Identification of the RGG Box Motif in Shadoo: RNA-Binding and Signaling Roles?

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Abstract: Using comparative genomics and in-silico analyses, we previously identified a new member of the prion-protein (PrP) family, the gene SPRN, encoding the protein Shadoo (Sho), and suggested its functions might overlap with those of PrP. Extended bioinformatics and conceptual biology studies to elucidate Sho’s functions now reveal Sho has a conserved RGG-box motif, a well-known RNA-binding motif characterized in proteins such as FragileX Mental Retardation Protein. We report a systematic comparative analysis of RGG-box containing proteins which highlights the motif’s functional versatility and supports the suggestion that Sho plays a dual role in cell signaling and RNA binding in brain. These findings provide a further link to PrP, which has well-characterized RNA-binding properties.

Keywords: prion protein, RGG motif, RNA-binding protein, Shadoo, comparative genomics, conceptual biology, methylation, phosphorylation

Introduction

In 2003 we discovered a new gene, SPRN, which codes for a 151-residue protein (including N- and C-terminal signal sequences) with topographical similarities unique to prion protein (PrP), and which is highly conserved between fish and mammals (for an analysis of the similarities between PrP and Sho see Premzl et al. 2003). We called this new protein Shadoo (Sho; shadow of prion protein). Like PrP, Sho is most abundant in brain (Premzl et al. 2003; Uboldi et al. 2006; Watts et al. 2007). Although the functions of Sho are as yet little characterized, it has been shown by gain and loss of function experiments (RNAi and overexpression) to be essential for CNS development in zebrafish (L. Sangiorgio, University of Milan, pers. comm.) and may have a neuroprotective effect similar to PrP (Watts et al. 2007). While PrP is notorious for its association with the transmissible spongiform encephalopathies such as Creutzfeldt Jacob Disease and Bovine Spongiform Encephelopathy (Mad Cow Disease) it has become clear in recent years that PrP has a range of normal functions, including in neurogenesis and neural plasticity (Kanaani et al. 2005; Moya et al. 2005; Santuccione et al. 2005; Steele et al. 2006). In defining the natural functions of Sho we are investigating links of the protein’s properties to those of other proteins, including PrP.

Here we report our finding that Sho has a conserved ‘RGG-box’ motif (Kiledjian and Dreyfuss, 1992) defined as a sequence of closely spaced Arg-Gly-Gly (RGG) repeats interspersed with other, often aromatic, amino acids. The RGG box proteins are one class of RNA-binding proteins (RBPs) involved in various aspects of RNA processing, including splicing, stabilizing, transport and translation of mRNAs (Burd and Dreyfuss, 1994). In addition to being an RNA-binding motif, the RGG box of some proteins is known to mediate interactions with other proteins; for a recent detailed example see Lukasiewicz et al. (2007).

The capacity to bind RNA constitutes another point of similarity with PrP which is known to bind RNA and DNA (Grossman et al. 2003). While it has been established that PrP is competent to bind nucleic acid, it is also known to bind many other ligands including polyanionic glycosaminoglycans (‘GAGs’). Given its propensity to bind polyanions, it is currently unclear whether the binding of nucleic acids is biologically relevant, that is, whether a normal function of PrP involves this type of interaction or whether binding observed experimentally may be a non-specific interaction. However, others have observed that PrP modifies DNA structure in a manner similar to proteins involved in transcriptional regulation (Bera et al. 2007) and have queried whether PrP may be involved in the biogenesis or transport of nucleic acid (Lima et al. 2006).
The approach we have used here is underpinned by a novel combination of comparative genomics (Hedges and Kumar, 2002) and conceptual biology (Blagosklonny and Pardee, 2002). By comparing Sho sequences from species ranging from fish to human and integrating these results with those of a comprehensive analysis of published sequence data and experimental findings, we have been able to put our observations into the broader context of RGG-box proteins. This has allowed us to formulate functional hypotheses for Sho.

Materials and Methods

The amino acid sequences of 12 Sho proteins ranging from fish to human were used in this study. Ten of these sequences are available from GenBank [Homo sapiens Np_001012526, Canis lupus familiaris CA43798, Bos taurus CAJ43799, Mus musculus NP_899790, Monodelphis domestica CAF43800, Gallus gallus CAJ43796, Xenopus tropicalis CAJ43801, Danio rerio CAD35503, Takifugu rubripes CAG34291, Tetraodon nigroviridis CAG30521]. The sequences for Ornithorhynchus anatinus (platypus), M. domestica (American opossum), G. gallus (chicken), and X. tropicalis were initially extracted from the genomic databases (N. Chakka, unpublished work of this group). The sequences for X. tropicalis and X. laevis were also verified experimentally (T. Vassilieva and N. Chakka, unpublished work of this group). The sequences were aligned using ClustalW (Chenna et al. 2003). Subsequent manual adjustments in the N-terminal region were made to the alignment.

The Swiss-Prot protein database was searched using the program Prosite (Hofmann et al. 1999) http://au.expasy.org/ for known motifs within the Sho sequences. We also searched Swiss-Prot for all proteins that have an RGG-box motif, which we defined as being a sequence of at least 3 RGG repeats with no more than 6 residues between the repeats. This search produced 10 archaeal, 229 bacterial, 14 viral and 1632 eukaryotic sequences, within which there are 607 fungal, 300 plant and 70 human sequences. Examination of the human sequences showed that some well-known RGG-box proteins had not been picked up by this search. The search was then broadened to include proteins with 2 RGG repeats separated by 9, 8, 7, 6 or 5 residues. The results were visually inspected and those proteins with at least one ‘RG’ between the RGG repeats were included in our list. All uncharacterized proteins or redundant sequences were excluded. The remaining human proteins are collected in Table S1. We have only recorded the sequence beginning and ending with an RGG repeat. It should be noted that the functional RGG box may extend beyond the sequence denoted in Table S1. The RGG sequences were subsequently aligned using ClustalW.

