Which Models Are Appropriate for Six Subtropical Forests: Species-Area and Species-Abundance Models

Shi Guang Wei¹, Lin Li¹*, Zhen Cheng Chen¹, Ju Yu Lian², Guo Jun Lin³, Zhong Liang Huang²*, Zuo Yun Yin⁴

¹Guilin University of Electronic Technology, Guilin 541004, China, ²South China Botanical Garden, the Chinese Academy of Sciences, GuangZhou, 510650, China, ³Changjiang Water Resources Protection Institute, WuHan, 430051, China, ⁴Guangdong Forest Research Institute, GuangZhou, 510520, China

Abstract

The species-area relationship is one of the most important topics in the study of species diversity, conservation biology and landscape ecology. The species-area relationship curves describe the increase of species number with increasing area, and have been modeled by various equations. In this paper, we used detailed data from six 1-ha subtropical forest communities to fit three species-area relationship models. The coefficient of determination and F ratio of ANOVA showed all the three models fitted well to the species-area relationship data in the subtropical communities, with the logarithm model performing better than the other two models. We also used the three species-abundance distributions, namely the lognormal, logcauchy and logseriesy model, to fit them to the species-abundance data of six communities. In this case, the logcauchy model had the better fit based on the coefficient of determination. Our research reveals that the rare species always exist in the six communities, corroborating the neutral theory of Hubbell. Furthermore, we explained why all species-abundance figures appeared to be left-side truncated. This was due to subtropical forests have high diversity, and their species number includes many rare species.

Introduction

The species-area relationship (SAR) is among the well known and most studied patterns in ecology [1,2,5,4,6], and it is important to both our basic understanding of biodiversity and ability to conserve biodiversity [7]. The SAR describes the increasing number of species (S) as area (A) increases; when plotted, the SAR is, in theory, a monotonically increasing curve whose slope is steep at first but gradually becomes nearly flat. In reality, SAR curves have a variety of shapes depending on different sampling strategies, biases, and community parameters. The SAR has been used to determine the minimal sampling size of a particular community [8,9], to estimate species richness [10,11] or species extinction as a result of habitat loss and fragmentation [12,13,14] and to calculate the effective size of natural reserves for long-term conservation of biodiversity [15,16].

The SAR has been explained by two ecological hypotheses: (1) the habitat heterogeneity hypothesis predicts that, as area increases, habitat heterogeneity increases, and the total number of species also increases; (2) the area per se or equilibrium hypothesis invokes colonization–extinction dynamics which postulates that, as area increases, relative rates of colonization increases while rates of extinction decreases, and therefore the total number of species increase with area [4]. However, habitat heterogeneity and area are often closely interdependent, and therefore their relative influences are difficult to disentangle [17,18]. Equal sample sizes, such as sampling equally sized areas, should reduce effects of area, which, in turn, may be correlated with habitat heterogeneity [19,10,20].

Traditionally, the functional forms of the SAR have been modeled with convex-upward functions lacking an asymptote, such as the power [21], exponential [22,23,24], logarithmic [22,23], and logistic [25,26,27] functions, as well as the random-placement model [28]. More convex and asymptotic models as well as sigmoid models have also been used [29]. Overall, it seems that the power model represents a reasonably good approximation of the SAR, and for intermediate spatial scales [21,4,30].

Conventional reasoning behind the SAR is based on another fundamental ecological pattern, the species–abundance distribution (SAD), which describes how individuals are distributed among species [31]. He and Legendre [27] pointed out that area, species-abundance (SA), spatial distribution, and species richness have been central components of community ecology, and mathematically explore the interrelationships among these components. Among these, the SA pattern is so fundamental that Sugihara referred to it as “minimal community structure” [32].

There are two main types of models that have been suggested to characterize the SA pattern in biological communities [33,34,35]. One type is the rank-abundance curve [36,37], or dominance-diversity curve [38,39], which is described mainly by the broken stick model or the random niche-boundary hypothesis of MacArthur and Wilson [13] as well as the geometric series [40]. The second type is the SA distribution, which has been commonly...
modeled by the logseries distribution [24] or the lognormal distribution [41]. Fisher et al. [24] found the logseries distribution by assuming that the parameter \( k \) of the negative binomial distribution is close to zero; it was subsequently found that a great deal of data also fitted the logseries model [42,43,3]. The lognormal distribution has been derived through multiple routes, most recently from neutral community theory assuming identical species [44,45]. The lognormal model is a more general model of SA patterns in natural communities [46] and can therefore be fitted to all data that also fit the logseries model well. Hubbell’s [45] theory suggested that Fisher’s logseries describes the SAD of a metacommunity, while Preston’s lognormal model describes the SAD of a local community. Yin et al. [35] tested two alternative models, the so-called log-cauchy and log-sech models which have simpler functional forms than the commonly used lognormal model [47]; these two alternative models fitted the observed SADs better than the lognormal model.

