Research Article

Black Wattle Tannin As Steel Corrosion Inhibitor

Rafael Silveira Peres,1 Eduardo Cassel,2 and Denise Schermann Azambuja1

1 Laboratório de Eletroquímica, Instituto de Química, UFRGS, Avenida Bento Gonçalves 9500, 90619-900 Porto Alegre, RS, Brazil
2 Departamento de Engenharia Química, Faculdade de Engenharia, PUCRS, Avenida Ipiranga 6681, 90619-900 Porto Alegre, RS, Brazil

Correspondence should be addressed to Denise Schermann Azambuja, denise@iq.ufrgs.br

Received 31 January 2012; Accepted 22 February 2012

Academic Editors: L. Bazzi, G. Bereket, A. Kalendová, and E. E. Oguzie

In order to reduce the environmental impacts caused by chemical substances harmful to the environment and human health, the black wattle tannin can be used as an environmentally friendly corrosion inhibitor in acid and near neutral media. This paper provides information on the application of black wattle tannin as an inhibitor against the corrosion of carbon steel. The inhibition was evaluated using potentiodynamic polarization (PP) and electrochemical impedance spectroscopy (EIS) at room temperature in aerated 0.1 mol L$^{-1}$ Na$_2$SO$_4$ (pH 6.0 and 2.5). The black wattle tannin when used as a corrosion inhibitor is more effective at acidic pH, its efficiency being dependent on its concentration. At the higher pH value (pH 6.0), a blue-black film (ferric tannate) with a short-term protection against corrosion was formed in aerated aqueous solution. At pH 2.5, this blue-black film was not observed.

1. Introduction

Tannins comprise two different classes of polyphenolic compounds; hydrolysable and condensed tannins. The condensed tannins are found in substantial concentration in the wood and bark of several trees, for instance, black wattle [1]. The tannin extracted from the bark of the black wattle tree contains flavonoid units such as (−)-robinetinidol, (+)-catechin, and (+)-gallocatechin [2]. The flavonoid units are tree-ring flavonols with fifteen carbons. The molecular weight of condensed tannins ranges from around 500 to over 20,000 [3–5].

The hydrolysable tannins are obtained from the fruit, pod, and wood of several trees (fruit of Terminalia chebula, pod of Caesalpinia coriaria, and wood of Castanea valonae) [1–4]. This class of tannin is made up mainly of gallic and digallic acids, which are often esterified to polyols [3], and have molecular weights of up to 3000 [1].

Due to the OH$^-$ groups in the orthoposition in aerated aqueous solution, a highly insoluble and blue-black complex (ferric tannate) is formed [7].

The application of tannins in corrosion studies has been investigated by several authors and their efficiency is controversial. According to Favre et al. the presence of hydrolysable tannin (gallic acid) inhibits the formation of magnetite when lepidocrocite is reduced [8, 9]. Electrochemical impedance spectroscopy (EIS) experiments performed by Galvan et al. showed no substantial increase in the polarization resistance ($R_p$) of steel due to treatment of a rusted surface with tannins [10]. On the other hand, some authors have developed a tannin primer that exhibited excellent anticorrosive properties [11–14]. Rahim et al. found that the mangrove tannin and its flavonoid monomers are potential corrosion inhibitors for steel in acidic medium [15]. According to Ross and Francis, some of the rust is converted to more stable, inert, and adherent products [16]. These contradictory results may be due to the diversity of material used in the different studies [17, 18].

Martinez investigated the mechanism of the adsorption of mimosa tannin onto low-carbon steel in sulphuric acidic solutions. At pH 1 and 2, the value of the free energy of adsorption suggests a chemisorption mechanism. This mechanism
occurs due to the formation of an adsorption bond between the oxygen lone-pair electrons of the tannin \( \cdot OH \) group and the metal surface. At \( pH \geq 3 \), ferric tannate is formed and the value of the free energy of adsorption suggests a physisorption mechanism of ferric tannate adsorption onto the steel surface [19].

