Effect of empagliflozin monotherapy on postprandial glucose and 24-hour glucose variability in Japanese patients with type 2 diabetes mellitus: a randomized, double-blind, placebo-controlled, 4-week study

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Abstract

**Background:** This study evaluated the effect of empagliflozin on postprandial glucose (PPG) and 24-hour glucose variability in Japanese patients with type 2 diabetes mellitus (T2DM).

**Methods:** Patients (N = 60; baseline mean [SD] HbA1c 7.91 [0.80]%; body mass index 24.3 [3.2] kg/m²) were randomized to receive empagliflozin 10 mg (n = 20), empagliflozin 25 mg (n = 19) or placebo (n = 21) once daily as monotherapy for 28 days. A meal tolerance test and continuous glucose monitoring (CGM) for 24 hours were performed at baseline and on days 1 and 28. The primary endpoint was change from baseline in area under the glucose concentration-time curve 3 hours after breakfast (AUC₁⁻₄h for PPG) at day 28.

**Results:** Adjusted mean (95%) differences versus placebo in changes from baseline in AUC₁⁻₄h for PPG at day 1 were −97.1 (−126.5, −67.8) mg · h/dl with empagliflozin 10 mg and −91.6 (−120.4, −62.8) mg · h/dl with empagliflozin 25 mg (both p < 0.001 versus placebo) and at day 28 were −85.5 (−126.0, −45.0) mg · h/dl with empagliflozin 10 mg and −104.9 (−144.8, −65.0) mg · h/dl with empagliflozin 25 mg (both p < 0.001 versus placebo). Adjusted mean (95% CI) differences versus placebo in changes from baseline in 24-hour mean glucose (CGM) at day 1 were −20.8 (−27.0, −14.7) mg/dl with empagliflozin 10 mg and −23.9 (−30.0, −17.9) mg/dl with empagliflozin 25 mg (both p < 0.001 versus placebo) and at day 28 were −24.5 (−35.4, −13.6) mg/dl with empagliflozin 10 mg and −31.7 (−42.5, -20.9) mg/dl with empagliflozin 25 mg (both p < 0.001 versus placebo). Changes from baseline in mean amplitude of glucose excursions (MAGE; CGM) were not significantly different with either empagliflozin dose versus placebo at either timepoint. Curves of mean glucose (CGM) did not change between baseline and day 1 or 28 with placebo, but shifted downward with empagliflozin. Percentage of time with glucose ≥70 to <180 mg/dl increased from 52.0% at baseline to 77.0% at day 28 with empagliflozin 10 mg and from 55.0% to 81.1% with empagliflozin 25 mg, without increasing time spent with hypoglycemia.

**Conclusion:** Empagliflozin for 28 days reduced PPG from the first day and improved daily blood glucose control in Japanese patients with T2DM.

**Trial registration:** Clinicaltrials.gov NCT01947855

**Keywords:** SGLT2 inhibitor, Continuous glucose monitoring, CGM
Background

The prevalence of diabetes in Japan is increasing [1]. Cardiovascular and all-cause mortality are increased in Japanese patients with diabetes [2].

Postprandial hyperglycemia is common in patients with type 2 diabetes (T2DM) [3,4]. Control of postprandial glucose (PPG) helps patients to achieve HbA1c goals [5,6], and some guidelines for the management of T2DM provide specific targets for PPG [7-9]. Postprandial hyperglycemia is an independent risk factor for cardiovascular disease [10,11], possibly due to the oxidative stress, endothelial dysfunction and overexpression of adhesion molecules triggered by acute hyperglycemia and glucose fluctuations [12,13]. Daily glucose fluctuations may also increase the risk of microvascular and macrovascular complications associated with T2DM [14,15] while severe hypoglycemia is associated with increased mortality [16,17].

Inhibition of the sodium glucose cotransporter 2 (SGLT2), located in the proximal tubule, reduces renal glucose reabsorption, leading to increased urinary glucose excretion and reduced hyperglycemia in patients with T2DM [18,19]. Empagliflozin is a selective and potent SGLT2 inhibitor [20]. In international Phase III trials in patients with T2DM, 24 weeks’ treatment with empagliflozin given as monotherapy or as add-on therapy for 24 weeks was well tolerated and significantly reduced glycated hemoglobin (HbA1c), fasting plasma glucose (FPG), body weight and systolic blood pressure (SBP) versus placebo [21-24]. In Japanese patients with T2DM, empagliflozin monotherapy for 52 weeks led to sustained reductions in HbA1c, FPG, body weight and SBP [25,26]. The effect of empagliflozin on 24-hour glycemic variability in patients with T2DM has not been assessed.

