Prognostic value of systemic inflammation response index in nasopharyngeal carcinoma with negative Epstein-Barr virus DNA

Xiaofei Yuan1†, Hua Yang1†, Fangfang Zeng1†, Shiyu Zhou2, Shuting Wu1, Yue Yuan1, Linchong Cui1, Huiru Feng1, Danfan Lin1, Zilu Chen1, Xiong Liu1*, Jing Chen3* and Fan Wang1*

Abstract

Background: Inflammatory parameters and Epstein–Barr virus (EBV) DNA status have been confirmed to be associated with prognosis in nasopharyngeal carcinoma (NPC) patients. However, there are few in-depth studies on the prognosis of NPC patients with negative EBV DNA. Our study aimed to look for inflammatory biomarkers that can identify disease progression in NPC patients with negative EBV DNA.

Methods: A total of 795 NPC patients were recruited, and ultimately 325 NPC patients with negative EBV DNA were included in this study (170 in training cohort and 155 in validation cohort). Kaplan–Meier method and log-rank test were used to analyze progression-free survival (PFS) and overall survival (OS). The multivariate analysis of Cox proportional hazards regression model was used to determine the independent prognostic factors. Receiver operating characteristic (ROC) curves were used to assess prognostic value. The logistic regression was used to evaluate the relationship between EBV DNA status and inflammatory parameters. The correlation between clinical characteristics was analyzed by the chi-squared test or the Fisher’s exact test.

Results: The optimal cutoff point for the SIRI was 1.12. The EBV DNA-negative NPC patients with high SIRI level had worse PFS and OS (all \(p < 0.001\)). In multivariate Cox proportional hazard models analysis, SIRI was an independent prognostic factor for PFS and OS (all \(p < 0.05\)), and had higher prognostic value than other indicators. Above results were found in the training cohort and confirmed in the validation cohort. In addition, EBV DNA status was not associated with any inflammatory parameters.

Conclusions: The SIRI can provide more accurate risk stratification and better prognostic prediction for NPC patients with negative EBV DNA.

Keywords: Nasopharyngeal carcinoma, Systemic inflammation response index, Epstein–Barr virus, Prognosis, Risk stratification

Introduction

Nasopharyngeal carcinoma (NPC) is an epithelial carcinoma originating from the nasopharyngeal mucosa that is apparently different from other head and neck cancers [1]. NPC has extremely unbalanced geographical global distributions and is prevalent in Southeast Asia, especially in Southern China [2]. Since NPC is sensitive to chemoradiation, the main treatment for NPC is...
intensity-modulated radiation therapy (IMRT) combined with or without chemotherapy [3]. However, approximately 30% of patients still have a poor survival rate due to local recurrence and distant metastasis [3, 4]. Risk stratification through reliable predictors is necessary for individualized treatment which has important clinical value for better prognosis and improved survival.

Epstein–Barr virus (EBV) infection has been regarded as one of the risk factors contributing to the development of NPC [5]. The detection of plasma EBV DNA has been proven to be a reliable biomarker for population screening, prognosis, precise treatment and posttreatment monitoring of NPC patients [6–9]. High plasma EBV DNA load can reflect advanced TNM classification, tumor burden and residual disease of NPC patients [10–12]. Many previous studies have demonstrated that plasma EBV DNA is closely related to the prognosis of NPC patients [7, 13, 14]. About 40% of NPC patients are negative for EBV quantification before treatment. The prognosis of EBV DNA-positive patients is much worse than that of EBV DNA-negative patients. However, some NPC patients with negative EBV DNA still have poor survival. According to our previous study [15], the 5-year PFS and OS of NPC patients with low EBV DNA level were approximately 80% and 90%, respectively. Therefore, looking for biomarkers that can identify disease progression in NPC patients with negative EBV DNA may bring prognosis benefits for these patients.

Systemic inflammation is one of the basic components of the tumor microenvironment and can affect the pathogenesis of cancer patients [16]. Inflammation and immune surveillance have been recently established as cancer hallmarks, showing an essential role in the progression, recurrence and metastasis of cancer [1]. In recent years, immune-inflammation indexes including the systemic immune-inflammation index (SII) based on three leukocytes (peripheral blood neutrophils, platelets and lymphocytes) and the systemic inflammation response index (SIRI) based on three leukocytes (peripheral blood neutrophils, monocytes and lymphocytes) were investigated in various cancers. In the study by Chen et al. [17], high SII levels were associated with worse overall survival (OS) and disease-free survival (DFS) in colorectal cancer. A previous study was followed by Zeng et al. [18], which reported that pretreatment SII in NPC patients were significantly higher than those in patients with chronic rhinitis, and could be used as prognostic indicators in NPC patients. A retrospective study involving 177 patients with pancreatic cancer after chemotherapy in a training cohort, showed that a high pretreatment SIRI was significantly related to poor time to progression (TTP) and OS, which was verified in the validation cohort [19]. Chen et al. [20] first demonstrated that SIRI was an important prognostic biomarker for patients with NPC in 2018, and an increase in the SIRI was associated with worse OS. As mentioned earlier, some NPC patients with negative EBV DNA also have recurrence and metastasis. In view of the important role of the inflammatory indicators in cancer prognosis, we tried to apply these indicators to NPC patients with negative EBV DNA to observe whether they can effectively distinguish high-risk patients.

