Increased expression of adenosine 2A receptors in metastatic renal cell carcinoma is associated with poorer response to anti-vascular endothelial growth factor agents and anti-PD-1/Anti-CTLA4 antibodies and shorter survival

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Abstract

Background Adenosine and its adenosine 2A receptors (A2AR) mediate the immunosuppressive mechanism by which tumors escape immunosurveillance and impede anti-tumor immunity within the tumor microenvironment. However, we do not know whether the adenosine pathway (CD39/CD73/A2AR) plays a role in renal cell carcinoma (RCC). Therefore, we studied the role of immunosuppression in RCC by assessing the adenosine pathway in patients with RCC treated with anti-vascular endothelial growth factor (anti-VEGF) agents or immune checkpoints inhibitors (ICIs) or both.

Methods In 60 patients with metastatic RCC, we examined the expression of CD39, CD73, A2AR, and programmed cell death 1 ligand 1 (PD-L1) immunohistochemically in surgically resected tumor tissues and studied the clinicopathological characteristics of these patients. Patients were treated by cytoreductive nephrectomy with systemic therapy with anti-VEGF agent or a combination of the ICIs anti-cytotoxic T-lymphocyte-associated antigen 4 (CTLA4) antibody and programmed cell death 1 (PD-1) antibody.

Results Increased expression of A2AR in the primary tumors was associated with metastatic profiles. Patients treated with anti–PD-1 antibody in monotherapy, a combination of anti-PD-1 and anti-CTLA4 antibodies, or anti-VEGF agent showed better response and longer overall survival if the primary tumor had higher PD-L1 expression and lower A2AR expression. In Cox multivariate regression analysis, higher expression of A2AR was associated with shorter overall survival.

Conclusions Our findings suggest that the expression of A2AR and PD-L1 in the primary tumors in RCC might predict the outcomes of treatment with anti-VEGF agents and ICIs and that the A2AR pathway might be a molecular target for immunotherapy.

Keywords Adenosine 2A receptor (A2AR) · CD39 · CD73 · PD-L1 · Renal cell carcinoma

Abbreviations

A2AR Adenosine 2A receptor
ADP Adenosine diphosphate
AMP Adenosine monophosphate
APC Antigen-presenting cell
ATP Adenosine triphosphate
ccRCC Clear cell renal cell carcinoma
cD Cluster of differentiation
CD39 Membrane-bound ectonucleoside triphosphohydrolase 1
CD73 Ecto-5′-nucleotidase
chRCC Chromophobe renal cell carcinoma
coRCC Collecting duct renal cell carcinoma
CTLA-4 Cytotoxic T-lymphocyte-associated antigen 4
DC Dendritic cells
HIF Hypoxia-inducible factor

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Introduction

Renal cell carcinoma (RCC) is immunogenic and proangiogenic. The introduction of anti-vascular endothelial growth factor (VEGF) therapies as a new treatment for metastatic RCC has improved the prospects of patients with this disease. However, many tumors eventually develop resistance to such therapies due to secondary mutation of the target protein or compensatory changes [1]. Accordingly, interest has grown in immunotherapy as an alternative to conventional treatment with cytokines such as interferon and interleukin two. The immune system is theoretically capable of suppressing tumor development and mediating tumor regression, but this process requires that tumor antigen-specific T cells are generated and activated. Thus, tumor cells can survive if they avoid being destroyed by the immune system, an ability that is now considered as one of the hallmarks of cancer [2].

The immune modulation mediated by the immune checkpoint pathway ensures that tissue is protected from collateral damage during an inflammatory response. The B7 family of immune-regulatory ligands, such as cytotoxic T-lymphocyte-associated antigen 4 (CTLA4) and programmed cell death 1 (PD-1)/PD-ligand 1 (PD-L1), are key players in immune checkpoints that positively or negatively regulate various immune responses, and tumor cells use this regulatory mechanism to evade a tumor-directed T-cell response by upregulating CTLA4 or PD-1/PD-L1 [3, 4]. Targeting the inhibitory CTLA4 or PD-1/PD-L1 with monoclonal antibodies has shown striking antitumor activity in patients with cancers, and a number of these objective responses seemed to be durable. Thus, immune checkpoint blockade may be a new standard for the treatment of cancer and have prospects for long-term clinical benefit, including advanced RCC [5, 6]. However, cancer cells develop innate or adaptive immune resistance and progress while being treated with anti-VEGF therapy or immune checkpoints inhibitors (ICIs), such as an anti-PD-1 antibody or an anti-CTLA4 antibody [3, 4].

Multiple immunosuppressive mechanisms impede anti-tumor immunity. These mechanisms include the accumulation of extracellular adenosine through the activation of purinergic receptors, a potent and widespread strategy exploited by tumors to escape immunosurveillance [7, 8]. Adenosine is a purine nucleotide released by injured tissue that decreases inflammation and protects tissue from immune-mediated damage [9–11]. Intracellular adenosine is metabolized to adenosine monophosphate (AMP) and inosine by adenosine kinase and adenosine deaminase. Adenosine triphosphate (ATP) and adenosine diphosphate (ADP) released outside the cell are dephosphorylated by membrane-bound ectonucleoside triphosphate diphosphohydrolase 1 (CD39) and ecto-5’-nucleotidase (CD73) localized on the cell surface and are converted to extracellular adenosine [8–11]. In recent years, ATP released by cytotoxicity was shown to cause an inflammatory response via ATP receptors on the cell membrane [12, 13]. Among four cell membrane adenosine receptors (A1, A2A, A2B, A3A), A2A adenosine receptor (A2AR) has the high affinity for adenosine and is present on cells of both the innate and the adaptive immune systems [14], and engagement of A2AR is a critical regulatory mechanism that protects tissues against excessive immune reactions [9–11]. In tumors, this pathway is hijacked and hinders anti-tumor immunity, promoting cancer progression [9–11]. Studies in animal models have shown that prior treatment with anti–PD-1 antibodies results in increased expression of A2AR and CD73, suggesting that the adenosine pathway may contribute to therapeutic resistance to immunotherapy [15, 16]. Accordingly, interest has been increasing in new immunotherapy modalities targeting the adenosine pathway (CD39/CD73/A2AR) for cancer treatment [9–11].

