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Ketones Prevent Oxidative Impairment of Hippocampal Synaptic Integrity through $K_{\text{ATP}}$ Channels

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Abstract

Dietary and metabolic therapies are increasingly being considered for a variety of neurological disorders, based in part on growing evidence for the neuroprotective properties of the ketogenic diet (KD) and ketones. Earlier, we demonstrated that ketones afford hippocampal synaptic protection against exogenous oxidative stress, but the mechanisms underlying these actions remain unclear. Recent studies have shown that ketones may modulate neuronal firing through interactions with ATP-sensitive potassium ($K_{\text{ATP}}$) channels. Here, we used a combination of electrophysiological, pharmacological, and biochemical assays to determine whether hippocampal synaptic protection by ketones is a consequence of $K_{\text{ATP}}$ channel activation. Ketones dose-dependently reversed oxidative impairment of hippocampal synaptic integrity, neuronal viability, and bioenergetic capacity, and this action was mirrored by the $K_{\text{ATP}}$ channel activator diazoxide. Inhibition of $K_{\text{ATP}}$ channels reversed ketone-evoked hippocampal protection, and genetic ablation of the inwardly rectifying $K^+$ channel subunit Kir6.2, a critical component of $K_{\text{ATP}}$ channels, partially negated the synaptic protection afforded by ketones. This partial protection was completely reversed by co-application of the $K_{\text{ATP}}$ blocker, 5-hydroxydecanoate (5HD). We conclude that, under conditions of oxidative injury, ketones induce synaptic protection in part through activation of $K_{\text{ATP}}$ channels.

Introduction

The ketogenic diet (KD), a proven treatment for medically intractable epilepsy [1], results in prominent production of ketones (notably, D-β-hydroxybutyrate [BHB] and acetacetate [ACA]). Increasingly, ketones have been shown to exert neuroprotective actions in models of neurodegenerative disorders (NDs), likely by restoration of impaired mitochondrial metabolism and antioxidant capacity [2,3]. A common pathogenic feature of NDs is oxidative stress...
which correlates closely with progressive tissue injury, and when the hippocampus is affected, results in cognitive and memory deficits. Conversely, a synthetic BHB ester-linked polymer, KTX-0101, and dietary ketosis are shown to mitigate memory deficits in patients of Alzheimer’s disease (AD) [4,5]. In line with this, ketones attenuate impairment of hippocampal cognitive function in a model of AD [6]. Further, in vitro studies have demonstrated a crucial role for ketones in preserving hippocampal synaptic integrity in the face of oxidative stress and mitochondrial dysfunction [3,7]. However, the specific mechanisms underlying their beneficial actions remain unclear.

ATP-sensitive potassium (K$_{ATP}$) channels act as metabolic sensors, coupling cellular metabolism with neuronal activity by enhancing K$^+$ efflux. It is well known that plasmalemmal or surface K$_{ATP}$ (sK$_{ATP}$) channels are activated under conditions of metabolic stress [8–10]. K$_{ATP}$ channels localized to the mitochondrial inner membrane (i.e., mitoK$_{ATP}$ channels) may regulate mitochondrial homeostasis by modulating electron transport and calcium buffering [11]. Both sK$_{ATP}$ and mitoK$_{ATP}$ channels have been implicated in models of tissue injury, notably in the heart and brain [11–14]. Activation of these channels may inhibit mitochondrial permeability transition (mPT), a critical determinant of cell death [15–18]. Recently, Yellen and colleagues reported that ketones inhibit spontaneous firing of substantia nigra pars reticulata neurons through sK$_{ATP}$ channels, likely by increasing their open probability [19,20]. In the present study, we sought to determine whether the functional neuroprotective effects of ketones against hydrogen peroxide (H$_2$O$_2$)-induced impairment of hippocampal long-term potentiation (LTP) are mediated through either sK$_{ATP}$ and/or mitoK$_{ATP}$ channels.

