RESEARCH ARTICLE

Evaluating human cancer cell metastasis in zebrafish

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Abstract

Background: In vivo metastasis assays have traditionally been performed in mice, but the process is inefficient and costly. However, since zebrafish do not develop an adaptive immune system until 14 days post-fertilization, human cancer cells can survive and metastasize when transplanted into zebrafish larvae. Despite isolated reports, there has been no systematic evaluation of the robustness of this system to date.

Methods: Individual cell lines were stained with CM-Dil and injected into the perivitelline space of 2-day old zebrafish larvae. After 2-4 days fish were imaged using confocal microscopy and the number of metastatic cells was determined using Fiji software.

Results: To determine whether zebrafish can faithfully report metastatic potential in human cancer cells, we injected a series of cells with different metastatic potential into the perivitelline space of 2 day old embryos. Using cells from breast, prostate, colon and pancreas we demonstrated that the degree of cell metastasis in fish is proportional to their invasion potential in vitro. Highly metastatic cells such as MDA231, DU145, SW620 and ASPC-1 are seen in the vasculature and throughout the body of the fish after only 24–48 hours. Importantly, cells that are not invasive in vitro such as T47D, LNCaP and HT29 do not metastasize in fish. Inactivation of JAK1/2 in fibrosarcoma cells leads to loss of invasion in vitro and metastasis in vivo, and in zebrafish these cells show limited spread throughout the zebrafish body compared with the highly metastatic parental cells. Further, knockdown of WASF3 in DU145 cells which leads to loss of invasion in vitro and metastasis in vivo also results in suppression of metastasis in zebrafish. In a cancer progression model involving normal MCF10A breast epithelial cells, the degree of invasion/metastasis in vitro and in mice is mirrored in zebrafish. Using a modified version of Fiji software, it is possible to quantify individual metastatic cells in the transparent larvae to correlate with invasion potential. We also demonstrate, using lung cancers, that the zebrafish model can evaluate the metastatic ability of cancer cells isolated from primary tumors.

Conclusions: The zebrafish model described here offers a rapid, robust, and inexpensive means of evaluating the metastatic potential of human cancer cells. Using this model it is possible to critically evaluate whether genetic manipulation of signaling pathways affects metastasis and whether primary tumors contain metastatic cells.

Keywords: Cancer cells, Zebrafish, Metastasis, Invasion, Mouse
Background
Metastasis is the primary cause of human cancer mortality, accounting for >90% of deaths due to cancer [1]. There is now abundant evidence that, independent of the process of cellular transformation, the metastasis phenotype is genetically controlled [2]. Metastasis is a multistep process that involves local tumor invasion followed by dissemination to, and re-establishment at, distant sites. Families of genes have been described which have no effect on cell proliferation but which can suppress or promote metastasis [3,4]. Thus, targeting which have no effect on cell proliferation but which can suppress or promote metastasis [3,4]. Thus, targeting metastasis may prove to be effective in reducing cancer mortality if specific targets can be identified that suppress or promote metastasis [3,4]. Thus, targeting metastasis may prove to be effective in reducing cancer mortality if specific targets can be identified that suppress this phenotype. Here, we present a robust in vivo system for rapidly and accurately evaluating the effectiveness of candidate suppressor molecules.

Much of the analysis of metastasis pathways is conducted in tightly controlled in vitro cell systems, usually involving overexpression or ablation of a particular gene. Assays such as wound healing, transwell motility, invasion assays and hanging drop assays have been developed which provide readouts of cellular phenotypes related to metastasis [5-7]. These assays, however, do not address the issue of intravasation of tumor cells into blood vessels and extravasation into distant organs, a process requiring an in vivo assay system. Typically, such assays are performed in mice using experimental or spontaneous metastasis models [8,9]. While it is ultimately necessary to demonstrate that a pathway identified in vitro also affects invasion and metastasis in vivo, mouse models have significant drawbacks: 1) it is difficult to study early stages of the process where it is necessary to rapidly evaluate whether a particular drug or genetic manipulation has affected the metastasis phenotype, 2) evaluating the complete process in mice can require up to 6 months (depending on the cell system), 3) these experiments are expensive, immunsuppressed mice are required to study human cells and per diem charges in barrier facilities are costly, 4) in vivo imaging of small metastatic lesions is not possible in the deep tissues of the mouse, thus typically requiring termination and autopsy, thus extrapolation across experimental populations to realize the result, 5) popular immunosuppressed mice such as, nude (nu/nu), the severe combined immunodeficiency (SCID), or mice null for the recombination activating gene (Rag), have residual immune competence, which can actually prevent metastasis and, 6) the cohort size in these experiments is often pragmatically limited by high costs, thus statistical verification of metastasis modulation cannot be adequately assessed when the effect is mild.

