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Molecular Typing and Antimicrobial Susceptibility of Methicillin-Resistant *Staphylococcus aureus* Isolated from Bovine Milk in Tanzania

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Methicillin-resistant *Staphylococcus aureus* (MRSA) in raw milk can be transmitted from animals to humans, and in Tanzania raw milk is sold in local markets and consumed as purchased. This study was performed to determine the molecular characteristics and antimicrobial susceptibility pattern of MRSA strains isolated from raw bovine milk sold at local markets in Tanzania. A total of 117 raw milk samples were cultured on Baird-Parker medium to isolate *S. aureus* and PCR was used for amplification of *gltB* gene for *S. aureus* identification and the presence of *mecA* gene for methicillin-resistant strains. Coagulase-negative (CN) *S. aureus* were reconfirmed using tube coagulase, DNase, and API Staph tests. MRSA isolates were *spa* typed whereas antimicrobial susceptibility testing was performed by the disc diffusion method. Forty-six coagulase positives (CP) and two CN *S. aureus* were identified. Most strains were resistant to penicillin (72%), and 3 isolates: 2 CN *S. aureus* and 1 coagulase-negative Staphylococci (CNS), were phenotypically resistant to vancomycin, oxacillin, and cefoxitin and were confirmed to carry *mecA*. Resistance to clindamycin, trimethoprim-sulfamethoxazole, and tetracycline was 23.9%, 30.4%, and 41.3%, respectively. Twelve isolates exhibited multidrug resistance; however, only one *mecA* positive strain among the three was typeable and belonged to *spa* type t2603. This study reports for the first time the presence of CN variant of MRSA, which was assigned the spa type t2603, and the presence of multidrug resistant *S. aureus* isolates from bovine milk in Morogoro, Tanzania.

1. Introduction

*Staphylococcus aureus* is an important opportunistic pathogen both in humans and in dairy cattle. It is also a common cause of mastitis in dairy cows [1], a primary reason for antibiotic use in dairy farms. The use of antimicrobial agents in dairy farms as well as in other food animal production systems is a major concern in the emergence of resistant zoonotic bacterial pathogens [2, 3]. Methicillin-resistant *Staphylococcus aureus* (MRSA) has emerged as a major cause of health care-associated (HA) and community-associated (CA) infections [4]. In addition to that, infection and colonization by MRSA have been well documented in several animal species and mostly caused by livestock-associated MRSA strains [5] and are frequently multidrug resistant (MDR). This can result in higher costs, longer treatment time, and higher rates of hospitalization and comorbidities [6].

The presence of MRSA in bovine milk and dairy environments poses potential risk to farm workers, veterinarians, and farm animals that are exposed to infected cattle. This
study was conducted in Morogoro, which is one of the leading regions in livestock keeping in Tanzania, to investigate the occurrence of MRSA in milk samples. In Morogoro, milk collected from dairy farms is distributed to various local sales’ points and markets, from which samples were taken for investigation.

2. Methods

2.1. Study Area and Study Design. This study was carried out between January and June 2015 in Morogoro Municipality, which had a total population of 316,603 persons [7]. The current study was a cross-sectional design that involved 18 of the 29 Wards. In each of the selected Wards, sales points and local shops where raw milk is sold were randomly selected. A total of 117 milk samples (4 to 8 samples from each Ward), of which had a total population of 316,603 persons [7]. The study was conducted in Morogoro, which is one of the leading regions in livestock keeping in Morogoro, to investigate the occurrence of MRSA in milk samples. In Morogoro, milk collected from dairy farms is distributed to various local sales’ points and markets, from which samples were taken for investigation.

2. Methods

Table 1: Summary of primers used in this study.

| Primer       | Sequence (5’-3’)                      | Amplicon size (bp) | Specificity | Reference(s) |
|--------------|---------------------------------------|--------------------|-------------|--------------|
| MecA147-F    | GTG AAG ATA TAC CAA GTG ATT            | 147                | meca        | [8]          |
| MecA147-R    | ATG GCG TAT AGA TTG AAA GGA           |                    |             |              |
| Sa442-1      | AAT CTT TGT CGG TAC ACG ATA TTC TTC ACG | 108                | Species-specific Target | [9]          |
| Sa442-2      | CGT AAT GAG ATT TCA GTA GAT AATACA ACA |                    |             |              |
| 1095-F       | AGA CGA TCC TTC GGT GAG C             | Variable           | Spa         | [10]         |
| 1517-R       | GCT TTT GGA ATG TCA TTT ACT G         |                    |             |              |

2.2. DNA Extraction. DNA was extracted using the boiling method as described by [11] with some modifications. Briefly, three to five bacterial colonies were added to 1.5 ml Eppendorf tubes containing 200 μl of nuclease-free water. The tubes were boiled in water bath at 99°C for 10 min. After centrifugation at 30,000 xg for 1 min, 3 μl of supernatant was used as template in a 20 μl PCR mixture.

