Protective Effect of N-Acetyl Cysteine on *Moringa Oleifera* Aqueous Leaf Extract-Induced Hepatotoxicity in Wistar Albino Rats

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A B S T R A C T

**Background:** *Moringa oleifera* (*M. oleifera*) is a commonly used medicinal and nutritive plant. The aqueous leaf extract from this plant contains a high concentration of alkaloids and they are toxic to body organs especially the liver leading to hepatotoxicity after prolonged use. However, if N-Acetyl cysteine (NAC) is administered together with *M. oleifera* extract, it may have a hepatotoxic protective effect.

**Objective:** To establish the protective effect of N-Acetyl cysteine against *M. oleifera* aqueous leaf extract-induced hepatotoxicity in the Wistar albino rats.

**Methods:** An experimental laboratory-based study was conducted at the department of Physiology, Makerere University, College of Health Sciences. Three treatment groups of six Wistar albino rats each, were dosed with *M. oleifera* extract, Paracetamol and NAC once a day for 28 days. Group I; negative control, received 8.05g/kg bwt of *M. oleifera* extract plus 1ml of normal saline (NS), Group II; test group, received 8.05g/kg bwt of *M. oleifera* extract plus 50mg/kg of NAC. Group III; positive control, received 750mg/kg bwt of Paracetamol plus 50mg/kg of NAC. On the 14th and 29th day, three animals were selected randomly from each group and sacrificed; blood samples were collected, the liver was harvested for histopathological analysis. Serum alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), bilirubin and total protein levels were determined.

**Results:** There was an increase in serum ALT, AST and ALP levels in the *M. oleifera* extract plus NS group which was a sign of hepatotoxicity. The *M. oleifera* extract plus NAC group showed normal serum ALT, AST and ALP levels with no significant changes in the bilirubin (P-value = 0.9089) and total protein levels (P-value = 0.8858).

**Conclusion:** The results have provided evidence that NAC administration with *M. oleifera* extract effectively prevents the occurrence of *M. oleifera* leaves extract-induced hepatotoxicity.

**Keywords:** *Moringa oleifera*, N-Acetyl-Cysteine, hepatotoxicity, Paracetamol, Wistar rats

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INTRODUCTION

*Moringa oleifera* is a plant found in many tropical and subtropical countries with significant medicinal and nutritional values. Different parts of *M. oleifera* (leaves, pods, seeds, bark and flowers) have been shown to exhibit wide range of pharmacological activities, with the leaves being the most commonly used. The biological activities of the plant are attributed to the
presence of many non-nutritive secondary metabolites which are bioactive in the leaves which include; polyphenols, simple sugars, tannins, vitamins, carotenoids, phytates, phenolic acids, flavonoids, alkaloids, isothiocyanates, saponins, oxalates and glucosinolates triterpenoids. In alternative medicine, the leaves of the *M. oleifera* are used to treat several ailments including parasitic diseases, cuts, typhoid fever, arthritis, malaria, diseases of the skin, genital urinary ailments, hypertension, diabetes mellitus and many others. Previous animal studies show that *M. oleifera* leaves are safe at low doses, however, the aqueous extract at a lethal dose LD (16.1 g/kg) and ½ LD50 (8.05g/kg) have been reported to cause mild liver toxicity in Swiss Albino rats, when given once daily for 30 days. The observed hepatotoxicity was shown by an elevation of alkaline phosphatase (ALP), alanine aminotransferase (ALT), aspartate aminotransferase (AST) and serum bilirubin and was confirmed by significant histopathological changes in the liver tissue.

