REGISTERED REPORT PROTOCOL

Meals, Microbiota and Mental Health in Children and Adolescents (MMM-Study): A protocol for an observational longitudinal case-control study

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Abstract

Recent studies indicate that the interplay between diet, intestinal microbiota composition, and intestinal permeability can impact mental health. More than 10% of children and adolescents in Iceland suffer from mental disorders, and rates of psychotropics use are very high. The aim of this novel observational longitudinal case-control study, “Meals, Microbiota and Mental Health in Children and Adolescents (MMM-Study)” is to contribute to the promotion of treatment options for children and adolescents diagnosed with mental disorders through identification of patterns that may affect the symptoms. All children and adolescents, 5–15 years referred to the outpatient clinic of the Child and Adolescent Psychiatry Department at The National University Hospital in Reykjavik, Iceland, for one year (n≈ 150) will be invited to participate. There are two control groups, i.e., sex-matched children from the same postal area (n≈ 150) and same parent siblings (full siblings) in the same household close in age +/-3 years (n<150). A three-day food diary, rating scales for mental health, and multiple questionnaires will be completed. Biosamples (fecal-, urine-, saliva-, blood samples, and buccal swab) will be collected and used for 16S rRNA gene amplicon sequencing of the oral and gut microbiome, measurements of serum factors, quantification of urine metabolites and host genotype, respectively. For longitudinal follow-up, data collection will be repeated after three years in the same groups. Integrative analysis of diet, gut microbiota, intestinal permeability, serum metabolites, and mental health will be conducted applying bioinformatics and
systems biology approaches. Extensive population-based data of this quality has not been collected before, with collection repeated in three years’ time, contributing to the high scientific value. The MMM-study follows the “Strengthening the Reporting of Observational Studies in Epidemiology” (STROBE) guidelines. Approval has been obtained from the Icelandic National Bioethics Committee, and the study is registered with Clinicaltrials.gov. The study will contribute to an improved understanding of the links between diet, gut microbiota and mental health in children through good quality study design by collecting information on multiple components, and a longitudinal approach. Furthermore, the study creates knowledge on possibilities for targeted and more personalized dietary and lifestyle interventions in subgroups.

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**Introduction**

**Mental disorders and intestinal permeability**

It has been estimated that up to 20% of children and adolescents worldwide experience mental disorders [1]. Symptoms from the gastrointestinal (GI) tract are common among children with mental disorders [2–4]. Inherent to the rationale of a gastrointestinal role in mental disorders, including imbalanced microbiota, is the idea of intestinal permeability (IP) of the intestinal epithelial barrier. Imbalanced microbiota can result in dysbiosis, which, if left untreated, can affect the intestinal epithelial paracellular pathway [5]. Increased IP allows food-borne chemicals, including toxins, and microbial components, to enter tissues beneath the intestinal lining, which can result in local and systemic inflammation caused by pro-inflammatory cytokines in the intestine as well as outside the intestinal tract [6]. Loss of intestinal barrier function may lead to neuroinflammation and neuroimmune disorders such as autism spectrum disorder (ASD) [1, 4, 7], chronic fatigue syndrome [8], major depressive disorders (MDDs) [9, 10], and schizophrenia [11, 12].

Zonulin, a family of structurally and functionally related proteins, is currently the only known physiologic modulator of intercellular tight junctions and affects mechanisms that regulate the intestinal epithelial paracellular pathway. Zonulin upregulation has been associated with several chronic inflammatory diseases, low-grade inflammation, and autoimmune diseases, including ASD and attention deficit hyperactivity disorder (ADHD), which might have an autoimmune component [1, 13]. There is growing evidence, in genetically susceptible individuals, on changes in the intestinal microbiome composition and function, i.e., intestinal dysbiosis, possibly causing functional changes in intestinal permeability (IP). Functional changes in IP may lead to increased IP, contributing to increased antigen trafficking and break of tolerance, increasing the probability of developing a chronic inflammatory disease [6, 14, 15].

Intestinal microbiota-derived antigen and endotoxin trafficking from the lumen to the lamina propria has been found to trigger innate- and immunoregulatory responses, which may cause a pro-inflammatory micro milieu. Over time, an adaptive immune response could be mounted, producing pro-inflammatory cytokines, which may cause further opening of the paracellular pathway for the passage of antigens. The specific host genetic background has been found to affect where the inflammatory process will be targeted, i.e., in which organ or tissue [6, 13, 14, 16–18]. Dysbiosis and the presence of inflammation, as well as neuroinflammation, have all been described in mental disorders [13, 19].
The gut-brain axis involves bidirectional communication between the central and the enteric nervous system [20]. Increased IP might affect the enteric nervous system, impacting both mood and behavior [12]. The emerging recognition of the relationship between the gut-brain axis and the neuro-immune system provides a novel approach for potentially better understanding and managing mental disorders. There is an urgent need to investigate these emerging interrelations further.

**Mental disorders in children**

The etiology of most mental disorders is unknown, but both genetic and environmental factors seem to play a role [21, 22]. Many mental disorders become visible during childhood and pose a challenge both to the children and their families, affecting everyday life and wellbeing. Research shows that up to 20% of children and adolescents may experience behavioral, mental, and neurodevelopmental disorders, including 10% with clinical diagnosis, resulting in disability in young people worldwide [23–25]. The most common mental disorders are ADHD, anxiety disorders, ASD, eating disorders, depressive disorders, disruptive disorders, tic disorders (TD) [26], and comorbidities are common. In the Diagnostic and Statistical Manual of Mental Disorders, 5th edition (DSM-V), neurodevelopmental disorders (ND) include ADHD, ASD, and TD. These disorders should be viewed as a spectrum rather than distinct disorders due to shared symptoms, genetics, and neuropathology [23, 27].

Recent studies that follow children from birth to adulthood indicate that most adult mental health disorders begin their origin in early childhood and adolescence [28, 29]. Despite evidence suggesting that overdiagnosis and overtreatment might exist to some extent, rates of diagnoses have increased substantially. A growing number of children and adolescents are now requiring treatments, including many types of interventions. In a systematic review of the literature that included 41 studies conducted in 27 countries from every world region [30], worldwide pooled prevalence of mental disorders was 13.4% (CI 95% 11.3–15.9). The most common diagnoses were: any anxiety disorder 6.5% (CI 95% 4.7–9.1), any depressive disorder 2.6% (CI 95% 1.7–3.9), ADHD 3.4% (CI 95% 2.6–4.5), and any disruptive disorder 5.7% (CI 95% 4.0–8.1). In comparison to the prevalence of other childhood chronic health conditions, such as obesity (16.8%) [31] and asthma (8.5%) [32], the high frequency of mental disorders and their associated negative consequences, render them major health priorities [30]. The burden of mental disorders among children aged 5–14 years has been studied in each of the six regions of the World Health Organization. Disability-adjusted life-years (DALYs) are the main indicator and are built from years of life lost (YLLs) and years of life lived with disability (YLDs). These studies showed that mental disorders in children are among the leading causes of YLDs and of DALYs in Europe and America [33].

A similar pattern is seen in Iceland at the outpatient Child and Adolescent Psychiatric Department (BUGL), where more than half of the children seen for the first time each year (n≈150) are diagnosed with ADHD, over 40% with anxiety disorders, over 30% with depressive disorders or disruptive disorders, and up to 25% with behavioral disorders or ASD, where overlapping of disorders is common [34, 35]. However, as mental disorders are thought to be products of multiple interacting causal factors rather than a single cause and are therefore multidimensional, mental disorders should preferably be evaluated dimensionally instead of categorically [36]. This has been introduced into the most recent Diagnostic and Statistical Manual of Mental Disorders (DSM-5), with more latitude to selecting severity of symptoms instead of a strictly categorical model where a disorder is either present or absent. In a research context, this approach is also considered more useful [37], especially as studies indicate that similar symptoms can have very different genetic origins [38, 39]. Clinical guidelines usually
do not recommend medication as the first line of treatment for mental disorders except in severe cases [40]. Indeed, medication can result in many unwanted side effects. Other standard therapeutic options are psychological treatments [41, 42] depending upon the service available at each location. However, it is urgent to consider also other aspects. For example, symptoms from the GI tract are common among children with mental disorders [2–4], and there are indications that these have a higher prevalence of atopic reactions [43–45]. Research shows that childhood ADHD is associated with atopic diseases and impetigo [44]. Atopic diseases seem to be associated with ADHD symptoms severity in children and adults [46]. However, there are possible limitations inherent to observational studies and further research is needed. It is necessary to study the underlying mechanisms as well as to evaluate preventative, diagnostic and therapeutic interventions.

**Diet, intestinal microbiota, and mental disorders**

In recent years, the association between diet, nutritional status, and the intestinal microbiota [47–49] has become a focus of interest, not least concerning mental health disorders [50–56]. The microbiota in children is affected by dietary habits and influences nutritional status. There is a link between the gut microbiota and malnutrition in children, as alteration in microbiota of severe acute malnourished children exhibits lower relative diversity when compared to healthy children. Moreover, the microbiota does include decreased levels of *Bacteroides* and increased levels of *Proteobacteria* in malnourished children [57]. The gut microbiota has been associated with obesity, a multifactorial chronic disorder, as is associated with other metabolic disorders and the gut microbiota is considered one of environmental regulators [58]. Several human and animal studies have revealed increased number of *Firmicutes* versus decreased number of *Bacteroidetes* in obesity [59–61]. Growth stimulation of the generally considered beneficial bacteria of Bifidobacterium and Lactobacillus by nutritional strategies with fiber or prebiotics may impact gastrointestinal health [62]. The microbiota can thus be affected by both pre- and probiotics [63–66] while dietary factors have been associated with both mental health and preexisting, existing, or other GI health problems [4, 67–69]. Furthermore, studies have indicated the importance of a healthy diet, i.e., healthy diet reflecting consumption of salads, fruit, fresh vegetables, and fish and the effects of whole-of-diet interventions rather than examining only individual foods/nutrients, as a potential factor in the treatment of anxiety [20], depression [70–72], and ADHD [73, 74]. In addition, single nutrients, e.g., optimal intake of vitamin D, omega-3 fatty acids [75–78] and iron and low exposure to heavy metals show positive outcome [79]. However, vitamin D, iron and low exposure to heavy metals did neither improve anxiety or depression except for omega-3 fatty acids which improved anxiety [78].

