Pre-existing humoral immunity to low pathogenic human coronaviruses exhibits limited cross-reactive antibodies response against SARS-CoV-2 in children

Nina Li1,2†, XueYun Li1,2†, Jiani Wu1,2, Shengze Zhang1,2, Lin Zhu1,2, QiQi Chen1,2, Ying Fan1,2, Zhengyu Wu1,2, Sidian Xie1,2, Qi Chen1,2, Ning Wang3, Nan Wu4, Chuming Luo1,2, Yuelong Shu1,2,5,6* and Huanle Luo1,2,6*

1School of Public Health (Shenzhen), Shenzhen Campus of Sun Yat-sen University, Shenzhen, China, 2School of Public Health (Shenzhen), Sun Yat-sen University, Guangzhou, China, 3Shenzhen Institute of Advanced Technology, Chinese Academy of Sciences, Shenzhen, China, 4Department of Epidemiology, Shenzhen Nanshan Center for Disease Control and Prevention, Shenzhen, China, 5Institute of Pathogen Biology, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China, 6Key Laboratory of Tropical Disease Control, Sun Yat-sen University, Ministry of Education, Guangzhou, China

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection causes asymptomatic or mild symptoms, even rare hospitalization in children. A major concern is whether the pre-existing antibodies induced by low pathogenic human coronaviruses (LPH-CoVs) in children can cross-react with SARS-CoV-2. To address this unresolved question, we analyzed the pre-existing spike (S)-specific immunoglobin (Ig) G antibodies against LPH-CoVs and the cross-reactive antibodies against SARS-CoV-2 in 658 serum samples collected from children prior to SARS-CoV-2 outbreak. We found that the seroprevalence of these four LPH-CoVs reached 75.84%, and about 24.64% of the seropositive samples had cross-reactive IgG antibodies against the nucleocapsid, S, and receptor binding domain antigens of SARS-CoV-2. Additionally, the re-infections with different LPH-CoVs occurred frequently in children and tended to increase the cross-reactive antibodies against SARS-CoV-2 in 658 serum samples collected from children prior to SARS-CoV-2 outbreak. We found that the seroprevalence of these four LPH-CoVs reached 75.84%, and about 24.64% of the seropositive samples had cross-reactive IgG antibodies against the nucleocapsid, S, and receptor binding domain antigens of SARS-CoV-2. Additionally, the re-infections with different LPH-CoVs occurred frequently in children and tended to increase the cross-reactive antibodies against SARS-CoV-2. From the forty-nine serum samples with cross-reactive anti-S IgG antibodies against SARS-CoV-2, we found that seven samples with a median age of 1.4 years old had detected neutralizing activity for the wild-type or mutant SARS-CoV-2 S pseudotypes. Interestingly, all of the seven samples contained anti-S IgG antibodies against HCoV-OC43. Together, these data suggest that children’s pre-existing antibodies to LPH-CoVs have limited cross-reactive neutralizing antibodies against SARS-CoV-2.

KEYWORDS
low pathogenic human coronaviruses, SARS-CoV-2, cross-reactive antibody, neutralizing activity, antigen specific antibody

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EDITED BY
Pedro A. Reche, Complutense University of Madrid, Spain

REVIEWED BY
Daniel Smrz, Charles University, Czechia
Stephanie Longet, University of Oxford, United Kingdom

*CORRESPONDENCE
Huanle Luo
luohle@mail.sysu.edu.cn
Yuelong Shu
shuylong@mail.sysu.edu.cn

†These authors have contributed equally to this work

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**Introduction**

Coronaviruses (CoVs) refer to a large family of viruses that cause illnesses ranging from the common cold to more severe diseases. There are seven identified coronaviruses causing human infections. The highly pathogenic human coronaviruses (HPH-CoVs) including severe acute respiratory syndrome coronavirus (SARS-CoV), middle east respiratory syndrome coronavirus (MERS-CoV), and SARS-CoV-2 belong to the betacoronaviruses, while two alphacoronaviruses (HCoV-229E, HCoV-NL63) and two betacoronaviruses (HCoV-HKU1, HCoV-OC43) are identified as low pathogenic human coronaviruses (LPH-CoVs). Prior to the huge morbidity and mortality caused by HPH-CoVs, the LPH-CoVs have long been circulating in humans and cause common cold with mild respiratory syndromes (1–5). The serological assays by detecting antibodies induced by LPH-CoVs are used to define the population’s herd immunity, and ≥90% of adults have antibody evidence against these four LPH-CoVs (6, 7). It is believed that the primary infection of LPH-CoVs commonly occurs in childhood, with repeated infection within 1-3 years and a higher infection frequency in children under 5 years old (8–11).

