Prevalence of Campylobacter Species on Cattle Breeding Farms in Zimbabwe

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1. Introduction

Campylobacteriosis is an infection caused by the bacteria Campylobacter spp. and is associated with a number of disease conditions in cattle, primarily related to poor fertility in breeding cows [1]. Campylobacter fetus subspecies fetus and Campylobacter fetus subspecies venerealis are most commonly associated with early embryonic death and abortion [2], the latter of which causes bovine genital campylobacteriosis (BGC). Reproductive diseases of cattle, which result in infertility and abortion, cause significant economic losses [3]. Financial losses arise from extended calving intervals, increased age at first calving, low pregnancy rates, and temporary infertility in bulls [4]. In countries that maintain reproductive management based on natural mating with bulls such as Zimbabwe, mainly in beef herds where natural breeding prevails, the disease may be prevalent. In a study by Swanepoel et al. [5], BGC emerged as one of the most common identified causes of infertility and abortion in Rhodesia (now Zimbabwe). In 1988, a prevalence of 100% was recorded on a farm in Harare, Zimbabwe [6].

In this study, the prevalence of Campylobacter spp. in breeding bulls was investigated in samples submitted to the Central Veterinary Laboratory for diagnostic testing.

2. Materials and Methods

2.1. Study Animals and Diagnostic Tests. The Central Veterinary Laboratory (CVL), Zimbabwe, provided veterinary diagnostic testing services to farmers and public and private veterinary practitioners. The diagnostic tests carried out include tests for bovine sexually transmitted infections (STIs) such as Campylobacter spp. One hundred and four bulls of different ages and breeds were sampled from extensive beef production farms between September 2021 and February 2022. The bulls came from a total of seven farms. The farms are located in different provinces of Zimbabwe, that is, Mashonaland East, Harare, and Matabeleland North. Veterinarians submitted samples for different reasons...
including prebreeding examinations, surveillance, low pregnancy rates in cows, and abortions.

Skilled veterinarians carried out preputial scraping of the bulls to obtain samples for diagnostic testing. The collected washes were put in a 10 ml container with phosphate-buffered saline (PBS-pH 7.0) and sent to the CVL in cooler boxes for analysis within 24 hours.

2.2. Sample Analysis

2.2.1. DNA Extraction, Polymerase Chain Reaction, Gel Electrophoresis, and Visualisation. DNA was extracted from the samples using the Zymo Quick DNA Miniprep Plus kit (Zymo, USA) according to the manufacturer’s instructions. The concentration and purity of extracted DNA were checked using a Nanodrop spectrophotometer (Serva electrophoresis, Denmark).

25 µl reaction containing 9.3 µl of RNA-free water, 8.7 µl of the master mix (One Taq 2X Master Mix with standard buffer, New England Biolabs), 1 µl each of Campylobacter spp. specific primers (primer sequence F-5′ GGC TGA TCT ACG ATT ACT AGC GAT 3′ and R-3′ GCG CGC ATT AGA TAC CCT AGT AGT CC 5′), and 5 µl of the DNA template was used in each PCR tube. Nuclease-free water was used as a negative control. The presence of Campylobacter spp. DNA was determined using PCR reaction undertaken as previously described [7], with modifications in the number of cycles, that is, addition of 4 minutes of denaturation at 94°C to cycles 1–5 and reduction of annealing temperature from 37°C to 30°C. Final elongation was carried out at 60°C for 7 minutes instead of 10 minutes.

The PCR test was carried out using a thermocycler (Eppendorf Mastercycler, Nexus Gradient Thermal Cycler). Gel electrophoresis was carried out using 10 µl PCR products alongside a 100 kb ladder on 2% agarose gel containing a fluorescent dye in 1X TAE buffer at 90 volts for 1 hour. Visualisation of the gel was performed using a gel documentation system (Kodak Gel Logic System 100). The expected band size for Campylobacter spp. was 600 base pairs (bp).

2.3. Data Analysis. Data were entered into MS Excel (Microsoft, USA), and animal prevalence was determined by calculating the number of positive bulls divided by the total number of sampled bulls. Herd prevalence was calculated by the number of herds, which had at least one bull testing positive for Campylobacter spp., divided by the total number of herds tested.

