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Inhibitory activity of FDA-approved drugs cetilistat, abiraterone, diiodohydroxyquinoline, bexarotene, remdesivir, and hydroxychloroquine on COVID-19 main protease and human ACE2 receptor: A comparative in silico approach

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ARTICLE INFO

Keywords:
Molecular docking
Dynamic simulation
COVID-19
ACE2 receptor
Main protease

ABSTRACT

By September 1, 2021, SARS-CoV-2, a respiratory virus that prompted Coronavirus Disease in 2019, had infected approximately 218,567,442 patients and claimed 4,534,151 lives. There are currently no specific treatments available for this lethal virus, although several drugs, including remdesivir and hydroxychloroquine, have been tested. The purpose of this study is to assess the activity of FDA-approved drugs cetilistat, abiraterone, diiodohydroxyquinoline, bexarotene, remdesivir, and hydroxychloroquine as potential SARS-CoV-2 main protease inhibitors. Additionally, this study aims to provide insight into the development of potential inhibitors that may inhibit ACE2, thereby preventing SARS-CoV-2 entry into the host cell and infection. To this end, remdesivir and hydroxychloroquine were used as comparator drugs. The calculations revealed that cetilistat, abiraterone, diiodohydroxyquinoline, and bexarotene inhibit main protease and ACE2 receptors more effectively than the well-known drug hydroxychloroquine when used against COVID-19. Meanwhile, bexarotene and cetilistat bind more tightly to the SARS-CoV-2 main protease and the ACE2 receptor, respectively, than remdesivir, a potential treatment for COVID-19 that is the first FDA-approved drug against this virus. As a result, the molecular dynamic simulations of these two drugs in the presence of proteins were investigated. The MD simulation results demonstrated that these drugs interact to stabilize the systems, allowing them to be used as effective inhibitors of these proteins. Meanwhile, bexarotene, abiraterone, cetilistat, and diiodohydroxyquinoline’s systemic effects should be further investigated in suitable ex vivo human organ culture or organoids, animal models, or clinical trials.

1. Introduction

SARS-CoV-2 (COVID-19) is a respiratory virus that originated in Wuhan, in 2019 [1,2]. This pandemic has spread aggressively throughout the world (Iran, Italy, India, Republic of Korea, United States of America, India, among others). Despite numerous significant efforts worldwide, there are currently no specific treatments for this lethal virus. Until September 1, 2021, this virus had infected 218,567,442 people worldwide and killed 4,534,151 people (https://www.worldometers.info/coronavirus/) due to lung failure caused by SARS-CoV-2 disease. COVID-19 attacks and destroys the protective proteins on T-lymphocytes in the human lung. This condition causes an accumulation of fluid in the lungs, resulting in breathing difficulties [3]. Numerous drug combinations have been used to treat this virus, including Ritonavir, Oseltamivir, and lopinavir [4], Darunavir, Remdesivir, Saquinavir [5], Sofosbuvir [6], Azithromycin and Hydroxychloroquine [7], and (Hydroxychloroquine and Chloroquine) [8–10]. The virus enters host cells via its surface spike glycoprotein and attacks the protein on the surface of human cells. It then begins its activity. Numerous drugs are directed against the main protease required for viral replication and the processing of polyproteins from the virus’s RNA [11]. Thus, inhibiting this protease results in virus replication is prevented [12]. The kidney, lung cells, intestines, heart, lung cells, and arteries all have an outer membrane that contains an enzyme named ACE2. This critical enzyme plays a key role in cardiovascular/renal diseases and hypertension by regulating blood pressure and body fluid.

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https://doi.org/10.1016/j.imu.2021.100745
Received 28 July 2021; Received in revised form 20 September 2021; Accepted 20 September 2021
Available online 22 September 2021

