Kinetics of Inhibition of Monoamine Oxidase Using Curcumin and Ellagic Acid

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INTRODUCTION

Parkinson’s disease (PD) is the most common neurodegenerative disorder, affecting 1% of the population at the age of 65 and 4–5% of the population of the age of 85.\(^1,2\) PD is pathologically characterized by the loss of dopaminergic (DAergic) neurons and presence of intraneuronal cytoplasmic inclusions, termed “Lewy Bodies” in the substantia nigra pars compacta as well as in certain other brain nuclei.\(^1\) The symptoms of the disease are tremor or trembling in hands, arms, legs, jaw, and face; rigidity or stiffness of the limbs and trunk; bradykinesia or slowness of movement; and postural instability or impaired balance and coordination.\(^3\) Despite the known pathology, the etiology of this disease and the mechanisms involved in the neurodegeneration are still elusive.\(^4\)

There are various proposed mechanisms for the appearance and development of PD, including infection and inflammation, mitochondrial or proteasomal dysfunction, abnormal protein structures, dysregulation...
of heavy metals, and oxidative stress.[6] Generation of reactive oxygen species (ROS) may be a leading factor causing cell death[7] and other pathological aspects of PD,[8] however, whether ROS is the cause of PD or a consequence of the disease process itself, is still unknown.[9] One reason is that free radicals/toxic particles normally deactivated in the body are responsible, which can be controlled by antioxidants as an adjunct with dopamine agonist or monoamine oxidase (MAO) inhibitors.[10]

MAO are responsible for the oxidative degradation of a variety of neurotransmitters (e.g., noradrenaline, dopamine, and serotonin) as well as different exogenous and endogenous amines (i.e., tyramine and benzylamine) to their corresponding aldehydes.[11] Identification of MAO inhibitors is of great interest in drug discovery. Recent efforts toward the development of MAO inhibitors are focused on selective MAO type-A (MAO-A) or MAO type-B (MAO-B) inhibitors.[12] Selective MAO-A inhibitors are effective in the treatment of depression, whereas MAO-B inhibitors are logical application to the DAergic deficit in PD, depression, and Alzheimer’s disease.[13,14] Evaluation of naturals, botanicals, and dietary supplements for the MAO inhibitory constituents is considered important for improving their use and supporting their traditional use in treatment of depression, PD, and other neuropsychiatric as well as neurological disorders.[15] The natural polyphenols and their combinations constituted a large group of phytochemicals in herbal beverages, foods, fruits, and vegetables worldwide are suggested to have the optimal neuroprotective effect. However, little information is available on the possible synergistic and/or antagonistic interactions of the various polyphenols and their combinations.[16]

Ellagic acid, 2, 3, 7, 8-tetrahydroxy-chromeno [5, 4, 3-cde] chromene-5, 10-dione, is a naturally occurring powerful bioactive compound found in a range of plant species, especially fruits. It has potential pharmacological and industrial applications.[17] Ellagic acid is present in plants in the form of hydrolysable tannins called ellagittannins. These are the structural components of the plant cell wall and the cell membrane. Ellagittannins are hydrolysable molecules with hydroxybenzoic acid components.[18] The highest concentration of ellagic acid is found in many berries including strawberries, raspberries, cranberries, and grapes. Other sources of ellagic acid include walnuts, pecans, and beverages. Current research has revealed that ellagic acid possesses wide range of pharmacological properties such as antimutagenic, antioxidant, and anti-inflammatory activity in bacterial and mammalian systems.[19]

Curcumin is the chief, active curcuminoid of the dietary spice found in the rhizomes of Curcuma longa (turmeric), a plant in the ginger family (Zingiberaceae). Turmeric has been consumed for medicinal purposes for thousands of years. Extensive research on curcumin over the past few decades has revealed the health benefits of this ingredient to the modern era. The curcuminoids are a mixture of three principal compounds: Curcumin (curcumin I; 77%), demethoxycurcumin (DMC; curcumin II; 17%), and bis DMC (BDMC; curcumin III; 3%). The chemical name of curcumin is 1, 7-bis-4-hydroxy-3-methoxyphenyl)-hepta-1,6-diene-3,5-dione; the chemical formula is C21H20O6, and pKa value is 8.54.[20,21]

