Evaluation of Zinc Solubilizing Potential of Maize Rhizosphere Bacterial Isolates

Mangala Devi Perumal1*, V. Subramanian1 and K.G. Sabarinathan2

1Department of Soil Science and Agricultural Chemistry, 2Department of Agricultural Microbiology, Tamil Nadu Agricultural University, Coimbatore, Tamil Nadu, India
*Corresponding author

**Abstract**

Zinc is an essential micronutrient which plays a macro role in the growth and productivity of the plants. Generally Indian soils are low in Zn and as much as half of the country soils are categorized to be Zn deficient. Hence, zinc fertilizer is applied in the soil as zinc sulphate. However the major portions of applied zinc sulphate become unavailable to plant in the soil. There was six zinc solubilizing bacterial strains were isolated from rhizosphere of maize. The bacterial isolate ZSB SM-1 was found to be effective in solubilizing the insoluble zinc substances viz., zinc oxide, zinc carbonate and slowly soluble Zn-EDTA. The insoluble Zn substance was effectively solubilized at 0.1 per cent concentration, compared to 0.2 per cent concentration. The solubilization might be due to production of acids by the culture, since the pH of the broth has been shifted from 7.0-7.3 to 3.0-4.8 after 10 days of inoculation.

**Keywords**

Solubilizing, Maize, Bacterial isolates.

**Introduction**

Micronutrients are important component for the growth of plants, animals and microbes. Among the micronutrients, Zinc is essential for the normal healthy growth and reproduction of plants, animals and humans and when the supply of plant-available zinc is inadequate, crop yields are reduced and the quality of crop products is frequently impaired. In plants, zinc plays a key role as a structural constituent or regulatory co-factor of a wide range of different enzymes and proteins in many important biochemical pathways and these are mainly concerned with: carbohydrate metabolism, both in photosynthesis and in the conversion of sugars to starch, protein metabolism, auxin metabolism, pollen formation. Zinc deficiency is identified as the most common occurring micronutrient problem in many parts of the country on a wide range of soil types.

In India, it has been estimated that more than 50 per cent of the soils are Zn deficient (Singh et al., 2005) and is predominant in the semi-arid tropical soils.

Maize is one of the most important high value cereals consumed in many countries. The crop is widely grown in semi-arid tracts of India,
which is characterised by extreme variation in rainfall pattern both in time and space. Besides this, the major constraint to sustainable maize production is the poor soil fertility. A bacterial based approach was devised to solve the micronutrient deficiency problem in cultivated soils. Among the bacterial species strains belonging to the genera Acinetobacter, Bacillus, Pseudomonas, Glucanoacetobacter have been reported (Sharma et al., 2012) Since Zn is a limiting factor in maize crop production, this study on zinc solubilization by bacteria has immense importance in zinc nutrition of maize crop.

Materials and Methods

Collection of soil samples

 Soil samples were collected from seven different maize fields in and around Thoothukudi district, Tamil Nadu, India. Samples were randomly collected from the rhizosphere of young plants at a depth of 6-15 cm. then the soil samples were used for isolation.

Isolation and purification of zinc solubilizers

Isolation of zinc solubilizing bacteria present in the maize rhizosphere soil samples were done by serial dilution plate count method using carbon rich Tris-minimal salt medium (Fasim et al., 2002). The carbon rich agar medium containing insoluble zinc compounds such as zinc carbonate, zinc oxide and slowly soluble zinc EDTA was used to select the bacterial isolates possessing zinc solubilizing ability. The petriplates were incubated at room temperature (30±1 °C) for three days and the colonies exhibiting clear zones were selected, purified by four way streak plate method. The diameter of zone of solubilization was measured and the selected isolates were preserved on agar slants for further use.

Solubilization of insoluble Zn compounds - Plate assay

The medium used to assess the zinc solubilizing ability of bacteria in the study was Tris-minimal salt medium. The medium was prepared by incorporating insoluble Zn sources zinc oxide (Zn O), zinc carbonate (ZnCO₃), and slowly soluble Zinc-EDTA (Zinc-Ethylene Diamine Tetra Acetate) at 0.1 % and 0.2 % with the carbon source glucose at 10 %. The pH was adjusted to 6.0 after sterilization, with sterilized medium was added to petriplates. Care was taken for uniform distribution of the insoluble zinc source into the petriplates and the plates were prepared without air bubbles. The agar plates were allowed to cool and 10µl of bacterial (6x10⁶ cfu ml⁻¹) suspension was placed on the agar surface. After placing the culture on the agar surface, the plates were kept undisturbed for 10 min to get absorbed in the agar medium without spreading. Then the plates were incubated at 29 °C ± 1 °C and observed for solubilization zones up to 5 days. The diameter of the solubilization was measured and expressed in cm. Two replicates were maintained for each treatment.

