Health Benefits and Pharmacological Properties of Carvone

Abdelhakim Bouyahya 1,*, Hamza Mechchate 2, Taoufiq Benali 3, Rokia Ghchime 4, Saoulajan Charfi 5, Abdelali Balahbib 6, Pavel Burkov 7, Mohammad Ali Shariati 8, Jose M. Lorenzo 9,10,* and Nasrreddine El Omari 11

Abstract: Carvone is a monoterpene ketone contained in the essential oils of several aromatic and medicinal plants of the Lamiaceae and Asteraceae families. From aromatic plants, this monoterpene is secreted at different concentrations depending on the species, the parts used, and the extraction methods. Currently, pharmacological investigations showed that carvone exhibits multiple pharmacological properties such as antibacterial, antifungal, antiparasitic, antineuraminidase, antioxidant, anti-inflammatory, and anticancer activities. These studies were carried out in vitro and in vivo and involved a great deal of knowledge on the mechanisms of action. Indeed, the antimicrobial effects are related to the action of carvone on the cell membrane and to ultrastructural changes, while the anti-inflammatory, antidiabetic, and anticancer effects involve the action on cellular and molecular targets such as inducing of apoptosis, autophagy, and senescence. With its multiple mechanisms, carvone can be considered as natural compounds to develop therapeutic drugs. However, other investigations regarding its precise mechanisms of action as well as its acute and chronic toxicities are needed to validate its applications. Therefore, this review discusses the principal studies investigating the pharmacological properties of carvone, and the mechanism of action underlying some of these properties. Moreover, further investigations of major pharmacodynamic and pharmacokinetic studies were also suggested.

Keywords: carvone; pharmacological properties; mechanism of action

1. Introduction

Carvone is a monoterpene ketone (2-methyl-5-(1-methylethenyl)-2-cyclohexen-1-one) (C\textsubscript{10}H\textsubscript{14}O) (Figure 1) with a boiling point of 230 °C, which has an asymmetric carbon.
Chemically, carvone exists in two forms (enantiomers (+)-carvone and (-)-carvone) with the same chemical and physical properties and which differ only in their rotatory power. This monoterpene is present in the essential oils of some plant species, including *Mentha* spp., *Origanum* spp., *Rosmarinus* spp., *Thymus* spp., and many others [1–8]. The concentration of this volatile compound differs according to the species and is related to parameters such as the species, the geographical location, the phenological stages, and the parts of the plant. It is synthetized and secreted as a secondary metabolite from essential oils, and its major role in the plant remains unclear. Currently, many studies have proven that carvone has promising pharmacological properties. Indeed, it has shown neuroprotective effects, and can therefore be developed as a drug against certain disorders such as depression, sedation, nociception, and seizure [9–12]. This molecule has also demonstrated an antidiabetic effect, through its role in the prevention of obesity and metabolic problems associated with high-fat diets, achieved by improving glycoprotein component abnormalities and controlling glucose metabolism [13–15]. The use of carvone as an antifungal has also been investigated against various fungi strains (*Candida* spp.), mycotoxigenic fungi (*Fusarium* spp., *Aspergillus* spp., and *Penicillium* spp.), and dermatophytes (*Trichophyton* spp., *Epidermophyton floccosum*, and *Microsporum* spp.) [16–21]. The action of carvone on germ tube formation and fungi biofilm was reported. Additionally, carvone can be used as an antibacterial agent against many strains of bacteria, including methicillin-resistant *Staphylococcus aureus* (MRSA) [20,22–28]. Its antibacterial effects are often related to its capacity to penetrate into bacterial cells thus inducing an increase in cell permeability and a decrease in cell membrane integrity. It also exhibited an antibiofilm effect against *S. aureus*. On the other hand, carvone had anticancer activity against different cancer cell lines, including myeloma and melanoma cells, and breast cancer cells [29–32]. The antiproliferative mechanism of action has also been elucidated. Moreover, some studies proved the anti-inflammatory property of carvone and investigated the underlying molecular mechanism [33–36]. Anticancer mechanisms of carvone are due to its different actions against checkpoints of cancer cells such as inducing apoptosis and cell cycle arrest. Other pharmacological activities have also been reported, including antioxidant activity [37], neuraminidase inhibitory activity against the influenza virus [38], and antiparasitic activity against *Culex quinquefasciatus*, *Cx. pipiens*, *Aedes aegypti*, and *Haemonchus contortus* [39–42]. All these properties have enabled the use of carvone in other fields, such as the disinfection of food packaging and medical devices. In addition, to improve its action and extend its industrial use, carvone has been included in poly(lactic acid) films [16], antibacterial coating (ppCar) [23], and poly (lactic-co-glycolic acid) (PLGA) nanoparticles [25]. Therefore, the main objective of this paper is to report the pharmacological properties of carvone, and to highlight the mechanism(s) of action responsible for these activities.

![Chemical structure of carvone](image)

**Figure 1.** Chemical structure of carvone.

2. Research Methodology

Literature data for all carvone studies was collected using different scientific search engines, including Scopus, Wiley Online, Web of Science, Scifinder, Google Scholar, PubMed, ScienceDirect, and SpringerLink. They were organized in tables and then analyzed, highlighted, and discussed. The chemical structure of carvone was drawn using ChemDraw Pro 8.0 software.
3. Results and Discussion

3.1. Natural Sources of Carvone

Carvone is the major compound of the essential oil (EO) of many species of the Lamiaceae family; *Mentha* [43–46], *Mentha spicata* [22,43,44,47–60], *Mentha × villoso-nervata* [43], *Mentha piperita* L. [48,53,61], *Mentha crispa* L. [61,62], and *Mentha cardiaca* L. [60]. Moreover, carvone is the major compound of the EO of *Anethum graveolens* [50,53,63–68], *Thymus vulgaris* [45], *Majorana hortensis* [45], *Carum carvi* [50,52,63,66,67,69–73], *Anethum sowa* [22], and *Solanum tuberosum* L. [74,75]. In addition, other species belonging to the Orchidaceae family have been characterized by their richness in carvone; *Catasetum discolor*, *Catasetum longifolium*, *Catasetum integerrimu*, *Catasetum macroglossu*, *Catasetum tabular*, *Catasetum veracruz*, *Catasetum viridiflavu* [76], and *Lippia alba* [18,28,77–81]. Secondary metabolites in EOs are variable depending on certain factors, including geographic origin, stages of development, and parts of the plant. Indeed, some studies have proven this variability between geographic locations and suggest the fluctuation of these phytochemicals in plants for responding to environmental situations [18,77,78].

3.2. Pharmacological Properties of Carvone

With its varied actions on subcellular, cellular, and molecular actions, carvone exhibits several biological properties such as antimicrobial, anticancer, anti-inflammatory, antidiabetic, neurological, and many other pharmacological effects (Figure 2).

![Figure 2. Pharmacological properties of carvones. Carvone exhibits multiple biological activities including antidiabetic, anti-inflammatory, anticancer, neurological, antimicrobial, antiparasitic, antiarthritic, anticonvulsant, and immunomodulatory effects.](image)

3.2.1. Neurological Activity

Several authors have attempted to study the effect of the use of carvone on some neurological disorders such as depression, sedation, nociception, seizure, local anesthesia, as well as its effect on some receptors and on the action potential [9–12]. To study these effects, different models are used such as Swiss mice [9], Wistar rats [11], frog’s sciatic
nerve [10], and cortical neurons prepared from the cerebral cortices of rat fetuses [12] (Table 1).

Table 1. Neurological activities of carvone.

| Molecules                  | Origins       | Models Used                  | Experimental Approaches                                      | Key Results                                                                 | References |
|----------------------------|---------------|------------------------------|----------------------------------------------------------------|-----------------------------------------------------------------------------|------------|
| (S)-(+-)Carvone and        | Purchased     | Male Swiss mice             | Pentobarbital-induced sleeping time                           | LD_{50} = 484.2 mg/kg for (S)-(+-)-carvone                                  | [9]        |
| (R)-(−)-carvone            |               |                              | Locomotor activity assessed in an activity cage                | LD_{50} = 426.6 mg/kg for (R)-(−)-carvone                                   |            |
|                            |               |                              | PTZ-induced convulsions                                        | Both enantiomers induced depressive effects                                 |            |
|                            |               |                              | Pentobarbital-induced hypnosis                                 | Both enantiomers significantly reduced ambulation                          |            |
|                            |               |                              | PTZ-induced seizure                                            | At 100 mg/kg, (R)-(−)-carvone was more effective than (S)-(+-)-carvone       |            |
|                            |               |                              | PIC-induced seizure                                            | LD_{50} = 484.2 mg/kg for (S)-(+-)-carvone                                  |            |
|                            |               |                              |                                                                 | Both enantiomers induced depressive effects                                 |            |
|                            |               |                              |                                                                 | At 200 mg/kg, (S)-(+-)-carvone improved the latency of convulsions produced by PTZ and PIC |            |
|                            |               |                              |                                                                 | (S)-(+-)-carvone has depressant effects in the CNS                         |            |
|                            |               |                              |                                                                 | (R)-(−)-carvone has anticonvulsant-like activity                            |            |
| (+)-carvone, (−)-carvone   | Not reported  | The sciatic nerve of the frog (Rana ridibunda) from both sex | Three-chambered recording bath for the assessment of local anesthetic activity | Both carvone enantiomers elicited comparable responses                     | [10]       |
|                            |               |                              |                                                                 | The action potential of the evoked compound was abolished in 6 to 7 min and had an immediate recovery of 83% to 87% |            |
|                            |               |                              |                                                                 | Both carvones acted in the same way as lidocaine (10 mM)                    |            |
|                            |               |                              |                                                                 | No recovery of the action potential of the elicited compound, when nerves have been exposed to carvones for more than 6–7 min |            |
|                            |               |                              |                                                                 | The unusual neurotoxic effect of C+ and C− may be a disadvantage for their use in clinical practice |            |
| (+)-carvone, (−)-carvone   | Purchased     | Adult male Wistar rats       | Sucrose-gap apparatus (ex vivo assay) for CAP-inhibitory effect | C- was less potent (IC\textsubscript{50} = 10.7 ± 0.07 mM) in reducing nerve excitability than C+ (IC\textsubscript{50} = 8.7 ± 0.1 mM) | [11]       |


Table 1. Cont.

| Molecules                  | Origins     | Models Used                                      | Experimental Approaches | Key Results                                                                 | References |
|----------------------------|-------------|-------------------------------------------------|--------------------------|------------------------------------------------------------------------------|------------|
| (R)-(−)-carvone and (S)-(+)carvone | Purchased   | Cultures of cortical neurons prepared from the cerebral cortices of fetal rats | [3H] Flunitrazepam Binding Cell viability assay | Both isomers blocked GABA-induced activation of [3H] Flunitrazepam binding The doses required to produce negative receptor modulation were not lethal The insecticidal effect of carvones can be explained by their interaction with the GABA receptor at its non-competitive blocker region | [12]       |

To assess the effect of carvone on disorders related to the central nervous system (depression, convulsion), De Sousa et al. [9] conducted their study using different enantiomers of carvone ((S)-(+)carvone and (R)-(−)-carvone). The LD₅₀ values of the enantiomers varied between 400–500 mg/kg, and both enantiomers demonstrated depressive effects, expressed as decreased ambulation and responsiveness to touch, as well as increased sedation, palpebral ptosis, and antinociceptive effects. In addition, (S)-(+) and (R)-(−)-carvone reduce ambulation significantly. At 0.5 and 2.0 h after administration, (R)-(−)-carvone appeared to be more effective than its equivalent enantiomer [9]. However, at 1 h, (S)-(+)carvone was slightly more powerful. To increase pentobarbital sleep duration, (R)-(−)-carvone (100 mg/kg) was more effective than (S)-(+)carvone but was less effective at 200 mg/kg compared with its enantiomer, suggesting induction of a sedative effect. At a dose of 200 mg/kg, (S)-(+)carvone substantially improved the latency of convulsions produced by PTZ and PIC, while (R)-(−)-carvone was ineffective against these convulsions. These findings indicate that (S)-(+) and (R)-(−)-carvone have a CNS depressant effect, with an anticonvulsant property in (S)-(+)carvone [9].

Faliagkas et al. [10] tested two enantiomers of carvone ((+) and (−)-carvone) for their local anesthetic activity at two concentrations, 10 and 20 mM. The authors used a nerve preparation based on the frog’s sciatic nerve. They found that both enantiomers induce similar responses. When rinsed out of the nerve preparation and replaced with normal saline, they completely abolished the action potential of the evoked compound within 6–7 min, with an instantaneous recovery of 83–87%. Both carvones acted in the same way as 10 mM lidocaine (a standard local anesthetic), although they were 3–4 times less active in terms of reaction time. There was no recovery of the elicited compound action potential when the nerves were exposed to carvones for more than 6–7 min, suggesting a neurotoxic effect. In conclusion, the unique neurotoxic action of (+)- and (−)-carvone may be a disadvantage in therapeutic practice.