Tropical forests are generally considered to be ecological systems well suited for species-area and spatial-aggregation research [48,28]. Subtropical forests display a relatively high diversity of tree species, therefore providing suitable conditions for ecological analyses. Moreover, it is urgent to study their ecology, because the area of subtropical forests has recently been rapidly reduced while the survived forests have been degraded and fragmented [49]. Studying trees has the obvious advantage, because their sedentary life history makes them relatively easy to relocate, and most them can also be identified because their taxonomy is rather well-resolved.

Based on a detailed data set from six subtropical forests, in this study we intend (1) to examine the validity of the random-placement, power law and logarithm model for subtropical forests, to fit the corresponding SAR relationships and then compare their fitted results to find the appropriate model to express the SAR of subtropical forests; (2) to focus on the second type of SAR models, i.e., to fit the lognormal, logseries and log-cauchy distributions, and to find the suitable SAD models for subtropical forests; and (3) to compare species richness among different communities, and based on SAR models to explain ecological changing processes of the subtropical forests based on fitted SAR models. We also discuss the influence of different sampling sizes and sampling methods on model fitting. This study thus contributes to our understanding of SAR and SAD in subtropical forests.

**Methods**

**Ethics Statement**

No specific permits were required for the described field studies. Our study site (six 1-ha plots in the subtropical forests of southern China) is owned by the Chinese government and managed by South China Botanical Garden, Chinese Academy of Sciences. We can do our research works freely in these plots under the Regulations of the People’s Republic of China on Nature Reserves. Our field studies did not involve endangered or protected species.

**Sampling Tree Communities**

For this study, we censused trees in six 1-ha plots in the subtropical forests of southern China (Table 1). In each community, each woody stem ≥1 cm diameter at breast height (DBH) was identified to species. Community A was sampled within the Maoershansh National Nature Reserve (110°30′E, 25°56′N), which belongs to the monsoon evergreen broad-leaved forest. Community B was sampled within Nanling Nature Reserve (112°56′–113°4′E, 24°30′–24°48′N), which belongs to broad-leaved forest. Communities C and D were sampled within the Dinghushan Biosphere Reserve (112°30′–112°33′41″E, 23°09′21″–23°11′30″ N), which contains monsoon evergreen broad-leaved, needle and broad-leaved mixed forests. Communities E and F were sampled within the Jinggangshan Nature Reserve (114°05′–114°25′E,26°22′–26°48′ N), which contains broad-leaved, needle and broad-leaved mixed forests.

**SAR Models**

The empirical study of SAR dates back to H. C. Watson, who presented the first known species-area curve for Great Britain’s vascular plants in 1859 [50]. Watson found a linear relationship between the logarithm of the number of species present and the logarithm of the area sampled, over areas ranging from a square mile to all of Great Britain. This is the most common pattern found on regional scales within relative homogeneous landscapes [22,4]. The relationship is given by the so-called power function:

\[
S(A) = c A^z
\]

where \( S(A) \) is the total number of species encountered in the geographical area \( A \), \( c \) is the parameter characterizing the specific biogeographical region and \( z \) is the parameter which is specific to the sampled community.

The second SAR model is the logarithm model, which is transformed from the model mentioned by Buys et al. [51]. Liu et al. [52] used the data of five plots located in the temperate zone to fit ten species-area models and showed that the following logarithm model provided excellent fits:

\[
S(A) = (a + b \cdot \ln A)^c
\]

where \( S(A) \) is the total number of species encountered in the geographical area \( A \), \( a \) is the species number in the area unit, \( b \) is a parameter independent from area, which is considered a measure of spatial heterogeneity and \( c \) is a fitted constant.

