Standardized Electrolyte Supplementation and Fluid Management Improves Survival During Amphotericin Therapy for Cryptococcal Meningitis in Resource-Limited Settings

Nathan C. Bahr,1,2 Melissa A. Rolfes,1,2 Abdu Musubire,3 Henry Nabeta,3 Darlisha A. Williams,1,2,3 Joshua Rhein,1,2,3 Andrew Kambugu,1,2,3 David B. Meya,1,2,3,4 and David R. Boulware1,2

1Division of Infectious Diseases and International Medicine, Department of Medicine, and 2Center for Infectious Diseases and Microbiology Translational Research, University of Minnesota, Minneapolis; 3Infectious Disease Institute, and 4School of Medicine, College of Health Sciences, Makerere University, Kampala, Uganda

Background. Amphotericin B is the preferred treatment for cryptococcal meningitis, but it has cumulative severe side effects, including nephrotoxicity, hypokalemia, and hypomagnesemia. Amphotericin-induced severe hypokalemia may predispose the patient to cardiac arrhythmias and death, and there is very little data available regarding these toxicities in resource-limited settings. We hypothesized that standardized electrolyte management during amphotericin therapy is essential to minimize toxicity and optimize survival in sub-Saharan Africa.

Methods. Human immunodeficiency virus-infected, antiretroviral therapy naive adults with cryptococcal meningitis were prospectively enrolled at Mulago Hospital in Kampala, Uganda in 3 sequential cohorts with amphotericin B deoxycholate induction treatment. Intravenous fluid use was intermittent in 2001–2002, and universal in 2006–2012. In 2001–2009, serum potassium (K+) was monitored on days 1, 7, and 14 of treatment with replacement (K+, Mg2+) per clinician discretion. In 2011–2012, K+ was measured on days 1, 5, and approximately every 48 hours thereafter with universal electrolyte (K+, Mg2+) supplementation and standardized replacement. Clinical outcomes were retrospectively compared between fluid and electrolyte management strategies.

Results. With limited intravenous fluids, the 14-day survival was 49% in 2001–2002. With universal intravenous fluids, the 30-day survival improved to 62% in 2006–2010 (P = .003). In 2011–2012, with universal supplementation of fluids and electrolytes, 30-day cumulative survival improved to 78% (P = .021 vs 2006–2010 cohort). The cumulative incidence of severe hypokalemia (<2.5 mEq/L) decreased from 38% in 2010 to 8.5% in 2011–2012 with universal supplementation (P < .001).

Conclusions. Improved survival was seen in a resource-limited setting with proactive fluid and electrolyte management (K+, Mg2+), as part of comprehensive amphotericin-based cryptococcal therapy.

Keywords. amphotericin; cryptococcal meningitis; HIV/AIDS; potassium; side effect.

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Correspondence: Nathan C. Bahr, MD, MA, Infectious Diseases Institute, PO Box 22418, Mulago Hospital Complex, Kampala, Uganda (bahrx026@umn.edu).

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Cryptococcal meningitis is the most common cause of adult meningitis in sub-Saharan Africa [1–3], and among persons infected with human immunodeficiency virus (HIV) it accounts for 20%–25% of acquired immune deficiency syndrome-related mortality in Africa [4–7]. Standard treatment is combination therapy with amphotericin and flucytosine (or with fluconazole when flucytosine is unavailable) [8, 9]. Although amphotericin-based regimens have superior clinical efficacy over fluconazole monotherapy [10], amphotericin has side effects including nonlife-threatening infusion-related reactions (eg, rigors, fevers, chills, nausea, and vomiting) as well as more significant cumulative toxicities such as nephrotoxicity, anemia, hypokalemia, and hypomagnesemia [11–13].
In persons receiving amphotericin B deoxycholate for >10 days, hypomagnesemia and hypokalemia are near universal (96% and 100%, respectively) [14]. Yet unlike in high-income countries, potassium (K\textsuperscript+) and magnesium (Mg\textsuperscript2+) monitoring and electrolyte replacement are limited in low- and middle-income countries. In many settings, electrolyte monitoring is much more limited or absent, and electrolyte replacement is more sporadic and physician-dependent. Little data exist on how best to optimize management of electrolytes in patients given amphotericin in resource-limited settings. In the second month (January 2011) of the Cryptococcal Optimal Antiretroviral Therapy Timing (COAT) Trial (clinicaltrials.gov NCT01075152), an association was noted between low serum K\textsuperscript+ and in-hospital mortality. A standardized electrolyte supplementation protocol was thereafter implemented as a quality improvement initiative. Although electrolyte abnormalities are a common amphotericin toxicity, before February 2011 electrolyte supplementation had been given only as reactive, physician-dependent ad hoc responses to laboratory abnormalities.

