Effects of Rosuvastatin on Apolipoprotein J in Balloon-Injured Carotid Artery in Rats

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Abstract

Background: Restenosis after percutaneous coronary intervention in coronary heart disease remains an unsolved problem. Clusterin (CLU) (or Apolipoprotein [Apo] J) levels have been reported to be elevated during the progression of postangioplasty restenosis and atherosclerosis. However, its role in neointimal hyperplasia is still controversial.

Objective: To elucidate the role Apo J in neointimal hyperplasia in a rat carotid artery model in vivo with or without rosuvastatin administration.

Methods: Male Wistar rats were randomly divided into three groups: the control group (n = 20), the model group (n = 20) and the statin intervention group (n = 32). The rats in the intervention group were given 10mg /kg dose of rosuvastatin. A 2F Fogarty catheter was introduced to induce vascular injury. Neointima formation was analyzed 1, 2, 3 and 4 weeks after balloon injury. The level of Apo J was measured by real-time PCR, immunohistochemistry and western blotting.

Results: Intimal/medial area ratio (intimal/medial, I/M) was increased after balloon-injury and reached the maximum value at 4 weeks in the model group; I/M was slightly increased at 2 weeks and stopped increasing after rosuvastatin administration. The mRNA and protein levels of Apo J in carotid arteries were significantly upregulated after rosuvastatin administration as compared with the model group, and reached maximum values at 2 weeks, which was earlier than in the model group (3 weeks).

Conclusion: Apo J served as an acute phase reactant after balloon injury in rat carotid arteries. Rosuvastatin may reduce the neointima formation through up-regulation of Apo J. Our results suggest that Apo J exerts a protective role in the restenosis after balloon-injury in rats. (Arq Bras Cardiol. 2018; 111(4):562-568)

Keywords: Coronary Artery Disease; Percutaneous Coronary Intervention; Rosuvastatin Calcium; Apolipoprotein J; Coronary Reestenosis; Rats.

Introduction

Coronary heart disease (CHD) is one of the most common cardiovascular diseases with high morbidity and mortality. Major effective techniques for myocardial revascularization are percutaneous coronary intervention (PCI) and coronary bypass surgery. Percutaneous transluminal coronary angioplasty (PTCA) is an effective treatment for CHD, but its effect in long-term is influenced by a high restenosis rate. Although drug eluting stents (DES) combined with dual antiplatelet therapy greatly reduce the occurrence of restenosis, the incidence rate still exceeds 10%. The mechanism underlying restenosis after PCI has been widely studied worldwide, but effective cellular or molecular targets for the treatment of restenosis after PCI urgently needs to be identified.

Clusterin (CLU), or Apolipoprotein (Apo) J, is a heterodimeric glycoprotein, which is composed of α and β subunits linked by disulfide bond. The coding gene of Apo J is located on chromosome 8p21-p12, mainly encoding two isoforms – secretory CLU (sCLU) and nuclear CLU (nCLU). Apo J has been reported to be induced during the progression of postangioplasty restenosis and atherosclerosis. However, the role of Apo J in neointimal hyperplasia is still controversial.

Methods

Animals

Male Wistar rats weighing 350-400 g were randomly divided into three groups: control group (n = 20), model group
Balloon injury

The rats were weighed on the day of operation, and randomly divided into three groups. The rats in the intervention group were given 10 mg/kg dose of rosuvastatin. A 2F Fogarty catheter was introduced to induce vascular injury as previously reported. Briefly, the rats were anesthetized after intraperitoneal injection of 10% chloral hydrate at a dose of 0.3 mL/100 g body weight. A 2F balloon catheter was inserted into aortic outlet of carotid artery. The balloon was then inflated and pulled back 3 times to denude the endothelium.

