The study of the pharmacological action of the dry extract from *Prunus domestica* fruits on the model of comorbid functional constipation in the combined alcoholic liver damage in rats

The search for the most effective hepatoprotective agents is a topical issue of modern medicine. As a rule, the plant raw material is a promising object for studying medicinal properties. Based on the literature data, we are interested in the horticultural crop of *Prunus domestica*, the family Rosaceae, which is sufficiently cultivated and attracts attention by its pharmacoeconomic component.

**Aim.** In the previous experimental studies, the *Prunus domestica* extract containing fibers was found to have the laxative, hepatoprotective and probiotic effects. The foregoing facts provided an opportunity to study the *Prunus domestica* extract containing fibers for activity in comorbidity of some relevant pathologies and experimentally substantiate the prospects of its application as a medicine of choice for constipation in patients with pathologies of the digestive system.

**Materials and methods.** The study object was the dry extract from *Prunus domestica* fruits containing fibers. The experiment was performed in stages. At the first stage, the experimental subacute alcoholic liver damage was caused in animals. At the second stage of the experiment, functional constipation was modeled in animals. In the course of subacute hepatitis and during the second phase of the experiment animals were introduced the test extract in the dose of 200 mg/kg and the reference drug – a hepatoprotector *Silybor* in the dose of 25 mg/kg. At the second stage of the study, a laxative Senadexin in the dose of 14 mg/kg was added to the therapy. All substances were introduced orally into the body of the experimental animals. To assess the motor activity of the intestine, the method of Sagar et al., modified by Choi et al., was used. The motor activity of the small intestine was determined according to the corresponding calculation formula. The functional state of the liver was assessed by the biochemical parameters in the blood serum: the total protein, urea and ALT activity.

**Results.** The analysis of the experimental data has shown that the test extract exhibits a moderate laxative effect. In animals of the intact group, the relevant signs (a pronounced laxative effect) indicate the development of diarrhea, which is not an adequate solution to the development of constipation. When using the extract from *Prunus domestica* fruits during the whole period of the experiment the normalization of all indicators characterizing the intestinal motility was observed. The analysis of the experimental data has demonstrated that the phytoobject studied has a soft laxative effect, mainly due to the improvement of the intestinal motility. When studying the functional state of the liver against the background of the introduction of the extract studied the positive changes in the content of markers of liver damage (decrease in the urea concentration and decrease in the activity of ALT in the animal serum) were detected compared to the control pathology group of animals No. 2.

**Conclusions.** The experimental data obtained on the study of the correlation mechanisms of the hepatoprotective and laxative activity of the extract from *Prunus domestica* fruits containing fibers indicate the presence of a soft laxative effect that occurs by intensifying the intestinal motility, as well as the pronounced hepatoprotective action. These effects of the extract studied were revealed when using it in the treatment of constipation against the background of the experimental subacute liver damage, thus not being inferior the effects studied in the complex application of the hepatoprotective and laxative reference drugs. The key conclusion is that the extract studied, in contrast to the reference drug Senodexin, does not cause signs of diarrhea in its laxative effect, which can be a distinctive, positive feature in further clinical studies. This herbal object can be promising and rational when using it with the single-stage complex therapy as a hepatoprotective and laxative agent in the treatment of comorbid conditions in gastroenterology associated with diseases of the liver and congestive intestinal phenomena.

**Key words:** *Prunes Domestica* fruits; hepatoprotectors; comorbid functional constipation; subacute liver damage; urea; ALT
Исследование фармакологического действия сухого экстракта плодов Сливы на модели коморбидного функционального запора на фоне комбинированного алкогольного поражения печени у крыс

Посы наиболее эффективных гепатопротекторных средств является актуальной проблемой современной медицины. Как правило, перспективными объектами для изучения лечебных свойств является растительное сырье. Исходя из литературных данных нас заинтересовала садовая культура Слива домашняя (лат. Prunus domestica), семейства Rosaceae, которая достаточно культуривается, а также привлекает внимание своей фармакоэкономической составляющей.

Цель исследования. В предыдущих экспериментальных исследованиях экстракт Сливы домашней, который содержал волокна, был установлен как слабительный, гепатопротекторный и пробиотический эффекты. Высказанное дало возможность исследовать экстракт Сливы домашней, содержащих волокна, на активность при коморбидизации нескольких соответствующих патологий и экспериментально обосновать перспективу его применения в качестве средства выбора при запорах у пациентов с патологиями травной системы.

