The host micro-RNA cfa-miR-346 is induced in canine leishmaniasis

Gloria Buffi, Aurora Diotallevi, Marcello Ceccarelli, Federica Bruno, Germano Castelli, Fabrizio Vitale, Mauro Magnani and Luca Galluzzi

Abstract

Background: Leishmaniasis includes a group of anthropo-zoonotic parasitic diseases caused by a protozoan belonging to the Leishmania genus, which can have different hosts, like humans, dogs, or wild animals [1, 2]. According to WHO data, leishmaniasis shows a worldwide distribution, involving humans, dogs, or wild animals [1, 2]. According to WHO data, leishmaniasis shows a worldwide distribution, involving...
at least 98 countries, with the highest number of cases in developing countries [3]. Italy, in particular the southern regions (including Sicily and Sardinia), is a country highly endemic for leishmaniasis caused by *Leishmania (L.) infantum*, although cases of canine leishmaniasis (CanL) and human leishmaniasis are largely underestimated due to the lack of notification of new cases, and the difficulty in diagnosing or misdiagnosis [4, 5]. Although a subclinical or self-limiting form was described, CanL represents a very critical problem in veterinary medicine, because the disease can be severe and with fatal exits [6].

*Leishmania (L.) infantum* (syn. *L. (L.) chagasi* in South America) is responsible for both VL and CL in humans [7], while in dogs is the most important aetiological agent of CanL in the Mediterranean Basin, Central America, while in dogs is the most important aetiological agent of CanL in the Mediterranean Basin, Central America, South America and parts of Asia [8].

*Leishmania* is a dimorphic parasite that develops as promastigotes in the midgut of the sand-fly and as intracellular amastigotes in phagocytic cells, mainly macrophages, of the vertebrate host. *Leishmania* promastigotes enter the host macrophages, where they survive and replicate within the phagolysosomal environment, subverting the innate immune response and metabolic pathways of the host [9, 10]. Among different mechanisms activated by *L. (L.) infantum* to survive and spread the infection in the host cell, we previously reported the mild induction of the Unfolded Protein Response (UPR) in human monocyctic cell lines (U937 and THP-1) and murine primary macrophages [11]. The mild UPR could represent a common pathogenic mechanism among the different species of *Leishmania* (*Leishmania*), also considering the role of X-box binding protein 1 (XBP1) in the *L. amazonensis* infection highlighted by Dias Teixeira et al. [12]. The unconventional splicing of XBP1 mRNA, induced by the endoplasmic reticulum (ER) transmembrane protein IRE1, originates a spliced mRNA (sXBP1) encoding an active transcription factor that leads to the expression of chaperones, proinflammatory cytokines [13], and genes involved in the autophagic response [14]. Few years ago, Bartoszewski et al. showed that, during ER stress, sXBP1 can induce the expression of miR-346 in both human and murine cells, whose target genes include the major histocompatibility complex (MHC) class I gene products and interferon-induced genes [15]. We recently investigated the expression of mir-346 in a human cell infection model, showing the significant induction of miR-346, in both U937 and THP-1 human cell lines following ER stress elicited by infection with four different strains of *L. (L.) infantum* and a human clinical isolate of *L. (V.) braziliensis* [16].

MicroRNAs have gained more and more relevance in the last years in the context of host–pathogen interaction. In fact, it is known that many intracellular parasites are able to modify the miRNA expression profiles of host cells [17]. In particular, in the context of CanL, Bragato et al. reported a differential modulation in the expression of miR-150, miR-451, miR-192, miR-194, and miR-371 in peripheral blood mononuclear cell (PBMC) of symptomatic dogs naturally infected with *L. infantum*. These miRNAs can target genes involved in the immune response and pathogenesis, such as NF-kB, TNF-α, CD80, and IFN-γ [18].

In this study, we investigated the UPR and the induction of *Canis lupus familiaris* cfa-miR-346 in a canine macrophage-like cell line (DH82) infected with *Leishmania* spp. Furthermore, the presence of cfa-miR-346 in the plasma of non-infected and infected dogs and its potential role as a marker of infection was analyzed.

**Results**

**Tunicamycin treatment induces the expression of ER stress markers but not cfa-miR-346 in a canine macrophage-like cell line**

Previously it has been shown that *L. (L.) infantum* could induce a mild but significant increase in ER stress expression markers, including the spliced form of XBP1 (sXBP1), in human and murine infected macrophages [11].

To further explore these pathways in a canine cell line, we first determined whether cfa-miR-346 could be induced following ER stress in DH82 cells treated with tunicamycin, an inducer of ER stress through the inhibition of protein N-linked glycosylation. After 4 h treatment, the gene expression analysis revealed a significant induction of the ER stress markers (i.e., ATF3, ATF4, CHOP, HSPA5, sXBP1) compared to vehicle DMSO (Unpaired t-test with Welch’s correction *p* < 0.05) (Fig. 1A). Notably, the unspliced form of XBP1 (uXBP1) resulted downregulated, confirming previous findings in human and murine models. On the contrary, the expression of cfa-miR-346 did not change significantly in response to treatment with Tunicamycin, suggesting a lack of correlation between the upregulation of sXBP1 and the induction of cfa-miR-346 at least at the time point tested (Fig. 1B).

**ER stress response induction by *Leishmania* infection in canine macrophage-like cell line**

The ER stress response and induction of cfa-miR-346 were investigated in DH82 cells infected with promastigotes of four *L. (L.) infantum* strains (i.e., MHOM/TN/80/IPT1, MHOM/FR/78/LEM75, MHOM/IT/08/31U and the canine clinical isolate 42) and a *L. (V.) braziliensis* human clinical isolate (i.e., AN1) for 24 h and 48 h, as
described in methods. The infection indexes are reported in Table 1.

