Shape Tuning and Size Prediction of Millimeter-Scale Calcium-Alginate Capsules with Aqueous Core

Jinchao Zhao 1,3, Qing Guo 1, Wei Huang 1, Teng Zhang 2, Jing Wang 1, Yu Zhang 2, Leping Huang 3,* and Youhong Tang 3,*

1 Hubei Biomass Fibers and Eco-dyeing & Finishing Key Laboratory, School of Chemistry and Chemical Engineering, Wuhan Textile University, Wuhan 430200, China; jczhao@wtu.edu.cn (J.Z.); whiteguo0524@163.com (Q.G.); huangwei19950904@163.com (W.H.); jing0123wang@163.com (J.W.)
2 School of Material Science and Engineering, Wuhan Textile University, Wuhan 430200, China; 1715043028@mail.wtu.edu.cn (T.Z.); zhangyuer126@126.com (Y.Z.)
3 Institute for NanoScale Science & Technology, College of Science and Engineering, Flinders University, South Australia 5042, Australia
* Correspondence: lphuang@wtu.edu.cn (L.H.); youhong.tang@flinders.edu.au (Y.T.);
Tel.: +86-27-59367580 (L.H.); +61-8-82012138 (Y.T.);
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Abstract: Controllable feature and size, good mechanical stability and intelligent release behavior is the capsule products relentless pursuit of the goal. In addition, to illustrate the quantitative relationship of structure and performance is also important for encapsulation technology development. In this study, the sphericity and size of millimeter-scale calcium sodium alginate capsules (mm-CaSA-Caps) with aqueous core were well tuned by manipulating the viscosity, surface tension, and density of CaCl2/carboxyl methyl cellulose (CMC) drops and sodium alginate (SA) solution. The well-tuned mm-CaSA-Caps showed significant mechanical and control-releasing property effects. The results showed that the prepared mm-CaSA-Caps were highly monodispersed with average diameter from 3.8 to 4.8 mm. The viscosity of the SA solution and the viscosity and surface tension of the CaCl2/CMC solution had significant effects on the mm-CaSA-Caps sphericity. Uniform and spherical mm-CaSA-Caps could be formed with high viscosity CaCl2/CMC solution (between 168.5 and 917.5 mPa·s), low viscosity SA solution (between 16.2 and 72.0 mPa·s) and decreased surface tension SA solution (by adding 0.01 wt.% poloxamer 407). The diameter of the mm-CaSA-Caps could be predicted by a modified Tate’s law, which correlated well with the experimental data. The Caps with sphericity factor (SF) < 0.07 had better mechanical stability, with the crushing force 2.91–15.5 times and the surface Young’s modulus 2.1–3.99 times higher than those of the non-spherical Caps (SF > 0.07). Meanwhile, the spherical Caps had a more even permeation rate, which was helpful in producing uniform and sustained releasing applications in foodstuff, medicine, agriculture and chemical industry.

Keywords: calcium-alginate capsules; size; sphericity; mechanical stability; drug release

1. Introduction

Fish-egg-like calcium sodium alginate (CaSA) matrix capsules (Caps) with an aqueous core at the millimeter scale prepared using the extrusion dripping method have been studied in food [1], agriculture [2] and biomedical industries [3,4]. Sodium alginate (SA), an anionic linear polysaccharide of 1,4-linked-α-L-guluronic acid and β-D-mannuronic acid residues that are widespread in seaweeds, can produce gel in the presence of divalent and trivalent cations and form non-toxic and biocompatible mini-dose containers by dripping SA into a calcium salt coagulation bath under very
mild crosslinking conditions [5]. The active compounds, from water to oil, encapsulated in the CaSA hydrogel matrix can be protected by a semipermeable gel membrane with controlled release and shielding functions [6,7]. For core-shell structures, CaCl2 droplets containing the active molecule and non-gelling polymer used to modulate viscosity and density are dropped into a SA bath to form the gel membrane in the interface of alginate molecules and calcium cations [8]. This inverted extrusion dripping method is popular due to its simple setup. As well, it ensures that bioactive materials to be encapsulated never come into contact with the ionotropic gel-forming polymers and remain within their original environment [8]. Moreover, this method is more likely to obtain microcapsules within a narrow dimensional range and with high encapsulation efficiency, thereby proving a good choice for industrialized production [9].

Sphericity of diameter and mono-dispersion are required for the production of high-quality mm-CaSA-Caps. It has been reported that capsules are easily deformed during the dripping process [5,10]. In the production of traditional CaSA solid core beads with a desired size and spherical shape [8], SA solution is dripped into a Ca2+ coagulation bath and some trial-and-error attempts are often required with regard to aspects of the liquid formulation and experimental setup, e.g., solution viscosity or surface tension, tip size and collecting distance, aspects that are important for mm-CaSA-Caps preparation. Lee et al. [11] discussed the influence of process variables of the method on the capsule size and shape by optimizing preparation conditions, notably the concentration of SA and CaCl2, gelation time, dripping tip diameter, gelation solution height and stirring rate of the gelation bath. Messaoud et al. [12] reported that the inner polymer thickening agent had an impact on membrane stability and could act as an internal coating or provide mechanical reinforcement that could effectively prevent breakage and leakage of capsules.

Efficiency of mass transfer and mechanical stability are vital for capsule applications. The capsules should not only control sustained release but also protect the inner fluid from mechanical stresses [13], especially with regard to their biological function. For example, red blood cells, which are natural microcapsules, can squeeze through capillaries with only 10% of their diameter without any damage due to their lowest bending resistance membranes [14]. Researchers have kept the focus on membrane modification to prepare capsules with specified releasing and mechanical properties that can meet different needs in various environments [15–21], but few researchers have paid attention to capsule shape and size manipulation and their effects on the final properties.

