Morphometric analysis and three-dimensional computed tomography reconstruction of the long bones of femoral and crural regions in Van cats

O. Yılmaz¹, İ. Demircioğlu²

¹Department of Anatomy, Faculty of Veterinary Medicine, Van Yüzüncü Yıl University, Van, Turkey
²Department of Anatomy, Faculty of Veterinary Medicine, Harran University, Şanlıurfa, Turkey

[Received: 21 June 2020; Accepted: 6 September 2020]

Background: This study was conducted to determine the morphometric features and to make the three-dimensional (3D) reconstruction of the images obtained by computed tomography (CT) of the main long bones (femur, tibia, fibula) in Van cats, and to reveal the biometric differences between these measurement values of both sexual dimorphism and homotypic variations.

Materials and methods: A total of 16 adult Van cats, 8 male and 8 female, were used in the study. The pelvic limb long bones of the cats, which were anesthetized with the ketamine-xylazine combination, were scanned with a CT device and their images were obtained. These images were converted to a 3D structure by means of MIMICS 20.1 (The Materialise Group, Leuven, Belgium) software programme. Then, morphometric, volumetric, surface area measurements of these bones were calculated and statistical analyses were performed.

Results: In the analysis, it was observed that right greatest length (GL), left GL, right greatest length from caput ossis femoris (GLc), left greatest breadth of the distal end (Bd), right volume (V), and left V measurement values of the femur; left GL1, right greatest breadth of the proximal end (Bp), right Bd, left Bd, right smallest breadth of the diaphysis (SD), right V, left V, and right surface area (SA) measurement values of the tibia; right V, left V, right SA, and left SA measurement values of the fibula were statistically significantly higher in male cats than in female cats (p < 0.05). The volume and surface area measurement values of the femur, tibia and fibula of male and female cats were determined as 8.57 ± 1.33 cm³ and 7.00 ± 0.49 cm³, 64.28 ± 6.72 cm² and 59.42 ± 4.72 cm²; 7.56 ± 1.28 cm³ and 6.15 ± 0.52 cm³, 56.89 ± 6.47 cm² and 52.72 ± 3.15 cm²; 1.10 ± 0.17 cm³ and 0.83 ± 0.05 cm³, 14.18 ± 1.61 cm² and 11.95 ± 0.48 cm², respectively.

Conclusions: The statistical differences between the sexes in terms of the measurement parameters of the femur, tibia and fibula in adult Van cats were determined. We believe that these differences can be used in determining morphological variations of Van cats. In addition, it is thought that the data obtained from the present study would be beneficial to veterinary physicians in the clinical application areas in order to evaluate the pathological conditions related to these bones and to the studies in the field of zooarchaeology.

Key words: computed tomography, femur, fibula, three-dimensional reconstruction, tibia, Van cat
INTRODUCTION

Living around and named after the Van Lake region of Turkey, Van cats are an attractive breed of domestic cats with their distinctive eye coloration (one eye can be yellow and the other can be blue, both can be yellow, or both can be blue), soft fur, affection for swimming, hunting capabilities, cleanliness, and intelligence. In 22 April 2006, Van cats were licensed and registered with the communiqué no. 2006/16 as a breed of domestic cat with all their distinctive properties, and are now officially under protection of the Republic of Turkey [6]. There are only a handful of studies in literature, however, regarding the osteological properties of these cats [46–48].

Known also as “the thigh bone”, femur (os femoris) is the strongest of all the long bones. It usually has a cranioventral bearing along the skeletal structure, and it plays a significant role during the forward motion of the body [14]. Meanwhile, “the osa cruris” structure is another formation of the long bones of the pelvic limbs, and consists of the combination of the tibia and fibula bones. Tibia is a long, tubular bone and is the stronger of the two as it shoulders the weight of the body. It is located towards the medial surface of the leg, and has joints with the femur at the proximal end and with osa tarsi at the distal end. Located towards the lateral surface, the fibula is weaker and does not have a joint with femur at the proximal end. On the distal end, however, it has a limited amount of contact with osa tarsi [4]. In ruminants, the proximal end of the fibula makes a joint with tibia, but the distal end only reaches half-way down the range of tibia where it ends with a pointy edge. Sus and carnivore species have a fully developed fibula, but it’s separated from tibia with a formation known as “spatium interosseum cruris”. This gap, which is enclosed with soft tissue, goes almost all the way down to the distal edge in Sus species, while it’s only limited to the proximal region in carnivores [15].

In recent years, medical imaging systems, computed tomography (CT), and three-dimensional (3D) reconstruction software become commonly utilized tools when trying to determine the morphological properties of the anatomical structures in the skeletal systems of animals, particularly those of small, pet animals like cats and dogs. These systems are also being used to obtain osteometric and volumetric measurement results, to evaluate any potential developmental, metabolic, inflammatory, or degenerative anomalies, and to investigate numerous other pathological situations like traumas, dislocations, fractures, or neoplasia [26, 45]. Anthropological studies also commonly use these methods and 3D modelling in tandem to determine the morphological properties of long bones and to perform various measurements on them [8, 17, 21].

