Accuracy in Trace Analysis

Table 1. Pork irradiation experimental design

| Temperature | Radiation dose (kGy) |
|-------------|---------------------|
| °C          | 0.5  | 1.75 | 3.5  | 5.25 | 7.0  |
| -20°C       | 6    | 6    |      |      |      |
| -10°C       | 6    | 6    |      |      |      |
| 0°C         | 10   | 6    | 10   | 6    | 6    |
| +10°C       | 6    |      | 6    |      |      |
| +20°C       | 6    |      | 6    |      |      |

Each replicate contained seven pork chops; one-half of the replicates were fried after irradiation.

The data obtained from the analyses of the samples described above were used to develop response surface equations which would predict the effect of the irradiation dose and temperature on each vitamin over the entire area covered by the design. From the equation for the response surface for thiamin loss the predicted losses in pork chops irradiated at 0°C and then cooked were as follows: 0 kGy, -1.5% (-0.04%); 0.50 kGy, -10.1% (14.3%); 3.5 kGy, -48.7% (54.3%); and 7.0 kGy, -65.9% (69.7%). The values given in parentheses represent the average of the actual observed values. There was no loss of thiamin in the control on a sample weight basis upon cooking, but there was an overall weight loss of about 30%. The effect of temperature on the degradation of thiamin can be illustrated by comparison of the predicted results obtained at a dose of 3.5 kGy, losses of 32.7 (34.9%) and 63.3% (60.2%) at -20°C and +20°C respectively. The fit of the predicted values to the measured values is indicated by a $R^2$ value of 0.90 for the equation for the response surface.

The effect of radiation on thiamin was different in poultry from that observed in pork. The response surface equation for thiamin loss in chicken breasts cooked after irradiation predicted the following losses of thiamin at 0°C: 0 kGy, +3.2%; 1.0 kGy, -0.38%; 2.0 kGy, -2.2%; 3.0 kGy, -5.9%; 3.5 kGy, -7.4%; and 7.0 kGy, -34.9%. Thus, in the range of greatest interest for the control of salmonella contamination (3.0 kGy) the loss of thiamin was very low especially when compared to losses in pork chops irradiated and cooked in the same manner.

References

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2. Contamination of Foods by Migration from Plastic Packaging Materials

Plasticizers such as di-(2-ethylhexyl)adipate (DEHA), and various phthalates are present in both flexible films and in coatings which come into contact with foods during retail packaging or the use of plasticized “cling-film” in the home. Epoxidized soybean oil (ESBO) is used as a plasticizer and secondary heat stabilizer in food packaging and recently so-called polymeric plasticizers have been introduced. These components can migrate from the packaging materials into the foods for which monitoring is therefore necessary. Stable isotope dilution GC/MS methods have proved to be successful for quantification of the levels of migration.

The stable isotope internal standards d₄-DEHA, d₄-dibutylphthalate, d₄-dicyclohexylphthalate and d₄-diethylphthalate can be readily synthesized from their respective commercially available isotopically-labelled acids, and thereby obtained with high label incorporation and in high chemical purity. Food sample preparation for the determination of these plasticizers involves extraction with acetone/hexane, size-exclusion chromatographic clean-up (automated) in dichloromethane/hexane solvent and a final capillary GC/MS selected ion monitoring quantitative step. For DEHA determinations [1], ions at m/z 129 and 133 are monitored for d₄- and d₄-DEHA respectively, whilst for the phthalates m/z 149 and 153 provide common ions for monitoring a number of unlabelled phthalates and their respective internal standards [2]. The approach has been employed for monitoring migration in a large number of food samples of different types such as cheese, fresh and cooked meat, vegetables, confectionary products and even complete microwave cooked meals [3,4]. In no instance was there any evidence of interference from food components, and the limit of detection of the procedure was 0.1 mg/kg with a RSD of between 1 and 3% [1,2].

