A Systematic Review of Candidate miRNAs, Its Targeted Genes and Pathways in Chronic Myeloid Leukemia–An Integrated Bioinformatical Analysis

Marjanu Hikmah Elias1*, Syarifah Faezah Syed Mohamad1,2 and Nazefah Abdul Hamid1

1 Faculty of Medicine and Health Sciences, Universiti Sains Islam Malaysia, Nilai, Malaysia, 2 Faculty of Applied Sciences, Universiti Teknologi MARA Cawangan Pahang, Jengka, Malaysia

Chronic myeloid leukaemia is blood cancer due to a reciprocal translocation, resulting in a BCR-ABL1 oncogene. Although tyrosine kinase inhibitors have been successfully used to treat CML, there are still cases of resistance. The resistance occurred mainly due to the mutation in the tyrosine kinase domain of the BCR-ABL1 gene. However, there are still many cases with unknown causes of resistance as the etiopathology of CML are not fully understood. Thus, it is crucial to figure out the complete pathogenesis of CML, and miRNA can be one of the essential pathogeneses. The objective of this study was to systematically review the literature on miRNAs that were differentially expressed in CML cases. Their target genes and downstream genes were also explored. An electronic search was performed via PubMed, Scopus, EBSCOhost MEDLINE, and Science Direct. The following MeSH (Medical Subject Heading) terms were used: chronic myeloid leukaemia, genes and microRNAs in the title or abstract. From 806 studies retrieved from the search, only clinical studies with in-vitro experimental evidence on the target genes of the studied miRNAs in CML cells were included. Two independent reviewers independently scrutinised the titles and abstracts before examining the eligibility of studies that met the inclusion criteria. Study design, sample size, sampling type, and the molecular method used were identified for each study. The pooled miRNAs were analysed using DIANA tools, and target genes were analysed with DAVID, STRING and Cytoscape MCODE. Fourteen original research articles on miRNAs in CML were included, 26 validated downstream genes and 187 predicted target genes were analysed and clustered into 7 clusters. Through GO analysis, miRNAs’ target genes were localised throughout the cells, including the extracellular region, cytosol, and nucleus. Those genes are involved in various pathways that regulate genomic instability, proliferation, apoptosis, cell cycle, differentiation, and migration of CML cells.

Keywords: chronic myeloid leukemia, genes, pathways, microRNA, BCR-ABL1
INTRODUCTION

Chronic myeloid leukaemia (CML) is a proliferative disorder of pluripotent stem cells. CML is linked to a specific genetic disorder involving BCR and ABL1 gene translocation, resulting in the Philadelphia chromosome. BCR-ABL1 fusion gene encodes an active tyrosine kinase BCR-ABL which activates several molecular pathways that cause abnormal cell adhesion, increase cell proliferation, and inhibit apoptosis. Nevertheless, tyrosine kinase plays an essential role in many signalling cascades, including biological processes such as cell growth, differentiation, metabolism, and apoptosis (1). Numbers of studies have revealed the mechanisms of CML pathogenesis which involved several key signalling pathways, including the MAPK, JAK-STAT, PI3K/AKT, EGFR, ERBB, TGF-β and tumour protein p53 pathways (2–4).

Thus, specific treatment for CML has been developed by targeting the BCR-ABL1 gene. The first molecularly targeted therapy, Imatinib, is a small molecule known as tyrosine kinase inhibitor (TKI) that directly targets BCR-ABL1 tyrosine kinase activity. Despite the success of Imatinib as the front-line therapy for CML, there were reports on drug resistance that is primarily due to the presence of mutations in the BCR-ABL1 tyrosine kinase domain (TKD) (5–7). Tyrosine kinase inhibitors are not able to fully prevent the progression of CML cells with BCR-ABL TKD mutated CML cells. Therefore, research into alternative treatments for CML remains clinically essential. In recent years, microRNA (miRNA) has been widely studied in human malignancies and chemical compounds. They have received widespread attention as essential regulators of gene expression in leukemogenesis and are linked to resistance to BCR-ABL TKIs (3, 8).

MiRNA is a short, non-coding RNA that regulates gene expression at the post-transcriptional level. It inhibits translation by binding to the 3’untranslated (3’UTR) region of specific target mRNA. MiRNA has been linked to disease pathogenesis in CML and is known to play an essential role in tumorigenesis (9). MiRNA’s roles and functions have been highlighted in several studies in a variety of circumstances and scenarios. For instance, miRNAs expression profiles were used as biomarkers and therapeutic tools (10–12). Their expression differences were used in several studies to improve response prediction in diseases, particularly CML. Aberrant miRNA expression is linked to stem cell survival, cell renewal and sensitivity or resistance to TKI therapy, all of which contribute to disease progression (1, 13, 14). Additionally, miRNAs have been discovered to affect genes in signalling pathways involved in cell proliferation, apoptosis, leukemogenesis, and tumour suppression (15–17). Hence, the current study aims to identify and screen differentially expressed miRNAs in CML patients from previous literatures. The application of integrated bioinformatics analysis is essential in predicting miRNA target genes, gene ontology and pathways, and protein-protein interaction networks. Findings from this study will help researchers to better understand the role of miRNA in CML pathogenesis and treatment resistance.

METHODS

Search Strategy

A comprehensive search of information was done using PubMed, Scopus, EBSCOhost MEDLINE, and Science Direct to identify relevant research publications with an unlimited starting publication date until 1st April 2021. The Medical Subject Heading (MeSH) terms like chronic myeloid leukaemia, genes and microRNAs were used as the keywords in the title or abstract. The search strategy involved a combination (“AND”) of the following two sets of keywords (1): "chronic myelo* leukemia” OR CML OR “BCR*ABL*positive” and (2) mi*RNA. Synonyms for keywords were generated through MeSH terms from the Cochrane Library. Additional text terms were discovered by reviewing collected review articles. Additional references were discovered from the bibliographies of the retrieved studies.

Inclusion Criteria

Case-control and prospective observational studies with abstracts investigating the differentially expressed miRNAs on Philadelphia chromosome-positive chronic myeloid leukemia patients in chronic, accelerated, or blast phases were included. In addition, only clinical studies that have further in vitro experimental evidence on the target genes of the studied miRNAs in CML cells were included in this review. Due to limited resources, only manuscripts written in English were included.

