Nitrite and Nitric Oxide as Potential Diagnostic Markers in Acute Vascular Diseases

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Introduction

Nitric oxide (NO) is a signaling molecule involved in numerous biological pathways, including multiple disease and injury processes involving ischemia, infection, and inflammation. The biological role of the NO molecule was discovered in the 1980’s, and its discoverers, Robert Furchgott, Louis Ignarro and Ferid Murad, shared the 1998 Nobel Prize in Physiology or Medicine. A quick Medline search for “nitric oxide” will yield over 100,000 references, indicating its ubiquity and importance. NO is known to be a key molecule regulating arterial blood flow [1,2], and there is a significant body of literature regarding therapeutic applications of NO and pharmaceutical agents for delivery of NO. One of the most notable examples of a therapeutic compound which acts to increase the signaling capacity of NO (by inhibiting the consumption of cyclic guanosine monophosphate (cGMP)) is sildenafil citrate (Viagra). Inhaled NO is used therapeutically for respiratory complications in premature infants. As a diagnostic, exhaled NO is used primarily on the potential utility of monitoring plasma nitrite, which is circulating storage pool for NO, rapidly producing NO under ischemic conditions, with multiple potential therapeutic applications now under investigation. The ability to monitor nitrite levels in real time could prove important for the therapeutic use of nitrite. This review will focus primarily on the potential utility of monitoring plasma nitrite, which is under investigation as a therapeutic agent, and on the prognostic and diagnostic value of nitrite for various diseases.

Nitric Oxide role in vascular biology

NO is produced by the conversion of L-arginine to L-citrulline by one of three enzymes, the nitric oxide synthases (NOS’s) including endothelial NOS (eNOS) neuronal NOS (nNOS) and inducible NOS (iNOS). The former two enzymes are referred to as constitutive NOS’s, and are dependent on calcium ions for activity, while iNOS is calcium-independent. nNOS is predominantly expressed in neurons and muscle, whereas eNOS is predominantly expressed in endothelial cells, and iNOS is expressed by macrophages. A variety of cell types and tissues express these isoforms, with many tissues expressing more than one isoform [5]. There are 5 enzyme co-factors for the NOS enzymes, including flavin adenine dinucleotide (FAD), flavin mononucleotide (FMN), heme, tetrahydrobiopterin (BH4) and calmodulin, so NO is tightly regulated under normal physiological conditions. NOS proteins first bind to the cofactors FAD and FMN. The additions of L-arginine, BH4 and heme allow the NOS protein to form dimers. eNOS and nNOS dimers formed this way are inactive, and depend on calmodulin binding stimulated by increases in intracellular calcium. In contrast, iNOS dimers bind calcium/calmodulin and are active even at low (resting intracellular) concentrations of calcium. Thus, nNOS and eNOS are activated by a transient increase in intracellular calcium concentration, whereas iNOS is activated at the level of transcription [5]. The different isoforms of NOS are found in a variety of cell types and tissues, including neuronal cells and vascular endothelial cells.

eNOS is of particular importance for cardiovascular physiology, as it maintains basal vascular tone through release of low levels of NO.
NO generated by eNOS diffuses from the endothelium to smooth muscle cells where it binds to and activates soluble guanylate cyclase (sGC) resulting in increased levels of cGMP [7]. cGMP activates protein kinase G (PKG) causing relaxation of smooth muscle cells by opening K+ channels and/or by reducing the sensitivity of the contractile machinery to Ca2+ [8,9]. This pathway is considered the predominant mechanism for NO-mediated dilation of cerebral arteries and arterioles. The regulation of eNOS is multifaceted. Calcium is an important regulator of eNOS, and many agonists activate eNOS by increasing intracellular Ca2+ [10]. Calcium complexes with calmodulin and activates eNOS by displacing it from cave-1 [11]. Additionally, eNOS in cerebral arteries and arterioles appears to be directly regulated by heat shock protein (HSP90), serine/threonine phosphorylation, and tyrosine phosphorylation. HSP90 and eNOS co-localize within endothelial cells [12]. Geldanamycin, an inhibitor of HSP90, reduces cGMP levels and constricts cerebral arteries to a similar extent as NO inhibition [12].

Nitric oxide production is altered in many pathological states. Endothelial cells, neurons, glia, and invading leukocytes all produce reactive oxygen species (ROS) such as superoxide anion (O2•−) and peroxynitrite (ONOO−) which can scavenge NO, cause endothelial dysfunction, and alter vascular tone [13]. Ironically, under conditions of limited substrate availability, the conversion of L-arginine to NO, superoxide anion, and peroxynitrite mediates the damage secondary to both brain trauma and ischemia/reperfusion injury [15-17].

Recently, an alternative pathway for generation of NO has been discovered [18]. The anion nitrite was previously considered physiologically inert, a stable end product of nitric oxide (NO) metabolism, and a biomarker for NO activity. Nitrite has been shown to be a circulating storage pool for NO [18,19] that the body can rapidly activate when needed. Rapid conversion of nitrite to NO can occur under ischemic conditions (low pH, low partial pressure of oxygen (pO2)) [20]. There are several enzymes which may act to reduce nitrite to NO, including deoxyhemoglobin, deoxymyoglobin, xanthine oxidoreductase (XOR), neuroglobin, eNOS, and components of the mitochondrial electron transport chain [22]. The deoxyhemoglobin state of hemoglobin has now been characterized as an allosterically regulated nitrite reductase. The different nitrite reductase “enzyme” systems operate along a range of physiological and pathological hypoxia, with hemoglobin reducing nitrite at an oxygen tension from 60 mm Hg down to 20 mm Hg, myoglobin active below 4 mm Hg, and xanthine oxidoreductase and acidic reduction nitrite at zero oxygen and low pH [21,22]. This allows for graded nitrite reduction to NO along the circulating and metabolic oxygen gradient [23]. In addition, as pH decreases, the rate of the reaction increases [24]. Maximal physiological dilatation via this mechanism is proposed to occur at around 50% hemoglobin oxygen saturation (HbO2 sat (%)) [25]. Conversion of nitrite to NO by deoxyhemoglobin can occur in as little as 30 seconds to several minutes [26].

NO generated by NOS during normoxia may be chemically stored in a nitrite reservoir and re-generated during hypoxia and ischemia when oxygen-dependent NOS function is limited, and in areas in which increased blood flow would be desirable [27,28]. Nitrite represents the largest known source of bioavailable NO in the circulation, and can be considered an ischemic NO reservoir. There is an oxygen-dependent balance between NO and nitrite centered on hemoglobin in red blood cells. Nitrite ions produced in plasma from oxidation of intravascular NO by ceruloplasmin can be taken up by erythrocytes. Nitrite uptake into RBCs involves both diffusion of protonated nitrite (HNO3) and facilitated diffusion of NO2− via anion-exchanger-1 (AE1) [29]. Following its entry into the RBCs, nitrite reacts with oxy- and deoxyhemoglobin and undergoes oxidative and reductive reactions. As hemoglobin within erythrocytes becomes deoxygenated, intracellular nitrite may be catalytically reduced to NO, which may then be released from the cell. In the presence of oxyhemoglobin, intraerythrocytic nitrite, which constitutes the majority of intravascular nitrite, reacts to produce methemoglobin and nitrate. NO in plasma is largely destroyed by reaction with trace amounts of plasma hemoglobin or may diffuse into erythrocytes to react with oxyhemoglobin to produce nitrate and methemoglobin. When NO reacts with deoxyhemoglobin, it produces primarily iron-nitrosyl-hemoglobin (NO-Hb) [2].

