Laminin γ2 fragments are increased in the circulation of patients with early phase acute lung injury

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Abstract

Objective: Laminin-5, a cell adhesive molecule expressed solely by epithelium, is known to enhance epithelial cell migration and repair of injured epithelium, after its essential component γ2-chain is processed proteolytically. Our previous study revealed circulating levels of amino-terminal fragment of laminin γ2-chain (G2F) reflect epithelial tumor invasiveness in carcinoma patients, but its physiological role in alveolar epithelial injury remains unknown. Design: Sampling of epithelial lining fluids or pulmonary edema fluids from patients with acute lung injury (ALI) or related diseases was performed. Plasma samples were obtained from them at the time of disease onset or later. G2F concentrations were determined by immunoassay constructed by ourselves. Results: We found a significantly higher amount of G2F in pulmonary edema and epithelial lining fluids of patients with ALI, as compared with those with the other respiratory diseases. Their plasma levels were also elevated significantly early at the onset of ALI (mean ± SD; 147 ± 82 ng/ml in non-surviving and 90 ± 56 in surviving patients) as compared with those in the patients with cardiogenic pulmonary edema (59 ± 36) or idiopathic pulmonary fibrosis (37 ± 17), indicating alveolar epithelium rapidly secretes laminin-5 in ALI. At 5 days after onset, non-surviving patients maintained higher plasma concentrations (152 ± 84), but in contrast, the levels in surviving patients declined (71 ± 35), suggesting secretion of laminin-5 was suppressed, associated with recovery from ALI. Conclusion: Circulating G2F may be a biomarker for alveolar laminin-5 secreted early at disease onset in ALI, potentially regulating alveolar epithelial repair.

Keywords Laminin · Circulation · Acute lung injury · Alveolar epithelial repair
Introduction

The alveolar epithelium provides a primary defensive barrier to the external environment, and thus is frequently injured by inflammatory or physical stimuli in acute respiratory distress syndrome (ARDS) or acute lung injury (ALI) [1]. Repair of damaged alveolar epithelia is an indispensable step in restoring the lung to its normal architecture [2, 3]. The repair process is initiated by the proliferation, migration and spreading of alveolar epithelial cells from the intact margins into the damaged areas [4, 5]. Proliferation and differentiation of alveolar type-II cells are also important mechanisms in the reconstitution of extracellular matrix (ECM) that can promote alveolar epithelial repair [6, 7].

Laminins (LMs) are a family of high molecular weight ECM proteins, deposited in the basement membranes, also involved in cellular adhesion, growth and differentiation [8, 9]. There are at least 15 types of LM isoforms, resulting from a variety of combinations of α, β and γ chains [9, 10]. Laminin-5 (LM5), also recently designated as laminin-332 [10], is a heterotrimer composed of α3, β3 and γ2 chains. LM5 is secreted solely by epithelial cells and plays an important role in epithelial cell migration especially during tumor invasion, metastasis or remodeling of epithelial tissue [11–14]. LM5 enhances cellular migration when it is simultaneously secreted into the ECM and activated via cleavage of its essential constituent γ2-chain by matrix metalloproteinases (MMPs) [15, 16]. This proteolysis can release the amino-terminal fragment of γ2-chain (G2F), which is not deposited in the ECM, and can be subsequently detected in the peripheral blood [17]. We have developed an immunoassay for quantification of G2F in biological fluids and demonstrated that G2F is present in normal circulation. G2F concentrations in circulation correlated with the degree of carcinoma invasion in vivo, suggesting the potential clinical value of this circulating fragment as a biomarker for epithelial invasion induced by LM5 [17].

LM5 is potentially involved in normal epithelial repair [9, 18], and also expressed during epidermal wound healing at the leading edge of the dermal–epidermal junction by keratinocytes within several hours after mechanical injury [19]. Previous studies have shown that migration of alveolar epithelial cells is necessary during the repair process of ALI, which requires several hours for initiation and several days to be completed [3, 20]. However, little is known about the cell migration factors produced or activated during injury, proliferation and differentiation of alveolar epithelium [21]. Therefore, we examined the relationship between disease status and alveolar LM5 accumulation during the exudative phase in patients with ALI, as reflected by circulating G2F levels in several biological fluids.

