miR-21 inhibition of LATS1 promotes proliferation and metastasis of renal cancer cells and tumor stem cell phenotype

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Abstract. MicroRNA (miR)-21 has many regulatory functions in the cell, including activities in cancer cells and cancer stem cells. Large tumor suppressor gene 1 (LATS1) is a target of miR-21 that could mediate several of these phenotypes. This study explored the effect of miR-21 silencing in renal cancer cell function and LATS1 expression. Silencing of miR-21 in Caki-2 cells reached an efficiency of 55‑60%. This was sufficient to detect decrease in Caki-2 cell proliferation and migration invasion capacity. miR-21 silencing increased LATS1 expression at both mRNA and protein levels. The number of tumor spheres formed by cells expressing si-miR-21 was significantly reduced and the expression of tumor stem cell markers Nanog and CT3/4 were significantly downregulated. miR-21 seems to regulate LATS1 expression in renal cancer Caki-2 cells, resulting in reduced proliferation, invasion, and cancer stem cell phenotype. miR-21 may promote malignant phenotype of tumor cells through LATS1 silencing, which can be regarded as a new target candidate gene for renal cancer treatment.

Introduction

Renal cell carcinoma (RCC) originates from the malignant tumor of the epithelium system of urinary tubule of renal parenchyma. RCC has become the malignant tumor with the highest mortality in the urinary system, accounting for 2-3% of adult malignant tumors (1,2). The incidence of RCC shows an upward trend each year, ranking 10 in the incidence rate of male malignant tumors in China in 2008. The causes and pathogenesis of RCC are still unclear.
Materials and methods

Cell line and main reagents. Human renal carcinoma Caki-2 cells were from American Type Culture Collection (ATCC; Manassas, VA, USA), Dulbecco's modified Eagle's medium (DMEM) culture and fetal bovine serum from Gibco Life Technologies (Carlsbad, CA, USA), methyl thiazolyl tetrazolium (MTT) kit from Sigma (St. Louis, MO, USA), miR isolation extraction and separation kit, RPMI-1640, Lipofectamine® 2000 and TRIZol from Invitrogen Life Technologies (Carlsbad, CA, USA). The quantitative (qPCR) (qPCR) Mix kits were from Takara Bio, Inc. (Otsu, Japan) and mouse monoclonal LATS1 antibody (dilution, 1:500; cat. no. sc-398560), mouse monoclonal Nanog antibody (dilution, 1:500; cat. no. sc-293121), mouse monoclonal OCT3/4 (dilution, 1:500; cat. no. sc-5279), and mouse monoclonal GAPDH antibody (dilution, 1:500; cat. no. sc-293335) were from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA). Primers were designed and synthesized by Shanghai Jima Pharmaceutical Technology Co., Ltd. (Shanghai, China). si-miR-21 and si-negative control were synthesized by Shanghai Jima Pharmaceutical Technology. si-miR-21 forward, 5'-GATCCAGUGAAGUGACTT-3'; reverse, 5'-UGCUCUTUGACGUAUGGATT-3'; si-negative control forward, 5'-UUCACCGUACUCUCAGUGT-3'; reverse, 5'-ACUGGAACCUCUCGCGGAAT-3'; LATS1 forward, 5'-AAATGCCCACATCCGGGAA-3' and reverse, 5'-ACCT GGCTCTCCCCCTAAACA-3'.

si-miR-21 and si-negative control transfection of Caki-2 cells. The cells in the logarithmic growth phase were divided into three groups based on treatment: miR-21, si-miR-21, and si-negative control. When the cell fusion reached 80-90%, transfection was conducted with Lipofectamine® 2000 according to instructions. At 48 h after transfection, miR was extracted following the manufacturer's instructions. We used light photometer to detect RNA absorbance and calculate RNA concentration and purity. The samples with a rate from 1.8 to 2.1 were selected for further experiments. qPCR was used to conduct comparative analysis of miR-21 mRNA in Caki-2 in the three groups after transfection.

qPCR. Total RNA was extracted from the three groups and purified according to the TRIZol kit instructions; 1 µg RNA template was converted into cDNA by reverse transcription according to reverse transcription kit. The reaction system was 10 µl, at 37°C for 15 min and heated at 85°C for 30 sec. The amplification condition was: 95°C for 5 min, then 95°C for 30 sec, 58°C for 30 sec, 72°C for 30 sec, a total of 30 cycles, and finally 72°C for 5 min. Analysis of dissolution curve was conducted after the reaction.

Western blotting. Total protein from cells of the three groups was extracted by RIPA lysis solution plus 1% PMSF, and the protein concentration was detected by BCA (all from Biosharp, Hefei, China). A 1/4 volume of the protein sample buffer was added, denatured at 100°C for 10 min. SDS-polyacrylamide separating gel (Keygen, Nanjing, China) and stacking gel were prepared, Tris glycine electrophoretic buffer solution and equal amount of denatured protein of ~50 µg per well were added, proteins were transferred to PVDF membrane after electrophoresis and blocked in 3% BSA for 2 h. Primary antibodies (anti-LATS1 1:1,000; anti-GAPDH 1:1,000) were added at 4°C overnight, TBST membrane was washed 3 times/10 min, then horseradish peroxidase-labeled secondary antibodies (1:5,000) were added at 37°C for 2 h, TBST membrane was washed 3 times/10 min, ECL development, and development in vilber lourmat in the dark.

