Concomitant in Vivo Voltammetric and Electrophysiological Analysis Indicate that Nociceptin/Orphanin FQ Affects Dopamine and then Serotonin Activities in Brain Substancia Nigra.

Francesco Crespi¹,*

¹Biology, GSK, Verona, Italy

Abstract

Nociceptin/orphanin-FQ (NOCI) together with its receptor NOP are widely expressed in cortical and subcortical motor areas and it is known that NOCI acts as an anxiolytic attenuating the behavioral inhibition of animals acutely exposed to stressful/anxiogenic conditions.

Influence of NOCI upon the dopaminergic system has been observed in the ventral tegmental area and in the nucleus accumbens as well as an inhibitory action of NOCI is described upon serotoninergic mechanisms at cells and terminal levels. In particular, it is known that serotoninergic fibers from the raphe system project to the substantia nigra (SN) and that this modulation is behaviourally relevant.

In the present work, the effect of exogenous NOCI injected into the SN upon DA and 5-HT levels have been analyzed by means of differential pulse voltammetry and nafion-carbon fiber microelectrodes. Electrophysiological monitoring of multicell activity was concomitantly performed with the same microsensor.

It appeared that both levels of these biogenic amines were specularly altered, with possibly a driving influence of the DA activity upon the serotoninergic function(s).

Corresponding author: Francesco Crespi, Biology, GSK, Verona, Italy, Email: fm.crespi@libero.it

Keywords: Nociceptin/orphanin-FQ, substantia nigra, dopamine, serotonin, in vivo voltammetry-electrophysiology

Received: Apr 06, 2019  Accepted: Apr 19, 2019  Published: Apr 23, 2019

Editor: Mozhgan Torabi, PhD. in Animal Physiology, Department of Biology, Faculty of Science, Shahid Chamran University of Ahvaz, Iran.
Introduction

Nociceptin/orphanin-FQ (NOCI) is an opioid-like neuropeptide that activates a G-protein coupled receptor: the NOP receptor [1]. NOCI and its receptor are widely expressed in cortical and subcortical motor areas [2].

In 1997 Jenck and Coll. [3] demonstrated that NOCI acts as an anxiolytic, attenuating the behavioral inhibition of animals acutely exposed to stressful/anxiogenic conditions although the anxiolytic mechanism of NOCI is at present not completely clarified.

It has been reported that treatment with NOCI reduces the firing activity of dopamine (DA) cells in the ventral tegmental area (VTA) [4] and inhibits DA release in the nucleus accumbens [5]. This is resulting in altered regulation of motor control [1, 6].

Concerning serotonin (5-HT), it is known that serotonergic fibers from the raphe system project to the substantia nigra (SN) and that this modulation is behaviourally relevant [7]. In particular, an inhibitory action of NOCI is described upon serotonergic mechanisms exerted at two different levels:

1. On dorsal raphe nucleus (RDN) neurons, where NOCI causes inhibition by increasing K+ conductance [8], and
2. On cortical serotonergic nerve terminals, where NOCI inhibits 5-HT release [9].

In the present work, the influence of NOCI upon DA and 5-HT release in SN is analyzed in vivo, in situ and in real time by means of electrochemical (voltammetric) experiments using nafion-coated carbon fiber microelectrodes (mCFE) for selective measurement of these two neurotransmitters [10]. In addition, the same mCFE is used for concomitant electrophysiology and voltammetric measurements in SN as described earlier [11].

Methods and Results

Animals

Experiments were performed on adult male Wistar rats (260-300 g, Harlan, Italy). All procedures were carried out in accordance with the Italian law (Legislative Decree no.116, 27 January 1992), which acknowledges the European Directive 86/609/EEC, and were fully compliant with GSK policy on the care and use of laboratory animal and codes of practice.

In vitro experiments using differential pulse voltammetry (DPV) with mCFE have been performed to analyze at first the electrochemical characteristics of NOCI. The mCFE was prepared and DPV was performed as already described [10] (figure 1).

In vitro data showed that this peptide (17 amino acids) is not electroactive up to a concentration 1 µM dissolved in artificial cerebral spin fluid (aCSF) at pH 7.4. This is useful information as it allows performing local injection of the peptide in SN without alteration of the voltammetric analysis.

In vivo experiments have been performed in anesthetized (urethane 1.5g/kg i.p.) adult male rats with the nafion-mCFE stereotaxically inserted in the SN following Paxinos and Watson coordinates [12]. A stainless-steel injector was placed into the SN near the mCFE (less than 1mm).

