A new endoscopic closure method for gastric mucosal defects: feasibility of endoscopic hand suturing in an ex vivo porcine model (with video)

Background and study aims: More secure endoscopic closure techniques for iatrogenic gastric defects are required for safe endoscopic surgery. We developed a novel endoscopic suturing method, endoscopic hand suturing (EHS), of gastric mucosal defects and determined its feasibility and efficacy ex vivo.

Materials and methods: We created 24 mucosal defects (each 2 cm in diameter) by endoscopic submucosal dissection. The following three techniques were tested: EHS with a 3–0 barbed suture that was grasped with biopsy forceps (n=6) or a prototype through-the-scope needle holder (n=6) by endoscopy, looping with endoloops (n=6) by endoscopy, and clipping with hemoclips (n=6) by hand. The mucosal edges were attached to each other at three points. The closure strength was compared among the three groups, and the procedural duration was compared between the EHS and looping groups.

Results: All 12 lesions were completely closed by EHS. The median strength of the closure, measured with a spring scale, was significantly greater in the EHS group (0.74 kg) than in the looping group (0.33 kg, P=0.0012) or clipping group (0.07 kg, P=0.0009). The median procedural duration did not significantly differ between the EHS and looping groups (19.7 vs. 19.8 minutes, P=1.0000). The use of the needle holder significantly reduced the procedural duration compared with the biopsy forceps.

Conclusion: Mucosal defects can be firmly closed with EHS, which may be helpful for establishing a safer and more secure endoscopic surgery.

Introduction

Endoscopic submucosal dissection (ESD) is accepted, particularly in Asian countries, as a minimally invasive endoscopic approach to curing early stage cancer without organ resection [1–5]. Delayed bleeding is a major complication after gastric ESD that occurs in approximately 5% of patients [6–8]. Delayed perforation can also develop after successful resection [9]. Although the definitive causes of these complications are unknown, the exposure of an iatrogenic mucosal defect to gastric acid may be an important factor. When such defects are created in patients at risk for complications, closure of the defect with hemoclips or a detachable snare is sometimes attempted [10–12]. Suturing with a needle and suture thread can yield a secure closure when the alimentary tract is surgically anastomosed. If it is possible to firmly close an iatrogenic defect by suturing the mucosa endoscopically, delayed bleeding after ESD or anastomotic leakage after endoscopic transluminal surgery can be more effectively prevented. Therefore, we designed an endoscopic continuous suturing method, endoscopic hand suturing (EHS), in which a needle and suture are used to optimally close iatrogenic mucosal defects. Here, we investigated the feasibility of this method and compared its performance with that of conventional looping and clipping in a porcine model ex vivo.

Materials and methods

Preparation

We obtained 10 isolated stomachs from pigs that had been previously slaughtered for food. A single endoscopist created 24 mucosal defects (each 2 cm in diameter) by ESD in these stomachs, which were then placed on an ESD training unit after the insides had been rinsed several times with tap water. Several dots were circumferentially marked by placing 2-cm plastic disks on the mucosal surface. Normal saline with a small
A small amount of indigo carmine was injected to create a submucosal cushion. A GIF-2TQ260M multibending scope with two working channels (Olympus Medical Systems Corp., Tokyo, Japan) was used for ESD and endoscopic closure. The closure methods described below were then applied by one endoscopist and one assistant (Fig. 1).

