Fluorescent polymer films based on photo-induced electron transfer for visualizing water†

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As fluorescent materials for visualization, detection, and quantification of a trace amount of water, we have designed and developed a PET (photo-induced electron transfer)-type fluorescent monomer SM-2 composed of methyl methacrylate-substituted anthracene fluoresphore-(aminomethyl)-4-cyanophenylboronic acid pinacol ester (AminoMeCNPhenylBPin) and achieved preparation of a copolymer poly(SM-2-co-MMA) composed of SM-2 and methyl methacrylate (MMA). Both SM-2 and poly(SM-2-co-MMA) exhibited enhancement of the fluorescence emission with the increase in water content in various solvents (less polar, polar, protic, and aprotic solvents) due to the formation of the PET inactive (fluorescent) species SM-2a and poly(SM-2-co-MMA)a, respectively, by the interaction with water molecules. The detection limit (DL) of poly(SM-2-co-MMA) for water in the low water content region below 1.0 wt% in acetonitrile was 0.066 wt%, indicating that poly(SM-2-co-MMA) can act as a PET-type fluorescent polymeric sensor for a trace amount of water in solvents, although it was inferior to that (0.009 wt%) of SM-2. It was found that spin-coated poly(SM-2-co-MMA) films as well as 15 wt% SM-2-doped polymethyl methacrylate (PMMA) films produced a satisfactory reversible fluorescence off–on switching between the PET active state under a drying process and the PET inactive state upon exposure to moisture, which is demonstrated by the fact that the both the films are similar in hydrophilicity to each other from the measurement of the water contact angles on the polymer film surface. Herein we propose that PET-type fluorescent polymer films based on a fluorescence enhancement system are one of the most promising and convenient functional dye materials for visualizing moisture and water droplets.

Introduction

In recent years, concern has been raised about the development of fluorescent sensors and their functional materials such as polymer films and sensor-immobilized membranes for visualizing water in solutions, solids, and gas or on material surfaces, from the viewpoint of their potential applications to environmental and quality control monitoring systems and industry, as well as fundamental study in photochemistry, analytical chemistry, and photophysics.1–23 Several investigations have been conducted on the design and synthesis of organic fluorescent sensors and polymers for the detection of water based on ICT (intramolecular charge transfer),24–34 ESIP (excited state intramolecular proton transfer),35–38 PET (photo-induced electron transfer),39–46 or solvatochromism47–52 and the elucidation of the optical sensing properties based on changes in wavelength, intensity, and lifetime of fluorescence emission depending on the water content. It was demonstrated that most of ICT- and ESIP-type fluorescent sensors and fluorescent conjugated polymers exhibited attenuation of the fluorescence emission, that is, fluorescence quenching (turn-off) systems with the increase in water content in solvents, and were suitable for the detection and quantification of a trace amount of water (below 1–10 wt% in almost every case) in solvents. However, one can see that the fluorescence quenching (turn-off) systems make it difficult to visually confirm the presence of water in samples and on material surfaces. On the other hand, the PET-type fluorescent sensors are based on a fluorescence enhancement (turn-on) system and showed the increase in the fluorescence intensity with the increase in water content in solvents, so that it allowed us to visually confirm the presence of water in samples and on material surfaces. Thus, we have focused on the design and development of PET-type fluorescent sensors for the detection and quantification of water and the preparation of their functional materials for visualizing water. In our continuous work for the improvement of the sensitivity and accuracy of PET-type fluorescent sensors for water during the past decade, we have demonstrated that anthracene-(aminomethyl)-4-cyanophenylboronic acid pinacol ester (AminoMeCNPhenylBPin) OF-2 and its derivative SM-1 having a hydroxymethyl group on the anthracene skeleton were highly...
sensitive PET-type fluorescence sensors for the detection and quantification of a trace amount of water in polar, less polar, protic, and aprotic solvents (Fig. 1a and b). In each sensor, the PET takes place from the nitrogen atom of the amino moiety to the photoexcited anthracene fluorophore in the absence of water, leading to quenching of the fluorescence. When water was added to the solution of OF-2 or SM-1, the nitrogen atom of the amino moiety was protonated or strongly interacted with water molecules to form the PET inactive (fluorescent) species OF-2a or SM-1a, and as a result, a drastic enhancement of the fluorescence emission was observed due to the suppression of PET. Indeed, the detection limits (DLs) and quantitation limits (QLs) of OF-2 and SM-1 for water in acetonitrile were, respectively, 0.009 wt% and 0.026 wt% and 0.004 wt% and 0.013 wt%, which were equivalent or superior to those of the (previous work), (c) quenching of the fluorescence quenching (turn-off) based on the reported ICT-type and ESIPT-type fluorescent sensors. Thus, the PET method based on the fluorescence enhancement (turn-on) system makes it possible to visualize, detect, and determine a trace amount of water in solvents.

Meanwhile, under the Coronavirus Disease 2019 (COVID-19) situation, face shields made of polyester or polycarbonate films and partitions made of acrylic resin are one of convenience and commercially available protective goods for reducing the risk of droplet infection. Therefore, if we can visually confirm the presence of droplets containing infectious viruses on the face shields and partitions, this allows us to accurately remove the viruses by wiping away the droplets. However, the virus-containing droplets are generally 5 μm or more which is too small for our eyes to see. Nevertheless, because over 90% of the droplets is composed of water, functional materials as well as techniques and methods capable of visualizing water are undoubtedly useful for detecting the virus-containing droplets. In our previous work, for this purpose, we have achieved the preparation of various types of fluorescent polymer films (polystyrene, poly(4-vinylphenol), polyvinyl alcohol, and polyethylene glycol) doped with the PET-type fluorescent sensor OF-2 or SM-1. It was found that the OF-2- or SM-1-doped polymer films exhibited a reversible switching of the fluorescent color between feeble green excimer emission in the PET active state under a drying process and intense blue monomer emission in the PET inactive state upon exposure to moisture or water droplets. Our previous work is the first to achieve the preparation of PET-type fluorescent sensor-doped polymer films for water, while ICT-type and ESIPT-type fluorescent sensor-doped polymer films and fluorescent conjugated polymers for water based on a fluorescence quenching (turn-off) system have been reported. However, the reversibility of the fluorescence intensity of OF-2- or SM-1-doped polymer films between the excimer and monomer emissions in the dry–wet process were not fully satisfactory for the practical use for the visualization and detection of water on materials surfaces, due to destruction of the films during the dry–wet process.

