Genome-wide Association Studies for Osteoporosis: A 2013 Update

Yong-Jun Liu¹, Lei Zhang¹,², Christopher J. Papasian³, Hong-Wen Deng¹,²

¹Center for Bioinformatics and Genomics, Department of Biostatistics, School of Public Health and Tropical Medicine, Tulane University, New Orleans, LA, USA
²Center of System Biomedical Sciences, University of Shanghai for Science and Technology, Shanghai, PR, China
³University of Missouri - Kansas City, School of Medicine, Kansas City, MO, USA

In the past few years, the bone field has witnessed great advances in genome-wide association studies (GWASs) of osteoporosis, with a number of promising genes identified. In particular, meta-analysis of GWASs, aimed at increasing the power of studies by combining the results from different study populations, have led to the identification of novel associations that would not otherwise have been identified in individual GWASs. Recently, the first whole genome sequencing study for osteoporosis and fractures was published, reporting a novel rare nonsense mutation. This review summarizes the important and representative findings published by December 2013. Comments are made on the notable findings and representative studies for their potential influence and implications on our present understanding of the genetics of osteoporosis. Potential limitations of GWASs and their meta-analyses are evaluated, with an emphasis on understanding the reasons for inconsistent results between different studies and clarification of misinterpretation of GWAS meta-analysis results. Implications and challenges of GWAS are also discussed, including the need for multi- and inter-disciplinary studies.

Key Words: Genome-wide association study, Osteoporosis

INTRODUCTION

In the past 4 years, genome-wide association studies (GWASs), assaying hundreds of thousands of single nucleotide polymorphisms (SNPs) in thousands of individuals, have identified a number of promising genetic variants that are associated with osteoporosis and related traits. The first published whole genome sequencing study for osteoporosis and fractures was published, reporting a novel rare nonsense mutation. This article reviews the current status of GWASs of osteoporosis and their meta-analyses with an emphasis on prominent results, approaches, and problems with these studies. The major findings of the first whole genome sequencing study for osteoporosis and fractures are also briefly discussed. We focus primarily on bone mineral density (BMD), the most important risk trait for osteoporosis, and on osteoporotic fracture (OF), the most severe clinical outcome of osteoporosis. We address how to interpret the discordance of research findings between individual GWASs and meta-analyses, and between different meta-analy-
GWASs AND META-ANALYSIS ON OSTEOPOROSIS AND OF

Osteoporosis is the most common metabolic skeletal disorder in humans. It predisposes people to fragility fractures and confers substantial morbidity and mortality, affecting over 200 million people worldwide.[1,2] Osteoporosis is mainly characterized by low BMD, a highly heritable trait with heritability ranging from 0.5 to 0.8.[3-6] OF, as an endpoint clinical outcome of osteoporosis, also has moderate heritability, of approximately 0.5-0.7.[7,8] To date, GWASs and their meta-analyses have identified over 60 genes/loci associated with variations in BMD and more than 20 genes/loci associated with risk of OF. In addition, a few recently published whole-genome sequencing study identified a rare nonsense mutation novel within a novel gene LGR4 that was strongly associated with low BMD and OF.[9]

The majority of published GWASs have focused on BMD using SNP data. A recent review by Richards et al.[10] summarized the major findings from SNP-based GWASs, which will not be repeated here. Instead, we highlight prominent genes or loci identified in SNP based GWASs with a focus on consistency and inconsistency of results. We address issues related to interpretation of meta-analysis results and replication of study findings among GWASs and meta-analyses. A few GWASs have focused on genome wide analyses of copy number variants (CNV) and biological pathways. The major findings from these latter studies are briefly introduced and discussed.

1. GWASs based on SNPs

To date, a total of 19 GWASs have been published for osteoporosis. Of these, 14 are individual GWASs, and 5 are GWAS meta-analyses. The significant genes and loci identified from these studies, along with information regarding study design (e.g., individual GWASs or meta-analysis) and phenotypes (e.g., hip and/or spine BMD or OF) are summarized in tables 1 and 2.

1) GWAS meta-analyses

The Genetic Factors of Osteoporosis (GEFOS) consortium published two large-scale GWAS meta-analyses.[11,12] Their first meta-analysis (GEFOS-1), which included 19,195 subjects of European descent, identified 20 BMD loci.[11] Their second GWAS meta-analysis (GEFOS-2), the largest one to date in the bone field, included 32,961 individuals in the discovery phase and was replicated in 50,933 independent subjects.[12] The study subjects included Europeans and East Asians. GEFOS-2 identified a total of 56 BMD loci at the genome-wide significance level. Of these 56 loci, 32 were novel and the remaining 24 were genes/loci that were previously known to affect bone mass regulation and metabolism (e.g., receptor activator of nuclear factor (NF)-kappaB (κB) [RANK], RANK ligand [RANKL], and lipoprotein receptor-related protein 5 [LRP5]). Notably, multiple loci achieved highly significant associations in GEFOS-2, with the magnitude of P values reaching $10^{-4}$ for 2 loci, $10^{-5}$ for 1 locus, $10^{-6}$ for 2 loci, $10^{-7}$ for 6 loci, $10^{-8}$ for 11 loci, and $10^{-10}$ for 27 loci. GEFOS-2 also revealed 14 loci associated with risk of fractures (Table 2).

Koller et al.[17] carried out a meta-analysis of GWASs restricted to premenopausal white women from four cohorts (n = 4,061, aged 20 to 45 years), a subset of GEFOS-2, and identified two loci (wingless-type MMTV integration site family, member 16 [WNT16] and estrogen receptor 1 [ESR1]/C6orf97) influencing peak bone mass at the lumbar spine and femoral neck. Only 4 of the 56 loci detected in the joint female GEFOS analysis[12] were observed to have P values below $5 \times 10^{-5}$ in this study.

Zhang et al.[18] conducted a three-stage GWAS meta-analysis in 27,061 study subjects. Stage 1 meta-analyzed seven GWA samples and 11,140 subjects for BMDs at the lumbar spine, hip and femoral neck, followed by a Stage 2 in silico replication of 33 SNPs in 9,258 subjects, and by a Stage 3 de novo validation of three SNPs in 6,663 subjects. Combining evidence from all the stages, two novel loci were identified at the genome-wide significance level:
### Table 1. Putative bone mineral density genes identified in genome-wide association studies and meta-analyses

| Gene      | Full name                                                                 | Most significant \( P \) value | Initial discovery study design | Reference                           | Initial discovery population | Within-study replication | Between-study replication                        |
|-----------|---------------------------------------------------------------------------|----------------------------------|--------------------------------|-------------------------------------|-----------------------------|--------------------------|-----------------------------------------------|
| ABCF2     | ATP-binding cassette, sub-family F 2.56×10(-5)                            | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 | Caucasians               |
| ADAMTS18  | ADAM metallopeptidase with thrombospondin type 1 motif, 18 2.56×10(-5)  | GWAS                             | Xiong et al.[30]                | US Caucasians                      | Chinese                     |                         |
| ALDH7A1   | Aldehyde dehydrogenase 7 family, member A1 2.08×10(-9)                  | GWAS                             | Guo et al.[37]                  | Chinese                            | Caucasians                 | ALS                      |
| ANAPC1    | Anaphase promoting complex subunit 1 1.5×10(-9)                          | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| ARHGAP1   | Rho GTPase activating protein 1 4.0×10(-9)                               | Meta-analysis                     | Rivadeneira et al. [11]         | Caucasians of Northern European    | Estrada et al.[12]         | in Caucasians and Asians |
| AXIN1     | Axin 1 2.2×10(-8)                                                        | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| C7orf58   | Chromosome 7 open reading frame 58 1.1×10(-9)                            | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| C12orf23  | Chromosome 12 open reading frame 23 9.6×10(-10)                          | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| C18orf19  | Chromosome 18 open reading frame 19 4.9×10(-8)                           | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| CDKAL1    | CDK5 regulatory subunit associated protein 1-like 1 1.8×10(-8)           | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| CLCN7     | Chloride channel, voltage-sensitive 7 1.5×10(-16)                        | GWAS                             | Duncan et al.[26]               | Postmenopausal Caucasian women     | Estrada et al.[12]         | in Caucasians and Asians |
| CLDN14    | Claudin 14 4.15×10(-9)                                                   | Meta-analysis                     | Zhang et al.[18]                | Caucasians                         | Caucasians                 | Estrada et al.[12]       | in Caucasians and Asians |
| CPN1      | Carboxypeptidase N, polypeptide 1 9×10(-10)                              | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| CRHR1     | Corticotropin releasing hormone receptor 1 1.4×10(-8)                    | Meta-analysis                     | Rivadeneira et al. [11]         | Caucasians of Northern European    | Estrada et al.[12]         | in Caucasians and Asians |
| CTNNB1    | Catenin (cadherin-associated protein), beta 1, 8.1×10(-10)              | Meta-analysis                     | Rivadeneira et al. [11]         | Caucasians of Northern European    | Estrada et al.[12]         | in Caucasians and Asians |
| CYLD      | Cylindromatosis (turban tumor syndrome) 6.2×10(-8)                      | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| DCDC5     | Doublecortin domain containing 5 2.3×10(-9)                              | Meta-analysis                     | Rivadeneira et al. [11]         | Caucasians of Northern European    | Estrada et al.[12]         | in Caucasians and Asians |
| DHH       | Desert hedgehog 1.2×10(-15)                                              | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| DNM3      | Dynamin 3 8.5×10(-15)                                                    | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |
| DOK6      | Docking protein 6 8.87×10(-7)                                             | GWAS                             | Hsu et al.[27]                  | US Caucasians                      | Caucasians of Northern European |
| ERC1      | ELKS/RAB6 interacting/CAST family member 1 5.6×10(-12)                   | Meta-analysis                     | Estrada et al.[12]              | Caucasians                         | Caucasians                 |                         |

