Abstract

Root and stem wood anatomy of *C. myrianthum* (Verbenaceae) from a semideciduous seasonal forest in Botucatu municipality (22°52'20"S and 48°26'37"W), São Paulo state, Brazil, were studied. Growth increments demarcated by semi-ring porosity and marginal bands of axial parenchyma were observed in the wood of both root and stem. Many qualitative features were the same in both root and stem: fine helical thickenings, and simple and multiple perforation plates in vessel elements; large quantities of axial parenchyma in the growth rings, grading from marginal bands and confluent forming irregular bands in earlywood to lozenge aliform in latewood; axial parenchyma cells forked, and varied wall projections and undulations; separte fibres; forked and diverse fibre endings. Quantitative features differing between root and stem wood were evaluated using student’s t-test, and vessel frequency, vessel element length, vessel diameter, ray height, and vulnerability and mesomorphy indices differed significantly. Root wood had lower frequency of vessels, narrower and longer vessel elements, and taller rays than wood of the stem. The calculated vulnerability and mesomorphy indices indicated that *C. myrianthum* plants are mesomorphic. Roots seem to be more susceptible to water stress than the stem.

Key words: secondary xylem, semideciduous seasonal forest, wood anatomy.

Introduction

*Citharexylum myrianthum* Cham., a deciduous tree, reaches 8 to 15 m in height and 20 to 40 cm in breast height diameter, and has a wide occurrence in Brazilian forest formations (latitude 14°45’–31°50’S). It occurs in Dense Ombrophilous Forest (Atlantic Rain Forest *sensu stricto*), semideciduous seasonal forest, riparian areas (Carvalho
and also in lowland dry tropical forest (Tabarelli 1992). The rapid growth makes *C. myrianthum* an important species for restoration of degraded areas (Sansevero *et al.* 2009).

General descriptions of wood anatomy of the genus *Citharexylum* have already been reported by Metcalfe and Chalk (1950). For *C. myrianthum*, Gomes *et al.* (1989) studied stem wood anatomy from trees that occurred in eastern part of Paraná state (South Brazil) and it was the first time that radiate perforation plates in vessels were mentioned in this species. Barros *et al.* (2001) described the wood anatomy of trees from a seasonally flooded forest of the Reserva Biológica de Poço das Antas (Rio de Janeiro state, Southern Brazil). Cambial activity and the seasonal formation of secondary xylem in stems of *C. myrianthum* occurring at São Paulo State were studied by Marcati (2000) and annual growth rings were observed in the wood. Seasonal presence of acicular calcium oxalate crystals in the cambial zone, with greater abundance in dry than in wet periods, was reported for this species (Marcati 2000; Marcati & Angyalossy 2005). Across this rich body of work, only stem wood was studied.

Perhaps not surprisingly, given the comparative difficulty of collection specimens, root wood anatomy, in general, has received much less attention than stem wood anatomy (e.g. Gasson & Cutler 1990). When studied, physiological and anatomical research demonstrated that within a species, root xylem has wider vessels than stem xylem and that root wood is more vulnerable to embolism than stem wood (Alder *et al.* 1996; Ewers *et al.* 1997; Machado *et al.* 1997; Kavanagh *et al.* 1999; Kolb & Sperry 1999; McElrone *et al.* 2004; Psaras & Sofroniou 2004).

The objectives of this study were to compare root and stem wood anatomy of *Citharexylum myrianthum* qualitatively and quantitatively and to relate any anatomical differences between the organs to their function and ecological adaptations.

**Material and Methods**

*Citharexylum myrianthum* were collected in a semi-deciduous seasonal forest in the municipality of Botucatu (22°52′20″S, 48°26′37″W), São Paulo state, south-eastern Brazil. The average annual rainfall is about 1300 mm, with a mean annual temperature of 20°C. July is the driest and coldest month with mean temperature of 15°C, and January is the wettest and warmest month with mean temperature of 25°C. The dry season typically extends from May to September (Fig. 1). Climate data were obtained from the Estação Meteorológica of the Faculdade de Ciências Agrônomicas, UNESP, Botucatu, São Paulo State. The Estação Meteorológica was about 11 km from the study site.

