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Precision Medicine for Neonatal Sepsis

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Neonatal sepsis remains a significant cause of morbidity and mortality especially in the preterm infant population. The ability to promptly and accurately diagnose neonatal sepsis based on clinical evaluation and laboratory blood tests remains challenging. Advances in high-throughput molecular technologies have increased investigations into the utility of transcriptomic, proteomic and metabolomic approaches as diagnostic tools for neonatal sepsis. A systems-level understanding of neonatal sepsis, obtained by using omics-based technologies (at the transcriptome, proteome or metabolome level), may lead to new diagnostic tools for neonatal sepsis. In particular, recent omic-based studies have identified distinct transcriptional signatures and metabolic or proteomic biomarkers associated with sepsis. Despite the emerging need for a systems biology approach, future studies have to address the challenges of integrating multi-omic data with laboratory and clinical meta-data in order to translate outcomes into precision medicine for neonatal sepsis. Omics-based analytical approaches may advance diagnostic tools for neonatal sepsis. More research is needed to validate the recent systems biology findings in order to integrate multi-dimensional data (clinical, laboratory and multi-omic) for future translation into precision medicine for neonatal sepsis. This review will discuss the possible applications of omics-based analyses for identification of new biomarkers and diagnostic signatures for neonatal sepsis, focusing on the immune-compromised preterm infant and considerations for clinical translation.

Keywords: systems biology, diagnosis, infection, neonate, sepsis, preterm infant

INTRODUCTION

Neonatal sepsis, a bacterial bloodstream infection associated with inflammation and life-threatening organ dysfunction, is classified as early-onset sepsis (EOS, <72 h after birth) or late-onset sepsis (LOS, >72 h) (Bateman and Seed, 2010). Prompt and accurate diagnosis based on clinical and laboratory findings remains challenging. The complex and dynamic disease pathophysiology often results in clinical signs that are subtle, non-specific and overlap with non-infectious conditions (Camacho-Gonzalez et al., 2013). Consequently, there is no consensus definition for neonatal sepsis (Wynn, 2016). Further, the current "gold standard" test, microbiological culture, lacks sensitivity and has slow turnaround times (24–48 h). Adjunct tests such as hematological indices and inflammatory markers either have poor sensitivity and specificity or require serial measurements (Delanghe and Speeckaert, 2015). Simple, rapid and accurate
diagnostic tests that can guide treatment of septic infants are urgently needed (Dong and Speer, 2015). Markers with high negative predictive value may allow empiric antibiotic treatment of uninfected infants to be withheld and reduce unnecessary antibiotic exposure associated with increased adverse short- and long-term outcomes (Kuppala et al., 2011; Arboleya et al., 2015).

Neonatal sepsis pathophysiology involves multiple organ systems; highlighting the need for a systems biology approach to capture the complex interactions between biological systems during disease (Smith et al., 2014; Alyass et al., 2015). Advances in genomics, transcriptomics, proteomics and metabolomics inform us of the genetic predispositions to sepsis; transcriptional changes in host responses during sepsis; protein expression altered by sepsis; and metabolites produced as a result of sepsis (Fanos et al., 2013). Integrating findings from these tools into our understanding of disease pathophysiology will enable future translation into precision medicine, where patients are identified and treated based on genetic, cellular and molecular markers that relate to the underlying causes of their disease instead of common phenotypic signs of sepsis (Flores et al., 2013; van Karnebeek et al., 2018).

This review discusses the potential of current “omics” approaches to characterize sepsis pathophysiology and allow the discovery of new biomarkers and neonatal sepsis signatures. The necessary considerations for translating these omics-based approaches from bench-to-bedside are also considered.

**Transcriptomics of the Host Response to Sepsis**

Changes in hematological markers during bacterial infections, such as in immature-to-total neutrophil ratios and white blood cell (WBC) counts, have been used for decades as adjunct tests for neonatal sepsis diagnosis. However, the clinical utility of tests remains limited by wide ranges of specificity (31–100%) and sensitivity (17–90%), especially early in sepsis onset, and by the considerable influence of common variables such as gestational and postnatal age (Schmutz et al., 2008; Chirico and Loda, 2011; Polin, 2012; Sharma et al., 2017).