The aim of this study was to evaluate the anticorrosive properties of tannin extracted from the bark of the black wattle tree (Acacia mearnsii De Wild.) in 0.1 mol L\(^{-1}\) \( Na_2SO_4 \) solutions: \((□)\) \( pH \) 2.5 medium and \((●)\) \( pH \) 6 medium.

2. Experimental Procedure

2.1. Material. The tannin sample used in this work was supplied by TANAC (Montenegro, Brazil). The chemical composition of the carbon steel samples is given in Table 1. All solutions and samples were prepared with analytical grade reagent. Sulphuric acid (Synth, Brazil) was used to adjust the pH value. Sodium sulphate (VETEC, Brazil) was employed in the preparation of the electrolyte solutions.

2.2. Sample Preparation and Experimental Setup. Disc-working electrodes with an area of approximately 1.0 cm\(^2\) were prepared from the steel samples by cold resin (epoxy) embedding. The surface was prepared by grinding with silicon carbide paper up to grade number 1200 followed by degreasing with an acetone/chloroform mixture (1:1) and drying under a hot air stream.

A saturated calomel electrode \((E = 0.241 \text{V/NHE})\) was used as the reference electrode, and all potentials are referred to it. The auxiliary electrode was a Pt wire. The experiments were carried out under naturally aerated conditions at 25°C.

The electrochemical measurements were performed using a potentiostat (AUTOLAB PGSTAT 30, (Echo Chemie, The Netherlands)) coupled to a frequency response analyser (FRA 2). The software used for analysis of impedance spectra was FRA 4.9 (Echo Chemie, The Netherlands). The potentiodynamic polarization curves were recording at a scan rate of 1 mVs\(^{-1}\). The corrosion potentials \((E_{corr})\) and the polarization resistances \((R_p)\) were calculated at a scan rate of 0.1 mVs\(^{-1}\) with potential range of \(E_{corr} ± 20 \text{ mV}\). This method was based on ASTM G59 [20] and ASTM G102 [21] standards.

The EIS measurements were performed in potentiostatic mode at the open circuit potential, OCP. The OCP after potential stabilization is referred to in this study as the corrosion potential, \(E_{corr}\). The amplitude of the EIS perturbation signal was 10 mV, and the frequency range studied was from \(10^2\) to \(10^{-2}\) Hz.

Fourier transform infrared (FTIR) analysis was performed with a Perkin Elmer Spectrum 1000 spectrometer.

3. Results and Discussion

3.1. Influence of Black Wattle Tannin Concentration. Figure 1 shows the polarization resistance \((R_p)\) for carbon steel samples immersed for 1 day in aerated 0.1 mol L\(^{-1}\) \( Na_2SO_4 \) solutions (\( pH \) 2.5 and 6) with different concentrations of black wattle tannin. The \(R_p\) values were obtained from stepwise potentiostatic polarization using a single small potential step \((\Delta E)\) of \(±20 \text{ mV}\).

At concentrations of up to 2 g L\(^{-1}\) of tannin, the \(R_p\) values of the steel samples increase for both mediums (\( pH \) 2.5 and 6). Concentrations above 2 g L\(^{-1}\) were not tested due to the presence of nondissolved black wattle tannin particles in \( Na_2SO_4 \) solutions at room temperature. Thus, in this study, the concentration of tannin used as an inhibitor was 2 g L\(^{-1}\) in aerated 0.1 mol L\(^{-1}\) \( Na_2SO_4 \) solution. This concentration is not very different from that reported by Rahim et al. who tested the efficiency of mangrove tannin [15, 22].

3.2. Study at pH 6.0. After one hour of immersion in aerated 0.1 mol L\(^{-1}\) \( Na_2SO_4 \) solution (\( pH \) 6.0), a nonadherent blue-black film on the steel surface and a blue-black precipitate on the bulk of the electrolyte were observed. Some authors have attributed the formation of these blue-black products to ferric-tannate formation [23]. Fourier transform infrared (FTIR) analysis was performed in order to verify whether these blue-black products were ferric tannate.

