Glycemic Responses, Glycemic Index, and Glycemic Load Values of Some Street Foods Prepared from Plantain (Musa spp., AAB Genome) in Côte d’Ivoire

Camille Adam Kouamé 1,2,* 1, Nestor Kouakou Kouassi 1,2, Jacko Rhedoor Abodo 3, Kingsley Kwadwo Asare Pereko 4, Maria Cristina Casiraghi 5, Denis Yao N’dri 2 and Georges N’guessan Amani 2

1 Food Biochemistry and Tropical Products Technology Laboratory, Nutrition Section, Department of Food Science and Technology, University Nangui Abrogoua, P.O. Box 801 Abidjan 02, Cote D’Ivoire; nestorkksi@yahoo.fr
2 Food Biochemistry and Tropical Products Technology Laboratory, Biochemistry and Food Technology Section, Department of Food Science and Technology, University Nangui Abrogoua, P.O. Box 801 Abidjan 02, Cote D’Ivoire; ndri_denis@yahoo.fr (D.Y.N.); amanigeorges@yahoo.fr (G.N.A.)
3 Endocrinology Diabetes Nutrition, CHU Yopougon, P.O. Box 632 Abidjan 23, Cote D’Ivoire; sfadabidjan@yahoo.fr
4 Department of Community Medicine, School of Medical Sciences, University of Cape Coast, Cape Coast, Ghana; kpereko@gmail.com
5 Department of Food, Environmental and Nutritional Sciences (DeFENS), Human Nutrition Unit, University of Milan, Via Celoria n. 2, 20133 Milan, Italy; maria.casiraghi@unimi.it
* Correspondence: kadamcamille@gmail.com; Tel.: +225-0776-5139 or +39-38-8851-1528

Received: 13 June 2017; Accepted: 26 July 2017; Published: 16 September 2017

Abstract: The glycemic index (GI) and glycemic load (GL) of four culinary preferences including five local street dishes prepared from three varieties of plantain at different maturity stages was determined. The GI was obtained following ISO/FDI 26642:2010 protocol, and the GL was calculated from test foods’ GI, considering the amount of available carbohydrate in the traditional portion size. GI values were 44 for Klaclo (with Ameletiha variety at all black stage), 39 for Aloco (with Agnrin variety at full yellow stage), 39 for Aloco (with Agnrin variety at full yellow with black spots stage); 45 for Chips (with Ameletiha variety at green stage) and 89 for Banane braisée (with Afoto variety at light green stage). GI values were inversely correlated with the total sugar and carbohydrate in foods (p < 0.01), and no relationship existed between the GI values and the amount of protein (p = 0.89). Except for Chips (GL = 12), the GLs of the others foods were high (GL > 20). Contrary to Banane braisée, the consumption of Klaclo, Aloco, and Chips may promote the control of postprandial glucose response. Data provides the first GI published values of plantain-based foods commonly consumed in the urban area of Abidjan (Côte d’Ivoire).

Keywords: diabetes mellitus; glycemic load; glycemic index; plantain cultivar; snack foods

