Novel biological and technological platforms for dental clinical use

Subjects: Dentistry
Created by: Thimios Mitsiadis

Abstract

Human teeth have a limited capacity to regenerate and thus biological reconstruction of damaged or lost dental tissues remains a significant challenge in modern dentistry. Recent efforts focus on alternative therapeutic approaches for partial or whole tooth regeneration that complement traditional dental treatments using sophisticated materials and dental implants. These multidisciplinary approaches are based on the combination of stem cells with advanced tissue engineer products and computing technology, and they hold great promise for future applications in dentistry. The administration to patients of dynamic biological agents composed by stem cells and scaffolds will certainly increase the regenerative capacity of dental pathological tissues. The design of innovative materials for tissue restoration, diagnostics, imaging and targeted pharmaceutical treatment will significantly improve the quality of dental care and will have a major societal impact. This review depicts the current challenges in dentistry and describes the possibilities for novel and successful therapeutic applications in the near future.

Giovanna Orsini¹, ², Pierfrancesco Pagella¹, Angelo Putignano², Thimios A. Mitsiadis¹*

¹ Orofacial Development and Regeneration, Institute of Oral Biology, Centre of Dental Medicine, Faculty of Medicine, University of Zurich, Zurich, Switzerland;
² Department of Clinical Sciences and Stomatology, Polytechnic University of Marche, Ancona, Italy.

* Correspondence:
Thimios A. Mitsiadis, Orofacial Development and Regeneration, Institute of Oral Biology, Centre for Dental Medicine, Plattenstrasse 11, 8032 Zürich, Switzerland.

Email: thimios.mitsiadis@zzm.uzh.ch

Introduction - The Tooth Organ

The tooth organ is composed by a unique combination of hard and soft tissues. The outermost layer is constituted by enamel, the most mineralized tissue of the human body, which guarantees protection to the inner elements of the tooth (Figure 1). Enamel displays unique physical characteristics, such as complex three-dimensional organization and extremely long hydroxyapatite crystallites, to resist large masticatory forces and continual attacks by acids from food and bacterial sources (Boyde, 1997). Ameloblasts, which are the epithelial cells responsible for enamel formation, and their precursors are lost upon tooth eruption, making human adult teeth inapt of enamel regeneration. The great complexity of enamel, together with the absence of appropriate cells in adult patients, make therapies aiming to enamel regeneration an exciting challenge.

Due to its extremely high mineral content, enamel is very brittle. This property is compensated by dentin, a less mineralised, elastic, avascular tissue (Figure 1). Dentin encloses the dental pulp, a soft connective
tissue that conveys vascularisation and innervation, representing the vital core of the tooth organ (Figure 1). The vascular system provides oxygen, nutrients and metabolites, while sensory innervation is fundamental for pain perception and control of biting strength. In the peripheral boundary of the dental pulp are situated mesenchymal-derived odontoblasts, which produce and maintain dentin. Dentin is characterized by closely packed tubules traversing its thickness and containing the cytoplasmatic extensions of odontoblasts, as well as sensory nerve terminals, which render dentin highly sensitive to external stimuli. More importantly, dentin can repair itself, due to the activation of the existing odontoblasts or the newly formed odontoblasts derived from pulp stem cells that produce a reactionary mineralized matrix upon injury. However, pulp reaction is not sufficient in case of severe tooth injury and/or extensive infection, and this healing failure often leads to pulp irreversible inflammation followed by necrosis (DeRosa, 2006).

The tooth is anchored to the alveolar bone by the roots, constituted by dentin and cementum. Roots are connected to the alveolar bone by a specialized connective tissue, the periodontal ligament, which ensures tooth stability and absorbs mechanical stresses during chewing (Figure 1). Periodontal disease is the most frequent cause of tooth loss, making periodontal regeneration a pressing need for the dental field (Mitsiadis et al., 2015).

The structural hallmarks of dental hard tissues are strictly dependent on tightly regulated and long developmental processes that cannot be easily reproduced within acceptable therapeutic time frames. Moreover, the oral cavity constitutes a challenging environment for any regenerative approach, as it is constantly exposed to chemical, mechanical and bacterial insults. Despite these difficulties, recent technological advancements are becoming an inherent aspect of dental practice, improving effectiveness of treatments. Similarly, the continuous developments in stem cell research and nanotechnology are paving the way for regenerative approaches in dentistry.

**Innovation in current dental treatments: From materials to tooth regeneration**

The great improvements in computing-related technologies and materials has widened the options to alternative and more precise dental treatments (Beuer et al., 2008; Hancocks, 2017), and helped in establishing more reliable diagnostic tools and therapeutic plans (Levato et al., 2015; Lynch, 2017). Numerous advancements have been made with the advent of novel imaging techniques such as computer-aided design and manufacturing (CAD/CAM) technology, optimized intraoral imaging, digital radiography, and computer aided implant surgery (Levato et al., 2015); (Hammerle et al., 2009; Zhou et al., 2018). Apart from its use as a diagnostic tool, imaging contributed to the improvement of the daily dental practice, since treatments benefited from high definition microscopes that permit the detailed visualization of the operative dental field (Del Fabbro et al., 2015).

Material sciences have led the way for the development of therapeutic approaches aiming to substitute damaged or lost dental tissues. Despite limitations in functionality and longevity, biomaterials are still present in dental treatments since nanotechnology has remarkably improved their performance and the clinical outcome of certain procedures. The combination of nanomaterials with advanced technologies has upgraded prosthetic and aesthetic dentistry, which are fields aiming to optimize the functional and aesthetic appearance of dentition. 3D printing systems represent the most innovative next-step technology, aiming to manufacture customised products based on computer-designed digital tools (Yang et al., 2018). Pain management has also enormously benefited from the advent of these novel technologies (Banerjee et al., 2011).

However, the most important development of the last decade is the rise of a new dental discipline that is based on the capacity of stem cells to repair or regenerate various impaired tissues. Stem cell-based regenerative dentistry is linked to advanced tissue engineering products and nanotechnology, which have created an important clinical shift toward the functional repair and regeneration of damaged dental tissues.
Combining stem cell biology and nanotechnology for regenerating dental tissues

Stem cells are characterized by their potential to self-replicate and their capacity to differentiate into a vast variety of cell populations (Mitsiadis and Graf, 2009). Epithelial and mesenchymal stem cell populations are present in almost all adult human tissues and organs, including teeth. A variety of dental mesenchymal stem cells (DMSCs) populations have been isolated from both deciduous and permanent teeth, characterized, and tested for their potential applications in regenerative dentistry (Gronthos et al., 2000; Gronthos et al., 2002; Miura et al., 2003). Adult DMSCs localized in the dental pulp and periodontal tissue ensure human tooth homeostasis and regeneration (Bluteau et al., 2008), and therefore represent optimal clinical tools for the repair of damaged dental tissues. Actual efforts are oriented towards pulp and periodontal tissue repair, where these tissues can be regenerated by transplantation of stem cells alone or in combination with functionalized scaffolds. More challenging and problematic is, however, the regeneration of tooth enamel using epithelial cells, since neither dental epithelial stem cells (DESCs) nor ameloblasts are present in the crown of adult functional teeth (Mitsiadis et al., 2015; Orsini et al., 2015).

More exiting, but greatly perplexing, is the perspective to generate entire brand-new teeth by mixing DESCs and DMSCs. Although very difficult to be realized, several attempts towards this direction have been pursued in animal models (Oshima and Tsuji, 2014).