The third model is Coleman’s “zeroth-order”, random-placement theory of species-area curves. The random-placement model is suitable for natural random distribution that is underlying processes. It describes the slow-rapid slow accumulation of new species in a region. In other words, it is of the logistic type [53]. Consider a region of total area \( A_0 \) within which individuals of various species are located. Assume that there are \( N \) species in the area \( A_0 \), and that the \( i \)-th species is represented by \( n_i \) individuals. Consider any sub-region of area \( A < A_0 \). Under the assumption of independent, random placement of individuals, the probability that a given member of the \( i \)-th species does not reside in a sub-region of size \( A \) is simply \((1-A/A_0)\). Similarly, the probability that all members of species \( i \) lie outside of \( A \) is given by \((1-A/A_0)^n_i\). Thus, the probability that at least one member of the \( i \)-th species resides in the sub-region \( A \) is \((1-A/A_0)^{n_i}\) which, in turn, yields an expression for the mean number of species in \( A \), denoted \( S(A) \) [53]:

\[
S(A) = N - \sum_{i=1}^{N} (1-A/A_0)^{n_i}
\]

**SAD Models**

Preston [41] adopted the lognormal model to describe a biological community with few dominant and rare species, but with numerous medium-abundant species. His model is:
\[ S(R) = S_m \exp[-\alpha^2(R - R_m)^2] \]

where \( S(R) \) is the species number of the \( R \)th octave; \( S_m \) is the modal number of species in the \( R \)th octave; \( R_m \) is the “height” of the distribution curve. \( \alpha \) is a parameter describing the amount of spread of the distribution [41,54]. In order to fit the field data to the lognormal distribution, we adopted the linear regression model \( \ln S(R) = \ln S_m - \frac{1}{2}(R - R_m)^2 \) to calculate the parameters.

The second SAD model is the *logseries* distribution model [35]:

\[ S(R) = S_m/[1 + \alpha^2(R - R_m)^2] \]

The means of parameters are the same as in the lognormal model.

The third SAD model is the *logcauchy* distribution model. Fisher et al. [24] first used the logseries model to fit the SAD of Malayan butterflies and Lepidoptera. It is one of the two well-known SAD models (the other being the lognormal model). The number of species with \( n \) individuals in a community is:

\[ S(n) = \alpha n^a / n \]

where \( a \) and \( x \) are two parameters, \( a > 0 \) and \( 0 < x < 1 \) (in most cases \( x > 0.9 \)). However, these two parameters are not independent. The parameter \( a \) is called Fisher’s \( x \) diversity index. It was widely used in 1960’s and 1970’s as a diversity index and has found renewed interest through the recently developed neutral theory of biodiversity [45]. In Hubbell’s theory, \( a \) is a fundamental biodiversity parameter. He and Hu [55] further showed that Hubbell’s fundamental biodiversity parameter is a function of Simpson’s diversity.

**Results**

**Tree Density and Diversity**

Measures of tree density and diversity per hectare varied widely between the six communities (Table 1), ranging from 1440 to 2400 stems (mean ± S.E. = 2100 ± 10). 69 to 142 observed species (mean ± S.E. = 104.0 ± 14.1), and 34 to 229.7 estimated species (mean ± S.E. = 140.8 ± 20.5).

Observed and estimated species richness resulted in broadly similar rankings of the six communities which is reflected in these two diversity measures being significantly correlated (Spearman rank correlation, \( n = 6, Z \)-value = 2.08, \( P \)-value = 0.04). However, there were interesting exceptions: first, communities C and F were...
tied in observed species richness, but community C had a much higher estimated species richness than community F and, second, community A had a higher observed species richness but lower estimated species richness than community D. These inconsistencies and the fact that the estimated values are significantly higher than the observed values (One-sample sign test, \( n = 6 \), \( P \)-value = 0.03) further support the view that observed species richness is a negatively biased measure of total species richness (see Discussion). Therefore, we do not consider observed species richness from hereupon.

Ranking of communities by estimated species richness was not associated with forest type, with the ranking from lowest to highest estimated species richness being: monsoon evergreen broad-leaved forest, needle and broad-leaved mixed forest, needle and broad-leaved mixed forest, monsoon evergreen broad-leaved forest, broad-leaved forest and broad-leaved forest. Given this particular sequence and the same sample size, no trend can possibly be discerned between forest types.

Likewise, ranking of communities by number of stems per ha was not associated with forest type, with the ranking from lowest to highest tree density being: needle and broad-leaved mixed forest, monsoon evergreen broad-leaved forest, monsoon evergreen broad-leaved forest, needle and broad-leaved mixed forest. Given this particular sequence and the same sample size, no trend can possibly be discerned between forest types.

