Mammalian Osmolytes and S-Nitrosoglutathione Promote ΔF508 Cystic Fibrosis Transmembrane Conductance Regulator (CFTR) Protein Maturation and Function*

Marybeth Howard‡§, Horst Fischer¶, Jeremie Rouxçi, Bento C. Santos**, Steven R. Gullans‡‡, Paul H. Yancey§§, and William J. Welch¶¶†

From the ‡Department of Surgery and the ‡Department of Anesthesia, Surgical Research Laboratory, University of California, San Francisco, California 94110, the §Children’s Hospital Oakland Research Institute, Oakland, California 94609, the ¶Renal Division, Department of Medicine, Escola Paulista de Medicina, UNIFESP, 04023-900 Sao Paulo, Brazil, the §§Laboratory of Functional Genomics, Brigham and Women’s Hospital and Harvard Medical School, Boston, Massachusetts 02115, the ¶¶Department of Biology, Whitman College, Walla Walla, Washington 99362, and the ‡‡Department of Medicine and Physiology, University of California, San Francisco, California 94110

In cystic fibrosis, the absence of functional CFTR results in thick mucous secretions in the lung and intestines, as well as pancreatic deficiency. Although expressed at high levels in the kidney, mutations in CFTR result in little or no apparent kidney dysfunction. In an effort to understand this phenomenon, we analyzed ΔF508 CFTR maturation and function in kidney cells under conditions that are common to the kidney, namely osmotic stress. Kidney cells were grown in culture and adapted to 250 mM NaCl and 250 mM urea. High performance liquid chromatography analysis of lysates from kidney cells adapted to these conditions identified an increase in the cellular osmolytes glycerophosphorylcholine, myo-inositol, sorbitol, and taurine. In contrast to iso-osmotic conditions, hyperosmotic stress led to the proper folding and processing of ΔF508 CFTR. Furthermore, three of the cellular osmolytes, when added individually to cells, proved effective in promoting the proper folding and processing of the ΔF508 CFTR protein in both epithelial and fibroblast cells. Whole-cell patch clamping of osmolyte-treated cells showed that ΔF508 CFTR had trafficked to the plasma membrane and was activated by forskolin. Encouraged by these findings, we looked at other factors common to the kidney that may impact ΔF508 maturation and function. Interestingly, a small molecule, S-nitrosoglutathione, which is a substrate for gamma glutamyltranspeptidase, an abundant enzyme in the kidney, likewise promoted ΔF508 CFTR maturation and function. S-Nitrosoglutathione-corrected ΔF508 CFTR exhibited a shorter half-life as compared with wild type CFTR. These results demonstrate the feasibility of a small molecule approach as a therapeutic treatment in promoting ΔF508 CFTR maturation and function and suggest that an additional treatment may be required to stabilize ΔF508 CFTR protein once present at the plasma membrane. Finally, our observations may help to explain why ΔF508 homozygous patients do not present with kidney dysfunction.

The cystic fibrosis transmembrane conductance regulator (CFTR) protein is a cAMP-regulated chloride channel expressed in epithelial cells. Mutations within this protein cause cystic fibrosis, a disease of altered electrolyte transport and dehydration of secretions. The hallmark of CF is the accumulation of thick mucous secretions and frequent colonization of Pseudomonas bacterium within the lungs. Lung function deteriorates with age, and lung pathology is the leading cause of morbidity and mortality. Other organs affected by this disease are the pancreas, the intestines, and the liver. Interestingly, whereas CFTR is expressed in the kidney, CF patients show little or no kidney dysfunction. Indeed, Dr. di Sant‘Agnese, who first identified abnormally high sodium chloride in the sweat of CF patients, also noted “the disassociation between increased electrolytes in the sweat and the low sodium and chloride levels in urine during the periods of low salt diet or heat waves appears to be a unique characteristic of CF” (1). In other words, the ability of the kidney to regulate sodium chloride levels in the urine of CF patients appeared normal.

The most prevalent mutation associated with the disease is the deletion of a phenylalanine residue at position 508 (ΔF508 CFTR) (2). Loss of this single amino acid results in a failure of the newly synthesized protein to move out of the endoplasmic reticulum (ER) to its final locale at the plasma membrane (3). This failure is due to the synthesis of a protein that apparently cannot fold properly. Consequently, the protein is retained in the ER and degraded by the ubiquitin-proteosome pathway (4, 5). Validation for the idea that the ΔF508 CFTR mutation leads to improper folding is supported by the facts that: 1) misfolding of the protein can be corrected by lowering the temperature of cells expressing the mutant protein (6), 2) correct folding of the protein can occur upon addition of protein stabilizing agents, such as glycerol and TMAO (7, 8), and 3) once corrected, the ΔF508 protein can traffic to the plasma membrane and function as a cAMP-regulated chloride channel (6–9).

During osmotic stress, yeast and sharks increase the intracellular levels of glycerol and TMAO, respectively, two cellular osmolytes known to protect proteins from denaturation under harsh environmental conditions and as mentioned above, effec-

* This work was supported by National Institutes of Health Grants GM35351, DK51829, and 1F50HL06088, by the American Lung Association, and by the Cystic Fibrosis Research Institute. The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked “advertisement” in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

† To whom correspondence should be addressed: Dept. of Surgery, University of California at San Francisco, 1001 Potrero Ave., Bldg. 1, Rm. 210, San Francisco, CA 94110. Tel.: 415-206-6884; Fax: 415-206-6997; E-mail: mhb1@tsa.ucsf.edu.

