Background

Despite complete resection of gastrointestinal stromal tumors (GIST), recurrent and/or metastatic disease occurs, often depending on the grade of malignancy. As such, markers are needed that accurately predict patients at high risk for recurrence. Previously our group reported Pfetin as a prognostic biomarker for GIST. In order to create an approach for predicting risk of recurrence, we incorporated Pfetin expression with clinicopathological data to produce a predictive model.

Object

Forty-five patients with localized primary GIST were treated with complete gross surgical resections surgically at our institution between 1995 and 2010 were included. The majority of tumors originated in the stomach (38 cases), as well as small intestine (6 cases) and rectum (1 case).

Method

(1) We performed retrospective analysis of the connection between Pfetin expression, clinicopathological data, and incidences of recurrence, using bivariate and multivariate analyses. (2) The reactivity of the monoclonal antibody against Pfetin was examined by immunohistochemistry. Pfetin. We have reported Pfetin, identified microarray technology, and compared between statistically different GISTs for good and poor prognoses and for prognostic marker. Results. There were 7 cases of recurrences. (1) By univariate analysis, tumor size, mitoses, exposure to abdominal cavity, and complete tumor removal predicted risk of recurrence. (2) Pfetin-negative cases were significantly related to recurrence ($P = 0.002$).

Conclusions

This analysis demonstrates that lack of Pfetin expression is an additional predictor of recurrence in resected GIST. Further study may determine the role of this variable added to the current predictive model for selection of adjuvant therapy.

1. Background

Gastrointestinal stromal tumor (GIST) is the most common mesenchymal tumor of the digestive tract. The precise cellular origin of GIST recently has been proposed to be the interstitial cell of Cajal, an intestinal pacemaker cell [1–3].

Although it comprises a low incidence (2/100,000 people/year) in Japan, their rates of recurrence and mortality remain high (30–40%) [4, 5].

These tumors are almost 90% associated with mutations of c-KIT [6, 7]. And recently c-kit negative cases have been identified as harbor activating mutations of platelet-derived growth factor receptor alpha (PDGFRα).

Most frequently, GISTs are located in the stomach (more than 60%) and the small bowel (30%) but can arise anywhere from the esophagus to the rectum and in the omentum and peritoneum [8, 9].

Before imatinib mesylate (inhibitor of tyrosine kinase) was developed, there were no effective treatments. GISTs uniquely metastasize by hematogenous spread and peritoneal seeding. It is most frequently recurrent in the liver, omentum, and peritoneum [8, 10]. The 5-year survival rate was 11–30%. In 2001, the first reported case of imatinib mesylate yielding striking effects in the treatment of GISTs was reported [11]. Imatinib dramatically changed the mean survival time of GIST (from 32–39 to 58 months) [8].

Despite complete resection, GISTs sometimes recur or metastasize, according to the degree of the malignancy. Viewed in this light, there are many arguments for adjuvant therapy. Treatment for recurrent and/or metastatic GIST with
Imatinib has shown effectiveness. The recent Z9001 trial in the USA that tested 1 year of adjuvant treatment by imatinib reported reduction of the recurrences and metastases (Lancet) [8]. As such, the standard of care for high-risk patients after complete resection is one year of adjuvant therapy.

There are also risks associated with imatinib treatment which include costs and drug resistant from long time treatment by imatinib. One of the imatinib induced complications is the GI bleeding, such as GAVE (define). It can be due to the presence of residual tumor or to other less common etiologies [12, 13]. A multitude of dermatological toxicities also occurs, from various acute rashes to Steven-Johnson syndrome [14].

As with any therapy, balancing risks and benefits is paramount. Thus, it is necessary to diagnose the high-risk group for recurrences and/or metastases accurately. A sensitive and specific marker is sought to select patients at high risk for recurrence. Previously our group has reported Pftin as a prognostic biomarker for GIST, identified using a proteomic approach [15]. Pftin was originally cloned as a gene highly expressed in the fetal cochlea and brain [16]. Pftin is also a member of KCTD family [16] and contributes to carcinogenesis and cancer progression.

The aim of this study was to determine the role of Pftin expression in predicting the incidence of recurrence, in particular related to incorporation of this marker into the current model.