### Endoscopic hand suturing

Defects were closed with 15-cm, 3–0 absorbable barbed V-Loc 180 (VLOCL0604) sutures (Covidien, Mansfield, MA, USA) (Fig. 2a). This type of unidirectional, knotless suture was originally developed to facilitate skin suturing, and it has subsequently been applied to gastrointestinal surgeries [13–15]. The short barbs are oriented along the suture to face away from the needle, preventing the suture from sliding backward after the tissues have been tightened. A loop on the tail and the continuous small part of the suture without barbs was cut and anchored with an EZ clip (HX-610–90; Olympus). In closing six of the lesions, the needle was grasped with biopsy forceps (FB-220U; Olympus). A 10-mm SD-210L-10 electrocautery snare (Olympus) was inserted into a separate working channel to tightly grasp the neck of the biopsy forceps for stabilization (Fig. 2b). For the next six lesions, a prototype through-the-scope needle holder (Olympus) was used (Fig. 2c–e) [16,17]. This device has one swinging lateral jaw and one fixed straight jaw, both of which firmly close when the handle slider is grasped. A button on the handle unlocks and opens the jaws. Irrespective of the shape of the endoscope, the tip of the device can simultaneously rotate as the handle rotates, which accelerates suturing, even inside the stomach. To insert the needle and string gently through the overtube, we grasped the tail of the needle and the suture near the anchor clip with biopsy forceps. We grasped the tail of the needle with the needle holder and placed the anchor clip into another working channel in a retrograde manner. The tip of the needle was or-

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**Fig. 1** Comparison of three mucosal lesion closure methods. All mucosal defects were closed at three points. **a** Endoscopic hand suturing. The defect was continuously sutured endoscopically by using a barbed suture with hemoclips at the end. **b** Looping. Endoloops were attached to mucosal edges with hemoclips and closed endoscopically. **c** Clipping. Mucosal edges were affixed to each other with hemoclips by hand.

**Fig. 2** Tools used for endoscopic hand suturing. **a** A barbed suture (V-Loc 180) with a hemoclip anchor at its end. **b** Biopsy forceps fixed with a 10-mm electrocautery snare. **c, d** The tip of the prototype through-the-scope needle holder. **e** The grip of the prototype needle holder. The jaws were tightly closed by gripping the slider, and the button unlocked and opened the jaws.
intended toward the endoscope to avoid potential injury to the mucosa. The needle was released in the stomach and grasped again, and then the mucosal edges and part of the submucosa were continuously sutured to each other longitudinally, from the distal to the proximal sides of the endoscope. After some amount of suturing, the slack part of the suture was tightened by directly grabbing and pulling it with biopsy forceps or the needle holder. After three sutures had been made, one mucosal edge slightly distal to the third suturing point was sutured as a lock. The excess suture and the needle were cut away with a loop cutter (FS-5L-1; Olympus), the end of the suture was clipped to the mucosa, and the needle was removed through the overtube by grasping the tip of the needle with biopsy forceps or the needle holder (Fig. 1a). Video 1 shows EHS with the needle holder.

**Looping**

An MAJ-340 endoloop, 20 mm in diameter (Olympus), was opened above a mucosal defect and placed at each mucosal edge with an EZ clip (HX-610–90 or HX-610–90L; Olympus) [11,12]. Thereafter, the loop was slowly closed by heading a locking unit. The loop was firmly closed and detached. The excess loop was cut and removed with a loop cutter. We spaced three loops along the line to divide the closure seam into four equidistant segments (Fig. 1b).

**Clipping**

We applied three HX-610–90 or HX-610–90L EZ clips per lesion. To stitch the mucosal edges to each other without slippage, the procedure was performed by hand after the stomach had been opened as described below. Clips were spaced along the line to divide the closure seam into four equidistant segments (Fig. 1c). We closed six experimentally created mucosal defects under each condition (EHS with biopsy forceps, EHS with the needle holder, looping, and clipping) (Fig. 3, Fig. 4a–c). We created two artificial lesions in the gastric corpus of nine stomachs for both the EHS and looping groups. For a forward approach, one lesion was located at the posterior wall, and for a retroflex approach, one lesion was positioned at the lesser curvature. An additional six lesions in total were created for the clipping group by using the remainder of the two stomachs and one additional whole stomach. Endoscopic closure of the lesion on the lesser curvature was initially achieved by a retroflex approach and repeated again on the posterior wall to avoid potential mechanical injury to the sutured lesion on the posterior wall while the endoscope was handled in a retroflex approach. After closure by endoscopy, the stomach was removed from the unit, cut along the greater curvature, and opened with the mucosal surface facing upward to assess the strength of closure. The clipping method was performed on two of the opened stomachs. We also directly cut one additional stomach and opened it for the clipping method after the creation of four mucosal defects.

