Andean Root and Tuber Crops: Underground Rainbows

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The Andean region is recognized today as one of the most important centers of crop origin and diversity in the world (National Research Council, 1989). Many of our most important food crops worldwide, most notably potatoes, were domesticated in this system (National Research Council, 1989). A unique feature of the Andean agricultural system is a taxonomically diverse group of Andean root and tuber crops (ARTC), coupled to the commonly found complement of grains and legumes, such as maize (Zea mays L.), chenopodium, beans, and lupines. The ARTC played a vital role in allowing sustained intensive cultivation and nourishment of a population that reached 10–12 million people at the height of the Incan Empire (1450–1535 A.D.) (King, 1987). About 17 species of root and tuber crops were domesticated in the Andes, making this the largest known geographical concentration of underground crops (Hernandez-Bermejo and Leon, 1992; Tapia, 1993).

The ARTC evolved in extremely inhospitable areas for agriculture, and Pre-Columbian people made extremely efficient use of what by today’s agricultural standards would be considered marginal land (King, 1987). The steep slopes of the Andes are prone to constant erosion, extreme fluctuations in rainfall and temperature, and contain relatively poor soils. Crops grown in this environment were selected for their ability to cope with long periods of drought, freezing temperatures, and high UV irradiation. The Andean farmers took advantage of natural plant adaptations to extreme environments to domesticate a unique cohort of crops, and combined their cultivation with the use of complex irrigation canals, crop rotation, intercropping techniques, and soil preservation practices (Flores and Flores, 1997). Underground storage organs are among the most common and efficient structures evolved by plants for survival in challenging environments. Root and tuber crops can also exhibit some of the highest yields for calories produced per area of cultivation; thus their adaptation and diversification in the Andes made both ecological and economic sense.

The ARTC today are common staples for an estimated 25 million people in the Andean highlands, and are at least an occasional food source for another 100 million people in Ecuador, Colombia, Peru, Bolivia, Argentina, and Chile. The traditional agricultural system of the Andes faces many challenges and uncertainties. Therefore, the need for promoting and preserving ARTC is immediate such that studies are necessary to improve pest management and crop productivity to motivate farmers towards the cultivation of these crops. Furthermore, the susceptibility of some of the more important ARTC such as mashua and ulluco to viral diseases warrants the need to provide disease-free accessions to minimize annual crop losses due to disease. The potential uses of the Andean root and tuber crops are discussed in this paper as we review their biology and biochemistry.

THE ANDEAN ROOT AND TUBER CROPS

The ARTC span no fewer than nine plant families: Asteraceae, Basellaceae, Brassicaceae, Leguminosae, Nyctaginaceae, Oxalidaceae, Solanaceae, Tropaeolaceae, and Umbelliferae. The genetic diversity and potential of these species is remarkable. Large germplasm collections are available for the three major tuber crops, oca, mashua, and ulluco, with about 8000 accessions combined. Thus, substantial ARTC biodiversity still persists in situ and ex situ. Oca, ulluco, and mashua alone could significantly expand the range of nutritional properties represented in the commonly cultivated and consumed Solanum sp. The ARTC represent a vast and mostly untapped pool of variation in type and content of starch, amino acid composition, other nutritional factors, and natural pesticides. The great adaptability of the ARTC favors their potential cultivation outside their area of origin. For example, oca is currently grown in Australia and New Zealand (National Research Council, 1989), while other species have recently been introduced to Mexico, Central America, Brazil, Europe, and Australia.

Tuber crops

Oca (Oxalis tuberosa, Oxalidaceae). After potato, oca is the most common tuber crop in the Andean region (Pulgar-Vidal, 1981). Oca is a prostrate herb with a compact growth habit of 20–30 cm in height with cylindrical succulent stems varying in color from green to red (Fig. 1), and the tubers range from cylindrical to ovoid, averaging 7 to 11 cm in length. The texture of the tuber skin varies from smooth to rough, and the color is white, yellow, pink, red, purple, or black (Seminario, 1988). Oca is widely adapted to divergent environments, and it is commonly cultivated in areas with elevations between 2800 and 4000 m and a range of precipitation of 570–2500 mm (Seminario, 1988). It grows in moderately cool temperatures down to 5 °C, and poor soil conditions with a pH range of 5.3–7.8 (Leon, 1964; Seminario, 1988). This crop is grown in greatest abundance in the highlands of Ecuador, Peru, and Bolivia, but is also found in regions of Chile, Argentina, Colombia, and Venezuela. Oca has been commercialized in New Zealand, Australia, Mexico, France, and Great Britain either as a food staple or as a home-garden ornamental (King and Gershoff, 1987; National Research Council, 1989).

