Novel insights into the role of immune cells in skin and inducible skin-associated lymphoid tissue (iSALT)

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Summary

The skin is equipped with serial barriers that provide rapid and efficient protection against external intruders. Beneath the epidermal physical barriers of the stratum corneum and the tight junctions, the integrated immune systems in both the epidermis and the dermis act in a coordinated manner to protect the host. This “immunological” barrier is composed of various cells, including skin-resident cells, such as keratinocytes, dendritic cells, tissue-resident macrophages, resident memory T cells, mast cells, and innate lymphoid cells. Additionally, infiltrating memory T cells, monocytes, neutrophils, basophils, and eosinophils are recruited in support of the host immunity.

In addition to discussing the role of each of these cellular populations, we describe the concept of skin-associated lymphoid tissue (SALT), which reminds us that the skin is an important component of the lymphatic system. We further describe the newly discovered phenomenon of multiple cell gathering under skin inflammation, which can be referred to as inducible SALT (iSALT). iSALT contributes to our understanding of SALT by highlighting the importance of direct cell-cell interaction in skin immunity.

Cite this as Ono S, Kabashima K. Novel insights into the role of immune cells in skin and inducible skin-associated lymphoid tissue (iSALT). Allergo J Int 2015; 24: 170–9
DOI: 10.1007/s40629-015-0065-1

Key words

iSALT – SALT – dermal dendritic cell – macrophage – clustering – T cell

Introduction

The skin is one of the largest organs in the human body and the primary interface with the external environment. Highly sophisticated protective systems have evolved in the skin to defend the body against external intruders. The protective components of mammalian skin include both physical and immunological barriers.

The stratum corneum (SC) and the tight junctions (TJs) work at the epidermal level as the most important “physical” barriers [1]. Beneath them, the integrated innate and acquired immune systems in both the epidermis and the dermis, which are activated in a coordinated manner to neutralize the intruders, are the most important “immunological” barriers. The immunological barrier consists of skin-resident cells, such as keratinocytes (KCs), dendritic cells (DCs), tissue-resident macrophages, mast cells, resident memory T cells, and innate lymphoid cells (ILCs). In addition, infiltrating memory T cells, monocytes, neutrophils, basophils, and eosinophils are recruited to the skin under inflammation to support and coordinate host immunity.

In this review, we present the current knowledge on immune cells in the skin, which play important roles to establish or diminish the skin inflammation. We also revisit the established concept of skin-associated lymphoid tissue (SALT) [2], which reminds us that the skin is not only a peripheral site of inflammation but also an essential component of the lymphatic system.

Finally, we introduce the recently proposed concept of inducible SALT (iSALT), which has yielded novel insights into the roles of dermal DCs (dDCs).
and macrophages in contact allergy by clarifying the details of cellular interactions in a murine contact hypersensitivity (CHS) model [3]. The concept of iSALT sheds light on the importance of the sequential cell–cell interactions among immune cells in situ for the establishment of skin inflammation.

**Epidermis**

**Keratinocytes**

The human epidermis consists of two major populations: KCs and Langerhans cells (LCs). It has gradually been discovered that KCs play important immunological roles in addition to their structural role [4]. KCs are able to sense various exogenous harmful pathogens and endogenous cellular damages through pattern recognition receptors (PRRs), which exist both on the cellular membranes and in the cytosol, to mediate immune responses. First, KCs express various Toll-like receptors (TLRs): TLR1, TLR2, TLR4, TLR5, and TLR6 on cell surfaces, and TLR3, TLR7, and TLR9 on cell surfaces, and TLR3, TLR7, and TLR9 in endosomes [5, 6, 7, 8, 9]. Pathogen-associated molecular patterns (PAMPs) of microbial origin, such as lipopolysaccharide (LPS), peptidoglycan, flagellin, and nucleic acids are sensed by some of these TLRs. Signals from these receptors facilitate the secretion of numerous cytokines, including interleukin (IL)-1 family (IL-1α, IL-1β, and IL-1β), IL-6, and tumor necrosis factor (TNF). Immune responses generated by different TLRs seem to be distinct, and numerous chemokines are secreted to attract various leukocytes depending on the type of inflammation. Activated KCs are also capable of conditioning LCs or DCs to promote predominant T helper (Th)1-, Th2-, or Th17-type immune responses through TLR signaling [10] or the secretion of different types of cytokines [11].