This is an Open Access article under the CC BY license. This paper is available on line at http://www.jbc.org

1 The abbreviations used are: CFTR, cystic fibrosis transmembrane conductance regulator; CF, cystic fibrosis; ER, endoplasmic reticulum; TMAO, trimethylamine N-oxide; GPC, glycerophosphorylcholine; GSNO, S-nitrosoglutathione; IMCD, inner medullary collecting duct; ANOVA, analysis of variance; HPLC, high performance liquid chromatography; GGT, γ-glutamyltranspeptidase.
tive in correcting ΔF508 CFTR protein folding (10, 11). Similarly, mammalian systems have their own set of osmolytes, including sorbitol, myo-inositol, betaine, taurine, and glycerophosphorylcholine (GPC) (12, 13). These compounds typically accumulate to high concentrations in the kidney medulla as part of the normal physiological process of concentrating urine. In contrast to other organs of epithelial origin, the kidney is considered unaffected in ΔF508 CFTR homogzygotes. Therefore, we hypothesized that osmolytes that are synthesized and accumulated within kidney medulla might promote the proper folding and function of the ΔF508 CFTR protein and therefore help explain why ΔF508 CF patients do not present with kidney dysfunction.

The isosmotic renal cortex also expresses CFTR, but unlike the renal medulla, the absence of disease phenotype cannot be explained by elevated osmolality. Recently, Zaman et al. (14) reported that S-nitrosoglutathione (GSNO) promotes the maturation of endogenous wild type and ΔF508 CFTR maturation, although functional data were not presented. Interestingly, GSNO is a substrate for γ-glutamyltranspeptidase, (15) a transmembrane protein expressed at high levels in the lumen of the renal cortical proximal kidney (16–18). Here we show that GSNO promotes ΔF508 maturation and restores function while having no effect on wild type CFTR maturation. Thus, our results point to two possible mechanisms, intracellular osmolyte accumulation and hydrolysis of GSNO, carried out by the kidney, as potential explanations why CF patients exhibit little or no kidney dysfunction.

**EXPERIMENTAL PROCEDURES**

**Cell Culture and Adenovirus Infections—**All cells were maintained at 37 °C and 5% CO₂. Mouse inner medullary collecting duct cells (mIMCD3) isolated from the terminal third of the IMCD (19) were cultured in Dulbecco's modified Eagle's medium/F-12 supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin. 3T3 cells stably transfected with either the wild type or ΔF508 CFTR plasmid (20) were cultured in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum and 1% penicillin and streptomycin. For osmotic adaptation experiments, cells were gradually adapted to increasing concentrations of equimolar NaCl and urea over a 3-week period to a final concentration of 250 mM NaCl and 250 mM urea. Cells were maintained at this concentration for an additional 2–3 weeks before the initiation of experiments. For cellular osmolyte treatments, the compounds were added directly to the growth medium of the cells at the concentrations indicated in the text. Expression of either wild type or ΔF508 CFTR using recombinant adenovirus (Ref. 21; ΔF508 adenovirus, purchased from Dr. J. Wilson, University of Pennsylvania) was carried out as described previously (21). Twenty-four hours post-infection, 2 mM sodium butyrate was added to the cells to increase transcription of the CFTR cDNA. Eighteen hours later, cells were either harvested for biochemical analysis or used for whole-cell patch-clamping.

**Immunoblotting—**Cells were washed twice in calcium free phosphate-buffered saline and lysed in 50 mM Tris-HCl, pH 7.4, 1% Nonidet P-40, 0.25% sodium deoxycholate, 150 mM NaCl, 1 mM EDTA supplemented with protease inhibitors (Calbiochem). The lysates were clarified at 14,000 × g at 4 °C. An aliquot of the supernatant was sampled for protein determination (BCA, Pierce), whereas the remaining portion was adjusted to 1X Laemmli sample buffer (40 mM Tris-HCl, pH 6.8, 1% SDS, 50 mM dithiothreitol, 7.5% glycerol) and heated at 37 °C for 20 min. Equal amounts of total protein were loaded on 7.5% polyacrylamide gels. Following SDS-PAGE, the proteins were transferred to nitrocellulose and probed with a monoclonal antibody specific for the CFTR protein (1:1,000, Mab721, Upstate Biotechnology) and an anti-rat IgG 1:5000. CFTR protein was visualized by chemiluminescence. Amido black staining was performed on a 7.5% gel. Proteins were visualized by fluorography. All cells were maintained at 37 °C as described previously (22). On the stage of an inverted microscope cells were bathed (in mm): 136 N-methyl-D-glucamine, 127 HCl, 1.7 CaCl₂, 1 MgCl₂, 10 Heps, 10 glucose, 500 mannitol, pH 7.3. Patch pipettes were filled with (in mm): 130 N-methyl-D-glucamine, 130 HCl, 10 EGTA, 1 MgCl₂, 2 Heps, 1 glucose, 500 mannitol, 5 Mg-ATP, 0.1 Li-GTP, pH 7.3. Single isolated cells were randomly selected for patch clamping. Only seals >10 GΩ were used. After establishing the whole cell configuration the membrane potential (V_m) was continuously clamped to −40 mV. The access resistance (R_a) and the cell membrane capacitance (C_m) were measured using the current transients caused by a 10-mV voltage pulse. R_a = 15 ± 2 mΩ (n = 104) and was not affected by the different treatments. Current-voltage (I-V) step protocols before and after addition of 20 μM forskolin were applied, from a resting potential of −40 mV, from −100 mV to +40 mV, and the resulting currents were recorded. Larger voltage jumps reduced the success rate of experiments due to seal breakage and were generally not performed. Whole cell conductance (G_w) was calculated as the slope between −40 and −20 mV. For the calculation of the specific membrane conductance (G_m) in pico Siemens/picofarad) G_w was corrected for R_a and normalized to C_m. Quantitative effects of different treatments were compared with factorial ANOVAs followed by Bonferroni-corrected t tests. p < 0.05 was considered significant. Calculations were done with StatView, version 4.5 (Abacus Concepts, Berkeley, CA).