Based on multiple previous reports regarding the insecticidal activity of carvone and its effects on the nervous system, Sánchez-Borzone et al. [12] studied the effect of both carvone enantiomers on the GABAA receptor as a major insecticidal target by determining their effects on the recognition sites of benzodiazepines (BZD), belonging to the group of sedatives and anxiolytics, using primary neuronal cultures. Both isomers were able to block GABA-induced stimulation of [3H] flunitrazepam binding, suggesting that they act as negative allosteric modulators on the GABAA receptor. Their action was equivalent to that of thujone in this study, with the (R)-(−)-carvone stereoisomer being the most potent. The unusual configuration of the isopropenyl group at position five appears to be important for receptor engagement, whereas the structure of carvone does not appear to be important for receptor recognition. In a mouse neuron culture system, the doses required to produce negative receptor modulation were not lethal. These findings support the theory that carvones’ insecticidal effect is explained, at least in part, by their interaction with the non-competitive blocker site of the GABAA receptor.
Gonçalves et al. [11] previously demonstrated that the in vivo antinociceptive activity of (−)-carvone is impaired by a decrease in nerve excitability. In their study, they attempted to investigate and reveal the neuropharmacological effect of carvone to explain the observed effect (compound action potential (CAP) inhibitory effect) [11]. Using a modified single sucrose-gap technique (ex vivo), the effects of (+)- and (−)-carvone on CAP properties were evaluated. The study findings showed that (−)-carvone was less potent (IC$_{50}$ = 10.70.07 mM) in reducing nerve excitability than its enantiomer, (+)-carvone (IC$_{50}$ = 8.70.1 mM), despite having a similar mode of action, as their effects were partially counteracted by nerve washing and also by a reduction in depolarization velocity, most likely due to voltage-gated sodium channel blockades. These findings suggest that monoterpenes suppression of CAP conduction in peripheral nerves may further enhance knowledge about the pharmacology of natural bioactive substances. Furthermore, changing the chemical structures of such molecules may be used to activate or inhibit neuronal excitability [11].

3.2.2. Antidiabetic Activity

Several studies showed the antidiabetic effects of volatile compounds including carvone [82]. Indeed, three separate studies were interested in the potential antidiabetic activity of carvone by revealing its overall activity on in vivo models such as C57BL/6 mice with high-fat diet-induced obesity and streptozotocin (STZ)-induced diabetes in Wistar rats. Those studies have also focused on the main underlying mechanism of action exhibited by this molecule following multiple biochemical, hematological, and histopathological analyses [13–15] (Table 2).

| Molecules | Origins | Models Used | Experimental Approaches | Key Results | References |
|-----------|---------|-------------|-------------------------|-------------|------------|
| S-carvone | Purchased | C57BL/6 mice (male, ten weeks old) | GTT and ITT, Histological examination, Determination of hepatic triglyceride and serum lipid levels, Determination of insulin resistance, Gene expression analysis | Prevented weight gain, fat buildup in the liver, and insulin resistance, Increased expression of macrophage marker genes in white adipose tissue, including F4/80, Cd11b, Cd11c, Cd206, and Tnf-$\alpha$, Decreased expression of genes involved for lipid production and transport in the liver (Ppar2, Scd1, Cd36) | [13] |
| Carvone   | Purchased | Male Wistar rats weighing approximately 180–200 g | STZ-induced diabetes, Estimation of blood glucose and plasma insulin levels, Extraction and determination of glycoproteins | Improved glycemic status in a dose-dependent manner, in diabetic rats (30 mg/kg b.w.), Increased plasma insulin levels, Reduced plasma glucose levels, Restored the altered plasma and tissue glycoprotein levels, Restored the abnormal levels of plasma and tissue glycoprotein components | [14] |
To verify whether S-carvone can prevent obesity and metabolic problems caused by a high-fat diet, Alsanea and Liu [13] conducted a study on ten-week-old C57BL/6 male mice fed a high-fat diet and injected, intraperitoneally twice a week, with benzyl isothiocyanate (BITC), S-carvone, or vehicle for 8 weeks. Body weight, food consumption, and body composition were all monitored, and glucose tolerance and insulin tolerance tests were performed at the end of the study. Moreover, to determine the effects of BITC and (S)-carvone therapies on lipid and glucose metabolism and inflammatory responses, serum biochemistry, histology, and gene expression analyses were carried out. Therefore, (S)-carvone and BITC inhibited the weight gain induced by a high fat diet, as well as the insulin resistance and the accumulation of fat in the liver. The positive effects were related to increased expression of macrophage marker genes in white adipose tissue, including \( F4/80 \), \( Cd11b \), \( Cd11c \), \( Cd206 \), and \( Tnf-\alpha \), and decreased expression of genes involved in production and transport of lipids in the liver (\( Ppar2 \), \( Scd1 \), and \( Cd36 \)). In conclusion, this study suggests that BITC and (S)-carvone block high-fat diet-induced obesity and metabolic disorders and may be considered for the management of the obesity epidemic [13].

Muruganathan et al. [14] conducted their research to examine the impact of carvone on glycoprotein disruption in the STZ-induced diabetes model. A single intraperitoneal dose of STZ (40 mg/kg b.w.) induced diabetes in male Wistar rats. Glycoprotein levels were altered in experimental diabetes mellitus. Carvone was administered intragastrically to diabetic rats at doses of 25 mg/kg, 50 mg/kg, and 100 mg/kg for 30 days. Carvone’s effects on plasma glucose, insulin, plasma, and tissue glycoproteins were evaluated. In experimentally diabetic rats, oral treatment with carvone (50 mg/kg b.w.) for 30 days improved glycemic status in a dose-dependent manner, with a substantial increase in plasma insulin levels, and decrease in plasma glucose levels. The abnormal levels of plasma and tissue glycoprotein components were nearly normalized. Current results indicate that carvone, in addition to its antihyperglycemic action, may be able to improve glycoprotein component abnormalities in experimental diabetes. In view of these encouraging results, it is recommended to expand the scope of carvone usage in further studies to mitigate the negative consequences of diabetes [14].

Muruganathan et al. [14] investigated the impact of carvone on carbohydrate metabolic enzymes in the livers of normal and STZ-induced diabetic rats. A single intraperitoneal dose of STZ (40 mg/kg b.w.) was used to induce diabetes. STZ injection caused a substantial increase in plasma glucose and glycosylated hemoglobin (HbA1c), as well as a reduction in insulin and hemoglobin (Hb) levels. Carbohydrate metabolic enzymes, glycogen, enzymatic antioxidants in the pancreas, and hepatic marker levels have all been affected. Diabetic rats treated daily with a single oral dose of carvone (50 mg/kg b.w.) for 30 days, showed a substantial decrease in plasma glucose and HbA1c levels, as well as a significant improvement in Hb and insulin levels [14]. Administration of carvone restored the reversed activity of carbohydrate metabolic enzymes, enzymic antioxidants, and hep-

| Molecules | Origins | Models Used | Experimental Approaches | Key Results | References |
|-----------|---------|-------------|-------------------------|-------------|------------|
| Carvone   | Purchased Male Wistar rats (160–190 g) | STZ-induced diabetic rats | Biochemical analysis Histopathological study of liver and pancreas Immunohistochimical examination of the pancreas | Decreased plasma glucose and HbA1c levels (50 mg/kg b.w.) Improved Hb and insulin levels Restored the reversed activity of carbohydrate metabolic enzymes, enzymic antioxidants, and hepatic marker enzymes Decreased STZ-induced damage to hepatic and pancreatic cells Controlled glucose metabolism by enhancing important enzymes in the hepatic tissues of diabetic rats | [15] |
atic marker enzymes in diabetic rats to near-normal levels. The results were compared with gliclazide, a common oral hypoglycemic drug. Histopathological examination of the liver and pancreas, as well as immunohistochemistry of the pancreas, showed that carvone therapy decreases STZ-induced damage to liver and pancreatic cells. According to these findings, carvone controls glucose metabolism by improving enzymes important in the hepatic tissues of diabetic rats. Nevertheless, more research and safety studies are required to further verify carvone’s benefits [14].

3.2.3. Antifungal Activity

Carvone has emerged as a promising antifungal compound. Its application extends from the screening of basic properties against different fungi strains and mycotoxins, to an application designed in the food industry, in particular food packaging [16–21] (Table 3).

| Molecules         | Origins  | Strains Used                                                                 | Experimental Approaches                                                                 | Key Results                                                                 | References |
|-------------------|----------|-----------------------------------------------------------------------------|---------------------------------------------------------------------------------------|-----------------------------------------------------------------------------|------------|
| R-(-)-carvone     | Purchased| Poly (lactic acid) (PLA) films for food packaging applications                | Inclusion of R-(−)-carvone in the polymer matrix                                      | Lower Tg and Tm                                                             | [16]       |
|                   |          |                                                                             | Preparation and determination of film thickness                                       | Higher gas permeability                                                     |            |
|                   |          |                                                                             | Determination of remaining content                                                     | Lower tensile strength                                                     |            |
|                   |          |                                                                             | Determination of thermal, mechanical and barrier properties                            | Higher elongation at break                                                   |            |
|                   |          | Candida rugosa, Candida lusitaniae, Candida glabrata, Candida utilis, Candida | Planktonic anti-candida assay                                                           | Homogeneous and transparent antifungal films                                |            |
|                   |          | krusei, Candida guilliermondii, Candida tropicalis, Candida albicans, Candida |                                                                       |                                                                            |            |
|                   |          | parapsilosis, and Candida dubliniensis                                       |                                                                       |                                                                            |            |
| Carvone           | Purchased| Candida parapsilosis, Candida krusei, Aspergillus flavus, and Aspergillus    | Mic = 0.5 mg/mL. The concentration of 0.5 mg/mL inhibited at least 50% of the biofilm | Inhibited the polymorphism                                                  | [17]       |
|                   |          | fumigatus Broth macro-dilution method AFST-EUCAST method CLSI M38-A method   |                                                                       | up to 86% Changes in yeast cell envelope and cell viability were greater than 50% |            |
|                   |          | Determination of GM-MIC                                                     |                                                                       | Induced important antifungal activities                                   |            |
| Carvone chemotype | Naturel  | Candida parapsilosis, Candida krusei, Aspergillus flavus, and Aspergillus    | GM-MIC > 500 µg/mL against the different strains studied                            | No activity against selected clinical strains                               | [18]       |
| Molecules | Origins | Strains Used | Experimental Approaches | Key Results | References |
|-----------|---------|--------------|-------------------------|-------------|------------|
| Carvone   | Purchased | *Fusarium subglutinans*, *Fusarium cerealis*, *Fusarium verticillioides*, *Fusarium proliferatum*, *Fusarium oxysporum*, *Fusarium sporotrichioides*, *Aspergillus tubingensis*, *Aspergillus carbonarius*, *Alternaria alternata*, and *Penicillium* sp. | In vitro antifungal activity Evaluation of deoxynivalenol production Evaluation of inhibitory effects on plant seed germination | Induced toxic effects on the growth of the mycelium of all fungal species | [19] |
| Carvone   | Naturel (Mentha spicata) | *Cryptococcus neoformans*, dermatophytes (*Trichophyton* spp., *Epidermophyton floccosum*, and *Microsporum* spp.), and *Aspergillus* strains | In vitro antifungal activity Evaluation of the inhibitory activity of germ tube formation | *Mentha spicata* EO was effective against *Cryptococcus neoformans*, as well as the dermatophytes *Trichophyton rubrum* and *Trichophyton verrucosum* (0.32 µL/mL) Inhibited the germ tube development of *Candida albicans*, at concentrations below the MIC (0.16 µL/mL) | [21] |
| (+)-carvone (C+) (−)-carvone (C−) α,β-epoxycarvone (EP) (+)-hydroxydihydrocarvone (HC+) (−)-hydroxydihydrocarvone (HC−) | Purchased | *Candida parapsilosis*, *Candida tropicalis*, *Candida krusei*, and *Candida albicans* | Determination of MIC by microplate dilution method and MFC | Low antifungal activity against *Candida tropicalis* and *Candida parapsilosis* EP and C+ showed moderate activity against *Candida krusei* similar to C+ and C− against *Candida albicans* All the molecules tested showed fungistatic and fungicidal activity against *Candida* yeasts, and the most significant result was recorded with C+, C−, and EP | [20] |

In their ultimate goal to develop antifungal poly(lactic acid) (PLA) films for food packaging applications, Boonruang and collaborators [16] used (R)-(−)-carvone in their study. The molecule was incorporated into PLA-based polymer at 10%, 15%, and 20% by weight. The film conversion process consists of three steps, namely, melt blending, sheet extrusion, and biaxial stretching. The incorporation of antifungal compounds into the polymer matrix resulted in decreased Tg and Tm, increased gas permeability, reduced tensile strength, and increased elongation at the break of the antifungal PLA films. The antifungal films were homogeneous and transparent.