Finally, there was no correlation between number of stems and estimated species richness (Spearman rank correlation, \( n = 6 \), \( \zeta \)-value = 0.32, \( P \)-value = 0.75).

Species-area Relationships

Using the \( F \)-test, each of the three SAR models fitted highly significantly with the observed numbers by randomized sampling as well as sequential sampling (Tables 2 and 3, respectively). This good fit can also be observed graphically for each of the plotted species-area relationships (Fig. 1).

Using the adjusted coefficient of determination (\( R^2_a \)-value) for comparison (Tables 2 and 3), the logarithm model had the better fit among the three models (mean = 0.9878 for sequential sampling, mean = 0.9992 for randomized sampling), with three out of six of its \( R^2_a \)-values above 0.99 for randomized sampling. The power model was the second better model (mean = 0.9639 for sequential sampling, mean = 0.9842 for randomized sampling) while the random-placement model performed worst (no sequential sampling performed, mean = 0.9415 for randomized sampling). Because the random-placement model performed worst, if we average over all results, randomized sampling \( (n = 18, \text{ mean } = 0.9749) \) performed worse than sequential sampling \( (n = 12, \text{ mean } = 0.9758) \). However, once we exclude the random-placement model because it was not used for sequential sampling, we can compare the results from only the logarithm and power models; we then observe that randomized sampling \( (n = 12, \text{ mean } = 0.9917) \) performed better than sequential sampling \( (n = 12, \text{ mean } = 0.9758) \). We argue that this better performance is due to randomized sampling smoothing the resulting SA curves (see Discussion).

The SARs (Fig. 1) illustrate that communities D, E and F were fitted very well beyond area sizes of about 4000–5000 m\(^2\). Communities C and F fitted well using the random-placement model at all area sizes while the communities C, E and F were fitted well with the logarithm model. Community A fitted worse than the other communities for all three models. All communities fitted well to the random-placement model beyond area sizes of about 7000 m\(^2\), suggesting that the random-placement model describes SARs better at larger scales.

Species-abundance Distributions

All the SA data sets were statistically fitted using the three SAD models (Tables 4–5) with each fit also displayed graphically in Fig. 2. Using the coefficient of determination (\( R^2_a \)) for comparison, the mean \( R^2_a \)-values over the six communities were 0.738, 0.648...
and 0.586 for the logcauchy, lognormal and logseries model, respectively. This ranking is also evident in that the logcauchy had two $R^2$ values above 0.8 and the lognormal and logseries one above 0.8 and one above 0.7. Therefore, using $R^2$-values as criteria, the logcauchy model performed better than other two models.

The mean $R^2$-values of the six communities A–F were 0.579, 0.690, 0.729, 0.548, 0.642 and 0.556, respectively; thus, the best average fit for three SAD models was for community B and the worst for community D.

The three SAD models were graphically fitted using the SA data sets (Fig. 2). The results showed that the left side of each of the three distributions was apparently truncated for each of the six communities, which suggests that many rare species exist within these communities which were not detected within our samples.

**Discussion**

**Tree Density and Diversity**

The six sub-tropical forest communities displayed a wide range for stem density and estimated species richness (by a factor of 1.64 and 2.45, respectively). For comparison, species richness in three 1-ha plots of tropical seasonal rainforest in south-west China varied only by a factor of 1.26 and 1.61 for trees and treelets.
This large variation among the six plots cannot be explained by forest type as the largest variation for stem density is within the needle and broad-leaved mixed forest type (Table 1). Currently, we cannot ascribe any factors responsible for such large variation, but latitude, altitude, topography and development history of each community is different, therefore, a combination of habitat heterogeneity and/or evolutionary-historical factors may be responsible for these differences [63,64].

Surprisingly, neither stem density nor estimated species richness seemed to be influenced in any way by forest type. However, given the insufficient sample sizes for each forest type, not much should be read into this result. We are surprised that no correlation was found between stem density and estimated species richness, as there is often such a positive relationship between the number of individuals and the number of species [65].

Our results further support the view that observed species richness is a negatively biased measure of total species richness. This is the logical consequence of the shape of the SA curve if area is substituted for sampling effort. As sampling effort increases, species richness will increase, so it logically follows that insufficient sampling effort will lead to a negatively biased estimate of total species richness (also called observed species richness). Therefore, species richness estimators (as, for example, implemented in the program EstimateS or SPADE) need to be used to correct for this inherent sampling bias [66,29]. The resulting estimated species richness is generally much closer to the total species richness than the observed species richness.
Species-area Relationships

The question of the relationship between area and number of species is one of the oldest questions ecologists have posed [67], and the SAR is one of the most important relationships when investigating this relationship as well as related problems involving species diversity, conservation biology and landscape ecology [68,69,70,4,71]. Many models have been put forward to describe the SAR [29], three of which we applied in this study.

