Antitumor Effects of 8-Deoxylactucin in RB355 Human Retinoblastoma Cells Are Mediated via Apoptosis Induction, Reactive Oxygen Species Production, and Cell Cycle Arrest

Background: Retinoblastoma is a rare malignancy arising from the immature cells of the retina, generally in children up to the age of 3 years. Here, we assessed the anticancer effects of a natural sesquiterpene lactone – 8-deoxylactucin – on the growth of the retinoblastoma RB355 and normal RPE cells.

Material/Methods: Cell viability was assessed by 3-(4,5-dimethylthiazol-2-Yl)-2,5-diphenyltetrazolium bromide (MTT) assay, and apoptosis was assessed by DAPI staining and annexin V/propidium iodide (PI) assay. Reactive oxygen species (ROS) levels were determined by fluorescence microscopy. Flow cytometry was used to determine the cell cycle distribution. Protein expression was determined by Western blot analysis.

Results: The results showed that 8-deoxylactucin exerted selective and potent anticancer effects on the RB355 cells and exhibited an IC$_{50}$ of 25 µM. Nonetheless, the cytotoxic effects of 8-deoxylactucin on the normal RPE cells were comparatively lower, as evident from the IC50 of 65 µM. 8-Deoxylactucin increased the production of ROS and triggering apoptosis of RB355 cells. The induction of 8-deoxylactucin-induced apoptosis was also accompanied with increased cleavage of caspase 3, upregulation of Bax, and downregulation of Bcl-2. The 8-deoxylactucin-induced cell cycle arrest of RB355 cells was also associated with inhibition of cyclin A and B1 expression, as well as the inhibition of Cdc2 phosphorylation.

Conclusions: 8-Deoxylactucin inhibits the growth of RB355 cells by apoptosis, cell cycle arrest, and increased production of ROS.

MeSH Keywords: Antineoplastic Agents • Apoptosis • Cell Cycle • Genes, Retinoblastoma

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Background

Retinoblastoma is a rare malignancy, accounting for about 4% of all pediatric cancers [1]. It generally arises from the immature retinal cells and is most common in young children. Of the children with retinoblastoma who survive, most lose their vision [2]. It has been observed that most retinoblastoma is diagnosed by or before the age of 3 years [3]. Several factors have been shown to be responsible for the development of retinoblastoma, including the papilloma virus and loss of RB1 gene function [4]. The treatment for retinoblastoma involves surgical removal of the affected tissue, immediately followed by chemotherapy. However, the chemotherapeutic agents often have adverse effects and poor clinical efficacy [5]. Owing to these factors, research is now focussed on developing effective chemotherapy with no or minimal adverse effects so that the quality of the patient’s life is not impaired. Sesquiterpene lactones are a large and diverse group of natural metabolites synthesized by a number of plant species [6], and approximately 5000 sesquiterpene lactones have been characterized [7]. These metabolites have gained great attention over the last few decades due to their exceptional pharmacological potential and they have been shown to exhibit a wide array of bioactivities, ranging from antimicrobial to anticancer [8]. Several of the sesquiterpene lactones have now reached clinical trials due to their selective and potent anticancer activity [9]. 8-Deoxyactucin is one of the important sesquiterpene lactones with immense pharmacological potential. It is commonly extracted in its purest form from the plant Cichorium intybus [10]. Although sesquiterpenes have been shown to halt the growth of cancer cells [1], the anticancer property of 8-deoxyactucin has not been previously reported. The present study was designed to examine the anticancer effects of 8-deoxyactucin against the retinoblastoma cell line RB355. The results revealed that 8-deoxyactucin inhibits the growth of RB355 cells by induction of apoptosis and cell cycle arrest.

Material and Methods

Cell lines and culture conditions

The retinoblastoma cell line (RB355) and the normal retina cell line (RPE) were purchased from the American Type Culture Collection. All the chemicals and reagents were obtained from Invitrogen Life Technologies (MA, USA) unless stated otherwise. The cells were kept in Dulbecco’s modified Eagle’s medium. The medium was supplemented with 10% fetal bovine serum antibiotics (100 U/ml penicillin and 100 μg/ml streptomycin) and 2 mM glutamine. The cells were cultured in a CO2 incubator at 37°C with 98% humidity and 5% CO2.

MTT cell viability assay

The RB355 cells were seeded in 96-well plates and treated with 0–500 μM of 8-deoxyactucin for 24 h. After 24-h incubation, the cells were incubated with MTT for 4 h. After this, the medium was removed and the colored formazan product was solubilized by 200 μl of dimethyl sulfoxide. The viability of the RB355 and RPE cells was then determined by measuring absorbance at 570 nm using a spectrometer (BD Biosciences, San Jose, CA, USA).

DAPI (4',6-diamidino-2-phenylindole) and propidium iodide (PI) staining assays

The RB355 cells were grown in 6-well plates (0.6×106 cells/well) for 12 h. The cells were then subjected to 8-deoxyactucin treatment for 24 h at 37°C. We placed 15 μl of cell culture onto glass slides and stained it with a solution of DAPI. The slides were then covered with cover slips and examined with a fluorescent microscope. Annexin V/PI staining was performed as described previously [11].