1. Introduction

As a global commodity for food security, plantain (Musa paradisiaca normalis) has received attention in recent years by nutritionists, agronomists, and agriculturalists. Plantain has been recognized as an important food crop with high potential to improve food security globally considering its production, processing, and utilization [1], and there is a wide body of knowledge about the technology, chemistry, post-harvest physiology, and biochemistry of plantain [2–4]. Plantain is a staple food and an important source of carbohydrates for millions of people worldwide, particularly in sub-tropical countries, with global production estimated at 37.877 million 805 tons in 2013. It is the third food crop in Côte d’Ivoire,
with an annual production of more than 1.624 million 354 tons [5] and an estimated consumption of 120 kg/capita/year [6]. In addition to its high domestic demand, plantain has a high market value both sub-regionally and internationally. Mature plantain pulp and derivative products have a high nutritional value in terms of micronutrients content including calcium, carotenoids, ascorbic acid, and zinc [7]; this composition varies according to the variety and the cultivar, as well as to the stage of fruit maturation. In particular, studies on post-harvest physiology show that plantain composition changes dramatically during ripening; plantain starch is progressively converted into sugars as ripening progresses, likely because of the increased activities of several endogenous enzymes [8]. The fruits of plantain are generally subject to post-harvest culinary processes that take into account the variety, the stage of maturity, and the addition of other ingredients [1]. Some culinary preparations such as roasted or fried plantain, plantain chips, boiled plantain, or pounded plantain, obtained from different varieties, represent a common staple food in many West African Countries—particularly Côte d’Ivoire, Ghana, Nigeria, and Togo [9]. Other preparations found in certain countries include plantain pastry lined with beans or with green leafy vegetables, plantain fritters, plantain pulp cooked with water, palm oil, goat or meat, salt and various spices, etc. [7]. In Côte d’Ivoire, plantains are consumed at all stages of ripeness. Recent data gathered from representative samples of consumers during surveys in urban areas of Abidjan (Côte d’Ivoire) showed four preferred culinary patterns of plantain consumption, according to variety and maturity of fruit, such as fried plantain called “Aloco”, fritters plantain called “Klaclo”, plantain chips “Chips” and roasted plantain “Banane braisée” [10]. Plantain chips are deep-fried thin slices (1.2–0.8 mm thick) of fruits. Fried plantains as for Aloco are thick slices in cubes of peeled ripe fruits, salted and fried in vegetable oil. Fritters plantain (Klaclo) is prepared with blended over-ripe fruit of plantain mixed with corn flour (30–50%), salt, and other spices, modelled into balls and fried in vegetable oil. Aloco and Klaclo are very popular dishes made in the small roadside restaurants and now in good restaurants called “allocodrome” in Côte d’Ivoire. This dish is usually served with grilled fish or hard-boiled egg and onion-tomato sauce and consumed in the afternoon as snack. It has other names such as Dodo in Benin and Amadan in Togo or Dodo-Ikere in Nigeria. Banane braisée (or “blissi”, as it is locally called) consist of the entire pulp of unripe or half-ripe plantain, roasted on heated charcoal with regular turning over to prevent the plantain from charring [11]. Women mainly carry out the preparation of chips, Aloco, fritters, and roasted plantain on the streets, an activity that often represents their principal source of income [10]. From a nutritional point of view, the wide diffusion of these plantain-based meals could have a significant impact on glucose metabolism, thereby contributing to the long-term development of metabolic disorders such as diabetes mellitus, which has become an issue of public health concern in Côte d’Ivoire [12]. Although there have been several reports on the nutritional properties of plantain [4], no data are available about the glycemic index (GI) or glycemic load (GL) of these street foods. This research was therefore undertaken to evaluate the GI and GL of several street foods, prepared with traditional recipes, from three local varieties of plantain at different stages of fruit ripeness.

2. Materials and Methods

2.1. Setting

The study was performed at the Department of Food Sciences and Technology of the University Nangui Abroguia using internationally recognized GI methodology [13,14]. All clinical procedures were carried out at the Endocrinology and Diabetology Center, CHU Yopougon (Félix Houphouet Boigny University), Abidjan, Côte d’Ivoire. This study was conducted according to the principles of the Helsinki Declaration and approved by the Ethical Committee of the Félix Houphouet Boigny University Research, Cocody. Written informed consent was obtained from all subjects prior to participation. Participants were informed about the complete details of the study protocol and were given the opportunity to ask questions or to withdraw from the study at any time.
2.2. Materials

The reference food glucose (Glucose pur anhydre, COOPER, Place Lucien Anvert) was from Arts Pharmacy Limited (Abidjan-District, Côte d’Ivoire). Fresh and mature plantain fruit of three varieties—Afoto variety (Musa spp., AAB group, Queensland, Australia, cv False Horn), Agnrin variety (Musa spp., AAB group, cv French Horn), and Amelieha variety (Musa spp., AAB group, Queensland, Australia, cv French)—were used in this experiment. Fruits were purchased in the local market of Yopougon-Siporex, Abidjan-District (Côte d’Ivoire) in quantities sufficient to conduct all tests. In order to make samples more homogeneous, each batch of plantain was bought on a single day from the same seller and stored in the laboratory under the same conditions, at room temperature (28 ± 2 °C) and relative humidity of 80–90%.

2.3. Subjects

The study involved a group of 30 (23 men and 7 women) healthy non-obese and physically active volunteers. They were recruited from the University Nangui Abrogoua among the staff and students through advertisement and were selected based on age (18–40 years), BMI (19–26 kg/m²), and fasting blood glucose value (4–5.5 mmol/L). Glycated hemoglobin (HbA1c) was also evaluated, and subjects with an HbA1c <8% were included in the study. Smokers were excluded from the study. Anthropometric measurements were carried out for all the subjects using standardized methods before the start of the study. Height was recorded to the nearest centimeter using a Stadiometer (Seca Limited, Birmingham, West Midlands, UK) with the subjects standing erect without shoes. Body weight was recorded using the Tanita BC-418 MA (Tanita UK Limited, Yiewsley, Middlesex, UK) with the subjects wearing light clothing and no shoes, and blood pressure was measured with an automatic device (A & D Company Ltd., Tokyo, Japan). Determination of HbA1c was assessed with a Bio-Rad DiaSTAT™ Hemoglobin A1C Analyzer (Bio-Rad Laboratories Inc., Hercules, CA, USA). Subjects’ characteristics are summarized in Table 1. During the study, subjects were advised to maintain their habitual daily activities without any change in their physical activities.