The introduction of microarrays and next-generation sequencing (NGS) technologies, particularly RNA-Sequencing (RNA-Seq), has transformed our ability to monitor gene-expression changes occurring at cellular level during sepsis. Transcriptional profiling allows simultaneous measurement of expression levels of thousands of genes, where differentially expressed genes associated with sepsis could lead to the discovery of novel cell-specific gene signatures for early and accurate diagnosis of septic infants. Additionally, visualization of gene interaction networks and identification of enriched pathways associated with disease could improve our understanding of the relationship between innate, adaptive and metabolic responses during neonatal sepsis (Chaussabel et al., 2010; Skibsted et al., 2013; Smith et al., 2014; Xia et al., 2015). This section focuses on transcriptomic studies in the neonatal population (Table 1).

Smith et al. analyzed blood samples from preterm and term infants taken when investigated for suspected infection using microarray. The study identified a 52-gene network comprising of genes from innate, adaptive and metabolic pathways that could distinguish bacterial infections from uninfected infants with 98% accuracy. This combined immune-metabolic network performed better compared to individual gene sets (65–84%) (Smith et al., 2014). Cernada et al showed that microarray-based gene expression profiling could discriminate between very low birth-weight infants with bacterial sepsis and controls, with good overall sensitivity (100%) but lower specificity (68%). The differences observed between septic and non-septic controls were associated with 554 differentially expressed genes mainly linked to tumor necrosis factor and cytokine signaling (Cernada et al., 2014). A microarray-based study by Wynn et al. demonstrated that infants with EOS or LOS had different transcriptomes to non-septic infants. However, early and late septic responses differed significantly and were associated with postnatal age at the time of sepsis. These findings underscore the importance of controlling for postnatal age in neonatal sepsis transcriptome studies (Wynn et al., 2015).

Recent transcriptomic studies have also explored the diagnostic and prognostic potential of micro-ribonucleic acids (miRNA) in neonatal sepsis (Table 1). miRNAs are more stable than messenger (m)RNA and increasing evidence supports their importance in sepsis pathophysiology and potential as sepsis markers (Wang et al., 2015; Inal et al., 2016).

Chen et al. identified 10 miRNAs significantly altered during neonatal sepsis in preterm and term infants using microarray, which were confirmed using quantitative real-time reverse transcription-polymerase chain reaction (qRT-PCR). The identified miRNAs were linked to genes and proteins involved in pathogen recognition, inflammation, immune cell activation, release of pro-inflammatory cytokines and apoptosis (Chen et al., 2014). Wang et al. assessed the utility of adult sepsis miRNA biomarkers (miR-15a/15b/16/223) using blood samples collected during septic screens from term neonates. qRT-PCR analysis showed only miR-15a and miR-16 was significantly up-regulated in neonatal sepsis patients compared to controls, with higher area under the curve (AUC) values compared to miR-15b and miR-223. Both miR-15a and miR-16 were found to play a pivotal role in regulating lipopolysaccharide (LPS)-induced inflammatory responses during sepsis (Wang et al., 2015). Separately, in LPS stimulated neonatal leukocytes derived from cord blood of infants delivered by healthy mothers, miRNA let-7b-5p expression was found to be significantly lower compared to adults (Yu et al., 2016).

The mRNA and miRNA studies reviewed observed transcriptional profile differences between infected and control cases that were consistently linked to functions of the innate immune system. Collectively, these studies demonstrate the potential of microarray-based approaches to identify new gene signatures for improved pathophysiological understanding of neonatal sepsis. DNA microarrays, though limited by the number of probes available, remain the most common method to determine transcriptional expression level changes during neonatal sepsis for both mRNA and miRNA (Skibsted et al., 2013; Cernada et al., 2014; Chen et al., 2014; Smith et al., 2014; Wynn et al., 2015).
| Reference          | Methodology                           | Study population and case definitions                                                                 | Type of sample | Principal findings                                                                                                                                                                                                 | Study limitations                                                                                                                                 |
|--------------------|---------------------------------------|----------------------------------------------------------------------------------------------------------------|----------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------|
| Smith et al., 2014 | Microarray with 48,802 human gene probes | 62 preterm and term neonates (24–42 weeks gestation); infected (n = 27) and controls (n = 35)                  | Peripheral blood | - Identified a 52-gene immune-metabolic network associated with sepsis  
- The individual innate, adaptive and metabolic pathway markers had accuracy of 84, 65, and 74%, respectively; combined use of three pathway markers had the highest accuracy of 98% for predicting bacterial infections in neonates | The robustness of the 52-gene immune-metabolic network for predicting bacterial infections not yet validated in large-scale studies across multiple neonatal units |
| Cernada et al., 2014 | Microarray with >28,000 human gene probes | 36 VLBW infants; septic (n = 17) and controls (n = 19) | Peripheral blood | - Genome-wide expression profiles could discriminate between septic infants and controls with 100% sensitivity and 68% specificity  
- Showed 554 genes were differentially expressed between neonates with bacterial sepsis and matched controls, with 66 genes associated with tumor necrosis factor and 56 genes with cytokine signaling | Limited number of patients in the study  
Authors consider study as "starting point to perform strongly powered, prospective collaborative studies in the neonatal population" |
| Wynn et al., 2015   | Microarray with 20,533 human gene probes | 68 preterm and term neonates (23–42 weeks gestation; with EOS (n = 6), LOS (n = 9), clinical sepsis (n = 22) and uninfected (n = 31) | Peripheral blood | - Identified significant differences in transcriptome of infants with EOS or LOS, and showing importance of accounting for timing of sepsis episode when investigating transcriptional profiles | Limited sample size especially for EOS and LOS groups  
- Whole blood analyses without cell typing. This limits examination of cell-specific gene expression |
### TABLE 1 | Continued

