A phase I study of BI 811283, an Aurora B kinase inhibitor, in patients with advanced solid tumors

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Abstract

Purpose This phase I study investigated the maximum tolerated dose (MTD), safety, pharmacokinetics, pharmacodynamics, and antitumor activity of the Aurora B kinase inhibitor BI 811283 in patients with advanced solid tumors.

Methods BI 811283 was administered via 24-h infusion on Days 1 and 15 of a 4-week cycle (schedule A) or Day 1 of a 3-week cycle (schedule B) in a modified 3+3 dose-escalation design. Pharmacodynamic assessments included immunohistochemistry for phosphorylated histone H3 (pHH3) on skin biopsies to determine Aurora B kinase inhibition and plasma concentrations of caspase-cleaved CK-18 (apoptosis marker).

Results A total of 121 patients were treated. The MTDs of BI 811283 were 125 mg (schedule A) and 230 mg (schedule B). Dose-limiting toxicities were primarily hematological (febrile neutropenia and grade 4 neutropenia); the most common drug-related adverse effects included neutropenia, fatigue, leukopenia, nausea, alopecia, diarrhea, and decreased appetite. A trend toward a decrease in pHH3 was observed, with increasing BI 811283 doses, indicating target engagement; there was no consistent trend regarding caspase-cleaved CK-18 levels. No objective response was observed although 19 patients in each schedule achieved clinical benefit (stable disease).

Conclusions BI 811283 demonstrated a generally manageable safety profile and disease stabilization in some patients.

Trial registration EudraCT No: 2007-000191-17, ClinicalTrials.gov Identifier: NCT00701324.

Keywords Aurora B kinase · BI 811283 · Phase I · Mitosis modulators · Solid tumors

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Background

The Aurora kinases belong to a family of three serine/threonine protein kinases (A, B, and C) that have a prominent role in regulating cell division [1, 2]. Aurora B kinase is part of the chromosome passenger complex (CPC), a group of proteins identified by its change in cellular localization over the duration of the cell cycle [3, 4]. Aurora B kinase has an important role in regulating many aspects of mitotic cell division, such as chromosome bi-orientation and sister chromatid cohesion, where Aurora B kinase is associated with the centromeres [5, 6], and spindle disassembly and cytokinesis following re-localization of the CPC to the midbody [7]. Furthermore, Aurora B kinase assists with chromosome condensation via phosphorylated histone H3 (pHH3) [8, 9].

There is considerable evidence linking Aurora B kinase to tumorigenesis. An increased expression of Aurora B kinase promotes chromosome instability and aneuploidy in vitro [10], and cells overexpressing Aurora B kinase have been shown to form aggressive tumors in nude mice [11]. In vivo, overexpression of Aurora B kinase is associated with several different tumor types when compared with matched normal tissue, such as brain, thyroid, breast, lung, colorectal, and prostate cancers, and is linked with characteristics including genetic instability, disease progression, and poor outcome in these patients [12–19]. In vitro, cells lacking Aurora B kinase cannot complete cytokinesis, become tetraploid and subsequently die [20]. This, together with the essential functions of Aurora B kinase during cell replication and its elevated expression in many cancer cell types, suggests that this mitotic enzyme is a valid target for therapeutic intervention.

There are multiple Aurora kinase inhibitors in preclinical studies or clinical trials. These agents are either pan-Aurora kinase inhibitors (targeting Aurora A, B, and occasionally C), or specific for either Aurora A or Aurora B [21].

BI 811283 is an adenosine triphosphate-competitive, reversible, and potent inhibitor of Aurora B kinase (half-maximal inhibitory concentration [IC₅₀], 9 nM) [22, 23]. In a preclinical, in vitro study, BI 811283 exhibited broad antiproliferative activity in 24 tumor cell lines of different tissue origin (half-maximal effective concentration [EC₅₀], <14 nM) [22, 23]. Within 1 h of treatment, pHH3 was reduced, indicating Aurora B kinase inhibition. Polyploidy was observed in up to 80% of cells 48 h following treatment, with a subset of cells subsequently becoming senescent and going through apoptosis [22, 23]. In studies utilizing in vivo xenograft models of human non-small cell lung cancer, colon carcinoma, and pancreas carcinoma, inhibition of Aurora B kinase with BI 811283 resulted in a dose-dependent inhibition of tumor growth, and tumor regression was observed in some cases. A decrease in pH3 was also observed, acting as a marker of Aurora B kinase inhibition [24].