Materials and methods

Collection of epithelial lining fluid (ELF) and pulmonary edema fluid (PEF) specimens

ELF specimens were prepared according to the published procedure [22], as shown schematically in Fig. 1. The protocol was briefly described. Following routine pre-medication, a flexible fiberoptic bronchoscope (Olympus Co., Japan) was inserted into the lungs. The fiberoptic was wedged in a segmental bronchus of the right middle lobe. The micro-sampling probe, with the cotton probe as absorptive material, was inserted into the lungs through the bronchoscope, and the inner probe was slowly advanced into the distal airway for absorbing the ELFs. The inner probe was removed, and the tip was placed in a pre-weighed tube and weighed at −80°C. The diluted solution for measurements of biochemical constituents was prepared by adding 3 ml of saline to the tube containing the triplicate probes. The probe was then dried and weighed again to measure the ELF volume recovered, and the dilution factor was calculated.

Undiluted PEF samples were collected as previously described [1, 23]. Briefly, the PEF sample was obtained within 15 min of intubation and mechanical ventilation. A 14-French suction catheter (Becton–Dickinson, NJ) was passed through the endotracheal tube and wedged into the distal airways; then, gentle suction was applied to obtain
PEF (at least 1–2 ml), which was collected into a specimen trap, and 10–100 U of heparin was added to each sample. Then, the heparinized samples were centrifuged, and supernatants were stored at −80°C.

Subjects for analysis of G2F levels in ELFs and plasma specimens

Twenty-seven patients with ALI who were admitted between 1999 and 2001 to the Intensive Care Unit of Kyoto Prefectural University of Medicine Hospital or Keio University Hospital were enrolled into the first part of the study. ALI and ARDS were diagnosed using the American European Consensus Conference definitions [24]. Eligible patients met consensus conference oxygenation and radiographic criteria for either ALI or ARDS, and were followed until death or hospital discharge (survival). The lung injury score was calculated according to the published algorithms [25].

Plasma anticoagulated with EDTA was collected at onset (day 0) and at fixed intervals afterwards (days 1, 3, 5 and 7). ELF specimens were obtained only at day 0 as described above. Control plasma samples were collected from 15 healthy volunteers (11 males, 4 females), while control ELF samples were obtained from 13 males and 2 females who underwent bronchoscopy to identify causes of hemoptysis, or to examine small, solitary, peripheral pulmonary nodules. Normal and control samples were also obtained between 1999 and 2001 at the above hospitals.

Plasma and ELF specimens were also collected from 5 patients with cardiogenic pulmonary edema (CPE) and 11 patients with idiopathic pulmonary fibrosis (IPF) who were recruited from the Tokyo Women’s Medical University Hospital or the Keio University Hospital, respectively. Plasma specimens were separately obtained from 20 sepsis patients without ALI, recruited from Iwate Medical University Hospital, diagnosed according to published guidelines [26]. Additionally, ELF specimens were also obtained from 12 patients with chronic obstructive pulmonary disease (COPD), recruited from the Hokkaido University of Medicine Hospital. All of these samples were collected between 1999 and 2006. The institutional review board of each center in Japan approved the study protocol, and written informed consent was obtained from either the patient or each patient’s next of kin or legal representative before enrollment.

Subjects for analysis of G2F levels in PEF specimens

In the second part of the study, 21 patients with ALI/ARDS as defined above and 11 control patients with CPE were included in this study [24, 27]. In order to allow comparison of mean concentrations of G2F in ELFs and PEFs, samples of undiluted PEF were obtained only at the onset of respiratory failure. These patients were enrolled at the Moffitt Long University and San Francisco General Hospitals in San Francisco between 1990 and 2005. The institutional review board at University of California at San Francisco approved the study protocol, and written informed consent was obtained from either the patient or each patient’s next of kin or legal representative before enrollment. The lung injury score was also calculated [25].