Every one of the tested three SAR models (power, logarithm, random-placement) fitted very well to the subtropical community data, with minor differences (see Results). Comparing the fits of these three models using mean $R_a^2$-values showed slightly better average fits for the logarithm model than for the power and the random-placement model, a result supported by Wei et al. [57].

Plotting the SAR for each community and visually comparing the fitted models gives us some interesting insights into the effect of sampling size (in our case, size of sampled area) on the goodness of fit of each model (Fig. 1). While there are examples where the respective model fits the real data very well for all sampling sizes (e.g., community F fitted by the logarithm model), which is reflected by very high corresponding $R_a^2$-values, other examples reveal that the fit is only good for some sampling sizes, with $R_a^2$-values also varying among different communities. In most cases, the fit is better for intermediate to high sampling sizes and worse for small sampling sizes. Random sampling effects usually have a greater proportional effect at small sampling sizes, which may explain why the real data curves do not conform so well to the model curves at lower sampling sizes. Randomization of sampling order usually removes these sampling effects, and therefore the resulting smoothed curves usually display better model fits even at low sampling sizes.

Several authors have argued that the performance of SAR models is dependent on the sampling size [26,72]. For example, He & Legendre [27] suggested the power model best fits for small to intermediate sampling sizes, the exponential model best fits for small samples, and the logistic model is the best for small to large scales (250,000 m²). And consequently, that there is no model that is universally best, but that each model’s performance depends on sampling scales and SADs [26]. Because the underlying SADs are different for different communities, Tjørve [73] claims that one should not expect the same SAR model to be the best fit for both sample area (census patches) data sets and isolate (habitat patches or islands) data sets [73], or across different scales [see also [74]].

The SAR curves in this study showed similarity to those in Condit’s [75] study of tropical forests in that there was no asymptote in the SAR curves (Fig. 1). Though, former studies found the distribution of 90% species (abundance≥20) in Dinhu plot (200000 m²) were affected by the terrain factors [76]. However, based on the habitat partitioning theory, there should be an asymptote in the SAR curve, so our results show the habitat

| Model   | Community | Parameter | $R_a^2$ |
|---------|-----------|-----------|---------|
| Lognormal | A        | 18.220    | 1.103   | 4  | 0.805 |
|         | B        | 37.521    | 0.756   | 3  | 0.895 |
|         | C        | 16.516    | 0.869   | 3  | 0.653 |
|         | D        | 10.121    | 0.879   | 1  | 0.453 |
|         | E        | 19.527    | 1.045   | 4  | 0.602 |
|         | F        | 17.067    | 0.988   | 4  | 0.481 |
| Logcauchy | A      | 16.789    | 0.373   | 4  | 0.709 |
|          | B      | 39.7640   | 0.420   | 2  | 0.942 |
|          | C      | 20.693    | 0.230   | 1  | 0.838 |
|          | D      | 16.681    | 0.344   | 1  | 0.727 |
|          | E      | 18.224    | 0.168   | 1  | 0.620 |
|          | F      | 15.204    | 0.219   | 2  | 0.591 |

| Model   | Community | Parameter | $x$     | $a$     | $R_a^2$ |
|---------|-----------|-----------|---------|---------|---------|
| Logseries | A        | 0.994     | 17.379  | 0.223  |
|         | B        | 0.989     | 37.889  | 0.833  |
|         | C        | 0.995     | 17.883  | 0.697  |
|         | D        | 0.997     | 12.335  | 0.463  |
|         | E        | 0.996     | 21.477  | 0.704  |
|         | F        | 0.993     | 18.811  | 0.595  |

Table 4. The fitted parameters and results for the lognormal and logcauchy models (see Methods for details).

doi:10.1371/journal.pone.0095890.t004

Table 5. The fitted parameters and results of the $\chi^2$ tests for the logseries model (see Methods for details).

doi:10.1371/journal.pone.0095890.t005
partitioning theory is not play leading role in the subtropical forest for small to intermediate scales. We think the lack of an asymptote is the prediction of a spatially explicit, zero-sum, and community drift model that is incorporated with the speciation and the SAR pattern [77,78]. According to Hubbell’s description of the community, there will always be rare species [75], as, for example, illustrated by the SAD graphs in Fig. 2. As a result, the number of species in the first octaves (number of rare species) are always larger than 1. As in Hubbell’s description [75], rare species exist in our communities, this also corroborating Hubbell’s theory. But our study can’t approve relative rate of colonization increases while rate of extinction decreases when area increases. So our result can’t prove area per se or equilibrium hypothesis.

