Medicinal Chemistry

Cancer-Cell-Specific Drug Delivery by a Tumor-Homing CPP-Gossypol Conjugate Employing a Tracelessly Cleavable Linker

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In memory of Prof. Dr. Carsten Schmuck 1968–2019

Abstract: Tumor-targeted drug delivery is highly important for improving chemotherapy, as it reduces the dose of cytotoxic agents and minimizes the death of healthy tissues. Towards this goal, a conjugate was synthesized of gossypol and a MCF-7 cancer cell specific CPP (cell penetrating peptide), thus providing a selective drug delivery system. Utilizing the aldehyde moiety of gossypol, the tumor homing CPP RLYMRYSPTTRRYG was attached through a semi-labile imine linker, which was cleaved in a traceless fashion under aqueous conditions and had a half-life of approximately 10 hours. The conjugate killed MCF-7 cells to a significantly greater extent than HeLa cells or healthy fibroblasts.

To date, cancer is one of the leading causes of global death due to the difficulties associated with tumor selective therapy, such as inefficient drug accumulation, cancer cell heterogeneity, and drug resistance.[1] The nonspecific toxicity of anticancer agents towards healthy tissues is a major challenge in conventional chemotherapeutic treatments.[2] Thus, targeted drug delivery, envisioned by Paul Ehrlich as a "magic bullet," is a long-standing research objective.[3] Towards this goal, significant advancements were achieved by exploiting the advantages of different cancer specific vectors,[4] such as antibodies,[5] aptamers,[6] folic acid derivatives,[7,8] and cell penetrating peptides.[9] Tumor-homing peptides, which are small oligopeptides (3 to 15 residues) identified through sophisticated techniques (phage display, mRNA display), are evolving as specific vectors for cancer cells.[10] The design of such targeted drug delivery systems relies majorly on the conjugation of anticancer drugs to the vectors via a cleavable linker. Synthetic organic chemistry aims towards developing suitably cleavable linkers, which release unmodified drugs over a multi-hour timeframe under physiological conditions.[11] Compared to antibodies and aptamers, tumor homing peptides are relatively easy to modify chemically and can be conjugated to drugs through cleavable linkers. Therefore, conjugating novel anticancer agents to tumor homing peptides in order to understand and harness their therapeutic potential is of major interest. Gossypol (AT101), an aldehyde containing phenol derived from the cotton plant, was initially explored as a male antifertility drug. It exhibited promising anticancer activities[12] towards various tumors through different mechanisms including proliferation inhibition and apoptosis induction.[13,14] Its antiproliferative effect is caused by regulating cycline D1,[15] autophagy,[16] and inhibition of aldehyde dehydrogenase.[17] Furthermore Gossypol is studied for combination therapy in addition with other therapeutic agents against glioblastoma,[18] pancreatic cancer cells,[19] and non-small-cell lung carcinoma.[20] It has also shown cytotoxicity against breast cancer cells by inhibiting the expression of mouse double minute 2 (MDM2) and vascular endothelial growth factor (VEGF).[21] Currently gossypol is in phase II clinical trials as an anticancer drug. However, gossypol, as many other conventional anticancer drugs, faces a number of obstacles including bad water solubility, poor cellular uptake and a lack of selectivity. Therefore, reversible attachment of gossypol to a vector that enables cell membrane penetration, increases solubility and allows for addressing cancer cells selectively, seemed highly advantageous.

