Optimal myelin elongation relies on YAP activation by axonal growth and inhibition by Crb3/Hippo pathway

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Fast nerve conduction relies on successive myelin segments that electrically isolate axons. Segment geometry—diameter and length—is critical for the optimization of nerve conduction and the molecular mechanisms allowing this optimized geometry are partially known. We show here that peripheral myelin elongation is dynamically regulated by stimulation of YAP (Yes-associated protein) transcription cofactor activity during axonal elongation and limited by inhibition of YAP activity via the Hippo pathway. YAP promotes myelin and non-myelin genes transcription while the polarity protein Crb3, localized at the tips of the myelin sheath, activates the Hippo pathway to temper YAP activity, therefore allowing for optimal myelin growth. Dystrophic Dy²⁄² mice mimicking human peripheral neuropathy with reduced internodal lengths have decreased nuclear YAP which, when corrected, leads to longer internodes. These data show a novel mechanism controlling myelin growth and nerve conduction, and provide a molecular ground for disease with short myelin segments.
Fast nerve conduction relies on successive myelin segments that electrically isolate axons between nodes of Ranvier. The myelin geometry—diameter and internodal length—is therefore a critical parameter of the nerve conduction velocity and changes in this geometry underlie numerous functional properties of the brain and nerves. Moreover, reduced internodal length, which strongly impairs nerve conduction velocity, is linked to human neuropathies such as multiple sclerosis in the central nervous system and CMT4F (Perxin gene mutations) and congenital neuro-muscular dystrophy 1A (z2 laminin gene mutations) in the peripheral nervous system. While the molecular mechanisms responsible for the homogeneity of myelin thickness in Schwann cells along the same axon have been uncovered, the mechanisms responsible for myelin elongation and the formation of regular myelin lengths remain unknown. Using the remarkable homogeneity of myelin internodes along axons of the peripheral nervous system, we investigated these mechanisms in vivo and in vitro.

Recently, the Hippo pathway and YAP (Yes-associated protein) transcription cofactor have been shown to be perturbed in Schwannomas, suggesting that they play an unknown role in Schwann cells. Here we show that myelin elongation in Schwann cells is dynamically regulated by the stimulation of YAP activity during the nerve elongation that occurs with postnatal body growth. This elongation is finely tuned as YAP activity is downstream of Dystrophin D2/25, mice that mimic a human peripheral neuropathy with reduced internodal lengths. We found that the shortened myelinating Schwann cells (mSCs) have depleted nuclear YAP which when corrected, by lifting the Crb3/Hippo inhibition, leads to longer internodes. We conclude that the Crb3/Hippo/YAP pathway is a major regulator of myelination in peripheral nerves and we propose that defects in YAP activation underlie peripheral neuropathies with reduced internodal length. The modulation of this pathway can be used to correct myelination and myelin geometry.

Results
Crb3 negatively regulates myelin sheath elongation. We found that the apical polarity protein Crb3, one of the mammalian homologues of drosophila Crumbs, is expressed in the peripheral nerve and specifically distributes to the microvilli of mSCs. These microvilli are fine cellular protrusions that originate from the outer membrane of the Schwann cell, mix with microvilli of neighbouring cells and contact the node of Ranvier. The relevance of YAP was confirmed in dystrophic D2/25 mice that mimic a human peripheral neuropathy with reduced internodal lengths. We found that the shortened myelinating Schwann cells (mSCs) have depleted nuclear YAP which when corrected, by lifting the Crb3/Hippo inhibition, leads to longer internodes. We conclude that the Crb3/Hippo/YAP pathway is a major regulator of myelination in peripheral nerves and we propose that defects in YAP activation underlie peripheral neuropathies with reduced internodal length. The modulation of this pathway can be used to correct myelination and myelin geometry.

Hippo pathway and YAP mediate Crb3 effects. Crb3 can activate the Hippo pathway, via Merlin and/or Willin (also termed FRMD6) to prevent the shuttling of YAP transcription cofactor to the nucleus thereby inhibiting tissue growth due to nuclear YAP activity. We found that, in addition to Merlin, Willin, Mst1/2 and Lats1/2, and YAP were also expressed in the myelinated mouse peripheral nerve. We then examined the Merlin localized to Cajal's bands, while Willin localized to the Schwann cell microvilli, showing that Willin but not Merlin is the target of Crb3 in mSC microvilli. In contrast YAP was mostly nuclear, while endogenous levels of Mst1/2 and Lats1/2 were too low to be detected by immunohistochemistry. As Willin is known to activate the Hippo pathway, we therefore silenced Willin in Schwann cells in vivo using the viral approach described previously. This silencing increased the myelin internodal length at two months compared with control shRNA (Fig. 2b), showing that Willin and the Hippo pathway participates in the control of myelin elongation. It is worth noting that Willin silencing affected more significantly thin fibres versus large fibres at the opposite to Crb3 (Supplementary Fig. 8), suggesting that additional molecules are involved. We then examined the consequence of silencing the final target of this pathway, YAP. Without YAP (Supplementary Fig. 7c,e), internodal length remained shorter than controls (Fig. 2c) and myelinated fibre diameter was also significantly thinner (6.6 ± 0.23 versus 7.4 ± 0.27 μm; P < 0.05). In addition YAP silencing, such as Crb3 and Willin silencing, did not affect SC proliferation.
Supplementary Fig. 7f) suggesting that silenced cells are not more abundant and the reduced internodal length is not due to competition for axonal space. So we conclude that YAP is required for optimal myelin length and diameter in mSC in vivo.

To determine if Crb3 indeed controls YAP activity, we measured the amount of nuclear YAP in Crb3-silenced cells and found that these cells had more nuclear YAP (Fig. 2d,e) than neighbouring non-silenced cells. In addition, using co-infection with both Crb3 shRNA and YAP shRNA adenoviruses, we reduced YAP expression in the over-elongating Crb3-silenced cells. In this condition, mSCs over-elongated but significantly less than when Crb3 alone was silenced (Fig. 2f,g), showing that Crb3 required YAP to mediate to a full extent its effect on myelination. We conclude that Crb3 controls myelin sheath elongation via the Hippo pathway that negatively regulates YAP.

