Simultaneous Extraction and Determination of Volatile Organic Compounds and Semi-volatile Organic Compounds in Indoor Air using Multi-bed Solid Phase Extraction Device

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Abstract

Simultaneous extraction and determination of indoor volatile compounds including volatile organic compounds (VOCs) and semi-volatile organic compounds (SVOCs) was developed using multi-bed solid phase extraction (SPE)-type collection device. The collection device was prepared by packing styrene-divinylbenzene polymer particles and activated carbon particles. The collected analytes were completely desorbed by passing 7 mL of acetone, and the solvent was then injected into gas chromatograph-mass spectrometry without concentration process. Because the proposed method does not require ultrasonication and a concentration process of eluted solvent, a quantitative determination of relatively volatile compound was achieved. The total recovery including extraction and elution recoveries for all the investigated analytes were in the range from 91.6 to 109%. The limit of quantifications were less than 4.0 ng L\(^{-1}\) for all the investigated analytes, and relative standard deviations of the peak area of the analytes in indoor air were less than 12%. The collection device could be reused over 50 time samplings.

**Keywords** Volatile organic compounds, semi-volatile organic compounds, sick house syndrome, gas chromatography
Introduction

Volatile organic compounds (VOCs) are organic chemical substances that emitted into air at room temperature. VOCs are found in indoor air, and exposure to high concentration VOC or long-term exposure to low level VOCs may cause health effects, such as eye irritation and headaches.¹,² These adverse health effects are called multiple chemical sensitivity or sick building syndrome. To prevent these illnesses, the guidelines and standard measurement methods for the VOCs in indoor air have been defined by the relevant agencies as well as the World Health Organization (WHO).³ In Japan, the Ministry of Health, Labour and Welfare (MHLW) adopted reference values and standard measure method for 13 VOCs in indoor air, such as toluene, ethylbenzene, dibutyl phthalate (DBP), and di-(2-ethylhexyl) phthalate (DEHP).⁴ In 2017, MHLW announced a revision of the reference values and the addition of 2-ethylhexanol (2-EH), 2,2,4-trimethyl-1,3-pentanediol monoisobutyrate (texanol) and 2,2,4-trimethyl-1,3-pentanediol diisobutyrate (TXIB) into the regulated substance.⁵

WHO classifies VOCs with boiling points from 0°C to 50–100°C as very volatile organic compounds (VVOCs), 50–100°C to 240–260°C as VOC, and greater than 250°C as semi-volatile organic compounds (SVOCs). Based on this classification, tetradecane, DBP, DEHP, texanol, and TXIB are categorized by SVOCs, and others are VOCs.

Gas chromatograph coupled with mass spectrometry (GC-MS) analysis is mostly used for determination of VOCs. GC-MS showed higher selectivity and sensitivity, although an
adequate sample preconcentration process is needed for determining trace VOCs in air. Solid adsorption / thermal desorption (TD) and solid adsorption / solvent extraction are widely employed for preconcentration of VOCs in air samples except for formaldehyde and acetaldehyde, and MHLW also recommended these two sample preparation methods. Formaldehyde and acetaldehyde are typically determined by high-performance liquid chromatography after the derivatization with 2,4-dinitrophenylhydrazine due to its high volatility and low detector response. TD method is a solvent-free analytical method, and it has been widely used in recent environmental analysis. However, in most cases an expensive automated TD system is needed, and insufficient desorption from the adsorbent could occur especially for SVOCs. To determine wide variety of volatile compounds (including VOCs and SVOCs), multi-bed-type adsorbents has been employed. In solvent extraction method, an activated carbon-based porous particle is typically employed as adsorbent, and a toxic solvent of carbon disulfide (CS₂) is used as elution solvent. The method requires a long extraction time, and evaporation of volatile compounds during extraction process is considered.

Our research group has developed solid phase extraction (SPE)-type collection devices for extraction of SVOCs in air samples. Recently, a styrene-divinylbenzene (Sty-DVB) polymer particles of Sunpak-H was developed for precise determination of airborne SVOCs. The Sunpak-H-packed SPE-type collection device exhibited rapid and quantitative elution
recovery of collected SVOCs by passing few amount of organic solvent. Because collected analytes can easily eluted from the adsorbent by just passing an organic solvent, the collection device can be repeatedly used by drying the adsorbent that packed in a glass cartridge.

