Preparation and Optimization of Esomeprazole Nanosuspension using Evaporative Precipitation–Ultrasonication

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Abstract

Purpose: To prepare and optimize esomeprazole nanosuspension to enhance drug dissolution rate.

Methods: Esomeprazole nanosuspensions were prepared by evaporative precipitation-ultrasonication method using F68 (Poloxamer 188) and F127 (Poloxamer 407) as stabilizers. Formulation and process variables (concentration of stabilizers and drug, power input and duration of ultrasonication) affecting the characteristics of nanosuspensions were optimized. The nanosuspensions were characterized for particle size, shape, zeta potential, stability and in vitro drug release study.

Results: For optimization of esomeprazole nanosuspension, the effect of some important parameters, including concentration of F68, concentration of esomeprazole, precipitation temperature, duration of ultrasonication and power input, on particle size was investigated, and the optimal values were 0.4% w/v, 3.5 mg/ml, 4°C, 20 min and 60% W, respectively. Particle size was in the range of 125 - 184 nm with good zeta potential (15.9 - 25.5 mV). In vitro dissolution rate of esomeprazole was enhanced 4-fold (100% in 60 min) compared with crude esomeprazole (24% in 60 min), and this was due to decrease in particle size. The stability results indicate that nanoformulations stored at 4°C for two months showed maximum stability.

Conclusion: The results indicate the suitability of evaporative-precipitation-ultrasonication method for preparation of nanosuspensions of poorly soluble drugs with improved in vitro dissolution rate, thus potentially capable of enhancing fast onset of therapeutic activity, and bioavailability.

Keywords: Agglomeration, Esomeprazole, Ultrasonification, Nanosuspension, Drug release, Solubility, Stability

INTRODUCTION

Nanosuspension is the colloidal dispersion of solid drug particles in a liquid phase having particle size below 1µm [1,2]. At present time about 40% of the drugs in the development pipelines and 60% of the drugs coming directly from synthesis are poorly soluble [3]. These poorly soluble drugs require some innovative formulation approaches by which they can reach up to a sufficiently high bioavailability. There are many formulation approaches used for poorly soluble drugs, nanosuspension is also one of them. Preparation of drug in the form of nanosuspension was shown a technically simpler alternative, particularly for poorly soluble drugs [4-6]. With the help of this technique, the drugs are dispersed in water and the fineness of...
dispersed particles force them to dissolve more quickly owing to their higher dissolution pressure and leads to an increased saturation solubility [7]. In nanoformulation it is not only necessary that the drug particles be rendered into nanosize domains, but they must also be stabilized and formulated rigorously to retain the nature and properties of the nanoparticles [8].

There are mainly two approaches uses to prepare the nanosuspension: 'top down' and 'bottom up' technologies. The top down technology basically depends on mechanically attrition to render large crystalline particles into nanoparticles by various means such as wet milling and bottom up technology start from molecules, which are dissolved and precipitate them by adding the solvent to a non-solvent [9, 3]. In this research work bottom up technology is used to prepare the nanosuspension due to several advantages. These include high drug entrainment efficiency for poorly soluble drugs, narrow particle size distribution, high batch to batch reproducibility, no homogenization required, simplicity, ease of scale up and low cost equipment required [10].

The main focus of this research work is on the dissolution enhancement of poorly soluble drugs using suitable nanosuspension technology with a view to surmounting the problems associated with drug release, manufacture and stability of the drug product.

EXPERIMENTAL

Materials

Esomeprazole magnesium was obtained as a gift from Unichem laboratories, Ghaziabad, India. Pluronic F-68 and Pluronic F-127 were purchased from Sigma Aldrich Company, St. Louis, USA. PVA, PVP K-30 and HPLC grade methanol were obtained from Rankem, New Delhi, India. HPLC grade water was used throughout the study.

