K-ras mutations and HLA-DR expression in large bowel adenomas

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Summary A total of 72 sporadic colorectal adenomas in 56 patients were studied for the presence of point mutations in codons 12 and 13 of the K-ras gene and for HLA-DR antigen expression related to clinicopathological variables. Forty K-ras mutations in 39 adenomas were found (54%): 31 (77%) in codon 12 and nine (23%) in codon 13. There was a strong relationship between the incidence of K-ras mutations and adenoma type, degree of dysplasia and sex. The highest frequency of K-ras mutations was seen in large adenomas of the villous type with high-grade dysplasia. Fourteen out of 15 adenomas obtained from 14 women above 65 years of age carried mutations. HLA-DR positivity was found in 38% of the adenomas, large tumours and those with high-grade dysplasia having the strongest staining. Coexpression of K-ras mutations and HLA-DR was found significantly more frequently in large and highly dysplastic adenomas, although two-way analysis of variance showing size and grade of dysplasia to be the most important variable. None of the adenomas with low-grade dysplasia showed both K-ras mutation and HLA-DR positivity (P=0.004). K-ras mutation is recognised as an early event in colorectal carcinogenesis. The mutation might give rise to peptides that may be presented on the tumour cell surface by class II molecules, and thereby induce immune responses against neoplastic cells.

Keywords: K-ras; HLA-DR; large bowel adenoma; dysplasia; polymerase chain reaction-restriction fragment length polymorphism; immunohistochemistry

The adenoma–carcinoma sequence has for several years been accepted as the main pathway for the development of most neoplasias in colorectal carcinogenesis (Muto et al., 1975). This is thought to be a multistep process involving genetic instability (Nowell, 1976), clonal expansion (Nowell, 1976), and acquisition of malignant phenotype (Vogelstein et al., 1988). Several abnormal genes have been found, with mutations within the ras gene family being among the first to appear (Bos et al., 1987; Forrester et al., 1987). Mutations of K-ras, located on the short arm of chromosome 12, are frequent in colorectal neoplasms. In adenocarcinomas mutations in codons 12 and 13 of K-ras have been identified in 39–65% of tumours (Bos et al., 1987; Vogelstein et al., 1988; Burmer and Loeb, 1989; Breivik et al., 1994), and 80–90% of them are present in codon 12 (Bos et al., 1987; Vogelstein et al., 1988; Breivik et al., 1994).

Vogelstein et al. (1988) found ras mutations in 47% of carcinomas, in 58% of adenomas greater than 1 cm, and in less than 10% of adenomas 1 cm or smaller, indicating that K-ras mutations are a relatively early event in colorectal carcinogenesis. Miyaki et al. (1990) demonstrated that K-ras mutations are frequently detected in adenomas with severe dysplasia.

Whereas K-ras mutations in familial adenomatous polyposis (FAP) adenomas are found with an overall frequency of about 18%, sporadic adenomas showed higher values, but with considerable variation in frequency, ranging from 15% to 75% (reviewed by McLellan et al., 1993). Furthermore, evidence for very early involvement of ras mutations in up to 70% of aberrant crypt foci of the colon has been provided (Pretlow et al., 1993; Yamashita et al., 1995).

The expression of HLA-antigens has been studied in a relatively small series of lesions, both in sporadic and FAP

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Histopathological diagnosis was based upon 1–6 sections from each adenoma, 110 sections altogether. The tumours were categorized according to size, histological type and grade of dysplasia (Table II).

Epithelial dysplasia was graded as low grade and high grade by histopathological criteria according to the WHO classification, in which low grade includes mild and moderate dysplasia, and high grade corresponds to the former severe dysplasia (Jass and Sobin, 1989). Histopathological evaluations were done by two independent observers (SNA and TOR).

Tumour material

The mucosal specimens were placed in 0°C, isotonic phosphate-buffered saline (PBS), pH 7.6, immediately after resection. Within 1 h samples for immunohistochemistry were excised and placed in 96% ethanol at 4°C and further processed for paraffin embedding (Brandtzaeg, 1974). Serial sections cut at 6 μm were dewaxed and subjected to immunofluorescence staining at room temperature. One section from each series was stained by a trichrome routine method (HAS) containing haematoxylin, azofloxin and saffron. The next sections from each paraffin block were cut at 20 μm for analysis of K-ras mutations in codons 12 and 13. The blade of the microtome was changed and thoroughly cleaned twice in xylol and once in ethanol after each sample to avoid cross-contamination.

