Nonhuman Primate Models for SARS

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Animal Models for SARS

Development of animal models for SARS-CoV infection of humans is of utmost importance to elucidate the pathogenesis of SARS and to develop intervention strategies against the infection. A wide range of animal species is susceptible to experimental infection with SARS-CoV, including rodents (mice and hamsters), carnivores (ferrets and cats), and nonhuman primates (cynomolgus and rhesus macaques, common marmosets, and African green monkeys) [3–11]. Adult mice infected with varying doses of SARS-CoV in the respiratory tract show no clinical signs of disease, although the virus replicates in respiratory tissues, peaking early after infection, with viral titres in the lungs reaching relatively high levels. The infection is accompanied by only mild inflammatory changes of the respiratory tract. On the other hand, aged mice, as well as hamsters and ferrets, do show signs of clinical disease (weight loss and ruffled fur), albeit, in most cases, in the absence of the typical lung lesions seen in humans with SARS [4–6].

In contrast, SARS-CoV inoculation in the respiratory tract of cynomolgus macaques causes infection of bronchial epithelial cells, type-1 and type-2 pneumocytes at one to four days postinfection, followed by extensive type-2 pneumocyte hyperplasia in the lungs at four to six days postinfection [3,11]. Moreover, multiple foci of acute DAD are observed, characterised by flooding of alveoli with protein-rich oedema fluid mixed with variable numbers of neutrophils and rare syncytia (Figure 1), and by extensive loss of epithelium from alveolar and bronchiolar walls. These lesions are quite similar to those observed in humans in the acute stages of SARS. Although clinical signs (respiratory distress and general malaise) were observed, they were not further studied in the initial experiments.

Studies from different laboratories confirmed that nonhuman primates could be infected experimentally with SARS-CoV, although the severity of

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Abbreviations: ARDS, acute respiratory distress syndrome; DAD, diffuse alveolar damage; SARS, severe acute respiratory syndrome; SARS-CoV, SARS coronavirus

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Figure 1. Histologic Lesions in Lungs from Cynomolgus Macaques Experimentally Infected with SARS-CoV

Syncytia (indicated by the arrowhead) in the lumen of a bronchiole (A) and expression of SARS-CoV antigen by a syncytium in the lumen of an alveolar duct (B).

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The induced lung pathology varied and clinical signs were generally not reported. So far, factors underlying these
differences have not been delineated, but virus strain, age, microbiological state, genetic background, and origin of the animals could have played a role.

In this issue of *PLoS Medicine*, James Lawler, Jason Paragas, and colleagues provide further support for the use of the cynomolgus macaque model to study the pathogenesis of SARS and to test intervention strategies [12]. They show that infection of these macaques with a molecularly cloned SARS-CoV resulted in a mild-to-moderate symptomatic illness, indistinguishable from the illness induced with wild-type SARS-CoV. Chest radiographs from several infected animals showed unifocal or multifocal pneumonia that peaked between eight to ten days postinfection. The authors point out that the results are reminiscent of the milder syndrome observed in younger humans infected with SARS-CoV.

Molecular modification of the cloned virus may now further allow deciphering of the role of different viral gene products in the pathogenesis of SARS. An important result of this present study is the observation that one group of animals inoculated via the nasal and ocular route exhibited significant faecal shedding of SARS-CoV. Thus far, efforts to evaluate the efficacy of SARS-CoV vaccines have largely aimed at protecting the respiratory tract.