Regarding the adenosine pathway in RCC, studies have shown that high CD73 expression correlates with poor prognosis [17] and that the ADORA2A (A2AR) and NT5E (CD73) genes are both highly expressed in the disease [18]. Recently, a phase I clinical trial with a small-molecule A2AR antagonist showed that this molecule could safely block adenosine signaling in a cohort of 68 RCC patients who had progressed on PD-1/PD-L1 inhibitors [18]. Thus, the adenosine pathway may play roles in
the tumor microenvironment in RCC, and may be attractive for an immunotherapy. Therefore, we studied the relationship between the expression levels of PD-L1, CD39, CD73, and A2AR and clinicopathological features in patients with metastatic RCC who underwent cytoreductive nephrectomy and treated by anti-VEGF therapy or with an anti–PD-1 antibody monotherapy or a combination of anti-CTLA4 and anti–PD-1 antibodies. Such information should shed light on the role and biological significance of the adenosine pathway in RCC.

Materials and methods

Patients

This study was performed with data and surgical samples from 60 patients (43 men and 17 women; median age, 65 years; range 17–79 years) with histopathologically diagnosed metastatic RCC treated by cytoreductive nephrectomy at our center from 2011 to 2019. The types of RCC were as follows: clear cell RCC (ccRCC), 50 patients; papillary RCC type 2 (pRCC2), four patients; collecting duct RCC (cdRCC), two patients; papillary RCC type 1 (pRCC1), one patient; chromophobe RCC (chrRCC), one patient; sarcomatoid RCC, one patient; and RCC with Xp11 translocation involving TFE3 gene fusion (TFE-3 RCC), one patient. We also resected ten metastatic lesions during cytoreductive nephrectomy in ten patients, including seven distant organs and three lymph nodes.

All 60 patients treated by cytoreductive nephrectomy underwent preoperative computed tomography or magnetic resonance imaging or both for staging of the primary tumor. After cytoreductive nephrectomy, 49 patients were treated with sunitinib or pazopanib as first-line systemic VEGF-targeting therapy for metastases and 11 patients were treated with a combination of anti-CTLA4 and anti–PD-1 antibodies. Of the 49 patients treated with an anti-VEGF agent, 20 were subsequently treated with anti–PD-1 antibody monotherapy as a second- or third-line treatment.

We also examined primary renal tumor samples from five patients with metastatic RCC (two ccRCC, two pRCC2, and one coRCC) who did not undergo cytoreductive nephrectomy and were treated with a combination of anti-CTLA4 and anti–PD-1 antibodies as first-line therapy. The primary renal tumors in these five patients were assessed by core needle biopsies.

Treatment effects were assessed according to the Response Evaluation Criteria in Solid Tumors (RECIST) criteria. Follow-up ranged from 3 to 77 months, with a median of 29 months. To assess metastatic lesions, computed tomography or magnetic resonance imaging or both was performed every 2–4 months.

To obtain the data for this study, we reviewed all participants’ medical records in March 2020. The study was conducted in accordance with the Declaration of Helsinki and was approved by the ethical review board of Dokkyo Medical University Hospital. Each patient provided written informed consent by signing a consent form that was approved by our institutional Committee on Human Rights in Research.

Immunohistochemistry

Antibodies against CD8 (PA0183, Leica Biosystems Newcastle Ltd), PD-L1 (OptiView PD-L1 [SP142]; Ventana Medical Systems, Inc.), CD39 and CD73 (D7F9A; Cell Signaling Technology), and A2AR (SA654; ENZO Life Science) were used in formalin-fixed, paraffin-embedded tissues for immunohistochemical staining with the automated BOND system (Leica BOND-III system, Leica Biosystems Newcastle Ltd), as described previously [19]. Three of the authors independently examined 1500–2000 cancer cells and tumor-infiltrating lymphocytes (infiltrating immune cells that have a small, rounded cell, and a large dark-stained nucleus with little eosinophilic cytoplasm) in 7–10 microscopic fields of 2–3 slide sections of the eosin-stained slides and the identical immunostaining slides in a high-power view. The investigators rated the level of expression of PD-L1, CD39, CD73, and A2AR in the tumor cells (TC) and infiltrating immune cells (IC) on a score of 0–3, as follows: < 1% staining, TC-0/IC-0; 1–5% staining, TC-1/IC-1; 5–30% staining, TC-2/IC-2; and > 30% staining, TC-3/IC-3. Subsequently, the tumors were divided into two groups: a low expression group in which TCs and ICs were negative or weakly positive for the antibodies (< 5% of both of TCs and ICs were positive, TC-1 or IC-1; TC-0 or IC-0), and a high expression group in which TCs or ICs showed strong positivity for the antibodies (> 30% of either TCs or ICs, or both were positive, TC-3 or IC-3; TC-2 or IC-2), as performed in our previous study [20]. Accordingly, we evaluated the response rate and overall survival in the high expression and low expression groups.

We also examined the expression of PD-L1, CD39, CD73, and A2AR in 10 resected metastatic tumors from ten patients.

Statistical analysis

We used Fisher’s exact test to determine whether the two categorical variables were associated with each other. The Kaplan–Meier method was used to create overall survival curves, and differences between the curves were assessed with the log-rank test. We examined prognostic factors affecting survival by Cox regression analysis. In all analyses,
\( P < 0.05 \) indicated statistical significance. Analyses were performed with commercial software.

**Results**

Many of the cells that showed a positive reaction for anti–PD-L1, anti-CD8, anti-CD39, anti-CD73, and anti-A2AR antibodies were immune cells (Figs. 1, 2, 3, Supplementary Figs. 1, 2), although some tumor cells also showed a positive reaction for these antibodies, in particular for anti-A2AR antibody (Figs. 2, 3, 4, Supplementary Figs. 1, 2).

Increased expression of A2AR in the primary tumors was significantly associated with metastatic profiles for several organs (\( P = 0.0002 \)) but was not significantly associated with regional lymph node involvement (\( P = 0.0744 \), Table 1). There was no relationship between expression of A2AR and histological grade or local invasion (Table 1). Compared with ccRCC, non-clear cell RCC (non-ccRCC) had a higher expression of A2AR (\( P = 0.0071 \)) and lower expression of PD-L1 (\( P = 0.0109 \)); however, we found no difference between RCC histological types and expressions for CD39 (\( P = 0.4058 \)) or CD73 (\( P = 0.2963 \)) (Fig. 3, Supplementary Figs. 3–5).

