EFFECT OF NOVEL 2-AMINO-5-BENZYLTHIAZOLE DERIVATIVE ON CELLULAR ULTRASTRUCTURE AND ACTIVITY OF ANTIOXIDANT SYSTEM IN MURINE LYMPHOMA CELLS

Ya. R. Shalai¹, M. V. Popovych¹, O. R. Kulachkovskyy¹, V. P. Hreniukh¹, S. M. Mandzynets¹, N. S. Finiu̇k¹,², A. M. Babšky¹

¹ Ivan Franko National University of Lviv, Biology Faculty, 4, Hrushevskyi St., Lviv 79005, Ukraine
e-mail: yarunash@gmail.com

² Institute of Cell Biology, NAS of Ukraine, 14–16, Drahomanov St., Lviv 79005, Ukraine

Shalai Ya.R., Popovych M.V., Kulachkovskyy O.R., Hreniukh V.P., Mandzynets S.M., Finiu̇k N.S., Babšky A.M. Effect of novel 2-amino-5-benzylthiazole derivative on cellular ultrastructure and activity of antioxidant system in murine lymphoma cells. Studia Biologica, 2019: 13(1); 51–60 • DOI: https://doi.org/10.30970/sbi.1301.591

The influence of newly synthesized thiazole derivative N-(5-benzyl-1,3-thiazol-2-yl)-3,5-dimethyl-1-benzofuran-2-carboxamide on cellular ultrastructure and antioxidant system activity in lymphoma cells was studied in vitro. A pronounced cytotoxic action of the newly synthesized thiazole derivative on the tumor cells in vitro was reported earlier. However, no cytotoxicity of this substance was detected toward non-cancerous cells. In addition, it was shown that the scavengers of active forms of Oxygen significantly reduced a cytotoxic effect of the studied compound. It was found that thiazole derivatives at concentrations of 10 and 50 µM affected the level of lipid peroxidation products and superoxide radicals. The purpose of this work was to investigate the effect of N-(5-benzyl-1,3-thiazol-2-yl)-3,5-dimethyl-1-benzofuran-2-carboxamide on the ultrastructure of lymphoma cells and the activity of enzymes of the antioxidant defense.

The influence of the thiazole derivatives on cellular ultrastructure of lymphoma cells was investigated. Electron microscopy study show that control lymphoma cells have a special subcellular formations such as a relatively large nucleus, and specific plasma membrane filaments. The effects of thiazole derivative at concentrations of 10 and 50 µM revealed apoptotic and necrotic manifestations of cytotoxicity, such as a deformation and disintegration of nucleus, an increased area and number of lysosomes, a destruction of the plasma membrane and a change of mitochondria shape.

The effect of the thiazole derivative on the activity of key enzymes of the antioxidant system in lymphoma cells was investigated. The studied compound at concentrations of
10 and 50 µM activate superoxide dismutase and lead to a decrease of the activity of catalase and glutathione peroxidase. This effect suggests that the change in the activity of enzymes leads to accumulation of H₂O₂ in the lymphoma cells. Such results indicate that the studied substance can realize its cytotoxic effect through the action of the antioxidant system. The obtained data can be used to carry out further preclinical studies of the thiazole derivatives as potential antitumor drugs.

Keywords: lymphoma, thiazole derivative, cellular ultrastructure, antioxidant system, lysosomes

INTRODUCTION

Intracellular structure reflects the physiological condition in cells. The ultrastructure investigations detected the effect of damaging factors on different components of the cell. Electron microscopy along with the study of functional processes makes it possible to establish cause-effect relationships between different pathological conditions and ultrastructural changes in the cell [16]. Pathological states in the organism can be conditioned to distortion of the oxidative-reducing homeostasis. Under these conditions, the ratio of formation of free radicals and the activity of enzymes of antioxidant defense is often distorted [9]. Various factors such as antitumor drugs may affect this ratio.

The thiazole derivatives are heterocyclic compounds that exhibit in addition to antitumor activity also antibacterial, antifungal, antiviral, anti-inflammatory, anticonvulsant, and antidepressant activities [11, 14, 15]. A cytotoxic effect of newly synthesized thiazole derivatives on some lines of tumor cells has been established [1, 2]. It was found that scavengers of the reactive Oxygen species (ROS) significantly reduced a cytotoxic effect of the investigated substances [2]. Further investigations of lymphoma cells revealed that the levels of lipid peroxidation and superoxide radicals were affected by the thiazole derivatives [12]. The purpose of this work was to investigate the effect of thiazole derivative (TD: N-(5-benzyl-1,3-thiazol-2-yl)-3,5-dimethyl-1-benzofuran-2-carboxamide) on the ultrastructure of lymphoma cells and the activity of enzymes of the antioxidant defense.