**Implications for Future SARS Research**

The persistent threat of a possible new introduction of SARS-CoV or of a related virus in the human population necessitates further refinement of animal models to elucidate the pathogenesis of, and intervention strategies against, SARS. Although none of the current animal models have fully reproduced all features of SARS, the most important aspects of SARS are observed in experimentally infected nonhuman primates. Such models have already demonstrated the protective effect of pre- and postexposure use of pegylated human interferon-α, of candidate SARS-CoV vaccines, and of convalescent sera from SARS patients (unpublished results; [10,11]). In the absence of ongoing human SARS-CoV infections, candidate SARS vaccines may have to be approved on the basis of animal data alone, preferably in two different species (the “two animal rule”). But in order for such approval, the pathophysiological mechanisms underlying SARS pathogenesis and the preventive efficacy of such vaccines need to be well understood. The present study further supports the use of nonhuman primate SARS models for pathogenesis studies and for the development of intervention strategies. Infection of cynomolgus macaques with recombinant SARS-CoV containing defined modifications may allow further elucidation of SARS.

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**Glossary**

**Type-1 Pneumocytes:** These cells are responsible for alveolar gas exchange. They cannot replicate and are susceptible to a large number of toxic insults.

**Type-2 Pneumocytes:** These cells are responsible for producing and secreting surfactant. They can replicate in the alveoli to replace damaged type-1 pneumocytes.

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**References**

1. Poon LL, Guan Y, Nicholls JM, Yuen KY, Peiris JS (2004) The aetiology, origins, and diagnosis of severe acute respiratory syndrome. *Lancet Infect Dis* 4: 663–671.
2. Fouchier RA, Kuiken T, Schutten M, van Amerongen G, van Doornum GJ, et al. (2003) Koch’s postulates fulfilled for SARS virus. *Nature* 425: 240.
3. Kuiken T, Fouchier RA, Schutten M, Rimmelzwaan GF, van Amerongen G, et al. (2003) Newly discovered coronavirus as the primary cause of severe acute respiratory syndrome. *Lancet* 362: 263–270.
4. Roberts A, Vogel L, Guanier J, Hayes N, Murphy B, et al. (2005) Severe acute respiratory syndrome coronavirus infection of golden Syrian hamsters. *J Virol* 79: 503–511.
5. Roberts A, Paddock C, Vogel L, Butler E, Zaki S, et al. (2005) Aged BALB/c mice as a model for increased severity of severe acute respiratory syndrome in elderly humans. *J Virol* 79: 5835–5838.
6. Martina BE, Haagmans BL, Kuiken T, Fouchier RA, Rimmelzwaan GF, et al. (2003) SARS virus infection of cats and ferrets. *Nature* 425: 915.
7. McAuliffe J, Vogel L, Roberts A, Falde G, Fischer S, et al. (2004) Replication of SARS coronavirus administered into the respiratory tract of African Green, rhesus and cynomolgus monkeys. *Virology* 350: 8–15.
8. Rowe T, Gao G, Hogan RJ, Crystal RG, Yoss TG, et al. (2004) Macaque model for severe acute respiratory syndrome. *J Virol* 78: 11401–11404.
9. Greenough TC, Carville A, Codere J, Somasundaran M, Sullivan JL, et al. (2005) Pneumonitis and multi-organ system disease in common marmosets (*Callithrix jacchus*) infected with the severe acute respiratory syndrome-associated coronavirus. *Am J Pathol* 167: 455–465.
10. Bukreyev A, Lamirande EW, Bucholz UJ, Vogel LN, Elkins WR, et al. (2004) Mucosal immunisation of African green monkeys (*Cercopithecus aethiops*) with an attenuated parainfluenza virus expressing the SARS coronavirus spike protein for the prevention of SARS. *Lancet* 363: 2122–2127.
11. Haagmans BL, Kuiken T, Martina BE, Fouchier RA, Rimmelzwaan GF, et al. (2004) Pegylated interferon-alpha protects type I pneumocytes against SARS coronavirus infection in macaques. *Nat Med* 10: 290–293. E-pub 22 February 2004.
12. Lawler JV, Endy TP, Hensley LE, Garrison A, Fritz EA, et al. (2006) Cynomolgus macaque as an animal model for severe acute respiratory syndrome. *PLoS Med* 3: e194. DOI: 10.1371/journal.pmed.0030149