A significant induction of ER stress expression markers—including sXBP1—was evident after 48 h infection with *L. braziliensis*, *L. infantum* MHOM/TN/80/IPT1 and isolate 42. However, a significant upregulation of cfa-miR-346 was detected at both 24 and 48 h of infection with all *Leishmania* strains (Unpaired t-test with Welch’s correction *p* < 0.05.), suggesting the hypothesis that, in the canine cell model, the induction of cfa-miR-346 could not be related to the induction of ER stress expression markers (Fig. 2).

**Cfa-miR-346 induction is related to Leishmania spp. infection, and it is independent from GRID1 expression**

To deeply analyze the induction of miR-346 following the infection, and therefore to exclude that its regulation was not related to the phagocytosis mechanism of the macrophage, the infection was performed with both live and heat-killed (HK) parasites as described in methods.

A significant induction of miR-346 was evident at 6, 24 h, and 48 h in cells infected with live parasites (One-way ANOVA with Tukey’s Multiple Comparison Test. *p* < 0.05). After 6 h the induction of mir346 was not significantly different between cells infected with live or HK parasites, probably due to an initial response mediated by macrophage phagocytosis [19, 20]; however, at 24 and 48 h the mir346 expression in cells treated with HK parasites was significantly lower, dropping close to the values of non-infected controls (Fig. 3). These results further confirm that the induction of miR-346 at 24 h and 48 h post-infection is related to the interaction with the vital pathogen.

Cfa-miR-346 is located in intron 2 of the glutamate ionotropic receptor delta type subunit 1 (GRID1) gene. To investigate if mir-346 upregulation could be due to induction of GRID1, the expression level of GRID1 was evaluated by real-time PCR in infected and non-infected DH82 cells. The real-time PCR assays showed no amplification or amplification with Ct > 33 in some technical

### Table 1 Infection indexes in DH82 cells

| Leishmania strain/isolate                  | Infection indexa 24 h | Infection indexa 48 h |
|-------------------------------------------|-----------------------|-----------------------|
| *L. (L.) infantum* MHOM/TN/80/IPT1        | 143.3 ± 31.1          | 168.9 ± 58.8          |
| *L. (L.) infantum* MHOM/FR/78/LEM75       | 100.2 ± 22.1          | 92.7 ± 29.7           |
| *L. (L.) infantum* MHOM/IT/08/31U         | 130.0 ± 10.1          | 167.8 ± 25.8          |
| *L. (V) braziliensis* human clinical isolate AN1 | 185.2 ± 31.2          | 94.0 ± 26.3           |
| *L. (L.) infantum* canine clinical isolate 42 | 189.9 ± 31.9          | 245.7 ± 50.5          |

*a calculated by multiplying the percentage of infected macrophages by the average number of parasites per macrophage, means ± SD*
replicates, leading to the conclusion that GRID1 gene was not expressed in either infected or non-infected DH82 cells (data not shown).

Cfa-miR-346 expression in plasma of infected dogs
To explore the role of miR-346 in dogs naturally infected with *Leishmania* spp., a study was conducted on 29 mixed breed dogs from Pantelleria island and Marche region (central Italy), both endemic areas for leishmaniasis. All plasma samples used in this study were collected for diagnostic purposes during routine examination. Each dog was subjected to a physical examination to assess the presence of clinical signs compatible with leishmaniasis, to a blood sampling and/or to a lymph node aspirate. The diagnosis of CanL was based on the presence of clinical signs (weight loss, skin, and eye signs, lymphadenomegaly), serological and/or qPCR-based methods, performed on blood and/or lymph node aspirates. In two cases (dog IDs 42 and 64) the isolation of the parasite was also obtained. Following the diagnostics evaluation, the dogs were divided into two groups, negative and positive, for *Leishmania* infection, and the relative expression of miR-346 was evaluated in plasma samples obtained from all dogs. Despite the partial overlap between the distribution of miR-346 expression, the results showed that the levels of miR-346 in plasma of dogs naturally infected with *L. (L.) infantum* is significantly higher compared to non-infected dogs, confirming the previous finding obtained in vitro and suggesting the induction of miR-346 following active infection (Fig. 4).
Discussion

Many different mechanisms activated by *Leishmania* spp. to survive within the host cell have been described, e.g. counteracting the antigen presentation and interfering with the macrophage signaling cascade [10]. Among them, the role of the ER stress and Unfolded Protein Response (UPR), including the activation of IRE1-XBP1 and PERK/eIF2α/ATF4 arms of UPR, have also been investigated [11, 12, 21].

![Figure 3](image3.png)

**Fig. 3** miR-346 induction is related to *Leishmania* infection. cfa-miR-346 expression in DH82 cells infected with *L. (L.) infantum* MHOM/TN/80/IPT1 (A) and canine clinical isolate 42 (B) for 6 h, 24 h, 48 h. The graph shows the fold changes in comparison to non-infected cells (mean ± SD of two independent experiments). One-way ANOVA with Tukey’s Multiple Comparison Test. n.s.: not significant, **p < 0.01, ***p < 0.001

![Figure 4](image4.png)

**Fig. 4** Relative amount of cfa-miR-346 in plasma of non-infected (*n* = 11) and infected dogs (*n* = 18). The scatter plot graph indicates the mean ± SD. Unpaired t-test with Welch’s correction. ***p < 0.001
Although dogs are considered of primary importance in the epidemiology of leishmaniasis, the majority of the current knowledge about *Leishmania*-macrophage interaction comes from cell cultures of murine or human origin. In fact, a limited number of works have been conducted using the canine macrophage-like cell line DH82 as a cell model to study *Leishmania* infection. The DH82 cells can be considered an M2 subtype macrophage model for leishmaniasis [22], however, these cells were less permissive to *L. infantum* infection and showed a lower number of amastigotes per macrophages compared to mouse macrophages or human U937 cell line [23]. This was evident also in the present work, since the infection indexes were rather low compared to human or murine models used in our previous publications, where infection indexes were approximately 150–600 [11, 16].

