LETTER TO THE EDITOR

Intranasal application of polyethyleneimine suppresses influenza virus infection in mice

Biao He¹, Yuhong Fu¹, Shuai Xia¹, Fei Yu¹, Qian Wang¹, Lu Lu¹ and Shibo Jiang¹,²

Emerging Microbes and Infections (2016) 5, e41; doi:10.1038/emi.2016.64; published online 27 April 2016

Dear Editor,

Emerging and reemerging viruses that cause respiratory infectious diseases, such as severe acute respiratory syndromes coronavirus (SARS-CoV), Middle East respiratory syndrome coronavirus (MERS-CoV), and influenza viruses, are a significant threat to public health worldwide. Although vaccines are the most effective strategy to prevent viral infection, vaccine development is a long process and may be effective only against the corresponding virus. Therefore, it is essential to develop antiviral agents for intranasal administration as nonspecific prophylaxis against infection by emerging or reemerging respiratory viruses with epidemic or pandemic potential.

Cholera toxin (CT), which acts as a mucosal adjuvant to stimulate mucosal and systemic immune responses, has the potential for intranasal application as an immunotherapeutic against infection by respiratory viruses.¹ However, CT can exacerbate lung pathology after influenza virus infection through the induction of IL-17, a potent proinflammatory cytokine, because co-administration of an IL-17RA neutralizing antibody with CT attenuates lung pathology and increases protection against influenza virus infection.² Therefore, we hypothesized that intranasal application of a mucosal stimulant that induces antiviral cytokines, except IL-17, may be effective against influenza virus infection.

In this regard, polyethyleneimine (PEI), a mucosal adjuvant, exhibits stronger mucosal adjuvanticity but induces much lower IL-17 expression than cholera toxin subunit B (CTB).³ Therefore, for the first time, we tested PEI for its potential protective effects against influenza virus infection. Ten-week-old specific-pathogen-free (SPF) female Balb/c mice were anesthetized with pentobarbitalum natricum. Then, 50 μL of PBS containing 20 μg of 25 kD linear PEI (Polysciences, Warrington, PA) or 2.5 μg of CTB (Sigma-Aldrich, St Louis, MO, USA) or PBS alone as a control was intranasally administered twice at 24 and 48 h before challenge with 5 LD₅₀ influenza virus H1N1 (A/PR/8/34). Mouse body weights were monitored every day, and those with greater than 25% loss of their initial body weight were euthanized as described.⁴

As shown in Figure 1A, the body weight of the mice in the PEI-pretreated group remained stable from day 1 to day 5 after H1N1 challenge and then gradually decreased until day 11, for a total loss of 18%, before recovering. By contrast, the body weights of the mice in the CTB- and PBS-pretreated groups decreased significantly beginning on day 2 and reached losses of more than 25% by day 8 and 10, respectively, after H1N1 challenge. The final survival rate of the mice in the PEI-pretreated group was 60%, whereas that of the mice in the CTB- and PBS-pretreated groups was 0% (Figure 1B). We then examined the viral titers in mouse lungs as previously described⁵ and found that PEI significantly reduced lung viral titers on day 2 after H1N1 challenge, whereas the viral titers in the lungs of mice in the CTB- and PBS-pretreated groups showed no significant differences (Figure 1C). Subsequently, we examined lung sections stained with hematoxylin and eosin as previously described.² On day 2 after H1N1 infection, the pulmonary alveoli were relatively intact, and only a few inflammatory cells were observed in the lung tissues of mice in the PEI-pretreated group. However, the lungs of mice in the CTB- and PBS-pretreated groups were filled with abundant inflammatory cells (Figure 1D). These results suggest that intranasal application of PEI has efficacy in protecting mice from challenge by influenza virus H1N1.

To determine the efficacy of PEI against another subtype of influenza virus, we pretreated mice with PBS containing PEI or PBS alone and challenged them with 5 LD₅₀ influenza virus H3N2 (A/Guizhou/54/89). Mice in the PEI-pretreated group were fully protected against H3N2 challenge, showing no significant body weight loss (Figure 1E) and a 100% survival rate, whereas those in the PBS group lost more than 25% of their body weight and showed a 0% survival rate on day 6 post-challenge (Figure 1F). Therefore, PEI-mediated protection against influenza virus infection is not subtype-specific.

