Immunity to HIV in early life

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INTRODUCTION
In the dynamic developmental period from fetal life to adolescence, the human organism is more susceptible and vulnerable to chronic viral infections (1). HIV pathogenesis in early life is characterized by faster disease progression and shorter time to AIDS and death, compared to adults. The innate and adaptive arms of the immune system are more tolerogenic soon after birth and fail to control viral replication in infancy. Persistent viremia, reactivation of co-infections, and microbial translocation during chronic infection drive HIV pathogenesis through increased immune activation, resulting in immune dysregulation and functional immune exhaustion that exacerbate disease progression in a positive feedback loop (2–4) as visualized in Figure 1.

However, the recent report of a child who apparently achieved "functional cure" for more than 2 years following early therapeutic intervention (5) has raised major interest in the development of therapeutic strategies that could potentially be applied for cure even beyond the pediatric setting. In this review, we will discuss immunity to HIV in early life and how it affects pathogenesis and disease outcome, but also offers unique opportunities for prophylactic and therapeutic interventions in pediatric HIV infection.

PEDIATRIC HIV INFECTION: EPIDEMIOLOGY
Of the estimated 35 million people living with HIV today, 3.3 million are children (UNAIDS 2013 report). Administration of combination antiretroviral therapy (ART) can reduce risk of transmission to approximately 1–2% (6) and WHO now recommends the use of triple-drug ART regimens in all pregnant and breastfeeding women regardless of CD4 count, either lifelong (formerly referred to as option B+) or only during pregnancy and breastfeeding (formerly referred to as option B) (7). Despite increasing availability of ART, many women are diagnosed with HIV late in pregnancy and prevention of mother-to-child transmission (PMTCT) coverage rates remain suboptimal in many countries, resulting in 260,000 children becoming newly infected with HIV in 2012.

VERTICAL HIV TRANSMISSION
Without PMTCT, the overall risk of HIV transmission is up to 40% in sub-Saharan Africa; infection can occur in utero (5–10%), intrapartum (15%), or post-partum via breastfeeding (15%) (8). Interestingly, MTCT rates are much lower (<7%) in the natural hosts of SIV infection (such as sooty mangabeys), compared to humans (30–40%) or rhesus macaques (25–75%) (9).
During breastfeeding, no transmissions have been observed in sooty mangabeys, despite high plasma and breast milk viral loads (10). This has been attributed to paucity of CCR5+ CD4+ target cells for SIV infection (11), since the vast majority of vertically transmitted SIV/HIV-strains are R5-tropic and therefore depend on CCR5 as a co-receptor to infect CD4+ T-cells (12). Indeed, a recent study has shown very low to absent populations of CCR5+ CD4+ T-cells in the peripheral blood compartment and especially in the intestinal tract of sooty mangabeys compared to rhesus macaques (13). In human infants, CCR5+ CD4+ T-cells are virtually absent in cord blood (14). However, CCR5+ CD4+ T-cells are abundant in the intestinal mucosa and these cells are highly susceptible to HIV infection (15). The increased availability of target cells in the intestinal tract, in combination with the effectiveness of the placental barrier prior to labor, are both likely to contribute to the higher risk of intra- and post-partum, compared to in utero, transmission.

**CLINICAL COURSE**

Once infection is established, the course of disease progression in vertically infected infants is generally faster than in adults, with a rapid decline of CD4 cells and onset of recurrent infections, failure to thrive, and delayed neurodevelopment. In adult HIV infection, survival times decrease with increasing age at seroconversion, with a median time to AIDS and death of approximately a decade without ART (16). In contrast, mortality among vertically infected children in Africa exceeds 50% by the age of 2 years in the absence of ART (17). However, precise timing of transmission is an important determinant of survival: in a recent meta-analysis, 1-year mortality was 26% among children infected postnatally through breastfeeding, compared to 52% if infected perinatally (18). Another study that distinguished in utero, intrapartum and postnatal infection found median survival times from infection of 208, 380, and >500 days, respectively (19). Furthermore, lower mortality rates have been observed that the later HIV is acquired postnatally via breastfeeding (20). Interestingly, a cohort of children with hemophilia, who acquired HIV infection between the age of 5 and 14 years, had significantly longer AIDS-free survival times than adults (16). Long-term survival rates in vertically infected untreated children are estimated to be higher than initially appreciated with a probability of 20–30% surviving up to 10 years if infected perinatally and 16 years if infected via breastfeeding (20, 21).
Absolute CD4 count and viral load levels, which are well established clinical markers of disease progression in adults, have to be interpreted differently in children. The absolute CD4 count is three- to fourfold higher in the newborn compared to adults and gradually declines with age to reach adult levels after about 6 years or later (22, 23). The percentage of CD4+ T-cells is less variable with age and often used as an indicator of CD4 depletion in young children. HIV-1 plasma RNA levels are generally higher in vertically infected children than in adults and peak viremia (in the region of 5–7 log copies per milliliter) is observed around 3 months of age (24). Peak viremia is higher in children infected intrapartum than in utero (25) and lower in postnatally infected children, suggesting improved viral control by the more mature immune system (26, 27). Whereas in adult infection, a steady state viral set point is reached within several weeks of infection (28, 29), viremia persists at high levels, and only declines slowly until a quasi-set point is reached after several years in children who survive (30). This gradual control of viral replication has been associated with increasing maturation and development of the immune system (1).

INNATE IMMUNE SYSTEM

In the initial response to infections, the innate immune system performs two broad roles. On the one hand, it directly inactivates or attenuates invading pathogens, and on the other hand, it instructs the adaptive arm of the immune system to generate appropriate effector responses. Innate immune cells, such as macrophages, monocytes, and dendritic cells interact with pathogen-associated molecular patterns (PAMPs) through pattern recognition receptors (PRRs) that have evolved as substrate-specific receptors that can discriminate between self- and non-self and constantly sample the intra- and extracellular milieu [reviewed in Ref. (31)]. PRRs include the Toll-like receptors (TLRs), which are mainly located within the membranes of the cell surface and endosomes, and intracellular sensors such as RIG-1 like receptors (RLRs). HIV is principally detected by the innate immune system through the recognition of viral nucleic acids that are present in the cytosol of productively infected cells during the viral replication cycle and in the endosomes of phagocytosing cells. For example, cell-free virus that is phagocytosed by dendritic cells is recognized in the endosome by TLR-7 and TLR-9, which bind single-stranded RNA and DNA, respectively. Most PAMP/PRR interactions activate signaling pathways that converge in transcriptional activation of pro-inflammatory cytokines and type-1 interferons (IFNs). Type 1 IFNs bind to receptors of infected and neighboring cells and induce expression of IFN-stimulated genes (ISGs) that synergistically inhibit viral replication and spread (32).

The importance of these innate immune mechanisms in viral infections has been demonstrated in children with gene mutations in the TLR3 signaling pathway, who are prone to severe herpetic simplex encephalitis (33). Certain polymorphisms in the TLR-9 gene were associated with increased risk of mother-to-child transmission of HIV in one study (34), but further independent studies on innate immune signaling in pediatric HIV infection are required. Genetic polymorphisms of APOBEC3G, a host restriction factor that is induced by type-1 IFNs and directly inhibits replication of HIV, are associated with increased viral loads (35), lower CD4 counts (36), and more rapid disease progression in HIV-infected children (37).