Imunoassay

The obtained specimens were stored at −80°C until usage. The concentration of soluble G2Fs was determined as described previously [17].

Statistical analysis

Between-group statistical comparisons were performed on mean G2F concentration data using one-way analysis of variance (ANOVA), followed by a Bonferroni/Dunn post-hoc test. Time course data of plasma G2F levels were also analyzed using repeated measures ANOVA. Statistical differences for mean PEF concentrations of G2F between CPE and ALI patient groups were analyzed using non-parametric statistics (Mann–Whitney U test). Differences were regarded as statistically significant when \( P < 0.05 \). All calculations were performed using StatView 5.0 software (SAS Institute Inc., Cary, NC).

**Results**

**G2F levels in ELF samples of the ALI patients**

In the first part of the study, we enrolled 27 patients with ALI. Nineteen patients survived to yield a hospital mortality rate of 30%. Baseline clinical features of the surviving or non-surviving patients are shown in Table 1. ELF samples were collected from the 25 ALI patients (including 19 survivors and 6 non-survivors) at onset (day 0). One-way ANOVA revealed a significant difference in the ELF levels of G2Fs among patient and control groups (\( F = 6.3, P < 0.0004 \)). G2F levels in ELF from these patients were ten-fold higher (mean ± SD; 6,034 ± 6,245 ng/ml) than those from 15 control subjects (813 ± 807 ng/ml, \( P < 0.0003 \)) or compared to 12 COPD patients (350 ± 442 ng/ml, \( P < 0.0003 \)), indicating a high degree of secretion of G2F into the air spaces in the early phase of ALI (Fig. 2). The ELF levels in these 25 ALI patients were also significantly elevated compared to 5 control patients with CPE (1,237 ± 807 ng/ml,
In order to compare PEF levels of G2F to ELF levels, we examined G2F concentrations in PEFs of 11 CPE patients and 21 ALI patients. These patient characteristics were shown in Table 2. PEF levels in the ALI subjects were significantly higher (mean ± SD; 3,242 ± 6,813 ng/ml) than those in the CPE patients (897 ± 798 ng/ml, *P* < 0.05), as shown in Fig. 3.

We quantified G2F in plasma obtained from 27 patients with ALI (Table 1). One-way ANOVA demonstrated a significant difference in plasma levels of G2Fs among patients and control groups (*P* = 5.6, *P* < 0.0001). At day 0, plasma levels of G2F in 8 non-surviving ALI patients were significantly elevated (mean ± SD; 147 ± 82 ng/ml, *P* < 0.0001) compared with 15 control subjects (35 ± 47 ng/ml).

### Table 1 Physiological characteristics of the ALI patients studied in monitoring plasma G2F levels

| Clinical outcome (n of patients) | Survived (n = 19) | Non-survived (n = 8) | Total subjects (n = 27) |
|---------------------------------|------------------|---------------------|------------------------|
| Age - yr                        | 67 ± 9           | 70 ± 11             | 68 ± 10                |
| Sex - no. (%)                   |                  |                     |                        |
| Male                            | 15 (79)          | 6 (75)              | 21 (78)                |
| Female                          | 4 (21)           | 2 (25)              | 6 (22)                 |
| Days after disease onset (n of patients) | 0 (n = 17) | 1 (n = 18) | 3 (n = 18) | 5 (n = 14) | 7 (n = 10) | 0 (n = 8) | 1 (n = 7) | 3 (n = 7) | 5 (n = 6) | 7 (n = 5) |
| Tidal volume (ml)               | 546 ± 78         | 495 ± 104.9         | 459 ± 108             | 441 ± 92               | 433 ± 107 | 469 ± 104 | 555 ± 53 | 462 ± 131 | 486 ± 140 | 505 ± 150 |
| PaO2/FiO2                       | 184 ± 83         | 230 ± 86            | 243 ± 72              | 295 ± 102              | 310 ± 69  | 120 ± 47  | 218 ± 67 | 251 ± 93  | 221 ± 114 | 257 ± 122 |
| Lung injury scoreb              | 2.12 ± 0.82      | 2.03 ± 0.95         | 1.62 ± 0.73           | 1.43 ± 0.67            | 1.27 ± 0.41| 3.00 ± 0.66| 2.36 ± 0.56| 2.36 ± 0.61| 2.31 ± 0.52| 2.40 ± 0.60|

