Lower Incidence of Parvovirus-B19 Infections in Dutch Blood Donors during SARS-CoV-2 Pandemic

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During the SARS-CoV-2 pandemic several studies reported a lower incidence of respiratory infections other than COVID-19 compared to previous years, but bias due to changes in uptake and capacity of non-COVID care cannot be excluded (1, 2). The demand for blood did not decrease during the SARS-CoV-2 outbreak, and Dutch donors responded en masse to the request to continue donating blood. Therefore, the observed incidence of infectious diseases with routes of transmission similar to SARS-CoV-2 among blood donors may not suffer from these potential testing biases. In fact, the number of blood donations per month in 2020 and 2021 was comparable to the years before (719,211 donations in 2020 compared to min-max range of 682,315 [2019] to 721,183 [2013]). Parvovirus B19 (B19V), a member of genus Erythroparvovirus, causes the childhood disease erythema infectiosum, or fifth disease, but many B19V infections remain asymptomatic, especially in immunocompetent adults. Transmission of B19V occurs primarily via the respiratory route but also via transfusion of blood components and plasma products (3, 4). In the past, transmission of B19V by plasma-derived medicines occurred frequently because nonenveloped viruses are difficult to inactivate by heat or detergents during the plasma fractionation process. To comply with the European guidelines that plasma pools for the production of certain plasma-derived products may not exceed 10,000 IU/ml B19V DNA Sanquin Blood Supply Foundation routinely screens donated plasma for fractionation for B19V to exclude donations with a high B19V load (>1.2 × 10^6 IU/ml) before fractionation.

From 1 January 2013 to 30 June 2021, Sanquin screened 5,974,061 plasma donations for B19V DNA, as described previously (5). The standard test pool size was 480, but donations used for production of solvent-detergent-treated plasma were screened in pools of 96. When a pool of 480 or 96 donations contained an unacceptable level of B19V DNA (>5,000 IU/ml or >1,000 IU/ml for pool size of 480 or 96 donations, respectively), the pool was deconstructed to identify the donation(s) containing a high load of B19V DNA. During the study period, 200 donations were identified which contained >1.2 × 10^6 IU/ml of B19V DNA. As shown in our previous study (6), high-level viremia is a hallmark of acute B19V infection, while persistent low levels of B19V DNA (>1 × 10^5 IU/ml) are often associated with the detection of viral remnants. Figure 1 shows the number of highly viremic donations per 10,000 donations for B19V to exclude donations with a high B19V load (>1.2 × 10^6 B19V DNA IU/ml) before fractionation.

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donations were identified, and 0 were identified in the first 6 months of 2021, compared to 13 (2015) to 41 (2019) donations with loads of $>1.2 \times 10^6$ IU/ml in the years before (Fig. 1). The number of donations with high B19V DNA loads during the first 4 months of 2020 were comparable to the first months of low-prevalence B19V years 2015 and 2016. However, from May 2020 until 30 June 2021, zero B19V highly viremic blood donations were detected. This sharp decrease in highly viremic donations coincides with, and probably is causally related to, the COVID-19 epidemic. From 15 March to 11 May 2020 and from 15 December 2000 to 15 March 2021, all child daycare facilities, schools, cafes, restaurants, and sports clubs were closed in the Netherlands, and people had to work from home if possible. Additional measures including 1.5-m distancing, the obligation to wear face masks in public areas, more emphasis on hand hygiene, and limiting the number of social contacts were also practiced on a large scale between and after the lockdowns. All measures aimed to reduce the spread of SARS-CoV-2 but may have had similar effects on the spread of B19V in The Netherlands. For B19V, closing daycare facilities and schools probably prevented many transmission events between (young) children and subsequently the onward spread to their parents (potential blood donors).

On top of the seasonal cycle, symptomatic B19V infections are known to peak every 3 to 4 years (7). This interepidemic cycle is less obvious in viremic blood donors (see Fig. 1). As previously described, the absence of this 3- to 4-year cycle can be related to the fact that infected blood donors are asymptomatic and feel well enough to donate blood (8); the number of notified B19V cases is often limited to children with symptomatic fifth disease.

