Research Paper

Plasma Levels of Endothelial Cell-Specific Molecule-1 as a Potential Biomarker of Oral Cancer Progression

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Abstract

In Taiwan, oral cancer is the fourth most common cancer and the most common malignancy with a poor prognosis. Endothelial cell-specific molecule-1 (ESM-1) is secreted by vascular endothelial cells in the liver, lungs, kidneys, and gastrointestinal tract. ESM-1 expression is associated with tumor prognosis, metastasis, and angiogenesis in many cancers. However, few studies have examined the association of plasma ESM-1 levels with oral squamous cell carcinoma (OSCC) progression. We measured the plasma ESM-1 levels of 438 male OSCC patients through a commercial enzyme-linked immunosorbent assay. The Cancer Genome Atlas (TCGA) dataset was also used to analyze the ESM-1 levels in 328 OSCC patients and 33 normal tissues. Our results revealed that the plasma ESM-1 levels of OSCC patients were significantly associated with the tumor (T) status but not with the lymph node status, metastasis, and cell differentiation. TCGA bioinformatics database analysis revealed that ESM-1 expression was significantly higher in OSCC patients than in normal individuals (p < 0.05). In addition, the examination revealed similar results for the ESM-1 expression levels and pathological stage in OSCC. In conclusion, plasma ESM-1 is a novel biomarker for predicting the T status in OSCC patients.

Key words: ESM-1, oral squamous cell carcinoma, biomarker.

Introduction

Oral cancer, which involves malignant tumors that affect any region of the oral cavity, lip, salivary glands, and pharyngeal regions, is a major problem concerning human health worldwide. Oral squamous cell carcinoma (OSCC) is the most frequently (approximately 90%) occurring malignant oral cancer, and the incidence and prevalence have increased in recent years, especially in Asian countries [1-3]. The major risk factors for oral cancer are smoking and alcohol, which have been noted in 90% of oral cancer cases [4]. Surgery is the preferred treatment approach for OSCC. The recurrence of OSCC is due to local metastasis and invasion, leading to a poor prognosis [5, 6]. Therefore, more accurate and acceptable biomarkers must be developed for early cancer detection and to predict OSCC progression.

Endothelial cell-specific molecule-1 (ESM-1) or endocan is a 50-kDa secretory proteoglycan composed of a 165-amino acid mature protein core (20 kDa) and approximately 30 kDa of a unique dermatan sulfate chain linked to serine residues [7-9]. It is predominantly secreted by vascular endothelial cells from the liver, lungs, gastrointestinal tract, and kidneys [10, 11]. ESM-1 production is highly
regulated by the environment. Angiogenic factors and inflammatory cytokines, such as interleukin (IL)-1β and tumor necrosis factor-α (TNF-α), induce ESM-1 expression and are strongly downregulated by IL-4 and interferon-γ [7, 8, 10]. ESM-1 is expressed not only in normal tissues but also in various types of tumors, with differential expression levels in glioblastoma [12], non–small cell lung cancer [13], gastric cancer [14], colorectal cancer [14], renal cell cancer [15, 16], bladder cancer [17], ovarian cancer [18], hepatocellular carcinoma [19], and acute myeloid leukemia [20]. ESM-1 plays an influential role in angiogenesis and tumor growth. In tumor tissues, angiogenesis is crucial for progression and is an indicator of poor prognosis. ESM-1 levels are associated with increased proangiogenesis growth factors, such as fibroblast growth factor-2 (FGF-2), vascular endothelial growth factor (VEGF), and hepatocyte growth factor/scatter factor, and are regulated by VEGF [8, 21]. ESM-1 is also overexpressed at the mRNA and/or protein levels in cancer tissues, and it has been related to tumor progression and poorer survival [22, 23]. Moreover, a meta-analysis study which including 15 eligible studies of 1,464 patients also mentioned that high ESM-1 expression predicts poor overall survival in gastrointestinal and hepatocellular carcinoma [24].

Previously, ESM-1 has been observed not only in the serum of septic patients [25] but also as a potential serum biomarker in gastric cancer [26], bladder cancer [27], colorectal cancer [28] and hepatocellular carcinoma [29] patients. Furthermore, higher levels of serum ESM-1 are correlated with a poor prognosis [30]. However, whether serum ESM-1 levels are elevated in OSCC patients remains unclear, and the association between the serum levels of ESM-1 and prognosis of OSCC patients has not been elucidated. The present study examined whether serum ESM-1 can be used as a biomarker for the early diagnosis of OSCC.

Materials and Methods

Patient Specimens

In total, 438 OSCC male patients (mean age, 55.23 years) from 2008 to 2012 were recruited from Chung Shan Medical University Hospital in Taichung and Changhua Christian Hospital in Changhua, Taiwan. Data of the tumor stage, tumor, node, and metastasis (TNM) status and cell differentiation were obtained from the medical records of the OSCC patients. The OSCC patients were diagnosed according to the TNM classification, which is described in the American Joint Committee on Cancer (AJCC) Staging Manual, seventh edition.

