Salt-inducible kinase 2 regulates fibrosis during bleomycin-induced lung injury

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Idiopathic pulmonary fibrosis is a progressive and normally fatal disease with limited treatment options. The tyrosine kinase inhibitor nintedanib has recently been approved for the treatment of idiopathic pulmonary fibrosis, and its effectiveness has been linked to its ability to inhibit a number of receptor tyrosine kinases including the platelet-derived growth factor, vascular endothelial growth factor, and fibroblast growth factor receptors. We show here that nintedanib also inhibits salt-inducible kinase 2 (SIK2), with a similar IC50 to its reported tyrosine kinase targets. Nintedanib also inhibited the related kinases SIK1 and SIK3, although with 12-fold and 72-fold higher IC50s, respectively. To investigate if the inhibition of SIK2 may contribute to the effectiveness of nintedanib in treating lung fibrosis, mice with kinaseinactive knockin mutations were tested using a model of bleomycin-induced lung fibrosis. We found that loss of SIK2 activity protects against bleomycin-induced fibrosis, as judged by collagen deposition and histological scoring. Loss of both SIK1 and SIK2 activity had a similar effect to loss of SIK2 activity. Total SIK3 knockout mice have a developmental phenotype making them unsuitable for analysis in this model; however, we determined that conditional knockout of SIK3 in the immune system did not affect bleomycin-induced lung fibrosis. Together, these results suggest that SIK2 is a potential drug target for the treatment of lung fibrosis.

Pulmonary fibrosis is a serious condition characterized by scarring of the lungs because of excessive extracellular matrix deposition that occurs over time, leading to an irreversible reduction in lung function. It has been associated with several risk factors including exposure to silica or asbestos dust, infection, autoimmune disease, ionizing radiation, and some chemotherapy drugs. Idiopathic pulmonary fibrosis (IPF) is a form of pulmonary fibrosis for which there is no clear cause and is estimated to account for approximately 20% of interstitial lung disease (reviewed in Refs. (1–4)). The global prevalence of IPF has recently been reported to be between 0.33 and 4.51 cases per 10,000 (1), and the median age at diagnosis is around 65 years old (2, 3, 5). Disease progression is variable, and the reported median survival times from diagnosis range from 2 to 4 years. Treatments for IPF are limited but include pulmonary rehabilitation, supplemental oxygen, and two disease-modifying drugs, nintedanib and pirfenidone. These may slow disease progression but are not curative, and so improved therapies are needed (2, 3, 5).

The molecular mechanisms underlying the development of IPF are complex and not fully understood. It is proposed that repeated microinjuries to the epithelia results in an excessive tissue repair response that leads to the formation of fibroblastic loci and the differentiation of myofibroblasts that result in excess deposition of extracellular matrix (2, 3, 5). This process is accompanied by inflammation; however, it has been debated if inflammation is critically involved in driving disease pathology or if it is a secondary phenomenon. Classical immunosuppressive therapies have proven to be ineffective in IPF; however, this does not exclude a more nuanced role for distinct subsets of immune cells (4). One immune cell that has been proposed to play a role is the macrophage. Macrophages are plastic cells, and their roles vary depending on context; early in inflammation, macrophages adopt a proinflammatory phenotype, sometimes termed “M1” or classically activated macrophages. In contrast, during resolution or tissue repair, macrophages can adopt an anti-inflammatory and potentially profibrotic phenotype. This was originally described in response to activation by the Th2 cytokine interleukin-4 (IL-4) and termed “M2a” or alternatively activated macrophages. This “M2a”-like activation has been linked to fibrosis in different pathologies (6, 7), including IPF (8, 9).

At present, there is no single animal model that fully recapitulates all the features of IPF. However, several models, including bleomycin-induced fibrosis, have been used to evaluate potential targets and therapies in a preclinical setting (10). Bleomycin, an antibiotic made by Streptococcus verticillus, has been used as a chemotherapeutic agent in several
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conditions, including head and neck squamous cell carcinomas, testicular carcinomas, ovarian cancer, and lymphomas (11). Its clinical use is however limited by lung toxicity, and it can cause fibrosis reminiscent of that found in interstitial lung diseases including IPF; this observation led to its use in rodents as a lung fibrosis model (12). Multiple studies have implicated a role for macrophages in bleomycin-induced lung injury. For example, depletion of macrophages with clodronate reduces bleomycin-induced lung fibrosis (8, 13), whereas macrophage colony-stimulating factor 1 (M-CSF1) knockout mice (which have greatly reduced macrophage numbers) show a similar protection (14). More recently, it has been suggested that the differentiation of monocye-derived alveolar macrophages, rather than pre-existing tissue resident alveolar macrophages, is required for bleomycin-induced fibrosis (15, 16). Knockout of proteins that promote an M2/IL-4-like activation phenotype has been reported to protect against bleomycin-induced fibrosis and reduce expression of markers associated with IL-4-induced activation of macrophages in the lung (17–21). Furthermore, myeloid-specific knockout of the phosphatases PTEN and Shp2 resulted in increased fibrosis and promoted an M2/IL-4-like activation state in macrophages (22, 23).

Salt-inducible kinases (SIKs) form a subgroup of the calmodulin-dependent kinase subfamily of protein kinases, and three isoforms, SIK1, SIK2, and SIK3, exist in mammalian cells (24–26). One of the main physiological functions ascribed to SIKs so far is the regulation of cAMP response element-binding protein (CREB)–dependent gene transcription. SIKs phosphorylate the CREB-regulated transcriptional coactivators (CRTC), resulting in their binding to 14-3-3 proteins and retention in the cytoplasm (24, 25, 27, 28). Inhibition of SIKs results in the dephosphorylation of CRTC, allowing their binding to the transcription factor CREB, promoting transcription of CREB-dependent immediate early genes. In innate immune cells, such as macrophages, CREB is required for the efficient transcription of the anti-inflammatory cytokine IL-10 (29, 30). In line with this, inhibition of SIKs promotes IL-10 production in macrophages. In addition, SIKs repress the production of proinflammatory cytokines, such as tumor necrosis factor (TNF) (31, 32). SIK2 and SIK3 are the major SIK isoforms expressed in macrophages (33). Macrophages with a kinase-inactive knockin mutation in SIK2 or SIK3 produced elevated levels of IL-10 and lower levels of proinflammatory cytokines in response to Toll-like receptor agonists compared with wildtype macrophages. SIK-inactive knockin macrophages also showed elevated levels of mRNA encoding Sphk1, Arg1, and Light, genes associated with an “M2b-like” proresolution phenotype in macrophages (33).

