Pulsatile Delivery of Methylphenidate Hydrochloride Pulsincap by Box-Behnken Design

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

The objective of this study was to prepare and evaluate Methylphenidate hydrochloride pulsatile drug delivery system by applying Box-Behnken design. The drug is a central nervous system (CNS) stimulant used in the treatment of attention deficit hyperactivity disorder (ADHD). Pulsincap system was prepared by using formaldehyde cross linked capsules. Capsules were filled with methylphenidate hydrochloride granules and hydrogel plug made of HPMC K100M is placed over granules to achieve desired drug release after lag time. The untreated cap was then fitted and sealed using 5% ethyl cellulose ethanolic solution to the formaldehyde treated capsule body. Granules were prepared by wet granulation technique using two polymers Ethyl cellulose and Eudragit RS100. Box-Behnken design was applied for optimization in which three independent variables, X1 = Drug; polymer ratio, X2 = Polymer: polymer ratio (Ethyl cellulose: Eudragit RS 100) and X3 = Plug weight were selected. Two dependent variables Y1 = lag time and Y2 = percent release were selected. The empty formaldehyde treated capsules were evaluated for physical appearance, solubility, capsule dimensions and formaldehyde content. Hydrogel plugs were evaluated for hardness & thickness of the plug, lag time and swelling index. Granules were evaluated for percentage yield, assay and flow properties. The prepared pulsincap formulations were evaluated for weight variation, content uniformity, capsule lock length, in-vitro dissolutions studies, drug kinetics and stability studies. Contour plots and Response surface plots indicated that with the increase in X1 and X3 there is increase in Lag time and decrease in %

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that delivers extended release followed by CONCERTA dopamine is scientific studies. Dopamine exhibited persistent, inattention for at least seven years. Neurological imbalances symptoms by ADHD patients are because of dopamine imbalances, wherein higher levels of dopamine in brain are seen in the afternoon time. Certain neurotransmitters are low in quantity or lacking in patients with ADHD is shown in scientific studies. Methylphenidate hydrochloride is a central nervous system (CNS) stimulant used in the treatment of Attention Deficit Hyperactivity Disorder (ADHD). The drug blocks dopamine transporters and increases the neurotransmitters (dopamine and noradrenalin) at synapse. Methylphenidate hydrochloride like RITALIN and CONCERTA provides both immediate release followed by a delayed release of the drug. Ritalin LA® is a methylphenidate hydrochloride extended-release capsule showing bi-modal release profile [3]. CONCERTA® is an extended-release tablet of methylphenidate HCl USP and delivers drug at a controlled rate using osmotic pressure. Ritalin LA® uses the SODAS® (Spheroidal Oral Drug Absorption System) technology. From research envisaged we found that the pulsincap technology was not used. So, in the present study Methylphenidate HCl capsules using “pulsincap technique” were formulated to obtain delayed release of the drug (after lag time). In this ADHD condition, there is a necessity for the drug to release in afternoon time during which the symptoms are highly observed. When administered to a child in the morning around 8 to 9 A.M, prepared pulsincap releases in the afternoon around 12 to 1 P.M, reducing the symptoms of ADHD in children. This avoids dosing during day time which is highly beneficial for school children. Generally, for children drug dose cannot be given accurately at afternoon time. So, the drug can be given in the morning hours which tend to release in the afternoon time. Pulsatile drug delivery of Methylphenidate hydrochloride is required as the symptoms of ADHD are highly observed in the afternoon time. “Pulsicap” technique is used to prepare Methylphenidate hydrochloride pulsatile-release capsules by applying Box-Behnken Design. “Formaldehyde treated capsule bodies are used for preparing pulsicap system. Highly retarding polymers like Eudragit RS 100 and Ethyl cellulose N22 are chosen to prepare the methylphenidate HCL granules. HPMC K100M and HPMC K4M are chosen to prepare hydrogel plugs. The pulsicap is formulated by filling the methylphenidate HCL granules in to formaldehyde cross-linked capsule bodies and by placing a hydrogel plug upon it. Lag time is influenced by the polymers and hydrogel plug used” [4].

1. INTRODUCTION

Pulsatile drug delivery systems (PDDS) are gaining popularity as they ensure spatial, temporal and, smart delivery by delivering drugs at the right place in right amount at right time thereby improving patient compliance [1]. These devices are meant to provide drugs in accordance with disease’s circadian behaviour. This means that these systems will deliver drug at a point in the circadian cycle when disease is at its most severe and fatal (24 hrs.). The pulsatile drug delivery technique is most useful for drugs administered to treat disorders with a chronopharmacological pattern [2].