The success and efficacy of any stem cell-mediated therapy can be evaluated by a set of modern nanotechnology tools, since they allow tracking the migration, fate and regenerative impact of stem cells in vivo. For example, transplanted stem cells can be tracked for long periods with non-invasive imaging techniques using fluorescent dyes (Arbab et al., 2009; Gera et al., 2010), and with magnetic nanoparticles that can be traced by MRI and provide information about their kinetics and fate during dental tissue regeneration (Jimenez-Rojo et al., 2012). This knowledge could be used for designing appropriate scaffolds that will host stem cells before transplantation. Furthermore, it will allow evaluating the therapeutic efficacy of precise dental stem cell populations that have been exposed to specific microenvironments. Indeed, artificial microenvironments, which may direct stem cells towards a precise fate and function, can be achieved through nanotechnology (Bluteau et al., 2008). A big variety of nanoscale biodegradable structures with specific size, surface chemistry and shape can be used for the creation of microenvironments that are adapted for the needs of regenerative dentistry (Mitsiadis et al., 2012). Such biodegradable scaffolds, once transplanted, may act as temporary niches that control stem cell behaviour and guide dental tissue repair (Iwatsuki et al., 2006).

It is obvious that the range of dental disciplines that can benefit from the recent advances of stem cell biology, material sciences and nanotechnology is extremely wide. The present mini-review covers current and future therapeutic approaches for managing the (1) damage of the tooth crown, including the harm of enamel and/or dentin-pulp tissues, (2) periodontal insults, and (3) tooth loss.

Tooth crown damage - Current restorative treatments

Enamel and dentin of the tooth crown are most often the first tissues to be affected following traumatic injuries or carious lesions (Figure 1).
Figure 1. Tooth structure in physiological and pathological conditions. A) Histological section of a human premolar (blue colour: toluidine blue). B) Histological sections of human carious teeth. Left side: ground unstained section, showing mild carious injuries (red arrows) affecting only enamel. Right side: decalcified section stained with hematoxylin and eosin, showing a severe carious injury (red arrows) with bacterial invasion (asterisks: bacterial front within dentin). Abbreviations: d, dentin; e, enamel; p, pulp; td, tertiary dentin.

Prompt and efficient repair of enamel and/or dentin is fundamental to prevent infection and damage extending towards dental soft tissues (i.e., dental pulp, periodontium) and alveolar bone. The most used
approach for treating enamel and dentin harm is the substitution and restoration of the destroyed or lost dental hard tissues by sophisticated composite materials. However, traditional adhesive systems are unstable and fail over time, thus leading to marginal leakage and poor retention of the restoration in the tooth (Breschi et al., 2018). Therefore, a major task of nanotechnology in dentistry is to develop novel durable materials and adhesive systems with improved enamel- and/or dentin-bonding performance in order to increase the longevity of the restorations and prevent repeated treatments. Indeed, the introduction of novel materials such as phosphine oxide initiators and monomethacrylate diluents has led to dental composites with satisfactory and adequate properties (Kilambi et al., 2009). The introduction of nanofillers and nanomaterials led to even more significant advances in terms of optimizing the properties and performance of the composites (Ilie et al., 2013; Goracci et al., 2014; Monterubbianesi et al., 2016). These nanotechnology-based strategies using cross-linking agents and Ca- and P-releasing means, which mimic the process of natural dentin mineralization, have also reduced the degradation of the resin-dentin bonded interface (Mazzoni et al., 2018).

Ceramic-based materials are privileged by dentists for the restoration of damaged tooth crowns, mainly because of their superior aesthetic appearance and biocompatibility (Wittneben et al., 2017; Ozcan and Jonasch, 2018). To overcome the fact that ceramic materials are brittle and prone to cracks propagation, several transformation/toughening mechanisms have been developed, leading to higher aging resistant-ceramics (Zhang et al., 2017) such as zirconia with exceptional toughness and flexural strength (Guazzato et al., 2004).

Nanomodified materials could be also designed for controlling oral microbiota and the formation of dental plaque, and, furthermore, for enhancing the mineralization process in the cases of enamel wear and/or dentin hypersensitivity due to the extensive consumption of acidic drinks (Orsini et al., 2013; Lelli et al., 2014). Indeed, the use of synthetic nanohydroxyapatite particles and other Ca-based nanomodified materials in dentifrices may offer a protective nanostructured coating on the tooth surface that simultaneously restores the lost minerals from enamel (Orsini et al., 2013; Lelli et al., 2014).

The preservation of the dental pulp, which is a living tissue ensuring tooth physiological function, is of prime importance during the treatment of a damaged tooth crown. In very severe tooth injury, the pulp is also affected and may lose its vitality. Therefore, the endodontic therapy (i.e., pulp tissue removal) is imposed in order to prevent further bacterial progression and damage of the surrounding alveolar bone. This is followed by disinfection of the dental root canals and the replacement of the pulp tissue with inorganic materials. Devitalized teeth are more fragile than normal teeth and consequently are predisposed to postoperative fractures (DeRosa, 2006).

**Challenges in dentin-pulp regeneration**

Regenerative endodontics aims at reforming the original pulp tissue morphology and physiology based on tissue engineering principles (Murray and Garcia-Godoy, 2006; Diogenes and Hargreaves, 2017). Nanomaterials can be used either alone or implemented with growth factors and stem cells in order to stimulate and enhance the regenerative capacity of the pulp tissue. Adjustment of biomaterials for dental specific purposes would require adjustments at a nanoscale level, thus allowing multifunctionality within a given small surface, increasing the quality of targeting, and better controlling bioactive molecules delivery (Fioretti et al., 2011; Diogenes and Ruparel, 2017). Nanomaterials developed for endodontic purposes can deliver antibacterial and anti-inflammatory molecules, as well as growth factors that will guide the behaviour (e.g., cell migration, proliferation, and differentiation) of the various dental pulp cell populations (e.g., pulp fibroblasts, endothelial cells, neuronal cells, immune cells). Biomimetic scaffolds composed of natural molecules, such as type I collagen, hyaluronic acid and chitosan, combined with nanoassembled materials possessing anti-inflammatory capabilities have been generated to stimulate pulp tissue regeneration and to prevent inflammation (Fioretti et al., 2011). Although such nanofibrous and microporous membranes have provided promising results, significant improvements are still needed to
create scaffolds that promote proper pulp regeneration (Yamauchi et al., 2011; Albuquerque et al., 2014). Numerous attempts using human DMSCs have been made in a variety of animal models in order to achieve complete dental pulp regeneration (Figure 2A), a process that also requires neovascularization and re-innervation of this tissue. Experiments have shown that human DMSCs are capable to differentiate into odontoblasts and to form dentin-like structures when transplanted together with a ceramic powder in immune-compromised mice ex vivo (Gronthos et al., 2000; Gronthos et al., 2002). Similar studies have revealed that human DMSCs seeded on poly-D, L-lactide/glycol scaffolds are able to regenerate vascularized pulp tissue when transplanted into an empty mouse tooth root canal (Volponi et al., 2010; Hayashi et al., 2015). Recently, new experimental strategies have been elaborated, where DMSCs-seeded scaffolds combined with bioactive molecules fulfil the empty pulp chamber immediately after pulp removal (Albuquerque et al., 2014; Piva et al., 2014). Pilot studies in humans have demonstrated the safety and efficacy of DMSCs for complete dental pulp regeneration and new dentin formation (Nakashima and Iohara, 2017; Nakashima et al., 2017). Bone morphogenetic proteins (BMPs) have been commonly used for accelerating and enhancing the production of dentin during dental pulp regeneration (Luiz de Oliveira da Rosa et al., 2017). While these stem cell-based procedures appear to improve pulp tissue regeneration, their effectiveness for achieving accurate, precise and long-lasting therapies is still unclear. As a matter of fact, most approaches aiming at dental pulp regeneration led to the formation of fibrotic tissues that can undergo degeneration over time or be replaced with bone. The possibility to decellularise healthy human dental pulps (Song et al., 2017) opens new horizons in regenerative dentistry since these decellularised tissues could serve as natural scaffolds for supporting transplanted autologous DMSCs. Decellularised pulps represent ideal biomaterials for hosting stem cells and guiding neovascularization and re-innervation within the regenerating tissues.

