Review Article

Dermatological Manifestations in the Intensive Care Unit: A Practical Approach

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Dermatological problems are not usually related to intensive medicine because they are considered to have a low impact on the evolution of critical patients. Despite this, dermatological manifestations (DMs) are relatively frequent in critically ill patients. In rare cases, DMs will be the main diagnosis and will require intensive treatment due to acute skin failure. In contrast, DMs can be a reflection of underlying systemic diseases, and their identification may be key to their diagnosis. On other occasions, DMs are lesions that appear in the evolution of critical patients and are due to factors derived from the stay or intensive treatment. Lastly, DMs can accompany patients and must be taken into account in the comprehensive pathology management. Several factors must be considered when addressing DMs: on the one hand, the moment of appearance, morphology, location, and associated treatment and, on the other hand, aetiopathogenesis and classification of the cutaneous lesion. DMs can be classified into 4 groups: life-threatening DMs (uncommon but compromise the patient’s life); DMs associated with systemic diseases where skin lesions accompany the pathology that requires admission to the intensive care unit (ICU); DMs secondary to the management of the critical patient that considers the cutaneous manifestations that appear in the evolution mainly of infectious or allergic origin; and DMs previously present in the patient and unrelated to the critical process. This review provides a characterization of DMs in ICU patients to establish a better identification and classification and to understand their interrelation with critical illnesses.

1. Introduction

Dermatological problems have received limited attention within the field of intensive care medicine, probably because they are considered within a specialty with a large outpatient component and rarely are serious and because of the limited impact they have on the prognosis of critical patients. Less than 0.5% of patients requiring admission to an intensive care unit (ICU) do so because of a primary dermatological problem [1]. However, and more generally, dermatological manifestations (DMs) in critical patients are relatively frequent and can be markers of extracutaneous pathology. In a previous study, we observed that 10.4% of patients admitted to an ICU had some type of DM [2], and a study by Agrawal observed DMs in up to 42.2% of critically ill patients [3]. Identifying these manifestations can be key in the recognition of an underlying disease. Examples include skin manifestations in meningococcal sepsis, skin as an expression of disseminated candidiasis, or echyma gangrenosum in Pseudomonas aeruginosa infection [4, 5]. Other times, DMs reflect secondary effects of drugs or devices [6, 7], problems derived from procedures [8], or simply a patient’s critical situation [9, 10]. DMs can be presented in a highly varied way, and a definitive diagnosis often requires the collaboration of a dermatologist.

The objective of this review is to describe the most frequent DMs found in critically ill patients. Thermal injuries and necrotizing fasciitis, which constitute differentiated entities, are not included. This review consists of several sections: we will analyse the differentiating characteristics and the classification of DMs in critical patients, review
published series, describe schematically the different types of DMs, and close with the conclusions.

1.1. Differentiating Characteristics of Dermatological Manifestations in Critical Patients. The management of critical patients requires a multidisciplinary evaluation and management as a fundamental part of their treatment. Rarely are patients admitted due to a dermatological disease; however, the DMs that appear during the evolution of the patients and that require treatment are relatively frequent [11]. Prolonged immobilization, insufficient nutritional support, impaired tissue perfusion, changes in body temperature, hygiene challenges, use of multiple drugs, and invasive processes may facilitate the development of DMs in ICU patients [12].

To these circumstances, we must add the lack of symptoms due to an altered state of consciousness, either by the use of sedative drugs or by neurological impairment, which makes communication with the patients difficult. There is little literature that specifically addresses DMs in the critical patient context, and there is no homogenous classification available.

Patients respond to the aggression of critical illnesses with profound changes at the immunological, endocrine, and vascular levels that result in impairments in tissue perfusion and changes in the microcirculation of the skin that alter the signs and symptoms of cutaneous lesions [13].

DMs that can be identified in Intensive Care Medicine have been classified in different ways. In some cases, a classification based on morphology or lesions [14] has been used according to whether the DM required the care of a dermatologist or only an intensivist [15]. We believe, concurring with other authors, that the classification that should be used in critical patients must have a dermatological base, but it should also include groups with differentiating characteristics in the diagnosis, treatment, and evolution during admission to the ICU [16–18].

In Table 1, a classification is proposed that includes the particularities of ICU patients. There are 4 main groups: life-threatening DMs (corresponding in the literature with the term primary lesions); DMs associated with systemic diseases (may indicate an underlying disease); DMs secondary to the management of the critical patient (divided into infectious, drug reactions, and secondary to devices); and previous DMs (named coincidental in previous studies [16, 17]) that, if not identified, could hinder or confuse the main diagnosis of the patient.

1.2. Review of Published Series on Dermatological Manifestations in the ICU. Table 2 shows the published works that include the follow-up of series of patients admitted to the ICU where the main DMs are analysed. Some studies focus on DMs associated with ICU treatment [15, 19]; others, such as studies on paediatric patients, include a morphological classification attributable to the importance of exanthematic childhood diseases.

It is difficult to compare different publications due to the lack of homogeneity in the DM classification; therefore, we have adapted the results obtained to the classification devised by Dunn et al. [16] in a classic study that, with some modifications, has been maintained in our publications and in those made by Awa et al. [17, 18]. The advantage offered by this classification is that it groups DMs based on the situation of the critical patients without ignoring the origin of the DMs. Different studies show that dermatological admission is infrequent but that DMs usually accompany the evolution of critical patients either in the context of a systemic disease or adding to their main pathological process, influencing stay, and mortality.

The differences attributed to the country of origin of the different articles determine the type of lesions identified, as is the case in the study by Prashanth and Pai, in which 64% of patients had Dengue haemorrhagic fever [20]. We also verified the importance of the year of publication of the study. In our case, we went from 12 patients with DMs secondary to meningococcal meningitis in 1999 to 1 case in 2013, a situation that we attribute to the broad use of a meningococcal vaccine [2].

As limitations of the different studies, except for George [1], the studies were carried out in a single centre and were mainly retrospective, which hinders the generalization of the results. Furthermore, the type of DM studied differs among the authors. Some studies included necrotizing fasciitis [1] or pressure ulcers [18], which we exclude from classification because they have very well-defined characteristics and are treated independently in most cases.

All the studies described collaboration with a dermatologist specialist, either as a principal physician or as a consultant.