Материалы и методы. Объектом исследований был выбран сухой экстракт плодов Сливы домашней, содержащий волокна. Эксперимент проводили поэтапно. На первом этапе у тварин вживили экспериментальное підстрко алкогольне ураження печінки на другому етапі експерименту тваринам вводили функціональну запору. У період перебігу підострого гепатозу та впродовж другої фази експерименту тваринам вводили досліджувані екстракти у дозі 200 мг/кг і препарат порівняння вітчизняний гепатопротектор сілібор у дозі 25 мг/кг.

Результаты. Отмечены экспериментальные данные, которые указывают на развитие диареи, что не является адекватным решением развития запоров. У животных интактной группы соответствующие признаки (выраженный слабительный эффект) указывают на то, что исследуемый экстракт проявляет умеренное слабительное действие. У животных экспериментальных групп соответствующие признаки (выраженный слабительный эффект) указывают на то, что исследуемый экстракт проявляет умеренное слабительное действие. У животных экспериментальных групп соответствующие признаки (выраженный слабительный эффект) указывают на то, что исследуемый экстракт проявляет умеренное слабительное действие.
The search for the most effective hepatoprotective agents is a topical issue of modern medicine. As a rule, the plant raw material is a promising object for studying medicinal properties [1]. Based on the literature data, we are interested in the horticultural crop of Prunus domestica, the family Rosaceae, which is sufficiently cultivated and attracts attention by its pharmacoeconomic component [2, 3].

In the previous studies, the qualitative and quantitative chemical composition of the Prunus domestica fruits was determined. The presence of anthocyanins, rutin, gallic acid, sugars, organic acids (malate, citrate, chlorogenic, neochlorogenic, carboxylic acids), dietary fibers and hydroxycinnamic acids in the raw material was proven. [4-7].

Since in the previous experiments the dry extract from Prunus domestica fruits containing fiber (DECF) demonstrated the presence of both the hepatoprotective and laxative effects, it was expedient to study its activity in comorbidity of some relevant pathologies and experimentally substantiate the prospects of its application as a medicine of choice for constipation in patients with pathologies of the hepatobiliary system [8, 9].

It should be noted that functional constipation is often observed in patients with liver disease, namely alcoholic and non-alcoholic fatty liver disease, liver toxicity, liver failure, liver fibrosis and cirrhosis; in turn, this constipation is a factor in the progression of the underlying disease and adversely affects on the quality of life of such patients [10-13].

Materials and methods

Experimental animals. The study was conducted on 40 white non-linear male rats with the body weight of 200-220 g randomized to minimize differences in the body weight according to the EU Council Directive 2010/63/EU on compliance with the laws, regulations and administrative provisions of the EU Member States on the protection of animals used for experimental and other scientific purposes [14]. Prior to the experiment, the animals were housed under standard conditions in the Central Research Laboratory of the National University of Pharmacy (NPhU) in natural daylight and day-night mode with free access to water and food [15].

The experiment design. Animals were evenly divided into 5 experimental groups (n=8) according to the following design scheme of the experiment:

- the first group – animals without any pathology during the experiment (intact control (IC));
- the second group – animals without liver pathology at the first stage of the experiment, but induced functional constipation at the second stage (control pathology No. 1 (CP-1));
- the third group – animals with induced liver pathology at the first stage of the experiment and induced functional constipation at the second stage (control pathology No. 2 (CP-2));
- the fourth group – animals with induced liver pathology at the first stage of the experiment and induced functional constipation at the second stage (test group). As treatment within the experiment these animals received DECF;
- the fifth group – animals with induced liver pathology at the first stage of the experiment and induced functional constipation at the second stage (reference group (RG)). As a treatment the animals received Silybor as a hepatoprotector and Senedaxin as a laxative.

At the first stage of the experiment the experimental subacute combined alcoholic liver damage was induced in animals of the corresponding experimental groups. Animals from the corresponding groups, in which the pathology was modeled, were subcutaneously injected 50 % oily solution of tetrachloromethane in the dose of 4 ml/kg for 4 days, followed by the intravenous injection of 40 % aqueous ethanol solution in the dose of 13 ml/kg in 3 hours. During the induction of this model the hepatoprotectors studied were administered 1 hour before the introduction of hepatotoxins. Synergy with combined administration of tetrachloromethane...
and ethanol allows inducing a steady damage of the liver parenchyma, which is similar to chronic alcoholic hepatitis, in the short term [14, 15]. The advantage of the subacute liver damage model compared to the chronic and subchronic ones in this case is in avoiding alcohol-associated diarrhea with the prolonged use of ethanol, which could affect the interpretation of the results of the second stage of the experiment [16].

