Abstract. Novel quinazolinone compounds have been studied in the field of drug discovery for a long time. Among their broad range of pharmacological effects, certain compounds effectively inhibit cancer cell proliferation. MJ-33 is a quinazolinone derivative with proposed anticancer activities that was synthesized in our laboratory. The present study aimed to evaluate the anticancer activity of MJ-33 in fluorouracil (5FU)-resistant colorectal cancer cells (HT-29/5FUR) and to investigate the underlying molecular mechanisms. The cell viability assay results indicated that HT-29/5FUR cell viability was inhibited by MJ-33 treatment in a concentration-dependent manner compared with the control group. The cellular morphological alterations observed following MJ-33 treatment indicated the occurrence of apoptosis and autophagy, as well as inhibition of cell proliferation in a time-dependent manner compared with the control group. The acridine orange, LysoTracker Red and LC3-green fluorescent protein staining results indicated that MJ-33 treatment significantly induced autophagy compared with the control group. The DAPI/TUNEL dual staining results demonstrated increased nuclear fragmentation and condensation following MJ-33 treatment compared with the control group. The Annexin V apoptosis assay and image cytometry analysis results demonstrated a significant increase in apoptotic cells following MJ-33 treatment compared with the control group. The western blotting results demonstrated markedly decreased Bcl-2, phosphorylated (p)-BAD, pro-caspase-9 and pro-caspase-3 expression levels, and notably increased cytochrome c and apoptotic peptidase activating factor 1 expression levels following MJ-33 treatment compared with the control group. Moreover, the expression levels of autophagy-related proteins, including autophagy related (ATG)-5, ATG-7, ATG-12, ATG-16, p62 and LC3-II, were increased following MJ-33 treatment compared with the control group. Furthermore, MJ-33-treated HT-29/5FUR cells displayed decreased expression levels of p-AKT and p-mTOR compared with control cells. The results suggested that MJ-33-induced apoptosis was mediated by AKT signaling, and subsequently modulated via the mitochondria-dependent signaling pathway. Therefore, the results suggested that suppression of AKT/mTOR activity triggered autophagy in the HT-29/5FUR cell line. In summary, the results indicated that MJ-33 inhibited HT-29/5FUR cell viability, and induced apoptosis and autophagy via the AKT/mTOR signaling pathway. The present study may provide novel insight into the anticancer effects and mechanisms underlying MJ-33 in 5FU-resistant colorectal cancer cells.

Introduction

Colorectal cancer (CRC) is a serious malignant disease, typically initiated by aberrant epithelial cell proliferation in the colon or rectum (I). CRC is the third most frequently diagnosed...
type of cancer worldwide (2), and is characterized by a high and continuously increasing mortality rate, estimated to reach 60% by 2035 (3). The treatment options for CRC currently include chemotherapy, targeted therapy and immunotherapy (4). The chemotherapeutic agents most widely used for CRC include 5-fluorouracil (5FU), irinotecan and oxaliplatin (4,5). 5FU is a first-line drug that has been extensively used as a monotherapy or as part of a combination regimen for the treatment of CRC (4,5). However, the proportion of patients with advanced CRC who respond to 5FU is limited to 10-15% (6), whereas ~50% of patients with metastatic CRC display resistance to 5FU-based chemotherapies (7,8). Therefore, the development of novel therapeutic strategies for 5FU-resistant CRC is important.

Several quinazolinone compounds have been developed and approved as novel anticancer agents, including nolatrexed, idelalisib and raltitrexed (9-11). In our laboratory, several quinazolinone compounds (such as MJ-29, MJ-33 and LJJ-10) have been designed, synthesized and studied to investigate their anticancer activities (12,13). For example, the inhibitory effect of MJ-33 on AKT-mediated DU145 prostate cancer cell metastasis was previously reported (14). However, the cytotoxicity and mechanisms underlying MJ-33 in chemoresistant cells are not completely understood. Therefore, the present study aimed to investigate the anticancer activities of MJ-33 in a CRC cell line with acquired resistance to 5FU, namely HT-29/5FUR.

AKT is an important cancer-related regulator that is responsible for cancer cell survival, proliferation and migration (15,16). Moreover, AKT overactivation has been reported to serve as a biomarker of tumorigenesis, tumor growth, metastasis and resistance to cancer therapies (17). AKT/mTOR signaling-mediated control of autophagy and apoptosis in chemoresistant cells has been previously reported (18). In addition, inhibiting the PI3K/AKT signaling pathway restores the sensitivity of HT-29 cells to 5FU (19). A previous study also demonstrated the inhibitory effects of MJ-33 on AKT phosphorylation, resulting in antimetastatic activity (14).