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The essential step in the infection of the host cell by SARS-CoV-2 is binding to the ACE2 receptor. Therefore, the ACE-2 receptor is a key target for researchers looking for therapeutic opportunities to combat this lethal virus. Anti-COVID-19 therapeutics that employ various strategies, including experimental and computational approaches, is critical for containing this pandemic disease [7,13]. Drug repurposing using a computer-aided drug design (CADD) approach, i.e., a computational drug repurposing study, is an appealing method for exploring potential therapeutic opportunities and new medications against COVID-19 that requires less time and expense. Multiple types of research have employed in silico approaches to investigate a promising candidate with a high binding affinity and interaction with the ACE-2 receptor and SARS-CoV-2 proteins. Bhardwaj et al. described the inhibitory activity of bioactive molecules derived from tea plants against SARS-main CoV-2’s protease [14] and RNA-dependent RNA polymerase [15]. In another study, Bhardwaj et al. used a comparative in silico approach to evaluate the inhibitory activity of acridinedione analogs against the SARS-CoV-2 main protease [16]. Singh et al. [17] and Sharma et al. [18] recently reported on an in silico investigation of bioactive compounds from tea as potential inhibitors of SARS-CoV-2 nonstructural protein 15 and 16. In another research, Singh et al. [19] described a series of molecules based on 3α,7α-himachalenes scaffolds derived from a plant (Cedrus deodara) as potential SARS-CoV-2 nonstructural protein 1 inhibitors. Hussien et al. investigated the drug brincidofovir, which targets the ACE2 receptor and the main protease of SARS-CoV-2 [20]. Kalhor et al. used in silico strategies to determine the effect of the approved small molecule on the SARS-CoV-2 chimeric receptor-binding domain complexed with ACE2 [21]. Yuan et al. used a robust two-tier screening system in vitro to demonstrate the inhibitory activity of FDA-approved drugs cetilistat, abiraterone, diiodohydroxyquinoline, and bexarotene against COVID-19 infection [22]. A docking simulation approach was employed in this paper to investigate the interaction of four FDA-approved drugs, bexarotene (antineoplastic retinoid), abiraterone (synthetic androstan steroid), diiodohydroxyquinoline (anti-parasitic), and cetilistat (anti-pancreatic lipase), with the SARS-CoV-2 main protease and spike receptor domain complexed with ACE2 (ACE2-RBD). The results were compared to those obtained with hydroxychloroquine (an antimalarial medication) [7] and remdesivir (an antiviral drug). Additionally, the dynamic simulation elucidates the molecules’ dynamic behavior. Several clinical trials indicated that antiviral, antimalarial, and anti-HIV medications were effective against the COVID-19 virus. Among them, remdesivir (Veklury®) is the first FDA-approved drug to treat COVID-19 disease in hospitalized adults and children aged 12 years and older who weigh at least 40 kg [23]. According to Nguyen et al., remdesivir may target RNA polymerase and the main protease, which may help explain why this drug is effective against COVID-19 [24]. Chloroquine and hydroxychloroquine were previously considered the most likely candidates for reducing the frequency of human SARS-CoV-2 infections [7]. Chloroquine was the first drug to be used on the front lines in China and abroad to treat severe SARS-CoV-2 infections.

While in vitro and preclinical evidence for these drugs was promising, clinical benefit has been limited, and the researchers concluded that hydroxychloroquine and chloroquine do not appear to be effective in reducing the mortality rate of COVID-19 patients hospitalized. Another solidarity trial, the French discovery trial, supported the findings (https://who.int/news-room/q-a-detail/coronavirus-disease-covid-19-hydroxychloroquine). Despite the study’s limitations, researchers did not find any effect of hydroxychloroquine or chloroquine, which were used in several countries to treat COVID-19. Hydroxychloroquine, has a better clinical safety profile than chloroquine (over a long period), allows for a higher daily dose [25], and poses fewer concerns about drug-drug interactions [26]. Baildia et al. [27] and Beura et al. [28] employed an in silico approach to demonstrate the inhibitory activity of hydroxychloroquine on the COVID-19 main protease and the human ACE2 receptor. The drug-protein interaction was simulated in this study to better understand the mechanism of the proposed SARS-CoV-2 drugs and investigate the inhibitory effect of bexarotene, abiraterone diiodohydroxyquinoline, and cetilistat on the SARS-CoV-2 main protease. Additionally, this study aims to provide insight into the development of potential inhibitors that can inhibit ACE2 and thus prevent viral infection. The findings of this research can be used to introduce new candidate agents for ex vivo and in vivo surveys and provide necessary details for future research.

2. Methods

2.1. Docking of cetilistat, abiraterone, diiodohydroxyquinoline, bexarotene, remdesivir, and hydroxychloroquine

2.1.1. Receptors (proteins) preparation

AutoDock Vina [29] with MGL tools 1.5.4 was used to perform blind docking calculations. AutoDock Vina runs faster than AutoDock software and also makes more precise docking calculations [30]. The SARS-CoV-2 main protease and the ACE2-RBD were obtained from the Protein Data Bank (PDB ID: 6LU7, 6LZG) [31], and inhibitors were selected and removed. AutoDock Tools (v.1.5.4) was used to prepare the receptors. MGL Tools (v.1.5.4) combined non-polar hydrogen atoms and added Kollman charged atoms to protein crystal structures. SARS-CoV-2 main protease and ACE2-RBD coordinate files were then saved in PDBQT format. The 6LU7 was surrounded in a 55 × 68 × 65 box direction with a grid spacing of 1.00 Å and grid set centers of −26.02, 12.57, and 59.17, while the 6LZG was surrounded in a 64 × 74 × 110 box direction with a grid spacing of 1.00 Å and grid set centers of −25.41, 18.43, and −6.37.

2.1.2. Ligands (drugs) preparation

The three-dimensional SDF structures of cetilistat, abiraterone, diiodohydroxyquinoline, bexarotene, remdesivir, and hydroxychloroquine were obtained from PubChem (https://pubchem.ncbi.nlm.nih.gov) and converted to PDB format. MGL Tools (v.1.5.4) was then used to save the drugs in PDBQT format, and docked results were visualized through the BIOVIA Discovery Studio software.

