Molecular screening of phytochemicals from *Amelanchier Alnifolia* against HCV NS3 protease/helicase using computational docking techniques

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**Abstract:**
Hepatitis C is serious health concern worldwide caused by HCV. It causes liver cirrhosis and hepatocellular carcinoma. Development of prevention solutions is under progress. Meanwhile, the treatment of the viral disease using compounds isolated from natural medicinal plants is promising. The traditional use of photo-chemicals from medicinal plants like *Amelanchier alnifolia* for viral treatment is hopeful. Therefore, it is of interest to screen for flavonoids from *Amelanchier alnifolia* against protein targets of HCV. Hence, we assessed the binding of flavonoids to HCV NS3/4A protease and helicase proteins. Results show that Quercitin 3-galactoside and 3-glucosides showed good binding score with protease and helicase respectively. Their interaction/binding sites are documented in this report. This data provide insights for the consideration of flavonoids as potential inhibitors of HCV/NS3/4A protease and helicase.

**Background:**
Hepatitis C Virus (HCV) is an endemic worldwide problem which affects 170 million people globally and 10 million people in Pakistan [1]. It causes acute and chronic hepatitis and is a main cause of liver cirrhosis and hepatocellular carcinoma [2]. HCV belongs to *Flaviviridae* family with a positive-sense single-stranded RNA genome which encodes three structural proteins (Core, E1, E2) and six non-structural proteins (NS2, NS3, NS4A, NS4B, NS5A & NS5B) [3]. Among all HCV proteins, NS3/4A serine protease and helicase are effective drug targets to develop anti-HCV agents [4]. The basic role of NS3/4A is the proteolytic processing at NS4A/4B, NS4B/5A, and NS5A/5B sites, and it shows a vital role in HCV replication. As it is involved in viral replication, it has worked as a reliable drug target for HCV. Till today, no vaccination is available for treatment of HCV and present standard of care is a combination therapy of Pegylated interferon alpha (PegIFN-α) injections with oral antiviral nucleoside analogue ribavirin (RBV) which leads to treatment of HCV in 50% genotype 1 cases and 80% of genotype 2 cases but this treatment no rapid response and side effects in genotype 1a and 1b patients [5-7]. Present treatment is expensive. Less effective and has numerous side effects thus, there is a need of developing antiviral agents that are less harmful and hasability to target all genotypes of HCV with the same competence. Recently, two NS3 protease inhibitors have been approved as triple therapy (PEG-IFN- α, ribavirin and Boceprevir or Telaprevir) against HCV [8]. But still there is a strong need to develop specific compounds that can target important factors of the HCV life cycle [9]. Several Medicinal plants are tested and many of them are proved to have antiviral effect in their phytochemicals. Medicinal plants are cost-effective, multiple target activities, minor side-effects and thus,
preferred over conventional treatment [10-14]. Phytochemicals such as alkaloids, organosulfur compounds, limonoids, lignans, furyl compounds, polyynes, thiophenes, proteins, peptides, flavonoids, terpenoids, sulphides, polyphenolics, coumarins, saponins, chlorophyllins have functions like scavenging, antioxidant activities, hindering viral entry, DNA and RNA replication against numerous viruses [15]. Recently, our group reported that phytochemicals Amelanchier alnifolia showed novel inhibition of HCV titer in infected liver cells [16]. Therefore, this study was planned to screen phytochemical of Amelanchier alnifolia against HCV NS3/4A protease and helicase using in-silico approaches.

Figure 1: Chemical structures of Amelanchier alnifolia phytochemicals constructed using ChemDraw Software.

Methodology:
This study was designed to dock Amelanchier alnifolia phytochemicals against HCV NS3/4A protein with the following communications. Intel (R) xenon (R) CPU E5620@2.40GHz system having 3.8GB RAM with the open 11.4 (X 86, 64) operating platform. Protein-ligand docking was carried out using the MOE (Molecular Operating Environment) software package. Interactions between HCV NS3/4A protease and ligands were imagined using ligPlot feature of MOE software [17].
Ligand preparation

A literature was searched to find Amelanchier alnifolia phytochemical structures. Chemical structures of phytochemicals were drew using ChemDraw software (Figure 1) and were transformed into their respective 3D structures. 3D structures were optimized by addition of hydrogens through MOE software package. Energies of selected molecules were minimized with parameters (gradient: 0.05, Force Field: MMFF94X, Chiral Constraint and Current Geometry). Molecules were then saved in .mdb format and were used as input file for MOE-Docking.