KCs also sense endogenous damage-associated molecular patterns (DAMPs), which are released from stressed host cells as a result of exogenous insults including infection, intoxications, and traumas. Adenosine triphosphate (ATP), heat shock pro-

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**Abbreviations**

| Abbreviation | Full Form |
|--------------|-----------|
| AD           | Atopic dermatitis |
| Ags          | Antigens |
| AMPs         | Anti-microbial peptides |
| APCs         | Antigen presenting cells |
| ASC          | Apoptosis-associated speck like protein containing a caspase recruitment domain |
| ATP          | Adenosine triphosphate |
| CCL5         | Chemokine C-C motif ligand 5 |
| CHS          | Contact hypersensitivity |
| CXCL2        | Chemokine C-X-C motif ligand 2 |
| CX3CR1       | C-X3-C chemokine receptor 1 |
| CXCR2        | Chemokine C-X-C motif receptor 2 |
| DAMPs        | Damage-associated molecular patterns |
| cDCs         | Dermal conventional DCs |
| dDCs         | Dermal DCs |
| DCs          | Dendritic cells |
| dLNs         | Draining lymph nodes |
| DN           | Double negative |
| dSEARCH      | Dendrite surveillance extension and retraction cycling habitude |
| DTA          | Diphtheria toxin subunit A |
| DTR          | Diphtheria toxin receptor |
| HMGB1        | High-mobility group box protein 1 |
| HSV-2        | Herpes simplex virus |
| IFN          | Interferon |
| IL           | Interleukin |
| ILCs         | Innate lymphoid cells |
| iSALT        | Inducible SALT |
| KCs          | Keratinocytes |
| LCs          | Langerhans cells |
| LPS          | Lipopolysaccharide |
| M1           | Classically activated macrophage |
| M2           | Alternatively activated macrophage |
| MALT         | Mucosa-associated lymphoid tissue |
| Mgl2         | Macrophage galactose C-type lectin type 2 |
| MHC          | Major histocompatibility complex |
| MLCs         | Memory lymphocyte clusters |
| NLR          | Nucleotide-binding domain and leucine-rich repeat containing family |
| PAMPs        | Pathogen-associated molecular patterns |
| pDCs         | Plasmacytoid dendritic cells |
| PRRs         | Pattern recognition receptors |
| SALT         | Skin-associated lymphoid tissue |
| SC           | Stratum corneum |
| Th           | T helper |
| TJs          | Tight junctions |
| TLRs         | Toll-like receptors |
| TNF          | Tumor necrosis factor |
| TSLP         | Thymic stromal lymphopoietin |
| XCR1         | XC-chemokine receptor 1 |
teins, hyaluronan, monosodium urate, galectins, adenosine, high-mobility group box protein 1 (HMGB1), IL-1α, and IL-33 have been identified as DAMPs [12]. DAMPs are sensed through a cytosolic inflammasome complex, which is formed by NLR (nucleotide-binding domain and leucine-rich repeat containing family), ASC (apoptosis-associated speck like protein containing a caspase recruitment domain), and pro-caspase-1 in the cytoplasm [13, 14]. Activation of the inflammasome complex leads to subsequent activation of proinflammatory cytokines such as IL-1β and IL-18 [13]. Although it remains unclarified which DAMPs are sensed by KCs, KCs are reported to express NLR and thus considered to sense these DAMPs [4, 15]. In addition to being a sensor, KCs can also be a source of DAMPs, such as ATP [16], HMGB1 [17], IL-1, and IL-33 during inflammation [18, 19].