Exclusion Criteria

Publications that did not have primary data, such as editorials, case reports, conference proceedings, and narrative review articles, were excluded. In silico, in vitro, and in vivo studies were excluded. The review focus on the outcome of the differentially expressed miRNAs in CML patients. Therefore, studies that involved responses toward tyrosine kinase inhibitors treatment or any other intervention studies on a new treatment for CML patients were excluded from consideration. These selection criteria were used to achieve the objective of this systematic review in determining the typical miRNA expression signature in CML patients, the miRNAs target genes, and related pathways that could potentially be involved in the pathogenesis of CML.

Screening of Articles for Eligibility

Articles retrieved from all resources were screened in three phases. All articles with titles that did not match the inclusion criteria were excluded, and duplicates were removed in the first phase. The abstracts of the remaining articles were screened, and any articles that did not meet the inclusion criteria were excluded in the second phase. Finally, the full texts of the remaining articles were read and assessed thoroughly. Systematic reviews, meta-analyses, and articles that did not meet the inclusion criteria were excluded in this third phase. All the authors were involved in the screening, selection, and data extraction phase. Any differences in opinions were resolved by discussion between
the authors. All data extraction was performed independently using a data collection form to standardize the data collection, and records on reasons for rejection were kept. Figure 1 shows the flow chart that summarizes the article selection process and the reasons for article exclusion.

Data Extraction and Study Quality
Data were extracted from the studies that fulfilled inclusion criteria. Data collected from these studies include (1) author name (2), study design (3), study objective (4), study population (5), type of sample used (6), method used in gene expression analysis and experiments performed to validate their targeted genes (7), results (upregulated and downregulated miRNAs, their validated targeted genes and downstream effected genes), and (8) conclusion. The extracted details are listed in Table 1.

The analysis details of each report were discussed thoroughly among the reviewers to assess the quality of each study. The authors focused on the reported list of miRNAs as well as their targeted genes. Bias was excluded by adhering to the inclusion criteria. Pairs of reviewers with adequate reliability worked independently to determine the validity of each study.

Prediction of miRNA Target Genes
The miRNAs listed were further analysed using four different bioinformatics tools that include 1) DIANA-microT web server v5.0 with MiTG scores being set at more than 0.95, 2) TargetScan release 7.2 with Cumulative weighted context++ score of more than -0.5, 3) miRDB with a target score more than 90, and 4) mirDIP v5.0.2.3 with score class set at “very high”. Genes that are predicted by more than two bioinformatics tools were selected for further analysis.

Gene Ontology and Pathway Enrichment Analysis
Two groups of analysis that include 1) a group of validated target genes and downstream genes of the miRNAs extracted from the studies and 2) a group of predicted target genes of the reported miRNAs was done. These two groups of genes were analyzed using Database for Annotation, Visualization, and Integrated Discovery (DAVID). DAVID was performed to determine the cluster of genes that displayed significant functional annotation enrichment related to CML’s pathogenesis. The contribution of genes in the pathway related to CML was based on the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway, the Biological Biochemical Image database (BBID), BIOCARTA pathway database, and Reactome.

Protein–Protein Interaction (PPI) Network
The targeted genes were further analyzed at the protein level using protein-protein interaction network functional enrichment analysis through STRING (Protein-Protein Interaction Network Functional Enrichment Analysis) (https://string-db.org/). Results from STRING were further analyzed using Cytoscape to visualize molecular interaction networks and integrating gene

---

**FIGURE 1** | PRISMA flow diagram for studies selection in this systematic review.
TABLE 1 | Summary of selected studies.

| Title (References) | Study design | Samples(Type) | Methodology | Findings |
|--------------------|--------------|---------------|-------------|----------|
| MIR-152-3p promotes the development of chronic myeloid leukemia by inhibiting p27 (18). | Case-control study | CML (n=40) Controls (n=40) | qPCR | Luciferase assay | miR-152-3p (upregulated) |
| MIR-362-5p promotes the malignancy of chronic myelocytic leukemia via downregulation of GADD45α (19) | Case-control study | CML (n=40) Controls (n=26) | qPCR | Luciferase assay | miR-362-5p (upregulated) |
| miR-140-5p induces cell apoptosis and decreases Warburg effect in chronic myeloid leukemia by targeting SIX1 (20) | Case-control study | CML (n=30) Controls (n=30) | qPCR | Luciferase assay | miR-140-5p (downregulated) |
| MiRNA-409-5p dysregulation promotes imatinib resistance and disease progression in children with chronic myeloid leukemia (21) | Case-control study | CML (n=42) Controls (n=40) | qPCR | Luciferase assay | miR-409-5p (downregulated) |
| MiR-570 inhibits cell proliferation and glucose metabolism by targeting IRS1 and IRS2 in human chronic myelogenous leukemia (22) | Cross sectional study | CML (n=15) Controls (n=15) | qPCR | Luciferase assay | miR-570-3p (downregulated) |
| MicroRNA-320a acts as a tumor suppressor by targeting BCR/ABL oncogene in chronic myeloid Leukemia (23) | Case-control study | CML (n=90) Controls (n=90) | qPCR | Luciferase assay | miR-320a (downregulated) |
| Restoration of miR-424 suppresses BCR-ABL activity and sensitizes OML cells to imatinib treatment (24) | Case-control study | CML (n=18) Controls (n=10) | qPCR | Luciferase assay | miR-424-5p (downregulated) |
| The malignancy suppression role of miR-23a by targeting the BCR/ABL oncogene in chronic myeloid leukemia (25) | Case-control study | CML (n=70) Controls (n=25) | qPCR | Luciferase assay | miR-23a-3p (downregulated) |
| Low Expression of miR-196b Enhances the Expression of BCR-ABL1 and HOXA9 Oncogenes in Chronic Myeloid Leukemogenesis (26) | Case-control study | CML (n=16) Controls (n=10) | qPCR | Luciferase assay | miR-196b (downregulated) |
| Decreased microRNA-30a levels are associated with enhanced ABL1 and BCR-ABL1 expression in chronic myeloid leukemia (27) | Case-control study | CML (n=16) Controls (n=10) | qPCR | Luciferase assay | miR-30a (downregulated) |
| miR-29b suppresses CML cell proliferation and induces apoptosis via regulation of BCR/ABL1 protein (28) | Case-control study | CML (n=5) Controls (n=3) | qPCR | Luciferase assay | miR-29b (downregulated) |
| Targeting of the signal transducer Smo links microRNA-326 to the oncogenic Hedgehog pathway in CML stem/progenitor cells (29) | Case-control study | CML (n=6) Controls (n=4) | qPCR | Luciferase assay | miR-326 (downregulated) |
| BCR-ABL mediated repression of miR-223 results in the activation of MEF2C and PTEN in chronic myeloid leukemia (30) | Cross sectional study | CML (n=35) Controls (n=26) | qPCR | Luciferase assay | miR-223 (downregulated) |
| Down-Regulation of hsa-miR-10a in Chronic Myeloid Leukemia CD34+ Cells Increases USF2-Mediated Cell Growth (31) | Case-control study | CML (n=6) Controls (n=6) | qPCR | Luciferase assay | miR-10a (downregulated) |

