Tacrolimus dose adjustment is not necessary in dose to dose conversion from a twice daily to a prolonged release once daily dose form

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Twice daily TAC (BID TAC) and prolonged released once daily dose tacrolimus (OD TAC) have different pharmacokinetic (PK) profiles in kidney transplant (KT) recipients. Precise dose adjustment recommendations when converting from BID TAC to OD TAC remain inconclusive. A single center, PK study was conducted in stable KT recipients taking constant doses of TAC, mycophenolic acid, and prednisolone. The area under the concentration–time curve (AUC) 0–24 and Ctrough were measured before and 4 weeks after 1:1 conversion from BID TAC to OD TAC without subsequent dose adjustment. A 90% confidence interval (CI) of geometric mean ratio (GMR) of OD TAC/BID TAC within the range of 0.9–1.11 was utilized to indicate equivalence of the narrow therapeutic index drugs. The roles of CYP3A5 genotypic polymorphism on PK parameters were also assessed. There were 20 patients with median time since transplantation of 18 months. The mean of CKD-EPI eGFR was 60.7 ± 16.43 mL/min/1.73 m². The median total daily TAC dose of 0.058 mg/kg/day. The geometric means (%CV) of AUC 0–24 of OD and BID TAC were 205.16 (36.4%) and 210.3 (32.5%) ng/mL × h, respectively, with a GMR of 0.98 (90%CI 0.91–1.04). The geometric means (%CV) of Ctrough of OD TAC and BID TAC were 5.43 (33.1%) and 6.09 (34.6%) ng/mL, respectively. The GMR of Ctrough was 0.89 (90%CI 0.82–0.98), which was below 0.9. The newly calculated target Ctrough level of OD TAC was 4.8–6.2 ng/mL. The best abbreviated AUC0–24 was AUC = 0.97(C0) + 5.79(C6) + 18.97(C12) − 4.26. The GMR AUC 0–24 was within the range of 0.9–1.11 irrespective of CYP3A5 genotypic polymorphism while the GMR of Ctrough was below 0.9 only in the CYP3A5 expressor patients. The 1:1 conversion from BID TAC to OD TAC without subsequent dose adjustment provided similar AUC0–24 regardless of CYP3A5 genotypic polymorphism. However, the Ctrough was lower in the CYP3A5 expressor group. Therefore, it is not necessary to routinely increase the OD TAC dose after conversion.

Trial registration: Thai Clinical Trials Registry (TCTR20210715002).

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Tacrolimus (TAC) is one of the main immunosuppressive drugs used to prevent allograft rejection after kidney transplantation (KT). There are two available oral forms: (1) Prograf®, a TAC that is administered twice daily (BID TAC), and (2) Advagraf®, a newer prolonged released once daily (OD TAC). Even though the formulation and pharmacokinetics (PK) of both drugs are different, yet they both have comparable efficacy in preventing rejection and have similar adverse event rates1,2,3. OD TAC is more convenient to administer and improves patient compliance4. However, in clinical practice, PK monitoring of the TAC levels is mandatory because TAC has a narrow therapeutic index5. Aside from that, the PK results have shown that there are high inter-patient variabilities5. Individual TAC PK can be affected by several factors, including CYP3A5 genotypic polymorphism6,7.

Measuring area under the concentration–time curve (AUC) is the gold standard for monitoring TAC exposure and accurately represents the total daily exposure for each patient while the trough level concentration (C\text{trough}) is more practical and preferred in clinical practice even though it only provides TAC exposure just before the morning dose. Scientifically, to maintain the same level of TAC exposure between BID TAC and OD TAC, both drugs should have an equivalent level of AUC\text{0-24} rather than C\text{trough}. Despite this crucial pharmacokinetic knowledge, several earlier PK studies pertaining 1:1 conversion from BID TAC to OD TAC monitored C\text{trough} instead of doing a full AUC\text{0-24} and showed that the C\text{trough} level in OD TAC was lower than BID TAC8–13. Such finding suggests that the C\text{trough} level in the maintenance phase of KT recipients treated with OD TAC should be set at the same level as recommendation for BID TAC, which should be within the range of 5 to 7 ng/mL1,4–16. Therefore, the total daily dose is generally increased by 10–15% to achieve the same C\text{trough} as that of BID TAC when BID TAC is switched to OD TAC16–17,21. With this strategy of the current practice, dose adjustment based on the C\text{trough} level may lead to unnecessary incrementation of OD TAC dose, and unexpectedly high TAC exposure as a consequence4.

As a matter of fact, several previous prospective PK studies have compared the AUC\text{0-24} as well as C\text{trough} of BID TAC and after 1:1 conversion to OD TAC in adult KT recipients17,22–25. However, there were some considerations regarding these previous reports. Most of these works were supported by pharmaceutical companies. Moreover, OD TAC dose adjustment after conversion was allowed in many studies, which resulted in an increase in mean TAC dosage at the end of the study. The values of equivalence ratio used in some studies were between 0.8 and 1.25 while the most appropriate values utilized in monitoring the drug with a narrow therapeutic index such as TAC should be 0.9 and 1.11. In addition, the findings from these studies are controversial. Therefore, there is a need to assess if 1:1 conversion from BID TAC to OD TAC without subsequent dose adjustment could effectively yield comparable AUC\text{0-24} levels or not.