YAP promotes myelin and non-myelin genes transcription.

YAP is a transcription cofactor that regulates gene expression through its binding to transcription factors including TEA domain factors, TEAD21,22. To understand how YAP regulates Schwann cell myelination we first investigated whether dominant active YAP S127A changes myelin protein expression in rat primary Schwann cells (RSCs) differentiated in vitro. Indeed early growth response protein 2 (EGR2), the key transcription factor required to promote myelination, and myelin-associated glycoprotein (MAG) were upregulated after YAP overactivation (Fig. 3a,b), while myelin protein zero (MPZ) was downregulated and myelin basic protein (MBP) remained undetectable. In addition, we found that YAP overactivation leads to the upregulation of Rab11 and Laminin γ 1 expression (Fig. 3a,b), two proteins not specific to myelin but required for the myelination process23,24.

We then investigated if YAP can directly control myelin gene transcription at the promoter level focusing on MPZ, the main peripheral myelin protein, and EGR2, a key myelin gene regulator that controls MPZ expression25. Cells with either Mpz or Egr2 promoters driving the expression of luciferase were transfected

### Figure 1 | Crb3 negatively regulates myelin sheath elongation.

(a) Crb3 immunostaining (red) localizes as discrete lines typical of microvilli at the extremities of mSC (GFP, green). (b) Crb3 (red) partially colocalizes with microvilli marker moesin (green) in a typical band pattern. (c) Confocal images Z axis projection of Crb3 immunostaining (red) reveals a typical microvilli ring around axonal node marker Ankyrin G (green). (d) Representative mSCs infected with Crb3 (Crb3 sh, DsRed2) or control shRNAs (Cont sh, GFP) lentiviruses. (e) Lengths versus diameters of mSCs infected with Crb3 (black square, n = 83, slope line 2) or control (white circle, n = 60, slope line 1) shRNA viruses. Two independent shRNAs against Crb3 yielded similar results (results from only one is shown). Three animals for control and Crb3 sh virus. (f) Average length (left) and diameter (right) of mSCs described in e. (g) mSC infected with Crb3 shRNA virus (Crb3sh, DsRed2) is longer than the next non-infected mSC (NI) on the same axon. Nodal regions (arrows) are visualized using Kv1.2 immunostaining (green) and nuclei using DAPI (blue). Statistical two-tailed Student’s t-tests: ***P<0.001; NS, P>0.05 not significant. Scale bars, 100 μm (a,d,g), 5 μm (b,c). Error bars show s.e.m.
with control GFP or different YAP and TEAD constructs and the impact of these factors on luciferase expression was measured. YAP alone was able to stimulate the Egr2 promoter but not the Mpz promoter (Fig. 3c). However, when cells co-expressed YAP and TEAD1, the Mpz promoter was strongly stimulated and the Egr2 promoter activity was maximal. Co-expressing YAP with another TEAD family member, TEAD4, also stimulated the Egr2 promoter but not Mpz (Fig. 3c), suggesting that effects of YAP on the Egr2 and Mpz promoters partially rely on TEAD isoforms. We verified that TEAD isoforms are indeed expressed in the mouse sciatic nerve (Supplementary Fig. 9). Taken together these results indicate that effects of YAP on the Egr2 and Mpz promoters partially rely on TEAD isoforms.

Myelin elongation slows down when YAP is phosphorylated. The rate of myelin sheath growth is not uniform during peripheral nerve myelination3,26,27, so obtaining a regular myelin sheath along a single axon requires a finely tuned developmental regulation. We therefore investigated whether Crb3 and YAP could participate in this process. Firstly, the effect of Crb3 silencing was analysed at different ages. At 15 dPN a small but significant change could be detected versus control shRNA (Fig. 4a). By 30 dPN the internodal length had more than doubled in Crb3-silenced cells (Fig. 4a), with a growth rate three times higher than in control cells (19.7 versus 6.0 μm per day). By 60 dPN, while the internodal length of Crb3-silenced cells remained higher than controls cells (Fig. 4a), both growth rates decreased to similar values (3 versus 2.8 μm per day). These data indicate that Crb3 silencing increases myelin elongation by boosting the growth rate without altering the timeframe of the elongation. As Crb3 is still significantly expressed in adult mouse nerve (Fig. 1a; Supplementary Fig. 1a), this suggested that Crb3 inhibition of myelin elongation is not active at adult age. Consistently the expression of Crb3 target, YAP, in the nucleus of...
mSCs steadily decreased between 15 dPN and 60 dPN (Fig. 4b,c, left axis), paralleling the decrease of the mean internodal extension rate (Fig. 4c, right axis). This suggested that nuclear YAP amount is the limiting factor for myelin elongation. We then investigated the total levels of YAP and phosphorylated YAP, the inactive form generated by the Crb3/Hippo pathway, in sciatic nerve by western blot. We found that while total YAP levels remained constant between 15 and 60 dPN, the level of phospho-YAP strongly increased after 30 dPN (Fig. 4d,e), indicating that the Crb3/Hippo pathway controls the amount of active nuclear YAP during myelination. Taken together our data confirms that the Crb3/Hippo pathway controls the amount of active nuclear YAP during myelination.

Nerve stretching triggers myelin elongation via nuclear YAP.

