Design, Synthesis and Antitubercular Activity of Novel Isoniazid‑Cyclic‑Amine‑Azachalcones Hybrids

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In this work, it is described the design of twenty-four heterocyclic amine-azachalcones compounds through molecular hybridization of chalcone scaffold and fragments of isoniazid, fluoroquinolones, and linezolid with antituberculosis potential. The new compounds were synthesized via Claisen-Schmidt condensation, providing yields of 36-95%. Fifteen compounds showed antituberculosis activity against Mycobacterium tuberculosis H37Rv strain. Two amine-azachalcones 15 and 17 showed relevant biological activity with minimum inhibitory concentration (MIC) values of 6.62 and 4.85 µM, respectively. Compound 12 showed the best profile of antitubercular activity with MIC = 9.54 µM and selectivity index (SI) = 9.33. It was found that morpholine group is important to increase potency of antimycobacterial activity but also to add some toxicity to the chalcone molecular framework. The results described herein would be a guide in the designing of novel and optimized antitubercular derivatives based on the chalcone scaffold.

Keywords: isoniazid, heterocyclic amines, molecular hybridization, antitubercular activity, aldol condensation

Introduction

Tuberculosis (TB) disease is classified as one of the top ten leading causes of death in the world, due to its aggressive infection promoted by a unique agent. Moreover, one-fourth of the global population was detected with latent tuberculosis, occurring without any disease symptoms.1 In 2017, ten million people manifested TB and the situation aggravated even more with 1.3 million deaths.1

The first-line treatment for TB involves the use of five different drugs such as isoniazid, pyrazinamide, streptomycin, ethambutol, and rifampicin, which can be used in association.2 When the first-line drugs cannot be used, it is recommended the second-line treatment with levofloxacin or moxifloxacin, bedaquiline, and linezolid.3 Failures in chemotherapy are related to several aspects such as high complexity (association of three or more drugs); long period of treatment (6 to 9 months or up to 24 months);2,4 countless adverse effects (nausea, vomiting, and epigastric pain),1,2 and bacterial resistance.
Bacterial resistance is a cause of alarm for the health systems throughout the world. Bacterial strains of TB have been classified into two types, TB multidrug-resistant (resistance to the first-line drugs), and TB extremely resistant (resistance to first and second-line drugs). In this context, it is reinforced the relevance to search into new antituberculosis drug candidates.

Natural products remain a fundamental source of inspiration in the discovery and development of new bioactive compounds. Through secondary metabolism, natural products are able to create several types of substances, with multiple biological activities.

Among the secondary metabolites, our research group is especially interested in chalcone scaffold, as a synthetic target. Chalcones are considered a privileged structure and have been widely used as a template in medicinal chemistry studies. Due to its structural diversity, chalcones have shown great potential in the molecular modification process, leading to the synthesis of compounds containing different pharmacophore groups, which can enhance its pharmacological effect.

Recent reports aimed at chalcones as compounds with a wide range of biological activities such as anticancer, cancer-preventative effects, anti-inflammatory, antibacterial, antidiabetic, antioxidant, antimicrobial, antiviral, antimalarial and neuroprotective effects. It is noteworthy that chalcone compounds are emerging as potential candidates for new antitubercular drugs (Figure 1). Structure-activity relationship (SAR) and quantitative structure-activity relationship (QSAR) information has demonstrated that the nitrofuran scaffold as a substituent in chalcones improved its antitubercular activities.

Firstly, it was necessary to obtain p-aminated acetophenones with the appropriate substitution patterns (Table 1). The synthesis of acetophenones occurred via aromatic nucleophilic substitution of 4-fluoroacetophenone in the presence of cyclic amines, using K$_2$CO$_3$ and dimethyl sulfoxide (DMSO) as a solvent. Products were obtained in 71 to 99% yields; the reaction times were observed for 24 or 48 h until the complete consumption of starting materials (Scheme 2). Amine-azachalcones were obtained via Claisen-Schmidt condensation of p-aminated acetophenones and carbaldehydes using a solution of NaOH in ethanol as solvent, which was added dropwise, providing the compounds with yields ranging from 36 to 95% (Table 1).

Structural modifications intended to increase the complexity of cyclic amines bonded to the aromatic ring, in order to generate a variation in the molecular volume in R'. Thus, compounds were obtained from simple N-heterocyclic amine templates, as pyrrolidine, to more complex molecules like 1-(pyrimidin-2-yl)piperazine. When positional isomers were compared, pyridine compounds substituted with nitrogen at position 4 showed...

Figure 1. Structures of new chalcone antitubercular agents 1-4.
the worst yields (11, 14, 17, 20, 23, 26, 32) except for the compound 29.

Unknown synthetic compounds 9-32 were characterized by $^1$H and $^{13}$C nuclear magnetic resonance (NMR) and high-resolution mass spectrometry (HRMS).

**Antitubercular activity**

The antitubercular activity of compounds 9-32 was evaluated against *Mycobacterium tuberculosis* H37Rv strain (Table 2). Minimum inhibitory concentrations (MICs) were defined as the lowest concentration that resulted in 90% of *M. tuberculosis* growth inhibition. Fifteen compounds of the series demonstrated some level of activity against *M. tuberculosis*.