Attention Deficit Hyperactivity Disorder (ADHD) is a disorder that is characterised by a pattern of inattention (inability to concentrate) sometimes combined with hyperactivity-impulsivity that is persistent, developmentally inappropriate, and occurs in two different settings before seven years age. Neurological imbalances symptoms exhibited by ADHD patients are because of dopamine imbalances, wherein higher levels of dopamine in brain are seen in the afternoon time. Certain neurotransmitters are low in quantity or lacking in patients with ADHD is shown in scientific studies. Methylphenidate hydrochloride is a central nervous system (CNS) stimulant used in the treatment of Attention Deficit Hyperactivity Disorder (ADHD). The drug blocks dopamine transporters and increases the neurotransmitters (dopamine and noradrenalin) at synapse. Methylphenidate hydrochloride like RITALIN and CONCERTA provides both immediate release followed by a delayed release of the drug. Ritalin LA® is a methylphenidate hydrochloride extended-release capsule showing bi-modal release profile [3]. CONCERTA® is an extended-release tablet of methylphenidate HCl USP and delivers drug at a controlled rate using osmotic pressure. Ritalin LA® uses the SODAS® (Spheroidal Oral Drug Absorption System) technology. From research envisaged we found that the pulsincap technology was not used. So, in the present study Methylphenidate HCl capsules using “pulsincap technique” were formulated to obtain delayed release of the drug (after lag time). In this ADHD condition, there is a necessity for the drug to release in afternoon time during which the symptoms are highly observed. When administered to a child in the morning around 8 to 9 A.M, prepared pulsincap releases in the afternoon around 12 to 1 P.M, reducing the symptoms of ADHD in children. This avoids dosing during day time which is highly beneficial for school children. Generally, for children drug dose cannot be given accurately at afternoon time. So, the drug can be given in the morning hours which tend to release in the afternoon time. Pulsatile drug delivery of Methylphenidate hydrochloride is required as the symptoms of ADHD are highly observed in the afternoon time. “Pulsicap” technique is used to prepare Methylphenidate hydrochloride pulsatile-release capsules by applying Box-Behnken Design. “Formaldehyde treated capsule bodies are used for preparing pulsicap system. Highly retarding polymers like Eudragit RS 100 and Ethyl cellulose N22 are chosen to prepare the methylphenidate HCL granules. HPMC K100M and HPMC K4M are chosen to prepare hydrogel plugs. The pulsicap is formulated by filling the methylphenidate HCL granules in to formaldehyde cross-linked capsule bodies and by placing a hydrogel plug upon it. Lag time is influenced by the polymers and hydrogel plug used” [4].

2. MATERIALS AND METHODS

2.1 Chemicals and Reagents

Methylphenidate Hydrochloride was obtained from Dr. Reddy’s laboratories, Hyderabad, India. Size “0” capsules were obtained from S.D. Fine Chemicals Ltd, Hyderabad, India. Eudragit RS 100 was obtained from Mumbai, Maharashtra, India. Ethyl cellulose N22 was obtained from Balaji Drugs, Hyderabad, India. HPMC K100M and HPMC K4M were obtained from Yarrow Chemical Products Mumbai, Maharashtra, India. All the chemicals utilised were of analytical standards.
2.2 Methods

2.2.1 Formulation of Pulsincap drug delivery system

Includes various steps.

2.2.1.1 Cross linking of empty capsules

“In order to prepare a pulsincap system size ‘0’ capsules were chosen and the solubility of these gelatin capsules is modified by cross linking them with formaldehyde. Hard gelatin capsules of size ‘0’ were taken and their bodies were separated from caps. 25 ml of 15% (v/v) formaldehyde was prepared, taken into petri plate and placed at the bottom of the desiccator. Capsules bodies were evenly spread on the mesh and the mesh is placed above the petri plate containing formaldehyde. The desiccators were tightly closed and empty bodies of capsules were exposed to formaldehyde vapours. The reaction time is then optimised by removing capsule bodies at different time intervals from desiccator i.e., capsule bodies were collected at every 1 hour until 6 hours. They are dried at 50°C for 30 minutes after being removed from the desiccator to ensure that the reaction between gelatin and formaldehyde vapours is completed. Residual formaldehyde was removed by drying capsules bodies at room temperature. These capsule bodies were sealed with untreated caps and stored in self-sealing covers” [5,6].

2.2.1.2 Preparation of hydrogel plug

Two different swellable hydrophilic polymers HPMC K100M and HPMC K4M were initially selected as they can control the lag time. Required amounts of HPMC K100M and HPMC K4M were weighed and compressed using ‘6mm punch’ in tablet compression machine [5].

2.2.1.3 Preparation of granules

Wet granulation method was used for preparation of Methylphenidate HCl granules using PVPK30 as granulating agent. The wet mass was passed through a 20#mesh to obtain granules and dried at room temperature.

2.2.1.4 Filling of granules in capsule

Granules equivalent to 40mg of the drug dose were weighed and filled into formaldehyde treated capsule body and locked by hydrogel plug and fixed with untreated cap [5].

2.2.1.5 Sealing of capsules

The cap was sealed with the body using 5 % ethyl cellulose ethanolic solution [5].

2.2.2 Drug-excipient compatibility study

2.2.2.1 FTIR studies

The spectrum analysis of pure drug and physical mixture of drug with different excipients were studied by FTIR. Potassium bromide disk was mounted in a holder and IR spectrum was recorded from 4000cm to 500 cm in a scan time of 12 minutes using a shimadzu (Koyo, Japan) IR spectrophotometer (model-8400S). The spectra were observed for the presence of characteristic peaks.