2.3. PCR Detection of S. aureus. The detection of S. aureus was performed using primers (Macrogen Inc., Seoul, South Korea) which were species specific for S. aureus (Table 1). The PCR mixture contained an aliquot of 3 μl bacterial DNA template, primers, and distilled water to a total volume of 20 μl into AccuPower® PCR PreMix tubes (Bioneer Inco., South Korea). The mix contained 1 U Taq DNA polymerase and 250 μM each of dNTP, 10 mM Tris-HCl (pH 9.0), 30 mM KCl, 1.5 mM MgCl₂, stabilizer, and tracking dye. Each primer concentration was 0.4 μM derived from a chromosomal DNA specific for S. aureus amplifying 108 bp product, which codes for the enzyme glutamate synthase (gltB). The PCR mixtures were incubated in a TAKARA PCR Thermal Cycler Dice Gradient TP600 (Takara Bio, Tokyo, Japan). PCR conditions were as described by [9] with initial denaturation step at 95°C for 5 mins, and 35 cycles of amplification at 95°C for 30 sec, with annealing at 55°C for 30 sec, extension at 72°C for 30 sec, and final extension at 72°C for 5 min and a hold at 4°C.

2.5. PCR Detection of mecA Gene. The detection of mecA gene was carried out as a single target PCR amplification using the primer pairs listed in Table 1. All S. aureus and CNS isolates were screened for mecA gene for the genotypic identification of MRSA. The primer and PCR conditions were obtained from [8] with some modifications. The initial primer concentration was 0.046 μM and amplicon size was 147 bp. The PCR was run in 20 μl of AccuPower PCR PreMix tubes (Bioneer Inco., South Korea) containing 3 μl template DNA, with cycling parameters beginning with an initial denaturation step at 95°C for 5 min followed by 35 cycles of 95°C for 30 sec, 52°C for 45 sec, and 72°C for 30 sec, ending with a final extension step at 72°C for 7 min and a hold at 4°C.

2.6. Spa Typing of mecA Carrying Isolates. For spa gene PCR, primer pair in Table 1 used for typing were derived from [10]. The PCR mixture contained 20 μl of AccuPower PCR PreMix with 3 μl bacterial DNA and concentration of 1 μl each of the primers with variable product size (bp). PCR conditions were 94°C for 3 min; 35 cycles each of 94°C for 30 sec, 50°C for 30 sec, and 72°C for 60 sec; and a final extension at 72°C for 5 min. PCR products were purified using GeneJET purification kit (Thermo scientific). Samples were sequenced with the same primers used in PCR. Sequencing reactions used BigDye v3.1 sequencing mix (Applied Biosystems) and were cycled using 30 cycles of 96°C for 10 sec, 50°C for 5 sec, and 60°C for 2 min. Products were purified and separated on an ABI 3730 DNA Analyzer (Applied Biosystems). Chromatograms were analyzed using Ridom StaphType v2.2.1 software (Ridom GmbH).

2.7. Antimicrobial Susceptibility Test. The Kirby-Bauer Disk Diffusion Susceptibility test was used to obtain the antimicrobial resistance profile of the isolates for clindamycin (2 μg), vancomycin (30 μg), trimethoprim-sulfamethoxazole (25 μg), tetracycline (30 μg), penicillin G (10 IU), oxacillin (1 μg), and cefoxitin (30 μg). Isolates were considered to
Table 2: The *S. aureus* isolated from milk in the Morogoro Municipality.

| Wards (Codes) | Samples analyzed | Positive samples | % of positive sample |
|---------------|------------------|------------------|----------------------|
| Boma (1)      | 7                | 1                | 2.1                  |
| Mazimbu (2)   | 8                | 3                | 6.3                  |
| Mwebesongo (3)| 8                | 6                | 12.5                 |
| Msamvu (4)    | 8                | 8                | 16.7                 |
| Kihonda (5)   | 8                | 1                | 2.1                  |
| Kichangani (6)| 4                | 2                | 4.2                  |
| Kilakala (7)  | 6                | 4                | 8.3                  |
| Mafiga (8)    | 6                | 2                | 4.2                  |
| Kiwanjacha Ndege (9) | 8 | 3 | 6.3 |
| Sabasaba (10) | 5                | 3                | 6.3                  |
| Chamwino (11) | 7                | 3                | 6.3                  |
| Mafisa (12)   | 6                | 3                | 6.3                  |
| Mbuyuni (14)  | 5                | 2                | 4.2                  |
| Mji Mpya (15) | 6                | 2                | 4.2                  |
| Kingolvira (16)| 4               | 1                | 2.1                  |
| Tungi (17)    | 9                | 1                | 2.1                  |
| Mkundi (18)   | 4                | 2                | 4.2                  |
| Magadu (19)   | 8                | 1                | 2.1                  |

Total number of *S. aureus* isolated from Municipality (*n* = 48) and the proportion of isolates from each Ward.