Evidence suggests that the cultivated oca is an octoploid species, 2n = 8x = 64 chromosomes (Arbizu and Tapia, 1994). For practical purposes, the crop is sterile and thus maintained vegetatively. The ex-situ oca germplasm is distributed amid several genebanks in South America, with 482 accessions cataloged at the International Potato Center–Peru (CIP), 1696 at Instituto Nacional de Investigacion Agropecuaria–Peru (INIA), 912 at Universidad Nacional San Antonio–Peru (UNSAAC), 680 at Instituto Boliviano de Tecnologia Agropecuaria–Bolivia (IBTA). However, Andean...
farmers primarily maintain the biodiversity of oca through on-farm propagation (Arbizu, 2000; Arbizu et al., 1997), and only a small fraction of the total germplasm has been exploited by plant breeders to develop new cultivars (Hodge, 1985).

Oca tubers show high variability in nutritional levels between genotypes, and they are a good source of carbohydrates, calcium, iron, fats, and fiber (Table 1). Among different morphotypes, protein levels vary widely, ranging from 1% to 9% on a dry-weight basis. The bitter morphotypes contain oxalic acid at concentrations up to 500 µg·kg⁻¹ (National Research Council, 1989). As is the case for most Andean root and tuber crops, there are very few reports on the basic biology, agronomy, and biochemistry of oca.

**Mashua** (*Tropaeolum tuberosum, Tropaeolaceae*). Mashua is an herbaceous perennial plant, and a close relative of the garden nasturtium. The plant habit is prostrate or climbing and it grows over 1 m in diameter and 0.5 m in height, and produces slender leaves (Fig. 1). The tubers vary in color and shape. The tuber skin is white, yellow, or occasionally purplish or red. Some tubers are mottled or striped with red or purple, particularly below the “eyes,” and as in potato and oca, small leaf scales border the deep-set “eyes.” The flesh of the tuber is generally yellow irrespective of the morphotype (National Research Council, 1989). Mashua can be propagated vegetatively, but it also produces a large quantity of viable seed (Arbizu and Tapia, 1994).

Although mashua is grown from 2400 to 4300 m, the best production of mashua tubers occurs at elevations around 3000 m, with maximum yields of 30,000 kg·ha⁻¹. Mashua is usually cultivated once a year because tuber formation occurs only during short photoperiods of ~11–13.5 h of daylight. The life cycle of 5–6 months is relatively short compared with other root and tuber crops (National Research Council, 1989; Zela et al., 1997). Without traditional irrigation practices, mashua requires annual precipitation amounts between 700 and 1600 mm, and the best soil type for this crop is a fertile, organic soil with pH values from 5.3 to 7.5 (Torres et al., 1992).

Mashua is cultivated in Argentina, Bolivia, Colombia, Ecuador, Peru, and Venezuela, and has recently been introduced to New Zealand. Andean farmers recognize a number of different mashua morphotypes based on color, form, and taste of the tubers. Domesticated mashua is thought to be comprised of more than 400 different morphotypes (Sperling and King, 1990). Mashua accessions in genebanks, distributed among different institutions around the Andean region, include 101 at Cuzco (Peru), 146 at Ayacucho (Peru), 125 at Universidad Nacional Mayor de San Marcos–Peru (UNMSM), 55 at Programa de Investigacion de Papa–Bolivia (PROIMPA), and 43 at Instituto Nacional de Investigacion Agropeciaria–Ecuador (INIAP).

The majority of these accessions are maintained through in vitro propagation.

