Amelioration of oxidative and inflammatory changes by *Peganum harmala* seeds in experimental arthritis

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**Abstract**

**Background:** The present study was designed to investigate the therapeutic effects against oxidative stress and alleviative effects of *Peganum harmala* seeds (PH) in rats with Complete Freund’s Adjuvant (CFA) induced arthritis.

**Methods:** The extract was evaluated for its phytoconstituents, antiarthritic and antioxidant properties. The action of chloroform (PHC) and ethanolic (PHE) extracts of PH was evaluated in adult Lewis rats (150-200 g) with CFA induced arthritis. Arthritic rats received PH extracts 100 mg/kg orally for 28 consecutive days (Prophylactic treatment) and from 14th day of CFA injection (Therapeutic treatment).

**Results:** PHE significantly suppressed the arthritis severity in rats than PHC in 28 days. All complications shown significant reduction (*p* < 0.05) in arthritic rats including paw volume (63.09%), body weight loss, decreased locomotor activity, erythrocyte sedimentation rate and synovial/hepatic tissues lipid peroxidation and increase in cellular antioxidants superoxide dismutase (U/mg) activities and hemoglobin counts. The results showed the presence of alkaloids and flavonoids in PHE. Histology and radiographic analysis of arthritic ankle joints indicated abnormal changes. Marked reduction in inflammation and arthritic changes were observed after treatment with PHE.

**Conclusion:** Therefore, the investigation suggests that PHE at 100 mg/kg will be useful in the management of rheumatoid arthritis complications which may possibly be due to boosting the intracellular antioxidant defense.

**Keywords:** Arthritis, *Peganum harmala*, Antioxidant, Freund’s adjuvant, Lipid peroxidation

**Background**

Rheumatoid arthritis (RA), a systemic inflammatory autoimmune disease that afflicts 1–2% of the global population, is characterized by immune-mediated inflammatory sinusitis, which leads to hyperplasia, vasculogenesis, cartilage and bone destruction, joint malformation, functional impairment [1, 2]. Extra-articular complications of RA includes weight loss, oxidative tissue damage, chronic fatigue, and hematological disorders which are associated with the swelling around the joint, pain, abnormal increase in number of synovial cells, morphological changes, and pannus formation [3]. This chronic progressive autoimmune disorder of unknown cause is characterized by pain, swelling and stiffness of synovial joints [4]. The disease affected approximately about 0.75% in India and 1% adult population in the world and the responsible causative factors for the development of it are unknown [5].

An inflammatory reaction, imbalance between pro-inflammatory and anti-inflammatory state, increased cellularity of synovial tissue, joint inflammation and joint damage are the pathological hallmarks of RA [6–8]. The other most common extra-articular complication in RA is anemia [9]. Extra-articular can also be considered as a predictor of mortality in patients with RA [10]; these complications arose throughout the onset of disease and its progression which are mostly related to...
excessive inflammation and cellular oxidative stress induced by inflammatory mediators and free radicals, respectively [11]. The detrimental effects of reactive oxygen species (ROS) and nitrogen species (RNS) like free radicals on synovial joint and vital organs is also evident and have potential role in development of RA. Those complications can be reduced by scavenging the free radicals by antioxidants and ameliorating the radicals [12, 13].

The treatment strategy for the disease involves the use of non-steroidal anti-inflammatory drugs (NSAIDs), disease modifying anti-rheumatic drugs (DMARDs), immunomodulators, biological agents and corticosteroids. This conventional approach has shown effective results with ameliorating symptoms but the adverse effects cannot be neglected [14]. However, despite an increase in use, evidence for effectiveness and safety of other complementary therapies with them is limited [15].

**Peganum harmala** (PH), also known as harmala or Syrian rue, is an herbaceous perennial plant of the family Nitrariaeaceae; is used as a traditional herbal remedy for various diseases [16].

In traditional medicine, it has been used for the treatment of different conditions, like asthma, lumbar colic, jaundice and to stimulate menstrual flow. However, in recent pharmaceutical studies, PH found to have an antispasmodic, antimicrobial, emmenagage and abortive effects, blocking different types of intestinal calcium channels, mono amine oxidase inhibition and antidepressant effect, analgesic [17].

