Decreased Serum Level of Interleukin-22 Correlates with Hepcidin in Patients with Hidradenitis Suppurativa

Małgorzata PONIKOWSKA¹, Łukasz MATUSIAK¹, Monika KASZTURA², Ewa JANKOWSKA¹,² and Jacek C. SZEPIETOWSKI¹
¹Department of Dermatology, Venereology and Allergology, Wrocław Medical University, ²Department of Food Hygiene and Consumer Health Protection, Faculty of Veterinary Medicine, Wrocław University of Environmental and Life Sciences

Current understanding of the underlying pathophysiology of hidradenitis suppurativa (HS) links the disease with proinflammatory activation and autoimmune processes. This study investigated serum levels of interleukin (IL)-22, a cytokine critically involved in epithelial homeostasis, in the context of the broad clinical spectrum of patients with HS. The study also assessed the relationship between serum IL-22 and pro-inflammatory activation (as evidenced by serum level of IL-6) and serum hepcidin (central regulator of systemic iron homeostasis). Serum concentrations of IL-22 were assessed in 74 patients with HS and 15 healthy subjects. Compared with healthy controls, patients with HS demonstrated decreased levels of serum IL-22 (median; interquartile range (IQR): 12.4 pg/ml (9.8; 23.5) vs 34.8 pg/ml (24.8; 39.8), \( p < 0.001 \) vs controls). Disease severity (assessed both with Hurley staging and Hidradenitis Suppurativa Severity Index) did not differentiate IL-22 levels (\( p > 0.1 \) in both comparisons). Serum levels of IL-22 and IL-6 did not correlate with each other (\( R = -0.17, p = ns \)). In a subgroup of 24 patients with HS with pro-inflammatory activation, the mean level of IL-22 was similar to that of the remaining patients (median (IQR): 9.8 pg/ml (8.5; 15.0) vs 12.0 pg/ml (9.4; 16.3), \( p = ns \)). Patients with HS demonstrated a decreased level of hepcidin (mean: 31.3 ± 25.9 pg/ml, which correlated with the levels of IL-22 (\( R = 0.36, p < 0.05 \)). Patients with HS demonstrated significantly decreased levels of serum IL-22, which was neither correlated with pro-inflammatory status nor associated with disease severity, but correlated modestly with serum hepcidin.

Key words: hidradenitis suppurativa; cytokines; interleukin-22; hepcidin.

Accepted Sep 8, 2021; Epub ahead of print Sep 13, 2021

Acta Derm Venereol 2021; 101: adv00558.

Corr: Jacek C. Szepietowski, Department of Dermatology, Venereology and Allergology, Wrocław Medical University, ul. Chałubińskiego 1, PL-50-368 Wrocław, Poland. E-mail: jacek.szepietowski@umed.wroc.pl

H idradenitis suppurativa (HS) is a chronic skin disease with a recurrent nature and debilitating course, with a prevalence of 0.05–4.1% of the population (1, 2). HS significantly impairs patients’ quality of life and remains a challenge in terms of treatment and management (3). The pathophysiology of HS is multi-factorial and not fully understood. Recent data strongly indicate the generalized character of the disorder, with a pivotal role of immunological abnormalities and pro-inflammatory activation (4, 5). Subclinical inflammation presents within the skin of patients with HS before the appearance of clinical lesions leads to aberrant secretion of cytokines and abnormal production of antimicrobial proteins (AMP) (6). AMPs are produced continuously by the keratinocytes as part of innate immunity in response to microbial infection and inflammation. In addition to antimicrobial properties, AMPs exhibit multiple immunomodulatory functions; namely stimulation of production of cytokines (6, 7). Interleukin (IL)-22, a member of the IL-10 family, is released by various types of lymphoid and non-lymphoid cells, and its production is regulated by an interplay of cytokines and transcription factors (8). IL-22 interacts directly with IL-22 receptors expressed in tissue cells (broadly distributed in the skin, digestive and respiratory system), preserving the integrity of homeostasis in the boundary organs and tissues against intestinal pathogens and commensal bacteria (9). In the epithelial cells IL-22 stimulates AMPs, therefore preventing bacterial infections and interacting with inflammatory processes (10). The role of upregulated IL-22 production has been reported in psoriasis (11). Little is known about downstream IL-22 signalling in HS. It appears that HS

