Experimental approach, theoretical investigation and molecular docking of 2-chloro-5-fluoro phenol antibacterial compound

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A B S T R A C T
The molecular structural dimerization of biologically potent 2-chloro-5-fluoro phenol (2C5FP) is optimized. A combined experimental and theoretical characteristics of vibrational spectral determinations (NMR, FT-IR and Raman) on 2-chloro-5-fluoro phenol (2C5FP) were used at DFT-B3LYP/6-31+G(d,p) level of computation. A close coherence is achieved when experimentally observed wave numbers are compared with calculated wave numbers by refinement of the scale factors. Calculated values of global chemical descriptors of the present molecule reveal significant molecular stability and chemical reactivity. Non-Linear optical (NLO) property of the present molecule is investigated by determining the second order non linear parameter of β. Moreover, hydrogen bond and thermodynamic parameters at various temperatures are determined and discussed. Investigated compound 2C5FP possesses a better antibacterial activity against Echerichia coli, Streptococcus aureus and Staphylococcus aureus, respectively. The title molecule is subjected to molecular docking studies with two different proteins, namely Staphylococcus aureus Tyrosyl-tRNA synthetase (PDB ID: 1JIL) and human dihydroorotate dehydrogenase (hDHODH) (PDB ID: 6CJF). The results of molecular docking analysis support the antibacterial activity and demonstrate a strong interaction with the DHODH inhibitor.

1. Introduction

The antioxidant potential of phenolic compounds exhibit potent activities in the treatment of cancer. These compounds are considered as secondary metabolities, which are most significant bioactive agent in chemo preventive and chemotherapeutic effects in cancer [1]. Phenolic compounds, ubiquitous in edible plants like, flavonoids, chromones, tanins, lignans, quinines, phenolic acids and many more. Polyphenols have reported to many industrial applications. For instance, they can be used as natural colorants, food preservatives and in the production of paints, paper and cosmetics [2]. One of the most important polyphenolic flavonoids found in fruits and vegetables [3] (Apple, grapes, tomato, green tea pine and many others) act as antioxidants due to their redox potential possessing anticancer activities on different cancer cell lines as in, MCF-7 breast cancer and HeLa cervical cancer cells, A549 lung cancer cell, Miapaca-2, Panc-1 and SNU-213 human pancreatic cancer cells [4].

More than 20 phenols were investigated that displayed inhibitory activities of the β-class carbonic anhydrase from the fungal parasite Malassezia globoasa (MgCA). All tested phenols exhibited better efficacy in inhibiting MgCA than the clinically used sulfonamide acetazolamide [5]. In particular, when humans regularly consume high levels of fresh fruits and vegetables, flavonoids have been associated with prevention of cancer and heart diseases. Phenyl sulfonifuroxan NO-donor phenols are preventive agents for metabolic syndrome since metabolic syndrome (Mets) give rise to dyslipidemia, raised fasting glucose, renal and liver diseases, for the development of cardiovascular diseases (CVD) and type-2 diabetic mellitus (T2DM) [6].

The hydroxyl group of aromatic ring from the phenolic compounds displays antimicrobial, anti-arthrogenic, anti-inflammatory, cytotoxic, and pharmacological activities [7], respectively. Flavonoids, consisting of two aromatic rings A and B, joined by a 3-carbon bridge have low molecular weight compounds with fifteen carbon atoms arranged in

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C6-C3-C6 configuration [2]. Flavonoids are commonly found in phytochemicals which can be used to protect the plant against UV light, herbivores, fungal parasites, oxidative cell injury and hypo lipidemic [8]. Due to low toxicity fluorine derivatives of phenols have wide variety of applications in agricultural chemicals, dyes biocides and potential therapy of skin disorders [9], respectively. Chlorophenols used for many applications in the photochemical industry is due to the substituted donor (OH) and acceptor (CL) group of atoms in the benzene ring [10].

Considering the above remarkable significant applications of phenol derivatives in pharmaceutical industry, the antibacterial activity and molecular docking have been reported for title compound 2-chloro-5-fluoro phenol. The vibrational spectroscopic studies of various phenol derivatives have been reported in earlier literatures [11, 12]. N. Sunderaganesan et. al and D. Mahadevan et. al were investigated the vibrational analysis of 2,3- difluoro phenol and 3- bromo phenol [13, 14]. The substitution of halogen atoms on the title compound influences the drug activity and binding affinity by molecular docking. The previous study reports that the spectroscopic investigations and DFT studies of 2-chloro-5-fluoro phenol have not been reported so far. Hence, detailed theoretical studies and an extensive experimental investigation on IR, Raman and NMR spectra of the title molecule have been performed. Besides, intra and intermolecular hydrogen bond interaction, global chemical parameters are determined from HOMO-LUMO energy gap and NLO activities of the title compound have been carried out. Finally, due to potential biological activities of 2C5FP molecule is subjected to antibacterial activity.

2. Materials and methods

2.1. Experimental

The fine sample 2C5FP was procured from Lancaster chemical company, UK. The FT-IR spectrum was recorded in BRUKER IFS 66V spectrometer and FT-Raman spectrum was recorded using FRA-106 Raman module equipped with Nd: YAG Laser source operating at 1.064μm width with 200mW power. Both FT-IR and FT-Raman spectra were measured in the region of 4000–100cm–1 and 3500-100cm–1, respectively. The spectral wave numbers of all bands are accurate to ± cm–1. The carbon and proton (13C–1H) NMR spectra were recorded in Acetone using TMS as an internal standard on Bruker NMR spectrometer at 400 MHz. The spectral measurements were recorded at IIT (SAIF), Chennai, India.

2.2. Antibacterial activity

The isolates of bacterial strains viz, Streptococcus aureus, Staphylococcus aureus, (Gram – positive bacteria), Escherichia coli and Pseudomonas aureus (Gram – negative bacteria) were obtained from the Department of Microbiology, Srimad Andavan College, Trichy, Tamilnadu, India.

2.2.1. Disc diffusion method

The antibacterial activity of the title compound 2C5FP is performed by the in-vitro disc diffusion technique described by Kirby and Bauer [15]. The antibacterial activity is determined over the active compound (2C5FP) against two gram – positive isolates and against two gram – negative isolates. In the disc diffusion method, the compound solution was prepared by dissolving the test compound (2C5FP) in 90% ethanol in the concentration of 1 mg/ml. The sterile paper discs with 6mm in diameter loaded with test compound were placed in test bacteria spread agar plates. The bacterial plates are incubated at 37 °C [16, 17]. Standard antibiotic amoxicillin was used as positive control with concentration of 1 mg/ml. The results obtained for the tested compound as the average diameter of inhibition zone of bacterial growth around the discs in mm after 24h incubation period.

