Angiotensin(1-7) and ACE2, “The Hot Spots” of Renin-Angiotensin System, Detected in the Human Aqueous Humor

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Abstract: Background: The main purpose of the study was to establish whether essential components of the renin-angiotensin system (RAS) exist in the human aqueous humor.

Methods: Forty-five patients ≥ 60 (74±7) years of age undergoing cataract surgery at Tampere University Hospital were randomly selected for the prospective study. The exclusion criterion was the use of oral antihypertensive medicine acting via renin-angiotensin system. Aqueous humor samples were taken at the beginning of normal cataract extraction. The samples were frozen and stored at -80 °C. The concentrations of intraocular endogenous RAS components Ang(1-7), ACE2, and ACE1 were measured using ELISA.

Results: Concentration medians of Ang(1-7), ACE2, and ACE1 in the aqueous humor were: Ang(1-7) 4.08 ng/ml, ACE2 2.32 ng/ml and ACE1 0.35 ng/ml. The concentrations were significantly higher in glaucomatous than in non-glaucomatous eyes, ACE1 (p=0.014) and Ang(1-7) (p=0.026) vs non-glaucomatous eyes.

Conclusions: Ang(1-7), ACE2 and ACE1 are found in the human aqueous humor. The observations are consistent with the conception that local tissue-RAS exists in the human eye and it might have a role in the control of intraocular pressure.

Keywords: Angiotensin (1-7), angiotensin converting enzyme 2, angiotensin II, angiotensin converting enzyme 1, aqueous humor, glaucoma, renin-angiotensin system.

INTRODUCTION

The systemic renin-angiotensin system (RAS) controls fluid volume, electrolyte balance and blood pressure (BP) homeostasis [1]. RAS is also regarded as a tissue-specific regulatory system accounting for local effects and long-term changes in different organs [2, 3]. Many peptides and enzymes of RAS have already been detected in the human eye [2, 4, 5] and they are even suggested to have a role in the pathogenesis of different ocular diseases including glaucoma [6]. One of the major known risk factors for glaucoma is increased intraocular pressure (IOP) [7-9] which is a net sum of homeostatic balance between aqueous humor formation and outflow.

It has recently been reported that orally administered antihypertensive drugs can also reduce IOP [10]; for example, oral angiotensin converting enzyme (ACE) inhibitor (captopril) [11] and the AT1- receptor blocker (ARB) [12] (losartan) have been shown to lower IOP in both non-glaucomatous and glaucomatous patients. In animal studies, ACE inhibitors [13, 14] ARBs [15, 16], renin inhibitors [17] and angiotensin (1-7) [18] have been reported to lower IOP, even when they are locally administered. These findings imply that a local intraocular RAS may be involved in the regulation of IOP [19, 20]. Recently, other broader theories have been published on RAS involvement in the pathogenesis of glaucoma [6, 21]. So far, only AngII and ACE1 have been identified in the human aqueous humor [6, 22]. RAS is known to consist of over twenty peptidases, close to twenty angiotensin peptides, and at least six receptors [23]. AngII-ACE1-angiotensin 1 receptor type (AT1R)-axis together with Ang(1-7)-ACE2-Mas-receptor (MasR)-axis are seen as the main pathways of RAS that may mediate therapeutic benefits. The main purpose of this study was to detect the levels of Ang(1-7), ACE2 and ACE1 qualitatively and quantitatively in the human aqueous humor.