While significant efforts have been produced so far, regenerative procedures have to be further investigated in order to ultimately provide evidence of functional dental pulp regeneration in vivo (Figure 2A) (Torabinejad and Faras, 2012; Diogenes and Ruparel, 2017).
Challenges in enamel regeneration

*De novo* formation of enamel in humans is one of the greatest challenges in regenerative dentistry, since amelogenesis is a very complex process and dental epithelial stem cells (DESCs) that could regenerate enamel are very rare in adult human teeth. Very few dental epithelial cells with stem cell properties have been isolated from the periodontal tissue (i.e., epithelial rests of Malassez, ERM). Experiments using porcine ERM have demonstrated that these cells can differentiate into ameloblasts when co-cultured with dental pulp cells *in vitro* and can form enamel structures after their transplantation *in vivo* (Shinmura et al., 2008). Although ERM is a potential stem cell source for enamel regeneration, availability of these cells in human teeth is scarce, making thus necessary the identification of other epithelial stem cell populations of non-dental origin that could differentiate into enamel-producing ameloblasts.

Another key issue in generating new enamel is time. The accomplishment of proper enamel formation requires many years, a time frame clearly incompatible with clinical needs. Moreover, mild disturbances during this process could lead to the generation of defective enamel (Cantu et al., 2017). Therefore, any procedure and technique that will be able to considerably accelerate the process of amelogenesis will be of benefit to the patients and dental community.
Periodontal diseases - Current periodontal therapies

Periodontium is a common site of pathologies that severely affect not only the structure of the surrounding tissues (i.e., dental root, alveolar bone) but also tooth functionality. Severe inflammation to the periodontium leads to significant alterations in both the structure and quantity of the alveolar bone, a process that ultimately may cause tooth loss (Lindhe et al., 1983). Contemporary, periodontal therapies include a wide range of surgical procedures along with use of bone grafts as tissue substitutes, barrier membranes for protecting the healing area from undesirable epithelial tissues (Howell et al., 1997; Aghaloo and Moy, 2007), and growth factors for enhancing the healing capacity of the harmed tissues (Lynch et al., 1991b). Bone grafting materials, aiming to stimulate bone augmentation and periodontal regeneration, include intraoral or extraoral autografts, freeze-dried and fresh-frozen bone allografts, animal-derived bone deproteinised xenografts, and hydroxyapatite and beta-tricalcium phosphate alloplasts (Pilipchuk et al., 2015; Sheikh et al., 2017). These grafting materials could be used alone or in association with various growth factors. It has been shown that application of these regenerative methods in clinics allowed the formation of novel osseous tissues with similar to the pre-existent native bone characteristics (Scarano et al., 2006; De Angelis et al., 2011; Danesh-Sani et al., 2016; Clark et al., 2018). Even though, these approaches do not always ensure a predictable and desirable outcome of periodontal regeneration and often result in healing with epithelial lining rather than new periodontal tissue formation (Lynch, 1992).

Challenges in periodontal regeneration

A fundamental goal in regenerative dentistry is to reconstruct a functional periodontium consisting of new cementum, alveolar bone and periodontal ligament around the tooth root damaged area (Figure 2B). DMSCs isolated from the periodontal space (i.e., periodontal ligament stem cells, dental follicle stem cells) of human teeth can differentiate into the various cell types of the periodontium in vitro when combined with different scaffolds or dentin matrix (Takahashi and Yamanaka, 2006; Washio et al., 2010; Arakaki et al., 2012; Yang et al., 2012). These stem cell populations have been shown to improve periodontal regeneration when transplanted into immunocompromised animals ex vivo, indicating their great potential for future stem cell-based therapies in dentistry (Seo et al., 2004; Caton et al., 2011). A variety of growth factors have been also used for improving the regenerative efficacy of stem cells in the periodontium. Diverse experiments have demonstrated that platelet-derived growth factors (PDGFs) stimulate periodontal tissue regeneration (Lynch et al., 1991b; Howell et al., 1997; Clark et al., 2018), while BMPs enhance alveolar bone and cementum production (Lynch et al., 1989; Howell et al., 1997; Selvig et al., 2002). However, excessive bone formation that results in tooth ankylosis can be a frequent side effect following the use of BMPs, since these molecules favour and direct stem cells differentiation towards the osteogenic fate.

Clinical studies have demonstrated that enamel matrix derivatives also assist and promote periodontal tissue regeneration (Miron et al., 2016b; Miron et al., 2017). Advanced new bone grafting materials with improved physicochemical properties have been used as carriers of enamel protein derivatives in order to further improve their clinical performance (Miron et al., 2016a; Miron et al., 2016b). Nevertheless, despite the very encouraging clinical outcomes, the mechanism of action of these enamel matrix molecules is not yet clear.

TOOTH LOSS

Current dental implant treatments

The use of dental implants has become a common and successful treatment for replacing missing teeth for pathologic, traumatic and genetic causes (Figure 3) (Esposito et al., 2014).
A typical dental implant is composed of a metal screw part that interfaces and integrates within the alveolar bone, and another part where tooth crown substitutes are placed. The retention of a dental implant requires its close contact with the alveolar bone, a process termed osseointegration. Despite their large and regular usage in dental clinics, implants still need significant improvements, particularly in their capacity to stimulate cellular events at the implantation site that would guarantee their long-term integration and retention (Variola et al., 2009; Variola et al., 2011). The use of nanotechnology has improved the osseointegration of implants by modifying their surfaces, thus allowing the shortening of the healing period (Barbucci et al., 2003; Mendonca et al., 2008). Indeed, zinc-modified calcium silicate coatings, nanohydroxyapatite-blasted surfaces, nanotextured blasted titanium surfaces, as well as gold nanoparticles coated surfaces have considerably enhanced the adhesive properties of implants and therefore their osseointegration (Coelho et al., 2016; Heo et al., 2016; Bezerra et al., 2017; Yu et al., 2017). However, there is a major risk of infection of tissues surrounding the implant, a pathology termed peri-implantitis (Singh, 2011). In vitro and in vivo studies have shown that the incorporation of antibacterial agents to dental implants (e.g., silver nanoparticles) could partly prevent the growth of bacteria and therefore decrease the percentage of implant treatments failure (Godoy-Gallardo et al., 2016; Pokrowiecki et al., 2017). It has been also demonstrated that gallium-modified chitosan/poly (acrylic acid) bilayer coatings might improve titanium implant performances by limiting bacterial adhesion and proliferation (Bonifacio et al., 2017). Dental implants have also benefited from regenerative technologies using scaffolds, stem cells and growth factors that contribute to enhanced osseointegration and host tissue response (Pilipchuk et al., 2015). Despite a good number of preclinical studies in large animal models for guided bone and periodontal regeneration around implants using growth factors and protein delivery systems (Lynch et al., 1991a; Selvig et al., 2002; Sauerbier et al., 2011; Alvarez et al., 2012; Larsson et al., 2016), and the evident clinical advantages, well-conducted human randomized clinical studies that will definitively validate these approaches are still lacking (Caton et al., 2011) (Seo et al., 2004; Rickert et al., 2011; Sauerbier et al., 2011). To date, only few randomized clinical trials have been performed and therefore it is absolutely necessary the realization of larger trials (Kaigler et al., 2013; Kaigler et al., 2015).

