Cyatta abscondita: Taxonomy, Evolution, and Natural History of a New Fungus-Farming Ant Genus from Brazil

Jeffrey Sosa-Calvo1,2, Ted R. Schultz2, Carlos R. F. Brandão3, Christiana Klingenberg4, Rodrigo M. Feitosa5, Christian Rabeling6, Maurício Bacci Jr.7, Cauê T. Lopes8, Heraldo L. Vasconcelos8

1 Maryland Center for Systematic Entomology, Department of Entomology, University of Maryland, College Park, Maryland, United States of America, 2 Department of Entomology, National Museum of Natural History, Smithsonian Institution, Washington, District of Columbia, United States of America, 3 Museu de Zoologia, Universidade de São Paulo, São Paulo, São Paulo, Brazil, 4 Abteilung Entomologie, Staatliches Museum für Naturkunde, Karlsruhe, Karlsruhe, Germany, 5 Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, Paraná, Brazil, 6 Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, United States of America, United States of America, 7 Centro de Estudos de Insetos Sociais, Universidade Estadual Paulista, Rio Claro, São Paulo, Brazil, 8 Instituto de Biologia, Universidade Federal de Uberlândia, Uberlândia, Minas Gerais, Brazil

Abstract

Cyatta abscondita, a new genus and species of fungus-farming ant from Brazil, is described based on morphological study of more than 20 workers, two dealate gynes, one male, and two larvae. Ecological field data are summarized, including natural history, nest architecture, and foraging behavior. Phylogenetic analyses of DNA sequence data from four nuclear genes indicate that Cyatta abscondita is the distant sister taxon of the genus Kalathomyrmex, and that together they comprise the sister group of the remaining neoattine ants, an informal clade that includes the conspicuous and well-known leaf-cutter ants. Morphologically, Cyatta abscondita shares very few obvious character states with Kalathomyrmex. It does, however, possess a number of striking morphological features unique within the fungus-farming tribe Attini. It also shares morphological character states with taxa that span the ancestral node of the Attini. The morphology, behavior, and other biological characters of Cyatta abscondita are potentially informative about plesiomorphic character states within the fungus-farming ants and about the early evolution of ant agriculture.

Introduction

New insect species are being discovered at a rate that is at least twice the historical average [1–3]. As we might expect, many newly discovered species are cryptic species (i.e., multiple species previously thought to comprise a single species) and/or are easily referable to well-known genera or species groups. A significant number of newly discovered species, however, are phylogenetically important, i.e., they are the sole representatives of previously unknown, anciently diverged lineages and, as such, sources of new information about early insect evolution. New supraspecific taxa (e.g., new genera, families, etc.) are commonly erected for such species. For example, a new order of insects, the Mantophasmatodea, was described as recently as 2002 [4–8] (currently considered a suborder of Notoptera [9]).

Despite their status as a relatively well-studied group, ants (Formicidae) are no exception to this trend of increasing species discovery [10]. Over 12,700 ant species in 308 genera have so far been described [11,12], yet most ant systematists estimate that there are twice as many extant species [13,14], which would make ants the most speciose family of social insects [12]. In recent years, more than 19 new extant and ca. 17 extinct ant genera and one subfamily (Martialinae) have been described [15–36].

Fungus-farming "attine" ants are exclusively New World in distribution, ranging from the United States in the North to Argentina in the South [37–39] with six genera (Acromyrmex, Atta, Cyphomyrmex, Mycetophylax, Mycocepurus, and...
Trachymyrmex) also present in the Caribbean [30,39,40]. To date, 256 valid species have been described in 15 extant genera and in one ichnogenus (Attaichnus Lazza) [11,30]. Because attine ants participate in complex associations with their cultivated fungi and other microbial symbionts, they have become model systems for the study of symbiosis and coevolution [41–61]. Attine ants have additionally become model systems for comparative evolutionary studies of mating frequency/polyandry, gyne number, parthenogenesis, social parasitism, caste polyethism and polymorphism, nesting behavior, foraging behavior, and diverse microbial symbioses [47,48,57,62–70]. Leaf-cutter ants in particular are the subjects of a century of applied research due to their status as serious pests of agriculture in Central and South America [71–85].

Here we describe the sole representative species of a new genus, Cyatta abscondita gen. n. et sp. n., in the fungus-farming ant tribe Attini (Formicidae: Myrmicinae) and document its presence in the Brazilian Cerrado, a global biodiversity hotspot [86]; in the Caatinga, a xeric shrub-land and thorn forest in northeastern Brazil; and in Atlantic semi-deciduous forest, considered a transitional zone between humid Atlantic forests and the drier biomes of the Caatinga and Cerrado [87]. We describe the morphology, behavior, fungal associations, nest architecture, and other biological characters of C. abscondita that are potentially informative about plesiomorphic character states within the tribe Attini and within the informal clade Neoattini and, consequently, about the early evolution of ant farming behavior.

Materials and Methods

Material examined

The specimens examined have been deposited in the following institutions:

• BLME Coleção Entomológica, Bacci Laboratory of Molecular Evolution, São Paulo State University (UNESP), Rio Claro, Brazil.
• CRC C. Rabeling Collection, Cambridge, MA, U.S.A.
• DZUP Coleção Entomológica “Pe. Jesus Santiago Mourê”, Departamento de Zoologia, Universidade Federal do Paraná, Curitiba, PR, Brazil.
• MCZ Museum of Comparative Zoology, Harvard University, Cambridge, MA, U.S.A.
• MZSP Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil.
• MBC–UFU Museu de Biodiversidade do Cerrado, Universidade Federal de Uberlândia, Uberlândia, Minas Gerais, Brazil.
• USNM United States National Museum of Natural History, Washington, DC, U.S.A.

Morphological measurements and specimen preparation

All measurements were taken to the nearest 0.001 mm and, unless otherwise noted, are in millimeters. Images of worker and gyne were generated at the USNM Ant Lab using a JVC KY–F75U digital camera mounted on a Leica Z16 APO stereomicroscope attached to a Dell Optiplex GX620 computer. Composite images were assembled using Auto-Montage Pro® (Version 5.03.0061 BETA) software (Synoptics Ltd.). Images of the male were generated at the MCZ using a Leica DFC 420 digital camera mounted on a Leica M216 dissecting scope. Composite images were assembled using Leica Application Suite (Version 4.0) and Helicon Focus (Version 5.3) software packages. The only two larvae collected were dehydrated sequentially through a series of ethanol concentrations to 100% absolute and then critical-point dried in a Balzers CPD–030 using liquid CO₂ at the Scanning Electron Microscopy (SEM) Lab in the SI–NMNH. Once the ethanol was replaced with CO₂ the samples were slowly heated to the critical point, slowly depressurized back to atmospheric pressure, dried, and mounted on aluminum stubs. The two prepared larvae and an adult worker Paratype were sputter-coated with 60:40 wt% Gold:Palladium alloy on a Cressington Scientific 108 auto/SE sputter coater to a thickness of 20–25 nm. Scanning Electron Micrographs (SEMs) of these specimens were generated using a Philips XL–30 ESEM with Lanthanum Hexaboride (LaB6) source and with a backscatter detector. All images were cropped and edited using Photoshop CS5® (Version 12.0) (Adobe Inc.).

The measurements, indices, abbreviations, and morphological terminology utilized throughout follow Gauld & Bolton [88], Klingenberg & Brandão [30], Rabeling et al. [89], Serna & Mackay [90], and Sosa-Calvo & Schultz [91] and literature cited therein, with modifications where noted. Characters and terminology used in the description of the larvae are based on Schultz & Meier [92]. The following abbreviations are used in the description: w= worker, dg= dealate gyne, m= male.

Latitude and longitude coordinates were converted to decimal degree when needed by using the Earth Point Web Site (http://www.earthpoint.us/Convert.aspx). In cases where coordinates were not documented in the specimen label, the coordinates were estimated using Google Earth v7.0 (http://www.google.com/earth/index.html) and are presented within brackets. The distribution map of Cyatta abscondita was generated using the software ArcGIS v10.1 (Esri, Redlands, CA).

Field observations and nest excavations

Field work was conducted at Fazenda Água Limpa from 20–27 February 2009 (JSC, TRS), 12–18 April 2010 (JSC, TRS, CTL), and 14–22 September 2011 (JSC, TRS, CTL); and at the Broa preserve from 21–28 July 2011 (CR, MB). Fazenda Água Limpa (FAL) is a 4490 ha experimental farm and conservation area of the Universidade de Brasilia located at S15.94938° W47.87754°, ~30 km from Brasilia, DF, Brazil, at an altitude of 1048–1160 m. The Broa preserve, one of the southernmost Cerrado preserves of São Paulo state, is located at S22.18517° W47.93567°, ~9.5 km northwest of Itirapina in São Paulo state, Brazil, at an altitude of 530 m.

At FAL, nest entrances were located by following foragers carrying bait (Cream of Rice cereal liberally distributed on the ground) as they returned to the nest. Nests were excavated by first digging a trench about 1 meter wide and 1.5 meters deep and located about 1 meter from the nest entrance, and then by
Carefully shaving away soil in the direction of the nest entrance until either tunnels or nest chambers were encountered; see 93.94 for a detailed description of the methodology. During the course of the excavation, the trench was deepened as necessary and the dimensions, depths, and relative positions of tunnels and chambers were measured, photographed, and sketched. Fungus gardens were transferred from subterranean chambers to plastic containers with flame-sterilized spoons, knives, and/or forceps. Colonies were maintained in live culture in plastic containers, the bottoms of which were lined with plaster saturated with water. At Broa, nest contents were preserved in 95% ethanol after a few days, once the ants had reconstituted and restabilized their fungus gardens and removed all soil particles. Two FAL nests (JSC110920-01 and JSC110919-02) were maintained in live culture for up to four months (nest 4, see Table 1).