The presence of a six-membered hetero-aromatic ring bonding directly to the cyclic amine group did not contribute to the antituberculosis activity (MIC > 250 µM) since the bulkiest azachalcones 27-32 were inactive.

Hybrid pyrrolidine substituted derivatives 9 and 10 were also inactive against *M. tuberculosis* (MIC > 250 µM). However, compound 11 showed good activity (MIC = 20.22 µM).

Compounds 12-14, for instance, presented different biological activities. The isomers 12 and 14 exhibited values of MIC = 9.54 and 24.52 µM, respectively. On the other hand, isomer 13 was inactive (MIC > 250 µM).

Thiomorpholine derivatives 18-20 showed MIC values ranging from 34.79 to 111.13 µM. Bioisosteric substitution of sulfur atom (in thiomorpholine derivatives) by oxygen (morpholine nucleus) improved the biological activity. Compounds 15, 16 and 17 were considered the most active positional isomers in the series, exhibiting an MIC of 6.62, 14.03, and 4.85 µM, respectively.

Additionally, N-alkyl piperazine derivatives 21-23 (MIC = 14.15, 26.64 and 16.72 µM, respectively) displayed good biological activities when compared to pyrazinamide (MIC = 26.80 µM).

Regarding SAR of positional isomers (2-pyridine, 3-pyridine, 4-pyridine), it was observed different MICs in the same structural group, indicating the importance of this factor in the biological activity. Derivatives containing 4-pyridines were more active than their 2- and 3- positional congeners (Table 2, compounds 11, 17, 20, 26). Positional isomers with 2-pyridine were the second most active (Table 2, compounds 12 and 21). Compounds with a 3-pyridine nucleus were the least active (Table 2, compounds 10, 13, 16, 22, 25).

In summary, the compounds with MIC < 10 µM were considered good hits in a series of antituberculosis drug...
Table 1. Synthesis of the heterocyclic amine-azachalcones 9-32

Reaction conditions: 35a-35h (2 mmol), 36a-36c (4 mmol), ethanol (10 mL) and NaOH (30 mmol solubilized in 25 mL of ethanol added dropwise), room temperature, 4 h, inert atmosphere.
candidates. Azachalcones 12, 15 and 17 with MIC = 9.54 (selectivity index (SI) = 9.33), 6.62 (SI = 1.39) and 4.85 µM (SI = 3.49) showed higher activity than the reference drug tested, pyrazinamide (MIC = 26.80 µM). These derivatives also showed relevant antituberculosis profile when compared to ciprofloxacin (MIC = 9.43 µM). Figure 2 shows the SAR highlights of azachalcones.

### Table 2. In vitro antituberculosis activity of azachalcones 9-32 against *M. tuberculosis* H37Rv strain

| Compound | H37Rv MIC<sup>a</sup> (µg mL<sup>−1</sup>) | H37Rv MIC<sup>b</sup> (µM) | NIH/3T3 IC<sub>50</sub><sup>c</sup> (µg mL<sup>−1</sup>) | SI<sup>d</sup> |
|----------|---------------------------------|----------------|----------------|--------|
| 9        | > 250                           | > 250          | 17.39          | 0.07   |
| 10       | > 250                           | > 250          | 32.21          | 0.13   |
| 11       | 5.63                            | 20.22          | 64.78          | 11.51  |
| 12       | 2.79                            | 9.54           | 26.04          | 9.33   |
| 13       | > 250                           | > 250          | 32.87          | 0.13   |
| 14       | 7.17                            | 24.52          | 2.87           | 0.40   |
| 15       | 1.95                            | 6.62           | 2.71           | 1.39   |
| 16       | 4.13                            | 14.03          | 2.94           | 0.71   |
| 17       | 1.43                            | 4.85           | 4.99           | 3.49   |
| 18       | 34.50                           | 111.13         | 79.16          | 2.29   |
| 19       | 15.30                           | 49.28          | 5.28           | 0.34   |
| 20       | 10.80                           | 34.79          | 2.78           | 0.26   |
| 21       | 4.35                            | 14.15          | 2.88           | 0.66   |
| 22       | 8.19                            | 26.64          | 20.73          | 2.53   |
| 23       | 5.14                            | 16.72          | 2.77           | 0.54   |
| 24       | 3.62                            | 11.26          | 2.71           | 0.75   |
| 25       | 7.76                            | 24.14          | 23.39          | 3.01   |
| 26       | 3.38                            | 10.51          | 2.60           | 0.77   |
| 27       | > 250                           | > 250          | 32.43          | 0.13   |
| 28       | > 250                           | > 250          | 361.58         | 1.44   |
| 29       | > 250                           | > 250          | 32.78          | 0.13   |
| 30       | > 250                           | > 250          | 23.21          | 0.09   |
| 31       | > 250                           | > 250          | 26.94          | 0.10   |
| 32       | > 250                           | > 250          | 29.42          | 0.11   |
| INH<sup>e</sup> | 0.015                         | 0.10          | NC<sup>e</sup> | NC<sup>e</sup> |
| RMP<sup>e</sup> | 0.059                        | 0.065         | NC<sup>e</sup> | NC<sup>e</sup> |
| PYR<sup>e</sup> | 3.3                          | 27.80         | NC             | NC<sup>e</sup> |
| DOX<sup>h</sup> | NC<sup>e</sup>             | 2.50          | NC             | NC<sup>e</sup> |

