The MUC5B promoter polymorphism and telomere length in patients with chronic hypersensitivity pneumonitis

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RESEARCH IN CONTEXT

Evidence before this study

We searched Pubmed for studies published before January 14, 2017 with the search terms “MUC5B”, “TOLLIP”, “telomere length”, “hypersensitivity pneumonitis”, and “alveolitis, extrinsic allergic”. No studies were identified reporting associations with the mucin 5B (MUC5B) rs35705950 or toll interacting protein (TOLLIP) rs5743890 single nucleotide polymorphisms (SNPs) and hypersensitivity pneumonitis (HP). Two studies reported peripheral blood leukocyte (PBL) telomere length in HP, but did not examine associations with clinical features or outcomes.

Added value of this study

This is the first study to demonstrate that the MUC5B SNP rs35705950 minor allele, but not the TOLLIP SNP rs5743890, is associated with risk of chronic HP and extent of fibrosis. It also demonstrates that shorter PBL telomere length is associated with extent of fibrosis, microscopic honeycombing, and reduced survival in chronic HP.

Implications of all the available evidence

The MUC5B SNP rs35705950 minor allele and shorter PBL telomere length are associated with greater extent of fibrosis, and shorter PBL telomere length is associated with histopathologic findings typical of usual interstitial pneumonia and reduced survival in patients with chronic HP. These findings suggest their potential role in risk stratification of patients with chronic HP, and potentially shared pathobiology between IPF and chronic HP.
SUMMARY

**Background:** Patients with hypersensitivity pneumonitis (HP) may develop lung fibrosis, which is associated with reduced survival. Families with pulmonary fibrosis can present with members diagnosed with idiopathic pulmonary fibrosis (IPF) or HP, suggesting that fibrotic HP may share risk factors with IPF.

**Methods:** In an observational study of two independent cohorts of patients with chronic HP (cHP) (UCSF n=145, UTSW n=72), we measured two common single nucleotide polymorphisms associated with IPF (MUC5B rs35705950 & TOLLIP rs5743890) and peripheral blood leukocyte telomere length and evaluated their associations with cHP disease, survival, and clinical-radiograph-pathologic features.

**Findings:** The frequency of the MUC5B minor allele, but not the TOLLIP minor allele, was significantly increased in cHP patients in both cohorts (UCSF MAF 24.4% & UTSW MAF 32.3%) compared to healthy controls (MAF 10.7%; p-values for comparison = <0.0001 for both cohorts) and similar to IPF (UCSF MAF 33.3% & UTSW MAF 32.0%, p-values for comparison=0.10 & 0.95, respectively). The MUC5B minor allele (adjusted OR 1.91, p=0.045) and shorter telomere length (adjusted OR 0.23, p=0.002) were associated with extent of radiographic fibrosis and other measures of lung remodeling and fibrosis in the combined cHP cohorts. Shorter telomere length had a significant association (adjusted HR 0.18, p=0.001) with reduced survival in the combined cHP cohorts.
Interpretation: The MUC5B promoter polymorphism rs35705950 and shorter telomere length are associated with extent of fibrosis in cHP. Shorter telomere length is associated with histopathology findings typical of usual interstitial pneumonia and reduced survival in cHP.

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INTRODUCTION

Hypersensitivity pneumonitis (HP) is an inflammatory lung disease caused by inhalational exposure to a variety of organic antigens that has a heterogeneous clinical presentation and natural history.\(^1\) Heterogeneity of HP is represented most commonly by a clinical classification system that uses the temporal designations of acute, sub-acute, and chronic HP.\(^1\) Lung fibrosis is the feature that most clearly identifies patients with HP that will have a chronic, progressive course and a poor prognosis.\(^2\) In fact, fibrotic HP often has a clinical presentation similar to idiopathic pulmonary fibrosis (IPF), the most deadly form of idiopathic interstitial pneumonia, including progressive lung fibrosis and reduced survival.\(^2,3\)

The development of lung fibrosis in HP is thought to be due to persistent lung injury resulting from ongoing antigen exposure and inflammation. Epidemiologic studies confirm that only a subset of individuals exposed to an inciting antigen develop HP, suggesting a possible genetic contribution to the disease.\(^4-7\) However, the genomic factors that predispose individuals with HP to developing lung fibrosis are not known.\(^1\) Several single nucleotide polymorphisms (SNPs) have been associated with predisposition to IPF,\(^8-12\) and two of these, the mucin 5B (MUC5B) rs35705950 and toll interacting protein (TOLLIP) rs5743890 SNPs, have also been associated with survival in established IPF.\(^8,9\) In addition, variants in genes associated with telomere maintenance have been linked to IPF,\(^11,13-19\) and short telomere lengths measured in peripheral blood leukocytes (PBLs) have been associated with poorer survival in patients with IPF.\(^20\) Families with pulmonary fibrosis often include members
diagnosed clinically with chronic HP, suggesting that telomere dysfunction may be a risk factor for the development of chronic fibrotic HP.\textsuperscript{13-19,21}

We propose here that genomic risk factors associated with the development and progression of IPF may also be associated with the development of fibrosis and reduced survival in chronic HP (cHP). To test this, we measured two SNPs that have been associated with a predisposition to IPF and survival in IPF (MUC5B rs35705950 and TOLLIP rs5743890) as well as PBL telomere length in two well-characterized cohorts of patients with cHP, and specifically examined their association with (1) cHP (compared to IPF and controls), (2) radiographic and histopathologic features of remodeling and fibrosis in cHP, and (3) survival in cHP.

