Diagnosis, clinicopathological characteristics and prognosis of pulmonary mucinous adenocarcinoma

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Abstract. Primary pulmonary mucinous adenocarcinoma (PPMA) is an uncommon subtype of lung adenocarcinoma. The present study attempted to clarify the diagnosis, clinicopathological characteristics, and pathologic significance of epithelial growth factor receptor (EGFR) and Kirsten rat sarcoma viral oncogene (KRAS) mutations and the prognosis of PPMA. A total of 29 patients with PPMA from among 1,469 surgically resected patients with lung adenocarcinoma were enrolled. All of the tumours expressed CK7 and 5 cases exhibited co-expression with CK20. A total of 8 cases expressed EGFR, 14 cases expressed P53 and 2 cases expressed CEA. The majority of mucinous adenocarcinomas expressed thyroid transcription factor 1, Napsin A, Villin and Cam5.2 proteins. KRAS mutations were observed in 62% of patients and were more prevalent in the lower lung lobe and in patients with invasive mucinous adenocarcinoma. A total of 2 cases exhibited an EGFR mutation, and the co-mutation of KRAS and EGFR was only detected in 1 case. The relapse-free and overall survival rates at 5 years were 70.4, and 81.5%, respectively. The results may assist to identify a molecular target and supply important information for a therapeutic strategy for patients with PPMA.

Introduction

Lung carcinoma is the most common type of cancer and is the leading cause of cancer-associated mortality in humans worldwide; non-small-cell lung cancer (NSCLC) accounts for 80% of all lung cancer cases (1). Adenocarcinoma is the most common pathological subtype of NSCLC and has demonstrated an increasing prevalence (2). Primary pulmonary mucinous adenocarcinoma (PPMA) includes a group of subtypes of mucin-producing adenocarcinomas in the lung, and was first identified in 1978 as a mucous cyst tumour of the lung (3); other terms including cystic mucinous adenocarcinoma, colloid adenocarcinoma and mucinous bronchioloalveolar carcinoma were subsequently described (4,5). A novel classification for lung adenocarcinoma was proposed by an international multidisciplinary expert panel from World Health Organization (WHO) in 2015 (6) and invasive mucinous adenocarcinoma (IMA) was listed as an independent variant subtype. The new classification system of the concept of mucinous adenocarcinoma has been devised and is generally accepted. Referring to all of the subtypes in the lung with the mucin-producing phenotype, PPMA includes IMA, mucinous minimally invasive adenocarcinoma (mMIA) and mucinous adenocarcinoma in situ (mAIS). Compared with other lung adenocarcinoma subtypes, IMA exhibits different immunohistochemical and molecular features, and a different prognosis (7,8).

A total of 5-10% of lung adenocarcinoma cases exhibit features of mucinous differentiation (9), which provide specific challenges for pathological diagnosis. PPMA may be indistinguishable from metastatic neoplasms of gastrointestinal origin in terms of histology, and lung mucinous adenocarcinomas are different from non-mucinous adenocarcinomas in terms of mucin production. PPMA may require the use of immunohistochemical profiles, including the expression of cytokeratin 20 (CK20), caudal type homeobox 2 (CDX2), and hepatocyte nuclear factor 4α and the lack of expression of thyroid transcription factor 1 (TTF-1) and Napsin A (9).

The accumulation of genetic abnormalities is considered to induce the development of malignant neoplasms. Kirsten rat sarcoma viral oncogene (KRAS) and epidermal growth factor receptor (EGFR) mutations are frequently detected in patients with lung cancer (10,11). The KRAS mutation is the most common driver mutation in human cancer, although the prevalence of mutations and the affected codons differ according to the type of cancer. The majority of the available studies that included all types of adenocarcinoma and employed low sensitive and single-gene technologies have established that KRAS mutations occur in 61% of IMAs, and are associated with poor prognosis and a lack of response to chemotherapy (12). In addition, the EGFR mutation has been an area of particular interest on account of the identification that the application of EGFR-tyrosine kinase inhibitors in clinical tests resulted in a rapid response among patients with pulmonary adenocarcinoma who carried a mutation in EGFR (1). Furthermore, mutations in EGFR and KRAS are usually mutually exclusive (13).

