Acquired Resistance of *Mycobacterium tuberculosis* to Bedaquiline

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**Abstract**

Bedaquiline (BDQ), an ATP synthase inhibitor, is the first drug to be approved for treatment of multi-drug resistant tuberculosis in decades. *In vitro* resistance to BDQ was previously shown to be due to target-based mutations. Here we report that non-target based resistance to BDQ, and cross-resistance to clofazimine (CFZ), is due to mutations in *Rv0678*, a transcriptional repressor of the genes encoding the MmpS5-MmpL5 efflux pump. Efflux-based resistance was identified in paired isolates from patients treated with BDQ, as well as in mice, in which it was confirmed to decrease bactericidal efficacy. The efflux inhibitors verapamil and reserpine decreased the minimum inhibitory concentrations of BDQ and CFZ *in vitro*, but verapamil failed to increase the bactericidal effect of BDQ in mice and was unable to reverse efflux-based resistance in vivo. Cross-resistance between BDQ and CFZ may have important clinical implications.

**Introduction**

BDQ (Sirturo, TMC207) was recently granted accelerated approval as part of combination therapy to treat adults with multi-drug resistant pulmonary tuberculosis (TB) when an effective treatment regimen cannot otherwise be provided [1]. Its mechanism of action, inhibition of ATP synthase, was discovered through whole genome sequencing of resistant mutants that were selected *in vitro* [2]. As BDQ will now be used in clinical practice, it is important to understand any additional, non-target based mechanism(s) that lead to decreased sensitivity or resistance, so that appropriate drug susceptibility testing methods can be developed.

Several target-based resistance mutations in the *atpE* gene have been described previously [2–4], but when a Luria-Delbruck fluctuation assay was used to generate a larger set of BDQ-resistant mutants, target-based mutations were identified in only 30% (15/53) of the isolates, suggesting additional mechanism(s) of resistance [3]. Although the mutation rates observed in the above study suggested a genotypic rather than a phenotypic mechanism of resistance, whole genome sequencing of two non-*atpE* mutants initially did not reveal genomic changes.

Here we selected *Mycobacterium tuberculosis* isolates with increased minimal inhibitory concentrations (MICs) for BDQ *in vitro*, in mice and in patients, and studied these for non-target based mechanisms of resistance. After identifying the efflux system MmpS5-MmpL5 as being responsible for non-target based resistance, we studied the effect of efflux pump inhibitors (EPIs) on BDQ’s *in vitro* MICs and *in vivo* efficacy in mice. Hartkoorn *et al.* recently described the *in vitro* isolation of CFZ-resistant strains, with cross-resistance to BDQ, by up-regulation of the MmpS5-MmpL5 efflux system [5]. Both CFZ and BDQ can therefore select for efflux-based resistance, leading to cross-resistance between the two drugs.

**Results**

Non-target based resistance to BDQ is linked to mutations in *Rv0678*

Non-*atpE* mutant strains EH 3.2 and EH3.6 were re-analyzed by 454 sequencing for the presence of mutations and/or insertions. In non-*atpE* mutant strain EH 3.2, a non-synonymous single nucleotide polymorphism was observed in *Rv0678* gene (A202G leading to S68G), while in strain EH 3.6, a non-synonymous single nucleotide polymorphism was observed in *ppsC* gene (G5408T leading to G1803V). The *ppsC* gene is involved in phenolphthiocerol and phthiocerol dimycocerosate biosynthesis [6] and has not been linked to antibiotic resistance. While *Rv0678* gene has been implicated in decreased sensitivity to azoles [7] and an investigational compound with antitubercular activity [8], a focused analysis and
search for variants in the candidate gene (Rv0678 from mutant EH3.2) in mutant EH3.6 revealed an additional inconsistency in the read assembly, reflecting a large insertion at position 272 in Rv0678 gene. Although the full length sequence of this insertion could not be revealed by the 454 sequencing analysis, the beginning and end of the nucleotide sequence of this insert suggested an insertion of mobile element IS6110, resulting in a disruption at the protein level from amino acid 91 onwards. These findings suggested that in both non-\(\text{atpE}\) mutant strains Rv0678 was a candidate gene responsible for non-target based resistance to BDQ.

To confirm the results of 454 sequencing, a fragment containing Rv0678 gene and the region 143 bp upstream was Sanger sequenced (Fig. 1). One additional in vitro isolate (EH3.3) and 3 isolates from mice that had been treated with combinations including BDQ, CFZ, levofloxacin and amikacin (BCLA2, BCA4 and BCA8) were also included. All additional isolates had genomic changes in Rv0678, either a missense mutation (in EH 3.2, BCLA2, BCA4 and BCA8), or a single nucleotide insertion (EH 3.5) or an IS6110 insertion (EH 3.6) (Fig. 1, Table 1).

