Review Article
Role of the Synthetic B1 Vitamin Sulbutiamine on Health

Bernardo Starling-Soares,1 Pedro Carrera-Bastos,2 and Lucien Bettendorff3

1Extreme Sports Nutrition Institute, Belo Horizonte, Minas Gerais, Brazil
2Center for Primary Health Care Research, Lund University/Region Skåne, Skåne University Hospital, Malmö, Sweden
3Laboratory of Neurophysiology, GIGA-Neurosciences, University of Liege, 4000 Liege, Belgium

Correspondence should be addressed to Bernardo Starling-Soares; bernardostarling@gmail.com

Received 22 May 2019; Accepted 4 February 2020; Published 30 April 2020

Copyright © 2020 Bernardo Starling-Soares et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Sulbutiamine is a thiamine derivative developed in Japan in the mid-60’s as a beriberi treatment drug. Since then, different potential applications have been described. For instance, there is some evidence that sulbutiamine can have anti-fatigue, nootropic, and antioxidant effects, which led to its use as a sport supplement (although some authors argue it is actually a masking doping strategy). Moreover, this molecule has been proposed as a possible treatment for some microsporidial infections and even for certain types of cancer. Despite these potential effects, sulbutiamine is still a relatively unknown molecule, which justifies the present review, where we discuss its history and the existing literature on its health applications. We conclude that there is a great potential for sulbutiamine use, well beyond its first described function (to increase thiamine tissue concentration). Indeed, new mechanisms of action have been found, mainly associated with its derivatives. Nevertheless, and although the research on sulbutiamine started 50 years ago, only a limited number of studies were conducted during this time frame. As so, methodological concerns need to be addressed and new studies are necessary, especially randomized controlled trials. Only then will the full potential of this versatile molecule be identified.

1. Introduction

Sulbutiamine (Figure 1) results from the fusion of two thiamine (Figure 1) molecules linked by a disulfide bridge after the opening of their respective thiazolium rings and esterification of the primary alcohol of each half-molecule by an isobutyryl group [1]. These modifications increase the lipophilic characteristic of this derivative compared to thiamine. Regarding its chemical names, these are: isobutyrylthiamine disulfide, [4-[(4-amino-2-methylpyrimidin-5-yl)methyl-formyl-amino] -3-[2-[(4-amino -2-methyl-pyrimidin-5-yl)methyl-formyl-amino]-5-(2-methylpropanoyloxy) pent- 2-en-3-yl]disulfanyl-pent-3-enyl 2-methylpropanoate), bisibutiamine, and thiamine di(2-methylpropionate) disulfide [1–5]. It is sold under the brand names Arcalion (Laboratoires Servier) and Enerion (Egis Pharmaceuticals) [6, 7] and is commonly mistaken for butyrylthiamine, thiamine propyl disulfide, butyryl thiamine disulfide, and benfotiamine (thiamine precursor with different molecular structures and chemical/biological properties, Figure 1) and even for Vitaberin (actually a berberin-based complement-Ellab Laboratory). Sulbutiamine is also different from thiamine tetrahydrofurfuryl disulfide (Fursultiamine, Figure 1), another pharmacologically active thiamine disulfide compound known to promote voluntary activity through dopaminergic activation in the medial prefrontal cortex [8].

The synthesis of sulbutiamine and the first controlled studies on this molecule were made in Japan in the mid-60s (Tanabe Pharmaceutical-O-isobutyrylthiamine disulfide) [9]. It was mainly developed, along with other lipid soluble thiamine derivatives, for the treatment of beriberi, which was a public health problem in Japan until the 2nd World War [10, 11]. Then, in the 70s, a French pharmaceutical laboratory launched sulbutiamine under the brand name Arcalion labeled as an anti-asthenia drug, notwithstanding the fact that previous work had been focused on intestinal, urological, and cardiac actions of the near cousin of
sulbutiamine, BuTDS (O-butyrylthiaminedisulfide) [12–17].
Until that time, there were only assumptions about the intracellular (intraerythrocytary) metabolism of sulbutiamine based on Kawasaki’s studies with thiamine propyl disulfide [9, 16, 17]. But since then, it has been established that the uptake of thiamine derivatives by erythrocytes runs in parallel with blood retentivity, and that among the O-acyl thiamine disulfides tested, O-butyrylthiamine was the most efficient. Indeed, after parenteral administration of O-butyrylthiamine, an increase in the half-life of thiamine seems to be the result of its uptake by red blood cells [9]. This observation seems to have been the initial argument for the use of sulbutiamine in thiamine deficiency disorders.

Later, in vivo experiments proved that sulbutiamine can be transformed into thiamine derivatives after the reduction of the disulfide bond and regeneration of the closed thiazolium rings, yielding mainly two molecules of isobutyrylthiamine (Figure 2). Chromatographic behavior of plasma thiamine after the injection of sulbutiamine proved to be exactly the same as for "genuine" thiamine, implying that the isobutyryl groups are removed. This conclusion is more clearly supported by the verified formation of ThMP (thiamine monophosphate), since the isobutyryl group would block the phosphate acceptor group (alcohol group) essential for thiamine phosphate derivatives formation [1].