(Continued to the next page)
Table 1. Putative bone mineral density genes identified in genome-wide association studies and meta-analyses

| Gene   | Full name                        | Most significant P value | Initial discovery study design | Reference | Initial discovery population | Within-study replication | Between-study replication |
|--------|----------------------------------|--------------------------|--------------------------------|-----------|------------------------------|-------------------------|----------------------------|
| ESR1   | Estrogen receptor 1              | 6.1 × 10^{-11}           | GWAS                           | Styrkarsdottir et al.[24] | Icelandic Caucasians       | Caucasians of western Europe. | Rivadeneira et al.[11] and Richards et al.[23] in Caucasians. Koller et al.[31] in premenopausal women. |
| FAM9B  | Family with sequence similarity 9, member B | 1.2 × 10^{-8}           | Meta-analysis                   | Estrada et al.[12] | Caucasians and Asians        | Caucasians and Asians     | Estrada et al.[12] in Caucasians and Asians |
| FLJ42280 | Putative uncharacterized protein | 9.4 × 10^{-12}           | Meta-analysis                   | Rivadeneira et al.[11] | Caucasians of Northern European | Caucasians of Northern European | Estrada et al.[12] in Caucasians and Asians |
| FMN2/GREM2 | Formin 2/ gremlin 2          | 1.9 × 10^{-9}           | GWAS                           | Paternoster et al.[19] | Caucasians of Northern European | Caucasians of Northern European | Estrada et al.[12] in Caucasians and Asians |
| FOXL1  | Forkhead box L1                 | 1.7 × 10^{-8}           | Meta-analysis                   | Rivadeneira et al.[11] | Caucasians of Northern European | Styrkarsdottir et al.[70] in East-Asians. Estrada et al.[12] in Caucasians and Asians |
| FUBP3  | Far upstream element (FUSE) binding protein 3 | 3.1 × 10^{-8}           | Meta-analysis                   | Estrada et al.[12] | Caucasians and Asians        | Caucasians and Asians     | Caucasians and Asians |
| GALNT3 | UDP-N-acetly-alpha-D-galactosamine:polypeptide N-acetlygalactosaminyl-transferase 3 | 2.3 × 10^{-5}           | GWAS                           | Duncan et al.[28] | Postmenopausal Caucasian women | Caucasians | Estrada et al.[12] in Caucasians and Asians |
| GPATCH1 | G patch domain containing 1      | 9.2 × 10^{-9}           | Meta-analysis                   | Estrada et al.[12] | Caucasians and Asians        | Caucasians and Asians     | Caucasians and Asians |
| GPR177/ |                                  |                          |                                 |           |                              |                         |                             |
| WLS    | Wntless homolog [Drosophilia]   | 3.3 × 10^{-13}           | Meta-analysis                   | Rivadeneira et al.[11] | Caucasians of Northern European | Styrkarsdottir et al.[70] in East-Asians. Hsu et al.[27] in US Caucasians. Duncan et al.[26] in Postmenopausal Caucasian women. Estrada et al.[12] in Caucasians and Asians |
| HDAC5  | Histone deacetylase 5           | 1.7 × 10^{-8}           | Meta-analysis                   | Rivadeneira et al.[11] | Caucasians of Northern European | Caucasians of Northern European | Estrada et al.[12] in Caucasians and Asians |
| IBSP   | Integrin-binding sialoglycoprotein | 7.6 × 10^{-7}           | GWAS                           | Duncan et al.[26] | Postmenopausal Caucasian women | Caucasians | Estrada et al.[12] in Caucasians and Asians |
| IDUA   |                                  |                          |                                 |           |                              |                         |                             |
| IL21R  | Interleukin 21 receptor          | 2.31 × 10^{-6}          | GWAS                           | Guo et al.[37] | Chinese                       | Caucasians and Asians     | Chinese |
| INSSG2 | Alpha L-iduronidase              | 1.2 × 10^{-10}          | Meta-analysis                   | Estrada et al.[12] | Caucasians and Asians        | Caucasians and Asians     | Caucasians and Asians |
| JAG1   | Jagged 1                        | 5.27 × 10^{-10}         | GWAS                           | Kung et al.[71] | Southern Chinese women        | Estrada et al.[12] in Caucasians and Asians | Estrada et al.[12] in Caucasians and Asians |
| KIAA2018 |                                  |                          |                                 |           |                              |                         |                             |
| KCNMA1 | Potassium large conductance calcium-activated channel, subfamily M, alpha member 1 | 1.5 × 10^{-12}          | Meta-analysis                   | Estrada et al.[12] | Caucasians and Asians        | Caucasians and Asians     | Caucasians and Asians |

(Continued to the next page)
Table 1. (Continued from the previous page) Putative bone mineral density genes identified in genome-wide association studies and meta-analyses

| Gene            | Full name                                      | Most significant P value | Initial discovery study design | Reference                        | Initial discovery population | Within-study replication | Between-study replication |
|-----------------|------------------------------------------------|--------------------------|--------------------------------|---------------------------------|-----------------------------|--------------------------|--------------------------|
| LACTB2          | Lactamase, beta 2                              | 4.7 × 10(-8)             | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| LKR1            | Leucine, glutamate and lysine rich 1           | 4.5 × 10(-12)            | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| LIN7C           | Lin 7 homologue C                              | 4.9 × 10(-8)             | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| LRP4            | Low density lipoprotein receptor-related protein 4 | 4.0 × 10(-9)             | Meta-analysis                   | Rivadeneira et al.[11]          | Caucasians of Northern Euro- pean | Estrada et al.[12] in Caucasians and Asians |                           |
| LRP5            | Low density lipoprotein receptor-related protein 5 | 6.3 × 10(-12)            | GWAS                           | Richards et al.[23]             | Caucasians                  | Caucasians of western Europe | Rivadeneira et al.[11] in Caucasians of Northern European. Estrada et al.[12] in Caucasians and Asians | |
| LRR4C           | Leucine rich repeat containing 4C              | 8.89 × 10(-7)            | GWAS                           | Hsu et al.[27]                  | US Caucasians               | Caucasians of Northern European |                          |
| MARK3           | MAP/microtubule affinity-regulating kinase 3   | 1.8 × 10(-9)             | GWAS                           | Styrkarsdottir et al.[72]       | Icelanders                  | Caucasians of European descent | Rivadeneira et al.[11] in Caucasians of Northern European. Estrada et al.[12] in Caucasians and Asians | |
| MBL2            | Mannose-binding lectin (protein C) Z, soluble  | 1.6 × 10(-12)            | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| MEF2C           | Myocyte enhancer factor 2C                     | 1.3 × 10(-13)            | GWAS                           | Duncan et al.[26]               | Postmenopausal Caucasian women | Caucasians               | Rivadeneira et al.[11] in Caucasians of Northern European. Estrada et al.[12] in Caucasians and Asians | |
| MEPE            | Matrix extracellular phosphoglycoprotein       | 4.0 × 10(-9)             | Meta-analysis                   | Rivadeneira et al.[11]          | Caucasians of Northern Euro- pean. | Styrkarsdottir et al.[70] in East-Asians |                           |
| MPP7            | Membrane protein, palmi- coatylated 7 (MAGUK p55 subfamily member 7) | 2.9 × 10(-9)             | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| OSBP1A          | Oxysterol binding protein-like 1A              | 4.22 × 10(-7)            | GWAS                           | Hsu et al.[27]                  | US Caucasians               | Caucasians of Northern European |                          |
| PTHLH           | Parathyroid hormone-like hormone               | 1.9 × 10(-12)            | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| RAP1A           | RAS-related protein RAP1A                      | 2.80 × 10(-7)            | GWAS                           | Hsu et al.[27]                  | US Caucasians               | Caucasians of Northern European |                          |
| PKDC3           | Protein kinase domain containing, cytoplasmic homologue | 1.3 × 10(-9)             | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| NTAN1           | Ribosomal protein S6 kinase, 90kda, polypeptide 5 | 1.7 × 10(-11)            | Meta-analysis                   | Estrada et al.[12]              | Caucasians and Asians       | Caucasians and Asians    |                          |
| RSP03           | R-spondin 3                                   | 2.2 × 10(-9)             | GWAS                           | Duncan et al.[26]               | Postmenopausal Caucasian women | Estrada et al.[12] in Caucasians and Asians |                          |
| RUNX2           | Runt-related transcription factor 2            | 5.6 × 10(-11)            | Meta-analysis                   | Caucasians and Asians           | Caucasians and Asians       | Caucasians and Asians    |                          |