SIGNIFICANCE

Although interleukin-22 plays an important role in various inflammatory skin disorders, there are very limited data regarding its status in hidradenitis suppurativa, which is a rare, chronic and highly debilitating disease. The aim of this study was to assess serum levels of interleukin-22 in patients with hidradenitis suppurativa and to connect it with proinflammatory activation, potential derangements in iron status, and the clinical severity of the disease. The results showed that circulating levels of interleukin-22 were significantly lower in patients with hidradenitis suppurativa in comparison with healthy controls, and that these levels correlated with decreased levels of hepcidin, which is the central regulator of iron storage. Surprisingly, there was no connection between interleukin-22 levels and either proinflammatory activation or severity of hidradenitis suppurativa.

This is an open access article under the CC BY-NC license. www.medicaljournals.se/acta

Society for Publication of Acta Dermato-Venereologica
doi: 10.2340/00015555-3928
Acta Derm Venereol 2021; 101: adv00558
is characterized by relative deficiency of IL-22 expression in the lesional skin compared with other chronic inflammatory skin diseases, such as psoriasis or atopic dermatitis (12).

The aims of this study were to evaluate the serum level of IL-22 in a broad clinical spectrum of patients with HS, and to assess the relationship between serum IL-22 and serum hepcidin, which is central regular of systemic iron homeostasis. Our previous study found dysregulated iron homeostasis in HS (13), and IL-22 is known to regulate iron availability through induction of hepcidin (14).

MATERIALS AND METHODS

Study cohort
The study was conducted following approval from the local ethics committee (number KB-336/2017 issued by Wrocław Medical University Ethics Committee). Seventy-four patients (37 women and 37 men, mean ± standard deviation (SD) age: 37 ± 14 years) with diagnosed HS (according to well-established clinical criteria) (15) were included in this study. The mean duration of the disease was 9.7 ± 8.8 years. Subjects with any acute or chronic illness that might have influenced pro-inflammatory activation (including infection, known malignancy, chronic kidney disease, chronic cardiovascular diseases and haematological diseases) were excluded. Since the study evaluated iron status-related biomarkers, those patients who received treatment for anaemia or iron deficiency in the previous 12 months were also excluded. None of the patients received any biologic treatment before entering the study and they all were biologic naïve. None of the patients took any immunomodulatory medications (including antibiotics) for at least 6 months prior to inclusion. The clinical evaluation of the patients was performed with Statistica 10.

The control group comprised 15 subjects (8 women and 7 men, mean ± SD age: 40 ± 7 years) with no acute disease (occurring in the previous 6 months) or chronic disease and related therapy. Laboratory controls were recruited from the hospital staff and patients’ relatives

| Variable                              | Patients (n = 74) | Controls (n = 15) |
|---------------------------------------|------------------|------------------|
| Age, years, mean ± SD                | 37 ± 14          | 40 ± 7           |
| Sex (female/male), n                  | 37/37            | 8/7              |
| BMI, kg/m², mean ± SD                 | 30.0 ± 7.0       | 29.1 ± 3.7       |
| Smoking status, smokers/non-smokers, %| 42/58            | 33/67            |
| Laboratory findings                  |                  |                  |
| Creatinine, mg/dl, mean ± SD         | 0.79 ± 0.15      |                  |
| AST, IU/l, mean ± SD                  | 17.4 ± 10.5      |                  |
| ALT, IU/l, mean ± SD                  | 15.9 ± 12.1      |                  |
| Duration of HS, years, mean ± SD      | 9.7 ± 8.8        |                  |
| Hurley I/II/III, %                    | 30/52/18         |                  |
| HSSI, points, mean ± score ± SD      | 10.4 ± 4.1       |                  |
| DLQI, points, mean ± score ± SD      | 13.0 ± 7.6       |                  |
| SD: standard deviation; BMI: body mass index; AST: aspartate transaminase; ALT: alanine transaminase; HSSI: Hidradenitis Suppurativa Severity Index; DLQI: Dermatology Life Quality Index. |