3. Results

3.1. Prevalence of Campylobacter spp. The animal-level prevalence of Campylobacter spp. was 25.96%, and all the positive samples came from four (57.14%) of the 7 herds. Only three (42.9%) herds out of 7 herds had all the bulls testing negative for Campylobacter spp. (Table 1).

4. Discussion

The cattle population in Zimbabwe is estimated to be 5.2 million (Director, Division of Veterinary Technical Services). In recent years, farmers have lost cattle due to drought and diseases. The current thrust of the Zimbabwean government is to replenish and increase the national herd [8]. One of the ways the government has been advocating for this increase is through improvement of cattle breeding through natural breeding and artificial insemination. However, these efforts are being hampered by reproductive diseases such as those caused by Campylobacter spp. The disease is known to hamper economic growth where farmers incur huge losses [9]. The literature on the prevalence of Campylobacter spp. in Southern African and other developing countries is scarce.

The current prevalence and impact of campylobacteriosis in Zimbabwe are unknown. The last known study on prevalence of Campylobacter fetus subspecies venerealis was performed in 1989 [6], where 100% of the bulls had BGC. In recent years, veterinarians and farmers have reported existence of infertility problems in cows, and this is suspected to be due to sexually transmitted infections such as BGC. Furthermore, in the past years, movement of cattle increased due to the land reform program, resettlements, and social and cultural reasons [10]. Premovement permits were focusing mainly on foot and mouth disease, while production and reproductive diseases were not being checked. This could spread diseases such as campylobacteriosis across the country.

In this study, only 3 herds out of 7 had all bulls testing negative for Campylobacter spp. The high prevalence of Campylobacter spp. infection noted in this study warrants further research on campylobacteriosis in Zimbabwe. It is important to note that the positive Campylobacter spp. samples identified in the CVL laboratory were the first in over ten years (Personal Communication, Technologist-in-charge, CVL, Zimbabwe). In the past decade, farmers and animal movement regulatory authorities did not focus much on bovine sexually transmitted infections, and this may have resulted in the introduction of the infection into herds unknowingly through cattle movement, loaning, and sharing of bulls [9]. The difference in prevalence across the herds in the country could be due to management systems. Previous studies have shown that prevalence of Campylobacter spp. is affected by climate change, locations, animal breeds, and age.

As STIs decrease productivity of cattle by inducing reproductive losses, reduced conception rates, extended calving seasons, and increased costs, replacing bulls and preventing loss of genetic potential due to culling the disease are of great economic importance. A comprehensive study should be carried out in Zimbabwe to determine the epidemiology of Campylobacter spp. with the aim of developing intervention to reduce such losses.
5. Conclusion and Recommendation

The study reports a high prevalence of Campylobacter spp. in bull herds in Zimbabwe. There is a need for further surveillance of Campylobacter spp. in the country using advanced molecular techniques, which will be able to elucidate the subspecies responsible. Veterinary authorities should enforce strategies which can minimize the spread of cattle STIs in the country.

Data Availability

Raw data supporting this article are available from the corresponding author upon request.

Disclosure

The research was carried out as part of the work performed at the Central Veterinary Laboratory, Zimbabwe.

Conflicts of Interest

The authors declare no conflicts of interest.

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Table 1: Results of samples tested for Campylobacter spp. in the study.

| Herd | Number of *Campylobacter* spp. positive samples | Number of *Campylobacter* spp. negative samples | Total number of samples submitted | Animal prevalence (%) |
|------|-----------------------------------------------|-----------------------------------------------|--------------------------------|-----------------------|
| 1    | 4                                             | 13                                            | 17                             | 30.77                 |
| 2    | 0                                             | 8                                             | 8                              | 0                     |
| 3    | 8                                             | 11                                            | 19                             | 42.11                 |
| 4    | 0                                             | 20                                            | 20                             | 0                     |
| 5    | 0                                             | 2                                             | 2                              | 0                     |
| 6    | 1                                             | 18                                            | 19                             | 0.05                  |
| 7    | 14                                            | 5                                             | 19                             | 73.68                 |
| Total| 27                                            | 77                                            | 104                            | 25.96                 |