MATERIALS AND METHODS

Experiments were conducted using white wild-type male mice (20–30 g) with grafted NK/Ly lymphoma. Animals were kept in standard vivarium conditions at constant temperature on a mixed ration. All manipulations with animals were conducted in accordance with “General Ethical Principles of Experimentation on Animals” approved by the First National Congress on Bioethics (Kyiv, Ukraine, 2001) and “European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes” (Strasbourg, France, 1985). The bioethical examination of the experiments carried out at the Biological Faculty of Ivan Franko National University of Lviv was executed according to a protocol No. 10042019 of April 1, 2019.

Ascites tumor cells were passaged by intraperitoneal inoculation of 10–15×10⁶ cells to mice. Ascite was drained from the abdominal cavity of anaesthetized mice with sterile syringe on 7–10 day after the inoculation. Lymphoma cells were washed in 0.2 mol/L cacodylate buffer and fixed in a solution containing 1.5% glutaraldehyde in the cacodylate buffer for 2 h and 1% solution of osmium tetroxide (2 h) in the same buffer. Subsequently, the specimens were transferred in a 1.5% aqueous uranyl acetate solution.
for 12 h. Fixed specimens were washed and dehydrated at room temperature at concentrations of ethanol from 70 to 100%. The dehydrated specimens were placed in a pure epoxy resin for 48 h at 40 and 60 °C. Microscopic sections were cut on a UTMP6 ultramicrotome; then, the specimens were contrasted with a 1.5% solution of uranylacetate (prepared in 70% ethanol) and photographed using a PEM100 transmission electron microscope (Electron-SELMI, Ukraine) [4].

The thiazole derivative was synthesized at the Department of Organic Chemistry of Ivan Franko National University of Lviv. The steps of synthesis were described in detail in our previous publication [1].

The thiazole derivative was dissolved in the dimethylsulfoxide DMSO (final concentration 5%) and was added to the lymphoma homogenate at a final concentrations of 1, 10 and 50 µM duration of incubation 10 min.

To measure the activity of enzymes, lymphoma cell samples were frozen in a freezer chamber to -20 °C and subsequently used for investigation. Superoxide dismutase activity was measured by the method described by V. Kostyuk et al. [6]. The activity of SOD was expressed as unit SOD / mg protein. Catalase activity was measured spectrophotometrically by the method described by M. Korolyuk et al. [5] for an absorption wave length of 410 nm. The enzymatic activity was expressed in nmoles of H$_2$O$_2$ / min×mg of protein. Glutathionperoxidase (GPO) activity was measured by the method of Moin [10]. The enzymatic activity was expressed in µM of G–SH / min×mg of protein. Protein concentration in each specimen was determined by the method of O. Lowry et al. [7].

On the basis of electronic microphotographs, the following data were calculated: the relation of the nucleus and cytoplasm area, the ratio of the area of nucleus and cytoplasm (N/C ratio), a number and area of mitochondria, a number and area of lysosomes, as well as a qualitative assessment of the form of cells, nuclei, mitochondria and their cristae.

Statistical analysis of the obtained results was carried out using the MS Excel-2013 program. To assess a reliability of difference between statistical characteristics of two alternative sets of data, Student's coefficient was calculated. The difference was found to be significant at P < 0.05.

**RESULTS AND DISCUSSION**

Qualitative analysis of pictures of control lymphoma NK/Ly cells showed that nucleus (1) takes up a large part (~40%) of the cell area (Fig. 1). High nucleus-cytoplasm ratio is typical for tumor cells where an intensive synthetic and proliferative processes occur. Well notable nucleolus (3) and mitochondria (4) of different sizes and shapes are presented. The mitochondrial matrix is electronically dense. The plasma membrane contains the filamentary looking buds (6) that, most likely, play an important role in adhesion with other cells (Fig. 1, B) or with surfaces. In addition, there are a number of high-density lysosomes localized in the cytoplasm (5) (Fig. 1).

