Discrepancy between short-term and long-term effects of bone marrow-derived cell therapy in acute myocardial infarction: a systematic review and meta-analysis

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Abstract

Background: Bone marrow-derived cell therapy has been used to treat acute myocardial infarction. However, the therapeutic efficacy of this approach remains controversial. Here, we performed a systematic review and meta-analysis to evaluate short-term and long-term effectiveness of bone marrow-derived therapy.

Methods: We searched eight databases (Ovid-Medline, Ovid-EMBASE, Cochrane Library, KoreaMed, KMBASE, KISS, RISS, and KisTi) up to December 2014. Demographic characteristics, clinical outcomes, and adverse events were analyzed. We identified 5534 potentially relevant studies; 405 were subjected to a full-text review. Forty-three studies with 2635 patients were included in this review.

Results: No safety issues related to cell injection were reported during follow-up. At 6 months, cell-injected patients showed modest improvements in left ventricular ejection fraction (LVEF) compared with the control group. However, there were no differences between groups at other time points. In the cardiac MRI analysis, there were no significant differences in infarct size reduction between groups. Interestingly, mortality tended to be reduced at the 3-year follow-up, and at the 5-year follow-up, cell injection significantly decreased all-cause mortality.

Conclusions: This meta-analysis demonstrated discrepancies between short-term LV functional improvement and long-term all-cause mortality. Future clinical trials should include long-term follow-up outcomes to validate the therapeutic efficacy of cell therapy.

Keywords: Cell therapy, Acute myocardial infarction, Survival, Bone marrow
between the cell therapy and control groups. To overcome the limitations of individual studies and increase statistical power, several meta-analyses have been performed. A meta-analysis in 2012 showed that intracoronary BM cell therapy after AMI significantly improved the left ventricular ejection fraction (LVEF) at 6 months after treatment; however, the absolute value of improvement was modest (2.87 %) [6]. In contrast, a meta-analysis in 2014 reported that there was no detectable therapeutic benefit with regard to major adverse cardiac event rates after BM cell therapy after a median follow-up duration of 6 months [7], and a recent meta-analysis in 2015, using individual patient data from 12 randomized trials, demonstrated no benefit for 1-year follow-up LVEF and clinical outcomes [8]. Taken together, these studies suggest that there could be a discrepancy between the improvement of LV function and clinical outcomes. Furthermore, long-term follow-up results are lacking. Accordingly, we conducted the largest meta-analysis of this topic reported to date, including 43 randomized trials with 2635 patients, and focused on follow-up results at 6 months, 1 year, 3 years, and 5 years in order to evaluate short-term and long-term effects of the cell therapy in patients with AMI. We found the significant difference between cardiac magnetic resonance imaging (MRI) and echocardiographic measurements of LV functional improvement and infarction size reduction. Furthermore, we revealed the discrepancy between the LV function and clinical outcomes. Cell therapy showed the long-term mortality benefits at 5-year follow-up, although the effects of LV functional improvement and infarct size reduction were modest in short-term analyses.

Methods
Information sources and search strategy
We searched eight databases (Ovid-Medline, Ovid-EMBASE, Cochrane Library, KoreaMed, KMBASE, KISS, RISS, and KisTI) up to December 2014. To ensure a sensitive search, we designed strategies that included medical subject headings (MeSH) keywords, such as “myocardial infarction”, “acute MI”, “MI”, “STEMI”, “coronary heart disease”, “angina”, “ischemic heart disease”, “ischemic cardiomyopathy”, “heart failure”, “bone marrow cell (BMC)”, “mononuclear cell (MNC)”, “mesenchymal stem cell”, “mesenchymal stromal cell”, “MSC”, “granulocyte colony-stimulating factor (G-CSF)”, and all possible combinations.

Inclusion and exclusion criteria
The inclusion criteria were as follows: (1) randomized clinical trials (RCTs), (2) published original articles, (3) written in English, (4) stem cell or cell therapy studies, and (5) included proper outcomes (follow-up LV function, mortality, etc.). The exclusion criteria were as follows: (1) nonhuman studies, (2) preclinical studies, (3) gray literature, (4) non-RCTs, and (5) duplicated reports. Two reviewers screened studies according to the selection criteria. First, we removed duplicate studies and performed title and abstract screening. Second, we selected potentially pertinent studies and reviewed the full text. Finally, we chose 43 randomized clinical trials and analyzed 2635 patients [9–51] (Fig. 1).

Data extraction and quality assessment
Two independent reviewers assessed studies using a standardized form. Information extracted from the chosen studies included authors, publication year, country, design, results, and funding. Clinical outcomes consisted of complications such as arrhythmia, heart failure, ischemic heart disease (IHD) recurrence, restenosis, revascularization, stroke, and all-cause mortality. Effectiveness outcomes were improvement of left ventricular ejection fraction (LVEF) and reduction of infarct size at each follow-up point. Assessment of the quality of articles was made according to the Cochrane Handbook [52]. Two authors (Lee and Hong) evaluated all of the studies including methodological quality according to the risk of bias for randomized controlled trials.