For this intensive PK study, we applied the paradigm of bioequivalence testing to the narrow therapeutic index drugs using all of the PK parameters to compared the AUC\text{0-24} and other PK parameters, before and after switching from BID TAC to OD TAC using a 1:1 dose conversion without subsequent dose adjustment in stable KT recipients. The newly calculated value of C\text{trough} for OD TAC was identified. By using TAC concentrations at multiple time points instead of a single time point concentration to improve the predictive power of the C\text{trough}, to estimate the AUC\text{0-24}, we aimed to propose abbreviated AUC\text{0-24} equations that would accurately predict AUC\text{0-24} in our study population. The roles of CYP3A5 genotypic polymorphism on PK parameters following the 1:1 conversion without subsequent dose adjustment were also evaluated.

Methods

Study design and patients. A single center, open-labeled PK study was conducted at the King Chulalongkorn Memorial Hospital. The patients were consecutively enrolled from our kidney transplant clinic. The inclusion criteria were: (1) KT recipients aged ≥ 18 years, (2) on BID TAC (Prograf, Astellas, Tokyo, Japan) with mycophenolate mofetil (MMF; Cellcept, Roche, Basel, Switzerland) or enteric coated-mycophenolate sodium (EC-MPS; Myfortic, Novartis, Basel, Switzerland) and prednisolone, (3) had low to moderate risk for acute rejection1, (4) have stable kidney function (baseline serum creatinine < 3.0 mg/dL), and (5) had KT ≥ 6 months. Patients with a history of rejection or active infections were excluded from the study.

Sample size. The sample size was estimated based on one of the bioequivalence criteria for drugs with a narrow therapeutic index, with a 90% confidence interval (90% CI) for the geometric mean ratio (GMR) of OD TAC/BID TAC falling within the range of 0.9–1.1126. Assuming log-normally distributed data with GMR of 1 in paired measurements, a correlation between the BID and OD AUC\text{0-24} of 0.45 and a pooled coefficient of variation of 15%, 19 participants would provide 80% power for the equivalence test using the two one-sided test approach, with a significance level of 0.0526,67. We increased the sample size by 5% to account for potential loss of the participants during the follow-up period. Sample size calculations were performed using SAS 9.4 (Cary, NC, USA).
**Tacrolimus measurement.** KT recipients on stable doses of BID TAC and had C\textsubscript{trough} between 5 to 7 ng/mL\textsuperscript{10,11} were admitted to the Chulalongkorn Clinical Research Center (CRC) for a 24-h PK study. Serial whole blood samples were collected immediately before administration (pre-dose), and at 0.5, 1, 2, 3, 4, 6, 8, 12, 12.5, 13,14, 15, 16, 18, 20, and 24 h after dose administration for patients taking BID TAC\textsuperscript{10}. The patients were then switched from BID TAC to OD TAC at a ratio of 1:1 mg for 4 weeks to achieve a steady state without any subsequent dosage adjustment. Blood samples for OD TAC were obtained at pre-dose, 1, 2, 3, 4, 6, 9, 12, 15, and 24 h after dose administration\textsuperscript{10}. The BID TAC was administered at 7:00 and 19.00 while The OD TAC dose was at 7:00. Other medications apart from TAC, including known CYP450 interaction medications were maintained at the same dose throughout the study. All patients were given a standard calorie-controlled meal that was served at the same time during the intensive PK days to minimized the effects of food on the TAC absorption\textsuperscript{11} All three meals were scheduled at 8.00, 12.00, and 20.00.

TAC whole blood concentrations were measured by a chemiluminescent microparticle immunoassay (ARCHITECT\textsuperscript{12} tacrolimus assay, ABBOTT Park, IL, USA) using 2 mL of whole blood from EDTA tubes. Each blood sample was stored at 4 to 6 °C until the assay was performed on the following day. A linear trapezoidal method was used to calculate the AUC\textsubscript{0-24}.

**Outcomes.** The primary outcome was the AUC\textsubscript{0-24} of both formulations after the 1:1 conversion. Secondary outcomes were other PK parameters, abbreviated AUC equations of OD TAC, the incidence of adverse reactions, and allograft function by estimated glomerular filtration rate (eGFR by CKD-EPI) at 1 and 3 months after conversion.

**Statistical analysis.** The following PK parameters were determined utilizing non-compartmental methods: AUC\textsubscript{0-24}, C\textsubscript{trough}, C\textsubscript{max}, and time to maximum concentration. The data were analyzed by SPSS statistics version 18.0 (SPSS Inc., Chicago, Illinois, USA) and Stata 16.1 (StataCorp, College Station, TX). Descriptive statistics were used to summarize the participant characteristics at the first intensive PK assessment (baseline). The data that have been Ln-transformed such as AUC\textsubscript{0-24}, C\textsubscript{max} and C\textsubscript{trough} are reported as geometric mean (% coefficient of variation [%CV]), and time to C\textsubscript{max} as median (IQR). Generalized estimating equations were utilized to calculate the GMR value of AUC\textsubscript{0-24}, C\textsubscript{trough}, and C\textsubscript{max} in the OD TAC arm against the BID TAC arm as a reference with 90% CI. P-values were calculated based on 95% CI. Comparisons between the time to C\textsubscript{max} were performed using a Wilcoxon sign rank test. For both BID TAC and OD TAC forms, linear regression models were utilized to assess the proportion of the variance in AUC\textsubscript{0-24} explained by plasma concentrations at single time point, or combinations of time points using the R\textsuperscript{2} or adjusted R\textsuperscript{2} as appropriate.