Laboratory assessments
In both groups (HS patients and healthy controls) venous blood samples were taken in the morning after an overnight fast, following at least 15 min of supine rest. After centrifuging, the plasma and serum were collected and frozen at ~70°C until further analysis.

Serum level of IL-22 was measured using a commercially available enzyme-linked immunoassay (ELISA) (R&D Systems, Minneapolis, MN, USA). Serum level of IL-6 (pg/ml), as a marker of proinflammatory activation, was measured using a commercially available ELISA (R&D Systems) (19). The upper limit of normal in our laboratory for IL-6 was 1.48 pg/ml. We prospectively defined HS patients with IL-6 above 1.48 pg/ml as having proinflammatory activation (20).

Serum hepcidin (ng/ml) was measured using a commercially available ELISA (BACHEM, Bubendorf, Switzerland). This ELISA method was validated against a gold standard for hepcidin assessment, namely liquid chromatography mass spectrometry developed at King’s College London, confirming a strong correlation between the measurements performed using the liquid chromatography mass spectrometry and the BACHEM assay in patients with chronic kidney diseases and healthy subjects (21). Increased levels of hepcidin may occur in the circulation due to the state of inflammation (22).

Basic laboratory biomarkers indicating liver and kidney function were measured: creatinine (mg/dl), aspartate transaminase (AST), and alanine transaminase (ALT) (IU/l).

Statistical analysis
The normality of the distributions of continuous variables was tested using the Kolmogorov–Smirnov test. Continuous variables with a normal distribution were expressed as means ± SD. The remaining continuous variables with a skewed distribution were expressed as medians with upper and lower quartiles (interquartile range; IQR). Categorical variables were expressed as numbers with percentages. The statistical significance of differences between the groups was tested using Student’s t-test, Mann–Whitney U test, or the χ² test, where appropriate. The associations between variables were assessed using Spearman’s rank correlation coefficients. All statistical analyses were performed with Statistica 10 (Statsoft, Tulsa, OK, USA). A value of p < 0.05 was considered statistically significant.

RESULTS
The clinical severity of the disease was assessed with Hurley staging with the following distribution: 22 patients (30%) were classified as stage I, 39 (52%) as stage II and 13 (18%) as stage III, respectively. HS severity assessed with HSSI was 10.4 ± 4.1 points (range 4–18 points). Patients’ quality of life, evaluated with Dermatology Life Quality Index (DLQI), was impaired with mean score of 13.0 ± 7.6 points (range 0–28 points).

Serum interleukin-22 in patients with hidradenitis suppurativa
Compared with healthy controls, patients with HS demonstrated a significantly decreased level of serum IL-22 (median [IQR]: 12.4 pg/ml [9.8; 23.5] vs 34.8 pg/ml [24.8; 39.8], p < 0.001 vs controls) The IL-22 level neither differed across Hurley stages nor across HSSI (p > 0.05) (Table II).
Decreased serum IL-22 correlates with hepcidin in patients with HS

Relationship between serum interleukin-22 and pro-inflammatory activation in patients with hidradenitis suppurativa

Patients with HS demonstrated elevated levels of IL-6 (median [IQR]: 0.05 [0.05; 2.6] pg/ml). There was no correlation between serum level of IL-6 and disease severity expressed as HSSI (R = 0.22, p = 0.06).

There was no correlation between levels of IL-22 and IL-6 in the entire study population (R = –0.166, p = 0.16).