expression profiles to identify clusters of protein interaction that are highly related to the pathogenesis of CML. The gene interaction relationship was downloaded in the “TSV” format file and was imported into the Cytoscape software (http://www.cytoscape.org/) for further analysis and clustering. The Cytoscape MCODE plug-in was employed to perform module analysis of the target network and protein clustering. The module selection criteria were as follows: degree cutoff = 2, node score cutoff = 0.2, node density cutoff = 0.1, K-score = 2, and max depth = 100. The list of genes in the cluster was then analyzed again using DAVID for significantly enriched ontology terms.

RESULTS

A total of 806 potentially relevant titles were identified from the database search. EndNote X9 software by Clarivate Analytics (Philadelphia, USA) was used as the reference manager. Upon filtering the titles, 372 articles were identified as duplicates, and two other articles were withdrawn from publication. A total of 432 articles were retrieved for abstract reviewing. Upon screening titles and abstracts, 263 articles were removed, resulting in the selection of 159 potentially relevant articles for full-text review. However, three articles could not be retrieved as...
the full text are not in English. Then, 147 potentially relevant articles’ full text was thoroughly reviewed, and 130 articles were eliminated based on our inclusion and exclusion criteria. Finally, 17 articles were selected to be included in the present systematic review. All studies were original research articles published between the year 2008 to 2019. Homogeneity of the selected studies was ensured by adhering to the defined inclusion and exclusion criteria to prevent sampling bias. Notably, all the studies performed real-time polymerase chain reaction (qPCR) for miRNA expression analysis. A confirmatory method was done to validate the miRNA binding site on their selected genes. Sample sizes for each study varied from 8 to 180 samples for miRNA expression analysis. The characteristics of these studies are highlighted in Table 1.

Patient Recruitment and Sampling
Samples collections were described briefly in most of the study. Six studies collected peripheral blood samples, while eight studies collected bone marrow tissues (Table 1). The inclusion criteria listed in the study by Nie et al. (20) include diagnosis via bone marrow morphology, immunology, molecular biology, and cytogenetic result, with no chemotherapy treatment before the specimen collection (20). In most studies, samples were collected at diagnosis; thus, most patients were in the chronic phase during sample collection. However, the study by Babashah et al. (29) collected samples from CML patients in the blast crisis phase at diagnosis (29).

Effect of miRNA on Cells
The effect of miRNA on cells, including proliferation, apoptosis, cell cycle, migration, and invasion, were adequately reported in the studies. Among miR-409-5p, miR-424, miR-29b, miR-570, miR-320a, miR-23a, miR-196b, miR-30a, miR-326, miR-223, miR-10a that were downregulated in CML clinical samples, the low expression of miR-409-5p, miR-424, miR-29b miRNAs in CML cells were reported to be the cause of the increase in CML cell viability. The low expression of miR-570, miR-320a, miR-23a, miR-196b, miR-30a, miR-326, miR-223, miR-10a in CML cells contribute to high proliferation rate. On the other hand, overexpression of miR-152-3p, miR-362-5p were reported in CML samples and from the functional analysis done, these miRNAs contributed to the increase in proliferation rate. In the cell cycle analysis, the overexpression of miR-152-3p was reported to reduce the percentage of cells in G0/G1 phase when compared with G2SM phase. However, overexpression of miRNA-409-5p, miR-362-5p, miRNA-196b, miR-30a, miR-29b arrested cell cycle in G0/G1 phase and S phase (21). Thus, downregulation of miRNA-409-5p, miR-362-5p, miRNA-196b, miR-30a, miR-29b increase the cell cycle activities in CML cells.

From the selected studies, overexpression of miRNAs like miR-140-5p, miR-320a, miR-570 induced apoptosis, but overexpression of miRNAs like miR-362-5p reduced apoptosis. Thus, in CML cells, a high level of miR-362-5p and a low level of miR-140-5p, miR-320a, miR-570 contribute to low apoptosis activities. Overexpression of miR-320a was proved in the in vitro studies to inhibit CML cell migration and invasion, but overexpression of miR-362-5p increased CML cell migration and invasion. Therefore, in CML cells, upregulation of miR-362-5p and downregulation of miR-320a were proposed in cell migration and invasion pathways. Furthermore, synthetic overexpression of miR-570 suppresses glucose metabolism and reduced ATP generation in CML cells. Thus, in clinical samples, downregulation of miR-570 increases glucose metabolism and ATP generation, producing high available energy for cell growth (22).

Targeted Gene Validation
The targeted gene validation of all miRNAs was adequately reported in all studies by co-transfecting cell lines with the targeted gene 3’-UTR reporter vector and miRNA mimic. In all studies, miRNA mimics used in the luciferase assay reveal decreased luciferase activity in wild-type targeted gene 3’UTR, suggesting that each of the miRNAs could bind to their respected genes. Mutant-type targeted gene 3’UTR did not show significant changes in luciferase activities after miRNA mimic transfection in all studies, suggesting a specific target of the miRNAs.