3.2.1. Results of Fourier Transform Infrared (FTIR) Analysis. The FTIR spectrum of the black wattle tannin (Figure 2(a)) shows a broad absorption band with a maximum absorbance.

| Table 1: Chemical composition of carbon steel sample. |
|------------------------------------------------------|
| Element | C     | Mn    | P     | S     | Cu    | Cr    |
| wt.%    | 0.103 | 0.46  | 0.013 | 0.096 | 0.01  | 0.18  |

The FTIR spectrum of the black wattle tannin (Figure 2(a)) shows a broad absorption band with a maximum absorbance.
Figure 2: (a) FTIR spectrum of the black wattle tannin and (b) FTIR spectrum of the blue-black precipitate formed in the bulk of the electrolyte after one hour of immersion of steel in aerated 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 6.0) in the presence of acacia tannin.

at 3413 cm$^{-1}$ which is due to the presence of hydroxyl groups [15]. Peaks occurring between 1600 and 1450 cm$^{-1}$ are characteristic of aromatic compounds [15]. Various peaks in the 600–1300 cm$^{-1}$ correspond to substituted benzene rings [15].

Figure 2(b) shows the FTIR spectrum of the blue-black precipitate. The reduced intensity (11.90% to 20.02% of transmittance) of the broad peak at around 3413 cm$^{-1}$ shows the reduction of free OH groups [24]. These hydroxyl groups in aromatic rings enable the tannins to form ferric-tannate complexes. In this way, the formation of ferric tannate was detected. The peaks occurring at 1110 cm$^{-1}$ and 619 cm$^{-1}$ are characteristic of sulphate groups (electrolyte solution) [25, 26].

3.2.2. Potentiodynamic Polarization. The potentiodynamic polarization curves of steel immersed in aerated 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 6.0), in the presence and absence of tannin, were obtained from 600 mV up to $E_{corr}$ and 300 mV down to $E_{corr}$ after different periods of immersion (Figure 3).

According to Figure 3, the anodic current densities decrease in the presence of tannin in all immersion times, while the cathodic current densities show no significant modification. The presence the black wattle tannin shifts the $E_{corr}$ to more positive values in whole immersion times.

The corrosion current densities ($j_{corr}$) in Table 2 were calculated from the extrapolation of the anodic and cathodic Tafel lines (Figure 3). The inhibition efficiencies IE (%) were calculated by the polarization resistance according to (1) [15]:

$$\text{IE} (%) = \frac{R_{p\text{tannin}} - R_P}{R_{p\text{tannin}}} \times 100,$$

where $R_{p\text{tannin}}$ and $R_P$ are the polarization resistance value with the presence and absence of black wattle tannin, respectively.

Figure 3: Potentiodynamic polarization curves of steel immersed in aerated 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 6.0) in the presence of tannin (1) and absence of tannin (2) for (a) 1 day, (b) 3 days, and 7 days.
Figure 4: Nyquist plots for steel immersed in 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 6.0) for 1 day (closed triangle), 3 days (closed circle), and 7 days (closed star) in the presence of tannin; and for 1 day (open square), 3 days (open lozenge), and 7 days (open circle) in the absence of tannin.

Table 2: Polarization parameters for the corrosion of carbon steel immersed in 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 6.0), in the presence and absence of tannin, for different periods.

| Period       | $j_{corr}$ (μA cm$^{-2}$) | $E_{corr}$ (V) | $R_p$ (Ω cm$^2$) | IE (%) |
|--------------|--------------------------|----------------|------------------|--------|
| 1 day        | 39.66                    | −0.73          | 486.46           | 91.06  |
| 1 day with tannin | 0.883                     | −0.50          | 5446.0           |        |
| 3 days       | 20.52                    | −0.73          | 461.37           | 93.03  |
| 3 days with tannin | 1.512                     | −0.72          | 6622.2           |        |
| 7 days       | 13.58                    | −0.75          | 481.21           | 75.74  |
| 7 days with tannin | 3.241                     | −0.72          | 1984.3           |        |

The $j_{corr}$ value decreases significantly in the presence of tannin during the first three days of immersion. In the seventh day, the difference between the blank $j_{corr}$ and the inhibitor $j_{corr}$ decreases. The inhibition efficiency remains almost constant for the first and third days, though decreased on the seventh day of immersion.

