Research on stress-induced apoptosis of natural killer cells and the alteration of their killing activity in mouse liver

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Abstract

AIM: To investigate the stress-induced apoptosis of natural killer (NK) cells and the changes in their killing activity in mouse livers.

METHODS: A restraint stress model was established in mice. Flow cytometry was employed to measure the percentage of NK cells and the changes in their absolute number in mouse liver. The cytotoxicity of hepatic and splenic NK cells was assessed against YAC-1 target cells via a 4 h 51Cr-release assay.

RESULTS: The restraint stress stimulation induced the apoptosis of NK cells in the liver and the spleen, which decreased the cell number. The number and percentage of NK cells in the spleen decreased. However, the number of NK cells in the liver decreased, whereas the percentage of NK cells was significantly increased. The apoptosis of NK cells increased gradually with prolonged stress time, and the macrophage-1 (Mac-1)+ NK cells were more susceptible to apoptosis than Mac-1− NK cells. Large numbers of Mac-1− NK cells in the liver, which are more resistant to stress-induced apoptosis, were observed than the Mac-1− NK cells in the spleen. The stress stimulation diminished the killing activity of NK cells in the spleen was significantly decreased, but the retention of numerous Mac-1− NK cells in the liver maintained the killing ability.

CONCLUSION: Significant stress-induced apoptosis was observed among Mac-1+ NK cells, but not Mac-1− NK cells in the mouse liver. Stress stimulation markedly decreased the killing activity of NK cells in the spleen but remained unchanged in the liver.

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Key words: Restraint stress; Natural killer cells; Cell apoptosis; Killing activity

Core tip: Hepatic natural killer (NK) cells are classified into macrophage-1 (Mac-1) + and Mac-1− cells, and the different functional characteristics of Mac-1+ or Mac-1− NK cells in response to stress stimulation are confirmed. This study further proves the heterogeneity of NK cell function, and the results provide a reference for preventing the immune system damage caused by stress.

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INTRODUCTION

In the modern society, the acceleration of work and life style and the deterioration of the environment, as well as natural disasters and frequent traffic accidents, expose people to increasing stress. Prolonged or intense stress that overwhelms autoregulation, causes nervous, endocrine, and immune system dysfunction\textsuperscript{[4-6]}, as well as the apoptosis of lymphocytes such as natural killer (NK) cells, B cells and T cells. Immune system dysfunction is the direct cause of infectious diseases, cancers and self-deterioration\textsuperscript{[10]}. NK cells are an important type of lymphocytes, accounting for 10\% to 15\% of total lymphocytes and play a crucial role in body resistance to against infections and tumours, as well as immune and hematopoietic regulation\textsuperscript{[7,8]}. This study aims to determine the effects of persistent and intense stress on the apoptosis of hepatic and splenic NK cells and change in their function in mice.

MATERIALS AND METHODS

Experimental animals

Eight-week-old clean grade C57BL/6 mice were purchased from the Animal Centre of The 3\textsuperscript{rd} Affiliated Hospital of Harbin Medical University.

Mouse model preparation

The protocol used has been described in the published literature\textsuperscript{[9]}. The mice were placed in a 50 mL Falcon tube with 4 to 5 drilled holes at the bottom to maintain ventilation. Sufficient amounts of absorbent cotton were placed inside the tube to immobilise the mouse, and then the lid was screwed shut. The tube was kept at room temperature for 24 h in RPMI 1640 containing 200 mL/L FCS. YAC-1 cells were collected at exponential phase and counted for 10\% to 15\% of total lymphocytes and play a crucial role in body resistance to infections and tumours, as well as immune and hematopoietic regulation\textsuperscript{[7,8]}. The cytotoxicity of NK cells was assessed against YAC-1 target cells. Target cells were continuously cultured for 3 h and intense stress on the apoptosis of hepatic and splenic NK cells and change in their function in mice.

