Analgesic Effect of Valerian Root and Turnip Extracts

Afshin Zare¹, Zabihollah Khaksar², Zahra Sobhani¹, Masood Amini¹*

ABSTRACT

BACKGROUND

Medicinal plants are considered as one of the important sources of chemical substances with therapeutic effects. This study aimed to compare the analgesic effects of alcoholic extract of valerian root and turnip in rats.

METHODS

Fifty female Wistar rats weighing 190 g were divided into 5 equal groups of control (subcutaneous injection of 2.5% formalin in the right foot), sham (subcutaneous injection of 2.5% formalin+distilled water), experimental 1 (subcutaneous injection of 2.5% formalin+200 mg/kg turnip extract), experimental 2 (subcutaneous injection of 2.5% formalin+200 mg/kg valerian root extract) and experimental 3 (subcutaneous injection of 2.5% formalin+200 mg/kg turnip extract+200 mg/kg valerian root extract). The time duration of 0-5 and 16-60 minutes after injection of formalin were respectively considered as acute and chronic phases. Injection of distilled water and the extracts was conducted 30 minutes before assessing the analgesic effects.

RESULTS

A significant decrease in pain score in the acute phase was observed in the group received valerian root extract compared to the control group. Also, a significant reduction in pain score was noted in the acute and chronic phases of the group receiving simultaneous administration of valerian root and turnip extracts when compared to the control group.

CONCLUSION

Simultaneous use of valerian root and turnip extracts is recommended for analgesic effects in both acute and chronic phases of the pain.

KEYWORDS

Analgesia; Valerian root; Turnip; Formalin test; Pain; Rat

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INTRODUCTION

Since control of pain and its complications are important issue in surgical interventions, several and various researches were performed worldwide to find the best way to deal with pain reduction. The pathways involved in induction of pain are largely...
known, while a large number of medications have been used to control these pathways and among them, opioid analgesics have been widely used. Of course it should be borne in mind that these medicines induce tolerance and dependences too, and can cause incidence of unwanted effects; so necessary caution should be undertaken when they are taken by a patient.

These medicines control pain by affecting the central parts of the body. Unfortunately, opioids have their own side effects too and cause gastrointestinal complications such as gastrointestinal bleeding denoting to their hazards when are used. That is why finding new medicines to alleviate pain and to have fewer side effects than existing medications is very important and essential. Medicinal herbs are the most important and under investigation resources for pain relief that have been traditionally recognized as the important sources of treatment by human being. Nowadays due to the ease of access to these medicines, there is high focus on their use and research.

A medicinal plant which has been received much more attention in traditional medicine is turnip. This valuable plant has long had a special place in traditional medicine and in treatment of diseases. Turnip with scientific name of *brassica rapa* is from *Brassicaceae* family. Effective ingredients in this plant are riboflavin, phenols, flavonoids, ascorbic acid, vitamins and potassium. It has therapeutic properties in respiratory, renal, cutaneous, digestive and joint disorders too. It has been stated in other studies that turnip extract has anti-allergic, anti-inflammatory, anti-microbial and anti-cancer properties.

Another herb which has also many therapeutic effects in traditional medicine is valerian root. This plant with the scientific name of *Valeriana officinalis* is an herbaceous plant. Among different parts of the plant, the root and rhizomes are used. Valerian root with valerencic acids and flavonoids is used as a sedative and antispasmodic plant and has been widely used in France, Sweden and Germany for its anti-spasticity and analgesic properties.

The effects of flavonoids available in the extract of valerian and turnip on the synthesis of prostaglandins were clearly indicated. In the process of prostaglandin synthesis from arachidonic acid which occurs in response to inflammatory stimuli, flavonoids were shown to inhibit cyclooxygenase (COX) enzyme preventing sensitization of pain receptors and reduce the pain sensation associated with the response. So this study aimed to compare the analgesic effects of alcoholic extracts of valerian root and turnip in female Wistar rats using formalin test.