| Reference          | Methodology           | Study population and case definitions | Type of sample | Principal findings                                                                 | Study limitations                                                                 |
|--------------------|-----------------------|----------------------------------------|----------------|------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------|
| **Sepsis (EOS/LOS) cases** |                       | • Positive blood culture for Gram-positive or Gram-negative bacteria |                | **Principal findings**                                                             | Study limitations                                                                 |
|                    |                       | • Presence of persistent (>2 days) abnormal clinical signs such as ill appearing and respiratory or cardiovascular signs |                |                                                                                  | • Minimal mortality in cohort limited comparisons between survivors and nonsurvivors |
|                    |                       | • Presence of abnormal laboratory results showing systemic inflammation (e.g., CRP > 45 mg/L within 48 h of evaluation) |                |                                                                                  |                                                                                  |
|                    |                       | **Clinical sepsis cases**               | Peripheral blood | • Identified significant up-regulation of miR-101/122/185 and down-regulation of miR-96/182/141/143/181a/1184 in infants with neonatal sepsis |                                                                                  |
|                    |                       | • Negative blood culture                |                | **Uninfected cases**                                                              |                                                                                  |
|                    |                       | • Presence of persistent (>2 days) abnormal clinical signs such as ill appearing and respiratory or cardiovascular signs |                | • Identified up-regulated miR-15a and miR-16 in neonatal sepsis, with AUC values of 0.85 and 0.86, respectively, for neonatal sepsis diagnosis |                                                                                  |
|                    |                       | • Presence of abnormal laboratory results showing systemic inflammation (e.g., CRP > 45 mg/L within 48 h of evaluation) |                | • The miRNAs identified were not validated in large-scale studies in multiple neonatal units |                                                                                  |
| Chen et al., 2014  | miRNA microarray and qRT-PCR | 48 neonates; preterm (<37 weeks gestation, n = 5) and term (>37 weeks gestation, n = 43); with Gram-positive (n = 12), Gram-negative sepsis (n = 12) and uninfected (n = 24) | Peripheral blood |                                                                                  |                                                                                  |
| Wang et al., 2015  | qRT-PCR               | 87 term neonates; with sepsis (n = 46) and controls (n = 41) | Peripheral blood |                                                                                  |                                                                                  |