In all 60 patients, neither histological grade nor pT stage influenced the effects of the treatments; however, patients with regional lymph node involvement had a poorer response in distant metastatic organs (\( P = 0.0132 \), Table 1). Regardless of whether patients were treated only with an anti-VEGF (\( n = 29 \) patients) agent, with an anti-VEGF agent followed by anti–PD-1 antibody monotherapy (\( n = 20 \)), or only with anti–PD-1 and anti-CTLA4 antibodies (\( n = 11 \)), patients with lower A2AR expression in the primary tumors had better response (complete response, partial response, or stable disease) to anti-VEGF agents (\( P < 0.0001 \)) and ICIs (\( P = 0.0002 \) (Table 1) and longer survival (\( P = 0.0001 \)) (Fig. 5).

Expressions of PD-L1, CD39, and CD73 were not related to histological grade, local invasion, regional lymph node involvement, or metastatic profiles (Table 1).

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**Fig. 1** Immunohistochemistry of a case of better response to anti-VEGF therapies and longer survival. 73 y.o. male of left clear cell renal cell carcinoma with Fuhrman grade 2, pT2bN0M1 (PUL, LYM). CD8; high expression, PD-L1; low expression, CD39; high expression, CD73; low expression, A2AR; low expression. After cytoreductive nephrectomy, this patient had received sunitinib as first-line anti-VEGF therapy for 17 months with best response of partial response. After refractory to sunitinib, he has received axitinib as second-line anti-VEGF therapy with alive with disease for 29 months.
The expression levels of PD-L1 in the primary tumors did not relate to the response to anti-VEGF therapies for metastatic lesions or survival in any of the 60 patients (Table 1 and Fig. 5); however, among the 31 patients who received ICIs, 13 patients with higher PD-L1 expression showed better response to ICIs \( (P = 0.0130) \) (Table 1) and a weak tendency towards longer survival \( (P = 0.1581) \). The primary tumors of 37 of the 60 patients showed higher expressions of CD39, but expression levels of CD39 had no relation to the response to the therapies or survival (Table 1 and Fig. 5). In contrast, the primary tumors in 44 patients had lower expression of CD73, which was weakly related to longer survival \( (P = 0.0570, \text{Fig. 5}) \), but not treatment response (Table 1). Although CD8-positive T cells were seen in many tumor microenvironments, expression of CD8 had no influence on histological grade, local invasion, regional lymph node involvement, or response to anti-VEGF agents or ICIs. On the other hand, our findings indicated that pulmonary metastatic lesions might be more closely linked to lower A2AR expression in primary renal tumor \( (P = 0.0002) \) and better response to anti-VEGF agents \( (P < 0.0001) \) and ICIs \( (P = 0.0002) \) than other metastatic lesions are (Table 1).

Our analysis of the expression of PD-L1, CD39, CD73, and A2AR in the 10 resected metastatic lesions showed a positive relation of the expression levels in the primary tumor and metastatic lesion for A2AR \( (P = 0.0157) \) but not for PD-L1 \( (P = 0.3894) \), CD39 \( (P = 0.5271) \), or CD73 \( (P = 0.1967) \). We found that 7 of these patients had a higher expression of A2AR in both the metastatic lesion and the primary renal tumor. In six of these seven patients, other nonresectable metastatic lesions showed poorer response to anti-VEGF agents and subsequent anti–PD-1 antibody monotherapy or a combination of anti–PD-1 and anti-CTLA4 antibodies, regardless of the expression of PD-L1 in the primary tumor and metastatic lesion (Fig. 4, Supplementary Fig. 6, 7); in contrast, the 3 patients with a lower expression of A2AR in the metastatic lesions and/or primary tumor

![Image](Immunohistochemistry of a case of worse response to first-line anti-VEGF therapy and second-line nivolumab and shorter survival. 64 year old male of right clear cell renal cell carcinoma with Fuhrman grade 3, pT2aN0M1 (PUL, HEP). CD8; high expression, PD-L1; high expression, CD39; high expression, CD73; high expression, A2AR; high expression. After cytoreductive nephrectomy, this patient had received pazopanib as first-line anti-VEGF therapy for 3 months, while the metastatic disease progressed. Then the patient received nivolumab as second-line therapy for 3 months. The best response was partial response for pulmonary and liver lesions, however, the lesions progressed rapidly and the patient died.)

\( \text{Fig. 2} \) Immunohistochemistry of a case of worse response to first-line anti-VEGF therapy and second-line nivolumab and shorter survival. 64 year old male of right clear cell renal cell carcinoma with Fuhrman grade 3, pT2aN0M1 (PUL, HEP). CD8; high expression, PD-L1; high expression, CD39; high expression, CD73; high expression, A2AR; high expression. After cytoreductive nephrectomy, this patient had received pazopanib as first-line anti-VEGF therapy for 3 months, while the metastatic disease progressed. Then the patient received nivolumab as second-line therapy for 3 months. The best response was partial response for pulmonary and liver lesions, however, the lesions progressed rapidly and the patient died.
had a higher PD-L1 expression in the metastatic lesions and showed better treatment response (Supplementary Fig. 8).

In the five patients with metastatic RCC treated with a combination of anti-CTLA4 and anti–PD-1 antibodies, we found no relationship between CD8-positive T cells in the tumor microenvironment in core-needle biopsy for primary renal tumors and treatment response. Two of the patients had lower A2AR expression, and their renal tumors and metastatic lesions showed partial remission or stable disease after treatment (Supplementary Fig. 9). The remaining three patients had higher A2AR expression, and two of these three patients showed stable response in some lesions or poorer response in others (Supplementary Fig. 9).

To study the roles of PD-L1 and A2AR, we combined the expression levels of PD-L1 and A2AR in the primary tumor and subsequently classified the patients into four groups: type I, PD-L1 low and A2AR high; type II, PD-L1 high and A2AR high; type III, PD-L1 low and A2AR low; and type IV, PD-L1 high and A2AR low. Type IV expression was associated with longer survival (Fig. 5e), in particular in patients who received ICIs (Fig. 5f). In contrast, patients with higher A2AR expression in the primary tumor showed worse response (progressive disease) to the anti-VEGF agents or ICIs ($P = 0.0002$, Table 1), and poorer overall survival ($P = 0.0001$, Fig. 5), regardless of expression of PD-L1.

