Comparison of the choice of animals for re-sequencing in two maternal pig lines

Christina M. Dauben1, Christine Große-Brinkhaus1*, Esther M. Heuß1, Hubert Henne2 and Ernst Tholen1

Abstract

Next-generation sequencing is a promising approach for the detection of causal variants within previously identified quantitative trait loci. Because of the costs of re-sequencing experiments, this application is currently mainly restricted to subsets of animals from already genotyped populations. Imputation from a lower to a higher marker density could represent a useful complementary approach. An analysis of the literature shows that several strategies are available to select animals for re-sequencing. This study demonstrates an animal selection workflow under practical conditions. Our approach considers different data sources and limited resources such as budget and availability of sampling material. The workflow combines previously described approaches and makes use of genotype and pedigree information from a Landrace and Large White population. Genotypes were phased and haplotypes were accurately estimated with AlphaPhase. Then, AlphaSeqOpt was used to optimize selection of animals for re-sequencing, reflecting the existing diversity of haplotypes. AlphaSeqOpt and ENDOG were used to select individuals based on pedigree information and by taking into account key animals that represent the genetic diversity of the populations. After the best selection criteria were determined, a subset of 57 animals was selected for subsequent re-sequencing. In order to evaluate and assess the advantage of this procedure, imputation accuracy was assessed by setting a set of single nucleotide polymorphism (SNP) chip genotypes to missing. Accuracy values were compared to those of alternative selection scenarios and the results showed the clear benefits of a targeted selection within this practical-driven approach. Especially imputation of low-frequency markers benefits from the combined approach described here. Accuracy was increased by up to 12% compared to a randomized or exclusively haplotype-based selection of sequencing candidates.

Background

Next-generation sequencing (NGS) experiments provide data that are of increasing importance in animal genetics with the potential to improve knowledge of the genetic background of complex traits with societal and economical interest (e.g., [1]). In addition, human genetics can benefit from insights into the porcine genome due to physiological similarities [2].

Until now, many of the genetic analyses in pigs are based on single nucleotide polymorphism (SNP) genotype data from chip arrays (e.g., [3, 4]). However, as costs for re-sequencing decrease [5], NGS becomes increasingly important [6]. The ratio between genetic gain and costs needs to be considered [7], as costs are a central issue in pig breeding [8].

Available NGS data are used to enhance information content of routinely phenotyped and genotyped animals, especially in terms of imputation from a lower to a higher marker density. An efficient approach is to select a highly informative and representative subset of animals for re-sequencing and to use it as reference panel for imputation of a population that is genotyped at a lower marker density (e.g., [9]). Several studies have described how the final selection criteria were determined for selecting the animals to be resequenced, e.g., how frequently they...
are used for artificial insemination and the availability of sampling material or special phenotypes (e.g., [10–12]). Within the 1000 Bull Genomes Project, selection based on pedigree-based algorithms has been described [13, 14].

In a simulated data set, five approaches to select candidates for re-sequencing were discussed by Druet et al. [13]. Results were evaluated by considering minor allele frequency (MAF) and effects on imputation accuracy and genomic prediction [13].

NGS data and imputed whole-genome sequence data are assumed to include causative variants and to enable their detection (e.g., [1, 14, 15]). In this context, the composition of the resequenced subset is of high importance [15]. Verification, validation and fine-mapping of quantitative trait loci (QTL) derived from genome-wide association studies can benefit from imputed sequence data [14], although the relatively high level of linkage disequilibrium in pig breeding populations makes detection of causative variants more difficult. Combining statistical, bioinformatic, and functional information, such as genetic associations, linkage disequilibrium, annotation, and functional genomic data, might optimize such detection (e.g., [16–18]) and, in addition, will increase the number of QTL and the proportion of variance explained by these QTL, compared to the use of a lower marker density [19].