<sup>a</sup>MIC: minimum inhibitory concentration (lowest concentration that results in 90% inhibition of growth of *M. tuberculosis*);<sup>b</sup>IC<sub>50</sub>: half maximal inhibitory concentration on fibroblast cells (NIH/3T3);<sup>c</sup>SI: selectivity index (IC<sub>50</sub> on fibroblast cells / MIC on *M. tuberculosis* H37Rv strain);<sup>d</sup>INH: isoniazid, positive control for *M. tuberculosis*; NC: values not calculated;<sup>e</sup>RMP: rifampicin, positive control for *M. tuberculosis*;<sup>f</sup>PYR: pyrazinamide, positive control for *M. tuberculosis*;<sup>g</sup>DOX: doxorubicin, positive control for fibroblast cells.

Figure 2. SAR of the hybrids azachalcones 9-32.
Conclusions

In conclusion, twenty-four heterocyclic amine-azachalcones 9-32 were obtained from moderate to good yields. Antituberculosis activity was evaluated against the M. tuberculosis H37Rv strain. Although the higher potency of compounds 15 and 17 (MIC = 6.62 and 4.85 µM, respectively) towards antituberculosis activity, both analogues were considered more cytotoxic (SI = 1.39 and 3.49, respectively). However, azachalcone 12 showed the best profile of activity and selectivity index (MIC = 9.54 µM; SI = 9.33), which provide important information about the SAR.

Morpholine group as a substituent in molecular template was related to higher potency and relevant biological activity, but on the other hand it also increased the cytotoxicity of azachalcones. Further research might explore new structural modifications, in order to provide more information about SAR of azachalcones, and also, optimize the hit-like compounds of this series.

Experimental

General remarks

All solvents were purchased from Synth®, São Paulo, Brazil, and distilled before use according to the standard procedure. All reactions were performed under an atmosphere of dry nitrogen and monitored by thin-layer chromatography (TLC) using prepared plates (silica gel 60 F254 on aluminum purchased from Sigma-Aldrich®, St. Louis, MO, USA). The chromatograms were examined under both 254 and 360 nm UV light or with the developing agent ethanolic vanillin and heat. Flash column chromatography was performed on silica gel 60 (particle size 200-400 mesh ASTM, purchased from Sigma-Aldrich®, St. Louis, MO, USA) and eluted with hexane or hexane/ethyl acetate in different ratios. Melting points were determined using Fisatom 430D equipment. The 1H and 13C NMR spectra were recorded in CDCl₃ solutions using a Bruker 75 or 300 MHz spectrometer. Chemical shifts (δ) were expressed as parts per million (ppm) downfield from tetramethylsilane as the internal standard. High-resolution electrospray ionization mass spectrometry (HR-ESI-MS) measurements were carried out on a quadrupole time-of-flight instrument (UltrOTOF-Q, BrukerDaltonics, Billerica, MA, USA). General reagents as 4-fluoro-acetophenone (UltrOTOF-Q, BrukerDaltonics, Billerica, MA, USA). General procedure for the preparation of p-aminated acetophenone (35a-35h)

To a solution of compounds 33 (10 mmol), 34a-34h (17 mmol) in dimethyl sulfoxide (25 mL), K₂CO₃ (17 mmol) was added. The reaction was stirred at 90 °C for 24 h (35a, 35c, 35e, 35f) or 48 h (35d, 35g, 35h). After that, a saturated aqueous NaCl solution (50 mL) was added. Then, the reaction was extracted with ethyl acetate (2 × 25 mL). The organic phase was washed with saturated aqueous NaCl solution (10 × 50 mL). The aqueous phase was retro-extracted with ethyl acetate (2 × 25 mL). Organic phases were dried over anhydrous MgSO₄ and the solvent was removed under reduced pressure. Compounds 35a, 35c, 35e, 35f did not require further purification. Compounds 35d, 35g, 35h were purified by column chromatography on silica gel using hexane/ethyl acetate 8:2 as eluent.

1-(4-(Pyrrolidin-1-yl)phenyl)ethanone (34a)

Yield: 99%; light brown solid; 1H NMR (300 MHz, CDCl₃) δ 1.98-2.03 (m, 4H, 2CH₂), 2.47 (s, 3H, CH₃), 3.33 (m, 4H, 2CH₂), 6.48 (d, 2H, J 9.0 Hz, Ar-H), 7.83 (d, 2H, J 9.0 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 25.4, 25.8, 26.1, 50.3, 113.6, 127.2, 130.6, 152.9, 196.4.