**Challenges in entire tooth regeneration**

Regeneration of entire brand-new teeth for the replacement of missing or lost teeth is the most ambitious goal in dentistry and requires the use and recombination of dental mesenchymal and epithelial stem cells (Papagerakis, 2013; Otsu et al., 2014). DMSCs can form all mesenchymal components of the tooth organ and the surrounding tissues such as dentin, cementum and alveolar bone, while DESCs are essential for the generation of enamel. Since most of the dental epithelial cell populations disappear shortly after tooth eruption and DESCs are limited in human adult teeth, current knowledge on DESCs has been obtained mainly from rodents, where they contribute to the renewal of the enamel in the continuously growing incisors (Mitsiadis and Harada, 2015).
Two main strategies have been elaborated for constructing whole new teeth (Mitsiadis and Papagerakis, 2011; Otsu et al., 2014; Mitsiadis and Harada, 2015). One approach consists in recombining and culturing DESCs and DMSCs in vitro until they will form a tooth germ that subsequently will be transplanted into the alveolar bone. It is expected that this tooth germ will further develop and grow, erupt and finally become a functional tooth. Another approach relies to tooth-shaped polymeric biodegradable scaffolds that are filled with both DESCs and DMSCs and implanted into the alveolar bone, expecting that will finally give rise to functional teeth. The three-dimensional structure of the scaffolds should drive the differentiation of the transplanted stem cells into odontoblasts and ameloblasts (Bluteau et al., 2008). Indeed, several experiments in mice using bioengineered approaches have revealed that functional teeth with appropriate crowns, dental pulp, roots and periodontal ligament can be formed following their implantation in mandibles (Jimenez-Rojo et al., 2012; Oshima and Tsuji, 2014; Otsu et al., 2014; Mitsiadis and Harada, 2015). However, similar results have not yet been obtained with human cells, due mainly to the limited number of adult DESCs and the significantly elongated time period that is needed for proper human tooth development. Penury of DESCs within human adult teeth might be successfully addressed by differentiating patient-specific inducible pluripotent stem cells (iPSCs) into DESCs. Certain studies have shown that iPSCs technology could be successfully used in regenerative dentistry, since re-aggregation of human iPSC-derived mesenchymal cells and mouse dental epithelium resulted in the formation of entire teeth ex vivo (Takahashi and Yamanaka, 2006; Arakaki et al., 2012; Otsu et al., 2014; Mitsiadis and Harada, 2015). Although promising, this approach also needs further investigation, as effective protocols for the differentiation of human DESCs from iPSCs are not available yet.

**Novel Platforms for Tooth Modelling, Drug Discovery and Diagnostics**

**Use of organoids and organ-on-chip devices in dentistry**

Appropriate systems for modelling human organs and pathologies represent a constant need in all branches of biomedical research and practice, included dentistry. Animal models and two-dimensional (2D) human cell culture systems have been traditionally used for most pre-clinical studies aiming at the development of novel cell-based and pharmaceutical therapies. However, translation of preclinical results into effective treatments remains poor (Weeber et al., 2017), highlighting the need for accurate human-emulation systems (Skardal et al., 2016). In this context, great expectations are accompanying the recent developments on spheroids, organoids, microfluidics and organ-on-chip technologies.

Spheroids and organoids are 3D culture systems, obtained by primary stem cells and tissues, which are increasingly used to model and understand tissue-specific physiology (Figure 4). The 3D structure of both systems allows establishment of complex cell-cell interactions and gradients of oxygen, nutrients and soluble signals that generate tissue-specific heterogeneous cell types. Organoids provide additional features compared to spheroids, as they are able of self-organization, exhibit similar architecture to the tissue of origin and exert tissue-specific complex functions (Yin et al., 2016).
Dental spheroids or dentospheres have been successfully generated from both mouse and human dental epithelial and mesenchymal (e.g., pulp and periodontium) tissues (Berahim et al., 2011; Bonnamain et al., 2011; Miquel, 2011; Natsiou et al., 2017). Epithelial dentospheres formed from mouse incisors and molars, upon modulation of their culture conditions, have either demonstrated strong stem cell capabilities or generated differentiation gradients (Natsiou et al., 2017). Human mesenchymal spheroids consistently displayed higher expression of odontoblast- and periodontal-specific differentiation markers when compared to 2D culture systems (Berahim et al., 2011; Bonnamain et al., 2011). These aspects make spheroids valuable tools for studying cytodifferentiation events in human dental tissues in vitro, and might be a source of stem cells for personalized dental regenerative approaches. Indeed, genetic diseases are often associated with dental anomalies (Mitsiadis and Luder, 2011; Klein et al., 2014), which could be properly modelled and investigated in patient-specific dental spheroids. Similarly, such spheroids represent novel tools for studying the behaviour of definite human dental cell populations to novel materials and drugs. However, despite their incontestable advantages, it is not yet clear to what extent spheroids and organoids could faithfully represent the in vivo dental status. Organoids and spheroids in fact lack many features that are critical for the function of any organ, such as vasculature, innervation, mechanical cues, and immune responses (Ingber, 2016).

These limitations are the basis for the rise of microfluidic “organ-on-chip” systems. Organ-on-chips are microfluidic or nanofluidic devices composed of different chambers, where organ-specific elements such as epithelial, mesenchymal, endothelial and neuronal cells and/or tissues are cultured (Figure 4) (Bhatia and Ingber, 2014). Porous membranes allow the passage of molecular cues between the different chambers, while blood circulation is simulated by the regulated flow of enriched and specific media. These devices can incorporate mechanical forces to recreate physiological movements and stresses (Bhatia and Ingber, 2014), as well as electrical stimuli, allowing the modelling and analysis of complex organ-specific
physiological and pathological processes. Importantly, circulating immune cells and even living microbiomes can be integrated in these devices to mimic complex organ-level responses (Bhatia and Ingber, 2014; Ingber, 2016). Microfluidic devices involving dental tissues have been used for the first time for analysing the crosstalk of tooth germs and DMSCs with trigeminal innervation (Pagella et al., 2014; Pagella et al., 2015). These pioneer studies have shown that microfluidics can faithfully imitate and reproduce the *in vivo* dental situation and thus reinforce the options to study dental tissues in “organ-on-chip” systems. Results obtained from these devices contribute to successfully emulate human- and patient-specific dental tissues *in vitro*. The most ambitious goal of these microfluidic devices consists in the modelling of the functional interconnection between different human organs, by the realization of so-called “bodies-on-chip”. In fact, “organ-on-chip” devices can be interconnected via microfluidic tubes, which emulate systemic blood circulation. Via such emulated vasculature, molecular cues as well as immune responses can propagate to all organs, allowing the study of body-level responses to organspecific events (Ingber, 2016). Such approach is already being used and optimized for modelling of pharmacokinetic and pharmacodynamics of systemic human drug responses (Prantil-Baun et al., 2018). With these platforms, it will be possible to study body-level responses to the various dental pathologies. While it is long known that oral diseases are strongly associated with a plethora of systemic disorders, including atherosclerosis, stroke, and systemic infections (Slavkin and Baum, 2000), the mechanisms underlying these connections and thus their therapeutic relevance are far from being understood. A human “body-on-chip” system would finally allow understanding how dental and systemic health are correlated, thus testing how the treatment of dental diseases affects general physiology.

Microfluidic devices could be also employed for the detection of both specific metabolites (Wu et al., 2017) and particular bacterial strains (Czilwik et al., 2015) that are involved in chronic diseases. Within the dental field, microfluidic devices have been used for the detection of pathogenic bacteria that lead to periodontitis and carious diseases. Recent technological developments allowed the significant shortening of this process via optimization of fully automated and integrated DNA extraction, multiplex PCR preamplification and species-specific real-time PCR (Figure 4) (Chen et al., 2007; Czilwik et al., 2015). These systems allow a fast processing of samples, without loss of sensitivity and complex laboratory instrumentation.

Recent nanotechnology tools permitted the detection of single oral bacteria *in situ* via graphene-biosensors equipped with electrodes and antennae, which were printed onto enamel as “temporary-tattoos” (Figure 4) (Mannoor et al., 2012). These wearable devices are thus capable of monitoring bacteria present in the mouth and more specifically on the tooth surface (Mannoor et al., 2012). The same principle has been applied very recently for detecting and identifying ingested food and liquids (Tseng et al., 2018). These mounted onto enamel nanodevices could be optimized to sense a wide variety of properties of drinks, such as alcohol content, salinity, sugars, pH, and temperature (Tseng et al., 2018). Although still in their experimental phase, such sensors represent excellent tools for the refined control and understanding of oral environment that will greatly help the field of preventive dentistry.