(Continued to the next page)
| Gene      | Full name                                      | Most significant P value | Initial discovery study design | Reference                      | Initial discovery population | Within-study replication | Between-study replication |
|-----------|-----------------------------------------------|--------------------------|--------------------------------|--------------------------------|-----------------------------|--------------------------|---------------------------|
| SALL1/ CYLD | Sal-like 1 (Drosophila)                       | 5.0 × 10(-12)            | Meta-analysis                  | Estrada et al.[12]             | Caucasians and Asians       | Caucasians and Asians     |                           |
| SLC25A13  | Solute carrier family 25                      | 8.1 × 10(-48)            | Meta-analysis                  | Estrada et al.[12]             | Caucasians and Asians       | Caucasians and Asians     |                           |
| SMG6      | Smg-6 homolog, nonsense mediated mrn decay factor (C. Elegans) | 1.7 × 10(-8)            | Meta-analysis                  | Estrada et al.[12]             | Caucasians and Asians       | Caucasians and Asians     |                           |
| SMOC1     | SPARC related modular calcium binding 1       | 3.98 × 10(-13)           | Meta-analysis                  | Zhang et al.[18]               | Caucasians and Asians       | Caucasians and Asians     | Estrada et al.[12] in Caucasians and Asians |
| SOST      | Sclerostin                                    | 2.1 × 10(-8)             | GWAS                           | Styrkarsdottir et al.[72]      | Icelanders                  | Caucasians of European decent | Estrada et al.[70] in East-Asians, Estrada et al.[12] in Caucasians and Asians |
| SOX4      | SRY (sex determining region Y)-box 4          | 1.8 × 10(-8)             | GWAS                           | Duncan et al.[26]              | Postmenopausal Caucasian women | Caucasians               | Estrada et al.[12] in Caucasians and Asians |
| SOX6      | SRY (sex determining region Y)-box 6          | 6.4 × 10(-10)            | Meta-analysis                  | Rivadeneira et al.[11]         | Caucasians of Northern European |                           | Estrada et al.[12] in Caucasians and Asians, Liu et al.[48] in US Caucasians |
| SOX9      | SRY-box containing gene 9                    | 3.7 × 10(-8)             | Meta-analysis                  | Estrada et al.[12]             | Caucasians and Asians       | Caucasians and Asians     |                           |
| SP7       | Sp7 transcription factor 7                    | 9.9 × 10(-11)            | GWAS                           | Styrkarsdottir et al.[71]      | Icelanders                  | Caucasians of European decent |                           |
| SPP1      | Secreted phosphoprotein 1                     | 6.0 × 10(-8)             | GWAS                           | Duncan et al.[26]              | Postmenopausal Caucasian women | Caucasians               |                           |
| SPP2      | Secreted phosphoprotein 2, 24 kda             | 4.64 × 10(-7)            | GWAS                           | Hsu et al.[27]                 | US Caucasians               | Caucasians of Northern European | Estrada et al.[12] in Caucasians and Asians |
| SPTBN1    | Spectrin, beta, non-erythrocytic 1            | 1.6 × 10(-8)             | Meta-analysis                  | Rivadeneira et al.[11]         | Caucasians of Northern European |                           | Estrada et al.[12] in Caucasians and Asians |
| STARD3NL  | STARD3 N-terminal like                        | 1.1 × 10(-9)             | Meta-analysis                  | Rivadeneira et al.[11]         | Caucasians of Northern European |                           | Estrada et al.[12] in Caucasians and Asians |
| TBC1D8    | TBC1 domain family, member 8 (with GRAM domain) | 1.48 × 10(-7)            | GWAS                           | Hsu et al.[27]                 | US Caucasians               | Caucasians of Northern European |                           |
| TGFB3     | Transforming growth factor, beta receptor III | 2.13 × 10(-8)            | GWAS                           | Xiong et al.[30]               | US Caucasians               | Chinese                  |                           |
| TNFSF11/ RANKL | Tumor necrosis factor (ligand) superfamily, member 11 | 2.0 × 10(-14)            | GWAS                           | Richards et al.[23]            | Caucasians                  | Caucasians of western Europe | Duncan et al.[26] in Postmenopausal Caucasian women, Styrkarsdottir et al.[24] in Caucasians, Estrada et al.[12] in Caucasians and Asians |

(Continued to the next page)
GWASs for osteoporosis

14q24.2 (rs227425, \(P = 3.98 \times 10^{-13}\), SPARC related modular calcium binding 1 [SMOC1]) and 21q22.13 (rs170183, \(P = 4.15 \times 10^{-9}\), claudin 14 [CLDN14]). These two SNPs were also significant in GEFOS-2.[12] This study independently confirmed 13 previously reported loci.[18] Further gene expression analysis in osteogenic cells implied potential functional association of the two novel candidate genes (SMOC1 and CLDN14) in bone metabolism.

Most studies have focused on areal BMD (aBMD) obtained from a 2-dimensional projectional scan with dual energy X-ray absorptiometry (DXA). Although aBMD is the gold standard for diagnosing osteoporosis, it fails to provide a detailed information necessary to discern traits such as trabecular volumetric BMD (vBMD), cortical vBMD and bone microstructural parameters. Quantitative computed tomography (QCT) analysis has the advantage to reveal unique information about these bone traits. Paternoster et al.[19] published the first GWAS to identify genetic loci associated with cortical and trabecular bone microstructural parameters in European Caucasians. Their cortical vBMD GWAS meta-analysis (n=5,878) followed by replication (n=1,052) identified genetic variants in four separate loci (RANKL, rs1021188, \(P = 3.6 \times 10^{-14}\); LOC285735, rs271170, \(P = 2.7 \times 10^{-12}\); osteoprotegerin [OPG], rs7839059, P = 1.2 x 10^{-6}; and ESR1/C6orf97, rs6909279, \(P = 1.1 \times 10^{-5}\)). The trabecular vBMD GWA meta-analysis (n=2,500) followed by

### Table 1. (Continued from the previous page) Putative bone mineral density genes identified in genome-wide association studies and meta-analyses

| Gene                      | Full name                                         | Most significant P value | Initial discovery study design | Reference                                      | Within-study replication | Between-study replication |
|---------------------------|---------------------------------------------------|--------------------------|--------------------------------|-----------------------------------------------|--------------------------|---------------------------|
| TNFRSF11A/RANK            | Tumor necrosis factor receptor superfamily, member 11a, NFKB activator | 9.4 x 10(-9)            | GWAS                           | Styrkarsdottir et al.[24]                    | Icelandic Caucasians     | Caucasians of European descent | Richards et al.[23] in Caucasians of western Europe. Rivadeneira et al.[11] in Caucasians of Northern European. Estrada et al.[12] in Caucasians and Asians. |
| TNFRSF11B/OPG             | Tumor necrosis factor receptor superfamily, member 11b | 3.5 x 10(-16)            | GWAS                           | Styrkarsdottir et al.[24]                    | Icelandic Caucasians     | Caucasians of European descent | Richards et al.[23] in Caucasians of western Europe. Rivadeneira et al.[11] in Caucasians of Northern European. Estrada et al.[12] in Caucasians and Asians. |
| WNT16                     | Wingless-type MMTV integration site family, member 16 | 1.4 x 10(-16)            | Meta-analysis                   | Estrada et al.[12]                          | Caucasians and Asians    | Caucasians and Asians       | Koller et al.[17] in premenopausal women |
| WNT4                      | Wingless-type MMTV integration site family, member 4 | 9.6 x 10(-11)            | Meta-analysis                   | Rivadeneira et al.[11]                      | Caucasians of Northern European | Duncan et al.[26] in postmenopausal Caucasian women. Styrkarsdottir et al.[70] in East-Asians. Estrada et al.[12] in Caucasians and Asians. |
| XKR9                      | XK, Kell blood group complex subunit-related family, member 9 | 4.7 x 10(-8)             | Meta-analysis                   | Estrada et al.[12]                          | Caucasians and Asians    | Caucasians and Asians       | Duncan et al.[26] in postmenopausal Caucasian women. Styrkarsdottir et al.[70] in East-Asians. Estrada et al.[12] in Caucasians and Asians. |
| ZBTB40                    | Zinc finger and BTB domain containing 40          | 3.2 x 10(-10)            | Meta-analysis                   | Rivadeneira et al.[11]                      | Caucasians of Northern European | Duncan et al.[26] in postmenopausal Caucasian women. Styrkarsdottir et al.[70] in East-Asians. Estrada et al.[12] in Caucasians and Asians. |

Genes/loci in bold are those with evidence of cross study replication.

