Factors associated with livebirth in couples undergoing their first in vitro fertilization cycle: An internally validated prediction model

Abstract

Objective: The aim of the study is to create a new model to predict successful outcome in assisted reproductive techniques.

Materials and Methods: A retrospective cohort study was conducted in tertiary fertility center between 2010 and 2017. Nulliparous women younger than 45 years-old undergoing in vitro fertilization/intracytoplasmic sperm injection (IVF/ICSI) for the first time were included; frozen embryo transfers, canceled induction cycles, freeze-all cycles were excluded. Two prediction models were built using multivariate logistic regression with a subset of the dataset and then were internally validated using bootstrapping methods.

Results: Four hundred eighty eight women were included with 136 (27.9%) live births. The basal model was built using variable age, antral follicle count (AFC), and basal luteinizing hormone (LH) levels. Age over 37 years [odds ratio (OR): 0.07, 95% confidence interval (CI): 0.00-0.36] and AFC below 5 (OR: 0.15, 95% CI: 0.02-0.53) was associated with poorer outcomes whereas an LH level above 6 mIU/mL (OR: 2.24, 95% CI: 1.27-3.94) was associated with better outcomes. Optimism adjusted area under the curve (AUC) of this model was 0.68 (95% CI: 0.62-0.74). Combined model in addition to basal model variables included the length of induction cycle, the endometrial thickness at the day of transfer, grade and count of the transferred embryo. Cycles lasting more than ten days (OR: 2.23, 95% CI: 1.17-4.42), an endometrial thickness greater than 9 mm (OR: 2.07, 95% CI: 1.00-4.53) were associated with better outcomes. Optimism adjusted AUC of this model was 0.76 (95% CI: 0.70-0.81). Calibration of both models was good according to Hosmer Lemeshow test (p=0.979 and p=0.848, respectively).

Conclusion: This internally validated prediction model has good calibration and can be used predicting outcomes in first time IVF/ICSI cycles with modest sensitivity.

Keywords: Prediction models, assisted reproductive techniques, live birth, in-vitro fertilization

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Introduction

Subfertile couple counseling is one of the most important parts of assisted reproductive technology (ART) treatment. Several prediction models in the literature provided a data-driven perspective to both clinicians and patients\(^{(1-3)}\). Live birth is the ultimate goal of ART treatment, and the most common and recognized models by Nelson and Lawlor\(^{(4)}\) and Templeton et al.\(^{(5)}\) used a live birth as the primary outcome. However, both models underestimated the live birth rate based on external validation studies\(^{(6)}\). Another external validation study concluded that a better calibration was achieved for both models after adjustments based on current trends of in vitro fertilization (IVF) success; however, the Templeton model underestimated and the Nelson model overestimated the fertilization (IVF) success; however, the Templeton model models after adjustments based on current trends of in vitro study concluded that a better calibration was achieved for both models. The study was approved by the Institutional Clinical Research Ethics Committee (date: April 25, 2016; number: 08-341-16).

Women under 45 years old with fresh embryo cycles were included. All included cycles underwent ICSI. The frozen embryo cycles, women with prior IVF/ICSI cycles, patients with secondary infertility, canceled cycles due to a nonviable sperm during testicular sperm extraction, cycles with >2 transferred embryos, and patients with donor sperm or egg, were excluded. Hospital records from patient files were used to create an anonymous dataset for internal validation. These records were searched manually by E.K. and A.G. Age, infertility duration, hysterosalpingography evaluation notes, body mass index (BMI), infertility indication, and ovarian reserve assessment at day 3 of menstrual cycle were used as patient demographics, and total gonadotropin dose, cycle duration, drugs used for ovarian induction, and sonographic assessments of the follicles and endometrium were used as cycle characteristics. Endometrial thickness, embryo quality, and embryo age were used as transfer characteristics.