**Concluding remarks**

The important advances in stem cells and materials sciences are driving innovative approaches in dentistry. These progresses hold a great promise for the development of efficient and personalized treatments in the near future. At the same time, optimization of sophisticated systems for the modelling and monitoring of human tissues is leading to unprecedented possibilities for the study of diseases, diagnostics, and drug testing. Although extremely exciting, most of these approaches are not yet applicable in dental clinics. Stem cell-based dental regenerative approaches still lack reliable techniques that allow controlling stem cell behaviour upon transplantation. Similarly, state-of-the-art diagnostic systems still need to be validated in proper clinical settings. Nevertheless, these innovative approaches offer exciting perspectives to regenerative dentistry and might prove fundamental for the long-sought regeneration of fully functional dental tissues.
Acknowledgements

This work was supported by institutional funds from University of Zurich. The authors contributed to the writing, reading, and editing of the present review article. The authors confirm that there are no conflicts of interest associated with this work.

This work has been published in Frontiers in Physiology, 08 August 2018
doi: 10.3389/fphys.2018.01102

Conflict of Interest Statement

The authors declare that the research was conducted in the absence of any commercial or financial relationship that could be construed as a potential conflict of interest.

References

Aghaloo, T.L., and Moy, P.K. (2007). Which hard tissue augmentation techniques are the most successful in furnishing bony support for implant placement? Int J Oral Maxillofac Implants 22 Suppl, 49-70.

Albuquerque, M.T., Valera, M.C., Nakashima, M., Nor, J.E., and Bottino, M.C. (2014). Tissue-engineering-based strategies for regenerative endodontics. J Dent Res 93, 1222-1231.

Alvarez, P., Hee, C.K., Solchaga, L., Snel, L., Kestler, H.K., Lynch, S.E., and Hollinger, J.O. (2012). Growth factors and craniofacial surgery. J Craniomaxillofac Surg 23, 20-29.

Arakaki, M., Ishikawa, M., Nakamura, T., Iwamoto, T., Yamada, A., Fukumoto, E., Saito, M., Otsu, K., Harada, H., Yamada, Y., and Fukumoto, S. (2012). Role of epithelial-stem cell interactions during dental cell differentiation. J Biol Chem 287, 10590-10601.

Arbab, A.S., Janic, B., Haller, J., Pawelczyk, E., Liu, W., and Frank, J.A. (2009). In Vivo Cellular Imaging for Translational Medical Research. Curr Med Imaging Rev 5, 19-38.

Banerjee, A., Thompson, I.D., and Watson, T.F. (2011). Minimally invasive caries removal using bio-active glass air-abrasion. J Dent 39, 2-7.

Barbucci, R., Pasqui, D., Wirsen, A., Affrossman, S., Curtis, A., and Tetta, C. (2003). Micro and nano-structured surfaces. J. Mater. Sci.-Mater. Med. 14, 721-725.

Beraahim, Z., Moharamzadeh, K., Rawlinson, A., and Jowett, A.K. (2011). Biologic interaction of three-dimensional periodontal fibroblast spheroids with collagen-based and synthetic membranes. J Periodontol 82, 790-797.

Beuer, F., Schweiger, J., and Edelhoff, D. (2008). Digital dentistry: an overview of recent developments for CAD/CAM generated restorations. Br Dent J 204, 505-511.

Bezerra, F., Ferreira, M.R., Fontes, G.N., Da Costa Fernandes, C.J., Andia, D.C., Cruz, N.C., Da Silva, R.A., and Zambuzzi, W.F. (2017). Nano hydroxyapatite-blasted titanium surface affects pre-osteoblast morphology by modulating critical intracellular pathways. Biotechnol Bioeng 114, 1888-1898.

Bhatia, S.N., and Ingber, D.E. (2014). Microfluidic organs-on-chips. Nat Biotechnol 32, 760-772.

Bluteau, G., Ludger, H.U., De Bari, C., and Mitsiadis, T.A. (2008). Stem cells for tooth engineering. Eur Cell Mater 16, 1-9.
Bonifacio, M.A., Cometa, S., Dicarlo, M., Baruzzi, F., De Candia, S., Gloria, A., Giangregorio, M.M., Mattioli-Belmonte, M., and De Giglio, E. (2017). Gallium-modified chitosan/poly(acrylic acid) bilayer coatings for improved titanium implant performances. Carbohydr Polym 166, 348-357.

Bonnamain, V., Neveu, I., and Naveilhan, P. (2011). In vitro analyses of the immunosuppressive properties of neural stem/progenitor cells using anti-CD3/CD28-activated T cells. Methods Mol Biol 677, 233-243.

Boyde, A. (1997). Microstructure of enamel. Ciba Found Symp 205, 18-27; discussion 27-31.

Breschi, L., Maravic, T., Cunha, S.R., Comba, A., Cadenaro, M., Tjaderhane, L., Pashley, D.H., Tay, F.R., and Mazzoni, A. (2018). Dentin bonding systems: From dentin collagen structure to bond preservation and clinical applications. Dent Mater 34, 78-96.

Cantu, C., Pagella, P., Shajiei, T.D., Zimmerli, D., Valenta, T., Hausmann, G., Basler, K., and Mitsiadis, T.A. (2017). A cytoplasmic role of Wnt/beta-catenin transcriptional cofactors Bcl9, Bcl9l, and Pygopus in tooth enamel formation. Sci Signal 10.

Caton, J., Bostanci, N., Remboutsika, E., De Bari, C., and Mitsiadis, T.A. (2011). Future dentistry: cell therapy meets tooth and periodontal repair and regeneration. J Cell Mol Med 15, 1054-1065.

Chen, Z., Mauk, M.G., Wang, J., Abrams, W.R., Corstjens, P.L., Niedbala, R.S., Malamud, D., and Bau, H.H. (2007). A microfluidic system for saliva-based detection of infectious diseases. Ann N Y Acad Sci 1098, 429-436.

Clark, D., Rajendran, Y., Paydar, S., Ho, S., Cox, D., Ryder, M., Dollard, J., and Kao, R.T. (2018). Advanced platelet-rich fibrin and freeze-dried bone allograft for ridge preservation: A randomized controlled clinical trial. J Periodontol 89, 379-387.

Coelho, P.G., Zavanelli, R.A., Salles, M.B., Yeniyol, S., Tovar, N., and Jimbo, R. (2016). Enhanced Bone Bonding to Nanotextured Implant Surfaces at a Short Healing Period: A Biomechanical Tensile Testing in the Rat Femur. Implant Dent 25, 322-327.

Czilwik, G., Messinger, T., Strohmeier, O., Wadle, S., Von Stetten, F., Paust, N., Roth, G., Zengerle, R., Saarinen, P., Nittymaki, J., Mcallister, K., Sheils, O., O'leary, J., and Mark, D. (2015). Rapid and fully automated bacterial pathogen detection on a centrifugal-microfluidic LabDisk using highly sensitive nested PCR with integrated sample preparation. Lab Chip 15, 3749-3759.

Danesh-Sani, S.A., Loomer, P.M., and Wallace, S.S. (2016). A comprehensive clinical review of maxillary sinus floor elevation: anatomy, techniques, biomaterials and complications. Br J Oral Maxillofac Surg 54, 724-730.

De Angelis, N., Felice, P., Pellegrino, G., Camurati, A., Gambino, P., and Esposito, M. (2011). Guided bone regeneration with and without a bone substitute at single post-extractive implants: 1-year post-loading results from a pragmatic multicentre randomised controlled trial. Eur J Oral Implantol 4, 313-325.

Del Fabbro, M., Taschieri, S., Lodi, G., Banfi, G., and Weinstein, R.L. (2015). Magnification devices for endodontic therapy. Cochrane Database Syst Rev, CD005969.

Derosa, T.A. (2006). A retrospective evaluation of pulpotomy as an alternative to extraction. Gen Dent 54, 37-40.