**Ethics approval.** The study was registered in the Thai Clinical Trials Registry (TCTR20210715002). All procedures in this study were approved by the Institutional Review Board of the Research Ethics Review Committee for Research Involving Human Research Participants, Health Sciences Group, Faculty of Medicine, Chulalongkorn University (Institutional Review Board number 538/62), in compliance with the ethical principles described in the Declaration of Helsinki and its later amendments. All participants provided written informed consent before enrollment into the study.

**Results** Twenty patients [mean (± SD) age was 46 (± 12.1) years; 60% were males] completed the study. The mean body mass index (BMI) was 22.8 (± 3.95) kg/m\textsuperscript{2}. Median time since transplantation was 18.5 (IQR = 11.6–36.6) months. Baseline serum creatinine was 1.34 (± 0.32) mg/dL. Median total daily TAC dose was 0.058 (IQR = 0.038–0.096) mg/kg/day (Table 1). Fourteen participants (70%) were on statin, and 9 (45%) were on diltiazem, a calcium channel blocker. All patients received constant doses of sulfamethoxazole trimethoprim and acyclovir.

The concentration–time curves of OD TAC and BID TAC are shown in Fig. 1. The geometric mean (%CV) AUC\textsubscript{0-24} of OD TAC and BID TAC were 205.16 (36.4%) and 210.3 (32.5%) ng/mL x h, respectively (Table 2). The GMR (90%CI) of the AUC\textsubscript{0-24} for OD TAC versus BID TAC was 0.98 (90%CI 0.91–1.04), which fell within the range of equivalence ratio. The geometric mean (%CV) C\textsubscript{trough} of OD TAC and BID TAC were 5.43 (33.1%) and 6.09 (34.6%) ng/mL, respectively. The GMR of C\textsubscript{trough} of OD TAC versus BID TAC was 0.89 (90% CI 0.82–0.98), which fell outside the equivalence ratio, indicating that, at the same AUC\textsubscript{0-24} exposure, the C\textsubscript{trough} of OD TAC was lower than the C\textsubscript{trough} of BID TAC. The geometric mean (%CV) C\textsubscript{max} of OD TAC and BID TAC were 15.43 (42.0%) and 18.53 (44.3%) ng/mL, respectively, with a GMR of 0.83 (90% CI 0.78–0.89) which also fell outside the equivalence ratio.

There was a good correlation between C\textsubscript{trough} and AUC\textsubscript{0-24} in both BID TAC (R\textsuperscript{2} = 0.71) and OD TAC (R\textsuperscript{2} = 0.80) (Fig. 2). However, the equations for AUC prediction by C\textsubscript{trough} derived from the regression plot of BID TAC and OD TAC were different. The equation for AUC\textsubscript{0-24} prediction by using the C\textsubscript{trough} of BID TAC was AUC\textsubscript{0-24} = 55 + 25.7(C\textsubscript{trough}), while the equation for OD TAC was AUC\textsubscript{0-24} = 10 + 36.2(C\textsubscript{trough}).

Since OD TAC and BID TAC have different formulations and PK, thus, OD TAC should have its own specific target C\textsubscript{trough} level and it should not be the same as the target C\textsubscript{trough} level of BID TAC. According to the targeted C\textsubscript{trough} level of BID TAC (5 to 7 ng/mL), the AUC\textsubscript{0-24} can be calculated from the equations presented in Fig. 2 which was between 183.5 and 234.9 ng/mL x h. By aiming for the same level of AUC\textsubscript{0-24} as BID TAC, the new target C\textsubscript{trough} of OD TAC can be calculated from the OD TAC equation and ranged from 4.8 to 6.2 ng/mL.

To achieve more accuracy than the single timepoint monitoring but less complicated measurement than the full AUC\textsubscript{0-24}, the abbreviated AUC\textsubscript{0-12} of OD TAC and AUC\textsubscript{0-12} of BID TAC derived from two- and three-time point regression equations were detailed in Tables 3 and 4, respectively. The abbreviated AUC\textsubscript{0-12} equation derived from C0, C6, and C12 had the highest correlation with AUC\textsubscript{0-24}. A Bland–Altman plot of AUC\textsubscript{0-24} is depicted in
Fig. 3. The average difference between the linear prediction based on C0, C6, and C12 and the actual AUC 0-24 was 0.0 (SD ± 8.4) ng/mL × h, with a 95% limit of agreement extending from −16.47 to 16.47 ng/mL × h. The scatter of the individual points showed no evidence of bias across the range of the AUC 0-24. Lin’s concordance correlation coefficient was 0.99.

In addition, we further investigated the effects of CYP3A5 genotypic polymorphism on AUC 0-24 and Ctrough after converting from BID TAC to OD TAC. The CYP3A5 gene alleles were identified in the whole blood by real-time reverse transcription polymerase chain reaction by using forward and reverse primers (F5′-CAT GAC TTA GTA GAC AGA TGA-3′, R 5′-GGT CCA AAC AGG GAA GAA ATA-3′). A fluorescent TaqMan probe was utilized to identify the allelic variant of CYP3A5 (rs776746). The patients were then categorized according to CYP3A5 genotypic polymorphism: (1) expressor (CYP3A5 *1/*1 or CYP3A5 *1/*3) and (2) non-expressor (CYP3A5 *3/*3).

