Hematological reference intervals among full-term newborns in Ethiopia: a cross-sectional study

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Abstract

**Background:** Hematological reference intervals are used for medical decision tools for interpretation of numerical test results. Establishing of hematological interval among newborn babies is very important for the diagnosis of malignancy, anemia, bleeding disorders, and various infections. There is no locally established hematological reference intervals in Ethiopia. Thus, the aim of this study is to established locally determined hematological reference interval among full-term newborns.

**Methods:** A cross sectional study was conducted from February to May 2019 among 151 apparently healthy full-term newborns at Gondar University Hospital. About 3ml of cord blood was obtained for analysis of Hematological parameters and determined by using Sysmex KX-21N (Sysmex Corporation Kobe, Japan) automated analyzer. Median, 2.5th and 97.5th percentiles were computed.

**Result:** Males to female's ratio was almost equal. All hematological parameter had no statistically significant difference between males and females. The delivery types were not influenced its hematological values. The reference interval of white blood cells, red blood cells, platelets, hemoglobin, hematocrit, mean cell volume, and mean cell hemoglobin were (7.64-22.16) x10^9/l, (3.69-5.47)x10^12/l, (132.74-413.4) x10^9/l, (13.32-19.64) g/dl and (39.42-58.06)%,(91.6-113.22)fl, and (30.48-38.02)pg, respectively.

**Conclusion:** All hematological reference interval was established from full-term newborns at University of Gondar hospital was different from other studies in Nigeria, Iraq, Pakistan, Nepal, Saudi Arabia and Iran. Therefore, own determined reference value is very important for the clinicians to correctly diagnosis the patients at health facility levels.

Introduction

**Background**

Umbilical cord blood (UCB) is the blood remaining after in the excluding of the placenta. It is considered as biological waste products. However, it has many advantages like stem cell transplantation (rich in hematopoietic progenitor and stem cells), Rhesus factor (Rh) blood typing of the newborns, assessment of the neonatal thrombocytopenia or thrombosis, screening and diagnosis of anemia, polycythemia, bacterial sepsis and determination of hematological reference interval (RI) (1-3). The other main advantage of UCB is to minimize blood draw from very ill newborns that cause increased morbidity and mortality during neonatal life. The UCB is the alternative source to eliminate unwanted repeated blood drawn for laboratory tests. The collection of UCB is safe for the patients and technically easy as compared to other blood collection sites (4).

The UCB is composed of white blood cells (WBC), red blood cells (RBC), plasma, platelets and is also rich in hematopoietic stem cells, which have immense potential to cure the malignant and genetic disorder. Therefore, establishing of hematological RI among newborns from UCB is very important for
neonatologist for the diagnosis RBC disorders (like anemia and polycythemia), hematological malignancy, platelet disorders (like thrombosis and thrombocytopenia), autoimmune disease, certain genetic disorders problems, monitoring of the patients during in a therapeutic regimen and diagnosis of various bacterial infections (5-7). It is widely known that newborn hematological parameters are different from those of infants or adults. Therefore, separated hematological RI is mandatory for newborns (8).

The RI is the range between values from the lower reference limit to the upper reference limits (9). They are ideally defined on apparently healthy individuals and should be distinguished from clinical decision limits that are derived from known diseased patients. It is a distribution of numerical test results expected in a representative population of healthy individuals. It is mainly used to make medical decisions about hematological disease diagnosis, treatment & monitoring of disease prognosis of the patients (10). According to Clinical Laboratory Standards Institute guideline, the RIs are defined in relation to a healthy population to include the values in which 95% of apparently healthy individuals would fall and in which 2.5% of results in the lower range are out of the RI and 2.5% of values in the upper range will be out of the RI (11, 12).

Hematological RIs are the most common medical decision-supporting tools used for the interpretation of numerical hematological test results. Accurate patient results interpreting within the correct RI is mandatory to minimize the patient risks from disease, increase recovery rates, and improve monitoring therapy of various hematological disorders (13). Almost 80% of physicians’ medical decisions are based on information provided by laboratory reports. Therefore, locally established hematological RI among newborns is very important for the physician to correctly diagnosis and early treatment of the patients (14).