This could be explained by the fact that DH82 cells were not activated by PMA prior to the addition of the parasites; therefore, DH82 cells can multiply during the infection, influencing the infection index. Moreover, little is known about the surface molecules presented in these cells involved in parasite recognition and uptake. The low infection index could explain the late induction of ER stress markers, which was evident only at 48 h post-infection, in particular with *L. infantum* MHOM/TN/80/IPT1 and canine clinical isolate 42.

A number of recent studies demonstrated that infections caused by several *Leishmania* species can induce alteration of miRNA profile in human, murine, and canine macrophages [17]. We have recently investigated mir-346 expression in human U937 and THP-1-derived macrophages infected by *L. infantum* or *L. braziliensis*, finding a significant upregulation of this miRNA during infection [16].

In this work, we extended these findings to a canine macrophage-like cell line (DH82) by analyzing the induction of cfa-miR-346 after ER stress induction or *Leishmania* infection.

First, we showed that treatment of DH82 cells with tunicamycin significantly induced ER stress markers (including sXBP1); however, the mir-346 expression did not change at the time point tested, accounting for a sXBP1-independent mechanism of mir-346 induction in this cell line, contrary to what happened in humans [15, 16]. Although the phylogenetic relatedness between humans and dogs, the pathogenesis and the clinical manifestation of the disease is different, probably related also to a Th1/Th2 alternative activation and balance. In fact, while dogs with a chronic disease show a prevalence of Th2 response, there is not a clear Th1/Th2 dichotomy in the T-cell response in human leishmaniasis [24]. Moreover, a recent work evidenced the variability of immune response between humans and dogs related to differences in complement proteins amount [25], which could play an important role in parasite inactivation due to the different lytic properties, affecting the clinical manifestations [26]. Nevertheless, the differences in immune responses related to ER stress and miRNA dysregulation have not been investigated yet. It is noteworthy, the ER stress markers induction with tunicamycin resulted mild compared to U937, THP1 or murine macrophages (in which for example, sXBP1 was 20–100 fold upregulated) [11, 16]. In the same way, Nadaes et al. showed general low responsiveness in DH82 stimulated by LPS compared with RAW264.7 cells in terms of reactive oxygen species (ROS) and nitric oxide (NO) production, underlining a different activation mechanism in this cell line [22]. A significant induction of ER stress expression markers -including sXBP1 was observed after 48 h post-infection. Similar results have been obtained from the transcriptome analysis in dogs naturally infected where an over-expression of XBP1 and some other genes involved in ER stress and UPR have been evidenced in lymph nodes [27]. On the other hand, the cfa-mir-346 was always significantly upregulated after infection with all *Leishmania* strains and at all times tested, including 24 h from infection, when the expression of ER stress markers was still unchanged. Notably, we also showed that induction of cfa-miR-346 at early time of infection (6 h) can be elicited also by HK parasites, probably due to an initial response to macrophage-mediated phagocytosis; however, cfa-miR-346 upregulation was maintained at later time points (24 h and 48 h) only in cells infected with viable parasites. Taken together, these data account for a mechanism of cfa-miR-346 induction in canine macrophage-like cells i) depending on infection with live parasites and ii) not necessarily related to the induction of the ER stress marker sXBP1.

Afterward, we investigated the presence of cfa-miR-346 in the plasma of dogs naturally infected by *L. infantum* in comparison with plasma obtained from non-infected dogs. Our data showed a significant increase of the cfa-miR-346 expression levels in the plasma of dogs infected with *L. infantum* in comparison with plasma obtained from non-infected dogs. The circulating cfa-miR-346 was maintained at later time points (24 h and 48 h) only in cells infected with viable parasites. Taken together, these data account for a mechanism of cfa-miR-346 induction in canine macrophage-like cells i) depending on infection with live parasites and ii) not necessarily related to the induction of the ER stress marker sXBP1.

**First, we showed that treatment of DH82 cells with tunicamycin significantly induced ER stress markers (including sXBP1); however, the mir-346 expression did not change at the time point tested, accounting for a sXBP1-independent mechanism of mir-346 induction in this cell line, contrary to what happened in humans [15, 16]. Although the phylogenetic relatedness between humans and dogs, the pathogenesis and the clinical manifestation of the disease is different, probably related also to a Th1/Th2 alternative activation and balance. In fact, while dogs with a chronic disease show a prevalence of Th2 response, there is not a clear Th1/Th2 dichotomy in the T-cell response in human leishmaniasis [24]. Moreover, a recent work evidenced the variability of immune response between humans and dogs related to differences in complement proteins amount [25], which could play an important role in parasite inactivation due to the different lytic properties, affecting the clinical manifestations [26]. Nevertheless, the differences in immune responses related to ER stress and miRNA dysregulation have not been investigated yet. It is noteworthy, the ER stress markers induction with tunicamycin resulted mild compared to U937, THP1 or murine macrophages (in which for example, sXBP1 was 20–100 fold upregulated) [11, 16]. In the same way, Nadaes et al. showed general low responsiveness in DH82 stimulated by LPS compared with RAW264.7 cells in terms of reactive oxygen species (ROS) and nitric oxide (NO) production, underlining a different activation mechanism in this cell line [22]. A significant induction of ER stress expression markers -including sXBP1 was observed after 48 h post-infection. Similar results have been obtained from the transcriptome analysis in dogs naturally infected where an over-expression of XBP1 and some other genes involved in ER stress and UPR have been evidenced in lymph nodes [27]. On the other hand, the cfa-mir-346 was always significantly upregulated after infection with all *Leishmania* strains and at all times tested, including 24 h from infection, when the expression of ER stress markers was still unchanged. Notably, we also showed that induction of cfa-miR-346 at early time of infection (6 h) can be elicited also by HK parasites, probably due to an initial response to macrophage-mediated phagocytosis; however, cfa-miR-346 upregulation was maintained at later time points (24 h and 48 h) only in cells infected with viable parasites. Taken together, these data account for a mechanism of cfa-miR-346 induction in canine macrophage-like cells i) depending on infection with live parasites and ii) not necessarily related to the induction of the ER stress marker sXBP1.**
cardiac hypertrophy [31]. Regarding the infectious diseases in humans, similar to our findings, Nishimura et al. showed that hsa-miR-346 is secreted from macrophages infected by the intracellular pathogen *Mycobacterium avium* complex—which has been demonstrated to elicit ER stress-induced apoptosis in infected macrophages [32], and proposed that its serum levels could be a potential biomarker of *Mycobacterium avium* complex pulmonary disease activity [33]. In the same way, hsa-miR-346 was found to be a potential marker in monitoring the efficacy of treatments against *Mycobacterium tuberculosis* in patients with tuberculosis [34]. Since the expression of miR-346 can be influenced by various diseases, in both humans and dogs, further clinical studies will be necessary to identify the increase of cfa-miR-346 expression as a specific marker of CanL and exclude other infectious diseases that are common in dogs.