The aim of this study was to evaluate alternative strategies for the choice of animals for re-sequencing in the maternal Landrace (LR) and Large White (LW) pig lines. Selection was conducted by applying several theoretical concepts and approaches under practical conditions. Selection steps were built on existing pedigree and real SNP genotype information within the pigFit project which focuses on piglet survival and immunocompetence [20, 21].

Performance of the different selection strategies was assessed based on the accuracy of imputation of masked genotypes from SNP chip data. The resulting NGS data are expected to enable a highly accurate imputation within the LR and LW population and to discover the causative genetic variants for previously detected QTL regions.

**Methods**

**Animals and data sets**

The aim of the project ‘pigFit-Molecular genetic and immunological analysis of survival and postnatal growth of piglets’ is to identify biological relevant regions associated with survivability, health and immune traits in piglets and growing pigs. Within this project, phenotyping and genotyping were performed in the two maternal pig lines LR and LW. Blood and tissue samples were collected from piglets and their biological dams. SNP genotype data, based on the PorcineSNP60v2 BeadChip (Illumina Inc., San Diego, CA, USA), were used to conduct the animal selection process. In total, SNP genotype information for 944 LR and 800 LW animals that were genetically linked to the pigFit data set was available. Before starting the analysis, breed-specific quality control was performed for SNPs on autosomes. Markers showing a high linkage disequilibrium ($r^2 > 0.8$) within a region of 3 kb and markers with a low MAF (<1%) or a low call rate (<95%) were excluded. In addition, breed-specific pedigree information for 2871 LR animals and 1965 LW animals was available.

**Theoretical principles for selection of animals and implementation**

Druet et al. [13] have explained the impact of different strategies to choose individuals for sequencing on imputation accuracy and genomic prediction based on simulated sequence data. These approaches can be summarized as pedigree-based, haplotype-based, and random selection of individuals.

Based on the principles and results presented by Druet et al. [13], the current study applied a combined, heuristic approach (C) to select animals for re-sequencing. This approach combined haplotype- and pedigree-based methods according to their theoretical benefits, which are described below.

**Random selection**

Compared to targeted selection, random selection has been demonstrated to be a non-competitive approach [13]. In the current study, random selection of animals for sequencing was used for comparative purposes in validation steps.

**Haplotype-based (H) approach**

Haplotype-based selection aims at maximizing the diversity of haplotypes with or without weighting the haplotype frequency [13]. Our haplotype-based selection strategy made use of 60k SNP genotype information. SNP genotypes were processed using the software AlphaPhase [22] to construct haplotypes by phasing. Phasing was performed by breed and chromosome. Chromosomes were divided into fragments of 100 consecutive SNPs (GeneralCoreLength), as recommended for a 60k SNP density [22]. Hickey et al. [22] demonstrated that the highest percentage of alleles correctly phased were obtained for this setting. The average percentage of genotypes phased was >99%, indicating that phased genotypes can be used in subsequent analyses.

In general, the aim of the H approach is to cover the maximum proportion of haplotype diversity within the population. The results obtained with simulated data led
to the assumption that these animals will enable accurate imputation from chip genotype data to sequence data [13].

To carry out the selection step, we used the AlphaSeqOpt-Method 1 [23]. In a nutshell, haplotype libraries were constructed and animals with the most high-frequency haplotypes in the investigated population were identified as candidates for re-sequencing to enable exact and complete imputation [23]. Appropriate haplotypes were masked and 100-fold repetition was performed.

The settings for the AlphaSeqOpt software were modified according to the porcine genome structure with 18 autosomes and to the chip structure with a varying number of SNPs per chromosome. The call rate of SNPs and individuals had to exceed 90% to be included in the construction of the haplotype library. Information on the number of cores and the core length was adapted to the GeneralCoreLength used in the AlphaPhase software. Since the H approach aimed at covering a large amount of haplotypes, the threshold for haplotype frequency in the population was set to 0 to take all the haplotypes into account, regardless of their frequency. In both lines, the availability of sample material was a key limiting factor, particularly in previous generations. Thus, the target number of animals to select for re-sequencing was set to 100. Animals were added to the candidate set in an iterative manner. Although the AlphaSeqOpt software includes an additional approach, Method 2 [24], this method was not implemented because of uncertainties in defining the cost parameters due to technical innovations and because of known difficulties in the analysis of specific regions in the porcine genome.