The crosstalk between apoptosis and autophagy is a novel therapeutic target in cancer (20). In the later stage, when autophagy no longer serves a cytoprotective role, it leads to apoptosis activation to induce programmed cell death (21). In a previous study, a novel quinazolinone derivative induced autophagy-related apoptotic cell death in lymphoblastic leukemia MOLT-4 cells (22). Therefore, it was hypothesized that the mechanism underlying MJ-33-induced anticaner activity was associated with the induction of apoptosis and autophagy. The present study investigated the cytotoxic effects and mechanisms underlying MJ-33, and explored the involvement of the AKT downstream signaling pathway in HT-29/5FUR cells.

### Materials and methods

**Chemicals and reagents.** MJ-33 was synthesized in our laboratory at the School of Pharmacy, China Medical University. DAPI, 3-Methyladenine (3-MA), chloroquine (CQ), Bafilomycin A1 (Baf.A1), AKT activator (SC-79), Minimum Essential medium and MTT were purchased from Sigma-Aldrich (Merck KGaA). L-glutamine, RPMI-1640 medium, penicillin, streptomycin, trypsin-EDTA, acridine orange (AO), Lysotracker Red, LC3-green fluorescent protein (GFP) and FBS were purchased from Thermo Fisher Scientific, Inc. Pan-caspase inhibitor (z-VAD-fmk), caspase-9 inhibitor (z-LEHD-fmk) and caspase-3 inhibitor (z-DEVD-fmk) were purchased from Merck KGaA. 5FU was obtained from Pharmacia & Upjohn.

**Cell lines and cell culture.** The human colorectal cancer HT-29 cell line (23) was obtained from the Bioresource Collection and Research Center, Food Industry Research and Development Institute. The cell line was authenticated via STR profiling (Mission Biotech Co., Ltd. Cells were cultured in 75-cm² tissue culture flasks in RPMI-1640 medium supplemented with 2 mM L-glutamine, 10% FBS, 100 U/ml penicillin and 100 µg/ml streptomycin at 37°C with 5% CO₂. 5FU-resistant colorectal cancer cells (HT-29/5FUR) were established according to the following protocol. Parental HT-29 cells were exposed to an initial dose of 0.5 µg/ml (3.843 µM) at 37°C and surviving cells were cultured to 90% confluence for four passages (4 weeks). Surviving cells were exposed to 1 µg/ml 5FU (7.688 µM) for four passages (4 weeks) and then 1.5 µg/ml 5FU (11.530 µM) for four passages (4 weeks). Finally, surviving cells were exposed to 2 µg/ml 5FU (15.372 µM), the clinically relevant plasma concentration, for four passages (4 weeks). Surviving resistant cells were named 5FU-resistant colorectal cancer cells (HT-29/5FUR) (24,25). The doubling time of parental HT-29 cells was 17 h and the doubling time of HT-29/5FUR cells was 34 h.

Prior to MJ-33 treatment, HT-29/5FUR cells were treated with 10 nM 3-MA, 100 µM CQ or 10 nM Baf.A1 at 37°C for 1 h, 10 µM SC-79 at 37°C for 30 min, or 15 µM z-VAD-fmk, 15 µM z-LEHD-fmk or 15 µM z-DEVD-fmk at 37°C for 2 h.

The normal human colon CDD 841 CoN cell line (CRL-1790; American Type Culture Collection) was cultured in Minimum Essential medium supplemented with 10% FBS, 100 U/ml penicillin and 100 µg/ml streptomycin at 37°C with 5% CO₂.

**Cell viability assay.** HT-29/5FUR and CDD 841 CoN cells were seeded (1x10⁴ cells/well) into 96-well plates and treated with different concentrations of MJ-33 (25, 50, 75 and 100 µM). Control cells were treated with DMSO. Positive control cells were treated with 30 µM oxaliplatin (Sanofi S.A.). Cells were incubated with each treatment at 37°C for 24 or 48 h with 5% CO₂. Then, cell viability was assessed by performing the MTT assay as previously described (18,26).

**Cell death and morphological changes on microscopic observation.** HT-29/5FUR cells (1x10⁴ cells/100 µl) were treated with 25, 50, 75 and 100 µM MJ-33 or DMSO at 37°C for 48 h with 5% CO₂. Subsequently, cultured cells were observed using a phase-contrast light microscope (Leica Microsystems GmbH; magnification, x400) to visualize morphological alterations characteristic of apoptotic or autophagic cell death.