Zebrafish provide an experimentally and genetically tractable animal model of a wide variety of human diseases [10]. Recent studies have demonstrated that zebrafish form spontaneous tumors with similar histopathological and gene expression profiles as human tumors [11-13]. The zebrafish-cancer model overcomes the drawbacks of murine xenograft models and offers alternative options for studying human tumor angiogenesis and metastasis [14-21]. Following early reports of the application of zebrafish to evaluate metastasis [22], we now tested whether metastasis in fish faithfully reports the metastatic potential of a broad range of cancer cells. To do so, we correlated in vitro invasion efficacy to in vivo metastasis metrics following manipulation of the metastatic phenotype. Without exception, we show that gene manipulations that affect in vitro invasion, alter metastasis in fish in a corresponding manner, demonstrating that the zebrafish is a tractable model to assay metastatic potential of human cancer cells. We also show that primary human cancer cells can metastasize in fish and that this ability can be used to predict metastatic potential in a clinical setting.

Results
The endogenous metastasis phenotype of human cancer cells is maintained in zebrafish
We first investigated whether human cancer cells, with known invasion/metastasis potential, could disseminate throughout the zebrafish body. To minimize the possibility that cells were introduced directly into the vasculature in error during the injection process, the fish were examined after 12 hours and those showing cells already in the vasculature were removed from further analysis. MDA-MB-231 breast cancer cells, for example, are highly invasive in vitro and metastasize in experimental and spontaneous murine models in vivo [8]. CM-Dil stained MDA-MB-231 cells were injected into the perivitelline cavity of 48 hours post fertilization (hpf) embryos and analyzed at 30 hours post injection (hpi) using confocal microscopy, where the spread of cancer cells could be seen throughout the body of the fish. In contrast, the T47D breast cancer cells, which do not typically invade or metastasize [7], did not show evidence of metastasis in the fish (Figure 1a). In parallel, the invasion potential of these cell lines was assessed in vitro using standard transwell assays, which was concordant with their ability to metastasize in the fish (Figure 1a). During our analysis of non-metastatic cell lines (e.g. T47D and (see below) HT29), we noted that, although the majority of fish did not show any disseminated cells within the fish body after the first 48 hour period, there were infrequent cases where cells were detected in the body of the fish. Quantitation of these cells revealed that in these cases there were never more than 5 cells outside the yolk sac region. We therefore set the cut off of >5 cells to indicate the definition of metastasis. Where dissemination was seen for the highly metastatic cells, however, there were significantly more than 5 cells within the fish body.
We have also shown previously that DU145 prostate cancer cells invade in vitro and metastasize in vivo in mouse models [9] compared with LNCaP cells which do not. As shown in Figure 1b, DU145 cells metastasize throughout the body of the fish at 30 hpi but LNCaP cells do not. The same correlation was seen for invasive SW620 and non-invasive HT29 colon cancer cells (Figure 1c), where again the correlation between invasion and metastasis in the fish was observed. Finally we studied the pancreatic ductal adenocarcinoma cells AsPC-1 and BxPC3 (Figure 1d). In this case, the AsPC-1 cells showed higher invasion potential than BxPC3 in the fish model which was consistent with in vitro invasion assays (Figure 1d). The extent of metastasis in fish for the highly invasive cell lines was readily apparent with large numbers of cells throughout the body of the fish (>25) after 24–48 hours. In contrast, the cells with low metastatic potential rarely showed cells in the body of the fish. These correlations were consistent within the cohort of fish used for each experiment. To evaluate the metastatic potential, therefore, we determined the number of fish that showed metastasis in the four different cancer cell systems, compared with the number that did not. In the highly invasive cell lines,
MDA231 (n = 58), DU145 (n = 72), SW620 (n = 63) and ASPC-1 (n = 57), metastasis was seen in 97%, 89%, 92% and 84% of fish respectively (Figure 1e). In contrast, in the poorly invasive cells, T47D (n = 64), LNCaP (n = 53), HT29 (n = 84) and BxPC3 (n = 42) metastasis was seen in only 9%, 15%, 14% and 26% of fish respectively (Figure 1e). We then performed quantitative invasion assays (see Methods) for all of these cell lines (Figure 1e) where the relative proportion of invading cells mirrored the distribution of metastasis in the in vivo fish model.

To demonstrate that cells from the perivitelline cavity could intravasate into the circulation and extravasate into the fish body, we used the Tg(kdrl:EGFP) transgenic zebrafish which highlights the vasculature (Figure 1f). In this study, using invasive MDA231 cells, for example, we could clearly see CM-Dil-labeled human cancer cells both in the vasculature and in the body of the fish adjacent to the vasculature (Figure 1f). This analysis demonstrated that the human cancer cells showed the range of phenotypes associated with metastasis. In contrast, the non-invasive T47D cells, were never seen in the vasculature of the host fish, nor in tissues distant from the injection site (Figure 1a). Thus, these data suggest that zebrafish can robustly report metastasis potential of different types of human cancer cells.

