Proteomic analysis of stage I primary lung adenocarcinoma aimed at individualisation of postoperative therapy

J Maeda1, T Hirano*,1, A Ogiwara2,3, S Akimoto2,3, T Kawakami2, Y Fukui4, T Oka4, Y Gong1, R Guo1, H Inada1, K Nawa1, M Kojika1, Y Suga1, T Ohira1, K Mukai1 and H Kato1

1Department of Surgery, Tokyo Medical University, 6-7-1 Nishi-shinjuku, Shinjuku-ku, Tokyo 160-0023, Japan; 2Clinical Proteome Center, Tokyo Medical University, 2-6-1 Nishi-shinjuku, Shinjuku-ku, Tokyo 163-0217, Japan; 3Medical ProteoScope Co. Ltd, 2-6-1 Nishi-shinjuku, Shinjuku-ku, Tokyo 163-0217, Japan; 4Taiho Pharmaceutical Co. Ltd, Tokyo, Japan; 5Department of Pathology, Tokyo Medical University, 6-7-1 Nishi-shinjuku, Shinjuku-ku, Tokyo 160-0023, Japan

Although postoperative adjuvant chemotherapy (PAC) with uracil–tegafur significantly improves the prognosis of patients with stage I lung adenocarcinoma, subset analysis has revealed that only 11.5% of patients with stage IB derive actual benefit from such therapy. Therefore, it is extremely important to identify patients for whom adjuvant chemotherapy will be beneficial. We performed comprehensive protein analysis of 24 surgically resected specimens of stage I adenocarcinoma using liquid chromatography-tandem mass spectrometry (LC-MS/MS), followed by bioinformatical investigations to identify protein molecules. Furthermore, we carried out immunohistochemical studies of 90 adenocarcinoma specimens to validate the results of LC-MS/MS. We detected two kinds of protein molecules (myosin IIA and vimentin) by LC-MS/MS. We confirmed their immunohistochemical expression and distribution, and evaluated the relationship between the expression of these proteins and prognosis after adjuvant chemotherapy. Patients with no expression of either myosin IIA or vimentin showed a significantly better outcome regardless of PAC using uracil–tegafur. However, we were unable to select responders to uracil–tegafur using these proteins. Cases of adenocarcinoma lacking expression of either myosin IIA or vimentin show a good outcome without PAC, and therefore do not require such treatment.

British Journal of Cancer (2008) 98, 596–603. doi:10.1038/sj.bjc.6604197 www.bjcancer.com

© 2008 Cancer Research UK

Keywords: myosin IIA; vimentin; postoperative adjuvant chemotherapy; responder to uracil–tegafur; stage I lung adenocarcinoma

Death due to lung cancer is still increasing in most industrialised countries, including Japan, despite improvement of various diagnostic and therapeutic modalities. Even though the opportunities to detect lung cancer at an early stage are increasing, approximately 60 000 patients with lung cancer die every year in Japan, usually due to distant metastasis. Distant metastasis, including intrapulmonary metastasis, frequently occurs in patients with advanced-stage non-small cell lung cancer (NSCLC) who undergo only surgical resection, because in such cases micrometastases probably exist at the time of surgery. The concept of postoperative adjuvant chemotherapy (PAC) for control of micrometastasis does not conflict with the improved prognosis of NSCLC patients. However, the efficacy of PAC in patients after complete resection of NSCLC was a matter of controversy in the 1990s. Even as recently as 2003, the efficacy of PAC could not be demonstrated (Scagliotti et al, 2003). In 2004, however, some studies demonstrate a beneficial effect of PAC (Arriagada et al, 2004; Strauss et al, 2004; Winton et al, 2005). We have also reported that PAC with oral uracil–tegafur (DPD Inhibitory Fluoropyrimidine, Taiho Pharmaceutical Co. Ltd, Tokyo, Japan) provided better survival than surgical treatment alone in patients with stage I adenocarcinoma of the lung (Kato et al, 2004). The combination of uracil and tegafur (also referred to as UFT) at a molar ratio of 4:1 is an oral anticancer agent with good absorption in the small intestine (Fuji et al, 1979). Tegafur is a prodrug that is gradually converted into fluorouracil in the liver by the cytochrome P-450 enzyme system. Uracil enhances the serum concentration of fluorouracil by competitive inhibition of dihydropyrimidine dehydrogenase, the enzyme responsible for fluoro-uracil catabolism (Ikenaka et al, 1979). Oral uracil–tegafur generates a higher maximal plasma level of fluorouracil than intravenously infused fluorouracil at a dose that is equimolar to the amount of tegafur in uracil–tegafur (Ho et al, 1998).

