Viral microRNAs Target a Gene Network, Inhibit STAT Activation, and Suppress Interferon Responses

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Kaposi's sarcoma-associated herpesvirus (KSHV) encodes 12 pre-microRNAs during latency that are processed to yield ~25 mature microRNAs (miRNAs). We were interested in identifying cellular networks that were targeted by KSHV-miRNAs and employed network building strategies using validated KSHV miRNA targets. Here, we report the identification of a gene network centering on the transcription factor signal transducer and activator of transcription 3 (STAT3) that is targeted by KSHV miRNAs. KSHV miRNAs suppressed STAT3 and STAT5 activation and inhibited STAT3-dependent reporter activation upon IL6-treatment. KSHV miRNAs also repressed the induction of antiviral interferon-stimulated genes upon IFNα-treatment. Finally, we observed increased lytic reactivation of KSHV from latently infected cells upon STAT3 repression with siRNAs or a small molecule inhibitor. Our data suggest that treatment of infected cells with a STAT3 inhibitor and a viral replication inhibitor, ganciclovir, represents a possible strategy to eliminate latently infected cells without increasing virion production. Together, we show that KSHV miRNAs suppress a network of targets associated with STAT3, deregulate cytokine-mediated gene activation, suppress an interferon response, and influence the transition into the lytic phase of viral replication.
STAT3 is a latent transcription factor that is activated in response to cytokine-induced stimuli. Cytokine engagement to their receptors activates the receptor-associated Janus kinases (JAK) that phosphorylate STAT3 on a critical tyrosine residue (Y705). The Y705-phosphorylated STAT3 (pY705-STAT3) molecules dimerize, translocate into the nucleus where they bind to promoter regions of their target genes and activate their transcription. STAT3 also gets phosphorylated on its C-terminal serine residue (pS727-STAT3); this process can be mediated by many kinases like PKCδ and IRAK1. STAT3 is also a key player in the innate immune response involving type-I interferons (IFN) like IFN-α/β to counteract viral infections. Binding of IFNs to their receptors leads to the phosphorylation of various STAT proteins including STAT1 and STAT3. This is followed by the formation of STAT complexes that can promote the expression of inflammatory cytokines and antiviral interferon-stimulated genes (ISGs).

This network of KSHV miRNA targets centering on STAT3 also contained several novel KSHV miRNA targets like erythropoietin receptor (EPOR), hepatocyte growth factor receptor (MET), the antiapoptotic protein-baculoviral IAP repeat containing 5 (BIRC5), growth arrest and DNA damage 45 family member (GADD45β) and protein kinase C-δ (PRKCD or PKCδ). STAT3 was one of the most down-regulated proteins in proteomic screens performed to identify KSHV miRNA targets. Other validated KSHV miRNA targets such as interleukin-1 receptor-associated kinase 1 (IRAK1) and GRB2 were also present in this network. Further, we observed that HUV-ECs that were transfected with KSHV miRNAs that repress STAT3 demonstrated a weakened response to cytokines such as IL6 and the antiviral interferon, IFN-type A (also known as IFN-α or IFN-α2A). STAT3 repression using siRNAs or with chemical inhibitors in latently infected BCBL-1 cells promoted entry into the lytic phase of the viral life cycle. Together, our results show that KSHV miRNAs suppress gene expression in this STAT3 network, inhibit cytokine-mediated gene activation, defend against antiviral interferon responses, and regulate lytic reactivation.

Results

miRNA-target analysis reveals canonical and non-canonical base-pairing interactions in the 3′UTRs. Using a combination of microarray and proteomic approaches with gain and loss of viral miRNA activity, we have previously reported the identification of numerous mRNA targets of KSHV-encoded miRNAs (BCLAF1, TNFRSF12A, IRAK1, STAT3, TPM1). Using these previous expression datasets, we integrated the data using a rank sum method as described previously in order to select genes with the highest expression changes in response to gain or loss of viral miRNA function. Importantly, in this analysis, we ignored miRNA seed-matching information and ranked potential miRNA targets solely by their expression changes in response to individual KSHV miRNAs and KSHV infection. We then cloned 49 different full-length 3′UTRs of these predicted miRNA target mRNAs (genes changing the most in the microarray analysis described above) downstream of a renilla luciferase gene into a reporter plasmid (pDEST-765) and performed reporter assays in cells in the presence of various KSHV miRNAs. Luciferase activity was normalized to control assays that used parental luciferase reporters (lacking inserted human 3′UTR sequences) and non-targeting negative control miRNA mimics. These data demonstrated that 28 of 49 3′UTRs were repressed by at least one viral miRNA, when using a t-test and a p-value cutoff of 0.05 (Fig. 1A, Complete data in Table S1). We did not use an arbitrary fold-change cut-off value to denote repression, but used a standard t-test to assess the mean fold-change and standard deviation within the same test. However, this screening is likely to be an underestimate of miRNA targets, since we did not test all combinations of all individual viral miRNAs and 3′UTRs. There were also several instances of a single 3′UTR being repressed by multiple KSHV miRNAs (e.g. IRAK1, TPM1), indicating a high level of redundancy in KSHV miRNAs mediated targeting of cellular mRNAs. Importantly, these data also validate dozens of new miRNA targets of viral miRNAs.

There were at least 50 instances of a miRNA repressing a 3′UTR (green boxes, Fig. 1A), but the majority (28 of 50) of these target interactions were missed using a common bioinformatic tool for predicting canonical miRNA targets. Here we use the definition of canonical miRNA sites to include “7 mer-m8”, “7 mer-A1”, and “8 mer” sites. These results suggested that some miRNA:target repression events may function through non-canonical sites by many kinases like PKCδ and IRAK1. STAT3 also gets phosphorylated on its C-terminal serine residue (pS727-STAT3); this process can be mediated by many kinases like PKCδ and IRAK1. STAT3 is also a key player in the innate immune response involving type-I interferons (IFN) like IFN-α/β to counteract viral infections. Binding of IFNs to their receptors leads to the phosphorylation of various STAT proteins including STAT1 and STAT3. This is followed by the formation of STAT complexes that can promote the expression of inflammatory cytokines and antiviral interferon-stimulated genes (ISGs).