Here we report the findings of a phase I, dose-escalation trial of BI 811283 that was performed in patients with advanced solid tumors. Two dosing schedules were examined, with the maximum tolerated dose (MTD) determined for each. The safety, pharmacokinetics, pharmacodynamics, and antitumor activity of BI 811283 are also described.

Methods

Patient selection

Eligible patients were aged 18 years or older presenting with advanced, non-resectable and/or metastatic solid malignant tumors, who were either not amenable to established forms of treatment or for whom no therapy of proven efficacy was available, and with a life expectancy of ≥6 months. Further eligibility criteria included an Eastern Cooperative Oncology Group performance status (ECOG PS) of ≤2; recovery from reversible toxicities of previous anticancer therapies; evaluable tumor deposits; secure central venous access; adequate bone marrow, liver, and renal function [absolute neutrophil count ≥1500/mm³, platelet count ≥100 000/mm³, bilirubin ≤1.5 mg/dL (≤26 μmol/L, SI unit equivalent), aspartate amino transferase and/or alanine amino transferase ≤2.5 × upper limit of normal (ULN; if related to liver metastases then ≤5 × ULN), and serum creatinine ≤1.5 mg/dL (≤132 μmol/L, SI unit equivalent)]; no chemo-, radio-, immuno-, hormone-, or investigational therapy within 2 weeks prior to the start of treatment with the trial drug; no known brain metastases or second malignancy requiring therapy, and no serious illness or concomitant disease which could compromise patient safety (including clinically significant cardiovascular disease, left ventricular ejection fraction <50%, myocardial infarction within the last 6 months prior to inclusion and/or symptomatic coronary artery disease). Written informed consent was obtained from all study participants.

Study design and dose escalation

This was an open-label, first-in-human, phase I, dose-escalation trial of BI 811283 in patients with advanced solid tumors, conducted at two sites in Germany (EudraCT No: 2007-000191-17, ClinicalTrials.gov Identifier: NCT00701324). The primary endpoint was determination of the MTD of BI 811283 administered as a 24-h continuous infusion in a 4- or 3-week schedule. Several secondary endpoints were examined concurrently: the incidence and intensity of adverse events [AEs; determined by the common terminology criteria for AEs (Common Terminology Criteria for Adverse Events (CTCAE) version 3.0)], incidence of dose-limiting toxicity (DLT), objective response
The trial was conducted in accordance with the principles laid down by the Declaration of Helsinki and approved by the independent ethics committees and/or institutional review boards of the participating centers.

**Definition of dose-limiting toxicity**

A DLT was recorded if one or more of the following events occurred during the first treatment cycle: (1) drug-related grade $\geq 3$ non-hematological toxicity (except untreated nausea, vomiting, or diarrhea), (2) drug-related grade 4 neutropenia for $\geq 7$ days or febrile neutropenia, or (3) drug-related grade 4 thrombocytopenia or anemia. Additionally, for schedule A only, the following events on Day 15, if drug related, also constituted a DLT: (1) grade $\geq 2$ non-hematological toxicity (excluding alopecia and untreated nausea, vomiting, or diarrhea), or (2) grade $\geq 3$ neutropenia, thrombocytopenia, or anemia. The MTD was defined on the basis of DLT observed during the first treatment cycle only. However, DLTs observed after that time period were considered along with the type, number, and intensity of AEs to indicate how well BI 811283 was tolerated.

**Study assessments**

**Safety**

All treated patients were included in the safety evaluation. Key safety measures included evaluation of the incidence and intensity of AEs using the medication dictionary for regulatory activities (MedDRA) and classified according to CTCAE version 3.0, assessment of vital signs and laboratory parameters, and 12-lead electrocardiogram (ECG) and echocardiography measurements.

