Macrophages play an important role in clearing necrotic myocardial tissues, myocardial ischemia–reperfusion injury, and ventricular remodeling after myocardial infarction. M1 macrophages not only participate in the inflammatory response in myocardial tissues after infarction, which causes heart damage, but also exert a protective effect on the heart during ischemia. In contrast, M2 macrophages exhibit anti-inflammatory and tissue repair properties by inducing the production of high levels of anti-inflammatory cytokines and fibro-progenitor cells. Interleukin (IL)-38, a new member of the IL-1 family, has been reported to modulate the IL-36 signaling pathway by playing a role similar to that of the IL-36 receptor antagonist, which also affects the production and secretion of macrophage-related inflammatory factors that play an anti-inflammatory role. IL-38 can relieve myocardial ischemia–reperfusion injury by promoting the differentiation of M1 macrophages into M2 macrophages, inhibit the activation of NOD-like receptor thermal protein domain-associated protein 3 (NLRP3) inflammasome, and increase the secretion of anti-inflammatory cytokines, such as IL-10 and transforming growth factor-β. The intact recombinant IL-38 can also bind to interleukin 1 receptor accessory protein-like 1 (IL-1RAPL1) to activate the c-Jun N-terminal kinase/activator protein 1 (JNK/AP1) pathway and increase the production of IL-6. In addition, IL-38 regulates dendritic cell-induced cardiac regulatory T cells, thereby regulating macrophage polarization and improving ventricular remodeling after myocardial infarction. Accordingly, we speculated that IL-38 and macrophage regulation may be therapeutic targets for ameliorating myocardial ischemic injury and ventricular remodeling after myocardial infarction. However, the specific mechanism of the IL-38 action warrants further investigation.

**Keywords:** interleukin-38, macrophages, myocardial infarction, myocardial ischemia–reperfusion injury, ventricular remodeling, inflammation

**INTRODUCTION**

Myocardial infarction (MI), an ischemic injury of the myocardium following the obstruction of coronary artery flow, is one of the leading causes of death worldwide and is closely related to inflammation. After MI, various inflammatory mediators released by the ischemic myocardium are involved in tissue repair and related adaptive responses (1–4). The development of necrotic
myocardium after MI can be categorized into 3 overlapping stages: the inflammatory stage, the proliferative stage, and the mature stage. During the inflammatory phase, the activated cascades of cytokines, including the Nuclear factor kappa B (NF-kB) system and the complement system, enable the recruitment of neutrophils and macrophages to the infarcted area and clear the necrotic cells. During the proliferation phase, activated macrophages release cytokines and growth factors, such as transforming growth factor (TGF)-β and interleukin (IL)-10, resulting in the formation of highly vascularized granulation tissues. Meanwhile, the expression of pro-inflammatory mediators is inhibited, and fibroblasts and endothelial cells begin to proliferate. The activated myofibroblasts produce extracellular matrix proteins that form extensive microvacular networks. After entering the mature stage, fibroblasts and vascular cells undergo apoptosis, leading to collagen scarring, namely, ventricular remodeling (5, 6). Furthermore, the continuous excessive inflammatory response increases collagen deposition and matrix degradation, thereby aggravating ventricular remodeling and seriously affecting cardiac functions later in life. The inflammatory cascade induced by ischemia–myocardial reperfusion therapy after MI can also damage the myocardial tissue to a certain extent (7). Research evidence suggests that effective inhibition of an excessive inflammatory response after MI can reduce myocardial ischemia injury and improve cardiac function (5, 8). As a newly discovered inflammatory factor, IL-38 has been extensively studied for its anti-inflammatory effects. In our previous study, plasma IL-38 levels were significantly increased in a time-dependent manner in patients with ST-segment elevation MI (9). Animal experiments showed that IL-38 can exhibit an anti-inflammatory effect by regulating macrophage function (10, 11). Therefore, the regulation of the effect of IL-38 on macrophages in myocardial ischemic injury might be considered a new direction for the treatment of MI and the improvement of its prognosis.