We next examined what initially drives YAP activation and localization to the nucleus. As internodal length strongly correlates with nerve and limb elongation27 we hypothesized that axonal elongation, which occurs by stretching with postnatal body growth in vivo28, is required to drive YAP nuclear localization and myelin elongation. To test this hypothesis we used a femoral distraction technique to mechanically elongate the sciatic nerve of adult mice (Fig. 5a). This technique enabled nerve extension by ~30% over 15 days in living mice (Fig. 5b), without damaging axons29. As previously observed in rat and rabbit29,30, nerve stretching in adults steadily stimulated myelin sheath elongation (Fig. 5c,d, two first columns). This correlated with increased YAP expression in the nucleus of myelinating cells (Fig. 5e,f). When cells were silenced for YAP myelin did not elongate following nerve extension (Fig. 5d, two middle columns). In addition we observed that these YAP-silenced cells had thinner extremities and showed a heterogeneous structure (Supplementary Fig. 10b,c). This suggested that the loss of YAP under elongation conditions prevents the mSC from ‘keeping up’ with the imposed artificial extension. While Crb3 loss has no effect on the internodal length of mature mSCs (Supplementary Fig. 10a), the re-engagement of the Crb3/Hippo/YAP pathway in artificially extended adult nerves, allowed Crb3 silencing to increase nuclear YAP (Fig. 5f,g) and myelin elongation (Fig. 5c,d, two last columns) at even higher levels than in control cells of similarly extended nerves. This indicated that Crb3 can still play an inhibitory role on myelin elongation in adult nerves when axonal stretching stimulates YAP nuclear activity. We conclude that myelin elongation during development is initiated by the postnatal limb elongation and axonal stretching that recruits YAP to mSC nucleus; it is then negatively controlled by Crb3, via the Hippo pathway, to allow harmonious growth of all myelin segments along the axon (Fig. 6).

Crb3-Hippo dissociates longitudinal from radial growth.

Axonal neuregulin 1 type III (Nrg1) has been shown to be essential to control myelin growth. Indeed Nrg1 overexpression in axons leads to an abnormal thickening of the myelin sheath.
due to the overactivation of the PI3K-AKT pathway\(^1^2\), but no change in the internodal length\(^3\). On the opposite Crb3 silencing affected myelin length but not diameter and YAP silencing affected both parameters. To understand the interplay between Crb3-Hippo-YAP and Nrg1-PI3K-AKT pathways we first expressed constitutively active YAP in differentiated SC in presence of soluble Nrg1. YAP reduced AKT expression but strongly promoted its phosphorylation (Fig. 7a,b), indicating that YAP promotes AKT activity in SC. We then collected sciatic nerves of transgenic mice overexpressing Nrg1 in neurons\(^1\) and compared YAP and phospho-YAP expressions with littermate controls. Mutant mice expressed more YAP but also more phospho-YAP (Fig. 7c,d). As Crb3 expression did not change (Fig. 7e,f), this suggested that the Crb3-Hippo pathway was efficiently inactivating the excess of YAP. As a result, active nuclear YAP increased only slightly in SC of transgenic mice (Fig. 7g,h). Finally, internodal length increased also in mutant mice but this increase remained small and nearly non-significant (Fig. 7i; Student \(t\)-test \(P\) value \(= 0.035\)). Taken together these data indicates that axonal Nrg1 signalling upregulates YAP but this increase does not affect much the internodal length because YAP is largely inactivated by the Crb3-Hippo pathway.

**Lack of nuclear YAP leads to shorter myelin in Dy\(^2j/2j\) mice.** Reduced internodal length strongly impairs nerve conduction velocity\(^2\),\(^4\) and some human peripheral neuropathies have been
shown to be linked to reduced internodal length such as CMT4F (Periaxin gene mutations)6–8, and congenital muscular dystrophy 1A (α2 laminin gene mutations)9. To check whether the Crb3/Hippo/YAP mechanism is involved in these diseases we investigated a naturally occurring mouse line, Dy2j/2j, mutated on α2 laminin gene that displays a peripheral neuropathy with reduced internodal length 32 (Fig. 8a) mimicking the human congenital neuro-muscular dystrophy 1A (ref. 9). We found that mutant mice sciatic nerves had less YAP (Fig. 8b) than control littermates but no change in phospho-YAP or Crb3 expression (Fig. 8c; Supplementary Fig. 11). Moreover mutant mSCs had significantly less nuclear YAP (Fig. 8d,e). Taken together this suggested that decreased nuclear YAP may be responsible for the reduced internodal length. To confirm this we sought to restore nuclear YAP level in mSCs of Dy2j/2j mice. Because YAP overexpression was deleterious and lifting the Hippo pathway inhibition on YAP via Crb3 silencing increased nuclear YAP amount in wild-type mice (Fig. 2d,e), we silenced Crb3 in mutant mSCs. As expected this increased the amount of nuclear YAP (Fig. 8f,g) and accordingly increased the internodal length of mutant cells (Fig. 8h,i), confirming that the lack of nuclear YAP contributes to the peripheral nerve defect in these mice. Our data

Figure 5 | Nuclear expression of YAP is regulated by nerve elongation (a) Schematic of the technique we used to mechanically distract the adult (11 to 13 weeks old) mouse femur and to artificially extend the sciatic nerve (blue) that is attached to the spinal cord (device not to scale, see Online Methods for more details). (b) An entire extended sciatic nerve (E) and the non-extended contralateral nerve (NE) of the same mouse are shown after femoral distraction using reflected white light. The extended sciatic nerve is 30% longer than the contralateral nerve. (c) Representative mSCs infected with control (Cont sh) or Crb3 (Crb3 sh) shRNA virus in an extended (E) or a non-extended nerve (NE). (d) Quantification of internodal lengths of cells infected with control, YAP shRNA or Crb3 shRNA viruses in extended (E) or non-extended nerves (NE). Between 106 and 171 cells (from three animals) were measured for each condition. (e) Nuclear YAP (red) immunostaining in mSC nuclei (arrows) is higher in artificially extended nerves (E, right panel) than in non-extended nerves (NE, left panel). (f) Quantification of YAP immunostaining in nuclei of mSCs infected with control or Crb3 shRNA viruses in extended (E) or non-extended nerves (NE). Nuclei quantified: 52 (NE Cont sh), 131 (E Cont sh), 19 (E Crb3 sh). Three animals for each condition. (g) In an extended nerve, the nucleus (arrows) of a cell expressing Crb3 shRNA (red) has more YAP (green) than the nucleus (DAPI, blue) of a nearby non-infected cell (NI). a.u., arbitrary unit. Statistical two-tailed Student’s t-tests: *P<0.05; **P<0.01; ***P<0.001. Scale bars, 1 mm (b), 100 μm (c), 20 μm (e,g). Error bars show s.e.m.
Figure 6 | Schematic illustrating the model of myelin elongation in mSC. Top: A myelinating Schwann cell is exposed to input (green arrow) from the stretching axon in the elongating nerve which stimulates the translocation of YAP to the nucleus (green dotted line). YAP nuclear activity promotes myelin gene expression (blue arrow), which increases myelination and myelin elongation (blue dotted arrows). YAP nuclear translocation is negatively regulated by Crb3/Hippo pathway signalling originating from the cells extremities (long red arrow) which phosphorylates YAP, sequestering it in the cytoplasm which consequently limits myelin elongation (short red line). Bottom: the combination of signals promoting YAP activity (green arrows) and inhibiting it (red lines) allows the harmonious growth (blue dotted lines) of all myelin segments along the axon.

