Nutrition During Pregnancy and the Effect of Carbohydrates on the Offspring’s Metabolic Profile: In Search of the “Perfect Maternal Diet”

Irene P. Tzanetakou¹,*, Dimitri P. Mikhailidis² and Despina N. Perrea¹

¹Department for Experimental Surgery and Surgical Research “N.S. Christeas”, Medical School, National and Kapodistrian University of Athens, Greece, 15B Agiou Thoma Street, 11527, Athens, Greece
²Department of Clinical Biochemistry (Vascular Disease Prevention Clinics), Royal Free Hospital Campus, University College London Medical School, University College London (UCL), London, UK

Abstract: Fetal growth and development is primarily dependent upon the nutritional, hormonal and metabolic environment provided by the mother. A wartime famine study in Holland first showed that a low food intake reduces the glucose offered to the fetus and thus produces smaller size infants at birth. Maternal glucose regulation is however affected by numerous factors including physiological changes of pregnancy (e.g. insulin resistance [IR]), pathological conditions (e.g. gestational diabetes mellitus) and maternal nutrition. Maternal glucose is substantially influenced by the type of carbohydrates in the diet through its direct effect on glycemia. The rate at which each carbohydrate raises blood glucose levels after ingestion, can be measured via the dietary glycemic index (GI). Carbohydrate type and the GI of the diet enhance or inhibit abnormal hyperglycemia during pregnancy caused by either pathological conditions or the inability of the mother to cope with the physiological IR of pregnancy. In turn, maternal gestational hyperglycemia may be involved in the pathogenesis of IR, impaired glucose tolerance, type 2 diabetes mellitus, the Metabolic Syndrome and subsequent cardiovascular diseases in adult offspring. A low GI maternal diet has been associated with measurable benefits to the offspring. These include a positive effect on altering maternal blood glucose production, insulinemia and reduced adiposity as well as fetal and placental insulin and glucose regulation, fetal growth, birth weight and offspring adiposity.

We review the possible links between dietary carbohydrate in health during pregnancy and the effect of maternal carbohydrates in the diet through its direct effect on glycemia. The rate at which each carbohydrate raises blood glucose levels after ingestion, can be measured via the dietary glycemic index (GI). Carbohydrate type and the GI of the diet enhance or inhibit abnormal hyperglycemia during pregnancy caused by either pathological conditions or the inability of the mother to cope with the physiological IR of pregnancy. In turn, maternal gestational hyperglycemia may be involved in the pathogenesis of IR, impaired glucose tolerance, type 2 diabetes mellitus, the Metabolic Syndrome and subsequent cardiovascular diseases in adult offspring. A low GI maternal diet has been associated with measurable benefits to the offspring. These include a positive effect on altering maternal blood glucose production, insulinemia and reduced adiposity as well as fetal and placental insulin and glucose regulation, fetal growth, birth weight and offspring adiposity.

We review the possible links between dietary carbohydrate in health during pregnancy and the effect of maternal carbohydrate ingestion on programming the offspring’s metabolic profile.

Keywords: Pregnancy, nutrition, carbohydrates, metabolic profile, glycemic index.

INTRODUCTION

Over-nutrition, under-nutrition or unbalanced nutrition is considered a major cause of ill-health worldwide. The prevalence of overweight is high among children in most developed countries [1]. This increase in type 2 diabetes mellitus (DM) and the metabolic syndrome (MetS) in children possibly implies a combination of factors, the majority of which are related to nutrition during the life cycle [2]. However, whether and to what extent early fetal diet influences the achievement of long-term health requires more investigation.

The biological importance of maternal nutrition is well-established. Not only is it the sole way for the fetus to receive the required nutrients but it also affects the maternal metabolic adjustment capacity to the hormones secreted by the placenta that affect the metabolism of all nutrients.

Nutrition during early development is associated with the offspring’s growth, organ development, body composition and body functions. It also exerts long-term effects on health, morbidity and mortality risks in adulthood, as well as on the development of neural functions and behavior, a phenomenon called 'metabolic programming' [3].

The capacity of the offspring to respond to environmental information during early development was first suggested by the “Thrifty phenotype” hypothesis [4], further indicating that these early-life metabolic adaptations, promote the animal’s survival capability [5]. An increased susceptibility of the offspring to chronic diseases including obesity and DM in later life has been described in sub-optimal fetal environments (i.e. over-nutrition or malnutrition) in both humans and animal models [4, 6-8].

The response of the fetus to these insults in the environment during the prenatal period has short-term survival advantages, but may lead to long-term disadvantages. This is because it is associated with increased susceptibility of the offspring to cardiovascular disease, hypertension, type 2 DM and obesity [9]. Indeed, particular growth patterns in early life implicate early-life nutrition as the underlying mechanism for human diseases in adulthood [7, 10, 11]. Insulin resistance (IR), impaired glucose tolerance (IGT) [12-14], type 2 DM and MetS, as well as subsequent cardiovascular diseases in the adult offspring, can be induced from fetal and/or early postnatal hyperglycemia or hyperinsulinism (and also hyperleptinism) as a result of maternal...
gestational hyperglycemia and/or early postnatal overfeeding [12, 15-17].

The composition of the diet during pregnancy also modifies certain maternal risk factors (e.g. IGT, IR, hyperlipidemia etc) and may decrease the risk of developing gestational DM (GDM) [17]. It may also inhibit early metabolic changes in the offspring, associated with abnormal leptin and insulin levels [18].

