Pleiotropic Roles for the \textit{Plasmodium berghei} RNA Binding Protein UIS12 in Transmission and Oocyst Maturation

Katja Müller\textsuperscript{1,2,*}, Olivier Silvie\textsuperscript{3}, Hans-Joachim Mollenkopf\textsuperscript{4} and Kai Matuschewski\textsuperscript{1,2}

\textsuperscript{1} Department of Molecular Parasitology, Institute of Biology, Humboldt University, Berlin, Germany, \textsuperscript{2} Parasitology Unit, Max Planck Institute for Infection Biology, Berlin, Germany, \textsuperscript{3} Centre d’Immunologie et des Maladies Infectieuses, INSERM, CNRS, Sorbonne Université, Paris, France, \textsuperscript{4} Core Facility Microarray/Genomics, Max Planck Institute for Infection Biology, Berlin, Germany

Colonization of the mosquito host by \textit{Plasmodium} parasites is achieved by sexually differentiated gametocytes. Gametocytogenesis, gamete formation and fertilization are tightly regulated processes, and translational repression is a major regulatory mechanism for stage conversion. Here, we present a characterization of a \textit{Plasmodium berghei} RNA binding protein, UIS12, that contains two conserved eukaryotic RNA recognition motifs (RRM). Targeted gene deletion resulted in viable parasites that replicate normally during blood infection, but form fewer gametocytes. Upon transmission to \textit{Anopheles stephensi} mosquitoes, both numbers and size of midgut-associated oocysts were reduced and their development stopped at an early time point. As a consequence, no salivary gland sporozoites were formed indicative of a complete life cycle arrest in the mosquito vector. Comparative transcript profiling in mutant and wild-type infected red blood cells revealed a decrease in transcript abundance of mRNAs coding for signature gamete-, ookinete-, and oocyst-specific proteins in \textit{uis12(-)} parasites. Together, our findings indicate multiple roles for UIS12 in regulation of gene expression after blood infection in good agreement with the pleiotropic defects that terminate successful sporogony and onward transmission to a new vertebrate host.

Keywords: malaria, \textit{Plasmodium}, RNA binding protein (RBP), translational regulation, RNA, gametocyte, oocyst

INTRODUCTION

During the complex life cycle of \textit{Plasmodium}, malarial parasites switch between different host cells, extra- and intracellular life cycle forms, and an invertebrate and a vertebrate host. As the parasites progress in their developmental program, and particularly after switching host, adaptation to new environmental conditions, including temperature and immune defense, is central. For instance, \textit{Plasmodium} gametocytes must remain infectious in the blood of the vertebrate host until they are eventually transmitted to the vector during a blood meal. The gametocytes need to quickly adapt to new environmental factors, leave the digestive environment of the blood meal, and colonize the...
mosquito midgut (Billker et al., 1998; Billker et al., 2004). To swiftly accomplish a host switch, transcription and translation are tightly coordinated.

Transcriptional reprogramming is controlled by the apicomplexan Apetala-2 (Api-AP2) transcription factors (Balaji et al., 2005; Painter et al., 2011). Many genes needed for gametocytogenesis and early events of mosquito colonization are transcribed by the interplay of the Plasmodium transcription factors AP2-G and AP2-I, while asexual gene expression is repressed by AP2-G2 and AP2-FG controls female-specific gene regulation (Sinha et al., 2014; Yuda et al., 2015; Josling et al., 2020; Yuda et al., 2020). This transcriptional switch leads to the expression of mRNAs which are required for the multiple steps from gametocytogenesis to zygote formation until the motile ookinete stage is reached and the next checkpoint in transcription control is initiated by the AP2-O 1-4 family (Kaneko et al., 2015; Modrzynska et al., 2017). Therefore, a tight fine-tuning is essential to synthesize tailored proteins at the right time.

Translational repression occurs during host switch to keep gametocytes and sporozoites in a latent state. Accordingly, translation of pre-synthesized mRNAs, which are required for mosquito colonization, occurs only after transmission and constitutes a mechanism to be prepared for the new host environment. In gametocytes a large number of genes have been described to be translationally repressed (Le Roch et al., 2004; Mair et al., 2006; Lasonder et al., 2008; Mair et al., 2010; Zhang et al., 2010; Lasonder et al., 2016). Translational repression is particularly dominant in female gametocytes and affects a wide range of mRNAs, including those that encode surface proteins, e.g., P25, P28, and CCP2, transcriptional regulators, e.g., AP2-O, and proteases, e.g., plasmepsin IV. A conserved U-rich 47-nucleotide-long cis-acting consensus motif in the 5’ and 3’ UTRs of a subset of translationally repressed transcripts in P. berghei gametocytes was shown to be important in translational regulation in gametocytes (Hall et al., 2005; Braks et al., 2008). In the female gametocyte, the DX6-class DEAD box RNA-helicase DOZI (development of zygote impaired) is bound to its target transcripts in a large storage mRNP complex and silences their translation (Mair et al., 2006; Mair et al., 2010; Guerreiro et al., 2014). The Pumilio-family RNA binding protein PuF2 increases proliferation and decreases stage differentiation. As a consequence, deletion of P. berghei and P. falciparum PuF2 resulted in higher gametocyte rates and premature transformation of sporozoites in the mosquito salivary glands (Miao et al., 2010; Gomes-Santos et al., 2011; Muller et al., 2011; Lindner et al., 2013). In P. falciparum parasites, an up-regulation of many gametocyte specific transcripts, mostly independent of those that are regulated in dozil(-)/cith(-), was observed (Miao et al., 2010). Specific binding of pfs25, pfs28, and Plasmodium VI mRNA by the PfPuF2 protein was shown by RNA immunoprecipitation (Miao et al., 2013).

During host switch to the mosquito vector PuF2 negatively regulates gametocyte formation, and a post-transcriptional activator of gametocyte formation still remains elusive. In a complex with other proteins DOZI control of mRNA translation occurs much later, shortly after fertilization of female gametocytes inside the mosquito. Therefore, it is likely that additional factors participate in the regulation of post-transcriptional control of gene expression during mosquito transmission. In the present study we characterized an RNA binding protein, originally identified as up-regulated in infectious sporozoites gene 12 (UIS12), which is also highly expressed in gametocytes. A suppression subtractive hybridization screen aimed at discovering genes which are up-regulated in salivary gland sporozoites first described the P. berghei RNA binding protein UIS12 (PBANKA_0506200) (Matuschewski et al., 2002). As expected, protein prediction programs indicate an intracellular localization. Comparison of transcriptome and proteome data indicate that P. berghei UIS12 is expressed in sporozoites, but the corresponding protein is not detectable, suggesting that UIS12 expression is regulated post-transcriptionally (Lasonder et al., 2008; Le Roch et al., 2004; Lindner et al., 2019). Due to the presence of the two RRM motifs, UIS12 is likely involved in post-transcriptional control of gene expression in gametocytes and pre-erythrocytic stages. Employing experimental genetics, the role of UIS12 for parasite life cycle progression was analyzed with an emphasis on a potential function in post-transcriptional regulation.