The shift in macrophages to a proresolution phenotype in SIK knockin mice suggests that SIK inhibitors may act as immunomodulatory drugs. Interestingly, several clinically approved tyrosine kinase inhibitors including dasatinib and bosutinib, have been reported to also inhibit SIKs (34). Tyrosine kinase inhibitors have attracted attention as antifibrotic treatments with several showing protective effects in animal models of lung fibrosis (reviewed in Refs. (35, 36)). One of these, nintedanib, was successful in phase 3 trials for IPF (37) and gained Food and Drug Administration approval for this disease in 2014 (38, 39). The finding that dasatinib protects against bleomycin-induced lung fibrosis in mice (40) together with its ability to inhibit SIKs and the immune-modulatory functions of SIKs led us to investigate the role of SIKs in lung fibrosis. We show here that loss of SIK2 activity protects against bleomycin-induced lung fibrosis in mice, and that the clinically approved IPF drug nintedanib is a potent inhibitor of SIK2.

Results

Nintedanib inhibits SIK2 in vitro and in cells

Nintedanib was developed as a tyrosine kinase inhibitor and has been approved for the treatment of IPF (38, 39). Nintedanib has been suggested to be effective in IPF because it inhibits platelet-derived growth factor receptor (PDGFR), vascular endothelial growth factor receptor (VEGFR), and fibroblast growth factor receptor (FGFR), but it also inhibits some nonreceptor tyrosine kinases, such as Src, Yes, and Lck (41). Like these tyrosine kinases, members of the SIK subfamily of protein kinases contain an amino acid with a small side chain (threonine) at the gatekeeper site of the ATP-binding pocket, and for this reason, several tyrosine kinase inhibitors are also potent SIK inhibitors (34). We therefore investigated whether nintedanib also inhibits SIKs. Initially, nintedanib was profiled at 0.1 and 1 μM against a panel of 140 kinases, which included SIK2 and SIK3 (Table S1). Analysis of the tyrosine kinases in the panel showed that, as expected, nintedanib inhibited PDGFR, VEGFR, and FGFR as well as the Src family kinases Src, Yes, and Lck (Fig. 1A and Table S1). Nintedanib also inhibited SIK2 to a similar degree as its known tyrosine kinase target. To confirm this result, the IC₅₀ values for inhibition of SIK1, SIK2, and SIK3 were determined (Fig. 1, B–D). Nintedanib inhibited SIK2 with an IC₅₀ of 25 nM (95% confidence interval [CI]: 19–37 nM). This compares favorably with its established targets for which the published IC₅₀ values range from 13 to 108 nM (41). In contrast, nintedanib was less effective against SIK1 and SIK3 with IC₅₀ values of 308 nM (95% CI: 221–655 nM) for SIK1 and 1824 nM (95% CI: 732–3953 nM) for SIK3. As loss of SIK2 activity in macrophages is known to increase IL-10 production in response to lipopolysaccharide (LPS) (33), we next tested if nintedanib could also increase IL-10 production. Nintedanib increased IL-10 production by approximately threefold in LPS-stimulated bone marrow–derived macrophages (BMDMs) with a maximal effect seen between 0.3 and 1 μM (Fig. 1E). The increase in IL-10 production was however not as great as seen with either of the pan-SIK inhibitors, HG-9-91-01 and MRT199665 (Fig. 1E). Genetic studies have shown that there is compensation between different SIK isoforms in macrophages, and that loss of the activity of all three SIKs is required for the maximal effect on IL-10 (33). Loss of SIK2 activity has also been reported to reduce LPS-stimulated TNF production (33). Consistent with the effects of nintedanib on IL-10, nintedanib also reduced TNF production by macrophages with the
Maximal effect reached between 0.3 and 1 μM and an EC50 of 0.15 μM (Fig. 1). For comparison, published EC50 values for the inhibition of proliferation of VEGFR-, PDGFR-, or FGFR-dependent cell lines range from 0.007 to 0.3 μM (41, 42). To confirm that nintedanib was affecting IL-10 and TNF production primarily via the inhibition of SIK2, the effects of 3 μM nintedanib treatment were compared in wildtype macrophages and macrophages from knockin mice in which SIK2 was replaced by a kinase-inactive mutant. These experiments showed that nintedanib did not affect IL-10 or TNF secretion in SIK2 knockin macrophages and that the levels of IL-10 and TNF secretion were similar in the nintedanib-treated macrophages from wildtype mice and non-inhibitor treated macrophages from SIK2 knockin mice following LPS stimulation (Fig. S1). For some kinases, mutation of the gatekeeper site in the ATP pocket to a larger residue is able to block the binding of some ATP competitive inhibitors without reducing kinase activity. An example of this is mutation of Thr96 to glutamine.

Figure 1. Nintedanib inhibits SIK2 in vitro and in cells. A, nintedanib was screened against a panel of 140 kinases at 0.1 μM. The percent inhibition for the top 39 kinases is shown. Results for all 140 kinases are given in Table S1. B–D, nintedanib was tested for its ability to inhibit SIK1 (B), SIK2 (C), and SIK3 (D) in vitro at a range of concentrations from 3 nM to 100 μM. IC50 values were calculated as described in the Experimental procedures section. E, as (E) but levels of TNF were measured following 6 h of LPS stimulation (one-way ANOVA, F = 81.3, p < 0.001).

BMDM, bone marrow-derived macrophage; DMSO, dimethyl sulfoxide; IL, interleukin; LPS, lipopolysaccharide; SIK, salt-inducible kinase; TNF, tumor necrosis factor.
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in SIK2, which has previously been found to prevent the binding of HG-9-91-01 (32). In RAW264.7 cells expressing a wildtype version of SIK2, both nintedanib and HG-9-91-01 were able to increase IL-10 secretion following LPS stimulation relative to cells in the absence of inhibitor, similar to what was observed in BMDMs. In contrast, in cells expressing a T96Q mutation of SIK2, IL-10 production was not promoted by either nintedanib or HG-9-91-01 (Fig. 1G). Together, these results suggest that nintedanib selectively inhibits SIK2 in macrophages and not SIK1 and SIK3 when used at 3 μM.

Loss of SIK2 activity does not prevent the expression of IL-4-induced markers in BMDMs

Given that lung fibrosis in mice has been linked to increased polarization of macrophages to an IL-4-induced "M2a" phenotype, the effect of the SIK2 kinase-inactive knockin was tested on IL-4 and LPS-induced markers on macrophages. LPS induces the expression of CD64, which has been used as a marker of inflammatory M1 macrophages (43). Analysis of CD64 expression by flow cytometry showed that CD64 was expressed on BMDMs prior to stimulation and was upregulated following LPS stimulation (Fig. 2A). While SIK2 knockin decreases proinflammatory cytokine production in response to LPS, unexpectedly, it increased CD64 expression relative to wildtype macrophages following LPS stimulation (Fig. 2A). IL-4 is known to induce the expression of CD206 and CD301a in macrophages (44, 45). CD206 expression was low in wildtype BMDMs but was increased following IL-4 stimulation (Fig. 2B). SIK2 knockin resulted in a higher expression of CD206 relative to wildtype cells, both in unstimulated cells and following IL-4 stimulation (Fig. 2B). CD301a expression was also increased by IL-4 treatment in wildtype BMDMs (Fig. 2C). However, in contrast to CD206 expression, SIK2 knockin resulted in lower CD301a expression compared with wildtype cells, both before and after IL-4 stimulation (Fig. 2C).