Another important observation was that sulbutiamine increases the levels of thiamine and thiamine phosphate esters in the brain, thus having the potential to impact asthenia and chronic fatigue [1, 7, 18, 19]. In addition to thiamine, at least four phosphorylated thiamine derivatives have been described in living organisms: thiamine monophosphate (ThMP), thiamine diphosphate (ThDP, Figure 1), thiamine triphosphate (ThTP), and the recently discovered thiamine adenine nucleotide [20, 21]. But the main forms of vitamin B1 in plasma are thiamine and its derivative ThMP, which are transported into cells by specific transporters. ThMP can then be hydrolyzed to thiamine and subsequently pyrophosphorylated to ThDP by thiamine pyrophosphokinase (EC 2.7.6.2). Of interest, ThDP is the most researched thiamine derivative, since earlier studies have clearly demonstrated its role as a cofactor for mitochondrial pyruvate and alpha-ketoglutarate dehydrogenases, as well as for cytosolic transketolase, thereby affecting multiple functional systems [20].

Despite its importance, thiamine transport across cell membranes is a relatively slow process, hence the rationale
for the development of lipid soluble thiamine precursors able to freely diffuse into the cells and, especially, across the blood brain barrier. Indeed, because of the role of ThDP as a coenzyme in oxidative metabolism, brain is particularly affected during thiamine deficiency. In this regard, studies conducted in animal models (e.g., rats) have suggested that there are at least two different ThDP pools in the brain: a small pool of high turnover, precursor for ThMP and ThTP, and the larger coenzyme pool with increased half-life (probably because of its binding to apoenzymes) [20]. It is not clear whether these two pools are the result of a compartmentmentation between different cell types (e.g., neuronal and glial cells) or whether they coexist in the same cell type. As it has been shown that in cultured cells of neuronal origin, two ThDP pools (cytosolic and mitochondrial) coexist in the same cell line, the latter is the most probable [20].

ThTP is another thiamine derivative found in many cell types, including bacteria, but it is generally of minor abundance compared to ThDP. Interestingly, administering sulbutiamine to rats leads to increased levels of ThTP in brain tissue. ThTP seems to be preferentially associated with plasma, mitochondrial, and nuclear membranes in neurons, its synthesis previously being related to an “essentially and still not described specific membrane-associated enzyme in the brain” and presenting a role in membrane permeability modulation [1, 22]. Bettendorff and colleagues have shown that mitochondria isolated from rat brain cells synthesize ThTP from ThDP and P1 using the proton-motive force. This shows that (1) $F_0F_1$-ATPase is a necessary gear of this process and (2) ThTP synthesis is not ATP-dependent but actually $P_1$-dependent [22, 23]. ThTP is also synthesized by cytosolic adenylate kinase in skeletal muscle and electric organ and in non-excitable tissues, where this enzyme is particularly abundant [24, 25].

Sulbutiamine can also have neuroprotective effects, mainly because of its thiol content, which can affect antioxidant status. It is well known that the global thiol concentration is important in the regulation of the cellular redox status, since it provides a large antioxidant pool, consisting of (1) thiols bound to proteins, (2) sulfur bound in the disulfide bridges, and (3) free thiol groups, mainly in the form of reduced glutathione (GSH) [26]. Interestingly, several thiol containing compounds, including sulbutiamine, have been shown to upregulate GSH [27], which could prevent oxidative stress in brain cells. However, the mechanisms of action remain controversial.

Sulbutiamine is also a potent inhibitor of triosephosphate isomerase of the protozoan Encephalitozoon intestinalis causing microsporidiosis, as shown by Garcia-Torres and colleagues [28]. Sulbutiamine specifically inactivates the protozoan enzyme through interaction with cysteine residues, without affecting the human enzyme. This raises the possibility of using sulbutiamine in the treatment of microsporidiosis [28]. Since its development, sulbutiamine has been tested for the treatment of several diseases, with mixed results. In retrospect, sulbutiamine was first used in neurochemical studies [29, 30] and, more recently, it has been shown to improve fatigue in patients with multiple sclerosis (MS) who were already medicated for the disease (e.g., Fingolimod; β interferons; Glatiramer acetate; Azathiopurine) [31]. Despite the methodological limitations of this study, this is an important observation and opens the field for sulbutiamine use in various conditions, particularly in those characterized by an impaired intracellular and central nervous system thiamine metabolism [31, 32]. Overall, sulbutiamine appears to have various potential effects, such as improvement of asthenic syndromes, modulation of the wakefulness state (in extreme high doses), enhancement of long-term memory formation, chronic postinfectious fatigue amelioration, anti-ischemic effect, psycho-behavioral inhibition in major depression, modulatory effect on glutaminergic and dopaminergic cortical transmission, improvement of diabetic polyneuropathy and psychogenic erectile dysfunction parameters, reduction of amnesic effects of drug-induced schizophrenia, modulation of Alzheimer’s disease, and treatment for microsporidiosis caused by the intracellular parasite Encephalitozoon intestinalis (Supplementary Table S1) [3, 6, 7, 19, 28–30, 33–39].

2. Studies Included in This Review

The purpose of this review is to thoroughly describe sulbutiamine and its health applications, and to give directions for future research. As such, a web-based literature search was performed to identify all papers until January 2020 citing the following search terms: Sulbutiamine or Sulbuthiamine; Isobutyrylthiamine Disulfide; Arcalion; Enerion; Bisibutiamine or Bisibulthiamine; [4-[(4-amino-2-methyl-pyrimidin-5-yl) methyl-formyl-amino]-3-2-[(4-amino-2-methyl-pyrimidin-5-yl)methyl-formyl-amino]-5-(2-methylpropionoloxyl)pent-2-en-3-yldisulfanyl-pent-3-etyl]2-methylpropanoate; bis-(isobutyrylxy-2-ethyl)-1-N-(amino-4-methyl-2-pyr-imidyl-5) methyl formamido-2-propene-1-yl disulfide. The databases used were PubMed/MEDLINE and ScienceDirect. References of the eligible articles were then verified manually seeking potential additional articles that could have been missed by the electronic search. A total of 85 papers/book chapters/posters were found, but 67 of these were classified as “not related to the health and medical uses of sulbutiamine.” As a result, only 20 citations were included in the results of this review (Supplementary Table S1), although several papers related to sulbutiamine were presented here.