Some consider the diet’s glycemic index (GI) as an independent factor that can cause obesity and increase the risks of DM and heart disease in animals and humans [19-22]. The GI describes the way food, a meal, or a diet affects blood glucose levels during the postprandial period. The effect of the pre- and post-natal dietary profile, as measured by the GI, on the offspring’s health has not been thoroughly investigated.

In this article we review the evidence linking periconceptional, pregnancy and lactating diet, in terms of dietary carbohydrate, and the metabolic effects on the offspring’s health.

MATERNAL NUTRITION AND OFFSPRING HEALTH

There are different means of achieving fetal malnutrition. The most well-established are 2 models of maternal malnutrition (protein and caloric restriction) and 1 model of maternal over-nutrition (fat and/or sucrose over-nutrition). These maternal dietary models, which are typical of the Western diet, lead to a disturbed fetal nutritional environment [23]. This in turn has been linked to a disturbed metabolic profile, such as IR and glucose intolerance, in the adult offspring [24].

A possible mechanism predisposing to IR, glucose intolerance and DM in adult offspring occurs in the presence of repeated maternal hypoglycemia. This leads to subsequent inhibition of normal endocrine pancreas development and abnormal β-cell mass and β-cell dysfunction at birth [24]. There is further evidence suggesting that the particular phenotype of IR and glucose intolerance is “predominant” enough to be transmitted to a second generation without any further environmental modification [23].

Periconceptional and pregnancy nutrition is important in offspring developmental and functional adverse health offspring outcomes. There is evidence suggesting that maternal undernutrition during the periconceptional period and pregnancy results in altered fetal hypothalamic-pituitary-adrenal axis (HPAA) development and function through increased concentrations of active glucocorticoids in utero and offspring altered expression of glucocorticoid receptors, lower fetal capacity to modulate glucose transport into the muscle (increased expression of glucose transporter type 4 [GLUT4]), increased adiposity, dysfunctional cardiovascular regulation, increased rate of premature birth, altered fetal pancreatic function and insulin signaling through dysfunctional pancreatic β-cell development (reduced β-cell proliferation, islet size, and islet vascularization) and dysfunctional amino acid metabolism [25, 26]. The postnatal consequences of these changes however have not been thoroughly investigated.

Besides periconceptional undernutrition, a developmental modification at any stage can lead to adverse outcomes. A mismatched pre- and post-natal nutrient environment induces a defective cardiovascular function in adult offspring [27, 28].

The developmental stages are important for the fate of the offspring. Nutrition during pregnancy and lactation is of particular importance, as supported by animal experiments. In a striking example, a low fat diet during pregnancy and lactation was successful in protecting from metabolic disturbances in the offspring of obese mice with type 2 DM [29]. The offspring’s benefits were reduced hyperphagia, increased leanness, normal insulin sensitivity and glycemia.

To what extent pre and periconceptional maternal dietary status affects the health of the mother and the offspring remains to be elucidated. The pre-pregnancy body mass index (BMI) has been argued to be of greater importance than maternal nutrition per se in developing GDM [30]. Indeed, maternal obesity is a risk factor for the development of maternal GDM and strongly predisposes the offspring to obesity, glucose intolerance and IR in successive generations [31]. Nevertheless, GDM is only 1 of the 3 described models, along with protein restriction and caloric restriction, of a fetal environment leading to disturbed fetal development and offspring metabolic profile [23].

A prospective experiment on rats made a considerable effort to clarify the effect of issues surrounding gestational and lactating nutrition on the offspring’s eating behavior, adiposity, circulating glucose and insulin levels, triglyceride and cholesterol concentrations from birth to adolescence [28]. A maternal “junk-food diet” (i.e. rich in fat, sugar and salt) produces rat offspring with increased adiposity, raised circulating glucose, insulin, triglyceride and cholesterol by the end of adolescence in comparison with the offspring of mothers feeding on a balanced chow diet during pregnancy and lactation. The effects are intensified when one considers that both groups of offspring were given free access to junk-food from weaning onwards [28]. The junk-food diet is characterized by high-fat content and high GI and resembles the typical Western diet. These experiments render the carbohydrate quality an important constituent of the maternal diet.

PREGNANCY GLYCEMIA AND INSULINEMIA

A number of metabolic adaptations occur during pregnancy in order for the mother to provide herself as well as the developing fetus a relatively small increase in food intake. Physiologically, progressive IR and compensatory hyperinsulinemia take place to increase the storage of nutrients in maternal fat. Also, the process of slowing glucose uptake into maternal tissues serves to provide nutrients to the fetus. However, pathologic glucose intolerance may appear in the third trimester, without the presence of DM, if the maternal pancreatic β-cells are not able to compensate for their inherent as well as the physiological IR of pregnancy [32].

The fetal pancreas starts developing early in pregnancy, while by mid-gestation and until late gestation it is capable of gradually producing considerable amounts of insulin [33]. The glucose-stimulated insulin secretion of the fetus is
down-regulated by constant hyperglycemia, but enhanced by pulsatile hyperglycemia [34-37]. Recent cell and molecular studies have similarly supported the notion that chronic hyperglycemia down-regulates both fetal glucose tolerance and insulin sensitivity [38].