INTERLEUKIN-38 AND MACROPHAGES

IL-38, the 10th member of the IL-1 family, is widely expressed in human organs and tissues such as the placenta, spleen, thymus, and tonsil. It is most abundantly involved in B-cell proliferation in the skin and tonsil, whereas it is present in low levels in the heart and other non-immune organs (12). Close to the genes encoding IL-1 receptor antagonist (IL-1Ra) and IL-36 receptor antagonist (IL-36Ra), the gene encoding IL-38 is located on human chromosome 2Q13-14.1 (13). This cytokine is reported to have 37% (13) and 41% homology with IL-1Ra and 43% homology with IL-36Ra (14, 15). Van de Veerdonk et al. (16) suggested that the biological function of IL-38 is similar to that of IL-36Ra. In other words, it competitively binds to the IL-36 receptor (IL-36R) and inhibits the binding of IL-36 cytokines [IL-1F6 (IL-36α), IL-1F8 (IL-36β), IL-1F9 (IL-36γ)] with IL-36R and thereby affects the subsequent activation of NF-κB and Mitogen-activated protein kinase (MAPK), thus exhibiting certain anti-inflammatory effects (16, 17) (Figure 1). Previous studies on the association between IL-38 allele polymorphism and diseases have reported a relationship between IL-38 in compulsive arthritis and psoriatic arthritis (18, 19). Recent studies have reported IL-38 in human coronary atherosclerotic plaques (20). Moreover, polymorphisms in IL-38 are associated with coronary artery disease (21) and C-reactive protein (CRP) concentrations in humans (22). These results suggested that IL-38 may play an important role in coronary artery disease.

Macrophages, which are the main cells in inflammation, are functionally divided into classically activated macrophages (M1) associated with inflammatory responses and alternatively activated macrophages (M2) associated with injury repair and regeneration (23). At the onset of the inflammatory phase of MI, M1 macrophages, activated by tumor necrosis factor (TNF)-α, interferon (IFN)-γ, and lipopolysaccharide (LPS), are the major subtypes in the removal of dead cells and extracellular matrix debris (24, 25). In the proliferative stage of MI, M2 macrophages promote the repair and regeneration of damaged heart tissue (26). Notably, recent studies have reported that IL-38 can affect the production and secretion of inflammatory factors related to macrophages, thus playing a relevant role in systemic lupus erythematosus (27), arthritis (28), mandatory spondylitis (29), and cardiovascular diseases (9). Therefore, IL-38 is expected to be a new therapeutic target for ischemic heart injury via the regulation of macrophages.

ROLE OF INTERLEUKIN-38 IN MYOCARDIAL INFARCTION

MI is one of the cardiovascular events associated with high mortality worldwide. After MI, inflammatory mediators are released from the myocardium as a response to injury, post-injury repair, and adaptation (1–4). Our previous study reported that plasma IL-38 levels in patients with ST-segment elevation
MI are positively correlated with CRP, cardiac troponin I (cTNI), and N-terminal pro-brain natriuretic peptide (NT-proBNP) but weakly negatively correlated with left ventricular ejection fraction (9). cTNI has been widely studied as a diagnostic and prognostic marker of the acute coronary syndrome (ACS), and a previous study has reported that patients with ACS with anti-cTNI autoantibodies will eventually trigger severe autoimmune inflammation if mistaken by the immune system for antigens rather than self-protein (30). Thus, cTNI can not only represent the degree of cardiac injury but also predict autoimmune inflammation after Acute myocardial infarction (AMI). Because of its positive correlation with cTNI levels, IL-38 levels may be an indicator of myocardial injury or autoimmunity. Increased serum CRP levels are a nonspecific but sensitive marker of an acute inflammatory response, and high levels of CRP after acute MI can predict infarct expansion, cardiac rupture, and mortality (31, 32). Altogether, plasma IL-38 levels may be a new indicator to determine whether percutaneous coronary intervention is successful or not and can reflect the degree of the anti-inflammatory response in vivo. In addition, Zare Rafie et al. (33) showed that increased serum IL-38 levels were observed 24 h after tissue plasminogen activator treatment in patients with ischemic stroke, and a significant correlation was observed between changes in IL-38 concentration and patients’ prognosis at 3 months. The mechanisms underlying ischemic stroke and MI are blood flow obstruction and subsequent tissue inflammation. These results suggested that changes in serum IL-38 levels may be a new early predictor of the prognosis of ischemic disease.