**AO, LysoTracker Red and LC3-GFP staining.** HT-29/5FUR cells (1x10⁴ cells/ml) were treated with 75 µM MJ-33 or DMSO for 24 h at 37°C. After harvesting using trypsin-EDTA, cells were fixed in 4% paraformaldehyde on ice for 15 min. Subsequently, cells were stained with 1 µg/ml AO,
LysoTracker Red or LC3-GFP for 20 min at room temperature. Stained cells were visualized using ImageXpress Micro Confocal High-Content Image System (Molecular Devices, LLC; magnification, x400) and analyzed using MetaXpress (version 5.3.0.4; Molecular Devices, LLC) to detect acidic vesicular organelles (for AO staining), lysosomal function (for LysoTracker Red staining) or typical punctate pattern (for LC3-GFP staining).

**DAPI and TUNEL dual staining.** HT-29/5FUR cells were seeded (1x10⁵ cells/ml) into 12-well plates and treated with 75 µM MJ-33 or DMSO for 48 h at 37°C with 5% CO₂. Cells were harvested and fixed with absolute ethanol at room temperature for 10 min. Subsequently, cells were stained with 1 µg/ml DAPI solution for 30 min at room temperature as previously described (27). To detect DNA breaks, the In Situ Cell Death Detection Kit, Fluorescein (Roche Diagnostics GmbH) was used according to the manufacturer's protocol. After applying mounting medium (10% glycerol in PBS), stained samples were observed using a fluorescence microscope (Leica Microsystems GmbH; magnification, x400) as previously described (28). For each coverslip, 3 fields of view with similar numbers of cells were photographed for counting and further analysis.

**Caspase-3 and caspase-7 activity assays.** The fluorochrome-labeled inhibitor of caspases assay (FLICA) method was applied using the FAM-FLICA® Caspases 3 & 7 Assay Kit (Immunochemistry Technologies, LLC; cat. no. 93) and the NucleoCounter NC-3000 (ChemoMetec A/S) according to the manufacturer's protocol. All samples were analyzed using NucleoView NC-3000 software (version 1.4; ChemoMetec A/S). Briefly, HT-29/5FUR cells were seeded (1x10⁶ cells/well) into 6-well plates, treated with 75 µM MJ-33 or DMSO for 12 h at 37°C. Cells were resuspended in each well using 0.5 ml PBS. Samples were incubated with diluted FLICA reagent and Hoechst 33342 for 1 h at 37°C. Following washing twice with apoptosis wash buffer, cells were resuspended in 100 µl apoptosis wash buffer supplemented with PI. Subsequently, 30 µl cell suspension was immediately loaded into a 2-chamber slide (NC-Slide A2; ChemoMetec A/S) for analysis on the NucleoCounter NC-3000 using the built-in caspase assay program.

**Annexin assay and image cytometry analysis.** Experiments were conducted using the Annexin V-FITC Apoptosis Detection Kit (cat. no. AVK050; Strong Biotech Corp.) and the Counter NC-3000 cytomter (ChemoMetec A/S) according to the manufacturer's protocol and the previously described protocol (29). Briefly, HT-29/5FUR cells were seeded (total 1x10⁶ cells/well) into 6-well plates. Samples were treated with 75 µM MJ-33 or DMSO at 37°C for 6 h. Cells were resuspended in each well with 0.5 ml PBS. Subsequently, samples were incubated with 100 µl Annexin V binding buffer as previously described (29). Then, samples were incubated with 2 µl Annexin V-CF 488A conjugate and 2 µl Hoechst 33342 (10 µg/ml) at 37°C for 15 min. Following centrifugation at 200 x g at 37°C for 5 min, samples were washed with Annexin V binding buffer. Cell pellets were resuspended using 100 µl Annexin V binding buffer supplemented with PI at room temperature for 15 min. All samples were immediately analyzed using NucleoView NC-3000 software (version 1.4; ChemoMetec A/S). Subpopulations of stained cells were determined using scatterplots: Healthy cells (Annexin V/PI); early apoptosis (Annexin V+/PI); late apoptosis (Annexin V+/PI); and necrosis (Annexin V+/PI).

**Cell confluence assay.** The IncuCyte S3 ZOOM System instrument (Essen BioScience) was used to conduct the cell confluence assay. HT-29/5FUR cells were seeded (1x10⁶ cells/well) into a 96-well plate with 50 µM MJ-33 or DMSO for 48 h at 37°C. Cells were visualized and photographed every 2 h as previously described (30).