Mouse lymphocyte preparation

The protocol used has been described in the published literature\textsuperscript{[9]}. The mice was anaesthetised with ether and sacrificed via heart puncture. Subsequently, the mouse liver and spleen were collected and minced. The tissue sample was washed with phosphate-buffered saline (PBS) and filtered with 200 mesh strainer, and then the cell suspension was collected. After gradient centrifugation, the cells were lysed with 0.83\% NH\textsubscript{4}Cl-Tris buffer (pH 7.6). The resulting cell suspension was collected and the concentration was adjusted to 1.0 \times 10^5/mL.

Immunofluorescence labelling

Lymphocytes were isolated from mouse liver and spleen, and then double or triple immunofluorescence staining was performed to identify the CD\textsubscript{3}NK\textsubscript{1.1} cells as NK cells. Fluorescein-isothiocyanate (FITC)-labelled antibodies: CD\textsubscript{3} (145-2C11 clone); PE-labelled antibody: NK\textsubscript{1.1} (PK136 clone); Biotin-labelled antibody: macrophase-1 (Mac-1) (M1/70 clone) CD\textsubscript{8} (H1.2F3 clone), Ly49C/I (5E6 clone). All the monoclonal antibodies were purchased from BD Biosciences Pharmingen in San Diego, United States.

Cell suspension was transferred in centrifuge tube (cell number < 2 \times 10^5). After 2 min of centrifugation at 2500 r/min and 4 \textdegree C, the supernatant was removed, followed by vibration. Then, 10 \mu L of 2.4 G2 was added (anti-FcyRII/III). After incubation at 4 \textdegree C for 10 min, 10 \mu L of various monoclonal antibodies (CD\textsubscript{3}, NK\textsubscript{1.1}, Mac-1, CD\textsubscript{8}, and Ly49C/I) were added, accordingly. After vortex and incubation at 4 \textdegree C for 20 min, the cells were washed once with PBS (2500 r/min at 4 \textdegree C). For double staining, the cells were diluted with 0.5 mL of PBS and filtered with nylon mesh, and then 5 \mu L of propidium iodide (PI) was added for flow cytometry analysis. When subjected for triple staining, cells were incubated with 10 \mu L of biotin-labelled secondary antibody at 4 \textdegree C for 20 min, then washed with PBS once (2500 r/min at 4 \textdegree C). After diluting with 0.5 mL of PBS and filtration with a nylon mesh, the stained cells were analyzed via flow cytometry\textsuperscript{[11]}. Flow Cytometer was FAC sort from BD-United States and software was Cell Quest 3.0.

Detection of killing activity of NK cells

The cytotoxicity of NK cells was assessed against YAC-1 target cells. Target cells were continuously cultured for 24 h in RPMI 1640 containing 200 mL/L FCS. YAC-1 cells were collected at exponential phase and counted through trypan blue staining. Viable cells were considered as targets cells when their percentage exceeded 95\%. The cell concentration was adjusted to 1 \times 10^5/mL with RPMI 1640. After incubation with \textsuperscript{51}Cr for 2 h, the cells were washed three times with RPMI 1640 to remove free \textsuperscript{51}Cr. The target cell concentration was adjusted to 1.0 \times 10^4/mL or 2.0 \times 10^4/mL. The cells were divided into three groups: NK cell group, target cell maximum release group, and target cell spontaneous release group. Subsequently, the cells were seeded in U-bottom microplates (96-well). Lymphocytes in the mouse liver and spleen were utilized as effector cells, which were added into the U-bottom microplates at an effector:target ratio of 50:1, 25:1, and 12.5:1 in a volume of 100 \mu L/vial. The cells were incubated at 37 \textdegree C with 5\% CO\textsubscript{2} for 4 h and the microplates were centrifuged at 1500 r/min for 5 min. About 100 \mu L of supernatant was collected from each well, and its radioactivity (CPM) was measured with gamma counter\textsuperscript{[12]}. The specific killing rate was calculated using the following formula: specific killing rate (\%) = (experimental cell release - target cell spontaneous release)/(target cell maximum release - target cell spontaneous release) \times 100\%.

