Is it possible to govern the movement of the transcription bubbles of DNA by constant and periodic external fields?

Abstract

The influence of constant and periodic external fields on the living systems can lead to various changes in their functional properties. At the molecular level, these changes can be modeled as the changes in the dynamic properties of biomolecules. In this paper we investigate the effect of constant and periodic fields on the movement of transcription bubbles being small unwound regions (~10bp) in the DNA double helix, which result from the interaction of the RNA polymerase with the DNA promoter region at the initiation stage of the process of transcription. The bubbles are modeled mathematically by kinks being one–soliton solutions of the modified sine–Gordon equation. To obtain these solutions and calculate the dynamic characteristics of the bubbles including their velocity, coordinate, phase portrait, total energy and size, we apply the method of McLaughlin and Scott and quasi–homogeneous approximation. For definiteness all of the calculations were made for the pTTQ18 sequence. The results were used to analyze the bubbles dynamic behavior and to answer the question: is it possible to govern the movement of the transcription bubbles of DNA by constant and periodic external fields? Possible consequences on the gene expression and biological activity of cells are discussed.

Keywords: transcription bubble, DNA torque, periodic fields, sine–Gordon equation, method of McLaughlin and Scott, quasi–homogeneous approximation, nonlinear DNA dynamics

Abbreviations: DNA, deoxyribonucleic acid; AOT, angular optical trap; MTT, magnetic tweezers technique

Introduction

The action of constant and periodic fields on the living systems is one of the hotly discussed themes of modern biophysics because of the growing amount and variety of electronic devices and their influence on the basic, fundamental life processes and, consequently, on human health. It is assumed, in particular, that external periodic fields with the frequency of terahertz diapason can cause the changes in the physicochemical properties of DNA, which in turn can lead to the changes in the transcription and thus to the changes in the gene expression and cell differentiation. However, until now the mechanisms of the action of these fields remain unclear. Special attention is also paid to the influence of constant fields because of significant progress in the experimental studies of the dynamics of single molecules of DNA and by the development of new techniques that allows to measure directly the DNA torque. The method of the angular optical trap (AOT) and the method of magnetic tweezers technique (MTT) are among them. However, these and other currently existing experimental methods do not allow measure the DNA torque directly in the biological processes in which the DNA molecule is involved.

In this paper we apply the methods of mathematical modeling to investigate the action of constant and periodic fields on the movement of transcription bubbles of DNA which are small unwound regions (~10bp) in the DNA double helix resulting from the interaction of the RNA polymerase with the DNA promoter region at the initiation stage of the process of transcription (Figure 1). Our aim is to clarify the mechanisms of the influence of the fields on the bubbles dynamics, to find the relationship between the DNA torque and the bubble velocity, to estimate the value of the DNA torque necessary for the moving of the bubbles with the velocity of the process of transcription, to find the relation between the dynamic behavior of the bubbles and their initial velocities, and to give an answer to the question is it possible to govern the movement of the transcription bubbles of DNA by constant and periodic external fields (Figure 1).

Figure 1 Locally unwound region (bubble) in the DNA double strand, which is formed at the initial stage of transcription.

In physics, the DNA molecule is considered as a complex dynamic system consisting of a large number of coupled atoms and atomic groups which are arranged in a certain way in space. Such a system is not static, but movable. It has a large amount of internal motions...
caused by the effect of temperature, collisions with the molecules of the solution, and interactions with proteins. Among the variety of the internal motions of DNA there are the translational movements of transcription bubbles that can be considered as translational movements of quasi–particles in the potential field of the DNA.

