HIV can be partially contained by host immunity and understanding the basis of this may inform vaccine design. The importance of B-cell function in long-term control is poorly understood. One method of investigating this is in vivo cellular depletion. In this study, we take advantage of a unique opportunity to investigate the role of B cells in an HIV-infected patient. The HIV-1+ patient studied here was not taking antiretroviral drugs and was treated for pre-existing low-grade lymphoplasmacytoid lymphoma by depletion of CD20+ B cells using rituximab. We demonstrate that B-cell depletion results in a decline in autologous neutralizing antibody (nAb) responses and a 1.7 log_{10} rise in HIV-1 plasma viral load (pVL). The recovery of NAbs results in a decline in pVL. The HIV-1 sequences diversify and NAb-resistant mutants are subsequently selected. These data suggest that B-cell function can contribute to the long-term control of pVL, and that NAbs may be more important in controlling chronic HIV-1 infection than previously suspected.
**Results**

**Clinical course of the study subject.** The patient, a Caucasian man, was diagnosed with HIV seroconversion illness at the age of 58 years. Three years earlier, he was diagnosed with low-grade lymphoplasmacytoid lymphoma, for which he did not receive treatment. Histologically, this was defined as a small lymphocytic B-cell lymphoma with plasmacytoid differentiation mostly expressing CD20 and CD79. The tumour was IgM secreting. His only other medical history of note was an acute hepatitis B infection approximately 30 years earlier. Following HIV-1 seroconversion, the patient was recruited into a prospective, non-randomized observational study of early treatment and received 3 months of highly active antiretroviral therapy. After 30 months, he developed a rising paraproteinemia, attributed to his lymphoma, and received thalidomide treatment without clinical effect. He initiated rituximab therapy 1,075 days after HIV seroconversion and received four weekly doses (375 mg). He felt unwell 20 weeks after the first dose of rituximab with malaise and fever associated with a marked rise in HIV pVL (increase of 1.7 log₁₀; Fig. 1). In addition, he developed viraemia, in the absence of antiretroviral therapy (ART).

**Figure 1 | Autologous neutralizing antibody titres over time.** (a) Pseudotyped virus particles were constructed using full-length envelope clones that represent majority quasi-species at 336 or 1,042 days after seroconversion. Plasma VL over time is indicated (red line). The brief dip in viral load over the first 100 days represents the period of initial ART as part of the short-course ART trial (shaded grey). The neutralizing effects (y-axis) of patient sera are shown for autologous viruses constructed with cloned env genes derived 336 days (black squares) and 1,042 days (crosses) after seroconversion. Results from a single clone (tested in triplicate) representative of three clones are shown. Serum was sampled sequentially before and after rituximab therapy (1,075–1,106 days, shaded grey), and assayed at concentrations ranging from 1:20 to 1:4,370; the neutralizing effects of sera diluted 1:80 are shown here. Error bars show standard deviations of triplicate tests. (b) CD4 T lymphocyte count (dashed line) is shown over time.
biochemical hepatitis (attributed to reactivation of hepatitis B) that resolved with only supportive treatment. Antiretrovirals were initiated 1,392 days after seroconversion.

**Impact of B-cell depletion on NAB titres.** Following rituximab therapy, the subject's HIV-1 pVL, previously stable, increased by more than 1.7 log10, peaking at 737,400 copies per ml 4 months after the final dose of rituximab (Fig. 1). The pVL subsequently returned to baseline levels in the absence of ART. We therefore analysed the impact of rituximab therapy on NAB titres in relation to viral dynamics.