Selection of stabilizer

0.1% (w/v) solution of PVA, PVP K-30, HPC, F-68, F-127 were used as candidate to select the appropriate stabilizers for preparing esomeprazole nanosuspension. The results show that F68 and F127 allowed the production of submicron-sized particles with the small diameter of 380 and 385 nm respectively. These two stabilizers also showed the narrow particle size distribution, in comparison of other stabilizers. So in this current study, F68 and F127 were employed as stabilizers.

Preparation of nanosuspension

Nanosuspensions were prepared by Evaporative precipitation ultrasonication method. Esomeprazole was dissolved in methanol at room temperature to form a series of organic solutions having 2, 2.5, 3, 3.5, 4, 4.5, 5, 5.5, 6, 7, 8, 9, 10 mg/ml of drug. Pluronic F-68 or Pluronic F-127 was dissolved in water to form a series of anti-solvents ranging in concentration from 0.1 - 1.5 %w/v. Both solutions were filtered through a 0.22 µm syringe filter (RanDisc nylon syringe filter). 10 ml of organic solution was injected slowly and drop wise with a syringe into the 20 ml of Pluronic F-68/Pluronic F-127 solution and kept at a low temperature (4°C) in an iced water bath. During injection, the mixture was stirred continuously by the mechanical stirrer at 1000 rpm for 2.5 hr. After the antisolvent precipitation, the volatile solvent was evaporated by subsequently stirred the mixture at 300 rpm for next 3 hrs then sample was transferred to a test tube having the capacity of 30 ml. Each of samples was treated with an ultrasonic probe (Bandelin Sonoplus, Berlin, Germany) at various cycles ranging from 2 x 10 - 7 x 10 %, and power input ranging from 30 – 80 %W) and for various duration (5 - 40 min). The probe (tip diameter, 8 mm) was immersed about 10-12 mm in the liquid in which waves was travelled downwards and reflected upwards. During ultrasonication the temperature was also controlled by using ice water bath.

Determination of particle size analysis and polydispersity nanosuspension

Particle size and polydispersity index (PDI) were determined by dynamic light scattering (DLS) using a Malvern Zetasizer instrument (Malvern, UK), which allows the sample measurement in the range of 0.1 to 10000 nm. Each measurement was performed on the diluted suspension in low volume disposable sizing cuvette at 25 °C in triplicate. Instrument gave the size distribution on the basis of intensity and volume. The dispersant was water, and the complex refractive index used for particles was 1.33.

Evaluation of zeta potential of nanosuspension

The zeta potential of the nanosuspensions was measured by using an additional electrode in the same instrument used for particle size analysis (Malvern Zetasizer). For Zeta Potential determination samples of formulations were diluted with water and placed in the electrophoretic cell. Each sample was measured
three times at 25 °C and average values were employed for measuring the response. In this study, dispersant was water. Refractive index and dielectric constant of dispersant were 1.33 and 78.5 respectively.

**Morphological characterization**

Nanosuspensions shown are yellowish transparent relative to the HPLC grade water, which shows the transparency but without color. The nanosuspension particles were subjected to morphological examination using scanning electron microscopy (SEM, FEI Quanta 200F with oxford-EDS system IE 250 X Max 80, Netherlands). For this purpose, samples were prepared by transferring one drop of nanosuspension on a metal grid and dried under vacuum pressure, coated with gold palladium and the morphological characteristics of the particles were observed at suitable magnification (10,000 to 100,000 x).