Studies have provided substantiated evidence in support of docosahexanoic acid (DHA), an omega-3 fatty acid, being a beneficial bioactive compound for brain function but it’s possible effects on depression disorders are still unclear [80]. Preliminary data point towards beneficial effects of omega-3 fatty acids on pediatric depression [81–86]. However, sufficient evidence is lacking to support/rule out that omega-3 fatty acids are potential treatments for depression as large-scale studies are needed to validate these results.

It is well known that eating is an integral part of the behavioral challenges in many mental disorders, with monotonic food choices and food aversions being common, which in turns affects the microbiota [87, 88]. The intestinal microbiota among children with disorders such as ASD and ADHD has been found to have a different composition compared to both community controls and healthy full siblings [89–93] living in the same household. The studies reveal a significant increase in the Bacteroidetes/Firmicutes ratio and *Clostridium* levels in children diagnosed with ASD and reduced *Bifidobacterium* levels in children diagnosed with ASD or
ADHD. However, these studies often include few participants as well as being methodologically heterogeneous, making it hard to generalize the results [94–96].

The symbiotic relationship with the microbiota has evolved, and studies indicate that the host’s genetic composition might be related to host-microbiota composition [97]. Research has shown that the intestinal microbiota might influence the host genomic organization and gene regulation, i.e., gene expression in the gastrointestinal tract, and the mechanisms of gene regulation in cells of the immune system [98]. Cell components from the microbiota including their metabolites seem to be involved in the transcriptional response of the host to microbial colonization [99, 100]. Research shows that the intestinal microbiome may remodel host chromatin, cause differential splicing, directly interrupt host signaling cascades, and alter epigenetic landscape [101]. However, mechanistic understanding of this relationship is still in its infancy, although the phenomenology of the cross-regulation of gene expression between the intestinal microbiota and the cells of the immune system has been established [102]. Host genetics may differentially influence distinct members of the microbiota. Research has shown that abundances of certain taxa is different between monozygotic twin pairs when compared with dizygotic twin pairs [98].

Furthermore, genome-wide association studies have identified numerous single-nucleotide polymorphisms (SNPs) associated with mental disorders [103], which can be used to calculate individual polygenic risk scores for study participants. However, these SNPs are associations and mechanisms are still unknown.

Microbial imbalance, i.e., intestinal dysbiosis can be defined as qualitative and quantitative changes in the intestinal microbiota, the local distribution, and metabolic activity [5]. Dysbiosis has long been implicated in the development or exacerbation of mental disorders [104]. It has been suggested that the intestinal microbiota and the brain are linked in a bidirectional relation, i.e., the microbiome–gut–brain axis [105]. The advances in research and in understanding of the role of the microbiome in neurodevelopment and mental disorder have been remarkable in the last few years. Influences of the microbiome and how it might contribute to dysregulation of brain function is an area of growing interest [52, 106–110]. Diet-induced dysbiosis substantially influences brain function through shaping the intestinal microbiome [104, 105, 111].

Studies have also established a potential link between salivary microbiome alterations and disease initiation [112–114]. Wingfield et al. [113] revealed that the composition of the oral microbiome is associated with depression in young adults and Quing et al. [114] provided hypothesis of how the oral microbiota could influence schizophrenia. Differences in alpha and beta diversity of the salivary microbiome have been observed in both these studies, with clear separation into distinct clusters between cases and healthy controls [113, 114]. More research is needed on the oral-brain axis to clarify the biological links and interconnections between the oral microbiome and the pathophysiology of mental disorders.

Unhealthy dietary pattern may increase the risk of developing depression or anxiety, whereas a healthy dietary pattern may decrease it [105]. Diet is one of the most significant factors that influences the human intestinal microbiota structure and function, and the association found between diet and mental health might partially be through the microbiota. An adequate supply of micronutrients and macronutrients is essential to well-being and provides the foundation for microbiome health, low inflammation as well as for the efficacy of other psychotherapeutic and psychopharmacological interventions [115, 116].

Dietary effects on the gut microbiome are evident, even as early as infancy. Intestinal microbiome differences have been observed between breast-fed infants with a greater prevalence of Bifidobacteria compared to formula-fed infants [117]. Existing human studies of depression and intestinal microbiota report depression-specific findings regarding
proportions of microbiota [56, 118–120]. Moreover, research shows an enrichment of pro-
inflammatory bacteria as well as a depletion of specific anti-inflammatory butyrate-producing bacteria in subjects diagnoses with anxiety, bipolar disorder, depression, and schizophrenia [121].

There is growing evidence on changes in the intestinal microbiota composition and function, i.e., intestinal dysbiosis, which may cause functional changes in intestinal permeability (IP) in genetically susceptible individuals. Functional changes in IP may lead to increased IP, which could contribute to increased antigen trafficking and break of tolerance, and therefore, increasing the probability of developing a chronic inflammatory disease [6, 14, 15].

Loss of intestinal barrier function may lead to subsequent increased serum levels of microbiota-derived molecules and, in turn, activation of the immune system, which may lead to neuroinflammation. This has been described in several neuroimmune disorders such as ASD [1, 4, 7], chronic fatigue syndrome [8], major depressive disorders (MDDs) [9, 10], and schizophrenia [11, 12].

It is vital to study the intestinal microbiota and IP further regarding diet and mental health [56]. It is important to overcome previous limitations and increase the number of subjects, coordinate strategies, and assess confounding factors, e.g., the severity of the mental disorders and problems from the GI tract, and secure quality collection [94, 122] as well as lifelong medication use. Biosamples will be collected and analyzed with the latest technology factors in addition to the microbiota, that can shed a better light on research questions and potential mechanisms. As data on the gut–brain axis demonstrates association between intestinal microbiota composition and mental disorders such as depression, we will collect fecal samples within this study for microbiome analysis. However, longitudinal studies are lacking, and therefore in this present study, collection of data will be repeated in three years. These are the basis of the current study on Meals, Microbiota and Mental Health in Children and Adolescents, i.e., the MMM study.

Our hypothesis for the current study is the following: There are differences between cases and controls regarding dietary intake, nutritional status and/or the intestinal microbiota and related metabolic factors, in physical symptoms from the digestive tract and in intestinal permeability. In addition, we hypothesize that there are differences in other potentially relevant background factors and therefore comparisons with full siblings will give insight into family variability.

Aim

The aim of this observational longitudinal case-control study is to compare diet, intestinal permeability, intestinal microbiota, and related metabolic factors in children and adolescents diagnosed with mental disorders and control groups to identify potential patterns that may contribute to the symptoms.

Methods

Study type and design

The MMM study is a double-blinded observational longitudinal case-control study (Fig 1). Adherence to the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) guidelines was pursued throughout the study design.

**Study population and participants, recruitment, and sample size.** All children and adolescents between 5 to 15 years of age referred to the outpatient clinic of the Child and Adolescent Psychiatry Department (BUGL) over one year, will be invited to participate (n≈150). There are two control groups, i.e., age and sex-matched children from the same postal area
(n≈150) and full siblings (same parent siblings, i.e., brothers and sisters) living in the same household and close in age (+/- 3 years; n<150).

**Study group.** Interested patients of the Child and Adolescent Psychiatry Department (BUGL) at The National University Hospital in Reykjavik the guardians and child will receive invitation letters with information about the MMM-study. In a follow-up phone call questions about the study can be answered and more information given. If interested to participate, an informed consent is signed, including a letter to the child’s teacher allowing the researcher to contact the teacher. For the healthy control groups (group I and II), a randomly selected child (matched by gender and age and within the same zip-code) will be contacted by the Social Sciences Research Institute (SSRI) and invited to participate similarly (control group I). Parents will be asked if a potential full sibling (+/- 3 years) living in the same household could be included in the study (control group II) as we hypothesize that there are differences in other potentially relevant background factors comparisons with full siblings, and therefore, this will give insight into family variability. However, this is not a prerequisite for participation.

A kit including tools for collecting biosamples with good instructions and a booklet for the three-day dietary recording will be provided. In addition, links will be sent for demonstrating
the specimen collection, however medical auxiliary will be available for assistance by request. Participants will be asked to be present for a blood test at the Landspitali-University Hospital in Reykjavik or Akureyri-Hospital, in Iceland, based on their residency. Eligibility criteria for the study and control groups are outlined in Table 1.

Sample size. Power calculations were performed for detecting mean difference (independent sample t-test) intestinal permeability using unmatched case-control with an $\alpha$ of 0.05 and $\beta$ of 20% (i.e., power of 80%) with 1:1 ratio of cases vs. controls [123, 124]. The prevalence of IP among cases was calculated at 30%. However, assuming 10% prevalence in controls, the minimum number of children in each arm of the study is 62. However, as our case group covers a relatively broad spectrum of mental disorders (often overlapping individually) that will be examined in smaller case groups, the aim is set at a minimum of 100 participants in each arm (70% participation rate). The current study is an observational and non-intervention study, and therefore it is not possible to calculate power for all the factors that will be compared between the cases and controls. Corrections will be made for multiple comparisons.

Data collection

The project has been registered by the Icelandic National Bioethics Committee (VSN19-225) in Iceland and clinicaltrials.gov (NCT04330703). The MMM-study adheres to the “Strengthening the Reporting of Observational Studies in Epidemiology” (STROBE) guidelines. All participants, i.e., cases and controls, will complete a three-day food diary, validated questionnaire on frequency of food and supplement intake according to the ICEFFQII, which is designed for the Icelandic food environment [125, 126] as well as a validated questionnaire on physical ailments or diagnoses, based on a version of the ROME IV [127–129]. Questionnaires on atopic diseases, according to the iFAAM [130–132] and medication and other essential background factors, will be completed. The mental health questionnaires used in the study have all been validated internationally as well as in Iceland. These are teacher and parent reports and self-reports for children >10 years and are listed with other questionnaires in Table 2.

