Recent advances in the diagnosis and management of primary myelofibrosis

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Primary myelofibrosis (PMF) is a myeloproliferative neoplasm (MPN) in which dysregulation of the Janus kinase/signal transducers and activators of transcription (JAK/STAT) signaling pathways is the major pathogenic mechanism. Most patients with PMF carry a driver mutation in the JAK2, MPL (myeloproliferative leukemia), or CALR (calreticulin) genes. Mutations in epigenetic regulators and RNA splicing genes may also occur, and play critical roles in PMF disease progression. Based on revised World Health Organization diagnostic criteria for MPNs, both screening for driver mutations and bone marrow biopsy are required for a specific diagnosis. Clinical trials of JAK2 inhibitors for PMF have revealed significant efficacy for improving splenomegaly and constitutional symptoms. However, the currently available drug therapies for PMF do not improve survival. Although allogeneic stem cell transplantation is potentially curative, it is associated with substantial treatment-related morbidity and mortality. PMF is a heterogeneous disorder and decisions regarding treatments are often complicated, necessitating the use of prognostic models to determine the management of treatments for individual patients. This review focuses on the clinical aspects and outcomes of a cohort of Japanese patients with PMF, including discussion of recent advances in the management of PMF.

Keywords: Primary myelofibrosis; Prognostic score; Long-term outcome

INTRODUCTION

Primary myelofibrosis (PMF), classified as a clonal myeloproliferative neoplasm (MPN), is characterized by the progressive proliferation of mainly granulocytic and megakaryocytic cells in the bone marrow. This condition leads to bone marrow fibrosis, resulting in subsequent extramedullary hematopoiesis and splenomegaly [1]. Typical clinical features of PMF include progressive anemia, symptomatic splenomegaly, and various constitutional symptoms [2-4]. Transformation to acute leukemia occurs in up to 20% of patients and this, along with progressive disease and infection, is one of the major causes of PMF-related death [2-4]. PMF is associated with a poor prognosis and a marked reduction in life expectancy [4-6], with median survival ranging from 3.5 to 6 years [7-11]. The molecular pathogenesis of PMF is characterized by dysregulation of the Janus kinase/signal transducers and activators of transcription (JAK/STAT) signaling pathways, which are crucial for normal cytokine-mediated cell responses [12-14]. Approximately 85% of patients with PMF carry driver mutations in genes in these pathways [13], including JAK2V617F [15-18], MPLW515 [19,20], and calreticulin (CALR) [21,22]. In addition, there may be mutations in epigenetic regulators and RNA splicing genes that co-exist with driver mutations and play critical roles in disease progression [12,13]. In 2016, the World Health Organization revised...
The diagnostic criteria for MPN so that the diagnosis of PMF now requires screening for driver mutations in \textit{JAK2}, \textit{CALR}, and myeloproliferative leukemia (\textit{MPL}) along with bone marrow biopsy (Table 1) [23]. If a patient has none of the driver mutations, which is a condition referred to as triple-negative disease, screening for non-driver mutations such as in additional sex combs like 1 (\textit{ASXL1}), enhancer of zeste 2 polycomb repressive complex 2 subunit (\textit{EZH2}), tet methylcytosine dioxygenase 2 (\textit{TET2}), isocitrate dehydrogenase 1/2 (\textit{IDH1/2}), serine and arginine rich splicing factor 2 (\textit{SRSF2}), and splicing factor 3b subunit 1 (\textit{SF3B1}) is suggested. Another major revision was the identification of prefibrotic myelofibrosis as a new disease category separate from PMF and essential thrombocythemia (\textit{ET}); distinguishing among these entities may have prognostic significance.

Current drug therapies for PMF, such as erythropoiesis-stimulating agents, hydroxyurea, and immunomodulatory drugs, do not improve survival. Although allogeneic stem cell transplantation (alloSCT) is potentially curative [3,24-26], it is associated with substantial treatment-related morbidity and mortality and is therefore only used in a minority of cases. Two phase III trials revealed that the JAK2 inhibitor ruxolitinib exhibits sustained efficacy and improves survival [27]. Decisions regarding the treatment of patients with PMF are often complicated, particularly with respect to the timing of alloSCT or the participation in clinical trials for novel drugs. Treatments are usually planned based on each patient’s estimated prognosis. Although several studies have addressed survival and prognostic factors in cohorts of patients with PMF, there is little such research in Asia to guide therapeutic decision-making. Accordingly, we started an annual nationwide survey of PMF cases in Japan in 1999, enrolling a total of 780 patients over a 17-year study period [28]. This review focuses on