Investigations of K-ras mutations

Enriched PCR amplification K-ras mutations in codons 12 and 13 were identified by a sensitive technique (modified from Kahn et al., 1991), which will be outlined in detail elsewhere (J. Breivik et al., in preparation). Sections 20 μm-thick from the ethanol-fixed, paraffin-embedded tissue blocks were used directly as templates in the initial polymerase chain reaction (PCR) amplification (PCR A1). The material was put into 500 μl tubes with 50 μl of PCR amplification buffer (50 mm potassium chloride, 10 mM Tris HCl, 1.5 mM magnesium chloride), 25 μM of each deoxynucleoside triphosphate, 0.28 μg Taq Start Antibody (Clontech, Palo Alto, CA, USA), 1.25 U of Taq DNA polymerase (Promega, Madison, WI, USA) and 0.04 μM and 0.2 μM of the oligonucleotide primers (Genosys Biotechnology, TX, USA) 5K1 and 3K1 respectively:

5′-CCANNNNNNNTGG-3′

The 5K1 primer was designed (modified, mismatching bases are underlined) to induce two restriction sites in the wild-type (WT) amplification product (N = any base):

5′-CCANNNNNNNNNTGG-3′

is recognised by endonuclease BstXI, and involves WT codon 12.

5′-CCANNNNNNNNNTGG-3′

is recognised by endonuclease XcmI, and involves WT codon 13.

The amplification was performed in a GeneAmp PCR System 9600 (Perkin-Elmer, CT, USA). The reaction was run for ten cycles consisting of 30 s denaturation at 94°C, 1 min annealing at 50°C, and 2 min elongation at 72°C. After the last cycle the tubes were kept at the elongation temperature for 8 min.

An aliquot of 10 μl of the amplification product was incubated with either 10 U of BstXI or 2 U of XcmI. Digestion was performed as recommended by the supplier of the endonucleases (both New England Biolabs, MA, USA), in a volume of 20 μl.

An aliquot of 5 μl of the digested amplification product was used directly as a template in ten cycles of a second amplification reaction identical to PCR A1, designated PCR A2. A 5 μl aliquot of this amplification product was digested with the same endonuclease and under identical conditions as previously described.

Once again 5 μl of the digested amplification product was used directly as a template in an amplification reaction (PCR B). This time 5K1 was used in combination with 0.2 μM of a nested and modified 3′ primer, amplifying a segment of 152 bp:

3K2: 5′-GGATGGTCCTCACCAGTAATATGGATATTA-3′

3K2 contains both the restriction sites included in WT amplification products by 5K1. A total of 30–40 cycles were performed depending on the yield of the amplification. Reaction conditions were as described previously.

| Table I | Clinicopathological characteristics of the patients |
|---------|--------------------------------------------------|
| Number of patients | 56 | 4 | 9 |
| Sex | Male | 30 | 3 | 4 |
| | Female | 26 | 1 | 5 |
| Median age (years) | 65 | 32 | 59 |
| Range | 28–92 | 20–46 | 36–77 |
| Number of polyps | 72 | 12 | 15 |
| Histological type | Tubular | 30 | 7 | 11 |
| | Tubulovillous | 24 | 1 | 2 |
| | Villous | 18 | 0 | 0 |
| Grade of dysplasia | Low grade | 32 | 7 | 12 |
| | High grade | 38 | 6 | 6 |
| Localization | Right colon | 22 | 3 | 8 |
| | Left colon* | 24 | 3 | 8 |
| | Rectum | 26 | 2 | 2 |
| Size | < 10 mm | 24 | 16 | 12 |
| | 10–20 mm | 16 | 3 | 3 |
| | > 20 mm | 32 | 1 | 0 |

* Descending and sigmoid.