**MATERIALS AND METHODS**

Fifty adult female Wistar rats weighing approximately 1905 g prepared from Laboratory Animal Center of Shiraz University of Medical Sciences were divided to 5 equal groups. The control group just underwent formalin test without any other intervention and the sham group received 1 ml of distilled water intraperitoneally, 30 minutes before intraperitoneal injection of formalin. The 1st experimental group received 200 mg/kg of alcoholic extract of turnip root intraperitoneally, 30 minutes before injection of formalin. The 2nd experimental group received 200 mg/kg of alcoholic extract of valerian root intraperitoneally, 30 minutes before injection of formalin. The 3rd experimental group received 200 mg/kg of alcoholic extract of valerian root together with 200 mg/kg of alcoholic extract of turnip root intraperitoneally. 30 minutes before injection of formalin. All animals were kept under identical conditions at 22 °C and a period of 12 hours of light and 12 hours of darkness and fed with adequate water and food as described before.

To prepare turnip extract, 100 g of turnip roots were thoroughly washed and cut, and then ethanol was used three times in the extraction process. The resulting was filtered and completely dried with a vacuum rotary device. The extract was kept in refrigerator before use as reported before. To provide valerian root extract, the roots were changed into powder using electrical mill based on Soxhlet method adding 200 ml of related solvent containing water and ethanol to every 10 g of valerian root powder. The solution was then transferred to Soxhlet device and using a Rota vapor device. The solvent was finally removed from the extract at the end of the process.

All tests were performed between 7:30 AM to 16 PM. Half an hour prior to each experiment, the rats were transferred from cages to glass chambers for formalin test to adopt with their environment. Each animal was tested only once.
and euthanized by diethyl ether at the end of the experiment. For formalin test, 0.05 ml of 2.5% formalin was injected subcutaneously into the right hind paw of the rats as explained before.23

The animals were immediately placed in the test chamber and pain behavior was assessed using a mirror embedded at 45 degree angle to the horizon surface in the lower section of the chamber. Every 15 seconds, the behavioral response was recorded. Animal pain intensity was defined as 4 degrees based on the conventional category as Zero: When the animal had complete balance in walking and the weight was distributed equally on each leg; One: When the animal did not tolerate the body weight on the injected paw and/or when a problem existed with walking; Two: When the animal raised the painful toe and had no contact with the floor of the chamber; and Three: When the animal licked and/or severely shackled the painful paw.

Recording of behavioral responses began immediately after injection of formalin and was continued for 60 minutes. Pain score was calculated as 12 blocks of 5 minutes during 60 minutes period of the experiment according to the formula. Where, T0, T1, T2 and T3 are the number of 15 seconds in which the animal shows, respectively 0, 1, 2 and 3 behaviors in a period of 5 minutes. The time duration of 0-5 and 16-60 minutes were considered as acute and chronic phases for all groups, respectively.

SPSS software (version 18, Chicago, IL, USA) using ANOVA was used for statistical analysis. The statistical difference between groups was considered as P<0.05.

RESULTS

According to the results, a reduction in pain score for both acute and chronic phases was observed in the group receiving turnip extract when compared to the control group. This difference was not statistically significant. A significant reduction in the pain score in acute phase was noted in the group receiving valerian extract when compared to the control group. The simultaneous use of both valerian and turnip extracts could significantly reduce the pain score in both acute and chronic phases when compared to other groups (Table 1).

DISCUSSION

Our results showed a reduction in pain score in the group receiving turnip extract and the group receiving valerian root extract in comparison to the control group. The findings show the analgesic effects of the two extracts. When both extracts were simultaneously used, the analgesic effect was more prominent. It was demonstrated that nitric oxide (NO) plays a crucial role in mediating of many functions in nervous system including the memory formation, sexual feeling, aggressive and nutritional behaviors.24 Nitric oxide as a free radical messenger has high ability of emission by a number of cells. It increases membrane permeability and its role in pain has also been taken into consideration as the level of nitric oxide increases in the injured area following the damage in nervous system.25,26

Nitric oxide is considered as an active messenger in induction of pain. It is synthesized by nitric oxide synthase from L-arginine, while three types of nitric oxide synthase have been identified till now.27 The mechanism of action of nitric oxide in pain process based on activation of neuronal nitric oxide due to tissue damage or inflammation.28 It has been suggested that turnip extract has active biological compounds such as (i) flavonoids including isorhamnetin, kaempferol and quercetin glycosides; (ii) phenylpropanoid derivatives; (iii) indole alkaloids; (iv) sterol

| Group                  | Acute pain     | Chronic pain  |
|------------------------|----------------|---------------|
| 2.5% Formalin          | 2.24±0.097     | 1.72±0.114    |
| 2.5% Formalin+Distilled water | 2.29±0.165   | 1.70±0.039    |
| 2.5% Formalin+Turnip extract (200 mg/kg) | 1.9±0.083   | 1.47±0.112    |
| 2.5% Formalin+Valerian root extract (200 mg/kg) | 1.66±0.133*  | 1.46±0.101    |
| 2.5% Formalin+Turnip extract (200 mg/kg)+Valerian root extract (200 mg/kg) | 1.45±0.132*  | 1.12±0.054*   |