**Measuring outcomes and statistical parameters**

The primary outcome measured in the three groups was the strength of closure, which was assessed with a digital spring scale (Fig. 4d). The scale was attached to an 18-gauge needle that was sewn parallel to the suture line on one side of the mucosa and then pulled to the opposite side of the suture line while the other side of the mucosa was held by hand (Fig. 4e). The weight (in kilograms) indicated on the monitor when the closure collapsed was taken as a surrogate marker of closure strength. The secondary outcome measured was the procedural duration, which we defined as the time elapsed from the first insertion of the needle to the placement of a clip at the end of the suture in the EHS group, and from the first clipping step to place the first loop at the mucosal edge to cutting the excess of the third loop in the looping group. Differences in closure strength and procedural duration between the grasping devices used (biopsy forceps or needle holder) and the approach used (forward or retroflex) were also analyzed in the EHS group. Strength and procedural duration were analyzed with the Mann-Whitney U test. Statistical significance was set at a P value of <0.05. All data were statistically analyzed with JMP version 9.0 software (SAS Institute, Cary, NC, USA).

**Results**

All 24 procedures were completed, including EHS procedures with biopsy forceps (6 of 6, 100%) or the needle holder (6 of 6, 100%). We measured the median strength in the three groups (Fig. 5a) and found that closures were significantly tighter in...
the EHS group (0.74 kg [range 0.49 – 1.16 kg]) than in the looping group (0.33 kg [range 0.21 – 0.54 kg], \( P = 0.0012 \)) or clipping group (0.07 kg [range 0.02 – 0.31 kg], \( P = 0.0009 \)). Moreover, the strength was significantly weaker in the clipping group than in the looping group (\( P = 0.0200 \)).

The median procedural duration did not significantly differ between the EHS and looping groups (19.7 minutes [range 12.5 – 27.1 minutes] vs. 19.8 minutes [range 15.5 – 37.4 minutes], \( P = 1.0000 \)(\( \text{Fig. 5b} \)). The median procedural duration for stitching in the EHS group (excluding time required for locking, cutting, and clipping the end of the suture) was 13.6 minutes (range 7.9 – 20.2 minutes), indicating a median time of 4.5 minutes (range 2.6 – 6.7 minutes) per stitch.

We found no significant difference in median strength between EHS with biopsy forceps or with the needle holder (0.77 kg [range 0.63 – 1.02 kg] vs. 0.74 kg [range 0.49 – 1.16 kg], \( P = 1.0000 \)), whereas the median procedural duration was significantly shorter with the needle holder than with the biopsy forceps (17.0 minutes [range 12.5 – 21.2 minutes] vs. 22.7 minutes [range 16.8 – 27.1 minute], \( P = 0.0161 \)(\( \text{Fig. 6} \)).

The median stitching durations with the biopsy forceps and the needle holder were 16.4 minutes (range 11.3 – 20.2 minutes) and 10.3 minutes (range 7.9 – 17.1 minutes), respectively. The time required per stitch tended to be shorter with the needle holder than with biopsy forceps, but the difference was not statistically significant (median 3.4 minutes [range 2.6 – 5.7 minutes] vs. 5.5 minutes [range 3.8 – 6.7 minutes], \( P = 0.0538 \)).

The median strength and median procedural duration did not significantly differ between the forward and retroflex approaches in the EHS group (0.72 kg [range 0.60 – 1.16 kg] vs. 0.79 kg [range 0.49 – 1.06 kg], \( P = 0.6884 \) and 21.6 minutes [range 12.5 – 27.0 minutes] vs. 17.5 minutes [range 12.8 – 27.1 minutes], \( P = 0.6884 \), respectively)(\( \text{Fig. 7} \)).