(Continued)
| Reference | Methodology | Study population and case definitions | Type of sample | Principal findings | Study limitations |
|-----------|-------------|----------------------------------------|----------------|-------------------|------------------|
| Controls | • Blood collected from patients with upper respiratory infection or pneumonia | | | | |
| Yu et al., 2016 | miRNA microarray and qRT-PCR | Neonates (n = 5) and adults (n = 5) for isolation of majority of leukocyte subpopulations; and neonates (n = 31) and adult (n = 19) for isolation of pDC | Cord blood | • Identified decreased miRNA let-7b-5p expression in cord blood leukocytes | • Neonatal leukocyte responses were compared to adults instead of age-matched controls, with a small neonatal sample size used particularly for isolation of leukocyte subpopulations |
| | | | | • Let-7b-5p miRNA inhibits LPS-induced IL-6 and TNF-α production in monocytes | |
| | | | | | • Only specific leukocyte populations were used for analyses instead of whole blood |
| | | | | | • The leukocyte responses were assessed using cord blood from neonates born to healthy mothers and may not be reflective of miRNA-mediated regulation of immune responses in neonates with culture-proven bacterial sepsis, the population most in need of improved sepsis management |
| Kim et al., 2015 | Magnetic multiplexed nano-biosensor platform for MMP-7 and EpCAM | 20 preterm neonates (24–35 weeks gestation); with sepsis (n = 5), NEC (n = 10) and controls (n = 5). | Peripheral blood plasma | • MMP7/EpCAM ratio has high diagnostic accuracy for differentiating infants with NEC or sepsis from control infants with AUC values of 1.00 and 0.96 respectively | • Pilot study with small sample size and no power analysis performed |
| | | Sepsis • Positive blood culture • Absence of uniform serum CRP elevation NEC • Pneumatosis intestinale in pathognomonic abdominal radiographic result | | | • Authors concede that controls may yield some false negatives for sepsis or NEC |
| | | Controls • Absence of sepsis and NEC | | | • The biosensor platform not yet validated in large-scale multicenter studies |
| Ng et al., 2010 | MALDI-TOF MS and protein chip arrays | Biomarker discovery cohort 74 very preterm neonates (<32 weeks gestation), with sepsis/NEC (n = 37) or non-sepsis (n = 37) | Peripheral blood plasma | • ApoSAA score capable of differentiating infants with sepsis and NEC from control infants | • Mix of bacterial and fungal infections precludes comparisons with studies containing only bacterial sepsis |
| | | Case-control validation cohort 80 neonates; with sepsis/NEC (n = 40) and non-sepsis (n = 40) | | | • The algorithm not yet evaluated in large-scale multicenter bacterial sepsis studies |
| | | Prospective validation cohort 104 preterm neonates (<35 weeks gestation); with sepsis/NEC (n = 42), probable clinical sepsis (n = 13) and non-sepsis (n = 49) | | | |
| | | Case-control validation cohort 80 neonates; with sepsis and NEC at a 0.75 cut-off value; with 90% sensitivity and 95% specificity | | | |
| | | Prospective validation cohort | | | |
| | | ApoSAA score at 0.75 cut-off value showed 84% specificity and 89% sensitivity | | |
| (Continued) | | | | | |
| Reference | Methodology | Study population and case definitions | Type of sample | Principal findings | Study limitations |
|-----------|-------------|---------------------------------------|----------------|--------------------|------------------|
| **Sepsis cases** | • Positive blood culture for bacterial or fungal infection | **NEC cases** | • Stage II or above in Bell’s classification | **Non-sepsis cases** | • Negative blood culture with definitive diagnosis unrelated to sepsis/NEC |
| **Probable clinical sepsis** | • Negative blood culture | • Presenting >3 clinical signs and symptoms | • Evidence of hematologic or metabolic derangement | • Strong circumstantial background of sepsis based on clinical course and laboratory results |
| Buhimschi et al., 2011 | **Discovery phase** Two-dimensional gel electrophoresis and MS | **Validation phase** Immunoassay, western blotting and latent-class analysis | 180 preterm neonates (<37 weeks gestation) total for discovery and validation phases | **Cord blood serum** | **Discovery phase** Identified 19 proteins involved in immunity and defense, protease/extracellular matrix, and transfer/carrier pathways |
| | Discovery phase | 6 neonates; with clinical EOS \((n=3)\) and gestational age-matched control \((n=3)\) | **Validation phase** 174 neonates; with clinical EOS \((n=45)\) and no clinical EOS \((n=129)\). | **Hp & HpRP significantly increased in EOS** | **Need to be validated in a large-scale multicenter cohort to ensure Hp & HpRP can be used across different neonatal units to improve EOS diagnosis** |
| | Clinical EOS cases | • Positive blood culture | • Cord blood IL-6 > 90 pg/mL |  |  |
| | | • Histological chorioamnionitis stages II (chorionic inflammation) or III (full-thickness inflammation of both amnion and chorion) | **Controls** |  |  |
| | | • Negative blood culture | • Cord blood IL-6 < 9 pg/mL |  |  |
| | | • No histological chorioamnionitis | **Validation phase** |  |  |
| | | **Clinical EOS** | 174 neonates; with clinical EOS \((n=45)\) and no clinical EOS \((n=129)\). |  |  |
| | | • Positive blood culture and/or positive hematological indices \((≥2\) of following: absolute neutrophil count of <7,500 or >14,500 cells/mm\(^3\); absolute band count >1,500 cells/mm\(^3\); immature/total neutrophil \((I:T)\) ratio >0.16; platelet count <150,000 cells/mm\(^3\)) | **No clinical EOS** | • Negative blood culture |  |
| | | **No clinical EOS** | • Negative hematological indices |  |  |
RNA-Seq is emerging as a powerful tool for transcriptome-wide profiling that is independent of pre-identified probe sequences, thus allowing discovery of novel gene transcripts to generate sepsis-related gene signatures (Chaussabel et al., 2010). Although there have been no published studies using RNA-Seq to identify transcriptional signatures for diagnosis of neonatal sepsis to date, the method has shown potential in adult studies. Pena et al. found that adult sepsis was associated with an endotoxin tolerance signature that was useful in differentiating true from suspected sepsis prior to clinical sepsis confirmation and organ dysfunction prognosis (Pena et al., 2014). RNA-Seq allows for hypothesis-free assessment of the transcriptome for unbiased discovery-based identification of gene signatures associated with neonatal sepsis. Presently, the feasibility of translating NGS-based approaches into bedside tools for neonatal sepsis remains unknown. The challenges faced are both downstream, where extensive sample processing is required for nucleic acid extraction and library preparation for sequencing; and upstream, data storage (up to 150GB per whole-genome sequencing experiment).