2.3. Methods of computation

All Computations were accomplished with Gaussian 09 software package of programs for the title compound using DFT with Becke's three parameter exchange functional in combination with the Lee-Yang-Parr correlation functional (B3LYP) with standard 6-31++G (d, p) level of computations. In order to minimize the energy, the geometric optimization was performed along with all geometrical parameters without imposing molecular symmetric constraints. After geometric optimization, the vibrational assignments were made on the basis of potential energy distributions using MOLEVIB Program (version V7.0-G77) written by Sundis [18, 19]. Calculated theoretical wave numbers were quite close to experimental wave numbers when they were scaled using MOLEVIB. The scale factors were determined using linear correlation method between the theoretical and experimental fundamental modes. The global chemical indices namely ionization potential, electron affinity, hardness, softness, chemical potential, and electrophilicity index have been obtained from HOMO-LUMO analysis performing at B3LYP/6-31++G (d, p) basis set.

2.3.1. Molecular docking

The crystal structures of staphylococcus aureus tyrosyl-tRNA synthetase (PDB ID: 1JIL) and the hDHODH inhibitor (PDB ID: 6CFJ) are downloaded from the RSCB Protein Data Bank (PDB). The optimized molecular structure using DFT is useful to prepare the ligand PDB format. The ligand is prepared by minimizing the energy at B3LYP/6-31++G (d,p) level of computation. The protein is prepared by eliminating the water, co-factors and co-crystalline ligands. By using the Auto Dock Tools (ADT) graphical user interface is used to compute the Kollman charges and polar hydrogen. The bioactive conformation is simulated using Autodock 4.2 software package and the corresponding results are analyzed using PyMol software [20, 21]. Partial charges are computed by Gasteiger method and the receptor grids are generated using 90 × 90 × 90 A0 grid points in XYZ coordinates.
Table 1. Calculated structural parameters of 2-Chloro-5-fluoro phenol at B3LYP/6-31++G (d,p) method.

| Bond Length (Å) | Parameters | Monomer | Dimer | Exp |
|-----------------|------------|---------|-------|-----|
| C1-C2           | 1.41       | 1.41    | 1.41  |
| C1-C6           | 1.40       | 1.40    | 1.38  |
| C1-O12          | 1.36       | 1.35    | 1.38  |
| C3-C3           | 1.39       | 1.39    | 1.41  |
| C2-C10          | 1.75       | 1.75    | 1.74  |
| C3-C4           | 1.40       | 1.40    | 1.42  |
| C5-H7           | 1.08       | 1.08    | 1.09  |
| C4-C5           | 1.39       | 1.39    | 1.04  |
| C4-H8           | 1.08       | 1.08    | 1.09  |
| C5-C6           | 1.39       | 1.39    | 1.38  |
| C5-F11          | 1.36       | 1.36    | -     |
| C6-H9           | 1.09       | 1.09    | 1.09  |
| O12-H13         | 0.97       | 0.97    | 0.99  |

| Bond Angle (°) | Parameters | Monomer | Dimer | Exp |
|----------------|------------|---------|-------|-----|
| C6-C1-C6-C5    | 119.24     | 118.92  | 118.0 |
| C6-C1-O12-C10  | 122.40     | 122.51  | 124.0 |
| C5-C2-C3-C4    | 120.28     | 120.56  | 121.0 |
| C1-C2-C10-C110 | 119.70     | 119.60  | -     |
| C5-C2-C10-C110 | 120.02     | 119.83  | -     |
| C3-C2-C3-C4    | 120.91     | 120.82  | 120.0 |
| C2-C3-C4-C5    | 118.97     | 119.02  | -     |
| C4-C3-C4-C5    | 120.12     | 120.14  | -     |
| C5-C4-C5-C6    | 117.76     | 117.65  | 119.0 |
| C5-C4-H8-C9    | 121.72     | 121.77  | -     |
| C5-C4-H8-C9    | 120.51     | 120.58  | -     |
| C4-C5-C6-C7    | 122.83     | 123.03  | 123.0 |
| C4-C5-F11-C111 | 118.93     | 118.93  | -     |
| C6-C5-C6-C7    | 118.04     | 118.05  | -     |
| C1-C1-C6-C5    | 120.23     | 120.87  | -     |
| C1-C1-C6-C5    | 119.79     | 120.13  | -     |
| C1-O12-H13-C13 | 110.13     | 111.12  | -     |

| Dihedral Angle (°) | Parameters | Monomer | Dimer |
|--------------------|------------|---------|-------|
| C6-C1-C2-C3-C4     | 0.00       | 0.00    |
| C6-C1-C2-C10-C110  | 180.00     | -180.0  |
| O12-C1-C2-C3-C4    | 180.00     | 180.00  |
| O12-C1-C2-C10-C110 | 0.00       | -0.10   |
| C2-C1-C6-C5-C6     | 0.00       | 0.00    |
| C2-C1-C6-C5-C6-H9  | -180.00    | -180.00 |
| O12-C1-C6-C5-C5    | 180.00     | -180.00 |
| O12-C1-C6-C6-H9    | 0.00       | 0.50    |
| C2-C1-O12-H13-C13  | 180.00     | -179.09 |
| C6-C1-O12-H13-C13  | 180.00     | -138.17 |
| C1-C2-C3-C4-C5     | 180.00     | -0.07   |
| C1-C2-C3-H7-C8     | 0.00       | 180.00  |
| C1-C2-C3-H7-C8     | 0.00       | 180.00  |
| CL10-C2-C3-C4-C5   | 180.00     | -0.10   |