Available mean in each square which has an asterisk (*) indicates significant difference with the control group (p<0.05).
As valerian root extract has flavonoids and phenolic compounds, flavonoids are responsible for inhibition of nitric oxide synthesis as well as synthesis of protein kinase C and prostaglandin E2. The flavonoids were shown to prevent production of NO after injection of formalin leading to an analgesic activity. Since NO is a hyperalgesia mediator, if declines, it would lead to an analgesic property. Similar medicinal compounds and plants with flavonoids were shown to decline inflammatory responses of formalin test by influencing on synthesis of prostaglandins (E2) and their decrease.

The reduction of pain, inflammation and pain in acute phase would decline the plasticity; and practical changes in dorsal branch of the spinal cord and would reduce the release of transmitters such as P substance and excitatory amino acids from the end of nerve fibers similar to our findings. In another study conducted on plants with flavonoids, it was shown that available flavonoids in the plants inhibit pain. As inflammation is associated with production of pain in the second phase of formalin test, so a part of analgesic effects of flavonoids is related to the anti-inflammatory properties of them.

In the current study, valerian root and turnip extracts lead to a reduction in pain score using formalin test, that can be due to the presence of flavonoid compounds and the influence on prostaglandins as well as their anti-inflammatory properties. Similar study on plants with similar ingredients has shown that plants with flavonoids can inhibit cyclooxygenase (COX) in human monocytes and reduce the synthesis of prostaglandin E or thromboxane B2. It was demonstrated that voltage-gate calcium channels play an important role in control of cellular function in various tissues such as heart, blood vessels and nervous system. The evidences indicate that pharmacological blocking of calcium channels may have analgesic effects and be useful in treatment of visceral and somatic pains due to a decline in calcium crossing.

Calcium interferes with release of neurotransmitters and other materials which promote pain and inflammation. The activation of calcium channels is dependent on depolarization of membrane and neurotransmitters, and various materials are released by entrance of calcium causing the pain behavior. Calcium channel blockers are recently taken into consideration as analgesic factors and it has suggested that some blockers of calcium channels have analgesic effect in pre-clinical and clinical models of pain. Previous studies have suggested that flavonoids reduce intracellular concentration of calcium by inhibiting the activity of N-methyl-D-aspartate receptors, activities of nitric oxide synthase enzyme and calcium-dependent phospholipase A2 that are reduced. So flavonoids exert their analgesic effect by reduction in concentration of NO and prostaglandins. The results of our study also denote to a decline in the pain score in the groups received the extracts of valerian root and turnip when compared to the control group. In the case of simultaneous use of the extracts, this effect was more prominent.

Based on the results of our study, the hydroalcoholic extracts of valerian root and turnip in the experimental groups lead to a significant reduction in pain score when compared to the control group using the formalin test. The extracts of valerian root and turnip had anti-inflammatory and analgesic properties that can be due to presence of flavonoid compounds in these plants, the influence of prostaglandins as well as cyclooxygenase enzyme and the decline in intracellular calcium.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