Various studies have been performed to determine the anatomical and osteometrical properties of the main long bones of the pelvic limbs (femur, tibia, fibula) of different animal species, like mole-rats (Spalax leucodon Nordmann) [34], African giant rats (Cricetomys gambianus Waterhouse) [27], grasscutter rodents (Thryonomyos swinderianus, Temminck-1827) [29], guinea pigs (Cavia porcellus) [37], feral pigs (Sus scrofa) [19], martens (Martes fonia) [3], badgers (Meles meles) [32], squirrels (Sciurus vulgaris) [31], hedgehogs hedgehog (Erinaceus europaeus L.) [35], porcupines (Hystrix Cristata) [49], chinchillas (Chinchilla lanigera) [10], lomholtz tree-kangaroo (Dendrolagus lumholtzi) [40], Anatolian bobcats (Lynx lynx) [30], New Zealand rabbits (Oryctolagus cuniculus) [1, 16, 36], marmosets (Callithrix jacchus) [9], brown bears (Ursus arctos) [13], deers (Hippocamelus bisulcus) [41], sheeps (Ovis aries) [2], gazelles (Gazella subgutturosa) [12], foxes (Vulpes vulpes and Alopex lagopus) [24], dogs (Canis lupus familiaris) [28, 43], and domestic cats (Felis catus and Felis domestica) [5, 16]. No study was found when the existing literature was searched for long bones of the pelvic limbs of Van cats, however. This study was, therefore, performed with the aim of creating 3D models of the pelvic limb long bones of Van cats using CT scans and imaging of different anatomical structures of these bones, and to obtain osteometric and volumetric measurements and to determine any potential biomeirical difference in these measured values with regards to sexual dimorphism and homotypic variations.

MATERIALS AND METHODS

The total of 16 adult Van cats (8 males and 8 females) obtained from Van Yuzuncu Yil University Van Cat Research and Application Centre was used in this study. The ages of the cats varied between 3 and 8 years, and their live body masses (W) varied between 5810 and 8050 grams. Ad libitum cat feed and tap water was provided to these cats until the day before the study. The study received the approval of the Van Yuzuncu Yil University Van Cat Research and Application Centre's institutional ethics committee.
University Local Ethical Board for Animal Experimentation (Decision no: 2020/02-27.02.2020). The cats included in the study were numbered, and were not provided any feed starting the day before the study. Anaesthetisation of the cats for the scanning process was performed using a combination of ketamine (15 mg/kg, IM, Ketasol® 10% injectable, İnterhas Veterinary Medicines, Ankara) and xylazine (1–2 mg/kg, IM, Alfazyne® 2% injectable, Ege – Vet Veterinary Medicines, İzmir).

A 16-section multi-sequential CT device (Somatom Sensetion 16; Siemens Medical Solutions, Erlangen, Germany) located in Van Yuzuncu Yil University Medical Faculty Radiology Department was used to obtain the CT scans of the cats. A disposable sheet was placed onto the gantry of the device, and the cats were laid onto this sheet head-first in prone position (ventral decubitus) to achieve symmetry. The device parameters for the CT scan were as follows: KV/Effective mAs/Rotation time (sec) 120/120/0.75; gantry rotation period 420 ms; physical detector collimation, 16 × 0.6 mm; section thickness 0.4 mm; final section collimation 32 × 0.63 mm; feed/rotation 6 mm; Kernel, U90u; increment 0.5 mm; and resolution 512 × 512 pixels. Setting of the dosage parameters and the scanning process itself were performed on the basis of standard protocols established by the literature [18, 39]. The images obtained were saved in DICOM format.

These images were then transported to the MIM-ICS 20.1 (The Materialise Group, Leuven, Belgium) software and were reconstructed. Osteometric measurements were then performed on the 3D models of the femur,ibia, and fibula bones. The measurement points specified in the literature were used during the morphometric measurements [44]. Once the morphometric measurements were complete, the surface area and volume of each of the bones were calculated. The definitions and abbreviations for the measured osteometric parameters are presented in Table 1. Nomina Anatomica Veterinaria [25] was used as the terminology basis throughout the study. Weighing of the cats included in the study was performed using a digital scale (TESS®, RP-LCD, Çomak Terazi, İstanbul).

### Statistical analysis

Van cat pelvic limb long bones (femur, tibia, and fibula) were modelled in 3D environment based on the CT images of the cats obtained as part of study, and Shapiro-Wilk test (n < 50) was used to determine if the value distribution of the data obtained from these models was normal. Since the distribution of the values

