Lake sedimentary biogenic silica from diatoms constitutes a significant global sink for aluminium

Dong Liu1,4, Peng Yuan1,4*, Qian Tian1,2, Hongchang Liu1, Liangliang Deng1, Yaran Song1,2, Junming Zhou1,2, Dusan Losic3, Jieyu Zhou1,2, Hongze Song1,2, Haozhe Guo1,2 & Wenxiao Fan1,2

Diatoms play an important role in marine biogeochemical cycle of aluminum (Al), as dissolved Al is taken up by diatoms to build their siliceous frustules and is involved in the sedimentation of diatomaceous biogenic silica (BSi). The Al incorporation in BSi facilitates decreasing the dissolution of marine BSi and thus substantially influences the biochemical processes driven by diatoms, such as CO2 sequestration. However, the role of lake BSi in the terrestrial biochemical Al cycle has not been explored, though lakes represent the second-largest sink for BSi. By identifying the previously unexplored high Al/Si atomic ratios (up to 0.052) in lake BSi, here we show lake BSi is a large terrestrial Al pool due to its high Al content, and lake sedimentary BSi constitutes a significant global sink for Al, which is on the same magnitude as the Al sink in global oceans.
Aluminium (Al) is the third most abundant element in the Earth’s crust. Its biogeochemical cycling on land, in water, and in the atmosphere is a central topic of discussion in many geochemical, environmental science, and biological studies due to the active role of Al in geochemical processes (e.g., weathering and reverse weathering) as well as the ecological and environmental impacts of Al (e.g., potential ecotoxicity to organisms)\textsuperscript{2,3}. The marine biogeochemical cycling of Al is strongly associated with the cycling of silicon (Si) through marine diatoms\textsuperscript{4–10}. Diatoms account for ~40% of marine primary production\textsuperscript{11,12}, and the gross production of biogenic silica in the ocean reaches ~2.40 × 10\textsuperscript{14} mol year\textsuperscript{−1} (refs. 13,14). Diatoms control the biogenic cycling of Si throughout the world’s oceans because they use Si to build siliceous frustules\textsuperscript{11,15}. The inorganic part of diatom frustum is diatomaceous biogenic silica (denoted BSi hereafter)\textsuperscript{16}. The coupled Al and Si biogeochemical cycle occurs primarily during BSi biosynthesis, in which Al is incorporated into the structure of BSi with Al/Si atomic ratios of up to ~0.008 (ref. 5). The marine BSi subsequently settles, and ~3% of the total marine BSi is eventually preserved in the seafloor (with a burial rate of ~6.30 × 10\textsuperscript{12} mol year\textsuperscript{−1})\textsuperscript{14}. This process exports organic carbon to the deep sea through the marine biological pump and is therefore important for CO\textsubscript{2} sequestration\textsuperscript{12}. Furthermore, the process also delivers a large amount of dissolved Al from marine waters to the seafloor\textsuperscript{17,18}. The magnitude of Al deposition associated with marine BSi is nearly equivalent to that of the riverine input of dissolved Al into the oceans\textsuperscript{17}, and constitutes an important part of the marine biogeochemical Al cycle\textsuperscript{18}.

While marine Al cycle in relation to marine BSi has been well studied\textsuperscript{19}, surprisingly, no reports have considered diatoms-driven Si and Al co-cycles in lakes, even though lakes represent an important part of the marine biogeochemical Al cycle that is currently not considered.

In this study, we show that lake BSi has high Al content in lake BSi, and lake’s sedimentary BSi constitutes a significant global sink for Al. This large Al retention due to lake BSi needs to be acknowledged in the modelling of the global biogeochemical Al cycle that is currently not considered.