**Western blotting.** HT-29/5FUR cells were lysed with Trident RIPA Lysis Buffer (GeneTex, Inc.). Protein concentrations were determined using a Bio-Rad protein assay system (Bio-Rad Laboratories, Inc.). Proteins were separated as previously described (18,31,32). Briefly, proteins (35 µg) were separated via 10-12% SDS-PAGE and transferred using the iBlot Dry Blotting System (Invitrogen; Thermo Fisher Scientific, Inc.) to PVDF membranes. The membranes were blocked with PBS containing 0.1% Tween-20 and 5% skimmed dry milk for 2 h at room temperature. Subsequently, the membranes were incubated overnight at 4°C with primary antibodies (all purchased from Cell Signaling Technology, Inc.) targeted against: Bcl-2 (cat. no. 4223; 1:1,000), Bax (cat. no. 5023; 1:1,000), BAD (cat. no. 9292; 1:1,000), phosphorylated (p)-BAD (cat. no. 5284; 1:1,000), cytochrome c (cat. no. 4280; 1:1,000), apoptotic peptide activating factor-1 (Apaf-1; cat. no. 8969; 1:1,000), caspase-9 (cat. no. 9508; 1:1,000), caspase-3 (cat. no. 9662; 1:1,000), autophagy related (ATG)-5 (cat. no. 12994; 1:1,000), ATG-7 (cat. no. 8558; 1:1,000), ATG-12 (cat. no. 4180; 1:1,000), ATG-16 (cat. no. 8098; 1:1,000), p62 (cat. no. 23214; 1:1,000), LC3 (cat. no. 12741; 1:1,000), AKT (cat. no. 9272; 1:1,000), p-AKT (cat. no. 4060; 1:1,000), mTOR (cat. no. 2972; 1:1,000), p-mTOR (cat. no. 5536; 1:1,000) and β-actin (cat. no. 8457; 1:1,000). Following primary incubation, the membranes were incubated for 4 h at room temperature with anti-rabbit IgG (cat. no. 7074; 1:10,000; Cell Signaling Technology, Inc.) and anti-mouse IgG (cat. no. 7076; 1:10,000; Cell Signaling Technology, Inc.) HRP-conjugated secondary antibodies. Protein bands were visualized using Immobilon Western HRP Substrate (Merck KGaA) for 1 h.

**Statistical analysis.** Data are presented as the mean ± SD (n=3). Comparisons among multiple groups were analyzed using one-way ANOVA followed by Dunnett’s or Tukey’s post hoc test. Statistical analyses were performed using SPSS software (version 16.0; SPSS, Inc.). P<0.001 was considered to indicate a statistically significant difference.

**Results**

MJ-33 selectively exerts cytotoxic effects on HT-29/5FUR cells, which are associated with apoptosis and autophagy. The MITT assay was performed to examine the antiproliferative effects of MJ-33 on HT-29/5FUR cells in vitro. The results demonstrated that cell viability was significantly inhibited by MJ-33 in a concentration-dependent manner compared with MJ-33 alone. The MTT assay was performed to examine the antiproliferative effects of MJ-33 on HT-29/5FUR cells in vitro. The results demonstrated that cell viability was significantly inhibited by MJ-33 in a concentration-dependent manner compared with MJ-33 alone.
the control group (Fig. 1A and B). As the positive control, oxaliplatin also significantly inhibited HT-29/5FUR cell viability compared with the control group. In CCD 841 CoN cells, cell viability was not significantly altered by MJ-33 treatment, except for in the 100 µM MJ-33 for 48 h group, compared with the control group (Fig. IC and D).

The sensitivity of HT-29/5FUR cells and its parental cell line (HT-29) to MJ-33, 5FU and Ox are presented in Table I. The IC₅₀s of 5FU and Ox in HT-29/5FUR cells were higher compared with in HT-29 cells. Conversely, the IC₅₀ of MJ-33 in HT-29/5FUR cells was lower compared with HT-29 cells. Based on the IC₅₀ values obtained, 75 µM MJ-33 was selected for subsequent experiments. To further examine the effects of MJ-33 treatment on HT-29/5FUR cell morphology, light microscopy and the IncuCyte S3 ZOOM system were used. Compared with the control group, MJ-33 treatment induced morphological alterations, including cell shrinkage, and nuclear fragmentation and condensation, and cell death (Fig. 2A, B and Video S1) suggested apoptotic phenomena. In addition, autophagic cells with increasing volume of autophagic vesicles were observed following MJ-33 treatment (Fig. 2B). The results suggested that MJ-33-induced cell death was mediated via mechanisms involving apoptosis and autophagy.

