Plasma miR-136 can be used to screen patients with knee osteoarthritis from healthy controls by targeting IL-17

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Received February 5, 2018; Accepted August 7, 2018

DOI: 10.3892/etm.2018.6625

Abstract. The current study mainly evaluated the plasma level of microRNA (miR)-136 in knee osteoarthritis (KOA) patients and determined if miR-136 could be used as a potential biomarker to screen KOA patients from healthy controls. It was demonstrated that plasma miR-136 was significantly decreased in the plasma of KOA patients. Moreover, the reduction in plasma miR-136 negatively correlated with the severity of KOA. Additionally, the increase in the serum interleukin (IL)-17 level positively correlated with the severity of KOA. More importantly, dual luciferase assays and western blot assays indicated that IL-17 was a target gene of miR-136. Further analysis showed that plasma miR-136 could be used as a biomarker to screen KOA patients from healthy controls. In summary, for the first time, the present study revealed that plasma miR-136 could be used as a potential biomarker to screen KOA patients from healthy controls. Moreover, postoperative rehabilitation treatment requires a long time to ensure the recovery of knee joint function in patients. Hence, early detection and prevention of KOA are of great importance for high-risk patients.

Interleukin (IL-17) is a novel cytokine family secreted mainly by CD4+ T helper 17 cells (Th17), including IL-17A, IL-17B, IL-17C, IL-17D, IL-17E and IL-17F (7,8). By binding the corresponding receptors on the cell surface, IL-17 is widely involved in bone metabolism (9,10). Multiple studies have confirmed that IL-17 is the main factor leading to osteoarthritis bone injury (9-11). Clinical data also indicate that the levels of IL-17 and IL-17R are higher in the synovial fluid of patients with arthritis (11). Furthermore, high levels of IL-17 and TNFα are correlated with poor prognosis for osteoarthritis patients (12,13). However, the underlying mechanism by which IL-17 is highly upregulated in osteoarthritis has been poorly understood.

MicroRNAs (miRNAs/miRs) are small noncoding RNAs with approximately 23 nucleotides that extensively participate in posttranscriptional regulation by binding the 3' untranslated region (3'UTR) of mRNA (14,15). Due to the ability to be resistant to RNase, increasing evidence has suggested the use of circulating miRNAs as potential biomarkers for various diseases (16). For instance, increased levels of miR-29c, miR-93, and miR-126 are shown to be related to the progression of osteoarthritis (16).

Abnormal expression of miR-136 has been widely reported in the development of tumors and metabolic diseases (17,18). However, we still lack knowledge regarding whether miR-136 is involved in the progression of KOA. The current study explored the plasma level of miR-136 in KOA patients and attempted to determine if it could be used as a potential biomarker to screen KOA patients from healthy controls.

Materials and methods

Patient samples. This study was approved by the Ethics Committee of Rizhao People's Hospital (Rizhao, China) and performed in accordance with the Declaration of Helsinki. All participants gave their informed consent. In total, 50 patients with knee OA were recruited from February 2016 to December 2016 at the Department of Joint Surgery, Rizhao People's Hospital, according to the 1986 classification of osteoarthritis of the knee in the diagnostic criteria.
of the American Rheumatism Association (19). Diagnosis of osteoarthritis was performed according to criteria of the American College of Rheumatology (19). Exclusion criteria were: i) Symptoms suggestive of any other chronic inflammatory disease; ii) diabetes; iii) history of corticosteroid treatment; iv) any other form of arthritis; v) cancer; and vi) family history of osteoarthritis. X-ray examinations were carried out to divide patients into early, middle and late KOA stages. All participants were examined with bilateral knee plain radiographs (bilateral anteroposterior and lateral) using an SD 3000 Synchro Stand. Two independent radiologists evaluated radiographic changes related to OA using the K/L grading system (20). KOA was defined as a K/L grade ≥2 in both knees. In addition, health volunteers who were 35 years of age and gender matched at the Department of Physical Examination were enrolled in the study as health controls (HCs). The details of KOA patients and healthy controls are listed in Table I.