During the second stage of the experiment, functional constipation was induced in animals by daily intragastric administration of loperamide hydrochloride (PhC "Zdorovya", Ukraine) in the dose of 3 mg/kg for 6 days 1 hour prior to the introduction of the test samples. Loperamide-induced retention of feces in the large intestine in rats is equivalent to the state of spastic constipation and corresponds to functional constipation with a decrease in peristalsis and absorption of fluid in humans [17, 18].

During induction of subacute hepatitis and the second stage of the experiment animals were treated with hepatoprotectors, which were administered daily for 10 days at 10:00-11:00. As in the previous experiments, the hepatoprotector Silybor (PhC "Zdorovya", Ukraine) was used as a reference drug in the dose of 25 mg/kg, which was equivalent to ED_{30} of this medicine [19]. Since in the previous studies the maximum hepatoprotective effect of DECF was demonstrated in the dose of 200 mg/kg, in this experiment a similar dose was used to obtain both specific effects. At the second stage of the experiment the rats of the reference group received additionally Senadexin (PHC "LUBNIFARM", Ukraine) in the dose of 14 mg/kg (calculated with reference to the content of calcium sennosides A and B). The animal-equivalent dose was calculated from the average daily dose for humans, the interspecies weight difference, and the body surface area [20]. Taking into account the lack of data on the drug interaction of both reference drugs both samples were administered within one period (10.00-11.00) in order to maintain equal conditions of the experiment; however, due to the phased introduction of medications between receiving drugs it took 30 minutes. In addition, such administration regimen allows us to reproduce the peculiarities of the patients’ compliance in polypragmasy against the background of comorbid states [21].

All drugs studied in this experiment were dissolved or suspended in 1 ml of purified water and injected with a special metallic enteral probe. Animals of control groups received the corresponding amount of physiological saline solution.

On day 5 of the administration of loperamide fecal boluses of rats were collected, and their parameters were determined. The day before, animals were placed in individual cages with free access to food and drinking water according to the hygienic requirements of the experiment. After collecting feces from each animal the day before the average number was calculated, and the moisture content was determined. Boluses were dried in a dry oven until a complete evaporation of moisture. The percentage moisture content was calculated according to the formula:

\[ C_m(\%) = \left( \frac{M_m - M_d}{M_m} \right) \times 100, \]

where \( C_m \) – is the fecal water content, %; \( M_m \) – is the fecal wet mass, g; \( M_d \) – is the fecal dry mass, g.

18 hours before the last administration of the drugs studied animals were deprived of free access to food, but consumption of drinking water remained ad libitum. To assess the motor activity of the intestine in rats, the method proposed by Sagar et al., modified by Choi et al., and adapted for this species of laboratory animals [17, 22] was used. In 10 min after the last administration of the samples all animals were given a black contrast mass in the form of 3 % suspension of activated charcoal in 0.5 % aqueous methylcellulose solution of 1 ml per animal.

In 30 min after the introduction of the contrast mass animals were euthanized by decapitation under inhalation anesthesia, their blood was collected to obtain the serum, the body was dissected, and the small intestine was removed from the pyloric sphincter to the caecum. The intestine was unfolded into a straight line, its total length, as well as the distance that the contrast mass passed, were measured. The motor activity of the small intestine was determined by the percentage indicator of the distance of the contrast mass accumulation according to the formula:

\[ L_{sp}(\%) = \left( \frac{L_p}{L_t} \right) \times 100, \]

where \( L_{sp} \) – is the indicator of the relative distance that the contrast mass passed, %; \( L_p \) – is the total length of the small intestine, cm. \( L_p \) – is the path passed by the contrast mass along the intestine, cm.

Together with this, the colon was removed in animals, and the total number of fecal boluses remained in the lumen of the intestine was measured.

The condition of the liver was assessed by biochemical parameters in the blood serum – the content of total protein, urea, and alanine aminotransferase (ALT) activity were determined. These indicators allow assessing the functional state of the liver in relation to its main functions (protein synthesizing, detoxifying), as well as the degree of damage.

The content of total protein in the serum was determined quantitatively by the Biuret method using the standard set of reagents "Total protein (Biuret with a calibrator)" HP010.01 ("Filitsit-Diagnostika", Ukraine) according to the instructions for use.
The content of urea in the serum was determined by the diacetylmiooxim method using the standard reagent set "Urea-D (Diacetylmiooxymium with a calibrator)" HP018.01 ("Filitsit-Diagnostika", Ukraine) according to the instructions for use.

The activity of ALT was determined by the colorimetric method using the standard set of reagents "ALT-KIN (kinetic method)" HP001.02 ("Filitsit-Diagnostika", Ukraine) according to the instructions for use.

The experimental data were processed by the standard program package "Statistica 6.0" using the ANOVA (t-criterion) algorithm for dispersion analysis. Verification of samples for normality and comparability was performed within the framework of the program algorithm [23].