**METHODS**

**Study Design and Populations**

*Hypersensitivity pneumonitis cohorts*

This is an observational cohort study of patients with cHP drawn from two academic interstitial lung disease (ILD) centers: the University of California San Francisco (UCSF) and the University of Texas Southwestern (UTSW). Patients seen at UCSF and UTSW were prospectively enrolled into separate longitudinal ILD cohort studies from October 2001-April 2016 (UCSF) and from August 2005-August 2016 (UTSW). Baseline clinical information and blood samples were collected at the time of enrollment. The diagnosis in both cohorts were made by in-person multidisciplinary team discussion (MDD) according to available guidelines.\textsuperscript{22,23} All patients with a
MDD diagnosis of cHP who had a DNA sample collected from peripheral blood were included in the study. A diagnosis of cHP required a compatible clinical presentation and either (1) HRCT features of sub-acute/chronic HP\textsuperscript{24,25} AND identification of a plausible exposure or (2) surgical lung biopsy histopathology most consistent with cHP.\textsuperscript{26-28} In all cases, an alternative connective tissue disease was excluded.

Clinical information collected from both cohorts included demographics, pulmonary function tests, and radiologic and histopathologic studies. Ethnicity was self-reported. The date of death was recorded for both cohorts and confirmed using the United States Social Security Death Index for the UCSF cohort. Lung transplants and date of lung transplants were recorded.

Control cohorts

SNP frequencies in cHP patients were compared to two control populations: (1) patients with IPF identified from UCSF and UTSW and (2) the European population available from the 1000 Genomes Project Phase3, version 1.\textsuperscript{29} We compared telomere length in cHP patients to IPF patients from UCSF and UTSW.\textsuperscript{20}

Measurements

DNA

PBL genomic DNA was isolated in the UCSF cohort using the Gentra Puregene cell kit and in the UTSW cohort using Autopure LS (both from Qiagen, Valencia, CA, USA). MUC5B rs35705950 and TOLLIP rs5743890 SNPs were measured using the Taqman
SNP Genotyping Assays (Applied Biosystems, Foster City, CA, USA) independently at both institutions. PBL telomere length was measured for both cohorts at UTSW using quantitative PCR as previously described; samples were excluded for pre-specified DNA quality and concentration or sample volume criteria.\textsuperscript{15,20,30} Telomere length was expressed as the natural logarithm-transformed ratio of the telomere to single gene copy (log T/S), the difference between the observed and expected for age, and the age-adjusted percentile.\textsuperscript{15,20,30}

\textit{Radiology}

All available high-resolution computed tomography images of the chest (HRCTs) were reviewed by expert chest radiologists (B.M.E. or T.S.H. for UCSF and K.B. for UTSW). Radiologists scored HRCTs for pattern (definite, possible, or inconsistent with usual interstitial pneumonia)\textsuperscript{22}; semi-quantitative extent of fibrosis as the percentage of total lung volume involved (none, mild [$<10\%$], moderate [10-50\%], and severe [$>50\%$])\textsuperscript{31}; and presence or absence of honeycombing, traction bronchiectasis, upper/mid-lung predominance of abnormalities, peribronchovascular distribution of abnormalities, ground glass opacities outside of areas of fibrosis, consolidation, profuse micronodules, cysts away from areas of honeycombing, and significant (defined as three or more lobes) mosaic perfusion and/or air-trapping.\textsuperscript{22,32}

\textit{Histopathology}
In the UCSF cohort, surgical lung biopsies are reviewed prospectively in MDD at the time of diagnosis and scored by an expert lung pathologist (K.J.D.) using a structured pathology data collection form (see supplemental Figure 1). In the UTSW cohort, surgical lung biopsies were reviewed and scored by a separate expert lung pathologist (J.T.) specifically for this study using the same form.

**Statistical Analysis**

Bivariate associations of genotype (coded as a binary response variable for any minor allele vs. homozygous wild-type) with binary clinical, radiologic, and histopathologic variables were evaluated using the Chi-squared or Fisher’s exact test where appropriate; and with continuous clinical variables and telomere length (log T/S) using Student’s t test. Telomere length (log T/S) association with binary clinical, radiologic, and histopathologic variables was evaluated using Student’s t test; telomere length association with continuous clinical variables was evaluated with Pearson’s correlation. Statistical significance (i.e., alpha level) was determined using the Hochberg procedure for multiple testing within categories of comparison (e.g., MUC5B and radiographic features).33

For the SNP genotype association analyses, minor allele frequency (MAF) was calculated in each population as the total number of minor alleles divided by the total number of alleles in the population. MAF in the HP populations was compared to IPF and control populations using the Chi-squared test. For the primary analysis, comparisons were made in the population restricted to non-Hispanic white
individuals to control for confounding by ethnicity (i.e. population stratification). All associations were considered significant at alpha < 0.05 without correction for multiple comparisons. MAFs were reported along with exact binomial 95% confidence intervals (CIs).