**Determination of Cellular Osmolyte Concentration—**Control cells (mIMCD3) and hyperosmolar-adapted cells (HT500) were grown on 10-cm plates. Intracellular osmolyte concentrations were determined via HPLC analysis using a modified protocol as described by Rauchman et al. (19). Briefly, cells were lysed in 7% perchloric acid and the lysates centrifuged at 14,000 × g for 20 min. The acid soluble pellet was resuspended in 0.5 mM NaOH solution and used for subsequent protein analysis using the BCA assay. The acid soluble material was adjusted to neutral pH and centrifuged at 10,000 × g for 30 min. The supernatant was serially filtered through a C₈ Sep-Pak column and a 0.45-μM HPLC filter, lyophilized, and resuspended in HPLC mobile-phase solution and loaded onto an HPLC column. Osmolytes were measured by refractive index.

**Chemicals and Reagents—**Forskolin (Calbiochem) was made as a 20 mM stock in Me₂SO and used at 20 μM. GSNO (Sigma), prepared as a fresh stock each time, was dissolved in water at 10 μM and used at 10 μM. Osmolytes (Sigma) were dissolved in medium at 1 M and used at 300 μM.

**RESULTS**

**Hyperosmotic-adapted Kidney Cells Accumulate Osmolytes—**Mouse inner medullary collecting duct (mIMCD3) cells were cultured under iso-osmotic or hyperosmotic conditions. Cells were stepwise adapted to hyperosmotic conditions by the addition of a 50 mM solution of sodium chloride and urea every 4 days. The final concentration of sodium chloride and urea added was 250 mM each, and therefore, the cells are referred to as HT500 (hypertonic 500 mM, 750 mosmol). The cells retained a number of phenotypic characteristics typical of the kidney, such as tolerance to hyperosmotic conditions for long periods of time with minimal adverse effects on DNA, RNA, and protein synthesis (19, 23). Adaptation to the hyperosmotic conditions resulted in a significant increase in the intracellular levels of several osmolytes in the kidney cells, in particular, sorbitol, glycerophosphorylcholine, taurine, and myo-inositol (Fig. 1A, filled bars). These changes in the levels of the osmolytes are consistent with levels seen in the kidney medulla, a segment of
the kidney routinely exposed to hyperosmolarity (13). Interestingly, no increase in betaine was observed in mIMCD cells adapted to 750 mosmol. On the other hand, Rauchman et al. (19) observed an increase in betaine in mIMCD cells adapted to 910 mosmol, while Peterson et al. (24) reported that betaine does not increase in the inner medulla of rats subjected to a high protein diet e.g. a diet that increases urea levels in the kidney.

**Hyperosmotic-adapted Kidney Cells Exhibit Proper Maturation and Functional ΔF508 CFTR**—Having established conditions that increase osmolyte levels in the mIMCD3 cells, we tested whether the accumulation of these osmolytes would have any effect on the maturation of the ΔF508 CFTR protein. The rationale based on previous studies showing that osmolytes such as glycerol and TMAO, when added to cells, correct the folding and trafficking of ΔF508 CFTR (7, 8). Control mIMCD3 cells maintained in iso-osmotic growth medium, and HT500 cells maintained in hyperosmotic growth medium were infected with a recombinant adenovirus expressing either wild type or ΔF508 CFTR. Thirty-six hours post-infection the cells were treated with 2 mM sodium butyrate and then harvested 18 h later. Equal amounts of total protein were separated by SDS-PAGE, and CFTR protein expression was determined by Western blot analysis. Lanes 1 and 4, mock-infected cells; lanes 2 and 5, wild type CFTR protein expression; lanes 3 and 6, ΔF508 CFTR protein expression. B demarcates the immature core-glycosylated CFTR protein; C demarcates the fully glycosylated mature form of CFTR protein. C, representative recordings of currents elicited by voltage steps (from −100 mV to +40 mV) in mock- and ΔF508 CFTR adenovirus-infected HT500 cells after stimulation with 20 μM forskolin. D, current-voltage relationship of currents recorded in C. Open circles, mock-infected cells; filled circles, ΔF508 CFTR-infected HT500 cells. E, specific conductance (Gm) for mock- and ΔF508 CFTR adenovirus-infected HT500 cells. ΔF508 CFTR protein expressing cells showed a significantly higher Gm (p < 0.001, t test). Average capacitance of these cells was 53 ± 14 picofarads (n = 10) and was not different between mock- and ΔF508 CFTR adenovirus-infected cells.
Fig. 2. Sorbitol, myo-inositol, and taurine promote the maturation of ΔF508 CFTR and result in cAMP-regulated chloride transport in mouse fibroblast cells. A, osmolytes were added to a final concentration of 300 mM to the extracellular medium of 3T3 cells stably expressing ΔF508 CFTR over a 24-h period as described under "Experimental Procedures." The cells were maintained in the medium containing
glycosylated C band (Fig. 1B, lanes 2 and 5). Expression of the ΔF508 CFTR protein in mIMCD3 cells resulted in the appearance of only the core-glycosylated protein (band B), as is normally seen in cells expressing the ΔF508 mutant (Fig. 1B, lane 3). In the ΔF508 CFTR-infected HT500 cells, however, an increase in the amount of the core-glycosylated B band, as well as the processing of a portion of the protein to the mature form (C band) was observed (Fig. 1B, lane 6). Interestingly, ΔF508 CFTR expressed in the HT500 cells exhibited multiple electrophoretic species as evidenced by a smear of bands with slower mobility as compared with the immature B band. At the present time, the exact basis for this heterogeneous collection of CFTR species is not clear. Others, however, have similarly shown the mature CFTR protein to exist as a “broad smear” (5, 25, 26). Thus, it appears that hyperosmotic conditions promote the proper processing of at least a portion of the mutant ΔF508 CFTR protein, but have no apparent effect on the maturation of wild type CFTR.