Table II | Samples of sporadic adenomas subjected to K-ras mutation analysis and HLA-DR study |
|----------------|--------------------------------------------------|
| Grade of Dysplasia | Low grade (n = 34) | High grade (n = 38) |
| Size | < 10 mm | 15 | 2 | 17 |
| | 10–20 mm | 3 | 5 | 6 |
| | > 20 mm | 1 | 4 | 2 |
| Total number | 22 | 8 | 14 |

T, tubular; TV, tubulovillous; V, villous.
**DNA sequencing**

For all mutation-positive samples the product of PCR B was sequenced in order to identify the exact base substitution. Direct sequencing was done by a modified dideoxy chain termination method (Sanger et al., 1977), using the Sequenase PCR Product Sequencing Kit (United States Biochemical, Cleveland, OH, USA), under conditions recommended by the manufacturer. A nested sequencing primer was used:

3K3: 5'-TATTTACAAAAGGTTAC-3'

**Controls**

DNA from colon carcinoma cell line SW480 (Clontech, Palo Alto, CA, USA) and HCT116 (American Type Culture Collection, ATCC, Rockville, MD, USA) with known K-ras mutations at codon 12 (GTT) and codon 13 (GAC) respectively, were used as positive controls in each of the parallel procedures. Negative controls, without DNA, were run as controls of contamination. All mutation detection experiments were confirmed using a second independent amplification product.

**HLA-DR immunohistochemistry**

A murine monoclonal antibody to a non-polymorphic human HLA-DR antigen, clone L234 (IgG2a, Beckton Dickinson, Mountain View, CA, USA) was applied (1:20 for 20 h) in an indirect three-step immunofluorescence method (Brandtzaeg and Rognum, 1983), including affinity purified biotinylated horse anti-mouse IgG (0.05 g IgG1-1, 3 h) and fluorescein isothiocyanate (FITC)-labelled avidin (0.05 g 1-1, 30 min), both purchased from Vector Laboratories (Burlingame, CA, USA).

Two or more samples were examined from most of the larger adenomas. Thirty-nine of the sections showed varying amounts of normal adjacent mucosa. Embedded biopsies from five normal persons were included, as well as tissue from five resections owing to colorectal carcinoma, taken at least 10 cm away from the tumour.

Observations were done in a Leitz Aristoplan fluorescence microscope equipped with an Osram Hg 100 W lamp for fluorescence (green emission). Narrowband excitation and selective filtration (Leica I3 513683) of the fluorescence colour were obtained with a Ploem-type epi-illuminator.

**Evaluation**

The epithelial staining for HLA-DR antigens in adenomatous mucosa was evaluated according to the percentage of positive cells; 0, 1–5%, 5–25%, >25% respectively. Epithelial staining in adjacent normal mucosa was also noticed. The same investigator (SNA) was responsible for the fluorescence scoring throughout the study. Reproducibility studies of a similar fluorescence scoring system have been published previously (Norheim Andersen et al., 1993).

Blind tests for intra- and interobserver reproducibility of histological type and grade of epithelial dysplasia of the adenomas were performed, and proportions of agreement (P) and Kappa values were estimated in the analysis of observer variability (Landis and Koch, 1977).

**Statistical analysis**

The $\chi^2$ test was used for statistical evaluation to relate the size of polyps, type, degree of dysplasia, site and sex to K-ras mutations and HLA-DR antigen expression. When the numbers were small, Yates’ correction was used. When relating K-ras mutations, HLA-DR expression and classical variables, two-way analysis of variance was used. P-values less than 0.05 were regarded as statistically significant.

**Results**

**Frequency of K-ras mutations**

Forty mutations were found in 39 of 72 sporadic adenomas (54%), 31 (77%) in codons 12 and 9 (23%) in codon 13. One small adenoma (<1 cm) with high-grade dysplasia contained both a codon 12 (Val) and a codon 13 (Asp) mutation. In another small adenoma with high-grade dysplasia it was technically impossible to amplify DNA for K-ras mutation analysis.

The frequency of K-ras mutations tended to increase with size ($P=0.07$), was significantly higher in adenomas with high-grade dysplasia compared with low-grade ($P=0.001$), and was related to type of adenomas with higher frequency in villous/tubulovillous tumours than in tubular ($P<0.001$) (Table I).

Location at the right side vs left side (descending, sigmoid and rectum) of the large bowel did not show any significant differences. Significantly more women than men showed K-ras mutation ($P=0.001$) (Figure 1). Among the 15 adenomas from 14 older women only one adenoma, <10 mm and with low-grade dysplasia, did not show a point mutation. The same patient, however, had a tubulovillous adenoma between 10 and 20 mm in diameter and with low-grade dysplasia, containing a codon 12 GTT (Val) mutation.

