COMMUNICATION

Ratiometric sensing of fluoride ions using Raman spectroscopy

William J. Tipping, a Liam T. Wilson, b Sonja K. Blaseio, a, 1 Nicholas C. O. Tomkinson, a, b Karen Faulds a, 2 and Duncan Graham a, 3

Received 00th January 20xx, Accepted 00th January 20xx

DOI: 10.1039/00xx00000x

Ratiometric Raman spectroscopy represents a novel sensing approach for the detection of fluoride anions based on alkyne desilylation chemistry. This method enables rapid, anion selective and highly sensitive detection of fluoride in a simple paper-based assay format using a portable Raman spectrometer.

Raman microscopy is a powerful method for chemical analysis and biological imaging because it provides a spectral fingerprint with chemical specificity in a non-destructive manner. 1 An emerging approach for chemical sensing is alkyne-tag Raman imaging (ATRI) which has enabled in situ biomolecular detection with high sensitivity, and multiplex analysis using low molecular weight Raman labels and sensor moieties. 2 Alkynes have discrete vibrational modes within the cellular-silent region of the Raman spectrum (1800-2800 cm⁻¹) which greatly facilitates their detection in biological samples. 3 Early examples of ATRI reported metabolic labelling of intracellular biomolecules with alkyne-labelled precursors for the detection of nascent DNA, 4 glycans 5 and proteins 6 amongst others. Since then, functionalisation of small molecules with alkyne labels has enabled the intracellular visualisation of drugs, 7 natural products 8, 9 and lipids 10 using Raman and stimulated Raman scattering (SRS) microscopy.

An unexplored application of ATRI is the development of small molecule sensors for the detection and quantification of endogenous chemicals, ions and biomolecules. Two isolated examples make use of oligoyne scaffolds for intracellular hydrogen sulphide sensing 11 and for pH quantification 12 (Figure 1). In each of these reports, the ratiometric Raman sensor relies on analyte-induced changes of the sensing group resulting in a change in the Raman shift and/or scattering properties of the alkyne moiety. As such, ratiometric sensing approaches are advantageous because there is an effective internal referencing that increases sensitivity and improves quantification. 13, 14

Fluoride is a biologically important anion that is readily absorbed by the body but is excreted slowly, and as such, overexposure to fluoride may result in gastric and kidney problems, bone diseases and fluorosis. 15 There has been much interest in the development of fluorescence-based sensors for fluoride detection (Figure S1). 16 notably, using the fluoride-mediated desilylation of Si-O groups 17-19 and Si-C (alkyne) groups 20-26 as prominent examples for military and cellular applications.

Despite 20 years of developing fluorescent fluoride sensors based on Si-F chemistry, significant challenges have been identified. 27

![Figure 1](https://example.com/figure1.png)

Figure 1 Previous reports of sensing and quantification using alkyne-tag Raman imaging. (i) A previous example of an irreversible sensor for hydrogen sulfide (H₂S) detection 11 and (ii) a reversible sensor for quantifying intracellular pH. 12 This work reports an extremely low molecular weight Raman sensor (1) for ratiometric detection of fluoride ions.

---

a Centre for Molecular Nanometrology, WestCHEM, Department of Pure and Applied Chemistry, Technology and Innovation Centre, University of Strathclyde, 99 George Street, Glasgow G1 1RD, UK. Email: karen.faulds@strath.ac.uk; duncan.graham@strath.ac.uk
b Department of Pure and Applied Chemistry, WestCHEM, University of Strathclyde, 295 Cathedral Street, Glasgow G1 1XJ, UK. Email: nicholas.tomkinson@strath.ac.uk
cPresent address: Technische Universität Braunschweig, Institut für Technische Chemie, Franz-Liszts-Str. 35a, 38106 Braunschweig, Germany.

