Umbellula pomona sp. nov., a new sea pen from Mar del Plata Submarine Canyon (Cnidaria: Octocorallia: Pennatulacea)

Jessica RISARO 1, Gary C. WILLIAMS 2, Daniela PEREYRA 3 & Daniel LAURETTA 4

1, 3, 4 Museo Argentino de Ciencias Naturales “Bernardino Rivadavia” – CONICET, Av. Ángel Gallardo 470, Buenos Aires, Argentina.
2 Department of Invertebrate Zoology and Geology, California Academy of Sciences, Golden Gate Park, San Francisco, California 94118, USA.

*Corresponding author: jessicarisaro@gmail.com
1 Email: gwilliams@calacademy.org
2 Email: mdr.pereyra@gmail.com
3 Email: dlauretta@gmail.com

Abstract. Sea pens (Cnidaria: Anthozoa: Pennatulacea) constitute a distinctive group of colonial marine invertebrates. They inhabit the world’s oceans, from shallow to deep waters. Studies about this group in Argentina are scarce, and no species have been described in the area in over a decade. Based on samples collected in Mar del Plata Submarine Canyon at about 3000 m deep we describe a new species of sea pen, *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. This is a spiculate *Umbellula* that differs from other species of *Umbellula* with sclerites, by the number, development and distribution of the autozooids in its terminal cluster, as well as the shape of its axis. Molecular data also distinguishes it from other known species. Of the forty-three described species approximately ten are considered valid for the genus *Umbellula*, four of them are registered for the South Atlantic Ocean and only three are described for the Antarctic region. Since sampling efforts in this area have been scarce, the number of species of sea pens from the region is likely to increase substantially in the coming years.

Keywords. Argentina, benthos, octocorals, taxonomy, biodiversity.

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Introduction

Pennatulaceaans, known as sea pens, are a distinctive group of octocorals, in the order Pennatulacea Verrill, 1864 (Cnidaria, Anthozoa). They are formed from a single large primary polyp with (usually)
lateral buddings of the body wall that give rise to all secondary zooids that comprise the colony. Sea pens are almost exclusively adapted for soft substrata (Williams 1992) with a few exceptions (Williams & Alderslade 2011). Hitherto, of the approximately 450 described species, at least 200 are estimated to be valid (Williams 1995), grouped in thirty-seven genera and fourteen families of living pennatulaceans (Williams 2011, 2015; García-Cárdenas et al. 2019). Despite the fact that only a few works deal with the phylogenetic relationships of the group, recent data using mitochondrial genes (mtMutS and ND2) confirmed the Pennatulacea as a monophyletic group (McFadden et al. 2006; Dolan et al. 2013; Kushida & Reimer 2019).

Only a few taxonomic works dealing with sea pens from the southwestern Atlantic Ocean (SAO) (from the area of Argentina) have been made so far (Kölliker 1880; Acuña & Zamponi 1992; Zamponi & Pérez 1995; Pérez & Zamponi 2004; Williams 2011). This is the first description of a new species for the area in fifteen years because of the low number of studies made in this region, so we estimate that while more expeditions can be made, the biodiversity of this group will increase considerably and nowadays its biodiversity is likely to be underestimated. *Umbellula* Cuvier, 1798 is the only genus described within the family Umbellulidae Lindahl, 1874. It is characterized by an elongate and slender rachis with autozooids restricted to the distal region of the colony, usually forming a dense umbellate cluster. Siphonozooids are present at the base of the autozooids or on bare parts of the rachis. Most of the species lack sclerites, but when present, they are rods, spindles or needles, three-flanged, and usually round in cross-section (Williams 1990).

The Mar del Plata Submarine Canyon is located at the continental margin of Argentina at about 38º S latitude. The geomorphology of the external shelf and the submarine canyon of the Argentinian continental margin are strongly influenced by the Malvinas Current, a branch of the Antarctic Circumpolar Current that runs towards the northeastern region of the Argentinian continental margin (Piola & Matano 2001). The Malvinas Current transports cold subantarctic water and collides with the Brazil Current that carries warm waters along the continental slope of South America. This collision generates one of the most energetic regions of the world ocean (Piola & Matano 2001).

Although submarine canyons are known to be hotspots of benthic biomass and are globally numerous, these environments are very poorly sampled (Del Río Iglesias et al. 2012), therefore it is very important to study their biodiversity. Based on the specimens collected during the expeditions to the canyon (2012 and 2013), several new records and new species of Cnidaria Verrill, 1865, Mollusca Linnaeus, 1758 and Echinodermata Bruguière, 1791 have already been published (Cerino & Lauretta 2013; Martínez et al. 2014; Farias et al. 2015; Signorelli & Pastorino 2015; Maggioni et al. 2016; Pastorino & Sánchez 2016; Pastorino 2016, 2019; Martínez & Penchaszadeh 2017; Pereira & Doti 2017; Bernal et al. 2018). Here, we describe a new species of *Umbellula* from Mar del Plata Submarine Canyon based on specimens collected between 2934 m and 3282 m deep, using both morphological and molecular data.