Antioxidants such as the endogenous tripeptide glutathione, N-acetyl cysteine (NAC), vitamin C, vitamin K and lipoic acid, are protective against drug-induced toxicity. All these can directly neutralize free radicals, but they cannot replenish the cysteine required for Glutathione synthesis. Therefore, it is not surprising that NAC, which supplies the cysteine are necessary for glutathione synthesis, has proven effective in treating conditions associated with reactive free radicals that cause organ damage such as the paracetamol-induced hepatotoxicity. Previous studies have reported that NAC can protect the liver from various drug-induced toxicities including anti-tuberculosis drug-induced hepatotoxicity, endosulfan-induced liver and kidney toxicity in rats and acrylamide-induced oxidative stress. However, there is limited information related to the protective effect of NAC on *M. oleifera* aqueous leaf extract-induced hepatotoxicity. The fact that people are taking *M. oleifera* leaves to treat many common ailments without a prescribed dose, it was found necessary to identify a drug that can be protective to the liver. This study established the protective effects of NAC on *M. oleifera* aqueous leaf extract-induced hepatotoxicity in Wistar Albino rats.

**MATERIALS AND METHODS**

**Study design and setting**

*M. oleifera* leaves were harvested during the dry season from Mukono district in central Uganda where the plant is grown by most families. The identity of the plant was confirmed by a plant taxonomist from Makerere University, Kampala, Uganda and a voucher specimen number 41302 was deposited at the Makerere University herbarium.

**Extracts preparation**

The collected leaves were air-dried at room temperature in the department of Physiology, Makerere University, College of Health Sciences until constant weight was attained. The dried material was pulverized to coarse powder using a motor and pestle, followed by soaking 300g of the coarse powder in 3.0L of hot water (70°C) to prevent being attacked by fungi while shaking it at 2 hours intervals for 12 hours. The resulting suspension was filtered using a Whatman No.1 filter paper in a Buchner funnel. The filtrate was freeze-dried at 32 Pausing a freeze dryer (Genesis 12 ES) in the department of Chemistry laboratory, Makerere University with an original temperature set at -47°C and maintained at 0°C to dry the extract. The dry extract obtained was stored in an air-tight bottle and wrapped in a silicon paper to prevent moisture and fungal attack. The stock solution of the extract was prepared by dissolving 100g of the dry aqueous extract in 100ml of distilled water to make a stock solution from which the daily doses were calculated according to the individual rat’s body weight.

**Selection of experimental laboratory animals**

The experiment was performed using male disease-free Wistar albino rats, aged 8 weeks, obtained from Makerere University, College of Veterinary Medicine, Animal resources and Biosecurity. The rats were housed in cages with 6 rats each. All the experimental animals were kept under standard laboratory conditions of temperature (25±1°C), relative humidity (45-55%) and light/dark cycle (12hr light: 12hr dark cycle). Standard rat food pellets and tap water were provided *ad libitum*. Rats were acclimatized in the experimental room for one week before beginning the experiment at the physiology laboratory, Makerere University, College of Health Sciences.

**Drugs**

N-acetyl cysteine (NAC) purchased from NOWFOODS, 244 Knollwood Dr, Bloomington, IL. 60108 USA was used as the antioxidant for the experiment. Paracetamol and normal saline were purchased from Friecca Pharmacy, plot 160 Wandegeya, 8472, Kampala, Uganda.

**Experimental procedure**

The Wistar albino rats (n=18) were divided into three groups. Group I, Group II and Group III. Each group comprised of six rats (n=6). After an overnight fast, Group I rats were administered orally using an intra-gastric tube with single dose of 8.05 g/kg body weight (bwt) *M. oleifera* leaf extract plus 1ml Normal saline. Group II rats received *M. oleifera* leaf extract at a dose of 8.05 g/kg bwt NAC. Group III rats received 750mg/kg bwt Paracetamol plus 50 mg/kg bwt NAC. The drugs were given once day for 28 days. On day 14, three rats from each group were sacrificed by carbon dioxide asphyxiation. At the end of intervention period (28 days), the three remaining rats from each group were sacrificed by carbon dioxide asphyxiation. Blood was collected by heart puncture, serum harvested, liver tissue was removed for histopathology analysis.

**Blood analysis**

The blood samples were collected in non-heparinized vacutainers, centrifuged at 500 gram force for 10 minutes, serum was harvested and analyzed using an automated clinical chemistry analyzer Roche diagnostics, Germany, using COBAS-e-411 Clinical chemistry analyzer at the Clinical chemistry laboratory, Mulago National Referral Hospital, Kampala, Uganda. The Alanine amino transferase (ALT), Aspartate amino transferase (AST), Alkaline Phosphatase (ALP), Bilirubin and Total protein levels were measured.