Primary survival analyses were evaluated as time-to-death from any cause with right censoring for alive at end of study or lung transplantation using Cox proportional hazards models. Transplant-free survival (where lung transplantation is considered equivalent to death in Cox models) and Fine-Gray competing risks regression models (where lung transplant is considered a competing risk) were performed to evaluate sensitivity of results to the method of handling of lung transplantation. In the individual cohorts, survival associations were evaluated both unadjusted and adjusted for age, sex, baseline forced vital capacity (FVC) % predicted, and diffusing capacity of the lung for carbon monoxide (DLCO) % predicted. For the combined cohort, a stratified Cox proportional hazards model was used to evaluate survival and transplant-free survival; for the competing risks analyses, cohort was included as a covariate. Kaplan Meier plots were constructed comparing survival and transplant-free survival by telomere length using an age-adjusted percentile cut-off of 10%, and groups were compared using the log-rank test. The 10% cutoff was chosen because the vast majority (>80%) of patients with telomere-maintenance machinery mutations have telomere length <10th percentile, and this cutoff represents a biologically relevant group of patients in which manifestations of telomeropathies are more prevalent. To evaluate the
independent association of telomere length and MUC5B SNP with radiographic extent of fibrosis in the combined cohort, a multivariate ordinal logistic regression model was constructed with the outcome variable extent of radiographic fibrosis (0-none, 1-mild = 0-10%, or 2-moderate to severe = >10%) and predictor variables age (in years), sex (male vs. female), MUC5B SNP (any minor allele vs. homozygous wild-type), telomere length (in log T/S), and cohort (UCSF vs. UTSW). Interactions were assessed by testing the statistical significance of product terms between cohort and MUC5B or telomere length.33

RESULTS

Cohort characteristics

In the UCSF cohort, 145 patients with cHP were genotyped for the MUC5B rs35705950 and TOLLIP rs5743890 SNPs; telomere length was measured in 129 (89%) after exclusions for pre-specified quality criteria. In the UTSW cohort, 72 patients with cHP were genotyped for MUC5B rs35705950 and TOLLIP rs5743890 SNPs; all had PBL telomere length measured. The mean age was nearly 60 years in both cohorts, 61% were female in the UCSF and 47% in the UTSW cohort, and nearly half were ever-smokers in both cohorts (Table 1). The majority of patients identified as non-Hispanic white in both cohorts (85% in UCSF and 90% in UTSW). A family history of ILD was more common in the UTSW cohort (23.6%) than in the UCSF cohort (10.3%). Diagnosis was informed by either transbronchial lung biopsy or surgical lung biopsy in 70% or more of the cases, more than two-thirds had a potential antigen exposure identified, and nearly all patients had a HRCT pattern
inconsistent with usual interstitial pneumonia (UIP). Of those with available HRCT scans for review, 79% and 94%, had radiographic signs of fibrosis in the UCSF and UTSW cohorts, respectively.

**MUC5B, TOLLIP, and Telomere Length Associations with Clinical Features**

The presence of any MUC5B rs35705950 minor allele (genotype TT or GT compared to GG) was associated with older age at diagnosis in both cohorts (UCSF: 65.6 vs. 61.7 years, p=0.037; UTSW: 63.6 vs. 57.2 years, p=0.004), but not sex, smoking history, or an identifiable exposure. The presence of any TOLLIP rs5743890 minor allele (genotype GG or AG compared to AA) was not associated with age, sex, smoking history, or identifiable exposure. Shorter telomere length was significantly associated with older age in the UCSF cohort (r = -0.31, p=0.0003) but not significantly in the UTSW cohort (r = -0.19, p=0.10). Telomere length was not associated with sex, smoking history, exposure identification, or MUC5B rs35705950 genotype in either cohort.

**MUC5B and TOLLIP SNP Association Analysis**

In non-Hispanic white patients with cHP, the MUC5B rs35705950 minor allele frequency was increased in both the UCSF (MAF = 24.4% [95%CI 19.2-30.3]) and UTSW (MAF = 32.3% [95%CI 24.4-41.1]) cohorts compared to a publically available European healthy control population (MAF = 10.7%, p = < 0.0001 for comparison in both cohorts) and was comparable to patients with IPF (Table 2). The TOLLIP rs5743890 MAF was not significantly different in cHP patients compared to IPF or
healthy controls (Table 2). Results for SNP MAF associations were similar for the entire cohort (i.e. not restricted to the non-Hispanic white population, see supplemental Table 1).

**Telomere Length**

The mean telomere length was longer in the UCSF HP cohort compared to the UTSW HP cohort (mean log T/S 1.53+/−0.37 vs. 1.30+/−0.33, p = < 0.0001; mean observed-expected for age difference 0.026+/−0.358 vs. -0.203+/−0.0328, p = <0.0001, see Table 1). Compared to IPF, telomere length was longer in the combined cohort of HP patients (mean log T/S in HP 1.45+/−0.37 vs. in IPF 1.32+/−0.35, p = 0.0005; mean observed-expected for age difference in HP -0.058+/−0.364 vs. in IPF -0.166+/−0.360, p = 0.0034).

