Exploration of CYP21A2 and CYP17A1 polymorphisms and preeclampsia risk among Chinese Han population: a large-scale case-control study based on 5021 subjects

Bo Hou 1†, Xuewen Jia 1,2†, Ziwen Deng 3,4, Xin Liu 5, Huitang Liu 6, Haichu Yu 1* and Shiguo Liu 3,4*

Abstract

Background: Several genome-wide association studies have identified single-nucleotide polymorphisms (SNPs), such as rs4409766, rs1004467, and rs3824755 in CYP17A1 and rs2021783 in CYP21A2, as new hypertension susceptibility genetic variants in the Chinese population. This study aimed to look into the relationship between preeclampsia (PE) and these SNPs in Chinese Han women.

Methods: Overall, 5021 unrelated pregnant women were recruited, including 2002 patients with PE and 3019 normal healthy controls. The real-time PCR (TaqMan) method was applied to genotype these four polymorphisms.

Results: A statistically obvious difference in the allelic frequencies was observed in CYP21A2 rs2021783 between cases and controls ($\chi^2 = 7.201$, $P_c = 0.028$ by allele), and the T allele was associated with the occurrence and development of PE (OR = 1.151, 95% CI 1.039–1.275). We also found a significant association between rs2021783 and the development of early-onset PE ($P_c = 0.008$ by genotype, $P_c = 0.004$ by allele). For rs1004467 and rs3824755, the distribution of allelic frequencies differed markedly between mild PE and control groups ($\chi^2 = 6.843$, $P_c = 0.036$; $\chi^2 = 6.869$, $P_c = 0.036$), and patients with the TT genotype of rs1004467 were less easy to develop mild PE than were those carrying the CT or CC genotype ($\chi^2 = 7.002$, $P_c = 0.032$, OR = 1.306, 95% CI 1.071–1.593). The GG genotype of rs3824755 appeared to have a protective effect on the occurrence of mild PE (OR = 0.766, 95% CI 0.629–0.934).

Conclusions: CYP21A2 rs2021783 appears to be closely related to PE susceptibility, and CYP17A1 rs1004467 and rs3824755 seem to be closely associated with mild PE in Han women.

Keywords: Preeclampsia, GWAS, CYP17A1, CYP21A2, Susceptibility

* Correspondence: haichuyu@126.com; liushiguo2002@126.com
† Bo Hou and Xuewen Jia contributed equally to this work.
1 Department of Cardiology, The Affiliated Hospital of Qingdao University, Qingdao, China
3 Medical Genetic Department, The Affiliated Hospital of Qingdao University, Qingdao, China

Full list of author information is available at the end of the article

© The Author(s). 2020 Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.
Background

Preeclampsia (PE) is a special complication of pregnancy that is characterized by de novo proteinuria and hypertension after 20 gestational weeks. It affects 2 to 8% of all pregnant women worldwide and is the leading cause of maternal and fetal morbidity and mortality [1, 2], especially in developing countries. Furthermore, women with PE are more likely to develop cardiovascular disease (CVD), hypertension, and chronic kidney disease in later life [3, 4]. As a special type of hypertension, PE is a complex multifactor disease, which is the result of the interaction between genetic factors and environmental factors. Therefore, factors that affect blood pressure may be one of the primary mechanisms underlying PE. Although many environmental risk factors are associated with the formation of hypertension, including higher sodium intake, smoking, excessive alcohol consumption, obesity, activation of the sympathetic nervous system, and endothelial dysfunction [5], genetic factors also play an important role in the maintenance of hypertension [6]. Furthermore, multiple studies have shown that many candidate hypertension genes [7–9], such as STOX1, IL-1, IL6, and COMT, were also shown to be linked to PE [10–13].

Recent genome-wide association studies (GWAS) identified many genetic variants or chromosomal regions that are associated with hypertension, such as ATP2B1, CYP17A1, CYP1A2, SH2B3, CACNB2, TGFβ2, MTHFR, CYP11B2, and ULK4 [14–17]. Furthermore, ATP2B1, MTHFR, and CYP11B2 have been validated to have associations with the risk of PE in different ethnic groups [18–20]. Two large GWAS meta-analyses suggested that rs1004467 (P = 1.28 × 10^-10) and rs3824755 (P = 1.21 × 10^-6) in CYP17A1 were significantly associated with the risk of hypertension [14, 16]. Subsequently, Lu et al. found that rs4409766 had the strongest correlation with CYP17A1 (P = 7.33 × 10^-13) and CYP21A2 (P = 3.53 × 10^{-11}) in a large-scale hypertension GWAS, of which 80, 962 subjects were Chinese Han nationality [21].