Twelve of the 20 patients were CYP3A5 expressor while the remaining patients were non-expressor. In the CYP3A5 expressor group, the geometric means (%CV) of AUC 0-24 were 234.5 (26.3%) and 238.5 (23.5%) ng/mL × h for OD TAC and BID TAC, respectively (Table 5). The GMR (90%CI) was 0.98 (0.91–1.05). The geometric mean (%CV) of Ctrough for OD TAC and BID TAC were 5.77 (24.7%) and 6.74 (25.8%) ng/mL, respectively, with a GMR (90%CI) of 0.86 (0.79–0.93) which fell outside the equivalence ratio (Fig. 4). In the CYP3A5 non-expressor group,

| Variables | Value |
|-----------|-------|
| Age in years, mean (± SD) | 46 (± 12.1) |
| Gender, male/female, n (%) | 12 (60%)/8 (40%) |
| Body weight, kg, mean (± SD) | 61.6 (± 2.86) |
| BMI, kg/m², mean (± SD) | 22.8 (± 3.95) |
| Type of kidney transplant, DKT/LKT, n (%) | 13 (65%)/7 (35%) |
| HLA mismatch, n (%) | 0 (20%) |
| 1–5 | 15 (75%) |
| 6 | 1 (5%) |
| PRA, n (%) | 0–10 |
| 11–50 | 1 (5%) |
| ≥ 50 | 0 (0%) |
| Duration after transplantation, months, median (IQR) | 18.5 (11.6–36.6) |
| Etiology of ESRD, n | DN 4 |
| CGN, IgAN | 3 |
| Obstructive uropathy | 1 |
| Analgesic nephropathy | 1 |
| Hypertension | 2 |
| ADPKD | 1 |
| Unknown | 8 |
| Creatinine, mg/dL, mean (± SD) | 1.34 (± 0.32) |
| eGFR CKD-EPI, mL/min/1.73 m², mean (± SD) | 60.7 (± 16.43) |
| Hemoglobin, g/dL, mean (± SD) | 13.4 (± 1.27) |
| Albumin, g/dL, mean (± SD) | 4.5 (± 0.22) |
| CYP3A5 polymorphism, n (%) | Expressors [*1/*1, *1/*3] 12 (60%) |
| Non-expressors [*3/*3] | 8 (40%) |
| Total daily dose of tacrolimus | mg/day, median (IQR) 4.0 (2.38–5.75) |
| mg/kg/day, median (IQR) | 0.058 (0.038–0.096) |
| Dose in expressors [*1/*1, *1/*3], mg/day, median (IQR) | 5.5 (4.5–7) |
| Dose in non-expressors [*3/*3], mg/day, median (IQR) | 1.75 (1.25–2.75) |
| Mean BID TAC Ctrough, ng/mL, mean (± SD) | 6.03 (± 1.49) |

Table 1. Characteristics of study participants at first intensive PK assessment. SD: standard deviation; HLA: human leukocyte antigens; DKT: deceased donor kidney transplantation; LKT: living donor kidney transplantation; PRA: panel reactivity antibody; DN: diabetic nephropathy; CGN: chronic glomerulonephritis; IgAN: immunoglobulin A nephropathy; ADPKD: autosomal dominant polycystic kidney disease; eGFR CKD-EPI: estimated glomerular filtration rate by chronic kidney disease epidemiology collaboration equation; CYP3A5: cytochrome P450 family 3 subfamily A member 5; BID: twice daily; TAC: tacrolimus.
the geometric mean (%CV) of AUC$_{0-24}$ for OD TAC and BID TAC were 167.9 (41.1%) and 174.1 (35.8%) ng/mL × h, respectively, with a GMR (90%CI) of 0.96 (0.85–1.09). The geometric means (%CV) of C$_{\text{trough}}$ of OD TAC and BID TAC were 4.96 (43.6%) and 5.21 (41.8%) ng/mL, respectively, with a GMR C$_{\text{trough}}$ of 0.95 (0.80–1.13). The GMR OD TAC/BID TAC of both AUC$_{0-24}$ and C$_{\text{trough}}$ in the non-expressor group fell within the equivalence ratio.

There were no major adverse reactions including acute rejection, during the dose conversion period and 3 months after the conversion period. The CKD-EPI eGFR remained stable throughout the study period (Fig. 5).

**Discussion**

The results in the present PK study have demonstrated that 1:1 dose conversion in drug with a narrow therapeutic index such as TAC, from BID TAC to OD TAC without subsequent dose adjustment in stable adult KT recipients who received constant immunosuppressive regimens had a GMR of AUC$_{0-24}$ of OD TAC/BID TAC of
0.98 (90%CI 0.91–1.04) which fell within the range of equivalence ratio (90%CI = 0.9–1.1) while GMR of C\textsubscript{trough} was 0.89 (90%CI = 0.82–0.98) which fell outside the equivalence ratio. The regression plot of AUC\textsubscript{0-24} and C\textsubscript{trough} found that, at the same AUC\textsubscript{0-24} level, OD TAC had lower C\textsubscript{trough} level compared with BID TAC. Patients in the CYP3A5 expressor group exhibited comparable AUC\textsubscript{0-24} despite significantly decreased C\textsubscript{trough} after 1:1 conversion while the non-expressor group showed similar AUC\textsubscript{0-24} and C\textsubscript{trough}.