Assessment of the metastatic potential of primary human cancer cells in zebrafish

In the experiments described above, we have demonstrated that cancer cells with different in vitro invasion phenotypes show a parallel metastasis phenotype in zebrafish in all cases examined. These observations raised the issue whether metastatic potential in primary human cancer cells could also be assessed in this system. In a pilot proof-of-principle study, therefore, we prepared primary cultures of human lung cancer cells and maintained them for 10–15 days to collect sufficient cells for injection. Cells from only two different primary cultures were available to us and these were injected into zebrafish as described above and at 60 hpi tumor cell spread was assessed. The cells from tumor #8 showed a clear dissemination in the fish body, comparable with other highly invasive cell lines described above (Figure 2a). On the other hand, the cells from tumor #9 did not show any dissemination (Figure 2a). Histopathology of the primary tumors in these two cases, as part of routine diagnosis, showed that tumor #8 was derived from an undifferentiated squamous cell carcinoma, whereas tumor #9 was a well-differentiated adenocarcinoma. Quantitation of metastasis as defined above showed 81% (n = 42) of fish showed metastasis for cells from tumor #8 compared with only 7% (n = 57) for cells from tumor #9 (Figure 2b). Transwell invasion assays for cells from these primary cultures demonstrated that tumor #8 was highly invasive but that tumor #9 was not (Figure 2b). Thus, the relationship between invasion in vitro and metastasis in vivo in zebrafish is maintained in this analysis. Taken together, our data suggest that zebrafish may be a reliable in vivo model for assessment of the metastatic potential of primary human cancer cells.

The genetic regulation of tumor metastasis is maintained in zebrafish

WASF3 (or WAVE3) is a member of the Wiskott-Aldrich syndrome protein family. The structural motifs in the WASF3 protein predict that it orchestrates the reorganization of the actin cytoskeleton, which leads to the development of invadapodia and lamellipodia which facilitate invasion [4,23]. We have previously shown that WASF3 is highly expressed in primary human cancer and fundamentally important for metastatic spread of cancer cells [7-9,24,25]. To relate metastasis in fish with metastasis in mice, we analyzed DU145 cells in both models. As shown in Figure 3a-b, the control DU145 cells (shGFP) which have high WASF3 expression levels were highly invasive in vitro. Consistently, the control DU145 cells (shGFP) showed metastasis in the fish model at 60 hpi, whereas the stable WASF3-deficient cells (shW3-1 and shW3-2) show significantly reduced metastasis (Figure 3c). Quantitation of metastasis as defined above showed 94% (n = 51) of fish showed metastasis for control cells (shGFP) compared with only 12% (n = 72, shW3-1) and 18% (n = 66, shW3-2) for WASF3 knockdown cells.
cells (Figure 3d). In a mouse model of metastasis (Figure 3e), lung surface tumors and large tumor foci throughout the lungs were seen after 3 months, while the WASF3 knockdown cells clearly showed a significant reduction in both invasion potential or metastasis capability (no detectable tumors on the lung surface and very small tumor foci throughout the lung). Thus, assays evaluating metastatic potential of cells are consistent between mouse and zebrafish.

Quantitation of metastasis in zebrafish using Fiji algorithms

The transparent zebrafish embryos make imaging metastatic cells relatively straightforward using both conventional and confocal microscopy. Fish can be lightly anesthetized and photographed allowing repeat analysis of the same fish after different time intervals. Although the experiments described above show a clear distinction between the ability to metastasize and not, in certain circumstance it may be necessary to quantify the metastatic potential. To achieve this in a realistic time frame, we have modified the open source Fiji software [26] to quantitate the number of cells that have left the perivitelline space as a measure of metastasis. The photographs of the fish are used in this analysis and an example is shown in Figure 4a. In this system, the CM-Dil stained cells are imaged and counted throughout the fish. The cells remaining in the yolk sac are masked off and eliminated, and the number of cells in the fish body used as an estimate of the metastatic capability of the cells. In the example shown in Figure 4, MDA231 cells were imaged at 48 hpi (Figure 4b), where there was no significant difference in the number...
of metastatic cells (Figure 4c, left). At 120 hpi, however, there was a significant decline in the number of cells (Figure 4b-c). To streamline the study, we repeated the analysis where we counted only cells in the tail region, distal to the yolk sac (Figure 4b, right). Again, we saw no difference in the metastatic spread of the cells during early stages but saw a reduced number of cells at 120 hpi (Figure 4c, right). This analysis has advantages in that there is no need to assess the number of cells that remained in the yolk sac region or that had metastasized to the head region where the large area occupied by the eye presents challenges in quantitation.

Zebrafish show variation in metastasis potential related to tumor cell progression

To further evaluate the relationship between in vivo cell metastasis in zebrafish and in vitro cell invasion through matrigel, we studied the MCF10A system described by Tang et al. [27] where HRAS oncogene transformation and in vivo selection identified cells that have undergone progressive development into highly metastatic cells. The parental MCF10A cells (M-I), which originated from normal breast epithelial cells have been immortalized, but not transformed, and these cells do not metastasize in fish at 60 and 120 hpi (Figure 5a-b).
MCF10A cells, in which the activated HRAS oncogene was expressed, are described as oncogenically initiated MCF10AT cells [28]. These cells were then passaged through mice (M-II) and can form simple ducts that can progress to benign hyperplastic lesion and occasionally carcinomas. When M-II cells were challenged in the zebrafish assay, they showed modest spread in the fish body (Figure 5a-b). Cells (M-III) that were selected from one of the carcinomas that arose from the M-II cells predominantly form well differentiated carcinomas in mice. When these cells were introduced into the zebrafish model, their low grade phenotype is reflected in the poor ability to metastasis (Figure 5a-b). During the development of these low grade tumors, however, variants arose that were more aggressive, formed carcinomas and had acquired a metastatic phenotype in mice. When these cells (M-IV) were injected into the fish model, they showed the highest level of metastasis (Figure 5a-b). We next quantified the metastatic cells in the tail region using Fiji as described above. Twelve fish per group were monitored and confocal images were taken at 60 and 120 hpi, respectively. Quantitative analysis showed that relative metastatic ability of these cells in fish was defined as M-IV > M-II > M-III > M-I (Figure 5c), which is consistent with findings from in vitro invasion assays (Figure 5d).

