Development of a Click Beetle Luciferase Reporter System for Enhanced Bioluminescence Imaging of Listeria monocytogenes: Analysis in Cell Culture and Murine Infection Models

Sadeeq Ur Rahman1,2,3, Michael Stanton4, Pat G. Casey1, Angela Spagnuolo5, Giuliano Bensi5, Colin Hill1,2, Kevin P. Francis6, Mark Tangney1,4,7 and Cormac G. M. Gahan1,2,7,8*

1 APC Microbiome Institute, University College Cork, Cork, Ireland, 2 School of Microbiology, University College Cork, Cork, Ireland, 3 College of Veterinary Sciences and Animal Husbandry, Abdul Wali Khan University Mardan, Mardan, Pakistan, 4 Cork Cancer Research Centre, University College Cork, Cork, Ireland, 5 GSK Vaccines S.r.l., Siena, Italy, 6 PerkinElmer, Alameda, CA, United States, 7 SynBio Centre, University College Cork, Cork, Ireland, 8 School of Pharmacy, University College Cork, Cork, Ireland

Listeria monocytogenes is a Gram-positive facultative intracellular pathogen that is widely used as a model organism for the analysis of infection biology. In this context, there is a current need to develop improved reporters for enhanced bioluminescence imaging (BLI) of the pathogen in infection models. We have developed a click beetle red luciferase (CBR-luc) based vector (pPL2CBRopt) expressing codon optimized CBR-luc under the control of a highly expressed Listerial promoter (PHELP) for L. monocytogenes and have compared this to a lux-based system expressing bacterial luciferase for BLI of the pathogen using in vitro growth experiments and in vivo models. The CBR-luc plasmid stably integrates into the L. monocytogenes chromosome and can be used to label field isolates and laboratory strains of the pathogen. Growth experiments revealed that CBR-luc labeled L. monocytogenes emits a bright signal in exponential phase that is maintained during stationary phase. In contrast, lux-labeled bacteria produced a light signal that peaked during exponential phase and was significantly reduced during stationary phase. Light from CBR-luc labeled bacteria was more efficient than the signal from lux-labeled bacteria in penetrating an artificial tissue depth assay system. A cell invasion assay using C2Bbe1 cells and a systemic murine infection model revealed that CBR-luc is suited to BLI approaches and demonstrated enhanced sensitivity relative to lux in the context of Listeria infection models. Overall, we demonstrate that this novel CBR reporter system provides efficient, red-shifted light production relative to lux and may have significant applications in the analysis of L. monocytogenes pathogenesis.

Keywords: Listeria monocytogenes, click beetle luciferase, bioluminescence, in vivo imaging, pathogen, virulence, clinical
INTRODUCTION

*Listeria monocytogenes* is an opportunistic facultative intracellular pathogen which is capable of withstanding harsh environmental conditions, including stresses encountered during the processing, packaging and storage of foods (Gahan and Hill, 2005). Following consumption of contaminated foods *L. monocytogenes* survives the conditions encountered in the gastrointestinal tract and can progress to systemic infection (listeriosis) (Gahan and Hill, 2014). In humans, the clinical manifestations of listeriosis include meningitis, encephalitis, and septicemia predominately in the immune-compromised host, or late-term spontaneous abortion in pregnant individuals. The mortality rate can reach 30% during common-source outbreaks in susceptible hosts (Mead et al., 2000). *L. monocytogenes* infection in mice has been used as an important model for the analysis of intracellular parasitism and subsequent immunity and this research has benefited from the development of molecular tools to analyze bacterial and host responses *in vivo* (Cossart and Toledo-Arana, 2008; Lebreton et al., 2016).

Bioluminescence imaging (BLI) is a non-invasive technology that permits the molecular analysis of cells through the expression of proteins that emit visible light (Byrne et al., 2013). The development of sensitive light detection systems has permitted the use of BLI for the visualization of cell activity or localization in small animal models and other model systems (Contag and Bachmann, 2002). The technology is particularly useful for the *in vivo* tracking of bacterial infectious agents in real time in small animal models (Cronin et al., 2012a,b). Engineering of bacteria to produce light has typically involved cloning and expression of a gene system (the bacterial lux system) that is naturally found in species of *Vibrio* and *Photorhabdus* (Gahan, 2012). However, eukaryotic luciferases from natural sources (e.g., firefly, click beetle, sea pansy) can also be expressed in bacteria and may offer alternatives to the use of bacterial Lux for bacterial imaging studies (Gahan, 2012; Tangney and Francis, 2012; Chang et al., 2014; Karimi et al., 2016).

The bacterial lux-based bioluminescent reporter systems are based on the expression of the bacterial lux operon (*luxCDABE*) to produce light (Belkin et al., 1996). This system does not require the addition of substrate as the LuxCDE proteins generate endogenous substrate in the bacterial cytoplasm (Gahan, 2012). In contrast, the use of eukaryotic bioluminescence systems (Luc enzymes) requires the addition of exogenous luciferin as substrate (Gahan, 2012; Vorobyeva et al., 2015). The emission spectra of these systems also varies, with the insect luciferase enzymes emitting light in the green-red range of the visible light spectrum, compared with the bacterial luciferase system that emits blue-green light (Contag and Bachmann, 2002). This has implications for whole body imaging experiments, as blue-green light is of a relatively short wavelength (400–490 nm) and therefore has a reduced ability to penetrate host tissues compared with light emitted in the red region of the spectrum (Rice et al., 2001; Zhao et al., 2005).

To date, BLI vectors utilized in *L. monocytogenes* have successfully exploited the bacterial lux operon for *in vivo* localization studies (Hardy et al., 2004, 2009; Disson et al., 2008; van Pijkeren et al., 2010; Poulson et al., 2011; Cronin et al., 2012a; Bergmann et al., 2013) and for *in situ* analysis of microbial gene expression (Riedel et al., 2009; Sleator et al., 2009; Joyce and Gahan, 2010; Quereda et al., 2016). In the current study, we created a bioluminescence reporter system based upon a red-optimized click beetle luciferase enzyme (CBR-luc) for use in *L. monocytogenes* and compare this with a previously described lux system (Riedel et al., 2007). The CBR-luc system utilizes a *Listeria*-specific chromosomal integration vector (pPL2) (Lauer et al., 2002) and constitutive expression of the click beetle luciferase was achieved using the highly expressed listerial synthetic promotor (P*HEL*P) described previously (Riedel et al., 2007). We envisage that this vector will have significant applications in the study of *L. monocytogenes* in the context of both *in vivo* and *in vitro* experimental systems.