In Ethiopia there is no locally determined hematological RI among healthy full-term newborns in the country, particularly is in the study area. They used in Western and American RI. Since hematological RI is varying through age, race, ethnicity, altitude, sex, drug intake, time of sampling and socioeconomic status. It is commonly known that newborn hematological parameters are different from those of neonates, infants or adults (8). It need locally determined RI. Due to these reason we motivate to conduct this study. The aim of this study was to establish hematological RI by using UCB of apparently healthy, full-term newborns at University of Gondar compressive specialized hospital, northwest Ethiopia.

**Method**

**Study Setting, design and population**

An institution-based cross-sectional study was conducted from February 1 to May 30, 2019, at the gynecology and obstetrics department of University of Gondar comprehensive specialized hospital, Northwest Ethiopia. The hospital is located in Gondar town Amhara regional state, northwest Ethiopia. The town is located 2,133 meters’ elevation above sea level. The hospital provides both teaching and referral center in the region, which services more than five million people. Currently, the hospital holds
more than 550 beds, and it handles approximately 8000–9000 deliveries per year. The hospital has a range of specialties including pediatrics, surgery, gynecology, psychiatry, human immunodeficiency virus (HIV) care, and an outpatient clinic.

The newborns were selected based on the following criteria: Birth at full-term (39-42 weeks of gestation), and absence of any congenital anomalies. All selected newborns were physically examined at birth and found normal and apparently healthy. The premature newborn (delivered less than 37 weeks of gestation), twin newborns, the pregnancy complicated with diabetics, preeclampsia, hypertension, HIV/AIDS, chronic kidney, liver disease, malaria, anemia, and hematological malignancy were excluded from the study. On the other hand, a mother who had bleeding during pregnancy, maternal drinking of alcohol during pregnancy, cigarette smoking during pregnancy, and no antenatal care also were excluded from being sampled. A systematic random sampling technique was employed to select study participants. The sample size is determined based on national Committee for Clinical Laboratory Standards, International Federation of Clinical Chemistry and Clinical Laboratory Standards Institute guideline recommendations, a minimum size of 120 observations is required for determination of reference intervals (15). Study participants were selected every three intervals based on flow of delivery to give equal allocation of study participants. The excluded study participants were substituted with the next consecutive study participants. A total of 202 study participants were selected during the study periods. However, 43 were excluded due the presence of maternal anemia, 5 were excluded due to twins and the remained 3 newborns were still birth. Finally, a total of 151 newborns with their respective mothers were included and their results were analysis in SPSS.

Data collection method

Socio-demographic and clinical data collection

A pre-tested structured questionnaire prepared in English and translated to the local language (Amharic) was used to obtain newborn gender, birth weight, and presence of bleeding during pregnancy, alcohol consumption habits, and cigarette smoking during pregnancy via face-to-face interviews. The presence of maternal complications like malignancy, hypertension, diabetics, tuberculosis, HIV/AIDS, chronic kidney, and liver disease were retrieved from maternal medical records with the aid of data extraction sheet.

Blood collection and laboratory analysis

About 3ml of UCB specimen were obtained from each study participants after delivery from the clamped umbilical cord. The two trend Midwifery professionals collected the cord blood sample from the clamped cord through excluding of the placenta. The collected sample was immediately poured into tri-potassium ethylene diamine tetra acetic acid (K3-EDTA) test tube and gently mixed to prevent blood clotting. In addition, 3ml of venous blood was collected from the mother after delivery with a sterile and disposable
syringe. Hematological parameters: total white cell count (WBC), differential white cell count (neutrophils, lymphocytes and mixed which contains eosinophils, monocytes and basophiles), platelet count, red blood cell count (RBC), Hgb, hematocrit (%), mean cell volume (MCV), mean cell hemoglobin (MCH), mean cell hemoglobin concentration (MCHC), and red cell distribution width (RDW) were determined by using the using Sysmex KX-21N (Sysmex Corporation Kobe, Japan) automated hematological whole blood analyzer based on direct current principle. Three experienced laboratory technologist performs the complete blood count (CBC) by strictly adhering standard operating procedures.