Discussion

We found that myelin elongation in mSCs is driven by the stimulation of YAP transcription cofactor activity during the axonal elongation of postnatal body growth. The localization of YAP in nucleus, and therefore its ability to promote myelin gene transcription, is regulated by Crb3 and Willin, upstream controllers of the Hippo pathway, as evidenced by the accumulation of phospho-YAP coincident with decreased nuclear YAP at 30–60 dPN during the slowing of myelin extension. Thus YAP nuclear activity is stimulated by nerve elongation during body growth and conversely tempered by Crb3 signalling originating at the longitudinal extremities of the mSC. Taking these opposing forces into account we propose a model that accounts for optimal and homogeneous longitudinal growth of adjacent myelin segments (Fig. 6).

The Nrg1-Pi3K-AKT pathway has been shown to be critical during myelination and in particular to control the myelin geometry10–12. However manipulations of this pathway revealed that other mechanisms were likely to be involved in the promotion of myelination in Schwann cells and in oligodendrocytes, the myelinating glia of the central nervous system33,34. The YAP pathway may represent one of these supplementary mechanisms. Indeed YAP silencing reduced myelination and increasing YAP activity via Crb3 silencing increased longitudinal myelination.

How YAP, a co-transcription factor, stimulates myelination is not completely clear but our data gives some indications. First, YAP activation in cultured SC positively (EGR2, MAG) or negatively (MPZ) regulates myelin protein expression and it controls myelin gene transcription at the promoter level. This is nevertheless not limited to myelin genes as YAP also promotes expression of non-myelin proteins that are critical for myelination such as Rab11 or Laminin γ1. However YAP does not act alone and our data suggest that its functional interactions with TEAD transcription factors to promote myelin gene transcription are complex. So, additional studies will be required to fully understand the role of YAP on gene transcription in mSC.

Second, YAP can promote myelination via the Nrg1-Pi3K-AKT pathway. Indeed constitutively active AKT strongly increases AKT activation in differentiated SC in culture in presence of Nrg1. It has been reported that YAP promote the expression of the catalytic subunit of Pi3K, p110γ, which activates AKT35, but whether this is occurring in mSC remains to be shown. In any case Nrg1 and YAP pathways strongly interact in mSC. Indeed we found that Nrg1 overexpression in neurons, which increases myelination, strongly stimulates YAP expression. How this occurs is unclear but this is consistent with our finding that nuclear YAP expression is promoted by axonal clues during development.

The most intriguing part is the dissociation between the longitudinal and the radial growth of myelin. Indeed while during development myelin diameter and length increases harmoniously and are proportionate to the axon diameter, it has been shown that internodal length is actually more significantly related to the axon length27. This dissociation is reflected by human peripheral nerve diseases as some of them show short myelin segment with a correct diameter9. In addition, overexpression of axonal Nrg1 (refs 11,31) and activation of the Pi3K-AKT pathway10 in mSC increase radial but not longitudinal myelin growth. On the reverse Crb3 silencing and artificial axonal elongation promote only longitudinal myelin growth. Using mice overexpressing axonal Nrg1, we found that while YAP is upregulated the Crb3-Hippo pathway phosphorylates and inactivates it. So nuclear YAP does not increase much and myelin longitudinal growth remains close to normal, while the upregulation of the Pi3K-AKT pathway increases radial growth. This suggests that the dissociation results from the inhibition of YAP activity via Crb3 and the Hippo pathway.

It has been reported that AKT can phosphorylate and inactivate YAP36, which could also explain why the majority of YAP is phosphorylated in Nrg1 overexpressing mutant. However if AKT was a major regulator of YAP phosphorylation then nuclear YAP should be much lower in growing pups because at that age AKT activity is high10. In addition when we silenced AKT1 or AKT2 in mSC we obtained shorter myelin10, which is inconsistent with lower YAP phosphorylation. So we believe that AKT is not likely to be the main regulator of YAP activity in mSC and this role is essentially played by the Crb3-Hippo pathway.

It remains unclear how Crb3-silenced mSCs can elongate their myelin sheath without making it larger, while during the same developmental timeframe Dlg1-silenced cells enlarge it without making it longer10. This suggests that, beyond the production of myelin and interactions between YAP and Nrg1 pathways, Crb3 and YAP are involved in mechanisms that control the longitudinal deposition of myelin. We propose that multiple local points of myelin synthesis exist along the myelin sheath, most likely at the outside edge of incisures and of paranodal loops (Supplementary Fig. 2e), with local enrichment in phospho-AKT and eventually mTOR complexes. Crb3 and YAP could promote the formation of these myelinosynthesis points and therefore promote myelin elongation. In a second mechanism YAP could also promote the trafficking along the myelin sheath and therefore favour the longitudinal deposition of myelin. We envision these two mechanisms may co-exist and speculate that while radial myelination could be due to more myelin production
only, longitudinal myelination may be due to both more myelin production and more myelinogenenic points along the internode. It would be interesting to examine the consequence of relieving Crb3 inhibition while increasing myelination via the Nrg1-AKT pathway. In this case the excess of myelin might be deposited longitudinally instead of making the myelin thicker. Finally it can be noted that both longitudinal and radial myelination pathways are finely regulated by negative signalling coming from the stabilization of Dlg1 and PTEN proteins\(^\text{10}\) and from the Crb3/Hippo pathway (this study), respectively. This combination of positive and negative conflicting regulations may be the golden key for the optimization of the myelin sheath geometry in PNS and CNS.