This study was conducted to evaluate the effect of empagliflozin 10 mg and 25 mg once daily as monotherapy for 28 days on PPG and 24-hour glycemic variability in Japanese patients with T2DM.

Methods

This was a randomized, double-blind, placebo-controlled, parallel-group study conducted at two sites in Japan. The clinical trial protocol was approved by the Institutional Review Boards of the participating centers, and complied with the Declaration of Helsinki in accordance with the International Conference on Harmonisation Harmonised Tripartite Guideline for Good Clinical Practice. All patients provided written informed consent. The trial was registered with ClinicalTrials.gov (NCT01947855).

Patients

Japanese patients with T2DM aged ≥20 and ≤74 years, with a body mass index (BMI) ≤40 kg/m², who were on a diet and exercise regimen and were drug-naïve (no anti-diabetes agents for ≥12 weeks prior to consent) or treated with 1 oral anti-diabetes agent (except a sulfonylurea at > half maximum approved daily dose, or a thiazolidinedione) at an unchanged dose for ≥12 weeks prior to consent, were eligible for inclusion. At screening, drug-naïve patients were required to have HbA1c ≥ 7% and ≤ 10% and patients treated with 1 oral anti-diabetes agent were required to have HbA1c ≥ 7% and ≤ 9.5%. All patients were required to have HbA1c ≥ 7% to ≤ 10% at the start of the placebo run-in period.

Exclusion criteria included uncontrolled hyperglycemia (glucose level > 240 mg/dl) after an overnight fast confirmed by a second measurement; acute coronary syndrome, stroke or transient ischemic attack ≤ 20 weeks prior to randomization; indication of liver disease (alanine aminotransferase, alkaline aminotransferase or alkaline phosphatase levels > 3 times the upper limit of normal during screening, washout or run-in); impaired renal function (estimated glomerular filtration rate [eGFR] < 60 ml/min/1.73 m² according to Japanese estimation equation [27]) during screening, washout or run-in; gastrointestinal surgeries that induce chronic malabsorption; treatment with insulin, glucagon-like peptide-1 (GLP-1) analogues, sulfonylurea at > half the daily maximum approved dose or thiazolidinedione within 12 weeks prior to consent; treatment with anti-obesity drugs within 12 weeks prior to consent; use of any treatment at screening leading to unstable body weight; treatment with systemic steroids at time of consent; change in dosage of thyroid hormones within 6 weeks prior to consent; alcohol or drug abuse within 12 weeks of consent; investigational drug intake in another trial within 30 days of consent.

Randomization and interventions

All patients underwent a 2-week, open-label, placebo run-in period. Patients pre-treated with an oral anti-diabetes agent underwent a 2-week washout period prior to the placebo run-in. Following the run-in period, eligible patients were randomized (in a 1:1:1 ratio) to receive empagliflozin 10 mg, empagliflozin 25 mg, or placebo for 28 days. Patients were monitored at the trial site from days −2 to 2 and days 27 to 29. Blinded 24-hour continuous glucose monitoring (CGM) and a meal tolerance test (MTT) were performed at day −1, day 1 (of treatment) and day 28 (Figure 1). Patients were assigned to test meals providing 1440, 1600, or 1840 kcal/day, based on patient’s standard weight (Additional file 1: Table S1). Test meals contained 50–60% carbohydrate, 15–21% protein, and 21–35% fat (Additional file 1: Table S1). Plasma glucose profiles were determined at the timepoints shown in Figure 1.

