In vitro inhibitory and biofilm disruptive activities of ginger oil against Enterococcus faecalis [version 1; peer review: 1 approved with reservations, 1 not approved]

Shahida Mohd-Said1, Wee Wee Kweh2, Chong Yi Than2, Zamirah Zainal-Abidin3, Siti Noor Adnalizawati Adnan4, Safura Anita Baharin1, Eason Soo1

1Centre for Restorative Dentistry, Faculty of Dentistry, Universiti Kebangsaan Malaysia, Jalan Raja Muda Abdul Aziz, Kuala Lumpur, 50300, Malaysia
2Oral Health Division, Ministry of Health Malaysia, Muar, Johor, 84000, Malaysia
3Centre for Craniofacial Diagnostics and Biosciences, Faculty of Dentistry, Universiti Kebangsaan Malaysia, Jalan Raja Muda Abdul Aziz, Kuala Lumpur, 50300, Malaysia
4Faculty of Dentistry, Universiti Sains Islam Malaysia, Ampang, Kuala Lumpur, 55100, Malaysia

Abstract

Background: This study investigated the antibacterial effect of ginger (Zingiber officinale) oil against a common resistant root canal pathogen known as Enterococcus faecalis. The aim of the study was to determine the inhibition of E. faecalis growth in culture suspension and ability to inhibit growth of bacteria through disruption of pre-formed monospecies biofilm.

Methods: Ginger rhizome oil was prepared in two-fold concentration series from 0.04 to 5.00 mg/mL and mixed with brain heart infusion broth inoculated with E. faecalis in anaerobic condition. Among the antibacterial tests performed were the minimum inhibitory concentrations (MIC) and minimum bactericidal concentrations using microdilution assays, and anti-biofilm assay on 3-day old pre-form monospecies biofilm on a 94 well-plate. Ampicillin was used as a positive control.

Results: The result showed an in vitro dose-dependent bacteriostatic activity towards E. faecalis in suspension broth (MIC 0.04mg/mL) but no bactericidal activity within the tested concentration range. It was also found that the ginger oil inhibitory activity against E. faecalis was comparably less in anti-biofilm activity than against bacteria cultured in suspension solution.

Conclusion: The study suggests that at determined concentrations, ginger oil has the potential to be used as an antibacterial agent in the management of root canal infections particularly where newly formed E. faecalis is involved.

Open Peer Review

Reviewer Status

| Invited Reviewers | 1 | 2 |
|-------------------|---|---|
| version 1 | report | report |

1. Lara Thieme, Jena University Hospital, Jena, Germany
Mathias W. Pletz, Jena University Hospital, Jena, Germany

2. Fathilah Binti Abdul Razak, University of Malaya, Kuala Lumpur, Malaysia

Any reports and responses or comments on the article can be found at the end of the article.
Keywords
Enterococcus, biofilm, endodontics, Zingiber officinale

Corresponding author: Shahida Mohd-Said (shahidams@ukm.edu.my)

Author roles: Mohd-Said S: Conceptualization, Data Curation, Formal Analysis, Investigation, Methodology, Project Administration, Resources, Supervision, Validation, Writing – Original Draft Preparation, Writing – Review & Editing; Kweh WW: Formal Analysis, Investigation, Visualization, Writing – Original Draft Preparation; Than CY: Formal Analysis, Investigation, Visualization, Writing – Original Draft Preparation; Zainal-Abidin Z: Conceptualization, Formal Analysis, Methodology, Validation, Writing – Review & Editing; Adnan SNA: Validation, Writing – Review & Editing; Baharin SA: Conceptualization, Validation, Writing – Review & Editing; Soo E: Conceptualization, Funding Acquisition, Project Administration, Resources, Supervision, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: This study was funded by the Universiti Kebangsaan Malaysia Young Researchers’ Incentive Grant (GGPM-2013-096).

Copyright: © 2018 Mohd-Said S et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. Data associated with the article are available under the terms of the Creative Commons Zero "No rights reserved" data waiver (CC0 1.0 Public domain dedication).

How to cite this article: Mohd-Said S, Kweh WW, Than CY et al. In vitro inhibitory and biofilm disruptive activities of ginger oil against Enterococcus faecalis [version 1; peer review: 1 approved with reservations, 1 not approved] F1000Research 2018, 7:1859 https://doi.org/10.12688/f1000research.16851.1

First published: 28 Nov 2018, 7:1859 https://doi.org/10.12688/f1000research.16851.1
Introduction

Enterococcus faecalis is an opportunistic facultative anaerobe that is well recognised as an oral pathogen associated with persistent apical periodontitis and is highly prevalent in failed root filled teeth. The ability of E. faecalis to enter and survive in root canals, as well as its resistance to root canal medicaments, makes it one of the toughest pathogens to eradicate in endodontics. It has been reported that this pathogen is associated with a high percentage of endodontic failures, which is about one third of the canals of root-filled teeth with persistent periapical lesions. Inevitably, repeated use of calcium hydroxide and sodium hypochlorite as the two most commonly used root canal medicaments and irrigation solution, respectively, has been said to allow E. faecalis to adapt to the sub-lethal environment.