**Histopathology of the liver**


The harvested liver tissue was exposed to histopathology examination done at the pathology laboratory, Makerere University, College of Health Sciences. Tissues were fixed in 10% buffered formalin and then processed. tissue sections (4-5mm thickness) were prepared, stained with hematoxylin and eosin (H & E) and examined by light microscopy (Olympus CX21, Japan).

Statistical analysis

Statistical analysis was performed using Graph Pad prism version 8.0a Software (Graph Pad Software Inc., California, USA). Comparisons between groups were done by two-way analysis of variance (ANOVA) using Dunnett’s multiple comparison’s test. The level of significance was fixed at P-value=0.05.

Results

The study findings were; an increase in serum liver enzymes for the rats that were treated with a toxic dose of 8.05g/kg bwt M. oleifera extract plus 1ml of normal saline. However, the rats that received 8.05g/kg of M. oleifera extract plus 50mg/kg bwt of NAC and those that received 750mg/kg bwt of Paracetamol plus 50mg/kg of NAC showed normal serum liver enzymes, bilirubin and total protein levels at the end of the intervention period.

Serum ALT levels

The rats that were treated with M. oleifera plus NS had mean serum ALT 47.80 ± 7.00 U/L on day 0, 64.35 ± 9.85 U/L on day 14 and increased to 82.40 ± 21.80 U/L on day 28. However, the rats that were treated with M. oleifera extract plus NAC, had mean serum ALT 54.40 ± 9.90 U/L on day 0, 64.65 ± 0.65U/L on day 14 and reduced to 51.50 ± 4.30U/L (P-value=0.0694) on day 28 (Figure 1).

Serum AST levels

The rats that were treated with M. oleifera plus NS had mean serum AST 127.85 ± 3.65 U/L on day 0, 105.70 ± 6.50U/L on day 14 and reduced to 90.90 ± 16.50 U/L on day 28. However, the rats that were treated with Paracetamol plus NAC, had mean serum AST 128.70 ± 12.80 U/L on day 0, 134.20 ± 35.70 U/L on day 14 and increased to 130.05 ± 32.00 U/L on day 28 (Figure 2).

Serum ALP levels

The rats treated with M. oleifera plus NS had mean serum ALP 98.10 ± 1.40 U/L on day 0, 111.60 ± 22.70 U/L on day 14 and further increased to 130.05 ± 32.05 U/L on day 28. However, the rats that were treated with M. oleifera plus NAC, had mean serum ALP of 92.10 ± 4.10 U/L on day 0, 111.60 ± 22.70 U/L on day 14 and increased to 130.05 ± 32.05 U/L on day 28 (Figure 3).

Serum Bilirubin levels

The rats treated with M. oleifera plus NS had mean serum bilirubin 0.12 ± 0.00 µmol/L on day 0, increased to 0.05 ±
0.02 µmol/L on day 14 and decreased to 0.07 ± 0.01 µmol/L on day 28. However, the rats that were treated with *M. oleifera* plus NAC, had mean serum bilirubin 0.07 ± 0.05 µmol/L on day 0, increased to 0.20 ± 0.13 µmol/L on day 14 and reduced to 0.05 ± 0.03 µmol/L (P-value = 0.9089) on day 28. The rats treated with Paracetamol plus NAC, had mean serum bilirubin 0.05 ± 0.02 µmol/L on day 0, increased to 0.17 ± 0.02 µmol/L on day 14 and decreased to 0.08 ± 0.04 µmol/L (P-value = 0.9846) on day 28 (Figure 4).

**Figure 4**: Mean serum bilirubin

### Serum Total protein levels

The mean serum total protein was within normal levels in all treatment groups on day 0, 14 and 28 that is 67.90 ± 0.40 g/L to 69.35 ± 2.15 g/L (Figure 5).