2.1. Sulbutiamine and Thiamine Phosphate Metabolism.

After chronic intraperitoneal injection of sulbutiamine in rats (52 mg/kg for 14 days), ThDP was found to be the most abundant thiamine compound in the renal cortex, the medulla oblongata (including the pons), the cortex (telencephalon), the cerebellum, and the hippocampus [1]. In brain extracts, its proportion ranged from 55.6% of total thiamine in the cytosolic fraction to 90.4% in the homogenate [20]. Interestingly, a more recent study from Sambon and colleagues reported that the maximum ThDP content in cultured neuroblastoma cell line grown in a thiamine-restricted medium is achieved more rapidly with the addition of sulbutiamine, when compared to thiamine and benfotiamine [40]. As for ThTP, it exists in
2.2. Neurochemical Actions of Sulbutiamine. Using very high doses (300 mg/kg/day) of sulbutiamine in Rhesus Monkeys (*Macaca mulatta*), Balzamo & Vuillon-Cacciuttolo have observed an increase in the vigilance pattern [29]. Nevertheless, it is not entirely clear whether these effects were due to increased brain thiamine phosphate levels or to another compound [20]. Whatever the exact mechanism, however, other studies have confirmed the neurological effects of sulbutiamine. For instance, when applied intraperitoneally in mice, at doses close to human therapeutic doses, sulbutiamine had modulatory effects on glutamatergic and dopaminergic central transmission changing the turnover of both receptors and neurotransmitters. It should, however, be mentioned that these effects appear to depend on whether it is applied chronically or acutely. As an example, after administering this molecule to rats for 5 days, there was a significant increase in the density of dopamine D1 receptor binding sites in prefrontal and anterior cingulate cortex (+26% and +34%, respectively), as well as a significant decrease in the density of kainate binding in the cingulate cortex, the nucleus accumbens (~36% and ~28%, respectively), striatum, and hippocampus. On the other hand, the acute administration of sulbutiamine decreased the density of kainate binding sites in prefrontal and cingulate cortex, reduced 3,4-dihydroxyphenylacetic acid (DOPAC) levels (~30%) in the prefrontal cortex, and decreased dopamine (DA) and DOPAC levels (~34% and ~26%) in the cingulate cortex [36].

2.3. Memory and Sulbutiamine. Chronic administration of moderate to high doses of sulbutiamine to BALB/c mice increased sodium-dependent high affinity choline uptake by 10% in hippocampal neurons and improved performance in a partial acquisition task but not the capacity of acquisition in a single test (learning rate) [30]. Since the hippocampal cholinergic system is associated with long-term memory formation, these results prove that moderate to high doses of sulbutiamine are capable of enhancing long-term memory formation in mice. Again, it is not yet known whether these properties are mediated through the action of thiamine phosphate derivatives or are the result of thiamine disulfide specific reactions [20]. Independently of the exact mechanism, when tested for its capacity to interfere with the detrimental effects of dizocilpine-induced amnesia, on acquisition, consolidation, and memory retention of rodents, the chronic use of sulbutiamine showed memory improvement in an object recognition task and reduced the disruption of working memory and the deficit of acquisition of information stored in episodic memory [38]. And in one of the few human trials ever conducted that evaluated memory parameters, sulbutiamine (400–600 mg/day), in association with a cholinesterase inhibitor treatment (donepezil 5–10 mg/day), improved episodic memory, attention, and daily activities, when compared with donepezil alone. But when comparing the effects of sulbutiamine and donepezil separately, a similar improvement between groups was seen only in the test evaluating attention. This was a multicentric, randomized, and double-blind trial engaging early-stage Alzheimer disease patients [39].

2.4. Fatigue and Sulbutiamine. Since the beginning of its commercialization, sulbutiamine has been considered a nootropic molecule. Following Micheau’s observation that sulbutiamine exhibits a cholinergic hippocampal modulation and that this could be linked to partial memory improvement in mice, this drug started to be prescribed to treat fatigue conditions [30, 31]. But Tiev and colleagues showed, in 1999, in a randomized, double-blind, parallel group, placebo-controlled study, that although an improvement was observed on the 7th day, doses of 600 mg/day did not have a persistent effect at the end of 28 days, in women suffering from post-infection asthenia [19]. This is in contrast to an Indian prospective uncontrolled, non-randomized, commercial, observational study, where 52% of the patients presented complete resolution of infection-induced fatigue when sulbutiamine was used in combination with an anti-infective treatment [33]. Although the authors suggested that increases in acetylcholine transmission and consequently the level of activation of the ascending reticular formation level (potentially produced by sulbutiamine or some derivative) may be related to the pathogenesis of asthenia and the improvement seen, there is a need to evaluate to what extent asthenias related to acute infections may be naturally improved with the amelioration of the disease. Notwithstanding these conflicting results, a recent retrospective observational study, conducted by Sevim et al., concluded that sulbutiamine use was associated with a
reduction in the total score of Fatigue Impact Scale (FIS) and in all other three subscales assessing physical, cognitive, and psychosocial functioning in multiple sclerosis patients who were on some kind of disease modifying treatment. Nevertheless, in patients who were not on any treatment, 400 mg/day of sulbutiamine did not show any improvement [31].