| Major criteria | PrePMF | PMF |
|---------------|--------|-----|
| 1. Megakaryocytic proliferation and atypia, without reticulin fibrosis > grade 1, accompanied by increased age-adjusted BM cellularity, granulocytic proliferation and often decreased erythropoiesis | 1. Presence of megakaryocytic proliferation and atypia, accompanied by either reticulin and/or collagen fibrosis grades 2 or 3 |
| 2. Not meeting the WHO criteria for BCR-ABL1+ CML, PV, ET, myelodysplastic syndromes, or other myeloid neoplasms | 2. Not meeting WHO criteria for ET, PV, BCR-ABL1+ CML, myelodysplastic syndromes, or other myeloid neoplasms |
| 3. Presence of \textit{JAK2}, \textit{CALR}, or \textit{MPL} mutation or in the absence of these mutations, presence of another clonal marker or absence of minor reactive BM reticulin fibrosis | 3. Presence of \textit{JAK2}, \textit{CALR}, or \textit{MPL} mutation or in the absence of these mutations, presence of another clonal marker or absence of reactive myelofibrosis |

| Minor criteria | PrePMF | PMF |
|---------------|--------|-----|
| Presence of at least 1 of the following, confirmed in 2 consecutive determinations: | Presence of at least 1 of the following, confirmed in 2 consecutive determinations: |
| a. Anemia not attributed to a comorbid condition | a. Anemia not attributed to a comorbid condition |
| b. Leukocytosis > 11 × 10^9/L | b. Leukocytosis > 11 × 10^9/L |
| c. Palpable splenomegaly | c. Palpable splenomegaly |
| d. LDH increased to above upper normal limit of institutional reference range | d. LDH increased to above upper normal limit of institutional reference range |
| e. Leukoerythroblastosis | e. Leukoerythroblastosis |

Diagnosis | All three major criteria, and at least one minor criterion |

Adapted from Arber et al., with permission from American Society of Hematology [23].

WHO, World Health Organization; prePMF, prefibrotic primary myelofibrosis; PMF, primary myelofibrosis; BM, bone marrow; BCR-ABL1, breakpoint cluster region-Abelson 1; CML, chronic myeloid leukemia; PV, polycythemia vera; ET, essential thrombocythemia; JAK2, Janus kinase 2; CALR, calreticulin; MPL, myeloproliferative leukemia; LDH, lactate dehydrogenase.
the clinical aspects and outcome of patients with PMF in the Japanese registry and discusses recent advances in the management of PMF.

CLINICAL FEATURES OF PMF

Clinical features at presentation
Most patients with PMF are aged > 60 years at initial diagnosis, and the reported median age at diagnosis ranges from 69 to 79 years [29]. The incidence of PMF in several registries ranges from 0.1 per 100,000 individuals per year to 1 per 100,000 per year [29,30]. The clinical manifestations of PMF vary. Most patients present with anemia, splenomegaly, and constitutional symptoms [2], but up to 30% of patients are asymptomatic at diagnosis. Splenomegaly can cause abdominal pain, abdominal discomfort, and early satiety. Constitutional symptoms include low-grade fever, night sweats, and weight loss. Some patients present with headache, inactivity, fatigue, insomnia, pruritus, bone pain, or thrombosis [31,32]; these symptoms significantly lower the patient’s quality of life (QOL). Various scales (e.g., European Organization for the Research and Treatment of Cancer [EORTC] QLQ-30, Functional Assessment of Cancer Therapy-Lymphoma [FACT-Lym], and the modified Myelofibrosis Symptom Assessment Form [MF-SAF]) are used to assess symptoms and QOL [33-35].

Driver mutation status and cytogenetic analysis
As noted above, most patients with PMF carry one of three mutually exclusive somatic driver mutations [13]; JAK2V617F in up to 60%, CALR in up to 20%, and MPL in up to 5% of patients. Additional non-driver mutations associated with epigenetic modification (TET2, ASXL1, EZH2, IDH1/2), RNA splicing (SRSF2, SF3B1, U2AF1 [U2 small nuclear RNA auxiliary factor 1]), JAK/STAT signaling (CBL [Casitas B-cell lymphoma], LNK), and DNA repair (TP53 [tumor protein p53]) are found in 1% to 10% of patients. In cytogenetic analyses, chromosomal abnormalities such as complex karyotypes and single or double abnormalities, including +8, del(7)/7q-, 12p-, inv(3), and 11q23 rearrangements, are defined as unfavorable karyotypes [36].

Clinical features of PMF in the Japanese registry cohort
As previously reported [28], questionnaires were sent annually to approximately 500 hematology departments of board-certified member institutes of the Japanese Society of Hematology. Patients newly diagnosed with PMF between 1999 and 2015 were entered into the registry and followed annually to collect information on outcome. The average response rate to the questionnaires was 48.6% (range, 44.7% to 49.7%). Approximately 50 patients were entered per year, yielding an eventual total of 780 patients with PMF in the cohort. The median follow-up at the time of analysis was 23 months and the median age at diagnosis was 66 years (range, 19 to 96) [28]. Low blood cell counts or other abnormal laboratory results were the most frequent reasons for initial consultation with a hematologist. Only 20% of patients initially presented with symptoms such as fatigue, weight loss, palpitations, shortness of breath, dizziness, abdominal fullness, fever, or abdominal pain. Splenomegaly was present in 75% of patients, and up to 70% had anemia, defined as a hemoglobin concentration < 10 g/dL. More than half of the patients had leukoerythroblastosis and a circulating blast frequency ≥ 1%. The clinical features, age distribution, and male-to-female ratios observed in our cohort were similar to those reported in European and North American studies [2,9,36,37]. Since items other than weight change for symptom assessment were not included in our registry data, symptom burden was not evaluated using recent scoring systems in our analysis.