The most frequent mutations were G→A transitions (20/40) and G→T transversions (17/40). Only three G→C transversions were observed. Mutations in position 2 of a codon occurred three times more frequently than in position 1 (75% vs 25%), (Figures 2 and 3).

Two patients had two separate adenomas containing a K-ras mutation. One elderly woman 78 years of age showed codon 12 G→A transition (Asp) in one adenoma and G→T transversion (Val) in another. A man aged 61 years had a codon 13 G→A transition (Asp) in both adenomas.

Among the 12 adenomas from FAP patients only one showed a K-ras mutation, found in codon 12 (Asp) in a tubulovillous adenoma greater than 20 mm and with high-grade dysplasia. This gives a mutation frequency of 8%, which is significantly less frequent than in sporadic ones ($P<0.01$).

None of the hyperplastic polyps showed any K-ras mutation.

**Table III** K-ras mutation in sporadic adenomas according to clinicopathological variables

| Size (mm) | K-ras+ (%) | K-ras− (%) | Level of significance |
|----------|------------|------------|----------------------|
| <10      | 11 (15)    | 13 (18)    | $P=0.07$             |
| 10–20    | 6 (8)      | 10 (14)    |                      |
| >20      | 22 (31)    | 10 (14)    |                      |

| Type     |            |            |                      |
|----------|------------|------------|----------------------|
| Tubular  | 8 (11)     | 22 (31)    |                      |
| Tubulovillous | 15 (21) | 9 (13) | $P<0.001$             |
| Villous  | 16 (22)    | 2 (3)      |                      |

**Dysplasia**

|            | 11 (15)    | 23 (32)    |                      |
| Low grade  |            |            |                      |
| High grade | 28 (39)    | 10 (14)    | $P=0.001$             |

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**HLA-DR expression**

Mononuclear cells in lamina propria were strongly positive and served as internal controls in each section.

Thirty-nine of the 72 sporadic adenomas showed epithelial staining for HLA-DR to a varying extent (54%) (Figure 4). In 14 adenomas (19%) more than 25% of the neoplastic cells were positive for HLA-DR, whereas 13 adenomas (18%)

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**Figure 1** Distribution of adenomas and mutations. O, No mutation; △, G→A transition; ●, G→T transversion; ▲, G→C transversion; *, mutations in both codons 12 and 13. Patients were divided by sex and by whether younger or older than median age (65 years). Percentage gives frequency of adenomas with mutation in each group. Significantly more women than men had K-ras mutations in their adenomas (P<0.001), and women above 65 years of age had significantly more K-ras than those below (P<0.001).

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**Figure 2** Frequency of the different base substitutions among adenomas with mutation. The number of lesions is indicated above the columns, and the base substitution and corresponding amino acid are marked underneath.

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**Figure 3** K-ras sequences from four adenomas showing mutations. (a) 12 Val (GTT). (b) 12 Asp (GAT). (c) 12 Arg (CGT). (d) 13 Asp (GAC). (G band in 3 B due to wild-type sequence.)

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**Figure 4** Villous adenoma > 2 cm with high-grade dysplasia. (a) HAS staining (original magnification x 130). Detail of epithelium with high-grade dysplasia to the right (original magnification x 520). (b) Adjacent section stained green for HLA-DR determinants (original magnification x 130).
Table IV  HLA-DR staining in sporadic adenomas according to clinicopathological variables

| Size (mm) | Area of HLA-DR positivity (%) | Level of significance |
|-----------|--------------------------------|-----------------------|
|           | 0% (n) | 1–5% (n) | 5–25% (n) | >25% (n) | |
| <10       | 17 (24) | 2 (3)    | 4 (6)    | 1 (1)    | P=0.013 |
| 10–20     | 8 (11)  | 3 (4)    | 4 (6)    | 1 (1)    |           |
| >20       | 8 (11)  | 7 (10)   | 5 (7)    | 12 (17)  |           |
| Type      |         |          |          |          |           |
| Tubular   | 15 (21) | 7 (10)   | 4 (6)    | 4 (6)    |           |
| Tubulovillous | 9 (13) | 4 (6)    | 5 (7)    | 6 (8)    | NS       |
| Villous   | 9 (13)  | 1 (1)    | 4 (6)    | 4 (6)    |           |
| Dysplasia |         |          |          |          |           |
| Low grade | 23 (32) | 4 (6)    | 3 (4)    | 4 (6)    |           |
| High grade| 10 (14) | 8 (11)   | 10 (14)  | 10 (14)  | P=0.018  |

*a The groups with no staining and minimal staining (1–5%) were pooled.

showed HLA-DR staining in 5–25% of the cells. Scattered staining in 1–5% of the cells was seen in 12 adenomas (17%), and these were pooled with the totally negative adenomas.