Results and Discussion

Sho—RGG box

A sequence alignment of the N-terminal segment from residue 25 to 42 (the mature protein starts at residue 24) of Shos from different species (Fig. 1) reveals a strictly conserved arginine methylation site (GGRGG) (Lee and Bedford, 2002) at the beginning of a cluster of RGG repeats.

![Figure 1](image-url) Alignment of the RGG-box sequence at the N-terminal end of Shos from fish to mammals. LHS are sequence numbers. Mdl, Monodelphis domestica; XI, Xenopus laevis; Xi, Xenopus tropicalis; Danio, Danio rerio; Fugu, Fugu rubripes, Tetraodon, Tetraodon nigroviridis. Note that region starts with completely conserved KGG triplet. Complete RGG triplets are bolded.
In Shos from human and most other Eutherian mammals there are three RGG repeats, with the first and third separated by 9 residues (RGGARG-SARGVRRGG). Thus, the RGG box of human Sho consists of 15 residues: 4 positively charged Arg residues, 7 Gly residues—6 of them dipeptides ‘GG’ which give the sequence a large degree of flexibility—, and 4 small intervening residues. This pattern diverges slightly in other species; the first RGG repeat is conserved but the second and third RGG repeats are truncated to RG in some cases. In summary, although there is some variability in the number of Gly residues in the Sho RGG box, for species from fish to human there is conservation of Lys25 and the following 3 Arg residues which are regularly spaced with 3 intervening residues between each Arg. The increased prevalence of Gly-Gly dipeptides in the higher Eutherian mammals could suggest evolutionary pressure for increased flexibility in this domain.

Comparative analysis of RGG-box proteins—structure and composition
Proteins with an RGG-box motif, as defined for the purpose of this study (Methods), are presented in Supplementary Information Table S1. Most (#2–#34) are known to have an RNA-binding function. The subset of proteins highlighted in this paper is presented in Table 1. Analysis of all the proteins listed in Table S1 reveals that the RGG box is generally found at the end of the protein sequence, particularly at the C-terminus (Fig. 2A) and is mostly 10–19 residues in length (Fig. 2B). We found a slight preference for RGG repeats to be separated by 9 intervening residues (RGG-X9-RGG), as in Sho, but overall the spacing is variable (Fig. 2C).

The amino acid composition of the sequences was analysed by calculating the proportion of basic (Arg, Lys and His), acidic (Glu and Asp), aromatic (Phe, Trp and Tyr), polar (Ser, Thr, Asn, Gln and Cys), Gly and the other non-polar amino acids (Ala, Val, Leu, Ile, Met and Pro) which make up each sequence and then producing a frequency distribution for the entire set of proteins (Fig. 2D). As expected, a majority of sequences is Gly rich, with peaks in frequency at 50%–60% Gly composition while basic residues peak at 20%–30%. Although a significant number of sequences do not contain an aromatic acid between the RGG repeats, it is possible that there are aromatic residues in close sequence or spatial proximity to this domain. Very few sequences contain acidic residues.

The Sho RGG sequence conforms to these general structural and compositional parameters. It is found at the end of the protein (N-terminus), is 15 residues long and is comprised of 47% Gly, 27% basic, 20% non-polar and 7% polar residues, and has no acidic or aromatic residues. We aligned the RGG sequence of Sho against other sequences with RGG-X9-RGG spacing in order to identify those most similar to Sho (Fig. 3). Several sequences have 50% or more residues identical to those in the Sho RGG box. Experimental studies have demonstrated that the Fragile X Mental Retardation Protein (FMRP) (#32, Table S1) (Zanotti et al. 2006) and the Herpes Simplex protein ICP27 (Mears and Rice, 1996) bind RNA with their RGG boxes which, like Sho, consist of 2 RGG repeats separated by 9 residues.

Overall, our comparative analysis supports the prediction that the RGG box of Sho is competent to bind RNA.

Sho—predicted arginine methylation and phosphorylation sites
The Arg methylation site in Sho is completely conserved in all species from fish to human, suggesting functional importance. Arginine methylation is a common post-translational modification in RGG-box domains (Liu and Dreyfuss, 1995) which affects protein-protein interactions (Boisvert et al. 2005) and RNA binding (Dolzhanskaya et al. 2006). It influences diverse cellular processes, including cellular location of proteins (Passos et al. 2006) transcription, processing and transport of mRNAs (Yu et al. 2004) and signaling pathways (Boisvert et al. 2005).

Phosphorylation is another common post-translational modification found in RBPs. Methylation and phosphorylation mechanisms co-regulate a number of RGG-box proteins, possibly including Sho; again for a detailed example see Lukasiewicz et al. (2007). We identified 3 potential protein kinase C (PKC) phosphorylation sites (SAR (34–36 huSho), SLR (63–65 huSho) and SYR (119–121 huSho)) for Sho. One of these, SAR34–36, is within the RGG box and is found in all the Eutherian mammal sequences analysed (Fig. S1 in Supplementary Information). Phosphorylation of Ser34 would have a direct affect on the structure of the RGG box and most likely affect its function. Although the phosphorylation-site motifs are patterns with a high probability of random
Table 1. Subset of RGG-box proteins (see Table SI in Supplementary Information for full list).