Though the LPH-CoVs induced antibody response is short-lasting and has limited protection from hosts infected by the same or different common cold with mild respiratory syndromes (1–5), it is hypothesized that the cross-reactive antibodies response from prior LPH-CoVs exposure could have reduced the susceptibility and possibility of developing severe clinical syndrome on SARS-CoV-2 infection in children (12, 13). However, studies exploring whether the pre-existing antibodies induced by LPH-CoVs can cross-react with SARS-CoV-2 generate conflicting results. Some data showed that the pre-existing antibodies response in uninfected populations, especially in children and teenagers exhibited specific neutralizing activity against SARS-CoV-2 (14), and high levels of pre-existing immune responses against LPH-CoVs were associated with mitigating the disease severity of coronavirus disease 2019 (COVID-19) (15–17) or reduced the duration of symptom (18). Yet, other studies suggested a lack of SARS-CoV-2 cross-neutralization activity although antigen-specific antibodies response was detected from pre-pandemic serum samples of SARS-CoV-2 (11, 19, 20). These conclusions vary greatly in different cohorts which commonly include adults, and the cross-reactive antibodies against SRAS-CoV-2 in children with pre-existing LPH-CoVs humoral immunity need to be elucidated.

Here, we investigated the seroprevalence of LPH-CoVs in 658 serum samples obtained from hospitalized children prior to the SARS-CoV-2 pandemic and measured the cross-reactive antibodies against SARS-CoV-2. We observed that 40% to 60% of the serum samples contained spike (S)-specific immunoglobulin (Ig) G antibodies for the different LPH-CoVs. Higher levels of the nucleocapsid (N)-, S-, and receptor binding domain (RBD)-specific IgG antibodies against SARS-CoV-2 were found in the LPH-CoVs exposed group, and re-infections with different LPH-CoVs appeared to increase the antigen-specific cross-reactive antibodies. However, limited neutralizing activity existed even for the samples with cross-reactive S-specific IgG antibodies against SARS-CoV-2.

**Materials methods**

**Samples**

A total of 658 pre-COVID-19 serum samples of children with respiratory infection symptoms (aged 0-15 years) collected between May 27 and December 15, 2019 were obtained from Guangzhou Women and Children’s Medical Center. 28 serum samples from SARS-CoV-2 patients with strongly neutralization activity against SARS-CoV-2 WT spike in micro-neutralization assay were obtained from Sun Yat-sen University (2020–034). The experiments were performed in compliance with and under the approval of the biomedical research ethics committee, the public health school (Shenzhen) of Sun Yat-Sen University.

**Plasmid and proteins**

The env-deficient HIV-1 (pnl4-3.luc.R.-E-) plasmid expressing the luciferase reporter was constructed in our laboratory. The pcDNA3.1.SARS-CoV-2 WT spike plasmid was kindly provided by Dr. Yaoqing Chen of Sun Yat-sen University. cDNA of SARS-CoV-2 B.1.351(Delta) spike and SARS-CoV-2 P.1(Gamma) spike were synthesized and cloned into pcDNA3.1 vector. The proteins used in this study were purchased from Sino Biological (Beijing, China).

**Enzyme-linked immunosorbent assay**

Antigen-specific IgG antibodies to SARS-CoV-2 N/S/RBD and HCoV-229E S/HCoV-NL63 S/HCoV-HKU1 S/HCoV-OC43 S were detected using a standard enzyme-linked immunosorbent assay (ELISA) (21). The optimal coating concentration of antigen and serum were 250 ng/well of SARS-CoV-2 N and S, 150 ng/well of SARS-CoV-2 RBD, 50 ng/well of HCoV-OC43 S, 30 ng/well of HCoV-229E S, HCoV-NL63 S and HCoV-HKU1 S and 1:500. A positive control (serum sample FS B26 with strongly neutralization activity in micro-neutralization assay) was kindly provided by Guangdong Provincial Center for Disease Control and Prevention (21). The optimal coating concentration of antigen and serum were 250 ng/well of SARS-CoV-2 N and S, 150 ng/well of SARS-CoV-2 RBD, 50 ng/well of HCoV-OC43 S, 30 ng/well of HCoV-229E S, HCoV-NL63 S and HCoV-HKU1 S and 1:500. A positive control (serum sample FS B26 with strongly neutralization activity in micro-neutralization assay) was kindly provided by Guangdong Provincial Center for Disease Control and Prevention (21). The optimal coating concentration of antigen and serum were 250 ng/well of SARS-CoV-2 N and S, 150 ng/well of SARS-CoV-2 RBD, 50 ng/well of HCoV-OC43 S, 30 ng/well of HCoV-229E S, HCoV-NL63 S and HCoV-HKU1 S and 1:500. A positive control (serum sample FS B26 with strongly neutralization activity in micro-neutralization assay) was kindly provided by Guangdong Provincial Center for Disease Control and Prevention (21).
the detected values. Besides, one serum from mice was used as a negative control on each plate to check the consistency of each plate.