Rituximab therapy led to a 30% reduction in total IgG levels, which reached their lowest peak at the point of virus load (10.8 g l−1; normal range 8–16 g l−1) and returned to pre-rituximab levels thereafter (Supplementary Table S1). To evaluate NAB activity throughout this period, we generated pseudoviruses derived from autologous envelope (env) sequences. Serum obtained before the first dose of rituximab potently neutralized virus pseudotyped with autologous envelope derived from plasma acquired 2 years previously (day 336 after seroconversion) (Table 1). The same serum neutralized contemporaneous autologous Env pseudotypes, although to a lesser extent, with 1:60 serum dilutions effectively blocking more than 90% infection. Heterologous NAB activity was also observed against HXB2 and 8/12 Tier 2/3 clade-B reference strains (NIH, AIDS) (Table 2). Following rituximab therapy, we observed a marked decrease in neutralizing activity characterized by a drop of more than threefold in IC50 serum titres against day 336 Env pseudotypes, which recovered over a 4-month period. Strikingly, these changes coincided with the increase in pVL. Recovery of NAB responses correlated with control of viraemia (Fig. 1).

**Analysis of sequence evolution in HIV-1 envelope over time.** To evaluate sequence evolution over this period, env products were generated by reverse transcription (RT)–PCR from plasma viral RNA, before and after rituximab therapy (days 1,075–1,392; Fig. 2a). We identified clear sequence diversification as the virus peaked during rituximab treatment (mean genetic distance = 0.02 versus 0.004; P < 0.0001). This diversification manifested as an increase in effective population size as measured using BEAST77 (fivefold change; Fig. 2b). To assess whether these diverse strains represented a re-emergence of archived provirus, we used an identical sequencing strategy to analyse stored peripheral blood mononuclear cell (PBMC) samples available from the time of seroconversion up to day 1,042; these sequences were placed on a time-structured tree (Supplementary Fig. S1). Viruses detected after rituximab treatment did not represent re-emergence of early archived strains; those strains present as pVL peaked were most closely related to strains present in plasma at day 1,042, with a subset related to those archived in PBMC at day 1,042.

**Immune selection and neutralization of HIV-1 envelope.** The apparent in vivo impact of the NABs in this individual prompted a search for the targets of binding. To identify potential NAB targets, we first analysed immune selection using CODEML, analysing the ratio of non-synonymous to synonymous mutations (dN/dS) at each site. We observed two sites under selection (Fig. 3; Supplementary Fig. S1)—position 339 (dN/dS = 7.6, P < 0.0001) and position 363 (dN/dS = 6.977, P = 0.003). Residue 363 lies between the β14 and β15 anti-parallel strands of gp120 and is exposed on the surface of the outer domain in close proximity to the CD4 binding loop80. Computational modelling of atomic level envelope structures has shown that mutations proximal to the CD4 binding loop are critical in conferring resistance to CD4bs antibodies89. Furthermore, site-directed mutagenesis of site 363 has previously been shown to affect neutralization of the CD4bs antibodies M-14 and b12100,101. Before the first dose of rituximab (day 1,042 after seroconversion), position 363 was occupied by an arginine in all viral RNA sequences, whereas all proviral DNA clones sampled up to and including day 1,042 had a glutamine at the same position. Following rituximab therapy, R363 variants were temporarily replaced with Q363 variants, which reverted to arginine residues as NAB titres rebounded and viraemia was controlled.

Before rituximab therapy, an asparagine at site 339 formed part of an N-linked glycosylation site. This residue is located within the α3 helix, which is exposed on the surface of the protein according to atomic level structures. A previous study showed that site 339 is involved in recognition by 2G12, a monoclonal antibody the binding of which is carbohydrate dependent101. However, in addition to this, mutations at site 339 have been reported to affect binding by monoclonal antibody b12: this effect is observed despite site 339 being some distance away from the CD4 binding site, and potentially occurs through conformational perturbation80. No other sites in the envelope were found to be under positive selection, indicating that mutations at these positions affected NAB binding.