MATERIALS AND METHODS

**Strains, Media, and Bacterial Growth**

The strains and plasmids used in this study are listed in Table 1. *E. coli* TOP10 (Invitrogen, Paisley, United Kingdom) were used as a cloning host for the creation of pPL2CBRopt. *E. coli* was grown aerobically at 37°C in Luria-Bertani broth (LB) medium. *L. monocytogenes* EGDe (Glaser et al., 2001) and murinized *L. monocytogenes* EGDe (Monk et al., 2010) were grown aerobically at 30°C in brain heart infusion (BHI) medium (Oxoid, Basingstoke, United Kingdom). Clinical strains of *L. monocytogenes* strains 130029, 140030, and 150008 were obtained from clinical cases of listeriosis in Ireland (manuscript in preparation). The pH of the LB and BHI medium was buffered with 100 mM 3-(N-morpholinopropanesulfonic acid (pH 7.4) where outlined in the text. When required, antibiotics were added to the media as follows: for *E. coli*: chloramphenicol (30 µg/ml), ampicillin (100 µg/ml) and for *L. monocytogenes*: chloramphenicol (7.5 µg/ml). For measurement of growth curves, *L. monocytogenes* strains were grown overnight and diluted in fresh medium until the OD₆₀₀ reached 0.05. Subsequently strains were grown shaking at 30°C and readings were taken using a spectrophotometer. To measure bioluminescence during growth over time, overnight cultures were diluted in fresh medium to OD₆₀₀ 0.05 and subsequently grown at 30°C. At selected time points, 50 µl of culture was mixed with 50 µl luciferin (PerkinElmer) with a final concentration of 150 µg/ml as described by the manufacturer.

**Synthesis, Cloning, and Integration of CBR**

CBR was codon-optimized (CBRopt) for expression in *Listeria* and synthesized by GenScript (GenScript United States Inc., Piscataway, NJ, United States). The CBRopt gene (Gene Accession No. KY628997) was synthesized with an intact *L. monocytogenes* upstream constitutive Listerial promoter, highly expressed Listerial promotor (P*HEL*P) (Riedel et al., 2007) and the fragment was sub-cloned into the pPL2 backbone (Lauer et al., 2002) in *E. coli* using *Xho*I and *Pst*I restriction enzymes resulting in pPL2CBRopt (see Figure 1). Plasmid DNA
### TABLE 1 | Strains and plasmids used in this study.

| Strain/plasmid | Features | Source |
|----------------|----------|--------|
| **E. coli TOP10** | Cloning host | Invitrogen |
| **L. monocytogenes strains** | | |
| EGDe | Wild type of serotype 1/2a with known genome sequence | Glaser et al., 2001 |
| Murinized EGDe<sup>m</sup> | Mouse adapted EGDe strain of serotype 1/2a expressing modified internalin A (InlA) protein | Monk et al., 2010 |
| EGDe::pPL2lux | A wild type EGDe strain integrated with pPL2lux vector at tRNA<sub>Arg</sub> locus, which is a vector backbone and carrying Cm<sup>r</sup> marker | Bron et al., 2006 |
| EGDe::pPL2lux<sup>PHELP</sup> | A wild type EGDe strain with pPL2lux<sup>PHELP</sup> integrated at tRNA<sub>Arg</sub> expressing bacterial lux operon under the influence of a listerial highly expressed promoter (P<sub>HELP</sub>) and carrying Cm<sup>r</sup> marker | Riedel et al., 2007 |
| EGDe::pPL2CBR<sup>opt</sup> | A wild type EGDe strain with pPL2CBR<sup>opt</sup> integrated at tRNA<sub>Arg</sub> expressing click beetle red luciferase under the influence of a listerial highly expressed promoter (P<sub>HELP</sub>) and carrying Cm<sup>r</sup> marker | This study |
| **L. monocytogenes-4b-130029** | A clinical strain of serotype 4b isolated from the CSF of a female patient with pPL2CBR<sup>opt</sup> integrated at tRNA<sub>Arg</sub> expressing click beetle red luciferase under the influence of a listerial highly expressed promoter (P<sub>HELP</sub>) and carrying Cm<sup>r</sup> marker | This study |
| **L. monocytogenes-4b-140030** | A clinical strain of serotype 4b isolated from the CSF of a male patient with pPL2CBR<sup>opt</sup> and integrated at tRNA<sub>Arg</sub> expressing click beetle red luciferase under the influence of a listerial highly expressed promoter (P<sub>HELP</sub>) and carrying Cm<sup>r</sup> marker | This study |
| **L. monocytogenes-1/2a-150008** | A clinical strain of serotype 1/2a isolated from the CSF of a male patient and integrated with pPL2CBR<sup>opt</sup> integrated at tRNA<sub>Arg</sub> expressing click beetle red luciferase under the influence of a listerial highly expressed promoter (P<sub>HELP</sub>) and carrying Cm<sup>r</sup> marker | This study |
| Murinized EGDe::pPL2lux | A murinized wild type EGDe strain with pPL2lux vector integrated at tRNA<sub>Arg</sub> locus, which is a vector backbone and carrying Cm<sup>r</sup> marker | Bron et al., 2006 |
| Murinized EGDe::pPL2lux<sup>PHELP</sup> | A murinized wild type EGDe strain with pPL2lux<sup>PHELP</sup> integrated at tRNA<sub>Arg</sub> expressing bacterial lux operon under the influence of a listerial highly expressed promoter (P<sub>HELP</sub>) and carrying Cm<sup>r</sup> marker | Riedel et al., 2007 |
| Murinized EGDe::pPL2CBR<sup>opt</sup> | A wild type EGDe strain with pPL2CBR<sup>opt</sup> integrated at tRNA<sub>Arg</sub> expressing click beetle red luciferase under the control of a listerial highly expressed promoter (P<sub>HELP</sub>) and carrying Cm<sup>r</sup> marker | This study |
| Plasmids | | |
| pPL2lux | A derivative of pPL2 listerial integrative vector backbone with PSA listeriophage integrase and attachment, Cm<sup>r</sup> marker, bacterial lux operon without promoter | Bron et al., 2006 |
| pPL2lux<sup>PHELP</sup> | A derivative of pPL2 listerial integrative vector backbone with PSA listeriophage integrase and attachment site, bacterial lux operon with P<sub>HELP</sub> promoter and Cm<sup>r</sup> marker | Riedel et al., 2007 |
| pPL2CBR<sup>opt</sup> | A derivative of pPL2 listerial integrative vector backbone with PSA listeriophage integrase and attachment site and Cm<sup>r</sup> marker | This study |