The expression of HLA-DR increased with size of polyps (Table IV). Significantly more adenomas measuring 20 mm or more were positive for HLA-DR, compared with smaller polyps (P=0.013). The expression of HLA-DR also increased with grade of dysplasia (P=0.018). Among 34 adenomas with low-grade dysplasia, 31% were definitely positive for HLA-DR, whereas 74% of the 38 adenomas with high-grade dysplasia showed positive staining in >5% of the epithelial cells.

The staining was always heterogeneous. Areas with high-grade dysplasia often stained positive, but not invariably.

There was no relation between HLA-DR expression and histological type of adenomas (Table IV, NS).

Adenomas from the right side of the colon showed significantly more HLA-DR positivity than left-sided ones (P=0.003), whereas there were no statistically significant differences in staining according to sex (P=0.25).
In the 12 adenomas from FAP patients, only one (8%) – with high-grade dysplasia and measuring between 10 and 20 mm – was strongly positive (>25%) for HLA-DR. The rest showed either scattered staining in <5% of the cells or no staining at all, which is not significantly different from sporadic adenomas (P = 0.12).

The ten sections with normal control mucosa and the 15 hyperplastic polyps investigated did not express HLA-DR antigens in the epithelial cells, but mononuclear cells in lamina propria were also strongly positive here (Figures 5 and 6).

Furthermore, sections from 39 of a total of 84 adenomas included adjacent mucosa. Thirty-seven of these had histologically normal mucosa, being without evidence of epithelial HLA-DR expression. In the other two adenomas, both above 2 cm, with high-grade dysplasia, and showing strong HLA-DR staining, the epithelial cells in the adjacent mucosa were weakly DR-positive. There were, however, more inflammatory cells than normal in the lamina propria.

Intraobserver reproducibility was 'substantial', both with respect to grade of dysplasia (kappa = 0.74) and histological type (kappa = 0.63) of the adenomas (Figure 7a–d). Intraobserver reproducibility showed 'substantial' agreement regarding histological type (kappa = 0.73), and was 'almost perfect' (kappa = 0.83) when evaluating grade of dysplasia, the second observations being performed blindly 12 weeks after the first.

**K-ras mutations and HLA-DR expression**

K-ras mutations were present in about half of the adenomas, both the HLA-DR-positive and the HLA-DR-negative ones. However, when dividing the adenomas into subgroups, the picture became more complex. Among adenomas with high-grade dysplasia and K-ras mutation, the majority also expressed HLA-DR, whereas none of the low-grade adenomas were both K-ras and HLA-DR positive (P = 0.004) (Figure 8). When using two-way analysis of variance the relationship between K-ras and HLA-DR is partly explained by size (P = 0.02, Figure 9, Table V) and grade of dysplasia (P = 0.03, Figure 8, Table V), but not by type (P = 0.55, Figure 10, Table V).

Nevertheless, the traditional markers of increasing malignant potential, adenoma size and grade of dysplasia, are the most important characteristics for both K-ras and HLA-DR separately.

**Discussion**

In the present study the polyps were tested using a new, highly sensitive technique allowing detection of one mutated K-ras codon 12 and 13 allele in the presence of $10^5$–$10^6$ copies of the wild-type allele. Since relatively few tumours are reported to contain mutations in K-ras codon 61 or in N-ras, these mutations were not examined.

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**Figure 7** Scatter diagrams of interobserver and intraobserver reproducibility tests. The degree of accordance for histological grade (a) between the first observer (SNA) and the second one (TOR) (P = 0.87 (62/71), kappa = 0.74) and (b) between two separate observations by the same observer (SNA) (P = 0.92 (77/84), kappa = 0.83). The degree of accordance for histological type (c) between the two observers (P = 0.78 (55/71), kappa = 0.63) and (d) between two separate observations by the same observer (SNA) (P = 0.83 (70/84), kappa = 0.73).