Overall, therefore, in this progression model of human cancer cells, the zebrafish model accurately reports the relative level of metastatic potential in the different cells.

Suppression of oncogenic kinases affects metastasis in zebrafish

One important application of a rapid and robust in vivo metastasis assay is the ability to quickly evaluate whether a particular genetic manipulation in cancer cells leads to loss of metastatic potential. JAK1 and JAK2, for example, have been implicated in promoting invasion and metastasis

Figure 5 Relative metastatic potential in human cancer cells during progression from low to high grade tumors. When immortal, non-transformed MCF10A cells (M-I) were introduced into the zebrafish metastasis model (a), no spread was seen in any of the fish analyzed at 60 hpi. The cells (M-II) recovered from low grade hyperplastic lesions in mice from MCF10AT cells (MCF10A cells transformed with activated HRAS) show increased metastatic potential (arrows). M-III cells, which were derived from well differentiated carcinomas derived from M-II cells, show reduced metastatic potential, whereas M-IV cells, recovered from aggressive adenocarcinomas derived from M-II cells, show enhanced metastatic potential. The same relationship was seen at 120 hpi (b) (shown as z-stack images using 40x magnification). Quantitation of metastatic cells throughout the fish body using Fiji (c) confirms the relationship between tumor grade and metastatic potential in fish (left). At 120 hpi, although the relative proportion of metastatic cells is seen in the M-II/M-III/M-IV comparison, the overall number of cells in each tumor grade was reduced. When the same comparison was performed using the in vitro invasion assay (d), the relative proportion of invading cells in the different sub groups was again maintained. Data are presented as the mean of three independent experiments (n = 3) ± SEM, **P < 0.01.
in certain cell types such as 2C4 human fibrosarcoma cells, which are highly invasive (Figure 6a). When wild-type 2C4 cells were injected into zebrafish, they show metastasis after 24 hours, which is sustained over a 5 day period (Figure 6b). The derivative U4C cells, which are genetically deficient for JAK1 [29,30], failed to spread throughout the fish body and showed virtually no invasion in vitro (Figure 6a). In fact, not a single cell was seen in the fish body, but sustained cell masses were seen in the perivitelline space demonstrating the persistence of viable cells in the fish over this period. In the derivative JAK2-deficient γ2A fibrosarcoma cells [31,32], low-level metastasis was still seen but significantly suppressed compared with wild-type 2C4 cells (Figure 6b). Again, the quantitation analysis using Fiji algorithms showed the relative metastasis potential of these three different cell lines in the zebrafish reflected in vitro invasion assays (Figure 6c).

**Discussion**

Dissection of the functional aspects of genes that impact the metastasis phenotype requires a robust assay for tumor spread. While it is accepted that, in the final analysis, rodent models should be used to evaluate metastasis, this approach is costly and inefficient as an up-front assay to determine whether a particular genetic manipulation affects the metastasis phenotype. The spontaneous metastasis assay [8,9] has shortcomings, since it cannot be used to evaluate intravasation into the blood vessels. The zebrafish model, on the other hand provides a solution to the time-consuming and costly mouse experiments, since in many cases the assay can be performed within 24–36 hours of xenotransplantation, in large cohorts of fish, providing statistical power to the results. Although we have only studied cancer cells from breast, pancreas, colon and sarcomas thus far, in all cases in vitro invasion ability correlated with the metastatic potential of tumor cells to spread in vivo. Importantly, we have shown that genetic manipulations of human cancer cells which affect invasion, also affect metastasis in fish. Although the technical dexterity needed to inject 48 hpf zebrafish can be demanding, the absence of an adaptive immune response for the first 14 days post fertilization (dpf) [33] avoids side effects and dosing issues related to using immunosuppressants [34,35]. Early studies targeting the yolk sac as a site of injection truly challenged cancer cells to enter the blood stream. The blood supply to the yolk sac is extensive since this sustains the fish for the first 5 dpf and maximizes the opportunity for intravasation. It has been shown using the cloche mutant fish, which do not develop a vasculature or circulation, that metastatic human cells injected into the yolk sac cannot metastasize in these fish, demonstrating the requirement for a functional circulatory system in this process [15]. Injection into the yolk sac, however, has complications apparently associated with poor resealing of the yolk sac membrane which leads to spillage of the cancer cells or yolk sac contents. The perivitelline space between the body of the fish and the yolk sac provides an alternative, which does not suffer from these associated problems. The technical challenge is successfully targeting the perivitelline space, and avoiding injection directly into the circulatory system. For this reason we examined fish after 12 hours for the presence of cells in the vasculature and excluded these fish from the analysis. Since the injection involves large numbers of fish, excluding those that have been compromised during the injection process does not have any impact on the final analysis.