Data quality control

In order to increase the reliability of data, training was given for the data collector prior to data collection. During laboratory data collection standard operating procedures were strictly followed and implemented from prior to specimen collection up to recording and interpreting of the laboratory results. The collected sample were immediately dispensed to the wall of the EDTA test tube slowly and properly mixed by inverting the tube gently 8-10 times to prevent hemolysis of blood samples. Then label of the sample and the request paper with the same identification number in order to avoid any mix up of errors. The expired date of reagent was checked before analysis of patient samples. Daily installations and background run were done to minimize any background errors. Repeated analysis of randomly selected samples for reproducibility check (delta check) was carried out three times a week to evaluate instrument performance consistently and precisely.

Statistical analysis

The data were cleaned, edited, checked for completeness, and entered into Epi-info version 7.2.1.0. Then it was exported into SPSS version 20 for analysis. Outliers were detected by using Z score and Tukey method. Before any analysis, normal distribution of numerical data was checked using the Kolmogorov-Smirnov and Shapiro-Wilk test. Since all data are not Gaussian distributed we used non-parametric tests to determine the hematological RI as recommended as the clinical laboratory standard institutes (12). Median with interquartile range (IQR), and 95% confidence interval were computed. The 97.5th percentile and 2.5th percentiles were the upper and the lower reference limit of the study populations, respectively. Mann-Whitney U test also used to test for the mean difference of variance between gender (male versus female), and delivery type (vaginal versus cesarean section). According to the guideline of clinical laboratory standard institutes, we determine the 95% hematological RI by computing 97.5th percentile and 2.5th percentile.

Results

Sociodemographic and obstetric data of study participants
A total of 151 healthy newborns (76 females and 75 males) were included in this study. Majority of study participant (87.4%) were born from mother came from urban residence with normal birth weight (90.1%). About 81.5% of study participants were born through spontaneous vaginal delivery route and the remaining were by cesarean sections (Table 1).

The minimum vs maximum value of cord WBC, cord RBC, and cord platelet among study participants were (3.6 vs 27) x10^6/L, (3.02 vs 5.7) x10^{12}/L, (132 vs 500) x10^9/L, respectively. As a result, summarized in table 2 sex and delivery type had no influence for cord hematological values (p-value >0.05). The median value of cord WBC, RBC, Hgb, and Hct among male newborns were 12.6, 4.7, 16.2 and 49.6, respectively. On the other hand, the median value WBC, RBC, Hgb, and Hct among female newborns were 13.15, 4.6, 15.75, and 48.65, respectively (Table 2).

Hematological reference interval

The newborns hematological RI is determined based on non-parametric estimation methods. Based on the SPSS statistical analysis of newborn laboratory data did not require partition by sex groups. Based on the finding, all hematological value did not have a statistically significant difference of means among sex groups as well as the route of delivery types (table 2). According to clinical laboratory approve guideline partitioning was required when the difference between the observed means of two subclasses is statistically significant at the 5% probability level (9). The median and 95% of RI of RBC, Hgb, Hct, MCV, MCH and MCHC were 4.64 (3.69-5.47 x10^{12}/l), 16 (13.32-19.64), 49.1% (39.42-58.06%), 105.85fl (91.6-113.22fl), 35pg (30.48-38.02pg) and 33.1g/dl (31.48-36.5g/dl), respectively. The lower and upper limit WBC and platelets were 7.64, 22.16, and 132.74, 413.4, respectively (Table 3).

Discussion

This study tries to find out the RI among newborns by using UCB which was the first time reported in Ethiopia. The current finding showed that there was no statistically significance difference of all hematological parameters between male and female newborns. which was consistent with a study reported in Sokoto, Northern Nigeria (16) and Lagos, Nigeria (17). The present study also showed that mode of delivery had no influence of hematological parameters of the newborns which agrees to a study done by Fady M et al reported that there was no significant difference in the MCV, MCH, MCHC, RDW, lymphocytes, and monocytes (21). However, this result is contrasting to a study done by L Glasser et al showed that mode of delivery had influenced to hematological test values (22). This variation might be due related to low sample size used in the current study (151 versus 10,287 study participants)

The of Hgb values (13.3-19.6g/dl) of this study almost agrees with a study reported in Saudi Arabia (15.7-19.7g/dl), however, the upper limit 97.5th percentiles of platelet value (413 x10^9/l) the present study was greater than as compared to the upper limit of Saudi reported (297 x10^9/l) (23). On the other hand, Our finding of the 95% RI of RBC (3.69-5.47) values were comparable to a study reported in Nepal (3.67-
4.93) (18), Iran (3.61-5.29) (19), and Nigeria (3.52-4.62) (20) reported. Similarly, the 95% RI of WBC (7.64-22.16 x10^9/l) in our study similar to the study conducted in Turkey (71.90–25.44 x10^9/l) (24).