**MUC5B, TOLLIP, and Telomere Length Associations with Radiographic Features**

For the two cohorts, 189 HRCTs were available for scoring by a radiologist (119/145 [82%] in the UCSF cohort and 70/72 [97%] in the UTSW cohort). In the combined cohort, the MUC5B rs35705950 minor allele was associated with moderate-severe radiographic fibrosis (p=0.009) and the presence of traction bronchiectasis (p=<0.001), but not a pattern consistent with definite or possible UIP or presence of radiographic honeycombing (Table 3). Among features inconsistent with UIP, the MUC5B minor allele was associated with the absence of diffuse ground glass opacities (p=0.012) but not other inconsistent features such as upper-mid lung
distribution, peribronchovascular distribution, micronodules, or mosaic perfusion/air-trapping.

In the combined cohort with HRCT scores and telomere length measurements (n=175), shorter telomere length was associated with moderate to severe radiographic fibrosis (<0.0001), a pattern consistent with definite or possible UIP (p=0.03), presence of honeycombing (p=0.002), and presence of traction bronchiectasis (p=0.0007). Among the inconsistent with UIP features, shorter telomere length was associated with the absence of diffuse ground glass opacities (p=0.004) but not other inconsistent features such as upper-mid lung distribution, peribronchovascular distribution, micronodules, or mosaic perfusion/air-trapping. The TOLLIP rs5743890 minor allele was not associated with any HRCT feature (data not shown). Results were consistent across individual cohorts (supplement Table 2). In the combined UCSF and UTSW HP cohorts, the MUC5B minor allele (adjusted OR 1.91, p=0.045) and shorter telomere length (adjusted OR 0.23, p=0.002) were independently associated with extent of radiographic fibrosis, after adjustment for age and sex (Table 4). There was no evidence for a significant interaction between MUC5B and telomere length, MUC5B and cohort, or telomere length and cohort on extent of radiographic fibrosis.

*MUC5B, TOLLIP, and Telomere Length Associations with Histopathologic Features*

A total of 75 surgical lung biopsies were available for histopathologic scoring (54/145 [37%] in the UCSF cohort and 16/72 [22%] in the UTSW cohort). In the
combined cohort, the MUC5B rs35705950 minor allele was not significantly associated with features of lung remodeling or fibrosis (Table 3), nor was it associated with any typical features of HP histopathology such as lymphocytic interstitial infiltrate (p=0.29), interstitial granulomas (p=0.08), or small airway disease (0.72).

In the combined cohort with histopathology scores and telomere length measurements (n=70), a shorter telomere length was associated with the presence of any histopathologic fibrosis (p=<0.0001), microscopic honeycombing (p=0.002), a heterogeneous distribution of fibrosis (p=0.0006), and a moderate to marked profusion of fibroblastic foci (p=0.003). Telomere length was not associated with any typical features of HP histopathology such as lymphocytic interstitial infiltrate (p=0.95), interstitial granulomas (p=0.09), or small airway disease (0.97). The TOLLIP rs5743890 minor allele was not associated with any of these histopathologic features (data not shown). Results were consistent across individual cohorts (supplement Table 2).

**MUC5B, TOLLIP, Telomere Length and Survival**

The presence of any minor allele for MUC5B rs35705950 SNP was of borderline statistical significance with worse survival (censoring for lung transplant) in the combined UCSF & UTSW cHP cohorts (adjusted HR 2.01, 95% CI 0.97-4.20, p=0.061) (Table 5). The TOLLIP rs5743890 SNP was not associated with survival in either cohort or the combined cohort. Shorter PBL telomere length was associated with
worse survival (censoring for lung transplant) in both cohorts of cHP patients and the combined cHP cohort (adjusted HR 0.18, 95%CI 0.06-0.51, p=0.001) (Figure 2). Results were similar for transplant-free survival and when treating lung transplant as a competing risk (supplemental Tables 3 & 4 and Figure 2). There were no significant interactions for MUC5B and telomere length, MUC5B and cohort, or telomere length and cohort on survival.

**DISCUSSION**

This is the first study reporting on MUC5B rs35705950 and TOLLIP rs5743890 single nucleotide polymorphisms and peripheral blood leukocyte telomere length associations with clinical features and outcomes in patients with HP. Shorter PBL telomere length and the MUC5B promoter variant rs35705950, but not the TOLLIP variant rs5743890, were found to be associated with fibrosis in cHP patients. Shorter PBL telomere length is also strongly associated with histopathology features typical of usual interstitial pneumonia and reduced survival in cHP patients. Prior to this study, the MUC5B SNP appeared to confer specific risk for idiopathic interstitial pneumonia and familial pulmonary fibrosis.\textsuperscript{34} We speculate that the MUC5B SNP and telomere dysfunction may predispose HP patients to development of lung remodeling and fibrosis, which in turn leads to worse outcomes.

