A summary of the molecular testing recommended in acute myeloid leukemia

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Abstract

Advances in Next-Generation Sequencing technologies (NGS) are revealing germline and somatic mutations that, together with karyotype, determine the diagnosis and subtype of Acute Myeloid Leukemia (AML). Molecular testing is also essential for the genetic risk stratification of patients with AML, in particular for those with normal karyotype AML (CN-AML), a large and highly heterogeneous group of patients. Patients determined to be at high risk could benefit from a more aggressive first-line therapy, or a more directed therapy, such as midostaurin (for FLT3-mutated AML) or ivosidenib (for IDH1-mutated AML). Here, we will summarize the molecular testing currently recommended in AML and introduce new mutations that may have prognostic value and clinical application in the near future.

Introduction

Acute Myeloid Leukemia (AML) is a complex hematological neoplasm, characterized by the World Health Organization (WHO) into multiple subtypes [1,2]. Cytogenetic analysis is essential for AML diagnosis since certain subtypes are characterized by numerical chromosome abnormalities or gross translocations, including loss of chromosome 5 or the common structural aberration t(8;21) (q22;q22), leading to production of the AML-ETO fusion gene, respectively. In fact, according to the National Comprehensive Cancer Network (NCCN), “karyotype represents the single most important prognostic factor for predicting remission rates, relapse risks and overall survival outcomes” [3].

In recent years, significant progress has been made in the biological understanding of AML [4]. In particular, as a result of advances in Next-Generation Sequencing technologies (NGS), genetic studies are revealing an ever-rising number of somatic mutations that help to determine both the phenotypic and prognostic heterogeneity of this pathology [5].

The increasingly widespread use of NGS technology with gene panels in clinics means that the simultaneous analysis of mutations in a high number of genes is both practical and economically feasible [5]. Moreover, the incorporation of molecular markers into current prognostic algorithms is permitting the more precise risk stratification of patients. Indeed, the European Leukemia Net (ELN) currently recommends the evaluation of mutations by NGS using a panel of genes commonly mutated in AML (or a generic myeloid neoplasm gene panel) at diagnosis and at relapse to inform the clinical management of patients with AML [6].

This review will summarize the mutations that currently serve as molecular markers of AML as well as introduce those that may have informative value in the diagnosis, risk stratification and follow-up of patients with AML in the near future.

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Genetic testing recommended in AML

To inform diagnosis

Advances in NGS technologies have driven the inclusion of several new AML subtypes according to their associated genetic alterations, highlighting the importance of molecular testing for a complete diagnosis. For instance, the WHO included the categories of “AML with mutated NPM1” and “AML with biallelic mutations of CEBPA” in 2008 [1], and “AML with mutated RUNX1” and “AML with BCR-ABL1” were provisionally recognized as new entities in the 2016 revision [2]. Although the latter entity is not frequent (less than 1% of all AML cases [7,8]), de novo AML patients identified as having this subtype may benefit from treatment with tyrosine kinase inhibitors [9,10].

Germline mutations

The presence of germline mutations in several genes has been shown to cause a predisposition to myeloid neoplasms, including AML. These observations led to the inclusion of a new category of myeloid neoplasms with germline predisposition (Table 1) in the 2016 revision of the WHO classification of hematological neoplasms [2].

Table 1: Classification of myeloid neoplasms with germline predisposition. Adapted from [2].

| With pre-existing platelet disorders and With other organ dysfunction.          |
|-----------------------------------------------------------------------------|
| AML with CEBPA germline mutation                                            |
| Myeloid neoplasms with germline mutation of DDX41                           |
| With pre-existing platelet disorders                                       |
| Myeloid neoplasms with germline mutation of RUNX1                           |
| Myeloid neoplasms with germline mutation of ANKRd26                        |
| Myeloid neoplasms with germline mutation of ETv6                           |
| With other organ dysfunction                                                |
| Myeloid neoplasms with germline mutation of GATA2                          |
| Myeloid neoplasms associated with bone marrow failure syndromes            |
| Myeloid neoplasms associated with telomere biology disorders               |
| Juvenile myelomonocytic leucemia associated with neurofibromatosis, Noonan syndrome or Noonan-syndrome like disorders |
| Myeloid neoplasms associated with Noonan syndrome                           |
| Myeloid neoplasms associated with Down syndrome                            |

When there is a suspicion that a patient may harbor congenital AML—due to a family history of neoplasia, an exceptionally early age of AML presentation, or the presence of various tumors—the ELN recommends the genetic analysis of the genes RUNX1, CEBPA, GATA2, DDX41, ANKRd26, and ETv6 [6]. Mutations in these genes are just some that bestow a predisposition to the development of AML; others include mutations in genes associated with a general susceptibility to cancer, such as the Fanconi anemia genes, or TP53 and BRCA1/BRCA2 [6,11-13], often mutated in therapy-related leukemias [12].

