Amyloid- and tau-PET imaging in a familial prion kindred

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Abstract

Objective
To study the in vivo binding properties of $^{18}$F-AV-1451 (tau-PET) and Pittsburgh compound B (PiB-PET) in a unique kindred with a familial prion disorder known to produce amyloid plaques composed of prion protein alongside Alzheimer disease (AD)–like tau tangles.

Methods
A case series of 4 symptomatic family members with the 12-octapeptide repeat insertion in the PRNP gene were imaged with 3T MRI, PiB-PET, and tau-PET in their fourth decade of life.

Results
There was significant neocortical uptake of the tau-PET tracer in all 4 familial prion cases. However, PiB-PET images did not demonstrate abnormally elevated signal in neocortical or cerebellar regions for any of the patients.

Conclusions
In vivo detection of molecular hallmarks of neurodegenerative diseases will be a prerequisite to well-conducted therapeutic trials. Understanding the in vivo behavior of these PET biomarkers in the setting of various neurodegenerative processes is imperative to their proper use in such trials and for research studies focused on the basic neurobiology of neurodegeneration. This study supports the high specificity of neocortical $^{18}$F-AV-1451 binding to AD-like tau and the lack of PiB binding to PrP plaques. It is uncertain how early in the disease course tau pathology appears in the brains of individuals who carry this PRNP gene mutation or how it evolves throughout the disease course, but future longitudinal $^{18}$F-AV-1451 imaging of symptomatic and asymptomatic individuals in this kindred will help address these uncertainties.

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Familial prion diseases are associated with prion protein (PrP) deposits and genetic mutations in the PRNP gene. In addition to the PrP deposits seen in all subtypes, other characteristic pathologies include the following: spongiform changes in familial Creutzfeldt-Jakob disease, PrP amyloid plaques in the cerebellum in Gerstmann-Strassler-Scheinker syndrome, and thalamic neuronal loss seen in fatal familial insomnia. We have reported on a familial prion kindred with a 12-octapeptide repeat insertion in the PRNP gene characterized by a frontotemporal dementia (FTD)-like presentation with mixed pathologic features. These cases were pathologically characterized by cerebellar plaques that were negative for Aβ40 and Aβ42 but positive for PrP. The neocortical pathology consisted of Alzheimer disease (AD)-like neurofibrillary tangles (NFT) that were present in addition to the PrP pathology. The tau pathology found in these cases is unusual for sporadic prion disease, but it has been reported in other familial prion cases, where it was composed of the same AD-like tau isoforms. We have recently shown that ex vivo and in vivo binding of 18F-AV-1451 (tau-PET) is specific for AD-like tau isoforms and that the regional distribution is distinct for AD dementia relative to familial FTD with mutations in the MAPT gene.

Pittsburgh compound B (PiB) is a derivative of thioflavin. Therefore, it is possible that PiB-PET could bind PrP plaques in this prion kindred. The cerebellar predominance of PrP plaques is distinct from the neocortical NFTs, allowing these aggregates to be differentiated spatially in vivo. In this study, we report the tau-PET and PiB-PET binding patterns in this kindred that has both tau aggregates and PrP plaques.

Methods

Standard protocol approvals, registrations, and patient consents

All participants or their designees provided written informed consent with approval of the Mayo Clinic Foundation and Olmsted Medical Center Institutional Review boards.

Participants and imaging

Participants were part of the Mayo Clinic AD Research Center in Rochester, Minnesota. Four symptomatic family members of the 12-octapeptide repeat insertion kindred were imaged with 3T MRI, amyloid-PET with (PiB-PET), and AV-1451 (tau-PET) in their fourth decade of life. These patients were all asymptomatic for 1–6 years and had a behavioral variant FTD-like presentation with minimal parkinsonism. Case 1 also had a prominent nonfluent aphasia. Detailed demographic information is not relevant to the objective of this study and may allow for patient identification. Therefore, we do not report additional metadata that can be linked to the cases. One patient with young-onset AD was also evaluated for comparison purposes. Given the potential for on-target PET tracer uptake in the cerebellum, which is commonly used as a reference region for quantification, we used visual assessment as the gold standard without normalizing signal intensity to a reference region. Once we verified that there was no regional cerebellar signal on either PET modality, we created intensity-normalized images using the gray matter in the cerebellar crus for display purposes only.

Data availability

The datasets analyzed in the current study are not publicly available due to restricted access, but further information about the datasets are available from the corresponding author on reasonable request.

Results

There was significant neocortical uptake of the tau-PET tracer in all 4 familial prion cases (figure 1, left). However, PiB-PET images did not demonstrate abnormally elevated signal in neocortical regions for any of the patients (figure 1, right). There was no abnormally elevated signal in the cerebellum on either PET modality. The global distribution of the elevated tau-PET signal (figure 2A) matched the known distribution of NFT in AD (figure 2B) in that the homotypic isocortex was severely affected with a stark sparing of the idiotypic cortex. However, there did seem to be relatively greater involvement of the frontal lobe compared with the precuneus in the familial prion participant.