Within-study replication means the study includes a follow-up replication component.

Between-study replication means the gene/loci showed significance in different studies (not necessarily totally independent because some large meta-analyses such as GEFOS-2 [Estrada et al. [12]] include the samples from other individual GWASs.

- LS, lumbar spine; FN, femoral neck; OF, osteoporotic fracture; HF, hip fracture.
Table 2. Putative osteoporotic fracture genes identified in genome-wide association studies and meta-analyses

| Gene          | Full name                                      | Phenotype       | Study design (GWAS, meta-analysis) | Reference              | Most significant P value |
|---------------|-----------------------------------------------|-----------------|-----------------------------------|------------------------|--------------------------|
| ADAMTS18      | ADAM metallopeptidase with thrombospondin type 1 motif, 18 | Hip-OF          | GWAS                              | Xiong et al.[30]       | 2.9 x 10(-2)             |
| ALDH7A1       | Aldehyde dehydrogenase 7 family, member A1    | Hip-OF          | GWAS                              | Guo et al.[37]         | 8.53 x 10(-9)            |
| C1orf53       | Chromosome 17 open reading frame 53           | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 4.1 x 10(-5)             |
| CTCNNB1       | Catenin (cadherin-associated protein), beta 1 | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 2.9 x 10(-7)             |
| DCD5          | Doublecortin domain containing 5              | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 3.3 x 10(-5)             |
| FAM210A       | Family with sequence similarity 210, member a  | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 8.8 x 10(-13)            |
| FUBP3         | Far upstream element (FUSE) binding protein 3 | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 3.5 x 10(-5)             |
| LRP5          | Low density lipoprotein receptor-related protein 5 | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 1.4 x 10(-8)             |
| MBL2/DKK1     | Lectin, mannose-binding, soluble, 2           | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 9.0 x 10(-9)             |
| MECOM         | MDS1 and EV1 complex locus                    | OF              | GWAS                              | Hwang et al.[38]       | 3.59 x 10(-8)            |
| MEPE/SPP1     | Matrix, extracellular, phosphoglycoprotein    | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 1.7 x 10(-8)             |
| RPS6KA5       | Ribosomal protein S6 kinase, polypeptide 5    | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 7.2 x 10(-5)             |
| SLC25A13      | Solute carrier family 25 (citrin), member 1   | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 5.9 x 10(-11)            |
| SOST          | Sclerostin                                    | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 6.9 x 10(-6)             |
| SPTBN1        | spectrin, beta, nonerythrocytic, 1            | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 2.6 x 10(-8)             |
| STAR03NL      | STAR03 N-terminal like                        | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 7.2 x 10(-5)             |
| TGFBR3        | Transforming growth factor, beta receptor III | Hip-OF          | GWAS                              | Xiong et al.[30]       | 2.13 x 10(-8)            |
| TNFRSF11A (RANK) | Tumor necrosis factor receptor superfamily, member 11a, NFKB activator | NV/V-Fracture | Meta-analysis                     | Richards et al.[23]   | 0.02                     |
| UGT2B17       | UDP glucuronosyltransferase 2 family, polypeptide B17 | Hip OF          | GWAS                              | Yang et al.[42]        | 5.0 x 10(-4)             |
| WNT4          | Wingless-related MMTV integration site 4      | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 1.4 x 10(-7)             |
| WNT16         | Wingless-related MMTV integration site 16     | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 2.7 x 10(-7)             |
| ZBTB40        | Zinc finger and BTB domain containing 40      | Any type of fracture | Meta-analysis                     | Estrada et al.[12]    | 3.6 x 10(-6)             |

Any type of fracture: consisting of low-trauma fractures at any skeletal site (except fingers, toes and skull). NV Fracture, Nonvertebral fracture; OF, Osteoporotic fracture; GWAS, genome-wide association study.

replication (n=1,022) identified one locus reaching genome-wide significance (formin 2 [FMN2]/ gremlin 2, DAN family BMP antagonist [GREM2], rs9287237, P=1.9x10(-7)). In addition, rs1021188 was associated with cortical porosity while rs9287237 was associated with trabecular bone fraction. The genetic variant in the FMN2/GREM2 locus was also associated with fracture risk in the MrOS Sweden cohort and GREM2 expression in human osteoblasts. Two of these (FMN2/GREM2 and LOC285735) are novel bone related loci, while the other three have previously been reported to be associated with aBMD. This study provided evidence that the genetic determinants of cortical and trabecular vBMDs differ. However, QCT has its limitations, including being not applicable to World Health Organization (WHO) definition of osteoporosis that is based on DXA measurement, being more expensive with a higher dosage of exposure to radiation and may not predict fractures better than DXA measurement.[20,21] Nevertheless, its advantages over DXA make QCT a complementary (but not necessarily replacement) approach to bone health assessment.[22]

Conducting these large-scale meta-analyses requires an extensive, concerted effort in collaboration and coordination among various research centers and groups, as well as centralization and standardization in data analyses. Thus, publication of these studies represents one of the most impressive achievements in the osteoporosis genetics field. Based primarily on the large samples involved in meta-analyses such as these, some researchers have begun to consider large meta-analyses as a gold standard for evaluating other individual GWASs, with an explicit assumption that validity of the findings from other studies of smaller samples needs to be confirmed in large meta-analyses such as GEfos-2.[12]
Comparing the results of the two published GEFOS meta-analyses, it is interesting to note that several loci identified in GEFOS-1 (e.g., corticotropin releasing hormone receptor 1 [CRHR1] and histone deacetylase 5 [HDAC5]) were not significant in GEFOS-2, despite the fact that the number of samples of GEFOS-2 almost tripled the number in GEFOS-1, and the majority of samples used in GEFOS-1 were included in GEFOS-2.

Further comparing the results of these meta-analyses with individual GWASs, it is intriguing that some significant loci identified in meta-analyses were found to be significant at the genome-wide level in individual GWASs, while others were not. Likewise, some significant loci identified in individual GWASs were replicated in meta-analyses, while many others were not. We will highlight some of these consistencies and inconsistencies in the following paragraphs and discuss how to interpret these findings.

2) Individual GWASs

(1) Single ethnicity

The first two individual GWASs were performed in human subjects of European ancestry [23,24] using the data from TwinsUK/Rotterdam and deCODE Genetic studies. These two studies identified a total of five loci at the genome-wide significance level ($P < 5 \times 10^{-8}$). The associated genes, including LRPS, OPG, RANK, RANKL and ESR1, are well known for their importance to bone and mineral metabolism, and have been well established in previous candidate gene studies.

The efficiency of extreme-truncate selection design for quantitative trait association studies has been well established.[25] Duncan et al. adopted this design by focusing on postmenopausal Caucasian women with extremely high or low hip BMD in the discovery stage.[26] Although the sample size of the discovery phase was moderate (1,955 subjects in discovery phase), the power of the study was still high due to the extremely discordant sample design. In addition to replicating 21 of 26 known BMD-associated genes, they identified six new associations in/around the genes chloride channel, voltage-sensitive 7 (CLCN7), UDP-N-acetyl-alpha-D-galactosamine:polypeptide N-acetylgalactosaminyltransferase 3 (GALNT3), integrin-binding sialoprotein (IBSP), latent transforming growth factor beta binding protein 3 (LTBP3), R-spondin 3 (RSPO3), and SRY (sex determining region Y)-box 4 (SOX4). Some of these genes (e.g., LTBP3) were not significant at the genome-wide significance level in GEFOS-2, although the samples used in this study were included in GEFOS-2.