As these results might suggest a change in macrophage polarization in vivo, the expression of CD206 was examined in alveolar macrophages. CD206 expression has been reported in alveolar macrophages and found to be increased following short-term bleomycin treatment (46). In line with this, CD206 was detected on CD45<sup>+</sup>/Siglec F<sup>+</sup>/CD11c<sup>+</sup> alveolar macrophages present in both the bronchoalveolar lavage fluid (BALF) (Fig. 3A) and in digests of lung tissue (Fig. 3B). These levels were increased 7 days post-treatment with bleomycin (Fig. 3, C and D) although the numbers of CD45<sup>+</sup>/Siglec F<sup>+</sup>/CD11c<sup>+</sup> macrophages that could be detected in either the BALF or in lung digests decreased following bleomycin treatment (Fig. 3, E and F). In contrast to BMDMs, the level of CD206 in alveolar macrophages was similar between wildtype and SIK1/2 knockin cells. The loss of SIK1 and SIK2 activity also did not affect the decrease in alveolar macrophage numbers seen following bleomycin treatment. While CD206 was detected on alveolar macrophages, it was only present at very low levels on CD45<sup>+</sup>/CD11c<sup>+</sup>/CD11b<sup>+</sup>/Ly6G<sup>+</sup> or CD45<sup>+</sup>/CD11c<sup>+</sup>/CD11b<sup>+</sup>/Ly6G<sup>+</sup> myeloid cells in the lungs of mice, and these levels were not affected by SIK1/2 knockin (Fig. S2).

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Bleomycin administration in the lungs of mice results in an initial inflammatory phase lasting 7 to 10 days that then resolves into a fibrotic phase. To examine a role for SIK2 in this initial inflammatory phase, mice were sacrificed at 7 and 10 days following bleomycin treatment, and the presence of immune cells and cytokines in the BALF was determined. TGFβ has been implicated in the development of bleomycin-induced fibrosis (47, 48). Levels of TGFβ were increased in the BALF following bleomycin treatment; however, no significant differences (p > 0.05, two-way ANOVA) were observed between wildtype, SIK2, and SIK1/2 knockin mice (Fig. 4A). IL-22 is another cytokine induced by bleomycin treatment, and knockout of IL-22 or its neutralization using a monoclonal IL-22 antibody reduced bleomycin-induced lung injury (49). Similar to TGFβ, induction of IL-22 was comparable between wildtype, SIK2, and SIK1/2 knockin mice (Fig. 4B). TNF is also increased following bleomycin treatment, and a role for TNF in promoting the resolution of inflammation by stimulating apoptosis in immune cells has been proposed in the bleomycin model (50, 51). The induction of TNF was however not affected by the loss of SIK2 or SIK1/2 activity (Fig. 4C). The levels of granulocyte–macrophage colony-stimulating factor and IL-10, cytokines implicated in limiting inflammation in the bleomycin model (52–54), were also increased following bleomycin treatment, but again, no significant differences were observed between wildtype, SIK2, or SIK1/2 knockin mice (Fig. 4, D and E). In addition to cytokines, several chemokines, including CCL2 and CXCL2, have been reported to be increased in mice following the administration of bleomycin and are required for the induction of bleomycin-induced pathology (14, 55, 56). Consistent with this, CCL2 and CXCL2 were found to be increased in the BALF of bleomycin-treated mice, but again, this was not affected by SIK2 or SIK1/2 knockin (Fig. S3, A and B). Bleomycin was also able to increase the number of CD45-positive immune cells present in the BALF (Fig. 4F). Both the SIK2 and SIK1/2 knockin mice showed a slight increase in immune cell recruitment 10 days after bleomycin treatment (Fig. 4F). While in naïve mice, the cells in the BALF were mostly alveolar macrophages, these decreased following bleomycin treatment, whereas a number of other immune cell types, including neutrophils, eosinophils, B cells, and both CD4 and CD8 T cells, were recruited to the BALF (Fig. S3, C–H). The percentages of these different cell types were however similar between the bleomycin-treated wildtype, SIK2, and SIK1/2 knockin mice (Fig. S3, C–H).

As SIK2 did not appear to affect the initial inflammatory phase of bleomycin-induced lung inflammation, we next examined its potential effects on fibrosis using a 22 day model. Following administration of bleomycin, wildtype mice lost weight with peak weight loss occurring 7 to 10 days after the administration of bleomycin, after which mice started to regain weight (Fig. 5). Mice that lost more than 20% of their body weight without showing signs of improvement within 3 days were considered to have reached the humane end point and were sacrificed. Four of the 13 bleomycin-treated wildtype animals reached this point between day 7 and 11 of the protocol.
SIK2 knockin mice showed a lower dropout rate, with one of eight mice reaching the humane end point before the end of the experiment (Fig. 5A). To estimate weight loss over the whole experiment, the area under the curve was calculated for each mouse relative to its starting weight. Using this measure, SIK2 single knockin mice lost significantly less weight ($p < 0.05$) than wildtype mice following bleomycin treatment (Fig. 5B). Consistent with this, SIK2 bleomycin-treated mice showed a lower peak weight loss relative to bleomycin-treated wildtype animals (Fig. 5C). Similar results were also obtained when SIK1/2 double knockin mice were analyzed, with the SIK1/2 knockin animals showing increased survival (Fig. 5D) as well as decreased weight loss (Fig. 5, E and F). The extent of fibrosis at day 22 was assessed by histological scoring of Masson’s trichrome-stained lung sections. As expected, extensive collagen deposition throughout the alveolar bed (stained blue in Fig. 6A) was apparent in the lungs from bleomycin-treated wildtype mice. Both the SIK2 (Fig. 6, A and B) and SIK1/2 (Fig. 6, A and C) knockin mice showed reduced levels of collagen deposition compared with the wildtype animals.

Figure 2. Effect of SIK2 knockin (Ki) on macrophage marker expression. BMDMs were generated from wildtype (blue circles) or SIK2 Ki (orange diamonds) mice. Cells were stimulated for 24 h with either 100 ng/ml LPS or 10 ng/ml IL-4. Cells were then stained for F4/80, CD301a, and either CD206 or CD64. Live F4/80+ve cells were gated, and the mean fluorescence intensity (MFI) for CD64 (A), CD206 (B), or CD301a (C) was determined. Graphs show mean and standard deviation with symbols representing individual mice. A $p$ value of less than 0.05 is indicated by *, less than 0.01 by **, and less than 0.001 by *** (two-way ANOVA followed by Sidak’s post hoc testing; $F$ and $p$ values for the ANOVAs are shown in Table S3). BMDM, bone marrow–derived macrophage; IL, interleukin; LPS, lipopolysaccharide; SIK, salt-inducible kinase.
Lung sections were also stained with H&E to assess inflammation and type II pneumocyte hypertrophy/hyperplasia (Fig. 6D). In addition to the reduction in fibrotic score relative to wildtype animals, at day 22, the lungs from bleomycin-treated SIK2 and SIK1/2 knockin mice showed lower inflammation scores than bleomycin-treated wildtype mice (Fig. 6, E and F). Type II pneumocyte hypertrophy/hyperplasia was also decreased in SIK2 knockin mice relative to wildtype (Fig. 6G); however, this change was not significant in SIK1/2 knockin mice (Fig. 6H, p > 0.05).