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We showed that 39 of the 72 sporadic adenomas contained a K-ras mutation. Only one of the 12 FAP adenomas was positive (8%). This is in general agreement with previous studies from the USA, reporting an incidence of 50% (Vogelstein et al., 1988) to 75% (Burmer and Loeb, 1989) K-ras mutations in sporadic adenomas. In FAP patients the incidence is lower, from 7% to 30% (Farr et al., 1988; Miyaki et al., 1990; Sasaki et al., 1990; Ando et al., 1991, 1992). This suggests that FAP adenomas follow another pathway in their tumorigenesis.

As in other investigations, most of the mutations were found in codon 12 (78%). Corresponding to studies from the USA, we detected mutations encoding aspartic acid and valine (17 and 12 tumours respectively) being most common. In contrast McLellan et al. (1993) found replacement of glycine by valine and arginine to be most frequent in British adenoma patients.

In an investigation of 251 colorectal carcinomas in a Norwegian population, Breivik et al. (1994) found G→A transitions in 57% and G→T transversions in 34%. This is in agreement with the figures in the present study. In both samples mutations in position 2 occurred three times more frequently than in position 1, with figures being identical (76% vs 24%). This might reflect the mutation profile in this ethnic population. Breivik et al. (1994) detected K-ras codon 12 and codon 13 mutations in 39% of the carcinomas with a method based on PCR amplification, blotting onto nylon
membranes and sequence-specific oligonucleotide hybridisation. These carcinomas are now being investigated, using this new technique (in preparation).

Most previous studies conclude that no K-ras mutations are found in normal mucosa (Forrester et al., 1987; Soh et al., 1993) except one in which two cases of codon 12 point mutations in normal mucosa immediately adjacent to carcinomas were found (Burmer and Loeb, 1989). In all reports the frequency of K-ras mutations is lower in colorectal carcinomas than in adenomas, but the figures vary somewhat with the sensitivity of the methods used.

We found that the frequency of K-ras mutations tended to increase with increasing size (P = 0.07), and were significantly more often found in high-grade epithelial dysplasia compared with low grade (Table III). Vogelstein et al. (1988) reported that the frequency of K-ras mutations was related to size, and Scott et al. (1993) found more codon 12 mutations with increasing size and grade of dysplasia. Furthermore, we confirmed that tubulovillous and villous adenomas had significantly higher mutation frequencies than tubular adenomas (Vogelstein et al., 1988). These observations are in contrast to McLellan et al. (1993), who did not find any correlation with size or with any other variables, except for a personal history of colorectal cancer. In the present study four patients with sporadic adenomas had a synchronous adenocarcinoma, but only one of them had a K-ras mutation in an adenoma. Two of the FAP patients had a synchronous carcinoma, but no K-ras mutation in the five adenomas investigated. Our sample is, however, too small to elucidate the question further.

One of the most surprising findings was the relation between K-ras mutation and sex. All female patients above 65 years of age showed adenomas with mutation, as compared with very few patients among elderly men (24%). Breivik et al. (1994) found a strikingly low frequency of K-ras mutations in colonic carcinomas in younger male patients compared with elderly women (more than 70 years). Perhaps the high frequency of mutation in females might be related to sex differences in faecal composition, bowel transit time and bile components.

Distinct expression of HLA-DR was present in 38% of the adenomas and increased with size and grade of epithelial dysplasia, but was not related to histological type. However, the amount of villous components is often a consequence of the size of the adenoma. Previous studies (Gutierrez et al., 1987, 1990; and Horie et al., 1990) have shown a correlation between HLA-DR expression and grade of dysplasia, the former study also reporting higher HLA-DR expression in villous over tubular adenomas.

Although some groups claim to find expression of HLA-DR in normal colorectal epithelium in humans (McDougall et al., 1990; Tsoulias et al., 1992; Mayer and Shlien, 1987), most investigations do not (Daar et al., 1982, 1984; Ghosh et al., 1986; Momburg et al., 1986; Gutierrez et al., 1987; Degener et al., 1988; Ruiz-Cabello et al., 1988; Norazmi et al., 1989; Bedossa et al., 1990; Horie et al., 1990). The discrepancy may be explained by the selection of normal controls, as slight inflammatory changes are known to induce DR-expression (Rognum et al., 1987). Another possibility may be different technical procedures (Mayer and Shlien, 1987), or binding of avidin to mucus in goblet cells.

