Case Report

Microscopic Polyangiitis Initially Presumed to Be Endocarditis

Takahiro Kaneko, Shunsuke Hino, Yosuke Iijima, and Norio Horie

Department of Oral and Maxillofacial Surgery, Saitama Medical Center, Saitama Medical University, Saitama, Japan

Correspondence should be addressed to Takahiro Kaneko; t_kaneko@saitama-med.ac.jp

Received 6 January 2020; Accepted 5 March 2020; Published 16 March 2020

Academic Editor: Ronald S. Brown

Copyright © 2020 Takahiro Kaneko et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The antineutrophil cytoplasmic antibody- (ANCA-) associated vasculitides (AAVs), which include fever of unknown origin (FUO), are rare diseases characterized by necrotizing inflammation of small blood vessels and the presence of ANCAs. Microscopic polyangiitis (MPA) is a subtype of the AAVs. Although the prevalence of AAVs has generally increased over the last 20 years, there have been rare reports from the dental and oral surgery field. In this article, we present a case of MPA suspected to be infective endocarditis (IE) following tooth extraction.

1. Introduction

Fever of unknown origin (FUO) can be caused by a wide group of diseases including infections, noninfectious inflammatory diseases, malignancies, and miscellaneous diseases [1]. However, diagnosing certain causes of FUO has remained difficult. The antineutrophil cytoplasmic antibody- (ANCA-) associated vasculitides (AAVs) are rare diseases included in FUO characterized by necrotizing inflammation of small blood vessels and the presence of ANCAs [2]. AAVs can be classified into three phenotypically distinct processes: microscopic polyangiitis (MPA), granulomatosis with polyangiitis (GPA), and eosinophilic granulomatosis with polyangiitis (EGPA). Two main types of ANCA can be detected in patients with AAV: leukocyte proteinase 3 (PR3) and myeloperoxidase (MPO) [2]. Patients with GPA are predominantly PR3-ANCA-positive, whereas those with MPA or EGPA are predominantly MPO-ANCA-positive [2]. These antibodies are checked not only at diagnosis but also in follow-up as disease activity markers [3].

The prevalence of AAV is estimated to be 46–184/million people, and the numbers of patients with AAV have generally increased [4]. There is a relatively high prevalence of AAV among middle-aged and elderly persons [5], affecting both sexes equally (quite unlike most systemic autoimmune diseases) [5]. There is a geographic difference in the occurrence of AAVs.

Recently considered risk factors for AAVs include ultraviolet radiation and silica, which is commonly inhaled [4]. Although there have been studies that suggested infection as a risk factor, the long-held view that there is an infectious trigger has not been borne out [4].

The clinical presentation of AAVs begins with fever and weight loss, followed by systemic vasculitis characterized by renal and respiratory dysfunction [6]. In cases of MPA, necrotizing glomerulonephritis occurs rapidly, and it frequently progresses to renal failure [6]. Laboratory tests show increased C-reactive protein (CRP) and serum creatinine levels with increased MPO-ANCA [7]. In GPA, hemorrhages of the lung and upper respiratory tract and destructive lesions of the ear, nose, and throat are prominent [8]. EGPA patients typically have a long-standing history of asthma and allergic rhinitis [9].

Induction therapy for AAVs includes treatment with high-dose glucocorticoids (GCs) in combination with another immunosuppressive agent such as cyclophosphamide (CYC) or rituximab (RTX) [3]. RTX is more effective than CYC in refractory AAV’s [3]. Patients with AAVs presenting with severe renal failure may be treated with pulsed CYC and GCs, with adjuvant plasma exchange [3].

Patients with FUO may visit dental or oral surgery clinics following recent dental treatment or have discomfort in the oral cavity. In this article, a rare case of MPA that was first referred to an oral surgery clinic for postextraction fever with
dysgeusia and numbness of the tongue was presented. Initially, the authors suspected that the patient had infective endocarditis (IE) and thus consulted with specialists in cardiology. However, IE was ruled out, with further detailed evaluations determining the presence of MPA.