Although the fetus is known to have a considerable capacity to metabolically adapt to acute and chronic changes in glucose supply, lower maternal blood glucose levels lead to reduced fetal growth rate and infant birth weight [39, 40] while a continuous high maternal blood glucose supply to the fetus may lead to fetal overgrowth and large for gestational-age (LGA) infants [41]. Indeed, numerous animal and human studies demonstrated an association between fetal growth and size at birth with maternal 24-h blood glucose levels [40-43].

Offspring born to GDM mothers with fasting and daily hyperglycemia have higher birth-weight and are frequently overweight in adolescence [44]. Compared with offspring of healthy mothers, offspring of type 2 diabetic mothers show decreased insulin sensitivity in adulthood even in the absence of IGT [45].

In Pima Indians - a population characterized by a high prevalence of type 2 DM - the offspring of type 2 diabetic mothers had a 3.7-fold higher risk of DM and a higher BMI than their siblings born before their mother developed DM [46]. This suggests an important role for the glycemia of the intrauterine environment beyond the possible attributable genetic factors.

The impact of both intrauterine hyperglycemia and genetic predisposition to type 2 DM was evaluated in a population of pregnant females with both GDM and type 1 DM [47]. The hyperglycemic intrauterine environment involved in both types of DM was associated with the pathogenesis of type 2 DM and pre-DM in adult offspring of women with both GDM and type 1 DM during pregnancy [47]. To support the above argument, an abnormal 2 h glucose level during an oral glucose tolerance test is present in the offspring of both type 2 diabetic and normal glucose-tolerant mothers previously exposed to mildly increased blood glucose levels in the third trimester [48, 49].

Even in the absence of abnormal fasting or 2 h glucose values, in utero hyperglycemia in children exposed to GDM increases the presence of metabolic markers of IR such as fasting plasma glucose (FPG) > 6.0 mmol/L, high density lipoprotein (HDL)-cholesterol < 1.03 mmol/L, triglycerides (TG) > 1.24 mmol/L, white cells (WC) > 90% for age and gender, and 2 h glucose > 7.8 mmol/L in these children [50]. At the cellular level, some findings suggest an impaired pancreatic β-cell mass and/or function, arising from possible defects in pancreatic angiogenesis and innervation, abnormal placental glucocorticoid concentrations or modification of parental imprinting [4]. A positive association between postprandial blood glucose levels and measures of fetal adiposity (1 h postprandial glucose and fetal abdominal circumference) is evident in women with normal glucose challenge tests even when the mean measured maternal postprandial glucose levels do not exceed the thresholds for good glucose control in pregestational diabetic pregnancies [51].

Further to the effect of glucose concentrations in the intrauterine environment, insulin itself may program the development of obesity and diabetes when occurring in elevated concentrations during perinatal life. This may occur due to a general increase of fetal food supply (e.g. in overweight pregnant women) and neonatal overfeeding [52].

**CARBOHYDRATES AND GLYCEMIC RESPONSE**

The role of maternal glycemia and insulinemia is well-established in terms of offspring development and growth, but the exact effect of dietary carbohydrates on the metabolic profile is not yet completely understood.

Conflicting evidence arises when examining the effect of total carbohydrate intake on insulin sensitivity [53]. In fact, a recent dietary intervention found that after 6 months on a low-carbohydrate and high-fat diet, insulin sensitivity improved among obese individuals [54]. The source and quality of dietary carbohydrates, however, may differentially affect insulin action and thereby affect the degree of IR.

Observational studies have indeed suggested that fasting insulin concentrations are lower among individuals reporting higher dietary fiber [55-57] or whole-grain intakes [58, 59] after adjustment for other lifestyle and also dietary factors. Furthermore, a number of studies show a beneficial effect on insulin sensitivity with a high consumption of dietary fiber [56] or whole-grain foods [60-63].

The GI is an index measuring the glycemic response to various carbohydrate-containing foods and has been used to qualitatively classify dietary carbohydrates [64], thus providing an insight into the glycemic and insulinemic effect of each type of carbohydrate. The GI is defined as the area under the curve of the glucose responses to a carbohydrate-containing food compared to either a specific glucose dose or a specific amount of white bread [65, 66]. The greater the particle size, the lower the glucose and insulin response. The greater the level of processing and refining, the higher the glycemic response of a particular carbohydrate (Table 1).

**Table 1. Factors Affecting GI of Foods**

| Factor                        |
|-------------------------------|
| Soluble fiber content         |
| Amylose content               |
| Particle size                 |
| Method of preparation         |

Indeed, low GI-resistant starch and soluble fiber together have an additive effect on glycemic response with soluble fiber having a selectively greater effect on postprandial insulin response. In contrast, resistant starch from high-amylase cornstarch greatly affects glucose reduction [67]. Data from older studies also suggest a relationship between high dietary GI and elevated triglyceride concentrations [68], low HDL cholesterol [69], accumulation of fat in liver (non alcoholic fatty liver disease), high adipose tissue and less lean body mass. Also, lower plasma adiponectin, and severe disruption of islet physiology and architecture have been reported for...
animals on high GI diets in comparison to low GI diets [19, 70, 71].