At 1, 2, 3 and 4 weeks after surgery, rats were anesthetized by intraperitoneal injection of 10% chloral hydrate at a dose of 0.3 mL/100 g body weight. Then, the animals were sacrificed by intravenous administration of 2-3 mL of potassium chloride solution via subclavian vein; 0.3 cm of the right carotid artery was fixed in 10% neutral formalin for pathological examination, and the other part was frozen immediately in liquid nitrogen and stored at -80°C for further use.

Hematoxylin-eosin (HE) staining

Vascular specimens were fixed in 10% formaldehyde solution for 3-4h. Routine dehydration and paraffin embedding were performed. The sections were cut evenly and the thickness was 4 μm. The injury of blood vessels was observed under light microscope.

Immunohistochemistry (IHC) assay

The level of Apo J was assessed by IHC in rat carotid artery. The primary antibody (polyclonal rabbit anti-human Apo J IgG) was purchased from Santa Cruz, Inc. (Cat No. sc-6354). The secondary antibody (labeled goat anti-rat/rabbit IgG polymer) was purchased from Maixin BioTech (Fuzhou, China). All photos were captured and saved using the iSCapture system, and data collection and analysis are performed using the Image Pro Plus 6 image processing software.

Enzyme-linked immunosorbent assay (ELISA)

Venous blood was collected and centrifugated at 3000r/min for 10 min. The supernatant was collected using a micropipette and stored in the refrigerator at -20°C for use. The samples were then thawed at room temperature for ELISA. ELISA was performed using a commercial kit (Rat Competitive ELISA for Apolipoprotein J A 252 SC), following the manufacturers‘ instructions.

Real-time polymerase chain reaction (PCR)

The mRNA level of Apo J was detected by real-time PCR in rat carotid artery. RNA was extracted by Trizol one-step extraction method, and reverse transcription was performed. Primers used for amplification for Apo J were as follows: Forward, TAA GGA GAT TCA GAA CGC GC; reverse, ATC CCT GGT GTC ATC TAG AG. Primers for the control GAPDH were as follows: Forward, GTG ATG CTG GTG CCG AGT AG; reverse, GGT GCC AGT GAT GCC GTG C. Real-time PCR reactions were prepared following the instructions of SYBR® Premix Ex Taq ™ system (Perfect Real Time). The mRNA levels in each sample were calculated by 2^-ΔΔCt.

Western blotting

Proteins were extracted from 30 mg of rat carotid artery. Briefly, proteins were separated using SDS-PAGE with 10% separation gel and 5% concentrated gel. Then the separated proteins were transferred into polyvinylidene difluoride (PVDF) membranes. The membranes were blocked and incubated with antibodies. Relative levels of Apo J were analyzed using Image Lab analysis software. β-actin was used as inner control. Bands were quantified using QUANTITY ONE software (Bio-Rad, Hercules, CA, USA).

Statistical analysis

Statistical analysis was performed using SPSS 20.0. Quantitative data were expressed by mean ± standard deviation (SD). The difference between two groups was compared using independent-samples t test; comparisons between three groups were analyzed using one-way ANOVA (analysis of variance). P < 0.05 was considered statistically significant.

Results

Survival and success rates of rat carotid artery model

Among the 52 rats of the model group and the intervention group, 2 died during operation by suffocation, and 2 died for arterial hemorrhage 12h after operation. Therefore, 47 rats survived with approximately 90% survival rate. The pathological examination showed intimal hyperplasia and thickening in the experimental group, suggesting that the model was successfully constructed. The mean operation time was 34.19 ± 6.09 min. The feasibility and success rate of this model can be highly reproducible if surgical procedures are properly performed.

Level of serum Apo J

There was no significant difference in serum Apo J level before and after operation in the intervention group (Table 1). There was no significant difference in the level of serum Apo J at 1, 2, 3 and 4 weeks before (F = 1.002, p = 0.408) of after (F = 0.189, p = 0.903) operation.