Within the past decade, herbal medicine has begun to offer some beneficial antibacterial activities against oral pathogens. More recently, studies also found the potential antibacterial properties of ginger on oral pathogens, including E. faecalis. In most instances, the effects of ginger extracts were studied on bacteria cultured in vitro and in cell suspensions, using calcium hydroxide or sodium hypochlorite as a control. Limited data is available on the activity of ginger oil in disrupting established E. faecalis biofilm and its comparable effect to antibiotics. Hence, the aim of our study was to explore the in vitro potential of ginger oil as an antibacterial agent against E. faecalis cultured in suspension and biofilm in comparison to ampicillin, as a common antibiotic used for oral infection.

Methods

Extraction and preparation of oil

Ginger oil was prepared from Zingiber officinale Roscoe rhizomes after fresh young ginger were finely sliced, dried and boiled in distilled water for 8h. The oil stock solution (500 mg/mL) was prepared by mixing 100 µL ginger oil with 200 µL dimethyl sulfoxide (DMSO; Sigma-Aldrich, St Louis, MO USA) in an Eppendorf tube. Next, 100 µL of the oil stock solution was pipetted and mixed with 100 µL DMSO in a second tube to produce two-fold dilution at 250 mg/mL. This procedure was repeated six times to produce an eight concentration series of ginger oil solution at 500, 250, 125, 62.5, 31.3, 15.6, 7.8, 3.9 mg/mL respectively. The solutions were vortexed each time after mixing to ensure a thorough mixture were produced. Following this, 20 µL of each oil series were transferred into a 980 µL brain heart infusion (BHI; Oxoid Ltd., Cheshire UK) broth in 2.5 mL universal bottles to produce eight working concentration series of ginger oil mixtures in broth microdilution test at 10, 5, 2.5, 1.25, 0.63, 0.31, 0.16 and 0.08 mg/mL respectively. The positive control ampicillin solution was prepared by mixing 500 mg ampicillin powder (Sigma-Aldrich) with 1 mL distilled water, and then eight series of two-fold solutions were prepared similarly to the technique mentioned above.

Culture and maintenance of bacteria

Enterococcus faecalis ATCC29212 (American Type Culture Collection, Virginia USA) used in this study was cultured in BHI (Oxoid Ltd., Cheshire UK) agar plates and regularly maintained in a 37°C incubator with 5% CO₂. Identification and purification of E. faecalis were done through Gram stain test as well as morphology and colony identification. The quantity and viable bacterial number of E. faecalis were determined by colony forming unit (CFU) and standardized with McFarland 0.5 turbidity (1.5 x 10⁸ cfu/mL) prior to the antibacterial assays. Cultures and broths were constantly checked for sterility and contamination. DMSO was used as solvent for ginger oil and ampicillin as the positive control.

Research and ethics approval was obtained from the Universiti Kebangsaan Malaysia Research and Ethics Committee (UKM 1.5.3.5/244/DD/2014/017) for methodology and dissemination of findings.

Broth microdilution assay

An antibacterial assay using the broth microdilution technique was done based on modified NCCLS guideline. A 96-well test plate was divided into two sections where half was inoculated with 50 µL E. faecalis suspension in each well and another section was not inoculated (50 µL broth/well only), as shown in Figure 1, plate rows from A to H. Sample solution series prepared earlier were mixed with the bacterial suspensions in wells and produce a final concentration series of oil mixtures at final concentrations of 5, 2.5, 1.25, 0.63, 0.31, 0.16, 0.08 and 0.04 mg/mL respectively. One hundred microliters sample series (ginger oil mixture or ampicillin solution) were dispensed into the prepared 96-well plate. Triplicates were done for each well and test plate. Plates were then incubated at 37°C for 24h. After this, the turbidity of wells on tested plates was measured at 595nm optical density using a microtiter plate reader (Thermo Scientific VARIOSKAN Flash, UK). Bacterial growth inhibition percentage was calculated using the following formula:

\[
\% \text{ Bacterial growth inhibition} = \left( \frac{\text{mean OD}_{595} (\text{control test})}{\text{mean OD}_{595} (\text{control})} \right) \times 100\%
\]

The reading generated by the device were analysed and the Minimum Inhibitory Concentration (MIC), defined as the lowest concentration of sample that inhibits visible growth of a microorganism after overnight incubation, was determined. Further to this, 20 µL of the mixture from each well that showed bacterial growth inhibition were cultured on BHI agar plates for 24h at 37°C in order to determine whether the inhibition is bacteriostatic or bactericidal in nature. The minimum bactericidal concentration (MBC), defined as the lowest concentration of samples that prevent the total growth of E. faecalis on test plates, was determined.

