Clinical impact of [18F]flutemetamol PET among memory clinic patients with an unclear diagnosis

Antoine Leuzy 1 · Irina Savitcheva2 · Konstantinos Chiotis1 · Johan Lilja3,4 · Pia Andersen5 · Nenad Bogdanovic5 · Vesna Jelic1,5 · Agneta Nordberg1,5

Received: 6 December 2018 / Accepted: 25 February 2019 / Published online: 26 March 2019
© The Author(s) 2019

Abstract

Purpose To investigate the impact of amyloid PET with [18F]flutemetamol on diagnosis and treatment management in a cohort of patients attending a tertiary memory clinic in whom, despite extensive cognitive assessment including neuropsychological testing, structural imaging, CSF biomarker analysis and in some cases [18F]FDG PET, the diagnosis remained unclear.

Methods The study population consisted of 207 patients with a clinical diagnosis prior to [18F]flutemetamol PET including mild cognitive impairment (MCI; n = 131), Alzheimer’s disease (AD; n = 41), non-AD (n = 10), dementia not otherwise specified (dementia NOS; n = 20) and subjective cognitive decline (SCD; n = 5).

Results Amyloid positivity was found in 53% of MCI, 68% of AD, 20% of non-AD, 20% of dementia NOS, and 60% of SCD patients. [18F]Flutemetamol PET led, overall, to a change in diagnosis in 92 of the 207 patients (44%). A high percentage of patients with a change in diagnosis was observed in the MCI group (n = 67, 51%) and in the dementia NOS group (n = 11; 55%), followed by the non-AD and AD (30% and 20%, respectively). A significant increase in cholinesterase inhibitor treatment was observed after [18F]flutemetamol PET (+218%, 34 patients before and 108 patients after).

Conclusion The present study lends support to the clinical value of amyloid PET in patients with an uncertain diagnosis in the tertiary memory clinic setting.

Keywords [18F]Flutemetamol · Amyloid PET · Alzheimer’s disease · Diagnostic change · Cholinesterase inhibitors

Introduction

Reliable biomarkers are a prerequisite for the early and accurate diagnosis of Alzheimer’s disease (AD) and other dementia disorders. Positron emission tomography (PET) imaging using the metabolic tracer 2-deoxy-2-[18F]fluoro-D-glucose ([18F]FDG) is a well-established method for the evaluation of functional changes in the brain of patients with dementia disorders, and has been shown to be useful in discriminating the typical pattern of hypometabolism seen in AD from those seen in other dementia disorders [1]. [18F]FDG PET shows low accuracy, however, in identifying patients with mild cognitive impairment (MCI) who will convert to different forms of dementia [1, 2]. According to the recent International Working Group 2 criteria [3], [18F]FDG PET should be considered a marker of disease progression, in contrast to amyloid-β (amyloid) PET, which is considered a pathophysiological marker.

Since the publication of the first in vivo PET study showing selective imaging of amyloid plaques in AD [4], amyloid PET has proven to be instrumental as a research tool [5]. Beyond this application, amyloid PET holds great potential as a diagnostic aid because of its ability to directly detect a core neuropathological feature of AD, a disease diagnosed with only moderate accuracy, even by expert clinicians [6].
Materials and methods

Participants

The study population consisted of 207 patients attending the Clinic for Cognitive Disorders, Theme Aging, Karolinska University Hospital, Stockholm, Sweden. Patients were seen between 2014 and 2018 and had been mainly referred by primary care physicians (GPs), but also from different hospital clinics, owing to different forms of cognitive problems. Some patients were referred from other memory clinics in Sweden to seek a second opinion. Many patients were relatively young (mean age <65 years). Most patients referred by GPs had undergone cognitive testing (e.g. Mini-Mental State Examination, MMSE), structural imaging (CT, or in a few patients, MRI) and blood analysis, while patients referred from other clinics had often undergone less thorough assessments.

At their first visit, patients underwent physical, neurological, psychiatric and cognitive assessments, and a detailed medical history was recorded. Most patients were accompanied by a close relative or friend. Patients were referred for neuropsychological testing, CT/MR imaging and CSF sampling, and some underwent apolipoprotein E (APOE) genotyping, electroencephalography, speech/language testing and [18F]FDG PET. Diagnoses were based on a consensus meeting between specialists in cognitive disorders, clinical neuropsychologists and specialist nurses. Diagnostic categories included MCI [19, 20], AD [21, 22], dementia of unclear aetiology (not otherwise specified, dementia NOS) [22], dementia due to a non-AD disorder, including dementia with Lewy bodies (DLB) [23], frontotemporal dementia (FTD) [24] and vascular dementia [25], and subjective cognitive decline (SCD) [26].

Neuropsychological assessments

Most patients completed a large battery of neuropsychological tests covering different cognitive domains [27]. These included the MMSE as well as components of the Wechsler Adult Intelligence Scale, Revised (WAIS-R; information and similarities, logical memory, block design and digit symbol), figure classification, subtest of the Synonyms Reasoning Block Test (SRB2), Rey Auditory Verbal Learning Test (RAVLT), copying and memory subtests of the Rey-Osterrieth Complex Figure Test (ROCFT), parts A and B of the Trail Making Test (TMT), and/or the Verbal Fluency Test (FAS).

Structural imaging

Structural CT or T1-weighted MR imaging was performed at various radiology departments in Stockholm using different platforms and protocols. Cerebral atrophy and white matter lesions were assessed clinically by experienced neuroradiologists at the Department of Radiology, Karolinska University Hospital, according to standard visual rating scales. Atrophy of the medial temporal lobe was evaluated using the medial temporal atrophy (MTA) scale [28]. Overall cortical atrophy was assessed using the global cortical atrophy (GCA) scale [29]. White matter hyperintensities were scored using the Fazekas scale [30].