Statistical analysis
We conducted the meta-analysis using Review Manager version 5.3 (Cochrane Collaboration). For dichotomous and continuous variables, the odds ratios (ORs) and weighted mean differences were calculated for groups using a random effects model and inverse variance weighting. Inverse variance weighting is a method of aggregating two or more random variables to minimize the variance of the weighted average [53]. For studies that did not report the actual change as the mean, the standard deviation was calculated using a standardized formula that was previously validated when the baseline and follow-up standard deviation were known. In this study, we reported results according to a random effects model considering heterogeneity, and the publication bias was tested using a Funnel plot [53, 54]. The Chi-square test with significance set at \( P < 0.10 \) was used to assess statistical heterogeneity of selected studies, and \( I^2 \) statistics were provided to quantify the heterogeneity.

Results
Search and selection of BM-derived cell therapy clinical trials
We identified 5534 potentially relevant studies and further screened these studies for eligibility. We selected 405 pertinent cell therapy studies for a full-text review.
The outcomes of interest included demographic characteristics, clinical outcomes, and effectiveness outcomes at each follow-up point. Of these 405 studies, 362 were excluded because they were not describing acute MI (n = 39), were not RCTs (n = 122), were gray literature (n = 1), were duplicate reports (n = 50), did not include clinical outcomes (n = 136), were not interventional studies (n = 10), and were not MI studies (n = 3). We finally included 43 studies with 2635 patients [9–51] (Fig. 1).

**Study characteristics**

The studies were published from 2004 to 2014. Table 1 lists the characteristics all of the included studies. The study sizes ranged from 8 to 204 patients, and the follow-up duration ranged from 3 to 108 months. Twenty-four of 41 studies were conducted in Europe, six studies were conducted in the USA, five studies were conducted in China, three studies were conducted in Korea, one study was conducted in Mexico, one study was conducted in Canada, one study was conducted in Thailand, one study was conducted in Brazil, and one study was conducted in Iran. Among the 2635 patients, 1399 patients received BM-derived cell therapy, and 1236 patients were in the control group. Most studies used freshly isolated BM-MNCs isolated by density gradient separation of autologous BM aspires. Three studies used BM-derived CD133+ cells by cell sorting using a specific antibody [20, 28, 30], and one study used CD34+ cells [24]. Two studies used granulocyte colony-stimulating factor (G-CSF)-mobilized peripheral blood mononuclear cells (PB-MNCs) by leukapheresis [17, 50], and two other studies used mesenchymal stem cells (MSCs), which were cultured from BM aspirates under attached conditions for 1 month [11, 35]. Most cell injections were performed through intracoronary infusion 2 to 7 days after percutaneous coronary intervention (PCI). One study infused cells intravenously after PCI [35] and another study injected cells intramuscularly through the epicardium during coronary artery bypass graft operation [28]. The total number of injected cells is listed in Table 1.