**SARS-CoV-2 pseudovirus neutralization assay**

HEK 293T cells were co-transfected with SARS-CoV-2 Spike plasmid and HIV-1 plasmid (pNL4-3.luc.R.-E-). Cell supernatant was collected at 48 hours post-transfection and virus titers were determined as described in detail previously (22). Diluted sera were added to a 96-well plate and incubated for 1 hour at 37°C. HEK 293T-ACE2 cells were added and plates were incubated for 48 hours. The luciferase activity level was assessed by Bright-Glo Luciferase Assay System (cat# E2620) purchased from Promega (USA), and 50% inhibitory concentration (IC₅₀) was determined as the last serum dilution at which the titration curve matches inhibition equal to or above 50% of the 100% assay (21).

**Statistics**

The finite mixture model was used to define the positive value of antibodies against the S antigen of LPH-CoVs. The individuals with a positive value of cross-reactive antibody against SARS-CoV-2 were defined by an OD value greater than the mean OD values minus two times the standard deviation of COVID-19 patients. The student’s t test was used to compare the differences between the two groups. A two-tailed P value <0.05 was considered statistically significant. The differences between male group and female group for each LPH-CoVs were compared by the χ² test. Pearson’s correlation was used to test the association between the S-specific antibodies against LPH-CoVs with the cross-reactive N, S, and RBD antigen-specific antibodies against SARS-CoV-2. Statistical analysis of the clinical data was performed using SPSS Statistics version 25 software (IBM, Armonk, NY, USA). All the experimental data were analyzed in GraphPad Prism software (version 8) and R Studio software.

**Results**

**Pre-existing antibodies response against LPH-CoVs in the children cohort**

In this retrospective study, 658 serum samples collected from hospitalized children with a median age of 3 years (IQR 0.8-4.2) including 402 boys and 256 girls in 2019 before the SARS-CoV-2 outbreak were enrolled. The cohort’s pathological clinical features were summarized in Supplementary Table 1. To evaluate the pre-existing antibodies against LPH-CoVs in this cohort, we analyzed IgG antibodies response against the S antigen of HCoV-229E, HCoV-NL63, HCoV-HKU1, and HCoV-OC43 via ELISA (Figure 1A). Besides, 28 serum samples with neutralizing antibodies against SARS-CoV-2 from convalescent COVID-19 patients were used as an independent group. By utilizing the finite mixture model (Supplementary Figures 1A-D), we have observed that 40.73%, 38.30%, 39.67%, and 61.55% of the children’s serum samples were classified as positive individuals with antibodies against the S antigen of HCoV-229E, HCoV-NL63, HCoV-HKU1, and HCoV-OC43 respectively, suggesting a higher prevalence of HCoV-OC43 in hospitalized children than the other three LPH-CoVs in our cohort (Figure 1B). Interestingly, the COVID-19 patients boosted comparable S-specific IgG antibodies against these four LPH-CoVs (Figure 1B). Overall, 499 samples (75.84%) from the cohort exhibited S-specific IgG antibodies response against at least one of the four LPH-CoVs. Importantly, re-infections with different LPH-CoVs occurred commonly, we have found that 140/113/107 cases (21.28%, 17.17%, 16.26%) contained S-specific IgG antibodies against two/three/four of the LPH-CoVs respectively (Figure 1C). Among the 139 serum samples showing S-specific IgG antibodies only to one of the four LPH-CoVs, we observed positive rates of 8.05% for HCoV-229E (24 cases), 5.37% for HCoV-NL63 (16 cases), 4.03% for HCoV-HKU1 (12 cases), and 29.19% for HCoV-OC43 (87 cases) (Figure 1D). Besides, we have found a higher seroprevalence rate (20.36%) for beta LPH-CoVs compared to alpha LPH-CoVs (7.60%) in this cohort (Figure 1E). These results indicate that a high prevalence of LPH-CoVs exists in the early phase of human life, however, the pre-existing antibodies response is short-lasting to protect host from the same or other LPH-CoVs infection.

**Pre-existing antibodies response against LPH-CoVs in different age groups of the children cohort**

We next compared the seroprevalence in different age groups for positively classified serum samples with antibodies against LPH-CoVs. As shown in Figure 2A, we observed that the majority of infants less than 3 months old had S-specific IgG antibodies against HCoV-229E (66.67%), HCoV-HKU1 (52.94%), and HCoV-OC43 (70.59%), while 23.53% of the samples were seropositive individuals for HCoV-NL63, indicating a maternal transient S-specific IgG antibody in infants. Without maternal antibody, the numbers of seropositive individuals in the age category of 3 to 9 months old decreased sharply for all the four types of LPH-CoVs, then started to keep a whole upward trend following the age increase (Figure 2A; Supplementary Figures 2A-D). In the 9-15 years old group, we have found a reduced seropositive rate for HCoV-
NL63, HCoV-OC43 and HCoV-HKU1, but a raised trend for HCoV-229E compared to the 6-9 years old group (Figure 2A).