So-called polymeric plasticizers are in fact oligomeric mixtures of components, and for example the adipate-based plasticizer “Reoplex R346” has a number average molecular weight some five times greater than the monomeric counterpart. The isotope dilution GC/MS procedure [5] for determining polymeric plasticizer levels in foods involves transmethylation of the polymeric plasticizer to dimethyladipate (DMA), utilizing d₄-DEHA as an internal standard which is in turn transmethylated to d₄-DMA. Using a similar clean-up to that for the other plasticizers outlined above, the GC/MS determination is based on monitoring of m/z 143 and 147 for d₄- and d₄-DMA respectively. The method has been demonstrated as applicable to a diverse range of food types, and to have a limit of detection of 0.1 mg/kg and a RSD of about 4%.

ESBO is a multi-functional additive used at levels of between 3 and 7% in a range of plastics. This material is determined in foods by transmethylation of the triglycerides and then derivatization of the epoxide groups in the fatty acid esters to form compounds with characteristics particularly suitable for GC/MS selected ion monitoring. The initial transmethylation is carried out under basic conditions (sodium methoxide/methanol) to ensure protection of the epoxide group, and derivatization involves formation of the 1,3-dioxolane by treatment with cyclopentanone (followed by BF₃/diethyl ether). The method has been used down to a limit of 0.1 mg/kg without necessitating prior separation of the epoxides from the other naturally occurring fatty acid esters derived from the food lipids. The choice of ions for selected ion monitoring depends on whether the mono-, di- or tri-epoxidized fatty acid esters are to be monitored, but for example for the monoepoxidized species m/z 367 and 396 are detected. The isotopically-labelled internal standard is easily synthesized from commercially available ¹³C-triolein, to give a method with an overall RSD of about 5%.

3. Pesticide Residues in Foods

The determination of pesticide residues in foods by stable isotope dilution presents some difficulties, firstly because of the large number of compounds that have to be simultaneously monitored, and secondly the high proportion of samples containing not detectable levels of pesticides which makes quantification the exception rather than the rule. For these reasons it is probably preferable to initially screen by another method and then re-analyse positive samples by GC/MS adding only the appropriate labelled internal standards for which quantification is required. For organochlorine pesticides at least 15 deuterated and/or ¹³C-labelled standards are commercially available, but these surprisingly have not been utilized for residue analysis and to date our own experience has been limited to the quantification of hexachlorobenzene.
(HCB) using the $^{13}$C$_6$-labelled standard [6]. For the analysis of eggs, sample preparation involves initial treatment with phospholipase, extraction with acetone/hexane and then clean-up of the lipid extract on a water deactivated alumina column. Selected ion monitoring is for $m/z$ 284 and 286 for HCB and 292 and 294 for $[^{13}$C$_6]HCB$. The GC/MS program involves switching between a number of different ions, grouped for different retention time windows to enable monitoring of altogether 10 organochlorine pesticides plus respective isomers. For the future it is intended to increase the number of isotopically labelled standards utilized which will further increase the complexity of the multiple ion monitoring program.

4. **Veterinary Drug Residues in Animal Tissue**

Veterinary drug residues in foods, although obvious candidates for isotope dilution approaches to analysis, do present the same logistical problems as pesticides with the same requirement for multi-residue monitoring. For the determination of the sulphonamide drug, sulphamethazine in kidney samples by isotope dilution GC/MS [7], after addition of d$_4$-sulphamethazine to an acetonitrile slurry of the kidneys and allowing for equilibration, the clean-up involves solvent partition, diazomethane treatment to form the methyl derivative and then HPLC fractionation as a further clean-up stage. In the trapped HPLC fraction GC/MS selected ion monitoring for $m/z$ 227 and 228 for sulphamethazine and 231 and 232 for the deuterated internal standard is carried out for quantification. The limit of detection of the method is around 0.05 mg/kg and the CV is between 3.7 and 5.7% for sulphamethazine spiking levels in the tissue from 0.2 to 1.2 mg/kg. Deuterated sulphamethazine had to be custom synthesized and other stable isotope-labelled drugs are not widely available commercially which hinders development of this approach, as does the fact that many drugs of interest are not amenable to GC. One possible way to overcome this latter difficulty is to maintain the use of isotope dilution but to use MS/MS which also confers advantages of reduced sample preparation and clean-up [8].

### References

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