Laboratory evaluation of four insecticides on the mosquito Culiseta longiareolata (Macquart) (Diptera: Culicidae).

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Abstract

The efficacy of four insecticides belonging to different groups, synthetic pyrethroid (lambdacyhalothrin), carbamate (Marshal), fungicide (Topas) and insect growth regulator (Neemrich) as well as their joint action were tested against 3rd larval instar of Culiseta longiareolata. Based on concentration mortality data LC50 and LC90 values, results obtained showed that LC50 values as observed were 0.09, 2.3, 9.0 and 25.0 ppm for lambdacyhalothrin, Marshal, Neemrich and Topas, respectively. The results showed that all mixtures consisted of a 1:1 (v/v) ratio of the LC25 of each compound indicated potentiating effect. The highest potentiating effect was achieved by a mixture of co-toxicity factor equaled +100 (lambdacyhalothrin + Neemrich). The lowest potentiating effects were obtained from mixtures of Marshal + Topas and Topas+Neemrich (co-toxicity factor equaled +40). The tested insecticides completely inhibited the emergence of adults till 0.0078, 0.0625, 0.25 and 1 ppm for lambdacyhalothrin, Marshal, Topas and Neemrich, respectively, and the emergence of adults was inversely proportional to the concentration.

Keywords: Insecticides, Culiseta longiareolata, Diptera Culicidae

1 Introduction

Cs. Longiareolata (Macquart) is a biting nuisance mosquito in Egypt.it is a widespread species and found in a high population density Kirkpartrick,(1925), Kenawy & Elsaid,(1989) and Teleb (1994). Cs.longiareolata is also incriminated as a vector of transmitting Plasmodium Reticulum, the causative organism of Malta fever Hewitt,(1940) and intermediate host of avian plasmodia, Gutsevich et al.,(1970). Chemical control is an effective strategy used extensively in daily life. The control of mosquito at the larval stage is necessary and efficient in the integrated approach to mosquito management. Resistance to carbamates has been noted in Cx.quinquefasciatus. The use of mixtures to a strategy of rotation over time of insecticides with different modes of action has already made it possible to prevent or to delay the appearance of resistance in the field Martin et al.,(2000). However, mixtures of appropriate dosages of unrelated compounds may have better prospects for managing resistance effectively than rotations of the types of compounds. The advantage of mixtures is that each insecticide eliminates most insects which are genetically susceptible to it, Barnes et al., (1995). However, many authors have already demonstrated the synergistic effect on insect pests of carbamates or organophosphates and pyrethroids ,Koziol & Witkowski, (1982),Ozaki et al., (1984) and Roberston & Smith, (1984) with insects of medical importance, a synergistic effect between pyrethroids and carbamates was reported on larvae of Cx.quinquefasciatus ,Corbel et al., (2003) and adults of Anopheles gambiae Corbel et al., (2002) susceptible to these insecticides. Insecticide mixtures have been proposed as an important tools for resistance management in different insect pests (Hemingway and Ranson, 2000). This type of potentiation or synergism is explained by the inhibition of esterases ,Bryne and Devonshire (1991) and Montella et al.,(2012) or monooxygenases activity ,Martin et al.,(2003). The present work was carried out to clarify the toxic effect of four insecticides which are regularly used in fields; lambda-cyhalothrin, Marshal, Topas and Neemrich. The interaction between them against the 3rd larval instar of Cs. Longiareolata and the effects of these insecticides on emergency of adults.
2 Materials and Methods

Mosquito culture
Cs.longiareolata larvae were collected from wells near Zagazig city in Sharkia Governorate and reared under laboratory conditions (25±2°C and 80±5% relative humidity) for several generations.

Insecticides
Commercial formulations of insecticides used for bioassays are: Pyrethroid (lambda-cyhalothrin), Carbosulfan (Marshal 25%), Topas (with the active compound Penconazole, is a systemic fungicide) and Neemrich. These chemicals obtained from Syngenta Agro Egypt Company - Egypt.

