Nectin/PRR: An Immunoglobulin-like Cell Adhesion Molecule Recruited to Cadherin-based Adherens Junctions through Interaction with Afadin, a PDZ Domain-containing Protein

Kenichi Takahashi,* Hiroyuki Nakanishi,* Masako Miyahara,* Kenji Mandai,* Keiko Satoh,* Ayako Satoh,* Hideo Nishioka,* Junken Aoki,‡ Akio Nomoto,‡ Akira Mizoguchi,§ and Yoshimi Takai*†

*Takai Biotimer Project, ERATO, Japan Science and Technology Corp., c/o JCR Pharmaceuticals Co., Ltd., Kobe 651-2241, Japan; ‡Department of Microbiology, Tokyo Metropolitan Institute of Medical Science, Tokyo 113-0033, Japan; §Department of Anatomy and Neurobiology, Faculty of Medicine, Kyoto University, Kyoto 606-8501, Japan; and †Department of Molecular Biology and Biochemistry, Osaka University Medical School, Suita 565-0871, Japan

Abstract. We have isolated a novel actin filament–binding protein, named afadin, localized at cadherin-based cell–cell adherens junctions (AJs) in various tissues and cell lines. Afadin has one PDZ domain, three proline-rich regions, and one actin filament–binding domain. We found here that afadin directly interacted with a family of the immunoglobulin superfamily, which was isolated originally as the poliovirus receptor–related protein (PRR) family consisting of PRR1 and -2, and has been identified recently to be the alphaherpes virus receptor. PRR has a COOH-terminal consensus motif to which the PDZ domain of afadin binds. PRR and afadin were colocalized at cadherin-based cell–cell AJs in various tissues and cell lines. In E-cadherin–expressing EL cells, PRR was recruited to cadherin-based cell–cell AJs through interaction with afadin. PRR showed Ca\textsuperscript{2+}–independent cell–cell adhesion activity. These results indicate that PRR is a cell–cell adhesion molecule of the immunoglobulin superfamily which is recruited to cadherin-based cell–cell AJs through interaction with afadin. We rename PRR as nectin (taken from the Latin word “necto” meaning “to connect”).

Key words: afadin • cadherin • poliovirus receptor–related protein • immunoglobulin superfamily • zonula adherens

Cell–cell adhesion plays essential roles in various functions, including the maintenance of the integrity of organized tissues, the control of cell growth, and tissue morphogenesis (Edelman, 1986; Takeichi, 1988, 1991, 1993; Albelda and Buck, 1990; Buck, 1992; Gumbiner, 1996; Barth et al., 1997). The functional units of cell–cell adhesion are typically composed of cell adhesion molecules (CAMs)\textsuperscript{1} and cytoplasmic peripheral membrane proteins (Gumbiner, 1996). CAMs mediate cell–cell adhesion at the extracellular surface by a homophilic or heterophilic interaction and determine the specificity of cell–cell recognition. At the intracellular surface, CAMs interact with cytoplasmic peripheral membrane proteins, which are linked to the cytoskeleton, to regulate the functions of CAMs and to transduce signals initiated by CAMs.

CAMs are classified into groups, including the cadherin family (cadherin), the Ig superfamily (IgCAM), the integrin family, and the selectin family. Of the CAM families, cadherin plays crucial roles in cell–cell adhesion of a majority of cell types. In polarized epithelial cells, cell–cell adhesion forms a specialized membrane structure, comprised of zonula occludens (ZO, tight junctions), zonula adherens (ZA; cell–cell adherens junctions [AJs]), and desmosome, which is known as the junctional complex. These junctional structures are typically aligned from the apical side to the basal side, although desmosome is independently distributed in other areas.

At ZA, classic cadherins interact with each other at the extracellular surface in a Ca\textsuperscript{2+}–dependent manner (Takeichi, 1988, 1991; Giger and Ginsberg, 1991; Tsukita et al.,

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1. Abbreviations used in this paper: AJs, adherens junctions; CAM, cell adhesion molecule; F-actin, actin filament; GST, glutathione S-transferase; PRR, poliovirus receptor–related protein; ZA, zonula adherens; ZO, zonula occludens.
The cytoplasmic region of cadherin interacts with β- and γ-catenins, and β-catenin interacts with α-catenin (Ozawa et al., 1989; Nagafuchi et al., 1991; Takeichi, 1991; Tsukita et al., 1992). α-Catenin interacts directly with actin filament (F-actin) (Rimm et al., 1995) and indirectly with it through α-actinin and vinculin, other F-actin-binding proteins (K nudsen et al., 1995; Weiss et al., 1998). ZA is distinguished from cadherin-based cell–cell AJ s in nonepithelial cells by having a thick F-actin belt, called the circumferential filament band. Furthermore, while ZA is defined as a junctional structure observed by electron microscopy, the cadherin-catenin system is not confined to ZA, but is distributed along the entire lateral membrane (Tsukita et al., 1992; Gumbiner, 1996). α-Acatinin is also broadly distributed (Geiger et al., 1981). In this study, therefore, we use “cell–cell AJs” s as cell–cell adhesion sites where cadherin is present, and distinguish “cell–cell AJs” s from “ZA.” Vinculin is more highly concentrated at ZA than the cadherin-catenin system (Geiger et al., 1981; Y onenura et al., 1995).