The beta carboline alkaloids, also called as harmal alkaloids (e.g. harmalol, harmine, harman and harmaline) and quinazoline alkaloids (vasicine and vasicinone) have potent pharmacological action [18]. They are mainly present in seeds of PH and are responsible for PH toxicity. The drug also shows antidepressant action by competitively and reversibly inhibiting the monoamine oxidases (MAO). Therefore, the present study aimed to evaluate the alleviative effects (from the day of arthritis induction and arthritis onset) of PH extracts on articular/ extra-articular complications (with special reference to cellular oxidative stress and anemia) in rats with CFA induced arthritis.

**Material and method**

**Instruments and chemicals**

Diclofenac sodium, Complete Freund’s adjuvant was obtained from Sigma Aldrich Chemical Co., Bengaluru, EDTA (Merck Laboratories Pvt. Ltd.), DTNB (Tokyo Chemical Industry Co. Ltd.), Thio-Barbituric Acid, Trichloro acetic Acid (Sunchem India). All other chemicals used in the study were of analytical grade. Cooling centrifuge (Remi-electrotechnic limited), UV Spectrophotometer (Pharmaspecs shimadzu), Actophotometer (Teknik), Hot plate (Lyzer India), Rotatory Flash Evaporator (IKA RV10), trinocular microscope, and other instruments were used.

**Plant material and extraction procedure**

The plant and seeds was collected from Mullu Mukhtar Husain Gulam Abbas, Jumerati Bazar, (M.P.) where it is used for commercial and medicinal purposes. The plant was authentified from Department Of Botany at Safia Science College, Bhopal (M.P.) by Dr. Zia Ul Hasan with a voucher Specimen no. 316/Bot/Safia/12.

The seeds were cleaned, dried in the shade so as to protect its chemical constituents not to get degrade at high temp. The moderately coarse powder of the PH seeds was subjected to successive soxhlet extraction by petroleum ether, chloroform, ethanol and water (Fig. 1). The last traces of solvent being removed under vacuum by rota-vaccum evaporator. The extracts obtained were weighed to a constant weight and percentage w/w basis was calculated and stored in cool dry place in airtight container for further use [19].

**Phytochemical analysis**

The extracts were subjected to phytochemical analysis for identification of alkaloid, saponin, tannins, phenolic compounds, carbohydrates, flavonoids, triterpenes and steroid detection according to method described by Harborne [20]. The chloroform extract of PH (PHC) and ethanolic extract of PH (PHE) extracts were selected for further studies after preliminary screening.

**Animals**

The experiment was carried out on healthy albino Lewis rats of either sexes weighing between 200 and 250 g, were used for the evaluation of acute oral toxicity test and anti arthritic activity. Animals were provided by the authorized animal house of Sapience Bioanalytical Research Lab, Bhopal, Madhya Pradesh, India. The animals were used after an acclimatization period of 10 days to the laboratory environment. The experiment was approved by the Institutional Animal Ethical Committee as per the committee for the, Purpose of control and supervision of experiments on animal’s guideline. (CPCSEA Reg. no.: 1413/PO/a/11). All animals were kept in standard cages and maintained at 25°C. Under 12 h dark/ light cycle. The animals were fed with standard rat feed and water was given after specific interval prior to experiment animal were kept for 12 h fasting.

**Experimental design**

**Acute oral toxicity study and selection of doses**

Oral dose of PH was determined through the acute oral toxic test in rats by Irwin Battery test at different doses. The dose level to be used as the starting dose is selected.
from one of four fixed levels, 5, 50, 300 and 2000 mg/kg body weight.

**Induction of arthritis**

Lewis rats of either sex weighing 180 ± 20 g was divided into seven groups of six animals each. Group I served as control (without treatment), Group II served as arthritic control (negative control), Group III was treated with Diclofenac sodium (positive control) (in 0.5%) the standard anti-arthritic control, Group IV received chloroform extract of *Peganum harmala* (PHC), Group VI received alcoholic extract of *Peganum harmala* (PHE) extract for 28 days from the induction day (0–28 days treatment), Group V and VII received chloroform extract of *Peganum harmala* (PHC) and alcoholic extract of *Peganum harmala* (PHE) for 14 days from the 14th day of induction (14th to 28th day treatment). The body weight changes were observed on every week. (Table 1).

