Portable low-cost open-source wireless spectrophotometer for fast and reliable measurements

Katrina Laganovska a,*, Aleksejs Zolotarjovs a, Mercedes Vázquez b, Kirsty Mc Donnell b, Janis Liepins d, Hadar Ben-Yoav c, Varis Karitans a, Krisjanis Smits a

a Institute of Solid State Physics, University of Latvia, Kengaraga Str. 8, Riga LV-1063, Latvia
b School of Chemical Sciences, National Centre for Sensor Research, Dublin City University, Glasnevin, Dublin 9, Ireland
c Department of Biomedical Engineering and Ilse Katz Institute of Nanoscale Science and Technology, Ben-Gurion University of the Negev, Beer Sheva 8410501, Israel
d Institute of Microbiology and Biotechnology, University of Latvia, Jelgavas Str. 1, Riga LV-1004, Latvia

A R T I C L E   I N F O

Keywords: Absorption Spectroscopy Mobile application Arduino Spectrometer

A B S T R A C T

We demonstrate a low-cost standalone portable spectrophotometer for fast and reliable measurement execution. The data acquired can be both displayed via a dedicated smartphone application or a computer interface, allowing users either to gather and view data on the move or set up a continuous experiment. All design and software files are open-source and are intended for the device to be easily replicable and further customizable to suit specific applications. The assembled device can measure absorption in the wavelength range from 450 nm to 750 nm with a resolution of 15 nm and is housed in a 90 × 85 × 58 mm casing. Validation of the device was carried out by assessing wavelength accuracy, dynamic range and the signal-to-noise ratio of the system, followed by testing in three different applications where limit of quantification, limit of detection and relative standard deviations were determined. The results indicated better performance than low-cost spectrophotometers, on average being comparable to moderate to high-cost spectrophotometers.

© 2020 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

Specifications table:

| Hardware name       | Open-Source Miniature Spectrophotometer |
|---------------------|-----------------------------------------|
| Subject area        | General                                 |
| Hardware type       | ● Measuring physical properties and in-lab sensors |
|                     | ● Field measurements and sensors        |
| Cost of hardware    | 225EUR                                  |
| Source file repository | All supporting software and descriptions are available at https://doi.org/10.17605/OSF.IO/RBFSE. |

* Corresponding author.
E-mail address: katrina.laganovska@cfi.lu.lv (K. Laganovska).

https://doi.org/10.1016/j.ohx.2020.e00108
2468-0672/© 2020 The Author(s). Published by Elsevier Ltd.
This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).
1. Hardware in context

Spectrophotometric systems are widely used in studies across many fields such as physics, materials science, chemistry, biochemistry, biomolecular chemistry, photovoltaics and more [7,3,18,2,17,14,8]. Many of these applications require highly precise and reliable data which often leads to the implementation of complex and expensive spectrophotometric systems. However, in addition to rendering impossible the range of applications which require portability or good time resolution (measuring the whole spectrum at once as opposed to scanning), when performing simpler applications such as day-to-day measurements or pilot studies, convenience can often outweigh the need for precision, making the complexity of classical systems an unnecessary and a time consuming obstacle.

There have been several projects that have focused on building a portable and compact spectrophotometer that could be used on-the-go [20,1,6,21,4], however, most of these projects have been aimed at creating very low-cost devices, usually to be used for educational purposes [1,6,21] or to obtain simple and limited information and not for acquiring scientifically significant data, which requires reproducible and accurate measurements.

Commercially available handheld mini-spectrophotometers on the other hand are mostly used in a reflective configuration [5,15] and obtain qualitative information about the samples. Although analyzing the reflected light has its advantages (no sample preparation, can measure solids etc.), it can not offer the quantitative results that are needed to acquire absolute values. This is due to the spectrometer not collecting all of the reflected light, as is achieved with the transmitted light in the transmission configuration.

In this work we describe the construction and performance of the portable, low-cost, customizable and fast (registers the whole wavelength range at once, allowing to obtain a full absorbance spectrum every second) open-source miniature spectrophotometer (OSMS).

