Bioinformatics Analyses of the Role of Vascular Endothelial Growth Factor in Patients with Non-Small Cell Lung Cancer

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Abstract

Purpose
This study was aimed to identify the expression pattern of vascular endothelial growth factor (VEGF) in non-small cell lung cancer (NSCLC) and to explore its potential correlation with the progression of NSCLC.

Methods
Gene expression profile GSE39345 was downloaded from the Gene Expression Omnibus database. Twenty healthy controls and 32 NSCLC samples before chemotherapy were analyzed to identify the differentially expressed genes (DEGs). Then pathway enrichment analysis of the DEGs was performed and protein-protein interaction networks were constructed. Particularly, VEGF genes and the VEGF signaling pathway were analyzed. The sub-network was constructed followed by functional enrichment analysis.

Results
Total 1666 up-regulated and 1542 down-regulated DEGs were identified. The down-regulated DEGs were mainly enriched in the pathways associated with cancer. VEGFA and VEGFB were found to be the initiating factor of VEGF signaling pathway. In addition, in the epidermal growth factor receptor (EGFR), VEGFA and VEGFB associated sub-network, kinase insert domain receptor (KDR), fibronectin 1 (FN1), transforming growth factor beta induced (TGFBI) and proliferating cell nuclear antigen (PCNA) were found to interact with at least two of the three hub genes. The DEGs in this sub-network were mainly enriched in Gene Ontology terms related to cell proliferation.
Conclusion

EGFR, KDR, FN1, TGFBI and PCNA may interact with VEGFA to play important roles in NSCLC tumorigenesis. These genes and corresponding proteins may have the potential to be used as the targets for either diagnosis or treatment of patients with NSCLC.

Introduction

Lung cancer ranks highest in both morbidity and mortality in most parts of the world [1, 2], and its absolute incidence is increasing dramatically [3]. Lung cancer can be categorized mainly into small cell and non-small cell histological subtypes. Among them, non-small cell lung cancer (NSCLC) is the most common form and accounts for almost 75% to 80% of lung cancer [2, 4]. Currently, about 70% newly diagnosed patients with either subtype of lung cancer suffer from local recurrence or metastatic lesions after resection, resulting in poor long-term survival rate [5]. Therefore, it is important to elucidate the mechanisms of lung cancer progression for the effective treatment of the disease.

Various studies have demonstrated that angiogenesis is essential for NSCLC growth and metastasis [6–8]. Vascular endothelial growth factor (VEGF), an angiogenic specific stimulator, has been found to regulate the growth of neoplastic angiogenesis and plays an important role in vascularization in different types of cancers [9, 10]. Bergers and Benjamin found that VEGF were highly expressed in the tumor microenvironment and strongly induced tumor angiogenesis [11]. Zhao et al. [12] suggested that angiogenesis and VEGF expression in lung cancer cells was attributed to increased activity of JAK2/STAT3 signal pathway which was associated with a decreased survival rate in cancers. In addition, Chen et al. [13] reported that the over-expression of VEGF was a poor prognostic factor in small-cell lung cancer. Therefore, strategies to inhibit tumor-associated angiogenesis may be promising in limiting NSCLC progression.

Despite progresses achieved in the management of NSCLC, identification of specific molecular criteria is still a challenge to increase response rates [14]. In the present study, we analyzed the gene expression profiles (GSE39345) of peripheral blood mononuclear cells (PBMC) in patients with advanced stage NSCLC. The differentially expressed genes (DEGs) between healthy controls and patients before chemotherapy were analyzed. To the best of our knowledge, the dataset has not been analyzed before. Particularly, we evaluated VEGF genes expression and analyzed the VEGF signaling pathway. The VEGFA sub-network was constructed and functional enrichment analysis were performed with its related DEGs. We aimed to identify the expression pattern of VEGF in NSCLC and to explore its potential correlation with the progression of NSCLC. Unlike previous studies that explored the function of VEGF through experimental methods, the bioinformatics analyses performed in study provided a comprehensive evaluation of VEGF related protein-protein interactions and could be used to predict the interaction relationships between VEGF and other genes.

Materials and Methods

Affymetrix Microarray Data

The microarray data GSE39345 used in our study was downloaded from the Gene Expression Omnibus (http://www.ncbi.nlm.nih.gov/geo/) database. This dataset analyzed the gene expression profiles of PBMC in patients with advanced stage NSCLC based on the platform of Illumina humanRef-8 v2.0 expression beadchip (GPL1604) (Affymetrix Inc., Santa Clara,
Californi, USA). The gene expression profiles consisted of 20 healthy controls (HC), 32 patients before chemotherapy and 17 patients after chemotherapy. In this study, the datasets from 20 HC and 32 NSCLC samples before chemotherapy were analyzed. The study was approved by the Institutional Review Board of Kaohsiung Chang Gung Memorial Hospital, Taiwan. Samples were collected after informed consent had been obtained from the patients. The patient records were de-identified prior to analysis [15].