### Materials and Methods

**Preparation of hippocampal slices and electrophysiology**

All animal handling protocols were approved by the Institutional Animal Care and Use Committees at the Barrow Neurological Institute (Protocol number 405/308) and the University of Calgary (AC11-0047). Transverse hippocampal slices (400-μm thickness) were prepared from brains of 5- to 6-week-old Sprague Dawley rats or Kir6.2KO mice (Kir6.2$^{-/-}$). Age-matched Kir6.2+/+ mice (wild-type mice; WT) were used as controls. Following decapitation, the whole brain was rapidly isolated and submerged in ice-cold oxygenated physiological saline (composition in mM: 124 NaCl, 1.8 MgSO$_4$, 4 KCl, 1.25 NaH$_2$PO$_4$, 26 NaHCO$_3$, 2.4 CaCl$_2$, and 10 D-glucose; pH: 7.4). Slices were cut using a vibratome (The Vibratome Company, St. Louis, MO), and then transferred to an incubation chamber containing physiological saline bubbled with 95% O$_2$/5% CO$_2$ at 35°C for 1 hr. Each slice was transferred to a submersion-type recording chamber affixed to a Zeiss AxioSkop FS2 microscope and superfused with warm (31 ± 1°C) physiological saline at a rate of 2–3 ml/min before the start of each experiment. To measure changes in synaptic transmission, extracellular population spikes (PS) were evoked by stimulation of Schaffer collaterals (SC) using a bipolar concentric electrode, and responses were recorded in stratum pyramidale of CA1 with a recording electrode (2–6 MΩ tip resistance, backfilled with 2 mM NaCl) connected to a Multiclamp 700A amplifier and digitized with a Digidata 1322A (Axon Instruments). The electrical stimulus was a 100 μs pulse with an intensity (20–100 μA) set at 50% of maximal PS amplitude. After obtaining a stable evoked PS, changes in PS amplitude following application of reagents were digitally stored for later off-line analysis. Upon SC stimulation, excitatory postsynaptic potentials (EPSP) were recorded at a control test frequency of 0.05 Hz (0.1ms, 20–100 μA) from stratum radiatum of CA1 hippocampus. An input-output curve (i.e., stimulus intensity vs. EPSP amplitude) was constructed and the baseline EPSP amplitude (over 1 mV) was set to 30–40% of the maximum response. To measure changes in synaptic plasticity, long-term potentiation (LTP) was evoked by
theta-burst stimulation (TBS) consisting of 5 spike trains delivered at 0.2 Hz (each train, five stimuli at 0.1ms, 100 Hz). LTP is a form of synaptic plasticity broadly used as an electrophysiological measure of learning and memory. LTP was expressed as the percent of mean baseline EPSP amplitude. Paired-pulse facilitation (PPF) was tested to measure changes in presynaptic functions [21]. The interval between two stimuli varied from 50 to 100, 200, and 300 ms. Following PPF, EPSP amplitude was normalized by indicating the ratio (second EPSP/first EPSP). Recorded data were filtered at 3 kHz, sampled at 10 kHz using pClamp, and analyzed with Clampfit.

**HT-22 cells and cell viability test**

Murine hippocampal HT-22 cells were provided as a gift by Dr. Richard Dargusch (The Salk Institute)[22] and fed with Dulbecco’s Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 100 U/mL penicillin, and 100 mg/mL streptomycin. Cells were maintained in a humidified incubator with 5% CO2 at 37°C. HT-22 cells were seeded at a density of 4,000 cells per well in 96-well cell culture plates and grown overnight. Subsequently, cells were pretreated with either ketone alone (BHB or ACA; each 1 to 3 mM) or a ketone cocktail (BHB and ACA; each 1 mM) overnight followed by the addition of H2O2 (200 μM to 1 mM) for 3, 6, 9, 12, and 24h. Cell viability after H2O2 administration was evaluated by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) reduction assay. Twenty μl of MTT (2.5 mg/ml) was added to the wells and incubated for 1h. After incubation, the medium was removed from the wells and 100 μl of dimethylsulfoxide (DMSO) was added to dissolve the formazan reagent. The absorbance was read using a microplate reader at 565 nm. Cell viability was expressed as a percentage of the control group.