Furthermore, if allowed by the guardians, information on prescription medication dispensed for the child from birth to the current date will be retrieved from the Icelandic Medicine Registry (IMR). During the recruitment period, the diagnosis of all patients coming to BUGL will be recorded to detect potential selection biases in data collection. If the recruitment is delayed, the advisory board will meet to discuss possible actions. Quality checks will be performed throughout the period, as well as simple descriptive statistical analysis. Longitudinal

| Participant type | Inclusion criteria | Exclusion criteria |
|------------------|--------------------|-------------------|
| Cases            | Children and adolescents 5–15 years, males, and females, referred for their first visit to the outpatient clinic at the Child and Adolescent Psychiatry Department (BUGL). Living in Reykjavik or Akureyri or surroundings. | Not speaking/understanding Icelandic language Recent use of antibiotics (within last 4 weeks) |
| Control group I: Healthy controls | Children and adolescents 5–15 years not diagnosed with mental disorders. Living in Reykjavik or Akureyri or surroundings. | Mental disorder diagnosis Recent use of antibiotics (within last 4 weeks) |
| Control group II: Parental siblings (full siblings, i.e., brothers and sisters) | Full siblings living in the same household close in age to the study subjects (+3y) not diagnosed with mental disorders. Living in Reykjavik or Akureyri or surroundings. | Mental disorder diagnosis Recent use of antibiotics (within last 4 weeks) |

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Data collection will take place three years after the first data collection. The participants will be contacted again to collect data from both cases and controls.

**Data management.** Questionnaires are completed electronically at home using an email link (in accordance with Icelandic data privacy protection for collection and storage), with each participant having codes used for unlocking the questionnaires. These have been divided into separate sessions to minimize the burden on the case participants. The user questionnaires are available in the SmartTrial platform [https://www.smart-trial.com/](https://www.smart-trial.com/) (research compliant and ethics approved) and are stored at a central database collection center. This procedure has been thoroughly quality-checked by study staff and accepted for privacy and ethics approval by the ethics committee and will secure proper storage of data. Study staff will review the completed surveys within one week of receipt and highlight any missing information. Removing of data will be considered when lacking key data on diet and microbiota or over 50% of data missing. Any issues will be discussed with the principal investigator, the SmartTrial advisory team, and/or the study team. The researchers’ staff will contact the participants no more than three times either via phone, email, or in person at a regular study visit (maximum three times) to remind them to complete questionnaires and acquire answers to missing items.

### Sample collection

All biosamples, except blood, will be collected at home by the participants, with help from their guardians when needed. A biosample kit will be delivered to the participants and collected by research staff to make sure the samples will be in good condition and usable. Participants are asked to collect the biosamples preferably towards the end of the 3-day food registration within the first month into the study and store the biosamples in a specific container in a cool dry place (4–8°C) according to the manufacturer’s guidelines. Commercial kits from DNA Genotek’s microbiome stool collection kits are used to collect and optimally stabilize stool samples. Fecal and saliva samples will be collected for microbiome 16S rRNA amplicon sequencing. A buccal swab will be collected for whole-genome genotyping of human DNA. Urine samples will be collected for targeted analysis of food-related peptides and untargeted metabolomics profiling. Blood will be collected either at Landspitali, University Hospital in Reykjavik, or Akureyri Hospital in Akureyri, depending on residence, for analyzing serum zonulin, inflammatory biomarkers, e.g., CRP, interleukins, and nutritional status, i.e., hemoglobin, ferritin, folic acid, Vitamin B₁₂, 25-hydroxy-vitamin-D, and fatty acids. The biosamples are preferable to be collected towards the end of the 3-day food registration according to

| Table 2. List of patient-answered validated mental health questionnaires and other validated and non-validated questionnaires. |
|--------------------------------------------------|
| Validated teacher and parent-reports               |
| The ADHD Rating Scales [133–135]                  |
| The Child Behavior Checklist (TRF and CBCL) [136–139] |
| The High-Functioning Autism Spectrum Screening Questionnaire (ASSQ) [140] |
| The Social Responsiveness Scale (SRS) [141–144]   |
| The Children’s Depression Inventory (CDI) [145–149] |
| Validated self-reports for children >10y          |
| The multidimensional Anxiety Scale for Children (MASC) [76, 150–155] |
| Other validated questionnaires                    |
| Food Frequency Questionnaires (ICFFQII) [125]     |
| ROME IV (shorter version) [127–129]               |
| Questionnaires on atopic diseases iFAAM [130–132] |
| Other non-validated questionnaires                |
| Questionnaires on medication and other essential background factors |

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standard procedure as mentioned above. The kit is then collected from both cases and controls by the researchers along with the food diary in Reykjavik, Akureyri, and near surroundings. Participants will be able to deliver the samples to a provided location if they prefer. For other locations in Iceland, the samples will be sent out, including guidelines on how to resend them to the researchers after their collection. The urine samples will be aliquoted into smaller tubes for storing at a laboratory unit at the University Hospital or Akureyri Hospital in a -80°C freezer along with the other biological samples.

Omics data and analyses

Microbiome data processing and analyses. PCR of 16S rRNA gene amplicons will be sequenced by an Illumina Miseq high-throughput platform and pair-end modality using standard protocols. Selected samples will be subjected to shotgun metagenomic library construction using the TruSeq DNA PCR-Free Library Preparation Kit by Illumina. The resulting 16S rRNA gene amplicon sequences will be analysed using the QIIME 2 microbiome bioinformatics platform [156].

For oral and intestinal microbiome data, we will perform principal coordinates analysis to investigate the compositional differences between samples. The cases and control groups will be compared concerning microbial diversity (alpha- and beta-diversity) and taxonomic abundances. Here methods developed for microbiome data to account for sparsity and under-sampling of the microbial community will be applied, such as implemented in the R package (phyloseq) [157, 158]. Correlations between microbial species will be mapped using sparCC [159], which is designed for compositional data such as microbial relative abundances and compared between case and control groups to reveal potential differences in the microbiome community structure. Shotgun metagenomic reads will be quality filtered to remove low-quality regions and screened for human reads. The taxonomic annotation will be performed using QIIME 2 [160]. Reads will be mapped against the human gut microbial integrated gene catalogue (IGC) [161].

Urine metabolomics processing and analyses. A morning urine sample is collected at home in a tube with peptide inhibitors. The urine is divided into two samples, one for targeted analysis of food-related peptides on an HPLC reverse phase (C-18) chromatography that gives the total peptide amount related to creatinine. The other urine sample will be used for untargeted metabolomics profiling (LC-ESI-qTOF-MS). All raw data will be extracted and processed using Agilent MassHunter Qualitative Analysis B.07.00 software. A list of peak areas, retention time and mass to charge (m/z) will be obtained and metabolites will be identified by comparing the data to selected databases. Multivariate statistical analysis will be performed using MetaboAnalyst 3.0 [162].

Serum measurements analyses. Concentrations of serum zonulin will be determined by ELISA assays according to the manufacturers’ protocols from Immundiagnostik AG (Bensheim, Germany) but performed at the Mucosal Immunology and Biology Research Center at the Massachusetts General Hospital in Boston. Inflammatory biomarkers will be determined by multiplex immunoassays based on the mesoscale discovery platform (e.g., CRP, interleukins) and nutritional status (hemoglobin, ferritin, folic acid, 25-hydroxy-vitamin-D, ω-3 fatty acids) by kits in the Landspitali laboratory in Reykjavik.

Nutritional analyses. For the nutritional data, the FFQs will be analyzed through PCA identifying dietary patterns in the groups. At the same time, the three-day registration will be reviewed by a dietitian and used to calculate a more exact intake of food items, calculated forward into nutrients (energy, macronutrients (protein, fat, carbohydrates/fibers)) and micronutrients (vitamins, minerals, and other bioactive compounds). Data with mean energy intake
below the Basal Metabolic Rate (BER) of the child will be excluded. Furthermore, the dietary data, as well as information on the nutritional status, inflammatory factors, and omics data, will be used to study associations with oral and fecal microbiota. The same analysis will also be made in the more significant subgroups, for example, children with ADHD or ASD.

**Genetic data and analyses.** Whole genome genotyping will be performed on human DNA isolated from buccal swab samples. Genotype data on study participants will be used to investigate how host gene anchors might predict or interact with particular microbiota. Here the focus will be on genetic variants previously associated with microbiota composition or psychiatric disorders.

**Integrative analysis of Meals, Microbiota, and Mental Health data.** For each type of omics- and biomarker data (nutrients, microbiota, metabolomics, food-related peptides, inflammatory factors), appropriate preprocessing, and quality control will be coordinated and performed by the researchers. All data will be appropriately normalized and transformed, checked for potential batch effects and outliers removed. The effects of potential confounders such as age, sex, body mass index, ROME IV (for intestinal microbiota), medication, or diet will be investigated. For each datatype cases will be compared to the two control groups, but also associations between the different variables will be looked for separately, as well as in relation to sub-groups, using multivariate models including adjustments for confounder variables. Using analysis of covariance (ANCOVA) the influence of potential confounders such as age, sex, body mass index, ROME IV (for intestinal microbiota), medication and diet will be investigated. Benjamin-Hochberg false discovery rate will be applied, correcting for multiple hypothesis testing.

Integrating different types of high-dimensional omics data is a challenging task, and multiple approaches can be undertaken. A computational framework for such analysis [163] has been suggested based on dimensionality reduction of omics data using co-abundance clustering, followed by cross-omics correlation analysis focused on features that differ between cases and controls. In addition, other types of omics integration approaches will be applied, as recently reviewed [164, 165], such as robust sparse canonical correlation, co-inertia, or Procrustes analysis, to identify shared patterns across omics and dietary data in the context of mental health.

