Comparative Chloroplast genomes of Gynura species: Sequence Variation, Genome Rearrangement and Divergence studies

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Abstract

Background

Some Gynura species were reported to be natural anti-diabetic plants. The chloroplast genomes of four Gynura species were sequenced for hybridizations to improve agronomic traits. There are only 4 genera of tribe Senecioneae have published chloroplast genome in Genbank up to now. The internal relationships of the genus Gynura and the relationship of the genus Gynura with other genera in tribe Senecioneae need further researches.

Results

The chloroplast genome of 4 Gynura species were sequenced, assembled and annotated. Comparing with other 12 Senecioneae species, the chloroplast genome features were detailedly analyzed. Subsequently, the differences of the microsatellites and repeats type in the tribe were found. By comparison, the IR expansion and contraction is conserved in the genera Gynura, Dendrosenecio and Ligularia. The region from 25,000 to 50,000 bp is relatively not conservative but the 7 ndh genes in this region are under purifying selection with small change in amino acids. The phylogenetic tree shows two major clades, same as the sequence divergence in region 25,000 to 50,000 bp. Based on the oldest Artemisia pollen fossil, the divergence time were estimated.

Conclusions

Sequencing of chloroplast genome of the 4 Gynura species help us to develop abundant genetic resources. The phylogenetic relationships and divergence time among 4 Gynura and 16 Senecioneae species were sorted out by comparing the chloroplast genomes. The phylogenetic relationship of the genera Gynura and Ligularia is different with former work and further morphology and genome-wide analysis are needed to clarify the genera relationship.
Background

*Gynura* is a genus of flowering plants in the tribe Senecioneae of family Asteraceae endemic to Asia, which contains 44 species in total [1]. Many species of the genus *Gynura* have been reported to have medicinal value to diabetes mellitus, such as *G. procumbens*, *G. divaricata* and *G. medica*. The aqueous extract from *G. procumbens* possessed a significant hypoglycemic effect in streptozotocin-induced diabetic rats [2] and it improved insulin sensitivity and suppressed hepatic gluconeogenesis in C57BL/KsJ-db/db mice [3]. Polysaccharide from *G. divaricata* could alleviate the hyperglycemia by modulating the activities of intestinal disaccharidases in streptozotocin-induced diabetic rats [4] and *G. divaricata*-lyophilized powder was effectively hypoglycemic by activating insulin signaling and improving antioxidant capacity in mice with type 2 diabetes [5]. Phenolic compounds isolated from *G. medica* inhibited yeast α-glucosidase *in vitro* [6].

Some plants in genus *Gynura* have also been used as vegetable and tea in people’s daily life of East and South Asia, thus the genus *Gynura* is a natural treasure trove to treat the increasingly diabetes problem. Although *Gynura* plants are seemingly useful and harmless, but some shortcomings need improvement such as the medicinal effect to diabetes, potential toxicity and oral tastes [7-8]. Large improvement relies on interspecific hybridizations to increase genetic diversity and introgression of valuable traits.

Phylogenetic relationship is useful information for the interspecific hybridizations, but the phylogenetic relationship of the species in genus *Gynura* is, as yet, unclear.

A whole chloroplast DNA ranges between 120 and 160 kb in size on the circular chromosome in most plants, composing of Large Single Copy (LSC), Small Single Copy (SSC), and two copies of an Inverted Repeat (IRa and IRb) [9-10]. Contrast to mitochondrial and nuclear genomes, chloroplast genomes are more conserved in terms of gene content, organization and structure [11]. The chloroplast genomes of angiosperms
generally show slow substitution rates under adaptive evolution [12]. Considering its small size, conserved gene content and simple structure, the chloroplast genome are valid and cost-effective to research phylogenetic relationships and evolution of plants in different taxa. Recently, Forage species of *Urochloa* [13], marine crop *Gracilaria firma* [14], Epilithic sister genera *Oresitrophe* and *Mukdenia* [15], Family *Adoxaceae* and *Caprifoliaceae of Dipsacales* [16] were sequenced the related complete chloroplast genomes to elucidate the diversity, phylogeny and evolution.