Many cytoplasmic peripheral membrane proteins, which directly or indirectly link CAMs to the actin cytoskeleton, have been identified, but little is still known about the mechanism of how these F-actin-binding proteins are involved in the establishment of the polarized junctional alignment in epithelial cells. It has neither been fully understood how these F-actin-binding proteins regulate the function of the adhesion molecules, nor how the adhesion molecule-initiated signals are transduced to the F-actin-binding proteins.

Recently, we have isolated a novel ZA component, named l-afadin (Mandai et al., 1997). l-Afadin has been isolated as an F-actin-binding protein from rat brain. l-Afadin has one PDZ domain following the PDZ domain, and one F-actin-binding domain at the COOH-terminal region. l-Afadin is ubiquitously expressed and specifically localized at ZA in epithelial cells and at cell–cell AJs in nonepithelial cells. l-Afadin has a splicing variant, named s-l-afadin, which has one PDZ domain but lacks the third proline-rich regions following the PDZ domain, and one F-actin-binding domain at the COOH-terminal region. l-Afadin is ubiquitously expressed and specifically localized at ZA in epithelial cells and at cell–cell AJs in nonepithelial cells. l-Afadin has a splicing variant, named s-l-afadin, which has one PDZ domain but lacks the third proline-rich region and the F-actin-binding domain (Mandai et al., 1997). s-l-afadin is abundantly expressed in neural tissue. s-l-afadin is the protein encoded by the AF-6 gene, which is originally found to be fused to the ALL-1 gene (Prasad et al., 1993), known to be involved in acute leukemia (Cimino et al., 1991).

The result that l-afadin is specifically localized at ZA in epithelial cells and at cell–cell AJs in nonepithelial cells suggests the presence of an integral membrane protein(s) which interacts with l-afadin. On this assumption, we have attempted here to isolate an l-afadin-binding integral membrane protein(s) specifically localized at ZA in epithelial cells and at cell–cell AJs in nonepithelial cells. We have identified it to be the poliovirus receptor–related protein (PRR) family members (Morrison and Racaniello, 1992; Eberlé et al., 1995; Lopez et al., 1995), identified recently as the alphaherpesvirus virus receptor (Geraghty et al., 1988; Warner et al., 1998). The PRR family belongs to the Ig superfamily and consists of PRR1 and -2, which are ubiquitously expressed (Morrison and Racaniello, 1992; Aoki et al., 1994; Eberlé et al., 1995; Lopez et al., 1995). PRR2 has two splicing variants, PRR 2α and -2b, shown to serve as homophilic CA Ms (Aoki et al., 1997; Lopez et al., 1998). We describe here that PRR is a cell–cell adhesion molecule of the Ig superfamily which is specifically localized at ZA in epithelial cells and at cell–cell AJs in nonepithelial cells and recruited to cadherin-based cell–cell AJs through interaction with l-afadin. Therefore, we rename PRR as nectin (taken from the Latin word “necto” meaning “to connect”).

### Materials and Methods

#### Yeast Two-Hybrid Screening and β-Galactosidase Assay

The bait vectors, pBTM116-l-afadin-1 (amino acid [aa] 1–1015) and -2 (aa 706–1425), were constructed by subcloning the inserts encoding the respective aa residues of l-afadin into pBTM116 (Yon et al., 1993). The yeast two-hybrid library constructed from an adult rat brain cdNA was screened using a mixture of pBTM116-l-afadin-1 and -2 as baits as described (Hata et al., 1996). β-Galactosidase assay was performed as described (Hata et al., 1996).