A comprehensive PK data from all previous prospective PK studies using 1:1 conversion from BID TAC to OD TAC are illustrated in Table 6. In an earlier study by Alloway et al., 20 of 66 patients who had completed PK

| Time point | Equations | R^2 |
|------------|-----------|-----|
| C0 (0 h)   | AUC = 24.68(C0) + 4.40(C2) + 18.87 | 0.85 |
| C0, C1, C2 | AUC = 26.60(C0) + 3.82(C2) + 0.25(C2) + 19.7 | 0.86 |
| C0, C4     | AUC = 11.64(C0) + 8.90(C4) + 32.68 | 0.92 |
| C0, C3, C4 | AUC = 10.47(C0) + 2.09(C3) + 7.23(C4) + 31.05 | 0.92 |
| C0, C4, C6 | AUC = 10.13(C0) + 3.42(C4) + 8.02(C6) + 24.59 | 0.94 |
| C0, C4, C10| AUC = 4.67(C0) + 5.29(C4) + 12.62(C10) + 9.15 | 0.97 |
| C0, C6, C9 | AUC = 6.20(C0) + 7.02(C6) + 11.00(C9) + 0.66 | 0.97 |
| C0, C6, C12| AUC = 0.97(C0) + 5.79(C6) + 18.97(C12) − 4.26 | 0.98 |

Table 3. The proportion of variance in OD TAC AUC\textsubscript{0-24} is explained by single TAC levels, or combinations of TAC levels at multiple time points.

| Time point | Equations | R^2 |
|------------|-----------|-----|
| C0 (0 h)   | AUC = 1.23(C1) + 3.88(C3) + 4.40 (C6) + 6.76 | 0.99 |
| C0, C2     | AUC = 12.25(C0) + 2.94(C2) + 0.025 | 0.90 |
| C0, C2, C3 | AUC = 5.22(C0) + 0.98(C2) + 4.69(C3) + 9.32 | 0.94 |
| C0, C2, C4 | AUC = 1.90(C0) + 2.05(C2) + 5.88(C4) + 10.54 | 0.95 |
| C0, C3, C4 | AUC = 1.99(C0) + 4.64(C3) + 2.74(C4) + 13.15 | 0.94 |
| C0, C3, C6 | AUC = 2.35(C0) + 4.95(C3) + 3.23(C6) + 10.18 | 0.95 |
| C0, C4, C6 | AUC = 3.17(C0) + 6.14(C4) + 2.48(C6) + 8.86 | 0.91 |
| C2, C3     | AUC = 0.65(C2) + 6.46(C3) + 20.93 | 0.93 |
| C2, C4     | AUC = 2.07(C2) + 6.21(C4) + 14.65 | 0.96 |
| C2, C3, C4 | AUC = 1.83(C2) + 0.83(C3) + 5.55(C4) + 14.41 | 0.96 |
| C2, C3, C6 | AUC = 1.47(C2) + 3.15(C3) + 4.46(C6) + 13.46 | 0.96 |

Table 4. The proportion of variance in BID TAC AUC\textsubscript{0-12} is explained by single TAC levels, or combinations of TAC levels at multiple time points.
Figure 3. Bland–Altman plot between observed and predicted TAC AUC$_{0-24}$ by C0, C6, and C12 equation.

Table 5. AUC$_{0-24}$ and C$_{\text{trough}}$ with GMR (90%CI) for the OD versus the BD regimen, by CYP3A5 expression.

| Variables | geometric mean (%CV) | BID TAC | OD TAC | GMR (90%CI) | p-value |
|-----------|----------------------|---------|--------|-------------|---------|
| AUC$_{0-24}$, ng/mL × h | CYP3A5 expressor (CYP3A5 *1/*1, n = 12) | 238.5 (23.5) | 234.5 (26.3) | 0.98 (0.91–1.05) | 0.70 |
| C$_{\text{trough}}$, ng/mL | 6.74 (25.8) | 5.77 (24.7) | 0.86 (0.79–0.93) | 0.003 |
| AUC$_{0-24}$/dose, ng/mL × h/ng/kg/day | 1,749 (74.6) | 1,719.9 (79.0) | 0.98 (0.90–1.07) | 0.70 |
| AUC$_{0-24}$/dose, ng/mL × h/ng/kg/day | CYP3A5 non-expressor (CYP3A5 *3/*3, n = 8) | 174.1 (35.8) | 167.9 (41.1) | 0.96 (0.85–1.09) | 0.62 |
| C$_{\text{trough}}$, ng/mL | 5.21 (41.8) | 4.96 (43.6) | 0.95 (0.80–1.13) | 0.64 |
| AUC$_{0-24}$/dose, ng/mL × h/ng/kg/day | 5,978.0 (24.7) | 5,763.2 (15.6) | 0.96 (0.83–1.11) | 0.62 |