Endpoints

The primary endpoint was the change from baseline (day −1) in the area under the glucose concentration-
time curve 3 hours after breakfast (AUC\textsubscript{1--4h} for PPG) at day 28. Other efficacy endpoints were change from baseline in AUC\textsubscript{1--4h} for PPG at day 1, change from baseline in AUC of glucose 3 hours after dinner (AUC\textsubscript{10--13h} for PPG) at day 1 and day 28, change from baseline in 2-hour PPG after each meal (breakfast, lunch, dinner) at day 1 and day 28, change from baseline in FPG at day 2 and 29 and change from baseline in AUC\textsubscript{1--4h} and AUC\textsubscript{10--13h} for postprandial insulin at day 1 and day 28. Endpoints measured from CGM at day 1 and day 28 were changes from baseline in 24-hour mean glucose, mean amplitude of glucose excursions (MAGE) \cite{28} and the percentage of time with glucose ≥180 mg/dl, ≥70 to <180 mg/dl and <70 mg/dl per day. MAGE was calculated as the arithmetic mean difference between consecutive blood glucose peaks (between meals) and nadirs (between the peaks) when differences were >1 standard deviation of the mean glucose value in the same 24-hour period. Change from baseline in HbA1c was measured at day 29. Change from baseline in urinary excretion of 8-iso-prostaglandin F2\alpha (8-iso-PGF2\alpha; a marker of oxidative stress) in the fasting state and in the 24 hours after study drug administration was measured at day 28. Safety endpoints included changes in vital signs, weight, and clinical laboratory parameters, and adverse events (AEs; preferred terms coded according to the Medical Dictionary for Drug Regulatory Activities [MedDRA] version 16.1). AEs included all events with an onset after the first dose and up to 7 days after the last dose of study medication. Confirmed hypoglycemic AEs were defined as AEs with plasma glucose ≤70 mg/dL and/or requiring assistance. Events consistent with urinary tract infection (UTI), genital infection, and volume depletion were identified using prospectively defined search categories using 77, 89 and 8 preferred terms, respectively.

### Statistical analysis

Efficacy analyses were performed on the full analysis set (FAS) which included patients treated with ≥1 dose of study drug who had a baseline value for AUC\textsubscript{1--4h} for PPG. Safety was assessed in the treated set (patients treated with ≥1 dose of study drug).

The primary endpoint was analyzed using an analysis of covariance (ANCOVA) model, with treatment, baseline eGFR and number of previous anti-diabetes medications as fixed effects and baseline HbA1c and baseline AUC\textsubscript{1--4h} for postprandial insulin as linear covariates. Missing data were not imputed. In the hierarchical testing procedure, the superiority of empagliflozin 25 mg versus placebo was to be tested first, followed by empagliflozin 10 mg versus placebo if the first test was significant. Other efficacy endpoints were analyzed using the ANCOVA model described for the primary endpoint, with the baseline value for the endpoint in question as an additional linear covariate.

Safety analyses were descriptive, except for changes in lipid parameters, free fatty acids and blood ketone bodies, which were analyzed using ANCOVA. Postprandial insulin data and triglyceride data were log-transformed prior to analysis.

A sample size of 20 patients per treatment group was required to provide power of 95% for the pair-wise comparison and an overall power of ≥90% to detect a 150 h·mg/dl treatment difference in AUC\textsubscript{1--4h} for PPG for each
empagliflozin dose compared to placebo, assuming a standard deviation of 120 h · mg/dl and a dropout rate of 2 patients per group.

**Results**

**Patients**

Of 78 patients screened, 60 patients were randomized and treated and comprised the FAS. One patient in the placebo group discontinued prematurely. Baseline characteristics were balanced across treatment groups (Table 1).

**Efficacy**

Compared with placebo, empagliflozin 10 mg and 25 mg led to significant reductions from baseline in AUC$_{1-4h}$ for PPG at day 1 and at day 28 (Figure 2A) and in AUC$_{10-13h}$ for PPG at day 1 and at day 28 (Figure 2B). Reductions in AUC$_{1-4h}$ and AUC$_{10-13h}$ for PPG at day 28 compared with placebo were greater with empagliflozin 25 mg than empagliflozin 10 mg (No statistical tests were performed on the differences between the empagliflozin 10 mg and 25 mg groups). Empagliflozin 10 mg and 25 mg reduced AUC$_{1-4h}$ and AUC$_{10-13h}$ for post-prandial insulin at day 1 and day 28, but changes in AUC$_{1-4h}$ with empagliflozin 10 mg at day 28 were not significantly different to placebo (Table 2).

Changes from baseline in 2-hour PPG were significantly greater with empagliflozin 10 mg and 25 mg compared with placebo after breakfast at day 1 and day 28 (Figure 2C). Changes from baseline in 2-hour PPG after lunch were significantly greater with empagliflozin 10 mg compared with placebo at day 1, but did not reach significance versus placebo with empagliflozin 10 mg at day 28 or with empagliflozin 25 mg at day 1 or day 28 (Figure 2C).

Changes from baseline in 2-hour PPG after dinner were significantly different with empagliflozin 10 mg and empagliflozin 25 mg compared with placebo at day 28 but not at day 1 (Figure 2C).

Empagliflozin 10 mg and 25 mg led to significant reductions from baseline in FPG compared with placebo at day 2 and at day 29 (Figure 3). Reductions from baseline in FPG at day 29 compared with placebo were greater with empagliflozin 25 mg than empagliflozin 10 mg.