Electronic Supplementary Information (ESI) available: Figures S1 – S10. See DOI: 10.1039/x0xx00000x
COMMUNICATION

First, the structural complexity of fluorescent sensors increases the synthetic effort and expense required to prepare the sensors, restricting translation to low-cost, in-field applications. Second, the use of fluorophores that are not compatible with NIR excitation, which is essential for biological compatibility due to reduced photodamage and enhanced imaging depth penetration. Third, the requirement of a surfactant (e.g., cetyltrimethylammonium bromide, CTAB) to enable detection in organic-aqueous solvent mixtures, once again restricting translation.28 Here we propose the use of Raman spectroscopy as a novel method to address these limitations; an extremely small ratiometric sensor molecule (<200 Da) is used to detect fluoride with NIR excitation in aqueous mixtures without the use of a solubilising agent (Figure 1). Our method enables rapid monitoring of fluoride ions in solution and in a test-paper assay.

To test the validity of a Raman-based approach to detect fluoride ions, we chose to study the fluoride-mediated desilylation of alkyne groups as a model reaction.29 Raman spectroscopy is well-suited to temporal monitoring of chemical reactions in a non-destructive manner,30 in particular, for reactions of alkyne.31 The cleavage of Si—C alkynyl bonds by fluoride is rapid and irreversible owing to the significant differences in the bond dissociation energies (Si—C ~290 kJ/mol; Si—F ~590 kJ/mol).15 We designed 4-[(trimethylsilyl)ethyl]benzonitrile 1 as an extremely simple molecular Raman sensor, which incorporated two bioorthogonal Raman groups and was readily synthesised in high yield (95%) in a single step from commercially available starting materials (see Supplementary Information for details). The reaction of 1 (50 mM) with tetra-n-butylammonium fluoride (TBAF, 1 equiv.) in THF resulted in complete desilylation of the alkyne group within 1 minute (thus forming compound 2), and resulted in a red-shifting of the alkyne stretching frequency from 2162 cm⁻¹ to 2109 cm⁻¹ (Figure 2). This observation is consistent with the Raman spectra of the neat compounds (Figure S2).

We proposed that, based on the 53 cm⁻¹ shift in the alkyne frequency, ratiometric sensing could be achieved to confirm the presence of fluoride ions in solution. Furthermore, upon the removal of the silyl group, there was a marginal shift in the nitrile peak (2231 cm⁻¹ → 2230 cm⁻¹) which offers a second motif for ratiometric sensing. In addition, the relative intensity of the phenyl C=C peak (~1600 cm⁻¹) decreased upon desilylation, which is consistent with the relative peak intensities in the Raman spectra of the solid samples. To demonstrate the versatility of this approach, we showed that desilylation of sensor 1 was also successful using inorganic fluoride sources, including NaF and CsF in a THF:water (1:1 v/v) mixture (Figure S3).

A series of control reactions were followed by Raman spectroscopy and validated using UV-visible spectrometry. Figure S4A demonstrates the control reaction of sensor 1 in the absence of fluoride which showed no change in the Raman spectra over 30 min. In addition, no reaction was observed upon addition of a range of different TBA⁺ and Na⁺ counter anions (Figure S4B–D respectively). The sensor is, however, unstable at high pH (> pH 10, Figure S4E) and in the presence of NaI, which is consistent with observations of previous sensors of this type.20 Figure S5A shows that the detection limit of sensor 1 in solution using Raman spectroscopy is ~250 μM, enabling sensitive detection of fluoride in solution (Figure S5B), whilst Figure S5C shows the reaction proceeds in THF: phosphate buffer (1:1 v/v) mixture. UV-visible spectroscopy showed a reduction of the λmax at ~280 nm upon addition of fluoride, which is in close agreement with related trimethylsilyl-protected alkyne sensors (Figure S6).20 Together, these data indicated the potential for extremely rapid and sensitive detection of fluoride in organic solution using Raman spectroscopy.