RESULTS

The RNA Binding Protein PbUIS12 Is Expressed at Multiple Life Cycle Stages

We initiated our analysis by an NCBI conserved domain database (CDD) search (Marchler-Bauer et al., 2015), which revealed two ~70 amino acid long RNA-recognition motifs (RRM), belonging to the class of RNA-binding domains (RBD) (Dreyfuss et al., 1988), in the P. berghei UIS12 protein sequence. The UIS12 gene, and particularly the two RRM domains, are highly conserved between different Plasmodium UIS12 orthologs (Figure 1A), while in a candidate Toxoplasma gondii ortholog (TGME49_268380) the conservation is limited to the two RRM domains only. In general, the amino- and carboxy-terminal regions are less conserved, and all Plasmodium ortholog proteins have similar sizes of ~1,300 amino acid residues. The PbUIS12 coding region has a total length of 4,070 base pairs and consists of two exons (Figure 2A). Of note, the two regions encoding the RRM domains are separated by the single intron, which also exhibits a degree of conservation.

We next measured UIS12 transcript abundance throughout the P. berghei life cycle by quantitative RT-PCR (Figure 1B). In good agreement with the previous notion of up-regulation in salivary gland sporozoites, UIS12 mRNA steady state levels are low in midgut oocysts and highest in salivary gland sporozoites. High UIS12 transcript levels were also detected in early (24h) liver stages. In mixed asexual blood stages UIS12 mRNA levels are low, whereas in schizonts, gametocytes and ookineties, considerable expression was detected. Our observations mostly corroborate published data (Howick et al., 2019; Otto et al., 2014). However, we did not specifically analyze ring stages or...
distinguished between male and female gametocytes (Yeoh et al., 2017). Together, UIS12 is expressed at multiple points of the *P. berghei* life cycle, indicative of multiple functions during life cycle progression and parasite stage conversion.

**Targeted Gene Disruption of the RNA-Binding Protein PbUIS12**

In order to study the gene function, we generated a UIS12 loss-of-function mutant. *PbUIS12* deletion was generated by double homologous recombination employing a replacement strategy, as described previously (van Dijk et al., 1995) (Figure 2A).

The UIS12 targeting construct (pΔPANKA_UIS12) comprised ~500 bp each of the 5' and 3' non-coding sequences flanking a *TgDHFR/TS* pyrimethamine resistance cassette for positive selection with the anti-malarial drug pyrimethamine. Upon a double homologous recombination event, the UIS12 gene is expected to be replaced by the selectable marker (Figure 2A). As recipient parasites we used *P. berghei* (strain: ANKA; clone: cl507), which expresses the fluorescent protein GFP under the control of the constitutive elf1a promoter (van Dijk et al., 1995). After transfection we recovered parental *uis12(-)* blood stage populations, from which clonal lines were generated by injection of limiting serial dilutions into mice. The desired recombination event was confirmed by diagnostic PCR, which confirmed the presence of 5' and 3' integration in the parental and the clonal *uis12(-)* populations, and by the absence of wild-type-specific PCR fragments in the clonal *uis12(-)* populations. During the course of our UIS12 knockout analysis and to further corroborate our findings we successfully generated an independent second *uis12(-)* knockout line (*uis12(-) clone 2*), in a different *P. berghei* ANKA recipient line, Bergreen, which constitutively expresses GFP under the control of the *HSP70* promoter (Kooij et al., 2012) (Figure 2B: clone 2; Figure S1A: clone 1).

At this stage, we could already conclude that the successful generation of clonal *uis12(-)* knockouts demonstrates that the gene is not essential for normal progression of asexual blood stages, although *UIS12* is expressed considerably in schizonts (Figure 1B).

**Impaired Gametocyte Development of *uis12(-)* Parasites**

After successful selection of *uis12(-)* parasites, we first examined asexual and sexual blood stage growth. To this end, C57BL/6 mice (*n*=5) were infected by intravenous injection of either 10,000 *uis12(-)* or WT-infected erythrocytes. Starting three days after infection the mice were monitored by daily microscopic examination of thin blood films, and parasitemia and gametocytemia were quantified (Figures 2C, D). Parasitemia increased daily in wild-type- and *uis12(-)*-infected mice to a similar extent. On day 7, wild-type and *uis12(-)*-infected mice started developing symptoms indicative of onset of experimental cerebral malaria, supporting the notion that *uis12(-)* parasites display virulence typically seen in infections with *P. berghei* ANKA parasites.

The blood films were also analyzed for the presence of male and female gametocytes (Figure 2D). This analysis revealed that, in contrast to normal asexual blood infection, gametocytemia was significantly reduced in *uis12(-)* parasites as compared to the wild-type. *UIS12* gametocyte numbers were already slightly reduced on day four after infection, when the first fully mature gametocytes became visible. The difference between WT and *uis12(-)* parasites increased further over time. Examination of size and morphology of male and female gametocytes and ookinetes revealed no difference between the two parasite lines (Figure 2E). The presence of ookinetes indicates that *uis12(-)* gametocytes are able to fertilize and subsequently form a zygote that develops into an ookinet.
Both, male and female gametocytes were found in the expected female biased proportions. Together, the data indicate that UIS12 is dispensable for asexual growth, but plays an important role during gametocytogenesis. Analysis of the second uis12(-) clone corroborated these findings (Figures S1B, C).

**Exflagellation Is Markedly Reduced in uis12(-) Parasites**

The potential of male gametocytes to produce motile microgametes is an active process and termed exflagellation. In both uis12(-) clonal lines, exflagellation events were detected very rarely by microscopic examination of undiluted blood from uis12(-) infected mice at any given time point. Yet, this already showed that uis12(-) male gametocytes are able to produce motile male gametes, albeit at very low frequencies. To quantify the exflagellation centers, C57BL/6 mice (n=4) were injected intravenously with a high dose of 10^7 blood stages. Four days later, parasitemia, gametocytemia and the number of exflagellation centers were enumerated (Figure 2F, uis12(-) clone 1). Interestingly, in this experiment, the parasitemia was slightly, but significantly higher in uis12(-)-infected mice. In
agreement with our first experiment (Figure 2D), gametocytemia was 8.6-fold reduced in uis12(-) parasite-infected mice. Exflagellation was not visible under these conditions, i.e., 1:25 diluted blood, in uis12(-)-infected mice (Figure 2F). Even after an extended incubation or when examined the following days, exflagellation was not detectable. In conclusion, male uis12(-) gametocytes retain the ability to progress to functional microgametes, but at a very low rate. The defect in exflagellation is more severe than the reduction in gametocytemia indicating additive defects in absence of UIS12 upon progression from blood infection to transmission to the insect vector.

