Spectrum of CFTR mutations in Chechen cystic fibrosis patients: high frequency of c.1545_1546delTA (p.Tyr515X; 1677delTA) and c.274G>A (p.Glu92Lys, E92K) mutations in North Caucasus

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Abstract

Background: Cystic fibrosis (CF; OMIM #219700) is a common autosomal recessive disease caused by pathogenic variants (henceforward mutations) in the cystic fibrosis transmembrane conductance regulator gene (CFTR). The spectrum and frequencies of CFTR mutations vary among different populations. Characterization of the specific distribution of CFTR mutations can be used to optimize genetic counseling, foster reproductive choices, and facilitate the introduction of mutation-specific therapies. Chechens are a distinct Caucasian ethnic group of the Nakh peoples that originated from the North Caucasus. Chechens are one of the oldest ethnic groups in the Caucasus, the sixth largest ethnic group in the Russian Federation (RF), and constitute the majority population of the Chechen Republic (Chechnya). The spectrum of CFTR mutations in a representative cohort of Chechen CF patients and healthy individuals was analyzed.

Methods: Molecular genetic analysis of 34 CFTR mutations (representing approx. 80–85% of mutations in multiethnic CF populations of the RF) was performed in 32 CF patients from 31 unrelated Chechen families living in Chechnya. One hundred randomly chosen healthy Chechens were analyzed for the 15 most common "Russian" mutations. The clinical symptoms in Chechen CF patients with different CFTR genotypes were investigated.

Results: High frequencies of c.1545_1546delTA (p.Tyr515X; 1677delTA) (52 out of 64 CFTR alleles tested; 81.3%) and c.274G>A (p.Glu92Lys, E92K) (8/64, 12.5%) mutations were found. Twenty patients were homozygous for the c.1545_1546delTA mutation, and eight were compound heterozygous for the c.1545_1546delTA and c.274G>A mutations. Three carriers of the c.1545_1546delTA mutation were also found in the cohort of 100 apparently healthy Chechens (frequency – 0.015). The c.1545_1546delTA and c.274G>A mutations are linked to the same haplotype (22–7–16–13) of intragenic Short Tandem Repeat markers, i.e., IVS1CA, IVS6aGATT, IVS8CA, and IVS17bCA. (Continued on next page)
Background
Cystic fibrosis (CF; OMIM # 219700) is a common autosomal recessive disease caused by pathogenic variants (henceforward mutations) in the CFTR gene (OMIM #602421). To date, over 2000 pathogenic, likely pathogenic, or benign CFTR variants have been identified [1]. The distribution of CFTR mutations varies widely in different populations [2]. Therefore, identification of the spectrum of the most common CFTR mutations in a given population can be used to optimize genetic counseling, foster reproductive choices, and facilitate implementation of mutation-specific therapies.

In this study, we have focused on the Chechen population that lives within the Chechen Republic of the Russian Federation (RF), located in the North Caucasus (see Fig. 1). Chechens are an ancient Nakh language speaking group that together with a related Ingush population are jointly termed “Vainakhs” [3, 4]. Both populations predominantly live in the Chechen and Ingush Republics of the RF (Fig. 1). Small portions of this distinct population also inhabit several neighboring districts of Dagestan and Georgia. Regarding their population size, the Chechens rank fourth in the Caucasus (after Azerbaijanis, Georgians, and Armenians) and sixth within the entire multiethnic RF (after Russians, Tatars, Ukrainians, Bashkirs, and Chuvashes). The total number of Chechens around the world ranges between 1.5–2 million, with the majority of them (i.e. 1,344,122 according All-Russia Population Census) living in Chechnya proper [5].

Available historical and linguistic studies describe a single ancestral population that had been living on the northern slopes of the Great Caucasian Range (i.e., between the Caspian and Black sea regions) already several thousand years ago [6]. This presumed historic population had been associated with a Western Asian culture, distinct from the East Caucasus populations. Recent population genetic studies utilizing Y-chromosome haplotypes have demonstrated a robust genetic delineation of the Nakh language speaking people of Chechnya, Dagestan, and Ingushetia from other populations of the North Caucasus [7]. In this regard, Vainakhs are considered one of the oldest autochthonous Northern Caucasus ethnic groups [8]. The struggle for an independent Chechnya on the basis of the Nakh cultural-linguistic uniqueness ended in the 17th and 18th centuries CE (Common Era) when the Vainakh peoples became citizens of the former Russian Empire. In 1810, the Ingushs accepted Russian citizenship, followed by Chechens in 1859 [9].

