The placenta shed from goats with classical scrapie is infectious to goat kids and lambs

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The placenta of domestic sheep plays a key role in horizontal transmission of classical scrapie. Domestic goats are frequently raised with sheep and are susceptible to classical scrapie, yet potential routes of transmission from goats to sheep are not fully defined. Sparse accumulation of disease-associated prion protein in cotyledons casts doubt about the role of the goat’s placenta. Thus, relevant to mixed-herd management and scrapie-eradication efforts worldwide, we determined if the goat’s placenta contains prions orally infectious to goat kids and lambs. A pooled cotyledon homogenate, prepared from the shed placenta of a goat with naturally acquired classical scrapie disease, was used to orally inoculate scrapie-naïve prion genotype-matched goat kids and scrapie-susceptible lambs raised separately in a scrapie-free environment. Transmission was detected in all four goats and in two of four sheep, which importantly identifies the goat’s placenta as a risk for horizontal transmission to sheep and other goats.

The transmissible spongiform encephalopathies (TSE) are a heterogeneous group of disorders differing in aetiology, pathology, host range, strain repertoire and efficient transmission routes. The transmissible agent, the prion, is unique amongst infectious disorders in that it is widely believed to principally consist of a misfolded protease-resistant isoform of a host-encoded protein, the prion protein (PrP) (Caughey et al., 2009; Soto, 2011). Classical scrapie is a prion disease of domestic sheep (Ovis aries) that causes significant economic burden to sheep industries worldwide. Scrapie eradication programmes are largely based on the observation that classical scrapie is efficiently transmitted through contact with the placenta shed by infected ewes (Pattison et al., 1972) and that susceptibility is limited by polymorphisms in the prion protein gene, PRNP (Goldmann, 2008). Eradication programmes focused on such factors have resulted in dramatic decreases in disease prevalence, but eventual eradication could be delayed or infection reintroduced if other reservoirs of scrapie prions are not identified and managed.

Classical scrapie also affects domestic goats (Capra hircus) (González et al., 2010; Hadlow et al., 1980; Konold et al., 2010; Pattison & Millson, 1961) but much less is known about its pathology, including potential routes of transmission to sheep and other goats. In sheep, the shed placenta contains abundant accumulation of disease-associated misfolded prion protein (PrPSc) (Andreóletti et al., 2002; Lacroux et al., 2007; Tuo et al., 2001, 2002), is infectious (Onodera et al., 1993; Pattison et al., 1972, 1974; Race et al., 1998), plays a key role in horizontal transmission (Hoinville et al., 2010; Touzeau et al., 2006) and contributes to environmental contamination (Andreóletti et al., 2002; Gough & Maddison, 2010). Since sheep and goats are sometimes co-housed during parturition, and since goats have been used as surrogate dams for orphaned lambs, one of the most likely scenarios for scrapie transmission from goats to sheep is during the post-partum period through oral exposure to parturient material. Given the sparse accumulation of PrPSc in the placenta of goats as compared to sheep (O’Rourke et al., 2011), intra- and interspecies transmission by this route is not certain. The primary purpose of this experiment was to determine if classical scrapie in goats (caprine scrapie) is orally transmissible to other goats and sheep via the placenta.

