A major emphasis in *Rhododendron* research has been to describe and explain intersectional hybrid failure (Creech, 1955; Kho and Baer, 1970; Williams et al., 1982). The selfing behavior of *Rhododendron* is also poorly understood. An extensive examination of selfing in three species of *Rhododendron*, including studies of self and cross pollen tubes, showed no difference in growth, rate of pollen tubes between selves and outcrosses. Penetration of pollen tubes through the ovular micropyle region occurred 4 to 7 days after pollination, regardless of pollen source. Embryogenesis was studied in pistils collected from forced greenhouse plants of the same population. All ovules appeared to develop for a short period before senescing. Percent capsule set data from both years’ diallel pollinations indicated that some active form of self-recognition and rejection was operating and that environmental stresses and resource allocation were also influential. Additional information gathered included ovule counts, seed count to capsule size correlations, and germination trials. These pointed to a reduction in reproductive success at each developmental stage. Self-incompatibility (SI), defined as inability to set seed following self-pollination, is clearly not applicable here. There are inherent difficulties in separating an active, late-acting self-recognition/rejection system from inbreeding depression, which is a passive accumulation of homozygous recessive lethal and sublethal genes.

**Materials and Methods**

*Rhododendron prinophyllum* accession no. 790912 is a full sibling population located at the Univ. of Minnesota Landscape Arboretum in the rhododendron and azalea trial plots. The two dozen plants comprising the population are F1 seedlings arising from a cross between accessions 790577 and 790580. While the specific origin of the parents is not recorded, it is known that neither parent would set self seed. They are not considered to be related.

**Year 1–1986**

*Field pollinations.* Nine plants from a population of *R. prinophyllum* were selected in May 1986. When the population had reached ≈50% anthesis, nine flower clusters from each plant were emasculated; we also removed already opened flowers as well as buds still physiologically immature. The clusters were not bagged, as prior work had shown that flowers with their corollas removed, accomplished during emasculation, are not visited by insects and remain unpollinated. Two days later, each of the plants was selfed and crossed with fresh pollen from every other individual. Only one pollen type was used on each flower cluster. Number of flowers pollinated per cluster ranged from as few as six to as many as 23. In all, 1013 pollinations were made.

*Pollen tube analysis.* Two days after pollination, one to three pistils were collected from each cross and fixed in FAA. The fixed material was then stored in 70% ethanol. The pistils were prepared and observed for pollen tube growth rates and stage of ovule development in both cross- and self-pollinated material with a Zeiss epifluorescence microscope (Lab. 16 model with epifluorescence condenser IUFl, Carl Zeiss, Oberkochen, Germany) and the filter combination: KP450, KP490, Ft 510, LP 520, as described by Kho and Baer (1968). Photographs were obtained with the aid of a Nikon At35 35-mm camera and Kodak Tungsten 160 film.

*Capsule collection.* Beginning in early October, capsules were collected as they matured, and the number of capsules was recorded for each cross. Three crosses were lost during the season and data on capsules were unavailable. Compatibility was analyzed from the 1986 crosses with the aid of SIGMAS (Self- Incompatibility Genetic Modeling Application Systems), a computer software developed by Liedl and Anderson (1987).

**Year 2–1987**

*Field/indoor pollinations.* Six plants chosen on the basis of 1986 results from the original nine plants were again labelled

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Abbreviations: SI, self-incompatibility.
and emasculated. The six plants were selected to include three plants that had set a range of self capsules and three plants that set no self capsules the previous summer. Two clusters were emasculated and pollinated for each cross for a total of 752 crosses.

To microscopically observe pollen tube growth rates following selfing and crossing, branches from a plant with no self capsules, no. 9, (Fig. 3) and from the highest self-capsule setting plant, no. 8, (Fig. 3) were cut and brought indoors for forcing where temperatures would be more uniform. As the flowers began to open, they were emasculated, and the plants were then selfed and crossed to one another. All pistils from these pollinations were collected at intervals of 6, 12, 18, 24, and 30 h after pollination and fixed in FAA for later microscopic analysis.