2. Case Report

A 75-year-old woman was referred to the oral surgery clinic for a suspected dental infection. Her medical history included tuberculosis, hypertension, hyperlipidemia, and neurogenic bladder. Her surgical history included hip joint surgery at the ages of 11, 27, and 68 years. Four weeks earlier, the patient visited an otolaryngologist for otorrhea of the right ear and was diagnosed with otitis media and treated with antibiotics. On the same day, the patient underwent extraction of the right mandibular canine and lateral incisor by the patient’s family dentist. Two weeks earlier, the patient had visited the primary care doctor for regular follow-up and complained of fatigue, fever, dysgeusia, swelling and numbness of the neck, and backache, which continued after the extraction. On laboratory examination, C-reactive protein (CRP) was 11.46 mg/dL, and the white blood cell count was 11,000/μL. Subsequently, chest X-ray and computed tomography (CT) showed no abnormal findings, and she tested negative for influenza virus. The patient then developed trismus and pain at the apex of the tongue and was referred to the oral surgery clinic.

On examination, the patient complained of fever and discomfort of the tongue with dysgeusia. Although usually both the maxilla and mandible cause dentures to wear, this does not occur when the teeth have been extracted. Salivary function was relatively good, and there was no xerostomia observed. No trismus was found; the teeth extraction sockets had healed well. None of the other residual teeth exhibited any prominent caries or periodontal disease. There were no abnormal findings in the orofacial region except for the dysgeusia and redness and numbness at the apex of the tongue (Figure 1). Orthopantomography revealed no abnormal findings, including maxillary sinuses and temporomandibular joints (Figure 2). On physical examination, the patient’s temperature was 38.5°C, and blood pressure was 125/75 mmHg. The serum CRP concentration was 19.4 mg/dL, and the WBC count was 14,700/μL (Table 1). Given the history of tooth extraction, the patient was immediately referred to the division of cardiology for suspected IE.

The cardiologist took blood cultures, which were negative for bacteria, and an ultrasound cardioagram and transesophageal echocardiography showed no vegetations or peridontal disease. There were no abnormal findings of necrotizing glomerulonephritis, the patient was finally diagnosed with MPA (fever, MPO-ANCA-positive, CRP elevation, proteinuria, and histopathological findings of renal biopsy) 40 days after the first visit. Steroid therapy was then started with 30 mg prednisolone/day without immunosuppressive agents, and four days later, the patient was discharged. The inflammatory symptoms improved well, and the dysgeusia and numbness at the apex of the tongue disappeared four weeks later. At 20-week follow-up, prednisolone was tapered to 10 mg/day, and she was MPO-ANCA-negative.

The protocol of the report has been approved by the ethical review board of the university.

3. Discussion

To finally diagnose cases of FUO, detailed medical examination with various investigations and a longer time is needed. Dentists and oral surgeons should refer patients with fever to a physician immediately when the odontogenic diseases do not appear related to the cause of the fever. In addition, other
differential diagnoses with FUO that may be initially encountered include Castleman’s disease and adult-onset Still’s disease [10].

In the present study, it seemed appropriate to initially diagnose the patient with suspected IE, because dentists are extremely well aware of the association between tooth extraction and IE. IE is defined as an infection of a native or prosthetic heart valve, the endocardial surface, or of an indwelling cardiac device. In this case, it was obviously important to refer the patient to a physician for meticulous examination. However, it is of note that there were oral symptoms that were overlooked and might suggest the occurrence of an AAV. The patient showed dysgeusia that had lasted for four weeks. AAVs frequently affect the peripheral and central nervous systems [11], and there was a case report of MPA that described multiple cranial neuropathies including dysgeusia [12]. As a matter of fact, diseases with both continuous fever and dysgeusia are few [13]. The patient felt numbness at the apex of the tongue. Numbness of the extremities and face has been reported in AAVs [14]. The numbness of the tongue may also be a sensory symptom caused by AAVs. In the present case, the dysgeusia and numbness of the tongue improved with the steroid therapy for MPA.

The patient had developed otitis media just before the tooth extractions. There are cases of AAVs that are complicated with otitis media, and the concept of otitis media with granulation resistant to antibiotics and insertion of tympanic ventilation tubes. It is unclear whether the otitis media in the present case could be considered OMAAV, because dentists are extremely well aware of the association between tooth extraction and IE. IE is defined as an infection of a native or prosthetic heart valve, the endocardial surface, or of an indwelling cardiac device. In this case, it was obviously important to refer the patient to a physician for meticulous examination.

The criteria of OMAAV include otitis media with exacerbation or granulation resistant to antibiotics and insertion of tympanic ventilation tubes. It is unclear whether the otitis media in the present case could be considered OMAAV, because it improved before steroid therapy.

GPA, which was formerly known as Wegener’s granulomatosis, has characteristic oral lesions of gingival swellings.