**Results**

**Identification of Al distribution and coordination in BSi.** Two model species of lake diatoms, *Cyclotella meneghiniana* (Bacillariophyceae: *Centraecae*) (denoted BSi-C) and *Nitzschia palea* (*Pennonatae*) (BSi-N), which are among the most widely distributed freshwater diatoms in the world\textsuperscript{25}, were used for studying the uptake of Al by lake BSi. The Si and Al distributions in the BSi structures were examined using FIB milling (see Supplementary Fig. 1 for the details of the FIB pretreatment of BSi-C as an example) combined with EDS mapping analysis (Fig. 1). This FIB milling process allows detection of the inner structure of the BSi, thus avoiding possible disturbances from non-structural Al on the external surface of the BSi. The FIB–EDS mapping images of the BSi-C (Fig. 1c) and BSi-N (Fig. 1f) show that Al is homogeneously distributed in the siliceous structure of lake BSi.

To evaluate the 3D structural Al distributions in BSi, TOF-SIMS analysis producing 3D reconstructions of architectural elements was used\textsuperscript{28}. The TOF-SIMS analysis result of a frustule globally is ~1.72 × 10\textsuperscript{9} kg year\textsuperscript{−1}, which is on the same order of magnitude as that of the marine Al burial rate (~1.36 × 10\textsuperscript{9} kg year\textsuperscript{−1}). Therefore, lake BSi is a large terrestrial Al pool due to the high Al content in lake BSi, and lake’s sedimentary BSi constitutes a significant global sink for Al. This large Al retention due to lake BSi needs to be acknowledged in the modelling of the global biogeochemical Al cycle that is currently not considered.
of BSi-C exemplifies the 3D elemental distribution. A typical disk-like C. meneghiniana frustule with a diameter of ~4.0–5.0 μm (determined by field-emission scanning electron microscope (FESEM) imaging: Fig. 2a) and a thickness of ~2.5 μm (Fig. 2b) was subjected to the 3D elemental characterization by TOF-SIMS. Figures 2c–f show the Si and Al distributions in the top view and side view of the frustule of BSi-C. Al is clearly homogeneously distributed throughout the structure of the BSi, which is further indicated by the visible 3D Si and Al distributions from all angles (see Supplementary Movie for a short video of the reconstructed elemental distributions). This result demonstrates the incorporation of Al into the structure of the BSi of lake diatoms.

Al K-edge X-ray absorption near-edge structure (XANES) spectroscopic analysis was used to reveal the coordination states of the structural Al in the lake BSi. The XANES spectra (Fig. 3) indicate that the Al in the lake BSi exists mainly in tetrahedral coordination, regardless of the diatom species, as evidenced by the 1566-eV signal of tetrahedral Al²⁹,³⁰. Isomorphous Si substitution by Al therefore occurs in natural lake BSi because the Si in BSi is tetrahedrally coordinated, verifying the incorporation of Al into lake BSi through Al–Si substitution. This incorporation mechanism of Al into BSi implies that the state of Al in BSi is stable because isomorphous substitution is significantly stronger than non-structural incorporation (such as by adsorption), as is well known.

Quantification of structural Al in BSi and in diatoms. FESEM with EDS spot analysis was used to determine the atomic percentages of Al and Si in lake BSi. Over 100 spots from different frustules of lake BSi for each diatom species were collected for the spot analysis. The results reveal high Al incorporation capacities for all of the lake BSi samples. Based on the atomic percentages of Al and Si in the BSi (Fig. 4a, b), the average lake Al/Si atomic ratios (Al/SiBSi) are 0.049 ± 0.003 and 0.052 ± 0.003 for BSi-C and BSi-N, respectively (Supplementary Fig. 2)—at least six times greater than the maximum Al/SiBSi ratio (~0.008)³ of natural marine BSi. The Al/Si ratios of the entire diatoms (i.e., the intact diatoms, in which the organics were not removed) are 0.105 ± 0.008 (C. meneghiniana) and 0.115 ± 0.012 (N. palea) (Supplementary Fig. 2). Positive correlations (R² ≥ 0.75) are observed between the Si and Al contents of BSi for the lake BSi samples (Fig. 4a, b), indicating the incorporation of Al into the BSi of natural lake diatoms. A similar positive correlation as an indicator of Fe incorporation into marine BSi has been clarified previously³¹.