* Refs. [13, 14, 20].
3. Result and discussion

3.1. Molecular geometry

The optimized monomer and dimer structure of present compound 2C5FP is obtained from GAUSSIAN 09W program package. The optimized structure of monomer is shown in Figure 1. Comparative optimized structural parameters such as bond length and bond angle for two different conformers of title molecule is performed using B3LYP method 6–31++G (d, p) basis set as summarized in Table 1. Theoretical results demonstrate that most of the bond lengths and bond angles of monomer and dimer structure are well consistent with the similar theoretical and XRD data reported earlier [13, 14]. The calculated results show that the aromatic ring of the title molecule is perturbed from the regular hexagon as a result of the electronic effects of chloro, fluoro and bromo group. This leads to variation in the charge distribution on the carbon atom of the benzene ring and consequently causes the structural, vibrational and electronic properties of the molecule. In addition, the sharing of \( \pi \) electrons between the oxygen atom from OH group and electro negativity of fluorine and chlorine atoms affects the structural perturbations within the ring. Calculated bond lengths and bond angles are slightly overestimate in the bond lengths of C1-C6, C1-O12, C2-Cl10, C5-C6 and bond angles of C2-C1-C6, C1-C6-C5 & C2-C3-C4. A small reduction is observed in the bond length of C1-O12, C2-C3, C3-C4, C3-H7, C4-H8, O12-H13 and bond angles of C6-C1-O12 & C1-C2-C3 when compared with XRD values [22]. These small deviations between observed and calculated entities are obtained from different environments. It is evident that the experimental predictions belong to solid phase and theoretical results belong to gaseous phase.

The variation in bond lengths and bond angles depends on the substitution of electron withdrawing and electron donating groups in the benzene ring and various factors such as hybridization, size of the atoms,

Figure 2. Correlation graphic between experimental and calculated bond lengths of 2-chloro-5-fluoro phenol (a) Monomer (b) Dimer.

Figure 3. Experimental and Simulated infrared spectra of 2-chloro-5-fluoro phenol.
delocalization of electrons, lone pair of electrons and electronegativity difference between the atoms. For instance, the bond length of C2-Cl10 (1.75 Å) is greater than the bond length of C5-F11 (1.36 Å) which reveals that the size of the atoms increases as bond length increases. The C-X (X = F, Cl, Br...) bond distance denotes a considerable rising when substituted in the benzene ring in the place of C-H. In the title molecule, bond lengths of C-Cl and C-F nearly well coincides with 1.75 Å and 1.36 Å which is in good coherence with calculated value in the earlier literature for P-chlorophenol and 2,3-difluoro phenylboronic acid, respectively [23, 24]. A remarkable reduction observed in the bond angle of C1-O12-H13 (110.13 Å) is due to the presence of a lone pair of electrons in the central oxygen atom.

The linearity between the observed and computed bond lengths and bond angles of monomer and dimer structure of 2C5FP can be estimated from plotting the computed data against experimental ones which is depicted in Figure 2 and are described through the following equations.

\[
Y_{\text{Cal}} = 1.0226 \times Y_{\text{Exp}} - 0.03621 \quad (R^2 = 0.99607) \quad \text{for Monomer} \quad (1)
\]

\[
Y_{\text{Cal}} = 1.02044 \times Y_{\text{Exp}} - 0.03396 \quad (R^2 = 0.99997) \quad \text{for Dimer} \quad (2)
\]

3.2. Interpretations of fundamental modes

According to theoretical computations, 2-chloro-5-fluoro phenol (2C5FP) has a planar structure with 13 atoms and 33 fundamentals modes of vibration along with C1 point group symmetry. The comparisons of observed and simulated FT-IR and FT Raman spectra of 2C5FP are shown in Figures 3 and 4, respectively. In the present investigation, the vibrational assignments were predicted on the basis of potential energy distribution (PED) results have been carried out from the MOLVIB [19, 25] output. Table 2 reported the complete vibrational analysis performed at 6–31+G (d,p) basis set.

### 3.2.1. C-O & O-H vibration

The C-O stretching vibrations in P-mono substituted phenols observed as a strong band in the region 1300-1200 cm\(^{-1}\) [26]. The C-O stretching band coupled with C-C stretching modes vibration. In the present compound, the C-O stretching mode is identified at 1258 cm\(^{-1}\) in the FT-IR spectrum. The C-O in-plane and out-of-plane modes are identified within their characteristic region and the corresponding values are summarized in Table 2.

The O-H stretching modes are delicate to hydrogen bonding. A non-hydrogen bonded group absorbs strongly in the region 3700-3500 cm\(^{-1}\). In a five- and six-benzene ring compound, the presence of intra molecular hydrogen bond reduces the hydroxyl stretching mode to the 3559-3200 cm\(^{-1}\) region. In the O-H region, due to the halogen substitution of the benzene ring stretching mode turns to the higher region. Accordingly in 2C5FP, the O-H stretching mode is found at 3524 cm\(^{-1}\) in the FT-IR spectrum. This is well consistent with the
literature data [23]. The O-H in-plane and out-of-plane bending modes are generally appeared in the region 1530-1200 cm\(^{-1}\) and 720-590 cm\(^{-1}\) [13], respectively. In 2C5FP, the O-H in-plane and out-of-plane vibrations are assigned at 1267 cm\(^{-1}\) in FT-Raman and 453 cm\(^{-1}\) in FT-IR, respectively.

3.2.2. C-H vibration

Aromatic compounds generally shows the multiple weak bands due to C-H stretching vibration in the region 3100-3000 cm\(^{-1}\). The C-H in-plane bending vibration coupled with C-C stretching modes which occur in the region 1500-1100 cm\(^{-1}\) and C-H out-of-plane bending modes appear in the region 1000-800 cm\(^{-1}\) [27]. In this region, the modes are not affected substantially by the substituents. Hence, in the present molecule of the title compound, the C-H stretching vibration is assigned at 3106, 2926 and 3079 cm\(^{-1}\) in FT-Raman and 453 cm\(^{-1}\) in FT-IR, respectively.

The bands at 796, 743 and 696 cm\(^{-1}\) are assigned to C-H in-plane bending modes and the bands at 570, 564, 532 and 508 cm\(^{-1}\) are identified to C-H out-of-plane bending vibrations. These wave members are in line with the similar compound of 3-Bromo Phenol [14].

3.2.3. Ring vibration

Most of the vibrational bands are affected by the substituents in the aromatic ring. All the carbon atoms within the ring undergo coupled vibrations, called skeletal modes of vibration and exhibit in the region 1660-1420 cm\(^{-1}\) [28]. In the present investigation, the bands for C-C stretching vibration are assigned at 1699, 1607, 1536, 1487, 1316 & 1297 cm\(^{-1}\) in FT-IR and 1613, 1321 & 1297 cm\(^{-1}\) in FT-Raman spectrum. These assignments coincide very well with the earlier reports. Due to O-H substitution in the benzene ring leads to higher C-C stretching vibrations for the phenol compounds.