**Pharmacokinetics**

Blood samples for the evaluation of pharmacokinetic parameters were collected 5 min before drug administration, at 1, 2, 4, 10, and 20 h during the infusion, at 23:59 (just before the end of infusion), and at the following timepoints after the start of infusion: 24:15, 24:30, 25:00, 26:00, 28:00 and 32:00 h (Day 2), 48:00 h (Day 3), 72:00 h (Day 4), and 120 h (Day 6). Further samples were obtained in each subsequent treatment cycle (up to Cycle 6) just prior to the start and at the end of the infusion. Urine samples were also obtained after the first and second infusions of BI 811283 (schedule A: Cycle 1 Days 1 and 15; schedule B: Cycle 1 Days 1 and 22). The concentration of BI 811283 in plasma and urine was determined by a validated high-performance liquid chromatography–tandem mass spectrometry (HPLC–MSMS) assay. Briefly, samples were subjected to protein precipitation with methanol. Then, the
samples were analyzed on a PerkinElmer Sciex API 5500 LC-MSMS system using a Zorbax ® Eclipse Plus C18 column. An electrospray ion source [atmospheric pressure ionization (API)] was used for ionization. Measurements were performed in the positive ionization mode. Preclinical assessments demonstrated that BI 811283 protein binding is dependent on the plasma concentrations of alpha-1-acid glycoprotein (AGP) (Boehringer Ingelheim, data on file). Therefore, AGP concentrations were determined in clinical plasma samples following BI 811283 dosing and were obtained during the first two treatment cycles at the same time points as those collected for pharmacokinetic analysis, with AGP concentration determined using a validated immunoturbidimetric assay. In the assay, AGP was precipitated using a specific antiserum and the resulting turbidimetric reading at 340 nm was used to determine its concentration. Pharmacokinetic analyses were summarized using descriptive statistics.

Pharmacodynamics

Target inhibition following BI 811283 dosing was assessed. Skin biopsies were analyzed for a reduction of pHH3, a marker of Aurora B kinase inhibition [26]. Three- to four-millimeter punch skin biopsies were obtained during screening and within 6 h after the end of infusion on Day 16 of schedule A or Day 2 of schedule B of the first treatment cycle. The skin biopsies were divided, with one half each for western blotting and immunohistochemistry (IHC). For western blotting, the sample was lysed in Lysis Matrix A (MP Biomedicals #6910-050) and lysis buffer [100 mM NaHCO3 pH 9.6, 1 mM dithiothreitol, Protease inhibitor cocktail (Serva 3910)]. Equivalent amounts of proteins from the cell lysates were applied to 12 % TRIS–glycine gels (Invitrogen NuPAGE Novex NP0341BOX) and blotted with phospho-H3 antibody (Cell Signalling #9701) and H3 antibody (Cell Signalling #9715) to determine the ratio of phospho-H3/total H3 protein. For IHC analysis, the sample was fixed in formalin, embedded into paraffin with 5-µm sections prepared. The slides were probed with phospho-H3 antibody (Cell Signalling #9701) and counterstained with Mayer’s hemalum (Merck 1.09249.1000). Epidermal areas were marked and measured, and positive nuclei counted with the aid of the NIKON image software. Quantification was done in a blinded fashion by two independent researchers.

Serum levels of caspase-cleaved cytokeratin-18 (CK-18) act as a marker of the cell apoptosis anticipated with Aurora B kinase inhibition by BI 811283 [27]. Therefore, plasma samples for quantifying the caspase-cleaved fragment of CK-18 were obtained pre-dose and 48, 72, and 120 h after the first two BI 811283 administrations in each schedule. Samples were also collected pre-dosing and 120 h post-dosing, up to Cycle 6. Plasma concentrations of caspase-cleaved CK-18 were determined using a validated ELISA.

Antitumor activity

Objective tumor response was evaluated using computed tomography (CT) or magnetic resonance imaging (MRI) scans at baseline and at the end of every other treatment cycle, according to RECIST criteria version 1.0 [25].