To test whether the hyperosmotic stress resulted in the synthesis and folding of a functional ΔF508 CFTR protein, the HT500 cells were whole cell patch-clamped to quantify forskolin-stimulated chloride transport. Whole-cell currents were measured under conditions selective for chloride currents in the presence of 20 μM forskolin (see “Experimental Procedures”). Fig. 1C shows typical examples of current step responses recorded from mock-infected and ΔF508 CFTR-infected HT500 cells. HT500 cells expressing ΔF508 CFTR showed large forskolin-activated chloride currents. Measured currents were time- and voltage-independent (Fig. 1C) and showed linear current-voltage relationships (Fig. 1D). Fig. 1E summarizes the calculated specific membrane conductances (G). Thus, whole-cell patch-clamping demonstrated that ΔF508 CFTR expressed in HT500 cells showed significant function, exhibiting typical CFTR-specific characteristics, i.e. chloride selectivity, activation by forskolin, and linear current-voltage relationships.

Individual Osmolytes in Fibroblast Cells Promote the Maturation of Functional of ΔF508 CFTR—Having shown that hyperosmotic stress both increases the levels of different osmolytes and promotes the maturation of ΔF508 CFTR, we tested whether the individual osmolytes themselves were effective in correcting ΔF508 misfolding, and, thereby, would restore chloride channel function. If so, this would indicate that the osmolytes themselves, rather than some other effect due to salt and urea adaptation of renal cells, can help promote ΔF508 CFTR protein maturation. For these experiments, we utilized 3T3 fibroblasts stably expressing ΔF508 CFTR under the control of the cytomegalovirus promoter (20). Using these cells, we previously showed that two other osmolytes, glyceraldehyde 3-phosphate dehydrogenase, and TMAO, were effective in correcting the misfolding of the ΔF508 CFTR mutant (7). Furthermore, Denng et al. (6) used these cells to show that incubation at low temperatures (e.g. 27 °C) also corrected the misfolding of ΔF508 CFTR protein. Cells were incubated in growth medium containing the individual osmolytes for 3 days at a concentration of 300 mM, after which the cells were lysed and CFTR expression examined by Western blot (Fig. 2A). Cells not incubated with osmolytes nor stimulated with the general transcriptional activator, sodium butyrate, and cells stimulated with sodium butyrate alone to increase CFTR transcription showed little or no ΔF508 protein in the cells maintained at 37 °C (Fig. 2A, lanes 1 and 2). Treatment with the osmolytes myo-inositol, sorbitol, or taurine resulted in a portion of the ΔF508 CFTR protein being correctly folded and processed to the mature form (Fig. 2A, lanes 4–6). Sorbitol treatment elicited the highest levels of the mature form of the protein (Fig. 2A, lane 5), while betaine had little or no effect on ΔF508 CFTR maturation (Fig. 2A, lane 3). Curiously, betaine itself proved to be somewhat toxic to the fibroblast cells with the cells appearing rounded after 3 days of incubation with 300 mM betaine (data not shown). This result is consistent with a previous study showing betaine is toxic to renal cells when added alone to the extracellular medium but is nontoxic when added in conjunction with urea (27).

Incubation of the cells at 27 °C along with the osmolytes revealed a synergistic effect on ΔF508 maturation. When incubated at 27 °C, low levels of the immature form of ΔF508 CFTR were now evident and increased upon addition of the general transcriptional activator, sodium butyrate (Fig. 2A, lanes 1 and 2). Note as well, and consistent with previous reports showing ΔF508 CFTR folding to be temperature-sensitive, the appearance of the mature form of the protein (band C) in those cells maintained at the lower temperature. When the cells growing at 27 °C now were treated with the various osmolytes, significant amounts of mature ΔF508 CFTR (band C) were observed (Fig. 2A, lanes 4–6, 27 °C). Similar to the situation at 37 °C, addition of betaine (300 mM) to the cells proved to be somewhat toxic after 3 days of incubation (Fig. 2A, lane 3). Thus, ΔF508 CFTR maturation in 3T3 cells can be rescued by either lowering the temperature of the cells or by the addition of various cellular osmolytes. Moreover, osmolyte treatment along with a lower growth temperature resulted in a synergistic enhanced effect on the maturation of the mutant protein.