Sample acquisition and RNA extraction. A 5 ml aliquot of blood was collected from all participants and directly into sodium citrate tubes. Total RNA was isolated with RNAsol LS (Vigorous, Beijing, China) according to the manufacturer's instructions for isolating small RNAs. The quality, quantity and integrity of RNA were monitored using a NanoDrop spectrophotometer (ND-1000; NanoDrop Technologies; Thermo Fisher Scientific, Inc., Wilmington, DE, USA).

qPCR validation. RNA was reverse transcribed into cDNA using a PrimeScript OneStep RT-qPCR kit (C28025-032; Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA). The detailed RT-qPCR procedure is described as follows: 95°C for 10 min; followed by 50 cycles of 95°C for 10 sec, 55°C for 10 sec, 72°C for 5 sec, 94°C for 1 sec, 59°C for 15 sec, and 95°C for 1 sec; and then cooled to 40°C. The relative expression levels were calculated with the 2^{ΔΔCt} method (21), and the experiments were repeated in triplicate. The specific primers used in the current study are listed as follows: miR-136-RT: GTCGTATATCCAGTGAGGTTCC GAGGTATTCGCAGCTAGCAACACAC; U6-RT: GTCGTATATCCAGTGAGGTTCC GAGGTATTCGCAGCTAGCAACACAC; U6-F: GCGCTGGAGTGT GACAATGGTG; U6-R: GCAGCTGGTGAACGGGTTC; Universal reverse primer: GTGCAGGTTCCAGGT.

Enzyme-linked immunosorbent assay (ELISA). The serum samples were used to quantify the level of IL-17 using an enzyme-linked immunosorbent assay according to the manufacturer's instructions (R&D Systems, Inc., Minneapolis, MN, USA). Samples were read at a 450 nm wavelength using a microplate reader (Model 3550; Thermo Fisher Scientific, Inc.).

Cell culture. 293 cells were seeded at a density of 1.5×10^4 cells/cm² and cultured in Dulbecco's modified Eagle's medium (HyClone; GE Healthcare Life Sciences, Logan, UT, USA) containing 10% heat-inactivated fetal calf serum (Gibco; Thermo Fisher Scientific, Inc.), streptomycin (100 mg/ml, Gibco; Thermo Fisher Scientific, Inc.), and penicillin (100 U/ml; Gibco; Thermo Fisher Scientific, Inc.) in an incubator at 37°C with 5% CO₂.

Table I. Comparison of patients baseline data between groups.

| Characteristic       | KOAs (n=74) | HCs (n=79) | P-value |
|----------------------|-------------|------------|---------|
| Age (mean ±SE, years) | 65.8±7.3    | 66.2±6.9   | 0.451   |
| Sex (female/male, n)  | 40/34       | 42/37      | 0.298   |
| BMI (mean ±SE, kg/m²) | 26.7±1.5    | 26.8±1.2   | 0.353   |
| K/L grade (2/3/4)     | 22/29/23    |            |         |
| BMI, body mass index  |             |            |         |

miRNA target prediction and dual-luciferase reporter assay. miRNA targets were predicted using TargetScan (https://www.targetscan.org). The 3'UTR of IL-17, which contains the predicted target site for miR-136, was cloned into the pmirGLO luciferase reporter vector (Promega Corporation, Madison, WI, USA), which was cleaved at SacI and XhoI sites. The details of the PCR procedures are described as follows: A hot start step at 95°C for 10 min, followed by 40 cycles of 95°C for 15 sec, 55°C for 45 sec and 72°C for 30 sec.

Prior to conducting the dual reporter assay, 5×10^4 293 cells/well were seeded in 24-well plates with 500 µl DMEM and cultured for 18 h. The cells were transfected with the modified firefly luciferase reporter vector (500 ng/µl) and mixed with VigoFect transfection reagent (Vigorous, Beijing, China) according to the manufacturer's protocol. After continuous exposure to miR-136/pmirGLO-IL-17-3'UTR or NC/pmirGLO blank vector for 48 h, the firefly and Renilla luciferase activities were measured with the Dual-Luciferase® Reporter Assay system (Promega Corporation) according to the manufacturer's protocol. Firefly luciferase activity was normalized to Renilla luciferase activity.

Cell transfection. miR-136 mimic, inhibitor and negative control (NC) were purchased from GenePharma Co., Ltd. (Shanghai, China). In brief, 6×10^4 cells were equally seeded in 6-well plates with 2 ml DMEM containing serum and antibiotics. At the same time, miR-136 mimic, inhibitor or NC was mixed with HiPerFect transfection reagent (Qiagen, Inc., Valencia, CA, USA) and incubated at room temperature for 10 min. The complex was then transfected into the cells for 48 h.