All animal use and care was approved by the Washington State University Institutional Animal Care and Use Committee. The experiment was conducted using a previously described source of tissue (O’Rourke et al., 2011). In brief, the placenta donor goat (3950; Nubian) originated from a naturally infected US goat herd [herd 2,
O’Rourke et al. (2011) reported to be without direct contact with sheep for at least 5 years and located on premises without prior history of scrapie disease. At the time of regulatory investigation, goat 3950 was not clinical but was determined to be scrapie-infected through antemortem rectal biopsy testing and standard scrapie immunohistochemistry (IHC). The goat was acquired at 20 months of age and by 34 months of age began showing clinical signs of classical scrapie disease including progressive weight loss, truncal scratching and ataxia. At 35 months of age, the goat gave birth to three kids and the placenta (G797) was immediately collected for processing. The donor goat was euthanized at 38 months of age for humane reasons associated with progression of scrapie disease. All three kids associated with this placenta developed clinical scrapie disease by 2.5 years of age. Additionally, the donor goat and all three kids had subclinical infection with small ruminant lentivirus (SRLV) as determined by repeated serological testing as conducted by the Washington Animal Disease Diagnostic Laboratory (Pullman, WA, USA) using competitive inhibition ELISA (Small Ruminant Lentivirus Antibody Test kit, cELISA; VMRD) (Herrmann et al., 2003a, b). The PRNP genotypes of all animals in this study were determined by DNA sequence analysis as previously described (O’Rourke et al., 2011). Donor goat 3950 and all three kids associated with this placenta were heterozygous for the central caprine PRNP haplotypes 1 and 2 (White et al., 2008), which only differ at codon 240 [respectively, proline (P) and serine (S)]. Both haplotypes are associated with susceptibility to classical scrapie disease (Vaccari et al., 2009).

All cotyledons from one fetal unit of placenta G797 were pooled, stored for 2 years at −80 °C, and then homogenized just days prior to use as inoculum. Pooled cotyledons (116 g total wet weight) were homogenized in a new Oster blender using the setting ‘mix’, first for 5 min neat and then as a final 83 % (w/v) homogenate in PBS for 2 min. Aliquots (~4 ml each) of the G797 cotyledon homogenate were briefly stored at −20 °C before use. As previously described (O’Rourke et al., 2011), accumulation of disease-associated PrP in G797 cotyledon homogenate was determined by Western blot analysis using mAb F99/97.6.1 and by scrapie ELISA (HerdChek CWD Ag Test; IDEXX Laboratories). As seen by Western blot (Fig. 1a), typical proteinase K-resistant PrP (PrP(res)) bands were readily detected in the obex hindbrain of donor goat 3950 (ob, lane 1; loading 450 μg tissue wet weight equivalent) and in a sodium phosphotungstic acid (Na-PTA) extract of 90 mg tissue wet weight equivalent of G797 cotyledon homogenate (cot, lane 2). No PrP(res) bands were observed in a Na-PTA extract of 90 mg tissue wet weight equivalent of a similarly prepared cotyledon homogenate derived from the shed placenta of scrapie-unexposed goat 4113 (Fig. 1a: cot, lane 3); this goat and cotyledon homogenate were also heterozygous for caprine PRNP haplotypes 1 and 2. Determination of PrPSc content in the obex and G797 cotyledons from goat 3950 was by scrapie ELISA using twofold serial dilutions of neat homogenates (Fig. 1b). The tissue equivalents loaded into each assay well were expressed in terms of total protein (BCA