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The purpose of the present study was to clarify the clinicopathological characteristics, KRAS and EGFR mutation status, and prognosis of patients with PPMA. The results may assist in identifying a molecular target and supplying important data for therapeutic strategy for patients with PPMA.

Materials and methods

Patient selection. Between January 2001 and May 2011, a total of 1,469 patients with lung adenocarcinoma were surgically treated in the Department of Cardiothoracic Surgery, Taizhou People's Hospital (Taizhou, China). From these, 29 patients (2%) with PPMA were selected. Inclusion criteria were as follows: (i) the presence of a mucinous pattern; (ii) the lack of a prior non-lung related cancer diagnosis; (iii) the presence of accurate clinical follow-up data, as well as the availability of residual tissue for genetic analyses. The study was reviewed and approved by the institutional review board of Taizhou People's Hospital (authorization number: KY201600301). Due to the retrospective nature of the present study, the requirement for written informed consent was waived.

Clinical information. All available clinical information was collected for analyses from the medical records and reports that were completed by the referring physicians. Sex, age at diagnosis, smoking status, tumour size, stage and location were included.

Histological classification. The surgically resected specimens from each patient were fixed for 24 h in 10% neutral buffered formalin solution in phosphate buffered saline (pH 7.4) at room temperature. Following fixation, the samples were washed with distilled water, dehydrated in graded alcohol and embedded in paraffin and sectioned at a thickness of 4 mm. Histological slides were reviewed and sections were stained with hematoxylin and eosin (H&E; Beijing Leagene Biotech Co., Ltd., Beijing, China) according to the manufacturer’s protocol. The tumor tissues were dissected from each patient were fixed for 24 h in 10% neutral buffered formalin solution at room temperature, washed in distilled water, dehydrated in graded alcohol, embedded in paraffin. Tumor tissues were dissected from unstained histological sections. DNA from the tumor tissues was isolated and purified using the DNA Tissue Isolation kit (Qiagen GmbH, Hilden, Germany) according to the manufacturer’s protocol. KRAS and EGFR mutations were subsequently identified using the Human KRAS and EGFR Mutation Detection kit (Amoy Diagnostics, Xiamen, China) according to the manufacturer’s protocol. Mutational analyses of EGFR gene exons 18, 19, 21, and 21 and KRAS exons 12 and 13 were performed using the loop-hybrid mobility shift assay and sequencing as described previously (15).

Statistical analysis. All analyses were performed using SPSS 19.0 software (IBM Corp., Armonk, NY, USA). Data are expressed as the mean ± standard deviation. The χ² test and Fisher’s exact test were used to compare the observed percentages. Fisher’s exact test was performed if there were ≤5 observations in a group. P<0.05 was considered to indicate a statistically significant difference.

Results

Clinicopathological characteristics. Of the 1,469 lung adenocarcinoma tumours, 29 tumours were PPMA. The clinicopathological data for these cases are summarised in Table I. Of these patients, 12 were male and 17 were female, and the median age was 69 years (ranging from 35 to 83 years old). Smokers (smoking index ≥200; smoking index equalled the average root number of cigarettes smoked per day multiplied by number years of smoking) accounted for 34.5% (10/29) of the patients. The maximum diameters of the tumours ranged from 0.9-13 cm. The predominant histologic patterns were IMA (72.4%, 21/29), AIS and MIA (27.6%, 8/29). A total of 31% (9/29) of patients presented with pleural invasion. Lymphatic invasion was observed in 10 cases (34.5%). No instances of pulmonary metastasis or vascular invasion were observed.