Species-abundance Distributions

Preston [41] first formulated the lognormal equation of the SAD by the method of “octaves”, using it to explain the division of resource use by species coexisting in the same community. Whittaker [40] also suggested that certain communities accorded with certain models. The lognormal model is supposed to describe communities where the total number of species is large, and the abundance is determined by many independent factors multiplicatively, whereas the logseries model describes the abundance of species whose diversity is lower or in which the species compete for a single resource [38,33,35]. The lognormal model can estimate the theoretical total number of species in the whole community using a truncated distribution [54], while the logseries model cannot be used for this purpose [3]. Using mean $R^2$-values for comparative performance evaluation, the logcauchy model had the better fit among the three models. Moreover, subtropical forests have high diversity, and their large species number includes many rare species (see Discussion above), which explains why all six communities appeared to be left-side truncated (Fig. 2).

Conclusions

To conclude, we found that our measures of tree density and diversity varied widely among the three forestry types, with the reasons remaining unclear, which should be subjected of further study. The three SAR models fitted most real data of subtropical forest trees very well for most sampling sizes. But due to intermediate sampling sizes, the logarithm model had the better fit among the three models, especially using randomized sampling. When fitting SADs, the logcauchy model had the better fit among the three tested models. Furthermore, our results suggest that many rare and undiscovered species remain undetected.

Acknowledgments

We thank Dinglushan Forest Ecosystem Research Station (DFERS), Nanling Nature Reserve, Maershan Nature Reserve and Jinggangshan Nature Reserve bureau for supporting our field investigations. We thank Bruno A. Walther for his helpful revision and good suggestion. We thank Professor Faungliang He of University of Alberta, Dr. I Fang Sun of National Dong Hwa University, Dr. Richard Condit of CTFS, Dr. Pierre Legendre of University of Montreal for training of data analysis. We also thank Dr. Wan hui Ye, Hong lin Cao, Jun Hui Shi, Xiao Yong Zhou, Qian Mei Zhang and Jiong Li for their help in field work.

Author Contributions

Conceived and designed the experiments: LL ZLH. Performed the experiments: JYL ZCC. Analyzed the data: SGW GJL. Wrote the paper: SGW ZYY.