Our data suggest that Crb3 act as a sensor in the microvilli for signals coming from neighbouring cells or from the axonal node of Ranvier (Supplementary Fig. 2e). Activated by these signals Crb3 could activate Willin and the Hippo pathway, phosphorylate and prevent YAP access to the nucleus and thereby inhibit

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**Figure 7** Crb3-Hippo pathway dissociates longitudinal from radial myelin growth. (a) Western blots showing expression of AKT and active phosphorylated AKT (P-AKT) in primary rat Schwann cells differentiated in culture with dibutyryl-cyclic AMP (+db) following infection with GFP control or constitutively active YAP S127A Flag adenovirus. GFP and YAPS127A panels confirm infection in three independent cultures of each condition. Actin is used as loading control. (b) Ratio active P-AKT on AKT is calculated from the quantifications of the western blots shown in a and after normalization over actin. Three independent experiments. NS: not significant. (c) Representative western blots showing expression of YAP and phosphorylated YAP (P-YAP) in sciatic nerves of transgenic Nrg1 mice (Mutant) and of control littermate. Actin is used as loading control. (d) Quantifications of western blots showing YAP and phosphorylated YAP levels in mutant mice (Mut, 5 mice) relative to control mice (Cont, 4 mice) normalized over actin levels. (e) Representative western blots showing expression of Crb3 in sciatic nerves of transgenic Nrg1 mice (Mutant) and of control littermate. Actin is used as loading control. (f) Quantifications of western blots showing Crb3 levels in mutant mice (Mut, 5 mice) relative to control mice (Cont, 4 mice) normalized over actin levels. (g) Representative nuclear YAP immunostaining on sciatic nerve longitudinal sections of transgenic Nrg1 mice (Mut) and littermate control (Cont). Scale bar, 50 μm. (h) Quantification of YAP immunostaining fluorescence intensity in nuclei of mSCs as shown in e. Nuclei quantified: 112 (Control), 270 (Mutant). Three animals for each condition. (i) Quantification of internodal lengths after teasing sciatic nerves of transgenic mice (Mut, n = 57 cells, three animals) and control littermate (Cont, n = 84 cells, three animals). Cells were stained with FluoroMyelin, nuclei with TOPRO3 and nodes immunostained with Kv1.2 antibodies. Mice were 5 months old. a.u., arbitrary units. Statistical two-tailed Student’s t-tests: *P < 0.05; **P < 0.01; NS: not significant. Error bars show s.d for b, d and f and s.e.m for h and i.
myelination. Crb3 features a short extracellular domain which may interact with the extracellular matrix or with other proteins extracellular domains in cis or in trans. Another possibility could be that Crb3 interacts with its transmembrane domain or its intracellular domain with other transmembrane proteins that will act as the sensor protein for extracellular signals. Willin was first detected in peripheral nerve tissues as an interacting protein for Neurofascin using two-hybrid technique and Neurofascin will act as the sensor protein for extracellular signals. Willin was found to interact with the extracellular matrix or with other proteins that might interact with the extracellular matrix. Another possibility is that Crb3 interacts with its transmembrane domain or extracellular domains in cis or in trans. This interaction might also be involved in the signal transduction. However, the link of these molecules with the cell polarity is less clear. Therefore, additional work is required to discover the complete mechanism that is engaged during the control of longitudinal myelination in peripheral nerves.

We found that in adult nerve relieving Crb3 inhibition on myelin elongation does not lead to longer cells, despite the continued expression of Crb3. To see the inhibitory effect of Crb3 in adult mSCs we have to re-engage axonal extension by...
artificially extending the nerve. This means that Crb3 is not functionally inactivated in adult mice but that its activity is silent because most of the YAP is already phosphorylated and inactive. When we re-engage axonal extension more unphosphorylated YAP is generated so the Crb3 activity can be seen again. This is supported by the strong YAP phosphorylation that occurs in adult transgenic Nrg1 mice as it suggests that Crb3 and the Hippo pathway are still very active in adult mice. This is also consistent with the proposal that Crb3 is activated by cell–cell interaction at the microvilli, because this does not stop at adult age.

A major implication of our data is that nerve extension promotes nuclear YAP localization and stimulates myelin sheath elongation. These data are consistent with empirical observations showing that myelin internodal length correlates with the limb growth during development. Nevertheless, the molecular mechanisms that underlie this process remain unclear. When nerves are extended axons are also stretched and elongate by inserting new material all along their length. This elongation mechanism is different from the axonal growth that occurs during embryonic development in vitro, which is mediated by the axonal growth cone. Our data suggest that the extending axons send a signal to the mSC. This signal might originate from axo-axonal junctions that link glial paranodal loops to paranodes of the node of Ranvier (Supplementary Fig. 2e). As it is lying directly on the axon, one way that Schwann cell has to detect axonal stretching is to anchor both glial extremities on the stretched axon and to use the glial cytoskeleton as a tension sensor. But this may not be the only one.

