Genetic Analysis of *Bordetella pertussis* Isolates from the 2008–2010 Pertussis Epidemic in Japan

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Abstract

A large pertussis epidemic occurred between 2008 and 2010 in Japan. To investigate epidemic strains, we analyzed 33 *Bordetella pertussis* isolates from the epidemic period by sequencing virulence-associated genes (*fim3*, *ptxP*, *ptxA*, and *prn*) and performing multilocus variable-number tandem repeat analysis (MLVA), and compared these results with those of 101 isolates from non-epidemic, earlier and later time periods. DNA sequencing of the *fim3* allele revealed that the frequency of *fim3B* was 4.3%, 12.8%, 30.3%, and 5.1% within isolates in 2002–2004, 2005–2007, 2008–2010, and 2011–2012, respectively. The isolation rate of the *fim3B* strain therefore temporarily increased during the epidemic period 2008–2010. In contrast, the frequencies of the virulence-associated allelic variants, *ptxP3*, *ptxA1*, and *prn2*, increased with time during overall study period, indicating that these variants were not directly involved in the occurrence of the 2008–2010 epidemic. MLVA genotyping in combination with analysis of allele types showed that the prevalence of an MT27d strain temporarily increased in the epidemic period, and that this strain carried virulence-associated allelic variants (*fim3B*, *ptxP3*, *ptxA1*, and *prn2*) also identified in recent epidemic strains of Australia, Europe, and the US. Phenotypic analyses revealed that the serotype Fim3 strain was predominant (≥87%) during all the periods studied, and that the frequency of adhesion pertactin (Prn) non-expressing *B. pertussis* decreased by half in the epidemic period. All MT27d strains expressed Prn and Fim3 proteins, suggesting that *B. pertussis* MT27d strains expressing Prn and Fim3B have the potential to cause large epidemics worldwide.

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Introduction

*Bordetella pertussis*, a highly communicable Gram-negative coccobacillus, is the cause of pertussis (whooping cough), a major acute respiratory infection resulting in severe childhood illness and infant death [1]. Although universal immunization programs have contributed to significant reductions in the morbidity and mortality rates associated with pertussis, the incidence of pertussis has increased in several countries despite high vaccination coverage [2-6]. In Japan, acellular pertussis vaccines (ACVs) were introduced in 1981 and are used to control pertussis with a schedule of 3 primary doses and single booster dose at ages 3, 4, 6, and 18–23 months, respectively. This vaccination schedule has been followed since 1994. The incidence of pertussis cases in adolescents and adults, however, has significantly increased since the early 2000s [7]. The waning of vaccine-acquired immunity and the decrease in opportunities of natural immune boosting owing to reduced levels of *B. pertussis* circulation have been proposed as explanations for the recent resurgences of pertussis [2,8,9]. Another possible underlying factor is the adaptation of the *B. pertussis* population to vaccine-induced immunity [2,10,11].

Antigenic and genetic shifts in *B. pertussis* circulating strains have been identified within virulence-associated genes encoding serotype 3 fimbriae (*fim3*), pertussis toxin S1 subunit (*ptxA*), pertactin (*prn*), and the pertussis toxin promoter (*ptxP*). Allele frequencies of the virulence-associated allelic variants, *fim3B*, *ptxA1*, *prn2*, and *ptxP3*, have increased within the *B. pertussis* population in several countries [10,12–17]. *B. pertussis* strains carrying *ptxP3* are more capable of producing pertussis toxin (PT) than are *ptxP1* strains, and the emergence of *ptxP3* strains was associated with pertussis resurgence in the Netherlands [18]. Similarly, a significant correlation was observed between an increase in *fim3B* strains and pertussis notifications in the US [13]. Strains with *fim3B* have a single amino-acid substitution (A87E) within the surface epitope of Fim3, which interacts with human serum [19,20]. Furthermore, multilocus variable-number tandem repeat analysis (MLVA) has revealed that the *B. pertussis* population has changed over the past 50 years worldwide. In Australia, the frequency of *B. pertussis* MLVA type 27 (MT27) and MT70 strains increased after the introduction of an ACV, and subsequently, the MT27 strain became predominant in 2008–2010 [21,22]. An increase in the frequency of *B. pertussis* MT27 strain was also observed in Europe and the US [13,16,23].