**Material and methods**

**Taxonomic data**

During the “Talud Continental III” expedition to Mar del Plata Submarine Canyon in September 2013, four specimens of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. were collected at stations Nº 45 (38º1.913′ S, 53º39.268′ W, 2934 m depth) and Nº 46 (38º5.310′ S, 53º39.988′ W, 3282 m depth) (Fig. 1) using trawls and preserved in ethanol 96%. All the studied specimens were deposited in the Argentinian Museum of Natural Sciences “Bernardino Rivadavia” (Museo Argentino de Ciencias Naturales “Bernardino Rivadavia”- MACN), Buenos Aires, Argentina.

The general morphology of the specimens, the distribution of sclerites and the shape of transverse sections of the axis were studied by the naked eye and using a stereoscopic microscope. For the sclerites’ shape
we followed the nomenclature of Bayer et al. (1983). Scanning electron microscopy (SEM) was used to examine the shape and length of the sclerites from the tentacles, polyp body and rachis of the colonies and the shape of the axis in transverse section. All measures of the peduncle (when possible), rachis, polyp’s body and tentacles were done with a digital caliper and correspond to the holotype. Photographs of the colonies were taken using a digital SLR Nikon D800 camera with a Nikkor 60 mm F2.8 macro lens. To separate and prepare the sclerites of rachis, polyp’s body and tentacles to examine them by SEM, a portion of each tissue was cut and treated with diluted sodium hypochlorite (commercial bleach) for fifteen minutes and then washed with distilled water. Finally, the residual of water was evaporated with ethanol 96% and the sclerites were placed on SEM tubs; for the axis sections, a portion of it was cut and all soft tissue was retired, then the same protocol of the sclerites was followed.

Molecular data
Small tissue fragments were obtained from the holotype of *Umbellula pomona* sp. nov. and one paratype, both preserved in 96% ethanol. The samples were preliminarily washed with deionized water, allowing removal of ethanol. Total genomic DNA was extracted following the salting-out method (Miller et al. 1988). Following previous published works (Dolan et al. 2013; Kushida & Reimer 2019) we amplified two mitochondrial regions: mtMuts (France & Hoover 2002; Sánchez et al. 2003) and ND2 (McFadden et al. 2004). The primer’s sequences and thermocycling profiles used to amplify each target gene are described in Table S1 (Supplementary File 1).

Amplifications were carried out in 20 µl reaction mixtures containing 1× PCR buffer mix (the buffer includes dNTPs and MgCl₂), 0.2 µM of each primer, 1 U of Taq polymerase (MyTaq DNA Polymerase, Bioline), 4 µg of Bovine Serum Albumin (BSA), 2 µl containing < 10 ng template DNA, and brought to final volume with dH₂O. The PCR reactions were performed in a T100TM thermocycler (Bio-Rad, USA), with negative controls included to verify the absence of contamination. The PCR products size was

Fig. 1. Distribution of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. in Mar del Plata Submarine Canyon.
confirmed via 1.5% agarose gel electrophoresis. PCR products were purified and sequenced in Macrogen, Korea.

**Molecular analysis**

We added our sequences to the data set used in Dolan et al. (2013) (Table 1), since we are dealing with a deep-sea species of *Umbellula* (which is the target genus of this work) and no new data on this genus was added in Kushida & Reimer (2019). All the sequences were downloaded from GenBank, although the mtMutS sequence of *Anthoptilum* sp. (KF313832) based on a primnoid (as noted by Kushida & Reimer 2019), so we did not include this sequence in our data matrix. The new sequences were edited using the software Geneious ver. 5.6.7 and checked using BLAST (Altschul et al. 1990) to rule out contamination and to confirm gene identity. The concatenated data set consisted of 41 taxa and 1257 bp. Sequence alignment was performed using MAFT (Multiple Alignment using Fast Fourier Transform) ver. 7 ([https://mafft.cbrc.jp/alignment/server/](https://mafft.cbrc.jp/alignment/server/)) using L-INS-i strategy and default parameters. Trees were built using Bayesian analysis (BA) and maximum likelihood (ML). To determine the evolution model (GTR+G and GTR+I+G for mtMuts and ND2, respectively), we implemented the Akaike information criterion (AIC) in PartitionFinder 2 software (Lanfear et al. 2016). Elliselidids were chosen as outgroup (Dolan et al. 2013; Kushida & Reimer 2019). The Bayesian tree was built using Mrbayes (Ronquist et al. 2012) on Cipres science gateway (Miller et al. 2010): number of runs: 2, number of chains: 4, number of generations: 100 000 000, chain sample frequency: 10 000. The first 25% of each search was discarded (burninfrac = 0.25). To ensure that the Markov chains reached stationarity (effective sample size values over 200) we used the software Tracer ver. 1.7 (Rambaut et al. 2018). Maximum likelihood analyses were performed using PhyML ver. 3.1 (Guindon et al. 2010) for each individual gene and as a complete set. Bootstrap support was calculated based on 1000 rounds.