**Reduced Oocyst Formation By uis12(-) Gametocytes, But Failure to Produce Sporozoites**

The severe defect in *in vitro* exflagellation implied a large impact in mosquito colonization. However, after transmission to *Anopheles stephensi* mosquitoes, uis12(-) parasites were able to establish midgut infections and produce oocysts, showing that early events of midgut colonization are not abolished *in vivo* (Figure 3). The mean mosquito infection prevalences were 54% (± 18%) for wild-type (*n* = 5) and 46% (± 23%) for uis12(-) (*n* = 5).

This finding prompted us to quantify oocyst maturation. To this end, midguts were isolated daily starting on day 4 after mosquito infection up until day 14. Oocyst numbers and morphology as well as their ability to produce sporozoites were analyzed by fluorescence microscopy (Figure 3, uis12(-) clone 2). In uis12(-)-infected mosquitoes, the number of oocysts per midgut was more than 10-fold reduced as compared to wild-type, consistent with reduced gametocyte numbers and the very low exflagellation rate (Figure 3B). Strikingly, in the absence of UIS12 oocysts were markedly smaller in size (Figures 3A, C). While up until day 4 the oocyst size was comparable between both groups, over time uis12(-) oocysts displayed an abnormal

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**Figure 3** Disruption of UIS12 leads to a reduced oocyst size and a defect in sporozoite formation. (A) Shown are representative live micrographs of uis12(-) and wild-type oocysts 7, 10 and 14 days after an infectious blood meal. GFP (green) is constitutively expressed under the HSP70 promoter (*P. berghei* ANKA Bergreen) (Kooij et al., 2012). Uis12(-) clone 2 was used for this experiment. nuclei were stained with Hoechst (blue). Note the reduced density and size of uis12(-) oocysts, and absence of sporozoites in uis12(-) oocysts. Magnification, 630-fold; scale bars, 10 µm. (B, C) Time course of oocyst numbers per infected mosquito midgut (B) and oocyst size (µm²) (C) at different time points after midgut infection (uis12(-) clone 2, red; WT, black). *p < 0.05 (multiple t-tests).*
growth pattern. Their size remained unchanged, whereas the WT oocyst diameter increased ~10-fold over the following ten days. This finding indicates that the early events of oocyst development, including transformation of the motile ookinete to the spherical oocyst, and the initial increase in biomass (Carter et al., 2007) are reduced, but not completely abolished. An additional defect leads to arrested oocyst maturation. The analysis of the second usi12(-) clone corroborated these findings (Figures S1D–F).

Upon close examination at later time points, usi12(-) oocysts displayed little or no nuclear staining, harbored many large vacuoles, and frequently were surrounded by an apparently wrinkled oocyst wall (Figure 3A, Figure S1F). As a consequence, no sporozoites were seen inside usi12(-) oocysts. This lack of sporozoite formation was confirmed by analysis of extracted midguts; no midgut- associated sporozoites were detectable. In WT-infected midguts sporoblast formation was observed inside the oocysts starting around day 11 after infection, and from day 12 onwards oocysts started to fill with sporozoites (Matuschewski, 2006) (Figure 3A). In contrast, usi12(-) midguts remained free of sporozoites even until late time points of mosquito infection, excluding delayed sporozoite formation.

In good agreement, initial microscopic screening of salivary gland extracts of usi12(-)-infected mosquitoes for presence of sporozoites did not return a single sporozoite. To obtain independent support for the complete absence of sporozoites in usi12(-)-infected mosquitoes, we inoculated C57BL/6 mice with salivary gland extracts of 20 usi12(-)-infected mosquitoes. In addition, we exposed C57BL/6 mice to ~100 usi12(-)-infected mosquitoes (Table 1, both clones). In sharp contrast to control infections with wild-type-infected mosquitoes under similar conditions, none of the attempts to transmit usi12(-) parasites resulted in blood stage infections (Table 1). This defect in transmission confirms that usi12(-) parasites are unable to complete their development inside the mosquito.

**Deregulation of Transcripts Important for Mosquito Transmission in usi12(-) Blood Stages**

We hypothesized that due to the presence of the two RRM motifs, UIS12 is likely involved in post-transcriptional control of gene expression. Reduced gametocyte production, exflagellation, and oocyst development detected in usi12(-) parasites suggests that UIS12 might be regulating target mRNAs and alter expression of the respective proteins, which in turn play important roles in these stages.

To profile the transcriptome of usi12(-) parasites, we performed a comparative microarray analysis on usi12(-) and wild-type mixed blood stage RNAs (Figure 4, Figure S2). Since the first defect is detected during gametocyte formation, we limited our analysis to mixed blood stages, that include the asexual blood stages, from which gametocytes originate. Attempts to retrieve sufficient usi12(-) gametocytes for a microarray failed. The microarray analysis was performed with two biological replicates (R1 and R2), both with usi12(-) clone 1. For both replicates, parasitemia of the samples were comparable (R1: WT 6.2%; usi12(-) 6.3%; R2: WT 6.4%; usi12(-) 8.7%). As expected, gametocytemia was reduced in usi12(-) parasites (R1: WT 0.2%; usi12(-) 0.06%). Each microarray experiment was performed by dual-color hybridizations, and a color-swap dye-reversal was included in order to compensate for dye-specific effects. In this microarray analysis, the -fold change of the expression levels of 2,890 P. berghei genes could be successfully compared (Figure 4A, Figures S2A, D).

Overall, absence of PbUIS12 correlated with a global perturbation of 30% of all mRNA expression levels (threshold, <2 or >2). More transcripts were down-regulated (646, 23%) than up-regulated (199, 7%). The mean down-regulation was -6.4-fold, whereas the mean up-regulation was +2.5-fold (GEO Series accession number: GSE152686). As expected, the UIS12-knockout was independently confirmed by the microarray analysis; US12 transcripts were down-regulated by ~22.3-fold (Table 2). Within the 25 most down-regulated transcripts known proteins required for the switch to the mosquito vector, CCP1 (LAP2), LAP5, P48/45, and P28, as well as numerous previously unrecognized proteins are represented (Table 2). The list of the 25 most up-regulated genes contains, among others, PIR proteins (Table S1). Remarkably, many transcripts that were down-regulated more than 10-fold in usi12(-) blood stages, are reported to be expressed in gametocytes (Hall et al., 2005). We also analyzed the proportions of up-, down-, and non-regulated transcripts and found distinctions between stage-specific RNAs, as defined by RNA sequencing data (Otto et al., 2014). For instance, gametocyte transcripts were defined by >2-fold up-regulation in gametocytes as compared to asexual blood.