**Results and discussion**

The use of loperamide hydrochloride in the dose of 3 mg/kg for 5 days resulted in virtually identical disorders in the work of the digestive tract in healthy animals (CP-1 group) and in animals with subacute alcoholic liver damage (CP-2 group). All animals in these groups experienced functional constipation, which was reflected in a decrease in the number of bowel movements per day – indicating inhibition of the intestinal motility, and the percentage of water in fecal boluses – indicating inhibition of absorption and/or increased water secretion in the intestinal cavity (p<0.05 versus the intact control).

When studying the effect of DECF on the defecation parameters in animals it was demonstrated that the test sample had a marked laxative effect without provoking secretory diarrhea. After application of the drug for 4 days against the background of induction of liver damage and for 5 days on the background of the introduction of loperamide all the defecation parameters studied probably did not differ from the similar values in the group of the intact control (p<0.05, Tab. 1).

In the reference group the introduction of Silybor in the dose of 25 mg/kg and Senodexin in the dose of 14 mg/kg to animals significantly changed the defecation parameters. The results obtained in this group indicate an expressive laxative effect (apparently due to Senodexin). The combined therapy in rats in the reference group significantly increased the number of defecations compared to animals of the control pathology No. 1 and control pathology No. 2 groups. At the same time, the overall average mass of fecal boluses, in particular due to the increased fluid content (p≤0.05 versus the intact control), excessively increased. It should be noted that the fecal masses in animals of the reference group were poorly formed and difficult to separate. The signs registered in animals of the reference group indicate the constipation correction due to the development of secretory diarrhea, which is not an optimal solution to the problem of the constipation development (Tab. 1).

In the experimental study of the parameters of the gastrointestinal motility it was shown that loperamide probably suppressed motility in the small intestine and caused the retention of the fecal masses in the large intestine in the groups of the control pathology No. 1 and control pathology No. 2. It should be noted that the combination of model pathologies into a complex disorder contributed to the moderate unlikely suppression of peristalsis compared to the isolated model of constipation. This fact correlates with the literature data [10] (Tab. 2).

The use of DECF in the dose of 200 mg/kg throughout the experiment contributed to the normalization of all indicators of the intestinal motility studied, which did not statistically differ from the similar parameters in the intact control group (p>0.05). In this case, according to the experimental data, it is obvious that the extract under research exhibits a mild laxative effect, mainly due to the improvement of the intestinal motility (Tab. 2).

---

**Table 1**

| Experimental group (n=8) | The number of fecal boluses | The fecal wet mass, g | The fecal dry mass, g | % of water |
|-------------------------|-----------------------------|----------------------|----------------------|------------------|
| Intact control          | 32.1±3.2                    | 6.51±0.64            | 3.98±0.44            | 39.50±2.27      |
| Control pathology No. 1 | 21.5±2.6*                   | 4.48±0.29*           | 3.31±0.21            | 25.93±1.31*     |
| Control pathology No. 2 | 18.5±2.3*                   | 5.01±0.35            | 3.70±0.29            | 26.43±1.48*     |
| DECF, 200 mg/kg         | 29.8±3.5*                   | 5.84±0.58            | 3.88±0.43            | 33.00±2.67**    |
| Silybor, 25 mg/kg + Senadexin, 14 mg/kg | 40.9±4.0**/3   | 13.69±1.38**/3      | 4.43±0.51            | 67.61±1.28**/3  |

Notes:
1) * – probable differences in relation to animals of the intact control group (p≤0.05);
2) ** – probable differences in relation to animals of the control pathology group No. 1 (p≤0.05);
3) # – probable differences in relation to animals of the control pathology group No. 2 (p≤0.05).
The combined therapy with a hepatoprotector and Senadexin also contributed to the improvement of the gastrointestinal motility. However, at the same time, the average indicator of the relative distance that the contrast mass passed through the small intestine was less than in the intact control, and a significantly smaller number of residual fecal boluses in the lumen of the large intestine were recorded. On the one hand, it may indicate episodic diarrhea against the background of motility inhibition in the small intestine, which is observed in some patients with liver cirrhosis [10]. On the other hand, the less motility of the small intestine may be due to the development of manifestations of tolerance to sennosides, and it is a characteristic feature of the long-term therapy. In addition, the probable cause may be an increase in the content of loperamide in the blood because of greater depression of the liver function. In any case, the mitigating effect of this therapy was characterized by the increased secretion and decreased reabsorption of the fluid in the large intestine; thus, secretory diarrhea (by the action of Senodexin) compensated for constipation. This was indicated by the most probable decrease in the number of fecal boluses in the lumen of the large intestine, even compared to the intact animals (p≤0.05), among all groups. In addition, it should be noted that the fecal masses in the large intestine of the reference group was amorphous, poorly textured and formed; they were separated only by peristalsis zones, which was also a functional sign of diarrhea (Tab. 2).