CSF biomarkers

CSF samples were collected via lumbar puncture from 152 out of the 207 patients (73%) under nonfasting conditions as part of routine memory assessment. The lack of CSF biomarkers in 55 patients was due to the use of anticoagulants in 18 patients, spinal stenosis or related problems in 6, refusal to undergo CSF sampling in 12, and technical problems in 19. The CSF samples were routinely analysed at the Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal, Sweden, for Aβ1-42, t-tau and p-tau, using commercially available ELISAs (INNOTEST; Fujirebio,
PET investigations

[18F]FDG and [18F]flutemetamol PET investigations were performed at the Department of Medical Radiation Physics and Nuclear Medicine Imaging, Karolinska University Hospital, Stockholm, Sweden, using a Biograph mCT PET/CT scanner (Siemens/CTI, Knoxville, TN).

[18F]FDG PET

[18F]FDG PET was performed as part of the clinical assessment in 78 out of the 207 patients (38%). These studies were performed prior to [18F]flutemetamol PET (median 5 months, interquartile range, IQR, 3–12 months) in 68 patients (87%) and after [18F]flutemetamol PET (median 2 months, IQR 0.5–3 months) in 9 patients. [18F]FDG PET was performed as a 10-min or 15-min list-mode scan 30 to 45 min after injection of 2–3 MBq/kg. All appropriate corrections, including point spread function, scatter and time-of-flight were applied, with a low-dose CT scan used for attenuation correction. Images were reconstructed using ordered subsets expectation maximization (OSEM; five iterations, 21 subsets, 2.0 mm gaussian filter), yielding an effective spatial resolution of 3.0 mm.

According to clinical routine assessment, summation images were visually analysed, supported by a semiquantitative analysis in the form of standardized uptake value ratios (SUVR) based on automatically generated regions of interest (ROI; cortical and subcortical regions, normalized to whole brain) and voxelwise Z-score stereotactic surface projection images (Siemens syngo via software). Metabolic patterns were classified as typical of AD, as suggestive but not typical of AD, as consistent with other non-AD neurodegenerative diseases (e.g. FTD or DLB) or as nonspecific in the case of minor patchy focal changes.

[18F]Flutemetamol PET

[18F]Flutemetamol PET was performed as a 20-min list-mode scan 90 min after injection of 185 MBq [18F]flutemetamol. Data were corrected and images reconstructed in an identical way to the [18F]FDG PET studies. [18F]Flutemetamol summation images were visually assessed as positive (abnormal) or negative (normal) by a board-certified nuclear medicine physician with later reassessment by either a neuroradiologist or a physician with experience in neuroimaging who had successfully completed an electronic training programme developed by GE Healthcare for the interpretation of [18F]flutemetamol images [31]. In cases of disagreement between raters, a consensus was achieved between readers during the re-read session.

In addition to visual reads, semiquantitative analysis of [18F]flutemetamol uptake was performed using an automated ROI-based approach (Hermes Medical Solutions) [32], with SUVR calculated using an isocortical composite ROI, comprising brain regions typically associated with high amyloid load in AD (frontal, lateral temporal, cingulate and parietal cortices), using the pons as the reference region. Amyloid-positivity was defined using an a priori SUVR cut-off value of 0.60, based on separation from cognitively normal controls [33].

Statistical analysis

Characteristics were compared between diagnostic groups using Kruskal Wallis analysis of variance (ANOVA) and the Wilcoxon signed ranks test for continuous variables. Categorical variables were assessed using Fisher’s exact test. Agreement between visual and SUVR-based [18F]flutemetamol classifications, as well as interrater agreement for visual readings, were assessed using percentage agreement and Cohen’s kappa. The proportions of patients showing changes in diagnosis and drug treatment were assessed using a one-sample proportions test. All statistical tests were performed using R v. 3.3.2 (The R Foundation for Statistical Computing), with two-sided p values <0.05 considered to indicate significance.

Results

The demographic, clinical and biomarker findings in the 207 patients in whom [18F]flutemetamol PET was performed due to diagnostic uncertainty are presented in Table 1. Most patients received an initial diagnosis of MCI (131, 63%), followed by AD (41, 20%), dementia NOS (20, 10%), non-AD (10, 5%) and SCD (5, 2%). Figure 1 shows the structural imaging-based ratings for atrophy and white matter changes in the different patient groups before amyloid PET. MTA scores of 0 or 1, indicating no or minimal atrophy, respectively, were predominant across the MCI, AD, dementia NOS and SCD groups. In contrast, MTA scores indicating mild atrophy were noted in half of the patients with non-AD disorders. In terms of GCA and white matter lesions, most patients showed mild changes. [18F]FDG PET imaging was performed in 78 of the 207 patients (38%; Table 1, Fig. 1d). A metabolic pattern suggestive but not typical of AD was the most common finding in those with a diagnosis of MCI or AD prior to [18F]flutemetamol PET (16 patients, 36%, and 9 patients, 56%, respectively); in the remaining groups, patterns not typical of AD or suggestive of a non-AD disorder were predominant (non-AD, 4 patients, 100%; dementia NOS, 10 patients, 91%), with only minor patchy changes seen in patients with SCD. Similar findings were obtained using the diagnoses
obtained after $^{[18F]}$flutemetamol PET, with only 11 AD patients (14%) showing a typical metabolic pattern.

CSF was sampled in 152 patients according to the Swedish clinical practice guidelines for memory assessment at specialist memory clinics. Overall, 103 patients (68%) showed abnormal CSF biomarkers: 24% showed only abnormal $\beta_1$-$42$, 21% showed abnormal $\beta_1$-$42$ in combination with elevated t-tau or p-tau, and 32% showed negative CSF; a further 23% showed only abnormal tau. The main reasons for performing amyloid PET were a clinical suspicion of AD in combination with either a negative or an unclear (i.e. isolated positive or borderline $\beta_1$-$42$ or tau) CSF profile (117 patients, 57%).