Mathematically the transcription bubbles can be described as the soliton–like solutions of the nonlinear differential equations imitating the internal DNA mobility. In the case of homogeneous synthetic DNA having the sequence of identical bases, Englander and co–authors showed that the bubble movement can be modeled by the sine–Gordon equation with constant coefficients, this equation having the exact one–soliton solutions in the form of kinks. In the work of Englander and co–authors as well as in the works of other authors developing the Englander’s idea, just these solutions were used to simulate the DNA open states or bubbles. To take into account effects of dissipation and the action of some external field, they modified the sine–Gordon equation by adding the following two terms: one to model effect of dissipation and the other to model influence of an external field. With the help of the method of McLaughlin and Scott the kink–like solutions of the modified sine–Gordon equation were found in the two particular cases: when the external field is constant and when the external field is periodic.

In the case of inhomogeneous DNA, the coefficients of the modified sine–Gordon equation are no longer constants, but depend on the sequence of bases (Table 1). However, in the quasi–homogeneous approximation, this equation can be reduced to the homogeneous one but with the changed coefficients the values of which depend on the concentrations of different types of bases. In this paper we apply the quasi–homogeneous approximation to solve the equation of that type and to calculate the main dynamic characteristics of the bubbles necessary to analyze the influence of constant and periodical fields on the bubble behavior.

Table 1 Coefficients of Eq. (1)

| Type of the n-th base | \(I_n\) \((10^{-14} \text{kg} \cdot \text{m}^2)\) | \(K\) \((\text{J}/\text{m}^3)\) | \(R_n\) \((10^{-10} \text{m})\) | \(V_\alpha\) \((10^{-20} \text{ J})\) | \(a\) \((10^{-18} \text{ m})\) | \(\alpha_c\) \((\text{ J} / \text{ c})\) |
|-----------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| adenine              | 7.61            | 6.75            | 5.8             | 2.09            | 3.4             | 4.25            |
| thymine              | 4.86            | 6.75            | 4.8             | 1.43            | 3.4             | 2.91            |
| guanine              | 8.22            | 6.75            | 5.7             | 3.12            | 3.4             | 4.10            |
| cytosine             | 4.11            | 6.75            | 4.7             | 2.12            | 3.4             | 2.79            |

**Models and methods**

Let us begin with the discrete version of the modified sine–Gordon equation:

\[
\frac{d^2 \phi_n(t)}{dt^2} + \frac{d\phi_n(t)}{dt} = \alpha \sin \phi_n(t) - \frac{K}{R_n} (\phi_{n+1}(t) - 2\phi_n(t) + \phi_{n-1}(t)) + V_\alpha \sin \phi_n(t) = M_0 + M_1 \cos(\Omega t).
\]

Here \(\phi_n(t)\) is the angular displacement of the \(n\)-th base, \(I_n\) is the moment of inertia of the \(n\)-th base, \(K\) is the stiffness of the sugar–phosphate chain, \(a\) is the distance between adjacent base pairs, \(V_\alpha\) is the factor characterizing the interaction between the complementary bases inside the \(n\)-th pair, \(n = 1, 2, \ldots, N\) is the number of bases in the sequence, \(\alpha_c = \frac{\alpha}{\lambda}\), \(\lambda\) is the dissipation factor, \(M_0\) is the constant torque, \(M_1 \cos(\Omega t)\) is a periodic field.

The values of the coefficients in the left side of the system of equations (1) are shown in Table 1. The values of the parameters of the external fields (\(M_0, M_1\) and \(\Omega\)) are yet arbitrary.

Let us assume that the desired solutions of Eqs. (1) are sufficiently smooth functions. Then, we can apply the continuum approximation:

\[
a \to 0, \quad z = na \to z, \quad \phi_n(t) \to \phi(z, t), \quad I_n = I(z_n) \to I(z), \quad V_\alpha = V(z_n) \to V(z), \quad R_n = R(z_n) \to R(z), \quad \alpha_c = \alpha(z_n) \to \alpha(z).
\]

As a result Eqs. (1) are transformed to:

