Tree Species Diversity, Composition and Aboveground Biomass Across Dry Forest Land-Cover Types in Coastal Ecuador

Xavier Haro-Carrión¹², Bette Loiselle²³, and Francis E. Putz²⁴

Abstract
Tropical dry forests (TDF) are highly threatened ecosystems that are often fragmented due to land-cover change. Using plot inventories, we analyzed tree species diversity, community composition and aboveground biomass patterns across mature (MF) and secondary forests of about 25 years since cattle ranching ceased (SF), 10–20-year-old plantations (PL), and pastures in a TDF landscape in Ecuador. Tree diversity was highest in MF followed by SF, pastures and PL, but many endemic and endangered species occurred in both MF and SF, which demonstrates the importance of SF for species conservation. Stem density was higher in PL, followed by SF, MF and pastures. Community composition differed between MF and SF due to the presence of different specialist species. Some SF specialists also occurred in pastures, and all species found in pastures were also recorded in SF indicating a resemblance between these two land-cover types even after 25 years of succession. Aboveground biomass was highest in MF, but SF and Tectona grandis PL exhibited similar numbers followed by Schizolobium parahyba PL, Ochroma pyramidale PL and pastures. These findings indicate that although species-poor, some PL equal or surpass SF in aboveground biomass, which highlights the critical importance of incorporating biodiversity, among other ecosystem services, to carbon sequestration initiatives. This research contributes to understanding biodiversity conservation across a mosaic of land-cover types in a TDF landscape.

Keywords
aboveground biomass, pasture, plantations, secondary forest, tree diversity, tropical dry forest

Introduction
Global biodiversity is declining at alarming rates (Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services [IPBES], 2019) and land-cover change is the main cause (Daskalova et al., 2020; IPBES, 2019). The relationship between land-cover change and biodiversity loss is particularly evident in biodiversity hotspots in tropical dry forests (TDFs e.g., Sloan et al., 2014). TDFs are among the most highly threatened ecosystems on Earth, with the largest remnants located in South America (Miles et al., 2006) which is particularly understudied relative to other TDF regions (Sanchez-Azofeia et al., 2005).

TDFs in South America were mostly cleared for agriculture, particularly cattle ranching (Aide et al., 2013; Miles et al., 2006). Despite a long history of land use change in the region, our understanding about the regeneration ecology and conservation value of fragments that remain, including old-growth and secondary forests, as well as the managed land-cover types that now characterize TDF landscapes is limited. Some studies suggest that species richness in regenerating TDF could reach similar values to those of old-growth forest. This regeneration could happen at similar or even greater rates than those observed in wet forests because of the
resprouting capacity and dominance of wind-dispersed species in TDF (Chazdon, 2014; Chazdon et al., 2007; Hilje et al., 2015; Kennard, 2002; Lebrija-Trejos et al., 2008). In Mesoamerica, TDF in the intermediate stages of regeneration from pasture were found to be richer in species than older forests (Hilje et al., 2015), and in Bolivia species richness in TDF reached mature forest levels a couple of decades after cessation of slash-and-burn agriculture (Kennard, 2002). In regards to stem densities, adult tree densities tends to decrease as regeneration progresses but that of saplings could show different pattern (Dupuy et al., 2012). While these findings are promising for TDF regeneration, other studies suggest that due to soil damage, TDF recovery after shifting cultivation can be slow (Lawrence et al., 2007). In TDF regenerating from pastures, persistent alien grasses can limit tree regeneration (Cabin et al., 2000; Lyons-Galante & Haro-Carrión, 2017). Rainfall patterns and the presence of established trees also affect succession rates and trajectories (Derroire et al., 2016; Kennard et al., 2002). Despite these differences in species recovery, there is agreement that, as in wet forests, species composition recovery lags behind changes in species densities and forest structural changes (e.g. biomass, stem density) in TDF (Chazdon, 2014; Chazdon et al., 2007). These findings indicate that documenting species richness and community composition in mature and secondary TDF is still needed to improve our knowledge of how diversity is maintained over a broad range of land-cover types and across various TDF landscapes within the Neotropics.

Few evaluations of biodiversity in TDF managed land-uses (e.g. agriculture, plantations) exist, probably because it is expected that most managed land-uses host fewer tree species than forest. A study by Mora et al. (2016) in Mexico reported trade-offs between tree species richness and fodder production, but observed no trade-off between tree species richness and aboveground biomass in old-growth forests and pastures. In landscapes where tree plantations occur, these biomass – species richness relationship could be different because plantations, like forests, accumulate high amounts of biomass but are less diverse than forests. Few studies have directly compared biomass in natural and plantation forests in TDF landscapes. One study in India indicates that plantation forests store more biomass than natural forests (Guha et al., 2019). In the Neotropics, comparisons across studies suggest a similar trend (Kirby & Potvin, 2007; Kraenzel et al., 2003; Read & Lawrence, 2003; Wishnie et al., 2007). Tree plantations can have a broad range of management objectives in TDF landscapes. Some could have restoration purposes and involve native species while others involve introduced species for timber production (Assis et al., 2013; Kirby & Potvin, 2007; Maneschy et al., 2010; Vleut et al., 2013; Wishnie et al., 2007). For instance, teak (Tectona grandis), an introduced species in the Americas, grows in TDF regions and has even been proposed as a climate mitigating strategy to sequester atmospheric carbon (Kirby & Potvin, 2007; Kraenzel et al., 2003). Because of these varied objectives, careful attention is needed to understand the ecological role of plantations. For example, if carbon sequestration is considered, other ecosystem services and biodiversity could be in jeopardy (Coomes et al., 2008). Given this complexity, it seems critical to document both biomass and the richness and composition of species over natural and managed land-cover types in TDF.

This research investigates variation in tree species diversity, community composition, and aboveground biomass across the main land-cover types of a highly fragmented TDF landscape in Ecuador. We aim to fill gaps in knowledge about biodiversity conservation over a broader range of land-cover types, including managed land-uses, and to provide critical information about floristic composition and species diversity for a TDF region less documented in the scientific literature.