**ATP assay**

Tissue samples were obtained as previously described from the CA1 region of rat brain slices (n = 12 in each experimental group from five rats) using 18-gauge needles [3]. Briefly, a mixture of each sample in 5% trichloroacetic acid (100-fold dilution, volume/weight) was sonicated for 10 min and then centrifuged at 14,000 g for 2 min at 4°C. Ten μl of supernatant from each sample was diluted 1,000-fold in phosphate-buffered saline and then mixed with the Enlighten ATP assay kit reagents (Promega, Medison, WI, USA) according to the manufacturer’s instructions. Light intensity after final reaction was measured in a TD-20/20 Luminometer (Turner Biosystems, Sunnyvale, CA, USA) and then compared to an ATP standard curve collected from serial dilutions of an ATP stock solution.

**Drugs and statistical analysis**

All reagents used in this study were purchased from Sigma-Aldrich (St. Louis, MO), unless stated otherwise. Hydrogen peroxide (H2O2) was made fresh for each trial from stock solution; D-β-hydroxybutyrate (BHB); acetocetate (ACA); and 5-hydroxydecanoate (5-HD) were also prepared immediately before each experiment and directly dissolved in physiological saline. While ACA is known to be spontaneously decarboxylated to acetone, once dissolved in saline, it is not volatile. Diazoxide (DZ) was added to physiological saline as a 20 mM stock solution dissolved in NaOH (0.1 M), after which the pH was adjusted to 7.4 with HCl. Glibenclamide (Glb) was dissolved in dimethyl sulfoxide (DMSO). Numerical data are expressed as the mean ± SEM for n slices. Student t-test or ANOVA were performed to determine the presence of significant variance in data from different experimental groups. p <0.05 was considered to be significant.
Results

Ketones preserve synaptic integrity against hydrogen peroxide (H_2O_2)

Hippocampal slices from normal rats displayed robust LTP following theta-burst stimulation (TBS) in physiological saline. The EPSP amplitude measured 256 ± 13% and 159 ± 5% at peak and 60 min post-TBS, respectively (Fig. 1A). Perfusion of 2 mM H_2O_2 in hippocampal slices gradually led to a decrease in baseline EPSP amplitude, and an eventual failure of TBS to induce LTP (Fig. 1A, D). Interestingly, while the baseline EPSP amplitude was not altered in response to 200 μM H_2O_2, LTP was still impaired (Fig. 1A). Thus, in hippocampal slices exposed to H_2O_2, the EPSP amplitude at 60 min post-TBS was significantly decreased compared to the control group (p<0.01, Fig. 1D), confirming that exogenous application of H_2O_2 disrupts hippocampal synaptic plasticity in a dose-dependent manner.

Having established the adverse effects of H_2O_2, we then examined the effects of ketones on oxidative impairment of hippocampal LTP. Since brain and serum ketone levels have been reported to be in the range of 0.5–5 mM during the suckling period in immature rodents and during treatment with the KD in humans [23–26], we used a similar concentration range (BHB or ACA; each 1 to 3 mM) in this study. In hippocampal slices exposed to ketones alone, no differences were seen in the EPSP amplitude between control and ketone-applied groups at 60 min post-TBS (Fig. 1A, B, D). When a ketone alone (BHB or ACA; 1 mM each) was co-applied with 200 μM H_2O_2, LTP was only partially inhibited; the EPSP amplitude of these two groups at 60 min post-TBS was significantly decreased compared to that of controls (p< 0.05, Fig. 1B, D). In contrast, LTP was sustained following co-application of a ketone cocktail (ACA and BHB, each 1 mM) and 200 μM H_2O_2; there was no difference between the EPSP amplitude of ketone-treated hippocampal slices compared to controls (Fig. 1C, D). And as expected, the deficit in LTP induced by 200 μM H_2O_2 was reversed by 3 mM ketone (Fig. 1D). Despite the slightly increased EPSP amplitude post-TBS compared to 2 mM H_2O_2 alone, LTP impairment was maintained in the presence of a cocktail of ketones and 2 mM H_2O_2 (Fig. 1C, D).