| No. | Name (ID) | RGG domain (residue numbers) | Other RNA-binding motifs\(^b\) | Functions/Comments R\(^d/P\)^f | (For details and references see Table SI) |
|-----|-----------|------------------------------|-------------------------------|--------------------------------|------------------------------------------|
| 1   | Shadoo    | RGGARGSARGGVRGGG (28–42)    |                               | PrP family member. Likely attached to cell membrane by GPI anchor. |                                           |
| 12  | hnRNP U   | RGGGHRGGRGFNMHRGGN-FRGGAPGNRGG (701–728) | RGG box first identified when a 26-residue sequence (MRGGNFRGGAPGNNRGG-YNRRGN) found to be sufficient for RNA binding. Expected involvement in splicing, pre-mRNA processing and stabilizes specific mRNAs. |                                           |
| 21  | HABP4     | RGGPRGGMRGRGRGG (185–199)   | Also constitutes a hyaluronan binding motif, (R/K–X(7)-R/K) where X is not acidic. Binds strongly and specifically to hyaluronan and weakly to RNA. Involved in mRNA transport, chromatin remodeling, regulation of transcription. |                                           |
| 26  | EWS       | RGGFDRGGMRSRGG-GRGGGMGSAGERGG (304–332) and RGGPGGMRGRGLMGRGPGGMFRG-GRGDGRGFRRGGMRGRMGRG (565–617) | 1 RRM\(^c\) Found on cell surface, nucleus and cytoplasm. Is a transcriptional activator but this activity can be repressed by RGG box. May be involved in pre-mRNA splicing and transport. Suggested that EWS protein acts as a receptor or binding protein for ligands on cell surface, such as nucleic acids, and thus might mediate extracellular and nuclear events. |                                           |
| 32  | FMRP      | RGGGGGRGQGGGRGGRGG (534–548) | 2 KH\(^d\) Binds many mRNA transcripts. Transports mRNA from nucleus to cytoplasm. Involved in neural plasticity through translational repression. |                                           |
| 33  | Nucleolin | RGGGRGFGGGRGGRGGRG-GRGGGMGRGGG (656–696) | 4 RRM Found on cell surface, nucleus and cytoplasm. RGG box is necessary for efficient RNA binding but the RRMs are required for specific RNA recognition. Duplex DNA, ssDNA and RNA are all effective ligands. Acts as cell surface receptor—binds cytokine MK and HB-19 through its RGG box. |                                           |
| 34  | G3BP1     | RGGLGGMRPGPRGG (435–449)    | 1 RRM Role in ras-signaling pathway affecting cell proliferation and survival as well as involved in RNA metabolism. Cleaves MYC mRNA and has helicase activity. Combining these functions, suggested to be member of novel sub-class of RBPs which act at level of RNA metabolism in response to cell signaling, thus allowing cell to rapidly control protein activity at a stage after transcription. |                                           |

\(^a\)number of protein as appears in Table S1.
\(^b\)RNA-binding motifs in addition to the RGG box.
\(^c\)RGG box first identified when a 26-residue sequence (MRGGNFRGGAPGNNRGG-YNRRGN) found to be sufficient for RNA binding. Expected involvement in splicing, pre-mRNA processing and stabilizes specific mRNAs.
\(^d\)K homology region as in hnRNP K.
\(^e\)RNA binding.
\(^f\)Protein binding.
occurrence it is interesting to note that the presence of at least one phosphorylation site has been experimentally confirmed in 70% of the RGG-box proteins surveyed (Table S2 in Supplementary Information). This is a high proportion even taking into account the over-representation of nuclear proteins in the phosphoproteome (Olsen et al. 2006) and leads us to suggest that phosphorylation is particularly prevalent in RGG-box proteins. The finding of potential methylation and phosphorylation sites in Sho is another point of similarity with other RGG-box proteins. The existence of phosphorylation sites within Sho raises the possibility that Sho may be involved in a signaling pathway that is regulated by phosphorylation.

Functional significance
Sho differs from most of the other proteins surveyed in that it has no other RNA-binding motifs. This is unusual but not unique as hnRNP U (#12; Table S1) has no other RNA-binding motif apart from the RGG box. The RGG box is typically associated with binding to single-stranded nucleic acids, (Zhang and Grosse, 1997) whereas additional RNA-binding motifs may allow binding of a broader range of RNA targets as is the case for nucleolin (#33, Table S1) (Ghisolfi et al. 1992). The inherent flexibility of the RGG box (Ramos et al. 2003) can also enable binding to several RNA targets, as has been shown for FMRP (Darnell et al. 2004) which binds to many RNA targets but an affinity for RNA that forms a stable G-quartet structure (Menon and Mihailescu, 2007; Ramos et al. 2003). As Sho lacks other RNA-binding motifs, we expect it to bind single-stranded nucleic acid, and potentially a range of such targets, as for FMRP.

The RGG box is a positively charged domain known to interact electrostatically with other proteins and anionic molecules. A well-characterized

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**Figure 2.** Frequency histograms of structural and compositional features of the 45 RGG sequences surveyed (Table S1). **A)** Position of the RGG box region in the proteins. N-terminal (within the first 35% of the protein sequence), C-terminal (last 35% of the protein), Middle, region between. **B)** Length of the RGG box. **C)** Spacing between RGG repeats where X may be any residue including Arg and Gly. **D)** Amino acid composition in terms of type: basic (R, K, H); acidic (D, E); Gly; aromatic (F, Y, W); non-polar amino acids (A, V, L, I, M, P) and polar (S, T, N, Q, C).
example is the RGG box of the yeast protein Npl3p (Lukasiewicz et al. 2007). A non-protein example is provided by the intracellular hyaluronan binding protein (HAPB4) (#21, Table S1) which has high sequence similarity to Sho (Fig. 3). The RGG domain of HAPB4 also constitutes a glycosaminoglycan (‘GAG’) binding motif (R/K–X(7)-R/K) (Yang et al. 1994) and has been found to bind strongly and specifically to hyaluronan and weakly to RNA (Huang et al. 2000). Although it is not surprising to find this motif in an Arg-rich sequence (in fact it is present in most of the proteins included in Table S1), it has particular relevance in the case of Sho, given its cellular location.