As expected, we have observed that re-infections of different coronaviruses increased with age under five years old and became stable afterward (Figure 2B).

Next, the effect of biological sex on antibody levels of S-specific IgG against LPH-CoVs was analyzed. Due to the number difference of serum samples between boys and girls, we calculated the seropositive rates for boys and girls, respectively. Interestingly, boys had a higher positive rate compared with girls for HCoV-229E infection. However, no obvious gender-related pattern was found for the other three coronaviruses (Figures 3A–D; Supplementary Table 2).

Identification of the cross-reactive antibodies against SARS-CoV-2 in the children cohort

To understand the cross-reactive antibody responses against SARS-CoV-2 induced by LPH-CoVs, we evaluated the IgG antibodies response against the N, S, and RBD antigens of SARS-CoV-2 in the serum samples using ELISA method. For the classified samples with only one of the four LPH-CoVs exposure, both HCoV-229E and HCoV-OC43 seropositive samples stimulated significantly higher levels of IgG antibodies response against the N, S, and RBD antigens of SARS-CoV-2 compared to the negative samples. The HCoV-NL63
seropositive samples contained elevated levels of IgG antibody response against the S and RBD proteins of SARS-CoV-2 rather than the N protein. Interestingly, no significant difference for the IgG antibodies response against the N, S, and RBD proteins of SARS-CoV-2 was found between the HCoV-HKU1 seropositive samples and the seronegative samples (Figure 4A). However, both alpha and beta coronaviruses seropositive samples did show enhanced IgG antibodies response against the N, S, and RBD proteins of SARS-CoV-2 compared with the negative samples (Figure 4B). As expected, we have found that the levels of IgG antibodies against the N, S, and RBD proteins of SARS-CoV-2 tended to increase with re-infections, and patients with all the four types of LPH-CoVs exposure had the highest levels of cross-reactive antigen-specific IgG antibodies against SARS-CoV-2 (Figure 4C). These results suggest that the repeated infections with LPH-CoVs may facilitate the host generating high levels of antigen-specific antibodies with cross-reactivity to SARS-CoV-2. We further determined the correlation between the S-specific IgG antibodies against LPH-CoVs and the cross-reactive antigen-specific IgG antibodies against SARS-CoV-2. For the serum samples with only one of the LPH-CoVs exposure, the positive correlations were noted between the cross-reactive antigens-specific IgG antibodies against SARS-CoV-2 with the S-specific IgG antibodies against each of the LPH-CoVs, especially with HCoV-OC43. (Figures 5A–D).

Interestingly, the cross-reactive IgG antibodies against the N and S antigens of SARS-CoV-2 peaked mostly in the individuals with an age category of 9–12 months old, while the cross-reactive IgG antibody against the RBD antigen of SARS-CoV-2 peaked mostly in children of 6–9 months old (Figures 6A–C).

**LPH-CoVs stimulate limited cross-reactive neutralizing antibodies against SARS-CoV-2**

Forty-nine individuals with a positive cross-reactive S-specific IgG antibodies response against LPH-CoVs were determined by an OD value higher than two times the average of the COVID-19 patients. To examine whether the antibodies boosted by LPH-CoVs in these samples have neutralizing activity for SARS-CoV-2, we measured the neutralizing activity utilizing wild-type (WT) and two variants of concern (VOC) including Gamma and Delta SARS-CoV-2 S pseudotypes. As shown in Figures 7A, C, we have found...
that five serum samples could neutralize WT SARS-CoV-2 though the majority of selected samples lack neutralizing activity. Out of the five serum samples, two samples could neutralize Delta SARS-CoV-2 and one contained the neutralizing activity against Gamma SARS-CoV-2, indicating an immune escape of the variants. Interestingly, only one serum sample was found having the ability of neutralizing Delta or Gamma SARS-CoV-2 respectively, but without neutralizing activity against WT SARS-CoV-2. Notably, all of the serum samples with neutralizing activity against SARS-CoV-2 were collected from individuals under 4 years old, with a median age of 1.4 years old (Figures 7B, C), indicating the individuals with younger age may be beneficial to the cross-reactive neutralizing activity against SARS-CoV-2.

Next, we found all of the seven serum samples with cross-reactive neutralization activity against SARS-CoV-2 were HCoV-OC43 seropositive (Figure 7C), suggesting the pre-existing antibody against HCoV-OC43 may play a role in reducing SARS-CoV-2 infection.