1 Schoenen J, Sandor PS. Textbook of pain. Edinburgh: Churchill Livingstone. 1999.
2 Meier ML, Stämpfli P, Humphreys BK, Vrana A, Seifritz E, Schweinhardt P. The impact of pain-related fear on neural pathways of pain modulation in chronic low back pain. Pain Rep 2017;2:e601.
3 Lechner SG. An update on the spinal and peripheral pathways of pain signaling. e-Neuroforum 2017;23:131-6.
4 Stone AL, Bruehl S, Smith CA, Garber J, Walker LS. Social learning pathways in the
relation between parental chronic pain and daily pain severity and functional impairment in adolescents with functional abdominal pain. *Pain* 2018;159:298-305.
5 Harrison LM, Kastin AJ, Zadina JE. Opiate tolerance and dependence: Receptors, G-proteins and antiopiates. *Peptides* 1998;19:1603-30.
6 Waldhoer M, Bartlett SE, Whistler JL. Opioid receptors. *Ann Rev Biochem* 2004;73:953-90.
7 Law PY, Loh HH, Wei LN. Insights into the receptor transcription and signalling: Implications in opioid tolerance and dependence. *Neuropharmacol* 2004;47:300-11.
8 Payan DG. Nonsteroidal anti-inflammatory drugs, nonopioid analgesics, drugs used in gout. In: Katzung BG, editor. Basic and clinical pharmacology. USA: Appleton and Lange; 1992; pp. 56-98.
9 Chou R, Gordon DB, de Leon-Casasola OA, Rosenberg JM, Bickler S, Brennan T, Carter T, Cassidy CL, Chittenden EH, Degenhardt E, Griffith S. Management of Postoperative Pain: a clinical practice guideline from the American pain society, the American Society of Regional Anesthesia and Pain Medicine, and the American Society of Anesthesiologists’ committee on regional anesthesia, executive committee, and administrative council. *J Pain* 2016;17:131-57.
10 Dowell D, Haegerich TM, Chou R. CDC guideline for prescribing opioids for chronic pain-United States, 2016. *JAMA* 2016;315:1624-45.
11 Huang KCH. The pharmacology of Chinese herb. 2nd ed. USA: CRC Press; 1999; pp. 125-8.
12 Hassanzadeh H. Vegetables growing in garden and home. The first printing, Agricultural-Technical Office, 1991; pp. 174-5.
13 Neal JM, Barrington MJ, Fettiplace MR, Gitman M, Memtsoudis SG, Mörwald EE, Rubin DS, Weinberg G. The third American Society of Regional Anesthesia and Pain Medicine practice advisory on local anesthetic systemic toxicity: executive summary 2017. *Reg Anesth Pain Med* 2018;43:113-23.
14 Toker G, Kupeli E, Memisoglu M, Yesilada E. Flavonoids with antinociceptive and anti-inflammatory activities from the leaves of Tilia argentea (silver linden). *J Ethanopharmacol* 2004;95:393-7.
15 Sahoo S. A review of some medicinal plants used for nervous disorders. *J Med Plant* 2018;6:220-4.
16 Mills S, Bone K. The essential guide to herbal safety. 1st ed. Philadelphia: Elsevier Health Sciences; 2005; p. 616.
17 Samsam Shariat H. Collection of medicinal herbs. 1st ed. Tehran: Char Bagh; 2007; p. 938.
18 DerMarderosian A, Beutler JA. The review of natural products: the most complete source of natural product information. 1st ed. Philadelphia: Facts and Comparisons; 2001; p. 609.
19 Ahmadianai A, Hosseiny J, Semnani S, Javan M, Saeedi F, Kamalinejad M. Antinociceptive and anti-inflammatory effects of Elaeagnus angustifolia fruit extract. *J Ethanopharmacol* 2000;72:287-92.
20 Katzung BG. Basic and clinical pharmacology,6th ed., New York, Conn Appleton and Lange Co. Norwalk, Connecticut. 1995; p. 466.
21 Amos S, Adamu M, Binda L, Edmond I, Kunle OF, Akah P, wambebe C, Gamaniel K. Preliminary studies on anti-inflammatory and anti-nociceptive effects of the aqueous extract of Chrysanthemum indicum. *Acta Pharm* 2002;52:213-18.
22 Mohajeri D, Doustar Y, Mousavi G. Protective and antioxidant activities of turnip root etanolic extract against cisplatin hepatotoxicity in rats. *Zahedan J Res Med Sci* 2011;13:36-44.
23 Stephanie N. Washburn, Brianne C. Patton, Adam R. Ferguson, Kara L. Hudson, James W. Grau. Exposure to intermittent nociceptive stimulation under pentobarbital anesthesia disrupts spinal cord function in rats. *Psychopharmacology (Berl)* 2007;192:243–52.
24 Warembourg M, Leroy D, Jolivet A. Nitric oxide synthase in the guinea pig preoptic area and hypothalamus: distribution, effect of estrogen, and colocalization with progesterone receptor. *J Comp Neurol* 1999;407:207-27.
25 Nilsson BO. Biological effects of aminoguanidin: an update. *Inflamm Res* 1999;48:509-15.
26 Chen D, Pan D, Tang S, Tan Z, Zhang Y, Fu Y, Lü G, Huang Q. Administration of chlorogenic acid alleviates spinal cord injury via TLR4/NFκB and p38 signaling pathway antiinflammatory activity. *Mol Med Rep* 2018;17:1340-6.
27 Munger BL, Bennett GJ, Kajander KC. An experimental painful peripheral neuropathy...
due to nerve constriction. Exp Neurol 1992;118:204-14.

