Development, genetic and cytogenetic analyses of genetic sexing strains of the Mexican fruit fly, *Anastrepha ludens* Loew (Diptera: Tephritidae)

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**Abstract**

**Background:** *Anastrepha ludens* is among the pests that have a major impact on México’s economy because it attacks fruits as citrus and mangoes. The Mexican Federal government uses integrated pest management to control *A. ludens* through the Programa Nacional Moscas de la Fruta (National Fruit Fly Program, SAGARPA-SENASICA). One of the main components of this program is the sterile insect technique (SIT), which is used to control field populations of the pest by releasing sterile flies.

**Results:** To increase the efficiency of this technique, we have developed a genetic sexing strain (GSS) in which the sexing mechanism is based on a pupal colour dimorphism (brown-black) and is the result of a reciprocal translocation between the Y chromosome and the autosome bearing the *black pupae* (*bp*) locus. Ten strains producing wild-type (brown pupae) males and mutant (black pupae) females were isolated. Subsequent evaluations for several generations were performed in most of these strains. The translocation strain named *Tapachula-7* showed minimal effect on survival and the best genetic stability of all ten strains. Genetic and cytogenetic analyses were performed using mitotic and polytene chromosomes and we succeeded to characterize the chromosomal structure of this reciprocal translocation and map the autosome breakpoint, despite the fact that the Y chromosome is not visible in polytene nuclei following standard staining.

**Conclusions:** We show that mitotic and polytene chromosomes can be used in cytogenetic analyses towards the development of genetic control methods in this pest species. The present work is the first report of the construction of GSS of *Anastrepha ludens*, with potential use in a future Moscafrut operational program.
Inbreeding the F₁ produced wild-type and mutant phenotypes at a ratio of 3:1 (Table 1). From these results, we conclude that the bp mutation is recessive and autosomal. The correct classification of the colour of the pupae was confirmed based on the pigmentation of the adults. The abdomen, thorax and wings of black pupae adults are darker than the wild-type flies. In addition, the black pupae phenotype can be identified in the larval stage based on the black coloration of their anal lobes (Figure 1).

Development of genetic sexing strains

Ten genetic sexing strains were detected after screening 2,600 single male families, corresponding to a success rate of 0.38%. These strains were maintained in the laboratory for several generations. Females consistently emerged from black pupae, while males emerged from brown pupae (Figure 1).

Each strain had a specific average value of egg hatching. The strains T(Y:bp⁺)-7 T(Y:bp⁺)-3, T(Y:bp⁺)-1, T(Y:bp⁺)-10, T(Y:bp⁺)-6, T(Y:bp⁺)-2, T(Y:bp⁺)-8 and T(Y:bp⁺)-5 had the highest values, and their hatching ranged from 67.91% to 34.41%. The strains with the lowest values, T(Y:bp⁺)-4 and T(Y:bp⁺)-9 showed a hatch rate of 14.4% and 28.81%, respectively (Table 2).

The ten strains analyzed also showed differences in the percentage of recombinant progeny. T(Y:bp⁺)-3, T(Y:bp⁺)-2 and T(Y:bp⁺)-8 had the highest values, with 2.78%, 1.74% and 0.72%, respectively. In contrast, the strains T(Y:bp⁺)-5, T(Y:bp⁺)-6 and T(Y:bp⁺)-7 did not show any recombinants at this level of rearing.

During this first evaluation, the strain T(Y:bp⁺)-7 showed a higher egg hatch as compared to the other GSS (Table 2).

Survival test and genetic stability

The comparative analysis of the survival showed significant differences between the wild-type strain and the GSS T(Y:bp⁺)-7. The primary differences were observed at egg hatching (F = 135.462, 27; P<0.0001) and in larval

| Parent | Cage | Wild-type | Black pupae | X² |
|--------|------|-----------|-------------|----|
| Male   | Female | (no.) | (no.) | (3:1) |
| Wild-type | black pupae | 1 | 2,804 | 952 | 0.24 |
| 2     | 2,343 | 738 | 1.80 |
| 3     | 1,866 | 610 | 0.17 |
| black pupae | Wild-type | 1 | 2,190 | 668 | 4.03 |
| 2     | 2,266 | 759 | 0.13 |
| 3     | 1,761 | 572 | 0.29 |