### Table 1

Demographic, clinical and biomarker data using diagnoses obtained prior to $^{[18F]}$flutemetamol PET

|                      | MCI | AD | Non-AD disorder | Dementia NOS | SCD |
|----------------------|-----|----|-----------------|--------------|-----|
| Number of patients   | 131 | 41 | 10              | 20           | 5   |
| Age (years), mean (SD) | 64.5 (8.6) | 65.2 (8.7) | 67.6 (5.2) | 62.3 (9.5) | 68.4 (8.3) |
| Sex, female/male (n/n) | 76/55 | 27/14 | 5/5            | 10/10        | 3/2 |
| $APOE\epsilon 4$    | 25.6 (3.7) | 24.5 (3.7) | 23.4 (3.8) | 22 (5.5) | 29 (1.2) |
| Number of patients with data available | 75 | 27 | 4              | 9            | 5   |
| Number (%) positive | 42 (56) | 15 (56) | 2 (50) | 2 (22) | 2 (40) |
| CSF biomarkers       |     |         |                |              |     |
| $A\beta_1$-$42$ (ng/L), mean (SD) |         |         |     |     |     |
| $p$-tau (ng/L), mean (SD) |     |     |     |     |     |
| $t$-tau (ng/L), mean (SD) |     |     |     |     |     |
| CT/MRI scales        |     |         |                |              |     |
| MTA ($n^d$)          |     |         |                |              |     |
| 0/1$^k$              | 76   | 20  | 2               | 10           | 5   |
| 2                    | 39   | 14  | 5               | 6            | 0   |
| 3                    | 11   | 5   | 2               | 2            | 0   |
| 4                    | 2    | 1   | 1               | 1            | 0   |
| GCA ($n^e$)          |     |         |                |              |     |
| 0                    | 32   | 8   | 1               | 4            | 0   |
| 1                    | 75   | 25  | 5               | 9            | 3   |
| 2                    | 20   | 7   | 3               | 5            | 2   |
| 3                    | 1    | 0   | 1               | 1            | 0   |
| Fazekas scale ($n^f$) |     |         |                |              |     |
| 0                    | 28   | 7   | 1               | 3            | 0   |
| 1                    | 68   | 21  | 6               | 10           | 4   |
| 2                    | 21   | 8   | 2               | 1            | 1   |
| 3                    | 11   | 4   | 1               | 5            | 0   |
| $^{[18F]}$FDG metabolic patterns$^g$ |     |         |                |              |     |
| Number of patients with data available | 44 | 16 | 4              | 11           | 3   |
| Typical of AD ($n$)  | 4    | 2   | 0               | 0            | 0   |
| Not typical of AD ($n$) | 16 | 9   | 0               | 5            | 0   |
| Non-AD ($n$)         | 13   | 3   | 2               | 5            | 0   |
| Nonspecific ($n$)    | 11   | 2   | 2               | 1            | 3   |
| $^{[18F]}$Flutemetamol |     |         |                |              |     |
| Number (%) positive on visual assessment$^h$ |     |         |                |              |     |
| Global SUVR, mean (SD)$^i$ |     |         |                |              |     |
| Number (%) positive by SUVR$^j$ |     |         |                |              |     |

*AD* Alzheimer’s disease, *GCA* global cortical atrophy, *MCI* mild cognitive impairment, *MMSE* Mini-Mental State Examination, *MTA* medial temporal atrophy, *NOS* not otherwise specified, *SUVR* standardized uptake value ratio

$^a$MMSE: dementia NOS < AD ($p < 0.01$); MCI, AD, non-AD, dementia NOS < SCI ($p < 0.01$)

$^b$ $p$-tau AD > MCI and dementia NOS ($p < 0.05$); SCI > dementia NOS ($p < 0.05$)

$^c$t-tau: AD > dementia NOS ($p < 0.01$)

$^d$MTA (0/1 none/minimal, 2 mild, 3 moderate, 4 severe): MCI vs. non-AD ($p < 0.01$); AD vs. non-AD ($p < 0.01$); non-AD vs. dementia NOS ($p < 0.05$)

$^e$GCA: 0 none, 1 mild, 2 moderate, 3 severe

$^f$Fazekas scale (white matter hyperintensities): 0 none, 1 mild, 2 moderate, 3 severe

$^g$ $^{[18F]}$FDG metabolic patterns: AD vs. SCI ($p < 0.05$), dementia NOS vs. SCI ($p < 0.01$)

$^h$Visual assessment: MCI > non-AD ($p < 0.01$); AD > non-AD ($p < 0.01$); non-AD > dementia NOS ($p < 0.05$)

$^i$SUVR: MCI > dementia NOS ($p < 0.05$), AD > MCI ($p < 0.01$), non-AD ($p < 0.01$), dementia NOS ($p < 0.001$)

$^j$SUVR positivity (SUVR defined using a global cortical cut-off value of >0.60): MCI > dementia NOS ($p < 0.01$), AD > non-AD ($p < 0.01$) AD > dementia NOS ($p < 0.001$)

$^k$MTA 0 and 1 are considered normal; these categories were therefore combined

---

Eur J Nucl Med Mol Imaging (2019) 46:1276–1286 1279
the absence of CSF samples (55 patients, 27%). A clinically unclear picture of memory decline in combination with a CSF profile indicating AD (low A\(_{\beta}1-42\) and one or both tau markers positive) was the third most common reason (32 patients, 15%). Table 2 and Fig. 2 show the relatively poor agreement between CSF biomarkers and \([^{18}\text{F}]\)flutemetamol PET (A\(_{\beta}1-42\), 66%; p-tau, 76%; t-tau, 77%).

There was high agreement between raters in the visual analysis of the \([^{18}\text{F}]\)flutemetamol PET scans (198/207, 96%; Cohen’s kappa = 0.92). In 37 visual readings, however, one of the two readers had some difficulty in defining the \([^{18}\text{F}]\)flutemetamol PET scan as positive or negative; this finding was similarly distributed between experienced and less experienced readers (20 and 17 readings, respectively). In four scans, both readers were uncertain. Overall, some uncertainties were observed in 11% of readings. Agreement between visual and semiquantitative assessment approaches was achieved in all cases. Using both visual and SUVR-based classifications, a higher proportion of patients were rated as \([^{18}\text{F}]\)flutemetamol-positive in the preamyloid PET MCI and AD patients than in the dementia NOS patients (\(p<0.01\)), and in the AD patients as compared with the non-AD patients (\(p<0.05\)).