**In vitro drug release studies**

*In vitro* release of ESP from different nanosuspension formulations were studied by dialysis method using a dialysis membrane of molecular weight cutoff of 12 KDa. The Freshly prepared phosphate buffer solution (pH 7.4) and Franz diffusion cells were used in this study. Before using the dialysis membrane it was pre-soaked in the dissolution medium overnight and then adapted to the terminal portion of the cylindrical donor compartment. The nanosuspension (5 mL) containing drug, sufficient for establishing sink conditions for the assay, was placed into the donor compartment. The receptor compartment contained 15 mL of phosphate buffer solution of pH 7.4 maintained at 37 ± 0.5°C under mild agitation using a magnetic stirrer. Samples were withdrawn from the receptor compartment at predetermined time intervals and replaced by an equal volume of the medium. The amount of dissolved drug was determined using HPLC (Cecil, UK) at λ max of 302 nm. The mobile phase was a mixture of phosphate buffer of pH 7.6 and acetonitrile (44 : 55), pumped from the solvent reservoir to the column (Spher ODS 5 µm) at a flow rate of 1 ml/min. Rf for esomeprazole was 3.42 min. To examine the limiting effect of the dialysis membrane on drug dissolution and for comparison, the release profile of pure drug was also determined. For this purpose, the equivalent amount of ESP was dispersed in 5 mL of phosphate buffer solution. Each release experiment was performed in triplicate.

**Stability studies**

Esomeprazole nanosuspensions were stored at 4, 25 °C and room temperature (approximately 35 °C) for two months, and average particle size determined using the zetasizer.

**Statistical analysis**

Statistical analysis of the data was carried out using Systat ®12 software (Systat Software, Inc, San Jose, CA, USA). All the tests were run in triplicate and the results given as mean ± standard deviation (SD). Mean values were compared using Student’s t-test. Differences were considered statistically significant at a level of p < 0.05.

**RESULTS**

Effect of drug and stabilizer concentration on particle size

The effect of the concentration of stabilizer on the particle size is shown in Table 1. The result exhibited that the particle size reduced with the increasing of pluronic F68 concentration and finally a plateau region was reached at the concentration of 0.25% w/v (batch ESP4) from where the particle size no remarkably changed. However, at the concentration of 0.6% w/v (batch ESP8) and above, there were again remarkable changes obtained in particle size. The student t-test showed that there was significant difference (p<0.05) in particle size of batch ESP4 compared with the batch ESP8. Similarly, in presence of Pluronic F127, plateau region was obtained at the concentration of 0.2% w/v (ESP13). But, at the concentration of 0.4% w/v (batch ESP16) and above again significant changes (p<0.05) in particle size was observed. After 15 days of nanosuspension preparation, batch ESP6 showed the least particle size changes. Thus, 0.4% w/v concentration of pluronic F68 was selected for formulating the other batches.

In this study, crystals were obtained at both lowest and highest drug concentration. At the lowest drug concentration (2.0 mg/ml), the crystal was grown to 468 nm after 15 days. However, at the highest drug concentration (10.0 mg/ml), comparatively larger crystal was obtained. These larger size crystals increased to 592 and 890 nm, after 15 and 30 days, respectively (Table 2, Figure 1).

When drug concentration was between 3.0 mg/ml and 5.0 mg/ml, the initial particle size was < 200 nm but grew to approximately 400 nm after 30 days. In this range of drug concentration, the
Table 1: Nanosuspension formulae for optimization of stabilizing agent (n = 3)