Thus, in this work, to improve the reversibility of the fluorescence intensity of PET-type fluorescent polymer films by giving strong durability during the dry–wet process, we have designed and developed a PET-type fluorescent monomer SM-2 having a methyl methacrylate group on the anthracene skeleton as a derivative of SM-1 and achieved preparation of a copolymer poly(SM-2-co-MMA) composed of SM-2 and methyl methacrylate (MMA) (Fig. 1c and d). It was found that spin-coated poly(SM-2-co-MMA) films as well as SM-2-doped polymethyl methacrylate (PMMA) films produced a satisfactory reversible fluorescence off–on switching between the PET active state under a drying process and the PET inactive state upon exposure to moisture, which is demonstrated by the fact that the both the films are similar in hydrophilicity to each other from the measurement of the water contact angles on the polymer film surface. Herein we propose that PET-type fluorescent polymer films based on a fluorescence enhancement system are one of the most promising and convenient functional dye materials for visualizing moisture and water droplets.

**Results and discussion**

The PET-type fluorescent monomer SM-2 was prepared by the reaction of SM-1 with methacryloyl chloride (Scheme 1). Then, polymerization was carried out by a ratio of SM-2 and MMA of 1 : 20 using 2,2′-azobis(isobutyronitrile) (AIBN) as a free radical initiator to give poly(SM-2-co-MMA) as a white solid (M<sub>n</sub> = 18 900, M<sub>n</sub>/M<sub>w</sub> = 2.08, 17% yield). As the result, the <sup>1</sup>H NMR spectrum indicated that the molar ratio (y/x) of MMA unit (y) and SM-2 unit (x) and the weight percentage (wt%) of SM-2 in poly(SM-2-co-MMA) are presented in Scheme 1. 

![Fig. 1 Mechanisms of PET-type fluorescent sensors](image_url) 

**Scheme 1 Synthesis of SM-2 and poly(SM-2-co-MMA).**
the obtained poly(SM-2-co-MMA) were determined to be ca. 40 and ca. 15 wt%, respectively.

The optical sensing ability of the PET-type fluorescent monomer SM-2 for water in solvents was investigated by photoabsorption and fluorescence spectral measurements in 1,4-dioxane and THF as less polar solvents, acetonitrile as a polar solvent, and ethanol as a protic solvent containing various concentrations of water (in the water content region below 10 wt%) (Fig. 2). As with the cases of OF-2 and SM-1, SM-2 in all the four solvents showed a vibronically-structured photoabsorption band in the range of 300 nm to 400 nm originating from the anthracene skeleton and did not undergo appreciable changes in the absorbance and shape upon the addition of water to the solutions (Fig. 2a, c, e and g). For the corresponding fluorescence spectra, SM-2 in the absolute solvents exhibited a feeble and vibronically-structured fluorescence band with a fluorescence maximum wavelength ($\lambda_{\text{max}}$) at around 420 nm in the range of 400 nm to 500 nm, which is attributed to the monomer emission originating from the anthracene fluorophore in the PET active state (Fig. 2b, d, f and h). On the other hand, in the low water content region below 1.0 wt%, the fluorescence band increased in the intensity with the increase in the water content in the solution, which is attributed to the formation of the PET inactive (fluorescent) species SM-2a by the addition of a water molecule, as with the cases of OF-2 and SM-1 (Fig. 1a–c). As shown in Fig. 3a, the acetonitrile solution of SM-2 without the addition of water did not show visual fluorescence emission but exhibited the blue fluorescence emission originating from the anthracene fluorophore upon the addition of water. In fact, to confirm the formation of the PET inactive species SM-2a by the interaction with a water molecule, we performed $^1$H NMR spectral measurements of SM-2 with and without the addition of water in the acetonitrile-d$_3$ solution (2.0 x 10$^{-5}$ M) (Fig. 4). The $^1$H NMR spectrum of the SM-2 solution (water content of 0.49 wt%) without the addition of water showed an obvious signal that can be assigned to a single chemical species with the SM-2 structure. On the other hand, some additional signals appeared in both the aliphatic and aromatic regions in the $^1$H NMR spectrum of the SM-2 solution with water content of 2.3 wt%, compared to that of the solution without the addition of water, indicating the existence of other chemical species as well as SM-2. Moreover, for the $^1$H NMR spectrum of the SM-2 solution with water content of 13 wt%, the chemical shifts of the methyl protons H$_3$ of boronic acid pinacol ester, the aminomethyl protons H$_6$, and the aromatic protons H$_7$ and H$_8$ of the anthracene skeleton showed considerably upfield shifts, while those of the methylene protons H$_8$ next to the anthracene skeleton and the aromatic protons H$_7$ and H$_8$ of the phenyl group showed considerably downfield shifts. Consequently, the
fact strongly indicates that the PET inactive species SM-2a interacted with water molecules occurred upon the addition of water to the SM-2 solution (Fig. 1c), as with the cases of OF-242 and SM-1.23

The sensitivity and accuracy of SM-2 for the detection of water in solvents were evaluated by the changes in the fluorescence peak intensity at around 420 nm and the plots against the water fraction in solvents (Fig. 5). The plots for SM-2 demonstrated that the fluorescence peak intensity increased linearly as a function of the water content in the low water content region below 1.0 wt% in all four solvents (Fig. 5a), while the fluorescence intensity leveled off when the water content reached 1.0 wt% as with the cases of OF-242 and SM-1.23 The results of the plots for SM-2 are as follows:

1.4-Dioxane: \( F = 288.6[H_2O] + 7.99 \) \( (R^2 = 0.974, [H_2O] = 0.0089–0.60 \) wt%\)

THF: \( F = 326.0[H_2O] + 5.32 \) \( (R^2 = 0.986, [H_2O] = 0.0247–0.40 \) wt%\)

Acetonitrile: \( F = 355.5[H_2O] + 2.10 \) \( (R^2 = 0.977, [H_2O] = 0.0173–0.32 \) wt%\)