![Fig. 1. Comparison of the disease-associated prion protein content in the hindbrain at the level of the obex and in shed cotyledons from donor goat 3950. (a) Proteinase K-resistant prion protein (PrP(res)) bands were detected by Western blot analysis in the obex (ob, lane 1) and cotyledon (cot, lane 2; Na-PTA precipitate) homogenates derived from the scrapie-infected donor goat 3950 but not in similarly prepared PRNP genotype-matched cotyledon homogenate from the scrapie-naive donor goat 4113 (cot, lane 3; Na-PTA precipitate). (b) Scrapie ELISA titrations of disease-associated prion protein (PrPSc) in shed cotyledons (open diamonds) and obex (open circles) from donor goat 3950. PrPSc content is expressed as background-corrected arbitrary units of optical density (corrected OD); tissue equivalents loaded at each twofold serial dilution are expressed in terms of total protein content (x-axis scaling, log2). As determined per manufacturer’s directions, the scrapie ELISA cut-off sensitivity was 0.228 and is shown as a horizontal line.](http://vir.sgmjournals.org)
Recipient goats consisted of four Saanen kids born to scrapie-unexposed does. One goat kid was homozygous for caprine \(PRNP\) haplotype 1 (4471); the other three were heterozygous for haplotypes 1 and 2. Recipient sheep consisted of four white- or mottled-faced lambs born to scrapie-unexposed ewes. Recipient lambs were homozygous for the scrapie-susceptible ovine \(PRNP\) haplotype, which codes for valine at position 136 (i.e. VV\(136\)) (Goldmann, 2008). Kids and lambs were born in a holding facility in which scrapie-infected animals had never been housed. Newborn animals nursed colostrum for 48 h and were then moved and raised by hand in scrapie-unexposed isolation rooms, one for kids and one for lambs. Newborns were inoculated at 48–72 h of age via the oral route by placing a single, partially thawed aliquot of G797 cotyledon homogenate (~3.3 g wet tissue weight equivalent) near the back of the tongue, immediately after which kids nursed a bottle of fresh cow’s milk and lambs nursed a bottle of lamb’s artificial milk replacer. Kids and lambs were later weaned onto a balanced ration of grass and alfalfa hay with access to appropriate mineral supplements. Since these animals were raised indoors, each received subcutaneous injections of 75 000 IU vitamin D3 at approximately three-week intervals (Vitamin A D injection; Agri Laboratories).

As depicted in a timeline (Fig. 2a), scrapie infection status was monitored antemortem by biopsy of the rectoanal mucosa-associated lymphoid tissue (RAMALT) (González lymphoid tissue. Images in which Pr\(P^S\)c was not detected (left column) or was detected (right column) are presented for each recipient species (goat, top row; sheep, bottom row). Age at time of biopsy is given as days post-inoculation (dpi). Accumulation of Pr\(P^S\)c is relatively sparse in goat 4479 as compared with sheep 4442. A higher power inset (bar=20 \(\mu\)m) of the region of interest (white box) is provided for goat 4479. All other bars=200 \(\mu\)m. (c) Detection of proteinase K-resistant prion protein (Pr\(P^{Pros}\)) bands by Western blot after Na-PTA extraction of retropharyngeal (rp) and ileocecal (ic) lymph node homogenates. The typical three Pr\(P^{Pros}\) glycoforms are only evident in the lymph nodes of recipient sheep 4440 (rp, lane 2; ic, lane 5) but not recipient sheep 4445 (lanes 3 and 6) or 4450 (lanes 4 and 7). Assay controls included the Na-PTA extractions of a dilute scrapie-positive obex homogenate (positive control: ob, lane 1) and of a retropharyngeal lymph node from a scrapie uninfected sheep (negative control: rp, lane 8).
et al., 2005) and in sheep, also by biopsy of the nictitating membrane (biopsies 3 and 4) (O’Rourke et al., 2000). Scrapie IHC using monoclonal antibody F99/97.6.1 was applied to formaldehyde-fixed, paraffin-embedded tissues as previously described (O’Rourke et al., 2011). Antemortem lymphoid accumulation of PrPSc was detected in three of four recipient goats but in only one (sheep 4442) of four recipient sheep (Fig. 2a, 2b). One recipient sheep (4440) was euthanized at 747 days post-inoculation (p.i.) due to development of an abomasal emptying disorder. Although PrPSc was not detected by scrapie IHC in the obex or in any of the lymphoid tissues examined from this sheep (Table 1, example shown in Fig. 2b), PrPres accumulation was evident by Western blot analysis after Na-PTA extraction of retropharyngeal and ileocecal lymph node homogenates (Fig. 2c, lanes 2 and 5). One recipient sheep (4440) was euthanized at 747 days p.i. due to development of an abomasal emptying disorder. Although PrPSc was not detected by scrapie IHC in the obex or in any of the lymphoid tissues examined from this sheep (Table 1, example shown in Fig. 2b), PrPres accumulation was evident by Western blot analysis after Na-PTA extraction of retropharyngeal and ileocecal lymph node homogenates (Fig. 2c, lanes 2 and 5). Recipient goat 4474 was euthanized at 784 days p.i. for comparison with recipient sheep 4440. Similar to sheep 4440, antemortem accumulation of PrPSc had not been detected in goat 4474 (last biopsy at 721 days p.i.; Fig. 2b). In contrast to sheep 4440, PrPSc and PrPres were readily detected in multiple post-mortem tissues of recipient goat 4474 by 784 days p.i. (Table 1), though still not in the RAMALT. One recipient sheep (4442) and three recipient goats (4470, 4471 and 4479) were eventually removed from isolation to await development of clinical signs. As summarized in Table 1, transmission of scrapie infection was confirmed in four of four recipient goats and in two of four recipient sheep.