Immunohistochemistry analysis. Immunohistochemical analysis for markers of pulmonary differentiation was performed on one slide of tumour tissue per case. The primary antibodies that were used were as follows: TTF-1 (1:200 dilution; cat. no. IR056; Dako; Agilent Technologies, Inc., Santa Clara, CA, USA), Napsin A (1:200 dilution; cat. no. NCL-L-Napsin A; Leica Microsystems, Inc., Buffalo Grove, IL, USA), cytokeratin 7 (CK7; 1:200 dilution; cat. no. IR619; Dako; Agilent Technologies, Inc.), CK20 (1:200 dilution; cat. no. IR777; Dako; Agilent Technologies, Inc.), EGFR (1:100 dilution; cat. no. NCL-L-EGFR; Leica Microsystems, Inc.), tumor protein (P)53 (1:200 dilution; cat. no. IR616; Dako; Agilent Technologies, Inc.), carcinoembryonic antigen (CEA; 1:200 dilution; cat. no. IR622; Dako; Agilent Technologies, Inc.), Cam5.2 (1:100 dilution; cat. no. RTV-CAM5.2-0.11; Leica Microsystems, Inc.) and Villin (1:200 dilution; cat. no. IR076; Dako; Agilent Technologies, Inc.) antibodies. Staining of <5% of tumour cells was interpreted as negative, staining of 5-80% of tumour cells was classified as mixed, and staining in >80% of tumour cells was interpreted as positive. Histological classification and the interpretation of the immunohistochemical results were based on a consensus review by three pathologists.

EGFR and KRAS mutation analysis. The tumor tissues were surgically resected for all patients and then fixed for 24 h in 10% neutral buffered formalin solution at room temperature, washed in distilled water, dehydrated in graded alcohol, embedded in paraffin. Tumor tissues were dissected from unstained histological sections. DNA from the tumor tissues was isolated and purified using the DNA Tissue Isolation kit (Qiagen GmbH, Hilden, Germany) according to the manufacturer’s protocol. KRAS and EGFR mutations were subsequently identified using the Human KRAS and EGFR Mutation Detection kit (Amoy Diagnostics, Xiamen, China) according to the manufacturer’s protocol. Mutational analyses of EGFR gene exons 18, 19, 21, and 21 and KRAS exons 12 and 13 were performed using the loop-hybrid mobility shift assay and sequencing as described previously (15).
in 18/29 (62.1%) cases. Positive CK7 expression was identified in all cases; 100% of tumour cells expressed this cytokeratin, but 5 (17.2%) tumours co-expressed CK20. A total of 8 (27.6%) cases expressed EGFR, 14 (48.3%) cases expressed P53, but only 2 (6.9%) cases expressed CEA. There were 9 (31%) cases that expressed Napsin A. In all of the cases, a large proportion of carcinoma cells expressed Villin and 11 (37.9%) cases expressed Cam5.2. The co-expression of CDX2 and TTF1 was not identified within single tumour cells (Fig. 1).

**EGFR and KRAS mutations.** The EGFR and KRAS mutation statuses of the patients are summarised in Table II. A total of 18 cases (62%) exhibited a KRAS mutation and an EGFR mutation was identified in 2 cases (6.9%). Exon 12 was mutated in all of the KRAS mutation cases, while the 2 EGFR mutations occurred in exon 21 (L858R) of EGFR. A KRAS and EGFR co-mutation was detected in 1 case. A total of 10 patients exhibited neither EGFR nor KRAS mutations.

**Association between the clinicopathological characteristics and KRAS and EGFR mutations.** In the 29 patients with PPMA, KRAS mutations were significantly more likely to be located in the lower lung lobe (73.9%, 17/23) compared with in the upper/middle lobe (16.7%, 1/6; P=0.036). Additionally, the number of tumours with a KRAS mutation was increased in patients with invasive mucinous adenocarcinoma (76.2%, 16/21) compared with in other subtypes (25%, 2/8; P=0.035). No significant associations were identified between KRAS or EGFR mutations and other clinicopathological parameters (Table III). There were no significant associations between histological features, including lymphatic permeation or pleural invasion, and KRAS or EGFR mutations (Table III).