**INTERLEUKIN-38 MODULATES MACROPHAGE FUNCTIONS AFTER MYOCARDIAL INFARCTION**

After MI, morphological changes occur in the infarcted and non-infarcted areas of the ventricle, which affect the pumping function of the heart and eventually lead to heart failure (34). In this process, inflammation plays an important role in the changes in myocardial tissue structure after MI (35, 36). As key cells involved in inflammation, M1 macrophages mainly secrete TNF-α, IL-1β, and IL-6 to promote inflammation, whereas M2 macrophages mainly secrete IL-10 and TGF-β, which are related to inflammation regression and tissue repair and remodeling (37, 38). Among the inflammatory markers, IL-6 is associated with adverse clinical outcomes as reported in previous clinical trials, whereas increased levels of the anti-inflammatory cytokine IL-10 are associated with more favorable outcomes (39, 40). When MI occurs, the ischemic myocardial tissue rapidly recruits inflammatory monocytes with a high expression of Ly6C and reaches a peak 3 days after MI. The recruited monocytes differentiate into inflammatory macrophages in response to chemokines and inflammatory cytokines (41, 42). Pro-inflammatory M1 macrophages secrete several inflammatory factors such as TNF-α, IL-1 β, and proteases that help clarify dead cells and debris from the infarcted areas. However, the long-term inflammatory response caused by these compounds can lead to extensive damage to the infarcted myocardium, thus affecting subsequent ventricular remodeling (43–45). From 4 to 14 days after MI, macrophages gradually change from inflammatory (M1) to anti-inflammatory or repair (M2). M2 macrophages establish an anti-inflammatory environment by downregulating inflammatory cytokines and upregulating anti-inflammatory cytokines such as IL-10, TGF-β, and vascular endothelial growth factor, and the inflammatory response is gradually replaced by angiogenesis and myofibroblast differentiation (46). Previous studies have reported that IL-38 plays an anti-inflammatory role by binding to the IL-36R and neutralizing the IL-36 inflammatory factor signaling pathway (16). *In vitro* culture of THP-1 macrophage cell line showed that IL-38 overexpression can effectively reduce the expression of IL-6, TNF-α, IL-23, and IL-10 induced by LPS but did not alter the expression of IL-1β (28). Ge et al. (11) observed that IL-38 promoted the differentiation of macrophages into anti-inflammatory M2 macrophages but prevented the differentiation into pro-inflammatory M1 macrophages after exposure to LPS (**Figure 2A**). IL-38 blocking significantly aggravated the inflammation and increased the differentiation from M2 to M1 macrophages (11). Interestingly, another study reported that dendritic cells (DCs) regulate the expression of inflammatory factors by affecting the infiltration of mononuclear macrophages in the infarcted heart during ventricular remodeling after MI (47).

NOD-like receptor thermal protein domain-associated protein 3 (NLRP3) inflammasome, an effective pro-inflammatory mediator of the innate immune system, is also a regulator of various inflammatory diseases such as MI, atherosclerosis, and diabetes (48–51). NLRP3 inflammasome consists of NLRP3 protein, ASC, and caspase 1. Once activated, cytokine precursors (i.e., promoting IL-1β) are converted to a biologically active form and trigger inflammatory responses (52). The NLRP3 inflammasome protein in macrophages cotreated with IL-38 and LPS was downregulated and returned to levels similar to those of control cells that were not treated with LPS (11). IL-38 can significantly inhibit the activation of NLRP3, thus showing a potent anti-inflammatory activity (11) (**Figure 2B**).

