HOLOPROSENCEPHALY: CHROMOSOMAL ABNORMALITIES IN THE ETIOPATHOGENESIS OF 127 ANTENATAL CASES

HOLOPROZENSEFALİ: 127 ANTENATAL OLGUNUN ETYOPATOGENEZİNDE KROMOZOM ANOMALİLERİ

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ABSTRACT

Objective: Holoprosencephaly (HPE, #MIM 236100) is the most common developmental defect of midline cleavage in the human forebrain. Environmental, genetic, and multifactorial causes are involved in its etiology. About half of the cases have chromosome aberrations such as trisomies 13 and 18, triploidy and structural imbalances. Single gene mutations have been shown to affect ~25% of cases. In this retrospective study, we aimed to determine the etiological factors related to HPE in 127 fetuses.

Material and Method: This study comprises 127 prenatally diagnosed fetal HPE samples from a period of 25 years, which were evaluated by karyotyping, fluorescence in situ hybridization (FISH) and aCGH investigation.

Results: A total of 64 (50.39%) chromosome aberrations were identified in this cohort. The predominant chromosomal abnormality was trisomy 13 (n=38), which was followed by trisomy 18 (n=8) and triploidy (n=5). Terminal 7q deletion was the most frequent structural anomaly (n=10, of which 5 were de novo deletion, 4 were unbalanced product of maternal translocations and one unknown in origin) and the deletion of 18p was detected in one case. In the remaining two cases, we detected trisomy 20 and pericentric inversion 11 coincidentally.

Conclusion: This study, indicates that in the presence of clinical findings suggesting HPE, cytogenetic and molecular cytogenetic studies should be performed. An aCGH study must also be done for submicroscopic chromosomal anomalies, to determine

ÖZET

Amaç: Holoprosensefali (HPE, #MIM 236100), ön beyin orta hat bölünmesinde en sık görülen gelişimsel bozukluktur. Etiyolojisinde, çevresel, genetik ve multifaktöriyal etkenler rol oynamaktadır. Vakaların yaklaşık yarısında, trizomi 13 başta olmak üzere, trizomi 18 ve triploidi gibi sayısal anomaliler ve yapısal kromozom anomalileri bulunuyordur. Olguların ~%25’inde tek gen mutasyonları göstermiştir. Bu retrospektif çalışmadan fetaal dönemde saptanan 127 fetuste HPE etiyolojisinde rol oynayan faktörlerin araştırılması planlandı.

Gereç ve Yöntem: Bu çalışma, 25 yıllık bir periyotta fetal HPE samples from a period of 25 years, which were evaluated by karyotyping, fluorescence in situ hybridization (FISH) and aCGH investigation.

Bulgular: Bu kohortta olgulanın 64 (%50,39)’unda bir kromozom anomali saptanmıştır. En sık görülen sayılal anomaliler trizomi 13 (n=38) idi, bunu sırasıyla trizomi 18 (n=8) ve triploidi (n=5) izlemiştir. Terminal 7q deletion was the most frequent structural anomaly (n=10, of which 5 were de novo deletion, 4 were unbalanced product of maternal translocations and one unknown in origin) and the deletion of 18p was detected in one case. In the remaining two cases, we detected trisomy 20 and pericentric inversion 11 coincidentally.

Sonuç: Bu çalışma, HPE klinik bulgularının varlığında sitogenetik ve moleküler sitogenetik çalışmaların birlikte veya tamamlayıcı olarak yapılması gerektiğini göstermektedir. Özellikle aCGH ça-

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their sizes, real breakpoints and identify possible novel genes that might play a role in HPE etiology.

**Keywords:** Holoprosencephaly, HPE, #MIM 236100, nervous system malformations, prenatal diagnosis

**INTRODUCTION**

Holoprosencephaly (HPE, #MIM 236100), resulting from failed forebrain separation during early embryogenesis, is the most common developmental defect in humans with an incidence of 1:250 in first trimester fetuses and 1:10,000 in newborns (1).