Figure 4. The mean (± SE) tacrolimus concentration–time curves by CYP3A5 genotype of both BID TAC and OD TAC.
profiles had TAC dose adjustment during the PK studies for various reasons. Of note, the values of equivalence ratio used in the study were 0.8–1.25. Despite TAC dose adjustment, the GMR value of AUC 0-24 between OD TAC and BID TAC was 0.95 which fell within the equivalence ratio while that of Ctrough was 0.87 which fell outside the equivalence ratio of narrow therapeutic index (Table 6). In another PK study conducted by Midtvedt et al., the GMR values of AUC 0-24 and Ctrough of OD TAC and BID TAC were 0.82 and 0.81, respectively. Moreover, the study allowed subsequent dose adjustments during the following 2–3 weeks post conversion in order to keep the Ctrough concentration within 5–10 ng/mL. Likewise, a study conducted by Stift et al., the TAC doses were subsequently increased by 1, 1.5, and 2 mg to reach a Ctrough greater than 4.0 ng/mL. The GMR value of AUC 0-24 was 0.98 while that of Ctrough was 0.89 which fell outside the equivalence ratio. Since all of these three PK reports were 1-way conversion studies, van Hooff et al., utilized a 4-period crossover replicate study design in 60 KT patients. Although TAC dose adjustments were prescribed in some of the patients, the analyses were performed in patients without dose modifications. The precise results showed that the values of GMR of AUC 0-24 and Ctrough between both TAC formulations fell within the equivalence ratio. However, it should be noted that the results from these PK studies of 1:1 dose conversion from BID TAC to OD TAC should be interpreted cautiously because there were subsequent dose adjustments and most of these earlier reports were pharmaceutical company-sponsored studies. In addition, the results were inconsistent across the studies.

Table 6. Prospective study of 1:1 mg conversion from BID to OD TAC in stable adult kidney transplant recipients with AUC 0-24 monitoring.

| Study | Alloway | Midtvedt | van Hooff | Stift | The present study |
|-------|---------|----------|-----------|-------|------------------|
| Population | N = 66 | N = 20 | N = 60 | N = 40 | N = 20 |
| Ethnicity | Mainly Caucasian | Caucasian | Mainly Caucasian | Caucasian | Asian |
| Trial design | Open label, 1:1 mg conversion | Open label, 1:1 mg conversion | Open label, 1:1 mg conversion | Open label, 1:1 mg conversion | Open label, 1:1 mg conversion |
| Dose adjustments allow | Yes (no dose adjustment in 18 patients) | Yes (analysis was made in patients without dose adjustment) | Yes | Yes | No |
| Pharmaceutical company sponsored | Yes | No | Yes | Yes | No |
| Mean Ctrough BID TAC = 6.6 ng/mL | Mean Ctrough OD TAC = 5.7 ng/mL | Equivalence ratio = 0.87 (90%CI 0.83–0.92) | Mean Ctrough BID TAC = 6.6±2.9 ng/mL | Mean Ctrough OD TAC = 5.4±1.4 ng/mL | Mean Ctrough BID TAC = 7.4 (7.0–7.7) ng/mL | Mean Ctrough OD TAC = 6.6 (6.2–7.0) ng/mL (p = 0.003) | Mean Ctrough BID TAC = 6.69 ng/mL (CV 34.6%) | Mean Ctrough OD TAC = 5.43 ng/mL (CV 33.1%) | GMR = 0.89 (0.82–0.98) |
| AUC BID vs OD TAC | AUC 0-24 BID TAC = 202.5 ng/mL×h | AUC 0-24 OD TAC = 192.3 ng/mL×h | Ratio = 0.94 (90%CI 0.90–0.99) | AUC 0-24 BID TAC = 217.5 ng/mL×h | AUC 0-24 OD TAC = 213.3 ng/mL×h | Ratio = 0.87 | AUC 0-24 BID TAC = 219.2 (208.1–230.9) ng/mL×h | AUC 0-24 OD TAC = 213.3 (202.6–224.5) ng/mL×h | GMR = 0.98 (0.91–1.04) |
| Conclusion | After dose adjustment, PK of OD TAC was equivalent to BID TAC | Ctrough decreased after conversion | Both AUC 0-24 and Ctrough of BID TAC and OD TAC are similar after conversion | After dose adjustment, AUC 0-24 are similar, but Ctrough was lower in OD TAC | After conversion, AUC 0-24 are the same, but the Ctrough of OD TAC is lower |

Figure 5. Allograft function by eGFR CKD-EPI at before conversion, one month, and 3 months after conversion; p-value by repeated ANOVA. (eGFR CKD-EPI; estimated glomerular filtration rate by chronic kidney disease epidemiology collaboration equation).
Our findings established that in the 1:1 conversion from BID TAC to OD TAC without subsequent dose adjustment, the TAC exposure remains similar despite being approximately 11% lower in the C_trough level (Table 2). Since the AUC_0-24 revealed a similar level of TAC exposure after conversion, the 10% to 15% incrementation of OD TAC dose to maintain the same level of C_trough currently performed in routine clinical practice is not necessary. As shown in Tables 3 and 4, the equations derived from the regression plot of C_trough for AUC prediction (abbreviated AUC) for both BID TAC and OD TAC are different, indicating that the target C_trough level of OD TAC used in real clinical practice should not be the same as that of BID TAC. For a targeted level of AUC_0-24 within the range of 180–240 ng/mL × h, the practical used C_trough level of BID TAC is 5 to 7 ng/mL, the C_trough levels specific for OD TAC should be 4.8 to 6.2 ng/mL. This should be beneficial to physicians in prescribing the dose of OD TAC and monitoring TAC exposure, particularly in the places where PK studies of TAC are not easily to performed.

As stated earlier, the abbreviated AUC of BID TAC is more accurate than C_trough and is less complicated than AUC_0-12 or full AUC_0-24 in therapeutic drug monitoring for KT recipients. Since BID TAC and OD TAC have different concentration profiles, the abbreviated AUC_0-24 equations of BID TAC should not be used to predict AUC_0-24 for OD TAC. In this regard, there is only one study of abbreviated AUC_0-24 for OD TAC published which was conducted in pediatric KT recipients. Herein, our full AUC_0-24 study of OD TAC provides abbreviated AUC equations derived from stable adult KT recipients (Table 3). Physicians can choose one of these equations to suit clinical practice by considering the number and timing of the blood draws. Moreover, for physicians who still mainly use AUC_0-12 for BID TAC adjustment, our study also provides abbreviated AUC_0-12 for BID TAC (Table 4).