Results

Patient demographics and disposition

A total of 129 patients were enrolled into the study which was performed between July 2007 and August 2011. Of these, 124 patients were entered to one of the two treatment schedules and 121 were treated with study medication (schedule A: 63 patients; schedule B: 58 patients). Patient demographics and disease characteristics are summarized in Table 1. Median patient age was 58 years (range 23–79 years), and 47.1 % of patients were male. The majority of patients (77.7 %) presented with stage IV disease at screening, with colorectal cancer being the most frequent tumor type (24.8 %). Most patients had undergone previous surgery (90.9 %) or systemic chemotherapy (92.6 %), and many were heavily pre-treated: 57 (47.1 %) had received ≥4 lines of chemotherapy (Table 1).

Dose-limiting toxicities, safety, and tolerability

Patients in the schedule A cohort were treated with BI 811283 at doses from 5 to 140 mg. First-cycle DLTs were primarily hematological events; one patient treated with 10 mg had raised liver function tests (Table 2). DLTs were seen in two of three patients treated with the 140 mg dose. The MTD was determined to be 125 mg; one patient in the initial cohort at this dose experienced grade 4 neutropenia on Day 15. Patients in the schedule B cohort were treated with BI 811283 starting at 13.5 mg and escalated to 300 mg. As with schedule A, DLTs were primarily hematological events; one patient treated with 105 mg had raised liver function tests (Table 2). DLTs were seen in two of three patients treated with the 140 mg dose. The MTD was determined to be 230 mg. During the data review meeting following the end of the study, five events of grade 4 neutropenia in schedule B were retrospectively identified as possible DLTs (105 mg, n = 2; 125 mg, n = 1; 180 mg, n = 1; 230 mg, n = 1). In all five patients, the initial hematology laboratory tests performed during Cycle 1 showed grade 4 neutropenia. However, there were no laboratory
tests performed on Day 7 or 8 after the start of the grade 4 neutropenia to show whether neutropenia was still present to qualify as a DLT (i.e., \( \geq 7 \) days), although in all of these cases the neutropenia had resolved by Day 8–10 after the start of the grade 4 event. Five additional DLTs [grade 4 febrile neutropenia (\( n = 2 \)), grade 3 febrile neutropenia (\( n = 2 \)), and grade 4 neutropenia \( \geq 7 \) days (\( n = 1 \))] were also identified in the 230-mg dose expansion cohort of seven patients. Therefore, although per protocol the MTD was determined as 230 mg for schedule B, based on retrospective DLTs from Cycle 1 and in patients enrolled in the expansion cohort, a lower dose may have been recommended for further study. Expansion cohorts included nine additional patients in schedule A, and seven additional patients in schedule B treated at the respective MTDs (125 and 230 mg). All patients in both treatment schedules had at least one AE during the treatment course, with 100 (82.6 %) patients having AEs that were considered drug related by the investigators [schedule A: \( n = 49 \) (77.8 %); schedule B: \( n = 51 \) (87.9 %)]. Drug-related AEs that occurred in \( \geq 10 \) % of all patients across all treatment cycles were neutropenia (\( n = 40 \), 33.1 %), fatigue (\( n = 38 \), 31.4 %), leukopenia (\( n = 35 \), 28.9 %), nausea (\( n = 29 \), 24.0 %), alopecia (\( n = 24 \), 19.8 %), diarrhea (\( n = 20 \), 16.5 %), and decreased appetite (\( n = 18 \), 14.9 %). Tables 3 and 4 summarize the most common drug-related AEs in schedules A and B by dose. Serious AEs (SAEs) were reported in 61 (50.4 %) patients [schedule A: \( n = 33 \) (52.4 %); schedule B: \( n = 28 \) (48.3 %)]. Thirteen of these patients had SAEs regarded as drug related [schedule A: \( n = 3 \) (4.8 %); schedule B: \( n = 10 \) (17.2 %)]. The most common drug-related SAE was febrile neutropenia (experienced by seven patients);
Table 2  DLTs observed during the first cycle in both treatment schedules