Figure 3. Expression of CD206 in wildtype and SIK1/2 knockin (Ki) alveolar macrophages. Wildtype and SIK1/2 Ki mice were given an oropharyngeal dose of 1.5 mg/kg bleomycin or an equivalent volume of PBS. Mice were culled on day 7, and cells in the BALF (A, C, and E) and lung digests (B, D, and F) were stained with CD45, CD11c, Siglec F, CD206, F4/80, Ly6G, and CD11b and analyzed by flow cytometry. Representative histograms for CD206 expression in CD45+ve/Siglec F+ve/CD11c+ve alveolar macrophages are shown in (A and B), and mean fluorescent intensity for CD206 in alveolar macrophages is shown in (C and D). The number of alveolar macrophages as a percentage of live cells is shown in (E and F). Four to 5 mice were analyzed per group, and graphs show mean and standard deviation with individual mice shown by symbols. Data were analyzed by two-way ANOVA with Sidak’s post hoc testing to compare between PBS and bleomycin (black) or wildtype and SIK1/2 Ki genotypes (purple). p < 0.05, 0.01, and 0.001 is indicated by *, **, and ***, respectively; ns indicates p > 0.05. F and p values for the ANOVAs are shown in Table S4. BALF, bronchoalveolar lavage fluid; SIK, salt-inducible kinase.
As well as SIK2, immune cells also express SIK3. Both SIK3 knockout and SIK3 kinase inactive knockin mice exhibit increased postnatal mortality and growth retardation (33, 57), making them poorly suited for the analysis of SIK3 in lung fibrosis. To address the role of SIK3 in immune cells in bleomycin-induced fibrosis, we generated a hemopoietic cell-specific knockout of SIK3 by crossing floxed SIK3 mice to a Vav-iCre transgene (58). Following bleomycin treatment, survival was higher in the SIK3 knockout (8 of 10) relative to the wildtype group (6 of 12, Fig. 7A) although this did not reach significance for the group size analyzed (p = 0.133, log-rank Mantel–Cox test). Despite this, no difference in weight loss was observed between wildtype and the immune cell-specific SIK3 knockout mice, either when individual time points (Fig. 7B) or the area under the curve (Fig. 7C) were compared. Histological analysis of the lungs at day 20 did not show significant differences between the wildtype and SIK3 knockout in the extent of fibrosis or inflammation (Fig. 7, D and E).

Discussion

IPF is a fatal condition that remains difficult to treat; however, the approval of nintedanib in 2014 was a significant advance in the management of this condition (39). Since then, nintedanib has also shown positive results in phase 3 trials for interstitial lung disease associated with systemic sclerosis (59, 60) and progressive fibrosing interstitial lung diseases (61, 62) and has been approved for these conditions.

Nintedanib is a tyrosine kinase inhibitor, and its success in IPF has been attributed to the inhibition of several tyrosine kinase receptors, including the VEGFR, PDGFR, and FGFR (36, 38, 41). It does however inhibit several other kinases that,
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Figure 5. Bleomycin (bleo)-induced weight change in wildtype, SIK2, and SIK1/2 knockin (Ki) mice. Mice were given an oropharyngeal dose of 2 mg/kg of bleo or an equivalent volume of PBS. Weight was monitored daily for 22 days. Mice that lost more than 20% of their body weight without showing signs of improvement within 3 days were considered to have reached the humane end point and were sacrificed at that point. A–C, wildtype and SIK2 Ki mice were treated with either PBS or bleo. The percentage survival is shown in (A). The area above or below the curve relative to the starting weight is shown in (B). Data show average values for each group, and error bars represent SEM, with initial group sizes of four and two for the PBS-treated wildtype and SIK2 Ki mice, respectively. D–F, as (A–C), except female wildtype and SIK1/2 double Ki mice were analyzed, with survival shown in (D), area under the curve in (E), and weight changes in (F). Initial group sizes were four for wildtype PBS-treated mice, two for SIK1/2 Ki PBS-treated mice, seven for wildtype bleomycin-treated mice, and nine for the SIK1/2 Ki bleomycin-treated group. For (B and D), p < 0.05 and p < 0.001 are shown by * and ** (two-way ANOVA and Tukey's post hoc testing). For (C and F), data were analyzed using a mixed-effects model and Tukey's post hoc testing. For (E and F), p < 0.001 are shown by * and *** (two-way ANOVA and Tukey's post hoc testing). For comparisons on individual days between wildtype and SIK2 or SIK1/2 Ki bleomycin-treated mice, * indicates p < 0.05, ** indicates p < 0.01, and *** indicates p < 0.001. Full statistical analysis is shown in Table S6. SIK, salt-inducible kinase.
therefore be required to determine the targets of SIK2 during the development of lung fibrosis. Bleomycin-induced fibrosis has been associated with an increase in expression of markers of IL-4-induced activation in macrophages (17–21). While this may be important in fibrosis, it is not straightforward to show a direct link between macrophage polarization and fibrosis. Transcriptomic profiling has shown that macrophages can adopt multiple heterogenous states \textit{ex vivo} depending on the signaling inputs they receive (66). With relation to bleomycin-induced fibrosis, transcriptional analysis of monocyte-derived fibrosis, transcriptional analysis of monocyte-derived and tissue resident alveolar macrophages show they both upregulate a subset of M1- and M2-associated genes, with no clear indication of an overall change in the M1/M2 ratio (16).