**FIGURE 1 | Cloning and expression of codon optimized click beetle luciferase.**

(A) Plasmid map of pPL2CBR<sup>opt</sup> expressing codon-optimized click beetle luciferase (CBR<sup>opt</sup>) from the Listerial highly expressed promoter (P<sub>HELP</sub>) (Riedel et al., 2007) in a pPL2 backbone (Lauer et al., 2002). (B) Growth rate of L. monocytogenes EGDe tagged with a bacterial lux (Riedel et al., 2007), CBR<sup>opt</sup> or an integrated vector control. Data and error bars represent mean and standard deviations, respectively, of triplicate samples. Each experiment was repeated at least three times in triplicate.
was isolated from *E. coli* using a QIAprep Spin Miniprep kit according to the manufacturer’s instructions (QIAGEN, Crawley, United Kingdom). The sequences were verified by sequencing through MWG Biotech AG (Ebersberg, Germany). Transformation of *L. monocytogenes* was performed as described (Park and Stewart, 1990). Successful integrants were screened for site-specific integration by PCR using template DNA of *L. monocytogenes* as described previously (Bron et al., 2006). Restriction endonucleases (Roche Diagnostic, Mannheim, Germany), T4 DNA ligase (Roche) and 2× PCR mixture (NEB, United Kingdom) were used as advised by the manufacturer. From each successful transformant one colony carrying the anticipated genotype was chosen for further bioluminescence analysis using an IVIS Lumina II system (PerkinElmer).

**In Vitro Tissue Depth Model**

*L. monocytogenes EGD::pPL2CBR<sup>op</sup>, L. monocytogenes EGD::pPL2lux (silent vector) or L. monocytogenes EGD::pPL2luxP<sub>HELP</sub> (expressing bacterial lux operon)** *(Table 1)* were grown over night in BHI medium. Subsequently, cultures were grown in fresh BHI medium to a specified OD<sub>600</sub>. At specified OD<sub>600</sub> (OD<sub>600</sub> 1 or 1.5) cells were transferred to 96-well round bottom plates for imaging. Luciferin (PerkinElmer) was added to *L. monocytogenes EGD::pPL2CBR<sup>op</sup> (for *in vitro* 150 µg/ml, for *in vivo* 150 mg/kg body weight) according to the manufacturer’s instructions 15 min prior to imaging. Measurements of bioluminescence were repeated with intervening layers of uniformly cut meat slices (one to five layers, thickness varied from 2 to 10 mm depending on meat type) tested over the surface of the plate. Three different meat types (to reflect different physical and chemical parameters), chicken, serrano ham, and fresh sliced raw beef steak were tested. Images were acquired with up to 3 min exposure and processed by Living Image Software v4.2.

**Cell Invasion**

All the cell culture procedures were performed as described previously (Bron et al., 2006). C2Bbe1 cells (CRC-2012; American Type Culture Collection), a clone of the Caco2 human adenocarcinoma cell line, were used for the cell invasion assay. Briefly, C2Bbe1 cells were maintained in Dulbecco’s modified Eagle’s medium (DMEM) with 4.5 g/l Glutamax (Gibco Laboratories, Grand Island, NY, United States), 10% fetal bovine serum (Gibco), 1% (vol/vol) non-essential amino acids (Gibco), 1% (vol/vol) penicillin–streptomycin (Gibco), and 0.01 mg/ml human transferrin (Calbiochem) at 37°C. For bacteria preparation and infection, overnight cultures of *L. monocytogenes* were pelleted by centrifugation (7000 × g for 5 min), washed twice with PBS, and used to infect mice (*n* = 24) intraperitoneally at 2 × 10<sup>5</sup> CFU in a total of 200 µl of PBS (Rea et al., 2004). The mice were followed for 72 h for imaging with 24 h intervals as described. After 24-, 48-, and 72-h post-infection, the infected mice were imaged following injection of luciferin (PerkinElmer, Inc., Boston, MA, United States) according to the manufacturer’s instructions at a dose of 150 mg/kg body weight administered via IP route as appropriate or imaged directly (mice infected with *L. monocytogenes EGD::pPL2luxP<sub>HELP</sub>). After whole body imaging, mice were sacrificed, and post-mortem resected organs were imaged using the IVIS system. Images were acquired with up to 5 min exposure and processed by Living Image Software v4.2 or v4.3.1.

**Ethics Statement**

All animal procedures were performed in accordance with the national ethical guidelines prescribed by the Health Products Regulatory Authority (HPRA). Protocols and all the murine experiments were approved by the animal ethics committee of University College Cork (AERR #2010/003 and #2012/015).

**Statistical Analysis**

All experiments were performed in triplicate at least two times. Data shown are for representative experiments. Statistical significance was based on either the Student’s *t*-test or ANOVA for comparison of groups (see Figures 2B, 6C). GraphPad Prism 5 or IBM-SPSS v22 was used for graphs and statistical analysis. Linear regressions were carried out using GraphPad Prism 5. In all analysis, *p* < 0.05 was considered as significant.
RESULTS AND DISCUSSION

Construction and Functional Analysis of a Click Beetle Red Luciferase Reporter System

The main purpose of the work reported herein was the construction and functional analysis of the pPL2CBRopt bioluminescence vector (Figure 1A) using the backbone of the pPL2 integrating plasmid. pPL2 contains a PSA listeriophage integrase gene and attachment site that directs site-specific single copy integration into tRNAAsp locus on the L. monocytogenes chromosome (Lauer et al., 2002). In our lab, we have previously successfully created pPL2-derivatives, pPL2lux and pPL2luxPHelp that allow BLI through expression of the bacterial lux operon (Bron et al., 2006; Riedel et al., 2007).

Here we used a similar approach to create a vector expressing a CBR-luc with potential applications in deep tissue whole body imaging.