Arthritis was induced in rats by the injection of 100 μl of Complete Freund’s Adjuvant (CFA) into the sub plan tar region of hind paw of the rats [21]. On 29th day, at the end of experiment, all animals were sacrificed by cervical decapitation and blood was collected in plain and EDTA containing tubes.

**Arthritic assessment**

To follow the course of the disease, swelling of the hind paw was determined with vernier calipers and body weight was measured by electronic weighing balance. The paw volume was periodically measured on 0th, 7th, 14th, 21st and 28th day (after injection of CFA) thereafter to confirm the reduction of RA. The locomotor activity was determined [22].

**Hematological and biochemical assessment**

Hemoglobin was measured in rat blood by the Erba auto analyzer before and after the induction of arthritis. After treatment with the standard and various extracts hemoglobin was measured and compared with the control group.

Blood was withdrawn on the day 7,14,21,28 from every rat through the orbital puncture into a blood collecting tube to measure erythrocyte sedimentation rate (ESR). The ESR was determined by the method of Huxtable [23]. The antioxidant parameters superoxide dismutase (SOD) and lipid peroxidation (LPO) was estimated in plasma by enzymatic testing kits [24].

**Histological and radiographic studies**

For histopathology, the proximal interphalangeal joints were removed, washed with saline and stored in 10% formalin on 29th day from two animals per group. The interphalangeal joints sections were obtained stained with eosin-haematoxylin stain and viewed under 100 X magnifications. The hind (arthritic induced) legs of the experimental rats were taken for X-ray, and examined for soft tissue swelling, bony erosions and narrowing of the spaces between joints.

**Statistical analysis**

The results were expressed as the mean ± standard error mean of the mean of 6 animals for each group. Statistical differences were evaluated using a One-way analysis of variance (ANOVA) followed by Dunnett’s test. Results were considered to be statistically significant at $p<0.05$ (or less).

| S. No. | Group Details |
|-------|---------------|
| 1. | Group I as control (GI) Sod. CMC 0.5% w/v in distilled water 5 ml/ kg, p.o |
| 2. | Group II as Arthritic control (GII) Mycobacterium butyricum in paraffin oil 10 mg/ml |
| 3. | Group III as standard drug (GIII) Diclofenac sodium suspended in 0.5% w/v sodium CMC, mg/ kg p.o. |
| 4. | Group IV as test (GIV) 100 mg/kg p.o. body weight of chloroform extract of *Peganum harmala* seeds (PHC) for 28 days. (Treatment-1) |
| 5. | Group V as test (GV) 100 mg/kg body weight of chloroform extract of *Peganum harmala* seeds for (PHC)14 days (Treatment-2) |
| 6. | Group VI as test (GVI) 100 mg/kg body weight of ethanolic extract of *Peganum harmala* seeds for (PHE) 28 days (Treatment-3) |
| 7. | Group VII as test (GVII) 100 mg/kg body weight of ethanolic extract of *Peganum harmala* seeds (PHE) for 14 days (Treatment-4) |
Results

Morphological analysis
Morphological features help in identification and confirmation of crude herbal drugs. The seeds are 3–4 mm in diameter, brown in color and triangular or conical in shape with micropyle at tip. (Fig. 2).

Preliminary phytochemical screening
Following percentage yield was obtained after successive solvent extraction of PH seeds: (Table 2).

Preliminary phytochemical tests had shown that petroleum ether extract contains fats; chloroform extract contains alkaloids, proteins and flavonoids. Ethanolic extract showed alkaloids, carbohydrates, phenolic compounds and flavonoids. Aqueous extracts has shown the presence of carbohydrates and phenolic compounds. The chloroform (PHC) and ethanolic extracts (PHE) were selected for further studies.

Modulatory effect of PHC and PHE on hind paw volume
The paw volume of the hind paws was measured by Vernier calipers on day 0,7,14, 21 and 28th day of the experiment. Erythema and edema were the inflammatory signs that developed over a 24 h period and increase thereafter in the present study in hind paws injected with CFA. The arthritic inflammation was significantly decreased ($p<0.01$) in arthritic rats treated orally from 0 day with PH extracts compared with those receiving vehicle and negative control (Figs. 3 and 4). Treatment group 3 has significant reduction in inflammation ($p<0.001$). All treatment groups of PHE extract shown marked decrease in arthritic inflammation ($p<0.01$). The PHE extract (0.31±0.06) was more effective than PHC (0.34±0.04) in mean changes in paw volume in 28 days. The effect of PH extracts on hind paw volume as percentage inhibition was in following order: GVI (63.09%) >GVII (61.90%) >GIV (60.71%) >GV (59.52%).