In the present study, we sequenced, assembled and annotated the chloroplast genomes of four *Gynura* species. Combined with other published chloroplast genomes of tribe Senecioneae, the structure features, repeat motifs, adaptive selection, phylogenetic relationships and divergence time were analyzed.

**Results And Discussion**

**Chloroplast genome features of 16 Senecioneae species**

In this study, we focused on the chloroplast genome features of tribe Senecioneae, which is the largest tribe of family Asteraceae. Although the tribe comprises about 500 genera and 3000 species [44], we only found that 4 genera of tribe Senecioneae had published chloroplast genome in Genebank. Five species of genus *Dendrosenecio*, one species of genus *Jacobaea*, five species of genus *Ligularia*, one species of genus *Pericallis* and four species of genus *Gynura* were used to find their similarities and differences. The whole sequence length ranges from 150,551 bp (*Dendrosenecio brassiciformis*) to 151,267 bp (*Pericallis hybrida*). With the typical quadripartite parts like most land plants, the chloroplast genome has one Large single copy (LSC), one Short single copy (SSC), two Inverted regions (IRa and IRb) (Fig 1). The LSC length ranges from 82,816 bp (*Jacobaea vulgaris*) to 83,458 bp (*Dendrosenecio cheranganiensis*), the SSC length ranges from
17,749 bp (D. brassiciformis) to 18,331 bp (P. hybrida) and the IRs length both range 24,688 bp (D. brassiciformis) to 24,845 bp (P. hybrida) (Table 1). Changes in each region are not consistent with changes in whole chloroplast genome. J. vulgaris has the shortest chloroplast genome in length but its SSR region is longer than 4 Gynura species. In addition, there are 95 coding genes in the chloroplast genome of P. hybrida and 87 coding genes in the J. vulgaris. GC content varies between 37.2% and 37.5%. Only the rRNA number is conserved in chloroplast genome of tribe Senecioneae, which is same as family Adoxaceae and Caprifoliaceae [16], but different from genus Oresitrophe and Mukdenia [15].

Microsatellites and Repeats type

Number of microsatellites with mono-, di- and tri-nucleotide repeat motifs varies in the tribe. D. brassiciformis, J. vulgaris and L. hogdsonii do not have tri-nucleotide repeat motifs while four Gynura species have 4 to 5 tri-nucleotide repeat motifs. The number of mono-nucleotide repeat motif is 28 to 38, accounting for the largest proportion (Fig 2a). The unit size of microsatellites is significantly different in four Urochloa species [13], which has tetra-nucleotide repeat motif and the tri-nucleotide is the largest in the proportion. The total number of repeats types is consistent with those in the four Gynura species, but the number of each repeat type is different. Palindromic repeats are the most abundant and complement repeats are secondary in 16 Senecioneae species (Fig 2b). Compared with the Oresitrophe species [15], the Senecioneae species have 5 to 12 reverse repeats, but the Oresitrophe species do not have reverse repeats. In addition to this, forward and palindromic repeats number is similar in the Oresitrophe species.

Contraction and Expansion of Inverted Repeats

The chloroplast genome is highly conserved in land plants, but the IR expansion and
contraction leads to the different genome sizes of different plants [41]. 16 species of Senecioneae tribe were analyzed the LSC/IRb/SSC/IRA/LSC border and adjacent genes (Fig 3). *rps19* and *rpl2* gene locate in the LSC/IRb and IRa/LSC border in pairs. In 16 Senecioneae species, the two copies of *rps19* have no change in position to the border and the two copies of *rpl2* are relatively conserved with 1-3 base position changes except the IRa/LSC border of *P. hybrida*. One copy of *ycf1* spans the border of LSC/IRb, another copy is different. The start position is just on the border in the four *Gynura* species, but the others also span the border of IRa/LSC. By comparison, the IR expansion and contraction is conserved in the genera *Gynura, Dendrosenecio* and *Ligularia*.