Even though PAC with uracil–tegafur has significantly improved the prognosis of patients with stage I primary lung adenocarcinoma, even subset analysis of stage IB has revealed that 11.5% of patients actually derive some benefit from the treatment (Kato et al, 2004). Nonresponders to uracil–tegafur, including relapse-free patients without any adjuvant therapy, gain no benefit from PAC. In this context, it is important to establish biomarkers for prediction of responders to uracil–tegafur, and/or for favourable prognosis without the use of PAC.

Clarification of the entire human genome is one of the most significant events in the history of bioscience, and has accelerated the comprehensive analysis of human genes and their protein
proteins. Many biologists recognise the importance of protein analysis, because proteins play central role various cellular functions. However, in cancer research, there has been a tendency for most researchers to avoid investigation of cancer-related proteins, because their structures are more complicated than those of the genes. Nevertheless, techniques for the comprehensive analysis of proteins have improved greatly in recent years. The concept of comprehensive protein analysis has been established, and the new research field of proteomics has been developed. One of the main purposes of clinical proteomics in the field of oncology is the development of new therapeutic strategies for cancer, centred on individualised therapy. In this study, we attempted to identify biomarkers for the selection of responders to uracil–tegafur and nonresponders, including relapse-free patients without any requirement for PAC, using clinical proteomics methodology.

MATERIALS AND METHODS

Institutional review board approval for this investigation

The institutional review board approved the use of proteomics analysis to explore biomarkers for selection of responders to oral uracil–tegafur (294/323/480/702).

Surgical samples of stage I lung adenocarcinoma for mass spectrometry

After obtaining written informed consent, lung cancer tissues were obtained from patients with pathologically confirmed stage I adenocarcinoma resected at Tokyo Medical University Hospital between 1995 and 2001. Tissues were kept frozen at −80 °C until use. We collected 11 lung adenocarcinoma specimens from 11 patients who subsequently underwent PAC using uracil–tegafur for more than 2 years. In 5 of these 11 patients, recurrent lesions were detected within 2 years after surgery (U1R1), and the remaining 6 were confirmed to be disease-free for 5 years after surgery (U1R0). Furthermore, 13 specimens of lung cancer were collected from patients receiving no adjuvant therapy after surgery. In 6 of these 13 patients, recurrence was recognised within 2 years after surgery (U1R1), but no recurrent lesions were detected during 5 years after surgery in the other 7 (U0R0).

Protein extraction

The surgically resected materials were suspended and homogenised in PBS supplemented with a protease inhibitor cocktail (Roche Diagnostics Inc., Basel, Switzerland) at 4 °C. The cell lysate was then fractionated by ultracentrifugation (52 000 g, 4 °C, 20 min). The resulting pellet containing plasma membranes from the cells, was solubilised in PBS containing 5% SDS with continuous ultrasonication. The resulting solution was taken as the insoluble fraction, whereas the supernatant from the ultracentrifugation, containing mainly cytosolic proteins, was taken as the soluble fraction. The total protein concentrations of both fractions were measured (Lowry et al, 1951) using bovine serum albumin as a standard.

Protein condensation with SDS-PAGE

We added 150 pmol egg white lysozyme (Sigma-Aldrich Inc., St Louis, MO, USA) to an aliquot containing 75 μg protein from each fraction, then dried it under vacuum. The mixture was then solubilised in sample buffer (Laemmli, 1970) with gentle stirring at 37 °C for 1 h. A two-third volume of the solution containing 50 μg protein and 100 pmol lysozyme was subjected to SDS-PAGE on 12.5% polyacrylamide gel 1-mm-thick. SDS-PAGE was carried out at a constant current of 20 A until the bromophenol blue marker passed the boundary between the stacking and separation gels. In this ‘halfway’ running, most proteins remained stacked in a small area of approximately 2 mm in height between the gel boundary and the blue marker. After electrophoresis, this small gel area was excised from the gel slab, and the proteins were fixed in the gel slice with an excess volume of aqueous solution containing 40% methanol and 10% acetic acid.

In-gel tryptic digestion of protein

The gel slice was subjected to an in-gel tryptic digestion process (Shevchenko et al, 1996), with minor modifications. Briefly, after S-carboxyamidomethylation of Cys residues with iodoacetamide, the gel slice was incubated in a small volume of 50 mM ammonium bicarbonate buffer solution containing 1 μg of trypsin (Promega Co., Madison, WI, USA). The resulting peptides were extracted from the gel matrix, and dried under vacuum.

