A Sparse Semi-Blind Source Identification Method and Its Application to Raman Spectroscopy for Explosives Detection

Yuanchang Sun *and Jack Xin*

Abstract

Rapid and reliable detection and identification of unknown chemical substances is critical to homeland security. It is challenging to identify chemical components from a wide range of explosives. There are two key steps involved. One is a nondestructive and informative spectroscopic technique for data acquisition. The other is an associated library of reference features along with a computational method for feature matching and meaningful detection within or beyond the library.

Recently several experimental techniques based on Raman scattering have been developed to perform standoff detection and identification of explosives, and they prove to be successful under certain idealized conditions. However data analysis is limited to standard least squares method assuming the complete knowledge of the chemical components. In this paper, we develop a new iterative method to identify unknown substances from mixture samples of Raman spectroscopy. In the first step, a constrained least squares method decomposes the data into a sum of linear combination of the known components and a non-negative residual. In the second step, a sparse and convex blind source separation method extracts components geometrically from the residuals. Verification based on the library templates or expert knowledge helps to confirm these components. If necessary, the confirmed meaningful components are fed back into step one to refine the residual and then step two extracts possibly more hidden components. The two steps may be iterated until no more components can be identified. We illustrate the proposed method in processing a set of the so called swept wavelength optical resonant Raman spectroscopy experimental data by a satisfactory blind extraction of a priori unknown chemical explosives from mixture samples.

*Department of Mathematics, University of California at Irvine, Irvine, CA 92697, USA.
1 Introduction

A critical problem in homeland security is reliable and rapid identification of unknown chemical and biological substances in the explosives. Due to the harmful environment caused by the release of the explosive chemicals, non-destructive spectroscopic techniques are typically used to record the optical spectrum without interfering with the samples. Ideally, a standoff detection is performed to acquire the spectral information. Then a search and matching procedure through a prepared spectral database will be carried out for identification. However such an approach would be unsuccessful if the explosives contained chemical compounds outside of the database, which is highly likely as hidden explosives are often unknown a-priori. In general, the samples may involve multiple unknown substances besides impurities. Conventional analysis routines are mostly based on least squares fitting whose residuals could remain mysterious. Further analysis calls for the development of blind identification methods to extract major components from the residuals.

Various recent experimental techniques (see [3, 6, 7] and the references therein) can identify pure chemicals with notable success. These methods are mainly based on Raman spectroscopy, a spectroscopic technique to study the chemical composition of the samples [12, 21]. Combined with other spectroscopic techniques such as Ultraviolet and Infrared spectroscopy, Raman spectroscopy has been widely used in materials science, biosciences, geosciences (gemology), forensic sciences, nano-technology, and pharmaceutical chemistry [3, 4, 5, 7, 13, 15, 17, 23]. For example, the Swept Wavelength Optical Resonant Raman Detector (SWOrRD) at the Naval Research Laboratory developed in 2009 can generate two dimensional spectral maps of biological agents and chemical substances. The resultant two dimensional signatures contain much more information than the single illumination wavelengths, which may result in a greater likelihood of successful identification even in complex mixtures [3, 7]. Raman spectroscopy is based on Raman scattering, an inelastic scattering process that shifts the frequencies of the incident photons. During the interaction of the incident light with a molecule, a scattered light of lower (Stokes Raman) or higher (anti-Stokes Raman) energy is emitted, allowing the measurements of the molecule’s vibrational modes. The appeals of this technique are non-destructiveness and fast sensing capability. It has become a promising tool for standoff distance detection for explosives at airports among other transportation centers.

In general, Raman spectra of a sample are composed of many substances, and they must be identified by an analysis software. If one has the complete knowledge of the kinds of substances in the sample, the least squares fitting can be used to retrieve their concentrations (or volumes) by a linear combination of the known spectra on a template. In most practical situations, one may have to identify the substances and quantify their concentrations at the same time. This becomes a blind source separation (BSS) problem, or recovering pure signal sources from their mixtures without a detailed knowledge of the mixing process. There have been several studies on the BSS of Raman spectra [11, 16] based on independent component analysis (ICA, [9]) and nonnegative matrix factorization (NMF, [10]). However these methods are non-convex and too general to be robust and reliable in real-world applications. The independence hypothesis of ICA does not hold if chemicals share some common structures and have
correlated Raman spectral line shapes. Moreover, these existing approaches do not address how to identify unknown substances from the fitting residuals when partial knowledge of the source signals is available. Such a semi-blind problem is more often encountered in applications and of great importance to practitioners.