**Sequence Variation and Adaptive selection**

The whole chloroplast genome sequences of 16 Senecioneae species were aligned by MAFFT program to find the sequence variation. The alignment result was used to calculate the DNA polymorphism by DnaSP program. The base sequence has a Pi value (nucleotide diversity) of 0.2-0.3 at 25,000 bp to 50,000 bp, and other positions below 0.1 (Fig 4A). This shows that this region is less conservative than other regions of chloroplast genome. For further analysis of the results, the chloroplast genome sequences of four *Gynura* species, *D. cheranganiensis*, *L. hodgsonii* and *P. hybrida* were realigned and visualized by mVISTA program. The overall result is consistent with the DNA polymorphism result (Fig S1). The four *Gynura* species are conserved in 25,000 bp to 50,000 bp region and similar with *L. hodgsonii*. In that region, *D. cheranganiensis* is close to *P. hybrida*, but different from the four *Gynura* species and *L. hodgsonii* (Fig 4B). That region has total 12 genes and 7 genes are coding the NAD(P)H dehydrogenase (NDH) complex subunit. The function of NAD(P)H dehydrogenase (NDH) complex is well-known in the photosystem I (PSI) cyclic electron flow (CEF) and chlororespiration [42-43], so the substitution of *ndh* genes was further studied. The ratio of non-synonymous (dN)/synonymous substitution (dS) rate were
calculated by PAML program. The ratio > 1 indicates positive selection, the ratio < 1 indicates purifying selection and the ratio = 1 indicates neutral evolution. All the dN/dS ratio of 7 genes below 1 indicates that they are under purifying selection and little amino acid change happened (Table 2). That means the functions of 7 *ndh* genes are conservative during evolution, although they are not located in a conservative area.

**Phylogenetic Relationships**

The sequence alignment of 16 Senecioneae species was used to construct the ML (Fig 5) and BI (Fig S2) tree. In ML tree, two major clades were constructed with 100% bootstrap value. One clade includes the genera *Gynura* and *Ligularia*, and the other clade includes the genera *Dendrosenecio, Pericallis* and *Jacobaea*. In genus *Gynura*, *G. bicolor* was the first to differentiate, followed by *G. divaricata*, at last were the *G. formosana* and *G. pseudochina*. The two clades divergence result is consistent with the sequence divergence in 25,000 to 50,000 bp region. The former systemic phylogenies of tribe Senecioneae based on ITS region (nuclear) and plastid fragment sequences show a significant different with the phylogenetic tree [44]. In the former phylogenetic tree, genus *Ligularia* belongs to *Tussilagininae* grade, which was in the earlier diverging lineage than other genus. The sequence between four *Gynura* species and five *Ligularia* species is relatively conserved and the Pi value of most sequence locations are below 0.1 (Fig S3), significantly lower than the 16 species alignment. From the perspective of whole chloroplast genomes, genus *Ligularia* is close to genus *Gynura*.

**Divergence Time Estimation**

For the divergence time estimation of the 16 Senecioneae species, *Artemisia gmelinii* and *Chrysanthemum boreale* (Tribe Anthemideae) were selected as outgroup due to oldest *Artemisia* fossil pollen [38-39]. The divergence time of 16 Senecioneae species were
estimated by BEAST2.0 program (Fig 6). The divergence clades of these genera are same with ML tree. The two major clades were expected to differentiate at 37.4 mya (late Eocene). Both Gynura and Ligularia differentiated at 5.8 mya (late Miocene). Dendrosenecio and Pericallis also differentiated at 5.8 mya. The divergence time of tribe Senecioneae and Anthemideae was at 51.39 mya (early Eocene) and the result was consistent with previous study on the evolution and phylogenetic of family Asteroideae basing on the plastid fragment sequences [39]. The traditional view on divergence time of the genus Gynura is in the Old World after the Atlantic opening. In that time, the Senecioioid species were transferred to South America and the divergence began[45]. The divergence time of Gynura species was about 0.3 mya and the result showed that the divergence time of genus Gynura was much earlier than the traditional views. The divergence time of genus Gynura could not start at hundreds or thousands years ago[45] and the divergence time estimated by BEAST program was in the same time period with other genera of land plants[13-16].