The objective of this project was to retrospectively assess 3 prospective cryptococcal cohorts: (1) intermittent use of intravenous (IV) fluids and rare ad hoc electrolyte supplementation, (2) standardized IV fluid but rare ad hoc electrolyte supplementation, and (3) standardized administration of IV fluids and universal electrolyte supplementation. Mortality was compared across cohorts to determine whether an aggressive approach to fluid and electrolyte management improves short-term (<30 days) survival after cryptococcal meningitis in persons receiving amphotericin-based therapy to understand how best to safely administer amphotericin in a limited-resource setting.

**METHODS**

**Study Population**

Three prospective studies of HIV-infected adults with cryptococcal meningitis were conducted at Mulago Hospital, the national tertiary referral hospital, in Kampala, Uganda. The first cohort enrolled from November 2001 through March 2002, before the availability of antiretroviral therapy (ART), as previously reported [15]. The second cohort enrolled from June 2006 through September 2009, after ART availability. During this time, ART was initiated at a median of 5 weeks [15–17], with additional enrollees from November 2010 to January 2011 from individuals screened for enrollment into the COAT trial (clinicaltrials.gov:NCT01075152) [18]. The third cohort consisted of individuals screened for enrollment into the COAT trial and a follow-on observational cohort that enrolled participants from February 2011 until November 2012. Figure 1 outlines the differences in the clinical management of the 3 cohorts. Inclusion and exclusion criteria are listed in the Supplementary Material, Appendix S1. Written informed consent was obtained. Study protocols were approved by the Institutional Review Boards of Makerere University, the University of Minnesota, and the Uganda National Council of Science and Technology.

Cryptococcal treatment consisted of standardized induction therapy of 14 days of amphotericin B 0.7–1.0 mg/kg in 500 mL 5% dextrose in water over 4 hours in all cohorts. Patient weights were measured in 2001–2002 then estimated until January 2011 when a weighing scale became available again. Lumbar punctures were completed on approximately days 1, 7, and 14 of amphotericin therapy. During 2010–2012 (this included patients from the COAT trial in cohorts 2 and 3), adjunctive oral fluconazole 800 mg/day was also included in the induction regimen [9, 19, 20]. In 2001–2009 (cohorts 1 and 2), after initial induction therapy, consolidation therapy consisted of 8 weeks of fluconazole 400 mg/day. During 2010–2012 (patients from the COAT trial in cohorts 2 and 3), enhanced consolidation therapy began with fluconazole (800 mg/day) until outpatient clinic registration (~3 additional weeks) and the 14-day cerebrospinal fluid (CSF) culture was known to be sterile, followed by ~9 additional weeks of fluconazole (400 mg/day) for a 12-week total consolidation. After consolidation therapy, all cohorts received secondary prophylaxis with fluconazole (200 mg/day).

Cryptococcus meningitis was diagnosed until April 2011 via latex agglutination and culture. Qualitative cultures were performed in 2001–2002. Quantitative cultures were performed first with a 10 mcL calibrated loop in 2006–2009 [15], and then 100 mcL serial 10-fold dilutions in 2010–2012 [21].
Fluid and Electrolyte Management
In 2001–2002 (cohort 1), IV fluids were of limited quantity and intermittently available. In 2006–2009 (cohort 2), all participants received 1 liter of 0.9% NaCl normal saline (NS) before amphotericin, supported via the Minnesota Medical Foundation. In these cohorts, serum electrolyte (Na+, K+) and creatinine monitoring was performed on days 1, 7, and 14, and electrolyte replacement was limited in supply. In 2010–2012 (COAT trial patients in cohorts 2 and 3), participants received 2 liters of NS daily, and after provision of informed consent (median day 5), subjects had additional laboratory safety monitoring with serum electrolyte (Na+, K+, HCO3) and creatinine measurement approximately every 48 hours. In 2010–2012, laboratory results were measured at the Makerere University-Johns Hopkins University (MU-JHU) laboratory using a Roche COBAS Integra 400 Plus Analyzer. The MU-JHU laboratory is a College of American Pathologists-accredited laboratory. In 2001–2009, laboratory tests were performed at the Mulago Hospital laboratory.