To elucidate the mechanism of action of PEI, we examined the RNA levels of IFN-α, IFN-β, IFN-γ, GM-CSF, IFITM3 and IL-17 in the lungs of mice pretreated with PEI, CTB, and PBS, respectively, before viral challenge using quantitative reverse transcription-PCR (qRT-PCR). As previously reported, some of these cytokines, such as interferon, GM-CSF and IFITM3, were effective in protecting against influenza infection.⁶⁻⁸ As shown in Supplementary Figure S1, PEI induced a significantly higher RNA level of IFN-α than CTB or PBS. Furthermore, the RNA levels of GM-CSF and IFITM3 elicited by PEI were similar to those induced by CTB but much higher than those

1Key Lab of Medical Molecular Virology of MOE/MOH, School of Basic Medical Sciences and Shanghai Public Health Clinical Center, Fudan University, Shanghai 200032, China
2New York Blood Center, Lindsley F. Kimball Research Institute, New York, NY 10065, USA
Correspondence: L Lu; SB Jiang
E-mail: lul@fudan.edu.cn; shibojiang@fudan.edu.cn
Received 23 March 2016; revised 6 April 2016; accepted 7 April 2016
induced by PBS. The RNA level of IL-17 in mice pretreated with CTB, an IL-17-inducing adjuvant, was approximately 7- and 42-fold higher than that in mice pretreated with PEI and PBS, respectively. These results suggest that IFN-α4, GM-CSF, and IFITM3 are ‘good cytokines’ because they act as protective mediators against influenza virus infection, whereas IL-17 is a ‘bad cytokine’ that exacerbates pathology, primarily in the lung, after influenza infection.

In summary, we demonstrated that PEI, a mucosal stimulant for topical intranasal administration, is highly effective in preventing influenza virus infection. Compared to the bacteria-produced toxin CTB, the chemically synthesized polymer PEI is safer for mucosal application in humans. PEI has been tested in several clinical trials for gene delivery in vivo, demonstrating a good safety profile. Moreover, its low cost of production and abundance makes PEI more suitable for urgent and widespread use during a time of influenza epidemic or pandemic.

ACKNOWLEDGEMENTS

We thank Dr Ze Chen at the Shanghai institute of biological products in China for providing influenza virus H3N2 (A/Guizhou/54/89). This work was supported by grants from the national nature science foundation of China (81590762 to Shibo Jiang, 81373456 to Lu Lu) and Shanghai Kai Star Project (16QA1400300) to Lu Lu.

Supplementary Information for this article can be found on the Emerging Microbes and Infections website (http://www.nature.com/emi)

1. Adkins I, Holubova J, Kosova M et al. Bacteria and their toxins tamed for immunotherapy. Curr Pharm Biotechnol 2012; 13: 1446–1473.
2. Gopal R, Rangel-Moreno J, Fallert Junecko BA et al. Mucosal pre-exposure to Th17-inducing adjuvants exacerbates pathology after influenza infection. Am J Pathol 2014; 184: 55–63.
3. Wegmann F, Garthlin KH, Harandi AM et al. Polyethyleneimine is a potent mucosal adjuvant for viral glycoprotein antigens. Nat Biotechnol 2012; 30: 883–888.
4. He B, Xia S, Yu F et al. Putative suppressing effect of IgG Fc-conjugated haemagglutinin (HA) stalk of influenza virus H7N9 on the neutralizing immunogenicity of Fc-conjugated HA head: implication for rational design of HA-based influenza vaccines. J Gen Virol 2016; 97: 327–333.
5. He B, Chang H, Liu Z et al. Infection of influenza virus neuraminidase-vaccinated mice with homologous influenza virus leads to strong protection against heterologous influenza viruses. J Gen Virol 2014; 95: 2627–2637.
6. Strayer DR, Carter WA, Stouch BC et al. Protection from pulmonary tissue damage associated with infection of cynomolgus macaques by low dose natural human IFN-alpha administered to the buccal mucosa. Antiviral Res 2014; 110: 175–180.
7. Huang FF, Barnes PF, Feng Y et al. GM-CSF in the lung protects against lethal influenza infection. Am J Respir Crit Care Med 2011; 184: 259–268.
8. Everitt AR, Clare S, Pertel T et al. IFITM3 restricts the morbidity and mortality associated with influenza. Nature 2012; 484: 519–523.
9. Neuberg P, Kühler A. Recent developments in nucleic acid delivery with polyethyleneimines. Adv Genet 2014; 88: 263–288.