However, recent studies have shown that IL-38 can also exhibit anti-inflammatory effects by binding to interleukin 1 receptor accessory protein-like 1 (IL-1RAPL1) on the surface of LPS-activated macrophages (28, 53). Interestingly, the intact recombinant IL-38 binds to IL-1RAPL1, activates the downstream JNK/AP1 pathway, and increases the production of IL-6 to act as a stimulator (**Figure 2C**). Conversely, truncated IL-38 reduces JNK/AP1 signaling and restricts Th17 activation by lowering IL-6 and IL-8 levels (53–55). Although most *in vitro* experiments have reported that IL-38 can regulate the function of macrophages and play an anti-inflammatory role, the specific binding sites of IL-38 on macrophages are not completely known. These results also indicated that IL-38 binding receptors are diverse; hence, how IL-38 acts on macrophages in the myocardium needs to be further studied and discussed.
REGENERATION OF INTERLEUKIN-38 IN MACROPHAGES HAS THERAPEUTIC POTENTIAL IN MYOCARDIAL ISCHEMIA–REPERFUSION INJURY

Reperfusion therapy for ischemia caused by MI, such as percutaneous coronary intervention and intravenous thrombolytic therapy, can improve myocardial ischemia, limit the MI size in a timely and effective manner, and quickly restore the blood supply to the heart. However, in the process of blood flow restoration of the ischemic myocardium, reperfusion injury may occur because of cellular swelling and myofibrillar contracture, thus resulting in poor prognoses such as local acute inflammatory reaction, and metabolic disorder, apoptosis or necrosis, and even cardiac insufficiency (7, 56, 57). As the main cell of an inflammatory response, macrophages play an important role in this process (58). As mentioned earlier, M1 macrophages, an inflammatory cell, damage cardiac tissues and cells by releasing inflammatory mediators and proteases in the early stage of reperfusion (59). A study has reported that in the myocardium, myocardial ischemia–reperfusion injury is mainly caused by macrophages polarized by Dectin-1 (60). Interestingly, soluble receptors for advanced glycation end products can improve cardiac function after ischemia–reperfusion injury in mice by significantly controlling pro-inflammatory cytokines and observably increasing the level of the anti-inflammatory cytokine IL-10 (61). Evidence has indicated that M2 macrophages have a protective effect on the myocardium after ischemia–reperfusion; hence, effectively restricting M1 and promoting differentiation of M2 macrophages may be a therapeutic target for improving myocardial ischemia–reperfusion injury in the future. The aforementioned promotion of IL-38 on M1-to-M2 transformation reflects the potential therapeutic effect of IL-38 on myocardial ischemia–reperfusion injury.