Antigen-presenting innate immune cells such as dendritic cells play a key role not only in inducing pathways that directly inhibit viral replication but also in regulating the host adaptive immune response to HIV infection. Upon TLR stimulation, neonatal dendritic cells and monocytes are less polyfunctional and produce less type-1 IFNs than adult cells (38). However, it has been noted that neonatal innate immune cells do not have a generalized deficit in secretion of pro-inflammatory cytokines, but rather display distinct cytokine profiles to shape differential adaptive immune responses compared to adults. The most consistently reported difference in neonatal innate immune cell cytokine patterns is a relatively low production of Th1-polarizing cytokines, such as Interleukin-12 and IFN-gamma, in favor of Th2- and Th17-inducing Interleukin-10, Interleukin-6, and Interleukin-23 (38, 39) [reviewed in Ref. (40)]. This immunoregulatory strategy has likely evolved for two reasons: first, to dampen pro-inflammatory Th1 responses that could be damaging in utero and after birth when the newborn is exposed to an abundance of newly encountered antigens; and second, to ensure protection against extracellular bacterial and fungal pathogens, which can be rapidly fatal soon after birth (40). However, as a trade-off, the skewed Th1/Th2 ratio leaves the infant more susceptible to intracellular pathogens such as HIV [reviewed in Ref. (11)]. Interestingly, HIV-exposed but uninfected children show enhanced pro-inflammatory cytokine responses upon stimulation of innate immune cells compared to unexposed children, indicating that altered innate immune ontogeny could contribute to the increased vulnerability of this population to morbidity and poor growth (41).

The intrinsically low level of Th1 responses induced by neonatal innate immune cells is reinforced by the dysregulation and depletion of dendritic cells upon HIV infection [reviewed in Ref. (42)]. Most studies have been undertaken in chronically HIV-infected adults, but perinatally infected children also have reduced frequencies and functionality of dendritic cells and other innate immune cell populations including natural killer (NK) cells (43–45).

Natural killer cells are innate lymphocytes that play a critical role in the control of viral infections through interactions with other host immune cells via the secretion of chemokines and cytokines or by direct cellular cytotoxicity against infected target cells [reviewed in Ref. (46)]. NK cells recognize infected cells either through interaction with antibodies bound to the target cell surface in a process termed antibody-dependent cellular cytotoxicity (ADCC) or through interactions of an intricate network of inhibitory and activating cell surface receptors with ligands on infected cells. For example, NK cells are activated by reduced expression levels of MHC-class-I-molecules on the cell surface of HIV-infected cells via signaling of the inhibitory killer cell Ig-like receptors (KIRs), which is an important complementary mechanism to CD8 T-cell responses that depend on MHC-class-I-recognition [reviewed in Ref. (47)].

Frequencies of neonatal NK cells are similar to adults (48), but there are differences in expression patterns of inhibitory and activating cell surface receptors (49) and reduced cytotoxic activity of cord blood NK cells has been reported consistently (50, 51) [reviewed in Ref. (52)]. NK cells isolated from vertically
HIV-infected children have decreased cytolytic activity compared to uninfected children (53) and exhibit reduced ADCC compared to adult cells (54), but neonatal NK cells have been shown to be capable of suppressing replication of CCR5-tropic HIV in vitro through a non-cytotoxic chemokine-mediated mechanism (55). Total numbers of NK cells in vertically HIV-infected children decline with age (56) and are only partially reconstituted upon initiation of ART (45). Vertical HIV infection is also associated with increased NK cell activation, differentiation, and loss of perforin expression, which might impair NK cell function and compromise their role in protecting the organism against HIV, co-infections, and cancer (56).

DEVELOPING IMMUNE SYSTEM: IMMUNE TOLERANCE AND TREGS

The maternal and fetal immune systems are finely adapted toward increased immunological tolerance to avoid controlled damage to the fetus by allogeneic T-cells from the mother on the one hand and pathological reactions to maternal- and self-antigens by the fetus on the other (1). There is immune adaptation away from Th1 and toward Th2 responses during pregnancy (57), although the concept of pregnancy as a generalized state of immunosuppression is no longer supported (58, 59); for example, pregnant women appear to respond well to vaccines during pregnancy (60–62), and even mount anti-fetal CD4+ T-cell response without pregnancy loss (63). The hyporesponsiveness of the fetal adaptive immune system to antigens was previously interpreted as a functional deficit of effector cells, whereas currently the induction of immune tolerance is thought to be an active process that is largely mediated by regulatory T-cells (Tregs) [reviewed in Ref. (64)]. After depletion of CD4+ CD25+ FoxP3+ Tregs, fetal CD4 and CD8 T-cells have been shown to be highly responsive to stimulation ex vivo (65). In utero exposure to maternal alloantigens induces the development of tolerogenic Tregs that suppress activity of effector T-cells and persist into adulthood (66). The abundant population of Tregs that peaks in the second trimester and gradually declines to adult levels around birth (67) has been shown to originate from fetal hematopoietic stem cells (HSC); these differ from adult HSC based on functional and transcriptional analyses and give rise to a distinct lymphocyte lineage (68).

The role of Tregs in the pathogenesis of HIV infection remains controversial, but several studies have proposed a modulating function in HIV-specific immune responses, chronic immune activation, and inflammation [reviewed in (69)]. Most studies have reported a decrease in absolute numbers of Tregs in chronic HIV infection; however, as the frequency of Tregs is often reported as a percentage of CD4+ T-cells, there appears to be a selective expansion of different Treg subsets within the CD4 population during the CD4 decline that characterizes chronic infection (70). In HIV-infected children, one study found a positive correlation between the proportion of Tregs and HIV viral load, and a negative correlation with CD4 count suggesting a selective expansion of Tregs in pediatric HIV infection (71). Interestingly, in the same study these increased frequencies of Tregs also correlated with the proportion of activated CD8+ T-cells, suggesting the ineffectiveness of these cells to limit immune activation. However, another study in vertically HIV-infected children showed a depletion of Tregs compared to HIV-negative children when measured as proportion of total T-cells (CD3+ lymphocytes) that was associated with increased levels of immune activation (72). Another study found an association between altered frequencies of Treg subsets and autoantibody production in HIV-positive children highlighting the potential role of Tregs in immunoregulatory pathways that may be disrupted in HIV infection (73). In HIV-exposed but uninfected infants, a significant increase in HIV-specific CD4+ and CD8+ T-cell responses was observed after depletion of Tregs in cord blood mononuclear cells in vitro (73, 74) indicating a suppressive function in pathogenic-specific adaptive immunity.

ADAPTIVE IMMUNITY: CD8 T CELLS

HIV-specific CD8+ T cells are thought to play a key role in control of viral replication based on studies in humans (75, 76) and SIV-infected monkey models (77, 78). In adults, the rapid decline in viral load during acute infection from several million copies/ml to a viral load set point 3 logs lower is temporally associated with the appearance of HIV-specific CD8+ T-cells (75, 76, 79). Cytotoxic CD8+ T cells interact directly with HIV-infected target cells via the T-cell receptor that recognizes viral peptides presented on the cell surface by MHC-class-I molecules. The HLA-class-I type of an individual affects the rate of disease progression: certain HLA alleles such as HLA-B35 are associated with more rapid progression to AIDS, whereas protective alleles such as HLA-B57 and HLA-B27 are associated with viremic control and slow disease progression (80). One of the main mechanisms proposed for the protective effect of these alleles is their peptide binding specificity, allowing CD8+ T cells to target epitopes located particularly in the Gag region, in which an escape mutation that would evade the immune response is associated with a high cost in viral fitness (81, 82). Transmission of viral variants with an accumulation of costly escape mutations that reduce viral replicative capacity facilitates containment of HIV in the new host and reduces viral load and loss of CD4 cells (83, 84).