#### Construction of Expression Vectors

The cdNA s of human nectin-1 and mouse nectin-2α and -2β were obtained by PCR using human and mouse brain cdNA s as templates, respectively. Nucleotide sequence analysis was performed by the dideoxy-nucleotide termination method using a DNA sequencer (model 373; Applied Biosystems Inc.). The nucleotide sequence of our isolated human nectin-1 cdNA was different from the originally published sequence (Lopez et al., 1995), but identical to the recently published sequence (Geraghty et al., 1998). Prokaryote and eukaryote expression vectors, bait vectors, and prey vectors were constructed in pGEX-KG (Guo and Dixon, 1991), pCMV-Myc (Nakanishi et al., 1997), pFLAG-CMV1 (Eastman Kodak Co.), pCAGGS (Niwa et al., 1991), pCAGGS-FLAG, pBTM116, and pVP16-3 (Hata and Südhof, 1995) using standard molecular biology methods (Sambrook et al., 1989). pCAGGS-FLAG was constructed by subcloning the insert encoding the preprotyrpsin-signal peptide and FLA epitope of pFLAG-CMV1 into pCAGGS. The glutathione S-transferase (GST)-fusion vectors, containing α-, β-, and γ-catenins, and the cytoplasmic region of E-cadherin (aa 734–884) (Itoh et al., 1997), were kindly supplied by Drs. M. Itoh and S. Tsukita (Kyo University, Kyoto, Japan). Various constructs of l-afadin and nectin shown in Fig. 1 were obtained by overlapping PCR using human and mouse brain cDNAs as templates, respectively. Nucleotide sequence analysis was performed by the dideoxy-nucleotide termination method using a DNA sequencer (model 373; Applied Biosystems Inc.).

#### In Vitro Binding of l-Afadin to Nectin

Afinity chromatographies were done as follows. Anti-Myc epitope Ab-coupled beads were prepared by cross-linking of a mouse anti-Myc mAb to Sepharose beads (Amersham-Pharmacia Biotech Ltd.). The purification GST-fusion protein to be tested was applied to the affinity column. After the column was washed extensively, the beads were used as an affinity column. The purified GST-fusion proteins were purified by use of glutathione-Sepharose beads (A meshrham-Pharmacica Biotech Ltd.).
were eluted by boiling the beads in an SDS sample buffer [60 mM Tris-Cl, pH 6.7, 3% SDS, 2% (vol/vol) 2-mercaptoethanol, and 5% glycerol]. The sample was then subjected to SDS-PAGE, followed by staining with Coomassie brilliant blue.

35S-labeled l-afadin blot overlay was done as described (Mandai et al., 1999). In brief, 32P-labeled l-afadin was generated using the TNT T7 quick coupled transcription/translation system (Promega Corp.) and used as a probe. The sample to be tested was subjected to SDS-PAGE and transferred to a nitrocellulose membrane. The membrane was blocked in PBS containing 5% defatted powder milk and 1% Triton X-100. The membrane was then incubated at 4°C for 16 h with 40 μl of the 32P-labeled l-afadin probe in 1 ml of PBS containing 5% defatted powder milk and 1% Triton X-100. After the incubation, the membrane was washed with PBS containing 5% defatted powder milk and 1% Triton X-100, followed by autoradiography using an image analyzer (Fuji BAS-2000II; Fuji Photo Film Co.).

**Antibodies**

Rabbit antisera against nectin-1, -2a, and -2b were raised against GST-nectin-1-CPN, GST-nectin-2a-CP, and GST-nectin-2b-CP, respectively. These antisera were separately affinity-purified by use of the respective GST-fusion proteins covalently coupled to NHS-activated Sepharose 4B (Amersham-Pharmacia Biotech Ltd.) and used as pAbs. A rat anti-l-afadin mAb was prepared as described (Aoki et al., 1997). The specificity of the anti-nectin-2 mAb used here was confirmed as follows. Immunofluorescence microscopic analysis showed that an HeLa cell line stably expressing mouse nectin-2a reacted with this mAb but wild-type HeLa cells did not (Aoki et al., 1997). In addition, when a cell lysate from mouse mammary tumor MTD-1A cells, which were metabolically radiolabeled with [35S]methionine, was subjected to immunoprecipitation with this mAb, followed by autoradiography as described (Yoshida and Takeichi, 1982), a radioactive band with a molecular mass of approximately 350 kD was specifically immunoprecipitated (data not shown). This band was recognized by another anti-nectin-2 pAb described above (data not shown). A rabbit anti-l-afadin pAb and a mouse anti-l-afadin mAb were prepared as described (Mandai et al., 1997; Sakisaka et al., 1999). Mouse and rat (ECCD 2) anti-l-afadin mAbs were purchased from Transduction Laboratories and TAKARA Shuzo, respectively. Mouse anti-Myc and anti-FLAG mAbs were from American Type Culture Collection and Easman Kodak Co., respectively.

**Immunoprecipitation**

The bile canaliculi-rich fraction was prepared from mouse liver (Tsukita and Tsukita, 1989). This fraction was sonicated in the lysis buffer described above and subjected to ultracentrifugation. After the supernatant was diluted with buffer A (20 mM Tris-Cl, pH 7.5, 150 mM NaCl, 1% Triton X-100, 1 mM EDTA, 10 μg/ml of leupeptin, 1 mM PMSF, and 1 μg/ml of pepstatin A) to make a final concentration of 0.2% deoxycholate, this sample was incubated with the anti-nectin-2 mAb at 4°C for 3 h. Protein G-Sepharose beads (Amersham-Pharmacia Biotech Ltd.) were added to this diluted sample, and incubation was further performed at 4°C for 1 h. After the beads were extensively washed with buffer A, the bound proteins were eluted by boiling the beads in the SDS sample buffer, and subjected to SDS-PAGE, followed by Western blot analysis.

