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Is a Higher Protein-Lower Glycemic Index Diet More Nutritious Than a Conventional Diet? A PREVIEW Sub-study

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High protein diets and low glycemic index (GI) diets have been associated with improved diet quality. We compared the changes in nutrient intakes of individuals at high risk of developing type-2 diabetes over 3 y who followed either a higher protein-lower GI diet (HPLG) or a conventional moderate protein-moderate GI diet (MPMG). This post hoc analysis included 161 participants with overweight and pre-diabetes from the Australian cohort of the PREVIEW study (clinical trial registered in https://www.clinicaltrials.gov/ct2/show/NCT01777893?term=NCT01777893&draw=2&rank=1) who were randomly assigned to a HPLG diet (25% energy from protein, dietary GI ≤ 50, n = 85) or a MPMG diet (15% energy from protein, dietary GI ≥ 56, n = 76). Food records were collected at 0-mo (baseline) and at 6-, 12-, 24-, and 36-mo (dietary intervention period). Linear mixed models were used to compare the differences in total energy, macro- and micronutrients, dietary GI, glycemic load (GL) and body weight between the two diet groups at the 4 dietary intervention time points. At 3 y, 74% participants from the HPLG diet and 74% participants from the MPMG diet completed the trial. The HPLG group showed significantly higher protein intake and lower dietary GI and GL than the MPMG group (group fixed effect $P < 0.001$ for all three parameters). By 6-, 12-, 24-, and 36-mo there was a 3.0, 2.7, 2.2, and 1.4% point difference in protein intake and 6.2, 4.1, 4.8, and 3.9 GI unit difference between the groups. The intake of energy and saturated fat decreased (mostly in the first 6-mo), while the intake of dietary fiber increased (from mo-0 to mo-12 only) in both diets, with no significant differences between the diets. The dietary intakes of zinc (group fixed effect $P = 0.05$), selenium ($P = 0.01$), niacin ($P = 0.01$), vitamin B12 ($P = 0.01$) and dietary cholesterol (group by time fixed effect $P = 0.001$) were higher in the HPLG group than in the MPMG group. Despite both diets being designed to be nutritionally complete, a HPLG diet was found to be more nutritious in relation to some micronutrients, but not cholesterol, than a MPMG diet.

Keywords: zinc, niacin, selenium, vitamin B12, dietary fiber, dietary cholesterol, pre-diabetes
INTRODUCTION

Weight gain resulting in overweight (BMI ≥ 25–30 kg/m\(^2\)) and obesity (BMI > 30 kg/m\(^2\)) is a major contributor to the risk of developing type 2 diabetes (T2D) (1). Sedentary behaviors and excessive dietary energy intake cause positive energy balance, which is the physiological mechanism behind weight gain (2). Therefore, in order to reduce the risk of chronic complications of obesity, it is critical for individuals with overweight to, firstly, decrease dietary energy intake to induce weight loss and, secondly, to maintain weight loss over time (3, 4).

Modifications in the quality of the habitual diet may also play a major role in the prevention of T2D and diabetes risk factors (5, 6). Diet quality is often defined as the ability to meet recommended nutrient intakes without excessive total energy consumption (7). Interestingly, Shay et al. showed that individuals with overweight and obese, who are at increased risk of developing T2D, had higher total energy intakes but consumed less nutritious foods compared to leaner people (8).

The current study was a secondary analysis of the Australian cohort from the PREVIEW (Prevention of diabetes through lifestyle intervention and population studies in Europe and around the World) intervention study, a large multinational study designed to assess the most effective lifestyle (diet and physical activity) strategy to prevent T2D in high-risk individuals (9). Based on the results from the DiOGenes (Diet, Obesity and Genes) study (10), which showed improved weight loss maintenance over 4-mo by following a HPLG diet, the PREVIEW trial hypothesized that a HPLG dietary intervention would be more effective than a conventional MPMG intervention in preventing T2D (9).

Both diets with a low dietary GI and/or low glycemic load (GL), and diets with higher percentage of total energy from protein have been previously found to be protective against T2D (6, 11). Additionally, low dietary GI and GL showed association with improved adherence to daily recommended nutrient intakes required for good health (12–14). This is likely due to some low GI foods (e.g., fruits, legumes, intact whole grains and dairy products) being particularly rich in micronutrients (13). Similarly, protein-dense foods have a high ratio of micronutrients to energy (15). Therefore, consuming a higher percentage of energy from protein-dense foods and lower dietary GI may enhance diet quality by optimizing nutrient adequacy without increasing the total energy intake (15). Improved diet quality may be a potential mechanism underlying the link between low dietary GI, higher protein intake and T2D prevention.

In the present study, we investigated the changes in nutrient intakes in a group of Australian individuals with overweight/obesity prescribed either a healthy HPLG diet or a healthy MPMG diet over a 3-y period. We hypothesized that a healthy HPLG dietary intervention would be associated with more positive changes in micronutrient density than a healthy MPMG dietary intervention.

SUBJECTS AND METHODS

This study was an analysis of the Australian data of the PREVIEW study, a 36-mo, multicenter, randomized clinical intervention trial. Fogelholm et al. have described the PREVIEW study protocol in full (9). Briefly, the PREVIEW study aimed to determine the effects of a healthy HPLG diet [protein 25% of energy intake (en%), carbohydrate (CHO) 45 en%, GI ≤ 50] and of a healthy MPMG diet (protein 15 en%, CHO 55 en%, GI ≥ 56) in combination with two exercise regimens (high-intensity or moderate-intensity exercise) on T2D prevention in males and females aged 25–70 y, with overweight (BMI ≥ 25 kg/m\(^2\)) and pre-diabetes (diagnosis based on the American Diabetes Association criteria) (16). The study comprised of a weight loss phase followed by a weight maintenance phase. During the 8-wk weight loss phase, the participants undertook a low energy diet (LED, 800 kcal/d) with the aim to lose ≥8% of 0-mo body weight to be eligible for the 34-mo weight maintenance phase.

The study participants attended 17 group sessions (8–20 individuals per group) delivered by research dietitians throughout both the weight loss phase and weight maintenance phase. With the support of a behavior modification tool, PREMIT (17), the group sessions provided information on how to adopt one of the two healthy intervention diets. In addition to the group counseling visits, dietary material with instructions on how to achieve the prescribed macronutrient composition and dietary GI, and recipes for each diet were provided to participants to encourage compliance.