X² tables = 3.84, P = 0.05, df = 1

Results

Genetic analysis of the black pupae mutation

The two reciprocal crosses, Cbp⁺ × Qbp and Cbp × Qbp⁺, produced wild-type offspring.
survival ($F = 32.372, 27; P < 0.0001$) while the differences in adult emergence were not significant ($F = 1.692, 27; P = 0.202$) (Table 3). A comparison of the egg to adult survival between the wild-type and the translocation strain shows that $T(Y:bp^+)-7$ is roughly 50% sterile (68.5% as compared to 39.3%, Table 3). Based on the results obtained with the medfly [20] this shows that $T(Y:bp^+)-7$ carries a simple, reciprocal Y-autosome translocation, i.e. only one autosome is linked to the Y chromosome.

In the first experiment to determine the genetic stability of $T(Y:bp^+)-7$, the strain was reared with 600 to 2500 parental pairs per generation and no recombinants were observed (Table 4). However, in the second experiment where the population was increased averages of 0.025% recombinant insects were found per generation (Table 5). A third, larger scale experiment (15,000 parental pupae per generation) was set up but this time recombinants were not removed. The highest level of recombinants reached 1.5% in the eighth generation (Table 6).

Cytogenetic analysis
Mitotic chromosomes. *Anastrepha ludens* has six pairs of acrocentric chromosomes, five autosomes and a pair of sex chromosomes (Figure 2a) [21]. The male is heterogametic (XY) [21]. The autosomes display the characteristic somatic pairing observed in Diptera and this is very useful for the identification of homologous chromosomes. The analysis of at least 80 cells from the GSS $T(Y:bp^+)-7$ indicates that the translocation involves the largest autosome (chromosome 2) and the Y chromosome. As it is evident from Figures 2b,c a small distal part of this autosome is joined to the Y chromosome, while a tiny part of the Y chromosome is connected to the autosomal breakpoint. The resulting 2-Y translocation chromosome is slightly shorter (10.58 ± 0.99%) than the wild-type chromosome 2 (11.93 ± 0.8%).

| GSS | Number of generations evaluated | Pupae (no.) | Adults (no.) | Adults (%) | Egg hatch (%) | Recombinants (%) |
|-----|---------------------------------|-------------|--------------|------------|--------------|------------------|
| bp  | bp+                             |            |              |            |              |                  |
|♂♀  |                                |            |              |            |              |                  |
| 1   | 16                              | 13,343     | 15,060       | 15,580    | 82           | 34               |
| 2   | 15                              | 9,031      | 6,712        | 7,914     | 119          | 125              |
| 3   | 8                               | 6,545      | 6,438        | 5,283     | 188          | 96               |
| 4   | 9                               | 2,074      | 1,973        | 1,889     | 1            | 0                |
| 5   | 6                               | 4,641      | 4,172        | 3,817     | 0            | 3,127            |
| 6   | 6                               | 7,362      | 4,742        | 5,893     | 0            | 3,882            |
| 7   | 7                               | 10,765     | 9,843        | 9,592     | 0            | 8004             |
| 8   | 5                               | 2,738      | 2,614        | 2,358     | 0            | 32               |
| 9   | 6                               | 1,367      | 2,059        | 1,171     | 1            | 4                |
| 10  | 5                               | 4,825      | 4,634        | 4,510     | 3            | 17               |

Table 2 Evaluation during multiple generations of ten genetic sexing strains of *Anastrepha ludens*