When studying the functional state of the liver by biochemical indicators it was demonstrated that double pathology contributed to a significant disorder of the protein synthesizing and detoxifying functions of the liver, which in turn, led to hepatocyte cytolysis. It should be noted that the use of loperamide in the isolated control pathology group No. 1 did not lead to these severe disorders of the liver, but the urea content in the serum of these animals was significantly higher than in the intact control (p≤0.05). The experimental data obtained correlate with the data concerning the partial excretion of

Table 2
Parameters of the intestinal motility (30 min) of the experimental animals with subacute liver damage against the background of loperamide-induced constipation, M±m

| Experimental group (n=8) | The total length of the small intestine, cm | The distance that the contrast mass passed, cm | The indicator of the relative distance that the contrast mass passed, % | The number of fecal boluses in the colon |
|-------------------------|-------------------------------------------|---------------------------------------------|----------------------------------------------------------------|----------------------------------------|
| Intact control          | 102.34±2.87                               | 71.41±3.48                                  | 69.70±2.66                                                      | 4.00±0.60                              |
| Control pathology No. 1 | 100.35±2.95                               | 54.66±2.41*                                 | 54.41±1.50*                                                    | 6.63±0.60*                             |
| Control pathology No. 2 | 104.25±3.82                               | 50.25±2.15*                                 | 48.33±1.57*                                                    | 5.88±0.58*                             |
| DECF, 200 mg/kg          | 100.63±2.95                               | 68.51±4.32**/#/                            | 67.91±3.33**/#/                                               | 3.63±0.50**/#/                         |
| Silybor, 25 mg/kg + Senadexin, 14 mg/kg | 103.50±2.93 | 63.65±3.27**/#/ | 61.58±2.84**/#/ | 1.88±0.40**/#/ |

Notes:
1) * – probable differences in relation to animals of the intact control group (p≤0.05);
2) ** – probable differences in relation to animals of the control pathology group No. 1 (p≤0.05);
3) # – probable differences in relation to animals of the control pathology group No. 2 (p≤0.05).

The content of markers of the general functional state of the liver in the serum of rats with subacute liver damage against the background of loperamide-induced constipation, M±m

| Experimental group (n=8) | Total protein, g/L | Urea, mmol/L | ALT, mkkat/L |
|-------------------------|--------------------|--------------|-------------|
| Intact control          | 73.14±4.10         | 6.78±0.32    | 0.61±0.03   |
| Control pathology No. 1 | 65.10±2.44         | 8.35±0.21*   | 0.63±0.06   |
| Control pathology No. 2 | 57.28±1.68**/##    | 13.92±0.53*  | 1.55±0.10**/## |
| DECF, 200 mg/kg          | 68.65±2.41*        | 9.18±0.69*   | 0.93±0.08**/## |
| Silybor, 25 mg/kg + Senadexin, 14 mg/kg | 66.08±2.23* | 10.93±0.60**/## | 1.10±0.09**/## |

Notes:
1) * – probable differences in relation to animals of the intact control group (p≤0.05);
2) ** – probable differences in relation to animals of the control pathology group No. 1 (p≤0.05);
3) # – probable differences in relation to animals of the control pathology group No. 2 (p≤0.05).
urea with feces and sweat (in addition to the main pathway – with urine) [24]; thus, the total rate of its excretion can decrease due to constipation (Tab. 3). The combined therapy and monotherapy with DECF probably contributed to the improvement of the functional state of the liver, which was mediated through positive changes in the content of markers of liver damage in the serum (ps0.05 versus the control pathology No. 2). However, it should be noted that the hepatoprotective effect of DECF in the dose of 200 mg/kg was more pronounced than in the reference group, and it was already shown in the previous experimental studies of hepatoprotective properties of different doses of the extract compared to Silybor in the dose of 25 mg/kg.

CONCLUSIONS
Thus, the experimental data obtained indicate the presence of a soft laxative effect that occurs by intensifying the intestinal motility, as well as the pronounced hepatoprotective action in the dry extract from Prunus Domestica fruits containing fibers in the dose of 200 mg/kg. These effects of DECF were revealed when using it in the treatment of constipation against the background of the experimental subacute liver damage, and were not inferior, and in some cases even surpassed, the effects studied in the complex application of the hepatoprotective and laxative reference drugs. It is important that DECF in its laxative effect, in contrast to Senodexin, did not cause signs of diarrhea in animals, which could be a beneficial feature of this drug for further clinical application. This herbal extract has shown that it can become a promising alternative to the single-stage complex therapy with plant hepatoprotectors and laxatives; it will avoid polypragmasy in the treatment of comorbid conditions in gastroenterology associated with diseases of the liver and congestive intestinal phenomena.