In the current study, only individuals from The University of Sydney cohort of PREVIEW were included. Participants were compared based solely on their assigned intervention diet, providing two separate groups for the analyses. Data were collected between August 2013 and March 2018. Measurements from 0-, 6-, 12-, 24-, and 36-mo were used for the analyses of the nutritional assessments. Because the weight maintenance phase commenced after an 8-wk weight loss period, the data assessed at 6-, 12-, 24-, and 36-mo correspond to 4-, 10-, 22-, and 34-mo following one of the two dietary interventions.

All study participants provided written informed consent prior to commencing screening measurements. The study protocol (No X14-0408) was reviewed and approved by the Sydney Local Health District Human Research Ethics Committee—Royal Prince Alfred Hospital (Sydney, Australia) and is in accordance with the Declaration of Helsinki of 1975 as revised in 2008.

Reported intakes of energy, macro- and micronutrients and dietary GI and GL values were assessed at 0-, 6-, 12-, 24-, and 36-mo using 4-d (3 weekdays and 1 weekend day) food diaries. Food records were reviewed by research dietitians during the study to assess the adequacy of the information. Research dietitians entered the individual food records in Research dietitians were involved in the individual food records in FoodWorks Professional version 8 (Xyris Software, Brisbane, QLD, Australia, 2015) which contained the Australian food composition data (AUSNUT 2011–2013, AUSFOODS 2015 and AUSBRANDS 2015) and GI values (glucose = 100 scale). Missing GI values were obtained from the University of Sydney's online database (http://www.glycemicindex.com). Overall dietary GI
and GI were calculated as previously described by Louie et al. (18). Marine-sourced long-chain n-3 polyunsaturated fatty acids (LC n-3 PUFAs) daily reported intake was calculated as the sum of eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) daily reported intakes obtained from the FoodWorks Professional analysis.

Energy, macro- and micronutrients and GI and GL values used for the analyses were the mean of the 4-d food diaries. Body weight and height were measured as described in the PREVIEW study protocol (9). Body fat percentage (BF%) and fat free mass percentage (FFM%) were obtained using dual X-ray absorptiometry (DEXA) machine (Discovery W model, Hologic Inc, Bedford, MA, USA).

Nitrogen excretion was measured from 24-h urine collections and was used as a marker of protein intake. Total volume (mL) and weight (g) were measured within a few hours of delivery, then samples were frozen at −80°C in 0.8 mL aliquots (3 tubes). The urea content was analyzed on the Beckman Coulter AU480 Clinical Chemistry autoanalyzer (Beckman Coulter Inc, Brea, California, USA) in 100 µL urine samples in duplicate (average coefficient of variation 1.6%).

### STATISTICAL ANALYSIS

All data are expressed as means (± SD) for 0-mo characteristics, or adjusted means (95% CI). Intention-to-treat analyses were calculated by including all available observations (0- to 36-mo) from study participants who completed a 4-d food diary at 0-mo. Data were analyzed under the assumption that missing data were missing at random. Analyses of nutritional assessments and weight loss were conducted by using analysis of covariance (ANCOVA) linear mixed models. In the model, the dependent variables were the dietary intake and body weight values measured at 6-, 12-, 24-, 36-mo. The independent categorical variables (fixed factors) were the diet group (HPLG and MPMG), time (6-, 12-, 24-, and 36-mo), and the interaction between diet group and time. Time was treated as a categorical variable and an unstructured model was applied for covariance matrix between the repeated measurements. The models were adjusted for 0-mo values, age and sex. If the group main effect was significant, the overall differences (a mean of four timepoints) between groups were compared. If both the group main effect and group-by-time interaction were significant, then between group differences at each timepoint were tested. For the purpose of this analysis, the overall differences between timepoints (a mean of 2 groups) were not compared when the main effect of the time showed significance. To show compliance to the assigned dietary interventions, linear mixed models were also used to report the changes from baseline in en% protein, 24-h nitrogen excretion and dietary GI at 6-, 12-, 24-, and 36-mo [data represented as adjusted means (SE)]. A sensitivity analysis was conducted by the per-study protocol at 6-mo, which excluded dropout participants and participants who did not complete a 4-d food diary at 6-mo. In this analysis, the differences between the groups were assessed using ANCOVA adjusted for 0-mo values, age and sex. All statistical analyses were performed using IBM SPSS version 24 (IBM Corporation, New York, NY, USA, 2016). Statistical significance was determined by an α level of 0.05.

### RESULTS

Between August 2013 and February 2015, 169 participants were randomly assigned to one of the two dietary interventions (Figure 1). During the 0-mo assessments, 8 participants did not complete the 4-d food diary and were excluded from the intention-to-treat analysis set (Figure 1). Therefore, the analysis of 0-mo data included 161 individuals of whom 85 were assigned to the HPLG diet and 76 to the MPMG diet (Figure 1). During the LED phase, eight participants (six from the HPLG group and two from the MPMG group) did not lose ≥8% body weight, 10 participants (HPLG = 4, MPMG = 6) dropped out of the study and two participants from the HPLG developed T2D. These participants did not continue in the weight maintenance phase (8-wk to 36-mo), leaving 73 participants in the HPLG group and 68 participants in the MPMG group (Figure 1). In total, 104 subjects completed year 3 (74% of those who started the weight maintenance phase), of whom 54 participants were assigned to the HPLG diet (74% of those who started the weight maintenance period) and 50 to the MPMG diet (74%) (Figure 1).

Baseline characteristics of the participants included in the intention-to-treat analysis are summarized in Table 1. In both groups, there were more females than males (69% female in the HPLG group and 71% in the MPMG group), more Caucasians (HPLG 82%, MPMG 83%) than other ethnicities (self-reported) and more individuals in the age group 51–70 y (HPLG 63%, MPMG 62%) than in the age group 25–50 y. The average BMI was 36.3 kg/m² (class II obesity, World Health Organization classification) (19) and the average age was 53 years. Both body fat percentage (BF%) and fat free mass percentage (FFM%) were on average higher in the MPMG (44.4 and 56.2%, respectively) than in the HPLG group (41.7 and 55.6%, respectively). The percentage of energy contributed by protein was 19% protein and daily dietary GI was 54 (moderate, GI Foundation classification) (20) in both groups.