**Results**

**Systematics**

Phylum Cnidaria Verrill, 1865  
Class Anthozoa Ehrenberg, 1834  
Subclass Octocorallia Haeckel, 1866  
Order Pennatulacea Verrill, 1864  
Family Umbellulidae Lindahl, 1874

**Type genus**

*Umbellula* Cuvier, 1798.

**Diagnosis** (adapted from Williams 1990, modifications in bold)

Rachis long and slender. Axis quadrangular to round in cross section. Autozooids restricted to the distal terminus, usually forming an umbellulate cluster. Siphonozooids are present on the rachis at the base of the autozooids or on bare parts of the rachis. Sclerites either present in peduncle, rachis, and terminal cluster, or totally absent. When present, sclerites are rods, spindles or needles, three-flanged round in cross-section.

**Included genera**

*Umbellula* Cuvier, 1798.

**Distribution**

Cosmopolitan, from 250 m to over 6200 m deep (Williams 2011).
### Table 1

GenBank accession numbers of the sequences analyzed in this study. New specimens/sequences in bold. – = data not available; * = data not used (see comment in text).

NOCS = National Oceanography Centre, Southampton (UK); a = Natural History Museum (London, UK); b = NIWA (New Zealand); c = MACN (Argentina).

| Taxon                        | Location                  | Depth | Catalogue# | mtMutS  | ND2       |
|------------------------------|---------------------------|-------|------------|---------|-----------|
| Anthoptilidae                |                           |       |            |         |           |
| Anthoptilum marrayi          | Tasman Sea, AUS           | –     | –          | –       | DQ302938  |
| Anthoptilum sp. 1            | Subantarctic              | –     | 2010.18a   | KF313832* | KF313805  |
| Anthoptilum sp. 2            | Sumatra, Indian Ocean     | 1714  | NOCS       | –       | KF313806  |
| Funiculinidae                |                           |       |            |         |           |
| Funiculina armata            | NE Atlantic               | 350   | 2010.11a   | KF313833 | KF313807  |
| Funiculina quadrangularis    | NE Atlantic               | 173   | NOCS       | KF313834 | –         |
| Halipteridae                 |                           |       |            |         |           |
| Halipteris finmarchica       | Tasman Sea, AUS           | –     | –          | DQ302868 | DQ302941  |
| Halipteris finmarchica       | New Zealand, W Pacific    | –     | 28801b     | KF313835 | KF313808  |
| Kophobelemnidae              |                           |       |            |         |           |
| Kophobelemnon macrospinum    | Tasman Sea, AUS           | –     | –          | DQ302865 | DQ302937  |
| Kophobelemnon pauciforum     | Crozet, S Atlantic        | 4189  | 2010.21a   | KF313836 | KF313809  |
| Kophobelemnon sp. 1          | Koster Channel, NE Atlantic| 70    | 2010.13a   | KF313837 | KF313810  |
| Kophobelemnon sp. 2          | Monterey, E Pacific       | 2456  | 2010.10a   | KF313838 | KF313811  |
| Kophobelemnon sp. 3          | New Zealand, W Pacific    | 1812  | 28827b     | KF313839 | KF313812  |
| Scleroptilidae               |                           |       |            |         |           |
| Pennatulidae                 |                           |       |            |         |           |
| Pennatula aculeata           | Monterey, E Pacific       | 2456  | 2010.15a   | KF313840 | KF313813  |
| Pennatula phosphorea         | Millport, NE Atlantic     | 55    | NOCS       | KF313841 | KF313814  |
| Pennatula murray             | Monterey, E Pacific       | 3208  | 2010.9a    | KF313842 | KF313815  |
| Pennatula sp.                | Tasman Sea, AUS           | –     | –          | DQ302870 | DQ302943  |
| Protoptilidae                |                           |       |            |         |           |
| Distichoptilum gracile       | Tasman Sea, AUS           | –     | –          | DQ302866 | DQ302939  |
| Distichoptilum gracile       | New Zealand, W Pacific    | 1211  | 28813b     | KF313843 | KF313816  |
| Distichoptilum gracile       | Monterey, E Pacific       | 2456  | NOCS       | KF313844 | KF313817  |
| Protoptilum sp.              | Monterey, E Pacific       | 3208  | 2010.20a   | KF313844 | KF313818  |
| Pterocididae                 |                           |       |            |         |           |
| Gyrophyllum sp.              | NE Atlantic               | 1580  | NOCS       | KF313845 | –         |
| Gyrophyllum sp.              | New Zealand, W Pacific    | 997   | 28779b     | KF313846 | KF313819  |
| Pterocides sp.               | Tasman Sea, AUS           | –     | –          | DQ302871 | DQ302944  |
| Renillidae                   |                           |       |            |         |           |
| Renilla muelleri             | GOM, Florida, USA         | –     | –          | DQ297432 | DQ297451  |
| Scleroptilidae               |                           |       |            |         |           |
| Scleroptilum grandiflorum    | Mid-Atlantic Ridge        | 2190  | 2010.14a   | KF313847 | KF313820  |
Remarks
Since sclerites were found in the rachis of *U. pomona* sp. nov., we added this character to the family diagnosis.