In this paper, we shall develop a convex semi-blind source separation method based on sparsity of the source spectra. We are concerned with the regime where the sample contains some known and some unknown components. In other words, we have knowledge of some of the components and their concentrations, which is the case of the SWOrRD data. We further assume that the upper bound of the concentrations of the known substances is available, say from experiments or prior knowledge, as is the case of SWOrRD data. Though our method here is designed for Raman spectra, it is applicable to spectra with similar line shapes, such as nuclear magnetic resonance (NMR) data, see [18, 19] where more general sparseness source conditions and post-processing methods have been studied. A similar semi-blind source identification approach has been developed for mysterious species of the atmospheric gas mixtures [20].

Our method in the paper may provide an assistive tool for practitioners to analyze the hidden structures from residuals of fitting complicated mixtures. The method contains two major steps. The first step decomposes Raman data $X$ into a linear combination of the reference spectra (known components) plus a remainder, or $X = A S + R$, where the columns of matrix $A$ are known reference spectra, the matrix $S$ contains the non-negative concentrations and has known upper bound. The first step is carried out by solving a constrained least squares problem. The constraints on $S$ help to maintain the nonnegativity of remainder matrix $R$. The second step performs a nonnegative blind source separation of the remainder matrix $R$ to extract the unknown components. We show that proper sparsity of source signals reduces the general non-convex problem to constrained convex programming permitting solutions with better mathematical properties. Sparse solutions to convex objectives are achieved through $\ell_1$ norm minimization and a fast iterative method (the linearized Bregman). The two steps may be iterated. If some of the components from step two are confirmed as chemically meaningful, they are fed back to step one to refine the residual for further extraction of hidden components in step 2.

The paper is organized as follows. In section 2, we introduce the source sparseness condition and our method. In section 3, we illustrate the proposed method in processing a set of SWOrRD experimental data, and show satisfactory numerical results. Conclusion and future works are discussed in section 4.

This work was partially supported by NSF-ADT grant DMS-0911277. The authors thank Naval Research Laboratory for the experimental Raman data and Dr. J. Grun for helpful discussions.

2 The method

In this section, we shall present our method for chemicals detection in Raman spectroscopy. Based on the assumption that part of the substances in the chemical mixture
and the upper bounds of their concentrations are known, the first step of the method fits these known chemicals to the mixtures. This step solves a constrained least squares optimization.

### 2.1 Constrained least squares

The following linear model is used for Raman spectra of mixtures

\[ X = A S + R, \]  

(2.1)

where the columns of matrix \( X \) are the measured Raman data, the columns of \( A \) contain reference spectra of the known chemicals, and those of \( S \) matrix contains their concentrations. Matrix \( R \) is the fitting residual which might contain hidden spectral structures, the instrument noise, etc. Matrix \( S \) is nonnegative on physical ground that its entries represent concentrations. Furthermore, the upper bound of \( S \) is known \textit{a priori}. Then for the estimation of \( S \), we minimize the following constrained objective function:

\[
\min_S \|X - AS\|_2^2, \quad \text{s.t. } 0 \leq S \leq c, \]  

(2.2)

where the vector \( c \) contains the upper bounds of the concentrations of the known substances in the sample. For the Raman data in our numerical experiments, we found that the linear constraint in (2.2) helps to maintain a nonnegative remainder \( R = X - AS \). In the residual \( R \), there might be spectral structures (one or many) of chemicals buried in noise, or just random noise. In either case, we factorize the residuals in a blind fashion due to the lack of knowledge of the hidden components. Conventional blind source separation methods such as NMF and ICA are non-convex optimization methods. These methods are for general purpose, yet often unreliable in real-world applications due to non-convexity or sensitivity of their working assumptions. For our problem, we show that proper sparsity of source signals reduces the general non-convex problem to constrained convex programming permitting solutions with better mathematical properties. Sparse solutions to convex objectives are achieved through \( \ell_1 \) minimization.