In control experiments two groups of rats (n=4 each) were treated with artificial cerebral spin fluid (aCSF: vehicle, 0.5µl), that was injected in SN in 8-10s. The values of DA and 5-HT remained constant during 90min. (figures 2 and 3).

In NOCI experiments local injection of NOCI (0.5µl, 0.1, 1 or 10 nanomoles, n=4 each dose) resulted in

- A rapid, significant increase of DA levels in the SN up to 270% of control values within 10min then stabilizing at approximately 230% of control values. In particular, this was observed following local injection of NOCI 1 nanomole (nm), a significant increase to approximately 150% of control values was also obtained following local injection of NOCI 0.1 nm. In contrast, the highest dose (10 nm) was resulting in a decrease of the DA signal (figure 2 top).

- A dose-dependent, rapid decrease of 5-HT levels in the SN.

In particular, NOCI 10 nm was significantly decreasing 5-HT levels to approx. 30% of control values within 10min and NOCI 1nm to approx. 60% of control values within 40-50 min.

Parallel electrophysiological monitoring performed in the same animals with the same mCFE as described earlier [11] also indicated a dose-dependent,
Figure 1. Top left: schematic representation of the mCFE: the protruding active tip of the carbon fiber (diam. 12µm) can be cut under the microscope at a length of 200–250µm for DPV - electrophysiology measurements. Top right: the tip of the mCFE before (left) and after nafion coating as described earlier (Crespi et al., 1988).

Bottom, DPV voltammograms obtained in vitro in aCSF solution containing NOCI 1 µm (line a) or containing nM DA and 5-HT (line b) or obtained in vivo in the SN (line c).

Note the absence of any oxidation signal in line a, while two oxidation peaks are obtained in vitro in presence of nM DA and 5-HT at approx. 8mV and 25mV, respectively (line b, peak 1 and peak 2. In this line h = size of the peak measured in nanoAmperes, nA).

Line c shows the voltammogram obtained when the nafion mCFE is inserted into SN. Note the presence of two in vivo oxidation signals nearly overlapping in vitro peak 1 and peak 2.
Figure 2. TOP: effect of local injection of vehicle (aCSF 0.5µl), or NOCI 0.1nm, 1nm or 10nm, respectively on DA levels and BOTTOM: effect of local injection of vehicle (aCSF 0.5µl), or NOCI 0.1nm, 1nm or 10nm, respectively on 5-HT levels monitored in the SN with DPV and nafion mCFE. N=4 each treatment. Data are presented as % of basal pre-treatment levels ± sem. Stats: 2wANOVA, *p<0.05 versus control (vehicle), Dunnett test.

NOCI 0.1 versus vehicle: F(11,44)=2.8959 p=0.0596
NOCI 1 versus vehicle: F(11,44)=5.5102 p=0.0002
NOCI 10 versus vehicle: F(11,44)=5.6706 p=0.0001
Figure 3. Electrophysiological monitoring performed in the SN with the same nafion mCFE used for parallel DPV measurements as described earlier (Crespi 2002). Briefly, DPV scans last 30 sec. and are repeated every 5 min. Electrophysiology is then performed in the time gap between each DPV scan. In such a way, these two types of in vivo recordings could be performed concurrently in situ, allowing direct comparison of cell firing and monoamine release in the same brain region and in real time. In addition, the influence of pharmacological treatments upon both electrochemical and electrophysiological signals is therefore studied concurrently in real time and in the same animal.

Note here the dose-dependent effect of local injection (arrow) of NOCI 1 nm and 10 nm, while NOCI 0.1 nm is having no effect on the signal as well as aCSF (control).

Data are presented as % of basal pre-treatment levels ± sem, (n=4 each treatment).

Stats are shown in the figure.
rapid decrease of (multi)-cell firing in the SN (figure 3).

Discussion

The present original in vivo data demonstrates that NOCI locally injected in the SN directly affects cell firing as well as DA and 5-HT levels in this brain region. Taken together with the observations from Calo’ et al [13] that NOCI is involved in:

(i) Inhibition of glutamate release/anti-epileptic action and disruption of spatial memory; (ii) Inhibition of serotonin release/anxiolytic action; (iii) Inhibition of mesolimbic dopaminergic transmission/anti-rewarding properties; (iv) Modulation of striatal dopamine and glutamate/effects on locomotor activity, the present data further support the wide role of NOCI in the CNS functions as a potent modulator of neurotransmitter activities. In particular, these data support the direct interaction of NOCI with the dopaminergic and the serotoninergic neurons in the SN.