The genotypes of goats used in this study included caprine PRNP haplotypes 1 and 2 (White et al., 2008), which differ in sequence only at codon 240. The mature PrP produced by haplotypes 1 and 2 are identical, however, since several C-terminal amino acids, including codon 240, are removed during post-translational maturation of the protein (Stahl et al., 1990). Thus, goats in this study all expressed the archetypal PrP of sheep and goats (referred to as ARQ), which codes for alanine (A) at codon 136, arginine (R) at codon 154 and glutamine (Q) at codon 171 (Goldmann, 2008). Transmission of scrapie to all four goat recipients indicates a placental prion titre high enough to efficiently infect ARQ/ARQ goat kids by the oral route. The recipient sheep used in this study were VRQ/VRQ, a genotype known to be at greatest risk for developing scrapie under field conditions (Baylis et al., 2004). However, a recent oral inoculation study in sheep demonstrates that transmission from an ARQ/ARQ donor is efficient in PrP homologous (i.e. ARQ/ARQ) recipients but results in significantly prolonged incubation in heterologous (ARQ/VRQ or VRQ/VRQ) recipients (González et al., 2012). Similarly, oral transmission of cattle-origin bovine spongiform encephalopathy prions is reduced in VRQ/VRQ sheep as compared with ARQ/ARQ sheep (McGovern et al., 2015; Tan et al., 2012). These findings may explain why transmission of caprine scrapie was only confirmed in two of four VRQ/VRQ sheep recipients in this study but, given constraints that limited the incubation time available for study, transmission to the other two sheep cannot be ruled

### Table 1. Summary of results for goats and sheep orally inoculated as neonates with homogenate prepared from a placenta shed from a goat with clinical scrapie disease

| Animal | Time to PrPSc detection (days p.i.) | Time to clinical disease (days p.i.) | Time to post-mortem examination (days p.i.) | Post-mortem immunoassay |
|--------|-----------------------------------|------------------------------------|---------------------------------------------|--------------------------|
|        |                                   |                                    | Obex | RPLN | ICLN | RAMALT |
|        |                                   |                                    | IHC  | IHC  | WB   | IHC  | WB   | IHC  |
| Goats  |                                   |                                    |      |      |      |       |      |      |
| 4474   | 784                               | NA                                 | 784  | POS  | POS  | POS  | POS  | SUS  |
| 4470   | 564                               | 994                                | 1057 | POS  | POS  | –     | POS  | POS  |
| 4471*  | 564                               | 995                                | 1057 | POS  | POS  | –     | POS  | POS  |
| 4479   | 661                               | 1032                               | 1056 | POS  | POS  | –     | POS  | POS  |
| Sheep  |                                   |                                    |      |      |      |       |      |      |
| 4440   | 747                               | NA                                 | 747‡ | ND   | ND   | [POS]‡| ND   | [POS] |
| 4450   | ND                                | NA                                 | 811  | ND   | ND   | [ND]  | ND   | [ND] |
| 4445   | ND                                | NA                                 | 815  | ND   | ND   | [ND]  | ND   | [ND] |
| 4442   | 387                               | 849                                | 908  | POS  | POS  | –     | POS  | POS  |

*Goat 4471 was homozygous for the caprine PRNP haplotype 1. The other three goats were heterozygous for haplotypes 1 and 2.

