Activity of terpenes derived from essential oils against *Sarcoptes scabiei* eggs

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**Abstract**

**Background:** The limited ovicidal activity of currently available acaricides is a significant obstacle to efficacious scabies treatment. Several essential oils or their respective components have proved to be active against the eggs of arthropods, mainly lice and ticks. Information on the activity of these oils and/or components against the eggs of mites remains very limited. The aim of this study was to assess the activity of six terpenes (carvacrol, eugenol, geraniol, citral, terpinen-4-ol and linalool) commonly found in essential oils against the eggs of *Sarcoptes scabiei*.

**Methods:** *Sarcoptes* eggs were exposed to paraffin oil containing 1, 2.5, or 5% of each terpene tested. After a 12-h exposure period, the eggs were washed and placed in paraffin oil for hatching. Embryonic development following treatment was assessed every day to determine the stage of developmental arrest.

**Results:** The median effective concentration to obtain 50% egg mortality (EC⁵₀) was 0.5, 0.9, 2.0, 4.8, 5.1 and 9.8% for carvacrol, eugenol, geraniol, citral, terpinen-4-ol and linalool, respectively. The microscopic images of eggs after each treatment indicated that these six terpenes may act by penetrating through the aeropyles on the egg surface.

**Conclusions:** In conclusion, carvacrol, eugenol and geraniol possess significant ovicidal activities, which should be considered as promising ovicidal agents for the treatment of scabies.

**Keywords:** *Sarcoptes scabiei*, Scabies, Essential oil, Terpenes, Ovicidal activity

**Background**

*Sarcoptes scabiei* is a parasitic mite responsible for the skin disease called scabies. With an estimated 200 million people infected worldwide, scabies is one of the most prevalent infectious skin diseases [1]. It remains a major public health issue in many resource-poor areas. In addition to being a highly contagious skin infestation, scabies can induce secondary bacterial infections and potentially life-threatening sequelae, including sepsis, post-streptococcal glomerulonephritis, rheumatic fever and heart disease [1].

Relatively few treatments are currently available for the management of human scabies; these include a number of topical agents, such as 5% permethrin (pyrethroid), 10–25% benzyl benzoate, crotamiton (aniline) or 0.5% malathion (organophosphorurate), and one oral treatment (ivermectin [macrocyclic lactone]) [2]. The limited ovicidal activity of these currently available acaricides has been and remains a significant obstacle for efficient scabies control. All treatments require a second administration of the agent after a 7- to 14-day interval to kill newly hatched mites, but poor patient compliance with repeated treatments can lead to treatment failure [3, 4]. Therefore, there is an urgent demand for novel drugs which can target the eggs of *S. scabiei*.

*Sarcoptes* eggs are oval in shape and exhibit numerous aeropyles on their external surface. The egg shell consists of two layers: an inner translucent layer and...
an outer layer of finger-like projections with rounded tips [5]. Bernigaud et al. [4] described two phenotypes of egg stage: (i) an early embryonic neurologically immature and immobile accumulation of cells with an absence of a differentiated nervous system; and (ii) a mature embryo, still inside the egg, with the same vital systems found in newly emerged larva, including nervous, respiratory and circulatory systems. The early embryonic stage of *Sarcoptes* eggs as drug targets appears to be unexplored so far.

Essential oils and their constituents represent an appealing alternative strategy against *S. scabiei*. Essential oils of tea tree (*Melaleuca alternifolia*) [6], lemon-grass (*Cymbopogon citratus*) [7], lavender (*Lavandula angustifolia* Mill.) [8], clove (*Eugenia caryophyllata*) [8, 9] and palmarosa (*Cymbopogon martini*) [8] have been shown to possess a strong activity against the mite and its stages (larvae, nymphs and adults) of *S. scabiei*, with supporting evidence that these terpenes may also be active against the eggs of mites [10]. The objective of the present study was to assess the activity of terpinen-4-ol and eugenol, were also effective against *Sarcoptes* mites [6, 9]. The terpene citral, linalool, geraniol and carvacrol were shown to have a significant acaricidal effect on *Psoroptes* mites and eggs [10, 11], with supporting evidence that these terpenes may also be active against the eggs of mites [10]. The objective of the present study was to assess the effect of terpinen-4-ol, citral, linalool, eugenol, geraniol and carvacrol on the eggs of *S. scabiei*.