The clinical relevance of mutations in Rv0678 gene was studied by sequencing 6 paired samples (baseline and post-baseline) with increased post-baseline MICs for BDQ (2 to 16 fold, see below) from a clinical trial (NCT00540449), in which MDR-TB patients were treated with BDQ and a background MDR-TB regimen. None of the post-baseline isolates had mutations in the ATP synthase operon, while all 6 did have genomic changes in the Rv0678 gene (Table 1). Three isolates had a missense mutation, and three had changes leading to the disruption of the Rv0678 protein: a single nucleotide deletion, a single nucleotide insertion (both resulting in a frameshift) and a transposon IS6110 insertion in position 172 of Rv0678 gene. In some cases, mixed populations of parent and mutant strains were observed, due to the fact that DNA was extracted directly from sputum cultures.

**Cross resistance with CFZ and effect on efflux pump inhibitors on MICS**

MICs of BDQ and CFZ of all preclinical and clinical strains, in the absence and presence of the EPIs verapamil and/or reserpine were determined (Tables 2 and 3).

In preclinical non-\(\text{atpE}\) mutants (Tables 1 and 3), mutations resulting in a disruption of the protein Rv0678 correlated with the higher BDQ MIC increases (8-fold), whereas the effects of the single amino acid changes were more moderate (4-8 fold). Rv0678 mutants were cross-resistant to CFZ and MIC increases for BDQ and CFZ were very similar. The EPIs verapamil and reserpine reduced the MICs of both BDQ and CFZ, but this effect decreased in mutants with higher initial MICs. The effect of these EPIs was therefore more pronounced for the wild type strains H37Rv and EH3.0. The EPIs had minor effects on the BDQ MICs of the preclinical \(\text{atpE}\) mutants BK12 and LV13. Isolate LV13 carries a double mutation (Rv0678 and atpE) and is therefore cross-resistant to CFZ.

For clinical isolates (Table 2), the increases in MIC ranged from 2-fold to 16-fold, depending on the mutation. Isolates with 8 and 16-fold MIC increases had a disrupted Rv0678 gene. Rv0678 mutants were cross-resistant to CFZ and MIC increases for BDQ and CFZ were again very similar. The EPI reserpine used at 3 \(\mu g/ml\) reduced the MICs of both BDQ and CFZ, but efflux pumps were less susceptible to reserpine when the MIC levels were high, resulting in smaller MIC decreases. Concentrations of reserpine greater than 3 \(\mu g/ml\) inhibited the growth of some clinical isolates. Some clinical strains were also tested with verapamil with similar MIC decreases for BDQ and CFZ (results not shown).

**Rv0678 mutations lead to upregulation of MmpSS-MmpL5 protein expression**

We performed whole mycobacterial quantitative proteome analysis comparing mutants EH3.2 and EH3.6 to the wild-type isogenic strain, EH 3.0. Using iTRAQ-labeling and a mass spectrometric (LC-MS/MS) approach, we identified from each sample approximately 2000 proteins of which 1453 could be quantified in terms of relative protein expression (Table S1). A small subset of 25 proteins was found to be deregulated in mutants versus wild-type strain. We regarded proteins showing an increase or decrease with a fold change >1.3 as deregulated. The top upregulated and downregulated proteins are shown in Table 4 and Table S2, respectively. In both EH3.2 and EH3.6 mutant strains, the highest upregulated protein identified was MmpSS (mycobacterial membrane protein small, Rv0677c), an outer membrane protein known to be involved in multi-drug transport [7]. MmpSS

| Table 1. Mutations in Rv0678 gene and MICs of BDQ of resistant preclinical strains. |
|----------------------------------------------------------|
| **Mutation in Rv0678** | **Strain** | **Description** | **DNA** | **Protein** | **BDQ MIC (\(\mu g/ml\))** |
|-------------------------|-----------|----------------|----------|-------------|--------------------------|
| H37Rv                   | wild type | M. tuberculosis strain | wt       | wt          | 0.063                    |
| BCLA 2                  | BDQ-R     | mutant, H37Rv-derived, no \(\text{atpE}\) mutations | A413G    | E138G       | 0.250 (\(<4\))           |
| BCA 4                   | BDQ-R     | mutant, H37Rv-derived, no \(\text{atpE}\) mutations | G281A    | R94Q        | 0.500 (\(<8\))           |
| BCA 8                   | BDQ-R     | mutant, H37Rv-derived, no \(\text{atpE}\) mutations | G281A    | R94Q        | 0.500 (\(<8\))           |
| BK12                    | BDQ-R     | mutant, H37Rv-derived, \(\text{atpE}\) mutation \([2]\) | no mutation | no mutation | 2.000 (\(<32\))          |
| LV13                    | BDQ-R     | mutant, H37Rv-derived, \(\text{atpE}\) mutation \([24]\) | Ins G 192–193 frameshift | 4.000 (\(<64\))        |
| EH 3.0                  | MDR       | M. tuberculosis strain \([3]\) | wt       | wt          | 0.125                    |
| EH 3.2                  | BDQ-R     | mutant, EH 3.0-derived, no \(\text{atpE}\) mutations \([3]\) | A202G    | S66G        | 0.500 (\(<4\))           |
| EH 3.6                  | BDQ-R     | mutant, EH 3.0-derived, no \(\text{atpE}\) mutations \([3]\) | IS6110 nt 272 disruption | 1.000 (\(<8\))         |
| EH 3.3                  | BDQ-R     | mutant, EH 3.0-derived, no \(\text{atpE}\) mutations \([3]\) | Ins A 38–39 frameshift | 1.000 (\(<8\))         |