### 2.2 Convex blind source separation

The remainder matrix \( R \) is factorized as:

\[ R = W M, \]

(2.3)

where the columns of the matrix \( W \) are the identified substances, \( M \) is their concentrations in the sample. All the matrices are nonnegative. For the purpose of illustration, we shall call \( W \in \mathbb{R}^{p \times n} \) the source matrix, \( M \in \mathbb{R}^{n \times m} \) the mixing matrix. The dimensions of the matrices are expressed in terms of three numbers: (1) \( p \) the number of available samples, (2) \( m \) the number of mixture signals, and (3) \( n \) the number of source signals. For the Raman data we considered in the paper, there are more mixtures than sources, i.e., \( m \geq n \). The goal is to recover \( W \) and \( M \) for a given \( R \). This is also known as an NMF problem. Various methods have been developed to solve BSS problems by exploiting the natures of source signals. For
example, independent component analysis (ICA) recovers statistically independent signals. However, the independence should not be assumed on the Raman spectra of the chemical substances when they share common structural features (the line shapes of their Raman spectra are similar). A better working assumption for the data is a so-called partial sparseness condition. Namely, the source signals are only required to be non-overlapping at some locations of acquisition variable. This sparseness condition was first known in the 1990s [1, 22] in the study of blind hyper-spectral unmixing of remote sensing, where the source condition is called pixel purity assumption (PPA) [2]. In 2005, Naanaa and Nuzillard [14] used this assumption to separate the signals in nuclear magnetic resonance spectroscopy. In fact, this condition is well suited to many chemical substances including those studied in this paper. Such a sparseness condition leads to a dramatic mathematical simplification of a general non-negative matrix factorization problem (2.3) which is non-convex. Geometrically speaking, the problem of finding the mixing matrix $M$ reduces to the identification of a minimal cone containing the rows of matrix $R$. The latter can be achieved by linear programming. In the context of hyper-spectral unmixing, the resulting geometric (cone) method is the so called N-findr [22], and is now a benchmark in hyper-spectral unmixing. Next we shall review the essentials of the partial sparseness condition and the geometric cone method.

Simply speaking, the key sparseness assumption on the source signals is that each source has a stand alone peak at some location of acquisition variable where the other sources are zero. More precisely, the source matrix $W \geq 0$ is assumed to satisfy the following condition:

**Assumption.** For each $j \in \{1, 2, \ldots, n\}$ there exists an $i_j \in \{1, 2, \ldots, p\}$ such that $w_{i_j,j} > 0$ and $w_{i_j,k} = 0$, $(k = 1, \ldots, i_j - 1, i_j + 1, \ldots, n)$.

Eq. (2.3) can be rewritten in terms of rows as

$$R_i = \sum_{k=1}^{n} w_{i,k} M^k$$

where $R_i$ denote the $i$th row of $R$, and $M^k$ the $k$th row of $M$. The source assumption implies that $R_i^j = w_{i,j} M^j$ $j = 1, \ldots, n$ or $M^j = \frac{1}{w_{i,j}} R_i^j$. Hence Eq. (2.4) is rewritten as

$$R_i = \sum_{k=1}^{n} \frac{w_{i,k}}{w_{i,k}} M^i_k,$$  \hspace{1cm} (2.5)

which says that every row of $R$ is a nonnegative linear combination of the rows of $M$. The identification of $M$’s rows is equivalent to identifying a convex cone of a finite collection of vectors. The cone encloses the data rows in matrix $R$, and is the smallest of such cones. Such a minimal enclosing convex cone can be found by linear programming methods. Mathematically, the following optimization problems are suggested to estimate the mixing matrix

$$\min \text{ score} = \| \sum_{i=1, i \neq l}^{p} R_i^l \lambda_i - R^l \|_2, \text{s.t. } \lambda_i \geq 0, l = 1, \ldots, p.$$  \hspace{1cm} (2.6)
A score is associated with each row of $R$. A row with a low score is unlikely to be a row of $M$ because this row is roughly a nonnegative linear combination of the other rows of $R$. On the other hand, a high score means that the corresponding row is far from being a nonnegative linear combination of other rows. The $n$ rows from $R$ with highest scores are selected to form $M$, the mixing matrix.