Immunoprecipitation experiments using cultured cells were done as follows. Mammary tumor MTD-1A cells were sonicated in buffer A. The lysate was then subjected to ultracentrifugation. The supernatant was subjected to immunoprecipitation with the anti-nectin-2 mAb as described above. For COS7 cells expressing the FLAG-G-tagged protein and/or the Myc-tagged protein, the cells were similarly subjected to immunoprecipitation with the anti-FLAG or anti-Myc mAb. For EL cells expressing the FLAG-tagged protein, the cells were also similarly subjected to immunoprecipitation with the anti-FLAG mAb.

**Cell Culture and DNA Transfection**

MDCk cells were kindly supplied by Dr. W. Birchmeier (Max Delbruck Center for Molecular Medicine, Berlin, Germany). EL and L cells were kindly supplied by Drs. A. Nagafuchi and S. Tsukita (K yoto University, Kyoto, Japan). EL cells were cloned by introduction of the exogenous E-cadherin cDNA to cadherin-deficient L cells (Nagafuchi et al., 1987). These cells were maintained in DME containing 10% FCS.

To prepare COS7 cells transiently expressing the FLAG-tagged protein and/or the Myc-tagged protein, COS7 cells were transfected with the pFLAG-CMV1 construct and/or the pCMV-Myc construct, respectively, using the DEAE-dextran method and cultured for 2 d (Hata and Südhof, 1990). To prepare EL cells transiently expressing the FLA G-tagged protein, EL cells were transfected with the pCAGGS-FLAG construct using Lipofectamine reagent (GIBCO BRL) according to the manufacturer's protocol. The cells were then cultured for 1 d, replated, and cultured for 4 d. A MDCk cell line stably expressing FLAG-nectin-1 was prepared as described (Furuse et al., 1998). In brief, MDCk cells were transfected with pCAGGS-FLAG-nectin-1 using Lipofectamine reagent (GIBCO BRL). The cells were then cultured for 1 d, replated, and selected by cultivating in the presence of 100 μg/ml of Geneticin (GIBCO BRL). A M D c k cell line stably expressing full-length nectin-1 (nectin-1-L cells) or nectin-2a (nectin-2a-L cells) was similarly prepared with pCAGGS-nectin-1-L and pCAGGS-nectin-2a-L, respectively, except that the concentration of Geneticin was increased to 500 μg/ml.

**Immunofluorescence and Immunoelectron Microscopy**

Immunofluorescence microscopy of cultured cells and frozen sections of various mouse tissues was done as described (Mandai et al., 1997). Immunoelectron microscopy of mouse intestine absorptive epithelial cells was done using the silver-enhancement technique as described (Mizoguchi et al., 1994; Mandai et al., 1997).

**Cell Aggregation Assay**

Cell aggregation assay was done according to the method described by Takeichi (1977) with slight modifications. To obtain a single-cell suspension, cells were washed with PBS, incubated with 0.2% trypsin and 1 mM EDTA at 37°C for 5 min, and dispersed by gentle pipetting. Cells were then suspended in HBS S in the presence of 1 mM CaCl₂ or 1 mM EDTA (M iura et al., 1992) (106 cells/ml), placed in 12-well plates precoated with BSA, and rotated on a gyratory shaker at 37°C for indicated periods of time. A aggregation was stopped with the addition of 2% glutaraldehyde. The extent of aggregation of cells was represented by the ratio of the total particle number at time t of incubation (Nt) to the initial particle number (No).

**Other Procedures**

Vinulin was purified from chicken gizzard as described (O’Halloran et al., 1986). Protein concentrations were determined with BSA as a reference protein (Bradford, 1976). SD SPAGE was done as described (Laemmli, 1970). The protein markers used were myosin (197 kD), BSA (78 kD), ovalbumin (50 kD), carbonic anhydrase (33 kD), and soybean trypsin inhibitor (28 kD).