Figure 1 Larvae, pupae and adults from the GSS Tapachula -7 based on the bp mutation. The males can be identified in the larval stage based on the brown coloration of their anal lobes compared to the black coloration of the females. a) wild-type males, b) black pupae females. Arrows point to the anal lobes.
Table 3 Survival of the wild-type and the *black pupae* strain compared to the genetic sexing strain T(Y:bp)\(^+\)-7 (mean ± SD)

| Strain          | Egg hatched (%) | Larval survival 3rd instar (%) | Larval to pupal survival (%) | Adult emergence (%) | Egg to adult survival (%) |
|-----------------|-----------------|-------------------------------|----------------------------|--------------------|--------------------------|
| Wild type       | 94.60 ± 3.20a   | 74.40 ± 11.73a                | 98.48 ± 1.12a              | 94.72 ± 4.30a       | 68.50 ± 12.00a           |
| bp/bp           | 91.80 ± 4.28a   | 76.20 ± 7.34a                 | 93.94 ± 5.74b              | 91.23 ± 4.44a       | 65.30 ± 8.12a            |
| T(Y:bp)\(^+\)-7 | 63.70 ± 6.00b   | 43.90 ± 5.91b                 | 98.66 ± 2.92a              | 91.02 ± 6.16a       | 39.30 ± 5.25b            |

The same letter after the mean values indicates that the differences between these lines are not significant.

Table 4 Progeny of the strain T(Y:bp)\(^+\)-7 during multiple generations (F) with different numbers of breeders

| F   | No. of pairs bp bp\(^+\) | Total pupae bp bp\(^+\) | Adults bp c\(\delta\) c\(\delta\) partly emerged | Adults bp\(^\ast\) c\(\delta\) c\(\delta\) partly emerged | Total insects | Recombinants (%) |
|-----|--------------------------|-------------------------|-----------------------------------------------|------------------------------------------------|---------------|------------------|
| 14  | 2,500                    | 2,481 2,913             | 5,394                                        | 1,625 c\(\delta\) c\(\delta\) partly emerged 0 | 2,219 c\(\delta\) c\(\delta\) partly emerged 0 | 3,844            |
| 15  | 1,500                    | 3,000 3,000             | 6,000                                        | 2,476 c\(\delta\) c\(\delta\) partly emerged 74 | 2,750 c\(\delta\) c\(\delta\) partly emerged 21 | 5,226            |
| 16  | 1,500                    | 2,600 2,800             | 5,400                                        | 2,059 c\(\delta\) c\(\delta\) partly emerged 85 | 2,492 c\(\delta\) c\(\delta\) partly emerged 19 | 4,551            |
| 17  | 1,500                    | 600 600                 | 1,390                                        | 610 c\(\delta\) c\(\delta\) partly emerged 0  | 605 c\(\delta\) c\(\delta\) partly emerged 0 | 1,215            |
| 18  | 1,500                    | 2,310 2,665             | 4,975                                        | 1,894 c\(\delta\) c\(\delta\) partly emerged 102 | 2,392 c\(\delta\) c\(\delta\) partly emerged 37 | 4,286            |
| 19  | 1,500                    | 2,607 2,659             | 5,266                                        | 2,222 c\(\delta\) c\(\delta\) partly emerged 76 | 2,415 c\(\delta\) c\(\delta\) partly emerged 24 | 4,637            |
| 20  | 1,500                    | 3,000 3,000             | 6,000                                        | 2,507 c\(\delta\) c\(\delta\) partly emerged 106 | 2,809 c\(\delta\) c\(\delta\) partly emerged 34 | 5,316            |

Table 5 Progeny of the strain T(Y:bp)\(^+\)-7 during eight generations of rearing with 3000 females and 2500 males per cage (recombinants were removed each generation)

| F   | Adults bp c\(\delta\) c\(\delta\) partly emerged | Adults bp\(^\ast\) c\(\delta\) c\(\delta\) partly emerged | Total insects | Recombinants (%) |
|-----|-----------------------------------------------|------------------------------------------------|---------------|------------------|
| 3549 | 0 123                                         | 2698 0                                             | 6281          | 0.00             |
| 3455 | 0 130                                         | 2802 1                                             | 6271          | 0.02             |
| 3916 | 0 64                                          | 2698 0                                             | 6635          | 0.00             |
| 3842 | 1 68                                          | 2755 0                                             | 6619          | 0.02             |
| 3365 | 3 89                                          | 2633 0                                             | 6037          | 0.05             |
| 3517 | 2 49                                          | 2600 1                                             | 6139          | 0.05             |
| 3487 | 1 96                                          | 2564 1                                             | 6110          | 0.03             |
| 3227 | 1 53                                          | 2457 1                                             | 5769          | 0.03             |