The analysis of germline mutations is critical to inform the selection of a suitable donor if the patient is a candidate for Hematopoietic Stem Cell Transplantation (HSCT). Germline mutations can also confer a higher risk of relapse in AML or a predisposition to the development of a secondary leukemia, including therapy-related leukemias [12]. For example, between 5% and 7% of patients with CEBPA-mutated AML harbor germline mutations [13] and although the majority reach complete remission following chemotherapy, recurrent disease is frequent [13].

If a positive result is detected, it is recommended that genetic counseling be offered to the patient’s family members for the prevention and/or early detection of germline myeloid neoplasms [11].

The importance of studying TP53

In the case of TP53, the study of mutations in this gene is of particular importance given that their presence predicts a particularly poor outcome for the patient [4,14,15]. Mutations in the TP53 gene locus may be point mutations or deletions of 17p13 of a range of sizes, and either somatic or germline [16].

TP53 mutations usually coincide with a complex karyotype [14,15]. However, their presence in conjunction with a monosomal karyotype or aneuploidies such as −5/5q and −7/7q—themselves associated with high risk—is independent and additive in terms of risk stratification, giving rise to an extremely adverse prognosis [4,15].

Recently, the presence of ASXL1 and RUNX1 mutations were also included in the high-risk category of AML due to their association with a worse prognosis and poor survival, respectively [2]. Germline mutations of the latter are associated with familial platelet disorder with associated myeloid malignancy, a disorder of abnormal hematopoiesis that confers affected individuals with a higher risk of AML [17]. Moreover, germline mutations in ASXL1 [18], have been reported as causing a predisposition to myelodysplastic syndrome and/ or AML. However, further studies are required to confirm the impact of germline mutations in ASXL1, as well as other germline mutations such as SRP72 [19], and CBFα2 [20] before their possible inclusion in congenital AML genetic testing recommendations in the future.

Risk stratification

The genetic risk stratification of patients with AML is especially useful to determine the response to therapy of patients with normal karyotype AML (CN−AML), a particularly heterogeneous group of patients representing 40%-50% of all new cases.

Analysis of FLT3 mutations

The analysis of mutations in the Fms-Like Tyrosine kinase 3 gene (FLT3), including Internal Tandem Duplications (ITD) and mutations in the Tyrosine Kinase Domain (TKD), is recommended by the WHO, NCCN and ELN for patients with CN−AML due to its prognostic value [2,3,6]. Specifically, numerous studies have reported higher relapse rates and worse...
Inhibitor (TKI) gilteritinib has also been approved by the FDA for the treatment of relapsed or refractory FLT3-mutated AML [27].

Despite this, a 2017 survey carried out among AML experts in Europe and the United States reported that only 51.4% of specialists evaluate FLT3-ITD in all patients [28]; while at the national level, a recent survey among AML experts in Spanish clinics revealed that 7.5% “never” or “sometimes” carried out the analysis of FLT3 mutations [29]. These data demonstrate that there is still a lack of understanding of the prognostic and therapeutic indications of the presence of FLT3 mutations.

### Molecular analysis for risk stratification

Since 2017, both the NCCN and the ELN recommend the routine detection of mutations in the genes \( CEBPA, NPM1 \) and \( RUNX1 \) at diagnosis to define the category of AML, as well as mutations in the genes \( KIT, FLT3 \) (ITD and TKD), \( NPM1, CEBPA, RUNX1, ASXL1 \) and \( TP53 \) (together with cytogenetic alterations) in order to refine to prognosis of patients with AML [3,6,30].

In the case of core binding factor AML (CBF-AML), the presence of mutations in the \( KIT \) gene reduces the prognosis from favorable to intermediate risk [31]. Meanwhile, in the case of CN-AML, the presence of mutations in \( NPM1 \) and \( CEBPA \) (in the absence of FLT3-ITD mutations) improves the prognosis from intermediate risk to favorable, although the presence of FLT3-ITD mutations modifies the risk to adverse (Table 2) [6,21–25]. Specifically, FLT3-ITD with a high allelic ratio (≥0.5) is associated with a poor prognosis (higher relapse rate and worse overall survival) [21–25]. However, AML patients mutated in \( NPM1 \) with a low FLT3-ITD allelic ratio (<0.5) have a similar prognosis to patients mutated in \( NPM1 \) without FLT3-ITD; in other words, a favorable prognosis [6,21–25].