**Discussion**

The impact of pre-existing LPH-CoV-specific antibodies with cross-reactivity against SARS-CoV-2 has been investigated since the beginning of SARS-CoV-2 pandemic. It is critical if the antibodies boosted by LPH-CoVs can neutralize SARS-CoV-2 due to the 90% herd immunity of humans against LPH-CoVs (23, 24), which usually occur repeatedly in early life. In this study, we detected a generally moderate level of seroprevalence in 658 children under 15 years old for the three human coronaviruses (HCoV-229E, HCoV-NL63, HCoV-HKU1) and a high positive rate for HCoV-OC43. Although the seroprevalence of LPH-CoVs varies greatly by reason of different antigens used, methodologies adopted, population with distinct ages, and other demographic characteristics (25–27), the seroprevalence acquired from this cohort is somewhat in line with the normal range within children (28–30). Besides, the obvious maternal transient passive IgG antibodies response against LPH-CoVs was observed for infants within 3-months, which is consistent with the previous stratified studies (29, 31).

Limited studies have investigated whether biological sex affects LPH-CoVs infection although several reports have indicated that male COVID-19 patients appeared to be more susceptible to SARS-CoV-2 and generated a higher level of antibodies (21, 32). In this study, we found that HCoV-229E boosted higher S-specific IgG antibodies in boys than girls after 3 months old while no obvious sex-based antibody trend was observed for the other three LPH-CoVs. A recent study has found a higher level of IgG antibody against HCoV-229E other than the other three coronaviruses in men compared with women in an adult cohort (older than 20 years of age) (33). These data indicate that sex may be a factor affecting the prevalence of HCoV-229E.

Most adults commonly contain antibodies against all the four LPH-CoVs since re-infections occur initially in children, causing the difficulty of evaluating the cross-reactive antibody response to every specific coronavirus. In this study, we selected samples which are positive with only one type of IgG antibody against LPH-CoVs to investigate the cross-reactive antibody response against SARS-CoV-2, HCoV-HKU1 seropositive

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**FIGURE 3**
The effect of biological sex on pre-existing antibodies against LPH-CoVs in different age groups (A–D). The seropositive rates of HCoV-229E (A), HCoV–NL63 (B), HCoV-HKU1 (C) and HCoV-OC43 (D) in male and female individuals with different ages.
samples appeared to have no cross-reactive antibodies against the N, S, and RBD antigens of SARS-CoV-2. However, HCoV-OC43 seropositive samples showed significantly higher levels of, even comparable antigen-specific antibodies response against SARS-CoV-2 with the COVID-19 patients. Previous studies investigating whether the pre-existing anti-HCoV-OC43 antibodies affect SARS-CoV-2 infection yielded contradictory results. Several investigations have indicated high levels of N-specific antibodies against HCoV-OC43 in COVID-19 adult patients were associated with mild disease (34, 35), indicating a potential protective role of the pre-existing antibodies against HCoV-OC43 for SARS-CoV-2. On the other hand, Guo et al. have found significantly higher anti-S of HCoV-OC43 IgG antibody titers in patients with severe disease than those in mild patients, indicating the cross-reactive antibody may enhance the severity of COVID-19 patients (36). Unluckily, we were not able to track these patients in our cohort to assess the susceptibility or disease severity for SARS-CoV-2 infection. However, it is notable that all of the seven serum samples from children ≤ 4 years old with neutralizing activity against SARS-CoV-2 contained S-specific IgG against HCoV-OC43, indicating there may be a role for antibodies boosted by HCoV-OC43 in the earlier age during SARS-CoV-2 infection.

It is valuable that the serum samples of young individuals under 1-year-old were enrolled in this study. A prior study has reported that about 44% (21 cases in 48 samples in total) of the
individuals had detectable S-specific IgG antibodies against SARS-CoV-2 in children ranging from 1-16 years via a flow cytometry-based method, most of which could neutralize SARS-CoV-2 (14). Another study has observed that 20% of SARS-CoV-2-free individuals contained antibodies against the N, S RBD antigens of SARS-CoV-2, but with very low or undetectable neutralizing antibodies in a cohort ranging from 1-90 years. Besides, they also found age did not affect the cross-reactive antibodies (11). Consistent with this report, we have found about 24.64% of individuals with pre-existing LPH-CoVs antibodies contained the cross-reactive antigen-specific antibodies against SARS-CoV-2, most of which contained undetectable neutralizing antibodies. However, all the seven samples with neutralizing activity against SARS-CoV-2 were collected from children under 4 years old in our cohort. Notably, four samples were acquired from infants born within 1 year old in a total of eleven samples with positive S-specific antibodies against SARS-CoV-2 were assessed by Pearson correlation test. The r values were exhibited for the correlation. A two-tailed P value<0.05 was considered statistically significant.