In pediatric HIV infection, the HLA type of both mother and child is of relevance for disease progression in the child. In mothers with protective HLA alleles, the virus accumulates immune escape mutations that are associated with a loss of viral fitness, thereby attenuating the transmitted virus, resulting in slower disease progression in the child (85). However, as the child inherits 50% of the HLA alleles from the mother, the virus can be partially pre-adapted to the shared alleles so that the recipient child cannot mount an effective immune response against the escaped epitopes and fails to contain viral replication (86). The beneficial effect of protective HLA alleles is therefore greatest if they are not shared between mother and child, and the child can employ them to mount effective immune responses (85). In a study of vertically HIV-infected children and adolescents, the protective allele HLA-B57 was overrepresented in the group of children with slow disease progression (87). In a recent genomic study of over 1000 vertically HIV-infected children, the protective HLA alleles HLA-B57, -B27, -B14, -Cw8, and -DRB1*10 were the SNPs most strongly associated with increased levels of immune activation (35).

The adaptive immune system starts to develop as early as 7–9 weeks of gestation when T-cell progenitor cells populate the thymus (88). In vertically HIV-infected children, HIV-specific
CD8+ T cell responses can be detected from birth (89, 90), but less frequently and at a lower magnitude than in older children or adults (90–92). However, these responses seem to be insufficient to control viremia, as there is typically no rapid decline in viral load in pediatric HIV infection as opposed to the decline in viral load after acute infection in adults [see above and Ref. (25)]. In infants, neither breadth nor magnitude of HIV-specific CD8 T-cell responses at 1 month of age correlate with viremic control or survival at 12 months (93), but Gag-specific CD8 T cell responses are higher in infants who survive to 1 year of age (94). The magnitude and breadth of Gag-specific CD8 T cell responses correlated negatively with viral load in one study (95), but not in another (96). Also, in a study of older children and adolescents who were categorized as progressors and non-progressors, there was no difference in magnitude and breadth of HIV-specific CD8 T cell responses between the two groups (87). However, magnitude and breadth of HIV-specific CD8 responses increase with age (90, 92, 96).

Broad Gag-specific CD8+ T cell responses have consistently been shown to be associated with reduced viral load (97, 98) in HIV-infected adults. But whereas Gag is the most immunogenic HIV-protein in adults, infants preferentially target epitopes in the more variable proteins Env and Nef (92, 95) with an increasing proportion of Gag-specific responses only later in life (95).

In addition to the specificity and magnitude, the effectiveness of a CD8 T cell response to control viral replication is also determined by its quality as measured by the ability to degranulate and produce different effector cytokines simultaneously (99, 100). The frequency of HIV-specific CD8 T cell that exhibit three different effector functions or more (CD107+, IL-2+, IFN-gamma+1) is reduced in children younger than 2 years but increases with age (95). In another study, the functional profile of HIV-specific CD8 T cell responses predicted the rate of subsequent disease progression in perinatally HIV-infected infants showing that more polyfunctional responses were associated with slower disease progression (101).

Other functional measures that have been described for effective CD8 T cell responses include the proliferative capacity of a T cell clone upon antigen stimulation in vitro (102). In children, the proliferative capacity of HIV-specific CD8 T cells has thus far not been studied in detail and published data are only available for CD4 T cells at present.

**ADAPTIVE IMMUNITY: CD4 T CELLS**

To maintain fully functional CD8+ T cell responses, the help of CD4+ T cells with the same antigen-specificity is required (103, 104). In the mouse model, depletion of CD4 T helper cells during acute infection results in ineffective CD8 T cell memory responses (105, 106). CD4+ T cells are the major target for HIV and massive depletion of CD4+ T cells results during acute infection in adults (107, 108), with ongoing preferential infection of activated and HIV-specific CD4 T cells in chronic infection (109). This depletion of HIV-specific CD4+ T helper cells has been proposed as a main mechanism for failure to control HIV successfully, given their central role in orchestrating diverse cellular and humoral immune functions (110).

In addition to providing help to CD8 T cells (111), HIV-specific CD4 T cells have been shown to have intrinsic direct cytotoxic activity against infected cells (112–114). Increased breadth, especially of Gag-specific CD4 T cell responses, is inversely correlated to viral load in chronically infected adults; elite or viremic controllers show a higher ratio of Gag- vs. Env-specific CD4 responses (115). Another study in a large cohort of ART-naïve HIV-1-clade-C infected adults also found that the magnitude of Gag-specific CD4 T cell responses was associated with higher CD4 counts and lower viral loads (116).

As discussed above in relation to innate immunity, there is a selective impairment of CD4 T cell responses in early childhood with a shift from Th1 toward Th2 type responses (117, 118) and poor generation of persistent Th1 memory responses (119). This has been attributed to low production of Interleukin-12 (120), which is required for Th1-polarization and maintenance of Th1 effector functions (121, 122), but does not reach adult levels until adolescence. The bias toward Th2 responses in early life is reinforced by the increased capacity of neonatal monocytes and dendritic cells to produce Interleukin-10, an immunomodulatory cytokine that induces a shift of T cell induction toward Th2 and Th17 profiles (38).

During fetal development in non-pathologic sterile conditions, the T cell compartment is considered to be mostly naive and tolerant, although a recent study highlighted the existence of CD4 T cell effector memory subsets with Th1, Th2, and Th17 functional profiles in cord blood of healthy newborn infants (123). HIV-specific CD4 T cell responses can be primed in utero (124), but are detected in infants at significantly lower frequencies than in chronically infected adults and increase with age (92, 95). These low levels of HIV-specific Th1 responses in early life coincide with the Th1/Th2 bias and the expanded population of suppressive regulatory T cells as described above. In the macaque model, SIV-specific CD4 T cell responses are suppressed in infants by regulatory T cells that show greater in vitro suppressive activity and are present at higher frequencies than in adults and have been associated with impaired immune control (125).

Gag-specific CD4 T cell responses were detectable at higher frequencies at 3 months of age in vertically HIV-infected children who survived to 12 months, compared to those who died, and Gag-specific responses at 3 and 6 months of age were negatively correlated with VL (94). Another study detected Gag-specific CD4 responses only in a minority of older vertically infected children (median age 9.9 years). Those with detectable responses had lower viral loads than non-responders and the magnitude of the response was inversely correlated with viral load; no association was observed between CD8 T cell responses and viral load (96).

The functional capability of CD4 T cells to produce IL-2 is causally linked to the proliferative capacity of both CD4 and CD8 T cell subsets, a parameter consistently correlated with immune control and slow disease progression [reviewed in Ref. (126)]. HIV-specific proliferative responses of CD4 T cells are detected only in a fraction of viremic chronically infected adults and are inversely correlated with VL if present (127, 128); IL-2 producing HIV-specific CD4 responses are associated with slow disease progression in adults (129, 130). Interleukin-2 is also important for the differentiation, activation, and proliferation of NK cells, Th1 and Th2 CD4 T cells, B cells, Tregs, and memory CD8 T cells,
highlighting the key immunoregulatory role of these CD4 T cells in the immune response (131).