**Methods**

The *Sarcoptes* eggs used in this analysis were collected from the crusts of naturally infested New Zealand White rabbits kept on a rabbit farm in Nanning, Guangxi Province, China. Prior to initiating the study, the farm owners were contacted and permission subsequently obtained to use the infected rabbits for the purpose of the study. The crusts were scraped from the skin of the infected rabbits with a scalpel, placed in Petri dishes and then transported to the laboratory within a few hours. *Sarcoptes* eggs were isolated individually using a needle for testing under a stereomicroscope (SMZ745; Nikon Corp., Tokyo, Japan; 2× magnification).

Six terpenes (terpinen-4-ol, citral, linalool, eugenol, geraniol and carvacrol) were purchased from Shanghai Macklin Biochemical Company (Shanghai, China). All compounds were of the highest purity available (purity 95–99%). To test the ovicidal activity of these compounds, *Sarcoptes* eggs at the early embryonic stage were placed on a microscope slide and exposed to paraffin oil containing 1, 2.5 or 5% of each terpene. After a 12-h exposure period, the eggs were washed and placed in paraffin oil for subsequent hatching. Eggs exposed to 25% benzyl benzoate (Aladdin, Shanghai, China) were the positive control group, and eggs exposed only to paraffin oil (Aladdin) were the negative control group. The eggs were incubated in a humidity chamber (≥70% relative humidity) at 35 °C for 5 days. Six replicates (each of 10 eggs) were performed for each terpene and each concentration.

To detect and describe changes in the development of the embryos, eggs at the early embryonic stage were exposed to 1% of each terpene and placed on a microscope slide. The development of these eggs in paraffin oil and in 25% benzyl benzoate was also assessed as controls, as was the effect of 25% benzyl benzoate separately. A normal development was obtained with eggs in paraffin oil. At least five eggs were used for each solution. The slides were incubated in a humidity chamber (≥70% relative humidity) at 35 °C for 72 h, following which the eggs were examined under a microscope (Eclipse 80i; Nikon Corp.; 40× magnification) and pictures taken every 12 h. During this process, the development of embryos in the eggs was observed. Eggs were considered dead if they failed to hatch after 72 h.

Results were analyzed using SPSS software version 20.0 (SPSS IBM Corp., Armonk, NY, USA). The median effective concentration to obtain 50% egg mortality (EC50) was calculated by probit regression analysis. Data were analyzed with one-way analysis of variance followed by least significant difference. Values of *P* < 0.05 were considered significant.

**Results**

At the test concentration of 5%, the ovicidal effect of carvacrol, eugenol, geraniol, citral, terpiene-4-ol and linalool was 100, 100, 91.7, 50.0, 48.3 and 36.7%, respectively (Fig. 1). A mortality rate of 81.7% and 7.8% was observed in the positive control group (eggs exposed to 25% benzyl benzoate) and the negative control group (non-treated eggs), respectively. Compared to the negative control, exposure to the six terpenes at the concentrations of 5% and 2.5%, as well as exposure to carvacrol and eugenol at the concentration of 1%, led to significant differences in *Sarcoptes* egg hatching rate (*P* < 0.01).
was no significant difference \( (P > 0.5) \) in terms of hatching rate between eggs in the positive control group and those exposed to all concentrations of carvacrol, eugenol at concentrations of 2.5% and 5% and geraniol at the 5% concentration. The \( EC_{50} \) value was 0.5, 0.9, 2.0, 4.8, 5.1 and 9.8% for carvacrol, eugenol, geraniol, citral, terpinen-4-ol and linalool, respectively (Table 1). Embryos in eggs exposed to 1% terpinen-4-ol (Table 2, images 8–11) those in eggs exposed to linalool (Table 2, images 16–19) developed normally and larvae hatched within 72 h. Embryos in eggs exposed to 1% citral (Table 2, images 12–15) developed to a certain stage but ultimately stopped developing; embryos in eggs exposed to 1% carvacrol (Table 2, images 26–28), 1% eugenol (Table 2, images 20–22), 1% geraniol (Table 2, images 23–25) and 25% benzyl benzoate (Table 2, images 5–7) did not develop by 24 h, and the eggs remained at the early embryonic stage (Table 2). Eggs shells in all treatment groups appeared to be intact.