Nitrite is found in both plasma and erythrocytes, and there is an arterial to venous gradient in the nitrite levels in plasma, erythrocytes, and whole blood [30,31]. Nitrite levels decrease with blood deoxygenation from artery to vein, consistent with a dynamic oxygen-dependent nitrite metabolism. Nitrite gradients have been taken to signify arterio-venous nitrite consumption eliciting vasodilatation. However, these gradients may also reflect the difference between nitrite production due to NOS activity, where the arterial contribution is greater, and nitrite consumption due to nitrite reductase activity, where the venous contribution is greater [25]. Nitrite has become widely accepted as a marker of nitric oxide, since there is an artery-to-vein gradient [3], indicating the consumption of the molecule during exercise stress in the forearm circulation, but other molecules such as NO3− and SNO-Hb are not consumed. Gladwin et al. [32] reported that when humans were exposed to 80 ppm inhaled NO gas, there was an increase in peripheral forearm blood flow that was only associated with increases in plasma nitrite. However, in many studies investigators report only combined values for nitrite (NO2−) and nitrate (NO3−) (together commonly referred to as NO3−). Given the utility of nitrite as a marker for NO3− this can sometimes make interpretation of results difficult.

Plasma nitrite concentrations vary little between different mammals, indicating that the formation of nitrite is a common feature within the circulation preserved throughout evolution. In mammalian physiology, three sources of nitrite have been identified. The first significant source is dietary nitrite ingested from food. The second source is nitrite released during the reduction of dietary nitrate (NO3−). Considerable quantities of nitrate are contained in nitrate-rich vegetables such as beetroot, spinach, or dark green leafy vegetables [33]. Nitrate reductase enzymes are found in commensal bacteria in the mouth or intestines [33]. Such bacterial nitrate reductases contribute significantly to the endogenous nitrite pool of the host organism. There is some disagreement about the predominance of either dietary sources of nitrite or endogenous nitrite produced by ceruloplasmin-dependent oxidation of intravascular NO produced by eNOS [34]. Measured nitrite levels in whole blood or plasma show significant variability due to differences in dietary habits, lifestyle (e.g. tobacco consumption) and physical exercise prior to testing. Other investigators report that in humans and other mammals, ~90% of the circulating plasma nitrite is derived from eNOS. In fasting humans, almost all nitrite in the vascular circulation originates from NO released by nitric oxide synthases (NOS) [2]. Plasma nitrite levels are reduced up to 70% in eNOS knockout mice and upon acute NOS inhibition in wild-type mice. Plasma nitrite reflects a constant proportion of the total NO synthesis and is now widely accepted as an index of NOS activity in vivo [35].
Whole blood nitrite levels are similar at 2 or 4-hour intervals under fasting and non-fasting conditions, and are similar when drawn on 3 consecutive days with a coefficient of variation of <8% [2,35]. In resting humans, various values for plasma nitrite and nitrate concentrations have been reported, ranging from 0.1 – 0.5 μM for nitrite and 12-50 μM for plasma nitrate. In one study, the nitrite levels measured in plasma, red blood cells (RBCs) and whole blood, of 15 healthy volunteers were 121 ± 9, 288 ± 47, 176 ± 17 nM respectively [2]. Nitrite in erythrocytes constitutes about two thirds of intravascular nitrite at concentrations approaching 300 nM [2]. Virtually all nitrite (94 ± 4%) in erythrocytes is located in the cytosol [2]. Nitrite levels detected in blood are normally <1 μM. However, the nitrite levels in the tissues are much higher, up to 10 μM [36].

Prognostic value of plasma nitrite

In a study of plasma nitrite levels as a prognostic indicator of cardiovascular risk, nitrite levels decreased with increasing cardiovascular risk load. Studies [2,35] demonstrate that plasma nitrite can be determined with sufficient reproducibility in humans. Further, Kleinbongard et al. showed that plasma nitrite levels progressively decrease with increasing cardiovascular risk load, and that the presence of endothelial dysfunction and its degree are reflected by the relative difference in plasma nitrite compared to controls [35].

These studies suggest the possibility that plasma nitrite may indicate not only acute changes in NO activity in animals and in healthy humans, but also chronic changes in NO activity in individuals with early stages of atherosclerosis [35]. Because of great inter-individual variation in concentrations, nitrite seems to be less suitable for primary diagnostic purposes but may be a sensitive marker for the individual follow-up of disease states associated with changes in endothelial function. Thus, determination of plasma nitrite may be suitable to assess endothelial dysfunction in patients and to monitor the efficacy of therapeutic interventions in future clinical trials [35].

Ischemia/reperfusion injury

Ischemia-reperfusion (IR) injury is a series of cellular events that takes place following the resumption of oxygen delivery to a tissue after a period of hypoxia. This injury may be severe enough to cause significant morbidity and mortality [37]. In the 1980’s, it was discovered that a series of short-duration ischemic events in advance of a longer-duration ischemic insult could confer significant protection. Preconditioning by ischemic tolerance was first identified in the heart, and was subsequently found to occur in the brain and a variety of organs including the liver, intestine, kidney, and lung [38]. Preconditioning stimuli include ischemia, low doses of endotoxin, hypoxia, hyperthermia and hyperthermia, cortical spreading depression, anesthetics, and 3-nitropropionic acid, among others. Preconditioning strategies have always proven to be effective in experimental models but the clinical relevance of such strategies is questionable. While it would be clinically dangerous and impractical to precondition at-risk patients with ischemia [38], an understanding of the protective mechanisms could lead to therapeutic approaches to improve patient outcomes.

Cardiac preconditioning requires a minimum of 2 minutes of ischemia and reperfusion to induce an immediate protective effect of ischemic preconditioning (IPC). IPC occurs in 2 phases: an “early” phase of protection, which develops within minutes after initial ischemia and lasts 2 to 3 hours, and a “late” phase of protection, which begins 12 to 24 hours later and lasts 3 to 4 days. Late IPC has protective effects against myocardial infarction and myocardial stunning, but early IPC only has an effect against myocardial infarction [39]. Ischemic preconditioning in the brain is accomplished globally by occlusion of the bilateral common carotid arteries, or focally by occlusion of one side of the middle cerebral artery for about 1 to 20 minutes. Preconditioning-induced neuroprotection is observed not only in terms of infarct volume but also in terms of neurological scores and behavior studies in animals [38].