Table 1. Baseline clinical and anthropometric characteristics (mean ± standard error of mean (SEM)) of subjects (n = 30) involved in the study.

| Parameters     | Mean | SEM  | Range    |
|----------------|------|------|----------|
| Age (years)    | 30   | 0.5  | 25–35    |
| Gender (male/female) | 23/7 | -    | -        |
| Body weight (kg) | 63.3 | 1.3  | 47–74    |
| Height (m)     | 1.7  | 0.0  | 1.6–1.9  |
| BMI (kg/m²)    | 21.2 | 0.3  | 18.1–24.6|
| Fasting glucose (mmol/L) | 4.6  | 0.1  | 4.1–5    |
| HbA1c (%)      | 3.0  | 0.1  | 2.1–4.0  |
| Systolic BP (mmHg) | 107.7 | 1.7  | 90–120   |
| Diastolic BP (mmHg) | 73.0 | 1.3  | 60–90    |

BMI = body mass index, HbA1c = hemoglobin A1c, BP = Blood pressure.

2.4. Test Foods—Collection of Samples and Preparation of Experimental Diets

Five local plantain-based dishes were prepared at the Food Biochemistry and Tropical Products Technology laboratory of University Nangui Abrogoua. These dishes were chosen on the basis of their frequency of consumption by the population of the urban areas of Abidjan [10]. They included fried plantain “Aloco” prepared from fruits at two stages of maturity—the full yellow stage (aag6) and the full yellow with black spots (aag7); fritters plantain “Klaclo” from fruit at the black stage of maturity (kam8); plantain chips from the green stage (Cam1); and charcoal-roasted “Banane braisée” from the
light green stage of maturity (raf2). These stages correspond to preferential uses in traditional culinary preparations. Samples of the various food products were prepared by roasting and frying according to the traditional methods as described by Aboua et al. [11]. Table 2 lists the plantain species employed in the study, their ripening stages, and cooking/processing methods applied to formulate the test foods.

Table 2. Composition and local cooking/processing methods applied to formulate the tested foods.

| Food Tests     | Code | Plantain Species (Musa spp., AAB Genome) | Ripening Stage at Use | Local Name | Cooking Method  | Major Ingredients                  |
|----------------|------|------------------------------------------|-----------------------|------------|----------------|-----------------------------------|
| Charcoal-roasted plantain | raf2 | Afofo | light green (stage 2) | Banane braisee | Roasting | - |
| Fried plantain | aag6 | Agranin | full yellow (stage 6) | Alcoco | Deep frying | Salt, refined palm oil |
| Fried plantain | aag7 | Agranin | full yellow with black spots (stage 7) | Alcoco | Deep frying | Salt, refined palm oil |
| Fritters plantain | kam8 | Ameletiha | all black (stage 8) | Klaclo | Deep frying | Salt, refined palm oil, corn flour (30%) |
| Chips plantain | cam1 | Ameletiha | green (stage 1) | Chips | Deep frying | Salt, refined palm oil |

2.5. Proximate Analysis

All foods were tested immediately after cooking. Moisture, ash, lipids, and protein were assessed by following AACC International approved Methods n. 44-15.02, 08-01.01, 30-10.01, and 46-12.01, respectively. The total dietary fiber content was evaluated in accordance with the method of Prosky [15]; available carbohydrates (AvCHO) were calculated by difference as suggested by FAO/WHO procedure [13]. Total sugars (TS) were determined using the 3,5-dinitrosalicylic acid method [16]. All the reported determinations were carried out in triplicate.

2.6. Glycemic Index Testing Procedures

The study was performed in accordance with the international standard GI testing protocol [14], in line with procedures recommended by the FAO/WHO Expert Consultation [13]. Subjects were invited to attend the test sessions on seven consecutive occasions with a 2-day interval between test days. Fifty (50) grams of anhydrous glucose powder dissolved in 250 mL water was used twice as the reference food; plantain dishes were prepared on the day of testing following the common practices used by the local food sellers in Côte d’Ivoire. All foods were tested in portions containing equivalent available carbohydrate amounts (50 g). On the day before a test, subjects were asked to restrict their intake of alcohol and caffeine-containing drinks and to avoid intense physical activity. The order of test foods was randomized, and all the foods were tested after a 12-h overnight fast. Blood glucose concentrations were measured in the capillary whole blood obtained by finger prick (Accu-Chek® Fastclix Lancing Device, Castle Hill, NSW, Australia) in the fasted state and at 15, 30, 45, 60, 90, and 120 min after the start of the meal. Blood glucose was measured using a calibrated Accu-Chek® Performa glucometer (Accu-Chek Performa, Roche Diagnostic, Castle Hill, NSW, Australia).

2.7. Calculation of Glycemic Index and Glycemic Load

The incremental area under the post-prandial blood glucose curve (iAUC), ignoring the area beneath the baseline, was calculated geometrically for each tested food [17], and the GI was evaluated as a percentage of the mean iAUC of the reference glucose solution consumed by the same subject (GI = iAUC test food/iAUC reference food × 100). When the individual GI values for any subject fell outside the range of values calculated as mean ± 2 SD (standard deviation), this result was considered as an outlier and was thus excluded from the mean GI calculation. The glycemic load (GL) of a specific serving of each food was calculated using the formula: GL = GI food/100 × g of available
carbohydrates in the portion. Based on the consumption habits observed in Côte d’Ivoire [10], the GL of the different tested foods was calculated considering the portions reported in Table 3.