2. Patients

We examined the primary tumor tissues of 45 GIST patients who underwent surgery with complete (R0) resection at our institution between 1995 and 2011. Tumors originated in the stomach (37 cases), duodenum (1 case), small intestine (6 cases), and rectum (1 case). There were 7 recurrences. Diagnosis was based on the World Health Organization classification system for soft-tissue tumors: tumor size, presence of necrosis, differentiation, mitotic rate, MIB-1 index, and presence of epithelioid cells. C-kit expression in all GIST samples was confirmed using immunohistochemical staining (CD117 antibody, DAKO Japan Corp., Tokyo, Japan). Clinicopathological features of the 45 GIST patients are listed in Table 1.

3. Pathologic Analysis

All tumors included in the study were rereviewed by one pathologist, and the diagnosis of GIST was confirmed by positive staining for KIT (CD117) protein, as previously described. Tumor morphology was classified as predominantly epithelioid or spindle-shaped. Mitotic rate was determined by counting the number of mitotic figures per 50 high power fields (HPF) and categorized as <5, 5–10, or >10. Mib-1.

4. Immunohistochemistry

Pftin expression was examined immunohistochemically using paraffin-embedded tissues. 4-mm-thick tissue sections were autoclaved in 10 mmol/L citrate buffer (pH 6.0) at 1218°C for 30 min and incubated with the antibody against Pftin (1:1000 dilution). Immunostaining was carried out according to the streptavidin-biotin peroxidase method using the Strept ABC complex/horseradish peroxidase kit (DAKO). More than 20% of tumor cells were stained with the anti-Pftin antibody which was considered to be Pftin positive.

In most cases, the difference was quite obvious and the two reviewers concurred with the results.

6. Results

6.1. Clinical Features. The median age of the population was 66.2 years (range: 38–87) and there were 24 (53.3%) males. Tumor locations included the stomach in 42 (82%), small bowel in 6 (13.3%), duodenum in 1 (2%), and rectum in 1 (2%) (Table 1). The median tumor size was 5.76 (0.6–15) cm.

| Table 1: Clinicopathological features of the 45 gastrointestinal stromal tumors (GIST) cases. |
|---------------------------------|------|------|------|------|
| Sex                            | Cases | Recurrence (%) | P value |
| Male                           | 24   | 3     | 12.5 | 0.613 |
| Female                         | 21   | 4     | 19.0 |       |
| Age 38–87 years old (mean 66.2) |      |       |      | 0.771 |
| Location                       |      |       |      | 0.635 |
| Stomach                        | 37   | 5     | 14.0 |       |
| Intestine                      | 6    | 2     | 25   |       |
| Duodenum                       | 1    | 0     | 0    |       |
| Rectum                        | 1    | 0     | 0    |       |
| Tumor size                     |      |       |      |       |
| 5–10 cm                       | 10   | 4     | 40   |       |
| >10 cm                        | 10   | 3     | 30   |       |
| Mitosis                        |      |       |      |       |
| <5/50 HPF                      | 32   | 2     | 6.3  | P = 0.035 |
| >5/50 HPF                      | 13   | 5     | 38.5 |       |
| MIB-1 index                    |      |       |      | 0.963 |
| ≥10%                           | 6    | 1     | 16.7 |       |
| <10%                           | 39   | 6     | 15.4 |       |

Tumor size and mitosis have significant correlations with recurrence. These factors are also Fletcher’s agents. These data lead Fletcher’s classification to be good collation to recurrences.
Table 2: Clinical malignant factors of the 45 gastrointestinal stromal tumors (GIST) cases.

|                      | Cases | Recurrence (%) | P value |
|----------------------|-------|----------------|---------|
| Dissemination (tumor explosion) | Positive | 9 | 4 | 44.4 | 0.088 |
|                      | Negative | 36 | 3 | 8.3 | |
| Invasion (other organs) | Positive | 4 | 3 | 75 | 0.094 |
|                      | Negative | 41 | 4 | 9.8 | |
| Damage on operation | Positive | 2 | 2 | 100 | 0.287 |
|                      | Negative | 43 | 5 | 11.6 | |
| Tumor necrosis       | Positive | 8 | 1 | 12.5 | 0.866 |
|                      | Negative | 37 | 6 | 16.2 | |

In clinical malignant factors, dissemination and invasion have a tendency for recurrence.