**Discussion**

Investigating the interaction between diet, intestinal microbiota, IP, and their associations with mental health and comparison to healthy controls is an original and novel study. The Child and Adolescent Psychiatry Department (BUGL) at Landspitali, the National University Hospital of Iceland is the only psychiatric department for children and adolescents. This is the first population-based study enrolling all children with the most severe mental disorders assessed both categorically and dimensionally for one year and matched controls. This approach will provide valuable information on the relationship between these three domains offering crosstalk between different disciplines, to identify novel associations and potential mechanisms involved in neurodevelopmental disorders. There is a call for more population-based multidimensional transdisciplinary studies designed for improving mental health and our study fits well with priorities recently put forward in the Roadmap for Mental Health Research in Europe [166]. Furthermore, the World Health Organization (WHO) has recently called for a stronger focus on adolescent’s health, with depression as the number one cause of illness and disability in this age group [167]. Along with increasing the amount and precision of clinical information collected, the longitudinal approach of the MMM study will improve both reliability and validity of psychiatric diagnoses in line with LEAD (Longitudinal, Expert,
Table 3. Strengths and limitations of the MMM study.

- This is the first observational study evaluating diet, microbiota, intestinal permeability, and mental health in children and adolescents in Iceland.
- Recognition of the relationship between the gut-brain axis and the neuro-immune system provides a novel approach for better understanding and managing mental disorders.
- More multidimensional transdisciplinary studies including longitudinal observational data have been called for as a basis for lifestyle treatment options for improving mental health and wellness.
- The study will use two control groups, i.e., full siblings living in the same household close in age and age and sex-matched children from the same postal area, which is another strength of the study.
- Limitations are mainly the willingness to participate as full participation, although priceless for science, is demanding.
- This study will highlight associations but not causation.
- Multiple variables being assessed which makes it more challenging to get at mechanisms.
- Mental health molecular changes may occur years before symptoms making it difficult to determine linear relationship.
- The longitudinal follow-up will help but as diet and microbiome sampling will be done 3 years out it will be even more difficult to fully state if any of the variables had an influence on mental health.

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All Data) procedure [168]. Furthermore, collection of multi-omics data and clinical correlates otherwise missing in this unique study population is of importance. Repeating data collection in the same groups in three years, when majority of children will be in their adolescence, gives a valuable opportunity to observe changes especially as studies on mental health are lacking in this age group [169, 170]. More multidimensional transdisciplinary studies including longitudinal observational data have been called for as a basis for lifestyle treatment options for improving mental health and wellness [169, 170]. Importantly, the longitudinal design also allows for determining if the baseline dietary, microbiome, and metabolome patterns are predictive of future outcomes, which may aid the definition of biomarker-based diagnostic tools and therapeutic interventions. The novelty of this longitudinal observational case-control study in children and adolescents resides in the new way of approaching mental disorders holistically. Using longitudinal experts diagnoses with simultaneous use of categorical and dimensional assessments using recognized and validated rating scales, identifying patterns between the leading players of meals, intestinal microbiota, IP, and mental health has never been done before. The strengths and limitations of this study are outlined in Table 3.

The project is a cooperation between the Unit for Nutrition Research, Faculty of Food Science and Nutrition, University of Iceland and BUGL, as well as the Department of Infectious Diseases, both at Landspítl University Hospital, but includes also national and international partners. Domestic co-operation between institutions is vital in this type of inter-disciplinary research, and all participating researchers are well-established experts in their fields. Consequently, these researchers possess complementary expertise necessary to complete the research project and answer the scientific questions put forward.

The data will give multiple possibilities for further scientific writings, as well as longitudinal data after three years. Information on publications from the study will be published on the MMM web page www.mmmrannsoknin.hi.is and on social networking sites for scientists and researchers, i.e., Akademia.edu (USA) and ResearchGate.net (Europe) and social media such as Twitter and LinkedIn. This extensive dataset will be able to answer many scientific questions and will be open to other researchers after the study period.

Conclusions

The social impact of the study is high. This study will enhance understanding of gut associated contributors to mental disorders among children and adolescents in Iceland. The multi-omics approach needs to be highlighted as it is a valuable contribution. The results could increase the
alternatives for treatment in mental health diseases through relevant lifestyle options. New knowledge will be obtained that could be further pursued, giving rise to new studies and influencing the quality of life and future of public health, as this group of children is vast and expanding. By diminishing symptoms and social impairment among children and adolescents, the overall school and social situation may benefit. Furthermore, increased wellbeing and decreased medical costs are of value to society.

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References

1. Asbjornsdottir B, Snorradottir H, Andresdottir E, Fasano A, Lauth B, Gudmundsson LS, et al. Zonulin-Dependent Intestinal Permeability in Children Diagnosed with Mental Disorders: A Systematic Review and Meta-Analysis. Nutrients 2020; 12:1982. https://doi.org/10.3390/nu12071982 PMID: 32635367
2. Levy RL, Olden KW, Naliboff BD, Bradley LA, Francisconi C, Drossman DA, et al. Psychosocial aspects of the functional gastrointestinal disorders. Gastroenterology 2006; 130:1447–58. https://doi.org/10.1053/j.gastro.2005.11.057 PMID: 16678558
3. Olden KW, Drossman DA. Psychologic and psychiatric aspects of gastrointestinal disease. Med Clin North Am 2000; 84:1313–27. https://doi.org/10.1016/s0025-7125(05)70288-1 PMID: 11026930
4. Rose DR, Yang H, Serena G, Sturgeon C, Ma B, Careaga M, et al. Differential immune responses and microbiota profiles in children with autism spectrum disorders and co-morbid gastrointestinal symptoms. Brain, Behavior, and Immunity 2018; 70:354–68. https://doi.org/10.1016/j.bbi.2018.03.025 PMID: 29571898
5. Holzapfel WH, Haberer P, Snel J, Schillinger U, Huis in’t Veld JHJ. Overview of gut flora and probiotics. International Journal of Food Microbiology 1998; 41:85–101. https://doi.org/10.1016/s0168-1605(98)00044-0 PMID: 9704859
6. Fasano A. All disease begins in the (leaky) gut: role of zonulin-mediated gut permeability in the pathogenesis of some chronic inflammatory diseases. F1000Res 2020;9. https://doi.org/10.12688/F1000RESEARCH.20510.1
7. Esnaafoglu E, Ormik S, Ayyıldız SN, Erdil A, Ertürk EY, Daglı A, et al. Increased Serum Zonulin Levels as an Intestinal Permeability Marker in Autistic Subjects. Journal of Pediatrics 2017; 188:240–4. https://doi.org/10.1016/j.jpeds.2017.04.004 PMID: 28502607
8. du Preez S, Corbitt M, Cabanas H, Eaton N, Staines D, Marshall-Gradsnik S. A systematic review of enteric dysbiosis in chronic fatigue syndrome/myalgic encephalomyelitis. Syst Rev 2018; 7:241. https://doi.org/10.1186/s13643-018-0909-0 PMID: 30572962
9. Alvarez-Mon MA, Gómez AM, Orozco A, Lahera G, Sosa MD, Díaz D, et al. Abnormal Distribution and Function of Circulating Monocytes and Enhanced Bacterial Translocation in Major Depressive Disorder. Front Psychiatry 2019; 10:812. https://doi.org/10.3389/fpsyt.2019.00812 PMID: 31803077
10. Stevens BR, Goel R, Seungbum K, Richards EM, Holbert RC, Pepine CJ, et al. Increased human intestinal barrier permeability plasma biomarkers zonulin and FABP2 correlated with plasma LPS and
altered gut microbiome in anxiety or depression. Gut 2018; 67:1555–7. https://doi.org/10.1136/gutjnl-2017-314759.

11. Maes M, Sirivichayakul S, Kanchanatawan B, Vodjani A. Upregulation of the Intestinal Paracellular Pathway with Breakdown of Tight and Adherens Junctions in Deficit Schizophrenia. Molecular Neurobiology 2019; 56:7056–73. https://doi.org/10.1007/s12035-019-1578-2 PMID: 30972627

12. Barber GS, Sturgeon C, Fasano A, Cascella NG, Eaton WW, McMahon RP, et al. Elevated zonulin, a measure of tight-junction permeability, may be implicated in schizophrenia. Schizophrenia Research 2019; 211:111–2. https://doi.org/10.1016/j.schres.2019.07.006 PMID: 3107855

13. Fasano A. Intestinal Permeability and Its Regulation by Zonulin: Diagnostic and Therapeutic Implications. Clinical Gastroenterology and Hepatology 2012; 10:1096–100. https://doi.org/10.1016/j.cgh.2012.08.012 PMID: 22902773

14. Fasano A. Zonulin and Its Regulation of Intestinal Barrier Function: The Biological Door to Inflammation, Autoimmunity, and Cancer 2011. https://doi.org/10.1152/physrev.00003.2008.

15. Fasano A. Zonulin, regulation of tight junctions, and autoimmune diseases. Ann N Y Acad Sci 2012; 1258:25–33. https://doi.org/10.1111/j.1749-6632.2012.06538.x PMID: 22731712

16. Fasano A. Leaky Gut and Autoimmune Diseases. Clinical Reviews in Allergy & Immunology 2012; 42:71–8. https://doi.org/10.1007/s12016-011-8291-x PMID: 22109896

17. Sturgeon C, Fasano A. Zonulin, a regulator of epithelial and endothelial barrier functions, and its involvement in chronic inflammatory diseases. Tissue Barriers 2016; 4:e1251384. https://doi.org/10.1080/21683870.2016.1251384 PMID: 28123927

18. Asmar Re, Panigrahi P, Bamford P, Berti I, Not T, Coppa G v., et al. Host-dependent zonulin secretion causes the impairment of the small intestine barrier function after bacterial exposure. Gastroenterology 2002; 123:1607–15. https://doi.org/10.1053/gast.2002.36578 PMID: 12402335

19. Fiorentino M, Sapone A, Senger S, Camhi SS, Kuziel SK, Bui TM, et al. Blood–brain barrier and intestinal epithelial barrier alterations in autism spectrum disorders. Molecular Autism 2016; 7. https://doi.org/10.1186/s13229-016-0110-z.