2.2. Molecular dynamics (MD) simulations

Molecular docking analysis revealed that bexarotene and cetilistat have the lowest energy in their interactions with the SARS-CoV-2 main protease and ACE2-RBD, respectively. As a result, the molecular dynamic simulations of these two drugs were investigated in the presence of proteins. Classical MD simulations [32,33] were conducted using the CHARMM27 force field [34,35]. SwissParam was used to generate the ligand topology [36]. TIP3P water [37] was used to solve the free protein and protein-ligand complexes in the cubic box with periodic boundary conditions in three directions. The solutes were positioned in the box’s center, with a minimum distance of 1.0 nm between the solutes’ surface and the box. Na+ ions were added to the system to neutralize the charge. The systems were balanced at 300 K and 1.0 bar after energy minimization through the steepest descent method. The Parrinello-Rahman barostat was used to maintain a pressure of 1.0 bar, and a temperature of 300 K was maintained using a modified Berendsen thermostat. The LINCS algorithm enabled the computation of bond lengths, while the particle-mesh Ewald scheme (PME) was used to compute long-range electrostatic forces (grid spacing 0.16 nm) [38]. The short-range nonbonded interactions were computed using cutoff ratios of 1.0 nm for van der Waals and Coulomb potentials. Finally, a 30 ns MD simulation with a time step of 2 fs was performed with random generation of velocities through a Maxwell distribution.

2.3. Interaction analysis by MM-PBSA binding energy

Using the Molecular Mechanics Poisson Boltzmann Surface Area (MM-PBSA) technique, the binding free energy of protein-ligand systems...
was calculated. This in silico method was a combined energy system described by the binding free energy, composed of electrostatic, SASA, van der Waals, and polar solvation energies. The MM-PBSA binding free energies were calculated using the GROMACS script g_mmpbsa [39].

Using the MM-PBSA method, the following equation was employed to determine the binding free energy of the interacting proteins:

$$\Delta G_{\text{binding}} = G_{\text{complex}} - (G_{\text{receptor}} + G_{\text{ligand}})$$

$$\Delta G_{\text{binding}}$$ denotes the total binding energy of the protein-ligand complexes; $$G_{\text{receptor}}$$ and $$G_{\text{ligand}}$$ denote the binding energy of the free receptor and unbounded ligand, respectively.

3. Results

3.1. Molecular docking studies

While significant efforts are being made to develop drugs and vaccines against COVID-19, only a few available therapeutic agents are currently available. One of the most effective strategies for resolving this issue and combating this lethal virus is drug repurposing or using drugs already in clinical use for a different therapeutic indication. In drug discovery, molecular simulation approaches (docking and MD simulation) significantly reduce experimental time and cost [40]. Several research groups recently reported discovering inhibitors of the SARS-CoV-2 main protease and the ACE2 receptor [41–44]. As is well known, the surface of the lung contains a large number of ACE2 receptors. Compounds that interact with the ACE2 receptor inhibit its function, decrease its surface area, and ultimately prevent virus entry into the host cell [45]. The main protease is a critical SARS-CoV-2 protein that plays a vital role in virus replication. Viral replication and infection may be inhibited by compounds that interact with this key protein [46]. Yuan et al. demonstrated that FDA-approved drugs cetilistat, abiraterone, diiodohydroxyquinoline, and bexarotene inhibited SARS-CoV-2 infection in vitro [22]. However, the question remains regarding the fundamental objective of this study. In this research, the binding affinity and interaction of cetilistat, abiraterone, diiodohydroxyquinoline, bexarotene, remdesivir, and hydroxychloroquine with the SARS-CoV-2 protease and ACE2 receptor were evaluated, which are responsible for virus replication and entry into the host cell, respectively. This computational study aids in clarifying the mechanism of the proposed SARS-CoV-2 drugs and investigates their inhibitory effect. As comparator drugs, remdesivir and hydroxychloroquine were used. Cetilistat is one of the pancreatic lipase inhibitors used to treat obesity. It inhibits fat digestion and absorption [47]. Diiodohydroxyquinoline is a quinoline derivative that is employed to treat amoebiasis as a luminal amebicide [48]. Diiodohydroxyquinoline is poorly absorbed into the circulatory system, and cetilistat is rapidly hydrolyzed into its metabolites when bile is present [47,49]. Approximately 15–20% of COVID-19 patients have been reported to exhibit gastrointestinal symptoms, with some also displaying infectious virus particles and detectable viral RNA [50]. The results of research on hamsters infected with SARS-CoV-2 revealed that their intestines expressed high levels of viral nucleocapsid protein and inflammation, as well as detectable viral RNA [51]. As with SARS, feces may be an essential source of SARS-CoV-2 infection [52,53], oral diiodohydroxyquinoline and cetilistat may be topical luminal antivirals in reducing viral shedding in the gastrointestinal tract. Bexarotene and abiraterone acetate are non-chemotherapeutic antineoplastic drugs with limited proven immunosuppressive effects [22]. Bexarotene is a third-generation retinoid that effectively treats breast cancer, non-small cell lung cancer, and cutaneous T cell lymphoma [54–57]. Abiraterone acetate inhibited the androgen synthesizing enzyme CYP17A1 when combined with a corticosteroid. This procedure results in the treatment of refractory prostate cancer via androgen deprivation [58,59]. Yuan et al. reported that tamibarotene and AM580, both members of the same drug class as bexarotene, demonstrated remarkable antiviral activity against influenza viruses, Zika virus, coronaviruses (SARS-CoV and MERS-CoV), adenovirus, and enterovirus A71 [60]. The docked drugs with the SARS-CoV-2 main protease are depicted in Figs. S1–S6. The docking score of cetilistat, abiraterone, diiodohydroxyquinoline, bexarotene, remdesivir, and hydroxychloroquine with the SARS-CoV-2 main protease was calculated to determine their docking power on the crystal structure of the main protease. The binding free energy of bexarotene, remdesivir, abiraterone, cetilistat, diiodohydroxyquinoline, and hydroxychloroquine was $$\Delta G_{\text{binding}} = -8.10$$, $$-7.80$$, $$-7.00$$, $$-6.60$$, $$-5.30$$, and $$-5.00 \text{ kcal/mol}$$, respectively (Table 1).

