The role of genetic testing in management of hereditary chronic pancreatitis

Nicholas Faure Walker • Oliver J Warren • Lynsey Gawn • Long R Jiao
HPB Surgery, Department of Surgery and Cancer, Hammersmith Hospital, Imperial College London, London, UK
Correspondence to: Long R Jiao. Email: l.jiao@imperial.ac.uk

This case of a young lady with chronic pancreatitis mimicking pancreatic cancer demonstrates how genetic testing helps surgical management.

Case history

A 26-year-old Gujarati lady, nine months pregnant with her first child, presented to her local emergency department with severe epigastric pain radiating to the back, vomiting and a fever. She had no history of cholelithiasis, was on no regular medication, had never drunk alcohol and had no family history of chronic pancreatitis, cystic fibrosis, hyperlipidaemia or chronic abdominal pain. Investigations showed raised amylase, but normal calcium, lipid profile and renal function. An abdominal ultrasound did not reveal any biliary calculi. Blood cultures were positive for Klebsiella. The patient underwent an emergency caesarean section, delivering a healthy child but required intensive care unit admission for 14 days. A computed tomography (CT) scan showed focal interstitial pancreatitis with extensive calcification, and a bulky inflammatory mass associated with an ectatic pancreatic duct at the tail of the pancreas. She was admitted again two months later with a further bout of acute pancreatitis and recurrent Klebsiella septicaemia, and following discharge experienced ongoing abdominal pain over the next year. When she presented with a third acute flare of pancreatitis she was referred to our regional Hepato-Pancreato-Biliary (HPB) unit for further investigation and management. Upon admission she had a normal full blood count, urea and electrolytes. Her C-reactive protein was 19, amylase 130, anti-neutrophil cytoplasmic antibodies (ANCA) PR3 40 (normal <26), anti-beta-2 glycoprotein 1, IgG 11 (normal <7). Malaria parasites, blood cultures, rheumatoid factor, anticyclic citrullinated peptide antibody (CCP), double-stranded DNA antibody (DSDNA) and HIV tests were all negative. C3 and C4 levels were normal. A CT scan of her abdomen showed signs of acute on chronic pancreatitis with extensive calcification at the body and head of pancreas, and a bulky inflammatory mass associated with an ectatic pancreatic duct at the tail (Figure 1). Endoscopic retrograde cholangiopancreatography (ERCP) was attempted (Figure 2) but it was not possible to insert a stent to bridge the disrupted pancreatic duct at the proximal body of the pancreas. Given the CT appearance, we were concerned that she might have developed pancreatic cancer (PC) at the tail of her pancreas on the background of chronic pancreatitis. Clinically, the patient continued to experience persistent severe epigastric pain refractory to pharmaceutical management. She underwent a laparoscopic distal pancreatectomy with spleen preservation. She was found to have a severely inflamed tail of the pancreas and a giant mass around the tail involving the colonic mesentery, the stomach and the splenic vessels. However, it was still possible to safely dissect the mass from the splenic vessels to preserve the spleen and prevent the sequelae of splenectomy as no definitive diagnosis of cancer was made preoperatively. Histology demonstrated the pancreatic parenchyma to have been completely replaced by fibrous tissue, but no malignant change was identified. Postoperative recovery was unremarkable and the patient was discharged on day 5. At outpatient follow-up six weeks later she was asymptomatic and taking no analgesia. Subsequent genetic testing revealed that she was...
posiive for the serine peptidase inhibitor Kazal type 1 N34S gene mutation (‘SPINK-1 N34S’) but negative for the cationic trypsinogen gene R122H mutation (‘PRSS1 N34S’).

Discussion

Chronic pancreatitis is a destructive inflammatory process which can lead to total destruction of the pancreas with subsequent malabsorption, secondary diabetes and severe unrelenting pain. Alcohol is the commonest causative factor for the development of chronic pancreatitis, accounting for 70% of cases in the UK. However, almost a third of cases are classified as idiopathic as no obvious cause is found. Increasingly, a genetic predisposition is believed to be contributory in this group of patients. The serine peptidase inhibitor Kazal type 1 (SPINK-1) N34S mutation, the cystic fibrosis transmembrane conductance regulator (‘CTFR’) gene and the R122H and N291 mutations in the cationic trypsinogen gene (PRSS1) have all been shown to be risk factors for the development of chronic pancreatitis. SPINK-1 is thought to lower the threshold for the development of pancreatitis by preventing premature accelerated autolysis of trypsin thus reducing trypsin activity by as much as 20%. The N34S mutation has been found in 13–37% of those with intrahepatic cholestasis of pregnancy (ICP) compared with 2.5–5.1% in normal populations. In Threadgold et al.’s study including 91 patients with ICP, 15 (16%) were positive for the N34S mutation. They found the mutation did not statistically affect age of onset, pancreatic exocrine deficiency, duration of illness, frequency of exacerbations, hospital admissions or the need for surgery. However Sandhu et al.’s study from 2011 including 35 ICP patients, of whom 37.1% were SPINK-1 N34S positive, showed that this subgroup were more likely to suffer acute flares compared with controls without the mutation (11.8 [±1.5] versus 4 [±0.98]) over a 9.6 year follow-up. No statistical difference in age of onset, exocrine insufficiency or magnetic resonance cholangiopancreatography or ERCP scoring was demonstrated and none of their cohort developed PC in the time of the study. Piepoli et al.’s 2006 case control study of 61 Italian patients with PC and 106 healthy controls found no relation between SPINK-1, UGT1A7, UGT1A9, ARP or CFTR gene polymorphisms and PC. SPINK-1 has not shown to be an independent risk factor for the development of PC to date.