To determine whether the osmolyte-corrected ΔF508 CFTR protein was in fact functional, the 3T3 cells were whole-cell patch-clamped. The cells were incubated in myo-inositol, taurine, TMAO, or sorbitol (300 mM each) for 3 days, and whole cell currents were measured under conditions selective for chloride currents (see “Experimental Procedures”). Fig. 2B shows examples of current step responses recorded from the unstimulated cells (top panels) or from the forskolin-stimulated cells (bottom panels) incubated in the absence or presence of the osmolytes. Wild type CFTR-expressing 3T3 cells showed typical CFTR-mediated chloride currents after forskolin stimulation (Fig. 2B, right panels). Untreated ΔF508 CFTR expressing cells, either without or with forskolin stimulation, showed no chloride currents (Fig. 2B, left panels). In contrast, 3 days of treatment with the different osmolytes resulted in the appearance of large forskolin-activated chloride currents. All measured currents were time- and voltage-independent (Fig. 2B) and showed linear current-voltage relations (Fig. 2C).

Fig. 2D summarizes the specific conductance (Gm) from 47 patch-clamped cells. Treatment with myo-inositol, but not any
Small Molecules and ΔF508 CFTR Maturation/Function

of the other osmoieties, resulted in a significant increase in $G_m$ in the absence of forskolin stimulation. As a control for the effects of myo-inositol on basal currents we used its isomer scyllo-inositol. Cells incubated with scyllo-inositol showed no increase of basal $G_m$, but upon forskolin stimulation did exhibit an increase in $G_m$ (data not shown). In cells treated with myo-inositol, TMAO, or sorbitol, stimulation with forskolin (20 μM) increased the $G_m$ to levels similar to that found for cells expressing wild type CFTR. Treatment with taurine resulted in the largest recovery of a forskolin-stimulated $G_m$, which was significantly greater than $G_m$ measured in the wild type CFTR expressing cells (Fig. 2D).

Osmolytes Promote ΔF508 Maturation in Epithelial Cells—Individual osmoieties proved to be effective in promoting ΔF508 CFTR maturation in fibroblasts after 3 days of treatment. However, because CFTR is expressed in epithelial cells, we tested whether kidney cells that are normally exposed to osmoieties would be even more effective in correcting ΔF508 misfolding. mIMCD3 cells were treated with the individual osmoieties for either 24 or 72 h. In the case of the 24-h treatments, the cells were first incubated with adenosine expressing ΔF508 and then incubated in myo-inositol, sorbitol, or taurine at a final concentration of 300 mM. For the 72-h treatments, the mIMCD3 cells were first incubated in myo-inositol, sorbitol, or taurine at a final concentration of 300 mM for 24 h and then infected with the ΔF508 CFTR adenovirus. The infected cells were further incubated with the individual osmoieties for another 48 h prior to cell lysis (B). Cells were stimulated with 2 mM sodium butyrate and harvested 18 h later. Equal amounts of total protein were separated by SDS-PAGE, and CFTR expression was determined by Western blot. Lane 1, no osmoieties; lane 2, myo-inositol; lane 3, sorbitol; lane 4, taurine.

Fig. 3. Osmolyte treatment of epithelial cells promotes processing of ΔF508 CFTR protein. mIMCD3 epithelial cells expressing ΔF508 CFTR were incubated with osmoieties for 24 or 72 h prior to cells lysis. In the case of the 24-h treatments, the mIMCD3 cells were first infected with adenosine expressing ΔF508 and then incubated in either myo-inositol, sorbitol, or taurine at a final concentration of 300 mM before cell lysis (A). For the 72-h treatments, the mIMCD3 cells were first incubated in myo-inositol, sorbitol, or taurine at a final concentration of 300 mM for 24 h and then infected with the ΔF508 CFTR adenovirus. The infected cells were further incubated with the individual osmoieties for another 48 h prior to cell lysis (B). Cells were stimulated with 2 mM sodium butyrate and harvested 18 h later. Equal amounts of total protein were separated by SDS-PAGE, and CFTR expression was determined by Western blot. Lane 1, no osmoieties; lane 2, myo-inositol; lane 3, sorbitol; lane 4, taurine.

notype in patients homozygous for ΔF508 CFTR, we searched for other aspects unique to the kidney that might affect CFTR maturation. In searching the literature, we found a study by Zaman et al. (14) reporting that GSNO treatment promoted the maturation of both wild type and ΔF508 CFTR. Although data demonstrating that the corrected ΔF508 could now function as a chloride channel were not presented, we were intrigued since GSNO is a substrate for γ-glutamyltranspeptidase (GGT), an enzyme that is expressed at high levels in the kidney (16–18). GGT hydrolyzes glutathione and S-substituted GSH derivatives within the kidney filtrate (28) for subsequent uptake into the cells by amino acid transporters. GSNO treatment has been shown to have modifying effects (e.g., nitrosylation) on other cellular proteins thereby impacting their function (29–32). Thus, we tested whether GSNO would have any effect on ΔF508 maturation and if so whether the corrected ΔF508 CFTR would exhibit cAMP-regulated chloride channel function.

Treatment of mIMCD3 cells expressing ΔF508 CFTR with 10 μM GSNO for either 1 or 2 h resulted in the proper maturation of the mutant protein (Fig. 4A, lanes 2–4). Treatment of the cells expressing wild type CFTR with GSNO had no obvious effect on the amounts of mature CFTR produced (Fig. 4B, lanes 2–4). Next, we examined whether the GSNO-corrected ΔF508 protein exhibited functional chloride channel activity. Whole-cell currents were recorded from forskolin-stimulated (20 μM) ΔF508 CFTR-expressing mIMCD3 cells pretreated with 10 μM GSNO and compared with untreated or mock-infected cells (Fig. 4C). The GSNO-treated cells, but not the untreated ΔF508 CFTR-infected mIMCD3 cells, showed linear time- and voltage-independent current characteristics (Fig. 4D). In comparison, mock-infected or untreated ΔF508 CFTR-infected cells showed very small $G_m$ values that were not statistically different from one another (Fig. 4E). Thus, short term GSNO treatment results in a portion of ΔF508 CFTR to mature into the C band, resulting in CFTR forskolin-stimulated chloride transport.