Cetilistat, abiraterone, diiodohydroxyquinoline, and bexarotene had lower binding free energy than hydroxychloroquine, which had in-vitro activity against COVID-19. The negative free energy values indicated a strong affinity for the binding pocket. These findings revealed that bexarotene formed a more stable complex with the SARS-CoV-2 main protease than remdesivir, the first FDA-approved drug against COVID-19, indicated by its lower free energy. In other words, the SARS-CoV-2 main protease favors bexarotene to the well-known drug remdesivir. The docking energies, atoms involved in bonding with ligands, the nature of interactions, and bond lengths are shown in Tables S1–S6. It is evident that hydrophobic interactions play a role in forming the complex between bexarotene and cetilistat and the SARS-CoV-2 main protease. Meanwhile, hydrophobic and hydrogen-bonding interactions play a significant role in forming the complex between abiraterone and hydroxychloroquine and SARS-CoV-2 protease. Additionally, electrostatic, hydrophobic, and hydrogen bonding interactions are dominant in the formation of the SARS-CoV-2 protease-diiodohydroxyquinoline and SARS-CoV-2 protease-remdesivir complexes. The amino acid residues surrounding drugs-SARS-CoV-2 protease systems are represented in Tables S1–S6. As illustrated in Fig. S2, the main protease system of bexarotene-SARS-CoV-2 is surrounded by the amino acids PHE294, ILE249, PRO252, PRO293, and VAL297. Overall, this research indicates that bexarotene with the highest binding free energy may inhibit viral replication and infection at levels comparable to those found in FDA-approved and clinically trialed drugs.

As illustrated in Tables S7–S12 and Figs. 1–6, docking results for cetilistat, bexarotene, abiraterone, diiodohydroxyquinoline, remdesivir, and hydroxychloroquine with ACE2-RBD were obtained.

According to molecular docking studies, cetilistat, bexarotene, abiraterone, diiodohydroxyquinoline, remdesivir, and hydroxychloroquine do not interact with SARS-CoV-2 spike protein but do bind to the ACE2 receptor, preventing SARS-CoV-2 spike protein from binding to ACE2. Cetilistat, bexarotene, abiraterone, diiodohydroxyquinoline, and hydroxychloroquine had binding free energies of $$-8.70$$, $$-8.40$$, $$-8.10$$, $$-8.00$$, $$-6.70$$, and $$-5.30 \text{ kcal/mol}$$, respectively (Table 2).

Cetilistat, bexarotene, and abiraterone had a lower binding free energy than remdesivir and hydroxychloroquine. Additionally, diiodohydroxyquinoline had a lower binding free energy than hydroxychloroquine. These findings indicated that cetilistat, bexarotene, and abiraterone form a more stable complex with ACE2 than remdesivir and hydroxychloroquine, indicated by their lower binding free energy values.

Meanwhile, the diiodohydroxyquinoline-ACE2 complex formed is

| Drugs                      | $\Delta G$ (kcal/mol) |
|----------------------------|-----------------------|
| Abiraterone                | -7.00                 |
| Bexarotene                 | -8.10                 |
| Cetilistat                 | -6.60                 |
| Diiodohydroxyquinoline     | -5.30                 |
| Remdesivir                 | -7.80                 |
| Hydroxychloroquine         | -5.00                 |
more stable than the hydroxychloroquine-ACE2 complex. These findings strongly suggest that cetilistat, which had the lowest binding free energy, interacts efficiently with ACE2 receptors and disrupts SARS-CoV-2 attachment. Tables S7–S12 summarize the results of docking simulations, including the nature of interactions, the atoms involved in bonding with drugs, docking energies, and bond lengths. As can be seen, hydrophobic interactions play a role in the formation of the cetilistat-ACE2 system. Furthermore, one hydrogen bond was formed between the cetilistat-ACE2 system. SER47 and TRP349 amino acid residues surrounded the cetilistat-ACE2 system. Both hydrophobic and hydrogen bonding interactions were dominant in the formation of a complex between bexarotene and ACE2. The bexarotene-ACE2 system was surrounded by amino acids ALA348, HIS401, ARG393, PHE40, and PHE390. PHE504, TYR510, TYR127, and HIS505 were involved in the formation of hydrophobic interactions between abiraterone and ACE2 of SARS-CoV-2. The diiodohydroxyquinoline - ACE2 system resulted in the formation of one hydrogen and halogen bond. Moreover, the formation of the diiodohydroxyquinoline - ACE2 complex involved hydrophobic interactions. Five amino acids influenced the hydrophobic interactions between diiodohydroxyquinoline and ACE2: THR434, HIS540, PHE438

Fig. 1. Molecular docking perspective of abiraterone – (ACE2-RBD).