2.5. Depression and Sulbutiamine. Although no antidepressant effect of sulbutiamine was reported in patients with major depressive episodes, a multicentric, randomized, double-blind, placebo-controlled trial conducted by Loo et al. has shown that the administration of 600 mg/day of sulbutiamine for 8 weeks significantly decreased social inhibition associated with the depressive event [35].

2.6. Diabetes Polyneuropathy and Sulbutiamine. Kiew and colleagues using 400 mg/day of sulbutiamine for 42 days in an open-label randomized controlled study showed improvement in nerve signal parameters in type 2 diabetes patients suffering from polyneuropathies. However, there was no significant change in metabolic biomarkers, such as blood glucose and HbA1 [6]. Moreover, this study had several methodological limitations. Therefore, further research is needed before a definitive conclusion can be reached.

2.7. Psychogenic Erectile Dysfunction and Sulbutiamine. In a Hungarian study, Dmitriev and colleagues evaluating the efficacy of sulbutiamine treatment for 30 days in patients with psychogenic erectile dysfunction showed improvement in specific parameters of this disorder. The International Index of Erectile Function increased from 17.5 to 25.8, a 50% improvement in patients suffering from cavernous arterial dysfunction. Moreover, 36% of the patients experienced normalization of erectile function [37]. This study was not a controlled trial and, to our knowledge, it has not been replicated, and as such these results should be viewed with caution.

2.8. Deprivation Models and Sulbutiamine. Using the premises that “serum deprivation procedure has been used in various experimental models to induce neuronal death” and “serum withdrawal reduces the levels of neurotransphins and antioxidants in the extracellular medium which eventually leads cells to “starve” from these neurotrophic factors,” Kang and colleagues presented the beneficial actions of sulbutiamine in serum-deprived transformed retinal ganglion cells [27]. These effects were mainly due to enhancement of the cellular antioxidant and anti-apoptotic capacities by sulbutiamine and/or its derivatives, emphasizing its neuroprotective properties. Although 50 μM was the most effective dose when cell viability in a serum-deprived context was measured, lower doses (1 and 10 μM) of sulbutiamine showed very similar and promising results [27]. In 2011, this same Korean group using an ischemic model (oxygen-glucose deprived serum) in rat hippocampal CA1 pyramidal neurons showed increased neuronal viability, intrinsic membrane input resistance, and excitatory synaptic transmission in a dose dependent manner. 50 μM was again the most effective dose [41]. Working with the same in vitro model (transformed retinal ganglion cell-RGC-5 cells), Majid (one of the researchers of the two previous studies) and colleagues showed a dose dependent increase in cell viability (0%, 18%, and 28% with, respectively, 10, 50, and 100 μM sulbutiamine), less apoptosis, less caspase-3 activity, and a decrease in ROS levels for the 100 μM dose. Sulbutiamine at 100 μM also produced increases in catalase, GSH, GST (glutatione-s-transferase), Nrf-2 (nuclear factor erythroid 2-related factor 2), and HO-1 (Heme oxygenase-1) levels in serum-deprived medium. Of notice, when an oxidant medium was created using glutamate/buthionine sulfoximine no difference was seen in the majority of the parameters evaluated. In this later model, N-acetylcysteine (which is a precursor of GSH) showed better results and thus may be a complementary strategy to sulbutiamine in conditions associated with elevated oxidative stress (e.g., ischemia, diabetes) [42]. Benfotiamine is another synthetic derivative of thiamine that has been pointed as a potent stimulant of the antioxidant system. In an elegant study, Tapias and colleagues showed that benfotiamine and its main derivatives, but not thiamine, increased the expression of Nrf2/antioxidant response element (ARE)-dependent genes in mouse brain. This suggests that the disulfide bridge is not necessarily essential for the antioxidant action of sulbutiamine [43]. A recent study using a mouse neuroblastoma cell line (Neuro2a) grown in thiamine-restricted medium for 7 days and evaluating cell survival rate using the MTT method presented that the protective effects of benfotiamine and sulbutiamine are directly related to thiamine levels when cells were stressed by exposure to paraquat and β-amyloid peptide. This suggests these compounds act through a coenzyme-independent and indirect ROS scavenging mechanism [44]. Multiple positive interactions and effects between thiamine and derivatives of its different analogs appear to be possible when distinct deprivation models are tested. Therefore, the design of new studies and the discovery of theses derivatives will allow us a better understanding through which mechanisms and pathways each related thiamine molecule can act.

2.9. Microsporidiosis and Sulbutiamine. In a very different field, Garcia-Torres and colleagues discovered that low concentrations of sulbutiamine can specifically inhibit Encephalitozoon intestinalis triosephosphate isomerase and as such have classified this molecule as “a promising drug in the treatment of brain microsporidiosis (encephalitis),” one of the complications of this parasite that lacks mitochondria and oxidative phosphorylation. This discovery could have important consequences, since the prevalence of this type of infection can reach 60% in developing countries, Mexico being the country with the highest rate [28]. However, and although these researchers mention the ability of sulbutiamine to cross the blood-brain barrier, this molecule has never been identified in animal brain in its natural chemical structure after oral or even parenteral administration. Thus,
it seems that other ways of administration will be needed to
direct sulbutiamine to the brain. Nevertheless, other ex-
planations are possible, namely, (1) after interacting with
Encephalitozoon intestinalis in the intestinal tract, sulbuti-
amine may decrease the infectious rate by some unknown
action; (2) some presently unidentified metabolites of sul-
butiamine reaching the brain could interact with the cyste-
teines of the enzyme thus inhibiting its activity.