**Conclusion**

In summary, miR-346 was found to be upregulated in the canine macrophage-like cell line DH82 infected with four different strains/isolates of *L. (L.) infantum*, as well as one *L. (V.) braziliensis* isolate, confirming previous findings in human cell lines and pointing to a possible common pathogenic mechanism in human and canine host cells. Moreover, miR-346 has been shown to be upregulated in the plasma of dogs naturally infected by *L. infantum*. Nevertheless, miR-346 dysregulation is associated with various diseases, infectious or non-infectious, in both humans and dogs. According to our results, an increase of cfa-miR-346 plasma level could represent a promising potential biomarker of leishmaniasis, even though further studies will be needed to understand its role during *Leishmania* infection and to evaluate its specificity as a possible biomarker of canine leishmaniasis.

**Methods**

**Leishmania parasites cultivation**

Three *L. (L.) infantum* reference strains (MHOM/TN/80/IPT1, MHOM/FR/78/LEM75 and MHOM/IT/08/31U), and one *L. (L.) infantum* canine clinical isolate (isolate 42) were provided by the OIE Reference Laboratory National Reference Centre for Leishmaniasis (C.Re. Na.L.) (Palermo, Italy). Moreover, a *L. (V.) braziliensis* clinical isolate (isolate AN1), identified at the species level by PCR–RFLP as described by Schönian et al. [35], was obtained from a pharyngolaryngeal biopsy during routine diagnosis of a human patient with suspect MCL, as previously described [16, 36]. *Leishmania* promastigotes were cultivated at 26–28 °C in Evans’ Modified Tobie’s Medium (EMTM) and/or RPMI-PY medium [37], and stationary promastigotes were transferred to fresh medium (ratio 1:5) every 5–7 days.

**Cell culture and infection**

The canine macrophage-like cell line DH82 (ATCC® CRL-10389™) was cultured in an incubator at 37 °C and 5% CO₂ in Eagle’s Minimum Essential Medium (EMEM) supplemented with 15% heat-inactivated Fetal Bovine Serum (FBS), 2 mM L-glutamine, 10 g/l Non-Essential Amino Acid, 1 mM sodium pyruvate, 100 μg/ml streptomycin, 100 U/l penicillin. For infection experiments, 2.5 × 10⁵ cells were seeded in 35 mm dishes for 24 h for cell adhesion. Then, cells were infected with stationary-phase promastigotes of *L. (L.) infantum* strains/isolate or *L. (V.) braziliensis* clinical isolate with a parasite-to-cell ratio of 10:1. To exclude the modulation of target genes by the phagocytosis mechanism of macrophages, 2.5 × 10⁶ promastigotes of *L. (L.) infantum* MHOM/TN/80/IPT1 and canine clinical isolate 42 were inactivated (heat-killed, HK) at 70 °C for 45 min and placed into DH82 culture medium. To promote and synchronize cell infection, dishes were centrifuged at 450 × g for 3 min at room temperature. After 24 h, non-internalized parasites were removed by medium replacement. As ER-stress positive control, cells were treated with 2 μg/ml of tunicamycin for 4 h or with vehicle dimethyl sulfoxide (DMSO) as control. All cell culture reagents were purchased from Sigma-Aldrich (St. Louis, MO). After 6 h, 24 h, and 48 h from infection, cells were directly lysed for gene expression analysis by adding QIAzol Lysis Reagent to the cell-culture dish as described below in the RNA isolation and cDNA synthesis section. Moreover, one dish for each time point was stained with Hoechst and Acridine Orange dyes, and the infection index was calculated by multiplying the percentage of infected macrophages by the average number of parasites per macrophage. At least 300 macrophages were counted for each condition.

**Study design**

No direct intervention was done on animals. All plasma samples used in this study were surplus material collected for diagnostic purposes during routine examination; oral informed consent was obtained from the owners of dogs.