CYP17A1 encodes the cytochrome P450 protein, which has 17-alpha-hydroxylase and 17,20-lyase activities and is one of the key enzymes in the synthesis of human steroid hormones. It had been investigated to play a role in multiple complex diseases, such as cardiovascular disease [22], hypertension [14], and fetal growth restriction [23]. CYP21A2 encodes 21-hydroxylase enzyme, and mutations in this gene may lead to congenital adrenal hyperplasia, which is characterized by hypertension, hypokalemia, and sexual infantilism [24]. Because they are candidate genes for blood pressure and play crucial roles in steroid hormone biosynthesis, thus having a further effect on hypertension, we designed and conducted this study to explore the relationship between genetic variations in CYP17A1 (rs4409766, rs1004467, and rs3824755) and CYP21A2 (rs2021783) and the sensitivity of Chinese Han women to PE.

Material and methods

Subjects

A total of 5021 subjects, including 2002 PE patients (mean age 30.09 ± 5.65 years) and 3019 controls (mean age 30.31 ± 4.01 years), were recruited from the Affiliated Hospital of Qingdao University, Linyi People’s Hospital, Liaocheng People’s Hospital, Yantai Yuhuangding Hospital, Heze Municipal Hospital, Binzhou Medical University Hospital, and the Maternal and Child Health Care of Zaozhuang between January 2012 and May 2016. All women had singleton pregnancies and were of Chinese Han ethnicity. The inclusion criteria for cases were established according to ACOG (American College of Obstetricians and Gynecologists, 2013), defined as hypertension (systolic blood pressure ≥ 140 mmHg or diastolic blood pressure ≥ 90 mmHg on two or more occasions at least 4 h apart) with proteinuria (0.3 g/24 h, 0.3 mg/dl, or 1+ by dipstick, without renal disease or infection) after 20 gestational weeks [25]. Exclusion criteria for both groups included chronic hypertension, a history of renal and endocrinal diseases, multifetal pregnancy, and autoimmune disorder. The control groups included normal healthy pregnant women without any pregnancy disorder or fetal disorders. All PE subjects were divided into the mild PE group (n = 454) and severe PE group (n = 1548). Severe PE was diagnosed if any following symptoms appeared in the case subjects: blood pressure ≥ 160/110 mmHg or progressive renal insufficiency (proteinuria ≥ 5 g/24 h, or cr > 1.1 mg/dl), new-onset visual or cerebral disturbances, impaired liver functions, and pulmonary edema [25]. Additionally, we divided the cases into early-onset PE (n = 937, before 34 weeks of gestation) and late-onset PE (n = 1065, after 34 weeks of gestation).

The questionnaire provided clinical data, including age, history of pregnancy and childbirth, blood pressure, gestational age, clinically relevant symptoms, and some laboratory examinations. All participants were provided with an explanation of the research and then gave written informed consent. Our study was approved by the ethics committee of the Affiliated Hospital of Qingdao University.

Genotyping

Qiagen DNA extraction kit (Qiagen, Hilden, Germany) was applied to extract genomic DNA from 2 ml peripheral venous anticoagulant blood, and then stored at −80 °C. SNP genotyping was determined by the TaqMan allele recognition real-time PCR. The probes and primers were designed and synthesized by Applied Biosystems by Life Technologies (ABI, NY, USA). The sequence of primers was provided in https://www.mysequenom.com/Tools. All polymerase chain reaction (PCR) amplification systems (thermocycler, C1000TM) were conducted in a total of 25 μl, containing DNase-free water, 11.25 μl of DNA, and 12.5 μl of 2x PCR Master Mix and 1.25 μl of 20x SNP Genotyping Assay. The cycling protocol was included in
an initial denaturation at 95 °C for 3 min, and then carried out 45 cycles of 95 °C for 15 s and 60 °C for 1 min. The fluorescent signals of VIC/FAM-labeled probes were detected in each cycle, and different genotypes were detected applying Bio-Rad CFX manager 3.0 software. We also selected 100 samples of each genetic variant to verify the genotype by Sanger sequencing.

**Statistical analysis**

All data were analyzed applying the software package SPSS 22.0 (SPSS Inc., Chicago, IL, USA). Clinical data and demographic were performed using Student’s *t* test or the chi-square test and were described by the mean ± standard error (SE) or percentage. A *P* value < 0.05 was considered statistically significant. Each control group polymorphism was tested for the Hardy-Weinberg equilibrium using the goodness-of-fit $\chi^2$ test. The distributions of genotypic and allelic frequencies between the case and control groups were analyzed by Pearson’s $\chi^2$ test. Ninety-five percent confidence intervals (CIs) and odds ratios (ORs) were applied to express the risk between the groups, and *P* < 0.05 was considered statistically significant; however, for four SNPs, a level of *P* < 0.0125 or *Pc* < 0.05 was considered significant when using Bonferroni’s correction. The genotypic-phenotype was analyzed by the one-way analysis of variance. The power analysis was calculated using the program Power and Sample Size Calculations (PS, version 3.1.2).