**Functional improvement of the LV and reduction of infarct size**

Analyses based on a random effects model for the difference in LVEF and infarct size are shown in Fig. 2. Cell therapy improved the LVEF at 6 months (2.75 % increase; \( P < 0.001 \)) and 1 year (1.34 % increase; \( P = 0.03 \)) as compared with the control group (Fig. 2a and b). However, at the 3- and 5-year follow-up, there were no significant differences in LVEF between the cell therapy and control groups (Fig. 2c and d). Infarct size in the cell therapy tended to decrease at 6 months compared with
| Author               | Year | Country | Timing of cell injection | Cell injection (n) | Control (n) | Total patient (n) | Cell type | Cell delivery route | Cell preparation | No. of injected cells | Follow-up (months) |
|----------------------|------|---------|--------------------------|-------------------|------------|------------------|-----------|--------------------|-------------------|---------------------|---------------------|
| Assmus et al. [9]    | 2014 | Germany | After PCI               | 101               | 103        | 204              | MNC       | IC                 | Ficoll gradient   | 1 × 10^6            | 60                  |
| Benedek et al. [10]  | 2014 | Romania | After PCI               | 9                 | 9          | 18               | MNC       | IC                 | Concentration by apheresis | 1.66 ± 0.32 × 10^9 | 48                  |
| Lee et al. [11]      | 2014 | Korea   | After PCI               | 30                | 28         | 58               | MSC       | IC                 | 1-month culture   | 7.2 ± 0.90 × 10^7  | 6                   |
| Robbers et al. [12]  | 2014 | Netherlands | After PCI           | 30                | 45         | 75               | BM-MNC/PB-MNC | IC | Lymphoprep™     | 296 ± 146 × 10^6 | 4                   |
| Gao et al. [13]      | 2013 | China   | After PCI               | 21                | 22         | 43               | MSC       | IC                 | 2-week culture    | 3.08 ± 0.52 × 10^6 | 6, 12, 24            |
| Surder et al. [14]   | 2013 | Switzerland | After PCI            | 65                | 67         | 132              | MNC       | Density gradient   | 5 × 10^7 - 5 × 10^8 | 4                   |
| Wohrle et al. [15]   | 2013 | Germany | After PCI               | 29                | 13         | 42               | MNC       | IC                 | Ficoll gradient   | 324 × 10^6          | 6, 12, 24, 36       |
| Jazi et al. [16]     | 2012 | Iran    | After PCI               | 16                | 16         | 32               | MNC       | IC                 | Ficoll gradient   | 24.6 ± 8.4 × 10^8  | 6                   |
| Kang et al. [17]     | 2012 | Korea   | After PCI               | 57                | 60         | 117              | PB-MNC    | IC                 | G-CSF/leukapheresis | 1.1 ± 0.5 × 10^9  | 6, 24, 60           |
| Skalicka et al. [18] | 2012 | Czech   | After PCI               | 17                | 10         | 27               | MNC       | IC                 | Gelfusine         | 26.4 ± 10^8        | 4, 24               |
| Traverse et al. [19] | 2012 | USA     | After PCI               | 79                | 41         | 120              | MNC       | IC                 | Sepax*            | 1.50 ± 10^6        | 6                   |
| Colombo et al. [20]  | 2011 | Italy   | After PCI               | 5                 | 5          | 10               | CD133+ MNC | IC | CD133 by CliniMACS® | 4.9-135 × 10^6 | 12                  |
| Hirsch et al. [21]   | 2011 | Netherlands | After PCI            | 69                | 65         | 134              | MNC       | IC                 | Lymphoprep™       | 296 ± 164 × 10^6   | 4                   |
| Pena-Duque et al. [22]| 2011 | Mexico  | After PCI               | 4                 | 4          | 8                | MNC       | IC                 | Sepax*            | 1 ~ 2 × 10^6 CD34 | 6                   |
| Plewka et al. [23]   | 2011 | Poland  | After PCI               | 40                | 20         | 60               | MNC       | IC                 | Safe Flow*        | 1.44 ± 0.49 × 10^8 | 24                  |
| Quyyumi et al. [24]  | 2011 | USA     | After PCI               | 16                | 15         | 31               | CD34+ MNC | IC | Isolex 300®      | 5 × 10^6           | 6, 12               |
| Roncalli et al. [25] | 2011 | France  | After PCI               | 52                | 49         | 101              | MNC       | IC                 | Ficoll            | 98 ± 8.7 × 10^6   | 3                   |
| Srimahachota et al. [26] | 2011 | Thailand | After PCI             | 11                | 12         | 23               | MNC       | IC                 | Isoprep*          | 420 ± 221 × 10^6   | 6                   |
| Turan et al. [27]    | 2011 | Germany | After PCI               | 38                | 18         | 56               | MNC       | IC                 | Point of Care system | 1 × 10^6           | 3, 12               |
| Yerebakan et al. [28]| 2011 | Germany | During CABG            | 20                | 20         | 40               | CD133+ MNC | IM | CD133 by CliniMACS® | 6.0 × 10^6 | 108                 |
| Grajek et al. [29]   | 2010 | Poland  | After PCI               | 31                | 14         | 45               | MNC       | IC                 | Ficoll gradient   | 4.1 ± 1.8 × 10^9  | 3, 6, 12            |
| Mansour et al. [30]  | 2010 | Canada  | After PCI               | 14                | 14         | 45               | CD133+ MNC | IC | CliniMACS®      | 10 × 10^6          | 4                   |
| Piepoli et al. [31]  | 2010 | Italy   | After PCI               | 19                | 19         | 38               | MNC       | IC                 | Ficoll gradient   | 418 ± 10^6         | 3, 6, 12            |
| Traverse et al. [32] | 2010 | USA     | After PCI               | 30                | 10         | 40               | MNC       | IC                 | Ficoll gradient   | 100 ± 10^6         | 6                   |
| Wohrle et al. [33]   | 2010 | Germany | After PCI               | 29                | 13         | 42               | MNC       | IC                 | Ficoll gradient   | 381 ± 130 × 10^6   | 6                   |
| Cao et al. [34]      | 2009 | USA     | After PCI               | 41                | 45         | 86               | MNC       | IC                 | Lymphoprep™       | 5 ± 1.2 × 10^7    | 48                  |
| Hare et al. [35]     | 2009 | USA     | After PCI               | 39                | 21         | 60               | MSC (allogeneic) | IV | 1-month culture | 0.5 ~ 5 × 10^6 | 6                   |
| Nogueira et al. [36] | 2009 | Brazil  | After PCI               | 14                | 6          | 20               | MNC       | IC                 | Ficoll gradient   | 10 × 10^6          | 6                   |
Table 1  Study characteristics (Continued)

