Induction of Ran GTP drives ciliogenesis

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ABSTRACT The small GTPase Ran and the importin proteins regulate nucleocytoplasmic transport. New evidence suggests that Ran GTP and the importins are also involved in conveying proteins into cilia. In this study, we find that Ran GTP accumulation at the basal bodies is coordinated with the initiation of ciliogenesis. The Ran-binding protein 1 (RanBP1), which indirectly accelerates Ran GTP → Ran GDP hydrolysis and promotes the dissociation of the Ran/importin complex, also localizes to basal bodies and cilia. To confirm the crucial link between Ran GTP and ciliogenesis, we manipulated the levels of RanBP1 and determined the effects on Ran GTP and primary cilia formation. We discovered that RanBP1 knockdown results in an increased concentration of Ran GTP at basal bodies, leading to ciliogenesis. In contrast, overexpression of RanBP1 antagonizes primary cilia formation. Furthermore, we demonstrate that RanBP1 knockdown disrupts the proper localization of KIF17, a kinesin-2 motor, at the distal tips of primary cilia in Madin–Darby canine kidney cells. Our studies illustrate a new function for Ran GTP in stimulating cilia formation and reinforce the notion that Ran GTP and the importins play key roles in ciliogenesis and ciliary protein transport.

INTRODUCTION Virtually all mammalian cells display cilia, and these organelles execute essential functions. Ciliary functions include motility (e.g., sweeping motion across respiratory tract epithelium), development of left–right asymmetry during embryogenesis, photosenescence (e.g., photoreceptors in the retina), mechanosensation (e.g., fluid flow in the renal tubules), chemosensation (e.g., odorant detection in the olfactory sensory epithelium), and initiation of some signal transduction cascades (e.g., Hh, Wnt, and PDGF; Nonaka et al., 2002; Rosenbaum and Witman, 2002; Huangfu et al., 2003; Haycraft et al., 2005; Schneider et al., 2005; Satir and Christensen, 2007; Lal et al., 2008; Scholery, 2008; Berbari et al., 2009; Gerdes et al., 2009; Johnson and Leroux, 2010). Ciliopathies have been recognized for several years and are often marked by a constellation of clinical features, including retinal degeneration, mental retardation, situs inversus, polyactyly, encephalocoele, and cystic disease of the kidney, liver, and pancreas (Badano et al., 2006; Fliegauf et al., 2007). Indeed, polycystic kidney disease and retinitis pigmentosa are two well-described ciliopathies.

Of interest, protein synthesis does not occur in cilia, and recent work has begun to explain how proteins are targeted to this organelle. Specifically, intraflagellar transport (IFT) is now appreciated as essential for assembly and maintenance of almost all flagella and cilia. IFT can function to transport membrane channels and signaling proteins into and along the cilia and flagella (Rosenbaum and Witman, 2002; Pollitt et al., 2006; Pazour and Bloodgood, 2008; Pedersen and Rosenbaum, 2008). In addition, various ciliary targeting sequences have been identified that appear to direct proteins into the cilia (Geng et al., 2006; Jenkins et al., 2006; Tao et al., 2009; Pollitt et al., 2010; Hurd et al., 2011). In addition, small G proteins and their regulators, such as Rab6, Rab8, Rab10, Rab11, Rab23, Arf4, Arf6, and Rabin8, were reported to mediate ciliary protein targeting from the Golgi, trans-Golgi network, and ciliary transition zone into cilia and also photoreceptors (Eggenschwiler et al., 2001; Deretic et al., 2005; Nachury et al., 2007; Yoshimura et al., 2007; Babbey et al., 2010; Knodler et al., 2010). Furthermore, elegant new studies demonstrate that ciliary transition zone proteins regulate the entry and exit of ciliary proteins (Craigie et al., 2010; Williams...
et al., 2011). Nevertheless, our view of ciliary transport is still evolving, and a complete understanding of the mechanisms that target proteins to cilia is elusive.

Recent studies from our group and others suggest that Ran GTP and the importins are involved in targeting proteins to primary cilia. As it is classically understood, the Ran/importin system is a nucleocytoplasmic shuttle (Harel and Forbes, 2004). In interphase cells, the steep gradient of Ran GTP between cytosol (low) and nucleus (high) promotes cargo-importin loading in the cytosol and induces cargo release in the nucleus (Gorlich et al., 2003; Kalab et al., 2006). In the cytosol, importin α binds both cargo protein and importin β, resulting in an importin α/β/cargo heterotrimer that is transferred across the nuclear pore. Once in the nucleus, Ran GTP binds the importins—releasing the nuclear cargo—and Ran GTP/cellular apoptosis susceptibility gene product/importin α and Ran GTP/importin β are separately recycled back to the cytosol. Once in the cytosol, the Ran GTP/importin complexes associate with Ran GTPase-activating protein 1 (RanGAP1) and Ran-binding protein 1 (RanBP1), converting Ran GTP to Ran GDP and releasing the importins (Harel and Forbes, 2004; Pemberton and Paschal, 2005; Yudin and Fainzilber, 2009). The cycle then begins anew. Of interest, knockdown of RanBP1 markedly increases cytosolic Ran GTP and disrupts the critical intracellular Ran GTP gradient (Tedeschi et al., 2007). During cell division, Ran GTP concentrates at the chromosomes and mitotic spindles. At the spindle poles, Ran GTP releases active aster-promoting activities—including TPX2 and NuMa—from the importin α/β heterodimer, and this event subsequently initiates the process of spindle assembly (Clarke and Zhang, 2001; Nachury et al., 2001).