**Prognosis following resection.** The clinical outcomes of patients with Mas were then examined. The median follow-up period for the surviving patients was 74.8 months (range, 34-132 months). No serious complications following the surgeries and no surgical mortalities were noted. A total of 1 patient was lost to follow-up, and none of the 3 patients with AIS or the 5 patients with MIA exhibited recurrences. A total of 8 patients exhibited recurrences. With regard to the site, the first recurrence was an ipsilateral pulmonary metastasis in 3 patients and bilateral pulmonary metastasis in 5 patients. Of these patients, 5 succumbed to recurrent lung cancer and 3 were alive with disease. The 11 patients with IMA were asymptomatic, and are currently being managed with close radiologic and clinical follow-up. The overall and relapse-free survival rates at 5 years were 70.4, and 81.5%, respectively.

**Discussion**

The 2015 WHO Classification of Lung Tumors: Impact of Genetic, Clinical and Radiologic Advances Since the 2004
Classification has been published recently (6). Compared with the 2004 WHO Classification, there are multiple major changes for common types of lung cancer, the majority of which follow the 2011 lung adenocarcinoma classification system that was sponsored by the International Association for the Study of Lung Cancer (IASLC)/American Thoracic Society/European Respiratory Society, which has essentially been adopted with minor changes (14). IMAs, which account for 2-10% of lung adenocarcinoma cases in East Asia, Europe, and the United States (16-18), were considered to demonstrate increased levels of malignancy compared with other common subtypes of lung adenocarcinoma, including lepidic and acinar subtypes (19).

Lung adenocarcinoma has been associated with poor recurrence-free survival and adverse prognosis, as the disease often presents with minimal or misleading clinical symptoms, and multifocal heterogeneous unresectable lesions (20). In the present study, PPMA was classified as AIS, MIA or IMA according to the new classification of lung adenocarcinoma by IASLC (14). The diagnosis of AIS, MIA or IMA cannot be made without the complete histological evaluation of the entire tumour. In the present study, the maximum diameters of the tumours ranged from 0.9-13 cm. Pleural invasion was observed in 9 cases (31%) and lymphatic invasion was present in 10 cases (34.5%).

In the present study, CK7 expression was identified in all of the PPMA tumours, most often combined with TTF1, EGFR, P53, Napsin A, Villin or Cam5.2 expression. Only a minority of tumours expressed CK20 and CEA. Geles et al (21) confirmed that the number of TTF1-negative mucinous adenocarcinomas was quite high, which may suggest differences between a Caucasian and an Asian population. In the study by Kunii et al (22), MUC2 and MUC5AC expression, together with HNF4α, was observed in cases that were negative for TTF1.

The presence of EGFR mutations has been demonstrated as a predictive factor for a good response to EGFR-tyrosine kinase inhibitors (15,23). Mucinous adenocarcinoma is associated with the absence of EGFR mutations and the presence of KRAS mutations (24). One previous study (25) indicated that EGFR mutations are not commonly detected in patients with mucinous adenocarcinoma. In the present study, it was indicated that invasive mucinous adenocarcinoma of the lung is genetically and clinically distinct. EGFR mutations were detected in 2/29 patients (6.9%), and these mutations have been previously described to range from 0 to 22% (25). KRAS mutations were detected in 18/29 patients (62%) in the present study, in concordance with the data of previous studies in which these mutations have been previously described to range from 42.5 to 86% (7,23,25). The results of the present study for EGFR and KRAS mutations were similar to those in previous studies. The KRAS mutations may be associated with the carcinogenesis of mucinous adenocarcinoma. In terms of specific KRAS mutations, exon 12 was most common in the cohort of the present study. KRAS exon 12 is most common in colorectal and pancreatobiliary carcinoma, suggesting that IMA of the lung may have more in common with pancreatobiliary, and intestinal tract cancer types (26).