Empagliflozin 10 mg and 25 mg led to significant reductions from baseline in 24-hour mean glucose compared with placebo at day 1 and at day 28 (Figure 4). Reductions from baseline in 24-hour mean glucose compared with placebo at day 28 were greater with empagliflozin 25 mg than empagliflozin 10 mg. Mean glucose levels over 24 hours by CGM at baseline, day 1 and day 28 are shown in Figure 5. A reduction from baseline (downward shift) in mean glucose levels at all timepoints over 24 hours was evident from day 1 with empagliflozin, and reductions from baseline

| Table 1 Patient demographics and baseline characteristics (full analysis set) |
|-----------------------------|---------------------|---------------------|
|                            | Placebo            | Empagliflozin 10 mg | Empagliflozin 25 mg |
| N                           | 21                 | 20                  | 19                   |
| Male                        | 17 (81.0)          | 14 (70.0)           | 16 (84.2)            |
| Age (years)                 | 60.7 (10.8)        | 64.8 (5.9)          | 62.6 (7.8)           |
| Time since diagnosis of type 2 diabetes |                     |                     |                      |
| ≤5 years                    | 9 (42.9)           | 4 (20.0)            | 5 (26.3)             |
| >5 to 10 years              | 9 (42.9)           | 8 (40.0)            | 7 (36.8)             |
| >10 years                   | 3 (14.3)           | 8 (40.0)            | 7 (36.8)             |
| Number of anti-diabetes medications |                     |                     |                      |
| 0                           | 18 (85.7)          | 16 (80.0)           | 17 (89.5)            |
| 1                           | 3 (14.3)           | 4 (20.0)            | 2 (10.5)             |
| Body weight (kg)            | 67.7 (10.0)        | 63.5 (10.6)         | 65.9 (12.1)          |
| Body mass index (kg/m$^2$)  | 24.9 (2.8)         | 24.1 (3.7)          | 24.0 (3.2)           |
| HbA1c (%)                   | 8.00 (0.82)        | 7.99 (0.83)         | 7.73 (0.75)          |
| Fasting plasma glucose (mg/dl)* | 154.5 (19.8)   | 151.0 (21.6)       | 151.9 (23.3)         |
| AUC$_{1-4h}$ for postprandial glucose (mg · h/dl) | 682.8 (91.2) | 680.4 (92.2) | 658.1 (116.2) |
| 24-hour mean glucose (mg/dl) | 184.1 (30.5)       | 181.3 (25.9)       | 178.4 (33.4)         |
| Mean amplitude of glucose excursions, MAGE (mg/dl) | 91.4 (26.4) | 94.1 (18.5) | 89.1 (29.7) |
| Estimated glomerular filtration rate (ml/min/1.73 m$^2$) (Japanese estimation equation [27]) | 82.6 (12.8) | 76.5 (11.1) | 80.7 (9.3) |
| Systolic blood pressure (mmHg)* | 119.8 (11.5) | 119.1 (15.9) | 124.0 (11.6) |
| Diastolic blood pressure (mmHg)* | 71.8 (7.7)     | 70.7 (10.7)       | 74.7 (8.0)           |

Data are n (%) or mean (standard deviation). *Baseline for these parameters is day 1.
Figure 2 Changes from baseline in (A) AUC\textsubscript{1-4h} for PPG, (B) AUC\textsubscript{10-13h} for PPG and (C) 2-hour PPG after each meal, based on analyses of covariance in the full analysis set. CI, confidence interval; PPG, postprandial glucose; SE, standard error.
Table 2 Changes in postprandial insulin after breakfast and dinner at day 1 and day 28