The lower limit WBC (7.64) value of this study was lower than a study reported in Saudi Arabia (10.9) (23), Nepal (10.49) (18) and Pakistan (9.7) (25). On the contrary, the lower limit of WBC was higher than a study reported in Sagamu, Nigeria (4.2) (20) and Iran (5.16) (19). However, the MCH RI (30.5-38.02) were similar to a study done in Saudi Arabia (30.2-40.6) (23), Nepal (31.6-36.2), (18), and Pakistan (32.2-35.4) (25). On the other hand, the Hct value (39.4-58.1) of the current study was comparable to a study conducted in Iran (39.6-56.9) (19). (This may be due to the variation in geographical location, and race or genetic factors may be contributing to the presence of this difference.

The lower limit of RBC (3.69 x10^9/l) in this finding was lower than studies conducted in Saudi Arabia (5.1 x10^9/l) (23) and Sagamu Nigeria (4.3 x10^9/l). On the contrarily, the higher limit of RI of Hgb value (19.6g/dl) of this study was consistent to a study reported in Saudi Arabia (19.7g/dl) (23) but higher than in a study reported in Logas Nigeria (14.8g/dl) (17), Pakistan (17.3g/dl) (25), Iraq (15.22g/dl) (26), and Nepal (17.2g/dl) (18). The difference of Hgb value might be due to method variation, inclusion criteria of the study participants and time of cord clamping. Similarly, the higher limit of 95th percentile RI of both WBC and platelet values were higher than in the study conducted in Nepal, Pakistan, Logas Nigeria, Iran and Iraq (17-19, 25, 26).

The platelet counts of this study range from (132.7-413) x10^9/L. Based on our finding less than 132 x10^9/L is thrombocytopenia and greater than 413 x10^9/L is thrombocytosis for newborns. The lower limit of platelet counts in the current study (132.7 x10^9/L) were consistent to a study reported from Mashhad, Iran (131 x10^9/L) (19). However, the higher limit (413 x10^9/L) of the current study was higher in this study reported by Marwaha et al (142x10^9/L) (27). The major limitation of this study was nucleated RBC is not corrected when total WBC was counted at the time of analysis.

**Conclusion**

there was no statistical significance of RI of all hematological value between male and female newborns. The of Hgb values of this study almost agrees with a study reported in Saudi Arabia. The RI of RBC of the current study was comparable to Nepal, Iraq and Nigeria findings. However, the lower limit of WBC was lower than a study reported in Saudi Arabia, Nepal and Pakistan. The current finding of RI is very helpful for the neonatologist to accurately diagnose and monitoring of therapies of the patients.

**Abbreviations**

CBC: Complete Blood Count, Hct: Hematocrit, Hgb: Hemoglobin, MCH: Mean Cell Hemoglobin, MCHC: Mean Cell Hemoglobin Concentration, MCV: Mean Cell Volume, MPV: Mean platelet volume, PDW: Platelet distribution width, RBC: Red Blood Cell, RDW: Red Cell Distribution Width, Rh: Rhesus factor, RI: Reference Interval, WBC: White Blood Cell.
Declarations

Ethical approval and consent to participate

The research was conducted after obtaining an ethical clearance letter from the School of Biomedical and Laboratory Science Ethical Review Committee. A permission letter was obtained from the University of Gondar compressive specialized hospital. Written informed consent was obtained from each selected mother after explaining the aim of the study with the potential adverse effects during sample collection. The results of the study participants were used without personal identifiers and only used for research purpose.

