Diagnostic Utility of HPV16 E6 mRNA or E7 mRNA Quantitative Expression for Cervical Cells of Patients with Dysplasia and Carcinoma

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Abstract
Current human papillomavirus (HPV)16 DNA testing has high sensitivity but low specificity, while mRNA testing (qualitative) improves the specificity. However, both techniques are not able to discriminate between transient and persistent infections. To overcome the disadvantages, we quantitatively detected E6 and E7 mRNAs by quantitative real-time polymerase chain reaction (qRT-PCR) in cervical brushing cells from 87 HPV16+ and 31 HPV16− patients. Our results showed that the expression levels of E6 mRNA or E7 mRNA were significantly increased in HPV16-positive cases than that in the negative cases. Furthermore, in HPV16+ cases, the expression levels of E6 mRNA were significantly increased in invasive cancer compared with high-grade squamous intraepithelial lesion (HSIL; p < 0.01), and HSIL compared with low-grade squamous intraepithelial lesion (LSIL; p < 0.01). There were no significant changes between LSIL and benign lesions. The expression levels of E7 mRNA presented no significant difference among the above-mentioned four groups. To test whether qRT-PCR can discriminate between transient and persistent infections, 57 HPV16+ patients were followed up for 1 year, and our results demonstrated that the expression levels of both E6 mRNA and E7 mRNA in the persistent infection group were significantly increased relative to the transient infection group (p < 0.01 or 0.05). Thus, a quantitative detection of the expression levels of E6 mRNA in cervical brushing cells may not only be used as an ancillary tool to cytological diagnosis of cervical neoplasia, but may also help to determine the severity of the lesions and the triage of transient infection.

Keywords
Human papillomaviruses, cervical lesions, E6 mRNA, E7 mRNA, cytological diagnosis

Introduction
It is well known that persistent infection of the high-risk type human papillomavirus (HPV)16 is closely related to the occurrence of cervical cancer1–3, and the introduction of HPV DNA testing into the cervical lesion diagnostic programs has improved the effectiveness of cervical screening. However, the current DNA testing has high sensitivity (97.6%) but low specificity (17%). The low specificity of HPV DNA testing is not able to discriminate transient infections, which usually clear within 2 years, from persistent infections. Numerous studies demonstrated that E6 and E7 proteins were key factors in the processing of HPV due to the above genomes being integrated into the human genome, which results in persistent infections4. The expression levels of E6 and E7 mRNA are low during transient infections, but increase once the viral genome integrates into the host. Furthermore, as major oncogenes, E6 and E7 proteins, are correlated with modifying the expression of cell cycle controllers and DNA repair regulations in the development and
progression of cervical cancer. Thus, it is necessary to introduce the detection of E6 and E7 mRNA expression in primary cervical screening programs in order to enhance the overall diagnostic accuracy and provide better long-term protection.

Recently, it has been reported that the detection of E6 and E7 mRNAs in high-risk HPV is superior to the detection of HPV DNA. However, the mRNA testing methods are only qualitative and will not be able to detect the severities of the lesions. Also, both current DNA and mRNA techniques cannot discriminate between transient and persistent infections. Quantitative real-time polymerase chain reaction (qRT-PCR) is the most sensitive and reliable method for detection and quantification of nucleic acids, including RNA. There are numerous applications for qRT-PCR in our studies and it is commonly used for both diagnostic and basic research.

In previous research, we used the qRT-PCR method to detect the mRNAs of both vascular endothelial growth factor (VEGF) and endostatin in cervical epithelial cells. Our results showed that the combined detection of VEGF and endostatin had important clinical application value, and the sensitivity and accuracy were significantly increased compared with cytological diagnosis alone. The primary objective of this study was to apply qRT-PCR to quantitatively detect the mRNAs of E6 and E7 in HPV16-positive and negative cervical epithelial cells and determine the possibility of this technique to overcome the disadvantages of current HPV DNA and HPV mRNA testing. Our E6 mRNA qRT-PCR results showed that qRT-PCR was not only able to identify transient from persistent infections in HPV-positive patients, but also to detect the severities of cervical intraepithelial lesions. Thus, quantitative detection of the expression levels of E6 mRNA in cervical brushing cells may not only be used as an ancillary tool to cytological diagnosis of cervical neoplasia, but may also help to determine the severity of the lesions and the triage of transient infection.