Next, we determined whether ketones would preserve normal hippocampal synaptic transmission when challenged with H_2O_2. Synaptic transmission was progressively depressed during perfusion of 2 mM H_2O_2; the PS amplitude measured 18 ± 3% of baseline after a 15 min infusion with 2 mM H_2O_2, and then recovered to 84 ± 5% of baseline following a 30 min washout (Fig. 1E). Despite a clear deficit in LTP produced by 200 μM H_2O_2, changes in PS amplitude were not seen in the presence of 200 μM H_2O_2 (Fig. 1E). Further, no change in the PS amplitude was found with an infusion of a ketone cocktail (Fig. 1E, F). Upon application of a ketone cocktail with 2 mM H_2O_2, ketones prevented the synaptic depression under conditions of H_2O_2 exposure; the evoked PS amplitude measured 72 ± 4% and 59 ± 5% of baseline at 15 min and at 30 min time-points, respectively. Additionally, a 30-min washout restored the PS amplitude to 92 ± 5.4% of baseline (Fig. 1E, F). The synaptic protection was potentiated in hippocampal slices with a 10 min pre-incubation with ketones (Fig. 1F). The PS amplitude of the ketone pre-incubated group was significantly different than that seen when a ketone cocktail and H_2O_2 were co-applied after a 30 min infusion (p< 0.05; Fig. 1F). These data indicate that physiological concentrations of ketones can mitigate hippocampal synaptic depression under conditions of oxidative stress.

Activation of K_{ATP} channels provides synaptic protection against H_2O_2

Diazoxide (DZ) is an activator of both sK_{ATP} and mitoK_{ATP} channels, and has been demonstrated to exert cellular protective effects in pre-ischemic preconditioning and against oxidative injury [16,27]. While both types of K_{ATP} channels have clearly been shown to mediate
Fig 1. Ketones ameliorate synaptic impairment caused by hydrogen peroxide (H$_2$O$_2$). (A) LTP recorded in rat CA1 hippocampal slices exposed to various concentrations of H$_2$O$_2$. Theta-burst stimulation (TBS) of schaffer collaterals led to robust LTP changes when either ACA or BHB (3 mM each) were perfused, but not with either 200 μM or 2 mM H$_2$O$_2$. Representative traces of excitatory post-synaptic potentials (EPSPs) at respective time-points (a, b, c) depicted on the right of each panel. Vertical arrows in this and following figures indicate the time-point of TBS initiation. Dotted line denotes the baseline field potential amplitude which is the mean EPSP during a 10 min physiological saline infusion. (B) Only partial inhibition of LTP was observed when 200 μM H$_2$O$_2$ was co-applied with either ACA or BHB (1 mM each). (C) Intact LTP formation was seen when a cocktail of ketones (BHB and ACA, 1 mM each) was used in conjunction with 200 μM H$_2$O$_2$; but not with 2 mM H$_2$O$_2$. (D) Summary bar graph indicating changes in EPSP amplitudes at 60 min after TBS amongst various treatment groups. Dose-dependent protection by ketones against LTP impairment by H$_2$O$_2$-induced oxidative stress was seen in these experiments. Each vertical bar represents the EPSP amplitude ± SEM (obtained in 10 slices from 5 rats). One way ANOVA followed by Tukey post-hoc analysis; *, p < 0.05; ** or ##, p < 0.01, NS, not significant. (E) Acute application of H$_2$O$_2$ results in depression of the population spike (PS) in stratum pyramidale of Cornu Ammonis (CA)1. Shown are changes in the mean (±SEM) PS amplitude before, during, or after H$_2$O$_2$ infusion, but only when 2 mM H$_2$O$_2$ was used. Representative traces of the PS alone (top), or co-applied with 2 mM H$_2$O$_2$ (middle) or 2 mM H$_2$O$_2$ (bottom) depicted on the right. Pre-incubation with ketones potentiated the restoration of PS amplitude compared to co-application of ketones with H$_2$O$_2$. (F) Summary of amplitude changes in the PS (reflected as % of baseline) during or after drug application. Each vertical bar represents PS amplitude ± SEM, and data were collected from 12 slices from 5 rats. The dotted line reflects the baseline (100%) control PS amplitude before drug treatment. Asterisks denote significant differences between control and treatment groups (*, p < 0.05; ***, p < 0.001), whereas # indicates significant differences between the ketones plus H$_2$O$_2$ group, and other groups in which pretreatment of ketones occurred before H$_2$O$_2$ application (p < 0.05).