Table 3. Proximate composition of tested meals.

| Food Samples (Code)        | Banane Braisée (raf2) | Klaclo (kam8) | Alococ (aag7) | Alococ (aag6) | Chips (cam1) |
|---------------------------|-----------------------|--------------|--------------|--------------|--------------|
| Dry matter (g/100 g)      | 44.5 ± 0.1 d          | 63.7 ± 0.1 b | 66.2 ± 0.0 c | 63.6 ± 0.1 b | 66.4 ± 0.1 a |
| Ash (g/100 g)             | 1.4 ± 0.0 c           | 1.1 ± 0.0 d  | 1.0 ± 0.0 e  | 1.5 ± 0.0 b  | 2.0 ± 0.0 a  |
| Proteins (g/100 g)        | 5.3 ± 0.0 c           | 6.1 ± 0.0 b  | 8.8 ± 0.0 c  | 4.4 ± 0.0 d  | 5.3 ± 0.0 c  |
| Lipids (g/100 g)          | 0.3 ± 0.1 e           | 14.1 ± 0.2 a | 12.4 ± 0.0 b | 11.6 ± 0.1 c | 10.9 ± 0.0 d |
| Total CHO (g/100 g)       | 93.1 ± 0.1 a          | 78.7 ± 0.2 d | 78 ± 0.0 e   | 82.6 ± 0.1 b | 81.9 ± 0.1 c |
| Total sugars (g/100 g)    | 6.5 ± 0.1 c           | 12.5 ± 0.3 a | 10.3 ± 0.3 b | 9.8 ± 0.1 b  | 4.0 ± 0.3 d  |
| Starch (g/100 g)          | 77.9 ± 0.2 a          | 99.6 ± 0.0 e | 60.9 ± 0.2 a | 65.5 ± 0.2 c | 70.1 ± 0.3 b |
| Total dietary fiber (g/100 g) | 1.7 ± 0.0 a         | 1.6 ± 0.0 b  | 1.6 ± 0.0 b  | 1.6 ± 0.0 b  | 1.7 ± 0.0 a  |
| Energetic Value (kcal/100 g) | 395.8 ± 0.4 e       | 465.7 ± 1 a  | 458.0 ± 0.3 b | 451.9 ± 0.4 c | 446.7 ± 0.2 d |

Data are expressed on dry matter basis. Data in the same line with different superscript letter are significantly different (p < 0.05) as assessed by Tukey’s test. Data are means (standard deviation) of three independent experiments.
* Calculated by difference of moisture content, ash, fiber, lipids and protein. ** Energetic value = 4 × %total carbohydrates + 4 × %protein + 9 × %lipids (kcal/100g)

2.8. Statistical Analysis

Data processing was carried out using SPSS software (version 17.0, SPSS, Chicago, IL, USA). Data are presented as means ± standard deviation (SD) or means ± standard error of mean (SEM), as indicated. Comparisons between the foods’ composition were made by using one-way analysis of variance (ANOVA) and Tukey’s multiple comparisons test. For GI data analysis, differences in postprandial blood glucose concentrations at any time points, iAUCs, GI, and GL values were evaluated using repeated-measures analysis of variance (RM-ANOVA). Spearman’s correlation coefficient was used to assess the relationship between GI values and macronutrient content of tested foods. Statistical significance was set at p < 0.05.