Effect of miRNAs in the Expression of Downstream Genes
Some studies also reported on the downstream genes that are differently expressed related to the expression changes of their studied miRNA. Overexpression of miR-140-5p was reported to increase BAX protein expression indirectly but decreased the BCL2 protein expression via SIX1 in CML cells (20). Overexpression of miRNA-409-5p in CML cells indirectly leads to downregulation of NUP43, leading to downstream downregulation of PCNA, c-Myc and cyclin D1 protein (21). The expression of genes associated with glucose metabolism, namely PGC1α, PCK1 and ABCA1 proteins, were indirectly suppressed by miR-570 overexpression via IRS1 and IRS2 (22). Inhibition of miR-362-5p indirectly increased P38 and JNK activity in CML cells via GADD45A (19). MiR-320a was reported to regulate the phosphorylation of PI3K, AKT and NF-κ B via BCR-ABL (23). Expression of p-Crk1 and p-STAT5 was reduced in the presence of miR-424 through BCR-ABL (1). Overexpression of miR-23a resulted in lower expression of PI3K, Akt and MMP-9, which are the downstream target of BCR-ABL (25). Overexpression miR-326 indirectly downregulates SMO expression, leading to downregulation of Bcl2 expression in CML cells (29). Significant downregulation of the survival gene Bcl-XL was reported to be associated with down-regulation of MEF2C and PTBP2 due to overexpression of miR-223-3p (30). Overexpression of miR-29b led to a significant increase in BCR-ABL expression that upregulates p21 and p27 expression in CML cells (28).

Gene Ontology Analysis of the Downstream Genes
A total of 26 downstream genes that were affected by the miRNAs was extracted from all the studies. The functions and pathway enrichment of these genes were analyzed using DAVID (https://david.ncifcrf.gov/home.jsp). A p-value of <0.05 was used as a cut-off standard. The gene listed were categorized into three functional
categories of gene ontology that include biological process (BP), cellular component (CC) and molecular function (MF), as shown in Table 2. In the CC group, the downstream genes are enriched in the intracellular component of cells, including nucleus, cytosol, cytoplasm, nucleoplasm, and mitochondria. In the BP group, the downstream genes are enriched in the regulation of transcription, cell proliferation, apoptosis, and drug response. The downstream genes are enriched in the DNA binding, protein binding, and protein heterodimerization activities in the MF group. The complete list for gene ontology cluster is included in 'Data S1'.

Signaling Pathway Enrichment Analysis of the Downstream Genes

The miRNA targeted genes and downstream genes signalling pathway enrichment analysis were conducted using DAVID with integrated KEGG PATHWAY, BBID, BIOCARTA, and Reactome. Concerning CML pathogenesis, the genes are mainly enriched in pathways related to cancer (hsa05200), microRNAs in cancer pathway (hsa05206), Hepatitis B (hsa05161), PI3K-Akt signalling pathway (hsa04151) and many other pathways with some directly associated with apoptosis, proliferation, and cell cycle pathways as reported in Table 3. The complete list of pathways is included in 'Data S2'.

| Term               | Description                                      | Count | p-value  |
|--------------------|--------------------------------------------------|-------|---------|
| MF.GO:0005515      | Protein binding                                  | 20    | 2.18E-04|
| CC.GO:0005634      | Nucleus                                          | 14    | 0.002201|
| CC.GO:0005829      | Cytosol                                          | 12    | 4.13E-04|
| CC.GO:0005737      | Cytoplasm                                        | 12    | 0.01868 |
| CC.GO:0005654      | Nucleoplasm                                      | 8     | 0.031381|
| BP.GO:0043524      | Negative regulation of neuron apoptotic process  | 6     | 5.12E-07|
| BP.GO:0042493      | Response to drug                                 | 6     | 3.02E-05|
| BP.GO:0006283      | Cell proliferation                               | 6     | 7.30E-05|
| MF.GO:0045665      | Sequence-specific DNA binding                    | 6     | 3.61E-04|
| BP.GO:0045944      | Positive regulation of transcription from RNA polymerase II promoter | 6 | 0.006252 |
| BP.GO:0006355      | Regulation of transcription, DNA-templated       | 6     | 0.034631|
| BP.GO:0043066      | Negative regulation of apoptotic process         | 5     | 0.002207|
| MF.GO:0046982      | Protein heterodimerization activity              | 5     | 0.002344|
| BP.GO:0008284      | Positive regulation of cell proliferation        | 5     | 0.002408|
| BP.GO:0006915      | Apoptotic process                                | 5     | 0.004870|
| BP.GO:000122       | Negative regulation of transcription from RNA polymerase II promoter | 5 | 0.011202 |
| MF.GO:0042802      | Identical protein binding                        | 5     | 0.012588|

Identification of Key Candidate Genes and Pathways in the Protein–Protein Interaction Network (PPI) and Modular Analysis of the Downstream Genes

Using STRING online database (http://string-db.org), a total of 26 proteins from miRNA targets and their downstream genes were...
filtered into a PPI network complex, containing 22 nodes and 75 edges (Figure 2) with a PPI enrichment p-value is 2.22e-16. At the same time, two other proteins did not fall into the PPI network. The results were transferred from STRING to Cytoscape for further analysis. Through Cytoscape MCODE, a significant module from the PPI network complex were found. Functional annotation clustering showed that this cluster (score = 8.909) consisted of 12 nodes and 49 edges (Figure 2). The cluster is mainly associated with protein binding (GO:0005515) as all the 12 proteins are involved in this molecular function. Ten of the proteins can be found in the cytosol (GO:0005829), and nine are involved in cancer pathways (hsa05200). Table 4 includes a functional annotation cluster with more than six proteins involved. The complete list for functional annotation cluster is included in ‘Data S3’.