---

**Table 1. Studied parameters and abbreviations (according to Von den Driesch, 1976 [44])**

| Parameter | Abbreviation | Definition |
|-----------|--------------|------------|
| **Measurement points and abbreviations of the femur in Van cats** | | |
| 1 | GL | Greatest length: distance between the proximal and the distal end [mm] |
| 2 | GLc | Greatest length from caput ossis femoris: distance between the caput ossis femoris and the distal end [mm] |
| 3 | Bp | Greatest breadth of the proximal end [mm] |
| 4 | Bd | Greatest breadth of the distal end [mm] |
| 5 | Dc | Greatest depth of the caput ossis femoris [mm] |
| 6 | SD | Smallest breadth of the diaphysis (Corpus ossis femoris) [mm] |
| 7 | V | Volume of the femur [cm³] |
| 8 | SA | Surface area of the femur [cm²] |

| **Measurement points and abbreviations of the tibia in Van cats** | | |
| 1 | GL1 | Greatest length: distance between the proximal and the distal end [mm] |
| 2 | Bp | Greatest breadth of the proximal end [mm] |
| 3 | Bd | Greatest breadth of the distal end [mm] |
| 4 | SD | Smallest breadth of the diaphysis (Corpus tibiae) [mm] |
| 5 | V | Volume of the tibia [cm³] |
| 6 | SA | Surface area of the tibia [cm²] |

| **Measurement points and abbreviations of the fibula in Van cats** | | |
| 1 | GL2 | Greatest length: distance between the proximal and the distal end [mm] |
| 2 | V | Volume of the fibula [cm³] |
| 3 | SA | Surface area of the fibula [cm²] |
weren't normal in general, a number of non-parametric tests were performed throughout the rest of the study. The defining statistics for the measured properties in this study are presented as mean, standard deviation, and minimum and maximum values. Wilcoxon test was used to compare the measurements for left and right side bones. Comparisons between the sexes were performed using Mann-Whitney-U test. The correlations between the measurements were investigated through Spearman Correlation coefficient calculations, and were separately performed for each sex. Statistical significance threshold was taken as ($\alpha$) 5%, and SPSS (IBM SPSS for Windows, Ver.23) package software was used for calculations.

**RESULTS**

The volume and surface area measurements of the Van cat pelvic limb femur, tibia, and fibula were performed as part of this study. The morphometric measurement parameters and 3D reconstructions are presented in Figures 1–3 with regards to the measurement point locations on bones. These results were then compared in terms of sexual dimorphism and homotypic variations, and are presented in Tables 2–7. Any statistically significant differences between the results ($p < 0.05$) were recorded.

Table 2 presents the defining statistics for femurs’ morphometric measurement results and the comparison of the sexes in terms of homotypic variations.

Accordingly, right greatest length (GL), left GL, right greatest length from caput ossis femoris (GLc), left greatest breadth of the proximal end (Bp), right greatest breadth of the distal end (Bd), left greatest breadth of the distal end (Bd), and left V values were found to be higher in males in a statistically significant manner, compared to females ($p < 0.05$). When the left and right femur defining statistics were compared within each sex, right smallest breadth of the diaphysis (SD)
values were found to be higher than left SD values in females and the difference was statistically significant (p = 0.036). Furthermore, male left Bd values were statistically higher than right Bd values (p = 0.017). No meaningful difference was determined for any other right and left side average femur values.
### Table 4. Descriptive statistics and homotypic variations of the biometric parameters of the tibia obtained from three-dimensional reconstruction of computed tomography images in Van cats

| Parameter | Male | Female | P*   |
|-----------|------|--------|------|
|           | Mean ± SD | Minimum | Maximum | Mean ± SD | Minimum | Maximum |
| Right GL1 | 113.27 ± 4.84 | 105.43 | 120.74 | 108.14 ± 3.06 | 103.27 | 112.80 | 0.059 |
| Left GL1  | 115.37 ± 5.48 | 105.09 | 123.31 | 107.86 ± 2.97 | 102.42 | 110.81 | 0.036 |
| Right Bp  | 19.12 ± 0.70 | 18.17 | 19.95 | 18.31 ± 0.54 | 17.37 | 19.10 | 0.021 |
| Left Bp   | 19.07 ± 0.90 | 17.41 | 20.21 | 18.37 ± 0.33 | 17.81 | 18.77 | 0.052 |
| Right Bd  | 15.02 ± 0.72 | 14.34 | 16.11 | 13.80 ± 0.61 | 12.79 | 14.58 | 0.006 |
| Left Bd   | 15.31 ± 0.67 | 14.33 | 16.40 | 13.85 ± 0.61 | 12.71 | 14.47 | 0.001 |
| Right SD  | 8.16 ± 0.68 | 7.17 | 9.16 | 7.40 ± 0.43 | 6.89 | 8.05 | 0.036 |
| Left SD   | 7.91 ± 0.79 | 7.12 | 9.40 | 7.58 ± 0.46 | 6.99 | 8.08 | 0.674 |
| Right V   | 7.55 ± 1.27 | 6.05 | 9.50 | 6.18 ± 0.51 | 5.36 | 6.91 | 0.036 |
| Left V    | 7.57 ± 1.28 | 6.02 | 9.62 | 6.11 ± 0.53 | 5.30 | 7.01 | 0.021 |
| Right SA  | 56.84 ± 6.48 | 42.70 | 63.21 | 52.53 ± 2.95 | 47.49 | 57.00 | 0.027 |
| Left SA   | 56.94 ± 6.46 | 43.05 | 62.66 | 52.91 ± 3.35 | 47.81 | 56.94 | 0.093 |