The Andean highlanders use mashua as a food crop and as a medicinal crop. All parts of the plant can be consumed, including the tubers, leaves, and blossoms, but the tuber is the most commonly consumed because of its flavor and nutritional value (National Research Council, 1989). Compared to other Andean root and tuber crops, mashua contains high levels of ascorbic acid (vitamin C), thiamin (vitamin B1), riboflavin (vitamin B2), as well as lipids (Table 1), and the tubers are relatively high in protein content and fiber. These nutritional characteristics make the mashua tuber an important component of an Andean highlander diet.

**Ulluco** (*Ullucus tuberosus, Basellaceae*). Ulluco is a low-growing herb with heart-shaped succulent leaves, and morphotypes vary from prostrate, semi-climbing vines to dense and compact bushes (Fig. 1). Tubers can be elongated or curved, with a thin, soft
Morphotypes differ in the time it takes to reach maturity, varying from 5–9 months and 10–13.5 h of daylight (Vega, 1997). Ulluco requires 700–800 mm of precipitation to sustain productive yield, but it is considered well adapted to dry conditions and slightly acidic soil (5.5 to 6.5) (Vega, 1997).

Ulluco tubers are an excellent source of protein, carbohydrates, and vitamin C (Busch et al., 2000) (Table 1). Because the tubers are rather perishable, pre-Columbian people processed the tubers using an environmental freezing and drying process, which takes advantage of the day–night temperature differential during the dry period (May through September). The long-lasting product of this process is called lingli or chuki, which is usually ground into flour and added to cooked foods. Similar “freeze-drying” processes have been used to preserve native potatoes, oca and mashua (National Research Council, 1989). The dried tubers are a major nutrient source in this crop, containing ≈75% to 80% starch and 6% to 14% sugar, most of which is glucose and sucrose (National Research Council, 1989). Achiara flour and starch are used for the production of specialty noodles and other products in South East Asian countries (International Potato Center, 1994).

**Root crops**

**Maca (Lepidium meyenii, Brassicaceae).** Maca, a member of the radish family, occupies a rather restricted ecological zone in central Peru at elevations of 3500 to 4500 m above sea level. Maca is possibly the only crop in these high mountains, where frost can occur at midday in summer, and survive (Bonnier, 1986). In this habitat, frost can be damaging to cereal grains such as maize, rice (Oryza sativa L.), and wheat (Triticum aestivum L.), and can be stored for several years. The major nutrients in this crop are calcium (258 mg) and iron (15.4 mg) per 100 g fresh weight (National Research Council, 1989). The dried hypocotyls contain up to 59% carbohydrate, 10% protein, 8.5% fiber, 2.2% lipid, and are also rich in starch, glucosides, alkaloids, and tannins (Dini et al., 1994). Total protein content may fluctuate between 10% and 14% depending upon the variety and soil fertility.

**Mauka (Mirabilis expansa, Nyctaginaceae).** Mauka is a relatively disease-resistant root crop, whose historical importance is just now coming to light as it was rediscovered only a few years ago. In the early 1960s, the Bolivian scientist Julio Rea first described mauka as a vital food source of the Maukallajita Indians inhabiting the high valleys of La Paz, Bolivia (Rea and Leon, 1965). Fifteen years later, the cultivation of mauka was confirmed in the cold, dry regions north of Quito, Ecuador, and later reported in Cajamarca, Peru (National Research Council, 1989).

Mauka is a low compact herbaceous plant not exceeding 1 m in height, and is a relatively disease-resistant crop that produces edible storage roots (Franco and Vejarano, 1996) (Fig. 1). The cultivation of this crop has been restricted to three small regions in Peru, Ecuador and Bolivia (National Research Council, 1989), and it is believed to face strong genetic erosion. It grows well at high elevations (2200–3500 m) and at temperatures between 4° and 29 °C (Franco and Vejarano, 1996) (Fig. 1). The underground storage roots are a reproductive phase in the following season (Tello et al., 1992). Estimated fresh yields of hypocotyls harvested from May to July are 14.7 t ha⁻¹, resulting in nearly 4.4 t ha⁻¹ of dried hypocotyls (Tello et al., 1992). Although highly regarded for its medicinal uses, maca is also valued for nutritional reasons. The dried hypocotyls are comparable in nutritive value to cereal grains such as maize, rice (Oryza sativa L.), and wheat (Triticum aestivum L.), and can be stored for several years. The major nutrients in this crop are calcium (258 mg) and iron (15.4 mg) per 100 g fresh weight (National Research Council, 1989). The dried hypocotyls contain up to 59% carbohydrate, 10% protein, 8.5% fiber, 2.2% lipid, and are also rich in starch, glucosides, alkaloids, and tannins (Dini et al., 1994). Total protein content may fluctuate between 10% and 14% depending upon the variety and soil fertility.