**Discussion**

Using an ex vivo porcine model, we found that EHS was feasible and offered firmer closure than conventional looping and clipping methods. Based on our findings, we speculate that EHS would make mucosal defect closure more secure than the other two methods.

Clipping is the most popular method in clinical practice because it is convenient and simple, particularly for patients at a high risk for delayed bleeding due to antiplatelet or antithrombotic medications [10]. However, the findings presented here imply that the mucosal edges attached by clipping are too fragile to stick together for long periods of time. Looping can be used particularly for large lesions that cannot be closed by clipping [11,12], although this method is somewhat tedious and more time-consuming.

Here, we found that looping was stronger than clipping, but EHS was more secure and did not take any longer than looping. Therefore, our data suggest that EHS is the most reliable of the three methods tested. Although in practice looping is accompanied by clipping, this combined process is still predicted to be weaker than EHS based on the data presented in this study. Because clipping is far weaker than looping, it would not significantly enhance the strength of looping.

In the looping and clipping groups, closure sites collapsed when the clips became detached as the mucosal edges parted. In contrast, closure sites dehisced when sutures in the EHS group tore.
the mucosa. This finding suggests that sites closed with EHS will remain intact unless affected by a strong force that is powerful enough to tear the mucosa, whereas mucosal sites closed by looping or clipping may easily dehisce in the presence of less powerful forces, such as food movement or peristalsis. Many reports have addressed the management of delayed bleeding, and an evidenced-based consensus is gradually forming [6–8, 18–20]; however, no definitive methodology can yet effectively prevent delayed bleeding. Furthermore, delayed perforation can occasionally occur after a successful procedure [9]. The precise cause of delayed perforation is unknown, but excessive thermal injury to the surface of a mucosal defect may be a risk factor. Exposure of a mucosal defect to gastric acid after ESD can also be a risk factor for delayed bleeding and delayed perforation because gastric acid slows the gastric ulcer healing process and stimulates nonbleeding visible vessels or nonperforated thin walls in an iatrogenic mucosal defect. EHS should prevent such complications by preventing the exposure of mucosal defects to gastric acid. Practically, antisecretory medicines, which keep the risk for delayed bleeding low, are used to accelerate the healing of mucosal defects. Therefore, EHS would be more effective in patients who have a high risk for delayed bleeding, such as those who are using antithrombotic agents, have a coagulopathy or liver cirrhosis, or are being treated for chronic renal failure by hemodialysis.

Over-the-scope clips [21] and the OverStitch Endoscopic Suturing System (Apollo Endosurgery, Austin, TX, USA) [22], which were principally developed to close transmural defects, can also effectively close mucosal defects. A previous study showed comparable strength when wall defects were closed with over-the-scope clips or surgical suturing [23]. The Eagle Claw (Olympus) [24] and Flexible Endostitch (Covidien) [25] are also promising suturing devices for wall closure, but the efficacy of mucosal closure with these devices is unknown, and a comparison with EHS is warranted. On the contrary, EHS may have potential application in the treatment of perforation or anastomosis. Although full-thickness suturing by EHS would be more difficult than mucosal suturing because of collapse of the stomach during the procedure, it might be worth trying in the future. Our study has some limitations. EHS requires a certain amount of suturing time, a skillful hand for endoscope maneuvering, and some experience with the technique. In addition, extant needles and sutures are not intended for endoscopic suturing of the mucosa. Other limitations of our study are as follows: a relatively small number of lesions closed per technique, the use of hand clipping instead of endoclipping, and the use of porcine stomachs ex vivo. The findings of this study should be considered preliminary at best. Thus, an in vivo porcine trial that included a histologic assessment of healed sutured lesions would be desirable to con-
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