The proliferative capacity of HIV-specific CD4 T cells is selectively impaired in vertically infected children with uncontrolled viral replication (124, 132), but, in contrast to adult infection, can be rescued with administration of ART (133). The magnitude of the response increases with age and is strongest in children maintaining viral suppression on ART (133). Interestingly, proliferative and IL-2 producing CD4 responses were also detected in HIV-exposed uninfected infants but were mostly absent in infected children (134). Polyfunctional HIV-specific CD4 T cell responses, especially Interleukin-2 producing cells, were less frequently detected in younger children, but if present, they were associated with low viral loads and remarkably slow disease progression (95). However, it remains unclear whether these responses actually mediate viral control or are simply a proxy for undisrupted immunoregulatory networks.

**IMMUNE EXHAUSTION**

Chronic HIV infection and persistent antigenic stimulation lead to more terminally differentiated T cell populations with loss of effector functions and proliferative capacity upon antigenic stimulation [reviewed in Ref. (3)]. This state of “Immune Exhaustion” is characterized by the upregulation of inhibitory co-receptors on CD4 and CD8 T cells such as PD-1 (135, 136), Tim-3 (137), CD160 (138), LAG-3 (139), and CTLA-4 (140) that correlate with markers of disease progression in chronic HIV infection (136, 137, 139–141). As immune functions can be partially restored upon antibody-mediated blockade of these receptors (136, 137, 139, 140), they present an attractive target for immunotherapeutic intervention in HIV infection, although immune-related adverse events remain a concern (142, 143).

In pediatric subjects, few studies have been conducted to elucidate the role of immune exhaustion in chronic HIV infection. One study found an expansion of Tim-3+ CD4 and CD8 T cells in vertically HIV-infected adolescents and a correlation of Tim-3 expression on CD8 T cells with viral load (144). In the same study, PD-1 expression only correlated with the frequency of activated CD8 T cells. This is consistent with another report that described a correlation of PD-1 expression on CD8 T cells with immune activation markers and the magnitude of HIV-specific CD8 T cell responses, but not with viral load, suggesting that immune exhaustion is driven more by chronic immune activation than directly by viral replication (96).

**IMMUNE ACTIVATION**

Persistent systemic immune activation plays a key role in the pathogenesis of HIV infection and is regarded as the driving force in generalized immune dysregulation, chronic inflammation (145, 146), depletion of CD4 T cells (147, 148), and progression to AIDS (149) [reviewed in (4)]. Recent studies suggest that much of the CD4 depletion is caused by a process called “pyroptosis,” whereby abortive HIV infection of CD4+ T cells causes a highly inflammatory form of cell death, which drives accumulation of further CD4 cells and an ongoing cycle of inflammation and cell death (150, 151). T cell immune activation, as commonly measured by expression of activation markers such as HLA-DR and CD38 on CD4 and CD8 T cells, is a stronger predictor of disease progression than viral load in adults (152). Similarly, in vertically HIV-infected infants, the subsequent rate of disease progression can be predicted by T cell activation levels at 1–2 months of age (153). Both CD4 and CD8 immune activation show a strong inverse correlation with CD4 percentage, but interestingly no correlation is observed between immune activation and viral load in children (72, 154). In vertically HIV-infected children who maintain high CD4 counts without ART (often referred to as pediatric slow progressors), immune activation levels are strikingly low despite high viral loads (Muenchhoff et al., unpublished data). This resembles the phenotype of non-pathogenic infection in the natural hosts of SIV, such as African Green Monkeys and Sooty Mangabeys (155), in whom CD4 counts are maintained despite high levels of viremia, due to attenuated immune activation (156). In acute infection, sooty mangabeys also show high levels of immune activation and a robust innate immune response that is associated with a generalized upregulation of Interferon Stimulated Genes (ISGs, see innate immunity), but this initial response is rapidly resolved, suggesting the involvement of active immunoregulatory mechanisms that dampen detrimental excessive immune activation (157).

After initiation of ART, immune activation levels in children and adults decrease (154) but usually remain elevated compared to HIV-negative controls and this residual immune activation despite ART is associated with incomplete immune reconstitution, increased non-AIDS comorbidities, and mortality (158–160) [reviewed in Ref. (161)]. Persistent immune activation is associated with increased levels of pro-inflammatory cytokines and elevated coagulation markers that have been linked to cardiovascular disease and cancer in adults [reviewed in Ref. (4, 162)], emphasizing a potential role for adjunctive immunomodulatory interventions together with ART to reduce residual immune activation in people living with HIV.

The cause of systemic immune activation in HIV infection has not yet been fully elucidated but appears to arise from a complex interplay between HIV and the host involving several molecular and cellular mechanisms including innate and adaptive immune responses that create a pro-inflammatory milieu, fibrosis of lymphoid tissue, uncontrolled co-infections, and breach of the intestinal barrier resulting in translocation of microbial products into the systemic circulation [reviewed in Ref. (161)].

**MICROBIAL TRANSLLOCATION**

From early stages of HIV infection, the gastrointestinal tract is a target for HIV-induced pathology, with severe depletion of mucosal CD4 T cells (in particular Th17 cells), dendritic cells, and innate lymphoid cells (107, 163, 164) disrupting mucosal immunity and the intestinal epithelial architecture [reviewed in (2, 165)]. This breach in the mucosal barrier results in translocation of microbial products from the gut into the systemic circulation leading to immune activation (166). A critical difference between non-pathogenic SIV infection (sooty mangabeys, African green monkeys) and pathogenic infection (rhesus macaques, pig-tailed macaques) is in preservation of CD4+ Th17 cells in the sooty mangabeys and African Green monkeys (167). Markers that are used to measure microbial translocation include direct microbial products like lipopolysaccharide (LPS), a component of the
Gram-negative bacterial cell wall, and conserved bacterial ribosomal 16sRNA, or indirect markers such as sCD14 as a soluble marker of monocyte activation (168).

The immaturity of the gut in early life, characterized by increased epithelial permeability (169) and the evolution of the gut microbiota following bacterial and viral colonization (170) might influence the degree of microbial translocation in pediatric populations. Plasma levels of LPS in uninfected children initially increase after birth to reach a plateau at about 6 months of age (171) and then decrease again after the age of 2 years (172).

In vertically HIV-infected infants, levels of microbial translocation are elevated compared to uninfected controls and persist at a higher level after initiation of ART (173). Persistent microbial translocation correlates with cellular and soluble markers of immune activation in children on ART suggesting an important role in immune reconstitution (174). In a recent study in untreated HIV-infected children, microbial translocation was strongly correlated with monocyte and T cell activation and PD-1 expression levels on CD4 and CD8 T cells demonstrating the connection between microbial translocation, immune activation, and immune exhaustion (175).