Capsule collection and seed counting. Capsules were again collected in fall as they matured. Total capsules per cross was recorded. The length and width of each capsule were measured using a Manostat caliper. The resulting measures were recorded and used to estimate the volume of each capsule using the formula: 

\[ \text{Volume} = \pi \times \text{length} \times \text{width} \times \frac{1}{2} \]

The number of seeds per capsule was counted to estimate reproductive success. All five locules were split away from the capsule using a no. 11 surgical blade (Bard-Parker, Becton Dickinson Co., Rutherford, N.J.), and seeds were removed through the placental groove with a needle probe. Contents of each locule were counted with the aid of an OptiVisor binocular/magnifier (Donegon Optical, Kansas City, Me.), lens no. 7 (magnification ×2.5). Several samples of open-pollinated capsules from plants in the study population and capsules collected from mature plants of other *R. prinophyllum* populations were also counted and similarly measured.

Seed germination. Two 50-seed samples from each cross were tested for germination capacity. If <100 seeds were available, they were divided equally for the two treatments. Milled sphagnum moss was moistened with distilled water and ≈25 ml placed into 100 x 15-mm petri dishes and teased to create a flat, uniform surface filling the plate. Seeds were soaked overnight in 1000 ppm (treatment 1) or 500 ppm (treatment 2) gibberellic acid (GA), drained, rinsed thoroughly, and placed on the medium. Samples were placed in germination chambers under lights at 24°C. Germinating seeds were observed with the OptiVisor, and counts were taken at 2-day intervals after germination began. The appearance of a protruding radicle was used as the start of germination. A final count was averaged between the two GA treatments to yield total percent germination.

Pollen tube analysis. Microscope examination of pollen tube growth following the 1987 crosses was made using the technique described by Williams et al. (1982) with the following modification: Under a Wild stereomicroscope, the fixed pistils were placed on a slide in ethanol, and the style and stigma were removed and returned to the collection vial. Retaining the pedicel as a handle, the ovary walls were carefully cut away from the ovules by first slicing between each locule with a no. 11 surgical blade, cutting across the base. The blade was then used to remove pieces of the ovular wall. The ovules were stored in 70% ethanol. Pollen tubes were examined following the same procedures as in 1986.

Estimating number of ovules. During dissecting away the ovary walls, a single intact locule was removed and transferred to a clean slide with potassium iodine stain, which darkened the starch and allowed easier examination of the ovules. The remaining four locules were returned to the respective collection vials along with the style. Stain was added to the dissecting slide, and any ovules lost in the dissecting process were counted. The remaining locule was halved along the placental groove, and ovules of each half were counted. The total was recorded and multiplied by five, to estimate the total number of ovules that could potentially develop into seed from each stigma pollinated.

Fertilized ovules. The remaining four locules, when cleared as described for pollen tubes, were gently teased away from the receptacle and pedicel, placed onto a clean slide with fresh aniline blue stain, and squashed under a coverslip. Ovule condition, whether fertilized or unfertilized, and development or degeneration were noted.

For additional study of ovule development following pollination, two plants from the same population were dug in Nov. 1987 before the ground had frozen. They were potted and stored in a below-ground storage facility to fulfill cold requirements, then forced into flower in a greenhouse. Upon reaching 50% anthesis, the two plants (designated A and C) were reciprocally crossed and selfed. Several branches were also emasculated but not pollinated on each plant to provide a control for assessing changes during development of the ovules. Pistils were harvested at 1, 2, 4, 7, 14, 21, and 28 days after pollination. The collections were treated similarly to the field material. Pollen tube growth and ovule development were observed and recorded.

Results and Discussion

Pollen tube and ovule observations. Germination of pollen tubes and penetration of the stigma surface had occurred between 1 and 2 days after pollination regardless of pollen source. No difference in growth rate between self and outcross pollen tubes was perceived.

The fixed pistils from the remaining 1987 field pollinations were first checked to be certain that pollen tetrads had germinated and the resulting pollen tubes had grown through the style. Microscopic evidence suggested that pollen tubes from self- and cross-pollinations reached the base of the style by the 3rd day. (Fig. 1 A, B, and C). Some ovules had been penetrated by pollen tubes by this time. These were readily distinguished from ovules lacking pollen tubes in that they had a linear fluorescent pollen tube segment running from the outer ovule surface, through the micropyle, and up to the embryo sac. In many instances, it was even possible to see the pollen tubes extending beyond the entrance to the ovule micropyle, making judgment of ovular condition even simpler (Fig. 1D).