When grade, pT stage, lymph node metastasis stage, PD-L1, CD39, CD73, A2AR and histology (ccRCC vs. non-ccRCC) were correlated in Cox univariate analysis, higher histological grade, higher pT stage, positive lymph node metastasis, and higher CD73 and A2AR were associated with shorter overall survival. In Cox multivariate analysis, A2AR and grade were significant ($P = 0.0006$, $P = 0.0062$, respectively, Table 2).

**Discussion**

The present study examined the expression levels of PD-L1, CD39, CD73, and Although the expression pattern of these four molecules was diverse in individual cases, we found...
that higher A2AR expression in the primary tumor was associated with worse response to both anti-VEGF agents and ICIs. A higher A2AR expression in the primary tumor was also an independent prognostic factor for shorter survival. Furthermore, patients with higher PD-L1 expression in the primary tumor who were treated with ICIs showed a more favorable response but not a longer survival. Last, lower CD73 expression correlated with longer survival but not with treatment response. CD39 levels were not associated with treatment response or survival. These findings suggest that not only PD-L1 but also A2AR might be required in the tumor microenvironment to allow the tumor to evade the host immune system and that blockade of the A2AR pathway might be a new anti-tumor response for advanced RCC.

Because cancer cells evade the immune system by hijacking immune cells [2], elucidating the orchestrating mechanism between tumor and immune cells in the tumor microenvironment might be essential to enable cancer immunotherapy. The expression of PD-L1 on tumor cells has been associated with good prognosis and sustained clinical responses in immunotherapeutic regimens based on PD-L1/PD-1/CTLA4 immune checkpoint blockade; however, the relative importance of PD-L1 expression on tumor cells versus immune cells in the tumor microenvironment is a matter of controversy. Expression of PD-L1, CD39, CD73, and A2AR were increased in immune cells in the present study, while some tumor cells showed positive expression of A2AR. Although the different roles of the expression of PD-L1, CD39, CD73, and A2AR in tumor and immune cells were unclear, our findings suggested that the influence of the A2AR pathway and the PD-1/PD-L1 axis on the interaction between the immune system and tumor cells in the tumor microenvironment might be linked with cancer progression in RCC.

Patients who are resistant or refractory to anti–PD-1/PD-L1 antibodies or who have predominantly PD-L1–negative tumors might not harbor immune-suppressed characteristics through the PD-1/PD-L1 axis [3, 4]. In the present
Table 1  Relationship between PD-L1/CD39/CD73/A2AR status and clinicopathological characteristics

|                  | Grade | pT 1,2 (n=12) | pT 3,4 (n=48) | pN 1,2 (n=6) | pN 3,4 (n=54) | Metastatic organs* | anti-VEGF | ICI | anti-VEGF+/− ICI |
|------------------|-------|---------------|---------------|-------------|-------------|-------------------|-----------|-----|----------------|
|                  |       | (n=12)        | (n=48)        | (n=6)       | (n=54)      | Pul               | Pul/Oth   | Oth | PR/SD          |
| Grade            | 1,2   | 6             | 8             | 3           | 2           | 2                 | 0         | 10  | 2              |
|                  | 3,4   | 8             | 18            | 15          | 21          | 11                | 18        | 24  | 24             |
| p value          |       | 0.0449        | 0.0210        | 0.0635      | 0.0518      |                   |           |     |                |
| pT               | 1,2   | 2             | 6             | 3           | 1           | 6                 | 3         | 0   | 5              |
|                  | 3,4   | 12            | 20            | 22          | 17          | 10                | 18        | 25  | 25             |
| p value          |       | 0.7059        | 0.3532        | 0.0635      | 0.2206      |                   |           |     |                |
| pN               | 0     | 13            | 22            | 12          | 14          | 12                | 9         | 12  | 12             |
|                  | 1,2   | 1             | 4             | 16          | 4           | 4                 | 9         | 7   | 14             |
| p value          |       | 0.0024        |               |             |             |                   |           |     |                |
| Metastatic organs| Pul   | 14            | 10            | 0           | 14          | 14                | 15        | 17  |                |
|                  | Pul/Oth | 10            | 11            | 6           | 15          | 15                | 13        | 5   | 13             |
|                  | Oth    | 2             | 13            | 11          | 14          | 14                | 9         | 4   | 19             |
| p value          |       | <0.0001       | 0.0306        | 0.0002      |             |                   |           |     |                |
| PD-L1            | low   | 7             | 12            | 3           | 13          | 14                | 16        | 4   | 14             |
|                  | high  | 5             | 4             | 24          | 19          | 9                 | 15        | 4   | 14             |
| p value          |       | 0.7560        | 0.4039        | 0.0768      | 0.1488      |                   |           |     |                |
| CD39             | low   | 6             | 12            | 2           | 21          | 15                | 8         | 6   | 5              |
|                  | high  | 6             | 15            | 4           | 33          | 24                | 13        | 8   | 16             |
| p value          |       | 0.5081        | 0.7906        | 0.9778      | 0.5455      |                   |           |     |                |
| CD73             | low   | 9             | 12            | 5           | 39          | 13                | 19        | 13  | 15             |
|                  | high  | 3             | 12            | 1           | 15          | 8                 | 8         | 2   | 9              |
| p value          |       | 0.8839        | 0.5593        | 0.2203      | 0.4634      |                   |           |     |                |
| A2AR             | low   | 9             | 13            | 4           | 31          | 26                | 9         | 13  | 18             |
|                  | high  | 3             | 13            | 2           | 23          | 13                | 12        | 1   | 10             |
| p value          |       | 0.9208        | 0.6625        | 0.0744      | 0.0002      |                   |           |     |                |
| PD-L1 + A2AR     | low-low | 3              | 9             | 0           | 13          | 4                 | 4         | 4   | 5              |
|                  | low-high | 4             | 14            | 2           | 16          | 10                | 8         | 1   | 7              |
|                  | high-low | 4           | 14            | 4           | 18          | 17                | 5         | 9   | 13             |
|                  | high-high | 1       | 14            | 6           | 0           | 7                 | 3         | 4   | 0              |
| p value          |       | 0.9561        | 0.2743        | 0.3075      | 0.0021      |                   |           |     |                |