Statin intervention inhibited intimal hyperplasia

Results of pathological examination showed that no intimal hyperplasia was observed in the control group (Figure 1). In the model group and intervention group, the intima was slightly thickened 1 week after operation, and further thickened 2 weeks after operation. During 3 weeks after operation, the degree of intimal hyperplasia was further aggravated in the model group; however, this change was not as marked as at 2 weeks after operation, and the cells gradually became paralleled. During 4 weeks after operation, the degree of intimal hyperplasia was further aggravated in the model group, but no significant changes were observed regarding the degree of intimal hyperplasia as compared with week 3 (Figure 2 and 3).
Table 1 – Serum levels of apolipoprotein J (Apo J) before and after operation in the statin intervention group

| Time points | Pre-operation | Post-operation |
|-------------|---------------|----------------|
|             | n  | Apo J    | n  | Apo J    | t'   | p*   |
| 1           | 7  | 13.498 ± 3.015 | 7  | 10.317 ± 3.567 | 1.802 | 0.097 |
| 2           | 7  | 14.062 ± 4.538 | 7  | 11.516 ± 1.762 | 1.383 | 0.192 |
| 3           | 8  | 11.234 ± 2.740 | 8  | 11.117 ± 3.104 | 0.08  | 0.937 |
| 4           | 8  | 14.143 ± 4.609 | 8  | 11.205 ± 3.579 | 1.424 | 0.176 |

F* 1.002
F* 0.408

*p value < 0.05 was considered statistically significant.

Intimal and medial membrane areas were measured using Image Pro Plus 6, and intimal/medial area ratio (intimal/medial, I/M) was used to indicate the degree of intimal hyperplasia. As shown in Table 2, the I/M was close to 0 in the control group and was significantly different from that in the model group and the intervention group at all time points (1, 2, 3 and 4 weeks). There were significant differences of I/M between different time points in the model group, and I/M reached the maximum at the fourth week. No significant difference of I/M was observed between 2, 3 and 4 weeks post-surgery in the intervention group, and I/M in the intervention group was significantly lower than that in the model group (Table 2). Taken together, our results suggest that rosuvastatin could significantly inhibit intimal hyperplasia in rats.

Figure 1 – Hematoxylin-eosin (HE) staining in the control group. Upper, magnification 40×; lower, magnification 400×.

Figure 2 – Hematoxylin-eosin (HE) staining in the model group and in intervention group 1 week (w), 2 weeks, 3 weeks and 4 weeks after balloon injury of rat carotid arteries; magnification 40×.
Level of Apo J in carotid arteries

The mRNA levels of Apo J were measured by real-time PCR. Our results showed that the Apo J mRNA level was strikingly increased 2 weeks after operation, reached to a peak at the 3rd week, and decreased at the 4th week post-surgery in the model group. In intervention group, the Apo J mRNA level was strikingly increased and reached to a peak at the 2nd week, and decreased at the 3rd and 4th week post-surgery in the intervention group. In addition, the mRNA level of Apo J was higher in the intervention group than in the model group at the 1st week after operation. At the 2nd week post-surgery, the mRNA level of Apo J was strikingly increased in both groups and was significantly higher in the statin-intervention group than the model group (Table 3). Similar results have been observed in the protein levels of Apo J as shown in Figure 4. Our results showed that rosuvastatin could significantly increase the expression level of Apo J in balloon-injured rat carotid arteries.

Discussion

In the present study, we found that I/M increased after balloon-injury and reached the maximum at 4w in the model group; also, I/M was slightly increased at 2w and stopped increasing after rosuvastatin administration. Our results suggest that rosuvastatin could significantly reduce the degree of intimal hyperplasia in balloon-injured carotid arteries in rats. The levels of Apo J mRNA and protein in carotid arteries were significantly upregulated after rosuvastatin administration as compared with the model group, and reached to maximum at 2 weeks, which was earlier than the in the model group. Our results suggest that rosuvastatin may inhibit intimal hyperplasia through upregulation of Apo J after balloon-injury in rats.