Collections 1 and 2 days after pollination of greenhouse plants A and C gave little new information. Pollen tetrads had germinated, and the tubes had penetrated the stigma surface and begun growth down the style. Ovules were unpenetrated.

At day 4, the greenhouse material was similar to the 1987 field collections. By day 7, the squashes showed nearly all ovules penetrated by pollen tubes from both self- and outcross pollination. Ovules from the unpollinated stigmas were unpenetrated and had begun to degenerate.

At day 14, ovules developing further had greatly enlarged (Fig. 2), while the nondeveloping ovules showed no size change; but there was an increasing fluorescence from callose accumulation, which is symptomatic of general degeneration (Kho and Baer, 1970; Williams et al., 1984a). The enlarged ovules were balloon-like, and it was difficult to discern the developing embryo. However, a globular embryo was found in several instances above the degenerating synergid at the end of the micropyle.

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Fig. 1. Comparison of the condition of fixed ovules collected from the field 3 days after pollination in 1987 from *R. prinophyllum*, Minnesota Landscape Arboretum accession no. 790912. (A) Ovules from the cross 8 × 9. Note pollen tube extending from the micropyle entrance to the region of the embryo proper in the ovule on the right (arrow). The left ovule appears unpenetrated. Capsule set: 88%. (B) Ovules of female 8, selfed. Pollen tube is obvious in the right-hand ovule (arrow). Capsule set: 22%. (C) Ovule of female 9, selfed, with entering pollen tube (arrow). Capsule set: 0. (D) Ovule from the cross 9 × 6. Pollen tube visible externally (arrow) as it enters the ovule. Capsule set: 79%.

(Fig. 2b). An area of brightly fluorescing callose between the synergid and the embryo fit the description of a callose special wall (Williams et al., 1984a), which is thought to function as a separation of the newly formed sporophyte from maternal tissue. Observations of styles from days 21 and 28, did not add significant information.

**Ovule counts.** Reproductive success in angiosperms can be expressed by percentages of viable seed produced (Aalders and Hall, 1961; Cope, 1958; Flaschenriem and Ascher, 1979; Lundqvist, 1954; Takahashi, 1973). The reproductive success or failure through seed set in *R. prinophyllum* requires an assessment of initial reproductive potential. Counts of ovules revealed that there were from 275 to 419 per ovary in *R. prinophyllum*. Consequently, the number of potential seeds per female differed significantly (*P* < 0.01). This factor must then be taken into consideration when evaluating success in seed production.

**Capsule set.** Self-pollination the first year resulted in a wide range of compatibility, with five clearly self-incompatible individuals (Fig. 3). There were also four plants that were partially self-compatible. This selfing behavior is in contrast to that reported by Whiting (1979) when seven *R prinophyllum* (*R. roseum*) plants (from a different seed source than used in this research) produced no seed from self-pollination. This variability is not unexpected, however, when viewed in light of past literature regarding self- and interspecific crossings in *Rhododendron* (Bowers, 1960; Kehr, 1966; Kho and Baer, 1970, 1973; Widrlechner et al., 1982).

Data showing that selfing in five of the nine individuals produced no capsules would suggest that a self-recognition system exists. Lack of capsule set seen in four of the outcross pollinations might also indicate the degree of relatedness among the full-sibling population. Crossing tags were lost on the pollinated flower clusters for three of the crosses, and these appear as × in Fig. 3 (crosses 5 × 7, 7 × 5, and 9 × 8).

If a SI system were functioning in *R. prinophyllum*, then the
four plants setting self-capsules could be explained as exhibiting pseudo-self-compatibility, which is the incidence of a range of seed set following self-pollination in plants known to have SI (Darwin, 1876). The 1986 data for capsule set also show large differences in the percentage of capsules set between reciprocal crosses (e.g., crosses 7 × 8 and 8 × 7). Reciprocal differences such as these point to sporophytic as opposed to gametophytic control of self-fertilization.

