Original Article

Screening of Korean Medicinal Plant Extracts for α-Glucosidase Inhibitory Activities

Shruti Sancheti\textsuperscript{a}, Sandesh Sancheti\textsuperscript{a}, Seung-Hun Lee\textsuperscript{a}, Jae-Eun Lee\textsuperscript{a} and Sung-Yum Seo\textsuperscript{a,b,*}

\textsuperscript{a}Department of Biology, Kongju National University, Kongju 314-701, Korea. \textsuperscript{b}Korean Collection of Herbal Extract, Inc., Kongju 314-701, Korea.

Abstract

Glycosidases are the enzymes involved in various biochemical processes related to metabolic disorders and diseases. Therefore, much effort has been focused on searching novel pharmacotherapy for the treatment of these ailments from medicinal plants due to higher safety margins. To pursue these efforts, the present study was performed to evaluate the α-glucosidase inhibitory activities of thirty Korean medicinal plant extracts. Among the plants studied, \textit{Euonymus sachalinensis}, \textit{Rhododendron schleippenbachii}, \textit{Aristolochia chinensis} and \textit{Juglans regia} showed the strongest α-glucosidase inhibitory activity with IC\textsubscript{50} values of 10, 20, 30 and 80 μg/mL, respectively. In addition, \textit{Meliosma oldhamii} and \textit{Symlocos chinensis} showed moderate α-glucosidase inhibition with IC\textsubscript{50} values of 150 and 220 μg/mL, respectively. Therefore, they might prove to be a potential natural source for the treatment of metabolic ailments such as, diabetes, and need further investigations.

Keywords: α-Glucosidase inhibitor; Korean plants; Screening; Glycosidases.

Introduction

Glycosidases are widespread in microorganisms, plants, and animals. They are a very important class of enzymes, which catalyze a hydrolytic cleavage of glycosidic bonds in oligosaccharides or glycoconjugates. Among these glycosidases, α-glucosidase is able to catalyze the cleavage of glycosidic bonds involving terminal glucose connected at the site of cleavage through α-linkage at the anomeric center (1-3).

Glycosidases are involved in several important biological processes (like: digestion, biosynthesis of glycoproteins and lysosomal catabolism of glycoconjugates) related to metabolic disorders and diseases, such as, diabetes, obesity, glycosphingolipid lysosomal storage disease, HIV infections, and tumors (1-3). These observations indicate that the inhibition of glycosidases would represent a novel pharmacological approach towards the treatment of the above mentioned complications, including diabetes.

Diabetes mellitus (DM) is a metabolic disorder characterized by chronic hyperglycemia that has a significant impact on the health, quality of life and life expectancy of patients, as well as health care system (1, 4). In clinical practice, the DM treatment is restricted to the use of oral hypoglycemic agents and insulin, where the former possesses serious side effects (5). Therefore, many traditional herbal remedies for treating diabetes used throughout the world as plant drugs and herbal formulations, are frequently considered to be free from side effects and less toxic than the synthetic one (6). Thus,
Table 1. α-Glucosidase inhibitory activities and IC\textsubscript{50} values of the studied Korean plant extracts.