RNA-Seq also involves computationally intensive analysis pipelines requiring specialized bioinformatics skills. One possible solution to this is to reduce the dimensionality of the data obtained from patients, focusing only on the minimal identified gene signature required for adequate diagnostic performance (Costa, 2014; Pena et al., 2014; Smith et al., 2014; Tebani et al., 2016b). Such genes could be detected using more rapid approaches such as nanostring and qRT-PCR. Nanostring allows detection of up to 800 different transcripts in a single reaction and is not influenced by pipetting errors, nonspecific enzymatic reactions or reference gene instability, enabling more accurate measurements of gene expression. However, it is limited by the time involved from sample collection to quantitation (16–48 h) compared to qRT-PCR (within 2 h). qRT-PCR has been routinely used for validation of gene expression in microarray and NGS-based sepsis studies, showing promise for clinical translation in childhood leukemia studies and possibility of using a minimal identified gene signature for neonatal sepsis diagnosis in the clinical setting. This approach provides a cost-efficient, rapid, less computationally and bioinformatically intensive alternative to

### Table 1 Continued

| Reference | Methodology | Study population and case definitions | Type of sample | Principal findings | Study limitations |
|-----------|-------------|---------------------------------------|----------------|-------------------|-------------------|
| Mickiewicz et al., 2013 | NMR Spectroscopy | 140 patients; of which neonates (n = 7), infants (n = 47), toddlers serum (n = 54) and school age children (n = 32) had either septic shock (n = 60), SIRS (n = 40) or were healthy controls (n = 40) | Peripheral blood, serum, urine | Identified increased levels of lactate, glucose, creatinine, 2-oxoisocaproate, 2-hydroxyisovalerate and 2-hydroxybutyrate; and decreased threonine, acetate, 2-aminoenurate and adipate in sepsis | Small sample size of neonates, with no age-matched controls |
| | | | | | Mixed case definition for septic shock consisting of Gram-positive, Gram-negative and polymicrobial infection |
| | | | | | The metabolites identified need to be validated in large-scale multicenter studies |
| Fanos et al., 2014 | GC-MS and NMR | 25 neonates (≤35 weeks mean gestation); with sepsis (n = 9) and healthy controls (n = 16) | Urine | Identified increased concentrations of glucose, lactate and acetate; and decreased ribitol, ribonic acid, pseudouridine, 2,3,4-trihydroxybutyrate and 3,4,5-trihydroxyacetanilide acid in sepsis | No criteria listed for case definitions |
| | | | | | Small sample size with no power analysis conducted |
| | | | | | Identified biomarkers not yet validated in large-scale multicenter studies |

Apo, Apolipoprotein; AUC, area under the curve; CRP, C-reactive protein; EOS, early-onset sepsis; EpCam, epithelial cell adhesion molecule; GC-MS, gas-chromatography mass-spectrometry; Hp, haptoglobin; HpRP, haptoglobin-related protein; IL, interleukin; LOS, late-onset sepsis; LPS, lipopolysaccharide; MALDI-TOF, matrix assisted laser desorption ionization time of flight; miRNA, micro-ribonucleic acid; MMP, matrix metalloproteinase; MS, mass spectrometry; NEC, necrotizing enterocolitis; NMR, nuclear magnetic resonance; pDC, plasmocytoid dendritic cell; PRISM III-APS, pediatric risk of mortality III-acute physiology; qRT-PCR, quantitative reverse-transcriptase polymerase chain reaction; SAA, serum amyloid A; SIRS, systemic inflammatory response syndrome; TNF, tumor necrosis factor; VLBW, very low birth weight.
measure gene signature expression for neonatal sepsis diagnosis (Hoffmann et al., 2006; Chauvel et al., 2010; Chen et al., 2014; Tsang et al., 2017).