On visual evaluation of the \([^{18}\text{F}]\)flutemetamol PET scans, amyloid positivity was found in 69 of 131 patients (53%) with an initial diagnosis of MCI, in 28 of 41 (68%) with AD, in 2 of 10 (20%) with a non-AD disorder, in 4 of 20 (20%) with dementia NOS and in 3 of 5 (60%) with SCD (Table 1). Amyloid status in the diagnostic groups before and after \([^{18}\text{F}]\)flutemetamol PET as well as changes in diagnosis are summarized in Table 3 and Fig. 3, respectively. As shown in...
Fig. 3, the majority of patients with a diagnosis of MCI after \([^{18}F]\)flutemetamol were amyloid-negative. The vast majority of patients with an initial diagnosis of MCI who were amyloid-positive received a diagnosis of prodromal AD or AD (54 of 60, 90%). In patients with an initial diagnosis of AD, the diagnosis was dismissed in seven amyloid-negative patients.

Finally, all patients with an initial diagnosis of dementia NOS group, and almost all patients with a non-AD disorder were amyloid-negative. Overall, \([^{18}F]\)flutemetamol PET led to a significant change in diagnosis (92 patients, 44%; \(p < 0.05\)). Among the patients with MCI, dementia NOS, AD and a non-AD disorder, the highest percentage change in diagnosis was observed in those with MCI (67 patients, 51%) as well as in those with dementia NOS (11

Table 2 Agreement between CSF positivity and \([^{18}F]\)flutemetamol PET positivity using dichotomized measures based on the diagnoses made before \([^{18}F]\)flutemetamol PET

| CSF biomarker | MCI (n = 86\(^c\)) | AD (n = 38\(^e\)) | Non-AD disorder (n = 9\(^f\)) | Dementia NOS (n = 15\(^g\)) | SCD (n = 4\(^i\)) |
|---------------|---------------------|-------------------|-------------------------------|-----------------------------|------------------|
| Positive      | \(\beta\)        | 25 (29)           | 19 (50)                       | 2 (22)                      | 2 (13)           |
|               | p-tau              | 30 (35)           | 19 (50)                       | 2 (22)                      | 2 (13)           |
|               | t-tau              | 29 (34)           | 20 (52)                       | 3 (33)                      | 2 (13)           |
| Negative      | \(\beta\)        | 11 (13)           | 3 (8)                         | 3 (33)                      | 2 (13)           |
|               | p-tau              | 3 (3)             | 4 (10)                        | 2 (22)                      | 3 (20)           |
|               | t-tau              | 3 (3)             | 3 (8)                         | 1 (11)                      | 3 (20)           |
| Positive      | \(\beta\)        | 23 (27)           | 6 (16)                        | –                           | 3 (75)           |
|               | p-tau              | 18 (21)           | 6 (16)                        | –                           | –                |
|               | t-tau              | 19 (22)           | 6 (16)                        | –                           | –                |
| Negative      | \(\beta\)        | 27 (31)           | 10 (26)                       | 4 (45)                      | 11 (74)          |
|               | p-tau              | 35 (41)           | 9 (24)                        | 5 (56)                      | 10 (67)          |
|               | t-tau              | 35 (41)           | 9 (24)                        | 5 (56)                      | 10 (67)          |

Concordance (both biomarkers positive or negative): CSF-positive/PET-positive and CSF-negative/PET-negative. Discordance (only one of two biomarkers positive): CSF-positive/PET-negative (isolated CSF positivity) and CSF-negative/PET-positive (isolated PET positivity).

\(a\) The cut-off value used for \([^{18}F]\)flutemetamol SUVR was 0.60, in combination with visual assessment.

\(b\) The cut-off values used to binarize CSF biomarkers were <550 ng/L for \(\beta\), >80 ng/L for p-tau and >400 ng/L for t-tau.

\(c\) Number of patients in whom CSF was sampled.

Fig. 2 Relationships between CSF biomarkers and isocortical composite \([^{18}F]\)flutemetamol SUVR. The vertical lines mark the cut-off value of 0.60 for isocortical composite \([^{18}F]\)flutemetamol SUVR; the horizontal lines mark the cut-off values for \(\beta\) (a <550 pg/mL), p-tau (b >80 pg/mL) and t-tau (c >400 pg/mL); the dashed lines indicate borderline zones (within 5% of the cut-off values); and the grey and white quadrants indicate concordance and discordance between biomarkers, respectively.
patients, 55%), while a smaller percentage change was seen in those with a non-AD disorder and those with AD (3 patients, 30%, and 8 patients, 20%, respectively). Among the five patients with SCD, in three with a positive [18F]flutemetamol scan the initial diagnosis was revised to MCI, prodromal AD and AD, respectively, on clinical follow-up.

The outcome of amyloid PET led not only to a change in diagnosis but also to more patients receiving treatment with cholinesterase inhibitors (ChEIs). Nine patients with MCI (seven amyloid-positive), 22 with prodromal AD/AD (16 amyloid-positive) and 3 with non-AD/dementia NOS (one amyloid-positive) and 3 with non-AD/dementia NOS (one amyloid-positive) were receiving treatment with various ChEIs prior to amyloid PET. After amyloid PET and revision of initial diagnoses, 92 patients with prodromal AD/AD (87 amyloid-positive), 8 with MCI (4 amyloid-positive) and 9 with non-AD/dementia NOS (2 amyloid-positive) received ChEI treatment. ChEI treatment was used in 34 patients prior to amyloid PET and in 109 patients after amyloid PET, an increase of 75 patients (+218%; \( p < 0.001 \)). Treatment was discontinued following [18F]flutemetamol PET in one amyloid-negative patient with