| Study                  | Year | Country | PCI Type | Timepoints (months) | MNC Type | IC Type     | Cells | Plus-minus value |
|------------------------|------|---------|----------|--------------------|----------|-------------|-------|------------------|
| Yao et al. [37]        | 2009 | China   | After PCI | 12, 12, 24        | MNC      | IC          | 1.9 ± 1.3 × 10^8 | 6, 12 |
| Huikuri et al. [38]    | 2008 | Finland | After PCI | 40, 40, 80        | MNC      | IC          | 4.02 ± 1.96 × 10^6 | 6    |
| Meluzin et al. [39]    | 2008 | Czech   | After PCI | 20, 20, 40        | MNC      | IC          | 1-10 × 10^7       | 3, 6, 12 |
| de Lezo et al. [41]    | 2007 | Spain   | After PCI | 10, 10, 20        | MNC      | IC          | 9 ± 3 × 10^8     | 3    |
| Ge et al. [42]         | 2006 | China   | After PCI | 10, 10, 20        | MNC      | IC          | 4 × 10^7         | 6    |
| Janssens et al. [43]   | 2006 | Belgium | After PCI | 33, 34, 67        | MNC      | IC          | 172 ± 10^6       | 4    |
| Lunde et al. [44]      | 2006 | Norway  | After PCI | 47, 50, 97        | MNC      | IC          | 68 × 10^6        | 6, 36 |
| Meluzin et al. [45]    | 2006 | Czech   | After PCI | 22, 22, 44        | MNC      | IC          | 1 × 10^7 - 1 × 10^8 | 3 |
| Schachinger et al. [46]| 2006 | Germany | After PCI | 101, 103, 204     | MNC      | IC          | 236 ± 1.74 × 10^6 | 24 |
| Karpov et al. [47]     | 2005 | Russia  | After PCI | 22, 22, 44        | MNC      | IC          | 88.5 ± 49.2 × 10^6 | 6 |
| Ruan et al. [48]       | 2005 | China   | After PCI | 9, 11, 20         | MNC      | IC          | Not described     | Not described |
| Chen et al. [49]       | 2004 | China   | After PCI | 34, 35, 69        | MSC      | IC          | 2-5 × 10^6       | 3, 6 |
| Kang et al. [50]       | 2004 | Korea   | After PCI | 6, 7, 13          | PB-MNC   | IC          | 1 × 10^9         | 6, 24 |
| Wollert et al. [51]    | 2004 | Germany | After PCI | 30, 30, 60        | MNC      | IC          | 24.6 ± 9.4 × 10^8 | 6, 18, 60 |

Included studies were all randomized clinical trials (RCTs). Plus-minus value indicates mean ± SE.

PCI: percutaneous coronary intervention, MNC: mononuclear cell, IC: intracoronary cell infusion, BM-MNC: bone marrow-derived mononuclear cell, PB-MNC: peripheral blood-derived mononuclear cell, G-CSF: granulocyte colony-stimulating factor, CABG: coronary artery bypass graft, IM: intramyocardial cell injection, IV: intravenous cell infusion, MSC: mesenchymal stromal cell.
Fig. 2 (See legend on next page.)
that in the control group; however, this difference was not statistically significant (−2.99%; 95% confidence interval [CI], −7.08, 1.11; P = 0.15; Fig. 3a). At the 1-year follow-up, infarct size was significantly reduced (−6.10%; 95% CI, −11.14, −1.05; P = 0.02; Fig. 3b). However, there were no significant differences in infarct size at the 3-year (1.10%; 95% CI, −9.24, 11.44; P = 0.83; Fig. 3c) and 5-year follow-up (1.10%; 95% CI, −9.24, 11.44; P = 0.83; Fig. 3d).

Taken together, these data indicated that the differences in LV systolic function and infarct size between the cell therapy and standard care control groups disappeared within 1 year of the initial treatment.

**Difference between measurement modalities**

It is possible for there to be differences in measurements between the modalities [12]. Accordingly, we performed subgroup analysis of the results from cardiac MRI and echocardiography (or LV angiogram). LVEF measured by cardiac MRI showed no significant difference between the cell therapy and control groups (0.51%; 95% CI, −1.20, 2.23; P = 0.56), whereas echocardiographic (or LV angiographic) measurement (non-MRI) revealed that cell therapy significantly increased LVEF compared with that in the control group (4.02%; 95% CI, 2.65, 5.39; P < 0.001; Table 2 and Fig. 4a).

To examine the effects of study design on maintaining blindness, we also analyzed bone marrow aspiration and placebo infusion in patients in the control group. Patients were separated into two groups according to whether each procedure was used in the control group; cells were isolated from the BM by aspiration and infused placebo in the control group (placebo procedure), or no procedure was performed in the control group (Table 2). The outcome was measured as the change in LVEF from baseline to 6 months. Significant effects were observed in the group in which the control group underwent aspiration and placebo infusion, as indicated by an LVEF of 1.86% (95% CI, 0.54, 3.19; P = 0.006). Additionally, the other group in which no procedures were performed showed significant differences in LVEF (3.20%; 95% CI, 1.84, 4.56; P < 0.001).

Regarding the reduction in infarct size (Table 2 and Fig. 4b), MRI measurement did not show any significant differences in infarct size (−3.96% 95% CI, −10.81, 2.90; P = 0.26) between the cell therapy and control groups. Moreover, non-MRI modalities (echocardiography or LV angiogram) also showed no significant differences (0.1%; 95% CI, −0.09, 0.29; P = 0.29). No significant treatment effect was found the group in which the control group underwent aspiration in infarct size (−7.93% 95% CI, −19.57, 3.72; P = 0.18). Additionally, lack of aspiration in the control group did not have any beneficial effects on infarct size reduction (0.55%; 95% CI, −0.62, 1.72; P = 0.36).