**Results**

**Demographic and clinical characteristics**

In Table 1, the demographic and clinical features of the two groups were shown. There were no obvious differences in the age, age of menarche, or number of abortions between the control group and the case group (*P* > 0.05). However, compared with normal controls, PE patients were more likely to have earlier gestational weeks at admission and gestational weeks at delivery, lower fetal birth weight, lower cholesterol, higher blood pressure, higher triglyceride, higher creatinine, elevated white blood cells, and higher blood urea nitrogen (*P* < 0.001).

**Genetic analysis**

The genotypic distributions of the following SNPs in the control groups were consistent with the Hardy-Weinberg equilibrium (for rs4409766, $\chi^2 = 0.091$, *P* = 0.763; for rs1004467, $\chi^2 = 1.203$, *P* = 0.272; for rs3824755, $\chi^2 = 0.206$, *P* = 0.886; for rs2021783, $\chi^2 = 0.040$, *P* = 0.841).

The genotypic and allelic frequencies of CYP17A1 (rs4409766, rs1004467, rs3824755) and CYP21A2 (rs2021783) polymorphisms are summarized in Table 2.

There was an obvious difference in the allele frequencies of CYP21A2 rs2021783 between the PE patients and control group ($\chi^2 = 7.201$, *P* = 0.007, *Pc* = 0.028). The T allele was related to the occurrence of PE (OR = 1.151, 95% CI 1.039–1.275). In the genotype (CC vs. CT vs. TT, *P* = 0.017; however, after Bonferroni’s correction, *Pc* = 0.068), the difference between the groups showed no statistical significance. In contrast, there were no obvious differences in the genotypic and allelic frequencies of rs4409766, rs1004467, and rs3824755 in CYP17A1 between the groups (for rs4409766, $\chi^2 = 2.518$, *P* = 0.284 by genotype, and $\chi^2 = 1.345$, *P* = 0.246 by allele; for rs1004467, $\chi^2 = 1.424$, *P* = 0.491 by genotype, and $\chi^2 = 1.414$, *P* = 0.234 by allele; and for rs3824755, $\chi^2 = 2.392$, *P* = 0.302 by genotype, and $\chi^2 = 2.386$, *P* = 0.122 by allele).

For rs2021783 in CYP21A2, Table 3 displays that there was no difference between the patients with mild/severe PE and control groups (*Pc* > 0.05). However, for rs1004467, the distribution of allelic frequencies differed significantly between the mild PE and control groups ($\chi^2 = 6.843$, *P* = 0.005).

### Table 1: Demographic and clinical characteristics of the case and control groups

|                          | Case          | Control        | *t*  | *P*  |
|--------------------------|---------------|----------------|------|------|
| Age (years)              | 30.09 ± 5.65  | 30.31 ± 4.01   | −1.478 | 0.140 |
| Age of menarche (years)  | 13.98 ± 1.19  | 14.02 ± 1.20   | −0.984 | 0.325 |
| Number of abortion       | 0.62 ± 0.95   | 0.62 ± 0.86    | −0.102 | 0.919 |
| Gestational age at admission (weeks) | 35.56 ± 3.48 | 39.16 ± 1.35   | −43.19 | 0.000* |
| Gestational age at delivery (weeks) | 36.21 ± 3.09 | 39.36 ± 1.22   | −41.25 | 0.000* |
| Fetal birth weight (g)   | 2600 ± 934    | 3387 ± 673     | −30.66 | 0.000* |
| Systolic blood pressure (mmHg) | 160.05 ± 18.51 | 115.88 ± 10.56 | 95.73 | 0.000* |
| Diastolic blood pressure (mmHg) | 104.12 ± 13.63 | 73.47 ± 7.32   | 91.24 | 0.000* |
| White blood cell (WBC)   | 9.61 ± 2.84   | 8.96 ± 2.41    | 8.26  | 0.000* |
| Triglyceride             | 3.88 ± 1.83   | 2.94 ± 1.25    | 11.85 | 0.000* |
| Cholesterol (CHO)        | 3.89 ± 1.87   | 6.11 ± 1.23    | −27.88 | 0.000* |
| Blood urea nitrogen (BUN) | 4.67 ± 1.98   | 3.18 ± 0.98    | 28.80 | 0.000* |
| Creatinine (CREA)        | 65.26 ± 27.60 | 51.46 ± 19.66  | 17.45 | 0.000* |

