EVALUATION OF PHYTOCONSTITUENTS, AND ANTIMICROBIAL AND ANTIOXIDANT ACTIVITY OF JUGLANS REGIA FROM KASHMIR REGION

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INTRODUCTION

Medicinal plants act as best basic material and a great source of phytoconstituents that have infinite application in medicine. Plants with medicinal value are used to cure several diseases since ancient times. Medicinal plants getting familiarized in various countries because they exist in nature with fewer side effects. *Juglans regia* is also included in this category. *J. regia* is a big deciduous tree found mainly in Iran, Baluchistan, Himalayan regions of India, Armenia, and several temperate regions [1]. In India, Kashmir occupies the largest position in the total production of walnut. It is highly beneficial and common plant of Kashmir where it is known as Doon. People in this region use different parts of this tree to treat several diseases. Many medicinal plants getting familiarized in various countries around the globe. Dependability on herbs has to continue to increase because they exist in nature with fewer side effects [2]. In addition to their anticarcinogenic activity, the walnut bark has antioxidant and metal chelating activity [3]. The bark of the walnut tree contains flavonoids, phenolic compounds, alkaloids, and steroids [4]. The bark from *J. regia* is used as a tooth cleaner. In dentistry, walnut stem bark is recognized treat infections initiated by oral pathogens. >760 species of bacteria are responsible for oral infection, of which 50% are unknown [5]. The worldwide requirement for alternative prevention, treatment choices, and products for oral infections that are harmless, efficient, and cost-effective comes from the elevation in disease occurrence usually in developing countries, rise in multidrug resistance by several pathogenic microorganisms to antibiotics and several drugs [6,7]. The World Health Organization in 2007 reported that community health expenditure specific to dental care was 5–10%. Tooth decay and oral infection treatment are probably very expensive that people have to struggle with for a lifetime [8]. Natural plant products are used even to correct numerous oral infections. In Burkina Faso, West Africa >62 species of plants belong to 29 families documented to cure oral infections. It is supposed that a quarter of recommended medicine contains constituents taken from the plants in industrialized countries [9]. Despite the great progress in medical sciences, plants are still thought to be a vital source of different drugs in various countries around the globe. Dependability on herbs has to turn out to be vital due to adverse effects possessed by chemical drugs such as tooth discoloration, change in taste, and formation of resistant microbes restricted its use.

METHODS

Sample collection
The bacterial strains *Streptococcus mutans* (MTCC-890), *Staphylococcus aureus* (MTCC-737), and *Pseudomonas aeruginosa* (MTCC-741) were obtained from IMTECH, Chandigarh. The clinical strains of *P. aeruginosa* and *S. aureus* were obtained from Molecular and Immuno-Parasitology Research Laboratory of Shoolini University. These strains were cultivated in broth (Nutrient broth) followed by incubation for 24 h at 37°C. The
bark of *J. regia* stem was collected in July from Kulgam district in Jammu and Kashmir.

**Plant material processing**

The plants collected were authenticated with farm receipt no. 048 and book no. 2915. The bark was surface sterilized with mercuric chloride and dried in shade for 14 days. The dried bark was then crushed into a powdered form and extracted with methanol for 2 days using orbital shaker method. The extracted material was separated using Whatman filter paper no.1, the methanol was evaporated in a water bath at temperature <40°C, and then, crude extract was kept in a refrigerator at 4°C for further use.

**Antibacterial activity**

The antibacterial property of plant extract was determined using well diffusion assay [10]. The 0.5 McFarland was used as a positive control. DMSO was also added in a well as negative control to confirm that DMSO does not possess any antibacterial activity. After 24 h of incubation, the inhibition zone was measured and compared with an inhibition zone of the standard antibiotic used.

**Phytochemical screening**

Methanolic extract (ME) was screened for the detection of phytocomponents. Results of phytoconstituents screening are summarized in Table 1. Phytochemical screening was performed in test tubes. The observations (Table 1) confirmed the presence of phytoconstituents such as phenols, flavonoids, terpenoids, tannins, glycosides, phlobatannins, and cardiac glycosides.

**Total phenolic estimation**

In alkaline medium, the reaction of phenols with the phosphomolybdic acid of Folin–Ciocalteu reagent takes place and forms blue-colored complexes that are determined spectroscopically [11]. The overall phenols present in the test tubes. The observations (Table 1) confirmed the presence of phytoconstituents such as phenols, flavonoids, terpenoids, tannins, glycosides, phlobatannins, and cardiac glycosides.

**Total flavonoid content (TFC) estimation**

The AlCl₃ colorimetric method was performed to know the concentration of flavonoids in the sample [12]. In this method (AlCl₃ colorimetric method), acid stable compounds are formed by AlCl₃ with C-4 keto and C-5 hydroxyl groups of flavonoids. Studies have described quercetin to be a proper standard to determine the concentration of flavonoids in a sample extract. Hence, quercetin solution with different concentrations was taken to create the calibration curve. 10 mg of quercetin was mixed in 10 ml of methanol and then diluted to different concentrations of 20, 40, 60, 80, and 100 µg using methanol. The assay was performed using 0.1 ml of extract from stock solution (mg/ml), 0.2 ml of 5% sodium nitrate was added in each test tube. After 5 min, 0.2 ml of 10% AlCl₃ was added to the mixture, and finally, after 6 min, 2 ml of 1 M NaOH was added. Absorbance was taken at 510 nm. A standard curve with several concentrations of quercetin was drawn on the basis, of which concentration of phenols in the sample was calculated and expressed as mg/g.