In this study, regulated compounds including VOCs and SVOCs in indoor air to avoid sick building syndrome were simultaneously extracted and determined by a novel multi-bed SPE-type collection device. After the optimization of extraction and elution conditions, the device was applied to the determination of VOCs in real indoor air samples.

**Experimental**

*Chemicals*

Toluene, ethylbenzene, xylene, styrene, \( p \)-dichlorobenzene (\( p \)-DCB), tetradecane, DBP, DEHP, texanol, and TXIB were purchased from Tokyo Kasei Kogyo (Tokyo, Japan). Acetone, and 2-EH were obtained from Kanto Chemical Co., Inc. (Tokyo, Japan). The abbreviation and boiling points of the investigated analytes are listed in Table 1.

*Collection device*

Sunpak-H (50/80 mesh) was prepared by Shinwa Chemical Industries Ltd. (Kyoto, Japan). Carboxen 1000 and Carbopack X were purchased from Sigma-Aldrich Japan (Tokyo, Japan).
Specific surface area of Sunpak-H, Carboxen 1000, and Carbopack X was 100–150 m\(^2\) g\(^{-1}\), 1200 m\(^2\) g\(^{-1}\), and 240 m\(^2\) g\(^{-1}\), respectively. First a stainless steel wire mesh (HORIBA STEC) and a glass filter (15 mm diameter, GA-200, Advantec Tokyo Kaisha, Ltd., Tokyo, Japan) was placed in a specially designed glass cartridge (14.9 mm i.d., 60 mm length, HORIBA STEC Co., Ltd., Kyoto, Japan). Then Sunpak-H (0.3 g) and Carboxen 1000 (0.1 g) or Carbopack X (0.1 or 0.2 g) were packed in the cartridge, and fixed by another glass filter and a PTFE O-ring. Packing length of the Sunpak-H and Carboxen 1000 (0.2g) were 4 mm and 1 mm, respectively. Figure 1 shows photograph and illustration of the developed multi-bed-type collection device. The device was washed with acetone and dried by N\(_2\) flow (5 L min\(^{-1}\)) for 10 min before use, and analytes were not detected on the method blank.

**Analytical method**

The collection recovery for the investigated analytes using the multi-bed-type collection device was calculated as follows. First, 100 μL of standard solution of the analytes (100 mg L\(^{-1}\) each dissolved in acetone) was spiked on the tip side of the device (spiked on the glass filter). Then, another collection device was connected to the first device in tandem, and a setting volume of clean air was sampled using a gas sampling pump (HORIBA STEC) through the two devices at 10 L min\(^{-1}\). After the air sampling, the collected analytes were eluted by passing acetone for respective collection device. The collection recovery was
calculated by the ratio of peak area detected on the former collection device (spiked device) and latter collection device. When any analytes were detected on the latter device, the collection recovery could be calculated to 100%. The elution recovery of the spiked analytes was calculated by sequential solvent elution. When any analytes were detected on the 2nd elution, the elution recovery could be calculated to 100%. The eluted solvent of 2 μL was injected into GC-MS without concentration process. The collection device was repeatedly used by drying the adsorbent with N₂ flow (5 L min⁻¹) for 10 min.

**GC-MS measurement**

A JEOL JMS-Q1000 GCMk-II system (JEOL, Tokyo, Japan) was used for all the GC-MS measurements. Helium (>99.999% purity) was used as the carrier gas at a head pressure of 100 kPa. The injector and interface temperature were set at 300 and 320°C, respectively. Sample solutions (2 μL) were injected by split mode (1:10). A fused silica capillary column of HP-5 (30 m × 0.25 mm, 0.25 μm film thickness, Agilent Technologies, Santa Clara, CA, USA) was used for analytes separation. The column temperature was held at 40°C for 3 min, then programmed to 300°C at a rate of 20°C min⁻¹. The mass spectrometer was operated in the selected ion monitoring mode. The selected ions were summarized in Table 1.