Proliferation assay by MTT. Cells from each group were inoculated in 96-well culture plate at a density of 2×10^3 cells/well. Three multiple pores were set and 2 ml of 5 mg/ml MTT was added to each pore after 24, 48, 72, 96 and 120 h. They were incubated for 4 h, then the supernatant was discarded, 150 µl of DMSO was added, mixed, dissolved and crystallized. The light density value [D (490)] in each pore was determined at 490 nm wavelength, and cell proliferation inhibitory rate (IR): was calculated IR = [1-experimental group D (490)/the control group D (490)] × 100.

Invasion ability of Caki-2 cells by Transwell. Experiments were conducted according to the BioCoat Matrigel invasion chamber kit instructions. Eight different visual fields (magnification, ×100) were counted under the microscope, repeating 3 times, and the percentage of the invasion cells was calculated.

Tumor sphere formation. Serum-free medium (SFM) was prepared with Ham's DMEM-F12 (1:1), B27 (1:50), EGF (20 mg/ml), and bFGF (20 ng/ml). The cells of the three groups were inoculated in DMEM with 10% fetal bovine serum until the cells were in the logarithmic growth phase, and they were washed with PBS, digested with 0.25% trypsin, and washed in PBS twice after digestion. Then the cells were placed in the low adhesion culture in suspension for 48 h and changes in cell morphology were observed under an inverted microscope. The formation of tumor spheres was observed and the numbers of tumor spheres were counted after 14 days.

Statistical analysis. SPSS 19.0 software (SPSS, Inc., Chicago, IL, USA) was used for plotting and statistical analysis, measurement data are expressed by mean ± standard deviation, comparison of rate was tested by χ², GraphPad Prism 5.0
(GraphPad Software, Inc., La Jolla, CA, USA) was used to carry out one-way ANOVA analysis and plotting for qPCR results. *P<0.05 was considered to indicate a statistically significant difference.

Results

Efficiency of siRNA transfection by qPCR. According to qPCR, transfection of Caki-2 cells with si-miR-21 resulted in significant reduction of miR-21 compared to transfection with si-negative control (F=71.90, P<0.05). The silencing efficiency was 55‑60% (Fig. 1).

LATS1 expression after transfection of miR-21 knockdown. Western blot analysis showed that the expression level of LATS1 protein was significantly higher in cells transfected with si-miR-21 compared to control cells and si-negative cells (Fig. 2A). qPCR showed that LATS1 mRNA expression was significantly higher in cells transfected with si-miR-21 compared to control and si-negative cells (F=108.5, P<0.05; Fig. 2B).

Proliferation of Caki-2 cells after miR-21 silencing. Following cell transfection, we collecting cell number data at 1, 24, 48, 72, 96 and 120 h. We found no significant differences at 24 and 48 h, but si-miR-21 showed lower proliferation after 72 h (Fig. 3). The proliferation rate at 72 h was: 40.5±11.6% for si-miR-21, 57.4±5.9% for si-negative control, and 58.3±4.3% for non-transfected control (t=2.375, P<0.05). Cell proliferation at 96 h was: 43.7±12.5% for si-miR-21, 75.6±7.5% for si-negative control, and 78.3±6.9% for non-transfected control (t=4.587, P<0.01). Cell proliferation at 120 h was: 48.9±12.3% for si-miR-21, 81.9±4.6% for si-negative control, and 85.2±3.8% for non-transfected control (t=5.698, P<0.01). Thus, si-miR-21 intervention inhibited the proliferation capacity of Caki-2 cells.

Caki-2 cell invasion after si-miR-21 transfection. The invasion ability of Caki-2 cells transfected with si-miR-21 significantly decreased compared to the si-negative control cells and the non-transfected cells (F=135.1, P<0.01; Fig. 4). Thus, miR-21 knockdown reduces the invasivity of Caki-2 cells.

Discussion

miRs are endogenous regulatory molecules that play roles as oncogenes or tumor suppressor genes in the development of tumors. As one of the most studied miRs in recent years, miR-21 is highly expressed in gastric cancer, breast cancer, colon cancer, and other tumors, and can be used as a marker to determine the prognosis of various tumors. Some studies suggest that miR-21 overexpression downregulate LATS1...
expression. A large number of studies have found that as one of the key factors in the hippo-Yap signaling pathway, LATS1 expression is reduced in a variety of tumors, which may be related to the high expression of miR-21 (27-29).

Here we showed that silencing miR-21 in Caki-2 cells decreased the proliferation ability after 72 h and continued to decrease until 120 h. Downregulation of miR-21 also resulted in elevated LATS1 expression, indicating that miR-21 has a regulatory effect on the expression of LATS1. miR-21 is known to regulate cell differentiation, proliferation, development, apoptosis, metabolism and cancerous transformation (30).

Tumor stem cells have self-renewal capacity and can produce heterogeneous tumor cells. They can sustain the vitality of the tumor cell population through self-renewal, unlimited proliferation, movement, and migration ability to promote the transfer of tumor cells. Cancer stem cells can be in a dormant state for a long time and are insensitive to a variety of chemotherapeutic drugs and external physicochemical factors killing tumors, resulting in antitumor drug resistance and reducing the treatment effect. To clarify the mechanism regulating tumor stem cell state is very important for the treatment of tumors. This study found that downregulation of miR-21 inhibited the phenotype of tumor stem cells.

Our study results show that silencing miR-21 expression inhibited the malignant activities of renal carcinoma cells. This inhibition may be induced by downregulation of LATS1 expression and miR-21 may be seen as a marker for diagnosis and prognostic analysis of RCC and LATS1 may become a new target for treatment.

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