**Discussion**

The results of the present investigation demonstrated for the first time the strong ovicidal activity of several terpenes extracted from essential oils against the eggs of *Sarcoptes scabiei*, of which carvacrol, eugenol and geraniol showed the highest activity. The ovicidal activity of carvacrol has been reported in ticks, with a hatching rate of 8.3% at 1% concentration [12]. The ovicidal activity of eugenol, geraniol, citral, terpinen-4-ol and linalool has been reported against *Psoroptes* eggs [10] and is consistent with the activity against *Sarcoptes* eggs demonstrated in the present study. The same terpenes have also been shown to have an effect on the eggs of lice [13, 14] and mosquitoes [15].

Many essential oils and/or their components have been shown to be neurotoxic, acting on different targets in the nervous system of arthropods. It has been proven that terpinen-4-ol, linalool, carvacrol and geraniol inhibit the activity of acetylcholinesterase in different insects (including head lice) and also ticks [16–18]; linalool and carvacrol interfere with \( \gamma \)-aminobutyric acid receptors in insects [17, 19]; eugenol, geraniol, carvacrol and citral block the octopamine receptor binding sites in insects [20, 21]; and eugenol interferes with cell membranes and organelles in epidermal and gut epithelia of *Sarcoptes* mites [9]. Information on the mechanisms of action of essential oils and/or their components on the eggs of arthropods is much more limited. In the present study, serial microscopic examination revealed that unhatched eggs, especially embryos in eggs exposed to carvacrol, eugenol and geraniol, ceased to develop within 0 and approximately 24 h and that the eggs remained in the early embryonic stage (Table 2). The ovicidal action of these terpenes is due to their ability to penetrate into the eggs, possibly through aeropyles on the surface of *Sarcoptes* egg shells [22]. Conversely, the lack of ovicidal activity of a chemical may be due to its poor penetration into the eggs. Active terpenes can kill eggs in the early embryonic stage. In our study, embryos in eggs exposed to those terpenes showing less ovicidal activity (linalool, citral, terpinen-4-ol) remained half-developed or were premature, but the eggs ultimately hatched, suggesting that these terpenes may not be neurotoxic to the mites.

In clinical practice, topical agents for the treatment of scabies should be applied onto the skin surface of patients and left for 8–12 h [23]. In the present study, we exposed the eggs to each solution for only 12 h, whereas the classical technique with a filter paper results in an exposure duration of up to several days. The classical technique consists of placing a filter paper at the bottom of a Petri dish and impregnating it with a test solution; eggs are then placed on the filter paper and exposed to the test solution during the whole process [10]. With a 12 h-exposure period, 19.3% of eggs were able to hatch when exposed to 25% benzyl benzoate, whereas only 8.3% of eggs finally hatched with the filter paper method [7]. In the present study, only carvacrol, eugenol or geraniol led to > 90% mortality rate of *Sarcoptes* eggs after 12 h exposure to the 5% solution. In preliminary tests with the filter paper method, we observed that all of the six terpenes (when used at the 5% concentration) were able to kill > 90% eggs (unpublished data). These findings suggest that carvacrol, eugenol and geraniol can act on eggs with a short contact time.

*Sarcoptes* mites are difficult to sample in large numbers from human patients, and no in vitro culture system has yet been established [3]. Mites from different hosts may exhibit minor differences in terms of morphology and host preference, but they share the same biology [24]. From previous studies, it can be postulated that there is no difference in the survival rate of *Sarcoptes* eggs from animal hosts and humans under the conditions tested [3].
The main limitation for the use of essential oils or related compounds (i.e. terpenes) is the risk of a skin reaction [25]. However, by limiting the dose and concentration, we can prevent essential oils from causing significant health risks. Tisserand and Young [25] recommend that the maximum dermal use level for carvacrol, eugenol and geraniol is 1, 0.5 and 5.3%, respectively. The EC\textsubscript{50} was 0.5, 0.9 and 2.0% for carvacrol, eugenol and geraniol, respectively. Therefore, the use of topical geraniol and carvacrol should be safe and be expected to have a satisfactory ovicidal effect against 	extit{S. scabiei}.

**Conclusions**

In conclusion, carvacrol, eugenol and geraniol possess significant ovicidal activities. The results of the present study provide a scientific basis for the application of essential oil and/or their constituents as a therapeutic agent for scabies. Further studies should consider testing combinations of terpenes to evaluate potential synergistic effects. In vivo evaluation (using an animal model of scabies) is also required.