2. Hardware description

The OSMS device is based on the Hamamatsu C12880MA spectrometer chip, used in a transmission configuration. Transmittance configuration was chosen over reflectance as it offers absolute values instead of relative values that are given by reflectance. The chip is controlled by simple and widely-available microcontrollers and data is transferred via Bluetooth to a mobile application. Connection to a PC via USB interface is also possible for more complex (absorbance-over-time, data accumulation etc.) data acquisition and analysis (e.g. kinetics measurements). The device can measure absorption in the range from 450 nm to 750 nm and uses a single warm white LED as the light source. The open-source miniature spectrophotometer (OSMS) proposed in this study can be adapted to suit specific fields such as food analysis, water quality, biological research, and others by adjusting the software or adding additional LED light sources.

The miniature spectrophotometer device consists of only 4 main components – an LED, a spectrometer, a microcontroller and a Bluetooth module (and a power source if used in portable mode) as can be seen in Fig. 1 and allows the user to acquire absolute absorbance values of the measured samples. The case accommodates for the insertion of a standard cuvette (10 mm × 10 mm) which contains the sample to be measured. Although the device is generally designed for liquid samples, solid samples can also be measured if attached to a quartz holder.

- The recorded data can be easily viewed and sent to a computer using the mobile application and be analyzed from thereon.
- The device registers the full spectrum at once, making it especially attractive to studies that are time sensitive.
- The size of the finished device is fitting for field applications where larger equipment would be difficult to use.

A note on why the visible range was chosen: the Hamamatsu miniature spectrometer chips are only available for visible range. Spectrometers for other ranges are significantly costlier (up to 20×) and also larger in size. The second reason is the light source. Nichia Optisolis was chosen as it emits in a wider range of spectrum than a regular white LED and we suggest others who build the device to choose similar LEDs. However, even with improved range of spectrum, the LED does not cover wavelengths over 780 nm. 750 nm was chosen as the limit in our device, even though the spectrometer chip itself can cover up to 850 nm (for the same reason we chose a lower limit of 450 nm, even though the limit of the spectrometer itself is 340 nm. As was already mentioned it is possible to add additional LEDs to cover other specific ranges.

3. Design files

3.1. Electronics

A schematic of the components was shown in Fig. 1 The device consists of the Hamamatsu C12880MA spectrometer, a white LED (Nichia Optisolis was used), an Arduino Nano microcontroller for control and readout of data, and a Bluetooth module for data transfer to the mobile application. The case of the spectrophotometer was 3D printed and a 10 mm × 10 mm (inner dimensions) cuvette is to be used for the samples. After the sample measurement is obtained,
The calculated absorption spectrum is sent to the smartphone via the custom mobile application and the data is displayed there. From the application, the data can then be forwarded via e-mail for further analysis.

The schematic in Fig. 2 shows the wiring between all elements. There are three main components: C12880MA spectrometer chip, HC-05 Bluetooth module, and an Arduino Nano.

The Hamamatsu C12880MA spectrometer chip is connected to Arduino Nano both for voltage supply and for readouts. Pins 4, 6 and 7 are Clock, Start Pulse, and Trigger pulse and are connected to the digital output pins of the Arduino and the video signal on pin 10 is read via the analog input pin A0.

The resistors $R_1$ and $R_2$ work as a voltage divider, transforming the 5 V signals coming from Arduino to 3.3 V which are needed according to the HC-05 module datasheet. The TX pin is left unmodified as the Arduino TTL levels will register a 3.3 V as a high state. The TX and RX of the HC-05 is connected to the digital pins D11 and D10 of Arduino, that are set up to act as a software serial port. Arduino itself is attached to a 9 V power source via a simple switch.

3.2. 3D printing

The 3D model was created in Autodesk Fusion 360 software. Overall dimensions of the assembled device are $90 \times 85 \times 58$ mm (W x D x H). The case is assembled of three different parts: main body, lid and top cap (lid for the cuvette) (Fig. 3a). Main body: allows simple assembly, three different spaces for LED diodes for future upgrades (Fig. 3-4), mounting point for the spectrometer unit (Fig. 3-6) as well as additional features: switch mount (Fig. 3-7), fixture hole (Fig. 3-8) and cuvette holding elements (Fig. 3-5). The lid contains holes for screws and light-tight mount for top cap an essential element for background-free measurement. When placed on a flat surface and the top cap lid is mounted, there is no significant amount of light entering the measuring device.