Table 1: Laboratory data of the first examination.

| Test                  | Normal range | Normal range |
|-----------------------|--------------|--------------|
| Complete blood cell count |              |              |
| WBC (×10⁹/µL)         | 14.7         | 3.25–8.59    |
| RBC (×10⁹/µL)         | 4.29         | 3.58–4.90    |
| Platelets (×10⁹/µL)   | 530          | 148.00–336.00|
| Blood chemistry       |              |              |
| TP (g/dL)             | 6.5          | 6.50–8.00    |
| CPK (U/L)             | 29           | 0.00–140.00  |
| ALT (U/L)             | 53           | 4.00–43.00   |
| ALP (U/L)             | 426          | 115.00–280   |
| Creatinine (mg/dL)    | 0.70         | 0.30–1.00    |
| UA mg/dL              | 1.4          | 2.00–6.20    |
| Na (mEq/L)            | 138          | 136.00–145.00|
| K (mEq/L)             | 3.6          | 3.30–5.00    |
| Zn (µg/dL)            | 57           | 65.00–110.00 |
| CH50                  | 60           | 31.60–57.60  |
| Procalcitonin (ng/mL) | 0.1          | 0.00–0.40    |
| Ferritin (ng/mL)*     | 471          | 0.00–120.00  |
| RF (IU/mL)*           | <11          | 0.00–15.00   |
| Immune tests          |              |              |
| β-D-Glucan (pg/mL)    | ≤5.0         | 0.00–10.90   |
| Cryoglobulin (-)      | (-)          | (-)          |
| MPO-ANCA (U/mL)*      | 124.0        | 0.00–3.40    |
| Viral tests           |              |              |
| CMV-CF                | 8            | 0.00–3.00    |
| Tumor markers         |              |              |
| CEA-FEIA (ng/mL)*     | 2.3          | 0.00–6.70    |

These data include results of both examinations performed at the Oral and Maxillofacial Clinic and the Internal Medicine Clinic. *Data obtained later (day six to day ten after the first visit). WBC: white blood cell (count); RBC: red blood cell (count); TP: total protein; ALB: albumen; CPK: creatine phosphokinase; AST: aspartate aminotransferase; ALT: alanine aminotransferase; LDH: lactate dehydrogenase; ALP: alkaline phosphatase; γ-GTP: glutamyl transpeptidase; BUN: blood urea nitrogen; UA: uric acid; eGFRcreat: estimated glomerular filtration rate based on serum creatinine; T-Bil: total bilirubin; CH50: 50% hemolytic complement activity; CRP: C-reactive protein; cTnI: cardiac troponin I; ASO: anti-streptolysin O antibody; RF: rheumatoid factor; ANA: antinuclear antibody; PR3-ANCA: protease 3-antineutrophil cytoplasmic antibody; MPO-ANCA: myeloperoxidase-antineutrophil cytoplasmic antibody; CMV-CF: cytomegalovirus complement fixation test; EB Anti-VCA/IgM-FA: Epstein-Barr antiviral capsid antigens/IgM fluorescence assay; CEA-FEIA: carcinoembryonic antigen fluorometric enzyme immunoassay; CA19-9: carbohydrate antigen 19-9.
and nonspecific ulcerations, which are identified in 6-13% of cases [16]. However, there has been no report describing the other characteristics of AAV in the oral cavity. The dysgeusia and numbness of the tongue in the present case might be manifestations of AAVs that can be found in the oral cavity.

The patient's medical history revealed there were several previous issues, including tuberculosis, hypertension, hyperlipidemia, and neurogenic bladder. However, there was no indication of any association between these diseases and the AAVs.

4. Conclusion

The prevalence of AAVs has generally increased over the last 20 years, and AAVs are associated with high morbidity or mortality, especially if not immediately diagnosed and treated [4, 17]. Therefore, dentists and oral surgeons should keep AAVs in mind. At the same time, there may be hidden oral signs, such as the dysgeusia and numbness of the tongue in the present case, that may help make the diagnosis.