‡Euthanized due to inter-current disease (abomasal emptying disorder).

§[Bracketed] result is for WB after Na-PTA extraction; corresponding standard WB result = ND.

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out. Nonetheless, these studies collectively suggest an equal or greater risk of transmission to ARQ/ARQ sheep.

It is unknown if co-infection of the donor goat with an SRLV had a confounding influence on the outcome of this experiment. Small ruminant lentiviruses are a highly related group of retroviruses with potential for interspecies transmission (Leroux et al., 2010), causing persistent infections that can result in several types of chronic inflammatory diseases (Blacklaws, 2012). Co-infection of sheep and goats with classical scrapie and an SRLV increases the peripheral distribution of PrPSc (González et al., 2010; Salazar et al., 2010) and the infectious titre of prions in the milk of ewes with SRLV-associated mastitis (Lacroux et al., 2008). In this study, direct effects on the recipient kids and lambs are unlikely since SRLV infection was not detected by 16 months of age. Although effects on the donor goat cannot be ruled out, the increased distribution of PrPSc that has been reported in co-infected animals was only in association with viral pathology in the respiratory tract or mammary gland; similar lesions are not reported in the placenta. Nevertheless, very little is known about the mechanisms underlying PrPSc accumulation at the placental feto-maternal interface (Alverson et al., 2006; Andréoletti et al., 2002; Lacroux et al., 2007; Tuo et al., 2001, 2002) or the basic mechanisms underlying enhanced cellular accumulation of PrPSc associated with SRLV co-infection (Stanton et al., 2008).

In conclusion, this study importantly demonstrates that the placenta of goats infected with classical scrapie can transmit scrapie to susceptible goat kids and lambs via a natural route of exposure despite relatively sparse accumulation of PrPSc within the goat’s placenta. Thus, like for sheep, the parturient materials and post-partum period of goats must be considered transmission risks for other susceptible small ruminants and environmental contamination.