### Prediction of miRNA’s Targeted Genes

Apart from the downstream genes reported from the articles, target genes of the miRNAs were also identified

![Figure 2](image-url)  
**FIGURE 2** | PPI network and modular analysis of downstream genes. From STRING online database analysis, a total of 26 proteins were filtered into a PPI network complex. The green nodes represent a functional annotation cluster that was identified from Cytoscape MCODE. This functional annotation clustering showed a cluster consisted of 12 proteins.

| Term                           | Description                                      | Count | p-value      |
|--------------------------------|--------------------------------------------------|-------|--------------|
| GO:0005515                     | protein binding                                  | 12    | 7.56E-04     |
| GO:0005829                     | cytosol                                          | 10    | 8.32E-06     |
| hsa05200                       | Pathways in cancer                               | 9     | 1.50E-08     |
| Up_Keywords: Phosphoprotein    | Phosphoprotein                                   | 9     | 0.029        |
| hsa05161                       | Hepatitis B                                      | 8     | 4.92E-10     |
| Up_Keywords: mutagenesis site  | mutagenesis site                                 | 8     | 4.06E-05     |
| GO:0005634                     | nucleus                                          | 8     | 0.020        |
| hsa05210                       | Colorectal cancer                                | 7     | 1.87E-10     |
| hsa04088                       | FoxO signaling pathway                           | 7     | 2.08E-08     |
| hsa04151                       | PI3K-Akt signaling pathway                       | 7     | 5.68E-06     |
| Up_Keywords: Nucleus           | Nucleus                                          | 7     | 0.037        |
| hsa05222                       | Small cell lung cancer                           | 6     | 1.12E-07     |
| hsa04722                       | Neurotrophin signaling pathway                   | 6     | 6.31E-07     |
| h_l2rbPathway                  | IL-2 Receptor Beta Chain in T cell Activation    | 6     | 1.42E-06     |
| hsa04932                       | Non-alcoholic fatty liver disease (NAFLD)        | 6     | 1.98E-06     |
| GO:0043066                     | negative regulation of apoptotic process         | 6     | 5.77E-06     |
| hsa05166                       | HTLV-I infection                                 | 6     | 2.54E-05     |
| hsa05206                       | MicroRNAs in cancer                              | 6     | 4.51E-05     |
| GO:0005739                     | mitochondrion                                    | 6     | 6.56E-04     |

Table 4 | Functional annotation clustering on the cluster identified from downstream genes.
Gene Ontology Analysis of the Predicted Target Genes

The functions and pathway enrichment of predicted target genes were analysed using DAVID (https://david.ncifcrf.gov/home.jsp). A p-value of <0.05 was used as a cut-off standard. The genes listed were categorised into three functional categories of gene ontology that include biological process (BP), cellular component (CC) and molecular function (MF), as shown in Table 5. In the CC group, the predicted target genes are enriched in the nucleus, proteinaceous extracellular matrix and perinuclear region of the cytoplasm. In the BP group, the genes are enriched in the regulation of transcription, extracellular matrix organisation and angiogenesis. In the MF group, the predicted target genes are enriched in the DNA binding. The list of predicted genes' GO is included in Data S5.

Signaling Pathway Enrichment Analysis of the Predicted Target Genes

The predicted target genes signalling pathway enrichment analysis were conducted using DAVID with integrated KEGG PATHWAY, BBID, BIOCARTA, and Reactome. The genes were found to be mainly enriched in the PI3K-Akt signalling pathway (hsa04151), focal adhesion (hsa04510), pathways in cancer (hsa05200), and many other pathways, with each pathway, involve from 13 to six predicted target genes (Table 6). The complete list for predicted pathways is included in Data S6.

Identification of Key Candidate Genes and Pathways in the Protein–Protein Interaction Network (PPI) and Modular Analysis of the Predicted Target Genes

Using STRING online database (http://string-db.org), a total of 187 proteins from predicted target genes were filtered into a PPI network complex, containing 116 nodes and 211 edges (Figure 3) with PPI enrichment p-value is less than 1.0E-16. At the same time, 51 other proteins did not fall into the PPI network.

The results were transferred from STRING to Cytoscape for further analysis. Through Cytoscape MCODE, six significant modules from the PPI network complex were found. Functional annotation clustering showed that cluster 1 (score = 9) consisted of 11 nodes and 45 edges (Figure 3). Cluster 1 is mainly located in the extracellular region and associated with extracellular matrix organisation and the collagen catabolic process. Cluster 2 (score= 5) consisted of five nodes and ten edges (Figure 3) associated with homeobox, sequence-specific DNA binding and

| Term          | Description                                                                 | Count | p-value       |
|---------------|-----------------------------------------------------------------------------|-------|---------------|
| MF_GO:0005515 | protein binding                                                             | 106   | 0.010477      |
| CC_GO:0005634 | nucleus                                                                     | 73    | 0.001961      |
| BP_GO:0006351 | transcription, DNA-templated                                                | 30    | 0.019201      |
| MF_GO:0003677 | DNA binding                                                                 | 26    | 0.033161      |
| BP_GO:0004594 | positive regulation of transcription from RNA polymerase II promoter        | 24    | 1.21E-04      |
| BP_GO:000122  | negative regulation of transcription from RNA polymerase II promoter        | 18    | 9.06E-04      |
| MF_GO:0002700 | transcription factor activity, sequence-specific DNA binding                | 17    | 0.036439      |
| MF_GO:0004566 | sequence-specific DNA binding                                               | 13    | 0.006885      |
| BP_GO:00030198| extracellular matrix organization                                          | 12    | 4.54E-06      |
| BP_GO:0001525 | angiogenesis                                                                | 12    | 1.54E-05      |
| CC_GO:0005578 | proteinaceous extracellular matrix                                          | 12    | 7.45E-05      |
| BP_GO:0006366 | transcription from RNA polymerase II promoter                               | 12    | 0.014271      |
| CC_GO:0004871 | perinuclear region of cytoplasm                                            | 12    | 0.04456       |
| BP_GO:00030574| collagen catabolic process                                                  | 10    | 1.39E-08      |
| CC_GO:0005788 | endoplasmic reticulum lumen                                                | 10    | 1.26E-04      |
| MF_GO:0000978 | RNA polymerase II core promoter proximal region sequence-specific DNA binding| 10    | 0.010699      |
| BP_GO:0006283 | cell proliferation                                                          | 10    | 0.01151       |
| BP_GO:00043066| negative regulation of apoptotic process                                    | 10    | 0.029644      |
transcription regulation. Cluster 3 (score = 3.333) consisted of four nodes and five edges (Figure 3) associated with the FoxO signalling pathway, pathways in cancer and mutagenesis site. Cluster 4 (score = 3.33) consisted of seven nodes and ten edges (Figure 3) associated with polymorphism, nucleus and DNA methylation. Cluster 5 (score = 3) consisted of three nodes and five edges (Figure 3) associated with coiled-coil structure and sodium ion transport channel. Cluster 6 (2.667) consisted of seven nodes and eight edges (Figure 3) located in the cytoplasm and nucleus and associated with protein binding, RNA binding and phosphatidylinositol-mediated signalling. Table 7 includes functional annotation clustering for all six clusters. The list of genes ontology of cluster 1 until 6 are included in ’Data S7’.