Western blotting. Cell protein was extracted using radioimmunoprecipitation lysis buffer (Beijing Solarbio Science & Technology Co., Ltd., Beijing, China) and was collected following centrifugation at 12,000 × g for 30 min at 4°C. A bicinchoninic protein assay kit (Pierce; Thermo Fisher Scientific, Inc.) was used to determine the protein concentration. A total of 15 µg protein was loaded per lane, separated by 10% SDS-PAGE and transferred to polyvinylidene difluoride membranes. The membranes were blocked with 8% non-fat dry milk at 4°C overnight. Following three washes with PBS with Tween 20 (5 min/wash), the membranes were incubated with the following primary antibodies at 4°C overnight: IL-17 (cat no. 13838; 1:1,000; Cell Signaling Technology, Inc., Danvers, MA, USA) and GAPDH (cat. no. 5174; 1:1,000;
Following several washes with TBST, the membranes were incubated with horseradish peroxidase (HRP)-conjugated goat anti-rabbit and anti-mouse Immunoglobulin G (IgG) or HRP-conjugated mouse anti-goat IgG (all 1:5,000; Zhongshan Gold Bridge Biological Technology Co., Beijing, China) for 2 h at room temperature and then washed. The blots were then incubated with horseradish peroxidase (HRP)-conjugated anti-IgG secondary antibody (1:5,000; OriGene Technologies, Inc., Beijing, China) for 2 h at room temperature and then washed, followed by detection with enhanced chemiluminescent substrate (EMD Millipore, Billerica, MA, USA). GAPDH was used as an internal control. ImageJ software (National Institutes of Health, Bethesda, MD, USA) was used for density analysis.

**Receiver operating characteristic (ROC) curves.** An ROC curve was established to interpret the ability of miR-136 in discriminating KOA patients from healthy controls. In brief, the fluorescence signals of each reaction well were collected quantitatively, and the Cq values were recorded. The expression of miR-136 in KOA patients relative to healthy control was expressed as $2^{-\Delta\Delta Cq}$. $\Delta Cq$ is the difference of Cq values of the target gene to the internal reference U6 in each sample, namely $\Delta Cq=(Cq$ miR-136$)-Cq$ U6. To perform ROC analysis, $2^{-\Delta\Delta Cq}$ was recorded parallel to the diagnostic results of gold standard, that is healthy control=0, KOA patients=1. Then, the area under the curve (AUC), sensitivity and specificity were computed in order to validate the diagnostic application of
miR‑136 as KOA biomarkers in contrast to the diagnostic results of gold standard (version 20.0, SPSS, Inc., Chicago, Illinois).

Statistics. The data are represented as the mean ± standard error. The two-tailed unpaired Student’s t-tests were used for comparisons of the two groups. For multiple groups comparisons, one-way analysis of variance followed by Tukey’s post hoc test was used. Statistical tests were performed using SPSS software (version 13.0; SPSS, Inc., Chicago, IL, USA). P<0.05 was considered to indicate a statistically significant difference.

Results

Reduced plasma miR-136 and increased serum IL-17 levels in KOA patients. First, we examined the plasma levels of miR-136 in KOA patients and healthy controls. Compared with healthy controls (1±1.07), the plasma levels of miR‑136 were significantly increased to 0.55±0.37, 0.26±0.27, and 0.08±0.17 for patients at the early, middle and late KOA stages, respectively (Fig. 1A). Next, serum IL-17 levels were determined using an ELISA kit. In contrast to healthy controls (3.23±2.86 ng/ml), the levels of IL-17 for patients at the early, middle and late KOA stages were 34.37±8.93, 69.65±15.41, and 96.42±21.45 ng/ml, respectively (Fig. 1B).

miR-136 was reduced along with increased K/L grades. Moreover, we examined miR-136 and IL-17 levels based on the K/L grade. Our data showed that the level of plasma miR-136 was 1±0.47 but was much lower in the K/L grade 2 (0.61±0.32), grade 3 (0.37±0.24), and grade 4 (0.11±0.13) groups (Fig. 2A). In addition, we determined the serum IL-17 levels
IL-17 is a target gene of miR-136. The above observations motivated us to explore the underlying mechanism by which miR-136 levels were negatively correlated with the serum levels of IL-17. TargetScan analysis showed a conserved binding site of miR-136 in the 3'UTR of IL-17 (Fig. 3A). Dual-luciferase reporter assay showed that miR-136 significantly suppressed the relative luciferase activity of pmirGLO-IL-17-3'UTR, compared with the blank pmirGLO plasmid (Fig. 3B). The western blot assay also indicated that overexpression of miR-136 significantly suppressed the protein level of IL-17 (Fig. 3C), while inhibition of miR-136 significantly increased the expression of IL-17 (Fig. 3D). These data validated that IL-17 was a target gene of miR-136.