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References
Alverson, J., O’Rourke, K. I. & Baszler, T. V. (2006). PrPSc accumulation in fetal cotyledons of scrapie-resistant lambs is influenced by fetus location in the uterus. J Gen Virol 87, 1035–1041.
Andréoletti, O., Lacroux, C., Chabert, A., Monnereau, L., Tabouret, G., Lantier, F., Berthon, P., Eychenne, F., Lafond-Benestad, S. & other authors (2002). PrP(Sc) accumulation in placentas of ewes exposed to natural scrapie: influence of foetal PrP genotype and effect on ewe-to-lamb transmission. J Gen Virol 83, 2607–2616.
Baylis, M., Chihota, C., Stevenson, E., Goldmann, W., Smith, A., Sivam, K., Tongue, S. & Gravenor, M. B. (2004). Risk of scrapie in British sheep of different prion protein genotype. J Gen Virol 85, 2735–2740.
Blacklaws, B. A. (2012). Small ruminant lentiviruses: immunopathogenesis of visna-maedi and caprine arthritis and encephalitis virus. Comp Immun Microbiol Infect Dis 35, 259–269.
Caughey, B., Baron, G. S., Chesebro, B. & Jeffrey, M. (2009). Getting a grip on prions: oligomers, amyloids, and pathological membrane interactions. Annu Rev Biochem 78, 177–204.
Goldmann, W. (2008). PrP genetics in ruminant transmissible spongiform encephalopathies. Vet Res 39, 30.
González, L., Jeffrey, M., Sisó, S., Martin, S., Bellworthy, S. J., Stack, M. J., Chaplin, M. J., Davis, L., Dagleish, M. P. & Reid, H. W. (2005). Diagnosis of preclinical scrapie in samples of rectal mucosa. Vet Rec 156, 846–847.
González, L., Martin, S., Hawkins, S. A., Goldmann, W., Jeffrey, M. & Sisó, S. (2010). Pathogenesis of natural goat scrapie: modulation by host PRNP genotype and effect of co-existent conditions. Vet Res 40, 48.
Gough, K. C. & Maddison, B. C. (2010). Prion transmission: prion excretion and occurrence in the environment. Prion 4, 275–282.
Haddow, W. J., Kennedy, R. C., Race, R. E. & Eklund, C. M. (1980). Virologic and neurohistologic findings in dairy goats affected with natural scrapie. Vet Pathol 17, 187–199.
Herrmann, L. M., Cheevers, W. P., Marshall, K. L., McGuire, T. C., Hutton, M. M., Lewis, G. S. & Knowles, D. P. (2003a). Detection of serum antibodies to ovine progressive pneumonia virus in sheep by using a caprine arthritis-encephalitis virus competitive-inhibition enzyme-linked immunosorbent assay. Clin Diag Lab Immunol 10, 862–865.
Herrmann, L. M., Cheevers, W. P., McGuire, T. C., Adams, D. S., Hutton, M. M., Gavin, W. G. & Knowles, D. P. (2003b). Competitive-inhibition enzyme-linked immunosorbent assay for detection of serum antibodies to caprine arthritis-encephalitis virus: diagnostic tool for successful eradication. Clin Diag Lab Immunol 10, 267–271.
Hoinville, L. J., Tongue, S. C. & Wilesmith, J. W. (2010). Evidence for maternal transmission of scrapie in naturally affected flocks. Prev Vet Med 93, 121–128.
Konold, T., Bone, G. E., Phelan, L. J., Simmons, M. M., González, L., Sisó, S., Goldmann, W., Cawthraw, S. & Hawkins, S. A. (2010). Monitoring of clinical signs in goats with transmissible spongiform encephalopathies. BMC Vet Res 6, 13.
Lacroux, C., Corbière, F., Tabouret, G., Lugan, S., Costes, P., Matthey, J., Delmas, J. M., Weisbecker, J. L., Foucras, G. & other authors (2007). Dynamics and genetics of PrPSc placental accumulation in sheep. J Gen Virol 88, 1056–1061.
Lacroux, C., Simon, S., Benestad, S. L., Maillet, S., Matthey, J., Lugan, S., Corbière, F., Cassard, H., Costes, P. & other authors (2008). Prions in milk from ewes incubating natural scrapie. PLoS Pathog 4, e1000238.
Leroux, C., Cruz, J. C. & Mornex, J. F. (2010). SRLVs: a genetic continuum of lentiviral species in sheep and goats with cumulative evidence of cross species transmission. Curr HIV Res 8, 94–100.
McGovern, G., Martin, S., Jeffrey, M., Bellworthy, S. J., Spiropoulos, J., Green, R., Lockey, R., Vickery, C. M., Thurston, L. & other authors (2015). Influence of breed and genotype on the onset and distribution of infectivity and disease-associated prion protein in sheep following oral infection with the bovine spongiform encephalopathy agent. J Comp Pathol 152, 28–40.

O’Rourke, K. I., Baszler, T. V., Besser, T. E., Miller, J. M., Cutlip, R. C., Wells, G. A., Ryder, S. J., Parish, S. M., Hamir, A. N. & others authors (2000). Preclinical diagnosis of scrapie by immunohistochemistry of third eyelid lymphoid tissue. J Clin Microbiol 38, 3254–3259.

O’Rourke, K. I., Zhuang, D., Truscott, T. C., Yan, H. & Schneider, D. A. (2011). Sparse PrP(Sc) accumulation in the placentas of goats with naturally acquired scrapie. BMC Vet Res 7, 7.

Onodera, T., Ikeda, T., Muramatsu, Y. & Shinagawa, M. (1993). Isolation of scrapie agent from the placenta of sheep with natural scrapie in Japan. Microbiol Immunol 37, 311–316.