Anti-biofilm assay

A monospecies biofilm was initially produced by inoculating 200 µL E. faecalis suspension culture at 1.5 x 10⁶ cfu/mL cell density with BHI broth in 96-well plates (modified from Azizan et al). The culture was checked daily for contamination for 4 days and cell density was measured at 595 nm optical density using a microtiter plate reader, and repeated in three separate tests. The findings from this monospecies biofilm development test was charted (Figure 2) and used to determine the age for a stable pre-formed biofilm for the anti-biofilm assay.
In the anti-biofilm assay, *E. faecalis* biofilm was developed and maintained for 3 days. On the third day, culture broth in the plate was pipetted out in slanting position to prevent disruption of biofilm and replaced with 100 µL of fresh broth. Then 100 µL ginger oil or ampicillin solutions at eight respective concentrations were dispensed into the wells and plates were incubated at 37°C for 24h. The turbidity of the bacterial cultures was measured using a microtiter plate reader at 595 nm optical density.

**Statistical analysis**
Each assay was conducted in triplicate and two independent experiments were done. The data collected were analysed with SPSS 21.0. Non-parametric Mann-Whitney test were used.
with significant level set as 0.05. Results were accepted as statistically significant if the p value was <0.05.

**Results**
The broth microdilution antibacterial assay showed a dose-dependent inhibition of *E. faecalis* growth when exposed to all tested concentration series of ginger oil solution (Figure 3), but no bactericidal effect of the oil was found within the range tested (0.04 – 5.0 mg/mL). While ampicillin showed almost 80% inhibition for all concentrations tested, ginger oil showed about 76% inhibition at 5.0 mg/mL and 50% and less at 2.50 – 0.04 mg/mL. The difference in means of inhibition between all ginger oil concentrations tested and between ginger oil and ampicillin was found to be statistically significant (p=0.002 and p=0.007, respectively).

In the anti-biofilm test, ginger oil also inhibited the growth of pre-formed biofilm (Figure 4). However, it was found that both ginger oil and ampicillin showed lesser inhibitory effect on pre-formed biofilm than on suspended cells in broth (Figure 3).

![Figure 3. Antibacterial effects of ginger oil and ampicillin against *Enterococcus faecalis* in suspension culture. * significant difference (p<0.05) between ginger oil and ampicillin at 5.00mg/mL.](image3)

![Figure 4. Anti-biofilm test of ginger oil and ampicillin against 3-day *Enterococcus faecalis* biofilm.](image4)
In this test, the anti-biofilm activity of ginger oil was observed as an increase between 1.25 to 5 mg/mL, yet the activity failed to achieve an acceptable level of inhibition of more than 50%. At 1.25 mg/mL, ginger oil was seen to have somewhat equal biofilm disruptive property as ampicillin, but the means of anti-biofilm activity between the two agents showed no statistically significant difference (p=0.052).

Discussion

It has been advocated that ginger, as the stem of Z. officinale plant, has strong antimicrobial properties including antibacterial and antifungal against pathogens, including bacteria from the oral cavity. In the present study, it was further discovered that ginger rhizome oil exhibited bacteriostatic activity on E. faecalis cultured in plates as monospecies suspension and causes disruption of the pre-formed biofilm. The dose-dependent effect of the oil was accepted as a positive outcome of the study and we predict that with higher concentration (more than 5 mg/mL), ginger oil would have bactericidal effect on E. faecalis.

The ability of ginger oil to disrupt 3-day old pre-formed monospecies E. faecalis biofilm provided us with a new insight on the antibacterial property of this herbal oil. Although the activity was not as potent as the effect on suspension culture, ginger oil still produced acceptable inhibitory activities against the pathogen in vitro. This study provides clearer appreciation of the E. faecalis resistance to antibacterial agents when they are in biofilm form and conforms to other evidence of biofilm resistance.