3.2.3. Electrochemical Impedance Spectroscopy (EIS). The EIS spectra for the steel in aerated 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 6.0), in the presence and absence of tannin, for different periods are shown in Figures 4 and 5.

After 1, 3, and 7 days of immersion in the presence of tannin the plots show two time constants, one at high frequency and one at low frequency with maximum phase angles of around $-20^\circ$ and $-45^\circ$, respectively. The evolution of the EIS spectra may be related to the adsorption of the inhibitor on the metal surface resulting in an increase in the resistance of the steel to polarization. The experimental data shown in Figures 4 and 5 were fitted using two different equivalent circuits (EC) depending on the immersion time as shown in Figure 6. The fitting quality was evaluated based on the error percentage associated with each component, showing errors smaller than 5%. The software used to simulate the EIS data was NOVA 1.7 (Echo Chemie, The Netherlands). The simulated data obtained from the fittings are given in Table 3.

The equivalent circuit (EC) proposed for fitting the EIS diagram after 1, 3, and 7 days of immersion in the presence of tannin is $R_s(Q_1[R_1(Q_2R_2)])$ where $R_s$ represents the ohmic resistance between the reference and working electrodes, $R_1$ and $R_2$ represent the resistance or charge transfer in physical meaning [27]. The $Q_1$ and $Q_2$ parameters are the impedance related to a constant phase element (CPE) and can be attributed electrode surface or adsorbed species [27]. The CPE impedance takes into account the phenomena due to the surface heterogeneities. The CPE impedance is given by (2) [27]:

$$\frac{1}{Z_{CPE}} = Q(j\omega)^n,$$  \hspace{1cm} (2)
the presence and absence of tannin.

| Immersion time | Chi-square | Equivalent circuit | Fitting parameters | n   |
|----------------|------------|-------------------|-------------------|-----|
| 1 day          | 1.3 × 10^{-3} | R_s(R_1Q_1)       | R_1 (kΩ cm²) | Q_1 (F cm²) | 0.80 |
| 3 days         | 0.9 × 10^{-3}  | 61.4              | 2.13             | 3.01 × 10^{-4} | 0.73 |
| 7 days         | 1.1 × 10^{-3}  | 58.6              | 2.30             | 2.24 × 10^{-4} | 0.78 |
| 1 day with tannin | 3.1 × 10^{-3} | 64.1              | 2.34 × 10^{-6}   | 0.64  | 0.369 | 1.84 × 10^{-4} | 0.57  | 53.7 |
| 3 days with tannin | 2.6 × 10^{-3} | 62.3              | 3.10 × 10^{-6}   | 0.60  | 0.378 | 1.57 × 10^{-4} | 0.63  | 66.5 |
| 7 days with tannin | 3.4 × 10^{-3} | 65.6              | 2.68 × 10^{-6}   | 0.60  | 0.112 | 7.83 × 10^{-5} | 0.73  | 6.56 |

$R_s$ represents the ohmic resistance between the reference and working electrodes; $R_1$ and $R_2$ represent the resistance or charge transfer; $Q_1$ and $Q_2$ are the impedance related to a constant phase element (CPE) and can be attributed to electrode surface or adsorbed species; $n$, $n_1$, and $n_2$ represent a CPE exponent. Capacitor for $n = 1$, a resistor for $n = 0$, and a diffusion process for $n = 0.5$.

Table 3: Fitting parameters used to simulate the EIS plots for steel immersed for 1, 3, and 7 days in 0.1 mol L⁻¹ Na₂SO₄ solution (pH 6.0) in the presence and absence of tannin.

![Figure 6](image)

Figure 6: Equivalent electric circuits proposed to simulate the experimental data of steel immersed in 0.1 mol L⁻¹ Na₂SO₄ (pH 6.0) for (a) 1, 3, and 7 days in the absence of tannin; for (b) 1, 3, and 7 days in the presence of tannin.