In this report, we describe the synthesis of cancer cell line specific peptide–gossypol conjugates and their cytotoxic effects. The aldehyde group of gossypol was utilized for conjugation, and thiazolidine (1a) as well as imine linkages (1b) were explored as traceless cleavable linkers (Figure 1). The cancer cell line specific cell penetrating peptide (CPP), RLYMRYSPTTRRYG was developed by Matsushita and co-workers.[9a] It is specific to MCF-7 breast cancer cells and was chosen as gossypol also showed anticancer activity on this cell line. This CPP is internalized into cells through a dynamin-dependent endocytic pathway. Conjugation of this CPP to gossypol increased solubility of the hydrophobic drug (in working buffers: Dulbecco’s
Modified Eagle’s Medium (DMEM) and RPMI medium supplemented with 10% FBS, streptomycin sulfate (0.1 mg mL$^{-1}$), penicillin (10 U mL$^{-1}$) and amphotericin B (0.25 μg mL$^{-1}$), at pH 7.4); this makes the use of potentially harmful solubilizing agents superfluous. Initial attempts to synthesize the thiazolidine linked conjugate 1a were based on standard procedures from the literature, which led to the formation of byproducts among which also was imine 1b. The inseparable mixture of conjugates 1a and 1b obtained from the above reaction exhibited specific toxicity to MCF-7 cells compared to HeLa cells. Time-dependent cytotoxicity assays revealed that imine 1b was much more efficient in killing MCF-7 cells compared to HeLa cells, whereas the thiazolidine-linked conjugate was inactive towards both cells at any time point. A HPLC-study of the cleavage processes revealed uncontrolled degradation of the thiazolidine linked conjugate 1a over time, which was probably caused by reactive oxygen species produced by gossypol. On the other hand, the imine-linked conjugate 1b, cleanly released gossypol over the course of multiple hours, which is an important prerequisite for targeted tumor delivery. Initially, a suitable linker between gossypol and the tumor-homing cell penetrating peptide RLYMYYSPTTRRYG had to be selected, which allows for efficient intracellular release of the anticancer drug. Although several cleavable linkers were reported for different applications, many of them are difficult to incorporate into peptide backbones, while others require specific cleavage conditions such as the involvement of enzymes, nucleophilic/basic or electrophilic/acidic reagents, reducing or oxidizing agents, photoirradiation, organometallic and metal reagents. Furthermore, these methods sometimes leave a residual moiety on the released cargo. Anticancer drugs such as doxorubicin or paclitaxel were linked through ester and amide bonds to peptides, but slow cleavage of these bonds in the cellular environment limits the activity of the drug. Disulfide linkages, which can be cleaved by glutathione in cells, were also reported. However, this strategy is not traceless and the release of the active drug can be inhibited due to drug dimer formation through disulfide bonds. The aldehyde functionality of gossypol opens up the possibility to insert a thiazolidine linkage (1a). In an earlier report a thiazolidine was
employed for traceless release of a drug from an antibody-drug conjugate. An imine-based linkage through an Ala-modified homing peptide was also explored. Gossypol is known to form comparably stable conjugates with amines as well as small peptides, as the resulting Schiff’s base is stabilized by two cooperative intramolecular hydrogen bonds formed by the ortho- and meta-hydroxyl groups.

For this work the tumor homing peptide specific to MCF-7 cells was synthesized by Fmoc-based solid-phase peptide synthesis with a cysteine residue attached at the N-terminus (Figure 2A). The methionine residue was substituted by its analogue, norleucine, to avoid oxidation. Initially, the ligation reaction was attempted between peptide 2a and an excess of gossypol to avoid reaction of the second aldehyde moiety. There-
fore, the peptide was dissolved in 6 M Gn-HCl buffer at pH 5, followed by its addition to five equivalents of gossypol in methanol and the solution was heated at 45 °C for two hours. Consumption of the peptide was observed by HPLC analysis with emergence of two new peaks, corresponding to diastereomers formed by the axially chiral rac-gossypol and the chiral peptide (Figure 2B). The peptide–gossypol conjugates were purified by preparative HPLC followed by evaporation of methanol and lyophilization. Mass spectrometry showed a similar m/z pattern for both peaks, the main signal, however, was 32 Da less than 1a, which corresponds to imine 1b (Figure 2C).

According to mass spectrometric analysis, the product obtained mainly contained 1b but also showed a mass corresponding to 1a. The cytotoxicity of both fractions was tested on MCF-7 cells as target cells and HeLa cells as a negative control. Gossypol itself was equally potent in killing both cell lines (Figure S13, Supporting Information), while the peptide 2a alone was nontoxic for both (Figure S13). Gratifyingly, a specific toxicity towards MCF-7 cells was observed for the CPP-Gossypol conjugate mixture. Cell viability for HeLa cells for mixture (Figure 2E) was higher than for the MCF-7 cells (Figure 2D).

Having evidence for the cell specific toxicity of peptide-gossypol conjugates of type 1, we turned our attention towards deducing the structure of the active conjugate. To exclude that some of the products arose from the simultaneous condensation of the second aldehyde functionality of gossypol with a nearby Arg residue (Figure S9, Supporting Information), we probed the gossypol conjugation reaction with a model tripeptide (CRL) derived from the N-terminus of the peptide 2a. However, only the expected thiazolidine linked conjugate was formed (Figure S10, Supporting Information) as confirmed by mass spectrometry analyses. We also performed a ligation experiment between gossypol and Fmoc-Arg-OH under similar conditions to exclude condensation of the guanidine moiety with the gossypol aldehyde. No conjugation product was observed as evident from analytical HPLC (Figure S11, Supporting Information).

The molecular mass loss of 32 Da compared to the parent Cys containing peptide 1a (Figure S12, Supporting Information) was attributed to desulfurization of the Cys residue to Ala. As reliable thiazolidine formation under these conditions had been described before for other aldehydes, desulfurization during the ligation reaction was most likely promoted by gossypol, which is known to produce reactive oxygen species (ROS).