Although zebrafish have been used as a model for metastasis previously, protocols between different groups were not consistent in terms of the number of cells injected, the site of injection, the age of the fish used and

![Figure 6](http://www.biomedcentral.com/1471-2407/13/453)
the method of quantitation of metastasis [18,22]. To evaluate this metastasis model more robustly, we have used a standardized protocol with a variety of different cancer cells and cell systems. In this report we clearly demonstrate that metastasis in the zebrafish correlates with in vitro invasion assays and in one case (DU145 cells, Figure 3) with metastasis potential in murine models of metastasis. We observed that metastatic spread in the fish was achieved as early as 24 hpi, and possibly sooner, and that the maximum tumor spread was achieved within 48 hours in most cases, without a significant increase over subsequent days. A reduction in the numbers of cancer cells, however, occurred when the analysis was extended to 5 dpi. The metastasis assay, however, if initiated at 2 dpi, can be completed before it is necessary to feed the fish (4 dpi) and, at least in the cells we tested, do not need to be followed for more that 2–3 days to evaluate metastasis. In prior studies, the number of cells injected into the fish varied between 50–2000 [14-17,19-22]. We observed that injecting too many cells can lead to mortality in our system and, following preliminary evaluation of the optimal number to demonstrate metastasis, we consistently injected cells ~300 cells per fish.

In many of our studies, the difference in the number of disseminated cells between the metastatic and non-metastatic tumors was usually striking. The same was observed in experimental cell systems where inactivation of a particular gene led to almost complete loss of invasion or metastasis, e.g. the JAK1 deficient 2C4 cells or the DU145 WASF3 knockdown cells. In these experiments, however, cells were seen outside the yolk sac region, which raised the issue of how metastasis is defined? In cell lines such as T47D, LNCAP and HT29, which are generally considered non-metastatic, our analysis showed that, even if there were disseminated cells, in the majority there were usually only between 1–5 cells in the body of the fish, where 5 cells was the exception. There are several reasons why small numbers of cells may appear in the fish body. Firstly, on rare occasions, the injection procedure could have inadvertently penetrated the vasculature and cells were introduced directly, although we screened all fish 12 hours after injection and excluded any that already showed cells outside the yolk sac region to overcome this being a major factor in the analysis. In practice this was only a very small number (<2%) for each cohort. It is also possible that cells defined as non-metastatic, are in fact weakly metastatic, and so occasional cells will disseminate into the fish. This is particularly true in experimental systems when, for example, shRNA knockdown of a particular gene is not complete, leaving some cells with gene expression levels above the threshold that will allow metastasis. It is important to note, however, that in many of our experiments involving apparently non metastatic cells, the presence of >5 disseminated cells was only seen in a minority of fish in the cohort. The main criterion for metastasis, therefore, is the presence of >5 cells in the majority of fish. In practice, however, the numbers of metastatic cells throughout the various cohorts for metastatic cells was far greater than 5 as shown in Figure 4 (35–55 cells after 48 hours). The number of disseminated cells becomes particularly important when defining relative metastatic potential, as seen in the MCF10A continuum. In this case the overall number of cells found outside the yolk sac area correlated with the invasiveness of the cells in vitro. We expect, however, most metastasis assays will want to determine whether ablation or overexpression of a gene leads to changes in metastatic potential, and following the protocol described here will facilitate this determination.

Although we were limited by the number of clinical samples available to us, the demonstration that primary human cancer cells can survive, grow, and metastasize in zebrafish provides a very encouraging proof-of-principle and opens opportunities to evaluate the metastatic potential in primary cells from biopsies or following surgery, which can have important advantages for clinical management of the patient. Even though it may take 2–3 weeks to establish the primary cell cultures, the presence of metastatic cells in a tumor for which there is a well-differentiated histopathology, could affect the future screening and management protocol. In addition, the well-established systems [36-38] for drug screening in zebrafish, opens up the possibility of identifying therapeutics that can target metastasis on a tumor-by-tumor basis, so providing a personalized approach to individual tumors.

Conclusions
In summary, we provide a side-by-side critical evaluation showing that zebrafish can evaluate the metastatic potential of human cancer cell lines and primary tumors. The ability to perform these analyses in large cohorts of fish allow for robust statistical analysis in a short time frame (24–120 hours) and provides a rapid means of evaluating whether genetic manipulation of cells, or cell origin, affects metastasis in vivo. Where it has been investigated, the metastasis phenotype in fish is identical to that in rodents with the advantages that individual cells can be imaged during the metastasis process.