*Significant difference vs. controls
The allele T may thus be a risk factor for mild PE (OR = 1.219, 95% CI 1.051–1.413), and the TT genotype was associated with the risk of developing mild PE compared to those subjects who had a CT or CC genotype ($\chi^2 = 7.002, P = 0.008, P_c = 0.032$). Similarly, the allelic frequencies of rs3824755 differed significantly between the mild PE group and the control group ($\chi^2 = 6.869, P = 0.009, P_c = 0.036$). Further, the GG genotype appeared to be a protective effect on the occurrence of mild PE (OR = 0.766, 95% CI 0.629–0.934). For rs4409766, there was no difference in the genotypic and allelic frequencies between the mild/severe PE and control groups ($P > 0.05$).

We also found that rs2021783 was related to the risk of early-onset PE ($P_c = 0.008$ by genotype, $P_c = 0.004$ by allele). No obvious differences were found in the genotypic and allelic frequencies of CYP17A1 (rs4409766, rs1004467, and rs3824755) between the control group and early and advanced PE patients (Table 4).

**Analysis of genotype-phenotype relationship**

A laboratory-based examination of PE among several genotypes is presented in Table 5. For rs2021783, patients carrying the TT genotype had a lower menarche age (13.55 ± 1.31 vs. 14.01 ± 1.09, or vs. 14.00 ± 1.22; $P = 0.027$) than did those with the CT or CC genotype. Furthermore, the level of urea nitrogen was lower in patients with the CT genotype than in those carrying the CC genotype (4.49 ± 1.72 vs. 4.73 ± 2.00, $P = 0.016$). However, the gestational age, birth weight of children, number of abortions, systolic/diastolic blood pressure, triglyceride, total cholesterol, and creatinine showed no statistical differences among three groups (all, $P > 0.05$).

**Discussion**

A normal pregnancy is a condition of increased extracellular fluid, plasma volume, renal blood flow, and glomerular filtration rate. In contrast, PE is characterized by hypertension, proteinuria, higher vascular resistance, and decreased intravascular volume, cardiac output, and uteroplacental...
Despite unremitting efforts that have been undertaken for several decades, the precise pathophysiology or pathogenesis of this human pregnancy disorder remains elusive. However, increasing evidence suggests that gene-environment interactions maybe play important roles in the pathogenesis of PE [6]. Because PE is a special type of hypertension, we hypothesize that the genetic factors that affect blood pressure may be the primary cause for the incidence of PE.

GWAS had been demonstrated to be an effective way for identifying genetic variants in many common traits and diseases [27]. The Framingham Heart Study enrolled 9400 participants across three generations in Framingham, and the Wellcome Trust Case Control Consortium, which examined 2000 cases and a shared set of 3000 controls of white Europeans in Great Britain, reported the first two GWAS of hypertension but did not observe genetic loci that attained genome-wide significance [28, 29]. Then, a study in 34,433 subjects of European ancestry found eight blood pressure loci, including rs11191548 in CYP17A1, rs17367504 in MTHFR, rs12946454 in PLCD3, rs1378942 in CYP1A2, rs11998073 in FG5, rs653178 in SH2B3, rs1530440 in c10orf107, and rs16948048 in ZNF652 [15]. Since then, more genetic loci, such as ATP2B1, CYP17A1, CYP1A2, SH2B3, CACNB2, MTHFR, CYP11B2, and UILK4, were identified in a large-scale GWAS associated with blood pressure [14, 15, 30, 31].

Some genetic variants in CYP17A1 were associated with hypertension in different populations, such as rs11191548 which was found to be associated with both the She ethnic minority in China and the Chinese Han population [32, 33]. rs11191416 and rs6163 were also identified to have association with blood pressure in 4178 European ancestry participants [34]. Furthermore, rs1004467 in CYP17A1 is a common intronic variant that is associated with hypertension; it has been reported as a significant locus in the genome-wide meta-analysis and was verified in a Chinese Han cohort [15]. A study of mean arterial pressure and pulse pressure among 26,600 East Asians followed by a repeat study of 28,783 participants suggested that CYP17A1 rs3824755 was associated with blood pressure (P = 1.2 × 10^{-6}) [16]. The other two genetic loci of CYP21A2 rs2021783 and CYP17A1 rs4409766 were only found to be associated with blood pressure in a Chinese population [21], which indicated that these SNPs were genetic marker variants of hypertension in Chinese populations.