Ethanol: \( F = 317.6[H_2O] + 76.7 \) \( (R^2 = 0.988, [H_2O] = 0.0352–0.31 \) wt%\)

The correlation coefficient \( (R^2) \) values for the calibration curves of SM-2 were 0.974–0.988, which indicates good linearity. A linear change in fluorescence intensity as a function of water content is one of the factors required for the practical use of a fluorescence sensor for water.23 The intercept values (2.1–76.7) demonstrated that the plots for 1,4-dioxane, THF, and acetonitrile fit straight lines passing through almost the origin, which also indicates the fluorescence enhancement due to the formation of the PET inactive species SM-2a with the increase in the water content. Meanwhile, it is considered that the enhanced fluorescence of SM-2 in absolute ethanol is attributed to the suppression of PET by the hydrogen bonding between the hydroxyl group of ethanol and the amino moiety of SM-2, as with the cases of OF-242 and SM-1.23 It is worth mentioning here that there was a little difference in the \( m_s \) values (288–355) for SM-2 between the four solvents, while the \( m_s \) values for SM-2 were equivalent to those for OF-2 but smaller than those for SM-1 (Table 1). The large \( m_s \) values for SM-1 relative to SM-2 and OF-2 can be attributed to the fact that the fluorescence emission property was improved by the introduction of a hydroxymethyl group to the anthracene fluorophore. Actually, fluorescence quantum yields \( (\Phi_0) \) of OF-2, SM-1, and SM-2 in absolute acetonitrile were below 2%, but in acetonitrile with 1.0 wt% water content, the \( \Phi_0 (20\%) \) of SM-1 was higher than those (13% and 12%, respectively) of OF-2 and SM-2. The DLs and QLs of SM-2 for water in the solvents were determined based on the following equations: \( DL = 3.3e/m_s \) and \( QL = 10\sigma/m_s \), where \( e \) is the standard deviation of blank sample and \( m_s \) is the slope of a calibration curve obtained from the plot of the fluorescence intensity as a function of water content.

Fig. 4 ¹H NMR spectra of SM-2 (2.0 × 10⁻² M) in acetonitrile-d₃ with 0.49 wt%, 2.3 wt%, and 13 wt% water content.

Fig. 5 Fluorescence peak intensity at around 420 nm of SM-2 (λₑₓ = 374 or 375 nm) as a function of water content below (a) 10 wt% and (b) 1.1 wt% in 1,4-dioxane, THF, acetonitrile, and ethanol. Fluorescence peak intensity at around 420 nm of SM-2 (λₑₓ = 374 nm) and poly(SM-2-co-MMA) (λₑₓ = 375 nm) as a function of water content below (c) 10 wt% and (d) 1.1 wt% in acetonitrile.
peak intensity at around 420 nm the water fraction in the low water content region below 1.0 wt% (Fig. 5b). The DLs and QLs of SM-2 for water were, respectively, 0.011 and 0.035 wt% in 1,4-dioxane, 0.01 and 0.03 wt% in THF, 0.009 and 0.028 wt% in acetonitrile, and 0.01 and 0.032 wt% in ethanol, which were equivalent to those of OF-2 but inferior to those of SM-1. Consequently, it was found that methyl methacrylate-substituted anthracene-AminoMeCNPhenylBPiN SM-2 can act as a PET-type fluorescent sensor for the detection and quantification of a trace amount of water in polar, less polar, protic, and aprotic solvents, as with the reported PET-type fluorescent sensors for water including OF-2 and SM-1.

To investigate the optical sensing ability of the copolymer poly(SM-2-co-MMA) for water, photoabsorption and fluorescence spectra of poly(SM-2-co-MMA) were measured in acetonitrile containing various concentrations of water (Fig. 6). As with the case of SM-2, poly(SM-2-co-MMA) in absolute acetonitrile exhibited a vibronically-structured photoabsorption band in the range of 300 nm to 400 nm and a feeble and vibronically-structured fluorescence band ($\lambda_{\text{max}}$ = ca. 420 nm) in the range of 400 nm to 500 nm originating from the anthracene skeleton in the PET active state. The photoabsorption spectra showed unnoticeable changes with the increase in the water content in the acetonitrile solutions. In contrast, the fluorescence intensity of the monomer emission band originating from the anthracene fluorophore increased almost linearly with the increase in the water content in the low water content region below ca. 1.0 wt% in the acetonitrile solutions due to the formation of the PET inactive species poly(SM-2-co-MMA) by the addition of water molecules (Fig. 1d), while the fluorescence intensity leveled off when the water content reached 1.0 wt% as with the cases of SM-2 (Fig. 5c). One can see that the anthracene solution of poly(SM-2-co-MMA) without the addition of water did not show any visual fluorescence emission but exhibited the blue fluorescence emission originating from the anthracene fluorophore upon the addition of water (Fig. 3b). Indeed, the plot of the fluorescence peak intensity at around 420 nm versus the water fraction in the low water content region below 1.0 wt% showed that the calibration curve had a good linearity with the $m_c$ value of 50, intercept value of 2.61, and $R^2$ value of 0.994 [$F = 50.4[H_2O] + 2.61$ ($R^2 = 0.994$, [H$_2$O] = 0.0379–0.40 wt%) (Fig. 5d). However, the $m_c$ value (50) for poly(SM-2-co-MMA) was much smaller than that (353) of SM-2 (Table 1). Based on the calibration curve, the DLs and QLs of poly(SM-2-co-MMA) for water in acetonitrile were estimated to be 0.066 and 0.2 wt%, respectively, which were inferior to those (0.009 and 0.028 wt%) of SM-2. The deterioration of the DL and QL values of poly(SM-2-co-MMA) compared with SM-2 may be attributed to the dynamic motion of the main chain, to which the anthracene skeleton was directly attached, and hydrophobic environment of the polymer chain, which can inhibit the interaction of the SM-2 moiety with water molecules, leading to the non-radiative decay of the photoexcited anthracene fluorophore. In fact, in acetonitrile with 1.0 wt% water content, the $\phi_f$ (%) of poly(SM-2-co-MMA) was lower than that (12%) of SM-2. Nevertheless, it was found that the copolymer poly(SM-2-co-MMA) composed of SM-2 and MMA can act as a PET-type fluorescent polymeric sensor for the detection and quantification of a trace amount of water in solvents.