### 2.3 Sparse source recovery

With the recovery of $M$, we solve for $W$ next. An existing method [14] is to directly compute $W = RM^+$, where $M^+$ is the pseudo-inverse of $M$. It is however sensitive to noise, and tends to introduce errors and artifacts of negative values. Although a nonnegative least squares can produce a nonnegative $W$, spurious peaks might be introduced in the results. To benefit from the sparseness source condition in the first step, a more reliable method is to solve a nonnegative $\ell_1$ optimization. Although the source signals (columns of $W$) are not sparse, the rows of $W$ possess sparsity. Hence, we seek the sparsest solution for each row $W^i$ of $W$.

\[
\min \| W^i \|_0 \quad \text{subject to} \quad W^i M = R^i, \quad W^i \geq 0.
\]  

(2.7)

Here $\| \cdot \|_0$ (0-norm) represents the number of nonzeros. Because of the non-convexity of the 0-norm, we minimize the $\ell_1$-norm as a convex relaxation:

\[
\min \| W^i \|_1 \quad \text{subject to} \quad W^i M = R^i, \quad W^i \geq 0,
\]  

(2.8)

which is in the form of linear programming. The fact that the data may in general contain noise suggests us to solve the following unconstrained optimization problem,

\[
\min_{W^i \geq 0} \mu \| W^i \|_1 + \frac{1}{2} \| R^i - W^i M \|_2^2,
\]  

(2.9)

for which Bregman iterative method [8, 24] with a proper projection onto non-negative convex subset is used to obtain a solution. In this paper, we shall use the linearized Bregman iteration to solve (2.9) due to its efficiency. For each row $W^i$ of $W$, we introduce $u = (W^i)^T, f = (R^i)^T, B = M^T$, then (2.9) is equivalent to

\[
\min_{u \geq 0} \mu \| u \|_1 + \frac{1}{2} \| f - B u \|_2^2.
\]  

(2.10)

The $l_2$ norm in (2.9,2.10) is to model the unknown measurement error or noise as Gaussian. When there is minimal measurement error, one must assign a tiny value to $\mu$ to heavily weigh the fidelity term $\| f - B u \|_2^2$ in order for $B u = f$ to be nearly satisfied. The linearized Bregman method can be written iteratively by introducing an auxiliary variable $v^j$:

\[
\begin{align*}
\{ & v^{j+1} = v^j - B^T (B v^j - f), \\
& u^{j+1} = \delta \cdot \text{shrink}_+(v^{j+1}, \mu), \}
\end{align*}
\]  

(2.11)

where $u^0 = v^0 = O, \delta > 0$ is the step size, and $\text{shrink}_+$ is for computing nonnegative solutions,

\[
\text{shrink}_+(v, \mu) = \begin{cases} 
 v - \mu, & \text{if } v > \mu, \\
 0, & \text{if } v < \mu.
\end{cases}
\]  

(2.12)
3 Numerical experiments

In this section, we shall apply our method to the so-called SWorRD Raman data provided by the Naval Research Laboratory, and present the computational results.

3.1 SWorRD Data

The Swept Wavelength Optical Resonant Raman Detector (SWorRD) at the Naval Research Laboratory is a spectroscopy system which is able to produce two-dimensional spectral data of biological agents and chemical substances. A target substance is illuminated with a specific laser wavelength and this generates a resonance Raman spectrum. When the laser wavelength is varied and the process is repeated, the resonance Raman spectra (one at each laser wavelength), forms a two-dimensional plot (signature) where one axis is the input laser wavelength and the other axis is the wavenumber of the Raman spectrum. Fig. 1 is an example of 2D SWorRD Raman spectra of a mixture of ethanol, ethylene glycol, acetonitrile, and water. The data matrix $X$ corresponds to this 2D spectra. Each column $X$ is a Raman spectrum of the mixture at a specific laser wavelength. For SWorRD data, there are much more mixtures than the number of sources. The spectral features may be slightly altered from one wavelength to another. To reduce this effect, we do not necessarily use all the data, and so we sample the spectra at a subset of input laser wavelengths.

3.2 Results

We report here the computational results from our proposed method. We tested our method on two sets of SWorRD data. The samples consist of several liquid substances, some of them are commonly used for synthesizing explosives. The first
Figure 2: A Raman spectrum of the mixture of methanol and two other liquid chemicals is shown on the left, while the Raman spectral reference of methanol is plotted on the right.