In this study, we analyzed the spectrum of CFTR mutations in a representative cohort of Chechen CF patients and healthy individuals. According to the best of our knowledge, this is the first study of this population, which also has relevance for the strong Chechen diaspora in the RF and beyond.

Methods
Our study included a representative cohort of 32 Chechen CF patients from 31 unrelated families. Except for a single case born in 1987, all patients were born between years 2004–2016. Patients and their families were self-reported ethnic Chechens living predominantly in the capital city Grozny. Cystic fibrosis was diagnosed based on consensus criteria for non-screened populations at the “Maternity Hospital” (Grozny, Chechnya) and in part at the Research and Clinical Department of Cystic Fibrosis of the Federal State Scientific Budgetary Institution “Research Center for Medical Genetics” or Federal State Autonomous Institution “National Medical Research Center of Children’s Health” of the Ministry of Health of the Russian Federation in Moscow.

We used a representative group of 100 unrelated apparently healthy Chechens as controls. The majority of them were also drawn from Grozny (67/100 of the entire control group), with the remaining volunteers originating from other regions of Chechnya. Ethnicity up to the third generation had been validated through a structured questionnaire filled under supervision. Healthy individuals, CF patients, or their legal representatives provided written informed consent for the study. This research project received approval from the Ethics Committee of the “Research Centre for Medical Genetics” (Moscow).

The “Wizard Genomic DNA Purification Kit” (Promega, USA) was used for DNA extraction from whole blood samples where EDTA was used as an anticoagulant. Initially, we examined the 34 most common CFTR mutations utilized for diagnosis of CF within the multiethnic
RF that account for over 85% of all CF-causing mutations [10]. In-house molecular genetic methods previously described [11], including amplified fragment length (AFLP) and restriction fragment length (RFLP) polymorphism techniques were utilized to detect insertion/deletion variants and nucleotide substitutions, respectively. The panel of tested CF-causing mutations currently includes mutations: c.54-5940_273+10250del21kb (p.Ser18ArgfsX16;
CFTR dele2,3), c.254G > A (p.Gly85Glu; G85E), c.262_263delITT (p.Leu88IlefsX22; 394delITT), c.274G > A (p.Glu92Lys; E92K), c.287C > A (p.Ala96Glu; A96E), c.350G > A (p.Arg171His; R171H), c.411_412insCTA (p.Leu138dup; L138ins), c.472dupA (p.Ser158LysfsX5; 604insA), c.489+1G > T (621+1G > T), c.1000C > T (p.Arg334Trp; R334W), c.1040G > C (p.Arg347Pro; R347P), c.1397C > G (p.Ser466X; S466X), c.1519_1521delATC (p.Ile507del; I507del), c.1521_1523delCTTT (p.Phe508del; F508del), c.1545_1546delTA (p.Tyr515X; 1677delTA), c.1585-1G > A (1717-1G > A), c.1624G > T (p.Glu92Lys, E92K), c.287C > A (p.Ala96Glu, A96E), c.411_412insCTA (p.Leu138dup; L138ins), c.1000C > T (p.Arg334Trp; R334W), c.3846G > A (p.Trp1282X; W1282X), c.3817delGT (p.Ser1273LeufsX28; 3944delTG), c.3844T > C (p.Ser1273ProfsX4; 3971delT), c.3718-2477C > T (3849+10kbC-T), c.3816_3821delT), c.3821AA > G (p.Trp1310_Gln1313del) were identified once in each case (Table 1).

Four intragenic short tandem repeats (STR) (IVS1CA, IVS6aGATT, IVS8CA, and IVS17bCA) were examined as previously described [12]. STR haplotypes were established by segregation analysis of given CFTR alleles within CF families. STR haplotype frequencies in healthy controls were analyzed for the 15 most common mutations: c.54-5940_273+10250del21kb (p.Ser18ArgfsX16; CFTR dele2,3), c.3817-2477C > T (3849+10kbC-T), c.3816_3817delGT (p.Ile507fsX28; 3944delTG), c.3844T > C (p.Trp1282Arg; W1282R), c.3846G > A (p.Tyr515X; 1677delTA), c.3821AA > G (p.Trp1282X; W1282X), and c.3909C > G (p.Asn1303Lys; N1303K) [10].