In pathological conditions, such as inflammation, dysplasia and carcinoma, the epithelium may express varying amounts of these antigens (Rognum et al., 1983, 1987; Ruiz-Cabello et al., 1988). The presence of HLA-DR on the surface membrane of the neoplastic cells is a prerequisite for recognition by activated CD4+ T cells, and it has been shown that the growth of colorectal cancer cells expressing class II following γ-interferon treatment is inhibited by activated CD4+ T-helper cells (Gedde-Dahl et al., 1991), indicating a direct cytotoxic effect (Gjertsen et al., personal communication). The activation of such T cells requires the presence of accessory molecules not usually present on tumour cells. It is therefore believed that specific T cells will be activated primarily by dendritic cells present in the tumour or in the draining lymph nodes. Such T cells will home to the tumour where release of cytokines, such as tumour necrosis factor-α and γ-interferon, may induce class II molecules in the tumour.

The hyperplastic polyps showed neither HLA-DR expression nor K-ras mutation. In agreement with the present study, Bedossa et al. (1990) found no expression of HLA-DR in the epithelium of 15 hyperplastic polyps, in contrast to 8 of 15 adenomas, which showed patchy positivity. Jen et al. (1994), however, found K-ras mutations in 5 of 22 hyperplastic polyps with diameter less than 10 mm. Our observation gives support to the view that hyperplastic polyps might be regarded as harmless lesions.

The coexpression of K-ras and HLA-DR in adenomas that have the most malignant potential, i.e. the largest and those with high-grade dysplasia, may be of interest, although the two-way analysis of variance shows that size and grade of dysplasia are of greatest importance for both K-ras mutation and HLA-DR positivity, separately. The dissociation of HLA-DR positivity and K-ras mutation in right-sided adenomas adds to the complexity of the relationship between the markers. Nevertheless, co-expression of K-ras and DR might be interpreted as a marker of carcinomatous development.

Previous studies (Jung and Schluesener, 1991; Gedde-Dahl et al., 1991; Shlien et al., 1991) have shown that a variety of different HLA class II molecules can bind present ras peptides to T cells. Fossum et al. (1994) described a specific immune response against a synthetic peptide carrying the 13Asp mutation in p21 ras in patient with colonic carcinoma, and the responding T cells cloned from the peripheral blood of the patient were of both CD4 and CD8 phenotype. The corresponding mutation was not detected in the cancer, and the authors therefore speculated that a specific T cell response resulted in eradication of the tumour cells harbouring the 13Gly→Asp mutation. It was observed that the DR7 molecule, which was able to present the 13Gly→Asp mutation, seemed to have a modulating effect on the cancers carrying this mutation, resulting in fewer tumours reaching advanced Dukes' stages when DR7 was present.

In our study none of the adenomas with low-grade dysplasia showed both K-ras mutation and HLA-DR positivity. This may perhaps indicate that adenomas at this stage carrying both a ras mutation and expressing HLA-DR are immunogenic, and therefore may be eliminated by the immune system.

The majority of the K-ras-positive high-grade and large adenomas were also immunogenic. Several authors (Vogelstein et al., 1988; Ando et al., 1991) have reported no incidence of ras mutations in large, colorectal adenomas as compared with carcinomas. This indicates that some ras-positive adenomas may either not develop into carcinomas or, alternatively, will lose the expression of mutant ras following some form of selective pressure.

It is tempting to speculate that in some of the adenomas, the DR expression may reflect the ongoing immune response directed at a tumour-specific antigen, such as mutated ras. This immune response may eventually lead to a selective elimination of cells expressing mutant ras. This scenario will offer an explanation for the fact that K-ras mutations are less frequent in carcinomas than in adenomas. Furthermore, this view would be compatible with the observation that class II expression is associated with a better prognosis in colorectal carcinomas (Michiels et al., 1991).

It is known that colorectal cancers expressing multiple mutations owing to DNA mismatch correction defects might be highly immunogenic, often showing dense lymphocytic infiltration (Kim et al., 1994). It has been speculated that this may explain the more favourable prognosis observed within this group of patients (Bodmer et al., 1994). In future studies we plan to relate infiltration of T lymphocytes and their subsets to K-ras and HLA-DR expression in both adenomas and carcinomas.
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