Electrolyte Supplementation Protocols
During the presupplementation period through January 2011 (cohorts 1 and 2), electrolyte management was at the treating physician’s discretion, and replacement was generally given in response to abnormal electrolyte levels. Replacement was very infrequently completed.

In February 2011, a routine electrolyte supplementation protocol was implemented with K+ and Mg2+ universally given starting on day 1 of amphotericin therapy, data from that point is termed the supplementation period and corresponds with the 2011–2012 cohort (cohort 3). Table 1 summarizes the management differences. The protocol included baseline K+ measurement in addition to the measurements on day 5, 7, 9, 11, and 14 with replacement to goal (K+ = 4.0 mEq/L) after all measurements. Potassium supplementation was 32–40 mEq K+ daily (primarily oral KCl 8 mEq tablets in divided doses). After 1 week of amphotericin, an additional 16 mEq KCl orally was added to baseline supplementation. If hypokalemia occurred despite the universal supplementation, the baseline supplementation was increased by one 8 mEq tablet twice daily in addition to one time replacement doses. The KCl replacement dose was standardized at 10 mEq K+ for each 0.1 mEq/L serum K+ below the target goal (K+ = 4.0 mEq/L). Intravenous replacement was via 40 mEq K+ mixed in 500 mL NS given over 4 hours.

Magnesium supplementation was also addressed in the 2011–2012 cohort (cohort 3), although Mg2+ measurement was unavailable on site. Magnesium is wasted with amphotericin use (as noted above), and hypomagnesemia has numerous deleterious effects; however, hypomagnesemia also interferes with the patient’s ability to properly replete potassium, thus making magnesium replacement crucial to adequate potassium replacement. Universal supplementation was with magnesium trisilicate (500 mg, 4 mEq) twice daily, which was the only oral Mg2+ locally available, initially. This was later changed to Slow Mag (MgCl2), with no standardized electrolyte management (as noted above), and hypomagnesemia has numerous deleterious effects; however, this interferes with the patient’s ability to properly replete potassium, thus making magnesium replacement crucial to adequate potassium replacement. Universal supplementation was with magnesium trisilicate (500 mg, 4 mEq) twice daily, which was the only oral Mg2+ locally available, initially. This was later changed to Slow Mag (MgCl2), MgSO4, Mg2+ supplementation

Table 1. Electrolyte Management Strategies During Amphotericin Therapy by Time Period

| Electrolyte Protocol Component | Presupplementation (2001–2010) | Universal Supplementation (2011–2012) |
|-------------------------------|-----------------------------|--------------------------------------|
| K+ monitoring                 | Day 1, 7, 14 (2001–2009)    | Day 1, 5, 7, 9, 11, 14, and as needed |
|                               | Day 5, 7, 9, 11, 14 (2010) | Goal: serum K+ of 4.0 mEq/L          |
| K+ supplementation            | In reaction to laboratory   | • Day 1–6: 32–40 mEq KCl daily       |
|                               | abnormalities               | • Day 7–14: 48–56 mEq KCl daily      |
|                               |                             | • Mild hypokalemia (<3.5 mEq/L):     |
|                               |                             | • Increase daily routine dose by +16 mEq KCl |
|                               |                             | • Replacement of 10 mEq per 0.1 mEq/L deficit to a target of 4.0 mEq/L with each measurement |
| Mg2+ monitoring               | None                        | None                                 |
| Mg2+ supplementation          | At physician discretion     | Day 1–14: 8 mEq Mg2+ daily           |
|                               |                             | MgSO4, 5 g IV, if K+ levels <3.0 mEq/L for 3 consecutive days despite adequate KCl replacement and continued supplementation until K+ normalized. |

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IV fluids and universal electrolyte supplementation (n = 142, February 2011–November 2012).

