Prevalence of *Helicobacter pylori* infection and antibiotic resistance profile in Armenia

Manik Gemilyan1*, Gagik Hakobyan1, Lucie Benejat2, Bujana Allushi2, Davit Melik-Nubaryan1, Harutyun Mangoyan3, Amandine Laur2, Emilie Daguerre2, Eduard Grigoryan4 and Francis Megraud2,5

Abstract

**Background:** The prevalence of *Helicobacter pylori* infection was never assessed in Armenia, nor was the prevalence of *H. pylori* resistance against the main antibiotics concerned, despite the fact that these data are fundamental to establish evidence-based recommendations for management of this infection. We aimed to fill this gap by assessing prevalence of *H. pylori* among adult population in Armenia and resistance of *H. pylori* strains to clarithromycin and levofloxacin.

**Methods:** *Helicobacter pylori* seroprevalence was determined in 217 asymptomatic adult subjects submitted to a health checkup using an ELISA. Molecular methods were used to detect *H. pylori* in gastric biopsies from 91 adult dyspeptic patients [55 (60.4%) were positive] as well as the mutations associated with clarithromycin resistance by real-time PCR and with levofloxacin by sequencing the *gyrA* QRDR.

**Results:** *Helicobacter pylori* seropositivity was found to be 41.5% globally and increased with age from 13.6% (age 18–25 years) to 83.3% (age > 65 years). Only two cases were found with a A2142/43G mutation causing clarithromycin resistance, and 6 cases showed mutations associated with levofloxacin resistance.

**Conclusions:** *Helicobacter pylori* infection is estimated to be about 42% among adults in Armenia and the low clarithromycin resistance allows the use of the standard triple therapy as a first line therapy.

**Keywords:** *Helicobacter pylori*, Seroprevalence, Gastric biopsies, Real-time PCR, Sequencing, Clarithromycin, Levofloxacin

**Background**

*Helicobacter pylori* (*H. pylori*) is a Gram-negative bacterium colonizing the stomach. Infection by *H. pylori* causes gastritis in all infected subjects [1, 2], as well as peptic ulcer disease and gastric cancer in a proportion of infected individuals [3].

The prevalence of *H. pylori* infection varies in different parts of the world, essentially due to the level of development which can partly be explained by promiscuity, low education and lack of hygiene facilities especially water [4]. According to a review that summarized studies involving populations with a large age span, among European countries, the percentage of population infected by *H. pylori* was lowest in Denmark (17%) and Switzerland (19%) and highest (> 80%) in Russia, Portugal and Estonia [5]. The prevalence of *H. pylori* infection in Africa, Asia and Latin America is generally higher than in Europe, and a trend towards decreasing prevalence was noted in several countries between 2009 and 2016 compared to 2000 and 2009 period [6]. However, Armenia has remained a “blank” spot on the *H. pylori* prevalence map, since no data have been published for the country.

Current guidelines on *H. pylori* management suggest to abandon the so-called standard triple therapy including a proton pump inhibitor (PPI) and two antibiotics: clarithromycin and amoxicillin or clarithromycin and metronidazole because of the high rate of clarithromycin resistance (>15%) in most European regions except Northern Europe, or to perform antimicrobial susceptibility testing before starting treatment [2].

---

*Correspondence: mgemilyan@yahoo.co.uk*

1 Yerevan State Medical University, 2 Koryunst, 0025 Yerevan, Armenia
Full list of author information is available at the end of the article
Levofloxacin-containing regimens can be used as a rescue therapy; however, resistance to fluoroquinolones is also frequent and is a major limiting factor for using such schemes. Given that no data were available on *H. pylori* resistance in Armenia, it was crucial to perform a survey to be able to make recommendations for first and second line treatments against this infection.

With current report we aimed to fill this gap by assessing prevalence of *H. pylori* among general adult population in Armenia and resistance of *H. pylori* strains to the main antibiotics known to be produce resistance, i.e. clarithromycin and levofloxacin.

**Methods**

The recruitment of study participants was done during the period of December 12, 2017 to January 11, 2018, in two medical centers: “Vardanants Center for Innovative Medicine” (VMC) and “Armenia Republican Medical Center”. Written, informed consent was obtained from each patient included in the study.

**Determination of *H. pylori* prevalence**

*Helicobacter pylori* prevalence was determined by serology. Adult subjects (age 18 years and older) presenting in VMC for routine health checkup according to their insurance plan were approached by medical volunteers who explained the study protocol and invite them to participate. In all subjects who signed the informed consent, 4 ml blood was drawn in addition to the amount intended for checkup. The blood was immediately centrifuged, and the serum decanted and maintained at +4 °C. At the end of each day the sera were transferred in an ice box and placed in a freezer at −80 °C. After collection of all gastric biopsies, they were transferred frozen by courier service to Bordeaux, France where they were processed.