Table 6 Progeny of the strain T(Y:bp)\(^+\)-7 during ten generations without removing recombinants

| F   | No. of pupae bp bp\(^+\) | Total pupae bp bp\(^+\) | Adults bp c\(\delta\) c\(\delta\) partly emerged | Adults bp\(^\ast\) c\(\delta\) c\(\delta\) partly emerged | Total insects | Recombinants (%) |
|-----|--------------------------|-------------------------|-----------------------------------------------|------------------------------------------------|---------------|------------------|
| P   | 935 1072                 | 2007 709                | 20 c\(\delta\) c\(\delta\) partly emerged 3 | 1009 c\(\delta\) c\(\delta\) partly emerged 9 | 1718          | 0.00             |
| F\(_1\) | 1058 1450              | 2508 1045               | 14 c\(\delta\) c\(\delta\) partly emerged 16 | 1311 c\(\delta\) c\(\delta\) partly emerged 15 | 2356          | 0.00             |
| F\(_3\) | 1299 1348              | 2647 1142               | 16 c\(\delta\) c\(\delta\) partly emerged 16 | 1262 c\(\delta\) c\(\delta\) partly emerged 16 | 2451          | 0.00             |
| F\(_5\) | 1185 1499              | 2684 1026               | 12 c\(\delta\) c\(\delta\) partly emerged 12 | 1410 c\(\delta\) c\(\delta\) partly emerged 12 | 2473          | 0.04             |
| F\(_8\) | 1178 1198              | 2376 1051               | 6 c\(\delta\) c\(\delta\) partly emerged 6 | 1159 c\(\delta\) c\(\delta\) partly emerged 6 | 2230          | 0.00             |
| F\(_10\) | 1135 1304             | 2439 856                | 12 c\(\delta\) c\(\delta\) partly emerged 12 | 1009 c\(\delta\) c\(\delta\) partly emerged 15 | 1912          | 0.00             |
| F\(_12\) | 1294 1405             | 2699 919                | 146 c\(\delta\) c\(\delta\) partly emerged 49 | 1023 c\(\delta\) c\(\delta\) partly emerged 60 | 2216          | 0.00             |
| F\(_14\) | 1134 1408             | 2542 849                | 33 c\(\delta\) c\(\delta\) partly emerged 7 | 1103 c\(\delta\) c\(\delta\) partly emerged 22 | 2023          | 0.00             |
| F\(_8\) | 8000 7000             | 15000 5016               | 260 c\(\delta\) c\(\delta\) partly emerged 116 | 4313 c\(\delta\) c\(\delta\) partly emerged 102 | 9885          | 0.30             |
| F\(_6\) | 834 968              | 1802 606                | 17 c\(\delta\) c\(\delta\) partly emerged 17 | 808 c\(\delta\) c\(\delta\) partly emerged 17 | 1512          | 1.50             |
| F\(_8\) | 831 1188              | 2019 641                | 37 c\(\delta\) c\(\delta\) partly emerged 17 | 1027 c\(\delta\) c\(\delta\) partly emerged 9  | 1743          | 0.57             |
| F\(_10\) | 938 1096             | 2034 713                | 12 c\(\delta\) c\(\delta\) partly emerged 3 | 986 c\(\delta\) c\(\delta\) partly emerged 6  | 1699          | 0.76             |
translocation can be observed. Although the Y chromosome cannot be identified in polytene nuclei using standard staining, we succeeded to identify the autosome involved in the translocation and map the autosomal breakpoint. *Anastrepha ludens* shows a characteristic ectopic pairing between the telomeres of the autosomes [21], a phenomenon that also was observed in other Tephritidae species as *Bactrocera oleae* [22] and *B. cucurbitae* [23].