**DISCUSSION**

Tyrosine kinase inhibitors that target BCR-ABL protein have been successfully used to treat CML. However, there are still many cases that does not response well to the treatment. In those cases, it is postulated that their CML pathogenesis does not only involves BCR-ABL oncogene, it involves other mechanisms. For many decades, numerous molecular and clinical studies involving chromosomal changes, DNA mutation, DNA methylation and mRNA expression have been done to understand the underlying mechanism of CML development and progression. Nevertheless, the complete mechanisms of CML remain unclear. Thus, since many years ago, miRNA has come into the picture and has been studied extensively since then. In this systematic review, we attempted to improve our understanding of the involvement of miRNAs in CML development from fourteen different reports. The studies’ dataset was divided into two datasets to make sure high coverage of target genes. Figure 4 shows the summary of the bioinformatical analysis findings. The analysis of the first dataset started with pooling of the miRNA and their validated downstream genes, followed by gene’s ontology analysis, signalling pathway enrichment analysis and finally, the
protein-protein interaction network and modular analysis. One cluster was revealed from the first dataset. However, a second dataset was constructed and analysed to have a broader view of miRNA involvement in CML pathogenesis. The analysis of the second dataset involved the pooling of the miRNAs, followed by prediction of their target genes, gene’s ontology analysis, signalling pathway enrichment analysis and finally, the protein-protein interaction network and modular analysis. Six significant clusters were revealed from the second dataset.

In this review, through integrated bioinformatical analysis, a cluster consisted of 12 nodes and 49 edges has been identified from the first dataset consisting of the reported target genes and downstream genes. All the 12 clustered proteins are enriched in the intracellular signalling through phosphorylation, and are involved in cancer pathogenesis. Seven of these proteins (MAPK8, CCND1, IRS1, MYC, BCL2, BAX, AKT1) are enriched in the cytosol and nucleus. Three proteins (PIK3CA, IRS2, BCL2L1) are enriched in the cytosol, and one protein is in the nucleus (GADD45A). MAPK8, CCND1, IRS1, GADD45A, MYC, BCL2, AKT1, IRS2, and BCL2L1 are phosphoproteins. Phosphoproteins are proteins that bind to the phosphate group that can be switched on or off. These proteins are essential in most cellular processes such as protein synthesis, cell division, signal transduction, cell growth, development, and ageing (32).

From the KEGG pathway, MAPK8, CCND1, PIK3CA, MYC, BCL2, BAX, AKT1, MMP9, and BCL2L1 are essential genes in cancer pathways that regulate cell cycle, proliferation, apoptosis, genomic instability, and block of differentiation. At the same time, MAPK8, CCND1, PIK3CA, IRS1, GADD45A, AKT1 and IRS2 are involved in FoxO signalling pathway that can affect the regulation of cell cycle, oxidative stress resistance and DNA repair.

The first dataset of those experimentally validated target genes shows a significant effect of miRNAs in CML pathogenesis. However, it is known that miRNA can target more than one target gene due to its seed sequence (33). Therefore, it is crucial to identify the other miRNA’s target genes and their pathways involved. This review identified the other miRNA’s target genes using four different in silico analyses. All the predicted target genes were then analysed using bioinformatical analysis. Six clusters were identified from the functional annotation clustering analysis of 187 proteins.

### TABLE 7 | Functional annotation clustering of cluster 1 to 6.

| Cluster | Term | Description | Count | p-value |
|---------|------|-------------|-------|---------|
| 1       | Up_Keywords: Extracellular matrix | Extracellular matrix | 11 | 8.05E-20 |
|         | CC.GO:0005576 | Extracellular region | 11 | 2.82E-11 |
|         | Up_Keywords: Secreted | Secreted | 11 | 6.17E-11 |
|         | Up_Keywords: Disulfide bond | Disulfide bond | 11 | 1.65E-08 |
|         | Up_Keywords: Signal | Signal | 11 | 1.13E-07 |
|         | Up_Keywords: Polymorphism | Polymorphism | 11 | 0.004699 |
| 2       | Up_Keywords: MF.GO:0043565 | Sequence-specific DNA binding | 5 | 8.77E-07 |
|         | Up_Keywords: DNA-binding | DNA-binding | 5 | 9.82E-05 |
|         | Up_Keywords: Transcription regulation | Transcription regulation | 5 | 1.64E-04 |
|         | Up_Keywords: Transcription | Transcription | 5 | 1.84E-04 |
|         | Up_Keywords: Nucleus | Nucleus | 5 | 0.004211 |
|         | CC.GO:0005634 | Nucleus | 5 | 0.007789 |
| 3       | hsa04068 | FoxO signaling pathway | 3 | 0.001116 |
|         | hsa06500 | Pathways in cancer | 3 | 0.006998 |
| 4       | Up_Keywords: Polymorphism | Polymorphism | 7 | 0.040122 |
|         | Up_Keywords: Nuclear | Nuclear | 6 | 0.005069 |
|         | GO:0005634 | nucleus | 6 | 0.010445 |
| 5       | Up_Keywords: Coiled coil | Coiled coil | 3 | 0.021754 |
|         | GO:0005248 | voltage-gated sodium channel activity | 2 | 0.001185 |
|         | GO:0001518 | voltage-gated sodium channel complex | 2 | 0.001536 |
| 6       | MF.GO:0005515 | Protein binding | 7 | 0.019848 |
|         | CC.GO:0005737 | cytoplasm | 6 | 0.008813 |
|         | CC.GO:0005634 | nucleus | 6 | 0.010445 |

![FIGURE 4](https://example.com/figure4.png) | Summary of the bioinformatical analysis findings.
Cluster 1 from the second dataset consisted of 11 proteins, including COL3A1, ADAMTS2, SPARC, COL1A2, COL1A1, COL4A4, COL5A3, COL7A1, COL5A2, COL9A1, and COL19A1. All these proteins are in the extracellular matrix organization. COL3A1, COL1A2, COL1A1, COL4A4, COL5A3, and COL5A2 are involved in ECM-receptor interaction pathways and focal adhesion pathways. This collagen genes family is essential as the component of tissues structure and can interact with cells via several receptor families and regulate cell’s proliferation, differentiation, and migration (34). Other than that, SPARC, another gene in cluster 1, is significantly downregulated in CML patients. In CML cells exposed to exogenous SPARC, the G0/G1 cell cycle arrest and reduced growth rate of the cells were reported (35). However, the mechanism involved in SPARC downregulation in CML is still unknown. Thus, in CML, inhibition of SPARC by miR-29b-3p could be one of the CML pathogenesis mechanisms and is worth investigating.