The MICs of BDQ-R preclinical strains derived from either the drug susceptible H37Rv or the MDR EH3.0 parent strains are shown. Fold-changes between brackets represent the difference between resistant and wild-type MICs. \(\text{wt}\): wild-type; \(\text{Ins}\): insertion; \(\text{nt}\): nucleotide; BDQ: bedaquiline; BDQ-R: bedaquiline-resistant.
Figure 1. Mutations in *Rv0678* gene of *M. tuberculosis* BDQ resistant strains. A. PCR fragment amplified for sequencing and mapping of mutations in *Rv0678* gene of BDQ resistant strains in (B) H37Rv-derived mutants and (C) EH 3.0-derived mutants. Codons START and STOP of *Rv0678* gene are underlined. The nucleotide positions are indicated on top of each mutation. Mutations are bold and colored: missense mutations are
expression was more than 18-fold and 4-fold upregulated in mutant strains EH3.6 and EH3.2 respectively, as compared to parent strain EH3.0. MmpS5 along with MmpL5, a transmembrane channel, forms the MmpS5-MmpL5 efflux pump complex, belonging to Resistance Nodulation Cell Division (RND) family of mycobacterial transporters [9]. Four out of the 13 mmp genes of M. tuberculosis are transcriptionally coupled with mmpS genes, and usually function as a complex to transport out different substrates. In both mutant strains, strong upregulation of MmpL5 protein was observed, about 6- and 2-fold in EH3.6 and EH3.2 respectively (Table 4). Other prominent differentially regulated proteins were Rs0559c and Rs0455c, whose function is not known as yet and belong to conserved hypothetical proteins family. Rs0559c gene is known to be in operon with Rs0560c, which was not found among the quantifiable proteins in our dataset.

The M. tuberculosis genome contains several efflux pump genes belonging to 5 families of transporters namely, ATP-Binding Cassette (ABC) superfamily, Major Facilitator Superfamily (MFS), RND family, Multidrug and Toxic Compound Extrusion (MATE) family and Small Multidrug Resistance (SMR) family. We further analyzed the expression of other efflux pump proteins that could be quantified in our whole mycobacteria proteome dataset. From this dataset, we could identify 13 proteins belonging to different efflux pump families (Table 5) and the only other protein which was slightly overexpressed in the mutant strains was DrrA (Daunorubicin-dim-transport resistance ATP-binding protein), an ABC transporter probably involved in daunorubicin resistance and phthiocerol dimycocerosate transport [6].

Overexpression strain of MmpS5-MmpL5 has increased BDQ and CFZ MICs

In order to confirm that overexpression of the MmpS5-MmpL5 efflux pump confers resistance to BDQ and CFZ, 3 strains overexpressing either MmpS5, MmpL5 or MmpS5-MmpL5 were generated. For this, plasmids derived from the replicative vector pSD5 [10] (Fig. S2), containing the corresponding genes under control of the strong promoter P_hsp65 were transformed in the wild type strain H37Rv. The MICs of BDQ and CFZ of MmpS5-MmpL5 overexpression strain were increased by 4-fold and 2-fold respectively when compared to the control strain H37Rv pSD5 (Table 6). In the presence of verapamil, the MIC levels of BDQ and CFZ decreased by 4-fold and 2-fold respectively. When MmpS5 and MmpL5 were overexpressed separately, no increase in the MIC was observed (Table 6). These results suggest that BDQ and CFZ are substrates of this efflux pump, and that the entire operon MmpS5-MmpL5 must be overexpressed in order to transport these drugs.

No fitness cost associated with mutations in Rv0678 gene

To determine lung CFUs. No differences in growth were observed (Fig. S1B).

Efflux-based resistance leads to a translational effect which cannot be rescued by verapamil

To assess the effect of increased MICs on the efficacy of BDQ, groups of 6 mice were infected IV with either wild-type H37Rv or the BDQ resistant (mouse) isolates BCLA2, BCA4 and BCA8. Treatment started at 12 days post infection with 6.25 mg/kg or 50 mg/kg of BDQ, in the absence or presence of verapamil dosed at 25 mg/kg. After 4 weeks of treatment, mice were sacrificed and lung CFU titers determined (Fig. 2). Although BDQ still reduced CFU counts after 4 weeks, all mutants were clearly less susceptible to the bactericidal activity of BDQ compared to the wild type strain H37Rv. The resistance of the least resistant strain BCLA2 (MIC increase 3-fold) could be overcome by increasing the BDQ dose 9-fold (from 6.25 mg/kg to 50 mg/kg), but could not completely overcome resistance of strains BCA4 and BCA8 (MIC increase 9-fold). Adding verapamil to BDQ slightly increased the bactericidal effect after 4 weeks in several groups of mice, but verapamil adjunctive therapy was never significantly better than BDQ monotherapy (p > 0.05).