Prior to 2009, only four isolated specimens had turned up in mass-collected ant samples. These include, from a leaf-litter sample, one specimen from riparian forest or Cerradão at Paineiras, Minas Gerais (MG), collected in 1999 and, from pitfall trap samples, one specimen from Caatinga at Reserva Particular do Patrimônio Natural (RPPN) Serra das Almas [330m; S05.16479° W40.67978°], Crateús, Ceará (CE), in crude Cyatta abscondita nests, including depths and dimensions of individual chambers, chamber contents, fungus garden morphology, ant demographics, and additional natural history information.

| CHAMBER DIMENSIONS (cm) |
|--------------------------|
| Chamber number | Depth (cm) | Height | Width | Depth | Demography, garde morphology, notes |
|-----------------|------------|--------|-------|-------|-----------------------------------|
| 1 FAL JSC100412-01 April 12-18, 2010 | 1 | 70 | 1.2 | 3 | N/A | 2 pendant garden filaments |
| 2 | 70 | 1.5 | 2.5 | N/A | empty; 3 polydesmid milipedes |
| 3 | 75 | 2.5 | 5 | 5 | 24 pendant garden filaments |
| 4 | 80 | 2.5 | 3.5 | N/A | pendant garden; 6 workers (1 callow) |
| 2 FAL JSC100416-01 April 17-18, 2010 | 1 | 40 | 0.5 | 1 | N/A | small pendant garden; no ants |
| 2 | 46 | 1.5 | 2.5 | N/A | small pendant garden; 2 workers |
| 3 | 50 | 0.5 | 1 | N/A | empty |
| 4 | 55 | 2 | 3 | N/A | small pendant garden; 2 workers |
| 5 | 56 | 2.6 | 3 | N/A | empty |
| 6 | 61 | 2.5 | 5.5 | N/A | empty |
| 7 | 71 | 2.5 | 3 | N/A | empty |
| 3 FAL JSC110919-02 September 19, 2011 | 1 | 29 | 1.5 | 5.5 | 1.3 | empty |
| 2 | 35 | 2 | 3 | 1.2 | pendant garden; 1 workers |
| 3 | 44 | 1.5 | 2 | 0.5 | small pendant garden; a couple of workers; dealate gyne |
| 4 FAL JSC110920-01 September 20, 2011 | 1 | 38 | 2 | 3 | 1 | small pendant garden |
| 2 | 42 | 3 | 2 | 2 | 2 workers |
| 3 | 43.5 | 2 | 2.5 | 3 | 2 workers |
| 4 | 46 | 2 | 3 | 1.5 | 15-20 pendant garden filaments |
| 5 | 50 | 1.5 | 1.5 | 1.5 | small pendant garden; 2 workers |
| 6 | 59.5 | 1.5 | 3.5 | 1.2 | large pendant garden; 7 workers |
| 7 | 82 | 1.8 | 2.5 | 3 | large pendant garden; 4 workers |
| 8 | 104 | 1.9 | 4 | 3.5 | large pendant garden; several workers; larvae; dealate gyne |
| 5 Broa CR110721-05 July 21, 2011 | 1 | 52 | 1.5 | 3 | N/A | fungus garden; 13 workers |
| CR110721-04 | 2 | 80 | 1.5 | 5 | N/A | ~50 pendant garden filaments; 10 workers; 1 male |
| CR110721-08 | 3 | 130 | 1 | 2.5 | N/A | fungus garden; 3 workers |
| CR110721-07 | 4 | 160 | ? | ? | ? | fungus garden; no ants; chamber collapsed during excavation |
| 6 Broa CR110726-09 July 26-28, 2011 | 1 | 102 | ? | ? | ? | fungus garden; 3 workers; chamber collapsed during excavation |
| CR110728-01 | 2 | 195 | 1 | 3 | N/A | fungus garden; 4 workers |

For each nest, chambers are arranged according to depth, descending from the shallowest to deepest chambers. Chamber height refers to the chamber’s vertical axis, chamber width to the horizontal axis parallel to the plane of excavation, and chamber depth to the horizontal axis perpendicular to the plane of excavation. FAL: fazenda Água Limpa, Brasília; Broa: Broa preserve, Itirapina; see text for more details.

doi: 10.1371/journal.pone.0080498.s001

Table 1. Colony demographies and nest measurements of six excavated Cyatta abscondita nests, including depths and dimensions of individual chambers, chamber contents, fungus garden morphology, ant demographics, and additional natural history information.
2003 and two specimens from Cerrado sensu stricto from the Reserva Ecológica do Instituto Brasileiro de Geografia e Estatística (IBGE) (1100m; S15.85000° W48.05000°), DF, Brasilia, in 2008. More recently, in 2011, four additional stray specimens were taken in pitfall traps in Cerrado sensu stricto at Reserva Particular de Patrimônio Natural (RPPN) do Acangau (671m; S17.179083° W 47.058305°), Paracatu, Minas Gerais (MG), and in fragments of semideciduous forest at Fazenda Águas Claras (494m; S21.4023° W48.6873°), Pindorama, São Paulo (SP), and at Estação Experimental (414m; S21.5222° W49.3013°), Sales, (SP).

Permits

Research and export permits for field work by JSC, TRS, CTL, and HLV were issued by the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq; Portarias No. 267, 359) and the Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis (IBAMA; permit numbers 14789–1, 147892, 14789–3). Dr. Yves P. Quinet and Francyregis Nunes allowed CRFB and RMF to study the only known specimen from the Caatinga and supported them during field work in Crateús. Prof. José Mauro da Silva Diogo, Director, allowed JSC, TRS, and CTL to work at FAL. The Fundação Acangau permitted HLV to collect at RPPN do Acangau.

Molecular phylogenetics

DNA extraction, amplification, and sequencing were carried out at the Laboratories of Analytical Biology (LAB) at the National Museum of Natural History, Smithsonian Institution, Washington, DC. Ant genomic DNA was extracted from a worker collected at FAL, Brasilia using the Qiagen DNAEasy Tissue Kit. Four nuclear protein-coding genes, elongation factor 1-alpha paralog F1 (EF1α F1), elongation factor 1-alpha paralog F2 (EF1α F2), wingless (wg), and long wavelength rhodopsin (LW Rh) were amplified and sequenced following MacClade 4.08 [97]. Data were partitioned and modeled as in Schultz & Brady [95] and analyzed using MrBayes 3.1.2 [98] to light brown. Body densely reticulate and covered with minute simple appressed hairs, more abundant on dorsum of head, waist segments, and gaster than on mesosoma. Palp formula 4,2. Anterior margin of clypeus produced into a convex, almost marginal inflated laterally and strongly notched medially. Tubercles on mesosomal dorsum short, attenuate, and blunt. Metapleura with two spiniform processes between mid and hind coxae. Propodeum armed with a pair of short triangular spines. Node of petiole high, well-developed. Gaster lacking

Divergence time estimation

The time of divergence of the new genus Cyatta was estimated using the Bayesian uncorrelated lognormal approach as implemented in the program BEAST v1.7.5 [102] using a normal prior distribution for the root age (as described in Schultz & Brady [95]). Beast XML files were generated using the complementary program BEAUti v1.7.4 (as implemented in the BEAST package). The results combine two runs of 100 million generations each. Burn-in, convergence, and mixing were assessed by examining time series plots and ESS values in Tracer 1.5.0 [101]. Based on this information, burn-in was set at 20 million generations for each run.

Nomenclatural acts

The electronic edition of this article conforms to the requirements of the amended International Code of Zoological Nomenclature ([103]), and hence the new names contained herein are available under that Code from the electronic edition of this article. This published work and the nomenclatural acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information viewed through any standard web browser by appending the LSID to the prefix "http://zoobank.org/". The LSID for this publication is: urn:lsid:zoobank.org:pub:1CF00B3D-CE25-456A-8D12-6DDA628BC110. The electronic edition of this work was published in a journal with an ISSN, and has been archived and is available from the following digital repositories: LOCKSS [http://www.lockss.org] and PubMed Central [http://www.ncbi.nlm.nih.gov/pmc].

Results

Taxonomy

Cyatta gen. n. urn:lsid:zoobank.org:act:0ED4A047-327E-482C-A6AF-E92136DB5697

Figures 1–4.

Type species. Cyatta abscondita sp. n., by present designation.

Worker. Small, monomorphic attine ant, total length (TL)= 2.29–2.56; Weber’s length (WL)= 0.58–0.65. Color pale yellow to light brown. Body densely reticulate and covered with minute simple appressed hairs, more abundant on dorsum of head, waist segments, and gaster than on mesosoma. Palp formula 4,2. Anterior margin of clypeus produced into a convex, almost triangular, smooth, shining flange, i.e., “clypeal apron,” with long unpaired median seta that originates closer to its posterior margin. Psammophore absent. Masticatory margin of mandibles 4-toothed. Antennal scrobes and preocular carinae absent. Antennae 11-segmented. Frontal lobes reduced, barely covering antennal insertions and diverging anteriorly. Frontal area subtriangular, distinct. In full-face view, posterior cephalic margin inflated laterally and strongly notched medially. Tubercles on mesosomal dorsum short, attenuate, and blunt. Metapleura with two spiniform processes between mid and hind coxae. Propodeum armed with a pair of short triangular spines. Node of petiole high, well-developed. Gaster lacking
Cerrado unique specimen identifier No. USNMENT00758173].

**Gyne.** Preocular carina absent. Mandible 4-toothed, apical tooth nearly twice as long as preapical tooth. Parapsidal lines inconspicuous.