**Pedigree-based (P) approaches**

One of the strategies discussed by Druet et al. [13] maximizes the expected genetic relationship between sequenced animals and the population based on pedigree data. For implementing the P approach in our study, the AlphaSeqOpt software (P1) [23] was used and the algorithm was as described by Goddard et al. [25]. Basic settings were comparable with those in the H approach but the parameter OptimisationMethod was set to Pedigree. A subset of 100 candidates from the genotyped animals was selected based on pedigree relationships [23, 25].

The ENDOG v.4.8 software [26] was used to determine the underlying population structure and to characterize each population. Unlike all other approaches, this software makes use of the entire pedigree of the pigFit population and available relatives, covering 17 generations. Ancestors (P2) were identified as the animals that best explained the genetic variability of both populations considering their unbalanced use in breeding [26, 27].

**Selection of animals for re-sequencing by the combined (C) approach**

As the starting point for the C approach, all animals from the H, P1, and P2 approaches were taken into account to select animals for re-sequencing. According to the available budget, the target number of animals was set to 57, divided into 28 LR and 29 LW animals. The C approach is practical-driven and aims at optimizing imputation accuracy of common and low-frequency variants by combining haplotype- and pedigree-based criteria. The findings shown by Druet et al. [13] provided an indication for the prioritization between the three approaches. As a consequence, we defined steps 1 to 4 for the final selection of candidates for re-sequencing. These steps were conducted successively and included the availability of sample material as a requirement.

- **Step 1**: Animals that were selected by the H approach and at least by one of the P approaches (P1, P2)
- **Step 2**: Animals that were selected in both P approaches (P1, P2)
- **Step 3**: Animals that were selected by the H approach and both parents, sire and dam, were identified as important ancestors (P2)
- **Step 4**: Animals that were selected by the H approach and one parent was identified as an important ancestor (P2)

**Validation**

Evaluation of alternative selection strategies was performed based on the 60k SNP genotype data in a 0, 1, 2 coding as obtained from the PorcineSNP60v2 BeadChip (Illumina Inc., San Diego, CA, USA). The workflow for the validation is shown in Fig. 1. Breed-specific data sets were split into a reference panel with high marker density (HD) and a target panel with low marker density (LD). The reference panel with high marker density included 28 LR animals and 29 LW animals. Three sets of candidates for re-sequencing were analyzed. The set comprising candidates selected by the C approach (combined sample) was compared to the set of the top candidates from the H approach (haplotype sample) and a set of randomly selected animals in a 30-fold repetition (random sample). Genotype data in the reference panel contained information on all markers that passed quality control (LR $n_{\text{SNPs}} = 43,325$, LW $n_{\text{SNPs}} = 43,248$).

The remaining animals (LR $n=777$, LW $n=668$) were allocated to the target panel. The assessed imputation scenarios are shown in Fig. 1. LD genotype data was simulated by setting 10,000 SNPs/ 23.1% (LD1) or half of the number of SNPs (LD2) to missing. Imputation was performed from LD to HD for each of the reference panels.
combined sample, haplotype sample, and random sample, using FImpute v3 [28]. Imputation accuracy was calculated as the correlation (r) between true and imputed genotypes. Mean accuracies were averaged over a 20-fold repetition of imputation.

**Results and discussion**

**Haplotype-based (H) approach**

The H approach resulted in 100 candidates per breed that maximized the diversity of haplotypes across the population. As shown in Fig. 2, in LR, 60% of the candidates belonged to the most recent generation. The remaining 40% were distributed among the three previous generations, with decreasing proportions. In LW, the proportion of animals in the most recent generation was slightly higher (66%). The remaining candidates had a distribution similar to that for LR.