All patients had complete resections. Seven cases had recurrence and/or metastasis after resection of the primary tumor. Six cases were gastric cases (14%) and 2 cases were intestinal cases (25%).

(1) Tumor Size and Mitosis are High-Risk Markers for GIST. On univariate analysis, tumor size and mitoses were significantly correlated with recurrence (Table 1). These factors were a significant correlation with Fletcher’s risk classification. These data led Fletcher’s classification to be a good correlation with recurrences. All 7 cases of recurrence were grouped as high risk according to Fletcher’s classification.

This demonstrated high accuracy. Dissemination and invasion also correlated with recurrence (Table 2).

(2) Pfetin-Negative Cases Were Significantly Related to Recurrence. Thirteen cases were Pfetin negative and 5/13 of these cases recurred (see Table 3). (P = 0.002). Clinicopathologically, Pfetin correlated with mitoses (P = 0.024).

These data were consistent with those in our previous study in which Pfetin expression was strongly correlated with recurrence and/or metastasis of GIST patients. Pfetin expression was strongly correlated with the prognostic value of GIST patients.

The Kaplan-Meier estimated disease-free survival curves revealed Pfetin expression to correlate significantly and inversely with recurrence. (Figure 1). Disease-free survival was dramatically longer in Pfetin-positive than in Pfetin-negative cases; Pfetin expression was strongly correlated with the prognostic value of GIST patients.

7. Discussion

Imatinib remains the standard of care for adjuvant therapy [10, 17, 18] after surgical resection for primary, localized GIST. Given significant recurrence, despite adjuvant imatinib, we need accurate predictive models for recurrence to guide adjuvant treatment. Current risk classification systems assist in determining the risk of disease recurrence in individual patients with GIST, so disease management can be personalized. However, risks of imatinib including cost, severe side effects, and inconvenience must be considered. Additional prognostic markers are urgently needed to improve decision-making regarding adjuvant therapy for these patients.

The current standard, NIH consensus classification system for GIST, classified patients into risk groups on the basis of tumor size and mitotic index. Subsequently, Mietteinen modified this into the pathology risk stratification system by including tumor location and histology. Gold et al. reported the importance of intestinal location [19, 20] showing that intestinal GISTs are more likely to recur those gastric locations.

Nomograms created by DeMatteo presented risk of recurrence as percentages on a continuous scale. The goal of our study was to consider the impact of the addition of Pfetin level on these models.

In this study, Pfetin expression also correlated with outcome. This protein was discovered by microarray analysis using proteomic technology in Professor Kondo’s lab. Pfetin’s exact role remains to be determined.

Regardless, in this study test set, we showed an inverse relationship between Pfetin expression and risk of recurrence. Future testing in a validation set is planned. If this predictive relationship is confirmed, one might consider including it in a revised model.

8. Conclusion

Pfetin is an independent predictor of recurrence/metastasis for completely resected primary, localized GIST. Further investigation of this role is planned.
Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

The authors would like to express their deepest appreciation to Professor Tadashi Kondo, Director of Proteome Bioinformatics Project, National Cancer Center Research Institute for providing Pfetin, constructive comments, and warm encouragement.

References

[1] M. Miettinen, M. Sarlomo-Rikala, and J. Lasota, “Gastrointestinal stromal tumours,” Annales Chirurgiae et Gynaecologiae, vol. 87, no. 4, pp. 278–281, 1998.

[2] J. A. Breiner, J. Meis-Kindblom, L.-G. Kindblom et al., “Loss of 14q and 22q in gastrointestinal stromal tumors (pacemaker cell tumors),” Cancer Genetics and Cytogenetics, vol. 120, no. 2, pp. 111–116, 2000.

[3] H. Yamamoto, Y. Oda, K.-I. Kawaguchi et al., “c-kit and PDGFRA Mutations in Extragastrointestinal Stromal Tumor (Gastrointestinal Stromal Tumor of the Soft Tissue),” The American Journal of Surgical Pathology, vol. 28, no. 4, pp. 479–488, 2004.

[4] G. D. Demetri, P. Reichardt, Y. K. Kang et al., “Efficacy and safety of regorafenib for advanced gastrointestinal stromal tumours after failure of imatinib and sunitinib (GRID): an international, multicentre, randomised, placebo-controlled, phase 3 trial,” The Lancet, vol. 381, no. 9863, pp. 295–302, 2013.