**TABLE 1 | No malaria transmission by usi12(-)-infected mosquitoes.**

| Parasites          | Route of injection | Infected/Injected | Pre-patency (days) |
|--------------------|--------------------|-------------------|-------------------|
| WT 507cl           | 10,000 sporozoites i.v. | 5/5               | 3                 |
| usi12(-) clone 1   | mosquito bite (~100 mosquitoes) | 0/5               | N.A.              |
|                   | salivary gland extract i.v. (20 mosquitoes) | 0/4               | N.A.              |
| WT Bergreen        | mosquito bite (~100 mosquitoes) | 3/3               | 3                 |
| usi12(-) clone 2   | mosquito bite (~100 mosquitoes) | 0/3               | N.A.              |
|                   | salivary gland extract i.v. (20 mosquitoes) | 0/3               | N.A.              |

1 C57BL/6 mice were exposed to the bites of ~100-infected mosquitoes for 15–20 min or injected intravenously (i.v.) with salivary gland extract isolated from 20 mosquitoes (usi12(-)) and, as a control, 10,000 sporozoites (WT). Data are pooled from two separate experiments.

2 The pre-patent period represents the days until detection of the first infected erythrocytes by microscopic thin blood film examination after sporozoite inoculation. N. A. not applicable. The colors were used to highlight different conditions.
In good agreement with normal replication rates, very few asexual blood stage-specific transcripts were down-regulated in *uis12(-)* parasites. Instead, the proportion of up-regulated transcripts was highest for asexual blood stage-specific mRNAs, which might reflect over-representation of these stages due to defects in gametocytogenesis. However, signature transcripts, like *AMA1* (-0.3), *HSP70* (-1.6), *MSP1* (1.7) and *MSP8* (1.5), were neither.

**Figure 4B** | Down-regulation of distinct mRNAs coding for signature gamete-, ookinete- and oocyst-specific proteins in *uis12(-)* blood stage parasites. Shown are mean values of microarray results of total RNA isolated from mixed infected erythrocytes from *uis12(-)* (clone 1) - and WT-infected mice from two biological replicates. (A) Shown is a volcano-plot illustrating the fold change of the expression levels and the negative log *p*-values of all analyzed 2,890 *P. berghei* genes. The dotted black line represents a *p*-value of 0.05 and all transcripts with a *p*-value <0.05 are shown in red. Exemplary transcripts are highlighted and labeled in blue. (B) Pie charts displaying the proportions of up- (green >=2), non- (grey), and down- (orange <=-2) regulated transcripts among all transcripts (upper left), blood stage-specific transcripts (center), and gametocyte-specific transcripts (upper right) (Otto et al., 2014). Exemplary transcripts are listed in the respective region of the chart. The number of transcripts analyzed is shown in a white circle inside the center. (C) Pie charts displaying the proportions of up- (green >=2), non- (grey), and down-(orange <=-2) regulated transcripts among female (left) and male (right) gametocyte-specific transcripts (Yeoh et al., 2017). The number of transcripts analyzed is shown in a white circle inside the center.

**Table 2** | The top 25 down-regulated genes in blood stages of *uis12(-)* vs. wild-type.

| Gene ID       | Product description                        | Mean -fold change R1<sup>a</sup> | Mean -fold change R2<sup>b</sup> | -fold change R1<sup>a</sup> | -fold change R2<sup>b</sup> |
|---------------|-------------------------------------------|----------------------------------|----------------------------------|-----------------------------|-----------------------------|
| PBANKA_0806800 | unknown                                   | -41.0                            | -37.5                            | -44.4                       | -47.0                       |
| PBANKA_1449000 | candidate microgamete surface protein MiGS| -36.2                            | -25.4                            | -36.3                       | -47.0                       |
| PBANKA_1349400 | candidate DNA replication complex GINS protein| -31.6                         | -29.0                            | -34.2                       | -35.9                       |
| PBANKA_1340400 | candidate lactate dehydrogenase           | -30.2                            | -24.2                            | -36.3                       | -38.3                       |
| PBANKA_1400400 | fam-b-protein                             | -28.8                            | -14.3                            | -43.4                       | -43.4                       |
| PBANKA_1361600 | E1-E2 ATPase                              | -25.9                            | -25.9                            | -25.9                       | -26.9                       |
| PBANKA_1040300 | fam-b-protein                             | -25.4                            | -18.5                            | -32.3                       | -32.3                       |
| PBANKA_1315300 | LCCL domain-containing protein (LAPS)      | -24.4                            | -27.4                            | -21.4                       | -21.4                       |
| PBANKA_0413000 | unknown                                   | -23.6                            | -29.3                            | -18.0                       | -18.0                       |
| PBANKA_1246600 | cleft-like protein 1                      | -22.9                            | -9.7                             | -36.2                       | -35.0                       |
| PBANKA_0506200 | RNA binding protein (UIS12)               | -22.3                            | -31.7                            | -12.8                       | -12.8                       |
| PBANKA_0514900 | ookinete surface protein P28              | -21.6                            | -15.5                            | -27.8                       | -27.8                       |
| PBANKA_1300700 | LCCL domain-containing protein (CCp1)      | -21.1                            | -20.7                            | -21.4                       | -21.4                       |
| PBANKA_0719700 | unknown                                   | -20.7                            | -19.2                            | -22.2                       | -22.2                       |
| PBANKA_0707100 | inner membrane complex protein 1i         | -20.7                            | -26.5                            | -15.0                       | -15.0                       |
| PBANKA_1334900 | unknown                                   | -20.4                            | -20.1                            | -20.7                       | -20.7                       |
| PBANKA_1353800 | unknown                                   | -20.1                            | -26.1                            | -14.1                       | -14.1                       |
| PBANKA_1432400 | perforin-like protein 2                   | -19.4                            | -16.9                            | -21.9                       | -21.9                       |
| PBANKA_0608600 | candidate EGF-domain protein              | -19.0                            | -23.9                            | -14.0                       | -14.0                       |
| PBANKA_1400900 | unknown                                   | -18.9                            | -20.3                            | -17.5                       | -17.5                       |
| PBANKA_1359600 | 6-cysteine protein (P48/45)               | -18.6                            | -21.0                            | -16.2                       | -16.2                       |
| PBANKA_1106000 | unknown                                   | -17.9                            | -22.2                            | -13.5                       | -13.5                       |
| PBANKA_1414800 | unknown                                   | -17.9                            | -19.6                            | -18.1                       | -18.1                       |
| PBANKA_1120400 | candidate inner membrane complex protein 1| -17.8                            | -18.9                            | -18.7                       | -18.7                       |
| PBANKA_1318600 | unknown                                   | -17.8                            | -15.9                            | -19.7                       | -19.7                       |