3. Results and Discussion

3.1. Chemical Composition of Meals

Energy and proximate composition of tested foods are shown in Table 3. The dry matter content of the meals varied from 44.5 g/100 g for Banane braisée to 66.4 g/100 g for Chips. As expected, plantain meals contain low amounts of proteins, varying in a range of values from 4.4 g/100 g dw (Alococ aag6) to 8.8 g/100 g dw (Alococ aag7), but a high lipid content. In particular, except for Banane braisée which showed a low lipid content (0.3 g/100 g dw), in Klaclo, Alococ (aag6 and aag7), and Chips, lipid levels were 14.1, 12.4, 11.6, and 10.9 g/100 g dw, respectively. Carbohydrates were the main nutrient present in meals, ranging from 78 to 93 g/100 g dw, with amounts of total sugar varying from 4 to 12.5 g/100 g dw. Total dietary fiber content was low and very similar among meals. These results are not surprising, since several previous studies have shown that the plantain is known for being very rich in starch and an excellent source of energy [18,19]. However, statistical analysis showed significant differences (p < 0.05) between the means of the variables estimated in the proximate composition of the test foods (Table 3). These differences could partly be attributed to certain inherent characteristics in each plantain variety, such as genotype, the conditions of cultures, and nature of soils [19], as well as the variability in the traditional processing/cooking methods used (Table 1). Numerous studies have reported that plantain is poor in fiber, lipid, and protein [18–20], and the nutritional composition of plantain is diversely affected by processing methods [11], as shown by our data (Tables 2 and 3). Moreover, as expected, fried plantain contained the highest amount of lipid and dry matter compared to the roasted plantain (Table 3). Alococ aag6 and aag7, Chips, and fritter plantain (Klaclo) probably absorbed great amounts of oil, whereas the unripe plantain (roasted plantain) may have lost lipid during the roasting process, as explained in an earlier study [11]. Furthermore, these authors suggested...
that palm oil frying presents some nutritional advantages due to a vitamin and lipid enriching effect on the plantain meals, increasing its dry matter unlike the roasting process. These facts are important, because a high moisture content in food has been shown to accelerate microbial growth and food spoilage [21]. The serving sizes calculated to contain 50 g available carbohydrate are shown in Table 4. The GI testing portion differed considerably with the smallest portion calculated for Chips (93.8 g) and the largest portion for the Banane braisée (122.9 g).

Table 4. Glycemic index and glycemic load values of tested product.

| Food Code | Experimental Portion (g) | Available CHO (g/100 g of Food Wet Weight) | GI (Mean ± SEM) | Category 1 | Serving Size | GL | Category 2 |
|-----------|-------------------------|-------------------------------------------|-----------------|-------------|--------------|----|------------|
| Banane braisée | raf2 | 41 | 122.9 | 28 | 88 ± 1.8 a | High | 178.8 | 64 | High |
| Klaclo | kam8 | 49 | 101.8 | 28 | 44 ± 0.3 c | Low | 227.0 | 46 | High |
| Alococ | aag6 | 50 | 99.0 | 29 | 39 ± 0.3 b | Low | 281.1 | 55 | High |
| Alococ | aag7 | 52 | 97.0 | 29 | 39 ± 0.5 b | Low | 281.1 | 55 | High |
| Chips | cam1 | 53 | 93.8 | 28 | 45 ± 0.3 c | Low | 49.4 | 12 | Medium |

a,b,c Data in the same column with different letter superscripts are significantly different (p < 0.05); n = number of values included after outliers analysis. GI = Glycemic Index; GL = Glycemic Load; CHO = Carbohydrate. 1 Glycemic indexes were classified as high (≥ 70), medium (56–69), and low (≤ 55); 2 Glycemic loads were classified as high (≥ 20), medium (11–19), and low (≤ 10) (www.glycemicindex.com).

3.2. Glycemic Responses, Glycemic Index, and Glycemic Load of Test Meals

The mean intra-individual CV of glycemic responses to the two 50 g glucose standard tests for the thirty subjects was about 4%, and the mean inter-individual CV in glycemic response to the standard tests was 11%. These values are consistent with reported data that low mean within-subject variation (reference CV < 30%) is required for accuracy [17].

The mean post-prandial blood glucose curves of each test meal and reference food (glucose solution) are shown in Figure 1. Blood glucose concentration increased to the maximum value at 45 min for all the test foods and then decreased until 120 min. There were no significant variations between subjects regarding the mean basal blood glucose concentration assessed before ingestion of the test meals in all seven occasions. Blood glucose levels evaluated at each time point after the consumption of Banane braisée were not significantly different from those elicited by the glucose standard meal, except for times 30 and 45 min, in which capillary whole-blood glucose levels assessed for this meal were significantly lower than those observed for the glucose reference meal (p < 0.05). Furthermore, no significant differences were evidenced between blood glucose values at 15, 30, 45, 60, and 120 min after consumption of Klaclo, Alococ aag6, Alococ aag7, and Chips (p > 0.05). The postprandial glucose peak was consistently observed at 45 min for all the tested meals; the magnitude of the peak was significantly lower in subjects that received the fried plantain (p < 0.05) than in those who consumed roasted plantain (Banane braisée)—an effect that was also observable and significant for the iAUCs calculated for these meals (p < 0.001). The mean iAUC evaluated for Alococ (aag7), Alococ (aag6), Klaclo, and Chips were 94.5, 93.9, 107.5, and 110.1 mmol × min/L, respectively, resulting significantly (p < 0.05) lower than the iAUC calculated for Banane braisée.