INTERLEUKIN-38 AMELIORATES MALIGNANT VENTRICULAR REMODELING AFTER MYOCARDIAL INFARCTION

Although an early coronary reperfusion strategy can significantly improve survival in patients with MI, reducing the risk of heart failure after MI is essential. The emerging understanding of the role of macrophages in inflammation after MI suggests that these
cells are necessary for wound healing and the production of stable scars for ventricular remodeling; however, excess pro-inflammatory macrophages are also harmful (8, 34, 45, 63). Therefore, timely suppression of excessive inflammation after MI is necessary to prevent malignant ventricular remodeling and ensure the optimal formation of a supportive scar in the infarct area. In our previous study, we reported that IL-38 mRNA expression was significantly increased in the boundary region of the infarcted heart in mice and reached a peak at 24 h. Double immunofluorescence experiments on major cells involved in the development of MI showed that IL-38 was mainly expressed in cardiomyocytes and also detected in CD68 macrophages 7 days after MI. Thus, both cardiomyocytes and macrophages can be the sources of IL-38 after MI (10). Combined with the previously observed increase in circulating IL-38 levels in ST-segment elevation MI patients, IL-38 may be implicated in the development of MI (9). The infiltration of CD45+ cells in the infarcted heart of mice treated with recombinant IL-38 on the seventh day was significantly reduced compared with that in the control group. Especially, the macrophages of CD45+CD11b+F4/80+ and CD45+CD11b+F4/80+CD86+ were reduced after IL-38 treatment, whereas the macrophages of CD45+CD11b+F4/80+CD86+ were not increased. At the same time, immunohistochemical staining results of mouse hearts showed that CD68+ macrophages in the IL-38 treatment group were significantly reduced compared with those in the control group. These results showed that IL-38 can ameliorate ventricular remodeling after MI by cutting down M1 macrophages to decrease the secretion of pro-inflammatory molecules, rather than increasing the number of M2 macrophages (10, 64). Furthermore, high IL-38 levels in ST-segment elevation MI patients may be an adaptive mechanism to prevent the progression of harmful ventricular remodeling after MI. B-cell lymphoma-2 (Bcl-2) family proteins are upstream regulators of mitochondrial cytochrome C release, and the ratio of Bcl-2-associated X (Bax) to Bcl-2 is an important determinant of cell apoptosis susceptibility (65). IL-38 overexpression in cultured cardiomyocytes reduced the mRNA level of the pro-apoptotic molecule Bax and increased the expression of the anti-apoptotic molecule Bcl-2 (10), suggesting that IL-38 may affect cardiomyocyte apoptosis by regulating the Bcl-2/Bax pathway (Figure 3A).

In addition to acting on cardiomyocytes and macrophages, IL-38 exhibits anti-inflammatory effects by regulating the function of DCs. Anzai et al. (47) reported that the infiltration of M2 macrophages of F4/80+CD206+ was reduced in the DC-deficient MI model, whereas the infiltration of M1 macrophages of F4/80+CD206+ was increased. Choo et al. (66) observed the expression of the cell markers of M1 and M2 macrophages in the MI model under tolerogenic dendritic cell (tDC) treatment. They found that the expression of M1 markers (IL-1β, IL-6, IL-12b, and TNF-α mRNA) decreased significantly by day 3. On the contrary, the expression of M2 markers (arginase-1, Mrcl1, Mgl1, and Mgl2) increased by day 5 (66). Moreover, regulatory T (Treg) cells may inhibit inflammation by controlling the secretion of anti-inflammatory factors such as IL-10 and TGF-β (67). Choo et al. (66) reported that tDCs induce the production of myocardium-directed Treg cells, which then orchestrate healing by affecting the polarization of macrophages to initiate repair programs and augmenting new angiogenesis in the infarcted heart. The abundance of Treg cells in mice with MI steered the macrophage differentiation toward the M2 type, thus resulting in favorable ventricular remodeling (Figure 3B). Excessive effective T cells and insufficient recruitment of Treg cells can lead to malignant ventricular remodeling after MI (68). The ability of IL-37-induced DCs to activate naive T cells and induce Treg cells has been reported (69), whereas tolerant DCs induced by IL-38 and TnI show stronger tolerance (10). We found that IL-38 inhibited LPS-induced maturation of DCs, thus resulting in lower expression of cell surface molecules and inflammatory factors than that in LPS-induced DCs that were not treated with IL-38 (10). These results suggested that IL-38 may be involved in ventricular remodeling after MI via the regulation of DCs to Treg cells and macrophage polarization. Therefore, IL-38 has the potential as a protective factor in ameliorating malignant ventricular remodeling after MI.