**Figure 5**: Mean serum total protein

### Histopathology results

the liver tissue for all the study rats that received *M. oleifera* plus NAC and 8.05g/kg *M. oleifera* extract plus ns were analyzed. The liver tissue had vasocongestion, lymphocyte infiltration around the portal triad (Figure 6). The liver tissue for the rats that received *M. oleifera* plus NAC showed normal appearance of the hepatocytes with no lymphocyte infiltrations around the portal triad (Figure 7) and the normal liver tissue showed normal hepatocytes around the portal triad (Figure 8). The liver tissue from rats that received a toxic dose of paracetamol plus NAC showed normal liver tissue with vasocongestion (figure 9).

**Figure 6**: Histopathological appearance of the liver tissue that received *M. oleifera* plus NS; H&E-(X10): Infiltration of lymphocytes around the portal triad as shown by the arrow.

**Figure 7**: Histopathological appearance of the liver tissue that received *M. oleifera* plus NAC; H&E-(X10): Normal appearance with no lymphocyte infiltrations around the portal triad as shown by the arrow.

**Figure 8**: Histopathological appearance of a normal liver tissue H&E-(X10): Normal appearance of the hepatocytes around the portal triad as shown by the arrow.
DISCUSSION

The findings for the rats that received 8.05g/kg M. oleifera leaves extract plus 50mg/kg NAC, and 750mg/kg Paracetamol plus NAC showed normal mean serum ALT, AST, ALP and bilirubin levels by day 28 (Figures 1,2,3,4), highly suggesting that NAC prevented development of M. oleifera leaf extract-induced hepatotoxicity in Wistar Albino rats. These results were in agreement with those from previous studies which have reported that NAC can protect the liver from various drug-induced toxicities including against anti-tuberculosis drug-induced hepatotoxicity, endosulfan-induced liver and kidney toxicity in rats, acrylamide-induced oxidative stress and Cisplatin-induced toxicity.

In the current study, the elevation in mean of serum ALT, AST, and ALP levels in the rats that received M. oleifera plus NS was possibly due to the inflammation of the liver tissue caused by ingestion of a toxic dose of M. oleifera aqueous leaf extract (Figure 1-4). The previous studies have shown that administration of toxic doses (8.05g/kg) of M. oleifera leaf extract to Swiss Albino rats, induces reversible or irreversible toxicity changes in the liver, kidney and heart. The aqueous leaf extract contains a high concentration of alkaloids and the plant-derived alkaloids, by function and chemical nature, are toxic to mammals. This could have contributed to leakage of hepatic cell enzymes into plasma which is a sign of hepatic tissue damage. However, rats that received the synthetic antioxidant NAC along with either M. oleifera or paracetamol were protected from hepatotoxic effects of either agent. Therefore, antioxidants, such as NAC should be administered along with M. oleifera leaves to prevent the toxicity effects of M. oleifera leaf extract.

It is also possible that NAC prevented the altered oxidative stress parameters related to exposure of the liver to a toxic dose (8.05g/kg) of M. oleifera aqueous leaf extract by inducing antioxidant mechanisms since NAC has strong antioxidant properties. Antioxidants appear to act against disease processes by increasing the levels of endogenous antioxidant enzymes and decreasing toxic products such as lipid peroxidation byproducts. The antioxidant effect of NAC is believed to arise from NAC-induced increases in local nitric oxide concentrations and promotion of microcirculatory blood flow, thereby enhancing local oxygen delivery to peripheral tissues. The microvascular effects of NAC therapy are associated with a decrease in morbidity and mortality, even when NAC is administered in the setting of established hepatotoxicity.

The mild non-significant elevation of ALT, AST and ALP levels noted on day 14 in rats that were co-administered with both M. oleifera and NAC (figure 1-4), was probably due to the low dose of NAC (50mg/kg) used in the current study due to a longer period of administration (28 days). Compared to a higher dose of NAC (150mg/kg) employed in earlier studies where there was shorter duration of administration (3-5 days). The mild elevation in serum liver enzymes could also be because of evidence of intestinal metabolism of NAC in the rat, which would further reduce NAC bioavailability after oral administration. Therefore, these results proved that NAC probably protected the integrity of the liver cells and preserved it from leakage as ALT, AST, ALP, bilirubin and total protein levels were kept within normal ranges.