Conclusion

This study analyzes the chloroplast genome of four Gynura species used as herbal medicine in parts of Asia. By comparing with other plants in tribe Senecioneae, the repeat motifs, detailed structure feature, phylogenetic relationships and divergence time estimation were identified. The phylogenetic relationship of genera Gynura, Ligularia and others are still in doubt. The tribe Senecioneae contains 155 genera and almost distributes throughout the world [44]. The phylogenetic relationship is difficult to determine by chloroplast fragment or genome. Morphology and genome-wide analysis are needed to further clarify the genera relationship. Determining interspecific relationships and intergeneric relationships will facilitate the hybrid breeding of Gynura species.
Methods

Plant materials, Genome sequencing and assembly

*Gynura bicolor*, cultivated plant, the voucher specimen (510918-1), collected from Nanjing Botanical Garden Mem. Sun Yet-Sen. *Gynura divaricata*, cultivated plant, the voucher specimen (510918-6), collected from Nanjing Botanical Garden Mem. Sun Yet-Sen. *Gynura formosana*, cultivated plant, the voucher specimen (512019-3), collected from Kunming Botanical Garden. *Gynura pseudochina*, wild plant, the voucher specimen (512019-8), collected from Wenshan Zhuang and Miao Autonomous Prefecture, Yunnan Province, all the plants were collected by Prof. Bingru Ren, and the specimens were deposited in the Herbarium of Institute of Botany, Jiangsu Province and Chinese Academy of Sciences.

The *Gynura bicolor, G. divaricata, G. formosana* and *G. pseudochina* plants grow in greenhouse with normal sunlight and temperature. The DNA was extracted from their fresh leaves by CTAB method [17] and DNA degradation and contamination was monitored on 1% agarose gels.

About 1.5μg DNA sample was fragmented by sonication to a size of 350bp, then DNA fragments were end polished, A-tailed, and ligated with the full-length adapter for Illumina sequencing with further PCR amplification. After PCR products purification (AMPure XP system), libraries were analyzed for size distribution by Agilent 2100 Bioanalyzer and quantified by using real-time PCR.

The libraries constructed above were sequenced by Illumina HiSeq X Ten platform and 150bp paired-end reads (PE150) were generated with insert size around 350bp. Quality control (QC) is removing reads with ≥10% unidentified nucleotides (N), > 50% bases having phred quality < 5 and > 10 nt aligned to the adapter, allowing ≤10% mismatches. The perl script NOVOPlasty 2.7.2 [18] was used to assemble the chloroplast genome
sequence with 50 K-mer. The chloroplast genome sequence of *Dendrosenecio cheranganiensis* (Tribe Senecioneae) was selected as the reference genomes. The family Asteraceae plants used in the study were downloaded from Genebank, as follows: *Dendrosenecio brassiciformis* (NC_037960.1), *Dendrosenecio cheranganiensis* (NC_037956.1), *Dendrosenecio johnstonii* (NC_037959.1), *Dendrosenecio kilimanjari* (NC_037957.1), *Dendrosenecio meruensis* (NC_037958.1), *Jacobaea vulgaris* (NC_037957.1), *Ligularia hodgsonii* (NC_039381.1), *Ligularia intermedia* (NC_039382.1), *Ligularia jaluensis* (NC_039383.1), *Ligularia mongolica* (NC_039384.1), *Ligularia veitchiana* (NC_039385.1), *Artemisia gmelinii* (NC_031399.1), *Chrysanthemum boreale* (NC_037388.1).

**Chloroplast genome annotation**

The whole chloroplast genome sequences were annotated by Dual Organellar Genome Annotator [19] and GeSeq [20] with default parameters. Chloroplast genome sequences of tribe Senecioneae plants *Dendrosenecio cheranganiensis* and *Pericallis hybrida* were used as reference sequences. Subsequently, all tRNAs were verified by ARAGORN v1.2.38 [21] and tRNAscan-SE v2.0 [22]. Four chloroplast genomes (*Gynura bicolor*, *G. divaricata*, *G. formosana* and *G. pseudochina*) with annotation were submitted into GenBank and the submission ID is 2222634. Schematic diagram of chloroplast genome with annotations were obtained by OGDRAW [23].