Pul lung metastasis only, Pul/Oth; metastases to lung and other organs, Oth other than lung metastases

Note: Metastatic organs* Pul lung metastasis only, Pul/Oth; metastases to lung and other organs, Oth other than lung metastases

*Grade 1,2 (n=12) Pul/Oth, Grade 3,4 (n=48) Pul/Oth, Grade 1,2 (n=6) Pul/Oth, Grade 3,4 (n=54) Pul/Oth, Grade 0 (n=39) Pul/Oth, Grade 1,2 (n=21) Pul/Oth

p value for grade, pT, pN, metastatic organs, anti-VEGF, ICI, anti-VEGF+/− ICI

| PD-L1 + A2AR | low-low | 3 | 10 | 0 | 13 | 9 | 4 | 4 | 5 | 4 | 9 | 4 | 2 | 2 | 9 | 4 |
|              | low-high | 4 | 14 | 2 | 16 | 10 | 8 | 1 | 7 | 10 | 0 | 9 | 3 | 12 | 6 | 12 |
|              | high-low | 4 | 18 | 4 | 18 | 17 | 5 | 9 | 13 | 0 | 17 | 6 | 7 | 0 | 18 | 4 |
|              | high-high | 1 | 6 | 0 | 7 | 3 | 4 | 0 | 3 | 4 | 0 | 4 | 1 | 4 | 1 | 6 |
| p value      |        | 0.9561 | 0.2743 | 0.3075 | 0.0021 | 0.0012 | 0.0009 | 0.0014 |
study, patients with higher expression of PD-L1 in the primary tumor showed a favorable response to ICIs regardless of the effects of previous anti-VEGF agents; however, they did not have a longer survival. In contrast, patients with higher expression of A2AR showed poorer response not only to ICIs but also to anti-VEGF agents and had a shorter
survival, regardless of the expression levels of PD-L1. The biological characteristics of primary and metastatic tumors are known to not always be identical [21]. In the present study, metastatic lesions were resected in only ten cases. Nevertheless, we found that the primary renal tumors with a higher expression of A2AR had metastatic lesions with a higher expression of A2AR, but we did not find such a positive relation for PD-L1, CD39, or CD73. The nonresectable metastatic tumors in patients with a higher expression of A2AR in the resected metastatic tumor showed worse response, regardless of the expression level of PD-L1 in the resected metastatic tumor. In the present study, 21 patients with regional lymph node metastasis showed a worse treatment response for other distant nonresectable metastatic lesions. This might be linked with the following findings based on three resected metastatic lymph nodes from three patients. Two of these patients had higher A2AR expressions in the resected metastatic lymph nodes and the primary renal tumor. They showed a worse response of the nonresectable distant metastatic lesions to anti-VEGF agents or ICIs, regardless of the expression level of PD-L1. The third patient had a lower expression of A2AR in both the metastatic lymph node and the primary renal tumor, and showed better treatment response. These findings indicate that even if metastatic lesions in patients with a higher expression of PD-L1 in the primary tumors or metastatic tumors or both show better response to ICIs in the short term, the prognosis for survival might be worse than we expected. The findings also highlight the impact of A2AR on cancer progression in RCC. In fact, in a recent phase I clinical trial a small-molecule A2AR antagonist safely blocked adenosine signaling in a cohort of 68 patients with RCC refractory to PD-1/PD-L1 inhibitors [18]. Moreover, treatment with anti–PD-1 has been reported to lead to increased expression of A2AR and CD73 and to be associated with enhanced tumor responses to A2AR blockade [16]. Accordingly, cancer cells are likely to exploit the A2AR pathway to promote metastasis and progression, and an increased expression of A2AR not only in the primary tumor but also in metastatic lesions probably has an anti-immune effect that enables cancer cells to survive and develop resistance to anti-tumor agents. Because many of the primary tumors in our sample were not well histologically differentiated (48/60 tumors) and were locally invasive (54/60 tumors), future studies should study the expression of PD-L1, CD39, CD73, and A2AR in primary tumors with well-differentiated, non-invasive, and non-metastatic profiles and in distant metastatic lesions to understand the differences in tumor microenvironments between primary tumors and their metastatic lesions. A diverse expression pattern of PD-L1, CD39, CD73, and A2AR was found not only in each patient but also in inter-individual metastatic lesions. If possible, it may be useful to study the expression of these molecules in individual metastatic lesions and compare their expression to examine the immunological difference in the tumor microenvironments and to determine individual patient treatment strategies in the future.

Both the hypoxia-inducible factor (HIF)-VEGF pathway and PD-1/PD-L1 axis play essential roles in the progression of RCC. A study performed in an effort to develop a tolerable and more effective combination regimen of an anti-VEGF agent and an anti–PD-1 antibody found improved outcome [22, 23]. Our findings that increased expression of A2AR correlated with poorer response to anti-VEGF agents, as well as ICIs, indicates that in RCC A2AR might be associated with angiogenesis by an unknown mechanism. Several lines of studies have suggested that adenosine stimulate VEGF production by immune cells through the stimulation of A2AR [14]. Thus, it is likely that the immunosuppressive A2AR pathway influences angiogenesis. Furthermore, at present we do not know whether A2AR-related resistance to anti-VEGF agents and ICIs exists at the time of tumor diagnosis or evolves as a resistance mechanism during the course of treatment. The mechanisms by which blocking of the VEGF pathway and PD-1/PD-L1 axis may influence the biological profiles of PD-1, PD-L1, CD39, CD73, and A2AR in the tumor microenvironment are also unclear. Future research should study the molecules of the HIF-VEGF, PD-1/PD-L1, and A2AR pathways in RCCs to investigate the spatial and temporal biological changes in the primary and metastatic lesions before and after treatment.