We investigated the influence of the Al concentration in the culture medium on the Al/SiBSi values of cultured diatoms by EDS spot analysis. The cultured diatom species are the same as the abovementioned natural lake diatoms. Six Al concentrations (1.0, 2.0, 5.0, 10.0, 50.0, and 100.0 μM) were applied, with Si concentrations remaining constant. The obtained Al/SiBSi values range between 0.045 and 0.055 for cultured C. meneghiniana and between 0.049 and 0.070 for cultured N. palea (grey columns in Fig. 5a, b). In line with the observations for natural lake BSi, the Al/SiBSi values of cultured lake BSi are also significantly higher than those of the BSi of cultured marine diatoms³⁴, which showed a maximum Al/SiBSi value of only ~0.007 (for the marine Lauerdiella annulata diatoms with an Al concentration of 400 nM in the culture medium)³⁵. In addition, similar to those of natural lake diatoms (Supplementary Fig. 2), the Al/SiBSi values of cultured diatoms are lower than the Al/Si values of the entire diatoms (Fig. 5). This pattern occurs because Al exists not only in BSi but also in the organic components of diatoms³⁴. This point is supported by the mass spectrometry (MS) data from TOF-SIMS (see Supplementary Fig. 3) analysis of the cultured C. meneghiniana, from which the Al species associated with the organic components of the Al-bearing lake diatoms were determined. Notably, the specific mechanisms, explaining why diatoms take

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**Fig. 2** Morphology and composition of a frustule of BSi-C. **a** Top view and **b** isometric view (tilted 55° from the top view) of the frustule of BSi-C in the field-emission scanning electron microscope (FESEM) images. **c** Top view and **d** side view of the three-dimensional (3D) Si distributions obtained by time-of-flight secondary ion mass spectrometry (TOF-SIMS). **e** Top view and **f** side view of the 3D Al distributions obtained by TOF-SIMS. The signal fading from left to right in the 3D images in (c) and (d) was caused by the edge effect; i.e., the primary ion beam came from the left side and resulted in unhomogenized signals due to the round shape of the frustule.⁴¹
SiBSi values of cultured and lake BSi are similar. Furthermore, two Al concentrations (1.0 and 2.0 μM) in the culture medium were selected for comparison. These concentrations do not further increase the incorporation of Al into lake BSi. From the perspective of the biogeochemical Al cycle, the abovementioned Al burial due to lake BSi accounts for ~9.1% of the total export of terrestrial Al in biomass to continental sediments (~1.90 × 10¹⁰ kg year⁻¹)⁶. Moreover, the magnitude of this Al sink is comparable to the magnitude ((0.4–2.9) × 10⁹ kg year⁻¹ (ref. 18)) of the input of dissolved Al into oceans by rivers globally. Based on the burial rate of Al due to BSi (~1.72 × 10⁹ kg year⁻¹) in global lakes and on an estimate that ~5–20% of total lake BSi is eventually buried in sediments⁷, the total Al uptake by the BSi of global lakes is on the order of (~0.86–3.44) × 10¹⁰ kg year⁻¹. Therefore, the Al occurrence in lake BSi, either in the waters or in the sediment, constitutes an important part of the global Al cycle.

The large Al pool due to Al incorporation in lake BSi, especially the Al sink through the burial of lake BSi, has not been considered in the scientific literature, but may have substantial effects on the global Al cycle. The scale of the terrestrial dissolved Al exported to oceans could be affected by the magnitude of Al burial in lakes, thereby mediating related marine geochemical processes, such as reverse weathering in which Al ions are active reactants. For modelling and interpretations of the global Al cycle, the Al uptake by lake BSi and its potential consequences must be taken into account.