**Methods**

**Study Area**

The study landscape encompasses an area of about 36,000 ha of semi-deciduous tropical vegetation between the towns of Pedernales (0°4’20”N 80°3’0”W) and Jama (0°12’07”S 80°15’49”W) (Figure 1), considered the largest remnant of this vegetation type in Ecuador (Neill, 1999). Lowland semi-deciduous tropical vegetation extends along the coast approximately 20 km inland with elevations of 100–400 m. This vegetation type is characterized by temperatures around 25°C with little seasonal variation, annual precipitation of about 1500–2500 mm with a dry season of around 4 months with precipitation as low as 3 mm per month (Josse & Balslev, 1994; Neill, 1999). The terrain includes steep slopes with valleys used for agriculture and cattle ranching. Semi-deciduous forest canopy cover varies; some forests have relatively open canopies and dense understories while others have more closed canopies with lower understory development. Some tree species are thorny and some lose their leaves during the dry season (e.g., Cochlospermum vitifolium and Tabebuia chrysantha). Species characteristic of this vegetation type include Gallesia integrifolia (Phytolacaceae), Triplaris cumingiana and Cuculoba mollis (Polygonaceae), Pseudolmea rigida (Moraceae), and Eugenia spp. (Myrtaceae) (Sierra, 1999).

The landscape is about 40% forested, with about half relatively well-preserved forests while the other half is either severely degraded (i.e. selectively logged) or
The major land-cover types in the study area include mature forest (MF), secondary (SF) and degraded forest, pasture, and forestry plantation (PL) (Haro-Carrión & Southworth, 2018) (Figure 1A). MF with many old-growth characteristics but some previous anthropogenic disturbances (e.g., light selective logging) accounts for about 15% of the landscape. Some MF occurs in slopes, likely because these areas are less suited for cattle, but some occur in relatively flat areas and by the coastline because of land-management choices. A couple are now protected in private reserves. SF results from pasture abandonment or pasture-fallow cycles. They are reported to be about 25 years old by landowners, which in the majority of cases was corroborated by remote sensing (Haro-Carrión & Southworth, 2018). SF includes secondary and degraded forests; the extent of SF in the landscape is not fully known because degraded and SF are difficult to differentiate using Landsat data, but together they account for about 25% of the landscape (Haro-Carrión & Southworth, 2018). Accounting for about 50% of the study area, pasture dominates the landscape and is planted with exotic grass species used in cattle ranching. Typically, pastures have scattered trees of several different species that provide shade for the cattle (Figure 1B and C). Even-aged mono-cultural PL of teak (*Tectona grandis*), pachaco (*Schizolobium parahyba*), and balsa (*Ochroma pyramidale*) cover about 10% of the landscape. Teak and pachaco PL were estimated to be 15–20, and balsa PL 10–15 years old. The study area is considered a biodiversity hotspot because of low cover of native vegetation and its exceptionally high species diversity and endemism (Myers et al., 2000).

**Field Data Collection**

Field inventories during the summers of 2010 and 2012 were used to estimate aboveground biomass and tree diversity in all major land-cover types. The entire study area is characterized by privately owned property. We attempted to sample across the entire study area, but site selection was constrained by the ability to secure permission to access and sample sites. An average distance of approximately 7,500 m separates MF and SF.
sites from one another, but some are less than 500 m apart. SF typically bordered both pastures and MF while PL of all types were typically embedded in the pasture matrix (Figure 1B).

In MF, SF, and pastures at random locations, we established 60 × 60 m plots to sample trees >20 cm DBH (stem diameter at 1.3m or above buttresses) with a nested 20 × 20 m plot for trees 10–20 cm DBH (modified after Magnusson et al., 2005; Phillips et al., 2003). Only the smaller plots were used to sample PL because tree species richness and variation in stem density was low and did not require a more complex sampling approach. To avoid the effect of edges, we located sampling plots at distances >50 m from edges, which was facilitated by characteristically sharp edges in most forest (both MF and SF) – pasture edges (Figure 1C). Less pronounced edges were found of some SF. In these cases, we tried to locate our plots in the middle of the patch. We sampled a total of eight MF, 13 SF, 10 pastures, four Tectona grandis PL, and three Schizolobium parahyba and Ochroma pyramidale PL. Species were categorized as endemic, native, or introduced to Ecuador and noted for their conservation status based on IUCN criteria. Taxonomy is based on https://www.tropicos.org/. Specimen processing was undertaken in the facilities of the QCA Herbarium and specialists assisted with species identification in the QCA and QCNE Herbaria in Quito. Voucher specimens were deposited at QCA and QCNE.

**Tree Species Diversity and Composition**

To compare land-cover types on the basis of tree diversity, we used individual-based rarefaction curves and the Mao Tao estimator computed with EstimateS software (V.8.2) to correct for differences in stem densities (e.g., Colwell et al., 2004; Gotelli & Colwell, 2001). The significance of observed differences in species richness between land-cover types (at P < 0.05) was evaluated by visually comparing rarefaction curves and their associated 95% confidence intervals (CIs). PL of Schizolobium parahyba, Ochroma pyramidale and pastures did not have enough individuals to be included in rarefaction analysis.

To assess tree community similarities of the eight MF and 13 SF, we examined patterns in assemblage composition using a multidimensional scaling analysis (NMDS; Saeed et al., 2018). To deal with rarely sampled species, the species abundance by site matrix was transformed using a Wisconsin double standardization (Bray & Curtis, 1957). Then, we used the transformed matrix to create a Bray-Curtis dissimilarity matrix and used this matrix to generate an NMDS of tree species composition by site. To understand separate sites in terms of their species composition, we report which tree species had high and low positive loadings on both axes of the NMDS.

Prior to conduction the NMDS, to examine if proximity of plots among sample sites influenced similarity of tree species compositions, we conducted a Mantel test (Dutilleul et al., 2000; Rossi, 1996). To do this, we generated distances matrices based on geographic distance (i.e. Euclidean distance) and tree composition matrices using the Jaccard index. We rejected the hypothesis that spatial distance affected similarities in tree species composition (Mantel statistic based on Pearson’s product-moment correlation = 0.09126; p = 0.157) and proceeded with the NMDS. The statistical significance of community composition differences between MF and SF was tested using a PERMAVOA (Permutational Multivariate Analysis of Variance) (McArdle & Anderson, 2001). We focused this analysis on MF and SF because species stem density and species richness were too low in pastures and PL to produce meaningful results.