In light of this nuclear transport model, our group and others discovered that some ciliary proteins contain nuclear localization signals (NLS), which mediate a direct interaction between these ciliary proteins and the importins. Of note, the Verhey group found that importin β2 (transportin) binds to a C-terminal NLS-like sequence in KIF17, a kinesin-2 motor that ferries cargo to the cilary tip. Mutation of the KIF17 NLS-like sequence abrogated its entry into the cilium, indicating that this sequence serves as a ciliary localization signal (CLS) for KIF17 (Dishinger et al., 2010). Similarly, our group demonstrated that importin β2 directs the retinitis pigmentosa protein 2 (RP2) to the primary cilium via an interaction with the M9 domain of RP2 (Hurd et al., 2011). The ciliary proteome reveals that many importins and Ran regulatory proteins reside in ciliary compartments, suggesting that Ran/importin might govern ciliogenesis in addition to controlling the localization of ciliary proteins (Andersen et al., 2003; Avidor-Reiss et al., 2004; Li et al., 2004; Blacque et al., 2005; Efimenko et al., 2005; Keller et al., 2005; Pazour et al., 2005; Stolc et al., 2005; Broadhead et al., 2006; Gherman et al., 2006; Liu et al., 2007).

Of interest, a fraction of cellular Ran GTP and importin β2 is found at the cilia and centrosomes during interphase and regulates the localization of some ciliary proteins (Keryer et al., 2003; Fan et al., 2007; Dishinger et al., 2010; Hurd et al., 2011). We hypothesize that the concentrated Ran GTP at the centrosomes serves as a docking platform to release ciliary proteins from importins and to allow entry past the newly formed basal bodies into the cilium. By this mechanism, we propose that Ran GTP regulates ciliary protein transport and ciliogenesis.

**RESULTS**

**Ran GTP localizes to cilia and basal bodies**

To explore the possible functions of Ran GTP in ciliogenesis, we generated polyclonal antibodies toward Ran GTP using a Ran peptide as previously described by the Macara laboratory (Richards et al., 1995). This antibody detected a single band around 25 kDa in Madin–Darby canine kidney (MDCK), murine inner medullary collecting duct 3 (IMCD3), and human telomerase-immortalized retinal pigment epithelial 1 cells (TERT RPE; Supplemental Figure S1A). Furthermore, this antibody immunoprecipitated Myc-RanQ69L (GTP-locked Ran) but not Myc-RanT24N (GDP-locked Ran) (Supplemental Figure S1B). Using this reagent, we extended our previous studies (Dishinger et al., 2010) and localized Ran GTP in additional mammalian cell types and tissues. We found that Ran GTP localized to primary cilia (Figure 1A, top) and to basal bodies (Figure 1A, bottom) in fully polarized MDCK cells. In differentiated human air tract

**FIGURE 1:** Ran GTP localizes to cilia and basal bodies. (A) Ran GTP localizes to primary cilia and basal bodies in MDCK cells. MDCK cells were grown on Transwell filters for 6 d postconfluence, fixed, and stained with anti–Ran GTP (green) and either anti–polyglutamylated tubulin (red; top) or anti–γ-tubulin (red; bottom). Nuclei were stained with 4′,6-diamidino-2-phenylindole (DAPI). (B) Ran GTP localizes to motile cilia in air tract epithelia. Air tract epithelia were fixed and stained with anti–Ran GTP (green) and either anti–polyglutamylated tubulin (red; top) or anti–γ-tubulin (red; bottom). Nuclei were stained with DAPI. (C) Ran GTP localizes to motile cilia and basal bodies in respiratory turbinates. GFP-centrin2 transgenic mouse respiratory turbinates were stained with anti–Ran GTP (red) and anti–γ-tubulin (blue). (D) Ran GTP localizes to olfactory sensory neuron cilia. GFP-centrin2 transgenic mouse olfactory epithelial sections were stained with anti–Ran GTP (green) and anti–acetylated tubulin (red). Scale bars, 10 μm.
epithelial cells, Ran GTP localizes to motile cilia and multiple basal bodies (Figure 1B). In green fluorescent protein (GFP)–centrin2 transgenic mouse sections, we determined that Ran GTP primarily localizes to the basal bodies and motile cilia of nasal respiratory epithelium (Figure 1C). We also demonstrated that Ran GTP accumulated at olfactory cilia in GFP-centrin2 mouse sections, similar to our previous findings in wild-type mouse (Dishinger et al., 2010; Figure 1D). We confirmed that Ran GTP antigenic peptides blocked our previous findings in wild-type mouse (Dishinger et al., 2010; Pugacheva et al., 2007). Usually, primary cilia are fully developed following 24-48 h of serum deprivation. Here, we cultured TERT RPE cells in complete media or serum-free media for 48 h and then stained with anti–acetylated tubulin and anti–Ran GTP to visualize the primary cilia. We did not detect primary cilia or Ran GTP staining in basal bodies when cells were grown in complete media (Figure 2C, left). However, we noted a dramatic enhancement of Ran GTP staining in the basal bodies of primary cilia following serum starvation (Figure 2C, right, and Supplemental Figure S2B, C). To rule out the possibility that ciliary Ran GTP might contaminate the centrosomal fractions, we did a deciliation time course of TERT RPE cells during nocodazole and cytochalasin B treatment. These agents are commonly used for centrosome isolation procedures. Cilia were completely disassembled after 90 min of treatment with nocodazole and cytochalasin B (Supplemental Figure S2A). In addition, centrosome isolation revealed that Ran GTP levels in the centrosomal fractions were significantly augmented when TERT RPE cells were cultured in serum-free media (Figure 2D). We also confirmed the apparent increase in centrosome-associated Ran GTP in TERT RPE cells after serum starvation by anti-Ran GTP immunoprecipitation (Supplemental Figure S2D). The optimized staining conditions that allow us to visualize centrosomal Ran GTP (cold acetone fixation) yield weak Ran GTP nuclear staining. However, we observed strong nuclear localization of Ran when we used a protocol adapted for nuclear staining: 4% paraformaldehyde fixation followed by 1% SDS permeabilization (Supplemental Figure S1C).