ADAPTIVE IMMUNITY: B CELLS AND ANTIBODIES

HIV-specific antibodies have been in the focus of intense research for several decades in the quest for a prophylactic HIV vaccine. Recently, hope has been fueled by the results of the RV144 vaccine trial, which showed modest vaccine efficacy (176) and identified antibodies against the V1V2 HIV-envelope region as a correlate of protection in vaccinees (177). Antibodies can mediate antiviral activities by binding either to free virus or to viral particles on infected cells. The Fc-region of the bond antibody can mediate phagocytosis of the virus or infected cells, activation of innate immune cells resulting in ADCC against infected cells (see innate immunity), or activation of the complement system leading to opsonization and lysis of the virus or infected cells [reviewed in Ref. (178)]. An important function of antibodies especially in regard to sterilizing immunity is the ability to neutralize the virus by binding to the viral surface and inhibit cellular infection. The surface of HIV consists of the gp41 trans-membrane protein and the heavily glycosylated gp120 surface protein, which are both highly variable between different viral variants. Therefore, antibodies elicited by an effective protective vaccine would need to be broadly neutralizing across different viral subtypes (179).

These broadly neutralizing antibodies (bNABs) have been shown to occur naturally in a minority (20%) of HIV-infected subjects over the course of several years post-infection (180–182) during a process of intense somatic hypermutation (183). A recent study identified several young children with HIV-specific antibodies that had surprisingly high neutralizing breadth (184). Interestingly, the proportion of children with high neutralization scores was greater than in large-scale cohort studies of chronically HIV-infected adults and high levels of neutralizing breadth were already reached at an age of ~2 years, suggesting accelerated affinity maturation. These findings were especially surprising as it had previously been shown that infants produce lower antibody titers with less diversified somatic mutation upon infection and vaccination shortly after birth [reviewed in Ref. (185)]. As higher set point, viral load levels are associated with increased emergence of bNABs in adults (186) and in these young children (184), it has been hypothesized that the high antigenic load in pediatric HIV infection boosts development of NAB breadth. Also other factors like the shift toward Th2 responses in early life (as discussed above) could promote B-cell function and somatic hypermutation. Differential proportions of IgG-subclasses in children with a preponderance of IgG1 and IgG3 (185) that have higher neutralizing potency (187, 188) could also contribute to this finding. CD4+ T follicular helper (Tfh) cells are thought to be crucial in providing help to B cells in the germinal center to support antibody maturation [reviewed in Ref. (110)], but there have been no studies of Tfh cells in HIV-infected children to date. Further investigation in this field could yield important findings for vaccine development.

MISSISSIPPI CHILD/RESERVOIR/CURE

The case report of an in utero HIV-infected child who appeared to have been “functionally cured” after early initiation of ART was received with much excitement. The child initiated ART within 30 h of life and stayed on ART until 18 months of age when ART was discontinued by the mother. In subsequent visits to the clinic half a year later and in the successive follow-up period, viral load remained undetectable in the absence of ART and no signs of ongoing viral replication or HIV-specific T-cell or antibody responses were detected (5), hence the child was labeled “functionally cured.” However, after more than 2 years of follow up with undetectable HIV-RNA levels after treatment cessation, there was a recent report of rebound viremia in this child.

The main obstacle to HIV eradication is a pool of long-lasting, treatment-resistant viral reservoirs that only decay slowly on ART and are a source for viral rebound after treatment cessation [reviewed in Ref. (189, 190)]. Different cell types and body compartments are thought to contribute to the viral reservoir, but major sources of replication-competent pro-viral DNA during effective ART are resting central and transitional memory CD4+ T cells (191). In this regard, children might be partially protected against the early establishment of viral reservoirs in these cell populations, because neonates have much lower frequencies of CD4 T cells of central memory phenotype than adults (22, 23).

The frequency of latently infected CD4 T cells in children strongly depends on early initiation of ART and the time to achieve virologic suppression (192). A cross-sectional study found low to undetectable levels of integrated pro-viral DNA in CD4 cells of children starting treatment before 6 months of age after more than 3 years of follow-up (193). A recent longitudinal study compared perinatally infected youth initiating treatment before or after 3 months of age (194). In contrast to the late treatment group, all children starting ART before 3 months had undetectable HIV-RNA plasma levels using highly sensitive assays but integrated pro-viral DNA could be detected by PCR in all subjects even after follow-up periods of up to 17 years. In the early-treated group, transitional memory CD4 T cells had a larger contribution to the pool of infected cells than longer lasting central memory CD4 T cells. These findings are consistent with a study of adult post-treatment controllers who initiated ART during acute HIV infection (195), indicating that early treatment can prevent seeding
of long-lived cellular reservoirs. It has to be noted that only a fraction of integrated viral DNA codes for replication-competent virus (196) and so the standard assay to measure the size of the functional latent viral reservoir at present is a limiting dilution viral outgrowth assay (197, 198). Using this assay, only one of the early-treated children in this study had inducible replication-competent virus, as opposed to all children receiving ART at a later stage, although limited sample availability in pediatric studies reduces the sensitivity of these assays. Caution also has to be taken as rebound viremia is observed even in patients with extraordinarily low viral burden in the peripheral blood compartment (199) and a large proportion of the viral reservoir is actually located within the gut and other tissues that are not accessed in most studies (200). Reliable biomarkers to predict successful drug-free remission upon treatment cessation are therefore needed.

Apart from timing of ART initiation, the size and constitution of the viral reservoir is affected by host immunological factors including persistent immune activation that fuels replenishment of the reservoir during ART [reviewed in Ref. (161)] and effective immune responses that can reduce the pool of infected memory cells (201, 202). To achieve a cure of HIV in a broader group of patients will therefore most likely require a multifaceted approach that could include induction of potent HIV-specific immunity prior to reversion of HIV-latency in combination with immunomodulatory interventions.

CONCLUSION

In summary, immunity to HIV in early life differs from that in adulthood not only in quantity but also in quality. This results from the adaptations of the immune system to the abundance of antigenic stimulation and pathogen exposure in utero and after birth. The initial ontogeny of the immune system is characterized by a more tolerogenic state of innate and adaptive immunity that prevents potentially harmful autoimmune and inflammatory responses, and is geared toward preferential protection against extracellular pathogens. As a consequence, vertically acquired HIV infection is poorly controlled and marked by rapid progression to extracellular pathogens. As a consequence, vertically acquired HIV infection is poorly controlled and marked by rapid progression to extracellular pathogens. As a consequence, vertically acquired HIV infection is poorly controlled and marked by rapid progression to extracellular pathogens. 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As a consequence, vertically acquired HIV infection is poorly controlled and marked by rapid progression to extracellular pathogens. As a consequence, vertically acquired HIV infection is poorly controlled and marked by rapid progression to extracellular pathogens. As a consequence, verte...
23. Schatorje EE, Gemen EF, Driessen GJ, Leunenik I, van Hout RW, de Vries E. Paediatric reference values for the peripheral T cell compartment. Scand J Immunol (2012) 75(4):436–44. doi:10.1111/j.1365-3082.2012.02671.x

24. European Collaborative S. Level and pattern of HIV-1 RNA viral load over age: differences between girls and boys? AIDS (2002) 16(1):97–104. doi:10.1097/00001731-200201040-00012

25. Mphatswe W, Blanckenberg N, Tudor-Williams G, Prendergast A, Thobakgale C, Mkhwanazi N, et al. High frequency of rapid immunological progression in African infants infected in the era of perinatal HIV prophylaxis. AIDS (2007) 21(12):1253–61. doi:10.1097/QAD.0b013e32813beec2

26. Obimbo EM, Wamalwa D, Richardson B, Mbori-Ngacha D, Overbaugh J, Emery S, et al. Proliferative HIV-1 in Kenya: pattern and correlates of viral load and association with mortality. J Acquir Immune Defic Syndr (2009) 51(2):219–25. doi:10.1097/QAI.0b013e31819c61d8