In our series, the JAK2V617F mutation was detected in 56% of the patients, but the MPL and CALR mutations were not found in most subjects. However, in a previously published study from our institution, the JAK2 mutation, an MPL exon 10 mutation, and a CALR exon 9 mutation were found in up to 60%, up to 2%, and 25% of the patients, respectively [38]. Ten percent of patients had no detectable mutations (i.e., triple-negative disease), and none had overlapping mutations. These results are consistent with data from larger cohort studies [12,13]. Bone marrow or blood karyotype analysis revealed abnormalities in 34% of cases. The most frequent abnormal karyotypes were del(20) and del(13). Unfavorable karyotypes were detected in 15% of the patients [28].

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OUTCOME OF PMF

Survival
Long-term survival data in PMF patients were reported by the Mayo Clinic-Italian collaborative group, with a median survival of 5.9 years among 267 patients [39]. Patients with PMF had a worse survival than those with polycythemia vera (PV) (median survival, 13.5 years) or ET (median survival, 19.8 years). A similar result was reported by a Swedish group. In their population-based study, patients with PMF had higher relative survival ratios (RSRs), defined as the observed survival in the patient group divided by the expected survival of a comparable group from the general population, compared with those with PV and ET; the 10- and 20-year RSRs were 0.21 and 0.06 for PMF, 0.64 and 0.32 for PV, and 0.68 and 0.44 for ET, respectively [10].

Survival and causes of death in the Japanese registry cohort
In the Japanese registry cohort, the median survival was 47 months (95% confidence interval [CI], 41 to 53) at the time of the analysis. The 3-year overall survival (OS) was 59% (95% CI, 55% to 63%) (Fig. 1) [28]. These values are worse than those reported previously [7-11]. However, as the registry cohort was based on a questionnaire survey, it might have included patients with more advanced disease. We did not observe a plateau in survival curves during follow-up, indicating a persistent mortality rate that was attributable to PMF. Unfortunately, no improvement in survival was observed over the 17-year study period. Of the patients in the registry for whom the final cause of death was known, infection and leukemic transformation were the most frequent causes, followed by bleeding, PMF progression without transformation to leukemia, heart failure, and other malignancies (Table 2) [28].

ASSESSING PROGNOSIS IN PMF

Prognostic scoring systems
Several prognostic scoring systems have been established for PMF. The International Prognostic Scoring System (IPSS), developed in 2009, utilizes five independent risk factors at the time of diagnosis to predict survival: age > 65 years; hemoglobin level < 10 g/dL; leukocyte count > 25 × 10⁹/L; circulating blast frequency ≥ 1%; and the presence of constitutional symptoms (Tables 3 and 4) [2]. The IPSS was subsequently refined to the Dynamic IPSS (DIPSS) in 2010 [37] and the DIPSS-plus in 2011 [36]. The DIPSS-plus incorporates information such as an unfavorable karyotype, need for red cell transfusion, and platelet count < 100 × 10⁹/L, and stratifies patients into four risk groups: low (no risk factors); intermediate-1 (one risk factor); intermediate-2 (two or three risk factors); and high (four or more risk factors), with median OS of 185, 78, 35, and 16 months, respectively. The advantage of DIPSS and DIPSS-plus is that they can be applied to patients at any time during the disease course. These scoring systems are based on easily assessable clinical characteristics and blood counts, and are currently the standard methods used to estimate the prognosis of individual patients.

More recently, a number of genetic risk factors independent of DIPSS-plus were identified, including driver gene mutational status [39-42], JAK2 allele burden [43], and the presence or number of other non-driver somatic mutations [44,45]. In most patients with PMF with one of the abovementioned three driver mutations, Tefferi
et al. [40] showed that the CALR mutation was associated with a better outcome than the other two driver mutations, independent of the DIPSS-plus estimated risk. In addition, patients with triple-negative disease had a much higher incidence of leukemic transformation and an unfavorable outcome. As for non-driver mutations, the presence of mutations in ASXL1, EZH2, SRSF2, or IDH1/2 is defined as high molecular risk, as these mutations are associated with poor prognosis and risk of leukemic transformation [44]. The number of these detrimental mutations is also an adverse risk factor, and the presence of two or more non-driver mutations predicts the worst survival [45]. Notably, these molecular risk factors predict a poor outcome independent of conventional prognostic scoring systems.