Achira is a perennial monocotyledon closely related to the ornamental canna grown in temperate and tropical zones. Achira grows to 1 m in height and produces rhizomes of ≈60 cm in length (Fig. 1). It is cultivated in the Andean region and in Mexico and the West Indies. In South America, achira is distributed from the Amazon basin through the Pacific coast to northern Chile and Argentina at altitudes ranging from 1000 to 2900 m (Flores and Flores, 1997; National Research Council, 1989). Compared to the other Andean tuber crops the biodiversity of achira is rather lacking; only 20 clones of achira for example have been maintained at CIP (International Potato Center, 1994). Cultivars vary in foliage color, stem height, rhizome size and coloration, and are propagated exclusively by vegetative means (Flores and Flores, 1997).

It is likely that achira originated in the tropics of South America (Ugent et al., 1984) and has long been domesticated and cultivated in Peru. Archeological remains of achira are found in Pre-Ceramic cultures in South America, dating back to 2250 BC (Ugent et al., 1984). Further evidence for the ancient use of achira rhizomes is found in the arts and crafts of the pre-Inca civilizations of Nasca and Chimú (Montaldo, 1977).

The rhizomes of achira are very rich in carbohydrates, containing ≈75% to 80% starch and 6% to 14% sugar, most of which is glucose and sucrose (National Research Council, 1989). Achiara flour and starch are used for the production of specialty noodles and other products in South East Asian countries (International Potato Center, 1994).

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Mauka provides a wealth of edible stems and storage roots with a comparatively high protein and carbohydrate content. Mauka roots contain \( \approx 87\% \) carbohydrate on a dry weight basis (Montenegro and Franco, 1988; National Research Council, 1989) and accessions from Bolivia and Peru have 7% and 5% protein content, respectively (Hernandez-Bermejo and Leon, 1992). Similarly, the calcium levels in mauka roots are nearly 1000% higher than the other ARTC, although they occur as calcium oxalate raphides in root cortex cells for the most part and thus are nutritionally unavailable. The calcium levels vary from 157 to 461 mg per 100 g of dry weight (Montenegro and Franco, 1988). Phosphorus levels are also elevated compared to the other ARTC; however, sodium and iron levels are low in mauka. The leaves contain \( \approx 17\% \) protein with a higher level of digestibility than other forages cultivated in the Andean highlands (National Research Council, 1989).

**Medicinal applications of ARTC**

The ARTC have recently gained worldwide interest due to their purported medicinal properties for humans. For centuries, plants have been used by civilizations for medicinal purposes. Roots from several plants have been used as sedatives, stimulants, antiminthimics, and anti-microbials, including mandrake (Mandragora officinalis L.), ginseng (Panax ginseng L., P. quinquefolium L.), ipecac (Cephalis ipecacuanha Brot.), and valerian (Valeriana officinalis L.) (Foster and Duke, 1990). Similarly, many of the ARTC have been employed by the natives of the Andean region to ease labor pains, treat kidney ailments, and to increase and/or decrease fertility and libido in males and females.