*p < 0.05; **p < 0.01; abbreviations — see Table 1

### Table 5. "r" values of Spearman’s rho nonparametric correlations coefficients between the morphometric measurements of the tibia in Van cats by gender

| Female | Age (A) | Body mass (W) | Right GL1 | Left GL1 | Right Bp | Left Bp | Right Bd | Left Bd | Right SD | Left SD | P* |
|--------|---------|---------------|-----------|----------|----------|---------|----------|---------|----------|---------|------|
|        | 0.957*** | 0.878** | 0.878** | 0.781* | 0.830* | 0.634 | 0.390 | 0.830* | 0.781* | 0.719* | 0.731* | 0.059 |
|        | 0.488  | 0.952** | 0.922** | 0.814* | 0.814* | 0.707 | 0.599 | 0.719* | 0.731* | 0.719* | 0.731* | 0.059 |
|        | -0.342 | 0.262 | 0.929** | 0.690 | 0.714* | 0.619 | 0.476 | 0.667 | 0.571 | 0.571 | 0.571 | 0.059 |
|        | 0.098  | 0.595 | 0.476 | 0.405 | 0.905** | 0.762* | 0.429 | 0.500 | 0.714* | 0.731* | 0.731* | 0.059 |
|        | 0.390  | 0.571 | 0.167 | 0.333 | 0.690 | 0.476 | 0.357 | 0.571 | 0.810* | 0.810* | 0.810* | 0.059 |
|        | 0.488  | -0.310 | -0.786* | -0.714* | -0.333 | 0.167 | 0.357 | 0.357 | 0.333 | 0.333 | 0.333 | 0.059 |
|        | 0.088  | -0.476 | -0.429 | -0.405 | -0.095 | 0.286 | 0.810* | 0.238 | 0.381 | 0.381 | 0.381 | 0.059 |
|        | -0.025 | 0.060 | 0.431 | 0.467 | 0.407 | 0.192 | -0.443 | -0.084 | 0.833* | 0.833* | 0.833* | 0.059 |
|        | 0.537  | 0.524 | -0.024 | 0.214 | 0.048 | 0.167 | -0.262 | -0.452 | 0.515 | 0.515 | 0.515 | 0.059 |

*p < 0.05; **p < 0.01; abbreviations — see Table 1

### Table 6. Descriptive statistics and homotypic variations of the biometric parameters of the fibula obtained from three-dimensional reconstruction of computed tomography images in Van cats

| Parameter | Male | Female | P* |
|-----------|------|--------|------|
|           | Mean ± SD | Minimum | Maximum | Mean ± SD | Minimum | Maximum |
| Right GL2 | 105.09 ± 5.08 | 97.92 | 113.15 | 100.78 ± 2.32 | 97.28 | 103.99 | 0.074 |
| Left GL2  | 105.67 ± 6.22 | 97.33 | 115.56 | 100.86 ± 2.64 | 96.84 | 104.18 | 0.074 |
| Right V   | 1.09 ± 0.16 | 0.90 | 1.29 | 0.83 ± 0.05 | 0.76 | 0.90 | 0.001 |
| Left V    | 1.10 ± 0.18 | 0.83 | 1.30 | 0.82 ± 0.05 | 0.73 | 0.90 | 0.003 |
| Right SA  | 14.07 ± 1.60 | 11.72 | 15.97 | 11.95 ± 0.43 | 11.15 | 12.60 | 0.012 |
| Left SA   | 14.28 ± 1.62 | 11.77 | 16.07 | 11.95 ± 0.53 | 11.21 | 12.56 | 0.012 |

*p < 0.05; Mann-Whitney U test; SD — standard deviation; other abbreviations — see Table 1
Table 3 shows the relationship between the morphometric values for femur based on sex. Accordingly, female cats have positive correlations between the following values (p < 0.05): age and left Bd; right GL with left GL, right GLc, and left GLc; right greatest breadth of the proximal end (Bp) with left Bd and left SD; right Bd with left Bd, right greatest depth of the caput ossis femoris (Dc), and left Dc; left Bd and right Dc; and finally, right SD and left SD. The male cats, on the other hand, were found to have positive correlations between the following values (p < 0.05): between ‘age and W’ with right GL, left GL, left GLc, left Dc, and right SD values; right GL with left GL1, right Bp, left Bp, right SD, and left SD values; between right GL1 with left GL1, right Bp and left Bp values; between left GL1 and left Bp values; between right Bp with left Bp, right Bd, and left SD values; and between left SD with left Bp and right SD values (p < 0.05).

The defining statistics for the morphometric values of tibia, and the comparisons in terms of homotypic variations, are provided in Table 4. Accordingly, male cats were found to have a higher left side SA value compared to the right side, and the difference was statistically significant (p = 0.012). No other statistical difference was determined between the right and left side tibia average values per sex (p > 0.05). That being said, right V, left V, right SA, and left SA values were found to be higher in males compared to females in a statistically significant manner (p < 0.05). Tibia average volume was determined as 7.56 ± 1.28 cm³ for males and as 6.15 ± 0.52 cm³ for females, while its SA was determined as 56.89 ± 6.47 cm² for males and as 52.72 ± 3.15 cm² for females, on average.