Stem and main root samples from three adult specimens (Tab. 1) were collected at 1.30 m and 30 cm distal from root collar, respectively. Wood samples from sapwood were fixed in 70% ethanol. Transverse, radial and tangential sections (10–18 μm) were cut using a sliding microtome. These sections were double stained with 1% aqueous solution of fuchsin and astra blue (Roeser 1972) and mounted on slides in Entelan® synthetic medium. Some root and stem materials were macerated in a mixture of equal volumes of acetic acid and hydrogen peroxide at 60°C (Johansen 1940) for 12 to 24 hours. The material was stained with 1% aqueous solution of safranin and mounted in glycerin. The presence of calcium oxalate was confirmed, since dilutions with hydrochloric acid (HCl) produced no effect (Chamberlain 1932).

![Figure 1](image-url) – Climate diagram following Walter (1986), constructed from data obtained at meteorological station located at Faculdade de Ciências Agrônomicas (FCA), UNESP, Botucatu, from 1995 to 2012, showing one well demarcated dry season per year.
Wood descriptions follow the microscopic features definitions of IAWA Committee (1989).

The vouchers and samples of root and stem wood were deposited, respectively, in the Herbarium (BOT) and in the Wood Collection (BOTw) (Tab. 1) of the Departamento de Recursos Naturais, Faculdade de Ciências Agronômicas, UNESP, São Paulo.

Quantitative data were based on 30 individual counts per specimen; the statistical requirements or minimum numbers of measurements were fulfilled: \( n = \left( \frac{t \text{ value}}{\text{sample variance}} \right)^2 \times \frac{10\% \times \text{sample mean}}{\text{accuracy of 10\% \times sample mean}}, \) following Freese (1967) and Eckblad (1991). The numerical values given in Table 2 are the means accompanied by standard deviation. Wood anatomical parameters were analysed statistically using the Student’s t-test and a threshold of \( p \leq 0.05 \) for significance (Zar 1996).

**Results**

Qualitative anatomical features of *Citharexylum myrianthum* did not vary between stem and root wood; differences between the organs were found only in quantitative features (Tab. 2).

Growth rings – distinct, delimited by marginal bands, semi-ring-porous wood (Fig. 2a-b), and slightly distended rays in earlywood, and radially flattened fibers with thick walls in the latewood. In addition, the axial parenchyma pattern varied through the growth ring, with marginal bands and confluent irregular bands in the earlywood, and confluent and lozenge-aliform in latewood (Fig. 2a-b).

Vessels: predominantly solitary (70–92%), in multiples of 2 (7–24%) and of 3–6 (0.8–9%); fine helical thickenings throughout the body of vessel element in both earlywood and latewood.

### Table 1 – Collection data from the *Citharexylum myrianthum* Cham. studied.

| Wood collection number | DBH (cm) | Height (m) | Root diameter (cm) |
|------------------------|---------|------------|-------------------|
| BOT, 1369              | 45      | 17         | 12,0              |
| BOT, 1370              | 29      | 18         | 6,0               |
| BOT, 1371              | 59      | 17         | 8,5               |

* DBH: diameter at breast height (1.3 m).  
** Root diameter: 30 cm from the base.
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**Table 2** – Mean and standard deviation of anatomical characteristics of *Citharexylum myrianthum* Cham. wood, by specimens. O = organs; S = stem; R = root. L = length in µm, D = diameter in µm, F = frequency (vessels/mm²) (rays/linear mm), I = index; v = vulnerability; m = mesomorphy. WT = wall thickness of fibres in µm. W = ray width in cell numbers, H = rays height in µm, IV = intervessel pit diameter in µm, RV = ray-vessel pit diameter in µm.