Determination of soluble zinc content (Broth assay)

The bacterial isolates were inoculated separately to Tris-minimal salt medium supplemented with 0.1 % insoluble zinc compounds. The solubilization of zinc from laboratory grade ZnO, ZnCO₃ and Zn-EDTA by ZSB was assessed. Tris-minimal salt medium (D-glucose -10 g; Tris HCl- 6.06 g; Sodium chloride-4.68 g; KCl- 1.49 g; NH₄Cl – 1.07 g; Na₂SO₄-0.43 g; MgCl₂.2H₂O-0.2 g; CaCl₂.2H₂O – 30mg; Distilled water-1000 ml; pH-7) was prepared, splitted in 50 ml aliquots
in 100 ml Erlenmeyer flasks and 0.1 % of insoluble Zn-compound added, steam sterilized for 30 minutes in an autoclave. Then the flasks were inoculated with 1ml suspension of the test culture with a cell load of $10^7$ cells ml$^{-1}$. Three flasks were maintained with an uninoculated control for each treatment. Experiments were done in triplicate. The samples were withdrawn at 0, 4, 8, 12 and 16 days intervals, centrifuged to remove the debris and cells. Ten ml of this solution was fed to Atomic Absorption Spectrophotometer (AAS) to determine the soluble zinc content.

**Determination of pH**

The pH of the zinc solubilizing bacteria (ZSB) culture filtrates and the uninoculated sample was determined at 0, 24, 36, 48, 72, 96 and 120 hrs after inoculation. The culture was filtered using Whatman No.1 filter paper. The pH was estimated using pH meter (Elico pH meter).

**Zinc tolerance by ZSB isolates**

Zinc is a nutrient at low concentration but toxic at higher concentration. The solubilization of zinc might limit the growth of the bacteria at higher level. Unless the cultures tolerate a higher level of zinc, its solubilization may not continue. Therefore the ability of selected isolate to tolerate solubilized zinc was determined under *in vitro* condition in nutrient broth containing different concentrations of soluble zinc (ZnSO$_4$). The nutrient broth was prepared and splitted in 100 ml aliquots in Erlenmeyer flasks. The ZnSO$_4$ was incorporated into the broth in such a way that the final concentration of zinc was 25, 50, 100, 200, 300, 400 and 500 mg kg$^{-1}$. These solutions were divided into 10 ml quantities in test tubes, sterilized at 15 psi for 15 minutes and inoculated with 0.1 ml of ZSB. An uninoculated control was also maintained. The total ZSB population was also maintained by plating on Tris-minimal salt medium. The growth of bacteria in the zinc containing medium indicated their tolerance to zinc.

**Results and Discussion**

There are six isolates were isolated from maize rhizospher soil. These selected isolates were identified based on their morphological and biochemical characters test. The zinc solubilization potential of ZSB cultures in different concentration of insoluble zinc with glucose as carbon source was assessed. The results are presented in Figure 1. The six cultures (ZSB SM-1, 2, 3, 4, 5, 6) established growth at both 0.1 per cent and 0.2 per cent concentration of ZnO, ZnCO$_3$ and Zn EDTA. Among the zinc concentrations, 0.1 percent supported more solubilization compared to 0.2 per cent irrespective of carbon sources and cultures. Among the cultures tested, ZSB SM-1 showed higher solubilization.

Maximum solubilization was observed at 0.1 per cent ZnO in glucose medium (4.2 cm) by ZSB SM-1. The solubilization on Zn-EDTA supplemented medium was comparatively less. With glucose as carbon source at 0.1 % zinc concentration, ZSB SM-1 exhibited the solubilization zone of 4.2, 3.8 and 2.5 cm with zinc oxide, zinc carbonate and zinc EDTA, respectively. Zinc tolerance potential also varied with isolates ZSB SM-1 and ZSB SM-3 (Table 1). The ZSB SM-1 culture had shown a cell count of $1 \times 10^4$ on the 8$^{th}$ day after inoculation in a 500 mg kg$^{-1}$ concentration of zinc, whereas the ZSB SM-3 was completely inhibited at 500 mg kg$^{-1}$ concentration after 8 days of inoculation. Based on the solubilization zone ZSB SM-1 used to further experiment.
Table 1: Population of ZSB SM-1 and ZSB SM-3 on Tris minimal salt medium containing different concentration of Zn (mg g⁻¹)

| Zinc concentration (mg kg⁻¹) | ZSB-SM 1 |          |          |          | ZSB-SM 3 |          |          |          |
|-----------------------------|----------|----------|----------|----------|----------|----------|----------|----------|
|                             | Mean     | Mean     | Mean     | Mean     | Mean     | Mean     | Mean     | Mean     |
|                             | 1st DAI  | 3rd DAI  | 5th DAI  | 8th DAI  | 1st DAI  | 3rd DAI  | 5th DAI  | 8th DAI  |
| 0                           | 212      | 200      | 193      | 186      | 207      | 192      | 183      | 173      |
| 25                          | 117      | 90       | 7        | 6        | 115      | 85       | 5        | 3        |
| 50                          | 158      | 20       | 18       | 14       | 154      | 19       | 13       | 9        |
| 100                         | 189      | 100      | 9        | 8        | 181      | 97       | 6        | 2        |
| 200                         | 60       | 30.5     | 16       | 11       | 57       | 30       | 12       | 8        |
| 300                         | 31       | 26.5     | 2        | 10       | 29       | 25       | 2        | 1        |
| 400                         | 18       | 14       | 1.5      | 0.75     | 17       | 9        | 1        | 0        |
| 500                         | 11       | 6.5      | 2.5      | 1        | 11       | 4        | 0        | 0        |
| Sed                         | 14.7     | 20.3     | 9.6      | 4.6      | 14.8     | 18.3     | 1.9      | 11.5     |
| CD (p = 0.05)               | 34.7     | 48.1     | 22.7     | 10.8     | 35.1     | 43.2     | 4.4      | 27.2     |