Therefore, we conducted this retrospective study to evaluate the prognostic value of inflammatory parameters in NPC patients with negative EBV DNA and to validate all results. This study aimed to strengthen the survival prediction, risk stratification and precise treatment of such patients.

Materials and methods

Case selection

Between January 2005 and December 2015, we retrospectively recruited 795 patients who were diagnosed with NPC at Nanfang Hospital of Southern Medical University. All patients were confirmed by pathological examinations and staged according to the eighth edition of the American Joint Committee on Cancer (AJCC) staging system. This retrospective study was approved by the Ethics Committee of Nanfang Hospital of Southern Medical University (Ethical review approval no.: NFEC-2017–165).

Inclusion and exclusion criteria

The inclusion criteria in this study were as follows: (a) patients with NPC confirmed by histopathology; (b) patient with complete medical treatment time records and history records; (c) patients with at least one complete record of a peripheral blood count before treatment; and (d) patients with at least one EBV DNA test before treatment. The exclusion criteria were as follows: (a) patients with prior malignancy; (b) patients with a history of previous antineoplastic therapy; (c) patients with non-WHO pathological types; (d) patients with uncontrolled infection; or (e) patients with positive EBV DNA.

Finally, a total of 325 NPC patients with negative EBV DNA were included. All these patients were divided into the training cohort and validation cohort by random number.

Hematological examination

The peripheral blood cells of all patients were collected and tested for leukocyte, neutrophil, lymphocyte, monocyte, and platelet counts within 1 week before therapy. The measurements of plasma EBV DNA were performed within 1 month before therapy. The SII was defined as (neutrophil*platelet)/lymphocyte; the SIRI is defined as
(neutrophil*monocyte)/lymphocyte. All peripheral blood cell and EBV DNA assessments were performed in the Laboratory Medicine Center of Nanfang Hospital, Southern Medical University according to standard operating procedures. The cutoff level chosen to classify the patients into the negative and positive EBV DNA groups was 500 copies/mL before treatment, referring to the threshold of Laboratory Medicine Center, Nanfang Hospital, Southern Medical University, in this study (The detection method for EBV DNA in Supplementary file 1.).

**Treatment with guideline**

All patients developed treatment plans according to the then latest NCCN (National Comprehensive Cancer Network) guidelines. The general principles of treatments were as follows: all patients received IMRT, patients with stage I received IMRT alone, patients with stage II received concurrent chemoradiotherapy, patients with stage III/IV received concurrent chemoradiotherapy, induction chemotherapy, and/or adjuvant chemotherapy.

The general principles of radiation therapy were as follows: all patients were treated with 2.12–2.24 Gy each time, 5 times a week, with IMRT for 6–8 weeks. The total prescribed IMRT doses were 70–74 Gy to the gross tumor volume of the nasopharynx (GTVn), 66–70 Gy to the positive neck lymph node area (GTVnd), 60–62 Gy to the high-risk sites defined as clinical target volume (CTV1), and 50–56 Gy to the low-risk sites defined as clinical target volume (CTV2).

The principles of chemotherapy were as follows: induction chemotherapy (IC) (1–2 cycles) and adjuvant chemotherapy (AC) (1–4 cycles) included TP, TPF, and PF. The TP regimen was paclitaxel 135 mg/m²/d or docetaxel 60 mg/m²/d on day 1 and cisplatin 25 mg/m²/d on days 1 to 3, the TPF regimen was paclitaxel 135 mg/m²/d or docetaxel 60 mg/m²/d on day 1, cisplatin 25 mg/m²/d on days 1 to 3, and 5-fluorouracil 600 mg/m²/d on days 1 to 5, and the PF regimen was cisplatin 25 mg/m²/d on days 1 to 3 and 5-fluorouracil 600 mg/m²/d on days 1 to 5. The concurrent chemoradiotherapy (CCRT) (1–2 cycles) included cisplatin monotherapy and TP. The cisplatin monotherapy regimen was cisplatin 25 mg/m²/d on days 1 to 3, and the TP regimen was paclitaxel 135 mg/m²/d or docetaxel 60 mg/m²/d on day 1 and cisplatin 25 mg/m²/d on days 1 to 3. All chemotherapy regimens were administered every 21 days as a complete cycle.