To detect *H. pylori* and its resistance to clarithromycin, an in-house real-time PCR was used. Briefly after DNA extraction the PCR was run in a LightCycler apparatus with specific primers and labelled probes [8]. For levofloxacin resistance, the Quinolone resistance determining region (QRDR) of the gyrA gene was amplified and sequenced by using the primers:

F-QRDR-Hpylo(GCGTATTTTGATGCGATGC) and R-QRDR-Hpylo(ACAAAAATCAATGGTGTCCTTTATCA).

Cycling conditions consisted of an initial denaturation at 95 °C for 3 min, 40 cycles of denaturation at 95 °C for 30 s, annealing at 58 °C for 30 s and elongation at 72 °C for 30 s, followed by a final extension at 72 °C for 5 min. PCR amplicons were examined by applying 5 µl on a 2% agarose gel and then purified using the ExoSAP-IT (Applied Biosystems, Foster City, CA, USA). Purified PCR products were sequenced with the BigDye Terminator Cycle Sequencing Reaction Kit v3.1 (Applied Biosystems) using the PCR primers as sequencing primers. Sequencing was accomplished with an ABI 3130 Genetic Analyzer (Applied Biosystems), and the obtained results were analysed with DNAbaser software (HeracleBiosoft, Germany).

**Statistics**

Results of analyses were de-identified, entered into an Excel spreadsheet and analyzed using SPSS 20.0 software.

**Results and discussion**

**Prevalence of *H. pylori* infection**

The total number of study participants was 230, with a mean age 39.2 years ±13.0 (range 18–74); 178 (77.4%) were female. Ninety subjects (39.1%) harbored *H. pylori* antibodies while 127 (55.2%) tested negative and 13 (5.7%) were in the grey zone. After eliminating doubtful results, among a total of 217 subjects the rate of positive antibody tests was 41.5%. The distribution of *H. pylori* seropositivity by age is presented in Fig. 1. The frequency of *H. pylori* seropositivity was 39.6% in males and 42.0% in females. The lowest rate (13.6%) was reported in the youngest age group (18–25 years), and the highest rate (83.3%) in the age group over 65 years. The prevalence steadily increased with age (Additional file 1).

It is interesting to see that the prevalence of *H. pylori* infection is less than 40% for those aged 26–45 year, and much lower than 90% for the higher age groups. However, this study has some limits, because a recruitment bias is possible since the individuals were recruited in only one center in the capital city Yerevan, and they
were part of an insurance check-up which may indicate a higher socioeconomic level. The age distribution between the groups was also not balanced. Furthermore, despite the fact that the ELISA used was one of the best, it was not validated in Armenia and so it is possible, although unlikely, that the Armenian strains have a different antigenic profile.

Prevalence of *H. pylori* resistance
The number of study participants providing antral biopsy specimens was 91, 64.8% were female. Analysis of gastric antral tissue specimens revealed that 55 (60.4%) of the patients with dyspeptic symptoms were infected by *H. pylori*. Of these, the mutations associated with clarithromycin resistance (A2142-43G) were found in 2 patients (3.6%), and with fluoroquinolone resistance (mutation N87K or D91Y) in 6 (12.8%), 3 of each (Additional file 2).

Conclusions
These data suggest that Armenia ranks among countries with a low *H. pylori* macrolide resistance rate as well as a medium fluoroquinolone resistance rate. This gives an excellent background to suggest standard triple therapy with PPI, amoxicillin and clarithromycin as a first-line treatment option for *H. pylori* infection in Armenia [2]. However, triple therapy containing levofloxacin should not be offered as a first-line treatment because it is known that resistance to this antibiotic group can develop rapidly and levofloxacin-containing regimens are generally discouraged from being used as a first-line option. Despite the fact that culture and antibiogram could not be carried out, the detection of mutations by molecular methods provides a reliable substitute in which we can be confident [9]. Furthermore resistance against other antibiotics, e.g. amoxicillin, tetracycline and rifabutin, is seldomly found and does not necessitate a routine checkup [10]. Resistance to metronidazole has not been studied in Armenian population. However, if detected in vitro, it can be overcome by a prolonged treatment and adding bismuth to the eradication scheme.