For this purpose, a codon optimized CBR-luc (CBRopt) gene for optimal expression in L. monocytogenes was synthesized under the influence of a constitutive synthetic promoter, the highly expressed Listeria promoter PHelp (Riedel et al., 2007). The synthesized construct was then sub-cloned into pPL2 using XhoI and PstI restriction enzymes (Figure 1A). The sequence of the insert was verified and the novel vector was then transformed and integrated into the L. monocytogenes EGDe wild type strain and an engineered L. monocytogenes variant (EGDeopt) expressing a murine-specific InIA (Monk et al., 2010). The pPL2 vector contains a PSA listeriophage integrase gene and attachment site that directs site-specific single copy integration into the tRNAAsp locus on the L. monocytogenes chromosome (Lauer et al., 2002). The successful integrants were then examined for integration using PCR and positive clones were screened phenotypically for light emission. A total of 10 candidate integrants were picked using PCR and positive clones were screened phenotypically for light emission. A total of 10 candidate integrants were picked using PCR and positive clones were screened phenotypically for light emission. A total of 10 candidate integrants were picked using PCR and positive clones were screened phenotypically for light emission.

The synthesized construct was then sub-cloned into pPL2 using the backbone of the pPL2 integrating plasmid. pPL2 contains a PSA listeriophage integrase gene and attachment site that directs site-specific single copy integration into tRNAAsp locus on the L. monocytogenes chromosome (Lauer et al., 2002). In our lab, we have previously successfully created pPL2-derivatives, pPL2lux and pPL2luxPHelp that allow BLI through expression of the bacterial lux operon (Bron et al., 2006; Riedel et al., 2007). Here we used a similar approach to create a vector expressing a CBR-luc with potential applications in deep tissue whole body imaging.

For this purpose, a codon optimized CBR-luc (CBRopt) gene for optimal expression in L. monocytogenes was synthesized under the influence of a constitutive synthetic promoter, the highly expressed Listeria promoter PHelp (Riedel et al., 2007). The synthesized construct was then sub-cloned into pPL2 using XhoI and PstI restriction enzymes (Figure 1A). The sequence of the insert was verified and the novel vector was then transformed and integrated into the L. monocytogenes EGDe wild type strain and an engineered L. monocytogenes variant (EGDeopt) expressing a murine-specific InIA (Monk et al., 2010). The pPL2 vector contains a PSA listeriophage integrase gene and attachment site that directs site-specific single copy integration into the tRNAAsp locus on the L. monocytogenes chromosome (Lauer et al., 2002). The successful integrants were then examined for integration using PCR and positive clones were screened phenotypically for light emission. A total of 10 candidate integrants were picked and screened for integration with results indicating that 9 (90%) were positive integrants. In parallel, we used the pPL2luxPHelp and pPL2lux (silent vector) (Riedel et al., 2007) as controls for integration and subsequent analysis of luminescence.

CBRopt Reporter Is Stably Integrated in L. monocytogenes and Does Not Impede Growth Rate

We tested the stability of the L. monocytogenes EGDe::pPL2CBRopt and EGDe::pPL2CBRopt strains in the absence of chloramphenicol. L. monocytogenes EGDe::pPL2CBRopt and EGDe::pPL2CBRopt were sub-cultured for at least 50 generations in BHI medium without antibiotic selection. Dilutions of the last passage of the cultures were plated and the presence of the integrated plasmids in the resulting colonies was evaluated by scoring chloramphenicol resistant colonies. Our results indicated that all 50 colonies tested after 50 generations were found to retain chloramphenicol resistance indicating that the integrating plasmid was stably maintained without antibiotic selection pressure. Furthermore, at least 10 random chloramphenicol-resistant colonies were also tested phenotypically for light emission and the results indicated that all 10 tested colonies were bioluminescent.

Previous work has indicated that integration of lux-expressing pPL2 constructs does not influence bacterial growth rate relative to wild type bacteria (Riedel et al., 2007). In the current study we evaluated the growth behavior of L. monocytogenes EGDe::pPL2CBRopt relative to EGDe::pPL2luxPHelp (Riedel et al., 2007) and EGDe::pPL2 (Bron et al., 2006). Strains were grown in parallel under standard conditions (Figure 1B). Our data indicated that the growth of L. monocytogenes EGDe::pPL2CBRopt was normal relative to the other strains tested (Figure 1B). Furthermore, the growth rates of L. monocytogenes EGDe::pPL2CBRopt were also not affected in BHI (in which pH was adjusted to 7.4) or LB medium (data not shown).

CBRopt Can Be Used for Tagging Clinical L. monocytogenes Strains

Listeria monocytogenes EGDe is the most commonly used laboratory strain for the analysis of listerial pathogenesis (Glaser et al., 2001). However, there is increasing interest in the analysis of wild type clinical isolates in order to determine strain specific differences in pathogenesis (Maury et al., 2016). For this purpose, three clinical strains (Table 1), all originating from patients who suffered meningitis and isolated from cerebrospinal fluid (CSF) were transformed with pPL2CBRopt. Our results indicated that all three wild type clinical strains could be successfully tagged with our newly developed plasmid (Supplementary Figure S1). After 30 passages of these strains in vitro 50 colonies were tested and all retained chloramphenicol resistance indicating that the system is as stable as observed in the EGDe laboratory strain. We also determined that the in vitro growth rate of these wild type field isolates in BHI broth was not affected by the presence of the integrated plasmid (data not shown). We transformed the clinical isolates with pPL2luxPHelp in order to provide a comparison with the CBRopt system. Results indicated that bioluminescent profile followed the same light emission pattern as observed for the EGDe strain as described below (Supplementary Figure S1).