Organic integration of information from different levels and disciplines may help identify genetic variants of modest or moderate effects. By integrating GWAS in Caucasians and expression signature profiling of various human tissues, Hsu et al.[27] identified three novel genes, member of RAS oncogene family (RAP1A), TBC1 domain family, member 8 (with GRAM domain) (TBC1D8), and oxysterol binding protein-like 1A (OSBPL1A), for BMD, and replicated a well-known gene, OPG. They also prioritized 16 suggestive genome-wide significant candidate genes, among which two candidate genes, G protein-coupled receptor 177 (GPR177) and SOX6, were replicated in GEFOS-2. The samples used in this study (mainly from the Framingham Osteoporosis Study) were included in GEFOS-1 and GEFOS-2. Therefore, it does not represent an independent replication. The three identified novel genes (i.e., RAP1A, TBC1D8, and OSBPL1A) were not significant at the genome-wide level in either GEFOS-1[11] or GEFOS-2.[12]

Gene expression profiling analyses in this study were conducted in lymphocytes and the liver whose expression profiles may not be directly related to bone. Moreover, it is the differential gene expression with regard to osteoporosis-related phenotypes, not simply expression of genes, which is more clinically relevant to risk of osteoporosis and OF.[28,29]

(2) Multi-ethnicities

To investigate potential population generality/specificity of genes/loci, several GWASs were conducted in populations of multiple ethnicities. Xiong et al.[30] used a US Caucasian cohort as the discovery sample followed by replication analyses in independent populations including those of African (from the Tobago study) and Asian ancestry. Two novel genes, ADAM metallopeptidase with thrombospondin type 1 motif, 18 (ADAMTS18) and transforming growth factor, beta receptor III (TGFBR3), were identified at the genome-wide significance level. These two genes were found to be significant in other individual GWASs conducted in completely independent samples by independent investigators in premenopausal US Caucasian women[31] and in postmenopausal women with extremely discordant BMD values (28). However, neither of these genes was
found to be significant in the two GEFOS meta-analyses. [11,12]

Focusing on subjects of a specific gender and/or age range may reduce the potential confounding effects of heterogeneity, and empirically increase the power of a study. Koller et al.[31] conducted a GWAS in 1,524 pre-menopausal US Caucasian women aged 20–45 years and 669 African American premenopausal women aged 20–44 years. A novel gene, catper channel auxiliary subunit beta (CATSPERB), was identified as being significant for peak BMD. Despite the fact that samples from this study were included in GEFOS-2,[12] CATSPERB was not found to be significant in the meta-analyses.

(3) Children

Although the majority of GWASs have been focused exclusively on adults, Timpson et al.[32] published the first GWAS in 1,518 children from the Avon Longitudinal Study of Parents and Children. They identified association with BMD at the Osterix (SP7) gene locus, a transcription factor responsible for regulating osteoblast differentiation. In another study, Medina-Gomez et al.[33] identified a novel gene, Wnt16 that was associated with total body and skull BMD variation in children and adults, which suggested a role in peak bone mass accrual which may impact the risk of osteoporosis later in life. Interestingly, this locus was also associated with cortical bone thickness, wrist BMD, bone strength, and risk of forearm fracture in adults,[34] peak bone mass in premenopausal women,[17] and BMD and fracture in elderly individuals of European descent.[12]

(4) OF

The majority of the published GWASs of osteoporosis have been focused on BMD, a major risk factor for OF. However, OF is an important clinical end-point of osteoporosis, and is the most clinically relevant trait. Earlier studies have demonstrated a high genetic determination for OF risk,[13] and have also shown that BMD and fracture may have different genetic determination,[7,8] indicating the necessity to perform genetic studies on OF per se. Over the past several years, OF, as an independent study phenotype, has received increased attention in the bone genetics field. [35,36] Currently, a major problem facing genetic studies of OF is the difficulty in recruiting sufficiently large and homogeneous samples of the same OF type.

The first published GWAS for OF used 700 elderly Chinese Han subjects (350 with hip OF and 350 healthy matched controls) in the discovery phase, followed by replication in an independent Chinese sample containing 390 cases with hip OF and 516 controls.[37] A novel gene, aldehyde dehydrogenase 7 family, member A1 (ALDH7A1), was identified to be significant for hip OF. Further analyses confirmed its relevance to hip BMD in both Chinese (sample size, 2,955) and Caucasian (sample size, 7,007) populations. Although the discovery sample size was relatively small, the study was conducted in a homogeneous sample of the same ethnicity and the same type of carefully clinically adjudicated OF, which may empirically increase the power of gene identification. Notably, ALDH7A1 was not found to be significant in GEFOS-2.

Hwang et al.[38] published a GWAS for OF in East Asian populations. They first conducted a GWAS in the discovery cohort which included 288 cases and 1,139 controls, followed by a two-stage de novo replication analyses which included 462 cases and 1,745 controls in stage 1 and 369 cases and 560 controls in stage 2. A new locus associated with OF (rs784288 in the MECOM gene) showed genome-wide significance ($P=3.59 \times 10^{-8}$; OR 1.39). Further RNA interference analysis revealed that a MDS1 and EVI1 complex locus (MECOM) knockdown suppresses osteoclastogenesis.

An alternative approach for studying the genetic determination of OF is to first identify BMD loci, and then test the relevance of those loci to OF. This approach has been used for individual GWASs[23,24,30] and for GEFOS-2 (Table 2).[12] Using this approach, GEFOS-2 identified 14 BMD loci that were associated with risk of OF. One potential problem with the GEFOS-2 OF analyses is that it combined OFs from a variety of clinically heterogeneous types of OFs, thus departing from the assumption of homogeneity made by meta-analysis for the phenotype. From perspective of bone biology, the proportions of cortical and cancellous bone differ at the different sites in the skeleton (e.g., hip, spine and wrist) where OFs frequently occur.[39] The differences in behavior of bone at the different sites are most likely caused by the different environments of the bone cells in cortical or cancellous bone.[39] Moreover, each type of fracture has its own unique risk factors.[2] For instance, fall is a major risk factor for hip and wrist fractures; however, it contributes little to the risk of vertebral fracture. Epidemiology data also show that the prevalence
of fracture at different sites is different in terms of age, gender and ethnicity.[2,40] So far, the majority of the genetic variants identified by GWASs or candidate gene studies for BMD at different skeletal sites do not overlap.[5,10] Given these evidences, it is likely that different skeletal sites may not share much common genetic component. Therefore, the mixing of phenotypes in GEFOS-2 meta-analysis complicates the interpretation of results, as increasing sizes of non-homogeneous samples may lead to false positive/negative results. Although meta-analysis can be very powerful, its application must be executed with caution.

2. GWASs based on CNVs

SNP-based GWASs, including GEFOS meta-analyses, have discovered genes/loci which, collectively, account for less than 6% of the risk for developing osteoporosis. Therefore, it is important to explore the possibility that some of the remaining undiscovered genetic factors that influence risk of osteoporosis are due to genomic mechanisms other than individual nucleotide changes, such as CNVs.

CNV is a common type of genomic variability, with variations in the size of DNA fragments ranging from 1 kilobase to several megabases. CNVs, which may include duplications or deletions, can influence gene expression by disrupting coding sequences, perturbing long-range gene regulation, or altering gene dosage, and these effects could contribute to phenotypic variations or disease risk.[41]

A CNV-based GWAS for osteoporosis was conducted by Yang et al.[42] in 700 elderly Chinese individuals comprising 350 cases with homogeneous hip OF and 350 matched controls. In that study, CNV 4q13.2 was strongly associated with OF. Further validation experiments identified a deletion variant of UDP glucuronosyltransferase 2 family, polypeptide B17 (UGT2B17) in CNV 4q13.2, and the association between CNV of UGT2B17 and OF was replicated in an independent Chinese sample containing 399 cases with hip OF and 400 controls.[42] Despite the relatively small sample sizes, the CNV focused GWAS identified novel genes/loci that were otherwise missed in SNP-based GWASs.

Oei et al.[43] performed a genome-wide CNV analysis in 5,178 individuals from Netherlands, including 809 OF cases, and followed by in silico lookups and de novo replication in several independent samples. A rare (population prevalence 0.14%) 210 kb deletion located on chromo-some 6p25.1 was associated with the risk of fracture. The prevalence of this deletion showed geographic diversity, being absent in samples from Australia, Canada, Poland, Iceland, Denmark, and Sweden, but present in the Netherlands (0.34%), Spain (0.33%), USA (0.23%), England (0.15%), Scotland (0.10%), and Ireland (0.06%). Larger and geographically restricted studies are needed to confirm this regional association.

3. GWASs based on biological pathways

Conventional GWASs consider the effects of individual genetic markers independently, and focus only on genes or markers of top-ranking statistical significance. However, genes and/or their products often work together, interacting in functional groups or pathways to contribute to phenotypic variation or susceptibility to disease. Therefore, GWASs which focus on individual genes/loci may not be optimally effective for identifying pathophysiologically significant genetic pathways underlying osteoporosis and OF. A major advantage of pathway-based GWASs is that they link the wealth of information embedded in GWAS data to our knowledge of functional biological pathways that is readily available in public databases.