2.10. Anticancer Potential of Sulbutiamine. Proliferation of
cancer cells is often related to high activity of the key mi-
 tochondrial metabolism enzyme pyruvate dehydrogenase
kinase (PDK), the so-called Warburg effect. Thiamine has
been cited as a potential inhibitor of tumor proliferation in vivo [44] presumably by inhibiting PDK function and consequently pyruvate dehydrogenase (PDH) and mitochondrial function. Therefore, it may be limited by
transporter saturation and restricted accumulation of thi-
amine within tumor cells. Aiming to circumvent this, the
lipophilic thiamine analog sulbutiamine and benfotiamine
were tested in cultured human cancer cell lines and its
cellular proliferation was determined using a crystal violet
assay. The sensitivity of various tumor cell lines (HCT 116
colorectal carcinoma), U-87 MG (glioblastoma), MDA-
MB-231 (metastatic breast adenocarcinoma)) determined by
(colorectal carcinoma), U-87 MG (glioblastoma), MDA-
MB-231 (metastatic breast adenocarcinoma) determined by
the crystal violet assay showed a decreased proliferation for
thiamine, sulbutiamine, and benfotiamine after 5 days of
treatment. The main difference was the concentration
necessary for the effect (IC50), being 30–50-fold lower for
sulbutiamine and benfotiamine than for thiamine for all
cancer cell lines tested. When enzymatic specific PDH ac-

tivation tests were done, no effect for thiamine, TMP, sulbu-
tiamine, or benfotiamine was observed, and only ThDP was
capable of decreasing the PDH inhibition made by PDK
isoforms. When in vivo tumor growth-induced mice (sub-
cutaneous xenograft model) were treated every second day
with pharmacological doses (250 mg/kg) of sulbutiamine or
benfotiamine, both compounds increased the ThDP content,
but only benfotiamine reduced the tumor volume and
weight. Neither sulbutiamine nor benfotiamine was detected
in the tumors, suggesting that the apoptotic response ob-
served following benfotiamine treatment is different from
sulbutiamine, probably by its specific derivatives and
independent of the PDH-PDK axis [45]. Although this novel
and preliminary study of Jonus and colleagues presents
sulbutiamine as an anticancer therapy in vitro but not in vivo,
two points cited by the authors have to be considered
before excluding its in vivo potential:

(1) Changes to PDH phosphorylation may have been
impactted by the terminal end point used for analysis
of tumor tissue and not indicative of the initial re-
sponse to treatment that would be more comple-
mentary to in vitro findings

(2) Alternatively, the increase in TPP achieved by in vivo
dosing paradigm may not have been substantial
enough to sufficiently inhibit PDK

Therefore, new studies need to be done using sulbti-
amine in cancer models to address the potential anticancer
effect of sulbutiamine.

2.11. Antidoping and Sulbutiamine. The sulbutiamine mass
spectrometry peak is cited as one of the “peaks identified as a
compound with a possible anabolic, or otherwise stimulating
or endurance enhancing effect” by Peters and colleagues [4].
In this study aimed at developing a novel technique to
identify anabolic steroids, herbal mixtures and sport sup-
plements were used as samples. Later, in the same year,
Sobolevsky & Rodchenkov, researchers of the Moscow
Antidoping Center, observed that 2% of 5,151 antidoping
urine samples contained sulbutiamine and alerted that
“sulbutiamine could hamper the detection of boldenone
metabolites due to the coelution when a single quadrupole
GC-MS instrument is used for the screening analysis” [5].
However, as an analytic study this paper gives no infor-
mation as to the route of administration and the dose used so
as to find a significant concentration in the urine (>500 ng/
ml or >0.7 μM). Previous attempts in rodents were unsuccess-
ful and it is not clear whether this is due to more
sensitive analytical tools or the fact that it was done in
humans and not in rodents. But this paper may be the
document proof of the existence of sulbutiamine in body
fluids.

3. Side Effects

Balzamo and Vuillon-Cacciuttolo reported hypersalivation in
2 out of 6 rhesus monkeys after administration of high doses
of sulbutiamine (300 mg/kg/day). Although these high doses
affected the sleep pattern, they did not alter the motor ability
of the animals [29]. Shah reported side effects occurring in
0.6% of the patients using sulbutiamine. In a total of 1,772
patients, 5 presented nausea, 1 headache, 1 insomnia, 1 di-
arrhea, 1 tremor, and 1 drowsiness [33]. Sevim and colleagues
reported no “serious” adverse effect in 26 relapsing-remitting
multiple sclerosis patients using a single dose of 400 mg of
sulbutiamine [31]. In accordance, in a sample of 300 patients
included in the study (approximately 100–400 mg sulbuti-
amine; 100–600 mg sulbutiamine; 100–placebo), Tiev and
colleagues reported that only 15 patients using sulbutiamine
presented side effects (headache, agitation, palpitations, di-
arrhea, cough, sinusitis, cystitis, bronchitis, osteoarthritic and
low back pain, palpitations, asthma, abdominal pain, and
nephrotic colic). However, these side effects may not be re-
lated to sulbutiamine as in the placebo group 12 patients
presented similar effects [19]. In another study, Ollat and
colleagues showed that 3 patients out of 40 (8%) using sul-
butiamine in a standalone therapy had headache/insomnia as
a side effect [39].