Apo J has been reported to be closely related to cardiovascular diseases, such as atherosclerosis and restenosis after angioplasty.14,15 Ishikawa et al.7 revealed the distribution of Apo J in the extracellular matrix of endarterium in human atherosclerotic aorta, and its potential protective role against human atherosclerosis by cholesterol transport from the aortic wall to the liver. It has been reported that Apo J is increased in tissue injury and cell stress, and plays vital role in protection against oxidative stress, cell lysis and apoptotic cell death.16-21 Additionally, ApoJ could be observed in active tissue remodeling. These findings indicate that ApoJ may act as an acute phase reactant. In the present study, we observed a marked neointimal thickening 2 weeks post-surgery, with proliferation and migration VSMCs observed by HE staining in the model group. The proliferation and migration of VSMC were the most active at week 3, and decrease at week 4. In the meantime, the mRNA and protein levels of Apo J were significantly increased at week 2, reached a peak at 3 weeks after operation, and then decreased at 4 weeks. The results showed high expression of Apo J in the phase of active proliferation and migration of VSMCs. Consistent with other studies, our results suggest that Apo J may be an acute phase reactant after balloon-injury in rat carotid arteries.

Table 2 – Intimal/medial (I/M) area ratio in the study groups

| Time points | Control group | Model group | Statin intervention group | t’ | p’ |
|-------------|---------------|-------------|---------------------------|----|----|
|             | n | I/M      | n | I/M      | n | I/M      |     |     |
| 1           | 5 | 0.04 ± 0.07 | 5 | 0.63 ± 0.40<sup>a</sup> | 5 | 0.42 ± 0.04<sup>a</sup> | 10.066 | < 0.001 |
| 2           | 5 | 0.01 ± 0.02 | 4 | 1.08 ± 0.29<sup>a</sup> | 4 | 1.29 ± 0.31<sup>a</sup> | 39.639 | < 0.001 |
| 3           | 5 | 0.03 ± 0.03 | 4 | 1.81 ± 0.11<sup>b</sup> | 4 | 1.47 ± 0.54<sup>b</sup> | 37.142 | < 0.001 |
| 4           | 5 | 0.05 ± 0.04 | 4 | 2.61 ± 1.12<sup>c</sup> | 4 | 1.50 ± 0.26<sup>c</sup> | 20.287 | < 0.001 |
| F<sup>v</sup> | 0.741 | 9.432 | 21.393 |<sup>✓</sup> |<sup>✓</sup> |<sup>✓</sup> |<sup>✓</sup> |
| P<sup>v</sup> | 0.543 | < 0.001 | < 0.001 |<sup>✓</sup> |<sup>✓</sup> |<sup>✓</sup> |<sup>✓</sup> |

<sup>1</sup> t test used to compare the differences between the two groups. <sup>1F one-way ANOVA (analysis of variance) to compare the difference between all four groups. <sup>p value < 0.05 was considered statistically significant</sup>
In-stent restenosis after interventional procedures has become one of the most urgent problems to be solved worldwide. Rosuvastatin, a potent hydroxymethylglutaryl coenzyme A (HMG-CoA) reductase inhibitor, has been reported to reduce neointimal thickening after vascular endothelial injury in rats. In the present study, the rats in the intervention group received intragastric administration of rosuvastatin (10 mg/kg/d). In accordance with other studies, we found that rosuvastatin significantly reduced the neointima formation.

It has been reported that secreted isoform of Apo J (sCLU) could inhibit the proliferation and migration of VSMCs. Kim et al. also found that Apo J could significantly inhibit neointimal hyperplasia using adenovirus-mediated overexpression of Apo J in rats. In the present study, we found that the mRNA and protein levels of ApoJ in carotid arteries were significantly upregulated after rosuvastatin administration as compared with the model group. Moreover, Apo J reached a maximum at week 2 after rosuvastatin administration, and that was earlier than the model group which reached peak expression at the third week. These results suggest that rosuvastatin may increase the level of Apo J in the balloon-injured carotid arteries, which indirectly indicates a protective role of Apo J against restenosis after balloon-injury in rats.