1-(4-Morpholinophenyl)ethanone (34c)

Yield: 79%; light brown solid; 1H NMR (300 MHz, CDCl₃) δ 2.51 (s, 3H, CH₃), 3.27-3.32 (m, 4H, 2CH₂), 3.81-3.86 (m, 4H, 2CH₂), 6.79 (d, 2H, J 9.0 Hz, Ar-H), 7.85 (d, 2H, J 9.0 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 26.2, 47.5, 66.6, 113.3, 128.1, 130.3, 154.2, 196.6.

1-(4-Thiomorpholinophenyl)ethanone (34d)

Yield: 85%; light yellow solid; 1H NMR (300 MHz, CDCl₃) δ 2.49 (s, 3H, CH₃), 2.65-2.70 (m, 4H, 2CH₂), 3.73-3.79 (m, 4H, 2CH₂), 6.85 (d, 2H, J 9.0 Hz, Ar-H), 7.88 (d, 2H, J 9.0 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 25.8, 26.1, 50.3, 113.6, 127.2, 130.6, 152.9, 196.4.

1-(4-(4-Methylpiperazin-1-yl)phenyl)ethanone (34e)

Yield: 90%; light brown solid; 1H NMR (300 MHz, CDCl₃) δ 2.38 (s, 3H, CH₃), 2.50 (s, 3H, CH₃), 2.58-2.63 (m, 4H, 2CH₂), 3.37-3.42 (m, 4H, 2CH₂), 6.86 (d, 2H, J 9.0 Hz, Ar-H), 7.86 (d, 2H, J 9.0 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 26.1, 45.9, 47.1, 54.6, 113.5, 127.8, 130.4, 153.9, 196.5.

1-(4-(4-Ethypiperazin-1-yl)phenyl)ethanone (34f)

Yield: 94%; beige solid; 1H NMR (300 MHz, CDCl₃) δ 1.09 (t, 3H, J 6.0 Hz, CH₃), 2.43 (q, 2H, J 6.0 Hz, CH₂), 2.48 (s, 3H, CH₃), 2.52-2.57 (m, 4H, 2CH₂), 3.32-3.37
phases were dried over anhydrous MgSO₄ and the solvent was retro-extracted with aqueous NaCl solution (3 × 50 mL). The aqueous phase (4 × 20 mL), the organic phase was washed with saturated tert NaCl and the product was extracted with -butanol (2 × 20 mL). Then, it was added 50 mL of a saturated solution of NaOH (2 h). After the complete addition of NaOH (2 h), the solution was solubilized in anhydrous ethanol (25 mL) at room temperature, under nitrogen atmosphere. To a solution of 1-(4-(4-(Pyridin-2-yl)piperazin-1-yl)phenyl)ethanone (34g) in anhydrous ethanol (10 mL), under nitrogen atmosphere, was added 50 mL of a saturated solution of NaOH (2 h). After the complete addition of NaOH (2 h), the reaction was kept at room temperature for two hours. Then, it was added 50 mL of a saturated solution of NaCl and the product was extracted with tert-butanol (4 × 20 mL). The organic phase was washed with saturated aqueous NaCl solution (3 × 50 mL). The aqueous phase was retro-extracted with tert-butanol (2 × 20 mL). Organic phases were dried over anhydrous MgSO₄ and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel using hexane/ethyl acetate (7:3, 6:4, 1:1) as eluent and/or by recrystallization.

To a solution of p-aminated acetophenone 35a-35h (2 mmol), carbaldehyde 36a-36e (4 mmol) in anhydrous ethanol (10 mL) at room temperature, under nitrogen atmosphere, and vigorously stirred, NaOH (30 mmol) solubilized in anhydrous ethanol (25 mL) was added dropwise. After the complete addition of NaOH (2 h), the reaction was kept at room temperature for two hours. Then, it was added 50 mL of a saturated solution of NaCl and the product was extracted with tert-butanol (4 × 20 mL). The organic phase was washed with saturated aqueous NaCl solution (3 × 50 mL). The aqueous phase was retro-extracted with tert-butanol (2 × 20 mL). Organic phases were dried over anhydrous MgSO₄ and the solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel using hexane/ethyl acetate (7:3, 6:4, 1:1) as eluent and/or by recrystallization.

General procedure for the preparation of chalcones 9-32

(E)-3-(Pyrindin-3-yl)-1-(4-(pyrrolidin-1-yl)phenyl)prop-2-en-one (10)

Yield: 85%; yellow solid; mp 185 °C; 1H NMR (300 MHz, CDCl₃) δ 2.01-2.06 (m, 4H, 2CH₂), 3.36-3.40 (m, 4H, 2CH₂), 6.56 (2H, J 9.0 Hz, Ar-H), 7.32 (dd, 1H, J 4.7, 2.7 Hz, Ar-H), 7.63 (d, 1H, J 15.7 Hz, CH), 7.73 (d, 1H, J 15.7 Hz, CH), 7.91 (dl, 1H, J 8.0 Hz, Ar-H), 7.98 (d, 2H, J 9.0 Hz, Ar-H), 8.57 (dd, 1H, J 4.7, 1.3 Hz, Ar-H), 8.84 (d, 1H, J 1.5 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 25.4, 47.6, 111.0, 123.8, 124.3, 125.0, 131.1, 131.5, 134.7, 138.2, 149.4, 150.1, 151.2, 186.8; HRMS (FTMS + pESI) m/z, calcd. for C₁₈H₁₈N₂O [M⁺]: 279.1497, found: 279.1516.