**Major adverse clinical events**

The safety of BM cell harvesting and delivery has been established. All-cause mortality was analyzed by the Mantel-Haenszel (M-H) random model, which is one of the most popular procedures for detecting differential item functioning [55]. If we assume a constant relative Odds (θi = μ or Δ2 = 0), then the Mantel-Haenszel statistic is optimal for testing H0: μ = 1, and there is considerable literature on efficient estimates of μ and on methods for testing H0: θ1 = θ2 = ... = θk [56].

The results demonstrated that there were no differences between groups at the 6-month (odds ratio [OR], 1.08; 95% CI, 0.42, 2.8; P = 0.87) and 1-year follow-up (OR, 0.89; 95% CI, 0.24, 3.32; P = 0.86; Table 3 and Fig. 5). Interestingly, mortality tended to decrease at the 3-year follow-up in the cell therapy group compared with that in the control group (OR, 0.58; 95% CI, 0.22, 1.56; P = 0.28). At 5 years, 458 patients were followed up from five studies. BM-derived cell therapy significantly decreased all-cause mortality, with a 55% relative risk reduction (OR, 0.45; 95% CI, 0.21, 0.97; P = 0.04). All-cause mortality occurred in 4.4% (10 deaths in 226 patients) in the cell therapy group and 9.5% (22 deaths in 232 patients) in the control group.

In the morbidity analysis, including heart failure, recurrence of IHD, repeated revascularization, and stroke (Table 3), the cell therapy group generally showed favorable outcomes; however, this difference was not statistically significant compared with the control.

Taken together, these data indicated that BM-derived cell therapy for patients with AMI produce clinical benefits in the long-term follow-up.

**Discussion**

Here, we conducted a systematic review and meta-analysis of comparative studies to evaluate effectiveness of BM-derived cell therapy for patients with AMI. Our study included the greatest number of publications describing cell therapy trials to date and reflects the latest clinical outcomes and long-term follow-up results. We included all types of BM-derived cell therapy trials and
**Fig. 3** Infarct size reduction. Forest plot and meta-analysis of infarct size at the 6-month, 1-year, 3-year, and 5-year follow-up. **a** Infarct size at 6 months after treatment. **b** Infarct size at 1 year. **c** Infarct size at 3 years. **d** Infarct size at 5 years.

### a. Infarct size at 6 months after treatment

| Study or Subgroup | Cell therapy | Control | Mean Difference IV, Random, 95% CI | Mean Difference IV, Random, 95% CI |
|-------------------|--------------|---------|-----------------------------------|-----------------------------------|
| Lundie 2006       | -0.7 ± 5.1   | 43      | 2.6 ± 5.2 17.9%                  | 1.90 (−0.28, 4.08)                |
| Melzini 2008      | 0.5 ± 0.3    | 20      | 0.2 ± 0.2 18.9%                  | 0.50 (−0.34, 0.68)                |
| Quyyumi 2011      | -5.2 ± 6.1   | 11      | -6.7 ± 5.4 14.8%                  | 1.50 (−3.42, 6.42)                |
| Traverse 2012     | 1.7 ± 5.5    | 75      | 2.6 ± 4.9 18.0%                  | -0.90 (−2.91, 1.11)               |
| Wohrlie 2010      | -8.3 ± 5.5   | 29      | 7.8 ± 4.4 17.0%                  | -16.10 (−19.22, −12.98)           |
| Wohrlie 2013      | -22.6 ± 19.5 | 29      | -16.9 ± 9.6 10.1%                | -6.70 (−15.51, 2.11)              |
| Woilert 2004      | 14.1 ± 13    | 10      | 16.2 ± 30.3 3.3%                 | -2.10 (−22.54, 18.34)             |
| Subtotal (95% CI) | 217          | 146     | —                      | -2.99 (−7.08, 1.11)               |

Heterogeneity: $\tau^2 = 23.14$, $\chi^2 = 114.74$, df = 6 ($P < 0.00001$); $I^2 = 95$

Test for overall effect: $Z = 1.43$ ($P = 0.15$)

Test for subgroup differences: Not applicable

### b. Infarct size at 1 year

| Study or Subgroup | Cell therapy | Control | Mean Difference IV, Random, 95% CI | Mean Difference IV, Random, 95% CI |
|-------------------|--------------|---------|-----------------------------------|-----------------------------------|
| Malliaras 2014    | -11.1 ± 4.6  | 17      | -2.2 ± 7.1 19.2%                 | -8.80 (−14.14, -3.52)             |
| Melzini 2008      | 0.2 ± 0.3    | 20      | 0.1 ± 0.3 24.6%                  | 0.10 (−0.09, 0.29)                |
| Turan 2011        | -9.6 ± 6.6   | 38      | 3 ± 6.6 21.7%                    | -12.00 (−15.70, −8.30)            |
| Wohrlie 2013      | -25.1 ± 21   | 29      | -16.3 ± 14.7 11.3%               | -8.80 (−19.88, 2.28)              |
| Yao 2009          | -6.2 ± 2.8   | 12      | -2.7 ± 3.3 12.3%                 | -3.50 (−5.95, -1.05)              |
| Subtotal (95% CI) | 116          | 71      | —                      | -6.10 (−11.14, -1.05)             |