In this work, carboxyl methyl cellulose (CMC) was added as thickener to CaCl2 solution and dripped into a SA bath to form mm-CaSA-Caps. The diameter and sphericity of the capsules was regulated by controlling viscosity, surface tension and density of SA and CaCl2/CMC solutions. Capsule diameter was predicted based on the modified Tate’s law. In particular, the sphericity factor (SF) effect on the mechanical properties and permeability of mm-CaSA-Caps is discussed.

2. Experimental

2.1. Materials

Sodium alginate (SA, 10 g/L, 20 °C, > 0.02 Pa·s) was dissolved in deionized water as a gelation bath. Sodium carboxymethyl cellulose (CMC, 20 g/L, 25 °C, 800–1200 mPa·s) was used as a non-gelling thickener agent to increase the viscosity of the aqueous core solution. Tween 80, a nonionic surfactant, was used to modify the surface tension of the CMC solutions. Dextrose monohydrate, glycerin and sodium hydroxide used were of analytic grade. The above reagents were purchased from Sinopharm Chemical Reagent Co. Ltd., Shanghai, China. Poloxamer 407 (Lutrol F127, average Mn~12600), a nonionic surfactant used to modify the surface tension of the SA solutions, was provided by BASF, Ludwigshafen, Germany. 3,5-dinitrosalicylic acid (98%) was purchased from Macklin, Shanghai, China. Deionized water was used throughout the experiments.

2.2. Preparation of mm-CaSA-Caps

The mm-CaSA-Caps were prepared at 20–25 °C by an extrusion dripping device [11] as shown in Figure 1a. In detail, a solution containing 1 wt.% CMC and 1.5 wt.% CaCl2 was loaded into a 20
mL plastic syringe, pumped dropwise with the flow rate of 0.5 mL/min by a dripping nozzle into a gelation bath containing 200 mL 0.5 wt.% SA solution. A hypodermic needle with the outer diameter of 1.06 mm (19G) was used as the dripping nozzle. The distance between the dripping nozzle and the surface of SA solution was fixed at 10 cm. After 10 min dripping process, the solution was stirred for 20 min, and the obtained capsules were then washed with deionized water three times. Then, an additional crosslinking process for the capsules was conducted for 15 min in 2.0 wt.% CaCl₂ solution. Finally, the mm-CaSA-Caps were stored in deionized water before measurement. As shown in Figure 1, the mm-CaSA-Caps with the diameter of 4.170 ± 0.072 mm have a beautiful round shape (Figure 1b). The transparent membrane of the capsules was formed of an even 0.20 mm film (Figure 1c). After freeze drying for 24 h, the mm-CaSA-Caps retained their spherical structure (Figure 1d). The cross section of the membrane (Figure 1e) had the typical layered porous structure of a common hydrogel after dehydration, as shown in the high magnification (Figure 1e) of a rectangular portion from Figure 1d.

Figure 1. (a) Schematic drawing of the preparation process of millimeter-scale calcium sodium alginate capsules (mm-CaSA-Caps); (b) mm-CaSA-Caps (top view); (c) microscope image of the mm-CaSA-Caps (in water); (d) SEM image of the mm-CaSA-Caps (after freeze drying) and (e) high magnification of the rectangular part in (d).

2.3. Experimentation

The viscosity and density of SA and CaCl₂/CMC solutions were adjusted by concentration of solution. The concentrations of CMC were 0.5, 0.75, 1.0, 1.25 and 1.5 wt.%. The concentrations of SA were 0.25, 0.5, 0.75, 1.0 and 1.25 wt.%. The surface tension of the SA solution was adjusted by adding poloxamer 407. The concentrations of poloxamer 407 were 0, 0.01, 0.05, 0.1 and 0.2 wt.%. The surface tension of the CMC/CaCl₂ solution was adjusted by adding Tween 80. The concentrations of Tween 80 were 0, 0.015, 0.025, 0.038, 0.075, 0.15 and 0.3 g/L.

2.4. Measurement of Solution Properties

The viscosities of SA solution and CaCl₂/CMC solution were determined by a rotary viscometer (NDJ-5S, Shanghai Pingxuan Scientific Instrument Co., Ltd., Shanghai, China) in accordance with ISO3219:1993. During the viscosity measurements, the temperature was kept at 25 °C. The data were the averages of 3 measurements. The surface tension was determined using a surface tension instrument (DCAT21, Dataphysics Instruments GmbH, Filderstadt, Germany) in accordance with ISO 304:1985. The data were the averages of 3 measurements. The density was measured using the
the pycnometer method in accordance with ISO 1675: 1985. The data were the averages of 3 measurements.

2.5. Capsule Characterization

Images of the capsules in air and water were captured by a digital camera (BL-SM500, Jinhua Oseelang Trade Co., Ltd., Jinhua, China). The diameter of the capsules (D, mm, using Dmin as the diameter D.) was measured with a digital micrometer and the shape of the capsules was quantified using the sphericity factor (SF) in Equation (1) [22,23]

\[ SF = \frac{(D_{\text{max}} - D_{\text{min}})}{(D_{\text{max}} + D_{\text{min}})} \]  

where \( D_{\text{max}} \) is the longest diameter length, and \( D_{\text{min}} \) is the shortest diameter length perpendicular to \( D_{\text{max}} \). The capsules were considered spherical when \( SF < 0.07 \). To determine \( D \) and \( SF \), 15 capsules for each condition were randomly chosen.