Methods
Cell culture and primary lung cancer cell isolation
Human breast cancer cells (MDA231 and T47D), prostate cancer cells (DU145 and LNCaP), pancreatic ductal adenocarcinoma cells (AsPC-1 and BxPC3) and colon cancer cells (SW620 and HT29) were obtained from the American Type Culture Collection (ATCC, Rockville,
MD, USA). The parental human fibrosarcoma cell line 2C4, the JAK1-deficient U4C and JAK2-deficient y2A derivatives were a gift from Dr. Ivo P. Touw (Erasmus University Medical Center, The Netherlands) and were cultured in DMEM supplemented with 10% (v/v) fetal bovine serum (FBS). The normal MCF10A breast epithelial cell line (M-I) and its derivatives M-II, M-III and M-IV [27,39], were a gift from Dr. Shuang Huang (Georgia Regents University, USA). The M-III and M-IV cells were maintained in DMEM/F-12 supplemented with 5% horse serum. The culture medium for the M-I and M-II was similar but also included 20 ng/ml recombinant human EGF (R&D Systems, Minneapolis, MN, USA), 0.5 μg/ml hydrocortisone (Sigma-Aldrich, St. Louis, MO, USA), 10 μg/ml insulin (Sigma-Aldrich) and 100 ng/ml cholera toxin (Sigma-Aldrich). Primary lung cancer cells were obtained immediately after resection, disaggregated with trypsin and cultured in DMEM for 5–10 days.

In vitro invasion assay and experimental metastasis mouse model
To measure cell invasion potential, matrigel invasion assays were performed as described previously [7,25] using Transwells (BD biosciences, San Diego, CA, USA) with 8-μm pore size filters. The invading cells were fixed in 3.7% paraformaldehyde and stained with 0.5% crystal violet in 2% ethanol. The lower surface of the filter was photographed and the invading cells were counted from six fields at 200× magnification. Each experiment was performed in triplicate on at least three occasions. For experimental metastasis, 6-week-old male SCID mice were injected with 1×10⁶ WASF3 knockdown DU145 cells or the knockdown control cells through the lateral tail vein [9]. Mice were sacrificed 3 months after injection and the lung tissues were processed for hematoxylin and eosin (HE) staining. All experimental procedures were approved by the Animal Care and Use Committee of Georgia Regents University.

Cell preparation and transplantation
For cell labeling, cells were incubated with cell tracker CM-Dil (Invitrogen, Carlsbad, CA, USA) at a final concentration of 2.5 μg/ml for 4 min at 37°C followed by 15 min at 4°C. To remove unincorporated dye, cells were rinsed twice with phosphate-buffered saline (PBS), and then resuspended at a higher concentration (5×10⁶ cells/ml). Before injection, CM-Dil labeled cells were assessed for viability using trypan blue exclusion and only samples in which there was >90% viability were used. We also evaluated the cells for uniform red staining and membrane integrity using a Zeiss Axiovert microscope (Zeiss, Thornwood, NY, USA) before being transplanted into the fish.

Zebrafish husbandry and the metastasis model
Zebrafish were maintained using established temperature and light cycle conditions as previously described [40,41]. All experimental procedures were approved by the Animal Care and Use Committee of Georgia Regents University. The Tg(kdrl:EGFP) transgenic fish line was a gift from Dr. Daniel Wagner (Department of Biochemistry and Cell Biology, Rice University, USA). For zebrafish xenotransplantation, 48 hpf wild-type AB or Tg(kdrl:EGFP) strains of transgenic zebrafish embryos were dechorionated and anaesthetized in 0.3× Danieau’s solution containing phenthiourea (PTU, Sigma-Aldrich) and 0.04 mg/ml tricaine (Sigma-Aldrich) before human cell injection. Approximately 300 CM-Dil labeled human cells were injected into the perivitelline cavity of each embryo, and zebrafish were maintained in 0.3× Danieau’s solution containing PTU for 1 h at 28°C. After confirmation of a visible cell mass at the injection site, zebrafish were transferred to an incubator and maintained at 34°C.

Confocal imaging
Living zebrafish embryos were anesthetized using 0.04 mg/ml tricaine and were then embedded in a lateral orientation in 0.5% agarose. Serial sections were captured using an Olympus FLUOVIEW™ FV1000 laser scanning confocal microscope (Olympus, Tokyo, Japan) and 2.5 μm z-step intervals. Low magnification (×4 objective) was used to provide an overview of the tumor cell metastasis pattern throughout the fish, and higher magnification (×20 objective) was used to define the precise localization of metastatic cells and foci within the zebrafish. Z-stack images were processed using ImageJ/Fiji as previously described [40,42].

Automated cell counting and statistical analysis
The custom Fiji (ImageJ2) software package [26] was used for automated cell counting. Briefly, a 190–255 intensity threshold was set to select cells and the ‘analyze’ particle tool was used with default selection of cell size and cell shape during counting. A Fiji macro was generated using the ‘record’ function to streamline analyses and remove bias. To count the metastatic cell number in the tail, the fish tails were selected for cell counts using the polygon selection tool. A P-value of 0.05 or less was considered to be statistically significant and determined by the Student’s t-test. Values of three or more experiments were given as mean ± SEM.

Competing interests
The authors declare no competing financial interests.

Authors’ contributions
YT and JC designed the experiments; YT performed the molecular and cellular experiments; YT, XX and DW performed the fish husbandry and imaging, YT and SW performed the image analysis; JM provided input into
zebrafish experiments; YT, JC and JM wrote the manuscript. All authors read and approved the final manuscript.

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References
1. Siegel R, Naishadham D, Jemal A: Cancer statistics. Cancer J Clin 2012, 62:10–29.
2. Yokota J: Tumor progression and metastasis. Carcinogenesis 2000, 21:497–503.
3. Steeg PS, Ouatas T, Halverson D, Palmieri D, Salerno M: 3 Cancer Center, Georgia Regents University, Augusta, GA, USA. 2 Department of Cellular Biology and Anatomy, Georgia Regents University, Augusta, GA, USA. 3 Vision Discovery Institute, Georgia Regents University, 1120 15th Street, Augusta, GA, 30912, USA.