Heterogeneity: $\tau^2 = 26.97$, $\chi^2 = 62.12$, df = 4 ($P < 0.00001$); $I^2 = 94$

Test for overall effect: $Z = 2.37$ ($P = 0.02$)

Test for subgroup differences: Not applicable

### c. Infarct size at 3 years

| Study or Subgroup | Cell therapy | Control | Mean Difference IV, Random, 95% CI | Mean Difference IV, Random, 95% CI |
|-------------------|--------------|---------|-----------------------------------|-----------------------------------|
| Wohrlie 2013      | -28.5 ± 14.4 | 29      | -27.6 ± 16.4 100.0%               | 1.10 (−9.24, 11.44)               |
| Subtotal (95% CI) | 29           | 13      | —                      | 1.10 (−9.24, 11.44)               |

Heterogeneity: Not applicable

Test for overall effect: $Z = 0.21$ ($P = 0.83$)

Test for subgroup differences: Not applicable

### d. Infarct size at 5 years

| Study or Subgroup | Cell therapy | Control | Mean Difference IV, Random, 95% CI | Mean Difference IV, Random, 95% CI |
|-------------------|--------------|---------|-----------------------------------|-----------------------------------|
| Woilert 2004      | 15 ± 12.2    | 10      | 13.7 ± 24.5 100.0%               | 1.30 (−15.66, 18.26)             |
| Subtotal (95% CI) | 10           | 10      | —                      | 1.30 (−15.66, 18.26)             |

Heterogeneity: Not applicable

Test for overall effect: $Z = 0.15$ ($P = 0.88$)

Test for subgroup differences: Not applicable
analyzed 2635 patients. Among 43 studies, the majority of trials used uncultured fresh autologous BM aspirates and applied a density gradient to separate MNCs. Two trials employed G-CSF-mobilized peripheral blood MNCs, and four trials utilized CD133+ or CD34+ BM-MNCs separated by antibody-based cell sorting. Two trials cultured BM aspirates for 1 month to obtain MSCs. In terms of cell delivery, most studied applied intracoronary infusion through the balloon catheter.

Although the feasibility of cell therapy has been demonstrated in clinical trials, safety should be considered as a priority in future clinical studies. Malignant tumor formation after transplantation of BM MSCs into animal hearts suggests that the unstable chromosomal status of cultured cells could induce serious adverse events [57]. Furthermore, the calcifying activity of uncultured BM cells has also been reported in preclinical animal studies [58]. These data suggest that careful monitoring of cells before transplantation and long-term follow-up for safety should be performed in clinical trials.

The individual studies were not sufficiently powered to detect significant differences in major adverse clinical events between the cell therapy and control group. Therefore, meta-analyses can be used to evaluate appropriate results of each study and may provide insights into clinical outcomes and benefits of cell therapy. In this meta-analysis, we focused on follow-up results at 6 months, 1 year, 3 years, and 5 years in order to evaluate short-term and long-term effects of the cell therapy. We found that at 6 months, the cell-injected group showed a modest or insignificant improvement in LVEF, depending on the measurement modality. However, there was a tendency toward decreased mortality at 3 years. At the 5-year follow-up, cell injection significantly decreased all-cause mortality (55 % relative risk reduction) as compared with that in the control group, indicating a discrepancy between the short-term LV functional improvement and long-term all-cause mortality.

We further reviewed the specific mode of death of all-cause mortality. The REPAIR-AMI study at the 5-year follow-up demonstrated that BM-derived cell therapy tended to reduce cardiac death (4 deaths/100 patients) compared with that of the placebo treatment (14 deaths/100 patients) [9]. Additionally, the MAGIC-Cell study at the 5-year follow-up showed that G-CSF-mobilized cell therapy decreased cardiac death (no deaths/57 patients) compared with that of the control treatment (2 deaths/60 patients) [17]. These data suggested that the effects of cell therapy on mortality could be primarily driven by the reduction of cardiac death after MI.

Functional improvement of LV systolic function was modest, as compared to the standard care of revascularization and medical treatment of MI, BM-derived cell therapy showed better clinical outcomes and survival benefits in the long-term follow-up analysis. We would like to suggest several putative explanations for the significant clinical benefit after cell therapy. First, LV functional improvement and infarct size reduction at the early phase after revascularization with cell therapy could affect long-term outcomes according to the "legacy effect", which was described in a prior diabetes trial; the difference in glycosylated hemoglobin (HbA1c) levels between intensively and conventionally treated patients disappeared within 1 year of the completion of the trial. Nevertheless, outcomes continued to favor the intensively treated group [59]. This meta-analysis suggests that LVEF at short-term follow-up after cell therapy could affect long-term cardiovascular outcomes. REPAIR-AMI long-term follow-up at 2 and