The controversy begins with studies indicating an inverse effect between GI and appetite, energy and food intake. In humans, inclusion of an ingredient containing increased soluble fiber and amylose has been argued to increase subsequent energy intakes despite its reduced glycemic and insulinenic effects [72] Furthermore, carbohydrates with a high GI (glucose, polycose and sucrose) fed in humans suppress subjective appetite and food intake in the short term, in contrast to those with a low GI (amylose and amylopectin) [73]. Whether these studies indicate a missing link between obesity and GI or interpretation of these intervention studies is confounded by the comparison of differing patient populations (e.g. obese non-diabetic subjects, type 2 diabetic subjects), or the differences in age ranges is inconclusive but they do suggest a need for future studies incorporating information such as the variation of the “second meal effect (SME)”.

The SME represents a meal’s ability (carbohydrate content and quality) to diminish the glucose response (reduction of glycemia) to carbohydrates eaten during the following meal. Fermentable carbohydrates (such as fiber and resistant starch) independent on their effect on a food’s GI regulate postprandial responses to a second meal by reducing non esterified fatty acid (NEFA) competition for glucose disposal, and increasing colonic fermentation in humans [74]. In accordance with the above evidence, an association of gut microbial metabolism and key factors associated with IR has been provided, whereby the composition of indigestible carbohydrates of the evening meal affect glycemic fluctuations and related metabolic risk variables at the next-day breakfast meal through a mechanism involving colonic fermentation [75].

Regardless of the timing of the meals, the amount of carbohydrate tested and the mixture of carbohydrate with other macronutrients in complex meals are also important determinants of the level of response. The glycaemic load (GL) is a measure developed to account for a complex meal’s glycemic response [76, 77]. It has been argued to provide more accurate reflections of the meal’s effect on blood glucose levels than the GI but with questionable validity [78, 79].

Another point to consider is the specific characteristics of the subjects to be tested which are important in determining the level of reduction that can be achieved. Therefore, older, less slim, more glucose intolerant subjects have the capacity for greater improvement in glucose and insulin responses than do young, fit, slim subjects.

METABOLIC PROGRAMMING, CARBOHYDRATES AND GI

Maternal glycemia and insulinemia are 2 modifiable factors that are extremely important for the developing fetus. Protein is an important element of the fetal diet, in terms of the effect on the fetus’s metabolic profile [4, 80]. Various experimental studies showed that a high-protein and a low-carbohydrate maternal diet can cause harmful effects, such as raised blood pressure and increased cortisol secretion in response to psychological stress in the offspring [81], outlining the importance of balance, but also introducing carbohydrates as an essential element in the search for the “optimum” pregnancy diet [82, 83]. This is strengthened by the fact that the preferred fuel for fetal growth and development is glucose, a substrate primarily affected by the amount and type of carbohydrate in the maternal diet. In rats, differences in amniotic fluid concentrations of glucose, uric acid and urea nitrogen respond to differing amounts of carbohydrate in the maternal diet [84]. Few studies on the other hand have explored the type of carbohydrate in response to its glycemic response when studying the fetal origins of disease hypothesis [83, 85].

A few studies have supported the notion that the type of carbohydrate in the diet influences glycosylated hemoglobin and maternal plasma glucose levels as well as fetal glucose levels, fetal growth and infant birth weight [22, 85]. In sheep, increased birth weight offspring with a faster growth rate in early postnatal life are produced with maternal trans- sient high glycemic intakes in the third trimester of preg- nancy [86]. Also, high compared with low GI diets during pregnancy were associated with heavier infants who had a higher birth centile, a higher ponderal index and a higher prevalence of large-for-gestational age, all being strong pre- dictors of chronic diseases in later life [22, 85, 87]. In contrast, lighter infants with a 2-fold increased risk of being born small-for-gestational-age were delivered by women on a lower-GI diet [22]. However, the dietary type of carbohydrate was not thoroughly recorded in the lower GI diets group, thus confounding the true effect of the diet’s GI on the delivery outcome.

Reductions of insulin sensitivity in women consuming Western diets in late pregnancy may be due to the diet itself. Low GI maternal diets with a high fiber content even as an addition to a typical high fat and high sucrose Western diet may reduce levels of insulin resistance [88], a finding with therapeutic implications for the supplementation of low GI products on a Western-high-GI diet.

In addition to the effect of malnutrition in pregnancy on the metabolic profile of the offspring, a study on rats attempted to provide proof on a novel property of low GI diets on the fetal origins of disease hypothesis. In this study, nutri- tional recovery of adult offspring exposed to a protein defi- ciency during intrauterine life was evident when they were lactated with a low GI diet (soybean flour) [89]. This can be considered as a dispute of the original idea that a mismatched pre- and post-natal environment would evoke a disturbed metabolic profile and obesity, but appears positively promising in terms of its therapeutic potential.

CONCLUSIONS

Fetal exposure to maternal diabetes and hyperglycemia may contribute considerably to the worldwide diabetes epi- demic. Knowledge of the exact environmental influences that affect the diseases of aging may allow us to find novel ways to increase our life-span and quality [90]. From a clinical point of view, universal screening and therapy for all types of metabolic profile disturbances during pregnancy is recommended. These measures might serve as causal approaches to primary prevention. This may be possible by the acquisition of knowledge of the exact dietary factors that
influence maternal and offspring glucose and insulin metabolism, especially in high-risk populations. This may provide the basis for health interventions targeting individuals who have increased susceptibility to a disturbed metabolism before pregnancy or have already been exposed to a diabetic environment in utero [91].