Al incorporation into amorphous silica has long been known to be able to reduce silica solubility⁵,⁷–⁹,³⁸. The decrease in solubility of the BSi of cultured marine diatoms with Al incorporation was also confirmed. The mechanism of the abovementioned dissolution–inhibition effect has been ascribed to the production of negative charges by Al incorporation in silica, which repels OH⁻ and thereby retards the dissolution of silica⁷–⁹,³⁸. The Al/SiBSi values of the BSi of marine diatoms normally remain at a level of 10⁻⁴–10⁻³ in marine waters with generally low Al concentrations. The low extent of Al incorporation in marine BSi is thought to be unable to obviously affect the dissolution kinetics or the solubility of marine BSi⁶,⁹. In contrast, the noticeable Al incorporation in lake BSi substantially decreases its solubility.
In a 10-day dissolution test aiming at examining the dissolution–inhibition effect of Al-bearing lake BSi, the Al-scarce BSi showed high dissolution extent, 90.2% (wt%) for the BSi from the cultured *C. meneghiniana* and 79.6% for that from the cultured *N. palea* (details in Supplementary Fig. 5). However, the 10-day dissolution extents of the Al-bearing BSi with different Al/Si values decreased to 49.6% for *C. meneghiniana* diatoms and 67.7% for *N. palea* diatoms. Energy-dispersive X-ray spectroscopy (EDS) spot analysis was used for the determination, and the data of more than 100 spots for each sample. Error bounds were obtained on the basis of 95% confidence interval for the analysis on Al/Si atomic ratios.

The consequences of such a distinct dissolution–inhibition effect of lake BSi deserve to be considered in the preservation of lake BSi. The annual burial rate of lake BSi normalized to the total area of global lakes is 18.2 g m\(^{-2}\) year\(^{-1}\) (ref. \(^{20}\)). This value is ~11.5 times greater than that of marine BSi (1.58 g m\(^{-2}\) year\(^{-1}\)) normalized to the area (~2.40 × 10\(^9\) km\(^2\)) of BSi-containing oceanic regions\(^{14}\). Given the key role of diatoms in the closely coupled Si and C cycles through the biological pump\(^{12}\), this result suggests that the biological pump driven by diatoms in lakes is more efficient than its counterpart in oceans, making the former biological pump more conducive to CO\(_2\) sequestration. The pronounced dissolution–inhibition effect likely contributes to the high efficiency of sedimentation and preservation of lake BSi, although quantifying this contribution is difficult at the present stage due to the lack of data of other variables, such as the scale of biological scavenging\(^{5}\). The role of lake BSi for carbon sequestration over geological history or during the present, and the

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**Fig. 4** Al and Si atomic percentages in frustules and entire diatoms. a The data of Al and Si atomic percentages for BSi-C; b the data for BSi-N; c the data for the entire *Cyclotella meneghiniana* diatoms; and d the data for the entire *Nitzschia palea* diatoms. Energy-dispersive X-ray spectroscopy (EDS) spot analysis was used for the determination, and the data of more than 100 spots (specified in the brackets of a–d) from over 35 entire diatoms or their BSi were collected. \(R^2\) is the correlation coefficient between the atomic percentages of Al and Si (the linear fitting formulas are given in Supplementary Table 1). Source data are provided as a Source Data file.

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**Fig. 5** Al/Si atomic ratios of cultured diatoms and their frustules. a The average Al/Si atomic ratios of the entire *Cyclotella meneghiniana* diatoms (black columns) cultured in media with various Al concentrations and of their biogenic silica (BSi) (grey columns). b The data of the entire *Nitzschia palea* diatoms (black columns) and of their BSi (grey columns). The energy-dispersive X-ray spectroscopy (EDS) spot analysis for the determination of Al/Si atomic ratios is based on over 100 spots for each sample. Error bounds were obtained on the basis of 95% confidence interval for the analysis on Al/Si atomic ratios. Source data are provided as a Source Data file.
The carbon contents of the BSi extracted from the diatoms were all below the X-ray spectrometer. The samples for SEM characterization were pre-treated by 6N HNO3 for 2 h, and a different collector was used for N. palea. The living diatoms were picked from the collected specimens using an Olympus Microscope (CX31) at 1000× magnification. The identification of the species of the living diatoms was achieved by determining the rbcL gene sequences of the diatoms (Supplementary Fig. 8). Before the analysis such as determination of Al/Si atomic ratios through EDS analysis, diatom samples were cleaned in 0.1 N nitric acid for 72 h and rinsed three times with deionized water, and the products were subjected to centrifugation and subsequent freeze-drying.