2. Classification of Dermatological Manifestations in Intensive Medicine

2.1. Life-Threatening Dermatological Manifestations.
| Publication year | 1994 | 1999 | 2008 | 2011 | 2012 | 2012 | 2013 | 2013 | 2013 | 2014 | 2017 | 2017 | 2018 | 2018 |
|-----------------|------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| Period (years)  | 1    | 2    | 11   | 3    | 2    | 1    | 3    | 1    | 3    | 1    | 3    | 3    | 2    | <1   |
| Country         | UK   | Spain| UK   | Holland | India | Germany | Spain | Turkey | India | Guatemala | Turkey | Turkey | India | India |
| Type of study   | Retros. | Prosp. | Retros. | Prosp. | Retros. | Prosp. | Retros. | Prosp. | Prosp. | Prosp. | Prosp. | Prosp. | Prosp. |
| Type of patient | Adult | Adult | Adult | Pedia. | Adult | Adult | Adult | Adult | Adult | Adult | Adult | Adult | Adult | Adult |
| No. of patients | 27   | 46   | 2406  | 42   | 318  | 154  | 90   | 427  | 47   | 73   | 312  | 50   | 312  | 50   |
| No. of lesions  | 27   | 46   | 2406  | 42   | 318  | 154  | 90   | 427  | 47   | 73   | 312  | 50   | 312  | 50   |
| Type of lesions (N) | | | | | | | | | | | | | |
| Life-threatening | 5    | 0    | 199   | 3    | 3    | 7    | 5    | —    | | | | | | |
| Systemic       | 10   | 14   | 61    | 18   | 284  | —    | 26   | —    | 29   | —    | —    | 0    | 20   | 2    |
| Associate       | 9    | 34   | 12    | 15   | 8    | 20   | 45   | 61   | 159  | 38   | —    | 0    | 109  | 1    |
| Previous        | 3    | 3    | 191   | 2    | 14   | 8    | —    | —    | 67   | —    | 96   | 60   | 121  | 36   |
| Other           | —    | —    | 1943  | 8    | 9    | 20   | 14   | 29   | 94   | —    | 13   | 2    | 46   | 1    |
| Acute skin failure (N) | 199 | | | | | | | | | | | | | |
| APACHE II       | 17.2 | 18   | 17.4  | | | | | | | | | | | |
| Length of stay (IQR) | 19   | —    | 16    | 23   | 5–57 | 10 (13) | 17 | (3–43) |
| Mortality ICU (%) | 33   | 26   | 27.5  | 16   | 21.8 | 46.1 | 31.1 | 46.6 | 11.2 |
| Mortality HOSP (%) | 39.6 | | 38.3  | | | | | | |

Prosp.: prospective study. Retros.: retrospective study. UNI: unicenter study. MULTI: multicenter study.
These DMs require admission to the ICU. They are rare (as seen in Table 2) but severe and are associated with high mortality. They cause loss or deterioration of cutaneous functions, leading to an imbalance and failure of the cutaneous barrier, defined as acute skin failure [21]. Destruction of the stratum corneum, the outermost layer of the epidermis, causes the loss of fluids, proteins, and electrolytes, immunological dysfunction with the consequent risk of infection, thermoregulatory impairment, and a hypercatabolic state.

Within this group, we include various entities such as immunobullous diseases, generalized pustular psoriasis, erythroderma, severe cutaneous adverse reactions (SCARs), and infectious diseases such as scalded skin syndrome.

2.1.1. Immunobullous Skin Diseases. Immunobullous diseases represent a heterogeneous group of entities in which the typical lesion is a blister due to the loss of adhesion between intraepidermal cells or between the basal keratinocytes to the basement membrane; according to the degree of systemic repercussions, these diseases may require critical treatment, with pemphigus vulgaris as its potentially most serious form. The bullous pemphigoid, more common in the elderly, usually has a self-limited course with low mortality rate.

The paraneoplastic pemphigus (PNP) is a variety of pemphigus with differential characteristics. The PNP is an autoimmune blistering syndrome that can affect multiple organs other than skin and is closely related with benign or malignant tumours especially lymphomatoid and hematologic malignances. Patients with PNP can develop life-threatening restrictive bronchiolitis obliterans that is irreversible with lung transplant being the only possible alternative. PNP treatment should be focused on the eradication of the associated tumour process. Cutaneous manifestations usually have a poor response to treatment, being better than those associated with benign processes such as thymoma or Castleman’s disease [22].

The pemphigus group has desmogleins as the main antigen. Autoantibodies directed against these proteins inhibit their adhesive function, causing loss of the intercellular binding of keratinocytes.

The pemphigus group of blistering autoimmune diseases is characterized by mucocutaneous blisters that break easily and become erosions. Upon examination, a positive Nikolsky’s sign (skin detachment when applying lateral pressure on the skin near an ampoule) is typical, and the diagnosis is made by skin biopsy. Treatment should combine skin care with anti-inflammatory and immunosuppressive aetiological treatment with corticosteroids, immunoglobulins, and, in recent years, mycophenolate mofetil; biological therapies such as rituximab have been added with satisfactory results [23].

2.1.2. Generalized Pustular Psoriasis. Generalized pustular psoriasis (GPP), also known as Von Zumbusch disease, is an inflammatory skin disorder with multisystemic and potentially fatal affection. Recently, a genetic mutation in \( \alpha \) or other involved interleukins [26, 27].

2.1.3. Erythroderma. Erythroderma is an exfoliative dermatitis characterized by erythema that affects more than 90% of the skin surface with a variable degree of desquamation. It is a cutaneous manifestation of various skin diseases (psoriasis and atopic dermatitis), drug reactions (allopurinol, carbamazepine, ciprofloxacin, etc.), or malignant processes (T-lymphoma or paraneoplastic phenomena) [28]. Patchy areas of erythroderma are observed and evolve to affect most of the skin surface accompanied by a variable degree of scaling. The skin appears as intense red, bright, dry, and scaly and is often accompanied by pruritus and ectropion. Frequently, the clinical presentation indicates the underlying disease, with psoriasis being the most frequent cause followed by drug reactions [29]. Skin biopsy may be useful to determine the aetiology, although sometimes the findings are nonspecific and may require multiple biopsies. Erythroderma treatment is based on controlling the underlying disease, removing external contributing factors, and providing supportive therapy. At the systemic level, adequate hydration must be maintained with fluid replacement, electrolyte correction, and meticulous skin care [30].

