Linking Inflammation to the Brain-Liver Axis

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Although much remains to be explored, these findings collectively highlight a crucial role of ER stress and inflammation in liver and fat to impair insulin signaling and dysregulate glucose homeostasis in obesity and diabetes. The key question that remains to be addressed is whether overnutrition/obesity induces ER stress and inflammation in the central nervous system to disrupt the ability of insulin to control glucose homeostasis. If this is the case, do any of the key players that are highlighted above play a role in this dysregulation?

In fact, high-fat feeding induces ER stress and UPR as well as the IKK-β/NF-κB proinflammatory pathway in the hypothalamus of rodents (14,15). The activation of hypothalamic ER stress and inflammation impair the ability of central insulin and leptin to inhibit appetite. TNF-α induces ER stress in the hypothalamus (16), while fatty acids activate hypothalamic TLR4 to impair the anorectic effect of central leptin (17). In fact, hypothalamic leptin’s ability to inhibit food intake is restored in mice with neuronal-specific knockout of the TLR adaptor protein MyD88 (18), while anti-inflammatory cytokines such as interleukin-10 reduce hypothalamic inflammation and mediate the ability of exercise to improve the anorectic control of central insulin and leptin in diet-induced obese rats (19). Although mounting evidence indicates that high-fat feeding induces hypothalamic ER stress and inflammation, the metabolic consequence has been limited to the dysregulation of food intake.

In this issue of Diabetes, Milanski et al. (20) have linked hypothalamic inflammation to a disruption of the brain-liver axis that controls glucose homeostasis in obese rodents through well-designed and executed experiments. The authors first confirm that consumption of a high-fat diet increased hypothalamic expression of the inflammatory cytokines TNF-α and IL-1β in rats, then demonstrate that pretreatment with central anti-TLR4 antibody or an anti-TNF-α monoclonal antibody significantly reduced expression of these cytokines and inhibited NF-κB in the hypothalamus. Neutralization of hypothalamic TLR4 or TNF-α in obese rats improved glucose tolerance (as assessed by intraperitoneal glucose tolerance test), and this was associated with improved hepatic insulin signal transduction (insulin receptor substrate 1 → Akt → FoxO1). Next, the authors reproduced and earlier findings that TLR4 and TNF-α receptor 1 knockout mice were protected against diet-induced insulin resistance. This was further confirmed by the fact that both TLR4 and TNF-α receptor 1 knockout mice were protected from hypothalamic fatty acid–induced hepatic insulin resistance, suggesting that hypothalamic events may represent an important portion of the total body phenotype of TLR4 and TNF-α receptor 1 knockout mice.

To assess whether changes in hepatic insulin signaling are responsible for the improved glucose tolerance, the authors performed a pyruvate tolerance test, a hyperinsulinemic-euglycemic clamp, and assessed changes in hepatic gluconeogenic gene expression (PEPCK and
glucose-6-phosphatase [G6Pase]) in obese rats with hypothalamic inflammatory neutralization. Inhibition of hypothalamic inflammation reduced pyruvate-induced gluconeogenesis and normalized insulin-mediated suppression of glucose production and hepatic PEPCK and G6Pase gene expression in obese rats. In addition, both vagotomy and pharmacological inhibition of muscarinic receptors reversed the metabolic benefits resulting from hypothalamic anti-inflammation, indicating a brain-liver neural axis is required to restore hepatic insulin sensitivity.

Although pharmacological inhibition of hypothalamic TLR4—but not TNF-α—led to a substantial weight loss in obese rats, inhibition of hypothalamic TLR4 and TNF-α both restored the ability of insulin to inhibit glucose production through the brain-liver axis, suggesting that enhancement of hepatic insulin sensitivity was not secondary to weight loss. Neutralization of hypothalamic TLR4 or TNF-α in obese rats also reduced hepatic steatosis, which could have led to an enhancement in hepatic insulin sensitivity. However, despite the fatty liver phenotype of LDL receptor knockout mice, inhibition of hypothalamic inflammation in LDL receptor knockout mice fed a high-fat diet showed improved hepatic insulin signal transduction. Thus, these data collectively suggest that independent of changes in body weight and hepatic lipid accumulation, inhibition of diet-induced hypothalamic inflammation restores the ability of insulin to stimulate hepatic signal transduction and suppress glucose production in obese rodents.

Of note, insulin triggers signaling cascades and activates ATP-sensitive potassium (K_{ATP}) channels in the hypothalamus to inhibit glucose production (21), while central leptin similarly enhances insulin-mediated inhibition of glucose production in normal rats (22). Thus, it remains to be assessed whether inhibition of hypothalamic inflammation restores the central ability of insulin and/or leptin signaling to activate a brain-liver circuit to inhibit glucose production in obese rodents. In addition, it would be important to investigate the mechanistic links between hypothalamic TLR4 or TNF-α signaling and central insulin/leptin resistance, and to further assess whether hypothalamic activation of ER stress, UPR, JNK, and/or NF-κB impair central insulin/leptin signaling to dysregulate glucose production (Fig. 1).

Lastly, obesity is not only associated with hypothalamic inflammation and gliosis in rodents but also induces gliosis in the hypothalamus of humans (23). Given that intranasal insulin delivery lowers plasma glucose levels in humans (24,25) while activation of K_{ATP} channels in the brain of humans is implicated to lower glucose production (26), a possibility remains that obesity induces insulin resistance in the brain and consequently dysregulates glucose homeostasis in humans as in rodents.

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