Cultivation of diatoms. Cultured diatoms of the same two species as the abovementioned living diatoms were provided by the Freshwater Algae Culture Collection at the Institute of Hydrobiology (FACHB), Chinese Academy of Sciences. After successive culturing and harvesting over many generations in an Al-free medium, these diatoms contain very little Al (Al/Si atomic ratios <0.001). These Al-scarce diatoms were used as control samples for comparison. The diatoms were grown in the culture medium (details of the components are given in Supplementary Table 3) at a temperature of 25°C under a 12/12 light/dark cycle at an intensity of 71 umol photons m−2 s−1 for 480 h (one diatom life cycle). The pH of the diatom culture medium was adjusted to 8.0 using a solution of NaOH. This pH is close to the pH of the abovementioned lakes where the natural lake diatoms were collected. The pH of the culture medium during culture process is in the range of 8.0–8.4. Al-bearing diatoms were obtained by adding AlCl3 to the diatom medium to reach designated Al concentrations of 1.0, 2.0, 5.0, 10.0, 50.0, and 100.0 μM. The diatoms cultured in the above-mentioned weakly alkaline condition of this work have a high level of Al tolerance, unlike the pronounced ecotoxicity of Al to organisms shown at the acidic or circumneutral conditions. The growth inhibition of the diatom cells in the presence of Al was found to be minor, even for the high Al concentrations of 50.0 and 100.0 μM (Supplementary Fig. 9). The cultured diatoms were treated according to the same procedures described above for further analysis, such as microscopic characterization.

Extraction of BSI. Two chemical treatments were used to obtain the BSI samples from the natural lake diatoms and the cultured diatoms, including immersion in a 30% hydrogen peroxide solution for 72 h to remove the organic matrix of the diatoms, and then washing in 0.1 N acetic acid for 72 h to remove non-structural Al. These steps were repeated until complete removal of the organic components was achieved. To confirm the removal of organics, carbon elemental analysis on the BSI was performed using a Vario EL III elemental analyzer, in which samples were oxidized in a combustion tube in the presence of oxygen at high temperature (≥1150°C). Samples of ~2.00 mg were placed in tin boats with a size of 6 × 12 mm. The carbon contents of the BSI extracted from the diatoms were all below the detection limit (<40 ppm).

Determination of Al/Si atomic ratios. The Al/Si atomic ratios in the natural lake diatoms and the cultured diatoms and their BSI were determined by an EDS method using a Hitachi SU9000 FESEM attached to an AMETEK energy dispersive X-ray spectrometer. The samples for SEM characterization were pre-treated by carbon coating. Over 100 spots from more than 35 entire diatoms and their BSI of each diatom species were detected using FESEM–EDS spot analysis with a voltage of 15 kV and a current of 20 μA. For the cultured diatoms and their BSI, 103 spots of EDS spot analysis for each sample were collected.

FIB-EDS mapping analysis. FIB-EDS mapping analyses of the natural lake BSI were carried out to reveal the Al and Si distributions in the BSI samples. This process was achieved using a FIB-SEM (GAIA3, Tescan) equipped with a Trilag FESEM column, a Coba FIB column, a multichannel gas injection system, and an Oxford Instrument X-Max 80 Atecs energy-dispersive X-ray spectrometer. A single frustule of BSI selected for characterization was picked using a nanomanipulator (Oxford OmniProbe 400), and the sample surface was milled using a focused gallium ion (Ga+) beam with an accelerating voltage of 30 kV, a current of 1.0 nA, and a total milling time of 1 min.