Liquid chromatography-tandem mass spectrometry

The peptide mixture (1 μg) was analysed using a liquid chromatography-tandem mass spectrometry (LC-MS/MS) system in a fully automated manner (Kawakami et al, 2005). Briefly, reversed-phase peptide separation was performed on a C18 capillary LC column (Michrom BioResources Inc., Auburn, CA, USA) at a flow rate of 1 μl/min. For gasification of the protonated peptides, the LC effluent was directly interfaced with an electrospray ionisation (ESI) source in a positive ion mode modified on a Finnigan LTQ linear ion trap mass spectrometer (Thermo Fisher Scientific Inc., Waltham, MA, USA) (Schwartz et al, 2002). The ESI used a Teflon-coated spray needle (20 μm id, AMR Inc., Tokyo, Japan). The ESI-MS/MS operation and continuous data acquisition of full MS scan and subsequent three MS/MS scans were carried out on an Xcalibur system controller (Thermo Fisher Scientific).

Semiquantitative analysis

All full MS data were investigated using an i-OPAL semiquantitative LC-MS data analysis system (i-OPAL algorithm: Patent no. WO 2004/090526 Al). First, the signal intensity of the full MS scan was normalised so that the total signal intensity of each sample became the same value. Several standard signals derived either from the injected egg white lysozyme or from sample intrinsic common proteins were selected as i-OPAL alignment markers. The i-OPAL alignment programme was used to align the nonlinearly fluctuating LC retention time axis of all LC-MS data to finally generate a single combined LC-MS data set for the soluble and the insoluble fractions, respectively. Analysis of variance (ANOVA) was applied for each peak signal in the final combined LC-MS data set to select candidate marker signals whose intensity differed significantly in a particular patient group. ANOVA was carried out using a Spotfire DecisionSite package.

Database searches

All MS/MS data were investigated using the Mascot search engine (Matrix Science Ltd., London, UK, http://www.matrixscience.com) against the Homo sapiens (human) subset of the Swiss-Prot and the RefSeq protein sequence databases. The database searches were performed allowing for fixed modification of cysteine residues (S-carbamidomethylation, +57.0 Da) and variable modification of methionine residues (oxidation, +16.0 Da), peptide mass tolerance ±2.0 Da and fragment m/z tolerance ±0.8.

Surgical specimens of stage I lung adenocarcinoma for immunohistochemical staining

Sample set A for confirmation of LC-MS semiquantitative results To confirm the semiquantitative results of LC-MS, 23
formalin-fixed, paraffin-embedded specimens derived from the same cases as those used for LC-MS analysis were collected for immunohistochemical investigation. As one formalin-fixed specimen had already been exhausted for the previous investigations, the remaining 23 specimens were investigated.

**Sample set B for validation** To validate the expression of the protein molecules on lung adenocarcinoma cells, 90 formalin-fixed, paraffin-embedded specimens from patients with lung adenocarcinoma, resected at Tokyo Medical University Hospital between 1995 and 2001, were used. All the patients had undergone curative resection of lung cancer, and after surgery, a pathologically definitive diagnosis of stage I adenocarcinoma had been obtained. We evaluated recurrence after surgery using chest roentgenography and serum tumour markers (CEA, CA19-9 and SLX) every 3 months and computed tomography of the head and body, and bone scintigraphy, every 6 months. When it was difficult to evaluate roentgenographically whether the lesion was recurrent or not, either cytological or pathological examinations were performed to obtain a definitive diagnosis (Table 1).

Of the 90 patients, 51 underwent PAC using uracil-tegafur. These 51 cases included 24 recurrences (U1R1) within 5 years and 27 cases without recurrence (U1R0) within 5 years after surgical treatment. The remaining 39 patients did not receive any adjuvant chemotherapy. These 39 patients included 17 with recurrence (U0R1) and 22 without recurrence (U0R0). The clinicopathological backgrounds of the 90 patients with lung adenocarcinoma are summarised in Table 2.

**Immunohistochemical staining of surgically resected specimens of stage I lung adenocarcinoma**

Four-micrometer-thick tissue sections were prepared from formalin-fixed, paraffin-embedded surgical specimens and collected on glass slides. The sections were stained immunohistochemically by the ABC method using either anti-myosin IIA mouse monoclonal antibody (clone ab24762, abcam, Cambridge, CB4 0FW, UK) (diluted 1:500) or anti-vimentin antibody (Dako monoclonal antibody (clone ab24762, abcam, Cambridge, CB4). The specimens were reacted with 0.06% 3,3’-diaminobenzidine tetrahydrochloride and 0.03% hydrogen peroxide in Tris-buffered saline to visualise the positive areas. Meyer’s haematoxylin was used for counterstaining.

**Evaluation of immunohistochemically stained preparations**

Cells showing cytoplasmic staining were evaluated as positive. For myosin IIA immunohistochemical staining, we evaluated a case as positive when more than 50% of the cells were stained. Also, for vimentin immunostaining, cases in which more than 25% of the cells were stained were evaluated as positive. For both kinds of staining, normal alveolar epithelium served as an internal negative control.

