Genetic predisposition to male breast cancer in Poland

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Abstract

Background: Breast cancer in men accounts for fewer than 1% of all breast cancer cases diagnosed in men and women. Genes which predispose to male breast cancer include BRCA1 and BRCA2. The role of other genes is less clear. In Poland, 20 founder mutations in BRCA1, BRCA2, CHEK2, PALB2, NBN, RECQL are responsible for the majority of hereditary breast cancer cases in women, but the utility this genes panel has not been tested in men.

Methods: We estimated the prevalence of 20 alleles in six genes (BRCA1, BRCA2, CHEK2, PALB2, NBN, RECQL) in 165 Polish male breast cancer patients. We compared the frequency of selected variants in male breast cancer cases and controls.

Results: One of the 20 mutations was seen in 22 of 165 cases (13.3%). Only one BRCA1 mutation and two BRCA2 mutations were found. We observed statistically significant associations for PALB2 and CHEK2 truncating mutations. A PALB2 mutation was detected in four cases (OR = 11.66; p < 0.001). A CHEK2 truncating mutation was detected in five cases (OR = 2.93; p = 0.02).

Conclusion: In conclusion, we recommend that a molecular test for BRCA1, BRCA2, PALB2 and CHEK2 recurrent mutations should be offered to male breast cancer patients in Poland.

Keywords: BRCA1, BRCA2, PALB2, CHEK2, NBN, RECQL, Mutation, Male breast cancer

Background

Breast cancer in men accounts for fewer than 1% of all breast cancer cases diagnosed in men and women [1]. The risk of breast cancer is elevated in men with a BRCA1 or BRCA2 mutation [2–4]. The prevalence of BRCA2 mutations in men with breast cancer ranges from 4 to 40% in different populations [5–13]. The prevalence of BRCA1 mutations in men with breast cancer is lower; estimates range from 0 to 7% [8, 14–18]. There are reports of male patients with mutations in PALB2 and CHEK2 [12, 16, 19–22]. The risk of male breast cancer in carriers of PALB2 mutations is increased by approximately 7-fold [12, 16, 19, 21, 23–26]. The common truncating allele of CHEK2 (1100delC) was associated with a 4.5-fold increased risk of the male breast cancer in the Scandinavian population [22]. Poland is a genetically homogeneous population. Twenty founder alleles in BRCA1, BRCA2, CHEK2, NBN, PALB2 and RECQL genes are responsible for the majority of hereditary breast cancers in Polish women. We identified one of these 20 mutations in about half of women with familial breast cancer (the mean number of breast cancers per family was 3.6) [27]. This panel of 20 founder mutations in BRCA1, BRCA2 CHEK2, NBN, PALB2 and RECQL genes included about 85% of all mutations detected in high-risk families in Poland [27]. One
of these founder mutations is present in 12.5% in Polish women with breast cancer diagnosed before age 51, and in 17% in Polish women diagnosed at age 40 or below [28–33].

Founder mutations in the BRCA, CHEK2 and PALB2 genes have been described in the literature in other ethnic populations. Founder BRCA mutations have been detected in the population of Ashkenazi women, in Russia, Latvia, Slovenia, Slovakia, Germany, Finland, the Czech Republic, Tunisia, Hungary and the Bahamas, and accounted for 20 to 90% of all detected mutations in these genes [34–45].

The CHEK2 1100delC mutation is present in Eastern and Northern European populations with a frequency of 0.2–1.4% in the general population [46, 47]. Two other truncating mutations of CHEK2 (c.444 + 1G > A, del5395(ex10-11del)) are predominantly detected in Eastern European countries, (c.444 + 1G > A in Poland, Belarus, Russia; and del5395(ex10-11del) in Poland, Belarus, Czech Republic, Slovakia) [35, 48]. Both alleles are also present at lower frequencies in Germany (0.1% del5395(ex10-11del) and 0.3% for c.444 + 1G > A) [49]. Recurrent PALB2 mutations have been detected in Finland, Argentina, Spain, China and Canada with a frequency of 0.6–3.6% [50–55].

In the current study, we evaluated the frequency of 20 recurrent mutations in six genes (BRCA1, BRCA2, CHEK2, PALB2, NBN and RECQL) in 165 men diagnosed with breast cancer in Poland.