The MUC5B promoter polymorphism rs35705950 minor allele has been repeatedly shown to occur more frequently in both sporadic and familial forms of IPF and thus far is the strongest identified genetic risk factor for IPF.\textsuperscript{8,10-12,34} How this
polymorphism contributes to IPF pathogenesis remains unclear. The minor allele causes increased MUC5B production by distal bronchiolar epithelial cells, and MUC5B protein accumulates in the honeycomb cysts of IPF lungs.\textsuperscript{10,35,36} The prevailing theories are that over-production of MUC5B impairs mucociliary clearance, contributes to lung injury or epithelial cell stress and/or disrupts reparative mechanisms in the distal lung, leading to lung fibrosis.\textsuperscript{37}

The MUC5B rs35705950 SNP was not associated with radiographic or histopathologic features common to HP such as air trapping on HRCT, or histopathologic quantification of airway centered inflammation or fibrosis, granulomas, or lymphocytic interstitial inflammation. In contrast, the SNP was associated with changes characteristic of IPF including HRCT evidence of moderate-severe fibrosis and traction bronchiectasis. We also observed a borderline association of the MUC5B minor allele with microscopic honeycombing in cHP, which is interesting considering the localization of MUC5B in honeycomb cysts from IPF lungs.\textsuperscript{35,36} This observation, though not statistically significant after multiple testing correction, could suggest that MUC5B confers susceptibility to lung remodeling across different clinical forms of fibrotic ILD. In contrast to IPF, where the MUC5B minor allele is associated with better survival,\textsuperscript{9} a statistical trend toward poorer survival was found among patients with cHP. We hypothesize that the mechanisms by which MUC5B promotes pulmonary fibrosis in its idiopathic forms are similarly active in patients who develop cHP, and that the combination of injury
from inflammation in HP in the context of excess MUC5B production increases the risk of developing lung remodeling and fibrosis in HP.

Telomere dysfunction has been implicated in the pathogenesis of both sporadic and familial forms of IPF. Mutations in several telomere-associated genes have been identified in sporadic and familial IPF, and telomeres are short in the alveolar epithelial cells (AEC) of these patients. Telomere dysfunction isolated to AECs causes lung remodeling and fibrosis in mouse models. In addition to typical IPF presentations, many families contain members with a clinical diagnosis of chronic HP, suggesting shared pathobiology of telomere dysfunction in IPF and cHP. Consistent with this possibility, shorter PBL telomeres were associated with fibrosis and poor survival in cHP. Similar to the MUC5B SNP, short PBL telomere length was associated with radiographic and histopathologic changes characteristic of IPF including radiographic severity of fibrosis, traction bronchiectasis and honeycombing, and histopathologic evidence of microscopic honeycombing, subpleural distribution, and fibroblast foci. Similar to IPF, shorter telomere length was a strong predictor of poor survival in cHP. Given these findings, we speculate that persistent inflammation in HP could be a driver of accelerated AEC turnover, contributing to telomere shortening in these cells, and, in susceptible individuals (i.e. those born with shorter telomeres or with an impaired ability to maintain telomeres), eventually leads to elements of lung remodeling and fibrosis similar to those seen in IPF. This hypothesis requires experimental examination.
No association was found between the TOLLIP rs5743890 SNP minor allele and predisposition, fibrosis, or survival in patients with cHP. In contrast to prior studies, there was no association between TOLLIP rs5743890 in IPF compared to controls. However, the smaller sample size of this study may have limited the statistical power for this comparison.

Our findings are associations, and do not prove a causative role for MUC5B or telomere dysfunction in cHP. There may be alternative explanations for the findings. Recent studies have highlighted the problem of misclassification of HP and IPF diagnoses. Therefore, it is possible that some patients diagnosed with cHP actually had IPF, leading to an enrichment in the MUC5B minor allele and shorter telomere lengths in the cHP study cohorts. However, this is unlikely considering diagnoses were established by multidisciplinary consensus, most were supported by histopathology, had identified exposures, had HRCT features inconsistent with a UIP pattern, and were consistent across two ILD centers. Even if true, this explanation would call into question the validity of clinical classification of ILD’s and would argue for the importance of developing molecular classification systems, at least in regards to risk stratification. It should also be noted that we observed numeric differences in MUC5B and TOLLIP MAFs between our two cHP cohorts. While we believe these differences are related to random variation and differences in distribution of fibrotic severity between the two cohorts, we cannot exclude the
possibility of a contribution from different MAFs in the local non-Hispanic white populations.

In regards to the telomere length findings, shorter telomeres in PBLs in cHP could alternatively be a marker of telomere attrition in PBLs resulting from persistent inflammation rather than a surrogate marker of AEC telomere length. The link between inflammation and telomere attrition is complex. However, since telomere length was measured in PBLs originating from bone marrow, and it is unlikely that the bone marrow was affected by localized pulmonary inflammation, we believe it is more likely that PBL telomere length represents a surrogate marker for the “starting point” of AEC telomere length. Future studies should examine this possibility as well as the telomere lengths in various cell types of HP lungs to establish a causative role. Finally, because our genotype frequency analysis was restricted to the non-Hispanic white population, our results may not be generalizable to other racial/ethnic groups.

In conclusion, the MUC5B promoter polymorphism and shorter peripheral blood leukocyte telomere length are associated with fibrosis and reduced survival in cHP. These findings may have a role in risk stratification of patients with cHP, and provide insights into its pathogenesis. Future studies are needed to elucidate the role of MUC5B and telomere biology in the pathogenesis of cHP.
Authors’ Contributions: BJL, CAN, CKG, and PJW conceived of the study and study design. BJL, CAN, IA, BME, TSH, JAG, KDJ, KB, and JT collected data for the study. BJL, CAN, and EV performed statistical analyses. All authors contributed to data interpretation. BJL and CAN wrote the original draft of the manuscript. All authors contributed to manuscript review, editing, and final approval for submission.