Fig. 2. Molecular docking perspective of bexarotene – (ACE2-RBD).
ILE291, and PRO415. By blocking the angiotensin-converting enzyme-2 receptor, the proposed FDA-approved drugs cetilistat, bexarotene, abiraterone, and diiodohydroxyquinoline may inhibit the attachment of SARS-CoV-2 to this receptor and thus neutralize viral entry into the host cell. As a result, these drugs may be promising candidates for inhibiting covid-19 infection at concentrations comparable to those found in FDA-approved and clinically tested drugs such as remdesivir and hydroxychloroquine. On the whole, these drugs interacted with both the main protease and the ACE2 receptor, indicating that they could be used as multitarget drugs against COVID-19, and their systemic effects should be further investigated in suitable ex vivo human organ culture or organoids, animal models, or clinical trials. The researchers hope that this study will aid in the development of potential SARS-CoV-2 vaccines and therapeutics.

3.2. Molecular dynamics simulations

3.2.1. Radius of gyration (Rg)

The radius of gyration (Rg) of both ACE2-cetilistat and protease-bexarotene complexes quantifies the molecule’s overall extension...
during a 30ns MD run (Fig. 7). A low Rg value demonstrates better structural entirety and folding treatment [61]. Throughout the 30ns MD simulation, two complexes maintain a stable mean Rg of 2.2 nm for 6LU7-Bex and 3.2 nm for 6LZG-Ceti. During simulation, a slight enhancement in the Rg value of the 6LZG-Ceti complex was observed, indicating its structural integrity. The MD simulation results entirely support that bexarotene and cetilistat form stable complexes with 6LU7 and 6LZG, indicating their inhibitory properties for the main protease and ACE2 receptor, respectively.

3.2.2. RMSD

RMSD analysis revealed insights and structural changes in the protein that confirm the protein’s stability and equilibrium during simulation. The RMSD plot of the backbone atoms in the 6LU7 complex with
RMSD was calculated for the 6LU7-Bex and 6LZG-Ceti structures that converged during the 30ns MD simulation. The results indicated that both structures stabilized after 1000ps. 6LU7-Bex averaged 0.37 Å°, while 6LZG-Ceti averaged 0.36 Å° throughout the 30ns simulation. The low RMSD values indicated that bexarotene and cetilistat were stable in MD simulations with proteins.

3.2.3. RMSF

RMSF analysis was used to determine the flexibility of the total protein concerning its average structure. Low RMSF values demonstrated narrowed movements, whereas high RMSF values demonstrated increased flexibility [62]. Ligand binding poses energy, and interaction has a direct correlation with residual fluctuation (RMSF) values. The RMSF plots for each residue in the 6LU7 complex with bexarotene and 6LZG complex with cetilistat are shown in Fig. 9. The RMSF values for the 6LU7-Bex and 6LZG-Ceti complexes were extremely low; as a result, they exhibited minimal movement, indicating that both complexes were stable. RMSF values were 0.29 and 0.28 Å° on average for 6LU7-Bex and 6LZG-Ceti during a 30 ns simulation. During the simulation, it was observed that residues in the loop region of 6LU7 were more fluctuated than those in the alpha-helix and beta-sheet regions. This indicated that the protein remained stable throughout the 30ns simulation period [31]. Except for the loop region, the residues S84 in alpha-helix and I31 in beta-sheet region of 6LZG exhibited significant variation up to 0.48 Å° and 0.45 Å°, respectively. Overall, the RMSF values for both proteins indicated that the complexes 6LU7-Bex and 6LZG-Ceti were stable [63].

3.3. MM-PBSA binding free energy

MmPbSaStat.py was used to calculate the average free binding energy of the 6LU7-Bex and 6LZG-Ceti (Table 3). The average free binding energy and the standard deviation/error of the files were calculated using the information obtained from g_mmpbsa. The binding free energy

Table 2

| Drugs                  | ΔG (KJ/mol) |
|------------------------|-------------|
| Abiraterone            | -8.10       |
| Bexarotene             | -8.40       |
| Cetilistat             | -8.70       |
| Diodohydroxyquinoline  | -6.70       |
| Remdesivir             | -8.00       |
| Hydroxychloroquine     | -5.30       |

bexarotene and the 6LZG complex with cetilistat is shown in Fig. 8. RMSD was calculated for the 6LU7-Bex and 6LZG-Ceti structures that converged during the 30ns MD simulation. The results indicated that both structures stabilized after 1000ps. 6LU7-Bex averaged 0.37 Å°, while 6LZG-Ceti averaged 0.36 Å° throughout the 30ns simulation. The low RMSD values indicated that bexarotene and cetilistat were stable in MD simulations with proteins.

Fig. 7. Radius of gyration (Rg) for (A) bexarotene – CoV-2 main protease and (B) cetilistat – (ACE2-RBD) during 30 ns MD simulation.