**Table 1** Probit regression analysis of the activity of the six terpenes against 	extit{Sarcoptes scabiei} eggs

| Compounds     | EC\textsubscript{50} (95% confidence limits) | Pearson Chi-square |
|---------------|---------------------------------------------|-------------------|
| Terpinen-4-ol | 5.1% (3.8–12.2)                             | 15.8              |
| Citral        | 4.8% (3.5–11.5)                             | 15.2              |
| Linalool      | 9.8% (9.1–1.4)                              | 7.6               |
| Eugenol       | 0.9% (0.1–1.4)                              | 9.2               |
| Geraniol      | 2.0% (1.4–2.4)                              | 18.5              |
| Carvacrol     | 0.5% (0.7–1.1)                              | 9.2               |

\* Activity of tested compound was determined based on the median effective concentration to obtain 50% egg mortality (EC\textsubscript{50})

\*b 95% confidence limits were not available

**Fig. 1** Mortality rate of 	extit{Sarcoptes} eggs exposed to six terpenes at a concentration of 1, 2.5 and 5%, respectively. Asterisk above each bar indicates a significant difference (P<0.01) in egg mortality between the indicated concentration of terpene and paraffin oil (negative control). Hash sign above bar indicates a significant difference (P<0.01) in egg mortality between the indicated concentration of terpene and 25% benzyl benzoate (positive control).
**Table 2** Microscopic aspect of *Sarcoptes* eggs at different time points after exposure to terpinen-4-ol, citral, linalool, eugenol, geraniol and carvacrol over time

| Treatment groups | 0 h | 24 h | 48 h | 72 h |
|------------------|-----|------|------|------|
| Paraffin oil (negative control) | ![Image](image1.png) | ![Image](image2.png) | ![Image](image3.png) | ![Image](image4.png) |
| 25% Benzyl benzoate (positive control) | ![Image](image5.png) | ![Image](image6.png) | ![Image](image7.png) | ![Image](image8.png) |
| Terpinen-4-ol | ![Image](image9.png) | ![Image](image10.png) | ![Image](image11.png) | ![Image](image12.png) |
| Citral | ![Image](image13.png) | ![Image](image14.png) | ![Image](image15.png) | ![Image](image16.png) |
| Linalool | ![Image](image17.png) | ![Image](image18.png) | ![Image](image19.png) | ![Image](image20.png) |
| Eugenol | ![Image](image21.png) | ![Image](image22.png) | ![Image](image23.png) | ![Image](image24.png) |
| Geraniol | ![Image](image25.png) | ![Image](image26.png) | ![Image](image27.png) | ![Image](image28.png) |
| Carvacrol | ![Image](image29.png) | ![Image](image30.png) | ![Image](image31.png) | ![Image](image32.png) |
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Authors' contributions
ML, SL and ZY carried out the tests, data collection and statistical analysis. FF conceived the study and drafted the manuscript. CB and JG revised the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials
The data supporting the results of this paper are included in the paper.

Declarations

Ethics approval and consent to participate
The study protocol was approved by the ethics committee of Guangxi University (approval no: GXU2019-019).