For prototyping purposes, the case is optimized for fused deposition modeling (FDM) 3D printing minimal amount of supports is needed (only for holes and switch mount). All three parts of the design are printed separately. Material choice any thermoplastic filament. For better results and to lower the reflectivity of the internal surfaces a black color is desirable. Layer height 0.2 mm or lower, print time around 10 h depending on the precision and printer. Minimal infill (20%) is acceptable. Apart from the top cap, the design does not contain any moving components.

3.3. Software and the application

The device can be controlled in two ways. For simple and portable measurements, an Android application has been created (see Fig. 4), which offers the user to either acquire the whole absorption spectrum with the option to send it via e-mail.
for a more detailed analysis (Fig. 4a) or a specific wavelength can be set and the absorption value will be displayed on the screen (Fig. 4b).
The Python-based software for interfacing with a computer via a USB port offers the same functionality as the Android application with the additional options to automatically record and observe absorption data over time, set an extended accumulation time and export the data directly to the computer (Fig. 4c).

### 3.4. Design files summary

| Design filename          | File type    | Open source license                  | Location of the file                      |
|-------------------------|--------------|---------------------------------------|-------------------------------------------|
| Schematic               | figure       | GNU General Public License (GPL) 3.0  | available at https://doi.org/10.17605/OSF.IO/RBFSE |
| Android mobile application | .apk file    | GNU General Public License (GPL) 3.0  | available at https://doi.org/10.17605/OSF.IO/RBFSE |
| MIT App Inventor project file | .aia file    | GNU General Public License (GPL) 3.0  | available at https://doi.org/10.17605/OSF.IO/RBFSE |
| Python executable       | .exe file    | GNU General Public License (GPL) 3.0  | available at https://doi.org/10.17605/OSF.IO/RBFSE |
| Python source code      | .py file     | GNU General Public License (GPL) 3.0  | available at https://doi.org/10.17605/OSF.IO/RBFSE |
| 3D design files         | .stl file    | GNU General Public License (GPL) 3.0  | available at https://doi.org/10.17605/OSF.IO/RBFSE |

**Schematic:** The electronic schematic of the device with all the necessary components and their connections.

**Android mobile application:** The ready-to-install Android application file containing the application shown in Fig. 4.

**MIT App Inventor project file:** In order to enable the customization of the application for a wider range of users it was created using the MIT App Inventor. This is the project file which can be easily edited once opened via the MIT App Inventor.

**Python executable:** The executable file for desktop use. Offers accumulation functions as well as a continuous mode of measurements and file saving.

**Python source code:** The code behind the Python executable. Users can create changes here if need for customization arises.

**3D design files:** These are the 3D design files we used for our final container and lid printing.
4. Bill of materials

| Designator | Component                  | Number | Cost per unit (currency) | Total cost | Source of materials | Material type |
|------------|----------------------------|--------|--------------------------|------------|---------------------|---------------|
| U1         | Arduino Microcontroller    | 1      | 20                       | 20         | Arduino             | Electronics   |
| U2         | C12880MA Spectrometer      | 1      | 190                      | 190        | Hamamatsu           | Electronics   |
| U3         | Bluetooth module HC-05     | 1      | 7.5                      | 7.5        | Amazon              | Electronics   |
| L1         | LED Nichia Optisolis       | 1      | 2                        | 2          | Nichia Optisolis    | Electronics   |
| R1         | Resistor 1 k               | 1      | 0.02                     | 0.02       | Local hardware store| Electronics   |
| R2         | Resistor 2.2 k             | 1      | 0.02                     | 0.02       | Local hardware store| Electronics   |
| R3         | Resistor 4.7 k             | 1      | 0.02                     | 0.02       | Local hardware store| Electronics   |
| S1         | Switch                     | 1      | 0.10                     | 0.10       | Local hardware store| Electronics   |
| PS1        | 9 V battery                | 1      | 2                        | 2          | Local hardware store| Electronics   |
| Container  | 3D printed container       | 1      | 1.10                     | 1.10       | Fillamentum         | 3D printing   |
| Lid        | 3D printed lid             | 1      | 0.4                      | 0.4        | Fillamentum         | 3D printing   |

5. Build instructions

The electronic parts should be soldered together as was shown in Fig. 2. Mind that the spectrometer C12880MA as well as the light source LED need to have longer wires since they are a bit farther away from the micro-controller. The 3D design was purposefully built more robust as a first prototype and could be made smaller if mounted on a PCB.