GSNO-corrected ΔF508 CFTR Is Unstable Compared with Wild Type CFTR—The relatively low concentration required and the fast kinetics of GSNO-induced ΔF508 CFTR protein maturation highlighted this naturally occurring compound as a potential therapeutic agent. To pursue this possibility further, we investigated the stability of GSNO-corrected ΔF508 CFTR protein. The impetus here being based on Sharma et al. (26), who reported that ΔF508 CFTR, corrected either by low temperature or by glycerol, was unstable when the cells expressing the corrected ΔF508 CFTR were returned to 37 °C. mIMCD3 cells expressing either wild type or ΔF508 CFTR were pulse-labeled with [35S]methionine for 4 h in either the absence or presence of 100 μM GSNO. The medium containing the radiolabel was removed; fresh medium was added to the cells and further incubated for 4 h (i.e., chase) either in the absence or in the presence of GSNO prior to quantitative immunoprecipitation (Fig. 5). Immediately following the pulse labeling, significant amounts of the mature form (C band) of the ΔF508 CFTR protein were observed. The ΔF508 CFTR protein corrected by GSNO treatment proved to be unstable and little of the radiolabeled protein was detected after the 4-h chase period, either in the absence or in the presence of GSNO. Thus, while our Western blot analysis showed the corrected ΔF508 CFTR, presumably due to continuous maturation of nascent ΔF508 CFTR protein in the presence of GSNO, our pulse-chase studies showed that once ΔF508 CFTR protein had folded and trafficked to the plasma membrane, the protein was unstable.

DISCUSSION

Cystic fibrosis is a disease affecting epithelial cell function in various organs such as lung, intestines, pancreas, and liver as
CFTR-infected mIMCD cells after stimulation with 20 CFTR expressing cells, untreated; lane 1, wild type CFTR: lane 1.

Resulted in forskolin-stimulated time- and voltage-independent currents. Small currents were recorded in untreated cells. Treatments with GSNO filled circles treated cells; lane 4, 2-h treatment with 10 M GSNO (mIMCD cells. GSNO-treated significantly higher currents elicited by voltage steps (from -100 mV to +40 mV) in ΔF508 CFTR-infected mIMCD cells after stimulation with 20 μM forskolin. Very small currents were recorded in untreated cells. Treatments with GSNO resulted in forskolin-stimulated time- and voltage-independent currents. D, current-voltage relations of currents recorded in C. Open circles, untreated cells; filled circles, GSNO-treated cells. E, specific conductance (Gₛ) for GSNO-treated and untreated ΔF508 CFTR- or mock-infected mIMCD cells. GSNO-treated ΔF508 CFTR-infected cells showed a significantly higher Gₛ (p = 0.033, factorial ANOVA).

FIG. 4. GSNO promotes the maturation and function of ΔF508 CFTR but has no effect on wild type CFTR maturation. mIMCD cells were infected with either ΔF508 or wild type CFTR recombinant adenoivirus as described previously. Cells were left untreated or were treated with 10 μM GSNO for 1 or 2 h prior to cell lysis. Equal amounts of total protein were separated by SDS-PAGE, and CFTR expression was determined by Western blot. A, ΔF508 CFTR: lane 1, mock-infected parental cells; lane 2, ΔF508 CFTR expressing cells, untreated; lane 3, 1-h treatment of 10 μM GSNO; lane 4, 2-h treatment with 10 μM GSNO. B, wild type CFTR: lane 1, mock-infected parental cells; lane 2, wt CFTR expressing cells, untreated; lane 3, 1-h treatment of 10 μM GSNO; lane 4, 2-h treatment with 10 μM GSNO. C, representative recordings of currents elicited by voltage steps (from -100 mV to +40 mV) in ΔF508 CFTR-infected mIMCD cells after stimulation with 20 μM forskolin. Very small currents were recorded in untreated cells. Treatments with GSNO resulted in forskolin-stimulated time- and voltage-independent currents. D, current-voltage relations of currents recorded in C. Open circles, untreated cells; filled circles, GSNO-treated cells. E, specific conductance (Gₛ) for GSNO-treated and untreated ΔF508 CFTR- or mock-infected mIMCD cells. GSNO-treated ΔF508 CFTR-infected cells showed a significantly higher Gₛ (p = 0.033, factorial ANOVA).

FIG. 5. GSNO-corrected ΔF508 CFTR protein is less stable than wild type CFTR protein. mIMCD3 cells were infected with either wild type or ΔF508 CFTR recombinant adenovirus. Twenty-four hours post-infection, cells were treated with 2 mM sodium butyrate. Fourteen hours post-induction, 50 μCi/ml of [35S]methionine was added to the medium either in the absence or presence of 100 μM GSNO. After 4 h, cells were washed and either harvested immediately (pulse-labeled) or the medium was removed and the cells washed and further incubated in medium supplemented with 1 mM cold methionine for 4 h either in the absence or presence of GSNO before being harvested (pulse-labeled and chase). The amount of radiolabeled CFTR present in the lysates was determined by quantitative immunoprecipitation as described under “Experimental Procedures.” Shown is a fluorogram of the gel. Treatment of the cells without (-) or with (+) GSNO during the pulse and chase is indicated at the top.