The use of systemic antibiotics such as ampicillin in acute endodontic infections has raised many concerns over the increase in the prevalence of bacterial resistance in dentistry. On the other hand, the current use of antibacterial irrigation solutions such as sodium hypochlorite, chlorhexidine and iodine as well as calcium hydroxide as canal medicament reduces the needs for systemic antibiotics. However, some degree of toxicity towards vital tissues and allergy reactions to some patients have been reported. Through our study, we found that ginger oil has lower but comparable antibacterial efficacy against E. faecalis compared to ampicillin. Further studies to investigate the effects of combined oil-antibiotics may perhaps provide some useful information on the range of its antibacterial activities. Studies have shown that combination of herbal oils with antibiotics does produce adequate synergistic and additive effects against microorganisms.

Lastly, the results of this study demonstrated that at determined concentrations, ginger oil has the potential to be used as an antibacterial agent in the management of failed root canal therapy. Our recent study on the effect of ginger oil on tooth dentine microhardness also provide promising usage of this herbal oil in future endodontics as it offers comparable changes to dentine structure when used as irrigation solution in contrast to sodium hypochlorite and ethylenediaminetetraacetic acid. These two findings may help to access infection deep within the root canal system. Nevertheless, the current clinical scenario does not offer immediate and accurate chairside information on the type of bacterial species involved in specific endodontic infection; hence this finding may not be readily useful for purpose as yet. More studies will be required to evaluate the clinical efficacy of the delivery of ginger oil as an antibacterial agent for endodontic infections.

Conclusion

Within the concentration tested (0.04 – 5.0 mg/mL), ginger oil showed an acceptable dose-dependent in vitro inhibitory activity against E. faecalis (MIC 0.04mg/mL), while its antibacterial activity towards bacteria in suspension broth was comparatively better than its anti-biofilm activity.

Data availability

F1000Research: Dataset 1. File containing raw data for biofilm development test, anti-biofilm assay and suspension assay.

Grant information

This study was funded by the Universiti Kebangsaan Malaysia Young Researchers’ Incentive Grant (GGPM-2013-096).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgements

Authors wished to thank Miss Nurulnua Kamal Baharin of Faculty of Dentistry UKM Microbiology Lab, for her assistance in this study.

References

1. Portenier I, Waltimo TMT, Haapasalo M: Enterococcus faecalis- the root canal survivor and ‘star’ in post-treatment disease. Endod Topics. 2003; 6(1): 136–159. Publisher Full Text
2. Chivatxaranukul P, Dashper SG, Messer HH: Dentinal tubule invasion and adherence by Enterococcus faecalis. Int Endod J. 2008; 41(10): 873–882. PubMed Abstract | Publisher Full Text
3. Duggan JM, Sedgley CM: Biofilm formation of oral and endodontic Enterococcus faecalis. J Endod. 2007; 33(7): 815–818. PubMed Abstract | Publisher Full Text
4. Gajan EB, Aghazadeh M, Abasov R, et al.: Microbial Flora of Root Canals of Pulpathy-infected Teeth: Enterococcus faecalis a Prevalent Species. J Dent Res Dent Clin Dent Prospects. 2009; 3(1): 24–27. 
PubMed Abstract | Free Full Text

5. Liu H, Wei X, Ling J, et al.: Cytotoxicity and anti-biofilm formation capability of Enterococcus faecalis cells in starvation phase and its susceptibility to sodium hypochlorite. J Endod. 2010; 36(4): 630–635. 
PubMed Abstract | Publisher Full Text

6. Mohammad Z, Palazzo F, Giardino L, et al.: Microbial biofilms in endodontic infections: an update review. Biomed J. 2013; 36(2): 59–70. 
PubMed Abstract | Publisher Full Text

7. Svensater G, Bergenholtz G: 

8. Roth KA, Friedman S, Lévesque CM, et al.: Microbial biofilm proliferation within sealer-root dentin interfaces is affected by sealer type and aging period. J Endod. 2012; 38(9): 1253–1256. 
PubMed Abstract | Publisher Full Text | Free Full Text

9. Evans M, Davies JK, Sundqvist G, et al.: Mechanisms involved in the resistance of Enterococcus faecalis to calcium hydroxide. Int Endod J. 2002; 35(3): 221–228. 
PubMed Abstract | Publisher Full Text | Free Full Text

10. Maekawa LE, Valera MC, Oliveira LD, et al.: Effect of Zingiber officinale and propolis on microorganisms and endotoxins in root canals. J Appl Oral Sci. 2013; 21(1): 25–31. 
PubMed Abstract | Publisher Full Text | Free Full Text

11. Panpati VV, Tattan S, Kota N, et al.: In vitro evaluation on antioxidant and antimicrobial activity of spice extracts of ginger, turmeric and garlic. J Pharmacogn Phytochem. 2013; 2(3): 143–148. 
Reference Source