Giovanä et al. [17] were interested in finding an effective antifungal drug in the fight against candidiasis, an infection caused by *Candida* spp. which has developed significant resistance to current therapies. Since it has already been documented that *Mentha* spp. has antifungal properties, the authors of this research chose four main components present in its EO, including carvone. They evaluated growth suppression by microdilution, biofilm breakdown by electron microscopy, and germ tube formation inhibition by optical microscopy. The compounds tested had an antifungal activity with a MIC of 0.5 mg/mL, at
least 50% biofilm inhibition at the 0.5 mg/mL concentration, polymorphism inhibition at 86%, and changes in the cell envelope of yeast (SEM) and cell viability greater than 50% among the Candida strains tested. Due to the potential antifungal capacity of carvone, as well as its low cytotoxicity, it was considered a viable candidate to supplement antifungal regimens.

Mesa-Arango et al. [18] selected two carvone chemotypes from Colombian L. alba (Mill.) EOs to investigate their antifungal activity against multiple strains such as Candida parapsilosis, Candida krusei, Aspergillus flavus, and Aspergillus fumigatus strains using standardized protocols. According to the research results, the GM-MIC values were greater than 500 g/mL against the various strains tested, suggesting a weak antifungal activity.

In their study, Morcia et al. [19] selected certain natural EO compounds (including carvone) on 10 species of mycotoxigenic fungi involved in several plant diseases, namely, Fusarium subglutinans, Fusarium cerealis, Fusarium verticillioides, Fusarium proliferatum, Fusarium oxysporum, Fusarium sporotrichioides, Aspergillus tubingensis, and Aspergillus carbonarius. Carvone and the other chemicals examined had a toxic effect on mycelium development in vitro on all fungal species, albeit at varying levels of activity, prompting additional research on these compounds in the field of mycotoxins.

In a recent study, Piras et al. [21] examined the antifungal efficacy of Mentha spicata L. EO, containing 62.9% carvone. Their primary objective was to test its effectiveness on the virulence factors of Candida albicans, especially the suppression of germ tube development, as well as their impact on other strains. Consequently, M. spicata EO showed a superior effect against Cryptococcus neoformans and the dermatophytes Trichophyton rubrum and Trichophyton verrucosum (0.32 µL/mL) and also inhibited germ tube formation in Candida albicans up to 80% at concentrations eight times lower than the MIC. The results of the study support and validate the use of this plant EO in traditional medicine.

Given the reported activity of carvone in the literature, Moro et al. [20] conducted a research to assess the antifungal activities of (+)- and (−)-carvone, (+)- and (−)-hydroxydihydrocarvone, and α,β-epoxycarvone. (+)-Hydroxydihydrocarvone (HC+), (−)-Hydroxydihydrocarvone (HC−), and, α,β-epoxycarvone (EP) were synthesized from (+)-carvone (C+) or (−)-carvone (C−). The antifungal activity (MIC and MFC) was tested against Candida parapsilosis, Candida tropicalis, Candida krusei, and Candida albicans. All compounds showed modest antifungal efficacy against Candida tropicalis and Candida parapsilosis. Moreover, EP and C+ had modest antifungal activity against C. krusei. The results indicate that carvones and their derivatives may be used as antifungal drugs against Candida yeasts.

3.2.4. Antibacterial Activity

The antibacterial activity of carvone has been studied by several authors against multiple strains such as Escherichia coli, S. aureus, Streptococcus faecalis, and Pseudomonas aeruginosa. Different enantiomers were used in comparative studies to assess the structure–function relationship. Some studies were interested in the antibacterial effect of the encapsulated form (carvone loaded PLGA nanoparticles), others in its microbial transformation, while others were interested in manufacture of carvone biofilms (antibacterial coating) to prevent bacterial colonization of medical devices [20,22–28] (Table 4).
**Table 4. Antibacterial activity of carvone.**

| Molecules                  | Origins                                      | Model Used                  | Experimental Approaches                          | Key Results                                                                 | References |
|---------------------------|----------------------------------------------|-----------------------------|--------------------------------------------------|------------------------------------------------------------------------------|------------|
| (S)-(−)-carvone           | Naturel (Mentha spicata and Anethum sowa)    | Bacillus subtilis, Enterobacter aerogenes, Enterococcus Faecalis, Klebsiella pneumoniae, Pseudomonas aeruginosa, Staphylococcus aureus, Streptococcus mutans, Yersinia enterocolitica, Salmonella typhi, Escherichia coli, Staphylococcus epidermis, and Mycobacterium smegmatis | Disk diffusion assay, Broth dilution assay | The activity of carvone was comparable with the bioactivity of their original oils Active against a broad spectrum of human pathogenic bacteria (R)-(+)−limonene showed comparable bioactivity profile over the (S)-(−)-isomer | [22]       |
| (R)-(+)−carvone           | Purchased                                   | Staphylococcus aureus       | Single-step plasma polymerization, Plasma polymerization of carvone, Surface characterization, Antibacterial activity, Live-dead fluorescence assay, Crystal violet assay, Morphology of bacteria by field emission scanning electron microscope (FE-SEM) | Polymerization provided a hydrophobic antibacterial coating (ppCar) with an average roughness < 1nm ppCar had a static water contact angle of 78° Reduced effectively Escherichia coli (86%) and Staphylococcus aureus (84%) Broken bacterial membrane | [23]       |
| Carvone                   | Purchased                                   | Staphylococcus aureus       | Live-dead fluorescence assay, Crystal violet assay, Morphology of bacteria by field emission scanning electron microscope (FE-SEM) | Antibacterial activity | [23]       |
| (−)-Carvone, (+)-Carvone   | Purchased                                   | Absidia glauca, Staphylococcus aureus, Escherichia coli, Pseudomonas aeruginosa, Enterobacter aerogenes, Proteus vulgaris, and Salmonella typhimurium | Biotransformation Semi-preparative scale, Biotransformation and isolation, GC-MS | Biotransformation of carvone into diol 10-hydroxy-(-)-neodihydrocarveol by Absidia glauca Both molecules showed antimicrobial activity against all strains tested | [24]       |
| Molecules                                                                 | Origins                      | Model Used                                                                 | Experimental Approaches                                                                 | Key Results                                                                                                                                                                                                 | References |
|--------------------------------------------------------------------------|------------------------------|----------------------------------------------------------------------------|-----------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------|
| Semicarbazone and thiosemicarbazone of R-(-) carvone                    | Synthetized                  | Escherichia coli, Staphylococcus aureus, Pseudomonas aeruginosa, and Enterococcus faecalis | Determination of MIC                                                                    | Inhibitory activity on *Pseudomonas aeruginosa* for thiosemicarbazone (MIC = 78.1 µg/mL) and for semicarbazone (MIC = 312.5 µg/mL).
Thiosemicarbazone was active on *Staphylococcus aureus* (MIC = 39 µg/mL).
Thiosemicarbazone exerted interesting inhibitory activity on *Staphylococcus aureus* and *Pseudomonas aeruginosa*.                                                                                           | [26]       |
| Carvone                                                                  | Purchased                    | Staphylococcus aureus and Enterococcus coli                               | Nanoparticles preparation Determination of drug loading and entrapment efficiency In vitro carvone release from nanoparticles Antibacterial properties of the carvone-loaded nanoparticles | Production of small nanoparticles (126 nm), with high drug loading (12.32%) and good inhibition of microbial growth.
Carvone-loaded nanoparticles inhibited *Staphylococcus aureus* (MIC = 182 mg/mL) and *Enterococcus coli* (MIC = 374 mg/mL).                                                                                   | [25]       |
| (+)-carvone, (-)-carvone, (+)-hydroxy-dihydrocarvone, (-)-hydroxy-dihydrocarvone, α,β-epoxycarvone | Synthetized/purchased       | Escherichia coli and Staphylococcus aureus                               | Determination of MIC by microplate dilution method and MBC                              | C- and HC- showed low activity against *Escherichia coli*
EP, C+, and HC+ did not inhibit the growth of the bacterial strains tested.                                                                                                                                 | [20]       |
| R-carvone, S-carvone                                                     | Purchased                    | Methicillin-resistant *Staphylococcus aureus* (MRSA)                      | Broth micro-dilution method Time-kill assay                                              | MIC values for R- and S-carvone against six different strains of *Staphylococcus aureus* ranged between 500 and 1000 µg/mL.
R-carvone + gentamicin and S-carvone + gentamicin exhibited significant synergistic activity against MRSA.
The combined treatment improved the effectiveness of carvone.                                                                                                                                           | [27]       |
Aggarwal et al. [22] investigated the antibacterial effects of *Mentha spicata* (containing (S)-carvone as main component (56%) and *Anethum sowa* (containing (R)-carvone as major component (50.4%)). Evaluation of the in vitro bioactivity of the separated oily components showed that both optical isomers of carvone were active against a wide range of microorganisms tested. The activity of these monoterpene enantiomers was found to be similar to the bioactivity of the oils in which they were discovered.

Since current coating techniques, such as immobilization of antimicrobial compounds, time-releasing antibiotic agents, and silver nanoparticles, require multiple processing steps and have low efficacy and stability, Chan et al. [23] proposed a single-step plasma polymerization of carvone to produce a moderately hydrophobic antibacterial coating (ppCar) with an average roughness $< 1$ nm. Even after 10 days of air aging, ppCar maintained a static water contact angle of $78^\circ$ and remained stable for 24 h in LB broth immersion. ppCar performed well in the live/dead fluorescence test and the crystal violet assay. The biofilm test effectively reduced *E. coli* (86%) and *S. aureus* (84%) bacteria. For its bactericidal actions, it has also been shown that ppCar perforates the bacterium membrane. The cytotoxicity test revealed that the coating is not harmful to human cells. This work would be of interest to researchers interested in creating a bacteria-resistant and biocompatible coating on various substrates at low cost.

In a novel research, the microbial transformation of C- was investigated (metabolized by the phytopathogenic fungus *Absidia glauca*) by Demirci et al. [24]. The diol 10-hydroxy-(+)-neodihydrocarveol was produced after 4 days of incubation. X-ray diffraction and spectroscopy methods were used to determine the absolute arrangement and structure of the crystalline material (MS, IR, and NMR). Human pathogenic bacteria were used to test the antimicrobial activity of the substrate and metabolite. The main results of the study showed that the inhibitory action of the metabolite is modest.

In their research, Fatondji et al. [26] synthesized semicarbazone and thiosemicarbazone from R-(-)-carvone via direct condensation of semicarbazide or thiosemicarbazide in an acidic medium and evaluated their antibacterial activity. The purity of the synthesis products was determined by thin layer chromatography (TLC) after recrystallization, and their structures were verified by IR spectroscopy, nuclear magnetic resonance (NMR), and mass spectrometry (MS). The compounds were evaluated on *S. aureus*, *S. faecalis*, *P. aeruginosa*, and *E. coli* strains. Therefore, the chemicals inhibited *P. aeruginosa* growth, with MIC values of 78.1 and 312.5 $\mu$g/mL for thiosemicarbazone and semicarbazone, respectively. With a MIC of 39 $\mu$g/mL, thiosemicarbazone was also active against *S. aureus*. On the other hand, *P. aeruginosa* and *S. aureus* have become increasingly resistant to antibiotics such as oxacilline (MIC = 1.5 $\mu$g/mL for *S. aureus*) and cefixime (MIC < 1 $\mu$g/mL). Both compounds showed poor antibacterial activity against *E. coli* and *E. faecalis*. Furthermore, the thiosemicarbazone showed intriguing action against *S. aureus* and *P. aeruginosa*.

In order to extend the antibacterial action of carvone, Esfandyari-Manesh et al. [25] attempted to create poly (lactic-co-glycolic acid) (PLGA) nanoparticles. These nanoparticles
were created using emulsification solvent evaporation (ESE) and nanoprecipitation techniques. Nanoparticles were studied for their shape, size and size distribution, drug loading, entrapment efficiency, release profile, and antibacterial efficacy. This allowed to generate nanoparticles with smaller size (126 nm), narrower size distribution (PDI of 0.08–0.2), higher drug loading (12.32 percent carvone), as well as better microbial growth suppression than ESE. Carvone drug release experiments in vitro at 37 °C for 4 days revealed an early burst (36%). The MIC of carvone-loaded nanoparticles against *S. aureus* and *E. coli* was 182 and 374 mg/mL, respectively. The nanoparticles created in this research were of the appropriate size and shape, and according to the antimicrobial research, EO-loaded PLGA nanoparticles may be useful in medicinal and culinary applications.