**Conclusion**

Our results showed that Apo J served as an acute phase reactant after balloon-injury in rat carotid arteries. Rosuvastatin may reduce the neointima formation through further up-regulation of Apo J. Our findings suggest that Apo J exerts a protective role against restenosis after balloon-injury in rats.

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**Author contributions**

Conception and design of the research and Writing of the manuscript: Yang N, Qin Q; Acquisition of data: Yang N, Dong B, Yang J, Li Y, Kou L, Liu Y; Analysis and interpretation of the data: Yang N, Dong B, Yang J, Li Y, Kou L; Statistical
analysis: Yang N, Dong B; Critical revision of the manuscript for intellectual content: Yang N, Dong B, Yang J, Li Y, Qin Q.

Potential Conflict of Interest
No potential conflict of interest relevant to this article was reported.

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References

1. Sharma PK, Chhatravallia AK, Cohen DJ, Jiang JS, Baweja P, Gosch K, et al. Predicting long-term bleeding after percutaneous coronary intervention. Catheter Cardiovasc Interv. 2017;89(2):199-206.
2. Lee JY, Park DW, Kim YH, Yun SC, Kim WJ, Kang SJ, et al. Incidence, predictors, treatment, and long-term prognosis of patients with restenosis after drug-eluting stent implantation for unprotected left main coronary artery disease. J Am Coll Cardiol. 2011;57(12):1349-58.
3. Shannan B, Seifert M, Boothman DA, Tilgen W, Reichrath J. Clusterin and DNA repair: a new function in cancer for a key player in apoptosis and cell cycle control. J Mol Histol. 2006;37(5-7):183-90.
4. Tougakos IP, Gonos ES. Regulation of clusterin/apolipoprotein J, a functional homologue to the small heat shock proteins, by oxidative stress in ageing and age-related diseases. Free Radic Res. 2006;40(12):1324-34.
5. Park S, Mathis KW, Lee IK. The physiological roles of apolipoprotein J/clusterin in metabolic and cardiovascular diseases. Rev Endocr Metab Disord. 2014;15(1):45-53.
6. Gelissen IC, Hochgrebe T, Wilson MR, Easterbrook-Smith SB, Jessup W, Dean RT, et al. Apolipoprotein J (clusterin) induces cholesterol export from macrophage-foam cells: a potential anti-atherogenic function? Biochem J. 1998;331(Pt 1):231-7.
7. Ishikawa Y, Akasaka Y, Ishii T, Komiyama K, Masuda S, Asuwa N, et al. Distribution and synthesis of apolipoprotein J in the atherosclerotic aorta. Arterioscler Thromb Vasc Biol. 1998;18(4):665-72.
8. Navab M, Anantharamaiah GM, Reddy ST, Van Lenten BJ, Wagner AC, Hama S, et al. An oral apol peptide renders HDL antiinflammatory in mice and monkeys and dramatically reduces atherosclerosis in apolipoprotein E-null mice. Arterioscler Thromb Vasc Biol. 2005;25(9):1932-7.
9. Miyata M, Biro S, Kaieda H, Eto H, Orihara K, Kihara T, et al. Apolipoprotein J/clusterin is induced in vascular smooth muscle cells after vascular injury. Circulation. 2001;104(12):1407-12.
10. Millis AJ, Luciani M, McCue HM, Rosenberg ME, Moulson CL. Clusterin regulates vascular smooth muscle cell nodule formation and migration. J Cell Physiol. 2001;186(2):210-9.
11. Shirasawa T, Miyata M, Eto H, Hamada N, Akasaka Y, Miyauchi T, et al. Deficiency of clusterin inhibits neointimal hyperplasia after vascular injury. J Atheroscler Thromb. 2009;16(6):772-81.