data set includes 21 mixed Raman spectra at different excitation wavelengths. This sample is known to contain methanol and its concentration is below 1/3. There are also two \textit{a-priori} unknown liquid chemicals. The method first fits the known reference spectrum of methanol to the Raman data. Then it identifies two unknown chemicals from the fitting residuals. The results are shown in a series of plots, Fig. 2-5. From all the data available, we found that the 5 spectra at consecutive laser wavelengths of (248, 250, 252, 254, 256) nm produce the best results. The theoretical underpinning of optimal selection of the data is under further study. Fig. 2 shows the line shape of the spectrum of the mixture, and the spectral reference of methanol. The residual after fitting methanol to the data is plotted in Fig. 3, where some structure can be seen. Then further identification of the two hidden chemicals was made by the convex BSS method, and the results are presented in Fig. 4. Compared to the ground truth, the results are satisfactory in that the recovered spectral structures are recognizable as ethanol and acetonitrile. Conventional method such as NMF tends to lose reliability in dealing with the data, the NMF results are also presented as a comparison. In the second example, we try to identify two hidden chemicals from a mixture of four chemicals. The known chemicals are methanol and ethanol. The total concentration of the two is below 1/2, which is known from the sensing hardware. The computational results are presented in Fig. 6-9. Although no apparent Raman spectral structure can be identified from the first plot in Fig. 6, the second structure is easily recognizable as acetonitrile upon comparison with reference spectrum. Next we subtract the confirmed acetonitrile from the residual by solving the constrained least squares in step 1, then feed the new residual back to step 2 for extracting more hidden chemicals. The extracted structure in Fig. 9 can be easily identified as ethylene glycol comparing to the ground truth. Computations are performed on a Dell laptop with 6G RAM and 1.6 GHz i7 CPU. The cpu time for example 1 is 7.613 seconds, and it is 10.327 seconds for example 2. The method proves to be efficient and can be of use in rapid detection of chemical substances.
Figure 3: One fitting residual from the constrained least squares data fitting.

Figure 4: Left column is the two identified spectral structures, right column shows the Raman spectral references for ethanol and acetonitrile. The value of $\mu$ used in step 2 is 0.09.
Figure 5: Left column is the results computed by NMF, right column shows the Raman spectral references for ethanol and acetonitrile.

Figure 6: A Raman spectrum of the mixture of methanol, ethanol and two other chemicals is shown on the left, while the Raman spectral references of methanol and ethanol are plotted on the right. $\mu = 0.09$ is used for the source recovery in step 2.
Figure 7: One fitting residual from the constrained least squares data fitting.

Figure 8: Left column is the two identified spectral structures, right column shows the Raman spectral references for acetonitrile and ethylene glycol.
A semi-blind sparse and convex source identification method is developed to extract meaningful spectral structures from mixture data of Raman spectroscopy. The method is designed to identify potentially hidden chemicals after fitting the known spectral references to the data. Our method can be useful for extracting unknown source signals from the residuals after known reference spectra have been first deployed to fit the data. The major strength of the technique is its ability to be used either with known reference spectra for quantification or without reference spectra for identification of unknown/hidden chemical substances. Numerical results on SWOrRD data showed the promising potential of our method on explosives detection.

The model considered in the paper is a linear and stationary model which assumes no shift/squeeze in the spectral lines. A future line of work will study how to build this nonlinear effect into the identification model. Given that the shift/squeeze amount is small, one may explore the idea of image registration. We also plan to study more reliable and efficient methods for the residuals decomposition, as their success highly depends on a viable working assumption on the pure signals.

The semi-blind source identification problem we addressed here also has analogues in detecting atmospheric trace gases with the so called differential optical absorption spectroscopy. Analysis of the fitting residuals for mysterious species arises there as well, and is non-convex in general. Recently the authors achieved some success in this direction based on a similar semi-blind framework, although the details of fitting and blind source identification process are quite different [20].
References

[1] J. Boardman, Automated spectral unmixing of AVIRIS data using convex geometry concepts, in Summaries of the IV Annual JPL Airborne Geoscience Workshop, JPL Pub. 93-26, Vol. 1, pp 11–14 ( 1993).

[2] C-I Chang, ed., Hyperspectral Data Exploitation: Theory and Applications, Wiley-Interscience, 2007.