Discussion

During our search for alternative mechanisms of resistance to BDQ, we observed that non-atpE mutants had reduced MICs when tested in the presence of the EPIs reserpinie and verapamil. While this observation suggested that efflux could be involved in emergence of resistance, it was subsequently observed by ourselves and others [11] that MICs of wild-type, non-resistant strains were also significantly reduced when tested in the presence of EPIs: MIC levels for BDQ and CFZ indeed decrease 16-fold in the presence of verapamil [11] (Table 3). An efflux-based mechanism is therefore implied in intrinsic resistance to BDQ, in addition to a possible role in acquired resistance.

A detailed re-analysis of non-atpE mutants led to the identification of genomic changes in Rv0678, initially in 2 strains with increased MICs, isolated in vitro. We subsequently identified mutations in Rv0678 of all 10 additional strains investigated, including 6 from patients treated with BDQ in a clinical trial.

Drug efflux has recently been highlighted as an important acquired resistance mechanism in M. tuberculosis [12]. Mutations or/and insertions in Rv0678 have previously been described to contribute to resistance to antifungal azoles [7], to an oxazole with antituberculosis activity [8] and to CFZ [5]. The Rv0678 gene and the mmpS5-mmpL5 operon are located in opposite orientations in the genome (Fig. 3). The respective-10 consensus boxes of their promoters are placed in a palindromic region where the protein Rv0678 hypothetically binds, preventing transcription of both the Rv0678 gene and the mmpS5-mmpL5 operon [7]. Since Rv0678 is a negative regulator of MmpS5 and MmpL5, mutations in this gene lead to an increased expression of this efflux pump [7]. All non-atpE mutants resistant to BDQ carried a different mutation in Rv0678, regulating, including missense mutations, single nucleotide insertions, deletions, and IS6110 insertion sequences. Acquisition of resistance due to IS6110 insertion was described recently [9], but it has not been reported to disrupt the Rv0678 gene. Different mutations had different effects on the MICs, presumably because
Table 2. MICs of BDQ and CFZ of clinical isolates, in the absence and presence of reserpine.

| Patient | Isolate | DNA     | protein | Mutation in Rv0678 | MICs (µg/ml) | MICS (µg/ml) | MICS (µg/ml) |
|---------|---------|---------|---------|-------------------|-------------|-------------|-------------|
|         |         |         |         |                   | BDQ         | BDQ+RES     | CFZ         |
|         |         |         |         |                   |             |             |             |
|         |         |         |         |                   |             |             |             |
| Patient A | Baseline | wt       | wt       |                   | 0.125       | 0.016 (:4)  | 1.000       |
|          | Post-baseline | T124C    | W42R     |                   | 0.250 (:2)  | 0.125 (:2)  | 2.000 (2)   |
| Patient B | Baseline | wt       | wt       |                   | 0.125       | 0.031 (:4)  | 0.500       |
|          | Post-baseline | A97G     | T33A     |                   | 0.500 (:4)  | 0.250 (:2)  | 2.000 (:4)  |
| Patient C | Baseline | wt       | wt       |                   | 0.063       | <=0.002 (:>32) | 0.500       |
|          | Post-baseline | C107T    | A36V     |                   | 0.500 (:8)  | 0.125 (:4)  | 4.000 (:8)  |
| Patient D | Baseline | wt       | wt       |                   | 0.063       | 0.016 (:4)  | 0.500       |
|          | Post-baseline | Del C 212 | frameshift |                   | 0.500 (:8)  | 0.250 (:2)  | 1.000 (:2)  |
| Patient E | Baseline | wt       | wt       |                   | 0.063       | 0.008 (:8)  | 0.250       |
|          | Post-baseline | Ins I56110 nt 172 | frameshift |                   | 0.500 (:8)  | 0.031 (:16) | 1.000 (:4)  |
| Patient F | Baseline | wt       | wt       |                   | 0.016       | 0.008 (:2)  | 0.500       |
|          | Post-baseline | Ins C 141–142 | frameshift |                   | 0.250 (:16) | 0.125 (:2)  | 4.000 (:8)  |

MICs and mutations in Rv0678 gene are shown for baseline and post-baseline isolates of each patient. For MICs without reserpine, fold-changes between brackets represent the difference between resistant and wild-type MICs. For MICs in the presence of reserpine (3 µg/ml), fold-changes between brackets represent the differences between MIC with and without reserpine. wt: wild-type; Ins: insertion; Del: deletion; nt: nucleotide; BDQ: bedaquiline; CFZ: clofazimine; RES: reserpine.

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they result in different conformational changes of Rv0678 protein, or in its complete disruption. Frameshift mutations led to the highest increases in MICs (3- or 16-fold), compared to amino acid changes (2 to 8-fold). Proteome data showed that the mutant EH 3.2 (carrying an amino acid substitution) and EH 3.6 (which has an IS6110 insertion) have MmpS5 and MmpL5 overexpressed. For these strains, the MICs are similar, but the effect of EPIs was less pronounced for the mutant with the disrupted regulator (EH 3.6) and the largest overexpression of MmpS5-MmpL5.