One of the most frequent ectopic pairing has been observed between the polytene elements III and V (Figure 3f). Polytenes chromosomes from the strain T(Y:bp+)-7 are shown in Figure 3a-e. The ectopic pairing of the two telomeres, chromosome III and V are shown, but interestingly only one homolog of the polytene element III is involved in this. In all cases, a part of a single chromosome III is connected with its telomere to the telomere of chromosome V. This region should represent the distal part of the chromosome element involved in the Y-autosome translocation. In this ectopic pairing either the telomere of the intact chromosome III is taking part (Figure 3a); or the fragment of the chromosome that is connected to the Y chromosome (Y-A) (Figure 3b-e) is pairing. Based on these observations as well as on the mitotic karyotype of this strain, we can conclude that polytene chromosome III corresponds to the

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**Figure 2**

a) *Anastrepha ludens* mitotic metaphase chromosomes from brain ganglia of third instar male larva of the wild-type strain. b-c) Mitotic metaphases of the GSS *Tapachula-7*. d) Graphic representation of the Y-autosome translocation observed in the *Tapachula-7* strain and its comparison with the wild-type. e-f) Mitotic metaphases of the second GSS, *Tapachula -I*. X, Y: represent the sex chromosomes, 2: represents the intact free autosome, Y-2: represents the translocation fragment with the Y centromere, 2-Y: represents the reciprocal translocation fragment carrying the autosomal centromere. Centromeres are indicated by circles in 2d. Bar = 10µm

**Figure 3**

Polytene chromosomes from the strain T(Y:bp+)-7. Thick arrows show the ectopic pairing between the telomeres of the V and III chromosomes. a) Thin arrow shows the autosome breakpoint, in this case the intact chromosome III is taken part in the ectopic pairing. b-e) The autosome fragment of the Y-A chromosome is taken part in the ectopic pairing. f) Normal ectopic pairing between chromosomes V and III from the wild-type strain. Bar = 10µm
longest chromosome of the mitotic karyotype, i.e. chromosome 2. Taking into account both the mitotic karyotype and polytene chromosomes analyses of this GSS, we can propose the structure of the T(Ybp)-7. According to the chromosome maps of A. ludens [21] the chromosome III has a breakpoint at the beginning of the region 25 (Figure 4). This breakpoint produces two autosomal fragments of different size: The short fragment includes the distal part of the chromosome from the telomere to 25 chromosomal region (24.67 μm) while the second fragment consisting of the rest of the chromosome element III. The short fragment is joined to that part of the Y chromosome that carries the centromere Y-2 (Figure 2d). The second autosomal fragment, with the autosomal centromere, is linked to remaining region of the Y chromosome resulting in the 2-Y chromosome (Figure 2d) that is slightly shorter than the wild-type chromosome element III. Each of these new chromosomes, 2-Y and Y-2 has its centromere and telomere sufficient for their stability.

Discussion

We have developed a GSS for Anastrepha ludens using the black pupae mutation as a selectable marker. In this strain, males emerge from wild-type (brown) pupae, whereas females emerge from mutant (black) pupae. This particular mutation has additional benefits because it is also expressed in the larval and the adult stage. This characteristic confers various benefits to this system, including greater reliability, avoiding the difficulties in the clear discrimination of the colours as it was reported by Rössler [24] for the genetic sexing system based on the marker dark pupae in Ceratitis capitata. Undoubtedly, the genetic marker white pupae found in other fruit flies provide better separation due to the greater contrast with the wild-type pupae [25-30].

However, despite significant efforts, this marker has not been found yet in Anastrepha ludens, neither in field populations nor in inbred laboratory colonies.

During this study, different doses of irradiation were used: 25 Gy, 30 Gy and 40 Gy. The first dose was too low, and no translocations were obtained. The highest dose induced such a high sterility that it was impossible to breed the insects. The optimal dose was 30 Gy, and the success rate was 0.38% (10/2600). The doses used to induce translocations were lower than those used in Ceratitis capitata. Franz et al., (1994) [31] reported a success rate of 7.1% at 50 Gy, and Kerremans and Franz, (1995) [17] reported a success rate of 4.3% at 40 Gy. These differences between C. capitata and A. ludens indicate that it is necessary to determine the optimal dose for each species individually. The optimal dose can be influenced by specific biological characteristics, such as the size of the Y chromosome. The Y chromosome of C. capitata is significantly longer than that of A. ludens and this might be responsible for a greater probability for inducing Y-autosome translocation compared to A. ludens.