Baseline participant characteristics were compared between the cohorts using analysis of variance or 2-sample t tests to compare means and the Fisher exact test to compare frequencies. Serial serum K+ levels were evaluated using a repeated measures model with levels at days 1, 7, and 14 of amphotericin therapy. The occurrence of severe hypokalemia (K+ < 2.0 mEq/L) or mild hypokalemia (K+ 2.0–2.4 mEq/L) was compared with the Fisher exact test. Cox proportional hazard regression compared survival between cohorts with adjustment, when possible, for different baseline characteristics. The survival analysis was restricted to the pre-ART time period only. Participants contributed time from cryptococcal meningitis diagnosis to one of the following: 30-day survival, death, or ART initiation. COAT trial participants randomized to early ART were right-hand censored for the survival analysis at COAT trial randomization (n = 8 in 2010, n = 49 in 2011–2012). Thus, no persons received ART in the 30-day survival analysis, to make all 3 cohorts comparable. For determination of cumulative incidence of hypokalemia, all participants were included. Statistical analysis was conducted using SPSS version 21 (IBM Corporation, Armonk, NY) and evaluated against type I error α < 0.05.

**RESULTS**

**Patient Characteristics**

Ninety-two subjects were included in cohort 1, 195 in cohort 2 (174 in 2006–2009 and 21 in November 2010–January 2011), and 142 in cohort 3. Table 2 displays baseline demographics. Demographics were similar, except for the following: proportion of persons with altered mental status with a Glasgow Coma Scale (GCS) < 15 was less in cohort 1 than cohorts 2 and 3 (7.6%, 28%, and 31%, P < .001); the mean HIV viral load was slightly higher in cohort 3 than cohort 2 (5.5 vs 5.2 log10 copies/mL, P = .006); and viral load and CD4 counts were not available for cohort 1. Time from hospital admission to definitive diagnosis also improved over time (median 3 to ≤1 days), yet demographics were overall similar.

**Incidence of Severe Hypokalemia**

Figure 2 displays the time to severe or life-threatening (Grade ≥3) hypokalemia (K+ < 2.5 mEq/L) by cohort. The incidence of severe electrolyte abnormalities in cohort 1 during 2001–2002 was negligible. Among those surviving to day 7, only 1.8% (1 of 56) had severe hypokalemia, and zero of 46 who survived to day 14 had severe hypokalemia. Electrolyte data were not prospectively recorded for cohort 2 between 2006 and
but similarly negligible incidence of severe hypokalemia at day 7 and day 14 was recalled (D. B. M., A. M.). However, in the 2010 period of cohort 2 when electrolytes were measured starting at day 5 approximately every 48 hours, the cumulative incidence of severe hypokalemia (K+ < 2.5 mEq/L) was 38% (8 of 21). After the implementation of universal supplementation (February 21, 2011) and enhanced attention to weight-based dosing of amphotericin (cohort 3), the incidence of severe hypokalemia declined to 8.5% (12 of 142, P < .001 compared to without supplementation). No persons developed clinically significant hyperkalemia with electrolyte supplementation. The majority of the hypokalemia occurs during the second week of amphotericin therapy, thus with minimal monitoring in cohort 1, the lack of detected hypokalemia does not indicate the absence of hypokalemia. Severe hypokalemia rarely occurs before day 7. Without intensive K+ monitoring, absence of hypokalemia at day 14 likely may represent a survival bias. Abbreviation: IV, intravenous.

2009, but similarly negligible incidence of severe hypokalemia at day 7 and day 14 was recalled (D. B. M., A. M.). However, in the 2010 period of cohort 2 when electrolytes were measured starting at day 5 approximately every 48 hours, the cumulative incidence of severe hypokalemia (K+ < 2.5 mEq/L) was 38% (8 of 21), with 19 severe hypokalemic events occurring in 8 participants in the first 14 days of amphotericin. Severe hypokalemia was evenly distributed between the trial’s randomization arms. After the standardized electrolyte protocol was implemented in February 2011, 15 severe hypokalemic events occurred among 12 participants for a cumulative incidence of 8.5% (12 of 142) (relative risk = 4.5; 95% CI, 2.1–9.7; P < .001). Thus, a lack of severe hypokalemia observed in 2001–2002 likely reflects unrecognized severe hypokalemia because of lack of testing between day 8 and 13 as well as a survival bias, namely those who survived to 14 days were more likely the patients without severe hypokalemia during amphotericin therapy. Unintentional missed doses of amphotericin are unlikely because the medication was administered by study nurses and monitored by study physicians.