The accumulation of Ran GTP at the centrosome is linked to ciliogenesis

The localization of Ran GTP with cilia and basal bodies prompted us to examine whether Ran GTP fills a functional role in ciliogenesis. First, we assayed the presence of Ran GTP in centrosomes during MDCK polarization, a lengthy process that culminates in cilia formation. We cultured MDCK cells on Transwell filters for various lengths of time and stained with γ-tubulin and Ran GTP antibodies. At day 1, when MDCK cells were not yet polarized, we did not strongly detect Ran GTP at the centrosomes (Figure 2A, top). However, Ran GTP significantly accrued at the centrosomes by days 3 and 6 (Figure 2A, middle and bottom). We also biochemically confirmed the finding that increased levels of Ran GTP were associated with centrosomes by centrosomal fractionation (Figure 2B). We used full-length glutathione S-transferase (GST)–importin β1 to pull down Ran GTP from the isolated centrosomes of fully polarized MDCK cells and probed with a mouse anti-Ran antibody. We verified that Ran GTP is clearly enriched in the centrosomal (γ-tubulin positive) fractions (Figure 2B). In nonpolarized MDCK cells, we weakly detected Ran GTP in the centrosomal fractions (data not shown).

In fibroblasts and some epithelial cells such as TERT RPE and IMCD3, serum starvation initiates primary ciliogenesis (Schneider et al., 2005; Pugacheva et al., 2007). Usually, primary cilia are fully developed following 24-48 h of serum deprivation. Here, we cultured TERT RPE cells in complete media or serum-free media for 48 h and then stained with anti–acetylated tubulin and anti–Ran GTP to visualize the primary cilia. We did not detect primary cilia or Ran GTP staining in basal bodies when cells were grown in complete media (Figure 2C, left). However, we noted a dramatic enhancement of Ran GTP staining in the basal bodies of primary cilia following serum starvation (Figure 2C, right, and Supplemental Figure S2B, C). To rule out the possibility that ciliary Ran GTP might contaminate the centrosomal fractions, we did a deciliation time course of TERT RPE cells during nocodazole and cytochalasin B treatment. These agents are commonly used for centrosome isolation procedures. Cilia were completely disassembled after 90 min of treatment with nocodazole and cytochalasin B (Supplemental Figure S2A). In addition, centrosome isolation revealed that Ran GTP levels in the centrosomal fractions were significantly augmented when TERT RPE cells were cultured in serum-free media (Figure 2D). We also confirmed the apparent increase in centrosome-associated Ran GTP in TERT RPE cells after serum starvation by anti-Ran GTP immunoprecipitation (Supplemental Figure S2D). The optimized staining conditions that allow us to visualize centrosomal Ran GTP (cold acetone fixation) yield weak Ran GTP nuclear staining. However, we observed strong nuclear localization of Ran GTP when we used a protocol adapted for nuclear staining: 4% paraformaldehyde fixation followed by 1% SDS permeabilization (Supplemental Figure S1C).