well as the reproductive system. However, even though CFTR is expressed in the kidney (33, 34), cystic fibrosis patients present with little or no kidney dysfunction. The absence of kidney disease suggests that either other chloride channels can complement the function of CFTR in the kidney (e.g. there is a redundancy in the kidney with regards to chloride channels) or that some other mechanisms are operative in the kidney that prevail over the particular CFTR mutation. In this regard, studies by Morales et al. (34) reported a unique isoform of CFTR expressed in the renal medulla comprising only the first transmembrane domain, the nucleotide binding domain, and the R domain. The authors suggested that the generation of this unique functional isoform of CFTR in the kidney might be one mechanism by which the kidney is protected from functional defects of CF. In their study, the authors demonstrated that this unique isoform functioned as a cAMP-regulated chloride channel and was effective in correcting the regulation of the outwardly rectifying chloride channel. Thus, for CF patients with disease-associated mutations in the carboxy half of CFTR, expression of this alternate isoform expressing only the amino-terminal half of CFTR would maintain chloride transport necessary for normal kidney function. Alternatively, the kidney, by the very nature of its function (i.e. to concentrate urine), is routinely exposed to hyperosmotic conditions, specifically in the medulla. While similar hyperosmotic conditions likely would be deleterious to most cell types, certain cells within the kidney are able to adapt to and continue their normal physiologic functions even in the presence of high osmotic stress. Under these conditions, kidney homeostasis is preserved, at least in part, by the accumulation of cellular osmolytes that help prevent general protein denaturation (discussed further below).

In our studies here, we adapted mIMCD3 kidney cells to hyperosmotic conditions by the addition of 250 mM sodium chloride and 250 mM urea. By HPLC analysis, we found that the hyperosmotically adapted mIMCD3 cells accumulated a number of intracellular osmolytes, including GPC, myo-inosi-
tol, sorbitol, and taurine. The accumulation of the osmolytes was accomplished by an ability of the cells to produce a functional form of the ΔF508 CFTR mutant. Furthermore, when these different osmolytes were added individually to the medium of ΔF508 CFTR expressing cells, a portion of ΔF508 CFTR was processed to its mature form and resulted in the cells exhibiting cAMP-regulated CFTR chloride channel function. Interesting, myo-inositol activated CFTR channels in the absence of agonist. While not within the scope of this study, signaling roles of inositol in both calcium release from the ER as well as activation of PKC may be involved. In this regard, others have shown that PKC has a modulating effect on CFTR channel activity (35, 36).

The mechanisms by which cellular osmolytes impact protein folding and/or stability have been an active area of research (for reviews, see Refs. 37 and 38). Early work by Timasheff and colleagues (39) concluded that osmolytes help stabilize proteins in their native conformation due to their preferential exclusion from the immediate vicinity of the protein. Preferential exclusion of the osmolytes leads to an increase in the local concentration of water surrounding the protein. The resultant increase in surface tension would then favor a decrease in surface area of the protein, thereby promoting the folded state (i.e. one with the lowest energy conformation). Subsequent studies by Bolen and others (40–42) concluded that in the presence of osmolytes, unfavorable interactions of the peptide backbone (but not the R groups) with the osmolytes enhances proper protein folding, thereby helping to minimize these unfavorable interactions. In other words, the same general principles that govern protein folding events under iso-osmotic conditions, in particular the hydrophobic effect, are likely amplified in solutions containing high concentrations of the osmolytes. Thus, in the case of ΔF508 CFTR where a potentially critical hydrophobic amino acid phenylalanine is absent, we suspect that an osmolyte enhancement of the hydrophobic effect may help drive (and/or stabilize) an early folding intermediate requiring hydrophobic interactions in and around position 508. As a consequence, at least a portion of the folding intermediates continues to fold to the native state and thereby escape the quality control pathway operative in the endoplasmic reticulum. Subsequent maturation in the Golgi followed by localization at the plasma membrane results in a ΔF508 CFTR protein that appears functional as assayed by forskolin-mediated chloride transport.

Our results demonstrating that osmotic stress-induced osmolyte accumulation, or the addition of individual osmolytes to the medium of cells results in the rescue of ΔF508 CFTR maturation, confirms and extends earlier work. Specifically, previous studies have shown that high concentrations of either glycerol or TMAO, when added to fibroblast cells in culture, restored wild type-like ΔF508 CFTR localization and chloride channel function (7, 8). Moreover, both glycerol and TMAO were found to be effective in promoting the proper folding of a variety of other temperature-sensitive protein folding mutants, including the tumor suppressor p53, the viral oncoprotein protein pp60src, the ubiquitin activating enzyme E1 (43), the water channel protein aquaporin-2 (44), α1-antitrypsin (45), and 3-ke-toacid decarboxylase (46). In addition, cellular osmolytes appeared effective in slowing and even preventing the formation of the pathogenic form of the so-called scrapie prion protein (47). Based on these observations we have collectively referred to the cellular osmolytes as “chemical chaperones” to reflect their wide range of action in promoting the proper folding of proteins, which due to mutation, oftentimes fail to reach the native state.