The cellular location of Sho will determine its opportunities to bind RNA and whether this is its primary function. We originally predicted Sho to be a GPI-anchored protein (Premzl et al. 2003). This has now been confirmed in mouse (Watts et al. 2007) and for a Sho-like protein (Sho2) (Premzl et al. 2004; Strumbo et al. 2006) in zebrafish (Miesbauer et al. 2006). However, some GPI-anchored proteins, including PrP, undergo anchor cleavage (‘shedding’), (Parkin et al. 2004; Zhang et al. 2005) resulting in formation of soluble proteins which can relocate to other cellular destinations and are capable of performing multiple functions (Campana et al. 2005). While the cell surface is one likely location for Sho, it may be a multifunctional protein found in other cellular locations as well, as for PrP. If Sho sheds its GPI anchor or undergoes proteolytic cleavage before attachment to the cell membrane (Watts et al. 2007), the RGG-box domain would be available for functional roles intracellularly. Other RGG-box proteins are known to have multiple cellular locations, for example, nucleolin and the Ewing Sarcoma (EWS) protein (#26, Table S1) are found on the cell surface as well as in the nucleus and cytoplasm. In fact, there is growing evidence that some RNA-binding proteins have additional roles as cell surface receptors (Bajenova et al. 2003; Belyanskaya et al. 2003; Hirano et al. 2005) and in signaling pathways as noted for the ras GTPase activating protein binding protein 1 (Kennedy et al. 2001).

Attached to the cell surface, Sho would be positioned to act as a receptor for ligands found at the cell surface, including nucleic acids, as suggested for EWS (Belyanskaya et al. 2001). Sho may, therefore, have a role in cell signaling, similar to PrP which binds the neural cell adhesion molecule and thus participates in the tyrosine kinase fyn signaling pathway leading to neurite outgrowth (Santuccione et al. 2005). Alternatively, in this location Sho may bind other anionic ligands such as the GAG, hyaluronan, which is known to bind another GPI-anchored protein, brevican, and is involved in the structural plasticity of neural tissue.

### Table S1: Alignments of Protein Sequences with RGG Box

| Protein       | Start | End | #  | Sequence |
|---------------|-------|-----|----|----------|
| Q5BIV9_HUMAN  | 28    | 42  | 10 | RGGARRGGA SARGGGGGG VGGGGG |
| HABP4_HUMAN   | 185   | 199 | 10 | RGGARRGGG MRRGGGRRGG |
| FUS_HUMAN     | 491   | 505 | 10 | RGGARRGGG GGGGGGGGG |
| K1C9_HUMAN    | 478   | 492 | 9  | RGGARRGGG SGGGGGYGGRS |
| G3BP1_HUMAN   | 435   | 449 | 9  | RGGARRGGG LGGGGGMRGPP |
| LS14A_HUMAN   | 406   | 420 | 9  | RGGARRGGG YGGGGGGLFG |
| NOLA1_HUMAN   | 173   | 187 | 9  | RGGARRGGG RGGGGGGG |
| NOLA1_HUMAN   | 185   | 199 | 9  | RGGARRGGG RGGGGGGG |
| EWS_HUMAN     | 309   | 323 | 8  | RGGARRGGG RGGGGGGG |
| NOLA1_HUMAN   | 30    | 44  | 8  | RGGARRGGG GGGGGGGG |
| FBRL_HUMAN    | 24    | 38  | 8  | RGGARRGGG GGGGGGGG |
| BRWD3_HUMAN   | 1699  | 1713| 8  | RGGARRGGG TTRGRGGG |
| RBP56_HUMAN   | 337   | 351 | 8  | RGGARRGGG GGGGGGGG |
| THOC4_HUMAN   | 38    | 52  | 8  | RGGARRGGG AQAARGVR |
| FBRL_HUMAN    | 15    | 29  | 7  | RGGARRGGG GGGGGGGG |
| FA9A8_HUMAN   | 352   | 366 | 7  | RGGARRGGG GGGGGGGG |
| PP1RA_HUMAN   | 726   | 740 | 7  | RGGARRGGG GGGGGGGG |
| HNRPG_HUMAN   | 113   | 127 | 7  | RGGARRGGG GGGGGGGG |
| FMR1_HUMAN    | 534   | 548 | 7  | RGGARRGGG GGGGGGGG |
| HNRP3_HUMAN   | 714   | 728 | 7  | RGGARRGGG GGGGGGGG |
| CA077_HUMAN   | 128   | 142 | 6  | RGGARRGGG GGGGGGGG |

**Figure 3.** Alignment of the RGG box of proteins with RGG-X9-RGG spacing. The number of the residue at the start and end of the sequence is given, as well as the total number of exact residue matches (#) to Sho.
(Rauch, 2004). It is interesting to note that PrP also binds GAGs including hyaluronan and heparin (Pan et al. 2002) and that GAGs may facilitate the conversion of the normal cellular PrP to the isofrom found in prion disease (Yin et al. 2007).

If Sho were to shed its GPI anchor and re-enter the cell or if a segment of the N-terminal region incorporating the RGG domain was cleaved off prior to expression at the cell surface, the RGG box would be available to interact with cellular RNA. Indeed, as a small protein of no more than 123 residues, Sho would be capable of diffusing in and out of the nucleus (Cyert, 2001) and shuttling RNA from the nucleus to the cytoplasm. This is a function normally performed by RNA-binding proteins involved in neural plasticity, which participate in the biogenesis of mRNA, its transport to dendrites and repression of translation pending appropriate neural stimulation (Ule and Darnell, 2006).