Statistical analysis

The data are presented as mean \pm SD and percentage.

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Results

Stress stimulation and the change of NK cell number

The percentage of splenic NK cells in total lymphocytes did not change significantly (3.9% ± 1.2% vs 2.6% ± 1.1%, P > 0.05), whereas the percentage of hepatic NK cells increased (8.6% ± 1.3% vs 14.9% ± 1.5%, P < 0.05; Figure 1).

After restraint stress stimulation, the numbers of lymphocytes in the experimental and control groups were (53.1 ± 9.7) × 10^5 vs (19.7 ± 4.6) × 10^5 (n = 6, P < 0.05) in liver, (87.7 ± 9.6) × 10^5 vs (36.4 ± 7.1) × 10^5 (n = 6, P < 0.05) in spleen. The number of NK cells in the experimental and control groups were (57.4 ± 8.9) × 10^5 vs (24.6 ± 7.3) × 10^5 (n = 6, P < 0.05) in the liver, (29.7 ± 6.5) × 10^6 vs (8.6 ± 1.4) × 10^6 (n = 6, P < 0.05) in the spleen (Figure 2).

Number of Mac-1^+ and Mac-1^- NK cells

NK cells were isolated from mouse liver and spleen after 24 h of restraint stress stimulation, followed by immunofluorescence staining with FITC: Mac-1 and PE: NK1.1. The cells were analyzed via flow cytometry. The results showed that the percentage of hepatic Mac-1^- NK cells in the experimental group was significantly higher than that of the control group (77.2% ± 1.7% vs 33.9% ± 1.1%, P < 0.05, Figure 1B), whereas the percentage of hepatic Mac-1^- NK cells relatively increased. The percentage of splenic Mac-1^- NK cells was slightly decreased (75.1% ± 1.1% vs 68.5% ± 1.6%, P > 0.05).

Stress stimulation and apoptosis of NK cells

NK cells were isolated from mouse liver after 24 h of restraint stress stimulation, followed by immunofluorescence staining with Annexin V-FITC and PI. The cells were analyzed via flow cytometry. The results revealed significant apoptosis of NK cells (9.5% ± 1.4% vs 19.3% ± 1.3%, P < 0.05), especially the Mac-1^- NK cells (5.2% ± 1.8% vs 19.3% ± 1.4%, P < 0.05), whereas the apoptosis of Mac-1^- NK cells did not significantly change (13.4% ± 1.3% vs 7.4% ± 1.7%, P > 0.05; Figure 4).

Stress stimulation and the change of killing activity mediated by NK cells

The killing activity of NK cells was markedly decreased in the spleen (16.7% ± 1.4% vs 8.9% ± 1.1%, P < 0.05), but was sustained in the liver even after 24 h of restraint stress stimulation (Figure 5).

Discussion

With the development of modern medical technology, increasing attention has been focused on the relationship between stress and health. The response of the body to stress is a dynamic balance, which allows the body to recover to its original state through autoregulation after stress reaction. However, persistent stress may overwhelm...
Numerous studies have demonstrated that acute stress stimulation causes distinctly reduced number of lymphocytes in the thymus, spleen, peripheral blood, and liver, as well as disrupt the function of T, B and NK cells. Moreover, the decrease in lymphocytes mainly results from apoptosis.

Our research shows that the number of splenic lymphocytes was significantly decreased, but their percentage did not change significantly. We speculate that various lymphocytes in spleen decreased with the same percentage, which is consistent with the results of previous studies\[10,15\]. Moreover, the results showed that the number of hepatic lymphocytes significantly declined but the percentage of NK cells dramatically increased (Figures 1A and 2). Therefore, the number of NK cells in the liver and the spleen were determined. Although both the liver and the spleen had fewer NK cells, the number of NK cells was relatively high in the liver, dramatically increasing the percentage of NK cells in the liver (Figure 2). These results indicate that the hepatic NK cells differed from splenic NK cells after stress stimulation. A recent study reported that NK cells have organ specificity, which allows liver to be considered as immune organ and NK cells possess unique functional characteristics\[16,17\].