**Statistical analysis**

Statistical analysis was carried out using the SPSS program. Statistical significance of the relationship between recurrence and immunohistochemical reactivity was evaluated using χ² test. Disease-free survival curves were calculated from the day of surgery using the Kaplan–Meier method, and the significance of differences in survival rates between the patient groups was calculated by the log-rank test. In all statistical analyses, a P-value of <0.05 was taken to indicate a statistically significant difference.

**RESULTS**

**LC-MS data analysis**

After i-OPAL alignment and peak detection, we obtained 13136 signal peaks from the soluble fraction and 14984 peaks from the insoluble fraction. Using Spotfire, we restricted the candidate signal peaks on the basis of the following conditions:

1. A Mascot search result with a score equal to or more than 50.
2. An ANOVA P-value equal to or less than $1 \times 10^{-5}$ (for the soluble fraction) or $1 \times 10^{-6}$ (for the insoluble fraction).

**Table 1** Clinical features of lung adenocarcinoma cases subjected to LC-MS/MS

| Characteristic | (%) |
|---------------|-----|
| Age (year)    |     |
| Median        | 65.0 |
| Range         | 32–78 |
| Gender        |     |
| Male          | 51 cases (56.7%) |
| Female        | 49 cases (43.3%) |
| Pathological stage |     |
| IA            | 41 cases (45.6%) |
| IB            | 49 cases (54.4%) |
| Presence of recurrence |     |
| (+)           | 11 cases (45.8%) |
| (−)           | 13 cases (54.2%) |
| PAC           |     |
| (+)           | 11 cases (45.8%) |
| (−)           | 13 cases (54.2%) |

**Table 2** Clinical features of lung adenocarcinoma cases as revealed by immunohistochemical staining

| Characteristic | (%) |
|---------------|-----|
| Age (year)    |     |
| Median        | 64.8 |
| Range         | 45–82 |
| Gender        |     |
| Male          | 54 cases (60.0%) |
| Female        | 36 cases (40.0%) |
| Pathological stage |     |
| IA            | 33 cases (36.7%) |
| IB            | 57 cases (63.3%) |
| Existence of recurrence |     |
| (+)           | 41 cases (45.6%) |
| (−)           | 49 cases (54.4%) |
| PAC with urasil-tegafur |     |
| (+)           | 51 cases (56.7%) |
| (−)           | 39 cases (43.3%) |

PAC = postoperative adjuvant chemotherapy with oral uracil–tegafur.
As the peptide compositions of the soluble and the insoluble fractions differed, we applied different criteria to obtain approximately the same number of candidate signals. As a result, we were able to restrict the number of candidate signals to 23 and 28 for the soluble and insoluble fractions, respectively. From the restricted candidate signals, we selected several myosin IIA and vimentin signals as final candidate biomarker signals, because these two candidate biomarkers were identified by more than one distinct peptide sequence, and almost all of these signals had similar patterns of intensity (Figure 1A and B; Table 3).

Table 3 lists the amino-acid sequences from the selected candidate biomarker signals described above. These sequences were identified from MS/MS data using Mascot software.

As the peptide compositions of the soluble and the insoluble fractions differed, we applied different criteria to obtain approximately the same number of candidate signals. As a result, we were able to restrict the number of candidate signals to 23 and 28 for the soluble and insoluble fractions, respectively. From the restricted candidate signals, we selected several myosin IIA and vimentin signals as final candidate biomarker signals, because these two candidate biomarkers were identified by more than one distinct peptide sequence, and almost all of these signals had similar patterns of intensity (Figure 1A and B; Table 3).

Table 3 lists the amino-acid sequences from the selected candidate biomarker signals described above. These sequences were identified from MS/MS data using Mascot software.

Figure 1A shows the distribution of the signal intensity of several peptide ions derived from myosin IIA, and Figure 1B shows the signal intensity distribution of vimentin-derived peptide ions. For most signals, the intensity for group U1R1 patients showed patterns that differed significantly (i.e., were markedly higher) from those of the other patient groups.

**Immunohistochemical staining of myosin IIA and vimentin**

Representative staining of myosin IIA and vimentin is shown in Figure 2, and a summary of the immunohistochemical data is presented in Table 4. Cytoplasmic staining was observed in cases...
positive for myosin IIA and vimentin. We evaluated cases in which more than 50% of the cells showed immunohistochemical reactivity for myosin IIA, considered as overexpressing (positive). We also evaluated overexpressing (positive) cases in which more than 25% of the cells showed immunohistochemical reactivity for vimentin. On the basis of these criteria, we evaluated sample sets A and B.

Immunohistochemical evaluation of sample set A (Table 4A): all patients with cancers lacking expression of both myosin IIA and vimentin showed relapse-free survival at 5 years. On the other hand, all patients with cancers showing positive expression of both myosin IIA and vimentin suffered disease recurrence.