Plus–minus values are mean ± SD. Surviving or non-surviving were judged 3 weeks after disease onset. ELF collections were performed only at day 0. Tidal volumes, PaO2/FiO2 and lung injury scores were observed individually at 5 days.

* The analyses according to variables involved fewer than 19 surviving or 8 non-surviving patients, because data for some patients were missing.

* Calculated according to the published methods [25]. Higher scores indicate more severe illness.

![Fig. 2 Levels of G2Fs in ELF of subjects in the early phase of ALI.](image)
Tidal volumes, PaO2/FiO2 and lung injury scores were observed the PaO2/FiO2 143 ±
Tidal volume (ml) 677
Lung injury score a 2.50 ±
Plus–minus values are mean ± SD
Hospital mortality, no. (%) 2 (18) 15 (68)

Days of unassisted ventilation 19 ±
Sepsis, no. (%) 0 (0) 11 (50)
Current smoker, no. (%) 1 (9) 7 (32)
Caucasian, no. (%) 6 (55) 11 (50)

Sex, no. (%)  
Age (years) 45 ±

Disease (no. of patients) CPE (n = 11) ALI (n = 21)

Age (years) 45 ± 20 42 ± 14
Sex, no. (%)  
Male 6 (54) 12 (59)
Female 5 (46) 9 (41)
Caucasian, no. (%) 6 (55) 11 (50)
Current smoker, no. (%) 1 (9) 7 (32)
Sepsis, no. (%) 0 (0) 11 (50)
Days of unassisted ventilation 19 ± 9 7 ± 10
Tidal volume (ml) 677 ± 166 620 ± 220
PaO2/FiO2 143 ± 90 74 ± 27
Lung injury score a 2.50 ± 0.70 3.10 ± 0.50
Hospital mortality, no. (%) 2 (18) 15 (68)

Plus–minus values are mean ± SD
Tidal volumes, PaO2/FiO2 and lung injury scores were observed the onset of the diseases
a Calculated according to the published methods [25]. Higher scores indicate more severe illness

Fig. 3 Comparison of G2F levels in PEF of the patients with CPE and ALI. The box–whisker plots show the 25th and 75th percentiles, the median (horizontal line within the box), and the 10th and 90th percentiles (whiskers). Individual values below the 10th percentile and above the 90th percentile are shown as open circles. The ALI patients were classified into the five groups (day 0, 1, 3, 5 or 7 after disease onset), and each of the groups was further divided into survivors (white box) and non-survivors (shadowed box). The plasma G2F levels of 15 control subjects were indicated as a black box in the left end. Statistical differences between survivors and non-survivors are indicated as follows: *P < 0.02, **P < 0.003, or ***P < 0.0002. The levels in the patients with CPE or IPF were not significantly higher than those in the control subjects (P < 0.0001 on day 0, 5 or 7; P < 0.0004 for day 1; P < 0.002 for day 3). In contrast, plasma G2F concentrations in the surviving patient groups at day 0, 1 or 3 were moderately elevated (P < 0.006 for day 0; P < 0.002 for day 1; P < 0.007 for day 3 vs. control group), but no difference could be detected at day 5 or 7 (P = 0.08 or 0.33 vs. control group, respectively). Plasma levels of G2Fs were not significantly elevated in the patients with CPE (59 ± 36 ng/ml) or IPF (37 ± 17 ng/ml), as compared with those in control subjects (Fig. 4).