Pattison, I. H. & Millson, G. C. (1961). Scrapie produced experimentally in goats with special reference to the clinical syndrome. J Comp Pathol 71, 101–109.

Pattison, I. H., Hoare, M. N., Jebbett, J. N. & Watson, W. A. (1972). Spread of scrapie to sheep and goats by oral dosing with foetal membranes from scrapie-affected sheep. Vet Rec 90, 465–468.

Pattison, I. H., Hoare, M. N., Jebbett, J. N. & Watson, W. A. (1974). Further observations on the production of scrapie in sheep by oral dosing with foetal membranes from scrapie-affected sheep. Br Vet J 130, lxv–lxvi.

Race, R., Jenny, A. & Sutton, D. (1998). Scrapie infectivity and proteinase K-resistant prion protein in sheep placenta, brain, spleen, and lymph node: implications for transmission and antemortem diagnosis. J Infect Dis 178, 949–953.

Salazar, E., Monleón, E., Bolea, R., Acín, C., Pérez, M., Alvarez, N., Leginaoikoa, I., Juste, R., Minguiñón, E. & other authors (2010). Detection of PrPSc in lung and mammary gland is favored by the presence of Visna/maedi virus lesions in naturally coinfected sheep. Vet Res 41, 58.

Soto, C. (2011). Prion hypothesis: the end of the controversy? Trends Biochem Sci 36, 151–158.

Stahl, N., Baldwin, M. A., Burlingame, A. L. & Prusiner, S. B. (1990). Identification of glycoinositol phospholipid linked and truncated forms of the scrapie prion protein. Biochemistry 29, 8879–8884.

Stanton, J. B., Knowles, D. P., O’Rourke, K. I., Herrmann-Hoesing, L. M., Mathison, B. A. & Baszler, T. V. (2008). Small-ruminant lentivirus enhances PrPSc accumulation in cultured sheep microglial cells. J Virol 82, 9839–9847.

Tan, B. C., Blanco, A. R., Houston, E. F., Stewart, P., Goldmann, W., Gill, A. C., de Wolf, C., Manson, J. C. & McCutcheon, S. (2012). Significant differences in incubation times in sheep infected with bovine spongiform encephalopathy result from variation at codon 141 in the PRNP gene. J Gen Virol 93, 2749–2756.

Touzeau, S., Chase-Topping, M. E., Matthews, L., Lajous, D., Eychenne, F., Hunter, N., Foster, J. D., Simm, G., Elsen, J. M. & Woolhouse, M. E. (2006). Modelling the spread of scrapie in a sheep flock: evidence for increased transmission during lambing seasons. Arch Virol 151, 735–751.

Tuo, W., Zhuang, D., Knowles, D. P., Cheevers, W. P., Sy, M. S. & O’Rourke, K. I. (2001). Prp-c and Prp-Sc at the fetal-maternal interface. J Biol Chem 276, 18229–18234.

Tuo, W., O’Rourke, K. I., Zhuang, D., Cheevers, W. P., Spraker, T. R. & Knowles, D. P. (2002). Pregnancy status and fetal prion genetics determine PrPSc accumulation in placentomes of scrapie-infected sheep. Proc Natl Acad Sci U S A 99, 6310–6315.

Vaccari, G., Panagiotidis, C. H., Acin, C., Peletto, S., Barillet, F., Acutis, P., Boissers, A., Langeveld, J., van Keulen, L. & other authors (2009). State-of-the-art review of goat TSE in the European Union, with special emphasis on PRNP genetics and epidemiology. Vet Res 40, 48.

White, S., Herrmann-Hoesing, L., O’rourke, K., Waldron, D., Rowe, J. & Alverson, J. (2008). Prion gene (PRNP) haplotype variation in United States goat breeds (Open Access publication). Genet Sel Evol 40, 553–561.