Although we cannot entirely exclude the possibility that additional efflux pumps may be involved in non-target based acquired resistance to BDQ, the fact that mutations in Rv0678 gene were found in all investigated isolates, together with the proteome data demonstrating overexpression of MmpS5-MmpL5, support the conclusion that the MmpS5-MmpL5 efflux pump is responsible for non-target based acquired resistance. Moreover, the increase in BDQ MIC of the overexpression strain H37Rv pCSCVA30 suggests that BDQ is a substrate of MmpL5. This RND pump may also be responsible for the observed intrinsic resistance to BDQ as described above. CFZ is apparently also a substrate of the same efflux pump, as all non-aptE strains with increased MICs for BDQ were cross-resistant to CFZ, and the effects of EPIs on the MICs of CFZ and BDQ were very similar.

As efflux inhibition may theoretically increase the bactericidal and sterilizing efficacy of existing drugs in a regimen, the addition of EPIs to TB regimens has the potential to enhance mycobacterial killing in vivo [11,13]. Mouse efficacy data obtained with rifampicin indeed suggest increased bactericidal and sterilizing efficacy of a regimen including verapamil [13]. The excellent potentiating effect of verapamil on the bacteriostatic effect [MIC] of BDQ was very promising, and raised the expectation that the combination of BDQ with verapamil would also result in a significantly better bactericidal effect in vivo. In this context, the results shown in figure 2 are disappointing and the reasons for this minor effect on killing in mice need to be further investigated. Different expression levels of efflux pumps in vivo versus in vitro, a different mechanism of bacteriostatic [MIC] and bactericidal (in mice) effects of BDQ and drug-drug interactions may have contributed to the lack of in vivo efficacy of the combination. Gupta et al. described accelerated bactericidal activity by addition of verapamil in one

| Table 3. MICs of BDQ and CFZ of preclinical strains with EPIs. |
|---|---|---|---|---|---|---|
| **Strain** | **BDQ (µg/ml)** | **BDQ+VER** | **BDQ+RES** | **CFZ (µg/ml)** | **CFZ+VER** | **CFZ+RES** |
| H37Rv | 0.063 | 0.004 (±16) | 0.016 (±4) | 0.250 | 0.031 (±8) | 0.125 (±2) |
| BCLA 2 | 0.250 (±4) | 0.031 (±8) | 0.125 (±2) | 0.500 (±2) | 0.125 (±4) | 0.500 (±4) |
| BCA 4 | 0.500 (±8) | 0.125 (±4) | 0.250 (±2) | 1.000 (±4) | 0.500 (±2) | 1.000 (±2) |
| BK12 | 2.000 (±32) | 1.000 (±2) | 2.000 (±4) | 0.250 (±2) | 0.031 (±8) | 0.063 (±4) |
| LVI3 | 4.000 (±64) | 2.000 (±2) | 4.000 (±4) | 1.000 (±4) | 0.500 (±2) | 1.000 (±2) |
| EH 3.0 | 0.125 | 0.008 (±16) | 0.016 (±8) | 0.250 | 0.016 (±16) | 0.031 (±8) |
| EH 3.2 | 0.500 (±4) | 0.016 (±32) | 0.250 (±2) | 1.000 (±4) | 0.016 (±64) | 0.125 (±8) |
| EH 3.6 | 1.000 (±8) | 0.125 (±4) | 0.500 (±2) | 2.000 (±8) | 1.000 (±2) | 1.000 (±2) |
| EH 3.3 | 1.000 (±8) | 0.250 (±4) | 0.500 (±2) | >2.000 (±x8) | 2.000 (±1) | 2.000 (±1) |

MICs of BDQ and CFZ of preclinical strains, in the absence and presence of verapamil (40 µg/ml) or reserpine (3 µg/ml). Each value is the average of 3 independent experiments. For MIC without EPIs fold-changes between brackets represent the difference between resistant and wild-type MICs. For MICs in the presence of EPIs, fold-changes between brackets represent the differences between MIC with and without EPI. BDQ: bedaquiline; CFZ: clofazimine; RES: reserpine; VER: verapamil.