Both the HPLG group and the MPMG showed an overall increase from 0-mo (mean of 4 timepoints) in en% protein (Figure 2A). The main effect of the group was significant \( P < 0.001 \) indicating that the HPLG group consumed a significantly higher percentage of energy from protein than the MPMG group throughout the intervention. The HPLG group met the 25 en% of protein target only at the 6-mo timepoint and the MPMG group exceeded the 15 en% of protein target at all timepoints (Table 2). Dietary GI showed an overall decrease from 0-mo (mean of four timepoints) in both groups but it was significantly lower in the HPLG group than the MPMG group throughout the intervention (group fixed effect \( P < 0.001 \)) (Figure 2B).

There was a significant group-by-time effect \( P = 0.03 \) in 24-h nitrogen excretion (a marker of protein intake), but the main effect of the group did not reach statistical significance \( P = 0.16 \).
Screening

595 screened (447 females, 148 males)

401 excluded by the criteria
25 withdrew consent

Randomization

169 randomly assigned (119 females, 50 males)

89 assigned to HPLG diet and completed baseline measurements (63 females, 26 males)

80 assigned to MPMG diet and completed baseline measurements (56 females, 24 males)

0-mo

85 completed baseline food diary (59 females, 26 males)

76 completed baseline food diary (54 females, 22 males)

0-mo

4 dropped out during LED
6 did not lose 8% of baseline bodyweight
2 diagnosed with T2D

6 dropped out during LED
2 did not lose 8% of baseline bodyweight

8-wk

73 eligible to start weight maintenance phase (54 females, 22 males)

68 eligible to start weight maintenance phase (48 females, 20 males)

16 dropped out during weight maintenance phase
3 diagnosed with T2D

16 dropped out during weight maintenance phase
2 diagnosed with T2D

36-mo

54 completed 36-mo intervention (37 females, 17 males)

50 completed 36-mo intervention (33 females, 17 males)

FIGURE 1 | Screening, randomization and follow-up of study participants. Intention-to-treat analysis included all participants who were randomly assigned to the HPLG group or MPMG group and completed the food record during the baseline assessments. HPLG, higher protein-lower glycemic index; MPMG, moderate protein-moderate glycemic index.
TABLE 1 | Baseline characteristics of the intention-to-treat population (n = 161)*.

|                      | HPLG group  | MPMG group  |
|----------------------|-------------|-------------|
|                      | (n = 89)    | (n = 76)    |
| **Ethnicity**        |             |             |
| Caucasian            | 70 (82%)    | 63 (83%)    |
| Asian                | 6 (7%)      | 5 (7%)      |
| Other                | 9 (11%)     | 8 (10%)     |
| **Sex**              |             |             |
| Females              | 59 (69%)    | 54 (71%)    |
| Males                | 26 (31%)    | 22 (29%)    |
| **Age, years**       |             |             |
| 25–50                | 31 (37)     | 29 (38)     |
| 51–70                | 54 (63)     | 47 (62)     |
| **Screening height, cm** | 167.1 (8.3) | 167.9 (8.8) |
| **Weight, kg**       | 99.1 (18.6) | 102.2 (21.7) |
| **BMI, kg/m²**       | 36.3 (7.1)  | 36.3 (7.2)  |
| **BF , %**           | 41.7 (11.2) | 44.4 (14.4) |
| **FFM, %**           | 55.6 (11.0) | 56.2 (10.8) |
| **Energy intake, kJ/d** | 9,292 (2,640) | 9,165 (2,712) |
| **Protein, en%**     | 19.5 (3.5)  | 19.7 (4.7)  |
| **Fat, en%**         | 34.3 (5.7)  | 35.0 (6.3)  |
| **Saturated fat, en%** | 11.9 (2.9)  | 12.6 (3.1)  |
| **Carbohydrate, en%** | 40.3 (7.0)  | 40.2 (7.2)  |
| **Dietary fiber, g** | 26.7 (7.8)  | 26.0 (7.4)  |
| **Dietary GL**       | 54.1 (5.1)  | 54.3 (4.5)  |
| **Dietary GL**       | 119.1 (42.3)| 118.4 (45.0)|
| **Alcohol, en%**     | 3.5 (4.8)   |             |

*Values are expressed as n (%) for ethnicity, sex and age and as mean (SD) for all other measurements. BF, body fat; FFM, fat free mass; HPLG, higher protein-lower glycemic index; MP MG, moderate protein-moderate glycemic index.

(Figure 3). Nitrogen excretion (24-h), was higher in the HPLG group than in the MP MG group at 6-, 12-, and 24-mo, but higher in the MP MG than in the HPLG at 36-mo. Table 2 shows body weight and the reported intake of macronutrients, dietary GL, dietary GL, Zinc, Selenium, Niacin and Vitamin B12 at 0-, 6-, 12-, 24-, and 36-mo in the intention-to-treat population. Body weight and energy intake decreased in both the HPLG and MPLG diet groups (with the greatest difference between the groups at 6-mo) and there were no significant differences between the groups during the intervention (group fixed effect $P = 0.91$ and $P = 0.84$, respectively). Fixed effects of the group were significant for the changes in the groups’ absolute intake (g/d) of protein ($P < 0.001$), en% carbohydrate ($P < 0.001$), absolute intake (g/d) of carbohydrate ($P = 0.009$), starch ($P < 0.001$) and dietary GL ($P < 0.001$) (Table 2). There was a significant group-by-time effect ($P = 0.02$) but not a significant group effect ($P = 0.52$) for dietary fiber, with both diets showing an increase in intake from 0- to 12-mo. At 12-mo, the increase in dietary fiber from 0-mo was greater in the HPLG group than in the MP MG group but the difference in intake was not significant between the diets.

The percentage of energy contributed by total fat and by saturated fat showed a decrease in both groups at 6-mo, but the decrease was attenuated at 12-, 24-, and 36-mo. The main effect of the group for the en% total fat and en% saturated fat did not reach statistical significance ($P = 0.97$ and $P = 0.84$, respectively). At 6-mo, there was a significant difference ($P < 0.001$) in groups’ cholesterol intake (adjusted means: HPLG 350.2 mg (95% CI: 322.8, 377.5), MP MG 225.8 mg (196.7, 254.8)). The differences remained statistically significant ($P < 0.05$), although they were less pronounced at 12-, 24-, and 36-mo. The fixed effect of the group was significant for en% alcohol ($P = 0.03$) but not the group-by-time interactions, therefore the effect of the group did not differ between timepoints (Table 2).