In the present study, we found CD8-positive T cells in many tumor microenvironments, but expression of CD8 did not correlate with pathological status, response to anti-VEGF agents or ICIs, or survival. Antigen-specific CD8 T cells are known to play an important role in controlling cancer, but persistent antigen stimulation results in T-cell exhaustion, suggesting that some T cells are exhausted or inhibited after chronic antigen stimulation or metabolic reprogramming [24–28]. Exhausted CD8 T cells have decreased effector function and proliferative capacity, partly caused by overexpression of inhibitory receptors such as PD-1 [24–28]. Blockade of PD-1 reinvigorates T cell responses. Thus, elucidating roles of T-cell exhaustion in tumor microenvironments may be needed to understand the immune-suppressive mechanisms of tumors and develop immunotherapeutic interventions. In contrast, a recent phase I clinical trial with an A2AR antagonist found that long-term clinical benefit is associated with increased recruitment of CD8-positive T cells into the tumor [18].

As part of the present study, five patients with metastatic RCCs who did not undertake cytoreductive nephrectomy were treated with a combination of anti-CTLA4 and anti–PD-1 antibodies as the first-line therapy. In two patients with lower A2AR expression in the core-needle biopsies from the primary renal tumors, both the primary renal tumors and the metastatic lesions showed better response,
whereas two of three patients with higher A2AR expression showed a poorer response. However, CD8-positive T cells in the tumor microenvironment were not related to treatment response. Thus, although the expression levels of CD8 had no association with histological grade, local invasion, regional lymph node involvement, or the effects of treatment, future studies should examine immune effector cytotoxic CD8-positive T cells and immune-suppression exhausted CD8 T cells before and after treatment with anti-VEGF agents and ICIs.

Adenosine signaling has been reported not only to directly dampen T-cell immunity but also to shift the balance away from T effector responses towards myeloid suppressor recruitment [9–11]. Furthermore, adenosine has been reported to drive naive CD4 T cells toward a regulatory phenotype, CD4-positive (CD4 + CD25 + Foxp3 +) regulatory T cells (Tregs), via induction of FoxP3, which directs macrophage differentiation towards an M2 phenotype, inhibits natural killer cell function and diminishes antigen presentation by dendritic cells [11, 29]. Tregs infiltrate tumors, and tumor cells that express CD39 and CD73 at high levels produce adenosine and suppress the tumor immune response through A2AR of effector T cells and natural killer cells. Adenosine produced by Tregs activates the A2AR on CD4 + CD25 effector T cells and suppresses the immune response [30]. On the other hand, anti-VEGF therapy can also decrease Treg, either by inhibiting accumulation of myeloid-derived suppressor cell (MDSC) and immature dendritic cell (DC) in the tumor microenvironment or directly through VEGF pathway inhibition on Treg [31–33]. MDSCs are involved with the differentiation of antigen-presenting cells (APCs), such as macrophages and DCs. MDSCs, Tregs, DCs, and tumor-associated macrophages (TAMs) may enhance the tumor-promoting immune response and control cancer growth [34]. Therefore, further research is needed on the expression of A2AR in Treg, MDSCs, DCs, and TAMs to shed light on the tumor-promoting immune response of A2AR in these immune cells infiltrating tumor tissues and also to explore the interrelations between them in the tumor microenvironment.

RCCs have unique features that make them attractive for therapeutic approaches targeting components of the immune system. In the present study, patients with increased A2AR expression in the primary lesion showed worse response to anti-VEGF agents and ICIs and shorter survival. Although the spatial and temporal molecular biological differences between the primary and metastatic lesions are unknown, our findings might highlight the therapeutic potential of blocking adenosine-mediated immunosuppression to strengthen anti-tumor immunity. To elucidate how the CD39/CD73/A2AR pathway and PD-L1/PD-L1 axis interact in immunosuppressive signaling in RCC, we need to study the mechanisms underlying anti-tumor immunity, including immune cell infiltration and angiogenesis. Without such information, we cannot effectively utilize PD-L1- or A2AR-mediated immunosuppression for cancer immunotherapy.

The limitations of the present study were its relatively small patient population, and short follow-up period, which did not allow us to draw definite conclusions from the results. As described above, another limitation was the lack of any mechanistic insight into how A2AR enhances anti-tumor immunity and the mechanism of suppressing T cell immunity through extracellular adenosine production and after binding to A2AR. T cells are activated by dendritic cells presenting non-self antigens, and the activated T cells migrate to target organs and damage tissues [3, 4]. Future studies should investigate the pharmacokinetics of adenosine, AMP, cyclic AMP, ADP, and ATP in tumor cells and immune cells to understand how the CD39/CD73/A2AR pathway in the tumor microenvironment plays a role in how cancer cells survive and escape the anti-tumor immune response. On the other hand, since tumor-related inflammation plays a vital role in the development and progression of cancers [35, 36], we should study the relationship between expression of CD39/CD73/A2AR and serum inflammatory markers, including the neutrophil-to-lymphocyte ratio (NLR), the platelet-to-lymphocyte ratio (PLR), the monocyte-to-lymphocyte ratio (MLR), C-reactive protein, and the systemic inflammation response index (SIRI). We studied only 10 cases of non-ccRCC, so future studies should also examine the roles of CD39/CD73/A2AR in papillary, chromophobe, and collecting duct RCC in larger cohorts. In addition, in this study we evaluated the association only of adenosine with RCC, so future studies should also evaluate the roles of other nucleotides, such as guanine and pyrimidine nucleosides, in RCC. The results of such studies might be able to shed light on immunosuppressive signaling in human RCC and on how to select patients with tumors that are immune suppressed through the A2AR axis and who might respond to therapeutic blockade of this axis.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval This study was conducted according to the tenets of the Declaration of Helsinki and was approved by the institutional ethics review board of Dokkyo Medical University Hospital (approval no. R-31-10 J).

Informed consent Before surgery, written consent regarding the use of surgical samples fixed in buffered formalin and medical records, as well as data publication was requested, and protection of patient privacy in future studies was guaranteed through a signed consent form approved by our institutional Committee on Human Rights in Research. Consequently, each patient in this cohort signed the consent form approved by our institutional Committee on Human Rights in Research prior to nephrectomy. Furthermore, surviving patients provided written consent for analysis of PD-L1, CD39, CD73, and A2AR of the surgical samples, through a signed form approved by our institutional Committee on Human Rights in Research (approval no. R-31-10 J). In cases of patient death, consent for publication was sought orally from next of kin of the participant. All samples were anonymized before analysis to protect patient privacy.