12. Ainiati BA, Diab AH, Essa SA, et al.: Antibacterial Activity of the Ethanolic Extracts of Allium sativum L., Bulbus and Zinger officinalis Roscoie Rhizomes as Irrigating Solutions. World J Pharm Sci. 2014; 3(6): 324–337. 
Reference Source

13. Azizi A, Aghayan S, Zaker S, et al.: In Vitro Effect of Zinger officinalis Extract on Growth of Enterococcus faecalis and Streptococcus gansuensis. Int J Dent. 2015; 489842. 
PubMed Abstract | Publisher Full Text | Free Full Text

14. Giriraju A, Yurus G: Assessment of antimicrobial potential of 10% ginger extract against Streptococcus mutans, Candida albicans, and Enterococcus faecalis: an in vitro study. Indian J Dent Res. 2013; 24(4): 397–400. 
PubMed Abstract | Publisher Full Text

15. Shabahsia S, Mughimipour M, Shamszadeh N, et al.: Evaluation of antibacterial effect of Eucalyptus and ginger on dentinal tubules Enterococcus faecalis in the presence of smear layer. Bioscience Biotechnology Research Communications. 2017; 19(2): 225–232. 
Publisher Full Text

16. Valera MC, Oliveira SA, Maekawa LE, et al.: Action of Chlorhexidine, Zinger officinalis, and Calcium Hydroxide on Candida albicans, Enterococcus faecalis, Escherichia coli, and Endotoxin in the Root Canals. J Contemp Dent Pract. 2016; 17(2): 114–118. 
PubMed Abstract | Publisher Full Text

17. Zainal-Abidin Z, Abdul-Wahab NA, Ghazi-Ahmad MK, et al.: In vitro Antibacterial Activity of Zinger officinalis and Orthosiphon stamineus on Enterococcus faecalis. J Agri Sci. 2017; 9(13): 112–121. 
Publisher Full Text

18. Clinical and Laboratory Standards Institute: M11-A8 Methods for Antimicrobial Susceptibility Testing of Anaerobic Bacteria: Approved Standard - Eight Edition. 2012. 
Reference Source

19. Azizzan N, Mohd Said S, Zainal Abidin Z, et al.: Composition and Antibacterial Activity of the Essential Oils of Orthosiphon stamineus Benth and Ficus deltoidea Jack against Pathogenic Oral Bacteria. Molecules. 2017; 22(12): pi: E2135. 
PubMed Abstract | Publisher Full Text

20. Helal MM, Osman MY, Gholabash MOI, et al.: Study of some biological activities of aqueous extract of ginger (Zinger officinalis). Egypt Pharmaceut J 2014; 13(2): 144–150. 
Publisher Full Text

21. Bhaskar A, Velumungan N, Sumitha, et al.: Efficacy of passive ultrasonic irrigation with natural irritants (Morinda citrifolia juice, Aloe Vera and Propolis) in comparison with 1% sodium hypochlorite for removal of E. faecalis biofilm: An in vitro study, Indian J Dent Res. 2013; 24(1): 35–41. 
PUBMED Abstract | PUBisher Full Text

22. Jantan IB, Yassin MS, Chin CB, et al.: Anti fungal Activity of the Essential Oils of Nine Zingiberaceae Species. Pharm Bioll. 2003; 41(5): 392–397. 
PUBisher Full Text

23. Mueller M, Hobiger S, Jungbauer A: Anti-inflammatory activity of extracts from fruits, herbs and spices. Food Chem. 2010; 122(4): 987–996. 
PUBisher Full Text

24. Patel RV, Thakar VT, Patel VK: Antimicrobial activity of ginger and honey on isolates of extracted carious teeth during orthodontic treatment. Asian Pac J Trop Biomed. 2011; 11(1): 558–561. 
PUBisher Full Text

25. Sasidhran I, Nirmala Menon A: Comparative Chemical Composition and Antimicrobial Activity Fresh & Dry Ginger Oils (Zinger officinalis Roscoie). Int J Curr Pharm Res. 2010; 2(4): 4–7. 
Reference Source

26. Sebiso A, Awofodu AD, Awosanya AO, et al.: Comparative studies of antibacterial effect of some antibiotics and ginger (Zinger officinalis) on two pathogenic bacteria. J Microbial Antimicrob. 2011; 3(1): 18–22. 
Reference Source

27. Sivasothy Y, Chong WK, Hamid A, et al.: Essential Oils of Zinger officinalis var. rubrum Theilade and their Antibacterial Activities. Food Chem. 2011; 124(2): 514–517. 
PUBisher Full Text