Zinc, selenium, niacin and vitamin B12 showed significant group effects ($P = 0.05$ for zinc and $P = 0.01$ for the other three micronutrients), with the HPLG group overall reportedly consuming higher intakes of these micronutrients than the MP MG group. However, the group fixed effects were not significant suggesting that the effect of the group did not differ between timepoints.

Supplementary Tables 1, 2 show the micronutrient (minerals and vitamins, excluding zinc, selenium, niacin and vitamin B12) reported intakes at 0-, 6-, 12-, 24-, and 36-mo. Calcium, iron, potassium, sodium, phosphorus, magnesium, iodine, thiamine, riboflavin, vitamin B6, dietary folate equivalent (DFE), vitamin A, vitamin C and vitamin E did not show any significant group-by-time effects nor group effects.

Compared to the intention-to-treat analysis, in the perstudy protocol analysis, the HPLG group showed a greater consumption of $\alpha$-linolenic acid (ALA) and of LC n-3 PUFA at 6-mo than the MP MG group (Supplementary Table 3). However, significant differences were no longer found between the groups’ reported intakes of zinc and niacin (Supplementary Table 3).

**DISCUSSION**

To the best of our knowledge, this is the first study comparing the nutrient density of a healthy HPLG diet and of a healthy MP MG diet prescribed for a 3-y period. We found that, with comparable reductions in energy intake between the two groups during the intervention, individuals following the HPLG diet consumed higher intakes of some micronutrients than those assigned to the MP MG diet. Specifically, the HPLG group had higher intakes of zinc, selenium, niacin and vitamin B12 compared to the MP MG group. These results support our hypothesis that a HPLG diet is more micronutrient-dense than a conventional diet. Conversely, participants in the MP MG group showed lower intake of dietary cholesterol than those prescribed to the HPLG diet.

Previous studies have reported that higher protein diets and lower GI/GL diets improved diet quality and protection against chronic conditions (13, 15, 21, 22). Zinc, selenium, niacin and vitamin B12, for which reported intake was higher in the HPLG group compared to the MP MG group, are predominantly found in animal sources of protein. Some studies have linked high protein, particularly animal protein to increased risk of chronic diseases, especially when the protein sources are of poor quality (23). Since the en% contributed by total fat and saturated fat decreased during the intervention, it suggests that the HPLG...
intervention achieved the prescribed 25 en% from protein by favoring the consumption of lean meats (e.g., skinless poultry and fish) to meats higher in saturated fat, as recommended. This was also supported by the tendency to increased reported intake of dietary LC n-3 PUFA (predominantly found in oily fish) in the HPLG group compared to the MPMG group until the 2-y analysis. The consumption of lean animal sources of proteins has been previously linked with increased nutrients of particular concern, including vitamin B12, zinc, and selenium (15). In the PREVIEW study Australia, the majority of participants (63 and 62% in the HPLG and MPMG group, respectively) were aged 51–70 y, consistent with the higher prevalence of pre-diabetes in the older population. Older adults have increased risk of vitamin B12 deficiency due to the reduced physiological ability to absorb vitamin B12 with aging (24). Moreover, poor zinc status has been linked with decreased insulin secretion and insulin insensitivity (25) and low blood selenium concentrations with an increased incidence of cardiovascular disease (CVD) (26). Therefore, in the long-term, individuals with pre-diabetes may particularly benefit from a HPLG diet which is more likely to provide adequate intakes of vitamin B12 [Estimated Average Requirements (EAR) for Australia and New Zealand = 2.0 µg/day for both males and females] (27), zinc (EAR = 12 and 6.5 mg/day for males and females, respectively) (28) and selenium (EAR = 60 and 50 µg/day for males and females, respectively) (29).

Dietary cholesterol, which is abundant in protein-dense foods, such as eggs, and low GI foods such as dairy products, was not surprisingly higher in the HPLG intervention. Despite some evidence suggesting that dietary cholesterol increases the risk of CVD (30, 31), the link between dietary cholesterol and cardiometabolic health remains controversial (32). Furthermore, any potential cardiovascular risk derived from higher dietary cholesterol in the HPLG diet may have been attenuated by the greater increase in dietary fiber observed at 12-mo and 36-mo in the HPLG group compared to the MPMG group and by the reduction in en% contributed by saturated fat in both dietary groups. Indeed, dietary fiber has been proposed to inhibit the absorption of cholesterol and improve control of cholesterol concentration (33) and a lower intake of saturated fat has shown to be cardioprotective for people with pre-diabetes, a population at higher risk of cardiovascular complications (34).

While the differences in dietary protein, carbohydrate, GI and GL between the two groups were statistically significant, by 3 y, the participants were not achieving the targeted separation of 10% points for protein and carbohydrate reported intake and of 6 GI units between the two interventions. This is related to the MPMG group consuming more protein (5 percentage points higher intake) and less dietary GI (2 GI units) than the targeted 15 en% from protein and dietary GI ≥ 56. In both groups, the baseline values of protein (∼20 en%) were higher and the baseline values of carbohydrate (∼40 en%) were lower than the averages observed in the 2011–2012 Australian National Nutrition and Physical Activity Survey (protein 18–19 en% and carbohydrate 42–45 en%) (35). This pattern was consistent with that of a moderately high protein-lower carbohydrate diet. Therefore, more effort was required by individuals in the MPMG group to make changes to their habitual diet, which may have accounted for the failure of the group to comply with the prescribed dietary targets. Furthermore, the GI concept is well-known in Australia due to efforts of non-governmental organizations to educate the population about GI (20). “Low-GI” labeled foods are also widely available in supermarkets (36). For these reasons, individuals in the MPMG group may have been less willing to choose higher GI food options to increase their dietary GI. These findings should be considered in the planning of future studies.