Within the kidney, the inner medullary collecting ducts are routinely exposed to hypotonic stress and thus the accumulation of intracellular osmolytes could help to explain the lack of adverse phenotype in individuals homozygous for ΔF508 CFTR. The renal cortex, another segment of the kidney that expresses CFTR, is typically exposed to iso-osmotic conditions and therefore osmolyte accumulation would not likely explain any lack of adverse phenotype. During the course of our studies however, Zaman et al. (14) reported that another small molecule, GSNO, acted to promote the proper maturation of both the wild type and ΔF508 forms of CFTR. Consistent with their report we found that the addition of GSNO to the medium of ΔF508-expressing IMCD3 cells resulted in the proper maturation of ΔF508 CFTR. In addition, we showed that the GSNO-treated cells expressing the ΔF508 CFTR mutant now were capable of carrying out forskolin-mediated chloride transport.

Recently, Anderson et al. (48) reported similar effects of GSNO on ΔF508 CFTR expressing cells using a fluorescent-based assay. Our data showed that the maturation of ΔF508 CFTR occurred using rather low concentrations (e.g. 10 μM) of GSNO and was very rapid (within 1 h the mature form of the ΔF508 CFTR protein was readily apparent by Western blot analysis).

The mechanism by which ΔF508 CFTR maturation and subsequent function are rescued by GSNO treatment is currently unknown. GSNO (a thiol derivative of glutathione) is a reservoir of both nitric oxide (NO) and glutathione. NO at low concentrations can act as a signaling molecule, activating guanylate cyclase and the subsequent formation of the second messenger cGMP (for review, see Ref. 49). cGMP in turn is known to activate various ion channels and kinases (49). Higher concentrations of NO can lead to the modification of cysteine thiols (nitrosylation) leading to either activation or inactivation of the protein target and thus affect various cellular functions, including transcription, ion channel function, chaperone function, and proteolysis (reviewed in Ref. 50). Contradictory evidence of the effect of NO on CFTR channel function and biogenesis has been reported. For example, NO has been shown to activate CFTR in human T lymphocytes (51), while Ruckes-Nilges et al. (52) reported that NO had no effect on CFTR or any other chloride channel activation in primary nasal epithelial cells. GSNO, on the other hand, was found to activate chloride channels in lung epithelial cells (53). Finally, other studies have shown that NO inhibits CFTR protein expression (54, 55). In the case of glutathione, this tripeptide is an important antioxidant necessary for the maintenance of the redox state of the cell. While others have suggested that the redox state of the cell is important for the activation of CFTR channels (56), in our study, the low concentration of GSNO required to promote ΔF508 CFTR maturation likely would not impact glutathione levels. Thus, we suspect that the operative mechanism by which GSNO promotes ΔF508 CFTR maturation is by modification of either cysteine thiols of the nascent ΔF508 CFTR protein or cysteine thiols of a cellular protein(s) involved in the monitoring of quality control mechanisms of protein folding.

While GSNO levels in the blood of healthy individuals has been questioned, it may be possible that plasma GSNO is formed in vivo (57–59) due to inflammation as is common in CF patients. In addition, intracellularly synthesized GSNO could be exported out of the cells by the MRP2 transporter, which is expressed in the proximal tubule lumenal membrane of the kidney (60). Whatever the case, extracellular glutathione and S–substituted GSH derivatives are a substrate for γ-glutamyltranspeptidase, an enzyme expressed at high levels in the lumenal membrane of the kidney. Hydrolysis of GSNO would produce glutamate and S-substituted cysteinylglycine, in the kidney filtrate (15, 28). Subsequent uptake of the S-substituted cysteinylglycine by amino acid transporters and further protein
modifications by S-nitrosylation could be one mechanism by which ΔF508 CFTR protein maturation is rescued. Studies are under way to test this and other possibilities.

Our pulse-chase studies demonstrated that once ΔF508 CFTR was corrected by GSNO, the mature form of the protein was unstable. Whether ΔF508 CFTR is corrected by temperature, glycerol (26), or GSNO, the protein appears to be unstable relative to its wild type counterpart. Thus, the ΔF508 CFTR protein is not a typical temperature-sensitive folding mutant, since most temperature-sensitive protein folding mutants, once properly folded, usually exhibit temperature-dependent stability profiles similar to the wild type protein (61). This then indicates that ΔF508, whether acquiring a functionally competent conformation via low temperature, osmolyte, or GSNO treatments, presumably does not achieve a conformation identical to that of the wild type protein. Our results together with the results of Sharma and colleagues highlight the possibility that a second drug may be required to stabilize the ΔF508 CFTR protein once it has exited the ER and Golgi and trafficked to the plasma membrane.

In summary, our studies point to two mechanisms that may help to explain the absence of kidney dysfunction in CF patients; osmolyte accumulation and GSNO catabolism. In addition, our work further demonstrates the feasibility of a small molecule approach as a clinical therapy to correcting ΔF508 misfolding, while at the same time realizing a second issue that needs to be addressed in the search for therapeutic candidates, i.e. the stability of the fully “mature” ΔF508 CFTR protein. While using high concentrations of osmolytes may not be clinically desirable, GSNO, a substrate for GGT, an enzyme also expressed in the lung (62), is one potential therapeutic candidate. In this regard, a study of nine CF patients receiving one dose of aerosolized GSNO showed that the compound was a candidate. In this regard, a study of nine CF patients receiving one dose of aerosolized GSNO showed that the compound was a candidate. In this regard, a study of nine CF patients receiving one dose of aerosolized GSNO showed that the compound was a candidate. In this regard, a study of nine CF patients receiving one dose of aerosolized GSNO showed that the compound was a candidate. In this regard, a study of nine CF patients receiving one dose of aerosolized GSNO showed that the compound was a candidate. In this regard, a study of nine CF patients receiving one dose of aerosolized GSNO showed that the compound was a candidate.