Furthermore, KCs secrete anti-microbial peptides (AMPs), namely, β-defensins and cathelicidins. The secretion of these AMPs is increased in a Th17 cell-derived cytokine-dependent manner, with IL-17A and IL-22 having a particularly strong effect [20]. These peptides act to protect against infectious microbes by direct killing of the pathogens, recruitment of host immune cells, and modulation of cytokine production. AMPs are essential for host defense as demonstrated by sterile inflammation and decreased skin infections in psoriatic skin lesions, wherein high expression of AMPs is noted, regardless of a dysfunctional epidermal barrier [21–23]. Likewise, decreased AMPs and increased susceptibility to infection in atopic dermatitis (AD) patients has been indicated [24]. On top of its role in host defense, AMPs seem to be involved in the pathogenesis of psoriasis, because cathelicidin activates plasmacytoid DCs (pDCs), an important chaperon in psoriasis, after being coupled with self-DNA [25].

**Langerhans cells**

LCs are highly specialized antigen presenting cells (APCs) that account for approximately 2–4% of the total epidermal cell population [26]. They are characterized as the intraepithelial subpopulation of DCs, and their functions are distinct from those of dermal DCs.

Architecturally, LCs form dense cellular networks in the basal and suprabasal layers of the epidermis. Various surface markers characterize human and murine LCs (Tab. 1). Langerin/CD207 is the lectin receptor that binds mannose and related sugars found on a variety of microorganisms. CD205 is the essential molecule for antigen capturing and processing. Human LCs also highly express CD1a, which is capable of presenting microbial lipid antigen to T cells [27].

After sensing inflammatory signals, LCs undergo subsequent morphological and physiological changes: they increase in size and begin a motion of the dendritic processes known as dendrite surveillance extension and retraction cycling habitude (dSEARCH) [28]. Importantly, a recent report has demonstrated that LCs elongate their dendrites to penetrates the TJs to capture external antigens (Ags) that have violated the SC barrier [29]. After capturing Ags, LCs undergo maturation and then migrate from the epidermis to the draining lymph nodes (dLNs). Therein, LCs present the Ags to T cells, enabling epicutaneous sensitization of protein Ags.

The importance of LCs in the sensitization of protein Ags has been exhibited in an ovalbumin-induced murine AD model, in which LC deficiency reduced clinical symptoms and Ag-specific IgE production [30]. These findings support the idea that LCs are primarily responsible for capturing protein Ags, which cannot penetrate the physical epidermal barriers.

The role of LCs in the CHS reaction is a matter of debate. First, LC-deficient murine model of transgenic Langerin-DTA (diphtheria toxin subunit A) exhibited enhanced CHS and proposed a regulatory rather than a stimulatory role of LCs in the sensitization phase against hapten, although there remained the possibility that constitutive LC deficiency might result in abnormal compensatory effect of T cells in the model [31, 32]. With an additional observation of the Langerin-specific IL-10 deletion model, the authors concluded that the suppressive effect of LCs requires their IL-10 production for inducing CD4+ regulatory T (Treg) cells [34]. Because IL-10 from Langerin+ DC is also likely to have a similar suppressive effect in this model, further investigation is expected. The protective role of LCs in CHS by tolerizing CD8 T cells and activating Treg cells has also been demonstrated [34]. Besides, the observations of inducible LC-specific ablation strategy using Langerin-DTR (diphtheria toxin receptor)-knocked-in mice and of mice lacking LCs due to independent molecular defects show that LCs and dDCs seem to have a functional redundancy [35, 36, 37, 38, 39]. Accumulating literatures support the concept that rather the hapten dose determines the requirement for LCs for efficient T cell priming, and the number of skin DCs, but not the type of DCs, regulates the strength of the CHS reaction [31, 36, 40].

Studies of other skin inflammation models, such as cutaneous leishmaniasis, indicate a tolerogenic function of LCs in vivo [41]. Likewise, in the absence of antigen presentation by dDCs, antigen presentation to CD4 T cells by LCs alone is unable to mediate T cell differentiation in vivo [42, 43]. In addition, a distinct function of LCs from dDCs has
been indicated from the recent studies of the skin *Candida albicans* infection model. Ag presentation by LCs is essential for the generation of Ag-specific Th17 cells, whereas Langerin⁺ dDCs are required for the generation of Ag-specific cytotoxic T cells and Th1 cells [44]. Intriguingly, the morphology of *Candida albicans* seems to be important for driving such distinct T cell responses [45].