| Specimens | BOT~w~1369 | BOT~w~1370 | BOT~w~1371 |
|-----------|-------------|-------------|-------------|
| O | S | R | S | R | S | R |
| L | 268.1 ± 42.7 | 306.0 ± 46.0 | 283.4 ± 35.0 | 302.4 ± 39.3 | 267.0 ± 44.1 | 315.5 ± 30.5 |
| Vessels | D | 148.6 ± 27.2 | 127.1 ± 31.4 | 144.8 ± 47.7 | 133.0 ± 43.4 | 162.7 ± 38.8 | 141.1 ± 40.7 |
| F | 13.1 ± 2.5 | 7.9 ± 2.1 | 12.0 ± 2.5 | 9.4 ± 3.7 | 11.0 ± 3.2 | 8.1 ± 2.7 |
| I | v | 11.7 | 17.0 | 12.8 | 14.9 | 14.6 | 18.3 |
| m | 3067.1 | 5146.0 | 3500.4 | 4343.4 | 3643.4 | 5613.6 |
| L | 1055.1 ± 299.0 | 1121.6 ± 206.9 | 1060.7 ± 270.2 | 1127.9 ± 204.3 | 1066.7 ± 270.0 | 1111.0 ± 238.0 |
| Fibres | D | 28.8 ± 4.4 | 31.5 ± 2.8 | 28.6 ± 4.3 | 29.2 ± 4.6 | 28.7 ± 4.7 | 28.7 ± 3.8 |
| WT | 6.1 ± 1.0 | 6.0 ± 0.9 | 6.2 ± 1.1 | 6.2 ± 1.2 | 6.1 ± 1.2 | 6.2 ± 1.1 |
| W | 3.7 ± 0.5 | 3.8 ± 0.6 | 3.8 ± 0.7 | 3.7 ± 0.6 | 3.9 ± 0.6 | 3.9 ± 0.6 |
| Rays | H | 282.7 ± 86.0 | 327.1 ± 145.2 | 315.7 ± 120.5 | 367.9 ± 116.8 | 295.6 ± 97.0 | 341.2 ± 160.1 |
| F | 4.4 ± 0.9 | 4.5 ± 0.7 | 4.3 ± 0.9 | 4.5 ± 0.9 | 4.4 ± 1.0 | 4.5 ± 0.8 |
| Pits | IV | 3.6 ± 0.7 | 4.0 ± 0.5 | 3.8 ± 0.8 | 3.8 ± 0.7 | 3.8 ± 0.9 | 3.8 ± 0.5 |
| RV | 3.8 ± 0.6 | 3.9 ± 0.7 | 3.9 ± 0.9 | 3.9 ± 0.6 | 4.0 ± 0.5 | 3.9 ± 0.7 |

(Fig. 3a); vessel element tails of different shapes in one or both tips; simple perforation plates (Fig. 3b), radiate (Fig. 3d), and foraminite-reticulate perforation plates (Fig. 3c,e), transverse (Fig. 3c-e) or inclined (Fig. 3a); intervessel pits alternate, circular (Fig. 3a,b); vessel-ray pits similar to intervessel pits in shape and in size. Rays: predominantly multiseriate (Fig. 3f), heterocellular, body ray cells procumbent with one row of upright and/or square marginal cells (Fig. 3g). Axial parenchyma: paratracheal aliform, confluent forming irregular bands, and initial marginal bands (Fig. 2a,b); variable from 3–4 cells per strand mostly, occasionally more than 6 cells per strand; cells forked (Fig. 3i), with projections (Fig. 3j), and undulations (Fig. 3h). Fibres: thin-to thick-walled (Fig. 2a,b); simple to minutely bordered small pits only in radial walls; septate fibres (Fig. 3k) and nonseptate fibres present; fibres with wall interruptions (Fig. 3l); forked and with a wide range of tip shapes (Fig. 3m-o). Mineral inclusions: acicular calcium oxalate crystals in ray parenchyma cells (procumbent, upright and/or square cells).