2.2.2.2 DSC studies

The instrument was calibrated with indium standard 3-5mg samples were weighed and placed in a closed, hermetic sample pans with pin hole. Thermograms were produced by heating the sample at a constant rate 10°C/min from 0°C to 210.0°C. A dry purge of nitrogen gas of 50 ml/min was used for all runs. Samples were heated. The heat of fusion, melting point, appearance of any new peak, disappearance of the crystalline sharp peak and peak shape were recorded. The thermogram of the optimized Methylphenidate HCl formulation was superimposed with that of pure drug.

2.3 Evaluation of Empty Formaldehyde Treated Capsules

2.3.1 Physical examination

Capsules were visually examined for any defects after 6 hrs. of formaldehyde cross linking.

2.3.2 Solubility studies

“The solubility of capsule bodies which were cross linked using 15% v/v formaldehyde solution was checked in orbital shaker bath. This was performed in order to optimize the crosslinking time. Capsules were collected at the end of every 1 hour of cross linking and checked for their solubility in 0.1N HCL. Deformation of capsule body shape is considered as end point” [5,7,8].

2.3.3 Measurement of dimensions of capsule bodies

“Capsules are subjected to measurement of dimensions in order to compare the differences
between plain capsules and formaldehyde cross-linked capsules. Dimensions like total capsule length, capsule body diameter and capsule body length of both plain capsules and formaldehyde cross-linked capsules were measured by using screw gauge and compared [6,8].

2.3.4 Quantitative test for free formaldehyde content

“To prove that the formaldehyde content in the cross-linked capsules is within the limits a quantitative test was employed. The sample was accurately weighed (about 3 g of formaldehyde treated capsules) and added to a mixture of H₂O₂ (25ml) and 1M sodium hydroxide (50ml) in a conical flask. Heated on the water bath until effervescence ceases (usually about 30 mins), cooled and excess of alkali was titrated with 1M hydrochloric acid, using phenolphthalein as indicator” [9].

Equivalent factor: 0.03003g of HCHO ≡ 1ml of M NaOH

2.4 Evaluation of Hydrogel Plugs

2.4.1 Hardness and thickness

The hardness and thickness of 50mg, 75mg and 100mg HPMC K 100M hydrogel plug were measured by using Monsanto hardness tester and screw gauge respectively [5].

2.4.2 Lag time

Lag time was determined during in vitro dissolution studies of formulations. The prepared hydrogel plugs were plugged to capsule bodies containing formulated granules and the caps were closed. The lag time test was conducted using USP I dissolution testing apparatus using 0.1N HCl for 2 hours, followed by 4.5 pH phosphate buffer for 3 hours and then followed by 6.8 pH phosphate buffer for 3 hrs. The drug release was observed. The graphs were plotted by taking time on x-axis vs. % drug release on y-axis and by the extrapolation of drug release line on to the x-axis, lag time was obtained [7].

2.4.3 Swelling index

Individually weighed plugs (W1) were placed in glass beakers with 200 ml of 0.1 N HCl and incubated at 37°C±1°C. At regular 1 hour time intervals until 4 hrs the plugs were removed from the beakers, and the excess surface liquid was removed carefully using the filter paper. The swollen plugs were weighed again (W2) and swelling index (SI) was calculated using the formula [10].

\[
\text{Swelling Index} = \frac{W2-W1}{W1}
\]

2.5 Evaluation of Granules

2.5.1 Percentage yield

The prepared granules were collected and weighed (practical yield). The percentage yield of granules was calculated using formula [11].

\[
\text{Percentage yield} = \frac{\text{Practical yield}}{\text{Theoretical yield}} \times 100
\]

2.5.2 Assay

Granules were accurately weighed equivalent to the drug dose and dissolved in 10ml of acetone. After required dilutions the drug was analysed by UV-Visible spectrophotometric method at 208nm [12].

2.5.3 Flow properties

Angle of repose, Bulk density, Tapped density, Compressibility index, and Hausner’s ratio were determined.

2.5.4 Angle of repose

Funnel method was used to determine angle of repose. Granules were accurately weighed and placed in a funnel, with the funnel’s height adjusted so that its tip touched the apex of the granules inside. The granules were allowed to pass through the funnel and drop on the surface. The diameter of the pile of the granules was measured and the angle of repose was calculated using the equation:

\[
\tan \theta = \frac{h}{r}
\]

Where, \( \theta \) = angle of repose, \( h \) = height of the heap (in cm) and \( r \) = radius of the base (in cm) [13].

2.5.5 Bulk density (\( pb \))

It is the mass of the powder divided by the bulk volume [13].
2.5.6 Tapped density (pt)

It is the mass of the powder divided by the tapped volume [13].

2.5.7 Compressibility index

Carr’s index was calculated from the following equation using the values of bulk density (pb) and tapped density (pt) [13].

\[ C = (pt - pb / pt) \times 100 \]

2.5.8 Hausner’s ratio

It is an indirect index of ease of powder flow. It is calculated using formula [12-14]

\[ \text{Hausner’s ratio} = \frac{pt}{pb} \]

pt is tapped density and pb is bulk density.