20. Yang B, Wei J, Ju P, Chen J. Effects of regulating intestinal microbiota on anxiety symptoms: A systematic review. General Psychiatry 2019; 32:e100056. https://doi.org/10.1136/gpsych-2019-100056 PMID: 31179435

21. Thapar A, Cooper M, Eyre O, Langley K. What have we learnt about the causes of ADHD? J Child Psychol Psychiatry 2013; 54:3–16. https://doi.org/10.1111/j.1469-7610.2012.02611.x PMID: 22963644

22. Lesch K-P. Editorial: Illuminating the dark matter of developmental neuropsychiatric genetics—strategic focus for future research in child psychology and psychiatry. J Child Psychol Psychiatry 2014; 55:201–3. https://doi.org/10.1111/jcpp.12223 PMID: 24552481

23. Hansen BH, Oerbeck B, Skirbekk B, Petrovski BE, Kristensen H. Neurodevelopmental disorders: prevalence and comorbidity in children referred to mental health services. Nordic Journal of Psychiatry 2018; 72:885–91. https://doi.org/10.1080/08039488.2018.1444087 PMID: 29488416

24. Stein DJ, Szatmari P, Gaebel W, Berk M, Vieta E, Maj M, et al. Mental, behavioral and neurodevelopmental disorders in the ICD-11: an international perspective on key changes and controversies. BMC Medicine 2020; 18:21. https://doi.org/10.1186/s12916-020-1495-2 PMID: 31983345

25. McGinnity A, Meltzer H, Ford T, Goodman R. Mental health of children and young people in Great Britain, 2004. 2005.

26. National Institute of Mental Health. NIMH + Treatment of Children with Mental Illness n.d. https://www.nimh.nih.gov/health/publications/treatment-of-children-with-mental-illness-fact-sheet/index.shtml.

27. Kern JK, Geier DA, King PG, Sykes LK, Mehta JA, Geier MR. Shared Brain Connectivity Issues, Symptoms, and Comorbidities in Autism Spectrum Disorder, Attention Deficit/Hyperactivity Disorder, and Tourette Syndrome. Brain Connect 2015; 5:321–35. https://doi.org/10.1089/brain.2014.0324 PMID: 25602622

28. Caspi A, Houts RM, Ambler A, Danese A, Elliott ML, Harriri A, et al. Longitudinal Assessment of Mental Health Disorders and Comorbidities Across 4 Decades Among Participants in the Dunedin Birth Cohort Study. JAMA Network Open 2020; 3:e203221. https://doi.org/10.1001/jamanetworkopen.2020.3221 PMID: 32315069

29. Dalsgaard S, Thorsteinsson E, Trabjerg BB, Schullehner J, Plana-Ripoll O, Brikell I, et al. Incidence Rates and Cumulative Incidences of the Full Spectrum of Diagnosed Mental Disorders in Childhood and Adolescence. JAMA Psychiatry 2020; 77:155–64. https://doi.org/10.1001/jamapsychiatry.2019.3523 PMID: 31746968

30. Polanczyk G v Salum GA, Sugaya LS, Caye A, Rohde LA. Annual research review: A meta-analysis of the worldwide prevalence of mental disorders in children and adolescents. J Child Psychol Psychiatry 2015; 56:345–65. https://doi.org/10.1111/jcpp.12381 PMID: 25649325
31. Ogden CL, Carroll MD, Curtin LR, Lamb MM, Flegal KM. Prevalence of High Body Mass Index in US Children and Adolescents, 2007–2008. JAMA 2010; 303:242. https://doi.org/10.1001/jama.2009.2012 PMID: 20071470
32. Roper WL, Hamburg MA, King Holmes DK, Deborah Holtzman W, John Iglehart GK, Maki DG, et al. Morbidity and Mortality Weekly Report Centers for Disease Control and Prevention Editorial and Production Staff. 2007.
33. Baranne ML, Falissard B. Global burden of mental disorders among children aged 5–14 years. Child and Adolescent Psychiatry and Mental Health 2018; 12:19. https://doi.org/10.1186/s13034-018-0225-4 PMID: 29682005
34. ödärson Ö, Èvarsson FM, Helgadóttir S, Lauth B, Wessman I, Sigurjónsdóttir SA, et al. Icelandic translation and reliability data on the DSM-5 version of the schedule for affective disorders and schizophrenia for school-aged children–present and lifetime version (K-SADS-PL). Nordic Journal of Psychiatry 2020; 74:423–8. https://doi.org/10.1080/08039488.2020.1733660 PMID: 32134350
35. Johannesdóttir A. Tilvisanir á gönguþeld og í bráðaþjónustu BUGL. University of Iceland, 2017.
36. Bornstein RF. Toward a Multidimensional Model of Personality Disorder Diagnosis: Implications for DSM–5. Journal of Personality Assessment 2011; 93:362–9. https://doi.org/10.1080/00223891.2011.577474.
37. Kessler RC. The categorical versus dimensional assessment controversy in the sociology of mental illness. J Health Soc Behav 2002; 43:171–88. PMID: 12096698
38. Major Depressive Disorder Working Group of the Psychiatric GWAS Consortium S, Ripke S, Wray NR, Lewis CM, Hamilton SP, Weissman MM, et al. A mega-analysis of genome-wide association studies for major depressive disorder. Mol Psychiatry 2013; 18:497–511. https://doi.org/10.1038/mp.2012.21 PMID: 22472876
39. Power RA, Tansey KE, Buttenschøen HN, Cohen-Woods S, Bigdell T, Hall LS, et al. Genome-wide Association for Major Depression Through Age at Onset Stratification: Major Depressive Disorder Working Group of the Psychiatric Genomics Consortium. Biological Psychiatry 2017; 81:325–35. https://doi.org/10.1016/j.biopsych.2016.05.010 PMID: 27519822
40. Common mental health problems: Common mental health problems: identification and path identification and pathways to care vs to care 2011.
41. Solomon CG, Feldman HM, Reiff ML. Attention Deficit–Hyperactivity Disorder in Children and Adolescents. N Engl J Med 2014; 370:838–46. https://doi.org/10.1056/NEJMcp1307215.
42. Galanter CA. Limited Support for the Efficacy of Nonpharmacological Treatments for the Core Symptoms of ADHD. American Journal of Psychiatry 2013; 170:241–4. https://doi.org/10.1176/appi.ajp.2012.1212156 PMID: 23450282
43. Ravid NL, Annunziato RA, Ambrose MA, Chuang K, Mullarkey C, Sicherer SH, et al. Mental health and quality-of-life concerns related to the burden of food allergy. Psychiatr Clin North Am 2015; 38:77–89. https://doi.org/10.1016/j.psc.2014.11.004 PMID: 25725570
44. Hak E, de Vries TW, Hoekstra PJ, Jick SS. Association of childhood attention-deficit/hyperactivity disorder with atopic diseases and skin infections? A matched case-control study using the General Practice Research Database. Annals of Allergy, Asthma & Immunology 2013; 111:102–106.e2. https://doi.org/10.1016/j.anai.2013.05.023 PMID: 23866227
45. Dzidic M, Abrahamsson TR, Artacho A, Björkstén B, Collado MC, Mira A, et al. aberrant IgA responses to the gut microbiota during infancy precede asthma and allergy development. J Allergy Clin Immunol 2017; 139:1017–1025.e14. https://doi.org/10.1016/j.jaci.2016.06.047 PMID: 27531072
46. Chuang Y-C, Wang C-Y, Huang W-L, Wang L-J, Kuo H-C, Chen Y-C, et al. Two meta-analyses of the association between atopic diseases and core symptoms of attention deficit hyperactivity disorder. Scientific Reports 2022; 12:3377. https://doi.org/10.1038/s41598-022-0732-1 PMID: 35232975
47. Singh RK, Chang H-W, Yan D, Lee KM, Ucmak D, Wong K, et al. Influence of diet on the gut microbiome and implications for human health. Journal of Translational Medicine 2017; 15:73. https://doi.org/10.1186/s12967-016-1175-y PMID: 28388917
48. Catinean A, Neag MA, Muntean DM, Bocsan IC, Buzoianu AD. An overview on the interplay between nutraceuticals and gut microbiota. PeerJ 2018; 6:e4465. https://doi.org/10.7717/peerj.4465 PMID: 29576949
49. Flint HJ, Duncan SH, Louis P. The impact of nutrition on intestinal bacterial communities. Current Opinion in Microbiology 2017; 38:59–65. https://doi.org/10.1016/j.mib.2017.04.005 PMID: 28486162
50. Logan AC, Jacka FN, Craig JM, Prescott SL. The Microbiome and Mental Health: Looking Forward with Lessons from Allergic Diseases. Clin Psychopharmacol Neurosci 2016; 14:131–47. https://doi.org/10.9755/cpn.2016.14.2.131 PMID: 27121424
51. Yoo BB, Mazmanian SK. The Enteric Network: Interactions between the Immune and Nervous Systems of the Gut. Immunity 2017; 46:910–26. https://doi.org/10.1016/j.immuni.2017.05.011 PMID: 28636959

52. Liu RT. The microbiome as a novel paradigm in studying stress and mental health. American Psychologist 2017; 72:655–67. https://doi.org/10.1037/amp0000058 PMID: 29016169

53. McIntyre RS, Subramaniappillai M, Shekotikhina M, Carmona NE, Lee Y, Mansur RB, et al. Characterizing the gut microbiota in adults with bipolar disorder: a pilot study. Nutritional Neuroscience 2019; 1–8. https://doi.org/10.1080/1028415X.2019.1612555 PMID: 31132957

54. Dalile B, van Oudenhove L, Vervliet B, Verbeken K. The role of short-chain fatty acids in microbiota–gut–brain communication. Nature Reviews Gastroenterology & Hepatology 2019. https://doi.org/10.1038/s41575-019-0157-3 PMID: 31123355

55. Hwang Y-H, Park S, Paik J-W, Chae S-W, Kim D-H, Jeong D-G, et al. Efficacy and Safety of Lactobacillus Plantarum C29-Fermented Soybean (DW2009) in Individuals with Mild Cognitive Impairment: A 12-Week, Multi-Center, Randomized, Double-Blind, Placebo-Controlled Clinical Trial. Nutrients 2019; 11:305. https://doi.org/10.3390/nu11020305 PMID: 30717153

56. Bear TLK, Dalziel JE, Coad J, Roy NC, Butts CA, Gopal PK. The Role of the Gut Microbiota in Dietary Interventions for Depression and Anxiety. Advances in Nutrition 2020. https://doi.org/10.1093/advances/nmaa016 PMID: 32149335

57. Iddrisu I, Monteagudo-Mera A, Poveda C, Pyle S, Shahzad M, Andrews S, et al. Malnutrition and Gut Microbiota in Children. Nutrients 2021; 13. https://doi.org/10.3390/nu13082727.