**FIGURE 5**
Correlations between the S-specific IgG antibodies against LPH-CoVs and the cross-reactive antigens-specific IgG antibodies against SARS-CoV-2 for the serum samples with only one of the LPH-CoVs exposure (n=298) (A-D) The correlations between the S-specific IgG antibodies against HCoV-229E (A), HCoV-NL63 (B), HCoV-HKU1 (C), HCoV-OC43 (D) and the cross-reactive IgG antibodies against the N, S, RBD antigens of SARS-CoV-2 were assessed by Pearson correlation test. The r values were exhibited for the correlation. A two-tailed P value<0.05 was considered statistically significant.
FIGURE 6

The cross-reactive IgG antibodies against SARS-CoV-2 in different age groups with pre-existing antibodies against LPH-CoVs (A–C). The prevalence (line) and OD values (dots) of cross-reactive IgG antibodies against the N (A), S (B), and RBD (C) of SARS-CoV-2 in different age groups.
studies have found that vaccination of measles-mumps-rubella or tetanus-diphtheria-pertussis provided a protective role against the severe COVID-19 by activating the cross-reactive T cell response, which is a possible explanation for children with reduced susceptibility and severe clinical syndrome in SARS-CoV-2 infection (39–41). Thus, the cross-reactive T cell response should be an important point for future study since poor cross-reactive neutralizing antibodies are stimulated by LPH-CoVs. Due to the lack of COVID-19 children, the SARS-CoV-2 positive samples used here as control were collected from adult COVID-19 patients. Although comparable S-specific IgG antibodies against all the four LPH-CoVs were boosted in SARS-CoV-2 patients, we could not get a solid conclusion about the memory antibodies since the baseline of antibodies against LPH-CoVs has not been measured before they were infected with SARS-CoV-2.

Overall, our data demonstrate that the pre-existing antibody response boosted by LPH-CoVs has a moderate to high seroprevalence in children under 15 years old. However, the majority of pre-existing antibodies lack the neutralizing activity against SARS-CoV-2. Besides, HCoV-OC43 has a higher prevalence and may boost the cross-reactive neutralizing antibody in children under four years old against SARS-CoV-2, providing an insight into immunogen design and vaccine development.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.
 Ethics statement

All the experiments were performed in compliance with and under the approval of the biomedical research ethics committee, the public health school (Shenzhen) of Sun Yat-Sen University (2020-034).

Author contributions

HL and YS conceived and designed the study. NL and XL performed experiments, analyzed data, and wrote the manuscript. JW, SZ, and LZ assisted in laboratory experiments. LZ and QiC contributed to the data collection, formal analysis, and data interpretation. YF and SX collected clinical samples and data. QiC and NaW prepared experimental review materials. ZW, NaW, and CL provided experimental technical guidance. HL and YS approved the final version of the manuscript. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2022.1042406/full#supplementary-material

References

1. Jiang S, Du L. Effect of low-pathogenic human coronavirus-specific antibodies on SARS-CoV-2. Trends Immunol (2020) 41:853–4. doi: 10.1016/j.it.2020.08.003
2. Zhu Z, Lian X, Su X, Wu W, Marraro GA, Zeng Y. From SARS and MERS to COVID-19: a brief summary and comparison of severe acute respiratory infections caused by three highly pathogenic human coronaviruses. Respir Res (2020) 21:224. doi: 10.1186/s12931-020-01479-w
3. Vabret A, Mourez T, Gouarin S, Petitjean J, Freymuth F. An outbreak of coronavirus OC43 respiratory infection in Normandy, France. Clin Infect Dis (2003) 36:985–9. doi: 10.1086/374222
4. Chiu SS, Chan KH, Chu KW, Kwan SW, Guan Y, Poon LL, et al. Human coronavirus NL63 infection and other coronavirus infections in children hospitalized with acute respiratory disease in Hong Kong, China. Clin Infect Dis (2005) 40:1721–9. doi: 10.1086/430301
5. Uddin SMI, Englund JA, Kuppers JY, Chu HY, Steinhoff MC, Khatry SK, et al. Burden and risk factors for coronavirus infections in infants in rural Nepal. Clin Infect Dis (2018) 67:1507–14. doi: 10.1093/cid/ciy317
6. Goree GJ, Patel GB, Vitale IN, CYConnor TZ. Prevalence of antibodies to four human coronaviruses is lower in nasal secretions than in serum. Clin Vaccine Immunol (2010) 17:1875–80. doi: 10.1128/CVI.00278-10
7. Saizol A, Perlman S. Lessons for COVID-19 immunity from other coronavirus infections. Immunity (2020) 53:248–63. doi: 10.1016/j.immuni.2020.07.005
8. Edridge AWD, Kaczorowska J, Hoste ACR, Bakker M, Klein M, Loens K, et al. Seasonal coronavirus protective immunity is short-lasting. Nat Med (2020) 26:1691–3. doi: 10.1038/s41591-020-1083-1
9. Galanti M, Shaman J. Direct observation of repeated infections with endemic coronaviruses. J Infect Dis (2020) 223:409–15. doi: 10.1093/infdis/jiaa392
10. Aldridge RW, Lever D, Beale S, Johnson AM, Zamison M, Hayward AC, et al. Seasonality and immunity to laboratory-confirmed seasonal coronaviruses (HCoV-NL63, HCoV-OC43, and HCoV-229E): results from the flu watch cohort study. Wellcome Open Res (2020) 5:52. doi: 10.12688/wellcomeopenres.15812.2
11. Anderson EM, Goodwin EC, Verma A, Arevalo CP, Bolton MJ, Werick ME, et al. Seasonal human coronavirus antibodies are boosted upon SARS-CoV-2 infection but not associated with protection. Cell (2021) 184:1858–64. doi: 10.1016/j.cell.2021.02.010
12. Davies NG, Klepac P, Liu Y, Prem K, Ift M, Pearson CAB, et al. Age-dependent effects in the transmission and control of COVID-19 epidemics. Nat Med (2020) 26:1205–11. doi: 10.1038/s41591-020-0962-9
13. Waterlow NR, van Leeuwen E, Davies NG, Group CC-W, Flasche S, Eggo RM. How immunity from and interaction with seasonal coronaviruses can shape...
SARS-CoV-2 epidemiology. Proc Natl Acad Sci USA (2021) 118:e2108395118. doi: 10.1073/pnas.2108395118