Table 3: Phenotypic and genotypic identification of methicillin-resistant isolates.

| Isolates | Phenotype | Genotype mecA gene |
|----------|-----------|--------------------|
| S. aureus | 6.52% (*n* = 3) | n = 2 |
| CNS      | 19.05% (*n* = 8) | 2.38% (*n* = 1) | n = 1 |

be resistant to methicillin if they were resistant to both oxacillin and cefoxitin, with particular emphasis to cefoxitin which is a better inducer of *mecA* gene. Moreover, cefoxitin disk diffusion tests give clearer endpoints and are easier to read than oxacillin [12]. *S. aureus* ATCC 29213 was used as reference strain and interpretation was done according to standard guidelines of Clinical and Laboratory Standards Institute [13].

3. Results

3.1. Prevalence of *S. aureus* in Raw Milk Samples. A total of 117 raw milk samples were analyzed, of which 75 (64%) yielded coagulase-positive staphylococci (CPS) and 42 (36%) coagulase-negative staphylococci (CNS) presumptive isolates. PCR detected 46 *S. aureus* (Figure 1) among CPS and 2 *S. aureus* among the CNS, giving a 41% prevalence of *S. aureus* in raw milk in the Morogoro Municipality (Table 2). The two CN *S. aureus* were reconfirmed with tube coagulase, DNAse, and API Staph tests.

3.2. Antimicrobial Susceptibility Test. The overall resistance to clindamycin, vancomycin, trimethoprim-sulfamethoxazole, tetracycline, penicillin G, oxacillin, and cefoxitin was 23.9%, 2.2%, 30.4%, 41.3%, 71.7%, 6.5%, and 4.4%, respectively. Resistance to both oxacillin and cefoxitin was seen in three isolates: 2 CN *S. aureus* and 1 CNS (Table 3) and they were confirmed to carry *mecA* (Figure 2).

Twelve (26.1%) of the CP *S. aureus* isolates exhibited multidrug resistance (MDR), whereas none of the CNS isolates were MDR (Table 4).

3.3. Staphylococcus Protein A (spa) Typing. Among the 3 isolates containing *mecA*, 1 CN *S. aureus* contained a *spa* gene and the primers produced a band of 1150 bp based on
Table 4: Multidrug resistance pattern of S. aureus and CNS isolated from raw milk.

| Number of antibiotic agents | S. aureus isolates (N = 46) | CNS isolates (N = 42) |
|-----------------------------|-----------------------------|----------------------|
| Number | Percent | Number | Percent |
| 0 | 9 | 19.6% | 33 | 78.6% |
| 1 | 12 | 26.1% | 5 | 4.8% |
| 2 | 13 | 28.3% | 4 | 11.9% |
| 3 | 9 | 19.6% | 0 | 0% |
| 4 | 2 | 4.4% | 0 | 0% |
| 5 | 1 | 2.2% | 0 | 0% |

4. Discussion

This study found the prevalence of S. aureus in raw milk to be 41.0%, with samples from Msamvu (16.7%) and Mwembesongo (12.5%) being the most contaminated while those from Tungi (2.1%), Magadu (2.1%), Kihonda (2.1%), and Boma (2.1%) were the least contaminated. This frequency of contamination is similar to that reported in studies conducted in Morocco, Brazil, Ethiopia, and Kenya, which found frequencies of 40%, 68%, 48.7%, and 30.6%, respectively [14–17].

The milk samples were collected from sale points and open markets; therefore, the high frequency of contamination could be related to poor hygiene practices in handling milk at various stages from farms to the markets. It is also possible that the health status of the animals may have contributed to the occurrence of some of the isolates recovered, as previous studies have associated this with mastitis and other animal infections [18, 19]. However, this study did not investigate the health status of the dairy cows.

