Commentary: Oil degradation and biosurfactant production by the deep sea bacterium *Dietzia maris* As-13-3

Pattanathu K. S. M. Rahman¹* and Kamaljeet K. Sekhon Randhawa²

¹ Technology Futures Institute, School of Science and Engineering, Teesside University, Middlesbrough, UK, ² Section for Sustainable Biotechnology, Aalborg University, Copenhagen, Denmark

Keywords: genus *Dietzia*, di-rhamnolipids, biosurfactant, pathogenesis, alkanes, biosynthetic pathway

A commentary on

Oil degradation and biosurfactant production by the deep sea bacterium *Dietzia maris* As-13-3 by Wang, W., Cai, B., and Shao, Z. (2014). Front. Microbiol. 5:711. doi: 10.3389/fmicb.2014.00711

Surfactants or surface active agents find broad industrial applications as additives—in household detergents, cleaning products, toothpastes, paints, coatings, leather, paper, cosmetics, pharmaceuticals, food products, and agriculture. Irrespective of their interesting applications, all surfactants end up their journey in sewers or drains—from where they directly or indirectly affect the flora, fauna, and water bodies causing environmental concerns.

Biosurfactants—plant based or produced by microbes are considered the holy grail of "natural" products. The versatility of biosurfactants is due to their amphiphilic property—a sought-after attribute—which gives them a double affinity for both hydrophilic and lipophilic compounds. Biosurfactants are therefore "detergents" which help lower the surface tension of a liquid or interfacial tension between two liquids. Their eco-friendly origin makes them stand tall among their synthetic or petroleum based counterparts. Regardless of the technology that has been employed to produce biosurfactants their sky-high prices are still a reason of concern for the biosurfactant research community anticipating more intensive research. Apart from the innovative production technologies tried so far, there are a number of microbial strains which have also been harnessed in these technologies for cost-effective generation of biosurfactants.

A detailed and effective study is conducted by Wang et al. (2014) showing the complete pathway of the di-rhamnolipid synthesis process in the genus *Dietzia* isolated from extreme environments. This short account will present an annotation on the highlights of the study with some critical analysis. In this study the authors have explored the possibility of biosurfactant production by *Dietzia maris* As-13-3, which was originally isolated from deep sea hydrothermal field environment of Southwest Indian Ocean (Chen and Shao, 2009). Grown on 2% (v/v) n-hexadecane as the sole carbon source this deep sea bacterium was shown to produce di-rhamnolipid as biosurfactant. Analysing *Dietzia maris* As-13-3 for various activities, the authors claim that the strain could play significant role in oil degradation, biosurfactant production, and marine oil removal.

Rhamnolipids are the most studied biosurfactants and there are now companies entirely dedicated to their research and production such as AGAE Technologies Ltd., Jeneil Biosurfactants, and Paradigm Biomedical Inc., all based in USA (Sekhon et al., 2012). Nevertheless, there is enormous scope for investigating new and extreme environments so as to look for novel strains which can mass produce these biosurfactants using relatively cheap substrates. Microbial consortia ERCPII-2 isolated from heavy crude oil contaminated site in Iran, *Virgibacillus salarius* isolated
from soil and seawater samples from Saudi Arabia, 10 thermohalophilic isolates of genus *Bacillus* isolated from Iranian petroleum reservoirs are few examples of extremophilic microorganisms that produce biosurfactants (Darvishi et al., 2011; Zargari et al., 2014; Elazzazy et al., 2015). Although alkanes are considerably not very cheap but interestingly *Dietzia maris* As-13-3 could utilize three different alkanes such as n-tetradecane, n-hexadecane, and pristane, for biosurfactant production. The strain could utilize short-chain and middle-chain n-alkanes from C₉ to C₂₀, and vigorous degradation was reported for C₁₄ to C₁₈ and pristane (Wang et al., 2014). It is also quite fascinating how the authors summarized that n-hexadecane serves as the main raw material for production of biosurfactant by this strain and revealed a complete pathway for the biodegradation of alkanes by terminal oxidation—mentioning the genes including alkB, cyp153, algC, rmlA, rmlB, rmlC, rmlD, rhlA, rhlB, and rhlC which are categorically induced by alkanes for biosurfactant production.

The rhamnolipids produced by *Dietzia maris* As-13-3 at a CMC of 120 mg L⁻¹ lowered the surface tension of water from 74 ± 0.2 to 38 ± 0.2 mN m⁻¹ (Wang et al., 2014), which was not as good as 29 mN m⁻¹ with a CMC value in range of 5-60 mg L⁻¹ by *Pseudomonas aeruginosa* (Van Dyke et al., 1993) but better than 42 mN m⁻¹ at a CMC value of 225 mg L⁻¹ by *Burkholderia thailandensis* (Dubeau et al., 2009). The comparison with surface tension values of rhamnolipids produced by other significant strains gives a very clear picture of the effectiveness and efficacy of these biosurfactants—efficient biosurfactants have a low CMC value. Future studies in the field would, however, have benefitted from a comparative analysis of the total yield of rhamnolipids produced per liter of the media by this deep sea bacterium and throwing light on the prospective commercialization of these rhamnolipids.

Genus *Dietzia* is little unknown but contains clinically significant organisms. This emerging actinomycetes has been misidentified as *Rhodococcus equi* to which it has remarkable similarities—for instance other species of this genus including *Dietzia maris* showed 98.1% similarity to *Rhodococcus* sp. (Koerner et al., 2009). *Dietzia maris* taxon, initially proposed for two strains isolated from flatfish halibut, was classified as "F.maris" (Harrison, 1929). Surprisingly, there are three reports of *Dietzia maris* causing infections in humans (Koerner et al., 2009) and has been clearly implicated as potential opportunistic human pathogen (Bemer-Melchior et al., 1999; Pidoux et al., 2001; Reyes et al., 2006). Having said that, to date there has been no attempts to deeply investigate the pathogenicity factors of *Dietzia maris*—which apparently colonizes human skin as it excretes n-alkanes (Koerner et al., 2009). In view of these studies it will be too early and hasty to say and claim that *Dietzia maris* As-13-3 is a “non-pathogenic” strain and the rhamnolipids are suitable for commercial production and are industrially safe specially in the food and pharmaceutical sector—without thoroughly inspecting the virulence factors of this deep sea bacterium.