Survival
When limited IV fluids were available and intermittent shortages occurred, the 14-day survival was 49% (45 of 92) in 2001–2002. With universal provision of IV fluids in 2006–2010, the 30-day cumulative survival was 62% (log rank P = .003 vs 2001–2002 cohort). In 2011–2012 (cohort 3), with universal IV fluids and electrolyte supplementation, 30-day cumulative survival improved to 78% (95% CI, 70%–85%; P = .021 vs 2006–2010 cohort, P < .001 vs 2001–2002 cohort). Right-hand censoring occurred at time of antiretroviral therapy (ART) initiation (including n = 8 in 2010; n = 49 in 2011–2012 randomized to early ART; n = 3).
Implementation of a comprehensive electrolyte management protocol was associated with reduced severe hypokalemia and improved survival when added to standard amphotericin treatment and IV fluids for HIV-associated cryptococcal meningitis in a resource-limited setting. Current Infectious Diseases Society of America guidelines for cryptococcal treatment mention that if facilities do not have sufficiently rapid or reliable K+ monitoring, amphotericin use may not be safe [8]. However, further guidance is not provided, and magnesium is not mentioned [8, 22].

In the high-income countries, electrolytes are checked frequently and replaced rapidly in hospitalized patients; however, in the many resource-limited settings, unique barriers exist for the close monitoring of electrolytes. Laboratory facilities may not be reliably available or not able to return results rapidly enough to be acted upon in a clinically useful fashion. Electrolyte replacements themselves may be unavailable in many settings, and cost, although low relative to many other medications, can be a barrier. Thus, although a comprehensive electrolyte management strategy would be ideal, this may not be realistic in all settings. Yet, amphotericin-induced electrolyte wasting is a universal expectation, and so the issue must be addressed. In many illnesses, this may be inconsequential; however, for any illness treated with electrolyte wasting medications, such as amphotericin, K+ and Mg2+ replacement becomes extremely important [23–25]. Stakeholders who influence health policy in low- and middle-income countries should view electrolyte management as part of the package of care for proper cryptococcal treatment.

Multiple alternatives to the comprehensive electrolyte replacement strategy detailed above are possible. Amphotericin predictably depletes potassium and magnesium [14, 26]. Thus, scheduled supplementation only without additional measurement or replacement doses would be feasible. Although there is some risk of hyperkalemia due to replacement, the predictable nature of electrolyte wasting allows for scheduled replacement without significant fear of hyperkalemia. Furthermore, because amphotericin’s effect on electrolyte wasting is known to be cumulative and dose dependent [11, 14, 26], increasing the potassium supplementation during the second week should be well tolerated. This strategy would reduce overall hypokalemia but likely leave some small percentage of patients (~10%) severely hypokalemic; moreover, routine replacement would have an acceptable, potential risk of hyperkalemia should even routine monitoring be difficult to obtain.

A second strategy would be to use shorter 1-week courses of amphotericin. In a prospective study in Uganda by Muzoora et al [27], 5 days of amphotericin (1.0 mg/kg per day) with adjunctive fluconazole (1200 mg/days) was well tolerated with approximately 75% of the rate of microbiologic clearance (ie, early fungicidal activity) compared to 14 days of amphotericin (1.0 mg/kg per day) but without electrolyte abnormalities. In that study, routine potassium (40 mEq/day) supplementation was given. Similar high levels of efficacy without toxicity were also reported in a similar 7-day randomized trial of amphotericin (1.0 mg/kg per day) with fluconazole (1200 mg/day) in Malawi [28]. An unresolved clinical question, raised by Thomas Harrison and colleagues, St. George’s University, London [27, 28], is the ideal length of amphotericin induction, and this question is currently being tested in a phase III trial (ISRCTN45035509).