**Proteomics of Sepsis Samples**

Overall, no single protein biomarker has emerged with sufficient sensitivity, specificity and reproducibility to accurately diagnose sepsis. This has prompted the need to move from single protein markers, such as CRP or procalcitonin, to the use of panels of biomarkers to improve diagnostic performance (Ludwig and Hummon, 2017). Proteomic studies on body-fluids and tissues in both human adults and animal models assessing the mechanism of both adult and neonatal sepsis have previously been reviewed (Cao and Robinson, 2014; Delanghe and Speeckaert, 2015). This section focuses on recent proteomic studies aimed at identifying biomarkers for human neonatal sepsis.

Necrotizing enterocolitis (NEC) shares similar manifestations with neonatal sepsis, including increased CRP levels, making it difficult to distinguish from sepsis. Several proteins have been identified as potential candidates to differentiate both sepsis and/or NEC from uninfected neonates (Table 1). Kim et al. employed a multiplexed nano-biosensor proteomic platform on plasma samples from 20 preterm neonates to formulate a ratio of protein levels based on matrix metalloproteinase (MMP)-7 and epithelial cell adhesion molecule (EpCAM) concentrations. The MMP-7/EpCAM ratio differentiated NEC from sepsis and healthy controls with high diagnostic accuracy, although the study was limited by a small sample size and lack of information on how neonates were diagnosed for sepsis (Kim et al., 2015). Ng et al. assessed plasma samples from preterm infants with or without NEC/sepsis using ProteinChip array and matrix assisted laser desorption ionization-time of flight (MALDI-TOF) mass spectrometry (MS). The protein concentrations identified from diagnostic proteomic peaks were measured by immunoassay. Multivariate logistic regression analysis determined Apolipoprotein (Apo)C2 and serum amyloid A (SAA) as the most promising markers based on immunoassay concentrations, and were used to construct an ApoSAA score that was capable of differentiating sepsis/NEC cases from non-sepsis/NEC cases. Downstream validation in separate case-control and prospective cohort studies of preterm infants showed high diagnostic performance. Further confirmation in large multicenter trials will be necessary before the ApoSAA score can be translated for use to diagnosis sepsis/NEC cases (Ng et al., 2010). Separately, Buhimschi et al. profiled the serum proteome from venous cord blood of preterm neonates with and without EOS. Gel and MS-based proteomics identified 19 proteins with differential abundance between cases and controls. Downstream validation involving neonates with and without EOS showed that Haptoglobin (Hp) and Haptoglobin-related protein (HpRP) immunoreactivity were significantly elevated in both clinical and culture-confirmed EOS neonates, and that a combination of Hp & HpRP, IL-6 and neonatal hematological indices could improve the clinical EOS diagnosis (Buhimschi et al., 2011).

Together, these proteomic studies highlight the potential to use a panel of biomarkers for more effective diagnosis. Importantly, the use of MS-based technologies for proteomic profiling in these studies underscore the capacity to discover new protein biomarker combinations through a hypothesis-free unbiased approach (Ng et al., 2010; Buhimschi et al., 2011). Compared to traditional hypothesis-driven approaches, where proteins are pre-selected for analysis, MS-based omics can screen thousands of protein abundances and post-translational modifications in a single acquisition, allowing unprecedentedly wide coverage for discovery of novel biomarkers for panel construction. Proteomic biomarkers identified through this approach have potential to be translated into use on bench-top mass spectrometers with optimized and validated assays that are ready for clinical use, allowing timely analysis of multiple proteins in very low abundances close to the bedside (Honour, 2003; Rifai et al., 2006; Ludwig and Hummon, 2017). However, translating a MS-based approach for diagnosis of neonatal sepsis in the clinic has not been assessed. Instrumentation costs will need to be reduced and a standardized protocol for sample collection, preparation, processing, analysis and reporting that is not time-intensive (<12 h) needs to be developed before MS-based technologies can be feasibly utilized by the bedside (Tebani et al., 2016a). Multiplex immunoassays can be developed from identified multi-biomarker panels and performed using available technologies such as Luminex™. A point-of-care protein-microarray device for quantification of multiple serum proteins using minimal sample volume from neonates has previously been developed, supporting potential to translate validated multi-protein biomarker panels for bedside diagnosis (Buchegger et al., 2012; Tighe et al., 2015).