Fig. 4 Levels of G2Fs in plasma of clinical subjects. Number of the subjects in each column was indicated in parentheses. The box–whisker plots show the 25th and 75th percentiles, the median (horizontal line within the box), and the 10th and 90th percentiles (whiskers). Individual values below the 10th percentile and above the 90th percentile are shown as open circles. The ALI patients were classified into the five groups (day 0, 1, 3, 5 or 7 after disease onset), and each of the groups was further divided into survivors (white box) and non-survivors (shadowed box). The plasma G2F levels of 15 control subjects were indicated as a black box in the left end. Statistical differences between survivors and non-survivors are indicated as follows: *P < 0.02, **P < 0.003, or ***P < 0.0002. The levels in the patients with CPE or IPF were not significantly higher than those in the control subjects (P < 0.0001 on day 0, 5 or 7; P < 0.0004 for day 1; P < 0.002 for day 3). In contrast, plasma G2F concentrations in the surviving patient groups at day 0, 1 or 3 were moderately elevated (P < 0.006 for day 0; P < 0.002 for day 1; P < 0.007 for day 3 vs. control group), but no difference could be detected at day 5 or 7 (P = 0.08 or 0.33 vs. control group, respectively). Plasma levels of G2Fs were not significantly elevated in the patients with CPE (59 ± 36 ng/ml) or IPF (37 ± 17 ng/ml), as compared with those in control subjects (Fig. 4).

By using repeated measures ANOVA, we also found that time course data of plasma G2F levels in non-survivors were statistically different from those in survivors (P < 0.02). Plasma G2F concentrations of the 20 patients with non-ALI sepsis were not high (32 ± 18 ng/ml, not shown in Fig. 4), similar to levels in the controls (35 ± 14 ng/ml).

Discussion

It is well understood that the degree of alveolar epithelial injury is an important predictor of the outcome in ARDS/ALI [20, 28]. However, there are currently no reliable clinical or biological markers of the extent of alveolar epithelial injury. Therefore, we tested the pathogenetic significance of a novel biological molecule expressed in specific response to the early onset of ALI.
Recent experimental data suggest that accelerating re-
epithelialization of the alveolar barrier leads to the reso-
lution of ALI [3, 21, 28]. Re-epithelialization can occur
even in the absence of the proliferation of epithelial cells
or the recruitment of stem cells immediately after the
injury [3, 4]. Previous studies hypothesized that most of
alveolar repair is mediated by type-II pneumocytes that
migrate and differentiate into type-I cells under stimula-
tion of some component of ECM proteins in newly
deposited basement membranes [29]. We prospectively
expected LM5 may be expressed simultaneously with
type-II pneumocyte migration during the early phase
alveolar epithelial repair after the onset of ALI.

The alveolar basement membrane contains several
isofoms of LMs, and several functions for these adhesion
molecules have been elucidated [6, 30]. LM5 attracted
our interest because it is specifically expressed in epithelium
and stimulates cell migration, especially keratinocyte
remodeling or carcinoma cell invasion [11–14]. Our prior
experimental studies suggest that physiological epithelial
cell migration can be estimated by the circulating levels of
G2F, derived from the essential component γ2-chains of
LM5 [17]. Thus, we hypothesized circulating G2F levels
are associated with alveolar epithelial migration to repair
the injured lung.