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| Table 4. Top 10 upregulated proteins of the BDQ-resistant strains EH3.6 and EH 3.2 compared to EH3.0. |
|---|---|---|---|
| **Rv number** | **gene** | **Description** | **Fold change EH3.6/EH3.0** | **Fold change EH3.6.2/EH3.0** |
| Rv0677c | mmpS5 | Possible conserved membrane protein MmpS5 | 18.7 | 4.6 |
| Rv0676c | mmpL5 | Probable conserved transmembrane transport protein MmpL5 | 5.9 | 2.3 |
| Rv0559c | Rv0559c | Possible conserved secreted protein | 5.5 | 1.9 |
| Rv0455c | Rv0455c | Conserved protein | 3.3 | 1.4 |
| Rv2936 | ddrA | Daunorubicin-dim-transport ATP-binding protein ABC transporter DmrA | 1.8 | 2.0 |
| Rv2610c | pinA | Alpha-mannosyltransferase PimA | 1.8 | 1.7 |
| Rv1426c | lipO | Probable esterase LipO | 1.8 | 1.4 |
| Rv0064A | vapB1 | Possible antitoxin VapB1 | 1.7 | 1.7 |
| Rv3836 | Rv3836 | Conserved hypothetical protein | 1.7 | 2.1 |
| Rv0736 | rslA | Anti-sigma factor RslA | 1.7 | 1.5 |

Proteins displaying the highest differential upregulation factors (top 10 proteins) are indicated. The descriptions of the genes were retrieved from TubercuList (http://tuberculist.epfl.ch/).

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mouse strain while the activity in a different mouse strain was very modest [14].

Disruption of MmpL5 by transposon insertion leads to attenuated virulence in mice, at least in some cases [15]. Overexpression of MmpL5 did not lead to increased fitness in vitro or increased virulence in vivo, but it did not result in a loss of fitness either. This increases the likelihood that non-target based resistance will be used by mycobacteria as a stepping stone leading to higher level, target-based resistance. Induction of efflux pumps was shown to be the first step in a general pathway to resistance to azithromycin, eventually leading to high-level chromosomal-mutation-related resistance as ordered events in an “antibiotic resistance arrow of time” [16]. The frequency of target-based and non-target based mutations for BDQ is very similar in vitro [3], but the latter are more likely to emerge first in vivo, as efflux-based mutations generally lead to lower levels of resistance (2 to 16-fold higher MICs) than target based mutations (16-1000 fold higher MICs). Indeed so far only non-

| Table 5. Regulation of efflux pumps proteins in EH3.6 and EH3.2. |
|---------------------------------------------------------------|
| Rv number | Gene | Efflux Pump Family | Description | Fold change EH3.6/EH3.0 | Fold change EH3.2/EH3.0 |
|-----------|------|-------------------|-------------|------------------------|------------------------|
| Rv0676c   | mmpL5 | RND               | Probable conserved transmembrane transport protein MmpL5 | 5.9 | 2.3 |
| Rv1183    | mmpL10| RND               | Probable conserved transmembrane transport protein MmpL10 | 1.0 | 0.8 |
| Rv0206c   | mmpL3 | RND               | Possible conserved transmembrane transport protein MmpL3 | 1.0 | 1.0 |
| Rv1877    | Rv1877| MFS               | Probable conserved integral membrane protein | 1.2 | 1.1 |
| Rv0341    | iniB  | Membrane protein  | Isoniazid inducible gene protein IniB | 1.0 | 1.4 |
| Rv0342    | iniA  | Membrane protein  | Isoniazid inducible gene protein IniA | 1.0 | 1.3 |
| Rv2936    | dnaA  | ABC               | Daurorubicin-dlm-transport ATP-binding protein ABC transporter DnaA | 1.8 | 2.0 |
| Rv1747    | Rv1747| ABC               | Probable conserved transmembrane ATP-binding protein ABC transporter | 1.1 | 1.1 |
| Rv1463    | Rv1463| ABC               | Probable conserved ATP-binding protein ABC transporter | 1.1 | 1.0 |
| Rv1458c   | Rv1458c| ABC              | Probable unidentified antibiotic-transport ATP-binding protein ABC transporter | 1.0 | 1.0 |
| Rv2477c   | Rv2477c| ABC              | Probable macrolide-transport ATP-binding protein ABC transporter | 1.0 | 1.0 |
| Rv2688c   | Rv2688c| ABC              | Antibiotic-transport ATP-binding protein ABC transporter | 0.9 | 1.0 |
| Rv1218c   | Rv1218c| ABC              | Probable tetronasin-transport ATP-binding protein ABC transporter | 0.6 | 1.0 |

Efflux pump proteins were selected from the complete dataset of the proteome analysis (Table S1). The descriptions of the genes were retrieved from TubercuList (http://tuberculist.epfl.ch/).

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| Table 6. MICs of MmpS5 and MmpL5 overexpression strains. |
|----------------------------------------------------------|
| Strain:plasmid | Description | MIC (µg/ml) |
|---------------|-------------|-------------|
|               |             | BDQ | BDQ+VER | CFZ | CFZ+VER |
| H37Rv         | wild type M. tuberculosis strain | 0.063 | 0.008 | 0.250 | 0.063 |
| H37Rv::pSD5    | H37Rv strain containing the control plasmid pSD5 | 0.063 | 0.004 | 0.250 | 0.031 |
| H37Rv::pCVGA30 | H37Rv-derived strain overexpressing MmpS5-MmpL5 | 0.250 | 0.063 | 0.500 | 0.250 |
| H37Rv::pCVGA26 | H37Rv-derived strain overexpressing MmpS5 | 0.063 | 0.004 | 0.250 | 0.031 |
| H37Rv::pCVGA28 | H37Rv-derived strain overexpressing MmpL5 | 0.063 | 0.008 | 0.250 | 0.063 |

MICs of strains overexpressing MmpS5, MmpL5 or the operon MmpS5-MmpL5 in the absence and presence of verapamil (40 µg/ml) are indicated. Each value is the average of at least 2 independent experiments. BDQ: bedaquiline; CFZ: clofazimine; VER: verapamil.
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between BDQ and CFZ is of great concern because of its clinical implications. Our findings suggest that BDQ and CFZ will not protect each other against the emergence of resistance. In patients, it is crucially important to use the combination BDQ + CFZ only if protected from emergence of resistance by additional antibiotics, that are not a substrate of the MmpL5 efflux pump.