Table 5 presents the relationship between the morphometric values of tibia per sex. Accordingly, a positive correlation with 92.9% ratio was determined between the right GL1 and left GL1 values of the female cats, while a positive correlation with 81% ratio was determined between their right Bd and left Bd values (p < 0.05). Meanwhile, negative correlations for females between right Bd with right GL1 and left GL1 values were determined with 78.6% and 71.4% ratio, respectively (p < 0.05). For male cats, a positive correlation was shown between age and W values with right GL1, left GL1, right Bp, left Bp, right SD, and left SD values; between right GL1 with left GL1, right Bp and left Bp values; between left GL1 and left Bp values; between right Bp with left Bp, right Bd, and left SD values; and between left SD with left Bp and right SD values (p < 0.05).

The defining statistics for the morphological values of fibula per sex and homotypic variations are provided in Table 6. Accordingly, male cats were found to have a higher left side SA value compared to the right side, and the difference was statistically significant (p = 0.012). No other statistical difference was determined between the right and left side tibia average values per sex (p > 0.05). That being said, right V, left V, right SA, and left SA values were found to be higher in males compared to females in a statistically significant manner (p < 0.05). Fibula average volume was determined as 1.10 ± 0.17 cm³ for males and as 0.83 ± 0.05 cm³ for females, while its surface area was determined as 14.18 ± 1.61 cm² for males and as 11.95 ± 0.48 cm² for females.

Table 7 presents the relationship between the measured morphometric values for fibula bones. Accordingly, a positive correlation in male cats between age with W, right GL2 and left GL2 values with 95.7%, 78.1%, and 87.8% ratio, respectively, while a positive correlation was found between W with right GL2 and left GL2 values with 83.8% and 92.2% ratios, and between right GL2 and left GL2 values with 97.6% ratio (p < 0.05). Similarly, a positive correlation between right GL2 and left GL2 values was determined for females with 95.2% as well (p < 0.01).

**DISCUSSION**

Studies on the morphology of animal bones yield important data for a variety of developmental, evolutionary, and forensic sciences. Studies on the morphometric and volumetric analyses and morphology of the long bones, in particular, are of great help in
determining crucial behavioural activities that differ among animal species, such as foraging, hunting, evading hunters, and migration; as well as their phylogeny, allometry and postures. In addition, these measurements hold an important place in determining morphological variations and taxonomic classifications among different species of the same genus; as well as identifying the differences between sexes [5, 23, 38]. For this purpose, many studies have been conducted on humans [21, 22, 42] and animals [2, 5, 9, 10, 14, 16, 29, 33, 36] that involved obtaining osteometric measurements on the long bones of their pelvic limbs. In our study, the morphometric, volumetric and surface area measurements of the femur, tibia, and fibula of adult Van cats were obtained via CT scans and 3D modelling, in order to identify the differences between the two sexes. According to these statistics, the right GL, left GL, right GLc, left Bd, right V, and left V values of the femur, the left GL1, right Bp, right Bd, left Bd, right SD, right V, left V, and right SA values of the tibia, and the right V, left V, right SA, and left SA values of the fibula are higher in male cats than that of female cats in a statistically significant fashion (p < 0.05). Most of the other measurements were likewise found to be higher in male cats. We can, therefore, surmise that the femur, tibia and fibula of the male cats are larger than that of the female cats. We think that the larger bones of the pelvic limbs in male Van cats are correlates to the body mass as well as the bones in thoracic limbs according to data obtained from the previous studies by Yilmaz [48].

In the study conducted by Pazar and Kahveciolgu [36] on 15 male and 15 female New Zealand rabbits, the researchers found no significant difference between the morphometric properties on the long bones of the right and left pelvic limbs (p > 0.05). In the present study, although no statistically significant difference was found between the morphometric values of the right and left tibia and fibula of different sexes (p > 0.05), it has been observed that in female cats, the femur right SD value is higher than the left SD value, and in male cats, the femur left Bd value is higher than the right Bd value in a statistically significant manner (p < 0.05). These results are thought to be important for the evaluation of unilateral studies of the pelvic limb long bones in Van cats.

In their study, Boonsri et al. [5] report that the GL values of the femur, tibia and fibula of cats with the dolichocephalic and mesaticephalic skulls are 108.95 ± 5.14 and 95.37 ± 12.39 mm, 110.12 ± 5.39 mm and 96.88 ± 12.69 mm, and 101.75 ± 6.01 and 84.53 ± 17.50 mm, respectively. In the same study, the measurements of these bones belonging to male and female domestic cats were reported to be 103.64 ± 12.80 mm and 101.22 ± 10.74 mm, 105.65 ± 12.48 mm and 102.07 ± 10.57 mm, and 94.79 ± 17.13 mm and 93.21 ± 12.07 mm, respectively. In Turkish Van cats, these values in male and female cats were, respectively, 108.84 ± 4.93 mm and 103.31 ± 2.93 mm, 113.42 ± 5.16 mm and 108.00 ± 3.02 mm, and 105.38 ± 5.65 mm and 100.72 ± 2.48 mm. It is thought that in general, the marginal differences among these values that are otherwise in compliance with literature, and the difference can be attributed to factors such as age, length, body weight, breed and measuring techniques. In addition, according to these values, the Van cats are similar to cats with a dolichocephalic skull shape in terms of pelvic limb long bone dimensions.