[5] T. Nishida, K. Shirao, A. Sawaki et al., “Efficacy and safety profile of imatinib mesylate (STI571) in Japanese patients with advanced gastrointestinal stromal tumors: a phase II study (STI571B202),” International Journal of Clinical Oncology, vol. 13, no. 3, pp. 244–251, 2008.

[6] H. Joensuu, C. Fletcher, S. Dimitrijevic et al., “Management of malignant gastrointestinal stromal tumours,” The Lancet Oncology, vol. 3, no. 11, pp. 655–664, 2002.

[7] M. Miettinen and J. Lasota, “Gastrointestinal stromal tumors (GISTs): definition, occurrence, pathology, differential diagnosis and molecular genetics,” Polish Journal of Pathology, vol. 54, no. 1, pp. 3–24, 2003.

[8] R. P. DeMatteo, K. V. Ballman, C. R. Antonescu et al., “Adjuvant imatinib mesylate after resection of localised, primary gastrointestinal stromal tumour: a randomised, double-blind, placebo-controlled trial,” The Lancet, vol. 373, no. 9669, pp. 1097–1104, 2009.

[9] R. Rajendra, S. M. Pollack, and R. L. Jones, “Management of gastrointestinal stromal tumors,” Future Oncology, vol. 9, no. 2, pp. 193–206, 2013.

[10] M. Miettinen, M. Majidi, and J. Lasota, “Pathology and diagnostic criteria of gastrointestinal stromal tumors (GISTs): a review,” European Journal of Cancer, vol. 38, supplement 5, pp. S39–S51, 2002.

[11] J. Berman and T. J. O’Leary, “Gastrointestinal stromal tumour workshop,” Human Pathology, vol. 32, no. 6, pp. 578–582, 2001.

[12] K. Pornsuk Siri, S. Chewatanakornkul, S. Kanngurn et al., “Clinical outcomes of gastrointestinal stromal tumor in southern Thailand,” World Journal of Gastrointestinal Oncology, vol. 4, no. 11, pp. 216–222, 2012.

[13] M. Valadão and E. Linhares, “The role of the surgeon in the management of GIST,” Revista do Colegio Brasileiro de Cirurgioes, vol. 36, no. 3, pp. 261–265, 2009.

[14] D. T. Alexandrescu, C. A. Dasanu, H. Farzanmehr, and C. L. Kaufman, “Persistent cutaneous hyperpigmentation after tyrosine kinase inhibition with imatinib for GIST,” Dermatology Online Journal, vol. 14, no. 7, article 8, 2008.

[15] Y. Suehara, T. Kondo, K. Seki et al., “Pfetin as a prognostic biomarker of gastrointestinal stromal tumors revealed by proteomics,” Clinical Cancer Research, vol. 14, no. 6, pp. 1707–1717, 2008.

[16] B. L. Resendes, S. F. Kuo, N. G. Robertson et al., “Isolation from cochlea of a novel human intronless gene with predominant fetal expression,” Journal of the Association for Research in Otolaryngology, vol. 5, no. 2, pp. 185–202, 2004.

[17] C. B. Tan, W. Zhi, G. Shahzad, and P. Mustacchia, “Gastrointestinal stromal tumours: a review of case reports, diagnosis, treatment, and future directions,” ISRN Gastroenterology, vol. 2012, Article ID 595968, 16 pages, 2012.

[18] A. Rammohan, J. Sathyanesan, K. Rajendran et al., “A gist of gastrointestinal stromal tumours: a review,” World Journal of Gastrointestinal Oncology, vol. 5, no. 6, pp. 102–112, 2013.

[19] J. S. Gold, M. Gönen, A. Gutiérrez et al., “Development and validation of a prognostic nomogram for recurrence-free survival after complete surgical resection of localised primary gastrointestinal stromal tumour: a retrospective analysis,” The Lancet Oncology, vol. 10, no. 11, pp. 1045–1052, 2009.

[20] S. Grover, S. W. Ashley, and C. P. Raut, “Small intestine gastrointestinal stromal tumours,” Current Opinion in Gastroenterology, vol. 28, no. 2, pp. 113–123, 2012.