According to the common folklore of the Andean highlands, maca is an aphrodisiac that improves male and female fertility and is believed to improve the energy and vitality and to improve the reproductive capabilities of the domesticated animals at high elevations (Sanchez Leon, 1964). During the conquest of the Incan Empire, the Spaniards fed maca to their soldiers, horses, chickens, and pigs to enhance their energy and vitality and to improve the reproductive capabilities of the domesticated animals at high elevations (Sanchez Leon, 1996). Recent clinical studies with rats (Ciceri et al., 2001, 2002) and adult men (Gonzalez et al., 2001) have supported the fertility-enhancing properties of maca. The reported gains in female fertility are likely due to an increase in the development of Graafian follicles, the fluid-filled vesicles of the mammalian ovary containing maturing ova (Chacon, 1990; Rea, 1992). Nutraceutical companies in the U.S. now market maca in pill form as a supplement reputed to enhance stamina and fertility, but as for many such products there is a wide variation in quality control. Chemical analysis of maca indicates that the fertility-enhancing properties may be attributed to the presence of biologically active aromatic isothiocyanates, in particular benzyl isothiocyanate and \( \text{p-} \) methoxybenzyl isothiocyanate (Johns, 1981), or possibly due to the existence of prostaglandin-like compounds and phytoestrogens in the hypocotyls (Dini et al., 1994).

The domestication of mashua also has been associated with its numerous medicinal uses in the folk medicine of the Andean region (Johns et al., 1982). Mashua is considered an antiproliferative, believed to cause impotence and infertility in males. Experimental studies on male rats that were fed a diet of mashua tubers showed a 45% decrease in testosterone/dihydrotestosterone levels (Johns et al., 1982). In females, mashua is proposed to increase fertility due to \( N,N\text{-di-(methoxy-4-benzyl)} \) thiourea contained in the tubers, a compound that competitively inhibits estradiol binding and increases estrogenic activity (Johns et al., 1982). Mashua tubers have been reported in Andean folk medicine as an antibiotic and diuretic and have been used to remedy multiple kidney ailments and diseases, as well as to eliminate bladder and kidney stones (Johns et al., 1982). In modern medicinal practices in Bolivia, mashua is employed as an emmenagogue, and is thought to stimulate menstruation (Johns et al., 1982). Many of the reported medicinal properties of mashua experimentally appear to result mainly from \( \text{p-methoxybenzyl isothiocyanate} \) (Johns and Towers, 1981). The reported antibiotic, antitumor, and diuretic properties of isothiocyanates support the many uses of mashua in Andean folk medicine (Fahey et al., 1997; Johns et al., 1982; Talalay and Fahey, 2001).

Ulluco in the highlands is used in medicinal applications for pregnant Andean women. Women consume ulluco to ease pain during pregnancy, and the severity of labor pains during childbirth (National Research Council, 1989), but no scientific research has been conducted to identify the bioactive principles. Additionally, little is known about the medicinal properties of mauka and och; however, \( \text{Mirabilis} \) species native to the American Southwest have been utilized by the Hopi Indians in North America for medicinal purposes (Moore, 1988; Seminario and Seminario, 1995).

**ETHNOBOTANICAL CONNECTIONS**

Andean farmers developed a sophisticated repertoire of practices to cope with rugged planting terrain, diverse environmental conditions, and indigenous diseases, which create an ongoing challenge to grow and maintain successful plots of ARTC each year. The severely sloping and eroding mountain slopes posed a serious challenge to the area’s agriculture. To alleviate the problem of soil erosion and to provide suitable growing plots, the ancient Andean farmers constructed elaborate terraces across the steep slopes, perched one above another along the mountainsides (Fig. 2). The highlands of Peru contain nearly 60,000 ha of terraced farmland, and tubers such as oca, potato, ulluco, and mashua are still planted in some of these plots today (National Research Council, 1989).

The environmental conditions in the highlands of Peru are perhaps the most extreme and varied in the world. For instance, at the base of a single terraced slope, moderate to hot temperatures and accompanying rainfall may prevail, while near the summit cold temperatures and frost predominate (Fig. 2). Andean farmers utilized knowledge of these microclimates to protect against crop failure, understanding that crop or variety adaptation may be different at each elevation. Vertically diversified farming practices encouraged the development and planting of several different crop varieties, each with slightly different tolerances to temperature, moisture, soil type, and other factors (National Research Council, 1989). The rationale for these practices is that they served as a form of insurance in the event that one or more morphotypes failed, and the different growth cycles at differing elevations allowed the work to be staggered over a longer period of time, which permitted more land area to be cultivated (National Research Council, 1989).