|                      | Placebo (n = 20) | Empagliflozin 10 mg (n = 20) | Empagliflozin 25 mg (n = 19) |
|----------------------|------------------|-----------------------------|-----------------------------|
| **AUC_{1-4h}** for postprandial insulin, μIU · h/ml |                  |                             |                             |
| Baseline             | 66.4 (63.5)*     | 58.9 (55.6)                 | 65.3 (46.2)                 |
| Relative change from baseline at day 1 | 1.2 (1.1, 1.3)  | 1.0 (0.9, 1.1)              | 1.0 (0.9, 1.0)              |
| Difference vs placebo (95% CI)  | 0.8 (0.7, 0.9)  | 0.8 (0.7, 0.9)              | 0.8 (0.7, 0.9)              |
| p-value              | <0.001           | <0.001                      | <0.001                      |
| Relative change from baseline at day 28 | 1.0 (0.9, 1.1)  | 0.9 (0.8, 0.9)              | 0.8 (0.7, 0.9)              |
| Difference vs placebo (95% CI)  | 0.9 (0.8, 1.0)  | 0.8 (0.7, 0.9)              | 0.8 (0.7, 0.9)              |
| p-value              | 0.074            | 0.002                       | 0.002                       |
| **AUC_{10-13h}** for postprandial insulin, μIU · h/ml |                  |                             |                             |
| Baseline             | 73.8 (56.6)†     | 60.7 (60.5)                 | 70.0 (53.7)                 |
| Relative change from baseline at day 1 | 1.1 (1.0, 1.2)  | 0.9 (0.8, 1.0)              | 0.9 (0.8, 0.9)              |
| Difference vs placebo (95% CI)  | 0.8 (0.8, 0.9)  | 0.8 (0.7, 0.9)              | 0.8 (0.7, 0.9)              |
| p-value              | 0.002            | <0.001                      | <0.001                      |
| Relative change from baseline at day 28 | 1.0 (0.9, 1.0)  | 0.8 (0.8, 0.9)              | 0.8 (0.7, 0.9)              |
| Difference vs placebo (95% CI)  | 0.9 (0.8, 1.0)  | 0.8 (0.7, 0.9)              | 0.8 (0.7, 0.9)              |
| p-value              | 0.011            | 0.001                       | 0.001                       |

Log-transformed data. Baseline data are gMean (% gCV), change from baseline data are adjusted gMean ratio (95% CI) based on analysis of covariance (ANCOVA) in the full analysis set. *63.8 (65.5) for day 1 analysis (n = 21). †72.8 (55.5) for day 1 analysis (n = 21).

Figure 3 Change from baseline in FPG at day 2 and day 29 based on analyses of covariance in the full analysis set. CI, confidence interval; FPG, fasting plasma glucose; SE, standard error.
seemed to be slightly greater with empagliflozin 25 mg than empagliflozin 10 mg (Figure 5). At day 1, adjusted mean (SE) changes from baseline in MAGE were 15.1 (3.5), 11.0 (3.7) and 8.9 (3.7) mg/dl with placebo, empagliflozin 10 mg and empagliflozin 25 mg, respectively. At day 28, adjusted mean (SE) changes from baseline in MAGE were −4.7 (4.5), −3.7 (4.6) and −2.2 (4.7) mg/dl with placebo, empagliflozin 10 mg and empagliflozin 25 mg, respectively. Differences were not statistically significant with either empagliflozin dose compared with placebo at either time-point. Compared with placebo, empagliflozin 10 mg and 25 mg reduced the percentage of time with glucose ≥180 mg/dl (p < 0.01), and increased the percentage of time with normoglycemia (glucose ≥70 to <180 mg/dl) (p < 0.01) without significantly increasing the percentage of time with hypoglycemia (glucose <70 mg/dl) (Figure 6; Additional file 1: Table S2).

At day 29, adjusted mean (SE) changes from baseline in HbA1c were −0.11 (0.06)% with placebo compared with −0.46 (0.06)% with empagliflozin 10 mg (adjusted mean [95% CI] difference: −0.35% [−0.52, −0.19]; p < 0.001) and −0.63 (0.06)% with empagliflozin 25 mg (adjusted mean [95% CI] difference: −0.52% [−0.68, −0.35]; p < 0.001).

Consistent with reductions in PPG, the excretion of 8-iso-PGF2α, a marker of oxidative stress, was significantly reduced from baseline with empagliflozin 10 mg and 25 mg compared with placebo at day 28 in the fasting state (Table 3). Reductions from baseline in the excretion of 8-iso-PGF2α in the 24 hours after study drug administration were only significantly different with empagliflozin 25 mg compared with placebo at day 28 (Table 3).

### Safety

AEs were reported in 2 patients (9.5%) on placebo, 3 patients (15.0%) on empagliflozin 10 mg and 3 patients (15.8%) on empagliflozin 25 mg. No severe AEs, serious AEs, or AEs leading to discontinuation occurred. No hypoglycemic AEs were reported. One patient (on empagliflozin 25 mg) experienced an AE consistent with genital infection (bartholinitis). No AEs consistent with UTI or volume depletion were reported. No AEs of diabetic ketoacidosis or those related to abnormal ketone body levels were reported.

At day 29, weight was reduced from baseline by 0.9 kg, 1.7 kg and 2.1 kg with placebo, empagliflozin 10 mg and 25 mg, respectively (Additional file 1: Table S3). Acute changes in SBP and diastolic BP (DBP) (at day 2) with empagliflozin compared with placebo were small, and more pronounced reductions were observed at day 29 (Additional file 1: Table S3). In contrast, pulse rate appeared to increase with empagliflozin compared with placebo at day 2, but changes from baseline in pulse rate were similar between empagliflozin and placebo at day 29 (Additional file 1: Table S3).