Cluster 2 consisted of five proteins, including PBX3, HOXA7, HOXB7, HOXB6, and HOXA5, enriched in the nucleus. These proteins have sequence-specific DNA binding and are essential in transcription regulation. Interestingly, PBX3 is an important co-factor for the HOXA gene family (36). The HOX family is a group of highly conserved genes in mammals and are crucial in regulating cell differentiation and proliferation (36). HOXA5 impairs myelopoiesis, causing blockage of hematopoietic stem cells differentiation. The downregulation of HOXA5 is commonly related to DNA methylation (37). Apart from DNA methylation, the current review also found that miR-196b-5p could also regulate HOXA5 expression in CML cells.

Cluster 3 consisted of FOXG1, AKT3, VEGFA dan FOXO1. From KEGG pathway analysis, FOXG1, AKT3, and FOXO1 are enriched in the FoxO signalling pathway, while AKT3, FOXO1, and VEGFA are important in the cancer pathway having essential roles in apoptosis, proliferation, and angiogenesis. FOXO1 is a Forkhead box O (FoxO) family member. It plays a role in the regulation of differentiation and metabolism in tissues and organs. In CML, FOXO1 could increase the activation of CML cells (38). Thus, it will be an excellent move to inhibit FOXO1 in deactivating CML cells. In this review, it is suggested that miR-196b-5p could also regulate HOXA5 expression in CML cells.

Cluster 4 consisted of seven proteins, including WEE1, CCNE2, FBXW7, TDG, TET3, TET2, and TET1. Six proteins (WEE1, CCNE2, TDG, TET3, TET2, and TET1) are located in the nucleus, while four (TDG, TET3, TET2, TET1) are involved in DNA methylation. TET oncogene family that includes TET1, TET2, and TET3 plays a role in the DNA methylation process (39). Although there are no reports on the role of TET genes in CML pathogenesis, TET3 has been identified as a prognostic biomarker for acute myeloid leukemia (AML) (40), suggesting its involvement in myeloproliferative pathogenesis. Furthermore, the WEE1 gene in the cluster has been linked to the cell cycle and identified as a critical mediator of cell fate in AML (41). Meanwhile, high WEE1 kinase expression in acute lymphoblastic leukemia (ALL) has been identified as a poor prognostic factor that functions as a cancer-conserving oncogene which helps protect cancer cells from DNA damage (42). As a result of WEE1’s participation in the cell cycle and DNA damage repair, the WEE1 kinase family was identified as one of the most promising targets in the DNA damage response (DDR) pathway (43).

Cluster 5 consisted of three proteins, including SCN9A, NAV1, and SCN2A. These proteins are involved in the voltage-gated sodium channel complex. Although there are still no CML studies on these genes, ion channel signalling mechanisms are known to be involved in cancer cells migration, invasion, and metastasis (44). Thus, it will be very informative to study the effect of ion channel signalling through SCN9A and SCN2A via miR-301-5p in CML cells.

Cluster 6 consisted of seven protein-binding proteins, including KITLG, SOCS1, AGO4, TRA2B, PTEN, SRSF7, and QKI. KITLG is localised in the cytoplasm, TRA2B is in the nucleus, while SOCS1, AGO4, PTEN, SRSF7, and QKI can be found in both intracellular regions. KITLG, AGO4, PTEN are involved in phosphatidylinositols-mediated signaling that is crucial in regulating cancer cells’ survival, proliferation, invasion, and growth. SOCS1 plays an important role in regulating optimal JAK/STAT activity. However, regulation of SOCS1 via DNA methylation in CML patients is still uncertain as the findings are contradictory (45, 46). Thus, from the analysis, regulation of SOCS1 via miR-30a-5p in CML is suggested.

The studies included in this review only focused on their miRNA of interest and its few target genes. Therefore, this review are not able to rule out the entire networks of miRNAs and their target genes in CML. In silico analyses were done to improve the coverage of miRNA’s target genes. However, further in vitro analysis and clinical studies need to be done to validate the predicted mechanisms. Nevertheless, this review added new insight into the involvement of miRNA in CML pathogenesis for future studies.

CONCLUSION

Pathogenesis of CML at the molecular level involves a wide range of mechanisms that are still undiscovered. In this study, the function of miRNAs was found to be significant in the development of CML. The miRNA’s target genes are localised in the extracellular, cytosol and nucleus of CML cells. Thus, the importance of miRNAs cannot be denied as miRNAs are universally involved in various pathways that regulate genomic instability, proliferation, apoptosis, cell cycle, differentiation, and migration of CML cells. Therefore, from the identified miRNAs and their pathways involved in CML pathogenesis, potential new biomarkers for a better prognosis and new miRNA-based treatment for CML patients could be developed.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.
AUTHOR CONTRIBUTIONS

ME collected and/or assembled data and wrote the manuscript. SS wrote, reviewed, and edited the manuscript. NA confirm the authenticity of all raw data, proofread, edited and gave approval of the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This study was funded by the Fundamental Research Grant Scheme, awarded by Ministry of Higher Education of Malaysia, grant number FRGS/1/2018/SKK08/USIM/03/1.