TOP-SIMS analysis. A frustule of C. meneghiniana BSI (BSC-C) was analysed by TOF-SIMS to identify the 3D elemental distribution of Al and Si in the BSI. The SIMS instrument was equipped with a Ga+ TOF analyzer (GATAN X-TOF analyzer, Tescan); a liquid metal ion gun with 30 keV of Ga+, an ion beam current of 25 pA, and an angle of incidence to the normal of 55° for bombarding the surface; and an electron beam for imaging. Each layer of BSI-C is produced with an ion beam for surface analysis, and a sputtering ion beam is used to ablate the surface layer.

Al K-edge XANES analysis. Al K-edge XANES analysis was performed on the BL08U beamline of the Shanghai Synchrotron Radiation Facility (SSRF, Shanghai, China). A dispersion was prepared by mixing BSI with ultrapure water and subsequently shaking the mixture to disperse the samples. The dispersion was dropped onto tin foil and smeared evenly. After the dispersion was air-dried, the tin foil coated with BSI was positioned at a 90° angle to the incident X-ray beam. Al K-edge XANES spectra were recorded in total electron yield mode in a vacuum chamber (<10−3 torr, 25°C) from 1558 eV to 1576 eV across the Al K-edge with a step of 0.1 eV and a dwell time of 2 s, and the measurement was repeated three times at each energy level. The XANES spectra were then normalized with the IFEFFIT program.

Dissolution experiment of Al-bearing BSI. To determine the effect of Al incorporation on the solubility of BSI, the BSI samples extracted from the cultured C. meneghiniana and N. palea were used. A series of BSI samples with average Al/Si atomic ratios ranging from <0.001 (Al-scarce BSI) to 0.045–0.070 (Al-bearing BSI) were used for the dissolution experiment; 0.50 mg of BSI sample was dissolved in 100 mL of deionized water in a Teflon beaker at 25°C. After 10 days of dissolution, the resulting solution was centrifuged and 1.0 mL of solution from the supernatant was collected for the determination of Si concentration, which was performed using a Thermo Scientific iCAP 7000 inductively coupled plasma atomic emission spectrometer (ICP-AES). Before ICP-AES analysis, 1.0 mL of 2.0% HNO3 was added into the solution and the resultant extract solution was left overnight. The parameters of ICP-AES were as follows: a power of 1150 W, a plasma gas flow rate of 12 L min−1, a coating gas flow rate of 0.5 L min−1, and a nebulizer pressure of 200 kPa. Digestions of the blank sample were simultaneously carried out for calibration. The 10-day dissolution extent of BSI was calculated using the following equation:

$$D(%) = \frac{C_{0} - C_{t} \times 10^{3}}{C_{0} \times 10^{3}} \times 100 \times 10^{3} \times 0.5$$

where D (%) is the 10-day dissolution extent of BSI, M (mg L−1) is the concentration of Si detected by ICP-AES, and the atomic weights of Si and SiO2 are 28.1 and 60.1 g mol−1, respectively.

Reporting Summary. Further information on research design is available in the Nature Research Reporting Summary linked to this article.

Data availability
The authors declare that the main data supporting the findings of this study are available within the article and its Supplementary Information files. Extra data are available from the corresponding author upon request. The source data underlying Figs. 3, 4, and 5 and Supplementary Figs. 2, 3, and 9 are provided as a Source Data file.