Despite many pollinations having been made per cross in 1987, the overall number of capsules set was generally less than in 1986 (Fig. 3). Only female no. 8 set any self capsules. Also, the self set for female no. 8 was less than one-fourth (=22%) than the capsule set the previous season (=93%). However, the percentages of outcross set in 1987 were also much lower.

The overall reduction in capsule set in 1987 was not completely unexpected. Winter 1986-87 had virtually no snow cover, which is unusual for Minnesota. The open winter was preceded by a dry fall and followed by a dry spring and early summer. A hot, dry weather pattern characterized the breeding season. During the 1987 pollination period, temperatures reached 32°C.

A second reason for reduced capsule production for the 1987 season is addressed by Wiens (1984) in seed : ovule ratios (S : O). While annual species mature on average =85% of their ovules as seed, woody plants are reported as having a S : O ratio of only =33%.

Stephenson (1981) reviews literature pertaining to trends in reproduction of various species on a flower to fruit basis. He concluded that, within a species, the ratio of flowers : fruit can
vary among populations, among individuals in a population, and from year-to-year in perennial individuals. Seemingly a logical point, this still is important when data on reproductive success are analyzed. Overall fruit : flower ratios for the *R. prinophyllum* population were 51.5% in 1986 and 31.4% in 1987. Considering the harshness of weather conditions and related source/sink problems, it may well be that 31% is not really a poor performance.

**Seed counts.** Seed count data for 1987 were based on 232 capsules (Table 1). Percent seed per capsule is derived as the mean number of developed seeds per capsule divided by the standard number of ovules per capsule from that female. Percent seed per flower pollinated is similarly derived by using the mean number of developed seeds per pollinated flower, rather than per capsule, obtained; this percentage, therefore, more accurately depicts the overall level of reproductive success.

In a compatible cross, if the percentage of flowers that abort or fail to develop represents “normal” abortion of excess flowers, then percent seed per capsule should more accurately reflect the physiological interactions. If woody plants have a fixed S : O ratio of ≈33% as suggested by Wiens (1984), an interpretation of seed set failure due to any other causes, such as prezygotic SI or postzygotic abortion, should take into account this possible background rate of abortion.

**Seed germination.** Standard germination of azalea seed at the Univ. of Minnesota Landscape Arboretum involves surface-sowing the cleaned seed on commercially available acid peat and keeping them warm (20°C), moist, and in diffuse light until germination occurs. Germination procedures for *Rhododendron* by other researchers are similar. An acid-based medium of sphagnum peat or oak leaf mold and sharp sand is used (Cox, 1985; Hartmann and Kester, 1975; Kains and McQueston, 1942).

Trials using these methods were all done on seeds from the cross 8 × 6, as there was an abundance of seed available. None germinated.

While we found no literature stating that mature dried *Rhododendron* seed requires a moist-chilling treatment to overcome a secondary dormancy, there had to be some reason seed was not germinating. Arisumi et al. (1988) reported improved germinability in immature *R. simiarum* seed by a cold treatment to either capsules, seed, or synergistically, to both. It seems logical to suspect that some internal dormancy, imposed to prevent matured seed from germinating prematurely in adverse conditions, would be operating in *R. prinophyllum*.

Application of GA is often the next best means of overcoming dormancy (Bewley and Black, 1982; Ellis et al., 1983). Thus, two more 50-seed samples were taken from cross 8 × 6. One was treated with 500 ppm GA and the other with 1000 ppm, since we found no suitable rates in the literature. Germination occurred in both dishes =10 days after sowing, and a final count made 3 days later yielded an average of 17% for the two germination treatments. Compared to previous results, this one seemed to support the use of the growth regulator; an average of the two treatments compensates for the inability to replicate the trials for seed from each cross at several GA levels.