Finally, in resource-limited settings without access to KCl pharmaceutical preparations, foods rich in potassium may be readily available and could provide suitable potassium replacement while on amphotericin. For example, the average US avocado has ~6.5 mEq of potassium per 100 g, whereas a banana has approximately two thirds of that amount [29]. In a patient able to tolerate food or nasogastric feeding, this may be a reasonable alternative method of potassium supplementation. Defaulting to fluconazole monotherapy to avoid amphotericin toxicity is a poor strategy. The survival with fluconazole monotherapy is ~30% worse than with amphotericin, in cross-cohort comparisons [10]. Although the lure of less toxicity in fluconazole monotherapy may be appealing, as we have demonstrated here, the electrolyte toxicity of amphotericin is manageable, and acute kidney injury is relatively infrequent (~8%) with IV fluid prehydration [11]. As mentioned above, in 2001–2002 (cohort 1), IV fluids were of limited quantity and intermittently available. In 2006–2009 (cohort 2), all participants received 1 liter of 0.9% NaCl NS before amphotericin. In 2010–2012 (COAT trial patients in cohorts 2 and 3), participants received 2 liters of NS daily according to trial standard operating procedures. Persistent significant chronic kidney injury is rare: in the 2006–2009 Kampala cohort [17], 95% of survivors had a serum creatinine <2 mg/dL at 5 weeks after cryptococcal meningitis diagnosis, with 80% <1.5 mg/dL. In 2010–2012, 99% of survivors had creatinine <2 mg/dL at 5 weeks.

Cost is often considered as a barrier to therapeutic interventions. As an example of the costs of these medications; 1 non-profit medical wholesaler in Kampala, Uganda supplies the following prices of 1 vial of 10 mEq IV KCl for $1.04 and oral 8 mEq KCl for $3.30 per 100 tablets [30]. Magnesium prices are similar [30]. Although these purchases would require some resources, the cost is quite affordable when one considers the immediate 30-day survival benefit. Supporting local industry and local avocado or banana farmers may be a wiser investment than importing supplies.

The main limitation is the historical comparison of 3 cohorts over time. Although fluid and electrolyte management was the major change, other unseen bias influencing mortality may exist. Severity of illness was similar among cohorts and, if anything, increased over time with higher proportions with altered mental status and higher CRAG titers during 2006–2012.
Quantitative cultures were performed in cohorts 2 and 3; however, the method was different, and so direct comparison would not be accurate. One of the major improvements in clinical care during the trial of cohort 2 and 3 was improved safety monitoring with more frequent detection of laboratory abnormalities. In 2001–2002, K⁺ monitoring at day 1, 7, and 14 only detected 2% with hypokalemia at day 7. Yet lack of monitoring did not equate to absence of hypokalemia during the second week of amphotericin, based on a 38% incidence of hypokalemia in 2010 with more frequent monitoring. Likewise, monitoring alone without action did not decrease mortality or hypokalemia in 2010. One might also argue that caregiver or institutional experience may have been gained over time, which explained the survival benefit. However, we believe this explanation is also unlikely given that the medical officers and nurses caring for the patients directly changed with each study (A. K.; D. B. M./A. M.; H. N.).

Other specific changes were made. First, the cryptococcal antigen lateral flow assay (Immy, Norman, Oklahoma) was implemented in April 2011 as a point-of-care test to decrease time-to-diagnosis. Although exceedingly helpful, the severity of illness and demographics remained similar. Second, concomitant fluconazole 800 mg/day was given starting in 2010 during induction therapy and as enhanced consolidation until the CSF was known to be sterile. This fluconazole regimen was in place during the initial 2 months of the COAT trial when the investigators detected an association between hypokalemia and mortality. Based on a 2013 trial, the addition of fluconazole to amphotericin did not have a statistical survival benefit over 4 weeks of amphotericin alone [31]. The higher dose fluconazole (800 mg/day) “enhanced” consolidation therapy was used in 2010–2012, but this is of unproven significance in terms of any potential benefit. Although fluconazole adjunctive therapy with amphotericin leads to more rapid clearance of Cryptococcus from the CSF [15, 18], this alone is unlikely to explain the improved survival in the third cohort [31]. The 16% magnitude of 30-day survival difference between cohorts 2 and 3 was beyond the expected effect of added fluconazole (5% better 14-day survival) [31]. A further limitation is the lack of recording of electrolyte and creatinine data during 2006–2009, although the values would be expected to be similar to 2001–2002 or 2010.