4. Conclusions

In recent years, there has been an increase in the number of
studies on the health applications of sulbutiamine.
Experimental evidence, observational studies, and clinical trials suggest that sulbutiamine has various actions in human physiology through its ability to provide thiamine to less accessible tissues (such as the brain), by increasing antioxidant capacity and by modulation of protein action [7, 41]. However, caution is required in evaluating some of these results (e.g., post-infection fatigue will be necessarily ameliorated in the first month even in patients not using sulbutiamine, and as pointed by the authors “relief from asthenic symptoms cannot be attributed exclusively to sulbutiamine”) [33, 46, 47]. There is also evidence that sulbutiamine has, to some degree, a neuromodulator capacity (probably through some of its metabolites). Nevertheless, various questions and concerns must be answered by controlled studies before the effectiveness and clinical relevance of sulbutiamine can be attested.

The dosage and absorption rates seem to be another important question to address in future studies, as seen in Balzamo’s study, where sulbutiamine at approximately 10 times the usual dose interfered with the sleep/wake cycle. However, low doses (1–10 μM) used in in vitro studies showed general antioxidant effects, although 50 μM could have additional effect in neurons. One explanation for this could be that ThTP content generated by sulbutiamine could play a role in relation to membrane permeability modulation [1].

It is also important to remark that sulbutiamine seems to have the capacity to interfere with the action of central proteins, as in the case of caspase-3, bearing in mind that “the primary caspases involved in the apoptotic degeneration of BGC-5 after axotomy have been caspase-3 and caspase-9” [42]. Even more interesting, sulbutiamine, as presented by García-Torres and colleagues, may inactivate microsporidial triosephosphate isomerase, putting this molecule in a class of drugs and supplements with a broad spectrum of action [28]. And now it will probably gain attention in oncologic research as the most recent publication presented an anticancer effect in vitro [45].

Because of the great potential of this molecule, initially designed for the treatment of beriberi, gaps in the literature should not intimidate but challenge researchers to design and conduct high quality research. This review intends to present and discuss most of information available on sulbutiamine and possibly serve as a starting point for future studies. Specifically, randomized controlled trials are necessary to investigate the health potential of this molecule. Presently, the safest approach seems to lie in evaluating patients individually and carefully considering the possibility of using sulbutiamine in adequate doses for the clinical conditions already listed here. Also, it would be of interest to compare the effects of sulbutiamine with those of thiamine and benfotiamine (which is not a disulfide, but a thioester) both in vitro and in vivo. This would help answer the question whether sulbutiamine mainly acts via thiamine/thiamine phosphate derivatives or whether it has more specific pharmacological effects. Efforts should be undertaken to investigate whether, after administration of sulbutiamine, this compound or its metabolite thiamine disulfide can be found in the brain using modern mass spectrometry-based analytical methods.

Conflicts of Interest
L. Bettendorff is the Research Director of the “Fonds de la Recherche Scientifique-FNRS, Belgium. The authors declare that they have no conflicts of interest.

Authors’ Contributions
BFS drafted the manuscript. PCB and LB made critical revisions and gave suggestions during the preparation. All authors read and approved the final manuscript.

Supplementary Materials
Description of sulbutiamine studies and its health applications. (Supplementary Materials)

References
[1] L. Bettendorff, L. Weekers, P. Wins et al., “Injection of sulbutiamine induces an increase in thiamine triphosphate in rat tissues,” Biochemical Pharmacology, vol. 40, no. 11, pp. 2557–2560, 1990.
[2] G. E. Gibson, J. A. Hirsch, R. T. Cirio et al., “Abnormal thiamine-dependent processes in Alzheimer’s disease. Lessons from diabetes,” Molecular and Cellular Neuroscience, vol. 55, pp. 17–25, 2013.
[3] R. Balestrieri and S. Bertolini, “Etude de l’activité thérapeutique d’Arcalion 200 sur les manifestations psycho-physiques des syndromes asthéniques,” Viemed Healthcare Vie Med, vol. 18, 1981.
[4] R. J. B. Peters, J. C. W. Rijk, T. F. H. Bovee et al., “Identification of anabolic steroids and derivatives using bioassay-guided fractionation, UHPLC/TOFMS analysis and accurate mass database searching,” Analytica Chimica Acta, vol. 664, no. 1, pp. 77–88, 2010.
[5] T. Sobolevsky and G. Rodchenkov, “Sulbutiamine in sports,” Drug Test Analysis, vol. 2, no. 11-12, pp. 643–646, 2010.
[6] K. K. Kiew, W. W. Mohamad, A. Ridzuan et al., “Effects of sulbutiamine on diabetic polynuropathy: an open randomised controlled study in type 2 diabetics,” Malays Journal of Medical Sciences, vol. 9, no. 1, pp. 21–27, 2002.
[7] O. S. Levin and S. IuB, “The use of eiron in the treatment of asthenic disorders in patients after mild cranio-cerebral trauma,” Korsakov Journal of Neurology and Psychiatry, vol. 107, no. 5, pp. 44–48, 2007.
[8] M. Saiki, T. Matsui, M. Soya et al., “Thiamine tetrahydrofurfuryl disulfide promotes voluntary activity through dopaminergic activation in the medial prefrontal cortex,” Scientific Reports, vol. 8, no. 1, Article ID 10469, 2018.
[9] K. Harada, K. Kohno, I. Daira et al., “Retention in the body of various o-acylthiamine disulfides and several properties of o-butyryl-thiamine disulfide,” The Journal of Vitaminology, vol. 12, no. 3, pp. 221–230, 1966.
[10] L. Bettendorff and P. Wins, “Thiamin diphosphate in biological chemistry: new aspects of thiamin metabolism, especially triphosphate derivatives acting other than as cofactors,” FEBS Journal, vol. 276, no. 11, pp. 2917–2925, 2009.
[11] L. Bettendorff and P. Wins, “Thiamine derivatives in excitable tissues: metabolism, deficiency and neurodegenerative diseases,” Recent Research and Development Neurochemical, vol. 2, no. 1, pp. 37–62, 1999.
[12] K. Gotoh, T. Shinoda, Y. Kimura et al., “Clinical experience with BuTDS in urological field,” Hinyokikaiyō Acta Urologica Japonica, vol. 12, no. 7, pp. 709–712, 1966.