The GI/GL values and classifications for each test meal are given in Table 4. The mean GIs assessed for plantain fried meals were low, ranging from 38 to 45; on the contrary, as expected from its iAUC value, Banane braisée showed the highest GI value of 89. In terms of variability, CVs evaluated for GI data were highest for Banane braisée (11%), followed by Alococ aag7 (7%), Alococ aag6 (4%), Klaclo, and Chips (3%). When RM-ANOVA with the post-hoc Tukey’s multiple comparison tests was applied to the all experimental data, significant (p < 0.05) differences were found among GI. In particular, Klaclo (GI = 44) and Chips (GI = 45) meals showed significantly (p < 0.05) higher values than those of evaluated for Alococ aag6 and Alococ aag7 (GI = 39). The higher GI of Banane braisée prepared at light green stage of fruit maturity assessed in this study is in contrast with that reported in other studies in which low and/or intermediate GI were observed [22]. This contrast could be related...
...varietal differences of the species roasted. Nevertheless, in the case of Banane braisée, this food (the entire pulp of plantain) was processed on charcoal fire using dry heat. This may have resulted in the loss of water and the degradation of starches, thus increasing their digestibility. On the other hand, foods processed by frying in palm oil were found to have low GI (GI < 55) [23]. As reported in previous work [24,25], the lower GI observed upon frying compared to roasting could be attributed to the high lipid content, which promotes a slow rate of starch digestion as a consequence of the delay of gastric emptying. In fact, lipids are known to slow gastric emptying [26], corroborated by the significant correlation ($p < 0.05$) assessed in this study between the GI values and the lipid content of meals. Lipids delay the transition time of the stomach contents to the duodenum, thus reducing starch hydrolysis rate and the absorption of monosaccharides through the microvilli of the epithelial cells of the jejunum as well as in upper part of the ileum, leading to positive effects on the postprandial glycemic and insulin responses [27]. However, even though Klaclo, Chips, and Aloco may result in lower GI values than Banane braisée, they are processed with increasing amounts of palm oil, and thus their consumption should not be promoted [26].

![Glycemic response curves](image)

**Figure 1.** Glycemic response curves elicited by 50 g available carbohydrate portions of reference glucose, Banane braisée, Klaclo, Aloco aag6, Aloco aag7, and Chips. Values are the mean for 30 subjects with their SEM represented by vertical bars.

Although the dietary glycemic index is directly relevant in metabolic studies in which the total carbohydrate content is held constant, in free-living populations the amount of carbohydrate (e.g., as a percentage of energy) and its composition varies among individuals. Since the glucose and insulin responses depend on both the quantity and quality of the carbohydrate, the dietary glycemic load was used to represent both these traits of the carbohydrate intake. Indeed, high-GL diets increase the risk of diabetes by chronically increasing insulin demand, which in turn may lead to β-cell exhaustion, dysfunction, and apoptosis [28]. The GLs calculated for tested plantain foods appear high (GL > 20) [23], despite the fact that the majority of the meals (except for Banane braisée) had shown a low GI (GI < 55). The high GL values assessed can mainly be attributed to the large serving-size in which these plantain based meals are traditionally consumed. Considering the...
widespread consumption of these foods in Côte d’Ivoire, a suggestion to reduce the portion sizes of these street foods should therefore be recommended in the Ivoirian dietary guidelines, in an attempt to promote consumer behaviors which could favor a better control of glycemic metabolism. For optimal health, aim to keep a daily GL under 100 (www.glycemicindex.com). Thus, the portion sizes which are currently popular must be limited to 200 g for Banane braise, 300 g for Klaclo and Aloco, and 100 g for Chips. The relationships between meals composition and GI were investigated by bivariate correlations. As expected, correlation data analysis showed that the nutrient intake of the test meals influences their GI values. Lipid exhibited the strongest negative correlation with GI values (Spearman’s $\rho = -0.542$; $p < 0.01$), and no significant association was observed between the protein level of tested food and their GI values (Spearman’s $\rho = 0.126$; $p > 0.01$). This was expected, as previous findings from Henry et al. [29] indicated that a small amount of protein in foods—as observed in this study (4.4–8.8 g/100 g; Table 3)—does not significantly affect the glycemic response. At least 20–30 g dietary protein is needed to increase insulin responses sufficiently to reduce glycemic responses [30].

4. Conclusions

Our data set provides for the first time the GI values of some local plantain-based street foods commonly consumed in the urban area of Abidjan. Our results underline that meals such as Klaclo, Aloco, and particularly Chips may promote a low postprandial glucose response, in contrast with the opinion diffused among Ivorian population. However, despite their low GI, the GL assessed for this street food was quite high, suggesting a revision of portions commonly consumed. Moreover, these products are formulated with amounts of palm oil and thus should not be promoted. The importance and popularity of plantain in Côte d’Ivoire as a food crop dictate the need to characterize the glycemic impact (GI and GL) of all the foods prepared from plantain with the aim of better understanding their role in managing and/or preventing diseases related to glucose metabolism.

Acknowledgments: The Agricultural Productivity Program in West Africa (PPAAO/WAAPP 1B) supported this work by the FIRCA (Fonds Interprofessionnel pour la Recherche et le Conseil Agricoles; Don IDA N°6260 CI et Don N°TF 098014 CI). The authors are grateful to the participants of this study. The author would like to thank the following MSc students for the helpful assistance: N’Cho laeticia, Mogué Raoul and Ahoua Prudence.