**CONCLUSIONS**

Although IL-38 has not been known for a long time, it is actively studied in various inflammatory diseases. In the context of MI, IL-38 may regulate the release of inflammatory factors by regulating the macrophage phenotype (10, 11) and the function of DCs (10, 69) and play a similar function to IL-
36Ra by antagonizing the relevant receptors in the IL-36 pathway (16, 70). Thus, it regulates cardiac function and ventricular remodeling after MI, even myocardial injury after ischemia–reperfusion therapy. Most studies have reported that regulating the phenotypic transformation of M1 and M2 macrophages can promote the enhancement of cardiac angiogenesis, improve cardiac function, and reduce the infarct area after MI, thus facilitating a better prognosis (71–73). Therefore, the regulation of macrophages by IL-38 indicates its potential therapeutic value and requires more research. Although several studies have reported the role of IL-38 in autoimmune diseases and inflammation-related diseases, IL-38-related signaling pathways and anti-inflammatory mechanisms are unclear. A series of issues such as the production source of IL-38 in different environments and the main functional cells, the possible different inflammatory effects of different concentrations of IL-38, and whether IL-38 directly acts on macrophages under myocardial ischemia need to be further studied.

AUTHOR CONTRIBUTIONS

ZL, YD, and YP wrote the article. KY and QZ edited the article. YC, RZ, and JY finished the figures. QD, CP, and YZ provided feedback and guidance. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by the National Natural Science Foundation of China (grant numbers 82070310, 81770273, 81900400, 81900270, 82100339).