Conflict of Interest Statements:
None of the authors report conflicts of interest relevant to this study. Dr. Ley reports a speaking fee from Genentech. Dr. Wolters reports grants from Medimmune, personal fees from Roche, and grants from Genentech. Dr. Garcia reports personal fees from Pliant Therapeutics involving activities outside the submitted work.

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REFERENCES

1. Selman M, Pardo A, King TE, Jr. Hypersensitivity pneumonitis: insights in diagnosis and pathobiology. *American journal of respiratory and critical care medicine*. 2012;186(4):314-324.

2. Mooney JJ, Elicker BM, Urbania TH, et al. Radiographic fibrosis score predicts survival in hypersensitivity pneumonitis. *Chest*. 2013;144(2):586-592.

3. Kern RM, Singer JP, Koth L, et al. Lung transplantation for hypersensitivity pneumonitis. *Chest*. 2015;147(6):1558-1565.

4. Christensen LT, Schmidt CD, Robbins L. Pigeon breeders’ disease—a prevalence study and review. *Clin Allergy*. 1975;5(4):417-430.

5. Dalphin JC, Debieuvre D, Pernet D, et al. Prevalence and risk factors for chronic bronchitis and farmer’s lung in French dairy farmers. *Br J Ind Med.* 1993;50(10):941-944.

6. Hendrick DJ, Faux JA, Marshall R. Budgerigar-fancier's lung: the commonest variety of allergic alveolitis in Britain. *Br Med J*. 1978;2(6130):81-84.

7. Lalancette M, Carrier G, Laviolette M, et al. Farmer’s lung. Long-term outcome and lack of predictive value of bronchoalveolar lavage fibrosing factors. *The American review of respiratory disease*. 1993;148(1):216-221.

8. Noth I, Zhang Y, Ma SF, et al. Genetic variants associated with idiopathic pulmonary fibrosis susceptibility and mortality: a genome-wide association study. *The Lancet respiratory medicine*. 2013;1(4):309-317.

9. Peljto AL, Zhang Y, Fingerlin TE, et al. Association between the MUC5B promoter polymorphism and survival in patients with idiopathic pulmonary fibrosis. *JAMA : the journal of the American Medical Association*. 2013;309(21):2232-2239.

10. Seibold MA, Wise AL, Speer MC, et al. A common MUC5B promoter polymorphism and pulmonary fibrosis. *The New England journal of medicine*. 2011;364(16):1503-1512.

11. Fingerlin TE, Murphy E, Zhang W, et al. Genome-wide association study identifies multiple susceptibility loci for pulmonary fibrosis. *Nature genetics*. 2013;45(6):613-620.

12. Zhang Y, Noth I, Garcia JG, Kaminski N. A variant in the promoter of MUC5B and idiopathic pulmonary fibrosis. *The New England journal of medicine*. 2011;364(16):1576-1577.

13. Alder JK, Stanley SE, Wagner CL, Hamilton M, Hanumanthu VS, Armanios M. Exome sequencing identifies mutant TINF2 in a family with pulmonary fibrosis. *Chest*. 2015;147(5):1361-1368.

14. Armanios MY, Chen JJ, Cogan JD, et al. Telomerase mutations in families with idiopathic pulmonary fibrosis. *The New England journal of medicine*. 2007;356(13):1317-1326.

15. Diaz de Leon A, Cronkhite JT, Katzenstein AL, et al. Telomere lengths, pulmonary fibrosis and telomerase (TERT) mutations. *PloS one*. 2010;5(5):e10680.
16. Kannengiesser C, Borie R, Menard C, et al. Heterozygous RTEL1 mutations are associated with familial pulmonary fibrosis. *The European respiratory journal*. 2015;46(2):474-485.

17. Kropski JA, Mitchell DB, Markin C, et al. A novel dyskerin (DKC1) mutation is associated with familial interstitial pneumonia. *Chest*. 2014;146(1):e1-7.

18. Tsakiri KD, Cronkhite JT, Kuan PJ, et al. Adult-onset pulmonary fibrosis caused by mutations in telomerase. *Proceedings of the National Academy of Sciences of the United States of America*. 2007;104(18):7552-7557.

19. Stuart BD, Choi J, Zaidi S, et al. Exome sequencing links mutations in PARN and RTEL1 with familial pulmonary fibrosis and telomere shortening. *Nature genetics*. 2015;47(5):512-517.

20. Stuart BD, Lee JS, Kozlitina J, et al. Effect of telomere length on survival in patients with idiopathic pulmonary fibrosis: an observational cohort study with independent validation. *The lancet Respiratory medicine*. 2014.

21. Newton CA, Batra K, Torrealba J, et al. Telomere-related lung fibrosis is diagnostically heterogeneous but uniformly progressive. *The European respiratory journal*. 2016.

22. Raghu G, Collard HR, Egan JJ, et al. An official ATS/ERS/JRS/ALAT statement: idiopathic pulmonary fibrosis: evidence-based guidelines for diagnosis and management. *American journal of respiratory and critical care medicine*. 2011;183(6):788-824.

23. Travis WD, Costabel U, Hansell DM, et al. An official american thoracic society/european respiratory society statement: update of the international multidisciplinary classification of the idiopathic interstitial pneumonias. *American journal of respiratory and critical care medicine*. 2013;188(6):733-748.