It is desirable to detect fluoride in aqueous solutions for use in analysis of groundwater and drinking water samples, although the majority of fluorescent sensors have only been examined in neat organic medium.28 Water is only weakly Raman active, and therefore, generates minimal interference in Raman analysis.1 Having established the detection of fluoride anions by 1 in organic solvents, we examined our system in aqueous mixtures. We repeated the reaction, monitoring experiments with sensor 1 (50 mM) using an increased [TBAF] (75 mM, 1.5 equiv.) to compensate for the high enthalpy of hydration of fluoride in water (ΔH = -504 kJ/mol).33 Upon addition of excess fluoride to the reaction mixtures, Raman spectra were recorded every 0.5 s for rapid detection. The peak intensities of the starting material (2162 cm⁻¹), the desilylated product (2109 cm⁻¹) and the nitrile band (2230-2231 cm⁻¹) were
plotted as a function of time as the water content of the mixture varied from 0–25% (v/v). Figure 3A shows a representative example of a reaction conducted in 25% water in THF and indicates the desilylation reaction was extremely rapid, with the formation of desilylated product over the course of the experiment (~90% completion at 180 s). Similar plots for the other conditions are provided in Figure S7. Interestingly, the peak area for the nitrile band at ~2231 cm⁻¹ also varied throughout the reaction time course, confirming that the desilylation of the alkynyl group significantly influenced the electronic properties of the benzonitrile group. Figure 3B presents the integrated peak areas for the desilylated product at 2109 cm⁻¹ as a function of time when the reaction was performed using different initial %water. As the water content of the reaction mixture increased, the rate of product formation decreased. This was likely due to the hydration enthalpy of fluoride in water.33 In spite of this, these results indicate an extremely fast response time (<20 s) for fluoride detection in aqueous-organic mixtures which compares favourably to most fluorescent sensors.

We next sought to assess the application of Raman sensor 1 on different materials to demonstrate the versatility of the approach. Recently, the use of paper-based substrates for fluoride sensing with a fluorescent probe has been reported.44 To demonstrate the application of Raman spectroscopy in this area, we pre-treated filter paper test strips with sensor 1 (10 μL of 100 mM in THF) which were air-dried for 5 minutes before a 300 μL solution of TBAF in THF (0.125-5 mM) was applied directly onto the paper. Raman maps (20 μm × 20 μm) were acquired using identical conditions to Figure 3A. Peak annotations are in cm⁻¹.

Figure 4 Paper-based detection of fluoride using Raman sensor 1. A Filter paper was pre-treated with sensor 1 (100 mM in THF, 10 μL) before air-drying and subsequent direct addition of TBAF in THF (300 μL) at the indicated concentrations. Raman maps were acquired across 20 μm × 20 μm (1 μm pixel size; 400 spectra) using 785 nm laser excitation with a 20x objective lens (~180 mW) for 0.5 s. The maps represent the ratio 2105 cm⁻¹/2160 cm⁻¹. For the blank sample, 10 mM TBAF replaced TBAF. B Average Raman spectra from maps presented in A. Peak annotations are in cm⁻¹.

We next sought to assess the application of Raman sensor 1 on different materials to demonstrate the versatility of the approach. Recently, the use of paper-based substrates for fluoride sensing with a fluorescent probe has been reported.44 To demonstrate the application of Raman spectroscopy in this area, we pre-treated filter paper test strips with sensor 1 (10 μL of 100 mM in THF) which were air-dried for 5 minutes before a 300 μL solution of TBAF in THF (0.125-5 mM) was applied directly onto the paper. Raman maps (20 μm × 20 μm; 400 individual spectra) were acquired across the sample test strip after drying (~2 minutes). Representative Raman maps and average Raman spectra from the maps are presented in Figure 4. As the fluoride concentration increased, so too the ratio of 2105/2160 cm⁻¹ (alkyne/alkyne 1) increased, and the ratio 2231/2237 cm⁻¹ (nitrile 2/nitrile 1) arising from the benzonitrile group also increased (ratio images provided in Figure S8 and additional repeat in Figure S9). Interestingly, the Raman shift of the alkyne and nitrile group of the desilylated product 2 was slightly shifted when analysed on paper (2160 and 2237 cm⁻¹) compared to solution phase analysis (2162 and 2231 cm⁻¹), potentially due to differences in local intramolecular bonding in solution compared to solid phase. Using this simple ratiometric approach, fluoride detection at concentrations as low as 125 μM could be achieved (Figure 4 and Figure S8) which compares favourably to a recently reported colorimetric sensor (5.75 mg/L, ~300 μM in 3.7 v/v EtOH:water)35 and a coordination-based sensor (300 μM in aqueous buffer).36 In addition, we found that our low-cost paper-based sensing system was compatible with handheld, portable Raman detection (Figure S10), which demonstrates a potential translation of this method for in-field fluoride analysis. Together, these results highlight ratiometric Raman spectroscopy as a novel detection method for fluoride ions in solution using a simple paper-based assay format.
Conclusions