3.2.4. C-F and C-Cl vibrations

In the vibrational spectra of related molecules, the C-F stretching band which appeared over a wide range of frequency in the region 1360-1000 cm\(^{-1}\) is due to the influence of the adjacent substituents. For mono fluorinated compounds the C-F stretching modes appeared in the region 1110-1000 cm\(^{-1}\) in FT-IR and 1000-800 cm\(^{-1}\) in FT-Raman spectrum. These assignments coincide very well with the earlier reports. Due to O-H substitution in the benzene ring leads to higher C-C stretching vibrations for the phenol compounds.

### Table 2. Observed FT-IR, FT-Raman and calculated (unscaled and scaled) frequencies (cm\(^{-1}\)) of 2-Cloro-S-fluoro phenol at B3LYP/6-31++G(d,p) method.

| S.NO | C1 Symmetry | FT-IR | FT-Raman | Calculated frequency | Assignment with PED % |
|------|-------------|-------|----------|----------------------|----------------------|
|      |             |       |          | Unscaled             |                      |
| 1    | A           | 3524  | -        | 3829                | sOH (88)             |
| 2    | A           | 3106  | -        | 3233                | sCH (97)             |
| 3    | -           | 3079  |          | 3083                | sCH (95)             |
| 4    | A           | 2926  | -        | 3201                | sCH (88)             |
| 5    | A           | 1699  | -        | 1657                | sCC (88)             |
| 6    | A           | 1607  | 1613     | 1641                | sCH (78)             |
| 7    | A           | 1536  | -        | 1537                | sCC (70)             |
| 8    | A           | 1487  | -        | 1459                | sCC (72)             |
| 9    | A           | 1316  | 1321     | 1362                | sCC (79)             |
| 10   | A           | 1292  | 1297     | 1317                | sCC (70)             |
| 11   | A           | -     | 1267     | 1299                | sOH (76)             |
| 12   | A           | 1258  | -        | 1193                | sCO (72)             |
| 13   | A           | 1205  | -        | 1172                | sCH (69)             |
| 14   | A           | 1143  | -        | 1135                | sCH (79)             |
| 15   | A           | 1106  | -        | 1071                | sCH (71)             |
| 16   | A           | 1058  | 1059     | 991                 | sCH (69)             |
| 17   | A           | 970   | 974      | 941                 | sCH (62)             |
| 18   | A           | 933   | -        | 840                 | sCH (58)             |
| 19   | A           | 850   | -        | 806                 | sCH (53)             |
| 20   | A           | 796   | -        | 751                 | bCC (66)             |
| 21   | A           | -     | 743      | 693                 | bCC (68)             |
| 22   | A           | 729   | -        | 642                 | sCCL (70)            |
| 23   | A           | 696   | -        | 619                 | bCC (61)             |
| 24   | A           | 638   | 641      | 568                 | bCO (66)             |
| 25   | A           | 610   | -        | 512                 | bCF (61)             |
| 26   | A           | 564   | 570      | 459                 | bCCC (56)            |
| 27   | A           | 532   | -        | 380                 | bCCC (53)            |
| 28   | A           | 508   | -        | 348                 | bCCC (48)            |
| 29   | A           | 453   | -        | 322                 | sOH (52)             |
| 30   | A           | -     | 386      | 318                 | sCO (45)             |
| 31   | A           | -     | 338      | 234                 | bCCl (51)            |
| 32   | A           | -     | 244      | 226                 | bCCl (49)            |
| 33   | A           | -     | 73       | 119                 | bCCl (41)            |

Abbreviations: \(\\nu\) - stretching; \(\omega\) - in-plane bending; \(\omega\) - out-of-plane bending.
assignments C-Cl are identified and presented in Table 2. These vibrational assignments correlate well with the similar compounds of 2, 3, 4, 5-tetrachloro phenol [22].

The correlation between the observed and calculated frequencies is plotted and the corresponding graphics are given in Figure 5. The linear relationship is obtained between the experimental and theoretical wave numbers for IR and Raman, respectively by the following equations:

\[

\nu_{\text{Cal}} = 0.9979 \nu_{\text{Exp}} + 9.358 \quad (R^2 = 0.99997) \quad \text{(IR)}
\]

\[

\nu_{\text{Cal}} = 0.99633 \nu_{\text{Exp}} + 10.068 \quad (R^2 = 0.99997) \quad \text{(Raman)}
\]

As a result, the performance of B3LYP/6–31++G (d,p) method with respect to the determination of the wave numbers within the title compound for IR and Raman are quite close.

3.3. Error analysis of different fundamentals

It is a well known fact that the correlation between the observed wave numbers and theoretical wave numbers implies that the systematic errors are due to the regulation in inharmonic effects, lack of basis sets and electronic correlations. It is manifest and obvious that the evaluation of standard deviation (S.D) and scale factors provide fine agreement between the experimental and the calculated fundamental modes of vibrations. The RMS deviation and correlation coefficient for observed and calculated wave numbers at B3LYP/6–31++G (d,p) level have been carried out using the following equation [32, 33].

\[

\text{RMS} = \sqrt{\frac{1}{n-1} \sum (\nu_{\text{Exp}} - \nu_{\text{Calc}})^2}
\]

Where 'n' is the number of observed and calculated data. The RMS error of the frequencies between the unscaled and experimental wave numbers in IR and Raman is found to be 104.567 cm⁻¹ and for scaled frequencies it is 8.1557 cm⁻¹. In order to refine the observed fundamentals the scale factors were applied between the observed and calculated wave numbers and found to be 0.9858. The correlation graphics between the experimental and calculated wave numbers are quite close to the scaled frequencies as shown in Figure 5.

3.4. Electronic properties

In order to explore the molecular stability and reactivity, the energy gap of HOMO-LUMO is predicted. The key parameters of quantum chemistry is the highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO). HOMO energy exemplifies the donating electron while LUMO energy symbolizes the accepting electron ability. The energy gap is a crucial parameter for determining the electrical conductivity and other molecular properties of chemical systems.

| Molecular Properties (ev) | Homo Energy | Lumo Energy | Energy gap (ΔE) | Ionisation Potential (I) | Electron Affinity (A) |
|---------------------------|-------------|-------------|----------------|--------------------------|----------------------|
| 2-Chloro-5-fluoro phenol   | -6.67108    | -1.00786    | 5.6632         | 6.67108                  | 1.00786              |

Figure 5. Correlation graphic between the experimental and theoretical IR and Raman wave numbers of 2-chloro-5-fluoro phenol.