Next, to evaluate the possibility for the PET-type fluorescent sensor to function in polymer matrices for visualization and detection of water, we prepared spin-coated poly(SM-2-co-MMA) films on glass substrates, and photoabsorption and fluorescence spectra of the spin-coated poly(SM-2-co-MMA) films before and after exposure to moisture were repeatedly measured several times. In addition, PMMA films doped with OF-2 or SM-2 at 15 wt% as well as 50 wt% were prepared on glass substrates by spin-coating process for comparison with the spin-coated poly(SM-2-co-MMA) films, which contained ca. 15 wt% SM-2 unit (Fig. 7 and 8). The as-prepared 15 wt% and 50 wt% OF-2- or SM-2-doped PMMA films as well as the poly(SM-2-co-MMA) films (in dry process) showed a vibronically-structured photoabsorption band in the range of 300 nm to 400 nm originating from the anthracene skeleton (Fig. 7a, c, e and 8a, c). For the corresponding fluorescence spectra in dry process, the 15 wt% OF-2- or SM-2-doped PMMA films and the poly(SM-2-co-MMA)
flms exhibited a feeble and vibronically-structured fluorescence band with a $\lambda_{\text{max}}$ at around 415–430 nm in the range of 400 nm to 500 nm, which is attributed to the monomer emission originating from the anthracene fluorophore in the PET active state (Fig. 7b, d and f), but the 50 wt% OF-2- or SM-2-doped PMMA films showed a broad and feeble fluorescence band with a $\lambda_{\text{max}}$ at around 450 nm in the range of 400 nm to 600 nm attributable to the excimer emission originating from the anthracene fluorophore in the PET active aggregate state (Fig. 8b and d). When all the OF-2- or SM-2-doped PMMA films and the poly(SM-2-co-MMA) films were exposed to moisture (in wet process), the photoabsorption spectral shape did not undergo appreciable changes, although a slight change in the absorbance was observed due to the disturbance of the baseline in the photoabsorption spectra (Fig. 7a, c, e and 8a, c). For the 15 wt% OF-2- or SM-2-doped PMMA films and the poly(SM-2-co-MMA) films, the corresponding fluorescence spectra in wet process showed the enhancement of the vibronically-structured monomer emission band originating from the anthracene fluorophore in the PET inactive state (Fig. 7b, d and f). On the other hand, the 50 wt% OF-2- or SM-2-doped PMMA films showed an appearance of the monomer emission band with a $\lambda_{\text{max}}$ at around 415–430 nm and the enhancement of the excimer emission band with a $\lambda_{\text{max}}$ at around 450 nm, that is, the enhancement of the broad fluorescence band originating from the anthracene fluorophore in the range of 400 nm to 600 nm arising from the PET inactive state upon exposure to moisture (Fig. 8b and d). It is worth noting here that when all the OF-2- or SM-2-doped PMMA films and the poly(SM-2-co-MMA) films after exposure to moisture were dried in the atmosphere, the photoabsorption and fluorescence spectra recovered the original spectral shapes before exposure to moisture. Actually, one can see that the 15 wt% SM-2-doped PMMA films and the poly(SM-2-co-MMA) films initially exhibited visually imperceptible blue emission in the PET active state but the visual blue monomer emission in the PET inactive state upon exposure to moisture (Fig. 3c and d). Meanwhile, the 50 wt% SM-2-doped PMMA films showed feeble green excimer emission in the PET active state before exposure to moisture but the bluish green monomer and excimer emissions in the PET inactive state upon exposure to moisture (Fig. 9f). Therefore, for all the OF-2- or SM-2-doped PMMA films and the poly(SM-2-co-MMA) films, the reversibility of the fluorescence intensity at the $\lambda_{\text{max}}$ in the dry–wet process was investigated (Fig. 9a–e). It was found that the dry–wet cycles of the 15 wt% OF-2- or SM-2-doped PMMA films and the poly(SM-2-co-MMA) films showed a good reversible switching of the fluorescent intensity even in the five times dry–wet process (Fig. 9a–c). However, the 50 wt% OF-2- or
SM-2-doped PMMA films showed that the fluorescence intensity in the wet process was attenuated from the third time onward (Fig. 9d and e). The poor reversibility of the fluorescence intensity of the 50 wt% OF-2- or SM-2-doped PMMA films may be attributed to destruction of the films during the dry-wet process and/or promotion of aggregate formation of OF-2 and SM-2 after dry process. We also measured the water contact angles on the polymer film surfaces to investigate the hydrophilicity of the 15 wt% SM-2-doped PMMA films and the poly(SM-2-co-MMA) films (Fig. 10). The water contact angles on the polymer film surfaces were 68.3° and 68.2° for the 15 wt% SM-2-doped PMMA films and the poly(SM-2-co-MMA) films, respectively, clearly indicating that the hydrophilicity was similar to each other. Thus, the fact provides the evidence that the both films shows similar reversible switching in the fluorescence intensity in the dry-wet process. Consequently, this work demonstrated that PET-type fluorescent polymer films based on a fluorescence enhancement system produce a satisfactory reversible fluorescence off-on switching between the PET active state and the PET inactive state during the dry-wet process, and thus are one of the most promising and convenient functional dye materials to enable the visualization and detection of moisture and water droplets.

Conclusions

We have designed and developed the PET-type fluorescent monomer SM-2 composed of methyl methacrylate-substituted anthracene fluorophore-AminoMeCNPhenylBPin and the copolymer poly(SM-2-co-MMA) composed of SM-2 and MMA, as fluorescent sensors for visualization, detection, and quantification of a trace amount of water. It was found that both SM-2 and poly(SM-2-co-MMA) exhibited enhancement of the fluorescence emission with the increase in water content in various solvents (less polar, polar, protic, and aprotic solvents) due to the formation of the PET inactive (fluorescent) species SM-2a and poly(SM-2-co-MMA)a, respectively, by the interaction with water molecules. The detection limit of poly(SM-2-co-MMA) for water in acetonitrile was 0.066 wt%, indicating that poly(SM-2-co-MMA) can act as a PET-type fluorescent polymeric sensor for a trace amount of water in solvents, although it was inferior to that (0.009 wt%) of SM-2. Moreover, we have achieved the preparation of the spin-coated poly(SM-2-co-MMA) films as well as 15 wt% SM-2-doped PMMA films and demonstrated that both the polymer films produced a satisfactory reversible fluorescence off-on switching between the PET active state under a drying process and the PET inactive state upon exposure to moisture. Consequently, this work proposes that PET-type fluorescent polymer films are one of the most promising and convenient functional dye materials to enable the visualization and detection of moisture and water droplets.