Author Contributions: C.A.K. is the principle investigator for this research and prepared the manuscript. C.A.K. determined the incremental area under the curve, glycemic index and glycemic load of test foods and drafted the manuscript. N.K.K. carried out the collection of samples and description/preparation of experimental diets. N.K.K. assisted in the food processing and proximate analysis. D.Y.N. participated in the design of the study and performed the statistical analysis. M.C.C. has been involved in data analysis and critically revising the manuscript for important intellectual content K.K.A.P. assisted in the editing of script and making input in scientific writing. A.J.R. and M.B. carried out the blood glucose measurements and conducted the selection of volunteers. G.N.A. is the research coordinator and the guarantor. All the authors read and approved the final manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Cauthen, J.; Jones, D.; Gugerty, M.K.; Leigh Anderson, C. Banana and Plantain Value Chain: West Africa Evans School Policy Analysis and Research (EPAR). EPAR TECHNICAL REPORT #239, Brief No. 239. 2013, pp. 1–25. Available online: https://evans.uw.edu/sites/default/files/Evans_UW_Request_239_West_Africa_Bananas_and_Plantains_01_08_14_final_1.pdf (accessed on 21 August 2016).
2. Arisa, N.N.; Adelakan, A.O.; Alamu, A.E.; Oggunfowora, E.J. The effect of pretreatment of plantain (Musa paradisiaca) flour on the pasting and sensory characteristics of biscuit. Int. J. Food Nutr. Sci. 2013, 2, 10–23.
3. Odenigbo, A.M.; Asumugha, U.V.; Ubbor, S.; Nwazozor, C.; Otuonye, C.A.; Offia-Olua, I.B.; Princewili-Ogbonna, I.L.; Nzeagwu, O.C.; Henry-Uneze, H.N.; Anyika, J.U.; et al. Proximate Composition and Consumption Pattern of Plantain and Cooking-Banana. Br. J. Appl. Sci. Technol. 2013, 3, 1035–1043. [CrossRef]
4. Oladele, E.-O.P. Resistant Starch in Plantain (Musa AAB) and Implications for the Glycaemic Index. Ph.D. Thesis, School of Food Science and Nutrition, University of Leeds, United Kingdom, 2013; p. 281. Available online: http://etheses.whiterose.ac.uk/id/eprint/5239 (accessed on 21 August 2016).
5. FAOSTAT Data. Available online: http://fenix.fao.org/faostat/beta/fr/#data/QC (accessed on 21 August 2016).

6. CNRA (Centre Nationale de Recherche Agronomique). Programme Plantain, Bananes, Ananas. Actualité de Recherche. Available online: http://www.cnra.ci/descprog.php?id=13&prog=Plantain,%20Bananes,%20Ananas&act=present (accessed on 21 August 2016).

7. Ekesa, B.N.; Kimiywe, J.; Davey, M.W.; Dhuique-Mayer, C.; Van Den Bergh, I.; Karamura, D.; Blomme, G. Banana and plantain (Musa spp.) cultivar preference, local processing techniques and consumption patterns in Eastern Democratic Republic of Congo. Int. J. Agric. Sci. 2012, 4, 312–319.

8. Happi, E.T.; Andrianaivo, R.H.; Wathelet, B.; Tchango Tchango, J.; Paquot, M. Effects of the stage of maturation and varieties on the chemical composition of banana and plantain peels. Food Chem. 2007, 103, 590–600. [CrossRef]

9. Dzomeku, B.M.; Dankyi, A.A.; Darkey, S.K. Socioeconomic importance of plantain cultivation in Ghana. J. Anim. Plant Sci. 2011, 21, 269–273.

10. Kouamé, A.C.; Kouassi, K.N.; N’dri, Y.D.; Amani, N.G. Plantain (Musa spp., AAB genome) Cultivar Preference, Local Processing Techniques and Consumption Patterns of Plantain Based Foods Mostly Consumed in Urban Area of Abidjan, Côte d’Ivoire. Nat. Technol. 2015, 12, 117–129.

11. Aboua, F. Effect of home processing methods on the nutritional value of plantains in Ivory Coast. Trop. Sci. 1994, 34, 274–281.

12. Adoueni, K.V.; Anktche, A.; Azoh, P.; Sibailly, A.; Derbe, M.; Sanogo, F.; Acka, F.; Kofi, S.; Koupoukou, E. Prise en charge du patient diabétique en Côte d’Ivoire sur le mode de la décentralisation. Diabetes Metab. 2012, 38, 52. [CrossRef]

13. Joint FAO/WHO Expert Consultation. Carbohydrates in Human Nutrition; FAO Food and Nutrition Paper 66; FAO: Rome, Italy, 1998.

14. International Standards Organisation. Food Products-Determination of the Glycaemic Index (GI) and Recommendation for Food Classification; ISO 26642-2010; International Standards Organisation: Geneva, Switzerland, 2010.