References

1. Jaccard P (1902) Lois de distribution florale dans la zone alpine. Bull Soc Vaudoise Sci Nat 30: 69–130.
2. Jaccard P (1908) Nouvelles recherches sur la distribution florale. Bull Soc Vaudoise Sci Nat 44: 223–270.
3. Pielou EC (1977) Mathematics Ecology. New York: Wiley-Interscience. 320 p.
4. Rosenzweig ML (1995) Species diversity in space and time. Cambridge: Cambridge Univ. Press. 436 p.
5. Shen G, Yu M, Hu XS, Mi X, Ren H, et al. (2009) Species-area relationships explained by the joint effects of dispersal limitation and habitat heterogeneity. Ecology 90: 3033–3041.
6. Wang XG, Wiegand T, Wolf A, Howe R, Davies SJ, et al. (2011) Spatial patterns of species richness in two temperate forests. J Ecol 99: 1392–1393.
7. Turner VR, Tjørve E (2005) Scale-dependence in species-area relationships. Ecography 28: 721–730.
8. Barkman JH (1989) A critical evaluation of minimum area concepts. Plant Ecol 85: 89–104.
9. Hopkins B (1995) The concept of minimal area. J Ecol 45: 441–449.
10. Hubbell SP, Foster RB (1985) Diversity of canopy trees in a Neotropical forest and implications to conservation. Oxford: Blackwell Scientific Publications. 254 p.
11. Palmer MW (1990) The estimation of species richness by extrapolation. Ecology 71: 1195–1198.
12. Obeso JR, Arno C (1992) Plant-Species Richness and Extinction on Isolated Dunes along the Rocky Coast of Northwestern Spain. J Veg Syst. 3: 129–132.
13. Brooks TM, Pimm SL, Oyugi JO (1999) Time lag between deforestation and bird extinction in tropical forest fragments. Conserv Biol 13: 1140–1150.
14. Brooks TM, Tobias J, Balmford A (1999) Deforestation and bird extinctions in the Atlantic forest. Annu Conserv 2: 211–222.
15. MacArthur RH, Wilson EO (1967) The Theory of Island Biogeography. Princeton, NJ: Princeton University Press. 203 p.
16. Williamson MH (1981) Island populations. Oxford: Oxford University Press. 236 p.
17. Kohl DD, Walsh DW (1994) Plant species richness -the effect of island size and habitat diversity. J Ecol 82: 367–377.
18. Kochy M, Rydin H (1997) Biogeography of vascular plants on habitat islands, peninsulas and mainland in an east-central Swedish agricultural landscape. Nord J Bot 17: 215–223.
19. Kelly BJ, Bastow WJ, Mark AF (1989) Causes of the species-area relation:a study of islands in lake Manapouri, New Zealand. J Ecol 77: 1021–1028.
20. Krauss J, Klein AM, Steffen-Dreweser I, Tschurnke T (2004) Effects of habitat area, isolation, and landscape diversity on plant species richness of calcareous grasslands. Biodivers Conserv 13: 1427–1439.
21. Arrhenius O (1921) Species and area. J Ecol 9: 95–99.
22. Gleason HA (1922) On the relation between species and area. Ecology 3: 158–162.
23. Gleason HA (1925) Species and area. Ecology 6: 66–74.
24. Fisher R, Corbet A, Williams C (1983) The relation between the number of species and the number of individuals in random sample of an animal population. J Anim Ecol 12: 42–58.
25. Archibald EEA (1949) The Specific Character of Plant Communities: II. A Quantitative Approach. J Ecol 37: 274–288.
26. He FL, Legendre P (1996) On Species-Area Relations. Am Nat 115: 717–739.
27. He FL, Legendre P (2002) Species diversity patterns derived from species-area models. Ecology 83: 1185–1189.
28. Plotkin JB, Potts MD, Manokaran N, Lafrankie J, Ashton PS (2000) Species-area curves, spatial aggregation, and habitat specialization in tropical forests. J Theor Biol 207: 81–99.
29. Walther BA, Moore JL (2005) The concepts of bias, precision and accuracy, and their use in testing the performance of species richness estimators, with a literature review of estimator performance. Ecography 28: 815–829.
30. Suding AL, Storch D (2004) Power-law species-area relationships and self-similar species distributions within finite areas. Ecol Lett 7: 60–68.
31. Ovaskainen O, Hanski I (2003) The species-area relationship derived from species-specific incidence functions. Ecol Lett 6: 903–909.
32. Sugihara G (1988) Minimal community structure: an explanation of species abundance patterns. Am Nat 131: 770–787.
33. May RM (1975) Patterns of species abundance and diversity. In: Cody ML, Diamond JM, editors. Ecology and Evolution of Communities. Cambridge, MA: Belknap Press of Harvard University. 81–120.
34. Plotkin J, Muller-Landau H (2002) Sampling the species composition of a landscape. Ecology 83: 3344–3356.
35. Yin ZY, Peng SL, Ren H, Guo QF, Chen ZH (2004) LogCauchy, log -sech and lognormal distributions of species abundances in forest communities. Ecol Model 184: 329–340.
36. Newby MC (1999) Ecology:Concepts&Applications. New York: McGraw-Hill Science Engineering. 572 p.
37. Jackson CR, Churchill PF, Roden EE (2001) Successional development. Ecology 81: 555–556.
38. Whittaker RH (1965) Dominance and diversity in land plant communities. Science 147: 250–260.