28. Davies D: Understanding biofilm resistance to antibacterial agents. Nat Rev Drug Discov. 2003; 2(10): 114–122. 
PUBMED Abstract | PUBisher Full Text

29. Gupta A, Duhan J, Tewari S, et al.: Comparative evaluation of antimicrobial efficacy of Syzygium aromaticum, Ocimum sanctum and Cinnamomum zeylanicum plant extracts against Enterococcus faecalis: A preliminary study. Int Endod J. 2013; 46(8): 775–783. 
PUBMED Abstract | PUBisher Full Text

30. Neelakantan P, Subbarao C, Sharma S, et al.: Effectiveness of curcumigen against enterococcus faecalis biofilm. Acta Odontol Scand. 2013; 71(6): 1453–1457. 
PUBMED Abstract | PUBisher Full Text

31. Guivrach M, Orioni U, Ahmed HM, et al.: Sodium Hypochlorite Accident: A Systematic Review. J Endod. 2017; 43(1): 16–24. 
PUBMED Abstract | PUBisher Full Text

32. Levy SB, Marshall B: Antibacterial resistance worldwide: causes, challenges and responses. Nat Med. 2004; 10(12 Suppl): S122–S129. 
PUBMED Abstract | PUBisher Full Text

33. Matsura M, Nakazawa H, Hashimoto T, et al.: Combined antibacterial activity of amoxicillin with clavulanic acid against ampicillin-resistant strains. Antimicrob Agents Chemother. 1980; 17(6): 908–911. 
PUBMED Abstract | PUBisher Full Text

34. Williams EM: Synergy and other interactions in phytomedicines. Phytotherapy. 2001; 8(5): 401–409. 
PUBMED Abstract | PUBisher Full Text

35. Yost RA, Bergeron BE, Kirkpatrick TC, et al.: Evaluation of 4 Different Irrigating Systems for Apical Extrusion of Sodium Hypochlorite. J Endod. 2015; 41(9): 1530–1534. 
PUBMED Abstract | PUBisher Full Text

36. Zandi H, Rodrigues RC, Kristoffersen AK, et al.: Antibacterial Effectiveness of 2 Root Canal Irritants in Root-filled Teeth with Infection: A Randomized Clinical Trial. J Endod. 2016; 42(9): 1307–1313. 
PUBMED Abstract | PUBisher Full Text

37. Zainol SN, Mohd Said S, Zainal Abidin Z, et al.: Synergistic benefit of eugenia caryophyllata L. and cinnamonum zeylanicum blume essential oils against oral pathogenic bacteria. Chem Eng Trans. 2017; 56: 1429–1434. 
PUBisher Full Text

38. Mohd Zakia SK, Baharin SA, Qamanz Zaman J, et al.: The Effect of Zinger officinalis Roscoie (Ginger) on Dentin Microhardness: An in vitro Study. J Agric Sci. 2017; 9(13): 102–111. 
PUBisher Full Text

39. Mohd-Said S, Kweh WW, Than CY, et al.: Dataset 1: In vitro inhibitory and biofilm disruptive activities of ginger oil against Enterococcus faecalis. F1000Research. 2018. http://www.doi.org/10.5256/f1000research.16851.d225122
Fathilah Binti Abdul Razak
Department of Oral & Craniofacial Sciences, Faculty of Dentistry, University of Malaya, Kuala Lumpur, Malaysia

The study aimed to assess the antibacterial and antibiofilm properties of ginger oil on *E. faecalis* which is a pathogen often isolated from tooth root canals. *E. faecalis* used were in the planktonic (cell suspension) and monospecies biofilm forms. Tests performed include MIC, MBC and an antibiofilm assay.

Comments:

1. Introduction: Adequate

2. Methods:
   (i) OD readings from ELISA plate reader were used as a measurement of cell density in both the planktonic and biofilm state. This procedure should be revised as in the former, the cells are free in the suspension while in the latter, the cells are sedimented/attached to the bottom base, which blocks vertical light source from the bottom and may contribute to higher OD readings. A different set of readings would be obtained if using a spectrophotometer with horizontal light source.
   (ii) An inoculation of 20ul of the broth mixture on agar plates would not be able to indicate a bacteriostatic effect of a test agent, it can only confirm the bactericidal activity of a test agent i.e. when there is no colony growth.
   (iii) How was a stable 3-day preformed biofilm defined, and how can Fig. 2 explain this?
   (iv) The 3-day biofilm could have resulted from sedimentation since the biofilm was left static for 3 days. It is thus suggested that the biofilm is gently washed first before adding the test agent. The use of a shaking incubator is also suggested.