**Methods**

**Patients**

Patients were selected from a registry of breast cancer cases held at the Hereditary Cancer Center in Szczecin. We identified 186 men with breast cancer from a registry of 25,000 breast cancer cases diagnosed in Poland. Patients had a genetic consultation between 2000 and 2019 at the Hereditary Cancer Center in Szczecin. During the patient interview a written, informed consent and a blood sample were obtained from all subjects. All patients were of European ancestry and ethnic Poles.

Because of the quality of the DNA we were able to include 165 men in the study (88.7%). Clinical data including family history (cancer cases in first and second-degree relatives) and tumor characteristics were obtained.

Table 1: Clinical characteristics of breast cancers in mutation carriers and non-carriers

| Mutation       | BRCA1 | BRCA2 | PALB2 | CHEK2 truncating | CHEK2 c.470C > T | NBN | Non-carriers |
|---------------|-------|-------|-------|------------------|------------------|-----|--------------|
| Histology     |       |       |       |                  |                  |     |              |
| Ductal        | 1 1   | 1 1   | 1 1   | 1 1              | 1 1              |     |              |
| Lobular       | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Other         | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| T1/T2         | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| T3/T4         | 1 1   | 1 1   | 1 1   | 1 1              | 1 1              |     |              |
| ER positive   | 1 1   | 1 1   | 1 1   | 1 1              | 1 1              |     |              |
| PR positive   | 1 1   | 1 1   | 1 1   | 1 1              | 1 1              |     |              |
| HER2 positive | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Triple negative | 0 1 | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Lymph nodes positive | 0 1 | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Bilateral     | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Multiple primary cancers | 0 1 | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Family history positive | 1 1 | 1 1   | 1 1   | 1 1              | 1 1              |     |              |
| Breast        | 1 1   | 1 1   | 1 1   | 1 1              | 1 1              |     |              |
| Ovarian       | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Other         | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| Age of diagnosis |     |       |       |                  |                  |     |              |
| < 50          | 0 1   | 0 1   | 0 1   | 0 1              | 0 1              |     |              |
| ≥ 50          | 1 1   | 1 1   | 1 1   | 1 1              | 1 1              |     |              |
| Median age of diagnosis (range) | 74 | 69 (62–76) | 57 (42–77) | 62 (33–70) | 64 (34–72) | 62 | 63 (26–89) |
during an interview with the patient and from medical records (Table 1). All clinical data were obtained in more than 70% of cases.

Controls
The frequencies of *BRCA1*, *CHEK2*, *PALB2*, *NBN* in cancer-free controls were taken from previous studies [29, 30, 56]. The frequency of mutations in the *BRCA1* and *NBN* genes were determined in the group 4000 samples described by Lerner et al. [56]. The frequency of mutations in the *CHEK2* gene were determined in the group of 5496 samples described by Cybulski et al. [29]. The frequency of mutations in the *PALB2* gene were determined in the group of 4702 samples described by Cybulski et al. [30].

Genotyping
DNA was isolated from 5 to 10 mL of peripheral blood. Recurrent mutations of *BRCA1*, *BRCA2*, *CHEK2*, *PALB2*, *NBN*, *PALB2* and *RECQL* were genotyped as described previously [28–32]. The *BRCA1* 5382insC (c.5263_5264insC) and 4153delA (c.4035delA) mutations were detected using allele-specific oligonucleotide polymerase chain reaction (PCR) and C61G (c.181 T > G) was detected using restriction fragment length polymorphism PCR. The other mutations of *BRCA1*: 3819del5 (c.3770_3704delGTAAAA), 185delAG (c.68_69delAG), 5370C > T (c.5251C > T), 3875del4 (c.3756_3759delGTCT) and *BRCA2* (c.658_659delGT, c.3847_3848delGT, c.5239_5240insT, c.5946delT, c.7913_7917del5) were genotyped using TaqMan assays (Applied Biosystems/Life Technologies, Carlsbad, CA) on Roche LightCycler 480. The c.444 + 1G > A and c.470C > T in *CHEK2* were detected using restriction fragment length polymorphism PCR. The *CHEK2* del5395(ex10-11del) was tested by a multiplex PCR reaction and c.1100delC was detected by allele-specific oligonucleotide PCR. One mutation on *NBN* was detected using allele-specific oligonucleotide PCR. The two mutations of *PALB2* c.509_510delGA and c.172_175delTTGT and one c.1667_1667 + 3delAGTA of *RECQL* were analyzed using TaqMan assays (Applied Biosystems/Life Technologies, Carlsbad, CA) on Roche LightCycler 480. All mutations were confirmed by Sanger direct sequencing. Sequencing reactions were performed using a BigDye Terminator v3.1 Cycle Sequencing Kit (Life Technologies) according to the manufacturer’s protocol. Sequencing products were analyzed on the ABI Prism 3100 Genetic Analyzer (Life Technologies). The DNA quality allowed for analysis of all 20 alleles in 165 from 186 patients (88.7%). Statistical analysis comparing the frequency of mutations in the cases and controls was performed in this group of 165 cases.