| Sr. No. | Plant name            | Family         | Plant part      | % Inhibition | IC\textsubscript{50} Values (µg/mL)\textsuperscript{*} |
|---------|-----------------------|----------------|-----------------|--------------|------------------------------------------------------|
| 1       | Actinidia arguta      | Actinidiaceae  | Whole plant     | 33 ± 1       | --                                                   |
| 2       | Meliosma oldhamii     | Sabiaceae      | Whole plant     | 67 ± 3       | 150 ± 3                                              |
| 3       | Aster tataricus       | Asteraceae     | Stem            | n. a.        | --                                                   |
| 4       | Capsella bursa-pastoris | Brassicaceae | Whole plant     | 23 ± 1       | --                                                   |
| 5       | Allium macrostemon    | Alliaceae      | Whole plant     | 19 ± 2       | --                                                   |
| 6       | Rhododendron schlippenbachii | Ericaceae | Leaf            | 98 ± 1       | 20 ± 1                                               |
| 7       | Pyrola japonica       | Pyrolaceae     | Whole plant     | 29 ± 2       | --                                                   |
| 8       | Symlocos chinensis    | Symlocaceae    | Leaf            | 52 ± 2       | 220 ± 5                                              |
| 9       | Juglans regia         | Juglandaceae   | Whole plant     | 99 ± 1       | 80 ± 2                                               |
| 10      | Sinapis alba          | Brassicaceae   | Seed            | 22 ± 2       | --                                                   |
| 11      | Aster tataricus       | Asteraceae     | Root            | 17 ± 3       | --                                                   |
| 12      | Magnolia kobus        | Magnoliaceae   | Whole plant     | 4 ± 1        | --                                                   |
| 13      | Inula helenium        | Asteraceae     | Whole plant     | 9 ± 2        | --                                                   |
| 14      | Digitaria violascens  | Poaceae        | Whole plant     | n. a.        | --                                                   |
| 15      | Dendropanax morifera  | Araliaceae     | Whole plant     | 39 ± 3       | --                                                   |
| 16      | Sesamum indicum       | Pedaliaceae    | Whole plant     | 11 ± 2       | --                                                   |
| 17      | Alisma canaliculatum  | Alismataceae   | Rhizomes        | 22 ± 3       | --                                                   |
| 18      | Celtis sinensis       | Cannabaceae    | Whole plant     | 25 ± 1       | --                                                   |
| 19      | Astilbe chinensis     | Saxifragaceae  | Rhizomes        | 95 ± 2       | 30 ± 2                                               |
| 20      | Corydalis remota      | Papaveraceae   | Whole plant     | 23 ± 2       | --                                                   |
| 21      | Phlomis umbrosa       | Labiatae       | Whole plant     | 12 ± 2       | --                                                   |
| 22      | Curcuma zedoaria      | Zingiberaceae  | Whole plant     | 27 ± 2       | --                                                   |
| 23      | Gleditsia japonica    | Fabaceae       | Whole plant     | n. a.        | --                                                   |
| 24      | Miscanthus sinensis   | Poaceae        | Whole plant     | 30 ± 3       | --                                                   |
| 25      | Heracleum moellendorffii | Apiaceae   | Whole plant     | 17 ± 2       | --                                                   |
| 26      | Draba nemorosa        | Brassicaceae   | Whole plant     | 11 ± 1       | --                                                   |
| 27      | Vaccinium hirtum      | Ericaceae      | Whole plant     | 46 ± 1       | --                                                   |
| 28      | Smilax sieboldii      | Smilaceae      | Whole plant     | 24 ± 3       | --                                                   |
| 29      | Euonymus sachalinensis | Celastraceae | Leaf            | 99 ± 1       | 10 ± 1                                               |
| 30      | Sinomenium acutum     | Menispermaceae | Bark            | 28 ± 2       | --                                                   |

Note: The IC\textsubscript{50} values of 1,2,3,4,6-penta-O-galloyl-β-D-glucose (positive control) for α-glucosidase inhibitory activity was measured as 0.37 ± 0.03 µg/mL, respectively.
\textsuperscript{*} All results are represented as mean ± SD (n = 3).
n. a.: no activity.
--: Not determined (IC\textsubscript{50} values were only determined for the plant extracts with ≥ 50% inhibition at 5 mg/mL).

Many plants and crude drugs have recently been tested for their effects on α-glucosidase inhibition.

To pursue the findings, in this research, we screened thirty Korean medicinal plants. The details of the plants’ scientific names and families are listed in Table 1. A literature survey did not show any reference to a previous work on the...
Experimental

Plant material

The dried and matured plant parts of thirty Korean medicinal herbs were obtained from “Korean Collection of Herbal Extracts” a Biotech company in Korea. A collection of voucher specimen is available with the company (Korea Collection of Herbal Extracts, 2000).