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\[ I(z) \frac{d^2 \psi(z,t)}{dt^2} + \alpha(z) \frac{d \psi(z,t)}{dt} - KR(z) \alpha^{1/2} \left( \frac{d \psi(z,t)}{dt} \right)^{1/2} + V(z) \sin \psi(z,t) = 0 + M_0 + M_1 \cos(\Omega t). \]  

(2)

To simplify further calculations, we use the quasi–homogeneous approximation according to which we can replace the coefficients of the left–hand side of Eq. (2) by the averaged values:

\[ I(z) \rightarrow I_z = \frac{N_A}{N} + I_0 + \frac{N_C}{N} - I_0, \]
\[ R(z) \rightarrow R_z = \frac{N_A + N_T}{N} + R_0 + \frac{N_C}{N} - R_0, \]
\[ V(z) \rightarrow V_z = \frac{N_A + N_T}{N} + V_0 + \frac{N_C}{N} - V_0, \]
\[ \alpha(z) \rightarrow \alpha_z = \frac{N_A + \alpha_T}{N} + \alpha_0 + \frac{N_C}{N} - \alpha_0, \]

where \( N_A \) is the number of adenines, \( N_T \) is the number of thymines, \( \alpha_A \) is the number of guanines, \( N_C \) is the number of cytosines, and \( N = (N_A + N_T + N_G + N_C) \) is the total number of bases in the sequence.

After the averaging procedure, Eq. (2) takes the form similar to the sine–Gordon equation, but with the coefficients recalculated by formulas (3):

\[ \frac{\partial^2 \psi(z,t)}{\partial t^2} + \alpha(z) \frac{\partial \psi(z,t)}{\partial t} - K R(z) \alpha^{1/2} \left( \frac{\partial \psi(z,t)}{\partial t} \right)^{1/2} + \sin \psi(z,t) = M_0 + M_1 \cos(\Omega t), \]

(4)

where \( K = KR^2 \). Numerical values of the coefficients that are averaged over the entire sequence of plasmid pTTQ18 are given in Table 2.

Table 2 Coefficients of Eq. (4)

| Type of the sequence | \( R \) | \( R^1 \) | \( R^1 \) | \( \alpha \) |
|----------------------|----------|----------|----------|----------|
| pTTQ18              | 6.21     | 1.88     | 2.21     | 3.51     |

For convenience, let us introduce new (dimensionless) variables:

\[ \tau = \alpha \tau, \quad \zeta = \mu \tau, \]

where \( \alpha = (\sqrt{KR})^2, \mu = \alpha^{-1} \left( \sqrt{KR} \right)^2 \). In these variables, Eq. (4) takes the form:

\[ \varphi_{\mu} + \beta \varphi - \varphi_{\mu \mu} + \sin \varphi = f_0 + f_1 \cos(\alpha \tau), \]
\[ \varphi_{\mu} - \beta \varphi + \varphi_{\mu \mu} + \sin \varphi = f_0 + f_1 \cos(\alpha \tau), \]

(5)

where \( \beta = \alpha \left( \sqrt{KR} \right)^2, f_0 = M_0 / \sqrt{\alpha}, f_1 = M_1 / \sqrt{\alpha}, \alpha = \Omega / \left( \sqrt{KR} \right)^2 \).

If the dimensionless coefficient of dissipation is small (\( \beta < 1 \)), Eq. (5) has approximate one–soliton solution in the form of kink:

\[ \varphi = 4 \arctan \left[ \exp \left( \gamma \left( \zeta - \nu \tau - \zeta_0 \right) \right) \right], \]

(6)

The velocity of which is defined by the equation of McLaughlin and Scott:

\[ \frac{d \zeta}{d \tau} = - \alpha \zeta + \beta \varphi + \alpha \varphi_{\mu} + \beta \varphi_{\mu} + \beta \varphi_{\mu \mu} + \sin \varphi \]

(7)

Here \( \gamma = \mu / (1 + (\nu \tau)^2)^{1/2} \), \( \zeta_0 \) is an arbitrary constant.