Pertussis epidemics still occur worldwide, and epidemic strains have been characterized by Fim serotyping and/or genotyping within some regions [24–26]. In a Dutch epidemic, significant changes in Fim serotypes and MTs were observed during a period when the pertussis vaccine dose was lowered [24]. Besides phenotypic variants of Fim, *B. pertussis* variants that do not express adhesion pertactin (Prn) have been recently identified in Japan as
well as in other countries [27–30]. Since Prn is a component of ACVs, it is reasonable to hypothesize that Prn-negative variants have increased fitness in humans immunized with ACV. To date, the relationship between the prevalence of Prn-negative variants and pertussis epidemics has not been evaluated.

In Japan, a large pertussis epidemic occurred in 2008–2010 despite high vaccination coverage with ACVs. To elucidate the causes of the epidemic, we determined temporal trends in the frequencies of virulence-associated genes (fim3, ptxP, ptxA, and prn) and genotypes in the *B. pertussis* population from 2002 to 2012. In addition, phenotypes of epidemic isolates were characterized by their expression of Fim and Prn proteins.

**Materials and Methods**

**Pertussis surveillance data**

National surveillance data were obtained from the Ministry of Health, Labor and Welfare of Japan Infectious Disease Surveillance data. Each week, the number of pertussis cases was reported from approximately 3,000 sentinel clinics and hospitals within Japan. Diagnosis was based on bacterial culture, clinical symptoms, and/or the results of a serologic test. The reporting criteria did not change during the study period, 2002–2012.

**Bacterial strains**

We examined 134 clinical *B. pertussis* isolates collected in Japan from 2002 to 2012 (Table S1). Thirty-three of those isolates were collected during the 2008–2010 pertussis epidemic, while 101 isolates were epidemiologically unrelated cases of pertussis. The isolates were cultured on Bordet-Gengou agar (Difco) or cyclodextrin solid medium (CSM) agar [31], and incubated at 36°C for 2–3 days. DNA was extracted from *B. pertussis* isolates by boiling, and stored at −20°C.

**Sequence analysis of fim3, ptxP, ptxA, and prn**

Four virulence-associated genes, *fim3*, *ptxP*, *ptxA*, and *prn*, were analyzed using PCR-based sequencing [14,18,32,33]. Sequence reactions were carried out with a BigDye® Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems), and resultant products were sequenced using an Applied Biosystems 3130xl Genetic Analyzer or 3730 DNA Analyzer. Subsequent sequencing of the variable region 2 (R2) of *prn* was performed where necessary, to distinguish between *prn1* and *prn7* alleles. Primer sets used in this study are listed in Table S2.

**MLVA**

MLVA typing was performed as previously described [22,28]. MTs were assigned using the MLVA typing tool found at http://www.mlva.net. Novel MLVA types were assigned by the webmasters, Drs. H. van der Heide and F. Mooi, from the Centre for Infectious Disease Control Netherlands, within National Institute for Public Health and the Environment, in the Netherlands.

**Immunoblotting and serotyping analysis**

Prn expression in *B. pertussis* isolates was analyzed by immunoblotting [28]. Briefly, protein samples (1 μg) were first subjected to 10% SDS-PAGE, then transferred onto nitrocellulose membranes (Bio-Rad), and finally incubated with anti-Prn antiserum. Antigen-antibody complexes were visualized using a horseradish peroxidase-conjugated secondary antibody (Bio-Rad) and the Western Lightning ECL Pro (PerkinElmer, Inc.). Resultant blots were imaged using a LAS-3000 (Fujiﬁlm, Tokyo, Japan).