Fig. 8. RMSD plots for (A) bexarotene – CoV-2 main protease and (B) cetilistat – (ACE2-RBD) during 30 ns MD simulation.

Fig. 9. RMSF plots for (A) bexarotene – CoV-2 main protease and (B) cetilistat – (ACE2-RBD) during 30 ns MD simulation.
can evaluate the durability of the ligand-receptor interaction, which is a critical parameter in drug discovery. The lower the binding energy, the more effectively the ligand and protein bind [14]. Except for the polar solvation energy, the van der Waals, SASA, and electrostatic energies were used to bind 6LU7 and 6LZG to bexarotene and cetilistat. van der Waals energy augmentation to the overall binding free energy was greater than that of electrostatic contribution energy. The binding free energies of 6LU7-Bex and 6LZG-Ceti were determined to be $-63.40 \pm 11.70$ and $-95.92 \pm 12.04$ kJ/mol, respectively, indicating that the studied drugs interacted uniquely with the 6LU7 and 6LZG proteins.

Table 3

| System | $\Delta E_{\text{van der Waals}}$ (kJ/mol) | $\Delta E_{\text{electrostatic}}$ (kJ/mol) | $\Delta E_{\text{polar solvation}}$ (kJ/mol) | $\Delta E_{\text{SASA}}$ (kJ/mol) | $\Delta E_{\text{binding}}$ (kJ/mol) |
|--------|----------------------------------|----------------------------------|----------------------------------|----------------------------------|----------------------------------|
| 6LU7   | $-73.14 \pm 11.88$               | $32.02 \pm 10.40$               | $-63.40 \pm 11.70$               |                                 |                                  |
| Bex    | $11.10$                          | $17.51$                          | $21.65 \pm 1.45$                | $11.70$                          |                                  |
| 6LZG   | $-181.08 \pm 4.54$              | $110.38 \pm 20.98$             | $-95.92 \pm 10.40$             |                                 |                                  |
| Ceti   | $16.80$                          | $12.22$                          | $18.32 \pm 2.05$               | $12.04$                          |                                  |

4. Conclusion

The docking studies revealed that the binding free energies of FDA-approved drugs cetilistat, abiraterone, diiodohydroxyquinoline, and bexarotene are more potent than those of hydroxychloroquine, which has been used against COVID-19. In other words, these drugs formed more stable complexes with the main protease of SARS-CoV-2 and the ACE2 receptor. Additionally, cetilistat and bexarotene have a higher affinity for and interactions with the ACE2 receptor and the main SARS-CoV-2 proteases than remdesivir, the first FDA-approved drug target. The docking energy against time graphs for 6LU7-Bex and 6LZG-Ceti are depicted in Fig. S7. The findings above indicate that bexarotene and cetilistat are stable in the presence of proteins during MD simulation.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.imu.2021.100745.

References

1. Gorbalenya AE, Baker SC, Baric RS, de Groot RJ, Drosten C, Gulyaeva AA, et al. Severe acute respiratory syndrome-related coronavirus: the species and its viruses—a statement of the Coronavirus Study Group. bioRxiv 2020. https://doi.org/10.1101/2020.02.07.937862. In press.

2. Gautret P, Lagier J-C, Parola P, Hoang VT, Medlel L, Maillle M, et al. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. Int J Antimicrob Agents 2020;56:105949.

3. Chan JF, Lau SK, To KK, Cheng VC, Wong PC, Yuen KY. Middle East respiratory syndrome coronavirus: another zoonotic betacoronavirus causing SARS-like disease. Clin Microbiol Rev 2015;28:465–522.

4. Muradlilhara N, Sathivel R, Velumuragun D, Gromia MM. Computational studies of drug repurposing and synergism of lopinavir, oseltamivir and ritonavir binding with SARS-CoV-2 Protease against COVID-19. J Biomol Struct Dyn 2020;39:2607–14.

5. Khan SA, Zia K, Ashraf S, Uddin R, Ul-Haq Z. Identification of chymotrypsin-like protease inhibitors of SARS-CoV-2 via integrated computational approach. J Biomol Struct Dyn 2020;39:2607–14.

6. Elfyky AA. Ribavirin, Remdesivir, Sofosbuvir, Galidesivir, and Tenofovir against SARS-CoV-2 RNA dependent RNA polymerase (RdRp): a molecular docking study. Life Sci 2020;253:117592.

7. Gao J, Tian Z, Yang X. Breakthrough: chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies. Biosci. Trends 2020;14(1):72–3. https://doi.org/10.5582/bst.2020.01047. Epub 2020 Feb 19.

8. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. Cell Res 2020;30:269–71.

9. Khan SA, Zia K, Ashraf S, Uddin R, Ul-Haq Z. Identification of chymotrypsin-like protease inhibitors of SARS-CoV-2 via integrated computational approach. J Biomol Struct Dyn 2020;39:2607–14.

10. Yuan S, Chan JF, Chik KK, Chan CC, Tsang JO, Liang R, et al. Discovery of the FDA-approved drugs cetilistat, abiraterone, diiodohydroxyquinoline, and bexarotene as potential drugs for SARS-CoV-2 and ACE2 receptors.