Statistical analysis – The mean data by organ and the results of the student’s t-test are summarized in Tab. 3 and the mean differences between organs are graphically summarized in Fig. 4. Vessel element parameters including length, tangential diameter, and frequency, as
well as ray height were significantly different between root and stem. Root wood had lower vessel frequency, longer and narrower vessel elements when compared to stem wood. Root wood had taller rays than stem wood. Vulnerability and mesomorphy indices are significantly different between the two organs (Tab. 3 and Fig. 4).
a greater appreciation for the variability of the soil environment and its influence on plant development can be gained. The trees studied here grew in a semi-deciduous seasonal forest with a dry season of about 5 months. During the dry season, soil water availability is low (Borchert 1994) and thus the roots experience at least one form of distinct seasonality presumably sufficient to signal distinct growth increment formation in the roots.

Semi-ring porous wood and marginal bands of axial parenchyma delimiting growth rings have already been described by Metcalfe & Chalk (1950) in *Citharexylum*, and by Gomes et al. (1989) in *C. myrianthum* specifically, but only for stem wood. Metcalfe & Chalk (1950) stated that semi-ring porous wood is a common feature to Verbenaceae and, according to Carlquist (2001), this feature would provide functional advantage to the species as wide earlywood vessels would accommodate larger volumes of water in wet seasons while narrow latewood vessels maximize the resistance to cavitation during the dry season. If this rationale holds true, the presence of semi-ring porosity in the roots suggests that the seasonality of soil water availability may strongly influence root wood vessel diameter.

**Table 3** – Student’s t-test between root and stem measurements. * significant at p < 0.05. ± standard deviation. L = length in µm, D = diameter in µm, F = frequency (vessels/mm²) (rays/linear mm), I = index; v = vulnerability; m = mesomorphy. WT = wall thickness of fibres in µm. W = ray width in cell numbers, H = rays height in µm, IV = intervessel pit diameter in µm, RV = ray-vessel pit diameter in µm.

| Mean stem | Mean root | F-ratio |
|-----------|-----------|---------|
| Vessels   |           |         |
| L*        | 272.87 ± 41.06* | 307.96 ± 39.10* | 1.10* |
| D*        | 152.05 ± 39.18* | 133.73 ± 39.18* | 1.01* |
| F*        | 12.06 ± 2.87*  | 8.26 ± 2.96*   | 1.06* |
| I         |            |         |
| v*        | 13.47±5.30*  | 18.15±9.23*   | 3.03* |
| m*        | 3671.13±1591.13* | 5601.17±2996.87* | 1.15* |
| L         | 1061.11 ± 277.11 | 1120.18 ± 214.63 | 1.66 |
| Fibres    |           |         |
| D         | 28.75 ± 4.44  | 29.83 ± 3.97  | 1.25 |
| WT        | 6.153 ± 1.1129 | 6.18 ± 1.06   | 1.10 |
| W         | 3.79±0.63     | 3.78±0.58     | 1.18 |
| Rays      |           |         |
| H*        | 298.01±101.96* | 345.42±141.31* | 1.92* |
| F         | 4.37±0.83     | 4.48±0.81     | 1.31 |
| Pits      |           |         |
| IV        | 3.77±0.80     | 3.85±0.61     | 1.74 |
| RV        | 3.90±0.70     | 3.88±0.66     | 1.13 |

**Discussion**

Our observations of the stem wood of *Citharexylum myrianthum* are in concordance, in general, with previous descriptions of the genus (Metcalfe & Chalk 1950; Détienne & Jacquet 1983) and of the species (Gomes et al. 1989; Marcati 2000; Barros et al. 2001). The inclusion of root wood anatomy in this study broadens the literature on *C. myrianthum* and allows for within-plant organographic comparisons.