Consent

Informed consent was obtained from an individual participant included in the study.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

[1] B. A. Cunha, “Fever of unknown origin: focused diagnostic approach based on clinical clues from the history, physical examination, and laboratory tests,” Infectious Disease Clinics of North America, vol. 21, no. 4, pp. 1137–87, xi, 2007.
[2] D. Cornec, E. Cornec-Le Gall, F. C. Fervenza, and U. Specks, “ANCA-associated vasculitis: clinical utility of using ANCA specificity to classify patients,” Nature Reviews Rheumatology, vol. 12, no. 10, pp. 570–579, 2016.
[3] E. Ntatski, D. Carruthers, K. Chakravarty et al., “BSR and BHPR guideline for the management of adults with ANCA-associated vasculitis,” Rheumatology, vol. 53, no. 12, pp. 2306–2309, 2014.
[4] R. A. Watts, A. Mahr, A. J. Mohammad, P. Gatenby, N. Basu, and L. F. Flores-Suárez, “Classification, epidemiology and clinical subgrouping of antineutrophil cytoplasmic antibody (ANCA)-associated vasculitis,” Nephrology, Dialysis, Transplantation, vol. 30, supplement_1, pp. i14–i22, 2015.
[5] A. Mahr, L. Guilevin, M. Poissonnet, and S. Aymé, “Prevalences of polyarteritis nodosa, microscopic polyangiitis, Wegener’s granulomatosis, and Churg-Strauss syndrome in a French urban multiethnic population in 2000: a capture-recapture estimate,” Arthritis Care & Research, vol. 51, no. 1, pp. 92–99, 2004.
[6] J. C. Jannette and R. J. Falk, “Small-vessel vasculitis,” The New England Journal of Medicine, vol. 337, no. 21, pp. 1512–1523, 1997.
[7] M. A. Hamidou, A. Moreau, C. Toquet, D. Kouri, P. de Faucault, and J. Y. Grolleau, “Temporal arteritis associated with systemic necrotizing vasculitis,” The Journal of Rheumatology, vol. 30, no. 10, pp. 2165–2169, 2003.
[8] S. Lionaki, E. R. Blyth, S. L. Hogan et al., “Classification of antineutrophil cytoplasmic autoantibody vasculitides: the role of antineutrophil cytoplasmic autoantibody specificity for myeloperoxidase or proteinase 3 in disease recognition and prognosis,” Arthritis and Rheumatism, vol. 64, no. 10, pp. 3452–3462, 2012.
[9] C. Comarmond, C. Pagnoux, M. Khllef and al., “Eosinophilic granulomatosis with polyangiitis (Churg-Strauss): clinical characteristics and long-term followup of the 383 patients enrolled in the French Vasculitis Study Group cohort,” Arthritis and Rheumatism, vol. 65, no. 1, pp. 270–281, 2013.
[10] S. Hino, A. Nakamura, T. Kaneko, N. Horie, and T. Shimoyama, “Adult-onset Still’s disease initially thought to be an odontogenic infection: a case report,” Gerodontology, vol. 35, no. 2, pp. 139–142, 2018.
[11] J. H. Schirmer, M. N. Wright, R. Vonthin et al., “Clinical presentation and long-term outcome of 144 patients with microscopic polyangiitis in a monocentric German cohort,” Rheumatology, vol. 55, no. 1, pp. 71–79, 2016.
[12] A. Morinaga, K. Ono, K. Komai, and M. Yamada, “Microscopic polyangiitis presenting with temporal arthritis and multiple cranial neuropathies,” Journal of the Neurological Sciences, vol. 256, no. 1-2, pp. 81–83, 2007.
[13] E. Nwokolo, C. Ujuju, J. Anyanti et al., “Misuse of artemisinin combination therapies by clients of medicine retailers suspected to have malaria without prior parasitological confirmation in Nigeria,” International Journal of Health Policy and Management, vol. 7, no. 6, pp. 542–548, 2018.
[14] T. Kanda, H. Tanio, C. Wu, H. Nishihara, F. Nogaki, and T. Ono, “Churg-Strauss syndrome with severe granulomatous angitis and crescentic glomerulonephritis, which developed during therapy with a leukotriene receptor antagonist,” Clinical and Experimental Nephrology, vol. 14, no. 6, pp. 602–607, 2010.
[15] Y. Hisata, E. Sasaki, K. Ishimaru et al., “Do you know otitis media with ANCA-associated vasculitis (OMAAV)?,” Journal of General and Family Medicine, vol. 18, no. 6, pp. 428–431, 2017.
[16] F. P. Fonseca, B. M. Benites, A. Ferrari et al., “Gingival granulomatosis with polyangiitis (Wegener’s granulomatosis) as a primary manifestation of the disease,” Australian Dental Journal, vol. 62, no. 1, pp. 102–106, 2017.
[17] D. Geetha, Q. Jin, J. Scott et al., “Comparisons of guidelines and recommendations on managing antineutrophil cytoplasmic antibody-associated vasculitis,” Kidney International Reports, vol. 3, no. 5, pp. 1039–1049, 2018.