After an initial assessment of patients with a detailed historical examination, semen analysis based on the World Health Organization criteria, ovarian reserve, and tubal patency assessment; ovarian stimulation was started during days 3 and 5 of the menstrual cycle. The starting dose was individualized based on patient age, ovarian reserve, and BMI. Further adjustment was also individualized based on the ovarian response assessment. The planned antagonist protocols (Cetrotide, Merck-Serono) were started after 5 days of gonadotropin usage or at least a 12 mm diameter of follicles were seen. Patients with a high risk of ovarian hyperstimulation syndrome were triggered with a dual trigger method or Gonadotrophin-releasing hormone (GnRH) agonists. Vaginal progesterone at 90 mg/day (Crinone 8% gel; Merck-Serono, Istanbul, Turkey) was used for luteal phase support from the day of embryo transfer to 12 weeks of gestational age. The ongoing pregnancy was defined as a pregnancy completed >20 weeks of gestational age. Antenatal follow-ups were organized based on the Ministry of Health guidelines.

Materials and Methods

This was a retrospective cohort study conducted in a single tertiary infertility clinic in the Department of Obstetrics and Gynecology at Ankara University. Dataset was gathered from patients evaluated between January 2010 and January 2017. The study was approved by the Institutional Clinical Research Ethics Committee (date: April 25, 2016; number: 08-341-16). Descriptive statistics of all variables used in the study were investigated. The distribution properties of variables were evaluated with the Shapiro-Wilk test and were assumed with a normal distribution feature if the p value was >0.05. Theoretical quantile-quantile graphs of parameters with normal distribution properties (Shapiro-Wilk test p>0.05) were created.
and the distribution assumption was visually tested. The variable presentation was made in the form of median value and interquartile range, and specific presentation types were not used for distribution assumption. T-test or Wilcoxon rank-sum test was used following the distribution assumption of the examined variable for binary group comparisons. Logistic regression analysis was used to create the prediction model. First, all examined parameters were modeled alone and relative probability ratios, confidence interval (CI), and p-values were found. A variable selection model was applied to create the multiparameter model. Step models are prone to produce biased or incompatible clinical reality models, thus all combinations of clinically important parameters or parameters that are important in the univariate regression analysis were tested. The Akaike criterion was used as an aid in parameter selection, and the accuracy and calibration of the model were tested in each step. The accuracy of the created models was tested with receiver operating characteristic curves. Model calibration was tested with the Hoslem-Lemeshow test and calibration curves. A certain part of dataset was used to create the model in the study (60%) and all dataset was included in the validation stage (60% + 40%). The internal validation of the model was done using 10,000 different datasets created using the bootstrapping method. The deviation corrected CI of the parameters used for internal validation were found and operating characteristic curves were created with corrected optimism. All statistical analysis R for Windows: Software language for statistical computing (Version 3.1.3) and packages of the same program “pROC,” “ModelGood,” “rms,” “caret,” “boot,” “ggplot2,” and “ROC632” were used. Unless otherwise stated, 0.05 was accepted as the statistically significant p-value limit.

Results

A total of 488 women who started controlled hyperstimulation for their first embryo transfer were included in the present study. The missing values other than basic patient characteristics were below 1% in the whole dataset. No imputation was applied to the dataset. Among 488 cycles, 136 (27.9%) resulted in an ongoing pregnancy. The model was based on the comparison of the main characteristics of 305 patients [live birth (number): 85, failed cycles (number): 220], which were presented in Table 1. Age, antral follicle count, day 3 serum luteinizing hormone (LH) level, gonadotropin induction duration, mature oocyte total count, fertilization rate, endometrial thickness on the embryo transfer day, and transferred embryo grade were significantly different between patients with and without live birth.