**Biomass Estimation**

Aboveground biomass estimations for MF, SF, pastures and all three PL types were calculated using an equation for ‘moist forests’ based DBH and wood density. This equation was selected because it fits well the annual and seasonal precipitation patterns of the study area (Chave et al., 2005)

\[
AGB_{est} = px \exp \left( -1.499 = 2.148 \ln(D) \\ \quad + 0.207 (\ln(D))^2 - 0.0281 (\ln(D))^3 \right)
\]

where:

- **AGB** – Aboveground biomass
- **p** – Wood density
- **D** – Diameter at breast height (DBH)

For wood density we used the average of reported values for South America for each species (Zanne et al., 2009); if the species was not found in the database we used values for the genus; if the genus was not represented we used the average value for the family (1 species). For 4 species, values from other regions of the world were used because no reported data were found from South America. AGB estimations were scaled up to megagrams per hectare (Mg/ha) for easier interpretation and comparison among land-cover types. We compared land-cover types on the basis of AGB with one-way analysis of variance (ANOVA) after testing for normality with the Shapiro-Wilks method and checked graphically for homoscedasticity by plotting the residuals versus fitted values of the ANOVA model. Statistics were done using R 4.0.2 (R Core Team, 2020), with
‘vegan’ (Oksanen et al., 2019) for community composition, ‘tidyverse’ (Hadley Wickham et al., 2019) for data manipulation, and ‘ggplot2’ (H. Wickham, 2016) for graphing.

Results

Species Diversity

A total of 124 tree species were recorded, including 10 endemic to Ecuador and 15 with IUCN conservation status (Table 1). Species of conservation concern occurred mostly in MF, but half of the species present in MF also occurred in SF and one species was only found in SF. Tree species in pastures and PL were all native with the exception of Tectona grandis (Table 1).

Tree species richness was highest in MF followed by SF, pastures and PL (Table 2). The same sequence was found for rarefaction curve-based estimates of diversity, but excluding PL of Schizolobium parahyba and Ochroma pyramidale, and pastures that did not have enough individuals to be included (Figure 2). Species richness numbers indicate that a similar diversity pattern to that of Tectona grandis PL is expected in PL of the other two species. Pastures were richer in species than PL, with 10 species recorded among 30 trees sampled in 10 pasture sites. Stem densities of trees >10 cm DBH standardized to per hectare values were highest in PL, followed by MF, SF, and pastures (Table 2).

Community Composition

Tree species composition in MF and SF plots differed significantly (PERMANOVA F = 2.3345, p < 0.001). The NMDS produced a solution with a stress value of 0.13, indicating a good representation of the data when reduced to two dimensions. The NMDS indicates that community differences are due to the presence of unique species in each. NMDS MF plots clustered closely while SF were more dispersed, with one (SF13) more similar to MF than SF. This site was classified as MF through remote sensing analysis (Haro-Carrión & Southworth, 2018), so it is possible that it was just miss-reported by landowners as SF when it is actually a selectively-logged forest. On the first NMDS axis, rare MF species had the most negative loadings while rare SF species had the most positive loadings. On the second axis, rare SF species contributed the most to both the higher and lower ends of the axis (Figure 3).

Aboveground Biomass

AGB differed among land-cover types (F = 19.12, p < 0.001); a Tukey post-hoc revealed that MF supported higher AGB on average than any other land cover type. Twenty-five-year-old SF and 15-20-year-old Tectona grandis PL were similar in AGB followed by 15–20-year-old Schizolobium parahyba PL with lower but not significantly different values to those of SF or teak PL. Pastures supported the lowest AGB, but not significantly lower than 10–15-year-old Ochroma pyramidale PL (Figure 4).

Discussion

Land-cover change is driving massive losses of biodiversity globally (Daskalova et al., 2020, IPBES, 2019). Given the irreplaceable regulating, material, and spiritual services of biodiversity, understanding species diversity and community composition patterns across mosaics of land-cover types is of paramount importance, especially in landscapes such as tropical dry forests.

Our data from a TDF in Ecuador on tree species diversity, stem density and community composition indicate that it is possible that SF are in earlier or intermediate stages of succession compared to other TDFs of similar age. Although, it is difficult to make cross-study comparisons with the exiting literature because of different sampling methodologies (i.e. different minimum tree size sampled), findings from other regions could offer some insights. In Mesoamerica, 32-year-old SF regenerating from pastures were richer in species than forests both older and younger than them by decades (Hilje et al., 2015), and chronosequence data indicate species characteristic of MF could start dominating forest composition about 40 years of succession after agricultural abandonment (Lebrija-Trejos et al., 2008). While these findings might suggest that after an additional 5–15 years of succession, what were 25-year old SF at the time of this study could reach species richness numbers comparable to MF, tree stem density and community assemblages patterns indicate otherwise. We found lower tree stem density of trees >10 DBH in SF than in MF, which does not correspond to patterns expected for mid-succession SF (Chazdon, 2014; Chazdon et al., 2007; Dupuy et al., 2012). The absence of MF specialists in SF communities is also characteristic of early or intermediate stages of regeneration (Chazdon, 2014; Hilje et al., 2015; Lebrija-Trejos et al., 2008). However, we acknowledge that successional patterns of both stem density and species composition could vary when trees <10 cm DBH (not considered in this study) are included in the analysis (Dupuy et al., 2012). Finally, SF specialists, absent in MF communities, were found in pastures indicating a close resemblance between pasture and SF after 25 years of succession. Floristic inventory details show that all 10 species found in pastures were also recorded in SF in equal or higher numbers. In contrast, only seven of these 10 species were recorded in MF and in equal or lower numbers than in SF. Some of these species include Guazuma ulmifolia and Cochlospermum

Schizolobium parahyba.
### Table 1. Species Stem Density, Status in Ecuador Accompanied by IUCN Conservation Status If Available, and Wood Densities for Tree Species in Mature Forest (MF; N = 8), Secondary Forest (SF; N = 13), Pastures (Pa; N = 10), and Forestry Plantations (Pl; N = 10).