Given the reported activity of carvone in the literature, Moro et al. [20] conducted research to assess the antibacterial activity (MIC and MBC) of C+, C−, HC+, HC−, and EP against *E. coli* and *S. aureus*. In addition, HC+, HC−, and EP have been synthesized from C+ or C−. The results showed that C− and HC− possessed a weak antibacterial action against *E. coli*. In contrast, EP, C+, and HC+ had no effect on the bacterial strains examined.

The aim of Mun et al.’s [27] study was to evaluate the antibacterial activity of R- and S-carvone in combination with gentamicin (GET) against MRSA. The latter is a gram-positive bacterium which causes nosocomial pneumonia, abscesses, and surgical site infections. Multidrug resistance is common in nosocomial MRSA infections. The broth micro-dilution method was used in this research to evaluate the antimicrobial sensitivity of R- and S-carvone and GET. The MIC for R- and S-carvone against six distinct strains of *S. aureus* ranged from 500 to 1000 µg/mL. To explore the possible synergistic effects of various combinations of carvone enantiomers and GET, anti-MRSA activity was assessed using the checkerboard and time-kill tests. The results determined that R-carvone in combination with S-carvone, R-carvone in combination with GET, and S-carvone in combination with GET all showed substantial synergistic efficacy against MRSA. These results imply that the combination treatment successfully increases the anti-MRSA monotherapy activities of R-carvone, S-carvone, and GET. Carvone has been shown to be a possible adjuvant antibacterial agent in this research.

*L. alba* EO and its main components (citral and carvone) were tested in vitro by Porfirio et al. [28] for their antibacterial and antibiofilm properties against *S. aureus*. Hydrodistillation was used to extract the EOs from *L. alba* aerial parts, which were then evaluated by GC-MS. The microdilution method was used to determine the MIC and MBC. The biomass development in the biofilm was assessed using the microtiter plate technique with the assay of crystal violet and the analysis of the viability of bacterial cells. The results of the present study indicate that all of the oils and their major components have antibacterial action, and the lowest MIC and MBC values were 0.5 mg/mL when LA1EO and citral were used. Likewise, a potential 100% inhibition of *S. aureus* biofilm formation was observed at the concentration of 0.5 mg/mL of all EOs. In
contrast, the elimination of cells from the biofilm was confirmed at concentrations of 1, 2, 2, and 0.5 mg/mL for LA1EO, LA2EO, LA3EO, and citral, respectively.

3.2.6. Antiviral Activity
Carvone was also tested for its antiviral effects (Table 5).

| Molecules           | Models Used         | Experimental Approaches          | Key Results                                                                 | References |
|---------------------|---------------------|----------------------------------|-----------------------------------------------------------------------------|------------|
| Two analogues of    | In silico study     | Molecular docking               | All ligands showed strong binding affinity against active neuraminidase sites, ranging from −4.77 to −8.30 kcal/mol | [38]       |
| carvone             |                     | Molecular dynamics simulation    | Carvone derivatives could serve as potent neuraminidase inhibitors against the influenza virus |            |

Given the recent outbreaks of highly dangerous influenza viruses, it was found that it is imperative to develop new anti-influenza drugs. In their in-silico study, they designed 36 ligands to analyze how they bind to neuraminidase (NA) active sites. The design is based on structural resemblance to the commercial inhibitor, oseltamivir (OTV), ligand polarity, and amino acid residues in the NA active sites. Their study result suggests that one of the designed ligands had the lowest binding energy (Δ\text{G}_{\text{bind}}) (−8.30 kcal/mol), comparable with OTV (−8.72 kcal/mol), with seven hydrogen bonds formed. Since the stability analysis indicated that the A18-NA complex was stable, this study encourages further research to synthesize and evaluate this compound [38].

3.2.7. Antioxidant Activity
Oxidative stress is often implicated in several severe and chronic illnesses such as cancers, cardiovascular diseases, diabetes, and many others. Since antioxidant compounds can mitigate oxidative stress due to of their antiradical ability and/or reducing power, the search for new effective and safe antioxidants from plants has intensified in recent years [83]. As part of this research, carvone has been investigated for its antioxidant effect by various researchers. One of the first investigations on the antioxidant properties of carvone isolated from Mentha spicata was carried out by Elmasta¸s et al. [55]. The results of the total antioxidant activity test indicated that S-carvone possess high antioxidant activity compared with \( \alpha \)-tocopherol, used as a reference antioxidant.

Carvone was investigated for its antioxidant potential by various in vitro systems, including lipid peroxidation, 2,2-diphenyl-1-picrylhydrazyl (DPPH), and phosphomolybdenum assay [37]. In this study, carvone isolated from Z. alatum showed inhibitory activity against thiobarbituric acid reactive species (TBARS) induced by some pro-oxidants (10 \( \mu \)M FeSO\(_4\) and 5 \( \mu \)M sodium nitroprusside) in rat liver and brain homogenates. Carvone also caused the scavenging of the DPPH radical and the reduction of molybdenum, Mo(VI) to Mo(V). Galstyan et al. [84] documented the antioxidant property of a synthesis of carvone-derived 1,2,3-triazoles. The conjugates prepared demonstrated high antioxidant activity.

3.2.8. Anti-Inflammatory Activity
The anti-inflammatory effects of terpenoids compounds such as carvone were revealed by numerous studies [85]. Monoterpene \( \alpha,\beta \)-epoxy-carvone was evaluated by da Rocha et al. [33] for its anti-inflammatory properties. This was carried out using the acetic-acid-induced peritoneal capillary permeability test. The results showed that the intraperitoneal administration of \( \alpha,\beta \)-epoxy-carvone (300 mg/kg) inhibits the increase in vascular permeability caused by acetic acid. These findings suggest that \( \alpha,\beta \)-epoxy-carvone inhibits the acute inflammatory reaction.
The anti-inflammatory activity of cyane–carvone (CC), a monocyclic monoterpene, was evaluated by the methods of bradykinin, histamine, prostaglandin E\textsubscript{2} (PGE\textsubscript{2}), serotonin, and carrageenan-induced paw edema in mice [34]. It was found that in bradykinin, histamine, PGE\textsubscript{2}, and serotonin tests, 75 mg/kg CC significantly decreased paw edema (t = 30, 60, 90, and/or 120 min). While in the carrageenan test, 50 mg/kg and 75 mg/kg CC (t = 3 h and t = 4 h) and 25 mg/kg CC (t = 4 h) significantly decreased paw edema.

Zhao and Du [35] examined the anti-inflammatory and defensive role of \textit{D}-carvone on lipopolysaccharide (LPS)-initiated lung damage in mice. As a result, \textit{D}-carvone significantly attenuated the lung damage produced by LPS by reducing the lung wet-to-dry (W/D) ratio as well as the amount of total cells, macrophages, and neutrophils in BALF. An important reduction in serum TNF-\alpha, IL-1\beta, and IL-6 levels was observed in \textit{D}-carvone treated mice. This molecule also altered histopathological disorders due to LPS-initiated lung damage. The effects of \textit{D}-carvone were comparable with those of the positive control, dexamethasone. This study indicates that pre-treatment with \textit{D}-carvone significantly provided an anti-inflammatory and protective effect against LPS-instigated lung damage.

The molecular mechanism responsible for the anti-inflammatory properties of (\textit{S})-(\textit{+})-carvone has been highlighted by Sousa et al. [36]. The results of this study show that (\textit{S})-(\textit{+})-carvone is a novel Sirtuin-1 (SIRT1) activator with the potential to counteract the chronic low-grade inflammation characteristic of age-related diseases.

This substance appears to have various uses, this time following a study by da Rocha et al. [33] on mice, to examine the antinociceptive and anti-inflammatory activities of \textit{a} and \textit{\beta} Epoxy-carvone monomers, extracted from EOs of many plant species or obtained by organic synthesis. After intraperitoneal administration of this monomer at doses of 100 mg/kg, 200 mg/kg, or 300 mg/kg, a significant antinociceptive effect was observed, as shown by the abdominal contortion test induced by acetic acid, with a decrease in blood pressure. Nociception was induced by formalin in the first (300 mg/kg) and second phase (200 and 300 mg/kg). These results suggest that the \textit{a} and \textit{\beta} Epoxy-carvone monomer inhibits the acute inflammatory response, with a peripheral and central antinociceptive effect observed in mice which can be explained by the activation of the opioid system, responsible for the antinociceptive activity induced by this monomer.

Another work was carried out by Mogosan et al. [86] in order to have a qualitative and quantitative comparative analysis of the chemical composition and to evaluate the anti-inflammatory and antinociceptive effects of the EOs of three species of Mentha (\textit{Mentha piperita} L. var. pallescens (white peppermint), \textit{Mentha spicata} L. subsp. crispata (spearmint), and \textit{Mentha suaveolens} Ehrh. (pineapple mint)) grown in Romania. The anti-inflammatory activities of EOs were determined by the carrageenan-induced rat paw edema test, while the antinociceptive activity was assessed by the contortion test in mice, using a solution of 1% (\textit{v/v}) acetic acid administered intraperitoneally and by the hot plate test in mice. The data from this study showed that the EOs chemotype of \textit{M. spicata} L. (EOMSP) produced statistically significant and dose-dependent anti-inflammatory and antinociceptive effects.

### 3.2.9. Anticancer Activity

Similar to several other bioactives which showed promising anticancer properties [87–93], carvone has also been studied for its anticancer properties. Indeed, several in vitro investigations based on cell culture tests showed that this compound exhibits antiproliferative effects against various cancer cell lines. In this sense, the results of the study conducted by Ding et al. [29] demonstrated that carvone exerts significant antiproliferative effects against myeloma cancer cells in a dose-dependent manner. The anticancer activities were linked to the induction of apoptosis and the G\textsubscript{2}/M cell cycle arrest (Figure 3). Moreover, carvone could inhibit cell invasion and p-P38 protein expression at an IC\textsubscript{50} of 20 \mu M. In another study, Gopalakrishnan et al. [30] evaluated the chemopreventive efficacy of \textit{D}-carvone (at 10, 20, and 30 mg/kg b.w.) in vivo using 7,12-dimethylbenz(a)anthracene (DMBA)-induced skin carcinogenesis. The results showed that the tumor incidence of 100% in DMBA-painted animals as well as \textit{D}-carvone at a dose of 20 mg significantly prevented...
skin carcinogenesis. In addition, this study showed decreased levels of phase I enzymes (cytochrome P450 and cytochrome b5) with increased levels of phase II enzymes (GR, GST, and GSH) and increased expression of Bax, caspase-3, and caspase-9 with decreased expression of mutated p53 and Bcl-2 in animals treated with DMBA and D-carvone (20 mg).

In another experiment [31], carvone was investigated for its ability to reduce melanin content in melanoma cells as well as for its beneficial effects mediated by the cAMP pathway. This study found that carvone decreases the melanin content and inhibits melanoma cell proliferation in a dose-dependent manner. Likewise, it caused the inactivation of cell cycle-associated proteins such as cyclin-dependent kinase 1 (CDK1). It should be noted that the beneficial activities of carvone were abrogated by cAMP inhibition.

Furthermore, Patel and Thakar [32] evaluated the antiproliferative and apoptotic activity of L-carvone, and the underlying mechanism(s) of action on breast cancer (MCF 7 and MDAMB 231) and normal (MCF 10A) cell lines. Results showed that L-carvone exhibited a strong antiproliferative effect against MCF7 (IC_{50} = 1.2 mM) and MDA MB 231 cells (IC_{50} = 1.0 mM), inhibited the migration of breast cancer cell lines, and induced apoptosis. L-carvone exposure arrested MCF 7 cells in S phase of the cell cycle and caused DNA damage that was apparent from the increased tail movement, which could be caused by an increase in reactive oxygen species (ROS). Moreover, the glutathione levels were increased. Finally, p53 and caspase-mediated apoptosis was attributed to the increased level of p53, Bad, cleaved caspase 3, and cleaved PARP (Figure 3).

3.2.10. Antiparasitic Activity

Parasitic diseases are infectious diseases caused by parasites which, under favorable conditions, can be transmitted to humans [94], among them African trypanosomiasis, malaria transmitted by Plasmodium, schistosomiasis, and leishmaniasis. Parasitic diseases are still responsible for many health problems. These parasites are responsible for a high rate of mortality and morbidity each year in endemic countries [95], and are probably
responsible for more than 1 to 2 billion infections, which result in several million deaths each year [96].