Immunohistochemical evaluation of sample set B (Table 4B): among 90 cases, 75 (83.3%) showed overexpression of myosin IIA, and 48 (53.3%) showed overexpression of vimentin. There was no

---

Table 3 Amino-acid sequences from the selected peptide ion signals

| Name                          | Fraction | Sequence                          |
|-------------------------------|----------|-----------------------------------|
| Myosin, heavy polypeptide 9, non-muscle | MYH9_1 | Insoluble IRELESQISELQEDLESER     |
|                               | MYH9_2 | Insoluble KANLQIQINTDLNLER        |
|                               | MYH9_3 | Insoluble HEMPHIYAIIDTAYR         |
| Vimentin                      | VIM_1   | Insoluble ETNLDSLPLVDTHSK         |
|                               | VIM_2   | Insoluble NLQGAEENYVK             |
|                               | VIM_3   | Insoluble LGQLEEEENR              |
|                               | VIM_4   | Insoluble LLQDSVEPIEANTEFK        |
|                               | VIM_5   | Soluble SGDAAIVDMVPCESSYDYPLGR    |
|                               | VIM_6   | Soluble ILTVEDHYEGGIGEAVSAWGEPPITVTHLAVNR |

---

Figure 2 Immunohistochemical reactivity of representative cases using anti-myosin IIA antibody, ab24762 (abcam, Cambridge, UK) and anti-vimentin antibody (Dako Cytomation, Denmark A/S). Case A showed positive cytoplasmic staining for both myosin IIA and vimentin. Case B showed positive cytoplasmic staining for myosin IIA and negative cytoplasmic staining for vimentin. Case C showed negative cytoplasmic staining for myosin IIA and positive cytoplasmic staining for vimentin. Case D showed negative cytoplasmic staining for both myosin IIA and vimentin.
The non-relapse survival curves of all the studied cases, there were statistically significant differences between cases negative for myosin IIA and vimentin expression and cases that were positive for both (P = 0.006), and between cases positive for either myosin IIA or vimentin and cases that were negative for both (P = 0.029; Figure 4).

DISCUSSION

Lung cancer is the leading cause of cancer death in Japan, and its incidence is still increasing. Even if surgical resection involving either lobectomy or pneumonectomy accompanied by lymph node dissection is performed at a relatively early stage, distant metastasis often occurs within a few years. More than 20% of patients with stage 1 NSCLC suffer recurrence caused by distant metastasis. Distant metastasis is the most frequent mode of recurrence in patients who undergo surgical resection of lung cancer, and it is believed that in such patients, micrometastasis is
invariably present at the time of initial treatment. If an efficient PAC regimen could be devised for total control of micrometastasis, then the prognosis of patients with lung cancer would be markedly improved. A meta-analysis conducted in the 1990s showed that PAC using platinum-based agents had no effect on the survival of patients with NSCLC, even though previous studies had suggested a 5% increase in survival at 5 years (Non-Small Cell Lung Cancer Collaborative Group, 1995). At the 2004 ASCO meeting, the results of two randomised adjuvant trials showing the efficacy of platinum-based chemotherapy – the CALGB-9633 trial (carboplatin and paclitaxel) (Straus et al, 2004) and the JBR 10 trial (cisplatin and vinorelbine) – were reported. Furthermore, a recent large-scale randomised clinical trial involving meta-analysis conducted in patients assigned to cisplatin-based PAC had a significantly higher survival rate than those assigned to post-operative observation (44.5 vs 40.4% at 5 years; P<0.03) (Arriagada et al, 2004). Also our previous study showed that PAC with uracil–tegafur conferred a survival benefit for patients with resected stage I adenocarcinoma of the lung (Kato et al, 2004). Also, meta-analysis of PAC with tegafur–uracil supported this result (Hamada et al, 2005). However, even though a significant difference was found in this study, the 5-year-survival rate benefit of this therapy was 11.5% for stage IB adenocarcinoma (Kato et al, 2004). At present, although leading lung cancer experts appear to have reached a consensus concerning the effectiveness of PAC, none of the present PAC regimens are of benefit to more than 15% of patients with NSCLC. In this context, it is very important to predict the response to PAC and to select potential responders before carrying out PAC. Therefore, we attempted to identify biomarkers of either responders or nonresponders including relapse-free patients without PAC using uracil–tegafur, for selection of patients who would benefit from PAC using proteomic analysis of surgically resected specimens of stage I lung adenocarcinoma.