| Schedule A | Schedule B |
|------------|------------|
| **BI 811283 dose (mg)** | **n** | **Patients with DLT** | **DLT (n)** | **Patients with retrospectively identified potential DLT** | **DLT (n)** |
| 5          | 3        | 0                        | –              | –                        | –              |
| 10         | 7        | 1 Grade 3 AST increased and grade 3 ALT increased (1) | –              | –                        | –              |
| 13.5       | 3        | 0                        | –              | –                        | –              |
| 18         | 3        | 0                        | –              | –                        | –              |
| 24         | 4        | 0                        | –              | –                        | –              |
| 32         | 3        | 0                        | –              | –                        | –              |
| 43         | 5        | 0                        | –              | –                        | –              |
| 58         | 3        | 0                        | –              | –                        | –              |
| 78         | 7        | 1 Grade 3 neutropenia and grade 3 thrombocytopenia (1) | –              | –                        | –              |
| 105        | 6        | 1 Grade 3 hemoglobin decreased (1) | –              | –                        | –              |
| 125        | 16       | 1 Grade 4 neutropenia (1) | –              | –                        | –              |
| 140        | 3        | 2 Grade 3 neutropenia (1); grade 4 neutropenia (1) | –              | –                        | –              |

| BI 811283 dose (mg) | n | Patients with DLT | DLT (n) | Patients with retrospectively identified potential DLT | DLT (n) |
|--------------------|---|-------------------|---------|--------------------------------------------------------|---------|
| 13.5               | 3 | 0                 | –       | –                                                      | –       |
| 18                 | 3 | 0                 | –       | –                                                      | –       |
| 24                 | 3 | 0                 | –       | –                                                      | –       |
| 32                 | 4 | 0                 | –       | –                                                      | –       |
| 43                 | 3 | 0                 | –       | –                                                      | –       |
| 58                 | 3 | 0                 | –       | –                                                      | –       |
| 78                 | 3 | 0                 | –       | –                                                      | –       |
| 105                | 6 | 1c Grade 3 fatigue (1) | 2       | Grade 4 neutropenia ≥7 days (2); Grade 4 neutropenia ≥7 days and grade 4 leukopenia (1) | –       |
| 125                | 3 | 0c                | –       | 1                                                      | –       |
| 150                | 3 | 0                 | –       | –                                                      | –       |
| 180                | 3 | 0c                | –       | 1                                                      | –       |
| 230                | 3 | 0c                | –       | 1                                                      | –       |
| 300                | 3 | 2 Grade 4 febrile neutropenia (1); grade 3 febrile neutropenia (1) | –       | –                                                      | –       |
| 230                | 3 | 0                 | –       | –                                                      | –       |
| 270                | 5 | 2 Grade 4 neutropenia ≥7 days (2) | –       | –                                                      | –       |
| 230a Expansion cohort | 7 | 5 Grade 4 neutropenia ≥7 days (1); grade 4 febrile neutropenia (2); grade 3 febrile neutropenia (2) | –       | –                                                      | –       |

* ALT alanine aminotransferase, AST aspartate aminotransferase, DLT dose-limiting toxicity, MTD maximum tolerated dose

* Defined as the maximum tolerated dose

* Seven patients initially plus nine patients in the expansion cohort

* Upon data review at the end of the study, two events in the schedule B 105 mg cohort (grade 4 neutropenia ≥7 days, n = 2) and one event (grade 4 neutropenia ≥7 days) in each of the following schedule B cohorts: 125, 180, and 230 mg were retrospectively determined to be DLTs, although they were not considered to be DLTs during MTD determination. These events were considered to have potentially met the study protocol definition for a DLT, but there was not sufficient information to conclusively determine their exact duration (i.e., no laboratory tests were performed on Day 7 or 8 after the start of the grade 4 neutropenia). The investigators and sponsor used a worst case/conservative approach and assumed that these were DLTs.
other such events reported in more than one patient were leukopenia, neutropenia, vomiting, and diarrhea. A total of 22 (18.2%) patients had fatal AEs [schedule A: n = 10 (15.9%); schedule B: n = 12 (20.7%)]. None of the deaths were considered related to the study treatment. Overall, 19 (15.7%) patients had AEs that resulted in study drug discontinuation or a dose reduction [schedule A: n = 9 (14.3%); schedule B: n = 10 (17.2%); Supplementary Table 1]. In 10 patients (five patients in each dosing schedule), the AEs resulting in discontinuation or dose reduction were classified as significant according to the International Conference on Harmonization (ICH) E3.