No apparent trends of capsule membrane thickness were observed when the CMC concentration was increased [11]. Therefore, the effect of SF on the mechanical and diffusion properties of the capsules was investigated with the CMC concentration increased. Mechanical properties of the capsules were measured with a transducer rotational rheometer (Ar2000ex, TA Instruments, New Castle, DE, USA) and the force gap test was used to compress the capsules from 4.2 to 0.05 mm with the linear compression speed of 0.3 mm/s [12]. At least five replications were considered for each capsule type. The surface Young’s modulus (\( E_s \)), the measure of capsule stiffness, was estimated by analyzing the force–displacement curves in the range of small deformations and using Equation (2) [24]:

\[ F = \frac{4E_sh}{r\sqrt{3(1-\nu^2)}}d_d \]  

where \( d_d \) is the Caps displacement, \( F \) is the measured force, \( h \) is the membrane thickness, which is imported as a fixed value of 0.20 mm, \( r \) is the radius of the Caps and \( \nu \) is the surface Poisson ratio for which a value of 1/2 was assumed for the alginate hydrogel [25].

The permeability of mm-CaSA-Caps was measured by the diffusion of glucose from bulk solution into intra-hollow Caps. The concentration of glucose was measured by the dinitrosalicylic colorimetric method (DNS) [26]. In detail, 500 mm-CaSA-Caps was added into 100 mL glucose solution (20 mg/mL), of which 1 mL was taken at intervals into a 10 mL glass tube, followed by 1.5 mL DNS reagent and 1.5 mL deionized water, all shaken up and boiled for 5 min, cooled to room temperature, brought to volume in a 10 mL volumetric flask, and tested at 540 nm by an ultraviolet spectrophotometer. The variation of glucose concentration with time was ascertained against the glucose standard curve.

2.6. Prediction of the Diameter of mm-CaSA-Caps

The diameters of CaSA capsules (\( d_p \), mm) could be predicted using the modified Tate’s law mathematical model, given as Equation (3):

\[ d_p = K_{\text{LF}}(6d_t\gamma/\rho g)^{\frac{1}{3}} \]  

where \( K \) is the overall size correction factor, \( d_t \) is outer diameter of the needle (mm), \( \gamma \) is the surface tension of CaCl2/CMC solution (mN/m), \( \rho \) is the density of CaCl2/CMC solution (kg/m^3) and \( g \) is the gravitational force (9.81 m/s^2). Compared to the Tate’s law, the overall size correction factor \( K \) was introduced into the modified model to take into account the residual liquid at the dripping nozzle \( (K_{\text{SF}}) \) and the change of droplet diameter after gelation \( (K_{\text{SF}}) \). The \( K_{\text{LF}} \) and \( K_{\text{SF}} \) were calculated by Equations (4) and (5) [27].

\[ K_{\text{LF}} = 0.98 - 0.04d_t \]  
\[ K_{\text{SF}} = D/d_d \]
The outer diameter of the needle was 1.06 mm and \( K_{LF} \) was 0.9376 in this study. The diameter of the droplet \((d_d, \text{mm})\) was calculated by Equation (6) [27]:

\[
d_d = \sqrt[3]{\frac{6V}{\pi n}} = \sqrt[3]{\frac{6m}{\pi \rho n}}
\]  

(6)

where \( V \) is the volume of CaCl\(_2\)/CMC solution droplet (m\(^3\)), \( m \) is the mass of the CaCl\(_2\)/CMC solution droplet (kg) and \( \rho \) is the density of CaCl\(_2\)/CMC solution (kg/m\(^3\)). In this equation, it is assumed that the droplet is in the form of a sphere.

To evaluate the accuracy and reliability of the capsule diameter prediction model, average absolute deviation (AAD) and maximum absolute deviation (MAD) were introduced. AAD analysis indicates the average deviation of the experimental data and MAD analysis reflects the degree of deviation of the experimental data. AAD is given by Equation (7) [27] and MAD is defined as the maximum value of the absolute deviation between the experimental data and the reference values.

\[
\text{AAD} = \frac{\sum_{i=1}^{n} |d_p - D|}{n} \times 100
\]

(7)

where \( n \) is sample number of the capsules.

3. Results and Discussion

3.1. Solution Properties

The viscosities of the SA and CaCl\(_2\)/CMC solutions are shown in Figure 2. The viscosity of the SA solution increased from 16.2 to 516.0 mPa\( \cdot \)s in an exponential manner with the SA concentration increasing from 0.25 to 1.25 wt.\% (Figure 2a). These results were in good agreement with previous studies [9,28]. Similar trends were also evident in the CaCl\(_2\)/CMC solution. The viscosity of the CaCl\(_2\)/CMC solution increase from 20.0 to 917.5 mPa\( \cdot \)s as the concentration of CaCl\(_2\)/CMC increase from 0.5 to 1.5 wt.\% (Figure 2b).

![Figure 2. Viscosity of the solution as the function of SA (a) and CaCl\(_2\)/CMC (b) concentration.](image)

Two surfactants were used, poloxamer 407 and Tween 80, which reduced the surface tension of the SA and CMC solutions respectively. The critical micelle concentration of Tween 80 is 0.014 g/L, according to literature data [29]. As 0.1 wt.% poloxamer 407 was added into the SA solution, the surface tension decreased from 47.01 ± 0.246 to 41.71 ± 0.401 mN/m, as shown in Figure 3a. The surface tension of the SA/poloxamer continued to decrease when the concentration of poloxamer 407 increased. As shown in Figure 3b, the surface tension of the CaCl\(_2\)/CMC solution also obviously decreased with the addition of Tween 80. When 0.015 g/L Tween 80 was added into the CaCl\(_2\)/CMC solution, the surface tension of the mixed solutions decreased from 63.41 to 49.01. The surface tension continued to decrease with the increase in Tween 80 content, until it was close to a constant 37 mN/m at 0.075 g/L Tween 80. Comparison of Figure 3a,b shows that the surface tension of the SA solution with poloxamer 407 decreased gradually, unlike the obvious critical micelle concentration appearing...
in the CaCl₂/CMC/Tween 80 solution. Compared to the behavior of low-molecular-weight nonionic surfactants, the aggregation behavior of PEO-PPO-PEO surfactants like poloxamer 407 is complex, in that aggregation of poloxamer 407 occurs over a range of concentrations rather than at a unique critical micelle concentration [30].