As proteins play a role in both physiological and pathological functions, it is now recognised that investigation of proteins is essential to obtain an accurate grasp of cellular physiology. Recent advances in proteomic techniques, including two-dimensional polyacrylamide gel electrophoresis and MS, have brought hope that the pathogenesis of any type of malignant neoplasm will be ultimately clarified. We believe that the present concepts of proteomic analysis will prove to be extremely valuable as a means of clinical oncology, and will lead to the development of new therapeutic strategies. Liquid chromatography-tandem mass spectrometry enables simultaneous evaluation of a large number of polypeptides, and furthermore, MS/MS has made it possible to identify protein molecules by obtaining information about their amino-acid sequences. We attempted to identify proteins associated with the effectiveness of postoperative uracil–tegafur chemotherapy and the favourable prognosis of stage I adenocarcinoma, and detected two kinds of protein molecules (myosin IIA and vimentin) showing significantly high expression in the group that suffered recurrence despite administration of uracil–tegafur, in comparison with the other groups. Our semiquantitative results of LC-MS were confirmed by immunohistochemistry for myosin IIA and vimentin (Table 4A).

Nonmuscle myosin IIA is a major component of the actomyosin cytoskeleton and is generally considered to contribute to contraction of the cell posterior during migration (Ridley et al, 2003). However, there is still a profound lack of understanding of the exact mechanical roles of myosin IIA during cell migration. A recent clinical study of patients with NSCLC found a significant positive correlation between the expression levels of myosin light chain kinase (which activates myosin II) and the likelihood of disease recurrence and metastasis (Minamiya et al, 2005), indicating that myosin IIA activation could be a factor contributing to metastasis. A key role for myosin IIA in cancer cell metastasis has been further suggested, indirectly, by a number of published studies focusing on the small calcium-binding protein, metastasin-1. This protein is upregulated in many metastasis cell lines, and when overexpressed enhances metastatic behaviour (Davies et al, 1993). A major cellular target of metastasin-1 seems to be myosin IIA (Garrett et al, 2006). Although studies of metastasin-1 suggest critical roles for myosin IIA in metastasis, it remains completely unknown how myosin IIA contributes to metastasis, and which isoforms are important for this process.

Vimentin is the most ubiquitous intermediate filament protein and the first to be expressed during cell differentiation. All primitive cell types express vimentin, but in most nonmesenchymal cells, it is replaced by other intermediate filament proteins during differentiation. Vimentin is expressed in a wide variety of mesenchymal cell types and metastatic tumours, and is also in a number of other cell types derived from mesoderm, mesothelium and ovarian granulose cells. Epithelial–mesenchymal transition is a key mechanism operating in the normal development of multicellular organisms. During this process, epithelial cells progressively acquire a reversible or irreversible mesenchymal phenotype that is essential for organogenesis (Thiery, 2002). Morphogenetic epithelial–mesenchymal transition is aberrantly recapitulated during tumorigenesis in a variety of epithelial cancers, including those of the thyroid, liver, kidney, prostate, breast and lung (Arias, 2001; Thiery, 2002). The common signature of this process involves disruption of normal epithelial integrity, with loss of morphological features including polarised epithelia, and partial or total gain of mesenchymal markers with progressive acquisition of a motile and invasive phenotype (Islam et al, 1996). In addition to a disrupted epithelial morphology, dysregulation of adhesion and junctional molecules and aberrant expression of E-cadherin, epithelial–mesenchymal transition involves de novo expression of other mesenchymal markers, such as fibronectin and vimentin in epithelial cells. Aberrant expression of vimentin in tumours and transformed cell lines has been correlated with increased motility, invasive behaviour and poor prognosis (Gilles et al, 1996; Hendrix et al, 1997). Recently, it was reported that the presence of vimentin-positive tumour cells mainly in fibrotic areas is consistent with other studies that have shown a correlation between tumour fibrosis and epithelial–mesenchymal transition (Blanco et al, 2004).

These two molecules identified by proteomic analysis might reflect the cellular functions of metastasis and the mechanism of recurrence of malignant neoplasms. We attempted to validate the results of LC-MS/MS using immunohistochemistry of an additional sample set (sample set B: 90 surgically resected lung cancer specimens) with monoclonal antibodies against the two proteins. The results showed that cases lacking expression of the two proteins had a good prognosis, irrespective of whether the patients had undergone PAC. Therefore, these two proteins appear to be potentially useful biomarkers for the selection of patients who do not require PAC. In the cases positive for both of these proteins, the 5-year-survival benefit was approximately 19% in patients with adenocarcinoma who underwent PAC with uracil–tegafur. However, there was no significant difference between patients who did, and did not, undergo PAC with uracil–tegafur. Therefore, in this investigation, we failed to select patients who might benefit from this adjuvant chemotherapy. A larger-scale investigation is therefore needed to establish suitable biomarkers for the selection of patients who might benefit from PAC with uracil–tegafur, because a few per cent of patients with stage I adenocarcinoma do obtain such a benefit.