**Male.** Mandibles broadly triangular with apical and subapical teeth present. Anterior margin of clypeus (clypeal apron) convex, projecting over mandibles, and with a long median seta. Discal cell present in forewing.

**Etymology.** *Cyatta* is a neologism constructed in part from the Brazilian Tupi language word Cy, meaning “sister,” referring to its status, along with the genus *Kalathomyrmex*, as the sister clade to the remaining genera of the informal clade Neaattini, to which the genus *Atta*, the most conspicuous member of the Neoattini, belongs.

**Cyatta abscondita**, sp. n
urn:lsid:zoobank.org:act:3260572C-B7CE-429C-9440-27613BDBE69E

*Holotype, worker.* labeled: “BRAZIL: DF: Brasilia; Faz. Água Limpa; 1106 m; 47.9013° S 15.8500° W 20.viii.2011; (J. Sosa-Calvo, T.R. Schultz, C.T. Lopes); nest series; Cerrado sensu stricto; in ground; JSC110920-01” [MZSP, unique specimen identifier No. USNM00758173]

*Paratypes.* same data as holotype [3w, MZSP, USNM00758172, 00756921, 00758307], [1dg, 4w, USNM, USNM00758174, 00758223, 00758316–18] [3w, USNM, USNM00521881 (EtOH vial)]. Same data as holotype, but “19.viii.2011; JSC110919-02” [1dg, MZSP, USNM00758175], [2w, USNM, USNM00758176–77] [1w, USNM, USNM00521910 (EtOH vial)]. Same data as holotype, but “16.iv.2010; JSC100416-04” [1w, MZC, USNM00758180] [1w, USNM, USNM00521908 (EtOH vial)]. Same data as holotype, but “1099 m; 47.90129° W 15.95242° S ±3m; 23.i.2009; (J Sosa-Calvo & TR Schultz); nest series underground; CR110721-09” [3w, MZSP, USNM00758219, 1m, MZSP, USNM00758204], [1w, MZC, USNM00758221], [2w, USNM, USNM00758218], [1w, USNM, USNM00758206], [10w, BLME & CRC, USNM00758207], [1w, USNM, USNM00758205, 1w, BLME & CRC, USNM00758199–201]. Same data, but “15.v.2011; CR110721-05” [1w, MZSP, USNM00758217], [1w, MZC, USNM00758218], [1w, USNM, USNM00758206], [10w, BLME & CRC, USNM00758207], [1w, USNM, USNM00758205, 1w, BLME & CRC, USNM00758199–201]. Same data, but “26.vi.2011, CR110722-09” [3w, BLME & CRC, USNM00758199–201]. Same data, but “28.vii.2011, CR110728-01” [3w, BLME & CRC, USNM00758262–65], [1w, USNM, USNM00758205], [SP, Pindorama; Fazenda Águas Claras; 494m; 48.6873° W 21.4023° S; 16.viii.2011, (GA Castilho); Floresta Estacional Semidecidual; pitfall B1 [1w, MZSP], SP; Sales; Estação Experimental; 414m; 49.3013° W 21.5222° S; 17.vii.2011, (GA Castilho); Floresta Estacional Semidecidual; pitfall B1] [1w, MZSP], SP; Sales; Estação Experimental; 414m; 49.3013° W 21.5222° S; 17.vii.2011, (GA Castilho); Floresta Estacional Semidecidual; pitfall B1 [1w, MZSP];...
almost or as long as antennal pedicel, not reaching apex of mandible (Figures 2c,d); clypeus with pair of lateral transverse carinae, each extending from below frontal lobe to mandibular insertion. Medially these carinae developed into lamellae perpendicular to clypeal face, thus forming a wall that divides the clypeus laterally into anterior and posterior areas, very
likely homologous with clypeal morphology of closest relative, *Kalahathomyrmex emeryi* (Figure 2c); medially clypeus is not so divided, face extending posterad between frontal lobes. Frontal lobes reduced, convex, barely covering antennal insertions (Figures 1a, 2c). Frontal carina fading out posteriorly at midlength of compound eye (Figures 1a, 2c). Well marked triangular area with concave anterior margin between frontal lobes reticulate, bordered anteriorly by rounded finger of clypeus, which extends broadly posterad between frontal lobes. Compound eye set slightly before middle of head, with 7–9 ommatidia at maximum length and 6 ommatidia at maximum width (33–47 ommatidia in total). Antennal scape covered with minute, simple, appressed hairs; antennal scape wider at seven-tenths of its length, and slightly surpassing postero-lateral corners of head when laid back over head capsule; first funicular segment (pedicel) slightly longer than or as long as second and third funicular segments combined. In full-face view, cephalic margin deeply notched medially (i.e., at vertex) and rounded laterally (Figure 1a), shallow vertexal sulcus extending medially towards frontal lobes, fading at eye level; in lateral view, ventral face of head slightly convex. 

**Mesosoma.** Profile of promesonotal dorsum in lateral view distinctly tuberculate, tubercles attenuate and blunt (Figures 1c,e, 2a). In dorsal view, promesonotum with raised shield-like area, broad anteriorly and narrowing posteriorly, distinctly separated from lower, lateral promesonotum; raised area formed anteriorly by triangular lateral pronotal tubercles and two median low and approximately pronotal tubercles and posteriorly by eroded remnants of promesonotal tubercles; lower, lateral area of promesonotum in dorsal view subtriangular, broader and anterolaterally angled anteriorly; in lateral view, inferior corner of pronotum rounded, lacking spines or angles. Aneupisternum indistinctly separated from katepisternum. Metanotal groove relatively broad and strongly impressed, in lateral view extending to antero-ventral margin of metapleuron. Metapleuron ventrally with two spiniform processes between mid and hind coxae, best seen by removing hind legs. Basal (dorsal) face of propodeum in lateral view a low, rounded, protuberance posterior to metanotal groove; in dorsal view, basal face very small, raised above remainder of propodeum, and narrowing anteriorly; decolous face of propodeum behind protuberance concave; propodeal spines triangular (Figure 1c), obliquely directed upwards and strongly diverging in dorsal view; declivity of propodeum much longer than base (dorsum); propodeal spiracle opening in an angle of 45° in relation to main body axis; in lateral view, propodeal lobes rounded without posterior projections.

Peduncle of petiole vestigial; in lateral view, petiolar node well developed, subquadrate, with anterior face almost straight and vertical; dorsum of petiolar node short and almost flat, meeting vertical posterior face in slightly rounded angle; ventral face of petiolar node slightly concave or straight medially, lacking petiolar process (Figures 1c,e); in fronto-dorsal view, node of petiole shallowly V shaped, dividing node into a pair of rounded tubercles. Postpetiole robust, almost twice as long as petiole and slightly less than 0.5x gaster length; in dorsal view, postpetiole subtriangular, anterior margin rounded, lateral margins slightly diverging posteriorly; posterior margin with deep median impression, forming two distinct small lobes; in lateral view, anterior portion of postpetiole convex, postpetiole relatively compressed dorsoventrally; ventral projections absent (Figures 1c,e). In profile, gaster elliptical and dorsally finely reticulo-striate; in dorsal view, apical margin of pygidium (gastral segment IV, i.e., abdominal segment A7) medially emarginate, bilobed; gastral sternite IV (hypopygium, i.e., A7) covered with simple decumbent hairs; in lateral view, pygidium rounded, laterally overlapping and concealing the hypopygium; pygidium weakly reticulate and shiny; in ventral view, pygidium posteromedially emarginate (i.e., V-shaped), the triangular hypopygium fitting within the emargination of the pygidium (Figures 2e,f). Sting apparatus present, protruding through emargination on pygidium.

Color pale yellow to light brown; antennae, mandibles, and legs lighter than rest of body. Body integument areolate, with short appressed simple hairs, appearing almost hairless.

Gyne. As in worker description, but with caste-specific morphological differences as follows. All gynes studied are dealate. **Head**: Eyes large, with 10–11 ommatidia in maximum length and 9 ommatidia in maximum width, ~65 ommatidia total; median ocellus rounded, located in a median sulcus extending almost from the occipital carina in the back of the head to the middle of the frons; integument surrounding ocelli darker in color than elsewhere. Clypeus with unpaired median seta arising on short transverse wrinkle-like ridge that crosses clypeal apron; two to four short simple appressed hairs on clypeal apron on each side of median clypeal seta. In full-face view, cephalic border with median (vertexal) notch, not as deep as in worker. Mandibles dorsally coarsely rugose.

**Mesosoma.** Pronotal dorsum conspicuously areolate, lacking anterior pronotal tubercles; lateral pronotal tubercles present, blunt and small; humeral pronotal tubercles vestigial. Mesoscutum, in dorsal view, rounded to slightly ovate and overall reticulo-rugose; dorsum of mesoscutum, in profile, almost flat; mesoscutal sulcus, in dorsal view, short, not extending more than 1/4 length of mesoscutum; notauli absent; parapsidal lines short, inconspicuous, and extending nearly to lateral margin of mesoscutum; transscutal suture conspicuous. Scuto-scuteellar sulcus deep and with ~7 transverse carina; margin of axilla rounded, dorsally reticulo-rugose. Scutellum posteriorly weakly bidentate, dorsally rugose and with a shallow median longitudinal groove. Anapleural sulcus deep, with transverse carinae, dividing mesopleuron into anepisternum and katepisternum. Metanotal groove extended into a complete metanot-propodeal suture (sensu Serna & Mackay [90]). In profile, metanepisternum (sensu Serna & Mackay [90] present, small. In profile, metanotal groove conspicuous, continuous with mesometapleural suture. Ventral metapleural processes present as a pair of spiniform tubercles between the mid and hind coxae, similar to, but longer than, those present in worker. Propodeum with pair of short, right-angled denticles; dorsum, lateral margin, and declivity of propodeum reticulate. **Metasoma**: petiole as in worker. Dorsum of postpetiole reticulo-rugose. Dorsum of gastral tergite IV (A7) rugulose; pilosity as in worker; gastral tergite and sternite IV (pygidium and hypopygium; A7) as in worker.
Measurements. TL 3.27–3.32, WL 0.86–0.87, HL 0.65–0.66, HW 0.60–0.63, SL 0.56, ML 0.41–0.42, EL 0.16–0.17, PL 0.25–0.28, PPL 0.29–0.30, GL 0.79–0.81, CI 91–96, SI 89–93, MI 62–65, FLD 0.20–0.21 (n=2).