Table 4: Black wattle tannin inhibition efficiencies obtained from electrochemical impedance spectroscopy (EIS) and potentiodynamic polarization (PP) in 0.1 mol L⁻¹ Na₂SO₄ solution (pH 6.0).

| Potentiodynamic polarization (PP) | Electrochemical impedance spectroscopy (EIS) |
|----------------------------------|---------------------------------------------|
| IE (%)                           | IE (%)                                      |
| 1 day                            | 91.06                                       | 96.57                                       |
| 3 days                           | 93.03                                       | 96.81                                       |
| 7 days                           | 75.74                                       | 65.52                                       |

$Z_{CPE}$ is the impedance and $\omega$ is the angular frequency. The CPE represents a capacitor for $n = 1$, a resistor for $n = 0$, and a diffusion process for $n = 0.5$.

The EC proposed for 1, 3, and 7 days in the presence of tannin has two time constants. The first constant $R_1Q_1$ represents the time constant of high frequency which is relative to the charge transfer reactions and double layer. The second constant $R_2Q_2$ represents the time constant of low frequency which is relative to the adsorption processes. For all times of immersion in the absence of tannin, the EC proposed was $R_s(R_1Q_1)$, which contains one time constant which is related with charge transfer reactions and double layer.

The Nyquist diagram (Figure 4) shows a small variation in the polarization resistance during the first three days of immersion in the absence of tannin. On the seventh day, there is a small increase in the resistance value which can be attributed to deposition of corrosion products on the metal surface. The $R_p$ values were obtained from the simulated parameters considering that, in all cases, $R_p$ represents the overall resistance. Thus, for the $R_s(Q_1(R_1(R_2R_2)))$ circuit, this value was obtained from the sum of $R_1$ and $R_2$. The highest values for polarization resistance were found on the first and third day of immersion in the presence of tannin. Table 4 shows the inhibition efficiencies (IE) obtained from electrochemical impedance spectroscopy (EIS) and Potentiodynamic Polarization (PP) by (1). The IE values obtained from both methods are in agreement and show a decrease in corrosion resistance with longer immersion times. This fact can be attributed due to the inhibition mechanism at this pH value which provides a formation of nonadherent and porous ferric tannate complex layer.

3.3. Study at pH 2.5. In the case of the immersion in aerated 0.1 mol L⁻¹ Na₂SO₄ solution (pH 2.5), the formation of the blue-black ferric tannate on the steel surface and in the electrolyte was not observed. According to Martinez and Stērn [23], the formation of ferric tannate is not detected at this pH.

3.3.1. Potentiodynamic Polarization. The potentiodynamic polarization curves of steel immersed in aerated 0.1 mol L⁻¹ Na₂SO₄ solution (pH 2.5), in the presence and absence of
tannin, were obtained from 600 mV up to $E_{corr}$ and 300 mV down to $E_{corr}$ after different periods of immersion (Figure 7).

The anodic current density values decreased in the presence of tannin for all exposure periods tested. Accordingly, the corrosion potential ($E_{corr}$) of the metal was shifted to a more positive value and the polarization resistance ($R_p$) increased in the presence of tannin as observed in Table 5. The corrosion current densities ($j_{corr}$) in Table 5 were calculated from the extrapolation of the anodic and cathodic Tafel lines (Figure 7). The inhibition efficiencies IE (%) were calculated by the polarization resistance according to (1).

The $j_{corr}$ value decreases significantly in the presence of tannin during all days of immersion. The inhibition efficiency increases with the increases of exposure days.

3.3.2. Electrochemical Impedance Spectroscopy (EIS). The EIS spectra for the steel immersed in aerated 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 2.5), in the presence and absence of tannin, at OCP, for different periods are shown in Figures 8 and 9.

The Nyquist plots (Figure 8) show a decrease in the $R_p$ value for steel with and an increase in the time of immersion in the aerated 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 2.5). In the presence of tannin, the $R_p$ shows the opposite behaviour (increase with increased time of immersion). This can be attributed to the kinetics mechanism of the adsorption of tannin onto the metal surface and/or the formation of corrosion products from the reaction of iron with tannin. The phase angles in the presence of tannin are higher than those in the absence of tannin, reaching a maximum value of $-65^\circ$ which indicates an increase in the capacitive character of the film formed.