Statistical analysis
We estimated the odds ratios for selected recurrent mutations. We compared the prevalence of the ten different mutant alleles in *BRCA1*, *CHEK2*, *PALB2*, *NBN* in the study group and in control groups. Odds ratios (OR) were generated from two-by-two tables and statistical significance was assessed using Fisher exact test or Chi-squared test with Yates correction. ORs were used as estimates of relative risk.

Results
There were 165 men with breast cancer included in the study. The median age of diagnosis was 62 years (range 26–89 years). Twenty-five cases were diagnosed before age 50 (15.2%) (Table 1). The majority of cases were diagnosed with ductal carcinoma (89.6% of cases), followed by lobular (4.2% of cases) and papillary cancer (3.7% of cases). 91.4% were estrogen receptor (ER) positive and 82.7% were progesterone receptor (PR) positive. Triple negative receptor status was found in 8.9% cases. Bilateral breast cancer was diagnosed in two patients (1.2%) (Table 1).

Multiple primary cancers were diagnosed in 20 of 165 patients (12.1%). Prostate cancer was diagnosed in five cases (3.0%) and colorectal cancer was diagnosed in four patients (2.4%). Approximately one-half of the patients had a positive family history of cancer: 31.5% for breast and/or ovarian cancer and 36.4% for other cancers. The clinical features of breast cancer diagnosed in carriers of mutations and non-carriers are presented in Table 1.

We did not find statistically significant differences between mutation carriers and non-carriers (p > 0.5), except for a higher incidence of bilateral breast cancer in *PALB2* mutation carriers (p = 0.004).

One of twenty founder mutations was diagnosed in 22 of 165 men (13.3%) (Table 2). A *CHEK2* mutation was found in 14 patients (8.5%). The most commonly detected *CHEK2* allele was the missense c.470C > T mutation, which was found in nine subjects (5.5%). A protein truncating mutation in *CHEK2* gene was found in five patients (3.0%). A *PALB2* mutation was seen in four patients (2.4%). A *BRCA2* mutation was found in two patients (1.2%) and a *BRCA1* mutation was found in only one patient (0.6%). One mutation was found in *NBN* (0.6%) and no mutations were found in *RECQL*.

A mutation was found in 20% of men diagnosed before age 50 and in 12.9% of men diagnosed after age 50. A mutation was found in 9.7% of men with ductal carcinoma and 15.2% of men with other histopathological subtypes. Mutations were more common in familial cases (17.3%) than in non-familial cases (11.5%).

Compared to the frequency of mutations in cancer-free controls from our previous studies, we observed statistically significant associations for *PALB2* (OR = 11.66
In two men, three primary cancers were diagnosed (1.3%). Both of these patients were diagnosed with bilateral breast cancer and both carried a mutation in the \( \text{PALB2} \) gene.

\( ^a \) for \( \text{BRCA1}, \text{CHEK2}, \text{PALB2}, \text{NBN} \) mutation frequencies in cancer-free controls were from our previous studies [29, 30, 56].

**Discussion**

A founder mutation of \( \text{BRCA1}, \text{BRCA2}, \text{CHEK2}, \text{PALB2} \) and \( \text{NBN} \) was found in 13.3% of men with breast cancer in Poland. The mutation frequency was similar in familial and non-familial cases and in young men and older men.