Modulatory effects of PHC and PHE on body weight changes
The present study showed that inflammatory consequences in arthritic joints have an impact on weight loss and it correlates with the extent of joint inflammation to the grade of weight loss. With the induction and progression of arthritis after one week of adjuvant injection, significant weight loss was observed in the arthritic rats, which get reduced in subsequent weeks. The treatment group (1st group) and standard drug treated groups showed less weight loss. (Fig. 5) PHE (GV and GVII) and PHC (GV) significantly alleviated ($p<0.05$) and GIII ($p<0.001$), the body weight loss of arthritic rats during the treatment course. The improvement of body weight gain of arthritic rats by PH extracts from day 0 was concomitant with food intake.

Modulatory effect of PHC and PHE on Locomotor activity
Locomotor activity of all animals was assessed during experiment on the day of 0,7,14,21 and 28th day with the help of Actophotometer instrument. The PH extracts and standard drug shown significant improvement ($p<0.001$) on 21st and 28th day, in the movement of rats which indicates the recovery in arthritic complications (Fig. 6). PHE extract was more effective than PHC and showed alleviative effect in 28 days.

Modulatory effects of PHC and PHE on hemoglobin content
On the day 0, 7, 14, 21 and 28th day of experiment hemoglobin content was observed and significant elevation in Hb content was observed in arthritic rats when compared with others group rats (Fig. 7). Hb content was significantly decreased by 14.34±0.58 to 12.84±0.07 in arthritic rats. The PH extracts significantly alleviated anemic condition in arthritic rats when given from 0 to 28 days. This alleviative effect of PHC was comparative with the standard drug and greater than the PHE. The PHC and standard drug shown more significant results than PHE when compared with arthritic control ($p<0.001$). The Hb content observed in 28 days in different groups was: GII (12.84±0.07) < GV < GVII < GVI < GI < GIV < GIII < GI (14.47±0.16).
Modulatory effects of PHC and PHE on ESR
ESR was significantly decreased with both group of PHE treatment as compared to vehicle treated arthritic rats and PHC treated groups. There was decrease in the levels of ESR from 8.4 to 7.5 mm per hour (mm/h) in group VI and 8.7 to 8.0 (mm/h) in group VII of PHE treated groups ($p$ < 0.05). There is significant increase in ESR levels in negative control group from 4.2 to 13.8. (Fig. 8).

Modulatory effects of PHC and PHE on oxidative stress
In Treatment groups (GIII, GVI and GVII) SOD level elevated significantly as compared to negative control groupII. However, lipid peroxidation significantly reduced which was measured in the form of MDA in liver. This reduction in MDA level (an indicator of LPO) was statistically significant ($p$ < 0.001) as compared to arthritic control group of rats (Figs. 9 and 10). PHE (204±0.09) was more effective than PHC (196.1±0.01) when compared with the healthy control rats on 28th day. The level of SOD were found to be significantly decreased ($p$ < 0.001) in arthritis rats (81±0.05) compared to control group (272.60 ±0.04). On the contrary, *Peganum harmala* treatment to arthritis rats showed a significant increasing effect in superoxide dismutase levels in developing phase ($p$ < 0.001).

The level of lipid peroxidation (MDA) were found to be significantly increased ($p$ < 0.001) in arthritis rats (108.4±0.35) compared to the normol control rats (28.7±0.11) and decreased in GVI and GVII (PHE treated).

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**Fig. 3** CFA induced inflammation in rats

**Fig. 4** Effect of PH extracts on mean change in paw volume. Values are Mean ± S.E.M, $n$ = 6 animals in each group. Results are Mean ± S.E.M. ANOVA, Dunnett’s test ***$p$ < 0.001, **$p$ < 0.01 (treated groups Vs arthritic control group II)
On the contrary, *Peganum harmala* treatment to arthritis rats showed a significant lowering effect in lipid peroxidation levels in developing phase (42.1±0.01) (Fig. 10).