Serotyping of *B. pertussis* isolates was performed with indirect whole-cell ELISA using anti-Fim2 and anti-Fim3 monoclonal antibodies as previously described [34,35], with some minor modiﬁcations. Briefly, bacterial cells cultured on Bordet-Gengou agar were resuspended in phosphate-buffered saline (PBS) to an optical density of 0.01 at 620 nm, and then inactivated at 56°C for 1 h. The wells of 96-well ELISA plate (Nunc Immuno Plate Maxisorp) were coated with 100 μl of this suspension to each well and allowing it to evaporate overnight at 36°C. Anti-Fim2 (NIBSC 04/154) and anti-Fim3 (NIBSC 04/156) antibodies were used at a 1:1,000 dilution in PBS. Antibody binding to bacterial cells was detected following the addition of a 1:4,000 dilution of alkaline phosphatase-labeled goat anti-mouse IgG (Southern Biotechnology Associates, Inc.) and with the use of p-nitrophenylphosphate as a substrate. The optical density was measured at 405 nm with 650 nm as a reference using a Multiskan FC microplate reader (Thermo Fisher Scientific Inc.). *B. pertussis* strain 18323 that expresses both Fim2 and Fim3 was used as a positive control.

![Figure 1. Number of reported pertussis cases per week in Japan from 2002 to 2012. Pertussis cases are shown by the black line, with each value representing a week of the year. The percentage of adolescent and adult cases (≥15 years old) per year is shown in red circles. The data were obtained from the Ministry of Health, Labor and Welfare of Japan Infectious Disease Surveillance data. Data regarding the number of adolescent and adult cases in 2012 were not available.](https://www.plosone.org/figure1)
Statistical analysis

Squared Pearson’s correlation coefficient ($R^2$) was used to identify a linear dependence between allele frequency ($fim3B$, $ptxP3$, $ptxA1$, or $prn2$) and isolation periods. Fisher’s exact test was performed to analyze the distribution of $B. pertussis$ population. The Simpson’s diversity index (DI) and 95% confidence interval (CI) of MTs was calculated as described by Hunter and Gaston [36] and Grundmann et al. [37], respectively, using the online tool available at http://www.comparingpartitions.info/.

Results

Characteristics of the 2008–2010 pertussis epidemic in Japan

There were 17,349 reported pertussis cases between January 2008 and December 2010 in Japan. Within this pertussis epidemic period, 3 sharp peaks representing increases in case frequency were observed: 1 in late May 2008 (347 cases at week 22), 1 in mid-May 2009 (207 cases at week 20), and 1 in mid-June 2010 (289 cases at week 24) (Fig. 1). The number of reported cases per year in 2008–2010 was $2.7$ times higher than the previous 5-year average. Although the number of pertussis patients over 15 years of age steadily increased in the 2000s, alongside increases of adolescent and adult incidence rates (40.7%, 44.2%, and 52.9% of all reported cases in 2008, 2009, and 2010, respectively), in 2011, the number of those patients decreased and the incidence rate in adolescents and adults also decreased to 41%.

Temporal changes in the frequencies of $fim3$, $ptxP$, $ptxA$, and $prn$ alleles

Among the 134 $B. pertussis$ isolates tested, 2 $fim3$ ($fim3A$ and $fim3B$), 3 $ptxP$ ($ptxP1$, $ptxP3$ and $ptxP8$), 2 $ptxA$ ($ptxA1$ and $ptxA2$), and 4 $prn$ ($prn1$, $prn2$, $prn3$, and $prn9$) alleles were identified. Figure 2 shows the temporal trends of the allele frequencies. The frequency of the allele $fim3B$ temporarily increased during the epidemic period (2008–2010): it was 4.3% in 2002–2004, 12.8% in 2005–2007, 30.3% in 2008–2010, and 5.1% in 2011–2012 (Fig. 2A). In contrast, the frequencies of $ptxP3$, $ptxA1$, and $prn2$ increased with time from 2002 to 2012 (Fig. 2B-D). High correlations ($R^2 \geq 0.95$) were observed between these latter allele frequencies and the isolation periods.