2.1.4. Severe Cutaneous Adverse Reactions (SCARs). SCARs are unpredictable and infrequent but have high morbidity and mortality. Adverse drug reactions alter the immune response and cause extensive epidermolysis. The main conditions that encompass noninfectious SCARs are Stevens–Johnson syndrome (SJS), toxic epidermal necrolysis (TEN), and drug-induced hypersensitivity syndrome/drug reaction with eosinophilia and systemic symptoms (DISH-DRESS) [31].

SJS and TEN are considered the same entity, with different degrees of severity. They are characterized by erythema with extensive epidermal detachment and mucous membrane erosion. The classification is based on the percentage of affected skin area. Epidermolysis of less than 10% is defined as SJS, detachment between 10 and 30% is classified as SJS/TEN overlap, and detachment greater than 30% is classified as TEN [32].

The first clinical manifestation is usually fever and malaise followed by generalized erythema with the appearance of blisters that break easily and erosion in mucous membranes of the mouth, eyes, and genital area. In the most extensive forms, erosion and necrosis can affect the conjunctiva, bronchi, trachea, intestine, and kidney, with a mortality ranging between 25 and 30%. The severity of the
clinical picture can be determined using the SCORTEN severity scale, designed as a predictor of mortality from toxic epidermal necrolysis. It uses 7 easy-to-measure items: age, presence of malignancy, tachycardia, degree of skin detachment, and plasma levels of urea, blood glucose, and bicarbonate [33].

Some medications alter the immune response, together with a genetic susceptibility that causes the development of SCARs [32]. Sulfonamides are the most frequently implicated drugs, followed by cephalosporins, quinolones, imidazoles, and anticonvulsants such as carbamazepine and lamotrigine (Figure 1). In recent years, specific human leucocyte antigen (HLA) genotypes have been associated with the appearance of SCARs with the use of carbamazepine [34].

DISH-DRESS is a drug-induced, multisystemic, and infrequent reaction that occurs between 3 and 6 weeks after the administration of a drug and has been associated with the reactivation of herpes virus 6. The main causative agents are anticonvulsants (carbamazepine, phenytoin, phenobarbital, mexiletine, and lamotrigine), allopurinol, and sulfasalazine. It is defined by a generalized, long-lasting maculopapular rash or any other skin rash that progresses occasionally to erythroderma, accompanied by fever, lymphadenopathy, and impaired liver function as the most common organ involvement [35].

Diagnosis by skin biopsy is the only method to confirm the nature of the skin reaction [36]. The main differentiating diagnosis is scalded skin syndrome caused by Staphylococcus species and other cutaneous autoimmune skin diseases such as pemphigus vulgaris and pemphigoid of the mucous membrane. Treatment should include immediate withdrawal of the suspect drug and life-support therapy [37].

2.1.5. Toxic Shock Syndrome. Toxic shock syndrome (TSS) is a rare, acute Gram positive infection characterized by fever, malaise, gastrointestinal symptoms, confusion, weakness, and refractory hypotension associated with diffuse, red, macular rash that can involve both skin and mucous membranes, conjunctival hyperaemia, and “strawberry tongue.” Desquamation occurs 1-2 weeks after disease onset. TSS is typically caused by exotoxins produced by Staphylococcus aureus or group A Streptococcus [38]. Some cases have been related to the use of vaginal tampons and vaginal colonization by Staphylococcus facilitating the production of exotoxin. Nonmenstrual cases may result around surgical site, burns, nasal packing, postinfluenza pneumonia, postpartum infections, or insulin pump infusion sites, or no source may be identified. Patients should be managed in accordance with sepsis guidelines. Antibiotics should be initiated against S. aureus and S. pyogenes and an aggressive source control is mandatory, which may include surgical debridement, removal of invasive devices, or vaginal examination in the case of suspected menstrual TSS [39].

2.1.6. Staphylococcal Scalded Skin Syndrome. Staphylococcal scalded skin syndrome (SSSS) is a bullous disease that is potentially life-threatening due to infection with Staphylococcus aureus, which produces an exfoliative exotoxin [40]. The bacterium is located in mucous membranes, and its toxin spreads through the bloodstream, causing intraepidermal detachment due to breakage of the desmoglein 1 complex far from the initial focus (positive Nikolsky’s sign). This pathology can affect any age, but it is more frequent in children under 5 years of age; however, when it affects adults, it is usually more serious, with mortality up to 60%. It can be confused with TEN, but its main differences are that it does not usually affect mucous membranes, is not related to drug exposure, and has a different histological pattern. In the treatment, the initiation of the appropriate antibiotic drug early is essential [41].

2.2. Dermatological Disorders Associated with Systemic Diseases. DMs can precede, accompany, or follow the evolution of a systemic disease. A wide variety of diseases can present DMs. The DMs that we can most frequently find in the field of intensive care are secondary to peripheral vascular damage produced in the context of a systemic underlying disease or are manifestations of an immune disease such as vasculitis or connective tissue diseases.

2.2.1. Purpura Fulminans. Venous insufficiency with cutaneous infarction and disseminated intravascular coagulation can appear in the context of severe sepsis; however, it can also be due to a congenital deficit of protein C or protein
Calciphylaxis is a rare multifactorial cutaneous vascular disease with predilection for patients with end-stage renal disease characterized by occlusion of microvessels in the subcutaneous adipose tissue and dermis. Calciphylaxis causes painful cutaneous lesions and is frequently accompanied by tactile hyperesthesia. A dusky discoloration of the skin indicates and area of imminent necrosis [47]. Sepsis originating from these resultant wounds is considered the most common cause of death. Although skin manifestations dominate the clinical presentation, calciphylaxis is likely a systemic disorder and extraskeletal calcifications are often found on imaging studies. Skin biopsy is the standard method for confirmation suspected calciphylaxis. However, the biopsy is not needed in patients with typical lesions and end-stage renal disease. Treatment should be aimed at controlling pain and wound care with removal of necrotic tissue. Other therapeutic options such as sodium thiosulfate or bisphosphonates have proven effective in some cases [48].

2.2.4. Cutaneous Vasculitis. Vasculitis is a group of complex and disabling diseases that are characterized by an inflammatory infiltrate in or around blood vessels that causes vascular damage. Skin involvement is common, and its form of presentation depends on the size of the affected vessel. The classification of vasculitis, much debated in recent years, is based on the size of the affected vessel, the pathogenesis, and the main organ involved [49]. We distinguish 2 types of cutaneous vasculitis: small vessel vasculitis and single-organ vasculitis. Clinically, the typical lesion is palpable purpura; however, other lesions such as ulcers, blisters, or livedo reticularis may be associated. Palpable purpura is very suggestive of small vessel vasculitis and affects arterioles, capillaries, and postcapillary venules. Skin biopsy is the gold diagnostic standard and allows differentiation from other nonvasculitic processes with similar clinical manifestation due to vessel wall dysfunction or their occlusion, as observed in coagulation diseases or purpura fulminans.