**Dermis**

**Dermal dendritic cells**

Some foreign Ags, such as hapten, are small enough to penetrate the dermis across the TJ.s. When epidermal physical barriers are disrupted by cutaneous inflammation or scarring, larger molecules also become capable of entering the dermis directly. dDCs efficiently capture and present these Ags to initiate acquired immunity.

From the recent literatures [46, 47, 48], besides pDCs that are absent from the skin in the steady state, at least five DC populations are identified in the murine dermis: LCs in transit, monocyte-derived DCs that originate from Ly6C⁺ blood monocytes, and dermal conventional DCs (cDCs) that originate from blood-borne precursors. cDCs consist of XCR1⁺ cDCs, CD11b⁺ cDCs, and XCR1⁻CD11b⁻ double negative (DN) cDCs [49, 50, 51]. In detail, XCR1⁺ cDCs are composed of CD103⁺ cDCs and CD103⁻ cDCs. XCR1⁻ cDCs are the only subset to express high levels of Langerin/CD207 among dDCs, and are often called as Langerin⁺ dDCs. LCs and CD11b⁺ cDCs are also shown to express CD301b, also known as macrophage galactose C-type lectin type 2 (Mgl2) [45, 52]. Other several different surface markers are used to distinguish each population (Tab. 1). Similar DC subsets are also noticed in the human skin.

Compared to other dDC subsets, CD103⁺ cDCs are highly efficient at cross-presenting Ags to naive CD8⁺ T cells, and likely promote Th1 immune responses [44, 53, 54]. As mentioned above, Langerin⁺ dDCs are shown to induce Th1 cells during cutaneous *Candida albicans* infection [44]. On the other hand, in the sensitization phase of CHS, various literatures indicate the importance and redundancy of different DC types, including LCs, Langerin⁺ dDCs, and CD11b⁺ cDCs [45, 52, 55, 56]. The hapten dose and the number of DCs seem to be important, as discussed in the chapter of LCs.

The importance of CD11b⁺ dDC in the generation of Th2 immune responses has also been suggested [52, 57]. The function and characteristics of DN cDCs remain uninvestigated.

Which T cell responses (Th1-, Th2-, and/or Th17-type) are promoted in dLNs is considered to depend not only on dDC types, but also on the local skin conditions, under which dDCs are activated. For example, in AD, thymic stromal lymphopoietin (TSLP) is massively expressed from the epidermis in the presence of mechanical injury [58]. TSLP receptor signaling through LCs and dDCs activates and upregulates OX40L on those cells, leading to subsequent skewing into Th2-type responses [59]. Likewise, IL-33 is one of the IL-1 family members derived from KCs, and it also mediates Th2-type responses [19].

Psoriasis was classically defined as Th1- and Th17-type inflammation. KC-derived IL-1β, IL-6, and TNF-α and pDC-derived IFN-α are thought to activate local dDCs, leading to IL-23 secretion to prime naive T cells into Th1 and/or Th17 sets in dLNs [4, 60]. Recently, the significance of IL-23/IL-17/IL-22 axis in psoriasis has become more evident from genome wide studies, the efficacy of biologics targeting these cytokines, and reduction in these cytokines after disease amelioration [61, 62, 63]. It has attracted the attention again which DC subsets play a pivotal role in producing IL-23 and in

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**Tab. 1: Surface markers of Langerhans cells (LCs) and dermal dendritic cells (dDCs)**

| Occupation | Murine surface markers | Human surface markers |
|------------|-------------------------|-----------------------|
| **LCs**    |                         |                       |
| 2–4 % of epidermal cells | CD45⁺, MHC class II⁺, CD11c⁺, T/B/NK cell markers⁺ | CD45⁺, MHC class II⁺, CD11c⁺, T/B/NK cell markers⁺ |
| Monocyte derived dDCs | no data | E-cadherin⁺, EpCAM⁺, Langerin/CD207⁺, Birbeck granule⁺ |
| CD11b⁺ cDCs | 50–70 % of dDC pool | CD11b⁺, CD301b⁺, CCR2⁺, CD172a⁺, CXCR1mid⁰⁵⁰⁰⁰ |
| XCR1⁺ cDCs | 10–20 % of dDC pool | XCR1⁺, Langerin/CD207⁺, CD205⁺, CD11b⁺, CD301b⁺, CD172a⁺, CX3CR1⁻ |
| DN cDCs | no data | CD11b⁻, CX(CR1)⁻, CCR2⁺, CD172a⁺ |
Macrophages