Current chemotherapy is based on developed synthetic drugs [97]. However, unfortunately, a number of these chemotherapeutic drugs against parasites have serious side effects [98]. In addition, some parasites develop resistance to the treatments [99]. The majority of people who suffer from these parasitoses often live in developing countries and have only low incomes. It is therefore urgent to discover alternative treatments against these diseases. Numerous studies have elucidated the role of plants against anthelmintic in traditional medicine [100–102]. However, a wide range of molecules isolated and identified from plants have shown promising activity against multiple parasites [103–106].

A limited number of studies have reported the parasitic activity of carvone [39,42]. Pavela, [39] evaluated the efficacy of carvone and other aromatic compounds and their mutual binary combinations for acute toxicity against *Culex quinquefasciatus* larvae. The results show that carvone is one of the substances identified with the highest synergistic effect on larval mortality. Although these results are more important when combining two aromatic molecules, in this context, among the combinations that show a higher synergistic effect on larval mortality there are carvone and carvacrol, carvone and α-terpineol, and finally carvone and menthone [39].

The evaluation of larvicidal activity of *Mentha* species EOs and their isolated major components against the West Nile virus mosquito (*Cx. pipiens* 3rd–4th instar larvae) showed that *Mentha pulegium* EO and its major components have an important anti-mosquito activity. Carvone showed moderate larvicidal activity against *Cx. pipiens* larvae of biotype molestus, with LC50 (95% CL) and LC90 (95% CL) values of 84.58–106.77 and 137.02–228.97, respectively [40]. In another study, Lima et al. [42] evaluated the larvicidal activity of *Mentha x villosa* EO (MVEO) and its major constituent, rotundifolone, against larvae of *Aedes aegypti*. The results of the study showed that MVEO exhibited outstanding toxic effects against *Ae. aegypti* larvae (LC50 = 45.0 ppm). The results of the comparative study between rotundifolone and the molecules isolated from the plant showed that all tested compounds were less potent than rotundifolone (LC50 = 62.5 ppm), except D-limonene. Concerning carvone, it exhibited intermediate larvicidal activity with (+)-carvone epoxide (LC50 = 254.6 ppm) and (−)-carvone epoxide (LC50 = 217.5 ppm) [42].

Katiki et al. [41] assessed in vitro the anthelmintic activity of aromatic compounds present in plants, on eggs collected from sheep droppings infected with the multidrug-resistant strain of gastrointestinal nematode *Haemonchus contortus*. The results obtained show that carvone is one the five promising compounds with anthelmintic activity with an LC50 value of 0.085 mg/mL [41].

### 3.2.11. Anti-Arthritic Activity

In a study conducted on arthritic rats, Chen et al. [107] evaluated the anti-arthritic activity of D-carvone against arthritis induced by Freund’s complete adjuvant (FCA) in rats. Following oral administration of D-carvone for 25 days at doses of 30 mg/kg and 60 mg/kg against FCA-induced arthritic rats, certain changes were observed, namely, improvement in body weight, reduction in leg swelling, edema formation, and organ index in these arthritic rats. Other improvements were also recorded concerning the decrease in the levels of white blood cells, with an improvement in the levels of red blood cells and hemoglobin, a decrease in the levels of lipid peroxidation, with the observation of a significant increase in the levels of enzymatic and non-enzymatic antioxidants. The results of this study showed the crucial role of D-carvone in significantly modulating inflammatory cytokine levels and improving ankle joint pathology against FCA-induced arthritic inflammation implying significant antiarthritic activity in rats.

### 3.2.12. Anticonvulsant Activity

Many drugs block seizures, but with little effect in preventing or curing this disease. Thus, the pharmaceutical industries continue to develop new safe and effective therapeutic
alternatives to the management of epilepsy. In this perspective, and in a study conducted by Costa et al. [108] in a model of epilepsy induced by pilocarpine, CC showed an anticonvulsant effect. Indeed, administration of the 25 mg/kg, 50 mg/kg, or 75 mg/kg doses of CC resulted in a reduction of 16.7%, 33%, and 66.7%, respectively, against pilocarpine-induced seizures, and was effective to increase both the latency to first seizures and the percent survival, resulting in 33.3%, 67%, and 91.7% protection against seizure-induced death, respectively. Knowing that the reference drug atropine administered at a dose of 25 mg/kg creates significant 100% protection, likewise, the results of this study were marked by an increase in acetylcholinesterase activity in the hippocampus of mice after seizures induced by pilocarpine. These data suggest the clear association of the anticonvulsant capacity of CC and the increased activity of the enzyme acetylcholinesterase.

In addition, another study was conducted on models of epilepsy by Marques et al. [109] to study the effects of CC against seizures induced by pilocarpine (PILO), pentylentetrazole (PTZ), and picrotoxin (PTX) in mice. After acute CC treatment at repeated oral doses (25 mg/kg, 50 mg/kg, and 75 mg/kg) for 14 days, positive anticonvulsant effects in PILO, PTZ, and PTX epilepsy models were recorded. Furthermore, it was shown that this substance could act on an allosteric site of GABAA, different from the site of action of BZD.

3.2.13. Anxiolytic Activity

Another research work was performed on male Wistar rats by Hatano et al. [78], this time showing the anxiolytic effects of (R)-(−)-carvone, extracted from *L. alba* EO which is widely used in the regions of Central and South America as a tranquilizer. Data from this research suggests that *L. alba* may exert anxiolytic-like effects on a specific subset of defensive behaviors that have been implicated in generalized anxiety disorder, and suggest that carvone is one of the constituents of *L. alba* responsible for its action as a tranquilizer.

3.2.14. Immunomodulatory Activity

On mice, another study was carried out by Lasarte-Cia et al. [110] in order to explore the immunomodulatory capacity of a series of compounds representing each of the 10 categories or groups of odors, including that of carvone, while highlighting its potential as a therapeutic agent for diseases related to the CNS. During this research, the impact of each particular odor on the immune response was evaluated after immunization with the model ovalbumin antigen in combination with the TLR3 agonist poly I:C. As a result of this study, it was shown that some odors can behave as immunostimulating agents, while others could be considered as potential immunosuppressive odors. In this context, it was found that inhaling the odor of carvone can have an immunostimulating effect with improved memory capacity in BALB/c and immunosuppressive mice, consequently leading to a deterioration of the memory ability in C57BL/6J mice, while facilitating or altering viral clearance, respectively, in a model of viral infection with a recombinant adenovirus. This can be explained by a higher infiltration of CD3+ T lymphocytes in the hippocampus and an increased local expression of the mRNA encoding the cytokines IL-1β, TNF-α, and IL-6, and a decrease in the number of CD3 and an increase in IFN-γ.

3.2.15. Antispasmodic Activity

Another intestinal antispasmodic virtue of C- has been revealed in a study conducted by Souza et al. [111]. The terminal parts of the ileum were mounted for isotonic contraction recordings. The effect of C− has been compared with that of verapamil, which is known to be a classic calcium channel blocker. The results of the measurement of the contractile response caused by C- showed that this monoterpenic reduces the contraction induced by a high K+, with 100 times more potency than verapamil. This is a typical action of the classic type of calcium channel blocker.
3.2.16. Acaricidal Activity

In 2015, Peixoto et al. [77] demonstrated the acaricidal activity of EOs of *L. alba* genotypes and of its main monoterpenes (carvone, limonene, and citral) against *Rhipicephalus microplus* ticks. The data obtained showed that citral had the greatest efficacy against *R. microplus* larvae, with an LC$_{50}$ of 7.0 mg/mL, followed by R-(−)-carvone (LC$_{50}$ = 9.9 mg/mL), and S-(+)-carvone (LC$_{50}$ = 10.9 mg/mL). However, the limonene enantiomers reached LC$_{50}$ values of 31.2 mg/mL for R-(+)-limonene and 54.5 mg/mL for S-(−)-limonene. In this regard, these results suggest that carvone as a major component of this EO may constitute an ecological alternative in the control of ticks in livestock.

3.2.17. Antimanic Activity

In a study by Nogoceke, et al. [112], the antimanic activity of (R)-(−)-carvone and (S)-(+) -carvone was evaluated in mouse models of mania (with hyperlocomotion caused by methylphenidate (5 mg/kg) or sleep deprivation for 24 h). After pre-treatment with (R)-(−)-carvone (50–100 mg/kg), (S)-(+) -carvone (50–100 mg/kg), and lithium (100 mg/kg positive control), the results showed that these doses do not modify spontaneous locomotor activity in the experiments induced by methylphenidate, whereas (S)-(+) -carvone caused a decrease in spontaneous locomotor activity in the experiment of sleep deprivation, which explains the induction of a sedative effect. Likewise, a 21-day chronic treatment with (R)-(−)-carvone (100 mg/kg), (S)-(+) -carvone (100 mg/kg), and lithium also allowed to block the hyperactivity induced by methylphenidate.

4. Conclusion and Perspectives

Here we have reported the benefits and pharmacological properties of carvone. This compound exhibited remarkable biological effects in vitro and in vivo and therefore may be a key candidate in drug development. Indeed, its anticancer and anti-inflammatory activities are promising with different mechanisms of action which allow us to consider it as a potential agent for the development of new anti-inflammatory and anticancer drugs. However, the pharmacodynamic actions were not well understood and, therefore, further investigations should be carried out to elucidate its mechanisms. Moreover, the study of combination between carvone used drugs in chemotherapy as well as its capacity to induce the sensitivity towards chemotherapy should be investigated. The antimicrobial action of carvone is also expected to be well determined in subsequent studies, and therefore further works should be investigated to determine its mechanism at subcellular, cellular, and molecular levels. Future clinical applications of carvone as an anticancer, anti-inflammatory, and antimicrobial drug could be developed in further investigations. However, these studies should take into account the validation of several steps. Indeed, the pharmacokinetic parameters should also be examined to determine the absorption, bioavailability, metabolism, and elimination of carvone. Moreover, toxicological investigations should be performed to validate its safety for other pharmaceutical applications.

Author Contributions: Conceptualization, A.B. (Abdelhak Bouyahya), H.M., T.B., M.A.S. and N.E.O.; writing—original draft preparation, A.B. (Abdelhak Bouyahya), H.M., T.B., R.G., S.C., A.B. (Abdelaali Balahbib) and N.E.O.; writing—review and editing, P.B., M.A.S., N.E.O. and J.M.L.; supervision, A.B. (Abdelhak Bouyahya), N.E.O. and J.M.L. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: Thanks to GAIN (Axencia Galega de Innovación) for supporting this research.

Conflicts of Interest: The authors declare no conflict of interest.
Abbreviations

| Abbreviation | Description |
|--------------|-------------|
| Bel-2        | B-cell lymphoma 2 |
| BITC         | Benzyl isothiocyanate |
| BZD          | Benzodiazepines |
| CAP          | Compound action potential |
| CDK1         | Cyclin-dependent kinase 1 |
| CNS          | Central nervous system |
| CV           | Crystal violet |
| DPPH         | 2,2-diphenyl-1-picrylhydrazyl |
| EO           | Essential oil |
| ESE          | Evaporation solvent |
| FE-SEM       | Field emission scanning electron microscope |
| GC-MS        | Gas chromatography–mass spectrometry |
| GM-MIC       | Geometric means–minimal inhibitory concentration |
| GTT          | Glucose tolerance test |
| Hb           | Hemoglobin |
| HbA1c        | Glycosylated hemoglobin |
| ICS0         | Half-maximal inhibitory concentration |
| IFN-γ        | Interferon gamma |
| IL           | Interleukin |
| ITT          | Insulin tolerance test |
| LC50         | Lethal concentration 50% |
| LD50         | Medium lethal dose |
| LPS          | Lipopolysaccharide |
| MBC          | Minimum bactericidal concentration |
| MFC          | Minimum fungicidal concentration |
| MIC          | Minimum inhibitory concentration |
| MRSA         | Methicillin-resistant staphylococcus aureus |
| MS           | Mass spectrometry |
| NMR          | Nuclear magnetic resonance |
| PARP         | Poly adenosine diphosphate-ribose polymerase |
| PGE2         | Prostaglandin E2 |
| PLA          | Poly (lactic acid) |
| PLGA         | Poly (lactic-co-glycolic acid) |
| ROS          | Reactive oxygen species |
| SIRT1        | Siruin-1 |
| STZ          | Streptozotocin |
| TBARS        | Thio barbituric acid reactive species |
| TLC          | Thin layer chromatography |
| TNF-α        | Tumor necrosis factor-α |