(E)-3-(Pyrindin-4-yl)-1-(4-(pyrrolidin-1-yl)phenyl)prop-2-en-one (11)

Yield: 65%; yellow solid; mp 171-173 °C; 1H NMR (300 MHz, CDCl₃) δ 2.03-2.07 (m, 4H, 2CH₂), 3.38-3.42 (m, 4H, 2CH₂), 6.57 (d, 2H, J 9.0 Hz, Ar-H), 7.46 (d, 2H, J 9.0 Hz, Ar-H), 7.64 (d, 1H, J 15.7 Hz, CH), 7.73 (d, 1H, J 15.7 Hz, CH), 7.99 (d, 2H, J 9.0 Hz, Ar-H), 8.65 (d, 2H, J 5.5 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 25.4, 47.6, 111.0, 122.0, 124.9, 126.5, 131.2, 139.0, 142.9, 150.5, 151.3, 186.7; HRMS (FTMS + pESI) m/z, calcd. for C₁₈H₁₈N₂O [M⁺]: 279.1497, found: 279.1516.

(E)-1-(4-(Piperidin-1-yl)-phenyl)-3-(pyridin-2-yl)prop-2-en-one (12)

Yield: 85%; yellow solid; mp 156-158 °C; 1H NMR (300 MHz, CDCl₃) δ 1.65 (s, 6H, 3CH₂), 3.38 (s, 4H, 2CH₂), 6.86 (d, 2H, J 9.0 Hz, Ar-H), 7.23-7.27 (m, 1H, Ar-H), 7.44 (d, 1H, J 8.0 Hz, Ar-H), 7.68-7.74 (m, 2H, Ar-H and CH), 8.04 (d, 2H, J 9.0 Hz, Ar-H), 8.13 (d, 1H, J 15.2 Hz, CH), 8.66 (d, 1H, J 3.8 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 24.4, 25.4, 48.5, 113.2, 124.0, 125.3, 125.8, 126.9, 131.1, 136.8, 140.8, 150.0, 153.7, 154.5, 187.69; HRMS (FTMS + pESI) m/z, calcd. for C₁₉H₂₀N₂O [M⁺]: 293.1654, found: 293.1655.

(E)-1-(4-(Piperidin-1-yl)-phenyl)-3-(pyridin-3-yl)prop-2-en-one (13)

Yield: 90%; yellow solid; mp 179-181 °C; 1H NMR (300 MHz, CDCl₃) δ 1.66 (s, 6H, 3CH₂), 3.39 (s, 4H, 2CH₂), 6.88 (d, 2H, J 9.0 Hz, Ar-H), 7.32 (dd, 1H, J 3.6, 8.0 Hz, Ar-H), 7.61 (d, 1H, J 15.7 Hz, CH), 7.74 (d, 1H, J 15.7 Hz, CH), 7.92 (d, 1H, J 8.0 Hz, Ar-H), 7.96 (d, 2H, J 9.0 Hz, Ar-H).
Ar-H), 8.57 (d, 1H, J 3.6 Hz, Ar-H), 8.83 (s, 1H, Ar-H); 
$^{13}$C NMR (75 MHz, CDCl$_3$) δ 24.3, 25.3, 48.4, 123.7, 124.0, 126.6, 130.9, 131.2, 134.5, 138.8, 149.6, 150.5, 154.4, 187.0; HRMS (FTMS + pESI) m/z, calcd. for C$_{18}$H$_{18}$N$_2$O [M]+: 293.1654, found: 293.1653.

(E)-1-(4-(Piperidin-1-yl)phenyl)-3-(pyridin-4-yl)prop-2-en-1-one (14)

Yield: 90%; yellow solid; mp 149-150 °C; 'H NMR (300 MHz, CDCl$_3$) δ 2.03 (s, 6H, 3CH$_3$), 3.38 (s, 4H, 2CH$_2$), 6.55 (d, 2H, J 8.0 Hz, Ar-H), 7.46 (s, 2H, J 15.8 Hz, CH), 7.62 (d, 1H, J 15.8 Hz, CH), 7.72 (d, 1H, J 15.8 Hz, CH), 7.97 (d, 2H, J 8.0 Hz, Ar-H), 8.63 (s, 2H, Ar-H); $^{13}$C NMR (75 MHz, CDCl$_3$) δ 24.3, 25.3, 48.4, 113.2, 122.6, 126.3, 127.8, 131.1, 138.5, 145.0, 148.3, 154.5, 186.4; HRMS (FTMS + pESI) m/z, calcd. for C$_{19}$H$_{20}$N$_2$O [M]+: 293.1654, found: 293.1740.