**MJ-33 induces apoptosis via the mitochondrial intrinsic signaling pathway in HT-29/5FUR cells.** MJ-33-induced cell death was observed in HT-29/5FUR cells, thus the apoptotic bodies were subsequently quantified. By performing DAPI and TUNEL staining on HT-29/5FUR cells, the proportion of apoptotic cells after MJ-33 treatment was examined. Under a fluorescence microscope, the nuclear morphology of dead cells was observed, which displayed chromatin condensation, a well-defined hallmark of apoptosis (Fig. 3A). The TUNEL-stained cells emitted green fluorescence in response to DNA fragmentation, which is an indicator of late-stage apoptosis (33). The results demonstrated that MJ-33 treatment significantly increased TUNEL-positive staining compared with the control group (Fig. 3B). Image cytometry analysis was conducted to determine the apoptotic subpopulations of MJ-33-treated and control cells. The percentage of early apoptotic cells was increased from 11% in control cells to 39% in MJ-33-treated cells (Fig. 3C). Moreover, MJ-33 treatment significantly increased the number of Annexin-V-positive/PI-negative cells compared with the control group (Fig. 3D). Therefore, the results further indicated that HT-29/5FUR cell apoptosis was induced by MJ-33 treatment.

To improve the current understanding of MJ-33-induced apoptotic death in MJ-HT-29/5FUR cells, the possible regulatory signaling pathways were investigated. It was hypothesized that MJ-33 induced apoptotic cell death via the caspase-dependent signaling pathways. Therefore, cell viability was examined following MJ-33 treatment with or without pan-caspase inhibitor (z-VAD-FMK). Cell viability in the MJ-33-treated groups was significantly lower compared with the control group (Fig. 4). Furthermore, cell viability was significantly higher in the MJ-33 + z-VAD-FMK/MJ-33 group compared with the MJ-33 group, suggesting that MJ-33-induced HT-29/5FUR cell apoptosis was mediated via caspase activity. To determine the effect of specific caspase enzymes on MJ-33-induced apoptosis, cells were pretreated with z-LEHD-FMK (to inhibit caspase-9) or z-DEVd-FMK (to inhibit caspase-3). The cell viability assay results demonstrated MJ-33-induced cytotoxicity in HT-29/5FUR cells was significantly inhibited by caspase-9 and caspase-3 inhibitors. The results indicated that MJ-33-induced apoptosis was mediated via caspase-9 and caspase-3 (Fig. 4B and C). The aforementioned results suggested that MJ-33 induced apoptotic cell death via the intrinsic signaling pathway in HT-29/5FUR cells.

To identify the roles of proapoptotic proteins in the molecular mechanisms underlying MJ-33-induced apoptosis, the expression levels of proapoptotic proteins, including Bax, Bcl-2 and p-BAD, were investigated in MJ-33-treated HT-29/5FUR cells. Compared with the control group, Bax and BAD protein expression levels were markedly increased in a concentration-dependent manner following treatment with MJ-33 (Fig. 5A). By contrast, p-BAD protein expression levels were markedly decreased by MJ-33 treatment compared with the control group, and reversely associated with the protein expression levels of Bax. Compared with the control group, Bcl-2 expression levels were slightly increased following treatment with 50 µM MJ-33, but notably decreased following treatment with 75 and 100 µM MJ-33.

Similarly, the protein expression levels of cytochrome c, Apaf-1, pro-caspase-9 and pro-caspase-3 were examined in HT-29/5FUR cells following treatment with MJ-33. The protein expression levels of cytochrome c and Apaf-1 were notably increased by MJ-33 treatment in a concentration-dependent manner compared with the control group (Fig. 5B). Conversely, the protein expression levels of pro-caspase-9 and pro-caspase-3 were decreased by MJ-33 treatment compared with the control group. The results indicated that apoptotic cell death was promoted by MJ-33 treatment via the mitochondria-mediated apoptotic signaling pathway.

**MJ-33 activates an autophagic mechanism in HT-29/5FUR cells.** The aforementioned results prompted further investigation.
into MJ-33-induced autophagy in HT-29/5FUR cells. Cells were treated with 75 µM MJ-33 and then stained with AO, LysoTracker Red or LC3-GFP (Fig. 6A). The relative fluorescence intensity emitted in the red range indicated significantly increased uptake of AO in the MJ-33-treated group compared with the control group, which suggested increased formation of acidic vesicles (Fig. 6B). LysoTracker Red staining results suggested significantly increased lysosomal activity and autophagosomal maturation in the MJ-33 treatment group compared with the control group (Fig. 6C). Additionally, the green fluorescence emitted in the LC3-GFP stained group was significantly higher in the MJ-33 treatment group compared with the control group, indicating punctate formation, which is typically observed in autophagic cells (Fig. 6D). Collectively, the aforementioned results suggested that MJ-33 treatment activated an autophagic mechanism in HT-29/5FUR cells.