**Pharmacokinetics**

Plasma concentrations of BI 811283 rose rapidly for several hours after the start of the 24-h infusion, reaching maximum concentrations after 20–24 h (Fig. 1). This was followed by a biphasic decline in plasma BI 811283, which was initially very rapid such that most of the compound was eliminated within the first 6 h following the end of the infusion. The mean terminal half-life of BI 811283 was 11.4–30.5 h for schedule A and 10.1–27.0 h for schedule B. In general, pharmacokinetic parameters were comparable between the two schedules and there were no significant differences between the first and second doses given to patients, regardless of treatment schedule. Although there was high inter-subject variability for both treatment schedules, the maximum measured concentration in plasma ($C_{\text{max}}$) and the area under the concentration–time curve over the time interval from 0 extrapolated to infinity (AUC$_{0-\infty}$) values increased in a dose-dependent manner. The fraction of BI 811283 excreted in urine was low (ranging from 4 to 12% of the administered dose) and did not differ between the two treatment arms. AGP plasma concentrations did not change significantly with time after BI 811283 dosing and were not dependent on BI 811283 concentrations. There was a trend toward increased exposure to total BI 811283 (bound and unbound) in patients with higher AGP plasma concentrations, although there was high variability.

**Pharmacodynamics**

In total, 28 out of 63 patients in schedule A [5, 10, 18, 24, 43, 125, and 140 mg ($n = 2$ each); 58 mg ($n = 3$); 78 mg ($n = 4$); 105 mg ($n = 5$)], and 28 out of 58 patients in schedule B [32 and 300 mg ($n = 1$ each); 13.5, 18, 43, 58, 105, 125 ($n = 2$ each); 150 mg ($n = 3$); 270 mg ($n = 4$); 230 mg ($n = 7$)] provided skin biopsies for pharmacodynamic analyses. Two different types of analyses were performed: an IHC determination of cells with nuclei that were positive for pHH3 and a western blot analysis to determine the ratio of pHH3 to total histone H3 in cells. Western blot
analyses resulted in a low to undetectable pH3 band, sometimes in combination with a very dominant globin band, which interfered with the pH3 signal. IHC analyses using formalin-fixed paraffin-embedded sections revealed a trend for a reduction in the number of phospho-histone H3-positive nuclei in the epidermis with increasing doses of BI 811283 from 58 mg upward in schedule A, and from 43 mg upward in schedule B [Fig. 2 and example images from a patient receiving schedule A treatment (Supplementary Fig 1)], suggesting Aurora B kinase inhibition in this patient population [11, 28]. There was no consistent trend regarding the level of caspase-cleaved CK-18 in plasma after infusion of BI 811283, no consistency between the first and second infusions, and data obtained from the effect curves were extremely variable (Supplementary Fig 2 and 3).

Antitumor activity

No patients in either treatment schedule achieved an objective response. However, 19 (30 %) patients in schedule A and 19 (33 %) patients in schedule B achieved clinical benefit from treatment [defined as stable disease (SD; n = 37) or non-evaluable but clinically not progressive disease (n = 1)]. To confirm a status of SD, an overall response assessment that met the SD criteria was to be recorded at least 42 days after study entry. Ten patients with SD received ≥ 10 cycles of treatment [tumor types: melanoma (n = 3), thyroid or parathyroid (n = 2), sarcoma (n = 2), breast (n = 1), lung (n = 1), and colorectal cancer (n = 1)].

Discussion

Novel treatment strategies for patients with advanced solid tumors are urgently needed. Although antimitotic drugs in the form of microtubule-targeting agents (MTAs), such as the taxanes, are one of the most widely used classes of cancer therapeutics, the broad and often toxic effects of these agents, together with the emergence of MTA resistance, have triggered interest in new strategies that involve selective inhibition of enzymes involved in the regulation of mitosis [29, 30]. Aurora B kinase is one such enzyme that plays several important roles in mitosis [2, 31, 32]. This fact, coupled with its elevated expression across a wide range of tumor types suggests that this enzyme could be an effective therapeutic target in oncology [33].