6. Operation instructions

6.1. To use the device without making changes

1. Assemble the device.
2. Upload Arduino code to your Arduino microcontroller.
3. Install Android app or run the Python software.

6.2. To obtain an absorption spectrum

1. Connect to Bluetooth if using app (you will need to pair your HC-05 module with your phone first)
2. Measure the baseline (the spectrum of the LED without the sample in).
3. Insert sample.
4. Choose either “Single” or “Continuous” measurement in Desktop mode or “Capture” in the Android app.
5. Repeat from step 2 for additional samples (however it is recommended to re-measure the baseline every once in a while).

Notes:

- Accumulation and continuous measurements are only possible in Desktop mode
- When changing accumulation time, make sure to measure the baseline again.

7. Validation and characterization

7.1. Characteristics of the spectrometer

To assess the quality of the main component of the system – the spectrometer chip, we determined the parameters that are frequently applied to commercial spectrometers in order to gauge their performance. Wavelength accuracy, dynamic range and signal-to-noise ratios were determined and are described below.
7.1.1. Wavelength accuracy tests

The ready-built C12880MA spectrometer does not offer any calibration features and only comes with factory set wavelength positions for each pixel of the CMOS linear image sensor (that serves for accumulating the incoming light); therefore testing wavelength accuracy of the spectrometer itself was a key element in assessing the quality of future measurements. The accuracy was determined by measuring the emission spectra of an RGB LED with each color component turned on separately (Fig. 5) using the C12880MA spectrometer and a commercial Andor Shamrock B-303i spectrograph coupled with an Andor DU-401A-BV CCD camera. Slit width was kept the same (50 μm) for both spectrometers in order to acquire comparable results. The result is an average difference in the maximum emission wavelengths of 1.04 nm as compared to the Andor results. The shown data has not been corrected.

As the difference (step) between adjacent pixels of the CMOS sensor (for the OSMS) for R and G peaks are 2.09 nm, 2.34 nm (the pixel to wavelength conversion is not linear), and the wavelength differences in maxima are less than half of that, it can be deduced that the maxima of the OSMS are as close to those of the commercial system as hardware limitations allow and cannot be viewed as real differences in the peak positions. For peak B, the difference between adjacent pixels is 2.2 nm and the difference between the OSMS and commercial system maxima is 1.3 nm, showing a bias towards the longer wavelengths that is not explained by adjacent pixel differences of the OSMS, however, it is also explained if the adjacent pixel step of the commercial system are also taken into account (0.55 nm).

7.1.2. Signal-to-noise ratio and dynamic range

The dynamic range is a measure of the intensity resolution of a spectrometer and was 10 bits divided by the average of standard deviations for 50 measurements of dark noise with an exposure time of 200 μs.

\[
\text{Dynamic range} = \frac{2^{10} - 1}{\text{RMS}_{\text{dark noise}}} = \frac{1023}{11.5} = 89 \text{ dB}
\]

The signal-to-noise ratio varies depending on wavelength and the chosen light source accordingly as can be seen in Fig. 6 (normalized data). The signal-to-noise ratio was calculated as the average value of 50 measurements divided by the standard deviation of the same measurements, using an exposure time of 200 μs. For the signal at 540 nm (light source supply power kept constant, intensity at 550 nm resulting in about half maximum) the SNR value was 19 dB showing that the obtained spectra are reliable enough for medium quality measurements.

\[
\text{SNR} = \frac{\text{Signal}}{\text{RMS}_{\text{noise}}} = \frac{496.8}{6.4} = 77.6 = 19 \text{ dB}
\]