A group of 24 patients with HS (32%) with evidence of pro-inflammatory activation (defined as IL-6 level above the upper limit of normal in our laboratory) was identified. Serum levels of IL-22 did not differ between these 2 groups (median [IQR]: 9.8 pg/ml [8.5; 15.0] vs 12.0 [9.4; 16.3], patients with proinflammatory activation vs remaining patients, p > 0.2). Also, in the group of 24 patients with proinflammatory activation, no correlation was found between serum levels of IL-22 and IL-6 (R = 0.144, p > 0.1).

Relationship between serum interleukin-22 and hepcidin in patients with hidradenitis suppurativa

As reported previously, patients with HS demonstrated decreased levels of hepcidin (mean 31.3 ± 25.9 pg/ml), which did not correlate with the severity of the disease expressed by HSSI (R = 0.01, p = 0.98). Hepcidin serum levels in our patients with HS correlated with IL-22 (R = 0.36, p < 0.05; Fig. 1).

DISCUSSION

This study revealed decreased serum levels of IL-22 in a broad clinical spectrum of patients with HS, which neither correlated with pro-inflammatory activation nor with disease severity. There was a modest correlation between serum levels of IL-22 and hepcidin, suggesting a potential relationship between IL-22 signalling and iron status in HS.

A growing body of evidence indicates that deranged inflammatory mechanisms are involved in the underlying pathophysiology of HS (4). However, it remains unclear which immune pathways and cytokines are involved in pro-inflammatory signalling leading to disease progression. Among cytokines for which a pathogenic role appears to be well-established are IL-1, IL-17, IL-10, and tumour necrosis factor (TNF)-alpha, which have also

![Fig. 1. Modest correlation (R = 0.36, p < 0.05) between serum levels of interleukin (IL)-22 and hepcidin, suggesting potential interrelationship between IL-22 signalling and iron status in hidradenitis suppurativa.](image-url)
become therapeutic targets in HS (24–27). In contrast to up-regulated signalling comprising these cytokines, studies investigating cellular responses in skin lesions of patients with HS revealed reduced numbers of IL-22-secreting cells with down-regulation of the IL-22 pathway (28). Interestingly, IL22 does not directly interact with the immune cells, but targets cells in the outer-body barriers (e.g. the skin, digestive and respiratory systems) and stimulates the production of antibacterial proteins and chemokines (29). IL-22 deficiency in the skin lesions in HS may be associated with an altered pattern of production of AMPs with subsequently impaired skin protection against infections (29, 30). To the best of our knowledge this study is the first to investigate serum levels of IL-22 in a broad clinical spectrum of patients with HS. The results suggest that, in HS, not only is local expression of IL-22 in the skin lesions affected (compared with psoriasis and atopic dermatitis (12)), but also the serum level of this cytokine is significantly decreased, which may indicate more a generalized down-regulation of IL-22 signalling pathways in HS. A lack of interrelation was found between serum IL-22 and disease severity, which can be interpreted as indirect evidence of an early derangement of IL-22 signalling during the natural course of the disease. The results of a double-blind, randomized, placebo-controlled trial with IL-1 antagonist anakinra in patients with HS, demonstrated that IL-1 blockade was associated with improved clinical status, with a concomitant increase in IL-22 production by peripheral blood mononuclear cells (31). These findings do not contradict the therapeutic observations. The current study included patients with less severe disease (as evidenced by both Hurley stage and DLQI), and anti-IL-1 strategy may well result in clinical improvement and the up-regulation of IL-22 signalling across the whole disease spectrum. Interestingly, 24 out of total 74 patients with HF (32%) demonstrated generalized pro-inflammatory activation, as evidenced by an elevated level of serum IL-6. In these patients the serum level of IL-22 did not differ from that in the remaining patients, which may indicate that inflammatory-mediated mechanisms are not involved in IL-22 down-regulation, at least as detected in the blood.