**Pedigree-based (P) approaches**

The P1 and H approaches are based on the same animals originating from the most recent generations. Within the
P1 approach, generations 3 and 4 (relative to the most recent generation being generation 1) comprised about two thirds of the 100 candidates in each breed (see Fig. 2).

The P2 approach included 17 generations of pedigree for both populations. In total, 148 LR and 117 LW animals were selected based on pedigree relationships. Selected LR animals were from 13 of the 17 generations, with the largest number from generations 6 to 8 (see Fig. 2), which were not selected by the H and P1 approaches. LW candidates were more equally distributed in generations 6 and older.

Concordance between the two P approaches was observed for four LR animals and two LW animals (see Fig. 3). Strongly diverging results between the P1 and P2 approaches were expected due to the differences in databases and purposes of the methods. The P1 approach selected animals by focusing on the pedigree relationships in genotyped animals of recent generations, while the P2 approach identified animals that explained the genetic variability of both populations, based on pedigree information going back 17 generations.

**Subset of animals for re-sequencing selected by the combined (C) approach**

Figure 3 provides the number of selected animals from the H, P1, and P2 approaches. These results were condensed to the target number of 57 animals within our C approach with respect to optimizing imputation.

Selection steps 1 and 2 (see “Methods” section) are the overlaps of two approaches. Step 1 (H ∩ P1 or H ∩ P2) is based on both, haplotype- and pedigree-based approaches, whereas step 2 (P1 ∩ P2) is solely based on pedigree-based criteria. In total, steps 1 and 2 identified nine and four LR animals as well as nine and two LW animals for re-sequencing, respectively. In order to complete the target number, steps 3 and 4 were implemented. The resulting animals fulfilled the haplotype-based criteria and were progeny of animals which were selected by one of the pedigree-based approaches. In comparison to steps 1 and 2, the importance of the pedigree-based criteria is downgraded in steps 3 and 4, but still effective in the representation of the diversity within the LR and LW populations. The resulting subset of animals is hereafter denoted as combined sample.

The combined sample included 28 LR and 29 LW animals, divided into 30 males and 27 females. These animals were born between 2012 and 2016 and can be assigned to six generations (see Table 1).

**Validation**

Imputation accuracy was calculated to evaluate and quantify the performance of the selected animals for re-sequencing in a practical data set based on 60k SNP genotypes. The detailed results are in Table 2 with MAF grouped in the classes 1 to 3%, 3 to 5% and >5%. In summary, imputation accuracy was high using the combined sample as the reference panel. The mean accuracy ranged from 83.6 to 93.1% in LR and from 86.2 to 93.9% in LW across both imputation scenarios (LD1, LD2) and all MAF classes (1–3%, 3–5%, >5%). Imputation using the random sample (LR 74.1–87.8%, LW 78.7–89.9%) and haplotype sample (LR 81.2–94.1%, LW 83.4–94.0%) as reference panel had a lower mean accuracy.

Assessment of the imputation accuracy was performed by imputing masked genotypes from the PorcineSNP60v2 BeadChip (Illumina Inc., San Diego, CA, USA) in a 20-fold repetition and reflect the accuracy of imputation to a HD density in these populations. However, this study cannot make a conclusive statement regarding the accuracy for imputation to sequence data.

---

**Table 1** Distribution among breeds and generations of animals selected for re-sequencing using the combined (C) approach

| Generation | 1 | 2 | 3 | 4 | 5 | 6 |
|------------|---|---|---|---|---|---|
| Landrace   | 8 | 8 | 7 | 4 | 1 | 0 |
| Large White| 8 | 12| 5 | 2 | 1 | 1 |

Generation 1: most recent generation of animals
Overall, imputation of 10,000 masked SNPs (LD₁) led to higher accuracies compared to imputation of 50% of the SNP set (LD₂). Exceptions were found for the random sample in LR and the MAF class >5% in the combined sample in LR.