Diogenes, A., and Hargreaves, K.M. (2017). Microbial Modulation of Stem Cells and Future Directions in Regenerative Endodontics. J Endod 43, S95-S101.

Diogenes, A., and Ruparel, N.B. (2017). Regenerative Endodontic Procedures: Clinical Outcomes. Dent Clin
Esposito, M., Ardebili, Y., and Worthington, H.V. (2014). Interventions for replacing missing teeth: different types of dental implants. *Cochrane Database Syst Rev*, CD003815.

Fioretti, F., Mendoza-Palomares, C., Avoaka-Boni, M.C., Ramaroson, J., Bahi, S., Richert, L., Granier, F., Benkirane-Jessel, N., and Haikel, Y. (2011). Nano-odontontology: nanostructured assemblies for endodontic regeneration. *J Biomed Nanotechnol* 7, 471-475.

Gera, A., Steinberg, G.K., and Guzman, R. (2010). In vivo neural stem cell imaging: current modalities and future directions. *Regen Med* 5, 73-86.

Godoy-Gallardo, M., Manzanares-Cespedes, M.C., Sevilla, P., Nart, J., Manzanares, N., Manero, J.M., Gil, F.J., Boyd, S.K., and Rodriguez, D. (2016). Evaluation of bone loss in antibacterial coated dental implants: An experimental study in dogs. *Mater Sci Eng C Mater Biol Appl* 69, 538-545.

Goracci, C., Cadenaro, M., Fontanive, L., Giangrosso, G., Juloski, J., Vichi, A., and Ferrari, M. (2014). Polymerization efficiency and flexural strength of low-stress restorative composites. *Dent Mater* 30, 688-694.

Gronthos, S., Brahim, J., Li, W., Fisher, L.W., Cherman, N., Boyde, A., Denbesten, P., Robey, P.G., and Shi, S. (2002). Stem cell properties of human dental pulp stem cells. *J Dent Res* 81, 531-535.

Gronthos, S., Mankani, M., Brahim, J., Robey, P.G., and Shi, S. (2000). Postnatal human dental pulp stem cells (DPSCs) in vitro and in vivo. *Proc Natl Acad Sci U S A* 97, 13625-13630.

Guazzato, M., Albakry, M., Ringer, S.P., and Swain, M.V. (2004). Strength, fracture toughness and microstructure of a selection of all-ceramic materials. Part II. Zirconia-based dental ceramics. *Dent Mater* 20, 449-456.

Hammerle, C.H., Stone, P., Jung, R.E., Kapos, T., and Brodala, N. (2009). Consensus statements and recommended clinical procedures regarding computer-assisted implant dentistry. *Int J Oral Maxillofac Implants* 24 Suppl, 126-131.

Hancocks, S. (2017). What is digital about dentistry? *Br Dent J* 223, 305.

Hayashi, Y., Murakami, M., Kawamura, R., Ishizaka, R., Fukuta, O., and Nakashima, M. (2015). CXCL14 and MCP1 are potent trophic factors associated with cell migration and angiogenesis leading to higher regenerative potential of dental pulp side population cells. *Stem Cell Res Ther* 6, 111.

Heo, D.N., Ko, W.K., Lee, H.R., Lee, S.J., Lee, D., Um, S.H., Lee, J.H., Woo, Y.H., Zhang, L.G., Lee, D.W., and Kwon, I.K. (2016). Titanium dental implants surface-immobilized with gold nanoparticles as osteoinductive agents for rapid osseointegration. *J Colloid Interface Sci* 469, 129-137.

Howell, T.H., Fiorellini, J.P., Paquette, D.W., Ofenbacher, S., Giannobile, W.V., and Lynch, S.E. (1997). A phase I/II clinical trial to evaluate a combination of recombinant human platelet-derived growth factor-BB and recombinant human insulin-like growth factor-I in patients with periodontal disease. *J Periodontol* 68, 1186-1193.

Ilie, N., Kessler, A., and Durner, J. (2013). Influence of various irradiation processes on the mechanical properties and polymerization kinetics of bulk-fill resin based composites. *J Dent* 41, 695-702.

Ingber, D.E. (2016). Reverse Engineering Human Pathophysiology with Organs-on-Chips. *Cell* 164, 1105-1109.
Iwatsuki, S., Honda, M.J., Harada, H., and Ueda, M. (2006). Cell proliferation in teeth reconstructed from dispersed cells of embryonic tooth germs in a three-dimensional scaffold. *Eur J Oral Sci* 114, 310-317.

Jimenez-Rojo, L., Granchi, Z., Graf, D., and Mitsiadis, T.A. (2012). Stem Cell Fate Determination during Development and Regeneration of Ectodermal Organs. *Front Physiol* 3, 107.

Kaigler, D., Avila-Ortiz, G., Travan, S., Taut, A.D., Padial-Molina, M., Rudek, I., Wang, F., Lanis, A., and Giannobile, W.V. (2015). Bone Engineering of Maxillary Sinus Bone Deficiencies Using Enriched CD90+ Stem Cell Therapy: A Randomized Clinical Trial. *J Bone Miner Res* 30, 1206-1216.

Kaigler, D., Pagni, G., Park, C.H., Braun, T.M., Holman, L.A., Yi, E., Tarle, S.A., Bartel, R.L., and Giannobile, W.V. (2013). Stem cell therapy for craniofacial bone regeneration: a randomized, controlled feasibility trial. *Cell Transplant* 22, 767-777.

Kilambi, H., Cramer, N.B., Schneidewind, L.H., Shah, P., Stansbury, J.W., and Bowman, C.N. (2009). Evaluation of highly reactive mono-methacrylates as reactive diluents for BisGMA-based dental composites. *Dent Mater* 25, 33-38.

Klein, C., Le Goff, C., Topouchian, V., Odent, S., Violas, P., Glorion, C., and Cormier-Daire, V. (2014). Orthopedics management of acromicric dysplasia: follow up of nine patients. *Am J Med Genet A* 164A, 331-337.

Larsson, L., Decker, A.M., Nibali, L., Pilipchuk, S.P., Berglundh, T., and Giannobile, W.V. (2016). Regenerative Medicine for Periodontal and Peri-implant Diseases. *J Dent Res* 95, 255-266.

Lelli, M., Putignano, A., Marchetti, M., Foltran, I., Mangani, F., Procaccini, M., Roveri, N., and Orsini, G. (2014). Remineralization and repair of enamel surface by biomimetic Zn-carbonate hydroxyapatite containing toothpaste: a comparative in vivo study. *Front Physiol* 5, 333.

Levato, C.M., Farman, A.G., and Miles, D.A. (2015). The "inevitability" of digital radiography in dentistry. *Compend Contin Educ Dent* 36, 238, 240.

Lindhe, J., Haffajee, A.D., and Socransky, S.S. (1983). Progression of periodontal disease in adult subjects in the absence of periodontal therapy. *J Clin Periodontol* 10, 433-442.

Luiz De Oliveira Da Rosa, W., Machado Da Silva, T., Fernando Demarco, F., Piva, E., and Fernandes Da Silva, A. (2017). Could the application of bioactive molecules improve vital pulp therapy success? A systematic review. *J Biomed Mater Res A* 105, 941-956.

Lynch, C. (2017). Defining digital dentistry: A survey of recent literature. *J Dent* 59, 1.

Lynch, S.E. (1992). Methods for evaluation of regenerative procedures. *J Periodontol* 63, 1085-1092.

Lynch, S.E., Buser, D., Hernandez, R.A., Weber, H.P., Stich, H., Fox, C.H., and Williams, R.C. (1991a). Effects of the platelet-derived growth factor/insulin-like growth factor-I combination on bone regeneration around titanium dental implants. Results of a pilot study in beagle dogs. *J Periodontol* 62, 710-716.