Individualised chemotherapy for lung cancer patients is currently attracting attention, because the efficacy of systemic chemotherapy using any single agent is less than 30%. Therefore, it is extremely important to select patients who might benefit from chemotherapy. Until an ideal chemotherapy agent is established, we propose that rather than focusing only on improving the efficacy of chemotherapy regimes, we should also make efforts to identify patients who will show a good response to regimens that
are already established. This proposal is justified only on the basis of evidence-based medicine. In this situation, positive indicators for the effectiveness of PAC with uracil–tegafur are needed. However, in this study, we were unable to detect novel biomarkers for selection of good responders. The two protein molecules detected in this proteomic analysis were biomarkers indicative of good prognosis.

The ultimate purpose of clinical proteomics is to improve diagnostic procedures including the exact evaluation of biological characteristics of tumour cells and to understand the molecular pathogenesis of cancers to devise novel therapeutic strategies. We believe that proteomic analysis will become an integral tool for investigation of tumour biology. We conclude that negative expression of both myosin IIa and vimentin is an indicator of good prognosis for stage I lung adenocarcinoma without the need for PAC.

ACKNOWLEDGEMENTS

This study was supported by grants from the Japan Society for the Promotion of Science (18390382). We thank Mrs Mami Murakami for her experimental expertise and statistical analysis.

REFERENCES

Arias AM (2001) Epithelial-mesenchymal interactions in cancer and development. Cell 105: 425 – 431

Arriagada R, Bergman B, Dunant A, Le Chevalier T, Pignon JP, Vansteenkiste J (2004) Cisplatin-based adjuvant chemotherapy in patients with completely resected non-small-cell lung cancer. N Engl J Med 350: 351 – 360

Barbareschi M, Girlanda S, Mauri MF, Forti S, Eccher C, Nauri FA, Togni R, Barbareschi M, Girlanda S, Mauri MF, Forti S, Eccher C, Nauri FA, Togni R, Arias AM (2001) Epithelial mesenchymal interactions in cancer and metastasis. Lab Invest 84: 999 – 1012

Barbareschi M, Girlanda S, Mauri MF, Forti S, Eccher C, Nauri FA, Togni R, Barbareschi M, Girlanda S, Mauri MF, Forti S, Eccher C, Nauri FA, Togni R, Arias AM (2001) Epithelial mesenchymal interactions in cancer and metastasis. Lab Invest 84: 999 – 1012

Blelo D, Girlanda S, Elizegi E, Ifno I, Fragla MF, Estemler M, Saffioti U, LeCanda F, Montuenega L (2004) Altered expression of adhesion molecules and epithelial-mesenchymal transition in silica-induced rat lung carcinogenesis. Lab Invest 84: 999 – 1012

Blanco D, Vicent S, Elizegi E, Ifno I, Fragla MF, Estemler M, Saffioti U, LeCanda F, Montuenega L (2004) Altered expression of adhesion molecules and epithelial-mesenchymal transition in silica-induced rat lung carcinogenesis. Lab Invest 84: 999 – 1012

Clerici M, Cognetti F, Tonato M (2003) Randomized study of adjuvant chemotherapy for non-small-cell lung cancer. J Nat Cancer Inst 95: 1453 – 1461

Davies BR, Davies MP, Gibbes FE, Barraclough R, Rudland PS (1993) Induction of the metastatic phenotype by transfection of a benign rat mammary epithelial cell line with the gene for p9Ka, a rat calcium-binding protein, but not with the oncogene Eμ-raf-1. Oncogene 8: 999 – 1008

Davies BR, Davies MP, Gibbes FE, Barraclough R, Rudland PS (1993) Induction of the metastatic phenotype by transfection of a benign rat mammary epithelial cell line with the gene for p9Ka, a rat calcium-binding protein, but not with the oncogene Eμ-raf-1. Oncogene 8: 999 – 1008

Davies BR, Davies MP, Gibbes FE, Barraclough R, Rudland PS (1993) Induction of the metastatic phenotype by transfection of a benign rat mammary epithelial cell line with the gene for p9Ka, a rat calcium-binding protein, but not with the oncogene Eμ-raf-1. Oncogene 8: 999 – 1008

Fujii S, Kitano S, Ikenaka K, Shirasaka T (1979) Effect of coadministration of uracil or cytosine on the anti-tumor activity of clinical doses of l-2-tetradhydrofuryl-5-fluorouracil and level of 5 – fluorouracil in rodents. Gann 70: 209 – 214

Garrett SC, Varney KM, Weber DJ, Bresnick AR (2006) S100A4, a mediator of metastasis. J Biol Chem 281: 677 – 680

Gilles C, Polette M, Elizegi E, Delvigne AC, Thompson EW, Foidart JM, Birembaut P (1996) Vimentin expression in cervical carcinomas: association with invasive and migratory potential. J Pathol 180: 175 – 180