To evaluate the presence of cfa-miR-346 in clinical samples of non-infected and infected dogs, 21 plasma samples of mixed breed dogs from Pantelleria Island (Sicily, Italy), an endemic area for leishmaniasis, have been obtained by the C.Re.Na.L. in Palermo (Italy) [38]. Additional 8 plasma samples have been supplied by the veterinary clinic “Santa Teresa” in Fano (Pesaro-Urbino, Italy). To diagnose CanL, every dog was subjected to physical, clinical, and dermatological evaluation, including the anamnestic history, changes compatible with CanL (e.g., alopecia and desquamation, nodular or ulcerative lesions, exfoliative dermatitis weight loss,
lymphadenomegaly). In addition, serological [i.e. IFAT, SNAP test (IDEXX Laboratories, Westbrook, Maine, USA) or Speed Leish K test (BVT Groupe Virbac, La Seyne sur Mer, France] and qPCR-based tests [39] were performed on blood and/or lymph nodal aspirates. In two cases (IDs 42 and 64), *L. (L.) infantum* amastigotes isolation was obtained as previously described [40]. After diagnosis, dogs were divided into two groups: i) non-infected: negative for clinical signs and/or negative IFAT, negative qPCR, negative for SNAP test or Speed Leish K test; ii) infected: presence of clinical signs, IFAT $\geq 1:40$ and/or positive qPCR, and/or positive SNAP test or Speed Leish K test [41]. Details of samples are summarized in Table 2.

### Plasma collection

One mL of peripheral blood was collected from the jugular vein of each dog and placed into tubes containing ethylene diamine tetraacetic acid (EDTA). Blood was centrifuged at $200 \times g$ for 15 min at $4 \degree C$ to separate plasma from the corpuscular part. Plasma collected was further centrifuged at $800 \times g$ for 15 min at $4 \degree C$ to remove any cellular debris. The supernatant (150 µl) was transferred in a sterile tube and processed as described below.

### RNA isolation and cDNA synthesis

RNA extraction from DH82 infected and non-infected cells was performed with the miRNeasy Mini Kit (Qiagen, Hilden, Germany) after direct lysis with 700 µl of

---

**Table 2** Canine sample ID, summary of clinical signs, and results of diagnostic methods

| Sample ID | Clinical signs | IFAT  | SNAP TEST | qPCR (blood) | qPCR (Lymph node) | Parasite isolation | Diagnosis of leishmaniasis |
|-----------|---------------|-------|-----------|--------------|-------------------|--------------------|--------------------------|
| 24        | Lymphadenomegaly | Neg | n.a | Neg | Neg | n.a | - |
| 29        | Neg            | Neg | n.a | Neg | n.a | - | - |
| 35        | Neg            | Neg | n.a | Neg | n.a | - | - |
| 43        | Eye signs      | Neg | n.a | Neg | n.a | - | - |
| 46        | Neg            | Neg | n.a | Neg | n.a | - | - |
| 52        | Neg            | Neg | n.a | Neg | n.a | - | - |
| 54        | Neg            | Neg | n.a | Neg | n.a | - | - |
| GOA       | Neg            | Neg | n.a | n.a | n.a | - | - |
| GON       | Neg            | Neg | n.a | n.a | n.a | - | - |
| VI A      | Neg            | Neg | n.a | n.a | n.a | - | - |
| ZAR       | Neg            | Neg | n.a | n.a | n.a | - | - |
| 9         | Lymphadenomegaly | 1:160 | n.a | Neg | Pos | n.a | + |
| 11        | Lymphadenomegaly | 1:320 | n.a | Neg | n.a | - | - |
| 23        | Skin signs, Lymphadenomegaly | 1:320 | n.a | Neg | Pos | n.a | + |
| 30        | Skin signs, Lymphadenomegaly | 1:320 | n.a | Neg | Neg | Bacterial contamination | + |
| 31        | Skin signs     | 1:80 | n.a | Neg | Pos | n.a | - |
| 32        | Neg            | 1:320 | n.a | Neg | Neg | n.a | + |
| 40        | Skin signs     | 1:320 | n.a | Neg | Pos | n.a | + |
| 42        | Neg            | 1:5120 | n.a | Neg | Pos | Pos | + |
| 45        | Neg            | 1:40 | n.a | Neg | Pos | Bacterial contamination | + |
| 49        | Eye signs      | 1:160 | n.a | Neg | n.a | n.a | - |
| 51        | Neg            | Neg | n.a | Neg | Pos | n.a | + |
| 58        | Lymphadenomegaly | 1:2560 | n.a | Neg | Pos | n.a | + |
| 63        | Neg            | 1:80 | n.a | Neg | Pos | n.a | + |
| 64        | Skin and eye signs, lymphadenomegaly | 1:2560 | n.a | Neg | Pos | Pos | + |
| STA-VOI   | Mild periocular dermatitis, anemia, thrombocytopenia | n.a | Pos | n.a | n.a | n.a | + |
| MAI-EUI   | Alopecia, weight loss, mild onychogryphosis, lymphadenomegal, leucopenia, decline in performance | n.a | Pos | n.a | n.a | n.a | + |
| MIA-MAI   | Chronic anemia | n.a | Pos | n.a | n.a | n.a | + |
| KIO-MEI   | Neg            | n.a | Pos | n.a | n.a | + | + |

POS Positive  NEG Negative,  NA Data not available
QIAzol Lysis Reagent (Qiagen, Hilden, Germany). RNA extraction from plasma was carried out with Total RNA Purification Kit (Norgen Biotek Corporation, Ontario, Canada) according to the manual instructions specific for plasma. In both cases, extracted RNA was quantified with a NanoVue PlusTM spectrophotometer (GE Healthcare Life Sciences, Piscataway, NJ, United States).

For gene expression analysis in DH82 cells, total RNA (500 ng) was reverse-transcribed using PrimeScriptTM RT Master Mix (Perfect Real Time) (Takara Bio Inc.). For expression analysis of miR-16 and miR-346 in all samples, total RNA (1–5 ng/μl) was reverse-transcribed with the TaqMan™ MicroRNA Reverse Transcription Kit (Applied Biosystems).