Surprisingly, the en% from alcohol reported intake decreased significantly more in the MPMG group throughout the weight maintenance phase, despite both groups receiving the same
| Measure and Group | 0-mo | 6-mo | 12-mo | 24-mo | 36-mo | P-value for Fixed Effects<sup>d</sup> | Group | Time | Group by Time |
|------------------|------|------|-------|-------|-------|-------------------------------|------|------|---------------|
| **Weight, kg**    |      |      |       |       |       |                               |      |      |               |
| HPLG             | 99.1 (18.6) | 87.1 (85.9, 88.3) | 89.4 (87.7, 91.1) | 93.0 (91.1, 94.9) | 93.7 (92.0, 95.4) | 0.91 | <0.001 | 0.63 |
| MPMG             | 102.2 (21.7) | 87.3 (86.0, 88.6) | 89.9 (88.1, 91.7) | 92.6 (90.5, 94.7) | 93.8 (92.0, 95.6) |      |      |               |
| **Energy, kJ/d**  |      |      |       |       |       |                               |      |      |               |
| HPLG             | 9,296 (2,478) | 6,872 (6,523, 7,221) | 7,230 (6,832, 7,629) | 7,274 (6,749, 7,799) | 7,127 (6,685, 7,569) | 0.84 | 0.11 | 0.96 |
| MPMG             | 9,145 (2,472) | 6,921 (6,550, 7,292) | 7,167 (6,731, 7,602) | 7,143 (6,579, 7,708) | 7,078 (6,618, 7,539) |      |      |               |
| **Protein, g/d**  |      |      |       |       |       |                               |      |      |               |
| HPLG             | 19.5 (3.5) | 25.2 (24.2, 26.2) | 23.4 (22.5, 24.2) | 23.4 (22.4, 24.5) | 23.3 (22.1, 24.6) | <0.001 | <0.001 | 0.48 |
| MPMG             | 19.7 (4.7) | 22.0 (21.0, 23.1) | 20.6 (19.5, 21.6) | 21.1 (19.9, 22.2) | 21.7 (20.4, 23.1) |      |      |               |
| **CHO, g/d**     |      |      |       |       |       |                               |      |      |               |
| HPLG             | 104.8 (30.3) | 101.3 (96.6, 105.9) | 98.1 (93.1, 103.1) | 97.2 (91.9, 102.6) | 95.7 (89.7, 101.7) | <0.001 | 0.28 | 0.53 |
| MPMG             | 104.0 (35.9) | 88.3 (83.4, 93.2) | 85.7 (80.3, 91.2) | 86.6 (80.9, 92.3) | 89.0 (82.7, 95.3) |      |      |               |
| **Fat, g/d**     |      |      |       |       |       |                               |      |      |               |
| HPLG             | 34.3 (5.7) | 31.0 (29.5, 32.5) | 33.1 (31.6, 34.7) | 33.4 (31.8, 35.1) | 33.5 (31.7, 35.3) | 0.97 | <0.001 | 0.55 |
| MPMG             | 34.9 (6.3) | 30.4 (28.8, 32.0) | 32.6 (30.9, 34.3) | 33.1 (31.4, 34.9) | 34.8 (33.0, 36.7) |      |      |               |
| **Saturated fat, g/d** | 11.9 (2.8) | 10.1 (9.4, 10.8) | 10.5 (9.8, 11.2) | 10.8 (10.1, 11.8) | 10.9 (10.0, 11.7) | 0.84 | 0.01 | 0.55 |
| **Starch, g/d**  |      |      |       |       |       |                               |      |      |               |
| HPLG             | 127.0 (45.0) | 76.4 (69.3, 83.4) | 83.8 (77.1, 90.5) | 80.5 (72.9, 88.2) | 83.3 (74.8, 91.8) | <0.001 | 0.27 | 0.12 |
| MPMG             | 122.8 (46.7) | 100.3 (92.8, 107.8) | 100.7 (93.4, 108.1) | 97.9 (89.4, 105.9) | 90.0 (81.4, 105.9) |      |      |               |
| **Sugars, g/d**  |      |      |       |       |       |                               |      |      |               |
| HPLG             | 89.0 (36.1) | 69.4 (63.5, 75.3) | 70.8 (65.4, 76.1) | 68.7 (63.1, 74.2) | 67.8 (60.9, 74.7) | 0.82 | 0.34 | 0.89 |
| MPMG             | 91.4 (36.6) | 72.3 (66.0, 78.7) | 72.4 (66.6, 78.3) | 68.3 (62.3, 74.2) | 66.6 (59.5, 73.8) |      |      |               |
| **Dietary fiber, g/d** | 26.7 (7.7) | 26.7 (24.9, 28.5) | 28.8 (26.9, 30.8) | 25.2 (23.3, 26.9) | 26.4 (24.8, 28.0) | 0.53 | <0.001 | 0.02 |
| **Dietary GI**   |      |      |       |       |       |                               |      |      |               |
| HPLG             | 54.1 (5.6) | 47.1 (45.9, 48.3) | 48.9 (47.8, 50.0) | 49.4 (48.3, 50.4) | 49.4 (47.9, 50.8) | <0.001 | 0.04 | 0.10 |
| MPMG             | 54.3 (4.5) | 53.3 (52.0, 54.7) | 52.7 (51.4, 53.9) | 54.1 (53.0, 55.2) | 53.2 (51.6, 54.7) |      |      |               |
| **Cholesterol, mg/d** | 119.1 (42.3) | 70.7 (64.7, 76.7) | 76.6 (70.8, 82.3) | 74.8 (68.8, 81.0) | 76.9 (69.5, 84.3) | <0.001 | 0.49 | 0.07 |
| HPLG             | 118.4 (45.0) | 92.6 (86.2, 99.0) | 91.7 (85.4, 97.9) | 90.7 (84.1, 97.4) | 83.7 (76.1, 91.3) |      |      |               |
| **MUFAs, g/d**   |      |      |       |       |       |                               |      |      |               |
| HPLG             | 381.5 (178.5) | 352.0 (322.8, 377.5)** | 315.9 (286.3, 345.5)** | 337.3 (296.2, 278.4)** | 320.2 (282.2, 358.2)** | 0.01 | 0.34 | 0.001 |
| MPMG             | 363.3 (172.1) | 225.8 (196.7, 254.8)** | 264.9 (232.4, 297.4)** | 282.1 (238.1, 326.1)** | 294.8 (254.8, 335.0)** |      |      |               |
| **PUFAs, g/d**   |      |      |       |       |       |                               |      |      |               |
| HPLG             | 34.4 (13.0) | 22.9 (20.9, 24.9) | 26.9 (24.6, 29.2) | 26.5 (24.1, 29.0) | 26.3 (23.6, 29.1) | 0.96 | <0.001 | 0.58 |
| MPMG             | 33.5 (12.7) | 23.0 (20.9, 25.1) | 25.5 (23.0, 28.0) | 26.5 (23.9, 29.2) | 27.9 (25.0, 30.9) |      |      |               |