**Metabolic Phenotyping of Septic Infants**

Sepsis induces hypoxia, oxidative stress, and an increased demand for energy resulting in both glucose metabolism and oxidative metabolism of fatty acids; necessitating the monitoring of metabolome dysregulations during neonatal sepsis. Metabolomics can characterize thousands of intermediate to low molecular-weight carbohydrates, amino acids, lipids and other molecules generated by the interaction between host genome, the gut microbiome and environment. It is thus a useful tool to investigate metabolic perturbations related to neonatal sepsis for identification of novel biomarkers. The mainstream analytical technologies for metabolic profiling are nuclear magnetic resonance (NMR) and MS connected to capillary electrophoresis, gas chromatography (GC) or liquid chromatography (LC) separation methods. Whilst NMR enjoys relatively fast and straightforward metabolite annotation and preservation of samples, MS is more sensitive and can detect metabolites with lower abundance. Clinical and pharmaceutical applications of metabolomics in newborns and infants has been reviewed previously (Sumner et al., 2007; Antonucci et al., 2012; Dessi et al., 2014; Fanos et al., 2014). This section focuses on metabolomic studies in neonatal sepsis (Table 1).

Mickiewicz et al. used H-NMR analysis of serum samples from a mixed pediatric cohort consisting of neonates and children up to 11 years of age. Several metabolites, including lactate, glucose, adipate, 2-hydroxybutyrate and threonine were differentially regulated between septic and non-septic patients across age groups, and also distinguished those with systemic inflammatory
response syndrome (SIRS). The outcome models designed using orthogonal partial least square discriminant analysis (OPLS-DA) was predictive of mortality in pediatric patients with septic shock (Mickiewicz et al., 2013). Fanos et al. profiled the metabolome of non-invasively collected urine samples from preterm septic newborns and healthy controls using H-NMR and GC-MS, and found increased concentrations of several metabolites including glucose and lactate but decreased concentrations of metabolites including ribitol, pseudouridine and 2-ketogluconic acid. Urine samples from sepsis patients also had different levels of acetone, ketone bodies, likely due to the hypermetabolic responses occurring during sepsis. Interestingly, OPLS-DA of the samples showed a clear separation of control and sepsis samples, with ability to discriminate EOS from LOS (Fanos et al., 2014).

Overall, despite different sample types used, both metabolomic studies consistently found increased levels of glucose and lactate in septic patients. The inclusion of both metabolites with other biomarkers such as ribitol, pseudouridine, adipate, 2-hydroxybutyrate and threonine suggests potential to develop a panel of stable early predictors for neonatal sepsis. These studies demonstrate the ability to conduct hypothesis-free screening of the metabolome using metabolomics-based technologies (NMR and MS) to discover novel compositions of metabolites for neonatal sepsis diagnosis (Mickiewicz et al., 2013; Fanos et al., 2014). In particular, coupling of MS analyzers with different separation methods (GC or LC) for acquisitions could improve sensitivity, specificity, chemical coverage and dynamic range for untargeted metabolite discovery from various biological samples (May and McLean, 2016; Tebani et al., 2016b). With clear evidence of metabolic disturbances caused by sepsis or infection, metabolites possess high potential to serve as biomarkers or predictors for sepsis and infection during the neonatal period. Currently, only a limited number of small-scale studies have been carried out, and translation of identified metabolite biomarker panels for bedside diagnosis of neonatal sepsis remains unknown (Mickiewicz et al., 2013; Fanos et al., 2014). MS-based analyses remain time intensive and costly, requiring extensive sample processing in addition to specialized hardware and software for spectral data acquisition, preprocessing and data analysis. Despite this, MS-based technologies have routinely been used for screening of inborn errors of metabolism in newborns with commercial companies working to develop metabolite-based biomarker diagnostic tests, paving the way for future translation of a rapid and accurate metabolite-based biomarker test for neonatal sepsis diagnosis (Nagana Gowda and Raftery, 2013; Tebani et al., 2016b).

**Toward Precision Medicine for Neonatal Sepsis**

Despite limited research in the field of neonatal sepsis to date and limitations within each study (Table 1), available data highlights potential of omics-based approaches to interrogate sepsis pathophysiology for the discovery of novel biomarkers and diagnostic signatures for sepsis. Investigating a septic event at multiple levels, such as across the transcriptional and metabolic response, and at different times during sepsis should capture novel features that account for interactions between genes and biomolecules in a systems network. This can also identify the dynamic changes between networks of genes and biomolecules from different systems, facilitating the discovery of novel dynamical network biomarkers for neonatal sepsis. Identification of dynamical network biomarkers, unlike traditional markers such as the use of a single protein concentration, may allow precise stratification of patients by disease phenotype (such as severity), thereby allowing better prognosis and targeted use of therapeutics (Christaki and Giamarellos-Bourboulis, 2014; Li and Chen, 2014).