RanBP1 also localizes to cilia and/or basal bodies

RanBP1 stimulates RanGAP1 to hydrolyze Ran GTP to Ran GDP and also inhibits the activity of the Ran guanine nucleotide exchange protein. We cultured MDCK cells on Transwell filters for various lengths of time and stained with γ-tubulin and Ran GTP antibodies. At day 1, when MDCK cells were not yet polarized, we did not strongly detect Ran GTP at the centrosomes (Figure 2A, top). However, Ran GTP significantly accrued at the centrosomes by days 3 and 6 (Figure 2A, middle and bottom). We also biochemically confirmed the finding that increased levels of Ran GTP were associated with centrosomes by centrosomal fractionation (Figure 2B). We used full-length glutathione S-transferase (GST)–importin β1 to pull down Ran GTP from the isolated centrosomes of fully polarized MDCK cells and probed with a mouse anti-Ran antibody. We verified that Ran GTP is clearly enriched in the centrosomal (γ-tubulin positive) fractions (Figure 2B). In nonpolarized MDCK cells, we weakly detected Ran GTP in the centrosomal fractions (data not shown).
factor (GEF) RCC1 (Hayashi et al., 1995; Lounsbury and Macara, 1997). Thus RanBP1 is considered a negative regulator of Ran GTP, and it is predominantly localized to the cytoplasm. Of interest, the cilium/flagella combined proteome database reveals that several importin α and β isoforms, Ran GTPase, and Ran-binding proteins are found in these organelles (Gherman et al., 2006). In addition, a small amount of RanBP1 was reported to localize to centrosomes (Di Fiore et al., 2003). We investigated whether RanBP1 might regulate Ran GTP/GDP conversion in cilia and in the basal bodies that initiate ciliogenesis. To address this possibility, we tested the localization of endogenous RanBP1 in IMCD3 cells and air tract epithelial cells. Although RanBP1 distributes throughout the cytoplasm, we also detected RanBP1 containing along primary cilia with acetylated tubulin in IMCD3 cells (Figure 3A). Similarly, in fully polarized MDCK cells, we moderately expressed inducible RanBP1-FLAG and observed its colocalization with primary cilia upon immunostaining with anti-FLAG and anti–acetylated tubulin antibodies (Figure 3B). In air tract epithelial cells, RanBP1 puncta were distributed along motile cilia (Figure 3C). In TERT RPE cells, we detected endogenous RanBP1 concentrated in the basal bodies of primary cilia following serum starvation (Figure 3D). We also expressed inducible enhanced GFP (EGFP)–RanBP1 in RPE cells and costained with either γ-tubulin or acetylated tubulin antibodies following 48 h of 50 ng/ml doxycycline (Dox) treatment and serum starvation to induce ciliogenesis. As with endogenous RanBP1, we did not observe strong localization of EGFP-RanBP1 in cilia, and we noted that EGFP-RanBP1 is associated with γ-tubulin at centrosomes (Figure 3E) and with the basal bodies of primary cilia (Figure 3F).

**FIGURE 3:** RanBP1 localizes to the primary cilia and basal bodies. (A) RanBP1 localizes to primary cilia in IMCD3 cells. IMCD3 cells were cultured on chamberslides for 24 h in complete media and then serum starved for an additional 24 h. Cells were fixed and stained with anti–acetylated tubulin (red) and anti-RanBP1 (green). (B) RanBP1 localizes to primary cilium in MDCK cells. RanBP1-FLAG–inducible MDCK cells were grown on Transwell filters for 5 d and induced with 50 ng/ml DOX for 48 h. Cells were fixed and stained with anti-FLAG (green) and anti–acetylated tubulin (red). (C) RanBP1 localizes to motile cilia in air tract epithelia. Air tract epithelial cells were differentiated on Transwell filters for 3 wk and stained with anti-RanBP1 (green) and anti–acetylated tubulin (red). (D) RanBP1 localizes to the basal body in TERT RPE cells. RPE cells were seeded on chamber slides and cultured with serum-free media for 48 h. Cells were fixed with 4% paraformaldehyde and stained with anti-RanBP1 (green) and anti–acetylated tubulin (red). (E, F) EGFP-RanBP1 localizes to the basal body in TERT RPE cells. Inducible EGFP-RanBP1 RPE cells were seeded on chamberslides and induced with 50 ng/ml DOX for 48 h in serum-free media. Cells were fixed and stained with anti–γ-tubulin (red) in E or anti–acetylated tubulin (red) in F. Bottom, high-amplification insets from the boxed regions. Scale bars, 10 μm.

Because reduction of RanBP1 promotes ciliogenesis, we reasoned that overexpression of RanBP1 might impair primary cilium formation. To this end, we further characterized the TERT RPE cell lines stably expressing inducible EGFP-RanBP1. We induced EGFP-RanBP1 expression using various doses of Dox coupled with 24 h of serum starvation to maximize ciliary growth. EGFP-RanBP1 expression increased with higher doses of Dox. We observed that 50 ng/ml Dox induced a 1.5-fold increase in EGFP-RanBP1 compared with endogenous RanBP1, whereas 100 ng/ml or 200 ng/ml Dox induced approximate threefold to fourfold enhancement in EGFP-RanBP1 relative to endogenous RanBP1 (Figure 5A). EGFP-RanBP1–inducible cells treated with low levels of Dox (50 ng/ml) and all control cells (0–200 ng/ml Dox) were still able to form primary cilia; however, inducible EGFP-RanBP1 cells exposed to higher doses of Dox (100 and 200 ng/ml) lose almost all cilia formation following serum starvation (Figure 5B). To exclude the possibility that EGFP expression abrogates cilia formation, we made control EGFP TERT RPE cells. Similar levels of EGFP expression alone do not disrupt conditions promoting cell cycle progression (Figure 4B, right, arrow). Enhanced ciliogenesis in the RanBP1 knockdown cells is quantified in Figure 4C.
Figure 6B. Next we corroborated the presence of increased Ran GTP levels in control and RanBP1-knockdown cells is depicted in Figure 6A. KPNA2 (importin β2) serves as an authentic CLS. Mutation of the CLS barred KIF17 ciliary entry. In a separate report, we demonstrated that expression of RanG19V (GTP-locked Ran) abolished ciliary entry of KIF17. Because Ran GTP is significantly increased in the basal bodies of RanBP1-knockdown cells, we next analyzed the fate of KIF17 in these cells. RanBP1-knockdown IMCD3 cells were transiently transfected with KIF17-mCitrine, cultured in serum-free media for 24 h, fixed, and stained with anti-acetylated tubulin and anti-RanBP1. Consistent with previous reports, we found KIF17-mCitrine localized to the tips of primary cilia in control cells (Figure 7A, top). In addition, we noted that RanBP1 colocalizes with KIF17-mCitrine at the tips of primary cilia (Figure 7A, top). However, KIF17-mCitrine localization at ciliary tips was perturbed in RanBP1-knockdown cells, and KIF17-mCitrine was observed in a midway position along the body of the cilium (Figure 7A, bottom). The global disruption of proper ciliary tip KIF17-mCitrine localization among a field of RanBP1 knockdown cells (as compared with controls) is shown in Figure 7B. Quantification of the immunofluorescence data confirms that virtually all of the ciliary tips contain KIF17-mCitrine in control cells; however, nearly half of the ciliary tips are missing KIF17-mCitrine in RanBP1-knockdown cells (Figure 7C).