Figure 6. The frontier molecular orbitals of 2-chloro-5-fluoro phenol.
and it is also used for predicting the most reactive sites in \( \pi \)-electron systems of molecular species [34, 35]. The theoretically predicted energy gap between HOMO and LUMO for the title compound is found to be 5.6632 eV which reveals that the title compound has a stable structure. The energy difference between the HOMO and LUMO elucidates the possibility of eventual charge transfer interaction within the molecular species which influences the NLO activity of the compound. The molecular energy parameter values are given in Table 3. The frontier molecular orbitals of 2C5FP is shown in Figure 6.

### Table 4. Global Chemical Parameters of 2-Chloro-5-fluoro phenol at B3LYP/6-31++G (d,p) method.

| Molecular Properties (ev) | Electro Negativity | Chemical Potential | Hardness | Softness | Electrophilicity |
|--------------------------|--------------------|--------------------|----------|----------|-----------------|
|                          | \( \chi \)          | \( \mu \)           | \( \eta \) | \( S \)   | \( \omega \)    |
|                          | 3.8395             | -3.8395            | 2.832    | 0.3532   | 2.603           |

Figure 7. Load plot for the Energy parameters (HOMO-LUMO) and global chemical descriptors of 2-chloro-5-fluoro phenol.

3.5. Non-linear optical activity

The current research work on the second order non-linear optical (NLO) molecule based on organic materials has been embarked upon due to their potential applications in the field of optoelectronics (such as optical switching and dynamic usage processing) and information technology. Organic materials display a remarkable non-linear optical properties due to their high molecular hyperpolarizabilities. The first hyperpolarizability of donor acceptor aromatic molecules depends on the electronic charge communication between donor and acceptor groups that is useful in determining the intra-molecular charge transfer. Recently, vibrational spectroscopic determinations combined with quantum chemical calculations have been used as an efficient tool for the investigation of the structural characteristics responsible for the nonlinear optical properties of the molecular systems [39].

Theoretical evaluation of hyperpolarizability is most significant and aids in understanding the relationship between the molecular structure and nonlinear optical properties and is also useful as guideline to experimentalists for the design and synthesis of organic NLO materials. Anju Linda et. al studied the theoretical determination of NLO on naphthalene derivatives of p-nitroaniline by using DFT studies. The molecule N-[Naphthalene-5-lyl]-4- nitrobenzamide exhibits large hyperpolarizability and hence it can be recommended for photo voltaic device fabrication [40]. In the present investigation, the electronic dipole moment, molecular polarizability, first hyperpolarizability and second order hyperpolarizability are determined using the \( x, y, z \) components as follows. According to Kellinmann symmetry, first hyperpolarizability is a third rank tensor that can be described by a \( 3*3*3 \) matrix and the 27 components of the 3D matrix can be deduced to 10 components.

The complete equations for evaluating the magnitude of dipole moment \( \mu \), polarizability\( (\alpha) \), first hyperpolarizability \( (\beta) \) and second hyperpolarizability \( (\gamma) \) using the \( x, y, z \) component are as follows [41].

\[
\text{Dipole moment } \mu = \left( \mu_x^2 + \mu_y^2 + \mu_z^2 \right)^{\frac{1}{2}}
\]

\[
\text{Polarizability } \alpha = \frac{\alpha_x + \alpha_y + \alpha_z}{3}
\]

\[
\text{First hyperpolarizability } \beta = \left( \beta_x^2 + \beta_y^2 + \beta_z^2 \right)^{\frac{1}{2}}
\]

Table 5. Calculated Non Linear Parameters of 2-Chloro-5-fluoro phenol at B3LYP/6-31++G (d,p) method.

| Non Linear Parameters | Dipole Moment (\( \mu/D \)) | Polarizability (\( \alpha \)) | First Hyper Polarizability (\( \beta \)) | Second Order Hyper Polarizability (\( \gamma \)) |
|-----------------------|-----------------------------|-------------------------------|---------------------------------|---------------------------------|
|                       | 1.4083 Dbye                  | \( 10.1768 \times 10^{-30}\) esu | 2.0953 \( \times 10^{-30}\) esu | -2927.24 \( \times 10^{-30}\) esu |
Second order hyperpolarizability
\[ \beta \equiv \frac{\gamma_{xxxx} + \gamma_{yyyy} + \gamma_{zzzz} + 2(\gamma_{xxyy} + \gamma_{xxzz} + \gamma_{yyzz})}{5} \]

The Non–Linear optical (NLO) parameters of the title molecule have been computed by using B3LYP with 6-31+G (d,p) level and the corresponding values are given in Table 5. Since the values of the non linear optical parameters of Gaussian output are in atomic units (a.u) the calculated values are converted into electrostatic units (esu) \((\alpha: 1 \text{ a.u} = 0.1482\times10^{-24} \text{ esu}, \beta: 1 \text{ a.u} = 0.86399\times10^{-32} \text{ esu}, \gamma: 1 \text{ a.u} = 5.0372\times10^{-40} \text{ esu})\) [42]. The calculated first hyperpolarizability of the present molecule \((\beta = 2.0953\times10^{-30} \text{ esu})\) is about 7 times greater than urea \((\beta = 0.28166\times10^{-30} \text{ esu})\). The reported value of first hyperpolarizability of similar structure is \(\beta = 2.8065\times10^{-30} \text{ esu}\) [43].

Hence, from the results of the present molecule 2C5FP refined that it supports in the development of new effective material for NLO applications.