CBR Signals Are Brighter and Produce Greater Flux Than lux During In Vitro Growth

We determined the bioluminescence profile of L. monocytogenes EGDe::pPL2CBRopt over time. L. monocytogenes EGDe::pPL2-CBRopt was grown in BHI broth for 24 h to monitor bioluminescence (Figures 2A,B). No bioluminescence was detected at the initial stage of growth. The bioluminescence profile increased as bacteria grew and bioluminescence reached a peak following 6 h of bacterial growth and remained relatively high during stationary phase. We directly compared the bioluminescence of L. monocytogenes EGDe::pPL2CBRopt with the L. monocytogenes EGDe::pPL2luxPHelp strain as both bioluminescent vectors are constructed using the pPL2 backbone with expression of luciferase under the same PHelp promoter system (Riedel et al., 2007). Bioluminescence signals of bacterial luciferase (L. monocytogenes EGDe::pPL2luxPHelp) were higher during initial growth as compared to the isolate.
expressing CBR\textsuperscript{opt}. However, the bioluminescence signal from \textit{L. monocytogenes} EGD::pPL2\textit{luxP}\textsubscript{HELP} diminished quickly as bacteria entered the stationary phase and the overall bioluminescence expression pattern for this strain was similar to our previously reported data (Riedel et al., 2007). A decrease in the bioluminescence signal during stationary phase may be a feature of light expression from the \textit{luxABCDE} system as the phenomenon has also been described for other Gram-positive bacteria including \textit{Streptococcus pneumonia} (Francis et al., 2001) and \textit{Lactobacillus casei} (Oozeer et al., 2005). It is possible that the diminished Lux signal during stationary phase may be related to decreased metabolic recycling of FMN to FMNH\textsubscript{2} (an essential component for bacterial Lux activity), and ATP production (Francis et al., 2001; Andreu et al., 2010). This hypothesis is partially supported by a previous study which demonstrated a 100-fold increase in signal after providing a gene (\textit{frp}) encoding a NADPH-FMN oxidoreductase in-trans with \textit{luxAB} enzymes in a yeast model system (Szittner et al., 2003). However, this limitation would not influence the activity of CBR, which in our system exhibits relatively robust activity in late exponential phase and stationary phase.

In order to measure light intensity per cell, bioluminescence signals were measured in parallel with determination of the total number of viable bacterial CFU. Our results indicated that CBR\textsuperscript{opt} signals were directly correlated with CFU ($r^2 = 0.9379$; \textbf{Figure 2C}). A similar positive correlation was observed between the OD\textsubscript{600} and total flux during growth in medium (\textbf{Figure 2D}). The strong positive correlation between CFU and total flux

---

**FIGURE 2** | Luminescence of tagged \textit{L. monocytogenes} during growth in media. (A) Luminescence imaging of luciferase-tagged \textit{L. monocytogenes} strains. Luminescent images were taken using the PerkinElmer IVIS Lumina II system and processed using Living Image software as indicated in the text. The color bar represents bioluminescent signal intensity (in photons/s/cm\textsuperscript{2}). (B) Luminescence (flux) of luciferase-tagged \textit{L. monocytogenes} strains during in vitro growth in BHI broth. Data and error bars represent mean and standard deviation, respectively, of triplicate samples for each time point. A two-way ANOVA with Bonferroni post-test was used to compare groups. (C) Luciferase-tagged \textit{L. monocytogenes} strains were grown to log phase and then serially diluted. Each dilution was imaged and also plated for CFU counts. The obtained data of total flux and CFU count was correlated. Statistical analysis showed a positive linear correlation (with $r^2 = 0.94$) between total flux emitted by CBR\textsuperscript{opt} and the luminescent bacterial. (D) \textit{L. monocytogenes} EGD::pPL2\textit{CBR}\textsuperscript{opt} were grown in BHI medium and images were obtained by IVIS at given OD\textsubscript{600} points. The data obtained was then analyzed that indicated a strong positive correlation between OD\textsubscript{600} and total flux of the luminescent bacteria. Linear regression analysis was used to determine the relationship between flux and OD\textsubscript{600} values in (C,D).
indicates that luminescence from CBR-luc tagged bacteria can successfully be used for accurate quantification of bacterial numbers during growth.

In order to directly compare signal intensity each luminescent strain was serially diluted until no luminescence signals were detectable (Figures 3A, B) and the CFU/ml of each representative dilution was calculated in order to determine relationship of total flux/CFU. The bioluminescence profile of both lux and CBR-luc luciferase reporters were indeed dependent on the number of the luminescent bacteria. Bioluminescence intensity was correlated with the number of bioluminescent bacteria both for EGD::pPL2 luxP HELP and EGD::pPL2 CBR opt during growth over time in BHI medium (Figure 3B) and the overall analysis indicated that CBR opt produces greater flux/CFU when compared with bacterial luciferase during growth in medium (p > 0.05) (Figure 3C).

**Bioluminescence Profile of CBR opt in an In Vitro Model of Varied Tissue Depth**

CBR-luc produces light of a wavelength (~600 nm) that is within the red region of the visible light spectrum. As red light has greater penetrating power in animal tissues, we predicted that light generated by CBR-luc would penetrate mammalian tissues more effectively than that of lux (in the blue spectrum at around 480 nm). We compared the ability of the CBR opt and Lux luciferase signals to penetrate through varying depths of tissue using a previously described in vitro assay (Mason et al., 2016). Layers of cut meat slices were used to mimic varying levels of tissue depth. An aliquot of 50 µl of cells were deposited into a round bottom 96-well plate and light measurements were repeated following the addition of layers of meat slices to generate tissue depth as indicated in Figure 4.
As expected, a decrease in light detection was associated with an increase in tissue depth. Signal transmission and total flux of bacterial luciferase (L. monocytogenes EGDe::pPL2luxPHELP) quickly declined as the thickness of the tissue increased as compared to CBRopt (L. monocytogenes EGDe::pPL2CBRopt). In particular, L. monocytogenes EGDe::pPL2CBRopt was clearly visible through multiple layers of raw beef steak whereas the penetrating power of light generated in L. monocytogenes EGDe::pPL2luxPHELP was poor and below detectable limits (Figures 4A,B).

**BLI of CBRopt during a Cell Invasion Assay**

The pPL2luxPHELP vector has previously proved effective for the imaging of L. monocytogenes infection in cultured mammalian cells (Riedel et al., 2007). In the current study, we investigated the bioluminescence profile of L. monocytogenes EGDe::pPL2CBRopt during a cell invasion assay in order to determine the possibility of using this vector for monitoring and detection of EGDe in cell culture. For this purpose, L. monocytogenes EGDe::pPL2CBRopt, L. monocytogenes EGDe::pPL2luxPHELP and L. monocytogenes EGDe::pPL2lux were used to infect C2Bbe1 cells for 1 h and the levels of bioluminescence and number of the intracellular bacteria were determined (Figures 5A,B). The efficacy of invasion did not differ between the strains and bacteria were isolated at similar levels from each well. However, the bioluminescence intensity indicated that L. monocytogenes EGDe::pPL2CBRopt produced a consistently brighter signal and significantly higher flux than L. monocytogenes EGDe::pPL2luxPHELP in the context of in vitro infection (Figures 5A,B). This suggests that the pPL2CBRopt vector may have applications for BLI of Listeria during infection assays, perhaps with the potential for use in high throughput assay systems.