(Continued)
### Table 2 | Continued

| Measure and Group | 0-mo | 6-mo | 12-mo | 24-mo | 36-mo | p-value for Fixed Effects^d |
|-------------------|------|------|-------|-------|-------|---------------------------|
| **Group** | **Time** | **Group by Time** |
| **LC n-3 PUFA, mg/d** | | | | | | |
| HPLG | 436.9 (491.8) | 647.2 (506.9, 787.5) | 521.8 (398.2, 645.3) | 553.9 (403.0, 704.9) | 418.0 (271.3, 564.7) | 0.08 | 0.52 | 0.11 |
| MPMG | 393.1 (490.0) | 404.9 (255.5, 554.2) | 425.8 (289.7, 561.9) | 372.7 (209.9, 535.4) | 442.6 (287.6, 597.4) | | | |
| **LA, g/d** | | | | | | |
| HPLG | 11.9 (4.7) | 8.6 (7.8, 9.5) | 9.4 (8.3, 10.4) | 9.2 (8.2, 10.2) | 9.0 (8.1, 9.9) | 0.53 | 0.01 | 0.59 |
| MPMG | 11.5 (5.0) | 8.3 (7.4, 9.2) | 9.8 (8.7, 10.9) | 9.9 (8.9, 11.0) | 9.3 (8.4, 10.3) | | | |
| **ALA, g/d** | | | | | | |
| HPLG | 1.7 (0.7) | 1.6 (1.4, 1.9) | 1.7 (1.4, 1.9) | 1.5 (1.3, 1.7) | 1.5 (1.3, 1.7) | 0.07 | 0.69 | 0.20 |
| MPMG | 1.9 (1.3) | 1.2 (1.0, 1.5) | 1.4 (1.2, 1.7) | 1.4 (1.2, 1.6) | 1.4 (1.2, 1.6) | | | |
| **Alcohol, %** | | | | | | |
| HPLG | 3.5 (4.8) | 3.5 (2.6, 4.4) | 2.9 (2.1, 3.8) | 3.8 (2.5, 5.2) | 3.4 (2.3, 4.5) | 0.03 | 0.46 | 0.91 |
| MPMG | 3.9 (5.6) | 2.0 (1.0, 2.9) | 1.9 (1.0, 2.8) | 2.5 (1.1, 4.0) | 2.2 (1.1, 3.4) | | | |
| **Zinc, mg/d** | | | | | | |
| HPLG | 12.1 (4.1) | 11.7 (10.7, 12.6) | 11.7 (10.9, 12.4) | 11.0 (10.1, 11.8) | 11.2 (10.3, 12.1) | 0.05 | 0.23 | 0.44 |
| MPMG | 12.4 (5.1) | 10.9 (10.0, 11.9) | 10.3 (9.4, 11.1) | 10.0 (9.1, 10.9) | 10.8 (9.9, 11.8) | | | |
| **Selenium, µg/d** | | | | | | |
| HPLG | 108.1 (42.7) | 107.8 (94.5, 121.1) | 92.4 (84.4, 100.3) | 96.2 (88.8, 103.6) | 99.5 (85.7, 113.3) | 0.01 | 0.53 | 0.34 |
| MPMG | 100.9 (38.0) | 88.0 (73.8, 102.1) | 88.1 (79.5, 96.7) | 85.5 (77.3, 93.6) | 81.6 (66.9, 96.2) | | | |
| **Niacin (NE), mg/d** | | | | | | |
| HPLG | 26.4 (9.8) | 25.9 (23.7, 28.1) | 24.5 (22.7, 26.4) | 24.2 (22.4, 25.9) | 25.4 (23.0, 27.7) | 0.01 | 0.38 | 0.72 |
| MPMG | 26.0 (10.5) | 23.0 (20.6, 25.3) | 21.9 (19.9, 23.9) | 22.1 (20.2, 24.0) | 21.2 (18.8, 23.7) | | | |
| **Vitamin B12, µg/d** | | | | | | |
| HPLG | 5.3 (2.7) | 5.6 (5.1, 6.1) | 5.2 (4.7, 5.6) | 5.0 (4.5, 5.5) | 5.1 (4.4, 5.8) | 0.01 | 0.17 | 0.71 |
| MPMG | 5.4 (3.7) | 4.6 (4.1, 5.2) | 4.4 (3.9, 4.8) | 4.4 (3.8, 4.9) | 4.5 (3.8, 5.3) | | | |

^aValues are expressed as the mean (SD) or adjusted mean (95% CI). The adjusted means with a double asterisk or single asterisk indicate a statistically significant difference between the groups at the timepoint (**P < 0.001; *P < 0.05). GI, glycemic index; GL, glycemic load; HPLG, higher protein-lower glycemic index diet; MPMG, moderate protein-moderate glycemic index diet; NE, niacin equivalent.

^bNumber of participants 85, 68, 64, 60 and 54 at 0-, 6-, 12-, 24-, and 36-mo, respectively.

^cNumber of participants 76, 59, 52, 50 and 50 at 0-, 6-, 12-, 24-, and 36-mo, respectively.

^dP-values show the significance of the fixed effects for changes in a variable as assessed by linear mixed model adjusted for each measurement 0-mo values, age and sex.

Advice regarding limiting alcohol consumption. However, at 3 y, alcohol intake was still below 5% of dietary energy intake in both groups as recommended by the National Health and Medical Research Council of Australia to avoid weight gain, nutritional inadequacy and harm from alcohol consumption (37).

A notable strength of our study was the use of a behavior modification tool specifically developed for the PREVIEW trial (PREMIT, the PREVIEW behavior Modification Intervention Toolbox) (17). Because the two intervention diets were designed to be healthy, it is plausible that the behavior modification tool by itself was effective in imparting more positive nutrition habits in both groups. This is evidenced by the overall reduced energy intake, maintained weight loss and nutritional adequacy achieved by the entire cohort.