Whole peripheral blood samples were collected from OSCC patients and placed in EDTA tubes. After the centrifugation of the blood samples at 3000 rpm for 10 min, the supernatants were placed at −80°C. This study was approved by the Institutional Review Board of Chung Shan Medical University Hospital (CSMUH No: CS13214-1), and informed written consent for participation was obtained from all participants. Table 1 shows the clinical characteristics of the patients.

Table 1. Demographic characteristics and clinical features of OSCC patients.

| Variables            | OSCC (n = 438) |
|----------------------|---------------|
| Age (years)          | 55.23 ± 10.82 |
| Smoking status       |               |
| Yes                  | 392 (89.5%)   |
| No                   | 46 (10.5%)    |
| Drinking status      |               |
| Yes                  | 233 (53.2%)   |
| No                   | 205 (46.8%)   |
| Betel nuts chewing   |               |
| Yes                  | 352 (80.4%)   |
| No                   | 86 (19.6%)    |
| Cancer location      |               |
| Buccal mucosa        | 157 (35.8%)   |
| Tongue               | 130 (29.7%)   |
| Gingiva              | 68 (15.5%)    |
| Others               | 83 (18.9%)    |
| Stage                |               |
| I                    | 118 (27.0%)   |
| II                   | 79 (18.0%)    |
| III                  | 43 (9.8%)     |
| IV                   | 198 (45.2%)   |
| Tumor T status       |               |
| T1                   | 141 (32.2%)   |
| T2                   | 120 (27.4%)   |
| T3                   | 37 (8.4%)     |
| T4                   | 140 (32.0%)   |
| Lymph node status    |               |
| N0                   | 287 (65.5%)   |
| N1                   | 48 (11.0%)    |
| N2                   | 99 (22.6%)    |
| N3                   | 4 (0.9%)      |
| Metastasis           |               |
| M0                   | 436 (99.5%)   |
| M1                   | 2 (0.5%)      |
| Cell differentiation  |               |
| Well differentiated   | 59 (13.5%)    |
| Moderately or poorly |               |
| differentiated       | 379 (86.5%)   |

Quantitative Analysis of Plasma ESM-1 Levels

The ESM-1 levels in the plasma samples of OSCC were analyzed using an ESM-1 Human ELISA Kit (LIK-1205, Lunginnov, Lille, France). The prepared standards and samples were added to an ELISA plate, according to the manufacturer instructions. After reading the absorbance of each well at 450 nm in a microtest plate spectrophotometer (STENERGY/H4, BioTek Instruments, Inc., Winoosi, VT, USA), the
ESM-1 levels were quantified using a calibration curve, with the provided human ESM-1 as a standard.

**Expression Analysis of The Cancer Genome Atlas OSCC data**

The Cancer Genome Atlas (TCGA; URL: https://tcga-data.nci.nih.gov/tcga/) was used to obtain the ESM-1 normalized expression data and associated clinical data, which corresponds to the head and neck squamous cell carcinoma dataset (n = 528). After filtering the samples, we included only one of six oral cancer anatomic subtypes (alveolar ridge, base of tongue, buccal mucosa, floor of mouth, oral cavity, and oral tongue; filtered oral cancer dataset size: n = 328). Box plots for ESM-1 expression values were generated with respect to the tumor grade/stage and TNM status.

**Statistical Analysis**

The standard deviation of the mean was used to express the values. The statistical significance of the means for ESM-1 levels was determined using the Mann–Whitney Rank sum test between groups. In addition, alcohol consumption status, smoking status, and betel nuts chewing status were analyzed using the Chi-squared test. Student t-test was used to calculate the significances of the differences. Analyses were performed using SPSS 16.0 statistical software (SPSS Inc., Chicago, IL, USA). Statistical significance was set at p < 0.05.

**Results**

**Demographic Data**

In this study, only male OSCC patients (n = 438) were included. The demographic data presents the tumor stage, TNM status, and tumor differentiation status of these patients (Table 1). Among these patients, 89.5% smoked, 53.2% consumed alcohol, and 80.4% chewed betel nut. The tumors were located in the buccal mucosa (n = 157), tongue (n = 130), gingiva (n = 68), and other parts (n = 83).