**Follow-up and endpoint**

The follow-up schedule was conducted strictly in accordance with the following 3, 6 and 12 months in the first year after therapy, every 6 months in the second and third years, and once a year thereafter. Each follow-up

---

**Fig. 1** X-tile analysis of survival data of NPC patients. a The optimal cut-off value for the SII was 655.54 (chi square = 18.922, p < 0.001); b The optimal cut-off value for the SIRI was 1.12 (chi square = 23.425, p < 0.001)
Evaluation included head and neck physical examination, abdominal ultrasound, nasopharyngeal endoscopy, chest radiograph, peripheral blood examination, whole-body positron emission tomography-computed tomography (PET-CT) or emission computed tomography (ECT), and nasopharyngeal and neck magnetic resonance imaging (MRI). For cases with suspected recurrence of nasopharyngeal neck tumor or distant metastasis of cervical lymph nodes, biopsy or needle biopsy was performed on the suspected site to make a definitive diagnosis. The primary outcomes of this study were progression-free survival (PFS) and overall survival (OS). PFS was defined as the time from the initial pathological diagnosis of NPC to the date of disease progression or death from any cause. OS was defined as the time between the initial pathological diagnosis of NPC and all-cause death, or at the last follow-up.

**Statistical analysis**

The optimal cutoff values of the SII and SIRI were determined using X-tile 3.6.1 software (Robert L Camp, Yale University, New Haven, CT, USA) [21, 22]. The cutoff values were plotted by X-tile 3.6.1 software, and other figures were plotted by GraphPad Prism V8.0. The chi-squared test and the Fisher’s exact test were used to explore the relationship between clinical characteristics and different variables. Logistic regression was used to estimate the odds ratio (OR) and 95%CI to evaluate the association between inflammatory parameters and EBV DNA status (negative vs. positive). Survival curves were analyzed using the Kaplan–Meier method and compared using the log-rank test. Univariate and multivariate Cox proportional hazards regressions were conducted to evaluate the prognostic significance of each variable with respect to PFS and OS. ROC curve analysis was performed to compare the different prognostic values.

### Table 1

| Variables      | Training cohort (n = 170) | Validation cohort (n = 155) |
|----------------|--------------------------|-----------------------------|
|                | No.(%)                   | No.(%)                      |
| Gender         |                          |                             |
| male           | 112 (65.9)               | 117 (75.5)                  |
| female         | 58 (34.1)                | 38 (24.5)                   |
| Age ≤ 55       | 134 (78.8)               | 126 (81.3)                  |
| Age > 55       | 36 (21.2)                | 29 (18.7)                   |
| Smoke no       | 113 (66.5)               | 89 (57.4)                   |
| Smoke yes      | 57 (33.5)                | 66 (42.6)                   |
| AJCC stage (8th) |                        |                             |
| I              | 14 (8.2)                 | 23 (14.8)                   |
| II             | 37 (21.8)                | 25 (16.1)                   |
| III            | 44 (25.9)                | 50 (32.3)                   |
| IVa            | 69 (40.6)                | 55 (35.5)                   |
| IVb            | 6 (3.5)                  | 2 (1.3)                     |
| Tumor classification |                     |                             |
| T1             | 36 (21.2)                | 48 (31.0)                   |
| T2             | 37 (21.8)                | 36 (23.2)                   |
| T3             | 29 (17.1)                | 26 (16.8)                   |
| T4             | 68 (39.9)                | 45 (29.0)                   |
| Node classification |                   |                             |
| N0             | 30 (17.6)                | 39 (25.2)                   |
| N1             | 59 (34.7)                | 40 (25.8)                   |
| N2             | 75 (44.1)                | 63 (40.6)                   |
| N3             | 6 (3.5)                  | 13 (8.4)                    |
| Metastasis     |                          |                             |
| Non-metastasis | 164 (96.5)               | 153 (98.7)                  |
| Metastasis     | 6 (3.5)                  | 2 (1.3)                     |
| WHO pathologic type |                   |                             |
| TypeI          | 0 (0)                    | 1 (0.6)                     |
| TypeII         | 15 (8.8)                 | 17 (11.0)                   |
| TypeIII        | 155 (91.2)               | 137 (88.4)                  |
| Leukocytes     |                          |                             |
| ≤ 9.5          | 159 (93.5)               | 140 (90.3)                  |
| > 9.5          | 11 (6.5)                 | 15 (9.7)                    |
| Neutrophils    |                          |                             |
| ≤ 6.3          | 157 (92.4)               | 138 (94.8)                  |
| > 6.3          | 13 (7.6)                 | 17 (11.0)                   |
| Lymphocytes    |                          |                             |
| ≤ 3.2          | 164 (96.5)               | 147 (94.8)                  |
| > 3.2          | 6 (3.5)                  | 8 (5.2)                     |
| Monocytes      |                          |                             |
| ≤ 0.6          | 151 (88.8)               | 137 (88.4)                  |
| > 0.6          | 19 (11.2)                | 18 (11.6)                   |
| Platelets      |                          |                             |
| ≤ 300          | 143 (84.1)               | 132 (85.2)                  |