4. Sossey-Alaoui K, Li X, Ranalli TA, Cowell JK: WAVE3-mediated cell migration and lamellipodia formation are regulated downstream of phosphatidylinositol 3-kinase. J Biol Chem 2003, 278:21746–21755.
5. Kunapuli P, Chitta KS, Cowell JK: Suppression of the cell proliferation and invasion phenotypes in glioma cells by the LG1 gene. Oncogene 2003, 22:3985–3991.
6. Goodwin AM: In vitro assays of angiogenesis for assessment of angiogenic and anti-angiogenic agents. Microvasc Res 2007, 74:172–183.
7. Teng Y, Mei Y, Hawthorn L, Cowell JK: WASF3 regulates mit-200 inactivation by ZEB1 through suppression of KISS1 leading to increased invasiveness in breast cancer cells. Oncogene 2013, doi: 10.1038/onc.2012.565. [Epub ahead of print]
8. Sossey-Alaoui K, Safina A, Li X, Vaughan MM, Hicks DG, Bakin AV, Cowell JK: Down-regulation of WAVE3, a metastasis promoter gene, inhibits invasion and metastasis of breast cancer cells. Am J Pathol 2007, 170:2112–2121.
9. Teng Y, Ren MQ, Cheney R, Sharma S, Cowell JK: Inactivation of the WASF3 gene in prostate cancer cells leads to suppression of tumorigenicity and metastases. BJ Cancer Research 2010, 103:1066–1075.
10. Lieschke GJ, Currie PD: Animal models of human disease: zebrafish swim into view. Nat Rev Genet 2007, 8:353–367.
11. Amatruda JF, Shepard JL, Stern HM, Zon LI: Zebrafish as a cancer model system. Cancer Cell 2002, 1:229–231.
12. Bernghardt S, Jette C, Langenau D, Hsu K, Stewart R, Look T, Kanik JP: Making waves in cancer research: new models in the zebrafish. Biotechnology 2005, 23:227–237.
13. Lam SH, Gong Z: Modeling liver cancer using zebrafish: a comparative oncogenomics approach. Cell Cycle 2005, 4:573–577.
14. Lee LM, Selff EA, Bonde G, Cornell RA, Hendrix MJ: The fate of human malignant melanoma cells transplanted into zebrafish embryos: assessment of migration and cell division in the absence of tumor formation. Dev Dyn 2005, 233:1560–1570.
15. Marques U, Weiss FU, Veeken DH, Nitsche C, Bakkers J, Lagendijk AK, Partecke U, Heidecke CD, Lerch WM, Bajowska CP: Metastatic behaviour of primary human tumours in a zebrafish xenotransplantation model. BMC Cancer 2009, 9:128.
16. Topczewska JM, Postovit LM, Margaryan NV, Sam A, Hess AR, Wheaton WW, Nickoloff BJ, Topczewski J, Hendrix MJ: Embryonic and tumorigenic pathways converge via Nodal signaling: role in melanoma aggressiveness. Nat Med 2006, 12:925–932.
17. Lally BE, Geiger GA, Kridel S, Arcury-Quandt AE, Robbins ME, Kock ND, Wheeler K, Peddi P, Georgakias A, Kao GD, Kouremis P: Identification and biological evaluation of a novel and potent small molecule radiation sensitizers via an unbiased screen of a chemical library. Cancer Res 2007, 67:8791–8799.
18. Stoeletov K, Klemke R: Catch of the day: zebrafish as a human cancer model. Oncogene 2008, 27:4509–4520.
19. Lee SL, Rouhi P, Dahl Jensen L, Zhang D, J H, Hauptmann G, Ingham P, Cao Y: Hypoxia-induced pathological angiogenesis mediates tumor cell dissemination, invasion, and metastasis in a zebrafish tumor model. Proc Natl Acad Sci USA 2009, 106:14955–14960.
20. Zhang L, Zhou F, Drabch Y, Gao B, Snaar-Jagalska BE, Mickanin C, Huang H, Sheppard KA, Porter JA, Lu OK, ten Dijke P: USP4 is regulated by AKT phosphorylation and directly deubiquestinises TGF-B type 1 receptor. Nat Cell Biol 2012, 14:717–726.
21. Santahamar K, Judson EC, Elks PM, Mckee S, Elworthy S, van Rooijen E, Walmesley SS, Renshaw SA, Cross S, van Eeden FJ: A zebrafish model to study and therapeutically manipulate hypoxia signalling in tumorigenesis. Cancer Res 2012, 72:4017–4027.
22. Konantz M, Balci TB, Hartwig UF, Dellaire G, André MC, Berman JN, Lengerke C: Zebrafish xenografts as a tool for in vivo studies on human cancer. Ann N Y Acad Sci 2012, 1266:124–137.
23. Sossey-Alaoui K, Su G, Malaj E, Roe B, Cowell JK: WAVE3, an actin-polymyelization gene, is truncated and inactivated as a result of a constitutional t1;13 (q21;q12) chromosome translocation in a patient with ganglioneuroblastoma. Oncogene 2002, 21:5967–5974.
24. Teng Y, Liu M, Cowell JK: Functional interrelationship between the WASF3 and KISS1 metastasis-associated genes in breast cancer cells. Int J Cancer 2011, 129:2825–2835.
25. Teng Y, Ngoka L, Mei Y, Lessoon L, Cowell JK: HSP90 and HSP70 proteins are essential for stabilization and activation of WASF3 metastasis-promoting protein. J Biol Chem 2012, 287:10051–10059.
26. Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, Preibisch S, Rueden C, Saalfeld S, Schmid B, Tinevez JY, White DJ, Hartenstein V, Elciert K, Tomancak P, Cardona A: Fiji: an open-source platform for biological-image analysis. Nat Methods 2012, 9:676–682.
27. Tang B, Yu M, Booker T, Santner SJ, Miller FR, Arner MR, Wakefield LM: TGF-beta switches from tumor suppressor to prometastatic factor in a model of breast cancer progression. J Clin Invest 2003, 112:1116–1124.
28. Dawson PJ, Wolman SR, Talt L, Heppner GH, Miller FR: MCF10AT: a model for the evolution of cancer from proliferative breast disease. Am J Pathol 1996, 148:313–319.
29. Staerk J, Kallin A, Demoulin JB, Vainchenker W, Constantinescu SN: JAK1 and Tyk2 activation by the homologous polycythemia vera JAK2 V617F mutation: cross-talk with IgF1 receptor. J Biol Chem 2005, 280:41893–41899.
30. Flex E, Petrangeli V, Stella L, Chiarietti S, Homakova T, Knoops L, Ariola C, Fodale V, Clapper E, Paolini F, Martinelli S, Fragale A, Sanchez M, Tavolaro S, Messina M, Cazzaniga G, Camera A, Pizzolo G, Torinesello A, Vignetti M, Battistini A, Cavé H, Gef B, Renaud J, Brondi A, Constantinescu SN, Foni R, Tartaglia M: Somatically acquired JAK1 mutations in adult acute lymphoblastic leukemia. J Exp Med 2008, 205:751–758.
31. Burfoot MS, Rogers NC, Waterman DT, Smith JM, Pons S, Paonessaw G, Burfoot MS, Rogers NC, Watling D, Feng J, Guschin D, Briscoe J, Hartenstein V, Eliceiri K, Tomancak P, Cardona A: Fiji: an open-source platform for biological-image analysis. Nat Methods 2012, 9:676–682.
32. Witthuhn BA, Kotenko SV, Pestka S, Stark GR, Ihle JN: Activation of JAKs by the homologous polycythemia vera JAK2 V617F mutation: cross-talk with the IgF1 receptor. J Biol Chem 2005, 280:41893–41899.
33. Kohlhauer F, Rogers NC, Waterman DT, Feng J, Guschin D, Briscoe J, Witthuhn BA, Kotenivo SV, Petesta S, Stark GR, Hill JN, Kerr MW: A JAK1/JAK2 chimera can sustain alpha and gamma interferon responses. Mol Cell Biol 1997, 17:895–906.
34. Traver D, Herbtomel P, Patton EE, Briscoe J, White MF, Kerr IM: Janus kinase-dependent activation of the interferons. J Biol Chem 1997, 272:24183–24190.
35. Rubinstein AL, Doan TN, Dingledine R, Sandberg EM: Automated, quantitative screening assay for antiangiogenic compounds using transgenic zebrafish. Cancer Res 2007, 71:1138–11392.
36. Vomir M, Huang W, Yussuf S, Shim JS, Ferrante AA, Liu JC, Parsons MJ: Chemical screen identifies FDA-approved drugs and target pathways.
that induce precocious pancreatic endocrine differentiation. *Proc Natl Acad Sci* 2011, 108:19264–19269.