The strategy for selecting animals is very important, especially to enable an accurate imputation of low-frequency variants [13]. As expected from previous studies (e.g., [13, 29]), imputation tended to be more accurate as the MAF of markers increased. Although imputation of low-frequency markers is less accurate (e.g., [13, 29]), imputation using the combined sample as the reference panel reached the highest increase in accuracy for low-frequency markers, compared to the alternative reference panels (haplotype sample, random sample). This may be due to the prioritization of haplotype diversity in the selection process for the combined sample and to the imputation to 60k level. However, in terms of low-frequency variants, imputation to the sequence level remains a major challenge, especially with a small number of sequenced animals.

In addition, our study observed differences in imputation accuracy between LR and LW. For low-frequency (1–3%) and higher-frequency markers (>5%), imputation performance was better in LW. In addition to breed-specific genetic aspects, one possible explanation is the larger number of animals within the reference panel for LW. It is important to underline that both nucleus populations are genetically distinct and differ in their breeding objectives to a considerable extent. Moreover, genotyped animals were assumed to be representative samples of both nucleus populations.

Compared to random selection, targeted selection approaches achieved up to 12.4% higher imputation accuracy using the combined sample as reference panel for LR (see Fig. 4). The highest increase in imputation accuracy compared to random selection was reached with markers that had a MAF of 1 to 3%. The advantage of the combined sample compared to the random sample was more evident for LR than for LW except for markers with a MAF >5% in LD₁ and a MAF of 3 to 5% in LD₂. The gain in accuracy through targeted selection was mainly achieved when imputing a larger number of markers (LD₂).

In general, imputation accuracy varied a lot between replicates of randomly selected reference panels. This emphasizes the importance of the structure of reference panels for re-sequencing experiments, especially when resources are limited [15]. Our results are consistent with those of Druet et al. [13], who showed the advantages of haplotype-based selection over randomized selection in a simulated data set.

Assuming the availability of sample material from all prioritized candidates, the H approach was tested as an alternative approach, by considering haplotype-based criteria, only. It should be noted that the C approach is a heuristic, practical-driven approach and its results differ slightly from those using exclusively the H approach. For markers with a higher MAF (>5%), we observed no improvement in imputation accuracy of the C approach compared to the H approach. The latter achieved an even

| Breed | Reference panel | Imputation scenario | MAF 1–3% | MAF 3–5% | MAF >5% |
|-------|----------------|-------------------|-----------|-----------|---------|
|       |                |                    | Min.      | Max.      | Min.    |
| LR    | com            | LD₁               | 86.53%    | 83.97%    | 88.95%  |
|       |                | LD₂               | 83.63%    | 82.26%    | 85.23%  |
|       | ran            | LD₁               | 74.13%    | 63.52%    | 84.26%  |
|       |                | LD₂               | 75.35%    | 70.80%    | 80.38%  |
|       | hap            | LD₁               | 82.81%    | 80.38%    | 85.43%  |
|       |                | LD₂               | 81.21%    | 78.81%    | 83.31%  |
| LW    | com            | LD₁               | 92.68%    | 92.47%    | 92.85%  |
|       |                | LD₂               | 86.52%    | 85.22%    | 88.23%  |
|       | ran            | LD₁               | 83.30%    | 76.37%    | 88.20%  |
|       |                | LD₂               | 79.22%    | 73.35%    | 83.88%  |
|       | hap            | LD₁               | 84.59%    | 81.45%    | 87.05%  |
|       |                | LD₂               | 83.37%    | 81.34%    | 85.35%  |