This malformation of the forebrain is often associated with a wide spectrum of facial anomalies, from severe to mild, such as cyclopia with proboscis, hypoplasia of the nose with a single nostril, agenesis of the premaxilla, cleft lip/palate, midfacial hypoplasia, single maxillary central incisor and hypotelorism. According to central nervous system (CNS) findings, HPE is classified into 4 groups: (i) alobar, when the hemispheres are not separated and a “monoventricle” is formed; (ii) semilobar, when the hemispheres and ventricles are partly separated and the interhemispheric fissure is only posteriorly present; (iii) lobar, when the cerebral hemispheres and lateral ventricles are separated but variable regions may partly or mildly be fused, and the peripheral olfactory nerves and bulbs are absent, and (iv) the middle interhemispheric fusion variant (MIHV), when the posterior-frontal and parietal lobes are not separated and the corpus callosum is absent (2-5).

HPE is an etiologically heterogeneous condition, which can result from chromosome aberrations, single gene mutations or environmental factors. Chromosomal aberrations are the most common etiological factor, detected in about 50% of affected individuals. Numerical (trisomy 13, trisomy 18 and triploidy), as well as structural chromosomal anomalies including submicroscopic aberrations have been associated with HPE. Structural chromosomal anomalies led the way in identifying the genes and pathways related to HPE. After the identification of Sonic Hedgehog (SHH) at 7q36 in 1996, 14 genes and 35 related loci have been published (6-8). ZIC2 at 13q32, SIX3 at 2p21, TGIF1 at 18p11.3, PTCH1 at 9q22 are reported as major causative genes and mutations in those genes are identified in about 25% of chromosomally normal cases (9-12). Single gene mutations are mostly identified in postnatal than in prenatal cases since major chromosome aberrations are lethal (13, 14).

We report here the results of classical and molecular karyotyping in 127 fetuses with HPE detected antenatally by ultrasonographic or fetal MRI evaluation.

**MATERIAL AND METHOD**

This study consists of classical karyotyping, and molecular cytogenetic examination of 127 samples from fetal tissue following the prenatal diagnosis (PD) of HPE. The examinations were carried out at the Medical Genetics Department of Istanbul Faculty of Medicine, Istanbul University, during a time span of 25-years. Fetal ultrasonography (USG) and invasive procedures were performed at the Perinatology Division of the Obstetrics and Gynecology Department of the same faculty. Fetal tissue samples were obtained by chorionic villus sampling (n=34), amniocentesis (n=26) and fetal blood sampling (n=67). Karyotype analysis was performed using G-banded chromosomes at 500-550 band level (except for 4 cases, examined at 400-450 band-levels). After 2007, cases with normal karyotype or a deletion encompassing the 7q36 band were further investigated with FISH using subtelomeric probe 7q36.3 (n=36) (Cytocell, Cambridge, UK). After 2011, aCGH was performed in 29 cases using Agilent SurePrintG3 CGH+SNP Microarray Kit (4x180K), according to the manufacturer protocol. These works were a part of routine clinical genetic evaluation. Written informed consent was obtained from the parents for all genetic tests. This study was approved by the Istanbul University, Istanbul Faculty of Medicine research ethics committee (2011/1275-642, 27.07.2011).

**RESULTS**

This study consists of 127 fetal samples with antenatal suspicion of HPE from a single center over a period of 25 years and is the largest series for prenatal work-up of HPE with cytogenetic and molecular cytogenetic analysis. The mean maternal age at the diagnosis was 30.5 years (range 16 to 45). The male to female sex ratio of fetuses was 1:2.1. Postmortem physical examinations were performed in 55, and fetal autopsy in 18 fetuses, for confirmation of HPE. The mean gestational age at the time of USG diagnosis was 21.7 weeks (range 11-38).