REFERENCES

1. Xia M, Zhang C, Gu J, Chen J, Wang LC, Lu Y, et al. Impact of C-Reactive Protein on Long-Term Mortality in Acute Myocardial Infarction Patients With Diabetes and Those Without. Chin Clin Acta (2018) 480:220–4. doi: 10.1016/j.chca.2018.02.025
2. Go AS, Mozaffarian D, Roger VL, Benjamin EJ, Berry JD, Blaha MJ, et al. Heart Disease and Stroke Statistics–2014 Update: A Report From The American Heart Association. Circulation (2014) 129(3):e28–e292. doi: 10.1161/01.01.heart.2014.01.020
3. Gupta T, Haririkhoshan P, Kolte D, Khera S, Aronow WS, Mubij M, et al. Outcomes of Acute Myocardial Infarction in Patients With Hypertrophic Cardiomyopathy. Am J Med (2015) 128(8):879–87.e1. doi: 10.1016/j.amjmed.2015.02.025
4. Schoenfeld MS, Kassas I, Shah B. Transradial Artery Access in Percutaneous Coronary Intervention for ST-Segment Elevation Myocardial Infarction and Cardiogenic Shock. Curr Treat Options Cardiovasc Med (2018) 20(2):11. doi: 10.1007/s11936-018-0607-1
5. Frangogiannis NG. The Mechanistic Basis of Infarct Healing. Circ Res (2008) 58(2):88–111. doi: 10.1161/j.01.heart.2008.06.007
6. Frangogiannis NG. The Mechanistic Basis of Infarct Healing. Antioxid Redox Signal (2006) 8(11-12):1907–39. doi: 10.1089/ars.2006.8.1907
7. Yellon DM, Hausenloy DJ. Myocardial Reperfusion Injury. N Engl J Med (2007) 357(11):2409–10. doi: 10.1056/NEJMra071667
8. Frangogiannis NG. Regulation of the Inflammatory Response in Cardiac Repair. Circ Res (2012) 110(1):159–73. doi: 10.1161/CIRCRESAHA.111.243162
9. Zhong Y, Yu K, Wang X, Wang X, Ji Q, Zeng Q. Elevated Plasma IL-38 Concentrations in Patients With Acute ST-Segment Elevation Myocardial Infarction and Their Dynamics After Reperfusion Therapy. Mediators Inflamm (2015) 2015:490120. doi: 10.1155/2015/490120
10. Wei T, Lan Y, Zhong Y, Yu K, Xu W, Zhu R, et al. Interleukin-38 Alleviates Cardiac Remodelling After Myocardial Infarction. J Cell Mol Med (2020) 24(1):371–84. doi: 10.1111/jcmm.14741
11. Ge Y, Chen J, Hu Y, Chen X, Huang M. IL-38 Alleviates Inflammation in Sepsis in Mice by Inhibiting Macrophage Apoptosis and Activation of the NLRP3 Inflammasome. Mediators Inflamm (2021) 2021:6370911. doi: 10.1155/2021/6370911
12. Palomo J, Dietrich D, Martin P, Palmer G, Gabay C. The Interleukin (IL)-1 Cytokine Family—Balance Between Agonists and Antagonists in Inflammatory Diseases. Cytokine (2015) 78(1):25–37. doi: 10.1016/j.cyto.2015.06.017
13. Lin H, Ho AS, Haley-Vicente D, Zhang J, Bernal-Fussell J, Face AM, et al. Cloning and Characterization of IL-11HY2, a Novel Interleukin-1 Family Member. J Biol Chem (2001) 276(23):20597–602. doi: 10.1074/jbc.M010095200
14. Kumar S, McDonnell PC, Lehr R, Tierney L, Tsizmas MN, Grisswold DE, et al. Identification and Initial Characterization of Four Novel Members of the Interleukin-1 Family. J Biol Chem (2000) 275(14):10308–14. doi: 10.1074/jbc.275.14.10308
15. Bensen JT, Dawson PA, Mychaleckyj JC, Bowden DW. Identification of a Novel Human Cytokine Gene in the Interleukin Gene Cluster on Chromosome 2q12-14. J Interferon Cytokine Res (2001) 21(11):899–904. doi: 10.1089/107999001753289505
16. van de Veerdonk FL, Stoeckman AK, Wu G, Boeckermann AN, Azam T, Netea MG, et al. IL-38 BINDS to the IL-36 Receptor and Has Biological Effects on Immune Cells Similar to IL-36 Receptor Antagonist. Proc Natl Acad Sci USA (2012) 109(8):3001–5. doi: 10.1073/pnas.1121534109
17. Yuan X, Peng X, Li Y, Li M. Role of IL-38 and its Related Cytokines in Inflammation. Mediators Inflamm (2015) 2015:807976. doi: 10.1155/2015/807976
18. Chou CT, Timms AE, Wei JC, Tsai WC, Wordsworth BP, Brown MA. Replication of Association of IL1 Gene Complex Members With Ankylosing Spondylitis in Taiwanese Chinese. Ann Rheum Dis (2006) 65(8):1106–9. doi: 10.1136/ard.2005.046847
19. Rahman P, Sun S, Peddle L, Snegurov T, Melay W, Greenwood C, et al. Association Between the Interleukin-1 Family Gene Cluster and Psoriatic Arthritis. Arthritis Rheumatol (2006) 54(7):2321–5. doi: 10.1002/art.21928
20. Jha HC, Srivastava P, Varshani H, Singh LC, Bhengraj AR, Prasad J, et al. Chlamydia Pneumoniae Heat Shock Protein 60 Is Associated With Apoptotic Signaling Pathway in Human Atheromatous Plaques of Coronary Artery Disease Patients. J Cardiol (2011) 58(3):216–25. doi: 10.1016/j.jcc.2011.07.010
21. Sabater-Lleal M, Huang J, Chasman D, Naïtaa S, Dehghan A, Johnson AD, et al. Multiethnic Meta-Analysis of Genome-Wide Association Studies in >100 000 Subjects Identifies 23 Fibrinogen-Associated Loci But No Strong Evidence of a Causal Association Between Circulating Fibrinogen and Cardiovascular Disease. Circulation (2013) 128(12):1310–24. doi: 10.1161/CIRCULATIONAHA.113.002251
22. Dehghan A, Dupuis J, Barbulic M, Bis JC, Eiriksdottir G, Lu C, et al. Meta-Analysis of Genome-Wide Association Studies in >800 000 Subjects Identifies Multiple Loci for C-Reactive Protein Levels. Circulation (2011) 123(7):731–8. doi: 10.1161/CIRCULATIONAHA.110.948570
23. Zhang Z, Tang J, Cui X, Qin B, Zhang J, Zhang L, et al. New Insights and Novel Therapeutic Potentials for Macrophages in Myocardial Infarction. Inflammation (2021) 44(5):1696–712. doi: 10.1007/s10753-021-01467-2
Alternatively Activated Macrophages. *Mediators Inflamm.* (2015) 2015:563951. doi: 10.1155/2015/563951