Compared with placebo, there were no significant differences in changes from baseline in total cholesterol or
Figure 5 Mean glucose over 24 hours by CGM. CGM, continuous glucose monitoring.
LDL-cholesterol with empagliflozin 10 mg or 25 mg (Table 4). Compared with placebo, HDL-cholesterol was significantly increased with empagliflozin 10 mg and 25 mg, and triglycerides were significantly reduced with empagliflozin 10 mg and 25 mg, at day 29. There were significant increases from baseline in free fatty acids with empagliflozin 25 mg, but not with empagliflozin 10 mg, compared with placebo at day 29. There were significant increases from baseline in blood ketone bodies with empagliflozin 10 mg and 25 mg compared with placebo at day 29 (Table 4).

No clinically relevant changes in electrolytes (sodium, potassium, calcium, magnesium, phosphate) were observed in any group at the end of treatment (Additional file 1: Table S4). Changes from baseline in hematocrit and eGFR were generally small in all groups (Additional file 1: Table S4).

**Conclusions**

This study was conducted to evaluate the effect of empagliflozin as monotherapy for 28 days on PPG and 24-hour

**Table 3 Changes in urinary excretion of 8-iso-PGF2α at day 28**

|                | Placebo (n = 20) | Empagliflozin 10 mg (n = 20) | Empagliflozin 25 mg (n = 19) |
|----------------|-----------------|-----------------------------|-----------------------------|
| **Fasting state, pg/ml** |                 |                             |                             |
| Baseline       | 197.8 (27.0)    | 194.6 (29.4)                | 146.5 (18.5)                |
| Change from baseline at day 28 | 40.6 (22.6)    | −48.1 (23.3)                | −33.5 (23.7)                |
| Difference vs placebo (95% CI) | −88.6 (−154.6, −22.7) | −74.0 (−139.8, −8.2)         |                             |
| p-value        | 0.010           | 0.028                       |                             |
| **In 24 hours after drug administration, pg/ml** |                 |                             |                             |
| Baseline       | 115.5 (11.0)    | 138.3 (20.6)                | 148.6 (21.7)                |
| Change from baseline at day 28 | −3.7 (10.3)    | −28.4 (10.4)                | −46.8 (10.6)                |
| Difference vs placebo (95% CI) | −24.7 (−54.5, 5.2) | −43.1 (−72.9, −13.3)        |                             |
| p-value        | 0.013           | 0.006                       |                             |

Baseline data are mean (standard error [SE]), change from baseline data are adjusted mean (SE) based on analysis of covariance (ANCOVA) in the full analysis set.
glycemic variability in Japanese patients with T2DM. Significant reductions from baseline in AUC$_{1-4h}$ for PPG were observed after acute and subchronic treatment with empagliflozin, with 80–90% of the reduction in AUC$_{1-4h}$ for PPG already achieved at day 1.

At day 28, although the reductions from baseline in AUC for PPG with empagliflozin observed after dinner were of a lower magnitude than those observed after breakfast, the reductions observed after dinner were significant. These observations were consistent with reductions in 2-hour PPG. The sustained effect of empagliflozin on PPG from morning to evening support once-daily administration of empagliflozin.

Of note, the reduction in PPG in this study was accompanied by a reduction in postprandial insulin levels. In contrast to insulin secretagogues and incretins, empagliflozin’s mode of action is independent of beta-cell function and insulin secretion [18]. By increasing urinary glucose excretion, empagliflozin reduces plasma glucose levels leading to a reduction in plasma insulin levels [29].

CGM can provide valuable information on the magnitude and duration of glucose fluctuations [30]. In this study, empagliflozin improved daily blood glucose control measured using CGM, with the curves of mean 24-hour glucose lower at day 1 and day 28 than at baseline. Consistent with changes in FPG, PPG and HbA1c, slightly greater