The most typical presentation is leukocytoclastic vasculitis or small vessel vasculitis restricted to the skin. The most frequent manifestation is palpable purpura (approximately 2–10 mm in diameter) located in the lower extremities and buttocks with a symmetrical distribution. On occasion, arthralgias and/or arthritis may also appear with general symptoms such as fever, malaise, anorexia, and myalgias. Systemic involvement is rare, and if it occurs, it is of mild intensity [50].

The treatment of this vasculitic process is symptomatic. In cases with extensive skin involvement, the use of corticosteroids may be effective. Severe forms may require corticoids at higher doses, dapsone, and even azathioprine or rituximab [51].

Another type of small vessel vasculitis is ANCA-associated vasculitis (AAV) that includes granulomatosis with polyangiitis, microscopic polyangiitis (MPA), and eosinophilic granulomatosis with polyangiitis (EGPA). AAV can affect multiple organs systems and the patient may need to be admitted to the ICU. Granulomatosis with polyangiitis, previously known as Wegener’s granulomatosis, is defined as a necrotizing granulomatous inflammation of the upper and lower respiratory tracts with necrotizing vasculitis of small and medium-size vessels and usual renal involvement. MPA belongs to the systemic vasculitis and the major organs involved are the kidneys and the lungs. EGPA, previously called Churg–Strauss syndrome, typically occurs in patients with preexisting asthma and involves the skin, lungs, heart, and peripheral nerves.

The most common skin manifestation of ANCA-associated vasculitis is palpable purpura located on limbs. Other
manifestations include livedo reticularis, nodules, urticarial lesions, and skin ulcers with necrosis [52].

Cutaneous manifestations in vasculitis are not exclusive to those affecting the small vessel. Medium and large vessel vasculitis can show DMs even though the affected vessels are not in the skin [53]. In large vessel vasculitis such as Takayasu arteritis and giant-cell arteritis, Raynaud’s phenomenon, digital gangrene, or unilateral digital clubbing, is typical, although in Europe and North America it is more frequent to manifest as acute inflammatory nodules or erythema nodosum-like lesions. Two major types of medium vessel vasculitis are Kawasaki disease and polyarteritis nodosa (PAN). Kawasaki disease is characterized by a mucocutaneous lymph node and polymorphous exanthema varying from macular to maculopapular or morbilliform form. The most frequent DMs of PAN are, in the order of frequency, purpura, livedo, and nodules with distal gangrene being less frequent. The systemic form of PAN is the most severe and 28 to 60% are accompanied by skin manifestations. The cutaneous form of PAN, less frequent, may present nodules (74–80%), livedo reticularis (56–74%), and ulcerations (8–51%) [54].

2.2.5. Connective Tissue Diseases. Connective tissue diseases such as scleroderma, lupus erythematosus, and dermatomyositis within the group of idiopathic inflammatory myopathies are autoimmune diseases of unknown origin that can affect different organs and have characteristic DMs. These 3 pathologies may require admission to the ICU due to their impact on vital organs or complications derived from immunosuppressive treatment. However, some patients may present diagnostic criteria for more than one specific connective tissue disease, as in mixed connective tissue disease (MCTD), making specific diagnosis difficult.

Scleroderma is characterized by the thickening and adhesion of the tegument due to increased production of collagen in the dermis, hypodermis, and some organs which leads to fibrosis of the skin and underlying tissues. Localized or morphea forms exist, and systemic forms can include organ involvement.

Systemic lupus erythematosus (SLE) can affect any organ and tissue of the body. At the cutaneous level, the most typical manifestation is a malar rash or butterfly rash characterized by raised erythematous lesions at the malar level, along the nasolabial sulcus, that can be precipitated by sun exposure and leaves no scar. Moreover, skin lesions could be plaques with epidermal atrophy that can leave a hypopigmented area on sun-exposed and sun-protected areas [55].

The main manifestations of dermatomyositis are skin disorders and skeletal muscle weakness. One of the most typical lesions is Gottron's papules, present in more than 70% of cases, in the form of slightly raised lesions with a purplish appearance that appear on the back of the metacarpophalangeal and interphalangeal joints. Also characteristic is Gottron’s sign with macules or erythematous plaques that are located on the back of joints such as the elbow or knee. Other common lesions are heliotrope erythema in the form of violet-coloured or periorbital erythematous macules, periangual telangiectasias, and symmetrical violaceous erythema, which appear in exposed areas such as the front and back of the upper trunk, arms, or face [56]. The recognition of these DMs is of special interest due to their association in more than 30% of cases with neoplasms, interstitial pulmonary fibrosis, and heart disease. Autoantibodies in idiopathic inflammatory myopathies research will help us identify different clinical subgroups [57, 58].

MCTD is a rare systemic autoimmune rheumatic disease characterized by the association of clinical manifestations of SLE, systemic skin sclerosis, and dermatomyositis in the presence of high levels of anti-U1-RNP antibodies. Symptoms at the cutaneous level are varied [59]. Raynaud’s phenomenon is an early feature in more than 90% of patients. Sclerodactyly and nail fold vascular changes are also common. Skin rashes may resemble lupus or dermatomyositis. Digital infarcts may present in some patients and it is rare that it presents as truncal scleroderma. These symptoms are usually accompanied by poliarthritis, hand oedema, myositis, and esophageal hypomotility. Treatment is based on controlling symptoms and should be early in the face of potentially serious manifestations such as pulmonary hypertension. Immunosuppressive therapy and steroids remain the therapeutic mainstay for MCTD [60].

2.3. Dermatological Manifestations Secondary to the Situation and Management of Critical Patients. Most DMs appear during admission to the ICU and are due to factors surrounding critical patients. During the stay in the ICU, immobilization, malnutrition, impaired tissue perfusion, immune dysfunction, fluctuations in body temperature, or difficulties in maintaining hygiene are factors that affect the cutaneous barrier and favour the appearance of cutaneous lesions [20]. We include lesions of infectious origin, from drug reactions or secondary to the use of devices.