A number of mechanisms have been advanced to explain the cytoprotective effects of NO following IR injury. A thorough discussion of NO-mediated cytoprotection against IR injury is beyond the scope of this review, so we refer the reader to recent reviews of the subject [40-42]. NO has been extensively studied in the setting of myocardial IR injury. A review of the literature which investigated the role of NO in modulating the severity of IR injury in the non-preconditioned myocardium spanning the period from 1991 to 2001 found that 73% of the studies reported that NO (endogenous or exogenous) was cardioprotective, whereas 12% reported that NO was detrimental [40]. This very comprehensive analysis revealed that the discrepancies between these two opposing findings can be explained by the dose of NO investigated, as it was found that physiological levels (i.e., nanomolar) of NO promote cytoprotection and suprapharmacological levels (i.e., high micromolar and millimolar) mediate cellular necrosis and apoptosis [4].

NO may play a key role as a mediator of the neuronal ischemic preconditioning response, either in conjunction with or independent of NMDA receptor activation [38,43]. Administration of the NOS inhibitor (N-nitro-L-arginine methyl ester (L-NAME)) before and during the preconditioning protocol attenuated functional recovery [44]. Treatment with the INOS inhibitor aminoguanidine abolished the induced protection [38]. Exogenous NO was shown to elicit preconditioning-induced protection [44]. Scavenging of NO during preconditioning significantly attenuated the induced neuronal tolerance, and neither eNOS- nor nNOS-knockout mice showed protection following ischemic preconditioning. Pharmacological elevation in tissue cGMP levels by administration of NO donors S-nitroso-N-penicillamine (SNAP) or sodium nitroprusside (SNP) before sustained ischemia elicited cardioprotection similar to ischemic preconditioning [44]. Subsequent studies also emphasized the significance of cGMP and activation of PKG in the triggering of preconditioning [44]. Increases in the tissue cyclic nucleotides, cyclic adenosine monophosphate (cAMP) and cGMP were found to occur during a multi-cycle preconditioning protocol, suggesting roles for the beta-adrenergic signaling pathway and nitric oxide (NO) as triggers of cardioprotection [44]. The administration of inhaled NO gas therapy also significantly protects the myocardium [45]. Preconditioning by volatile anesthetics also appears to involve NO pathways [38,46].

NO possesses a number of physiological properties that makes it a potent cardioprotective signaling molecule. First, NO is a potent vasodilator in the ischemic myocardium [47] which allows for essential perfusion of injured tissue. Second, NO reversibly inhibits mitochondrial respiration [48]. The inhibition of mitochondrial respiration during early reperfusion leads to a decrease in mitochondrial-driven injury by extending the zone of adequate tissue cellular oxygenation away from vessels [49]. Third, NO is a potent inhibitor of neutrophil adherence to vascular endothelium [50]. Neutrophil adherence is an important event initiating further leukocyte activation and superoxide radical generation, which in turn leads to injury to the endothelium and perivascular myocardium. Fourth, NO prevents platelet aggregation, which together with the anti-neutrophil actions of NO attenuates...
Nitrite therapy for IR injury

Nitrite therapy has proven to be an effective preconditioning agent [21]. There are several limitations to the currently used NO-based therapies that preclude their widespread use, but which nitrite might address. Inhaled NO therapy and NO donor therapy have the potential to cause unwanted side effects, since both therapies rely on systemic delivery. For example, NONOates, a class of NO donors, release NO over a period of time depending on the half-life of the drug [4]. Since NO begins to be released as soon as the drug enters the blood stream, there is no way of ensuring that the NO is released only at the site of injury. As a result, higher concentrations of the drug have to be administered to achieve the desirable therapeutic effects. This in turn can result in unwanted systemic side effects, such as hypotension or NO-mediated cytotoxicity. Alternative means to effectively deliver NO to the site of injury are thus necessary to achieve the therapeutic potential of NO-based therapies [4].

Nitrite is a relatively stable molecule that can be transported in the circulation and stored in tissues. Nitrite releases NO under conditions that exist in injured tissue (ischemia, hypoxia, or low pH), which allows nitrite to preferentially target injured tissue and reduce the risk of systemic hypotension and other unwanted side effects. This is probably the most important aspect of nitrite therapy [4]. In addition, since nitrite has been used for many years as part of the cyanide antidote kit in humans there is a wealth of clinical data to support the safety of very high doses of sodium nitrite in critically ill patients [4].

During hypoxia and ischemia, nitrite not only promotes blood flow by conversion to nitric oxide, it also protects the mitochondria [24]. Nitrite acts to inhibit mitochondrial respiration by blocking the electron transport chain at both complex I and cytochrome c. Under ischemic conditions, myoglobin reduces nitrite to NO, which can bind reversibly to the catalytic center of cytochrome c oxidase. Nitrite can also inhibit complex I by S-nitrosation [21]. Partial inhibition of mitochondrial respiration can regulate tissue oxygen gradients and conserve oxygen, particularly in conditions of physiological hypoxia. Inhibition of the most actively respirating mitochondria and those closest to the oxygen source would allow oxygen to diffuse beyond these mitochondria and further into the tissue to those sections of the tissue that are more distant from the oxygen source. This extension of the oxygen gradient deeper into the tissue would also extend the NO gradient in the tissue, thereby increasing the apparent bioavailability of both oxygen and NO [49]. While reperfusion of the tissue is necessary to restore cellular energetics, reperfusion itself exacerbates mitochondrial damage and tissue injury. Inhibition of complex I by nitrite is thought to contribute to cytoprotection by blocking entry of electrons into the respiratory chain and thus attenuating the burst of reactive oxygen species generation associated with reperfusion.

Nitrite has recently been found to confer cytoprotection against IR injury in a number of animal models [23]. Although the mechanisms of this phenomenon are still being characterized, the reproducibility of this effect in multiple animal IR models suggests nitrite as a novel potential therapy for human ischemic diseases [4].

Current methods for measurement of NO and NO₂

Monitoring authentic NO is very challenging because this free radical has a half-life of only ~ 1 microsecond to 2 milliseconds in blood, and a diffusion distance of only about 0.1 μm depending on model assumptions [34]. The measurement of NO metabolic products nitrite (NO₂⁻) and nitrate (NO₃⁻), is less technically challenging than direct NO measurement. Table 1 provides a comparison of current methods for measuring NO and NO₂. Other proposed indices for NO include nitrosohemoglobin [20] and nitrosated and nitrosylated proteins, such as SNO-albumin [53].