Data Preprocessing and Differential Expression Analysis

The original array data were performed background correction and quartile data normalization. Then the DEGs between HC and NSCLC samples were identified based on the R/Bioconductor package limma [16]. The absolute value of log₂-fold change (log₂FC) ≥ 1.5 and p-value < 0.05 were considered as cutoff value.

Pathway Enrichment Analysis

Kyoto Encyclopedia of Genes and Genomes (KEGG, http://www.genome.jp/) [17] is a collection of online database composed by the known genes and their biochemical functionalities. The Database for Annotation, Visualization and Integrated Discovery (DAVID, http://david.abcc.ncifcrf.gov/) [18] is a comprehensive set of functional annotation tool for relating the functional terms with gene lists by clustering algorithm. In order to analyze the DEGs in functional level, KEGG pathway enrichment analysis was performed using the DAVID online tool. The p-value < 0.05 was set as the threshold value.

VEGF Genes and VEGF Signaling Pathway

VEGF family members play important roles in the progression of NSCLC. In the present study, the distribution of DEGs in VEGF signaling pathway was studied using the KEGGParser [19] plugin for cytoscape (www.cytoscape.org) [20].

Protein-protein Interaction Network Construction

We downloaded the comprehensive interaction information of human proteins from the Search Tool for the Retrieval of Interacting Genes (STRING) database (http://string-db.org/) [21]. Then the interaction relationships of NSCLC DEGs were extracted to construct the protein-protein interaction (PPI) network (combined score > 0.4) using cytoscape.

Sub-network Construction

Study has found that the VEGFR and EGFR pathways are linked in solid tumors, particularly with respect to angiogenesis [22]. In our study, we extracted EGFR, VEGFA and VEGFB, and their interactions with the other DEGs from the PPI network. Then the sub-network associated with EGFR, and VEGF was constructed. In addition, the DEGs in the sub-network were performed Gene Ontology (GO) enrichment analysis using the BiNGO [23] plugin for cytoscape (p-value < 0.05).

Results

Identification of DEGs

In total, 3028 DEGs were obtained from both NSCLC and HC samples. Thereinto, 1666 were up-regulated and 1542 were down-regulated.
Pathway Enrichment Analysis
The result of pathway enrichment showed that the up-regulated DEGs were mainly enriched in 11 pathways, such as olfactory transduction, metabolism of xenobiotics by cytochrome P450 and drug metabolism. The down-regulated DEGs were mainly enriched in the pathways associated with cancer, such as Wnt signaling pathway and Notch signaling pathway.

VEGF genes and VEGF Signaling Pathway
Among the DEGs, VEGFA and VEGFB were found to be the initiating factors of VEGF signaling pathway. After analysis of the distribution of DEGs in VEGF signaling pathway, down-regulated DEG of kinase insert domain receptor (KDR) was found to be a bottle neck factor of the pathway (Fig 1).

PPI Network Analysis
The constructed PPI network contained 113797 interaction pairs between 2511 DEGs and the PPI networks obeyed the scale-free attribution (correlation = 0.859, R-squared = 0.888). The top 10 hub genes were shown in Table 1, including jun proto-oncogene, proliferating cell nuclear antigen (PCNA), epidermal growth factor receptor (EGFR) and VEGFA.

Sub-network Construction
The EGFR and VEGF associated sub-network was shown in Fig 2. In the sub-network, KDR and fibronectin 1 (FN1) interact with all of the three genes. Transforming growth factor beta (TGFBI) and PCNA interact with EGFR and VEGFA. Additionally, the DEGs in this sub-network were mainly enriched in the GO biological process terms related to cell proliferation (Table 2).

Discussion
Highly treatment-resistant NSCLC is one of the major causes of cancer death all over the world and angiogenesis has become an integral process in promoting the growth and metastasis of NSCLC. As a result, therapy against VEGF, one of the major mediators of angiogenesis, may be an critical approach for regressing NSCLC [6]. In this study, VEGFA and VEGFB were found to be the initiating factors of VEGF signaling pathway. In addition, KDR was found to be a bottle neck factor of this pathway. Moreover, in the EGFR and VEGF associated sub-network, KDR, FN1, TGFBI and PCNA were found to interact with at least two of the three hub genes.

VEGF gene belongs to the platelet derived growth factor/VEGF growth factor family. Its protein product promotes endothelial cell proliferation and migration, and prevents cell apoptosis. Importantly, it is also involved in tumor-associated angiogenesis [24]. In our study, VEGFA was found to be the initiating factor of VEGF signaling pathway. Besides, it was a hub gene in the PPI network. Deng et al. [25] has reported that VEGFA is implicated in the carcinogenesis of numerous cancers including lung cancer. The VEGFA expression has been suggested to be related to the relapse and poor prognosis of NSCLC [26]. Specially, in NSCLC, the tumor progression and metastasis are considered to be associated with the mutations of some tumor suppressor genes, such as p53, and the mutation of p53 induces VEGF gene expression [27]. Interestingly, KDR, the receptor of VEGF, was found to be down-regulated in the VEGF signaling pathway, acting as a bottle-neck factor of this pathway. Thus, we speculate that KDR may negatively regulate VEGF signaling pathway. Taken together, VEGFA and KDR may be potentially involved in the progression of NSCLC.
Currently, many drugs that target either VEGFR or EGFR signaling pathways in advanced NSCLC have been clinically validated [28]. EGFR, a member of the epidermal growth factor family, plays diverse biological roles in promoting malignancy including regulation of cell survival or apoptosis, and cell motility or metastasis, which makes EGFR an attractive therapeutic target.