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multiple sites by multiple KSHV miRNAs, using both canonical and non-canonical interactions, demonstrating a high level of complexity in their regulation. These validated targets may be of interest in future studies of the functional roles of viral miRNAs.

Figure 1. 3’UTR luciferase assays for predicted targets of KSHV-encoded miRNAs. (A) 293 cells were transfected with 3’UTR luciferase reporter plasmids for the indicated genes along with KSHV miRNA mimics. Cells were harvested at 48 hpt and reporter activity was measured as described in Methods. The table highlights conditions where the luciferase reporter was repressed (p < 0.05) in green boxes. Gray boxes denote untested conditions and black boxes represent tested conditions where no significant repression was observed. (B) Potential non-canonical miRNA targets were identified using miRanda25 and PAR-CLIP sequences7. Mutated bases (“MUT”) are denoted in gray text and underlined. (C) Luciferase assays with the sites in (B) are shown. Activity with each miRNA was normalized to a negative control miRNA (Neg) with each reporter (denoted by joining horizontal line on x-axis). Error bars show standard deviation from three biological replicates. *Represents p < 0.05 and ** is p < 0.01.
A single miRNA can repress gene expression from tens to hundreds of target mRNAs. Further, a single 3′ UTR can have binding sites for multiple miRNAs. We speculated that the KSHV miRNAs could thus repress multiple mRNAs across many biological pathways. We were interested in identifying specific networks of cellular genes that were particularly enriched for viral miRNA targets. After combining validated miRNA targets identified by microarrays (Fig. 1) and proteomics (STAT3), we performed a network interaction analysis using 34 validated KSHV miRNA targets using MetaCore software. This analysis utilized manually-curated associations of genes obtained from peer-review literature. Specifically, we used the software to investigate only direct interactions (e.g. phosphorylation, transcription regulation) between miRNA target genes. These specific interactions and supporting references are detailed in Table S3.

KSHV miRNAs repress multiple mRNAs in the STAT3 signaling network and suppress STAT3 activation upon IL6 treatment. The largest network of directly associating proteins among the 34 miRNA targets consisted of 10 KSHV miRNA targets and centered on STAT3 (Fig. 2). All of the luciferase validations for these targets are included in Table S1. The interactions in this network are based on the publications shown in Table S3. Note this network of 10 genes is a subset of the genes described in Fig. 1 and did not contain the targets with non-canonical miRNA sites detailed in Fig. 1. The interaction network revealed that targets that stimulate STAT3 activity in normal cells- IRAK1, PKCδ, EPOR and MET, were also repressed by KSHV-encoded miRNAs (Fig. 1, Table S1). Some downstream targets of STAT3 such as BIRC5 and GADD45B were also repressed by KSHV miRNAs (Fig. 2). Since multiple activators of STAT3 activity and expression were identified, we hypothesized that KSHV miRNAs may regulate STAT3 activation.

To study how the repression of these different targets by miRNAs affected STAT3 function, we transfected various KSHV miRNAs into HUVECs and measured STAT3 activation upon IL6-treatment. Previously, we have demonstrated, that STAT3 is repressed by KSHV miRs-K6-5, -K8 and –K9*. HUVECs were transfected with miR-K6-5, -K8 and –K9*, following which they were treated with IL6 and levels of pY705-STAT3, pS727-STAT3 and unphosphorylated STAT3 were measured. In cells transfected with negative control miRNA, IL6 treatment resulted in ~10-fold increase in the levels of pY705-STAT3 over untreated controls (Fig. 3A). However, cells transfected with KSHV miRNAs, miR-K6-5, -K8 and –K9*, showed reduced levels of pY705-STAT3 upon IL6 treatment resulted in ~10-fold increase in the levels of pY705-STAT3 over untreated controls (Fig. 3A). Upon IL6-treatment, miR-K12-6-5 and miR-K12-8, also had a ~2-fold reduction in pS727-STAT3 levels (Fig. 3A, bottom panel). We note that we previously measured miRNAs associated with the RNA-induced silencing complex (RISC) as RISC-associated miRNAs are a more accurate measure of functional intracellular miRNA levels. We have previously demonstrated that the transfected KSHV miRNA-mimics are associated with RISC at levels that are comparable to those after de novo KSHV infection. Together, these results and previous STAT3 3′ UTR results demonstrated that KSHV miRNAs could repress STAT3 activation upon IL6-treatment, likely through a combination of targeting STAT3 mRNA and also currently unknown upstream activators of STAT3.

KSHV miRNAs repress Ser/Thr kinases that regulate STAT3. From the 3′ UTR luciferase assays (Fig. 1), we observed that miR-K6-5 strongly repressed the 3′ UTR of PKCδ, a Ser/Thr kinase can phosphorylate STAT3. We measured the levels of PKCδ in HUVECs transfected with miR-K6-5 and observed strong repression in its protein levels (Fig. 3A). It is possible that PKCδ repression contributes the ~2-fold reduction in pS727-STAT3 levels observed upon IL6-treatment. However, the exact mechanism of miR-K6-5-mediated suppression of STAT3 phosphorylation is unknown and may involve a number of miR-K6-5...
targets. We observed that miR-K8 expression resulted in ~2-fold increase in the protein levels of PKCδ (Fig. 3A; right panel). This is likely due to indirect effects of miR-K8 targeting an inhibitor of PKCδ. However, it is to be noted that in our microarray analysis, RNA levels of PKCδ were repressed by ~30% (average log2 value of −0.47) in the context of KSHV infection, when compared to uninfected controls13.