Macrophages are unique cells that have a strong phagocytic function for the clearance of unnecessary wastes, such as apoptotic cells, cellular debris, and pathogens. In addition to the clearance of waste disposal, they facilitate the induction of inflammation by Ag recognition via a wide array of receptors, and thus participate in innate and acquired immunity [68, 69, 70]. Moreover, they produce growth factors that stimulate capillary growth, collagen synthesis, and fibrosis, all of which are related to tissue repair. Thus, they play a fundamental role as the frontline of tissue defense and the organizer of tissue homeostasis.

Macrophages are functionally grouped into two classes: M1 (classically activated) and M2 (alternatively activated) [71, 72], according to their role and the cytokines needed for their activation. M1 and M2 statuses are defined by a cell’s response to interferon (IFN)-γ and TLR signaling and to IL-4 and IL-13, respectively. M1-type macrophages produce proinflammatory cytokines including IL-1 and destroy intracellular pathogens by means of an increased oxidative burst and nitric oxide production. Although the role of M2-type macrophages has been less well characterized, several functions have been noted, including involvement in protection against parasitic infections, promoting Th2 immune responses, damping excessive inflammation, tumor progression, angiogenesis, wound healing, tissue remodeling, and fibrosis [73, 74, 75].

In mice, lymphocyte antigen 6C (Ly6C)-positive inflammatory monocytes, which express high amounts of chemokine receptor type 2 (CCR2) but low amounts of C-X3-C chemokine receptor 1 (CX3CR1), enters from the circulation to the tissue and takes on the macrophage phenotype during inflammation. They are believed to act as M1-type macrophages [75]. On the other hand, Ly6C-negative macrophages, which express high amounts of CX-3CR1 but low amounts of CCR2, are considered as M2-type macrophages. Likewise, CD14 CD16+ monocytes are classified as M1-type and CD14+CD16+ macrophages as M2-type in humans [76, 77]. Tissue-resident macrophages are classified as M2-type in general [72]. Phenotype transition from M1-type into M2-type has been reported under the stimulation by basophil-derived IL-4 in the late phase of chronic allergic inflammation of the skin [77]. Although the different roles of macrophages derived from inflammatory monocytes and tissue-resident macrophages have not yet been accurately determined, each subset is believed to act in a coordinated manner in both inflammation and tissue-repair phases.

T cells

T cells in their naive state first encounter APCs in secondary lymphoid organs. They are primed through Ag presentation by DCs and then proliferate and differentiate to adopt a memory phenotype. Non-lymphoid tissues, including the skin, serve as sites for such memory T cell infiltration because they tightly regulate the expression of adhesion molecules and chemokine receptors for memory T cell homing [80]. Memory T cells migrate into the peripheral tissue robustly under active inflammation; however, memory T cells also enter the tissue to scan for Ags even in the absence of inflammatory stimuli [81, 82].

The skin infiltrating CD4 T cells consist of Th1, Th2, Th17, Th22, Th9, and Treg cell subsets. It is generally believed that Th differentiation fate of each CD4 T cell is determined by the nature of the APCs that prime naïve T cells in the dLNs, as discussed in the chapters on LCs and dDCs.