Studies show that there is a close relation between age and body weight with the evaluation of long bones of the hind legs; or the development of these bones, especially from a young age [2, 7, 20]. It has also been found that there is a generally positive correlation between the morphometric measuring parameters of these bones [5]. Our study shows, regarding the properties of the femur based on sex, that there is a significant positive relationship between the age and body mass (W) with right GL, left GL, left GLc, left Dc, and right SD values of male cats, as well as the age and left Bd values of female cats (p < 0.05). On the other hand, while there is a positive correlation between body mass and morphometric parameters, no significant relationship could be found (p > 0.05). Regarding the measurements of tibia based on sex, while a positive relationship between the age and body mass with right GL1, left GL1, right Bp, left Bp, right SD, and left SD values in male cats was observed, no statistically significant relationship between age and body mass and these values were noted in female cats (p > 0.05). Regarding fibula measurements and sex, although there was a positive relationship between age and body mass (W) with right GL2 and left GL2 in male cats; there was only a positive relationship of 95.2% in right GL2 and left GL2 values in female cats (p < 0.01). The correlation between the measurements of femur, tibia, and fibula of male and
female cats are provided in the Tables 3, 5, and 7. In general, we can interpret that there is a positive correlation between age and body mass with the measurement values of the long bones in the pelvic limbs in Van cats.

Thanks to the recent developments in CT and 3D reconstruction software, it has become possible to obtain images of any desired width from an anatomical section without any physical harm to the animal, and the morphometric, volumetric and surface area measurements can now be obtained in a fast and efficient way. In this manner, the efficacy of the diagnosis and treatment of various pathological conditions affecting the anatomical area of interest can also be evaluated [21, 26, 46]. A literature survey has shown that, while the volumetric and surface area measurements of the long bones of pelvic limbs belonging to humans [21], brown bears [13], chinchillas [33], guinea pigs [11], and gazelles [12] have been obtained using CT and 3D imaging software, no information regarding the long bones of pelvic limbs of cats could be found in literature in that regard. The mean volumetric values of the femur were 8.57 ± 1.33 cm³ in males and 7.00 ± 0.49 cm³ in females; while their mean surface areas were 64.28 ± 6.72 cm² in males and 59.42 ± 4.72 cm² in females. The mean volumetric values of the tibia were 7.56 ± 1.28 cm³ in males and 6.15 ± 0.52 cm³ in females; while their mean surface areas were 56.89 ± 6.47 cm² in males and 52.72 ± 3.15 cm² in females. The mean volumetric values of the fibula were 1.10 ± 0.17 cm³ in males and 0.83 ± 0.05 cm³ in females; while their mean surface areas were 14.18 ± 1.61 cm² in males and 11.95 ± 0.48 cm² in females. The average volume and surface area measurement values in pelvic limb long bones are higher in male Van cats. It is thought that this difference detected between the measurement values of these bones may be related to body mass.

CONCLUSIONS

In conclusion, the sex-based differences of the measured parameters of the femur, tibia and fibula belonging to adult Van cats were detected. We believe that the information gathered in this study can be utilised in determining morphological variations and the taxonomy of different lineages of Van cats. Moreover, this study has hopefully uncovered information that will be beneficial to the clinical studies of veterinarians that seek to treat Van cats’ pathological conditions of the osteological structure, and for zooarchaeologists.