Several studies have analysed the association between KRAS or EGFR mutations and the clinicopathological characteristics in pulmonary carcinoma. The results of the present study suggest that KRAS or EGFR mutations in
patients with PPMA were not significantly associated with age, sex, smoking index, tumour size, stage, pleural invasion and lymphatic permeation. In addition, it was identified that KRAS mutations were more often observed in invasive mucinous adenocarcinomas compared with AIS or MIA, and additionally more frequently observed in tumours that were localised to the lower lobe than in the upper or middle lobe of the lung.

Mucinous differentiation in pulmonary adenocarcinoma appears to occur early during carcinogenesis, and is most likely already present in the precursor lesion (21). Survival data for patients with PPMA have been limited due to its low incidence and the results of the small number of published studies have been conflicting (26). Shim et al (26) demonstrated that there was no significant difference in recurrence-free survival between patients with invasive mucinous adenocarcinoma and other patients. In the present study, all incidences of recurrence were limited to the lungs and extrapulmonary metastases were not detected in the pulmonary mucinous adenocarcinoma cohort, even though patients with invasive mucinous adenocarcinoma exhibited a tendency towards an improved recurrence-free survival.

In conclusion, primary pulmonary mucinous adenocarcinoma is a rare type of low grade malignant tumour. KRAS mutations may promote lineage-specific tumorigenesis in a pathological subtype of lung adenocarcinoma. The most effective treatment for PPMA is surgical resection, which demonstrated a tendency towards improved survival. The results of the present study may be crucial for the development of effective treatment for patients with PPMA. Additional validation studies are required to improve the targeted therapy strategies for mucinous adenocarcinoma.

Table III. Clinicopathological parameters of the 29 patients with primary pulmonary mucinous adenocarcinoma included in the present study.

| Clinicopathological parameters | KRAS | EGFR |
|--------------------------------|------|------|
|                               | Positive (n=18) | Negative (n=11) | P-value | Positive (n=2) | Negative (n=27) | P-value |
| Age (years)                   |      |      | 0.271 |      |      | 1 |
| <65                           | 10   | 3    | 1     | 12   | |
| ≥65                           | 8    | 8    | 1     | 15   | |
| Sex                           |      |      | 0.461 |      |      | 1 |
| Male                          | 6    | 6    | 0     | 12   | |
| Female                        | 12   | 5    | 2     | 15   | |
| Smoking index                 |      |      | 0.298 |      |      | 0.532 |
| <200                          | 10   | 9    | 1     | 19   | |
| ≥200                          | 8    | 2    | 1     | 8    | |
| Tumour size (cm)              |      |      |       | 0.135 |      | |
| ≤3                            | 10   | 8    | 0.596 | 0     | 18   | |
| >3                            | 8    | 3    | 2     | 9    | |
| Lobe                          |      |      | 0.036 |      |      | 0.377 |
| Upper/middle                  | 1    | 5    | 1     | 5    | |
| Lower                         | 17   | 6    | 1     | 22   | |
| Stage                         |      |      | 0.45  |      |      | 0.532 |
| I-II                          | 11   | 9    | 1     | 19   | |
| III-IV                        | 7    | 2    | 1     | 8    | |
| Histological subtype          |      |      | 0.035 |      |      | 0.483 |
| AIS and MIA                   | 2    | 6    | 1     | 7    | |
| IMA                           | 16   | 5    | 1     | 20   | |
| Pleural invasion              |      |      | 0.369 |      |      | 0.089 |
| Positive                      | 4    | 5    | 2     | 7    | |
| Negative                      | 14   | 6    | 0     | 20   | |
| Lymphatic permeation          |      |      | 0.298 |      |      | 0.111 |
| Positive                      | 8    | 2    | 2     | 8    | |
| Negative                      | 10   | 9    | 0     | 19   | |

AIS, adenocarcinoma in situ; MIA, minimally invasive adenocarcinoma; IMA, invasive mucinous adenocarcinoma.
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