**Results and discussion**
Evaluation of extraction and elution performances

As investigated in previous studies, SVOCs including DBP, DEHP, 2-EH, texanol, and TXIB were successfully extracted on Sunpak-H, and they were completely and quantitatively eluted by passing acetone for 7 mL or under.\textsuperscript{14,15} On the other hand, only Sunpak-H (0.3 g) packed device showed incomplete extraction of toluene, where the extraction recoveries of toluene at air sampling volume of 300 and 600 L were 98.4 and 94.5%, respectively. Therefore, another adsorbent that has higher extraction property than Sunpak-H was introduced on the latter side of Sunpak-H. First, Carboxen 1000 (0.1 g) was introduced into the collection device at the latter side of Sunpak-H (0.3 g). The Sunpak-H/Carboxen 1000 device showed complete extraction of all the investigated analytes, where any compounds were detected from 2nd extraction device. However, toluene was not successfully eluted from the device due to strong adsorption onto Carboxen 1000 even if 15 mL of acetone was used. The desorption efficiency for toluene using Sunpak-H/Carboxen 1000 device was about 98%. Then, Carbopack X was investigated as the 2nd adsorbent. The Sunpak-H (0.3 g)/Carbopack X (0.1 g) packed device demonstrated incomplete extraction efficiency of toluene at sampling volume of 600 L (60 min). Therefore, the amount of Carbopack X was increased to 0.2 g. The Sunpak-H (0.2 g)/Carbopack X (0.2 g) packed device exhibits complete extraction of all the investigated analytes up to sampling volume of 600 L. In addition, the extracted analytes were completely eluted from the device by passing of acetone. Table 2 showed the elution
efficiency of the investigated VOCs and SVOCs using Sunpak-H (0.2 g)/Carbopack X (0.2 g) packed device. All the investigated analytes were completely and rapidly eluted by using 8 mL of acetone. The elution time was approximately 3 min.

Evaluation of the method

The additional recovery of the proposed method was investigated by comparing the peak area obtained by standard solution to spiked sample. To prepare the standard solution, 100 µL of stock solution including all the investigated analyte at 100 mg L⁻¹ dissolved in acetone was dissolved in 8 mL of acetone. On the other hand, the same stock solution of 100 µL was spiked onto the Sunpak-H (0.2 g)/Carbopack X (0.2 g) packed device, and then 600 L of clean air was collected (16.7 ng L⁻¹ as in air sample). After that, spiked sample solution was obtained by eluting spiked analytes by 8 mL of acetone. The additional recoveries for all the investigated analytes were in the range from 90.1 to 109% (n = 3) (Table 3). The results clearly showed that all the investigated analytes including VOCs and SVOCs were successfully and quantitatively recovered by the proposed method.

The limit of detection (LOD) and the limit of quantification (LOQ) of the proposed method were summarized in Table 3. The LOD was defined as signal to noise ratio of 3.3, and the LOQ were that of 10. The results showed satisfactory sensitivity of the proposed method for all the investigated analytes. The relative standard deviations (RSDs) of the peak area
obtained by measuring the standard solution that prepared by spiking 100 µL of stock solution (100 mg L⁻¹) onto the collection device, collecting 600 L of clean air, and eluting the analytes by 8 mL of acetone were less than 12% for all the analytes.

Since quantitative recovery of the proposed method for all the investigated VOCs and SVOCs were confirmed, the method was applied to the determination of VOCs and SVOCs in indoor air and in-car air samples. The air samples were collected for 600 L (10 L min⁻¹ × 60 min) at a height of 1.0 m above the ground at ambient temperature in November 2019. The quantitative results of determined compounds are listed in Table 4. Typical chromatogram for the determination of VOCs and SVOCs spiked with standard solution (100 mg L⁻¹ solution 100 µL spiked) and clean room air are shown in Fig. 2(A) and 2(B), respectively. All of the detected analytes were determined without co-eluting compounds, and the results clearly indicated successful determination of trace VOCs and SVOCs in air sample with simple sample collection method.
Conclusions

Multi-bed SPE-type collection device packed with Sunpak-H and Carbopack X showed quantitative extraction and elution performances for indoor air VOCs and SVOCs. Because the extracted analytes could be completely elute by passing 8 mL of acetone, the eluted solvent can be directly injected into GC-MS without sample preconcentration with satisfactory sensitivity. In addition, the method does not require use of toxic CS2 that typically used as a desorption solvent in solvent extraction for VOC analysis. The developed collection device showed quantitative determination of wide range of VOCs in air samples. Therefore, the method could be useful for quantification of several types VOCs in gaseous samples.
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Figure captions

Figure 1  Photograph and illustration of the multi-bed-type SPE device.

Figure 2  Chromatogram for the determination of VOCs and SVOCs.