**CBRopt Facilitates Robust In Vivo Whole Body Imaging of Murine Infection**

Murine infection is widely used for the analysis of factors contributing to the pathogenesis of L. monocytogenes (Lecuit, 2007; Cossart and Toledo-Arana, 2008). A number of pathogens such as Trypanosoma brucei (Van Reet et al., 2014), Mycobacterium tuberculosis (Chang et al., 2014), and...
and Lactobacillus species (Daniel et al., 2015) have been previously tagged with CBR reporters that provided enhanced whole body images in murine infection model. Given the improved signal intensity mediated by CBR during in vitro growth, in particular during late exponential and stationary-phase, we examined the signal intensity from L. monocytogenes EGDe::pPL2CBRopt during murine infection. For this purpose, three groups of 12 mice were infected with murinized L. monocytogenes EGDe::pPL2CBRopt, murinized L. monocytogenes EGDe::pPL2luxPHELP, and murinized EGDe::pPL2lux with a standard inoculum (2 × 10^6 CFU/ml) via the intraperitoneal route. Four mice were left uninfected to serve as the control group. Four mice from each group were then imaged at 24 h intervals post-infection until 72 h. For the mice infected with L. monocytogenes EGDe::pPL2CBRopt luciferin was injected intraperitoneally 10 min prior to imaging. After imaging, mice were sacrificed and the resected organs (liver, spleen, and kidneys) were imaged for bioluminescence and further processed for counting the number of viable bacteria. Results of the whole body imaging of mice indicated that no bioluminescence was observed 24 h post-infection in any of the strains. No bioluminescence signals were detectable after dissecting the mice and imaging individual organs at this time point. This lack of bioluminescence correlated well with the lower number of viable bacteria recovered from the resected organs of the infected mice. Of these, the highest number of CFU was recovered from the spleen where CFU/ml were in the range of 1 × 10^3.

Notably, mice infected with L. monocytogenes EGDe::pPL2CBRopt produced detectable bioluminescence signals at 48 h post-infection, while no signals were detected from mice infected with L. monocytogenes EGDe::pPL2luxPHELP or the L. monocytogenes EGDe::pPL2lux control. We acknowledge that there was some variability in observable signals between individual mice (when imaged at the specified sensitivity) as noted in previous studies by other groups (Poulsen et al., 2011). The number of CFU in these animals reached approximately 1 × 10^6 CFU/ml in the spleen and liver. Importantly, there was no significant difference in CFU/ml among the infected strains recovered from the infected mice post-mortem in resected organs (results not shown). Interestingly, at 48 h infection bioluminescent signals were detected in the resected liver and spleen of mice infected with L. monocytogenes EGDe::pPL2luxPHELP, but no signals were detectable during whole body imaging, indicating that these signals were attenuated during whole body imaging, potentially as a consequence of the low penetrating power of the shorter wavelength light emitted by Lux. In contrast, bioluminescence signals were observed during whole body imaging of mice infected with L. monocytogenes EGDe::pPL2CBRopt at 48 h post-infection in addition to a clear signal from post-mortem resected organs (Figure 6).

In a tandem study using infection with wild type (non-murinized) EGDe we also noted a clearly enhanced signal from mice infected with CBRopt labeled bacteria relative to mice infected with Lux-tagged EGDe (Supplementary Figure S2). In this experiment, bacterial burdens in the internal organs were comparable between the groups though the infectious load in the organs was noticeably lower than mice infected with EGDe (not shown). Again this suggests that the signal from CBRopt may provide greater sensitivity at low bacterial loads.

Overall, the ability to detect luminescence signals in mice infected with L. monocytogenes EGDe::pPL2CBRopt at 48 h suggested that pPL2CBRopt provides for increased sensitivity in the context of whole body imaging. At higher bacterial numbers on day 3 of infection the difference in signal intensity in whole body imaging between the Lux and CBR-labeled bacteria was less evident (Figure 6).

CONCLUSION

In conclusion, we have constructed a system for the stable chromosomal integration of constitutively expressed CBR luciferase in L. monocytogenes. In comparison to a L. monocytogenes strain expressing bacterial luciferase (LuxAB), CBR provides a greater luminescence signal during the stationary phase of growth and better penetrating power in an in vitro tissue depth model. We also noted enhanced sensitivity during murine...
infection and in an *in vitro* cell infection assay. A limitation of using CBR is the necessity to add luciferin substrate to the system. This adds an extra variable, as the availability of substrate in complex systems would certainly affect signal intensity. In contrast bacterial *lux* systems encode the capacity to generate endogenous substrate thereby potentially eliminating this extra variable. Given the different emission spectra of each system there is significant potential to utilize both in combination in order to measure dual bacterial populations in complex systems as has been described recently (Daniel et al., 2015). Overall,
we suggest that the pPL2CBR<sup>opt</sup> vector may have significant applications for the study of *L. monocytogenes* in various model systems.

**AUTHOR CONTRIBUTIONS**

SUR and CG wrote the paper. SUR, MS, PC, AS, GB, CH, KF, MT, and CG proofread the paper. SUR, MS, PC, AS, GB, CH, KF, MT, and CG designed experiments. SUR, MS, PC, and AS carried out experiments. SUR, MS, PC, AS, GB, CH, KF, MT, and CG analyzed experiments.

**FUNDING**

This work was funded by an Industry Academia Partnerships and Pathways grant (Vaccines and Imaging Partnership) as part of the European Commission Seventh Framework Programme (PIAP-GA-2013-612219-VIP). The authors acknowledge the financial support of the APC Microbiome Institute funded through Science Foundation Ireland (SFI) under Grant Number SFI/12/RC/2273.

**REFERENCES**

Andreu, N., Zelmer, A., Fletcher, T., Elkington, P. T., Ward, T. H., Ripoll, J., et al. (2010). Optimisation of bioluminescent reporters for use with mycobacteria. *PLOS ONE* 5:e10777. doi: 10.1371/journal.pone.0010777

Belkin, S., Smulski, D. R., Vollmer, A. C., Van Dyk, T. K., and LaRossa, R. A. (1996). Oxidative stress detection with *Escherichia coli* harboring a katG::lux fusion. *Appl. Environ. Microbiol.* 62, 2252–2256.