To examine the effect of sequence changes on neutralization susceptibility, pseudoviruses mimicking Q363R and R339N/E were constructed by site-directed mutagenesis. The selected Q363/339E mutation reduced the IC50 titre of day 1,042 serum by more than twofold (IC50 from > 775 to 300). Confirming previous data, the Q363R mutation was also shown to inhibit neutralization by fivefold by antibody b12101, whereas N339E inhibited neutralization susceptibility by fourfold to the carbohydrate-specific antibody 2G12102,103. To further define the site of binding of the patient’s NABs, we performed a direct competition experiment using well-defined monoclonal antibodies and recombinant clade B envelope as the antigenic target (Fig. 4). This experiment showed strong binding competition between patient sera and antibodies b12, 4,8d (which targets an epitope overlapping with the co-receptor binding site) and 447-52D (which targets the V3 loop). No competition was observed against 2G12. The experiment also revealed a clear decline in competition for binding in samples taken during the peak of viraemia, which recovered as pVL declined, in good accordance with the data obtained from the neutralization experiments, but using an independent system of evaluation (Fig. 4b).

**Table 1 | IC50 neutralizing titres against autologous viruses.**

| Serum (day) | D336 pseudovirus | D1042 pseudovirus | HXB2 | VSV-G |
|-------------|------------------|------------------|------|------|
| 1,042       | > 775            | 150              | 850  | < 20 |
| 1,135       | > 775            | 225              | 1000 | N/T  |
| 1,239       | 220              | < 120*            | 875  | 20   |
| 1,295       | 250              | 150              | 325  | 20   |
| 1,308       | 360              | 375              | 1000 | < 20 |
| 1,353       | 600              | 425              | 3950 | N/T  |
| Negative serum | < 20            | < 20             | < 20  | N/T  |

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Table 1 | IC50 neutralizing titres against autologous viruses.

Abbreviations: N/T, virus–serum combinations that were not tested because of limited availability of sera; VSV-G, vesicular stomatitis virus control.

The mean half maximal inhibitory concentrations of sera (IC50, reciprocal dilutions) are given for three viruses pseudotyped with representative Env clones derived from d336 and d1042 plasma (autologous).

*IC50 was the highest concentration of day 1,239 serum that could be tested against day 1,042 virus due to limited serum availability.
Table 2 | IC₅₀ neutralizing titres against heterologous viruses.

| Serum (day) | HXB2 | 6535 | QH0692 | PVO | TRO | AC10.0 | pREJO 4541 | pTRJO 4551 | pRHPA 4259 |
|-------------|------|------|--------|-----|-----|--------|------------|------------|------------|
| 1,042       | 850  | 220  | 220    | 90  | 120 | 90     | 140        | 50         | 50         |
| 1,353       | 3950 | 300  | 170    | 90  | 130 | 160    | 220        | 140        | 120        |
| Negative    | < 20 | 20*  | < 20   | < 20| < 20| < 20   | < 20       | < 20       | < 20       |

The mean half-maximal inhibitory concentrations of sera (IC₅₀, reciprocal dilutions) are given for pseudoviruses derived from a standardized reference panel (NIAID, NIH) (heterologous). Reductions in infected TZM-bl cells by pooled HIV-negative sera (*) were attributed to cellular toxicity which was not observed at concentrations below 1:20.

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**Figure 2** | Diversification of HIV-1 env sequences after rituximab therapy. (a) A time-structured tree, showing divergence-time estimates among intra-host plasma HIV-1 clonal sequences before and after rituximab therapy (shaded grey), based on a Bayesian relaxed molecular clock applied to 969 nucleotides. The external nodes represent sequences taken from time points indicated by the colours given in the figure key (inset). The topology is of the highest likelihood calculated using BEAST, with branch lengths proportional to time estimates for the subtending nodes (summarizing 60 million Markov chain Monte Carlo (MCMC) simulations, generated after discarding 6 million as burn-in). All the parameters in the MCMC started from random values, except for substitution rates and population size, where the priors were imported from the combined proviral and plasma viral sequence data of the same host calculated by BEAST software. (b) The effective population sizes (Nₑ) were calculated using BEAST. The x axis (days after seroconversion) is the same as for Figure 2a.