Indeed physical interactions between the glial abaxonal plasma membrane, its underlying cytoskeleton and the extracellular matrix could also play a critical role in the detection of axonal stretching. The abaxonal membrane (Supplementary Fig. 2e) contains dystroglycans that interact with the laminin of the basal lamina. These dystroglycans interact with Periadin and Uropodhin, which are linked to the cytoskeleton. Mutations in periaxin gene lead to Charcot-Marie-Tooth disease 4F, a peripheral neuropathy with reduced internodal length, as observed in congenital muscular dystrophy patients with mutations in the laminin-2 gene. In addition mouse mutants for dystroglycans and Uropodhin display a reduced internodal length, showing that the interaction between the basal lamina and the glial cytoskeleton is required for myelin elongation. Therefore we suggest that the anchoring of the Schwann cell to the basal lamina via dystroglycans and the related cytoskeleton participates in the detection/sensing of axonal stretching by offering a stable reference to oppose to the extending axon.

Highlighting the role of the Crb3/Hippo/YAP pathway in the emerging picture of myelin formation confirms that this process is far from passive. It is highly controlled in space and time by a combination of positive and negative signalling and by the superposition of different signalling pathways. This growing complexity will give us the opportunity to build a detailed in silico cellular model in which we will be able to identify molecular targets for drug or gene therapies for peripheral neuropathies.

Note added in proof: a recent publication describes the role of YAP in axonal sorting by Schwann cells in the mouse peripheral nerve.

Methods

Mice. Pregnant (E15-16) and adult Swiss mice were purchased from Janvier (France). Adult CB17/SCID mice were purchased from Charles River Laboratories (France) and CB17/SCID pups were obtained from breeding in our animal facility. Sciatic nerves of transgenic Nrg1 (Thyl:2-Nrg1 type III; 5 months old) mice and control littermate were a kind gift of R. Feldrich and R. Stassart (Department of Neurogenetics, Max Planck Institute of Experimental Medicine, Göttingen, Germany). Heterozygous Lama2 Dyl+/- mice were purchased from The Jackson Laboratory (USA) and used to generate Dyl+/- and Dyl+/+ mice for experiments. As a point mutation in the Dyl allele contains a unique Nde1 restriction site, we used a genotyping strategy to PCR across the point mutation followed by Nde1 restriction digestion of PCR products as previously described. Briefly DNA was extracted from ear biopsies and PCR performed with Phusion HF Taq polymerase (New England Biolabs, France) using the following sense (S) and antisense (AS) primers: S: 5'-TCC TGC TGT GAA TCT TG-3; AS: 5'-CTC TAT TGC TGA ACT TTG GAT-3. PCR products were digested overnight with Nde1 (New England Biolabs, France) to produce an uncut 377 bp product in wild type and digested 164- and 109-bp fragments from Dyl+/+ mice. Heterozygous mice were identified by the presence of all three bands.

Animal surgery and injection. Animal use was approved by the Comité Régional d’Ethique du Languedoc –Roussillon, France and the veterinary office of the Canton of Zurich, Switzerland. Injection of viral constructs in pup and adult mice were performed as described previously. Briefly, pups (as indicated) or postnatal day 30 CB17/SCID mice were anesthetized with isoflurane inhalation and placed under a Zeiss stereomicroscope (Stemi 2000, Carl Zeiss Microscopy GmbH). The glutaeus superficialis and biceps femoris muscles were separated to reveal the cavity traversed by the sciatic nerve. A thin glass needle filled with coloured viral solution (3 μl for pups or 8 μl for adult injections) was introduced into the nerve with a micromanipulator (IM—3C, Nariushi Japan Group). This solution was injected over 10 min with short pressure pulses using a microinjector (Pneumatic Picup PVM820, World Precision Instruments) coupled to a 3 MHz function pulse generator (FG8215, Langlois). The location of the injection site could be filled. The nerve was replaced in the cavity, the muscles were readjusted, and the wound was closed with Histoacryl glue (Bbraun, Germany). When fully awake, pups were put back to the mother and the litter treated with antibiotics to prevent infection. Sacrifices were done by CO2 inhalation. Lentivirus production and purification. One 15 cm polylysine-coated dishes of 95% confluent HEK 293T cells was transfected with 13.5 μg pSPAX2, 9 μg pCMV2 and 12.5 μg of the lentiviral vector. The lentiviral supernatant was harvested 48 and 72 h later, pooled, centrifuged at 1,000 x g and filtered. The supernatant was centrifuged using the SW28 rotor, 21,000 r.p.m. at 11 °C (Beckmann Coulter) for 2 h. The resulting pellet was re-suspended in 1 ml DMEM 10% FCS with antibiotics and centrifuged again for 1 h at 16,000 r.p.m. and 4 °C using the T60 rotor (Beckmann Coulter). The supernatant obtained was finally precipitated obtained in 40 μl PBS with 0.001% fast green (Sigma), and aliquots stored at –80 °C. Titres were between 1010 and 1011 TU ml–1. These viral preparations were used to infect mSCs in vivo.

Adenovirus production and purification. Adenoviruses were produced from the pAdEasy adenoviral vector system by Sirion Biotech (Martinsried, Germany). Titres were between 1011 and 1012 TU ml–1. They were used in both in vivo infection of mSCs and for infection of myelinating co-cultures.

ShRNAs, plasmids and viral vectors construction. The lentiviral pLKO.1 vectors containing shRNA targeting mouse Crb3: 1- #TRCN0000124547, 5’TTGGGCGTTGGTGTGCGGAT; 2- #TRCN0000124548, 5’CCCTTAACGCCGGCCTT; 3- #TRCN0000124545, 5’CGGACCCCTTCTCACAATAGCA; 4- #TRCN0000124544, 5’CCAACAAACCTCCTGCTCCCTT; are from SIGMA. These vectors express puruymycin and were used for low titre lentiviral production and shRNA validation in mouse EpI4-J3B1A cell line. The Crb3 shRNA sequences 3 and 4 described above, the validated mouse shRNA sequences for YAP (shRNA1 5’ CGG TGG TAA CAA GAA GGA ATT; shRNA2 5’ GCC CAA AAT CAG TAG TAA AAG GA) and Willin (shRNA1 5’GAC TCG AAAGAT GAG GAA AAT; shRNA2 5’GAC TCTT ACT CAG AGG AAT) were cloned into the lentiviral pSICOR (Addgene 11579) vector using Hpal-Xhol sites. Titres were used for high titre lentiviral production. In addition Crb3 shRNA 3 and 4, and YAP shRNA 1 sequences were cloned in the adenoaviral pAd5 shRNA plasmid described as previously. Lentiviral and adenoviral vectors gave similar results. Crb3 shRNA sequences 3 and 4 were also cloned using the restriction sites described above in pSICOR vector where GFP was replaced with PLAP as described previously.