| Breed | Reference panel | Imputation scenario | MAF 1–3% | MAF 3–5% | MAF >5% |
|-------|----------------|-------------------|-----------|-----------|---------|
|       |                |                    | Min.      | Max.      | Min.    |
| LR    | com            | LD₁               | 86.30%    | 84.51%    | 87.73%  |
|       |                | LD₂               | 83.60%    | 82.00%    | 85.77%  |
|       | ran            | LD₁               | 75.86%    | 66.82%    | 81.95%  |
|       |                | LD₂               | 77.65%    | 73.68%    | 81.18%  |
|       | hap            | LD₁               | 84.64%    | 81.24%    | 86.83%  |
|       |                | LD₂               | 82.14%    | 80.74%    | 84.30%  |
| LW    | com            | LD₁               | 88.68%    | 86.37%    | 89.97%  |
|       |                | LD₂               | 86.20%    | 84.93%    | 87.47%  |
|       | ran            | LD₁               | 82.24%    | 76.15%    | 86.73%  |
|       |                | LD₂               | 78.68%    | 74.33%    | 83.88%  |
|       | hap            | LD₁               | 87.36%    | 85.79%    | 89.46%  |
|       |                | LD₂               | 85.46%    | 84.05%    | 86.54%  |

Table 2  Imputation accuracy (r) of masked chip genotype data from lower (LD₁, LD₂) to higher marker density in Landrace (LR) and Large White (LW) using different reference panels

Imputation accuracy (r): correlation between true and imputed genotypes, com: combined sample, ran: random sample, hap: haplotype sample, LD₁: 10,000 SNPs (23.1%) were set to missing, LD₂: 50% of the SNPs were set to missing, MAF: Minor allele frequency
slightly higher accuracy for LD1 in LR. Nevertheless, an increase of about 2.4 to 8.1% was found with low-frequency markers of the C over the H approach.

Targeted selection using the C approach with haplotype- and pedigree-based selection criteria, as described in this paper, shows that it is superior compared to alternative approaches to select animals for a re-sequencing experiment using data from the LR and LW populations investigated here.

The respective characteristics of the LR and LW populations are common for commercial maternal nucleus breeding populations. Moreover, we had to consider the practical constraints (data and sample availability, and budget, which lead to a trade-off between the number of animals and coverage) within our C approach. Against this background, our practical-driven approach and its results could be considered for similar situations in pig breeding using the data of nucleus herds.

Data from re-sequencing the combined sample will be used to supplement the pigFit project in future investigations. Making use of highly informative genotype data, genome-wide association studies (GWAS) that use highly informative genotypes are a highly efficient approach to map loci and to detect QTL and candidate genes for
complex traits (e.g., [1]). This approach enables fine mapping [19] and validation of previously identified regions in a GWAS for important traits (e.g., [21]).

Conclusions

Targeted animal selection for re-sequencing is recommended. Taking into account all available information on population structure, when selecting animals for re-sequencing, including pedigree information and additional SNP genotype information, increased the imputation accuracy of masked SNP chip genotypes, measured as the correlation between true and imputed genotypes, by an average of around 7.3 to 12.4% for low-frequency markers and up to 5.3% for markers with a MAF >5%. The combined use of pedigree- and genotype-based approaches in practical data sets is particularly suitable for improving imputation accuracies for low-frequency markers.

Acknowledgements

The authors thank the Bundeshybridzuchtprogramm (BH2P GmbH) for providing data sets and the staff members of the BH2P GmbH, especially Anne K. Appel, for sample collection. We also wish to thank Nadine Leyer for laboratory work and preparation of DNA samples.

Authors’ contributions

CMD set up the workflow and conducted the appropriate analyses, interpreted the results, selected animals and wrote the first draft of the manuscript. EH performed the ENDog analysis. CGB and ET developed the initial experimental design within the pigFit project. HPI provided the phenotype and genotype data sets and DNA samples. ET and CGB supervised the study, oversaw the analysis and contributed to the interpretation of the results. All authors contributed to the discussion of the results and development of the manuscript. All authors read and approved the final manuscript.