**Radiology of hind paw after 28 day of induction of CFA**

Radiological imaging is a useful tool which helps to understand the severity of the disease. In these studies there was no deformities observed in ankle joint whereas in negative control bony destruction, soft tissue welling and narrowing of the joint spaces was observed. (Fig. 11a-g) Group received standard drug had prevented this bony destruction and there is minor swelling of the joint, Treatment 1st to 4th groups have no bony destruction and also there is no swelling of the joint. Treatment-III (GVI) and Treatment-IV (GVII) group has shown significant prevention against bony destruction by showing less soft tissue swelling and narrowing of joint spaces when compared with
Treatment II (GV), I (GIV) and negative control group (GII).

**Histopathology of joints**
The sections of proximal interphalangesal joints collected from the experimental rats were studied to observe the histopathological changes. (Fig. 12a-g) The degree of pannus formation, synovial hyperplasia and erosion of bone and cartilage were observed. The rats of group-II (negative control) showed edema formation, synovial hyperplasia and degeneration of cartilage. The infiltration of inflammatory exudates in articular region with bone marrow destruction and pannus formation was also observed when compared with the normal control group (Group-Ia). The test groups (group VI and VII) (Fig. 12f and g) showed protective effect as less inflammatory features were observed after 28 days treatment. But are comparable to the standard drug treated group III (c) showing normal bone marrow and lesser inflammatory features.
exudates. The small quantity of cellular infiltrates, normal bone marrow and absence of edema formation in the test groups exemplifies their preventive role for inflammatory responses. The test drug treated both groups prevent the ankle joint from degeneration, comparative to the negative control group.

**Discussion**

Adult population suffering from RA is around 1% worldwide in which the causative factor is unknown and pathogenesis is poorly understood [25, 26]. The inflammatory mediators released in arthritis are responsible for the pain, destruction of bone and cartilage that affects the movement of 80% suffering persons and adversely affects their quality of life. Arthritis induced with an antigenic solution of CFA stimulates cellular mediators and potentiates the production of certain immunoglobulins. This makes it beneficial in evaluating drugs for RA treatment as well as for assessing the anti-arthritis potency and anti-inflammatory potential.
of drugs [27]. This model in rats closely mimics the clinical features of human RA which makes it appropriate for evaluation [28].

The extent of inflammation in CFA induced rats is measured as swelling in paw represented by an index. Reduction in paw swelling is used as an indicator for the antiarthritic potential of various drugs [29, 30], it was employed to determine the activity of PH extracts at the dose level 100 mg/kg p.o. Groups received PH extracts showed significant reduction in paw volume especially PHC than PHE when compared with the arthritic control group (Fig. 3-4). The alleviative effect of PH extracts (from zero day) on paw edema in arthritic rats may be due to its inhibitory action on the release of pro-inflammatory cytokines [e.g. tumor necrosis factor (TNF-α) and interleukin (IL-1β)]. Those mediators affect the release of free radicals and other inflammatory mediators such as NO and prostaglandin (PGE₂) via specific enzymes [29]. In the present investigation, PH extracts specially the PHE showed significant reduction in paw edema.

The decrease in weight gain may be due to appetite loss, the decrease in intestinal absorption capacity (arising from intestinal oxidative damage), and/or metabolic disorders resulting from excessive release of systemic pro-inflammatory mediators and free radicals (rheumatoid cachexia) (Fig. 5) [31, 32]. This was observed in negative control arthritic rats. The improvement in body weight gain is a good and simple indicator for arthritis recovery during the treatment course of CFA in rat models. The result of present investigation reveals the potential of PH extracts in protecting against the weight loss in arthritic conditions.

In present study the rats received PH extracts showed significant activity in actophotometer. It may be due to the immunomodulation effect of PH extracts. As there is...
significant decrease in locomotors count in CFA induced arthritic rats and this may be due to the stimulation of immune system against the invading antigens [33]. The lesser movement of activity of animals in actophotometer especially in arthritic groups was due to the stiffness and pain in joints. (Fig. 6).