Mac-1 was employed to distinguish the subtypes of NK cells with different functions. Mac-1 (CD11b/CD18) is an adhesion molecule of the integrin family, highly expressed in most myeloid hematopoietic cells, such as neutrophils, monocytes/macrophages, eosinophils and B cells. Mac-1 is closely correlated with cell phagocytosis and adhesion, as well as a marker for myeloid and lymphoid hematopoietic cells. NK cells are the only lymphoid cells that express Mac-1. Some researchers have considered Mac-1 as a marker for mature NK cells, which have cell killing activity and are able to secrete cytokines, whereas Mac-1 NK cells are immature, with limited cell killing activity and cytokine production\[18-20\]. Mac-1 is expressed by 80% to 90% of mature NK cells in the liver, spleen, and peripheral blood. Therefore, current research on NK cells is mainly focused on Mac-1 NK cells. Our previous study discovered numerous Mac-1 NK cells in the liver and demonstrated that Mac-1 NK cells and Mac-1 NK cells have different functional characteristics and cell phenotypes\[21\]. Therefore, hepatic NK cells were classified into Mac-1 positive and Mac-1 negative subtypes according to Mac-1 expression.

Our study shows that the number of Mac-1 and Mac-1 NK cells in the spleen decreases with same percentage after stress stimulation, which results in sustained Mac-1 expression in splenic NK cells. The number of hepatic Mac-1 NK cells significantly decreased, whereas the number of Mac-1 NK cells did not change significantly, which accounts for the reduced Mac-1 expression in hepatic NK cells (Figures 1B and 3). Therefore, we speculate that Mac-1 hepatic NK cells are resistant to the apoptosis induced by stress stimulation.

A large number of studies have demonstrated that acute stress stimulation causes distinctly reduced number of lymphocytes in the thymus, spleen, peripheral blood, and liver, as well as disrupts the function of T, B and NK cells. Moreover, the decrease in lymphocytes is mainly caused by the apoptosis of lymphocytes.

Intracellular Annexin V expression was measured to assess the degree of apoptosis of hepatic NK cells induced by stress stimulation\[23\]. The results revealed significant NK cell apoptosis, especially Mac-1 NK cells, whereas Mac-1 NK cell apoptosis remained unchanged (Figure 4).

To determine the effects of stress on NK cell function, we employed YAC-1 as target cells to measure killing activity of NK cells. The results imply that stress stimulation decreases the killing activity of splenic NK cells, which accords with the results of previous research. Our study also discovered that stress stimulation does not affect the killing activity of hepatic NK cells, which contrasts with the conclusion of previous investigations. We concluded that Mac-1 NK cells had stronger killing activity than that of Mac-1 NK cells\[23\]. We also believed that the number of hepatic Mac-1 NK cells declined because of cell apoptosis, which allowed apoptosis-resistant Mac-1 NK cells to survive, and to exhibit relatively strong cell killing activity.

In conclusion, our research preliminarily demon-
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**Figure 4** Apoptosis of natural killer cells after 24 h of restraint stress. A: Apoptosis of NK cells; B: Apoptosis of Mac-1+ NK cells; C: Apoptosis of Mac-1- NK cells. NK: Natural killer; Mac-1: Macrophage-1.
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Peer review
Authors investigated the stress-induced apoptosis of NK cells and the changes in their killing activity in mouse livers. NK cell is an important type of lymphocytes. The authors made an interesting research on NK cell. This study proves the anti-stress ability of Mac-1-hepatic NK cells. This finding suggests that Mac-1-NK cells maintain immune functional stability under stress conditions. Further studies should investigate how to characterize Mac-1-NK cells and utilize them for preventing the immune dysfunction caused by stress.

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