It was established that TD at concentration of 10 µM causes destructive changes in lymphoma cells mainly via apoptosis (Fig. 2). In particular, the cells decreased their size (shrinkage) and lost their elliptical shape. We observed a deformation of the nucleus, a decrease in their size (A1, B1, D1), and a destruction of plasma membrane (A7, C7, D7). Some mitochondria contain swollen cristae (B4), while other contain paralleled cristae (D4).

TD at 50 µM concentration caused even more destructive changes in the lymphoma cells through both apoptosis and necrosis (Fig. 3). Some cells shrunk (B and C) and lost...
their elliptical shape, while other cells swell (A and D). Many other cells demonstrate a destruction such as a deformation of the nucleus (B,1 and D,1), a damage of plasma membrane (B, C), and an increase in number of lysosomes (A, D, 5) (Fig. 3).

Quantitative analysis of the electronic pictures showed that N/C ratio in control lymphoma NK/Ly cells was 0.69 relative units (r. u.). Under the effect of studied compound at concentrations of 10 and 50 µM, the N/C ratio was decreased by 21 and 23%, respectively, however, a significance of these changes was low ($P>0.05$). A number of lysosomes increased by 60% (50 µM TD), while their area was increased by 28% (10 µM, $P<0.05$) and 53% (50 µM, $P<0.01$), respectively. The lysosomes act as the waste disposal system of the cell by digesting unwanted materials in a cytoplasm using hydrolytic enzymes. The number and area of mitochondria did not significantly change (Table).

Thus, the results of the electronic microscopy show that control cells of lymphoma have a typical shape of tumor cells: large nucleus, less amount of cytoplasm, specific

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**Fig. 1.** Electronic microscopy of lymphoma NK/Ly untreated cells. The arrows point to: 1 – nucleus; 2 – cytoplasm; 3 – nucleolus; 4 – mitochondria; 5 – lysosomes; 6 – buds of the plasma membrane

**Рис. 1.** Електронна мікроскопія клітин лімфоми NK/Ly (контроль). Стрілками позначено: 1 – ядро; 2 – цитоплазма; 3 – ядерце; 4 – мітохондрії; 5 – лізосоми; 6 – вип'ячування плазматичної мембрани.

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plasma membrane buds, etc. The effects of TD revealed both the apoptotic and necrotic manifestations of cytotoxicity: a deformation and a disintegration of the nucleus, a destruction of plasma membrane, a significant increase in area and number of lysosomes, etc.

The thiazole derivative may realize their cytotoxic effect through the effect on the antioxidant system activity [2, 12]. Since the level of lipid peroxidation and radicals is regulated by the activity of antioxidant enzymes, the second task of our work was to find out what is the effect of the newly synthesized TD on the activity of key enzymes of the antioxidant system in lymphoma cells.

Superoxide dismutase is one of key enzyme of the antioxidant protection in the cell. Its main function is the dissociation of $O_2^{-}$ to the Hydrogen peroxide that is reduced by the catalase or by the glutathione peroxidase.

The control level of SOD activity in lymphoma was 0.32–0.37 active units/min × mg of protein. Under the effect of TD during 15 min at concentrations of 10 and 50 µM, the activity of SOD increased by 35% ($P<0.01$) and 29% ($P<0.05$), respectively (Fig. 4).

**Fig. 2.** Electronic microscopy of NK/Ly lymphoma cells under the effect of the thiazole derivative at concentration of 10 µM. For further description, see in Fig. 1

Рис. 2. Електронна мікроскопія клітин лімфоми NK/Ly за дії похідного тіазолу в концентрації 10 мкМ. Інші пояснення див. рис. 1
Fig. 3. Electronic microscopy of lymphoma NK/Ly cells under the action of the thiazole derivative in concentration of 50 µM. For further explanations, see in Fig. 1

Fig. 4. Effects of the thiazole derivative at concentrations of 1, 10, 50 µM on the activity of superoxide dismutase in the lymphoma cells. Control level of the enzyme activity was assumed to be 100%. M ± m; n = 5. * – P<0.05; ** – P<0.01
A quantitate analysis of electronic microscopy data of studying lymphoma cells under the effect of the thiazole derivative (TD)