Extraction

The dried plant parts individually were chopped into small pieces and pulverized into a fine powder. The powdered plant materials (100 g, dry weight) were kept for extensive decoction in 80% methanol for 3 days at room temperature. The extracts were then concentrated using rotary vacuum evaporator at 20-30°C to obtain the dried crude extracts.

Reagents

α-Glucosidase (from Saccharomyces cerevisiae type I) and 4-nitrophenyl α-D-glucopyranoside were purchased from Sigma-Aldrich (St. Louis, MO, USA). Other commercially available reagents and solvents were used as received.

α-Glucosidase assay

The enzyme inhibition activity for α-glucosidase was evaluated according to the method previously reported by Shibano et al. (7) with minor modifications. The reaction mixture consisted of 50 μL of 0.1 M phosphate buffer (with pH of 7.0), 25 μL of 0.5 mM 4-nitrophenyl α-D-glucopyranoside (dissolved in 0.1 M phosphate buffer, with pH of 7.0), 10 μL of test sample and 25 μL of α-glucosidase solution (a stock solution of 1 mg/mL in 0.01 M phosphate buffer, with pH of 7.0 was diluted to 0.1 Unit/mL with the same buffer, with pH of 7.0 just before assay). This reaction mixture was then incubated at 37°C for 30 min. Then, the reaction was terminated by the addition of 100 μL of 0.2 M sodium carbonate solution. The enzymatic hydrolysis of substrate was monitored by the amount of p-nitrophenol released in the reaction mixture at 410 nm using microplate reader. Individual blanks were prepared for correcting the background absorbance, where the enzymes were replaced with buffer. Controls were conducted in an identical manner replacing the plant extracts with methanol. 1, 2, 3, 4, 6-penta-O-galloyl-β-D-glucose was used as positive control. All experiments were carried out in triplicates. The inhibition percentage of α-glucosidase was assessed by the following formula:

\[
\% \text{ α-glucosidase}\% = 100 \times \left( \frac{\Delta A_{\text{Control}} - \Delta A_{\text{Sample}}}{\Delta A_{\text{Control}}} \right)
\]

\[
\Delta A_{\text{Control}} = \Delta A_{\text{Test}} - \Delta A_{\text{Blank}}
\]

\[
\Delta A_{\text{Sample}} = \Delta A_{\text{Test}} - \Delta A_{\text{Blank}}
\]

Statistical analyses

All assays were performed at least three times with triplicate samples. All results are expressed as mean ± SD. IC50 values were only determined for the plant extracts with inhibition ≥ 50% at 5 mg/mL by plotting a percent inhibition versus concentration curve, in which the concentration of sample required for 50% inhibition was determined and expressed as IC50 value.

Results and Discussion

Plants have always been used as an exemplary source of drugs and many of the currently available drugs have been directly or indirectly derived from them (8). Many herbal extracts are being used in the preparation of advanced remedies for diabetes, in which α-glucosidase inhibitors play an important role by controlling postprandial blood glucose levels by means of retarding uptake of dietary carbohydrates (9). Therefore, in search of such potent α-glucosidase inhibitors from natural source, in the present study thirty Korean medicinal plant extracts have been evaluated for their α-glucosidase inhibitory activity and compared with 1, 2, 3, 4, 6-penta-O-galloyl-β-D-glucose as a positive control.

In this study, the α-glucosidase inhibitory activity of the thirty plant extracts was evaluated at 5 mg/mL concentration at the preliminary
level and the percentage inhibitions are shown in Table 1.