Let us determine the coordinates of the kink \( \zeta \) by formula:

\[ \frac{d \zeta}{d \tau} = \frac{8 \alpha \varphi_{\mu}}{\sqrt{1 - \nu^2(\tau)}}, \]

and the total energy and the size of the kink by the formulas:

\[ E(\tau) = 8 \alpha \varphi_{\mu}, \quad D(\tau) = \frac{1}{\sqrt{1 - \nu^2(\tau)}}. \]

Differentiating (9) with respect to \( \tau \), we get two additional differential equations:

\[ \frac{d \varphi}{d \tau} = \frac{8 \alpha \varphi_{\mu}}{\sqrt{1 - \nu^2(\tau)}}, \]
\[ \frac{d \varphi}{d \tau} = \frac{8 \alpha \varphi_{\mu}}{\sqrt{1 - \nu^2(\tau)}}. \]

(10, 11)

Eqs. (7), (8) and (10), (11) are a complete set of equations which are necessary to calculate the time dependencies of the velocity, coordinate, size and total energy of the transcription bubbles.

**Results and discussion**

Here we present the results of numerical calculations of the velocity, coordinate, size and total energy of the transcription bubbles. To better understand the dynamic behavior of the bubbles, we made for three different values of the external torsion moment and for three different values of the initial velocities of the bubbles.

**Bubbles dynamics under the action of constant external field**

This case corresponds to \( f_0 = 0, f_1 = 0 \). Dynamic characteristics of the bubbles, obtained for three different values of the torsion moment \( f_0 \), are shown in Figure 3A & 3B. The bubbles trajectories in the phase plane (\( \nu, \zeta \)) are shown in Figure 3C. The bubble total energy and size are shown in Figures 3D & 3E. The time required for calculation of the curves in Figures 3A & 3B and of the curve 1 in Figure 3(C), is equal to \( 2.5 \times 10^3 \). The time required for calculation of the curves 2 and 3 in Figure 3C is equal to \( 5 \times 10^3 \) and \( 5 \times 10^4 \), respectively (Figure 3).

The values of \( f_0 \) and \( f_1 \) are chosen arbitrarily. The value \( f_0 \) is assumed to be equal to \( f_0^{crit} \) which in turn is determined from Eq. (7) as \( f_0^{crit} = \frac{4 \beta \varphi_0}{\pi \sqrt{1 - \nu_0^2}} \).

(12)

From Figure 3A it is seen that at \( f_0 = f_0^{crit} \) (curve 2) the bubble velocity is constant and equal to the initial velocity \( \nu_0 \). At \( f_0 < f_0^{crit} \) (curve 3), the bubble kink velocity initially decreases and after a time period (T), equal approximately 500 (0.84ns), reaches the stationary value:

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\[
U_0^\text{tr} = \left[ 1 + \left( \frac{4\beta}{\pi f_0} \right)^2 \right]^{-1/2}.
\]

At \( f_0 > f_0^{\text{crit}} \) (curve 1), the kink velocity initially increases and after approximately the same time period \( T \), reaches the stationary value:

\[
U_0^\text{tr} = \left[ 1 + \left( \frac{4\beta}{\pi f_0} \right)^2 \right]^{-1/2} = 0.207.
\]

From Figure 3B it is seen that at \( f_0 = f_0^{\text{crit}} \) (line 2) the bubble coordinate is a completely straight line. At \( f_0 < f_0^{\text{crit}} \) (curve 3), the coordinate initially forms a small bend, and then after a certain period of time it transforms into a straight line. Similarly, at \( f_0 > f_0^{\text{crit}} \) (curve 1), the coordinate initially forms a small bend (in opposing direction), and then it also transforms into a straight line.