Subsequently, for a case in which one CFTR mutation remained unidentified we carried out direct Sanger DNA sequencing of the entire CFTR coding region, including adjacent splice sites and the 3′-untranslated CFTR region [10]. Positive cases were confirmed in parents to establish their linkage phase. Random controls were analyzed for the 15 most common “Russian” mutations: c.54-5940_273+10250del21kb (p.Ser18ArgfsX16; CFTR dele2,3), c.3817-2477C > T (3849+10kbC-T), c.3816_3817delGT (p.Ile507fsX28; 3944delTG), c.3844T > C (p.Trp1282Arg; W1282R), c.3846G > A (p.Tyr515X; 1677delTA), c.3821AA > G (p.Trp1282X; W1282X), and c.3909C > G (p.Asn1303Lys; N1303K) [10].

To compare the clinical course of CF in the studied cohort, the patients were divided into the two most prevalent groups: 17 homozygous for c.1545_1546delTA (p.Tyr515X; 1677delTA) – 52/64 CFTR alleles (81.3%), and c.274G > A (p.Glu92Lys, E92K) – 8/64 alleles (12.5%) (Table 1). Twenty patients were homozygous for c.1545_1546delTA (p.Tyr515X; 1677delTA), while 8 were compound heterozygous for the c.1545_1546delTA (p.Tyr515X; 1677delTA) and c.274G > A (p.Glu92Lys, E92K) mutations. In addition, c.287C > A (p.Ala96Glu, A96E), c.1000C > T (p.Arg334Trp; R334W), c.3846G > A (p.Trp1282X; W1282X), and a novel variant c.3925_3936delCAGTGAGGTGAT (p.Trp1310_Gln1313del) were identified once in each case (Table 1).

In the control group of 100 randomly chosen Chechen individuals, 3 carriers of the c.1545_1546delTA mutation were detected (1.5%), while none of the 14 remaining common “Russian” CFTR mutations were detected.

The c.1545_1546delTA and c.274G > A mutations were present on a single intra-CFTR short tandem repeat (STR) haplotype “22–7–16–13” (as a sequence of IVS1CA, IVS6aGATT, IVS8CA, and IVS17bCA STR markers), this suggesting their common ancestral origin.
age at last clinical examination, age at diagnosis, sweat Cl\(^-\) concentrations, or BMI values. None of the most common before-mentioned complications (meconium ileus, liver cirrhosis, diabetes, polyposis) were revealed in either group. Significant differences were observed only in terms of pancreatic insufficiency in that all patients from Group 1 had fecal elastase 1 concentrations below 50 \(\mu g/g\), while all patients from Group 2 had concentrations over 200 \(\mu g/g\) (\(p < 0.0001\)), this indicating lower degree of pancreatic exocrine dysfunction associated with the presence of c.274G > A. Similarly, the proportion of patients with chronic \(P.\ aeruginosa\) lung colonization was significantly higher in Group 1 compared to Group 2 (69.0% vs. 14.0%, respectively; \(p = 0.024\)). The differences in the other studied microorganisms were not significant. Overall, the presence of the c.274G > A mutation is associated with less severe course of the disease than in the c.1545_1546delTA homozygotes (Table 2).

### Discussion
To the best of our knowledge, this is the first study on the distribution of \(CFTR\) mutations in the Chechen population (Fig. 1). We have provided evidence that the c.1545_1546delTA and c.274G > A mutations (as validated by the www.cftr2.org database) account for the majority of \(CFTR\) mutations in this population. These CF alleles very likely have a common origin, since they are residing on a

### Table 1 Distribution of \(CFTR\) gene mutations in Chechen CF patients

| \(CFTR\) genotypes                                                                 | Number of patients (n = 32) | Frequency |
|-----------------------------------------------------------------------------------|----------------------------|-----------|
| c.[1545_1546delTA];[1545_1546delTA] (p.[Tyr515X];[Tyr515X]; 1677delTA/1677delTA) | 20                         | 0.625     |
| c.[1545_1546delTA];[274G > A] (p.[Tyr515X];[Glu92Lys]; (1677delTA/ E92K)      | 8                          | 0.251     |
| c.[1545_1546delTA];[287C > A] p.[Tyr515X];[Ala96Glu]; 1677delTA/A96E            | 1                          | 0.031     |
| c.[1545_1546delTA];[1000C > T] p.[Tyr515X];[Arg334Trp]; 1677delTA/R334W        | 1                          | 0.031     |
| c.[1545_1546delTA];[3846G > A] p.[Trp1282X]; 1677delTA/W1282X                   | 1                          | 0.031     |
| c.[1545_1546delTA];[3925_3936delCAGTGGAGTGAT] (p.[Tyr515X],[Trp1310_Gln1313del])| 1                          | 0.031     |