Recent studies have demonstrated that increasing whole grain intake in the population can result in improved glucose metabolism and delay or reduce the risk of developing type 2 DM. Whole grains can thus provide a substantial contribution to the improvement of the maternal diets, particularly those characterized by the model of western nutrition of high GI carbohydrates and high saturated fatty acid content. A number of different whole grain foods and grain fiber sources were beneficial in IR reduction and improvement in glycemic index (GI), which has not been studied in relation to nutrition [93] and may be a useful parameter describing the destiny of the offspring. Future studies should evaluate the importance of this parameter.

The importance of a balanced maternal diet is emphasized, especially in terms of carbohydrate quality in pregnancy and lactation, for the prevention of diet-induced adiposity and associated metabolic disruptions in the offspring. However, more longitudinal studies are required to ascertain which aspects of carbohydrate nutrition are linked to the development of the MetS. Additionally, further research is required to define the optimal range of maternal glucose, which results in a good pregnancy outcome.

Cost-effective screening strategies for early detection of children at risk, as well as long-term observational and interventional studies around conception and during pregnancy are needed. This will help provide appropriate knowledge to target susceptible mothers and offspring for metabolic disturbances in later life.

REFERENCES

[1] Cali AM, Caprio S. Obesity in children and adolescents. J Clin Endocrinol Metab 2008; 93: S31-6.
[2] Esposito K, Cirotola M, Maiorino MI, Giugliano D. Lifestyle approach for type 2 diabetes and metabolic syndrome. Curr Atheroscler Rep 2008; 10: 523-8.
[3] Jovanovic L. Nutrition and pregnancy: the link between dietary intake and diabetes. Curr Diab Rep 2004; 4: 266-72.
[4] Stocker CJ, Arch JR, Cawthorne MA. Fetal origins of IR and obesity. Proc Nutr Soc 2005; 64: 143-51.
[5] Hales CN. Fetal and infant growth and impaired glucose tolerance in adulthood: the "thrifty phenotype" hypothesis revisited. Acta Paediatr Suppl 1997; 422: 73-7.
[6] Jackson AA. Nutrients, growth, and the development of programmed metabolic function. Adv Exp Med Biol 2000; 478: 41-55.
[7] Wells JC. The thrifty phenotype as an adaptive maternal effect. Biol Rev Camb Philos Soc 2007; 82: 143-72.
[8] Cleal JK, Poore KR, Boullin JP, et al. Mismatched pre- and postnatal nutrition leads to cardiovascular dysfunction and altered renal function in adulthood. Proc Natl Acad Sci USA 2007; 104: 9529-33.
[9] Cameron N, Demerath EW. Critical periods in human growth and their relationship to diseases of aging. Am J Phys Anthropol 2002; Suppl 35: 159-84.
[10] Mead MN. You are what your mother ate. Environ Health Perspect 2007; 115: A492-3.
[11] McArdle HJ, Andersen HS, Jones H, Gambling L. Fetal programming: causes and consequences as revealed by studies of dietary manipulation in rats -- a review. Placenta 2006; 27 Suppl A: S56-60.
[12] Weiss PA, Scholz HS, Haas J, Tamussino KF, Seissler J, Borkenstein MH. Long-term follow-up of infants of mothers with type 1 diabetes: evidence for hereditary and nonhereditary transmission of diabetes and precursors. Diabetes Care 2000; 23: 905-11.
[13] Pettitt DJ, Baird HR, Alec KA, Bennett PH. Knowler WC. Excessive obesity in offspring of Pima Indian women with diabetes during pregnancy. N Engl J Med 1983; 308: 242-5.
[14] Dabelea D, Hanson RL, Lindsay RS, et al. Intrauterine exposure to diabetes conveys risks for type 2 diabetes and obesity: a study of discordant sibships. Diabetes 2000; 49: 2208-11.
[15] Clausen TD, Mathiesen ER, Hansen T, et al. High prevalence of type 2 diabetes and pre-diabetes in adult offspring of women with gestational diabetes mellitus or type 1 diabetes: the role of in utero hyperglycemia. Diabetes Care 2008; 31: 340-6.
[16] Silverman BL, Metzger BE, Cho NH, Loeb CA. Impaired glucose tolerance in adolescent offspring of diabetic mothers. Relationship to fetal hyperinsulinism. Diabetes Care 1995; 18: 611-7.

[17] Plagemann A, Harder T, Kohlhoff R, Rohde W, Dorner G. Glucose tolerance and insulin secretion in children of mothers with pregestational IDDM or gestational diabetes. Diabetologia 1997; 40: 1094-100.

[18] Radesky JS, Oken E, Rifas-Shiman SL, Kleinman KP, Rich-Edwards JW, Gillman MW. Diet during early pregnancy and development of gestational diabetes. Paediatri Perinat Epidemiol 2008; 22: 47-59.

[19] Pawlak DB, Kushner JA, Ludwig DS. Effects of dietary glycemic index on adiposity, glucose homeostasis, and plasma lipids in animals. Lancet 2004; 364: 778-85.

[20] Walker CG, Bryson JM, Phuyal JL, Caterson ID. Dietary modulation of circulating leptin levels: site-specific changes in fat deposition and ob mRNA expression. Horm Metab Res 2002; 34: 176-81.