Кількісний аналіз електронно-мікроскопічних фотографій клітин лімфоми за дії похідного тіазолу

| No | Parameters                          | Control | TD, 10 µM | TD, 50 µM | Remarks                                                                                                                                                                                                 |
|----|-------------------------------------|---------|-----------|-----------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| 1  | Cell area (%)                       | 100±26.3| 100±25.7  | 100±13.0  | The control cells have mostly oval shape. TD caused shrinkage (apoptosis) or swelling (necrosis) of cells.                                                                                               |
| 2  | Nucleus area in cell (%)            | 38.1±12.9| 32.8±9.75 | 34.2±11.8 | In control cells large nuclei of oval shape occupied ~ 40%. TD led to a deformation and a destruction of the nucleus.                                                                                     |
| 3  | Cytoplasm area in cell (%)          | 61.9±20.6| 67.2±21.6 | 65.8±9.56 |                                                                                                                                                                                                       |
| 4  | Nuclear / cytoplasmic ratio         | 0.69±0.36| 0.55±0.21 | 0.53±0.23 | TD caused a decrease in the ratio occur mostly due to a nucleus decrease or destruction.                                                                                                               |
| 5  | Lysosome area (%)                   | 100±11.2 | 127±7.2*  | 153±12.7**| TD caused an increase in the number and area of lysosomes. The visibly bigger number of lysosomes was observed in the treated cells without nucleus.                                                      |
| 6  | Number of lysosomes (per cell)      | 12±3     | 11±2      | 20±4      |                                                                                                                                                                                                       |
| 7  | Mitochondria area (%)               | 100±8.6  | 110±7.2   | 94±3.9    | Some a long-shape mitochondria appear under TD influence.                                                                                                                                              |
| 8  | Number of mitochondria (per cell)   | 22±4     | 20±2      | 26±3      |                                                                                                                                                                                                       |

Comments: In all experimental groups cellular area are taken as 100 %, while for lysosomes and mitochondria data only Control parameters is assumed to be 100 %. Cytoplasm area includes mitochondria and lysosomes. M±m. Statistically significant difference compared to Control: * – \( P<0.05 \); ** – \( P<0.01 \)

A large amount of \( \text{H}_2\text{O}_2 \), produced due to an increased SOD activity, is neutralized by CAT. CAT restores the Hydrogen peroxide to water [9]. Fig. 5 shows changes in the activity of CAT in the lymphoma cells under the action of TD. A control level of CAT activity was 0.23–0.35 nmol of \( \text{H}_2\text{O}_2 \)/min×mg of protein. It was established that the activity of the TD at a concentration of 10 µM reduced by 15% the activity of CAT (\( P<0.05 \)).

In addition to CAT-activity, the neutralization of the \( \text{H}_2\text{O}_2 \) is also carried out by the GPO. This enzyme has a greater affinity for Hydrogen peroxide than CAT. Glutathion-peroxidase function is more efficient at low concentrations of \( \text{H}_2\text{O}_2 \), whereas the CAT is more effective with high concentrations of the substrate (\( \text{H}_2\text{O}_2 \)) protecting the development of the oxidative stress [9].
Fig. 5. Effect of the thiazole derivative at concentrations of 1, 10, 50 µM on the activity of catalase in the lymphoma cells. The control level of the enzyme activity is assumed as 100 %. M ± m; n = 5. * – P<0.05

Рис. 5. Активність каталази у лімфомі за дії похідного тіазолу у концентраціях 1, 10, 50 мкМ. Контрольний рівень активності ферменту прийнятий за 100 %. M ± m; n = 5. * – Р<0,05

Fig. 6 shows changes in the activity of GPO under the action of the TD. Control level of the enzyme was 4.33–4.53 nmol GSH/min×mg protein. It was defined that the activity of GPO was decreased under the action of TD at concentrations of 10 and 50 µM on 29 % (P<0.01) and 27 % (P<0.05), respectively.

Рис. 6. Активність глутатіонпероксидази у лімфомі за дії похідного тіазолу у концентраціях 1, 10, 50 мкМ. Контрольний рівень активності ферменту прийнятий за 100 %. M ± m; n = 5. * – Р<0,05; ** – Р<0,01

TD action leads to an increase in SOD activity in the lymphoma, while TD reduces the activity of CAT and GPO that leads to accumulation of H₂O₂ in tumor cells. These effects are cytotoxic and cause DNA breaks, apoptosis, and reduce the intensity of glycolysis in tumor cells [9]. It was found that TD increased the level of primary products of lipid peroxidation and does not change the level of secondary products, such as, malonic dialdehyde [12]. It was established that the TD decrease the level of superoxide radicals. These results indicate that an increase in SOD activity might have a compensatory effect. The activity of antioxidant enzyme affect the level of primary and secondary products of lipid peroxidation that can be toxic to cancer cells. According to the obtained results, SOD, CAT, and GPO as key enzymes of the antioxidant defense, may be a target for antitumor drugs. Thus, it is assumed that such changes in the activity of the antioxidant enzymes by TD may be a part of the mechanism of increasing sensitivity of cancer cells to the antitumor agents.