| \(CFTR\) alleles                                                                 | Number (n = 64) | Frequency |
|---------------------------------------------------------------------------------|----------------|-----------|
| c.1545_1546delTA (p.Tyr515X; 1677delTA)                                          | 52             | 0.8130    |
| c.274G > A (p.Glu92Lys, E92K)                                                    | 8              | 0.1250    |
| c.287C > A (p.Ala96Glu, A96E)                                                    | 1              | 0.0155    |
| c.1000C > T (p.Arg334Trp; R334W)                                                 | 1              | 0.0155    |
| c.3846G > A (p.Trp1282X; W1282X)                                                 | 1              | 0.0155    |
| c.3925_3936delCAGTGGAGTGAT (p.Trp1310_Gln1313del)                                | 1              | 0.0155    |

### Table 2 Comparison of two groups of Chechen CF patients

| \(CFTR\) genotype                                                                 | Group 1 (n = 17) | Group 2 (n = 8) | \(p\)-value |
|-----------------------------------------------------------------------------------|-----------------|----------------|-------------|
| Age at last clinical examination (yrs)                                           | 5.66 ± 8.28 (0.29÷31.46) | 4.53 ± 4.13 (0.92÷11.92) | > 0.05     |
| Age at diagnosis (yrs)                                                           | 1.66 ± 0.91 (0.00÷20.18) | 1.07 ± 0.91 (0.16÷3.00) | > 0.05     |
| BMI (kg/m\(^2\))                                                                | 14.93 ± 3.12 (12.30÷24.88) | 15.92 ± 2.55 (13.00÷21.00) | > 0.05     |
| Sweat chloride (mM/L)                                                            | 120.25 ± 36.27 (100.00÷134.00) | 120.25 ± 15.62 (100.00÷134.00) | > 0.05     |
| FEV\(_1\) (% predicted)                                                          | 82.66 ± 26.85 (52.00÷102.00) | 91.50 ± 2.12 (90.00÷93.00) | > 0.05     |
| FVC (% predicted)                                                                | 93.66 ± 27.09 (68.00÷122.00) | 91.50 ± 2.12 (90.00÷93.00) | > 0.05     |
| Meconium ileus                                                                   | 0               | 0             |             |
| Liver cirrhosis                                                                  | 0               | 0             |             |
| CF-related diabetes mellitus                                                      | 0               | 0             |             |
| Fecal elastase 1 concentration ≥200 \(\mu g/g\)                                  | 0               | 8             | < 0.0001    |
| Fecal elastase 1 concentration < 200 \(\mu g/g\)                                 | 17              | 0             |             |
| \(S.\ aureus\) lung colonization                                                 | 44%             | 14%           | > 0.05      |
| \(P.\ aeruginosa\) lung colonization                                             | 69%             | 14%           | 0.024       |
common population-specific intra-CFTR STR haplotype that probably increased in frequency due to genetic drift.

The c.1545_1546delTA mutation was previously found to be common in populations neighboring or with historic links to the greater Black Sea region [2] (e.g., Bulgaria, Romania, Greece, Cyprus, and Turkey, including Northern Iran and Georgia [14]). It was also identified in other ethnic groups from the Northern Caucasus region (i.e., Ingush, Armenians, Ossetians, Dagestanis, etc.), albeit only in a small number of CF patients examined [10]. The fact that the c.1545_1546delTA mutation is found in autochthonous ethnic groups of the Caucasus region (Chechens, Ingush, Georgians, Armenians) and is linked to a single haplotype may point to a single source of origin (or penetration) of this mutation into the Caucasus region, and differences in frequencies seen in various populations may be due to gene drift.

Interestingly, the c.274G > A mutation was found at the highest frequency in Chechen CF patients (12.5%). This CF allele was previously found in Turkey [2], but also in Chuvash CF patients living in central parts of the RF (55.6%) [15]. According to the Russian Cystic Fibrosis Patient Registry (RCFPR), the prevalence of this allele in patients in the Volga-Ural region is 6.88% (Chuvash Republic – 53.19%, Udmurt Republic – 6.76%, Tatarstan – 2.38%, Bashkortostan – 1.37%, Samara region – 3.06%, Perm – 0.75%, and Orenburg regions – 1.96%), including the Khanty-Mansi autonomous region (Yugra) – 3.85%, as well as sporadically in many other RF regions [16]. Given its population distribution, the c.274G > A mutation could be associated with the historical resettlements of Turkic-speaking peoples in regions of RF mentioned above, including their migrations to the Caucasus and greater Black Sea geographical area.