[21] Scribner KB, Pawlak DB, Ludwig DS. Hepatic steatosis and increased adiposity in mice consuming rapidly vs. slowly absorbed carbohydrate. Obesity (Silver Spring) 2007; 15: 2190-9.

[22] Scholl TO, Chen X, Khoo CS, Lenders C. The dietary glycemic index during pregnancy: influence on infant birth weight, fetal growth, and biomarkers of carbohydrate metabolism. Am J Epidemiol 2004; 159: 467-74.

[23] Reusens B, Remacle C. Intergenerational effect of an adverse intrauterine environment on perturbation of glucose metabolism. Twin Res 2001; 4: 406-11.

[24] Reusens, B, Ozanne SE, Remacle C. Fetal determinants of type 2 diabetes. Curr Drug Targets 2007; 8: 935-41.

[25] Oliver MH, Jaquerey AL, Bloomfield FH, Harding JE. The effects of maternal nutrition around the time of conception on the health of offspring. Soc Reprod Fertil Suppl 2007; 64: 397-410.

[26] Husted SM, Nielsen MO, Tygesen MP, Kiani A, Blache D, Ingvarsens KL. Programming of intermediate metabolism in young lambs affected by late gestational maternal undernourishment. Am J Physiol Endocrinol Metab 2007; 293: E548-57.

[27] Cleal JK, Poore KR, Boullin JP, et al. Mismatched pre- and postnatal nutrition leads to cardiovascular dysfunction and altered renal function in adulthood. Proc Natl Acad Sci USA 2007; 104: 9529-33.

[28] Bayol SA, Simbi BH, Bertrand JA, Stickland NC. Offspring from mothers fed a 'junk food' diet in pregnancy and lactation exhibit exacerbated adiposity that is more pronounced in females. J Physiol 2008; 586: 3219-30.

[29] Gallo-Kabani C, Vige A, Gross MS, et al. Resistance to high-fat diet in the female progeny of obese mice fed a control diet during the periconceptual, gestation, and lactation periods. Am J Physiol Endocrinol Metab 2007; 292: E1099-100.

[30] Radesky JS, Oken E, Rifas-Shiman SL, Kleinman KP, Rich-Edwards JW, Gillman MW. Diet during early pregnancy and development of gestational diabetes. Paediatri Perinat Epidemiol 2008; 22: 47-59.

[31] de Campos KE, Sinzato YK, Pimenta Wde P, Rudge MV, Damasceno DC. Effect of maternal obesity on diabetes development in adult rat offspring. Life Sci 2007; 81: 1473-8.

[32] Buchanan TA. Glucose metabolism during pregnancy: normal physiology and implications for diabetes mellitus. Isr J Med Sci 1991; 27: 432-41.

[33] Aldoretta PW, Carver TD, Hay WW, Jr. Maturation of glucose-stimulated insulin secretion in fetal sheep. Biol Neonate 1998; 73: 376-86.

[34] Carver TD, Anderson SM, Aldoretta PA, Esler AL, Hay WW, Jr. Glucose suppression of insulin secretion in chronically hyperglycemic fetal sheep. Pediatr Res 1995; 38: 574-62.

[35] Aldoretta PW, Hay WW, Jr. Chronic hyperglycemia induces insulin resistance and glucose intolerance in fetal sheep. Pediatr Res 2001; 49: 307A, abstract 1758.

[36] Limesand SW, Jensen I, Hutton JC, Hay WW, Jr. Diminished beta-cell replication contributes to reduced beta-cell mass in fetal sheep with intrutaneous growth restriction. Am J Physiol Regul Integr Comp Physiol 2005; 288: 1297-305.

[37] Carver TD, Anderson SM, Aldoretta PW, Hay WW Jr. Effect of low-level basal plus marked “pulsatile” hyperglycemia on insulin secretion in fetal sheep. Am J Physiol 1996; 271: 865-71.

[38] Hay WWW Jr. Placental-fetal glucose exchange and fetal glucose metabolism. Transacnt Am Clin Climatolog Assoc 2006; 117: 321-39.

[39] Smith CA. Effects of maternal undernutrition upon the newborn infant in Holland (1944-45). J Pediatr 1947; 30: 229-43.

[40] Mellor DJ. Nutritional and placental determinants of fetal growth rate in sheep and consequences for the newborn lamb. Br Vet J 1983; 139: 307-24.

[41] Langer O, Anyaegbunam A, Brustman L, Divon M. Management of women with one abnormal oral glucose tolerance test value reduces adverse outcome in pregnancy. Am J Obst Gynecol 1989a; 161: 593-99.

[42] Langer O, Levy J, Brustman L, Anyaegbunam A, Merkatz R. Divon M. Glycemic control in diabetes mellitus – how tight is tight enough: small for gestational age versus large for gestational age? Am J Obst Gynecol 1989b; 161: 646-53.

[43] Jovanovic-Peterson L, Peterson CM, Reed GF, et al. Maternal postpartum glucose levels and infant birth weight: the Diabetes and Early Pregnancy Study: The National Institute of Child Health and Human Development-Diabetes in Early Pregnancy Study. Am J Obst Gynecol 1991; 164: 103-11.

[44] Buzinario, EF, Berchieri CB, Haddad AL, Padovani CR, Pimenta Wde P. Overweight in adolescent offspring of women with hyperglycemia during pregnancy. Arq Bras Endocrinol Metabol 2008; 52: 85-92.