<sup>a</sup>biological replicate 1.
<sup>b</sup>biological replicate 2.
The colors were used to highlight different conditions.
TABLE 3 | Examples for deregulation of signature vector-stage transcripts in uis12(-) parasites.

| Known blood stage transcripts | Known transmission stage transcripts |
|-------------------------------|-----------------------------------|
| Up-regulated                  |                                   |
| Pb235                         | 2.8                               |
| AMA1                          | -0.3                              |
| MSP8                          | 1.5                               |
| HSP70                         | -1.6                              |
| SERA3                         | 1.7                               |
| Unaffected                    |                                   |
| MSP1                          | 1.7                               |
| Down-regulated (< 10-fold)    |                                   |
| CITH*                         | -2.3                              |
| Actin II                      | -2.8                              |
| CDPK4                         | -3.1                              |
| MAPK 2                        | -3.3                              |
| GAP45                         | -6.1                              |
| AP2-0                         | -5.7                              |
| CDPK3                         | -6.8                              |
| Plasmepsin VI                 | -6.3                              |
| HAP2                          | -7.5                              |
| MDV1                          | -10                               |
| Down-regulated (> 10-fold)    |                                   |
| COp6*                         | -11.2                             |
| COp4*                         | -12.2                             |
| COp3*                         | -15                               |
| P47                           | -15                               |
| COp2*                         | -15                               |
| P25                           | -17.1                             |
| P48/45                        | -18.8                             |
| COp1*                         | -21.1                             |
| P28                           | -21.6                             |

Shown are exemplary blood and transmission stage transcripts. The mean -fold change represents steady-state mRNA levels in uis12(-) parasites compared to the corresponding WT parasite stages. *genes with a knock-out phenotype similar to uis12(-).

up- nor down-regulated in the absence of UIS12 (Figure 4B, Table 3). We also noted that the mRNA levels of many signature mosquito stage-specific proteins were down-regulated in uis12(-) mixed blood stages. In addition, we also compared the proportions of up-, down- and non-regulated transcripts and found distinctions between male- and female-specific transcripts, as defined by RNA sequencing data (Yeoh et al., 2017) (Figure 4C, Figures S2C, F). While 67% of the female gametocyte transcripts were down-regulated in absence of UIS12, only 43% of male-specific transcripts were down-regulated, indicating differential impact of UIS12 on male and female gametocyte-specific transcripts in mixed blood stages.

Next, we wanted to independently confirm the microarray results by quantitative RT-PCR of selected transcripts on blood stage and gametocyte mRNA from uis12(-) and WT samples (Figure 5). This analysis revealed a good agreement with the mean -fold changes determined in the microarray for gametocyte- and blood stage-specific transcripts. The gametocyte-expressed transcripts Puf1, DOZI, UIS1/IK2, SET, and Actin II (Mair et al., 2006; Pace et al., 2006; Deligianni et al., 2011; Gomes-Santos et al., 2011; Muller et al., 2011) showed similar -fold changes in both assays. Similarly, the blood stage-expressed transcripts AMA1 and MSP1 remained un-regulated. For some transcripts, e.g., male development factor (MDV1) (Lal et al., 2009), the reduction in uis12(-) parasites was more pronounced by quantitative RT-PCR in comparison to the microarray data. qRT-PCR on gametocyte mRNA samples were consistent with the data from asexual blood stages (Figure 5, uis12(-) clone 1). Overall, we could confirm the results of the microarray on these samples, indicating that the down-regulation of the gametocyte transcripts is a direct result of a specific UIS12-dependent regulation and not merely a consequence of the absence of gametocytes in uis12(-) mixed blood stages.

Gene Ontology Analysis of the Deregulated Transcripts in uis12(-) Parasites

We next performed a gene ontology (GO) enrichment analysis (Ashburner et al., 2000) (Tables S2 and S3). According to this analysis the 199 up-regulated and the 646 down-regulated transcripts are involved in 21 and 42 different biological processes, as described by distinct GO terms, respectively. Examples for GO terms of the set of down-regulated genes include reproduction, cell communication and phosphorylation (Table S2), in good agreement with the known roles of kinases in gametogenesis (Invergo et al., 2017). Examples for GO terms of the set of up-regulated genes include RNA processing and histone acetylation (Table S3), further indicative of perturbations in gene expression and mRNA translation in absence of UIS12.

A 10-Nucleotide Motif Shared By Up-Regulated Transcripts in uis12(-) Parasites

Since UIS12 contains two RNA recognition motifs and the combination of at least two RRM domains allows the continuous recognition of a 8-10 nucleotide-long sequence
motif with high affinity (Maris et al., 2005), we screened all ORFs, and 1,000 base pairs of the flanking 5’ UTR and 3’ UTR sequences of genes up-regulated >2.5-fold in replicate 1 for common 8-10 nucleotide-long motifs using MEME (Bailey et al., 2009) using the respective portions of the non-regulated transcripts as reference. While this search did not return a motif in either the 5’ or 3’ UTR sequences, we found a 10 nucleotide-long motif (TTTYTTTTTC) in the ORF (Figure 6A). This signature was found 751 times in 174 of the 220 up-regulated sequences (e-value: 4.2x10^-4). We restricted our search to up-regulated transcripts, since down-regulation can at least be partially attributed to fewer gametocytes in mixed usi12(-) blood stages.

**DISCUSSION**

In this study, deletion of the candidate RNA binding protein UIS12 led to the discovery of a potential post-transcriptional regulator of gametocytogenesis and mosquito transmission. Our results underscore the key roles of a fine-tuned transcriptional control and post-transcriptional regulation during transmission of the malaria parasite between hosts. When UIS12 is absent, fewer gametocytes are produced and in vitro exflagellation is severely impaired, yet not abolished. The defects of usi12(-) parasites during host switching cannot be compensated for in subsequent steps in the *Plasmodium* life cycle, and usi12(-) infection leads to fewer oocysts, which fail to undergo oocyst maturation. We can correlate these defects with a set of signature sequences (e-value: 4.2x10^-4). We restricted our search to up-regulated transcripts, since down-regulation can at least be partially attributed to fewer gametocytes in mixed usi12(-) blood stages.

**Figure 6** A motif is shared by up-regulated transcripts in *usi12(-)* parasites, and the comparison of dozi(-), cith(-) and usi12(-) target transcripts reveals possible overlapping functions. (A) Shown is a graphic display of the 10-nucleotide U-rich motif found in the ORF of transcripts that are up-regulated in absence of UIS12 in the microarray with biological replicate 1. The size of the depicted nucleotide represents its probability at the respective position. (B) Venn diagram comparing shared down-regulated transcripts of the gametocyte microarray analyses performed for dozi(-) and cith(-) with a -fold change lower than -1 (Mair et al., 2010) and down-regulated transcripts in mixed blood stages of *usi12(-)* with a mean -fold change lower than -2.
function in sporozoites and prepares them for transmission to the vertebrate host (Miao et al., 2010; Gomes-Santos et al., 2011; Muller et al., 2011). We note that UIS12 likely executes a second function, similar to the one analyzed in the present study, in pre-erythrocytic stages, the stage where it was originally identified (Matuschewski et al., 2002), but the complete block in sporogony prevented us from studying this by classical reverse genetics. It, thus, still remains unclear if UIS12 also plays a role in the host switch back to the mammalian host. Of note, in sporozoites translational repression of UIS12 was described, and the protein is only detectable in liver stages (Lindner et al., 2019). Accordingly, a stage-specific knockout, which depletes UIS12 during sporogony is required to study its cellular roles for sporozoite transmission and liver infection.