63. Nian M, Lee P, Khaper N, Liu P. Inflammatory Cytokines and Postmyocardial Infarction Remodeling. *Circ Res* (2004) 94(12):1543–53. doi: 10.1161/01.RES.0000130526.20854.fa

64. Colin S, Chinetti-Gbaguidi G, Staels B. Macrophage Phenotypes in Atherosclerosis. *Immunol Rev* (2015) 262(1):153–66. doi: 10.1111/imr.12218

65. Oltval ZN, Milliman CL, Korsmeyer SJ. Bcl-2 Heterodimerizes In Vivo With a Conserved Homolog, Bax, That Accelerates Programed Cell Death. *Cell* (1993) 74(4):609–19. doi: 10.1016/0092-8674(93)90509-O

66. Choo EH, Lee JH, Park EH, Park HE, Jung NC, Kim TH, et al. Infarcted Myocardium-Primed Dendritic Cells Improve Remodeling and Cardiac Function After Myocardial Infarction by Modulating the Treg and Macrophage Polarization. *Circulation* (2017) 135(15):CIRCULATIONAHA.116.023106. doi: 10.1161/CIRCULATIONAHA.116.023106

67. Li MO, Flavell RA. Contextual Regulation of Inflammation: A Duet by Transforming Growth Factor-Beta and Interleukin-10. *Immunity* (2008) 28(4):468–76. doi: 10.1016/j.immuni.2008.03.003

68. Tang TT, Yuan J, Zhu ZF, Zhang WC, Xiao H, Xia N, et al. Regulatory T Cells Ameliorate Cardiac Remodeling After Myocardial Infarction. *Basic Res Cardiol* (2012) 107(1):232. doi: 10.1007/s00395-011-0232-6

69. Zhu R, Zhang F, Pan C, Yu K, Zhong Y, Zeng Q. Role of IL-37- and IL-37-Treated Dendritic Cells in Acute Coronary Syndrome. *Oxid Med Cell Longev* (2021) 2021:6454177. doi: 10.1155/2021/6454177

70. Grenngt MS, Rosler B, Jacobs CW, Becker KL, Joosten LA, van der Meer JW, et al. The IL-36 Receptor Pathway Regulates Aspergillus Fumigatus-Induced Th1 and Th17 Responses. *Eur J Immunol* (2013) 43(2):416–26. doi: 10.1002/eji.201242711

71. Hasan AS, Luo L, Yan C, Zhang TX, Urata Y, Goto S, et al. Cardiosphere-Derived Cells Facilitate Heart Repair by Modulating M1/M2 Macrophage Polarization and Neutrophil Recruitment. *PloS One* (2016) 11(10):e0165255. doi: 10.1371/journal.pone.0165255

72. Zhao J, Li X, Hu J, Chen F, Qiao S, Sun X, et al. Mesenchymal Stromal Cell-Derived Exosomes Attenuate Myocardial Ischemia-Reperfusion Injury Through miR-182-Regulated Macrophage Polarization. *Cardiovasc Res* (2019) 115(7):1205–16. doi: 10.1093/cvr/cvr040

73. Dai Y, Wang S, Chang S, Ren D, Shali S, Li C, et al. M2 Macrophage-Derived Exosomes Carry microRNA-148a to Alleviate Myocardial Ischemia/Reperfusion Injury via Inhibiting TXNIP and the TLR4/NF-Kappab/NLRP3 Inflammasome Signaling Pathway. *J Mol Cell Cardiol* (2020) 142:65–79. doi: 10.1016/j.yjmcc.2020.02.007

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Li, Ding, Peng, Yu, Pan, Cai, Dong, Zhong, Zhu, Yu and Zeng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.