The presence of growth rings in root wood has been demonstrated for a variety of different species in diverse habitats, such as in *Styrox* (Styracaceae) (Machado et al. 1997), in *Clusia criuva* (Clusiaceae) (Esemann-Quadros 2001), and in *Lippia salviifolia* (Verbenaceae) (Goulart & Marcati 2008). These findings provide counterexamples to the assertions of Ledebenko (1962) and Brown (1971), who suggested that root wood is less likely to form growth rings because of the comparative uniformity of the environmental conditions in the soil. Esemann-Quadros (2001), Machado et al. (1997) and Goulart & Marcati (2008) suggested that the seasonal soil water availability might be an important factor affecting the formation of growth rings in the wood of roots and stems. We agree with these authors and further suggest that by tracking such soil parameters,
Figure 4 – Box & Whisker plots of the significant parameters that differ between root and stem according to the Student’s t-test (p = 0.05). □ = mean, □ = mean ± standard error, ▶ ▼ = mean ± 1.96 standard error.
common and were described in Vitex sp. (Metcalfe & Chalk 1950), Citharexylum myrianthum (Gomes et al. 1989; Marcati 2000; Barros et al. 2001), Gmelina arborea (Ohtani et al. 1989), and Lippia salviifolia (Goulart & Marcati 2008). According to Gomes et al. (1989), there are different multiple perforation plates in C. myrianthum wood, including radiate and also derivatives from foraminate and reticulate types, and that agrees with what was found in this study. Multiple perforation plates in wood may function in preventing the spread of air embolisms by retaining air bubbles (Wheeler & Baas 1991). With such a functional interpretation, multiple perforation plates and semi-ring porous wood would be important contributors to the physiology of this species and its growth in a seasonally dry environment.

Fine helical thickenings in vessel walls were also observed by Gomes et al. (1989) and Barros et al. (2001) to Citharexylum myrianthum. Carlquist (2001) pointed out that the helical thickenings on the vessel walls would increase the contact area between the vessel wall and water column, and might increase the mechanical resistance of the vessel walls. We observed no change in distribution of helical thickenings from earlywood to latewood in C. myrianthum in stem or root, suggesting that an important physiological role related to seasonal water stress is unlikely in this species.

The variation in the axial parenchyma through the increment zone, grading from marginal bands and confluent irregular bands in the earlywood to lozenge aliform in the latewood, and in some places rays slightly distended when crossing the marginal bands of axial parenchyma were described for the first time to this species. Marcati et al. (2006) reported distended rays delimiting growth rings in addition to other markers to some species from cerrado (Piptocarpa rotundifolia (Asteraceae), Annona coriacea and A. crassiflora (Annonaceae)). The distended rays delimiting growth rings might be advantageous in addition to the other growth markers helping the dendrochronologists in the correct distinction of the growth rings limits.

Septate fibres are common in Verbenaceae species, and they were observed in the root and stem wood of Citharexylum myrianthum. As they are living fibres, they could play a role in water and starch storage (Carlquist 2001, 2012), and this might be important in the seasonal environments where the species are growing.

Typically, root vessels are wider than stem vessels (Alder et al. 1996; Ewers et al. 1997; Machado et al. 1997; Kavanagh et al. 1999; Kolb & Sperry 1999; McElrone et al. 2004; Psaras & Sofroniou 2004). The opposite was observed in this study in Citharexylum myrianthum, Styrax forest species (Machado et al. 2007) and Leguminosae llianis (Ewers et al. 1997). Christensen-Dalsgaard et al. (2007) associated the smallest vessels and the lowest vessel frequency to parts of the root and stem subjected to the greatest mechanical stresses or strains, observing a trade-off between hydraulic conductivity and wood stiffness. The semi-ring porosity of the root wood combined with larger amounts of axial parenchyma surrounding the vessels may provide functional advantages to the species during the dry season. The axial parenchyma surrounding the narrower vessels may play a role in regulating the osmotic potential of the sap in the dry season, permitting the plants to take up water or refill embolized vessels.

In conclusion, Citharexylum myrianthum root and stem wood were different in quantitative but not qualitative features. The root wood had lower frequency of vessels, narrower and longer vessel elements, a greater abundance of paratracheal axial parenchyma in the latewood, and taller rays than the wood of the stem. The calculated vulnerability and mesomorphy indices (Carlquist 1977) indicated that C. myrianthum is mesomorphic, but by comparison, the roots are more susceptible to water stress than the stem. The overall physiognomy of the wood suggests anatomical specialization to seasonally dry environment.

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