| Family               | Species                          | Density (ind > 10 cm DBH/ha) | Wood Density (g/cm³) |
|----------------------|----------------------------------|------------------------------|----------------------|
|                      |                                  | MF  | SF  | Pa  | Pl  | Status                        |
| Achariaceae          | *Mayna odorata*                  | 1   | –   | –   | –   | Native                         | 0.61 |
| Anacardiaceae        | *Mauria suaveolens*              | 2   | 2   | <1  | –   | Native                         | 0.31 |
|                      | *Spondias morbin*                | 9   | 1   | –   | 1   | Native                         | 0.40 |
| Annonaceae           | *Annona muricata*                | <1  | 4   | –   | –   | Native and cultivated          | 0.32 |
|                      | *Klarobelia megalocarpa*         | 3   | –   | –   | –   | Endemic vulnerable             | 0.59 |
|                      | *Mosannona pacifica*             | 4   | 2   | –   | –   | Endemic vulnerable             | 0.59 |
|                      | *Rollinia mucosa*                | –   | 2   | –   | –   | Native                         | 0.32 |
| Apocynaceae          | *Rauvolfia littoralis*           | <1  | –   | –   | –   | Native                         | 0.48 |
| Arecaceae            | *Phytelaphas aequatorialis*      | 8   | 4   | –   | –   | Native                         | 0.43 |
| Asteraceae           | *Fulcaldea laurifolia*           | 20  | 3   | –   | –   | Native                         | 0.60 |
|                      | *Vernonanthera patens*           | –   | 13  | –   | –   | Native                         | 0.60 |
| Bignoniaceae         | *Tabebuia chrysantha*            | 2   | 1   | –   | 6   | Native                         | 1.00 |
|                      | *Tabebuia guayacan*              | 44  | 44  | –   | –   | Native                         | 0.82 |
|                      | *Tecoma castaneifolia*           | –   | 18  | –   | –   | Native                         | 0.79 |
| Bixaceae             | *Cochlospermum vitifolium*       | 6   | 17  | <1  | –   | Native                         | 0.22 |
| Burseraceae          | *Bursera graveolens*             | <1  | 2   | –   | –   | Native                         | 0.32 |
| Capparaceae          | *Capparidastrom cf. pachaca*     | 3   | –   | –   | –   | Native                         | 0.68 |
| Celastraceae         | *Maytenus octogona*              | 4   | –   | –   | –   | Native                         | 0.72 |
| Chrysobalanaceae     | *Licania sp.*                    | 1   | –   | –   | –   | Unknown                        | 0.82 |
| Cordiaceae           | *Cordia alliodora*               | 1   | 21  | –   | –   | Native                         | 0.52 |
|                      | *Cordia hebeclada*               | –   | 5   | –   | –   | Native                         | 0.52 |
| Erythroxylaceae      | *Erythroxylum ruizii*            | 4   | –   | –   | –   | Native                         | 0.79 |
| Euphorbiaceae        | *Adelia triloba*                 | 3   | –   | –   | –   | Native                         | 0.54 |
|                      | *Alchornea leptogyna*            | 13  | –   | –   | –   | Native near threatened         | 0.42 |
|                      | *Croton fraseri*                 | –   | 2   | –   | –   | Endemic endangered             | 0.46 |
|                      | *Croton glabellus*               | 54  | 2   | –   | –   | Native                         | 0.46 |
|                      | *Sapium lauriflorum*             | 2   | <1  | <1  | <1  | Native                         | 0.41 |
| Fabaceae             | *Acacia macracantha*             | –   | 2   | –   | –   | Native                         | 0.73 |
|                      | *Acacia sp.*                     | 3   | –   | –   | –   | Native                         | 0.66 |
|                      | *Bauhinia oculata*               | 3   | 2   | –   | –   | Native                         | 0.64 |
|                      | *Bauhinia sp.*                   | 25  | –   | –   | –   | Unknown                        | 0.64 |
|                      | *Brownnea coccinea subsp. angustiflora* | 1   | –   | –   | –   | Native                         | 1.21 |
|                      | *Caesalpinia sp.*                | 3   | –   | –   | –   | Native                         | 1.01 |
|                      | *Centrolobium ochroxylum*        | 5   | <1  | –   | –   | Native                         | 0.69 |
|                      | *Symmodia cf. bauhiniaformia*    | 6   | –   | –   | –   | Native                         | 0.84 |
|                      | *Erythrina smithiana*            | <1  | <1  | –   | –   | Endemic endangered             | 0.19 |
|                      | *Geoffroea spinosa*              | 1   | 1   | –   | –   | Native                         | 0.67 |
|                      | *Inga sp.*                       | 4   | 3   | –   | –   | Unknown                        | 0.58 |
|                      | *Leucaena triechodes*            | –   | 4   | –   | –   | Native                         | 0.65 |
|                      | *Lonchocarpus atropurpureus*     | 6   | 12  | –   | –   | Native                         | 0.73 |
|                      | *Machaerium millei*              | 20  | 17  | –   | –   | Native                         | 0.24 |
|                      | *Ormoria sp.*                    | <1  | –   | –   | –   | Native                         | 0.61 |
|                      | *Prosopis cf. pallida*           | 2   | 2   | <1  | –   | Native                         | 0.88 |
|                      | *Prosopis juliflora*             | –   | <1  | <1  | <1  | Native                         | 0.74 |
|                      | *Pseudosamanea guajapele*        | –   | 11  | <1  | <1  | Native                         | 0.51 |
|                      | *Pterocarpus cf. rohrii*         | 1   | –   | –   | –   | Native                         | 0.46 |
|                      | *Samanea saman*                  | –   | 2   | 1   | –   | Native                         | 0.50 |
|                      | *Schizolobium parahyba*          | –   | –   | 333 | –   | Native                         | 0.35 |
|                      | *Senna alata*                    | <1  | –   | –   | –   | Native and cultivated          | 0.56 |
|                      | *Senna spectabilis*              | –   | 3   | –   | –   | Native and cultivated          | 0.56 |
|                      | *Swartzia littei*                | 5   | 1   | –   | –   | Endemic endangered             | 0.83 |