58. Kim B, Choi H-N, Yim J-E. Effect of Diet on the Gut Microbiota Associated with Obesity. J Obes Metab Syndr 2019; 28:216–24. https://doi.org/10.7570/jomes.2019.28.4.216 PMID: 31909364

59. Greenhill C. Gut microbiota : Firmicutes and Bacteroidetes involved in insulin resistance by mediating levels of glucagon-like peptide 1. Nat Rev Endocrinol 2015; 11:254. https://doi.org/10.1038/nrendo.2015.40 PMID: 25781856

60. Mariat D, Firmesse O, Levenez F, Guimarães V, Sokol H, Doré J, et al. The Firmicutes/Bacteroidetes ratio of the human microbiota changes with age. BMC Microbiol 2009; 9:123. https://doi.org/10.1186/1471-2180-9-123 PMID: 19507820

61. Koliada A, Syzenko G, Moseiko V, Budovska L, Puchkov K, Perederiy V, et al. Association between body mass index and Firmicutes/Bacteroidetes ratio in an adult Ukrainian population. BMC Microbiology 2017; 17:120. https://doi.org/10.1186/s12866-017-1027-1 PMID: 28532414

62. Wegh CAM, Schoterman MHC, Vaughan EE, Belzer C, Benninga MA. The effect of fiber and probiotics on children’s gastrointestinal disorders and microbiome. Expert Rev Gastroenterol Hepatol 2017; 11:1031–45. https://doi.org/10.1080/17474124.2017.1359539 PMID: 28737484

63. Lai H-C, Young J, Lin C-S, Chang C-J, Lu C-C, Martel J, et al. Impact of the gut microbiota, prebiotics, and probiotics on human health and disease. Biomedical Journal 2014; 37:259. https://doi.org/10.4103/2319-4170.138314 PMID: 25179725

64. Druart C, Alligier M, Salazar N, Neyrinck AM, Delzenne NM. Modulation of the Gut Microbiota by Nutrients with Prebiotic and Probiotic Properties. Advances in Nutrition: An International Review Journal 2014; 5:624S–633S. https://doi.org/10.3945/an.114.005835.

65. Hemarajata P, Versalovic J. Effects of prebiotics on gut microbiota: mechanisms of intestinal immunomodulation and neuromodulation. Therap Adv Gastroenterol 2013; 6:39–51. https://doi.org/10.1177/1756283X12459294 PMID: 23320049

66. Lazar V, Ditu L-M, Pirclabioriu GG, Picu A, Petcu L, Cucu N, et al. Gut Microbiota, Host Organism, and Diet Triadology in Diabetes and Obesity. Frontiers in Nutrition 2019; 6:21. https://doi.org/10.3389/fnut.2019.00021 PMID: 30931309

67. Bressan P, Kramer P. Bread and Other Edible Agents of Mental Disease. Frontiers in Human Neuroscience 2016; 10:130. https://doi.org/10.3389/fnhum.2016.00130 PMID: 27065833

68. Shah E, Rezaie A, Riddle M, Pimentel M. Psychological disorders in gastrointestinal disease: epiphenomenon, cause or consequence? Ann Gastroenterol 2014; 27:224–30. PMID: 24974805

69. Portincasa P, Lembo A, de Bari O, di Palo DM, Maggio A, Cataldo I, et al. The role of dietary approach in irritable bowel syndrome. Current Medicinal Chemistry 2017; 24:1–1. https://doi.org/10.2174/092986732466170428102451.

70. Jacka FN, Cherbuin N, Anstey KJ, Butterworth P. Does reverse causality explain the relationship between diet and depression? Journal of Affective Disorders 2015; 175:248–50. https://doi.org/10.1016/j.jad.2015.01.007 PMID: 25658499

71. Firth J, Marx W, Dash S, Carney R, Teasdale SB, Solmi M, et al. The Effects of Dietary Improvement on Symptoms of Depression and Anxiety: A Meta-Analysis of Randomized Controlled Trials. Psychosom Med 2019; 81:265–80. https://doi.org/10.1097/PSY.0000000000000673 PMID: 30720698
72. LaChance LR, Ramsey D. Antidepressant foods: An evidence-based nutrient profiling system for depression. World J Psychiatry 2018; 8:97–104. https://doi.org/10.5498/wjp.v8.i3.97 PMID: 30254980
73. Holton KF, Nigg JT. The Association of Lifestyle Factors and ADHD in Children. J Atten Disord 2016. https://doi.org/10.1177/1087054716644652 PMID: 27125993
74. Cagigal C, Silva T, Jesus M, Silva C. Does Diet Affect the Symptoms of ADHD? Current Pharmaceutical Biotechnology 2019; 20:130–6. https://doi.org/10.2174/13892010196180925140733 PMID: 30255748
75. Watanabe N, Matsuoka Y, Kumachi K, Harazaki M, Furukawa TA. Omega-3 fatty acids for a better mental state in working populations—Happy Nurse Project: A 52-week randomized controlled trial. Journal of Psychiatric Research 2018; 102:72–80. https://doi.org/10.1016/j.jpsychires.2018.03.015 PMID: 29627596
76. Ólason DT, Sigvatsson MB, Smári J. Psychometric properties of the Multidimensional Anxiety Scale for Children (MASC) among Icelandic schoolchildren. Scandinavian Journal of Psychology 2004; 45:429–36. https://doi.org/10.1111/j.1467-9450.2004.00424.x PMID: 15535811
77. Ghasemi Fard S, Wang F, Sinclair AJ, Elliott G, Turchini GM. How does high DHA fish oil affect health? A systematic review of evidence. Critical Reviews in Food Science and Nutrition 2018; 8398:1–44. https://doi.org/10.1080/10408398.2018.1425978.
78. Su KP, Tseng PT, Lin PY, Okubo R, Chen TY, Chen YW, et al. Association of Use of Omega-3 Polyunsaturated Fatty Acids With Changes in Severity of Anxiety Symptoms: A Systematic Review and Meta-analysis. JAMA Netw Open 2018;1. https://doi.org/10.1001/jamanetworkopen.2018.2327 PMID: 30646157
79. Kim S, Arora M, Fernandez C, Landero J, Caruso J, Chen A. Lead, mercury, and cadmium exposure and attention deficit hyperactivity disorder in children. Environmental Research 2013; 126:105–10. https://doi.org/10.1016/j.envres.2013.08.008 PMID: 24034783
80. von Schacky C. Importance of EPA and DHA Blood Levels in Brain Structure and Function. Nutrients 2021; 13:1074. https://doi.org/10.3390/nu13041074 PMID: 33806218
81. Fristad MA, Vesco AT, Young AS, Healy KZ, Nader ES, Gardner W, et al. Pilot Randomized Controlled Trial of Omega-3 and Individual-Family Psychoeducational Psychotherapy for Children and Adolescents With Depression. J Clin Child Adolesc Psychol 2019; 48:S105–18. https://doi.org/10.1080/15374416.2016.1233500 PMID: 27819485
82. T J, H Z, B F, V M, W I, G I, et al. Emulsified omega-3 fatty-acids modulate the symptoms of depressive disorder in children and adolescents: a pilot study. Child Adolesc Psychiatry Ment Health 2017;11. https://doi.org/10.1186/s13034-017-0167-2.
83. Ginty AT, Corkin SM. Short-term supplementation of acute long-chain omega-3 polyunsaturated fatty acids may alter depression status and decrease symptomology among young adults with depression: A preliminary randomized and placebo controlled trial. Psychiatry Res 2015; 229:485–9. https://doi.org/10.1016/j.psychres.2015.05.072 PMID: 26188642
84. Nemets H, Nemets B, Aptek A, Bracha Z, Belmaker RH. Omega-3 treatment of childhood depression: a controlled, double-blind pilot study. Am J Psychiatry 2006; 163:1098–100. https://doi.org/10.1176/ajp.2006.163.6.1098 PMID: 16741212
85. Smith B, Rogers SL, Blissett J, Ludlow AK. The relationship between sensory sensitivity, food fussiness and food preferences in children with neurodevelopmental disorders. Appetite 2020; 150:104643. https://doi.org/10.1016/j.appet.2020.104643 PMID: 32105808
86. Koponen KK, Salosenmaa A, Ruuskanen MO, Havulina AS, Mannistö S, Joustra K, et al. Associations of healthy food choices with gut microbiota profiles. The American Journal of Clinical Nutrition 2021; 114:605–16. https://doi.org/10.1093/ajcn/nqab077 PMID: 34020448
87. Rosenfeld CS. Microbiome Disturbances and Autism Spectrum Disorders. Drug Metab Dispos 2015; 43:557–71. https://doi.org/10.1124/dmd.115.069326 PMID: 25852213
88. Tomova A, Husarova V, Lakatosova S, Bakos J, Vikova B, Babinkova K, et al. Gastrointestinal microbiota in children with autism in Slovakia. Physiol Behav 2015; 138:179–87. https://doi.org/10.1016/j.physbeh.2014.10.033 PMID: 25446201
91. Wang L, Conlon MA, Christoffersen CT, Sorich MJ, Angle MT. Gastrointestinal microbiota and metabolite biomarkers in children with autism spectrum disorders. Biomarkers in Medicine 2014; 8:331–44. https://doi.org/10.2217/bmm.14.12 PMID: 24712423

92. Pärty A, Kalliomäki M, Wacklin P, Salminen S, Isolauri E. A possible link between early probiotic intervention and the risk of neuropsychiatric disorders later in childhood: a randomized trial. Pediatr Res 2015; 77:823–8. https://doi.org/10.1038/pr.2015.51 PMID: 25760553

93. Zhang M, Ma W, Zhang J, He Y, Wang J. Analysis of gut microbiota profiles and microbe-disease associations in children with autism spectrum disorders in China. Sci Rep 2018; 8:13981. https://doi.org/10.1038/s41598-018-32219-2 PMID: 30228282

94. Buie T. Potential Etiologic Factors of Microbiome Disruption in Autism. Clin Ther 2015; 37:976–83. https://doi.org/10.1016/j.clinthera.2015.04.001 PMID: 26046240

95. Lacorte E, Gervasio G, Bacigalupo I, Vanacore N, Raucci U, Parisi P. A Systematic Review of the Microbiome in Children With Neurodevelopmental Disorders. Frontiers in Neurology 2019; 10:727. https://doi.org/10.3389/fneur.2019.00727 PMID: 31417479

96. McGuinness AJ, Davis JA, Dawson SL, Loughman A, Collier F, O'Hely M, et al. A systematic review of gut microbiota composition in observational studies of major depressive disorder, bipolar disorder and schizophrenia. Molecular Psychiatry 2022:1–16. https://doi.org/10.1038/s41380-022-01456-3.