Fifty-four of the fetuses (42.51%) presented with alobar, 12 (9.44%) with semilobar, four with lobar HPE (3.14%), one with MIHV (0.787%) and in 56 (44.09%) the HPE sub-classification was not performed (Table 1). Two fetuses with a normal result were monozygotic twins. One of the twin pair presented with HPE, although its partner did not display any USG findings of HPE, a minor form could not be excluded. Five families with a history of more than one affected fetus with HPE had normal cytogenetic results.
Chromosomal aberrations were found in 64 of 127 fetuses (50.39%) of which 62 (48.81) were considered to be causatively related to the brain malformation. The results are summarized in Figure 1. Of the 57 fetuses investigated by classical karyotyping between 1994 and 2007, 25 (43.85%) had visible chromosome anomalies (trisomy 13 in 17 cases, triploidy in two, trisomy 18 in one, a derivative chromosome 7 as an unbalanced product of a maternal translocation in three, de novo 7q terminal deletion in one). After 2007, 70 fetal samples were karyotyped, and chromosome anomalies were detected in 39 (55.71%). The detailed description of the anomalies were as follows; trisomy 13 in 21, trisomy 18 in seven, triploidy in three, a de novo terminal deletion of 7q in four (two being the MZ twin pair), a derivative chromosome 7 as an unbalanced product of a maternal translocation in one and deletion 18p detected in one case In addition to those chromosome aberrations which all involve at least one HPE gene, two further presumably unrelated anomalies were detected: one was a pericentric inversion of chromosome 11 inherited from healthy a father, and the other non-mosaic trisomy 20.

Five cases in 4 pregnancies (counted as 2 because one case was twin) maternal unbalanced translocations were identified after fetal karyotyping. All were related to the 7q36 band, and the reciprocal breakpoints were 3p23, 3q28, 15q26.2 and 21q23.3. All cases with normal karyotype (n=34) were further investigated with FISH using a subtelomeric 7q probe, and four were found to have a 7q microdeletion. One case had a 3.5 Mb deletion of 18p encompassing the TGIF1 gene with a MLPA P070 probe set. In 29 fetal samples with normal karyotype, aCGH study was performed, and four fetuses with 7q microdeletion were identified. We detected 7q deletion and 9p duplication in aCGH examination in one case (~11,5 Mb with coordinates 147,250,584-158,816,094 and 22,3 Mb with coordinates 0-22,266,593), karyotype and FISH analysis of this case were normal. Retrospectively, those two aberrations were missed by karyotyping due to the poor quality of the chromosome preparations (at 400-450 band level). All remaining 26 cases with normal karyotypes also had normal aCGH results. ACGH results revealed that the de novo 7q microdeletions had identical coordinates (158,816,094) at q terminal breakpoints, but proximal breakpoints varied and deletion sizes differed for each case except the monozygotic twins (147,250,584; 150,997,359; 152,468,942; and 153,380,709).

### DISCUSSION

HPE arises from developmental failure at the rostral end of the neural tube during early embryogenesis (15). The phenotypic spectrum shows inter and intrafamilial variability, from severe CNS and facial anomalies to clinically normal phenotypes (16, 17). It is estimated that only 70% of obligate carriers show minor clinical features of HPE, suggesting incomplete penetrance and variable expresssion (16). Thus, chromosomal abnormalities, including numerical, structural, and submicroscopic imbalances were the cause of half of all HPE cases in the present report. All instances, involved known HPE genes.