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neuroprotective actions under various pathological conditions, their effects on synaptic neurotransmission have yet to be fully elucidated. To investigate whether $K_{ATP}$ channels affect synaptic transmission, we tested the effects of DZ on the reduction in PS amplitude caused by $H_2O_2$. Upon co-application of 300 $\mu$M DZ plus 2 mM $H_2O_2$, the PS amplitude remained constant and similar to control values (Fig. 2A, B). Further, when DZ was pre-incubated with 300 $\mu$M $H_2O_2$ for 10 min, no differences in PS amplitude were seen compared to controls (Fig. 2A). Pre-incubation with 100 $\mu$M DZ also prevented PS depression caused by $H_2O_2$, but the recovery rate of the PS amplitude was significantly slower than that seen with 300 $\mu$M DZ.
The PS amplitude measured 62 ± 3% of control at 15 min post 100 μM DZ plus H2O2 co-application, and then recovered to 92 ± 6% of control after a 30 min washout (Fig. 2A, B). Taken together, these results show that DZ can exert protective effects against H2O2-induced impairment of the hippocampal PS.

To determine whether KATP channels can influence hippocampal synaptic plasticity during oxidative stress, we measured the effects of DZ (10 μM to 300 μM) on LTP impairment induced by 200 μM H2O2. Normal LTP was sustained at hippocampal synapses when exposed to 300 μM DZ alone or when 300 μM DZ and H2O2 were co-applied; the EPSP amplitude at 50 min post-TBS measured 178 ± 15% and 172 ± 10%, respectively (Fig. 2C). However, lower concentrations of DZ were unable to fully prevent H2O2-induced LTP impairment; the EPSP amplitude of the 100 μM DZ plus H2O2 group measured 162 ± 12% and 142 ± 7% at peak and at 50 min post-TBS, respectively (Fig. 2C). And 10 μM DZ had no effect on the reduced EPSP amplitudes caused by 200 μM H2O2. Paired-pulse facilitation (PPF) did not alter at hippocampal synapses when a KB cocktail (BHB and ACA; 1 mM each) was applied, compared to physiological saline alone (Fig. 2D). When 100 μM DZ was infused, the EPSP amplitude of the second response relative to the first resulted in a slight increase in facilitation. However, one-way ANOVA did not find any overall difference amongst the experimental groups. Collectively, these data indicate that the activation of KATP channels in response to DZ can suppress synaptic impairment in oxidative stress, leading to restoration of synaptic integrity.

Functional neuroprotection induced by ketones arises from KATP channels opening and ATP generation

A recent study has shown that ketones can control neuronal firing via KATP channels [28]. These observations, together with a previous finding that pharmacological blockade of KATP channels with 5-hydroxydecanoate (5HD) abolished DZ-mediated synaptic protection [29], led us to investigate whether the synaptic protection provided by ketones against oxidative stress could be reversed by 5HD. When hippocampal slices were exposed to 200 μM 5HD alone, no changes in the PS amplitude were seen (Fig. 3A). However, a serial infusion of 200 μM 5HD that included a cocktail of ketones (ACA and BHB, 1 mM each) and 2 mM H2O2 at 10 min intervals resulted in a tendency toward synaptic depression. 5HD exerted a similar blockade when 100 μM DZ was used, but increasing the DZ concentration to 300 μM negated this action of 5HD (Fig. 3A). After application of 200 μM 5HD, there was no detectable change in LTP formation (Fig. 3B). However, pre-treatment with 5HD abolished the ketone-induced preservation of LTP: LTP measured 181 ± 23% and 110 ± 9% at peak and at 60 min post-TBS, respectively. Further, LTP was not sustained when 5HD was added to 100 μM DZ and H2O2; LTP measured 115 ± 11% at 60 min post-TBS (Fig. 3B). These results differed significantly from both control conditions and with 200 μM H2O2 alone (p < 0.01).