Genetic stability is one of the most important aspects in the evaluation of a GSS and its usefulness for a practical application. Genetic stability depends directly on the structure of the translocation, i.e. it depends on the distance between the autosomal breakpoint of the translocation and the location of the genetic marker [31]. This distance is directly proportional to the frequency of recombination in this chromosomal region in the heterozygotic males of a sexing strain [13]. The recombinant flies show a reverse phenotype as compared to the normal, non-recombinant flies, i.e. recombination in the parental males produces black pupae males and brown pupae females. The percentage of recombinant flies during various generations was high in the strains T(Ybp)-2, T(Ybp)-3 and T(Ybp)-8. These strains were considered unstable. On the contrary, the strain T(Ybp)-7 showed no recombinants when reared at a smaller scale. However, the strain showed 0.025% of recombinants per generation when the scale of rearing was increased for several generations. Monitoring a larger number of insects over a longer period of time will provide additional results to predict the behaviour of this strain under mass-rearing conditions.

In the evaluation of a GSS, it is important to consider the productivity and the product quality, both of which are tightly linked to the structure of the translocation. The segregation behaviour of the translocation during male meiosis will determine the number of genetically balanced or unbalanced individuals in the next generation. During male meiosis simple Y-autosome translocations can segregate in two different ways if they follow the rule that homologous centromeres segregate: alternate or adjacent-1 segregation. In the former the two translocation chromosomes stay together and segregate from the X chromosome. In the latter the A-Y chromosome segregates together with the X chromosome while the Y-A chromosome segregates with the non-translocated autosomal homologue. Figure 5

![Figure 4 a](image4a.png) The short translocation fragment representing the Y-2 chromosome of mitotic metaphases or Y-III of polytene chromosomes. b) Reference map of chromosome III showing the region involved in translocation; arrow in the map shows the position of the breakpoint. Bar = 10μm.
shows, that after fertilization only alternate segregation results in genetically balanced offspring while adjacent-1 segregation results in offspring where the chromosomal segment between the translocation breakpoint and the tip of respective chromosome arm is either present in three copies or segregation only in a haploid condition. At least in the medfly such long deletions lead to early lethality which is manifested as reduced egg hatch. The triplication type adjacent-1 offspring can survive, depending on the length of the triplication, even to the adult stage, although at severely reduced numbers [12]. Only if the autosome breakpoint is very close to the tip, i.e. the triplication is very short, such flies mate and produce offspring (Franz, personal communication). Both forms of segregation are completely legitimate and that explains why they occur at equal frequency in most translocation strains, at least in medfly. Consequently, only half of the sperm produced by males carrying a Y-autosome translocation lead to viable, genetically balanced offspring, i.e. such males are 50% sterile. In a third form of segregation, adjacent-2, homologous centromeres do not segregate, i.e. this is a form of non-disjunction and therefore significantly less likely to occur.

The egg hatch in strain T(Y:bp)-7 is reduced due to the lethality of the deletion type adjacent-1 offspring. In addition, the triplication carrying adjacent-1 individuals should ideally die as early as possible to avoid wasting larval diet and to avoid confusions in the quality control. In the T(Y:bp)-7 strain, this seems to be the case as larval survival is the only parameter besides egg hatch that differs significantly from the wild-type level.

The cytogenetic studies performed using mitotic chromosomes showed that T(Y:bp)-7 carries a simple reciprocal translocation between the longest autosome, chromosome 2, and the Y chromosome. Analysis of polytene chromosomes permitted us to identify the polytene element involved in the translocation. Based on the specific ectopic pairing observed between the telomeres of autosomes in the A. ludens polytene nuclei [21], we show that chromosome 2 of the mitotic karyotype corresponds to polytene element III (Figure 6). In addition, this analysis permitted us to map the autosomal breakpoint on chromosome III despite the fact that the Y chromosome remains under-replicated in polytene nuclei and cannot be identified after standard staining. The autosomal breakpoint is close to the beginning of region 25 of chromosome III according to the polytene maps of the A. ludens (Figure 4). It should be emphasized that both translocation chromosomes possess centromeres and telomeres, required for stability during cell divisions.