Of note, individual TAC PK can be affected by several factors, including hemoglobin levels, serum albumin, drug interactions, ABCB1 or MDR1 gene expression, and CYP3A5 genotypic polymorphism. The CYP3A5 non-expressor recipient (*3/*3 genotype) requires a lower dose, while the expressor recipient (*1/*1 and *1/*3) needs a greater dose to attain the target TAC levels. Of interest, more than 80% of the Caucasians are non-expressor while approximately 50% of Asians are non-expressor. This disparity may affect the PK profile of TAC among different ethnicities.

There are sparse data regarding the role of CYP3A5 genotypic polymorphism on PK profile in 1:1 conversion from BID TAC to OD TAC. Following conversion from BID TAC to OD TAC, earlier retrospective PK studies by Wehland et al., and Jonge et al. demonstrated that C_trough was only significantly reduced in non-expressors. Unfortunately, both studies had a limited number of patients who were CYP3A5 expressor. In the study by Wehland et al., non-expressors were younger and more likely to receive kidneys from living donors, and also tended to have better renal function. Furthermore, AUC_0-24 was not performed in both PK studies. A following prospective PK study by Glowacki et al., showed that the C_trough levels were comparable in the non-expressor group but were significantly lower in OD TAC when compared with BID TAC in the expressor group. The AUC_0-24 values were comparable after 1:1 conversion from BID TAC to OD TAC in both expressor and non-expressor groups. However, when the values of AUC_0-24 were adjusted by TAC dose, the dose-adjusted AUC_0-24 in OD TAC were slightly but significantly lower than BID TAC in both CYP3A5 groups. Our study showed that AUC_0-24 and dose-adjusted AUC_0-24 were similar for both expressor and non-expressor groups following 1:1 conversion from BID TAC to OD TAC without subsequent dose adjustment (Table 3). The discrepancies between the present PK study and that by Glowacki et al., are still inconclusive. The aim of study from Glowacki et al., was to compare the PK profiles between expressor and non-expressor groups. The included participants were categorized into two groups at the start of the study while our work examined the roles of CYP3A5 in the second part of the study, possibly resulting in a less biased study. Nonetheless, both studies had a small sample size. Future studies with a larger sample size are warranted.

The strength of this study was that the 1:1 conversion from BID TAC to OD TAC was strictly controlled. All PK studies were conducted at the Clinical Research Center which provided perfect facilities for the clinical study. However, there were some limitations in this study. First, this was a small, cross-sectional study conducted specifically in Asian KT recipients. Second, 99% of the variability in AUC_0-24 was accounted for in the linear study. However, there were some limitations in this study. First, this was a small, cross-sectional study conducted specifically in Asian KT recipients. Second, 99% of the variability in AUC_0-24 was accounted for in the linear study. Moreover, for physicians who still mainly use AUC_0-12 for BID TAC adjustment, our study also provides abbreviated AUC_0-12 for BID TAC (Table 4).

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Conclusions

Conversion from BID TAC to OD TAC with a 1:1 daily dose without subsequent dose adjustment is appropriate and provides similar TAC exposure regardless of CYP3A5 genotypic polymorphism. Despite the decrease in C_trough of OD TAC, increasing the dose to aiming the same C_trough level as BID TAC is not necessary. The pharmacokinetics of both OD TAC and BID TAC are different. The differences in the target C_trough are acceptable when the AUC_0-24 of both drugs are comparable.

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References

1. Bia, M. et al. KDQOI US commentary on the 2009 KDIGO clinical practice guideline for the care of kidney transplant recipients. Am. J. Kidney Dis. 56, 189–218. https://doi.org/10.1053/j.ajkd.2010.04.010 (2010).
2. Vadcharavivad, S., Saengram, W., Phupradit, A., Poolsup, N. & Chancharoenthana, W. Once-daily versus twice-daily tacrolimus in kidney transplantation: A systematic review and meta-analysis of observational studies. Drugs 79, 1947–1962. https://doi.org/10.1007/s40265-019-01217-7 (2019).
5. Andrews, L. M.
6. Kim, I. W.
10. Hougardy, J. M.
8. Wu, M. J.
16. Wiebe, C.
17. Alloway, R.
19. Crespo, M.
24. van Hooff, J.
21. Staatz, C. E. & Tett, S. E. Clinical pharmacokinetics of once-daily tacrolimus in solid-organ transplant patients. *J. Am. Soc. Nephrol.* 14, 1889–1896. https://doi.org/10.1016/S0150-9161(03)03140-0 (2011).

12. Gallego-Valcarce, E.
14. Schiff, J., Cole, E. & Cantarovich, M. Therapeutic monitoring of calcineurin inhibitors for the nephrologist. *Clin. Transplant.* 23, 866–879. https://doi.org/10.1111/ctr.12310 (2011).

13. Iaria, G. et al. Conversion from Prograf to Advagraf among kidney transplant recipients results in sustained decrease in tacrolimus exposure. *Transplantation* 91, 566–569. https://doi.org/10.1097/TP.0b013e3182924d26 (2011).