The essential role of Th1 cells is to protect against intracellular pathogens and viruses by secreting IFN-γ. They dominate in the skin during the early phase of contact dermatitis [83]. Th2 cells play a central role in allergic disorders such as AD by regulating both acute- and late-phase allergic reactions, mediated by immunoglobulin E and eosinophils, re-
spectively. They function by secreting IL-4, IL-5, IL-13, IL-24, IL-25, and IL-31. Full activation of Th2 cells is achieved through the cytokine production by KCs, such as IL-33 and TSLP, and by subsequent activation of group 2 ILCs to produce IL-5, IL-9, and IL-13 [84]. Skin Th17 cells protect against bacteria and fungi, such as Staphylococcus aureus, Klebsiella pneumoniae, and Candida albicans, by controlling neutrophils at the mucocutaneous surfaces [45, 85]. Th17 and Th22 cells are well known for their role in psoriasis. In psoriasis, KC-derived IL-1β, IL-6, and TNF-α and plasmacytid DC-derived IFN-α lead to Th17 cell-mediated inflammation. In turn, Th17 cells secrete IL-17A, IL-17F, and IL-22, which stimulate proliferation and abnormal differentiation of KCs.

CD8 T cells are the principal effector cells that recognize Ags on major histocompatibility complex (MHC) class I molecules. They are considered to be involved in various skin disorders [82] including contact dermatitis, psoriasis, graft versus host disease, vitiligo, alopecia areata [86], and drug eruptions [87]. After being primed by DCs, CD8 T cells circulate around the tissue having assumed a cytotoxic memory phenotype. They possess a strong capacity for eradicating Ag-bearing cells, such as virally infected cells, tumor cells, and grafted allogeneic cells, upon re-exposure. Recent discoveries concerning the important functions of dDCs and macrophages in memory CD8 T cell activation in the skin will be discussed later in the chapter on iSALT.

In a steady state, skin CD4 and CD8 T cells are essentially resident memory T cells, which mainly reside in the dermal perivascular area and the epidermis, respectively [88, 89]. Resident memory T cells comprise a functionally distinct, non-migratory population and persist long-term in peripheral tissues to provide effective protection against local Ag re-challenge. Although the mechanisms how the memory T cell subset resides, and is maintained and re-activated in the peripheral tissue are not yet fully determined, they are considered to be derived from precursors that enter the tissue during the effector phase of immune responses [90]. The essential role of skin-resident memory T cells in local immunity remains an important issue to be clarified.

SALT and iSALT: role of dendritic cells and macrophages

In humans and rodents, lymphoid structures that consist of T and B cell areas can be observed in specialized submucosal areas. These structures are known as “mucosa-associated lymphoid tissue (MALT)” [9]. In MALT, the prompt and efficient antigen presentation to T cells and B cell class switching occur independently of secondary lymphoid organs.

In the 1980s, cutaneous immunologists introduced the term “skin-associated lymphoid tissue (SALT)” based on multiple studies that revealed the existence of T cells and DCs in the skin under inflammatory conditions [2, 92, 93]. In the classical concept of SALT, it is supposed that antigen presentation to naive T cells occurs in the skin as it does in secondary lymphoid tissues or in MALT. However, in SALT, it remains unclear whether and how T cells are activated in the skin in situ. In addition, no distinct lymphoid structures have been observed in the skin in a steady state or even under inflammatory conditions, such as contact dermatitis and AD, in contrast to MALT. Therefore, SALT has remained conceptual for more than 30 years.

Recently, dDC clustering (Fig. 1a) around the post-capillary venules has been discovered in the elicitation phase of the CHS reaction [3] in a murine model for contact dermatitis. In this clustering, dDCs contact the effector T cells, leading to their activation. Both KCs and perivascular macrophages seem to play essential roles in this phenomenon, given that dDC clustering and effector T cell activation were abrogated in the absence of IL-1 signaling from KCs and macrophage-depleted mice. In addition, Chemokine (C-X-C motif) ligand 2 (CXCL2) was the major chemokine expressed by macrophages under stimulation with IL-1. Blocking of chemokine (C-X-C motif) receptor 2 (CXCR2), a CXCL2 receptor, also inhibited dDC clustering. Therefore, an inducible structure formed of perivascular macrophages, dDCs, and effector T cells is identified. Because the formation of this structure is essential for efficient activation of effector T cells, these inducible leukocyte clusters may function as SALTs. This structure was named “inducible SALT (iSALT)” (Fig. 1b) in analogy with inducible bronchus-associated lymphoid tissue (iBALT), which is involved in the respiratory immunity [94]. This clustering of dDCs and T cells was also observed in the skin of a contact dermatitis patient, demonstrating that iSALT forms in humans as well.