The present data revealed that, six plant extracts demonstrated \( \alpha \)-glucosidase inhibition \( \geq \) 50\%, namely, *Euonymus sachalinensis*, *Rhododendron schlippenbachii*, *Aristolb chinensis*, *Juglans regia*, *Meliosma oldhamii* and *Symplocos chinensis* with \( IC_{50} \) values of \( 10 \pm 1, 20 \pm 1, 30 \pm 2, 80 \pm 2, 150 \pm 3 \) and \( 220 \pm 5 \mu \text{g/mL} \), respectively. The traditional uses of these plants are listed in Table 2. These active plants have no documentary evidence in the literature for their \( \alpha \)-glucosidase inhibitory potency.

In addition, in the present screening, eight plant extracts showed medium activity, ranging from 25 to 49\%, 13 plant extracts showed less than 25\% inhibition, and 3 plant extracts did not exhibit \( \alpha \)-glucosidase inhibitory activity (Table 1).

The \( \alpha \)-glucosidase inhibitory potential of the identified potent crude extracts was lower as compared to PGG (Table 1), since crude extracts contain non-active components along with the active ones. Therefore, the crude extracts of *Euonymus sachalinensis*, *Rhododendron schlippenbachii*, *Aristolb chinensis*, *Juglans regia*, *Meliosma oldhamii* and *Symplocos chinensis* seem to be relatively potent inhibitors, where the inhibitory activity could further be improved by separation and purification of the active components.

Thus, introduction of such innovative herbal extracts for the treatment of diabetes and other related metabolic disorders, where the \( \alpha \)-glycosidase inhibition plays a key role, may prove fortune. However, further *in-vivo* studies needed to be confirmed to provide strong biochemical rationale.

**Acknowledgement**

This research was financially supported by the Grant of the Korean Ministry of Education, science and Technology ( The Regional core Research Program/ Zero Energy Green village Technology center.

**References**

1. Sancheti S, Sancheti S and Soo SY. *Chaenomeles sinensis*: A potent \( \alpha \)-and \( \beta \)-glucosidase inhibitor. *Am. J. Pharm. Toxicol*. (2009) 4: 8-11.
2. De Melo EB, da Silveira Gomes A and Carvalho I. \( \alpha \)-and \( \beta \)-glucosidase inhibitors: chemical structure and biological activity. *Tetrahedron* (2006) 62: 10277-10302.
3. Kim JH, Ryu YB, Kang NS, Lee BW, Heo JS, Jeong IY and Park KH. Glycosidase inhibitory flavonoids from *Sophora flavescens*. *Biol. Pharm. Bull*. (2006) 29: 302-305.
4. Nickavar B and Yousefian N. Inhibitory effects of six *Allium* species on \( \alpha \)-amylase enzyme activity. *Iranian J. Pharm. Res*. (2009) 8: 53-57.
5. Ghosh T, Maiti TK, Sengupta P, Dash DK and Bose A. Antidiabetic and *in-vivo* antioxidant activity of ethanolic extract of *Bacopa monnieri* Limn. aerial parts: a possible mechanism of action. *Iranian J. Pharm. Res*. (2008) 7: 61-68.
6. Sivajothi V, Dey A, Jayakar B and Rajakpoor B. Antihyperglycemic, antihyperlipidemic and antioxidant effect of *Phyllanthus rheedia* on streptozotocin induced diabetic rats. *Iranian J. Pharm. Res*. (2008) 7: 53-59.
7. Shibano M., Kitagawa S, Nakamura S, Akazawa N and Kusano G. Studies on the constituents of *Broussonetia* species. II. Six new pyrrolidine alkaloids, broussonetine A, B, E, F and broussonetinone A and B, as inhibitors of glycosidases from *Broussonetia kazinoki* Sieb. *Chem. Pharm. Bull*. (1997) 45: 700-705.
8. Grover JK, Yadav S and Vats V. Medicinal plants of India with anti-diabetic potential. *J. Ethnopharmacol*. (2002) 81: 81-100.
9. Dwek RA, Butters TD, Platt FM and Zittmann N. Targeting glycosylation as a therapeutic approach. *Nat. Rev. Drug Discov*. (2002) 1: 65-75.