(E)-1-(4-Morpholinophenyl)-3-(pyridin-4-yl)prop-2-en-1-one (15)

Yield: 81%; yellow solid; mp 138-140 °C; 'H NMR (300 MHz, CDCl$_3$) δ 3.30-3.33 (m, 4H, 2CH$_2$), 3.82-3.85 (m, 4H, 2CH$_2$), 6.88 (d, J 9.0 Hz, 2H, Ar-H), 7.23-7.28 (m, 1H, Ar-H), 7.44 (d, J 7.7 Hz, 1H, Ar-H), 7.72 (m, 2H, Ar-H and CH), 8.06 (d, J 9.0 Hz, 2H, Ar-H), 8.13 (d, J 15.5 Hz, CH), 8.66 (d, 1H, J 3.9 Hz, Ar-H); $^{13}$C NMR (75 MHz, CDCl$_3$) δ 47.4, 66.5, 113.3, 124.1, 125.3, 125.7, 128.5, 130.9, 136.9, 141.2, 150.0, 153.6, 154.3, 188.0; HRMS (FTMS + pESI) m/z, calcd. for C$_{19}$H$_{20}$N$_2$O [M]+: 295.1446, found: 295.1455.

(E)-1-(4-Morpholinophenyl)-3-(pyridin-3-yl)prop-2-en-1-one (16)

Yield: 91%; yellow solid; mp 156-158 °C; 'H NMR (300 MHz, CDCl$_3$) δ 3.31-3.34 (m, 4H, 2CH$_2$), 3.83-3.86 (m, 4H, 2CH$_2$), 6.90 (d, 2H, J 9.0 Hz, Ar-H), 7.33 (dd, J 7.4, 4.5 Hz, 1H, Ar-H), 7.60 (d, 1H, J 15.7 Hz, CH), 7.75 (d, 1H, J 15.7 Hz, CH), 7.91 (d, 1H, J 9.0 Hz, Ar-H), 7.99 (d, 2H, J 9.0 Hz, Ar-H), 8.59 (s, 1H, Ar-H); $^{13}$C NMR (75 MHz, CDCl$_3$) δ 47.4, 66.5, 113.4, 123.7, 123.9, 128.3, 130.9, 143.6, 143.9, 149.7, 150.6, 154.4, 187.4; HRMS (FTMS + pESI) m/z, calcd. for C$_{19}$H$_{18}$N$_2$O$_2$ [M]+: 297.1516, found: 297.1512.

(E)-1-(4-Morpholinophenyl)-3-(pyridin-3-yl)prop-2-en-1-one (17)

Yield: 55%; yellow solid; mp 122-124 °C; 'H NMR (300 MHz, CDCl$_3$) δ 3.32-3.35 (m, 4H, 2CH$_2$), 3.83-3.87 (m, 4H, 2CH$_2$), 6.90 (d, 2H, J 9.0 Hz, Ar-H), 7.45 (dd, 2H, J 4.5, 1.6 Hz, Ar-H), 7.63 (d, 1H, J 15.8 Hz, CH), 7.69 (d, 1H, J 15.8 Hz, Ar-H), 7.99 (d, 2H, J 9.0 Hz, Ar-H), 8.65 (dd, 2H, J 4.5, 1.5 Hz, Ar-H); $^{13}$C NMR (75 MHz, CDCl$_3$) δ 47.3, 66.5, 113.3, 122.0, 126.1, 128.0, 130.8, 140.0, 142.6, 150.5, 154.4, 187.2; HRMS (FTMS + pESI) m/z, calcd. for C$_{19}$H$_{18}$N$_2$O$_2$ [M]+: 295.1446, found: 295.1610.
Design, Synthesis and Antitubercular Activity of Novel Isoniazid-Cyclic-Amine-Azachalcones Hybrids

**J. Braz. Chem. Soc.**

**1H NMR**

1H NMR (300 MHz, CDCl₃) δ 2.33 (s, 3H, CH₃), 2.53-2.56 (m, 4H, 2CH₂), 3.37-3.40 (m, 4H, 2CH₂), 6.88 (d, 2H, J 9.0 Hz, Ar-H), 7.44 (d, J 4.69 Hz, 2H, Ar-H), 7.61 (d, J 15.9 Hz, 1H, CH), 7.64 (d, 1H, J 15.7 Hz, CH), 7.96 (d, 2H, J 9.0 Hz, Ar-H), 8.63 (s, 2H, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 26.0, 46.9, 54.5, 113.5, 121.3, 122.0, 126.1, 127.4, 130.9, 139.8, 142.6, 150.4, 154.3, 187.1; HRMS (FTMS + pESI) m/z, calcd. for C₁₉H₂₁N₃O [M⁺]: 308.1763, found: 308.1759.