Ideally, a genetic sexing system allows the elimination of females during the early stages of development to decrease the cost during mass-rearing effectively and to increase the capacity of the production facility. However, the method of sex separation we report here is based on the coloration of the pupae and allows sexing only at the pupal stage. This is done mechanically using sorting machines available on the market. These machines can be adapted for this specific use by modifying their sorting characteristics so that brown and black pupae can be separated with the required speed and accuracy. In 1969, Whitten [32] suggested this type of mechanized separation for GSS based on the colour of pupae to obtain sufficient quantities to meet the demand of an area-wide control program. The advantage of this method of separation is that it is non-destructive, allowing to feed the females back into the rearing colony and,
thereby, increasing production capacity if required [33]. Using such a seed sorting machine, Robinson and Riva [34] separated white and brown pupae of a Mediterranean fruit fly GSS with a success rate of 100%.

Conclusions
We have isolated the first GSS in *A. ludens*, which was given the common name *Tapachula-7*. Its characteristics, as far as we have determined them in the research described here, indicate that this strain will be stable and productive enough for a practical application in the mass-rearing for field operations. However, further testing under mass-rearing conditions is required to come to a final conclusion. In addition, the results of the current study show that mitotic and polytene chromosomes of this species are suitable for cytogenetic studies and could support the development of improved control methods in this pest.

Materials and methods

Biological material
The colony of *Anastrepha ludens* used in this study was founded using flies collected in the wild in the state of Chiapas, México [35]. The insects were then kept in the laboratory for several generations with photoperiods of 12:12, a temperature of 26°C and a relative humidity of 70-80%.

**black pupae strain**
Pupae with black colour, distinct from the brown-coloured wild-type pupae, were isolated from the mass-rearing facility Moscafuit located in Metapa de Domínguez, Chiapas, México after screening approximately 7,700,000. Larval anal lobes were black and had noticeably darker cuticles and wing lines than the wild-type individuals. Larval anal lobes were black and easy to be differentiated from the wild-type [36].

Rearing the flies
Adults are maintained in laboratory conditions at 26°C and a relative humidity of 60-70% with *ad libitum* water and a mixture of sugar and hydrolyzed proteins (yeast hydrolysate enzymatic MP Biomedicals, LLC France). Eggs are collected and incubated in Petri dishes with saturated humidity for 4 days at 26°C in a bioclimatic chamber Binder KBF720. The larvae are fed with diet of powdered corncob (19% powdered corncob, 9.2% sugar, 5.3% corn flour, 7.0% yeast, 0.44% citric acid, 0.4% sodium benzoate, 0.2% nipagin and 0.1% guar gum) and are maintained under controlled temperature and humidity. Pupation is carried out over 14 days on Vermiculite Strong Lite® substrate.

Genetic analysis
The inheritance of the *black pupae* mutation was determined by crossing wild-type males to *black pupae* females and *black pupae* males to wild-type females (♂wt to ♀hp, ♀hp to ♀wt). Three cages were set up for each cross. Eggs were collected from each cage, and the breeding process was continued until the emergence of the F1 generation progeny. Five females and 5 males of the F1 generation were then inbred to obtain the F2 generation. F1 and F2 pupae were carefully checked and separated by colour, and the adults were observed under a Zeiss stereomicroscope to confirm their phenotype.

Development of genetic sexing strain
Wild-type pupae two days before emergence were irradiated with 30 Gray (Gy) using a cobalt-60 Gammacell 220 source. The irradiated males were then placed in 4 litre cages and crossed with virgin females homozygous for *black pupae (bp/bp)*.

The F1 males of that cross were singly backcrossed to mutant females. If the F1 males carry a translocation between the Y and the autosome bearing the selectable marker “bp”, all the males of a particular family will be wild-type, and all the females will be mutant (Figure 7). Where such a family was detected in the screen, the progeny was further propagated, and their true-breeding nature was ascertained and maintained in the laboratory for several generations. In each generation, the percentage of the hatched eggs, the number of pupae and their colour as well as the number of adults were recorded. When recombinants, i.e. females emerging from brown and males from black pupae, were detected they were counted and removed. The pupal colour separation was performed manually, and the correct classification was corroborated in adults using the dark coloration of the *black pupae* strain for differentiation.