In any farming system, disease and pests represent serious threats to valuable ARTC in the Andean highlands, but under traditional agricultural practices that utilize chemically derived herbicides and pesticides, Andean farmers rely on intercropping to control disease and pests. In a single field, an Andean farmer may grow up to 200 different kinds of potatoes (Flores and Flores, 1997). In the highlands, through generations of cultural knowledge and knowledge of mashua’s disease and pest resistance, farmers have continually planted mashua in field plots with oca, potatoes, and ulluco to manage disease and pests (Fig. 3) (National Research Council, 1989). Studies by Johns et al. (1982) have shown that mashua tubers are resistant to an array of bacteria, fungi, nematodes, and insects such as the Andean weevil, a finding which validates the importance of mashua as an intercropping species throughout generations of Andean farming practices.

**BIOCHEMISTRY AND BIOTECHNOLOGY OF ANDEAN ROOT AND TUBER CROPS**

In recent years, underground storage organs have been studied in an attempt to understand the functional link between underground processes and aboveground mechanism. Apart from the classical role of providing mechanical support and nutrient uptake, roots and tubers also perform specialized roles as chemical factories that can synthesize, store, and secrete a diverse array of biologically active compounds such as proteins and low-molecular-weight secondary metabolites (Flores et al., 1999). Plant callus, cell suspension, and root cultures are systems amenable for the large-scale production of pharmaceutical enzymes (Flores et al., 1999) and offer a viable method for the production of antimicrobial proteins. Agrobacterium-transformed root cultures of mashua have been established, and root cultures showed the presence of the 58 kDa B-1,3-glucanase-related protein found in mashua tubers. Addition of jasmonate and salicylic acid to the culture medium elicited increased levels of this protein (Guinarré, 2001). Similar results were found with \( M. \text{ expansa} \) cell cultures (Vivanco and Flores, 2000). We have recently established...
Agrobacterium-transformed hairy root cultures of oca and ulluco, which may provide insights about the significance of biologically active proteins and phytochemicals from the roots of these two crops.

A protein of 18 kDa present in all oca morphotypes, was found to be the most abundant protein in oca tubers (20% to 40% of total insoluble tuber protein) (Flores, 1999; Flores et al., 2002). This protein, ocatin, appears to be a tuber-specific protein localized in the parenchyma cells of the pith and the peridermis, representing a good source of amino acids for the tuber. In addition to its storage role, oca- tin is very active against the following fungal pathogens: Rhizoctonia solani, Phytophthora cinnamomi, Fusarium oxysporum, and Nectria haematococca. Anti-bacterial activity was also found against Agrobacterium tumefaciens, A. radiobacter, Serratia marcescens, and Pseudomonas aeruginosa, and the gene encoding ocatin was isolated and observed to have high homology with pathogenesis-related (PR) proteins found in a wide variety of species (Flores et al., 2002).

Antimicrobial proteins were also purified from tubers of mashua (Guimarães, 2001). These proteins, a β-1,3-glucanase (32 kDa) and an osmotin-like protein (22 kDa), acted synergistically to inhibit the growth and spore formation of Trichoderma harzianum; however, unlike ocatin, these two proteins are present in relatively small amounts and are restricted to select morphotypes (Guimarães, 2001). A polyclonal antibody raised against the glucanase cross-reacted with a related protein of about 58 kDa found mainly in the cell walls of roots (Guimarães, 2001).