REFERENCES

1. Moradi F, Babashah S, Sadeghizadeh M, Jalili A, Hajifathali A, Rosshandel H. Signaling Pathways Involved in Chronic Myeloid Leukemia Pathogenesis: The Importance of Targeting Musashi2-Numb Signaling to Eradicate Leukemia Stem Cells. *Iran J Basic Med Sci* (2019) 22(6):581–9.
2. Cilloni D, Saglio G. Molecular Pathways: BCR-ABL. *Clin Cancer Res* (2012) 18(4):930–7.
3. Machova Polakova K, Lopotova T, Klamova H, Burda P, Trneny M, Stopka T, et al. Expression Patterns of microRNAs Associated With CML Phases and Their Disease Related Targets. *Mol Cancer* (2011) 10:41.
4. Zhu X, Zhao H, Lin Z, Zhang G. Functional Studies of miR-130a on the Inhibitory Pathways of Apoptosis in Patients With Chronic Myeloid Leukemia. *Cancer Gene Ther* (2015) 22(12):573–80.
5. Hershkovitz-Rokah O, Modai S, Pasmanik-Chor M, Toren A, Shomron N, Raanani P, et al. Restoration of miR-424 Suppresses BCR-ABL Activity and Sensitizes CML Cells to Imatinib Treatment. *Cancer Lett* (2015) 360(2):245–56.
6. Huang T, Fu Y, Wang S, Xu M, Yin X, Zhou M, et al. Inhibition of BCR-ABL1 Oncogene in Chronic Myeloid Leukemia Blastic Transformation. *BioMed Pharmacother* (2019) 119:109413.
7. Li Y, Yuan Y, Tao K, Wang X, Xiao Q, Huang Z, et al. Inhibition of BCR/ABL Protein Expression by miR-203 Sensitizes for Imatinib Mesylate. *PloS One* (2013) 8(4):e61658.
8. Di Stefano C, Mirone G, Perna S, Marfe G. The Roles of microRNAs in the Pathogenesis and Drug Resistance of Chronic Myelogenous Leukemia (Review). *Oncol Rep* (2016) 35(2):614–24.
9. Bavaro L, Martelli M, Cavo M, Soverini S. Mechanisms of Disease Progression and Resistance to Tyrosine Kinase Inhibitor Therapy in Chronic Myeloid Leukemia: An Update. *Int J Mol Sci* (2019) 20(24).
10. Yurt M, Ayildiz O, Karakus A, Nursal AF. MicroRNAs Expression Profiles as Biomarkers and Therapeutic Tools in Turkish Patients With Chronic Myeloid Leukemia. *Bratisl Lek Listy* (2020) 122(12):159–63.
11. Martins JR, de Moraes LN, Curry SS, Dadalto J, Capanacci J, Carvalho RF, et al. Comparison of microRNA Expression Profile in Chronic Myeloid Leukemia Patients Newly Diagnosed and Treated by Allogeneic Hematopoietic Stem Cell Transplantation. *Front Oncol* (2020) 10:1544.
12. Klumper T, Bruckmueller H, Diewock T, Kaechler M, Haenisch S, Pott C, et al. Expression Differences of miR-142-5p Between Treatment-Naive Chronic Myeloid Leukemia Patients Responding and non-Responding to Imatinib Therapy Suggest a Link to Oncogenic ABL2, SRL, CKIT and MCL1 Signaling Pathways Critical for Development of Therapy Resistance. *Exp Hematol Oncol* (2020) 9:26.
13. Khalil NA, Desouky MN, Diab IH, Hamed NAM, Mannaa HF. MicroRNA 30a Mediated Autophagy and Imatinib Response in Egyptian Chronic Myeloid Leukemia Patients. *Indian J Hematol Blood Transfus* (2020) 36(3):491–7.
14. Zhang XT, Dong SH, Zhang JY, Shen B. MicroRNA-577 Promotes the Sensitivity of Chronic Myeloid Leukemia Cells to Imatinib by Targeting NUP160. *Eur Rev Med Pharmacol Sci* (2019) 23(16):7008–15.

ACKNOWLEDGMENTS

The authors would like to thank staffs and management of the Faculty of Medicine and Health Sciences Universiti Sains Islam Malaysia (USIM) for their help and contributions.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2022.848199/full#supplementary-material

Frontiers in Oncology | www.frontiersin.org 12 March 2022 | Volume 12 | Article 848199
Activation of MEF2C and PTBP2 in Chronic Myeloid Leukemia. *Leukemia* (2013) 27(7):1578–80.

31. Agirre X, Jiménez-Velasco A, San José-Ereniz E, Garate L, Bandrés E, Cordeu L, et al. Down-Regulation of hsa-miR-10a in Chronic Myeloid Leukemia CD34+ Cells Increases USF2-Mediated Cell Growth. *Mol Cancer Res MCR* (2008) 6 (12):1830–40.

32. Ardito F, Giuliani M, Perrone D, Troiano G, Lo Muzio L. The Crucial Role of Protein Phosphorylation in Cell Signaling and its Use as Targeted Therapy (Review). *Int J Mol Med* (2017) 40(2):271–80.

33. Kehl T, Backes C, Kern F, Fehlmann T, Ludwig N, Meese E, et al. About miRNAs, miRNA Seeds, Target Genes and Target Pathways. *Oncotarget* (2017) 8(63):107167–75.

34. Ricard-Blum S. The Collagen Family. *Cold Spring Harb Perspect Biol* (2011) 3(1):a004978.

35. Giallongo C, La Cava P, Tibullo D, Barbagallo I, Parrinello N, Cupri A, et al. SPARC Expression in CML is Associated to Imatinib Treatment and to Inhibition of Leukemia Cell Proliferation. *BMC Cancer* (2013) 13:60.

36. Li Z, Zhang Z, Li Y, Arnovitz S, Chen P, Huang H, et al. PBX3 is an Important Cofactor of HOXA9 in Leukemogenesis. *Blood* (2013) 121(8):1422–31.

37. Elias MH, Azlan H, Baba AA, Ankathil R. Aberrant DNA Methylation at HOXA4 and HOXA5 Genes is Not Associated With Resistance to Imatinib Mesylate Among Chronic Myeloid Leukemia Patients. *Cardiovasc Hematol Disord Drug Targets* (2018) 18(3):234–8.

38. Liu TC, Lin SF, Chang JG, Yang MY, Hung SY, Chang CS. Epigenetic Alteration of the SOCS1 Gene in Chronic Myeloid Leukaemia. *Br J Haematol* (2003) 123(4):654–61.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Elias, Syed Mohamad and Abdul Hamid. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.