14. Monto AS, Delonge PM, Calleap AR, Bazzi LA, Captiola SB, Malosh RE, et al. Coronavirus occurrence and transmission over 8 years in the HIVE cohort of households in Michigan. J Infect Dis (2022) 229:9–16. doi: 10.1093/infdis/jiaa161

15. Cugno M, Meroni PL, Contsondi D, Griffini S, Grovetti E, Novembrino C, et al. Effects of antibody responses to pre-existing coronaviruses on disease severity and complement activation in COVID-19 patients. Microorganisms (2022) 10(6):1191. doi: 10.3390/microorganisms10061191

16. Sagar M, Reiffer K, Rossi M, Miller NS, Sinha P, White LF, et al. Recent endemic coronavirus infection is associated with less-severe COVID-19. J Clin Invest (2021) 131(1):e145540. doi: 10.1172/JCI143380

17. Ortega N, Ribes M, Vidal M, Rubio R, Aguilar R, Williams S, et al. Seven-month kinetics of SARS-CoV-2 antibodies and role of pre-existing antibodies to human coronaviruses. Nat Commun (2021) 12:4740. doi: 10.1038/s41467-021-24979-9

18. Gouna S, Weizirk ME, Bolton MJ, Arevalo CP, Goodwin EC, Anderson EM, et al. Health care worker seromonitoring reveals complex relationships between common coronavirus antibodies and COVID-19 symptom duration. JCI Insight (2021) 6(16):el150449. doi: 10.1172/jci.insight.150449

19. Poston D, Wenzlum Y, Wise H, Templeton K, Jenks S, Hatzizisisou N, et al. Absence of severe acute respiratory syndrome coronavirus 2 neutralizing activity in prepandemic sera from individuals with recent seasonal coronavirus infection. Clin Infect Dis (2021) 73:1208–11. doi: 10.1093/cid/ciaa1803

20. Miyara M, Saichi M, Sterlin D, Anna F, Marot S, Mathian A, et al. Pre-COVID-19 immunity to common cold human coronaviruses induces a recall-type IgG response to SARS-CoV-2 antigens without cross-neutralisation. Front Immunol (2021) 13:790334. doi: 10.3389/fimmu.2021.790334

21. Lu H, Jia T, Chen J, Zeng S, Qiu Z, Wu S, et al. The characterization of disease severity associated IgG subclasses response in COVID-19 patients. Front Immunol (2021) 12:632814. doi: 10.3389/fimmu.2021.632814

22. Nie J, Li Q, Wu J, Zhao C, Hao H, Liu H, et al. Quantification of SARS-CoV-2 neutralizing antibody by a pseudotyped virus-based assay. Nat Protoc (2020) 15:3699–715. doi: 10.1038/s41596-020-0394-5

23. Ma Z, Li P, Ikram A, Pan Q. Does cross-neutralization of SARS-CoV-2 only relate to high pathogenic coronaviruses? Trends Immunol (2020) 41:851–3. doi: 10.1016/j.tit.2020.08.002

24. Abbasi J. COVID-19 and the common cold–preexisting coronavirus antibodies may hinder SARS-CoV-2 immunity. JAMA (2022) 327:609–10. doi: 10.1001/jama.2220326

25. Severance EG, Bossis J, Dickerson FB, Stallings CR, Orignon AE, Sullens A, et al. Development of a nucleocapsid-based human coronavirus immunoassay and estimates of individuals exposed to coronavirus in a U.S. metropolitan population. Clin Vaccine Immunol (2008) 15:1805–10. doi: 10.1128/CVI.00124-08