[3] G. Comanescu, C.K. Manka, J. Grun, S. Nikitin, D. Zabetakis, Identification of explosives with two-dimensional ultraviolet resonance Raman spectroscopy, Applied Spectroscopy, Vol. 62, No. 8, pp. 833-839 (2008)

[4] K. Faulds, W.E. Smith, and D. Graham, Evaluation of Surface-Enhanced Resonance Raman Scattering for Quantitative DNA Analysis, Anal. Chem., 76 (2), pp 412–417 (2004)

[5] C. A. Froud, I. P. Hayward, and J. Laven, Advances in the Raman depth profiling of polymer laminates, Applied Spectroscopy, Vol. 57, Issue 12, pp. 1468–1474 (2003)

[6] M. Gaft, L. Nagli, Standoff laser based spectroscopy for explosives detection, Proc. SPIE 6737, 673903 (2007).

[7] J. Grun, C.K. Manka, S. Nikitin, D. Zabetakis, G. Comanescu, D. Gillis, and J. Bowles, Identification of Bacteria from Two-Dimensional Resonant-Raman Spectra, Anal. Chem., 79, pp. 5489-5493 (2007).

[8] Z. Guo and S. Osher, Template matching via $\ell_1$ minimization and its application to hyperspectral target detection, Tech. Rep. 09-103, UCLA, www.math.ucla.edu/applied/cam/, 2009.

[9] A. Hyvärinen, J. Karhunen and E. Oja, Independent Component Analysis, John Wiley and Sons, New York, 2001.

[10] D. D. Lee and H. S. Seung, Learning of the parts of objects by non-negative matrix factorization, Nature, 401 (1999) pp. 788–791.

[11] S. Miron, M. Dosso, C. Carteret, S. Margueron, and D. Brie, Joint processing of the parallel and crossed polarized Raman spectra and uniqueness in blind nonnegative source separation, Chemometrics and Intelligent Laboratory Systems 105, 1 (2011) pp.7–18

[12] D. A. Long, The Raman Effect: A Unified Treatment of the Theory of Raman Scattering by Molecules, John Wiley & Sons. Ltd, 2002.

[13] J.V. Miller and E.G. Bartick, Forensic Analysis of Single Fibers by Raman Spectroscopy, Applied Spectroscopy, Vol. 55, Issue 12, pp. 1729-1732 (2001).

[14] W. Naanaa and J.–M. Nuzillard, Blind source separation of positive and partially correlated data, Signal Processing 85 (9) (2005), pp. 1711–1722.
[15] I. Notingher, S. Verrier, S. Haque, J. M. Polak, L.L. Hench, *Spectroscopic study of human lung epithelial cells (A549) in culture: living cells versus dead cells*, Biopolymers, Volume: 72, Issue 4, pp. 230-240 (2003)

[16] C.J. Rowlands and S.R. Elliott, *Improved blind-source separation for spectra*, Journal of Raman Spectroscopy, to appear.

[17] S. Šašić and D.A. Clark, *Defining a strategy for chemical imaging of industrial pharmaceutical samples on Raman Line-Mapping and global illumination instruments*, Applied Spectroscopy, Vol. 60, Issue 5, pp. 494-502 (2006).

[18] Y. Sun, C. Ridge, F. del Rio, A.J. Shaka, and J. Xin, *Postprocessing and Sparse Blind Source Separation of Positive and Partially Overlapped Data*, Signal Processing 91(8)(2011), pp 1838-1851.

[19] Y. Sun, J. Xin, *A Recursive Sparse Blind Source Separation Method and its Application to Correlated Data in NMR Spectroscopy of Biofluids*, Journal of Scientific Computing, to appear, 2011.

[20] Y. Sun, L.M. Wingen, B.J. Finlayson-Pitts, and J. Xin, *A Semi-Blind Source Separation Method for Differential Optical Absorption Spectroscopy of Atmospheric Gas Mixtures*, arXiv:1109.6370[math.NA], Sept 28, 2011.

[21] G. Turrell, *Infrared and Raman Spectra of Crystals*, Academic Press, 1972.

[22] M.E. Winter, *N-findr: an algorithm for fast autonomous spectral endmember determination in hyperspectral data*, in Proc. of the SPIE, vol. 3753, 1999, pp 266-275.

[23] Y. Yao, Q. Li, J. Zhang, R. Liu, L. Jiao, Y. T. Zhu and Z. Liu, *Temperature-mediated growth of single-walled carbon-nanotube intramolecular junctions*, Nature Materials 6, pp. 283 - 286 (2007)

[24] W. Yin, S. Osher, D. Goldfarb, J. Darbon, *Bregman iterative algorithm for $\ell_1$-minimization with applications to compressive sensing*, SIAM J. Image Sci, 1(143), pp 143-168, 2008.