This phase I clinical trial was designed as an open, parallel group, first-in-human study to define the MTD of BI 811283, an inhibitor of Aurora B kinase, in patients with advanced solid tumors. The MTD of BI 811283 was determined as 125 mg on Days 1 and 15 of a 4-week cycle (schedule A) and 230 mg on Day 1 of a 3-week cycle.
Fig. 1 Geometric mean plasma concentrations of BI 811283 following 24-h intravenous infusion in schedule A (a) and schedule B (b) (linear scale)

Fig. 2 Effect of BI 811283 following 24-h intravenous infusion on histone H3 phosphorylation measured by immunohistochemistry in schedule A (a) and schedule B (b)
(schedule B). The most common DLTs observed in both schedules were primarily hematological events, particularly neutropenia. One patient with metastatic soft tissue sarcoma and no liver metastases treated with 10 mg BI 811283 in schedule A experienced a DLT of grade 3 liver enzyme (alanine and aspartate aminotransferase) elevation in Cycle 1 which resolved spontaneously within 5 days. She again developed grade 3/4 liver enzyme elevation following treatment on Cycle 2 Day 15 which did not improve sufficiently for her to receive subsequent treatment, and she discontinued from the study. Given the recurrence of this AE upon rechallenge of the drug, it was deemed almost certainly drug related. This was the only occurrence of dose-limiting liver enzyme elevation in the study. Such DLTs do not appear to be typical of this class of compound, although DLTs of liver enzyme elevations were observed with the Aurora A kinase inhibitor MLN8054, and development of this compound was ceased in favor of alisertib (MLN8237) [34]. The retrospective identification of additional AEs in schedule B which possibly fulfilled the criteria for DLTs and the additional DLTs identified in the expansion cohort of 230 mg in schedule B suggests that the MTD and recommended phase II dose for further study may have been less than 230 mg with schedule B. However, due to the lack of promising efficacy data in this study and the development of an oral Aurora kinase inhibitor [35–37] offering improved convenience to the patient compared with BI 811283 (which requires 24-h continuous infusion through a central line), the clinical development of BI 811283 was halted.

No patients achieved an objective response with BI 811283 in this study. The best overall response was SD in both treatment schedules, observed in approximately 30 % of patients, all of whom had previously been heavily pretreated. This is not unexpected; however, as this outcome is consistent with the cytostatic mechanism of action of this class of drugs and comparable with the efficacy data observed in phase I trials of other Aurora kinase inhibitors [35, 36, 38–41].

The overall safety profile was as to be expected in a population of patients with advanced cancer. In addition to the AEs typically associated with the underlying disease, the most common toxicities were hematological. BI 811283 is known to cause a transient inhibition of the proliferation of normal dividing cells in the bone marrow, resulting in a temporary decrease of blood cells and platelets. Inhibition of mucosal proliferation can also occur with BI 811283, leading to gastrointestinal symptoms such as nausea, diarrhea, and abdominal pain. These AEs are not considered unusual following treatment with a compound targeting rapidly dividing cells, and similar toxicities were observed in phase I trials of other Aurora kinase inhibitors [35, 36, 41].

Pharmacokinetic analyses showed that plasma concentrations of BI 811283 rose rapidly in patients during the first few hours of the 24-h infusion, with maximum concentrations generally reached 20–24 h after the start of the infusion. This was followed by a rapid, biphasic decline. Pharmacokinetic parameters did not differ between the two treatment arms and there were no meaningful differences in pharmacokinetic parameters of BI 811283 between the first and second doses given to patients, regardless of treatment schedule. The fraction of BI 811283 excreted in urine was low and did not differ between the two treatment arms. Although there was high variability, exposure of total BI 811283 appeared to increase in patients with higher AGP concentrations. However, unbound BI 811283 was not determined in this study, so the relationship to AGP exposure in this case is not known.