A second Ser/Thr kinase, IRAK1, is also known to phosphorylate and activate STAT319. IRAK1 is also a target of KSHV miR-K921. To study the effect of IRAK1 suppression on STAT3 activation, HUVECs were transfected with miR-K9 and levels of pY705-STAT3 were measured upon IL6-treatment. We observed ~70% reduction in the levels of pY705-STAT3 when compared to miR-Neg (Fig. 3B). There was also ~40% repression in pS727-STAT3 levels upon IL6-treatment, which could be a result of IRAK1 repression.

**Figure 3.** KSHV miRNAs inhibit IL6-dependent STAT3 activation and inhibit STAT3-dependent reporter activation. (A) HUVECs were transfected with KSHV miRNAs (miR-K6-5, miR-K8 or miR-K9*) or control miRNAs (Neg). At 48 hpt, transfected cells were mock-treated or treated with IL6 for 20 minutes. Equal amounts of protein were probed for pY705-STAT3 and total STAT3 (top) and pS727-STAT3 and PKCδ (bottom). Relative levels of various proteins were normalized to actin, compared to the control miRNA (Neg), and plotted on the right (n = 4). (B) HUVECs were transfected with miR-K9 (that targets IRAK1) or siIRAK1 along with appropriate negative control RNAs. Lysates were probed for pY705-STAT3, pS727-STAT3, total STAT3 and IRAK1. Relative levels of various proteins were normalized to actin and plotted on the right (n = 4). (C) The changes in the levels of pY705-STAT3 relative to total STAT3 were measured from 3A and 3B and plotted (n = 3). (D) 293 cells were reverse-transfected with the 4xM67 pTATA-Luc, pRNL-TK control and KSHV miRNAs or siRNAs. Cells were left untreated or treated with IL6 for eight hours and relative luciferase levels were measured as described in the Methods (n = 3). Full-length blots are in Figure S2.
OAS2, MxA, ISG15, IFITM1, IRF1)\(^28\). To determine if KSHV miRNAs can suppress activation of IFN-α, we transfected HUVECs with KSHV miRs-K6-3, -K6-5 and –K10b and measured changes in the levels of pY701-STAT1 upon IFN-α treatment. In HUVECs transfected with miR-Neg control, IFN-α treatment resulted in ~20-fold activation in the levels of pY701-STAT1, compared to untreated controls (Fig. 4A, lanes 1 and 2). However, KSHV miRNA-transfected cells showed ~2-fold reductions in the levels of pY701-STAT1, when compared to miR-Neg controls (Fig. 4A), suggesting an inhibition of this pathway. IFN-α treatment also strongly activated STAT1 in HUVECs, however, KSHV miRNAs had less of an effect on overall pSTAT1 levels compared to pY705-STAT3, suggesting specificity of KSHV miRNAs (Fig. 6A). BIRC5 has been identified as a transcriptional target of STAT3\(^30–32\), but can also inhibit STAT3 activation upon IL6-treatment and inhibit its transcriptional activity.

**STAT3 repression by KSHV miRNAs contributes to inhibition of interferon-mediated antiviral responses.** IFN-α can activate STAT1 as well as STAT3 to elicit antiviral responses\(^27\). We were interested in determining if KSHV miRNAs can suppress IFN-α-mediated STAT3 activation and therefore, inhibit expression of ISGs. HUVECs were transfected with KSHV miRNA mimics following which they were treated with IFN-α for 20 minutes. We measured changes in the levels of pY705-STAT3 and total STAT3 upon IFN-α treatment. As an additional control, we also measured changes in the levels of pY701-STAT1 upon IFN-α treatment. In HUVECs transfected with miR-Neg control, IFN-α treatment resulted in ~20-fold activation in the levels of pY705-STAT3, compared to untreated controls (Fig. 4A, lanes 1 and 2). However, KSHV miRNA-transfected cells showed ~2-fold reductions in the levels of pY705-STAT3, when compared to miR-Neg controls (Fig. 4A), suggesting a strong inhibition of this pathway. IFN-α treatment also strongly activated STAT1 in HUVECs, however, KSHV miRNAs had less of an effect on overall pSTAT1 levels compared to pY705-STAT3, suggesting specificity in repression of STAT proteins (Fig. 4A, Figure S1).

IFN-α has potent antiviral activity and can activate chemokines (e.g. CXCL10) and antiviral proteins (e.g. OAS2, MxA, ISG15, IFITM1, IRF1)\(^28\). To determine if KSHV miRNAs can suppress activation of IFN-α – regulated genes via STAT3 repression, we transfected HUVECs with KSHV miRNAs and treated them for 4 hours with IFN-α and measured changes in the levels of antiviral genes using qRT-PCR. HUVECs transfected with control miRNA, miR-Neg, demonstrated strong increases in the mRNA levels of the aforementioned ISGs upon IFN-α treatment (Fig. 4C). However, upon IFN-α treatment in the presence of KSHV miRNAs, we observed strong decreases in the activation of several ISGs. CXCL10, a ~10 kDa chemokine, was found to be consistently down-regulated (~2-fold) by miR-K6-5 and –K9* (Fig. 4D). A ~2-fold decrease in activation of ISG15, IFITM1 and IRF1 miRNAs were also observed with other KSHV miRNAs that repress STAT3. KSHV miR-K6-5 also repressed the induction of antiviral ISGs- OAS2 and MxA. From Western blot analysis (Fig. 4A and B), it appeared that the KSHV miRNAs that repressed STAT3 did not affect STAT1 levels upon IFN-α treatment. We conclude that KSHV miRNAs can inhibit ISG activation in the presence of interferons.