**Materials and Methods**

**Patients Recruitment and Sample Collection**

The study was conducted according to the guidelines of the institutional review boards at the First Affiliated Hospital of China Medical University; we obtained internal review board approval and/or patients informed consent for this study. The cervical brushing cells from 87 patients with HPV16 who attended the laboratory of cytopathology at the First Affiliated Hospital of China Medical University during the period June to October 2015 were included in the study. Their ages ranged from 23 to 77 years, with an average age of 42.4 years. An additional 31 randomly selected patients without HPV16 were included in the study and used as a control group. All patients had cytological diagnoses and biopsy histological diagnoses, and the detailed diagnostic results are shown in Table 1. A total of 57 HPV16+ patients with LSIL and benign by histological diagnoses were followed up for 1 year.

**HPV16 DNA Testing**

HPV16 DNA was detected using an HPV GenoArray DNA Test Kit (HybriBro Ltd., Hong Kong, China) and the protocol was performed according to the manufacturer’s instructions. The detailed procedure of HPV16 DNA testing was described in reference with pubmed identification (PMID) 18322063.

**Cytology Test**

Liquid-based cytology was used for the cytological preparation. All slides were automatically prepared and stained with Papanicolaou method for all 118 cases (BD Tripath, Burlington, NC, USA). The cytological diagnosis was assessed by two independent cytopathologists. The results were interpreted according to the 2015 Bethesda System. Residual materials were available for E6 and E7 mRNA qRT-PCR analysis.

**qRT-PCR**

Total RNA was extracted from cells using TRIzol Reagent (Life Technologies, Carlsbad, CA, USA) and RNeasy RNA isolation kit (QIAGEN, Hilden, Germany). First-strand cDNA was synthesized from 500 ng total RNA using a high-capacity cDNA RT Kit (Applied Biosystems, Carlsbad, CA, USA). The qRT-PCR was performed using SYBER GreenMaster Mix on a 7900HT Fast Real-Time PCR system (Applied Biosystems). Non-template controls were prepared every time for each paired primer to detect non-specific amplification. Average threshold cycle (Ct) values for GAPDH (housekeeping gene) were used to normalize average Ct values of the genes of interest. These values were used to calculate the average Ct values between groups, and the relative quantity (power of $-\Delta \Delta Ct$) was used to calculate
fold change between each group. The detail information of the primers is listed in Table 2.

The amplified products of E6, E7, and GAPDH were confirmed by DNA gel with correct sizes. The products were extracted, purified from the gel, and sent for DNA sequencing. The sequencing results were 100% correct.

Colposcopy and Histological Diagnoses

All HPV16-positive women were examined by colposcopy and underwent cervical biopsy. Biopsy samples were obtained within 4 weeks after the initial HPV DNA tests. Histological diagnosis was made by two experienced pathologists. The histological biopsy results were categorized into four general groups: benign (including no pathologic alteration and benign or reactive changes), LSIL (CIN1), and HSIL (including CIN2/CIN3, squamous cell carcinoma in situ, and/or involving glands). CIN2 lesions were confirmed by immunohistochemical staining for p16 and Ki-67. In patients who had more than one tissue sample, the highest grade of diagnosis was recorded.

Statistical Analysis

The SPSS 16.0 statistical software package (SPSS, Inc. Chicago, IL, USA) was used for all analyses. A Student’s t-test was used to compare data from the densitometry analysis. The McNemar’s test was used to compare the expression levels of both E6 and E7 mRNA in cervical brushing cells from HPV16+ and HPV16− patients. Analysis of variance and the least significant difference test were employed for statistical analysis. The level of statistical significance was set at $p < 0.05$.

Results

The qRT-PCR results of cervical cells from all 118 patients are presented in Table 3. For the mean E6 mRNA expression in 87 HPV16-positive patients was 0.484 ± 0.099; and in 31 HPV16-negative patients was 0.005 ± 0.002. The mean E7 mRNA expression in 87 HPV16-positive patients was 0.210 ± 0.06; and in 31 HPV16-negative patients was 0.003 ± 0.001. The expression levels of both E6 and E7 mRNA were significantly increased in HPV16-positive patients relative to HPV16-negative patients with $p < 0.01$ respectively (Fig.1A and B).

The 87 HPV16-positive patients were further divided into four groups: invasive carcinoma; HSIL; LSIL, and benign according the histological diagnosis. The mean and E6 and E7 mRNA expression levels are presented in Table 3 and Fig. 1 separately. The expression levels of E6 mRNA were significantly increased in invasive carcinoma compared with HSIL ($p < 0.01$), and in HSIL compared with LSIL ($p < 0.01$). There were no significant changes between LSIL and benign groups ($p = 0.97$). The expression levels of E7 mRNA were not significantly different among invasive cancer, HSIL, and LSIL groups, but there was a significant increase in the invasive cancer and HSIL groups when they were compared with the benign group individually ($p < 0.05$).