### Stratification according to PMF prognostic scoring in the Japanese registry cohort

The registry cohort was divided into four prognostic groups according to the various prognostic models reported previously (Fig. 2) [28]. Both IPSS and DIPSS differentiated between low and intermediate-1 risk patients, but could not distinguish high risk from intermediate-2 risk. In contrast, the DIPSS-plus model accurately divided the population into four statistically different risk groups. The inclusion of the need for red cell transfusion and an unfavorable karyotype are likely the factors that account for the good performance

| Variable                                           | IPSS [2] | DIPSS [29] | DIPSS plus [30] |
|----------------------------------------------------|----------|------------|-----------------|
| Age > 65 years                                     | ✓        | ✓          | ✓               |
| Constitutional symptoms^[a]                         | ✓        |            |                 |
| Hb < 10 g/dL                                       | ✓        | ✓          | ✓               |
| WBC > 25,000/μL                                    | ✓        | ✓          | ✓               |
| Peripheral blood blasts ≥ 1%                       | ✓        | ✓          | ✓               |
| Platelets < 10 × 10^3/μL                           | ✓        |            |                 |
| Red cell transfusion need^[b]                      | ✓        |            |                 |
| Unfavorable karyotype^[c]                          | ✓        |            |                 |
| Point per variable                                  | 1 point each | 1 point each but Hb = 2 | 1 point each |

IPSS, International Prognostic Scoring System; DIPSS, Dynamic IPSS; DIPSS plus, Dynamic IPSS plus additional prognostic factors; Hb, hemoglobin; WBC, white blood cell.

^[a] Weight loss 10% of the baseline value in the year preceding primary myelofibrosis diagnosis and/or unexplained fever or excessive sweats persisting for more than 1 month.

^[b] Red blood cell (RBC) transfusion at the time of referral and those with history of RBC transfusions, for myelofibrosis-associated anemia.

^[c] Complex karyotype or single or two abnormalities including +8, -7/7q-, i(17q), -5/5q-, 12p-, inv(3), or 11q23 rearrangements.

### Table 2. Cause of death in patients with primary myelofibrosis in Japanese registry data [28]

| Events                               | No. of patients (%) |
|--------------------------------------|---------------------|
| Leukemic transformation              | 91 (24)             |
| Infection                            | 89 (24)             |
| Bleeding (brain or gut)              | 36 (10)             |
| Primary disease                      | 28 (7)              |
| Heart failure                        | 20 (5)              |
| Other malignancies                   | 14 (4)              |
| Liver failure                        | 12 (3)              |
| Multigener failure                   | 10 (3)              |
| Respiratory failure                  | 7 (2)               |
| Thrombosis                           | 6 (2)               |
| Renal failure                        | 2 (1)               |
| GVHD after transplantation           | 2 (1)               |
| Splenic rupture                      | 1 (1)               |
| Traffic accident                     | 1 (1)               |

Adapted from Takenaka et al., with permission from Springer Nature [28].

GVHD, graft-versus-host disease.

| Variable | IPSS [2] | DIPSS [29] | DIPSS plus [30] |
|----------|----------|------------|-----------------|
| Age > 65 years | ✓        | ✓          | ✓               |
| Constitutional symptoms^[a] | ✓        |            |                 |
| Hb < 10 g/dL | ✓        | ✓          | ✓               |
| WBC > 25,000/μL | ✓        | ✓          | ✓               |
| Peripheral blood blasts ≥ 1% | ✓        | ✓          | ✓               |
| Platelets < 10 × 10^3/μL | ✓        |            |                 |
| Red cell transfusion need^[b] | ✓        |            |                 |
| Unfavorable karyotype^[c] | ✓        |            |                 |

Point per variable 1 point each 1 point each but Hb = 2 1 point each
of DIPSS-plus, as these were the most significant predictors of shorter survival in our study [28]. Therefore, DIPSS-plus appears to be the optimal model for predicting survival in Japanese patients with PMF, although IPSS and DIPSS could also distinguish patients at higher versus lower risk of mortality. Moreover, the DIPSS-plus model could successfully identify patients with a poor prognosis at any time during the clinical course. This makes it useful for decision-making, for example when considering alloSCT. However, it had only modest accuracy for prognostication, suggesting that covariates included in DIPSS-plus alone are not sufficient, and that information regarding gene mutation status (e.g., CALR and ASXL1) should be included. We did not have data on mutations other than JAK2V617F in the registry. It is essential to collect precise genetic information to improve prognostication for patients with PMF throughout the disease course and subsequently inform the best treatment options.