### Table 3 Change in diagnosis following [18F]flutemetamol PET

| Initial diagnosis | Change | [18F]Flutemetamol PETa | Number (%) of those with change | Diagnosis after [18F]flutemetamol PET |
|------------------|--------|-------------------------|---------------------------------|--------------------------------------|
| MCI              | Positive | 58 (87%)               | 1 non-AD (DLB), 13 prodromal AD, 44 AD |
|                  | Negative | 9(13%)                  | 1 AD, 3 dementia NOS, 5 non-AD (4 VaD, 1 PSP) |
| AD               | Positive | 1 (12.5%)               | Non-AD (DLB)                    |
|                  | Negative | 7 (87.5%)               | 1 dementia NOS, 2 non-AD (DLB, FTD), 4 MCI |
| Non-AD           | Positive | 2 (67%)                 | AD                              |
|                  | Negative | 1 (33%)                 | MCI                            |
| Dementia NOS     | Positive | 4 (36%)                 | AD                              |
|                  | Negative | 7 (64%)                 | Non-AD (1 DLB, 3 VaD, 3 FTD)    |
| SCD              | Positive | 3 (100%)                | MCI, prodromal AD, AD           |
|                  | Negative | –                      | –                               |

DLB dementia with Lewy bodies, FTD frontotemporal dementia, PSP progressive supranuclear palsy, VaD vascular dementia

a [18F]Flutemetamol PET status (positive/negative) was based on visual assessment supported by SUVR findings

b In two non-AD patients with DLB the initial diagnosis was FTD

Fig. 3 Visual [18F]flutemetamol ratings in the various diagnostic groups before (a) and after (b) [18F]flutemetamol PET (the number at the top of each column indicates the number of patients). Red [18F]flutemetamol-positive, blue [18F]flutemetamol-negative
MCI due to cholinergic side effects. Of the 109 patients receiving ChEi treatment, 93 (85%) were amyloid-positive, and 92 of 99 patients (93%) had a diagnosis of prodromal AD/AD after [18F]flutemetamol PET, while only 8 of 72 patients with MCI and 9 of 36 (25%) with non-AD/dementia NOS (including those with DLB) received ChEi treatment.

Discussion

The rapid development of molecular imaging techniques has enabled the in vivo study of the pathophysiology underlying different neurodegenerative diseases, including AD. Amyloid imaging has reached clinical use in memory clinics. However, most research studies published to date have included selected research populations. The present study included an unselected cohort of 207 patients, in whom extensive neuropsychological and, in various subsets, biomarker-based assessments could not provide a sufficiently certain clinical diagnosis. The main reasons for amyloid PET in our cohort were the clinical suspicion of AD accompanied by unclear or negative CSF findings, the absence of CSF biomarkers (due to contraindications, patient refusal or technical difficulties) and an ambiguous clinical presentation in the context of a CSF profile indicative of AD (low Aβ1-42 and elevated tau).

Despite the fact that [18F]FDG PET is a well-established diagnostic tool in the work-up of dementia disorder patients, it did not provide an adequate differential diagnosis in the present cohort. We estimate that this cohort of patients, requiring complementary amyloid PET after an extensive clinical work-up including neuropsychological testing, MRI, CSF biomarker analysis and other additional examinations, represents approximately 10% of all new referrals to our academic memory clinic (approximately 500 to 600 per year).

In this study, we assessed the incremental value of amyloid PET in the work-up of patients being followed in a tertiary specialist setting due to cognitive impairment. The largest patient group was MCI patients of whom 69 (53%) were [18F]flutemetamol PET-positive. A positive [18F]flutemetamol PET scan in this group led mainly to a diagnosis of prodromal AD/AD, while a negative scan, with a few exceptions, led to a diagnosis of non-AD/dementia NOS, or (in a subset of MCI patients) to retention of the original diagnosis. Overall, amyloid PET led to a significant change in diagnosis in 44% of patients and an increase in the use of AD drug treatment, from 34 to 109 of 207 patients (16% to 53%). On the basis of our study design, in which amyloid PET results were used to revise initial diagnoses and treatment plans, our findings provide an estimate of the incremental value of this type of investigation in the clinical setting. Thus, this study fulfils the requirements of a phase 4 study as defined by the five-phase biomarker validation framework recently introduced to the field [34, 35]. This type of study is vital to assess the utility of amyloid PET and to accelerate the development of evidence-based guidelines for its clinical use [36].

In the present study, the concordance rates observed between [18F]flutemetamol PET and CSF Aβ1-42 were lower than those reported to date from the ADNI (Alzheimer’s Disease Neuroimaging Initiative) and the Swedish BioFINDER (Biomarkers for Identifying Neurodegenerative Disorders Early and Reliably) studies [37-41], in which the composition of the cohorts largely resembles that found in clinical trials. The difference in observations is due to the fact that in the present study only patients with an unclear diagnosis following extensive assessment were recruited for amyloid imaging. Although our study, by design, was predisposed to discordance, it draws attention to potential complications in implementing the recent AD biomarker classification scheme [42]; based on categorical and biomarker analyses, this framework assumes that measures of amyloid based on CSF analysis and PET are interchangeable. Although disagreement between imaging and CSF analysis may be a reflection of how cut-off values are established—with European laboratories generally using Aβ1-42 cut-off values that are too low [18]—the use of a more lenient cut-off value (647 ng/L) derived from a recent study using the same CSF INNOTEST assay [40] resulted in the same overall level of concordance; though an increase in concordant-positive subjects was seen, this was accompanied by an increase in the number of subjects showing isolated CSF positivity. There is evidence, however, to suggest that higher cut-off values for Aβ1-42 may prove appropriate. Further studies addressing this, including comparison with Aβ1-42/tau ratios, are warranted. While many subjects may show concordance in the long term [42], more work is required to integrate and compare CSF analysis and amyloid PET so as to develop more refined guidelines for how these biomarkers are to be used and interpreted in the diagnostic work-up of patients with dementia disorders [43], including considerations related to differences in amyloid processing and neurodegeneration seen across atypical forms of AD [44] and the value of Aβ1-42 ratios with shorter isoforms [45, 46] and tau/Aβ1-42 [47].