Data availability

Because the majority of work is performed using licensed software, these are not transferable. Additional scripts written by members of the group may be made available upon request.

Code availability

Most of the work is done with licensed software therefore those are not transferable. Other scripts written in the group could be made available on request.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgment

The authors thank the Razi University Research Council for support of this work.
Mohammadi S, Heidarizadeh M, Entesari M, Esmailpour A, Esmailpour M, Darden T, York D, Pedersen L. Particle mesh Ewald: an N ∙ log (N) method for Ewald summation. Ophthalmochemistry 2016;12:3:1386–94.

Yao X, Ye F, Zhang M, Cai C, Huang R, Niu P, et al. In vitro antiviral activity and projection of optimized dosing design of hydroxychloroquine for the treatment of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Clin Infect Dis 2020;71:732–9.

Baldiya N, Ghosh NN, Chattopadhyay AP. Inhibitory activity of hydroxychloroquine on COVID-19 main protease: an insight from MD-simulation studies. J Mol Struct 2020;12:81599.

Beura S, Chetti P. In-silico strategies for probing chloroquine based inhibitors against SARS-CoV-2. J Biomol Struct Dyn 2020:11:28876–91.

Trott O, Olson AJ. AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. J Comput Chem 2010;31:455–64.

Singh S, Srivastava HK, Kishor G, Singh H, Agrawal P, Raghava GP. Evaluation of protein-ligand docking methods on peptide-ligand complexes for docking small ligands to peptides. BioXiv 2017:212514.

Prasanth D, Murahari M, Chandramohan V, Panda SP, Atmakuri LR, Guntupalli C. In silico identification of potential inhibitors from Cinnamomum against main protease and spike glycoprotein of SARS-CoV-2. J Biomol Struct Dyn 2020:1–15.

Berendsen HJ, van der Spoel D, van Drunen R. GROMACS: a message-passing parallel molecular dynamics implementation. Comput Phys Commun 1995:91:45–54.

Abraham MJ, Murtola T, Schulz R, Petersen MI, Hess B, Ervasti JM, et al. GROMACS: high performance molecular simulations through multi-level parallelism from laptops to supercomputers. SoftwareX 2015;1:19–25.

Bjelkmar P, Larsson P, Cuendet MA, Hess B, Lindahl E. Implementation of the CHARMM force field in GROMACS: analysis of protein stability effects from correction maps, virtual interaction sites, and water models. J Chem Theor Comput 2010;6:459–66.

Vanommeslaeghe K, Hatcher E, Acharya C, Kundu S, Zhong S, Shim J, et al. CHARMM general force field: a force field for drug-like molecules compatible with the CHARMM all-atom additive biological force fields. J Comput Chem 2010;31:671–90.

Zoete V, Cuendet MA, Grosdidier A, Michielin O. SwissParam: a fast force field performance molecular simulations through multi-level parallelism from laptops to supercomputers. SoftwareX 2015;1:19–25.

Prasanth D, Murahari M, Chandramohan V, Panda SP, Atmakuri LR, Guntupalli C. In silico identification of potential inhibitors from Cinnamomum against main protease and spike glycoprotein of SARS-CoV-2. J Biomol Struct Dyn 2020:1–15.

Berendsen HJ, van der Spoel D, van Drunen R. GROMACS: a message-passing parallel molecular dynamics implementation. Comput Phys Commun 1995:91:45–54.

Abraham MJ, Murtola T, Schulz R, Petersen MI, Hess B, Ervasti JM, et al. GROMACS: high performance molecular simulations through multi-level parallelism from laptops to supercomputers. SoftwareX 2015;1:19–25.

Bjelkmar P, Larsson P, Cuendet MA, Hess B, Lindahl E. Implementation of the CHARMM force field in GROMACS: analysis of protein stability effects from correction maps, virtual interaction sites, and water models. J Chem Theor Comput 2010;6:459–66.

Vanommeslaeghe K, Hatcher E, Acharya C, Kundu S, Zhong S, Shim J, et al. CHARMM general force field: a force field for drug-like molecules compatible with the CHARMM all-atom additive biological force fields. J Comput Chem 2010;31:671–90.

Zoete V, Cuendet MA, Grosdidier A, Michielin O. SwissParam: a fast force field performance molecular simulations through multi-level parallelism from laptops to supercomputers. SoftwareX 2015;1:19–25.

Darden T, York D, Pedersen L. Particle mesh Ewald: an N ∙ log (N) method for Ewald summation. J Chem Phys 1987;89:926–35.

Darden T, York D, Pedersen L. Particle mesh Ewald: an N ∙ log (N) method for Ewald summation in large systems. J. Chem. Phys. 1993;98:10089–92.

Kumar R, Kumar R, Consortium OSDD, Lynn A, g.mpbsa. – A GROMACS tool for high-throughput MM-PBSA calculations. J Chem Inf Model 2014;54:1951–62.

Vardhan S, Sahoo SK. In silico ADMET and molecular docking study on searching potential inhibitors from limonoids and triterpenoids for COVID-19. Comput Biol Med 2020;124:103936.