15. Prosky, L.; Asp, N.G.; Schweizer, T.F.; DeVries, J.W.; Furda, I. Determination of insoluble and soluble dietary fiber in foods and food products: Collaborative study. J. AOAC Int. 1992, 75, 360–367.

16. Dubois, M.; Gilles, K.A.; Hamilton, J.K.; Rebers, P.A.; Smith, F. Colorimetric method for the determination of sugars and related substances. Anal. Chem. 1956, 28, 350–356. [CrossRef]

17. Wolever, T.M.S.; Brand-Miller, J.C.; Abernethy, J.; Astrup, A.; Atkinson, F.; Axelsen, M.; Bjorck, I.; Brighenti, F.; Brown, R.; Brynes, A.; et al. Measuring the glycemic index of foods: Interaboratory study. Am. J. Clin. Nutr. 2008, 87, 247–257.

18. Kouadio, K.A.; Coulibaly, S.; Atchibri, L.O.; Kouamé, G.; Meité, A. Évaluation nutritionnelle comparative des fruits de trois hybrides de bananiers (CRBP 39, FHIA 17 et FHIA 21) avec ceux de la variété Orishele. Tropiccultura 2012, 30, 49–54.

19. Badila, C.; Diateau, M.; Ellaly, G.G.; Nguyen, D. Mise au Point d’un Procédé de Fabrication de Farine de Banane Plantain et des Tubercules de Patate Douce I: Evaluation des Caractéristiques Chimiques des Farines; Annales de l’Université Marien NGOUABI; Annale des sciences et techniques: République du Congo, Brazzaville, 2009; Volume 10, pp. 57–63.

20. Pirasath, S.; Thayananthan, K.; Balakumar, S.; Arasaratnam, V. Including side dishes to traditional main meals alter the glycaemic index. Sri Lanka J. Diabetes Endocrinol. Metab. 2013, 3, 12–18. [CrossRef]

21. Egbebi, O.A.; Bademosi, A.T. Chemical compositions of ripe and unripe banana and plantain. Int. J. Trop. Med. Public Health 2012, 1, 1–5.

22. Ogbuji, C.A.; Odom, T.C.; Ndulaka, J.C.; Ogbodo, M.O. Effects of various processing methods of ripe and unripe plantain diets on blood glucose level. Eur. J. Biol. Med. Sci. Res. 2013, 1, 49–54.

23. The University of Sydney, Glycemic Index. Sydney University Glycemic Index Research Service (SUGIRS). Available online: www.glycemicindex.com (accessed on 21 August 2016).

24. Odenigbo, A.; Rahimi, J.; Ngadi, M.; Amer, S.; Mustafa, A. Starch digestibility and predicted glycemic index of fried sweet potato cultivars. Funct. Food Health Dis. FFHD 2012, 2, 280–289.

25. Bahado-Singh, P.S.; Riley, C.K.; Wheatley, A.O.; Lowe, H.I. Relationship between processing method and the glycemic indices of ten sweet potato (Ipomoea batatas) cultivars commonly consumed in Jamaica. J. Nutr. Metab. 2011. [CrossRef] [PubMed]
26. Forouhi, N.G.; Koulman, A.; Sharp, S.J.; Imamura, F.; Kröger, J.; Schulze, M.B.; Croew, F.L.; Huerta, J.M.; Guevara, M.; Beulens, J.W.; et al. Differences in the prospective association between individual plasma phospholipid saturated fatty acids and incident type 2 diabetes: The EPIC-InterAct case-cohort study. *Lancet Diabetes Endocrinol.* **2014**, *2*, 810–818. [CrossRef]

27. Gentilcore, D.; Reawika, C.; Karen, L.J.; Antonietta, R.; Christine, F.-B.; Judith, M.W.; Rayner, C.K.; Horowitz, M. Effects of fat on gastric emptying of and the glycemic, insulin, and incretin responses to a carbohydrate meal in type 2 diabetes. *J. Clin. Endocrinol. Metab.* **2006**, *91*, 2062–2067. [CrossRef] [PubMed]

28. Willett, W.; JoAnn, M.; Simin, L. Glycemic index, glycemic load, and risk of type 2 diabetes. *Am. J. Clin. Nutr.* **2002**, *76*, 274–280.

29. Henry, C.J.K.; Lightowler, H.J.; Strik, C.M.; Renton, R.; Hails, S. Glycaemic index and glycaemic load values of commercially available products in the UK. *Br. J. Nutr.* **2005**, *94*, 922–930. [CrossRef] [PubMed]

30. Manthou, E.; Maria, K.; Kalliopi, G.; Chariklia, K.D.; Dimitrios, K.; Yiannis, K.; Athanasios, Z.J. Glycemic Response of a Carbohydrate-Protein Bar with Ewe-Goat Whey. *Nutrients* **2014**, *6*, 2240–2250. [CrossRef] [PubMed]

© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).