**Identification of PRR as an l-Afadin–Binding Protein**

We first attempted to identify an l-afadin–binding protein(s) by use of the yeast two-hybrid method. We screened 2 × 10⁷ clones of a prey cDNA library from rat brain with a mixture of two bait constructs, pBTM 116-l-afadin-1 (aa 1–1015) and -2 (aa 706–1425). Three independent clones, pPrey 0135, pPrey 0139, and pPrey 0140, were obtained. We focused on pPrey 0135 because this clone encoded the cytoplasmic COOH-terminal region of PRR1. PpPrey 0135 encoded a 97-aa sequence which was identical to the originally identified human PRR1 (aa 421–518) except for the absence of a single aa, glutamate, at position 438 (A F091111; GenBank/EMBL/DDBJ). The clone is likely to encode a rat counterpart of human PRR1. PRR1 has been identified recently as the alphaherpes virus receptor, and PRR1 and -2 have been designed as HveC and HveB, respectively (Geraghty et al., 1998; Warner et al., 1998) (Table I). PPR1 was here renamed nectin. R- transformation of fresh yeast cells confirmed that pPrey 0135 bound to pBTM 116-l-afadin-2 containing the PDZ

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domain. Analysis of other clones will be described elsewhere.

l-Afadin has a PDZ domain and nectin has cytoplasmic regions with a COOH-terminal motif of 4 aa residues, E/A-X-Y-V (X indicates W, V, and M for nectin-1, -2α, and -2δ, respectively) (Table I). It has been shown that PDZ domains bind to unique COOH-terminal motifs of 4-aa residues of integral membrane proteins (Saras and Heldin, 1996; Songyang et al., 1997; Hata et al., 1998). We next examined whether l-afadin specifically binds to nectin through the PDZ domain and the COOH-terminal motif. For this purpose, we constructed pBTM116-l-afadin-PDZ containing only the PDZ domain, and pVP16-nectin-2α-CP and pVP16-nectin-2δ-CP, both of which contained the cytoplasmic regions with the COOH-terminal motif (Fig. 1). We also prepared pVP16-neurexin-2α (containing the cytoplasmic COOH-terminal region of neurexin-2α) and pBTM116-neurabin-II (containing the PDZ domain of neurabin-II) as controls. Neurexin-2α has a COOH-terminal motif similar to that of nectin, E-Y-Y-V, which has been shown to bind to the PDZ domain of CASK (Hata et al., 1996). Neurabin-II is an F-actin–binding protein with one PDZ domain localized at cadherin-based cell–cell AJs (Satoh et al., 1998; Sakisaka et al., 1999), although its PDZ domain–binding protein(s) has not yet been identified. We quantified the yeast two-hybrid interactions by measuring β-galactosidase transactivation. The PDZ domain of l-afadin bound to the cytoplasmic regions of nectin, but not to that of neurexin-2α (Table II). The PDZ domain of neurabin-II did not bind to any of these proteins.

Direct Binding of l-Afadin to Nectin In Vitro and In Vivo

We first examined the in vitro direct binding of l-afadin to nectin by affinity chromatography. A GST-fusion protein of the PDZ domain of l-afadin (GST-l-afadin-PDZ) bound to a Myc-tagged protein of the cytoplasmic region of nectin-1 (Myc-nectin-1-CP) immobilized on protein A–Sepharose beads through the anti-Myc mAb (Fig. 2 A). The stoichiometry of the binding of l-afadin to nectin-1 was ~1:1. Similar results were obtained with nectin-2α and -2δ (data not shown). To further examine the direct binding of l-afadin to nectin in vitro, GST-fusion proteins of the cytoplasmic regions of nectin were subjected to SDS-PAGE, followed by a blot overlay with 35S-labeled l-afadin. 35S-labeled l-afadin bound to the GST-fusion proteins (Fig. 2, B1 and B2). However, when the COOH-terminal motif of 4-aa residues of each GST-fusion protein, which were shown in bold characters in Table I, were deleted, 35S-labeled l-afadin did not bind to the GST-fusion proteins. Consistent with the earlier observation that PDZ domains bind to unique COOH-terminal motifs

### Table I. COOH-terminal Sequences of the Nectin and Neurexin Family Members

| Nectin family | Nectin-1 | Human PRR1/HveC | SPISKKEWYV |
| Nectin-2α | Human PRR2α/HveB | SLIRARRAVYV |
| Nectin-2δ | Human PRR2δ | GFVNRSMAYV |
| Consensus | E/AXYV |
| Neurexin family | Neurexin-2α | KNNKDKKEYYV |

### Table II. Interactions between the Cytoplasmic Regions of the Nectin Family Members and the PDZ Domain of l-Afadin

|          | Control (pVP16-3) | Nectin-1 (pPrey 0135) | Nectin-2α (pVP16-Nectin-2α-CP) | Nectin-2δ (pVP16-Nectin-2δ-CP) | Neurexin-2α (pVP16-Neurexin-2α) |
|----------|------------------|----------------------|-------------------------------|-------------------------------|---------------------------------|
| Control (pBTM116) | ND | ND | ND | ND | ND |
| l-Afadin (pBTM116-l-Afadin-PDZ) | ND | 22,810 ± 2,300 | 3,420 ± 40 | 2,650 ± 70 | ND |
| Neurabin-II (pBTM116-Neurabin-II) | ND | ND | ND | ND | ND |

The data list β-galactosidase activities of yeast strains harboring the respective bait and prey plasmids. The data shown are nanomoles of substrate hydrolyzed per minute per milligram of protein ± SD after background subtraction. The β-galactosidase activities were determined by three independent experiments. ND, no detectable activity.
of 4-aa residues of integral membrane proteins (Saras and Heldin, 1996; Songyang et al., 1997; Hata et al., 1998), these results indicate that the PDZ domain of l-afadin directly binds to nectin and that the COOH-terminal motif of 4-aa residues of nectin is essential for this binding.