However, in addition to genetic factors, metabolic factors such as abnormal steroid metabolism play important roles in the occurrence of PE [35]. In normal pregnancy, the aldosterone levels are elevated to contribute to sodium
Aldosterone is reduced in PE patients [35, 36]. Additionally, PE patients had increased serum progesterone or androgen levels and decreased estrogen levels [37]. CYP17A1 and CYP21A2 play important roles in the metabolic pathways of steroid hormones. CYP17A1 encoded the cytochrome P450 proteins, which had 17-alpha-hydroxylase and 17,20-lyase activities, which was the key step in the biosynthesis of mineralocorticoids, glucocorticoids, and sex-steroid biosynthesis [38]. CYP21A2 encoded 21-hydroxylase enzyme and converted 17-hydroxyprogesterone into 11-deoxycortisol or progesterone to deoxycorticosterone [24]. Aldosterone affected blood pressure and the blood volume in the body by regulating the salt retained by the kidneys. Therefore, the mutations in CYP17A1 and CYP21A2 may lead to a loss in enzyme activity and then affect the synthesis of steroid hormones.

In our study, we investigated the associations between rs4409766, rs1004467, and rs3824755 in CYP17A1 and rs2021783 in CYP21A2 and PE; several GWAS had identified these as the genetic variants associated with hypertension in China [16, 21]. To our knowledge, this is the first study to study the relationship between patients with PE and normal pregnant women of CYP17A1 and CYP21A2 in Chinese Han women. In the present study, the C allele of rs2021783 indicated an increased risk in patients with PE compared to the control group. The CC genotype was more susceptible to experiencing severe PE or early-onset PE. The CT genotype had the effect of protecting women from severe PE or early-onset PE. We also found, for rs1004467, that the T allele was the risk allele for the occurrence of mild PE and that patients with the TT genotype had a 1.219-fold risk of developing the PE compared to patients with CT or CC genotype in Chinese Han women. For rs3824755, the frequency of the G allele was associated with a higher risk of mild PE patients, and the GG genotype was identified as a risk factor in mild PE. However, Lim et al. investigated the association between polymorphisms

| Table 4 Genotype and allele distributions in the early-onset/late-onset PE and control groups |
|---------------------------------------------------------------|
| Control | Early-onset PE | $\chi^2$ | P value | Pc | OR | 95% CI | Severe PE | $\chi^2$ | P value | Pc | OR | 95% CI |
|---------|---------------|--------|--------|-----|----|--------|---------|--------|--------|-----|----|--------|
| N       | 3019          | 937    |        |     |    |        | 1065   |        |       |     |    |        |
| rs4409766 |              |        |        |     |    |        |        |        |       |     |    |        |
| TT      | 1509          | 460    |        |     |    |        | 558    |        |       |     |    |        |
| CT      | 1233          | 405    |        |     |    |        | 421    |        |       |     |    |        |
| CC      | 277           | 72     | 2.896  | 0.235 |    | 86     | 2.195  | 0.334  |       |     |    |        |
| T       | 4251          | 1325   |        |     |    |        | 1537   |        |       |     |    |        |
| C       | 1787          | 549    | 0.062  | 0.803 |    | 1.015  | 0.905–1.137 | 593    | 2.226 | 0.136 | 1.087 | 0.974–1.213 |
| rs1004467 |              |        |        |     |    |        |        |        |       |     |    |        |
| TT      | 1199          | 372    |        |     |    |        | 451    |        |       |     |    |        |
| CT      | 1401          | 442    |        |     |    |        | 478    |        |       |     |    |        |
| CC      | 419           | 123    | 0.385  | 0.825 |    | 136    | 2.460  | 0.292  |       |     |    |        |
| T       | 3799          | 1186   |        |     |    |        | 1380   |        |       |     |    |        |
| C       | 2239          | 688    | 0.083  | 0.773 |    | 1.016  | 0.912–1.131 | 750    | 2.374 | 0.123 | 1.084 | 0.978–1.202 |
| rs3824755 |              |        |        |     |    |        |        |        |       |     |    |        |
| GG      | 1232          | 385    |        |     |    |        | 471    |        |       |     |    |        |
| GC      | 1396          | 445    |        |     |    |        | 463    |        |       |     |    |        |
| CC      | 391           | 107    | 1.587  | 0.452 |    | 131    | 3.789  | 0.150  |       |     |    |        |
| G       | 3860          | 1215   |        |     |    |        | 1405   |        |       |     |    |        |
| C       | 2178          | 659    | 0.511  | 0.475 |    | 0.961  | 0.863–1.071 | 725    | 2.843 | 0.092 | 1.093 | 0.986–1.213 |
| rs2021783 |              |        |        |     |    |        |        |        |       |     |    |        |
| TT      | 122           | 35     |        |     |    |        | 38     |        |       |     |    |        |
| CT      | 961           | 242    |        |     |    |        | 325    |        |       |     |    |        |
| CC      | 1936          | 660    | 12.986 | 0.002a | 0.008a | 702    | 1.272  | 0.529  |       |     |    |        |
| T       | 1205          | 312    |        |     |    |        | 401    |        |       |     |    |        |
| C       | 4833          | 1562   | 10.099 | 0.001a | 0.004a | 1.248  | 1.088–1.431 | 1729   | 1.274 | 0.259 | 1.138 | 0.948–1.219 |
| CT      | 961           | 242    |        |     |    |        | 325    |        |       |     |    |        |
| CC+TT   | 2058          | 695    | 12.183 | 0.000a | 0.000a | 0.746  | 0.632–0.880 | 740    | 0.631 | 0.427 | 1.063 | 0.914–1.237 |
| CC      | 1936          | 660    |        |     |    |        | 702    |        |       |     |    |        |
| CT+TT   | 1083          | 277    | 12.622 | 0.000a | 0.000a | 1.333  | 1.137–1.562 | 363    | 1.101 | 0.294 | 0.924 | 0.798–1.071 |