2.6 Optimization Using Box-Behnken Design

“A three-factor, three-level Box-Behnken design (BBD) was selected for the optimization procedure to explore quadratic response surfaces and construct second order polynomial models using Design Expert 11 (Version 11; Stat-Ease Inc, Minneapolis, MN). The Box-Behnken design is an independent quadratic design used to optimize the formulation where, the treatment combinations are taken at midpoints of edges and at the center of the process space. Initial preliminary trials were carried out to evaluate the formulations and for the processing of pulsatile capsules”. It was observed that variations in the quantities of polymers affect the lag time and percent release. When the polymers ethyl cellulose and eudragit RS 100 were used in ratio the release of methylphenidate was delayed and the lag time was found to be increased and also when the 100mg of hydrogel plug (HPMCK100M) was used it was found that the lag time increased. Based on this, three independent variables, X1 = Drug: polymer ratio, X2 = Polymer: polymer ratio (Ethyl cellulose: Eudragit RS 100) and X3 = Plug weight were selected at three levels (low, medium, and high). The levels for these three parameters were determined from the preliminary trials. Y1 = lag time in hrs and Y2 = percent release after 8 hrs were selected as dependent factors [15]. The independent and dependent factors, are given in Table 1. A three level three factor Box-Behnken design (BBD) was generated using Design Expert 11 software. The design gives us the coded values which are converted in to the actual values for all the factor levels to obtain the formulations according to Box- Behnken design, this is given in Table 2.

2.7 Evaluation of Pulsincap

2.7.1 Weight variation

Twenty capsules were selected randomly and weighed collectively and individually. Average weight was calculated. The % weight variation was calculated [5].

2.7.2 Content uniformity

“Twenty capsules were randomly selected from each batch and their contents were removed and powdered. From this sample, 40mg of powder (equivalent to drug dose) was accurately transferred to 10ml volumetric flask. The volume was made up with acetone and sonicated for 30 mins. Then, 1ml of the above solution was transferred to 10ml volumetric flask and the volume was made up to the mark with 0.1N HCL. The resulted solution was filtered through Whatman filter paper, suitably diluted and the drug content was estimated spectrophotometrically by measuring absorbance at 208nm” [5].

Table 1. Factor and Factor levels of Box-Behnken experimental design

| Independent factors                              | Levels            |
|--------------------------------------------------|-------------------|
|                                                   | Low   | Medium | High  |
| X1 = Drug: polymer ratio                          | 1:1   | 1:2    | 1:3   |
| X2 = Polymer: polymer ratio (Ethyl cellulose: Eudragit RS 100) | 1:1   | 1:2    | 1:3   |
| X3 = Plug weight (mg)                             | 50    | 75     | 100   |

Responses (Dependent factors)

Y1 = lag time in (hrs)
Y2 = percent release of methylphenidate after 8hours (%)

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Table 2. Formulations table according to Box Behnken experimental design

| Formulation code | Drug (mg) | Polymer (mg) | Ethyl cellulose (mg) | Eudragit RS 100 (mg) | HPMC K 100M Plug (mg) |
|------------------|-----------|--------------|----------------------|----------------------|-----------------------|
| F1               | 40        | 80           | 40                   | 40                   | 100                   |
| F2               | 40        | 120          | 40                   | 80                   | 100                   |
| F3               | 40        | 120          | 30                   | 90                   | 75                    |
| F4               | 40        | 80           | 26.66                | 53.33                | 75                    |
| F5               | 40        | 80           | 40                   | 40                   | 50                    |
| F6               | 40        | 40           | 13.33                | 26.66                | 100                   |
| F7               | 40        | 40           | 20                   | 20                   | 75                    |
| F8               | 40        | 40           | 13.33                | 26.66                | 50                    |
| F9               | 40        | 80           | 26.66                | 53.33                | 75                    |
| F10              | 40        | 80           | 26.66                | 53.33                | 75                    |
| F11              | 40        | 80           | 20                   | 60                   | 100                   |
| F12              | 40        | 40           | 10                   | 30                   | 75                    |
| F13              | 40        | 120          | 40                   | 80                   | 50                    |
| F14              | 40        | 80           | 20                   | 60                   | 50                    |
| F15              | 40        | 120          | 60                   | 60                   | 75                    |

2.7.3 Measurement of capsule lock length

Lock length of capsules was measured using screw gauge and the values were noted.

2.7.4 In-vitro dissolution studies

In-vitro drug dissolution studies were carried out using USP type-I (basket type) apparatus. The release of methylphenidate HCl from the pulsincap system was studied using three different dissolution media (900ml) in order to simulate pH changes across the GI tract. 0.1 N HCl (1.2 pH Buffer) for 2hrs, 4.5 pH phosphate buffer for 3hrs and 6.8 pH phosphate buffer. The rotating basket stirrer is set at a stirring speed of 100 rpm and a temperature is adjusted to 37 ± 0.5°C. Samples were withdrawn at predetermined time intervals of 0.5, 1, 1.5, 2, 3, 4, 5, 6, 7 and 8hrs and replaced with 5ml of fresh dissolution medium. The withdrawn samples were assayed at 208 nm for drug content using a UV visible spectrophotometer [5,16].