Genus *Umbellula* Cuvier, 1798

**Type species**
*Isis encrinus* Linnaeus, 1758.

**Diagnosis**
Same characteristics of the family.

**Valid species** (based on Broch 1957; Williams 1995; López-González & Williams 2011)
*Umbellula encrinus* (Linnaeus, 1758), *Umbellula lindahl* Kölliker, 1874, *Umbellula thomsoni* Kölliker, 1874, *Umbellula carpenteri* Kölliker, 1880, *Umbellula durissima* Kölliker, 1880, *Umbellula huxleyi*
Kölliker, 1880, *Umbellula pellucida* Kükenthal, 1902, *Umbellula spicata* Kükenthal, 1902, *Umbellula hemigymna* Pasternak, 1975, *Umbellula monocephalus* Pasternak, 1964, *Umbellula pomona* sp. nov.

**Remarks**

Although in his revision of the family, Broch (1958) synonymized *Umbellula dura* Thomson & Henderson, 1906 with *Umbellula durissima* Kölliker, 1880 and *Umbellula rosea* Thomson & Henderson, 1906 with *Umbellula thomsoni* Kölliker, 1880, we consider that, based on the original descriptions and images of *U. dura* and *U. rosea*, those species have important morphological characters to compare with our specimens.

*Umbellula pomona* Risaro, Williams & Lauretta sp. nov. urn:lsid:zoobank.org:act:77FCDEF4-AAE3-4BB5-BAB2-1EDAA4B19439 Figs 2–7

**Differential diagnosis**

*Umbellula pomona* sp. nov. is a spiculated *Umbellula* with three autozooids in its terminal cluster, a central well-developed polyp and two lateral, symmetric and smaller ones. It presents large sclerites in all its tissues as well as siphonozooids all along the rachis. Its central axis is circular in cross section all along its extension, and does not vary throughout the colony.

**Etymology**

The species is named after the birthplace of the first author (JR), Pomona (Río Negro, Argentina). The word ‘pomona’ is used as a noun in opposition.

**Material examined**

**Holotype**

SW ATLANTIC OCEAN • one complete spec. (preserved in 96% ethanol); Mar del Plata Submarine Canyon, “Talud Continental III” exped., stn N° 45; 38°1.913′ S, 53°39.268′ W; 2934 m deep; Sep. 2013; Daniel Lauretta leg.; MACN-IN 42608.

**Paratypes**

SW ATLANTIC OCEAN • 3 specs (two adult, without the peduncle: paratypes A and B; and one juvenile like, complete: paratype C; preserved in 96% ethanol); Mar del Plata Submarine Canyon, “Talud Continental III” exped., stn N° 46; 38°5.310′ S, 53°39.988′ W; 3282 m deep; Sep. 2013; Daniel Lauretta leg.; MACN-IN 42609.

**Description** (holotype MACN-IN 42608)

The colony looks rugous in all its extension, especially the terminal cluster of autozooids. The color of the polyps, rachis and the peduncle is white or light yellow (preserved). The holotype is 214 mm in length. It has a terminal cluster of three autozooids with tentacles, one central and larger polyp growing on the distal-most region of the rachis, and two smaller but well-developed lateral polyps growing at the base of the central one (Fig. 2). The central polyp is 25.3 mm in length and the two lateral polyps’ lengths are 3.0 mm and 2.6 mm. The tentacles of the two types of polyps are larger than its body, the measures are 12.3 mm for the central autozooid, and 1.1 mm and 1.7 mm for the lateral ones. The axis is 178 mm in length and it is circular in cross-section, approximately 0.74 mm in diameter. The rachis is approximately 0.8 mm in diameter in the middle zone between the peduncle and the terminal cluster. The peduncle is 8.68 mm in length, and its appearance is soft and smooth. The autozooids of the cluster grow with a kind of orientation determining a ‘dorsal’ region where the axis inserts and a ‘ventral’
Fig. 2. A. General aspect of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. A. Holotype (MACN-IN 42608). B. Detail of the terminal cluster of paratype A (MACN-IN 42609), showing three autozooids that form the terminal cluster and amplifications of sclerites (up) and siphonozooids (down). Abbreviations: CP = central polyp; LP = lateral polyp; Pd = peduncle; R = rachis; T = tentacle; S = siphonozooids; Scl = sclerites.