**KSHV miRNAs repress EPOR mRNA to inhibit EPO-dependent STAT5 activation.** From HUVEC mRNA panels, we identified KSHV miRNAs- K6-3, -K6-5 and -K10b as miRNAs that could strongly repress the levels of the EPO mRNA (Fig. 5A). Binding of erythropoietin (EPO) to EPOR predominantly activates STAT5 as pY694-STAT5 (pSTAT5), but can also activate STAT3\(^28\). However, we were unable to demonstrate STAT3 activation upon EPO-treatment in endothelial cells (Figure S1). To test if KSHV miRNAs can inhibit STAT5 activation, we transfected KSHV miRNAs into HUVECs and treated these cells with EPO. STAT5 activation (as pSTAT5) was observed upon EPO-treatment in HUVECs transfected with miR-Neg (Fig. 5B). HUVECs transfected with KSHV miR- K6-3, -K6-5 and –K10b showed ~2-fold reduction in the levels of pSTAT5 compared to miR-Neg-transfected cells, upon EPO-treatment. However, the levels of unphosphorylated STAT5 protein and JAK2 in these cells did not change dramatically in the presence of these miRNAs. Assessing the ratio of pSTAT5 to total STAT5 demonstrated that the majority of the changes were to phosphorylation of STAT5.

STAT3 and STAT5 are both activated in response to type-I interferons like IFN-α and therefore can regulate antiviral responses\(^29\). It is interesting that KSHV inhibits STAT5 activation in cell types like HUVECs, where STAT3 activation was not observed with EPO. Together, our results suggest a mechanism by which KSHV utilizes miRNAs to inhibit cytokine responses either directly by repression of STAT proteins (e.g. STAT3) or indirectly by repression of their cognate receptors (e.g. EPOR).

**BIRC5 repression by KSHV miRNAs promotes KSHV infection.** In RNA samples from HUVECs transfected with individual KSHV miRNAs, we observed that BIRC5 was repressed by several KSHV-encoded miRNAs (Fig. 6A). BIRC5 has been identified as a transcriptional target of STAT3\(^30–32\), but can also inhibit STAT3 function in the nucleus\(^33\). To identify the miRNA that might directly repress BIRC5, we performed 3′UTR luciferase reporter assays using the vector pDEST-BIRC5 and KSHV miRNA mimics (Fig. 1A, last row; Fig. 6B). Luciferase assays revealed miR-K12-5 as a miRNA that repressed the BIRC5 3′UTR by ~40% relative to the miR-Neg control (Fig. 6B). A search for potential KSHV miRNA binding sites using miRanda revealed a 7-mer binding site for miR-K12-5 in the 3′UTR of BIRC5 (Fig. 6C). We mutated this site to generate pDEST-BIRC5-mut (Fig. 6C, underlined). Luciferase reporter assays performed with pDEST-BIRC5-mut and miR-12-5 did not find repression of the mutant reporter, suggesting that this site might be important for BIRC5 repression.

To understand the contribution of the various network proteins to KSHV infection, we knocked down individual target genes in HUVECs using siRNAs, performed de novo infections with KSHV and measured LANA (~40% repression with miR-K9). No repression of pY705- and pS727-STAT3 were seen with siRNAs against IRAK1, despite robust repression of IRAK1, suggesting that miR-K9 repressed STAT3 activation primarily in an IRAK1-independent mechanism. While repression of pY705-STAT3 levels as a result of KSHV miRNAs could also be an outcome of reduced STAT3 protein levels, we observed for some cases, like miR-K9, that the fraction of phosphorylated STAT3 relative to total STAT3 was reduced (Fig. 3C).

We also measured the effect of KSHV miRNAs on the transcriptional activity of STAT3. We utilized a STAT3-driven firefly luciferase reporter plasmid along with KSHV miRs- K6-5, -K8 or –K9* and transfected 293T cells. As additional controls, siRNAs against STAT3 were used. Transfected 293T cells were treated with 20 ng/ml IL6 and fold activation of the reporter was measured relative to untreated controls (Fig. 3D). In miR-Neg-transfected cells, IL6-treatment resulted in an 8-fold increase in reporter activation, while it was reduced to a 5-fold increase with miR-K6-5 and miR-K9*. However, miR-K8 did not affect reporter activation. siNeg-transfected cells showed ~5-fold increase in reporter activity upon IL6-treatment, while in the siSTAT3-transfected cells, the activity was reduced to ~2-fold. Together, these results suggest that KSHV encoded miRNAs inhibit STAT3 activation upon IL6-treatment and inhibit its transcriptional activity.
mRNA levels. BIRC5 knockdown in HUVECs prior to infection increased LANA levels by ~2-fold, while STAT3 knockdown had no effect on de novo infection (Fig. 6D). In HUVECs, knockdown of STAT3 with siRNAs had no effect on the levels of BIRC5, suggesting that this pathway is either not functional in primary endothelial cells or needs a specific cytokine-mediated stimulus. Together, these data may suggest that repression of BIRC5 influences KSHV infection.

Inhibition of STAT3 in latently infected cells induces KSHV lytic reactivation. In order to study the effects of STAT3 repression in the context of lytic replication, we shifted studies from endothelial cells to PEL cells, since they are a better system to study lytic replication. We reduced STAT3 levels in BCBL-1 cells using siRNAs and measured the levels of the KSHV lytic switch protein, ORF50 or RTA. Strong repression of STAT3 protein levels was observed in BCBL-1 cells at both 24 h and 48 h post-electroporation. Interestingly, in these cells, RTA protein levels were also upregulated at both time points (Fig. 7A). We also measured the mRNA levels of RTA and STAT3 using qRT-PCR and observed similar increases in the levels of RTA upon STAT3 repression. Further, we measured the levels of lytic mRNAs of KSHV, including ORF57, ORF59 and K8 and observed similar increases in their levels upon STAT3 repression (Fig. 7B). It is important to note that these increases in lytic mRNAs occurred in the absence of any chemical inducers of lytic reactivation and hence, were solely mediated by STAT3 knockdown in BCBL-1 cells.