Discussion

This study demonstrates 2 important properties of these widely used molecular PET ligands: (1) in vivo neocortical tau-PET binding can detect AD-like NFT in the setting of familial prion disease and (2) PiB-PET did not detect amyloid plaques composed of prion protein in this kindred. This has important implications for the use of these ligands in the study of AD. These findings also indicate that the tau-PET is a promising tool for investigating the molecular basis of neurodegeneration in this familial prion kindred and other hereditary prion diseases with coexisting AD-like NFT pathology.

Four members of the 12-octapeptide repeat insertion in the PRNP gene kindred did not have any abnormally elevated PiB-PET signal. The fact that no elevated in vivo PiB-PET signal was seen in a kindred known to harbor amyloid plaques composed of PrP suggests that PiB-PET may be specific for amyloid plaques composed of beta-amyloid. In the

Glossary

AD = Alzheimer disease; FTD = frontotemporal dementia; NFT = neurofibrillary tangles; PiB = Pittsburgh compound B.
absence of pathologic confirmation, an alternative explanation would be that no PrP amyloid plaques were present at this stage of the disease. This seems unlikely because all 4 cases were clearly symptomatic, including 1 patient who was more than 6 years into the disease course when molecular imaging was obtained.

These PiB-PET findings stand in stark contrast to the intense uptake on tau-PET imaging, with the global spatial distribution mirroring that seen in AD (figure 2). This distribution does not follow the known distribution of the PrP plaques and, therefore, this does not represent off-target binding of PrP plaques. The magnitude seen in these familial prion cases is similar to that seen in individuals with young-onset AD dementia, which supports the possibility that the tau-PET binding seen in these cases is related to the known NFT pathology that occurs in this family. Although, this would need to be confirmed by future pathologic evaluations, the finding suggests good sensitivity and specificity for AD-like tau aggregations.

In vivo detection of molecular hallmarks of neurodegenerative diseases will be a prerequisite to well-conducted therapeutic trials. Not only will such biomarkers be able to assess whether the pathology of interest is actually present, but these molecular PET biomarkers also have the potential to objectively index disease severity and response to therapeutic intervention. Understanding the in vivo behavior of these PET biomarkers under various neurodegenerative circumstances is imperative to their proper use in clinical trials and research studies on the basic neurobiology of neurodegeneration.

This study demonstrates neocortical AV-1451 binding of presumed AD-like NFT and the lack of PiB binding to PrP plaques in this kindred. Although the spatiotemporal dynamics of how this pathology evolves across the neocortex in
this kindred remains unknown, future longitudinal tau-PET studies of symptomatic and asymptomatic individuals will be able to address this question directly.

**Author contributions**
D.T. Jones: acquisition of data, analysis and interpretation of data, and manuscript preparation. R.A. Townley, J. Graff-Radford, H. Botha, D.S. Knopman, R.C. Petersen, C.R. Jack, V.J. Lowe, and B.F. Boeve: acquisition of data, analysis and interpretation of data, and critical revision of the manuscript for intellectual content.

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References
1. Mastrianni JA. The genetics of prion diseases. Genet Med 2010;12:187–195.
2. Kumar N, Boeve BF, Boot BP, et al. Clinical characterization of a kindred with a novel 12-octapeptide repeat insertion in the prion protein gene. Arch Neurol 2011;68:1165–1170.
3. Ishizawa K, Komiya T, Shimazu T, et al. Hyperphosphorylated tau deposition parallels prion protein burden in a case of Gerstmann-Straussler-Scheinker syndrome P102L mutation complicated with dementia. Acta Neuropathol 2002;104:342–350.
4. Jayadev S, Nochlin D, Poorkaj P, et al. Familial prion disease with Alzheimer disease-like tau pathology and clinical phenotype. Ann Neurol 2011;69:712–720.
5. Tranchant C, Sergeant N, Warter A, Mohr M, Warter JM, Delacourte A. Neurofibrillary tangles in Gerstmann-Straussler-Scheinker syndrome with the A117V prion gene mutation. J Neurol Neurosurg Psychiatry 1997;63:240–246.
6. Lowe VJ, Curran G, Fang P, et al. An autoradiographic evaluation of AV-1451 Tau PET in dementia. Acta Neuropathol Commun 2016;4:58.
7. Jones DT, Knopman DS, Graff-Radford J, et al. In vivo (18)F-AV-1451 tau PET signal in MAPT mutation carriers varies by expected tau isoforms. Neurology 2018;90:e947–e954.
8. Xia CF, Arteaga J, Chen G, et al. [(18)F]T807, a novel tau positron emission tomography imaging agent for Alzheimer’s disease. Alzheimers Dement 2013;9:666–676.