Thus, the investigated TD at concentration of 10 and 50 µM caused irreversible changes in the ultrastructure of lymphoma cells. The action of the substance led to a destruction of the plasma membrane, the deformation and destruction of the nucleus, an increase in the area and a number of lysosomes.

Thiazole derivatives are characterized by high lipophilicity due to the presence of a macrocycle ring in the molecular structure. This feature facilitates their penetration
into the cell and, in turn, it determines intracellular targets [8]. Products of the liperoxidations and the antioxidant enzymes could be among these targets.

Earlier, it was shown that the effectiveness of the tested substance toward tumor cells is mediated by an increased amount of $\text{H}_2\text{O}_2$. We assumed that toxic effect of the thiazole derivative toward tumor cells, and their destructive changes might be due to the effect of substance on the activity of the antioxidant enzymes. Since the tested substance does not affect normal cells [1, 13], we suggest that this thiazole derivative might be considered as a potential antitumor drug.

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ВПЛИВ ПОХІДНОГО ТІАЗОЛУ НА КЛІТИННУ УЛЬТРАСТРУКТУРУ І СТАН АНТИОКСИДАНТНОЇ СИСТЕМИ У КЛІТИНАХ ЛІМФОМІ НЕМЕТ–КЕЛНЕРА

Я. Р. Шалай1, М. В. Попович1, О. Р. Кулачковський1, В. П. Гренюх1, С. М. Мандзинець1, Н. С. Фінюк1,2, А. М. Бабський1

1 Львівський національний університет імені Івана Франка вул. Грушевського, 4, Львів 79005, Україна
e-mail: yarunash@gmail.com
2 Інститут біології клітини НАН України, вул. Драгоманова, 14/16, Львів 79005, Україна

Досліджено вплив новосинтезованого похідного тіазолу N-(5-бензил-1,3-тіазол-2-іл)-3,5-диметил-1-бензофуран-2-карбоксаміду на ультраструктуру клітин і активність ферментів антиоксидантної системи лімфоми Немет–Келнера in vitro. Раніше встановлено виражену цитотоксичну дію новосинтезованого похідного тіазолу на пухлинні клітини in vitro і не виявано виражену цитотоксичність цієї речовини щодо непухлинних клітин. Крім того, було досліджено, що зневажлював активних форм Оксигену знижуємо цитотоксичній ефект досліджуваної сполуки. Подальші експерименти на клітинах лімфоми виявили, що досліджувана речовина зумовлювала зниження рівня супероксидного радикалу. Метою цього дослідження було дослідити вплив похідного тіазолу на ультраструктуру клітин і активність ферментів антиоксидантної системи лімфоми Немет–Келнера. На першому етапі вивчали вплив похідного тіазолу на ультраструктуру клітин, на підставі електронно-мікроскопічних досліджень встановлено, що контрольні клітини лімфоми мали характерну для пухлинних клітин форму: велике ядро, відносно менша кількість цитоплазми, специфічні вип'язування плазматичної мембрани. За дії похідного тіазолу у концентраціях 10 і 50 мкМ виявлено різні прояви цитотоксичності: деформація та дезінтеграція ядра, руйнування плазматичної мембрани, апоптичні та некротичні структурні зміни. На другому етапі досліджували вплив похідного тіазолу на активність ключових ферментів антиоксидантної системи лімфоми Немет–Келнера. Досліджувана речовина у концентраціях 10 і 50 мкМ призводила до підвищення активності супероксиддисмутази, натомість знижувала активність каталази і глутатіонпероксидази, що може зумовлювати накопичення Н2О2 у пухлинних клітинах. Отже, структурні зміни клітин лімфоми та цитотоксичний ефект похідного тіазолу ймовірно реалізуються внаслідок порушення активності антиоксидантних ферментів. Результати роботи можуть бути використані для проведення подальших доклінічних досліджень похідного тіазолу як потенційного протипухлинного препарату.

Ключові слова: лімфома, похідні тіазолу, ультраструктура, антиоксидантна система, лізосоми

Одержано: 11.04.2019