There are some major issues to be clarified with regard to iSALT. It should be determined whether iSALT enables naive T cell priming or B cell class switching and antibody production in situ, as in other lymphoid-associated tissues.

It should also be investigated whether iSALT or iSALT-like structures are established by other cells, or in other inflammatory settings, such as infection, psoriasis and AD. The essential roles of various skin-associated cells, including KCs, DCs, macrophages, T cells, ILCs, mast cells, basophils, and eosinophils, in either establishment or cessation of cutaneous immune reactions have been revealed. Therefore, it is possible that other T cell subsets or ILCs utilize such structures for their activation in situ.

Further supporting this idea, the essential role of local macrophages for resident memory CD4 T cell maintenance and activation against herpes simplex
virus (HSV-2) in the genital mucosa has been reported [95]. Within the memory lymphocyte clusters (MLCs), secretion of basal levels of IFN-γ from resident memory CD4 T cells activates macrophages, and secretion of chemokine (C-C motif) ligand 5 (CCL5) from macrophages sustains resident memory CD4 T cells in turn. Upon Ag re-challenge, these T cells are activated to secrete high levels of IFN-γ. Even though the site of activation (the skin vs. the genital mucosa) and the activated cell types (CD8 vs. CD4 T cells; recruited vs. resident) are different between iSALT and MLCs, the two cluster-based structures are similar in several ways, and both emphasize the importance of local cell-cell interaction in acquired immune response.

Besides, DC-specific IL-10 receptor deficient mice failed to prevent exaggerated activation of memory T cells in the elicitation phase of CHS, which implicates the critical role of IL-10 control of DCs during T cell re-activation (= challenge) in the skin during CHS. This may also be an example of the importance of cellular crosstalk between Treg cells and DCs [96] in situ for the cessation of cutaneous immune reactions. It would be interesting to pursue whether Tregs are engaged in iSALT.

In addition, mature DC and T cell clustering has been proposed in psoriasis [97, 98], although its role in the pathogenesis of psoriasis remains uninvestigated. It is possible that such DC-T cell interaction in the skin is essential for memory Th1 or Th17 cell activation in psoriasis.

Moreover, it is of note that both basophils and ILC2 cells are reported to accumulate in the skin lesions of AD patients and a murine AD model, and that these two cells types tend toward locational proximity to each other [99]. This report shows that IL-4 produced by basophils is crucial for ILC2 cell proliferation at the site, suggesting the cross-regulation of ILC2 cells by basophils. Because basophils can act as APCs, the inducible basophil-ILC2 clusters in the skin suggest the possibility of another iSALT-like structure in the murine AD model.

Furthermore, distinct lymphoid structures other than iSALT have been reported under particular circumstances, such as cutaneous lupus erythematosus [100, 101], keloids (the aberrant overgrowth of scar tissue that occur at the skin injury site) [102], and pseudolymphoma [103, 104], although the underlying mechanisms remain to be clarified. The functional and pathological differences between iSALT and other skin lymphoid structures should be determined in the future.

**Conclusion**

Our understanding of the immunological role of each cellular subset in the skin and of their interactions has progressed steadily. Because each pathological condition involves various immune cell reactions and interactions, we should be careful to discriminate the immune reactions that occur in the peripheral organs from those that occur in the secondary lymphoid organs. This will help us to un-
understand the common and characteristic roles of each organ, including the skin. Much remains to [105] be clarified in the skin-resident immune network, especially with regard to iSALT.

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Conflicts of interest
The authors declare that there are no conflicts of interest.

Cite this as
Ono S, Kabashima K. Novel insights into the role of immune cells in skin and indoluble skin-associated lymphoid tissue (iSALT). Allergo J Int 2015; 24: 170–9

DOI: 10.1007/s40629-015-0065-1

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