Lynch, S.E., De Castilla, G.R., Williams, R.C., Kiritsy, C.P., Howell, T.H., Reddy, M.S., and Antoniades, H.N. (1991b). The effects of short-term application of a combination of platelet-derived and insulin-like growth factors on periodontal wound healing. *J Periodontol* 62, 458-467.

Lynch, S.E., Williams, R.C., Polson, A.M., Howell, T.H., Reddy, M.S., Zappa, U.E., and Antoniades, H.N. (1989). A combination of platelet-derived and insulin-like growth factors enhances periodontal regeneration. *J Clin Periodontol* 16, 545-548.
Mannoor, M.S., Tao, H., Clayton, J.D., Sengupta, A., Kaplan, D.L., Naik, R.R., Verma, N., Omenetto, F.G., and Mcalpine, M.C. (2012). Graphene-based wireless bacteria detection on tooth enamel. *Nat Commun* 3, 763.

Mazzoni, A., Angeloni, V., Comba, A., Maravic, T., Cadenaro, M., Tezvergil-Mutluay, A., Pashley, D.H., Tay, F.R., and Breschi, L. (2018). Cross-linking effect on dentin bond strength and MMPs activity. *Dent Mater* 34, 288-295.

Mendonca, G., Mendonca, D.B., Aragao, F.J., and Cooper, L.F. (2008). Advancing dental implant surface technology--from micron- to nanotopography. *Biomaterials* 29, 3822-3835.

Miquel, A. (2011). [In the 11-M terrorist tragedy in Madrid]. *Rev Clin Esp* 211, 158-162.

Miron, R.J., Chandad, F., Buser, D., Sculean, A., Cochran, D.L., and Zhang, Y. (2016a). Effect of Enamel Matrix Derivative Liquid on Osteoblast and Periodontal Ligament Cell Proliferation and Differentiation. *J Periodontol* 87, 91-99.

Miron, R.J., Sculean, A., Cochran, D.L., Froum, S., Zucchelli, G., Nemcovsky, C., Donos, N., Lyngstadaas, S.P., Deschner, J., Dard, M., Stavropoulos, A., Zhang, Y., Trombelli, L., Kasaj, A., Shirakata, Y., Cortellini, P., Tonetti, M., Rasperini, G., Jepsen, S., and Bosshardt, D.D. (2016b). Twenty years of enamel matrix derivative: the past, the present and the future. *J Clin Periodontol* 43, 668-683.

Miron, R.J., Zucchelli, G., Pikos, M.A., Salama, M., Lee, S., Guillemette, V., Fujioka-Kobayashi, M., Bishara, M., Zhang, Y., Wang, H.L., Chandad, F., Nacopoulos, C., Simonpieri, A., Aalam, A.A., Felice, P., Sammartino, G., Ghanaati, S., Hernandez, M.A., and Choukroun, J. (2017). Use of platelet-rich fibrin in regenerative dentistry: a systematic review. *Clin Oral Investig* 21, 1913-1927.

Mitsiadis, T.A., and Graf, D. (2009). Cell fate determination during tooth development and regeneration. *Birth Defects Res C Embryo Today* 87, 199-211.

Mitsiadis, T.A., and Harada, H. (2015). Regenerated teeth: the future of tooth replacement. An update. *Regen Med* 10, 5-8.

Mitsiadis, T.A., and Luder, H.U. (2011). Genetic basis for tooth malformations: from mice to men and back again. *Clin Genet* 80, 319-329.

Mitsiadis, T.A., Orsini, G., and Jimenez-Rojo, L. (2015). Stem cell-based approaches in dentistry. *Eur Cell Mater* 30, 248-257.

Mitsiadis, T.A., and Papagerakis, P. (2011). Regenerated teeth: the future of tooth replacement? *Regen Med* 6, 135-139.

Mitsiadis, T.A., Woloszyk, A., and Jimenez-Rojo, L. (2012). Nanodentistry: combining nanostructured materials and stem cells for dental tissue regeneration. *Nanomedicine (Lond)* 7, 1743-1753.

Miura, M., Gronthos, S., Zhao, M., Lu, B., Fisher, L.W., Robey, P.G., and Shi, S. (2003). SHED: stem cells from human exfoliated deciduous teeth. *Proc Natl Acad Sci U S A* 100, 5807-5812.

Monterubbianesi, R., Orsini, G., Tosi, G., Conti, C., Librando, V., Procaccini, M., and Putignano, A. (2016). Spectroscopic and Mechanical Properties of a New Generation of Bulk Fill Composites. *Front Physiol* 7, 652.

Murray, P.E., and García-Godoy, F. (2006). The outlook for implants and endodontics: a review of the tissue engineering strategies to create replacement teeth for patients. *Dent Clin North Am* 50, 299-315, x.

Nakashima, M., and Iohara, K. (2017). Recent Progress in Translation from Bench to a Pilot Clinical Study on Total Pulp Regeneration. *J Endod* 43, S82-S86.
Nakashima, M., Iohara, K., Murakami, M., Nakamura, H., Sato, Y., Ariji, Y., and Matsushita, K. (2017). Pulp regeneration by transplantation of dental pulp stem cells in pulpitis: a pilot clinical study. *Stem Cell Res Ther* **8**, 61.

Natsiou, D., Granchi, Z., Mitsiadis, T.A., and Jimenez-Rojo, L. (2017). Generation of Spheres from Dental Epithelial Stem Cells. *Front Physiol* **8**, 7.

Orsini, G., Jimenez-Rojo, L., Natsiou, D., Putignano, A., and Mitsiadis, T.A. (2015). In vivo administration of dental epithelial stem cells at the apical end of the mouse incisor. *Front Physiol* **6**, 112.

Orsini, G., Procaccini, M., Manzoli, L., Sparabombe, S., Tiriduzzi, P., Bambini, F., and Putignano, A. (2013). A 3-day randomized clinical trial to investigate the desensitizing properties of three dentifrices. *J Periodontol* **84**, e65-73.

Oshima, M., and Tsuji, T. (2014). Functional tooth regenerative therapy: tooth tissue regeneration and whole-tooth replacement. *Odontology* **102**, 123-136.

Otsu, K., Kumakami-Sakano, M., Fujiwara, N., Kikuchi, K., Keller, L., Lesot, H., and Harada, H. (2014). Stem cell sources for tooth regeneration: current status and future prospects. *Front Physiol* **5**, 36.

Ozcan, M., and Jonasch, M. (2018). Effect of Cyclic Fatigue Tests on Aging and Their Translational Implications for Survival of All-Ceramic Tooth-Borne Single Crowns and Fixed Dental Prostheses. *J Prosthodont* **27**, 364-375.

Pagella, P., Miran, S., and Mitsiadis, T. (2015). Analysis of Developing Tooth Germ Innervation Using Microfluidic Co-culture Devices. *J Vis Exp*, e53114.

Pagella, P., Neto, E., Jimenez-Rojo, L., Lamghari, M., and Mitsiadis, T.A. (2014). Microfluidics co-culture systems for studying tooth innervation. *Front Physiol* **5**, 326.

Papagerakis, P., Mitsiadis, T. (2013). *Development and structure of teeth and periodontal tissues.*: John Wiley & Sons, Inc.

Pilipchuk, S.P., Plonka, A.B., Monje, A., Taut, A.D., Lanis, A., Kang, B., and Giannobile, W.V. (2015). Tissue engineering for bone regeneration and osseointegration in the oral cavity. *Dent Mater* **31**, 317-338.

Piva, E., Silva, A.F., and Nor, J.E. (2014). Functionalized scaffolds to control dental pulp stem cell fate. *J Endod* **40**, S33-40.

Pokrowiecki, R., Zareba, T., Szaraniec, B., Palka, K., Mielczarek, A., Menaszek, E., and Tyski, S. (2017). In vitro studies of nanosilver-doped titanium implants for oral and maxillofacial surgery. *Int J Nanomedicine* **12**, 4285-4297.