Food diary entries were reviewed by research dietitians with the study participants. This added strength to our study because it improved completeness of the food diaries and ensured that evident mistakes were corrected before the data were entered into the nutritional database. Other major strengths include the randomized controlled intention-to-treat design, which minimized the potential of selection bias, and the long dietary intervention period (36-mo). There were similar dropouts in both diet arms. Indeed, any modification in the habitual diet is considered successful when implemented in the long-term.

This study also has some limitations. The MPMG group did not meet the protein, carbohydrate and dietary GI targets of the diet. Higher dietary adherence may have enabled stronger conclusions to be made. Despite 4-d food diaries being reasonably accurate, self-reported methods to measure nutritional intake are prone to under- and over-reporting and any food composition database is limited by inherent variation (38). However, in this study, measurements of urinary nitrogen excretion (higher in the HPLG group at 6-, 12-, and 24-mo) confirmed the differences in protein reported intake between the interventions when the compliance to the assigned diets was reported higher in 4-d food diaries.
(a strength). Finally, it could be argued that an increase in protein from 20 to 25 en% in the HPLG diet may/may not be clinically relevant to whole body amino acid and nitrogen maintenance.

In conclusion, in the Australian context, long-term adherence to a HPLG diet—denser in zinc, selenium, niacin and vitamin B12—may offer greater nutritional benefit than a conventional MPMG diet to individuals with pre-diabetes and may be linked with chronic disease prevention. Future research is needed to investigate the effects of improved diet quality through higher protein intake and lower dietary GI on biomarkers of optimum health. The environmental impacts of consuming diets with varying amounts and sources of protein must also be considered (39).

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Materials, further inquiries can be directed to the corresponding author/s.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Sydney Local Health District Human Research Ethics Committee – Royal Prince Alfred Hospital (Sydney, Australia). The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

AM, RM, FA, and JB-M: designed this research. AR and MF: designed the protocol for the overall PREVIEW intervention study. AM and RM: conducted the research and analyzed the data. AM: performed the statistical analysis, wrote the manuscript, and has primary responsibility for the final content. All authors contributed to the article and approved the submitted version.

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REFERENCES

1. Roglic G. World Health Organization. Global Report on Diabetes. Geneva: World Health Organization (2016). 86 p.

2. Williams EP, Moshfegh A, Bookstaver P, et al. Overweight and obesity: prevalence, consequences, and causes of a growing public health problem. Curr Obes Rep. (2015) 4:363–70. doi: 10.1007/s13679-015-0169-4

3. Paizczuk C, Dias CM, Jorge R, Carraca EV, Yannakoulia M, de Zwaan M, et al. Successful weight loss maintenance: a systematic review of weight control registries. Obes Res. (2020) 21:e13003. doi: 10.1111/obr.13003

4. Jansen MD, Ryan DH, Apovian CM, Ard JD, Comuzzie AG, Donato KA, et al. 2013 AHA/ACC/TOS guideline for the management of overweight and obesity in adults: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines and The Obesity Society. J Am Coll Cardiol. (2014) 63(25 Pt B):2985–3023. doi: 10.1016/j.jacc.2013.11.004

5. Bhupathiraju SN, Tobias DK, Malik VS, Pan A, Hruby A, Manson JE, et al. Glycemic index, glycemic load, and risk of type 2 diabetes: results from 3 large US cohorts and an updated meta-analysis. Am J Clin Nutr. (2014) 100:218–32. doi: 10.3945/ajcn.113.079533

6. Lwosey G, Taylor R, Livesey HF, Buyen AE, Jenkins D JA, Augustin LSA, et al. Dietary glycemic index and load and the risk of type 2 diabetes: a systematic review and updated meta-analyses of prospective cohort studies. Nutrients. (2019) 11:1280. doi: 10.3390/nu11061280

7. Freeland-Graves JH, Nitzke S, Academy of Nutrition and Dietetics. Position of the academy of nutrition and dietetics: total diet approach to healthy eating. J Acad Nutr Diet. (2013) 113:307–17. doi: 10.1016/j.jand.2012.12.013

8. Shay CM, Van Horn L, Stampler J, Dyer AR, Brown JJ, Chan Q, et al. Food and nutrient intake and their associations with lower BMI in middle-aged US adults: the International Study of Macro-/Micronutrients and Blood Pressure (INTERMAP). Am J Clin Nutr. (2012) 96:483–91. doi: 10.3945/ajcn.112.050256

9. Fogelholm M, Larsen TM, Westerterp-Plantenga M, Macdonald I, Martinez JA, Boyadjieva N, et al. PREVIEW: prevention of diabetes through lifestyle intervention and population studies in Europe and around the World. Design, methods, and baseline participant description of an adult cohort enrolled into a three-year randomised clinical trial. Nutrients. (2017) 9:632. doi: 10.3390/nu9060632

10. Larsen TM, Dalskov SM, van Baak M, Jebb SA, Papadaki A, Pfeiffer AF, et al. Diets with high or low protein content and glycemic index for weight-loss maintenance. N Engl J Med. (2010) 363:2102–13. doi: 10.1056/NEJMoa1007137

11. Rodríguez NR. Introduction to Protein Summit 2.0: continued exploration of the impact of high-quality protein on optimal health. Am J Clin Nutr. (2015) 101:1317S–95. doi: 10.3945/ajcn.114.083980

12. Louie JC, Godyn J, Turner N, Cochrane J, Wiggers J, Flood V. Dietary glycemic index and glycemic load among indigenous and non-indigenous children aged 10–12 years. Nutrition. (2012) 28:e14–22. doi: 10.1016/j.nut.2011.12.007

13. Louie JC, Markovic TP, Ross GP, Foote D, Brand-Miller JC. Higher glycemic load diet is associated with poorer nutrient intake in women with gestational diabetes mellitus. Nutr Res. (2013) 33:259–65. doi: 10.1016/j.nutres.2013.02.008

14. Goletzke J, Buyen AE, Louie JC, Moses RG, Brand-Miller JC. Dietary micronutrient intake during pregnancy is a function of carbohydrate quality. Am J Clin Nutr. (2015) 102:626–32. doi: 10.3945/ajcn.114.104836