**Repeat structure analysis**

The microsatellite regions is a tract of repetitive DNA in which certain DNA motifs (ranging in length from 1-6 or more base pairs) are repeated, typically 5-50 times [24-25]. The perl script Microsatellite identification tool (MISA, http://pgrc.ipk-gatersleben.de/misa/misa.html) were used to find the microsatellite regions of chloroplast
Considering the features of plant chloroplasts, the numbers of each unit of continuous DNA motifs is set to 1-6, and the minus DNA motifs of each unit is 1-10, 2-6, 3-5, 4-5, 5-5, 6-5. Forward, reverse, complement and palindromic repeats type were detected by online tool REPuter [26]. The Hamming distance was setting as 1 and the minimum repeat size was 30bp.

**Chloroplast genome analysis**

All the chloroplast genome sequences was aligned by MAFFT7.427 [27] on FFT-NS-2 module. Alignments of 7 selected genome sequences were visualized by mVISTA [28]. The DNA polymorphism (nucleotide diversity) was calculated by DnaSPv5 [29] based on alignment results.

The molecular evolutionary rates (\(\omega\)) between orthologous genes were estimated by calculating the ratio of non-synonymous (dN)/synonymous substitution (dS) rate. The coding gene sequences of selected region were extracted by using the Artemis [30]. The gene sequences of each species were aligned by Clustal X [31] with default parameters, and the alignment results (dnd format) were converted to PML format by DAMBE [32] for next analysis. The dN/dS value was calculated by the codeml module (seqtype=1, model=0, Nsites=1,7,8) in PAML4.9i [33]. Significant differences were calculated by Likelihood-ratio test.

**Phylogenetic analysis**

The 16 chloroplast genome sequences of tribe Senecioneae (family Asteraceae) were aligned by MAFFT and the results was used to analyze the phylogenetic relationships. RAxML8 [34] was used to build the Maximum likelihood tree with GTRGAMMAI module and 1000 bootstrap replicates. Mrbayes3.2.7a [35] was used to build the Bayesian inference tree. The parameter settings are as follows: nst=6, rates=invgamma, burnin=500,
Ngen=20000, Samplefreq=10, Printfreq=100. Both the results of ML tree and BI tree were visualized by FigTree V1.4.3 (http://tree.bio.ed.ac.uk/software/figtree/).

Divergence time estimation

The divergence time of 16 species was estimated by BEAST2 [36]. The oldest *Artemisia* fossil pollen has been recorded from the Eocene-Oligocene boundary [37-38]. The Asteraceae family plant *Artemisia gmelinii* and *Chrysanthemum boreale* were selected as outgroup and the node *Artemisia-Chrysanthemum* was constrained by using a lognormal distribution with an offset of 31 Ma and a mean and standard deviation of 0.5 [39]. The HKY nucleotide substitution model and priors tree Yule model was selected with strict clock. Each MCMC run had a chain length of 100,000,000 with sampling every 10,000 steps. Tracer [40] was used to read the ESS and Trace value of logged statistics to access the results. Then, the divergence time was accessed by Treeannotator program of BEAST2. The detailed settings are: Burnin percentage=50, Posterior probability limit=0.0, Target tree type=Maximum clade credibility tree, Node heights=Mean heights.

Declarations

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Availability of data and materials

All the data and materials are available from the corresponding authors upon request.

Authors’ contributions

Jiawei Li and Han Lv obtained the samples. Tianyu Han and Bingru Ren analyzed the data.
Mimi Li, Jian Chen and Tianyu Han conceived and wrote the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Not applicable.

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References

1. Vanijajiva O. A revision of Gynura (Asteraceae: Senecioneae). Journal of Systematics & Evolution, 2011, 49(4):285-314.

2. Hassan Z, Yam MF, Ahmad M, et al. Antidiabetic Properties and Mechanism of Action of Gynura procumbens Water Extract in Streptozotocin-Induced Diabetic Rats. Molecules, 2010, 15(12):9008-9023.

3. Choi SI, Lee HA, Han JS. Gynura procumbens extract improves insulin sensitivity and suppresses hepatic gluconeogenesis in C57BL/KsJ-db/db mice. Nutrition Research and Practice, 2016, 10.