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References
1. Driscoll, C. T. & Postek K. M. The chemistry of aluminum in surface waters. In The Environmental Chemistry of Aluminum 2nd edn (ed Spósito, G.) Ch. 9, 364–366 (CRC Press, Boca Raton, 1995).
2. Isson, T. T. & Planavsky, N. J. Reverse weathering as a long-term stabilizer of marine pH and planetary climate. Nature 560, 471 (2018).
3. Driscoll, C. T. & Schecher, W. D. The chemistry of aluminum in the environment. Environ. Geochim. Health 12, 28–49 (1990).
4. Moran, S. B. & Moore, R. M. Evidence from mesocosm studies for biological removal of dissolved aluminum from sea water. Nature 335, 796–798 (1988).
5. Van Cappellen, P., Dixit, S. & van Beusekom, J. Biogenic silica dissolution in the oceans: Reconciling experimental and field-based dissolution rates. Global Biogeochem. Cycles 16, 23–1–23–10 (2002).
6. Loucaides, S., Cappellen, P. V., Roubeix, V., Moriceau, B. & Ragueneau, O. Controls on the recycling and preservation of biogenic silica from biomineralization to burial. Silicon 4, 7–22 (2012).
7. Van Bennekom, A. J., Fred Jensen, J. H., van der Gaast, S. J., Van Iperen, J. M. & Pieters, I. Aluminium-rich opal: an intermediate in the preservation of biogenic silica in the Zaire (Congo) deep-sea fan. Deep Sea Res. A 36, 173–190 (1989).
8. Gehlen, M. et al. Unraveling the atomic structure of biogenic silica: evidence of the structural association of Al and Si in diatom frustules. Geochim. Cosmochim. Acta 66, 1601–1609 (2002).
9. Dixit, S., Van Cappellen, P. & van Bennekom, A. J. Processes controlling solubility of biogenic silica and pore water build-up of silicic acid in marine sediments. Mar. Chem. 73, 333–352 (2001).

10. Boyd, P. W., Ellwood, M. J., Tagliabue, A. & Twinning, B. S. Biotic and abiotic retention, recycling and remineralization of metals in the ocean. Nat. Geosci. 10, 167–173 (2017).

11. Armbrust, E. V. The life of diatoms in the world’s oceans. Nature 459, 185–192 (2009).

12. Tréguer, P. et al. Influence of diatom diversity on the ocean biological carbon pump. Nat. Geosci. 11, 27–37 (2018).

13. Conley, D. J. & Carey, J. C. Silica cycling over geologic time. Nat. Geosci. 8, 431–432 (2015).

14. Tréguer, P. J. & De La Rocha, C. L. The world ocean silica cycle. Annu. Rev. Mar. Sci. 5, 477–501 (2013).

15. Kröger, N., Deutzmann, R. & Sumper, M. Polycationic peptides from diatom biosilica that direct silica nanosphere formation. Science 286, 1129–1132 (1999).

16. Parker, A. R. & Townley, H. E. Biomimetics of photonic nanostructures. Nat. Nanotechnol. 2, 347–350 (2007).

17. Mackenzie, F. T., Stoffyn, M. & Wollast, R. Aluminium in seawater: control by biological activity. Science 199, 680–682 (1978).

18. Stoffyn, M. & Mackenzie, F. T. Fate of dissolved aluminium in the oceans. Mar. Chem. 11, 105–127 (1982).

19. Gehlen, M., Heinze, C., Maier-Reimer, E. & Measures, C. I. Coupled Al-Si pump. Science 286, 431–432 (1999).

20. Gehlen, M., Heinze, C., Maier-Reimer, E. & Measures, C. I. Rapid post-burial diagenesis of aluminosilicate minerals. Earth Planet. Sci. Lett. 205, 1260–1262 (2002).

21. Vrieling, E. G., Poort, L., Beelen, T. P. M. & Gieskes, W. W. C. Growth and chemical image using TOF-SIMS revealing the biopolymer component spatial distributions in biomass of freshwater diatom species. Protist 153, 261–273 (2002).

22. Finlay, B. J., Monaghan, E. B. & Maberly, S. C. Hypothesis: The rate and scale of dispersal of freshwater diatom species is a function of their global abundance. Prog. Oceanogr. 58, 2647–2660 (2011).

23. Schindler, D. W. et al. Long-term ecosystem stress: the effects of years of experimental acidification on a small lake. Science 228, 1395–1401 (1985).

24. Hydes, D. J. Aluminium in seawater: control by inorganic processes. Science 205, 1260–1262 (1979).

25. Orions, K. J. & Bruland, K. W. The biogeochemistry of aluminium in the Pacific Ocean. Earth Planet. Sci. Lett. 78, 397–410 (1986).