Figure 3. The surface tension of the solution as the function of (a) 0.5 wt.% SA solution with different concentrations of poloxamer 407 and (b) 1.5 wt.% CaCl₂ and 1 wt.% CMC solution with different Tween 80 concentrations. Inserts show the chemical structures of poloxamer 407 and Tween 80.

Figure 4 shows that the concentrations of SA and CaCl₂/CMC solution have limited impact on the density of the solution. The densities of SA (1.05–1.06 g cm⁻³) and CaCl₂/CMC (1.06–1.07 g cm⁻³) solution are close to that of water (1.00 g cm⁻³). According to previous researches [31], the density of alginate solution showed minimal impact on the size and shape of Ca-alginate beads formation because the increment degree of alginate solution's density is only marginal, which is close to that of water. So, we did not make further experiments.

Figure 4. The density of (a) SA solutions at concentrations of 0.25, 0.5, 0.75, 1.0 and 1.25 wt.% and (b) CaCl₂/CMC solutions at CMC's concentrations of 0.5, 0.75, 1.0, 1.25 and 1.5 wt.%

3.2. Tuning Mechanical and Permeation Properties by Controlling SF

The SF of mm-CaSA-Caps is detailed in Table 1 in relation to the designed experiments. Spherical mm-CaSA-Caps (SF < 0.07) were obtained within the shaded area, which conformed with the viscosity of SA solutions < 72.0 mPa·s and the viscosity of CaCl₂/CMC solutions > 56.7 mPa·s.

Table 1. Effects of viscosity of SA and CaCl₂/CMC solutions on the sphericity factor (SF) of mm-CaSA-Caps.
### Table

| Viscosity of CaCl$_2$/CMC (mPa·s) | Viscosity of SA (mPa·s) |
|----------------------------------|-------------------------|
| 16.2                             | 56.7                    |
| 33.6                             | 0.067 ± 0.026           |
| 72.0                             | 0.086 ± 0.029           |
| 148.5                            | 0.258 ± 0.011           |
| 265.5                            | 0.394 ± 0.016           |
| 20.0                             | /                       |
| 168.5                            | 0.032 ± 0.019           |
| 372.0                            | 0.018 ± 0.011           |
| 917.5                            | 0.153 ± 0.035           |
|                                  | 0.289 ± 0.012           |
|                                  | 0.323 ± 0.007           |

Images of the mm-CaSA-Caps produced by different viscosities of SA solution with the viscosity of CaCl$_2$/CMC solution at 917.5 mPa·s are shown in Figure 5a–e. The mm-CaSA-Caps changed from spherical pearl shapes to teardrop shapes when the viscosity of the SA solution increased from 16.2 to 265.5 mPa·s. Plotting the average SF of column data in Table 1 against the viscosity of the SA solution produced the curve shown in Figure 5f. The SF of the mm-CaSA-Caps increased from 0.029 to 0.289. Uniform and spherical capsules could be formed when the viscosity value of the SA solution was less than 52.58 mPa·s.

**Figure 5.** Effect of SA solution viscosity (a) 16.2 mPa·s, (b) 33.6 mPa·s, (c) 72.0 mPa·s, (d) 148.5 mPa·s and (e) 265.5 mPa·s on (a)–(e) morphologies and (f) SF of the mm-CaSA-Caps when the viscosity of the CaCl$_2$/CMC solution is 917.5 mPa·s. The concentrations of SA were 0.25, 0.5, 0.75, 1.0 and 1.25 wt.%. The concentrations of CaCl$_2$ was 1.5 wt.%. The concentrations of CMC was 1.5 wt.%.
viscosity. Taking the average SF of column data in Table 1 and plotting it against the viscosity of the CaCl₂/CMC solution produced the curve shown in Figure 6f. That curve shows that the viscosity of the SA solution must be below 72 mPa·s and the viscosity of the CaCl₂/CMC solution must be between 56.7 and 917.5 mPa·s to obtain spherical mm-CaSA-Caps with SF < 0.07.

Figure 6. The effect of CaCl₂/CMC solution viscosity (a) 20.0 mPa·s, (b) 56.7 mPa·s, (c) 168.5 mPa·s, (d) 372.0 mPa·s and (e) 917.5 mPa·s on morphology and (f) SF of mm-CaSA-Caps when the viscosity of the SA solution is 72.0 mPa·s. The concentrations of SA were 0.75 wt.%. The concentrations of CaCl₂ was 1.5 wt.%. The concentrations of CMC were 0.5, 0.75, 1.0, 1.25 and 1.5 wt.%. The concentrations of SA were 0.75 wt.%. The concentrations of CaCl₂ was 1.5 wt.%. The concentrations of CMC were 0.5, 0.75, 1.0, 1.25 and 1.5 wt.%. The effect of the surface tension of the coagulation bath, i.e., SA/poloxamer 407, on the shape of the mm-CaSA-Caps was investigated. From Figure 7, the mm-CaSA-Caps prepared in 0.5 wt.% SA solution without poloxamer 407 were pear-shaped and had an obvious “tail” appearance, as shown in Figure 7a, e with SF = 0.087, which was above 0.07 (Figure 7i). It has been proved that non-spherical shape CaSA beads not only reduced the gel strength, but also resulted in uncontrolled release rate of the encapsulant compared to that of spherical beads [31]. After the addition of a small amount of poloxamer 407, the sphericity of capsules obviously improved, although there were still individual irregular capsules, as shown in Figure 7b,f with SF decreased to 0.014 (Figure 7i). The sphericity and uniformity of the capsules improved continuously while the surface tension of SA/poloxamer 407 solution continued to decrease and the SF decreased to 0.011. However, the surfactant poloxamer 407 had a limited effect on the surface tension of the SA solution, as shown in Figure 3a. The capsule shape did not obviously change with the increase in the concentration of poloxamer 407 (Figures 7 and 8). It has been proved that the penetration depth of droplets is mainly affected by the viscosity and surface tension of the bath solution [1]. During the production of the CaSA capsules, a surfactant was added to the gelation bath to reduce surface tension, and hence the impact force could be reduced to obtain spherical capsules [31]. Thus, the surfactant addition enhanced penetration and prevented shape deformation of the CaSA capsules.