[45] Hunter WA, Candy T, Rabone D, et al. Insulin sensitivity in the offspring of women with type 1 and type 2 diabetes. Diabetes Care 2004; 27: 1148-52.

[46] Dabelea D, Hanson RL, Lindsay RS, et al. Intrauterine exposure to diabetes conveys risks for type 2 diabetes and obesity: a study of discordant siblings. Diabetes 2000; 49: 2208-11.

[47] Claesson TD, Mathiesen ER, Hansen T, et al. High prevalence of type 2 diabetes and pre-diabetes in adult offspring of women with gestational diabetes mellitus or type 1 diabetes: the role of intrauterine hyperglycemia. Diabetes Care 2008; 31: 340-6.

[48] Franks PW, Looker HC, Kobes S, et al. Gestational glucose tolerance and risk of type 2 diabetes in young Pima Indian offspring. Diabetes 2006; 55: 460-5.

[49] BPDPO Study Cooperative Research Group, Metzger BE, Lowe LP, Dyer AR, et al. Hyperglycemia and adverse pregnancy outcomes. New Engl J Med 2008; 358: 1991-2002.

[50] Keely EJ, Malcolm JC, Hadjiyannakis S, Gaboury I, Lough G, Lawson ML. Prevalence of metabolic markers of IR in offspring of gestational diabetes pregnancies. Pediatr Diabetes 2008; 9: 53-9.

[51] Parretti E, Mecacci F, Papini M, et al. Third-trimester maternal glucose levels from diurnal profiles in nondiabetic pregnancies: correlation with sonographic parameters of fetal growth. Diabetes Care 2001; 24: 1319-23.

[52] Plagemann A. A matter of insulin: Developmental programming of body weight regulation. J Matern Neonatal Med 2008; 21: 435-48.

[53] Daly ME, Vale CV, Walker M, Alberti KG, Mathers JC. Dietary carbohydrates and insulin sensitivity: a review of the evidence and clinical implications. Am J Clin Nutr 1997; 66: 1072-85.

[54] Samaha FP, Iqbal N, Seshardi P, et al. A low-carbohydrate as compared with a low fat diet in severe obesity. N Engl J Med 2003; 348: 2074-81.

[55] Ludwig DS, Pereira MA, Kroecken CH, et al. Dietary fiber, weight gain, and cardiovascular disease risk factors in young adults. JAMA 1999; 282: 1539-46.

[56] Marshall JA, Bessesen DH, Hamman RF. High saturated fat and low starch and fibre are associated with hyperinsulinemia in a non-diabetic population: the San Luis Valley Diabetes study. Diabetes 1997; 46: 1943-8.

[57] Feskens EJ, Loeber JG, Kromhout D. Diet and physical activity as non-diabetic population: the San Luis Valley Diabetes study. Diabetes 1997; 46: 1943-8.

[58] Dalay ME, Vale CV, Walker M, Alberti KG, Mathers JC. Dietary carbohydrates and insulin sensitivity: a review of the evidence and clinical implications. Am J Clin Nutr 1997; 66: 1072-85.

[59] Samaha FP, Iqbal N, Seshardi P, et al. A low-carbohydrate as compared with a low fat diet in severe obesity. N Engl J Med 2003; 348: 2074-81.

[60] Ludwig DS, Pereira MA, Kroecken CH, et al. Dietary fiber, weight gain, and cardiovascular disease risk factors in young adults. JAMA 1999; 282: 1539-46.
Nutrition During Pregnancy and the Effect of Carbohydrates

Sensitivity in healthy young and old adults. Am J Clin Nutr 1990; 52: 524-28.

Pereira MA, Jacobs DR Jr, Pins J, et al. Effect of whole grains on insulin sensitivity in overweight hyperinsulinemic adults. Am J Clin Nutr 2002; 75: 848-55.

Davy BM, Davy KP, Ho RC, Beske SD, Davrath LR, Melby CL. High-fiber oat cereal compared with wheat cereal consumption favorably alters LDL-cholesterol subclass and particle numbers in middle-aged and older men. Am J Clin Nutr 2002; 76: 351-8.

Juntunen KS, Laaksonen DE, Poutanen KS, Niskanen LK, Mykkänen HM. High fiber rye bread and insulin secretion and sensitivity in healthy postmenopausal women. Am J Clin Nutr 2003; 77: 385-390.

Jenkins DJ, Wolever TM, Taylor RH, et al. Glycemic index of foods. A physiological basis for carbohydrate exchange. Am J Clin Nutr 1981; 34: 362-6.

Reaven G, Miller R. Study of the relationship between glucose and insulin responses to an oral glucose load in man. Diabetes 1968; 17: 560-9.

Wolever TM, Jenkins DJ, Jenkins AL, Josse RG. The glycaemic index: methodology and clinical implications. Am J Clin Nutr 1991; 54: 846-54.

Behall KM, DJ Schoffield, Halfrisch JG, Liljeborg-Elnmahl HG. Consumption of both resistant starch and beta-glucan improves postprandial plasma glucose and insulin in women. Diabetes Care 2006; 29: 976-81.

Liu S, Manson JE, Stampfer MJ, et al. Dietary glycaemic load assessed by food-frequency questionnaire in relation to plasma high-density-lipoprotein cholesterol and fasting plasma triacylglycerols in postmenopausal women. Am J Clin Nutr 2001; 73: 560-66.

