for 18 h, and then two bottles were placed in each cage (one is water and the other is 1% sucrose solution). The animal were allowed to drink ad libitum for 24 h [49]. To avoid position preference, two bottles were interchanged every 6 h according to previous report [50]. The consumption of water and sucrose solution was recorded during the 24 h, and the sucrose preference of each mouse was calculated as: sucrose preference (%) = sucrose solution intake/total liquid consumption× 100%.

Tail suspension test (TST)
TST was conducted as previously described [51]. The mouse was individually suspended 15 cm above the floor by the tail with adhesive tape placed approximately 2 cm from tail tip. The short-term inescapable stress led to an immobile posture. The animal behavior was recorded by video for 6 min. The performance was analyzed during the last 5 min.

Forced swim test (FST)
The apparatus of FST is a glass cylinder (30 cm diameter × 66 cm height) and filled with water for 25 cm depth at 25 ± 1 °C in a quiet testing room. On FST day 1, animals were individually placed into the water of glass cylinder for 15 min to induce a state of helplessness. 24 h later, the mice were returned to the same condition for 6 min, and three types of behaviors were observed in the last 5 min: immobility, swimming, and struggling. After each FST trial, the mouse was towel-dried, kept warm, and then returned to its home cage [52]. Water was changed after each animal test to avoid any influence.

Enzyme linked immunoabsorbent assay (ELISA)
The levels of CRH, ACTH, CORT, and 5-HT in serum were determined with commercially available ELISA kit according to the manufacturer instructions. Briefly, the mice were anesthetized with diethyl ether, and then used ophthalmological forceps to remove the eyeballs. Blood (about 1.0 ml/mouse) was collected in the 1.5 ml centrifuge tubes (anticoagulation with heparin). Then tubes were centrifuged at 3000 rpm at 4 °C for 10 min to isolate serum, and the supernatant was collected for ELISA. Serum was stored at −80 °C until use.

Western blot
The expressions of proteins in the BLA were analyzed using Western blotting [13]. The BLA were quickly dissected from brain slices (300 mm) on an ice-cold plate under an anatomical microscope, then the BLA was immediately placed in the labeled 1.5 ml centrifuge tube, weighed, and homogenized in cold tissue lysate (containing 1% 10 mM PMSF). The homogenates were centrifuged at 12,000 rpm with 4 °C for 15 min, and 5 × loading buffer was added into supernatants and heated at 95 °C for 5 min. The samples were stored at −80 °C until use. After centrifugation, 50 μg protein were separated and electrotransferred onto PDVF membranes (Invitrogen), which were probed with antibody for TSPO (1:2000 dilution), anti-5-HT1A (1:500), GluN2A (1:500), GluN2B (1:500), GluA1 (1:300), P-GluA1-ser845 (1:1000), GABA_A-γ (1:500), GABA_A-α (1:500), and BDNF (1:500) with β-actin (1:10000) as the loading control. The membranes were incubated with secondary antibodies anti-rabbit/anti-mouse/anti-goat IgG for the primary antibodies. The quantity of band intensity was normalized by comparison with β-actin, and the density analysis of protein was conducted using an ECL system (Lightning Blot System, PerkinElmer, Waltham, MA, USA).

Statistical analysis
Experimental Data were analyzed using SPSS 13.0. Results were expressed as the mean ± SEM. Data passed the homogeneity test were analyzed by the one-way ANOVA
least significant difference (LSD) test, otherwise were analyzed by the one-way ANOVA Dunnett’s T3 test comparisons. \( P < 0.05 \) was considered statistically significant.

**Abbreviations**

S-HT: 5-hydroxytryptamine; ACTH: Adrenocorticotropin-hormone; ADR: Adrenaline; BDNF: Brain-derived neurotrophic factor; BLA: Basolateral amygdala; CNS: Central nervous system; CORT: Corticosterone; CREB: cAMP response element binding protein; CRH: Corticotropin-releasing hormone; E: Estrogen; EPM: Elevated plus maze; ER: Estrogen receptor; FST: Forced swimming test; GABA: y-aminobutyric acid; Glu: Glutamate; Glu: Glutamine; HPA: Hypothalamic-pituitary-adrenal; HPA: Hypothalamo-pituitary-gonadal; HPT: Hypothalamic-pituitary-thyroid; HSP: Hormone-simulated pregnancy; LTP: Long-term potentiation; NE: Noradrenaline; NMDA: N-Methyl-D-aspartic acid; OF: Open field test; OXN: Ovariectomy; P: Progesterone; PPD: Postpartum depression; SPT: Sucrose preference test; TSPO: Translocator protein 18 kDa; TST: Tail suspension test; WB: Western blot

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**Availability of data and materials**

The datasets supporting the conclusion of this article are included within this article.

**Authors’ contributions**

SB-L and XB-L conceived and designed the study. MG-Z assisted in designing the experiments. KZ and YM-W analyzed the data. XB-L and SB-L wrote the manuscript, and MG-Z proofread it. All authors read and approved the final manuscript.

**Ethics approval**

The animal procedures involved in this study were reviewed and approved by the Fourth Military Medical University Animal Care and Use Committee.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

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