Selegiline (SG [(R)-(−)-N, α-dimethyl-N-2-propynyl-phenethylamine]) is a potent, irreversible, and selective inhibitor of MAO-B. It has been widely prescribed alone or in combination with levodopa in the treatment of PD.[22]

The present study was intended to investigate effects of the ellagic acid and curcumin on MAO-B and their kinetic inhibition. The in vitro effects of these selected polyphenols on MAO activities in mitochondria isolated from rat brains were examined. Brain mitochondria were assayed for MAO-B using benzylamine as substrates. Finally, we studied changes in kinetic parameters of MAO activity in the presence of different concentrations of ellagic acid and curcumin to determine the mechanism of inhibition.

### MATERIALS AND METHODS

#### Chemicals

Benzylamine was obtained from S.D. Fine Chemicals Ltd. Mumbai; mannitol, ethylenediaminetetraacetic acid disodium salt (EDTA), dimethyl sulfoxide (DMSO), and SG were obtained from Sigma Chemical (St Louis, MO, USA). All other chemicals used were of AR grade and were purchased from commercial sources.

#### Preparation of rat brain mitochondrial monoamine oxidase

Crude rat brain mitochondria were isolated as per the earlier described method of Gorgun et al.[23] In brief, the brain tissue was homogenized in the buffer of pH 7.4 containing 0.3 M mannitol and 0.1 mM EDTA. Homogenate was centrifuged at 600 g for 10 min at −4°C (R-248Mof CPR-24 plus Instrument, Remi, India). To obtain the brain mitochondria, the supernatant was collected and centrifuged at 10,000 g for 10 min at −4°C. The obtained mitochondrial pellets were washed 3 times with 0.25 M sucrose buffer containing 0.1 mM EDTA, resuspended in 0.25 M sucrose buffer, pH 7.4, and stored at −4°C for further studies.

#### Determination of protein, assay, and activity measurements of monoamine oxidase

The protein concentration and assay of MAO activity measurement with different concentrations (10–300 nM) of ellagic acid and curcumin were performed.[24] In brief, the reaction mixture contained 0.1 M phosphate buffer of pH 7.2, 10 mM benzylamine, MAO preparation, and different concentration of curcumin and ellagic acid (10–300 nM) in a total reaction volume of 200 μL in a 96-well plate. After 15 min of incubations at room temperature, readings were taken in at 250 nm.[24]

The effect of the selected constituents on MAO-B was explored by using the benzylamine deamination assay on 96-well plates.[25] A fixed substrate concentration and varying inhibitor concentrations (10–300 nM) were used to determine the higher half maximal inhibitory concentrations (IC50) value at the point where 50% inhibition of the catalytic activity of the enzyme occurred. For MAO-B, the substrate concentration of 10 mM benzylamine was chosen. The assay was performed wi01th the addition of inhibitor under study. Inhibition was calculated as percent of product formation compared to the corresponding control (enzyme-substrate reaction) without the inhibitors. We also performed separate controls in which the enzyme preparation is replaced by substrate, at the concentration used in the assay. The reactions were carried out in 0.1 M potassium phosphate buffer at pH 7.2. Incubations mixtures contained 100 μL of MAO preparation. The inhibitor was dissolved in buffer. The total reaction volume was 200 μL yielding a final DMSO concentration of 1% in the reaction mixture. The reaction mixtures were preincubated for 10 min at 37°C followed by the addition of MAO-B to initiate the reactions. Reactions were incubated for 15 min at 37°C and were determined fluorometrically, a flexible monochromator-based multi-mode microplate reader (Synergy HT, Bio-Tek) at 250 nm by using the GraphPad Prism 5.0 Version for Windows, GraphPad Software (San Diego, CA, USA) data analysis program.