Larvicidal bioassay
For each insecticide seven concentrations were prepared by diluting the formulation product with distilled water in plastic cups (250ml) against the 3rd larval instar of Cs.longiareolata. Twenty five larvae were placed in each cup. The test was carried out at the same conditions of rearing. Larvae were left for 24 h and mortality was then recorded and compared with control. Moribund larvae were considered dead. Four replicates per concentration were used. WHO Technique (1996) was used for measuring the susceptibility of larvae to given insecticides. Mortality was corrected according to Abbott formula (1925).

Joint action study
Concentration – mortality curves were established and the LC25 values were determined. Binary mixtures were prepared in proportion to their toxicity equivalents of LC25. The combined action of each mixture was expressed as the co-toxicity factor (C.F), estimated according to the equation given by Sun and Johnson (1960):

\[
\text{C.F} = \frac{\% \text{observed mortality} - \% \text{expected mortality}}{100}
\]

A positive factor of 20 or more is considered potentiation, A negative (-20) or more is considered antagonism, and intermediate values ranging between -20 and +20 indicate only additive effect.

Pupicidal bioassay
Ten replicates of newly developed pupae were transferred into plastic cups 10 cm height (4/ cup) containing different concentrations of each of the tested insecticide. The number of emerging adults was observed, calculated daily and compared with control (untreated).

Data analysis
Data obtained from each concentration. Larvicidal bioassay (total mortality) were subjected to probit analysis (Finney, 1971) and LC25, LC50 and LC90 values were calculated. All results were expressed as mean ± standard error, and the data were analyzed using student T-test. Results with p<0.05 were considered to be statistically significant.

3 Results
Larvicidal bioassay
The results presented in table 1 showed that all insecticides induced mortality on the 3rd larvae of Cs. longiareolata. On the basis of LC50 values, the larvical toxicity of lambda-cyhalothrin was the most potent (LC50= 0.08ppm) followed by Marshal (LC50= 2.4 ppm), Neemrich(LC50= 8.7ppm) and Topas(LC50= 24.6 ppm). Respectively. Similar trend has also been observed for LC50 (0.3,11.7,51.7 and 106.3 for lambda-cyhalothrin, Marshal, Neemrich and Topas, respectively.

Joint action study
The interaction of binary mixtures of tested insecticides against the 3rd larval instar of Cs. longiareolata is shown in table 2. The calculated “co-toxicity factor” exceeded 20; a results accounting to “potentiation effect”. All the mixtures exhibited potentiation effect. The highest potentiating effect was for mixtures of co-toxicity factor equaled +100 (lambda-cyhalothrin+ Neemrich). The lowest potentiating effect (±40) was obtained from mixtures of Marshal + Topas and Topas + Neemrich.

Pupicidal bioassay
Data concerning pupicidal activity are shown in table 3. The results revealed that all insecticides caused 100% complete inhibition of the adult emergence at 0.0078 ppm, for Lambda cyhalothrin, 0.0625 for Marshal , 1.0 for Neemrich , 0.25 ppm for Topas, The adult emergence increased by dilution. Zidan et al., (1997) found that the synthetic pyrothroid ,cyphenothrin showed complete inhibition of adult emergence of Cx. pipiens pupae.

4 Discussion
Kawakami,(1989) tested permethrin against Cx. pipiens larvae and found that LC50 was 0.01 ppm. Basset et al., (1997) found that Cx.pipiens subjected to Lambda – cyhalothrin did not develop cross resistant to cypermethrin, this was in contradiction with Xia et al.,(1998) who reported that Cx.pipiens resistance to Lambda –cyhalothrin developed 41 and 28 fold resistance to permethrin and cypermethrin, respectively.