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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References
1. Chandler DJ, Fuller LC. A review of scabies: an infestation more than skin deep. Dermatology. 2019;235:79–90.
2. Bernigaud C, Samaranwikkrama G, Jones MK, Gasser RB, Fischer K. The challenge of developing a single-dose treatment for scabies. Trends Parasitol. 2019;35(11):931–43.
3. Bernigaud C, Fernando DD, Lu H, Taylor S, Hartel G, Guillot J, et al. In vitro ovicidal activity of current and under-development scabicides: which treatments kill scabies eggs? Br J Dermatol. 2020;182:511–3.
4. Bernigaud C, Samaranwikkrama GR, Jones MK, Gasser RB, Fischer K. The challenge of developing a single-dose treatment for scabies. Trends Parasitol. 2019;35:931–43.
5. Mazzini M, Baiocchi R. Fine morphology of the egg-shell of Sarcoptes scabiei (L.) (Acarina: Sarcoptidae). Int J Parasitol. 1983;13:469–73.
6. Walton SF, McNinnon M, Pizzutto S, Dougall A, Williams E, Currie BJ. Acaricidal Activity of Melaleuca alternifolia (tea tree) oil: in vitro sensitivity of Sarcoptes scabiei var hominis to Terpinen-4-ol. Arch Dermatol. 2004;140:563–6.
7. Li M, Liu B, Bernigaud C, Fischer K, Guillot J, Fang F. Lemongrass (Cymbopogon citratus) oil: a promising miticidal and ovicidal agent against Sarcoptes scabiei. PLoS Negl Trop Dis. 2020;14:e0008225. https://doi.org/10.1371/journal.pntd.0008225.
8. Fang F, Candy K, Melloul E, Bernigaud C, Chai L, Darmon C, et al. In vitro activity of ten essential oils against Sarcoptes scabiei. Parasit Vectors. 2016;9:594.
9. Pasay C, Mounsey K, Stevenson G, Davis R, Arlian L, Morgan M, et al. Acaricidal activity of eugenol based compounds against scabies mites. PLoS ONE. 2010;5(8):e12079. https://doi.org/10.1371/journal.pone.0012079.
10. Fang F, Li M, Jiang Z, Lu X, Guillot J, Shi H. Comparing acaricidal and ovicidal activity of five terpenes from essential oils against Psoroptes ovis in vitro and in vivo. Parasit Vectors. 2019;12:425.
11. Chen Z, van Mol W, Vanhecke M, Duchateau L, Claerebout E. Acaricidal activity of plant-derived essential oil components against Psoroptes ovis in vitro and in vivo. Parasit Vectors. 2019;12:425.
12. Tabari MA, Youssif MR, Maggi F, Benelli G. Toxic and repellent activity of selected monoterpenoids (thymol, carvacrol and linalool) against the castor bean tick, Ixodes ricinus (Acari: Ixodidae). Vet Parasitol. 2017;245:86–91.
13. Yang Y-C, Lee SH, Clark JM, Ahn Y-J. Ovicidal and adulticidal activities of Origanum majorana essential oil constituents against insecticide-susceptible and pyrethroid/malathion-resistant Pediculus humanus capitis (Anoplura: Pediculidae). J Agric Food Chem. 2009;57:2282–7.
14. Yang Y-C, Lee S-H, Lee W-J, Choi D-H, Ahn Y-J. Ovicidal and adulticidal effects of Eugenia caryophyllata bud and leaf oil compounds on Pediculus capitis. J Agric Food Chem. 2003;51:4884–8.
15. Tabari MA, Youssif MR, Esfandiarani A, Benelli G. Toxicity of β-citronellol, geraniol and linalool from Pelargonium rassum essential oil against the West Nile and filarialis vector Culex pipiens (Diptera: Culicidae). Res Vet Sci. 2017;114:36–40.
16. Anderson JA, Coats JR. Acetylcholinesterase inhibition by nootkatone and carvacrol in arthropods. Pestic Biochem Physiol. 2012;102:124–8.
17. Lope MD, Pascual-Villalobos MJ. Mode of inhibition of acetylcholinesterase by monoterpenoids and implications for pest control. Ind Crops Prod. 2010;31:284–8.
18. Mills C, Cleary BJ, Gilmer JF, Walsh JJ. Inhibition of acetylcholinesterase by Tea Tree oil. J Pharm Pharmacol. 2004;56:375–8.
19. Tong F, Coats JR. Effects of monoterpenoid insecticides on [3H]-TBOB binding in fly GABA receptor and 36Cl—uptake in American cockroach ventral nerve cord. Pestic Biochem Physiol. 2010;98:317–24.
20. Enan EE. Molecular response of Drosophila melanogaster tyramine receptor cascade to plant essential oils. Insect Biochem Mol Biol. 2005;35:309–21.
21. Price DN, Berry MS. Comparison of effects of octopamine and insecticidal essential oils on activity in the nerve cord, foregut, and dorsal unpaired median neurons of cockroaches. J Insect Physiol. 2006;52:309–19.
22. Mazzini M, Baiocchi R. Fine morphology of the egg-shell of Sarcoptes scabiei (L.) (Acarina: Sarcoptidae). Int J Parasitol. 1983;13:469–73.
23. Salavastru CM, Chosidow O, Boffa MJ, Janier M, Tiplica GS. European guideline for the management of scabies. J Eur Acad Dermatol Venereol. 2017;31:1248–53.
24. Arlian LG. Biology, host relations, and epidemiology of Sarcoptes scabiei. Annu Rev Entomol. 1989;34:139–61.
25. Tisserand R, Young R. Essential oil safety. Amsterdam: Elsevier; 2014.

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