DISCUSSION

Studies from our group and others suggest that the importins, traditionally thought of as nucleocytoplasmic shuttles, may function as ciliary transporters as well. Our earlier work demonstrated that a small transmembrane protein Crb3b (also called Crumbs3-CLPI) binds to importin β1. In addition, Crb3b colocalizes with importin β1 in the primary cilium, which suggests that importin β1 may target Crb3b to this organelle (Fan et al., 2007). In a separate report, we showed that importin β2 (also called transportin) directly interacts with RP2. Of note, knockdown of importin β2 blocked the ciliary localization of both endogenous RP2 and EGFP-RP2, implicating importin β2 as the ciliary targeting protein for RP2 (Hurd et al., 2011). This is in agreement with concurrent studies from our group that identified two conserved NLS sequences encoded in KIF17, a kinesin-2 motor. It was discovered that the C-terminal NLS sequence serves as an authentic CLS. Mutation of the CLS barred KIF17 cilary entry, and this observation led to the elucidation of a mechanism in RanBP1 knockdown inhibits ciliogenesis.

RanBP1 knockdown initiates primary cilium formation (Supplemental Figure S4, A and B). Thus we validated the finding that RanBP1 antagonizes cilia formation, and these results are quantified in Figure 5C (control vs. EGFP-RanBP1 cells, 200 ng/ml Dox). It has been reported that RanBP1 knockdown or overexpression can alter cell growth, as well as the cell cycle, and this could alter ciliogenesis (Battistoni et al., 1997; Guarguaglini et al., 1997, 2000; Di Fiore et al., 2003; Tedeschi et al., 2007). However, the manipulations of RanBP1 used in this study had only minor effects on the cell cycle (Supplemental Figures S3 and S4, C and D). Knockdown of RanBP1 did reduce cells in S and G2+M (Supplemental Figure S3), but this did not seem sufficient to explain the large increase in ciliated cells. Overexpression of RanBP1 had only minor effects in reducing S phase (Supplemental Figure S4, C and D) yet markedly perturbed ciliogenesis.

RanBP1 knockdown enhances Ran GTP in centrosomes

Previous studies demonstrated that RanBP1 knockdown increases cytosolic Ran GTP (Tedeschi et al., 2007). In the present investigation, we found that RanBP1 knockdown stimulates ciliogenesis independent of serum starvation. We inferred that RanBP1 suppression may drive ciliary growth by increasing Ran GTP concentrations at the basal bodies. To test this notion, we isolated centrosomes from RanBP1-knockdown and control cells and subjected the purified fractions to a GST-importin β1 pull-down experiment. Indeed, the RanBP1-knockdown cells demonstrate increased centrosome-associated Ran GTP levels (Figure 6A). KPNA2 (importin α1) served as a positive GST-importin β1 pull-down control. Quantification of Ran GTP levels in control and RanBP1-knockdown cells is depicted in Figure 6B. Next we corroborated the presence of increased Ran GTP at basal bodies in situ. We cultured control and RanBP1-knockdown cells in complete growth media for 48 h and costained with Ran GTP and acetylated tubulin or γ-tubulin antibodies. Indeed, we identified markedly enhanced Ran GTP staining at the basal bodies of primary cilia in RanBP1-knockdown cells. In sharp contrast, we detected neither primary cilia nor significant levels of Ran GTP at the centrosomes in control knockdown cells (Figure 6C and Supplemental Figure S5A). Quantification of Ran GTP staining in control and RanBP1-knockdown cells is displayed in Supplemental Figure S5B.