38. Etchin J, Kanki JP, Look AT. Zebrafish as a model for the study of human cancer. *Methods Cell Biol* 2011, 105:309–337.

39. Soule HD, Maloney TM, Wolman SR, Peterson WD Jr, Brenz R, McGrath CM, Russo J, Pauley RJ, Jones RF, Brooks SC. Isolation and characterization of a spontaneously immortalized human breast epithelial cell line, MCF-10. *Cancer Res* 1990, 50:6075–6086.

40. Teng Y, Xie X, Walker S, Rempala G, Kozlowski DJ, Mumm JS, Cowell JK. Knockdown of zebrafish Lgi1a results in abnormal development, brain defects and a seizure-like behavioral phenotype. *Hum Mol Genet* 2010, 19:4409–4420.

41. Teng Y, Xie X, Walker S, Saxena M, Kozlowski DJ, Mumm JS, Cowell JK. Loss of zebrafish Lgi1b leads to hydrocephalus and sensitization to pentylenetetrazol induced seizure-like behavior. *PLoS One* 2011, 6:e24596.

42. Ariga J, Walker SL, Mumm JS. Multicolor time-lapse imaging of transgenic zebrafish: visualizing retinal stem cells activated by targeted neuronal cell ablation. *J Vis Exp* 2010, 43:e2093.

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