Zhang et al.[44] performed the first pathway-based GWAS for osteoporosis in a population of US Caucasians, and identified the importance of the regulation-of-autophagy (ROA) pathway for ultradistal radius BMD. This is the first study implicating the ROA pathway’s importance to osteoporosis, though autophagy had previously been recognized to be important in endochondral ossification.[45] Another pathway-based GWAS conducted in 700 elderly Chinese Han subjects (350 with hip OF and 350 healthy matched controls) revealed the importance of the tumor necrosis factor (TNF)-related apoptosis-inducing ligand (TRAIL) pathway for hip OF.[46] In the pathway-based analyses, the maximum statistic for all SNPs near a gene was taken to represent the significance of the gene and use a permutation-based approach that shuffles the phenotypes to adjust for multiple testing.[47] Notably, most individual genes included in these significant pathways did not reach genome-wide significance in SNP-based GWASs, underscoring the importance of looking beyond the top significant SNPs/genes and searching for additional risk factors with moderate statistical significance.
4. Bivariate GWASs

Osteoporosis is highly correlated, both genetically and environmentally, with other human diseases/conditions such as obesity (characterized by high body mass index [BMI]) and sarcopenia (characterized by low lean mass and strength). GWASs, however, have typically been implemented in a univariate framework, analyzing different phenotypes (e.g., BMD or BMI) independently, and ignoring the potential genetic correlation between disease traits. Thus, this approach is not well suited for detecting pleiotropic genes, which may exist for two genetically correlated diseases. One potentially effective strategy for identifying pleiotropic genes is to analyze potentially correlated disease phenotypes simultaneously via a multivariate GWAS approach.[47] This new strategy, which utilizes GWAS data more efficiently than univariate approaches, may help identify pleiotropic genes underlying diseases of shared genetic susceptibility, thereby revealing interconnected pathophysiological networks for a spectrum of common human diseases.

The first bivariate GWAS analysis for osteoporosis and obesity, conducted in 1,000 unrelated US Caucasians, suggested the SOX6 gene’s importance in co-regulation of osteoporosis and obesity.[48] This gene was not significant at the genome-wide level in separate analyses for either BMD or BMI, highlighting the power of bivariate analyses in discovering novel pleotropic genes/loci. The SOX6 gene was independently found to be significant at the genome-wide level in GEFOS-2[12] and was suggestively significant at the genome-wide level in the study of Hsu et al.[27] Identifying genes with pleiotropic effects, such as SOX6, may offer deeper insights into the pathophysiology of osteoporosis, and potentially, may reveal key mechanistic links between osteoporosis and obesity, two major public health problems.

5. The first whole genome sequencing study

GWASs typically identify common variants important for common human disorders. Many susceptible rare variants may be missed in GWASs due to the limited power and/or resolution of genotyping.[49,50] Recent advances in next-generation sequencing technologies have greatly enhanced our ability to discover functional rare variants.[51] Styrkarsdottir et al.[9] published the first whole genome sequencing study for osteoporosis. Through whole-genome sequencing of 2,230 Icelandic individuals with discordant BMD values, they found a rare nonsense mutation within the leucine-rich repeat containing G protein-coupled receptor 4 (LGR4) gene that was strongly associated with low BMD and OF. This mutation leads to termination of LGR4 at position 126 and fully disrupts its function. A potential problem with OF analysis is that all types of fractures were grouped together, which may introduce false positive or negative results as aforementioned. The phenotype of carriers of the c.376C>T mutation overlaps that of Lgr4 mutant mice.[9] Interestingly, although this mutation was associated with a wide range of phenotypes across species (i.e., humans and mice), it was not present in Danish and Australians.[9] Therefore, its effects in other human populations need to be further evaluated and validated.

INTERPRETATION OF INCONSISTENT RESULTS

Through the identification of novel genes, CNVs, and biological pathways that are associated with osteoporosis, recent GWASs and large-scale meta-analyses have greatly advanced our understanding of the pathophysiology of osteoporosis and OF. As illustrated above, however, comparison of the results across individual GWASs and meta-analyses raises a number of critical questions.

1) Are individual GWASs still useful when meta-analysis, with a much larger sample size, is available?

2) Why did some genes/loci identified as being significant in independent individual GWASs (e.g., ADAMTS18 in Ref [30]) fail to attain significance in independent GEFOS-2 meta-analyses,[12] which had a much larger sample size?

3) Why did some genes/loci identified as being significant in individual GWASs (e.g., RAP1A, TBC1D8, and OSBP-L1A in Ref [27]) fail to attain significance at the genome-wide level in GEFOS-2, in circumstances where the samples used in the individual GWASs were included in GEFOS-2 meta-analyses?[12]

4) Why have inconsistent findings even been observed between meta-analyses (e.g., GEFOS-1[11] and GEFOS-2[12] ) whose samples overlapped to a large extent?

5) Should the meta-analysis with the largest number of individuals be considered as the gold standard to evaluate findings of other independent studies (especially relatively small GWASs)?
To address these critical questions, we performed a series of theoretical analyses using simulation studies and published our results elsewhere.[52] Our theoretical analyses, under ideal situations, demonstrated that:

1) Although the power of an individual GWAS study (of average sample size) to identify any particular locus (of average effect size) is limited, the power to identify at least one (any one) locus can be high. This may explain the observation that a number of previous individual GWASs have identified novel loci despite the relatively limited sample size of each of the studies. Given the anticipated large number of significant loci that have eluded detection thus far, individual studies are still valuable in identifying at least some of these underlying effect loci.

2) The number of loci that can be detected in meta-analyses greatly exceeds the number that can be detected in individual GWASs. However, the power of a meta-analysis to identify many independent loci simultaneously can still be limited.

3) The meta-analysis has rather limited power to replicate the findings of particular loci from individual GWASs at the genome-wide significance level, particularly for SNPs with small effects, implying inconsistent findings between independent individual GWASs and meta-analysis is not unexpected.

4) Adding heterogeneous samples into a subset of homogeneous samples can reduce power for a meta-analysis, rather than having the anticipated effect of increasing power due to increased sample size. This was clearly shown in our published theoretical study[52] which examined the effects of sample heterogeneity by simulating a subset of samples with true effects and the other subset of samples without effects in a meta-analysis (based on GEFOS-2). Figure (adapted from [52]) illustrates that heterogeneity results in power loss in both the mixed-effects and random-effects models. This effect was further demonstrated in our more comprehensive theoretical analyses which considered a wide range of situations and scenarios.[53] Therefore, selecting samples with homogeneous effects may be at least as important as enlarging sample size for meta-analysis.

This observation may help explain the inconsistent findings between GEFOS-1 [11] and GEFOS-2 [12] despite the fact that the latter included the majority of samples in the former, and approximately 3 times as many total samples.

Our theoretical analysis was not intended to denigrate the value and importance of GWAS meta-analysis. Large-scale meta-analysis clearly represents a powerful tool for identifying novel genetic variants for osteoporosis. Nevertheless, it is critical to recognize that meta-analysis may introduce false positive/negative results due to heterogeneity and other confounding factors. Consequently, it is important to exert caution when designing studies, and interpreting results generated by meta-analysis of GWASs. In particular, the results of meta-analysis should not be used to evaluate the validity of findings from individual GWASs. Well-designed individual GWASs, using homogenous samples, have significant potential to identify additional novel genes/loci and related biological pathways for osteoporosis and OF that may not necessarily be significant in meta-analysis with larger samples, of more variable heterogeneity.

**IMPLICATIONS AND PERSPECTIVES**

Over the past 4 years, individual GWASs and meta-analyses of GWASs have highlighted more than 70 genes/loci and related biological pathways that contribute to the pathophysiology of osteoporosis and/or OF. Some of these genes, such as WNT, RANK-RANKL-OPG, have known functional relevance to bone metabolism and endochondral ossification, and their contribution to osteoporosis has been well established in earlier candidate gene studies. [5,54,55] The function of these established genes, proteins
and related biological pathways has been reviewed elsewhere.[10] However, the purpose of GWASs is not merely to verify previous findings but, more importantly, to identify novel genes and pathways associated with complex diseases. It is notable that more than half of the genes/loci listed in Table 1 are novel, though their functional importance to bone metabolism awaits validation, ultimately through molecular functional studies.

Collectively, the genes/loci identified from individual GWASs and meta-analyses, to date, explain less than 6% of the variance in BMD variation. Therefore, further endeavors are needed to explore undiscovered genetic factors associated with BMD variation. At the DNA level, there are several possible paths towards uncovering these novel and elusive genes:

1) Well-designed, powerful individual GWASs. For such studies to be powerful, well-defined and homogeneous phenotypes measured with high data quality and accuracy, large and homogeneous samples, and comprehensive statistical and bioinformatical analyses should be needed.