24. Silva CI, Churg A, Muller NL. Hypersensitivity pneumonitis: spectrum of high-resolution CT and pathologic findings. *AJR Am J Roentgenol*. 2007;188(2):334-344.

25. Silva CI, Muller NL, Lynch DA, et al. Chronic hypersensitivity pneumonitis: differentiation from idiopathic pulmonary fibrosis and nonspecific interstitial pneumonia by using thin-section CT. *Radiology*. 2008;246(1):288-297.

26. Akashi T, Takemura T, Ando N, et al. Histopathologic analysis of sixteen autopsy cases of chronic hypersensitivity pneumonitis and comparison with idiopathic pulmonary fibrosis/usual interstitial pneumonia. *American journal of clinical pathology*. 2009;131(3):405-415.

27. Churg A, Muller NL, Flint J, Wright JL. Chronic hypersensitivity pneumonitis. *Am J Surg Pathol*. 2006;30(2):201-208.

28. Churg A, Sin DD, Everett D, Brown K, Cool C. Pathologic patterns and survival in chronic hypersensitivity pneumonitis. *Am J Surg Pathol*. 2009;33(12):1765-1770.

29. Genomes Project C, Auton A, Brooks LD, et al. A global reference for human genetic variation. *Nature*. 2015;526(7571):68-74.

30. Cronkhite JT, Xing C, Raghu G, et al. Telomere shortening in familial and sporadic pulmonary fibrosis. *American journal of respiratory and critical care medicine*. 2008;178(7):729-737.
31. Best AC, Meng J, Lynch AM, et al. Idiopathic pulmonary fibrosis: physiologic tests, quantitative CT indexes, and CT visual scores as predictors of mortality. *Radiology.* 2008;246(3):935-940.

32. Hansell DM, Bankier AA, MacMahon H, McLoud TC, Muller NL, Remy J. Fleischner Society: glossary of terms for thoracic imaging. *Radiology.* 2008;246(3):697-722.

33. Hochberg Y. A sharper Bonferroni procedure for multiple tests of significance. *Biometrika.* 1988;75(4):800-802.

34. Borie R, Crestani B, Dieude P, et al. The MUC5B variant is associated with idiopathic pulmonary fibrosis but not with systemic sclerosis interstitial lung disease in the European Caucasian population. *PloS one.* 2013;8(8):e70621.

35. Conti C, Montero-Fernandez A, Borg E, et al. Mucins MUC5B and MUC5AC in Distal Airways and Honeycomb Spaces: Comparison among Idiopathic Pulmonary Fibrosis/Usual Interstitial Pneumonia, Fibrotic Nonspecific Interstitial Pneumonitis, and Control Lungs. *American journal of respiratory and critical care medicine.* 2016;193(4):462-464.

36. Nakano Y, Yang IV, Walts AD, et al. MUC5B Promoter Variant rs35705950 Affects MUC5B Expression in the Distal Airways in Idiopathic Pulmonary Fibrosis. *American journal of respiratory and critical care medicine.* 2016;193(4):464-466.

37. Evans CM, Fingerlin TE, Schwarz MI, et al. Idiopathic Pulmonary Fibrosis: A Genetic Disease That Involves Mucociliary Dysfunction of the Peripheral Airways. *Physiol Rev.* 2016;96(4):1567-1591.

38. Cogan JD, Kropski JA, Zhao M, et al. Rare variants in RTEL1 are associated with familial interstitial pneumonia. *American journal of respiratory and critical care medicine.* 2015;191(6):646-655.

39. Alder JK, Chen JJ, Lancaster L, et al. Short telomeres are a risk factor for idiopathic pulmonary fibrosis. *Proceedings of the National Academy of Sciences of the United States of America.* 2008;105(35):13051-13056.

40. Kropski JA, Pritchett JM, Zoz DF, et al. Extensive phenotyping of individuals at risk for familial interstitial pneumonia reveals clues to the pathogenesis of interstitial lung disease. *American journal of respiratory and critical care medicine.* 2015;191(4):417-426.

41. Naikawadi RP, Disayabutr S, Mallavia B, et al. Telomere dysfunction in alveolar epithelial cells causes lung remodeling and fibrosis. *JCI Insight.* 2016;1(14):e86704.

42. Povedano JM, Martinez P, Flores JM, Mulero F, Blasco MA. Mice with Pulmonary Fibrosis Driven by Telomere Dysfunction. *Cell Rep.* 2015;12(2):286-299.

43. Okamoto T, Miyazaki Y, Tomita M, Tamaoka M, Inase N. A familial history of pulmonary fibrosis in patients with chronic hypersensitivity pneumonitis. *Respiration; international review of thoracic diseases.* 2013;85(5):384-390.

44. Morell F, Villar A, Montero MA, et al. Chronic hypersensitivity pneumonitis in patients diagnosed with idiopathic pulmonary fibrosis: a prospective case-cohort study. *The lancet Respiratory medicine.* 2013;1(9):685-694.
45. Walsh SL, Wells AU, Desai SR, et al. Multicentre evaluation of multidisciplinary team meeting agreement on diagnosis in diffuse parenchymal lung disease: a case-cohort study. *The Lancet Respiratory Medicine*. 2016;4(7):557-565.