26. Mulabbi EN, Twyongyire R, Wahwire-Mangen F, Mwouro E, Koechler J, Kibuuka H, et al. Seroprevalence of human coronaviruses among patients visiting hospital-based sentinel sites in Uganda. BMC Infect Dis (2021) 21:585. doi: 10.1186/s12879-021-06258-6

27. Owusu M, Sylverken AA, El-Duah P, Acheampong G, Mutocheluh M, Adu-Sarkodie Y. Sero-epidemiology of human coronaviruses in three rural communities in Ghana. Pan Afr Med J (2021) 38:244. doi: 10.11604/pamj.2021.38.244.26110

28. Dijkman R, Jebbink MF, El Idrissi NB, Pyrc K, Müller MA, Kuijpers TW, et al. Human coronavirus NL63 and 229E are not associated with severe COVID-19 disease in children. J Clin Microbiol (2008) 46:2368–73. doi: 10.1128/JCM.00553-08

29. Shao X, Guo X, Espar F, Weibel C, Kuhn J. Seroprevalence of group I human coronaviruses in children. J Clin Virol (2007) 40:207–13. doi: 10.1016/j.jcv.2007.08.007

30. Zhou W, Wang W, Wang H, Lu R, Tan W. First infection by all four non-severe acute respiratory syndrome human coronaviruses takes place during childhood. BMC Infect Dis (2013) 13:433. doi: 10.1186/1471-2334-13-433

31. Woudenberg T, Pelleau S, Anna F, Attia M, Donnadieu F, Gravet A, et al. Humoral immunity to SARS-CoV-2 and seasonal coronaviruses in children and adults in north-eastern France. ebioMedicine (2021) 70:103495. doi: 10.1016/j.ebiom.2021.103495

32. Meng Y, Wu P, Lu W, Liu K, Ma K, Huang L, et al. Sex-specific clinical characteristics and prognosis of coronavirus disease-19 infection in wuhan, China: A retrospective study of 168 severe patients. PLoS Pathog (2020) 16(4):e1008520. doi: 10.1371/journal.ppat.1008520

33. Lin C-Y, Wolf J, Brice DC, Sun Y, Locke M, Cherry S, et al. Pre-existing humoral immunity to human common cold coronaviruses negatively impacts the protective SARS-CoV-2 antibody response. Cell Host Microbe (2022) 30:83–96.e4. doi: 10.1016/j.chom.2021.12.005

34. Dugas M, Grote-Westrick T, Merle U, Fontenay M, Kremer AE, Hames F, et al. Lack of antibodies against seasonal coronavirus OC43 nucleocapsid protein identifies patients at risk of critical COVID-19. J Clin Virol (2019) 113:104847. doi: 10.1016/j.jcv.2019.104847

35. Dugas M, Grote-Westrick T, Vollenberg R, Lorentzen E, Brix T, Schmidt H, et al. Less severe course of COVID-19 is associated with elevated levels of antibodies against seasonal human coronaviruses OC43 and HKU1 (HCoV OC43, HCoV HKU1). Int J Infect Dis (2021) 105:304–6. doi: 10.1016/j.ijid.2021.02.085

36. Guo L, Wang Y, Kang L, Hu Y, Wang L, Zhong J, et al. Cross-reactive antibody against human coronavirus OC43 spike protein correlates with disease severity in COVID-19 patients: a retrospective study. Emerg Microbes Infect (2021) 10:664–76. doi: 10.1002/2221751.20190588

37. Shwani K, Sharma R, Krishnan M, Jones T, Mayora-Neto M, Cantoni D, et al. Detection of serum cross-reactive antibodies and memory response to SARS-CoV-2 in prepandemic and post-COVID-19 convalescent samples. J Infect Dis (2022) 224:1305–15. doi: 10.1093/infdis/jiaa133

38. Ma Z, Li P, Ikram A, Pan Q. Cross-reactivity towards SARS-CoV-2: the potential role of low-pathogenic human coronaviruses. Lancet Microbe (2020) 1:e151. doi: 10.1016/S2586-5247(20)30098-7

39. Monereo-Sánchez J, Luysk J, Pinzón-Espinosa J, Richard G, Motazedzi E, Westely LT, et al. Diphtheria and tetanus vaccination history is associated with lower odds of COVID-19 hospitalization. Front Immunol (2021) 12:749264. doi: 10.3389/fimmu.2021.749264

40. Reche PA. Potential cross-reactive immunity to SARS-CoV-2 from common human pathogens and vaccines. Front Immunol (2020) 11:586984. doi: 10.3389/fimmu.2020.586984

41. Miyose V, Cullere X, Settles ML, Ji X, Kattan MW, Deaards M, et al. Protective heterologous T cell immunity in COVID-19 induced by the trivalent MMR and tdap vaccine antigens. Med (N Y) (2021) 2:1050–71.e7. doi: 10.1016/j.medj.2021.08.004