| EBV DNA Epstein-Barr virus DNA, AJCC American Joint Committee on Cancer, WHO World Health Organization, SII Systemic immune-inflammation index, SIRI Systemic inflammation response index |
All variables reaching statistical significance in univariate analysis were included in multivariate Cox proportional hazards regression analysis. A two-tailed p value of less than 0.05 was considered statistically significant. All results were validated by the validation cohort. All statistical analyses were performed using the SPSS (Statistical Packages for Social Science) version 23.0 (IBM, Corporation).

Results
The optimal cutoff values of the SII, SIRI and the peripheral blood cells
The X-tile 3.6.1 software was used to evaluate the optimal cutoff values of the SII and SIRI for progression outcome in training cohort. The analysis demonstrated that the optimal cutoff values of the SII and SIRI were 655.54 (p < 0.001) and 1.12 (p < 0.001), respectively (Fig. 1). The optimal cutoff values of other peripheral blood cells were determined as the upper limits of the normal values as defined by the Laboratory Medicine Center of Nanfang Hospital, Southern Medical University (leukocytes: 9.5*10⁹/L; neutrophil: 6.3*10⁹/L; lymphocyte: 3.2*10⁹/L; monocyte: 0.6*10⁹/L; platelet: 300*10⁹/L).

Patients characteristics and follow-up
The clinical characteristics of the 170 NPC patients in training cohort and 155 NPC patients in validation cohort were listed in Table 1. In training cohort, 112 (65.9%) were male and 58 (34.1%) were female, the median age was 47 years old, the median follow-up duration was 62.0 months. During the long-term follow-up, 39 (22.9%) patients
experienced disease progression, 23 (13.5%) patients experienced locoregional relapse, 14 (8.2%) patients experienced distant metastasis and 20 (11.8%) patients died. The 5-year PFS and OS rates were 78.8% and 89.4%, respectively. In validation cohort, 117 (75.5%) were male and 38 (24.5%) were female, the median age was 46 years old, the median follow-up duration was 60.0 months. During the long-term follow-up, 31 (20.0%) patients experienced disease progression, 10 (6.5%) patients experienced locoregional relapse, 21 (13.5%) patients experienced distant metastasis and 20 (12.9%) patients died. The 5-year PFS and OS rates were 80.6% and 87.7%, respectively.

Prognostic significance of clinical features and inflammatory parameters
The Kaplan–Meier survival curves for survival analyses showed that both higher SII group and higher SIRI group were significantly associated to worse PFS and worse OS in training cohort (all $p<0.01$) (Fig. 2). In validation cohort, only the higher SIRI group was significantly associated to worse PFS ($p<0.001$) and worse OS ($p<0.001$) (Fig. 3).

In the univariate Cox regression model, platelets ($p=0.001$), SII ($p<0.001$) and SIRI ($p<0.001$) were significantly associated with PFS in training cohort. The age ($p=0.038$), SII ($p=0.007$) and SIRI ($p<0.001$) were significantly associated with OS in training cohort (Table 2). The age, leukocytes, neutrophils, monocytes and SIRI were significantly associated with PFS and OS in validation cohort (all $p<0.01$) (Table 3). All variables reaching statistical significance in univariate analysis were included in multivariate Cox proportional hazards regression analysis.
In the multivariate Cox regression model, only SIRI were associated with PFS and OS in training cohort (all \( p < 0.05 \)) (Table 2). In validation cohort, the age and SIRI were associated with PFS and OS (all \( p < 0.05 \)) (Table 3). Only SIRI remained an independent prognostic factor in both training cohort and validation cohort.

### ROC curves analysis

Since in the multivariate analysis of validation cohort, both age and SIRI could be considered as independent prognostic factors in NPC patients with negative EBV DNA, we included age and SIRI in the ROC curves to compare their prognostic value. The results demonstrated that the area under the curve (AUC) values of SIRI larger than the AUC values of age for PFS and OS in validation cohort. Although the age was not an independent prognostic for NPC patients with negative EBV DNA in training cohort, we finally included age and SIRI for the ROC analysis in training cohort, and the final results were consistent with the validation cohort (Fig. 4).