*DAI – Day After Inoculation

Table 2: Solubilization of insoluble zinc compounds by ZSB SM-1 (broth assay)

| Time interval | Zinc oxide |         | Zinc carbonate |         | Zn-EDTA |         | Mean |
|---------------|------------|---------|----------------|---------|---------|---------|------|
|               | Available zinc (mg g⁻¹) | Per cent Solubilized | Available zinc (mg g⁻¹) | Per cent Solubilized | Available zinc (mg g⁻¹) | Per cent Solubilized |       |
| 0h            | 10         | 1       | 12             | 1.2     | 9       | 0.9     | 10   |
| 24 h          | 120        | 12      | 100            | 10      | 70      | 7       | 96   |
| 48 h          | 260        | 26      | 220            | 22      | 110     | 11      | 196  |
| 72 h          | 340        | 34      | 230            | 23      | 220     | 22      | 263  |
| 7 days        | 390        | 39      | 350            | 35      | 360     | 36      | 366  |
| 10 days       | 440        | 44      | 410            | 41      | 430     | 43      | 426  |
| Mean          | 260        |         | 220            |         | 200     |         | 226  |

Table 3: Determination of pH in ZSB SM-1

| Different Zinc sources | pH |
|------------------------|----|
|                        | 0 h | 24 h | 36 h | 48 h | 72 h | 96 h | 120 h |
| Zn O                   | 7.3 | 5.8  | 5.4  | 4.4  | 3.9  | 3.5  | 3.0   |
| ZnCO₃                  | 7.3 | 6.0  | 5.8  | 5.6  | 5.5  | 5.0  | 4.8   |
| Zn-EDTA                | 7.3 | 5.9  | 5.3  | 4.7  | 4.0  | 3.8  | 3.8   |
The soluble zinc was assessed by broth assay using the same chemicals. The soluble Zn was in an increasing trend throughout the experiment. On the 10\textsuperscript{th} day of incubation, zinc oxide, zinc carbonate and zinc EDTA solubilized 440, 410 and 430 mg g\textsuperscript{-1} of zinc respectively (Table 2).

In all the three zinc sources, the zinc solubility was doubled after 48 h of inoculation as compared to 24 h. Among the zinc sources, maximum solubilization was observed with zinc oxide on 10\textsuperscript{th} day (44 per cent) followed by zinc EDTA (43 per cent). Among the zinc sources, zinc oxide was solubilized better than zinc carbonate and zinc-EDTA and was significantly different from each other.

The pH of the medium supplemented with different zinc sources behaved differently (Table 3). In the zinc oxide and zinc EDTA added medium, the pH reduced gradually. Also the gradual reduction in pH was noticed in zinc oxide added Tris minimal salt medium broth and it reached to as low a value of 3.0 after 120 hours.
From the results, it’s clear that available zinc levels increased in incubation period. The Zn use efficiency of zinc sulphate is 0.32-1.5 per cent, whereas, under organic based zinc nutrition the efficiency is 2-8 per cent.

Exogenous application of soluble Zn sources, similar to fertilizer application, has been advocated to various crops. This causes transformation of about 96-99 per cent of applied available Zn to various unavailable forms. By considering the resource constraints of the farmers and zinc fertilizers being costly, it is prudent to develop alternate technologies that are economically feasible, environmentally sustainable and zinc efficient. It is in this context, growing interest in the use of beneficial micro-organisms as bio-inoculant assumes significance to increase availability of Zn for assimilation by crop plants. The Zn thus made unavailable can be reverted back to available form by inoculating a bacterial strain capable of solubilizing it.

References

Fasim, F., N. Ahmed, R. Parsons and G.M. Gadd. 2002. Solubilization of zinc salts by a bacterium isolated from the air environment of a tannery. FEMS Microbiol. Lett., 213: 1-6.

Saravanan, V.S., Kalaiarasan, P. M. Madhaiyan, Thangaraju, M. 2007. Solubilization of insoluble zinc compounds by *Gluconacetobacter diazotrophicus* and the detrimental action of zinc ion (*Zn*<sup>2+</sup>) and zinc chelates on root knot nematode *Meloidogyne incognita*. Lett, Appl Microbiol. 44: 235–241.

Sharma, K. Sushil, M.P. Sharma, Aketi Ramesh, and Om P. Joshi. 2012. Characterization of Zinc-Solubilizing *Bacillus* isolates and their potential to influence zinc assimilation in soybean seeds. J. Microbiol. Biotechnol. 22(3), 352-359.

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