INTRODUCTION

Dengue is a major public health problem in tropical and subtropical regions worldwide and is caused by four serotypes of dengue virus (DENV-1, -2, -3, -4), a flavivirus transmitted to humans by Aedes mosquitoes (1). DENV infection can be asymptomatic or of dengue virus-infected cells.

NK CELLS IN VIRAL INFECTIONS

Natural killer cells are innate lymphocytes specialized in defense against viral and intracellular bacterial infections and tumors (15). NK cells share some characteristics with the adaptive immune system and may possess specific memory features against some viruses and antigens (16). They can be rapidly recruited into infected organs and tissues by chemoattractant factors produced by virus-infected cells and activated resident macrophages and DCs, which are a major source of the interferon IFNα/β that induces NK cell proliferation and activation (17, 18). Reciprocally, NK cells can shape DCs activation and subsequently the adaptive response (17). Once activated, NK cells fight infection by producing chemokines and anti-viral cytokines, mainly IFNγ and MIP1-β, and by recognizing and eliminating infected cells by antibody-dependent cell-mediated cytotoxicity (ADCC) or by direct recognition through their activating receptors (15). NK cells have activating and inhibitory receptors that allow them to recognize stressed cells, tumors, and pathogen-infected cells and to differentiate them from healthy cells (15, 19). Most of the inhibitory receptors recognize classic and non-classic major histocompatibility complex class I (MHC I) molecules, and many viruses decrease the expression of MHC I molecules in infected cells to escape the CD8+ T-cell response, thereby becoming more vulnerable to NK cell recognition (19–21).

Viruses-infected cells often induce or increase the expression of ligands at their surface, allowing for recognition by NK cell activating receptors, including NKG2D, DNAM-1, CD94-NKG2C; the NCR receptors Nkp46, Nkp30, Nkp44; and others (19, 20, 22). The ligands include host stress-induced molecules and viral
proteins (20). To date, the ligands for many of the activating receptors (for example the NCRs) are still unknown, and their expression has been detected indirectly by cell staining with recombinant receptors or by blocking of killing with receptor-specific antibodies. A better characterization of NK ligands is needed. NKG2D is the most well-characterized NK activating receptor, and ligand binding leads to target killing and cytokine production (22). NKG2D ligand expression is increased by "stress" conditions, including viral infections (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23). In humans, eight ligands have been described for NKG2D, including MICA, MICB, and ULBP1-6. In addition to expression of NKG2D ligands on the surface of infected cells, soluble isoforms can be released into the serum, although the physiological relevance of the soluble ligands is controversial (22, 23).

ROLE OF NK CELLS IN DENGUE DISEASE

Many studies suggest that NK cells play a role in the response against DENV infection, principally in the early infection stages by limiting DENV replication. A higher absolute number of NK cells was observed in patients with mild dengue fever (DF) compared with children who developed DHF (27–29). However, the percentage of NK cells and CD8+ T cells expressing CD69, a marker of activation, was higher early during infection of children who developed DHF (27, 30). Homchampa et al. found evidence of NK cell cytotoxicity against non-infected K562 target cells from children with acute dengue, and the cytolytic activity was increased on a per-cell basis in the early disease stages of dengue compared with healthy controls, and was even higher in the most severe form of the disease. It was suggested that this NK cell activity was associated with higher viremia in the more severe cases. Studies in adults showed that patients with mild disease had higher numbers of NK cells, with the majority of cells having increased expression of activation markers (CD69, CD38, and HLA-DR), adhesion molecules (CD11a and CD44) (Table 1), and markers of intracellular cytotoxic granules (TIA-1), in contrast to severe dengue where reduced NK cell numbers were observed (31). The authors suggest that higher NK cell percentages and activity might indicate a good prognosis of disease. In a genome-wide association study (GWAS), it was shown that the transcriptome of blood cells from children with DSS was characterized by decreased abundance of transcripts related to T and NK responses, probably not due to a difference in lymphocyte counts but to an impaired response (32). The differences observed in NK cell numbers and percentages in these studies may be explained by differences in age (children versus adults), ethnicity, time of infection, and the experimental methods to define NK cells and their activation. Nonetheless, all these observations point to the importance of NK cells and their activation during early DENV infection. The activation and phenotype of NK cells during dengue acute infection is further developed by Petitdemange et al., under the research topic “protective immune response to dengue virus infection and vaccines: perspectives from the field to the bench.”