In mice, the combination BDQ + CFZ leads to significantly better bactericidal and sterilizing efficacy, despite the cross-resistance between the two drugs [17]. It is important to note here that the 5 mouse mutants described above were isolated from mice that had been treated with BDQ + CFZ in combination with either amikacin alone, or amikacin and levofloxacin for 4 months, and then relapsed 3 months later. No resistant strains could be isolated from mice treated for 6 months with the same combinations, suggesting residual activity of the BDQ + CFZ combination, and/or sterilizing activity of the companion drug(s) between months 4 and 6. The emergence of resistant strains late in the treatment course, and their subsequent elimination by continuation of treatment, underlines the importance of treatment compliance and the risk of premature discontinuation of therapy.

There is an additional reason why premature discontinuation of treatment creates a high risk for the emergence of resistance to both BDQ and CFZ. As both drugs have very long half-lives [18,19] early discontinuation results in lingering low plasma levels of BDQ and CFZ, without protection of companion drugs with shorter terminal half-lives. Sub-therapeutic concentrations of some TB drugs produce tolerance in actively growing TB cultures [20], although current data do not suggest upregulation of the MmpL5 pump by BDQ [21]. Sequential use of BDQ and CFZ – or CFZ and BDQ may also increase the chance to develop efflux-based resistance.

Finally, our findings have implications for the development of rapid genotypic drug susceptibility tests (DST). To monitor for emergence of resistance to BDQ, both Rv0678 and atpE genes of baseline and post-baseline isolates should be sequenced. SNP based assays are less likely to become the method of choice for rapid DST testing, as SNPs have already been described in many positions of both genes. A database linking phenotypic changes in MIC with genotypic changes will need to be generated to allow for clinical decisions based on genotypic DST data.

**Methods**

1.1 Ethic statement

The animal study reported in this manuscript was carried out in compliance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All animal procedures were approved by the Johnson & Johnson ethical committee for the use of animals. Humane endpoints were introduced to minimize animal pain and suffering.
1.2 Bacterial strains, media and growth conditions

Preclinical *in vitro* isolates EH 3.2, EH 3.3 and EH 3.6 originated from the MDR TB strain EH 3.0 and were isolated as previously described [3]. Preclinical *in vivo* isolates BCA4, BCA8 and BCLA2 were isolated from mice infected with H37Rv which were first treated for 4 months with a combination of 25 mg/kg BDQ, 20 mg/kg CFZ, and 150 mg/kg amikacin (BCA4 and BCA8 isolate) or BDQ, CFZ, amikacin and 300 mg/kg levofloxacin (BCLA2 isolate). After this 4 months treatment period, mice were kept for 3 additional months without any treatment before being sacrificed and assessed for relapses. Lung homogenates were plated on 7 H11 agar plates without BDQ and emerging colonies were tested for BDQ sensitivity.

Clinical isolates studied (listed in Table 2) came from patients participating in study C209 (NCT00910871). All available paired (baseline and post-baseline) samples with increased post-baseline BDQ MICs from this trial were studied.

1.3 Antimicrobial sensitivity testing

MICs were determined using two-fold dilution in 7 H9 medium with the resazurin microtitre assay (REMA) as previously described [22]. The MIC was interpreted as the lowest concentration that prevented a change in color from blue to purple or pink.

1.4 DNA manipulations and sequence analysis

454 pyrosequencing. Genomic DNA from both the parent (EH 3.0) and two mutant (EH 3.2 and EH 3.6) strains was pyrosequenced using the GS FLX shotgun sequencing protocol from 454/Roche (GS-FLX, Roche Applied Science) following manufacturer’s instructions. Raw sequence data from the 454 sequencing experiments have been submitted to the Sequence Read Archive database (http://www.ncbi.nlm.nih.gov/Traces/sra/) with the accession numbers SRP042318, SRP042320 and SRP042321.