Gene mutations can also cause both syndromic and non-syndromic HPE. It has been suggested that 25% of HPE cases carry a mutation in a single gene (18). Teratogenic and environmental agents have also been reported as aetiological factors. Determining the etiology of HPE is important for genetic counselling, especially in estimating the recurrence risk. When numerical aberrations are detected, the recurrence risk is low; although, taking germ line mosaicism into account, foetal karyotyping should be recommended for future pregnancies (19). In cases with structural chromosomal abnormalities, parental studies must be performed to exclude balanced rear-

### Table 1: Distribution of HPE types among chromosomal abnormalities

|                      | n=127 | % ratio in all cases | % ratio in anomalies |
|----------------------|-------|---------------------|---------------------|
| Normal karyotype     | 63    | 49.6                |                     |
| Chromosome abnormalities | 64    | 50.4                |                     |
| Trisomy 13           | 38    | 29.9                | 59.4                |
| Trisomy 18           | 8     | 6.29                | 12.5                |
| Triploidy            | 5     | 3.93                | 7.8                 |
| Derivative chromosomes | 5 (4 mat+1 unknown) | 3.93                | 7.8                 |
| 7q deletion          | 5 (dn)| 3.93                | 7.8                 |
| 18p deletion         | 1     | 0.78                | 1.56                |
| Trisomy 20           | 1     | 0.78                | 1.56                |
| Pericentric inversion| 1     | 0.78                | 1.56                |
role of chromosomal abnormalities in HPE
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The proportion of chromosome anomalies in live births with HPE ranges from 25-45%, and it is distinctly higher in early pregnancy, since numerical abnormalities frequently result in a non-viable foetus. The rate of chromosomal abnormalities in our cohort was 50.4%, and we further confirmed that observation. The mean gestational age at the time of USG diagnosis was 20.1 weeks for cases with numerical chromosome anomalies, and this did not differ significantly from foetuses with structural imbalances and chromosomally normal foetuses (22.5 and 22 weeks, respectively). The detection rate of chromosomal imbalances was 43.9% in the first period of the study, when only classical karyotyping was available. After the implementation of molecular cytogenetic techniques in 2007, a total of 36 chromosomal imbalances were diagnosed in 70 foetuses with HPE (51.42%). Theoretically, if the karyotype resolution had been high (>550 band level) in all cases (which is unfortunately not the case in a diagnostic laboratory), with the exception of the 18p microdeletion and three other cases all cytogenetic aberrations in the cohort could have been identified.

According to previously published data, trisomy 13, including cryptic rearrangements, comprises up to 75% of the chromosome anomalies in HPE. It has also been reported that about approximately 20% of the anomalies had were triploidy and 1%-2% had trisomy 18 (21, 22). In our cohort, the detection frequencies of trisomy 13, trisomy 18, and triploidy were 59.4, 12.5 and 7.8%, respectively, with much lower rates for trisomy 13 and triploidy but a higher frequency of trisomy 18 than previously reported. This finding might be a coincidence, because targeted sequencing of HPE-related genes was not performed and the aberration is probably confined to extra-foetal tissues. De novo or familial 7q deletions were also observed at significant rates (7.8%-7.8% respectively) in our cohort, probably due to the targeted FISH analysis for 7q microdeletions. In our cohort, we detected two incidental chromosome abnormalities, one, of them with a paternally transmitted inv(11), and the other with trisomy 20. The inv(11) is probably not related to HPE in the foetus since it was also present in the healthy father as well, the region does not involve a known HPE gene (1-3). The significance of trisomy 20 in extra-embryonic cells is unclear, and to the best of our knowledge, no cases of HPE with prenatal diagnosis of trisomy 20 have been reported.