**Yield: 88%; yellow solid; mp 143 °C; **1H NMR (300 MHz, CDCl₃) δ 2.33 (s, 3H, CH₃), 2.53-2.56 (m, 4H, 2CH₂), 3.37-3.40 (m, 4H, 2CH₂), 6.88 (d, 2H, J 9.0 Hz, Ar-H), 7.44 (d, J 4.69 Hz, 2H, Ar-H), 7.61 (d, J 15.9 Hz, 1H, CH), 7.64 (d, 1H, J 15.7 Hz, CH), 7.96 (d, 2H, J 9.0 Hz, Ar-H), 8.63 (s, 2H, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 26.0, 46.9, 54.5, 113.5, 121.3, 122.0, 126.1, 127.4, 130.9, 139.8, 142.6, 150.4, 154.3, 187.1; HRMS (FTMS + pESI) m/z, calcd. for C₁₉H₂₁N₃O [M⁺]: 308.1763, found: 308.1759.

**Yield: 48%; yellow solid; mp 97 °C; 1H NMR (300 MHz, CDCl₃) δ 2.33 (s, 3H, CH₃), 2.53-2.56 (m, 4H, 2CH₂), 3.37-3.40 (m, 4H, 2CH₂), 6.88 (d, 2H, J 9.0 Hz, Ar-H), 7.44 (d, J 4.69 Hz, 2H, Ar-H), 7.61 (d, J 15.9 Hz, 1H, CH), 7.64 (d, 1H, J 15.7 Hz, CH), 7.96 (d, 2H, J 9.0 Hz, Ar-H), 8.63 (s, 2H, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 26.0, 46.9, 54.5, 113.5, 121.3, 122.0, 126.1, 127.4, 130.9, 139.8, 142.6, 150.4, 154.3, 187.1; HRMS (FTMS + pESI) m/z, calcd. for C₁₉H₂₁N₃O [M⁺]: 308.1763, found: 308.1759.

**Yield: 94%; yellow solid; mp 150 °C; 1H NMR (300 MHz, CDCl₃) δ 1.13 (t, 3H, J 6.0 Hz, CH₃), 2.48 (q, 2H, J 6.0 Hz, CH₂), 2.51-2.61 (m, 4H, 2CH₂), 3.40-3.43 (m, 4H, 2CH₂), 6.90 (d, 2H, J 9.0 Hz, Ar-H), 7.27-7.29 (m, 1H, Ar-H), 7.45 (d, 1H, J 7.7 Hz, Ar-H), 7.69-7.77 (m, 2H, Ar-H and CH), 8.06 (d, 2H, J 9.0 Hz, Ar-H), 8.13 (d, 1H, J 15.2 Hz, CH), 8.67 (d, 1H, J 4.0 Hz, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 12.0, 47.2, 52.3, 52.5, 113.3, 124.1, 125.3, 127.9, 131.0, 136.8, 141.1, 150.1, 153.6, 154.2, 187.9; HRMS (FTMS + pESI) m/z, calcd. for C₂₂H₂₃N₅O [M⁺]: 372.1824, found: 372.1844.

**Yield: 36%; yellow solid; mp 208-210 °C; 1H NMR (300 MHz, CDCl₃) δ 3.42-3.44 (m, 4H, 2CH₂), 3.93-3.96 (m, 4H, 2CH₂), 6.49 (t, 1H, J 4.7 Hz, Ar-H), 6.89 (d, 2H, J 9.0 Hz, Ar-H), 7.29 (dd, 1H, J 7.9, 4.9 Hz, Ar-H), 7.56 (d, 1H, J 15.7 Hz, CH), 7.71 (d, 1H, J 15.7 Hz, CH), 7.88 (dd, 1H, J 6.1, 1.9 Hz, Ar-H), 7.96 (d, 2H, J 9.0 Hz, Ar-H), 8.29 (2H, J 4.7 Hz, Ar-H), 8.55 (d, 1H, J 3.7 Hz, Ar-H), 8.80 (s, 1H, Ar-H); 13C NMR (75 MHz, CDCl₃) δ 43.2, 47.0, 110.4, 113.3, 123.7, 123.9, 127.9, 130.8, 134.5, 149.8, 150.7, 154.1, 178.0; HRMS (FTMS + pESI) m/z, calcd. for C₂₂H₂₃N₅O [M⁺]: 372.1824, found: 372.1834.
(E)-3-(Pyridin-4-yl)-1-(4-(4-(pyrimidin-2-yl)piperazin-1-yl)phenyl)prop-2-en-1-one (29)

Yield: 94%; yellow solid; mp 207-208 °C; 1H NMR (300 MHz, CDCl3) δ 3.48-3.52 (m, 4H, 2CH2), 3.70-3.74 (m, 4H, CH2), 6.60-6.66 (m, 2H, Ar-H), 6.92 (d, 2H, J 9.0 Hz, Ar-H), 7.27-7.28 (m, 1H, Ar-H), 7.44 (d, 1H, J 7.7 Hz, Ar-H), 7.47-7.53 (m, 1H, Ar-H), 6.86-7.76 (m, 2H, Ar-H and CH), 8.08 (d, 2H, J 9.0 Hz, Ar-H), 8.13 (d, 1H, J 15.2 Hz, Ar-H), 8.20 (dd, 1H, J 4.2, 1.1 Hz, Ar-H), 8.67 (d, 1H, J 3.3 Hz, Ar-H); 13C NMR (75 MHz, CDCl3) δ 44.7, 46.9, 107.1, 113.4, 113.7, 123.7, 123.9, 127.8, 130.8, 131.1, 134.5, 137.6, 139.3, 148.0, 149.8, 150.7, 154.0, 159.0, 187.3; HRMS (FTMS + pESI) m/z, calcd. for C23H22N4O [M]+: 372.1824, found: 372.1826.