2) Meta-analyses with even larger samples than GE-FOS-2, which must be executed with caution because of the power loss associated with between-study heterogeneity and other confounding factors.

3) Meta-analyses with smaller sample sizes, but with less between-sample heterogeneity, to reduce the probability of generating false positive and false negative results.

4) Utilizing available GWAS data to perform additional analyses such as CNV, pathway based, and multivariate analyses.

5) Performing genetic studies focused directly on OF, rather than BMD or other less critical osteoporosis phenotypes. For OF studies, large and homogeneous samples of the same type of OF are critically important and essential. Although it is tempting to mix different types of OFs to increase sample size, heterogeneity may contribute to false findings and results that complicate data interpretation.

Current evidence suggests that the genetic architecture of osteoporosis and OF is complex, involving both common and rare functional variants.[50,51] These findings are similar for other complex human diseases that represent significant public health problems (e.g., obesity, diabetes, and cancers). GWASs and meta-analyses are largely designed to identify common variants. Thus, many susceptible rare variants may be missed in GWASs due to the limited power and/or resolution of genotyping. Recent advances in next-generation sequencing technologies, however, have greatly enhanced our ability to discover functional rare variants.[51] Unfortunately, large-scale whole genome re-sequencing studies are currently prohibitively expensive. More economically feasible approaches might involve re-sequencing of targeted genes/loci, or exome sequencing studies. Another potential approach to identify rare variants is to perform imputation analyses based on publicly available genome sequence data. The recent 1,000 Genomes Project (http://www.1000genomes.org/) produced a comprehensive catalog of human genomic variants, in particular rare variants.[56,57] Thus, through imputation of currently available GWAS samples, it is now feasible to identify/infer the majority of known human genomic variants (including rare variants) for these association studies. As technological advances decrease the unit price for genome re-sequencing, it is expected that large and powerful genome-wide re-sequencing studies will become feasible in the near future, resulting in the identification of numerous rare genetic variants that effect osteoporosis and OF. It is critical to recognize that in order to identify rare variants with reasonable power, re-sequencing studies will require very large sample sizes due to the low minor allele frequencies of these rare variants.

GWASs and re-sequencing studies focus on DNA sequence variants alone. Recent studies, however, have shown that epigenetic regulation is also critical to the pathophysiology of many human complex diseases, and may partially account for the heritability that has not been accounted for based solely on DNA variants.[58] Epigenetics refers to reversible, heritable changes in gene regulation that occur without a change in DNA sequence.[59] Consequently, in order to identify additional and novel heritable factors contributing to BMD variation and osteoporosis risk, it has become necessary and timely to study epigenomic regulation in relevant bone-related cells, including DNA methylation, histone modification, microRNA and long non-coding RNAs.

Protein post-translational modifications, such as protein phosphorylation, have also been shown to play significant roles in regulating gene expression,[60,61] signal transduction,[62,63] and cellular functions[64] closely related to bone metabolism. Functional studies at the protein level, including protein expression[28,65] and post-translational
modification, may also contribute to our comprehensive identification and understanding of cellular and molecular mechanisms regulating bone metabolism.

GWASs are ultimately studies at the DNA level, and the above comments illustrate the importance of studying complex human diseases from a systems biology perspective.[66,67] Gene expression is a complex process that is regulated simultaneously and interactively at DNA, RNA, protein, epigenomic and environmental levels. Therefore, a genomic convergence or systems biology approach that organically integrates the information from GWASs, gene expression, proteomics, epigenomics, protein post-translational modification, and gene-environment studies may help facilitate the identification of key pathways that are globally involved in the pathogenesis of osteoporosis and OF. For example, using a systems genetic analytic approach, Calabrese et al. identified a physiologically relevant gene network and used it to discover novel genes and regulatory mechanisms involved in the function of osteoblast-lineage cells.[68] In another study, Deng et al. ascertained SOD2 as a susceptibility gene for osteoporosis in Chinese by integrating evidence from DNA, RNA, and protein levels.[69] Ultimately, the functional relevance of the identified variants needs to be confirmed by in vivo and/or in vitro molecular biology studies.