Frost G, Leeds AA, Dore CJ, Madeiros S, Brading S, Dornhorst A. Glycaemic index as a determinant of serum HDL-cholesterol concentration. Lancet 1999; 353: 1045-48.

Scribner KB, Pawlak DB, Ludwig DS, et al. Hepatic steatosis and increased adiposity in mice consuming rapidly vs. slowly absorbed carbohydrate. Obesity 2007; 15: 2190-99.

Kabir M, Rizkalla SW, Champ M, et al. Dietary amylose-amylopectin starch content affects glucose and lipid metabolism in adipocytes of normal and diabetic rats. J Nutr 1998; 128: 35-43.

Keogh JB, Lau CW, Noakes M, Bowen J, Clifton PM. Effects of meals with high soluble fibre, high amylose barley variant on glucose, insulin, satiety and thermic effect of food in healthy lean women. Eur J Clin Nutr 2007; 61: 597-604.

Anderson, GH, Catherine NL, Woodend DM, Wolever TM. Inverse association between the effect of carbohydrates on blood glucose and subsequent short-term food intake in young men. Am J Clin Nutr 2002; 76: 1023-30.

Brighenti F, Benini L, Del Rio D, et al. Colonic fermentation of indigestible carbohydrates contributes to the second-meal effect. Am J Clin Nutr 2006; 83: 817-22.

Nilsson AC, Ostman EM, Holst JJ, Bjorck IM. Including indigestible carbohydrates in the evening meal of healthy subjects improves glucose tolerance, lowers inflammatory markers, and increases satiety after a subsequent standardized breakfast. Am J Physiol 2008; 138: 732-9.

Salmeron J, Manson JE, Stampfer MJ, Colditz GA, Wing AL, Willett WC. Dietary fiber, glycemic load, and risk of non-insulin-dependent diabetes mellitus in women. JAMA 1997; 277: 472-7.

Galgani J, Aguirre C, Diaz E. Acute effect of meal glycemic index and glycemic load on blood glucose and insulin responses in humans. Nutr J 2006; 5: 22.

Wolever T. Comment on validity of the glycemic glucose equivalent. Eur J Clin Nutr 2004; 58: 1672-3.

Thomas D, Elliott EJ. Low glycemic load, diets for diabetes mellitus. Cochrane Database Syst Rev 2009; (1): CD006296.

Zambrano E, Bautista CJ, Deas M, et al. A low maternal protein diet during pregnancy and lactation has sex- and window of exposure-specific effects on offspring growth and food intake, glucose metabolism and serum leptin in the rat. J Physiol 2006; 571: 221-30.

Koski KG, Hill FW. Effect of low carbohydrate diets during pregnancy on parturition and postnatal survival of the newborn rat pup. J Nutr 1986; 116: 1938-48.

Reynolds RM, Godfrey KM, Barker M, Osmond C, Phillips DJ. Stress responsiveness in adult life: influence of mother's diet in late pregnancy. J Clin Endocrinol Metab 2007; 92: 2208-10.

Fergusson MA, Koski KG. Comparison of effects of dietary glucose versus fructose during pregnancy on fetal growth and development in rats. J Nutr 1990; 120: 1312-19.

Koski KG, Fergusson MA. Amniotic fluid composition responds to changes in maternal dietary carbohydrate and is related to metabolic status in term fetal rats. J Nutr 1992; 122: 385-92.

Moses RG, Luebeke M, Davis WS, et al. Effect of a low-glycemic-index diet during pregnancy on obstetric outcomes. Am J Clin Nutr 2006; 84: 807-12.

Smith N, McAuliffe F, Quinn K, Lonnergan P, Evans AC. Transient high glycemic intake in the last trimester of pregnancy increases offspring birthweight and postnatal growth rate in sheep: a randomised control trial. BJO 2009; 116: 975-83.

Roman M, Nuttens M, Vamberege A, et al. Higher carbohydrate intake is associated with decreased incidence of newborn macrosomia in women with gestational diabetes. J Am Diet Assoc 2001; 101: 897-902.

Fraser RB, Ford FA, Lawrence GF. Insulin sensitivity in third trimester pregnancy. A randomized study of dietary effects. Br J Obst Gynecol 1988; 95: 223-9.

de Arruda Oliveira E, Gomes Cheim LM, et al. Nutritional recovery with a soybean flour diet improves the insulin response to a glucose load without modifying glucose homeostasis. Nutrition 2008; 24: 76-83.

Bunout D, Cambiazo V. Nutrition and aging. Rev Med Chil 1999; 127: 82-88.

Fetita LS, Sobngwi E, Serradas P, Calvo F, Gautier JC. Consequences of fetal exposure to maternal diabetes in offspring. J Clin Endocrinol Metab 2006; 91: 3718-24.

Du H, Van der A DL, Feskens EJ. Dietary glycemic index: a review of the physiological mechanisms and observed health impacts. Acta Cardiol 2006; 61: 383-97.

Demerath EW, Cameron N, Gillman MW, Towne B, Siervogel RM. Telomeres and telomerase in the fetal origins of cardiovascular disease: a review. Hum Biol 2004; 76: 127-46.

Gardner JP, Li S, Srinivasan SR, et al. Rise in IR Is Associated With Escalated Telomere Attrition. Circulation 2005; 111: 2171-7.