Figure 7. The effect of SA/poloxamer 407 solution surface tension (a) 54 mN/m, (b) 52 mN/m, (c) 50 mN/m, (d) 48 mN/m, (e) 46 mN/m, (f) 44 mN/m, (g) 42 mN/m and (h) 40 mN/m on morphology and (i) SF of mm-CaSA-Caps when the SA concentration was 0.75 wt.%. The concentrations of CaCl₂ was 1.5 wt.%. The concentrations of CMC were 0.5, 0.75, 1.0, 1.25 and 1.5 wt.%.
Figure 7. Images of mm-CaSA-Caps prepared with different surface tensions of SA/poloxamer 407 solution (a) 47.01 mN/m, (b) 41.71 mN/m, (c) 39.6 mN/m, (d) 37.17 mN/m; (e)–(h) are higher magnification images of (a)–(d) respectively and (i) SF of the mm-CaSA-Caps. The concentration of SA was 0.5 wt.%. The concentration of CaCl₂ was 1.5 wt.%. The concentration of CMC was 1.0 wt.%. The concentrations of poloxamer 407 were 0, 0.01, 0.05 and 0.2 wt.%.

The effects of the surface tension of the inner layer solution on the size and shape of the mm-CaSA-Caps is shown in Figure 8a–g. It can be observed that uniform and spherical capsules were obtained at different surface tensions of CaCl₂/CMC/Tween 80 solution. As shown in Figure 8h, as the surface tension of the solution decreased from 63.41 to 36.63 mN/m, the SF of capsules increased but still remained below 0.07 and the D of capsules decreased from 4.18 to 3.88 mm. Spherical beads can be obtained with low impact force and high penetration depth, depending mostly on the surface tension of bath solution and the viscosity of droplets [1,31]. The surface tension of droplets had less impact on SF but a decrease in the surface tension of droplets was not beneficial to the form of spherical capsules.

Figure 8. Images of the mm-CaSA-Caps prepared at different surface tensions of CaCl₂/CMC/Tween 80 solution (a) 63.61 mN/m, (b) 49.07 mN/m, (c) 45.14 mN/m, (d) 39.85 mN/m, (e) 38.86 mN/m, (f) 38.29 mN/m, (g) 36.63 mN/m and (h) SF of the mm-CaSA-Caps. The concentration of SA was 0.5 wt.%. The concentration of CaCl₂ was 1.5 wt.%. The concentration of CMC was 1.0 wt.%. The concentrations of Tween 80 were 0, 0.015, 0.025, 0.038, 0.075, 0.15 and 0.3 g/L.

The force–displacement curve of mm-CaSA-Caps with different SF (Figure 9a–e) and the crushing force and the crushing displacement of the Caps (Figure 9f) are shown here. It can be seen that the mm-CaSA-Caps with SF = 0.258 ± 0.011 had no crushing point. They developed uneven capsule walls that functioned as stress concentrators, leading to breakage under a small force when the wall touched the plate of the rheometer and the touching force was beyond the rheometer’s detection limit. With the decrease in SF, the crushing point appeared and the crushing force and crushing displacement of the Caps clearly increased. However, the crushing force and crushing displacement appeared to show large errors when the SF of the mm-CaSA-Caps was above 0.07, as shown in Figure 9f. It was also evident that the mm-CaSA-Caps with SF > 0.07 had unstable mechanical properties. In comparison, the Caps with SF < 0.07 had good mechanical stability. The Caps with SF = 0.002 ± 0.002 had the highest crushing force of 18.1 N and the crushing displacement of 3.96 mm.
Figure 9. The effect of SF on the mechanical properties of mm-CaSA-Caps. The concentrations of CMC were 0.75 (a), 1.0 (b), 1.15 (c), 1.25 (d) and 1.5 (e) wt.%. The crushing force and the crushing displacement of the Caps with different SF (f). The concentration of SA was 0.75 wt.%. The concentration of CaCl$_2$ was 1.5 wt.%.

Force–displacement curves with the displacement of 0–1 mm were selected and linearly fitted to investigate the elastic deformation and surface elastic modulus ($E_s$) of mm-CaSA-Caps. As shown in Figure 10A, the slope of the initial force–displacement curve increased when the SF of Caps decreased. The calculated $E_s$ of the Caps showed similar results (Figure 10B). The spherical Caps (SF < 0.07) had good resistance to deformation, with the $E_s$ of Caps with SF of 0.002 and 0.026 being 146.1 and 139.4 N/m respectively, whereas the $E_s$ displayed a three-fold decrease of 50.1 N/m when the SF of Caps was 0.074. In general, compared with mm-CaSA-Caps with poor sphericity, Caps with good sphericity had better mechanical stability and higher crushing strength under the action of external force. When capsules had greater rigidity, a larger external force was required to achieve the same deformation. Good mechanical properties guaranteed long term stability for protection of the active ingredients, such as additives, fragrance, pesticides and decorative pearls used in food, home care products, agriculture and cosmetics areas [33].