Following these observations, we measured changes in the viability of murine hippocampal HT22 cells to investigate long-term effects of ketones against H2O2-induced oxidative stress. While H2O2 application for 3, 6, 12, and 24 hrs dose-dependently increased cell death, these effects were significantly ameliorated by pre-treatment with ketones (BHB or ACA; each 1 mM or 3 mM) (Fig. 3C). Consistently, using a live cell imaging system, HT22 cells exposed to H2O2 and a cocktail of ketones (BHB and ACA, 1 mM each) appeared healthy with preserved confluence, compared to cells treated with 200 μM H2O2 alone for 24 hrs (Fig. 3D). Although all ketone treatments enhanced cell viability against oxidative stress, pretreatment with ACA provided more effective protection than BHB alone. Co-application of H2O2 with a cocktail of ketones resulted in a reduction in cell death (Fig. 3E). In contrast, when 200 μM 5HD was co-applied with ketones, neuronal protection against oxidative stress was lost (Fig. 3E).
Consistent with our earlier finding that ketones were able to raise ATP levels [3], samples taken from ketone-treated hippocampal slices used in electrophysiological experiments exhibited a slight increase in ATP levels (Fig. 3F). Hippocampal slices exposed to 2 mM H$_2$O$_2$ showed a decrease in ATP levels to 69 ± 7.4% of baseline after 2 hrs exposure, whilst co-appllication of ketones and H$_2$O$_2$ restored ATP levels to 94 ± 6.6% of control. These beneficial effects were reversed by 5-HD; ATP levels measured 75 ± 7.1% of baseline (Fig. 3F). Overall, these findings strongly support the notion that the metabolic regulation and synaptic protection of ketones is tightly linked through their action on K$_{ATP}$ channels.

Both mitoK$_{ATP}$ channels and sK$_{ATP}$ channels are necessary for ketone-induced LTP protection

Despite clear evidence that K$_{ATP}$ channels are involved in ketone-mediated synaptic protection, our initial pharmacological experiments with DZ and 5HD did not delineate the relative contributions of sK$_{ATP}$ vs. mitoK$_{ATP}$ channels. Hence, to determine which, or if both, channels...
are involved in the protective effects of ketones, we utilized Kir6.2 knockout (KO) mice lacking the expression of sKATP channels, but still retain functional mitoKATP channels [30]. TBS produced robust LTP in slices from wild-type (WT or Kir6.2+/+) mice. Hippocampal slices from WT mice exposed to H2O2 showed impairment of LTP at 60 min following TBS. As with rat hippocampal slices, the LTP deficit induced by H2O2 in WT mice was reversed by co-application of either ketones or DZ (Fig. 4A, E). In Kir6.2 KO slices, TBS produced intact hippocampal LTP (8 slices from 4 KO mice; Fig. 4B, E), and H2O2 induced a degree of LTP impairment similar to that seen in WT slices treated with H2O2 (Fig. 4B, E). Application of either ketones or DZ along with H2O2 did not fully preserve LTP formation in hippocampal slices from Kir6.2–/– mice (8 slices from 4 mice; Fig. 4C, E). In agreement with our LTP data, working memory deficits in Kir6.2 KO mice were age-dependent manner; disruption was seen at 12 weeks of age but not at 5 weeks [31].

An earlier study showed that pharmacological blockade of sKATP channels with glibenclamide (Glb) ameliorated LTP formation after high-frequency stimulation [32]. We found that Glb has a tendency to slightly reduce LTP in hippocampal slices of WT mice, but no differences in EPSP amplitude were observed 60 min after TBS amongst these groups (WT, WT/H2O2/ke- toes, and WT/H2O2/DZ; Fig. 4D, E). Cotreatment of ketones and H2O2 was unable to fully preserve LTP when Glb was bath-applied (Fig. 4D, E), suggesting that the synaptic protection