**CONCLUSIONS**

In summary, survival was substantially improved with IV fluids coupled with universal electrolyte supplementation of K⁺ and Mg²⁺, electrolyte monitoring, and standardized electrolyte replacement. WHO Rapid Advice for cryptococcosis treatment published in December 2011 recommended intense monitoring and supplementation based on our earlier data [9, 32]. We believe that the data presented herein should cause stakeholders to emphasize 3 important issues unique to cryptococcal treatment. First, electrolyte management is important for improving survival with amphotericin B deoxycholate therapy. Second, routine, proactive potassium and magnesium supplementation is superior to a reactive approach of replacing electrolytes once a life-threatening deficiency has been identified. Third, more operational research is needed to determine whether potentially shorter courses of 5, 7, or 10 days of amphotericin have a more favorable risk/benefit in resource-limited regions compared to 2 weeks of amphotericin [33]. Using initial quantitative CSF culture burden to guide therapy duration may be more rational than giving all persons 14 days of amphotericin [33]. In conclusion, in comparing 3 cohorts of patients with cryptococcal meningitis treated with amphotericin in Kampala, Uganda, survival significantly improved with a comprehensive electrolyte monitoring and replacement strategy.

**Supplementary Material**

Supplementary material is available online at Open Forum Infectious Diseases (http://OpenForumInfectiousDiseases.oxfordjournals.org/).

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**References**

1. Jarvis JN, Meintjes G, Williams A, et al. Adult meningitis in a setting of high HIV and TB prevalence: findings from 4961 suspected cases. BMC Infect Dis 2010; 10:87.

2. Durski KN, Kuntz KM, Yasukawa K, et al. Cost-effective diagnostic checklists for meningitis in resource-limited settings. J Acquir Immune Defic Syndr 2013; 63:e101–8.

3. Cohen DB, Zijlstra EE, Mukaka M, et al. Diagnosis of cryptococcal and tuberculous meningitis in a resource-limited African setting. Trop Med Int Health 2010; 15:910–7.

4. Castelnuovo B, Manabe YC, Kiragga A, et al. Cause-specific mortality and the contribution of immune reconstitution inflammatory syndrome in the first 3 years after antiretroviral therapy initiation in an urban African cohort. Clin Infect Dis 2009; 49:965–72.

5. Park BJ, Wannemuehler KA, Marston BJ, et al. Estimation of the current global burden of cryptococcal meningitis among persons living with HIV/AIDS. AIDS 2009; 23:525–30.

6. French N, Gray K, Watter C, et al. Cryptococcal infection in a cohort of HIV-1-infected Ugandan adults. AIDS 2002; 16:1031–8.

7. Liechty CA, Solberg P,Were W, et al. Asymptomatic serum cryptococcal antigenemia and early mortality during antiretroviral therapy in rural Uganda. Trop Med Int Health 2007; 12:929–35.

8. Perfect JR, Dismukes WE, Dromer F, et al. Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the Infectious Diseases Society of America. Clin Infect Dis 2010; 50:291–322.