Two novel type I Ribosome Inactivating Proteins (RIP), designated ME1 and ME2, of 27 and 27.5 kDa, respectively, were purified from the storage roots of the mauka (Mirabilis expansa) (Vivanco et al., 1999b). The proteins showed additive antifungal activity against several fungi, including Pythium irregulare, Fusarium oxysporum, and Trichoderma harzianum. Antibacterial activity in both ME1 and ME2 was observed against Pseudomonas syringae, Agrobacterium tumefaciens, Agrobacterium radiobacter, and others. In addition to the antifungal and antimicrobial activities of ME1 and ME2, antiviral activity has also been confirmed for these proteins. Preventive applications of ME1 and ME2 on Gomphena globosa L. leaves inhibited mechanical infection by potato virus X (Vivanco et al., 1999a), suggesting a broad-spectrum defense function. The two proteins are localized in cell wall and parenchyma tissue of storage roots, and similar proteins were found in roots of other Mirabilis species, indicating the two proteins are genus-specific (Vivanco and Flores, 2000). The potential applications for the aforementioned findings are extensive, since these proteins can possibly be used as antimicrobial agents in transgenically modified crops or in simple crop protection methods in low input agricultural systems, such as spraying root extracts on leaves of various crops to prevent or control pathogen infection.

Andean root and tuber crops are not only a novel source of antimicrobial proteins, but of secondary metabolites as well. For instance, we recently characterized a fluorescent secondary metabolite in the root exudates of oca (Bais et al., 2002). The main fluorescent compound from oca’s root exudates was identified as harmine (7-methoxy-1-methyl-β-carboline), a widespread photoactive β-carboline with reported light-mediated activity against bacteria and insects (Larson et al., 1988). Harmine exhibits a strong purplish-blue fluorescence, which correlates to the fluorescence observed in oca’s medium (Bais et al., 2002). As discussed above, mashua is known to contain glucosinolates such as 4-methoxybenzyl-glucosinolate as the predominant glucosinolate (Johns, 1981). In some morphotypes, we have found that benzyl- and 4-methoxybenzyl-glucosinolates are the predominant forms of glucosinolates present in the plant, while traces of other glucosinolates may be found as well (Guimarães, 2001). The underground glucosinolates pattern also varies; tubers of mashua contain mainly 4-methoxybenzyl-glucosinolates and traces of 2-phenylethyl- and 2-hydroxy-2-phenylethyl-glucosinolates. Mashua hairy roots produced glucosinolates in both the roots and the exudates (Guimarães, 2001). Benzylisothiocyanate, the product of glucosinolate degradation by myrosinase, is active against a range of tumor cells (Hasegawa et al., 1992; Pintão et al., 1995). Production of benzylisothiocyanates using hairy root cultures has been achieved, and large-scale production seems highly feasible (Wielanek and Urbanek, 1999).

Traits of the Andean cultivated potatoes such as resistance to frost, virus, nematodes, and bacteria are appealing to breeders, and...
some of these properties are associated with the glycoalkaloid content. Potent molluscidal activity was found in roots of *Campanula L.*, but the actual compound causing it has not yet been identified (Tripathi and Singh, 2000). Saponins of ulluco and glucosinolates of maca may be associated with disease resistance (Flores and Flores, 1997). Franco et al. (1999) reported that some morphotypes of ulluco, mashua, and oca inhibited the egg hatching of cyst nematodes, but again further studies are needed to identify the active compounds.

**CONCLUSION**

The ARTC are a fascinating set of crops adapted to the extreme conditions of the Andean region. The six Andean tuber and root crops described and discussed in this review have been cultivated in the Andean region for centuries and continue to be important food crops in South American countries. They are a good source of nutrition and have strong aesthetic appeal due to their wide variation in form and color. Many of the diverse varieties of these species have been collected and are available for research and breeding purposes (Condesan-CIP, 1997). One of the species, *Lepidium meyenii*, has spread to Mexico and New Zealand where it is marketed and consumed in numerous dishes. Oca is currently being imported to the United States from New Zealand in increasing amounts (http://www.friedas.com/detail.cfm?id=192). Another ARTC, ulluco, is now processed in Australia and exported to the United States.

The potential for introducing several ARTC as food crops for other areas of the world is significant. Recently, there has been an effort to select adapted clones for trials in other regions of the world. These efforts are aimed at increasing the crop options available to farmers by increasing the diversity of food sources, and developing crops for marginal growing regions. Suitable areas include high-altitude regions in low latitudes and oceanic or insular climates with long, cool growing seasons. In mild climates of the southern and northern hemispheres, the ARTC could be grown as alternative winter crops. Proper selection of germplasm adapted to specific environments will determine the successful introduction of the ARTC. Factors such as the short days required for tuber formation may limit potential production areas; thus, there is a need for photoperiod-insensitive varieties. Steps taken to strengthen the biotechnological knowledge related to these root and tuber crops are commendable, although further effort is required to exploit their phytochemical and pharmacological properties. Finally, the commitment of international research to the ARTC will not only aid in the introduction of these crops to other global regions, but will enhance the current production of these crops in their native Andean region.