Prantil-Baun, R., Novak, R., Das, D., Somayaji, M.R., Przekwas, A., and Ingber, D.E. (2018). Physiologically Based Pharmacokinetic and Pharmacodynamic Analysis Enabled by Microfluidically Linked Organs-on-Chips. *Annu Rev Pharmacol Toxicol* **58**, 37-64.

Rickert, D., Sauerbier, S., Nagursky, H., Menne, D., Vissink, A., and Raghoebear, G.M. (2011). Maxillary sinus floor elevation with bovine bone mineral combined with either autogenous bone or autogenous stem cells: a prospective randomized clinical trial. *Clin Oral Implants Res* **22**, 251-258.

Sauerbier, S., Rickert, D., Gutwald, R., Nagursky, H., Oshima, T., Xavier, S.P., Christmann, J., Kurz, P., Menne, D., Vissink, A., Raghoebear, G., Schmelzeisen, R., Wagner, W., and Koch, F.P. (2011). Bone marrow concentrate and bovine bone mineral for sinus floor augmentation: a controlled, randomized, single-blinded clinical and histological trial–per-protocol analysis. *Tissue Eng Part A* **17**, 2187-2197.
Scarano, A., Degidi, M., Iezzi, G., Pecora, G., Piattelli, M., Orsini, G., Caputi, S., Perrotti, V., Mangano, C., and Piattelli, A. (2006). Maxillary sinus augmentation with different biomaterials: a comparative histologic and histomorphometric study in man. *Implant Dent* 15, 197-207.

Selvig, K.A., Soërensen, R.G., Wozney, J.M., and Wikesjo, U.M. (2002). Bone repair following recombinant human bone morphogenetic protein-2 stimulated periodontal regeneration. *J Periodontol* 73, 1020-1029.

Seo, B.M., Miura, M., Gronthos, S., Bartold, P.M., Batouli, S., Brahim, J., Young, M., Robey, P.G., Wang, C.Y., and Shi, S. (2004). Investigation of multipotent postnatal stem cells from human periodontal ligament. *Lancet* 364, 149-155.

Sheik, Z., Hamdan, N., Ikeda, Y., Grynpas, M., Ganss, B., and Glogauer, M. (2017). Natural graft tissues and synthetic biomaterials for periodontal and alveolar bone reconstructive applications: a review. *Biomater Res* 21, 9.

Shinmura, Y., Tsuchiya, S., Hata, K., and Honda, M.J. (2008). Quiescent epithelial cell rests of Malassez can differentiate into ameloblast-like cells. *J Cell Physiol* 217, 728-738.

Singh, P. (2011). Understanding peri-implantitis: a strategic review. *J Oral Implantol* 37, 622-626.

Skardal, A., Shupe, T., and Atala, A. (2016). Organoid-on-a-chip and body-on-a-chip systems for drug screening and disease modeling. *Drug Discov Today* 21, 1399-1411.

Slavkin, H.C., and Baum, B.J. (2000). Relationship of dental and oral pathology to systemic illness. *JAMA* 284, 1215-1217.

Song, J.S., Takimoto, K., Jeon, M., Vadakekalam, J., Ruparel, N.B., and Diogenes, A. (2017). Decellularized Human Dental Pulp as a Scaffold for Regenerative Endodontics. *J Dent Res* 96, 640-646.

Takahashi, K., and Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 126, 663-676.

Torabinejad, M., and Faras, H. (2012). A clinical and histological report of a tooth with an open apex treated with regenerative endodontics using platelet-rich plasma. *J Endod* 38, 864-868.

Tseng, P., Napier, B., Garbarini, L., Kaplan, D.L., and Omenetto, F.G. (2018). Functional, RF-Trilayer Sensors for Tooth-Mounted, Wireless Monitoring of the Oral Cavity and Food Consumption. *Adv Mater* 30, e1703257.

Variola, F., Brunski, J.B., Orsini, G., Tambasco De Oliveira, P., Wazen, R., and Nanci, A. (2011). Nanoscale surface modifications of medically relevant metals: state-of-the art and perspectives. *Nanoscale* 3, 335-353.

Variola, F., Vetrone, F., Richert, L., Jedrzejowski, P., Yi, J.-H., Zalzal, S., Clair, S., Sarkissian, A., Perepichka, D.F., Wuest, J.D., Rosei, F., and Nanci, A. (2009). Improving Biocompatibility of Implantable Metals by Nanoscale Modification of Surfaces: An Overview of Strategies, Fabrication Methods, and Challenges. *Small* 5, 996-1006.

Volponi, A.A., Pang, Y., and Sharpe, P.T. (2010). Stem cell-based biological tooth repair and regeneration. *Trends Cell Biol* 20, 715-722.

Washio, K., Iwata, T., Mizutani, M., Ando, T., Yamato, M., Okano, T., and Ishikawa, I. (2010). Assessment of cell sheets derived from human periodontal ligament cells: a pre-clinical study. *Cell Tissue Res* 341, 397-404.

Weeber, F., Ooft, S.N., Dijkstra, K.K., and Voest, E.E. (2017). Tumor Organoids as a Pre-clinical Cancer Model
Wittneben, J.G., Gavric, J., Belser, U.C., Bornstein, M.M., Joda, T., Chappuis, V., Sailer, I., and Bragger, U. (2017). Esthetic and Clinical Performance of Implant-Supported All-Ceramic Crowns Made with Prefabricated or CAD/CAM Zirconia Abutments: A Randomized, Multicenter Clinical Trial. J Dent Res 96, 163-170.

Wu, J., Dong, M., Santos, S., Rigatto, C., Liu, Y., and Lin, F. (2017). Lab-on-a-Chip Platforms for Detection of Cardiovascular Disease and Cancer Biomarkers. Sensors (Basel) 17.

Yamauchi, N., Yamauchi, S., Nagaoka, H., Duggan, D., Zhong, S., Lee, S.M., Teixeira, F.B., and Yamauchi, M. (2011). Tissue engineering strategies for immature teeth with apical periodontitis. J Endod 37, 390-397.

Yang, B., Chen, G., Li, J., Zou, Q., Xie, D., Chen, Y., Wang, H., Zheng, X., Long, J., Tang, W., Guo, W., and Tian, W. (2012). Tooth root regeneration using dental follicle cell sheets in combination with a dentin matrix-based scaffold. Biomaterials 33, 2449-2461.

Yang, W.F., Choi, W.S., Leung, Y.Y., Curtin, J.P., Du, R., Zhang, C.Y., Chen, X.S., and Su, Y.X. (2018). Three-dimensional printing of patient-specific surgical plates in head and neck reconstruction: A prospective pilot study. Oral Oncol 78, 31-36.

Yin, X., Mead, B.E., Safaee, H., Langer, R., Karp, J.M., and Levy, O. (2016). Engineering Stem Cell Organoids. Cell Stem Cell 18, 25-38.

Yu, J., Xu, L., Li, K., Xie, N., Xi, Y., Wang, Y., Zheng, X., Chen, X., Wang, M., and Ye, X. (2017). Zinc-modified Calcium Silicate Coatings Promote Osteogenic Differentiation through TGF-beta/Smad Pathway and Osseointegration in Osteopenic Rabbits. Sci Rep 7, 3440.

Zhang, F., Chevalier, J., Olagnon, C., Batuk, M., Hadermann, J., Van Meerbeek, B., and Vleugels, J. (2017). Grain-Boundary Engineering for Aging and Slow-Crack-Growth Resistant Zirconia. J Dent Res 96, 774-779.

Zhou, W., Liu, Z., Song, L., Kuo, C.L., and Shafer, D.M. (2018). Clinical Factors Affecting the Accuracy of Guided Implant Surgery-A Systematic Review and Meta-analysis. J Evid Based Dent Pract 18, 28-40.

**Keywords**
tooth; dental implants; dental treatment; stem cells; dental pulp; organ-on-chip; organoids

Retrieved from https://encyclopedia.pub/617