Fig. 3. Transversal section of the central axis of paratype B of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. (MACN-IN 42609). A. Transversal section of the central axis between the peduncle and the rachis. B. Transversal section of the central axis near the terminal cluster.
region towards which the polyps come together (Fig. 2). The siphonozooids are numerous and resemble small white dots, and are distributed all along the rachis from the base of the autozooids to the middle of the rachis. These polyps are circular and inconspicuous; and have a diameter of about 393 ± 93 μm (299–486 μm, N = 36).

Sclerites are conspicuous, along much of the length of the colony (polyps, rachis, tentacles and pinnules). They are translucent and colorless, rod-shaped and spindle-shaped with spines (Figs 4–7). Those present in the rachis are spindle-shaped, have triangular protuberances all along their length (Fig. 4) and their sizes are 333.5 ± 71.1 μm (262.4–404.6 μm, N = 10), the body wall of the polyps has different types of sclerites, two of them are rough and rod-shaped, but differ in their measurements, while the others are spindle-shaped with spines along their lengths and also have different sizes (Fig. 5). The largest type is rod-shaped and approximates 1446 ± 32.7 μm (1413.3–1478.7 μm, N = 5) in length (Fig. 5A); another type, smaller and spine-shaped, approximates 687 ± 45.6 μm (641.4–732.6 μm, N = 5) (Fig. 5B–C). The other types are much smaller, and one type is rod shaped, like the largest one (Fig. 5D), and the other

**Fig. 4.** Variability of sizes and ornamentations of the rachis’ sclerites of the holotype of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. (MACN-IN 42608).
type is spine-shaped (Fig. 5E–F); these two types of sclerites have similar sizes, length approximate 534 ± 43.4 μm (490.6–577.4 μm, N = 5). Finally, the tentacles have rod-shaped sclerites (Fig. 6) with two sizes: the larger ones are 1223.2 ± 280.1 μm (943.1–1503.3 μm, N = 5) (Fig. 6A–C) and the smaller ones are 632.8 ± 96.2 μm (536.6–729 μm, N = 5) in length (Fig. 6D–F), and those of the pinnules are rod-shaped with some protuberances along their lengths and their sizes are approximately 269.1 ± 50.3 μm (218.8–319.4 μm, N = 10) (Fig. 7). In both tissues the sclerites are placed along the main axis, not transversally.

Variability
Of the three paratypes (A, B and C), one (paratype C) is significantly smaller than the others, so we consider it could be a juvenile (Fig. 8). The color of the three colonies is white or light yellow when preserved, all of them have three autozooids with the same grade of development as the holotype (as it can be seen in Fig. 2B) and its central axis is round in all its extension (Fig. 3). The large paratypes (A and B) lack their peduncle. Their general aspect is just like the holotype, they are rugous in all their

Fig. 5. Variability of sizes and ornamentations of the polyps’ sclerites of the holotype of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. (MACN-IN 42608).
length because of the presence of conspicuous sclerites in the autozooids and rachis (Fig. 2B). The total length of the large paratypes is 134 mm and 231 mm, their central autozooids are 36 mm and 34 mm long (length of the tentacles: 20 mm and 12 mm, length of the columns: 16 mm and 22 mm, respectively) while the dimensions of the lateral ones are 4.8 mm and 5.6 mm in one paratype and 6.4 mm and 7 mm in the other. Their rachis are 97 mm and 200 mm in length and their diameters are 0.9 mm and 2.2 mm at their widest sections. Their axis’ diameter is 0.83 mm and 1.3 mm. Their siphonozooids are 313 μm and 370 μm (mean) in diameter and look like the siphonozooids of the holotype. Finally, the sclerites of these paratypes look alike and have similar sizes as the sclerites of the holotype. The juvenile paratype’s (C) total length is 110 mm, its central autozooid is 7.9 mm in length (column and tentacles are 4.4 mm and 3.4 mm long, respectively), and the length of the lateral ones is 2.9 mm (column and tentacles are 1.4 and 1.5 mm in length, respectively). The peduncle’s length is 4.6 mm and the rachis’ 95 mm, while its diameter is 0.41 mm at its widest section. Its central axis is 0.35 mm in diameter. Finally, the siphonozooids of this paratype are tiny spots with the same aspect and distribution as those on the holotype and the largest paratypes.