Selection of Parameters for Univariate Regression Analysis and Prediction Model

Patient demographics, cycle, and transfer day characteristics were put into the one-way regression analysis. All parameters with plausible associations (p<0.250) were tested. The cut-off values for age, infertility duration, induction duration, day 3 serum LH level, endometrial thickness, and the total number of retrieved oocytes were visually determined based on the probability distribution graphs. Significant changes were observed in over 37 years of age, over 10 years of infertility duration, over 6 mL/IU for serum LH level, over 9 mm of endometrial thickness, and below four retrieved oocytes. Limit values were used for further regression analysis. Among the parameters examined are age above 37 years (p=0.032), low antral follicle number (p=0.004), basal LH levels above 6 mIU/mL (p=0.001), stimulation cycle lasting longer than 10 days (p=0.003), <500 pg/mL estradiol level (p=0.077) on the triggering day, <4 collected oocytes (p=0.013), and grade B embryo transfer (p=0.005) The significance levels were not obtained in other parameters; however, parameters with known clinical effects on live birth were prioritized with the prediction model creation.

Multivariate Regression Analysis and Creation and Calibration of the Predictive Model

Two separate prediction models were created. In the basal model, only the patient demographics were used. In the combined model, cycle and embryo transfer characteristics were included in the patient demographics. One parameter was added or subtracted at a time. The model accuracy and calibration were tested at each stage. Parameters without a significant model consistency increase or that impair its calibration were excluded. The patient age, basal antral follicle count, and day 3 serum LH level were used in the basal model (Table 2). The probability of success decreased (odds ratio: 0.07, 95% CI: 0.00-0.36) in patients aged over 37 years, success rate decreased (odds ratio: 0.15, 95% CI: 0.02-0.53) in <5 antral follicles, and LH greater than 6 mIU/mL (odds ratio: 2.24, 95% CI: 1.27-3.94) was associated with success. The accuracy of the combined model revealed an area under the curve (AUC) of 0.68 (95% CI: 0.62-0.74) and model sensitivity of 0.28 (95% CI: 0.17-0.39) for a 10% fixed false-positive rate (Figure 1). The calibration curve of the basal model revealed that the observed probabilities were consistent with the predicted probabilities. The Hosmer-Lemeshow test revealed that the model calibration was good under this observation (p=0.979). In addition to the basal model parameters, the duration of the stimulation cycle, the endometrial thickness, and the number and grade of the embryo transferred were used in the combined model creation. A clinical and statistical interaction was found between the number and grade of embryos, thus, it was adapted to the model considering this feature. Cycles with 10 days or longer duration (odds ratio: 2.23, 95% CI: 1.17-4.42) and endometrial thickness wider than 9 mm (probability ratio: 2.07, 95% CI 1.00-4.53) were more successfully observed. The transferred embryo characteristics revealed a significantly increased successful single grade B embryo transfer, which was found as a negative effect (odds ratio: 0.07, 95% CI 0.00-0.39). The accuracy of the combined model revealed an AUC of 0.76.
Table 1. Comparison of basic variables used to establish prediction model

| Patient demographics | Live birth (n=85) | No live birth (n=220) | P†  |
|----------------------|------------------|-----------------------|-----|
| Age (years), median (IQR) | 29.0 (26.00-32.0) | 31.0 (27.0-36.0) | 0.004 |
| Body Mass Index (kg/m²), median (IQR) | 22.4 (21.1-26.2) | 23.8 (21.6-26.6) | 0.132 |
| Infertility duration (years), median (IQR) | 5.0 (3.0-6.0) | 5.0 (3.0-8.0) | 0.664 |

**Tubal occlusion**
- Unilateral, n (%) | 2 (2.4) | 7 (3.2) | 0.999 |
- Bilateral, n (%) | 4 (4.7) | 8 (3.6) | 0.743 |

**Infertility etiology**
- Male factor, n (%) | 28 (32.9) | 93 (42.3) | 0.152 |
- Female factor, n (%) | 14 (16.5) | 43 (19.5) | 0.624 |

**Antral follicle count, n (%)**
- ≥5 follicle | 62 (72.9) | 130 (59.1) | 0.025 |
- <5 follicle | 2 (2.4) | 38 (17.3) | <0.001 |
- Polycystic ovaries | 21 (24.7) | 52 (23.6) | 0.881 |