miR-136 could be used to screen KOA patients from healthy controls. We then evaluated whether miR-136 could be used as a potential biomarker for patients with KOA. ROC analysis showed that the plasma miR-136 level could be used to screen KOA patients from healthy controls, with an ROC curve area of 0.935 (95% confidence interval: 0.817–1.000; P<0.0001) (Fig. 4).

Discussion

KOA, a chronic progressive disease, is a common disease in the elderly. With the advent of an aging society, the incidence of this disease is increasing (22,23). Undoubtedly, it is of great significance to improve the quality of life of patients with KOA (5).

Increasing evidence has associated key roles of miRs with the progression of KOA (24,25). For example, miR-9 is shown to modulate the development of KOA via the NF-kappaB1 pathway in chondrocytes (24); miR-29a is also reported to suppress synovitis in the pathogenesis of KOA by targeting VEGF (25). Abnormal expression of miR-136-5p after spinal cord injury has been identified. He et al reported that overexpression of miR-136-5p effectively enhanced inflammatory factors and chemokines via activating NF-κB/A20 signaling in the IL-17-mediated inflammatory response both in vitro and in vivo (26). In IL-17-stimulated astrocytes, miR-136-5p is also found to increase inflammatory responses via suppressing the expression of A20 (27). However, whether miR-136 is involved in IL-17 induced inflammatory responses in KOA patients has never been explored. In the current study, we mainly focused on miR-136, which has been extensively studied in various diseases. However, there have been limited studies on the role of miR-136 in the pathology of KOA. Here, we showed novel data that the plasma miR-136 level was significantly decreased in the plasma of KOA patients. Moreover, the reduction in plasma miR-136 levels was negatively correlated with the severity of KOA.

As a potent inflammatory factor, IL-17 plays an important role in the development of a variety of inflammatory diseases, and it is also an important regulator in the development of bone metabolism and bone diseases (28). It is suggested that IL-17 can regulate osteoblast differentiation and bone salt deposition. Meanwhile, through the stimulation of osteoblasts, bone cells, and stromal cells, IL-17 can enhance osteoclast activity and promote bone degradation (12,29). Targeting IL-17 or IL-17 receptors (IL-17R) is becoming a hot spot for the treatment of inflammatory diseases and inflammatory bone disease (29,30). Chen et al conducted a retrospective study and reported that Serum IL-17 concentrations were much higher in Han Chinese patients with primary KOA (n=98) than those in healthy control (n=50) (29). Deligne et al explored the expression of IL-17 in the inflamed and the non-inflamed area of each synovium sample (n=20) from human osteoarthritic knee tissues (30). They demonstrated the expression of IL-17 is much higher in the the inflamed than that of the non-inflamed area (30). Both of them pointed out that IL-17 may be effective in the early prevention and therapy for KOA patients (29,30).

In the current study, we also evaluated the level of IL-17 in the serum of KOA patients and healthy controls. Our data showed that the increase in serum IL-17 levels positively correlated with the severity of KOA. The negative correlation between IL-17 and miR-136 promoted us to explore its underlying mechanism. Interestingly, we found that IL-17 is a target gene of miR-136. Further analysis showed that plasma miR-136 levels could be used as a biomarker to screen KOA patients from healthy controls.

In summary, for the first time, we found that plasma miR-136 levels were significantly decreased in KOA patients. More importantly, by targeting IL-17, miR-136 could be used as a potential biomarker for KOA patients.

Acknowledgements

Not applicable.

Funding

The present study was supported by a grant from Scientific Research Starting Foundation of Rizhao People's Hospital (grant no. 20160823).

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

WL performed the experiments and analyzed the data. ZQ, NG, XT, DC performed part of the RT-qPCR experiments. WS designed the experiments, analyzed the data and gave final approval of the version to be published. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was approved by the Research Ethics Committee of Rizhao People's Hospital (Rizhao, China) and
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