Quantitative real-time PCR
The expression of several ER stress markers was evaluated by qPCR as previously described [11] with some modifications. Briefly, the qPCR reactions were carried out in duplicate in a final volume of 20 μl using TB Green PreMix ex Taq II Master Mix (Takara Bio Europe, France) and 200 nM primers listed in Table 3. The amplification conditions were: 95 °C for 10 min, 40 cycles at 95 °C for 10 s and 60 °C for 50 s. The microRNA cfa-miR-16 was used as reference gene [42, 43]. All qPCR reactions were performed in a RotorGene 6000 instrument (Corbett life science, Sydney, Australia). A duplicate non-template control was included for each primer pair reaction as negative control. The relative expression levels were calculated using the 2−ΔΔCt method [44].

Statistical analysis
Statistical analysis was performed by unpaired t-test with Welch's correction or one-way ANOVA with Tukey’s multiple-comparison post-test. Values are expressed as mean ± standard deviation (SD). All statistical tests were performed using GraphPad Prism version 8.02 (GraphPad Software, Inc., La Jolla, CA, USA). A P value ≤ 0.05 was considered statistically significant.

Abbreviations
WHO: World Health Organization; VL: Visceral leishmaniasis; CL: Cutaneous leishmaniasis; MCL: Mucocutaneous leishmaniasis; PCR–RFLP: Polymerase chain reaction-Restriction Fragment Length Polymorphism; qPCR: Quantitative real-time PCR; BLAST: Basic local alignment search tool; IFAT: Immunofluorescence antibody test.

Acknowledgements
We would like to thank Dr. Emanuele Gasparini from the Santa Teresa Veterinary Clinic, Fano (PU) Italy, for providing canine clinical samples. We are also appreciative to Dr. Mauro De Santi (University of Urbino Carlo Bo) for assisting with cell laboratory activities.

Authors’ contributions
Conception and design of the study: GB and AD. Acquisition of data: GB, AD, MC. Analysis, and interpretation of data: GB, AD, MC, MM, and LG. Drafting the article: GB and AD. Revising the article critically for important intellectual content: MC, FB, GC, FV, MM, and LG. All authors read and approved the final manuscript.
Funding
This work was partially supported by funds from the Italian Ministry of Health (grant number Ricerca Corrente 2018 IZS Si 02/18 RC) and FANOATENEO. The funder had no role in study design, data collection and analysis, interpretation of the data, or writing the manuscript.

Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate
The study did not require the approval of an ethical committee since no direct intervention was done on animals. All plasma samples used were surplus material from the diagnostic tests during routine examination with no unnecessary invasive procedures, including parasitological confirmation of leishmaniasis. Oral informed consent was obtained from the owners of dogs at the time of clinical examination. After the examination, all dogs were returned to their owners.

Consent for publication
Not applicable.

Competing interests
The authors declare no competing interests for the submission of this article.

Author details
1 Department of Biomolecular Sciences, Section of Biotechnology, University of Urbino Carlo Bo, via Arco d'Augusto 2, 61032 Fano, PU, Italy. 2 Centro di Ricerca Nazionale Per le leishmaniosi (C.R.E.Na.L), OIE Leishmania Reference Laboratory, Istituto Zoonolettico Sperimentale della Sicilia A. Miriti, via G. Marinuzzi 3, 90129 Palermo, PA, Italy.