![Fig. 5. Spectra of an RGB LED with each color component turned on separately. Inset table shows the position of maxima and the difference when measured by the miniature spectrophotometer OSMS and the commercial system Com.](image-url)
7.2. Comparison with commercial spectrophotometers

To test the OSMS in practical applications, three studies were performed and altogether additional 3 commercial spectrophotometers of various quality (price range) were used. For vitamin B₁₂ and phosphate measurements the commercial spectrophotometer Shimadzu UVMini1240 was used (with a wavelength range from 190 nm to 1100 nm and a spectral bandwidth of 5 nm [16]. A bench-top photo-filter colorimeter (WPA colourwave CO7500, filter bandwidth 40 nm) and a high-quality (TECAN Infinite M200 PRO) (with a wavelength range from 230 nm to 1000 nm and a spectral bandwidth of <9 nm for >315 nm) were used for horseradish peroxidase activity measurements. The absorbance values presented are unmodified absorbance values obtained for the samples in a 10 mm cuvette. LOQ and LOD were calculated by the following formulas:

\[
\text{LOQ} = \frac{3.3 \times S_d}{b}
\]

\[
\text{LOD} = \frac{10 \times S_d}{b}
\]

where \( S_d \) is the standard deviation of the ordinate intercept and \( b \) is the slope of the regression line.

7.2.1. Vitamin B₁₂

The first set of absorption measurements were performed for various concentrations of vitamin B₁₂, starting at 3 ppm and going up to 24 ppm. Vitamin B₁₂ was chosen as the model analyte as it is water soluble and of high importance to the health of a human being [19,12], making the measurements both simple and of practical use. Absorbance values were registered at 550 nm.

The obtained results show linear dependence (Fig. 7) between the concentration of B₁₂ and the absorbance values in the range of concentrations tested. Similar \( R^2 \) values for both the OSMS and the commercial bench-top spectrophotometer are seen. The tests were repeated on 3 different days \((n = 3)\), performing 3 replicate measurements each day \((n = 3)\) in order to determine the inter-day and the intra-day precision, respectively. The relative standard deviation can be seen in Table 1 for day to day measurements (inter-day precision) and the series measurements (intra-day precision). The results show slight differences in day to day measurements for both spectrophotometers, with a maximum RSD value of 6.60% obtained for the OSMS at the lowest concentration in comparison with a maximum RSD of 4.76% obtained for the bench-top commercial spectrophotometer (also at the lowest concentration). RSD values for the intra-day precision of the OSMS were found to be in the range of 3.78% and 0.82% in comparison with the RSD range of 3.33% and 0.70% found for the bench-top commercial spectrophotometer. Same LODs and very similar LOQs were obtained with both spectrophotometers.

---

**Fig. 6.** The normalized SNR spectrum and the spectrum of the light source LED (Nichia Optisolis). SNR value for peak at 540 nm is approximately 19 dB.
7.2.2. Phosphate

Another set of measurements was performed for samples containing different levels of phosphate (up to 2 ppm) [9]. Phosphates are an essential nutrient and are most commonly used as a fertiliser, however, the unsystematic application of phosphates leads to the necessity of monitoring the amounts of phosphates entering the water system. Excessive amounts of phosphates can directly lead to the overpopulation of some organisms (such as phytoplankton), which in turn would affect the ecosystem as a whole if left unsupervised [22,13]. The concentration of phosphates in large water systems is constantly monitored by environmental health organizations across the world, however, smaller water reserves are often left unmonitored and can loose their biodiversity. Handheld portable devices such as the open-source miniature spectrophotometer could make the examinations easier and more routine, allowing for greater control over the health of the local ecosystems. The represented absorbance values have been measured at 700 nm.

Although the obtained results show linear dependence between absorbance and concentration in the range of concentrations tested (Fig. 8), the obtained regression lines showed poorer fit for the UVMini1240 spectrometer measurements. As with B₁₂, the tests were performed on different days (n = 2) for determination of inter-day precision and the corresponding SD values can be seen in Table 2.