The few remaining uis12(-) gametocytes were able to form male and female gametes, fertilize, and form ookinetes. Accordingly, numbers of oocysts are extremely low in uis12(-)-infected Anopheles mosquitoes. Complete life cycle arrest in uis12(-) infections occurs at an early time point after oocytes are formed, indicating that the events leading to oocyst formation are not impaired (Carter et al., 2007).

Our microarray analysis revealed a sharp drop in the steady state mRNA levels of numerous well-characterized Plasmodium genes. The P. berghei LCL1 protein family members CCP1 and lecin adhesive-like protein 5 (LAP5) are expressed in female gametocytes and ookinetes. PbLAP proteins are important for sporogenesis inside oocysts, yet known to be expressed already in gametocytes (Pradel et al., 2004; Trueman et al., 2004; Raine et al., 2007; Lavazec et al., 2009; Saeed et al., 2010). The surface proteins of the 6-cysteine repeat protein family P48/45 are essential for male gametocyte fertility (van Dijk et al., 2001). P25 and P28 are surface proteins of sexual stages, with distinct roles in ookinetes to oocyst transformation (Siden-Kiamos et al., 2000; Tomas et al., 2001). Interestingly, some transcripts that are severely depleted, including P28, AP2-O, CCP2, and CCP4, have been reported to be translationally controlled (Verpenne et al., 1994; Saeed et al., 2013; Lasonder et al., 2016). Strikingly, the function of some of these repressed targets extends far beyond the ookinete stage, and the corresponding proteins are important for oocyst and sporozoite formation, illustrating the complexity of regulation of post-transcriptional control of gene expression in gametocytes (Lasonder et al., 2016). A deregulation of mRNAs already in mixed blood stages in absence of UIS12 might partially explain the observed inability to complete oocyst development. However, the abundant UIS12 mRNA levels in ookinetes suggest UIS12 presence at that stage and a potential additional checkpoint of post-transcriptional control by UIS12 possibly after transcription of mosquito stage-specific target genes is orchestrated by a family of closely related Apetala-2 proteins, AP2-O 1-4, in ookinetes (Kaneko et al., 2015; Modrzynska et al., 2017). And this additional checkpoint in ookinetes might be the reason for the pleiotropic effects observed upon absence of UIS12, finally resulting in the aberrant maturation of oocysts. Due to the low UIS12 transcript levels in oocysts a direct role at this point in the life cycle is less likely. We, thus, propose that UIS12 exerts multiple regulatory roles at consecutive life cycle stages, and, hence, is together with Puf2 one of the first known Plasmodium RNA binding proteins with pleiotropic functions for life cycle progression (Gomes-Santos et al., 2011; Lindner et al., 2013; Miao et al., 2013; Miao et al., 2010; Muller et al., 2011).

The GO-term enrichment of down-regulated transcripts in uis12(-) shows perturbation of many kinases and GO-term enrichment of up-regulated transcripts revealed perturbation of gene expression. A MEME search in the ORFs, 5′ UTRs and 3′ UTRs of the up-regulated transcripts identified in the microarray did reveal a 10-nucleotide U-rich motif in the ORFs unique to the analyzed subsets when compared to sequences that were not perturbed in absence of UIS12. This 10 nucleotide-long U-rich motif might represent a signature of UIS12 interaction with target mRNAs during blood infection, which might be required for proper preparation for host switching. We wish to point out that down-regulation of gametocyte signature transcripts in uis12(-) blood stages can be largely attributed to the at least 3.5-fold reduction in gametocyte numbers, yet the levels of down-regulation ranged from a low (~2, e.g., Puf2) to more than 15-fold (e.g., CCP1 and CCP2). By qPCR we could confirm the down-regulation of some of the gametocyte specific transcripts such as P28 in a gametocyte enriched mRNA preparation of uis12(-) blood stage parasites, indicating that the down-regulation of gametocyte transcripts might not only be a consequence of the absence of gametocytes in uis12(-) mixed blood stages, but a direct result of a specific UIS12-dependent regulation.

Further analyses employing RNA-Seq or single cell transcriptomics on synchronized and purified stages, ranging from ring stages to ookinetes, are warranted to gain an in-depth understanding of the mRNA repertoire that is under UIS12 control. Since the shared signature motif was identified in up-regulated transcripts, this would argue against a role of UIS12 as an mRNA stabilizing factor. At least three alternative scenarios emerge (Figure 7). One potential, albeit unusual, mechanism of UIS12 could be binding and stabilizing of large set of target mRNAs that encode gametocyte- and mosquito stage-specific proteins. Alternatively, UIS12 acts on one or few mRNAs, which in turn drive expression control, for instance by stabilizing transcripts that encode checkpoint activators. A reverse, and perhaps more likely function would be an inhibitory role towards target mRNAs or a partner protein, which in turn modulate the half-lives of target mRNAs. According to such a scenario, up-regulation of transcripts in the absence of UIS12 might be a consequence of increased mRNA stability. We consider the identification of a nucleotide motif a good starting point for future work, and are hesitant to propose a molecular mechanism at this early stage of discovery.

In gametocytes, the conserved DOZI/CITH RNA storage complex (Mair et al., 2006; Mair et al., 2010; Guerreiro et al., 2014) is central to translational repression. For instance, the DOZI/CITH complex is involved in translationally repressing P25, P28, PlasmepsinVI, AP2-O, and GAP45. These transcripts are also contained on our list of down-regulated transcripts in uis12(-) blood stages. Thus, we were curious how many targets
overlap when we compared the reported 117 genes down-regulated in both dozi(-) and cith(-) as determined by microarray on gametocytes (Mair et al., 2010) with the 646 down-regulated transcripts in uis12(-) blood stages (≤2-fold change), we found a surprisingly high overlap, of 102 of the 117 transcripts (Figure 6B). The RNA-helicase DOZI was previously shown to be a key regulator of P25 and P28 transcript stability, as well as of 731 other transcripts, and they all share a conserved U-rich nucleotide motif mainly in the 5' and 3' UTR (Hall et al., 2005; Braks et al., 2008; Guerreiro et al., 2014). The abundance of U residues in the DOZI and UIS12 motifs could hint at a related mechanism. The key differences are that the U-rich motif in our study is found in the ORFs of target genes, while the U-rich motif bound by the DOZI complex is located in 5' and 3' regions of its targets (Braks et al., 2008). Also, the U-rich motif identified for DOZI binding has a length of 47 nucleotides, whereas the motif identified herein is only 10 nucleotides long. Despite these clear distinctions a potential functional link of these two proteins remains an attractive possibility, since UIS12 and DOZI have >100 overlapping down-regulated targets (Figure 6B).