Structural chromosomal anomalies associated with HPE have also been reported. In HPE, structural chromosome anomalies are mostly seen in 13q, 18p11.2, 7q36, 3p24, p21 and 21q22.3 regions. These regions are also the loci where autosomal dominant inherited HPE genes are localized (23). Structural chromosome anomalies have resulted in the identification of novel genes associated with HPE. In this cohort, the four identified unbalanced products of maternal reciprocal translocations had one breakpoint at 3p23, 3q28, 15q26.2, 21q22.3, and shared a terminal breakpoint at 7q36. In all these cases, the HPE phenotype is presumably related to monosomy of 7q36->qter, which encompasses the SHH locus. However, in one, the derivative chromosome had a concomitant duplication of 3p23->qter, which may have contributed to the phenotype, as the association between 3p duplication and HPE is well documented (24). Ginocchio et al. reviewed HPE cases with 3p duplications and almost all had alobar HPE (24). Our case also displayed alobar HPE. Another unbalanced offspring of a balanced reciprocal translocation was monosomic for 7q36->qter and trisomic for 21q22.3->qter. The minimal critical region of HPE1 locus is on 21q22.3 the the monosomy of this region is associated with HPE; however, HPE has rarely been reported in association with trisomy 21 (25). Only one case with partial duplication of 21q22.3 and monosomy 18p11.2 as an unbalanced product of a maternal translocation has been reported to display HPE, which can be attributed to the monosomy 18p, which includes TGIF1 gene (26). Another familial case reported by Balci et al. showed a clear association between HPE and with monosomy 18p (27). A de novo terminal deletion of 18p was identified in our cohort by multiplex ligation-dependent probe amplification, and confirmed by subtelomeric FISH studies, and HPE was associated with haploinsufficiency of the TGIF1 gene located at 18p11.

Bendavid et al. demonstrated that submicroscopic aberrations are a most frequent causes of HPE, with 17% of de novo anomalies detected by aCGH analysis (28). Performing aCGH in 29 cases with normal karyotype revealed one chromosome anomaly that should have been detected by karyotyping, but was missed due to poor quality chromosome preparations. All other CNVs identified were in known recurrent regions.

To date, more than 14 genes have been associated with non-syndromic HPE. SHH (HPE3) located on 7q36 was described first, and is the most frequent causative gene in nonchromosomal and nonsyndromic HPE, accounting for approximately 12% of cases (18). According
to Raam et al. in all non-syndromic HPE patients with normal chromosomes, molecular analysis should be performed for the most common genes SHH, ZIC2, and SIX3 as a first step (29). If no mutation is found, then TGIF1 and GLI family zinc finger 2 should be analysed. Mutations in the four major genes (SHH, ZIC2, SIX3, TGIF1) were identified in 25% of cases. The SHH, SIX3, and TGIF1 mutations were inherited in more than 70% of these cases, whereas 70% of ZIC2 mutations occurred de novo. There was a positive correlation between the severity of the brain malformation and facial features for SHH, SIX3 and TGIF1 mutations, but no such correlation was shown for ZIC2. The most severe HPE types were associated with SIX3 and ZIC2 mutations, whereas microforms were associated with SHH mutations (30). HPE was found in five families in our cohort in at least two pregnancies, and only one of these families was consanguineous (the parents were first cousins). In one case with normal karyotype, a physical examination of the parents revealed that the mother had a single maxillary central incisor, and in the consanguineous family, both parents had hypotelorism. Molecular studies were performed on the two families with normal karyotypes and causative genes were SHH in one family (Solomon’ personnel communication) and SIX3 in the other (Duborg’ personnel communication).

Families are understandably concerned about the recurrence risk of HPE; therefore, the delineation of HPE etiology is very important for genetic counselling and the monitoring of future pregnancies. If a genetic aetiology is found, prenatal diagnosis should be recommended in the first trimester of subsequent pregnancies. However, the wide spectrum of the phenotypic variation should be taken into consideration, and USG examination is crucial.

CONCLUSION

When HPE is diagnosed prenatally, physical examination and cytogenetic analysis of the parents is essential. If high-resolution chromosome analysis is performed, almost all aberrations, including specific microdeletions, can be detected in more than half of the patients. As the identification of submicroscopic aberrations aids in the identification of new loci and genes, aCGH should be performed as a secondary test in cases with normal karyotype. If aCGH studies also reveal normal results, sequencing of related genes should be the third test performed. However, incomplete penetrance and variable phenotypic presentation should be taken into consideration during genetic counselling.

We are planning to investigate single gene disorders in cases where chromosome anomalies are not detected by any available technique in this study.

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