| Batch  | Pluronic F-68 (% w/v) | Pluronic F-127 (% w/v) | Mean particle size (µm) | Mean particle size after ultrasonication (µm) |
|--------|-----------------------|------------------------|-------------------------|---------------------------------------------|
|        |                       |                        | 0 day           | 7 day             | 15 day          |
| ESP1   | 0.1                   | -                      | 14.33           | 2.16              | 3.28            | 5.12            |
| ESP2   | 0.15                  | -                      | 13.98           | 1.68              | 2.4             | 3.79            |
| ESP3   | 0.2                   | -                      | 14.10           | 0.847             | 1.64            | 2.78            |
| ESP4   | 0.25                  | -                      | 13.65           | 0.218             | 0.564           | 0.954           |
| ESP5   | 0.3                   | -                      | 13.14           | 0.147             | 0.212           | 0.343           |
| ESP6   | 0.4                   | -                      | 12.98           | 0.184             | 0.227           | 0.287           |
| ESP7   | 0.5                   | -                      | 12.87           | 0.226             | 0.282           | 0.378           |
| ESP8   | 0.6                   | -                      | 12.82           | 0.625             | 0.728           | 0.98            |
| ESP9   | 0.8                   | -                      | 12.94           | 0.89              | 0.98            | 1.12            |
| ESP10  | 1.0                   | -                      | 13.10           | 1.28              | 1.35            | 1.54            |
| ESP11  | -                     | 0.1                    | 12.94           | 1.30              | 1.58            | 1.97            |
| ESP12  | -                     | 0.15                   | 12.82           | 0.698             | 0.948           | 1.62            |
| ESP13  | -                     | 0.2                    | 12.65           | 0.265             | 0.382           | 0.518           |
| ESP14  | -                     | 0.25                   | 12.62           | 0.256             | 0.369           | 0.483           |
| ESP15  | -                     | 0.3                    | 12.67           | 0.264             | 0.387           | 0.514           |
| ESP16  | -                     | 0.4                    | 11.85           | 0.356             | 0.491           | 0.655           |
| ESP17  | -                     | 0.5                    | 11.54           | 0.564             | 0.854           | 1.28            |
| ESP18  | -                     | 0.6                    | 11.62           | 0.99              | 1.35            | 1.76            |
| ESP19  | -                     | 0.8                    | 11.91           | 1.04              | 1.34            | 1.85            |
| ESP20  | -                     | 1.0                    | 11.94           | 1.16              | 1.4             | 1.89            |
| ESP21  | 0.1                   | 0.1                    | 15.14           | 0.891             | 0.945           | 1.21            |
| ESP22  | 0.2                   | 0.1                    | 14.67           | 0.564             | 0.654           | 0.768           |
| ESP23  | 0.15                  | 0.15                   | 14.45           | 0.517             | 0.597           | 0.682           |
| ESP24  | 0.2                   | 0.15                   | 12.68           | 0.528             | 0.631           | 0.785           |
| ESP25  | 0.2                   | 0.2                    | 13.14           | 0.84              | 0.9             | 1.12            |

Each batch contained esomeprazole (50 mg), methanol (10 mg) and water (20 mg).

Nanosuspension formulations also showed better stability than the other nanosuspensions with drug concentration < 3.0 or > 5.0 mg/ml.

Effect of power input and duration of ultrasonication

Crystal size decreased with increase in ultrasonic power input. However, particle size did not show any significant difference (p>0.05) when power input was 60 and 65% W.

The duration of ultrasonication also affected particle size. When the power input was 60% W, the particle size of nanoformulations was reduced by increasing the ultrasonication time to 20 min. However beyond the 20 min, the time did not help the nanoformulations in reducing the particle size. When time length of ultrasonication was 5 min to 10 min, some crystals deposited at the bottom of the container after three days. However, 20 min time length was sufficient to reduce particle size and prevent the crystal growth efficiently.

Fig 1: SEM images (x 104 magnification) of nanosuspension particles at acceleration voltage of 10 KV. Note: A = particles after evaporative precipitation; B = particles after ultrasonication; C = particles agglomeration after 15 days; D = particles agglomeration after 30 days; E = esomeprazole nanosuspension (batch ESP6d)
Table 2: Optimization of drug concentration (n = 3)

| Batch   | Esomeprazole (mg) | Pluronic F-68 (% w/v) | Mean particle size (nm) |
|---------|-------------------|-----------------------|-------------------------|
|         |                   |                       | 0 day | 15 day | 30 day |
| ESP6a   | 20                | 0.4                   | 365   | 468    | 620    |
| ESP6b   | 25                | 0.4                   | 242   | 460    | 580    |
| ESP6c   | 30                | 0.4                   | 132   | 165    | 215    |
| ESP6d   | 35                | 0.4                   | 159   | 174    | 192    |
| ESP6e   | 40                | 0.4                   | 125   | 182    | 204    |
| ESP6f   | 45                | 0.4                   | 156   | 212    | 328    |
| ESP6g   | 50                | 0.4                   | 184   | 287    | 418    |
| ESP6h   | 55                | 0.4                   | 258   | 345    | 475    |
| ESP6i   | 60                | 0.4                   | 245   | 352    | 492    |
| ESP6j   | 70                | 0.4                   | 274   | 372    | 483    |
| ESP6k   | 80                | 0.4                   | 387   | 495    | 692    |
| ESP6l   | 100               | 0.4                   | 432   | 592    | 890    |