The percentage of patients with a change in diagnosis observed in this study (44%) was higher than the average of those found in previous studies [48], although somewhat lower than in other studies [10, 11]. This most likely reflects differences in the composition of the study cohorts; in the present study, diagnostic uncertainty remained despite extensive neuropsychological testing, imaging and biomarker investigations. Our study cohort therefore most likely included a higher proportion of patients with atypical disease, an assertion supported by the number of MCI and dementia NOS patients with an unchanged diagnosis. We consider, however, that the most important finding was the high percentage of amyloid-positive MCI patients who received a change in
FDG PET is not always useful, especially in those with this population of patients it has also been demonstrated that medical reasons, or patient refusal of lumbar puncture. In pling is not possible due to the use of anti-coagulants, other when CSF biomarkers are not consistent or when CSF sam-

In our cohort, amyloid PET led to significant changes in the management of patients, including treatment with ChEIs, with a more than threefold increase in the number of patients receiving ChEI treatment. This increase was primarily accounted for by amyloid-positive patients with a diagnosis after [18F]flutemetamol PET of prodromal AD/AD. Similarly, amyloid-positive patients with a diagnosis after [18F]flutemetamol PET of AD comprised the majority of those receiving ChEIs prior to undergoing amyloid PET. The small number of [18F]flutemetamol-positive MCI and non-AD (DLB) patients receiving cholinergic drugs probably reflects varying adherence by physicians to the diagnos-
tic codes (i.e. MCI, amyloid-positive, versus prodromal AD, a research term not currently recognized in dementia care guidelines) and evidence suggesting that this drug class is of benefit in DLB [52]. Among the few amyloid-negative patients, treatment, whether initiated prior to or after [18F]flutemetamol PET, was related to CSF Aβ1-42 positivity (seen in close to half of such patients) and/or physician-specific treatment beliefs.

The present study underlines the value of amyloid PET when CSF biomarkers are not consistent or when CSF sampling is not possible due to the use of anti-coagulants, other medical reasons, or patient refusal of lumbar puncture. In this population of patients it has also been demonstrated that [18F]FDG PET is not always useful, especially in those with MCI. A negative amyloid PET study is important information for an MCI patient and often results in a decrease in the use of resources due to fewer ancillary investigations. In patients with aetiologically unclear dementia or non-AD, amyloid PET could provide important diagnostic information, as was illustrated in the present study with a 55% change in diagnosis among dementia NOS patients. Although the fact that the generally rather extensive investigations undergone by the patients in this study help to establish the clinical usefulness of [18F]flutemetamol PET, it must also be underlined that our findings may not be generalizable to older patients or those attending less specialized clinical centres. Although incorporating cost-effectiveness parameters associated with amyloid imaging was beyond the scope of the present investigation, further studies addressing this, including the optimal stage (early versus late) for the use of amyloid PET in the diagnostic work-up of patients, are crucial.

Conclusion

In summary, our findings indicate that amyloid PET imaging with [18F]flutemetamol had a significant impact in terms of change of diagnosis, management and drug treatment when added to the work-up of patients with an uncertain diagnosis followed in the setting of a tertiary memory clinical. These findings highlight the clinical value of amyloid PET in pa-

Compliance with ethical standards

Conflicts of interest J.L. is an employee of Hermes Medical Solutions, Stockholm, Sweden. All other authors declare no conflicts of interest.

Ethical approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the principles of the 1964 Declaration of Helsinki and its later amendments or comparable ethical standards.

Informed consent Informed consent was obtained from all individual participants included in the study.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

1. Nobili F, Arbizu J, Bouwman F, Drzezga A, Agosta F, Nestor P, et al. European Association of Nuclear Medicine and European Academy of Neurology recommendations for the use of brain (18)F-fluorodeoxyglucose positron emission tomography in neuro-
degenerative cognitive impairment and dementia: Delphi consensi.

2. Smailagic N, Vacante M, Hyde C, Martin S, Ukoumunne O, Sachpekidis C, (18)F-FDG PET for the early diagnosis of Alzheimer’s disease dementia and other dementias in people with mild cognitive impairment (MCI). Cochrane Database Syst Rev. 2015:1;CD010632. https://doi.org/10.1002/ 14651858.CD010632.pub2.

3. Dubois B, Feldman HH, Jacova C, Hampel H, Molinuevo JL, Blennow K, et al. Advancing research diagnostic criteria for Alzheimer’s disease: the IWG-2 criteria. Lancet Neurol. 2014;13: 614–29. https://doi.org/10.1016/S1474-4422(14)70090-0.
4. Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, Holt DP, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh compound-B. Ann Neurol. 2004;55:306–19. https://doi.org/10.1002/ana.20009.

5. Rabinovici GD. The translational journey of brain beta-amyloid imaging: from positron emission tomography to autopsy to clinic. JAMA Neurol. 2015;72:265–6. https://doi.org/10.1001/jamaneurol.2014.4143.

6. Beach TG, Monsell SE, Phillips LE, Kukull W. Accuracy of the clinical diagnosis of Alzheimer disease at National Institute on Aging Alzheimer Disease Centers, 2005-2010. J Neuropathol Exp Neurol. 2012;71:266–73. https://doi.org/10.1097/NEN.0b013e31824b211b.

7. Klein EP, Kaye J. Dementia specialists and early adoption of amyloid imaging. J Alzheimers Dis. 2013;33:445–50. https://doi.org/10.3233/JAD-2012-121216.

8. Fantoni ER, Chalkidou A, O’Brien JT, Farrar G, Hammars A. A systematic review and aggregated analysis on the impact of amyloid PET brain imaging on the diagnosis, diagnostic confidence, and management of patients being evaluated for Alzheimer's disease. J Alzheimers Dis. 2018;63:783–96. https://doi.org/10.3233/JAD-171093.

9. Shea YF, Barker W, Greig-Gusto MT, Loewenstein DA, Duara R, DeKosky ST. Impact of amyloid PET imaging in the memory clinic: a systematic review and meta-analysis. J Alzheimers Dis. 2018;64:323–35. https://doi.org/10.3233/JAD-180239.

10. Ceccaldi M, Jonveaux T, Verger A, Krolak-Salmon P, Houzard C, Godefroy O, et al. Added value of [(18)F]-florbetaben amyloid PET in the diagnostic workup of most complex patients with dementia in France: a naturalistic study. Alzheimers Dement. 2018;14:293–305. https://doi.org/10.1017/jolz.2017.09.009.