![Graph showing absorbance vs concentration for phosphate measurements with R² values and OSMS vs Com comparison](image)

**Table 1**
Absorbance values and calculated RSD, R², LOD and LOQ for vitamin B₁₂ concentrations in ranges from 3 ppm to 24 ppm – OSMS is the miniature spectrophotometer and Com is the commercial one.

|          | Inter-day (n = 3) | Intra-day (n = 3) |
|----------|------------------|------------------|
|          | RSD, %           | OSMS             | Com              | RSD, %           | OSMS             | Com              |
| Absorbance |                  |                  |                  |                  |                  |                  |
| 3 ppm     | 0.030            | 0.033            | 6.60             | 4.76             | 3.78             | 3.33             |
| 5 ppm     | 0.056            | 0.062            | 2.85             | 3.45             | 1.40             | 1.68             |
| 10 ppm    | 0.081            | 0.089            | 3.14             | 3.48             | 0.82             | 1.43             |
| 15 ppm    | 0.109            | 0.119            | 2.80             | 3.47             | 2.17             | 1.05             |
| 20 ppm    | 0.158            | 0.173            | 4.00             | 3.15             | 1.67             | 0.91             |
| 24 ppm    | 0.189            | 0.200            | 1.90             | 2.07             | 1.59             | 0.70             |
| Average   | 0.988            | 0.989            | 3.55             | 3.39             | 1.90             | 1.52             |

K. Laganovska et al. / HardwareX 7 (2020) e00108
The RSD values for the obtained data can vary greatly when measuring low (null) absorbances (up to 220.06% for the OSMS and 120.44% for the benchtop spectrophotometer). Such measurements partially reflect the noise of the systems. Here standard deviations for the null measurement were 0.016 and 0.012 for the OSMS and the bench-top spectrophotometer, respectively. Combined with the lower absorbance value for the OSMS, the RSD value becomes significantly higher. The average standard deviation for all samples for both spectrophotometers differs only by 0.0006. Not taking into account the 0 ppm measurements, the average RSD values between the OSMS and the benchtop spectrophotometer differ by only 1–2% for both intra and inter-day measurements. RSD values for the intra-day measurements were found to be in the range of 5.20% to 21.89% for the OSMS and 5.47% to 25.69% for the commercial spectrophotometer. The OSMS showed a slightly lower LOD and LOQ.

7.2.3. Horseradish peroxidase

Horseradish peroxidase (HRP) is an active and robust enzyme widely used as a reporter for coupled biochemical or immunological reactions [11,10]. Many electron donors are used as substrates in peroxidase reactions, some of them changing their colour when donating an electron to an OH ion. 3,3,5,5 tetramethylbenzidine (TMB) is one of typical substrates used in HRP coupled reactions. When oxidised, the substrate changes its colour from colourless to turquise. In order to test the performance of the OSMS in determining HRP coupled reactions, spectrophotometric measurements of the reaction product (oxidised TMB) were performed. Low concentrations of H₂O₂ in the excess of enzyme and TMB were prepared to ensure that all H₂O₂ was oxidised to O₂ and H₂O and the blue colour present in the cuvette was directly proportional to the H₂O₂ concentration. The absorption of oxidised TMB was measured at 600 nm. Two different commercial spectrophotometers were used for comparison a simple bench-top WPA colourwave CO7500 (Com1) and the monochromator based spectrophotometer Infinite M200 PRO (Com2).

The obtained results show linear dependency for concentrations in the range from 0 μM to 132 μM, with the linear region starting to diverge at 264 μM (Fig. 9). Despite the deviation from linearity, all of the spectrophotometers showed an $R^2$ above 0.99 with $R^2$ for the OSMS being higher than that of Com1 and lower than $R^2$ of Com2. Three replicate measurements were performed to obtain intra-day standard deviation values ($n = 3$). RSD values are found to be in ranges from 6.99% to 21.07% for the OSMS, 0.00% to 8.33% for the bench-top photo filter colorimeter and 3.09% to 9.43% for the monochromator based spectrophotometer (Table 3). Similar to phosphate measurements, the rather high RSD value of 21.07% is explained by the absorption value being lower with standard deviations on average being a similar 0.037, 0.029 and 0.036 for the OSMS, Com1 and Com2 respectively. LOD and LOQ were also significantly higher for the OSMS due to the same bias toward lower absorption values in general than for the commercial spectrophotometers.