Pc P value corrected by Bonferroni’s method
*aSignificant difference
of -34T/C in the promoter in CYP17A1 and PE in 164 cases and 182 normal pregnancies in Korea but did not obtain significant results [39]. Additionally, Coto et al. found no relationship between CYP21A2 rs6471 polymorphism with PE in 250 cases and 250 controls from Asturias [40].

Although our study contained a sufficient number of samples, including 2002 PE patients and 3019 age-matched normal controls, and had sufficient statistical power for each SNP (rs1004467, 0.221; rs4409766, 0.214; rs3824755, 0.338) to draw conclusions, some limitations should be noted. All subjects were selected from Shandong Province of China, and an ethnic group may yield false positives due to unmeasured collinearity in the cohort. Thus, our findings should be expanded outside the Han ethnicity and be validated in different ethnicities and regions.

Although PE was generally considered a complex multifactorial disease resulting from genetic and environment risk factors, including dietary and lifestyle factors, we could not consider all of these factors. Additionally, whether other SNPs in CYP21A2 and CYP17A1 participate in the development of PE needs to be studied further.

Conclusion
CYP21A2 rs2021783 appears to be closely associated with PE susceptibility, and CYP17A1 rs1004467 and rs3824755 seem to be related to mild PE in Chinese Han women.

Table 5 Laboratory-based examination of PE among different genotypes

|                  | (1) TT | (2) CT | (3) CC | (1) vs. (2) | (2) vs. (3) | (1) + (2) vs. (3) |
|------------------|--------|--------|--------|-------------|-------------|------------------|
| P                | 0.894  | 0.817  | 0.962  | 0.644       | 0.679       | 0.917            |
| Age (years)      | 30.15 ± 6.13 | 29.99 ± 5.51 | 30.12 ± 5.50 | 0.012*      | 0.027*      | 0.027*           |
| Age of menarche (years) | 13.55 ± 1.31 | 14.01 ± 1.09 | 14.00 ± 1.22 | 0.981       | 0.984       | 0.952            |
| Number of abortion | 0.617 ± 0.90 | 0.615 ± 0.95 | 0.619 ± 0.95 | 0.700       | 0.963       | 0.768            |
| Gestational age at admission (weeks) | 35.62 ± 3.07 | 35.64 ± 3.65 | 35.49 ± 3.44 | 0.372       | 0.353       | 0.169            |
| Gestational age at delivery (weeks) | 36.12 ± 2.89 | 36.17 ± 3.32 | 36.24 ± 3.00 | 0.993       | 0.356       | 0.169            |
| Fetal birth weight (g) | 2603 ± 975 | 2616 ± 952 | 2593 ± 926 | 0.895       | 0.910       | 0.638            |
| Systolic blood pressure (mmHg) | 160.28 ± 17.38 | 159.80 ± 17.84 | 159.41 ± 17.84 | 0.854       | 0.830       | 0.662            |
| Diastolic blood pressure (mmHg) | 103.78 ± 12.58 | 103.62 ± 13.11 | 104.37 ± 13.90 | 0.533       | 0.927       | 0.718            |
| White blood cell (x10^9/l) | 9.56 ± 2.81 | 9.70 ± 2.99 | 9.72 ± 3.08 | 0.910       | 0.704       | 0.927            |
| Triglyceride (mmol/l) | 3.82 ± 1.87 | 3.88 ± 1.69 | 3.91 ± 1.95 | 0.929       | 0.822       | 0.738            |
| Total cholesterol (mmol/l) | 6.69 ± 1.70 | 6.71 ± 1.95 | 6.72 ± 1.92 | 0.980       | 0.960       | 0.862            |
| Urea nitrogen (mmol/l) | 4.54 ± 1.77 | 4.49 ± 1.72 | 4.73 ± 2.00 | 0.048*      | 0.832       | 0.016*           |
| Creatinine (μmol/l) | 61.69 ± 21.18 | 62.49 ± 25.48 | 65.39 ± 28.94 | 0.466       | 0.473       | 0.314            |