MATERIALS AND METHODOLOGY

The study was undertaken in the Department of Ophthalmology in the University Hospital of Tampere, Finland between 28th February and 8th April 2014. The study conforms to the World Medical Association Declaration of Helsinki and the Ethical Principles for Medical Research Involving Human Subjects, and received approval from the Regional Ethics Committee at Tampere (ETL R14010). Informed consent was obtained from all participants. Patients over 60 years of age undergoing cataract surgery were included. The patients were selected randomly from the cataract operation list and they were operated upon by the same surgeon. The only exclusion criterion was the use of oral antihypertensive medicine acting via renin-angiotensin system. Together, 45 patients were operated upon; 15 of...
them had been diagnosed with glaucoma according to Finnish Evidence-Based Guidelines [9]. All the glaucomatous patients were ocular normotensive because of continuous antiglaucoma medication or previous glaucoma surgery. Thus all the eyes independently of medication were normotensive, and the division to glaucomatous/non-glaucomatous was based on the history of the patients. Medical records (age, gender, medications and IOP) were registered during the preoperative visit. IOP was measured using a rebound tonometer (Icare®, Icare Finland Oy, Vantaa, Finland). Blood pressure was measured three consecutive times prior to surgery in sitting position and the average of the measurements was calculated.

An additional eleven patients were operated upon; six of them were taking oral ACE inhibitor medication for high blood pressure and five of them were on ARB medication. These patients were not included in the main study results, but their anterior chamber RAS component concentrations are shown in Fig. (1).

Aqueous humor samples (0.05-0.35 ml) were taken in the beginning of routine cataract surgery. The liquid was collected into Eppendorf tubes and immediately chilled in an icebox. After the operation, the collected samples were frozen and stored at -80°C within 2 h. No protease/peptidase inhibitors were added in the collection and storage of the samples. The samples were analyzed using commercially available enzyme-linked immunosorbent assays (ELISA) due to their applicability to simultaneously assess the target molecule concentrations in a large number of samples. Ang(1-7) was measured using the Human Angiotensin(1-7) Elisa kit (MyBioSource, San Diego, CA, USA) with a detection limit of 0.1 ng/ml. ACE2 was measured using the Human ACE2 ELISA kit (Boster Immunoleader, Pleasanton, CA, USA) with a detection limit of <10 pg/ml, ACE1 using the Human ACE ELISA kit (Boster Immunoleader) with a detection limit of <5 pg/ml. AngII was also measured using the Angiotensin II ELISA kit (Enzo Life Sciences, Farmingdale, NY, USA) with a detection limit of 4.6 pg/ml. The kits were designed for usage with human serum, plasma, cell culture supernates, body fluid or tissue homogenates. The assay procedures and assays were conducted according to manufacturers’ instructions. Only Angiotensin II ELISA kit has been shown to have cross-reactivity, but it is of no relevance. In other ELISA kits cross-reactions were negligible. All assays were done blinded. Results are shown in ng/ml.

The data from the ELISA assays were analyzed using Microsoft Excel and SPSS Statistics for Windows ver. 21.0. (IBM Corp, Armonk, NY, USA). Nonparametric Mann-Whitney test and Spearman’s Correlation were used for data with skewed distribution and the results are shown as median with upper and lower quartiles, whereas independent sample t-test and Pearson’s Correlation were used for normally distributed data. The results are shown as mean ±SD. The level of significance was set at <0.05 (two-tailed) in all statistical tests.

RESULTS

Forty-six aqueous humor samples were analyzed from 45 subjects with an average age of 74±7 (mean ± SD) years. Of these, 27 (60 %) were female and 18 (40 %) male. Fifteen were diagnosed with glaucoma and 30 were non-glaucomatous. Overall demographics of the glaucomatous and non-glaucomatous subgroups are presented in Table 1.

Table 1. Demographics and intracameral concentrations of Ang(1-7), ACE2 and ACE1. For normally distributed data results are shown as mean ±SD, for skewly distributed data median with upper and lower quartiles. The small volume of some samples did not allow all measurements.

|                | Non-Glaucomatous (n=30) | Glaucomatous (n=15) | p-Value |
|----------------|-------------------------|---------------------|---------|
| Age            | 74 ± 7                  | 75 ± 8              | 0.843   |
| Gender (M/W)   | 10/20                   | 8/7                 |         |
| IOP            | 15 ± 4                  | 16 ± 6              | 0.658   |
| Systolic BP    | 163 ± 21                | 166 ± 26            | 0.692   |
| Diastolic BP   | 88 ± 10                 | 92 ± 11             | 0.335   |
| Ang(1-7)       | (3.60-4.59)             | (4.03-6.21)         | 0.026*  |
| (n=31)         | (n=15)                  |                     |         |
| ACE2           | 2.26                    | 2.96                | 0.409   |
| (1.19-6.02)    | (1.91-12.33)            |                     |         |
| (n=20)         | (n=14)                  |                     |         |
| ACE1           | 0.27                    | 0.48                | 0.014*  |
| (0.23-0.39)    | (0.33-0.79)             |                     |         |
| (n=20)         | (n=12)                  |                     |         |