CONCLUSION

Findings have provided evidence that NAC administration with M. oleifera leaves extract effectively prevents the occurrence of M. oleifera leaves extract-induced hepatotoxicity.

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Disclosure of conflict of interest

None.

REFERENCES

1. Mbikay M. Therapeutic potential of Moringa oleifera leaves in chronic hyperglycemia and dyslipidemia: a review. Frontiers in pharmacology. 2012;3.
2. Reddy YRR, Lokanatha O, Ratnam K, Reddy CS, Raju IN, Reddy CD. Acute and sub acute toxicity of Moringa oleifera stem bark extract in Swiss albino mice. Int J Sci Biotech Pharm Res. 2013; 2(4):74-82.
3. Kasolo JN, Bimenya GS, Ojok L, Ochieng J, Ogwal-Okenj JW. Phytochemicals and uses of Moringa oleifera leaves in Ugandan rural communities. Journal of Medicinal Plants Research. 2010; 4(9):753-7.
4. Konny BB, Olounlade PA, Azando EBV, Hounzangbe-Adote SEG. A review on phytochemistry and pharmacology of Moringa oleifera leaves (Moringaceae). Journal of Pharmacognosy and Phytochemistry. 2016; 5(5):325.
5. Stoln SJ, Hartman MJ. Review of the safety and efficacy of Moringa oleifera. Phytotherapy Research. 2015; 29(6):796-804.
6. Kasolo JN, Bimenya GS, Ojok L, Ogwal-Okenj JW. Sub-acute toxicity evaluation of Moringa oleifera leaves aqueous and ethanol extracts in Swiss Albino rats. International Journal of Medicinal Plant Research. 2012; 1(6):075-81.
7. Akurri KR, Mantovani JJ, Herzenberg LA, Herzenberg LA. N-Acetylcycteine—a safe antidote for cysteine/glutathione deficiency. Current opinion in pharmacology. 2007; 7(4):355-9.
8. Baniasadi S, Eftekharpi P, Taharsi P, Fahimi F, Raoufy MR, Masjedi MR, et al. Protective effect of N-acetylcysteine on antituberculous drug-induced hepatotoxicity. European journal of gastroenterology & hepatology. 2010; 22(10):1235-8.
9. Beceren A, Sehirli AO, Omurtag ZG, Arbak S, Turan P, Sener G. Protective effect of N-Acetyl-L-Cysteine (NAC) on endosulfan-induced liver and kidney toxicity in rats. Int J Clin Exp Med. 2017;10(7):10031-9.
10. Alturfân Ei, Beceren A, SeHIRLI AO, Demiralp ZE, SENER G, OMURTAG ZG. Protective effect of N-acetyl-L-cysteine against acrylamide-induced oxidative stress in rats. Turkish Journal of Veterinary and Animal Sciences. 2012; 36(4):438-45.
11. Gordon K. The OECD guidelines and other corporate responsibility instruments. 2001.
12. Dickey DT, Muldoon LL, Doolittle ND, Peterson DR, Kraemer DF, Neuwelt EA. Effect of N-acetylcysteine route of administration on chemoprotection against cisplatin-induced toxicity in rat models. Cancer chemotherapy and pharmacology. 2008; 62(2):235-41.

13. Fakurazi S, Sharifudin SA, Arulselvan P. Moringa oleifera hydroethanolic extracts effectively alleviate acetaminophen-induced hepatotoxicity in experimental rats through their antioxidant nature. Molecules. 2012; 17(7):8334-50.

14. Saito C, Zwingmann C, Jaeschke H. Novel mechanisms of protection against acetaminophen hepatotoxicity in mice by glutathione and N-acetylcysteine. Hepatology. 2010; 51(1):246-54.