Experimental

General

Melting points were measured with an AS ONE ATM-02. IR spectra were recorded on a SHIMADZU IRTracer-100 spectrometer by the ATR method. 1H and 13C NMR spectra were recorded on a Varian-500 FT NMR spectrometer. High-resolution mass spectral data were acquired by APCI on a Thermo Fisher Scientific LTQ Orbitrap XL. Photoabsorption spectra were observed with a SHIMADZU UV-3600 plus. Fluorescence spectra were measured with a Hitachi F-4500 spectrophotometer. The fluorescence quantum yields were determined by a Hamamatsu C9920-01 equipped with a CCD using a calibrated integrating sphere system. The addition of water to 1,4-dioxane, THF, acetonitrile, or ethanol solutions containing SM-2 or acetonitrile solutions containing poly(SM-2-co-MMA) was made in terms of weight percent (wt%). The determination of water in solvents was done with MKC-610 and MKA-610 Karl Fischer moisture titrators (Kyoto Electronics Manufacturing Co., Ltd) based on Karl Fischer coulometric titration for below 1.0 wt% and volumetric titration for 1.0–10 wt%. Polymer number-average molecular weights (Mn) and...
molecular weight distributions \((M_w/M_n)\) were determined by size exclusion chromatography (SEC) at 40 °C using a SHIMADZU Prominence LC-2030 plus with a guard column (LF-G, Shodex), two series-connected columns (LF-804, Shodex), a UV detector, and a differential refractive index detector (RID-20A). THF was used as the eluent, and poly(methyl methacrylate) (PMMA) standards were used to calibrate the SEC system. Static water contact angles were measured at five different positions on a substrate by the sessile drop technique using a Kyowa Interface Science DMO-602 contact angle meter.

**Synthesis**

\(10-\{\text{(5-Cyano-2-\{4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl\) benzyl\}(methylamino)methyl}anthracen-9-yl\}methyl methacrylate (SM-2).\) A solution of SM-1 \((0.100 \text{ g}, 0.203 \text{ mmol})\), 4-dimethylaminopyridine \((0.003 \text{ g}, 0.025 \text{ mmol})\), and triethylamine \((0.17 \text{ mL}, 1.21 \text{ mmol})\) in dry THF \((15 \text{ mL})\) was stirred for 0.5 h at 0 °C under nitrogen atmosphere, and then, methacryloyl chloride was added slowly to the solution. After stirring for 18 h at room temperature, the reaction mixture was concentrated. The residue was chromatographed on alumina (methanol : dichloromethane = 1 : 100 as eluent) to give SM-2 \((0.064 \text{ g}, \text{yield 56\%})\) as a light yellow solid; m.p. 137–139 °C; FT-IR \((\text{ATR})\): \(\nu = 2978, 2230, 1713, 1449, 1344, 1315, 1271, 1142 \text{ cm}^{-1};\) \(\text{H NMR} \ (500 \text{ MHz}, \text{acetone-}d_6);\) 1.29 (s, 12H), 1.87 (s, 3H), 2.32 (s, 3H), 4.04 (s, 2H), 4.51 (s, 2H), 5.56 (s, 1H), 5.97 (s, 1H), 6.24 (s, 2H), 7.50–7.64 (m, 5H), 7.73 (s, 1H), 7.87 (d, \(J = 7.5 \text{ Hz}, 1H\)), 8.46 (d, \(J = 8.7 \text{ Hz}, 2H\)), 8.51 (d, \(J = 8.8 \text{ Hz}, 2H\)) ppm; \(\text{C NMR} \ (125 \text{ MHz}, \text{CDCl}_3);\) \(\delta = 18.49, 25.13, 42.97, 52.68, 59.45, 60.81, 84.11, 113.64, 119.18, 124.66, 125.54, 125.79, 126.18, 126.21, 129.70, 130.88, 131.28, 132.02, 132.16, 135.57, 136.24, 146.43, 167.72 \text{ ppm}\) (two aromatic carbon signals were not observed owing to overlapping resonances); HRMS \((\text{APCI})\): \(m/z\) (%): \([M^+]: C_{32}H_{24}N_{2}O_2B, 560.28409;\) found 560.28491.

**Preparation of poly(SM-2-co-MMA)**

A solution of SM-2 \((0.023 \text{ g}, 0.042 \text{ mmol})\), methyl methacrylate \((0.100 \text{ mL}, 0.949 \text{ mmol})\), and azobisisobutyronitrile \((0.811 \text{ mg}, 0.005 \text{ mmol})\) in toluene \((0.77 \text{ mL})\) was degassed with four freeze–pump–thaw cycles, and then, the solution was stirred for 18 h at 70 °C under nitrogen atmosphere. The reaction mixture was concentrated, and the resulting residue was dissolved in dichloromethane. The dichloromethane solution was poured into \(n\)-hexane and the resulting precipitate was collected to give poly(SM-2-co-MMA) \((0.020 \text{ g}, \text{yield 17\%})\) as a white solid; m.p. 160–200 °C; FT-IR (ATR): \(\nu = 2991, 2949, 2231, 1728, 1481, 1449, 1387, 1348, 1267, 1240, 1190, 1146 \text{ cm}^{-1};\) \(\text{H NMR} \ (500 \text{ MHz}, \text{CDCl}_3);\) 0.82 (br, C–CH\(_3\) for MMA), 0.82 (br, C–CH\(_3\) for MMA in SM-2 unit), 1.31 (s, CH\(_3\) for BPin), 1.81 (br, CH\(_3\) for MMA and MMA in SM-2 unit), 2.30 (s, N–CH\(_3\) for SM-2 unit), 3.60 (br, O–CH\(_3\) for MMA), 3.96 (s, N–CH\(_2\)–Ph for SM-2 unit), 4.50 (s, N–CH\(_2\)–An for SM-2 unit), 6.05 (br, O–CH\(_2\)–An for SM-2 unit), 7.45–7.60 (m, 4CH at 2,3,6,7-positions on An and CH at 6-position on Ph for SM-2 unit), 7.63 (br, CH at 4- position on Ph for SM-2 unit), 7.83 (br, CH at 3- position on Ph for SM-2 unit), 8.23–8.40 (m, 4CH at 1,4,5,8-positions on An for SM-2 unit) ppm, the molar ratio \(y/x\) of MMA unit \((y)\) and SM-2 unit \((x)\) and the weight percentage (wt%) of SM-2 in the obtained poly(SM-2-co-MMA) was determined to be \(ca. 40\) and \(ca. 15\%\), respectively, from the \(^1\text{H NMR}\) spectrum; \(\text{SEC}\) \(M_w/M_n = 2.08;\) UV-Vis \(\lambda_{\text{abs}} = 339, 356, 375, 395 \text{ nm}\); PL \(\lambda_{\text{fl}} = 401, 422, 447, 480 \text{ nm}\) (in acetonitrile).