Male. A medium-sized male with head large relative to size of the mesosoma. Mandibles broadly triangular, apical and subapical teeth present large; remaining tooth minute, indistinct; texture coarsely granulate. Palp formula 4.2. Clypeus broadly trapezoidal in frontal view; anterior margin convex, with

![Image of ant specimens](https://example.com/ant_images.png)

**Figure 2.** Paratype worker (USNMENT00758223) of *Cyatta abscondita*. (A) Habitus, lateral view. (B) Head, lateral view, indicating palp formula. (C) Head, full-face view. (D) Clypeal apron indicating origin of median unpaired seta. (E) Metasoma, posterior view. Pygidium (p) 'V'-shaped, hypopygium (hyp) triangular. (F) Metasoma, ventral view, showing the median emargination of the pygidium and the triangular shape of the hypopygium.

doi: 10.1371/journal.pone.0080498.g002

*Cyatta*: A New Genus of Fungus-Farming Ants
a long median seta (0.11 mm) originating at the anterior margin and projecting over the mandibles; in lateral view anterior margin of clypeus forming a lamella projecting over the mandibles. Frontal lobes triangular, only partly covering the condyloid bulbs of the scape in full face view. Antennae with 13 segments; scape surpassing the posterior border of the head by 1/3 of its length. Antennal funicular segment II (0.08 mm) almost as long as funicular segment I (pedicel; 0.11 mm) (Figures 3a,b). Eyes conspicuously large, at maximum diameter approximately half as long as the entire head, counting ~15 ommatidia in maximum width and ~23 ommatidia in maximum length. Ocelli large, elevated above the remainder of the head. Surface of head coarsely granulate, finely rugulose around the ocelli. Tergum of promesonotum not distinctly enlarged, giving the mesosoma a rather slender appearance in lateral view. In dorsal view, lateral pronotal teeth pyramidal, twice as wide at the base than high, with sharp tips. Propodeal node with a broadly rounded ventral lobe. Reticulate sculpture on other attine genera, including the closest relative of *abscondita* known to us, unique in the Attini; this character is absent in all other attine genera the: (iv) thoracic-abdominal articulation apparently absent; (v) thoracic intersegmental constrictions superficial; (vi) deep lateral depressions associated with abdominal spiracles absent; all states shared with other Attini. Remarkably, body hairs present dorsally and laterally, a condition otherwise common in the Myrmicinae but rare in the Attini, in which larvae usually lack dorsal and lateral hairs. In the Attini, such hairs are known to be present only in the larvae of *Mycocepurus goeldii* and *M. smithii* [92], where their presence may be plesiomorphic, and in *Sericomymex* and some *Acromyrmex* species, where their presence is likely secondarily derived. Predominant hair type bifurcate with "anchor tips" (Figures 4a,d). Two rows of dorsomedian, very long anchor-tipped hairs present (Figure 4e). Labrum monolobate, narrow, bulging. Anterior setae present as papillae. Mandibles typically attine: short, fleshy, subconical. A distinct, undivided apical mandibular tooth and no subapical teeth; spinules evenly distributed on all mandibular surfaces. Mandibular gnathobases absent. Basal portions of maxillae fused with head capsule. Maxillary palp widely removed laterad from galea, a synapomorphy for the Neoattini. Galea reduced, present as two sensilla surmounting a low protuberance, as in all Attini except for some *Myrmicrypta* species. Maxillary palp digitiform, maxillary accessory palpal sensillum absent. A single seta present laterad of maxillary palp, a character shared with *Mycocepurus* species. As in most attines, labium feebly protruding, lateral sericteral protuberances absent, labial palps digitiform. Labial spinules present on anterior surface slightly dorsal to the sericteries. Hypopharyngeal spinules multidentate and apparently densely distributed. On the head, genal lobes absent, a state in the Attini shared with *Myrmicrypta*, *Apterostigma*, *Mycocepurus*, and *Mycterotes* species. Supra-antennal setae present and abundant, a condition common in the subfamily but otherwise present in the lower Attini only in *M. goeldii*. Subantennal (genal) setal arrangement plesiomorphic for the tribe, consisting of around 12 setae on each gena. Supraclypeal setae present and setiform. Two clypeal setae present. Spinules absent on the head dorsad of the labrum, the state common to most attines. Due to the poor condition of specimens, most ventral thoracic/abdominal characters could not be studied, including the presence/absence of: leg vestiges, prothoracic food anchor, ventromedian protuberances on various segments, papilliform spinules, and hairs.

Two setal sockets occur ventral of the anal opening on abdominal segment IX (Figure 4f). No other setae are associated with the anal opening. Ventral anal lip absent.

**Comments.** The new genus *Cyatta* shares with other genera belonging to the tribe Attini: (i) the presence of a thick unpaired median seta arising from the clypeal apron (considered a synapomorphy for the tribe by Brandão & Mayhé-Nunes [105]; but, along with the clypeal apron, presumed secondarily lost in *Kalathomymex* [30]); (ii) 11 antennal segments in the worker and gyne, 13 in the male (the latter secondarily reduced in some Cyphomyrmex, *Mycetagroicus*, *Sericomymex*, *Trachymyrmex*, and social parasites); (iii) palpal formula 4,2 (plesiomorphic for the Attini, secondarily reduced in *Apterostigma* and some social parasites). Larvae of *Cyatta* share with the larvae of other attine genera the: (iv) thoracic-abdominal articulation apparently absent; (v) thoracic intersegmental constrictions superficial; (vi) deep lateral depressions associated with abdominal spiracles absent; (vii) short, narrow labrum; and (viii) fleshy, subconical mandibles. Behaviorally, *Cyatta* shares with other Attini the cultivation of fungi for food.
The genus *Cyatta* shares with other members of the neoattine clade: (i) the antennal scape of the male long, longer than the sum of the length of antennal funicular segments I–III; (ii) the first funicular segment (pedicel) of the antenna of the male longer than second funicular segment; (iii) the petiole in workers somewhat sessile; (iv) the lack of hypostomal teeth in workers and gynes; and (v) the maxillary palp of the larva widely removed laterad from the galea.

*Cyatta* shares with its sister genus, *Kalathomyrmex*, (i) the lack of a tubercle or spine on the inferior lateral margin of the pronotum, a symplesiomorphy shared with the paleoattine clade; (ii) the clypeus with a pair of lateral transverse carinae, each extending from the frontal lobe to the mandibular insertion and each medially developed into a lamella perpendicular to the clypeal face, thus forming a wall that divides the clypeus laterally into anterior and posterior areas; and (v) the maxillary palp of the larva widely removed laterad from the galea.

*Cyatta* differs from its sister genus *Kalathomyrmex*, however, by (i) having, on the forewing of the male (forewing of gyne unknown), a closed marginal cell (Figure 3d) (open in the forewings of both the male and gyne of *Kalathomyrmex* [Klingenberg and Brandão ([30]), therein as radial cell]); (ii) the mesoscutum of the male with strongly impressed notauli (absent in the male of *Kalathomyrmex*); (iii) the pronotum of the male with lateral pronotal tubercles present, pyramidal (the pronotum in the male of *Kalathomyrmex* lacks any tubercles); and (iv) the psammophore absent in the worker, the gyne, and the male.

In addition to the previously mentioned absence of an inferior pronotal tubercle in adult workers, shared with *Kalathomyrmex*, *Cyatta* differs from all or most other Neoattini in a number of larval character states shared with the Paleoattini, suggesting that they may be retained symplesiomorphies, including: (i) the apical and preapical teeth are the largest and have a multidentate (saw-like) margin.

*Cyatta* differs from its sister genus *Kalathomyrmex*, however, by (i) having, on the forewing of the male (forewing of gyne unknown), a closed marginal cell (Figure 3d) (open in the forewings of both the male and gyne of *Kalathomyrmex* [Klingenberg and Brandão ([30]), therein as radial cell]); (ii) the mesoscutum of the male with strongly impressed notauli (absent in the male of *Kalathomyrmex*); (iii) the pronotum of the male with lateral pronotal tubercles present, pyramidal (the pronotum in the male of *Kalathomyrmex* lacks any tubercles); and (iv) the psammophore absent in the worker, the gyne, and the male.

In addition to the previously mentioned absence of an inferior pronotal tubercle in adult workers, shared with *Kalathomyrmex*, *Cyatta* differs from all or most other Neoattini in a number of larval character states shared with the Paleoattini, suggesting that they may be retained symplesiomorphies, including: (i)
dorsal and lateral body hairs present and abundant, shared with Mycocepurus species; (ii) a single seta present laterad of the maxillary palp, shared with Mycocepurus species; (iii) supra-antennal setae present and abundant, shared with Mycocepurus goeldii; (iv) genal lobes absent, shared with the paleoattines and the neoattine genus Mycetarotes. Larval characters are unstudied in Kalathomyrmex. In addition, the worker and gyne of Cyatta differ from members of the neoattine clade by (v) the node of the petiole well developed, high (Figures 1c,d, 2a).