Temporal changes in the frequencies of Prn and Fim variants

The Prn and Fim expression phenotypes of the 134 $B. pertussis$ study isolates were determined with immunoblotting and indirect whole-cell ELISA, respectively. Figure 3A shows the temporal trend of the frequencies of the 2 identified Prn variants, Prn-expressing and Prn-negative strains, with the frequency of Prn-negative strains at 43.5%, 41.0%, 21.2%, and 25.6% during 2002–2004, 2005–2007, 2008–2010, and 2011–2012, respectively. A decreased frequency of Prn-negative strains was observed during the epidemic period (2008–2010); however, this decrease was not statistically significant ($P > 0.05$). All the Prn-negative strains carried the $prn1$ allele (Table S1). Prn-negative strains carrying $prn1$ were previously found in Finland [30], and those carrying $prn2$ were found in France and the US [27,29].
Temporal changes in MTs and genetic diversity

Among the 134 *B. pertussis* isolates, 24 different MTs were identified, of which 2 were novel (MT251 and MT253). Figure 4 shows minimum spanning trees that revealed the genetic diversity of the *B. pertussis* population during the time periods of 2002–2004, 2005–2007, 2008–2010, and 2011–2012. Eighteen *B. pertussis* isolates carrying fim3B were identified during the time periods, and these fim3B strains were divided into 4 MTs: MT26 (n = 1), MT27 (n = 14), MT32 (n = 1), and MT69 (n = 2). Although MT27 and MT186 were the predominant types during all the time periods, the fim3B strain did not belong to MT186. MT27 strains carrying fim3B were most frequent in MT27 during the epidemic period (2008–2010) at 56% (10/18), and their frequency decreased to 7% (2/27) in 2011–2012. The temporal increase of MT27 strains carrying fim3B was statistically significant (*P* < 0.05) when compared with non-MT27 strains carrying fim3B.

MT27 strains could be further classified into 5 subtypes (MT27a to MT27e) based on their allele types (Table 1). MT27a strains carried fim3A, ptxP3, ptxA1, and pnm2, and were the predominant subtype. Notably, they were collected in both non-epidemic and epidemic periods. In contrast, MT27d strains carrying fim3B, ptxP3, ptxA1, and pnm2 were predominantly isolated during the epidemic period, with 10 of 12 isolates (83%) being of this subtype in 2008–2010. An MT27c strain carrying fim3A, ptxP3, ptxA1, and pnm9 was also isolated during the epidemic period. The other MT27 subtypes, MT27b and MT27e, were found only in 2011–2012.

The genetic diversity of MTs and the 5 MT27 subtypes was subsequently determined. As shown in Figure 5, Simpson’s DI was 0.74 (95% CI, 0.58–0.90), 0.85 (0.78–0.92), 0.77 (0.70–0.84), and 0.59 (0.43–0.75) in 2002–2004, 2005–2007, 2008–2010, and 2011–2012, respectively. Therefore, the genetic diversity within *B. pertussis* population decreased after the 2008–2010 pertussis epidemic.

Relationship between MTs and phenotypes in 2008–2010

During the 2008–2010 pertussis epidemic, MT186 (33%), MT27a (21%), and MT27d (30%) were the predominant MTs, and most of them were serotype Fim3 strains (89%) (Table 2). Fim3 strains were also identified within other minor MTs (MT22, MT27c, MT187, and MT248), while a Fim2 strain belonged to MT34, the 2 Fim2,3 strains were typed as MT27d and MT186, and the Fim(−) strain belonged to MT27a. Of the 10 MT27d strains, 9 expressed Fim3 and 1 expressed both Fim2 and Fim3. On the other hand, Pnm-negative strains were all observed within MT186. Seven (64%) of the 11 MT186 strains did not express Pnm. All other MTs expressed Pnm.

Discussion

In the present study, we demonstrated that the prevalence of *B. pertussis* strains carrying fim3B, which were classified as MT27d, temporally increased during the 2008–2010 pertussis epidemic in
Japan. The MT27d strains of the epidemic period were collected in several areas (Table S1). All MT27d strains carried fim3B, ptxP3, ptxA1, and prn2, and expressed Prn and Fim3 proteins. Although Prn-negative strains have recently increased in their prevalence within Japan and in other countries [27–30], here, a lowered frequency of Prn-negative strains was observed during the epidemic period specifically, indicating that Prn-negative strain types were not involved in the epidemic. Similarly, we found that the prevalence of the virulence-associated allelic variants, ptxP3, ptxA1, and prn2, has increased with time from the early 2000s, indicating that the variants were also not directly associated with the epidemic. Taken together, our findings demonstrate that B. pertussis strains carrying fim3B (i.e., MT27d) were associated with the 2008–2010 pertussis epidemic.