(A) spiked sample (B) clean room air.
Fig. 1
Fig. 2
Table 1  Analyte VOCs and SVOCs in this study.

| Compound                        | Abbreviation | Boiling point / °C | Measured ion / m z⁻¹ |
|---------------------------------|--------------|--------------------|----------------------|
| Toluene                         | Tol          | 111                | 91, 92               |
| Ethylbenzene                    | EB           | 136                | 91, 106              |
| Xylene                          | Xy           | 138-144            | 91, 106              |
| Styrene                         | Sty          | 145                | 78, 104              |
| p-Dichlorobenzene               | p-DCB        | 174                | 111, 146             |
| 2-Ethyl hexanol                 | 2-EH         | 184                | 57, 70               |
| 2,2,4-Trimethyl-1,3-pentanediol| Texanol      | 254-260            | 56, 71               |
| monoisobutyrate                 |              |                    |                      |
| Tetradecane                     | C₁₄          | 254                | 57, 71               |
| 2,2,4-Trimethyl-1,3-pentanediol | TXIB         | 280-281            | 56, 71               |
| monoisobutyrate                 |              |                    |                      |
| Dibutyl phthalate               | DBP          | 340                | 149, 223             |
| Di(2-ethylhexyl) phthalate      | DEHP         | 385                | 149, 279             |
Table 2  Elution recovery of the analytes.

| Solvent volume / mL | Tol | EB  | m,p-Xy | o-Xy | Sty  | p-DCB | 2-EH | Texanol | C_{14} | TXIB | DBP | DEHP |
|---------------------|-----|-----|--------|------|------|-------|------|---------|-------|------|-----|------|
| 6                   | 99.9| 99.9| 99.9   | 100  | 99.8 | 99.8  | 100  | 100     | 99.7  | 99.8 | 100 | 100  |
| 7                   | 99.6| 99.8| 99.6   | 99.7 | 99.3 | 99.9  | 100  | 100     | 99.4  | 98.1 | 100 | 100  |
| 8                   | 100 | 100 | 100    | 100  | 100  | 100   | 100  | 100     | 100   | 100  | 100 | 100  |
Table 3  LOD, LOQ and additional recovery of the proposed method.

| Compound   | LOD / ng L⁻¹ | LOQ / ng L⁻¹ | Reference value / ng L⁻¹ | Additional recovery, % |
|------------|--------------|--------------|--------------------------|------------------------|
| Tol        | 0.13         | 0.40         | 260                      | 102                    |
| EB         | 0.13         | 0.40         | 3800                     | 94.5                   |
| m,p-Xy     | 0.67         | 2.0          | 200                      | 102                    |
| o-Xy       | 0.67         | 2.0          | 200                      | 91.6                   |
| Sty        | 0.67         | 2.0          | 220                      | 97.9                   |
| p-DCB      | 0.67         | 2.0          | 240                      | 109                    |
| 2-EH       | 1.3          | 4.0          | (130)a                   | 94.5                   |
| Texanol    | 1.3          | 4.0          | (240)a                   | 102                    |
| C₁₄        | 0.67         | 2.0          | 330                      | 105                    |
| TXIB       | 0.13         | 0.40         | (100)a                   | 97.4                   |
| DBP        | 1.3          | 4.0          | 17                       | 104                    |
| DEHP       | 0.67         | 2.0          | 100                      | 94.6                   |

a: Considered guideline values.
| Compound       | Concentration / ng L⁻¹ | Teachers room | Laboratory | Clean room | Car |
|---------------|------------------------|---------------|------------|------------|-----|
| Tol           | 0.65                   | 40.6          | 6.36       | 79.9       |     |
| EB            | 0.53                   | 4.54          | 1.06       | 4.31       |     |
| m,p-Xy        | <LOQ                   | 6.37          | 2.15       | 7.50       |     |
| o-Xy          | N.D.                   | 4.40          | <LOQ       | 4.44       |     |
| Sty           | <LOQ                   | 4.37          | <LOQ       | 4.47       |     |
| p-DCB         | <LOQ                   | 2.93          | <LOQ       | 2.91       |     |
| 2-EH          | 5.30                   | 8.85          | 8.03       | <LOQ       |     |
| Texanol       | N.D.                   | N.D.          | 4.40       | N.D.       |     |
| C₁₄           | <LOQ                   | <LOQ          | N.D.       | <LOQ       |     |
| TXIB          | N.D.                   | N.D.          | N.D.       | N.D.       |     |
| DBP           | N.D.                   | N.D.          | N.D.       | N.D.       |     |
| DEHP          | <LOQ                   | N.D.          | N.D.       | N.D.       |     |

N.D.; Not detected.