Although the c.3846G > A mutation was found at a very high frequency in the neighboring Karachay-Cherkessia (Fig.1) [11], it was only sporadically observed in Chechens, thereby substantiating the strong genetic delineation of Vainakh populations from other ethnic groups residing in the Northern Caucasus. The penetration of the c.3846G > A mutation into the territory of the Northeast Caucasian region could be related to the migration of Jews from Byzantium through the northern Black Sea region or Georgia in the early Middle Ages or from Persia (Iran) in the late Middle Ages [11].

The c.287C > A mutation was previously described in Turkish CF patients, where its frequency is at 2.6% [2]. In the RF this mutation was also found in 2 patients from Dagestan, which is bordering Chechnya (Fig.1).

Finally, the c.1000C > T mutation is in CF populations from the greater Mediterranean region, such as those from southern France (1.2%), Greece (1.1%), Portugal (0.7%), and Spain (1.2%) [2], while in the RF it was sporadically observed in various ethnic groups at the frequency of 0.8% [16].

The comparison of key clinical parameters in the two groups of Chechen patients with different genotypes demonstrated that the allele c.274G > A is associated with higher residual pancreatic function and lower chronic lung colonization with pathognomonic microorganisms, in accordance with the data of the CFTR2 database [1].

Conclusions
The distribution of CFTR mutations in the Chechen CF population is unique in terms of the high frequency of mutations c.1545_1546delTA (p.Tyr515X; 1677delTA) and c.274G > A (p.Glu92Lys, E92K), which account for more than 90% of the mutant alleles in the studied ethnic group. Testing of the two CF-causing mutations is thus recommended for Chechen CF patients, since it allows identification of one or both mutant CFTR alleles in more than 99% of patients suspected of being affected by CF. Furthermore, we have confirmed the genetic delineation of the Chechen population from other ethnic groups of the Northern Caucasus (e.g. by low prevalence of the c.3846G > A mutation, which is dominant in adjacent Karachay-Cherkessia; Fig.1), as well as the role of historic migrations of Turkic-speaking peoples from Central Asia to the Northern Caucasus with the c.274G > A mutation very likely being their “marker” [17]. Analysis of genotype-phenotype correlations in two groups of Chechen CF patients (i.e. c.1545_1546delTA homozygotes versus c.1545_1546delTA/c.274G > A compound heterozygotes) demonstrated that the presence of the c.274G > A mutation is associated with generally less severe course of the disease. Our data will improve genetic counselling and provide a basis for the introduction of mutation-specific therapies in the future.

Abbreviations
BMI: Body mass index; CF: Cystic Fibrosis; CFTR: Cystic Fibrosis Transmembrane Regulator; CFTR2: The Clinical and Functional TRanslation of CFTR (CFTR2) (http://cftr2.org); DNA: Deoxyribonucleic acid; FEV1: Forced expiratory volume at 1 s. (spirometric parameter); FVC: Forced vital capacity (spirometric parameter); RF: Russian Federation; STR: Short Tandem Repeats

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Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.
Authors’ contributions
NVP contributed to the design of the study, performed molecular genetic experiments, analyzed and interpreted genetic and patient data, and wrote the manuscript. NYK contributed to the design of the study, analysis and interpretation of patient data, and writing of the manuscript. DKS ensured patients’ attendance, provided written informed consent, reviewed clinical examination of the patients, and collected samples for DNA research in Chechnya. AVP, TA, and TAV performed molecular genetic experiments. OIS, YVG, EIK, VDS, and OGN contributed to analysis of patient data and preparation of the manuscript. AVM analyzed and interpreted genetic and patient data and participated in writing the manuscript. MM Jr supervised the project, interpreted genetic and patient data, and reviewed the manuscript. RAZ contributed to design of the study, provided expeditions to Chechnya, performed clinical and genetic counseling, collected samples for DNA research, participated in writing the manuscript, and supervised all aspects of the project. All authors read and approved the final manuscript.

Ethics approval and consent to participate
The clinical and molecular genetic study was performed in accordance with the Declaration of Helsinki and was approved by the Institutional Review Board of the Federal State Budgetary Institution "Research Centre for Medical Genetics," Moscow, Russia, with written informed consent obtained from each participant and/or their legal representative as appropriate.

Consent for publication
Consent for publication was obtained from the legal guardians of the patients. The Ethics Committee approved this procedure.

Competing interests
The authors declare that they have no competing interests.

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