2.3.1. Dermatological Disorders of Infectious Origin. Lesions secondary to an infectious agent are the most frequent DMs in the intensive setting and appear in the evolution of critical illness [61]. Bedding, humidity, obesity, etc., favour the appearance of inflammatory dermatitis in large
Angioedema consisting of oedema of the mucous membranes, including the respiratory and intestinal tracts. The drugs commonly associated are penicillins and derivatives and nonsteroidal anti-inflammatory drugs.

Treatment is based on the withdrawal of the suspect drug and antihistamines or steroids for symptom control [36].

2.3.3. Dermatological Disorders Secondary to Devices. Patients admitted to the ICU require advanced monitoring and treatment that require the use of various devices, both invasive and noninvasive, that can cause varying degrees of DMs. Complications from the use of devices have usually been related to their capacity to produce infection [64] or to problems related to their placement [65, 66]. Invasive procedures or some devices used in the monitoring or treatment of critical patients can cause iatrogenic injuries or inflammatory reactions that cause contact dermatitis [8].

When implanting a catheter or performing a tracheostomy, for example, the skin barrier is broken, which can cause injuries that become permanent; sometimes the injuries are imperceptible, but sometimes they leave an unsightly lesion.

Contact dermatitis is an eczematous disease due to the inflammatory reaction of the skin to an external agent such as antiseptics, self-adhesive electrodes, or other devices [67].

2.3.4. Dermatological Disorders-Related Pressure Injury. Critically ill patients are at high risk of developing pressure ulcers. Factors related to the development of these ulcers are prolonged stay, immobility, deep sedation, use of vasoactive drugs, hypotension, anasarca, organ dysfunction, and malnutrition [68]. There are various strategies aimed at preventing the appearance of ulcers such as the use of pressure redistribution equipment, intense physiotherapy, protective dressings, and trying to achieve an optimal state of general health. However, despite use of best practice measures, pressure ulcers can occur, and their identification and early treatment are essential for their good evolution [69].

2.3.5. Dermatological Disorders Induced by Vasopressor Drugs. Skin necrosis can appear as a complication of the infusion of vasopressor drugs. Classically, it is related to the use of high doses of dopamine and norepinephrine. The existence of disseminated intravascular coagulation and cardiocirculatory shock are risk factors for the development of ischemic skins lesions that can progress to symmetrical peripheral gangrene related to vasopressors [70]. Vasopressin, useful in the treatment of catecholamine-resistant vasodilatory shock, may also provoke cutaneous ischemia, due to its vasoconstrictor action on the arteriolar level. The ischemic skin lesions are normally localized in the distal area of the extremities and the trunk. When the administration of vasopressin is by a peripheral vein, it may provoke local cutaneous ischemia if it infiltrates into the subcutaneous tissue. To avoid the potential for ischemic lesions, administration of vasopressin should be by central venous catheter, and careful monitoring of the extremities to detect ischemic changes should be carried out. Early recognition and prompt

Figure 5: Candida intertrigo: genitocrural intertrigo in the inframammary fold infected by Candida albicans. Note. Satellite lesions.
treatment with systemic, local, or topical sympathetic blockers or vasodilators may be beneficial in treating ischemia and the definitive treatment that has been established for gangrene is amputation of ischemic limbs [71].

2.4. Previous Dermatological Disorders. Patients may present dermatological disorders that may need to be identified when admitted to the ICU (e.g., when a patient cannot provide his or her medical history or information is not available). Some chronic skin diseases such as psoriasis or connective tissue diseases must be recognized and could be exacerbated during admission, requiring specific treatment. Other processes, such as lichen planus and atopic eczema, may require symptomatic treatment. Other diseases, due to their characteristics, may involve chronic treatment that must be taken into account in the management of the admitted patient, or the patient may manifest lesions unrelated to the pathology of admission that do not require treatment but that may complicate the evaluation of the patient, such as cutaneous tumours, cutaneous infectious diseases, and pigmentation disorders; in these cases, it is necessary to consult a dermatology specialist (Figure 6).

3. Conclusions

The present review includes the DMs that may appear in the intensive care setting. In many cases, making it difficult to diagnose, the patient cannot manifest clinical aspects (pain, itching, etc.) because of sedation or neurological disorders. The algorithm proposed by Jack et al. can be used to identify the most frequent DMs in ICU. In this algorithm, DMs with vital compromise are first identified. In the next step, differentiate the processes that appear during or before ICU admission and determine the priorities of the consultation with the dermatology specialist [72].

The great variety of skin problems highlights the importance of including a systematic examination of the skin in ICU care routines [73]. The classification we have used defines different groups of skin pathologies.

Life-threatening DMs are infrequent, but they must be quickly recognized, and initiation of specific treatment and maintenance at an early stage is essential. DMs associated with systemic diseases offer the possibility of adding dermatological signs to skin examinations that help in the diagnosis of underlying disease. In addition, DMs that occur during the management of critical patients must be expected and diagnosed during daily patient care.

The complexity of critical patients requires the collaboration of different health professionals. Intensive care professionals should consult dermatologists. We must provide comprehensive care covering any medical and surgical pathology; therefore, detection and control of different skin problems should be incorporated into routine quality care [74].

Abbreviations

AAV: ANCA-associated vasculitis
DMs: Dermatological manifestations
DISH-DRESS: Drug-induced hypersensitivity syndrome/drug reaction with eosinophilia and systemic symptoms
EGPA: Eosinophilic granulomatosis with polyangiitis
GPP: Generalized pustular psoriasis
ICU: Intensive care unit
MPA: Microscopic polyangiitis
MCTD: Mixed connective tissue disease
MMP: Mucous membrane pemphigoid
PAN: Polyarteritis nodosa
PNP: Paraneoplastic pemphigus
SCARs: Severe cutaneous adverse reactions
SJS: Stevens–Johnson syndrome
SLE: Systemic lupus erythematosus
SSSS: Staphylococcal scalded skin syndrome
TSS: Toxic shock syndrome
TEN: Toxic epidermal necrolysis.

Data Availability

No data were used to support this study.

Additional Points

DMs in critical patients are relatively frequent, and they are predominantly consequent to critical illness, treatment, or marker of extracutaneous pathology. The great variety of skin problems highlights the importance of including a systematic examination of the skin in ICU routine quality care.

Consent

All images have consent for publication.
Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

MB and JT were major contributors in writing the manuscript. JMC provided his dermatological point of view. All authors read and approved the final manuscript.