Funding

Open Access funding enabled and organized by Projekt DEAL. The study was performed within the ‘pigFıt’ project which was supported by funds from the German Government’s Special Purpose Fund held at Landwirtschaftliche Rentenbank (FK228-RZ-3-72.038). The funding body played no role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Availability of data and materials

Data cannot be made publicly available, as they are owned by a third party, the BH2P GmbH. The data sets used and analysed during the current study are available from the corresponding author on reasonable request and with permission of the BH2P GmbH pig breeding company (henne@bhzp.de).

Declarations

Ethics approval and consent to participate

All applicable international, national, and institutional guidelines for the care and use of animals were followed. Animals were reared within the BH2P nucleus herds in compliance with national regulations pertaining to livestock production and according to the procedures approved by the German Animal Protection Law (TierSchG). These animals are not considered as experimental animals as defined in EU directive 2010/63 and the Animal Protection Law (TierSchG). Blood sampling was performed by veterinarians as part of the routinely health and hygiene monitoring of the BH2P nucleus herds according to the Pig Keeping Hygiene Requirements Ordinance (SchaltHygV) §7 (1)

Sentence 2: No ethical approval was required for this study.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

1 Institute of Animal Science, University of Bonn, Endenicher Allee 15, 53115 Bonn, Germany. 2 BH2P GmbH, An der Wassermühle 8, 21368 Dahlen-Ellringen, Germany.

Received: 25 May 2021 Accepted: 31 January 2022

Published online: 19 February 2022

References

1. Hayes BJ, Daetwyler HD. 1000 Bull Genomes Project to map simple and complex genetic traits in cattle: applications and outcomes. Annu Rev Anim Biosci. 2019;7:89–102.
2. Groenen MAM, Archibald AL, Uenishi H, Tuggle CK, Takeuchi Y, Rothschild MF, et al. Analyses of pig genomes provide insight into porcine demography and evolution. Nature. 2012;491:393–8.
3. Ponsuksili S, Reyer H, Trakooljul N, Murani E, Wimmers K. Single- and bayerian multi-marker genome-wide association for haematological parameters in pigs. PLoS One. 2016;11:e0159212.
4. Bovo S, Mazzoni G, Bertolini F, Schiavo G, Giamalberti G, Gallo M, et al. Genome-wide association studies for 30 haematological and blood clinical-biochemical traits in Large White pigs reveal genomic regions affecting intermediate phenotypes. Sci Rep. 2019;9:7003.
5. Wetterstrand KA. DNA sequencing costs: data from the NHGRI Genome Sequencing Program (GSP); 2021. https://www.genome.gov/sequencingcostsdata. Accessed 30 Mar 2021.
6. Davey JW, Hohenlohe PA, Etter PD, Boone JQ, Catchen JM, Blaxter ML. Genome-wide genetic marker discovery and genotyping using next-generation sequencing. Nat Rev Genet. 2011;12:499–510.
7. Gorjanc G, Cleveland MA, Houston RD, Hickey JM. Potential of genotyping-by-sequencing for genomic selection in livestock populations. Genet Sel Evol. 2015;47:12.
8. Knol EF, Nielsen B, Knap PW. Genomic selection in commercial pig breeding. Anim Front. 2016;6:15–22.
9. van Binsbergen R, Bink M, Calus MPL, van Eeuwijk FA, Hayes BJ, Huissegge I, et al. Accuracy of imputation to whole-genome sequence data in Holstein Friesian cattle. Genet Sel Evol. 2014:46:41.
10. van Son M, Agarwal R, Kent MP, Grove H, Grindflek E, Lien S. Exploiting whole genome sequence data to fine map and characterize candidate genes within a quantitative trait loci region affecting androstenone on porcine chromosome 5. Anim Genet. 2017;48:653–9.
11. van Son M, Kent MP, Grove H, Agarwal R, Hamalain H, Lien S, et al. Fine mapping of a QTL affecting levels of skatole on pig chromosome 7. BMC Genet. 2017;18:85.
12. Derks MFL, Gjusvland AB, Bosse M, Lopes MS, van Son M, Harlizius B, et al. Loss of function mutations in essential genes cause embryonic lethality in pigs. PLoS Genet. 2019;15:e1008055.
13. Druet T, Macleod IM, Hayes BJ. Toward genomic prediction from whole-genome sequence data: impact of sequencing design on genotype imputation and accuracy of predictions. Heredity (Edinb). 2014;112:39–47.
14. Daetwyler HD, Capitan A, Pausch H, Stothard P, Van Binsbergen R, Brandum RF, et al. Whole-genome sequencing of 234 bulls facilitates mapping of monogenic and complex traits in cattle. Nat Genet. 2014;46:658–67.
15. Pausch H, MacLeod IM, Fries R, Emmerling R, Bowman PJ, Daetwyler HD, et al. Evaluation of the accuracy of imputed sequence variant genotypes and their utility for causal variant detection in cattle. Genet Sel Evol. 2017;49:24.
16. Cannon ME, Mohlke KL. Deciphering the emerging complexities of molecular mechanisms at GWAS loci. Am J Hum Genet. 2018;103:637–53.
17. Schad DJ, Chen W, Larson NB. From genome-wide associations to candidate causal variants by statistical fine-mapping. Nat Rev Genet. 2018;19:491–504.
18. Johnsson M, Jungnickel MK. Evidence for and localization of proposed causative variants in cattle and pig genomes. Genet Sel Evol. 2021;53:67.