9. World Health Organization. Rapid advice: Diagnosis, prevention and management of cryptococcal disease in HIV-infected adults, adolescents and children. Geneva: World Health Organization, 2011.
10. Rajasingham R, Rolfs MA, Birkenkamp KE, et al. Cryptococcal meningitis treatment strategies in resource-limited settings: a cost-effectiveness analysis. PLoS Med 2012; 9:e1001316.
11. Girmenia C, Gentile G, Micoczi A, et al. Nephrotoxicity of amphotericin B deoxycholate. Clin Infect Dis 2001; 33:915–6.
12. Gallis HA, Drew RH, Pickard WW. Amphotericin B: 30 years of clinical experience. Rev Infect Dis 1990; 12:308–29.
13. Imhof A, Walter RB, Schaffner A. Continuous infusion of escalated doses of amphotericin B deoxycholate: an open-label observational study. Clin Infect Dis 2003; 36:943–51.
14. Mayer J, Doubek M, Vorlicek J. Must we really fear toxicity of conventional amphotericin B in oncological patients? Support Care Cancer 1999; 7:51–5.
15. Kambugu A, Meya DB, Rhein J, et al. Outcomes of cryptococcal meningitis in Uganda before and after the availability of highly active antiretroviral therapy. Clin Infect Dis 2008; 46:1694–701.
16. Boulware DR, Bonham SC, Meya DB, et al. Paucity of initial cerebrospinal fluid inflammation in cryptococcal meningitis is associated with subsequent immune reconstitution inflammatory syndrome. J Infect Dis 2010; 202:962–70.
17. Boulware DR, Meya DB, Bergemann TL, et al. Clinical features and serum biomarkers in HIV immune reconstitution inflammatory syndrome after cryptococcal meningitis: a prospective cohort study. PLoS Med 2010; 7:e1000384.
18. Boulware DR, Meya DB, Muzoora C, et al. Timing of antiretroviral therapy after diagnosis of cryptococcal meningitis. N Engl J Med 2014; 370:2487–98.
19. Loyse A, Wilson D, Meintjes G, et al. Comparison of the early fungicidal activity of high-dose fluconazole, voriconazole, and flucytosine as second-line drugs given in combination with amphotericin B for the treatment of HIV-associated cryptococcal meningitis. Clin Infect Dis 2012; 54:121–8.
20. Pappas PG, Chetchotisakd P, Larsen RA, et al. A phase II randomized trial of amphotericin B alone or combined with fluconazole in the treatment of HIV-associated cryptococcal meningitis. Clin Infect Dis 2009; 48:1773–83.
21. Bicanic T, Meintjes G, Wood R, et al. Fungal burden, early fungicidal activity, and outcome of antiretroviral-naive or antiretroviral-experienced patients treated with amphotericin B or fluconazole. Clin Infect Dis 2007; 45:76–80.
22. Kaplan JE, Benson C, Holmes KH, et al. Guidelines for prevention and treatment of opportunistic infections in HIV-infected adults and adolescents: recommendations from CDC, the National Institutes of Health, and the HIV Medicine Association of the Infectious Diseases Society of America. MMWR Recomm Rep 2009; 58:1–207.
23. Bamba AV, Jadhav MP, Prabhur R, et al. Refractory hypokalemia due to conventional amphotericin B in patients with leukemia. Indian J Cancer 2009; 46:76–7.
24. White MH, Bowden RA, Sandler ES, et al. Randomized, double-blind clinical trial of amphotericin B colloidal dispersion vs. amphotericin B in the empirical treatment of fever and neutropenia. Clin Infect Dis 1998; 27:296–302.
25. McChesney JA, Marquardt JF. Hypokalemic paralysis induced by amphotericin B. JAMA 1964; 189:1029–31.
26. Mayer J, Doubek M, Doubek J, et al. Reduced nephrotoxicity of conventional amphotericin B therapy after minimal nephroprotective measures: animal experiments and clinical study. J Infect Dis 2002; 186:379–88.
27. Muzoora CK, Kabanda T, Ortu G, et al. Short course amphotericin B with high dose fluconazole for HIV-associated cryptococcal meningitis. J Infect Dis 2012; 64:76–81.
28. Jackson AT, Nussbaum JC, Phulusa J, et al. A phase II randomized controlled trial adding oral flucytosine to high-dose fluconazole, with short-course amphotericin B, for cryptococcal meningitis. AIDS 2012; 26:1363–70.
29. Pennington JAT, Youngt B. Sodium, potassium, calcium, phosphorus, and magnesium in foods from the United States total diet study. J Food Compost Anal 1990; 3:145–65.
30. Joint Medical Store. Potassium chloride slow-release 600mg tablet (Product catalog). Available at: http://www.jms.co.ug/resources. Accessed 1 September 2013.
31. Day JN, Chau TT, Wolbers M, et al. Combination antifungal therapy for cryptococcal meningitis. PLoS Med 2013; 368:1291–302.
32. Bahr N, Rolfs MAR, Musubire A, et al. The impact of routine electrolyte supplementation during amphotericin induction therapy in resource-limited settings. In: 8th International Conference on Cryptococcus and Cryptococciosis. Charleston, SC, 2011.
33. Rhein J, Boulware DR. Prognosis and management of cryptococcal meningitis in patients with human immunodeficiency virus infection. Neurobehavioral HIV Med 2012; 4:45–61.