**TREATMENT OF PMF**

**Management of PMF**

As noted above, the prognosis for patients with PMF is dismal compared with other MPNs, and alloSCT is the only current potentially curative treatment. However, as the median age at onset of PMF is 66 years, very few patients are eligible for alloSCT. For those who are not, therapy is primarily symptomatic, such as treating anemia or splenomegaly [3,4,11,26]. Treatment should be determined on a case-by-case risk assessment and evaluation of QOL using the DIPSS-plus risk classification and MF-SAF (Fig. 3). In terms of DIPSS-plus risk, a “watch-and-wait” policy is advisable for asymptomatic patients in the low and intermediate-1 risk groups because long-term survival can be expected with supportive therapy alone. For patients with symptomatic anemia, splenomegaly, or moderate constitutional symptoms, specific treatment is aimed at each particular symptom. PMF-associated anemia is usually treated with androgens, prednisolone, and danazol, which achieves improvement in up to 15% to 25% of patients [46,47]. Hydroxyurea is the first-line therapy for symptomatic splenomegaly, yielding an overall response rate of up to 40% and lasting an average of 1 year [47]. Recently, JAK2 inhibitors have been used for splenomegaly instead of hydroxyurea (see below). Splenectomy is an alternative choice for symptomatic splenomegaly, but it has a perioperative mortality rate of up to 5% to 10% and a morbidity rate of 25% [48,49]. Splenic irradiation can be used for patients who are not candidates for a JAK2 inhibitor or splenectomy, but the response is transient and there is a risk of severe cytopenia [50]. For patients in the intermediate-2 and high-risk groups, alloSCT should be considered if an appropriate donor exists. For those not eligible for alloSCT, a JAK2 inhibitor or participation in a clinical trial can be considered.

**JAK2 inhibitors**

The JAK2 inhibitor ruxolitinib has been available in the United States for the treatment of intermediate-2 or high-risk PMF since 2011. Two randomized phase III studies in the United States (Controlled Myelofibrosis Study with Oral JAK Inhibitor Treatment I [COMFORT-I] trial) and Europe (COMFORT-2 trial) were conducted for patients in the intermediate-2 or high-risk groups with PMF or post PV/ET myelofibrosis,
with trial results published in 2012 [33,51]. In the COMFORT-1 trial, 309 patients were assigned to either ruxolitinib or placebo. The COMFORT-2 trial included 219 patients randomized to receive either ruxolitinib or the best available therapy. Ruxolitinib administration improved splenomegaly and general symptoms such as fever, malaise, weight loss, and decreased activity. In 3 years of follow-up reports, the reduction in spleen volume and improvement in QOL were maintained, and improved survival was also observed [27,52]. These benefits were achieved regardless of the JAK2 mutation status, because the JAK2 inhibitor effectively suppresses proinflammatory cytokines, which is a major factor involved in the progression of PMF. Recently, the results of a pooled analysis of OS in COMFORT-1 and -2 were reported after crossover from the control groups to the ruxolitinib group in both studies; most patients in the control groups crossed over to ruxolitinib [53]. The OS at 144 weeks was 78% in the ruxolitinib group, 61% in the intention-to-treat control group, and 31% in the control group corrected for crossover, confirming the survival advantage conferred by ruxolitinib compared with placebo or other treatment. Spleen size at baseline was a risk factor for poor OS, and reduction in spleen size with ruxolitinib correlated with longer survival. The major adverse events were anemia and thrombocytopenia. Most of the grade 3 and 4 cytopenias appeared within 6 months (especially the first 2 to 3 months) of the start of treatment. Sudden interruption or withdrawal of JAK2 inhibitor provoked acute symptoms due to cytokine release syndrome; therefore, the dose of ruxolitinib should be tapered over up to 7 to 10 days to avoid this withdrawal syndrome. Since ruxolitinib inhibits T-cell function, attention should also be paid to the possible development of opportunistic infections, including tuberculosis, reactivation of hepatitis B, herpes zoster, or urinary tract infection [54]. Even if a patient is eligible for alloSCT, pre-transplant therapy with JAK2 inhibitor may be beneficial as splenomegaly has a negative impact on engraftment and transplant outcomes. A poor performance status due to constitutional symptoms increases non-relapse mortality after transplant. Therefore, pre-transplant ruxolitinib may improve performance status, reduce splenomegaly, and decrease the grade of subsequent acute graft-versus-host disease, resulting in faster hematopoietic recovery, improved graft function,

Figure 2. Survival curves of 780 patients with primary myelofibrosis in a Japanese registry, stratified by prognostic scoring system risk groups at diagnosis. (A) International Prognostic Scoring System (IPSS), (B) Dynamic IPSS (DIPSS), (C) Dynamic IPSS plus additional prognostic factors (DIPSS plus). Adapted from Takenaka et al., with permission from Springer Nature [28].
and improved survival. However, the use of ruxolitinib before alloSCT also has potentially detrimental effects, including withdrawal syndrome when the drug is discontinued, increased risk of infections, delayed hematopoietic recovery, and potential tumor lysis syndrome. As clinical experience is limited at present, use of a JAK2 inhibitor before alloSCT should be limited to patients enrolled in clinical trials.