Figure 10. (A) Magnification of force–displacement curves of CaSA capsules filled with the different SF for linear regression and (B) surface Young’s modulus ($E_s$) calculation. The concentrations of CMC were 0.75 (a), 1.0 (b), 1.15 (c), 1.25 (d) and 1.5 (e) wt.%. The concentration of SA was 0.75 wt.%. The concentration of CaCl$_2$ was 1.5 wt.%.
Effect of SF on the permeability of mm-CaSA-Caps was investigated via measurement of the diffusion of glucose from the bulk solution into intra-hollow Caps. As shown in Figure 11, a decrease of glucose concentration produced different behaviors with different SF of mm-CaSA-Caps below and above 0.07. The mm-CaSA-Caps with SF > 0.07 showed a faster initial permeation rate and shorter equilibrium time than observed in the mm-CaSA-Caps with SF < 0.07. Unlike the uniform capsule wall thickness of the spherical Caps, the pear-shaped Caps had thin wall structure at their tip, leading to rapid permeation of glucose. The spherical Caps had a more even permeation rate, conducive to uniform and sustained release of a loaded drug. Furthermore, the millimeter-scale alginate capsules are extensively used to encapsulate cells and immobilize enzymes. It is also important to adjust and control the permeability of solutes across the Ca-alginate capsule membrane [7].

![Figure 11](image_url)

**Figure 11.** The effect of SF on the permeability of mm-CaSA-Caps. The concentrations of CMC were 0.75 (SF = 0.258 ± 0.011), 1.0 (SF = 0.153 ± 0.035), 1.25 (SF = 0.026 ± 0.022) and 1.5 (SF = 0.002 ± 0.002) wt.%. The concentration of SA was 0.75 wt.%. The concentration of CaCl₂ was 1.5 wt.%.  

### 3.3. Size Control and Prediction

The results in Table 2 show that the outer layer SA solution and the inner layer CaCl₂/CMC solution had different levels of effect on the diameter of capsules. As the viscosity of SA solution increased from 16.2 to 265.5 mPa·s, the diameter of capsules decreased significantly under the different CaCl₂/CMC concentrations. Taking the average D of column data in Table 2 and plotting it against the viscosity of SA solution produces the curve shown in Figure 12a. The average diameter of mm-CaSA-Caps decreased 27.3%, i.e., from 4.418 to 3.213 mm. As the CaCl₂/CMC solution was dripped into the SA solution, the chelate reaction occurred quickly on the interface of CaCl₂/CMC and SA. The speed and degree of the reaction were controlled by the bidirectional diffusion of SA molecules and Ca²⁺, mainly by the Ca²⁺ diffusion rate. Since the number of SA molecules per unit volume increased at high SA concentrations, the number of binding sites of −COO⁻ for Ca²⁺ also increased, leading to higher degrees of crosslinking than those formed with lower SA concentrations. The crosslinking structure thus formed could prevent the Ca²⁺ diffusion, while the high viscosity of the SA solution as the external phase also decreased the diffusion rate of Ca²⁺, as illustrated in Figure 13. Therefore, the capsule diameter no longer increased in a short time [6]. The diameters of capsules formed with high SA concentrations were lower than those formed with low SA concentrations.

| Viscosity of CaCl₂/CMC (mPa·s) | 16.2 | 33.6 | 72.0 | 148.5 | 265.5 |
|---------------------------------|------|------|------|-------|-------|
| Viscosity of SA (mPa·s)         |      |      |      |       |       |
| 20.0                            |      |      |      |       |       |
From Table 2, the diameter of the mm-CaSA-Caps decreased as the viscosity of the CaCl₂/CMC solution increased when the viscosity of the SA solution was ≤ 33.6 mPa·s. However, no apparent trend was observed when the CaCl₂/CMC concentration increased with the viscosity of the SA solution ≥ 72.0 mPa·s. It has been reported that the hydroxyl groups in CMC have the potential to form hydrogen bonding with alginate carboxyl groups to enhance the mechanical stability of capsules [12,34]. Thus, when the viscosity of SA was low, the crosslinking density of CaSA was low during the initial reaction period of the CaCl₂/CMC drop entering the SA base, facilitating easier bidirectional diffusion of CMC and SA molecular in the interface. As the viscosities of CaCl₂/CMC and SA solution were both low, the diameter of capsules was high because of the easier polymer chain diffusion and hydrogen bonding formation. With the increase in viscosity of the CaCl₂/CMC solution, even the diffusion of the CMC chain became easier, as the CMC and SA molecules would have more hydrogen bond connection, increasing the interface viscosity and preventing further diffusion of CMC.
molecular chains. Therefore, the capsule diameters decreased with the increase in the viscosity of CaCl₂/CMC solution under the low viscosity of SA solution. When the viscosity of the SA solution increased, the molecular diffusion was mainly affected by the high crosslinking density of CaSA and the high viscosity of the SA solution. Effects on the viscosity of CaCl₂/CMC solution were not apparent. The diameter of the mm-CaSA-Caps no longer changed. Taking the average D of row data in Table 2 and plotting it against the viscosity of the CaCl₂/CMC solution produced the curve shown in Figure 12b, demonstrating that no apparent trend was observed when the CaCl₂/CMC concentration increased, as reported by others [11,35], the reason being that the concentration of Ca²⁺ was not changed in the CaCl₂/CMC solution, further evidence that the diameter of the mm-CaSA-Caps was mainly controlled by the Ca²⁺ diffusion.