The modest, but significant, relationship between serum hepcidin and IL-22, is an intriguing novel finding. It has been well established that production of hepcidin, a key regulator of iron homeostasis in the body, is induced by proinflammatory cytokine IL-6. We have recently reported deranged iron status in HS, which can be characterized as iron deficiency with a concomitant low level of hepcidin (11). Smith et al. (14) described that IL-22 signalling induces hepcidin production independently of IL-6. Hepcidin limits iron uptake and availability by causing iron accumulation in the macrophages. Therefore, the local and generalized down-regulation of IL-22 found in HS, may be the mechanism underlying the low level of hepcidin, and might lead to reduced iron availability with resultant deterioration of processes involved in microbial defence. However, this hypothetical pathway needs further study.

In summary, patients with HS demonstrated significantly decreased levels of serum IL-22, which was neither related to pro-inflammatory status nor associated with disease severity, but modestly correlated with serum hepcidin. Further research into the role of IL-22 in HS is required.

ACKNOWLEDGEMENTS

The preliminary results of our research were presented at European Hidradenitis Suppurativa Foundation 2019 8th Conference.

Funding source: Project for Young Scientists (number STM. C260.20.097), Wroclaw Medical University.

The authors have no conflicts of interest to declare.

REFERENCES

1. Zouboulis CC, Desai N, Emtestam L, Hunger RE, Joannides D, Juhász I, et al. European S1 guideline for the treatment of hidradenitis suppurativa/ acne inversa. J Eur Acad Dermatol Venereol 2015; 29: 619–644.
2. Saunte DML, Jemec GBE. Hidradenitis suppurativa: advances in diagnosis and treatment. JAMA 2017; 318: 2019–2032.
3. Matusiak L, Bieniek A, Szepeitowski JC. Psychophysical aspects of hidradenitis suppurativa. Acta Derm Venereal 2010; 90: 264–268.
4. Kelly G, Prens EP. Inflammatory mechanisms in hidradenitis suppurativa. Dermatol Clin 2016; 34: 51–58.
5. Włodarek K, Ponikowska M, Matusiak L, Szepeitowski JC. Biologics for hidradenitis suppurativa: an update. Immunotherapy 2019; 11: 45–59.
6. Prens E, Deckers I. Pathophysiology of hidradenitis suppurativa: an update J Am Acad Dermatol 2015; 73: S8–S11.
7. Bechara FG, Sand M, Skrygan M, Kreuter A, Alltmeyer P, Gambichler T. Acne inversa: evaluating antimicrobial peptides and proteins. Ann Dermatol 2012; 24: 393–397.
8. Lin S, Yang X, Liang D, Zheng SG. Treg cells: a potential regulator for IL-22 expression? Int J Clin Exp Pathol 2014; 7: 474–480.
9. Sabat R, Ouyang W, Wolk K. Therapeutic opportunities of the IL-22-IL-22R1 system. Nat Rev Drug Discov 2014; 13: 21–38.
10. Shabgah AG, Navashenajg JG, Shabgah OG, Mohammadi H, Sahebkar A. Interleukin-22 in human inflammatory diseases and viral infections. Autoimmun Rev 2017; 16: 1209–1218.
11. Wolk K, Haugen HS, Xu W, Witte E, Waggie K, Anderson M, et al. IL-22 and IL-20 are key mediators of the epidermal alterations in psoriasis while IL-17 and IFN-gamma are not. J Mol Med 2009; 87: 523–536.
12. Wolk K, Warszawska K, Hoeflick J, Witte E, Schneider-Burrus S, Witte K, et al. Deficiency of IL-22 contributes to a chronic inflammatory disease: pathogenic mechanisms in acne inversa. J Immunol 2011; 186: 1228–1239.
13. Ponikowska M, Matusiak L, Kasztura M, Jankowska EA, Szepeitowski JC. Deranged iron status evidenced by iron deficiency characterizes patients with hidradenitis suppurativa. Dermatology 2020; 236: 52–58.
14. Smith CL, Arvedson TL, Cooke KS, Dickmann LJ, Forte C, Li H, et al. IL-22 regulates iron availability in vivo through the induction of hepcidin. J Immunol 2013; 191: 1845–1855.
15. Poli F, Jemec GB, Revuz J. Clinical presentation. In: Jemec GB, Revuz J, Leyden JJ, editors. Hidradenitis suppurativa. Heidelberg: Springer, 2006: p. 11–24.
Decreased serum IL-22 correlates with hepcidin in patients with HS

16. Sartorius K, Emtestam L, Jemec GB, Lapins J. Objective scoring of hidradenitis suppurativa reflecting the role of tobacco smoking and obesity. Br J Dermatol 2009; 161: 831–839.