From Figures 3D & 3E it is seen that at \( f_0 = f_0^{\text{crit}} \) (curve 2) the bubble energy and size are constants. At \( f_0 < f_0^{\text{crit}} \) (curves with number 3), the bubble energy before reaching the stationary value is reduced from the value \( e_{01} = 8.04 \) till the value \( e_{02} = 8 \), and the bubble size is reduced from \( D_{01} = 0.995 \) till \( D_{02} = 0.99 \). At \( f_0 > f_0^{\text{crit}} \) (curves with number 1), the bubble energy before reaching the stationary value is increased from the value \( e_{01} = 8.04 \) till the value \( e_{02} = 8.16 \), and the bubble size is increased from \( D_{01} = 0.995 \) till \( D_{02} = 1.01 \). Hence, by setting different values of constant external torsion field we can force the bubble to move with a certain fixed velocity. The opposite is true. If we know that bubble is moving at a given velocity, we can calculate the value of the torsion moment, which will ensure the movement of bubble at that velocity.

\[
M_0^\text{tr} = \frac{4\beta}{\pi \frac{U_{\text{tr}}^\text{crit}}{N_0}} \sqrt{\frac{\rho}{\rho^0}} \sqrt{1 - \left( \frac{U_{\text{tr}}^{\text{crit}}}{N_0} \right)^2}.
\]

If we assume that the velocity of transcription is equal to 100 base pairs per second \( U_{\text{tr}}^\text{crit} = 0.34 \times 10^{-7} \text{ m/s} \), then from formula (17) we find the desired estimate: \( M_0^\text{tr} = 0.49 \times 10^{-31} \text{ J} \). Figure 4 shows the results of numerical calculations of the dynamic characteristics of the bubbles, made for three different values of the initial velocity \( U_0^\text{tr} \) (Figure 4).

\[
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Figure 4A shows that for any value of the initial bubble velocity $u_0$, the values of stationary velocity are the same. Figure 4B shows that the slopes of the bubbles trajectories reach the same value. Figure 4C & 4D shows that for different values of initial velocities the bubbles energy and size also reach the same values. Thus, we can conclude that the stationary dynamic characteristics of the bubbles do not depend on the initial velocities. On the contrary, they depend only on the value of the torsion moment.

Figure 4A shows that for any value of the initial bubble velocity $u_0$, the values of stationary velocity are the same. Figure 4B shows that the slopes of the bubbles trajectories reach the same value. Figure 4C & 4D shows that for different values of initial velocities the bubbles energy and size also reach the same values. Thus, we can conclude that the stationary dynamic characteristics of the bubbles do not depend on the initial velocities. On the contrary, they depend only on the value of the torsion moment.

Figure 5 shows that for any value of the initial bubble velocity $u_0$, the values of stationary velocity are the same. Figure 4B shows that the slopes of the bubbles trajectories reach the same value. Figure 4C show that for different values of initial velocities the bubbles energy and size also reach the same values. Thus, we can conclude that the stationary dynamic characteristics of the bubbles do not depend on the initial velocities. On the contrary, they depend only on the value of the torsion moment.

Figure 4 (a) Velocity $u(t)$, (b) coordinate $z(t)$, (c) phase trajectory, (d) total energy $e(t)$ and (e) size $D(t)$ of the transcription bubble. Initial velocities:

- $u_{01} = 0.1$, $u_{02} = 0.5$, $u_{03} = 0.8$.
- Torsion moment $f_{01} = 2.429 \cdot 10^{-3}$.
- Dissipation factor $\beta = 0.009$.

**Bubbles dynamics under the action of periodic external field**

This case corresponds to $f_0 \neq 0$, $f \neq 0$, $\omega \neq 0$. It could be realized, for example, in an experiment with a single molecule at the work of atomic force microscope in an oscillating manner. Analyzing the graphs of coordinate and velocity presented in Figure 5A & 5B one can notice that at the beginning of the bubbles movement there exists a short period $T \sim 500$ where oscillations are established. After the end of the period $T$ the bubbles continues to oscillate with the constant frequency $\omega$. Figure 5D & 5E show the time dependence of the total energy and the size of the bubbles. It can be seen the doubling of the frequency of the stationary oscillations. Figure 6 shows the dynamic characteristics of the bubbles, obtained for three different values of the initial velocity: $u_{01}$, $u_{02}$, $u_{03}$ (Figure 6).