**Free radical scavenging activity**

The free radical scavenging property of extract was calculated using 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay [13]. Using

![Fig. 1: Inhibition zones of methanolic extracts of *Juglans regia* against oral bacteria](image)
methanol, 0.004% w/v of DPPH solution was prepared using methanol. Stock solution (1 mg/ml) of the sample and ascorbic acid were made using methanol. Several concentrations of sample and ascorbic acid were prepared in test tubes and 2.5 ml DPPH was added and tubes were kept in dark for 30 min. After incubation, the absorbance was taken at 517 nm using a spectrophotometer. A control sample was prepared using methanol and DPPH. Methanol was taken as a blank.

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\text{Absorbance of control} - \frac{\text{Absorbance of sample}}{\text{Absorbance of control}} \times 100
\]

**Statistical analysis**

All the results data have been expressed in the form of mean ± standard deviation.

**RESULTS**

**Antibacterial activity**

Antibacterial activity of ME of *J. regia* was investigated and the results revealed different inhibition zones as shown in Table 1 and Fig. 1. The result showed that the ME was found to be potential antibacterial agents against the oral pathogenic bacteria which are responsible for different types of oral infections.

**Phytochemical screening**

The result revealed that MEs of *J. regia* contain flavonoids, phenols, terpenoids, cardiac glycosides, saponins, and tannins, whereas carbohydrates, alkaloids, fats, and steroids were found absent. Preliminary phytochemical screening results were described in Table 2.

**Total phenols and flavonoids estimation**

Using standard curve equation of gallic acid and quercetin equivalents, the total phenolic content (TPC) and TFC of methanolic bark extract of *J. regia* was calculated and was found to be 43.35±0.079 mg/g and 17.28±0.125 mg/g, respectively (Figs. 2 and 3, Table 3).

**Free radical scavenging activity**

The free radical scavenging activity of methanolic bark extract of *J. regia* was evaluated using DPPH. It was seen that the scavenging activity of DPPH increased with the increase in concentration for both standard ascorbic acid and methanolic bark extract of *J. regia* interestingly (Fig. 4).

**DISCUSSION**

The main agent responsible for gum inflammation and dental cavities is a dental plaque. Some people avoid the use of chemical mouth rinse due to side effects such as tooth discoloration, change in taste, and formation of resistant microbes [14]. Hence, currently, the researchers are looking for medicinal plants to eliminate different kinds of infections. The current study showed that *J. regia* has a capability to serve as a potential antibacterial agent and thus can be used in oral hygiene products. According to our study, we indicated that ME of *J. regia* was effective in inhibiting the three bacteria *S. mutans*, *S. aureus*, and *P. aeruginosa*. The inhibition zones are shown in Table 1 and Fig. 1. Mohammed, 2012, reported the inhibition effect of ME of *J. regia* bark against *S. aureus*, *S. mutans*, and *P. aeruginosa* [15] which were less effective as compared to our methanolic bark extract. The antibacterial property possessed by the plant is due to the presence of phenolic compounds, flavonoids, tannins, and terpenoids [16]. Phenols are very essential plant components due to their capability to scavenge free radicals as they contain hydroxyl group groups in their structure; therefore, plant phenols may contribute directly to their antioxidant potential. Some studies were performed to know the amount of phenolics present in the bark extract of *J. regia*. Asha et al., 2010, reported that the presence of total phenols and flavonoids in bark extract of *J. regia*, the TPC and TFC, was 20.32 mg/g and 11.48 mg/g, respectively, of dry weight [17]. On the other hand, Ogunmoyole et al., 2011, reported the values of TPC and TFC ranging from 35.2±0.75 mg/g to 20.2±0.12 mg/g [18]. Interestingly, the result indicates higher phenolic and flavonoid content in our sample too.

![Graphical representation of the standard curve of gallic acid](image1)

![Graphical representation of the standard curve of quercetin](image2)

![DPPH scavenging activity of standard ascorbic acid and methanolic bark extract of *Juglans regia*](image3)
DPPH which dissolves in methanol has purple color and defined absorbance at 517 nm. When antioxidant donates protons to DPPH, the purple color of the solution changes to yellow color with the decrease of absorbance [19]. The absorbance is inversely proportional to free radical scavenging activity. The scavenging activity of *J. regia* bark extract was found to be greater at a low concentration ranging from 20 to 80 µg/ml (47.16–79.31%) while Kshitij et al, 2012, have reported at higher concentration of 50–500 µg/ml (20–80%).

**CONCLUSIONS**

The results obtained in the study clearly show that *J. regia* bark may be considered a good candidate for employment as an effective antimicrobial agent against oral bacteria which can cause many oral diseases. To elevate oral sanitation, medicinal plant-based mouthwash can be used as a mediator and acts as a part of efficient home care medication. *J. regia* bark may also be included as a good source of healthy compounds such as phenols and flavonoids, suggesting that it could be useful in the prevention of diseases in which free radicals are present.

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**CONFLICTS OF INTEREST**

The authors declare that they no conflicts of interest.

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