Day 3 serum FSH level (mIU/mL), median (IQR) | 7.0 (6.0-9.0) | 7.2 (6.0-9.1) | 0.864 |
Day 3 serum estradiol level (pg/mL), median (IQR) | 44.0 (29.5-56.0) | 40.5 (30.0-54.0) | 0.973 |
Day 3 serum LH level (mIU/mL), median (IQR) | 5.3 (3.7-7.0) | 5.0 (3.2-6.0) | 0.045 |

**Controlled ovarian hyperstimulation (COH) characteristics**
Initial gonadotropin dosage (IU), median (IQR) | 225.0 (225.0-300.0) | 225.0 (225.0-300.0) | 0.894 |

Down-regulation method
- Antagonist, n (%) | 82 (96.5) | 200 (90.9) | 0.144 |
- Agonist, n (%) | 3 (3.5) | 20 (9.1) | |

**Type of gonadotropin for ovarian stimulation**
- Only HMG, n (%) | 23 (27.1) | 73 (33.2) | 0.337 |
- Only recombinant FSH, n (%) | 12 (14.1) | 25 (11.4) | 0.558 |
- Recombinant FSH and HMG, n (%) | 50 (58.8) | 122 (55.4) | 0.609 |

**Ovarian trigger agent**
- Only GnRH agonist, n (%) | 7 (8.2) | 21 (9.6) | 0.827 |
- Only hCG, n (%) | 63 (74.1) | 167 (75.9) | 0.767 |
- GnRH agonist and hCG, n (%) | 15 (17.7) | 32 (14.5) | 0.485 |

Total gonadotropin dosage (IU), median (IQR) | 2400 (1800-3000) | 2400 (2025-2925) | 0.566 |

The number of >17 mm oocytes at trigger day, median (IQR) | 3.0 (2.0-4.0) | 3.0 (2.0-4.0) | 0.242 |
COH duration (day), median (IQR) | 12.0 (11.0-12.0) | 11.0 (10.0-12.0) | 0.003 |
Estradiol level at trigger day (pg/mL), median (IQR) | 2018 (1370-3196) | 1729 (993-2868) | 0.071 |
Progesterone level at trigger day (ng/mL), median (IQR) | 0.54 (0.41-1.00) | 0.67 (0.47-1.05) | 0.331 |
LH level at trigger day (mIU/mL), median (IQR) | 2.0 (1.2-3.5) | 2.0 (1.0-3.5) | 0.571 |

**OPU and ET characteristics**
Total picked up oocytes, median (IQR) | 10.0 (6.0-14.0) | 8.0 (5.0-13.0) | 0.007 |
Mature (MII) oocyte number, median (IQR) | 9.0 (4.0-12.0) | 5.0 (3.0-10.0) | <0.001 |
### Table 2. Odds ratios of models created based on multivariate logistic regression analysis

| Basal model (AUC: 0.68, 93% CI: 0.62-0.74) | Odds ratio (OR) (95% confidence interval (CI)) | P† |
|-------------------------------------------|-----------------------------------------------|----|
| **Age**                                   |                                               |    |
| ≤37 age                                   | Reference                                     |    |
| >37 age                                    | 0.07 (0.00-0.36)                              | 0.011 |
| **Basal antral follicle count**            |                                               |    |
| ≥5 follicle                                | Reference                                     |    |
| <5 follicle                                | 0.15 (0.02-0.53)                              | 0.012 |
| Polycystic ovaries                        | 0.60 (0.32-1.09)                              | 0.102 |
| Day 3 serum LH level >6.0 mIU/mL          | 2.24 (1.27-3.94)                              | 0.004 |
| **Combined model (AUC: 0.76, 95% CI: 0.70-0.81)** |                                               |    |
| **Age**                                   |                                               |    |
| ≤37 age                                   | Reference                                     |    |
| >37 age                                    | 0.10 (0.00-0.35)                              | 0.030 |
| **Basal antral follicle count**            |                                               |    |
| ≥5 follicle                                | Reference                                     |    |
| <5 follicle                                | 0.23 (0.03-0.96)                              | 0.026 |
| Polycystic ovaries                        | 0.50 (0.25-0.96)                              | 0.043 |
| Day 3 serum LH level >6.0 mIU/mL          | 2.36 (1.30-4.32)                              | 0.004 |
| **COH duration**                          |                                               |    |
| Shorter than 10 days                      | Reference                                     |    |
| 10 days or longer                         | 2.23 (1.17-4.42)                              | 0.055 |
| **Transferred embryo grade and number**   |                                               |    |
| Single, Grade A                           | Reference                                     |    |
| Double, Grade A                           | 0.90 (0.45-1.77)                              | 0.697 |
| Single, Grade B                           | 0.07 (0.00-0.39)                              | 0.014 |
| Double, Grade B                           | 0.39 (0.02-2.65)                              | 0.456 |

