A case of early neonate bovine tuberculosis in Ethiopia

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Abstract
This report illustrates that calves may be infected with bovine tuberculosis at early age under natural conditions and progression can be rapid. Thus, testing of calves needs to be considered in any control program to reduce the risk of transmission.

KEYWORDS
bovine neonate, ethiopia, Mycobacterium bovis, tuberculous lesion

1 | INTRODUCTION

Bovine tuberculosis is a chronic disease rarely observed in an early age. Our observation of tubercle lesions in the lungs of a three-week-old calf and confirmation of Mycobacterium bovis strains illustrate that the progression of tuberculosis in neonates can be rapid under natural conditions, contributing to transmission within herds.

Bovine tuberculosis (bTB) is a chronic disease, caused primarily by M bovis, affecting cattle, other domesticated animals, certain wildlife species, nonhuman primates, and humans.1-3 Cattle can be infected at any age, but tubercle lesion development and subsequent clinical signs may appear at a later stage. Calves can be born with tuberculosis congenitally when infected with M bovis either via the umbilical vein from an infected genital tract of the dam, from the infected placenta, or from infected amniotic fluid.4 After birth, neonates can also be infected even at an early stages following delivery, mainly by inhaling infected droplets and/or by ingesting raw milk or colostrum from infected cows. Other

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routes of infection such as cutaneous, though not common, are still important in situations where the prevalence of bTB is high.\(^5\) As the course of the disease is slow compared with many other infectious diseases, infected animals can spread bTB to other herd mates and newborns before it begins to manifest clinical signs. The transmission rate may be faster at late stages of infection as the number of organisms being shed is larger in this stage.\(^1\)

The observation of characteristic tubercle lesions in the lungs of a three-week-old male calf led us to raise questions like “how the calf could get the infection?” and “how fast is the disease progression under natural infection?” In order to get answers to these questions, further investigation was made and this case report was compiled from the pathological and molecular investigations conducted for the diagnosis of the case.

### 2 | CASE HISTORY

A dairy farm located in northern Ethiopia and with a herd size of 100 cattle (86 crossbreeds of Holstein-Frisian and Zebu breeds and 14 Zebu breeds) was screened for bTB by the single intradermal cervical comparative tuberculin (SICCT) test. As it was learned from its history, the farm was newly established just a year earlier through the purchase of cows and heifers from dairy farms and markets in Mekelle City and nearby towns, including Adigrat and Humera. The SICCT testing was conducted on the herd in 2016 by Mekonnen et al,\(^6\) and a prevalence of 12% (95% CI: 7%-19.8%) was recorded. One of the reactors was a three-week-old calf (ID 8A0597), and its skin induration was 8 mm according to the calculation of the SICCT test result. In the same week, the SICCT test was conducted, all positive reactors (including the calf) were culled following disclosure of the test results to make the herd clear from bTB. It was observed from the husbandry of the farm that all calves in the farm were sharing the same housing with the adults. Moreover, the calves were fed on pooled milk obtained from milking cows in the herd including one SICCT reactor cow and looked after by the same individuals who were looking after the adult cattle.

The three-week-old calf positive for the SICCT test drew our attention. The calf had moderate body condition, but it was depressed and characterized by hair loss on various parts of its body. The dam of the calf (ID 8A0417) was a 10 years old cross of Holstein-Frisian and Zebu breeds. The body condition of this cow was rated as “Good,” and it was negative for bTB by the SICCT test according to the interpretation recommended by OIE.\(^7\) In addition, nasal discharge was collected from the dam using a cotton swab designed for such purpose, and the PBS wash of the swab was centrifuged at \(-3100\) g for 15 min. Genomic DNA was extracted from the pellet using a Qiagen DNA extraction kit as per the manufacturer’s instruction,\(^8\) and the eluent tested by an RD4-based PCR assay for specific identification of \(M\) bovis. The primers defined by Brosch et al\(^9\) were used in the PCR assay procedure described by Berg et al\(^10\)

### 3 | INVESTIGATION

During postmortem inspection of the calf, multiple granulomas were observed in the apical lobe of the lungs (Figure 1). A single \(-20\) mm diameter of a loosely encapsulated granuloma-like structure was also observed in the diaphragmatic lobe. Lymph nodes and other vital organs appear normal.