We also inhibited STAT3 function using the STAT3 inhibitor, Stattic and observed a 5-fold increase in the lytic mRNA levels of RTA. Similar increases were also observed in the levels of other lytic markers like ORF57, ORF59 and K8 (Fig. 7B). Antivirals like ganciclovir (GCV) are phosphorylated to GCV-monophosphate by the late lytic proteins- ORF21 and ORF36 of KSHV, following which they can inhibit viral DNA polymerases and...
therefore, viral replication. However, since KSHV remains latent in a majority of PEL cells, these cells are not sensitive to GCV. In contrast, BCBL-1 cells treated with Stattic would be more sensitive to GCV due to an increase in lytic replication and the concomitant increase in viral kinases that phosphorylate GCV. We hypothesized that if we could stimulate latently infected BCBL-1 cells to reactivate with Stattic, then virion production could be inhibited using GCV. To test this hypothesis, we treated BCBL-1 cells with Stattic or a combination of Stattic and GCV. Virions from the cell-free supernatants were harvested two days post-treatment and used to infect HUVECs to determine viral loads via qRT-PCR for LANA mRNA. Consistent with Fig. 7B, we observed almost a 6-fold

Figure 5. KSHV miRNAs repress the levels of erythropoietin-receptor (EPOR) mRNA and reduce erythropoietin-dependent STAT5 activation in endothelial cells. (A) HUVECs were transfected with individual KSHV miRNA mimics along with control miRNAs and the levels of EPOR mRNA were measured 48 hpt by qRT-PCR. B. HUVECs were transfected with KSHV miRNAs (miR-K6-3, miR-K6-5, miR-K10b or miR-K12) or miR-Neg. At 48 hpt, transfected cells were mock-treated or treated with EPO for 10 minutes. Equal amounts of protein were probed for pY694-STAT5, JAK2 and total STAT5. “M” denotes the protein size marker loaded in the center lane. Relative levels of various proteins were normalized to actin and quantified below. Full-length blots are in Figure S3.
increase in virus production in the presence of Stattic when compared to DMSO-treated control cells (Fig. 7C). Drug-treatment of BCBL-1 cells with Stattic did not severely affect cell viability (decrease of viability with Stattic was 6.25% ± 0.5%, by trypan blue assay) and therefore, the increase in virus production is not likely due to cell death. Virus production with a combination of Stattic and ganciclovir was similar to uninduced cells, suggesting that ganciclovir inhibited virus production.

Discussion
We report a protein network centering on STAT3 that is targeted by KSHV-encoded miRNAs to inhibit the action of cytokines, suppress interferon responses and regulate lytic replication. The different members in this network are individually repressed by KSHV miRNAs using both canonical and non-canonical base-pairing interactions in their 3′UTRs. By suppressing the core member of this network- STAT3, KSHV miRNAs are able to inhibit a multitude of pathways involving cytokines like IL6, EPO and IFN. Further, we demonstrate that STAT3 repression (by siRNAs or with drugs) can trigger KSHV lytic replication in PEL cells, as a result of which they become more sensitive to antivirals like ganciclovir.