Our data showed a significant reduction of silent parvovirus B19 infections, from 4.0/100,000 donations in 2013 to 2019 to 0.56/100,000 and 0.00/100,00 in 2020 and 2021, respectively. The prevalence in 2020 was significantly lower than in previous years, including the year with the lowest prevalence, 2015 ($P = 0.04$ for 2015 versus 2020) (9). When comparing the 1.5-year period (January 2015 to June 2016) with the lowest B19 prevalence with the most recent 1.5-year period (January 2020 to June 2021), this difference is even more pronounced ($P = 0.0014$). Since our data set does not suffer from bias caused by changes in the uptake of health care or a decreased testing capacity for infections other than SARS-CoV-2, our findings confirm that public

**FIG 1** The number of parvovirus B19 highly viremic donations per month per 10,000 donations (donations with a B19V DNA load of $>1.2 \times 10^6$ IU/ml).
health measures taken to limit the spread of SARS-CoV-2 reduced the transmission of other respiratory infections, such as B19V.

REFERENCES

1. Olsen SJ, Azziz-Baumgartner E, Budd AP, Brammer L, Sullivan S, Pineda RF, Cohen C, Fry AM. 2020. Decreased influenza activity during the COVID-19 pandemic: United States, Australia, Chile, and South Africa, 2020. MMWR Morb Mortal Wkly Rep 69:1305–1309. https://doi.org/10.15585/mmwr.mm6937a6.

2. Nicolay N, Mirinaviciute G, Mollet T, Celentano LP, Bacci S. 2020. Epidemiology of measles during the COVID-19 pandemic, a description of the surveillance data, 29 EU/EEA countries and the United Kingdom, January to May 2020. Euro Surveill 25:2001390. https://doi.org/10.2807/1560-7917.ES.2020.25.31.2001390.

3. Hourfar MK, Mayr-Wohlfart U, Themann A, Sireis W, Seifried E, Schrezenmeier H, Schmidt M. 2011. Recipients potentially infected with parvovirus B19 by red blood cell products. Transfusion 51:129–136. https://doi.org/10.1111/j.1537-2995.2010.02780.x.

4. Juhl D, Ozdemir M, Dreier J, Gorg S, Hennig H. 2015. Look-back study on recipients of Parvovirus B19 (B19V) DNA-positive blood components. Vox Sang 109:305–311. https://doi.org/10.1111/vox.12295.

5. Molenaar-de Backer MW, de Waal M, Sjerps MC, Koppelman MH. 2016. Validation of new real-time polymerase chain reaction assays for detection of hepatitis A virus RNA and parvovirus B19 DNA. Transfusion 56:440–448. https://doi.org/10.1111/trf.13334.

6. Molenaar-de Backer MWA, Russcher A, Kroes ACM, Koppelman MHGM, Lanfermeijer M, Zaaijer HL. 2016. Detection of parvovirus B19 DNA in blood: viruses or DNA remnants? J Clin Virol 84:19–23.

7. Zaaijer HL, Koppelman MH, Farrington CP. 2004. Parvovirus B19 viraemia in Dutch blood donors. Epidemiol Infect 132:1161–1166. https://doi.org/10.1017/s0950268804002730.

8. Kooistra K, Mesman HJ, de Waal M, Koppelman MH, Zaaijer HL. 2011. Epidemiology of high-level parvovirus B19 viraemia among Dutch blood donors, 2003–2009. Vox Sang 100:261–266. https://doi.org/10.1111/j.1423-0410.2010.01423.x.

9. Janssen MP, Cator EA, van der Poel CL, Schasberg WP, Bonsel GJ, van Hout BA. 2009. Monitoring viral incidence rates: tools for the implementation of European Union regulations. Vox Sang 96:298–308. https://doi.org/10.1111/j.1423-0410.2009.01166.x.