Figure 1. Effects of MJ-33 on HT-29/5FUR and CCD 841 CoN cell viability. Cells were seeded (1x10³ cells/well) into 96-well plates and treated with 0, 25, 50, 75 and 100 µM MJ-33. HT-29/5FUR cell viability following treatment for (A) 24 or (B) 48 h. CCD 841 CoN cell viability following treatment for (C) 24 or (D) 48 h. Data are presented as the mean ± SD from three independent experiments. Data were analyzed using one-way ANOVA followed by Dunnett's post hoc test. ***P<0.001 vs. control. 5FUR, fluorouracil-resistant; Ox, oxaliplatin.

Figure 2. Effects of MJ-33 on HT-29/5FUR cell morphology. (A) Morphological alterations and cell death in HT-29/5FUR cells following treatment with MJ-33 (magnification, x200). (B) Apoptotic and autophagic cells were observed. 5FUR, fluorouracil-resistant (magnification, x200).
MJ-33 inhibits AKT/mTOR activity and elevates the expression of autophagy-related proteins in HT-29/5FUR cells. It was previously reported that MJ-33 treatment regulated the MAPK, AKT, NF-κB and activator protein-1 signaling pathways in DU145 cells (14). Since AKT/mTOR signaling may mediate both apoptosis and autophagy (16,34), the effect of MJ-33 treatment on AKT and mTOR protein expression levels in HT-29/5FUR cells was evaluated. The western
Western blotting results demonstrated that the ratios of p-AKT/AKT and p-mTOR/mTOR protein were notably decreased by MJ-33 treatment in a concentration-dependent manner compared with the control group (Fig. 7A). Therefore, the results suggested that the activity of the AKT/mTOR axis was inhibited by MJ-33 treatment in HT-29/5FUR cells.

According to previous studies, inhibition of mTOR activity may induce autophagy via upregulating the expression of the ATG protein family (15,18,35). To further elucidate the molecular mechanisms underlying MJ-33-induced autophagy in HT-29/5FUR cells, the expression levels of autophagy-related proteins, including ATG-5, ATG-7, ATG-12, and ATG-16, p62 and LC3-II, were examined in MJ-33-treated HT-29/5FUR cells. The protein expression levels of all ATG proteins and p62 were notably increased, whereas the ratio of LC3/LC3-II was markedly decreased by MJ-33 treatment in a concentration-dependent manner compared with the control group (Fig. 7B). The results suggested that MJ-33-induced autophagy activated the intrinsic apoptosis signaling pathway. Western blotting was performed to measure the expression levels of (A) proapoptotic and (B) intrinsic apoptosis signaling pathway-related proteins. The density of the bands compared with the control sample (set to 1.0) are presented above each band. p, phosphorylated; Apaf-1, apoptotic peptidase activating factor 1.
autophagy may be triggered at the vesicle nucleation step, resulting in ATG protein involvement, reductions in the LC3/LC3-II ratio and increased p62 expression, ultimately supporting autophagosome formation and the overall progression of autophagy.

Inhibition of autophagy enhances MJ-33-induced apoptosis in HT-29/5FUR cells. To further examine the contribution of autophagy signaling to MJ-33-induced cell death, HT-29/5FUR cells were treated with different autophagy inhibitors, including CQ, 3-MA and Baf.A1. HT-29/5FUR cells were treated with autophagy inhibitor and/or MJ-33. Combined treatments significantly decreased cell viability compared with treatment with MJ-33 alone (Fig. 8A-C). To clarify the effect of autophagy inhibitors on MJ-33-induced apoptosis, the effects of MJ-33 and 3-MA treatment on caspase-3 and caspase-7 activities were assessed by performing FLICAs. MJ-33 treatment alone significantly elevated caspase-3 and caspase-7 activities compared with the control group (Fig. 9A). However, the combination of MJ-33 and 3-MA treatment induced significantly higher caspase-3 and caspase-7 activities compared with the MJ-33 group. Collectively, the results suggested that the autophagy mechanism in HT-29/5FUR cells was associated with MJ-33-induced cell death, indicating that autophagy may serve a cytoprotective role by inhibiting the apoptosis mechanism.