3.6. Reduced density gradient (RDG)

RDG analysis is used to evaluate the weak interaction based on electron density as a dimensionless quantity developed by Johnson et. al [44] and its first gradient is given by

\[ \text{RDG} = \frac{1}{2(3\pi^2)^{1/3}} \frac{1}{\rho(r)^{1/3}} \sqrt{\frac{\Delta^2 p(r)}{\rho(r)^{2/3}}} \]

By analyzing the low density gradient values and the high density gradient values can be identify the weak and strong interactions. The plot of \(p(r)\) Vs \(\lambda_2\) describes the nature and strength of interaction. The sign \(\lambda_2\) which is the highest value of Lessian matrix of electron density helps to identify the nature of interaction by color code. The RDG analysis was performed by using multiwfn software [20] followed by VMD programme. The predicted RDG plot for the title molecule (2C5FP) is shown in Figure 8. In the present molecule (2C5FP), the blue color represents the strong hydrogen bond interaction, red and green colors represent the strong repulsion and Van der Waals (VDW) interaction, respectively. These interactions can be responsible for the stability of the molecular system.
3.7. NMR chemical shifts

A combined NMR spectroscopic analysis and computer simulation technique offer a unique way to intercept and predict the chemical structure of large biomolecules. For instance, $^{13}$C chemical shift is one of the most pioneer techniques in determining the presence and absence of specific atom in a specific position of a molecular system [45]. Chemical shifts of 2C5FP are performed using the Gauge invariant atomic orbital (GIAO) method at 6–31++G (d,p) level in the gaseous phase. The GIAO method is one of the most significant and common approaches for evaluating isotropic nuclear magnetic shielding tensors. The obtained theoretical and experimental $^1$HNMR and $^{13}$CNMR spectra of 2C5FP are shown in Figures 9 and 10, respectively. The experimental $^1$HNMR and $^{13}$CNMR were recorded using acetone solvent and their corresponding chemical shifts ($\delta$) for protons and carbon atoms were observed. The isotropic chemical shifts $\delta$ can be evaluated using isotropic shielding data with respect to Tetramethylsilane (TMS) [42]. The relationship between the chemical shift ($\delta$) and isotropic shielding ($\sigma$) for carbon atom is given by

$$\delta_{\text{iso}}(C) = \sigma_{\text{TMS}}(C) - \sigma_{\text{iso}}(C)$$  \hspace{1cm} (11)

Where $\delta_{\text{iso}}$ is the isotropic chemical shift, $\sigma_{\text{iso}}$ is the isotropic shielding and C is the carbon atom [46]. The isotropic shielding value of carbon atoms and hydrogen atom in Tetramethylsilane (TMS) at B3LYP/6–31++G (d,p) level of computation is found to be 195.272ppm and 32.362 ppm, respectively. The experimental and calculated chemical shifts data with respect to TMS of 2C5FP in gas phase and acetone, chloroform solvents are shown in Table 6.

Generally, the CNMR isotropic chemical shifts of organic molecules are lying in the region 10–200 ppm. In the aromatic molecule, the carbon ($^{13}$C) and proton ($^1$H) chemical shifts are observed in the range of 115–150 ppm and 7.00–8.00 ppm. In the present investigation, theoretical $^{13}$CNMR spectrum makes out that the largest deshielded signal of the C5 and C1 atom is found to be 162.74 (gas), 163.24(acetone), 163.10 ppm (chloroform) and 154.91 (gas), 154.63 ppm (chloroform) 154.51 ppm (acetone), respectively. These can be attributed to the bond formed with electro negative atoms. Such electro negative atom polarizes the charge distribution in its bond to the adjacent carbon atom and reduces the electron density at the bridge for the title molecule. In the present compound, the carbon atoms C1 and C5 attached to the oxygen and fluorine atoms, respectively. Particularly, the $^{13}$C chemical shifts of atom C5 are

![Figure 9.](image)

(a) Experimental and (b) Calculated (B3LYP/6–31++G (d,p)) $^{13}$C NMR spectrum of 2 chloro-5-fluoro phenol.
slightly over estimated than C1, because the electro negativity of fluorine atom is greater than the oxygen. The $^{13}$C NMR chemical shifts were experimentally recorded in acetone at the interval of 112.53–163.29 ppm, which extensively coherence with the theoretically performed $^{13}$CNMR chemical shifts in acetone. The DFT showed a smaller change in carbon and proton chemical shifts with solvent than the experimentally observed data. The experimental proton ($^1$H) chemical shifts of 2C5FP was recorded in the range of 5.42–8.52 ppm. The correlated computational chemical shifts of proton in acetone are 8.29, 7.62, 7.28 and 4.60 ppm. Proton next to electron-donating groups are shielded likewise electron-withdrawing groups are deshielded [47]. Due to this, proton numbered H7 (8.29), H8 (7.62) and H9 (7.28) ppm have the maximum chemical shift values, as it is near the electro negative atoms such as 10Cl, 11F and O12. These results

| Atom | Exp Acetone | Acetone | Chloroform | Gas phase |
|------|-------------|---------|------------|-----------|
| C1   | 154.52      | 154.51  | 154.63     | 154.91    |
| C2   | 123.76      | 123.75  | 124.20     | 125.41    |
| C3   | 131.01      | 130.97  | 131.02     | 131.07    |
| C4   | 112.53      | 111.45  | 111.22     | 110.65    |
| C5   | 163.29      | 163.24  | 163.10     | 162.74    |
| C6   | 106.83      | 106.87  | 106.45     | 105.46    |
| H7   | 8.52        | 8.47    | 8.42       | 8.29      |
| H8   | 7.82        | 7.84    | 7.78       | 7.62      |
| H9   | 7.75        | 7.71    | 7.59       | 7.28      |
| H13  | 5.42        | 5.41    | 5.21       | 4.60      |

Figure 10. (a) Experimental and (b) Calculated (B3LYP/6-31++G (d,p)) $^1$H NMR spectrum of 2- chloro-5-fluorophenol.

Table 6. Experimental and Theoretical $^{13}$C and $^1$H isotropic chemical shifts (with respect to TMS) for 2-Chloro-5-fluorophenol at B3LYP/6-31++G (d,p) method.
concede that the experimental and theoretical chemical shifts are well consistent with each other.

The linear relation between the experimental \(^{13}\text{C}\) and \(^{1}\text{H}\) chemical shifts and the gauge–including atomic orbital (GIAO) magnetic isotropic shielding tensors is described by the following equation

\[
\delta_{\text{exp}} = a + b\sigma_{\text{cal}}
\]  

where, the intercept (a) and slope (b) of the least squares correlation line is used to scale the theoretical (GIAO)/B3LYP/6-31++G(d,p) isotropic shielding tensors \(\sigma\) and to predict the chemical shifts. The correlation graphics between the experimentally and calculated chemical shifts of the title molecule is depicted in Figure 11. The linear equations obtained between the experimental and calculated chemical shifts are described as

\[
\delta_{\text{cal}} = 10016\delta_{\text{Exp}} - 0.51598 \quad (R^2 = 0.9998)
\]  

### 3.8. Hydrogen bond and thermodynamic properties

Hydrogen bond is extremely powerful in deriving information such as weak molecular interactions, molecular conformations and dynamics in

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**Figure 11.** Correlation graphic between the experimental and theoretical chemical shifts of 2-chloro-5-fluoro phenol.