### Relationship between clinical characteristics and SIRI

The SIRI levels were significantly correlated with AJCC stage and tumor classification in training cohort (all \( p < 0.05 \)).
However, these results were not confirmed in validation cohort. The SIRI levels were significantly correlated with gender and smoke in validation cohort (all \( p < 0.05 \)) (Table 4).

### Relationship between EBV DNA status and inflammatory parameters or clinical characteristics in all 795 NPC patients

To further confirm that SIRI could be used as a good prognostic indicator for NPC patients with negative EBV DNA, we analyzed the correlation between EBV DNA status and inflammatory parameters. In the overall population \((n = 795)\), EBV DNA status was not associated with any inflammatory parameters (Supplement Table 1). However, EBV DNA status was significantly associated with AJCC stage \((p < 0.001)\), Tumor classification \((p < 0.001)\) and Node classification \((p < 0.001)\), the NPC patients with positive EBV DNA had more advanced stages (Supplement Table 2).

### Discussion

Multiple reports have shown that pretreatment negative EBV DNA patients have better survival and prognosis than patients with high plasma EBV DNA levels [7, 13, 14]. However, many patients with negative EBV
DNA still have disease progression, recurrence and metastasis, and the prognostic factors of these patients have rarely been studied. Our findings demonstrated that the SIRI had prognostic value in NPC patients with negative EBV DNA and that high SIRI was associated with worse PFS and OS in these patients.

In this study, we found that high SIRI level was closely related to worse 5-year PFS and OS both in in training cohort and validation cohort, these findings were consistent with previous studies [19, 20]. Currently, inflammation is considered a hallmark of cancer and has the ability to activate invasion and metastasis, induce angiogenesis, and sustain proliferative signaling by many scholars [16, 23]. But, there is still no accurate explanation for the mechanism related to the prognosis of SIRI in cancer patients. For now, the prognostic role of SIRI can be explained by the function of its own components. The neutrophils can release a variety of cytokines, such as interleukin-10 (IL-10) and transforming growth factor-β (TGF-β), to inhibit T cell proliferation and activation [24]. Lymphocytes are often able to inhibit tumor proliferation and metastasis, and kill tumor cells through their immune function (secrete interferon gamma (INF-γ) and tumor necrosis factor (TNF-α)) [25]. Monocytes and monocyte-derived M2 macrophages play important roles in tumor growth, invasion and suppression of antitumor immunity and dissemination [26]. In addition, the number of monocytes and tumor burden are often closely related [27]. However, there was no prognostic value of SII in NPC patients with negative EBV DNA. The reason for this result may be due to the different functions between platelets and monocytes in cancer. Platelets are closely related to cancer metastasis, and the mechanism for this phenomenon is that platelets support cancer cell migration and metastasis formation by stimulating and aiding cancer cell adhesion and extravasation [28]. However, patients with negative EBV DNA have far fewer distant metastases than patients with positive EBV DNA, so the SII could not achieve a good predictive effect in negative EBV DNA patients. The function of monocytes not only promotes distant metastasis of cancer cells, but also plays an important role in inducing tumor resistance, mediating immune escape, and promoting neovascularization [29]. Therefore, this also explains why only SIRI has prognostic value in NPC patients with negative EBV DNA.

![Fig. 4](https://example.com/roc_curves.png)

**Fig. 4** ROC curves analysis for comparing the prognostic potential of independent prognostic factors. a Prediction of PFS in training cohort; b Prediction of OS in training cohort; c Prediction of PFS in validation cohort; d Prediction of OS in validation cohort
To further confirm inflammatory parameters could be considered as possible prognostic factors for NPC patients with negative EBV DNA, we conducted a correlation analysis for EBV DNA status and clinical characteristics or inflammatory parameters in all patients. We found that NPC patients with positive EBV DNA had advanced AJCC stage, Tumor classification and Node classification. This conclusion was consistent with Leung et al. [10] and Peng et al. [12], who found that EBV DNA levels were related to AJCC stage. In addition, EBV DNA load was significantly correlated with both circulating tumor cells (CTCs) and tumor burden [11, 30], which explained why NPC patients with positive EBV DNA had a worse prognosis. But in our study, EBV DNA status was not related to peripheral blood inflammatory cells and inflammatory parameters, which meant that EBV infection did not affect the change in systemic inflammation. Current studies have shown that peripheral plasma EBV DNA comes from tumor cells, and EBV infection usually changes local inflammation around the tumor tissue [31], but the systemic inflammatory changes in EBV-infected patients are not obvious. Therefore, we considered that SIRI was a prognostic factor not affected by EBV infection and had good prognostic value for NPC patients with negative EBV DNA, so it was more suitable as a prognostic factor for these patients.