[13] H. Tanaka, M. Mizoguchi, Y. Fujimoto et al., “Clinical use of BuTDS in urological field,” Acta Urologica Japonica, vol. 12, no. 8, pp. 811–822, 1966.

[14] M. Tsuchida and Y. Kimura, “Effects of BuTDS on postoperative intestinal paralysis,” Acta Urologica Japonica, vol. 12, no. 8, pp. 808–810, 1966.

[15] S. Murakami, T. Arakawa, and M. Nakagawa, “Therapeutic effect of O-butyrylthiamine disulfide (BuTDS) on cardiac function under general anesthesia with reference to improvement of ECG findings,” Nihon Geka Hokan, vol. 35, no. 5, p. 879, 1966.

[16] C. Kawasaki, I. Tomita, and S. Shinoda, “Penetration of modified thiamine compounds into erythrocytes,” The Journal of Vitaminology, vol. 10, no. 2, pp. 154–160, 1964.

[17] C. Kawasaki and S. Shinoda, “Penetration of modified thiamine compounds into erythrocytes II,” Journal of Vitaminology, vol. 29, no. 4, pp. 255–259, 1964.

[18] L. Bettendorf, “The compartmentation of phosphorylated thiamine derivatives in cultured neuroblastoma cells,” Biochimica et Biophysica Acta (BBA)-Molecular Cell Research, vol. 1222, no. 1, pp. 7–14, 1994.

[19] K. P. Tiev, J. Cabane, and J. C. Imbert, “Traitement de l’asthénie en période postinfectieuse: étude randomisée en double aveugle de deux doses de sulbutiamine (400–600 mg/j) versus placebo,” La Revue de Médecine Interne, vol. 20, no. 10, pp. 912–918, 1999.

[20] L. Bettendorf, P. Wins, and M. Lesourd, “Subcellular localization and compartmentation of thiamine derivatives in rat brain,” Biochimica et Biophysica Acta (BBA)-Molecular Cell Research, vol. 1222, no. 1, pp. 1–6, 1994.

[21] L. Bettendorf, B. Wirtzfeld, A. F. Makarchikov et al., “Discovery of a natural thiamine adenine nucleotide,” Nature Chemical Biology, vol. 3, no. 4, pp. 211-212, 2007.

[22] M. Gangolf, P. Wins, M. Thiry et al., “Thiamine triphosphate synthesis in rat brain occurs in mitochondria and is coupled to the respiratory chain,” Journal of Biological Chemistry, vol. 285, no. 1, pp. 583–594, 2010.

[23] T. Gigliobianco, M. Gangolf, B. Lakaye et al., “An alternative role of F0F1-ATP synthase in Escherichia coli: synthesis of thiamine triphosphate,” Scientific Report, vol. 3, no. 1, p. 1071, 2013.

[24] L. Bettendorf, “A non-cofactor role of thiamine derivatives in excitable cells?” Archives of Physiology and Biochemistry, vol. 104, no. 6, pp. 745–751, 1996.

[25] L. Bettendorf, B. Lakaye, G. Kohn et al., “Thiamine triphosphate: a ubiquitous molecule in search of a physiological role,” Metabolic Brain Disease, vol. 29, no. 4, pp. 1069–1082, 2014.

[26] P. L. Wood, M. A. Khan, and J. R. Moskal, “Cellular thiol pools are responsible for sequestration of cytotoxic reactive aldehydes: central role of free cysteine and cysteamine,” Brain Research, vol. 1158, pp. 158–163, 2007.

[27] K. D. Kang, A. S. Abdul Majid, K.-A. Kim et al., “Sulbutiamine counteracts trophic factor deprivation induced apoptotic cell death in transformed retinal ganglion cells,” Neurochemical Research, vol. 35, no. 11, pp. 1828–1839, 2010.

[28] I. García-Torres, G. Hernández-Alcántara, D. Molina-Ortiz et al., “First characterization of a microsporidial triosephosphate isomerase and the biochemical mechanisms of its inactivation to propose a new druggable target,” Scientific Reports, vol. 8, no. 1, p. 8591, 2018.