The protective role of NK cells in the response against DENV is supported in mice models of the disease. In immunocompetent A/J mice, the early activation of NK and B cells was associated with the control of the viral load and the prevention of disease (33, 34). In C57BL/6 mice, the recruitment of NK and NKT cells by mast cells to the site of infection was also crucial for viral clearance (35), underlining the importance of these cells during the host early response against DENV. The early recruitment of NK cells to the liver was induced in part by CXCL10 (IP-10), and in the liver NK cells produce effector molecules (perforin, granzyme A, and granzyme B) needed for viral clearance (36). On the other hand,

Table 1 | Summary of the published findings related to ligands for receptors on NK cells and dengue virus infection.

| Ligand                  | Study type | Observation                                                                 | Receptor on NK cells | Probable effect                  | Reference                           |
|-------------------------|------------|------------------------------------------------------------------------------|----------------------|----------------------------------|-------------------------------------|
| ICAM-1                  | Acute patients' lymphocytes | Adhesion molecules (ICAM-1 and LFA-1) are increased in NK cells in the acute phase of dengue disease | LFA-1                | adhesion                         | Azeredo et al. (31)                 |
| DENV-specific antibodies| In vitro with activated peripheral blood lymphocytes | NK cells are the principal cells active in the ADCC against DENV-infected cells | CD16                 | Activation, ADCC                | Kurane et al. (42, 43) and Laoprasopwattana et al. (44) |
| DENV ligands (probably: protein E) | In vitro activation of NK cell toward E protein and VLP of flaviviruses | Protein–protein interaction between nKp44 and cells expressing DENV-E proteins. Interaction of NKP44 with E from another flavivirus (WNV) induces killing and IFNγ production | NKP44                | Activation?                      | Hershkovitz et al. (55)             |
| MICA                    | Allele association | MICA alleles associated with dengue symptomatic infection                    | NKG2D                | Activation?                      | Garcia et al. (59)                 |
| MICB                    | Allele association, GWAS | MICB alleles associated with symptomatic infection and dengue shock syndrome | NKG2D                | Activation?                      | Garcia et al. (59), Khor et al. (60), Whitehorn et al. (61) |
NK cells appear to play not only a protective role in dengue disease, as it was shown that after intrahepatic infiltration, NK cells were responsible for cell death in the liver at the early phase of infection, whereas CD8+ T cells were responsible for damage later (37). The mechanism of this cell death has not been elucidated, but the authors suggested that NK cells were killing DENV-infected cells. It is possible that under some conditions, the elimination of DENV-infected cells by NK cells can be exacerbated in a way that the immune effector cells become responsible for organ injury. Similarly, NKT cells in some conditions could be detrimental during dengue pathogenesis, in part by inducing NK cells and neutrophils activation (38).

Natural killer cells may play an important role in early DENV infection in vivo also by producing together with γδ T cells, IL-22 and IL-17A, which may influence dengue disease outcome (39). NK, NKT, and T cells that can respond in a non-TcR-dependent fashion are a major source of IFNγ in immune responses induced by inactivated DENV (40); however, in this context, the frequency of IFNγ-producing cells observed was very low. Using C57BL/6 mice, a study confirmed the role of NK cells in the early IFNγ production induced by IL-12 and IL-18 in response to DENV, key for the control of viral load and DENV-2-associated disease severity and lethality (41).

Natural killer cells can mediate ADCC against DENV-infected cells (42, 43) and this mechanism may be important during secondary infections when antibodies to DENV are present. Indeed, ADCC activity in plasma obtained before secondary DENV-2 or DENV-3 infection correlated with serotype-specific neutralizing antibody titers, anti-DENV IgG1 levels, and a multitypic PRNT50 pattern (44). Interestingly, a higher level of ADCC activity measured before secondary DENV-3 infection was associated with lower subsequent viremia, which suggests a protective role for antibodies and NK cells; however, this association was not observed for secondary DENV-2 infection. In another study, ADCC was correlated with DENV surface antigen expression, suggesting recognition by anti-DENV antibodies (42), whose Fc region is then recognized by the activating low affinity CD16 (FcγRIII) receptor on NK cells (45).

Human NK cells (CD3−CD16+CD11b+ cells) can lyse DENV-infected cells to a greater level than uninfected cells even in the absence of antibodies, suggesting a mechanism of direct recognition as well (42, 43). These studies were performed using DENV-infected Raji cells as targets. The NK receptors and their ligands implicated in the direct recognition of DENV-infected cells have not been fully elucidated, indicating the need for future studies.

**Expression of NK Cell Ligands During Dengue Disease**

Viruses try to escape the immune response of the host. As NK cells are crucial players in the anti-viral response, many viruses induce the up-regulation of MHC I, which serve as ligands for NK inhibitory receptors, in order to dampen the NK cell response even if enhancing expression of MHC I might increase their recognition by CD8+ T cells (20). DENV and other flaviviruses induce the up-regulation of MHC I (46–51). Expression of flavivirus pr-M protein in hamster cells induced increased surface expression of MHC I, although the authors suggested this could be an incidental consequence of viral assembly rather than a specific mechanism of immune evasion (52). It has also been shown that human cell lines expressing the non-structural (NS) proteins of DENV up-regulate MHC I (HLA-A, -B, -C) expression at their surface by TAP-dependent and TAP-independent pathways (46). This resulted in a lower sensitivity to lysis by NK cells, probably due to recognition by the corresponding inhibitory receptors (e.g., KIR2DL1 for HLA-C) and the inhibitory receptor LIR-1 that recognizes all MHC I proteins (Table 1). Further studies are needed to determine if MHC I up-regulation by DENV is the response to a particular viral protein, viral replication itself, or type-I IFNs induced by viral infection, and whether it plays a role in immune evasion of NK cells in vivo. Neurotropic flaviviruses, such as West Nile virus (WNV), can transiently activate and then suppress NK cell activity (53). Future studies are needed to determine if this happens during DENV infection by analyzing NK cell activity at different time points post-infection.