We next examined the binding of l-afadin to nectin in vivo. Western blot analysis indicated that nectin-2α, but not nectin-2β, was detected in mouse liver (data not shown). A n extract from the bile canaliculi-rich fraction of mouse liver was subjected to immunoprecipitation with the anti-nectin-2 mAb, which recognizes the extracellular domains of both nectin-2α and -2β (Aoki et al., 1997), followed by Western blot analysis with the anti-l-afadin pAb. l-Afadin was coimmunoprecipitated with nectin-2α (Fig. 3 A). When a cell extract from mouse mammary tumor MTD-1A cells was similarly subjected to immunoprecipitation with the anti-nectin-2 mAb, l-afadin was coimmunoprecipitated with nectin-2α and -2β (Fig. 3 B). Myc-l-afadin and FLAG-nectin-1, -2α, or -2β were overexpressed in various combinations in COS7 cells and the cell extracts were subjected to immunoprecipitation with the anti-Myc or anti-FLAG mAb, followed by Western blot analysis with these mAbs. Myc-l-Afadin and FLAG-nectin-2α or -2β were also coimmunoprecipitated (data not shown). These results indicate that l-afadin binds to nectin in vivo.

**Colocalization of Nectin with l-Afadin at Cell–Cell AJs in Nonpithelial Cells**

We next examined whether nectin and l-afadin are colocalized in nonpithelial cells. When the frozen sections of heart were doubly stained with the anti-nectin-2 mAb and the anti-l-afadin pAb, both of the proteins were colocalized at intercalated discs (cell–cell AJs) and not observed at costameres (cell–matrix AJs) (Fig. 6, A1 and A2). This result suggests that nectin and l-afadin are colocalized at cell–cell AJs in nonpithelial cells. To confirm this result, we examined their colocalization in EL cells expressing E-cadherin. EL cells were cloned by introduction of the exogenous E-cadherin cDNA to cadherin-deficient L cells (Nagafuchi et al., 1987). We have shown previously that l-afadin is localized at cell–cell AJs in cultured EL cells (Mandai et al., 1997; Sakisaka et al., 1999). In this cell line, nectin-2 was also colocalized with l-afadin at cell–cell AJs (Fig. 6, B1 and B2). These results indicate that nectin is colocalized with l-afadin at cadherin-based cell–cell AJs in nonpithelial cells.

**Recruitment of Nectin to Cadherin-based Cell–Cell AJs through Interaction with l-Afadin in EL Cells**

We then examined the function of the interaction of nectin

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**Figure 2. Direct binding of l-afadin to nectin in vitro.** (A) A affinity chromatography. GST-l-afadin-PDZ (aa 1007–1125) or GST alone (20 μg of protein each) was applied to protein A–Sepharose beads on which Myc-nectin-1-CP (aa 379–518) was immobilized through the anti-Myc mAb. After the beads were extensively washed, the bound proteins were subjected to SDS-PAGE (12% polyacrylamide gel), followed by protein staining with Coomassie brilliant blue. Arrow, GST-l-afadin-PDZ; arrowhead, Myc-nectin-1-CP. (B) 35S-Labeled l-afadin blot overlay. GST-Nectin-1-CPC, aa 387–467; GST-Nectin-2-CPC, aa 403–526.

**Table 2.** Western blot analysis of nectin-2 expression in mouse intestine absorptive epithelia and mouse liver. Nectin-2α and -2β were colocalized at intercalated discs (cell–cell AJs) and not observed at costameres (cell–matrix AJs) (Fig. 6, A1 and A2). This result suggests that nectin and l-afadin are colocalized at cell–cell AJs in nonpithelial cells. To confirm this result, we examined their colocalization in EL cells expressing E-cadherin. EL cells were cloned by introduction of the exogenous E-cadherin cDNA to cadherin-deficient L cells (Nagafuchi et al., 1987). We have shown previously that l-afadin is localized at cell–cell AJs in cultured EL cells (Mandai et al., 1997; Sakisaka et al., 1999). In this cell line, nectin-2 was also colocalized with l-afadin at cell–cell AJs (Fig. 6, B1 and B2). These results indicate that nectin is colocalized with l-afadin at cadherin-based cell–cell AJs in nonpithelial cells.
with l-afadin. We prepared EL cells transiently expressing the FLAG-tagged full length of nectin-1 (FLAG-nectin-1-EL cells) or the FLAG-tagged, COOH-terminal 4 aa–deleted mutant of nectin-1 (FLAG-nectin-1-D_C-EL cells). Immunoprecipitation analysis revealed that l-afadin was coimmunoprecipitated with FLAG-nectin-1, but not with FLAG-nectin-1-D_C (Fig. 7 A). In FLAG-nectin-1-EL cells, FLAG-nectin-1 was colocalized with l-afadin and E-cadherin at cell–cell AJs (Fig. 7, B1–B6). In FLAG-nectin-1-D_C-EL cells, however, nectin-1-D_C was not recruited to cell–cell AJs where E-cadherin was localized (Fig. 7, B7–B12). Nectin-1-D_C was not colocalized with l-afadin. These results indicate that nectin is recruited to cell–cell AJs through interaction with l-afadin in EL cells.