Antitubercular assay

The strains of M. tuberculosis H37Rv (ATCC27294) were grown in the Ogawa-Kudoh medium for 10 days at 37 °C. For testing, a colony was removed and cultured in Middlebrook 7H9 broth (Difco®, Franklin Lakes, NJ, United States) supplemented with oleic acid, bovine serum albumin, dextrose and catalase (OADC enrichment BBL/Becton-Dickinson®, Franklin Lakes, NJ, United States), and it was also added 0.5% glycerol as carbon source and 0.5% Tween 80 (Sigma-Aldrich®, St. Louis, MO, USA) to prevent the appearance of lumps. The broth was maintained for 15 days at 37 °C. The bacterial suspensions were prepared and adjusted to the No. 1 McFarland scale. The stock solutions of tested extract, fractions, and isolated compounds were solubilized in DMSO (Sigma-Aldrich®, St. Louis, MO, USA) and diluted in Middlebrook 7H9 broth (Difco®, Franklin Lakes, NJ, United States) supplemented with bovine serum and OADC enrichment (oleic acid dextrose catalase) (BBL/Becton Dickinson®, Franklin Lakes, NJ, United States). Rifampicin and isoniazid were solubilized according to the manufacturer’s recommendations (Sigma-Aldrich®, St. Louis, MO, USA) and used as positive control drugs. The determination of antimycobacterial activity was performed using the resazurin microtiter assay. Briefly, 100 µL of Middlebrook 7H9 broth (Difco®, Franklin Lakes, NJ, United States) supplemented was dispensed in each well of a sterile flat-bottom 96-well plate, then serial dilutions were made of solutions in order to obtain different concentrations of tested compounds (0.98 to 250 µg mL⁻¹) and reference drug (0.004 to 1 µg mL⁻¹). After dilutions, 100 µL of bacterial suspension (5 × 10⁵ CFU mL⁻¹) was added to each well. Plates were incubated for 7 days at 37 °C, after this period, 50 µL of resazurin solution (Sigma-Aldrich®, St. Louis, MO, USA) diluted in 0.01% sterile water was added to each well and the samples were incubated for 24 h at 37 °C. The reading was performed based on the color change and the absorbance on a microplate reader TP-Reader (Thermo Plate®) at a wavelength of 492 nm. Each compound was analyzed in triplicate on alternate days. The MIC was defined as the lowest concentration that results in 90% inhibition of growth of M. tuberculosis. Several authors consider MIC values of...
plant samples less than 200 µg mL⁻¹ as being active against *M. tuberculosis*. However, for this to work, we suggest the following MIC values: inactive, ≥ 100 µg mL⁻¹; moderate, ≥ 10 and ≤ 100 µg mL⁻¹; and high, ≤ 10 µg mL⁻¹.

Cytotoxicity assay

Fibroblasts (NIH/3T3) obtained from Rio de Janeiro Cell Bank (Brazil) were seeded in 96-well plates (1 x 10⁴ cells mL⁻¹) and incubated with synthetic compounds at 37 °C, 5% CO₂ for 48 h at the concentrations of 0.25-250 µg mL⁻¹. Doxorubicin (Sigma-Aldrich®, St. Louis, MO, USA) was used as the reference drug at concentrations of 0.025-25 µg mL⁻¹. Cell growth was estimated by the sulforhodamine B colorimetric method (SRB). DMSO (Vetec®, Rio de Janeiro, RJ, Brazil) was estimated by the sulforhodamine B colorimetric method (SRB). DMSO (Vetec®, Rio de Janeiro, RJ, Brazil) was used as a negative control at the concentration necessary to solubilize the highest concentration of the test samples and did not interfere with cell viability. The percentage of growth was calculated as described by Monks et al. (half maximal inhibitory concentration on fibroblast cells) was determined by nonlinear regression analysis (Microcal Origin 6.0®33 and Microsoft Office Excel 2007®). Selectivity index (SI) was calculated according to de Medeiros et al.®

Supplementary Information

Supplementary data are available free of charge at http://jbcs.sbq.org.br as PDF file.

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Author Contributions

Adriano C. M. Baroni was responsible for the conceptualization, resources, writing original draft, review and editing; Júlio Croda for the resources and antimycobacterial experiments; Renata T. Perdomo for the resources and cytotoxicity experiments; Jefferson R. S. Oliveira for the synthesis of chalcones and writing original draft; Flora M. F. Moreira for the cytotoxicity experiments; Cristiane Y. K. Shiguemoto for the writing of the original draft; Amarith R. das Neves for the writing of original draft, review and editing; Sandro L. Barbosa and Palimécio G. Guerrero Jr. for the writing review and editing.

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