References

1. Bouyahya, A.; Chamkhi, I.; Benali, T.; Guouguauou, F.-E.; Balahbib, A.; El Omari, N.; Taha, D.; Belmehdi, O.; Ghokhan, Z.; El Menyiy, N. Traditional Use, Phytochemistry, Toxicology, and Pharmacology of *Origanum majorana* L. *J. Ethnopharmacol.* 2021, 265, 113318. [CrossRef] [PubMed]

2. Bouyahya, A.; Et-Touys, A.; Abrini, J.; Talbaoui, A.; Fellah, H.; Bakri, Y.; Dakka, N. *Lavandula stoechas* Essential Oil from Morocco as Novel Source of Antileishmanial, Antibacterial and Antioxidant Activities. *Biocatal. Agric. Biotechnol.* 2017, 12, 179–184. [CrossRef] [PubMed]

3. Bouyahya, A.; El Omari, N.; Elmenyiy, N.; Guouguauou, F.-E.; Balahbib, A.; El-Shazly, M.; Chamkhi, I. Ethnomedicinal Use, Phytochemistry, Pharmacology, and Toxicology of *Ajuga iva* (L.) Schreb. *J. Ethnopharmacol.* 2020, 258, 112875. [CrossRef]

4. Bouyahya, A.; Chamkhi, I.; Guouguauou, F.-E.; Benali, T.; Balahbib, A.; El Omari, N.; Taha, D.; El-Shazly, M.; El Menyiy, N. Ethnomedicinal Use, Phytochemistry, Pharmacology, and Food Benefits of *Thymus capitatus*. *J. Ethnopharmacol.* 2020, 259, 112925. [CrossRef]

5. Bouyahya, A.; Lagrouh, F.; El Omari, N.; Bourais, I.; El Jemli, M.; Marmouzi, I.; Salhi, N.; Faouzi, M.E.A.; Belmehdi, O.; Dakka, N.; et al. Essential Oils of *Mentha viridis* Rich Phenolic Compounds Show Important Antioxidant, Antidiabetic, Dermatoprotective, Antidermatophyte and Antibacterial Properties. *Biocatal. Agric. Biotechnol.* 2020, 23, 101471. [CrossRef]
6. Bouyahya, A.; Dakka, N.; Talbaoui, A.; Et-Touys, A.; El-Boury, H.; Abrini, J.; Bakri, Y. Correlation between Phenological Changes, Chemical Composition and Biological Activities of the Essential Oil from Moroccan Endemic Oregano (Origanum compactum Benth). Ind. Crops. Prod. 2017, 108, 729–737. [CrossRef]

7. Bouyahya, A.; Belmehdi, O.; El Jeniti, M.; Marmouzi, I.; Bourais, I.; Abrini, J.; Faouzi, M.E.A.; Dakka, N.; Bakri, Y. Chemical Variability of Centaurium erythraea Essential Oils at Three Developmental Stages and Investigation of Their In Vitro Antioxidant, Antidiabetic, Dermatoprotective and Antibacterial Activities. Ind. Crops Prod. 2019, 132, 111–117. [CrossRef]

8. Bouyahya, A.; Et-Touys, A.; Bakri, Y.; Talbaoui, A.; Fellah, H.; Abrini, J.; Dakka, N. Chemical Composition of Mentha pulegium and Rosmarinus officinalis Essential Oils and Their Antileishmanial, Antibacterial and Antioxidant Activities. Microb. Pathog. 2017, 111, 41–49. [CrossRef]

9. De Sousa, D.P.; De Farias Nobrega, F.F.; De Almeida, R.N. Influence of the Chirality of (R)-(−) and (S)-(+) Carvone in the Central Nervous System: A Comparative Study. Chirality 2007, 19, 264–268. [CrossRef]

10. Faliagkas, L.; Yokou, D.; Theophilidis, G. Local Anaesthetic Properties vs. Neurotoxicity for (+)-and (−)-Carvone: An Ex Vivo Electrophysiological Study. Planta Med. Lett. 2015, 2, e6–e9. [CrossRef]

11. Gonçalves, J.C.R.; de Alves, A.M.H.; de Araújo, A.E.V.; Cruz, J.S.; Araújo, D.A.M. Distinct Effects of Carvone Analogues on the Isolated Nerve of Rats. Eur. J. Pharmacol. 2010, 645, 108–112. [CrossRef] [PubMed]

12. Sánchez-Borzone, M.; Delgado-Marín, L.; García, D.A. Inhibitory Effects of Carvone Isomers on the GABA A Receptor in Primary Cultures of Rat Cortical Neurons: Effects of Carvone Isomers on Gaba A Receptor. Chirality 2014, 26, 368–372. [CrossRef]

13. Alsania, S.; Liu, D. BITC and S-Carvone Restrain High-Fat Diet-Induced Obesity and Ameliorate Hepatic Steatosis and Insulin Resistance. Pharm. Res. 2017, 34, 2241–2249. [CrossRef] [PubMed]

14. Muruganathan, U.; Srinivasan, S.; Indumathi, D. Antihyperglycemic Effect of Carvone: Effect on the Levels of Glycoprotein Components in Streptozotocin-Induced Diabetic Rats. J. Acute Dis. 2013, 2, 310–315. [CrossRef]

15. Muruganathan, U.; Srinivasan, S. Beneficial Effect of Carvone, a Dietary Monoterpene Ameliorates Hyperglycemia by Regulating the Key Enzymes Activities of Carbohydrate Metabolism in Streptozotocin-Induced Diabetic Rats. Biomed. Pharmacother. 2016, 84, 1558–1567. [CrossRef] [PubMed]

16. Boonruang, K.; Chinsirikul, W.; Hararak, B.; Kerddonfag, N.; Chonhenchob, V. Antifungal Poly (Lactic Acid) Films Containing Thymol and Carvone. MATEC Web Conf. 2016, 67, 06107. [CrossRef]

17. Giovana, C.B.; de Simone, N.B.F.; de Priscilla, L.S.; Paula, C.A.; Marcelo, F.G.B.; Yeop Majlis, B. Plasma Polymerized Carvone as an Antibacterial and Biocompatible Coating. Mem. Inst. Oswaldo Cruz 2009, 104, 878–884. [CrossRef] [PubMed]

18. Mohamed, M.; Malnati, M.; Terzi, V. In Vitro Antifungal Activity of Terpinen-4-Ol, Menthol, and Carvone from the Essential Oils of Colombian Lippia alba (Mill.) NE Brown: Composition, Cytotoxicity and Antifungal Activity. Mem. Inst. Oswaldo Cruz 2009, 104, 878–884. [CrossRef] [PubMed]

19. Morcya, C.; Malnata, M.; Terzib, V. In Vitro Antifungal Activity of Terpinen-4-Ol, Eugenol, Carvone, 1,8-Cineole (Eucalyptol) and Thymol against Mycotoxicogenic Plant Pathogens. Food Addit. Contam. Part A 2011, 5, 1–8. [CrossRef]

20. Moro, I.J.; Gondo, G.D.G.A.; Pierri, E.G.; Bah, R.C.L.R.; dos Santos, C.P.; de Sousa, D.P.; dos Santos, A.G. Evaluation of Antimicrobial, Cytotoxic and Chemopreventive Activities of Carvone, Menthone, Menthohuran and Pulegone from Mentha spp. Afr. J. Plant Sci. 2016, 10, 203–210. [CrossRef]

21. Muruganathan, U.; Srinivasan, S. In Vitro Antifungal Activity of Terpinen-4-Ol, Eugenol, Carvone, Thymol and Carvone Isolated from the Oils of Colombian Lippia alba (Mill.) NE Brown: Composition, Cytotoxicity and Antifungal Activity. Mem. Inst. Oswaldo Cruz 2009, 104, 878–884. [CrossRef] [PubMed]

22. Fang, Y.; Siow, K.S.; Ng, P.Y.; Gires, U.; Yeop Majlis, B. Plasma Polymerized Carvone as an Antibacterial and Biocompatible Coating. Mater. Sci. Eng. C 2016, 68, 861–867. [CrossRef]

23. Demirci, F.; Noma, Y.; Basera, K.H.C. Microbial Transformation of (−)-Carvone. Z. Natur. C 2004, 59, 389–392. [CrossRef]

24. Esfandyari-Manesh, M.; Ghaedi, Z.; Asemi, M.; Khanavi, M.; Manayi, A.; Jamalifar, H.; Atyabi, F.; Dinarvand, R. Study of the Antileishmanial, Antibacterial and Antioxidant Activities of (−)- and (S)-(+) Carvone: An Ex Vivo Study. Z. Natur. C 2016, 53, 861–871. [CrossRef]

25. Fatondji, H.R.; Gbaguidi, F.; Kpoviessi, S.; Sonounameto, E.; Lagnika, L.; Ambaliou, S.; Moudachi, M.; Poupaert, J.; Acquah, G. Thymol and Carvone. Z. Natur. C 2014, 51, 392–397. [CrossRef]

26. Fatondji, H.R.; Gbaguidi, F.; Kpoviessi, S.; Sonounameto, E.; Lagnika, L.; Ambaliou, S.; Moudachi, M.; Poupaert, J.; Acquah, G. Synthèse, Génése et Études de Propriétés Antimicrobien des Nervures de La Semicarbazone et de La Thiosemicarbazone de La Carvone. J. Soc. Ouest Afr. Chim. 2010, 30, 11–17.

27. Mun, S.-H.; Kang, O.-H.; Joung, D.-K.; Kim, S.-B.; Choi, J.-G.; Shin, D.-W.; Kwon, D.-Y. In Vitro Anti-MRSA Activity of Carvone with Gentamicin. Exp. Ther. Med. 2017, 7, 989–996. [CrossRef] [PubMed]

28. Porfiroiu, E.M.; Melo, H.M.; Pereira, A.M.G.; Cavalcante, T.A.; Gomes, G.A.; de Carvalho, M.G.; Costa, R.A.; Júnior, F.E.A.C. In Vitro Antibacterial and Antifibrosarcoma Activity of Lippia alba Essential Oil, Citral, and Carvone against Staphylococcus aureus. Sci. World J. 2017, 2017, 4962707. [CrossRef] [PubMed]

29. Ding, X.; Chen, H. Anticancer Effects of Carvone in Myeloma Cells Is Mediated through the Inhibition of P38 MAPK Signalling Pathway, Apotosis Induction and Inhibition of Cell Invasion. J. Buon. 2018, 5, 747–751.
30. Gopalakrishnan, T.; Ganapathy, S.; Veeran, V.; Namasingayam, N. Preventive Effect of D-Carvone during DMBA Induced Mouse Skin Tumorigenesis by Modulating Xenobiotic Metabolism and Induction of Apoptotic Events. *Biomed. Pharmacother.* 2019, 111, 178–187. [CrossRef] [PubMed]

31. Kang, W.; Choi, D.; Park, S.; Park, T. Carvone Decreases Melanin Content by Inhibiting Melanoma Cell Proliferation via the Cyclic Adenosine Monophosphate (cAMP) Pathway. *Molecules* 2020, 25, 5191. [CrossRef] [PubMed]

32. Patel, P.B.; Thakkar, V.R. D-Carvone Induces P53, Caspase 3 Mediated Apoptosis and Inhibits the Migration of Breast Cancer Cell Lines. *Nutr. Cancer* 2014, 66, 453–462. [CrossRef] [PubMed]

33. Da Rocha, M.L.; Oliveira, L.E.G.; Patricio Santos, C.C.M.; de Sousa, D.P.; de Almeida, R.N.; Araújo, D.A.M. Antinociceptive and Anti-Inflammatory Effects of the Monoterpene α-Epoxycarvone in Mice. *J. Nat. Med.* 2016, 67, 743–749. [CrossRef] [PubMed]

34. Marques, T.H.C.; Marques, M.L.B.G.C.; Medeiros, J.-V.R.; Silva, R.O.; dos Reis Barbosa, A.L.; Lima, T.C.; de Sousa, D.P.; de Freitas, R.M. Cyane-Carvone, a Synthetic Derivative of Carvone, Inhibits Antiinflammatory Response by Reducing Cytokine Production and Oxidative Stress and Shows Antinociceptive Effect in Mice. *Inflammation* 2014, 37, 966–977. [CrossRef] [PubMed]

35. Zhao, M.; Du, J. Anti-Inflammatory and Protective Effects of D-Carvone on Lipopolysaccharide (LPS)-Induced Acute Lung Injury in Mice. *J. King Saud. Univ. Sci.* 2020, 32, 1592–1596. [CrossRef]

36. Sousa, C.; Neves, B.M.; Leitão, A.J.; Mendes, A.F. Elucidation of the Mechanism Underlying the Anti-Inflammatory Properties of (S)+(−)-Carvone Identifies a Novel Class of Sirtuin-1 Activators in a Murine Macrophage Cell Line. *Biomedicines* 2021, 9, 777. [CrossRef] [PubMed]

37. Sabir, S.M.; Singh, D.; Rocha, J.B.T. In Vitro Antioxidant Activity of S-Carvone Isolated from *Zanthoxylum alatum*. *Pharm. Chem. J.* 2015, 49, 187–191. [CrossRef]

38. Jusoh, N.; Zainal, H.; Abdul Hamid, A.A.; Bunnori, N.M.; Abd Halim, K.B.; Abd Hamid, S. In Silico Study of Carvone Derivatives as Potential Neuraminidase Inhibitors. *J. Mol. Model.* 2015, 21, 93. [CrossRef]

39. Pavela, R. Acute Toxicity and Synergistic and Antagonistic Effects of the Aromatic Compounds of Some Essential Oils against *Culex quinquefasciatus* Say Larvae. *Parasitol. Res.* 2015, 114, 3853–3853. [CrossRef]

40. Michaelakis, A.; Papachristos, D.; Kimbaris, A.; Polissiou, M. Larvicidal Evaluation of Three *Mentha* Species Essential Oils and Their Isolated Major Components against the West Nile Virus Mosquito. *Hell. Plant Prot. J.* 2011, 4, 34–43.