Fig 4. Both mitochondrial- and surface- KATP channels are necessary for ketone-mediated synaptic protection. (A) TBS-evoked intact LTP in wild-type slices decayed to baseline levels 60 min after H2O2 application but was maintained in slices incubated with either ketones or DZ (n = 10 slices; 5 WT mice; *** p < 0.001). (B) Loss of functional Kir6.2 channels did not affect TBS-induced LTP. However, LTP in Kir6.2 KO slices was impaired by H2O2 to a basal level indistinguishable from WT slices exposed to H2O2. (C) Partial blockade of LTP in Kir6.2 KO mice. Changes in LTP from Kir6.2 KO slices were observed in the presence of either ketones (ACA and BHB, each 1 mM) or DZ (100 μM) with H2O2 (200 μM). In contrast, addition of 5-HD in this condition resulted in the complete blockade of LTP formation (n = 8 slices from 4 KO mice). (D) Pharmacological blockade of sKATP channels with glibenclamide (Glb; 10 μM) did not significantly alter LTP formation, although it did produce a slight reduction in the EPSP amplitude post-TBS. When Glb was bath-applied, LTP impairment induced by H2O2 was not fully reversed by ketone application. (E) Bar graph summarizing mean EPSP amplitudes (±SEM) at 60 min after TBS in various treatment groups involving KO mice and WT mice. ANOVA followed by Tukey test; * p < 0.05; ** p < 0.01. N.S. = not significant.

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afforded by ketones might partially involve sK\textsubscript{ATP} channels. Additionally, the mitoK\textsubscript{ATP} channel inhibitor, 5HD, completely abolished the partial LTP protection induced by either ketones or DZ in Kir6.2 KO mice (Fig. 4C, E). Collectively, these data indicate that both sK\textsubscript{ATP} and mitoK\textsubscript{ATP} channels mediate in part the synaptic protective effects of ketones against oxidative stress.

**Discussion**

The major finding of the current study is that ketones—at physiologically relevant concentrations—preserve hippocampal synaptic integrity against oxidative stress, likely in part through activation of both sK\textsubscript{ATP} and mitoK\textsubscript{ATP} channels. The protective effects of ketones against an exogenous H\textsubscript{2}O\textsubscript{2} challenge were mirrored by DZ, an activator of sK\textsubscript{ATP} and mitoK\textsubscript{ATP} channels, and genetic ablation of sK\textsubscript{ATP} channels in Kir6.2-/- mice resulted in only partial recovery of hippocampal LTP by ketones and DZ. While a direct action of ketones on K\textsubscript{ATP} channels could not be demonstrated, similar to earlier studies [19,20,33], our data indicate a key role for both types of ATP-sensitive potassium channels.

It is well known that oxidative stress is a critical factor for the pathogenesis of NDs [34,35]. Oxidative injury induces synaptic impairment, and ultimately cell death, through mitochondrial dysfunction and decreased ATP production, among other actions [3,7,36]. Further, *in vitro* application of H\textsubscript{2}O\textsubscript{2} results in dose-dependent but site-specific injury (i.e., stratum pyramidale vs. stratum radiatum) [37,38]. Although the reasons behind the differential localization-related effects of oxidative stress remain unclear, a major factor may be the resulting region-specific energy deficits that modulate synaptic plasticity [39]. In this context, we found that hippocampal ATP levels correlated well with both cell viability and synaptic integrity as assessed by an established measure of LTP. Specifically, the protective effects of ketones may be related to their ability to enhance ATP production, as well as their antioxidant properties as demonstrated earlier [3,40]. Both of these effects help stabilize the resting membrane potential through preserved Na\textsuperscript{+}-K\textsuperscript{+}-ATPase function and scavenging of reactive oxygen species (ROS) [41].

Importantly, we show in the present study that pharmacological blockade of K\textsubscript{ATP} channels with 5HD results in the elimination of ketone-induced synaptic protection and favorable bioenergetic changes in rat hippocampus. Similarly, we observed that 5HD negated the partial protective effects of both ketones and DZ in Kir6.2 KO mice. Collectively, our data support the notion that ketones, through interactions with both sK\textsubscript{ATP} and mitoK\textsubscript{ATP} channels, promote mitochondrial homeostasis by enhancing ATP generation and antioxidant activity, consistent with previous studies showing that KD treatment—and possibly the resulting ketosis—elevate cellular antioxidant capacity via improvement in mitochondrial redox status [42,43].