Consent for publication

Not applicable

Availability of data and material

The datasets used and/or analyzed during the current study available from the corresponding author on reasonable request.

Competing of interest

The authors declare that they have no competing interests.

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Authors contribution

TT involved in the study design, data collection, laboratory analysis, result interpretation and drafting of the manuscript. TK and SG involved in statistical analysis, editing and reviewing of the manuscript. All authors read and approved the final manuscript.

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**Tables**

Table 1: Sociodemographic characteristics of study participants at University of Gondar compressive specialized hospital, 2019 (N= 151)
| Variable                  | Frequency | Percentage (%) |
|--------------------------|-----------|----------------|
| Gender                   |           |                |
| Male                     | 75        | 49.7           |
| Female                   | 76        | 50.3           |
| Residence of mother      |           |                |
| Urban                    | 132       | 87.4           |
| Rural                    | 19        | 12.6           |
| Occupation of the mother |           |                |
| Employed                 | 54        | 35.8           |
| House wife               | 57        | 64.2           |
| Birth weight             |           |                |
| Normal (>=2.5kg)         | 136       | 90.1           |
| Under birth (<2.5kg)     | 15        | 9.9            |
| Gravidia                 |           |                |
| Primigravidia            | 65        | 43             |
| Multigravidia            | 86        | 57             |

Table 2: The influence of sex and delivery type on hematological parameter in newborns based on median value

| Hematological parameter | Sex          | Delivery type |
|-------------------------|--------------|---------------|
|                         | Male | Female | P-value | Vaginal | Cesarean | p-value |
| WBC (x10^9/l)           | 12.6 | 13.15  | 0.98    | 12.6    | 13.1     | 0.64    |
| RBC (x10^12/l)          | 4.7  | 4.6    | 0.51    | 4.61    | 4.73     | 0.27    |
| Hgb (g/dl)              | 16.2 | 15.75  | 0.47    | 15.8    | 16.6     | 0.09    |
| Hct (%)                 | 49.6 | 48.65  | 0.44    | 48.6    | 50.45    | 0.05    |
| Platelet (x10^9/l)      | 292  | 271    | 0.17    | 276     | 274      | 0.62    |
| Lymphocyte absolute (x10^9/l) | 4.5 | 4.6    | 0.12    | 4.6     | 4.55     | 0.91    |
| Neutrophil absolute ((x10^9/l) | 6.4 | 7.25  | 0.19    | 7       | 6.95     | 0.57    |

Table 3: Complete blood count parameter among full-term newborns at gynecology and obstetrics, University of Gondar compressive specialized hospital, 2019 (N=151)
| CBC parameter                      | Median (IQR)       | Lower limit (2.5\textsuperscript{th}) | Upper limit (97.5\textsuperscript{th}) | RI (95\textsuperscript{th} percentile) |
|-----------------------------------|--------------------|----------------------------------------|----------------------------------------|----------------------------------------|
| Total WBC (x10\textsuperscript{9}/l) | 12.8 (10.5-15.4)  | 7.64                                   | 22.16                                  | 7.64-22.16                             |
| RBC (x10\textsuperscript{12}/l)    | 4.64 (4.25-4.89)  | 3.69                                   | 5.47                                   | 3.69-5.47                              |
| Hgb (g/dl)                        | 16 (14.8-17)      | 13.32                                  | 19.64                                  | 13.32-19.64                            |
| Hct (%)                           | 49.1 (44.9-51.8)  | 39.42                                  | 58.06                                  | 39.42-58.06                            |
| MCV (fl)                          | 105.85 (102.6-109.6) | 91.6                                   | 113.22                                 | 91.6-113.22                            |
| MCH (pg)                          | 35 (33.7-35.7)    | 30.48                                  | 38.02                                  | 30.48-38.02                            |
| MCHC (g/dl)                       | 33.1 (32.5-33.5)  | 31.48                                  | 36.5                                   | 31.48-36.5                             |
| Platelet (x10\textsuperscript{9}/l) | 276 (233-314)     | 132.74                                 | 413.4                                  | 132.74-413.4                           |
| Lymphocyte percentage (%)         | 35.9 (30-41)      | 14.22                                  | 58.32                                  | 14.22-58.32                            |
| Mix percentage (%)                | 10.3 (7.9-12.5)   | 3.34                                   | 18.64                                  | 3.34-18.64                             |
| Neutrophil percentage (%)         | 53.5 (48.5-60.7)  | 36.74                                  | 79.48                                  | 36.74-79.48                            |
| Lymphocyte absolute (x10\textsuperscript{9}/l) | 4.6 (3.6-5.5)     | 2.16                                   | 10.3                                   | 2.16-10.3                              |
| Mix absolute (x10\textsuperscript{9}/l) | 1.3 (0.9-1.7)     | 0.4                                    | 2.94                                   | 0.4-2.94                               |
| Neutrophil absolute (x10\textsuperscript{9}/l) | 7 (5.4-8.9)       | 2.96                                   | 13.54                                  | 2.96-13.54                             |
| RDW-SD                            | 68.1 (64.7-72.1)  | 55.48                                  | 81.3                                   | 55.48-81.3                             |
| Platelet distribution width (PDW) | 11.3 (10.5-12.2)  | 8.8                                    | 15.68                                  | 8.8-15.68                              |
| Mean platelet volume (MPV)        | 9.5 (9.1-10)      | 7.88                                   | 11.04                                  | 7.88-11.04                             |