Number of patients: n=30, number of eyes: n=31. * p-value <0.05. Ang(1-7), angiotensin (1-7); ACE1, -2, angiotensin-converting enzyme 1, -2; BP, blood pressure; IOP, intraocular pressure; M, men; W, women.

Median aqueous humor (n=46) Ang(1-7), ACE2 and ACE1 concentrations were: 4.08 ng/ml (Q1-Q3; 4.00-4.92), 2.32 ng/ml (Q1-Q3; 2.58-7.53) and 0.35 ng/ml (Q1-Q3; 0.30-0.51), respectively. None of the samples showed measurable levels of AngII. The Ang(1-7) and ACE1 concentrations were significantly higher in glaucomatous than in non-glaucomatous eyes (p=0.026 and p=0.014). See Table 1. Individual aqueous humor concentrations of Ang(1-7), ACE2 and ACE1 in non-glaucomatous and glaucomatous patients are presented in Fig. (1).

Age, IOP- and BP-values did not differ between the two subgroups (Table 1). With one exception, no significant differences between men and women were found in Ang(1-7), ACE2 or ACE1 concentrations in the subgroups (Table 2). The significant correlations were in the non-glaucomatous group ACE1 vs age and ACE1 vs ACE2 both genders together. In glaucomatous patients no correlations were found.

Glaucoma patients (n=15) used different topically administered antiglaucoma drugs. Patients who used prostaglandin analogues had higher Ang(1-7) and ACE1 concentrations vs patients with no medication (p=0.012 and p=0.028 respectively). Also, the use of a combination of beta blocker and carbonic anhydrase inhibitor was associated with higher ACE1 concentration (p=0.035). No statistically
significant difference was detected between patients using other glaucoma medications.

DISCUSSION

The present study was aimed to detect the central components of RAS in the human aqueous humor. In addition possible differences between glaucomatous and non-glaucomatous eyes were studied. We showed that endogenous Ang(1-7) and ACE1 and ACE2 are present in the aqueous humor of the human eye. The glaucomatous eyes have higher levels of ACE1 vs non-glaucomatous eyes. This would suggest a role of ACE1 in IOP balance in the development of glaucoma. Furthermore, ACE1 levels were associated with higher ACE2 concentrations in non-glaucomatous eyes, this offsetting each other’s effects on IOP. Interestingly, in the glaucomatous subjects age did not correlate with the aqueous humor concentrations of any of the RAS components, while in non-glaucomatous eyes increasing age was with higher ACE1 concentrations. Aqueous humor Ang(1-7), ACE1 and ACE2 levels did not differ between the genders in either of the subgroups. BP and IOP values were not associated with higher RAS component concentrations. High BP values measured just before surgery were likely to result from the anxiety and fear that patients usually feel prior to an operation. On the other hand, all glaucomatous patients were under treatment and therefore had normal IOP values.

The lack of measurable levels of AngII in aqueous humor samples may be explained by the absence of protease/peptidase inhibitors in the collection and storage of the samples. Thus AngI (DRVYIHPFH-L) and AngII (DRVYIHPF) peptides can be cleaved to shorter angiotensin peptides [24]; to AngIII (RVYIHPF) or Ang(1-9) (DRVYIHFH) or Ang(1-7) (DRVYIHP) [2]. For example, prolyl endopeptidase and prolyl carboxypeptidase can hydrolyze AngII to Ang(1-7). These alternative pathways of angiotensin degradation system are possible. In previous studies [4] intraocular AngII detection measurements were performed in pools consisting of different samples. It is also possible that the sensitivity of the used AngII assay was too weak in this study. Unfortunately, there is no reference to literature showing a rigorous assessment of these kits. Usually, ELISA methods are qualitative and quantitative but they need highly specific and sensitive antibodies. Because the sensitivity of the assays is strictly limited by the affinity between antibodies, peptides, and proteins, it is not possible in practice to accurately validate or tune assays.