Fig. 8. Phosphate absorbance measurements and $R^2$ values for the open-source miniature spectrophotometer OSMS and the commercial one Com.
Table 2
Absorbance values and calculated RSD, R^2, LOD and LOQ for phosphate concentrations (expressed as total phosphorous) in ranges from 0.2 ppm to 2 ppm – OSMS is the open-source miniature spectrophotometer and Com is the commercial one.

| Concentration (ppm) | OSMS Absorbance | Com Absorbance | Inter-day (n = 2) | Intra-day (n = 3) |
|---------------------|-----------------|----------------|-------------------|------------------|
| 0 ppm               | 0.008           | 0.012          | 220.06            | 36.67            |
| 0.2 ppm             | 0.037           | 0.024          | 66.16             | 36.67            |
| 0.4 ppm             | 0.059           | 0.046          | 41.52             | 36.67            |
| 0.8 ppm             | 0.103           | 0.084          | 20.68             | 36.67            |
| 1.2 ppm             | 0.171           | 0.143          | 11.79             | 36.67            |
| 1.6 ppm             | 0.233           | 0.221          | 8.95              | 36.67            |
| 2 ppm               | 0.321           | 0.298          | 52.74             | 36.67            |
| Average             | 0.050           | 0.050          | 9.65              | 36.67            |

R^2 values:
- OSMS: 0.989
- Com: 0.975

LOD:
- OSMS: 0.2
- Com: 0.3

LOQ:
- OSMS: 0.5
- Com: 0.8

Fig. 9. Hydrogen peroxide absorbance measurements and R^2 values for the open-source miniature spectrophotometer OSMS and two commercial spectrophotometers – the low-cost (photo filter) Com1 and the high-cost (monochromator) Com2.

Table 3
Absorbance values and calculated RSD, R^2, LOD and LOQ for hydrogen peroxide concentrations in ranges from 22 μM to 264 μM – OSMS is the open-source miniature spectrophotometer and Com1 and Com2 are correspondingly the low-cost and high-cost commercial spectrophotometers.

| Concentration (μM) | OSMS Absorbance | Com1 Absorbance | Com2 Absorbance | Inter-day (n = 2) | Intra-day (n = 3) |
|--------------------|-----------------|-----------------|-----------------|-------------------|------------------|
| 0 μM               | 0.016           | 0.050           | 0.066           | 21.07             | 0.00             |
| 22 μM              | 0.137           | 0.183           | 0.189           | 9.25              | 3.09             |
| 44 μM              | 0.256           | 0.307           | 0.296           | 7.90              | 3.77             |
| 66 μM              | 0.414           | 0.457           | 0.433           | 7.15              | 3.34             |
| 88 μM              | 0.514           | 0.537           | 0.515           | 6.99              | 5.99             |
| 132 μM             | 0.744           | 0.753           | 0.727           | 7.11              | 5.03             |
| 264 μM             | 1.282           | 1.223           | 1.260           | 8.10              | 7.42             |
| Average            | 0.995           | 0.990           | 0.996           | 9.65              | 4.84             |

R^2 values:
- OSMS: 0.995
- Com1: 0.990
- Com2: 0.996

LOD:
- OSMS: 1.9
- Com1: 1.1
- Com2: 1.0

LOQ:
- OSMS: 5.9
- Com1: 3.5
- Com2: 3.2
Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This article is based upon COST Action CA16215 and supported by the ERDF, European-Union Project No.1.1.1.2/16/I/001.