**Fig. 6.** Variability of sizes and ornamentations of the tentacles’ sclerites of the holotype of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. (MACN-IN 42608).
Phylogenetic analysis

Both phylogenetics reconstructions (i.e., BA and ML) agree in the basic topology of the trees (for simplicity we only show BA). Both type specimens of *U. pomona* sp. nov. were grouped together with low support values within the same group (possibly because we only have one gene sequence for each species). *Umbellula* spp. were recovered in two clusters, *Umbellula* clade I including most of the included *Umbellula* species and a second one (*Umbellula* clade II) including only *U. monocephalus, Umbellula pomona* sp. nov. and *Umbellula* sp. 2 from Dolan et al. (2013) (Fig. 9).

Discussion

Regarding *Umbellula pomona* sp. nov.

As currently defined, our specimens agree with the current set of characters assigned to *Umbellula*. They have a long, slender rachis, a cluster of autozooids on their distal-most part and numerous siphonozooids all along the rachis, in addition to conspicuous, rod- or needle-like sclerites. *Chunella* Kükenthal, 1902

![Fig. 7. Variability of sizes and ornamentations of the pinnules’ sclerites of the holotype of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. (MACN-IN 42608).](image-url)
and *Amphiacme* Kükenthal, 1902 are other similar genera, but have several polyp clusters disposed in intervals along the rachis and do not have sclerites (Kükenthal 1915).

From a morphological point of view, there are two major species groups in the genus *Umbellula*, based on the presence/absence of sclerites. Up to date, of all described species, five are spiculate (of which only three are probably valid) whose morphology needs to be compared: *U. thomsoni*, *U. durissima*, *U. dura*, *U. rosea* and *U. monocephalus*. None of them share the diagnostic set of characters (arrangement of the polyp cluster, form of the central axis and sclerite sizes) of our specimens (Table 2). *Umbellula pomona* sp. nov. has a reduced number of polyps, forming a terminal cluster with one central autozooid and two smaller, lateral ones growing from its base, a cylindrical central axis, and the sizes of their sclerites range from $1446 \mu m$ to $269.1 \mu m$, depending on location in the tissues. *Umbellula thomsoni* has been reported with three to ten autozooids in its terminal cluster growing like a flower or an umbrella, with a similar grade of development among them. The axis of this species differs in its shape in cross section, as near the cluster it is quadrangular, while it becomes cylindrical near the peduncle. Its sclerites vary from approximately $1300 \mu m$ (pinnales and tentacles) to less than $300 \mu m$ (rachis and body wall of the polyps) (Kükenthal 1915; Williams 1990). *Umbellula monocephalus* has only one polyp in the distal region of the rachis (Pasternak 1964; Grasshoff 1972; Tiefenbacher 2001). *Umbellula durissima* has

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**Fig. 8.** General aspect of the unique specimen of a juvenile-like paratype of *Umbellula pomona* Risaro, Williams & Lauretta sp. nov. (MACN-IN 42609, paratype C). Abbreviations: CP = central polyp; LP = lateral polyp; R = rachis; PD = peduncle; T = tentacles.
Fig. 9. Phylogenetic reconstruction of the concatenated set of the two mitochondrial genes (mtMutS and ND2) based on the Bayesian analysis. Nodes include the Bayesian posterior probabilities. Arrows point to the new specimens of *Umbellula* Cuvier, 1798 included in the analysis.
Table 2 Comparative descriptions of the spiculated species of Umbellula Cuvier, 1798 and known distribution of each species.