Interestingly, subjects who used prostaglandin analogues as glaucoma medication had higher Ang(1-7) and ACE1 concentrations. In addition, the use of a combination of beta
Table 2. Correlations between RAS components (ng/ml), sex, age, IOP and BP. The small volume of some samples did not allow all measurements. For abbreviations see Table 1.

|                  | Non-Glaucomatous |                     | Glaucomatous |                     |
|------------------|------------------|---------------------|--------------|---------------------|
|                  | Ang(1-7) n = 31  | ACE2 n = 20         | Ang(1-7) n = 15 | ACE2 n = 14         |
|                  | ACE1 n = 20      |                     | ACE1 n = 12   |                     |
| ACE2             | p-value          | 0.156               | 0.077        | 0.347               |
|                  | Ce <i>           | 0.330               | 0.100        | 0.947               |
| ACE1             | p-value          | 0.361               | 0.025*       |                     |
|                  | Ce <i>           | 0.216               | 0.595        | -0.298              |
| Gender           |                  |                     |              |                     |
| Women            | p-value          | 0.210               | 0.119        | 0.052               |
|                  | Ce <i>           | 0.259               | -0.011       | 0.901               |
| Men              | p-value          | 0.128               | 0.960        | 0.027*              |
|                  | Ce <i>           | 0.259               | -0.011       | 0.901               |
| Age              | p-value          | 0.338               | 0.897        | 0.600               |
|                  | Ce <i>           | -0.178              | -0.029       | -0.125              |
| IOP              | p-value          | 0.823               | 0.875        | 0.502               |
|                  | Ce <i>           | -0.042              | -0.036       | 0.160               |
| Systolic BP      | p-value          | 0.769               | 0.452        | 0.332               |
|                  | Ce <i>           | -0.055              | 0.169        | -0.229              |
| Diastolic BP     | p-value          |                     |              |                     |
|                  | Ce <i>           |                     |              |                     |

Cc: correlation coefficient.
*p-value < 0.05.

blocker + carbonic anhydrase inhibitor was associated with higher ACE1 concentrations. Due to the limited number of patients using glaucoma medications in the present study, these observations require further confirmation.

CONCLUSION

Angiotensin(1-7) and ACE2, the “hot spots” in the renin-angiotensin system, are found in the human aqueous humor. This supports the assumption that intraocular RAS may be involved in the regulation of IOP. This theory is further strongly supported by our very recent observation [25] on the expression of Mas-receptors in the retina and especially in the anterior part of the human eye.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