RanBP1 knockdown disrupts localization of KIF17 to the distal tips of primary cilia

Recently we reported that Ran GTP and importin β2 regulate ciliary entry of KIF17, a kinesin-2 motor. It was demonstrated that expression of RanG19V (GTP-locked Ran) abolished ciliary entry of KIF17. Because Ran GTP is significantly increased in the basal bodies of RanBP1-knockdown cells, we next analyzed the fate of KIF17 in these cells. RanBP1-knockdown IMCD3 cells were transiently transfected with KIF17-mCitrine, cultured in serum-free media for 24 h, fixed, and stained with anti-acetylated tubulin and anti-RanBP1. Consistent with previous reports, we found KIF17-mCitrine localized to the tips of primary cilia in control cells (Figure 7A, top). In addition, we noted that RanBP1 colocalizes with KIF17-mCitrine at the tips of primary cilia (Figure 7A, top). However, KIF17-mCitrine localization at ciliary tips was perturbed in RanBP1-knockdown cells, and KIF17-mCitrine was observed in a midway position along the body of the cilium (Figure 7A, bottom). The global disruption of proper ciliary tip KIF17-mCitrine localization among a field of RanBP1 knockdown cells (as compared with controls) is shown in Figure 7B. Quantification of the immunofluorescence data confirms that virtually all of the ciliary tips contain KIF17-mCitrine in control cells; however, nearly half of the ciliary tips are missing KIF17-mCitrine in RanBP1-knockdown cells (Figure 7C).

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In this study, we expand upon the existing paradigm and explore the functional role of Ran GTP in the centrosome/basal body and its relationship to ciliogenesis. To this end, we tested the Ran GTP distribution in several cell lines and tissues that display different types of cilia. We found that Ran GTP is remarkably enriched in both cilia and the basal bodies. We recognized a tight association between cell culture conditions that favor cilia formation—for instance, polari-
zation of MDCK cells and serum starvation of TERT RPE cells—and a marked accumulation of Ran GTP at the centrosomes. In MDCK cells, ciliogenesis begins when the cells are fully polarized, and this process usually takes 5–7 d to complete. In fibroblasts and some epithelial cells, serum-free culture initiates primary cilia formation within 24–48 h. These findings prompted us to propose that up-regulation of Ran GTP may be a crucial effector for ciliogenesis.

If Ran GTP is indeed a required factor in ciliogenesis, then we imagined that adjusting intracellular Ran GTP levels should yield profound effects on cilia formation. Fortuitously, earlier work showed that Ran GTP regulatory proteins reside in the centrosomes, among them the well-described RanBP1 (Di Fiore et al., 2003). RanBP1 is not a catalytic partner that regulates GTP/GDP turnover of Ran. Rather, RanBP1 binds to RanGAP1, and RanGAP1 promotes the hydrolysis of Ran GTP to Ran GDP (Bischoff et al., 1995; Kuhlmann et al., 1997; Lounsbury and Macara, 1997). In addition, RanBP1 inhibits the activity of RCC1, a Ran GEF, and thereby suppresses the Ran GDP to Ran GTP conversion (Hayashi et al., 1995). In short, the presence of RanBP1 promotes the formation of Ran GDP.

Thus we focused our attention on RanBP1 as a molecular tool to regulate Ran GTP. We demonstrated that endogenous and epitope-tagged RanBP1 localizes to cilia and/or basal bodies in several ciliated cell lines. Next, we reduced RanBP1 levels by shRNA-knockdown techniques and evaluated its effects on cilia formation and intra-
cellular Ran GTP in TERT RPE cells. We found that RanBP1-knockdown RPE cells robustly initiated primary cilia formation independent of serum starvation, normally a required condition for ciliogenesis. We noted that Ran GTP is simultaneously increased in the centrosomes/basal bodies of RanBP1 RPE–knockdown cells. These experiments provide additional support for the concept that Ran GTP enrichment at the centrosome promotes cilia formation.

It is known that knockdown of RanBP1 can have small effects on the progression of the cell cycle and induce apoptosis (Tedeschi et al., 2007). In our system, we saw a slight, 4–6% decrease of the mitotic index in RanBP1-knockdown TERT RPE cells by fluorescence-activated cell sorting (FACS) analysis (Supplemental Figure S3, A and B); however, this could not explain the nearly 20- to 30-fold increase we saw in ciliogenesis. Similarly, in IMCD3 cells RanBP1 knockdown resulted in specific defects in cilia trafficking, independent of ciliogenesis. In addition, overexpression of RanBP1 has been reported to impair S phase entry and the mitotic cycle and to induce multipolar spindles (Battistoni et al., 1997; Guarguaglini et al., 1997, 2000; Di Fiore et al., 2003). We did not detect any spindle or centrosome cohesion defects in cells overexpressing EGFP-RanBP1 in TERT RPE cells. Our FACS results showed that overexpression of EGFP-RanBP1 did not block cell cycle exit from G1 to G0 after serum starvation (Supplemental Figure S4, A and B); however, this could not provide additional support for the concept that Ran GTP enrichment at the centrosome promotes cilia formation.

Although we do find Ran GTP and RanBP1 concentrated at the ciliary tips in our study, we do not yet understand their function in this specialized location. It is interesting to note that the specific localization in a Ran GTP–dependent manner (Dishinger et al., 2010). In collaboration with the Verhey laboratory, we also found that Ran GTP was enriched in the primary cilia of NIH 3T3 cells (Dishinger et al., 2010).