The effects of surface tension of the coagulation bath (SA/poloxamer 407) on the size of mm-CaSA-Caps were investigated. The capsules diameters decreased a little with the decrease of the surface tension of SA/poloxamer 407 solution, as shown in Figure 12c. It had been reported that decreasing the surface tension of a gelation bath was beneficial to decreasing capsule deformation but had no apparent influence on the size of capsules [31] and beads [1]. The effects of the surface tension of the inner layer solution on the size of mm-CaSA-Caps are shown in Figure 12d. The surface tension of droplets had a greater impact on the diameter of the capsules [36], which decreased as the surface tension of CaCl₂/CMC solution decreased.

The diameter of mm-CaSA-Caps was predicted by Equation (2). The predicted diameter was verified with experimental data to evaluate the accuracy and reliability of the capsule’s diameter prediction model for mm-CaSA-Caps. As presented in Figure 12e, the predicted diameter was in good agreement with the experimental data. Error analysis of the capsule’s diameter predicted by the model (Figure 14) shows that the AAD and MAD were < 4% and 15%, respectively. This result demonstrated that the modified prediction model could be used to predict the diameter of mm-CaSA-Caps within reasonable deviation.

![Figure 14. Error analysis of the diameter of mm-CaSA-Caps.](image-url)
4. Conclusion

Millimeter-scale calcium-alginate capsules (mm-CaSA-Caps) were produced by the extrusion dripping method based on the ionotropic gelation principle. The viscosity and surface tension of SA solution and CaCl2/CMC solution were considered as major factors for tuning the shape of mm-CaSA-Caps. The results indicated that viscosities of SA solution and CaCl2/CMC solution were both critical factors for the preparation of spherical mm-CaSA-Caps. Deformed capsules were formed at high SA viscosity or low CaCl2/CMC solution viscosity, due to impact forces that distorted the shape of CaCl2/CMC droplets when they hit the surface of the viscous SA solution. A small amount of surfactant in the CaCl2/CMC solution was helpful for decreasing the SF of the mm-CaSA-Caps. For the diameter of mm-CaSA-Caps, the ranking of influential parameters is the viscosity of SA solution > surface tension of CaCl2/CMC solution > surface tension of SA solution > viscosity of the CaCl2/CMC solution. The diameter of capsules was predicted based on the modified Tate’s law with an absolute deviation of less than 4%. The SF of mm-CaSA-Caps had a marked impact on the mechanical properties and permeability of mm-CaSA-Caps. Spherical Caps had superior mechanical stability, higher crushing strength and higher steady-state permeation rate, findings that have broad application prospects in the food and medical fields.

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References

1. Davarci, F.; Turan, D.; Ozcelik, B.; Poneclet, D. The influence of solution viscosities and surface tension on calcium-alginate microbead formation using dripping technique. Food Hydrocolloid 2017, 62, 119–127.
2. Rakmai, J.; Cheirsilp, B.; Prasertsan, P. Enhanced thermal stability of cyclodextrin glycosyltransferase in alginate–gelatin mixed gel beads and the application for β-cyclodextrin production. Biocatal. Agric. Biotechnol. 2015, 4, 717–726.
3. Lee, K.Y.; Mooney, D.J. Alginate: Properties and biomedical applications. Prog. Polym. Sci. 2012, 37, 106–126.
4. Park, J.K.; Chang, H.N. Microencapsulation of microbial cells. Biotechnol. Adv. 2000, 18, 303–319.
5. Bremond, N.; Santanach-Carreras, E.; Chu, L.Y.; Bibette, J. Formation of liquid-core capsules having a thin hydrogel membrane: Liquid pearls. Soft Matter 2010, 6, 2484–2488.
6. Blandino, A.; Santanach-Carreras, E.; Chu, L.Y.; Bibette, J. Hydrogel membrane capsules: Liquid pearls. Soft Matter 2010, 6, 2484–2488.
7. Wang, J.; Jin, Y.; Xie, R.; Liu, J.; Ju, X.; Meng, T.; Chu, L. Novel calcium-alginate capsules with aqueous core and thermo-responsive membrane. J. Colloid. Interface Sci. 2011, 353, 61–68.
8. Nigam, S.C.; Tsao, I.F.; Sakoda, A.; Wang, H. Techniques for preparing hydrogel membrane capsules. Biotechnol. Tech. 1988, 2, 271–276.
9. Wischnewski, C.; Zwar, E.; Rehage, H.; Kierfeld, J. Strong deformation of ferrofluid-filled elastic alginate capsules in inhomogenous magnetic fields. Langmuir 2018, 34, 13534–13543.
10. Meiser, I.; Müller, S.C.; Zimmermann, H.; Ehrhart, F. Quantitative 3d high speed video analysis of capsule formation during encapsulation processes. In Proceedings of World Congress on Medical Physics and Biomedical Engineering, Munich, Germany, 7–12 September 2009; pp. 204–207.
11. Lee, B.B.; Ibrahim, R.; Chu, S.Y.; Zulkifli, N.A.; Ravindra, P. Alginate liquid core capsule formation using the simple extrusion dripping method. J. Polym. Eng. 2015, 35, 311–318.
12. Messaoud, G.B.; Sánchez-González, L.; Probst, L.; Desobry, S. Influence of internal composition on physicochemical properties of alginate aqueous-core capsules. J. Colloid Interface Sci. 2016, 469, 120–128.
13. Carin, M.; Barthes-Biesel, D.; Edwards-Levy, F.; Postel, C.; Andrei, D.C. Compression of biocompatible liquid-filled HSA-alginate capsules: Determination of the membrane mechanical properties. Biotechnol. Bioeng. 2010, 82, 207–212.
14. Fery, A.; Weinkamer, R. Mechanical properties of micro- and nanocapsules: Single-capsule measurements. *Polym. 2007*, *48*, 7221–7235.