Cell Aggregation Activity of Nectin

It has been shown previously that nectin-2α and -2δ show cell aggregation activity (Aoki et al., 1997; Lopez et al., 1998). To first confirm this result and to then examine cell aggregation activity of nectin-1, we prepared L cells stably expressing full-length nectin-1 (nectin-1-L cells) and -2α (nectin-2α-L cells). By use of these cell lines, we examined cell aggregation activity of nectin as described (Takeichi, 1977). Nectin-1 as well as nectin-2α showed cell aggregation activity in a time-dependent manner (Fig. 8). This activity was not affected by the presence or absence of Ca^{2+} in the medium, indicating that cell–cell adhesion activity of nectin-1 and -2α is Ca^{2+} independent.

No Direct Binding of Nectin to Known Components of Cell–Cell AJs

In the last set of experiments, to understand how nectin is recruited to cadherin-based cell–cell AJs through interaction with l-afadin, we examined the in vitro binding of nectin to the known components of cell–cell AJs, including α-, β-catenins, vinculin, and E-cadherin, by affinity chromatography. Under the conditions where Myc-nectin-1-CP bound to GST-l-afadin-PDZ, it did not bind to vinculin or any GST-fusion protein of α-, β-catenins, and the cytoplasmic region of E-cadherin (data not shown). Similar results were obtained with nectin-2α and -2δ (data not shown). Recently, we found that l-afadin does not bind directly to α-, β-catenin, or the cytoplasmic region of E-cadherin (Sakisaka et al., 1999). Thus, although nectin is recruited to cadherin-based cell–cell AJs through interaction with l-afadin, the mechanism of this recruitment remains unknown.

Discussion

Nectin constitutes a family consisting of three members, nectin-1, -2α, and -2δ, and belongs to the Ig superfamily...
anti-E-cadherin mAb. They were visualized with FITC-conjugated anti-mouse IgG, rhodamine-conjugated anti-rabbit IgG, and Cy5-conjugated anti-rat IgG A bs. (B1 and B4) FLAG-nectin-1; (B2 and B5) l-afadin; (B3 and B6) E-cadherin; (B1–B3) junction-level view; (B4–B6) cross-sectional view. The cross-sectional view was generated by confocal microscopy. Arrow, apical level; arrowhead, basal level. Bars, 10 µm.

Figure 5. Ultrastructural localization sites of nectin-2 in mouse small intestine absorptive epithelial cells. Intestine absorptive epithelial cells were labeled with the anti-nectin-2 mAb using the silver-enhancement technique. Open arrow, ZO; closed arrow, ZA; asterisk, desmosome. Bars, 0.2 µm.
The interaction of α-catenin with vinculin is required for the organization of ZO (Watabe-Uchida et al., 1998). Furthermore, it has been shown that the junctional organization is impaired in vinculin-null F9 cells (Watabe-Uchida et al., 1998). The unique localization properties of nectin, l-afadin, ponsin, and vinculin suggest that the nectin-l-afadin system plays a role in the assembly of the junctional complex in cooperation with the cadherin-catenin system.

We have confirmed that nectin-2α has cell–cell adhesion activity as described (Aoki et al., 1997; Lopez et al., 1998) and have shown that nectin-1 also has this activity. In contrast to cadherin, most IgCAMs regulate cell–cell adhesion in a Ca²⁺-independent manner. Consistently, both nectin-1 and -2 show Ca²⁺-independent cell–cell adhesion. These results indicate that nectin is a Ca²⁺-independent CAM which is associated with l-afadin and specifically localized at ZA in epithelial cells and at cadherin-based cell–cell AJs in nonepithelial cells. Nectin-1 and -2 have been shown to be expressed in most tissues examined thus far (Morrison and Racaniello, 1992; Eberlé et al., 1995; Lopez et al., 1995). We have found here that the three members of the nectin family are expressed in MDCk cells. It remains to be clarified why the different nectin family members are expressed in the same cells, but the three members of the nectin family may be functionally redundant because of their common properties, including Ca²⁺-independent cell–cell adhesion activity, l-afadin-binding activity, and localization at cadherin-based cell–cell AJs.