Survival test and genetic stability
The following three types of crosses were set up: a) flies from the *black pupae* strain (100♀ and 100♂) b) wild-type (50♀ and 50♂) and c) the GSS *Tapachula-7* (100♀ and 100♂). Each one was placed separately in cages of 30x30x30 cm. When the insects reached sexual maturity, 1000 eggs were taken in batches of 100. The eggs were incubated for 4 days at 26 ± 1°C in a bioclimatic KBF720 Binder chamber, and the number of fertile eggs was quantified after 7 days. The larval development of 100 eggs was conducted in a Petri dish with the specified larval diet. The number of third-instar larvae was quantified after 10 days. The number of mature pupae and emerged adults was also recorded. The T(Y:bp)-7 was reared for some generations increasing the number of parents. Three experiments were carried out: the first was using a population in the cage of 600-2500 adults, evaluated during seven generations and in each generation the recombinants were removed and the pupae yielded were recorded. In the
second experiment the population used was 3000 females × 2500 males and recombinants were monitored during eight generations, removing the recombinants in each generation. In the third experiment we monitored ten generations of rearing with 15,000 parental pupae (mixed black and brown) per generation in large cages (the recombinants were not removed during this experiment). To determine the number of recombinants in each generation, a sample of 100 ml of pupae was analyzed in each generation.

Cytogenetic analysis
Mitotic and polytene chromosomes were prepared from the genetic sexing strain T(Y:bp<sup>+</sup>)-7. Metaphase chromosomes were prepared from the brain ganglia of third-instar larvae [20] while for the staining the application of the C-band technique was used [37].

The slides were immersed in hydrochloric acid 0.2 N for 90 s, washed with distilled water and immersed in 5% barium hydroxide at 40°C for 1 min. The slides were then washed in distilled water with a few drops of acetic acid and placed in 2xSSC at 60°C for 30 min. Finally, the slides were washed and stained using 25% Wright stain in Sörensen buffer for 1 min and 12% Giemsa stain for 1 hour. The samples were observed using a Zeiss Axioskop 40 microscope with 100x magnification objective, and images were acquired using the software Axiovision 4.8.2 by Carl Zeiss de México S.A. de C.V. The length of the chromosomes in 35 metaphases from the wild-type samples and 15 from the genetic sexing strain were measured with UTHSCSA Image tool 3.0 (web site http://compdent.uthscsa.edu/dig/dtdesc.html). Only well metaphase spreads with 12 chromosomes were used to measure the total length of each chromosome. The relative chromosomal length was estimated as the percentage of the length of each chromosome relative to the total length of the total diploid complement (%RL). Third instar male larvae were used for the salivary gland polytene chromosomes using the method described for C. capitata [18,38] and previously used for A. ludens by García-Martínez et al. [21]. The male larvae were identified based on the brown coloration of the anal lobes compared to the black coloration of the female larvae.

Data analysis
The overall fitness was estimated by multiplying the transformation frequency between the stages (proportion of eggs hatched, larvae survival, pupae survival and pupae to adult survival). The data were transformed by arcsine √x and analyzed by ANOVA. Medians were compared using Tukey’s test with α = 0.5. Both of these analyses were performed using the software JMP version 5.0 (web site http://www.jmp.com).

Competing interests
The authors declare that they have no competing interests.
Authors' contributions

CSZC directed the study, designed the protocol, selected the data to present, integrated and composed the manuscript and collaborated with VGMI to implement novel technical modifications for the study of mitotic chromosomes.

JSMH designed protocols for the construction and testing of the genetic sexing strains, integrated and analyzed the data, performed the statistical analysis and help to initiate the initial draft of the manuscript.

VGMI conducted the laboratory cytogenetic methods, including slide preparation and imaging, discussed the results of the cytogenetic analysis and revised the manuscript.

JIP coordinated the laboratory work for establishing protocols and evaluating the genetic sexing strains.

AZ participated in the interpretation of the cytogenetic analysis data and reviewing the manuscript.

GF designed the protocols for constructing the genetic sexing strains and reviewing the manuscript.

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