REFERENCES

1. Cummings SR, Kelsey JL, Nevitt MC, et al. Epidemiology of osteoporosis and osteoporotic fractures. Epidemiol Rev 1985;7:178-208.
2. Cummings SR, Melton LJ. Epidemiology and outcomes of osteoporotic fractures. Lancet 2002;359:1761-7.
3. Ralston SH. Genetics of osteoporosis. Rev Endocr Metab Disord 2001;2:13-21.
4. Recker RR, Deng HW. Role of genetics in osteoporosis. Endocrine 2002;17:55-66.
5. Xu XH, Dong SS, Guo Y, et al. Molecular genetic studies of gene identification for osteoporosis: the 2009 update. Endocr Rev 2010;31:447-505.
6. Livshits G, Deng HW, Nguyen TV, et al. Genetics of bone mineral density: evidence for a major pleiotropic effect from an intercontinental study. J Bone Miner Res 2004;19:914-23.
7. Deng HW, Mahaney MC, Williams JT, et al. Relevance of the genes for bone mass variation to susceptibility to osteoporotic fractures and its implications to gene search for complex human diseases. Genet Epidemiol 2002;22:12-25.
8. Michaëlsson K, Melhus H, Ferm H, et al. Genetic liability to fractures in the elderly. Arch Intern Med 2005;165:1825-30.
9. Styrkarsdottir U, Thorleifsson G, Sulem P, et al. Nonsense mutation in the LGR4 gene is associated with several human diseases and other traits. Nature 2013;497:517-20.
10. Richards JB, Zheng HF, Spector TD. Genetics of osteoporosis from genome-wide association studies: advances and challenges. Nat Rev Genet 2012;13:576-88.
11. Rivadeneira F, Styrkárdsdottir U, Estrada K, et al. Twenty bone-mineral-density loci identified by large-scale meta-analysis of genome-wide association studies. Nat Genet 2009;41:1199-206.
12. Estrada K, Styrkársdottir U, Evangelou E, et al. Genome-wide meta-analysis identifies 56 bone mineral density loci and reveals 14 loci associated with risk of fracture. Nat Genet 2012;44:491-501.
13. Deng HW, Chen WM, Recker S, et al. Genetic determination of Colles' fracture and differential bone mass in women with and without Colles' fracture. J Bone Miner Res 2000;15:1243-52.
14. Richards JB, Kavvoura FK, Rivadeneira F, et al. Collaborative meta-analysis: associations of 150 candidate genes with osteoporosis and osteoporotic fracture. Ann Intern Med 2009;151:528-37.
15. Fink HA, Harrison SL, Taylor BC, et al. Differences in site-specific fracture risk among older women with discordant results for osteoporosis at hip and spine: study of osteoporotic fractures. J Clin Densitom 2008;11:250-9.
16. Augat P, Reeb H, Claes LE. Prediction of fracture load at different skeletal sites by geometric properties of the cortical shell. J Bone Miner Res 1996;11:1356-63.
17. Koller DL, Zheng HF, Karasik D, et al. Meta-analysis of genome-wide studies identifies WNT16 and ESR1 SNPs associated with bone mineral density in premenopausal women. J Bone Miner Res 2013;28:547-58.
18. Zhang L, Choi HJ, Estrada K, et al. Multistage genome-wide association meta-analyses identified two new loci for bone mineral density. Hum Mol Genet 2014;23:1923-33.
19. Paternoster L, Lorentzon M, Lehtimäki T, et al. Genetic determinants of trabecular and cortical volumetric bone mineral densities and bone microstructure. PLoS Genet 2013;9:
20. Lee DC, Gilsanz V, Wren TA. Limitations of peripheral quantitative computed tomography metaphyseal bone density measurements. J Clin Endocrinol Metab 2007;92:4248-53.
21. Adams JE. Quantitative computed tomography. Eur J Radiol 2009;71:415-24.
22. Engelke K, Adams JE, Armbrecht G, et al. Clinical use of quantitative computed tomography and peripheral quantitative computed tomography in the management of osteoporosis in adults: the 2007 ISCD Official Positions. J Clin Densitom 2008;11:123-62.
23. Richards JB, Rivadeneira F, Inouye M, et al. Bone mineral density, osteoporosis, and osteoporotic fractures: a genome-wide association study. Lancet 2008;371:1505-12.
24. Styrkarsdottir U, Halldorsson BV, Gretarsdottir S, et al. Multiple genetic loci for bone mineral density and fractures. N Engl J Med 2008;358:2355-65.
25. Deng HW, Chen WM. QTL fine mapping, in extreme samples of finite populations, for complex traits with familial correlation due to polygenes. Am J Hum Genet 2000;67:259-62.
26. Duncan EL, Danoy P, Kemp JP, et al. Genome-wide association study using extreme truncate selection identifies novel genes affecting bone mineral density and fracture risk. PLoS Genet 2011;7:e1001372.
27. Hsu YH, Zillikens MC, Wilson SG, et al. An integration of genome-wide association study and gene expression profiling to prioritize the discovery of novel susceptibility Loci for osteoporosis-related traits. PLoS Genet 2010;6:e1000977.
28. Deng FY, Lei SF, Zhang Y, et al. Peripheral blood monocyte-expressed ANXA2 gene is involved in pathogenesis of osteoporosis in humans. Mol Cell Proteomics 2011;10:M111.011700.
29. Liu YZ, Dvornyk V, Lu Y, et al. A novel pathophysiological mechanism for osteoporosis suggested by an in vivo gene expression study of circulating monocytes. J Biol Chem 2005;280:29011-6.
30. Xiong DH, Liu XG, Guo YF, et al. Genome-wide association and follow-up replication studies identified ADAMTS18 and TGFBR3 as bone mass candidate genes in different ethnic groups. Am J Hum Genet 2009;84:388-98.
31. Koller DL, Ichikawa S, Lai D, et al. Genome-wide association study of bone mineral density in premenopausal European-American women and replication in African-American women. J Clin Endocrinol Metab 2010;95:1802-9.
32. Timpson NJ, Tobias JH, Richards JB, et al. Common variants in the region around Osterix are associated with bone mineral density and growth in childhood. Hum Mol Genet 2009;18:1510-7.
33. Medina-Gomez C, Kemp JP, Estrada K, et al. Meta-analysis of genome-wide scans for total body BMD in children and adults reveals allelic heterogeneity and age-specific effects at the WNT16 locus. PLoS Genet 2012;8:e1002718.
34. Zheng HF, Tobias JH, Duncan E, et al. WNT16 influences bone mineral density, cortical bone thickness, bone strength, and osteoporotic fracture risk. PLoS Genet 2012;8:e1002745.
35. Nguyen TV, Eisman JA. Genetic profiling and individualized assessment of fracture risk. Nat Rev Endocrinol 2013;9:153-61.
36. Chen Y, Shen H, Yang F, et al. Choice of study phenotype in osteoporosis genetic research. J Bone Miner Metab 2009;27:121-6.
37. Guo Y, Tan LJ, Lei SF, et al. Genome-wide association study identifies ALDH7A1 as a novel susceptibility gene for osteoporosis. PLoS Genet 2010;6:e1000806.
38. Hwang JY, Lee SH, Go MJ, et al. Meta-analysis identifies a MECOM gene as a novel predisposing factor of osteoporotic fracture. J Med Genet 2013;50:212-9.
39. Rosen CJ. Primer on the metabolic bone diseases and disorders of mineral metabolism. 7th ed. Washington, DC: American Society for Bone and Mineral Research; 2008.
40. Melton LJ, 3rd. The prevalence of osteoporosis: gender and racial comparison. Calcif Tissue Int 2001;69:179-81.
41. McCarroll SA, Altshuler DM. Copy-number variation and association studies of human disease. Nat Genet 2007;39:S37-42.
42. Yang TL, Chen XD, Guo Y, et al. Genome-wide copy-number-variation study identified a susceptibility gene, UGT2B17, for osteoporosis. Am J Hum Genet 2008;83:663-74.
43. Oei L, Hsu YH, Styrkarsdottir U, et al. A genome-wide copy number association study of osteoporotic fractures points to the 6p25.1 locus. J Med Genet 2014;51:122-31.
44. Zhang L, Guo YF, Liu YZ, et al. Pathway-based genome-wide association analysis identified the importance of regulation-of-autophagy pathway for ultradistal radius BMD. J Bone Miner Res 2010;25:1572-80.
45. Bohensky J, Shapiro IM, Leshinsky S, et al. PIM-2 is an independent regulator of chondrocyte survival and autophagy in the epiphyseal growth plate. J Cell Physiol 2007;
46. Zhang YP, Liu YZ, Guo Y, et al. Pathway-based association analyses identified TRAIL pathway for osteoporotic fractures. PLoS One 2011;6:e21835.
47. Wang K, Li M, Bucan M. Pathway-based approaches for analysis of genomewide association studies. Am J Hum Genet 2007;81:1278-83.
48. Liu YZ, Pei YF, Liu JF, et al. Powerful bivariate genomewide association analyses suggest the SOX6 gene influencing both obesity and osteoporosis phenotypes in males. PLoS One 2009;4:e6827.
49. Gorlov IP, Gorlova OY, Sunyaev SR, et al. Shifting paradigm of association studies: value of rare single-nucleotide polymorphisms. Am J Hum Genet 2008;82:100-12.
50. Bodmer W, Bonilla C. Common and rare variants in multifactorial susceptibility to common diseases. Nat Genet 2008;40:695-701.
51. Cirulli ET, Goldstein DB. Uncovering the roles of rare variants in common disease through whole-genome sequencing. Nat Rev Genet 2010;11:415-25.
52. Liu YJ, Zhang L, Pei Y, et al. On genome-wide association studies and their meta-analyses: lessons learned from osteoporosis studies. J Clin Endocrinol Metab 2013;98:E1278-82.
53. Pei YF, Zhang L, Papasian CJ, et al. On individual genomewide association studies and their meta-analysis. Hum Genet 2014;133:265-79.
54. Liu YJ, Shen H, Xiao P, et al. Molecular genetic studies of gene identification for osteoporosis: a 2004 update. J Bone Miner Res 2006;21:1511-35.
55. Liu YZ, Liu YJ, Recker RR, et al. Molecular studies of identification of genes for osteoporosis: the 2002 update. J Endocrinol 2003;177:147-96.
56. Abecasis GR, Altshuler D, Auton A, et al. A map of human genome variation from population-scale sequencing. Nature 2010;467:1061-73.
57. Mills RE, Walter K, Stewart C, et al. Mapping copy number variation by population-scale genome sequencing. Nature 2011;470:59-65.
58. Manolio TA, Collins FS, Cox NJ, et al. Finding the missing heritability of complex diseases. Nature 2009;461:747-53.
59. Rodenhiser D, Mann M. Epigenetics and human disease: translating basic biology into clinical applications. CMAJ 2006;174:341-8.
60. Teitelbaum SL. RANKing c-Jun in osteoclast development. J Clin Invest 2004;114:463-5.
61. Jimi E, Akiyama S, Tsurukai T, et al. Osteoclast differentiation factor acts as a multifunctional regulator in murine osteoclast differentiation and function. J Immunol 1999;163:434-42.
62. Heinrich PC, Behrmann I, Haan S, et al. Principles of interleukin (IL)-6-type cytokine signalling and its regulation. Biochem J 2003;374:1-20.
63. Ihnatko R, Kubes M. TNF signaling: early events and phosphorylation. Gen Physiol Biophys 2007;26:159-67.
64. Milani R, Ferreira CV, Granjeiro JM, et al. Phosphoproteome reveals an atlas of protein signaling networks during osteoblast adhesion. J Cell Biochem 2010;109:957-66.
65. Deng FY, Liu YZ, Li LM, et al. Proteomic analysis of circulating monocytes in Chinese premenopausal females with extremely discordant bone mineral density. Proteomics 2008;8:4259-72.
66. Farber CR. Systems-level analysis of genome-wide association data. G3 (Bethesda) 2013;3:119-29.
67. Farber CR. Systems genetics: a novel approach to dissect the genetic basis of osteoporosis. Curr Osteoporos Rep 2012;10:228-35.
68. Calabrese G, Bennett BJ, Orozco L, et al. Systems genetic analysis of osteoblast-lineage cells. PLoS Genet 2012;8:e1003150.
69. Deng FY, Lei SF, Chen XD, et al. An integrative study ascertained SOD2 as a susceptibility gene for osteoporosis in Chinese. J Bone Miner Res 2011;26:2695-701.
70. Styrkarsdottir U, Halldorsson BV, Gudbjartsson DF, et al. European bone mineral density loci are also associated with BMD in East-Asian populations. PLoS One 2010;5:e13217.
71. Kung AW, Xiao SM, Cherny S, et al. Association of JAG1 with bone mineral density and osteoporotic fractures: a genome-wide association study and follow-up replication studies. Am J Hum Genet 2010;86:229-39.
72. Styrkarsdottir U, Halldorsso BV, Gudjartsson S, et al. New sequence variants associated with bone mineral density. Nat Genet 2009;41:15-7.