| Outcome | Subgroup | Studies | Cell-injected group participants (n) | Control group participants (n) | Mean difference | I² (%) |
|---------|----------|---------|-------------------------------------|--------------------------------|----------------|--------|
|         |          |         | IV, random (%) 95 % CI |                       |               |        |
|         |          |         | 4.02 | 2.65, 5.39 | 94 |
|         |          |         | 1.86 | 0.54, 3.19 | 64 |
|         |          |         | 3.20 | 1.83, 4.56 | 94 |
|         |          |         | 2.63 | 1.59, 3.67 | 92 |
|         |          |         | 1.86 | 0.54, 3.19 | 64 |
|         |          |         | 3.20 | 1.83, 4.56 | 94 |
|         |          |         | 2.63 | 1.59, 3.67 | 92 |
|         |          |         | 3.20 | 1.83, 4.56 | 94 |
|         |          |         | 2.63 | 1.59, 3.67 | 92 |
|         |          |         | 3.20 | 1.83, 4.56 | 94 |
|         |          |         | 2.63 | 1.59, 3.67 | 92 |
|         |          |         | 3.20 | 1.83, 4.56 | 94 |
|         |          |         | 2.63 | 1.59, 3.67 | 92 |
|         |          |         | 3.20 | 1.83, 4.56 | 94 |
|         |          |         | 2.63 | 1.59, 3.67 | 92 |

IV inverse variance, CI confidence interval, I² inconsistency (across studies), MRI magnetic resonance imaging, LVEF left ventricular ejection fraction
5 years demonstrated favorable clinical outcomes, including cardiovascular death and rehospitalization for heart failure. Specifically, lower LVEF and increased LV end systolic volume at 4 months were associated with adverse long-term outcomes [9, 60]. These data indicated that the functional improvement of LV at the early stage after cell transplantation would induce long-term benefits, although there was no significant difference in LVEF at long-term follow-up. Furthermore, to overcome such limitations of a single cell transplantation procedure and to achieve better outcomes, repeated cell transplantation is currently under investigation [61].

Second, there is a possibility of sustained LV functional improvement with a small degree after cell therapy. Although this meta-analysis showed no significant differences in LV function and infarct size at the 3- and 5-year follow-up between the cell therapy and control groups, a previous report suggested that the improvement in LVEF may be maintained over a long-term follow-up of 1–5 years [6]. Third, microvascular improvement thorough enhanced angiogenesis and cardioprotective effects could be contributed to outcomes. However, an MRI study reported that cell therapy did not augment the perfusion of the ischemic myocardium [12]. Fourth, patients with recovered LV function after standard revascularization therapy, considered as a low-risk population, could dilute...
benefits of cell therapy. In the REPAIR-AMI trial, patients with an LVEF below 48% after the primary PCI showed LV functional improvement after cell infusion [62]. There was no difference between the cell therapy and control group in patients with LVEF above 48%. In addition, CD133+ BM-derived cell therapy during coronary artery bypass grafting (CABG) demonstrated that patients with a pre-operative LVEF of less than 40% gained more LV functional improvement after cell therapy as compared with that in patients with LVEF higher than 40% [28]. Taken together, these data suggested that patients with greater damage from AMI could receive greater benefits from BM-derived cell therapy.

More than a decade after the initial cell therapy trial for MI, even though considerable knowledge has been accumulated, there are still many challenges to achieving favorable clinical outcomes [63]. The inconsistent clinical results of autologous BM cells may be due to the considerable patient-to-patient variability related to a decrease in the number and potency of stem and progenitor cells. These discrepancies may also be caused by the limitations of functional LV measurement. Although cardiac MRI is considered the gold standard for measurement of the LVEF, volumes and infarct size, it is difficult to distinguish the infarct from myocardia edema using MRI. Moreover, the cell processing strategy is also a controversial factor that may yield inconsistent clinical results.

The mechanisms mediating the benefits of cell injection remain unclear. In molecular and cellular analyses of BM-derived cell therapy, recent accumulating data have revealed the limited therapeutic efficacy of this strategy. Although BM cells have shown cardiovascular differentiation after transplantation into infarcted hearts, transdifferentiation is considered a rare event [64]. In vivo cell tracking studies in patients using FDG-PET have demonstrated that less than 1% of delivered cells stay in the myocardium after 24 hours of cell injection, indicating poor engraftment [65]. Therefore, paracrine effects of injected BM cells via the release of several humoral factors have also been proposed as the main mechanism of action [66]. However, autologous cells from diseased and elderly patients impair cellular function for regeneration and repair of infarcted hearts. To overcome such limitations, clinical trials investigating resident cardiac progenitor cell-based therapy and allogeneic healthy donor-derived cell therapy as well as repeated cell administration are ongoing to determine the merits of these second-generation cell therapies.