**AlloSCT**

A retrospective analysis using the Center for International Bone Marrow Transplant Research database assessed transplant outcomes for 289 patients (median age, 47 years) with PMF [55]. The 5-year disease-free survival and 100-day treatment-related mortality rates were 33% and 18% for human leukocyte antigen (HLA) identical sibling donor transplants, 27% and 35% for unrelated donor transplants, and 22% and 19% for transplants from alternative related donors, respectively. Splenectomy was performed in 65 patients prior to transplantation but was not associated with better survival. Further research indicated that alloSCT can engraft despite severe bone marrow fibrosis, with an incidence of graft failure of < 10%; long-term survival is achieved in up to 30% to 50% of patients [56]. It was also reported that bone marrow fibrosis disappeared in more than half of patients with sustained engraftment. However, the incidence of transplant-related mortality may be as high as 30% to 50%. Risk factors for transplant-related mortality include huge splenomegaly, the need for frequent red blood cell transfusions, transplantation from HLA-mismatched donors, low performance status, and a high comorbidity index [57].

Recently, reduced-intensity conditioning regimens have shown promise, typically for the elderly and those with comorbidities. While such regimens are feasible, they do not appear to improve outcome [58-63]. No randomized controlled trials have compared myeloablative and reduced-intensity conditioning for alloSCT. Therefore, the optimal intensity of the conditioning regimen still needs to be defined. An alloSCT from a haploidentical related donor is also a consideration when HLA-identical donors are not available, but the results of haploidentical donor transplantation should be evaluated in clinical trials. A consensus report from an international working group suggested that patients aged < 70 years with intermediate-2 or high-risk PMF are candidates for alloSCT. In addition, they recommend that alloSCT be considered even for patients with intermediate-1 risk disease who are aged < 65 years, if they have transfusion-dependent anemia, > 2% peripheral blood blasts, or adverse cytogenetics [57]. There is
still much interest in the use of JAK2 inhibitors before alloSCT, and several clinical trials are ongoing to evaluate the efficacy and safety of JAK2 inhibitors for this purpose [41,64].

**Treatment in the Japanese registry cohort**
Red cell transfusions, androgens, hydroxyurea, and ruxolitinib were administered to 192 (25%), 72 (9%), 83 (11%), and 47 patients (6%), respectively [28]. Eleven patients (1%) underwent splenectomy and 21 (3%) had splenic radiation. An additional 43 patients (6%) received alloSCT at a median of 343 days (range, 23 to 4,066) after diagnosis and a median age of 52 years (range, 24 to 66). The stem cell sources for a first alloSCT included unrelated donor bone marrow (n = 21), related donor peripheral blood stem cells (n = 14), cord blood (n = 5), and related donor bone marrow (n = 2). Six patients received a second alloSCT for disease relapse, and one patient received a third alloSCT. The median follow-up after the first alloSCT was 36 months (range, 3 to 130), and the 3-year OS was 84% (95% CI, 68% to 93%). The estimated median survival after the first alloSCT was 134 months (range, 7 to not reached), and the median survival after diagnosis among all patients who underwent alloSCT was 207 months (range, 105 to not reached). The latter value is significantly longer than that calculated for patients who did not undergo alloSCT (median, 45 months; range, 38 to 49; p < 0.001). Although ours was not a controlled trial and the patients undergoing alloSCT were younger than those who did not, the registry data suggest that alloSCT could prolong OS and thereby affect the natural course of PMF. In contrast, no improvement in OS was observed in the 47 patients treated with a JAK2 inhibitor in our series, although this was likely due to the small number of patients and the short observation period from the introduction of the agent.

**CONCLUSIONS**
Based on recent advances in our understanding of the molecular basis of PMF and the revised diagnostic criteria for MPN, screening for the JAK2, CALR, and MPL driver mutations and assessing bone marrow morphology have become essential for the diagnosis of PMF. Several prognostic scoring systems have been developed for PMF based on clinical characteristics and blood counts. While these are the standard methods currently used to estimate prognosis, it appears that evaluating other molecular risk predictors may improve the assessment of individual patients independent of the conventional prognostic scoring system. Choice of treatment should be based on risk assessment and the evaluation of the QOL in each case, generally using the DIPSS-plus risk classification and MF-SAF. The JAK2 inhibitor ruxolitinib can reduce spleen volume, and improve QOL and survival. However, alloSCT is currently the only potentially curative treatment for PMF, although it continues to have high transplant-related mortality. Ongoing trials of JAK2 inhibitors before alloSCT may help clarify the value of this approach as we continue to search for the best options for the treatment of PMF.