Table 4: Comparison of the current study to other similar findings based on cord blood hematological parameters
| Hematological parameter | northwestern Ethiopia (N=151) | Saudi Arabia (N=2163) | (21) | Nepal (N=210) | Pakistan (N=316) | (23) | Lagos, Nigeria (N=130) | Mashhad, Iran (N=447) | (19) | Iraq (N=220) | Sagamu, Nigeria (N=108) |
|--------------------------|-----------------------------|----------------------|------|---------------|------------------|------|----------------------|----------------------|------|---------------|------------------------|
| Total WBC                | 7.64-22.16                  | 10.9-21.5            | 10.49-19.4 | 9.7-17.7 | 7.9-18.3 | 5.16-18.2 | 7.32-12.92 | 4.2-25.8 |
| RBC                      | 3.69-5.47                   | 5.1-16.3             | 3.67-4.93 | -         | 3.52-4.62 | 3.61-5.29 | 3.53-4.47 | 4.3-5.9  |
| Hgb                      | 13.3-19.6                   | 15.7-19.7            | 13.28-17.2 | 13.5-17.3 | 11.8-14.8 | 13-18.8  | 12.3-15.22 | 13-17.8  |
| Hct                      | 39.4-58.1                   | 46.9-59.5            | -     | -           | 39-50.6  | 39.6-56.9 | 39.7-49.2 | 39.1-53.5 |
| MCV                      | 91.6-113.2                  | 97.9-112.3           | 95.2-107.2 | 103.8-108 | 98.5-122.2 | 97.5-119.8 | 105.5-117.7 | 84.4-98.8 |
| MCH                      | 30.5-38.02                  | 30.2-40.6            | 31.6-36.2 | 32.2-35.4 | 28.5-36.7 | 31.7-40  | 32.1-36.8 | 27.9-33.1 |
| MCHC                     | 31.48-36.5                  | 27.40-2          | 31.7-34.9 | -         | 28.1-31.4 | 30.1-35.2 | 29.03-32.86 | 31.7-34.5 |
| Platelet                 | 132.7-413                   | 178.2-297           | 165.6-288.2 | 223-347 | 152.9-297.3 | 131-383 | 207-328 | 129-607  |
| Lymphocyte %             | 14.2-58.3                   | 16.5-37.1           | 23.7-46.7 | -         | -         | -         | 29.6-49.98 | 27.8-49.6 |
| Neutrophil percentage    | 36.7-79.5                   | 50.3-74.1           | 51.5-74.7 | -         | -         | -         | 39.8-62.2 | 45.9-67.7 |
| Lymphocyte absolute      | 2.16-10.3                   | -                   | -       | 3.3-6.9 | -         | 1.14-8.46 | -         | -        |
| Neutrophil absolute      | 2.96-13.54                  | -                   | -       | 4.7-10.7 | -         | 1.12-10.2 | -         | -        |

**Supplementary Files**

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- Additionalfile1.docx