15. Voora, S. & Adey, D. B. Management of kidney transplant recipients by general nephrologists: Core curriculum 2019. *Clin. Pharmacokinet.* 58, 1013–1019. https://doi.org/10.1007/s40262-018-0599-6 (2019).

20. Niederberger, K. & Tett, S. E. Comparative pharmacokinetics and pharmacodynamics of once-daily tacrolimus formulation: Monitoring of plasma levels, graft function, and cardiovascular risk factors. *Transplant. Proc.* 37, 2323–2327. https://doi.org/10.1016/j.transproceed.2009.06.070 (2009).

23. Midtvedt, K. et al. Conversion of stable kidney transplant recipients from a twice daily Prograf-based regimen to a once daily modified-release tacrolimus formulation: A pharmacokinetic and bioequivalence study. *Pediatr. Nephrol.* 29, 1081–1088. https://doi.org/10.1007/s00467-013-2724-0 (2014).

31. Julious, S. A. Sample sizes for clinical trials with normal data. *Stat. Med.* 20, 1921–1986. https://doi.org/10.1002/sim.1783 (2004).

34. Sholten, E. M. et al. AUC-guided dosing of tacrolimus prevents progressive systemic overexposure in renal transplant recipients. *Clin. Pharmacokinet.* 50, 451–459. https://doi.org/10.1007/s40262-015-0282-2 (2015).

39. Sweeney, J. et al. Comparison of pharmacokinetics and pharmacogenetics of once- and twice-daily tacrolimus in the early stage after renal transplantation. *Transplantation* 94, 1013–1019. https://doi.org/10.1097/TP.0b013e31826bc600 (2012).

42. Beker, A. Yehezkel, D., Dresler, D. & Melki, Q. A. Effect of low- and high-fat meals on tacrolimus absorption following 5 mg single oral doses to healthy human subjects. *J. Clin. Pharmacol.* 41, 176–182. https://doi.org/10.1177/0091270011422099 (2011).

47. Wong, K. M., Shen, S. C., Chau, K. F. & Li, C. S. Abbreviated tacrolimus area-under-the-curve monitoring for renal transplant recipients. *Am. J. Kidney Dis.* 53, 606–666. https://doi.org/10.1053/j.ajkd.2014.08.013 (2014).

51. Almeida-Paulo, G. N. et al. Limited sampling strategies for tacrolimus exposure (AUCO-24) prediction after Prograf(R) and Advagraf(R) administration in children and adolescents with liver or kidney transplants. *Transpl. Int.* 27, 939–948. https://doi.org/10.1007/s00293-013-1722-z (2014).

57. Min, K. J. et al. Application of the DDI prediction model to a large-scale clinical trial of tacrolimus in renal transplant recipients. *Clin. Pharmacokinet.* 53, 515–531. https://doi.org/10.1007/s40262-013-0141-8 (2014).

63. Vignati, P. et al. Clinical and genetic factors affecting tacrolimus trough levels and drug-related outcomes in Korean kidney transplant recipients. *Transplantation* 94, 2323–2325. https://doi.org/10.1097/TP.0b013e3182924d26 (2011).

69. Wehland, M. et al. Clinical and genetic factors affecting tacrolimus trough levels and drug-related outcomes in Korean kidney transplant recipients. *Transplantation* 94, 2323–2325. https://doi.org/10.1097/TP.0b013e3182924d26 (2011).

75. Thervet, E. et al. Impact of cytochrome p450 3A5 genetic polymorphism on tacrolimus pharmacokinetics and concentration-to-dose ratio in renal transplant recipients. *Transplantation* 76, 1233–1235. https://doi.org/10.1097/TP.0b013e31823b7a89 (2003).

81. Udomkarjananan, S. et al. The cytochrome P450 3A5 non-expressor kidney allograft as a risk factor for calcineurin inhibitor nephrotoxicity. *Am. J. Nephrol.* 41, 176–182. https://doi.org/10.1111/anj.12362 (2014).

87. Hesse, F. et al. A simple novel technique to estimate tacrolimus dosages during the early post kidney transplantation period. *Transplant. Proc.* 47, 2444–2447. https://doi.org/10.1016/j.transproceed.2015.08.013 (2015).

93. Thervet, E. et al. Impact of cytochrome p450 3A5 genetic polymorphism on tacrolimus pharmacokinetics and concentration-to-dose ratio in renal transplant recipients. *Transplantation* 76, 1233–1235. https://doi.org/10.1097/TP.0b013e31823b7a89 (2003).

99. Wehland, M. et al. Differential impact of the CY3P3A4 and CY3P3A5 alleles on pre-dose concentrations of two tacrolimus formulations. *Pharmaceutical Genet. Genomics* 21, 179–184. https://doi.org/10.1097/PPG.0b013e32835ea085 (2011).
Author contributions
K.P., Y.A., N.T. participated in research design. K.Ti., S.T., S.U., P.S., A.V., K.P., N.T. performed research. K.Ti., N.T. analyzed data. K.Ti., K.Ta., K.Tu., S.E., N.T. wrote the manuscript. S.K. advised on statistical analysis. S.K. edited manuscript. S.K. prepared Figs. 1, 2, 3, 4. K.Ti. prepared Fig. 5. S.K., S.T., S.U., P.S., A.V., K.P., K.Ta., K.Tu., S.E., K.P., Y.A., N.T. approved the final manuscript.

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Competing interests
The authors declare no competing interests.

Additional information
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