Hamada C, Tanaka F, Ohita M, Fujimura S, Kodama K, Imaizumi M, Wada H (2005) Meta-analysis of postoperative adjuvant chemotherapy with tegafur-uracil in non-small-cell lung cancer. J Clin Oncol 23: 4999 – 5006

Hendrix MJ, Seftor EA, Seftor RE, Trevor KT (1997) Experimental co-expression of vimentin and keratin intermediate filaments in human breast cancer cells results in phenotypic interconversion and increased invasive behavior. Am J Pathol 150: 483 – 495

Ho DH, Pazdur R, Covington W, Brown N, Huo YY, Lassere Y, Kuritani J (1998) Comparison of 5-fluorouracil pharmacokinetics in patients receiving continuous 5-fluorouracil infusion and oral uracil plus N1-(2-tetradhydrofuryl)-5-fluorouracil. Clin Cancer Res 4: 2085 – 2088

Ikenaka K, Shirasaka T, Kitano S, Fujii S (1979) Effect of uracil on metabolism of 5-fluorouracil in vitro. Gann 70: 353 – 359

Islam S, Carey TE, Wolf GT, Wheelock MJ, Johnson KR (1996) Expression of N-cadherin by human squamous carcinoma cells induces a scattered fibroblastic phenotype with disrupted cell – cell adhesion. J Cell Biol 135: 1643 – 1654

Kato H, Ichinose Y, Ohta M, Hata E, Tsubota N, Tada H, Watanabe Y, Wada H, Tsuboi M, Hamajima N, Ohta M (2004) A randomized trial of adjuvant chemotherapy with uracil-tegafur for adenocarcinoma of the lung. N Engl J Med 350: 1713 – 1721

Kawakami T, Tateishi K, Yamano Y, Ishikawa T, Kuroki K, Nishimura T (2005) Protein identification from product ion spectra of peptides validated by correlation between measured and predicted elution times in liquid chromatography/mass spectrometry. Proteomics 5: 856 – 864

Laemmli UK (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature 227: 680 – 685

Lowry OH, Rosebrough NJ, Farr AL, Randall RJ (1951) Protein measurement with the Folin phenol reagent. J Biol Chem 193: 265 – 275

Minamiya Y, Nakagawa T, Saito H, Matsuoka I, Taguchi K, Ito M, Ogawa J (2005) Increased expression of myosin light chain kinase mRNA is related to metastasis in non-small cell lung cancer. Tumour Biol 26: 153 – 157

Non-Small Cell Lung Cancer Collaborative Group (1995) Chemotherapy in non-small-cell lung cancer: a meta-analysis using updated data on individual patients from 52 randomized clinical trials. BMJ 311: 899 – 909

Ridley AJ, Schwartz MA, Burridge K, Firtel RA, Ginsberg MH, Borisy G, Parsons JT, Horwitz AR (2003) Cell migration: integrating signals from front to back. Science 302: 1704 – 1709

Scaglotti GV, Fossati R, Torri V, Crino L, Giaccone G, Silvano G, Martelli M, Clerici M, Cognetti F, Tonato M (2003) Randomized study of adjuvant chemotherapy for completely resected stage I, II, or IIIA non-small-cell lung cancer. J Nat Cancer Inst 95: 1453 – 1461

Schwartz JC, Senko MW, Syka JE (2002) A two-dimensional quadrupole ion trap mass spectrometer. J Am Soc Mass Spectrom 13: 659 – 669

Shevchenko A, Wilm M, Vorm O, Mann M (1996) Mass spectrometric sequencing of proteins silver-stained polyacrylamide gels. Anal Chem 68: 850 – 858

Strauss GM, Herndon J, Maddaus MA, Johnstone DW, Johnson EA, Watson DM, Sugarbaker DJ, Schilsky RL, Green MR (2004) Randomized clinical trial of adjuvant chemotherapy with paclitaxel and carboplatin following resection in stage IB non-small cell lung cancer (NSCLEC): report of cancer and leukemia Group B (CALGB) Protocol 9633. Proc Am Soc Clin Oncol 23: 621

Thiery JP (2002) Epithelial – mesenchymal transitions in tumour progression. Nat Rev Cancer 2: 442 – 454

Winton T, Livingston R, Johnson D, Rigas J, Johnston M, Butts G, Cormier Y, Goss G, Inculet R, Vallieres E, Fry W, Bethune D, Ayoub J, Ding K, Seymour L, Graham B, Tsaol MS, Gandara D, Kessler D, Demmy T, Shepherd F (2005) Vinorelbine plus cisplatin vs observation in resected non-small-cell lung cancer. N Engl J Med 352: 2589 – 2597