4. Deng YX, Chen YS, Zhang WR, et al. Polysaccharide from Gynura divaricata modulates the activities of intestinal disaccharidases in streptozotocin-induced diabetic rats. The
British journal of nutrition, 2011, 106(9):1323-1329.

5. Xu BQ, Yang P, Zhang YQ. Hypoglycemic activities of lyophilized powder of *Gynura divaricata* by improving antioxidant potential and insulin signaling in type 2 diabetic mice. Food & Nutrition Research, 2015, 59(1):29652.

6. Tan C, Wang Q, Luo C, et al. Yeast α-Glucosidase Inhibitory Phenolic Compounds Isolated from *Gynura medica* Leaf. International Journal of Molecular Sciences, 2013, 14(2):2551-2558.

7. Rosidah, Yam MF, Sadikun A, et al. Toxicology evaluation of standardized methanol extract of *Gynura procumbens*. Journal of Ethnopharmacology, 2009, 123(2):0-249.

8. Zhang F, Zhou Y, Yang X, et al. *Gynura Rhizoma* containing pyrrolizidine alkaloids induces the hepatic sinusoidal obstruction syndrome in mice via upregulating fibrosis-related factors. Acta Pharmacologica Sinica, 2018.

9. Palmer, DJ. Comparative Organization of Chloroplast Genomes. Annual Review of Genetics, 1985, 19(1):325-354.

10. Palmer JD, Jansen RK, Michaels HJ, et al. Chloroplast DNA variation and plant phylogeny. Annals of the Missouri Botanical Garden, 1988, 75(4):1180-1206.

11. Raubeson LA, Jansen RK, Henry RJ. Chloroplast genomes of plants. 2005.

12. Erixon P, Oxelman B. Reticulate or tree-like chloroplast DNA evolution in *Sileneae (Caryophyllaceae)*?. Molecular Phylogenetics & Evolution, 2008, 48(1):313-325.

13. Marco PF, Martins AM and Ferreira ME. Molecular dating of phylogenetic divergence between *Urochloa* species based on complete chloroplast genomes. BMC Genomics, 2017, 18(1):516-.

14. Ng PK, Lin SM, Lim PE, et al. Complete chloroplast genome of *Gracilaria firma* (*Gracilariaceae, Rhodophyta*), with discussion on the use of chloroplast phylogenomics in the subclass Rhodymeniophycidae. BMC Genomics, 2017, 18(1):40.
15. Liu L, Wang Y, He P, et al. Chloroplast genome analyses and genomic resource development for epilithic sister genera *Oresitrophe* and *Mukdenia* (*Saxifragaceaee*), using genome skimming data. BMC Genomics, 2018, 19(1):235.

16. Fang WB, Ying W, Jiao Y, et al. Comparative Chloroplast Genomics of *Dipsacales* Species: Insights Into Sequence Variation, Adaptive Evolution, and Phylogenetic Relationships. Frontiers in Plant Science, 2018, 9:689.

17. Doyle JJ, Doyle JL. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem Bull. 1987;19:11-5.

18. Dierckxsens N, Mardulyn P, Smits G. NOVOPlasty *de novo* assembly of organelle genomes from whole genome data. Nucleic Acids Research, 2016,45(4):e18.

19. Wyman SK, Jansen RK, Boore JL. Automatic annotation of organellar genomes with DOGMA. Bioinformatics, 2004, 20(17):3252-3255.

20. Tillich M, Lehwark P, Pellizzer T, et al. GeSeq - versatile and accurate annotation of organelle genomes. Nucleic Acids Research, 2017, 45(1):3.

21. Laslett, D. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Research, 2004, 32(1):11-16.

22. Lowe T M, Chan P P. tRNAscan-SE On-line: integrating search and context for analysis of transfer RNA genes. Nucleic Acids Research, 2016,44: W54-57.

23. Lohse M, Drechsel O, Bock R. OrganellarGenomeDRAW (OGDRAW): a tool for the easy generation of high-quality custom graphical maps of plastid and mitochondrial genomes. Current Genetics, 2007, 52(5-6):267-274.

24. Richard GF, Kerrest A, Dujon B. Comparative genomics and molecular dynamics of DNA repeats in eukaryotes. Microbiol Mol Biol Rev, 2008, 72(4): 686-727.