We evaluated the regional effect on B. pertussis population because of the low number of samples of isolates. When the number of isolates was compared, the difference in regional population between epidemic (2008–2010) and post-epidemic (2011–2012) periods was statistically significant (P < 0.01), possibly because of the high number (18/39) of isolates collected in the Kinki district (including Osaka) during the post-epidemic period. A sampling bias cannot be excluded from the analysis of the trend in 2011–2012. However, no significant difference was observed between pre-epidemic (2005–2007) and epidemic periods (P > 0.05). The regional effect was therefore small or negligible for detection of the emergence of strain MT27d in the 2008–2010 epidemic.

In the past decade, the prevalence of B. pertussis strains carrying fim3B has increased worldwide [12–14,25]. In Ontario, Canada, 1 predominant B. pertussis clone was identified in the 2000s [12]. This strain carried the same virulence-associated allelic variants (fim3B, ptxP3, ptxA1, and prn2) as our epidemic strains within MT27d and was identical to the strains involved in recent pertussis resurgences within Europe and Australia. Similarly, fim3B strains carrying ptxP3, ptxA1, and prn2 were predominant in the US during the 2000s, and most were genotyped to MT27 [13]. Interestingly, the pertussis resurgence in the US was correlated with the emergence and predominance of the fim3B allele, but not with the ptxP3 allele. On the other hand, in the Netherlands, the prevalence of fim3B strains temporarily increased in the early

### Table 1. Comparison of MTs and allele types between *Bordetella pertussis* isolates collected in non-epidemic and epidemic periods.

| Allele types | No. of isolates detected within the time period |
|--------------|-----------------------------------------------|
| MT fim3 ptxP ptxA prn | 2002–2004 | 2005–2007 | 2008–2010* | 2010–2012 |
| 12 A 8 2 1 1 | | | | |
| 16 A 3 1 2 1 | | | | |
| 22 A 3 1 2 1 | | | | |
| 26 B 3 1 2 1 | | | | |
| 27a A 3 1 2 5 9 7 24 | | | | |
| 27b A 3 1 3 1 | | | | |
| 27c A 3 1 9 1 | | | | |
| 27d B 3 1 2 2 10 | | | | |
| 27e B 3 1 9 2 | | | | |
| 28 A 1 1 1 1 1 | | | | |
| 32 B 3 1 2 1 1 | | | | |
| 34 A 1 1 1 1 | | | | |
| 69 B 3 1 2 2 | | | | |
| 95 A 3 1 2 1 | | | | |
| 186 A 1 2 1 11 12 11 8 | | | | |
| 187 A 1 2 1 1 1 1 1 | | | | |
| 194 A 1 2 1 4 | | | | |
| 224 A 1 2 1 1 1 | | | | |
| 225 A 1 2 1 1 1 | | | | |
| 226 A 1 2 1 2 | | | | |
| 227 A 1 2 1 1 | | | | |
| 229 A 1 2 1 1 | | | | |
| 230 A 1 1 1 1 1 | | | | |
| 233 A 1 1 1 1 | | | | |
| 234 A 1 1 1 2 | | | | |
| 248 A 1 2 1 1 1 | | | | |
| 251 A 1 2 1 1 1 | | | | |
| 253 A 1 2 1 1 | | | | |

*Pertussis epidemic period.
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2000s, although the strains disappeared in 2010 [14]. Likewise, MT27d strains (carrying fim3B) were not identified after the 2008–2010 epidemic period, and the reasons behind the disappearance of this strain type are unclear. Our findings along with those of previous studies, therefore, suggest that the MT27d strain is a recent epidemic strain that is found worldwide, and that this strain is not only associated with pertussis resurgence but can also be correlated with pertussis epidemics.