| Species                     | Polyp’s size (mm) | Maximum number of polyps in the cluster | Cluster shape (data taken from original descriptions) | Axis form in cross section | Measurements of sclerites | Distribution / depth (m) |
|-----------------------------|-------------------|-----------------------------------------|--------------------------------------------------------|----------------------------|---------------------------|--------------------------|
| Umbellula thomsoni*         | 15–20             | 10                                      | Polyps distributed in a circle, forming an ‘umbrella’.  | Quadrangular near the terminal cluster and becoming cylindrical to the peduncle. | Rachis, polyps’ column and base of the tentacles: < 300 μm. Terminal section of tentacles and pinnules: 500–1300 μm. Peduncle: 80–110 μm (ovals). | Cosmopolitan / 1300–6200 |
| Kölliker, 1874              |                   |                                         |                                                        |                            |                           |                          |
| Umbellula durissima*        | 21                | 5                                       | One terminal and large polyp, two undeveloped ones.   | Cylindrical in all its extension. | Polyps and tentacles: 2000–2800 μm. Distal zone of rachis: 190–260 μm. Proximal zone of rachis: 76–200 μm. | North Pacific Ocean, south of Tokyo Japan / 1000 |
| Kölliker, 1880              |                   |                                         | Two or three small and undeveloped polyps.            |                            |                           |                          |
| Umbellula dura*             | –                 | 3–6                                     | In juvenile, one central and smaller polyp and two lateral ones. In developed states, four large and two small autozooids. | Cylindrical in all its extension. | Polyps: 140–1500 μm. Rachis: 140–110 μm (X-shaped). | Indian Ocean / 1300     |
| Thomson & Henderson, 1906   |                   |                                         |                                                        |                            |                           |                          |
| Umbellula rosea*            | 12                | 5                                       | Two pairs of lateral polyps (2 and 2) and one terminal. | Quadrangular in all its extension. | Polyps: 220–300 μm. Rachis: 90–160 μm (ellipses). | Indian Ocean / 3200     |
| Thomson & Henderson, 1906   |                   |                                         |                                                        |                            |                           |                          |
| Umbellula monocephalus*     | 40                | 1                                       | Formed by a unique terminal polyp.                     | Cylindrical in all its extension. | In the original description of the species and different tabular keys don’t specify the form of its internal axis | Indian Ocean and Atlantic Ocean / 4635–4700 |
| Pasternak, 1964             |                   |                                         |                                                        |                            |                           |                          |
| Umbellula pomona sp. nov.   | 36                | 3                                       | One central terminal polyp and one pair of lateral, smaller polyps growing from the base of the central one. All of them are completely developed. | Cylindrical in all its extension. | Polyps: 490.6–1413.3 μm. Rachis: 262.4–404.6 μm. Tentacles: 536.6–1503.3 μm. Pinnules: 218.8–319.4 μm. | 38°5.310′ S, 53°39.988° W Southwestern Atlantic Ocean / 3300 |

*Some authors consider U. dura as a synonym of U. durissima and U. rosea as a synonym of U. thomsoni (Broch 1958)
numerous polyps forming an apical cluster of three autozooids, one terminal and well developed, two lateral and two or three less developed ones (without tentacles) below the principal cluster. Its central axis is cylindrical in all its length. The sclerites vary in length from 2800 μm (body wall of the polyps and tentacles) to approximately 200 μm (rachis), additionally, this species does not have siphonozooids along the rachis, but they are grouped between the autozooids in the terminal cluster (Kölliker 1880; Thomson & Henderson 1906; Kükenthal 1915). *Umbellula dura* has three autozooids growing at the same level forming a kind of circle in which the central autozooid is smaller than the other laterals in juvenile states, but in more developed states have six autozooids in total, four large and two very small, all disposed in three sets. Its central axis is cylindrical in cross section and the sclerites vary in length from 1500 μm to 300 μm in the autozooids and from 140 μm to 100 μm in the rachis, also, in these tissue has X-shaped sclerites (Thomson & Henderson 1906), that are absent in *U. pomona* sp. nov. *Umbellula rosea* has three autozooids growing at the same level forming a kind of circle in which the central autozooid is smaller than the other laterals in juvenile states, but in more developed states have six autozooids in total, four large and two very small, all disposed in three sets. Its central axis is cylindrical in cross section and the sclerites vary in length from 1500 μm to 300 μm in the autozooids and from 140 μm to 100 μm in the rachis, also, in these tissue has X-shaped sclerites (Thomson & Henderson 1906), that are absent in *U. pomona* sp. nov. *Umbellula pomona* sp. nov. inhabits the deep region of Mar del Plata Submarine Canyon, at a depth of about 3000 m. Sixty-four sampling stations were established during three expeditions, from 200 m to 3500 m deep, over 150 specimens of sea pens were collected but only four specimens of the new species were found. This low number of specimens is not rare because in the study area, many species report a very low abundance. Many deep-sea invertebrate species from Mar del Plata Submarine Canyon have been registered/described based on a few (and even only one) specimens (Pastorino 2016; Lauretta & Penchasazadeh 2017; Martinez et al. 2019). Since sampling efforts in this area have been almost non-existent until a few years ago, the number of species of sea pens (and other invertebrates) from Mar del Plata Submarine Canyon is likely to increase substantially in the coming years.

**Phylogenetic position of *Umbellula pomona* sp. nov.**

In all the phylogenetic reconstructions made, both specimens of *U. pomona* sp. nov. were grouped together (with low support, since we could only amplify one different gene for each specimen). They were grouped within a cluster composed by only two other species of *Umbellula*: *U. monocephalus* and *U. sp. 2* (from Dolan 2008; Dolan et al. 2013). Unfortunately, no molecular data of *U. rosea*, *U. durissima* or *U. dura* (the most morphologically similar species to *U. pomona* sp. nov.) were available, so it was impossible to compare the species from the molecular point of view.