The culturing result (Löwenstein-Jensen medium slant) from lung tubercles of the calf showed two strains of typical \(M\) bovis. Heat-killed colonies were tested by RD4-based PCR and confirmed that both strains were \(M\) bovis. Spoligotyping\(^11\) then classified these two strains into different spoligotype patterns, SB0134 and SB0133 (Figure 2). Genomic DNA extracted from the dam’s nasal discharge was positive for \(M\) bovis by the RD4-based PCR assay; however, spoligotyping was not sensitive enough to confirm the pattern, and the DNA concentration was too low for MIRU-VNTR typing or whole genome sequencing. Attempt to culture the agent from the nasal swab sample of the dam was not successful.

### 4 | DISCUSSION

*Mycobacterium bovis* strains identified in the present case, classified as spoligotypes SB0134 and SB0133, suggesting mixed infection. Although mixed infection has not been frequently described in Ethiopia, it is more likely to happen in herds which have established their stocks from different farms, as was the case for the present herd. Hence, the mixed infection observed in this calf could be due to transmission from more than one reactor, possibly recruited from different herds without prior testing for bTB.

The time-frame for development of tubercle lesions in the investigated calf is comparable with artificially infected calves via respiratory route under experimental conditions.\(^12\) However, under such conditions; the extent of pathology could vary depending on the infective dose and factors specific to the animals and husbandry systems.

Evidence has shown that the route of transmission of \(M\) bovis could be inferred from lesions distribution.\(^13\) In the present calf, tubercle lesions were localized in the lungs and no tubercle lesions were observed in other tissues and organs, which could suggest inhalation of infectious droplets as a possible route of infection while ingestion of infected colostrum or milk might seem less likely. Other possibilities of in utero transmission through hematogenous route and/or aspiration of \(M\) bovis contaminated amniotic fluid\(^14,15\) could not
be justified as there was no lesion suggestive of bTB in other vital organs, such as liver. Tuberculosis transmitted hematogenously is typically characterized by the presence of hepatic tubercles or more disseminated form of the disease,\textsuperscript{16} while through aspiration of infected amniotic fluid, the primary tubercle could be in the lungs and/or the gut.

The molecular test result implies that the dam was infected with \textit{M. bovis} even though it was not a reactor to the SICCT test (the increase at the PPD-B injection site was 3.2 mm while at the PPD-A site it was 2.9 mm). This could potentially be due to a development of an anergic status, in which the dam would either had been in a late disease stage or immunosuppressed attributable to the hormonal effects during pre- and post-parturition. An animal with a chronic stage of the disease is known to be highly infectious and would likely serve as a good source of infection to a neonate by transmission in several possible ways.\textsuperscript{17,18} Whether or not this was the case for the observed dam, it is still likely that her calf got infected in its first week of life, while suckling the colostrum and/or inhaling the aerosol produced by her. Attempt to generate strong evidence to establish transmission from the dam has not been successful due to failure to culture \textit{M. bovis} from the nasal swab sample; resampling of the dam was not possible as the animal was later culled without the knowledge of the authors. Despite the proximity and connection between the dam and its calf, the possibility of transmission from other neighboring infected animals could not be ruled out as all
reactors in this farm shared the same housing and husbandry system; this could also explain why two strains of different spoligotype patterns were isolated from its lungs.

In conclusion, although this is a single case, it illustrates that the progression of bTB in calves can be rapid under natural conditions, contributing to transmission within herds. This has important implications in the appropriate structure of models used to estimate the rate of transmission within herds. Calf can respond to the tuberculin test at a very early age and thus should be considered in the routine bTB screening to reduce transmission within herds.

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CONFLICT OF INTEREST
The authors declare that they have no competing interests.