25. Gulcher J. Microsatellite markers for linkage and association studies. Cold Spring Harbor Protocols, 2012, 7(4):425-432.
26. Kurtz S, Choudhuri JV, Ohlebusch E, et al. REPuter: the manifold applications of repeat analysis on a genomic scale. Nucleic Acids Research, 2001, 29(22):4633-42.

27. Nakamura T, Yamada KD, Tomii K, et al. Parallelization of MAFFT for large-scale multiple sequence alignments. Bioinformatics, 2018.

28. Frazer KA, Pachter L, Poliakov A, et al. VISTA: computational tools for comparative genomics. Nucleic Acids Res, 2004, 1:32 (Web Server issue): W273-9.

29. Librado P, Rozas J. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. Bioinformatics, 2009, 25(11):1451-1452.

30. Carver T, Harris SR, Berriman M, et al. Artemis: an integrated platform for visualization and analysis of high-throughput sequence-based experimental data. Bioinformatics, 2012, 28(4):464-469.

31. Larkin MA, Blackshields G, Brown NP, et al. Clustal W and Clustal X version 2.0. Bioinformatics, 2007, 23(21):2947-2948.

32. Xia X, Kumar S. DAMBE7: New and Improved Tools for Data Analysis in Molecular Biology and Evolution. Molecular Biology and Evolution, 2018.

33. Yang ZH. PAML 4: phylogenetic analysis by maximum likelihood. Molecular Biology & Evolution, 2007, 24(8):1586-1591.

34. Stamatakis A. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics, 2014, 30(9):1312-1313.

35. Ronquist F, Teslenko M, van der Mark P, et al. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Syst Biol. 2012;61(3):539-42.

36. Bouckaert R, Heled J, Denise K, et al. BEAST 2: A Software Platform for Bayesian Evolutionary Analysis. PLoS Computational Biology, 2014, 10(4): e1003537.

37. Wang WM. On the origin and development of *Artemisia* (Asteraceae) in the geological
past. Botanical Journal of the Linnean Society, 2004, 145(3):331-336.

38. Hobbs CR, Baldwin BG. Asian origin and upslope migration of Hawaiian Artemisia (Compositae—Anthemideae). Journal of Biogeography, 2013, 40(3):442-454.

39. Panero JL, Crozier BS. Macroevolutionary dynamics in the early diversification of Asteraceae. Molecular Phylogenetics and Evolution, 2016:S105579031600083X.

40. Rambaut A, Drummond AJ, Xie D, et al. Posterior Summarization in Bayesian Phylogenetics Using Tracer 1.7. Systematic Biology, 2018, 67(5):??-??.

41. Wang RJ, Cheng CL, Chang CC, et al. Dynamics and evolution of the inverted repeat-large single copy junctions in the chloroplast genomes of monocots. BMC Evolutionary Biology, 2008, 8(1):36-0.

42. Burrows PA, Sazanov LA, Svab Z, et al. Identification of a functional respiratory complex in chloroplasts through analysis of tobacco mutants containing disrupted plastid ndh genes. Embo Journal, 1998, 17(4):868-876.

43. Shikanai T. Cyclic Electron Transport Around Photosystem I: Genetic Approaches. Annual Review of Plant Biology, 2007, 58(1):199-217.

44. Pelser PB, Kennedy AH, Tepe EJ, et al. Patterns and causes of incongruence between plastid and nuclear Senecioneae (Asteraceae) phylogenies. American Journal of Botany, 2010, 97(5):856-873.