**Figure 12.** Dimer structure of 2-chloro-5-fluoro phenol.

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| Temperature (K) | Thermodynamic parameters | \(S_m\) (cal mol\(^{-1}\)K\(^{-1}\)) | \(H_m\) (k cal mol\(^{-1}\)) |
|-----------------|--------------------------|-------------------------------|------------------|
| 100             | 10.975                   | 63.888                        | 55.184           |
| 200             | 20.363                   | 75.77                         | 56.754           |
| 300             | 28.903                   | 86.465                        | 59.227           |
| 400             | 36.16                    | 96.399                        | 62.492           |
| 500             | 41.987                   | 105.563                       | 66.411           |
| 600             | 46.547                   | 114.001                       | 70.847           |
| 700             | 50.13                    | 121.762                       | 75.687           |
the solid state. Intra and inter-molecular interactions play a pivotal role in many disciplines such as host-guest chemistry, organic chemistry, pharmacology, biochemistry and molecular biology. For instance, the key factor is the breakage or dissociation of intramolecular and intermolecular hydrogen bond in the dissolution of cellulose [48, 49]. In the present investigation, the existence of intra and inter molecular hydrogen bonding is interpreted through the analysis of structural parameters. The dimer structure of the title molecule is shown in Figure 12. The intermolecular hydrogen bonded distances 12O ≈ 1.99 Å and 25O ≈ 1.99 Å are lesser than the sum of van der Walls radii (2.7 Å for O and H) which strongly validates the presence of intermolecular H-bond bridges [28].

3.8.1. Thermodynamic properties

On the basis of vibrational evaluation the standard statistical thermodynamic parameters such as heat capacity, entropy and enthalpy are computed using B3LYP/6-31+ +G (d,p) basis set. The total energy of a molecular system is the sum of translational, rotational, vibrational and electronic energies i.e. $E = E_t + E_r + E_v + E_e$. Thermodynamic quantities such as heat capacity at constant pressure ($C_p$), entropy ($S_m$) and enthalpy ($H_m$) for various ranges of temperature (100–700 K) are evaluated and the corresponding values are presented in Table 7. It can be noticed that the thermodynamic quantities increased with the increase of temperature from 100 to 700K. All thermodynamic computations have been carried out in gas phase. The correlation equations of heat capacity ($C_p$), entropy ($S_m$), and enthalpy ($H_m$) change to temperatures that are fitted by parabolic formula and the corresponding fitting factors ($R^2$) for these thermodynamic functions are found to be 0.9999, 0.9996 and 0.9999, respectively. The corresponding fitting equations of 2C5FP are as follows and the correlation graphics are depicted in Figure 13.

$$C_p = 0.0519 + 0.1147-6.6149 \times 10^{-5} T^2 \left( R^2 = 0.9999 \right)$$ (14)

$$S_m = 51.605 + 0.12797-3.9798 \times 10^{-5} T^2 \left( R^2 = 0.9996 \right)$$ (15)

$$H_m = 53.885 + 0.004-3.2706 \times 10^{-5} T^2 \left( R^2 = 0.9999 \right)$$ (16)

Hence, all these thermodynamic properties provide useful information for future study on 2C5FP [41, 50].

3.9. In – vitro antibacterial determination assay

3.9.1. Test bacterial samples

The selected bacterial strains were used for the title compound viz., Staphylococcus aureus, Staphylococcus aureus, (Gram – positive bacteria), Escherichia coli and Pseudomonas aeruginosa (Gram – negative bacteria), respectively. The bacterial strains were cultivated in Muller – Hinton agar plates and then inoculated plates were incubated at 37°C for 24h for antibacterial sensitivity assay.

3.9.2. Interpretation of antibacterial activity

The antibacterial activity of the title molecule 2C5FP is performed by the disc diffusion method. The most influencing factor exhibits significant antibacterial effect in phenol compounds which is due to the presence of rich plant extracts in phenolic compounds and the other factors are molecular weight, chemical reactivity, hydroxy group and electron negativity of atoms in the compound. The substitution of the electron-donating –OH moiety and electron-withdrawing groups such as fluorine (F) and oxygen (O) on the benzene ring of the title compound are the most significant that contributes to the antibacterial activity [51]. The presence of high electronegative atoms such as oxygen and fluorine atoms which are larger values of the negative charges on the molecule 2C5FP suggest the high electron density of the centers that would preferentially interact with micro-organisms then increases the antibacterial potential. On the other hand, the presence of the proton exchangeability of the phenolic –OH, attached to the electron delocalized system with high proton binding affinity contributes to the ability of penetration and damaging the cytoplasmic membranes, thus enhancing the antibacterial activity [52].

The HOMO-LUMO energy gap is the most significant tool for the chemical reactivity as well as the biological activity of the molecules. The presence of energy gap promotes the detachment of one electron from the molecule and can migrate inside the micro-organism producing free radicals that can damage and contribute to kill the pathogens. The distribution of the electron density or electrostatic potential surface area is an important factor to identify the electrophilic and nucleophilic site over the molecule and charged surface area in a compound that is considered as the best parameter to estimate the biological activity. According to the mechanism of the antibacterial activity of the compounds, the positive charge on the end of the molecules is responsible for the damage to the plasma membrane of pathogenic micro-organisms. In the present compound, the higher positive charged regions of oxygen and fluorine atoms are involved in the biologically active part in the molecule as well as used to inhibit the pathogens [53].