*Significant difference
Author details
1. Department of Cardiology, The Affiliated Hospital of Qingdao University, Qingdao, China. 2. Women's Health Research Institute, Boston, Massachusetts, USA. 3. Department of Obstetrics and Gynecology, University of California, San Francisco, California, USA. 4. Department of Medicine, University of California, San Francisco, California, USA. 5. Department of Laboratory Medicine, Zhongnan Hospital of Wuhan University, Wuhan, China.

Received: 27 December 2019 Accepted: 17 June 2020
Published online: 25 September 2020

References
1. Duley L. The global impact of pre-eclampsia and eclampsia. Semin Perinatol. 2009;33(3):130–7. https://doi.org/10.1053/j.semperi.2009.02.010 published Online First: Epub Date.
2. Umesawa M, Kobashi G. Epidemiology of hypertensive disorders in pregnancy: prevalence, risk factors, predictors and prognosis. Hypertens Res. 2017;40(3):213–20. https://doi.org/10.1038/hr.2016.126 published Online First: Epub Date.
3. Ahmed R, Dunford J, Mehran R, Robson S, Kunadian V. Pre-eclampsia and future cardiovascular risk among women a review. J Am Coll Cardiol. 2014;63(18):1815-22 doi: https://doi.org/10.1016/j.jacc.2014.02.529 [published Online First: Epub Date].
4. Amini M, Ramezani Tehrani F, Rahmati M, Behboudi-Gandevani S, Azizi F. Changes over time in blood pressure of women with preeclampsia compared to those with normotensive pregnancies: a 15-year population-based cohort study. Pregnancy Hypertens. 2019;17:94–9. https://doi.org/10.1016/j.preghy.2019.05.007 published Online First: Epub Date.
5. Whelton PK, He J, Appel LJ, et al. Primary prevention of hypertension - clinical and public health advisory from the National High Blood Pressure Education Program. JAMA. 2002;288(15):1882–8. https://doi.org/10.1001/jama.288.15.1882 published Online First: Epub Date.
6. Garcia EA, Newhouse S, Caulfield MJ, Munroe PB. Genes and hypertension. Nature. 2016;534(7606):332-9. https://doi.org/10.1038/nature19419 published Online First: Epub Date.
7. Allison SJ. Hypertension: IL-1 receptor-induced sodium reabsorption in hypertension. Nature reviews. Nephrology. 2013;12(5):212-22. https://doi.org/10.1038/nrneph.2016.1 published Online First: Epub Date.
8. Ittun NC, Miyaji K, Song YX, Ikeda S, Shimbo T, Muramatsu M. Association of the catechol-o-methyl transferase gene Val158Met polymorphism with blood pressure and prevalence of hypertension: interaction with dietary energy intake. Am J Hypertens. 2011;24(9):1022–6. https://doi.org/10.1038/ajh.2011.93 published Online First: Epub Date.
9. Singh S, McDonough CW, Gong Y, et al. Genome wide association study in Chinese identifies eight loci associated with blood pressure and hypertension. Nat Genet. 2009;41(6):677–87. https://doi.org/10.1038/ng.384 published Online First: Epub Date.
10. Newton-Chen C, Johnson T, Gavett V, et al. Genome-wide association study identifies eight loci associated with blood pressure. Nat Genet. 2009;41(6):666–7. https://doi.org/10.1038/ng.361 published Online First: Epub Date.
11. Mohajertehran F, Afshari JT, Rezaieyazdi Z, Ghomian N. Association of single nucleotide polymorphisms rs2681472 is associated with early-onset preeclampsia in northern Han Chinese women. Reprod Sci. 2014;21(11):1425–7. https://doi.org/10.1177/1044703013516498 published Online First: Epub Date.
12. Li J, Liu MC, Zong JB, et al. Genetic variations in IL1A and IL1RN are associated with the risk of preeclampsia in Chinese Han population. Sci Rep-Uk. 2015;5250. https://doi.org/10.1038/srep05250 published Online First: Epub Date.
13. Verdonk K, Visser W, van den Meiracker AH, Danser AHJ. The renin-angiotensin-aldosterone system in pre-eclampsia: the delicate balance between good and bad. Clin Sci. 2014;126(7-8):537–44. https://doi.org/10.1042/CS20130455 published Online First: Epub Date.
14. Bogacz A, Bartkowiak-Wlcekorek, J, Procyk D, et al. Analysis of the gene polymorphism of aldosterone synthase (CYP11B2) and atrial natriuretic peptide (ANP) in women with preeclampsia. Eur J Obstet Gyn R B 2016;197:11-15 doi: https://doi.org/10.1016/j.ejogrb.2016.11.012 published Online First: Epub Date.
15. Li X, Luo YL, Zhang QH, et al. Methyleneetahydrofibrate reduce gene C677T, A1298C polymorphisms and pre-eclampsia risk: a meta-analysis. Mol Biol Rep. 2014;41(8):5435–8. https://doi.org/10.1007/s11033-014-3415-z published Online First: Epub Date.
16. Kelly TN, Takeuchi F, Tabara Y, et al. Genome-wide association study meta-analysis reveals transnetnic replication of mean arterial and pulse pressure loci. Hypertension 2013;62(5):583-59 doi: https://doi.org/10.1161/Hypertensionaha.113.110148 published Online First: Epub Date.
17. Evangelou E, Wainer HR, Moxen-Ansorena D, et al. Genetic analysis of over 1 million people identifies 535 new loci associated with blood pressure traits. Nat Genet 2018;50(10):1412-25 https://doi.org/10.1038/s41588-018-0205-x published Online First: Epub Date.
18. Wan JP, Wang H, Li CZ, et al. The common single-nucleotide polymorphism rs2681472 is associated with early-onset preeclampsia in northern Han Chinese women. Reprod Sci. 2014;21(11):1425–7. https://doi.org/10.1177/1044703013516498 published Online First: Epub Date.
19. Liu C, Li H, Qi Q, et al. Common variants in or near FGF5, CYP17A1 and MTHFR are associated with blood pressure and/or hypertension in she ethnic minority of China. Atherosclerosis 2011;212(9):709-14 doi: https://doi.org/10.1016/j.atherosclerosis.2011.09.006 published Online First: Epub Date.
34. Morrison AC, Bis JC, Hwang SJ, et al. Sequence analysis of six blood pressure candidate regions in 4,178 individuals: the cohorts for heart and aging research in genomic epidemiology (CHARGE) targeted sequencing study. PLoS One. 2014;9(10):e109155. https://doi.org/10.1371/journal.pone.0109155 published Online First: Epub Date.