Non-invasive measurements of NO levels can be obtained using electron paramagnetic resonance (EPR) imaging to make real-time measurements by administration of spin-trapping agents. This technique is useful in either in vitro studies or in animal research, but human trials using EPR have yet to be reported (excluding analysis of blood samples extracted from patients). Although EPR is a very powerful imaging tool in NO research, the heavy instrumentation, high cost, small sample size, and the potential toxicity of the spin-trapping agents are major roadblocks for future clinical applications [54]. Magnetic resonance imaging (MRI) and positron emission tomography (PET) are also under investigation for monitoring changes in NO and NOS enzymes, respectively. The biggest disadvantage of MRI is its low sensitivity, requiring millimolar levels of NO, which will mean that its applications will be very limited. In addition, there are major concerns about the potential toxicity of the imaging agents. Relatively long data acquisition times are typically needed to generate a detectable MR signal, while the lifetime of NO is short, so the accuracy of MRI measurements of NO is questionable [54]. PET imaging has only been evaluated to detect NOS enzyme expression, but has been unsuccessful to date. First, the expression of NOS is in the cytoplasm, and the radiolabeled NOS inhibitors may not permeate the cell membrane efficiently which dramatically reduces tracer uptake. Second, the specificity of these NOS inhibitors may not be high enough for imaging applications; many of them can undergo nonspecific adsorption to other proteins such as albumin. Third, the stability of those tracers in vivo is a major concern [54].

NO-specific electrodes are commercially available (e.g., World Precision Instruments, Innovative Instruments). Their potential clinical uses are limited because both oxygenated and deoxygenated hemoglobin compete with the sensor for NO. Recently, Takarada et al. [55] reported the first successful intravascular measurement of NO in humans using an NO electrode mounted onto a catheter. The sensor was positioned in the coronary sinus, and an acetycholine stimulus was administered nearby in the left main coronary artery. The sensor could not measure the absolute level of circulating NO because of the differences between the calibration sites and the measuring site. There was a weak but significant correlation between the NO levels measured by the catheter-type NO sensor and the nitrite level from samples taken at the same time and in the same location, measured by HPLC with the Griess reaction (r = 0.41, p = 0.048) [55]. The weakness of the correlation may be because the reported detection limit for nitrite was only 10 nM, about the same concentration as the MR signal, while the lifetime of NO is short, so the accuracy of MRI measurements of NO is questionable [54]. PET imaging has only been evaluated to detect NOS enzyme expression, but has been unsuccessful to date. First, the expression of NOS is in the cytoplasm, and the radiolabeled NOS inhibitors may not permeate the cell membrane efficiently which dramatically reduces tracer uptake. Second, the specificity of these NOS inhibitors may not be high enough for imaging applications; many of them can undergo nonspecific adsorption to other proteins such as albumin. Third, the stability of those tracers in vivo is a major concern [54].
Use of a catheter-mounted sensor [56] requires fluoroscopic guidance, which is available in catheterization labs, but not typically in operating rooms or intensive care units. Intravascular use of NO electrodes for a prolonged period may also be limited by thrombosis. Most examples of NO electrodes in human use are limited to non-vascular applications, such as extracorporeal blood during hemodialysis [57], insertion into synovial fluid [58] and subcutaneous insertion into the forehead [59].

Monitoring exhaled NO is useful for applications involving airway inflammation such as asthma, but not for most other applications where NO from a distant source would convert to nitrite before it reached the lungs [60]. Exhaled NO has a very short half-life in the lung vasculature and should be incapable of distal transport in blood. This is because NO reacts in a nearly diffusion-limited reaction with both oxyhemoglobin (6 to 8x10^-7 mol/L/sec) and deoxyhemoglobin (2 to 6x10^-7 mol/L/sec) to form methemoglobin/nitrate and iron nitrosyl hemoglobin, respectively, which yield a half-life of ~1 µs and a diffusion distance of ~0.1 µm [34]. Monitoring nitrite using exhaled breath condensate (EBC) is another alternative [61], but analysis is complicated due to the substantial contribution of nitrite from the oropharyngeal tract during standard collection of EBC [62].

As mentioned above, nitrite (NO₂⁻) is a widely accepted surrogate marker for nitric oxide [3,4]. The circulating half-life of nitrite has been estimated to be from 11 to 42 minutes [34,63,64], but 5-8 hours for nitrate [65]. This decay of nitrite is dominated by the reaction with oxyhemoglobin to methemoglobin and nitrate. Thus, nitrite is an appropriate marker [66] because it is present long enough to be measured, yet changes rapidly enough to track variations in NO levels. Plasma nitrite levels closely track changes in whole blood nitrite and cardiac nitrite in a rat model of cardiac arrest [67].

A great deal of interest in the nitrite anion itself has recently emerged [32]. Given the tremendous research interest in this field, as well as the numerous potential clinical applications described below, it would be of considerable value to be able to monitor nitrite in blood in real time, on a continuous basis. This is because nitrite has a relatively short half-life, and sample preparation requires considerable care [63]. An issue with measuring NO is the widespread contamination of labware with NO levels that can confound experimental results [53,63]. One of the most commonly reported methods for measuring NO₂⁻ is ozone-chemiluminescence [63]. However, along with other current methods including HPLC [68], gas chromatography-mass spectroscopy (GC-MS) [28] and capillary electrophoresis [69], ozone-chemiluminescence requires multiple sample preparation steps that require specimens to be centrifuged, chemically stabilized, frozen and then transported to these instruments for later analysis, with results provided hours or days later, which does not provide a suitable response time for many clinical applications. The short half-life of nitrite in whole blood, demanding rapid separation of plasma from erythrocytes, is a considerable obstacle for plasma nitrite determination in clinical practice [2]. Methods such as ion-selective nitrite electrodes [70] are insensitively sensitive to monitor NO₂⁻ in the physiological regime (100 nM to 1 µM). While some investigators report a low detection limit for the Griess reaction combined with HPLC (~10 nM) [55] a summary of research performed with the Griess reaction showed many reports with a detection limit of ~1 µM [63]. GC-MS methods can also be insensitively sensitive for measurement of nitrite in plasma and whole blood [28].

We have previously described a technology that can enable measurement of physiological levels of nitrite in blood plasma in humans in real time with no sample preparation and has the potential to enable clinical decision-making [71]. Figure 1 illustrates an early prototype of the plasma nitrite monitor. In this method, blood is processed through a small hemodialysis-type filter with a surface area of 20 cm² and a molecular weight cutoff of 10 kD to exclude hemoglobin. A transmembrane pressure gradient is applied between the inlet and outlet of the filter, resulting in separation of the blood elements by molecular weight. The filtrate containing the free nitrite is then pumped from the filter and mixed with a solution of dilute sulfuric acid and potassium iodide. This process stoichiometrically reduces the nitrite to nitric oxide [141]. The filtrate is then monitored for NO levels using an NO-specific electrode. A limitation of this technology is the requirement to draw blood samples. Currently about 30 ml of blood are required for about 6 hours of monitoring, but this volume may be further reduced as the technology is improved. Because the blood is processed using clinically proven filter membranes, the potential exists to salvage the retentate, primarily erythrocytes, leukocytes, and larger proteins, for re-infusion into the patient. Figure 2 shows a calibration trace of nitrite in anticoagulated human blood from 100 nM to 5 µM. The system shows a linear response (r² = 0.99) over a nitrite concentration range from 10 nM to 5 µM in diluted blood (1 part blood to 4 parts normal saline) which surpasses the normal physiological