**AKT serves a pivotal role in the mechanism underlying MJ-33-induced cytotoxicity in HT-29/5FUR cells.** MJ-33 inhibits AKT activity in a concentration-dependent manner, thus further experiments to determine the role of AKT in MJ-33-induced cytotoxicity were performed. Following treatment with MJ-33 and/or SC-79 (AKT specific activator), HT-29/5FUR cell viability was assessed. SC-79 did not significantly alter cell viability compared with the control group (Fig. 9B). Conversely, SC-79 significantly restored HT-29/5FUR cell viability in MJ-33-treated cells. The results suggested that AKT may serve as a critical regulator of MJ-33-induced cytotoxicity in HT-29/5FUR cells.

Collectively, the results indicated that MJ-33 selectively induced cytotoxicity in HT-29/5FUR cells via mediating the AKT/mTOR signaling pathway. The proposed molecular mechanism underlying MJ-33 in HT-29/5FUR cells is presented in Fig. 10. The results indicated that MJ-33 inhibited AKT activity, which triggered apoptosis via the caspase-dependent signaling pathway and induced autophagy via the ATG-dependent pathway.

**Discussion**

Quinazolinone compounds may possess diverse bioactivities, including anticancer properties (22,28,36). The anticancer mechanisms underlying apoptosis induction and antiproliferative activity have been previously reported (28). However, the autophagy-associated mechanisms of action of these compounds, particularly in chemoresistant cancer cells, have not been extensively investigated (36,37). The present study examined the cytotoxic effect of MJ-33 on a chemoresistant CRC cell line. The results indicated that MJ-33-induced cytotoxicity was associated with apoptosis and autophagy. Since the antimetastatic effects of MJ-33 were previously reported (14), the findings of the present study provided further understanding of the effects of MJ-33, contributing to establishing the therapeutic profile of MJ-33, which may be useful for further exploration of the anticancer effects of MJ-33 in 5FU-resistant cancer cells.
Apoptosis is a programmed cell death mechanism that serves a key role in the inhibition of tumor growth by limiting the number of cells (33). 5FU is an apoptosis-inducing agent, which is widely utilized for CRC treatment, despite its low reported response rates (6-8). Chemoresistant cancer cells are generally characterized by the absence of response to treatment, maintaining their ability to survive and continuous proliferation under chemotherapy treatment (38). In cancer cells exhibiting resistance to 5FU, the inactivation of apoptosis was reported in several studies (39,40). In the present study, the results suggested that MJ-33 induced cytotoxicity by reactivating the apoptotic mechanism in HT-29/5FUR cells. DNA condensation and fragmentation were observed following MJ-33 treatment, suggesting induction of HT-29/5FUR cell apoptosis. The significantly increased number of early apoptotic cells (Annexin V positive/PI negative) in the MJ-33 treatment group compared with the control group further indicated that MJ-33 induced apoptosis. Moreover, the results suggested that
MJ-33-induced apoptosis was mediated via a caspase-dependent signaling pathway. Since AKT activity is responsible for cell survival and apoptosis (15), the effect of MJ-33 on AKT activity and possible downstream signaling pathways were evaluated in HT-29/5FUR cells. Compared with the control group, the p-AKT/AKT ratio was notably decreased following MJ-33 treatment, suggesting that AKT activity was inhibited via downregulation of AKT phosphorylation. Moreover, compared with the control group, AKT activator treatment alone did not enhance cell viability, but in combination with MJ-33 treatment, AKT activator significantly restored cell viability, suggesting that MJ-33-induced apoptosis may be controlled via AKT signaling. In addition, the Bcl-2/BAX and p-BAD/BAD ratios were markedly decreased in the MJ-33 treatment group compared with the control group. The Bcl-2/BAX ratio is a proapoptotic indicator and the activity of BAD is regulated via phosphorylation, both of which control the process of cell apoptosis (13,29). Moreover, compared with the control group, MJ-33 treatment markedly upregulated proapoptotic protein expression, promoting programmed cell death. Furthermore, the notably increased expression levels of cytochrome c and Apaf-1, and markedly decreased expression levels of pro-caspase-9 and pro-caspase-3 in the MJ-33 treatment group compared with the control group suggested that MJ-33-induced apoptosis was triggered and controlled via the intrinsic mitochondria-dependent signaling pathway.