REFERENCES

1. Ajayi IE, Shawulu JC, Zachariya TS, et al. Osteomorphometry of the bones of the thigh, crus and foot in the New Zealand white rabbit (Oryctolagus cuniculus). Ital J Anat Embryol. 2012; 117(3): 125–134, indexed in Pubmed: 23420943.
2. Alpak H, Onar V, Mutuş R. The relationship between morphometric and long bone measurements of the Mokraran sheep. Turk J Vet Anim Sci. 2009; 33(3): 199–207, doi: 10.3906/vet-0709-23.
3. Atalar O, Özdemir D. Macro-anatomical investigations on the skeletons of marten (Martes foina). II. Ossa Membri Pelvini. Fırat Univ J Health Sci. 2002; 16: 233–236.
4. Bahadir A, Yildiz H. Veteriner Anatomi: Hareket Sistemi & İç Organlar. Ezgi Kitabevi Bursa, Turkey 2014.
5. Boonsri B, Pitakarnnop T, Buddhachat K, et al. Can feline (Felis catus) flat and long bone morphometry predict sex or skull shape? Anat Sci Int. 2019; 94(3): 245–256, doi: 10.1007/s12565-019-00480-8, indexed in Pubmed: 30771106.
6. Cak B. Turkish Van Cat and Turkish Angora Cat: A Review. J Agric Sci Technol A. 2017; 7(3): 151–159, doi: 10.17265/2161-6256/2017.03.002.
7. Campbell KE, Marcus L. The relationships of hindlimb bone dimensions to body weight in birds. Nat Hist Mus Los Angeles Co Sci Ser. 1992; 36: 395–412.
8. Carew RM, Morgan RM, Rando C. A preliminary investigation into the accuracy of 3D modeling and 3D printing in forensic anthropology evidence reconstruction. J Forensic Sci. 2019; 64(2): 342–352, doi: 10.1111/1556-4029.13917, indexed in Pubmed: 30296344.
9. Casteleyen C, Bakker J, Breugelmans S, et al. Anatomical description and morphometry of the skeleton of the common marmoset (Callithrix jacchus). Lab Anim. 2012; 46(2): 152–163, doi: 10.1258/la.2012.011157, indexed in Pubmed: 22517992.
10. Çevik-Demirkan A, Özdemir V, Türkmenoğlu İ, et al. Anatomical and morphometric measurements of the limb skeleton of the chinchilla (Chinchilla lanigera). Acta Veterinaria Brno. 2007; 76(4): 501–507, doi: 10.2754/avb200776040501.
11. Dayan MO, Beşoluk K, Eken E, et al. Three-dimensional modelling of the femur and humerus in adult male guinea pigs (guinea pig) with computed tomography and some biometric measurement values. Folia Morphol. 2019; 78(3): 588–594, doi: 10.5603/FM.a2019.0002, indexed in Pubmed: 30644082.
12. Demircioğlu I, Gezer Ince N. Three-dimensional modelling of computed tomography images of limb bones in gazelles (Gazella subgutturosa). Anat Histol Embryol. 2020; 49(6): 695–707, doi: 10.1111/ahe.12564, indexed in Pubmed: 32319123.
13. Demircioglu I, Kirbaş Doğan G, Aksünger Karaavcı F, et al. Three-dimensional modelling and morphometric investigation of computed tomography images of brown bear’s (Ursus arctos) ossa cruris (Zeugopodium). Folia Morphol. 2020; 79(4): 811–816, doi: 10.5603/FM.a2019.0125, indexed in Pubmed: 31777947.
14. Dursun N. Veterinary anatomy I. Medisan Publisher, Ankara, Turkey 2008.
15. Dyce KM, Sack WO, Wensing CJG. Textbook of veterinary anatomy, 4th ed. Saunders Elsevier Inc., Missouri, United States 2010: 490–500.
16. El-Ghazali HM, El-Behery EI. Comparative morphological interpretations on the bones of the pelvic limb of New Zea-
land rabbit (Oryctolagus cuniculus) and domestic cat (Felis domestica). J Adv Vet Anim Res. 2018; 5(4): 410–419, doi: 10.5455/javaran.2018.e229, indexed in Pubmed: 31453151.

17. Imai N, Funayama K, Suzuki H, et al. Stature estimation formulae based on bony pelvic dimensions and femoral length. Homo. 2020; 71(2): 111–119, doi: 10.1127/ homo/2020/1116, indexed in Pubmed: 31944200.

18. Kalra MK, Maher MM, Toth TL, et al. Strategies for CT radiation dose optimization. Radiology. 2004; 230(3): 619–628, doi: 10.1148/radiol.2303021726, indexed in Pubmed: 14739312.

19. Karan M. Macro-anatomical study of ossa membr pelvini in the feral pigs (Sus scrofa). Firat Univ Vet J Hlth Sci. 2012; 26: 31–34.

20. Kumaş M, Ayaz D. Age determination and long bone histology in Stellagama stellio (Linnaeus, 1758) (Squamata: Sauria: Agamidae) populations in Turkey. Vertebr Zool. 2012; 26: 31–34.

21. Lee UY, Kim IB, Kwak DS. Sex determination using discriminant analysis of upper and lower extremity bones: New dimensional analysis of the morphological evolution and locomotor behaviour of the carnivor hind limb. BMC Evol Biol. 2014; 14: 129, doi: 10.1186/1471-2148-14-129, indexed in Pubmed: 24927753.

22. Mahakanakrua K, Khanpetch P, Prasitwattanseree S, et al. Stature estimation from long bone lengths in a Thai population. Forensic Sci Int. 2015; 253: 135.e1–135.e4, doi: 10.1016/j.forsciint.2015.05.017, indexed in Pubmed: 26177502.

23. Martin-Serra A, Figueirido P, Palmqvist P. A three-dimensional analysis of the morphological evolution and anatomical and radiographic study of bone of the pelvic limb of endangered patagonian huemul deer (Hippocamelus bisulcus). Int J Morphol. 2020; 38(3): 747–754, doi: 10.4067/s0717-95022020000300747.

24. O. Yılmaz, İ. Demircioğlu, A morphometric study on the hind limb long bones (femur, tibia, fibula) in Van cats (Cats domesticus). J Adv Vet Anim Res. 2018; 5(4): 410–419, doi: 10.10455/javaran.2018.e229, indexed in Pubmed: 31453151.

25. Nomina Anatomica Veterinaria. International committee on veterinary gross anatomical nomenclature (ICVGAN), Published by the Editorial Committee, Hannover, 2017.

26. Ohlerth S, Scharf G. Computed tomography in small animals--basic principles and state of the art applications. Vet J. 2007; 173(2): 254–271, doi: 10.1016/j.tvjl.2005.12.014, indexed in Pubmed: 16516508.

27. Olude MA, Olopade JO, Mustapha OA. Macro-anatomical investigations of the skeletons of the African giant rat (Cricetomys gambianus Waterhouse): Pelvic limb. Eur J Anat. 2009; 13(3): 127–131.