(continued)
| Family         | Species                        | Density (ind >10 cm DBH/ha) | Wood Density (g/cm³) |
|---------------|-------------------------------|-----------------------------|----------------------|
| Lamiaceae     | Aegiphila alba                | –                           | –                    |
|               | Tectona grandis               | –                           | –                    |
| Lauraceae     | Licaria cf. triandra          | <1                         | 963                  |
| Lecythidaceae | Gustavia angustifolia         | 3                           | –                    |
| Malpighiaceae | Bunchosia cf. cornifolia      | –                           | –                    |
| Malvaceae     | Ceiba trichistandra           | 5                           | –                    |
|               | Eriotheca ruizii              | <1                         | <1                   |
|               | Herrania balaensis            | <1                         | –                    |
|               | Guazuma ulmifolia             | 8                           | <1                   |
|               | Ochna pyramidal              | –                           | <1                   |
|               | Pachira rupicola              | 1                           | –                    |
|               | Pseudobombax millei           | 3                           | 1                    |
| Meliaceae     | Guarea glabra                 | 19                         | –                    |
|               | Ruagea glabra                 | <1                         | –                    |
|               | Trichilia hirta               | <1                         | –                    |
|               | Trichilia sp.                 | –                           | 4                    |
| Moraceae      | Brosimum alicastrum           | 37                          | 2                    |
|               | Castilla elastica             | 2                           | 4                    |
|               | Clarisia biflora              | 1                           | –                    |
|               | Ficus sp.                     | 1                           | <1                   |
|               | Maclura tinctoria             | <1                         | <1                   |
|               | Piratinera guianensis         | <1                         | –                    |
|               | Tropis racemosa               | 1                           | 2                    |
| Myristicaceae | Virola sebifera               | <1                         | –                    |
| Myrtaceae     | Calyptranthes fusca           | 1                           | –                    |
|               | Eugenia aff. floridiana       | 1                           | –                    |
|               | Eugenia aff. oerstediana      | 1                           | –                    |
|               | Eugenia cf. oerstediana       | 1                           | –                    |
|               | Eugenia floridiana            | 1                           | 6                    |
|               | Myrcia cf. fallax             | 1                           | –                    |
|               | Psidium guajava               | –                           | 1                    |
| Nymataginaceae| Neea sp.                      | 10                          | 2                    |
| Olacaceae     | Heisteria cf. acuminata       | <1                         | –                    |
| Petiveriaceae | Gallesia integrifolia         | 6                           | 1                    |
| Picramniaceae | Picramnia latifolia           | 3                           | –                    |
| Polygonaceae  | Coccoloba cf. densifrons      | <1                         | –                    |
|               | Coccoloba cf. obovata         | 13                          | –                    |
|               | Triplaris cumingiana          | 5                           | <1                   |
| Primulaceae   | Jacquinia sprucei             | –                           | <1                   |
| Rhamnaceae    | Ziziphus thyrsiflora          | 5                           | –                    |
| Rubiaceae     | Alibertia sp.                 | 3                           | –                    |
|               | Alsise eggersii               | 10                          | <1                   |
|               | Faramea occidentalis          | 3                           | –                    |
|               | Guettarda acreana             | 20                          | 4                    |
|               | Guettarda hirsuta             | <1                         | –                    |
|               | Joosia sp.                    | <1                         | –                    |
|               | Randia carlosiana             | <1                         | 2                    |
| Rutaceae      | Zanthoxylum martinicense      | <1                         | –                    |
|               | Zanthoxylum riedelianum       | 7                           | 1                    |
| Sabiaceae     | Meliosma occidentalis         | <1                         | –                    |

(continued)
vitifolium, two species previously reported as typical of early stages of post-agricultural succession (Hilje et al., 2015). We believe that given the continuous perturbations from cattle and the presence of exotic grasses in many of the SF we studied (Lyons-Galante & Harocarrion, 2017), succession is happening at a slower pace comparable to that found in other TDF with similar histories of grazing (Cabin et al., 2000).

As expected, tree species richness was lower in pastures and all PL types, but documenting this finding is important for assessing species richness across land-cover types and, in the case of PL, for analysing tree species richness in conjunction with aboveground biomass. Ochroma pyramidale is a native species used in restoration and reforestation projects, and in cases its use has been aligned with biodiversity conservation objectives (Vleut et al., 2013; Wishnie et al., 2007). The Ochroma pyramidale PL studied in this research were considered by landowners as examples of “reforestation” but our findings indicate low species richness and a limited carbon storage when compared to the other analysed land-cover types. Less is known about the use of Schizolobium parahyba for restoration in TDF landscapes, but a similar pattern to that of Ochroma pyramidale might be expected based on its use elsewhere in the Neotropics (Assis et al., 2013; Maneschky et al., 2010). Tectona grandis is an introduced species. Its cultivation has been analyzed in reforestation initiatives in other TDF landscapes (Kraenzel et al., 2003). In the studied landscape, landowners reported limited environmental benefits to its cultivation, did not associate it with carbon sequestration initiatives, and mostly gave

### Table 1. Continued.

| Family        | Species               | MF | SF | Pa | Pl | Status               | Wood Density (g/cm³) |
|---------------|-----------------------|----|----|----|----|----------------------|----------------------|
| Salicaceae    | Casearia sylvestris   | <1 | 2  | –  | –  | Native               | 0.72                 |
| Sapindaceae   | Cupania americana     | –  | 22 | –  | –  | Native               | 0.73                 |
|               | Exothea paniculata    | 5  | 9  | –  | –  | Native               | 0.73                 |
| Sapindus saponaria |                | <1 | 1  | –  | –  | Native               | 0.67                 |
|               | Talisia setigera      | 23 | –  | –  | –  | Native endangered    | 0.84                 |
| Sapotaceae    | Chrysophyllum aff. argenteum | <1 | 1  | –  | –  | Native               | 0.78                 |
|               | Chrysophyllum sp.      | <1 | –  | –  | –  | Native               | 0.76                 |
|               | Pouteria brevipetiola | <1 | –  | –  | –  | Endemic endangered  | 0.78                 |
|               | Pouteria cordiformis  | <1 | –  | –  | –  | Endemic             | 0.78                 |
|               | Pradosia montana      | 15 | –  | –  | –  | Native               | 0.73                 |
|               | Pradosia sp.          | 3  | –  | –  | –  | Native               | 0.73                 |
| Solanaceae    | Acnistus arborescens  | –  | 1  | –  | –  | Native and cultivated | 0.50                 |
|               | Solanum sp.           | –  | <1 | –  | –  | Unknown              | 0.50                 |
| Ulmaceae      | Ampelocera longissima | 1  | –  | –  | –  | Native least concern | 0.67                 |
|               | Ampelocera macphersonii | |     | –  | –  | Native               | 0.67                 |
| Unidentified  | <1                   | –  | –  | –  | –  | Unknown              | 0.61                 |
| Urticaceae    | Cecropia sp.          | –  | 8  | –  | –  | Unknown              | 0.35                 |
|               | Urera baccifera       | 6  | 2  | –  | –  | Native               | 0.17                 |
| Zamiaceae     | Zamia poepiggiana     | 1  | –  | –  | –  | Native least concern | 0.61                 |