While loss of SIK2 activity decreases the production of inflammatory cytokines in macrophages (33), it had contrasting effects on markers for IL-4 stimulation in BMDMs, with CD206 expression being elevated but CD301a expression slightly decreased (Fig. 2). Of note, in BMDMs, the effect of SIK2 inhibition on CD206 expression was reduced following IL-4 stimulation, suggesting it is dependent on the cytokine environment of the macrophages. In contrast to BMDMs, loss of SIK2 activity did not affect CD206 expression in alveolar macrophages. This could reflect the different cytokine environments seen between cultured macrophages and the cells in the lung or the different origins of the cells; BMDMs are derived \textit{ex vivo} from bone marrow precursors stimulated with...
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Figure 7. Bleomycin-induced fibrosis in wildtype and immune cell–specific SIK3 knockout (Ko) mice. Wildtype Vav-iCre+/− or immune cell–specific SIK3+/−/Vav-iCre+/− Ko mice were given an oropharyngeal dose of 1.5 mg/kg of bleomycin or a PBS control. Weight was monitored daily for 20 days. Mice that lost more than 20% of their body weight without showing signs of improvement in 3 days were considered to have reached the humane end point and sacrificed. The percentage survival is shown in (A). The average percentage weight change relative to the weight on day 0 before bleomycin treatment is shown in (B). Error bars represent SEM, with initial group sizes of three for the PBS-treated wildtype and immune cell–specific SIK3 knockout mice and 12 (six males/six females) and 10 (six males/four females) for the bleomycin-treated wildtype and immune cell–specific SIK3 knockout mice, respectively. Mixed-effects model analysis showed no significant difference between wildtype and SIK3 knockout bleomycin-treated mice (p > 0.05). The area above or below the curve relative to the starting weight is shown in (C). Positive values indicate increases in weight, and negative values indicate decreases in weight. Average and standard deviation are shown with symbols representing individual mice. Gray symbols indicate mice that did not reach the 20 day end point. p < 0.01 is indicated by ** (two-way ANOVA and Sidak’s post hoc testing). On day 20, mice were sacrificed, and lungs were fixed and stained with either H&E or Masson’s trichrome as described in the Experimental procedures section. Scoring for fibrosis is shown in (E), and scoring for inflammation is shown in (D). Graphs show mean and standard deviation with symbols showing individual mice. Two-way ANOVA testing did not identify significant (p > 0.05) differences between the wildtype and immune cell–specific SIK3 knockout mice. F and p values for the mixed-model analysis and ANOVAs are shown in Table S8. SIK, salt-inducible kinase.

M-CSF, whereas alveolar macrophages in naïve mice are seeded in embryonic and early postnatal development and require granulocyte–macrophage colony-stimulating factor (67, 68).

Both CD206 and CD301a are members of the C-type lectin receptor family. While increases in CD206 expression on macrophages following bleomycin treatment is a useful marker for macrophage activation, it is less clear whether it plays an active role in the development of pathology. A recent report suggests that peptides targeting CD206 are protective in bleomycin-induced fibrosis by promoting cell death in CD206+/− macrophages (69). Nanoparticles targeting TGFβ siRNA to CD206+/− macrophages can also reduce bleomycin-induced fibrosis (15). While these findings support a role for CD206+/− macrophages, it does not address if CD206 is directly important in the response to bleomycin or if it only acts as a marker for alveolar macrophages that allows them to be targeted in vivo. The role of CD301a has not been addressed in bleomycin-induced fibrosis, although knockout of CD301a has been found to reduce tissue remodeling in antigen-induced granuloma formation (70). While SIKs are able to inhibit the inflammatory functions of macrophages in culture, it is not clear if this is sufficient to explain their roles in lung fibrosis. Loss of SIK2 activity did not reduce the initial inflammatory phase following bleomycin treatment and did not promote IL-10 production in this model in vivo. This contrasts with multiple studies showing that SIK inhibition in isolated macrophages promotes IL-10 production via promoting CREB activity (31–33, 64). This could be due to the bleomycin-induced IL-10 in the lung being derived from a different cellular source, such as T cells or B cells where CREB does not seem to be critical for its production (29). It should also be noted that for SIK inhibition in cultured macrophages to have its maximal effect on either IL-10 or the phosphorylation of the CREB coactivator CRTC3, all three SIK isoforms have to be inhibited. In contrast to this, SIK2 appears to be the predominant isoform regulating lung fibrosis, although a contribution from SIK3 cannot be ruled out. Together, this suggests that during bleomycin-induced lung fibrosis, the main effects of SIK2 inhibition may not be due to its anti-inflammatory role in macrophages, and that it could potentially be acting via a novel substrate.
SIK2 appears to be ubiquitously expressed (33), and as the knockin mice used were not conditional, it is not possible to say in what cell types SIK2 functions during the development of fibrosis. Further work will be needed to resolve these issues.

Together, the data presented here identify SIK2 as a target of a clinically approved antifibrotic drug and show that the SIK2 kinase-inactive knockin mouse is protected in a bleomycin-induced model for lung fibrosis. While the efficacy of nintedanib was originally proposed to be due to its ability to inhibit tyrosine kinases, it is possible that at least some of its effects in the clinic are due to its ability to inhibit SIK2.

Experimental procedures

Animals

The generation of SIK1 and SIK2 kinase-inactive knockin mice has been described previously (33). A conditional SIK3 line was obtained from IMPC and crossed to a Vav-iCre transgenic line to generate mice lacking SIK3 in the immune system (58, 71). All animals were maintained on a C57Bl6/J background using mice from Charles River Laboratories. Mice were kept under specific pathogen-free conditions in individually ventilated cages and given free access to food (RM3 irradiated pelleted diet supplied by Special Diet Services) and water. Animal rooms were maintained at a temperature of 21 °C and a humidity of 45 to 55% and 12/12 h light cycle. All animal works were carried out under a UK Home Office license and subject to approval by the University of Dundee’s Welfare and Ethical Use of Animals Committee. Experiments were planned in accordance with the ARRIVE guidelines.

Kinase inhibitors

Nintedanib was purchased from Tocris Bioscience. The SIK inhibitors HG-9-91-01 and MRT199665 were synthesized by Dr Natalia Shpiro and can be obtained from MRC PPU Reagents and Services (https://mrcppureagents.dundee.ac.uk/). LPS (Escherichia coli strain O5:B55) was obtained from Alexis Bioscience, and recombinant murine IL-4 was from Peprotech.

In vitro kinase assays

An in vitro selectivity screen was carried out as described (72). Human SIK1, SIK2, and SIK3 were expressed in S9 insect cells. To determine the IC50 value for SIK1, SIK2, and SIK3, nintedanib was assayed on a 10-point concentration curve from 0.003 to 100 µM. Assays were carried out in a final volume of 25.5 µl containing 50 mM Tris–HCl (pH 7.5), 0.1 mM EGTA, 0.05% (v/v) β-mercaptoethanol, 300 µM substrate peptide (ALNRTSSDSALHRRR), 10 mM magnesium acetate, and 0.05 mM [33P-γ-ATP] (50–1000 cpm/pmol) and incubated for 30 min at room temperature. Assays were started by the addition of ATP and stopped by addition of 5 µl of 0.5 M (3%) orthophosphoric acid and then harvested onto P81 Unifilter plates with a wash buffer of 50 mM orthophosphoric acid. IC50 values were calculated in GraphPad Prism (GraphPad Software, Inc) by nonlinear regression using the model $y = y_{\text{max}} + (y_{\text{max}} - y_{\text{min}})/(1 + (x/IC_{50}))$, where $y_{\text{max}}$ and $y_{\text{min}}$ are the maximum and minimum values for percentage inhibition, respectively.