Autophagy is also a mechanism closely associated with cell death and survival (21). The interaction between cytotoxicity and autophagy displays dual roles as dependent on the level of the signal, stimuli or stress can induce autophagy as an offensive or defensive mechanism (41,42). The antiapoptotic effect associated with autophagy of several quinazolinone compounds was previously reported (22,43). In the present study, MJ-33-induced HT-29/5FUR cell autophagy was detected by observing morphological alterations via microscopic examination. Furthermore, the formation of acidic vesicles, increasing volume of autophagic vesicles, acidic vesicular organelles and increased lysosomal activity indicated autophagy activation following MJ-33 treatment. Moreover, the punctate patterns suggested autophagosome maturation in MJ-33-treated cells compared with control cells, which also indicated autophagy induction. The ATG12-ATG5 conjugate interacts with ATG16 to form a complex that catalyzes the expanding autophagosome membrane, promoting early-stage autophagy (18,41). The conversion of LC3 to LC3-II via lipidation is required for autophagosome membrane maturation during the process of autophagy (44). In addition, the p62 protein is an autophagy substrate and an indicator for cargo recognition, serving an important role in delivering ubiquitinated proteins to the proteasome for degradation (45). In the present study, compared with the control group, MJ-33 treatment markedly increased p62 protein expression levels, which indicated induction of later-stage autophagy via triggering of autophagy-related protein expression. Collectively, the results indicated that MJ-33-induced HT-29/5FU autophagy was triggered and processed via inhibiting mTOR phosphorylation, and subsequently upregulating the expression of autophagy-related proteins.

Alterations in AKT activity are crucial for cell survival and strongly regulate apoptosis (15). The effect of MJ-33 on the regulation of AKT in HT-29/5FUR cells was investigated. The results demonstrated a markedly reduced p-AKT/AKT ratio in MJ-33-treated cells compared with control cells, indicating that AKT activity was inhibited by suppressing its phosphorylation. It was inferred that MJ-33-induced apoptosis may be controlled via AKT signaling. mTOR is an important molecule downstream of AKT, which serves critical roles in autophagy pre-initiation and progression (15). As the main component of the mTOR complex 1, it inhibits Unc-51 like autophagy activating kinase (46) and also regulates lysosome activity by suppressing transcription factor EB (47). Therefore, the present study investigated the effects of MJ-33 treatment on AKT/mTOR activity in HT-29/5FUR cells, and the association between the effects and MJ-33-induced autophagy. Compared with the control group, MJ-33 treatment inhibited the AKT/mTOR axis, thereby promoting autophagy. In previous studies investigating quinazolinone compounds, the anticancer effects against CRC cells have been reported via a variety of mechanisms, including apoptosis, antiangiogenesis and antimitastasis mechanisms (48,49). The molecular mechanisms associated with the anticancer properties of these compounds have been reported, including thymidylate synthase (TS) inhibition (50) and PI3K inactivation (48,51). The findings of the present study were consistent with previously reported results on the inhibitory effect of MJ-33 on the AKT/mTOR/AMPK axis in DU145 cells (14), which resulted in NF-κB downregulation. Therefore, exploring the relationships among NF-κB activity, the mechanism underlying 5FU resistance and the activity of MJ-33 should be investigated in future studies. Constitutive nuclear NF-κB activity is highly activated in TS inhibitor-resistant cells (52), and inhibiting NF-κB translocation may lower TS levels in CRC (53), causing higher TS expression in 5FU-resistant HT-29 cells (54). Since TS is the primary target of 5FU, the aforementioned studies also indicated that MJ-33 may serve as a potential therapeutic agent among quinazolinone class compounds for developing drugs against 5FU-resistant CRC.

The interplay between autophagy and apoptosis is complicated, with multiple processes, connections and...
cells, and MJ-33 combination strategies. The proposed molecular mechanism underlying MJ-33 in HT-29/5FUR cells that was identified in the present study is presented in Fig. 10. Consistent with a previous study that investigated the antimetastasis effects of MJ-33 (14), the interaction of MJ-33 and the AKT/mTOR pathway was further indicated in the present study. The results of the present study may provide novel insights into the cytotoxicity of MJ-33, MJ-33-induced autophagy and apoptosis, as well as the underlying molecular mechanisms. Additional effects of MJ-33 on different hallmarks of cancer, such as angiogenesis, metastasis and molecular targets, require further investigation.

In conclusion, the present study demonstrated that MJ-33 treatment significantly inhibited HT-29/5FUR cell viability compared with the control group in vitro. Moreover, the results indicated that MJ-33-induced cytotoxicity in HT-29/5FUR cells was mediated via the inhibitory effect of MJ-33 on the AKT/mTOR signaling pathway, which subsequently triggered apoptosis and autophagy. Therefore, MJ-33 may serve as a promising anticancer drug for 5FU-resistant CRC.

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Availability of data and materials

The datasets generated during the current study are available from the corresponding author on reasonable request.

Authors' contributions

MJH, HAH and JSY contributed to designing the study. JHC, DTB, YNJ and YHL performed the experiments. FJT and JSY analyzed the data. HAH, JSY and FJT wrote and revised the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interest.
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