**Preparation of poly(SM-2-co-MMA) film**

A solution of poly(SM-2-co-MMA) \((8.0 \text{ mg})\) in toluene \((0.4 \text{ mL})\) was stirred for 3 h at room temperature, while poly(SM-2-co-MMA) has dissolved quickly. To prepare a polymer film, 150 \(\mu\text{L}\) of a poly(SM-2-co-MMA) solution was spin-coated \((1000 \text{ rpm} \text{ for } 30 \text{ s})\) on a glass substrate (MIKASA MS-A-100 Opticoat Spinncoater). The spin-coated films were dried in air. The resulting poly(SM-2-co-MMA) films were exposed to moisture for 60 s using a humidifier.

**Preparation of 15 wt% and 50 wt% OF-2- or SM-2-doped PMMA films**

A solution of PMMA \((8.5 \text{ mg} \text{ and } 5.0 \text{ mg} \text{ for } 15 \text{ wt\%} \text{ and } 50 \text{ wt\%}, \text{respectively})\) in toluene \((0.5 \text{ mL})\) was stirred for several hours at 60–70 °C until PMMA has dissolved, and then, OF-2 or SM-2 \((1.5 \text{ mg} \text{ and } 5.0 \text{ mg} \text{ for } 15 \text{ wt\%} \text{ and } 50 \text{ wt\%}, \text{respectively})\) was added to the solution. To prepare a polymer film, 150 \(\mu\text{L}\) of a OF-2-co-MMA solution or a SM-2-PMMA solution was spin-coated \((1000 \text{ rpm} \text{ for } 30 \text{ s})\) on a glass substrate (MIKASA MS-A-100 Opticoat Spincoater). The spin-coated films were dried in air. The resulting 15 wt% and 50 wt% OF-2- or SM-2-doped PMMA films were exposed to moisture for 60 s using a humidifier.

**Conflicts of interest**

There are no conflicts to declare.

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**Notes and references**

1. H. S. Jung, P. Verwilgh, W. Y. Kim and J. S. Kim, Fluorescent and colorimetric sensors for the detection of humidity or water content, Chem. Soc. Rev., 2016, 45, 1242–1256.
2. F. Wu, L. Wang, H. Tang and D. Cao, Excited State Intramolecular Proton Transfer Plus Aggregation-Induced Emission-Based Diketopyrrolopyrrole Luminogen: Photophysical Properties and Simultaneously Discriminative Detection of Trace Water in Three Organic Solvents, Anal. Chem., 2019, 91, 5261–5269.
3 W. Cheng, Y. Xie, Z. Yang, Y. Sun, M.-Z. Zhang, Y. Ding and W. Zhang, General Strategy for in Situ Generation of a Coumarin-Cu\textsuperscript{2+} Complex for Fluorescent Water Sensing, Anal. Chem., 2019, 91, 5817–5823.

4 S. Song, Y. Zhang, Y. Yang, C. Wang, Y. Zhou, C. Zhang, Y. Zhao, M. Yang and Q. Lin, Ratiosimetric fluorescence detection of trace water in organic solvents based on aggregation-induced emission enhanced Cu nanoclusters, Analyst, 2018, 143, 3068–3074.

5 L. Liu, Q. Zhang, H. Duan, C. Li and Y. Lu, An ethanethioate functionalized polythiophene as an optical probe for sensitive and fast detection of water content in organic solvents, Anal. Methods, 2021, 13, 3792–3798.

6 T. Maeda and F. Würthner, Halochromic and hydrochromic squaric acid functionalized perylene bisimide, Chem. Commun., 2015, 51, 7661–7664.

7 D. Wang, H. Zhao, H. Li, S. Sun and Y. Xu, A fluorescent “glue” of water triggered by hydrogen-bonding cross-linking, J. Mater. Chem. C, 2016, 4, 11050–11054.

8 S. Roy, S. Das, A. Ray and P. P. Parui, An inquisitive fluorescence method for the real-time detection of trace moisture in polar aprotic solvents with the application of water rancidity in foodstuffs, New J. Chem., 2021, 45, 4574–4583.

9 S. Mishra and A. K. Singh, Optical sensors for water and humidity and their further applications, Coord. Chem. Rev., 2021, 445, 214063.

10 J. Othong, J. Boonmak, F. Kielar and S. Youngme, Dual Function Based on Switchable Colorimetric Luminescence for Water and Temperature Sensing in Two-Dimensional Metal–Organic Framework Nanosheets, ACS Appl. Mater. Interfaces, 2020, 12, 41776–41784.

11 Y. Zhou, G. Baryshnikov, X. Li, M. Zhu, H. Ägren and L. Zhu, Anti-Kasha’s Rule Emissive Switching Induced by Intermolecular H-Bonding, Chem. Mater., 2018, 30, 8008–8016.

12 P. Kumar, R. Sakla, A. Ghosh and D. A. Jose, Reversible Colorimetric Sensor for Moisture Detection in Organic Solvents and Application in Inkless Writing, ACS Appl. Mater. Interfaces, 2017, 9, 25600–25605.