The experimental data presented in Figures 8 and 9 were fitted using the two proposed equivalent circuits (EC) shown in Figure 10. The software used to simulate the EIS data was NOVA 1.7 (Echo Chemie, The Netherlands). The simulated data obtained from the fittings are given in Table 6.

The $R_s$ represents the ohmic resistance between the reference and working electrodes, $R_1$ and $R_2$ represent the resistance or charge transfer; $Q_1$ and $Q_2$ are the impedance related to a constant phase element (CPE) and can be attributed to electrode surface or adsorbed species; $n$, $n_1$, and $n_2$ represent a CPE exponent. Capacitor for $n = 1$, a resistor for $n = 0$, and a diffusion process for $n = 0.5$.

### Table 6: Fitting parameters used to simulate the EIS plots for steel immersed for 1, 3, and 7 days in 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 2.5) in the presence and absence of tannin.

| Immersion time          | Chi-square | Equivalent circuit | Fitting parameters |
|-------------------------|------------|--------------------|--------------------|
|                         |            | $R_s$ ($\Omega$ cm$^2$) | $R_1$ (k$\Omega$ cm$^2$) | $Q_1$ (F cm$^2$) | $n$ |
| 1 day with tannin       | $1.9 \times 10^{-3}$ | 48.1 | 1.76 | $4.73 \times 10^{-6}$ | 0.91 |
| 3 days with tannin      | $2.1 \times 10^{-3}$ | 49.6 | 2.34 | $5.68 \times 10^{-6}$ | 0.91 |
| 7 days with tannin      | $1.4 \times 10^{-3}$ | 57.4 | 3.08 | $4.43 \times 10^{-6}$ | 0.89 |
| 1 day without tannin    | $2.3 \times 10^{-3}$ | 63.6 | 0.71 | $2.80 \times 10^{-5}$ | 0.76 |
| 3 days without tannin   | $2.1 \times 10^{-3}$ | 49.8 | 0.70 | $9.21 \times 10^{-5}$ | 0.66 |
| 7 days without tannin   | $4.5 \times 10^{-3}$ | $R_s (R_1Q_1)(R_2Q_2)$ | 47.2 | $1.53 \times 10^{-5}$ | 0.74 | 112 | $5.49 \times 10^{-4}$ | 0.8 | 0.68 |

$R_s$ represents the ohmic resistance between the reference and working electrodes; $R_1$ and $R_2$ represent the resistance or charge transfer; $Q_1$ and $Q_2$ are the impedance related to a constant phase element (CPE) and can be attributed to electrode surface or adsorbed species; $n$, $n_1$, and $n_2$ represent a CPE exponent. Capacitor for $n = 1$, a resistor for $n = 0$, and a diffusion process for $n = 0.5$.

### Table 7: Black wattle tannin inhibition efficiencies obtained from electrochemical impedance spectroscopy (EIS) and potentiodynamic polarization (PP) in 0.1 mol L$^{-1}$ Na$_2$SO$_4$ solution (pH 6.0).

| IE (%) | Electrochemical impedance spectroscopy (EIS) |
|--------|---------------------------------------------|
| Potentiodynamic Polarization (PP) | IE (%) |
| 1 day | 55.7 | 59.6 |
| 3 days | 62.0 | 70.1 |
| 7 days | 68.7 | 74.3 |
Figure 7: Potentiodynamic polarization curves of steel immersed in aerated 0.1 mol L\(^{-1}\) Na\(_2\)SO\(_4\) solution (pH 2.5) in the presence of tannin (1) and absence of tannin (2) for (a) 1 day, (b) 3 days, and 7 days.

Figure 8: Nyquist plots for steel immersed in 0.1 mol L\(^{-1}\) Na\(_2\)SO\(_4\) solution (pH 2.5) for 1 day (closed triangle), 3 days (closed circle), and 7 days (closed star) in the presence of tannin; for 1 day (open square), 3 days (open lozenge), and 7 days (open circle) in the absence of tannin.

Figure 9: Bode plots for steel immersed in 0.1 mol L\(^{-1}\) Na\(_2\)SO\(_4\) solution (pH 2.5) for 1 day (closed triangle), 3 days (closed circle), and 7 days (closed star) in the presence of tannin; for 1 day (open square), 3 days (open lozenge), and 7 days (open circle) in the absence of tannin.
in corrosion resistance with longer immersion times.