**Correlation of Plasma ESM-1 Levels and Clinicopathological Characteristics of OSCC Patients**

The mean plasma ESM-1 levels were significantly higher in stage II patients (2.58 ± 1.39 ng/mL) than in stage I (2.1 ± 1.32 ng/mL) patients (p = 0.016; Figure 1A). In addition, the mean plasma ESM-1 levels were significantly higher in patients with T2 status (2.47 ± 1.31 ng/mL) than in patients with T1 status (2.09 ± 1.32 ng/mL) (p = 0.021; Figure 1B). No significant differences were noted between the N statuses (Figure 1C). The correlation of plasma ESM-1 levels with the clinicopathological characteristics is presented in Table 2. High plasma ESM-1 levels were significantly associated with the T status. Compared with patients with low plasma ESM-1 levels, a higher proportion of patients with high plasma ESM-1 levels had T1-T3 status (77.3% vs. 62.2%; p = 0.001).

![Figure 1. ELISA-determined plasma ESM-1 level of OSCC patients. ESM-1 levels were compared according to stage (A), T status (B) and N status (C).](http://www.medsci.org)
**Correlation of ESM-1 Levels and Clinicopathological Characteristics of OSCC Patients From TCGA**

TCGA OSCC database was used to verify the findings of our study. The ESM-1 mRNA levels, pathological stage, pathological T and N status of OSCC, and normal tissues were assessed. The ESM-1 expression levels in different cancer types are shown in Figure 2A. Higher ESM-1 levels were detected in OSCC tissues than in normal tissues (p = 0.03; Figure 2B). Among the OSCC patients, the relative levels of ESM-1 mRNA were significantly higher in stage III patients than in stage I patients (p = 0.0221; Figure 2C). The relative levels of ESM-1 mRNA were significantly higher in patients with T3 status than in patients with T1 status (p = 0.0138; Figure 2D). However, the relative levels of ESM-1 mRNA were not significantly associated with the N status (Figure 2E).

**Table 2. Correlation between plasma levels of ESM-1 and clinicopathological parameters in 438 OSCC patients.**

| Variables                  | ESM-1 levels | p value |
|----------------------------|--------------|---------|
| Age (years)                |              |         |
| <55                        | Low levels (n=275) | 143 (52.0%) | 141 (86.5%) | 0.020* |
| ≥55                        | High levels (n=163)   | 66 (40.5%)  | 60 (36.4%)  |         |
| Smoking status             |              |         |
| No                         | Low levels (n=275) | 28 (10.2%)  | 33 (20.2%)  | 0.012* |
| Yes                        | High levels (n=163) | 18 (11.0%)  | 18 (11.0%)  |         |
| Drinking status            |              |         |
| No                         | Low levels (n=275) | 116 (42.2%) | 54 (33.2%)  | 0.080  |
| Yes                        | High levels (n=163) | 89 (54.6%)  | 48 (29.4%)  |         |
| Betel nuts chewing         |              |         |
| No                         | Low levels (n=275) | 53 (19.3%)  | 48 (29.4%)  | 0.196  |
| Yes                        | High levels (n=163) | 33 (20.2%)  | 33 (20.2%)  |         |
| Cancer location            |              |         |
| Buccal mucosa              | Low levels (n=275) | 109 (39.6%) | 76 (27.6%)  | 0.059  |
| Tongue                     | High levels (n=163) | 48 (27.4%)  | 54 (33.2%)  |         |
| Gingiva                    | Low levels (n=275) | 40 (14.5%)  | 40 (14.5%)  | 0.196  |
| Others                     | High levels (n=163) | 28 (17.2%)  | 28 (17.2%)  |         |
| Stage                      |              |         |
| I+II                       | Low levels (n=275) | 127 (46.2%) | 127 (46.2%) | 0.594  |
| III+IV                     | High levels (n=163) | 71 (43.6%)  | 71 (43.6%)  |         |
| Tumor T status             |              |         |
| T1                         | Low levels (n=275) | 104 (37.8%) | 104 (37.8%) | 0.001* |
| T2+T3+T4                   | High levels (n=163) | 37 (22.7%)  | 37 (22.7%)  |         |
| Lymph node status          |              |         |
| N0                         | Low levels (n=275) | 184 (66.9%) | 184 (66.9%) | 0.429  |
| N1+N2+N3                   | High levels (n=163) | 103 (63.2%) | 103 (63.2%) |         |
| Metastasis                 |              |         |
| M0                         | Low levels (n=275) | 275 (100.0%) | 275 (100.0%) | 0.066  |
| M1                         | High levels (n=163) | 161 (98.8%) | 161 (98.8%) |         |
| Cell differentiation       |              |         |
| Well differentiated        | Low levels (n=275) | 37 (13.5%)  | 37 (13.5%)  | 0.990  |
| Moderately or poorly       | High levels (n=163) | 22 (13.5%)  | 22 (13.5%)  |         |
| differentiied              |              |         |

*p < 0.05.