Expression of LM5 in type-II pneumocytes was reported
previously, but its function is unknown [30]. In this study,
for the first time, we report that significant increases of G2F
are found in the circulation of ALI patients at the onset of
acute respiratory failure (Figs. 2, 3, 4). Therefore, we
speculate that the alveolar epithelial cells acting to repair the
injured sites should prominently express migration-active
LM5, generated by proteolytic shedding of the G2F domain,
and that soluble G2F is consequently released from the
alveolar surface into the ELF and diffuses from the ECM
into the circulation of ALI patients. The degree of G2F
secretion at the onset of ALI may be independent of disease
severity, as migration of alveolar epithelial cells may reflect
the drive to repair damage per se, rather than the actual
extent of injury. As satisfactory repair of the injured epi-
thelium is achieved, recovery of function in patients
responding to therapy is associated with a decrease of cir-
culating G2Fs at days 5 and 7 (Fig. 4). In contrast, the plasma
G2F levels in ALI patients unresponsive to therapy
remained significantly elevated at days 5 or 7, suggesting
that some physiological signaling might operate to promote
continued LM5 synthesis and proteolytic cleavage of its γ2-
chain by type-II pneumocytes in the absence of adequate
tissue repair, since alveolar epithelial function was not
sufficiently normalized in these cases (Fig. 4). We also
found that mean G2F concentrations in ELF samples from
the ALI patients are comparable with those in PEF speci-
mens from another group of the ALI patients, supporting the
conclusion that G2F levels in ELFs obtained during routine
bronchoscopic monitoring can accurately reflect the quan-
tity of them released into alveolar epithelial surfaces.

Approximately half of the ALI patients usually
received pulse therapy of steroids, which may not affect
their G2F levels (data not shown), because none of
increased G2F was found in the patients with COPD or
IPF probably treated with any steroids.

We should not be regardless of potential bias in our
analysis using multiple specimens from six different
medical centers, because their diagnosis cannot be always
standardized. Our study is the initial step to estimate
significance of circulating G2F in respiratory disorders,
using rarely obtainable specimens such as ELF or PEF.
Increase of G2Fs in the ELF demonstrated they are
mainly derived from injured alveolar tissues, as well as
their increase in the PEF. Some further trials without
potential institutional bias will be recommended, proba-
ably assisted by our present study.

Immunohistochemical observation of LM5 γ2 chains in
IPF lung tissues has been already reported, demonstrating
that regenerating epithelial cells in IPF can actually syn-
thesize γ2 chains, but their expression levels in IPF are
significantly reduced than those in cryptogenic organizing
pneumonia [31]. This evidence suggested that alveolar
LM5 accumulation can be induced preferably at the early
phase of pulmonary injury rather than during the chronic
phase of IPF, corresponding with our present results that
G2F levels in ALI patients at the disease onset were sig-
nificantly higher than those in IPF patients (Figs. 2, 4). We
have also preliminarily detected LM5 expression in small
population of pneumocytes in ALI by immunohistochem-
ical technique (unpublished observation), predicting G2Fs
are endogenously secreted from injured alveolar epithe-
lium. Some further studies are needed to investigate tissue
expression of LM5 in the patients with ALI, whose circu-
lating G2F increased significantly.

Several proteases may be secreted from inflammatory
leukocytes as they transmigrate into injured lung tissue. If
LM5 is deposited in alveolar ECM, it may be degraded
into small fragments in ALI. However, circulating G2F
levels appear to be independent of such random proteol-
ysis, since G2F is unlikely to be deposited in epithelial
ECM [32]. Shedding of G2F from LM5 heterotrimmers in
vivo is critically mediated by membrane-type 1 MMP,
and LM5 processed by the other MMPs cannot promote
epithelial cell migration [16, 33]. We infer that the
physiological action of LM5 to initiate epithelial repair
may be imparted specifically by membrane-type 1 MMP
cleavage and not random proteolysis.

In conclusion, we report that circulating G2F increases
substantially at disease onset in ALI patients. In
patients recovering from ALI, G2F concentrations
decreased in parallel with the clinical improvement; in
contrast, prolonged elevation of G2F was associated with
poorer prognosis, potentially suggesting that circulating
G2F may be a useful biomarker to monitor the pneumo-
cyte re-epithelialization process during alveolar epithelial
repair in ALI/ARDS patients.
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Conflict of interest statement  None of the authors has a financial relationship with a commercial entity that has an interest in the subject of this manuscript.

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