AUTHORS’ CONTRIBUTION
GAM performed the investigation, analyzed the data, and wrote the report. BG, SB, AJKC, GA, and JLNW contributed to the study conception and design and reviewed the report.

ETHICAL APPROVAL
This study was under the framework of a research protocol approved by the National Research Ethics Review Committee - NRERC No. 3.10/800/07.

DATA AVAILABILITY STATEMENT
All data generated and/or analyzed during this study are not publicly available but are available from the corresponding author on reasonable request.

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REFERENCES
1. CFSPH. Bovine tuberculosis Fact sheet. Center for Food Security and Public Health (CFSPH). Iowa State University; 2009.
2. Cousins DV Mycobacterium bovis infection and control in domestic livestock. Rev Sci Tech Off Int Epiz. 2001;20:71-85.
3. More S, Bøtner A, Butterworth A, et al. Assessment of listing and categorization of animal diseases within the framework of the Animal Health Law (Regulation (EU) No 2016/429): bovine tuberculosis. EFSA - DOPTED: 30 June 2017; 2017. https://doi.org/10.2903/j.efsa.2017.4959
4. Ozzyigit MO, Senturk S, Akkoc A. Suspected congenital generalized tuberculosis in a newborn calf. Vet Rec. 2007;160:307-308.
5. Phillips CJ, Foster CR, Morris PA, Teverson R. The transmission of Mycobacterium bovis infection to cattle. Res Vet Sci. 2003;74:1-15.
6. Mekonnen GA, Conlan AJK, Berg S, et al. Prevalence of bovine tuberculosis and its associated risk factors in the emerging dairy belts of regional cities in Ethiopia. Prev Vet Med. 2019;168:81-89.
7. OIE. Bovine tuberculosis. OIE-manual of diagnostic tests and vaccines for terrestrial animals. Paris; 2009.
8. Qiagen. QIAamp DNA Mini and Blood Mini Handbook, 5th Edn. Sample and Assay Technologies; 2016. http://www.Qiagen.com. Accessed March 30, 2019
9. Brosch R, Gordon SV, Marmiesse M, et al. A new evolutionary scenario for the Mycobacterium tuberculosis complex. PNAS - USA. 2002;99:3684-3689.
10. Berg S, Firdessa R, Habtamu M, et al. The burden of mycobacterial disease in Ethiopian cattle: implications for public health. PLoS One. 2009;4:e5068. https://doi.org/10.1371/journal.pone.0005068
11. Kamerbeek J, Schouls L, Kolk A, et al. Simultaneous detection and strain differentiation of Mycobacterium tuberculosis for diagnosis and epidemiology. J Clin Microbiol. 1997;35:907-914.
12. Yao RR, Gravenor MB, Charleston B, Hope JC, Martin M, Howard CJ. Mycobacterium bovis shedding patterns from experimentally infected calves and the effect of concurrent infection with bovine viral diarrhoea virus. J R Soc Interface. 2007;4:545-551.
13. Francis J. Susceptibility to tuberculosis and the route of infection. Aust Vet J. 1971;47:414.
14. Vural SA, Tunca R. Generalized tuberculosis in a 45 day-old calf. Dtsch Tierarztl Wochenschr. 2001;108:468-470.
15. Espejel MDC, Del Rio JC, Medrano A, Santillan MA, Morales JF. Congenital tuberculosis in a 25-day-old female calf. Turk J Vet Anim Sci. 2018;42:135-138.
16. Tomar RPS, Gupta A, Prasad TR, Bhalla PJS, Murthy GSN. Congenital tuberculosis. Med J Armed Forces India. 2008;64:78-80.
17. Raj P, Sarin YK. Congenital tuberculosis in a neonate: a diagnostic dilemma. J Neonat Surg. 2014;3:49.
18. Yeh J-J, Lin S-C, Lin W-C. Congenital tuberculosis in a neonate: A case report and literature review. Front Pediatr. 2019;7:255.
19. Conlan AJK, McKinley TJ, Karolemeas K, et al. Estimating the hidden burden of bovine tuberculosis in Great Britain. PLoS Comput Biol. 2012;8:e1002730.

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