**Figure 3** | Tracking of selected envelope mutants. This figure shows the relative frequencies of polymorphisms at sites (a) 339, dN/dS = 7.61, P < 0.00001 and (b) 363 dN/dS = 6.98, P < 0.031. The individual bars along the x axis represent the different time points of sequencing (comparable to Fig. 2; Supplementary Fig. S1): these are taken either from proviral DNA samples (days 20–1,042) or from plasma samples (days 1,042–1,392). The time of rituximab therapy is indicated. Each bar shows the frequency of mutations at these positions, the key depicts the different amino-acid residues detected.

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**Analysis of other potential effects of rituximab.** We investigated other possible causes for the observations made, particularly the possibility of an indirect impact on cellular immunity. We analysed the sequence of key CTL epitopes over time, and carried out interferon-γ (IFN-γ) ELISPOT analyses. Although some mutations in pre-rituximab sequences were observed, the majority of the epitopes remained stable (Table 3). It is of importance that control over pVL was regained without further escape/reversion at these sites.

Second, immune activation through recrudescence of other infectious agents may have a role in this case. Concurrent with the pVL increase, the patient also developed elevated liver transaminases, associated with reactivation of previously controlled hepatitis B virus (HBV) infection. However, control of HIV was observed at a time when HBV load was still high, suggesting that HBV replication per se was not driving HIV replication (up to day 1,389; Supplementary Fig. S2) in this patient. In the setting of acute illness, there was no evidence of EBV, HHV-6, HHV-8 or HCV in blood. Cytomegalovirus was present only at low copy number (50 copies per ml). A bronchoalveolar lavage was tested for a broad panel of relevant viruses, all of which were negative, including VZV and HSV1/2. Clearly, these data do not exclude reactivation of a further latent infection, or concurrent superinfection, although the major herpesvirus infections appear unlikely to be contributing significantly.

**Discussion**

In this study, B-cell depletion led to a decline in NAb titres against HIV, concurrent with rising pVL. Recovery of NAb titres coincided with control over viral load. Multiple analytical techniques were used to longitudinally characterize the distinct virus population that emerged as NAbs returned. The common ancestor of this new virus population was defined by polymorphisms at two residues exposed on the surface of the virus envelope, both of which showed evidence of positive selection, with mutations affecting binding of NAbs.

There are no published studies on the impact of rituximab alone on HIV viraemia in humans. Studies on acute SIVmac infection in macaques treated with rituximab indicate a role for NAb responses in containing post-acute viral load. In contrast, a recent study...
using African Green monkeys did not reveal any effect of rituximab treatment during acute SIV<sub>pm</sub> infection; thus, data from human studies are important to resolve this issue.

It is possible that depletion of B cells had an indirect impact on viral load through modulation of CD4+ or CD8+ T-cell subsets. Although we cannot exclude this, the mechanism for such an indirect effect is not clear and, importantly, no impact on CD8+ T-cell responses has been observed in the SIV models quoted above. The delay of several weeks that is observed between rituximab treatment and the change in viral load is much more consistent with the observed decline in antibody titres than a direct B-cell/T-cell interaction, which would be expected to take effect immediately. In addition, the changes in the prevailing sequence (reversion and subsequent re-selection of a mutated NAb epitope; Figs 2, 3; Supplementary Fig. S1) are consistent with relaxation and re-application of NAb-mediated selection pressure.

The impact of other reactivating viruses on HIV load is complex. As in this case, rituximab treatment is known to have the potential to reactivate HBV<sup>44,45</sup>, even in those with previously controlled infection (HbsAg −, HBcoreAb +). This points to an important role for B cells in long-term control of HBV—a virus in which CD8+ T cells have an important role in acute disease<sup>46</sup>. The relationship between HBV pVL and HIV pVL is not fully understood, but overall these appear to be independent<sup>44,45</sup>. In this case, HBV and HIV pVL increased within a similar timeframe; however, control over HIV occurred spontaneously, whereas HBV load remained high, indicating that they were differentially regulated in this patient (Supplementary Fig. S2). However, this does not exclude a secondary impact of HBV through induction of liver inflammation or other consequences of immune activation. In addition, although we did not find other evidence of systemic reactivation of other major pathogens in this individual, it remains possible that local or low-level reactivation could also affect HIV pVL.