Compound 1 represents an extremely low molecular weight chemical sensor for fluoride detection which works using NIR excitation for ratiometric Raman spectroscopy. Advantages of NIR excitation are biological compatibility and operator safety, particularly given that many fluorescent sensors require cell-damaging UV excitation. The narrow resonances of Raman peaks (~40 cm⁻¹) enables clear, unambiguous and fully resolved detection of the starting material and product in a single Raman spectral acquisition. This represents a significant advantage to the “turn-on” fluorescent probes which are currently reported, as typically only the desilylated product generates a fluorescent signal. Furthermore, the narrow resonances of Raman shifts are advantageous over ratiometric fluorescent sensors, which typically have broad emission profiles (~1500 cm⁻¹), imposing a limit on the multiplexing capability of the sensing system.

The detection of the alkyne sensor in solution and on test paper strips highlights the versatility of Raman spectroscopy for small-molecule sensor detection. Our results indicate that fluoride concentrations as low as 125 μM generate a ratiometric output in our simple paper-based assay. This is below the level at which biological toxicity was observed in cells (extracellular [NaF] = 3 mM) and is within the maximum contaminant level defined by the U.S. Environmental Protection Agency (4.0 mg L⁻¹, 211 μM). We anticipate that the limit of detection of our sensing method could be improved by increasing the Raman scattering intensity of the alkyne sensor through conjugation to form a poly-yne, whilst different silicon capping groups could be investigated as alternative sensing motifs for improved pH stability in alkaline solutions.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank the University of Strathclyde, the EPSRC and GlaxoSmithKline for financial support.

Notes and references

The research data associated with this paper will become available from the University of Strathclyde at the following link: https://doi.org/10.15129/46582865.502a-43be-5977-bd-41032be1e1.