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References

1. Bergers G, Hanahan D (2008) Modes of resistance to anti-angiogenic therapy. Nat Rev Cancer 8:592–603. https://doi.org/10.1038/nrc2442
2. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144:646–674. https://doi.org/10.1016/j.cell.2011.02.013
3. Pardoll DM (2012) The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 12:252–264. https://doi.org/10.1038/ncr3239
4. Topalian SL, Drake CG, Pardoll DM (2015) Immune checkpoint blockade: a common denominator approach to cancer therapy. Cancer Cell 27:450–461. https://doi.org/10.1016/j.ccell.2015.03.001
5. Motzer RJ, Escudier B, McDermott DF, George S, Hammers HJ, Srinivas S, Tykodi SS, Somson JA, Procopio G, Plimack ER, Castellano D, Choueiri TK, Gurney H, Hammers D, Bono P, Wagstaff J, Gauler TC, Ueda T, Tomita Y, Schutz FA, Kollmannsberger C, Larkin J, Ravaud A, Simon JS, Xu LA, Waxman IM, Sharma P, CheckMate 025 Investigators (2015) Nivolumab versus everolimus in advanced renal-cell carcinoma. N Engl J Med 373:1803–1813. https://doi.org/10.1056/NEJMoa1510665
6. Motzer RJ, Rini BI, McDermott DF, Arén Frontera O, Hammers HJ, Carducci MA, Salama P, Escudier B, Beuselinck B, Amin A, Porta C, George S, Neiman V, Bracarda S, Tykodi SS, Barthélemy P, Leibowitz-Amit R, Plimack ER, Oosting SF, Redman B, Meliach B, Powles T, Nathan P, Oudard S, Pook D, Choueiri TK, Donskov F, Grimm MO, Gurney H, Heng DYC, Kollmannsberger CK, Harrison MR, Tomita Y, Duran I, Grünewald V, McHenry MB, Mekan S, Tannir NM; CheckMate 214 investigators (2019) Nivolumab plus ipilimumab versus nivolumab in first-line treatment for advanced renal cell carcinoma: extended follow-up of efficacy and safety results from a randomised, controlled, phase 3 trial. Lancet Oncol 20:1370–1385. https://doi.org/10.1016/S1470-2045(19)30413-9
7. Ohta A, Gorelik E, Prasad SJ, Ronchese F, Lukashev D, Wong MK, Huang X, Caldwell S, Liu K, Smith P, Chen JF, Jackson EK, Apasov S, Abrams S (2006) A2A adenosine receptor protects tumors from antitumor T cells. Proc Natl Acad Sci U S A 103:13132–13137. https://doi.org/10.1073/pnas.0605251103
8. Sitkovsky MV, Hatfield S, Abbott R, Belikoff B, Lukashev D, Ohta A (2014) Hostile, hypoxia-A2-adenosinergic tumor biology as the next barrier to overcome for tumor immunologists. Cancer Immunol Res 2:598–605. https://doi.org/10.1158/2326-6066.CIR-14-0075
9. Young A, Mittal D, Stagg SMJ (2014) Targeting cancer-derived adenosine: new therapeutic approaches. Cancer Discov 4:879–880. https://doi.org/10.1158/2159-8290.CD-14-0341
10. Allard B, Beavis PA, Darcy PK, Stagg L (2016) Immunosuppressive activities of adenosine in cancer. Curr Opin Pharmacol 29:7–16. https://doi.org/10.1016/j.coph.2016.04.001
11. Vijayan D, Young A, Teng MWL, Smyth MJ (2017) Targeting immunosuppressive adenosine in cancer. Nat Rev Cancer 7:709–724. https://doi.org/10.1038/nrc2017.86
12. Idzko M, Ferrari D, Eltzschig HK (2014) Nucleotide signaling during inflammation. Nature 509:310–317. https://doi.org/10.1038/nature13085
13. Junger WG (2011) Immune cell regulation by autocrine purinergic signaling. Nat Rev Immunol 11:201–212. doi 10.1038/nni2938
14. Antonioli L, Blandizzi C, Pacher P, Hasgö B (2013) Immunity, inflammation and cancer: a leading role for adenosine. Nat Rev Cancer 13:842–857. https://doi.org/10.1038/nrc3613
15. Allard B, Pommey S, Smyth MJ, Stagg J (2013) Targeting CD73 enhances the antitumor activity of anti-PD-1 and anti-CTLA-4 mAbs. Clin Cancer Res 19:5626–5635. https://doi. org/10.1158/1078-0432.CCR-13-0545
16. Beavis PA, Milenkovski N, Henderson MA, John LB, Allard B, Loi S, Kershaw MH, Stagg J, Darcy PK (2015) Adenosine receptor 2A blockade increases the efficacy of anti-PD-1 through enhanced antitumor T-cell responses. Cancer Immunol Res 3:506–517. https://doi.org/10.1158/2326-6066.CIR-14-0211
17. Yu Y, Wang W, Song L, Hu W, Dong C, Pei H, Zhou G, Yue Z (2015) Ecto-5′-nucleotidase expression is associated with the progression of renal cell carcinoma. Oncol Lett 9:2485–2494. https://doi.org/10.3892/ol.2015.3138
18. Fong L, Hotson A, Powderly JD, Szoln M, Heist RS, Choueiri TK, George S, Hughes BGM, Hellmann MD, Shepard DR, Rini BI, Kummard S, Weise AM, Riese MJ, Markham B, Emens LA, Mahadevan D, Luke JJ, Laport G, Brody JD, Hernandez-Aya L, Bonomi P, Goldman JW, Berim L, Renouf DJ, Goodwin RA, Munneke B, Ho P, Hsieh J, McCaffrey I, Kwei L, Willingham SB, Miller RA (2020) Adenosine 2A receptor blockade as an immunotherapy for treatment-refractory renal cell cancer. Cancer Discov 10:40–53. https://doi.org/10.1158/2159-8290.CD-19-0980
19. Nuku A, Kamai T, Araki K, Kijima T, Koyama T, Iikeya T, Hara I, Yoshida KI (2020) Association of cancer progression with elevated expression of programmed cell death protein 1 ligand 1 by upper tract urothelial carcinoma and increased tumor-infiltrating lymphocytes. Cancer Immunol Immunother (2021) 70:2009–2021
lymphocyte density. Cancer Immunol Immunother 69:689–702. https://doi.org/10.1007/s00262-020-02499-7