45. Davies FG. The Genus Gynura (Compositae) in Malesia and Australia. Kew Bulletin, 1980, 35(4):711-734.

Tables

Table 1 Overview of chloroplast genome of 16 Senecioneae species.
| Species                  | Size(bp) | LSC(bp) | SSC(bp) | IR(bp) | GC% | Protein | rRNA | tRNA | Total genes |
|--------------------------|----------|---------|---------|--------|-----|---------|------|------|-------------|
| Gynura bicolor           | 150930   | 83258   | 18128   | 24772  | 37.2| 91      | 8    | 35   | 134         |
| Gynura divaricata        | 150723   | 82998   | 18163   | 24781  | 37.2| 91      | 8    | 35   | 134         |
| Gynura formosana         | 151104   | 83368   | 18164   | 24786  | 37.2| 91      | 8    | 35   | 134         |
| Gynura pseudocchina      | 151023   | 83330   | 18131   | 24781  | 37.2| 91      | 8    | 35   | 134         |
| Dendrosenecio brassiciformis | 150551 | 83426   | 17749   | 24688  | 37.5| 89      | 8    | 37   | 134         |
| Dendrosenecio cheranganiensis | 150606 | 83458   | 17768   | 24690  | 37.5| 89      | 8    | 37   | 134         |
| Dendrosenecio johnstonii | 150607   | 83471   | 17756   | 24690  | 37.4| 89      | 8    | 37   | 134         |
| Dendrosenecio kilimanjaro | 150593  | 83457   | 17756   | 24690  | 37.5| 89      | 8    | 37   | 134         |
| Dendrosenecio meruensis  | 150587   | 83450   | 17757   | 24690  | 37.5| 89      | 8    | 37   | 134         |
| Jacobaea vulgaris        | 150689   | 82816   | 18277   | 24798  | 37.3| 87      | 8    | 37   | 132         |
| Ligularia hodgsonii      | 151136   | 83254   | 18218   | 24832  | 37.5| 94      | 8    | 36   | 138         |
| Ligularia intermedia     | 151152   | 83259   | 18233   | 24830  | 37.5| 94      | 8    | 36   | 138         |
| Ligularia jalauensis     | 151148   | 83264   | 18226   | 24829  | 37.5| 94      | 8    | 36   | 138         |
| Ligularia mongolia       | 151118   | 83245   | 18215   | 24829  | 37.5| 93      | 8    | 36   | 137         |
| Ligularia veitchiana     | 151253   | 83331   | 18248   | 24837  | 37.5| 94      | 8    | 36   | 138         |
| Pericallis hybrida       | 151267   | 83246   | 18331   | 24845  | 37.3| 95      | 8    | 36   | 139         |

Table 2 Molecular evolutionary rate of 7 *ndh* genes in 16 Senecioneae species.
| Gene | ndhA | ndhB | ndhC | ndhD | ndhE | ndhF | ndhG | ndhH | ndhI | ndhJ | ndhK |
|------|------|------|------|------|------|------|------|------|------|------|------|
| dN/dS | 0.05971 | 0.71728 | 0.16003 | 0.34997 | 0.00017 | 0.33121 | 0.39237 | 0.13607 | 0.00017 | 0.06077 | 0.34337 |

**Figures**
Figure 1

Chloroplast genome map of Gynura divaricata. Genes inside the circle are transcribed clockwise and genes outside are transcribed counter-clockwise. The ratio of light gray inside to dark gray outside represents the ratio of AT/CG content. The colors of different genes correspond to different functional groups in the legend.
Figure 2

The repeat motif statistics of 7 Senecioneae species. (a) Frequency of repeat types. (b) Frequency of unit size.
Schematic representation of the border positions of LSC, IRs and SSC in the chloroplast genome of 16 Senecioneae species.
Sequence divergence of chloroplast genome sequences in 7 Senecioneae species.
(a) The Pi value (nucleotide diversity) of the 7 chloroplast genome sequences. (b)
The sequence divergence from 25,000 bp to 50,000 bp visualized by mVISTA program. The vertical scale indicates percentage identity, ranging from 50% to 100%.
Figure 5

Maximum-likelihood (ML) phylogenetic tree obtained for 16 Senecioneae species based on the whole chloroplast genome sequences. Unlabeled nodes have bootstrap values of 100%. Noted nodes shows the bootstrap values of ML/BI.
Divergence time estimation of 18 Asteraceae species. Dotted lines shorten the proportional length. The left and right numbers in square brackets are 95% HPD upper and lower bound respectively.

Figure 6

Supplementary Files

This is a list of supplementary files associated with the primary manuscript. Click to download.

S3.bmp
S2.jpg
S1.jpg