Conflict of interests: authors have no conflict of interests to declare.

References
1. Гарник, Т. П. Сучасні технології виробництва фітозасобів та перспективи фітотерапії / Т. П. Гарник // Фітотерапія. Часопис. – 2008. – № 1. – С. 59–63.
2. Dietary Natural Products for Prevention and Treatment of Liver Cancer / Y. Zhou, Y. Li, T. Zhou et al. // Nutrients. – 2016. – № 3. – P. 156. https://doi.org/10.3390/nu8030156
3. Identification of a flavonoid isolated from plum (Prunus domestica) as a potent inhibitor of hepatitis C virus entry / M. Bose, M. Kamra, R. Mullick et al. // Sci Rep. – 2017. – Vol. 1. – P. 3965. https://doi.org/10.1038/s41598-017-04358-5
4. Упур, Л. В. Слива в кишечной фармакологической аптеке / Л. В. Упур // Лекарственное дело. – 2016. – № 4. – С. 42–48.
5. Mohammed Shahn Basim. (Prunus Domestica fruits – perspective row for drugs development / Mohammed Shahn Basim, L. V. Lenchyk, N. B. Caidov // Nauka i innovaciya. Sertiya estestvennyh nauk. – 2017. – № 4. – P. 42–48.
6. Upyr, T. Phytochemical and pharmaceutical study of polysaccharide complexes of Prunus Domesticu fruit / T. Upyr, Shahn Basim Mohammed, Bashar Al-Jabbar Ali Sahlanli, L. Lenchyk, I. Senyk // Sci. J. «ScienceRise: Pharmaceutical Science». – 2018. – № 3 (13). – P. 32–37. https://doi.org/10.15587/2519-4852.2018.135025
7. Celik, F. Determination of phenolic compounds, antioxidant capacity and organic acids contents of Prunus domestica L. table plum fruits by HPLC / F. Celik // Acta Chromatographica. – 2017. – № 29 (4). – P. 507–510. http://dx.doi.org/10.1556/1326.2017.00327.
8. Сенюк, І. В. Вивчення послаблюючої активності різних субстанцій, одержаних з плодів сливи домашньої Prunus domestica / I. V. Senyuk, Bashar Aljabbar Ali Sahlanli, L. V. Lenchyk // Українська біофармація. – 2017. – № 5 (52). – С. 21–25. https://doi.org/10.24959/ubphj.17.134
9. Сенюк, І. В. Вивчення гепатопротекторної дії екстрактів із плодів сливи домашньої Prunus domestica / I. V. Senyuk, Bashar Aljabbar Ali Sahlanli, Mykhaylo Shchepa Basim // Фармацевтичний часопис. – 2018. – № 4. – С. 57–61. https://doi.org/10.11603/2312-0967.2018.4.9692
10. Kalaitzakis, E. Gastrointestinal dysfunction in liver cirrhosis / E. Kalaitzakis // World J. of Gastroenterol. – 2014. – Vol. 20 (40). – P. 14686–14695. https://doi.org/10.3748/wjg.v20.i40.14686
11. Scaleria, A. What does irritable bowel syndrome have in common with non-alcoholic fatty liver disease? / A. Scaleria, M. N. Di Minno, G. Tarantino // World J. of Gastroenterol. – 2013. – Vol. 19, Issue 33. – P. 5402–5420. https://doi.org/10.3748/wjg.v19.i33.5402
12. Yoon, E. Acetaminophen-Induced Hepatotoxicity : a Comprehensive Update / E. Yoon, A. Babar, M. Choudary, M. Kutner, N. Pyrsopoulos // J. of Clinical and Translational Hepatol. – 2016. – Vol. 4 (2). – P. 131–142. https://doi.org/10.14218/jch.2015.00052
13. Relationship between patterns of alcohol consumption and gastrointestinal symptoms among patients with irritable bowel syndrome / K. W. Reding, K. C. Cain, M. E. Jarrett, M. D. Eugenio, M. M. Heitkemper // The American J. of Gastroenterol. – 2013. – Vol. 108, Issue 2. – P. 270–276. https://doi.org/10.1038/ajg.2012.414
14. Доклінічні дослідження лекарственних засобів : метод. рек. / под ред. А. В. Стефанова. – К. : Авиценна, 2002. – 528 с.
15. Біохімічна оцінка результатів експериментальної дії антигепатиту фітозасобів на основі екстрактів аріштову та порошку часнику / А. Таттіс, І. А. Зупанець, С. К. Шепеко та ін. // Ліки України Плюс. – 2016. – № 3 (28). – С. 63–67.
16. Pattern of alcohol consumption and its effect on gastrointestinal symptoms in inflammatory bowel disease / G. R. Swanson, S. Sedghi, A. Farhadi, A. Keshavarzian // Alcohol (Fayetteville, N.Y.). – 2010. – Vol. 44, Issue 3. – P. 223–228. https://doi.org/10.1016/j.alcohol.2009.10.019
References