% Inhibition = \( \frac{\text{(activity of treated sample – blank)}}{\text{(activity of control sample – blank)}} \times 100 \)

#### Kinetics of monoamine oxidase inhibition

To understand the nature of MAO inhibition, sets of three concentrations of curcumin and ellagic acid (200, 400, 600 nM) were used and...
compared with the kinetic behavior of a control set. For determination of $K_m$ and $V_{max}$ values for inhibition of MAO-B with curcumin and ellagic acid, the assays were carried out at different enzyme concentration and total volume of reaction mixture was kept constant but the substrate (benzylamine) concentration varied (5, 10, and 15 mM). Controls without inhibitor were also run simultaneously. The results are presented as double reciprocal Lineweaver–Burk plots and the kinetic data namely $K_m$ and $V_{max}$ values.

RESULTS

Determination of monoamine oxidase inhibition and higher half maximal inhibitory concentrations values

Curcumin and ellagic acid were evaluated in vitro for MAO-B inhibitory activity against rat brain MAO-B, whereby the ellagic acid demonstrated more potent MAO-B inhibitory activity than curcumin [Table 1]. The inhibition of MAO-B by curcumin (IC$_{50}$ 500.46 nM) and ellagic acid (IC$_{50}$ 412.24 nM) was found to be 10-fold less potent compared to the inhibition of standard SG (IC$_{50}$ 65.50 nM) [Table 1 and Figure 1].

Evaluation of inhibition mechanism and kinetics

Detailed in vitro studies were carried out to understand the kinetics and mechanism of inhibition of rat brain MAO-B by curcumin and ellagic acid. Both curcumin and ellagic acid were tested against MAO-B at varying concentrations of benzylamine, a selective substrate, to investigate the nature of inhibition of the enzymes. Based on dose-response inhibition, two concentrations of the inhibitors were selected, one below and another above IC$_{50}$ value for the inhibition. For each experiment, three sets of assays were done at varying concentrations of the substrate, one control without inhibitors and with two concentrations of the inhibitor. The enzyme kinetics data are presented as double reciprocal Lineweaver–Burk plots [Figure 2]. Binding of curcumin and ellagic acid with rat brain MAO-B increase the $K_m$ value (the Michaelis–Menten constant) with no apparent effect on the $V_{max}$ indicating that the inhibition of MAO-B by curcumin (noncompetitive) and ellagic acid (competitive) is mixed type of inhibition [Figure 2].

DISCUSSION

In recent years, increasing attention has been devoted to the management or prevention of PD by herbal medicines. Recent studies indicated that the active components of herbal medicines, herbal extracts, and herbal formulations have effect on different in vitro and in vivo PD models. The present advances in pharmacokinetics and new drug development have shown the possibility of selective inhibition of brain MAO as compared to that in periphery, thus increasing the clinical potential of these drugs. SG and rasagiline are propargyl derivatives that have potent neuroprotective action and selective MAO-B inhibitory properties. These drugs proposed to be main discussion about drug treatment of PD.

MAOs are flavoproteins which catalyze the oxidative deamination of primary, secondary, and tertiary amines. The primary functions of MAOs are the metabolism of exogenous amines and the regulation of neurotransmitter level and intracellular amine stores. MAO-B preferentially deaminates phenylethylamine, benzylamine, and dopamine. Thus, inhibition of MAO-B may alleviate symptoms of PD.

Curcumin, an active ingredient isolated from Curcuma longa (L.), has been showed to possess extensive pharmacological activities, including antcardiovascular disorders, antitumor activities, cholesterol-lowering, antidiabetic, anti-inflammatory, and myelodysplastic syndromes,

Table 1: Inhibition of monoamine oxidase type-B activity in brain crude mitochondrial fraction

|               | IC$_{50}$ (nM) | Mechanism of inhibition | n |
|---------------|---------------|-------------------------|---|
| Curcumin      | 500.46        | None-competitive         | 5 |
| Ellagic acid  | 400.24        | Competitive              | 5 |
| Selegiline    | 65.50         | Competitive              | 5 |

Values are means±SEM. IC$_{50}$: Drug concentration that is required for 50% inhibition of enzyme activity; n: Number of measurements; SEM: Standard error of mean.