27. Biggar RJ, Ianes M, Pilon R, Miotto P, Taha TE, Broadhead R, et al. Virus levels in untreated African infants infected with human immunodeficiency virus type 1. J Infect Dis (1999) 180(6):1383–48. doi:10.1086/315339

28. Lyles RH, Munoz A, Yamashita TE, Bazmi H, Detels R, Rinaldo CR, et al. Natural history of human immunodeficiency virus type 1 viremia after seroconversion and proximal to AIDS in a large cohort of homosexual men. Multicenter AIDS Cohort Study. J Infect Dis (2000) 181(3):872–80. doi:10.1086/313182

29. Saathoff E, Pritsch M, Geldmacher C, Hofmann O, Koehler RN, Maboko L, et al. Viral and host factors associated with the HIV-1 viral load setpoint in adults from Mbeya Region, Tanzania. J Acquir Immune Defic Syndr (2010) 54(3):324–30. doi:10.1097/QAI.0b013e3181e180b8a

30. McIntosh K, Shevitz A, Zakun D, Kornegay J, Nogues N, et al. Age- and time-related changes in extracellular viral load in children vertically infected by human immunodeficiency virus. Pediatr Infect Dis J (1996) 15(12):1087–91. doi:10.1097/00006455-199612000-00006

31. Rustagi A, Gale M Jr. Innate antiviral immune signaling, viral evasion and association with mortality. J Acquir Immune Defic Syndr (2002) 28:217–51. doi:10.1177/1087425402280000010

32. Stetson DB, Medzhitov R. Type I interferons in host defense. Annu Rev Immunol (2002) 20:163–94. doi:10.1146/annurev-immunol-20.092501.134942

33. Zhang SY, Jouanguy E, Ugolini S, Smahi A, Elain G, Romero P, et al. TLR3 deficiency in patients with herpes simplex encephalitis. J Immunol (2007) 179(5):3362–4. doi:10.4049/jimmunol.179.5.3362

34. Ricci E, Malacrida S, Zanchetta M, Mosconi I, Montagna M, Giaquinto C, et al. Defective dendritic cell response to TLR7/8 agonists in perinatally HIV-1-infected children. Pathogens Dis (2013) 69(3):184–93. doi:10.1111/2049-632X.12067

35. Qin M, Brummel S, Singh KK, Sibilly N, Laine L, et al. Skewed pattern of Tolt-like receptor 4-mediated cytokine production in interferon- alpha-treated children. J Innate Immun (2009) 60:111–17. doi:10.1159/000323934

36. Levy O, Innate immunity of the newborn: basic mechanisms and clinical correlates. Nat Rev Immunol (2007) 7(5):379–90. doi:10.1038/nri2075

37. Selvaraj A, Pilakka-Kanthikeel S, Bhavnani PK, Hanna LE, Pahwa S, Swaminathan S. Defective dendritic cell response to Toll-like receptor 7/8 agonists in perinatally HIV-infected children. Pathogens Dis (2013) 69(3):184–93. doi:10.1111/2049-632X.12067

38. Kollmann TR, Crabtree J, Rein-Weston A, Blimkie D, Thommai F, Wang XY, et al. Innate immunity of the newborn: basic mechanisms and clinical correlates. J Innate Immun (2009) 11:1750–60. doi:10.1097/QAI.0b013e31819e4843

39. Belderbos ME, van Bleek GM, Levy O, Blanken MO, Houben ML, Schuijff L, et al. Altered innate immune development in HIV-exposed uninfected infants. J Acquir Immune Defic Syndr (2008) 49(4):348–57. doi:10.1097/QAI.0b013e31818c16ff

40. Levy O. Innate immunity of the newborn: basic mechanisms and clinical cor-

41. Reikie BA, Adams RC, Leiglodwicz A, Ho K, Naidoo S, Ruck CE, et al. Altering innate immune development in HIV-exposed uninfected infants. J Acquir Immune Defic Syndr (2011) 56(3):245–55. doi:10.1097/QAI.0b013e318244ed20
61. Ohfuji S, Fukushima W, Deguchi M, Kawabata K, Yoshida H, Hatayama H, et al. Immunogenicity of a monovalent 2009 influenza A (H1N1) vaccine among pregnant women: lowered antibody response by prior seasonal vaccination. *J Infect Dis* (2011) 203(9):1301–8. doi:10.1093/infdis/jir026

62. Healy CM. Vaccines in pregnant women and research initiatives. *Clin Obstet Gynecol* (2012) 55(2):474–86. doi:10.1097/GRF.B013e31824E3acab

63. Lissauer D, Piper K, Goodyear O, Kilby MD, Moss PA. Fetal specific CD8+ cytotoxic T cell responses develop during normal human pregnancy and exhibit broad functional capacity. *J Immunol* (2012) 189(2):1072–80. doi:10.4049/jimmunol.1200544

64. Mold JE, McCane JM. At the crossroads between tolerance and aggression: revisiting the “layered immune system” hypothesis. *Chimerism* (2011) 2(2):35–41. doi:10.4161/chim.2.2.16329

65. Michaelsson J, Mold JE, McCane JM, Nixon DF. Regulation of T cell responses in the developing human fetus. *J Immunol* (2006) 176(10):5741–8. doi:10.4049/jimmunol.176.10.5741

66. Mold JE, Michaelsson J, Burt TD, Muench MO, Beckerman KP, Busch MP, et al. Maternal alloantigens promote the development of tolerogenic fetal regulatory T cells in utero. *Science* (2008) 322(5907):1562–5. doi:10.1126/science.1164511

67. Takahata Y, Nomura A, Takahara H, Ogga S, Furuno K, Hikino S, et al. CD25+CD4+ T cells in human cord blood: an immunoregulatory subset with naive phenotype and specific expression of forkhead box p3 (Foxp3) gene. *Exp Hematol* (2004) 32(7):622–9. doi:10.1016/j.exphem.2004.03.012

68. Mold JE, Venkatasaubramanyam S, Burt TD, Michaelsson J, Rivera JM, Galkina SA, et al. Fetal and adult hematopoietic stem cells give rise to distinct T cell lineages in humans. *Science* (2010) 330(6081):1695–9. doi:10.1126/science.1198659

69. Simonetta F, Bourgeois C. CD4+FOXP3+ regulatory T-cell subsets in human immunodeficiency virus type 1 infection. *Clin Exp Immunol* (2012) 176(3):373–80. doi:10.1111/j.1365-2249.2011.04383.x

70. Chevalier MF, Weiss L. The split personality of regulatory T cells in HIV infection. *BLOOD* (2013) 121(1):29–37. doi:10.1182/blood-2012-07-497555

71. Freguia R, Gianesin K, Mosconi J, Zanchetta M, Carmona F, Rampon O, et al. Regulatory T cells and chronic immune activation in human immunodeficiency virus 1 (HIV-1)-infected children. *Clin Exp Immunol* (2011) 164(3):373–80. doi:10.1111/j.1365-2249.2011.04383.x

72. Prendergast A, O’Callaghan M, Menson E, Hamadache D, Walters S, Klein N, et al. Factors influencing T cell activation and programmed death 1 expression in HIV-infected children. *AIDS Res Hum Retroviruses* (2012) 28(5):655–62. doi:10.1089/AID.2011.0081