17. Hurley HJ. Axillary hyperhidrosis, apocrine bromhidrosis, hidradenitis suppurativa and familial benign pemphigus. Surgical approach. In: Roenigk RK, Roenigk HH, editors. Röenigk & Röenigk’s dermatologic surgery: principles and practice (2nd edn). New York: Marcel Dekker, 1996: p. 623.

18. Finlay AY, Khan GK. Dermatology Life Quality Index (DLQI) – a simple practical measure for routine clinical use. Clin Exp Dermatol 1994; 19: 210–216.

19. Grant A, Gonzalez T, Montgomery MO, Cardenas V, Kerdel FA. Infliximab therapy for patients with moderate to severe hidradenitis suppurativa: a randomized, double-blind, placebo-controlled crossover trial. J Am Acad Dermatol 2010; 62: 205–217.

20. Jankowska EA, Malyszko J, Ardebali H, Koc-Zorawska E, Banasiak W, von Haehling S, et al. Iron status in patients with chronic heart failure. Eur Heart J 2013; 34: 827–834.

21. Schmidt PJ. Regulation of iron metabolism by hepcidin under conditions of inflammation. Biol Chem 2015; 290: 18975–18983.

22. Tanaka T, Narazaki M, Kishimoto T. IL-6 in inflammation, immunity, and disease. Cold Spring Harb Perspect Biol 2014; 4: 6.

23. Kelly G, Sweeney CM, Tobin AM, Kirby B. Hidradenitis suppurativa: the role of immune dysregulation. Int J Dermatol 2014; 53: 1186–1196.

24. Matusiak L, Bieniek A, Szepietowski JC. Increased serum tumour necrosis factor-alpha in hidradenitis suppurativa patients: is there a basis for treatment with anti-tumour necrosis factor-alpha agents? Acta Derm Venereol 2009; 89: 601–603.

25. van der Zee HH, de Ruiter L, van den Broecke DG, Dik WA, Laman JD, Prens EP. Elevated levels of tumour necrosis factor (TNF)-α, interleukin (IL)-1β and IL-10 in hidradenitis suppurativa skin: a rationale for targeting TNF-α and IL-1β. Br J Dermatol 2011; 164: 1292–1298.

26. Glowaczewska A, Szepietowski JC, Matusiak Ł. Severe hidradenitis suppurativa successfully treated with secukinumab. Dermatol Ther 2020; 16: e13845.

27. Hotz C, Bonirotto M, Guguin A, Surenaud M, Jean-Louis F, Tisserand P, et al. Intrinsic defect in keratinocyte function leads to inflammation in hidradenitis suppurativa. J Invest Dermatol 2016; 136: 1768–1780.

28. Sabat R, Ouyang W, Wolk K. Therapeutic opportunities of the IL-22-IL-22R1 system. Nat Rev Drug Discov 2014; 13: 21–38.

29. Jones D, Banerjee A, Berger PZ, Gross A, McNish S, Amdur R, Shannugam VK. Inherent differences in keratinocyte function in hidradenitis suppurativa: Evidence for the role of IL-22 in disease pathogenesis. Immunol Invest 2018; 47: 57–70.

30. Tzaniaktou V, Kanni T, Giatrakou S, Katsoulis A, Papadavid E, Netae MG, et al. Safety and efficacy of anakinra in severe hidradenitis suppurativa: a randomized clinical trial. JAMA Dermatol 2016; 152: 52–59.