References
1. Abbate JM, Arfuso F, Napoli E, Gaglio G, Giannetto S, Latrofa MS, et al. Leishmania infantum in wild animals in endemic areas of southern Italy. Comp Immunol Microbiol Infect Dis. 2019;67:101374. Available from: https://doi.org/10.1016/j.cimid.2019.101374. 2019;67:101374. Available from: https://doi.org/10.1016/j.cimid.2019.101374.
2. Ribeiro J, Oturdu M,Peres-Cutillas P, Goyena E, Maia C, Cortes S, et al. Epidemiological and genetic studies suggest a common Leishmania infection transmission cycle in wildlife, dogs and humans associated to vector abundance in Southeast Spain. Vet Parasitol. 2018;259:61–7. Available from: https://doi.org/10.1016/j.vetpar.2018.05.012.
3. Alvar J, Vélez ID, Bern C, Hierro M, Desjeux P, Cano J, et al. Leishmania Worldwide and Global Estimates of its Incidence. PLoS One. 2012;7(15):e55671. Available from: https://doi.org/10.1371/journal.pone.0055671.
4. Gaspari V, Ortalli M, Foschini MP, Baldiviño C, Lanzoni A, Cagarella R, et al. New evidence of cutaneous leishmaniasis in north-eastern Italy. J Eur Acad Dermatol Venereol. 2017;31(9):1534–40. Available from: https://doi.org/10.1111/jdv.14309.
5. Verso MG, Vitale F, Castelli G, Bruno F, Migliazzo A, Bongiorno MR, et al. Suspected cutaneous leishmaniasis in a sample of Western Sicilian residents: What correlation with occupation? Med del Lav. 2017;108(2):123–9. Available from: https://doi.org/10.23749/mdl.v108i2.5665.
6. Solano-Gallego L, Miro G, Koutinas A, Cardoso L, Pennisi MG, Ferrer L, et al. LeishVet guidelines for the practical management of canine leishmanioses. ParasitVectors. 2011;4:86. Available from: https://doi.org/10.1186/1756-3305-4-86.
7. Martín-Sánchez J, Rodríguez-Granger J, Morillas-Márquez F, Merino-Espinosa G, Sampredo A, Aliaga L, et al. Leishmania infantum: Integration of human, animal and environmental data through a One Health approach. Transbound Emerg Dis. 2020;67(6):2423–34. Available from: https://doi.org/10.1111/tbed.13580.
8. Ruiz-Postigo JA, Jain S, Mikhailov A, Maia-Elkhoury AN, Valadas S, Wauysavathana S, et al. Global leishmaniasis surveillance: 2019–2020, a baseline for the 2030 roadmap. Relev Epidemiolerep. 2021;35(86):401–19. Available from: https://www.who.int/publications/i/item/who-wer9635-401-491.
9. Arango Duque G, Descoteaux A. Leishmania survival in the macrophage: Where the ends justify the means. Curr Opin Microbiol. 2015;26:32–40. Available from: https://doi.org/10.1016/j.mib.2015.04.007.
10. Podinovskia M, Descoteaux A. Leishmania and the macrophage: A multifaceted interaction. Future Microbiol. 2015;10(1):111–29. Available from: https://doi.org/10.2217/fmb.14.103.
11. Galluzzi L, Diotallevi A, De Santi M, Ceccarelli M, Vitale F, Brandi G, et al. Leishmania infantum Induces Mild Unfolded Protein Response in Infected Macrophages. PLoS One. 2016;11(12):e0168339. Available from: https://doi.org/10.1371/journal.pone.0168339.
12. Dias-Teixeira KL, Calegari-Silva TC, Dos Santos GRRM, Dos Santos JV, Lima C, Medina JM, et al. The integrated endoplasmic reticulum stress response in Leishmania amazonensis macrophage infection: The role of X-box binding protein 1 transcription factor. FASEB J. 2016;30(4):1557–65. Available from: https://doi.org/10.1096/fg.15-281550.
13. Martinon F, Chen X, Lee AH, Glimcher LH. Toll-like receptor activation of XBP1 regulates innate immune responses in macrophages. Nat Immunol. 2010;11:411–8. Available from: https://doi.org/10.1038/ni.1857.
14. Margariti A, Li H, Chen T, Martin D, Vázquez-Barrena G, Alam S, et al. XBP1 mRNA splicing triggers an autophagic response in endothelial cells through BECLIN-1 transcriptional activation. J Biol Chem. 2013;288(2):859–72. Available from: https://doi.org/10.1074/jbc.M112.412783.
15. Bartoszewski R, Brewer JW, Rab A, Crossman DK, Bartoszewska S, Kapoor N, et al. The Unfolded Protein Response (UPR)-activated transcription factor X-box-binding protein 1 (XBP1) induces microRNA-346 expression that targets the human antigen peptide transporter 1 (TAP1) mRNA and governs immune regulatory genes. J Biol Chem. 2011;286(48):41862–70. Available from: https://doi.org/10.1074/jbc.M111.304956.
16. Diotallevi A, De Santi M, Buffi G, Ceccarelli M, Vitale F, Galluzzi L, et al. Leishmania infection induces MicroRNA hsa-miR-346 in human cell line-derived macrophages. Front Microbiol. 2018;9:1019. Available from: https://doi.org/10.3389/fmicb.2018.01019.
17. Acrua SM, Floeter-Winter LM, Muxel SM. MicroRNAs: Biological Regulators in Pathogen-Host Interactions. Cells. 2020;9(1):113. Available from: https://doi.org/10.3390/cells9010113.
18. Bragato JP, Melo LM, Venturini GL, Rebech GT, Garcia LE, Lopes FL, et al. Relationship of peripheral blood mononuclear cells mRNA expression and parasitic load in canine visceral Leishmaniasis. Parasitol Int. 2009;58(4):359–66. Available from: https://doi.org/10.1016/j.parint.2009.07.008.
19. Nadaes NR, Silva da Costa L, Santana RC, LaRocque-de-Freitas IF, de Carvalho-Obriani A, Soares DC, et al. DH82 Canine and RAW264.7 Murine Macrophage Cell Lines Display Distinct Activation Profiles Upon Interaction With Leishmania infantum and Leishmania amazonensis. Front Cell Infect Microbiol. 2020;10:247. Available from: https://doi.org/10.3389/fcimb.2020.01019.
20. Miramín-Mohammadi A, Javadi A, Eskandari SE, Nateghi-Rostami M, Khamisopour A. Immune Responses in Cutaneous Leishmaniasis: in vitro Thelper1/Thelper2 Cy-tokine Profiles Using Live Versus Killed Leishmania infantum. Front Immunol. 2018;9:1019. Available from: https://doi.org/10.3389/fimmu.2018.001019.
21. Nadaes NR, Silva da Costa L, Santana RC, LaRocque-de-Freitas IF, de Carvalho-Obriani A, Soares DC, et al. DH82 Canine and RAW264.7 Murine Macrophage Cell Lines Display Distinct Activation Profiles Upon Interaction With Leishmania infantum and Leishmania amazonensis. Front Cell Infect Microbiol. 2020;10:247. Available from: https://doi.org/10.3389/fcimb.2020.01019.
22. Wildt K, Nadaes NR, Silva da Costa L, Santana RC, LaRocque-de-Freitas IF, de Carvalho-Obriani A, Soares DC, et al. DH82 Canine and RAW264.7 Murine Macrophage Cell Lines Display Distinct Activation Profiles Upon Interaction With Leishmania infantum and Leishmania amazonensis. Front Cell Infect Microbiol. 2020;10:247. Available from: https://doi.org/10.3389/fcimb.2020.01019.
25. Tirado TC, Bavia L, Ambrosio AR, Campos MP, de Almeida Santiago M, Messias-Reason JJ, et al. A comparative approach on the activation of the three complement system pathways in different hosts of Visceral Leishmaniasis after stimulation with Leishmania infantum. Dev Comp Immunol. 2021;120:104061. Available from: https://doi.org/10.1016/j.dci.2021.104061.

26. Mendes-Sousa AF, Nascimento AAS, Queiroz DC, Vale VF, Fujiwara RT, Araújo RN, et al. Different host complement systems and their interactions with saliva from Lutzomyia longipalpis (Diptera, Psychodidae) and Leishmania infantum promastigotes. PLoS One. 2013;8(11)e79787. Available from: https://doi.org/10.1371/journal.pone.0079787.