Macrophage culture

Primary BMDMs were obtained by differentiating bone marrow from 8- to 12-week-old C57Bl6/J mice for 7 days in Dulbecco’s modified Eagle’s medium supplemented with 20% (v/v) L929 conditioned medium as a source of M-CSF, 2 mM L-glutamine, 10% (v/v) fetal calf serum, 1 mM pyruvate, non-essential amino acids, 10 mM Hepes, 50 µM β-mercaptoethanol, 50 U/ml penicillin, and 50 µg/ml streptomycin. Macrophages were differentiated on non–tissue culture–treated plastic, harvested, and replated at a density of 1,000,000 cells/cm² on tissue culture–treated plastic in fresh media prior to stimulation on day 8. Where indicated in the figure legends, cells were incubated with inhibitors for 1 h prior to stimulation.

RAW264.7 cells with stably integrated expression cassettes for wildtype and T96Q SIK2 under the control of a tetracycline-inducible promoter have been described previously (32). Cells were cultured in Dulbecco’s modified Eagle’s medium supplemented with 10% (v/v) fetal bovine serum (FBS), 100 U/ml penicillin, 100 µg/ml streptomycin, 2 mM glutamine, 3 µg/ml puromycin, and 1 mg/ml G418. To induce the expression of SIK2, cells were treated with 1 µg/ml doxycycline in the absence of G418 and puromycin for 16 h before treatment with inhibitors and stimulation with LPS.

Bleomycin-induced lung fibrosis

Mice were anesthetized with isoflurane and received a single dose of bleomycin (1.5 or 2 mg/kg; Sigma) via an oropharyngeal route. Mice were culled at 3, 7, 10, or 22 days following the dosing as indicated in the figure legends or when the humane end point was reached (the humane end point was reached when either mice lost more than 25% of their initial weight, mice remained for more than 3 consecutive days at a weight loss over 20% of their initial weight, when after starting to recover, they stopped recovering and stayed at a stable weight for more than 5 days with weight loss of over 10% or when body condition score reached a value of 1.5). Mice were monitored daily, with body weight measured at a fixed time every morning, starting 3 days before the dosing and continuing until the last day, and body condition was independently assessed by two people (with one scorer blind to the experimental treatment and genotype). For the duration of the experiment, mice received supplementary food in the form of rehydrated mash (same formulation as normal food) and a high fat supplement (peanut butter; Tesco brand).

Mice were culled using an overdose of anesthetic (Euthatal) followed by exsanguination via vena cava and blood collected in Minicollect serum collection tubes (Greiner Bio-One). For analysis of CD206 expression by flow cytometry, mice were culled via lethal dose of anesthetic (medetomidine HCl/ketamine HCl) followed by exsanguination. Lungs were washed three times in succession with 400 µl PBS. Supernatants were collected for BALF measurements, and cells were used for flow cytometry. The left lung was fixed in 4% (w/v)
paraformaldehyde in PBS, and the right lung was divided into two, with half snap frozen in liquid nitrogen and half processed for flow cytometry.

**Histology**

Fixed lung tissue was sectioned twice along the transverse axis, and the resulting three samples were processed to paraffin embedding. Serial sections were mounted and stained with Masson's trichrome and H&E. A multiparametric scoring system (73) was developed to assess microscopic changes in the lung. Fibrosis, inflammation, and type II pneumocyte hypertrophy/hyperplasia were evaluated in 15 microscopic fields (five fields per transverse section of the left pulmonary lobe) and scored with an ordinal grading scale (0–4) developed according to published guidelines and principles for valid histopathologic scoring in research (74). Four to five score levels are considered the optimal range to maximize detection and reproducibility for semiquantitative grading applied to histopathology (74). Scoring of the histological end points was done by a veterinary pathologist blinded to genotype and treatment. The defining criteria of the scoring system are detailed in Table S2.

**Lung digestions**

To isolate single-cell suspensions from lung tissue, individual lungs were finely cut and placed in 1 ml of lung digest mix containing 2 U/ml of Liberase (Sigma–Aldrich) and 160 U/ml of DNaseI (Sigma–Aldrich) in Dulbecco’s PBS enriched with magnesium and calcium (Gibco). Lung digest were incubated at 37 °C, shaking at 200 rpm, for 35 min. The Liberase/DNaseI was then inactivated with RPMI (Gibco) containing 10% (v/v) FBS (Thermo Fisher Scientific). Digested lungs were pushed through a 70 μm cell strainer to generate a single-cell suspension, and red blood cells were lysed using ACK lysing buffer (Gibco). Cells were then washed with RPMI containing 10% (v/v) FBS and resuspended at 1 × 10^7 cells/ml.

**Flow cytometry**

BALF-containing cells were centrifuged at 1500 rpm for 5 min at 4 °C. Supernatants were collected for BALF cytokine measurements, and cells were used for analysis by flow cytometry. Single-cell suspensions, diluted in PBS, were used to analyze total cell counts on BD FACSVerse (BD Biosciences). For analysis, cells were washed with fluorescence-activated cell sorting buffer (1% [w/v] bovine serum albumin in PBS), and Fc receptors were blocked for 15 min at 4 °C using Mouse BD Fc Block (BD Biosciences). For cultured macrophages, adherent cells were collected by incubation for 5 min with 0.48 mM EDTA in PBS on ice and then scraping to form a single-cell suspension. Cells were stained with fixable viability dye eFluor 450 (eBioscience) or Fixable Blue Live/Dead stain (Thermo Fisher Scientific), prior to incubation with Fc block. Cell surface markers were stained, and data were acquired on either BD FACSVerse II or LSR Fortessa (BD Biosciences) flow cytometers using FACSDiva software (BD Biosciences). Analysis was performed using FlowJo (Tree Star, Inc.). Antibodies used for flow cytometry are listed in Table 1, and representative gating strategies are shown in Figs. S4 and S5.

**Cytokine measurements**

Cytokines were measured using Bio-Plex Pro assay system (Bio-Rad Laboratories), Luminex 200 machine, and xPONENT 4.1 software (Luminex Corporation) according to the manufacturer’s protocols.

**Statistical analysis**

Statistical analysis was carried out using GraphPad Prism. For analysis of weight loss, the area under/above the curve was calculated relative to the weight on day 0 using GraphPad Prism. The specific statistical tests used are indicated in the figure legends. For ANOVA, significant differences from post hoc testing are indicated on the figures where appropriate. Details of the F and p values for all ANOVAs are given in the legend (Fig. 1) or supporting tables (Figs. 2–7).

**Data availability**

All data relevant to this work are contained within this article and the associated supporting information.