Figure 1: Concentration-dependent inhibition of rat brain monoamine oxidase-B by curcumin and ellagic acid. The activity was expressed as % inhibition. Each point represents mean ± standard deviation of triplicate values

Figure 2: Lineweaver-Burk plots of inhibition of rat brain monoamine oxidase-B with (a) curcumin (200–600 nM) and (b) ellagic acid (200–600 nM); V: nmoles/min/mg protein; S: substrate benzylamine concentration (mM)
and anti-nerve degeneration. However, molecular mechanisms of curcumin-mediated anti-PD are still unknown.[21] The present study reveals that curcumin and ellagic acid can inhibit MAO activity. The inhibitory potency and mechanism of inhibition of curcumin and ellagic acid toward rat brain MAO-B were assessed. The results indicate that some effects of these selected polyphenols on brain functions could be related to a nonreceptor modulation of monoaminergic neurotransmission.

Inhibitory and kinetic parameters calculated in this article result from total concentrations of curcumin and ellagic acid added to crude brain mitochondrial fraction. The situation in various tissues in vivo may be different from our experimental conditions. This was the reason why the effects of curcumin and ellagic acid on MAO's activity were compared with the effect of a well-known MAO inhibitor such as SG.[12] The significant MAO inhibitory activity of curcumin and ellagic acid as testified herein suggest additional potential utility of these two polyphenols for alleviation of neurological disorders associated with depletion of monoamines, the most important of which are depression and PD.[20]

The results of the kinetic studies of MAO inhibition using different concentrations (200–600 nM) of curcumin and ellagic acid were plotted as double reciprocal Lineweaver–Burk plot [Figure 2]. The mode of inhibition of both compounds toward MAO-B is mixed (competitive and uncompetitive) type of inhibition. The \( V_{\text{max}} \) remained constant while the \( K_m \) varied from 16.66 ± 1.2, 28.57 ± 1.1, and 40.0 ± 1.4 mM for 200–600 μg/ml concentration of ellagic acid as compared to control (9.53 ± 2.10 mM) while for curcumin, \( K_m \) (6.25 mM) remains constant and \( V_{\text{max}} \) varied from 11.76 ± 1.86, 5.26 ± 2.11, and 3.77 ± 1.26 mM for 200–600 μg/ml concentration of ellagic acid as compared to control (22.22 ± 1.34 mM). The kinetic constant \( K_m \) which is basically a substrate concentration signifies the extent of affinity of an enzyme with its substrate. The calculated \( K_m \) values of the present investigation also signify that with increase in the concentration of curcumin and ellagic acid, there is corresponding increase in \( K_m \) indicating the decrease in affinity of MAO for its substrate. The decrease in \( K_m \) in relation to increase in concentration of curcumin and ellagic acid also indicates the effective inactivation of MAO at higher concentrations. Inhibitors of MAO-B not only lead to enhanced DAergic neurotransmission but also prevent activation of toxin and free radical formation, and they alleviated the process of neuron denaturation. The phytochemicals showed that there exists a structural similarity between natural polyphenolic compounds and established cholinesterase inhibitors in terms of molecular weight, phenolic rings, and hydrophobic component are described as competitive inhibitors of MAO.[27,34,35]

CONCLUSIONS

In the present study, we have evaluated kinetics of inhibition of MAO using curcumin and ellagic acid. The results of present reveal that both curcumin and ellagic acid inhibit the MAO activity with both the competitive and noncompetitive type of inhibitions. Curcumin and ellagic acid can be considered as a possible source of MAO inhibitor used in the treatment of Parkinson’s and other neurological disorders. Although it is always difficult to extrapolate from in vitro studies to the clinical reality, the present results suggest that the examined polyphenols might contribute to an inhibitory effect on MAO, and a systematic in vivo investigation of the curcumin and ellagic acid effect on MAO activity is needed. The exact physiological role of MAO-B inhibition by curcumin and ellagic acid is not known; however, their effects on MAO activity may contribute to nonreceptor actions of curcumin and ellagic acid participating in modulation of monoamine neurotransmission in the brain.