Most notably, Cyatta differs from all other attine genera and species by the following autapomorphies: (i) mandible of the worker and gyne with four teeth (Figures 1b, 2c); (ii) in ventral view, metapleura of the worker and gyne with two spiniform...
processes between the mid and hind coxae, apparently absent in the male; (iii) apical margin of the pygidium medially emarginate, V-shaped (Figures 2e,f); and (iv) forewing of the male with a closed discal cell (Figure 3d).

Based on the extreme degree of morphological divergence and the results of the divergence dating analyses (see below), we have chosen to describe *Cyatta* as a new genus rather than to describe it as a species within the genus *Kalathomyrmex*.

**Discovery history.** In 2003, a single stray worker of *C. abscondita* was taken in a pitfall trap as part of an ant survey conducted at the Reserva Particular do Patrimônio Natural Serra das Almas, Crateús, CE, Brazil, a relatively undisturbed area of Caatinga, a biome characterized by deciduous thorny woodland vegetation [106]. The specimen was deposited in the MZSP ant collection, where it was at first associated with the *Myctophyllax* species group, but subsequently recognized as a new neotropical genus by CK and CRFB. This isolated specimen inspired the first attempt to locate *C. abscondita* in the field in Serra das Almas in 2009 by CRFB and RMF. Unfortunately, it was the end of the rainy season and the soil was covered by a dense layer of grass, impairing observations of all small and inconspicuous ants. Visual searching and leaf-litter extraction failed to locate additional specimens, as did subsequent surveys at the same locality.

In 2008, two workers were taken in pitfall traps in the Instituto Brasileiro de Geografia e Estatística (IBGE) Cerrado preserve, near FAL in Brasília, DF, Brazil. These specimens, deposited in the MZSP, inspired attempts by JSC, TRS, CTL, and HLV to locate the species at this locality beginning in 2009. The first such attempt yielded only the collection of a series of stray workers and an unsuccessful nest excavation; however, subsequent visits resulted in the excavations of multiple nests and collections of gynes, larvae, and cultivated fungi.

The only known male of the species was fortuitously collected in 2011 by CR and MB when they accidentally encountered two nests of *C. abscondita* while excavating a nest of *Mycocepurus goeldii* in the Broa Preserve, Itirapina, SP, Brazil.

The earliest known collection of *C. abscondita* was that of a stray worker taken in a leaf-litter sample in Paineiras, MG, in 1999, only recently discovered in the entomological collection at MZSP and recognized as belonging to this species. Most recently, in 2011, two workers of *C. abscondita* were recovered from pitfall traps in fragments of semideciduous forests in the Sales and Pindorama municipalities in northwestern São Paulo state. This history of discovery indicates that *C. abscondita* is rarely collected by traditional methods. The cryptic nature of foragers and of nest entrances makes it almost invisible to traditional hand collecting. The rarity of individuals in pitfall and leaf-litter samples remains puzzling, since the concentrations of nests encountered at FAL and Broa Preserve suggest that it is locally abundant. Now that the genus and species are recognized and described, we hope that additional specimens will be identified in unsorted material in collections as well as in newly collected material from ant surveys in Brazil and perhaps even elsewhere in South America.

**Natural history**

**Macrohabitat.** Most collections of *C. abscondita* are from Cerrado localities (Figure 5). These include Fazenda Água Limpa (FAL) near Brasilia, the Broa preserve in São Paulo, the IBGE Cerrado preserve in Brasilia, DF, the Fazenda Olho D’Água in Paineiras, MG, and the Reserva Particular do Patrimônio Natural (RPPN) do Acangau in Paracatu, MG, all of which are characterized by diverse Cerrado phytophysionomies, ranging from campo limpo to Cerrado sensu stricto [107]. The predominant habitat, Cerrado sensu stricto, is a low canopy arboreal woodland that is characterized by the presence of small trees with a canopy height of less than 7 meters, shrubs, and abundant ground vegetation [108–110]. Cerrado soil is typically a red-yellow latosol, largely composed of well-drained and nutrient-poor quartz sand with moderate clay content below 15% [107,111]. Both FAL and Broa have typical Cerrado climates with a marked dry season from May to September and with a mean annual temperature and precipitation of 23°C and ~1420 mm, respectively [110]. For a complete account of vegetation and soil compositions at FAL and Broa, see [112] and [113]. The label data associated with the *C. abscondita* worker from Fazenda Olho D’Água indicates that it was obtained from a leaf-litter sample from a Winkler extractor. This suggests that this worker was likely taken in either a riparian forest or in “Cerradão,” because it is in such areas that trees are dense enough to produce conspicuous accumulations of leaf litter.

Some specimens of *C. abscondita* have been taken outside the Cerrado biome (Figure 5). Most notably, a stray worker was recovered from a pitfall sample taken in 2003 in a relatively undisturbed area of Caatinga, a biome characterized by deciduous thorny woodland vegetation [106], in the RPPN Serra das Almas, Crateús, CE, Brazil. More recently, in 2011, two workers of *Cyatta abscondita* were taken in pitfall traps in fragments of semideciduous forests in northwestern São Paulo state. This region is considered a transition zone between Cerrado and endangered coastal Atlantic Forest [114].

**Microhabitat.** Four nests of *Cyatta abscondita* were excavated at FAL and two nests at Broa (summarized in Table 1). Nest entrances of four additional nests were located at FAL; however, attempted excavations of these nests failed. At FAL, seven of the excavated nests occurred on the side of a little-used dirt service road in Cerrado sensu stricto (Figure 6b) and the eighth (nest 1, Table 1) on the lawn of the FAL dormitories (Figure 6a). The roadside nests were directly exposed to sunlight for most of the day, whereas the lawn nest was shaded by the adjacent building in the morning and afternoon. At the Broa preserve, both nests occurred in the shade of trees in Cerrado sensu stricto. Both Broa colonies were excavated serendipitously during excavations of *Mycocepurus goeldii* nests and the *C. abscondita* nest entrances were not observed.

**Foraging behavior.** Foraging workers of *Cyatta abscondita* are difficult to locate because colony sizes are small, workers forage individually, and individuals are very small and cryptic (Figure 6c). In mid-September, which coincides with the beginning of the rainy season at FAL, three to four individuals...
from three different nests, including nests 3 and 4 (Table 1), were observed foraging at night between 22h and 23h. In February and April, at the end of the rainy season, ants from five FAL nests, including nests 1 and 2 (Table 1), were likewise observed foraging individually only after sunset. Only in the case of FAL nest 1 were workers observed to forage in the

Figure 5. Known distribution of *Cyatta abscondita*. doi: 10.1371/journal.pone.0080498.g005
Figure 6. Habitat of *Cyatta abscondita*. (A, B) Fazenda Agua Limpa (FAZ). (A) Excavation of nest JSC100412-01 in dormitory garden area. (B) Cerrado *senso stricto*, where colonies were found on the side of the road. (C,D) Nest entrance of *Cyatta abscondita* (white arrows). (C) Worker entering nest. (D) Nest entrance of *Cyatta abscondita*, consisting of an inconspicuous ~1mm diameter hole in the ground. (E) Chamber with pendant fungus garden. (F) Excavation of nest 4 (JSC110920-01). Black bars indicate two chambers, the lower one 104 cm below the surface.

doi: 10.1371/journal.pone.0080498.g006
early afternoon on two consecutive days in April between 13h and 15h during a time when the nest entrance was shaded from direct sunlight. Unlike the other nests, nest 1 occurred in a well-watered, human-managed grassy lawn. Nests 5 and 6 (Table 1), which were collected in July at the height of the dry season at the Broa preserve in São Paulo State, were located in the shade and, unfortunately, neither foraging nor nest-building activity was observed.

The entrance of one FAL nest (JSC090223-26) was located ~4.5 centimeters from the entrance of an adjacent Mycocepurus goeldii nest. At around 23h a Cyatta abscondita worker was observed lurking slightly inside the nest entrance while workers of M. goeldii foraged on bait (granules of Cream of Rice cereal) placed near the nest entrances. When M. goeldii workers were absent, the C. abscondita worker darted out to retrieve a piece of bait and quickly returned to its nest. This lurking and rapid foraging behavior was repeated until the supply of bait was depleted. In rare cases of contact between C. abscondita and M. goeldii workers, C. abscondita workers were observed to remain motionless. Aggressive interactions were not observed.

**Nest architecture.** At FAL, nest entrances of Cyatta abscondita consisted of a single, inconspicuous, hole in the ground of approximately 1 mm in diameter without any accompanying mound or turret (Figures 6c,d). As mentioned above, the entrance of one nest was located in the mound of a Mycocepurus goeldii colony ~4.5 cm from the M. goeldii nest entrance. At the Broa preserve, nests chambers of C. abscondita were encountered serendipitously while excavating M. goeldii nests and the nest entrances were not observed.

At FAL, excavations of eight nests were attempted. Four excavations (JSC090223-26, JSC110914-02, JSC100415-03, JSC100416-04) failed (i.e., neither chambers containing fungus gardens nor gynes were found, but workers were collected at their respective nest entrances), but chambers containing fungus gardens were located in four nests (Table 1). Nests contained three to eight chambers. In FAL nests 1 and 3, which contained 4 and 3 chambers respectively, chambers were roughly arranged vertically below the nest entrance (Table 1), although it is possible that additional, laterally dispersed chambers were missed during the excavations. At FAL, nests 2 and 4 contained 7 and 8 chambers, respectively, some of which occurred at the same depth but were laterally separated from each other (Figure 6f). The shallowest chamber encountered (nest 3, FAL) was 29 cm deep and the deepest chamber (nest 6, Broa) was 195 cm deep. Because no gynes were found during the nest excavation at Broa, it is entirely possible that additional chambers occurred below a depth of 2 meters. Chambers were elliptically shaped, 1–2.5 cm wide and 2–5 cm high (Figure 6e). The largest garden chamber encountered (nest 2, FAL) was 2.5 x 5.5 cm; at Broa, a similarly sized chamber (nest 5) contained ~50 hanging garden filaments (Figure 6e). Some chambers were empty; in one case, an empty chamber contained three polydesmid millipedes.