1. Gar'nyk, T. P. (2008). "Fitooterapiya. Chasopy's 1, 59–63.

2. Zhou, Y., Li, Y., Zhou, T., Zheng, J., Li, S., & Li, H.-B. (2016). Dietary Natural Products for Prevention and Treatment of Liver Cancer: Nutrients, 8(3), 156. https://doi.org/10.3390/nu8030156

3. Bose, M., Kamra, M., Mullick, R., Bhattacharya, S., Das, S., & Karande, A. A. (2017). Identification of a flavonoid isolated from plum (Prunus domestica) as a potent inhibitor of Hepatitis C virus entry. Scientific Reports, 7(1). https://doi.org/10.1038/s41598-017-04358-5

4. Upyr, L. V. (2010). "Farmaev'ty'chna ency'klopediya. Kyiv: Morion, 1290.

5. Mohammed Shahn Basim, Lenchik, L. V., Caidov, N. B. (2017). "Prunus Domestica fruits - perspective row for drugs development. Nauka i innovaciya. Seriya estestvennyh nauk, 4, 42–48.

6. Upyr, T., Basim Mohammed, S., Bashar, A.-J. A., Lenchik, L., Senyuk, I., & Kyslychenko, V. (2018). "Phytochemical and pharmaceutical study of polysaccharide complexes of prunus domestica fruit. ScienceRise: Pharmaceutical Science, 3 (13), 32–37. https://doi.org/10.15587/2519-4852.2018.135825

7. Cellik, F. (2017). "Determination of phenolic compounds, antioxidant capacity and organic acids contents of Prunus domestica L., Prunus cerasifera Ehrh. and Prunus spinosa L. fruits by HPLC. Acta Chromatographica, 29(4), 507–510. https://doi.org/10.1556/1326.2016.00327

8. Senyuk, I., Bashar, A., & Lenchik, L. (2017). "Investigation of different substances cathartic properties made from Prunus domestica. Ukrains'kij Biofarmacevtichnij Zurnal, 5(52), 21–25. https://doi.org/10.24959/ubphj.17.134

9. Senjuk, I. V., Al Sahlaní, B. J., & Basim, M. S. (2018). "Farmacev'ty'chny'chasopy's, 4, 57–61. https://doi.org/10.11603/2312-0967.2018.4.9692

10. Kalaitzakis, E. (2014). "Gastrointestinal dysfunction in liver cirrhosis. World Journal of Gastroenterology, 20(40), 14686. https://doi.org/10.3748/wjg.v20.i40.14686

11. Scaler, A., Di Minno, M. N., Tarantino, G. (2013). "What does irritable bowel syndrome share with non-alcoholic fatty liver disease? World Journal of Gastroenterology, 19(33), 5402–5420. https://doi.org/10.3748/wjg.v19.i33.5402

12. Yoon, E., Babar, A., Choudhary, M., Kuy, M., Pyrsopoulos, N. (2016). "Acetaminophen-Induced Hepatotoxicity: a Comprehensive Update. Journal of clinical and translational hepatology, 4(2), 131–142. https://doi.org/10.14218/jch.2015.00052

13. Reding, K. W., Cain, K. C., Jarrett, M. E., Eugenio, M. D., & Heitkemper, M. M. (2013). "Relationship Between Patterns of Alcohol Consumption and Gastrointestinal Symptoms Among Patients With Irritable Bowel Syndrome. The American Journal of Gastroenterology, 108(2), 270–276. https://doi.org/10.1038/ajg.2012.414

14. Stefanov, O. V. (2001). "Doklînichni doslidzhennia likar'skikh zasobiv. Kyiv: Avitsena, 528.

15. Tattis, A., Zupanecz’, I. A., Shebeko, S. K., Otrishko, I. A., Grinczov, Ye. F. (2016). "Lîby' Ukraîny' Plyus, 3 (28), 63–67.