Overall, our findings support previous suggestions of an evolutionary balance between virus and NAbs<sup>12,44</sup>. At first sight, these findings appear at odds with previous studies that fail to confirm the association between long-term virus control and NAb response, especially in elite controllers<sup>50</sup>. However, as shown here, the dynamics of immune responses are critical to their interpretation; therefore complex relationships could easily be missed in cross-sectional studies, especially analyses of those with favourable CD8+ T-cell responses<sup>43,44</sup>.

There is a clear relationship between the changes in NAb titre and pVL demonstrated in this study, although the mechanism of action of such NAb in vivo may be complex. Studies using engineered antibodies in which Fc binding was disrupted have shown that interactions of NAbs with infected cells, leading to engagement of effector cells, may provide an important component of the protective capacity in vivo<sup>45,46</sup>. It is also possible that other non-NAb specificities not defined using in vitro NAb assays might have a role in vivo<sup>47</sup>. It is noteworthy that as B-cell responses to HIV envelope

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**Figure 4 | Characterization of patient antibodies by competition ELISA.** (a) Serial dilutions of patient serum (x axes) collected on days 1,042 (filled diamonds), 1,270 (filled circles), 1,295 (filled triangles) and 1,353 (filled squares) after seroconversion reduced gp120 binding with NAb b12, 447-52D and 4.8d, thereby reducing colorimetric change and light absorption (A<sub>450</sub>) (y axes). Pooled serum collected from HIV-uninfected subjects (stars) did not compete with any NAb for gp120 binding. All sera failed to compete with 2G12 for gp120 binding. (b) The reciprocal dilutions of sera, sampled before and after rituximab therapy, which reduce gp120-NAb binding by 50% (IC<sub>50</sub>, right y axis). Hundred percent binding was taken as that observed in microtiter wells containing lgGb12 (empty squares), 4.8d (empty triangles) or 447D (empty circles) in the absence of patient sera. Corresponding results are shown by the red line (left y axis).

**Table 3 | Analysis of CD8+ T cell responses and epitope evolution over time.**

| Epitope | Autologous sequence | Response pre-rituximab Year 1 | Response pre-rituximab Year 3 | Mutation pre-rituximab | Mutation post-rituximab |
|---------|---------------------|-----------------------------|-----------------------------|------------------------|-------------------------|
| A11 gag TK8 | TLYCVHQK | 1,200 | — | K8N | Q7A |
| A11 pol AK11 | ACQGVGGPGDHK | 300 | — | — | G9A |
| A11 nef QK10 | QVPRLPMTYK | — | — | — | — |
| A11 pol AK9 | AIFQCSMTK | — | — | — | — |
| B8 gag p17 GK9 | GSSGYKQLK | 160 | 200 | — | — |
| B8 gag p17 EV9 | EKSLYNTV | 640 | 600 | — | — |
| B8 gag p24 DI8 | DICYKRWII | — | — | K3G/K6D | K3R |
| B8 gag p24 DL9 | DCKTLKAL | — | — | KSQ | KSQ |
| B8 nef WM9 | WPKVYRERM | — | — | N3T (242)* | N3T (242)* |
| B8 nef FL8 | FLKEGGGL | — | — | — | — |
| B57 gag TW10* | TSNLEQIEGW* | — | — | — | — |

CD8+ T cell responses were analysed using ex vivo IFN-γ ELISpot analysis (spot forming units/ml million PBMCs are shown). The HLA Class Ia and Class II genotypes are A*01, A*01 and B*0801, B*0801. Sequencing of the corresponding peptides was carried out at the time points shown, and mutating residues have been underlined.