During mitosis, histone H3 is phosphorylated by Aurora kinase B on serine 10 [11, 28]. Aberrant pH3 is associated with chromosome instability and carcinogenesis [11, 26]. A reduction in phosphorylation on serine 10 indicates Aurora kinase B inhibition. Indeed in preclinical studies, BI 811283 reduced histone H3 phosphorylation in multiple cancer cell lines [22, 24]. In this exploratory study, pH3 was measured using both western blotting and IHC. Results from the western blot analyses were not considered meaningful due to a low to undetectable pH3 band, sometimes in combination with a very dominant globin band, which interfered with the pH3 signal. In contrast, IHC, which relies on single cell analysis, had a signal that was highly concentrated in the rare mitotic cells resulting in a signal-to-noise ratio that was much improved. The IHC analyses revealed a trend for a reduction in the ratio of cellular pH3 to total histone H3 with increasing doses of BI 811283 in the skin as surrogate tissue, suggesting a dose-dependent inhibition of Aurora B kinase in this patient population. Among the 10 patients who achieved SD for ≥10 cycles, skin biopsy samples from only four patients were evaluable (two patients from each treatment arm). Of these four patients, the biopsy samples from only two patients showed a pharmacodynamics effect; therefore, it is difficult to conclude whether the pharmacodynamics effect had any correlation with clinical outcome. Overall, these results were not unexpected, due to differences in molecular and physiological characteristics between tumor versus skin tissue, and these analyses provide encouraging clinical data regarding BI 811283 target engagement. There was no consistent increase seen in caspase-cleaved CK-18, a marker of tumor cell death [27] in plasma samples of patients treated with BI 811283. Therefore, caspase-cleaved CK-18 was not an informative clinical biomarker of Aurora B kinase inhibition by BI 811283 in this study.

There are several Aurora kinase inhibitors currently in clinical development, which either specifically target
Aurora A or Aurora B kinase, or demonstrate some activity toward both kinases [42]. Preliminary evidence from a phase II study indicates that alisertib (MLN8237), an oral Aurora A kinase inhibitor, has a generally manageable safety profile and results in durable disease control in multiple cancer types [37]. The Aurora A kinase inhibitor danusertib has shown an acceptable safety profile and promising clinical activity in patients with advanced hematological malignancies [43], but limited activity in phase I/II studies in advanced solid tumors [44, 45]. The most widely tested Aurora B kinase inhibitor is barasertib (AZD1152), which has been evaluated in both patients with advanced solid tumors and those with hematological cancers. While barasertib demonstrated limited activity in solid tumors [38, 46], greater activity has been observed in patients with acute myeloid leukemia (AML) [47–50]. In these patients, barasertib has demonstrated clinical activity in several phase I/II trials as monotherapy (hematological response rate of 19–25 %) and in combination with low-dose cytosine arabinoside (LDAC; response rate of 45 %). Grade ≥3 neutropenia/febrile neutropenia was reported as a common AE in these studies. Barasertib was also compared to LDAC in a randomized phase II trial in elderly patients with AML, demonstrating higher complete response rates (35.4 vs 11.5 %); however, barasertib was much less tolerable than LDAC in these patients (71 vs 15 % stomatitis; 67 vs 19 % febrile neutropenia) [47]. Another Aurora B kinase inhibitor, BI 831266, has also been recently studied in a phase I trial of patients with advanced solid tumors. Similar to the agent investigated in the current study, BI 831266 treatment resulted in objective response in only one patient (cervical cancer), with 16 % of patients experiencing SD [39]. The BI 831266 trial was discontinued based on these data, as well as the limited activity displayed by BI 811283 in this population.

Although all Aurora kinases are involved in cell division, only Aurora A regulates centrosome maturation/separation and bipolar spindle assembly, whereas Aurora B and C are involved in the regulation of mitotic chromosome dynamics [43]. Based on these different intracellular functions, it has been hypothesized that inhibition of multiple Aurora kinases may provide better antitumor activity. Agents that target both Aurora A and B kinases are currently in early-stage clinical development (e.g., PF-03814735, AT9283) in both solid tumors and hematological malignancies; however, these agents have shown limited antitumor activity thus far in phase I trials [40, 43].

Conclusions

In summary, this phase I trial demonstrated an acceptable safety profile with BI 811283 administered on two different treatment schedules. MTDs were determined as 125 mg (Days 1 and 15 of a 4-week cycle) and 230 mg (Day 1 of a 3-week cycle); however, the retrospective identification of additional potential AEs in schedule B during a data review at the end of the study suggests that the MTD may have been less than 230 mg. The limited antitumor activity observed with BI 811283 does not support its continued development in solid tumors. Further biomarker studies would be beneficial to better understand the role of Aurora kinase inhibition in tumor development and anticancer therapy.

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