There are only two molecular phylogeny available for sea pens that include several species and families (i.e., Dolan et al. 2013; Kushida & Reimer 2019). Both were constructed using two mitochondrial genes (mtMutS and ND2), and the latter was an extension of the former, including shallow-water species. Both papers agree that *Umbellula* is a polyphyletic group, with two clearly separate clades (here ‘*Umbellula* clade I’ and ‘*Umbellula* clade II’). ‘*Umbellula* clade I’ includes most of the species of *Umbellula* with available molecular data, including *U. encrinus*, the type species of the genus. ‘*Umbellula* clade II’ includes only three species: *U. monocephalus* (an atypical *Umbellula* since it has only one terminal polyp), *U. pomona* sp. nov. (also an atypical species since the polyps have different sizes) and a non-identified *Umbellula* sp. (*Umbellula* sp2 from Dolan et al. 2013) with a typical polyp cluster (according to the figure in Dolan 2008). According to Dolan et al. (2013), ‘*Umbellula* clade II’ was characterized by
species with sclerites and a round axis, characters also presents in *U. pomona* sp. nov., which supports this clade from the morphological data. If both clades were to be separated in two genera, ‘*Umbellula* clade I’ would retain the generic name *Umbellula* (since it includes the type species of the genus) and a new genus would have to be proposed for ‘*Umbellula* clade II’. Pending a complete revision of the nominal species assigned to *Umbellula*, we prefer to be conservative and include our species within *Umbellula* until a such revision is done and a decision based on molecular and morphological data can be made.

**Valid species within *Umbellula***

Up to date, there are 43 nominal species within *Umbellula* (Williams 1995; Cordeiro *et al.* 2019; this paper). At least nine species were recognized as probably valid by Williams (1995) (*U. durissima*, *U. monocephalus*, *U. thomsoni*, *U. hemigymna*, *U. huxleyi*, *U. lindahlri*, *U. pellucida*, *U. spicata* and...
Later, López-González & Williams (2011) disagreed with some specimens’ identifications and shared the vision of Pasternak (1962) that *U. carpenteri* and *U. magniflora* Kölliker, 1880 are possibly the same species. On the other hand, Broch (1957) in his revision of the family considered *U. thomsoni* and *U. leptocaulis* Kölliker, 1880 as the same species. In this work, we consider the valid species according to the revision made by Broch (1957) and López-González & Williams (2011). Clearly, a complete revision of *Umbellula* spp. is needed, probably using both morphological and molecular data. Table 3 shows the valid species of *Umbellula* with their known distributions considered in this paper.

**On the distribution of *Umbellula* spp. of the South Atlantic Ocean (SAO)**

*Umbellula* is a cosmopolitan genus (Table 3). Specimens of this group have been reported from all over the world, from the equator to the poles. Reports of *Umbellula* in the South Atlantic are limited; according to the papers made by Broch (1958) and Williams (1995), only four species were reported in the SAO: *U. durissima*, *U. monocephalus*, *U. thomsoni* and *U. lindalii*, of which only *U. thomsoni* and *U. lindalii* correspond to the southwestern Atlantic Ocean (SAO), where our specimens are located. The diversity of the group in Antarctic waters is also low, with three species reported (*U. durissima*, *U. lindalii* and *U. carpenteri*) (Kölliker 1880; Broch 1958; López-González & Williams 2011). As mentioned before, the conditions in the sampled area are strongly influenced by the Malvinas Current and the Brazil-Malvinas Confluence, which may explain the presence of Antarctic species in the SAO deep sea. Given the depths where our specimens came from (2934 m and 3282 m), it is possible that this record is (or is near to) the southern limit distribution of the species, since in the confluence area at that depth the North Atlantic Deep waters are present (see Voig et al. 2013), which run southward from the northern hemisphere and in the confluence area divide the Circumpolar Deep Water vertically in two, staying at a depth of 2000–3000 m. A similar situation has also been proposed in the case of the gastropod *Theta lyronuclea* (Clarke, 1959), previously reported from the North Atlantic Ocean and recently found in the same station as *U. pomona* sp. nov. (Sánchez & Pastorino 2020).

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**Compliance with ethical standards**

**Conflict of interest**

The authors declare that they have no conflict of interest.

**Ethical approval**

All applicable international, national, and/or institutional guidelines for the care and use of animals were followed by the authors.

**Sampling and field studies**

All necessary permits for sampling and observational field studies have been obtained by the authors from the competent authorities and are mentioned in the acknowledgements.
Data availability

The datasets generated during the current study are available from the corresponding authors on reasonable request.

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Supplementary material
Supplementary file 1. Primer’s sequences and PCR conditions used to amplify partial regions of the mitochondrial mtMutS and ND2 genes. https://doi.org/10.20.720.1121.2933