Indeed, for what concern dopamine, two parallel ex vivo approaches, dual in situ hybridization (ISH) and neurotoxic lesions of DA neurons by using 6-hydroxydopamine (6-OHDA) were applied in the ventral tegmental area (VTA) and substantia nigra pars compacta (SNc) by Norton et al. [14]. This was done in order to verify whether NOCI acts directly on DA neurons i.e. upon NOCI-receptors (NOP), or indirectly by means of local circuitry, or both. It appears that NOCI mRNA was found largely on non-dopaminergic (i.e., GABA) neurons, whereas NOP mRNA was located on DA neurons. These data indicated that NOCI is in a position to influence DA neuronal activity by means of the NOP located on DA neurons. Accordingly, the present in vivo electrophysiological data support previous in vitro electrophysiological recordings observing that NOCI hyperpolarized the dopaminergic cells of the substantia nigra pars compacta and inhibited their firing activity [6].

Furthermore, in vivo microdialysis studies have shown a large increase of dopamine release (in the order of 300% of control values) in striatum when treating conscious rats with nociceptin at the micromolar concentration [15]. This evidence is in accord with the present observation of a large increase of DA levels following local injection of NOCI in SN.

In contrast, intra-cerebroventricular administration of NOCI at a probe concentration of 1 mM but not at 0.1 mM significantly reduced rat nucleus accumbens dialysate DA levels in studies using a dual-probe microdialysis experimental design. Similar data were obtained when NOCI was applied to the ventral tegmental area of anesthetized rats by reverse dialysis while extracellular DA was sampled with a second dialysis probe in the nucleus accumbens [5]. The observation that only the 1mM NOCI concentration but not the 0.1 mM concentration is acting upon DA levels may indicate that the technical approach used by these authors is not the most responsive. Nevertheless, this data is in accord with the present detection of reduced levels of DA in SN when the high dose of NOCI is injected (see figure 2).

It is known that serotonergic fibers from the raphe system project to the Substantia Nigra (SN) [16] and in particular that serotonin released from terminals in SN, derived from cell bodies in the raphe dorsalis nucleus, decreases the activity of the nigro-striatal dopaminergic system [17]. It has been also observed that this modulation is behaviourally relevant [7].

Additionally, the present data showing large decrease of 5-HT levels in SN following local treatment with NOCI and a parallel dose-dependent, rapid decrease of (multi)-cell firing in the SN are in accord with results showing reduction of electrical activity of 5-HT neurons in RDN [18] as well as 5-HT release i.e. in cerebral cortex [19] after NOCI injection. Also, the inhibitory effect of NOCI on the nigral release of 5-HT can be related to the altered motor activity monitored i.e. using the fixed-speed rotarod test [6].

When considering the present data in a whole, it appears that there is a rapid effect of NOCI upon both DA and 5-HT signals in SN. The timing is similar although the effect is the opposite, with a very large increase of DA levels and a parallel large decrease of 5-HT values. However, while DA is rapidly largely significantly affected by NOCI 1nm, 5-HT is modified similarly by NOCI 10nm while is significantly less sensitive to NOCI 1nm (see figure 2). Based on this observation one may propose that the rapid, large effect of NOCI 1nm on DA release may be possibly the driver of the comparatively slower and reduced change of serotonin levels, indicating that is dopamine that may be
primarily influenced by NOCI in the SN.

The interaction between these two aminergic systems has been already reported in such diverse functions as temperature regulation [20], sleep [21], sexual behavior [22] and extrapyramidal function [23] although many results are conflicting [24].

In particular, it has been already reported that electrical stimulation of SN resulted in a rapid increase of the catecholaminergic DPVoltammetric signal followed by a slower decrease of the serotoninergic peak, both recorded simultaneously in the rat striatum [25].

Accordingly, Kuhr and coll. [26] observed a rapid rise of DA levels in the caudate after electrical stimulation in SN. A similar and more consistent effect was observed when stimulating the medial forebrain bundle (MFB) and recording DA and 5-HT activities in the striatum [25, 27].

In conclusion, this work provides further evidence that DPV combined with mCFE is a valuable tool in the study of the in vivo effect of NOCI, an endogenous neuropeptide involved in a number of biological actions [28, 29, 30]. In particular, it proposes a multifaceted implication of this neuropeptide on both dopaminergic and serotoninergic functions in the SN, suggesting a primary influence upon dopamine activity followed by the serotoninergic response. This data may be of importance in the interpretation of the biological functions of NOCI as indeed the NOCI-NOP receptor system is widely represented throughout the CNS [31].

For instance, the influence upon motor behavior may be considered as an implication of both DA and 5-HT in physiological as well as pathological conditions i.e. Parkinson disease (PD) [32].