Preparation of Receptor Protein

HCV NS3/4A protease and helicase structures were downloaded from Protein Data Bank using PDB ID: 3P8N and 2fm2 and was optimized by removal of water molecules, 3D protonation and Energy minimization. Energy minimization was carried out using parameters (gradient: 0.05, Force Field: MMFF94X+Solvation and Chiral Constraint: Current Geometry). Energy minimization was stopped when the root mean square gradient falls below the 0.05. The minimized structures were used as the receptor protein for Docking.

Molecular Docking

MOE docking program with default parameters was used to bind the selected ligands with receptor protein and to find the correct conformation (with the rotation of bonds, structure of molecule is not rigid) of the ligand so as to obtain minimum energy structure. After docking, best conformations were analyzed for hydrogen bonding/π-π interactions.
Results & Discussion:

Docking Analysis

All ligands were docked with the pocket of NS3/4A protease and helicase enzyme and top ranked conformations of each ligand were saved. Compound 01 was ranked as top docked molecule on the basis of S-Score obtained from MOE docking algorithm. Thus, it can be concluded that this compound can serve as competitive inhibitor against NS3/4A protease and helicase. All other compounds were having S-Score close to each other. All compounds were studied in detail to obtain their interaction information that can be important for the inhibition of NS3/4A protease and helicase enzyme. Interaction diagrams were obtained by using MOE ligand interaction analysis feature.

Binding interactions of Ligands and Protein

The most active Compound 01 was ranked as first on the docking score. It is clear from (Figure 2) that this compound was bound deep into the binding cavity of NS3 protease making interactions with the residues His 57, Asp 81 and Asp 168. Similarly, Compound 01 ranked first on the docking score which showed deep binding with NS3 helicase cavity making interactions with the residues Ala 157, His 528. On the basis of docking score Compound 02, Compound 03, Compound 04, Compound 05, Compound 06, Compound 07, Compound 08, Compound 09, Compound 10, and Compound 11 were ranked close to each other as shown in Table 1 (see supplementary material). There were some visible interactions observed between the Compounds and the binding site residues Asp 81, His 57 and His 528. The solvent contact was found almost all over the ligand.

Conclusion:

The design and development of prevention tools for Hepatitis C viral infection has been a challenge. Therefore, improved strategies for the treatment of Hepatitis C are relevant to the context. The use of extracts from *Amelanchier alnifolia* containing flavonoids shows promising results intraditional treatment pipelines. Hence, it is interest to document thepotential binding of *Amelanchier alnifolia* derived flavonoids with HCVNS3/4A protease and helicase. Molecular docking and binding simulations ofsuch flavonoids with HCV target proteins show the good binding ability ofquercitin 3-galactoside and 3-glucoside with protease and helicase, respectively. These observation provide insights to consider *Amelanchier alnifolia* derived flavonoids as potential inhibitors of HCV target proteins.

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Supplementary material:

Table 1: Interaction of phytochemicals with HCV NS3 protease and helicase

| Phytochemicals | S Score | Rmsd Value | Interacting residues | Phytochemicals | S Score | Rmsd value | Interacting residues |
|----------------|---------|------------|----------------------|----------------|---------|------------|----------------------|
| Quercitin 3-galactoside | 20.4386 | 2.5218 | His 57, Asp 81, Asp 168 | 3-glucoside | 17.3470 | 2.0156 | Ala 157, His 528 |
| 3-glucoside | 19.8765 | 1.6555 | Asp 168, Arg 155 | Quercitin 3-galactoside | 13.5612 | 1.0159 | Val 524, Gln 526 |
| Epicatechin | 18.5096 | 2.5606 | Asp 81, Ala 157 | Chlorogenic acid | - | 8.0074 | His 528 |
| Hydroxybenzoic acid | 12.4426 | 1.2542 | Ser 139 | Flavanol glycosides | 10.9378 | 1.9460 | Asp 527 |
| Flavanol glycosides | 11.0442 | 2.3385 | - | epicatechin | 10.8113 | 1.8173 | Asp 79 |
| Chlorogenic acid | -9.9895 | 6.8368 | - | Hydroxybenzoic acid | -7.1618 | 1.6794 | - |