### Limitations

This study has several limitations. First, pooled data for the meta-analysis showed different baseline characteristics between the cell therapy and placebo groups, which likely reduces the comparability of the meta-analysis. Furthermore, according to the specific study protocol of

### Table 3 Major adverse events

| Event                          | Follow-up | Study (participants) | Cell therapy group | Control group | Odds ratio (M-H, random) CI |
|-------------------------------|-----------|-----------------------|--------------------|---------------|-----------------------------|
|                               | Total event | Participants          | Total event | Participants |                             |
| All-cause mortality           | 6 mo 18 (1284) | 10 707                | 6 577              | 1.08 | 0.42–2.81 |
|                               | 1 yr 8 (286) | 5 174                | 4 112              | 0.89 | 0.24–3.32 |
|                               | 3 yr 5 (432) | 9 228                | 11 204             | 0.58 | 0.22–1.56 |
|                               | 5 yr 5 (458) | 10 226               | 22 232             | 0.45 | 0.21–0.97 |
| Heart failure admission       | 6 mo 9 (970) | 10 512               | 15 458             | 0.56 | 0.24–1.31 |
|                               | 1 yr 3 (83)  | 2 42                 | 0 41               | 3.07 | 0.30–30.96 |
|                               | 2 yr 4 (333) | 4 179                | 14 154             | 0.15 | 0.04–0.50 |
|                               | 3 yr 6 (577) | 11 284               | 17 293             | 0.65 | 0.30–1.44 |
| Recurrence of ischemic heart disease | 6 mo 14 (1214) | 29 663               | 37 551             | 0.57 | 0.32–1.00 |
|                               | 1 yr 2 (81)  | 0 48                 | 1 33               | 0.14 | 0.01–3.68 |
|                               | 2 yr 6 (373) | 6 199                | 11 174             | 0.68 | 0.18–2.56 |
|                               | 3 yr 6 (528) | 9 262                | 11 266             | 0.81 | 0.33–2.00 |
| Revascularization/restenosis | 6 mo 14 (1212) | 98 662               | 92 550             | 0.87 | 0.63–1.22 |
|                               | 1 yr 4 (156) | 2 87                 | 4 69               | 0.37 | 0.07–1.89 |
|                               | 2 yr 6 (373) | 36 199               | 45 174             | 0.65 | 0.39–1.10 |
|                               | 3 yr 6 (577) | 62 284               | 76 293             | 0.77 | 0.52–1.15 |
| Cerebral vascular accident (CVA) | 1 yr 2 (150) | 1 68                 | 2 82               | 0.69 | 0.08–5.81 |
|                               | 2 yr 3 (500) | 5 248                | 10 252             | 0.5 | 0.16–1.50 |

M-H Mantel-Haenszel, CI confidence interval, mo months, yr years
Fig. 5 Forest plot and meta-analysis of all-cause mortality. 

**a** All-cause mortality at 6 months after treatment.

**b** All-cause mortality at the 1-year follow-up.

**c** All-cause mortality at the 3-year follow-up.

**d** All-cause mortality at the 5-year follow-up.
each study, there were several differences in procedures for cell harvesting, separation, mode of delivery, and timing of delivery. Second, the number of clinical events was small during the follow-up period, although patients suffered from AMI. In 5 years after enrollment, a total of 32 deaths occurred in 458 patients (4.4 % mortality rate in the cell therapy group and 9.5 % mortality rate in the control group). This low event rate could be explained by the exclusion of high-risk patients in most trials, such as those with severe LV dysfunction and heart failure after large myocardial infarction. Clinical events in heart failure patients who were hospitalized due to decompensation have been reported to be approximately 6 % for in-hospital deaths, and even in patients who were discharged alive, the 6-month all-cause mortality rate was 10 % [67]. Accordingly, cell therapy trials for high-risk patients with AMI should be performed to clearly address the clinical benefits of the new therapeutic modality.

Conclusions
We conducted a meta-analysis of BM-derived cell therapy for patients with AMI and showed that this therapy was associated with long-term mortality benefits, although the effects of LV functional improvement and infarct size reduction were modest in short-term analyses. In this regard, to validate the therapeutic efficacy of cell therapy, future randomized clinical trials should include sufficient sample sizes with greater power to detect long-term clinical outcomes rather than surrogate short-term hemodynamic and image-based variables.

Abbreviations
AMI: acute myocardial infarction; BM: bone marrow; CABG: coronary artery bypass grafting; G-CSF: granulocyte colony-stimulating factor; IHD: ischemic heart disease; LVEF: left ventricular ejection fraction; MNC: mononuclear cell; MRI: magnetic resonance imaging; MSC: mesenchymal stromal cell; PB: peripheral blood; PCI: percutaneous coronary intervention; RCT: randomized clinical trial

Acknowledgements
This study was funded by the Global Stem Cell and Regenerative Medicine Acceleration Center (GSRAC) and supported by a grant from the Innovative Research Institute for Cell Therapy, Seoul National University Hospital (A062260) sponsored by the Ministry of Health and Welfare, Republic of Korea.

Funding
This study was funded by Global Stem Cell and Regenerative Medicine Acceleration Center from the Ministry of Health and Welfare, Republic of Korea.

Availability of data and materials
There is no supporting or supplementary material.

Authors’ contributions
SHL, JHH, KHC, JWN, and HJC were involved in designing the study and drafting the manuscript. HJC made substantial contribution to the concept and design of the study, study supervision, and finalizing the manuscript for important intellectual content. SHL and JHH performed data analysis, and made substantial contributions to the acquisition of data. All authors read and approved the final manuscript.

Competing interests
The authors declare that they have no competing interests.

Consent for publication
All authors consent for publication.

Ethical approval and consent to participate
Ethical approval is not applicable. All authors consent to participation.

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Received: 11 July 2016 Revised: 18 September 2016 Accepted: 27 September 2016 Published online: 20 October 2016

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