73. Arguello RJ, Balbaryski J, Barboni G, Candi M, Gaddi E, Laucella S. Altered CD4+ T cell frequency and phenotype of CD4+ forkhead box protein 3+ T cells and CD8+ T cells in utero. *Front Immunol* (2012) 3:35–41. doi:10.3389/fimmu.2013.00215

74. Chevalier MF, Weiss L. The split personality of regulatory T cells in HIV infection. *Blood* (2013) 121(1):29–37. doi:10.1182/blood-2012-07-497555

75. Schmitz JE, Kuroda MJ, Santra S, Sasseville VG, Simon MA, Lifton MA, et al. Escape from the dominant HLA-B27-restricted cytotoxic T lymphocyte response in Gag is associated with a dramatic reduction in human immunodeficiency virus type 1 replication. *J Virol* (2007) 81(22):12382–93. doi:10.1128/JVI.01543-07

76. Prince JL, Clabonthe DR, Carlson JM, Schaefer M, Yu T, Lakh S, et al. Role of transmitted Gag CTL polymorphisms in defining ep recap capacity and early HIV-1 pathogenesis. *PLoS Pathog* (2012) 8(11):e1003041. doi:10.1371/journal.ppat.1003041

77. Goupret FA, Lumw M, Farmer P, Matthews P, Prendergast A, Carlson JM, et al. Transmission of HIV-1 Gag immune escape mutations is associated with reduced viral load in linked recipients. *J Exp Med* (2008) 205(5):1309–17. doi:10.1084/jem.20072457

78. Schneidewind A, Brockman MA, Yang R, Adam RI, Li B, Le Gall S, et al. Escape from the dominant HLA-B27-restricted cytotoxic T lymphocyte response in Gag is associated with a dramatic reduction in human immunodeficiency virus type 1 replication. *J Virol* (2007) 81(22):12382–93. doi:10.1128/JVI.01543-07

79. Rodriguez B, Sethi AK, Cheruvu VK, Mackay W, Busch RJ, Kataba M, et al. Predictive value of plasma HIV RNA level on rate of CD4 T cell decline in untreated HIV infection. *JAMA* (2006) 296(12):1498–506. doi:10.1001/jama.2006.1498

80. Goulder PJ, Walker BD. HIV and HLA class I: an evolving relationship. *Immunity* (2012) 37(3):426–40. doi:10.1016/j.immuni.2012.09.005
98. Goldmacher C, Currier JR, Herrmann EP, Haule A, Kuta E, McCutchan F, et al. CD8+ T-cell recognition of multiple epitopes within specific Gag regions is associated with maintenance of a low steady-state viremia in human immunodeficiency virus type 1-seropositive patients. J Virol (2007) 81(5):2440–8. doi:10.1128/JVI.01847–06

99. Akinsiku OT, Bansal A, Sabba S, Heath SL, Goepfert PA. Interleukin-2 production by polyfunctional HIV-1-specific CD8+ T cells is associated with enhanced viral suppression. J Acquir Immune Defic Syndr (2011) 58(2):132–40. doi:10.1097/QAI.0b013e318224d2e9

100. Betts MR, Nason MC, West SM, De Rosa SC, Migueles SA, Abraham J, et al. HIV nonprogressors preferentially maintain highly functional HIV-specific CD8+ T cells. Blood (2006) 107(12):4781–9. doi:10.1182/blood-2005-12-18180

101. Thobakgale CF, Streeck H, Mkhwanazi N, Mncube Z, Maphumulo L, Chonco T, et al. HIV preferentially infects HIV-specific CD4+ T cells. Science (2003) 300(5645):659–62. doi:10.1126/science.1088774

102. Grakoui A, Showery NH, Woolard DJ, Han JH, Hanson HL, Ghayeb J, et al. HCV persistence and immune evasion in the absence of memory T cell help. Science (2003) 302(5654):659–62. doi:10.1126/science.1088774

103. Lichterfeld M, Kaufmann DE, Yu XG, Mui SK, Addo MM, Johnston MN, et al. Loss of HIV-1-specific CD8+ T cell proliferation after acute HIV-1 infection and restoration by vaccine-induced HIV-1-specific CD4+ T cells. J Exp Med (2004) 200(6):791–12. doi:10.1084/jem.20041270

104. Shedlock DJ, Shen H. Requirement for CD4+ T cell help in generating functional CD8+ T cell memory. Science (2003) 300(5617):337–9. doi:10.1126/science.1083117

105. Sun JC, Bevan MJ. Defective CD8+ T cell memory following acute infection with out CD4+ T cell help. Science (2003) 300(5617):339–42. doi:10.1126/science.1083117

106. Guadalupe M, Reay E, Sankaran S, Prindiville T, Flamm J, McNeil A, et al. HIV-1-specific interleukin-21+ CD4+ T cell responses contribute to durable control of HIV infection and loss of memory CD4+ T cells in multiple tissues during acute infection. J Virol (2014) 88(7):3733–41. doi:10.1128/JVI.02030-10

107. Norris PJ, Moffett HF, Yang OO, Kaufmann DE, Clark MJ, Addo MM, et al. Beyond help: direct effector functions of human immunodeficiency virus type 1-specific CD4+ T cells. J Virol (2004) 78(16):8844–51. doi:10.1128/JVI.18.78.8844-8851.2004

108. Sohoghoan DZ, D’Souza MP, Littman DR, Crotty S. HIV-1 infection predicts disease outcome. J Virol (2003) 77(11):6376–85. doi:10.1128/JVI.77.11.6376-6385.2003

109. Sohoghoan DZ, D’Souza MP, Littman DR, Crotty S. HIV-1 infection predicts disease outcome. J Virol (2003) 77(11):6376–85. doi:10.1128/JVI.77.11.6376-6385.2003

110. Streeck H, D’Souza MP, Littman DR, Crotty S. Harnessing CD4(+) T cell function in pediatric HIV-1 infection. J Immunol (2003) 170(1):4730–7. doi:10.4049/jimmunol.170.1.4730

111. Ramduth D, Byu BA, Goerav P. The generation of Th memory in neonates versus adults: prolonged primary Th2 effector function and impaired development of Th1 memory effector function in murine neonates. J Immunol (2001) 166(2):918–25. doi:10.4049/jimmunol.166.2.918

112. Upham JW, Lee PT, Holt BJ, Heathon T, Prescott SL, Sharp MJ, et al. Development of interleukin-12-producing capacity throughout childhood. Infect Immun (2002) 70(12):6583–8. doi:10.1128/IAI.70.12.6583-6588.2002

113. Cleary AM, Tu W, Enright A, Giuffon T, Dweal-Malefyt R, Gutierréz K, et al. Impaired accumulation and function of memory CD4 T cells in human IL-12 receptor beta 1 deficiency. J Immunol (2003) 170(1):597–603. doi:10.4049/jimmunol.170.1.597

114. Soghoian DZ, Jessen H, Flanders M, Sierra-Davidson K, Cutler S, et al. HIV-1-specific CD4+ T cell responses in acutely infected infants. AIDS Res Hum Retroviruses (2008) 24(2):265–70. doi:10.1089/aim.2007.0096

115. Hartigan-O’Connor DJ, Abel K, McCune JM. Suppression of SIV-specific CD4+ T cells by infant but not adult macaque regulatory T cells: implications for SIV disease progression. J Med Virol (2007) 80(1):628–31. doi:10.1002/jmv.20768