2.7.5 Stability studies

Pulsincap formulations were tested for their stability in amber colored bottle containers. Optimized formulation was stored at accelerated stability conditions (40°C ±2°C /75%± 5%RH) as per ICH guidelines over a period of 1 month in a humidity chamber and the capsules were evaluated for physical appearance, drug content, lag time and in-vitro drug release every week [17].

3. RESULTS AND DISCUSSION

3.1 Drug-excipient Compatibility Study

FTIR study was done to verify if there was any interaction between the pure drug and the various excipients employed in the study. FTIR spectrum of the pure drug Fig. 1 showed characteristics peaks at 1820-1660, 3400-2400, 1600 and 1475, 3400, 1375 representing presence of carbonyl group, acid, aromatic ring, amide, methyl group respectively. The peaks identified in the pure drug were relatively same when compared with the blend, indicating drug polymer compatibility.

DSC thermograms of pure drug showed a sharp peak at 225°C and in optimized formulation peak was observed at 211.2°C (Fig. 2). A small peak observed in optimized formulation at 268°C is due to the polymer ethyl cellulose used in the formulation which indicates no interaction between drug and excipients.

3.2 Evaluation of Empty Formaldehyde Cross Linked Capsules

Physical appearance of formaldehyde treated capsules showed no significant changes after exposing to formaldehyde vapours for 6 hours [5].

The solubility of capsule bodies was checked in orbital shaker bath and it was found that 15% v/v formaldehyde solution and 6 hours exposure
Time was optimum as the capsule bodies formed a palpable mass in 4 hours 30 mins, it was deformed in 5 hours 30 mins and remained intact up to 8 hours in 0.1 N HCL. Capsules bodies collected from the desiccator at the end of each hour were checked for their solubility and the observation is given Fig. 3 [7].

**Fig. 1. FTIR graph of pure drug and optimized formulation**

**Fig. 2. DSC of pure drug and optimized formulation**
3.3 Evaluation of the Hydrogel Plug

Hardness and thickness of hydrogel plugs of three different weights were measured and the plugs were evaluated for their lag time and swelling index. The results are given in Table 3. Results showed that at all polymer levels, drug release from the higher viscosity grade, K100M was slower as compared to the lower viscosity grade, K4M. The low viscosity grade HPMC was not found to be so effective to sustain the release of drugs from their matrices since medium can penetrate easily. Upon increasing the viscosity grade of the polymer, i.e., by using HPMC K100M plug the release rate has been found to be sustained. This is because when the viscosity of the polymer increases, the penetration of medium decreases due to the increased viscosity, resulting in delay in drug release. Hence, HPMC K100M plug was optimized. It is considered as the ideal hydrogel plug and was used in the final formulation [18].

3.4 Evaluation of Granules

Flow properties were determined and the results are given in Table 4. The results indicate that all the formulations showed excellent flow properties [12-14]. The percentage yield of all the formulations were calculated and it was observed that formulation F2 showed the highest percentage yield of 85.91% and formulation F6 showed the lowest percentage yield of 52.92%. The rest of the formulations showed percentage yield in the range of 60% to 80% [12]. Assay of all formulations was found to be in the range of 91.70±0.20 to 100.0±0.11% [13].

3.5 Evaluation of Pulsincap System

Weight variation, Content Uniformity, Measurement of capsule lock length was done and results are shown in Table 5. The results were within pharmacopeial limits.
Table 3. Evaluation of the hydrogel plug

|                      | HPMC K100M 50 mg | HPMC K100M 75 mg | HPMC K100M 100mg | HPMC K4M 50 mg | HPMC K4M 75 mg | HPMC K4M 100mg |
|----------------------|-----------------|------------------|------------------|---------------|---------------|---------------|
| Hardness (kg/cm²)    | 1.5±0.05        | 5.8±0.06         | 6.1±0.11         | 1.5±0.04      | 5.5±0.05      | 6±0.15        |
| Thickness (mm)       | 1±0.13          | 1.5±0.01         | 2±0.07           | 1±0.21        | 1.5±0.06      | 2±0.14        |
| Lag time (hrs)       | 2.4±0.01        | 3±0.20           | 4.1±0.12         | 1.5±0.09      | 2±0.12        | 2.8±0.05      |
| Swelling Index (%)   |                 |                  |                  |               |               |               |
| 0.1 N HCL            | 135.4±0.18      | 101±0.24         | 125±0.13         | 57.4±0.15     | 98±0.11       | 120±0.14      |
| 4.5 pH phosphate buffer | 97.8±0.17     | 109±0.13         | 170±0.15         | 79.8±0.21     | 90±0.19       | 114±0.11      |
| 6.8 pH phosphate buffer | 100.6±0.19   | 117±0.23         | 190±0.16         | 85.45±0.19    | 92±0.24       | 120±0.17      |