28. Onar V, Belli O. Estimation of shoulder height from long bone measurements on dogs unearthed from the Van-Yen-catepe early Iron Age necropolis in Eastern Anatolia. Rev Med Vet (Toulouse). 2005; 156(1): 53–60, doi: 10.1007/s12555-019-00480-8.

29. Onwuama KT, Ojo SA, Hambolu JO, et al. Macro-anatomical and morphometric studies of the hindlimb of grasscutter (Thryonomyss swinderianus, Temminck-1827). Anat Histol Embryol. 2018; 47(1): 21–27, doi: 10.1111/ ahe.12319, indexed in Pubmed: 29139158.

30. Özgel O, Aykut M. Macroanatomical investigation on the ossa membri pelvini of Anatolian bobcat, Lynx lynx. Pakistan J Zool. 2015; 47(5): 1492–1494.

31. Özdemir D, Atalar O. Macro-anatomical investigations on the skeletons of squirrel (Sciurus vulgaris). I. Ossa Membr Pelvini. Firat Univ J Health Sci. 2003; 17: 151–154.

32. Özdemir D, Karan M. Macroanatomical investigations on the skeletons of badger (Meles meles). II. Ossa membri pelvini. Firat Univ J Health Sci. 2001; 15: 397–400.

33. Özkadi S, Varlık A, Kalaycı İ, et al. Morphometric evaluation of chinchillas (Chinchilla lanigera) femur with different modelling techniques. Kafkas Univ Vet Fak Derg. 2016; 22(6): 945–951, doi: 10.9775/kvfed.2016.15683.

34. Özkân Z. Macro-anatomical investigations on the hind limb skeleton of mole-rat (Spalax leucodon Nordmann). Vet Arhiv. 2002a; 72: 159–166.

35. Özkân Z. Macro-anatomical investigations on the skeletons of hedgehog (Erinaceus europaeus L.) II. Ossa membri pelvini. Vet Arhiv. 2002b; 72: 213–220.

36. Pazzan G, Kahvecioğlu KO. Studies on homotypic variation of forelimb and hindlimb long bones of rabbits. J Fac Vet Med Istanbul Univ. 2009; 35: 23–39.

37. Pazzan G, Kahvecioğlu KO. Studies on homotypic variations of forelimb and hindlimb long bones of guinea pigs. J Fac Vet Med Istanbul Univ. 2013; 39: 20–32.

38. Polly PD. Limbs in Mammalian Evolution. In: Fins into Limbs. Evolution, development and transformation, Chapter 15, Hall BK (ed.). University of Chicago Press, Chicago 2007: 245–268.

39. Prokop M. General principles of MDCT. Eur J Radiol. 2003; 45(Suppl 1): S4–10, doi: 10.1016/s0720-048x(02)00358-3, indexed in Pubmed: 12598021.

40. Saber A, K. B. Some morphological and radiological studies on the pelvic limb skeleton of Lumnoltz Tree-Kangaroo (Dendrolagus lumnoltzii). Journal of Veterinary Anatomy. 2019; 12(1): 19–34, doi: 10.21608/jva.2019.67223.

41. Salinas P, Arenas-Caro A, Núñez-Cook S, et al. Morphometric, anatomic and radiographic study of bone of the pelvic limb of endangered patagonian huemul deer (Hippocamelus bisulcus). Int J Morphol. 2020; 38(3): 747–754, doi: 10.4067/s0717-95022020000300747.

42. Saulsman B, Oxnard CE, Franklin D. Long bone morphometrics for human from non-human discrimination. Forensic Sci Int. 2010; 202(1-3): 110.e1–110.e5, doi: 10.1016/j.forsciint.2010.05.009, indexed in Pubmed: 20837249.

43. Shahar R, Milgram J. Morphometric and anatomical study of the hind limb of a dog. Am J Vet Res. 2001; 62(6): 928–933, doi: 10.2460/ajvr.2001.62.928, indexed in Pubmed: 11400852.

44. Von Den Driesch A. A guide to the measurement of animal bones from archaeological sites. Peabody Museum Bulletin 1. Harvard University, Cambridge, MA 1976.

45. Wisner ER, Zwingenberger AL. Atlas of small animal CT and MRI. Willey-Blackwell Publishing, USA 2015: 40–68.

46. Yilmaz O, Soyguder Z, Yavuz A, et al. Three-dimensional computed tomographic examination of pelvic cavity in Van Cats and its morphometric investigation. Anat Histol Embryol. 2020; 49(1): 60–66, doi: 10.1111/ahx.12484, indexed in Pubmed: 31486129.

47. Yilmaz O, Soyguder Z, Yavuz A. Three-dimensional investigation by computed tomography of the clavicle and scapula in van cats. Van Vet J. 2020; 31(1): 34–41, doi: 10.36483/vanvetj.644080.

48. Yilmaz O. Three-dimensional investigation by computed tomography of the forelimb skeleton in van cats. Van Yuzuncu Yil University, Institute of Health Sciences, Faculty of Veterinary, Department of Anatomy, Van, Turkey 2018.

49. Yilmaz S, Dinç G, Aydin A. Macro-Anatomical Investigations on the Skeletons of Porcupine (Hystrix Cristata) II. Ossa Membri Pelvini. Turk J Vet Anim Sci. 1999; 23(3): 297–300.