**Conflict of interest**
No potential conflict of interest relevant to this article was reported.

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**REFERENCES**
1. Vardiman JW, Thiele J, Arber DA, et al. The 2008 revision of the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia: rationale and important changes. Blood 2009;114:2937-951.
2. Cervantes F, Dupriez B, Pereira A, et al. New prognostic scoring system for primary myelofibrosis based on a study of the International Working Group for Myelofibrosis Research and Treatment. Blood 2009;113:2895-2901.
3. Vannucchi AM, Barbui T, Cervantes F, et al. Philadelphia chromosome-negative chronic myeloproliferative neoplasms: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2015;26 Suppl 5:v85-v99.
4. Tefferi A. Myeloproliferative neoplasms: a decade of
discoveries and treatment advances. Am J Hematol 2016;91:50-58.
5. Cervantes F, Passamonti F, Barosi G. Life expectancy and prognostic factors in the classic BCR/ABL-negative myeloproliferative disorders. Leukemia 2006;22:905-914.
6. Tefferi A, Guglielmelli P, Larson DR, et al. Long-term survival and blast transformation in molecularly annotated essential thrombocythemia, polycythemia vera, and myelofibrosis. Blood 2014;124:2507-2513.
7. Dupriez B, Morel P, Demory JL, et al. Prognostic factors in agnogenic myeloid metaplasia: a report on 195 cases with a new scoring system. Blood 1996;88:1013-1018.
8. Cervantes F, Pereira A, Esteve J, et al. Identification of ‘short-lived’ and ‘long-lived’ patients at presentation of idiopathic myelofibrosis. Br J Haematol 1999;107:635-640.
9. Cervantes F, Dupriez B, Passamonti F, et al. Improving survival trends in primary myelofibrosis: an international study. J Clin Oncol 2012;30:2981-2987.
10. Hultcrantz M, Kristinsson SY, Andersson TM, et al. Patterns of survival among patients with myeloproliferative neoplasms diagnosed in Sweden from 1973 to 2008: a population-based study. J Clin Oncol 2012;30:3995-3001.
11. Tefferi A. Primary myelofibrosis: 2014 update on diagnosis, risk-stratification, and management. Am J Hematol 2014;89:915-925.
12. Shammo JM, Stein BL. Mutations in MPNs: prognostic implications, window to biology, and impact on treatment decisions. Hematology Am Soc Hematol Educ Program 2016;2016:5352-536.
13. Passamonti F, Maffioli M. Update from the latest WHO classification of MPNs: a user’s manual. Hematology Am Soc Hematol Educ Program 2016;2016:533-542.
14. Spivak JL. Myeloproliferative neoplasms. N Engl J Med 2017;376:2168-2181.
15. Baxter EJ, Scott LM, Campbell PJ, et al. Acquired mutation of the tyrosine kinase JAK2 in human myeloproliferative disorders. Lancet 2005;365:1055-1061.
16. James C, Ugo V, Le Couedic JP, et al. A unique clonal JAK2 mutation leading to constitutive signalling causes polycythaemia vera. Nature 2005;434:1144-1148.
17. Kralovics R, Passamonti F, Buser AS, et al. A gain-of-function mutation of JAK2 in myeloproliferative disorders. N Engl J Med 2005;352:1779-1790.
18. Levine RL, Wadleigh M, Cools J, et al. Activating mutation in the tyrosine kinase JAK2 in polycythemia vera, essential thrombocythemia, and myeloid metaplasia with myelofibrosis. Cancer Cell 2005;7:387-397.
19. Pardanani AD, Levine RL, Lasho T, et al. MPL515 mutations in myeloproliferative and other myeloid disorders: a study of 1182 patients. Blood 2006;108:3472-3476.
20. Pikman Y, Lee BH, Mercher T, et al. MPLW515L is a novel somatic activating mutation in myelofibrosis with myeloid metaplasia. PLoS Med 2006;3:270.
21. Klampfl T, Gisslinger H, Harutyunyan AS, et al. Somatic mutations of calreticulin in myeloproliferative neoplasms. N Engl J Med 2013;369:2379-2390.
22. Nangalia J, Massie CE, Baxter EJ, et al. Somatic CALR mutations in myeloproliferative neoplasms with nonmutated JAK2. N Engl J Med 2013;369:2391-2405.
23. Arber DA, Orazi A, Hasserjian R, et al. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. Blood 2016;127:2391-2405.
24. Cervantes F. How I treat myelofibrosis. Blood 2014;124:2635-2642.
25. Geyer HL, Mesa RA. Therapy for myeloproliferative neoplasms: when, which agent, and how? Blood 2014;124:3529-3537.
26. Harrison CN, McLornan DP. Current treatment algorithm for the management of patients with myelofibrosis, JAK inhibitors, and beyond. Hematology Am Soc Hematol Educ Program 2017;2017:489-497.
27. Verstovsek S, Mesa RA, Gotlib J, et al. Efficacy, safety, and survival with ruxolitinib in patients with myelofibrosis: results of a median 3-year follow-up of COMFORT-I. Haematologica 2015;100:479-488.
28. Takenaka K, Shimoda K, Uchida N, et al. Clinical features and outcomes of patients with primary myelofibrosis in Japan: report of a 17-year nationwide survey by the Idiopathic Disorders of Hematopoietic Organs Research Committee of Japan. Int J Hematol 2017;105:59-69.
29. Moulard O, Mehta J, Fryzek J, Olives R, Iqbal U, Mesa RA. Epidemiology of myelofibrosis, essential thrombocythemia, and polycythemia vera in the European Union. Eur J Haematol 2014;92:289-297.
30. Deadmond MA, Smith-Gagen JA. Changing incidence of myeloproliferative neoplasms: trends and subgroup risk profiles in the USA, 1973-2011. J Cancer Res Clin Oncol 2015;141:2131-2138.
31. Geyer HL, Scherber RM, Dueck AC, et al. Distinct clustering of symptomatic burden among myeloproliferative neoplasm patients: retrospective assessment in 1470 pa-
32. Emanuel RM, Dueck AC, Geyer HL, et al. Myeloproliferative neoplasm (MPN) symptom assessment form total symptom score: prospective international assessment of an abbreviated symptom burden scoring system among patients with MPNs. J Clin Oncol 2013;31:1285-1292.