Bergmann, S., Rohde, M., Schughart, K., and Lengeling, A. (2013). The bioluminescent *Listeria monocytogenes* strain Xen32 is defective in flagella expression and highly attenuated in orally infected BALB/c mice. *Gut Pathog.* 5:19. doi: 10.1186/1757-4749-5-19

Bron, P. A., Monk, I. R., Corr, S. C., Hill, C., and Gahan, C. G. (2006). Novel luciferase reporter system for in vitro and organ-specific monitoring of differential gene expression in *Listeria monocytogenes*. *Appl. Environ. Microbiol.* 72, 2876–2884. doi: 10.1128/AEM.72.4.2876-2884.2006

Byrne, W. L., DeJille, A., Kuo, C., de Jong, J. S., van Dam, G. M., Francis, K. P., et al. (2013). Use of optical imaging to progress novel therapeutics to the clinic. *J. Control. Release* 172, 523–534. doi: 10.1016/j.jconrel.2013.05.004

Chang, M., Anttonen, K. P., Cirillo, S. L., Francis, K. P., and Cirillo, J. D. (2014). Real-time bioluminescent imaging of mixed mycobacterial infections. *PLOS ONE* 9:e108341. doi: 10.1371/journal.pone.0108341

Contag, C. H., and Bachmann, M. H. (2002). Advances in in vivo bioluminescence imaging of gene expression. *Annu. Rev. Biomed. Eng.* 4, 235–260. doi: 10.1146/annurev.bioeng.4.1.235

Cossart, P., and Toledo-Arana, A. (2008). *Listeria monocytogenes*, a unique model in infection biology: an overview. *Microbes Infect.* 10, 1041–1050. doi: 10.1016/j.micinf.2007.08.043

Cronin, M., Akin, A. R., Collins, S. A., Meganck, J., Kim, J. B., Baban, C. K., et al. (2012a). High resolution in vivo bioluminescent imaging for the study of bacterial tumour targeting. *PLOS ONE* 7:e30940. doi: 10.1371/journal.pone.0030940

Cronin, M., Stanton, R. M., Francis, K. P., and Tangney, M. (2012b). Bacterial vectors for imaging and cancer gene therapy: a review. *Cancer Gene Ther.* 19, 731–740. doi: 10.1038/cgt.2012.59

Daniel, C., Poiret, S., Denvin, V., Boutillier, D., Lacorre, D. A., Foligne, B., et al. (2015). Dual-color bioluminescence imaging for simultaneous monitoring of

**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: http://journal.frontiersin.org/article/10.3389/fmicb.2017.01797/full#supplementary-material

**FIGURE S1** | Bloluminescence profile of clinical listerial strains labeled with CBR<sup>opt</sup> reporter. EGDe-4b-130029:pPL2CBR<sup>opt</sup>, EGDe-4b-130003:pPL2CBR<sup>opt</sup>, and EGDe-4b-150008:pPL2CBR<sup>opt</sup> (see Table 1) were integrated with the newly developed codon optimized (CBR<sup>opt</sup>) reporter. The tagged strains were then grown in BHI medium and were recorded at different time intervals. The overall bioluminescence profile is shown. Data and error bars represent mean and standard deviation, respectively, of triplicate samples for each time point. Asterisks *<i>p</i>*<i> </i>*<i> </i>*<i> </i>*<i> </i>indicates *<i>p</i>* < 0.05 based upon Student's *t*-test.

**FIGURE S2** | In vivo BLI of mice infected with wild type *L. monocytogenes* EGDe. (A) Whole body luminescence image of BALB/c mice infected with wild type *L. monocytogenes* EGDe tagged with either bacterial Lux or CBR<sup>opt</sup>. Mice were infected ip with a dose of 2 × 10<sup>9</sup> bacteria and were imaged for luminescence at 72 h post-infection. A clear and robust signal is detectable from mice infected with CBR<sup>opt</sup> tagged bacteria at this timepoint. A luminescent signal that exceeded background (uninfected animals) was detected in every *L. monocytogenes* EGDe CBR<sup>opt</sup> infected mouse. The number of CFU/ml recovered from infected mice were not significantly different between both groups (not shown). (B) Graph showing the signal intensity from the whole body images obtained at 72 h from infected mice. Luminescence is shown as total flux (p/s/cm<sup>2</sup>/sr). Data and error bars represent mean and standard error.

Gahan, C., and Hill, C. (2005). Gastrointestinal phase of *Listeria monocytogenes* infection. *J. Appl. Microbiol.* 98, 1345–1353. doi: 10.1111/j.1365-2672.2005.02559.x

Hardy, J., Chu, P., and Contag, C. H. (2009). Foci of *Listeria monocytogenes* persist in the bone marrow. *Dis. Model Mech.* 2, 39–46. doi: 10.1242/dmm.000836

Hardy, J., Francis, K. P., DeBoer, M., Chu, P., Gibbs, K., and Contag, C. H. (2004). Extracellular replication of *Listeria monocytogenes* in the murine gall bladder. *Science* 303, 851–853. doi: 10.1126/science.1092712

Karimi, S., Ahl, D., Vagesjo, E., Holm, L., Phillipson, M., Jonsson, H., et al. (2016). In vivo and in vitro detection of luminescent and fluorescent *Lactobacillus reuteri* and application of red fluorescent mCherry for assessing plasmid persistence. *PLOS ONE* 11:e0151969. doi: 10.1371/journal.pone.0151969

Kim, H., Boor, K. J., and Marquis, H. (2004). *Listeria monocytogenes* contributes to invasion of human intestinal epithelial cells. *Infect. Immun.* 72, 7374–7378. doi: 10.1128/IAI.72.12.7374-7378.2004
Lauer, P., Chow, M. Y., Loessner, M. J., Portnoy, D. A., and Calendar, R. (2002). Construction, characterization, and use of two Listeria monocytogenes site-specific phage integration vectors. J. Bacteriol. 184, 4177–4186. doi: 10.1128/ JB.184.15.4177-4186.2002

Lebreton, A., Stavr, F., Brisse, S., and Cossart, P. (2016). 1926-2016: 90 Years of listeriology. Microbes Infect. 18, 711–723. doi: 10.1016/j.micinf.2016.10.009