Due to the SIRI was an independent prognostic factor in NPC patients with negative EBV DNA, we analyzed the correlation between SIRI and other clinical characteristics. The SIRI levels were correlated with AJCC stage and tumor classification in training cohort. However, these results were not confirmed in validation cohort. Therefore, the conclusions of this part still need a large sample of data for further verification.

Since both age and SIRI were independent prognostic factors for NPC patients with negative EBV DNA in validation cohort, we further evaluated the predictive value of age and SIRI by ROC curves. We found that the prognostic value of SIRI was greater than that of age in both training cohort and validation cohort. This confirmed that the SIRI in NPC patients with negative EBV DNA could serve as a strong independent prognostic factor. In previous study, scholars have always combined the pretreatment plasma EBV DNA with clinical features or other biomarkers such as age, sex, T classification, N classification, body mass index (BMI), lactate dehydrogenase.

| Variables                  | Training cohort (n = 170) | Validation cohort (n = 155) |
|----------------------------|--------------------------|----------------------------|
|                            | SIRI                     |                            | SIRI                     |                            |
|                            | ≤ 1.12                   | > 1.12                    | ≤ 1.12                   | > 1.12                    |
| Gender, No. (%)            |                          |                            |                          |                            |
| male                       | 81 (64.8)                | 31 (68.9)                 | 78 (70.9)                | 39 (86.7)                 |
| female                     | 44 (35.2)                | 14 (31.3)                 | 32 (29.1)                | 6 (13.3)                  |
| Age, No. (%)               |                          |                            |                          |                            |
| ≤ 55                       | 101 (80.8)               | 33 (73.3)                 | 88 (80.0)                | 38 (84.4)                 |
| > 55                       | 24 (19.2)                | 12 (26.7)                 | 22 (20.0)                | 7 (15.6)                  |
| Smoke                      |                          |                            |                          |                            |
| no                         | 83 (66.4)                | 30 (66.7)                 | 69 (62.7)                | 20 (44.4)                 |
| yes                        | 42 (33.6)                | 15 (33.3)                 | 41 (37.3)                | 25 (55.6)                 |
| AJCC stage (8th), No. (%)  |                          |                            |                          |                            |
| I-II                       | 43 (34.4)                | 8 (17.8)                  | 39 (35.5)                | 9 (20.0)                  |
| III-IVb                    | 82 (65.6)                | 37 (82.2)                 | 71 (64.5)                | 36 (80.0)                 |
| Tumor classification, No. (%) |                      |                            |                          |                            |
| T1-T2                      | 61 (48.8)                | 12 (26.7)                 | 61 (55.5)                | 23 (51.1)                 |
| T3-T4                      | 64 (51.2)                | 33 (73.3)                 | 49 (44.5)                | 22 (48.9)                 |
| Node classification, No. (%) |                      |                            |                          |                            |
| N0-N1                      | 70 (56.0)                | 19 (42.2)                 | 61 (55.5)                | 18 (40.0)                 |
| N2-N3                      | 55 (44.0)                | 26 (57.8)                 | 49 (44.5)                | 27 (60.0)                 |
| Metastasis, No. (%)        |                          |                            |                          |                            |
| Non-metastasis             | 121 (96.8)               | 43 (95.6)                 | 109 (99.1)               | 44 (97.8)                 |
| Metastasis                 | 4 (3.2)                  | 2 (4.4)                   | 1 (0.9)                  | 1 (2.2)                   |

EBV DNA Epstein-Barr virus DNA, AJCC American Joint Committee on Cancer, SIRI Systemic inflammation response index
(LDH) and SII, to provide more accurate prognostic prediction for patients with NPC [1, 32, 33]. However, the prognosis and survival of NPC patients with negative EBV DNA were often ignored. To the best of our knowledge, this is the first study to investigate inflammatory immune indicators in NPC patients with negative EBV DNA for survival. In the past, scholars have always focused on the survival and prognosis of NPC patients with positive EBV DNA. Patients with negative EBV DNA and high SIRI level should be given more attention, necessary pretreatment inspections and close follow-up monitoring, as well as the formulation of more precise treatment strategies according to the specific situation.

However, our study has some limitations. First, this was a retrospective study conducted in a single institution, and potential bias could not be avoided. Second, due to limited data on EBV DNA during or after treatment, a study of changes in EBV DNA and SIRI was not performed. Therefore, more large-scale prospective, multicenter, randomized clinical trials are still needed for further verification and research.

Conclusion

In summary, the results of our study demonstrated that the SIRI was a significant predictor for prognosis in NPC patients with negative EBV DNA. In addition, inflammatory parameters did not change with EBV DNA status. Therefore, more precise and individualized treatment management strategies can be provided for NPC patients with negative EBV DNA and high SIRI level.