3. Results:
   (i) What were the MIC and MBC for ginger and ampicillin towards *E. faecalis*?
   (ii) Figure 3 does not really show a dose-dependent antibacterial response of *E. faecalis* to ginger oil, or to ampicillin. This would be obvious if error bars were shown in the figures.
(iii) It is not accurate to claim that the anti-biofilm activity of ginger oil was observed to increase between 1.25 to 5 mg/ml. Referring to Fig. 4 the increase were insignificant, and again this would be obvious if the error bars were shown.

4. Discussion:
Cross referencing to more relevant journals/works to defend and justify the study outcome would help improve the discussion.

**Overall comment:**
The manuscript was poorly prepared and certain details such as error bars in the figures were missing. Many steps were over simplified. Analysis and presentation of results can be further improved to give meaningful outcome. I agree with Reviewer 1 that the conclusion was rather over-interpreted, and very weak considering only one species of bacterium was used. More experiments are required to come to such conclusion.

**Is the work clearly and accurately presented and does it cite the current literature?**
Partly

**Is the study design appropriate and is the work technically sound?**
No

**Are sufficient details of methods and analysis provided to allow replication by others?**
Partly

**If applicable, is the statistical analysis and its interpretation appropriate?**
Yes

**Are all the source data underlying the results available to ensure full reproducibility?**
Partly

**Are the conclusions drawn adequately supported by the results?**
No

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Oral environment, Oral biofilm, Antimicrobial agent

I confirm that I have read this submission and believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

Reviewer Report 16 January 2019

https://doi.org/10.5256/f1000research.18421.r42492
In this study, the authors assessed the inhibitory activities of ginger oil in comparison to ampicillin against *Enterococcus faecalis*, both against planktonic and biofilm cultures, with the clinical background of analyzing novel treatment ideas for root canal infections caused by *E. faecalis*. Ginger oil has been shown to exhibit antibacterial effects against *E. faecalis* in some *in vitro* studies, but its anti-biofilm activity has not been investigated yet. Although the authors came up with an in principle adequate experimental set-up, the main criticism concerns the fact that only one laboratory strain has been analyzed (ATCC 29212), but no clinical isolates recovered from root canal infections. Clinical isolates usually show a different behavior than laboratory strains concerning antimicrobial susceptibility patterns, making it absolutely necessary to include an adequate number of clinical isolates when drawing conclusion for therapeutic treatments of infections. Further, it remains questionable whether the analyzed concentrations of ginger oil apply as therapeutic dosages. The lowest concentration (range 40 mg/L to 5000 mg/L) analyzed for both ginger oil and ampicillin, the reference substance in this study, is still 40 times higher than the MIC of ampicillin of 1 mg/L for *E. faecalis* ATCC 29212. Unfortunately, the conclusion drawn from the presented results is in our opinion over-interpreted (mainly due to n=1) and not conclusive from the data presented. As the authors state in the result part, the biofilm inhibitory activity of ginger oil was not strong enough to make it a significant result (quote “activity failed to achieve an acceptable level of inhibition of more than 50%”), contradicting very much the conclusion of the discussion part (quote “Although the activity was not as potent as the effect on suspension culture, ginger oil still produced acceptable inhibitory activities against the pathogen in vitro”). Ampicillin, a common antibiotic for oral infections and the reference substance, showed stronger effects than ginger oil at all concentrations tested in both of the planktonic and biofilm experiments! Regarding the language, only the sentences in which the meaning was not clear were marked and suggestions for improvement were made. Typos, wrong choices of words and sentence constructions were neglected.

Introduction: approved
Methods: approved with reservations/not approved due to n=1
Results: approved
Discussion: approved

Major comments:
1. Methods: Please include at least three root canal *E. faecalis* isolates in your study. Further, please state some data/references about therapeutic doses of ginger oil to better understand the suitability of the tested concentrations for therapeutic treatment.
2. Method anti-biofilm assay: We are struggling with your method of choice (OD
measurement) for the establishment of the biofilm growth. Firstly, a measurement of the OD value rather assesses the planktonic but not the biofilm state of the bacteria and further does not discriminate between viable and dead cells. We recommend performing a CFU determination of the detached biofilms at day 1, 2, 3 and 4 of biofilm growth to determine the viable cell number. This will tell you how the OD measurement correlates with cell numbers. Also, you might want to consider changing the medium every 24hr to enhance biofilm formation. Secondly, we do not understand why you have chosen a 3-day old biofilm for your subsequent anti-biofilm assay as the highest OD value is observed on day 1 (see Figure 2). Please explain your choice. For the anti-biofilm assay, please also verify for at least one strain that lower OD values (compared to the control) correlate with lower cell numbers and therefore stronger inhibitory effects of ginger oil.