**Taxonomy based on Tropicos.org (2020).**

### Table 2. Total Tree Species Richness, Number of Endemics, Number of Species With IUCN Status, and Average Stem Density Among Analysed Land-Cover Types for Trees >10 cm DBH.

| Land-use          | Species richness | No. endemic species | No. species with IUCN status | Area sampled (ha) | Av. stem density (ind/ha) |
|-------------------|------------------|---------------------|-----------------------------|------------------|--------------------------|
| Mature forest     | 99               | 9                   | 14                          | 3.2              | 183 ± 49                 |
| Secondary forest  | 68               | 6                   | 7                           | 5.2              | 130 ± 57                 |
| Pasture           | 10               | 0                   | 0                           | 7.47             | 4 ± 4                    |
| Tectona grandis   | 2                | 0                   | 0                           | 0.16             | 969 ± 120                |
| Schizolobium parahyba | 1            | 0                   | 0                           | 0.12             | 333 ± 14                 |
| Ochroma pyramidale | 1                | 0                   | 0                           | 0.12             | 725 ± 189                |
Despite these characteristics, findings from this research indicate that *T. grandis* could store as much carbon in aboveground biomass as SF, indicating it may compete and possibly outperform SF in carbon-based initiatives. In other TDF landscapes, synergies between forest conservation and carbon storage have been reported (Mora et al., 2016). Our findings suggest that aboveground biomass in many of the studied PL types could reduce these synergies, potentially creating a trade-off between biodiversity conservation and carbon sequestration initiatives.
This research provides a critical understanding of tree diversity, composition and aboveground biomass across the major land-cover types in a dry forest landscape in coastal Ecuador. However, results should be interpreted cautiously considering that a full understanding of landscape characteristics including matrix structure and configuration, forest patch size and shape is lacking and represent a limitation of this research. Likewise, our floristic inventory provides a reliable comparisons of tree species richness, diversity and aboveground biomass across land-cover types, but longer temporal evaluations of community composition and aboveground biomass are needed to fully understand the dynamics of carbon storage and sequestration and community composition change over time. Incorporating seedling and sapling into future research could also help understand tree diversity and community composition differences across land-cover types and clarify patterns of secondary succession. Finally, perhaps the single most significant variable not accounted in this research is land tenure and associated land-management decisions. Many sites challenge documented landscape – species diversity patterns and rather reflect unique land management choices. Species-rich MF are found adjacent to pastures, SF occurs near roads and forest (MF and SF) are in areas less steep than many pastures. These are just some examples that reflect unique management choices with direct implications for biodiversity conservation that require further analysis.

Implications for Conservation

We analysed tree species diversity, community composition and aboveground biomass across mature forests (MF), secondary forests (SF), three types of plantations (PL), and pastures in a TDF landscape in Ecuador. The following are key findings related to forest and biodiversity conservation:

1. MF were more diverse than SF, but detailed analyses indicate that while most endemic and IUCN-categorized species were found in MF (15 total), half of them were also present in SF. This finding agrees with broader findings that recognize the importance of SF in biodiversity conservation, especially in highly fragmented landscapes (Chazdon, 2014; Chazdon et al., 2007).

2. It is critical to evaluate richness and diversity patterns across major land-cover types to fully understand tree diversity and forest succession patterns. In this research, SF resemble pastures in tree species composition, and succession is probably delayed because of the effects of pasture grasses, suggesting some challenges when planning for reforestation.

3. Aboveground biomass and tree diversity were the highest in MF, indicating potential synergies between forest conservation and carbon storage and sequestration initiatives. However, species-poor Tectona grandis PL could host as much as or more aboveground biomass as SF, raising awareness of the importance of
aligning biodiversity conservation (or other ecosystem services) and carbon storage and sequestration in reforestation efforts.

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ORCID iD
Xavier Haro-Carrión https://orcid.org/0000-0002-8048-5380