* This epitope is a B57 escape mutant, transmitted to, but not recognized by the recipient, and which shows reversion pre-rituximab therapy.
may be poorly sustained, it has been proposed that long-term plasma cell pools may not be established (reviewed in refs 48, 49). As plasma cells are spared the effects of rituximab36 (and total IgG levels therefore remain relatively less affected), it is suggested that the antiviral NAb identified here, and potentially elsewhere, are maintained by B-cell populations that express CD20.

In conclusion, this unique study suggests that B cells, and their secreted NAb, can affect HIV viral load in chronic infection. This evidence, derived directly from observations in man, may inform the rational design of future immunotherapies and HIV vaccines.

Methods

Construction of HIV-1 env expression cassettes. The study was approved according to local ethical review (IRB Number 99/IA/161E). Full-length virus env was PCR amplified from cDNA obtained by RT of plasma-derived viral RNA collected 336 and 1,042 days after seroconversion for construction of pre-rituximab Env pseudotyped virus. The latter time point represented virus present around the time of rituximab treatment. The primers used were env1A (outer sense, 5′-CACCAG-GCTTACGCACTCATCTGGAGGAA3′), envM1 (outer antisense and RT, 5′-TAGGCTTTTCAATCGCTCCCTTCTTTTA-3′)35, envF2 (inner sense, 5′-GGGCTGAGACCGGTGAGCAGAAGACAGTGGCAATG(AG+3′)-3′) and envR1/2 (inner antisense, 5′-AAATCTGAGGGGGCCCTTGGCACCATRTTAT(−TAGC)-3′). Bulk PCR products were restriction-cloned into pcDNA1.1 (Invitrogen) using sites ApaI and Xhol to result in pcDNA1.1 env, followed by sequencing. Of 20 genetically distinct clones that were most representative of the consensus sequence, 3 were selected for neutralization experiments.

Site-directed mutagenesis. Where stated, 100 ng pcDNA1.1 env was used as a template for PCR amplification using PuFlural UltraII HS (Stratagene) in the supplied buffer supplemented with 250 µM dNTPs and 0.1 unit of the corresponding restriction enzyme. Mutant plasmids were identified by restriction analysis and confirmed by nucleotide sequencing. Double mutants were constructed by sequential PCR amplifications.

Construction and neutralization of Env-pseudotyped virus particles. Autologous env clones and a heterologous panel of subtype-B reference strains were amplified from cDNA obtained by RT of plasma-derived viral RNA collected 18h and, after extensive washing with PBS, a biotinylated secondary mouse anti-human IFN-γ monoclonal antibody (0.05 µg ml−1; Mabtech) was added. The plates were developed using an alkaline phosphate-conjugating substrate AP colour reagent A + B (BioRad). After 15 min, the colorimetric reaction was stopped by washing with tap water. Plates were air dried and spots were counted using an automated ELISpot reader (AID ELISpot reader System). IFN-γ-producing T cells were expressed as spot-forming units per 1×106 cells. The number of specific IFN-γ-secreting cells was calculated by subtracting the unstimulated control value from the stimulated sample.

Sequence evolution and bioinformatics software. For sequence evaluation in evolutionary studies, PCR amplification of HIV env, followed by cloning and sequencing, was performed as previously described37. Analyses of evolution and site-specific selection were performed using BEAST38 and CODEML39. CODEML was used to identify sites that have been subjected to positive selection using site-specific codon models. Each analysis was performed with up to 10,000,000 generations and 10% burn-in, reviewing the log file every 1000 generations. The site frequency spectrum was calculated by subtracting the unstimulated control value from the stimulated sample.

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Author contributions
K.H.G., A.J.F. and P.J.G. performed the sequence and T-cell analysis. D.B., E.C.T. and M.O.M. performed and interpreted the NAb analysis. A.K. and O.P. performed the bioinformatics analyses. S.J.E., D.M. and J.N.W. performed the clinical studies. G.S.C. and P.K. coordinated the studies in Oxford and London and all authors contributed to the article preparation.

Additional information
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Supplementary Information
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