116. Imami N, Westrop SJ, Grageda N, Herasmitshuk AL. Long-term non-progression and broad HIV-1-specific proliferative T-cell responses. Front Immunol (2013) 4:58. doi:10.3389/fimmu.2013.00058

117. Barrios C, Brandt C, Berney M, Lambeth PF, Siegrist CA. Partial correction in utero. J Immunol (2014) 192(11):5786–83. doi:10.4049/jimmunol.170.1.597

118. Prescott SL, Macaubas C, Holt BJ, Smallacombe TB, Loh R, Py JD, et al. Transplacental priming of the human immune system to environmental allergens: universal skewing of initial T cell responses toward the Th2 cytokine profile. J Immunol (1998) 160(10):4730–7. doi:10.4049/jimmunol.160.10.4730

119. Kuhn L, Meddows-Taylor S, Gray G, Tiemessen C. Human immunodeficiency virus (HIV)-specific cellular immune responses in newborns exposed to HIV in utero. Clin Infect Dis (2002) 34(2):267–76. doi:10.1086/338135
173. Papasavvas E, Arzoni L, Foulkes A, Violari A, Cotton MF, Pistilli M, et al. Increased microbial translocation in <780 days old perinatally human immunodeficiency virus-positive infants as compared with human immunodeficiency virus-exposed uninfected infants of similar age. Pediatr Infect Dis J (2011) 30(10):877–82. doi:10.1097/INF.0b013e318244e41e

174. Pilakka-Kanthikeel S, Huang S, Fenton T, Borkowsky W, Cunningham CK, Pahwa S. Increased gut microbial translocation in HIV-infected children persists in virologic responders and virologic failures after antiretroviral therapy. Pediatr Infect Dis J (2012) 31(6):583–91. doi:10.1097/INF.0b013e31823a0d05

175. Pilakka-Kanthikeel S, Kris A, Selvaraj A, Swaminathan S, Pahwa S. Immune activation is associated with increased gut microbial translocation in treatment-naive, HIV-infected children in a resource-limited setting. J Acquir Immune Defic Syndr (2014) 66(1):16–24. doi:10.1097/QAI.0000000000000096

176. Rerks-Ngarm S, Pitisuttithum P, Nitayaphan S, Kaewkungwal J, Chiu J, Paris R, et al. Vaccination with A1VAC and AIDSVAX to prevent HIV-1 infection in Thailand. N Engl J Med (2009) 361(23):2209–20. doi:10.1056/NEJMoa0908492

177. Haynes BF, Gilbert PB, McElrath MJ, Zolla-Pazner S, Tomaras GD, Alam SM, et al. Immune correlates of an HIV-1 vaccine efficacy trial. N Engl J Med (2012) 366(14):1275–86. doi:10.1056/NEJMoa1113425

178. Burton DR. Antibodies, viruses and vaccines. Nat Rev Immunol (2002) 2(9):796–803. doi:10.1038/nri8991

179. Walker BD, Burton DR. Toward an AIDS vaccine. Science (2000) 288(5467):760–4. doi:10.1126/science.1152622

180. Sather DN, Armann J, Ching MK, Mavrantoni A, Sellhorn G, Caldwell Z, et al. Factors associated with the development of cross-reactive neutralizing antibodies during human immunodeficiency virus type 1 infection. J Virol (2009) 83(2):757–69. doi:10.1128/JVI.02036-08

181. Walker LM, Huber M, Doores KJ, Falkowska E, Pejchal R, Julien JP, et al. Broad neutralization coverage of HIV by multiple highly potent antibodies. Nature (2011) 477(7356):466–70. doi:10.1038/nature10373

182. Gray ES, Madiga MC, Hermanus T, Moore PL, Wibmer CK, Tumba NL, et al. Immune responses driven by protective human leukocyte antigen alleles in early life. Front. Immunol. (2014) 5:391. doi:10.3389/fimmu.2014.00391

183. Saez-Cionion A, Bacchus C, Hocquelou L, Avettand-Fenoel V, Girault I, Lecouroux C, et al. Post-treatment HIV-1 controllers with a long-term virological remission after the interruption of early initiated antiretroviral therapy ANRS VISCONTI Study, PLoS Pathog (2013) 9(3):e1003211. doi:10.1371/journal.ppat.1003211

184. Laird GM, Eisele EE, Rabi SA, Lai J, Chioma S, Blankson JN, et al. Rapid quantification of latently infected, resting CD4+ T-cells carrying replication-competent virus in HIV-1-infected individuals. Methods Mol Biol (2005) 304:3–15. doi:10.3920/5925-907-9003

185. Lilandr GM, Eisele EE, Rabi SA, Lai J, Chioma S, Blankson JN, et al. Rapid quantification of the latent reservoir for HIV-1 using a viral outgrowth assay, PLoS Pathog (2013) 9(5):e1003398. doi:10.1371/journal.ppat.1003398

186. Chui TW, Nicle D, Justens J, Meyers JH, Robb G, Hallahan CW, et al. Persistence of HIV in gut-associated lymphoid tissue despite long-term antiretroviral therapy. J Infect Dis (2008) 197(3):714–20. doi:10.1086/527324

187. Yuki SA, Giannella S, Sinclair E, Epling L, Li Q, Duan L, et al. Differences in HIV burden and immune activation within the gut of HIV-positive patients receiving suppressive antiretroviral therapy. J Infect Dis (2010) 202(10):1553–61. doi:10.1086/656722

188. Descours B, Avettand-Fenoel V, Blanc C, Amari S, Melard A, Supervin V, et al. Reduced HIV reservoirs after early treatment HIV-1 proviral reservoirs decay continuously under sustained virologic control in early-treated HIV-1-infected children. J Infect Dis (2014). doi:10.1093/infdis/jiu297

189. Persaud D, Palumbo PE, Ziemniak C, Hughes MD, Alvero CG, Luzuriaga K, et al. Dynamics of the resting CD4+ T-cell latent HIV reservoir in infants initiating HAART less than 6 months of age. AIDS (2012) 26(12):1483–90. doi:10.1097/QAD.0b013e3283536568

190. Ananworanich J, Puthanakit T, Suntarattiwong P, Chokephaibulkit K, Kerr SJ, Fromentin R, et al. Reduced markers of HIV persistence and restricted HIV-specific immune responses after early antiretroviral therapy in children. AIDS (2014) 28(7):1015–20. doi:10.1097/QAD.0000000000000178

191. Persaud D, Palumbo PE, Ziemniak C, Hughes MD, Alvero CG, Luzuriaga K, et al. Dynamics of the resting CD4+ T-cell latent HIV reservoir in infants initiating HAART less than 6 months of age. AIDS (2012) 26(12):1483–90. doi:10.1097/QAD.0b013e3283536568

192. Persaud D, Palumbo PE, Ziemniak C, Hughes MD, Alvero CG, Luzuriaga K, et al. Dynamics of the resting CD4+ T-cell latent HIV reservoir in infants initiating HAART less than 6 months of age. AIDS (2012) 26(12):1483–90. doi:10.1097/QAD.0b013e3283536568

193. Ananworanich J, Puthanakit T, Suntarattiwong P, Chokephaibulkit K, Kerr SJ, Fromentin R, et al. Reduced markers of HIV persistence and restricted HIV-specific immune responses after early antiretroviral therapy in children. AIDS (2014) 28(7):1015–20. doi:10.1097/QAD.0000000000000178