20. Rittmeyer A, Barlesi F, Waterkamp D, Park K, Ciardiello F, von Pawel J, Gadgeel SM, Hida T, Kowalski DM, Dols M, Cortinovis DL, Leach J, Polikoff J, Barrios C, Kabbainavar F, Frontera OA, De Marinis F, Turna H, Lee JS, Ballinger M, Polikoff J, Barrios C, Kabbinavar F, Frontera OA, De Marinis F, Turna H, Lee JS, Ballinger M, Kowanetz M, He P, Chen DS, Sandler A, Gandara DR; OAK Study Group (2017) Atezolizumab versus docetaxel in patients with previously treated non-small-cell lung cancer (OAK): a phase 3, open-label, multicentre randomised controlled trial. Lancet 389:255–265. https://doi.org/10.1016/S0140-6736(16)32517-X

21. Gerlinger M, Rowan AJ, Horswill S, Larkin J, Endesfelder D, Gronroos E, Martinez P, Matthews N, Stewart A, Tarpey P, Varela I, Phillimore B, Begum S, McDonald NQ, Butler A, Jones D, Raine K, Latimer C, Santos CR, Nohadani M, Eklund AC, Spencer-Dene B, Clark G, Pickering L, Stamp G, Gore M, Szallasi Z, Downward J, Futreal PA, Swanton C (2012) Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. New Engl J Med 366:883–892. https://doi.org/10.1056/NEJMoa1113205

22. Motzer RJ, Penkov K, Haanen JL, Rini BI, Plimack ER, Stus V, Gafanov R, Hawkins R, Nosov D, Poulot F, Alekseev B, Soulères D, Melichar B, Vynnychenko I, Bondarenko I, Azevedo SJ, Borchiellini D, Szczyluk C, Markus M, McDermott RS, Bedke J, Tartas S, Chang YH, Tamada S, Shou Q, Perini RF, Chen M, Atkins MB, Powles T; KEYNOTE-426 Investigators (2019) Pembrolizumab plus axitinib versus sunitinib for advanced renal-cell carcinoma. N Engl J Med 380:1116–1127. https://doi.org/10.1056/NEJMoa1816714

23. Rini BI, Plimack ER, Stus V, Gafanov R, Hawkins R, Nosov D, Poulot F, Alekseev B, Soulères D, Melichar B, Vynnychenko I, Kryzhanivska A, Bondarenko I, Azevedo SJ, Borchiellini D, Szczyluk C, Markus M, McDermott RS, Bedke J, Tartas S, Chang YH, Tamada S, Shou Q, Perini RF, Chen M, Atkins MB, Powles T; KEYNOTE-426 Investigators (2019) Pembrolizumab plus axitinib versus sunitinib for advanced renal-cell carcinoma. N Engl J Med 380:1116–1127. https://doi.org/10.1056/NEJMoa1816714

24. Joyce JA, Fearon DT (2015) T cell exclusion, immune privilege, and the tumor microenvironment. Science 348:74–80. https://doi.org/10.1126/science.aab6204

25. Sowell RT, Kaech SM (2016) Probing the diversity of T cell dysfunction in cancer. Cell 166:1362–1364. https://doi.org/10.1016/j.cell.2016.08.058

26. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A (2017) Primary, adaptive, and acquired resistance to cancer immunotherapy. Cell 168:707–723. https://doi.org/10.1016/j.cell.2017.01.017

27. Thommen DS, Schumacher TN (2018) T cell dysfunction in cancer. Cancer Cell 33:547–562. https://doi.org/10.1016/j.ccell.2018.03.012

28. Blank CU, Haining WN, Held W, Hogan PG, Kallies A, Lugli A, Lynn RC, Philip M, Rao A, Restifo NP, Schietinger A, Schumacher TN, Schwartzberg PL, Sharpe AH, Speiser DE, Wherry EJ, Youngblood BA, Zehn D (2019) Defining T cell exhaustion. Nat Rev Immunol 19:665–674. https://doi.org/10.1038/s41577-019-0221-9

29. Willingham SB, Ho PY, Hotson A, Hill C, Piccione EC, Hsieh J, Liu L, Buggy JJ, McCallery I, Miller RA (2018) A2AR antagonism with CPI-444 induces antitumor responses and augments efficacy to anti-PD-(L)1 and anti-CTLA-4 in preclinical models. Cancer Immunol Res 6:1136–1149. https://doi.org/10.1158/2326-6066.CIR-18-0056

30. Deaglio S, Dwyer KM, Gao W, Friedman D, Ushva A, Erat A, Chen JF, Enjoji Y, Linden J, Ouuka M, Kurkoo VK, Strom TB, Robson SC (2007) Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. J Exp Med 204:1257–1265. https://doi.org/10.1084/jem.20062512

31. Voron T, Marcheteau E, Pernot S, Colussi O, Tartour E, Taieb J, Termé M (2014) Control of the immune response by proangiogenic factors. Front Oncol 4:70. https://doi.org/10.3389/fonc.2014.00070

32. Lanitis E, Irving M, Coukos G (2015) Targeting the tumor vasculature to enhance T cell activity. Curr Opin Immunol 33:55–63. https://doi.org/10.1016/j.coi.2015.01.011

33. Lapeyre-Prost A, Termé M, Pernot S, Pointet AL, Voron T, Tartour E, Taieb J (2017) Immunomodulatory activity of VEGF in cancer. Int Rev Cell Mol Biol. 330:295–342. https://doi.org/10.1016/bs.ircmb.2016.09.007

34. Gabrilovich DI, Nagaraj S (2009) Myeloid-derived suppressor cells as regulators of the immune system. Nature Rev Immunol 9:162–174. https://doi.org/10.1038/nri2506

35. Diakos CI, Charles KA, McMillan DC, Clarke SJ (2014) Cancer-related inflammation and treatment effectiveness. Lancet Oncol 15:e493-503. https://doi.org/10.1016/S1470-2045(14)70263-3

36. Mantovani A, Allavena P, Sica A, Balkwill F (2008) Cancer-related inflammation. Nature 454:436–444. https://doi.org/10.1038/nature07205

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