A

| DOX (ng/ml) | 0 | 50 | 100 | 200 |
|------------|---|----|-----|-----|
| EGFP-RanBP1 | Biot: RanBP1 | Endogenous RanBP1 | Biot: RanBP1 | Biot: Actin |
| Control vector | EGFP-RanBP1/Acetylated tubulin/DAPI |

![Image](36x411 to 378x735)

FIGURE 5: Expression of EGFP-RanBP1 disrupts primary cilium formation. (A) TERT RPE cells were infected with an inducible EGFP-RanBP1 retroviral construct, and a stable pool was selected. After growing the cells on chamberslides or 6-cm culture dishes for 24 h, the cells were induced in serum-free media with 50–200 ng/ml Dox for the next 24 h. Cells were harvested, and 20-μg protein samples were electrophoresed on a Bis-Tris PAGE and blotted with RanBP1 and actin antibodies. (B) Cells were fixed and stained with anti–acetylated tubulin. (C) Quantification of the experiments depicted in B: 100 cells from each cell line were counted (EGFP-RanBP1 and control cells treated with 200 ng/ml Dox). Results represent the mean of three individual experiments ± SD. Scale bars, 10 μm.
GTP by RanBP1 overexpression was sufficient to block ciliogenesis. In contrast, increasing the level of Ran GTP by RanBP1 knockdown appears to promote ciliogenesis in RPE cells. In a polarized cell such as IMCD3, modulating Ran GTP levels did not appear to affect ciliogenesis but did affect trafficking of the Kif17 motor. Thus careful control of Ran GTP levels may be necessary for optimal protein trafficking into and within the cilia. Perturbation of trafficking induced by lowering Ran GTP levels may be sufficient to prevent ciliogenesis, especially in nonpolarized cells. However, other explanations are possible for Ran-mediated modulation of ciliogenesis. For example, in spindle formation, the Ran importin system releases spindle assembly factors such as HURP and TPX2 that promote formation of the spindles (Kalab and Heald, 2008). In a similar manner, Ran and importins could regulate ciliary microtubular dynamics during cilia formation in addition to having effects on ciliary transport. Regardless of mechanism, it is clear from our studies that there is a coordinated regulation of Ran GTP levels at the centrosome and cilia that is essential for proper cilia formation and function.

**MATERIALS AND METHODS**

**Constructs**

Human RanBP1 was amplified by PCR from a HeLa cDNA library and subcloned into pRetroX-Tight-Pur vector using BamHI and EcoRI. Later, a FLAG or EGFP tag was inserted at the N-terminus of RanBP1. Myc-tagged Ran was generated by PCR from a human expressed sequence tag (EST; American Type Culture Collection, Manassas, Virginia) and then subcloned into pcDNA3.1 Zeo(+) vector (Invitrogen, Carlsbad, CA) using BamHI and XhoI. Myc-RanQ69L and Myc-RanT24N were generated by PCR mutagenesis. GST-importin β was generated by PCR from a full-length EST (mouse importin β; Open Biosystems, Huntsville, AL) and cloned into pGEX-4T3 (GE Healthcare, Piscataway, NJ) using BamHI and XhoI. KIF17-mCitrine (mCIt) was developed by the Verhey laboratory (Dishinger et al., 2010).

**Cell culture**

HEK 293T and MDCK II cells were grown in DMEM/high-glucose medium; IMCD3 and TERT RPE cells were grown in DMEM/F12 medium. All culture media were supplemented with 10% fetal bovine serum (FBS), 100 U/ml penicillin, 100 μg/ml streptomycin sulfate, and 2 mM l-glutamine.

Human primary airway epithelial cells were isolated from the tracheobronchial segments of donor lungs obtained at the time of double-lung transplantation and were cultured on collagen-coated plates using bronchial epithelial growth media (BEGM) (Cambrex Bioscience, Walkersville, MD). To differentiate cultures into mucociliary epithelium, passage 1 cells were seeded on collagen-coated Transwell filters and submerged in BEGM until the cells were confluent. Cells were then shifted to an air–liquid interface and maintained in a 1:1 mixture of BEGM and DMEM for 3 wk (Sajjan et al., 2004; Schneider et al., 2005).

**FIGURE 6:** RanBP1 knockdown increases Ran GTP at centrosomes and initiates primary cilia formation. (A) RanBP1 knockdown increases Ran GTP at centrosomes. Control or RanBP1 knockdown TERT RPE cells were cultured in complete growth media for 48 h. Centrosome fractions were isolated and pulled down using full-length GST-importin β1. Proteins bound to GST-importin β1 were eluted and immunoblotted with the indicated antibodies. (B) Quantification of Ran GTP immunoblots (GE Healthcare Typhoon scanner) from the experiment shown in A. (C) Ran GTP is enriched at the basal bodies of primary cilia in RanBP1-knockdown cells. Control or RanBP1-knockdown TERT RPE cells were grown on chamberslides in complete media for 48 h. Cells were fixed and stained with anti–acetylated tubulin (red) and rabbit anti–Ran GTP (green). Scale bars, 10 μm.