26. Orions, K. J. & Bruland, K. W. Dissolved aluminium in the central North Pacific. Nature 316, 427–429 (1985).

27. Middag, R., van Slooten, C., de Baar, H. J. W. & Laan, P. Dissolved aluminium in the Southern Ocean. Deep Sea Res. Part II Top. Stud. Oceanogr. 58, 407 (1999).

28. Finlay, B. J., Monaghan, E. B. & Maberly, S. C. Hypothesis: The rate and scale of dispersal of freshwater diatom species is a function of their global abundance. Prog. Oceanogr. 58, 2647–2660 (2011).

29. Li, D. E., Bancroft, G. M., Fleet, M. E., Feng, X. H. & Pan, Y. M. Al K-Edge XANES spectra of aluminosilicate minerals. Am. Mineral. 80, 432–440 (1995).

30. Ildefonse, P. et al. Aluminium X-ray absorption near edge structure in model compounds and Earth’s surface minerals. Phys. Chem. Miner. 25, 112–121 (1998).

31. Ingall, E. D. et al. Role of biogenic silica in the removal of iron from the Antarctic seas. Nat. Commun. 4, 1–6 (2013).

32. Van Hulten, M. M. P. et al. On the effects of circulation, sediment resuspension and biological incorporation by diatoms in an ocean model of aluminium. Biogeoosciences 11, 3757–3779 (2014).

33. Vrieling, E. G., Poort, L., Beden, T. P. M. & Gieskes, W. W. C. Growth and silica content of the diatoms Thalassiosira weissflogii and Navicula salinarum at different salinities and enrichments with aluminium. Eur. J. Phycol. 34, 307–316 (1999).

34. Koning, E., Gehlen, M., Flank, A. M., Galas, G. & Epping, E. Rapid post-mortem incorporation of aluminium in diatom frustules: evidence from chemical and structural analyses. Mar. Chem. 106, 208–222 (2007).

35. Zhou, L., Tan, Y., Huang, L., Fortin, C. & Campbell, P. G. C. Aluminium effects on marine phytoplankton: implications for a revised iron hypothesis (Iron–Aluminium Hypothesis). Biogeochemistry 139, 123–137 (2018).

36. Rauch, J. N. & Pacyna, J. M. Earth’s global Ag, Al, Cr, Cu, Fe, Ni, Pb, and Zn cycles. Global Biogeochem. Cycles 23, GB2001 1–16 (2009).

37. Leng, M. J. & Barker, P. A. A review of the oxygen isotope composition of lacustrine diatom silica for palaeoclimate reconstruction. Earth Sci. Rev. 75, 5–27 (2006).

38. Iler, R. K. The occurrence, dissolution, and deposition of silica. In The Chemistry of Silica Solubility: Polymerization Colloid and Surface Properties and Biochemistry. (ed Iler, R. K.) Ch. 1, 40–58 (John Wiley & Sons Inc., Press, New York, 1979).

39. Couradeau, E. et al. An early-branaching microbavitae cyanobacterium forms intracellular carbonates. Science 336, 459–462 (2012).

40. Gensemer, R. W. & Playle, R. C. The bioavailability and toxicity of aluminium in aquatic environments. Crit. Rev. Environ. Sci. Technol. 29, 315–450 (1999).

41. Vering, G., Crone, C., Bijma, J. & Arlinghaus, H. F. TOF-SIMS characterization of planktonic foraminifera. Appl. Surf. Sci. 203–204, 785–788 (2003).

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Author contributions

D.Li and P.Y. designed the experiments and wrote the manuscript. Q.T., H.G., and W.F. collected the samples of lake diatoms. Q.T., Y.S., and J.Y.Z. carried out the culture of diatoms and EDS analysis. H.L. and Q.T. performed the XANES analysis. D.L. was involved in the discussion on the data analysis and revision of the draft. L.D., J.M.Z., J.Y.Z., H.S., and W.F. pretreated the lake or cultured diatoms and were involved in the TOF-SIMS and FIB analysis.

Competing interests

The authors declare no competing interests.

Additional information

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Correspondence and requests for materials should be addressed to P.Y.

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