1. W. J. Tipping, M. Lee, A. Serrels, V. G. Brunton and A. N. Hulme, Chem. Soc. Rev., 2016, 45, 2075-2089.
2. H. Yamakoshi, K. Dodo, A. Palonpon, J. Ando, K. Fujita, S. Kawata and M. Sodeoka, J. Am. Chem. Soc., 2012, 134, 20681-20689.
3. A. F. Palonpon, M. Sodeoka and K. Fujita, Curr. Opin. Chem. Biol., 2013, 17, 708-715.
4. H. Yamakoshi, K. Dodo, M. Okada, J. Ando, A. Palonpon, K. Fujita, S. Kawata and M. Sodeoka, J. Am. Chem. Soc., 2011, 133, 6102-6105.
5. S. Hong, T. Chen, Y. Zhu, A. Li, Y. Huang and X. Chen, Angew. Chem., Int. Ed., 2014, 53, 5827-5831.
6. L. Wei, F. Hu, Y. Shen, Z. Chen, Y. Yu, C.-C. Lin, M. C. Wang and W. Min, Nat. Methods, 2014, 11, 410-412.
7. M. M. Gaschler, F. Hu, H. Feng, A. Linkermann, W. Min and B. R. Stockwell, ACS Chem. Biol., 2018, 13, 1013-1020.
8. W. J. Tipping, M. Lee, A. Serrels, V. G. Brunton and A. N. Hulme, Chem. Sci., 2017, 8, 5606-5615.
9. J. Seidel, Y. Miao, W. Porterfield, W. Cai, X. Zhu, S.-J. Kim, F. Hu, S. Bhattacharai-Kline, W. Min and W. Zhang, Chem. Commun., 2019, 55, 9379-9382.
10. E. L. Jamieson, J. Greaves, J. A. McLellan, K. R. Munro, N. C. O. Tomkinson, L. H. Chamberlain, K. Faulds and D. Graham, Spectrochim. Acta A, 2018, 197, 30-36.
11. C. Zeng, F. Hu, R. Long and W. Min, Analyst, 2018, 143, 4844-4848.
12. T. L. Wilson, W. J. Tipping, L. E. Jamieson, C. Wetherill, Z. Hanley, K. Faulds, D. Graham, S. P. Mackay and N. C. O. Tomkinson, Analyst, 2020, 145, 5289-5298.
13. X. Huang, J. Song, B. C. Yung, X. Huang, Y. Xiong and X. Chen, Chem. Soc. Rev., 2018, 47, 2873-2920.
14. L. E. Jamieson, C. Wetherill, K. Faulds and D. Graham, Chem. Sci., 2018, 9, 6935-6943.
15. Y. Zhou, J. F. Zhang and J. Yoon, Chem. Rev., 2014, 114, 5511-5571.
16. Y. Jiao, B. Zhu, J. Chen and X. Duan, Theranostics, 2015, 5, 173-187.
17. S. Y. Kim, J. Park, M. Koh, S. B. Park and J.-I. Hong, Chem. Commun., 2009, 4735-4737.
18. D. Kim, S. Singh, T. Wang, E. Seo, J. H. Lee, S.-J. Lee, K. H. Kim and K. H. Ahn, Chem. Commun., 2012, 48, 10243-10245.
19. A. Roy, D. Kand, T. Saha and P. Talukdar, Chem. Commun., 2014, 50, 5510-5513.
20. M. Jo, J. Lim and O. Š. Miljanić, Org. Lett., 2013, 15, 3518-3521.
21. L. Fu, F.-L. Jiang, D. Fortin, P. D. Harvey and Y. Liu, Chem. Commun., 2011, 47, 5503-5505.
22. H. Lu, Q. Wang, Z. Li, G. Lai, J. Jiang and Z. Shen, Org. Biomol. Chem., 2011, 9, 4558-4562.
23. D. Buckland, S. V. Bhosale and S. J. Langford, Tetrahedron Lett., 2011, 52, 1990-1992.
24. M. R. Rao, S. M. Mobin and M. Ravikanth, Tetrahedron, 2010, 66, 1728-1734.
25. L. Fu, F.-F. Tian, L. Lai, Y. Liu, P. D. Harvey and F.-L. Jiang, Sens. Actuators B Chem., 2014, 193, 701-707.
26. M. Kaur, M. J. Cho and D. H. Choi, Dyes Pigm., 2014, 103, 154-160.
27. P. Chen, W. Bai and Y. Bao, J. Mater. Chem. C, 2019, 7, 11731-11746.
28. L. Gai, J. Mack, H. Lu, T. Nyokong, Z. Li, N. Kobayashi and Z. Shen, Coord. Chem. Rev., 2015, 285, 24-51.
29. G. L. Larson, Synthesis (Stuttgart), 2018, 50, 2433-2462.
30. N. E. Leadbeater and J. R. Schmink, Nat. Protoc., 2007, 3, 1.
31. W. J. Tipping, M. Lee, V. G. Brunton, G. C. Lloyd-Jones and A. N. Hulme, Faraday Discuss., 2019, 220, 71-85.
32. H. F. Shurvell and M. C. Southby, Vib. Spectros., 1997, 55, 137-146.
33. L. Trembleau, T. A. D. Smith and M. H. Abdelrahman, Chem. Commun., 2013, 49, 5850-5852.
34. X. Wu, H. Wang, S. Yang, H. Tian, Y. Liu and B. Sun, ACS Omega, 2019, 4, 4918-4926.
35. L. C. Murfin, K. Chiang, G. T. Williams, C. L. Lyall, A. T. A. Jenkins, J. Wenk, T. D. James and S. E. Lewis, Front. Chem., 2020, 8, 10.
36. W. Tipping, R. Yuste, V. W. Cornish and W. Min, Nature, 2017, 544, 465.
37. H. Matsui, M. Morimoto, K. Horimoto and Y. Nishimura, Toxicol. in Vitro, 2007, 21, 1113-1120.

Page 4 of 5
A simple ratiometric sensor based on Raman spectroscopy enables rapid fluoride ion detection in a paper-based assay using a portable spectrometer.