This study supports the recommendation that a panel test be offered to all Polish men with breast cancer. In a previous study we have shown that 20 founder mutations account for 85% of all mutations in Polish women with hereditary breast cancer and were detected in 46% cases [27].

We identified a strong association with \( \text{PALB2} \) mutation and male breast cancer (OR = 11.6). In previous reports from other ethnic groups, the frequency of \( \text{PALB2} \) mutations ranged from 0.8 to 6.4% [12, 19, 21, 23–26]. Two previous studies reported an association of \( \text{PALB2} \) mutations and breast cancer risk in men [21, 23]. Pritzlaff et al. reported a 6.6-fold increased risk \( (p = 0.013) \) in an international cohort of 715 male breast cancer patients [21].

Our study is the first that evaluated the frequency of mutations in the \( \text{PALB2} \) gene in men with breast cancer.

| Mutations | All patients \((n = 165)\) | Family History Positive \((n = 52)\) | Family History Negative \((n = 113)\) | Chi-square test |
|-----------|---------------------------|--------------------------|------------------|----------------|
| **Any \( \text{BRCA1} \)** | | | | |
| c.5263_5264insC | 1 | 0.6 | 1 | 1.9 | 0 | 0.0 | 0.14 |
| c.181 T > G | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| c.4035delA | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| c.3700_3704delGTAAA | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| c.68_69delAG | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| c.5251C > T | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| c.3756_3759delGTCT | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| **Any \( \text{BRCA2} \)** | 2 | 1.2 | 1 | 1.9 | 1 | 0.9 | 0.57 |
| c.658_659delGT | 1 | 0.6 | 1 | 1.9 | 0 | 0.0 | 0.00 |
| c.5946delT | 1 | 0.6 | 0 | 0.0 | 1 | 0.9 | 0.00 |
| c.3847_3848delGT | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| c.5239_5240insT | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| c.7913_7917del5 | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| **Any \( \text{PALB2} \)** | 4 | 2.4 | 3 | 5.8 | 1 | 0.9 | 0.57 |
| c.509_510delGA | 4 | 2.4 | 3 | 5.8 | 1 | 0.9 | 0.00 |
| c.172_175delTTGT | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |
| **Any \( \text{CHEK2} \)** | 14 | 8.5 | 4 | 7.7 | 10 | 8.8 | 0.80 |
| **CHEK2 truncating** | 5 | 3.0 | 2 | 3.85 | 3 | 2.7 | 0.68 |
| c.100delC | 3 | 1.8 | 1 | 1.9 | 2 | 1.8 | 0.00 |
| c.444 + 1G > A | 1 | 0.6 | 1 | 1.9 | 0 | 0.0 | 0.00 |
| del5395(ex10-11del) | 1 | 0.6 | 0 | 0.0 | 1 | 0.9 | 0.00 |
| c.470C > T | 9 | 5.5 | 2 | 3.85 | 7 | 6.2 | 0.00 |
| **\( \text{NBN} \). c.657_661delACAAA** | 1 | 0.6 | 0 | 0.0 | 1 | 0.9 | 0.50 |
| **\( \text{RECQL} \). c.1667_1667 + 3delAGTA** | 0 | 0.0 | 0 | 0.0 | 0 | 0.0 | 0.00 |

\( BC \) breast cancer, \( \text{OV} \) ovarian cancer
from Poland, which is populated by ethnic Slavs. In the current study two patients with a PALB2 mutation were diagnosed with bilateral breast cancer and they were diagnosed with the third primary cancer. One of these had esophageal cancer and the other had bladder cancer. In previous studies, three male breast cancer patients with a PALB2 mutation had a second primary cancer (thyroid cancer, melanoma, prostate) [16, 21]. Male breast cancer patients who carry a mutation in PALB2 gene may be candidates for screening for second primary malignancies.

We have previously reported relatively poor survival for women with breast cancer and a PALB2 mutation [30]. Female breast cancer patients who carried one of the two Polish PALB2 founder mutations had a 10-year survival of 48%, compared to 75% for patients with breast cancer without a PALB2 mutation (HR = 2.27, 95% CI 1.64–3.15; p < 0.0001). Studies from Finland and China also reported poor survival of PALB2-associated breast cancer patients [57, 58]. In our cohort, 3 of 4 men with breast cancer and a PALB2 mutation died of metastatic cancer. The first patient was diagnosed with bilateral breast cancer at age 52 and died at age 72 of esophageal cancer. The second was diagnosed with unilateral breast cancer at age 42 and died of metastatic breast cancer at age 53. The third patient had bladder cancer and bilateral breast cancer at age 77 and died of breast cancer at age 79. The fourth patient was diagnosed with breast cancer at age of 42 (in 2017) and is currently alive. This data suggests poor prognosis in men with breast cancer and a PALB2 mutation but larger studies are necessary.