Table 4. Flow properties of granules

| Formulation | Bulk density (gm/ml) | Tapped density (gm/ml) | Carr’s index (%) | Hausners ratio | Angle of repose (º) |
|-------------|----------------------|------------------------|------------------|---------------|---------------------|
| F1          | 0.463±0.11           | 0.530±0.18             | 12.6±0.21        | 1.14±0.27     | 15.12±0.16          |
| F2          | 0.592±0.12           | 0.632±0.11             | 6.32±0.12        | 1.06±0.22     | 13.70±0.12          |
| F3          | 0.617±0.10           | 0.722±0.16             | 14.4±0.32        | 1.17±0.11     | 13.80±0.28          |
| F4          | 0.432±0.16           | 0.480±0.19             | 11.1±0.11        | 1.12±0.16     | 14.80±0.28          |
| F5          | 0.601±0.14           | 0.632±0.12             | 4.90±0.18        | 1.05±0.30     | 13.60±0.21          |
| F6          | 0.484±0.10           | 0.518±0.20             | 6.67±0.23        | 1.07±0.16     | 11.72±0.14          |
| F7          | 0.475±0.15           | 0.554±0.13             | 14.2±0.11        | 1.16±0.29     | 12.26±0.25          |
| F8          | 0.460±0.20           | 0.525±0.15             | 12.3±0.37        | 1.14±0.23     | 13.83±0.12          |
| F9          | 0.432±0.16           | 0.480±0.19             | 11.1±0.11        | 1.12±0.16     | 14.80±0.28          |
| F10         | 0.432±0.16           | 0.480±0.19             | 11.1±0.11        | 1.12±0.16     | 14.80±0.28          |
| F11         | 0.529±0.11           | 0.597±0.22             | 11.4±0.20        | 1.12±0.19     | 13.70±0.22          |
| F12         | 0.417±0.21           | 0.476±0.12             | 12.5±0.31        | 1.14±0.22     | 15.28±0.16          |
| F13         | 0.483±0.19           | 0.544±0.13             | 11.2±0.10        | 1.12±0.38     | 13.3±0.15           |
| F14         | 0.505±0.14           | 0.544±0.18             | 7.14±0.20        | 1.07±0.14     | 12.63±0.20          |
| F15         | 0.568±0.17           | 0.640±0.13             | 11.25±0.18       | 1.12±0.10     | 14.31±0.32          |

Table 5. Weight variation, content uniformity, capsule lock length of pulsincap formulation

| Formulation | Weight variation (mg) | Content Uniformity (%) | Capsule lock length (mm) |
|-------------|-----------------------|------------------------|--------------------------|
| F1          | 312.00±2.65           | 92.30±0.47             | 20±0.01                  |
| F2          | 372.67±3.21           | 91.00±0.50             | 19±0.03                  |
| F3          | 333.00±3.61           | 95.19±0.57             | 19.5±0.01                |
| F4          | 302.33±2.08           | 92.67±0.53             | 20±0.07                  |
| F5          | 273.67±4.93           | 98.93±0.49             | 19±0.02                  |
| F6          | 305.00±3.00           | 94.59±0.57             | 19±0.05                  |
| F7          | 288.33±2.08           | 100.6±0.70             | 19.2±0.01                |
| F8          | 275.67±4.16           | 101.23±0.44            | 20±0.01                  |
| F9          | 302.33±2.08           | 92.67±0.53             | 19.7±0.08                |
| F10         | 302.33±2.08           | 92.67±0.53             | 19±0.06                  |
| F11         | 328.00±2.65           | 104.3±0.61             | 20±0.02                  |
| F12         | 293.33±2.52           | 99.89±0.59             | 20±0.05                  |
| F13         | 294.00±2.65           | 97.25±0.71             | 20±0.01                  |
| F14         | 273.67±2.52           | 93.84±0.58             | 19±0.04                  |
| F15         | 336.33±2.52           | 95.45±0.38             | 19±0.01                  |
In-vitro drug release studies were conducted for all the 15 formulations obtained by applying experimental design. The % drug release of all the formulations is shown in Fig. 4. [5,16,19] It is observed that 100% of drug release was obtained for pure drug within 1 hour whereas for all the formulations lag time ranging from 2.4 hours to 4.2 hours was observed after which drug release was seen.

Formulations F1, F2 and F11 prepared using higher drug: polymer ratio of 1:2 & 1:3 and higher plug weight of 100mg of HPMC K100M, showed higher lag time of 4 to 4.2 hours and lower drug release at end of 8 hours whereas for other formulations F3, F4, F5, F6, F7, F8, F9, F10, F12, F13, F14 and F15 which were prepared using lower drug: polymer ratios of 1:1 and lower plug weights of 50 mg and 75 mg of HPMC K100M, it was found that at end of 8 hours there is higher amount of drug release and even the lag time was found to be less (2.4 to 3.5 hrs).