Difficulty in assessing quality of azalea seed is increased because of the inclusion of seed with low vigor in the seed lots. Seeds of poor quality will be represented when randomly sampling lots for germination trials. The averaged results of the two germination treatments are listed as percent germination in Table 1. Summary of female reproductive success in 1987 by percent capsule set, percent seed set per capsule, percent seed set per flower pollinated, and percent germination in *Rhododendron prinophyllum*, Minnesota Landscape Arboretum accession no. 790912.

| Capsule set (%) | Seed/flower pollinated (%) | Germination (%) | Capsule set (%) | Seed/flower pollinated (%) | Germination (%) |
|----------------|---------------------------|----------------|----------------|---------------------------|----------------|
| Female 4       |                           |                | Female 7       |                           |                |
| x4 0           | 0                         | 0              | x4 19.4        | 24.29                     | 4.70           | 12.0           |
| x5 22.2        | 15.22                     | 3.38           | x5 38.9        | 23.26                     | 7.94           | 12.0           |
| x6 40.0        | 46.98                     | 18.80          | x6 40.1        | 26.53                     | 11.71          | 16.0           |
| x7 31.6        | 18.03                     | 5.69           | x7 0           | 0                         | 0              | 0              |
| x8 3.0         | 19.37                     | 5.88           | x8 5.9         | 8.66                      | 5.10           | 0              |
| x9 52.4        | 16.35                     | 8.57           | x9 25.0        | 17.44                     | 4.36           | 6.0            |
| Avg 24.9       | 19.04                     | 6.08           | Avg 21.9       | 17.11                     | 4.97           | 11.0           |
| Female 5       |                           |                | Female 8       |                           |                |
| x4 0           | 0                         | 0              | x4 80.0        | 22.67                     | 16.88          | 1.0            |
| x5 0           | 0                         | 0              | x5 33.3        | 13.92                     | 4.16           | 0              |
| x6 38.5        | 32.50                     | 11.54          | x6 90.0        | 73.13                     | 6.58           | 1.0            |
| x7 9.1         | 7.66                      | 6.96           | x7 86.7        | 36.09                     | 29.14          | 0              |
| x8 45.5        | 21.35                     | 9.71           | x8 21.7        | 96.84                     | 2.11           | 6.0            |
| x9 5.9         | 6.92                      | 4.08           | x9 88.0        | 38.75                     | 3.41           | 1.0            |
| Avg 16.5       | 11.07                     | 3.57           | Avg 66.6       | 31.88                     | 24.92          | 2.0            |
| Female 6       |                           |                | Female 9       |                           |                |
| x4 25.8        | 26.20                     | 6.76           | x4 56.5        | 14.39                     | 8.13           | 21.0           |
| x5 25.8        | 39.41                     | 10.17          | x5 57.9        | 14.51                     | 8.41           | 11.0           |
| x6 0           | 0                         | 0              | x6 78.9        | 25.18                     | 18.56          | 14.0           |
| x7 25.0        | 38.31                     | 9.57           | x7 21.7        | 78.14                     | 16.98          | 9.0            |
| x8 21.6        | 67.32                     | 14.18          | x8 57.1        | 17.88                     | 8.94           | 1.0            |
| x9 8.6         | 58.97                     | 5.05           | x9 0           | 0                         | 0              | 0              |
| Avg 17.7       | 38.87                     | 7.70           | Avg 45.4       | 13.31                     | 7.51           | 11.0           |
just 2.6% of initial ovules produced will reach the seedling stage. Germination in general was poor. Seed from cross 8 × 6 treatments were beneficial. Poor germination may reflect the inherent problems associated with the relatedness of a full-sibbing population.

Levels of reproductive success. Reproductive success was greatly reduced at each successive criterion measured (Table 1). The best germination from the cross 6 × 4 resulted in only 38% germination and this is, of course, no indication of the ultimate survival of those seedlings to reproductive maturity. With a mean of 374 ovules per capsule in female 6, this suggests that just 2.6% of initial ovules produced will reach the seedling stage. Although factors of cultural conditions for these plants certainly play a role in their ability to produce seed, this is still a very poor reproductive output if viewed by traditional measures.