**Table 4 Changes in fasting serum lipids and ketone bodies at day 29**

|                          | Placebo (n = 20) | Empagliflozin 10 mg (n = 20) | Empagliflozin 25 mg (n = 19) |
|--------------------------|-----------------|-----------------------------|-----------------------------|
| **Total cholesterol, mg/dl** |                 |                             |                             |
| Baseline                 | 212.8 (7.7)     | 200.1 (8.4)                 | 208.7 (9.1)                 |
| Change from baseline at day 29 | −0.8 (3.6)     | 3.2 (3.7)                   | 1.1 (3.7)                   |
| Difference vs placebo (95% CI) | 4.0 (−6.6, 14.6) | 1.9 (−8.6, 12.3)            |                             |
| p-value                  | 0.455           | 0.722                       |                             |
| **HDL-cholesterol, mg/dl** |                 |                             |                             |
| Baseline                 | 46.8 (2.2)      | 47.6 (2.8)                  | 46.8 (2.8)                  |
| Change from baseline at day 29 | 0.1 (1.2)      | 4.7 (1.3)                   | 7.4 (1.3)                   |
| Difference vs placebo (95% CI) | 4.5 (1.0, 8.1) | 7.2 (3.7, 10.8)             |                             |
| p-value                  | 0.014           | <0.001                      |                             |
| **LDL-cholesterol, mg/dl** |                 |                             |                             |
| Baseline                 | 128.7 (6.3)     | 124.9 (8.4)                 | 130.5 (8.5)                 |
| Change from baseline at day 29 | −1.3 (3.2)     | 3.4 (3.3)                   | 4.3 (3.3)                   |
| Difference vs placebo (95% CI) | 4.7 (−4.6, 14.0) | 5.7 (−3.5, 14.8)            |                             |
| p-value                  | 0.314           | 0.221                       |                             |
| **Triglycerides, mg/dl**  |                 |                             |                             |
| Baseline                 | 159.6 (51.0)    | 131.5 (34.9)                | 142.2 (48.0)                |
| Relative change from baseline at day 29 | 1.0 (0.9, 1.1) | 0.8 (0.7, 0.9)              | 0.7 (0.6, 0.8)              |
| Difference vs placebo (95% CI) | 0.8 (0.7, 1.0) | 0.7 (0.6, 0.8)              |                             |
| p-value                  | 0.037           | <0.001                      |                             |
| **Free fatty acids, mg/dl** |                 |                             |                             |
| Baseline                 | 10.8 (0.8)      | 9.6 (0.7)                   | 9.3 (0.6)                   |
| Change from baseline at day 29 | 3.1 (0.8)      | 4.8 (0.8)                   | 7.4 (0.8)                   |
| Difference vs placebo (95% CI) | 1.7 (−0.6, 4.1) | 4.3 (2.0, 6.7)              |                             |
| p-value                  | 0.149           | <0.001                      |                             |
| **Ketone bodies, μmol/l** |                 |                             |                             |
| Baseline                 | 86.8 (10.2)     | 78.1 (12.0)                 | 69.4 (10.8)                 |
| Change from baseline at day 29 | −12.2 (44.0)   | 139.5 (44.7)                | 4080.4 (45.5)               |
| Difference vs placebo (95% CI) | 151.7 (244, 2790) | 402.2 (2933, 5472)          |                             |
| p-value                  | 0.021           | <0.001                      |                             |

Unless otherwise indicated, baseline data are mean (standard error [SE]), change from baseline data are adjusted mean (SE) based on analysis of covariance (ANCOVA) in the treated set. Fasting measurements. *Log-transformed data; baseline data are geometric mean (%CV) and change from baseline data are adjusted geometric mean ratio (95% CI).
treatment with empagliflozin 25 mg as

Adverse event; ANCOVA: Analysis of covariance; AUC: Area under the

Table S2.

ketone bodies >3000

parable with levels of up to about 1300

in an individual patient was 1449

The highest level of ketone bodies observed in our study

empagliflozin 25 mg and placebo, respectively, at day 29.

levels in the range of physiological conditions, which is

bodies most likely reflects an adaptive change, with ketone

crease in ketone bodies was modest, with adjusted mean

range of 1 ± 2 mM [33,34]. In this study, the average in-

creased free fatty acid levels and ketogenesis. The most

glucagon-to-insulin ratio [29], leading to lipolysis, in-

increased free fatty acid levels and ketogenesis. The most

common causes of ketosis are physiological conditions, in

which mild to moderate elevations of circulating ketone

bodies occur in response to fasting or prolonged exercise,

with ketone body levels not uncommonly rising to the

range of 1 ± 2 mM [33,34]. In this study, the average in-

crease in ketone bodies was modest, with adjusted mean

levels of 218, 486 and 66 μmol/l for empagliflozin 10 mg,

empagliflozin 25 mg and placebo, respectively, at day 29.

The highest level of ketone bodies observed in our study

in an individual patient was 1449 μmol/l, which is com-

parable with levels of up to about 1300 μmol/l reported

for subjects without diabetes after an overnight fast [35].