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Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Khalil WK, Assaf N, EIShebinye SA, Salem NA. Neuroprotective effects of bee venom acupuncture therapy against rotenone-induced oxidative stress and apoptosis. Neurochem Int 2015;80:79-86.
2. Patel SP, Jain PD, Sancheti JS, Ghumarkar PJ, Tambe R, Satheye S. Neuroprotective and neuroprotective effects of apigenin and luteolin in MPTP induced Parkinsonism in mice. Neuropsychopharmacology 2014;46:192-202.
3. Alaf Y, Vaya J, Khalib S, Loboda Y, Finberg JP. Selective inhibition of monoamine oxidase A or B reduces striatal oxidative stress in rats with partial depletion of the nigro-striatal dopaminergic pathway. Neuropsychopharmacology 2013;38:48-57.
4. Khatri DK, Juvekar AP. Propensity of Hyoscyamus niger seeds medicinalth to alloy stereotaxically rotenone-induced Parkinson’s disease symptoms in rats. Orient Pharm Exp Med 2015;15:327-39.
5. Mitra S, Chafrabati N, Dutta SS, Roy S, Bhattacharya P, Sinha P, et al. Genderspecific brain regional variation of neurons, endogenous estrogen, neurotransmission and glial cells during rotenone-induced mouse model of Parkinson’s disease. Neuroscience 2015;292:46-70.
6. Nazari QA, Takada-Takatori Y, Hashimoto T, Imaizumi A, Izumi Y, Akaike A, et al. Potential protective effect of highly bioavailable curcumin on an oxidative stress model induced by microinjection of sodium nitroprusside in mouse brain. Food Funct 2014;5:984-99.
7. Khatri DK, Juvekar P, Juvekar A. Phytochemical investigation and in vitro antioxidant activities indigolida cordifolia seed extracts. Int J Pharm Pharm Sci 2013;5:71-5.
8. Goluk K. Oral supplements of aqueous extract of tomato seeds alleviate motor abnormality, oxidative impairments and neurotoxicity induced by rotenone in mice: Relevance to Parkinson’s disease. Neurochem Res 2014;39:1362-94.
9. Bepe GJ, Dongmo AB, Fayet HS, Tsiang N, Oteau Z, Cianoa O, et al. Memory-enhancing activities of the aqueous extract of Albizia adiantifoliate leaves in the 6-hydroxydopamine lesion rodent model of Parkinson’s disease. BMC Complement Altern Med 2014;14:132.
10. Tarazi FI, Sahl ZT, Wolny M, Mousa SA. Emerging therapies for Parkinson’s disease: From bench to bedside. PharmacolTher 2014;144:123-33.
11. Singh R, Ramakrishna R, Bhatena M, Bhatta RS. In vitro evaluation of Bacopa monniera extract and individual constituents on human recombinant monoamine oxidase enzymes. Phytother Res 2014;28:1419-22.
12. Fisar Z, Hrudová J, Raboch J. Inhibition of monoamine oxidase activity by antidepressants and mood stabilizers. Neuro Endocrinol Lett 2010;31:645-56.
13. Riedener P, Laux G. MAO-inhibitors in Parkinson’s disease. Exp Neurobiol 2011;20:1-17.
14. Cai Z. Monoamine oxidase inhibitors: Promising therapeutic agents for Alzheimer’s disease (Review). Mol Med Rep 2014;9:1533-41.
15. Chaurasiya ND, Ibraheem MA, Muhammad I, Walker LA, Tekwani BL. Monoamine oxidase inhibitory constituents of propolis: Kinetics and mechanism of inhibition of recombinant human MAO-A and MAO-B. Molecules 2014;19:8996-52.