**Conclusion**

In summary, we have observed that Sho has a conserved RGG-box domain with similar composition to other known RGG-box proteins. We predict that this domain has functional significance and may mediate some of the neural functions already indicated for Sho. Our analysis leads us to postulate that Sho is an RNA-binding protein which may also play a role in cell signaling. Our initial experiments to test the prediction have shown Sho RGG box peptide is competent to bind RNA but further work is required to characterize the interaction.

The discovery of the RGG box in Sho opens new avenues for investigating its function and potential functional overlap with PrP. It is known that PrP plays a role in neural plasticity through its involvement in neural signaling pathways. Here we suggest that Sho may bind mRNA directly and thus play a role in neural plasticity similar to other neural RBPs.

**Note**

Beck et al. (J. Med. Genet. online 19/9/08) have reported an association of a null allele of SPRN with variant CJD.

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**Disclosure**

The authors report no conflicts of interest.

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### Identification of the RGG Box Motif in Shadoo: RNA-Binding and Signaling Roles?

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### Supplementary Information

**Table SI.** Proteins with RGG-box domains Selected on Criteria explained in Methods.

| Database name, name (ID) | # AA | RGG domain (residue numbers) | Other RNA binding motifs\(^a\) | Functions/Comments | R\(^b\)/P\(^c\) |
|--------------------------|------|-------------------------------|---------------------------------|--------------------|-----------------|
| SHO_HUMAN, Shadoo (Q5BIV9) | 151  | RGGARGSARGGVRGG (28–42)      |  | PrP family member. Likely attached to cell membrane by GPI anchor (Premzl et al. 2003). | |
| ROA0_HUMAN, hnRNP A0 (Q13151) | 305  | RGGNFSGRGGFGGSRGG (192–202)  | 2 RRM\(^b\) | Found in splicesome C; expected involvement in splicing, pre-mRNA processing. Similar to hnRNP A/B but less abundant. Component of ribonucleosomes. (Jurica et al. 2002). | R |
| ROA1_HUMAN, hnRNP A1 (P09651) | 372  | RGGNFSGRGGFGGSRGG (218–234)  | 2 RRM | Found in splicesome C; expected involvement in splicing, pre-mRNA processing. Transport of poly (A) mRNA from nucleus to cytoplasm. (Biamonti et al. 1989; Jurica et al. 2002; Siomi and Dreyfuss, 1995). | R |
| ROA2_HUMAN, hnRNP A2 (P22626) | 353  | RGGNFSGRGGVRGG (203–230)      | 2 RRM | Found in splicesome C; expected involvement in splicing, pre-mRNA processing. Trafficking of RNAs containing the cis-acting A2 response element (A2RE). (Jurica et al. 2002). | R |
| ROA3_HUMAN, hnRNP A3 (P51991) | 378  | RGGGSNGFMGRGGNFGG (216–241)  | 2 RRM | Found in splicesome C; expected involvement in splicing, pre-mRNA processing. Trafficking of RNAs containing the cis-acting A2 response element (A2RE). (Jurica et al. 2002). | R |
| HNRPD_HUMAN, hnRNPD0 (Q14103) | 355  | RGGFAGRARGGG (272–284)        | 2 RRM | Binds to mRNA with AU-rich elements (AREs) in 3’-UTR. Transcription regulator; binds to ds and ss DNA sequences. Possibly Involved in translationally coupled mRNA turnover. (Kajita et al. 1995; Tay et al. 1992). | R |
| HNRPG_HUMAN, hnRNP G (P38159) | 391  | RGGSGGTRGPPSRGG (113–126)      | 1 RRM | Found in splicesome C; expected involvement in splicing, pre-mRNA processing. (Jurica et al. 2002). | R |

(Continued)
| Database name, name (ID) | # AA | RGG domain (residue numbers) | Other RNA binding motifs | Functions/Comments | $R^2/P^*$ |
|--------------------------|------|-------------------------------|--------------------------|-------------------|--------|
| HNRPK, hnRNP K (P61978)  | 463  | RGGFDRMPPGRRGGRPMPPSRRDYDDMSPPRGGGRRPGLPLPPPPPPPRGGG (258–318) | 3 KH$^c$ | Found in splicosome C; expected involvement in splicing, pre-mRNA processing. Major poly(C) RNA binding hnRNP. Also binds poly(C) ssDNA. (Jurica et al. 2002). | R |
| HNRPQ_HUMAN, hnRNP Q (060506) | 623 | RGGPGSARGVRGARGGAQQQRGRGRVRGARGGRGG (526–559) | 3 RRM | 3 isoforms. Pre-mRNA processing. Associated with splicing intermediates and mature mRNA. Interacts preferentially with poly(A) and poly (U) RNA sequences. Region 518–549 (RGRAGYSQRGPGSARGVRGR GGAQQQRGRG) sufficient to bind RNA. (Mourelatos et al. 2001). | R |
| HNRPR_HUMAN, hnRNP R (043390) | 633 | RGGRGPAQQQRGRGSRGSRGNRGG (543–567) | 3 RRM | Found in splicosome C; expected involvement in splicing, pre-mRNA processing. (Hassfeld et al. 1998; Jurica et al. 2002). | R |
| HNRPU_HUMAN, hnRNP U (Q00839) | 824 | RGGGHRGRGGFMRGRGGNFRGGAPGNNRGG (701–728) | 3 RRM | First discovered that hnRNP U contains a 26-residue peptide (MRGGFRGGRGPRG N) sufficient to account for its RNA-binding activity. This novel RNA-binding motif was defined as the RGG box. Found in splicosome C; expected involvement in splicing, pre-mRNA processing. Stabilizes specific mRNAs. Aka SAF-A; has high affinity for DNA with scaffold attachment regions (SAR). (Fackelmayer and Richter, 1994; Helbig and Fackelmayer, 2003; Jurica et al. 2002; Kiledjian and Dreyfuss, 1992; Yugami et al. 2007). | R |
| HNRL1, hnRNP U like protein 1 (Q9BUJ2) | 856 | RGGGGGFRGRGGGGGFFQRYENRGPPGGNRRGFFQNRGGGSGGGNYRGG (612–658) | Pre-mRNA processing and transport. Binds poly(G) and poly(C) RNA. Represses transcription driven by viral and cellular promoters. Associated with RBRD7, activates transcription. (Gabler et al. 1998). | R |
| PURG_HUMAN, Purine-rich element-binding protein gamma (Q9UJV8) | 347 | RGGGGGRGRRGGG (7–17) | In purine-rich element binding protein family (PUR). Binds ssDNA and RNA. Highly expressed in many tumor lines. (Liu and Johnson, 2002). | R |
### Functions of RGG box of Shadoo