**Discussion**

ESM-1 is a soluble proteoglycan and a freely circulating molecule in the blood; hence, it has high potential as a biomarker. This is the first study to investigate the potential roles of ESM-1 in the peripheral blood of OSCC patients and the association between the plasma ESM-1 levels and clinicopathological characteristics. In this study, the plasma ESM-1 levels of OSCC patients were significantly associated with the tumor stage and T status. Similar results were also obtained in TCGA bioinformatics database analysis.

Tumor progression is a complex process, and the microenvironment not only plays a crucial role in cancer progression but also has profound effects on metastasis and angiogenesis [31-34]. ESM-1 expression is regulated by various cytokines and cell factors, such as IL-1 and TNF-α, which may promptly induce increased serum ESM-1 expression to regulate tumor-associated inflammation [25]. Increased ESM-1 expression by endothelial cells is associated with certain angiogenic factors, such as FGF-2 and VEGF-A [35]. In human renal cancer, VEGF-A regulates ESM-1 expression through the PI3K/Akt signaling pathway and the protein kinase C activator [15]; VEGF-C and nuclear factor-kB (NF-kB) upregulate ESM-1. In ESM-1 knockout mice, vascular outgrowth was delayed, filopodia extension was reduced, phosphorylated Erk1/2 (extracellular signal-regulated kinases 1/2) expression in sprouting vessels was decreased, and VEGF-A-dependent processes were effected [35]. Thus, ESM-1 plays a role in angiogenesis and inflammation. Numerous studies have demonstrated that ESM-1 can be used as a potential biomarker for detection in gastric cancer [36], bladder cancer [37], colorectal cancer [28], hepatocellular carcinoma [38], pituitary adenoma [39], ovarian cancer [40], breast cancer [41], and acute leukemia [20], but ESM-1 has not been investigated in OSCC.

In various cancers, higher than normal ESM-1 expression levels have been detected. In addition, tumor size has been positively correlated with circulating ESM-1 expression levels. In colorectal cancer, knockdown ESM-1 expression may affect cell cycle arrest, decrease cyclin D1, and regulate metastatic process through the decline of NF-κB, phosphor-Akt, and GSK3α/3β expression [42]. In SK-Hep1 cells, ESM-1 siRNA expression indicates that ESM-1 may increase the survival rate through the NF-κB/1KB pathway [43]. Moreover, ESM-1 expression was associated with higher serum alpha fetoprotein levels and larger tumor in early hepatocellular carcinoma [44]. The metastatic process
is also inhibited through the regulation of matrix metalloproteinase (MMP)- and epithelial-to-mesenchymal transition–related genes in cells with lower ESM-1 expression [42]. In murine OSCC, ESM-1 mediates nerve growth factor receptor (NGFR)-induced metastasis and invasion. Cells with ESM-1 overexpression may exhibit enhanced cell invasion and migration in vitro. In severe combined immunodeficient mice with NGFR-induced tumors, ESM-1 knockdown cells may reduce tumor formation and metastasis [45]. However, our study did not examine the regulation of plasma ESM-1 levels. In the future, we will examine the possible mechanisms of ESM-1 expression in OSCC.

Figure 2. ESM-1 mRNA level of OSCC patients from TCGA database. (A) The different of ESM-1 mRNA level from The Broad Institute TCGA GDAC Firehose in different type of cancer. (B) ESM-1 levels were compared according to normal people and OSCC patients. (C) ESM-1 levels were compared according to stage. (D) ESM-1 levels were compared according to tumor T status. (E) ESM-1 levels were compared according to N status. SARC: sarcoma. LIHC: liver hepatocellular carcinoma. KIRC: kidney renal clear cell carcinoma. HNSC: head and neck squamous cell carcinoma. GBM: Glioblastoma multiforme. COADREAD: colorectal adenocarcinoma. COAD: colon adenocarcinoma. CESC: cervical and endocervical cancers. BRCA: breast invasive carcinoma. BLCA: bladder urothelial carcinoma.
Most studies have indicated that ESM-1 is overexpressed in various cancer types and is associated with tumor progression and poor prognosis. However, contrary results have been reported in prostate cancer cells. Knockdown ESM-1 enhanced cell viability, cell migration, and cell invasion in vitro and increased the tumorigenesis and metastasis of prostate xenograft tumors in vivo through the regulation of TIMP-1/MMP-9 expression. ESM-1 plays a role in tumor suppression in prostate cancer cells [46].

In conclusion, our study revealed that plasma ESM-1 levels are useful for the detection of disease progression in OSCC patients. This is the first study to analyze plasma ESM-1 levels in patients with OSCC. We demonstrated that plasma ESM-1 levels were significantly associated with the T status in OSCC patients. As a secreted protein, ESM-1 may act as a potential marker of OSCC during tumor progression and can accordingly be used for disease diagnosis; furthermore, it plays a critical role in carcinogenesis.

Acknowledgments

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Competing Interests

The authors have declared that no competing interest exists.

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