Earlier studies have indicated that ketones may exert neuronal inhibitory effects through sK\textsubscript{ATP} channels [19,20], likely by increasing their open probability. However, given our finding that ketones increase ATP levels (both in normal hippocampus as well as that exposed to oxidative stress) and that sK\textsubscript{ATP} channels are known to be inhibited by high ATP/ADP ratios [44], there is an obvious discrepancy that needs to be reconciled. There is abundant evidence in the literature supporting the view that reductions in intracellular ATP/ADP ratios activate sK\textsubscript{ATP} channels, leading to stabilization of the membrane potential, reduced synaptic excitability and enhanced mitochondrial redox status, all of which contribute to resistance against neuronal damage [19,45–48]. Further, it has been hypothesized that there may be compartmental differences in cellular ATP levels, such that the space immediately subjacent to sK\textsubscript{ATP} channels may actually have lower ATP/ADP ratios than elsewhere [19]. While the underlying basis for this discrepancy remains unclear, it is possible that ketones may directly bind to and activate K\textsubscript{ATP} channels, despite cellular ATP levels remaining in a normal or elevated range [20].
Our findings that ketone application in 10 mM glucose-dissolved physiological saline and culture media containing 5.5 mM glucose conferred neuronal protection and synaptic restoration through KATP channels was somewhat surprising since earlier studies demonstrated that the normal brain glucose detected range of 0.82 to 2.4 mM [49] and the hyperglycemia actually negated the open probability of KATP channels [50]. On the other hand, Lund et al [51] recently reported that neurons cultured in BHB resulted in a strong alleviation of glycolytic intervention, likely by suppressing glucose metabolism seen with ketones [52]. While there is no direct evidence to elucidate the functional interplay between ATP level and glycolytic activity within ketone application, we cannot exclude the possibility that the decreased ATP supply originating from the glycolysis pathway may be indirectly responsible for enhancing the opening probability of KATP channels. Alternatively, there is compelling data that KATP channels can also be activated by various hormones and neurotransmitters through G-protein coupled interactions, cAMP, and protein kinase A [8,9,53]. In this regard, BHB has been shown as an agonist of the G-protein coupled free fatty acid 3 receptor in rat sympathetic neurons [54].

While KATP channels have been extensively linked to models of NDs and neuroprotective actions, notably as a mechanism for pre-ischemic conditioning [55,56], the functional role of these channels in synaptic neurotransmission remain less clear. However, with respect to cognition, targeted deletion of Kir6.2 results in age-specific disruption of working memory [31]. In Kir6.2 KO mice aged 5–6 weeks, we found intact LTP responses to TBS, but neither ketones nor DZ could rescue LTP impairment caused by oxidative stress in these mice. Clearly, the precise relationships between sKATP channels and synaptic plasticity during brain development have yet to be clarified [57].

In conclusion, our findings demonstrate that ketones can protect against oxidative impairment of hippocampal LTP, likely through activation of both sKATP and mitoKATP channels. These actions subsequently help restore neuronal, synaptic, mitochondrial, and metabolic function. Further, our data underscore the therapeutic relevance of KTX-0101 (a highly ketogenic medium chain triglyceride formulation) in the treatment of AD [4], and of DZ in the long-term treatment of 3xTgAD mice which has been shown to improve learning and memory function, and to reduce anxiety [33]. KATP channels represent yet another target mechanism that can potently modulate synaptic function, especially under conditions of mitochondrial dysfunction and oxidative stress, both of which have been implicated in various NDs.

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Author Contributions

Conceived and designed the experiments: DYK HJD JMR. Performed the experiments: DYK MGA DO SHL. Analyzed the data: DYK RJT PGS. Contributed reagents/materials/analysis tools: DYK JMR. Wrote the paper: DYK JMR.

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