16. Nguonsoure VL, Schluesselner J, Moundipa PF, Schluesener HJ. Natural polyphenols binding to amylid: A broad class of compounds to treat different human amyloid diseases. Mol Nutr Food Res 2015;59:8-20.
17. Garcia-Niño WR, Zazueta C. Ellagic acid: Pharmacological activities and molecular mechanisms involved in liver protection. Pharmacol Res 2015;97:84-103.
18. Teixeira Lde L, Bertoldi FC, Lajolo FM, Hassimotto NM. Identification of ellagittannins and flavonoids from Eugenia brasilienses Lami. (Grunimixama) by HPLC-ESI-MS/MS. J Agric Food Chem 2015;63:5417-27.
19. Huerga-González V, Lage-Yusty MA, Lago-Crespo M, López-Hernández J. Comparison of methods for the study of ellagic acid in pomegranate juice beverages. Food Anal Methods 2015;8:2286-93.
20. Alexander A, Qureshi A, Kumar L, Vaishnav P, Sharma M, et al. Role of herbal bioactives as a potential bioavailability enhancer for active pharmaceutical ingredients. Fitoterapia 2014;97:1-14.
21. Prasad S, Gupta SC, Tyagi AK, Aggarwal BB. Curcumin, a component of golden spice: From bedside to bench and back. Biotechnol Adv 2014;32:1053-64.
22. Robakis D, Fahn S. Defining the role of the monoamine oxidase-B inhibitors for Parkinson's disease. CNS Drugs 2015;29:433-41.
23. Gorgun FM, Zhao M, Singh S, Englander EW. Neuroglobin mitigates mitochondrial impairments induced by acute inhalation of combustion smoke in the mouse brain. Inhal Toxicol 2014;26:361-9.
24. Gacche RN, Shaikh RU, Chapole SM, Jadhav AD, Jadhav SG. Kinetics of inhibition of monoamine oxidase using Cymbopogon martini (Roxb.) Watts: A potential antidepressant herbal ingredient with antioxidant activity. Indian J Clin Biochem 2011;26:303-8.
25. Kotelnikova RA, Smolina AV, Grigoryev VV, Faingold II, Mischenko DV, Rybkin AY, et al. Influence of water-soluble derivatives of[60]fullerene on therapeutically important targets related to neurodegenerative diseases. Med Chem Commun 2014;5:1664-8.
26. Li XZ, Zhang SN, Liu SM, Lu F. Recent advances in herbal medicines treating Parkinson's disease. Fitoterapia 2013;84:273-85.
27. Finberg JP. Update on the pharmacology of selective inhibitors of MAO-A and MAO-B: Focus on modulation of CNS monoamine neurotransmitter release. Pharmacol Ther 2014;143:133-52.
28. Coelho-Cerqueira E, Netz PA, do Canto VP, Pinto AC, Follmer C. Beyond topoisomerase inhibition: Antitumor 1, 4-naphthoquinones as potential inhibitors of human monoamine oxidase. Chem Biol Drug Des 2014;83:401-10.
29. Yang J, Song S, Li J, Liang T. Neuroprotective effect of curcumin on hippocampal injury in 6-OHDA-induced Parkinson's disease rat. Pathol Res Pract 2014;210:357-62.
30. Ademosun AO, Oboh G. Comparison of the inhibition of monoamine oxidase and butyrylcholinesterase activities by infusions from green tea and some citrus peels. Int J Alzheimers Dis 2014;2014:586407.
31. Viña D, Serra S, Lamela M, Delogu G. Herbal natural products as a source of monoamine oxidase inhibitors: A review. Curr Top Med Chem 2012;12:2131-44.

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