| Method | NO/NO₂⁻ Detection Limit* | Continuous or Discrete? | Direct Measure in Blood? | Lab Sample Prep Required? | Potential as rapid clinical diagnostic? |
|--------|--------------------------|------------------------|--------------------------|--------------------------|---------------------------------------|
| EPR    | ~10 nM [139]             | Continuous             | Yes                      | No                       | No                                    |
| MRI    | ~1 mM [54]               | Continuous             | Yes                      | No                       | No                                    |
| Griess Reaction | 10-1500 nM [63]        | Continuous             | Discrete                 | Yes                      | No                                    |
| Ozone-Chemiluminescence | 1 nM [63]         | Discrete               | No                       | Yes                      | No                                    |
| Ion-selective nitrite electrode | 0.5-10 µM [70] | Continuous             | Yes                      | No                       | Yes                                   |
| Nitric oxide electrode | 0.08-1 nM [55]        | Continuous             | No                       | Yes                      | Yes                                   |
| HPLC   | 10 µM [68]               | Discrete               | No                       | Yes                      | No                                    |
| Capillary Electrophoresis | 60 nM [69]            | Discrete               | No                       | Yes                      | No                                    |
| GC-MS  | 0.5 µM [28]              | Discrete               | Yes                      | No                       | No                                    |
| Exhaled NO | 0.5 ppb (~15 nM) [140]   | Continuous             | No                       | No                       | No                                    |
| Exhaled breath condensate | ~ 1 µM [61]        | Continuous             | No                       | No                       | No                                    |
| Silver Medical instrument | < 10 nM [71]       | Continuous             | Yes                      | No                       | Yes                                   |

Table 1: Comparison of current NO and NO₂⁻ detection methods. *Detection limits are approximate since these can vary in practice.
range (typically 100 nM to 1 uM) [19]. The initial response time to a 100 nM bolus of NO₂ is 1-2 minutes. Analysis can be performed either continuously or with discrete samples. Any background signal from potential contaminants can simply be measured using nitrite-free water or saline, and then subtracted from the sample. Blood samples can be placed directly into the instrument, or drawn from the subject through an existing catheter.

Specificity of NO or NO₂ as a biomarker

Nitric oxide and NO₂ are involved in numerous biological pathways, therefore the specificity of these markers for vascular pathologies will be dependent upon concentrations, kinetics, and localization [27,72]. Interpretation of NO or nitrite readings will require a differential diagnosis. For example, the rate of increase in NO levels generated by eNOS and/or nNOS (during ischemia) is from 10 to 50 times faster [73,74] than the rate of NO₂ generated by iNOS (during inflammation) which provides one method to distinguish between ischemia and inflammation or infection. Continuous or frequent discrete measurement of NO or an index marker should thus allow distinction between ischemic events superimposed on inflammatory processes. Location-specific measurements will also help to improve marker specificity. There is an arterio-venous gradient in plasma NO₂ levels, indicating that NO₂ is not at a constant level in the general circulation. For example, measurement using a central venous catheter in the superior vena cava (SVC) will be primarily indicative of nitrite in the upper body, including the brain, head and arms, and may be differentiated from more global changes by comparison with measurements in the inferior vena cava or from an arterial line. Alternatively, measuring NO₂ in the coronary sinus could be indicative of cardiac changes. Making measurements from multiple locations simultaneously would indicate if a particular organ was experiencing ischemia.

Several examples of the potential utility of monitoring plasma nitrite or nitric oxide in real time are discussed below, although there are numerous examples which are beyond the scope of this review, such as traumatic brain injury [75,76], cancer [77], or renal [78] or hepatic [37] ischemia-reperfusion injury, among many others.

In-hospital stroke

Stroke is the third leading cause of death in the US, behind heart disease and cancer, and is a leading cause of serious, long-term disability [79,80]. Out of 795,000 strokes each year, roughly 200,000 are fatal, and the five-year survival rate of non-fatal strokes is only 50% [81]. The estimated direct U.S cost of stroke for 2009 is $54.2 billion with the long-term costs for an individual estimated to be $140,000 [82]. Ischemic stroke accounts for approximately 87% of all strokes, while the remainder are hemorrhagic [81]. In over 60% of stroke patients, vascular thrombus deposition is responsible for the interruption of cerebral blood flow (CBF). Reduced CBF and severe oxygen deficiency leads to ischemia, and eventually to behavioral and functional deficits, morbidity and mortality [83,84]. Key factors in predicting stroke morbidity and mortality are severity and time to treatment, with earlier treatment equating with improved outcome. A landmark study documented that treatment within 60 minutes reduces disability by 30% [85]. Since stroke symptoms can be ambiguous and many patients are alone, sedated, in surgery, or asleep during stroke onset, delays in recognition and assessment are common [86], thus significantly limiting the ability to provide timely therapy.

Even in hospitals, where 6.5 to 15% of strokes occur, delayed detection, assessment, and treatment are common [87]. Common cardiac, vascular and intracranial surgical procedures have significant intra-operative stroke rates, yet patients are not adequately monitored. Nearly 750,000 such procedures are performed every year in the US, including 500,000 coronary artery bypass grafts (CABG), 130,000 carotid endarterectomies (CEA), and 100,000 heart valve replacements. The risk of intra-procedural stroke due to CABG is 6% [88], 6% for CEA [88,89], and up to 10% for heart valve replacements [90]. Cardiovascular and cerebrovascular surgery have a negative impact on brain function due to stoppage of blood flow during surgery. In fact, more than 25% of patients who receive coronary artery bypass surgery suffer from temporary or permanent memory loss. As a result, it is of premier importance to develop strategies to protect the brain either prior to vascular surgeries or in patients at high risk of stroke [38]. A rapid monitoring technique to aid in the detection of ischemic stroke in humans may prove useful in determining a treatment regimen to promote functional recovery.

Although NO is an important regulator of the cerebral circulation, the relative importance of the rapid generation of NO from nitrite discussed above, as compared to eNOS and nNOS, is unknown. Both eNOS and nNOS are activated somewhat more slowly, in about 10 minutes following ischemia [91]. In a middle cerebral artery (MCA) occlusion model of focal ischemia, nNOS knockout mice develop significantly smaller infarct sizes and have better neurological outcomes.
than wild-type mice [5]. nNOS knockout mice are also resistant to focal and global cerebral ischemia, consistent with a role for nNOS-derived NO in cellular injury following ischemia [5]. In contrast, eNOS knockout mice subjected to the MCA occlusion model develop larger infarct sizes compared to wild-type mice [5,92]. Several hours after stroke onset, iNOS is activated as part of the inflammatory processes. The concentrations of NO generated from iNOS are significantly greater than those produced by eNOS and nNOS. In rats, NO rises in jugular venous blood immediately following a stroke induced by middle cerebral artery occlusion and then returns to normal when ischemia is relieved [73]. Clinical research shows that NO is also significantly elevated in cerebrospinal fluid following ischemic stroke [93].