Control shRNAs were directed against DsRed2 (5’AGTCCGATCAAGGCT CAA) or GFP (5’CAAGTGGTGAGGATTGTC) depending on the fluorescent reporter used (pSICOR expresses GFP while the pAd5 shRNA expressed DsRed). Both control shRNAs gave similar results and the data was on myelin elongation between Figs 2a and 2c). As both Crb3 shRNAs (3 and 4), Willin shRNAs (1 and 2), or YAP shRNAs (1 and 2) give similar effects on myelin elongation between Figs 1a and 1b (Figs 1c and 1b), we used Crb3 shRNA5, Willin shRNA1 and YAP shRNA1 in the following experiments.
Electron microscopy and PLAP staining. Nerves infected with adenoviral vectors expressing Crb3 shRNA 3 or 4 and PLAP were fixed and processed for electron microscopy as described previously.40 Samples were analysed with a H8125 Hitachi H 7100 transmission electron microscope at the Centre Regional d’Imagerie Cellulaire, Montpellier, France. Statistical analysis was done with four samples of nerves infected with each virus. For each cell showing a PLAP staining and for 2–4 non-labelled surrounding cells, axonal and myelin perimeters were measured using ImageJ software to obtain their diameters. From these values, the g-ratio of each cell (axon diameter/myelin diameter) was calculated.

Teased fibres preparation and staining. Teasing of sciatric nerves has been published previously. Briefly, sciatric nerves were isolated from injected mice, fixed for 10 min in Zamboni’s fixative, washed in PBS and subsequently teased on glass slides. After to obtain a single neuron, the nerves were dried overnight at room temperature and immunostaining was performed. For immunostaining, the teased fibres were first incubated for 1 h at room temperature in blocking solution (5% goat serum, 0.3% Triton X-100 and 0.01% sodium azide in PBS), then incubated with primary antibodies in blocking solution overnight at 4 °C. Slides were washed in PBS and incubated for 1 h at room temperature with secondary antibodies diluted in the blocking solution. Finally, samples were washed in PBS and mounted in ProLong Gold Antifade Reagent with DAPI (Molecular Probes). Images were acquired at room temperature using a 20× objective and processed with Image J and compiled into figures in Photoshop (Adobe).

Antibodies for immunostaining. All primary antibodies used are listed as follows: Rabbit anti-Crb3 (D2, kind gift from Dr A.Lé Bivic, IBDMLE, France, 1:200); Mouse anti-Moesin (610401, BD transduction, 1:100); Mouse anti-AnkG (NB20, Calbiochem, 1:50); Rabbit anti-Gliomedin (KR34, Koch, 1:1000); Mouse anti-KV1.2 (75-008, Antibodies incorporated 1:500); Rabbit anti-pERM (3149, Cell Signaling, 1:100); Rabbit anti-pan-Neurofascin (563, kind gift of Dr G. Goutebroze, Institut du Cellulaire, Montpellier, France, 1:1000); Rabbit anti-Gloiadin (AP672-4.1, kind gift of SR Levinson, university of Colorado Health Center, USA, 1:200); Rabbit anti-Caspr/paranadin (LS1, kind gift from Dr G. Goutebroze, Institut du Cellulaire, Montpellier, France, 1:1000); Mouse anti-MBP (SMI94, Sternberger Monoclonal incorporated, 1:10,000); Rabbit anti-Wilxin (F3998-1, kind gift from M. Takeichi, RIKEN Center for Developmental Biology, Japan, 1:200); Rabbit anti-YAP (4912, Cell Signaling, 1:20000); Secondary antibodies for teased fibres immunostaining were used at 1:1000 dilutions. To visualize rabbit primary antibodies: Alexa Fluor donkey anti rabbit IgGs (H+L) 488, 594 or 647 (A21206, A21207, A31573 respectively, Molecular Probes) was used. To visualize mouse primary antibodies: Alexa Fluor F(ab’)2 fragments of goat anti mouse IgG (H+L) 488, 594 or 647 (A11017, A11020, A21237 respectively, Molecular Probes) was used.

Quantification of cell length. For quantification of the length and width of infected cells, nerves were first fixed and cryo-protected with successive glycerol baths. Nerves were then dissociated carefully to spread nerve fibres and allow clear visualization of the intact infected cells. These nerve spreads were cover-slipped in glycerol and imaged with confocal microscopy. This technique allowed optimal visualization of the intact infected cell while avoiding potential deformation that can result from teasing individual nerve fibres. The length and the diameter (mean value of 5–10 measures along the cell) of infected myelinating Schwann cells were measured according to the DsRed2 or GFP expression using ImageJ software. Demyelinating, non-myelinating or incomplete cells were not measured.