16. Swanson, G. R., Sedghi, S., Farhadi, A., & Keshavarzian, A. (2010). "Pattern of alcohol consumption and its effect on gastrointestinal symptoms in inflammatory bowel disease. Alcohol, 44(3), 223–228. https://doi.org/10.1016/j.jalcohol.2009.10.019

17. Choi, J.-S., Kim, J. W., Cho, H.-R., Kim, K.-Y., Lee, J.-K., Sohn, J. H., & Ku, S.-K. (2014). "Laxative effects of fermented rice extract in rats with loperamide-induced constipation. Experimental and Therapeutic Medicine, 8(6), 1847–1854. https://doi.org/10.3892/etm.2014.2430

18. Wintola, O. A., Sumonou, T. O., Afolayan, A. J. (2010). "The effect of Aloe ferox Mill. in the treatment of loperamide-induced constipation in Wistar rats. BMC gastroenterology, 10, 95. https://doi.org/10.1186/1471-230x-10-95

19. Drogovoz, S. M., Borody'nà, T. V., Dery' medvid', L. V. (1998). "Lîby', 5, 32–35.

20. Nair, A. B., Jacob, S. (2016). "A simple practice guide for dose conversion between animals and human. Journal of basic and clinical pharmacy, 7(2), 27–31. https://doi.org/10.4103/0976-0105.177703

21. Gnjidic, D. (2017). "Determining medication burden and polypharmacy: finding the perfect measure / D. Gnjidic, M. Tinetti, H. G. Allore // Expert Rev. of Clinical Pharmacol. – 2017. – Vol. 10, Issue 4. – P. 345–347. https://doi.org/10.1080/17512433.2017.1301206

22. Sagar, L. (2018). "Laxative effects of fermented rice extract in rats with loperamide-induced constipation / L. Sagar, R. Sehgal, S. Ojha // BMC Complement Altern Med. – 2005. – Vol. 5 (1). – P. 18. https://doi.org/10.1186/1472-6862-5-18

23. Hal'fan, A. A. "STATYSTYCA 6. STATYSTYCA ANALIZY DANYH: URBANIZATYOR. – 3-e izd. / A. A. Hal'fan – M.: OOO "Bînox-Prress", 2007. – 512 s.

24. Levitt, D. G. (2006). "A model of blood-ammonia homeostasis based on nitrogen metabolism in the multiple organs involved in the production, catabolism, and excretion of ammonia in humans / D. G. Levitt, M. D. Levitt // Clinical and Experimental Gastroenterology. – 2018. – Vol. 11. – P. 193–215. https://doi.org/10.2147/ceg.s160921

25. Deacon, R. M. Housing, husbandry and handling of rodents for behavioral experiments / R. M. Deacon // Nature Protocols. – 2006. – Vol. 1 (2). – P. 936–946. https://doi.org/10.1038/nprot.2006.120
22. Sagar, L., Sehgal, R., & Ojha, S. (2005). Evaluation of antimotility effect of Lantana camara L. var. acuelata constituents on neostigmine induced gastrointestinal transit in mice. *BMC Complementary and Alternative Medicine, 5*(1), 18. https://doi.org/10.1186/1472-6882-5-18

23. Khalafian, A. A. (2007). *STATISTICA 6. Statisticheskii analiz dannykh. Uchebnik.* Moscow: ООО «Binom-Press», 512.

24. Levitt, D., & Levitt, M. (2018). A model of blood-ammonia homeostasis based on a quantitative analysis of nitrogen metabolism in the multiple organs involved in the production, catabolism, and excretion of ammonia in humans. *Clinical and Experimental Gastroenterology, 11*, 193–215. https://doi.org/10.2147/ceg.s160921

25. Deacon, R. M. (2006). Housing, husbandry and handling of rodents for behavioral experiments. *Nature Protocols, 1* (2), P. 936–946. https://doi.org/10.1038/nprot.2006.6.120.

---

Information about authors / Відомості про авторів / Сведения об авторах

**Bashar Jabbar Ali Al-Sahlanee**, postgraduate student of the Department of Biological Chemistry, National University of Pharmacy. E-mail: al.sahlny82@yahoo.com

**Lytkin D. V.**, head of the Central Research Laboratory, National University of Pharmacy. E-mail: d.v.lytkin@gmail.com

**Senyuk I. V.**, Candidate of Pharmacy (Ph.D.), associate professor of the Department of Biological Chemistry, National University of Pharmacy. E-mail: citochrom@gmail.com

**Mailing address:** 12, Kulikovska str., Kharkiv, 61002, Ukraine, Department of Biological Chemistry, National University of Pharmacy. Tel.: +380631313100

---

Надійшла до редакції 25.04.2019 р.