IFNs play an important role in the establishment of innate and adaptive immunity against viruses. IFNs and the ISGs they activate are widely considered to be tumor-suppressive and therefore, inhibition of this major antiviral pathway can prove to be beneficial for both viral persistence and tumor establishment. Consequently, many viruses including KSHV have evolved elaborate mechanisms to overcome this pathway. KSHV encodes many...
proteins in its genome to counteract the actions of IFNs. For instance, the viral IRF-2 protein (vIRF2) inhibits the formation of the ISGF3 (interferon stimulated gene factor 3) complex upon IFN-α engagement with its receptors. The viral IL-6 protein (vIL6) also inhibits the ISGF3 binding to DNA elements to block the synthesis of ISGs (For review35). In this report, we demonstrate that multiple KSHV miRNAs that repress STAT3 can weaken the innate immune responses to type-I interferons and inhibit induction of antiviral ISGs like IRF1, IFITM1 and ISG15. Among KSHV miRNAs, miR-K11 has been demonstrated to inhibit IFN signaling upon sendai viral infection by targeting IKKε36. Viral miRNA-mediated inhibition of IFN-responses also has precedents in other viruses. For instance, the anneloviral miRNAs were demonstrated to inhibit type-I interferon signaling and promote cell survival under IFN-rich conditions37. KSHV miRNAs can similarly promote spindle cell persistence under high interferon levels that are normally observed in the KS tumor. It is to be noted that vIRF and vIL6 are primarily expressed during the lytic phase; on the other hand, viral miRNAs can inhibit antiviral responses during latency. Previously it was demonstrated that KSHV infection of endothelial cells resulted in a biphasic activation of STAT338. Punjabi et al., observed phosphorylation of STAT3 (at Y705) upon de novo infection of HUVECs. However, we note that HUVECs that were transfected with siSTAT3 prior to de novo KSHV infection, had LANA levels that were comparable to those from cells transfected with control siRNA (Fig. 6D). KSHV miRNAs may facilitate waning of pY705-STAT3 levels at certain time points that was reported after de novo KSHV infection of endothelial cells38. KSHV proteins vGPCR and kaposin B have been shown to increase STAT3 phosphorylation39,40. It is possible that in latently infected cells, proper regulation of IL6 signaling and STAT3 phosphorylation is important for viral persistence. This proper regulation is likely achieved through a mixture of positive and negative regulators. A recent study reported an indirect activation of STAT3 by KSHV-encoded miRNAs (including miR-K6)41. Previously, we tested sixteen KSHV miRNAs in primary endothelial cells and no miRNAs increased STAT3 protein levels35. At the moment, our results do not have information about the role of KSHV miRNAs and STAT3 signaling in B cells. While we acknowledge the differences between these previous studies and our current work, it is essential to note that in certain genetic contexts, e.g. the absence of PTEN, STAT3 can also function as a
tumor suppressor. While PTEN deletions are uncommon in KSHV infections, PTEN inactivation by phosphorylation in KSHV infections have been reported. STAT3 suppression when combined with PTEN inactivation might therefore accelerate cancer progression. The role of other causal mutations that occur in KSHV infection and how STAT3 repression plays into it would require further investigation. Although repression of STAT3 activation is novel for KSHV, it is not an entirely new concept. STAT3 inhibition is known to benefit many viruses. The mumps viral protein can target STAT3 to ubiquitin-mediated degradation and the V protein of measles forms a complex with STAT proteins to prevent nuclear translocation. The P protein of rabies virus associates with STAT3 in an oncostatin M-dependent manner to inhibit STAT3 activation. STAT3 is also inhibited by the protein hormone that is essential for erythropoiesis, acts on endothelial cells to stabilize the vascular matrix. KS of STAT5 activation by KSHV miRNAs in endothelial cells via EPOR repression. Erythropoietin (EPO), a glycoprotein hormone that is essential for erythropoiesis, acts on endothelial cells to stabilize the vascular matrix. KS involves the infection of endothelial cells by KSHV, following which the proliferating ‘spindle-cells’ lead to leaky blood vessels and extravasation of red blood cells at the site of the lesion. EPO repression in endothelial cells might contribute to the loss of vascular architecture that is characteristic of KS. Interestingly, one of the targets of EPO in endothelial cells is thrombospondin 1 (THBS1), an antiangiogenic factor that is a target of KSHV miRNAs. Therefore, loss of EPO expression in endothelial cells can accelerate loss of THBS1 expression due to KSHV miRNAs. Human CMV, a β-herpesvirus, was shown to reduce erythropoietin levels under hypoxic conditions. The observation that KSHV can repress EPOR-dependent STAT5 activation is novel and highlights the differences in the mechanisms adopted by related herpesviruses to inhibit the EPO pathway. We also identify and validate BIRC5 as a novel target of KSHV miRNAs and our data suggests that BIRC5 repression may increase the susceptibility of endothelial cells to KSHV infection. KS of STAT3 in PEL cells enhances KSHV lytic replication and correlates with our findings. We have also extended this to further demonstrate a potential strategy for elimination of latent cells using a combination of a STAT3 inhibitor and ganciclovir (Fig. 7C). It is also possible that modest regulation of STAT3 expression or activation during latency may contribute to the low level of basal lytic replication observed in KSHV infected B-cells. Patients with autosomal dominant hyper-IgE syndrome (AD-HIES or Job’s syndrome) have mutations in their STAT3 gene and this reduces the levels of functional STAT3 protein. EBV-infected cells from AD-HIES patients also have a higher degree of spontaneous lytic reactivation. AD-HIES patients have a higher risk for VZV (varicella zoster virus) reactivation and increased EBV viremia. Lytic reactivation of KSHV with valproate in the presence of antivirals like ganciclovir has shown to induce PEL cell apoptosis. In summary, we have identified several novel targets of KSHV miRNAs and demonstrated how viral miRNAs could regulate responses to cellular stimuli.

**Experimental Methods**

**Cell culture and reagents.** Human umbilical vein endothelial cells (HUVECs; Lonza, Walkersville, MD) were maintained for five passages in complete EGM-2 BulletKit (Lonza). The KSHV-latently infected cell line, BCBL-1, was maintained in RPMI 1640 supplemented with 10% FBS, 1X Penicillin-Streptomycin and 55 μM β-mercaptoethanol. 293 cells were maintained in DMEM supplemented with 10% FBS and 1X Penicillin-Streptomycin. miRVana miRNA mimics of various KSHV-encoded miRNAs were purchased from Ambion (Life Technologies Inc., Carlsbad, CA). ON-TARGETplus SMARTpool siRNAs targeting human STAT3, BIRC5, IRAK1 and the non-targeting pool control were obtained from Thermo Fisher Scientific (Waltham, MA). The STAT3 inhibitor- Stattic, was purchased from Sigma-Aldrich Inc, St. Louis, MO. The STAT3-dependent luciferase expression plasmid, 4xM67 pTATA TK-Luc, was a gift from Jim Darnell and was obtained through Addgene.

**miRNA transfections and Western blotting.** miRNA transfections of HUVECs were performed with 10 nM miRNAs or 16.5 nM siRNAs using Dharmafect (Thermo Fisher Scientific). The transfected cells were harvested 48-hours post-transfection (lpi) and their total protein extracted in radio-immunoprecipitation assay buffer (RIPA, Sigma) containing 1X Halt Protease and Phosphatase inhibitors (Thermo Fisher Scientific). The LI-COR Odyssey Infrared Imaging System (LI-COR Biosciences, Lincoln, NE) was utilized for measurement of protein levels. Primary antibodies were obtained from Cell Signaling Technology Inc., Danvers, MA, unless specified otherwise. Mouse anti-actin primary antibody (AC-74) was obtained from Sigma. The rabbit anti-RTA antibody was a gift from Don Ganem. Secondary antibodies conjugated to infrared fluorescing dyes- IRDye 800CW and IRDye 680, were obtained from LI-COR. Changes in protein levels were measured relative to actin.