41. Katiki, L.; Evangelista, A.; Carvone, E.; Piza, A.; Fornazari, B.; Araujo, R.; Louvandini, H.; Amarante, A.; Costa, R.; Bueno, M. Anthelmintic Activity of Anethole, Carvone, Carvacrol, Thymol, Linalool, Limonene, Eucalyptol, Vanillin, Cinnamaldehyde and Eugenol in In Vitro Tests. *Planta Med.* 2014, 80, P1L14. [CrossRef]

42. Lima, T.C.; da Silva, T.K.M.; Silva, F.L.; Barbosa-Filho, J.M.; Marques, M.O.M.; Santos, R.L.C.; de Holanda Cavalcanti, S.C.; de Sousa, D.P. Larvicidal Activity of *Mentha × Villosa hudson* Essential Oil, Rotundifolone and Derivatives. *Chemosphere* 2014, 104, 37–43. [CrossRef] [PubMed]

43. Kokkini, S.; Karousou, R.; Lanaras, T. Essential Oils of Spearmint (Carvone-Rich) Plants from the Island of Crete (Greece). *Biochem. Syst. Ecol.* 1995, 23, 425–430. [CrossRef]

44. Younis, Y.M.; Beshir, S.M. Carvone-Rich Essential Oils from *Mentha longifolia* (L.) Huds. *Ssp. Schimperi* Briq. and *Mentha spicata* L. Grown in Sudan. *J. Essent. Oil Res.* 2004, 16, 539–541. [CrossRef]

45. Abdel-Elgawad, M.M.; Omer, E.A. Effect of Essential Oils of Some Medicinal Plants on Phytonematodes. *Anz. Schädl. Pflanz. Umw.* 1995, 68, 82–84. [CrossRef]

46. Monfaread, A.; Nabid, M.R.; Rustaiyan, A. Composition of a Carvone Chemotype of *Mentha longifolia* (L.) Huds. from Iran. *J. Essent. Oil Res.* 2002, 14, 51–52. [CrossRef]

47. Da Porto, C.; Decorti, D. Ultrasound-Assisted Extraction Coupled with Under Vacuum Distillation of Flavour Compounds from Spearmint (Carvone-Rich) Plants: Comparison with Conventional Hydrodistillation. *Ultrasound. Sonochem.* 2009, 16, 795–799. [CrossRef] [PubMed]

48. Barba, C.; Toledano, R.M.; Santa-Maria, G.; Herrera, M.; Martínez, R.M. Enantiomeric Analysis of Limonene and Carvone by Direct Introduction of Aromatic Plants into Multidimensional Gas Chromatography. *Talanta* 2013, 106, 97–103. [CrossRef] [PubMed]

49. Morcia, C.; Tumino, G.; Ghizzoni, R.; Terzi, V. Carvone (Mentha spicata L.) Oils. In *Essential Oils in Food Preservation, Flavor and Safety*; Elsevier: Amsterdam, The Netherlands, 2016; pp. 309–316.

50. De Carvalho, C.C.; Da Fonseca, M.M.R. Carvone: Why and How Should One Bother to Produce This Terpene. *Food Chem.* 2006, 95, 413–422. [CrossRef]

51. De Gonçalves, N.D.; Pena, F.L.; Sartoratto, A.; Delamelina, C.; Duarte, M.C.T.; Antunes, A.E.C.; Prata, A.S. Encapsulated Thyme (*Thymus vulgaris*) Essential Oil Used as a Natural Preservative in Bakery Product. *Food Res. Int. 2017*, 96, 154–160. [CrossRef]

52. Aydin, E.; Türkeş, H.; Keleş, M.S. Potential Anticancer Activity of Carvone in *N2a* Neuroblastoma Cell Line. *Toxicol. Ind. Health* 2015, 31, 764–772. [CrossRef]

53. Rafii, F.; Shahverdi, A.R. Comparison of Essential Oils from Three Plants for Enhancement of Antimicrobial Activity of Nitrofurantoin against Enterobacteria. *Chemotherapy* 2007, 53, 21–25. [CrossRef]

54. Chauhan, R.S.; Kaul, M.K.; Shahi, A.K.; Kumar, A.; Ram, G.; Tawa, A. Chemical Composition of Essential Oils in *Mentha spicata* L. Accession [IIMJ (I) 26] from North-West Himalayan Region, India. *Ind. Crops Prod.* 2009, 29, 654–656. [CrossRef]

55. Elmastas, M.; Dermirtas, I.; Isildak, O.; Aboul-Enein, H.Y. Antioxidant Activity of S-Carvone Isolated from Spearmint (*Mentha spicata* L. Fam. Lamiaceae). *J. Liq. Chromatogr. Relat. Technol.* 2006, 29, 1465–1475. [CrossRef]
56. Telci, I.; Sahbaz, N.I.; Yilmaz, G.; Tugay, M.E. Agronomical and Chemical Characterization of Spearmint (Mentha spicata L.) Originating in Turkey. *Econ. Bot.* 2004, 58, 721–728. [CrossRef]

57. Tisserat, B.; Vaughn, S.F. Growth, Morphogenesis, and Essential Oil Production in *Mentha spicata* L. Plantlets In Vitro. *Vitro Cell. Dev. Biol. Plant* 2008, 44, 40–50. [CrossRef]

58. Taghavi Nezhad, M.; Alipour, D.; Torabi Goudarzi, M.; Zamani, P.; Khodakaramian, G. Dose Response to Carvone Rich Essential Oils of Spearmint (*Mentha spicata* L.): In Vitro Ruminal Fermentation Kinetics and Digestibility. *J. Agric. Sci. Technol.* 2011, 13, 1013–1020.

59. Khan, N.I.; Tisserat, B.; Berhow, M.; Vaughn, S.F. Influence of Autoclaved Fungal Materials on Spearmint (*Mentha spicata* L.) Growth, Morphogenesis, and Secondary Metabolism. *J. Chem. Ecol.* 2005, 31, 1579–1593. [CrossRef]

60. Chowdhury, J.U.; Nandi, N.C.; Uddin, M.; Rahman, M. Chemical Constituents of Essential Oils from Two Types of Spearmint (*Mentha spicata* L. and *M. cardiaca* L.) Introduced in Bangladesh. *Bangladesh J. Sci. Ind. Res.* 2007, 42, 79–82. [CrossRef]

61. Gherman, C.; Culea, M.; Cozar, O. Comparative Analysis of Some Active Principles of Herb Plants by GC/MS. *Planta Med.* 2011, 77, 227–233. [CrossRef]

62. Murray, M.J.; Reitsema, R.H. The Genetic Basis of the Ketones, Carvone, and Menthone in *Anethum graveolens* L. *J. Pharm. Pharmacol.* 1965, 17, 415–435. [CrossRef]

63. Zheng, G.; Kenney, P.M.; Lam, L.K. Anethofuran, Carvone, and Limonene: Potential Cancer Chemoprotective Agents from Dill Fruit. *Carvone Oxide: An Example of* Planta Med. *Weed Oil and Caraway Oil.* 1013–1020.

64. Lichtenstein, E.P.; Liang, T.T.; Schulz, K.R.; Schnoes, H.K.; Carter, G.T. Insecticidal and Synergistic Components Isolated from Dill Fruit Oil. *J. Agric. Food Chem.* 1974, 22, 658–664. [PubMed]

65. Callan, N.W.; Johnson, D.L.; Westcott, M.P.; Welty, L.E. Herb and Oil Composition of Dill (*Anethum graveolens* L.). *Chem. Planta Med.* 1974, 43, 612–613. [CrossRef]

66. Zheng, G.; Kenney, P.M.; Lam, L.K. Anethofuran, Carvone, and Limonene: Potential Cancer Chemoprotective Agents from Dill Weed Oil and Caraway Oil. *Planta Med.* 1992, 58, 338–341. [CrossRef] [PubMed]

67. Betts, T.J. Carvone in the Developing Fruits of *Anethum graveolens* L. *Anethum graveolens* L. *Planta Med.* 1994, 612–613. [CrossRef]

68. Chemat, S.; Esveld, E.D. Contribution of Microwaves or Ultrasonics on Carvone and Limonene Recovery from Dill Fruits. *Ind. Crops Prod.* 1995, 58, 157–164. [CrossRef]

69. Toxopeus, H.; Lubberts, J.H.; Neervoort, W.; Folkers, W.; Huisjes, G. Breeding Research and in Vitro Propagation to Improve Purity in Caraway, Dill and Spearmint Oils. *Ind. Crops Prod.* 1995, 5, 39–51. [CrossRef]

70. Bouwmeester, H.J.; Davies, J.A.; Smid, H.G.; Welten, R.S. Physiological Limitations to Carvone Yield in Caraway (*Carum carvi* L.). *Ind. Crops Prod.* 1995, 4, 33–38. [CrossRef]

71. Ravid, U.; Putievsky, E.; Katzir, I.; Weinstein, V.; Ikan, R. Chiral GC Analysis of (S)(+)-and (R)(−)-Carvone with High Enantiomeric Purity in Caraway, Dill and Spearmint Oils. *Flavour Fragr. J.* 1992, 7, 289–292. [CrossRef]

72. Toxopeus, H.; Lubberts, J.H. Effect of Genotype and Environment on Carvone Production of Caraway (*Carum carvi* L.). *Ind. Crops Prod.* 1995, 4, 25–28. [CrossRef]

73. András, C.D.; Salamon, R.V.; Barabás, I.; Volf, I.; Szep, A. Influence of Extraction Methods on Caraway (*Carum carvi* L.) Essential Oil Yield and Carvone/Limonene Ratio. *Envir. Eng. Manag. J. EEMJ* 2015, 14, 277. [CrossRef]

74. Şanlı, A.; Karadöğan, T. Carvone Containing Essential Oils as Sprout Suppressants in Potato (*Solanum tuberosum* L.) Tubers at Different Storage Temperatures. *Potato Res.* 2019, 62, 345–360. [CrossRef]

75. Oosterhaven, C.; Hartmans, K.J.; Huizing, H.J. Inhibition of Potato (*Solanum tuberosum*) Sprout Growth by the Monoterpenic S-Carvone: Reduction of 3-Hydroxy-3-Methylglutaryl Coenzym A Reductase Activity without Effect on Its MRNA Level. *J. Plant Physiol.* 1993, 141, 463–469. [CrossRef]

76. Whitten, W.M.; Williams, N.H.; Armbruster, W.S.; Battiste, M.A.; Strekowski, L.; Lindquist, N. Carvone Oxide: An Example of Convergent Evolution in Eudicote Pollinated Plants. *Syst. Bot.* 1986, 5, 222–228. [CrossRef]

77. Peixoto, M.G.; Costa-Júnior, L.M.; Blank, A.F.; da Silva Lima, A.; Menezes, T.S.A.; de Alexandria Santos, D.; Alves, P.B.; de Holanda Cavalcanti, S.C.; Bacci, L.; de Fátima Arrigoni-Blank, M. Acaricidal Activity of Essential Oils from *Lippia alba* Genotypes and Its Major Components Carvone, Limonene, and Citral against *Rhipicephalus microplus*. *Vet. Parasitol.* 2015, 210, 118–122. [CrossRef] [PubMed]