We recently demonstrated the ability to monitor changes in plasma nitrite in real time as an indicator of cerebral ischemia, in a rabbit embolic stroke model [71]. There was a significant increase in plasma nitrite that began immediately following large clot embolization, which reached statistical significance within 3 minutes [71]. This may have occurred due to conversion of intraerythrocytic nitrite to NO, followed by release from red blood cells, followed by rapid oxidation of plasma NO back to plasma nitrite. This would produce at least a transient increase in free nitrite as compared to intracellular nitrite, as measured in this experiment. However, Jeffers et al. [94] provide a theoretical argument that it is not NO which is released from red blood cells during ischemia, but another species, possibly N₂O₃, which as measured in this experiment. However, Jeffers et al. [94] provide a theoretical argument that it is not NO which is released from red blood cells during ischemia, but another species, possibly N₂O₃, which then decomposes to release NO outside the cell. Factors favoring the conversion of intracellular (rather than free) nitrite to NO are the high concentration of intracellular hemoglobin [2] (about 20 mM), the large fraction of nitrite which is stored within erythrocytes, and the short diffusion distance between intracellular nitrite and deoxyhemoglobin as compared to free nitrite.

Intra-operative monitoring for cerebral ischemia is becoming the standard of care [95]. Even using less sensitive monitoring methods for cerebral ischemia, such as near-infrared spectroscopy (NIRS), EEG, and transcranial Doppler (TCD), clinical researchers have shown dramatic improvements in patient outcomes as compared to unmonitored patients [96-98]. The US Food and Drug Administration recently allowed a label claim that monitoring improves patient outcomes [99]. The therapeutic response for early intra-operative ischemia will primarily involve providing adequate oxygenation to surrounding tissues by elevating the mean arterial pressure, increasing cardiac output or pump flow, altering arterial CO₂ tension, or decreasing cerebral oxygen demand using pharmaceutical agents such as propofol, actions that could be managed by the anesthesiologist.

The therapeutic potential of nitrite for stroke was evaluated in a rat model of middle cerebral artery occlusion. In this study, solutions of nitrite were infused intravenously at the time of reperfusion, and were found to reduce infarct volume and enhance local cerebral blood flow and functional recovery. Nitrite treatment was also shown to provide neuroprotection when its administration was delayed to as much as 3 h following ischemia [4].

Subarachnoid hemorrhage

Subarachnoid hemorrhage (SAH) is a rare, but particularly devastating, type of stroke produced by a ruptured intracranial aneurysm that affects about 30,000 people annually. While this represents only about 3% of all strokes and 5% of deaths from stroke, the relative youth of the affected individuals means that SAH is responsible for a quarter of all life-years lost as a result of stroke [100]. It leads to severe spasm of the cerebral arteries, which develops 4–9 days after SAH in 30–70% of patients. Of those individuals with delayed cerebral vasospasm, 20–30% experience delayed ischemic neurological deficits (DIND) and approximately half suffer severe permanent neurological dysfunction or death. The influence of NO on blood flow, the presence of nNOS and eNOS in the cerebral vessels, and the affinity of NO for the heme moiety all suggest that the depletion of NO is responsible for vasospasm [101]. Current research suggests that bilirubin oxidized fragments (BOXes) constrict cerebral arteries by elevating levels of asymmetric dimethyl arginine (ADMA), a competitive inhibitor for NOS enzymes, and thus inhibit basal NO production [102]. This depletion of basal NO levels occurs around the time of vasospasm. Continuous infusions of nitrite have been shown to prevent vasospasm in a primate model of SAH [101]. The ability to monitor nitrite levels either in blood or cerebrospinal fluid on a frequent or continuous basis could provide an extremely rapid indication of the occurrence of vasospasm, and allow much more rapid treatment than the current method – Transcranial Doppler – which is used at most once or twice daily.

Pediatric cerebral ischemia

More than a half million infants in the US were born preterm in 2004. The preterm birth rate has climbed 18 percent since 1990, to one of every eight live-born infants [103]. Of these children, more than 10% will sustain neurological injuries leading to significant learning disabilities, cerebral palsy, or mental retardation, with very low birth weight infants having an even higher incidence of brain injury [104]. The estimated lifetime costs in 2003 dollars are expected to total $51.2 billion for persons born in 2000 with mental retardation, and $11.5 billion for persons with cerebral palsy [105]. No effective clinical strategies have yet been developed to counteract this condition [106]. In addition to preterm infants, the most common birth defect is congenital heart disease (CHD) requiring approximately 36,000 surgeries per year [107]. Improvements in cardiopulmonary bypass (CPB), surgical technique, and intensive care have lowered hospital mortality for most neonatal procedures to <3%. Improvements in survival have stimulated a focus on neurological morbidity, which may approach 70% [108].

The two most important forms of brain injury in premature infants, which are believed to be primarily responsible for conditions such as mental retardation, cerebral palsy and epilepsy, are germinal matrix–intraventricular hemorrhage (GMH-IVH) (also known as periventricular–intraventricular hemorrhage (PVIVH)) and periventricular leukomalacia (PVL) [109]. Postnatally, most hemorrhages occur when the neonate is younger than 72 hours, with 50% of hemorrhages occurring on the first day of life. A large body of experimental and clinical observations indicates that disturbances in cerebral blood flow (CBF) are important in the pathogenesis of PVIVH and PVL. First, because of their very high incidence of respiratory disease and the need for mechanical ventilation, the complications of such ventilation, alterations in mean arterial blood pressure (MAP) are very common in such infants. Second, considerable experimental data and some clinical results indicate that cerebrovascular autoregulation, the mechanism by which CBF is maintained constant despite alterations in MAP, is either defective or absent in at least some infants” [110]. As with adults, nitric oxide is involved in ischemia in this population as well. Hyponxia-induced release of nitric oxide can account for much of the cerebral vasodilatation observed in response to hypoxia in the fetus [111].

Currently there are no methods for continuously monitoring hypoxic-ischemic insults in the neonatal brain. PVL and PVIVH are
Cardiac arrest

There are approximately 295,000 cases of out-of-hospital cardiac arrests each year in the US, and an individual’s overall chance of surviving until hospital discharge is 7%. The rates of survival to discharge after in-hospital cardiac arrest were 33% among children and 21% among adults [117]. Dezfulian et al. [67] showed that global ischemia due to cardiac arrest caused a rapid drop in plasma nitrite levels in mice. They also showed that administration of nitrite solution (as opposed to saline control) at the onset of cardiopulmonary resuscitation (CPR) following 12 minutes of asystole was cardioprotective. Nitrite therapy significantly improved survival compared with placebo (76% vs. 48%, p=0.033). Nitrite therapy also significantly improved post-arrest left ventricular ejection fraction (LVEF) compared with placebo treated mice (54.4±2.4% vs. 43.5±2.9%; p=0.007). The investigators found that therapeutic nitrite repletion and S-nitrosation in the heart were associated with transient, reversible inhibition of complex I, reduced mitochondrial reperfusion, ROS generation, and oxidative injury. Nitrite improved pulmonary gas exchange, cardiac contractility, and survival.