**3′UTR luciferase assays.** 3′UTR-dual luciferase reporter assays were performed in 293 cells as reported previously. Briefly, 3′UTRs of various genes were cloned downstream of renilla luciferase gene in the vector, pDEST-765 (Protein Expression Laboratory, Leidos, Frederick, MD) and the HSV-TK-promoter driven firefly luciferase served as an internal control. Upon cotransfection of 293 cells with pDEST-765-3′UTR and KSHV miRNA mimics, we normalized the renilla luciferase values to the internal firefly luciferase control reporter, a parental control reporter lacking cloned 3′UTRs, and a non-targeting negative control miRNA mimic. Cells were
harvested 24 or 48 hpt and dual luciferase reporter assays were performed using the Dual-Luciferase Reporter system (Promega, Madison, WI). Mutations of miRNA-binding sites in 3′UTRs were performed using the QuikChange II Site-Directed Mutagenesis kit (Stratagene, San Diego, CA).

**Cytokine treatments and measurement of phospho-STAT3.** HUVECs were transfected with miRNAs or siRNAs as described above. Forty-eight hours after transfection, the cells were treated with 10 ng/ml of IL6 (Cell Signaling Technology) or 1000 U/ml of IFNα (Sigma) for 20 minutes after which cells were washed once with ice-cold PBS and harvested in RIPA buffer. Erythropoietin treatments of HUVECs were performed with 250 U of EPO (Sigma) for 10 minutes. Equal amount of lysates were used for Western blot analysis to measure levels of pY705-STAT3, pS727-STAT3, STAT3, PKCα, IRAK1, pY701-STAT1, pY694-STAT5 or STAT5 and their levels normalized to that of actin. When necessary, blots were stripped of their primary antibody using the Newbloc Nitro Stripping buffer (LI-COR) and re-probed.

**STAT3 activation luciferase reporter assays.** 293 cells were reverse-transfected with KSHV miRNAs, the STAT3-reporter plasmid-4xM67 pTATA TK-Luc and a control renilla luciferase expression plasmid (pRNL) using Lipofectamine 2000 (Life Technologies). Sixteen hours post-transfection, media was changed to that containing 20 ng/ml IL6. Cells were harvested eight hours post-IL6 addition and luciferase assays were performed as described earlier. Fold activation in response to IL6 addition were calculated relative to untreated controls.

**KSHV purification and infection.** KSHV production was induced in BCBL-1 cells using 1 mM valproic acid for seven days. The Vivaflow 50 tangential flow filtration system (Sartorius Stedim Biotech, Goettingen, Germany) was used to concentrate the virions released into the culture supernatants. KSHV infection of HUVECs was performed in media containing 8 µg/ml polybrene for four hours at 37°C. Media was changed every two days and cells were harvested for analysis by Western blotting or real-time quantitative PCR (qRT-PCR).

**Real-time quantitative PCR analysis.** Total RNA was extracted using the miRNeasy mini kit (Qiagen) and converted to cDNA using the High Capacity cDNA Reverse Transcription kit (Applied Biosystems, Foster City, CA). The following primers were used for measurement of LANA mRNA levels: 5′-GTGACCTTTGGGAGATGCCTA-3′ and 5′-CAGGAGATGGAGAATGAGTA-3′. STAT3 mRNA levels were measured using 5′-GTTGTCTCCTGCTGTATC-3′ and 5′-GGTATTGCTGCAGGTCGT-3′. KSHV lytic miRNAs were measured using primer pairs for RTA, ORF57, ORF59 and K8 as described62. GAPDH mRNA was used as a normalization control. qPCR analysis was performed using FastStart Universal SYBR Green Master (ROX) (Roche Diagnostics GmbH). Taqman Gene Expression Assays targeting cellular genes were purchased from Applied Biosystems.

**BCBL-1 electroporations and measure of lytic replication.** Electroporation of siRNAs into BCBL-1 cells were performed using the Amaxa 4D-Nucleofector (Lonza) with CA-189 program in the Nucleofector buffer SF following manufacturer’s instructions. Cells were harvested 24 hours post-electroporation and the levels of STAT3 and KSHV lytic miRNAs measured using qRT-PCR. BCBL-1 cells were also treated with the STAT3 inhibitor- Static or DMSO as control, for 24 hours and the levels of STAT3 and lytic mRNAs were measured using qRT-PCR. For assessment of viral loads upon drug treatment, BCBL-1 cells were treated with the STAT3 activation luciferase reporter assay and then harvested 24 or 48 hpt and dual luciferase reporter assays were performed using the Dual-Luciferase Reporter system (Promega, Madison, WI). Mutations of miRNA-binding sites in 3′UTRs were performed using the QuikChange II Site-Directed Mutagenesis kit (Stratagene, San Diego, CA).

**Network Analysis.** Luciferase validated miRNA targets (34 in total, 28 from microarrays, 6 from proteomic screen) were input into the MetaCore analysis software (Thomson Reuters). The network was generated using the direct interactions algorithm. This created a network of only the 34 targets and their direct interactions based on the database of interactions in the MetaCore software.

**Statistical significance.** Statistical significance relative to corresponding control samples were analyzed using a Student’s t-test. Significance values of p < 0.05 are denoted by *p < 0.001 by and **p < 0.0001 by ***unless otherwise indicated. In some cases, ANOVA was used instead of t-tests.

**Data Reporting.** Expression data can be retrieved from NCBI GEO using accession numbers GSE12967, GSE40093, GSE24069, GSE65148.