**Demography.** Dealate gynes were collected in three of the eight excavated nests, suggesting that in five nests additional chambers remained undiscovered in the soil, or that gynes escaped into adjacent tunnels. In each of the three queenright nests (nests 2, 3, and 4), a single gynae was consistently encountered in the deepest chamber (see Table 1); however, additional chambers may have been present at greater depths, because excavations were generally terminated upon encountering the gynae. Brood was found only in FAL nest 4, which was collected in September, the beginning of the rainy season, suggesting that colonies of Cyatta abscondita reduce their reproductive activities during the dry season. The maximum number of workers encountered in colonies ranged from ~20 (FAL nest 4) to 26 (Broa nest 5). One male was collected in nest 5 at Broa on 21 July.

**Garden morphology.** Gardens were pendant and arranged in filamentous curtains suspended from the chamber ceiling (Figure 5e), similar to the fungus gardens of Mycocepurus species [70,93,115,116] and of Kalathomyrmex emeryi (TRS, JSC, pers. obs.). Single fungal curtains were 5–6 mm long and 1–2 mm wide and a maximum number of 50 curtains were found in a single chamber. Curtains were directly attached to the soil of the chamber ceiling rather than to rootlets. In nest 4, which was maintained in laboratory culture for three months, workers attached garden filaments to the plastic ceiling of the nest box and cultivated suspended gardens. The filaments were firmly attached to the plastic ceiling by an unknown mechanism.

**Phylogeny.** Results of molecular phylogenetic analyses incorporating four nuclear gene sequences from Cyatta abscondita confirm the previous finding [95] that the tribe Attini is divided by an ancient divergence into two major clades, the Paleoattini and the Neoattini (Figure 7). Cyatta abscondita occupies a relatively isolated position in the latter clade, distantly related to the monotypic genus Kalathomyrmex Klingenber & Brandão, the result of an early divergence in the Neoattini. Its phylogenetic position, nested well within the paraphyletic group of "lower attine ants," strongly supports the hypothesis that C. abscondita practices "lower attine agriculture" [58].

A relaxed-clock-divergence dating analysis conducted in BEAST using the Bayesian uncorrelated lognormal approach with a normal prior on the root node (as described in Schultz & Brady [95]), resulted in a chronogram in which Cyatta occupies a position identical to that in the MrBayes results shown in Figure 7. The BEAST chronogram indicates that Cyatta and Kalathomyrmex diverged 26 Ma (95% CI= 18–34) and that the earliest possible divergence of the clade (Cyatta + Kalathomyrmex) from the rest of the Neoattini occurred 44 Ma (95% CI= 37–51).

**Discussion**

The pursuit and discovery of phylogenetically informative new species are arguably among the most important enterprises in systematic biology. Numerous studies have demonstrated the significant effects of taxon representation on phylogenetic inference, including, in addition to tree topology, ancestral character state reconstruction, divergence time estimation, and inferences of evolutionary rates [117–119]. Ward et al. [118] showed that the exclusion of a single relict
Figure 7. Phylogenetic position of *Cyatta abscondita*. This phylogeny of 66 fungus-farming and 26 outgroup ant species results from a Bayesian analysis of four nuclear protein-coding genes (see text for details). Fungus-farming ant species are indicated by bold black branches; the branch subtending *Cyatta abscondita* is indicated in red. Gray branches correspond to non-fungus-farming taxa. Blue dots on branches represent Bayesian posterior probabilities of 100; orange dots represent Bayesian posterior probabilities of 95–99. Divergence time of *Cyatta* and *Kalathomyrmex* estimated at 26 Ma (95% CI= 18–34) and divergence of the clade (*Cyatta + Kalathomyrmex*) from the rest of the Neoattini estimated at 44 Ma (95% CI= 37–51).

doi: 10.1371/journal.pone.0080498.g007
addition, the recently discovered ant genus Martialis, a relict subterranean species known from the Amazon forest in Brazil, has been shown to be the only hitherto known representative of an early diverging branch of the ant tree of life [27]. Within the Attini, the recently described ant genus Myctegatroicus [105,120] was found to be the sister group to the higher attine ants (Sericomymnex, Trachymymnex, Acromymnex, and Atta) and thus to occupy a phylogenetic position transitional between lower and higher agriculture [95]; however, until very recently its fungal cultivar association remained unknown. Subsequent field work documented that Myctegatroicus cerradensis cultivates a lower attine fungus, suggesting that biological investigations of the genus are critical for understanding the evolutionary transition from lower to higher agriculture [94]. This strategy of reciprocal illumination (i.e., information gathered from the field informing phylogenies, and phylogenies guiding field work) plays a key role for reconstructing and understanding the macro- and micro-evolutionary processes driving the attine agricultural symbiosis.

The results reported here indicate that Cyatta abscondita possesses an intriguing mosaic of characters, some that are shared with paleoattines, others that are shared with neoattines, and at least one that is shared only with non-attine ants. Because the Neoattini, Paleooattini, and the non-attine Myrmicineae span the ancestral node of the tribe Attini, this combination of character states suggests that the morphology, behavior, fungal associations, nest architecture, and other biological characters of C. abscondita are potentially informative about plesiomorphic character states within the tribe and, consequently, about the early evolution of ant agriculture.

At least one character of C. abscondita, the presence of a closed discal cell in the forewing of the male, is unknown in all other Attini. If, as we suspect, this is a retained plesiomorphy rather than an autapomorphy, then the absence of the discal cell in other Attini must be the result of at least three parallel losses, one in the Paleooattini, one in the Neoattini, and one in Kalathomyrnx.

Another character previously unknown in the Attini is the presence of at least two rows of elongate anchor-tipped hairs (Figures 4a,e) on the mid-dorsum of the larva. The function of such anchor-tipped hairs has recently been studied in the non-fungus-farming ant Pheidole rhea, which utilizes these specialized setae to hang fourth-instar larvae from the nest walls [121]. The widespread presence of this character state in non-fungus-farming ants strongly suggests that it is plesiomorphic for the Attini and that its presumably derived absence in most Attini may be connected to the fact that larvae are usually more or less enveloped in mycelium deep within the fungus garden rather than hung from the chamber wall.

Presumed plesiomorphic adult character states shared by C. abscondita and the Paleooattini include the presence of a rounded inferior lateral margin of the pronotum, also retained in Kalathomyrnx emeryi. All other neoattine genera, from Mycetarotes to the leaf-cutter ants, have a denticle or tooth in this position. A number of larval character states are shared with species of the paleoattine genus Mycocepurus, including the presence of dorsal and lateral hairs, the presence of a single seta lateral of the maxillary palp, and abundant supra-antennal setae. The absence of genal lobes is shared with all Paleooattini as well as, in the Neoattini, with Mycetarotes species. (Larval characters of Kalathomyrnx have not yet been documented.) The striking pendant, curtain-like morphology of the fungus garden is a character state shared with Mycocepurus species as well as with Kalathomyrnx.

Presumed neoattine synapomorphies shared by C. abscondita and other Neoattini include, in adults, (i) the lack of hypostomal teeth, also secondarily lost in some species of Apterosigma; (ii) the antennal funicular segment II of males short, as long as or slightly longer than funicular segment I (pedicel), whereas in the males of the paleoattine genera the funicular segment II is long, almost twice as long than the pedicel; and (iii) the wide separation of the maxillary palp from the galea in the larvae.

Although both molecular and morphological data indicate that Cyatta abscondita is the sister species of Kalathomyrnx emeryi, it is a very distant sister, having diverged from their most recent common ancestor approximately 26 mya. As far as is currently known, C. abscondita shares with Kalathomyrnx two unique morphological characters, the form of the clypeus and the morphology of the mandibles in the male. In common with the paleoattines, but differing from all other neoattines, C. abscondita also shares with Kalathomyrnx a rounded inferior pronotal corner. In contrast to these shared character states, two synapomorphic and one symplesiomorphic, C. abscondita notably lacks the defining feature of Kalathomyrnx, the basket-like psammophore (Gr. kalathos = “basket”) and differs not only from Kalathomyrnx but from all other Attini in a number of striking characters, including the 4-toothed mandible, the presence of paired ventral pleural spiniform processes, and the presence of a discal cell in the wing of the male. In fact, the morphology of C. abscondita is a mosaic of characters of the paleoattine and neoattine clades as well as of closely related non-attine myrmicines. For these reasons we choose to recognize Cyatta abscondita as a distinct genus within the fungus-farming ants.