97. Jin D, Wu S, Zhang Y, Lu R, Xia Y, Dong H, et al. Lack of Vitamin D Receptor Causes Dysbiosis and Changes the Functions of the Murine Intestinal Microbiome. Clinical Therapeutics 2015; 37:996–1009.e7. https://doi.org/10.1016/j.clinthera.2015.04.004 PMID: 26046242

98. Levy M, Thaiss CA, Elinav E. Metagenomic cross-talk: the regulatory interplay between immunogenetics and the microbiome. Genome Med 2015; 7:120. https://doi.org/10.1186/s13073-015-0249-9 PMID: 26589591

99. Shapiro H, Thaiss CA, Levy M, Elinav E. The cross talk between microbiota and the immune system: metabolites take center stage. Current Opinion in Immunology 2014; 30:54–62. https://doi.org/10.1016/j.coi.2014.07.003 PMID: 25064714

100. Thaiss CA, Levy M, Suez J, Elinav E. The interplay between the innate immune system and the microbiota. Curr Opin Immunol 2014; 26:41–8. https://doi.org/10.1016/j.coi.2013.10.016 PMID: 24556399

101. Nichols RG, Davenport ER. The relationship between the gut microbiome and host gene expression: a review. Hum Genet 2021; 140:747–60. https://doi.org/10.1007/s00439-020-02237-0 PMID: 33221945

102. Thaiss CA, Elinav E. Exploring New Horizons in Microbiome Research. Cell Host & Microbe 2014; 15:662–7. https://doi.org/10.1016/J.CHOM.2014.05.016.

103. Sullivan PF, Daly MJ, O'Donovan M. Genetic architectures of psychiatric disorders: the emerging picture and its implications. Nat Rev Genet 2012; 13:537–51. https://doi.org/10.1038/nrg3240 PMID: 22777127

104. Rogers GB, Keating DJ, Young RL, Wong M-L, Licinio J, Wesselingh S. From gut dysbiosis to altered brain function and mental illness: mechanisms and pathways. Molecular Psychiatry 2016; 21:738–48. https://doi.org/10.1038/mp.2015.16 PMID: 27090305

105. Mörk S, Wagner-Skacel J, Lahousen T, Lackner S, Holasek SJ, Bengeser SA, et al. The Role of Nutrition and the Gut-Brain Axis in Psychiatry: A Review of the Literature. Neuropsychobiology 2020; 79:8–8. https://doi.org/10.1159/000492834.

106. Chen LL, Abbaspour A, Mkoma GF, Bulik CM, Rück C, Djurfeldt D. Gut Microbiota in Psychiatric Disorders: A Systematic Review. Psychosomatic Medicine 2021; 83:679–92. https://doi.org/10.1097/PSY.0000000000000959 PMID: 34117156

107. Lima-Ojeda JM, Rupprecht R, Baghai TC. “I am i and my bacterial circumstances”: Linking gut microbiome, neurodevelopment, and depression. Frontiers in Psychiatry 2017. https://doi.org/10.3389/fpsyt.2017.00153.

108. Skonieczna-Zydecka K, Marlicz W, Misera A, Koulouzidis A, Loniewski I. Microbiome—The Missing Link in the Gut-Brain Axis: Focus on Its Role in Gastrointestinal and Mental Health. Journal of Clinical Medicine 2018; 7:521. https://doi.org/10.3390/jcm7120521 PMID: 30544486

109. Deans E. Microbiome and mental health in the modern environment. J Physiol Anthropol 2016; 36:1. https://doi.org/10.1186/s40101-016-0101-y PMID: 27405349

110. O’Mahony SM, Clarke G, Dinan TG, Cryan JF. Early-life adversity and brain development: Is the microbiome a missing piece of the puzzle? Neuroscience 2017; 342:37–54. https://doi.org/10.1016/j.neuroscience.2015.09.068 PMID: 26432952

111. Brown K, DeCoffe D, Molcan E, Gibson DL. Diet-induced dysbiosis of the intestinal microbiota and the effects on immunity and disease. Nutrients 2012; 4:1095–119. https://doi.org/10.3390/nu4081095 PMID: 23016134
112. Maitre Y, Micheneau P, Delpierre A, Mahalli R, Guerin M, Amador G, et al. Did the Brain and Oral Microbiota Talk to Each Other? A Review of the Literature. J Clin Med 2020;9. https://doi.org/10.3390/jcm9123876.

113. Wingfield B, Lapsley C, McDowell A, Miliotis G, McLaффerty M, O’Neill SM, et al. Variations in the oral microbiome are associated with depression in young adults. Sci Rep 2021; 11:15009. https://doi.org/10.1038/s41598-021-94498-6 PMID: 34294835

114. Qing Y, Xu L, Cui G, Sun L, Hu X, Yang X, et al. Salivary microbiome profiling reveals a dysbiotic schizophrenia-associated microbiota. NPJ Schizophrenia 2021; 7:51. https://doi.org/10.1038/s41537-021-00180-1 PMID: 34711862

115. Granero R. Role of Nutrition and Diet on Healthy Mental State. Nutrients 2022; 14:750. https://doi.org/10.3390/nu14040750 PMID: 35215400

116. Marx W, Moseley G, Berk M, Jacka F. Nutritional psychiatry: the present state of the evidence. Proc Nutr Soc 2017; 76:427–36. https://doi.org/10.1017/S0029665117002026 PMID: 28942748

117. Ma J, Li Z, Zhang W, Zhang C, Zhang Y, Mei H, et al. Comparison of gut microbiota in exclusively breast-fed and formula-fed babies: a study of 91 term infants. Scientific Reports 2020; 10:15792. https://doi.org/10.1038/s41598-020-72635-x PMID: 32978424

118. Dinan TG, Cryan JF. Melancholic microbes: a link between gut microbiota and depression? Neurogastroenterology & Motility 2013; 25:713–9. https://doi.org/10.1111/nmo.12198 PMID: 23910373

119. Luna RA, Foster JA. Gut brain axis: Diet microbiota interactions and implications for modulation of anxiety and depression. Current Opinion in Biotechnology 2015; 32:35–41. https://doi.org/10.1016/j.copbio.2014.10.007 PMID: 25448230

120. Simpson CA, Schwartz OS, Elilty D, Butler CA, Huang K, O’Brien-Simpson N, et al. Bugs and Brains, the Gut and Mental Health Study: a mixed-methods study investigating microbiota composition and function in anxiety, depression and irritable bowel syndrome. BMJ Open 2021; 11:e043221. https://doi.org/10.1136/bmjopen-2020-043221 PMID: 33722869

121. Nikolova VL, Hall MRB, Hall LJ, Cleare AJ, Stone JM, Young AH. Perturbations in Gut Microbiota Composition in Psychiatric Disorders. JAMA Psychiatry 2021; 78:1343. https://doi.org/10.1001/jamapsychiatry.2021.2573.

122. Cao X, Lin P, Jiang P, Li C. Characteristics of the gastrointestinal microbiome in children with autism spectrum disorder: a systematic review. Shanghai Arch Psychiatry 2013; 25:342–53. https://doi.org/10.3969/j.issn.1002-0829.2013.06.003 PMID: 24991177

123. Grimes DA, Schulz KF. Compared to what? Finding controls for case-control studies. Lancet 2005. https://doi.org/10.1016/S0140-6736(05)66379-9 PMID: 15836892

124. Rigby AS, Robinson MB. Statistical methods in epidemiology. IV. Confounding and the matched pairs odds ratio. Disabil Rehabil 2000; 22:259–65. https://doi.org/10.1080/096382800296719 PMID: 10864128

125. Eyestinsdottir T, Thorsdottir I, Gunnarsdottir I, Steingrimsdottir L. Assessing validity of a short food frequency questionnaire on present dietary intake of elderly Icelanders. Nutrition Journal 2012; 11:12. https://doi.org/10.1186/1475-2891-11-12 PMID: 22413931

126. Walter Willett. Nutritional epidemiology. Oxford University Press; 2013.

127. Baber KF, Anderson J, Puzanova M, Walker LS. Rome II versus Rome III classification of functional gastrointestinal disorders in pediatric chronic abdominal pain. J Pediatr Gastroenterol Nutr 2008; 47:299–302. https://doi.org/10.1097/MPG.0b013e31816c4372 PMID: 18728525

128. Caplan A, Walker L, Rasquin A. Development and preliminary validation of the questionnaire on pediatric gastrointestinal symptoms to assess functional gastrointestinal disorders in children and adolescents. J Pediatr Gastroenterol Nutr 2005; 41:296–304. https://doi.org/10.1097/01.mpg.0000172748.64103.33 PMID: 16131984

129. Hyams JS, di Lorenzo C, Saps M, Shulman RJ, Staiano A, van Tilburg M. Childhood functional gastrointestinal disorders: Child/adolescent. Gastroenterology 2016. https://doi.org/10.1053/j.gastro.2016.02.015.