References

[1] S. M. George, D. A. Harrison, C. A. Welch, K. M. Nolan, and P. S. Friedmann, "Dermatological conditions in intensive care: a secondary analysis of the intensive care national audit & research centre (ICNARC) case mix programme database," Critical Care, vol. 12, no. 1, p. S1, 2008, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2607109/.

[2] M. Badia, L. Serviá, J. M. Casanova, N. Montserrat, J. Vilanova, and E. Víctor, "Classification of dermatological disorders in critical care patients: a prospective observational study," Journal of Critical Care, vol. 28, p. 220, 2013.

[3] P. Agrawal, J. V. Peter, and R. George, "Dermatological manifestations and relationship to outcomes of patients admitted to a medical intensive care unit: a study from a tertiary care hospital in India," Postgraduate Medical Journal, vol. 89, no. 1055, pp. 501–507, 2013.

[4] T. Kugai and H. Nakagawa, "Evolution of purpura fulminans," New England Journal of Medicine, vol. 376, no. 22, p. 2182, 2017.

[5] J. Pedraz, Y. Delgado-Jiménez, S. Pérez-Gala, S. Nam-Cha, J. Fernández-Herrera, and A. García-Diez, "Cutaneous expression of systemic candidiasis," Clinical and Experimental Dermatology, vol. 34, no. 1, pp. 106–110, 2009.

[6] M. M. Campos-Fernández, S. Ponce-de-León-Rosales, C. Archer-Dubon, and R. Orozco-Topepe, "Incidence and risk factors for cutaneous adverse drug reactions in an intensive care unit," Revista de Investigación Clínica, vol. 57, pp. 770–774, 2005.

[7] M. O. Visscher, C. C. White, J. M. Jones, T. Cahill, D. C. Jones, and B. S. Pan, "Face masks for noninvasive ventilation: fit, excess skin hydration, and pressure ulcers," Respiratory Care, vol. 60, no. 11, pp. 1536–1547, 2015, http://rc.rcjournal.com/content/60/11/1536/tab-pdf.

[8] M. Badia, J. Trujillano, L. Serviá, J. March, and A. Rodríguez-Pozo, "Skin lesions after intensive care procedures: results of a prospective study," Journal of Critical Care, vol. 23, no. 4, pp. 525–531, 2008.

[9] E. H. De Laat, P. Pickkers, L. Schoonhoven, A. L. Verbeek, T. Feuth, and T. Van Amercberg, "Guideline implementation results in a decrease of pressure ulcer incidence in critically ill patients.\" Critical Care Medicine, vol. 35, no. 3, pp. 815–820, 2007.

[10] C. Spampinato and D. Leonardi, "Candida infections, causes, targets, and resistance mechanisms: traditional and alternative antifungal agents," BioMed Research International, vol. 2013, p. L1-L3, 2013, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3708393/pdf/BMR2013-204237.pdf, Article ID 204237.

[11] M. Badia, J. Trujillano, E. Gascó, J. M. Casanova, M. Alvarez, and M. León, "Skin lesions in the ICU," Intensive Care Medicine, vol. 25, no. 11, pp. 1271–1276, 1999.

[12] K. Curry, M. Kutash, T. Chambers, A. Evans, M. Holt, and S. Purcell, "A prospective, study of characteristics associated with skin failure in critically ill adults," Ostomy Wound Manage, vol. 58, pp. 40–43, 2012.

[13] S. Bourcier, J. Joffre, V. Dubée et al., "Marked regional endothelial dysfunction in mottled skin area in patients with severe infections," Critical Care, vol. 21, no. 1, p. 155, 2017, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5481873/pdf/13054_2017_Article_1742.pdf.

[14] J. H. Sillevis Smitt, J. B. M. van Woensel, and A. P. Bos, "Skin lesions in children admitted to the paediatric intensive care unit: an observational study," European Journal of Pediatrics, vol. 170, no. 10, pp. 1263–1265, 2011, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3175034/pdf/431_2011_Article_1436.pdf.

[15] S. R. Pektaş and A. K. Demir, "Prospective analysis of skin findings in surgical critically ill patients intensive care unit," Indian Journal of Dermatology, vol. 62, pp. 297–303, 2017, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5448265/.

[16] M. G. Dunnill, S. E. Handfield-Jones, D. Treacher, and D. H. McGibbon, "Dermatology in the intensive care unit," The British Journal of Dermatology, vol. 132, no. 2, pp. 226–235, 1995.

[17] G. Awall and T. Kaur, "Dermatological manifestations in the ICU," Journal of the Egyptian Women’s Dermatologic Society, vol. 15, no. 2, pp. 94–99, 2018.

[18] M. P. Agrawal and V. Sharma, "Purpura fulminans caused by meningococcemia," Canadian Medical Association Journal, vol. 182, p. E18, 2010, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2802628/pdf/182e018.pdf.

[19] P. Chang, Y. Sanchez, and R. Rodriguez Pellecerc, "Dermatoses found in an intensive care unit," Dermatologia CMQ, vol. 12, pp. 165–171, 2014, https://www.medigraphic.com/pdfs/cosmetica/dcm-2014/dcm1433.pdf.

[20] G. P. Prashanth and V. V. Pai, "A retrospective cohort study of dermatological problems observed in paediatric intensive care unit," Journal of the European Academy of Dermatology and Venereology, vol. 26, no. 9, pp. 1105–1108, 2012.

[21] M. Badia, J. Trujillano, J. M. Casanova, R. Egidio, M. Vallverdú, and M. León, "Fatal pemphigus vulgaris. acute skin failure concept," Medicina Intensiva, vol. 18, pp. 51–52, 1994.

[22] S. Kartan, Y. V. Shi, A. K. Clark, and L. S. Chan, "Paraneoplastic pemphigus and autoimmune blistering diseases associated with neoplasms: characteristics, diagnosis, associated neoplasms, proposed pathogenesis, treatment," American Journal of Clinical Dermatology, vol. 18, no. 1, pp. 105–126, 2017.

[23] M. Cholera and N. Chainani-Wu, "Management of pemphigus vulgaris," Advances in Therapy, vol. 33, no. 6, pp. 910–938, 2016, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4920839/pdf/12325_2016_Article_343.pdf.

[24] F. Capon, "IL36RN mutations in generalized pustular psoriasis: just the tip of the iceberg?" Journal of Investigative Dermatology, vol. 133, no. 11, pp. 2503–2504, 2013.