Even if DENV NS proteins induce MHC I up-regulation, NK cells can kill DENV-infected cells (28, 42, 43), suggesting that the signals for NK cell activation overcome the inhibitory signals. As described above, NK cells can kill DENV-infected cells by ADCC mediated by CD16 (42, 44) (Table 1), which is expressed on resting human NK cells and can induce a strong activating signal leading to cytolyis (20, 54). Interestingly, NK cells can also kill DENV-infected cells in the absence of antibodies (42), implicating a direct recognition by NK cell activating receptors. It has been reported that the activating receptor NKP44 can interact with the envelope protein (E) of DENV (55) (Table 1). NKP44 has also been implicated in killing WNV-infected cells after blocking the inhibitory receptor LIR-1 on NK cells, and this also induced IFNγ production. Further studies are necessary to determine if recognition of DENV-E by NKP44 on NK cells can induce a similar response against DENV-infected cells. NKP44 is expressed on activated, but not resting, NK cells (56) and can trigger NK cell killing of both tumor and virus-infected cells (57, 58).

Recently, a sequencing-based typing method and genotyping of asymptomatic DF and DHF patients in Cuba uncovered an association of certain alleles of the MICA and MICB genes (MICA*008 and MICB*008) with symptomatic DENV infection (59) (Table 1). The importance of MICB in dengue susceptibility was also indicated by GWAS with a large number of pediatric cases in Vietnam, where certain MICB and PLCE1 alleles showed a significant association with DSS (60). These results were confirmed by a study showing that MICB rs3132468 and PLCE1 risk genotypes were also associated with less severe clinical phenotypes of dengue in adults as well as with DENV infection in infants (61). This strongly suggests a role for this MICB variant in susceptibility to overall clinically apparent dengue disease. It still has not been determined whether the association between these NKG2D ligands and dengue clinical responses is directly due to the function of MICA/B molecules in dengue pathogenesis. Nonetheless, the importance of NKG2D ligands in the NK cell response against other viral infections (22) supports this hypothesis. Given the role of MICB in activation of NK, NKT, and CD8+ T cells through the NKG2D receptor, these findings support a central role for these cell types in shaping the outcome of DENV infection. It
is plausible that the MICB risk-associated phenotype is associated with an impaired NK cell response, potentially resulting in a higher in vivo virus titer and an increased risk of developing both symptomatic and severe dengue. Furthermore, inefficient induction of cytokines secreted by NK cells might result in dysregulated T-cell responses that may also shape the clinical phenotype (62). NKG2D ligand (MICA/B) expression on DENV-infected cells may allow direct recognition by NK cells, which might be important for the early innate immune response against DENV infection, leading to either more effective control of viral infection or alternatively contributing to the disease pathology. The expression pattern of these NKG2D ligands in DENV-infected cells in vitro and in vivo needs to be determined. It is also necessary to determine if in acute viral infections such as DENV, the production of soluble NKG2D ligands can also be observed and if this impacts dengue clinical manifestations. Because ligands of other NK activating receptors, such as DNAM-1, can be induced by "stress" (23), further studies are needed to characterize all the NK receptor ligands induced during DENV infection, and the results may depend on the cell-type analyzed, as well as the DENV serotype. A complete characterization of DENV-infected cell recognition by NK cells (Figure 1) is crucial to better understanding the role of these cells in dengue disease.

**CONCLUSION**

The innate immune response, and particularly type-I IFN and NK cells, plays a key role during the early infection events due to its ability to rapidly limit viral dissemination and to affect the antigen-specific, adaptive immune responses to effectively clear pathogens (8, 9). More studies in animal models and human populations will enable deciphering NK cell responses during DENV infection in vivo. Furthermore, in vitro experiments will also be needed to determine which NK receptors and ligands are implicated in DENV-infected cell recognition and NK cell-mediated killing and cytokine production. To date, only the upregulation of MHC I and the induction of a putative NKP44 ligand (DENV-E protein) have been reported in DENV-infected cells. Are there other ligands that play a role in NK cell recognition induced during DENV infection? Genetic studies suggest an important role for the NKG2D ligands MICA and MICB; however, as yet no functional experiments have validated this hypothesis. There is still much to do to determine if ligands of other NK receptors, for example HLA-E for CD94/NKG2A-C and the ligands for the activating receptors 2B4 and DNAM-1, are induced or repressed during DENV infection, and whether this has an implication in the immune response to DENV and its clinical outcome. Finally, the characterization of NK receptor ligands and the NK cell phenotype in patients’ blood cells will provide insights into the NK cell subsets activated during dengue disease. It will also establish possible associations between NK cell activity or NK cell ligand expression and protection from disease and/or increased dengue severity. Whether DENV interferes with innate anti-viral immunity mediated by NK cells at early times of infection and whether DENV virulence might be associated with its ability to counter the host cell defenses are critical issues that remain to be elucidated.
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