Table 3: Optimized particle data (mean ± SD, n = 3) for selected esomeprazole batches

| Batch   | Mean particle size (nm) | Zeta potential (mV) | Polydispersity index |
|---------|------------------------|---------------------|----------------------|
| ESP6c   | 132 ± 20               | 20.3 ± 1.8          | 0.54 ± 0.05          |
| ESP6d   | 159 ± 16               | 25.5 ± 3.3          | 0.47 ± 0.08          |
| ESP6e   | 125 ± 12               | 18.7 ± 2.4          | 0.57 ± 0.11          |
| ESP6f   | 156 ± 18               | 15.9 ± 2.1          | 0.63 ± 0.11          |
| ESP6g   | 184 ± 20               | 16.4 ± 2.4          | 0.61 ± 0.13          |

Zeta potential was determined as a function of physical stability. The stabilizer was adsorbed onto the surfaces of the generated nanoparticles, which gave the zeta potential ranging from -15.9 to -25.5 mV (Table 3). Batch ESP6d showed the best zeta potential value (-25.5 mV).

Morphological characteristics

The particles of esomeprazole nanosuspension were spherical to oval in shape (Figure 1). Many particles in batches ESP6d and ESP6e showed smooth surface while those of batches ESP6f and ESP6c had a rough surface. The nanoparticle size as observed by SEM also correlated with size measured by Zetasizer.

In-vitro drug release

The in-vitro release of drug from stabilized nanosuspensions was studied by dialysis membrane diffusion technique. It is a very common method to estimate the % release of drug from colloidal suspension.

Figure 2 shows the release profile of esomeprazole from different selected batches and pure drug in phosphate buffer of pH 7.4. The release profile of pure drug (control solution of esomeprazole) indicates very slow diffusion of esomeprazole with nearly 24% release in 60 min., while nanosuspensions showed a significantly enhanced release rate, 65% of the drug diffused in 30 min and 100% diffused in the 60 min test period. No significant (P > 0.05) difference in the in vitro release profile of selected batches (ESP6c, ESP6d, ESP6e, ESP6f and ESP6g) was observed.

Fig 2: In-vitro drug release from selected esomeprazole nanosuspension batches esomeprazole solution (control) in phosphate buffer (pH 7.4). Note: ESP6c = ◊; ESP6d = □; ESP6e = △; ESP6f = ×; ESP6g = +; esomeprazole solution = ●
Short-term stability of nanosuspension

The physical appearance of ESP6d nanosuspension did not change when it was stored at 4°C for 2 months. A loose, thin layer of sediment was observed when nanosuspension was stored at 25°C for 2 months. However, the sediment disappeared with slight manual shaking. At room temperature storage for 2 months, particle size increased to 390 nm. Mean particle diameter was 158 nm and 240 nm after storage at 4 and 25°C, respectively, compared to 159 nm prior to performing the stability study. Thus the nanosuspension batch, ESP6d, was stable after 2 months of storage at 4°C.

DISCUSSION

The evaporative precipitation ultrasonication method used for the production of esomeprazole nanosuspension can be described as a simple process comprising organic solvent, solvent, stabilizer, stirring and ultrasonication. In this method, the high energy generated as a result of the ultrasound led to erosion of the crystals and drug particles into nanoparticles. However, optimization of drug nanosuspension using this method is a complex process since it includes a lot of variables that affects the nanosuspension characteristics. Thus, this research work helped in identifying the different variables, such as the concentration of drug, concentration of stabilizer, duration of ultrasound and input power which may affect the drug release, stability of drug product and manufacturing process of nanosuspension. From this study, it was found that these significant variables also play a vital role in controlling the mean particle size and zeta potential in nanosuspension and after optimizing these variables it may be possible to prepare the nanosuspension having mean particle size of less than 200 nm.