Quantification of nuclear YAP immunostaining. Teased fibres immunostained for YAP were co-stained with nuclear DAPI or TOPRO3 (Thermo Fisher, France) due to clearly define nuclei and to allow quantification of nuclear YAP immunostaining. Confocal Z-series of nuclear YAP immunostaining, DAPI/TOPRO3, and DsRed2 or GFP when cells were infected, were obtained using a Leica SP5 and a 40× objective, ensuring no saturation of signal to allow subsequent co-staining. Stacked images were acquired in Z-stacks, the colour channels split and the scale set according to the embedded scale bar. The image containing DAPI was threshold corrected to clearly identify the full nucleus and a mask of all nuclei was generated. Using the ImageJ image calculator, the YAP staining channel AND the DAPI/nuclei mask were superposed, generating an image appropriate for the quantification YAP staining that was within the nuclear area. All the nuclear-restricted YAP immunostaining was outlined (ROI) and the raw integrated densities and areas measured for each nucleus. The raw integrated densities were then normalized to the ROI areas to obtain an integrated YAP nuclear fluorescence density per µm² for each nucleus. Using the channel containing DsRed2 or GFP, indications were then attributed to the quantified YAP nuclear intensity to infected or non-infected cells on the same image. The presence of infected cells and non-infected cells nuclei on the same image ensured minimal variability in the quantification following immunochemistry. Quantifications were done with at least three infected nerves and at least three independent immunohistochemistry staining experiments. All the analyses was conducted with relevant controls (no primary antibody included, no secondary included, control samples).

Western blot. Sciatric nerves were dissected from mice (C57 B6/J, Janvier) at specified developmental ages. After removal of the epineurium and perineurium, the nerves were homogenized with a chilled mortar and pestle in standard RIPA lysis buffer. For protein extracts from whole sciatric nerves or RSCs, the protein concentration was measured using a BCA protein assay kit (Pierce). Between 10 and 25 µg proteins were directly analysed by western blot. Samples were processed using standard SDS–PAGE and western blotting protocols. Blots were digitized and analysed by densitometry with Quantity One software (BioRad). Images in Figs 3a, 4a and 7a-c,e have been cropped for presentation and full size images are presented in Supplementary Figs 16–19 respectively. Images in Supplementary Figs 1, 3, 10, 11 have been cropped for presentation and full size images are presented in Supplementary Figs 16–19 respectively.

Luciferase gene reporter assay. HEK 293T cells (27 × 10⁴) per well were seeded in a 96-well plate in DMEM 10%FBS. Forty-eight hours later, cells were transfected in OptiMEM medium (Invitrogen) with 0.5 µl Lipofectamine 2000 (Invitrogen) and 0.6 µg DNA: 150 ng Renilla luciferase construct (pRL-CMV-Renilla, Promega), 150 ng Firefly luciferase constructs (pGL3 MPZ intron1 Addgene 21260) and pGL3KroD2 promoter Addgene 21260, 150 ng expressing construct 1 (pEGFP Clontech used as control, p2XFlGhAP1, Addgene 17791) and 150 ng of overexpressing construct 2 (pBluescript plasmid as carrier: pRKS-Myc-TEAD1 Addgene 33109; Myc-TEAD4 Addgene 24638). The Lipofectamine and DNA was incubated separately in OptiMEM medium (Invitrogen) for 5 min at 20–25 °C before mixing the two solutions. The mix was incubated for 20 min at 20–25 °C before being added to the cells. After 4–6 h the mix is removed and fresh DMEM 10% FBS is added. Twenty-four hours after transfection, cells were lysed in 20 µl passive lysis buffer (Promega) and assayed for luciferase activity (Firefly and Renilla luciferase) using automated Varioskan Flash (Thermoscientific). Lysates were subjected to two consecutive injections of 75 µl of each different luciferase substrate and values were recorded after an integration time of 8 ms. Luciferase activity was normalized to Renilla activity. Results are expressed as a percentage of the GFP control value. Results shown are from three independent experiments in triplicates.

Bone distraction. Adult SCID mice (Janvier, France) from 11 to 13 weeks old were first injected with 8 µl of adenovirus expressing Crb3 sh3 or YAP sh1 in the sciatic nerve as described above. One week later these animals were anaesthetized again with isoflurane inhalation and the skin shaved and disinfected as described for the viral injection. A skin incision was made following the femur and muscles were also cut along this line to visualize the femur. A microTRACK alveolar drill (KLSMartin) was then used to create a 2 mm diameter hole and the YAP vector was applied in this cavity by injection of the plasmid so only two screw holes remained on each side and slightly bent to conveniently adapt to the bone. The adjusted distraction apparatus was then fitted to the bone and...
fixed using two drill-free auto-blocking screws (diameter 0.9 mm; 5 mm long) placed on the last screw holes of the plate. The bone was then severed between the two screws using a Gigli saw (0.22 mm, RISystem, Davos, Switzerland). The bone was then extended 0.6 mm apart using a special screwdriver (KLS Martin, 51-555-95-07). Each full turn of the screwdriver moved the distractor 0.3 mm. The muscles and the skin were then successively closed using absorbable sutures. Animals were treated with buprenorphine analgesics twice a day for 2 weeks. Every day, at regular hours, animals were anesthetized using isoflurane and the distractor extended a further 0.6 mm. According to published works30,57 the rate of elongation we used did not induce any damage to the nerve and we did not detect any axonal thinning or axonopathy using immunostaining. At the end of the experiment, animals were killed by CO2 inhalation and both elongated and non-elongated contralateral nerves were dissected and fixed in Zamboni’s fixative for 10 min.

This protocol was adapted from previous publications29,30,57. We are grateful for RISystem (Davos, Switzerland) for their very useful online videos: (http://www.risystem.com/Standardized_Implant_for_Research/Training.html).

Statistics. All results are presented as mean ± s.e.m or s.d. All statistical analysis indicates t-test two-tailed P values. Significance: *P<0.05, **P<0.01, ***P<0.001; NS: not significant P>0.05.

Data availability. The authors declare that the data supporting the findings of this study are available within the article and its Supplementary Information file.

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Author contributions
Conceptualization: R.N.F. and N.T.; methodology: R.N.F., L.C. and N.T.; investigation: R.N.F., L.C., C.P.-T., J.B., S.B., J.P. and N.T.; writing—original draft: R.N.F. and N.T.; writing—review and editing: R.N.F., S.G., U.S. and N.T.; funding acquisition: R.N.F., U.S. and N.T. Supervision: N.T.

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