**TABLE:**

| Input | GST-importin β1 |
|-------|-----------------|
| Control shRNA | RanBP1 shRNA-1 |
| RanBP1 shRNA-1 | RanBP1 shRNA-2 |

**FIGURE CAPTION:** Figures 6A and C were generated using ImageJ software (Wayne Rasband, National Institutes of Health).

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**AUTHOR CONTRIBUTIONS**

K. L. L., S. F., B. L. M., and S. A. W. designed the experiments. K. L. L., S. F., B. L. M., E. W., and J. L. L. conducted the experiments. K. L. L. and S. A. W. analyzed the data. K. L. L. and S. A. W. wrote the paper. K. L. L. and S. F. managed the project.

**SUPPLEMENTARY MATERIAL**

Supplementary Material is available at G3: Genes | Genomes | Genetics online.

**SUPPLEMENTARY DATA**

The data that support the findings of this study are available from the corresponding author upon reasonable request.
purchased from BD Biosciences (San Diego, CA). Rabbit anti-RanBP1 and rabbit anti-Myc were from Bethyl Laboratories (Montgomery, TX), and mouse anti-Myc (clone 4A6) was from Millipore (Billerica, MA). Goat anti-mouse and goat anti-rabbit secondary antibodies for immunofluorescence were from Molecular Probes (Invitrogen). Horseradish peroxidase (HRP)–conjugated antibodies for immunoblots were purchased from GE Healthcare.

**Protein knockdown**

Human RanBP1 sequences GGCGAAAACT-GTTCGGATTGG and GGCGCATCGC-CTCCTCATGC were selected as RanBP1 shRNA targeting sequences for TERT RPE cells. Mouse RanBP1 sequences GTG-CAACACGTTCCGGTTGTTG and GGACCATCCGCTTCTATTAGA were chosen for RanBP1 knockdown in IMCD3 cells. These sequences were ligated into pSIREN RetroQ (Clontech). A luciferase shRNA construct served as the control.

**Immunofluorescence and confocal microscopy**

We performed immunofluorescence as described previously. In brief, cells were fixed with 4% paraformaldehyde, followed by 0.25% Triton X-100 permeabilization. For ciliary staining, 0.05% Triton X-100 was used to maintain delicate ciliary architecture. For γ-tubulin staining, we used cold acetone to fix the cells for 4 min. We blocked the samples in 3% goat serum/phosphate-buffered saline (PBS) for 30 min. Primary antibody dilutions were as follows: 1:500 anti-Ran GTP, 1:500 anti-Ran GTP, 1:2000 anti–acetylated tubulin and anti–polyglutamylated tubulin, 1:1000 anti-γ-tubulin, and 1:1000 anti-FLAG. Secondary antibodies were diluted 1:2000. Parafomaldehyde-fixed mouse respiratory turbinate andolfactory sections were permeabilized with 0.1% Trion X-100 for 20 min before blocking and staining as described earlier. Most images were obtained using an inverted epifluorescence microscope (Eclipse TE2000U; Nikon, Melville, NY). In some cases, images were acquired using a Meta Laser Scanning Confocal Microscope (LSM 510; Carl Zeiss MicroImaging, Thornwood, NY).

**Immunoprecipitation and Western blotting**

MDCK and TERT RPE cells were lysed in Triton X-100 lysis buffer (1% Triton X-100, 20 mM Tris-HCl, pH 7.4, 150 mM NaCl, 1.5 mM MgCl$_2$, 10% glycerol). After centrifugation, supernatants were collected. For anti-Ran GTP immunoprecipitation, 2 μl of rabbit anti-Ran GTP serum was added to cleared lysates and incubated overnight at 4°C. Then 40 μl of 50% protein A-Sepharose was added and incubated for another 2 h at 4°C. Immune complexes were washed three times with HNTG buffer (20 mM 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid, pH 7.5, 150 mM NaCl, 0.1% Triton X-100, 10% glycerol) and once with PBS. Samples were eluted in LDS (lithium dodecyl sulfate) sample buffer and electrophoresed on 10% Bis-Tris Novex gels (Invitrogen). After transferring proteins...
to polyvinylidene fluoride, membranes were blocked with 5% bovine serum albumin–Tris-buffered saline (TBS) and incubated with primary antibodies for 1–4 h in blocking buffer. HRP goat anti-rabbit secondary antibodies were diluted in 5% nonfat milk–TBS and incubated with the blots for 1 h. For GST pull-down experiments, 20 μl of a 50% slurry containing full-length GST- importin β1 was added to the centrosomal extracts and incubated for 2 h at 4°C. The GST-importin β1 beads were washed, eluted, and electrophoresed on Bis-Tris gels as described earlier.

Centrosome fractionation

Centrosomes were isolated from fully polarized MDCK cells as described previously (Hurd et al., 2011). To isolate centrosomes from TERT RPE cells, cells were grown in either regular culture media or serum-free media for 48 h at intermediate cell density. Cells were incubated in 5 μg/ml nocodazole and 5 μg/ml cytochalasin B at 37°C for 90 min, washed, lysed, and centrifuged as described previously (Hurd et al., 2011). Centrosomes were extracted with Triton X-100 lysis buffer. GST-importin β1 pull-down experiments and anti–Ran GTP immunoprecipitations were performed as described earlier.

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