35. Verdonk K, Saleh L, Lankhorst S, et al. Association studies suggest a key role for endothelin-1 in the pathogenesis of preeclampsia and the accompanying renin-angiotensin-aldosterone system suppression. Hypertension. 2015;65(6):1316–U274. https://doi.org/10.1161/Hypertensionaha.115.05267 published Online First: Epub Date.

36. Escher G, Mohaupt M. Role of aldosterone availability in preeclampsia. Mol Asp Med 2007;28(2):245-254 doi: https://doi.org/10.1016/j.mam.2007.03.002 [published Online First: Epub Date].

37. Salas SP, Marshall G, Gutierrez BL, Rosso P. Time course of maternal plasma volume and hormonal changes in women with preeclampsia or fetal growth restriction. Hypertension. 2006;47(2):203–8. https://doi.org/10.1161/01.HYP.0000200042.64517.19 published Online First: Epub Date.

38. Auchus RJ. The genetics, pathophysiology, and management of human deficiencies of P450C17. Endocrinoi Metab Clin N Am. 2001;30(1):101–19 vii.

39. Lim JH, Kim SY, Kim DJ, et al. Genetic polymorphism of catechol-O-methyltransferase and cytochrome P450c17alpha in preeclampsia. Pharmacogenet Genomics 2010;20(10):605-10 doi: https://doi.org/10.1097/FPC.0b013e32833df033 [published Online First: Epub Date].

40. Coto E, Tavira B, Marin R, et al. Functional polymorphisms in the CYP3A4, CYP3A5, and CYP21A2 genes in the risk for hypertension in pregnancy. Biochem Biophys Res Commun 2010;397(3):576-9 doi: 10.1016/j.bbrc.2010.06.003 [published Online First: Epub Date].

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.