| No. | Accession | Name, Description                                                                 | RGG Box Sequence | Additional Information                                                                 |
|-----|-----------|-----------------------------------------------------------------------------------|------------------|---------------------------------------------------------------------------------------|
| 15  | DDX4_HUMAN, DEAD box protein 4 (Q9NQI0) | In DEAD box helicase family. Helicase activity, RNA unwinding, needed in splicing, ribosome biogenesis and RNA degradation. (Castrillon et al. 2000; Luking et al. 1998). | RGGRGFSRGRGG (147–159) | ![ ] | R |
| 16  | THOC4_HUMAN, Tho complex subunit 4 (Q86V81) | In THO/TREX complex, promotes transcriptional activation, recruited to RNA polymerase during elongation. Associated with spliced mRNA; roles in mRNA export and decay. May mediate interactions of proteins and/or RNA. (Strasser et al. 2002; Virbasius et al. 1999). | RGGGAQAARVNRGG (38–52) | ![ ] | R/P |
| 17  | NOLA1_HUMAN, Nucleolar protein family A member 1 (Q9NY12) | Aka GAR1. Required for ribosome biogenesis and telomere maintenance. Processing or intranuclear trafficking of TERC, the RNA component of the telomerase reverse transcriptase (TERT). RGG box accessory to RNA binding. Interaction with SMN1 requires at least one of the RGG-box regions. (Bagini and Lapeyre, 1998; Whitehead et al. 2002). | RGGGRGFNRRGGGGF NRGNSSNHFRGGGGGGG GNFRGGGRGGFGRRG | ![ ] | R |
| 18  | SFPQ_HUMAN, Splicing factor proline- and glutamine-rich (P23246) | Pre-mRNA splicing factor. Binds to intronic polypyrimidine tracts. Possible role in nuclear retention of defective RNAs. Regulates basal and cAMP-dependent transcription. (Patton et al. 1993). | RGGGGGGFHRRGGGGG RGG (9–27) | ![ ] | R |
| 19  | FBRL_HUMAN, Fibrillarin (P22087) | Involved in pre-rRNA processing. Component of box C/D small nucleolar ribonucleoprotein (snoRNP) particles. (Aris and Blobel, 1991; Jansen et al. 1991). | RGGGFGGGFGFDGGG RGGGRGGGGGGRGGG GFRGRGRGG (8–47) | ![ ] | R |
| 20  | HABP4_HUMAN, Hyaluronan binding protein (HAPB4, Ki-1/57) (Q5JVS0) | This sequence also constitutes a hyaluronan binding motif, (R/K-X(7)-R/K) where X is not acidic (Yang et al. 1994). This domain within HABP4 has been found to bind strongly and specifically to hyaluronan and weakly to RNA. Involved in mRNA transport, chromatin remodeling, regulation of transcription. Interacts with chromodomain DNA helicase binding protein 3(CHD3). (Kobarg et al. 1997; Lemos et al. 2003; Passos et al. 2006). | RGGPRRGGMRGRG | ![ ] | R/P |
| 21  | PAIRB_HUMAN, Plasminogen activator inhibitor 1 RNA binding protein (Q8NC51) | Aka CG1–55. Regulation of mRNA stability/decay. Interacts with CHD3, similar to HABP4. (Lemos et al. 2003). | RGGGRGGG | ![ ] | R/P |

(Continued)
Table SI. (Continued)