We further tried to differentiate transient from persistent infections in LSIL and benign patients. We followed up a total of 57 patients for 1 year and divided them into two groups: the persistent infection group (18 patients) and transient infection group (39 patients). Each group was further divided into LSIL and benign groups according to histological diagnosis. Our results are shown in Table 4 and Fig. 2. The expression levels of both E6 and E7 mRNA were significantly increased in the persistent infection group relative to the transient infection group ($p < 0.01$). The follow-up results showed that 10 patients with LSIL and 29 patients

| Histology    | n   | E6     | E7     |
|--------------|-----|--------|--------|
| HPV16 (+)    | 87  | 0.484 ± 0.099 | 0.210 ± 0.06 |
| Carcinoma    | 10  | 1.610 ± 0.410 | 0.470 ± 0.149* |
| HSIL         | 20  | 0.628 ± 0.124 | 0.339 ± 0.217* |
| LSIL         | 18  | 0.143 ± 0.346 | 0.272 ± 0.115 |
| Benign       | 39  | 0.141 ± 0.034 | 0.048 ± 0.025 |
| HPV16 (−)    | 31  | 0.005 ± 0.002 | 0.003 ± 0.001 |

Benign: no intraepithelial lesion; carcinoma: invasive carcinoma; E6: HPV16 E6; E7: HPV16 E7; HPV: human papillomavirus; HSIL: high-grade squamous intraepithelial lesion; LSIL: low-grade squamous intraepithelial lesion. * $p < 0.05$ as compared with benign.
with benign lesions had a natural outcome in transient infection group after 1 year.

Discussion

HPV is a cyclic double stranded DNA virus and it is well known that the high-risk type HPV is closely related to the occurrence of cervical cancer. Because of its important causal role, HPV is used as a biomarker for the detection of cervical intraepithelial neoplasia and invasive cervical cancers. Accumulated evidence had shown that HPV DNA testing provides higher sensitivity (97.6%) than cytology. Thus, it has been recommended as the primary cervical cancer screening in many countries. However, DNA testing is not able to discriminate the transient from the persistent infection with a low specificity (17%). Furthermore, more than 90% of women have been infected with HPV infection in their lifetime, while most of the infections are transient infections and result in spontaneous recovery within 2 years.

Therefore, HPV DNA testing alone may increase the psychological burden for the positive patients and may also lead to excessive colposcopies.

HPV16 E6 mRNA and E7 mRNA are two critical onco-genic transcriptional factors in the early stages of the viral life cycle. Persistent HPV infection is a consequence of increased E6 and E7 mRNA expression in host cells to promote cellular proliferation and malignant transformation. HPV mRNA tests have higher specificity (76%) than DNA tests (17%) with comparable sensitivity. It was approved by the United States Food and Drug Administration (US FDA) for the screening of women older than 21 years old with cytology results as atypical squamous cells of undetermined significance, or for women older than 30 years old. However, these testing are qualitative tests only and E6 and E7 mRNA are cocktailed together. E6 plays an important role in the degradation of p53 and abrogates cell growth arrest, while E7 binds and deactivates retinoblastomas protein (pRb) to interfere with cell cycle regulation. In the current study, qRT-PCR was applied to quantitatively detect the mRNA expression of E6 and E7 individually in HPV16-positive and negative cervical epithelial cells. Our results demonstrated that quantitative detection of E6 mRNA and E7 mRNA expression levels was able to identify the transient from persistent infections in HPV-positive patients. Only E6 mRNA expression levels were able to be used for interpreting the severity of cervical intraepithelial lesions. Our small sample size study of E6 and E7 biomarker expression results were promising, and these results need be used to predict the severities of cervical intraepithelial lesions and identify transient from persistent infections in HPV-positive patients in large sample size studies. Additional clinical trials are needed to determine the true clinical value of this assay.
Conclusion

In summary, the current DNA testing of cervical brushing cells has high sensitivity but low specificity, while the mRNA testing (qualitative) improves specificity. However, both techniques cannot discriminate between transient and persistent infections. In the current study, qRT-PCR was applied to quantitatively detect the mRNA of E6 and E7 individually in HPV16-positive and negative cervical epithelial cells. Our results demonstrated that quantitative detection of the expression levels of E6 mRNA in cervical brushing cells may not only be used as an ancillary tool to cytological diagnosis of cervical neoplasia, but also help to determine the severity of the lesions and the triage of transient infection.

Ethical Approval

This study was approved by our institutional review board.

Statement of Human and Animal Rights

Human specimens were tested in accordance with our institutional review board guidelines [2018]2018-10-2.

Statement of Informed Consent

We had obtained the patient informed consent for this study.

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

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