[25] K. Sugira, "The genetic background of generalized pustular psoriasis: IL36RN mutations and CARD14 gain-of-function variants," Journal of Dermatological Science, vol. 74, no. 3, pp. 187–192, 2014.

[26] C. Leonard, R. Matheson, C. Zachariaie et al., "Anti-interleukin-17 monoclonal antibody ixekizumab in chronic plaque psoriasis," New England Journal of Medicine, vol. 366, no. 13, pp. 1190–1199, 2012, https://www.nejm.org/doi/pdf/10.1056/NEJMoa1109997?articleTools=true.

[27] S. K. Mahil, F. Capon, and J. N. Barker, "Genetics of psoriasis," Dermatologic Clinics, vol. 33, no. 1, pp. 1–11, 2015.
[28] C. Okoduwu, W. Lambert, R. Schwartz et al., “Erythroderma: review of a potentially life-threatening dermatosis,” *Indian Journal of Dermatology*, vol. 54, no. 1, pp. 1–6, 2009, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2800861/.

[29] A. Cesar, M. Cruz, A. Mota, and F. Azevedo, “Erythroderma. a clinical and etiological study of 103 patients,” *Journal of Dermatology Case Reports*, vol. 31, pp. 1–9, 2016, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4844528/pdf/jdcr-10-001.pdf.

[30] N. Mistry, A. Gupta, A. Alavi, and R. G. Sibbald, "A review of the diagnosis and management of erythroderma (generalized red skin),” *Advances in Skin & Wound Care*, vol. 28, no. 5, pp. 228–236, 2015.

[31] M. F. Struck, P. Hilbert, M. Mockenhaupt, B. Reichelt, and M. Steen, "Severe cutaneous adverse reactions: emergency approach to non-burn epidermolytic syndromes,” *Intensive Care Medicine*, vol. 36, no. 1, pp. 22–32, 2010.

[32] R. A. Schwartz, P. H. McDonough, and B. W. Lee, "Toxic epidermal necrolysis. part I. introductions, history, classification, clinical features, systemic manifestations, aetiology, and immunopathogenesis,” *Journal of the American Academy of Dermatology*, vol. 69, pp. 173e1–173e13, 2013.

[33] R. A. Schwartz, P. H. McDonough, B. W. Lee, and M. Lohitnavy, "Toxic epidermal necrolysis. Part II. Prognosis, sequelae, diagnosis, differential diagnosis, prevention, and treatment,” *Journal of the American Academy of Dermatology*, vol. 69, pp. 187.e1–187.e16, 2013.

[34] W. Tangamornsuksan, N. Chaiyakunapruk, R. Somkrua, R. A. Schwartz, P. H. McDonough, B. W. Lee, and M. Lohitnavy, "Cutaneous drug reactions in inpatients,” *Clinical Reviews in Allergy & Immunology*, vol. 28, no. 3, pp. 293–305, 2010.

[35] H. Watanabe, "Recent advances in drug-induced hypersensitivity syndrome/drug reaction with eosinophilia and systemic symptoms,” *Journal of Immunology Research*, vol. 18, 2018 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5878892/pdf/JIR2018-5163129.pdf, Article ID 5163129.

[36] C. Bigham and A. Elsey, "Cutaneous drug reactions in intensive care,” *British Journal of Hospital Medicine*, vol. 74, no. 6, pp. 340–346, 2013.

[37] R. Wolf and B. Davidovici, "Severe cutaneous adverse drug reactions: who should treat, where and how?: facts and controversies,” *Clinics in Dermatology*, vol. 28, no. 3, pp. 344–348, 2010.

[38] J. P. Burnham and M. H. Kollef, "Understanding toxic shock syndrome,” *Intensive Care Medicine*, vol. 41, no. 9, pp. 1707–1710, 2015.

[39] M. Gottlieb, B. Long, and A. Koyfman, "The evaluation and management of toxic shock syndrome in the emergency department: a review of the literature,” *The Journal of Emergency Medicine*, vol. 54, no. 6, pp. 807–814, 2018.

[40] V. Lamand, O. Dauwalder, A. Tristan et al., "Epidemiological data of staphylococcal scalded skin syndrome from France from 1997 to 2007 and microbiological characteristics of *Staphylococcus aureus* associated strains,” *Clinical Microbiology and Infection*, vol. 18, no. 12, pp. E514–E521, 2012.

[41] M. Z. Hadler and R. A. Schwartz, "Staphylococcal scalded skin syndrome: diagnosis and management in children and adults,” *Journal of the European Academy of Dermatology and Venerology*, vol. 28, pp. 1418–1423, 2014.

[42] A. Endo, A. Shiraiishi, J. Aiboshi, Y. Hayashi, and Y. Otomo, "A case of purpura fulminans caused by Hemophilus influenzae complicated by reversible cardiomyopathy,” *Journal of Intensive Care*, vol. 2, pp. 2–13, 2014, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4267704/pdf/40560_2013_Article_27.pdf.

[43] E. F. Alvarez, K. E. Olarte, and M. S. Ramesh, "Purpura fulminans secondary to Streptococcus pneumoniae meningitis,” *Case Reports in Infectious Diseases*, vol. 2012, 2012 https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3336247/pdf/CRIMID.2012-508503.pdf, Article ID 508503.

[44] Ch. Dalugama and I. B. Gawarammana, "Rare presentation of rickettsial infection as purpura fulminans: a case report,” *Journal of Medical Case Reports*, vol. 12, p. 145, 2018, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3970498/pdf/13256_2018_Article_1672.pdf.

[45] O. Join-Lambert, H. Lecuyer, F. Miller et al., "Meningococcal interaction to microvasculature triggers the tissular lesions of purpura fulminans,” *The Journal of Infectious Diseases*, vol. 208, no. 10, pp. 1590–1597, 2013.

[46] K. Sethi, J. Buckley, and J. de Wolff, "Splinter haemorrhages, Osler’s nodes, Janeway lesions and Roth spots: the peripheral stigmata of endocarditis,” *British Journal of Hospital Medicine*, vol. 74, no. 9, pp. c139–c142, 2013.

[47] S. U. Nigwekar, "Calciniphylaxis,” *Current Opinion in Nephrology and Hypertension*, vol. 26, no. 4, pp. 276–281, 2017.

[48] J. A. Garcia-Lozano, J. Ocampo-Candiani, S. A. Martinez-Cabiales, and V. Garza-Rodriguez, "An update on calciphylaxis,” *American Journal of Clinical Dermatology*, vol. 19, no. 4, pp. 599–608, 2018.