Lecuit, M. (2007). Human listeriosis and animal models. Microbes Infect. 9, 1216–1225. doi: 10.1016/j.micinf.2007.05.009

Mason, E. A., Lopez, R., and Mason, R. P. (2016). Wavelength shifting of chemiluminescence using quantum dots to enhance tissue light penetration. Opt. Mater. Express 6, 1384–1392. doi: 10.1364/OME.6.001384

Maury, M. M., Tsai, Y. H., Charlier, C., Touchon, M., Chenal-Francisque, V., Leclercq, A., et al. (2016). Uncovering Listeria monocytogenes hypervirulence by harnessing its biodiversity. Nat. Genet. 48, 308–313. doi: 10.1038/ng.3501

Mead, P. S., Slutsker, L., Dietz, V., and McCaig, L. F. (2000). Food-related illness associated with Listeria monocytogenes. J. Bacteriol. 182, 7117–7122. doi: 10.1128/JB.182.13.7117-7122.2000

Park, S. F., and Stewart, G. S. (1990). High-efficiency transformation of Listeria monocytogenes by electroporation of penicillin-treated cells. Gene 94, 129–132. doi: 10.1016/0378-1119(90)90479-B

Poulsen, K. P., Faith, N. G., Steinberg, H., and Czuprynski, C. J. (2011). Pregnancy reduces the genetic resistance of C57BL/6 mice to Listeria monocytogenes infection by intragastric inoculation. BMC Microbiol. 10:318. doi: 10.1186/1471-2180-10-318

Oozeer, R., Furet, J., Goupil-Feuillerat, N., Anba, J., Mengaud, J., and Corthier, G. (2005). Differential activities of four Lactobacillus casei promoters during bacterial transit through the gastrointestinal tracts of human-microbiota-associated mice. Appl. Environ. Microbiol. 71, 1356–1363. doi: 10.1128/AEM.71.13.1356-1363.2005

Park, S. F., and Stewart, G. S. (1990). High-efficiency transformation of Listeria monocytogenes by electroporation of penicillin-treated cells. Gene 94, 129–132. doi: 10.1016/0378-1119(90)90479-B

Poulsen, K. P., Faith, N. G., Steinberg, H., and Czuprynski, C. J. (2011). Pregnancy reduces the genetic resistance of C57BL/6 mice to Listeria monocytogenes infection by intragastric inoculation. BMC Microbiol. 10:318. doi: 10.1186/1471-2180-10-318

Quereda, J. J., Dussurget, O., Nahori, M. A., Ghozlane, A., Volant, S., Dillies, M. A., et al. (2016). Bacteriocin from epidemic Listeria strains alters the host intestinal microbiota to favor infection. Proc. Natl. Acad. Sci. U.S.A. 113, 5706–5711. doi: 10.1073/pnas.1523899113

Rea, R. B., Gahan, C. G., and Hill, C. (2004). Disruption of putative regulatory loci in Listeria monocytogenes demonstrates a significant role for Fur and PerR in virulence. Infect. Immun. 72, 717–727. doi: 10.1128/IAI.72.2.717-727.2004

Rice, B. W., Cable, M. D., and Nelson, M. B. (2001). In vivo imaging of light-emitting probes. J. Biomed. Opt. 6, 432–440. doi: 10.1117/1.1415210

Riedel, C. U., Monk, I. R., Casey, P. G., Morrissey, D., O’Sullivan, G. C., Tangney, M., et al. (2006). Improved luciferase tagging system for Listeria monocytogenes allows real-time monitoring in vivo and in vitro. Appl. Environ. Microbiol. 73, 3091–3094. doi: 10.1128/aeom.2004-09

Riedel, C. U., Monk, I. R., Casey, P. G., Waidmann, M. S., Gahan, C. G., and Hill, C. (2009). AgrD-dependent quorum sensing affects biofilm formation, invasion, virulence and global gene expression profiles in Listeria monocytogenes. Mol. Microbiol. 71, 1177–1189. doi: 10.1111/j.1365-2958.2008.06589.x

Sleator, R. D., Watson, D., Hill, C., and Gahan, C. G. (2009). The interaction between Listeria monocytogenes and the host gastrointestinal tract. Microbiology 155(Pt 8), 2463–2475. doi: 10.1099/mic.0.030205-0

Szittner, R., Jansen, G., Thomas, D. Y., and Meighen, E. (2003). Bright stable luminescent yeast using bacterial luciferase as a sensor. Biochem. Biophys. Res. Commun. 309, 66–70. doi: 10.1016/S0006-291X(03)01530-4

Tangney, M., and Francis, K. P. (2012). In vivo optical imaging in gene & cell therapy. Curr. Gene Ther. 12, 2–11. doi: 10.2174/156652312799789299

Van Pijkeren, J. P., Morrissey, D., Monk, I. R., Cronin, M., Rajendran, S., O’Sullivan, G. C., et al. (2010). A novel Listeria monocytogenes-based DNA delivery system for cancer gene therapy. Hum. Gene Ther. 21, 405–416. doi: 10.1089/hum.2009.022

Van Reet, N., Van de Vyver, H., Pyana, P. P., Van der Linden, A. M., and Buscher, P. (2014). A panel of Trypanosoma brucei strains tagged with blue and red-shifted luciferases for bioluminescent imaging in murine infection models. PLOS Negl. Trop. Dis. 8:e3054. doi: 10.1371/journal.pntd.0003054

Vorobyeva, A. G., Stanton, M., Godinat, A., Lund, K. B., Karateev, G. G., Francis, K. P., et al. (2015). Development of a bioluminescent nitroreductase probe for preclinical imaging. PLOS ONE 10:e0131637. doi: 10.1371/journal.pone.0131637

Zhou, H., Doyle, T. C., Coquoz, O., Kalish, F., Rice, B. W., and Contag, C. H. (2005). Emission spectra of bioluminescent reporters and interaction with mammalian tissue determine the sensitivity of detection in vivo. J. Biomed. Opt. 10, 41201. doi: 10.1117/1.2032388

Conflict of Interest Statement: AS and GB are permanent employees of the GSK group of companies. GB reports ownership of GSK restricted GSK shares. The other authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Ur Rahman, Stanton, Casey, Spagnuolo, Bensi, Hill, Francis, Tangney and Gahan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.