Genome assembly and variant detection. For each isolate, all individual sequences were assembled with high stringency onto the *M. tuberculosis* H37Rv reference sequence (GenBank NC_000962), using CLCbio genomics workbench 6.0.1 (CLC, Denmark). Following read assembly, a consensus sequence (representing the genome sequence of the sample) was retrieved for each of the 3 samples. Next, both mutant genome sequences were aligned to the genome of the parent strain and positional differences (conflicts) were calculated using CLCbio. Resulting variants per mutant strain were compared with the parent strain. Each of the identified variants was checked individually in the

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**Figure 3. Mechanism of BDQ and CFZ resistance in *Rv0678* mutants.** A. Regulation of *mmpS5* and *mmpL5* transcription by wild type *Rv0678* repressor. *Rv0678* protein binds to the intergenic region located between *Rv0678* and *mmpS5*, which contains the 10 consensus boxes of promoters for both *Rv0678* and *mmpS5*. This prevents the RNA polymerase to start transcription, resulting in the decrease of expression of *MmpS5, MmpL5* and *Rv0678* proteins. In response to an unknown stimulus, the regulator detaches from DNA and transcription can be resumed. B. Lack of regulation in *Rv0678* mutants. The strains carrying frame shifting mutations in *Rv0678* will not produce a functional repressor, thus the transcription of *mmpS5, mmpL5* and *Rv0678* will be increased. If the mutation results in an amino acid polymorphism, the protein may still be functional, but with reduced DNA-binding ability depending on the location of the mutation. In either case, the final consequence will be an increase in the expression levels of the proteins *MmpS5, MmpL5* and *Rv0678*.

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original read assembly of the sample in order to distinguish genuine variants from errors introduced during the sequencing process or assembly.

**PCR amplification and Sanger sequencing.** A 649pb DNA fragment containing Rv0678 gene and 143 bp upstream Rv0678 was amplified by PCR using primers CV010 and CV017 (Table S3). In the case of EH 3.6, this PCR fragment is 2007 bp, so additional PCRs with primers Rv0678 Fwd 2, Rv0678 Rev 2, IS6110_F and IS6110_R were necessary for sequencing the product. The PCR products were gel purified using QiAquick Gel Extraction Kit (QIAGEN), and further sequenced with the same primers used for amplification. For analysis of the sequences, Rv0678 sequence from *M. tuberculosis* H37Rv was taken as a reference [23] (http://tuberculist.epfl.ch).

1.5 Proteome analysis

Protein extraction and analysis were performed as previously described [21].

**Sample preparation.** *M. tuberculosis* strains were cultured in 7H9 broth to mid-log phase. Cell lysate was prepared in 2 ml lysis buffer and passed 3 times through the French Press at 14000 Pa in a small cell. Cell debris was removed by centrifugation and supernatant was collected.

**Digestion and iTRAQ Reagent Labeling.** The proteins were cleaned up by cold ethanol/acetone precipitation and digested with trypsin. After digestion, the peptides were labelled using iTRAQ labeling method. The samples were mixed and dried down using vacuum centrifugation.

**Nano-LC MS/MS.** Peptide mixtures were loaded on a HPLC Agilent 1200 system. After elution, they were further analysed by an EASY nano-HPLC system connected to an LTQ Orbitrap XL mass spectrometer.

**Data analysis.** The raw MS/MS data were analysed using Proteome Discoverer v1.3 beta. MS/MS spectra were converted to .mgf files and searched against the NCBI database (taxonomy: *Mycobacterium tuberculosis* H37Rv). The protein ratios were calculated based on the peptide ratios of the individual proteins. The peptide ratios were log2 transformed and normalized against the median. To find regulated proteins, all outlier peptides were removed and the protein ratio recalculated. The standard deviation (σ) of the protein ratios were calculated after removing outliers. Every protein was accepted as regulated with a log ratio of 

\[
\pm 2\sigma, \quad (2\sigma = 95.4\% \text{ confidence interval})
\]

1.6 Generation of overexpression strains

Replicative plasmids for overexpression of *mmpS5, mmpL5* or the operon *mmpS5-mmpL5* were cloned by GeneArt Gene Synthesis (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies). The three inserts, of 444 bp, 2910 bp and 3335 bp respectively, were synthesized (Life Technologies).

Replicative plasmids derived from pSD5 used for overexpression either *mmpS5, mmpL5* or the complete operon *mmpS5-mmpL5* in *M. tuberculosis* H37Rv. The inserts were cloned in the restriction sites *NdeI* and *MluI* under control of the strong promoter *P_\text{Hsp65}*. (TIF)

**Figure S2**

**Supporting Information**

**Table S1** Regulation of the proteome of EH 3.2 and EH 3.6 compared with the parent strain 3.0. The relatively quantified...
expression of the complete dataset (1455 proteins) is shown. Proteins lacking the Rv number in the list could not be found in *M. tuberculosis* H37Rv strain, and are referred to other mycobacterial species.

**Table S2** Top 9 downregulated proteins of the BDQ-resistant strains EH 3.6 and EH 3.2 compared to EH 3.0. Proteins displaying the highest differential downregulation factors (top 9 proteins) are indicated.

**Table S3** Oligonucleotides used in this study.

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**Author Contributions**

Conceived and designed the experiments: KA KT NL AK. Performed the experiments: CV NG KT TG LV. Analyzed the data: KA KT NL BCdJ AK. Contributed to the writing of the manuscript: KA CV AK.