12. Kim HJ, Yoo EK, Kim JY, Choi YK, Lee HJ, Kim JK, et al. Protective role of clusterin/apolipoprotein J against neointimal hyperplasia via anti-proliferative effect on vascular smooth muscle cells and cytoprotective effect on endothelial cells. Arterioscler Thromb Vasc Biol. 2009;29(10):1558-64.
13. Preusch MR, Vanakaris A, Bea F, Jeronimakis N, Shimizu T, Konstandin M, et al. Rosuvastatin reduces neointima formation in a rat model of balloon injury. Eur J Med Res. 2010;15(11):461-7.
14. Garcia-Rodriguez S, Arias-Santiago S, Perandres-Lopez R, Orgaz-Molina I, Castellote L, Buendia-Eisman A, et al. Decreased plasma levels of clusterin in patients with psoriasis. Actas Dermosifiliogr. 2013;104(6):497-503.
15. Vanni AE, Agroggiannis G, Chekans C, Perrea D. Clusterin/Apolipoprotein J immunolocalization on carotid artery is affected by TNF-alpha, cigarette smoking and anti-platelet treatment. Lipids Health Dis. 2014 Apr 23;13:70.
16. Witte DR, Aronow BJ, Stauderman ML, Stuart WD, Clay MA, Gruppo RA, et al. Platelet activation releases megakaryocyte-synthesized apolipoprotein J, a highly abundant protein in atheromatous lesions. Am J Pathol. 1993;143(3):763-73.
17. Sivamurthy N, Stone DH, Logero FW, Quist WC. Apolipoprotein J inhibits the migration, adhesion, and proliferation of vascular smooth muscle cells. J Vasc Surg. 2001;34(4):716-23.
18. Foglio E, Puddighin G, Fasanaro P, D’Arcangelo D, Perrone GA, Mocini D, et al. Apolipoprotein J (clusterin) induces cholesterol export from macrophage-foam cells: a potential anti-atherogenic function? Biochem J. 1998;331(Pt 1):231-7.
19. Van Dijk A, Vermond RA, Krijnen PA, Juffermans LJ, Hahn NE, Makker SP, et al. Intravenous clusterin administration reduces myocardial infarct size in rats. Eur J Clin Invest. 2010;40(10):893-902.
20. Lee YN, Shin YJ, Kang BH, Park JH, Min BH. Over-expression of human clusterin increases stress resistance and extends lifespan in Drosophila melanogaster. Biochem Biophys Res Commun. 2012;420(4):851-6.
21. Pereira RM, Mekary RA, da Cruz Rodrigues KC, Anaruma CP, Ropelle ER, da Silva AS, et al. Protective molecular mechanisms of clusterin against apoptosis in cardiomyocytes. Heart Fail Rev. 2018;23(1):123-9.
22. van der Hardt P, Cironnevecchen HC, Roks AJ, Balkema H, Zijlstra F, van Gilst WH, et al. Rosuvastatin attenuates angiotensin II-induced neointimal formation after stent implantation in the rat. Coron Artery Dis. 2008;19(1):47-53.
23. Kappert K, Leppanen O, Paulsson J, Furuhashi M, Carlsson MA, Heldin CH, et al. Highly active antiretroviral therapy attenuates re-endothelialization and alters neointima formation in the rat carotid artery after balloon injury. J Acquir Immune Defic Syndr. 2006;43(4):383-92.
24. Luan Z, Chase AJ, Newby AC. Statins inhibit secretion of metalloproteinases-1, -2, -3, and -9 from vascular smooth muscle cells and macrophages. Arterioscler Thromb Vasc Biol. 2003;23(5):769-75.
25. Miwa Y, Takahashi-Yanagi F, Morimoto S, Sasaguri T. Involvement of clusterin in 15-deoxy-delta-12,14-prostaglandins [2-induced vascular smooth muscle cell differentiation. Biochem Biophys Res Commun. 2004;319(1):163-8.