The discovery, description, and mapping of biological diversity is essential for devising strategies for protecting biodiversity hotspots, i.e., areas that support high concentrations of endemic species and that are threatened due to the rapid loss of habitat as a result of years of unsustainable human exploitation [86], [122]. The Brazilian Cerrado is one such biodiversity hotspot. With only 20% of the original primary habitat remaining, and with only 6.2% of that habitat protected, the Brazilian Cerrado sustains more than four thousand species of endemic plants and more than a hundred species of endemic vertebrates [86]. The Caatinga is even less protected than the Cerrado [123]. The discovery of Cyatta abscondita in the Cerrado and in the poorly explored Caatinga habitats of Brazil suggests that increasing the search for cryptic and inconspicuous species will lead to discoveries that will fundamentally alter our understanding of insect evolutionary history.
Acknowledgements

We thank the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and the Instituto Brasileiro do Meio Ambiente e dos Recursos Naturais Renováveis (IBAMA) for permission to conduct fieldwork in Brazil (permit numbers 14789–1, 14789–3). We are extremely grateful to two anonymous reviewers for providing thoughtful and constructive suggestions and comments. The manuscript is much improved as a result. Thanks to Dr. Yves P. Quinet and Francyregis Nunes for allowing us to study the only known specimen from the Caatinga and for the support to CRFB and RMF during field work in Crateús. We thank Dr. Rogerio R. Silva for calling our attention to the São Paulo specimens. We are indebted to Dr. Helena C. Morais (Universidade de Brasília), Dr. Karen Camargo (CNPq), and Jonas Maravalhas (UFU) for logistics during our work in Brasília and FAL and to Prof. José Mauro da Silva Diogo (FAL Director) for permission to work at FAL. We are thankful to Fundação Acangau for granting permission to HLV to collect at RPPN Acangau. We thank Scott Whittaker (NMNH) for help with preparation of the larvae, Ms. Jessica Louton and Dr. Natasha Mehdibadi (NMNH) for help with molecular work, Dr. Sean Brady (NMNH) for help with dating analyses, Ms Claudia Ortiz (Universidad Nacional de Colombia) for help with ArcGIS, and Ms. Eugenia Okonski (NMNH) for help with specimen preparation.

Author Contributions

Conceived and designed the experiments: JSC TRS CRFB CK RMF MB CTL CR. Contributed reagents/materials/analysis tools: JSC TRS CRFB CK RMF CR MB. Analyzed the data: JSC TRS CRFB CK RMF CR. Contributed reagents/materials/analysis tools: JSC TRS CRFB CK RMF CR MB CTL CR. Obtained permissions for collecting specimens: HLV MB CRFB.

References

1. Westwood JO (1833) On the probable number of species of insects in the creation; together with descriptions of several minute Hymenoptera. Mag of Nat Hist & J of Zoology, Bot, Mineralogy, Geology, and Meteorol 6: 116–123.
2. Stute B (2009) Finding new species: the Golden Age of discovery.
3. ISE (2011) State of Observed Species. Tempes, AZ: International Institute for Species Exploration.
4. Zompro O (2001) The Phasmatodea and Raptaphasmina n. gen., Orthoptera incertae sedis Baltic amber (Insecta: Orthoptera). Mitteilungen aus dem Geologisch-Paläontologischen Institut der Universität Hamburg 85: 229–261
5. Cameron SL, Barker SC, Whiting MF (2006) Mitochondrial genomics and the new insect order Mantophasmatodea. Mol Phylogenet Evol 38: 274–279. doi:10.1016/j.ympev.2005.09.020. PubMed: 16321547.
6. Klass K-D, Picker MD, Damgaard J, van Noort S, Tojo K (2003) The taxonomy, genitalic morphology, and phylogenetic relationships of Southern African Mantophasmatodea (Insecta). Entomol Abh 61: 3–67.
7. Klass K-D, Zompro O, Kristensen NP, Adis J (2002) Mantophasmatodea: a new insect order with extant members in the Afrotopics. Science 296: 1456–1459. doi:10.1126/science.1069397. PubMed: 11964441.
8. Picker MD, Colville JF, van Noort S (2002) Mantophasmatodea now in South Africa. Science 297: 1475. doi:10.1126/science.297.5586.1475a. PubMed: 12211240.
9. Arillo A, Engel MS (2006) Rock crawlers in Baltic amber (Notoptera: Mantophasmatodea). Am Museum Nov. 1–10.
10. Bebbere DP, Marriott FH, Gaston KJ, Harris SA, Scotland RW (2007) Predicting unknown species numbers using discovery curves. Proc R Soc Lond B 274: 1651–1658. doi:10.1098/rspb.2007.0464. PubMed: 17456460.
11. Bolton B (2013) AntCat. An online catalog of the ants of the world.
12. Ward PS (2007) Phylogeny, classification, and species–level taxonomy of ants (Hymenoptera: Formicidae). Zootaxa 1686: 549–563.
13. Holldobler B, Wilson EO (1990) The ants. Harvard University Press. 732 pp.
14. Ward PS (2010) Taxonomy, phylogenetics, and evolution. In: L Lach Cl ParKl ParrAbb. Ant Ecology. New York: Oxford University Press. pp. 3–17.
15. Dlussky GM, Brothers DJ, Rasnitsyn AP (2004) The first Late Cretaceous ants (Hymenoptera: Formicidae) from southern Africa, with comments on the origin of the Myrmicinae. Insect Syst Evol 35: 1–13. doi:10.1631/vj.2004.187631204789064727.
16. Dubovikov DA (2004) A new species of the genus Gesomyrmex Mayr, 1868 (Hymenoptera: Formicidae) from Vietnam. Tr Russkogo Entomologicheskogo Obshestva 75: 219–221.
17. Dubovikov DA (2004) Nebrothromyrmex majeri gen. et sp. n., a new genus and species of ants (Hymenoptera, Formicidae, Dolichoderinae) from Australia. Entomol Obzorzenie 83: 487–489.
18. Engel MS, Grimaldi DA (2005) Primitive new ants in Cretaceous amber from Myanmar, New Jersey, and Canada (Hymenoptera: Formicidae). Am Museum Nov. 1–24.
19. Fernandez F (2003) A new myrmicine ant genus from Malaysia with uncertain affinities (Hymenoptera: Formicidae). Zootaxa 341: 1–6.
20. Fernandez F (2004) Adelomyrmecini new tribe and Cryptomyrmex new genus of myrmicine ants (Hymenoptera: Formicidae). Sociobiology 44: 325–336.
21. Fisher BL (2006) Boloopenhagenia vicans gen. n. and sp. n. and two new species of the Electroterena genus group (Hymenoptera: Formicidae). Myrmecological Nachrichten 8: 111–118.
22. Bolton B, Fisher BL (2008) Afrotropical ants of the ponerine genera Centromyrmex Mayr, Promyopias Santachi gen. rev. and Feroponea gen. n., with a revised key to genera of African Ponereinae (Hymenoptera: Formicidae). Zootaxa 1929: 1–37.
23. Bolton B, Fisher BL (2012) Taxonomy of the cerapachyine ant genera Simopone Forel, Vicinopone gen. n. and Taniopone gen. n. (Hymenoptera: Formicidae). Zootaxa 3263: 1–101.
24. Cover S, Deyrup M, Station AB (2007) A new ant genus from the southwestern United States 80. Memoirs of the American Entomological Institute. pp. 89–99.
25. Fisher BL (2009) Two new dolichoderine ant genera from Madagascar: Aptonoma gen. n. and Ravavy gen. n. (Hymenoptera: Formicidae). Zootaxa 2118: 37–52.
26. Wild AL, Cuezzo F (2006) Rediscovery of a fossil dolichoderine ant lineage (Hymenoptera: Formicidae: Dolichoderinae) and a description of a new genus from South America. Zootaxa 1142: 137–145. doi:10.11603/2005.09.020. PubMed: 16321547.
27. Rabeling C, Brown JM, Verhaagh M (2008) Newly discovered sister lineage sheds light on early ant evolution. Proc Natl Acad Sci U S A 105: 14913–14917. doi:10.1073/pnas.0806187105. PubMed: 18794530.
28. Braantstickter MG (2009) The ant genus Stenamma Westwood (Hymenoptera: Formicidae) redefined, with a description of a new genus Propodilobus. Zootaxa: 41–57.
29. Fernandez F, Delabie JHC, Do Nascimento IC (2009) Diaphoromyrma, a new myrmicine ant genus (Hymenoptera: Formicidae) from North Eastern Brazil. Zootaxa: 55–62.
30. Klingenberg C, Brandão CRF (2009) Revision of the fungus-growing ant genera Mycetophylax Emery and Paracyctophylax Kusnezov rev. stat., and description of Kalathomyrmex n. gen. (Formicidae: Myrmicinae: Attini). Zootaxa 2052: 1–31.
31. Terayama M (2009) A synopsis of the family Formicidae of Taiwan (Insecta, Hymenoptera). The Research Bulletin of Kanto Gakuen University 17: 81–266.
32. Lapolla JS, Kallal RJ, Brady SG (2012) A new ant genus from the Greater Antilles and Central America, Zatania (Hymenoptera: Formicidae), exemplifies the utility of male and molecular character systems. Syst Entomol 37: 200–214. doi:10.1111/j.1365-3113.2011.00605.x.
33. Archibald SB, Johnson KR, Mathewes RW, Greenwood DR (2011) Intercontinental dispersal of giant thermophilic ants across the Arctic during early Eocene hyperthermals. Proc R Soc Lond B 278: 3679–3686. doi:10.1098/rspb.2011.0729. PubMed: 21543354.
34. Silva RR, Feltoas RM, Brandão CRF, Diniz JL (2009) Tropidomyrmex \textit{alveus}, a new myrmicine ant genus and species from Brazil, tentatively assigned to Solenopsisidini (Hymenoptera, Formicidae). Zootaxa 2052: 32–48.
35. Yamane S, Bui TV, Eguchi K (2008) Oparomyra hanguvung, a new genus and species of ant related to Apomyrma (Hymenoptera: Formicidae: Amblyoponini). Zootaxa 1767: 55–63.
36. Barden P, Grimaldi D (2013) A new genus of highly specialized ants in Cretaceous Burmese amber (Hymenoptera: Formicidae). Zootaxa 3681: 405–412. doi:10.11646/zootaxa.3681.4.5.
37. Weber NA (1958) Evolution in fungus-growing ants. Proceedings of the Tenth International Congress of Entomology 2: pp. 459–473.
38. Weber NA (1966) Fungus-growing ants. Science 153: 587–604. doi:10.1126/science.153.3736.587. PubMed: 17757227.
39. Wheeler WM (1907) The fungus-growing ants of North America. Bull Am Museum Nat Hist 23: 669–807.
40. Weber NA (1945) The biology of the fungus-growing ants. Part 8. The Trinidad, B.W.I. species. Rev de Entomol 16: 1–88.