AUC: Area under curve, CI: Confidence interval
†Multivariate logistic regression
Prediction of success at first IVF (95% CI: 0.70-0.81) and model sensitivity of 0.31 (95% CI: 0.20-0.42) for a 10% fixed false-positive rate (Figure 2). The consistency of the combined model was statistically significantly higher than the baseline model (AUC: 0.76 vs AUC: 0.68, p<0.001 De Long test, respectively). The calibration curve of the combined model revealed that the observed probabilities were consistent with the predicted probabilities. The Hosmer-Lemeshow test revealed a good model calibration following this observation (p=0.848).

Nomograms were created for the practical application of the models (Supplementary Figure 1,2). The values from the lines next to the parameters are marked first to use the nomogram. Each parameter score is calculated with the lines drawn perpendicular to the score curve above. After the scores are collected, the total score is marked in the total score line below and the possibility of live birth is read with the perpendicular line drawn below.

The Comparison Between the Templeton and the Present Models

A comparison was made with the Templeton model to show the practical benefit of the basal model. The Templeton model parameters were adapted to our dataset, and receiver operating characteristic curves were created for both the models. The AUC of the Templeton model was 0.60 (95% CI: 0.53-0.67) (Figure 1. Receiver operating characteristic curve of the basal model using parameters of age, antral follicle count, and luteinizing hormone level)

Supplementary Figure 1. The nomogram of the basal model for clinical use. The probability of live births corresponding to the total score observed in the lowest row after the corresponding scores are added on the upper score sheet for each parameter

Supplementary Figure 2. The nomogram of the combined model for clinical use. The probability of live births corresponding to the total score observed in the lowest row after the corresponding scores are added on the upper score sheet for each parameter.
3). The accuracy of the Templeton model was lower (p=0.062, DeLong test) than that of the basal model (Figure 3). The sensitivity of the Templeton model for a fixed 10% false positivity rate was very low for clinical use (0.10, 95% CI: 0.03-0.19).

Discussion

The prediction models in the present study have acceptable precision and good calibration. The baseline model was used in the pretreatment phase while informing patients or making treatment decisions. In addition, static (Supplementary Figures 1,2) nomograms are available for practical use of the model, especially for clinicians.

More than 30 prediction models were presented in the literature, wherein most used similar parameters for model creation. Infertility duration and infertility type were not found to be associated with the odds of live birth in this model compared with more recognizable models. One of the reasons is the time censored nature of the infertility duration, which was inevitably affected by the patients’ age and the unpredictable exact duration. A recent meta-analysis by van Loendersloot et al. found a weak association between the infertility duration and live birth (odds ratio: 0.99 95% CI: 1.5 0.98-1.00), which also concluded that among 21 external validation studies, only the model could be generalized, which includes female age, number of retrieved oocytes, developmental stage score, and morphology score of two best embryos. In the present prediction model, in addition to these parameters, day 3 serum LH level, and endometrial thickness were found significantly associated with the odds of live birth.

The most recent prediction model in the literature aimed to calculate “the number of mature oocytes required to obtain at least one euploid embryo”. This model was externally validated and revealed >80% positive predictive values with all the predicted used possibilities by the authors. The primary ending for the IVF/ICSI cycle outcome was different than our model; however, female age, sperm source used for ICSI, and the number of mature oocytes were used as parameters of the predictive model. In this study, testicular sperms were used as a sperm source in all of the included cycles to overcome the negative effects of the malefactors.