Diabetic ketoacidosis is typically accompanied by levels of

ketone bodies >3000 μmol/l [33] and develops almost ex-

clusively in states of absolute insulin deficiency. In con-

trast, the lowering of insulin levels with empagliflozin is

probably secondary to the reduction in plasma glucose

levels via increased urinary glucose excretion, which is ac-

companied by an improvement in beta-cell function [29].

Therefore, the empagliflozin-induced increase in ketone

bodies most likely reflects an adaptive change, with ketone

levels in the range of physiological conditions, which is

unlikely to put patients at risk of ketoacidosis in the ab-

sence of absolute (endogenous or exogenous) insulin defi-

ciency or extreme (ketogenic) diets.

Patients with T2DM have an increased risk of develop-

ing cardiovascular events compared with the general

population [36], which is related to the prevalence of the

classical cardiovascular risk factors of hypertension and

dyslipidemia, in addition to other important factors such

as glycemic control, oxidative stress, and obesity [37]. El-

evated PPG is an independent risk factor for cardiovas-

cular disease [10,11]; however, improvements in PPG

have not been shown to translate into reduced risk of

cardiovascular disease [38]. Empagliflozin improves gly-

cemic control with a low risk of hypoglycemia, leads to

weight loss and reduces blood pressure, possibly due to

diuretic effects, weight loss, or direct vascular effects

[21-26,39,40]; further, as demonstrated in this study,

empagliflozin reduces PPG and 8-iso-PGF2α, a marker of

oxidative stress that is an independent risk marker for

cardiovascular disease [41]. A cardiovascular outcome

trial (EMPA-REG OUTCOME; NCT01131676) is in-

vestigating the effect of empagliflozin in patients with

T2DM and high cardiovascular risk [42].

In conclusion, empagliflozin 10 mg or 25 mg as mono-

therapy for 28 days significantly reduced PPG and FPG

and improved daily blood glucose control in Japanese

patients with T2DM, without increasing time spent with

a hypoglycemic blood glucose level.

Additional file

Additional file 1: Table S1. Test meals. Table S2. Changes in

percentage of time with glucose level ≥180 mg/dl, ≥70 to <180 mg/dl,

and <70 mg/dl. Table S3. Changes in blood pressure, pulse rate and

weight. Table S4. Laboratory measurements.

Abbreviations

AE: Adverse event; ANCOVA: Analysis of covariance; AUC: Area under

glucose concentration-time curve; BMI: Body mass index; CGM: Continuous

glucose monitoring; CI: Confidence interval; DBP: Diastolic blood pressure;

eGFR: Estimated glomerular filtration rate; FAS: Full analysis set; FPG: Fasting

plasma glucose; HbA1c: Glycated hemoglobin; HDL-cholesterol: High-density

lipoprotein cholesterol; LDL-cholesterol: Low-density lipoprotein cholesterol;

MAGE: Mean amplitude of glucose excursions; MedDRA: Medical Dictionary

for Drug Regulatory Activities; MTT: Meal tolerance test; PPG: Postprandial

glucose concentration-time curve; BMI: Body mass index; CGM: Continuous

glucose monitoring; CI: Confidence interval; DBP: Diastolic blood pressure;

eGFR: Estimated glomerular filtration rate; FAS: Full analysis set; FPG: Fasting

plasma glucose; HbA1c: Glycated hemoglobin; HDL-cholesterol: High-density

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MAGE: Mean amplitude of glucose excursions; MedDRA: Medical Dictionary

for Drug Regulatory Activities; MTT: Meal tolerance test; PPG: Postprandial

glucose. SD: Standard deviation; SE: Standard error; SGLT2: Sodium glucose
cotransporter 2; T2DM: Type 2 diabetes mellitus; UTI: Urinary tract infection.

Competing interests

RN has received research support from Japan Diabetes Foundation,

Boehringer Ingelheim, Daichi-Sankyo and Astellas; has participated in

speaker’s bureau/advisory panels for Novo Nordisk, Eli Lilly, Sanofi,

Astellas, Boehringer Ingelheim, Daichi-Sankyo, Tanabe-Mitsubishi, Astra,

Zeneca, Kowa, Ono, Johnson & Johnson, Medtronic, Takeda and Astellas,

and served as a consultant for Boehringer Ingelheim and Eli Lilly. KK, YK,

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Authors’ contributions
RN contributed to the study design, acquisition and interpretation of data and reviewed/edited the manuscript. KK, YT, KI, TH, AS, SSL and UCB contributed to the study design and interpretation of data and reviewed/edited the manuscript. All authors read and approved the final manuscript.

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