| Database name, name (ID) | # AA | RGG domain (residue numbers) | Other RNA binding motifs$^a$ | Functions/Comments | R$^d$/P$^e$ |
|--------------------------|------|------------------------------|-----------------------------|--------------------|-------------|
| 23 FUS_HUMAN, RNA binding protein FUS (P35637) | 526 | RGGGRGGRGGMGGS | 1 RRM | Component of nuclear riboprotein complexes. Binds ds and ss DNA. Promotes annealing of complementary ssDNAs. (Rabbits et al. 1993). | R/P |
| 24 | | RGGGNRGRGRGGPMG RGG (377–396) | | | |
| 25 | | RGGRRGGRGGYRGRGG DRGGFRGGRGGDRGGG (473–505) | | | |
| 26 EWS_HUMAN, Ewing sarcoma (EWS) protein (Q01844) | 656 | RGGFDRGEMSRGRGGGG RGGMGSAGERGG (304–332) | 1 RRM | Found on cell surface as well as in the nucleus and cytoplasm. Binds RNA. Is a transcriptional activator but this activity can be repressed by the RGG box. May be involved in pre mRNA splicing and transport. It has been suggested that EWS protein may act as a receptor or binding protein for ligands on the cell surface, such as nucleic acids, and thus might mediate extracellular and nuclear events. Interacts with PTK2B/FAK2 then relocates from cytoplasm to ribosomes. (Belyanskaya et al. 2003; Belyanskaya et al. 2001; Ohno et al. 1994; Plougastel et al. 1993). | R/P |
| 27 | | RGGPGGMRGGGRGLMD RGGFRGGRGMDRGGFGRGG RRRGG (565–617) | | | |
| 28 RB56_HUMAN, TATA-binding protein-associated factor 2N (Q92804) | 592 | RGGYRGGGFQRRGG (337–351) | 1 RRM | Binds RNA and ssDNA. Transcription regulation. In RNA polymerase II transcriptional multiprotein complex. Similar to EWS and FUS/TLS. (Morohoshi et al. 1996). | R/P |
| 29 | | RGGGYDDGRGGGYYGD RGGGYGGDRGGYGD RGGGYGDRGGYGD RGGGYGDGRGGYGD GGGDRSRGGYGDGRGG (459–537) | | | |
| 30 CIRPB_HUMAN, Cold-inducible RNA-binding protein (Q14011) | 172 | RGGSAGGRGFFRGGRGRRR RGFSRGG (94–118) | 1 RRM | Cold-induced suppression of cell proliferation. Activates the ERK pathway. (Nishiyama et al. 1997). | |
| Gene Symbol       | Protein Name                                                          | RGG Box Sequence                                                                 | RGG Box Number | Other Characteristics                                                                                                                                                                                                                                                                                                                                 |
|-------------------|-----------------------------------------------------------------------|---------------------------------------------------------------------------------|----------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| PP1RA_HUMAN       | Serine/threonine-protein phosphatase 1 regulatory subunit (Q96QC0)   | Aka p99. Binds mRNA, ssDNA, poly(A) and poly(G). Inhibits phosphatase activities when phosphorylated. (Kreivi et al. 1997; Totaro et al. 1998). | RGGPGPGPGYPYHRGRGG RGNEPPPPPPPPFRGA RGGRSGGGPNNRGGG (693–740) |                                                                                                                                                                                                                                                                                                                                                          |
| FMR1_HUMAN        | Fragile X Mental Retardation Protein (FRMP) (Q06787)                 | Binds many mRNA transcripts. Transports mRNA from nucleus to cytoplasm. Involved in neural plasticity through translational repression. (Bagni and Greenough, 2005; Darnell et al. 2001; Ule and Darnell, 2006; Zalfa et al. 2003). | RGGGGRGQGGGRGRGG (534–548) | 2 KH                                                                                                                                                                                                                                                                                                                                                       |
| NUCL_HUMAN        | Nucleolin (P19338)                                                   | Found on cell surface as well as in the nucleus and cytoplasm. RGG box is necessary for efficient RNA binding and possibly operates by unstacking RNA bases, but the RRM are required for specific RNA recognition. Duplex DNA, ssDNA and RNA are all effective ligands for nucleolin. Associated with intranucleolar chromatin and preribosomal particles. Binds to histone HI to induce chromatin decondensation. When attached to the cell surface, nucleolin binds the proteins cytokine MK and HB-19 through its RGG box and acts as cell surface receptor. (Ghisolfi et al. 1992; Hirano et al. 2005; Said et al. 2002). | RGGGRGGFGGRGGGRGG RGFFGGGRGGFGGGRGGGFGGRGG (656–696) | 4 RRM                                                                                                                                                                                                                                                                                                                                                     |
| G3BP1_HUMAN       | Ras GTPase-activating protein-binding protein 1 (Q13283)             | G3BP has a role in the ras-signaling pathway affecting cell proliferation and survival as well as being involved in RNA metabolism. Cleaves MYC mRNA And has Helicase activity—unwinds DNA/DNA, RNA/DNA and RNA/RNA. Combining these two functions, it has been suggested the G3BPs are members of a novel subclass of RNA-binding proteins which act at the level of RNA metabolism in response to cell signaling allowing the cell to rapidly control protein activity at a stage after transcription. Also involved in formation of stress granules. (Irvine et al. 2004; Kennedy et al. 2001; Tourriere et al. 2003; Tourriere et al. 2001). | RGGLGGGMRGPPRGG (435–449) | 1 RRM                                                                                                                                                                                                                                                                                                                                                     |

(Continued)
| Database name, name (ID) | # AA | RGG domain (residue numbers) | Other RNA binding motifs | Functions/Comments | R^2/P^* |
|--------------------------|------|------------------------------|--------------------------|--------------------|---------|
| RGMC_HUMAN, Hemojuvelin (precursor) (Q6ZVN8) | 426 | RGGSSGALRGGGGGG RGG (54–72) | | In repulsive guidance molecule (RGM) family; RGMa and RGMb involved in neural development. GPI anchored. Interacts with neogenin which regulates shedding of GPI anchor. Binding cytokines BMP2 and BMP4 affects BMP signaling pathway and expression of hepcidin. Function of RGG domain unknown. (Matsunaga and Chedotal, 2004; Zhang et al. 2007; Zhang et al. 2005). | P |
| ZNH14_HUMAN, Zinc finger HIT domain-containing protein 4 (Q9C086) | 343 | RGGRGGARGERRGG (238–251) | | Aka PAPA-1. Induces growth and cell cycle arrest at GI phase. Interacts with splicing factors altering pre-mRNA splicing. Complexes with other nucleolar proteins. (Kuroda et al. 2004; Maita et al. 2004). | P |
| K1C9_HUMAN, Keratin type 1 cytoskeletal 9 (P35527) | 623 | RGGSYSYGRGSRGG (478–492) | Cytoskeletal and microfibrillar keratin. Function in mature or developing palmar and plantar skin (Langbein et al. 1993). | | |
| MRE11_HUMAN, Double strand break repair protein MRE 11A (P49959) | 708 | RGGRGQNSASRGG (577–589) | In MRN complex, role in dsDNA repair, recombination, maintenance of telomere integrity and meiosis. (Petrini et al. 1995). | | |
| WBP7_HUMAN, WW domain-binding protein 7 (Q9UMN6) | 2715 | RGGQSSRGGRRGG RGRGRRG (281–299) | WW domain-binding (Trithorax homolog 2). Possible transcriptional regulator. (FitzGerald and Diaz, 1999). | | |
| BRWD3_HUMAN, Bromodomain and WD repeat-containing protein 3 (Q6RI45) | 1802 | RGGGGTRGRGRRGG (1699–1713) | In WD repeat protein family involved in cell-cycle progression, signal transduction, apoptosis, gene regulation. Possible transcription factor with 2 bromodomains and 9 WD repeats. May be involved in Jak/Stat pathway. (Vodermaier, 2001). | | |
| CA077_HUMAN, Uncharacterised protein Clorf77 (Q9Y3Y2) | 248 | RGGVRGRGGPGRGG (153–166) | | NA |
|   | Accession       | Position | RGG Box                  | Description                                      |
|---|----------------|----------|--------------------------|--------------------------------------------------|
| 42| FA98A_HUMAN    | 519      | RGGHEQGGGRGG            | NA                                               |
|   | Protein FAM98A|          | YDHGGRGG                |                                                  |
|   |                |          | (352–374)               |                                                  |
| 43| (Q8NCA5)       | 519      | RGGGRGGGRGGGRGG         | NA                                               |
|   | FA98A_HUMAN    |          | (458–473)               |                                                  |
| 44| LS14A_HUMAN    | 463      | RGGYRGRGGGGLGF          | Putative alpha synuclein binding protein.        |
|   | LSM14 protein  |          | RRGRGRGGGRGG            |                                                  |
|   | homolog A      |          | (406–429)               |                                                  |
|   | (Q8ND56)       |          |                          |                                                  |