Additional files

Additional file 1. Contains all data on serological study results for *H. pylori* antibodies in serum of 230 subjects.

Additional file 2. Contains all data on macrolide and fluoroquinolone resistance mutation testing in gastric biopsy specimens of 91 patients.

Abbreviations
AFSSAPS: French Health Products Safety Agency; ELISA: enzyme-linked immunosorbent assay; *H. pylori*: *Helicobacter pylori*; PCR: polymerase chain reaction; PPI: proton pump inhibitor; QRDR: quinolone resistance determining region; VMC: Vardanants Center for Innovative Medicine.

Acknowledgements
Not applicable.

Authors’ contributions
MG: study supervision, liaison with France collaborators, writing and submitting manuscript, statistical analysis. GH: study design, ethical approval, data collection, writing manuscript. LB: study design, writing manuscript. BA: performing PCR tests. DM-N: study design, sample transportation to France. HM: data collection, manuscript writing. AL: performing serologic tests. ED: performing serologic tests. EG: subject recruitment, data entry, writing manuscript. FM: study design, writing manuscript. All authors read and approved the final manuscript.

Funding
This work was supported by United European Gastroenterology Activity Grant in the framework of the project “Adapting European guidelines on Helicobacter pylori management and faecal microbiota transplantation to use in Armenia as a model for low-resource countries”.

Availability of data and materials
All data generated or analysed during this study are included in this published article and its Additional files.

Ethics approval and consent to participate
The study protocol conforms to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in a priori approval by the Ethical Committee of the National Center for AIDS Prevention on 11 December, 2017.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

Author details
1 Yerevan State Medical University, 2 Koryunst, 0025 Yerevan, Armenia. 2 French National Reference Centre for Helicobacters, Bacteriology laboratory, Pellegrin Hospital, Bordeaux, France. 3 Vardanants Center for Innovative Medicine, Vardanants 18/1, 0010 Yerevan, Armenia. 4 Armenian Association of Gastroenterology, 5B Mashtotsave, 0015 Yerevan, Armenia. 5 INSERM 1053, University of Bordeaux, Bordeaux, France.

Received: 7 January 2019 Accepted: 29 May 2019
Published online: 08 June 2019
References

1. Correa P, Piazuelo MB. Natural history of Helicobacter pylori infection. Dig Liver Dis. 2008;40:490–6.
2. Malfertheiner P, Megraud F, O’Morain CA, Gisbert JP, Kuipers EJ, et al. Management of Helicobacter pylori infection—the Maastricht V/Florence consensus report. Gut. 2017;66:6–30.
3. Schistosomes, liver flukes and Helicobacter pylori. IARC Working Group on the evaluation of carcinogenic risks to humans. Lyon, 7–14 June 1994. IARC Monogr Eval Carcinog Risks Hum. 1994;61:1–241.
4. Hunt RH, Xiao SD, Megraud F, Leon-Barua R, Bazzoli F, et al. Helicobacter pylori in developing countries. World Gastroenterology Organisation global guideline. J Gastrointest Liver Dis. 2011;20:299–304.
5. Roberts SE, Morrison-Rees S, Samuel DG, Thorne K, Akbari A, et al. Review article: the prevalence of Helicobacter pylori and the incidence of gastric cancer across Europe. Aliment Pharmacol Ther. 2016;43:334–45.
6. Zamani M, Ebrahimtabar F, Zamani V, Miller WH, Alizadeh-Navaei R, et al. Systematic review with meta-analysis: the worldwide prevalence of Helicobacter pylori infection. Aliment Pharmacol Ther. 2018;47:868–76.
7. Burucoa C, Delchier J-C, Courillon-Mallet A, de Korwin J-D, Méraud F, et al. Comparative evaluation of 29 commercial Helicobacter pylori serological kits. Helicobacter. 2013;18:169–79.
8. Oleastro M, Ménard A, Santos A, Lamoulisatte H, Monteiro L, et al. Real-time PCR assay for rapid and accurate detection of point mutations conferring resistance to clarithromycin in Helicobacter pylori. J Clin Microbiol. 2003;41:397–402.
9. Ducournau A, Bénéjat L, Sifré E, Bessède E, Lehours P, et al. Helicobacter pylori resistance to antibiotics in 2014 in France detected by phenotypic and genotypic methods. Clin Microbiol Infect. 2016;22:715–8.
10. Megraud F, Coenen S, Versporten A, Kist M, Lopez-Brea M, et al. Helicobacter pylori resistance to antibiotics in Europe and its relationship to antibiotic consumption. Gut. 2013;62:34–42.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.