We observed a correlation between the three truncating mutation in the CHEK2 gene (c.1100delC, c.444 + 1G > A and del5395(ex10-11del)) and male breast cancer in Poland (OR = 2.93; p = 0.02). The 1100delC mutation in CHEK2 has been evaluated in several other studies [59–63] and four of these showed an increased risk of male breast cancer. In Finland, Hallamies et al. reported an odds ratio of 4.5 for men with the 1100delC allele (p = 0.02) [22]. A study in Netherlands confirmed the increased risk (OR: 4.1, p = 0.005) [20]. In the largest study of 715 male breast cancers the odds ratio associated with the 1100delC mutation in the CHEK2 gene was 3.8 (p = 0.002) [21]. The frequency of the c.1100delC allele in CHEK2 gene varies widely in different populations; the frequency is high in Finland (1.1–1.4%) and the Netherlands (1.3–1.6%) but is lower in Poland (0.2%). As a consequence the contribution of this allele to the burden of male breast cancer varies widely from country to country [20, 22, 56].

We found that the frequency of the 1100delC founder mutation in the CHEK2 gene to be higher in male breast cancer cases (1.8%) than in females with breast cancer (0.6%) [29]. A similar result was obtained by Hallamies at al. in Finland. The observed frequency of this mutation was higher in male breast cancer patients (5.9%) then among female patients (2.0%) [22, 64].

We did not find a significant association between the c.470C > T mutation in the CHEK2 gene and the risk of male breast cancer (p = 0.84). However, multiple primary cancers were diagnosed in three of nine carriers of this mutation (33.3%)(rectal cancer, myeloid leukemia and skin squamous cell carcinoma) In a previous study we

| Gene     | Mutation                          | All patients | Control* | OR    | 95%CI     | p-value |
|----------|-----------------------------------|--------------|----------|-------|-----------|---------|
| BRCA1    | c.5263_5264insC                   | 1/165        | 0.6%     | 12/4000 | 0.32%     | 1.87    | 0.24–14.38 | 0.54 |
|          | c.181 T > G                       | 0/165        | 0.0%     | 3/4000  | 0.08%     | –       | –         |      |
|          | c.4035delA                        | 0/165        | 0.0%     | 1/4000  | 0.03%     | –       | –         |      |
|          | Any BRCA1                         | 1/165        | 0.6%     | 17/4000 | 0.42%     | 1.43    | 0.19–10.80 | 0.73 |
| PALB2    | 509_510delGA                     | 4/165        | 2.4%     | 7/4702  | 0.1%      | 16.70   | 4.83–57.50 | <0.001 |
|          | 172_175delITTGTT                  | 0/165        | 0.0%     | 3/4702  | 0.05%     | –       | –         |      |
|          | Any PALB2                         | 4/165        | 2.4%     | 10/4702 | 0.15%     | 11.66   | 3.62–37.57 | <0.001 |
| CHEK2    | 1100delC                          | 3/165        | 1.8%     | 12/5496 | 0.2%      | 8.46    | 2.37–30.29 | 0.001 |
|          | c.444 + 1G > A                    | 1/165        | 0.6%     | 22/5496 | 0.4%      | 1.52    | 0.20–11.32 | 0.68 |
|          | del5395 (ex10-11del)              | 1/165        | 0.6%     | 24/5496 | 0.4%      | 1.39    | 0.19–10.34 | 0.75 |
| CHEK2    | truncating                        | 5/165        | 3.0%     | 58/5496 | 1.1%      | 2.93    | 1.16–7.40  | 0.02 |
|          | c.470C > T                        | 9/165        | 5.5%     | 264/5496| 4.8%      | 1.28    | 0.59–2.17  | 0.84 |
|          | Any CHEK2                         | 14/165       | 8.5%     | 321/5496| 5.8%      | 1.45    | 0.87–2.42  | 0.21 |
| NBN      | c.657_661delACAAA                 | 1/165        | 0.6%     | 22/4000 | 0.55%     | 1.10    | 0.15–8.23  | 0.92 |
reported a correlation between the c.470C > T mutation in the CHEK2 gene and multi-organ cancer susceptibility (breast, colon, kidney, prostate and thyroid cancer) with odds ratios ranging between 1.5 and 2.0 [65].