19. van den Berg S, Vandenplas J, van Eeuwijk FA, Bouwman AC, Lopes MS, Veerkamp RF. Imputation to whole-genome sequence using multiple pig populations and its use in genome-wide association studies. Genet Sel Evol. 2019;51:2.

20. Heuß EM, Pröll-Cornelissen MJ, Neuhoff C, Tholen E, Große-Brinkhaus C. Invited review: Piglet survival: benefits of the immunocompetence. Animal. 2019;13:214–24.

21. Dauben CM, Pröll-Cornelissen MJ, Heuß EM, Appel AK, Henne H, Roth K, et al. Genome-wide associations for immune traits in two maternal pig lines. BMC Genomics. 2021;22:717.

22. Hickey JM, Kinghorn BP, Tier B, Wilson JF, Dunstan N, van der Werf JHJ. A combined long-range phasing and long haplotype imputation method to impute phase for SNP genotypes. Genet Sel Evol. 2011;43:12.

23. Gonen S, Ros-Freixedes R, Battagin M, Gorjanc G, Hickey JM. A method for the allocation of sequencing resources in genotyped livestock populations. Genet Sel Evol. 2017;49:47.

24. Ros-Freixedes R, Gonen S, Gorjanc G, Hickey JM. A method for allocating low-coverage sequencing resources by targeting haplotypes rather than individuals. Genet Sel Evol. 2017;49:78.

25. Goddard ME, Hayes BJ, Meuwissen THE. Using the genomic relationship matrix to predict the accuracy of genomic selection. J Anim Breeds Genet. 2011;128:409–21.

26. Gutiérrez JP, Goyache F. A note on ENDOG: a computer program for analysing pedigree information. J Anim Breeds Genet. 2005;122:172–6.

27. Boichard D, Maignel L, Verrier É. The value of using probabilities of gene origin to measure genetic variability in a population. Genet Sel Evol. 1997;29:5–23.

28. Sargolzaei M, Chesnais JP, Schenkel FS. A new approach for efficient genotype imputation using information from relatives. BMC Genomics. 2014;15:478.

29. Ma P, Brandum RF, Zhang Q, Lund MS, Su G. Comparison of different methods for imputing genome-wide marker genotypes in Swedish and Finnish Red cattle. J Dairy Sci. 2013;96:4666–77.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.