13 H. Yan, S. Guo, F. Wu, P. Yu, H. Liu, Y. Li and L. Mao, Carbon Atom Hybridization Matters: Ultrafast Humidity Response of Graphdiyne Oxides, Angew. Chem., Int. Ed., 2018, 57, 3922–3926.

14 K. Tanaka, K. Nishino, S. Ito, H. Yamane, K. Suegata, K. Hashimoto and Y. Chuo, Development of solid-state emissive o-carboranes and theoretical investigation of the mechanism of the aggregation-induced emission behaviors of organoboron “element-blocks”, Faraday Discuss., 2017, 196, 31–42.

15 H. Mori, K. Nishino, K. Wada, Y. Morisaki, K. Tanaka and Y. Chuo, Modulation of luminescence chromic behaviors and environment-responsive intensity changes by substituents in bis-o-carborane-substituted conjugated molecules, Mater. Chem. Front., 2018, 2, 573–579.

16 K. Nishino, H. Yamamoto, J. Ochi, K. Tanaka and Y. Chuo, Time-Dependent Emission Enhancement of the Ethynylpyrene-o-Carborane Dyad and Its Application as a Luminescent Color Sensor for Evaluating Water Contents in Organic Solvents, Chem.–Asian J., 2019, 14, 1577–1581.

17 Y.-C. Liu, G.-D. Lu, J.-H. Zhou, J.-W. Rong, H.-Y. Liu and H.-Y. Wang, Fluoranthene dyes for the detection of water content in methanol, RSC Adv., 2022, 12, 7405–7412.

18 W.-E. Lee, Y.-J. Jin, L.-S. Park and G. Kwak, Fluorescent Actuator Based on Microporous Conjugated Polymer with Intramolecular Stack Structure, Adv. Mater., 2012, 24, 5604–5609.

19 D.-C. Han, Y.-J. Jin, J.-H. Lee, S.-I. Kim, H.-J. Kim, K.-H. Song and G. Kwak, Environment-Specific Fluorescence Response of Microporous, Conformation-Variable Conjugated Polymer Film to Water in Organic Solvents: On-line Real-Time Monitoring in Fluidic Channels, Macromol. Chem. Phys., 2014, 215, 1068–1076.

20 Q. Deng, Y. Li, J. Wu, Y. Liu, G. Fang, S. Wang and Y. Zhang, Highly sensitive fluorescent sensing for water based on poly(n-aminobenzoic acid), Chem. Commun., 2012, 48, 3009–3011.

21 J. Lee, M. Pyo, S. Lee, J. Kim, M. Ra, W.-Y. Kim, B. J. Park, C. W. Lee and J.-M. Kim, Hydrochromic conjugated polymers for human sweat pore mapping, Nat. Commun., 2014, 5, 3736.

22 T. Fumoto, S. Miho, Y. Mise, K. Imato and Y. Ooyama, Polymer films doped with fluorescent sensor for moisture and water droplet based on photo-induced electron transfer, RSC Adv., 2021, 11, 17046–17050.

23 S. Miho, T. Fumoto, Y. Mise, K. Imato, S. Akiyama, M. Ishida and Y. Ooyama, Development of highly sensitive fluorescent sensor and fluorescent sensor-doped polymer films for trace amounts of water based on photo-induced electron transfer, Mater. Adv., 2021, 2, 7662–7670.

24 I. M. Resta and F. Galindo, Phenol-based styrylpyrylium dyes for trace water detection via chromogenic and fluorogenic responses, Dyes Pigm., 2022, 197, 109908.

25 Z. Li, Q. Yang, R. Chang, G. Ma, M. Chen and W. Zhang, N-Heteroaryl-1,8-naphthalimide fluorescent sensor for water: Molecular design, synthesis and properties, Dyes Pigm., 2011, 88, 307–314.

26 W. Chen, Z. Zhang, X. Li, H. Ägren and J. Su, Highly sensitive detection of low-level water content in organic solvents and cyanide in aqueous media using novel solvatochromic AIEE fluorophores, RSC Adv., 2015, 5, 12191–12201.

27 S. Tsumura, T. Enoki and Y. Ooyama, A colorimetric and fluorescent sensor for water in acetonitrile based on intramolecular charge transfer: D–π-A\textsubscript{2}-type pyridine-boron trifluoride complex, Chem. Commun., 2018, 54, 10144–10147.

28 T. Enoki and Y. Ooyama, Colorimetric and ratiometric fluorescence sensing of water based on 9-methyl pyrido[3,4-b]indole-boron trifluoride complex, Dalton Trans., 2019, 48, 2086–2092.

29 K. Imato, T. Enoki and Y. Ooyama, Development of an intramolecular charge transfer-type colorimetric and fluorescence sensor for water by fusion with a juloidine structure and complexion with boron trifluoride, RSC Adv., 2019, 9, 31466–31473.
30. S. Tsumura, K. Ohira, K. Imato and Y. Ooyama, Development of optical sensor for water in acetonitrile based on propeller-structured BODIPY-type pyridine–boron trifluoride complex, *RSC Adv.*, 2020, **10**, 33836–33843.

31. C.-G. Niu, P.-Z. Qin, G.-M. Zeng, X.-Q. Gui and A.-L. Guan, Fluorescence sensor for water in organic solvents prepared from covalent immobilization of 4-morpholinyl-1, 8-naphthalimide, *Anal. Bioanal. Chem.*, 2007, **387**, 1067–1074.

32. Z.-Z. Li, C.-G. Niu, G.-M. Zeng and P.-Z. Qin, Fluorescence Sensor for Water Content in Organic Solvents Based on Covalent Immobilization of Benzothioxanthen, *Chem. Lett.*, 2009, **38**, 698–699.

33. D. Citterio, K. Minamihashi, Y. Kuniyoshi, H. Hisamoto, S. Sasaki and K. Suzuki, Optical determination of low-level water concentrations in organic solvents using fluorescent acridinyl dyes and dye-immobilized polymer membranes, *Anal. Chem.*, 2001, **73**, 5339–5345.

34. C.-G. Niu, A.-L. Guan, G.-M. Zeng, Y.-G. Liu and Z.-W. Li, Fluorescence water sensor based on covalent immobilization of chalcone derivative, *Anal. Chim. Acta*, 2006, **577**, 264–270.