We note that, in marked contrast to UIS12, gene deletion of DOZI leads to a developmental arrest of the fertilized female gamete, a process that is not abolished in the absence of UIS12. In all three knockout lines, uis12(-), dozi(-), and puf2(-) parasites the mRNA levels of P25 and P28 were perturbated. While in dozi(-) and uis12(-) the transcripts were down-regulated, in puf2(-) an up-regulation was described, indicating involvement in distinct shared and opposing regulatory processes. On the other hand, PlasmepsinVI is translationally repressed by PfPuf2 (Miao et al., 2013) and reduced in uis12(-) parasites, similar to the meiotic recombinase DMC1 (Mlambo et al., 2012). Both knockouts as well as the CCP knockouts resemble the defects of uis12(-). Clearly, further work is warranted to decipher the molecular mechanisms that orchestrate the sequential post-transcriptional control events and contributions of the regulatory molecules.

In conclusion, our experimental genetics analysis of the Plasmodium berghei candidate RNA binding protein UIS12 uncovered broad perturbations of mRNA expression during blood infection, including up-regulation of several hundred transcripts, which share a U-rich 10 nucleotide-long motif in their open reading frames. The observed mRNA deregulation correlates with pleiotropic defects after blood stage propagation during sexual stage development, mosquito midgut colonization and oocyst maturation, ultimately leading to a complete block in the Plasmodium life cycle inside the Anopheles vector.
METHODS

Ethics Statement
All animal work conducted in this study was carried out in strict accordance with the German “Tierschutzgesetz in der Fassung vom 18. Mai 2006 (BGBl. I S. 1207)”, which implements the directive 86/609/EEC from the European Union and the European Convention for the protection of vertebrate animals used for experimental and other scientific purposes. The protocol was approved by the ethics committee of MPI-IB and the Berlin state authorities (LAGeSo Reg# G0469/09, G0294/15).

Parasites and Experimental Animals
P. berghei ANKA cl507 parasites that constitutively express GFP under the PbEF1a promoter and P. berghei ANKA Bergreen parasites that express the green fluorescent protein GFP under the PbHSP70 promoter throughout their life cycle have been used in our experiments (Franke-Fayard et al., 2004; Kooij et al., 2012).

All mouse experiments were performed with female NMRI or C57BL/6 mice purchased from Charles River Laboratories (Sulzfeld, Germany) or bred in house. NMRI mice were used for transfection experiments, blood stage infections, and transmission to Anopheles stephensi mosquitoes. C57BL/6 mice were used for sporozoite and stage blood infections, exflagellation assays and ookinete cultures.

Gene Deletion of P. berghei UIS12
The P. berghei UIS12 gene deletion was performed by double crossover homologous recombination of a replacement pB3D-knockout plasmid (van Dijk et al., 1995). The gene knockout construct targeting the endogenous UIS12 locus comprised a 647 bp UIS12 5’ UTR and a 524 bp UIS12 3’ UTR sequence flanking a T. gondii DHFR/TS pyrimethamine-resistance cassette. Table S4 lists the oligonucleotides used for generation of the knockout construct. Prior to transfection, plasmids were linearized by restriction digest. A digest with SacII and KpnI produced a fragment consisting of the pyrimethamine resistance cassette flanked by the UIS12 5’ and 3’ UTRs. Transfection by electroporation of cultured schizonts was performed as previously described (Janse et al., 2006b). Two independent parasite lines, one in P. berghei Bergreen and one in P. berghei cl507 recipient lines were generated and phenotypically characterized. For genotyping of recombinant parasites, knockout- and wild-type-specific PCR fragments were amplified from genomic DNA of parental and clonal lines with the oligonucleotides listed in Table S4.

Quantitative Real-Time PCR (qRT-PCR)
Quantitative real-time PCR was used to quantify UIS12 gene expression and for comparing gene expression in uis12(-) and wild-type (WT) mixed blood stages and gametocytes. For mixed blood stage analyses, C57BL/6 mice were infected i.v. with 10,000 iRBC, and 7 days after infection whole blood was isolated. The gametocyte samples were generated by infecting phenylhydrazine-treated NMRI mice with 10⁶ iRBC. Three to four days later, when gametocytemia reached the highest levels, 12.5 mg sulfadiazine was supplemented to 1 liter of drinking water. Within 48 h the sulfadiazine treatment eliminated the asexual stages, which was confirmed by a Giemsa-stained thin blood film. Next, the infected blood was harvested. To isolate mixed blood stages or gametocytes, whole blood was cleaned from white blood cells and erythrocytes using cellulose-glass bead columns and saponin (0.3%; Sigma-Aldrich) lysis. Parasites were stored in 1 ml TRizol (Invitrogen) or RLT-Buffer with β-mercaptoethanol (Qiagen). Parasite RNA was isolated with the RNasy Kit (Qiagen) or TRizol-Reagent (Sigma-Aldrich). DNase treatment with Qiagen RNase free DNase Set or Ambion Turbo DNA free Kit was performed to remove residual genomic DNA.

Reverse transcription to cDNA was done using the RETROscript Kit (Ambion). The qPCR was performed on parasite cDNA that was generated from total RNA of selected P. berghei life cycle stages. Quantitative real-time PCR was performed as previously described (Silvie et al., 2008) using Power SYBR Green PCR Master Mix and the StepOnePlus Real-Time PCR System (both Applied Biosystems). All procedures were performed according to the manufacturer’s instructions. Life cycle expression data were normalized to GFP that is constitutively expressed under the EF1α promoter in P. berghei 507cl. Mixed blood stage and gametocyte expression data for comparison of knockout and WT parasites were normalized to PbHSP70. The relative transcript abundance was determined by the 2⁻ΔΔCT method. For oligonucleotide sequences refer to Table S4.

Plasmodium Life Cycle: Parasite Asexual and Sexual Growth, Mosquito Infection, and Sporozoite Transmission
For asexual blood stage and gametocyte growth curves, C57BL/6 mice were injected intravenously with 10,000 infected red blood cells. From day 3 to 7, Giemsa (VWR)-stained thin blood films were microscopically examined to determine parasitemia and gametocytemia. Parasitemia and gametocytemia are defined as the percentage of red blood cells infected with asexual stages or gametocytes, respectively.