15. Aguero, L.; Zaldivar-Silva, D.; Pena, L.; Dias, M.L. Alginate microparticles as oral colon drug delivery device: A review. *Carbohydr. Polym.* *2017*, *168*, 32–43.

16. Du, Y.; Huang, G.; Wang, H.; Xiao, J. Effect of high coacervation temperature on the physicochemical properties of resultant microcapsules through induction of Maillard reaction between soybean protein isolate and chitosan *J. Food. Eng.* *2018*, *234*, 91–97.

17. He, F.; Mei, L.; Ju, X.; Xie, R.; Wang, W.; Liu, Z.; Chu, L. pH-responsive controlled release characteristics of solutes with different molecular weights diffusing across membranes of Ca-alginate/protamine/silica hybrid capsules. *J. Membrane Sci.* *2015*, *474*, 233–243.

18. Mei, L.; Xie, R.; Yang, C.; Ju, X.; Wang, J.; Zhang, Z.; Chu, L. Bio-inspired mini-eggs with pH-responsive membrane for enzyme immobilization. *J. Membrane Sci.* *2013*, *429*, 313–322.

19. Mou, C.; Wang, W.; Li, Z.; Ju, X.; Xie, R.; Deng, N.; Chu, L. Trojan-Horse-Like Stimuli-Responsive Microcapsules. *Adv. Sci.* *2018*, *5*, 1700960.

20. Ribeiro, C.; Borges, J.; Costa, A.M.S.; Gaspar, V.M.; Bermudez, V.Z.; Mano, J.F. Preparation of well-dispersed chitosan/alginate hollow multilayered microcapsules for enhanced cellular internalization. *Molecules* *2018*, *23*, 625–641.

21. Wang, J.; Yu, H.; Xie, R.; Ju, X.; Yu, Y.; Chu, L.; Zhang, Z. Alginate/protamine/silica hybrid capsules with ultrathin membranes for laccase immobilization. *Aiche J.* *2013*, *59*, 380–389.

22. Chan, E.S. Preparation of Ca-alginate beads containing high oil content: Influence of process variables on encapsulation efficiency and bead properties. *Carbohydr. Polym.* *2011*, *84*, 1267–1275.

23. Balanč, B.; Kalušević, A.; Drvenica, I.; Coelho, I.; Djordjević, V.D.; Sousa, I.; Moldão-Martins, M.; Rakić, V.; Nedović, V.; et al. Calcium-alginate-inulin microbeads as carriers for aqueous carveja extract. *J. Food. Sci.* *2016*, *81*, E65–E75.

24. Leick, S.; Kott, M.; Degen, P.; Henning, S.; Päsler, T.; Suter, D.; Rehage, H. Mechanical properties of liquid-filled shellac composite capsules. *Phys Chem Chem Phys* *2011*, *13*, 2765–2773.

25. Degen, P.; Zwar, E.; Schulz, I.; Rehage, H. Magneto-responsive alginate capsules. *J. Phys. Condens. Mat.* *2015*, *27*, 194105.

26. Chai, Y.; Mei, L.; Lin, D.; Yao, S. Diffusion coefficients in intrahollow calcium alginate microcapsules. *J. Chem. Eng. Data* *2004*, *49*, 475–478.

27. Deng, H.; Zhang, C.; Xu, G.; Tao, Z.; Zhang, B.; Liu, G. Density measurements of endothermic hydrocarbon fuel at sub- and supercritical conditions. *J. Chem. Eng. Data* *2011*, *56*, 2980–2986.

28. Watanabe, H.; Matsuyama, T.; Yamamoto, H. Experimental study on electrostatic atomization of highly viscous liquids. *J. Electrostat.* *2003*, *57*, 183–197.

29. Katakam, M.; Bell, L.N.; Banga, A.K. Effect of surfactants on the physical stability of recombinant human growth hormone. *J. Pharm. Sci-U*. *1995*, *84*, 713–716.

30. Bohorquez, M.; Koch, C.; Trygstad, T.; Pandit, N. A study of the temperature-dependent micellization of pluronic f127. *J. Colloid Interf. Sci.* *1999*, *216*, 34–40.

31. Lee, B.B.; Ravindra, P.; Chan, E.S. Size and shape of calcium alginate beads produced by extrusion dripping. *Chem. Eng. Technol.* *2013*, *36*, 1627–1642.

32. Deng, Q.; Anilkumar, A.V.; Wang, T. The phenomenon of bubble entrapment during capsule formation. *J. Colloid Interf. Sci.* *2009*, *333*, 523–534.

33. Martins, E.; Poncelet, Denis.; Renard, D. Oil encapsulation in core-shell alginate capsules by inverse gelation. I: Dripping methodology. *J. Microencapsul.* *2017*, *34*, 82–90.

34. Ren, H.; Gao, Z.; Wu, D.; Jiang, J.; Sun, Y.; Luo, C. Efficient Pb(II) removal using sodium alginate-carboxymethyl cellulose gel beads: Preparation, characterization, and adsorption mechanism. *Carbohydr. Polym.* *2016*, *137*, 402–409.

35. Chai, Y.; Mei, L.; Wu, G.; Lin, D.; Yao, S. Gelation conditions and transport properties of hollow calcium alginate capsules. *Biotechnol. Bioeng.* *2004*, *87*, 228–233.

36. Chan, E.S.; Lee, B.B.; Ravindra, P.; Poncelet, D. Prediction models for shape and size of ca-alginate macrobeads produced through extrusion-dripping method. *J. Colloid Interf. Sci.* *2009*, *338*, 63–72.