From Figure 6 it is seen that after some short period $T$ the velocity and coordinate of the bubbles have different initial velocities at the “start-up, begin to oscillate with constant amplitude and frequency. It can be seen also the doubling of the frequency of the oscillations of the bubbles energy and size.

**Conclusion**

Transcription bubbles have been modeled as quasi–particles – kinks, moving in the potential field of DNA. The influence of constant and periodic fields on the movement of the kinks was studied by the method McLaughlin and Scott. The time dependences of the kinks velocity, coordinate, total energy and size were obtained.
When calculating we used the following dimensionless values of the initial bubbles velocity: $v_{01}=0.1$, $v_{02}=0.5$, $v_{03}=0.8$. The dimensional units these values correspond to: $v_{01}=187 \text{ m/s}$, $v_{02}=935 \text{ m/s}$, $v_{03}=494 \text{ m/s}$. To model the constant torsion moment, we used the dimensionless values: $f_{01}=2.429 \times 10^{-3}$, $f_{02}=1.215 \times 10^{-3}$, $f_{03}=1.215 \times 10^{-3}$; corresponding to the dimensional values: $M_{01}=5.37 \times 10^{-23} \text{ J}$, $M_{02}=2.68 \times 10^{-23} \text{ J}$, $M_{03}=2.68 \times 10^{-24} \text{ J}$. To simulate the amplitude and frequency of the periodic external field, we used dimensionless values: $f_1=2.429 \times 10^{-3}$, $f_2=1.215 \times 10^{-3}$, $f_3=1.215 \times 10^{-3}$; corresponding to the dimensional values: $M_{11}=5.37 \times 10^{-23} \text{ J}$, $M_{12}=2.68 \times 10^{-23} \text{ J}$, $M_{13}=2.68 \times 10^{-24} \text{ J}$, $\Omega_0=0.119 \times 10^{11} \text{ s}^{-1}$.

It was shown that in the case of constant torque, there is a small time period $T\approx 500$ (0.84 ns), after which the bubble velocity reaches the stationary value. This stationary value does not depend on the initial bubble velocity and is completely determined by the value of constant torque. By setting this or that value of the constant torsion moment we can force the bubble to move at a predetermined velocity. We used this result to estimate theoretically the value of the DNA torque which can not be measured by currently existing experimental methods. We obtained that this value is equal to $0.49 \times 10^{-11} \text{ J}$ if the velocity of transcription process is equal to 100 base pairs per second. Moreover we derived formula relating DNA torque and the velocity of transcription.

It was also shown that in the case of periodic field there is almost the same time period after which the bubble begins to oscillate with constant amplitude and frequency the value of which does not depend on the initial bubble velocity. This can lead to the destruction of the transcription bubble.

Summarizing all the results obtained, it can be concluded that external influences, such as constant torsion moment which is induced and regulated by topoisomerases, as well an external periodic field, have a significant effect on the movement of the transcription bubbles. Thus, in the frameworks of model investigations described above, we obtained positive answer on the question concerning a possibility to regulate till some extent the gene expression and biological activity of cells.

It is necessary to note, however, that all these results have been obtained in the frameworks of a rather simple model, which simulates angular oscillations of nitrous bases in one of the two polynucleotide chains, the other chain being modeled as an averaged external field. But we expect that further improvement of the model due to taking into account the transverse and longitudinal displacements of nitrous bases, the mobility of bases in the other polynucleotide chain, the helicity of the DNA structure will not change the main conclusions about the role of the constant and the periodic field in the bubble dynamics.

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**Conflict of interest**

Author declares that there is no conflict of interest.

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