There are several studies on mixture toxicities (particularly pyrethroids with other compounds) in different dipteran insect pests worldwide. Since pyrethroids and organophosphates have different modes of action, their mixtures have commonly been in practice against a variety of pests worldwide for the last many years Ahmad, (2009). Previously it has been assumed that organophosphates, when used in combination with pyrethroids, inhibit the enzymes responsible for metabolic detoxification in different insect pests Martin et al., (2003) Corbel et al.,(2003) showed that propoxur at LC50 significantly enhanced the insecticidal activity of permethrin. Ali Khan et al., (2013) showed that most of the insecticide mixtures
Table (1) Efficacy of the four insecticides on the 3rd larval instar of *Cs. Longiareolata*

| Insecticide                  | Lc values (ppm) | Slope function |
|------------------------------|-----------------|----------------|
|                              | Lc25            | Lc50           | Lc90           |                            |
| Lambda-cyhalothrin           | 0.04 (0.25-0.45)| 0.08 (0.04-0.09)| 0.3 (0.25-0.42)| 3.6 ± 0.1                  |
| Marshall                     | 0.5 (0.1-0.8)   | 2.4 (2.0-3.0)  | 11.7 (10.0-13.0)| 9.4 ± 0.6                  |
| Neemrich                     | 3.7 (3.2-4.0)   | 8.7 (8.0-0.9)  | 51.7 (45.0-60.0)| 22.5 ± 0.8                 |
| Topas                        | 11.0 (9.0-13.0) | 24.6 (22.0-27.0)| 106.3 (102.0-108.0)| 3.6 ± 0.1                  |

Values between brackets are 90% fiducial limits of the corresponding toxicity values. The latter values are estimated from their respective regression lines (LC-P lines).

Table (2) The joint action of four insecticides on the 3rd larval instar of *Cs. Longiareolata*

| Binary mixtures              | Lc25/ppm For Compound | Observed mortality | Co-Toxicity Factor |
|------------------------------|------------------------|---------------------|--------------------|
|                              | Compound 1             | Compound 2          | %                  |                      |
| Lambda-cyhalothrin + Marshall | 0.04 + 0.5             | 90                  | +80                |
| Lambda-cyhalothrin + Neemrich | 0.04 + 3.7             | 100                 | +100               |
| Lambda-cyhalothrin + Topas   | 0.04 + 11.0            | 85                  | +70                |
| Marshall + Neemrich          | 0.5 + 3.7              | 80                  | +60                |
| Marshall + Topas             | 0.8 + 11               | 70                  | +40                |
| Topas + Neemrich             | 11 + 3.7               | 70                  | +40                |

like one pyrethroid with other compounds significantly increased the toxicity of pyrethroids in the field population of house flies, *Musca domestica* L.

In addition, insecticides from pyrethroid and organophosphate classes may be potential or competitive substrates for the same oxidase, as demonstrated by Kulkarni and Hodgson (1980) thus potentiating the toxicity of the insecticide mixture. The pupicidal activity of insecticides was also studied by Fournet et al., (1993), and Trayler et al.,(1994).

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Table (3) Pupicidal activity of four insecticides on *Cs.Longiareolata* pupae

| Conc.(ppm) | % Emergency of adults | (Mean S.E.)* |
|------------|-----------------------|--------------|
|            | Lambda cyhalothrin    | Marshal      | Neemrich | Topas |
| 1          | 0.0                   | 0.0          | 0.0      | 0.0  |
| 0.5        | 0.0                   | 0.0          | 12.5 ± 3.5 | 0.0 |
| 0.25       | 0.0                   | 0.0          | 31.3 ± 3.5* | 0.0 |
| 0.125      | 0.0                   | 0.0          | * 50.0 ± 5.1 | 12.5 ± 3.5* |
| 0.0625     | 0.0                   | 0.0          | 75.0 ± 5.1* | 31.3 ± 3.1* |
| 0.0312     | 31.3 ± 3.1*           | 100.0±0.0  | 4305±2.4* |
| 0.0156     | 56.3 ± 3.1*           | —            | 56.3±3.1* |
| 0.0078     | 68.8 ± 5.5*           | —            | 75.0 ± 3.5* |
| 0.0039     | 25.0 ± 5.1*           | 87.5±3.4*   | —        | 87.5±3.4* |
| 0.0018     | 56.3±5.1*             | 100.0±0.0   | —        | 100.0±0.0 |
| 0.0009     | 83.3±3.3*             | —            | —        | —    |
| 0.00045    | 100.0±0.0             | —            | —        | —    |
| Control    | 100.0±0.0             | 100.0±0.0   | 100.0±0.0 | 100.0±0.0 |

* Significant P=0.05
**Results are the means of ten replicate

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