[49] J. C. Jennette, R. J. Falk, P. A. Bacon et al., “2012 revised international chapel hill consensus conference nomenclature of vasculitides,” *Arthritis & Rheumatism*, vol. 65, no. 1, pp. 1–11, 2012, https://onlinelibrary.wiley.com/doi/epdf/10.1002/art.37715.

[50] S. Alberti-Violetti, E. Berti, and A. V. Marzano, "Cutaneous and systemic vasculitides in dermatology: a histological perspective,” *Giornale Italiano di Dermatologia e Venereologia*, vol. 153, pp. 185–193, 2018.

[51] M. R. Goeser, V. Lanioux, and D. A. Wetter, "A practical approach to the diagnosis, evaluation, and management of cutaneous small-vessel vasculitis,” *American Journal of Clinical Dermatology*, vol. 15, no. 4, pp. 299–306, 2014.

[52] A. V. Marzano, M. G. Raimondo, E. Berti, P. L. Meroni, and F. Ingegnoli, "Cutaneous manifestations of ANCA-associated small vessels vasculitis,” *Clinical Reviews in Allergy & Immunology*, vol. 53, no. 3, pp. 428–438, 2017.

[53] E. Shavit, A. Alavi, and R. G. Sibbald, "Vasculitis-what do we have to know? a review of literature,” *The International Journal of Lower Extremity Wounds*, vol. 17, no. 4, pp. 218–226, 2018.

[54] F. Chasset and C. Francès, "Cutaneous manifestations of medium- and large-vessel vasculitis,” *Clinical Reviews in Allergy & Immunology*, vol. 53, no. 3, pp. 452–468, 2017.

[55] J. N. Stannard and J. M. Kahlenberg, "Cutaneous lupus erythematosus,” *Current Opinion in Rheumatology*, vol. 28, no. 5, pp. 453–459, 2016, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4695280/pdf/nihms795478.pdf.

[56] P. Sena, A. Gianatti, and D. Gambini, "Dermatomyositis: clinico-pathological correlations,” *Giornale Italiano di Dermatologia e Venereologia: OrganUfficiale, Societa Italiana di Dermatologia e Sifilografia*, vol. 153, no. 2, pp. 256–264, 2018.

[57] J. Damaoiseaux, J.-B. Vulsteke, C.-W. Tseng et al., “Autoantibodies in idiopathic inflammatory myopathies: clinical associations and laboratory evaluation by mono- and multispecific immunoassays,” *Autoimmunity Reviews*, vol. 18, no. 3, pp. 293–305, 2019.
K. Mariampillai, B. Granger, D. Amelin et al., "Development of a new classification system for idiopathic inflammatory myopathies based on clinical manifestations and myositis-specific autoantibodies," *JAMA Neurology*, vol. 75, no. 12, pp. 1528–1537, 2018.

J. Martinez-Barrio, L. Valor, and F. J. López-Longo, "Facts and controversies in mixed connective tissue disease," *Medicina Clinica (English Edition)*, vol. 150, no. 1, pp. 26–32, 2018.

N. C. Ciang, N. Pereira, and D. A. Isenberg, "Mixed connective tissue disease-enigma variations?" *Rheumatology (Oxford)*, vol. 56, pp. 326–333, 2017.

M. Fisher, T. William, and J. Wohlrab, "Skin diseases in intensive care medicine," *Journal der Deutschen Dermatologischen Gesellschaft*, vol. 7, pp. 108–115, 2009.

M. G. Kalra, K. E. Higgins, and B. S. Kinney, "Intertrigo and secondary skin infections," *American Family Physician*, vol. 89, no. 7, pp. 569–573, 2014, https://www.aafp.org/afp/2014/0401/p569.pdf.

A. V. Marzano, A. Borghi, and M. Cugno, "Adverse drug reactions and organ damage: the skin," *European Journal of Internal Medicine*, vol. 28, pp. 17–24, 2016.

F. Chaves, J. Garnacho-Montero, J. L. Del Pozo et al., "Diagnosis and treatment of catheter-related bloodstream infection: clinical guidelines of the Spanish society of infectious diseases and clinical microbiology and (SEIMC) and the Spanish society of Spanish society of intensive and critical care medicine and coronary units (SEMICYUC)," *Medicina Intensiva*, vol. 42, no. 1, pp. 5–36, 2018, http://www.medintensiva.org/es-pdf-S021056911730284X.pdf.

A. V. Wong, N. Arora, O. Olusanya et al., "Insertion rates and complications of central lines in the UK population: a pilot study," *Journal of the Intensive Care Society*, vol. 19, no. 1, pp. 19–25, 2018, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5810875/pdf/10.1177_1751143717722914.pdf.

J. M. Smit, R. Raadsen, M. J. Blans, M. Petjak, P. M. Van de Ven, and P. R. Tuinman, "Bedside ultrasound to detect central venous catheter misplacement and associated iatrogenic complications: a systematic review and meta-analysis," *Critical Care*, vol. 22, p. 65, 2018, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5851097/pdf/13054_2018_Article_1989.pdf.

A. N. Weitz, C. T. Lauren, J. A. Weiser et al., "Chlorhexidine gluconate-impregnated central access catheter dressings as a cause of erosive contact dermatitis," *JAMA Dermatology*, vol. 149, no. 2, pp. 195–199, 2013.

L. E. Edsberg, D. Langemo, M. M. Baharestani, M. E. Posthauer, and M. Goldberg, "Unavoidable pressure injury," *Journal of Wound, Ostomy and Continence Nursing*, vol. 41, no. 4, pp. 313–334, 2014.

N. Tayyib and F. Coyer, "Effectiveness of pressure ulcer prevention strategies for adult patients in intensive care units: a systematic review," *Worldviews on Evidence-Based Nursing*, vol. 13, no. 6, pp. 432–444, 2016.

M. W. Dünser, A. J. Mayr, A. Tür et al., "Ischemic skin lesions as a complication of continuous vasopressin infusion in catecholamine-resistant vasodilatory shock: incidence and risk factors∗," *Critical Care Medicine*, vol. 31, no. 5, pp. 1394–1398, 2003.

N. Ruffin, C. V. Vasa, S. Breakstone, and W. Axman, "Symmetrical peripheral gangrene of bilateral feet and unilateral hand after administration of vasopressors during septic shock," *BMJ Case Reports*, vol. 2018, Article ID bcr2017223602, 2018.