3. Results Bactericidal effects (MBC): If you plated the samples on agar plates to determine the MBC as described in the method section, it would be interesting to see the CFU numbers to see the correlation between OD values and viable bacteria (see above)!

4. Discussion: Disruption is the wrong word because you were analyzing biofilm growth inhibitory, but not biofilm eradicative effects! Also, “we found that ginger oil has lower but comparable antibacterial efficacy compared to ampicillin” is a very contradicting statement and in terms of your results wrong. Ampicillin showed stronger effects than ginger oil at all concentrations tested in both of the planktonic and biofilm experiments!

5. Conclusion: Why do you state 0.04 mg/mL as the MIC of ginger oil for ATCC 29212? At this concentration, only approximately 5% growth inhibition was observed (Fig. 3).

Minor comments:
1. Abstract Methods: You state that *E. faecalis* was grown in anaerobic conditions but this is not described in the method part of the main paper. For MIC testing according to CLSI guidelines, enterococci are grown aerobically (so at 37°C and 5 % CO₂ as you stated correctly in the method section).

2. Abstract methods: Please consider revision of the sentence: “Among the antibacterial tests performed were the minimum inhibitory concentrations (MIC) and minimum bactericidal concentrations using microdilution assays, and anti-biofilm assay on 3-day old pre-form monospecies biofilm on a 96 well-plate.” E.g. “Among the antibacterial tests performed were the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) microdilution assay as well as an anti-biofilm assay on 3-day old pre-formed biofilms on 96 well-plates.”

Further, it would be helpful for the reader to clarify which parameter was assessed in your anti-biofilm assay. If I understood your experiments correctly, you assessed the so-called minimum biofilm inhibitory concentration (MBIC) in contrast to biofilm eradicative effects (aka MBEC).

3. Abstract results: Please consider revision of the sentence. “It was also found that the ginger oil inhibitory activity against *E. faecalis* was comparably less in anti-biofilm activity than against bacteria cultured in suspension solution.” E.g. “It was also found that the inhibitory activity of ginger oil was lower in biofilm cultures compared to planktonic cultures of *E. faecalis*.”

4. Introduction: Your introduction is precise and on point. Maybe you could add 1-2 sentences about the definition/meaning and clinical problem of biofilm formation.

5. Introduction: First paragraph, last sentence: “Inevitably, repeated use of calcium hydroxide and sodium hypochlorite as the two most commonly used root canal medicaments and irrigation solution, respectively, has been said to allow *E. faecalis* to adapt to the sub-lethal
environment”. Please re-write this sentence as I do not get the meaning. Do you mean E. faecalis is tolerant to sodium hypochloride and calcium hydroxide treatment? Also, please change “Within the past decade, research has begun to explore some beneficial antibacterial activities of herbal medicine against oral pathogens.”

6. Methods: Please state the source of the ginger root as the quality and therefore the reproducibility of the experiments might vary depending on the growth conditions.

7. Method, part culture and maintenance of bacteria: The last sentence (DMSO was used as solvent for ginger oil...) is redundant and does not fit in this paragraph.

8. Method, part broth microdilution assay: This sentence is rather confusing because the two sections you describe are not visible in Figure 1. Please clarify your pipetting scheme.

9. Results: Please make the captions below the figures more informative by including more details about how the results presented in the figure were created (e.g. Figure 2: which strain is shown, how were the biofilms grown, ...).

References
1. EUCAST: EUCAST Quality Control. [accessed 15th Jan 2019]. Reference Source
2. Thieme L, Klinger-Strobel M, Hartung A, Stein C, et al.: In vitro synergism and anti-biofilm activity of ampicillin, gentamicin, ceftaroline and ceftriaxone against Enterococcus faecalis. J Antimicrob Chemother. 2018; 73 (6): 1553-1561 PubMed Abstract | Publisher Full Text
3. Macià MD, Rojo-Molinero E, Oliver A: Antimicrobial susceptibility testing in biofilm-growing bacteria. Clin Microbiol Infect. 2014; 20 (10): 981-90 PubMed Abstract | Publisher Full Text

Is the work clearly and accurately presented and does it cite the current literature? Yes

Is the study design appropriate and is the work technically sound? No

Are sufficient details of methods and analysis provided to allow replication by others? Yes

If applicable, is the statistical analysis and its interpretation appropriate? Partly

Are all the source data underlying the results available to ensure full reproducibility? Yes

Are the conclusions drawn adequately supported by the results? No

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: infectious diseases, clinical microbiology, antibiotic resistant bacteria, biofilms, enterococci,
We confirm that we have read this submission and believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however we have significant reservations, as outlined above.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com