Abbreviations

AC: Adjuvant Chemotherapy; AJCC: American Joint Committee On Cancer; AUC: Area Under The Curve; BMI: Body Mass Index; CCRT: Concurrent Chemoradiotherapy; CTCs: Circulation Tumor Cells; CTV: Clinical Target Volume; EBV: Epstein–Barr Virus; ECT: Emission Computed Tomography; GTVnc: Gross Tumor Volume Of The Nasopharynx; IC: Induction Chemotherapy; IMRT: Intensity-Modulated Radiation Therapy; LDH: Lactate Dehydrogenase; MRI: Magnetic Resonance Imaging; NCCN: National Comprehensive Cancer Network; NPC: Nasopharyngeal Carcinoma; OR: Odds Ratio; OS: Overall Survival; PET-CT: Positron Emission Tomography-Computed Tomography; PFS: Progression-Free Survival; SIRI: Systemic Inflammation Response Index; SII: Systemic Immune-Inflammation Index; TTP: Time To Progression.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12885-022-09942-1.

Acknowledgements

The authors thank all the staff in the Department of Otorhinolaryngology Head and Neck Surgery of NanFang Hospital for their support during the study.

Authors’ contributions

Study concepts: Fan Wang. Study design: Xiong Liu, Jing Chen. Data acquisition: Shiyu Zhou, Shuting Wu, Linchong Cui, Danfan Lin, Zilu Chen, Yue Yuan. Quality control of data and algorithms: Hua Yang, Fangfang Zeng, Shiyu Zhou. Data analysis and interpretation: Xiaofei Yuan, Hua Yang, Fangfang Zeng, Shuting Wu. Statistical analysis: Xiaofei Yuan, Shiyu Zhou, Huiru Feng. Manuscript preparation: Xiaofei Yuan. Manuscript editing: Xiaofei Yuan, Xiong Liu, Jing Chen. Manuscript review: Fan Wang. All authors read and approved the final manuscript.

Funding

This work was supported by the National Natural Science Foundation of China [grant number 81902774], the Natural Science Foundation of Guangdong Province [grant number 2020A1515010176], and the Natural Science Foundation of Guangdong Province [grant number 2020A1515010596].

Availability of data and materials

The data used to support the findings of this study are available from the corresponding author upon request.

Declarations

Ethics approval and consent to participate

This study was approved by the Ethics Committee of Nanfang Hospital of Southern Medical University (Ethical review approval no.: NFEC-2017–165). However, because the few patients we followed up came from various provinces in China, were far away or had died, it was impossible to obtain the relevant consent from the patients. We consulted the ethics committee of our hospital, the ethics committee agreed that we would query relevant information in the case system to conduct a retrospective study, because there was no privacy and tissue samples or blood samples in this study. All methods were performed in accordance with the relevant guidelines and regulations.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

1Department of Otolaryngology-Head and Neck Surgery, Nanfang Hospital, Southern Medical University, Baiyun District, Jingxi Street, Guangzhou 510515, People’s Republic of China. 2Department of Biostatistics, School of Public Health, (Guangdong Provincial Key Laboratory of Tropical Disease Research), Southern Medical University, Guangzhou 510515, Guangdong, China. 3School of Traditional Chinese Medicine, Southern Medical University, Guangzhou 510515, Guangdong, China.

Received: 13 April 2022 Accepted: 26 July 2022
Published online: 05 August 2022

References

1. Chen YP, Chan A, Le QT, Blanchard P, Sun Y, Ma J. Nasopharyngeal carcinoma. Lancet. 2019;394(10192):64–80.
2. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin. 2021;71(3):209–49.
3. Lee AWM, Ma BBY, Ng WT, Chan ATC. Management of Nasopharyngeal Carcinoma. Current Practice and Future Perspective. J Clin Oncol. 2015;33(29):3356–64.
4. Wu F, Wang R, Lu H, Wei B, Feng G, Li G, et al. Concurrent chemoradiotherapy in locoregionally advanced nasopharyngeal carcinoma: Treatment outcomes of a prospective, multicentric clinical study. Radiother Oncol. 2014;112(1):106–11.

5. Farell PJ. Epstein-Barr Virus and Cancer. Annu Rev Pathol. 2019;14:29–53.

6. Chan K, Woo J, King A, Zee B, Lam W, Chan SL, et al. Analysis of Plasma Epstein-Barr Virus DNA to Screen for Nasopharyngeal Cancer. N Engl J Med. 2017;377(6):513–22.

7. Wang WY, Twu CW, Chen HH, Jan JS, Jiang RS, Chao YJC, et al. Plasma EBV DNA Clearance Rate as a Novel Prognostic Marker for Metastatic/Recurrent Nasopharyngeal Carcinoma. Clin Cancer Res. 2010;16(3):1016–24.