78. Hatano, V.Y.; Torricelli, A.S.; Giassi, A.C.C.; Coslopo, L.A.; Viana, M. de B. Anxiolytic Effects of Repeated Treatment with an Essential Oil from *Lippia alba* and (R)-(−)-Carvone in the Elevated T-Maze. *Braz. J. Med. Biol. Res.* 2012, 45, 238–243. [CrossRef] [PubMed]

79. Peixoto, M.G.;Costa-Júnior, L.M.; Blank, A.F.; da Silva Lima, A.; Menezes, T.S.A.; de Alexandria Santos, D.; Alves, P.B.; de Holanda Cavalcanti, S.C.; Bacci, L.; de Fátima Arrigoni-Blank, M. Acaricidal Activity of Essential Oils from *Lippia alba* Genotypes and Its Major Components Carvone, Limonene, and Citral against *Rhipicephalus microplus*. *Vet. Parasitol.* 2015, 210, 118–122. [CrossRef] [PubMed]

80. Orleto-Verbel, J.; Gonzalez-Cervera, T.; Guéte-Fernandez, J.; Jaramillo-Colorado, B.; Stashenko, E. Chemical Composition and Antioxidant Activity of Essential Oils Isolated from Colombian Plants. *Rev. Bras. Farmacogn.* 2010, 20, 568–574. [CrossRef]

81. Ehlerh, P.A.D.; Chaves, F.C.M.; Ming, L.C.; da SILVA, M.A.S. Effect of Substrata on the Development of Stem Cuttings of *Lippia alba* (Mill.) NE BR.-Limonene-Carvone Chemotype. In Proceedings of the International Conference on Medicinal and Aromatic Plants, Budapest, Hungary, 8–11 July 2001; pp. 259–262.
82. Bouyahya, A.; El Omari, N.; Elmenyiy, N.; Guaougouaou, F.-E.; Balahbib, A.; Belmehdi, N.; Salhi, N.; Mrabti, H.N.; El-Shazly, M. Moroccan Antidiabetic Medicinal Plants: Ethnobotanical Studies, Phytochemical Bioactive Compounds, Preclinical Investigations, Toxicological Validations and Clinical Studies; Challenges, Guidance and Perspectives for Future Management of Diabetes Worldwide. *Trends Food Sci. Technol.* 2021, 28, 2652.

83. Benali, T.; Habbadi, K.; Khabbach, A.; Marmouzi, I.; Zengin, G.; Bouyahya, A.; Chammki, I.; Chthibi, H.; Aanniz, T.; Achbani, E.H.; El Omari, N.; Bakha, M.; Imtara, H.; Guaougouaoua, F.-E.; Balahbib, A.; Zengin, G.; Bouyahya, A. Antiinflammatory and Antimicrobial Activities of Achillea odora Subsp. Pectinata and Ruta montana Essential Oils and Their Potential Use as Food Preservatives. *Foods* 2020, 9, 668.

84. Galstyan, A.S.; Martiryany, A.I.; Grigoryan, K.R.; Ghazaryan, A.G.; Samvelyan, M.A.; Ghochikyan, T.V.; Tenadjenko, V.G. Synthesis of Carvone-Derived 1, 2, 3-Trizoles of Their Antioxidant Properties and Interaction with Bovine Serum Albumin. *Molecules* 2018, 23, 2991. [CrossRef] [PubMed]

85. Bouyahya, A.; Guaougouaou, F.-E.; El Omari, N.; El Menyiy, N.; Balahbib, A.; El-Shazly, M.; Bakry, Y. Anti-Inflammatory and Analgesic Properties of Moroccan Medicinal Plants: Phytochemistry, in Vitro and in Vivo Investigations, Mechanism Insights, Clinical Evidences and Perspectives. *J. Pharm. Anal.* 2021, 7, 266. [CrossRef]

86. Mogosan, C.; Vostinaru, O.; Oprean, R.; Hughes, C.; Filip, L.; Balica, G.; Moldovan, R. A Comparative Analysis of the Chemical Composition, Anti-Inflammatory, and Antioncicitive Effects of the Essential Oils from Three Species of Mentha Cultivated in Romania. *Molecules* 2017, 22, 263. [CrossRef] [PubMed]

87. Balahbib, A.; El Omari, N.; Hachlafi, N.E.; Lakhdar, F.; El Menyiy, N.; Salhi, N.; Mrabti, H.N.; Bakrim, S.; Zengin, G.; Bouyahya, A. Health Beneficial and Pharmacological Properties of p-Cymene. *Food Chem. Toxicol.* 2021, 11, 2259. [CrossRef]

88. Bouyahya, A.; Belmehdi, O.; Benjouad, A.; El Hassani, R.A.; Amzazi, S.; Dakka, N.; Bakry, Y. Pharmacological Properties and Mechanism Insights of Moroccan Anticancer Medicinal Plants: What Are the next Steps? *Ind. Crops Prod.* 2020, 147, 112198. [CrossRef]

89. El Hachlafi, N.; Lakhdar, F.; Khouchlaa, A.; Bakrim, S.; El Omari, N.; Balahbib, A.; Shariati, M.A.; Zengin, G.; Fikri-Benbrahim, K.; Orlando, G. Health Benefits and Pharmacological Properties of Hinokitiol. * Processes* 2021, 9, 1680. [CrossRef]

90. El Omari, N.; El Menyiy, N.; Zengin, G.; Goh, B.H.; Calvo, D.; Naviglio, D.; Bouyahya, A. Anticancer and Anti-Inflammatory Effects of Tomentosin: Cellular and Molecular Mechanisms. *Separations* 2021, 8, 207. [CrossRef]

91. El Omari, N.; Bakha, M.; Elmenyiy, N.; Guaougouaou, F.-E.; Balahbib, A.; Zengin, G.; Bouyahya, A. Anticancer Mechanisms of Phytochemical Compounds: Focussing on Epigenetic Targets. *Environ. Sci. Pollut. Res.* 2021, 6, 47869–47903. [CrossRef]

92. Hachlafi, N.E.; Aanniz, T.; Menyiy, N.E.; Baaboua, A.E.; El Omari, N.; Balahbib, A.; Shariati, M.A.; Zengin, G.; Fikri-Benbrahim, K.; Bouyahya, A. In Vitro and in Vivo Biological Investigations of Camphene and Its Mechanism Insights: A Review. *Food Rev. Int.* 2021, 51, 1–28. [CrossRef]

93. Omari, N.E.; Bakrim, S.; Bakha, M.; Lorenzo, J.M.; Rebezov, M.; Shariati, M.A.; Baboulaghra, S.; Balahbib, A.; Khayrullin, M.; Bouyahya, A. Natural Bioactive Compounds Targeting Epigenetic Pathways in Cancer: A Review on Alkaloids, Terpenoids, Quinones, and Isothiocyanates. *Nutrients* 2021, 13, 3714. [CrossRef]

94. Villeneuve, A.; Polley, L.; Jenkins, E.; Schurer, J.; Gilleard, J.; Kutz, S.; Conboy, G.; Benoit, D.; Seewald, W.; Gagné, F. Parasite Prevalence in Fecal Samples from Shelter Dogs and Cats across the Canadian Provinces. *Parasit. Vectors* 2015, 8, 281. [CrossRef] [PubMed]

95. Gushulak, B.D.; MacPherson, D.W. Globalization of Infectious Diseases: The Impact of Migration. *Clin. Infect. Dis.* 2004, 38, 1742–1748. [CrossRef] [PubMed]

96. Peters, W.; Pasvol, G. Atlas of Tropical Medicine and Parasitology: Text with CD-ROM; Elsevier Health Sciences: Amsterdam, The Netherlands, 2006.

97. Nussbaum, K.; Honek, J.; Cadmus, C.; Effert, T. Trypanosomatid Parasites Causing Neglected Diseases. *Curr. Med. Chem.* 2010, 17, 1594–1617. [CrossRef]

98. Dhorda, M. Molecular Parasitology and Diagnosis of Malaria in Pregnancy. Ph.D. Thesis, University Paris, Paris, France, June 2010.

99. Gehrig, S.; Effert, T. Development of Drug Resistance in *Trypanosoma brucei rhodesiense* and *Trypanosoma brucei gambiense*. Treatment of African Trypanosomiasis with Natural Products. *Int. J. Mol. Med.* 2008, 22, 411–419.

100. Hammond, J.A.; Fielding, D.; Bishop, S.C. Prospects for Plant Anthelmintics in Tropical Veterinary Medicine. *Vet. Res. Commun.* 1997, 21, 213–228. [CrossRef] [PubMed]

101. Carvalho Barros, J.; Munekata, P.E.; de Carvalho, F.A.L.; Pateiro, M.; Barba, F.J.; Domínguez, R.; Trindade, M.A.; Lorenzo, J.M. Use of Tiger Nut (*Cyperus esculentus*) Oil Emulsion as Animal Fat Replacement in Beef Burgers. *Foods* 2020, 9, 44. [CrossRef] [PubMed]

102. Acharya, J.; Hildreth, M.B.; Reese, R.N. In Vitro Screening of Forty Medicinal Plant Extracts from the United States Northern Great Plains for Anthelmintic Activity Against *Haemonchus contortus*. *Vet. Parasitol.* 2014, 201, 75–81. [CrossRef] [PubMed]

103. Baranova, A.; Sergiev, V.; Morozova, L.; Turbina, N.; Morozov, E. Imported Plasmodium vivax Malaria in the Russian Federation from Western Sub-Saharan Africa. *J. Trop. Med.* 2019, 2019, 16–27. [CrossRef] [PubMed]

104. Barrau, E.; Fabre, N.; Fouraste, I.; Hoste, H. Effect of Bioactive Compounds from Sainfoin (*Oxobrychus vicifolia* Scop.) on the in Vitro Larval Migration of *Haemonchus contortus*: Role of Tannins and Flavonol Glycosides. *Parasitology* 2005, 131, 531–538. [CrossRef] [PubMed]
105. Akendengué, B.; Champy, P.; Nzamba, J.; Roblot, F.; Loiseau, P.M.; Bories, C. Antifungal and Anthelmintic Activities of Cleistopholis patens (Annonaceae). *Planta Med.* 2009, 75, 1143–1145. [CrossRef] [PubMed]

106. Stepek, G.; Buttle, D.J.; Duce, I.R.; Lowe, A.; Behnke, J.M. Assessment of the Anthelmintic Effect of Natural Plant Cysteine Proteinases against the Gastrointestinal Nematode, Heligmosomoides polygyrus, In Vitro. *Parasitology* 2005, 130, 203–211. [CrossRef] [PubMed]

107. Chen, G.; Song, Y.; Ma, F.; Ma, Y. Anti-Arthritic Activity of α-Carvone against Complete Freund’s Adjuvant-Induced Arthritis in Rats through Modulation of Inflammatory Cytokines. *Korean J. Physiol. Pharmacol.* 2020, 24, 453–462. [CrossRef] [PubMed]

108. Costa, A.G.; Bertolucci, S.K.V.; Chagas, J.H.; Ferraz, E.O.; Pinto, J.E.B.P. Biomass Production, Yield and Chemical Composition of Peppermint Essential Oil Using Different Organic Fertilizer Sources. *Ciênc. Agrotecnol.* 2013, 37, 202–210. [CrossRef] [PubMed]

109. Marques, T.H.C.; Branco, M.L.B.G.C.; Medeiros, J.-V.R.; Lima, T.C.; de Sousa, D.P.; de Freitas, R.M. Anticonvulsant Effects of Acute Treatment with Cyane-Carvone at Repeated Oral Doses in Epilepsy Models. *Pharmacol. Biochem. Behav.* 2014, 124, 421–424. [CrossRef] [PubMed]

110. Lasarte-Cia, A.; Lozano, T.; Pérez-González, M.; Gorraiz, M.; Iribarren, K.; Hervás-Stubbis, S.; Sarobe, P.; Rabal, O.; Cuadrado-Tejedor, M.; García-Osta, A.; et al. Immunomodulatory Properties of Carvone Inhalation and Its Effects on Contextual Fear Memory in Mice. *Front. Immunol.* 2018, 9, 68. [CrossRef] [PubMed]

111. Souza, F.V.M.; da Rocha, M.B.; de Souza, D.P.; Marçal, R.M. (−)-Carvone: Antispasmodic Effect and Mode of Action. *Fitoterapia* 2013, 85, 20–24. [CrossRef] [PubMed]

112. Nogoceke, F.P.; Barcaro, I.M.; de Sousa, D.P.; Andreatini, R. Antimanic-like Effects of (R)-(−)-Carvone and (S)-(+) Carvone in Mice. *Neurosci. Lett.* 2016, 619, 43–48. [CrossRef] [PubMed]