**Literature Cited**

Agricultural Research Service. Dr. Duke’s phytochemical and ethnobotanical database. Cited from the World Wide Web 6 Jun. 2000.
Rea, J. and J. Leon. 1965. La mauka (Mirabilis expansa) un aporte de la agricultura prehispanica de Bolivia, p. 38–41. In: Abales Cientificos de la Universidad Agraria La Molina, Lima.
Sanchez Leon, A. 1996. Que Rica Maca! Somos 495:34–36.
Seminario, J. 1988. El chago mauka (Mirabilis expansa) en Cajamarca, p. 251–264. In: Anales del VI Congreso Internacional de Cultivos Andinos, Quito, INIAP.
Seminario, J. and A. Seminario. 1995. Coleccion de germoplasm regional de raizes andinas. Boletin de Lima 98:27–47.
Sperling, C.R. 1987. Systematics of the Basellaceae. PhD Diss., Dept. of Organism and Evolutionary Biology, Harvard Univ., Cambridge, Mass.
Sperling, C.R. and S.R. King. 1990. Andean tuber crops: Worldwide potential, p. 428–435. In: J. Janick and J.E. Simon (eds.). Advances in new crops. Timber Press, Portland.
Talalay, P. and J.W. Fahey. 2001. Phytochemicals from cruciferous plants protect against cancer by modulating carcinogen metabolism. J. Nutr. 11:3027S–3033S.
Tapia, M.E. 1993. Semillas Andinas: El banco de oro, p. 76. Consejo Nacional de Ciencia y Tecnologia (concytec), Lima, Peru.
Tello, J., M. Hermann, and A. Calderon. 1992. La maca (Lepidium Meyenii Walp.): Cultivo alimenticio potencial para las Zonas Altoandinas. Boletin de Lima 81:59–66.
Toreles, O.M., M. Perea-Dallos, and T.J. Fandino. 1992. Micropropagation of Cubio (Tropaeolum tuberosum). Biotechnol. Agr. For. Berlin 19:160–171.
Tripathi, S.M. and D.K. Singh. 2000. Molluscicidal activity of Punica granatum bark and Canna indica root. Braz. J. Med. Biol. Res. 33(11): 1351–1355.
Ugent, D., S. Pozorski, and T. Pozorski. 1984. New evidence for ancient cultivation of Canna edulis in Peru. Econ. Bot.
Vega, C.P. 1997. Cultivo de Olluco, p.38–41. IX Congreso Internacional de Cultivos Andinos, Cusco, Peru.
Vivanco, J.M. and H.E. Flores. 2000. Control of root formation by plant growth regulators, p. 1–25. In: A.S. Basra (ed.). Plant growth regulators in agriculture and horticulture: Their role and commercial uses. Food Products Press, an imprint of The Haworth Press, New York.
Vivanco, J.M., L.F. Salazar, and M. Querci. 1999a. Antiviral and antiviroid activity of MAP-containing extracts from Mirabilis jalapa roots. Plant Dis. 83(12):1116–1121.
Vivanco, J.M., B.J. Savary, and H.E. Flores. 1999b. Characterization of two novel type I ribosome-inactivating proteins from the storage roots of the Andean crop Mirabilis expansa. Plant Physiol. 119(4):1447–56.
Wielanek, M. and H. Urbanek. 1999. Glucotropaeolin and myrosinase production in hairy root cultures of Tropaeolum majus. Plant Cell, Tissue and Organ Culture 57:39–45.
Zela, G.M., H.C. Bravo, G. Zela, and V. Gonza. 1997. Cultivo de la maswa. IX Congreso Internacional de Cultivos Andinos, Cusco, Peru, p. 42–51.