References

[1] D.R. Albert, M.A. Todt, H.F. Davis, A low-cost quantitative absorption spectrophotometer, J. Chem. Educ. 89 (11) (2012) 1432–1435, https://doi.org/10.1021/ed0891111.
[2] C.C. Allain, L.S. Poon, C.S.G. Chan, Enzymatic determination of total serum cholesterol, Clin. Chem. 20 (4) (1974) 470–475.
[3] A. Barth, The infrared absorption of amino acid side chains, Prog. Biophys. Mol. Biol. 74 (3-5) (2000) 141–173, https://doi.org/10.1016/S0079-6107(00)00021-3.
[4] N. Chaianantakul et al, Development of mini-spectrophotometer for determination of plasma glucose, Spectrochim. Acta Part A 204 (2018) 670–676, https://doi.org/10.1016/j.saa.2018.06.107.
[5] A.J. Das et al, Ultra-portable, wireless smartphone spectrometer for rapid, non-destructive testing of fruit ripeness, Scientific Rep. 6 (2016), https://doi.org/10.1038/srep25004.
[6] E.K. Grasse, M.H. Torcasio, A.W. Smith, Teaching UV-vis spectroscopy with a 3D printable smartphone spectrophotometer, J. Chem. Educ. 93 (1) (2016) 146–151. https://doi.org/10.1021/acs.jchemed.5b00654.
[7] J. Hodgkinson, R.P. Tata, Optical gas sensing: a review, Meas. Sci. Technol. 24 (1) (2013), https://doi.org/10.1088/0957-0233/24/1/012004.
[8] J.-A. Jeong, Y.-S. Park, H.-K. Kim, Comparison of electrical, optical, structural, and interface properties of IZO-Ag-IZO and IZO-Au-IZO multilayer electrodes for organic photovoltaics, J. Appl. Phys. 107 (2) (2010), https://doi.org/10.1063/1.3294605.
[9] J. Murphy, J.P. Riley, A modified single solution method for the determination of phosphate in natural waters, Anal. Chim. Acta 27 (C) (1962) 31–36, https://doi.org/10.1016/0003-2670(00)88444-5.
[10] T.T. Ngo, Peroxidase in chemical and biochemical analysis, Anal. Lett. 43 (10-11) (2010) 1572–1587, https://doi.org/10.1080/00032711003653874.
[11] K.H. Petersen, Novel horseradish peroxidase substrates for use in immunohistochemistry, J. Immunol. Methods 340 (1) (2009) 86–89, https://doi.org/10.1016/j.jim.2008.09.018.
[12] H. Refsum et al, Homocysteine and cardiovascular disease, Annu. Rev. Med. 49 (1998) 31–62, https://doi.org/10.1146/annurev.med.49.1.31.
[13] K.C. Ruttenber, The Global Phosphorus Cycle. vol. 10. 2013, pp. 499–558.https://doi.org/10.1016/B978-0-08-095975-7.00813-5.
[14] R. Santbergen, R.J.C. van Zolingen, The absorption factor of crystalline silicon PV cells: a numerical and experimental study, Sol. Energy Mater. Sol. Cells 92 (4) (2008) 432–444, https://doi.org/10.1016/j.solmat.2007.10.005.
[15] SCiO, https://www.consumerphysics.com/scio-for-consumers.
[16] Shimadzu, https://toshvin.com/wp-content/uploads/2014/05/UV-Mini-1240.pdf.
[17] C.M. Stoscheck, Quantitation of protein, Methods Enzymol. 182 (C) (1990) 50–68, https://doi.org/10.1016/0076-6879(90)82008-P.
[18] L.P. Vernon, Spectrophotometry determination of chlorophylls and pheophytins in plant extracts, Anal. Chem. 32 (9) (1960) 1144–1150, https://doi.org/10.1021/ac60165a029.
[19] I. Volkov, V.R. Preedy, Why food fortification with vitamin B12 is needed? vol. 2, 2013, pp. 237–45 https://doi.org/10.1007/978-1-4614-7110-3_19.
[20] E. Widiatmoko et al, A simple spectrophotometer using common materials and a digital camera, Phys. Educ. 46 (3) (2011) 332–339, https://doi.org/10.1088/0031-9120/46/3/014.
[21] M.V. Wilson, E. Wilson, Authentic performance in the instrumental analysis laboratory: building a visible spectrophotometer prototype, J. Chem. Educ. 94 (1) (2017) 44–51, https://doi.org/10.1021/acs.jchemed.6b00515.17.
[22] J.-C. Yan et al, Development of a portable setup using a miniaturized and high precision colorimeter for the estimation of phosphate in natural water, Anal. Chim. Acta 1058 (2019) 70–79, https://doi.org/10.1016/j.aca.2019.01.030.