The shift from empirical evidence-based medicine to stratified, omics-led medicine for neonatal sepsis remains in its infancy and faces several challenges (Figure 1). Firstly, neonatal sepsis is a heterogeneous clinical syndrome without a consensus definition, therefore available studies have used differing case definitions (Table 1). Multiple factors contribute to heterogeneity of disease, including the specific pathogen involved (type, load and site of invasion) and the maturational state and capacity of the host immune system (influenced by gestational and postnatal age and associated comorbidities). The inability to stratify patients by disease state based on a clear and consistent definition hinders our ability to compare findings between studies, impeding progress of validating identified biomarkers or gene signatures that can universally improve diagnostic and prognostic tests for neonatal sepsis (Chen et al., 2014; Wynn, 2016). Secondly, we lack representative animal models to investigate and validate neonatal sepsis biomarkers and investigate the dynamics of sepsis pathophysiology. Thirdly, novel biomarkers discovered will need to be selected from an appropriate biological sample that can be pragmatically translated for clinical use (Thongboonkerd, 2013). Given the limited number of studies that have identified biomarkers capable of translating into a bench-side test for neonatal sepsis, analyzing a wide range of longitudinal samples, including whole blood, plasma, serum and urine would be the most comprehensive way of finding novel biomarkers and diagnostic gene signatures (Willis and Lord, 2015). The robustness of the identified markers will need to be validated in appropriately powered multicenter studies to account for instrumental, technical, biological and physiological variations. Finally, whilst our ability to generate omics-based data for neonatal sepsis has significantly improved in the last 10 years with the reducing cost and increasing speed of high-throughput technologies, it has led to a bottleneck with data analysis and interpretation.

Theoretically, discovery of biomarkers and/or sepsis signatures through integration of multi-omic data has potential to stratify neonates with sepsis for improved treatment and prognosis. However, despite increasing access to multi-omics profiles, our ability to analyze and interpret high-dimensional data remains limited by our understanding of the complex interactions between the genome, transcriptome, proteome and metabolome. The complexity of sepsis pathophysiology, heterogenic nature of clinical data, along with bioinformatical and statistical limitations hinder our ability to integrate
multi-omic data for interpretation of relationships between “networks” and “networks of networks” (Vucic et al., 2012; Li and Chen, 2014; Alyass et al., 2015). Data storage and management solutions that can integrate each patient’s clinical records, laboratory results and omics data need to be developed and implemented before we can transition into precision medicine by the bedside. These solutions will need to be robust enough to deal with the gigabytes of multiple omic platform-specific data, from a range of biological sample types, and across varying instrumental technologies (Gullapalli et al., 2012; Tebani et al., 2016b).

As we move from stratified medicine into the era of precision medicine, the gap between biological science and medicine needs to be bridged in order to translate a patient-specific bedside test for neonatal sepsis. This will require open collaborations and knowledge sharing across multiple disciplines from laboratory scientists to statisticians, computational biologists and clinicians. Further, the translation will require investment into informatics infrastructures that can meet the computationally intensive pipelines for integrative multi-omics, clinical and laboratory data analytics and visualization (Figure 1). Additionally, the legal and ethical framework ensuring confidentiality and privacy of patient information will need to be established. The storage and secure sharing of data through cloud computing solutions will also need to be addressed in order to provide informative digital health reports that guide clinicians for personalized management of sepsis in neonates (Costa, 2014; Alyass et al., 2015; Willis and Lord, 2015; Tebani et al., 2016b).

The use of precision medicine is highly relevant for neonatology. The heterogeneous nature of neonatal sepsis means that our current approaches to diagnosis are too simplistic and imprecise to identify affected individuals accurately (Wynn, 2016). The nature of neonatal medicine, with its highly intensive and frequent monitoring of infants, often over protracted periods and from birth, makes it ideal for translation to the precision medicine setting. Neonatal precision medicine will allow a more predictive and preventive approach, where septic infants can be identified ahead of clinical disease phenotype expression, ensuring prompt and effective antibiotic use. Ultimately, the era of precision medicine for neonatal sepsis, built on the platforms of omics technologies, holds promising potential to ensure quicker and more accurate diagnosis of neonatal sepsis for personalized treatment and improve prognosis of septic neonates (Thongboonkerd, 2013; Christaki and Giamarellos-Bourboulis, 2014).

**AUTHOR CONTRIBUTIONS**

SN, PJ, and TM researched the topic and wrote draft manuscripts. TS, PS, and AC contributed to discussions and drafting of the manuscript.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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