References
1. Pfeffer, S. et al. Identification of microRNAs of the herpesvirus family. *Nat Methods* **2**, 269–276 (2005).
2. Cai, X. et al. Kaposis sarcoma-associated herpesvirus expresses an array of viral microRNAs in latently infected cells. *Proc Natl Acad Sci USA* **102**, 5570–5575 (2005).
3. Samols, M. A., Hu, J., Skalsky, R. L. & Renne, R. Cloning and identification of a microRNA cluster within the latency-associated region of Kaposis sarcoma-associated herpesvirus. *J Virol* **79**, 9301–9305 (2005).
4. Fabian, M. R., Sonenberg, N. & Filipowicz, W. Regulation of mRNA translation and stability by microRNAs. *Annu Rev Biochem* **79**, 351–379 (2010).
5. Loeb, G. B. et al. Transcripencode-wide miR-155 binding map reveals widespread noncanonical microRNA targeting. *Mol Cell* **48**, 760–770 (2012).
6. Chi, S. W., Hannon, G. J. & Darnell, R. B. An alternative mode of microRNA target recognition. *Nat Struct Mol Biol* **19**, 321–327 (2012).
7. Gottwein, E. et al. Viral MicroRNA Targetome of KSHV-Infected Primary Effusion Lymphoma Cell Lines. *Cell Host Microbe* **10**, 515–526 (2011).
8. Ramalingam, D., Kieffer-Kwon, P. & Ziegelbauer, J. M. Emerging themes from EBV and KSHV microRNA targets. *Viruses* **4**, 1687–1710 (2012).
48. Reitsma, J. M., Sato, H., Nevels, M., Terhune, S. S. & Paulus, C. Human cytomegalovirus IE1 protein disrupts interleukin-6 signaling by sequestering STAT3 in the nucleus. J Virol 87, 10763–10776 (2013).

49. Mitzel, D. N., Jaramillo, R. J., Stout-Delgado, H., Senf, A. P. & Harrod, K. S. Human metapneumovirus inhibits the IL-6–induced JAK/STAT3 signaling cascade in airway epithelium. J Gen Virol 95, 26–37 (2014).

50. Reitsma, J. M., Sato, H., Nevels, M., Terhune, S. S. & Paulus, C. Human cytomegalovirus IE1 protein disrupts interleukin-6 signaling by sequestering STAT3 in the nucleus. J Virol 87, 10763–10776 (2013).

51. Mitzel, D. N., Jaramillo, R. J., Stout-Delgado, H., Senf, A. P. & Harrod, K. S. Human metapneumovirus inhibits the IL-6–induced JAK/STAT3 signaling cascade in airway epithelium. J Gen Virol 95, 26–37 (2014).

52. Reitsma, J. M., Sato, H., Nevels, M., Terhune, S. S. & Paulus, C. Human cytomegalovirus IE1 protein disrupts interleukin-6 signaling by sequestering STAT3 in the nucleus. J Virol 87, 10763–10776 (2013).

53. Mitzel, D. N., Jaramillo, R. J., Stout-Delgado, H., Senf, A. P. & Harrod, K. S. Human metapneumovirus inhibits the IL-6–induced JAK/STAT3 signaling cascade in airway epithelium. J Gen Virol 95, 26–37 (2014).

54. Reitsma, J. M., Sato, H., Nevels, M., Terhune, S. S. & Paulus, C. Human cytomegalovirus IE1 protein disrupts interleukin-6 signaling by sequestering STAT3 in the nucleus. J Virol 87, 10763–10776 (2013).

55. Mitzel, D. N., Jaramillo, R. J., Stout-Delgado, H., Senf, A. P. & Harrod, K. S. Human metapneumovirus inhibits the IL-6–induced JAK/STAT3 signaling cascade in airway epithelium. J Gen Virol 95, 26–37 (2014).
50. Lundquist, A. et al. Kaposi sarcoma-associated viral cyclin K overrides cell growth inhibition mediated by oncostatin M through STAT3 inhibition. Blood 101, 4070–4077 (2003).

51. Fiser, D. & Bahlmann, F. H. Erythropoietin and the endothelium—a promising link? Eur J Clin Invest 38, 457–461 (2008).

52. Butler, L. M. et al. Human cytomegalovirus inhibits erythropoietin production. J Am Soc Nephrol 25, 1669–1678 (2014).

53. Chugh, P. E. et al. Systemically circulating viral and tumor-derived microRNAs in KSHV-associated malignancies. PLoS Pathog 9, e1003484, doi: 10.1371/journal.ppat.1003484 (2013).

54. Daigle, D. et al. Upregulation of STAT3 marks Burkitt lymphoma cells refractory to Epstein-Barr virus lytic cycle induction by HDAC inhibitors. J Virol 84, 993–1004 (2010).

55. Hill, E. R. et al. Signal transducer and activator of transcription 3 limits Epstein-Barr virus lytic activation in B lymphocytes. J Virol 87, 11438–11446 (2013).

56. Zeng, Y. et al. Intracellular Tat of human immunodeficiency virus type 1 activates lytic cycle replication of Kaposi’s sarcoma-associated herpesvirus: role of JAK/STAT signaling. J Virol 81, 2401–2417 (2007).

57. King, C. A., Li, X., Barbachano-Guerrero, A. & Bhaduri-McIntosh, S. STAT3 Regulates Lytic Activation of Kaposi’s Sarcoma-Associated Herpesvirus. J Virol 89, 11347–11355 (2015).

58. Minegishi, Y. et al. Dominant-negative mutations in the DNA-binding domain of STAT3 cause hyper-IgE syndrome. Nature 448, 1058–1062 (2007).

59. Holland, S. M. et al. STAT3 mutations in the hyper-IgE syndrome. The New England journal of medicine 357, 1608–1619 (2007).

60. Siegel, A. M. et al. A critical role for STAT3 transcription factor signaling in the development and maintenance of human T cell memory. Immunity 35, 806–818 (2011).

61. Klass, C. M., Krug, L. T., Pozharskaya, V. P. & Offermann, M. K. The targeting of primary effusion lymphoma cells for apoptosis by inducing lytic replication of human herpesvirus 8 while blocking virus production. Blood 105, 4028–4034 (2005).

62. Scholz, B. A. et al. Abortive lytic reactivation of KSHV in CBF1/CSL deficient human B cell lines. PLoS Pathog 9, e1003336, doi: 10.1371/journal.ppat.1003336 (2013).

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Author Contributions
D.R. and J.M.Z. designed the study, performed experiments, and wrote the paper. All authors analyzed the results and approved the final version of the manuscript.

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