PALB2 and CHEK2 are clear breast cancer susceptibility genes. PALB2 encodes a BRCA2-binding protein that acts as a linker between BRCA1 and BRCA2 to form a BRCA1-associated genome surveillance complex. The complex is essential for the homologous DNA break repair [66]. Homozygous mutations of PALB2 cause Fanconi anemia, a rare recessive chromosomal breakage syndrome characterized by physical abnormalities, bone marrow failure and a high risk of malignancy. Heterozygous carriers of PALB2 mutations are at increased risk of breast and pancreatic cancers [67, 68].

CHEK2 is involved in the p53 pathway of DNA damage responses. CHEK2 interacts with many different proteins. Upon ionizing radiation-induced DNA damage, CHEK2 is activated by ataxia telangiectasia mutated (ATM) and is in turn capable of phosphorylating several substrates including Cdc25A, Cdc25C, p53, and BRCA1, leading to cell cycle arrest, apoptosis and DNA repair [69].

In contrast to our findings in female breast cancer patients, there were relatively few mutations found in BRCA1 and BRCA2. Several previous studies reported an association of BRCA1 and BRCA2 mutations with breast cancer risk in men [5–18]. In these studies, the frequency of detected mutations in the BRCA2 gene was higher than seen in the BRCA1 gene [14–16, 18, 21].

There are several limitations to this study. Patients were selected from a registry of breast cancer cases held at the Hereditary Cancer Center in Szczecin which could have over-represented patients with a positive history of breast and ovarian cancer. The molecular analysis was based on recurrent mutations which may underestimate the total mutation frequency.

Conclusions
Our study shows that mutations in the CHEK2 and PALB2 genes are important risk factors for male breast cancer in Poland. We recommend that a simple test for Polish founder mutations in BRCA1, BRCA2, PALB2 and CHEK2 should be offered to all male breast cancer patients in Poland.

Abbreviations
ASA-PCR: allele-specific amplification PCR; ATM: ataxia telangiectasia; BRCA1: breast cancer 1 gene; BRCA2: breast cancer 2 gene; Cdc25: M-phase inducer phosphatase 1; CHEK1: checkpoint kinase 1 gene; CHEK2: checkpoint kinase 2 gene; CI: confidence interval; ER: estrogen receptor; ERCC: human epidermal growth factor receptor 2; NBN: Nijmegen breakage syndrome gene; OR: odds ratio; PALB2: partner and localizer of BRCA2 gene; PCR: polymerase chain reaction; PR: progesteron receptor; RECQL: RecQ protein-like gene; RFLP-PCR: restriction fragment length polymorphism PCR

Supplementary Information
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Additional file 1.

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Authors’ contributions
M.S.Z. designed the study, analysed study data and drafted the manuscript. W.K., D.W., selected and prepared DNA samples for genotyping and performed genotyping. M.S.Z., J.T.-S.Z., C.C., T.H., R.S., M.W., J.G., K.O., H.G., enrolled patients and controls for the study, and collated phenotypic data for the study. K.O., and H.G., performed statistical analyses. S.A.N., J.T.-S.Z., J.G., and J.L. analysed study data, assisted in coordination of the study and in drafting the manuscript. T.H., C.C., R.S., M.W., conceived, designed and coordinated the study and assisted with drafting the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials
All data generated or analysed during this study are included in this published article [and its supplementary information files].

Declarations
Ethics approval and consent to participate
All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional research committee and with the 1964 Helsinki Declaration and its later amendments. The study was approved by the Ethics Committee of the Pomeranian Medical University in Szczecin (IRB BN-001/174/05). All individual participants included in the study provided written informed consent. A confidential ID number was assigned for further identification to each participant and to the corresponding data. Both hard and soft copy of the data kept in a safe place.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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