The EIS data demonstrated that at pH 2.5 a distinct behaviour was detected in the presence of tannin, with the formation of a more protective film. The inhibition mechanism of tannin at this pH value should be initially adsorbed on metal surface by electron rich centers of molecule. With increase of immersion time, a layer with adherent nature was formed. That might be a reason of increasing of inhibition efficiency with the increase of immersion time at pH 2.

4. Conclusions

(i) The inhibition action of black wattle tannin towards the corrosion of steel is dependent on the concentration of tannin added and the pH value of the electrolyte.

(ii) A nonadherent ferric tannate complex was formed on the steel surface at pH 6 in the presence of black wattle tannin.

(iii) Ferric tannate was not formed on the steel surface at pH 2.5 in the presence of black wattle tannin.

(iv) The black acacia tannin showed the formation of a layer with a short-term protection against corrosion at pH 6.

(v) The black acacia tannin showed the best performance as a corrosion inhibitor at pH 2.5. The inhibition efficiency increased in the presence of tannin.

(vi) Through EIS, the characteristics and behaviour of the films formed on the steel surface can be identified more clearly. The porosity of the ferric-tannate film and the stronger capacitive character of the film formed in acid medium were identified.

Acknowledgments

The financial support for this work provided by the Brazilian Government Agency CNPq and the Laboratory of Polymer Materials of the Federal University of Rio Grande do Sul (UFRGS), which carried the FTIR analysis, is gratefully acknowledged.

References

[1] A. G. Brown and H. C. Ko, “Black wattle and its utilisation—abridged english edition,” RIRDC 97/72, Rural Industries Research and Development Corporation, Barton, Australia, 1997.

[2] D. G. Roux, E. A. Mailhs, and E. Paulus, “Condensed tannins. 9. Distribution of flavonoid compounds in the heartwoods and barks of some interrelated wattles.” The Biochemical Journal, vol. 78, pp. 834–839, 1961.

[3] P. J. Hernes, R. Benner, G. L. Cowie, M. A. Goi, B. A. Bergamashchi, and J. I. Hedges. “Tannin diagenesis in mangrove leaves from a tropical estuary: a novel molecular approach,” Geochimica et Cosmochimica Acta, vol. 65, no. 18, pp. 3109–3122, 2001.

[4] A. Pizzi, Wood Adhesives: Chemistry and Technology, Marcel Dekker, New York, NY, USA, 1st edition, 1983.

[5] J. Mabrouk, M. Aksira, M. Aziz et al., “Effect of vegetal tannin on anodic copper dissolution in chloride solutions,” Corrosion Science, vol. 46, no. 8, pp. 1833–1847, 2004.

[6] S. Yahya, A. M. Shah, A. A. Rahim, N. H. A. Aziz, and R. Roslan, “Phase transformation of rust in the presence of various tannins,” Journal of Physical Science, vol. 19, no. 1, pp. 31–41, 2008.

[7] J. Gust and J. Suwalski, “Use of Mossbauer spectroscopy to study reaction products of polyphenols and iron compounds,” Corrosion, vol. 50, no. 5, pp. 355–365, 1994.

[8] M. Favre and D. Landolt, “The influence of gallic acid on the reduction of rust on painted steel surfaces,” Corrosion Science, vol. 34, no. 9, pp. 1481–1494, 1993.

[9] M. Favre, D. Landolt, K. Hoffman, and M. Stratmann, “Influence of gallic acid on the phase transformation in iron oxide layers below organic coatings studied with Mossbauer spectroscopy,” Corrosion Science, vol. 40, no. 4-5, pp. 793–803, 1998.

[10] J. J. C. Galvan, J. Simancas, M. Morcillo, J. M. Bastidas, E. Almeida, and S. Feliu, “Effect of treatment with tannic, gallic and phosphoric acids on the electrochemical behaviour of rusted steel,” Electrochimica Acta, vol. 37, no. 11, pp. 1983–1985, 1992.