3.6 Optimization by Box-Behnken Design

Three independent variables (factors) X1 = Drug: polymer ratio, X2 = Polymer: polymer ratio (Ethyl cellulose: Eudragit RS 100) and X3 = Plug
weight were selected at three levels (low, medium, and high). Two dependent factors $Y_1 = \text{lag time in hrs}$ and $Y_2 = \text{percent release after 8 hrs}$ were selected.

It is observed that the $Y_1$ response, i.e., lag time followed three models: linear model, quadratic model, 2FI model whereas, $Y_2$ response for all formulations followed a linear model. On overall linear model is selected for both the responses as the suitable statistical model for formulation optimization since it is the suggested model by the design expert software. This is shown in Table 6.

Lack of fit is an undesirable characteristic for a model. If the model does not fit the data well, the test will show a significant lack of fit. For a well-fitted model, lack of fit will be insignificant ($P > 0.10$). In this case, lack of fit was insignificant, so the model fits the data generated. Fit summary given in Fig. 5 suggests linear model for both the responses by the design expert software [15].

A mathematical relationship between factors and responses were generated using multiple linear regression analysis in the form of equations. These equations represent the quantitative effect of variables ($X_1$, $X_2$, and $X_3$) and their interactions on the response $Y$. Coefficients with more than one factor term represent interaction terms whereas, those with higher order terms represent quadratic relationships. A positive sign represents a synergistic effect and a negative sign indicates an antagonistic effect [15].

Linear model equation for $Y_1$:
$$Y_1 = 3.1933 + 0.55X_1 + 0.15X_2 + 0.42X_3$$
Linear model equation for $Y_2$:
$$Y_2 = 86.89 - 9.47X_1 + 2.40X_2 - 0.73X_3$$

### Table 6. ANOVA summary for responses $Y_1$ and $Y_2$

| Response          | Model  | F-value | P-value | R squared |
|-------------------|--------|---------|---------|-----------|
| $Y_1$(lag time)   | Linear | 9.36    | 0.0023  | 0.7186    |
|                   | Quadratic | 7.07    | 0.0211  | 0.9272    |
|                   | 2FI    | 3.89    | 0.0404  | 0.7448    |
| $Y_2$(%drug release) | Linear | 1.48    | 0.27    | 0.2874    |

### Fit Summary

**Response 1: $R_1$ (Lag time)**

| Source | Sequential p-value | Lack of Fit p-value | Adjusted $R^2$ | Predicted $R^2$ |
|--------|--------------------|---------------------|----------------|-----------------|
| Linear | 0.0023             | 0.6418              | 0.4171         | Suggested       |
| 2FI    | 0.8428             | 0.5553              | -0.2761        |                 |
| Quadratic | 0.0790        | 0.7961              | -0.1653        |                 |
| Cubic  | 1.0000             |                     |                |                 |

**Response 2: $R_2$ (% drug release)**

| Source  | Sequential p-value | Lack of Fit p-value | Adjusted $R^2$ | Predicted $R^2$ |
|---------|--------------------|---------------------|----------------|-----------------|
| Linear  | 0.2742             | 0.0830              | -0.5056        | Suggested       |
| 2FI     | 0.6069             | -0.0039             | -1.9756        |                 |
| Quadratic | 0.5407            | -0.0811             | -5.1779        |                 |
| Cubic   | 1.0000             |                     |                |                 |

Fig. 5. Fit summary for responses $Y_1$ and $Y_2$
“To study the effect of independent variables on dependent variables 2-D contour plot and 3-D response surface analysis was done using software Design expert 11. These plots provide information about effect of two independent variables on one dependent variable at a time by keeping third independent variable at middle level” [20].

3.6.1 Effect on Lag time (R1)

“2-D contour plot (Fig. 6) shows that, as the level of X1 (Drug: Polymer ratio) was increased from -1 to 1 at centre level of X3 (Plug weight), the lag time increased from 2.6 to 3.8 hours and as X2 (Polymer: Polymer ratio) was increased from -1 to 1 at centre level of X3 (Plug weight), the lag time was found to be moderately affected. Response surface plot as shown in Fig. 7 depicts similar agonistic effect of X1 and X3 on lag time. Thus, it was observed that with the increase in Drug: Polymer ratio (X1) there was increase in Lag time and with the increase in Polymer: Polymer ratio (X2) the lag time was at moderate level. With further increase in X3 (Plug weight) i.e., at higher level of X3, the Lag time was observed to be increased up to 4.1 hours” [20].

3.6.2 Effect on % drug release (R2)

Fig. 8 depicts that as the level of X1 (Drug: Polymer ratio) was increased from -1 to 1 at centre level of X3 (Plug weight) the % drug release was found to be decreased from 95% to 80%. And with the increase in X2 (Polymer: Polymer ratio), the % drug release was found to be increased as seen through the contour plot. Thus, it was observed that with the increase in Drug: Polymer ratio (X1) there was decrease in % drug release whereas, with the increase in Polymer: Polymer ratio (X2) also the % drug release was increased. With further increase in X3 (Plug weight) i.e., at higher level of X3, the % drug release was observed to be decreased up to 75%. 3-D response surface plot as shown in Fig. 7 also shows similar declining trend of % drug release with increase in X1 (Drug: Polymer ratio) and X3 (Plug weight) [21].