11. Boccardi M, Altmare D, Ferrari C, Festari C, Guerra UP, Paghera F, et al. Assessment of the incremental diagnostic value of florbetapir F18 imaging in patients with cognitive impairment: the Incremental Diagnostic Value of Amyloid PET with [18F]-Florbetapir (INDIA-FBP) Study. JAMA Neurol. 2016;73:1417–24. https://doi.org/10.1001/jamaneurol.2016.3751.

12. Bensaidane MR, Chalkidou A, O’Brien JT, Farrar G, Hammars A. A systematic review and aggregated analysis on the impact of amyloid PET brain imaging on the diagnosis, diagnostic confidence, and management of patients being evaluated for Alzheimer's disease. J Alzheimers Dis. 2013;33:445–50. https://doi.org/10.3233/JAD-2012-121216.

13. Zwan MD, Bouwman FH, Konijnenberg E, van der Flier WM, Verwer JL, et al. A consecutive case series experience with [18F] florbetapir PET imaging in patients with amnestic mild cognitive impairment. JAMA Neurol. 2018;75:1062–70. https://doi.org/10.1001/jamaneurol.2018.1346.

14. Zwan MD, Rinne JO, Hasselbalch SG, Nordberg A, Lleo A, Herukka SK, et al. Use of amyloid-PET to determine cutpoints for CSF markers: a multicenter study. Neurology. 2016;86:50–8. https://doi.org/10.1212/WNL.0000000000002081.

15. Petersen RC, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E. Mild cognitive impairment: clinical characterization and outcome. Arch Neurol. 1999;56:303–8.

16. Winblad B, Palmer K, Kivipelto M, Jelic V, Fratiglioni L, Wahlund LO, et al. Mild cognitive impairment—beyond controversies, towards a consensus: report of the International Working Group on Mild Cognitive Impairment. J Intern Med. 2004;256:240–6. https://doi.org/10.1111/j.1365-2796.2004.01380.x.

17. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's disease. Neurology. 1984;34:939–44.

18. World Health Organization. The ICD-10 classification of mental and behavioural disorders: clinical descriptions and diagnostic guidelines. Geneva: World Health Organization; 1992.

19. McKee IG, Galasko D, Kosaka K, Perry EK, Dickson DW, Hansen LA, et al. Consensus guidelines for the clinical and pathologic diagnosis of dementia with Lewy bodies (DLB): report of the Consortium on DLB International Workshop. Neurology. 1996;47:1113–24.

20. Neary D, Snowden JS, Gustafson L, Passant U, Stuss D, Black S, et al. Frontotemporal lobar degeneration: a consensus on clinical diagnostic criteria. Neurology. 1998;51:1546–54.

21. Roman GC, Tattemich TK, Erkinjuntti T, Cummings JL, Masdeu JC, Garcia JC, et al. Vascular dementia: diagnostic criteria for research studies. Report of the NINDS-AIREN International Workshop. Neurology. 1993;43:250–60.

22. Jessen F, Amariglio RE, van Boxtel M, Breteler M, Ceccaldi M, Chetelat G, et al. A conceptual framework for research on subjective cognitive decline in preclinical Alzheimer's disease. Alzheimers Dement. 2014;10:84–52. https://doi.org/10.1016/j.jalz.2014.01.001.

23. Garcia-Ptaeck S, Cavallin L, Kareholt I, Kramberger MG, Winblad B, Jelic V, et al. Subjective cognitive impairment subjects in our clinical practice. Dement Geriatr Cogn Dis Extra. 2014;4:419–30. https://doi.org/10.1159/000366270.

24. Scheltens P, Leys D, Barkhof F, Huglo D, Weinstein HC, Vermersch P, et al. Atrophy of medial temporal lobes on MRI in “probable” Alzheimer's disease and normal ageing: diagnostic value and neuropsychological correlates. J Neurol Neurosurg Psychiatry. 1992;55:967–72.

25. Pasquier F, Leys D, Weerts JG, Mounier-Vehier F, Barkhof F, Scheltens P. Inter- and intraobserver reproducibility of cerebral atrophy assessment on MRI scans with hemispheric infarcts. Eur Neurol. 1996;36:268–72.

26. Fazekas F, Chawluk JB, Alavi A, Hurtig HI, Zimmerman RA. MR signal abnormalities at 1.5 T in Alzheimer's dementia and normal ageing: diagnostic value and neuropsychological correlates. J Nucl Med. 1987;28:12–9. https://doi.org/10.2967/jnumed.118.207811.

27. Buckley CJ, Sherwin PF, Smith AP, Wolber J, Weick SM, Brooks JC, Garcia JH, et al. Vascular dementia: diagnostic criteria for research studies. Report of the NINDS-AIREN International Workshop. Neurology. 1993;43:250–60.

28. Winblad B, Palmer K, Kivipelto M, Jelic V, Fratiglioni L, Wahlund LO, et al. Mild cognitive impairment—beyond controversies, towards a consensus: report of the International Working Group on Mild Cognitive Impairment. J Intern Med. 2004;256:240–6. https://doi.org/10.1111/j.1365-2796.2004.01380.x.

29. Petersen RC, Smith GE, Waring SC, Ivnik RJ, Tangalos EG, Kokmen E. Mild cognitive impairment: clinical characterization and outcome. Arch Neurol. 1999;56:303–8.

30. Winblad B, Palmer K, Kivipelto M, Jelic V, Fratiglioni L, Wahlund LO, et al. Mild cognitive impairment—beyond controversies, towards a consensus: report of the International Working Group on Mild Cognitive Impairment. J Intern Med. 2004;256:240–6. https://doi.org/10.1111/j.1365-2796.2004.01380.x.

31. Bensaidane MR, Beauregard JM, Poulin S, Buteau FA, Guimond J, et al. Subjective cognitive impairment subjects in our clinical practice. Dement Geriatr Cogn Dis Extra. 2014;4:419–30. https://doi.org/10.1159/000366270.
disease and mild cognitive impairment: a phase 2 trial. Ann Neurol. 2010;68:319–29. https://doi.org/10.1002/ana.22068.