**Supporting information**—This article contains supporting information.

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alveolar macrophages drive lung fibrosis and persist in the lung over the life span. J. Exp. Med. 214, 2387–2404.
17. Yao, Y., Wang, Y., Zhang, Z., He, L., Zhai, J., Zhang, M., et al. (2016) Chop deficiency protects mice against bleomycin-induced pulmonary fibrosis by attenuating M2 macrophage production. Mol. Ther. 24, 915–925
18. Karmouty-Quintana, H., Philip, K., Acero, L. F., Chen, N. Y., Weng, T., Molina, I. G., et al. (2015) Deletion of ADORA2B from myeloid cells dampens lung fibrosis and pulmonary hypertension. FASEB J. 29, 50–60
19. Larson-Casey, J. L., Deshane, J. S., Ryan, A. J., Thannickal, V. J., and Carter, A. B. (2016) Macrophage Akt1 kinase-mediated mitophagy modulates apoptosis resistance and pulmonary fibrosis. Immunity 44, 582–596
20. Ballinger, M. N., Newstead, M. W., Zeng, X., Bhan, U., Mo, X. M., Kunkel, S. L., et al. (2015) IRAK-M promotes alternative macrophage activation and fibrillogenesis in bleomycin-induced lung injury. J. Immunol. 194, 1894–1904
21. Guo, X., Li, T., Xu, Y., Xu, X., Zhu, Z., Zhang, Y., et al. (2017) Increased levels of Gαb1 and Gαb2 adaptor proteins skew interleukin-4 (IL-4) signaling toward M2 macrophage-driven pulmonary fibrosis in mice. J. Biol. Chem. 292, 14003–14015
22. Tao, B., Jin, W., Xu, J., Liang, Z., Yao, J., Zhang, Y., et al. (2014) Myeloid-specific disruption of tyrosine phosphatase Shp2 promotes alternative activation of macrophages and predisposes mice to pulmonary fibrosis. J. Immunol. 193, 2801–2811
23. Kral, J. B., Kuttke, M., Schrattmaier, W. C., Birnecker, B., Warszawska, J., Wernig, C., et al. (2016) Sustained PI3K Activation exacerbates BLM-induced Lung Fibrosis via activation of pro-inflammatory and pro-fibrotic pathways. Sci. Rep. 6, 23034
24. Sakamoto, K., Bultot, L., and Goransson, O. (2018) The salt-inducible kinases: emerging metabolic regulators. Trends Endocrinol. Metab. 29, 872–840
25. Wein, M. N., Foretz, M., Fisher, D. E., Xavier, R. J., and Kronenberg, H. M. (2018) Salt-inducible kinases: physiology, regulation by cAMP, and therapeutic potential. Trends Endocrinol. Metab. 29, 723–735
26. Darling, N. J., and Cohen, P. (2021) Nuts and bolts of the salt-inducible kinases (SIKs). Biochem. J. 478, 1377–1397
27. Katoh, Y., Takemori, H., Min, I., Muraoka, M., Doi, J., Horike, N., et al. (2004) Salt-inducible kinase-1 represses cAMP response-element-binding protein activity both in the nucleus and in the cytoplasm. Eur. J. Biochem. 271, 4307–4319
28. Scroat, R. A., Conkright, M. D., Katoh, Y., Best, J. L., Canettieri, G., Jeffries, S., et al. (2004) The CREB coactivator TORC2 functions as a calcium- and cAMP-sensitive coincidence detector. Cell 119, 61–74
29. Sutavani, R. V., Phair, I. R., Barker, R., McFarlane, A., Shiipo, N., Lang, S., et al. (2018) Differential control of Toll-like receptor 4-induced interleukin-10 induction in macrophages and B cells reveals a role for p90 ribosomal S6 kinases. J. Biol. Chem. 293, 2302–2317
30. Ananieva, O., Darragh, J., Johansen, C., Carr, J. M., McIlrath, J., Park, J. M., et al. (2008) The kinases MSK1 and MSK2 act as negative regulators of Toll-like receptor signaling. Nat. Immunol. 9, 1028–1036
31. MacKenzie, K. F., Clark, K., Naqi, S., McGuire, V. A., Noehren, G., Kristariyanto, Y., et al. (2013) PGE2 induces macrophage IL-10 production and a regulatory-like phenotype via a protein kinase A-SIK-CRTC3 pathway. J. Immunol. 190, 565–577
32. Clark, K., MacKenzie, K. F., Petkevicius, K., Kristariyanto, Y., Zhang, J., Choi, H. G., et al. (2012) Phosphorylation of CRTC3 by the salt-inducible kinases controls the interconversion of classically activated and regulated macrophages. Proc. Natl. Acad. Sci. U. S. A. 109, 16986–16991
33. Darling, N. J., Toth, R., Arthur, J. S., and Clark, K. (2017) Inhibition of SIK2 and SIK3 during differentiation enhances the anti-inflammatory phenotype of macrophages. Biochem. J. 474, 521–537
34. Ozanne, J., Prescott, A. R., and Clark, K. (2015) The clinically approved drugs dasatinib and bosutinib induce anti-inflammatory macrophages by inhibiting the salt-inducible kinases. Biochem. J. 465, 271–279
35. Iwamoto, N., Distler, J. H., and Distler, O. (2011) Tyrosine kinase inhibitor effects of animal models in IPF research. Pulm. Pharmacol. Ther. 24, 137–148
36. Distler, O., and Distler, J. H. (2011) Tyrosine kinase inhibitor effects of animal models in IPF research. Pulm. Pharmacol. Ther. 24, 137–148
37. Sutavani, R. V., Phair, I. R., Barker, R., McFarlane, A., Shiipo, N., Lang, S., et al. (2018) Differential control of Toll-like receptor 4-induced interleukin-10 induction in macrophages and B cells reveals a role for p90 ribosomal S6 kinases. J. Biol. Chem. 293, 2302–2317
38. Ananieva, O., Darragh, J., Johansen, C., Carr, J. M., McIlrath, J., Park, J. M., et al. (2008) The kinases MSK1 and MSK2 act as negative regulators of Toll-like receptor signaling. Nat. Immunol. 9, 1028–1036
39. MacKenzie, K. F., Clark, K., Naqi, S., McGuire, V. A., Noehren, G., Kristariyanto, Y., et al. (2013) PGE2 induces macrophage IL-10 production and a regulatory-like phenotype via a protein kinase A-SIK-CRTC3 pathway. J. Immunol. 190, 565–577
40. Clark, K., MacKenzie, K. F., Petkevicius, K., Kristariyanto, Y., Zhang, J., Choi, H. G., et al. (2012) Phosphorylation of CRTC3 by the salt-inducible kinases controls the interconversion of classically activated and regulated macrophages. Proc. Natl. Acad. Sci. U. S. A. 109, 16986–16991
41. Darling, N. J., Toth, R., Arthur, J. S., and Clark, K. (2017) Inhibition of SIK2 and SIK3 during differentiation enhances the anti-inflammatory phenotype of macrophages. Biochem. J. 474, 521–537
42. Ozanne, J., Prescott, A. R., and Clark, K. (2015) The clinically approved drugs dasatinib and bosutinib induce anti-inflammatory macrophages by inhibiting the salt-inducible kinases. Biochem. J. 465, 271–279