46. Wong JY, De Vivo I, Lin X, Fang SC, Christiani DC. The relationship between inflammatory biomarkers and telomere length in an occupational prospective cohort study. *PloS one*. 2014;9(1):e87348.
Table 1. Characteristics of patients with chronic hypersensitivity pneumonitis

| Characteristic                       | UCSF (n=145) | UTSW (n=72) | p-value |
|--------------------------------------|--------------|-------------|---------|
| Age, mean (SD)                       | 63.3 (11.2)  | 60.6 (9.8)  | 0.081   |
| Female Sex, n (%)                    | 89 (61.4)    | 34 (47.2)   | 0.048   |
| Race, n (%)                          |              |             |         |
| White, non-Hispanic/Latino           | 123 (84.8)   | 65 (90.3)   | 0.27    |
| Hispanic or Latino                   | 13 (9.0)     | 4 (5.6)     |         |
| Black                                | 2 (1.4)      | 2 (2.8)     |         |
| Asian                                | 4 (2.8)      | 1 (1.4)     |         |
| Other/unknown                        | 3 (2.1)      | 0           |         |
| Ever-smoker, n (%)                   | 75 (51.7)    | 36 (50.0)   | 0.81    |
| Family history, n (%)                | 15 (10.3)    | 17 (23.6)   | 0.009   |
| Lung biopsy, n (%)                   |              |             |         |
| Surgical lung biopsy                 | 114 (78.6)   | 51 (70.8)   | 0.21    |
| Transbronchial lung biopsy           | 94 (64.8)    | 36 (50.0)   | 0.036   |
| Pulmonary Function Tests, mean (SD)  |              |             |         |
| FVC, % predicted                     | 67 (17)      | 66 (19)     | 0.82    |
| DLCO, % predicted                    | 48 (17)      | 50 (18)     | 0.57    |
| Antigen identified, n (%)            |              |             |         |
| Avian                                | 100 (69.0)   | 54 (75.0)   | 0.36    |
| Bird                                 | 73 (50.3)    | 31 (43.1)   | 0.31    |
| Down                                 | 41 (28.3)    | 21 (29.2)   |         |
| Mold                                 | 21 (14.5)    | 10 (13.9)   |         |
| Other                                | 2 (1.4)      | 7 (9.7)     |         |
| Unknown                              | 21 (14.5)    | 11 (15.3)   | 0.88    |
| Multiple                             | 45 (31.0)    | 18 (25)     |         |
| Telomere length measured, n (%)      | 129 (89.0)   | 72 (100)    | <0.001  |
| mean log T/S (SD)                    | 1.53 (0.37)  | 1.30 (0.33) |         |
| MUC5B genotype, n (%)                |              |             |         |
| GG                                   | 85 (58.6)    | 34 (47.2)   | 0.141   |
| GT                                   | 55 (37.9)    | 32 (44.4)   |         |
| TT                                   | 5 (3.4)      | 6 (8.3)     |         |
| TOLLIP genotype, n (%)               |              |             |         |
| AA                                   | 109 (75.2)   | 48 (66.7)   | 0.337   |
| AG                                   | 33 (22.8)    | 23 (31.9)   |         |
| GG                                   | 3 (2.1)      | 1 (1.4)     |         |
| HRCT available for scoring, n (%)    | 119 (82)     | 70 (97)     |         |
| Fibrosis extent, semi-quantitative, n (%) |          |             | <0.001  |
| None                                 | 25 (21)      | 4 (6)       |         |
| Mild (< 10%)                         | 39 (33)      | 21 (30)     |         |
| Moderate (10-50%)                    | 49 (41)      | 27 (39)     |         |
| Severe (> 50%)                       | 6 (5)        | 18 (26)     |         |
| Honeycombing, n (%)                  | 18 (15)      | 24 (34)     | 0.002   |
| UIP Pattern, n (%)                  | Definite | Possible | Inconsistent | p-value |
|------------------------------------|----------|----------|-------------|---------|
| Definite                           | 4 (3.4)  | 5 (7)    | 105 (88.2)  | 0.129   |
| Possible                           | 10 (8.4) | 54 (16)  | 11 (77)     |         |
| Inconsistent                       |          |          |             |         |

| Inconsistent Features, n (%)       |          |          |             |         |
|------------------------------------|----------|----------|-------------|---------|
| Upper-mid lung predominance        | 44 (37)  | 26 (37)  | 63 (53)     | 0.98    |
| Peribronchovascular distribution   | 63 (53)  | 36 (51)  | 52 (44)     | 0.84    |
| Diffuse ground glass               | 52 (44)  | 34 (49)  | 12 (10)     | 0.52    |
| Micronodules                       | 12 (10)  | 7 (10)   | 69 (58)     | 0.99    |
| Mosaic perfusion/air-trapping      | 69 (58)  | 40 (57)  |             | 0.91    |

**Abbreviations:** UCSF = University of California San Francisco; UTSW = University of Texas Southwestern; FVC = forced vital capacity; DLCO = diffusing capacity of the lung for carbon monoxide; HRCT = high-resolution computed tomography; UIP = usual interstitial pneumonia; log T/S = Telomere length expressed as the natural logarithm of the telomere to single gene copy ratio