This study has certainly outdone several other studies in terms of the extensive new insights into the genetics and physiology of the genus Dietzia—however a few more significant tests could have really elaborated how efficient and successful this deep sea bacterium *Dietzia maris* As-13-3 is in the production and commercialization of di-rhamnolipids. Such studies undoubtedly bring into forefront the less explored genus, thereby increasing our knowledge in terms of the amount of work that still can be undertaken for search of efficacious strains from extreme and unexploited environments.

**AUTHOR CONTRIBUTIONS**

PR initiated the research topic and co-ordinated the entire editorial process. There are 11 manuscripts accepted for publication in this research topic contributed by 55 authors from UK, Denmark, Greece, Germany, South Africa, India, Brazil, Bahrain, Portugal, and China. He has initiated peer review process by inviting experts from Germany, Spain, USA, Trinidad and Tobago, India, Denmark, China, Bahrain, Malaysia, Japan, and UK. The reviewers co-operation and timely responses to complete the research topic is highly commendable. One of the articles, submitted for the research topic by Wang et al. (2014), Oil degradation and biosurfactant production by the deep sea bacterium *Dietzia maris* As-13-3 is selected for General commentary. KS has contributed as Co-author in this Editorial commentary. She is an expert on this topic and has been working specifically in this field, she has her adept inputs in conceptualizing the commentary, analysing the data, interpreting and drafting the commentary. KS has given her final approval of the version to be published and agrees to be held accountable for all aspects of the work related to the accuracy and integrity.

**REFERENCES**

Bemer-Melchior, P., Haloun, A., Riegel, P., and Drugeon, H. B. (1999). Bacteraemia due to *Dietzia maris* in an immunocompromised patient. *Clin Infect Dis.* 29, 1338–1340. doi: 10.1086/313490

Chen, S., and Shao, Z. (2009). Isolation and diversity analysis of arsenic-resistant bacteria in communities enriched from deep-sea sediments of the Southwest Indian Ocean Ridge. *Extremophiles* 13, 39–48. doi: 10.1007/s00792-008-0195-1

Darvishi, P., Ayatollahi, S., and Mowla, D. (2011). Biosurfactant production under extreme environmental conditions by an efficient microbial consortium, ERCPPI-2. *Colloids Surf. B Biointerfaces* 84, 292–300. doi: 10.1016/j.colsurfb.2011.01.011

Dubeau, D., Deziel, E., Woods, D. E., and Lepine, F. (2009). *Burkholderia thailandensis* harbors two identical rhl gene clusters responsible for the biosynthesis of rhamnolipids. *BMC Microbiol.* 9:263. doi: 10.1186/1471-2180-9-263

Elazzazy, A. M., Abdelmonem, T. S., and Almaghribi, O. A. (2015). Isolation and characterization of biosurfactant production under extreme environmental conditions by alkali-halo-thermophilic bacteria from Saudi Arabia. *Saudi J. Biol. Sci.* 22, 466–475. doi: 10.1016/j.sjbs.2014.11.018

Harrison, F. C. (1929). The discoloration of halibut. *Can. J. Res.* 1, 214–239. doi: 10.1139/cjr29-013

Koerner, R. J., Goodfellow, M., and Jones, A. L. (2009). The genus Dietzia: a new home for some known and emerging opportunistic pathogens. *FEMS Immunol. Med. Microbiol.* 55, 296–305. doi: 10.1111/j.1574-695X.2008.00513.x
Pidoux, O., Argenson, J. N., Jacomo, V., and Drancourt, M. (2001). Molecular identification of a *Dietzia maris* hip prosthesis infection isolate. *J. Clin. Microbiol*. 39, 2634–2636. doi: 10.1128/JCM.39.7.2634-2636.2001

Reyes, G., Navarro, J. L., Gamallo, C., and de las Cuevas, M. C. (2006). Type A aortic dissection associated with *Dietzia maris*. *Interact. Cardiovasc. Thorac. Surg*. 5, 666–668. doi: 10.1510/icvts.2006.135640

Sekhon, K. K., Khanna, S., and Cameotra, S. S. (2012). Biosurfactant production and potential correlation with esterase activity. *J. Pet. Environ. Biotechnol*. 3:133. doi: 10.4172/2157-74631000133

Van Dyke, M. I., Couture, P., Brauer, M., Lee, H., and Trevors, J. T. (1993). *Pseudomonas aeruginosa* UG2 rhamnolipid biosurfactants: structural characterization and their use in removing hydrophobic compounds from soil. *Can. J. Microbiol*. 39, 1071–1078. doi: 10.1139/m93-162

Wang, W., Cai, B., and Shao, Z. (2014). Oil degradation and biosurfactant production by the deep sea bacterium *Dietzia maris* As-13-3. *Front. Microbiol*. 5:711. doi: 10.3389/fmicb.2014.00711

Zargari, S., Ramezani, A., Ostvar, S., Rezaei, R., Niazi, A., and Ayatollahi, S. (2014). Isolation and characterization of gram-positive biosurfactant producing halothermophilic *Bacilli* from Iranian petroleum reservoirs. *Jundishapur J. Microbiol*. 7:e10981. doi: 10.5812/jjm.10981

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.