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REFERENCES

[1] Fyhrquist F, Saijonmaa O. Renin-angiotensin system revisited. J Intern Med 2008; 264: 224-36.
[2] Paul M, Poyan Mehr A, Kreutz R. Physiology of local renin angiotensin systems. Rev Physiol Rev 2006; 86: 747-803.
[3] Kramkowski K, Mogiehniki A, Buczek W. The physiological significance of the alternative pathways of angiotensin II production. J Physiol Pharmacol 2006; 57: 529-39.
[4] Damsen AHJ, Derks FHM, Adriaan P1 et al. Angiotensin levels in the eye. Invest Ophthalmol Vis Sci 1994; 35: 1008-18.
[5] Savaskan E, Loffler KU, Meier F, et al. Immunohistochemical localization of angiotensin-converting enzyme, angiotensin II and AT1 receptor in human ocular tissues. Ophthalmic Res 2004; 36: 312-20.
[6] Giese MJ, Speth RC. The ocular renin-angiotensin system: A therapeutic target for the treatment of ocular disease. Pharmacol Ther 2014; 142: 11-32.
[7] Wu SY, Nemesure B, Leske MC. Observed versus indirect estimates of incidence of open-angle glaucoma. Am J Epidemiol 2001; 15: 184-7.
[8] Kroese M, Burton H, Vardy S, Rimmer T, McCarter D. Prevalence of primary open angle glaucoma in general ophthalmic practice in the United Kingdom. Br J Ophthalmol 2002; 86: 978-80.
[9] Tuulonen A, Airaksinen PJ, Erola E, et al. The Finnish evidence-based guideline for open-angle glaucoma. Acta Ophthalmol Scand 2003; 81: 3-18.
[10] Khawaja AP, Chan MP, Broadway DC, et al. Systemic medication and intraocular pressure in a British population: the EPIC-Norfolk Eye Study. Ophthalmology 2014; 121: 1501-7.
[11] Costagliola C, Di Benedetto R, De Caprio L, Verde R, Mastropasqua L. Effect of oral captopril (SQ 14225) on intraocular pressure in man. Eur J Ophthalmol 1995; 5: 19-25.
[12] Costagliola C, Verolino M, de Rosa ML, et al. Effect of oral losartan potassium on intraocular pressure in normotensive and glaucomatous human subjects. Exp Eye Res 2000; 71: 167-71.
[13] Watkins RW, Baum T, Cedeno K, et al. Topical ocular hypotensive effects of the novel angiotensin converting enzyme inhibitor SCH 33861 in conscious rabbits. J Ocul Pharmacol 1987; 3: 295-307.
[14] Shah GB, Sharma S, Mehta AA, Goyal RK. Oculohypotensive effect of angiotensin-converting enzyme inhibitors in acute and chronic models of glaucoma. J Cardiovasc Pharmacol 2000; 36: 169-75.
[15] Wang RF, Podos SM, Mittag TW, Yokoyoma T. Effect of CS-088, an angiotensin AT1 receptor antagonist, on intraocular pressure in glaucomatous monkey eyes. Exp Eye Res 2005; 80: 629-32.

[16] Inoue T, Yokoyoma T, Mori Y, et al. The effect of topical CS-088, an angiotensin AT1 receptor antagonist, on intraocular pressure and aqueous humor dynamics in rabbits. Curr Eye Res 2001; 23: 133-8.

[17] Giardina WJ, Kleinert HD, Ebert DM, et al. Intraocular pressure lowering effects of the renin inhibitor ABBOTT-64662 diacetate in animals. J Ocul Pharmacol 1990; 6: 75-83.

[18] Vaajanen A, Vapaatalo H, Kautiainen H, Oksala O. Angiotensin (1-7) reduces intraocular pressure in the normotensive rabbit eye. Invest Ophthalmol Vis Sci 2008; 49: 2557-62.

[19] Vaajanen A, Lahtila S, Oksala O, Vapaatalo H. Does the renin-angiotensin system also regulate intraocular pressure? Ann Med 2008; 40: 418-27.

[20] Vaajanen A, Vapaatalo H. Local ocular renin-angiotensin system - a target for glaucoma therapy? Basic Clin Pharmacol Toxicol 2011; 109: 217-24.

[21] Kuchtey J, Kuchtey RW. The microfibril hypothesis of glaucoma: implications for treatment of elevated intraocular pressure. J Ocul Pharmacol Ther 2014; 30: 170-80.

[22] Osusky R, Nussberger J, Ansmutz C, Flammer J, Brunner HR. Individual measurements of angiotensin II concentrations in aqueous humor of the eye. Eur J Ophthalmol 1994; 4: 228-33.

[23] Ferrão FM, Lara LS, Lowe J. Renin-angiotensin system in the kidney: What is new? World J Nephrol 2014; 3: 64-76.

[24] Shiota N, Saegusa Y, Nishimura K, Miyazaki M. Angiotensin II-generating system in dog and monkey ocular tissues. Clin Exp Pharmacol Physiol 1997; 24: 243-8.

[25] Vaajanen A, Kalesnykas G, Vapaatalo H, Uusitalo H. The expression of Mas-receptor of the renin-angiotensin system in the human eye. Graefes Arch Clin Exp Ophthalmol 2015 [Epub ahead of print].

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