Fimbriae of *B. pertussis* are composed of Fim2 and/or Fim3 and FimD. The minor fimbrial subunit FimD forms the adhesive tip [38]. Fim2 and Fim3 are encoded by the single-copy genes *fim2* and *fim3*, respectively [39,40], and are serologically distinct [41]. Fim resulting from the *fim3B* strain is Fim3B, which differs from Fim3A by a single amino-acid substitution (A87E) at the surface epitope. To date, the biological differences between Fim3A and Fim3B are unknown. In an effort to address this issue, we recently observed *B. pertussis* clinical strains and found that most strains carrying *fim3B* had strong autoagglutination capability, unlike those carrying *fim3A*, following the suspension of CSM agar cultures into saline (Otsuka and Kamachi, unpublished data).

Surprisingly, autoagglutination was not observed when the strains were cultured on Bordet-Gengou agar. Bacterial autoaggregation is a phenomenon associated with pathogen virulence in many Gram-negative bacteria [42–46], suggesting Fim3B strains are more virulent than Fim3A strains because of their ability to autoagglutinate. Further study is necessary to fully elucidate the relationship between autoagglutination and Fim3B. Attempts to identify the molecular mechanism that regulates autoagglutination within *fim3B* strains are currently underway.

In many countries, a shift from serotype Fim2 to Fim3 in *B. pertussis* circulating strains was observed after mass vaccination, and antigenic differences (in Fim, PT, and Prn) have been noted between *B. pertussis* vaccine strains and circulating strains. In Japan, the *B. pertussis* strain Tohama carrying ptsA2, ptn1, and fim2 has been used as a vaccine strain to produce ACVs from 1981. Among the 4 currently used Japanese ACVs, 2 contain Fim2 and all do not contain Fim3 [47]. A recent study demonstrated that Fim2 and Fim3 are immunogenic antigens, and that individuals recently infected with pertussis had greater anti-Fim3 IgG concentrations than anti-Fim2 IgG concentrations, consistent with the current predominance of Fim3 strains [41]. Based on this finding, there is a clear need for the improvement of currently used ACVs. Specifically, the inclusion of Fim3 (Fim3A and/or Fim3B) in ACVs may be an effective way to reduce the number of current circulating *B. pertussis* strains, including Fim3B strains. Although Fim2 has been shown to be a protective antigen, the protective immunogenicity of Fim3 is still unknown [48]. Further study of this topic will be required to evaluate the effectiveness of Fim3 as a protective antigen.

In the present study, the genetic diversity of the *B. pertussis* population decreased after the 2008–2010 pertussis epidemic. This decrease reflects the expansion of the MT27a strain type and the disappearance of MT27d strains. The MT27a strains carried the same *ptsP3, ptsA1*, and *prn2* alleles as the MT27d strains, but additionally carried *fim3A*. Significant changes in the *B. pertussis* population were previously observed in a pertussis epidemic in the Netherlands, and this study suggested that strain surveillance may be an early detector of pertussis epidemics [24]. Here, we observed significant changes within the *B. pertussis* population during the 2008–2010 epidemic, lending further support for an early warning system of future pertussis epidemics.
In conclusion, the prevalence of *B. pertussis* MT27d strains temporarily increased during the 2000–2010 pertussis epidemic in Japan. The MT27d strains carried the same virulence-associated allelic variants (*fim3B, ptxP2*, *ptxM1*, and *fim2*) as recent epidemic strains observed in Australia, Europe, and the US. *B. pertussis* MT27d strains, therefore, likely have the potential to cause large epidemics in other countries, and, hence, further study and strain surveillance of the MT27d strain type is warranted.

**Supporting Information**

Table S1 Characteristics of *Bordetella pertussis* strain isolates. (XLSX)

Table S2 PCR primers in this study. (XLSX)

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**Author Contributions**

Conceived and designed the experiments: YM NO KK. Performed the experiments: YM NO HT-A. Analyzed the data: YM NO KK. Contributed reagents/materials/analysis tools: NO KS KK. Wrote the paper: YM KK.

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