The risk of developing PC with chronic pancreatitis is difficult to estimate but Malka et al. estimated their cohort (85% with alcoholic chronic pancreatitis) to have a standardized incidence ratio (SIR) of developing PC of 19.0 although earlier studies have shown the SIR to...
be lower. Lowenfels et al.\textsuperscript{8} found in 2006 that the SIR of developing PC in patients with hereditary chronic pancreatitis (HCP) was 50 representing a risk of one in 1066 person years. The EUROPAC study is the largest study of patients with hereditary pancreatitis to date with 527 subjects: 82% of the 399 genetically tested had a PRSS mutation and the R122H was the commonest. They found the cumulative risk of developing PC was 1.5% at 20 years, 2.5% at 30, 8.5% at 40, 14.6% at 50, 25.3% at 60 and 44% at 70 years after the onset of HCP but the risk was not significantly altered by the PRSS mutation status even though 78% of their cohort had a PRSS mutation.\textsuperscript{9}

Chronic pancreatitis is clearly a risk factor in the development of PC but chronic pancreatitis can also present with weight loss, abdominal pain, nausea, diarrhoea and steatorrhoea which mimic PC. PC five-year survival is less than 5% so a low threshold for operative intervention is needed. It can be very difficult to distinguish between malignant and inflammatory masses from chronic pancreatitis on CT. Twelve percent of isolated pancreatic duct strictures identified on CT have been found to be malignant.\textsuperscript{10} The EUROPAC study also found that pancreatic calcification was found more often in those with HCP who went on to develop PC.\textsuperscript{9} The decision to operate is further complicated by the relatively high operative morbidity and mortality.

This was the first patient with chronic pancreatitis to be found positive for the SPINK-1 N34S mutation at our tertiary referral HPB unit in London. The SPINK-1 and PRSS1 gene mutations have been well investigated and should now be offered to patients with chronic pancreatitis of unknown aetiology as they help quantify the lifetime risk of developing PC and hence the need for total pancreatectomy in patients with chronic pancreatitis (Table 1). In this case, the patient was spared TP as she was found to be SPINK-1 N34S positive and PRSS1 gene mutation negative.

---

### Table 1

Summary of gene mutations and their associated risks of pancreatic cancer

| Gene mutation(s) | Risk of pancreatic cancer |
|------------------|---------------------------|
| Serine peptidase inhibitor Kazal type 1 (SPINK-1) N345 | Similar to non-hereditary chronic pancreatitis |
| Cationic trypsinogen gene (PRSS1) R122H and N291 | May represent an increased risk if the patient has hereditary chronic pancreatitis and is over 50\textsuperscript{9} |

---

References

1. Kocher HM, Kadaba R. Chronic pancreatitis. Clin Evid 21 Dec 2011. pii: 0417 (online reference).
2. Whitcomb DC. Inflammation and cancer V. Chronic pancreatitis and pancreatic cancer. Am J Physiol Gastrointest Liver Physiol 2004;287:G315–9
3. Sandhu B, Vitaszka P, Ferreira-Gonzalez A, et al. Presence of SPINK-1 variant alters the course of chronic pancreatitis. J Gastroenterol Hepatol 2011;26:965–9
4. Rinderknecht H, Rinderknecht H. In: Go VLW, ed. The Exocrine Pancreas: Biology, Pathobiology and Diseases. New York: Raven Press, 1986:163–83. (Cited by: Bini L, et al. JOP. J Pancreas [Online] 2007; 8(2):151–155. [Reference 2]). 1986
5. Threadgold J, Greenhalf W, Ellis I, et al. The N34S mutation of SPINK1 (PSTI) is associated with a familial pattern of idiopathic chronic pancreatitis but does not cause the disease. Gut 2002;50:675–81
6. Piepoli A, Gentile A, Valvano MR, et al. Lack of association between UGT1A7, UGT1A9, ARP, SPINK1 and CFTR gene polymorphisms and pancreatic cancer in Italian patients. World J Gastroenterol 2006;12:6343–8
7. Malka D, Hammel P, Maire F, et al. Risk of pancreatic adenocarcinoma in chronic pancreatitis. Gut 2002;51:849–52
8. Lowenfels AB, Maisonneuve P, DiMagno EP, et al. Hereditary pancreatitis and the risk of pancreatic cancer. J Natl Cancer Inst 1997;89:442–6
9. Howes N, Lerch MM, Greenhalf W, et al. Clinical and genetic characteristics of hereditary pancreatitis in Europe. Clin Gastroenterol Hepatol 2004;2:252–61
10. Kalady MF, Peterson B, Balie J, et al. Pancreatic duct strictures: identifying risk of malignancy. Ann Surg Oncol 2004;11:581–8

© 2013 The Author(s) 
This is an open-access article distributed under the terms of the Creative Commons Attribution Non-commercial License (http://creativecommons.org/licenses/by-nc/2.0/), which permits non-commercial use, distribution and reproduction in any medium, provided the original work is properly cited.

J R Soc Med Sh Rep 2013;4:6. DOI 10.1258/shorts.2012.012071