33. Verstovsek S, Mesa RA, Gotlib J, et al. Effect of ruxolitinib therapy on myelofibrosis-related symptoms and other patient-reported outcomes in COMFORT-I: a randomized, double-blind, placebo-controlled trial. J Clin Oncol 2013;31:1285-1292.

34. Harrison CN, Mesa RA, Kiladjian JJ, et al. Health-related quality of life and symptoms in patients with myelofibrosis treated with ruxolitinib versus best available therapy. Br J Haematol 2013;162:229-239.

35. Gangat N, Caramazza D, Vaidya R, et al. DIPSS plus: a refined dynamic International Prognostic Scoring System for primary myelofibrosis that incorporates prognostic information from karyotype, platelet count, and transfusion status. J Clin Oncol 2011;29:392-397.

36. Tefferi A, Mesa RA, Nagorney DM, Schroeder G, Silverstein MN. Splenectomy in myelofibrosis with myeloid metaplasia: a single-institution experience with 223 patients. Blood 2000;95:2226-2233.

37. Barbui T, Thiele J, Passamonti F, et al. Survival and disease progression in essential thrombocythemia are significantly influenced by accurate morphologic diagnosis: an international study. J Clin Oncol 2011;29:3179-3184.

38. Elliott MA, Chen MG, Silverstein MN, Tefferi A. Splenic irradiation for symptomatic splenomegaly associated with myelofibrosis and myeloid metaplasia. Br J Haematol 1998;103:505-511.

39. Harrison C, Kiladjian JJ, Al-Ali HK, et al. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis: long-term findings from COMFORT-II, a phase 3 study of ruxolitinib versus best available therapy. Leukemia 2016;30:1701-1707.

40. Vannucchi AM, Kantarjian HM, Kiladjian JJ, et al. A pooled analysis of overall survival in COMFORT-I and COMFORT-II, 2 randomized phase III trials of ruxolitinib for the treatment of myelofibrosis. Haematologica 2015;100:1139-1145.
55. Ballen KK, Shrestha S, Sobocinski KA, et al. Outcome of transplantation for myelofibrosis. Biol Blood Marrow Transplant 2010;16:358-367.
56. Guardiola P, Anderson JE, Bandini G, et al. Allogeneic stem cell transplantation for agnogenic myeloid metaplasia: a European Group for Blood and Marrow Transplantation, Societe Francaise de Greffe de Moelle, Gruppo Italiano per il Trapianto del Midollo Osseo, and Fred Hutchinson Cancer Research Center Collaborative Study. Blood 1999;93:2831-2838.
57. Kroger NM, Deeg JH, Olavarria E, et al. Indication and management of allogeneic stem cell transplantation in primary myelofibrosis: a consensus process by an EBMT/ELN international working group. Leukemia 2015;29:2126-2133.
58. Merup M, Lazarevic V, Nahi H, et al. Different outcome of allogeneic transplantation in myelofibrosis using conventional or reduced-intensity conditioning regimens. Br J Haematol 2006;135:67-373.
59. Patriarca F, Bacigalupo A, Sperotto A, et al. Allogeneic hematopoietic stem cell transplantation in myelofibrosis: the 20-year experience of the Gruppo Italiano Trapianto di Midollo Osseo (GITMO). Haematologica 2008;93:1514-1522.
60. Gupta V, Kroger N, Aschan J, et al. A retrospective comparison of conventional intensity conditioning and reduced-intensity conditioning for allogeneic hematopoietic cell transplantation in myelofibrosis. Bone Marrow Transplant 2009;44:317-320.
61. Kroger N, Holler E, Kobbe G, et al. Allogeneic stem cell transplantation after reduced-intensity conditioning in patients with myelofibrosis: a prospective, multicenter study of the Chronic Leukemia Working Party of the European Group for Blood and Marrow Transplantation. Blood 2009;114:5264-5270.
62. Abelsson J, Merup M, Birgegard G, et al. The outcome of allo-HSCT for 92 patients with myelofibrosis in the Nordic countries. Bone Marrow Transplant 2012;47:380-386.
63. McLornan DP, Mead AJ, Jackson G, Harrison CN. Allogeneic stem cell transplantation for myelofibrosis in 2012. Br J Haematol 2012;157:413-425.
64. Ballinger TJ, Savani BN, Gupta V, Kroger N, Mohty M. How we manage JAK inhibition in allogeneic transplantation for myelofibrosis. Eur J Haematol 2015;94:115-119.