8. Chan A, Hui EP, Ngan R, Tung SY, Cheng A, Ng WT, et al. Analysis of Plasma Epstein-Barr Virus DNA in Nasopharyngeal Cancer After Chemoradiation to Identify High-Risk Patients for Adjuvant Chemotherapy: A Randomized Controlled Trial. J Clin Oncol. 2018;36(31):3091-100.

9. Hsu C, Chan S, Chang K, Lin T, Lin C, Hsieh C, et al. Clinical scenario of EBV DNA follow-up in patients of treated localized nasopharyngeal carcinoma. Oral Oncol. 2013;49(6):620–5.

10. Leung SF, Zee B, Ma BB, Hui EP, Mo F, Lai M, et al. Plasma Epstein-Barr viral deoxyribonucleic acid quantitation tumor-node-mesta-sis staging prognostication in nasopharyngeal carcinoma. J Clin Oncol. 2006;24(34):5414–8.

11. Lu L, Li J, Zhao C, Xue W, Han F, Tao T, et al. Prognostic efficacy of combining tumor volume with Epstein-Barr virus DNA in patients treated with intensity-modulated radiotherapy for nasopharyngeal carcinoma. Oral Oncol. 2016;60:18–24.

12. Peng L, Yang Y, Guo R, Mao Y, Xu C, Chen Y, et al. Relationship between pretreatment concentration of plasma Epstein-Barr virus DNA and tumor burden in nasopharyngeal carcinoma: An updated interpretation. Cancer Med-Us. 2018;7(12):5988–98.

13. Zhao FP, Liu X, Chen XM, Liu J, Yu BL, Tian WD, et al. Levels of plasma Epstein-Barr virus DNA prior and subsequent to treatment predicts the prognosis of nasopharyngeal cancer. Oncol Lett. 2015;10(5):2888–94.

14. Huang C, Sun Z, Guo R, Liu X, Mao Y, Peng H, et al. Plasma Epstein-Barr Virus DNA Load After Induction Chemotherapy Predicts Outcome in Locoregionally Advanced Nasopharyngeal Carcinoma. Int J Radiat Oncol Biol Phys. 2019;104(2):355–61.

15. Li W, Yang C, Lv Z, Li J, Li Z, Yuan X, et al. Integrating pre- and post-treatment Plasma Epstein-Barr Virus DNA levels for better prognostic prediction of Nasopharyngeal Carcinoma. J Cancer. 2021;12(9):2715–22.

16. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646–74.

17. Chen J, Zhai E, Yuan Y, Wu K, Xu J, Peng L, et al. Systemic immune-inflammation index for predicting prognosis of colorectal cancer. World J Gastroenterol. 2017;23(4):6261.

18. Zeng X, Liu G, Pan Y, Li Y. Development and validation of immune inflammation–based index for predicting the clinical outcome in patients with nasopharyngeal carcinoma. J Cell Mol Med. 2020;24(15):8326–49.

19. Qiao Q, Zhuang L, Shen Y, Geng Y, Yu S, Chen H, et al. A novel systemic inflammation response index (SIRI) for predicting the survival of patients with pancreatic cancer after chemotherapy. Cancer-AM Cancer Soc. 2016;122(14):2158–67.

20. Chen Y, Jiang W, Xi D, Chen J, Xu G, Yin W, et al. Development and validation of nomogram based on SIRI for predicting the clinical outcome in patients with nasopharyngeal carcinomas. J Invest Med. 2016;67(3):691–8.

21. Lolli C, Caffo O, Scarpi E, Aletta M, Coneduca V, Maines F, et al. Systemic Immune-Inflammation Index Predicts the Clinical Outcome in Patients with mCRPC Treated with Abiraterone. Front Pharmacol. 2016;7:376.

22. Camp RL, Dolled-Filhart M, Rimm DL. X-tile: a new bio-informatics tool for biomarker assessment and outcome-based cut-point optimization. Clin Cancer Res. 2004;10(21):7252–9.

23. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature. 2008;454(7203):436–44.

24. Müller I, Munder M, Kropf P, Hänsch GM. Polymorphonuclear neutrophils and T lymphocytes: strange bedfellows or brothers in arms? Trends Immunol. 2009;30(11):522–30.

25. Marra P, Mathew S, Grigoraidis A, Wu Y, Kyle-Cesar F, Watkins J, et al. IL15RA Drives Antagonistic Mechanisms of Cancer Development and Immune Control in Lymphocyte-Enriched Triple-Negative Breast Cancers. Cancer Res. 2014;74(17):4908–21.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.