43. Iwamoto, N., Distler, J. H., and Distler, O. (2011) Tyrosine kinase inhibitors in the treatment of systemic sclerosis: from animal models to clinical trials. Curr. Rheumatol. Rep. 13, 21–27
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36. Grimminger, F., Gunther, A., and Vancheri, C. (2015) The role of tyrosine kinases in the pathogenesis of idiopathic pulmonary fibrosis. Eur. Respir. J. 45, 1426–1433
37. Richeldi, L., do Bois, R. M., Raghu, G., Azuma, A., Brown, K. K., Costabel, U., et al. (2014) Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis. N. Engl. J. Med. 370, 2071–2082
38. Liu, F., Bayliss, G., and Zhuang, S. (2019) Application of nintedanib and other potential anti-fibrotic agents in fibrotic diseases. Clin. Sci. (Lond) 133, 1309–1320
39. McCormack, P. L. (2015) Nintedanib: first global approval. Drugs 75, 129–139
40. Yilmaz, O., Oztoy, F., and Kayalar, O. (2015) Dasatinib attenuated bleomycin-induced pulmonary fibrosis in mice. Growth Factors 33, 366–375
41. Hilberg, F., Roth, G. J., Krassak, M., Kautschitsch, S., Sommergruber, W., Tontsch-Grunt, U., et al. (2008) BIBF 1120: triple angiokinase inhibitor with sustained receptor blockade and good antitumor efficacy. Cancer Res. 68, 4774–4782
42. Hilberg, F., Tontsch-Grunt, U., Baum, A., Le, A. T., Doebele, R. C., Lieb, S., et al. (2018) Triple angiokinase inhibitor nintedanib directly inhibits tumor cell growth and induces tumor shrinkage via blocking oncogenic receptor tyrosine kinases. J. Pharmacol. Exp. Ther. 364, 494–503
43. Ambarus, C. A., Krausz, S., van Eijk, J., Hamann, J., Radstake, T. R., Reedquist, K. A., et al. (2012) Systematic validation of specific phenotypic markers for in vitro polarized human macrophages. J. Immunol. Met. 375, 196–206
44. Stein, M., Keshav, S., Harris, N., and Gordon, S. (1992) Interleukin 4 potently enhances murine macrophage mannose receptor activity: a marker of alternative immunologic macrophage activation. J. Exp. Med. 176, 287–292
45. Raes, G., Blys, L., Dahal, B. K., Brandt, I., Grooeten, J., Brombacher, F., et al. (2005) Macrophage galactose-type C-type lectins as novel markers for alternatively activated macrophages elicited by parasitic infections and allergic airway inflammation. J. Leukoc. Biol. 77, 321–327
46. Misharin, A. V., Morales-Nebreda, L., Mutlu, G. M., Judinger, G. R., and Perlman, H. (2013) Flow cytometric analysis of macrophages and dendritic cell subsets in the mouse lung. Am. J. Respir. Cell Mol. Biol. 49, 503–510
47. Li, M., Krishnaveni, M. S., Li, C., Zhou, B., Xing, Y., Banfalvi, A., et al. (2011) Epithelium-specific deletion of TGF-beta receptor type II protects mice from bleomycin-induced pulmonary fibrosis. J. Clin. Invest. 121, 277–287
48. Zhao, J., Shi, W., Wang, Y. L., Chen, H., Bringas, P. J., Datto, M. B., et al. (2002) Smad3 deficiency attenuates bleomycin-induced pulmonary fibrosis in mice. Am. J. Physiol. Lung Cell Mol. Physiol. 282, L585–593
49. Sonnenberg, G. F., Nair, M. G., Birnbaum, J. I., Zaph, C., Fouser, L. A., and Artis, D. (2010) Pathological versus protective functions of IL-22 in airway inflammation but does not affect inflammatory phenotype. J. Leukoc. Biol. 99, 711–721
50. Nefa, M., Darling, N. J., van Gijssel Bonollo, M., Cohen, P., and Arthur, J. S. C. (2011) Salt-inducible kinases 2 and 3 are required for thymic T cell development. J. Exp. Med. 213, 955–961
51. Nefa, M., Darling, N. J., van Gijssel Bonollo, M., Cohen, P., and Arthur, J. S. C. (2011) Salt-inducible kinases 2 and 3 are required for thymic T cell development. J. Exp. Med. 213, 955–961
52. Xue, J., Schmidt, S. V., Sander, J., Drahfeh, A., Krebs, W., Quester, L., et al. (2014) Transcriptome-based network analysis reveals a spectrum model of human macrophage activation. Immunity 40, 274–288
53. Böttger, M., De Klee, I., Henri, S., van der Vlist, L., Dijkstra, S., et al. (2013) Alveolar macrophages develop from fetal monocytes that differentiate into long-lived cells in the first week of life via GM-CSF. J. Exp. Med. 210, 1977–1992
54. [preprint] Ghebremedhin, A., Salam, A. B., Adu-Addai, B., Noonan, S., Stratton, R., Ahmed, M. S. U., et al. (2020) A novel CD206 targeting peptide inhibits bleomycin induced pulmonary fibrosis in mice. bioRxiv. https://doi.org/10.1101/2020.07.27.218115
55. Sato, K., Imai, Y., Higashi, N., Kumatomo, Y., Onami, T., Hidrick, S. M., et al. (2005) Lack of antigen-specific tissue remodeling in mice deficient in the macrophage galactose-type calcium-type lectin 1/CD301a. Blood 106, 207–215
56. Boer, J., Williams, A., Skavdiss, G., Harker, N., Coles, M., Tolaini, M., et al. (2007) The selectivity of protein kinase inhibitors: a further update. Biochem. J. 408, 297–315
57. Klop, S. K., Imai, Y., Higashi, N., Kumatomo, Y., Onami, T., Hidrick, S. M., et al. (2005) Lack of antigen-specific tissue remodeling in mice deficient in the macrophage galactose-type calcium-type lectin 1/CD301a. Blood 106, 207–215
58. de Boer, J., Williams, A., Skavdiss, G., Harker, N., Coles, M., Tolaini, M., et al. (2007) The selectivity of protein kinase inhibitors: a further update. Biochem. J. 408, 297–315
59. Klop, S. K., Imai, Y., Higashi, N., Kumatomo, Y., Onami, T., Hidrick, S. M., et al. (2005) Lack of antigen-specific tissue remodeling in mice deficient in the macrophage galactose-type calcium-type lectin 1/CD301a. Blood 106, 207–215
50. Klop, S. K., Imai, Y., Higashi, N., Kumatomo, Y., Onami, T., Hidrick, S. M., et al. (2005) Lack of antigen-specific tissue remodeling in mice deficient in the macrophage galactose-type calcium-type lectin 1/CD301a. Blood 106, 207–215
60. Ratto, K., Imai, Y., Higashi, N., Kumatomo, Y., Onami, T., Hidrick, S. M., et al. (2005) Lack of antigen-specific tissue remodeling in mice deficient in the macrophage galactose-type calcium-type lectin 1/CD301a. Blood 106, 207–215