The observed data of antibacterial activity of the title molecule (2C5FP) and the standard drug are shown in Table 8 and the corresponding diagram depicted in Figures 14 and 15. From the results, it is

| S.NO | Cultures               | Diameter of the Zone of Inhibition in mm |
|------|------------------------|----------------------------------------|
|      |                        | 10μl | 20μl | 30μl | 40μl | Control | Antibiotic |
| 1    | Escherichia coli       | -    | -    | 8    | 12   | Nil     | 17        |
| 2    | Staphylococcus aureus  | -    | 8    | 10   | 12   | Nil     | 16        |
| 3    | Pseudomonas aeruginosa | -    | 8    | 12   | 13   | Nil     | 16        |
| 4    | Streptococcus aureus   | -    | 8    | 12   | 13   | Nil     | Nil       |

Figure 13. Variation of thermodynamic functions with temperature of 2-chloro-5-fluoro phenol.
observed that the title compound is recognized to be the potential antibacterial scaffold with the average zone of inhibition 8–13 mm against bacterial strains. The title compound is compared with Amoxicillin as standard antibiotic. The title molecule 2C5FP exhibits good antibacterial activity against Gram-positive bacteria such as *Streptococcus aureus*, *Staphylococcus aureus* and exhibits low activity against Gram-negative bacteria such as *Escherichia coli* and *Pseudomonas aureus*. Particularly, the molecule 2C5FP exhibits very effective activity in higher concentration (40 μL) for all selected pathogens. Among the selected bacterial strains, the title compound (2C5FP) exhibits better activity against *streptococcus aureus* when compared to standard antibiotic. The tested compound exhibits better activity against the Gram-Positive bacteria rather than Gram-negative bacterial strains.

3.9.3. Molecular docking analysis

The molecular docking technique is used recently in the biological study for drug discovery and to get insight into the exact binding location of the protein and ligands. It predicts the interaction between two molecules with overall minimum energy and the best orientation of protein-ligand [54]. The molecular docking analysis was performed on the most active compound 2C5FP against *staphylococcus aureus* tyrosyl-tRNA

![Figure 14. Disk diffusion method of 2-chloro-5-fluoro phenol at different concentrations; (a) *Escherichia coli* (b) *Pseudomonas aureus* (c) *Streptococcus aureus*; (d) *Staphylococcus aureus*.](image)

![Figure 15. A bar diagram plot for the antibacterial activity of 2-chloro-5-fluoro phenol.](image)

Table 9. Molecular docking results of 2-Chloro-5-fluoro phenol molecule with 1JIL and 6CJF protein targets.

| Drug | Protein (PDB ID) | Binding Energy (kcal/mol) | RMSD (Å) | No. of Hydrogen Bond | Bonded Residues | Bond Distance (Å) |
|------|----------------|--------------------------|----------|----------------------|----------------|------------------|
| 2C5FP | 1JIL          | -3.80                    | 112.20   | 2                    | Protein: A: ASN 29 | 1.7              |
|       |               |                          |          |                      | Protein: A: ASN 3 | 2.3              |
| 6CJF  |               | -4.33                    | 29.74    | 2                    | Protein: A: THR 63 | 2.8              |
|       |               |                          |          |                      | Protein: A: GLY 66 | 2.1              |
According to obtained docking results, it is conceded that the docked ligand 2C5FP form a stable complex with the receptors that the docked ligand 2C5FP form a stable complex with the receptors (Figure 16). In Figure 16, yellow dotted line represents the formation of intermolecular hydrogen bond with benzene ring. The docking computed results confirm that ASN 29 and ASN 3 amino acids form hydrogen bond interaction with a bond length of 1.7 Å and 2.3 Å, respectively, for 1JIL protein. Likewise, GLY 66 and THR 63 amino acids form a hydrogen bond interaction with a bond distance of 2.1 Å and 2.8 Å, respectively, for 6CJF protein. These hydrogen bonds play a positive role in strengthening the binding effect between the ligand and protein [60]. In consequence, the molecular docking analysis demonstrated an enhancement of the binding effect between the ligand and protein [60]. 

Figure 16. Docking and hydrogen bond interaction of ligand 2-chloro-5-fluoro phenol with two proteins (a) 1JIL (b) 6CJF.

4. Conclusion

The title molecule 2C5FP was characterized by various spectroscopic techniques (FT-IR, Raman and NMR). The optimized structural parameters such as bond lengths and bond angles of monomer and dimer of the title molecule compared with X – ray crystallographic data of related compound is found to have reliable agreement. The complete vibrational analysis has been performed and it is conceded that the relation between experimental and calculated wave numbers is found to be quite close. Non linear optical behavior of the title compound has been investigated by the first and second hyperpolarizability. Theoretically predicted 13C and 1 H NMR chemical shifts of title molecule is compared with experimental data in acetone solvent enhances a very good agreement. The correlations between the statistical thermodynamics and temperature are obtained and discussed. The charge transfer mechanism is attained with in a molecule by determining the energy gap between HOMO and LUMO and reactivity parameters. The title molecule 2C5FP exhibited better antibacterial activity against Gram-positive bacteria rather than Gram-negative bacterial strains. Finally, from the molecular docking results, the low binding energy of -3.8 kcal/mol confirm that the title compound 2C5FP is a good antibacterial compound.

Declarations

Author contribution statement

M. Arivazhagan: Conceived and designed the experiments; Contributed reagents, materials, analysis tools or data.
V. Vidhya: Analyzed and interpreted the data; Wrote the paper.
A. Austine: Conceived and designed the experiments; Performed the experiments.

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Declaration of interests statement

The authors declare no conflict of interest.

Additional information

No additional information is available for this paper.

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Hydrogen bonds are another significant factor that influences protein stability and contributes to the stability of proton-ligand binding interactions. An intermolecular hydrogen bond is formed with donor atom (D) and a hydrogen atom (H) in one compound and an acceptor atom (A) in the other compound. The donor group in a hydrogen bond is a strongly electronegative atom such as O, N and F that is covalently bonded to a hydrogen bond. The hydrogen acceptor is an electronegative atom of a neighboring compound that contains a lone pair that involves in the hydrogen bond [59]. The docking computation confirms that the O-H moiety is engaged in hydrogen bonding with the amino acids and shows π-anion interaction with benzene ring. The docking computed results confirm that ASN 29 and ASN 3 amino acids form hydrogen bond interaction with a bond length of 1.7 Å and 2.3 Å, respectively, for 1JIL protein. Likewise, GLY 66 and THR 63 amino acids form a hydrogen bond interaction with a bond distance of 2.1 Å and 2.8 Å, respectively, for 6CJF protein. These hydrogen bonds play a positive role in strengthening the binding effect between the ligand and protein [60]. In consequence, the molecular docking analysis demonstrated an enhancement of the ligand that has good pharmacological properties with the proteins.
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