To confirm the formation of the imine linked conjugate 1b through desulfurization, we synthesized 1b through simple ligation of gossypol and the tumor homing peptide derivative 2b, which was modified with an alanine residue at the N-terminus instead of cysteine. The reaction was also conducted in MeOH and aqueous Gn-HCl-Buffer. It afforded the peptide–drug conjugate 1b with a stable imine linkage as confirmed by HPLC and mass spectrometry (Figure 3, Figure S4). As in this
study preparation of CPP-gossypol conjugates of type 1 was conducted by SPPS, the available material was limited to sub mg amounts. Therefore, imine formation was confirmed by reaction of a model tripeptide NH₂-Ala-Arg-Leu-CONH₂ (ARL) and gossypol. The formation was monitored by HPLC/mass analyses (Figure S5, Supporting Information) and time-dependent ¹H NMR experiments (MeOD/deuterated Gn·HCl buffer). The proton signal at \( \delta = 11.05 \) ppm, which corresponds to the aldehyde moiety, gradually disappeared and a new signal at \( \delta = 9.91 \) ppm, corresponding to imine, appeared after three hours (Figure S7, Supporting Information). The resulting imine was isolated and fully characterized. Furthermore its structure was confirmed by HSQC (¹H-¹³C heteronuclear single quantum coherence spectroscopy, Figure S7, Supporting Information). As mentioned above, the unusual stability of such Schiff’s bases resulting from nucleophilic addition of Gossypol with amines has been described before and can be attributed to stabilization through intramolecular hydrogen bonds as indicated in Figure 3.[25, 26]

The question remained, whether it was the thiazolidine conjugate 1a or its imine congener 1b, which was responsible for the encouraging selective cell toxicity observed in the initial experiments. To prevent desulfurization of the Cys residue during ligation of gossypol to peptide 2a, the reaction was repeated under an argon atmosphere at lower temperature (37°C), which yielded a clean sample of 1a after purification by HPLC (Figure 3c).

The cytotoxic effects of the two differently linked peptide–drug conjugates 1a and 1b were investigated in a time-dependent cytotoxicity assay. Cell viabilities were studied after 12, 24 and 48 hours (Figure S15, Supporting Information). As shown in Figure 4, the imine linked conjugate 1b reduced the cell viability of MCF-7 cells to 26 % after 48 h, whereas both HeLa cells (derived from cervical cancer) and Wi-38 cells (normal human fibroblasts) had a much higher viability of 67 and 74 %, respectively. This underlines the cancer type selectivity of this compound. In contrast the thiazolidine linked conjugate 1a was inactive to both MCF-7 and HeLa cells at any time point. Moreover, the other diastereomeric conjugates separated by HPLC for both linkages showed a similar trend of cytotoxicity, that is, the imine linked conjugate of type 1b was potent to selectively kill the MCF-7 cells, while the thiazolidine linked conjugate of type 1a was not (Figure S14, Supporting Information).

The time-dependent gossypol release of both, the active imine based conjugate 1b and the inactive thiazolidine based conjugate 1a were studied in 6 mM Gn·HCl buffer at pH 7 and 37°C through time dependent HPLC analysis. The imine linked conjugate 1b had a half-life of approximately 10 hours and cleanly released gossypol as well as the homing peptide 2b (Figure 5). On the other hand, the thiazolidine linked conjugate 1a decomposed to several unidentified species after two hours (Figure S17, Supporting Information). This observation is consistent with the observed inactivity of the thiazolidine linked conjugate in both cell lines. Assuming a similar stability for 1b in the cytoplasm, the measured half-life of 10 h offers a sufficient period for intracellular accumulation of the peptide–drug conjugate 1b before the controlled release of the anticancer drug within MCF-7 cells. This leads to substantial cell death after 24 and 48 h.
In summary, we have developed a cancer cell specific delivery system for gossypol by using simple Schiff's base ligation chemistry to generate a semi-labile conjugate of Gossypol and a cancer type specific cell penetrating peptide as a vector. Imine formation between gossypol with the Alanine functionalized CPP 2b resulted in a potent conjugate, which killed specifically MCF-7 breast-cancer cells. Furthermore, the solubility of gossypol was improved, which made handling of the conjugates for cellular studies very convenient, as potentially harmful solubilizing agents like DMSO were not necessary. Importantly, the presented drug delivery strategy does not rely on any external stimulus to initiate drug release and activation. A particular advantage of this imine linkage is the convenient half-life of 10 hours in aqueous media. Hopefully these results accelerate the applicability of gossypol particularly in tumor targeted chemotherapy. In future research cell line-derived xenograft (CDX) mouse models should be established to evaluate the in vivo efficacy of the CPP-gossypol conjugate 1b. In a broader sense, the reported approach demonstrates that cell penetrating peptides with tumor homing properties can be easily ligated to gossypol without the need for an additional linker. Therefore, it should be easy to expand the scope of this approach to other cancer types, for which appropriate homing peptides can be identified.

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Conflict of interest

The authors declare no conflict of interest.

Keywords: cleavable linker • drug delivery • gossypol • imine linkage • tumor-homing peptide

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