**Fig 1. The differentially expressed genes (DEGs) distribution in vascular endothelial growth factor (VEGF) signaling pathway.** Red color represents up-regulated DEGs and green represents down-regulated DEGs.

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**Table 1. The top 10 hub genes in the protein-protein interaction network.**

| Node   | DE_state | Degree |
|--------|----------|--------|
| JUN    | -1       | 140    |
| PCNA   | -1       | 134    |
| EGFR   | 1        | 128    |
| CD4    | 1        | 127    |
| VEGFA  | 1        | 122    |
| IL6    | -1       | 117    |
| MAPK1  | -1       | 117    |
| CTNNB1 | -1       | 115    |
| GAPDH  | 1        | 109    |
| PIK3R1 | -1       | 107    |

DE_state represents differential expression state; 1 represents up-regulated differentially expressed genes (DEGs); -1 represents down-regulated DEGs.

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**Fig 2.** The vascular endothelial growth factor (VEGF) and epidermal growth factor receptor (EGFR) associated sub-network. Red color represents up-regulated differentially expressed genes (DEGs) and green represents down-regulated DEGs.

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**Table 2.** The top 10 significant Gene Ontology (GO) biological process terms enriched by the differentially expressed genes in sub-network.

| GO-ID  | Corrected p-value | Description                                      |
|--------|-------------------|--------------------------------------------------|
| 42127  | 7.95E-28          | regulation of cell proliferation                 |
| 8284   | 4.60E-27          | positive regulation of cell proliferation        |
| 48522  | 6.95E-27          | positive regulation of cellular process          |
| 48518  | 2.83E-25          | positive regulation of biological process        |
| 42221  | 9.97E-23          | response to chemical stimulus                    |
| 48513  | 8.15E-22          | organ development                                 |
| 51272  | 2.15E-21          | positive regulation of cellular component movement|
| 48856  | 2.15E-21          | anatomical structure development                 |
| 48731  | 2.15E-21          | system development                                |
| 50793  | 9.28E-21          | regulation of developmental process              |

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target [29, 30]. De Luca et al. [31] reported that activation of EGFR by TGF-α can increase the production of VEGF in human cancer cells.

In this study, we also found that EGFR was also involved in the VEGF and EGFR associated sub-network, in addition to the GO terms related to cell proliferation. In this sub-network, KDR, FN1, TGFBI and PCNA were found to interact with 2 or 3 of the 3 hub genes. FN1 encodes fibronectin, a glycoprotein in extracellular matrix (ECM), which has been implicated in the development of many types of cancer. In lung cancer, fibronectin can promote lung cancer cell migration by activating FAK signaling [32]. TGFBI is an extracellular matrix (ECM) protein induced by transforming growth factor-β (TGF-β) in human cancer cells [33]. TGFBI has been discovered as a gene induced in the lung cancer cell line A549 by TGF-β [34]. Park et al. [35] has reported that TGFBI stimulates adhesion, proliferation and migration of renal proximal tubular epithelial cells. TGFBI can sustain cell adhesion to substrates, which may increase cell motility and metastases [36]. In addition, TGFBI-derived proteolytic peptides are pro-apoptotic in several cell types, which may be due to the activation of cell-specific proteases capable of degrading TGFBI [37]. Therefore, TGFBI may be used as a potential target for NSCLC treatment.

Furthermore, the down-regulated DEG of PCNA was identified in the PPI network and sub-network. In normal cells, the proliferation and apoptosis are under the control of cell cycle regulatory systems. The uncontrolled cell proliferation is considered to be the hallmark of cancer cells, thus, the proliferative potential of cancer cells may be an important prognostic factor [38]. Some antibodies including PCNA have been used to investigate the cell proliferation [39]. Zienolddiny et al. [40] has reported that PCNA is a marker for the evaluation of cell proliferative activity in lung cancer. Therefore, PCNA may participate in the regulation network of VEGFA to play an important role in NSCLC tumorigenesis and serve as a potential molecular marker associated with NSCLC.

In conclusion, our study provides a comprehensive bioinformatics analysis of the expression of VEGF in NSCLC. DEGs such as VEGFA, VEGFB, EGFR, FN1, TGFBI, KDR and PCNA. Their related GO terms may play important roles in NSCLC tumorigenesis and progression and have the potential to be used as targets for NSCLC diagnosis and treatment. Further genetic and experimental studies with larger sample size are needed to confirm our results.

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Author Contributions
Conceived and designed the experiments: SW. Performed the experiments: YW LH YJ YY LL AB MF. Analyzed the data: YW LH SW. Wrote the paper: YW LH SW.

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