We have analyzed here the binding regions of l-afadin and nectin and found that the PDZ domain of l-afadin and the cytoplasmic regions of nectin directly interact with each other. PDZ domains are modular domains that bind to specific COOH-terminal peptide sequences (Saras and Heldin, 1996; Ponting et al., 1997; Hata et al., 1998). M any PDZ domain–containing proteins and their binding partners have been isolated recently, and peptide sequences for various PDZ domains have been reported. Using the oriented peptide library technique, PDZ domains are assigned into classes according to their peptide-binding specificities (Songyang et al., 1997). The PDZ domain of A F-6 (s-afadin) is classified as the class II, selecting peptides with hydrophobic or aromatic aa residues at position -2 relative to the COOH terminus. The PDZ domain binds preferentially to a peptide which terminates in the sequence, E-F-Y-V (Songyang et al., 1997). Nectin termi-
Figure 7. Recruitment of nectin to cadherin-based AJs through interaction with l-afadin. (A) Immunoprecipitation from EL cells expressing FLAG-nectin-1 or FLAG-nectin-1-ΔC. FLAG-nectin-1 or FLAG-nectin-1-ΔC was transiently expressed in EL cells. Each cell extract was subjected to immunoprecipitation with the anti-FLAG mAb. The immunoprecipitate was then subjected to SDS-PAGE (8 or 12% polyacrylamide gel), followed by Western blot analysis with the anti-FLAG or anti-l-afadin mAb. (B) Immunofluorescence microscopy of FLAG-nectin-1-EL cells and FLAG-nectin-1-ΔC-EL cells. FLAG-nectin-1-EL cells and FLAG-nectin-1-ΔC-EL cells were doubly stained with the mouse anti-FLAG mAb and the rat anti-E-cadherin mAb or the rabbit anti-l-afadin pAb. They were visualized with FITC-conjugated anti-mouse IgG and rhodamine-conjugated anti-rat or anti-rabbit IgG Abs. There was nuclear staining with this anti-l-afadin pAb, but its significance is not clear. (B1-B6) FLAG-nectin-1-EL cells; (B7-B12) FLAG-nectin-1-ΔC-EL cells; (B1 and B4) FLAG-nectin-1; (B7 and B10) FLAG-nectin-1-ΔC; (B2 and B8) E-cadherin; (B5 and B11) l-afadin; (B3, B6, B9, and B12) merge. Arrows, cell-cell AJs. Bars, 10 μm.
nates in the sequence, E/A-X-Y-V (X indicates W, V, and M for nectin-1, -2a, and -2b, respectively). Our finding is consistent with these earlier observations (Songyang et al., 1997), but we have shown here by the yeast two-hybrid assay that the PDZ domain of l-afadin does not bind to neu-rexin-2a which terminates in the sequence, E-Y-Y-V. A recent study of the third PDZ domain of PSD-95/SA P90 indicates that X residues at position -1 in the consensus sequence (X-S/T-X-V) and the upstream residues of the tetrapeptide determine the specificity and affinity for the binding of the PDZ domain to its binding partner (Niet-hammer et al., 1998). By analogy, unique aromatic or hydrophobic X residues at position -2 in the sequence (E/A-X-Y-V), such as W, V, and M, may be necessary to bind to the PDZ domain of l-afadin. It is also possible that the upstream residues of the tetrapeptide are crucial for the specificity and affinity for the PDZ domain. It has been shown recently that the PDZ domain of A-F-6 (s-afadin) binds to neu-rexin as well as the Eph receptor tyrosine ki-nase family members (Hock et al., 1998), but this result is not consistent with ours and the reason for this discrepancy is not known at present.

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Figure 8. Cell aggregation activity of nectin. (A) Ca2+-independent aggregation activity of nectin-1 and -2a. L cells stably expressing nectin-1 or nectin-2a were treated with trypsin in the presence of EDTA and then dispersed by pipetting to obtain a single-cell suspension. Each single-cell suspension was rotated in HBSS in the presence of 1 mM CaCl2 or 1 mM EDTA for 15, 30, and 60 min. The extent of aggregation of cells was represented by the ratio of the total particle number at time t of incubation (Nt) to the initial particle number (No). (A1) Nectin-1-L cells. (Filled circles and open circles) Wild-type L cells; (filled triangles and open triangles) nectin-1-L cells. (A2) Nectin-2a-L cells. (Filled circles and open circles) Wild-type L cells; (filled triangles and open triangles) nectin-2a-L cells. These results are representative of three independent experiments.

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