35. W. Liu, Y. Wang, W. Jin, G. Shen and R. Yu, Solvatochromogenic flavone dyes for the detection of water in acetone, *Anal. Chim. Acta*, 1999, **383**, 299–307.

36. J. S. Kim, M. G. Choi, Y. Huh, M. H. Kim, S. H. Kim, S. Y. Wang and S.-K. Chang, Determination of Water Content in Aprotic Organic Solvents Using 8-Hydroxyquinoline Based Fluorescent Probe, *Bull. Korean Chem. Soc.*, 2006, **27**, 2058–2060.

37. H. Mishra, V. Misra, M. S. Mehta, T. C. Pant and H. B. Tripathi, Fluorescence Studies of Salicylic Acid Doped Poly(vinyl alcohol) Film as a Water/Humidity Sensor, *J. Phys. Chem. A*, 2004, **108**, 2346–2352.

38. A. C. Kumar and A. K. Mishra, 1-Naphthol as an excited state proton transfer fluorescent probe for sensing bound-water hydration of polyvinyl alcohol, *Talanta*, 2007, **71**, 2003–2006.

39. Y. Ooyama, M. Sumomogi, T. Nagano, K. Kushimoto, K. Komaguchi, I. Imae and Y. Harima, Detection of water in organic solvents by photo-induced electron transfer method, *Org. Biomol. Chem.*, 2011, **9**, 1314–1316.

40. Y. Ooyama, A. Matsugasako, K. Oka, T. Nagano, M. Sumomogi, K. Komaguchi, I. Imae and Y. Harima, Fluorescence PET (photo-induced electron transfer) sensors for water based on anthracene–boronic acid ester, *Chem. Commun.*, 2011, **47**, 4448–4450.

41. Y. Ooyama, A. Matsugasako, Y. Hagiwara, J. Ohshita and Y. Harima, Highly sensitive fluorescence PET (photo-induced electron transfer) sensor for water based on anthracene–bisboronic acid ester, *RSC Adv.*, 2012, **2**, 7666–7668.

42. Y. Ooyama, K. Furue, K. Uenaka and J. Ohshita, Development of highly-sensitive fluorescence PET (photo-induced electron transfer) sensor for water: anthracene–boronic acid ester, *RSC Adv.*, 2014, **4**, 25330–25333.

43. Y. Ooyama, M. Hato, T. Enoki, S. Aoyama, K. Furue, N. Tsunoji and J. Ohshita, A BODIPY sensor for water based on a photo-induced electron transfer method with fluorescence enhancement and attenuation systems, *New J. Chem.*, 2016, **40**, 7278–7281.

44. Y. Ooyama, R. Sagisaka, T. Enoki, N. Tsunoji and J. Ohshita, Tetraphenylethene– and diphenylbenzofulvene–anthracene-based fluorescence sensors possessing photo-induced electron transfer and aggregation-induced emission enhancement characteristics for detection of water, *New J. Chem.*, 2018, **42**, 13339–13350.

45. D. Jinbo, K. Imato and Y. Ooyama, Fluorescent sensor for water based on photo-induced electron transfer and Förster resonance energy transfer: anthracene–(aminomethyl)phenylboronic acid ester-BODIPY structure, *RSC Adv.*, 2019, **9**, 15335–15340.

46. D. Jinbo, K. Ohira, K. Imato and Y. Ooyama, Development of fluorescent sensors based on a combination of PET (photo-induced electron transfer) and FRET (Förster resonance energy transfer) for detection of water, *Mater. Adv.*, 2020, **1**, 354–362.

47. L. Ding, Z. Zhang, X. Li and J. Su, Highly sensitive determination of low-level water content in organic solvents using novel solvatochromic dyes based on thioxanthone, *Chem. Commun.*, 2013, **49**, 7319–7321.

48. Y. Zhang, D. Li, Y. Li and J. Yu, Solvatochromic AIE luminogens as supersensitive water detectors in organic solvents and highly efficient cyanide chemosensors in water, *Chem. Sci.*, 2014, **5**, 2710–2716.

49. Y. Mise, K. Imato, T. Ogì, N. Tsunoji and Y. Ooyama, Fluorescence sensors for detection of water based on tetraphenylethene–anthracene possessing both solvatofluorochromatic properties and aggregation-induced emission (AIE) characteristics, *New J. Chem.*, 2021, **45**, 4164–4173.

50. N. Zhao, Z. Yang, J. W. Y. Lam, H. H. Y. Sung, N. Xie, S. Chen, H. Su, M. Gao, I. D. Williams, K. S. Wong and B. Z. Tang, Benzoazolium-functionalized tetraphenylethene: an AIE luminogen with tunable solid-state emission, *Chem. Commun.*, 2012, **48**, 8637–8639.

51. X. Y. Shen, Y. J. Wang, H. Zhang, A. Qin, J. Z. Sun and B. Z. Tang, Conjugates of tetraphenylethene and diketopyrrolopyrrole: tuning the emission properties with phenyl bridges, *Chem. Commun.*, 2014, **50**, 8747–8750.

52. F. Khan, A. Ebkote, S. M. Mobin and R. Misra, Mechanochromism and Aggregation-Induced Emission in Phenanthroimidazole Derivatives: Role of Positional Change of Different Donors in a Multichromophoric Assembly, *J. Org. Chem.*, 2021, **86**, 1560–1574.

53. Q. Wu, H. Xiong, Y. Zhu, X. Ren, L.-L. Chu, Y.-F. Yao, G. Huang and J. Wu, Self-Healing Amorphous Polymers with Room-Temperature Phosphorescence Enabled by Boron-Based Dative Bonds, *ACS Appl. Polym. Mater.*, 2020, **2**, 699–705.

54. W. Wu, N. Shi, J. Zhang, X. Wu, T. Wang, L. Yang, R. Yang, C. Ou, W. Xue, X. Feng, L. Xie and W. Huang, Electrospray fluoresce sensors for the selective detection of nitro explosive vapors and trace water, *J. Mater. Chem. A*, 2018, **6**, 18543–18550.

55. W. Wang, S. Gao and B. Wang, Building Fluorescent Sensors by Template Polymerization: The Preparation of a Fluorescent Sensor for d-Fructose, *Org. Lett.*, 1999, **1**, 1209–1212.