[11] G. Matamala, W. Smetlzer, and G. Droguett, “Use of tannin anticorrosion reaction primer to improve traditional coating systems,” Corrosion, vol. 50, no. 4, pp. 270–275, 1994.

[12] G. Matamala, W. Smetlzer, and G. Droguett, “Comparison of steel anticorrosive protection formulated with natural tannins extracted from acacia and from pine bark,” Corrosion Science, vol. 42, no. 8, pp. 1351–1362, 2000.

[13] O. R. Pardini, J. I. Amalvy, A. R. Di Sarli, R. Romagnoli, and V. F. Vetere, “Formulation and testing of a waterborne primer containing chestnut tannin,” Journal of Coatings Technology, vol. 73, no. 913, pp. 99–106, 2001.

[14] M. J. Kassim, A. A. Rahim, and M. A. Ismail, “Anti-Corrosive Performance Of Wash Primer Based On Mangrove Tannin,” in Proceedings of the 15th Symposium of Malaysian Chemical Engineers (SOMChe’01), pp. 323–327, Johor Bahru, Malaysia, September 2001.

[15] A. A. Rahim, E. Rocca, J. Steinmetz, M. J. Kassim, R. Adnan, and M. Sani Ibrahim, “Mangrove tannins and their flavanoid monomers as alternative steel corrosion inhibitors in acidic medium,” Corrosion Science, vol. 49, no. 2, pp. 402–417, 2007.

[16] T. K. Ross and R. A. Francis, “The treatment of rusted steel with mimosa tannin,” Corrosion Science, vol. 18, no. 4, pp. 351–361, 1978.
[17] O. Lahodny-Š and F. Kapor, “Corrosion inhibition of carbon steel in the near neutral media by blends of tannin and calcium gluconate,” *Materials and Corrosion*, vol. 53, no. 4, pp. 264–268, 2002.

[18] C. A. Barrero, L. M. Ocampo, and C. E. Arroyave, “Possible improvements in the action of some rust converters,” *Corrosion Science*, vol. 43, no. 6, pp. 1003–1018, 2001.

[19] S. Martinez, “Inhibitory mechanism of mimosa tannin using molecular modeling and substitutional adsorption isotherms,” *Materials Chemistry and Physics*, vol. 77, no. 1, pp. 97–102, 2003.

[20] ASTM G59-97: *Standard Test Method for Conducting Potentiodynamic Polarization Resistance Measurements*, 2009.

[21] ASTM G102-89: *Standard Practice for Calculation of Corrosion Rates and Related Information from Electrochemical Measurements*, 2010.

[22] A. A. Rahim, E. Rocca, J. Steinmetz, and M. Jain Kassim, “Inhibitive action of mangrove tannins and phosphoric acid on pre-rusted steel via electrochemical methods,” *Corrosion Science*, vol. 50, no. 6, pp. 1546–1550, 2008.

[23] S. Martinez and I. Ġtern, “Inhibitory mechanism of low-carbon steel corrosion by mimosa tannin in sulphuric acid solutions,” *Journal of Applied Electrochemistry*, vol. 31, no. 9, pp. 973–978, 2001.

[24] J. Gust, “Application of infrared spectroscopy for investigation of rust phase component conversion by agents containing oak tannin and phosphoric acid,” *Corrosion*, vol. 47, no. 6, pp. 453–457, 1991.

[25] J. Bandekar, R. Sethna, and M. Kirschner, “Quantitative determination of sulfur oxide species in white liquor by FT-IR,” *Applied Spectroscopy*, vol. 49, no. 11, pp. 1577–1582, 1995.

[26] X. Wang, Y. Du, L. Fan, H. Liu, and Y. Hu, “Chitosan- metal complexes as antimicrobial agent: synthesis, characterization and structure-activity study,” *Polymer Bulletin*, vol. 55, no. 1-2, pp. 105–113, 2005.

[27] G. W. Walter, “A review of impedance plot methods used for corrosion performance analysis of painted metals,” *Corrosion Science*, vol. 26, no. 9, pp. 681–703, 1986.
Submit your manuscripts at http://www.hindawi.com