Furthermore, the involvement of serotonergic mechanisms in the development of Levodopa-induced dyskinesias (LIDs) via aberrant processing of exogenous levodopa and release of dopamine as a false neurotransmitter in PD patients has been recently confirmed [33].

Additionally, the implication of NOCI within the regulation of feeding, body weight homeostasis, stress, the stress-related psychiatric disorders of depression and anxiety, and in drug and alcohol dependence, pathological situations involving both DA and 5-HT, has been described (for a review see [34]).

Therefore, the present data proposing a complex interaction between NOCI, DA and 5-HT systems may be of help in the interpretation of physiological as well as pathological states and consequent development of therapeutical approaches.

Acknowledgment
To M. Manzalini for skillful technical assistance and to Prof. P. Nuthall for english language

References
1. Cox BM, Chavkin C, Christie MJ, Civelli O, Evans C et al.(2000) Opioid Receptors. In: The IUPHAR Compendium of Receptor Characterization and Classification. (Girdlestone D ed): IUPHAR Media Ltd, London.
2. Darland T, Heinricher MM, Grandy DY (1998) Orphanin FQ/nociceptin: a role in pain and analgesia, but so much more. Trends Neurosci., 21, 215-221.
3. Jenck F, Moreau JL, Martin JR, Kilpatrick GJ, Reinscheid RK et al. (1997) Orphanin FQ acts as an anxiolytic to attenuate behavioral responses to stress. Proc. Natl. Acad. Sci. U.S.A., 94, 14854 – 14858.
4. Zheng F, Grandy OK, Johnson SW (2002) Actions of orphanin FQ/nociceptin on rat ventral tegmental area neurons in vitro. Br. J. Pharmacol.. 136, 1065-1071.
5. Murphy NP, Maidment NT (1999) Orphanin FQ/nociceptin modulation of mesolimbic dopamine transmission determined by microdialysis. J. Neurochem. 73, 179-186.
6. Marti M, Mela F, Veronesi C, Guerrini R, Salvadori S et al. (2004) Blockade of Nociceptin/Orphanin FQ Receptor Signaling in Rat Substantia Nigra Pars Reticulata Stimulates Nigrostriatal Dopaminergic Transmission and Motor Behavior. Journal of Neuroscience 24 (30), 6659-6666; DOI: https://doi.org/10.1523/JNEUROSCI.0987-04.2004.
7. Dray A, Davies J, Oakley NR, Tongroach P, Vellucci S (1978) The dorsal and medial raphe projections to the substantia nigra in the rat: electrophysiological, biochemical and behavioural observations. Brain Research 151(3), 431-442.
8. Vaughan CW, Christie MJ (1996) Increase by the ORL1 receptor (opioid receptor-like1) ligand,
nociceptin, of inwardly rectifying K conductance in dorsal raphe nucleus neurones. Br. J. Pharmacol. 117, 1609 – 1611.

9. Werthwein S, Bauer U, Nakazi M, Kathmann M, Schlicker E (1999) Further characterization of the ORL1 receptor-mediated inhibition of noradrenaline release in the mouse brain in vitro. Br. J. Pharmacol. 127, 300 – 308.

10. Crespi F, Martin KF, Marsden CA (1988a) Measurement of extracellular basal levels of serotonin in vivo using nafion-coated carbon fibre electrodes combined with differential pulse voltammetry, Neuroscience 27, 885-96.

11. Crespi F (2002) In vivo voltammetry and concomitant electrophysiology at a single biosensor to analyse ischaemia, depression and drug dependence. Journal of Neuroscience Methods 119, 173-184.

12. Paxinos G, Watson C (1982) The Rat Brain in Stereotaxic Coordinates. Academic, Sydney

13. Calo’ G, Guerrini R, Rizzi A, Salvadori S, Regoli D (2000) Pharmacology of nociceptin and its receptor: a novel therapeutic target. British Journal of Pharmacology 129,1261-1283.

14. Norton CS, Neal CR, Kumar S, Akil H, Watson SJ (2002) Nociceptin/orphanin FQ and opioid receptor-like receptor mRNA expression in dopamine systems. J. Comp. Neurol. 444, 358–368, Wiley-Liss, Inc. https://doi.org/10.1002/cne.10154

15. Konya H, Masuda H, Itoh K, Nagai K, Kakishita E, Matsuoka A (1998) Modification of dopamine release by nociceptin in conscious rat striatum. Brain Research 788 (1–2), 341-344.