*RNA-binding motifs in addition to the RGG box.

RRM = 80–90 amino acid sequence containing RNP-1 (octapeptide) and RNP-2 (6 amino acid) consensus sequences.

*K homology region as in hnRNP K.

RNA binding.

Protein binding.
Table S2. Phosphorylation sites in RGG box proteins surveyed in this studya.

| Protein        | Id       | PKCb | CK2c | TYRd | Expt\* |
|----------------|----------|------|------|------|--------|
| SHO_HUMAN      | Q5BIV9   | 3    | 0    | 0    |        |
| ROA0_HUMAN     | Q13151   | 4    | 3    | 0    | 2      |
| ROA1_HUMAN     | P09651   | 10   | 10   | 0    | 9      |
| ROA2_HUMAN     | P22626   | 9    | 4    | 0    | 6      |
| ROA3_HUMAN     | P51991   | 9    | 7    | 0    | 6      |
| HNRPD_HUMAN    | Q14103   | 9    | 6    | 1    | 7      |
| HNRPG_HUMAN    | P38159   | 18   | 16   | 1    | 7      |
| HNRPK          | P61978   | 7    | 12   | 1    | 6      |
| HNRQ_HUMAN     | O60506   | 7    | 3    | 2    | 2      |
| HNRPR_HUMAN    | O43390   | 5    | 5    | 2    |        |
| HNRPU_HUMAN    | Q00839   | 10   | 5    | 0    | 5      |
| HNRL1          | Q9BUJ2   | 6    | 9    | 0    | 3      |
| PURG_HUMAN     | Q9UJV8   | 6    | 2    | 0    | 1      |
| DDX4_HUMAN     | Q9NQI0   | 16   | 14   | 0    |        |
| THOC4_HUMAN    | Q86V81   | 4    | 5    | 0    | 1      |
| NOLA1_HUMAN    | Q9NY12   | 3    | 1    | 0    |        |
| SFPQ_HUMAN     | P23246   | 7    | 4    | 2    | 1      |
| FBRL_HUMAN     | P22087   | 5    | 3    | 0    |        |
| HABP4_HUMAN    | Q5JVS0   | 5    | 8    | 2    | 2      |
| PAIRB_HUMAN    | Q8NC51   | 6    | 10   | 0    | 12     |
| FUS_HUMAN      | P35637   | 7    | 6    | 1    |        |
| EWS_HUMAN      | Q01844   | 4    | 5    | 0    |        |
| RB56_HUMAN     | Q92804   | 7    | 12   | 2    | 1      |
| CIRPB_HUMAN    | Q14011   | 4    | 2    | 1    |        |
| PP1RA_HUMAN    | Q96QC0   | 10   | 12   | 2    | 4      |
| FMR1_HUMAN     | Q06787   | 9    | 12   | 1    | 1\*    |
| NUCL_HUMAN     | P19338   | 8    | 23   | 0    | 14     |
| G3BP1_HUMAN    | Q13283   | 2    | 6    | 1    | 5      |
| RGM_HUMAN      | Q6ZVN8   | 11   | 2    | 0    |        |
| ZNH14_HUMAN    | Q9C086   | 3    | 0    | 0    |        |
| K1C9_HUMAN     | P35527   | 7    | 14   | 3    |        |
| MRE11_HUMAN    | P49959   | 16   | 17   | 0    | 4      |
| WBP7_HUMAN     | Q9UMN6   | 40   | 36   | 3    | 4      |
| BRWD3_HUMAN    | Q6RI45   | 34   | 42   | 4    | 4      |
| CA077_HUMAN    | Q9Y3Y2   | 4    | 2    | 0    | 1      |
| FA98A_HUMAN    | Q8NCA5   | 5    | 9    | 1    |        |
| LS14A_HUMAN    | Q8ND56   | 4    | 7    | 1    | 11     |

aSearches were conducted using the ScanProsite program available on the ExPASy Proteomics Server of the Swiss Institute of Bioinformatics website http://au.expasy.org/.
bNumber of protein kinase C phosphorylation sites (PS00005).
cNumber of casein kinase II phosphorylation sites (PS00006).
dNumber of tyrosine kinase phosphorylation sites (PS00007).
eAs annotated in the SwissProt database.

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