To analyze male exflagellation, C57BL/6 mice were intravenously injected with 10⁶ infected red blood cells (iRBC). After four days, parasitemia and gametocytemia were determined. Next, five microliters of tail blood were diluted 1:25 in ookinete medium (RPMI 1640, 10% FCS and 50 mM xanthurenic acid, pH 8.0) and incubated for 12 min at room temperature. For the next 6 min exflagellation centers were counted with help of a Neubauer chamber.

Ookinete cultures were set up as previously described (Weiss et al., 1997). Four days after the intravenous injection of 10⁶ iRBC into C57BL/6 mice pre-treated one day earlier with phenylhydrazine 97% (Sigma-Aldrich, 100µl of 6.18 µl/ml) 1 ml of fresh mouse blood was mixed with 10 ml of ookinete medium. After 20 h incubation at 20°C, at 80% humidity the culture was examined for development of ookinetes and ookinetes were purified with anti-P28-coupled magnetic beads.

Anopheles stephensi mosquitoes were kept at 28°C (non-infected) or 20°C (infected) under a 14 h light/10 h dark cycle.
at 80% humidity. Four days after the transfer of $10^7$ iRBC, when exflagellation reached a maximum, mosquitoes were infected as described (Vanderberg, 1975). After 4 to 14 days, midguts were isolated and oocytes development analyzed by fluorescent microscopy (Zeiss AxioObserver Z1). For visualization of nuclei, HOECHST 33342 (Invitrogen) was added at a 1:1,000 dilution. Images were analyzed using the FIJI processing (oocyst diameter) and/or Photoshop (oocyst numbers) software.

Midguts and salivary glands were analyzed at day 14–18 and day 18–30, respectively, for presence of sporozoites.

To test *Plasmodium* transmission, age-matched female C57BL/6 mice were either subjected to the bites of ~100 infected mosquitoes or inoculated with 10,000 sporozoites, or for *uis12(-)* the salivary gland extract of 20 mosquitoes, in RPMI intravenously. After three days Giemsa-stained thin blood films were analyzed for presence of infected red blood cells.

**Microarray Expression Profiling of *P. berghei* Blood Stages**

The microarray was performed on two biological replicate samples of *uis12(-)* and wild-type with comparable parasitemia. One replicate was generated by infecting one C57BL/6 mouse each with 10,000 *uis12(-)* or wild-type blood stages intravenously (R1, biological replicate 1). After 7 days of infection, mixed blood stages were isolated from whole blood. For both samples, the parasitemia was comparable (*uis12(-)* 6.3%, WT 6.2%). The other replicate (R2, biological replicate 2) was generated by injecting one NMRI mouse with $10^7$ *uis12(-)* or wild-type blood stages and mixed blood stage parasites were harvested after 3 days (*uis12(-)* 8.7%, WT 6.4%).

Whole blood was isolated and cleaned from white blood cells and erythrocytes using cellulose-glass bead columns and saponin (0.3%; Sigma-Aldrich) lysis. About $10^8$ parasites were stored in 1ml TRizol (Invitrogen). The amount of RNA was determined by OD 260/280 nm measurement using a 'NanoDrop 1000' (peQlab) spectrophotometer. The RNA size, integrity and the amount of total RNA were measured with a Bioanalyzer 2100 (Agilent Technologies). Raw microarray image data were extracted and analyzed with the ‘G2567AA Image Analysis/Feature Extraction software’ (Version A.10.5.1.1, Agilent Technologies). On reporter and sequence level, the extracted MAGE-ML files were further analyzed with the ‘Rosetta Resolver Biosoftware’ (7.2.2.0 SP131). All subsequent data analysis was performed with the Microsoft Excel, Prism and the PlasmoDB (http://plasmodb.org/plasmo/) database. *P. berghei* microarrays were designed on the OligoWiz (http://www.cbs.dtu.dk/services/OligoWiz/) server that included annotated open reading frames (ORF) from the whole genome of *P. berghei*. ORF specific probes were designed as sense oligonucleotides of the coding strand (Wernersson et al., 2007). The aimed oligonucleotide length was between 50 and 60 bases. To avoid self-hit, the maximal homology was 97%, the cross hybridization maximum length was at 80% and random primed position preference scores within OligoWiz were chosen. Depending on these parameters and the sequence input length, each ORF was covered by different numbers of specific oligonucleotide probes. In total 41,133 specific probe sequences were uploaded to eArray (Agilent Technologies, https://earray.chem.agilent.com/earray/) as expression application with a customer specified feature layout.

The microarray data discussed in this publication have been deposited in NCBI's Gene Expression Omnibus (Edgar et al., 2002) and are accessible through GEO Series accession number GSE152686.

**Statistical Analysis and Online Bioinformatic Tools**

Statistical significance was determined with the help of Graph Pad Prism software, using Mann-Whitney test for comparison of parasitemia, gametocytemia, and exflagellation dot-plots and multiple t-tests (one per row) for all growth curves.

The identification and positioning of the two RRM domains of the UIS12 orthologous proteins has been performed by NCBI WEB batch-CD Search Tool (Marchler-Bauer et al., 2015). Protein comparison was done by NCBI protein blast (blastp) with default settings, no compositional adjustment and without low complexity filter. Sequence coverage of all *Plasmodium* orthologues was higher than 90%.

Gene ontology (GO) enrichment analysis (Ashburner et al., 2000) was performed with help of the Gene Ontology enrichment analysis tool from PlasmoDB, with the default settings and the redundant GO terms were removed with help of the REVIGO software.

To screen for common 8-10 nucleotide-long motifs we used the MEME webtool (Bailey et al., 2009), screened all ORFs, 5'UTR sequences and 3'UTR sequences (1,000 nucleotides per gene for ORFs) of transcripts up-regulated >2.5-fold and compared them to the respective gene parts of the non-regulated (-1.5 to -1.5) transcripts from our microarray analysis of biological replicate 1.

Parts of the illustrations used in Figure 7 were obtained from Servier Medical Art, (http://www.servier.fr/servier-medical-art) licensed under a Creative Commons Attribution 3.0 Unported License.
DATA AVAILABILITY STATEMENT

The microarray data discussed in this publication have been deposited in NCBI’s Gene Expression Omnibus (Edgar et al., 2002) and are accessible through GEO Series accession number GSE152686 (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE152686).

ETICS STATEMENT

The animal study was reviewed and approved by the ethics committee of MPI-IB and the Berlin state authorities (LAGeSo Reg# G0469/09, G0294/15).

AUTHOR CONTRIBUTIONS

KMa, OS, and KMü designed the experiments. KMü performed experiments and analyzed data. H-JM performed and analyzed the microarray. KMa and KMü wrote the paper. All authors commented on and revised the manuscript. All authors contributed to the article and approved the submitted version.

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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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