28 Zimmermann M. Pathobiology of neuropathic pain. Eur J Pharmacol 2001;429:23-37.

29 Schonhof I, Krumbein A, Bruckner B. Genotypic effects on glucosinolates and sensory properties of broccoli and cauliflower. Nahrung 2004;48:25-33.

30 Mukherjee P, Cinelli MA, Kang S, Silverman RB. Development of nitric oxide synthase inhibitors for neurodegeneration and neuropathic pain. Chem Soc Rev 2014;43:6814-38.

31 Chen W, Feng L, Shen Y, Su H, Li Y, Zhuang J, Zhang L, Zheng X. Myricitrin inhibits acrylamide-mediated cytotoxicity in human caco-2 cells by preventing oxidative stress. Biomed Res Int 2013;2013:724183.

32 Mehmet O, Yagiz U, Mehmet G. Comparison of the effects of specific and nonspecific inhibition of nitric oxide synthase on morphine analgesia, tolerance and dependence in mice. Life Sci 2003;72:1943-51.

33 Thompson D, Eling T. Mechanism of inhibition of prostaglandin H synthase by eugenol and other phenolic peroxidase substrates. Mol Pharmacol 1989;36:809-17.

34 Dohi T, Anamura S, Okamoto H, Tsujimoto A. Inhibition of lipoxygenase of rat dental pulp and human platelets by phenolic dental medicaments. Dent Jap 1990;27:45-9.

35 Willis WD. Role of neurotransmitters in sensitization of pain responses. Ann NY Acad Sci 2001;933:142-56.

36 Terayama R, Guan Y, Dubner R, Ren K. Activity induced plasticity in brain stem pain modulatory circuitry after inflammation. Neuroreport 2000;11:1915-19.

37 Ramezani M, Amin Gh, Jalili E. Antinociceptive and anti-inflammatory effects of hydroalcoholic extract of Vitex agnus castus fruit in mice. J Shahrekord Univ Med Sci 2010;11:46-51.

38 Lam AN, Demasi M, James MJ, Husband AJ, Walker C. The effect of red clover isoflavones on cox-2 activity in murine and human monocyte/macrophage cells. Nutr Cancer 2004;49:89-93.

39 Todorovic SM, Pathirathna S, Meyenburg A, Jevtovic-Todorovic V. Mechanical and thermal anti-nociception in rats after systemic administration of verapamil. Neurosci Lett 2004;360:57-60.

40 Nouri MH, Mahmoudi J, Hosseinchi Ghareh Aghaji MR. The effect of verapamil, nefedipine and diltiazem on formalin-induced pain response in laboratory small mice. J Vet Med Islam Azad Univ 2009;3:555-63.

41 Davidson EM, Coggeshal RE, Carlton SM. Peripheral NMDA and non-NMDA glutamate receptors contribute to nociceptive behaviors in the rat formalin test. Neuroreport 1997;8:641-6.

42 Kerckhove N, Pereira B, Soriot-Thomas S, Alchaar H, Deleens R, Hieng VS, Serra E, Lanteri-Minet M, Arcagni P, Picard P, Lefebvre-Kuntz D, Maindet C, Mick G, Balp L, Lucas C, Creach C, Letellier M, Martinez V, Navez M, Delbrouck D, Kuhn E, Piquet E, Bozzolo E, Brosse C, Lieter B, Marcaillou F, Hamdani A, Leroux-Bromberg N, Perier Y, Vergne-Salle P, Gov C, Delage N, Gillet D, Romettino S, Richard D, Mallet C, Bernard L, Lambert C, Dubray C, Duale C, Eschalier A. Efficacy and safety of a T-type calcium channel blocker in patients with neuropathic pain: A proof-of-concept, randomized, double-blind and controlled trial. Eur J Pain 2018;22:1321-30.