15. Phillips SM, Fulgoni VL III, Heaney RP, Nicklas TA, Slavin JL, Weaver CM. Commonly consumed protein foods contribute to nutrient intake, diet quality, and nutrient adequacy. Am J Clin Nutr. (2015) 101:13465–52S. doi: 10.3945/ajcn.114.084079

16. Mayer-Davis EJ, Sparks KC, Hirst K, Costacou T, Lovejoy JC, Regensteiner JG, et al. Dietary intake in the diabetes prevention program cohort: baseline and 1-year post randomization. Ann Epidemiol. (2004) 14:763–72. doi: 10.1016/j.annepidem.2004.02.004

17. Kuhlert D, Unyi-Reichzer A, Stratton G, Meinert Larsen T, Fogelholm M, Raben A, et al. PREVIEW behavior modification intervention toolbox (PREMIT): a study protocol for a psychological element of a multicenter project. Front Psychol. (2016) 7:1136. doi: 10.3389/fpsyg.2016.01136

18. Louie JCY, Flood VM, Atkinson FS, Barclay AW, Brand-Miller JC. Methodology for assigning appropriate glycaemic index values to an Australian food composition database. J Food Compos Anal. (2015) 38:1–6. doi: 10.1016/j.jfca.2014.06.002

19. World Organization. Obesity. Available online at: https://www.who.int/topics/obesity/en/ (accessed August 18, 2020).

20. Glycemic Index Foundation. What is GI? Available online at: https://www.glycsymbol.com/about/ (accessed August 18, 2020).

21. Louie JC, Buyen AE, Brand-Miller JC, Flood VM. The link between dietary glycemic index and nutrient adequacy. Am J Clin Nutr. (2012) 95:694–702. doi: 10.3945/ajcn.111.015271

22. Barakatun Nisak MY, Ruzita AT, Norimah AK, Gilbertson H, Nor Azmi K. Improvement of dietary quality with the aid of a low glycemic index diet in Asian patients with type 2 diabetes mellitus. J Am Coll Nutr. (2010) 29:161–70. doi: 10.1080/07315724.2010.10719830

23. Vinknes KJ, de Vogel S, Amanay K, Nurr E, Drevon CA, Gjesdal CG, et al. Dietary intake of protein is positively associated with percent body fat in middle-aged and older adults. J Nutr. (2011) 141:440–6. doi: 10.3945/jn.110.133710

24. Stabler SP, Allen RH. Vitamin B12 deficiency as a worldwide problem. Ann Rev Nutr. (2004) 24:299–326. doi: 10.1146/annurev.nutr.24.012003.132440

25. Jansen J, Karges W, Rink L. Zinc and diabetes–clinical links and molecular mechanisms. J Nutr Biochem. (2009) 20:399–417. doi: 10.1016/j.jnutbio.2009.01.009

26. Lubos E, Sinning CR, Schnabel RB, Wild PS, Zeller T, Rupprecht HJ, et al. Serum selenium and prognosis in cardiovascular disease: results from the Atherosclerose study. Atherosclerosis. (2010) 209:271–7. doi: 10.1016/j.atherosclerosis.2009.09.008

27. Nutrient Reference Values. Vitamin B12. Available online at: https://www.nrv.gov.au/nutrients/vitamin-b12 (accessed August 18, 2020).

28. Nutrient Reference Values. Zinc. Available online at: https://www.nrv.gov.au/nutrients/zinc (accessed August 18, 2020).

29. Nutrient Reference Values. Selenium. Available online at: https://www.nrv.gov.au/nutrients/selenium (accessed October 27, 2020).

30. Houston DK, Ding J, Lee JS, Garcia M, Kanaya AM, Tylavsky FA, et al. Dietary fat and cholesterol and risk of cardiovascular disease in older adults: the Health ABC Study. Nutr Metab Cardiovasc Dis. (2011) 21:430–6. doi: 10.1016/j.numecd.2011.10.007

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnut.2020.603801/full#supplementary-material
32. Veronese N, Solmi M, Caruso MG, Giannelli G, Osella AR, Evangelou E, et al. Dietary fiber and health outcomes: an umbrella review of systematic reviews and meta-analyses. *Am J Clin Nutr.* (2018) 107:436–44. doi: 10.1093/ajcn/nqz082

33. Berger S, Raman G, Vishwanathan R, Jacques PF, Johnson EJ. Dietary cholesterol and cardiovascular disease: a systematic review and meta-analysis. *Am J Clin Nutr.* (2015) 102:276–94. doi: 10.3945/ajcn.114.100305

34. De Souza RJ, Mente A, Maroleanu A, Cozma AI, Ha V, Kishibe T, et al. Intake of saturated and trans unsaturated fatty acids and risk of all cause mortality, cardiovascular disease, and type 2 diabetes: systematic review and meta-analysis of observational studies. *BMJ.* (2015) 351:h3978. doi: 10.1136/bmj.h3978

35. Australian Bureau of Statistics. *Australian Healthy Survey: Users’ Guide, 2011–2013.* Available online at: https://www.abs.gov.au/ausstats/abs@.nsf/mf/4363.0.55.001 (accessed August 18, 2020).

36. Barclay AW, Petocz P, McMillan-Price J, Flood VM, Prvan T, Mitchell P, et al. Glycemic index, glycemic load, and chronic disease risk—a meta-analysis of observational studies. *Am J Clin Nutr.* (2008) 87:627–37. doi: 10.1093/ajcn/87.3.627

37. National Health and Medical Research Council. *Australian Guidelines to Reduce Health Risks From Drinking Alcohol.* Available online at: https://www.nhmrc.gov.au/health-advice/alcohol (accessed August 18, 2020).

38. Biro G, Hulshof KE, Ovesen L, Amorim Cruz JA, Group E. Selection of methodology to assess food intake. *Eur J Clin Nutr.* (2002) 56 (Suppl. 2):S25–32. doi: 10.1038/sj.ejcn.1601428

39. Ridoutt BG, Handrie GA, Noakes M. Dietary strategies to reduce environmental impact: a critical review of the evidence base. *Adv Nutr.* (2017) 8:933–46. doi: 10.3945/an.117.016691

**Conflict of Interest:** FA and JB-M manage The University of Sydney’s glycemic index testing service and are directors of the Glycemic Index Foundation, a not-for-profit health promotion charity. JB-M is an author of books in The New Glucose Revolution series (De Capo, Cambridge, MA).

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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