Theoretical considerations. SI has been assumed to be the cause of reproductive failure in other attempts at selfing in various Rhododendron spp. (Cox, 1985; Yamaguchi, 1980). However, evidence from microscopic examination of pollen–pistil interaction in the present research does not support the operation of a genetic prefertilization barrier to selfing. Therefore, by definition, genetic SI cannot be operating in R. prinophyllum. However, this does not suggest that there is no self-recognition event. A discrimination between self-matings and sib matings exists that preferentially allows a degree of sibbing over levels of selfing. It is also apparent that the level of reproduction attained for a given self or cross for each female (or male) and for the population as a whole is greatly influenced by the environment and resources available in a particular season. The combination of these factors, along with evidence from ovule development following self- and cross-pollinations, points rather conclusively to what in the literature has been termed late-acting SI or postzygotic abortion; both of which are thought to be polygenic in nature (Cope, 1962 Crowe, 1971; Seavey and Bawa, 1986; Williams et al., 1984a, 1984b).

The nature of this late-acting system in R. prinophyllum is difficult to define because the system operates beyond formation of a zygote. It is difficult to pinpoint the precise moment, either microscopically or macroscopically, of any active controlled breakdown or to distinguish it from inbreeding depression caused by the passive buildup of recessive deleterious or lethal genes. Seavy and Bawa (1986) suggested the late-acting system to occur under temporal failure from selfing, genetic segregation expressed before germination, and success of embryo rescue. Failure of postzygotic development at a range of stages and failure of embryo rescue suggest inherent genetic failure. Data from capsule set might be interpreted to indicate that there is a reproductive barrier to selfing in R. prinophyllum. The fact that a continuum of failure exists in the crosses and selfs suggests that inbreeding also exerts an influence.

Any genetic system, pre- or postzygotic, that results in a certain level of heterozygosity, will function to preserve genetic diversity and to ensure ultimate survival of a population. Survival during periods of isolation requires some level of tolerance to inbreeding. With Rhododendron, the large number of potential seeds, preferentially combined with postzygotic abortion, might function this way.

Classic SI is based on recognition. The ultimate gene products that can inhibit incompatible pollen tubes or, perhaps less likely, promote compatible pollen tubes function to select paternal genotypes. Sexual selection, where competition and genetic differences result in different levels of reproductive success among viable individuals of the same sex (Darwin, 1876; Marshall and Ellstrand, 1986; Willson, 1979), is considered a primary evolutionary force. It is invoked when differentiating between success rates of various male parents in mixed pollinations (Hill and Lord, 1986). In a late-acting system, where incoming pollen tubes are not discriminated against, mate selection must occur at the level of the zygote or embryo via selective abortion. This pattern has invariably been viewed as wasteful due to sterilization of ovules in which there has already been a considerable maternal investment (Lewis, 1979) and has thus been considered anomalous or maladaptive. However, the number of species in which late-acting systems have been described is increasing despite the difficulties in proving their operation experimentally (Bookman, 1984; Brandham and Owens, 1978; Cope, 1958, 1962; Crowe, 1971; Miri and Bubar, 1965; Spiss and Paolillo, 1969).

If late-acting phenomena are considered maladaptive, then what are the implications for evolutionary survival of Rhododendron? First, this method of sexual selection may not be as inefficient as originally thought (Seavey and Bawa, 1986; Stephenson, 1981). The production of excess flowers to attract pollinators, increase male function, and optimize mate selection would be immediately wasteful if the amount of maternal investment were excessive. Instead, a selective investment might function to reduce the commitment of resources to offspring that are less likely to succeed (Westoby and Rice, 1982).

The Ericaceae is very diverse, and plants in this family tend to occupy nutrient-poor acidic environments. Resource availability affecting selection could result in cyclical seasons of “boom” and “bust” reproduction and might indeed argue for a selective advantage in retention of zygotes until a balance in investment and resources could be achieved. Evolution to adapt to periodic extremes of available resources might well result in a deferment of mate selection through polygenic late-acting systems.

Another way of looking at late-acting systems, evolutionarily, is as a stepping stone from inbreeding depression, which is completely wasteful in that it produces progeny destined to fail, to the more advanced SI systems. As such, Rhododendron, and perhaps the Erica family, might be viewed as representing an increment in the evolutionary ladder of reproductive mode. So, while somewhat wasteful, the late-acting systems still succeed on a certain level. Or, it might be that these plants are relics that, while maladaptive, still survive well enough within their own environment. In this event, it may be that any selective advantage would then depend heavily on the high reproductive potential of Rhododendron spp. that flower profusely for many years and produce hundreds of seeds per capsule (Williams et al., 1984b).

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