It is important that mutational analysis is carried out promptly so that patients identified as high risk can benefit from a more aggressive first-line therapy, or a more directed therapy in some cases. For example, patients with a positive result for FLT3-ITD (with a ratio > 0.5) may be candidates for the addition of the TKI midostaurin to their chemotherapy regime or for HSCT.

### Patient follow-up

#### Relapsed and refractory AML

In addition to its role in the accurate diagnosis of patients with AML, mutational analysis is also very informative in cases of relapsed or refractory AML. The repetition of mutational analysis (because a patient’s profile of mutations may change over time or as the result of chemotherapy or other therapies) may identify possible targets against which there may be an approved directed treatment or an agent undergoing clinical trial. For example, gilteritinib, an inhibitor of FLT3 is approved to treat refractory AML or relapsed patients with mutated FLT3 [27].

Currently, there is a difference between the NCCN and ELN recommendations on molecular testing in AML. Specifically, in

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**Table 2:** Genetic risk stratification according to the NCCN and ELN guidelines. Adapted from [3,6].

| Risk          | Cytogenetics                      | Molecular abnormalities                                      |
|---------------|-----------------------------------|-------------------------------------------------------------|
| Favorable     | CBF: inv(16) or t(16;16); CBFB-MYH11 (t(15;17)(q22;q21); RUNX1-RUNX1T1) | Normal cytogenetics: Mutated NPM1 without FLT3-ITD or FLT3-ITD<sup>TKD</sup> or biallelic CEBPA mutation |
| Intermediate  | Normal cytogenetics: t(9;11); MLLT3-KMT2A; t(8;21), inv(16), t(16;16) | CBF with c-KIT mutation NPM1 mutated with FLT3-ITD<sup>TKD</sup> NPM1 wild-type without FLT3-ITD or with FLT3-ITD<sup>TKD</sup> (without adverse genetic lesions) |
| Adverse       | Complex<sup>x</sup> or monosomal karyotype -5, -5q, -7, -7q t(v,11q23.3); reordered KMT2A; inv(3) or t(3;3); GATA2, MECOM(EVI1); t(6;9); DEK-NUP214; t(9;22); BCR-ABL1 | Normal cytogenetics: NPM1 wild-type with FLT3-ITD<sup>TKD</sup> TP53 mutated RUNX1 mutated ASXL1 mutated |

CBF: Core Binding Factor; ITD: Internal Tandem Duplications; †: high allelic ratio ≥0.5, low allelic ratio <0.5; *: ≥3 chromosomal abnormalities

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the 2017 update the ELN did not include a recommendation for the analysis of mutations in IDH1 and IDH2 in the evaluation of AML at diagnosis due to lack of evidence [6]. However, shortly after its publication, the FDA approved two targeted therapies, ivosidenib (AG-120, Tibsovo®) against IDH1-mutated AML andanidasibdenib (AG–221, IDHIFA®) against IDH2-mutated AML [32, 33]. Thus, in 2018 the NCCN included the recommendation for the analysis of mutations in the IDH1 and IDH2 genes because of the availability of FDA-approved directed therapies [30]. Nevertheless, the detection of DTA mutations in AML patients in remission is not associated with relapse [43].

It still remains to be determined whether the presence of different mutations in patients aged over 65 years of age has the same prognostic value as in patients aged under 65 years, although several groups are actively investigating this matter. For instance, preliminary data from the Alliance group indicates that mutations in the splicing factor SF1 may refine the prognosis of NPM1-mutated AML patients aged over 60 years [44].

Conclusion

Molecular testing at diagnosis, remission and relapse can provide large amounts of data to guide the individualized clinical management of patients with AML. In addition, the analysis of mutations is particularly useful for informing the treatment choice. The ELN recommends the evaluation of mutations in patients with AML at diagnosis and at relapse using NGS technology with gene panels, with which it’s possible to analyze a high number of genes simultaneously and at an ever-decreasing cost.

These panels generate more data than is currently recommended for the diagnosis and prognosis of patients with AML. However, with the continual advancements in our understanding of the impact of somatic mutations and the complex interactions between them, it may be possible to utilize this data in the future to optimize and personalize therapy for patients and in this way maximize their possibilities of reaching complete remission. As such, it is very important to store the samples of patients with AML in biobanks because, although the presence of a certain mutation is not prognostic in the current day, a new targeted therapy may be developed in the future.

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