41. Chapela IH, Rehner SA, Schultz TR, Mueller UG (1994) Evolutionary history of the symbiosis between fungus-growing ants and their fungi. Science 266: 1691–1694. doi:10.1126/science.266.5191.1691. PubMed: 17775630.
42. Currie CR, Bot AN, Boomsma JJ (2003) Experimental evidence of a tripartite mutualism: bacteria protect ant fungus gardens from specialized parasitoids. Oikos 101: 91–102. doi:10.1034/j.1600-0706.2003.12036.x.
43. Currie CR, Mueller UG, Malloch D (1999) The biological pathology of ant fungus gardens. Proc Natl Acad Sci U S A 96: 7988–8002. doi:10.1073/pnas.96.14.7998. PubMed: 10393936.
44. Currie CR, Wong B, Stuart AE, Schultz TR, Rehner SA et al. (2003) Ancient tripartite coevolution in the attine ant-microbe symbiosis. Science 299: 386–389. doi:10.1126/science.1078155. PubMed: 12532015.
45. Green AM, Mueller UG, Adams RM (2002) Extensive exchange of fungal cultivars between sympatric species of fungus-growing ants. Mol Ecol 11: 191–195. doi:10.1046/j.1365-294X.2002.01433.x. PubMed: 11856421.
46. Hinkle G, Wetterer JK, Schultz TR, Sogin ML (1994) Phylogeny of the attine ant fungi based on analysis of small subunit ribosomal RNA gene sequences. Science 266: 1695–1697. doi:10.1126/science.7992052. PubMed: 7992052.
47. Mueller UG (2002) Ant versus fungus versus mutualism: ant-cultivar conflict and the deconstruction of the attine ant-fungus symbiosis. Am Nat 160: S67–S98. doi:10.1086/342084. PubMed: 18707454.
48. Mueller UG, Rehner SA, Schultz TR (1998) The evolution of agriculture in ants. Science 281: 2034–2038. doi:10.1126/science.281.5385.2034. PubMed: 9748164.
49. Munkaci A, Pan JJ, Villesen P, Mueller UG, Blackwell M et al. (2004) Convergent coevolution in the domestication of coral mushrooms by fungus-growing ants. Proc R Soc Lond B 271: 1777–1782. doi:10.1098/rspb.2004.2759.
50. Villegas P, Mueller UG, Schultz TR, Adams RM, Bouck AC (2004) Evolution of plant-cultivar specialization and cultivar-switching in \textit{Apterostigma} fungus-growing ants. Evolution 58: 2252–2265. doi:10.1554/03-203. PubMed: 15562688.
51. Cáforo MJ, Currie CR (2005) Phylogenetic analysis of mutualistic filamentous bacteria associated with fungus-growing ants. Can J Microbiol 51: 441–446. doi:10.1139/w05-023. PubMed: 16121221.
52. Gerardo NM, Jacobs SR, Currie CR, Mueller UG (2006) Ant genetic associations maintained by specificity of chemotaxis and diversity. Annu Rev Entomol 51: 177–190. doi:10.1146/annurev-ento-120811-153710. PubMed: 1707513.
53. Autuori M (1941) Contribución para el conocimiento de la saúva (\textit{Acromyrmex spinigerus}) I: Evolución del saúveiro (\textit{Acromyrmex spinigerus}) I. Arq Instituto Biológico 12: 197–233.
54. Bell T (1874) The naturalist in Nicaragua. John Murray. 403 pp.
55. Autuori M (1942) Contribuição para o conhecimento da saúva (\textit{Atta cephalotes}). III. Escavação de um saúveiro (\textit{Acromyrmex spinigerus}) I. Arq Instituto Biológico 13: 137–149.
56. Blanton C, Ewel J (1985) Leaf-cutting ant herbivory in successional and agricultural tropical ecosystems. Ecology 66: 861–869. doi:10.2307/1904548.
57. Barnett JM, Peregrine DJ (1976) A review of the status of leaf-cutting ants and their control. Ann Appl Biol 84: 124–128. doi:10.1111/j.1744-7348.1976.tb01741.x.
58. Fowler HG (1979) The larger hornetiforms of the Paraguay of the géneros \textit{Atta} Fabrícios and \textit{Acromyrmex} Mayr: bionomía, distribución y sistemática. Informes Científicos. Asunción: Universidad Nacional de Asunción 2: 30–70.
59. Delabie J, Nascimento Id Fonseca Ed; Sgrillo RB, Soares P et al. (1997) Biogeography of leaf-cutting ants (Hymenoptera; Formicidae; Myrmicinae; \textit{Atta}) of the Amazon: a historical, biogeographical and ecological analysis. Bees 30: 17–24.
60. Hoelldobler B, Wilson EO (1985) Nest area expansion and recognition in leafcutter ants (\textit{Atta cephalotes}). J Insect Physiol 31: 143–150. doi:10.1016/0022-1910(86)90133-2.
61. Hoelldobler B, Wilson EO (2010) The leafcutter ants: civilization by agriculture. Belknap Press.
62. Wirth R, Meyer ST, Almeida WR, Araujo MV, Barbosa VS et al. (2007) Increasing densities of leaf-cutting ants (\textit{Atta spp.}) with proximity to the edge in a Brazilian Atlantic forest. J Trop Ecol 23: 501–505. doi:10.1017/S0266467407004221.
83. Lopes BC (2005) Recursos vegetais usados por Acromyrmex striatus (Roger) (Hymenoptera, Formicidae) em restinga da Praia da Joaquina, Florianópolis, Santa Catarina, Brasil. Rev Bras Zool 22: 372–382. doi: 10.1590/S1019-71702005000200011.

84. Varón EH, Eigenbrode SD, Bosque Pérez NA, Hilje L (2007) Effect of farm diversity on harvesting of coffee leaves by the leaf-cutting ant Atta cephalotes. Agric Forest Entomol 9: 47–55. doi: 10.1111/j.1461-9563.2006.00320.x.

85. Wirth R, Heiz H, Ryal RJ, Beyschlag W, Hölldobler B (2003) Herbivory of leaf-cutting ants: a case study on Atta colombica in the tropical rainforest of Panama. Berlin: Springer-Verlag.

86. Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GA, Kent J (2000) Biodiversity hotspots for conservation priorities. Nature 403: 853–858. doi:10.1038/35020501. PubMed: 10708275.

87. Lopes SF, Schiavinai I, Oliveira AP, Vale VS (2011) An Ecological Comparison of Floristic Composition in Seasonal Semideciduous Forest in Southeast Brazil: Implications for Conservation. Int J Forestry Res 2012.

88. Gauld I, Bolton B (1988) The Hymenoptera. Oxford University Press in association with British Museum (Natural History). 332 p.

89. Rabeling C, Cover S, Johnson R, Mueller UG (2007) A review of the North American species of the fungus-gardening ant genus Trachymyrmex (Hymenoptera: Formicidae). Zootaxa 1664: 1–53.

90. Serna F, Mackay W (2010) A descriptive morphology of the ant genus Procyrtocerus (Hymenoptera: Formicidae). J Insect Sci (Online) 10: 111.

91. Sosa-Calvo J, Schultz TR (2010) Three remarkable new fungus-growing ant species of the genus Myrmicocrypta (Hymenoptera: Formicidae), with a reassessment of the characters that define the genus and its position within the Attini. Ann Entomol Soc Am 103: 181–195. doi:10.1603/AN09108.

92. Schultz TR, Meier R (1995) A phylogenetic analysis of the fungus-growing ants (Hymenoptera: Formicidae, Attini) based on morphological characters of the larvae. Syst Entomol 20: 337–370. doi: 10.1111/j.1365-3113.1995.tb01000.x.

93. Rabeling C, Verhaagh M, Engels W (2007) Comparative study of nest architecture and colony structure of the fungus-growing ant, Mycocepurus smithii (Hymenoptera, Formicidae). J Nat Hist 39: 1735–1743. doi:10.1080/00222930400027462.

94. Rabeling C, Bacci M (2010) A new workerless inquiline in the Lower Attini (Hymenoptera: Formicidae), with a discussion of social parasitism in fungus-growing ants. J Insect Sci (Online) 10: 129–138. doi:10.1093/sysbio/syq012, PubMed: 20525640.

95. Soares AERA, Schrago CGC (2012) The influence of taxon sampling and tree shape on molecular dating: an empirical example from Mammalian mitochondrial genomes. Bioinformatics BIol Insights 6: 129–143. PubMed: 22693422.

96. Brandão CRF, Mayhè-Nunes AJ (2008) A new species of the fungus-farming ant genus Mycetagoecrus Brandão & Mayhè-Nunes (Hymenoptera, Formicidae, Attini), Rev Bras Entomol 52: 349–352.

97. Penick CA, Copple RN, Mendez RA, Smith AA (2012) The role of anchor-tipped larval hairs in the organization of ant colonies. PLOS ONE 7: e41595. doi: 10.1371/journal.pone.0041595. PubMed: 22848539.

98. Guénavard B, Weiser MD, Dunn RR (2012) Global models of ant diversity suggest regions where new discoveries are most likely are under disproportionate deforestation threat. Proc Natl Acad Sci USA 109: 7368–7373. doi: 10.1073/pnas.1113867109. PubMed: 22523955.

99. Silva JD; Tabarelli M, Fonseca, Lins (2004) A biodiversidade da Caatinga: áreas e ações prioritárias para a conservação. DF: Brasília. p. 382.