The large surface area of nanoparticles creates high total surface energy, which is thermodynamically unfavorable, resulting in a tendency to agglomerate to minimize the surface energy. Agglomeration can develop a variety of problems such as crystal growth, rapid settling, creaming [11]. So for overcome these problems Pluronic F-68 and/or Pluronic F-127 were used as a stabilizing agent in this study. The use of these stabilizers is often considered to have advantageous effect in formation of drug nanosuspension. The type and amount of stabilizers also produced a significant effect on the physical stability of nanosuspension. Thus stabilizers were used in different concentrations for managing the stability of different formulations (Table 1). The results revealed that batch ESP6 has better stability than the other batches and hence an appropriate concentration (0.4% w/v) of stabilizing agent (F68) was used for each drug concentration which enabled total crystal surface to become covered, thus providing the enough steric repulsion between the crystals. High concentration of stabilizing agent enhanced the viscosity of the suspension which produced the hindrance in transmission of ultrasonic vibrations and the diffusion between the solvent and anti-solvent during precipitation, thus lead an increase in particle size.

Particle size formation can be explained by the theory of crystallization. This theory includes several steps like particle nucleation, molecular growth and agglomeration or aggregation. The driving force of this process is supersaturation which is useful in determining the nucleation rate and diffusion controlled growth rate. Higher the supersaturation leads the faster crystal growth [12].

The best result was obtained, when drug concentration was lies in between 3.0 mg/ml to 5.0 mg/ml. However, at the higher drug concentration, due to greater supersaturation, a higher diffusion-controlled growth and agglomeration rate were achieved, resulting in larger initial crystal.

The mean particle size of nanosuspension decreased with the increase in ultrasonication power input. This phenomenon attributed to the increase erosion effect on the surface of large particles and particle agglomerates. However, the input power beyond the 60% W was not beneficial for reducing the particle size. In addition, the duration of ultrasonication also showed the similar effect on the particle size as of power input, and exhibited that longer time did not help for reducing the particle size. Hence, the optimal values of ultrasonic power input and duration were selected as 60% W and 20 min, respectively.

After optimization of different variables, submicron sized particles with an average diameter in range of 100 - 500 nm were successfully produced. The results showed a dramatically decrease in particle size of esomeprazole nanoparticles compared with crude esomeprazole which has a mean particle size of 12.1 µm.

Following ultrasonication process, the diffusion rate of selected batches was dramatically improved. The diffusion rate of esomeprazole nanosuspensions was enhanced by more than 4-fold compared with control (drug solution).
Enhancement of release rate might due to the higher surface area of the nanosized drug particles available for dissolution as well as lowered diffusion layer thickness.

The sample stored at 4°C, exhibited optimum stability due to the very low temperature. At this temperature, the solubility of esomeprazole decreased, leading to increased level of supersaturation and rapid nucleation. Since the number of nuclei increased, the solute on each nuclei decreased, and hence the potential for smaller crystals. Furthermore, low temperature decreases diffusion and growth kinetics at the particle boundary layer interface [13].

CONCLUSION

Esomeprazole nanosuspension has been successfully prepared by evaporative precipitation – ultrasonication method. The size of particles present in nanosuspension is highly dependent on process and formulation variables. Optimized formulations can be obtained in the nanoparticle size range of 100 - 200 nm. The nanoformulations has good stability when Pluronic F68 concentration is 0.4% w/v. Thus, evaporative precipitation- ultrasonication is a suitable method for the preparation of nanosuspension of poorly water-soluble drugs, such as esomeprazole, for enhancement of dissolution rate.

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