16. Dray A, Gonie TJ, Tanner T (1976) Evidence for the existence of a raphe projection to the substantia nigra of the rat. Brain Res. 113, 45-57.

17. Fibiger HC, Miller JJ (1977) An anatomical and electrophysiological investigation on the 5-HT projection from the DRN to the SN in the rat. Neurosci. 2, 975-987.

18. Vaughan CW, Connor M, Jennings EA, Marineih S, Alien RG, Christie MJ (2001) Actions of nociceptin/orphanin FQ and other prepronociceptin products on rat rostral ventromedial medulla neurons in vitro. J. Physiol. 534, 849-859.

19. Mela F, Marti M, Ulazzi L, Vaccai E, Zucchini S et al. (2004) Pharmacological profile of nociceptin/orphanin FQ receptors regulating 5-hydroxytryptamine release in the mouse neocortex. Eur. J. Neurosci. 19, 1-8.

20. Feldberg W, Lotti VJ (1967) Temperature response to monoamines and an inhibitor of MAO injected into the cerebral ventricles of rats. Br. J. Pharmacol. Chemother. 31, 152-168.

21. Jouvet M (1969) Biogenic amines and the state of sleep. Science 163, 32-41.

22. Gessa GL, Tagliamonte A (1974) Possible role of brain serotonin and dopamine in controlling male sexual behaviour. Adv. Biochem. Psychopharmacol. 11, 217-228.

23. Hornykiewicz O (1974) Neurohumoral interactions and basal ganglia function and dysfunction. In M.D. Yahr (Ed.), The Basal Ganglia, Vol. 55, Raven Press, New York, pp. 269-278.

24. Lloyd KG (1978) Neurotransmitter interactions related to central dopamine neurons. In M. Youdim, W. Lovenberg, D. Sharman and J. Lagnado (Eds.), Essays in Neurochemistry and Neuropharmacology Vol. 3, Wiley, New York, pp. 129-208.

25. Crespi F, Martin KF, Marsden CA (1988b) Simultaneous in vivo voltammetric measurement of striatal extracellular DOPAC and 5-HIAA levels: Effect of electrical stimulation of DA and 5HT neuronal pathways. Neuroscience Letters 90, 285-291.

26. Kuhr WG, Ewing AG, Caudill WL, Wightman RM (1984) Monitoring the Stimulated Release of Dopamine with In Vivo Voltammetry. I: Characterization of the Response Observed in the Caudate Nucleus of the Rat j. Neurochem. 43 (2t 1984 560-569

27. Crespi F, Paret J, Keane PE, Morre M (1984) An improved differential pulse voltammetry tech-ique allows the simultaneous analysis of dopaminergic and serotonergic activities in vivo with a single carbon-fibre electrode. Neurosci. Lett. 52, 159-164.

28. Meunier JC, Mollereau C, Toll L, Suaudeau C, Moisand C et al. (1995) Isolation and structure of the endogenous agonist of opioid receptor-like ORL1 receptor. Nature 377, 532-535.

29. Reinscheid RK, Nothacker HP, Bourson A, Andati A,
Henningsen RA et al. (1995) Orphanin FQ: a neuropeptide that activates an opioid-like G protein-coupled receptor. Science 270, 792-794.

30. Mogil JS, Pasternak GW (2001) The molecular and behavioral pharmacology of the orphanin FQ/Nociceptin peptide and receptor family. Pharmacol. Rev. 53, 381-415.

31. Slowe SJ, Clarke S, Lena I, Goody RJ, Lattanzi R et al. (2001) Autoradiographic mapping of the opioid receptor-like 1 (ORL1) receptor in the brains of mu-, delta- or kappa-opioid receptor knockout mice. Neuroscience 106, 469-80.

32. Politis M, Oertel WH, Wu K, Quinn NP, Pogarell O et al. (2011) Graft-induced dyskinesias in Parkinson’s disease: High striatal serotonin/dopamine transporter ratio. Movement Disorders 26 (11), 1997-2003.

33. Politis M, Wu K, Loane C, Brooks DJ, Kiferle L et al. (2014) Serotonergic mechanisms responsible for levodopa-induced dyskinesias in Parkinson’s disease patients. J Clin Invest. 124(3), 1340-1349. https://doi.org/10.1172/JCI71640.

34. Witkin JM, Statnick MA, Rorick-Kehn LM, Pintar JE, Ansonoff M et al. (2014) The biology of Nociceptin/Orphanin FQ (N/OFQ) related to obesity, stress, anxiety, mood, and drug dependence Pharmacology & Therapeutics 143 (3), pages 351 https://doi.org/10.1016/j.pharmthera.2013.10.011