34. Boccardi M, Gallo V, Yasui Y, Viece P, Padovani A, Mosimann U, et al. The biomarker-based diagnosis of Alzheimer's disease: 2–lessons from oncology. Neurobiol Aging. 2017;52:141–52. https://doi.org/10.1016/j.neurobiolaging.2017.01.021.

35. Frisoni GB, Perani D, Bastianello S, Bernardi G, Porteri C, Boccardi M, et al. Biomarkers for the diagnosis of Alzheimer's disease in clinical practice: an Italian intersocietal roadmap. Neurobiol Aging. 2017;52:119–31. https://doi.org/10.1016/j.neurobiolaging.2016.02.033.

36. Chiotis K, Saint-Aubert L, Boccardi M, Gietl A, Picco A, Varrone A, et al. Clinical validity of increased cortical uptake of amyloid ligands on PET as a biomarker for Alzheimer's disease in the context of a structured 5-phase development framework. Neurobiol Aging. 2017;52:214–27. https://doi.org/10.1016/j.neurobiolaging.2016.07.012.

37. Fagan AM, Head D, Shah AR, Marcus D, Mintun M, Morris JC, et al. Decreased cerebrospinal fluid Abeta(42) correlates with brain atrophy in cognitively normal elderly. Ann Neurol. 2009;65:176–83. https://doi.org/10.1002/ana.21559.

38. Fagan AM, Roe CM, Xiong C, Mintun MA, Morris JC, Holtzman DM. Cerebrospinal fluid tau/beta-amyloid(42) ratio as a prediction of cognitive decline in nondemented older adults. Arch Neurol. 2007;64:343–9. https://doi.org/10.1001/archneur.64.3.noce04123.

39. Palmqvist S, Mattsson N, Hansson O. Alzheimer's disease neuroimaging I. Cerebrospinal fluid analysis detects cerebral amyloid-beta accumulation earlier than positron emission tomography. Brain. 2016;139:1226–36. https://doi.org/10.1093/brain/aww015.

40. Palmqvist S, Zetterberg H, Blennow K, Vestberg S, Andreasson U, Brooks DJ, et al. Accuracy of brain amyloid detection in clinical practice using cerebrospinal fluid beta-amyloid 42: a cross-validation study against amyloid positron emission tomography. JAMA Neurol. 2014;71:1282–9. https://doi.org/10.1001/jamaneurol.2014.1358.

41. Mattsson N, Insel PS, Donohue M, Landau S, Jagust WJ, Shaw LM, et al. Independent information from cerebrospinal fluid amyloid-beta and florbetapir imaging in Alzheimer's disease. Brain. 2015;138:772–83. https://doi.org/10.1093/brain/awv367.

42. Jack CR Jr, Bennett DA, Blennow K, Carrillo MC, Dunn B, Haerlelein SB, et al. NIA-AA research framework: toward a biological definition of Alzheimer's disease. Alzheimers Dement. 2018;14:535–62. https://doi.org/10.1016/j.jalz.2018.02.018.

43. Ilhan-Gala I, Pegueroles J, Montal V, Vilaplana E, Carmona-Iragui M, Alcolea D, et al. Challenges associated with biomarker-based classification systems for Alzheimer’s disease. Alzheimers Dement Diagn Assess Dis Monit. 2018;10:346–57.

44. Paterson RW, Toombs J, Slattery CF, Nicholas JM, Andreasson U, Magdalinou NK, et al. Dissecting IWG-2 typical and atypical Alzheimer's disease: insights from cerebrospinal fluid analysis. J Neurol. 2015;262:2722–30. https://doi.org/10.1007/s00415-015-7904-3.

45. Leuty A, Chiotis K, Hasselbalch SG, Rinne JO, de Mendonca A, Otto M, et al. Pittsburgh compound B imaging and cerebrospinal fluid amyloid-beta in a multicentre European memory clinic study. Brain. 2016;139:2540–53. https://doi.org/10.1093/brain/aww160.

46. Janelidze S, Zetterberg H, Mattsson N, Palmqvist S, Vanderstichele H, Lindberg O, et al. CSF Abeta42/Abeta40 and Abeta42/Abeta38 ratios: better diagnostic markers of Alzheimer disease. Ann Clin Transl Neurol. 2016;3:154–65. https://doi.org/10.1002/acn3.274.

47. Hansson O, Seibyl J, Stomrud E, Zetterberg H, Trojanowski JQ, Bittner T, et al. CSF biomarkers of Alzheimer's disease concord with amyloid-beta PET and predict clinical progression: a study of fully automated immunoassays in BioFINDER and ADNI cohorts. Alzheimers Dement. 2018;14:1470–81. https://doi.org/10.1016/j.jalz.2018.01.010.

48. Barthel H, Sabri O. Clinical use and utility of amyloid imaging. J Nucl Med. 2017;58:1711–7. https://doi.org/10.2967/jnumed.116.185017.

49. Johnson KA, Minoshima S, Bohnen NI, Donohoe KJ, Foster NL, Herscovitch P, et al. Appropriate use criteria for amyloid PET: a report of the Amyloid Imaging Task Force, the Society of Nuclear Medicine and Molecular Imaging, and the Alzheimer's Association. Alzheimers Dement. 2013;9:E1–16. https://doi.org/10.1016/j.jalz.2013.01.002.

50. Donohue MC, Sperling RA, Petersen R, Sun CK, Weiner MW, Aisen PS, et al. Association between elevated brain amyloid and subsequent cognitive decline among cognitively normal persons. JAMA. 2017;317:2305–16. https://doi.org/10.1001/jama.2017.6669.

51. Prichep LS, John ER, Ferris SH, Rausch L, Fang Z, Cancro R, et al. Prediction of longitudinal cognitive decline in normal elderly with subjective complaints using electrophysiological imaging. Neurobiol Aging. 2006;27:471–81. https://doi.org/10.1016/j.neurobiolaging.2005.07.021.

52. Stinton C, McKeith I, Taylor JP, Laforêt L, Mioshi E, Mak E, et al. Pharmacological management of Lewy body dementia: a systematic review and meta-analysis. Am J Psychiatry. 2015;172:731–42. https://doi.org/10.1176/appi.ajp.2015.14121582.

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.