[29] E. Balzamo and G. Vuillon-caccitutto, “Facilitazione de l’etat de veille d’un traitement semi-chronique par la sulbutiamine (Arcalion) chez Macaca mulatta,” Revue d’Electroencephalographie et de Neurophysiologie Clinique, vol. 12, no. 4, pp. 373–378, 1982.

[30] J. Micheau, T. P. Durkin, C. Destrade et al., “Chronic administration of sulbutiamine improves long term memory formation in mice: possible cholinergic mediation,” Pharmacology Biochemistry and Behavior, vol. 23, no. 2, pp. 195–198, 1985.

[31] S. Sevim, H. Kaleağas, and B. Taşdelen, “Sulbutiamine shows promising results in reducing fatigue in patients with multiple sclerosis,” Multiple Sclerosis and Related Disorders, vol. 16, pp. 40–43, 2017.

[32] A. Costantini, A. Nappo, M. I. Pala et al., “Case report: high dose thiamine improves fatigue in multiple sclerosis,” BMJ, London, UK, 2013.

[33] S. N. Shah, “Adjuvant role of vitamin B analogue (sulbutiamine) with anti-infective treatment in infection associated asthma,” The Journal of the Association of Physicians of India, vol. 51, p. 891, 2003.

[34] D. Morin, T. Hauet, M. Spedding et al., “Mitochondria as target for anti-ischemic drugs,” Advanced Drug Delivery Reviews, vol. 49, no. 1-2, pp. 151–174, 2001.

[35] H. Loo, M. F. Poirier, H. Ollat et al., “Effects of sulbutiamine (Arcalion 200) on psycho-behavioral inhibition in major depressive episodes,” Encephale, vol. 26, no. 2, pp. 70–75, 2000.

[36] F. Trovero, M. Gobbi, J. Weil-Fuggaza et al., “Evidence for a modulatory effect of sulbutiamine on glutamatergic and dopaminergic cortical transmissions in the rat brain,” Neuroscience Letters, vol. 292, no. 1, pp. 49–53, 2000.

[37] D. G. Dmitriev, S. I. Gamidov, and O. V. Permiakova, “Clinical efficacy of the drug eneron in the treatment of patients with psychogenic (functional) erectile dysfunction,” Urologiya, vol. 1, pp. 32–35, 2005.

[38] J.-C. Bizot, A. Herpin, S. Pothion et al., “Chronic treatment with sulbutiamine improves memory in an object recognition task and reduces some amnesic effects of dizocilpine in a spatial delayed-non-match-to-sample task,” Progress in Neuro-Psychopharmacology and Biological Psychiatry, vol. 29, no. 6, pp. 928–935, 2005.

[39] H. Ollat, B. Laurent, S. Bakchine et al., “Effets de l’association de la Sulbutiamine à un inhibiteur de l’acétylcholinestérase dans les formes légères à modérées de la maladie d’Alzheimer,” L’Encéphale, vol. 33, no. 2, pp. 211–215, 2007.

[40] M. Sambon, A. Napp, A. Demelonne et al., “Thiamine and benfotiamine protect neuroblastoma cells against paracetamol and β-amylod toxicity by a coenzyme-independent mechanism,” Helyon, vol. 5, no. 5, Article ID e01710, 2019.

[41] J. Kwag, A. S. A. Majid, and K. D. Kang, “Evidence for neuroprotective effect of sulbutiamine against oxygen-glucose deprivation in rat hippocampal CA1 pyramidal neurons,” Biological & Pharmaceutical Bulletin, vol. 34, no. 11, pp. 1759–1764, 2011.

[42] A. S. A. Majid, Z. Q. Yin, and D. Ji, “Sulphur antioxidants inhibit oxidative stress induced retinal ganglion cell death by scavenging reactive oxygen species but influence nuclear factor (erythroid-derived 2)-like 2 signalling pathway differently,” Biological and Pharmaceutical Bulletin, vol. 36, no. 7, pp. 1095–1110, 2013.

[43] V. Tapias, S. Jainuddin, M. Ahuja et al., “Benfotiamine treatment activates the Nrfl2/ARE pathway and is
neuroprotective in a transgenic mouse model of tauopathy,” *Human Molecular Genetics*, vol. 27, no. 16, pp. 2874–2892, 2018.

[44] B. Comín-Anduix, J. Boren, S. Martinez et al., “The effect of thiamine supplementation on tumour proliferation: a metabolic control analysis study,” *European Journal of Biochemistry*, vol. 268, no. 15, pp. 4177–4182, 2001.

[45] H. C. Jonus, C. C. Byrnes, J. Kim et al., “Thiamine mimetics sulbutiamine and benfotiamine as a nutraceutical approach to anticancer therapy,” *Biomedicine & Pharmacotherapy*, vol. 121, p. 109648, 2020.

[46] B. Cameron, M. Bharadwaj, J. Burrows et al., “Prolonged illness after infectious mononucleosis is associated with altered immunity but not with increased viral load,” *The Journal of Infectious Diseases*, vol. 193, no. 5, pp. 664–671, 2006.

[47] S. Khalilpour, G. Behnammanesh, A. M. S. Abdul Majid et al., “Assessment of neuroprotective properties of *Rhus coriaria* L. ethanol extract in an in vitro model of retinal degeneration,” *Journal of Herbal Medicine*, vol. 10, pp. 45–52, 2017.

[48] A. Nair and S. Jacob, “A simple practice guide for dose conversion between animals and human,” *Journal of Basic and Clinical Pharmacy*, vol. 7, no. 2, p. 27, 2016.