A significant variability was noted in susceptibility of S. aureus to the tested antibiotics, with lowest resistance to oxacillin (6.5%) and cefoxitin (4.4%), whereas the highest resistance was to penicillin (71.7%). Isolates that were methicillin-resistant, hence resistant to both oxacillin and cefoxitin, were 2 (4.4%) S. aureus and 1 (2.4%) CNS. These variations are most probably related to frequency of use, which is associated with cost and availability. Compared to CNS, S. aureus had slightly higher levels of resistance for all five classes of antibiotics tested. In Morogoro the most frequently used antibiotics in the livestock industry are oxytetracycline and sulphur-based antibiotics [20, 21]. These drugs are cheap and available over the counter and are frequently used inappropriately. The high level of resistance to the tested antibiotics has also been reported in human population residing in areas where the study was conducted [22], prompting suggestion of potential transmission of antimicrobial resistance genes between bacteria found in humans and animals. In Morogoro, a number of factors compound the problem of antimicrobial resistance in zoonotic infections. These include (i) tendency for animal owners to stock drugs in their houses and engaging unskilled people such as farmers/peasants themselves and animal attendants to treat animals [23] and (ii) high degree of drugs abuse/misuse by livestock keepers through failure in observing the recommended therapeutic doses and arbitrary drug combinations and nonobservance of withdraw periods [24]. Others include lack of basic knowledge of the concept of antibiotic resistance among livestock keepers [21].

In the present study PCR was conducted to detect mecA gene in all isolates. In CP S. aureus it was found that none of the isolates contained the gene while 1 CNS and 2 CN S. aureus harboured it. This study revealed the prevalence of CN-MRSA and MRCNS to be 4.2% and 2.4%, respectively. This finding is in accordance with previous studies conducted elsewhere, 4.0% [6], 4.8% [25], and 0 to 7.4% [26]. Identification of MRSA in milk in this study emphasizes the need for increased public awareness regarding safe food handling to help prevent cross-contamination [27] and urges for public health interventions, including decreasing the use of antibiotics [28, 29]. The 2 CN-MRSA showed resistance to only one class of antimicrobials although mecA gene is believed to confer resistance to most currently available beta-lactam antibiotics [30]. However, not all mecA positive clones are resistant to methicillin, and overall resistance level in a population of MRSA depends on efficient production of PBP-2a, which is modulated by a variety of chromosomal and extrachromosomal factors [31]. This explains why MRSA resistance levels range from phenotypically susceptible to highly resistant [30]. According to Tavares [32], the resistance to antibiotics is determined by not only the presence of
resistance genes, but also the expression of these genes. The *S. aureus* may be pathogenic or nonpathogenic with the pathogenic strains usually exhibiting coagulase-positivity and causing disease in their hosts [33].

In conclusion, the prevalence of CN-MRSA and MRCNS in bovine milk was found to be 4.2% and 2.4%, respectively. The study reports for the first time the presence of presumptive coagulase-negative variant of MRSA and MRCNS in raw milk in Morogoro, Tanzania. Among the three *meca* positive isolates, hence, only one of the coagulase-negative variants of MRSA was typeable and was assigned the spa type t2603. Based on the results of the current study, it is important to characterize further the CN-MRSA and MRCNS isolates to determine whether the isolates were clonally related. Furthermore, future studies for MDR *S. aureus* strains should be screened for detection of the novel *mecA* homologue *mecC* gene and other antibiotic resistance genes such as *mecB*. Further studies are also required to identify the origin of these MRSA strains to figure out whether they originated from milk sellers (during handling) or the animals as livestock-associated MRSA (e.g., from mastitis).

**Abbreviations**

| Abbreviation | Description |
|--------------|-------------|
| CLSI         | Clinical Laboratory Standards Institute |
| CN-MRSA      | Coagulase-negative MRSA |
| CNS          | Coagulase-negative staphylococci |
| DNA          | Deoxyribonucleic acid |
| MRCNS        | Methicillin-resistant coagulase-negative staphylococci |
| MRSA         | Methicillin-resistant *Staphylococcus aureus* |
| PBP-2a       | Penicillin-binding protein 2a |
| PCR          | Polymerase chain reaction |
| SUA          | Sokoine University of Agriculture |

**Ethical Approval**

The study protocol was reviewed and approved by the SUA’s College of Veterinary and Medical Sciences’ Research, Ethics and Publications Committee. Before participating in the study, the village authorities and villagers were given detailed explanations about the objectives and methodologies. Participation was voluntary and participants could withdraw from the study at any time. Participants were also informed that their identities and personal information would be kept strictly confidential.

**Consent**

If participants agreed to participate, their consent was taken in written form (signed) or in thumb-printed form for those who were illiterate.

**Conflicts of Interest**

The authors declare that they have no conflicts of interest.

**Authors’ Contributions**

All authors contributed equally to this paper from proposal development up to implementation. The final manuscript was read and approved by all authors.

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