39. Guo Q, Brown JH, Valone TJ (2002) Long-term dynamics of winter and summer annual communities in the Chihuahuan Desert. J Veg Sci 13: 575–584.
40. Whittaker RH (1972) Evolution and measurement of species diversity. Taxon 21: 213–251.
41. Preston FW (1948) The Commonness, And Rarity, of Species. Ecology 29: 254–283.
42. Cohen JE (1968) An alternative derivation of a species abundance relation. Am Nat 102: 165–171.
43. Kempton RA, Taylor LR (1974) Log-series and log-normal parameters as diversity discriminants for the Lepidoptera. J Anim Ecol 43: 381–399.
44. Bell G (2001) Neutral macroecology. Science 293: 2413–2418.
45. Hubbell SP (2001) The unified neutral theory of biodiversity and biogeography. Princeton: Princeton University Press. 34–36 p.
46. Krebs CJ (2009) Ecology: the experimental analysis of distribution and abundance. San Francisco: Pearson Benjamin Cummings.
47. Stohlgren TJ, Otsuki Y, Villa CA, Lee M, Belnap J (2001) Patterns of Plant Invasions: A Case Example in Native Species Hotspots and Rare Habitats. Biol Invasions 3: 37–50.
48. Batista JLF, Maguire DA (1998) Modeling the spatial structure of tropical forests. Forest Ecol Manag 110: 293–314.
49. Laurance WF (2007) Forest destruction in tropical Asia. Curr Sci India 93: 1544–1550.
50. Williams CB (1964) Patterns in balance of nature and related problems in quantitative ecology. New York: Academic Press. 324 p.
51. Buys MH, Marita JS, Boucher C (1994) A model for species-area relationships in plant communities. J Veg Sci 5: 63–66.
52. Liu CR, Ma KP, Yu SL (1999) Plant community diversity in Dongling Mountain, Beijing, China – the fitting and assessment of species-area curves. Acta Phytoecol Sin 23: 490–500.
53. Coleman BD (1981) On random placement and species-area relations. Math Biosci 54: 191–215.
54. Preston FW (1962) The canonical distribution of commonness and rarity. Ecology 43: 105–125.
55. Williams CR (1965) Plant community diversity in accordance with Simpson's diversity index. Ecol Lett 8: 386–390.
56. Colwell RK (2009) EstimateS: Statistical estimation of species richness and shared species from samples. Version 8.2. User’s Guide and application. http://purl.oclc.org/estimates.
57. Wei SG, Li L, Walther BA, Ye WH, Huang ZL, et al. (2010) Comparative performance of species-richness estimators using data from a subtropical forest tree community. Ecol Res 25: 93–101.
58. He FL, Hu XS (2005) Hubbell’s fundamental biodiversity parameter and the Simpson diversity index. Ecol Lett 8: 386–390.
59. Lomolino MV (1989) Interpretations and comparisons of constants in the species-area relations: an additional caution. Am Nat 133: 277–280.
60. Coleman BD (1981) On random placement and species-area relations. Math Biosci 54: 191–215.
61. Preston FW (1962) The canonical distribution of commonness and rarity. Ecology 43: 105–125.
62. Guo Q, Brown JH, Valone TJ (2002) Long-term dynamics of winter and summer annual communities in the Chihuahuan Desert. J Veg Sci 13: 575–584.
63. Condit R, Latham R, Qian H (1999) Global patterns of tree species richness in moist forests: distinguishing ecological influences and historical contingency. Oikos 86: 369–373.
64. Batista JLF, Maguire DA (1998) Modeling the spatial structure of tropical forests. Forest Ecol Manag 110: 293–314.
65. Cole BD (1981) On random placement and species-area relations. Math Biosci 54: 191–215.
66. Williams CR (1964) Patterns in balance of nature and related problems in quantitative ecology. New York: Academic Press. 324 p.
67. Buys MH, Marita JS, Boucher C (1994) A model for species-area relationships in plant communities. J Veg Sci 5: 63–66.
68. Liu CR, Ma KP, Yu SL (1999) Plant community diversity in Dongling Mountain, Beijing, China – the fitting and assessment of species-area curves. Acta Phytoecol Sin 23: 490–500.
69. Coleman BD (1981) On random placement and species-area relations. Math Biosci 54: 191–215.
70. Preston FW (1962) The canonical distribution of commonness and rarity. Ecology 43: 105–125.
71. Williams CR (1965) Plant community diversity in accordance with Simpson’s diversity index. Ecol Lett 8: 386–390.
72. Colwell RK (2009) EstimateS: Statistical estimation of species richness and shared species from samples. Version 8.2. User’s Guide and application. http://purl.oclc.org/estimates.
73. Wei SG, Li L, Walther BA, Ye WH, Huang ZL, et al. (2010) Comparative performance of species-richness estimators using data from a subtropical forest tree community. Ecol Res 25: 93–101.
74. He FL, Hu XS (2005) Hubbell’s fundamental biodiversity parameter and the Simpson diversity index. Ecol Lett 8: 386–390.
75. Condit R, Latham R, Qian H (1999) Global patterns of tree species richness in moist forests: distinguishing ecological influences and historical contingency. Oikos 86: 369–373.