One-way ANOVA was performed in order to determine the effect of factor on the responses. The results of the ANOVA were applied to identify insignificant factors. Values of Probability less than 0.0500 indicates factors are significant and have a significant effect on the responses [16,20]. One-way ANOVA for the factors A, B, C versus Lag time was performed. P value for Factor A Drug: Polymer ratio was found to be 0.017 which indicates significant effect on lag time whereas P value for Factor B Polymer: Polymer ratio and Factor C Plug weight was found to be 0.588 and 0.089 respectively, indicating insignificant factors on the response on lag time.

In the similar way, One-way ANOVA was performed for the factors A, B, C versus % Drug release in order to determine the effect of the factors on % Drug release. P value for Factor A Drug: Polymer ratio was found to be 0.042 which indicates significant effect on % Drug release whereas P value for Factor B Polymer: Polymer ratio and Factor C Plug weight was found to be 0.857 and 0.746 respectively, indicating insignificant factors on the response % drug release. From, this generated data, we can hypothesize that as the Drug: Polymer ratio increases, lag time increases and % drug release decreases.

The polynomial equation for dependent and independent variables was generated, the formulation was optimized for the responses. The optimum variables were obtained by the numerical analysis based on the criterion of desirability. The graphs obtained for Predicted vs. Actual responses are shown in Fig. 8. [21]. Based on the predicted values obtained for responses by considering highest lag time and least % drug release at the end of 8 hours, three formulations F1, F2 and F11 were selected and the in vitro dissolution studies were carried out again to confirm the validity of the optimization procedure and the results were given in the following Table 7. As the results obtained for in vitro dissolution studies were close to the predicted responses, it was proved that the design applied is significantly fitting the data and thus the design is validated.

The lag time of 4 to 5 hours and minimum % drug release at the end of 8 hrs from the pulsatile capsules was selected as target response for the optimization. Based on this, F11 was selected as optimized formulation as it showed the highest lag time of 4.2±0.27 hours and least % drug release of 72±0.09% at the end of 8 hours.

Model dependent kinetics of optimized Formulation F11 was found to follow zero order drug release kinetics and korsemeyer-peppas drug release mechanism.
Fig. 6. Contour plot for the effect of Drug: polymer ratio and Polymer: polymer ratio on Lag time and % Drug release at centre level of X3 (Plug weight).

Fig. 7. Response surface plot showing influence on Lag time and % Drug release.

Fig. 8. Predicted vs. Actual response for R1 (Lag time) and R2 (% Drug release)
Table 7. Validation

| Formulation | R1 (Lag time in hrs) | R2 (% drug release) |
|-------------|----------------------|---------------------|
|             | Predicted            | Obtained            | Predicted            | Obtained            |
| F1          | 3.468                | 3.2±0.19            | 83.76                | 81.79±0.23          |
| F2          | 4.16                 | 4.1±0.04            | 84.615               | 85.34±0.16          |
| F11         | 3.768                | 4.2±0.27            | 76.04                | 72±0.09             |

The Stability studies were conducted for one month. The formulation was found to be stable with no significant change in physical appearance, drug content, lag time and in-vitro drug release [17].

4. CONCLUSION

Methylphenidate HCl used in ADHD has been formulated as pulsatile drug delivery system using ‘pulsincap technique’. FTIR and DSC studies indicated drug & excipient were compatible. Formaldehyde treated capsule bodies of 6 hrs formaldehyde exposure time were optimized as they were found to be intact for up to 8 hours and their formaldehyde content was found to be within official limits (50mg/day). HPMC K100M and HPMC K4M were initially selected as hydrogel plugs as they are swellable hydrophilic polymers, from their evaluation, the results showed that HPMCK100M plug has higher lag time of up to 4 hours so, HPMC K100M plug was selected for experimental design. Percentage yield of granules was observed in the range of 52–85%. Assay of all the formulations were within the acceptable limits (95-100%). Flow properties were performed and it showed that all the formulations exhibited good to excellent flow properties. In-vitro dissolution was performed for all the formulations in comparison with the pure drug and the formulations F1, F2 and F11 showed the lag times of 3.2±0.19hrs, 4.1±0.04hrs & 4.2±0.27 hours and showed % drug release of 81.79±0.23%, 85.34±0.16% and 72±0.09% at the end of 8 hours respectively. The F11 formulation is the best optimized formulation as it showed the highest lagtime of 4.2 hours and least % drug release of 72% at the end of 8 hours. Drug excipient compatibility studies showed that the drug and excipients were compatible with each other. Stability studies were conducted for the optimized formulation F11 and it was found to be stable. In conclusion, this study illustrated that that pulsatile delivery of Class-I drug methylphenidate hydrochloride can be obtained by using highly retarding polymers like ethyl cellulose, eudragit RS100 and formaldehyde cross linked capsules.

DISCLAIMER

The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

CONSENT AND ETHICAL APPROVAL

It is not applicable.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

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