García-Fernández R, Ziegelmüller P, González L, Mansur M, Machado Y, Redecke L, et al. Two variants of the major serine protease inhibitor from the sea anemone Stichodactyla helianthus, expressed in Pichia pastoris. Protein Expr. Purif. 2016;123:42–50.

Verma D, Kapoor S, Das S, Thakur K. Potential inhibitors of SARS-CoV-2 main protease (M pro) identified from the library of FDA approved drugs using molecular docking studies. 2020.

Mohammadi S, Heidarizadeh M, Entesari M, Esmailpour A, Esmailpour M, Moradi R, et al. In silico investigation on the inhibiting role of nicotine/caffeine by blocking the S protein of SARS-CoV-2 versus ACE2 receptor. Microorganisms 2020;8:1600.

Nguyen HL, Thai NQ, Truong DT, Li MS. Remdesivir strongly binds to both RNA-dependent RNA polymerase and main protease of SARS-CoV-2: evidence from MD-simulation studies. J Phys Chem B 2020;124:1337–46.

Marmor MF, Kellner U, Lai TY, Melles RB, Mieler WF. Recommendations on screening for chloroquine and hydroxychloroquine retinopathy (2016 revision). Ophthalmology 2016;123:1386–94.

Yao X, Ye F, Zhang M, Cai C, Huang R, Niu P, et al. In vitro antiviral activity and projection of optimized dosing design of hydroxychloroquine for the treatment of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Clin Infect Dis 2020;71:732–9.

Padwal R. Getlistat, a new lipase inhibitor for the treatment of obesity. Curr Opin Drugs 2008;9:414–21.

Cheung KS, Hung IF, Chan PP, Lung K, Tso E, Liu R, et al. Gastrointestinal manifestations of SARS-CoV-2 infection and virus load in fecal samples from the Hong Kong cohort and systematic review and meta-analysis. Gastroenterology 2020;159:81–95.

Chan JF-W, Zhang AJ, Yuan S, Poon VK-M, Chan CC-S, Lee AC-Y, et al. Simulation of the clinical and pathological manifestations of Coronavirus Disease 2019 (COVID-19) in golden Syrian hamster model: implications for disease pathogenesis and transmissibility. Clin Infect Dis 2020;71:2428–46.

Cheng VC, Lau SK, Woo PC, Yuen KY. Severe acute respiratory syndrome coronavirus as an agent of emerging and reemerging infection. Clin Microbiol Rev 2007;20:660–94.

Huang I, Cheng V, Wu A, Tang B, Chan K, Chu C, et al. Viral loads in clinical specimens and SARS manifestations. Emerg Infect Dis 2004;10:1550.

Phillip MM, Zhang L, Zhi L, McGurp MB, Berger E, Wagenen M, et al. Design and synthesis of potent retinoid X receptor selective ligands that induce apoptosis in leukemia cells. J Med Chem 1995;38:3146–55.

Gneideck R, Ansal C, Bagot M, Dummer R, Ducvic M, Knobler R, et al. The optimal use of hexabronate in cutaneous T-cell lymphoma. Br J Dermatol 2007;157:433–40.

Dragnev KH, Petty WJ, Shah SJ, Lewis LD, Black CC, Memoli V, et al. A proof-of-principle clinical trial of hexabronate in patients with non-small cell lung cancer. Clin Cancer Res 2002:7:1794–800.

Esteva FJ, Glaspie J, Zaidas N, Laufman L, Hutchins L, Dickler M, et al. Multicenter phase II study of oral hexabronate for patients with metastatic breast cancer. J Clin Oncol 2003;21:999–1006.

Duc I, Boffet P, Duranti V, Cardinelli S, Riviere A, De Giovanni A, et al. In vitro and in vivo models for the evaluation of potent inhibitors of male rat 17α-hydroxylase/C17,20-lyase. J Steroid Biochem Mol Biol 2003;84:537–42.

Arasarathnam M, Crumbaker M, Bhatnagar A, McKay MJ, Molloy MP, Gurney H. Inter-and intra-patient variability in pharmacokinetics of abiraterone acetate in metastatic prostate cancer. Canc. Chemotherapy. Pharmacol 2019;84:139–46.

Yuan S, Chu H, Chan JF-W, Ye Z-W, Wen L, Yan B, et al. SREBP-dependent lipidomic reprogramming as a broad-spectrum antiviral target. Nat Commun 2019;10:1–5.

Erva RR, Rajputaliya SB, Durtih CP, Bhatia M, Pola M. Molecular dynamic simulations of Escherichia coli L-asparaginase to illuminate its role in deamination of asparagine and glutamine residues. J Biotech 2016;62.

Priya R, Samitha R, Doss CGP, Rajasekaran G, Babu S, Sreemavan R, et al. Molecular docking and molecular dynamics to identify a novel human immunodeficiency virus inhibitor from alkaloids of Toddalia asiatica 2015;11: S414.

Basti A, Ali T, Rehman SI. Truncated human angiotensin converting enzyme 2: a potential inhibitor of SARS-CoV-2 spike glycoprotein and potent COVID-19 therapeutic agent. J Biomol Struct Dyn 2021;39(10):3605–14. https://doi.org/10.1080/07391102.2020.1764150.