Results of blood examinations e.g. hemoglobin, white blood cell count and pack cell volume reveals the functioning of bone marrow [34]. In our investigation on hemoglobin count in CFA induced rats, haemoglobin level was decreased in arthritic rats which represents the anemic condition. This was supported by reports on similar arthritis model. This may due to the two reasons, abnormal storage of iron in the reticuloendothelial system and synovial tissue, and the failure of bone marrow to respond to anemia [35] (Fig. 7). The hematological changes in arthritic rats indicated anemia which may be due to gastrointestinal bleeding (due to oxidative damage of gastrointestinal bleeding) and/or inhibition of bone marrow erythropoiesis synthesis by pro-inflammatory cytokines, especially TNF-α, IL-β and IL-6) [36]. The PH extracts especially PHE significantly alleviated the condition in arthritic rats.

The increase in ESR count is a clinical feature of chronic arthritic conditions. In our study PH extracts especially PHE after 28 days shown significant results by improving the conditions when compared to negative control group. PH extracts and standard drug counteracted on the increased ESR in arthritic rats and improved the conditions. (Fig. 8).

In RA conditions the reactive oxygen species are produced (especially superoxide anion and hydroxyl radicals) by phagocytic cells due to inflammatory events, which results in decrease in cellular antioxidants. During oxidative stress membrane lipid peroxidation and lysosomal destruction causes cellular lysis and supersaturation of antioxidants depletes them. The elevated level of myeloperoxidase (MPO) and MDA causes oxidative damage of the gut mucosal membrane [37]. In our present investigation the levels of MDA (an end product of peroxidized lipid decomposition) was increased significantly in CFA treated rats as compared to control rats. Treatment with PHE resulted in significant reduction in MDA levels in plasma whereas the SOD level is increased in rats. Excessive oxidative stress (ROS) could damage the synovial fluid and joint collagen by depleting SOD through oxygen radicals [38]. However administration of PHE to arthritic rats resulted in a significant increase in SOD activity and reduces the LPO level near to normal control rats which show ROS scavenging or antioxidant property. (Figs. 9 and 10) The phytochemical screening of PHE revealed the presence of alkaloids, carbohydrates, phenolic compounds and flavonoids. The presence of alkaloids, flavonoids and phenolic compounds may be responsible for antioxidant and anti-inflammatory action.

The histological architecture of rat joints was found normal in histological studies. The PH treated both groups and standard drug has shown protective effect on the joint structure. They tend to ameliorate the normal histological architecture with cartilage protection. The overall prevention of the inflammatory signs of the rat ankle joints was significant in 28 days drug treated group compared with 14 days drug treated group. Degeneration of the ankle joint was not observed in any of the drug treated groups when compared with the negative control. (Fig. 12).

Radiological studies also supported the anti-arthritic activity of PH. PHE treatment for 28 days have shown significant prevention against bony destruction by showing less soft tissue swelling and narrowing of joint spaces when compared with 14 days PHE treated group.

**Conclusion**
PHE at the dose of 100 mg/kg has a significant therapeutic effect than PHC in alleviating the articular/ extra-articular oxidative damage in CFA induced rat model especially when the treatment started from the day of arthritis induction. The presence of alkaloids and flavonoids might have potentiated the anti oxidant and antiarthritic activity. Further, experimentation is required for characterization and quantitative determination of bioactive phyto-constituents which are responsible to maintain the therapeutic efficacy.

**Supplementary Information**
The online version contains supplementary material available at https://doi.org/10.1186/s40816-020-00243-3.

**Additional file 1.**

**Abbreviations**
CFA: Complete Freund’s Adjuvant; PH: Peganum harmala; PHC: Chloroform extract of PH; PHE: Ethanic extract of PH; RA: Rheumatoid arthritis; ROS: Reactive oxygen species; RNS: Reactive Nitrogen species; NSAI-D’s: Non-steroidal anti-inflammatory drugs; DMARD’s: Disease modifying anti-rheumatic drugs; DTNB: (5−5′-dithiobis [2-nitrobenzoic acid]); MDA: Malondialdehyde; SOD: Superoxide dismutase; LPO: Lipid peroxidase

**Acknowledgements**
Authors are thankful to the Sophisticated Instrumentation Center (SIC), Dr. Harisingh Gour Vishwavidyalaya, Sagar for sophisticated instrumentation facilities supported under DST-PURSE (II).

**Authors’ contributions**
Both authors planned the study, designed all experiments, summarized, discussed and interpreted the results. The author(s) read and approved the final manuscript.

**Funding**
This research received no specific grant from any funding agency in the public, commercial or not for profit sectors.
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