Sepsis

In the United States there are ~750,000 cases of sepsis or septic shock each year, and that number was expected to have reached 934,000 by 2010 [118]. With a mortality rate of 25–56% [92], approximately 215,000 patients die each year. In the US, the estimated cost of ICU care for patients with sepsis exceeds $20 billion annually [119].

NO is generally believed to be harmful in sepsis, due to its effects on the macrocirculation (causing arterial hypotension). A phase III multicenter, randomized, double-blind, placebo-controlled trial of the nonspecific NOS inhibitor NG-methyl-L-arginine (L-NMMA) was stopped early because of increased mortality in the NO inhibition group, which seemed to be caused by cardiac failure [120] even though hypotension was improved [121]. Precisely why this clinical trial failed is not entirely certain, but concerns have been expressed about the dosage, patient selection, titration to maintain blood pressure, as well as the nonspecific nature of the NO inhibitor used. Despite the lack of benefit of NO inhibition in this large clinical trial, studies of NO in sepsis continue to yield important pathophysiologic insights.

Sepsis is increasingly recognized as a ‘disease of the microcirculation’, in which enhanced vasoconstriction and mitochondrial dysfunction cause irreversible damage and organ failure. There are multiple causes of organ failure in sepsis. First, redistribution of blood flow as well as microvascular failure, constriction, obstruction, and permeability changes cause tissue hypoxia and ischemia, which is a failure of oxygen delivery. Second, mitochondrial damage and dysfunction cause metabolic hypoxia which is a failure of oxygen use. Third, reactive oxygen and nitrogen species exert direct cytotoxic effects as well, damaging membranes, lipids, nucleic acids, and proteins [120].

Although inhibition of NOS activity is clearly effective at raising arterial pressure in sepsis, it can simultaneously worsen the impairment of microcirculatory perfusion and oxygen transport to tissues. In the healthy state and under pathologic conditions, NO maintains microcirculatory homeostasis by regulating microvascular tone, leukocyte adhesion, platelet aggregation, microthrombi formation, and microvascular permeability. Blocking NO production in sepsis worsens leukocyte adhesion, platelet aggregation and microthrombosis, and microvascular permeability, causing decreased myocardial blood flow.
flow and defects in tissue oxygenation that do not recover with fluid resuscitation alone. It has been suggested that vasodilators could be used therapeutically to improve tissue oxygenation in sepsis by opening the microcirculation (arterioles, capillaries, and venules < 100 mm diameter) [120].

Vasodilation, improvement of peripheral flow, and oxygenation might not be the only solution, as the problem might lie in cellular oxygen utilization rather than in oxygen delivery. Impaired oxygen utilization, also called 'cytopathic' or 'metabolic' hypoxia, is mainly due to reduced activities of mitochondrial respiratory enzyme complexes. Interestingly, mitochondrial dysfunction and reduced concentration or activity of complex I seem to be directly associated with organ failure and mortality in septic patients [122,123]. In a study in patients with septic shock, skeletal muscle biopsies demonstrated similar ATP depletion and respiratory chain complex I inhibition [113]. Complex I activity had a significant inverse correlation with norepinephrine requirements (a proxy for shock severity, p=0.0003) and nitrite/nitrate concentrations (p=0.0004), and a significant positive correlation with concentrations of reduced glutathione (p=0.006) and ATP (p=0.03) [122]. In this clinical setting, the degree of mitochondrial dysfunction correlated with the severity of illness and with eventual outcome [122,123]. NO contributes to mitochondrial dysfunction in sepsis, in part by nitrating regulatory proteins. Very high levels of NO can cause opening of the mitochondrial permeability transition pore, with uncoupling of oxidative phosphorylation, dissipation of membrane gradients, membrane swelling, and cell death. This implies that treatments that protect mitochondrial function or stimulate mitochondrial biogenesis and recovery might be useful for preventing organ failure and morbidity in sepsis [120].

The upregulation of inducible nitric oxide synthase (iNOS) in sepsis is heterogeneously expressed between and within organ systems and NO can be consumed by reactive oxygen species, giving the potential for localized areas of relative NO deficiency in microvascular beds despite a state of excess total body NO. Oxidative stress contributes as well, in which NO-induced mitochondrial injury causes inefficient mitochondrial respiration, with increased production of reactive oxygen and nitrogen species.

**Sepsis diagnosis**

While there are clear guidelines as to how physicians should treat septic patients once they are diagnosed [118], making the diagnosis of sepsis is very difficult even among intensivists [124]. The early stage of this disease is known as systemic inflammatory response syndrome (SIRS) and represents an assortment of symptoms, many of which are non-specific, so a diagnosis of SIRS is difficult. As the patient makes the transition from SIRS to sepsis, they are often physically normal, so it is difficult to know when they are making that transition. This is the key stage at which to catch the patients. Currently, sepsis diagnosis is made on the basis of meeting two of four SIRS criteria (including 2 or more of: 1) Temperature > 38°C or < 36°C; 2) Heart rate > 90 beats/min unless the patient is taking medications to reduce the rate (a beta-blocker or calcium channel blocker) or the heart is paced; 3) Respiratory rate > 20 breaths/min (or PaCO₂ < 32 torr) or mechanically ventilated; 4) Leukocyte count > 12,000/µL or < 4,000/µL, or > 10% immature band forms) as well as low blood pressure, and elevated lactate levels.

Early detection of sepsis can improve patient outcomes by either guiding antibiotic therapy or fluid resuscitative therapy. The association of timely and appropriate antibiotic therapy with improved morbidity and mortality has been established in the ICU setting [125]. Early goal-directed therapy (EGDT) in sepsis and septic shock is now standard of care [126], and is cost-beneficial, as these patients have a shorter length of hospital stay than those receiving standard care [118].

Carrigan et al. [127] provide an excellent review of diagnostic tools for sepsis. “We propose that current diagnostic methods could be complemented by including real time monitoring of proteic biomarkers during the course of treatment. Existing and novel methods alike require a ‘wait and see’ approach in their diagnostic methods because of their incubation cycles. Although traditional ELISAs (2–3h), immunoluminometric PCT assays (3h), and DNA detection by PCR (5–6h) provide more rapid results than culture testing (24–48h), these methods are incapable of monitoring the exponential changes in biomarkers occurring in both sepsis and experimental *in vivo* endotoxemia.” Further, according to Gao et al. [128], “Current standard microbiological techniques identify infecting organisms after culture of a clinical isolate in conditions suitable for replication of the infectious agent. This may be difficult with fastidious organisms or if patients have received antibiotics. Preliminary classification is usually possible within 24 hours, with full species identification and antimicrobial sensitivity data becoming available 48 to 72 hours after blood sampling. The slowness of the investigation usually mandates the use of ‘best guess’, and often broad spectrum, antibiotics while awaiting results.”