References
Aide, T. M., Clark, M. L., Grau, H. R., López-Carr, D., Levy, M. A., Redo, D., Bonilla-Moheno, M., Riner, G., Andrade-Núñez, M. J., & Muñiz, M. (2013). Deforestation and reforestation of Latin America and the Caribbean (2001–2010). Biotropica, 45(2), 262–271. https://doi.org/10.1111/j.1744-7429.2012.00908.x
Assis, G. B., de Suganuma, M. S., de Melo, A. C. G., & Durigan, G. (2013). Uso de especies nativas e exóticas na restauração de matas ciliares no Estado de São Paulo [Use of native and exotic species in the restoration of riparian forests in the state of São Paulo] (1957—2008). Revista Árvore, 37(4), 599–609. https://doi.org/10.1590/S0100-6762201300400003
Bray, J. R., & Curtis, J. T. (1957). An ordination of the upland forest communities of Southern Wisconsin. Ecological Monographs, 27(4), 325–349. https://doi.org/10.2307/1942268
Cabin, R. J., Weller, S. G., Lorence, D. H., Flynn, T. W., Sákiá, A. K., Sandquist, D., & Hadway, L. J. (2000). Effects of long-term ungulate exclusion and recent alien species control on the preservation and restoration of a Hawaiian tropical dry forest. Conservation Biology, 14(2), 439–453. https://doi.org/10.1046/j.1523-1739.2000.99006.x
Chao, A., Chazdon, R. L., Colwell, R. K., & Shen, T.-J. (2004). A new statistical approach for assessing similarity of species composition with incidence and abundance data: A new statistical approach for assessing similarity. Ecology Letters, 8(2), 148–159. https://doi.org/10.1111/j.1461-0248.2004.00707.x
Chao, A., Gotelli, N. J., Hsieh, T. C., Sander, E. L., Ma, K. H., Colwell, R. K., & Ellison, A. M. (2014). Rarefaction and extrapolation with Hill numbers: A framework for sampling and estimation in species diversity studies. Ecological Monographs, 84(1), 45–67. https://doi.org/10.1890/13-0133.1
Chave, J., Andalo, C., Brown, S., Cairns, M. A., Chambers, J. Q., Eamus, D., Fölster, H., Fromard, F., Higuchi, N., Kira, T., Lesure, J.-P., Nelson, B. W., Ogawa, H., Puig, H., Riéra, B., & Yamakura, T. (2005). Tree allometry and improved estimation of carbon stocks and balance in tropical forests. Oecologia, 145(1), 87–99. https://doi.org/10.1007/s00442-005-0100-x
Chazdon, R. L. (2014). Second growth: The promise of tropical forest regeneration in an age of deforestation. The University of Chicago Press.
Chazdon, R. L., Letcher, S. G., van Breugel, M., Martinez-Ramos, M., Bongers, F., & Finegan, B. (2007). Rates of change in tree communities of secondary neotropical forests following major disturbances. Philosophical Transactions of the Royal Society B: Biological Sciences, 362(1478), 273–289. https://doi.org/10.1098/rstb.2006.1990
Colwell, R. K., Mao, C. X., & Chang, J. (2004). Interpolating, extrapolating, and comparing incidence-based species accumulation curves. Ecology, 85(10), 2717–2727. https://doi.org/10.1890/03-0557
Coomes, O. T., Grim, F., Potvin, C., & Sima, P. (2008). The fate of the tropical forest: Carbon or cattle? Ecological Economics, 65(2), 207–212. https://doi.org/10.1016/j.ecolecon.2007.12.028
Daskalova, G. N., Myers-Smith, I. H., Bjorkman, A. D., Blowes, S. A., Supp, S. R., Magurran, A. E., & Dornelas, M. (2020). Landscape-scale forest loss as a catalyst of population and biodiversity change. Science, 368(6497), 1341–1347. https://doi.org/10.1126/science.aba1289
Derroire, G., Tigabu, M., Odén, P. C., & Healey, J. R. (2016). The effects of established trees on woody regeneration during secondary succession in tropical dry forests. Biotropica, 48(3), 290–300. https://doi.org/10.1111/btp.12287
Dupuy, J. M., Hernández-Steфанoni, J. L., Hernández-Juárez, R. A., Tetetla-Rangel, E., López-Martínez, J. O., Leyequén-Abarca, E., Tun-Dzul, F. J., & May-Pat, F. (2012). Patterns and correlates of tropical dry forest structure and composition in a highly replicated chronosequence in Yucatan, Mexico: Tropical dry forest structure and composition. Biotropica, 44(2), 151–162. https://doi.org/10.1111/j.1744-7429.2011.00783.x
Dutilleul, P., Stockwell, J. D., Frigon, D., & Legendre, P. (2000). The mantel test versus Pearson’s correlation analysis: Assessment of the differences for biological and
environmental studies. *Journal of Agricultural, Biological, and Environmental Statistics*, 5(2), 131. https://doi.org/10.2307/1400528

Gotelli, N. J., & Colwell, R. K. (2001). Quantifying biodiversity: Procedures and pitfalls in the measurement and composition of species richness. *Ecology Letters*, 4(4), 379–391. https://doi.org/10.1046/j.1461-0248.2001.00230.x

Guha, S., Pat, T., Nath, D. S., & Das, B. (2019). Comparison of biomass in natural and plantation dry forests in India. In B. Pradhan (Ed.), *GCEC 2017* (Vol. 9, pp. 995–1006). Springer Singapore. https://doi.org/10.1007/978-10-8016-6_69

Haro-Carrion, X., & Southworth, J. (2018). Understanding land cover change in a fragmented forest landscape in a biodiversity hotspot of coastal Ecuador. *Remote Sensing*, 10(12), 1980. https://doi.org/10.3390/rs10121980

Hilje, B., Calvo-Alvarado, J., Jiménez-Rodríguez, C., & Sánchez-Azofeifa, A. (2015). Tree species composition, breeding systems, and pollination and dispersal syndromes in three forest successional stages in a tropical dry forest in Mesoamerica. *Tropical Conservation Science*, 8(1), 76–94. https://doi.org/10.1080/19400829150100109

Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. (2019). *Summary for policymakers of the global assessment report on biodiversity and ecosystem services (summary for policy makers)*. Zenodo. https://doi.org/10.5281/ZENODO.3553579

Josse, C., & Balslev, H. (1994). The composition and structure of a dry, semideciduous forest in western Ecuador. *Nordic Journal of Botany*, 14(4), 425–434. https://doi.org/10.1111/j.1756-1051.1994.tb00628.x

Kennard, D. K. (2002). Secondary forest succession in a tropical dry forest: Patterns of development across a 50-year chronosequence in lowland Bolivia. *Journal of Tropical Ecology*, 18(1), 53–66. https://doi.org/10.1017/S0266467402002031

Kennard, D. K., Gould, K., Putz, F. E., Frederixsen, T. S., & Morales, F. (2002). Effect of disturbance intensity on regeneration mechanisms in a tropical dry forest. *Forest Ecology and Management*, 162(2–3), 197–208. https://doi.org/10.1016/S0378-1127(01)00056-0

Kirby, K. R., & Potvin, C. (2007). Variation in carbon storage among tree species: Implications for the management of a small-scale carbon sink project. *Forest Ecology and Management*, 246(2–3), 208–221. https://doi.org/10.1016/j.foreco.2007.03.072

Kraenzel, M., Castillo, A., Moore, T., & Potvin, C. (2003). Carbon storage of harvest-age teak (*Tectona grandis*) plantations, panama. *Forest Ecology and Management*, 173(1–3), 213–225. https://doi.org/10.1016/S0378-1127(02)00002-6

Lawrence, D., D’Odorico, P., Diekmann, L., DeLonge, M., Das, R., & Eaton, J. (2007). Ecological feedbacks following deforestation create the potential for a catastrophic ecosystem shift in tropical dry forest. *Proceedings of the National Academy of Sciences*, 104(52), 20696–20701. https://doi.org/10.1073/pnas.0705005104

Lebrero-Trejos, E., Bongers, F., Pérez-García, E. A., & Meave, J. A. (2008). Successional change and resilience of a very dry tropical deciduous forest following shifting agriculture: Tropical very dry forest secondary succession. *Biotropica*, 40(4), 422–431. https://doi.org/10.1111/j.1744-7429.2008.00398.x

Lyons-Galante, H. R., & Haro-Carrion, X. (2017). Effect of distance from edge on exotic grass abundance in tropical dry forests bordering pastures in Ecuador. *Journal of Tropical Ecology*, 33(02), 170–173. https://doi.org/10.1017/S026647417000062