Endometrial preparation for successful implantation was another key phase of any cycle’s endpoint. Several factors revealed the functionality of the endometrium. The optimal endometrial thickness was 10 mm at the Vaeger’s prediction model, and the impact of endometrial thickness on the models was also similar to ours. Other than the endometrial thickness, the duration of gonadotropin induction showed significance on our model and is a possible indicator of ovarian and endometrial response. Day 3 serum LH level was an important parameter in this study. In two recent studies, basal serum LH level was highly associated with an ovarian response especially agonist protocols. Lower LH levels during cycles are also related to a lower ongoing pregnancy rate, and ongoing pregnancy rates are higher at protocols that are supported with a recombinant LH based on a Cochrane review.

Stimulation durations longer than 10 days were associated with better outcomes in the first cycles, which is a reflection of the ovarian reserve and its effect on cycle success. Poor responders usually have a short stimulation duration due to already high endogenous follicle-stimulating hormone levels and asynchronous follicle growth. These patients have poorer outcomes compared to normo- and high-responders and our results reflect this mechanism.

Finally, transferred embryo grade and number were associated with ongoing pregnancy rates. This is an expected finding and was established in the literature.

Study Limitations

Several limitations were encountered in this study. Firstly, the number of included patients was below the average from similar studies in the literature. However, considering the patient volume of the clinic where the study was conducted and the included patient group, the number of patients was kept as high as possible and a wide range of years was chosen. In addition, the number of live births (n=85) in the cohort in which the model was developed is above the minimum number (n=10) per parameter in the logistic models. Therefore, the problem is not encountered in terms of statistical power. In addition, a possibility of selection bias is due to its retrospective nature. The possibility of a selection bias is never completely excluded although restrictive exclusion criteria were not set since the data source of the research was the patient files with
complete records. Another limitation was the indicator used for an ovarian reserve. The only parameter was the number of antral follicles and some studies reported that the serum anti-Müllerian hormone (AMH) level reflects the ovarian reserve better. The predictive value of the AMH level was not evaluated since AMH was not a routine parameter recorded in the years in which research records were obtained in our clinic. The internal validation of the developed model was made by developing a mixed-method due to the limited number of patients. The reserved patient population is mixed with the cohort in which the model was developed, and the validation study performed with the bootstrapping method is more insufficient than the studies using the external cohort. Finally, some interventional procedures were reported with IVF success association, and the relationship of these factors was not studied in our patient population.

The main strength of this study was the patients treated with current IVF protocols and techniques. Given that the prediction models perform better in populations with characteristics similar to the developed cohorts, our model was expected to perform better in external validation studies compared to its historical counterparts. Since the parameters used in the model were easily measured and generally recorded in IVF cycles, no technical problems were expected in external validation studies. In addition, during the creation and testing of the model, the highest standard statistical practices were adhered to, and the model was created with careful attention to technical principles. The value of the area (0.76) remaining in the high curve quotation observed in our study is the result of careful parameter selection and good statistical practice. Finding static and dynamic nomograms for the practical use of our model was another strong aspect.

**Conclusion**

The present created model was well-calibrated and easily interpretable to routine IVF/ICSI cycles. The combined model aid in the informed decision phase of the fertility-seeking couples; however, external validation is necessary with a large-sized prospective cohort to confirm the clinical usage.

**Ethics**

**Ethics Committee Approval:** The study was approved by the Institutional Clinical Research Ethics Committee (date: April 25, 2016; number: 08-341-16).

**Informed Consent:** Retrospective study.

**Peer-review:** Externally peer-reviewed.

**Authorship Contributions**

Surgical and Medical Practices: Y.E.Ş., B.Ö., M.S., C.B., R.A., Concept: E.K., B.B., Design: E.K., B.B., Data Collection or Processing: E.K., C.B., A.G., Analysis or Interpretation: E.K., B.B., Literature Search: E.K., C.B., A.G., Y.E.Ş., Writing: E.K., C.B., B.B.

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