130. Keil T, McBride D, Grimshaw K, Niggemann B, Xepapadaki P, Zannikos K, et al. The multinational birth cohort of EuroPrevall: Background, aims and methods. Allergy: European Journal of Allergy and Clinical Immunology 2010; 65:482–90. https://doi.org/10.1111/j.1398-9995.2009.02171.x PMID: 19793062

131. Sigurdardottir ST, Jonasson K, Clausen M, Lilja Bjornsdottir K, Sigurdardottir SE, Roberts G, et al. Prevalence and early-life risk factors of school-age allergic multimorbidity: The EuroPrevall-IFAAAM birth cohort. Allergy 2021; 76:2855–65. https://doi.org/10.1111/all.14857 PMID: 33934363
132. Grabenhenrich L, Trendelenburg V, Bellach J, Yürek S, Reich A, Fiandor A, et al. Frequency of food allergy in school-aged children in eight European countries—The EuroPrevall-IFAAM birth cohort. Allergy 2020; 75:2294–308. https://doi.org/10.1111/all.14290 PMID: 32219884

133. Barkley RA, Murphy KR. Attention-deficit hyperactivity disorder: a clinical workbook. Guilford Press; 2006.

134. Magnússon P, Smári J, Grétarsdóttir H, Prángardóttir H. Attention-Deficit/Hyperactivity symptoms in Icelandic schoolchildren: assessment with the Attention Deficit/Hyperactivity Rating Scale-IV. Scand J Psychol 1999; 40:301–6. https://doi.org/10.1111/1467-9450.40413 PMID: 10658514

135. Magnússon P, Smári J, Sigurðardóttir D, Baldursdóttir G, Sigmundsson J, Kristjánsson K, et al. Validity of Self-Report and Informant Rating Scales of Adult ADHD Symptoms in Comparison With a Semi-structured Diagnostic Interview. Journal of Attention Disorders 2006; 9:494–503. https://doi.org/10.1177/1087054705283650 PMID: 16481666

136. Child Behavior Checklist/4-18. University Associates in Psychiatry 1991.

137. Jensen PS, Watanabe HK, Richters JE, Roper M, Hibbs ED, Salzberg AD, et al. Scales, diagnoses, and child psychopathology: II. Comparing the CBCL and the DISC against external validators. J Abnorm Child Psychol 1996; 24:151–68. https://doi.org/10.1007/BF01441482 PMID: 8743242

138. Hannesdóttir H, Einarsdóttir S. The Icelandic child mental health study. An epidemiological study of Icelandic children 2–18 years of age using the Child Behavior Checklist as a screening instrument. European Child & Adolescent Psychiatry 1995; 4:237–48. https://doi.org/10.1007/BF01980488.

139. Hannesdottir Helga, Andre Sourander ASJPASJ P. Comparison of behavioral problems between two samples of 2- to 3-year-old children in Finland and Iceland. Nordic Journal of Psychiatry 2000; 54:13–7. https://doi.org/10.1080/080394800427528.

140. Ehlers S, Gillberg C, Wing2 L. A Screening Questionnaire for Asperger Syndrome and Other High-Functioning Autism Spectrum Disorders in School Age Children. Journal of Autism and Developmental Disorders 1999;29.

141. Constantino J. N. & Gruber CP. Social Responsiveness Scale. Western Psychological Services 2005. https://www.carautismroadmap.org/social-responsiveness-scale/?print=pdf.

142. Social Responsiveness Scale (SRS) SRS Total Score Results n.d.

143. Magnússon P. An Evaluation of the Psychometric Properties of the Social Responsiveness Scale in Two Groups of Children in Iceland. International Meeting for Autism Research, Reykjavik: 2008.

144. Constantino JN, Davis SA, Todd RD, Schindler MK, Gross MM, Brophy SL, et al. Validation of a brief quantitative measure of autistic traits: comparison of the social responsiveness scale with the autism diagnostic interview-revised. J Autism Dev Disord 2003; 33:427–33. https://doi.org/10.1023/a:1025014929212 PMID: 12959421

145. Kovacs M. The Children’s Depression, Inventory (CDI). Psychopharmacol Bull 1985; 21:995–8. PMID: 4089116

146. Kovacs M. Childrens Depression Inventory (CDI) manual. New-York: 1992.

147. Kazdin AE. Childhood depression. J Child Psychol Psychiatry 1990; 31:121–60. https://doi.org/10.1111/j.1469-7610.1990.tb02276.x PMID: 2179245

148. Craighead WE, Curry JF IS. Relationship of children’s depression inventory factors to major depression among adolescents. Psychological Assessment 1995; 7:171–6.

149. Amarsön EÖ, Smari J, Einarsdóttir H JE. The Prevalence of Depressive Symptoms in Pre-adolescent School Children in Iceland. Scand J Behav Ther 1994; 23:121–30.

150. MARCH JS, PARKER JDA, SULLIVAN K, STALLINGS P, CONNERS CK. The Multidimensional Anxiety Scale for Children (MASC): Factor Structure, Reliability, and Validity. Journal of the American Academy of Child & Adolescent Psychiatry 1997; 36:554–65. https://doi.org/10.1097/00004583-199704000-00019.

151. Anderson ER, Jordan JA, Smith AJ, Inderbitzen-Nolan HM. An examination of the MASCI social anxiety scalein a non-referred sample of adolescents. Journal of Anxiety Disorders 2009; 23:1098–105. https://doi.org/10.1016/j.janxdis.2009.07.013 PMID: 19643571

152. DIERKER LC, ALBANO AM, CLARKE GN, HEIMBERG RG, KENDALL PC, MERIKANGAS KR, et al. Screening for Anxiety and Depression in Early Adolescence. Journal of the American Academy of Child & Adolescent Psychiatry 2001; 40:929–36. https://doi.org/10.1097/00004583-200108000-00015 PMID: 11501693

153. Ivarsson T. Normative data for the Multidimensional Anxiety Scale for Children (MASC) in Swedish adolescents. Nordic Journal of Psychiatry 2006; 60:107–13. https://doi.org/10.1080/0803948060588067 PMID: 16635928
154. van Gastel W, Ferdinand RF. Screening capacity of the Multidimensional Anxiety Scale for Children (MASC) for DSM-IV anxiety disorders. Depression and Anxiety 2008; 25:1046–52. https://doi.org/10.1002/da.20452 PMID: 18833579

155. Skarphedinsson G, Villabø MA, Lauth B. Screening efficiency of the self-report version of the Multidimensional Anxiety Scale for Children in a highly comorbid inpatient sample. Nordic Journal of Psychiatry 2015; 69:613–20. https://doi.org/10.3109/08039488.2015.1026841 PMID: 25828764

156. Bolyen E, Rideout JR, Dillon MR, Bokulich NA, Abnet CC, Al-Ghalith GA, et al. Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. Nat Biotechnol 2019; 37:852–7. https://doi.org/10.1038/s41587-019-0209-9 PMID: 31341288

157. Paulson JN, Stine OC, Bravo HC, Pop M. Differential abundance analysis for microbial marker-gene surveys. Nat Methods 2013; 10:1200–2. https://doi.org/10.1038/nmeth.2658 PMID: 24076764

158. McMurdie PJ, Holmes S. Phyloseq: a bioconductor package for handling and analysis of high-throughput phylogenetic sequence data. Pac Symp Biocomput 2012:235–46. PMID: 22174279

159. Friedman J, Alm EJ. Inferring Correlation Networks from Genomic Survey Data. PLoS Computational Biology 2012; 8:e1002687. https://doi.org/10.1371/journal.pcbi.1002687 PMID: 23028285

160. Estaki M, Jiang L, Bokulich NA, McDonald D, González A, Koscielak T, et al. QIIME 2 Enables Comprehensive End-to-End Analysis of Diverse Microbiome Data and Comparative Studies with Publicly Available Data. Current Protocols in Bioinformatics 2020; 70:e100. https://doi.org/10.1002/cpbi.100 PMID: 32343490

161. Li J, Jia H, Cai X, Zhong H, Feng Q, Sunagawa S, et al. An integrated catalog of reference genes in the human gut microbiome. Nat Biotechnol 2014; 32:834–41. https://doi.org/10.1038/nbt.2942 PMID: 24997786

162. Xia J, Sinelnikov I v., Han B, Wishart DS. MetaboAnalyst 3.0—making metabolomics more meaningful. Nucleic Acids Research 2015; 43:W251–7. https://doi.org/10.1093/nar/gkv380.

163. Pedersen HK, Forslund SK, Gudmundsdottir V, Petersen AØ, Hildebrand F, Hyötyläinen T, et al. A computational framework to integrate high-throughput ‘omics’ datasets for the identification of potential mechanistic links. Nature Protocols 2018; 13:2781–800. https://doi.org/10.1038/s41596-018-0064-z PMID: 30382244

164. Knight R, Vrbancac A, Taylor BC, Aksenov A, Callewaert C, Debelius J, et al. Best practices for analysing microbiomes. Nature Reviews Microbiology 2018; 16:410–22. https://doi.org/10.1038/s41579-018-0029-9 PMID: 29795328

165. Chong J, Xia J. Computational Approaches for Integrative Analysis of the Metabolome and Microbiome. Metabolites 2017;7. https://doi.org/10.3390/metabo7040062 PMID: 29156542

166. Wykes T, Haro JM, Belli SR, Obradors-Tarragó C, Arango C, Ayuso-Mateos JL, et al. Mental health research priorities for Europe. The Lancet Psychiatry 2015; 2:1036–42. https://doi.org/10.1016/S2215-0366(15)00332-6 PMID: 26404415

167. WHO | WHO calls for stronger focus on adolescent health. WHO 2014. https://www.who.int/mediacentre/news/releases/2014/focus-adolescent-health/en/.

168. Kranzler HR, Kadden RM, Babor TF, Rounsaville BJ. Longitudinal, expert, all data procedure for psychiatric diagnosis in patients with psychoactive substance use disorders. J Nerv Ment Dis 1994; 182:277–83. https://doi.org/10.1097/00005053-199406000-00005 PMID: 10678309

169. Haro JM, Ayuso-Mateos JL, Bitter I, Demotes-Mainard J, Leboyer M, Lewis SW, et al. ROAMER: roadmap for mental health research in Europe. Int J Methods Psychiatr Res 2014; 23 Suppl 1:1–14. https://doi.org/10.1002/mpr.1406 PMID: 24375532

170. Fiorillo A, Luciano M, del Vecchio V, Sampogna G, Obradors-Tarragó C, Maj M. Priorities for mental health research in Europe: A survey among national stakeholders’ associations within the ROAMER project. World Psychiatry 2013. https://doi.org/10.1002/wps.20052 PMID: 23737426