It is widely recognized that an increase in systemic nitric oxide (NO) is a marker for infection. In experimental models of sepsis, NO produced by either eNOS or iNOS begins to increase at 2 hrs following lipopolysaccharide injection, and is maximal at 4-6 hours after initial infection [129,130]. iNOS generates much larger quantities of NO than either nNOS or eNOS. NO₂ levels are significantly elevated [131] in patients in the order: septic shock >> sepsis >>non-infectious SIRS or normal, healthy controls. NO levels in sepsis and septic shock are also much greater than in other infectious diseases, such as bacterial pneumonia [132]. In one study [133] NO₂ >54 µM showed 87% sensitivity and 77% specificity for sepsis. The sensitivity and specificity of NO₂ as a biomarker for sepsis might be dramatically improved by inclusion of kinetic data along with concentration data as part of the diagnostic criteria. In preliminary (unpublished) analysis of the data reported by Spack [133], patients could be identified with sepsis hours or days earlier by inclusion of a kinetics-based criteria such as an increase in NO₂ > 5 µM over the preceding measurement, rather than requiring an absolute level to be reached. This could be done by monitoring NO₂ on a much more frequent or potentially continuous basis, which would also dramatically improve the time to diagnosis. In addition, monitoring nitrite continuously might improve the timing at which other biomarkers, such as procalcitonin (PCT) and C-reactive protein (CRP) [127], might also be sampled, thus further improving the sensitivity and specificity of those markers, alone or in combination with NO₂. Longer term, earlier identification of sepsis study subjects using NO₂, in order to reduce heterogeneity of underlying conditions and disease course would be extremely useful in the identification and validation of new pharmaceutical targets.

**Sepsis therapies**

While the current standard of care, EGDT, is not implemented until a drop in blood pressure occurs, development of new therapies are likely to be significantly advanced by earlier detection of changes in NO levels that may ultimately lead to the drop in blood pressure which will allow a diagnosis of sepsis. While recombinant activated protein C (APC, also known as Drotrecogin alpha-activated) is currently the only FDA-approved therapy for sepsis, it is only indicated in patients...
at the late stage of severe sepsis [134]. Ultimately, the most successful therapies will in all probability be those that can be implemented as early as possible.

As discussed above, nitrite has been shown to be an NO donor specifically in conditions of low oxygen tension and low pH, which can be expected in the septic microvasculature. This is important in areas where blood flow should be increased but where NOS-derived and thus oxygen-dependent NO production is compromised [120]. Treatment with nitrite has been shown to protect mice against progressive hypothermia, mitochondrial dysfunction, organ damage, and even death induced by a lethal challenge with tumor necrosis factor (TNF) or lipopolysaccharide [135]. Nitrite treatment reduces mitochondrial dysfunction, organ damage, and mortality in shock [120], and thus may be a candidate for the treatment of shock [120]. Nitrite administration could prevent organ damage and multiple organ failure (MOF) by restoring or improving peripheral microvascular perfusion and/or mitochondrial respiration [120].

Summary and future directions

The potential utility for plasma nitrite as a diagnostic marker is extremely broad, with multiple potential clinical applications, including intra-operative stroke, subarachnoid hemorrhage, cerebral ischemia in premature infants, sepsis, and many others not discussed, such as renal or hepatic ischemia, traumatic brain injury and organ transplantation. Since nitrite levels have been shown to be relatively stable within an individual, the ability to monitor changes may provide an excellent method for diagnosing various diseases. It may be a prognostic indicator of cardiovascular health, similar to c-reactive protein (CRP). Frequent or continuous measurements of nitrite levels may be useful for intra-operative stroke monitoring.

One of the key clinical challenges in the development of nitrite as a diagnostic is that because nitrite and NO are involved in so many biological pathways, the specificity of the marker will be imperfect. The keys to improved specificity will be concentration, kinetics, and location. Gradients between two different sites can help to determine the origins of any changes that are detected. However, in some clinical settings such as cardiovascular surgery, the organs most at risk will be known, and sensitivity will be a key driver. As another example, in pediatric patients, if ischemia is detected and it is not cerebral ischemia, its source would still be identified and treated. Thus, in this setting, sensitivity is far more important than specificity. In settings where specificity is of greater importance, there may be complementary tests that can be used to improve the specificity of the results. For example, testing IL-6 levels will help to confirm whether elevated nitrite levels are due to inflammation. The ability to monitor nitrite levels on a continuous basis will also help to determine the timing of additional tests. Nitrite monitoring is similar to pulse oximetry in that the marker detected is global. Therefore, differential diagnoses will likely be required in the clinical setting to determine the probable cause of changes in nitrite. Chronic inflammatory conditions are likely to show high levels of NO, but are unlikely to show rapid changes, as occurs with ischemia. Sepsis will likely show levels of NO that are much higher than in ischemia.

The therapeutic potential of nitrite as a delivery vehicle for NO under ischemic conditions is considerable. A key requirement for the therapeutic use of nitrite in areas such as stroke, acute myocardial infarction, cardiac arrest, and others discussed above will be the development and validation of appropriate pharmacokinetic and pharmacodynamic models. This will require the ability to make frequent (preferably continuous) measurements of changes in nitrite and nitrate levels in the body. An initial clinical safety study for the continuous infusion of nitrite in healthy volunteers was recently published [28]. Fortunately, the long history of nitrite therapy for treatment of cyanide poisoning provides a substantial clinical safety profile [4]. A pharmacokinetic model [64] of nitrite in healthy volunteers was recently developed. Models of nitrite behavior in disease states such as cerebral ischemia or sepsis may be of significant pharmacologic benefit, so the ability to measure NO and/or nitrite in real time will help to establish the therapeutic value of nitrite.

The use of nitrite as a therapy for IR injury in humans may have some important limitations [23]. Hypotension due to nitrite-mediated vasodilation [18] may worsen outcomes in situations where hyperperfusion is already present. The production of NO in the setting of reperfusion may worsen oxidative and nitrosative stress both directly, in a dose-dependent manner [40], or by formation of highly reactive intermediates such as peroxynitrite [136]. It is unclear whether a therapy like nitrite will remain effective in humans with other co-morbidities, such as diabetes mellitus and renal insufficiency, in which biological processes such as eNOS uncoupling could adversely chemically “interact” with nitrite and produce unexpected consequences. Such unexpected effects of the NOS–NO axis has been observed in conditions associated with NO donor therapy and NOS uncoupling [137,138].

Finally, as clinical investigations are initiated for nitrite therapy for indications such as cardiac arrest, sepsis, and acute myocardial infarction, among others, the ability to monitor nitrite in real time has the potential to significantly improve the efficacy of the trials, as titration of nitrite could be performed, unlike with conventional measurement techniques. In addition, real time measurements can help to identify any unexpected problems with nitrite delivery. It remains to be seen whether nitrite monitoring will become a clinical requirement along with nitrite therapies as the results of these investigations become known.

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