Magnusson, W. E., Lima, A. P., Luizão, R., Luizão, F., Costa, F. R. C., de Castillo, C. V., & Kinupp, V. F. (2005). RAPELD: A modification of the Gentry method for biodiversity surveys in long-term ecological research sites. *Biota Neotropica*, 5(2), 19–24. https://doi.org/10.1556/S1676-0603200500300002

Maneschy, R. Q., de Santana, A. C., & da Veiga, J. B. (2010). Viabilidad Económica de Sistemas Silvipastorís com Schizolobium parahyba var. Amazonicum e Tectona grandis no Pará [Economic variability of silvopastoral systems with Schizolobium parahyba var. Amazonicum and Tectona grandis in Pará]. *Pesquisa Florestal Brasileira*, 0(60), 45. https://doi.org/10.4336/2009.pfb.06.40.49

McArdle, B. H., & Anderson, M. J. (2001). Fitting multivariate models to community data: A comment on distance-based redundancy analysis. *Ecology*, 82(1), 290–297. https://doi.org/10.1890/012-9658(2001)082[0290:FMMTCD2.0.CO;2

Miles, L., Newton, A. C., DeFries, R. S., Ravilious, C., May, I., Blyth, S., Kapos, V., & Gordon, J. E. (2006). A global overview of the conservation status of tropical dry forests. *Journal of Biogeography*, 33(3), 491–505. https://doi.org/10.1111/j.1365-2699.2005.01424.x

Mora, F., Balvanera, P., García-Frapolli, E., Castillo, A., Trilleras, J. M., Cohen-Salgado, D., & Salmerón, O. (2016). Trade-offs between ecosystem services and alternative pathways toward sustainability in a tropical dry forest region. *Ecology and Society*, 21(4), art45. https://doi.org/10.5751/ES-08691-210445

Myers, N., Mittermeier, R. A., Mittermeier, C. G., Da Fonseca, G. A., & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403(6772), 853.

Neill, D. A. (1999). Vegetación. In P. M. Jorgensen & S. León-Yánez (Eds.), *Catalogue of the vascular plants of Ecuador* (Vol. 75, pp. 13–25). Missouri Botanical Garden Press, St. Louis.

Oksanen, J., Blanchet, F. G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Michin, P. R., O’Hara, R. B., Simpson, G. L., Solymos, P., Stevens, M. H. H., Szoecs, E., & Wagner, H. (2019). *Vegan: Community ecology package*. R package version 2.5-6. https://CRAN.R-project.org/ pack age= vegan

Phillips, O. L., Martínez, R. V., Vargas, P. N., Monteagudo, A. L., Zans, M.-E. C., Sánchez, W. G., Cruz, A. P., Timaná, M., Yli-Halla, M., & work(s), S. R. R. (2003). Efficient plot-based floristic assessment of tropical forests. *J Trop Ecol*, 19(6), 629–645.

R Core Team. (2020). *R: A language and environment for statistical computing*. R Foundation for Statistical Computer. https://www.R-project.org/

Read, L., & Lawrence, D. (2003). Recovery of biomass following shifting cultivation in dry tropical forests of the
Yucatan. *Ecological Applications*, 13(1), 85–97. https://doi.org/10.1890/1051-0761(2003)013[0085:ROBFSC]2.0.CO;2

Rossi, J. -P. (1996). Statistical tool for soil biology. XI. *Autocorrelogram and Mantel test*, 32(4), 10.

Saeed, N., Nam, H., Haq, M. I. U., & Muhammad Saqib, D. B. (2018). A survey on multidimensional scaling. *ACM Computing Surveys*, 51(3), 1–25. https://doi.org/10.1145/3178155

Sanchez-Azofeifa, G. A., Quesada, M., Rodriguez, J. P., Nassar, J. M., Stoner, K. E., Castillo, A., Garvin, T., Zent, E. L., Calvo-Alvarado, J. C., Kalacska, M. E. R., Fajardo, L., Gamon, J. A., & Cuevas-Reyes, P. (2005). Research priorities for neotropical dry forests1. *Biotropica*, 37(4), 477–485. https://doi.org/10.1111/j.1744-7429.2005.00066.x

Sierra, M. R. (Ed.). (1999). *Propuesta preliminar de un sistema de clasificación de vegetación para el Ecuador continental [A preliminary proposal for a vegetation classification system for continental Ecuador]*. Proyecto INEFAN/GEF-BIRF y EcoCiencia.

Sloan, S., Jenkins, C. N., Joppa, L. N., Gaveau, D. L. A., & Laurance, W. F. (2014). Remaining natural vegetation in the global biodiversity hotspots. *Biological Conservation*, 177, 12–24. https://doi.org/10.1016/j.biocon.2014.05.027

Tropicos.org. (2020). *Missouri Botanical Garden*. http://www.tropicos.org

Vleut, I., Levy-Tacher, S. I., de Boer, W. F., Galindo-González, J., & Ramirez-Marcial, N. (2013). Can a fast-growing early-successional tree (Ochroma pyramidale, Malvaceae) accelerate forest succession? *Journal of Tropical Ecology*, 29(2), 173–180. https://doi.org/10.1017/S0266467413000126

Wickham, H. (2016). *ggplot2: Elegant graphics for data analysis*. Springer Verlag.

Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L., François, R., Grolemund, G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T., Miller, E., Bache, S., Müller, K., Ooms, J., Robinson, D., Seidel, D., Spinu, V., … Yutani, H. (2019). Welcome to the Tidyverse. *Journal of Open Source Software*, 4(43), 1686. https://doi.org/10.21105/joss.01686

Wishnie, M. H., Dent, D. H., Mariscal, E., Deago, J., Cedeño, N., Ibarra, D., Condit, R., & Ashton, P. M. S. (2007). Initial performance and reforestation potential of 24 tropical tree species planted across a precipitation gradient in the Republic of Panama. *Forest Ecology and Management*, 243(1), 39–49. https://doi.org/10.1016/j.foreco.2007.02.001

Zanne, A. E., Lopez-Gonzalez, G., Coomes, D. A., Ilic, J., Jansen, S., Lewis, S. L., Miller, R. B., Swenson, N. G., Wiemann, M. C., & Chase, J. (2009). Data from: Towards a worldwide wood economics spectrum (Version 5, p. 204748 bytes) [Data set]. Dryad. https://doi.org/10.5061/DYRAD.234