27. Sanz CR, Miró G, Sevane N, Reyes-Palomares A, Dunner S. Modulation of Host Immune Response during Leishmania infantum Natural Infection: A Whole-Transcriptome Analysis of the Popteal Lymph Nodes in Dogs. Front Immunol. 2022;12:794627. Available from: https://doi.org/10.3389/fimmu.2021.794627.full.

28. Sang J, Lu X, Zhang C, Zhang X, Li G. Circ-TOP2A acts as a ceRNA for miR-346 and contributes to glioma progression via the modulation of sushi domain-containing-2. Mol Med Rep. 2021;23(4):255. Available from: https://doi.org/10.3892/mmr.2021.11894.

29. Yi B, Huang J, Zhang W, Li AM, Yang SK, Sun J, et al. Vitamin D receptor down-regulation is associated with severity of albuminuria in type 2 diabetes patients. J Clin Endocrinol Metab. 2016;101(1):439–404. Available from: https://doi.org/10.1210/je.2016-1516.

30. Wroblewska JP, Lach MS, Ustaszewski A, Kulcenty K, Ibbis M, Jagiełło I, et al. The potential role of selected miRNAs in uveal melanoma primary tumors as early biomarkers of disease progression. Genes. 2020;11(3):271. Available from: https://doi.org/10.3390 GENES11030271.

31. Woong-Bin R, Kang MH, Song DW, Lee SH, Park HM. Expression Profile of Circulating MicroRNAs in Dogs With Cardiac Hypertrophy: A Pilot Study. Front Vet Sci. 2021;8:652224. Available from: https://doi.org/10.3389/fvets.2021.652224.

32. Go D, Lee J, Choi J, Cho S, Kim S, Son S, et al. Reactive oxygen species-mediated endoplasmic reticulum stress response induces apoptosis of Mycobacterium avium-infected macrophages by activating regulated IRE1-dependent decay pathway. Cell Microbiol. 2019;21(12). Available from: https://doi.org/10.1111/cmi.13094.

33. Nishimura T, Tamizu E, Uno S, Uwamino Y, Fujiwara H, Nishio K, et al. hsa-miR-346 is a potential serum biomarker of Mycobacterium avium complex pulmonary disease activity. J Infect Chemother. 2017;23(10):703–8. Available from: https://doi.org/10.1016/j.jiac.2017.07.015.

34. Uno S, Nishimura T, Nishio K, Kohsaka A, Tamizu E, Nakano Y, et al. Potential biomarker enhancing the activity of tuberculosis, hsa-miR-346. Tuberculosis. 2021;129:102101. Available from: https://doi.org/10.1016/j.tube.2021.102101.

35. Schönian G, Nasereeddin A, Dinse N, Schumann N, Fuchs-HOF, Preisner B, et al. PCR diagnosis and characterization of Leishmania in local and imported clinical samples. Diagn Microbiol Infect Dis. 2003;47(1):349–58. Available from: https://doi.org/10.1016/S0732-8893(03)00093-2.

36. Cecarelli M, Buffi G, Diotallevi A, Andreoni F, Bencardino D, Vitale F, et al. Evaluation of a qDNA-Based qPCR Assay for the Detection and Quantification of Old World Leishmania Species. Microorganisms. 2020;8(12):2006. Available from: https://doi.org/10.3390/MICROORGANISMS8122006.

37. Castelli G, Galante A, Verde Lo V, Migliazzo A, Reale S, Lupo T, et al. Evaluation of two modified culture media for leishmania infantum cultivation versus different culture media. J Parasitol. 2014;100(2):228–30. Available from: https://doi.org/10.1655/13-253.1.

38. Vitalle F, Bruno F, Migliazzo A, Galante A, Vullo A, Graziano R, et al. Cross-sectional survey of canine leishmaniasis in Pantelleria Island in Sicily. 2020; 56(2):91-3. Available from: https://www.veterinarniaitaliana.izs.it/index.php/revista/view/2059/719.

39. Castelli G, Bruno F, Reale S, Catanzaro S, Valenza V, Vitalle F. Molecular Diagnosis of Leishmaniasis: Quantification of Parasite Load by a Real-Time PCR Assay with High Sensitivity. Pathogens. 2021;10(7):865. Available from: https://doi.org/10.3390/pathogens10070865.

40. Castelli G, Bruno F, Caputo V, Fiorella S, Sammarco I, Lupo T, et al. Genetic tools discriminate strains of leishmania infantum isolated from humans and dogs in Sicily, Italy. PLoS Negl Trop Dis. 2020;14(7):1–16. Available from: https://doi.org/10.1371/journal.pntd.0008046.

41. Paltrinieri S, Solano-Gallego L, Fondati A, Lubas G, Gradoni L, Castagnaro M, et al. Guidelines for diagnosis and clinical classification of leishmaniasis in dogs. J Am Vet Med Assoc. 2010;236(11):1184–91. Available from: https://doi.org/10.2460/javma.236.11.1184.

42. Heishima K, Ichikawa Y, Yoshida K, Iwasaki R, Sakai H, Nakagawa T, et al. Circulating microRNA-214 and -126 as potential biomarkers for canine neoplastic disease. Sci Rep. 2017;7:2301. Available from: https://doi.org/10.1038/s41598-017-02607-1.

43. Hulanicka M, Garncarz M, Parzeniecka-Jaworska M, Janik M. Plasma miRNAs as potential biomarkers of chronic degenerative valvular disease in Dachshunds. BMC Vet Res. 2014;10(1):205. Available from: https://doi.org/10.1186/s12917-014-0205-8.

44. Pfaffl MW. A new mathematical model for relative quantification in real-time RT-PCR. Nucleic Acids Res. 2001;29(9):e45. Available from: https://doi.org/10.1093/nar/29.9.e45.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.