Determination of reactive oxygen species (ROS)

The RB355 retinoblastoma cells were cultured in 96-well plates and treated with different concentrations of 8-deoxyactucin for 24 at 37°C. This was followed by harvesting of cells by centrifugation and treatment with 5 μM DCH-DA. The ROS levels were determined by observing cells under fluorescence microscopy.

Cell cycle analysis

The RB355 retinoblastoma cells were incubated with varied concentrations of 8-deoxyactucin (0, 12.5, 25, and 50 μM) for 24 h. Cells were washed with phosphate-buffered saline (PBS) and then stained with PI, and the distribution of the cells in each phase of the cell cycle was determined using a FACS flow cytometer.

Western blotting

The exponentially growing RB355 cells were subjected to treatment with varied concentrations of 8-deoxyactucin for 48 h. The cells were harvested by centrifugation at 400 g for 5 min at 4°C and then washed with PBS and lysed in RIPA buffer. Cells were next incubated on ice for 30 min and vortexed and supernatant containing proteins was collected by centrifuging at 13 000 g for 20 min. The protein concentration in each lysate was then determined by BCA assay. Equal amounts of proteins (50 μg) were loaded and resolved on SDS-polyacrylamide gel. Following gel electrophoresis, proteins were transferred to polyvinylidene difluoride (PVDF) membranes. The membranes were blocked in blocking buffer containing 5% non-fat milk for...
1 h at room temperature and blotted with respective mouse anti-human primary antibodies (purchased from Santa Cruz Biotechnology) overnight at 4°C. Blots were washed in TBS, incubated with horseradish peroxidase-conjugated secondary antibody for 1 h and washed 3 times with TBS, and chemiluminescence was captured on Hyperfilm™ after incubating the blots in ECL Plus solution.

**Statistical analysis**

The experiments were performed in triplicate and results are expressed as mean ±SD. Statistical analysis was done using the t test (for comparisons between 2 samples) and one-way ANOVA followed by Tukey test (for comparisons between more than 2 samples). GraphPad prism 7 software was used to carry out the statistical analysis. Values of $p<0.05$ were regarded as indicative of a significant difference.

**Figure 1.** Chemical structure of 8-deoxylactucin.

**Figure 2.** MTT assay showing the effects of 8-deoxylactucin on the viability of RB355 retinoblastoma and normal RPE cells. The experiments were performed in triplicate and results are shown as mean ±SD (* $p<0.05$).

**Figure 3.** DAPI and PI staining showing induction of apoptosis in the retinoblastoma RB355 cells at the indicated concentrations of 8-deoxylactucin. The experiments were performed in triplicate.
8-Deoxylactucin exerts growth-inhibitory effects on RB355 retinoblastoma cells

To investigate the anticancer effects of 8-deoxylactucin (Figure 1) on RB355 retinoblastoma cells, MTT assay was performed at concentrations ranging from 0 to 500 µM. 8-Deoxylactucin was found to concentration-dependently suppress the viability of the RB355 cells (Figure 2). The IC$_{50}$ of 8-deoxylactucin against the RB355 cells was 25 µM. The toxicity of the 8-deoxylactucin on the normal RPE cells was also determined by MTT assay at 0 to 500 µM concentrations. 8-Deoxylactucin showed comparatively lower cytotoxicity against the normal RPE cells. The IC$_{50}$ of 8-deoxylactucin against the normal RPE cells was 65 µM (Figure 2).

Figure 4. Annexin V/PI assay showing the percentage of RB355 apoptotic cells at the indicated concentrations of 8-deoxylactucin. The experiments were performed in triplicate.

Results

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8-Deoxylactucin induces apoptotic cell death of RB355 cells

DAPI and PI staining were used to explore the underlying mechanisms for the anticancer effects of 8-deoxylactucin, clearly showing that 8-deoxylactucin caused nuclear shrinkage and increased apoptosis (Figure 3). Moreover, PI staining showed that 8-deoxylactucin was more abundant in the PI-stained (Figure 3). The percentage of apoptotic RB355 cells was determined by flow cytometry following annexin V/PI staining. The apoptotic RB355 cells were found to be increased significantly by 8-deoxylactucin treatment with 2.12% in control and 28.99% at 50 µM (Figure 4).

8-Deoxylactucin affects the expression apoptosis-related proteins

The 8-deoxylactucin-treated RB355 cells were lysed and proteins were extracted. The protein samples were then subjected to Western blot analysis. The results showed that 8-deoxylactucin increased cleavage of caspase-3 in a concentration-dependent manner. Moreover, the expression of Bax was increased and expression of Bcl-2 was decreased upon 8-deoxylactucin treatment (Figure 5).

8-Deoxylactucin enhances ROS production

To determine whether 8-deoxylactucin induces apoptosis via production of ROS in RB355 cells, the ROS levels of the RB355
cells were examined by fluorescence microscopy after treatment with 0, 12.5, 25, and 50 µM concentrations of 8-deoxylactucin. We found that the numbers of green fluorescent cells increased concentration-dependently, indicative of increased ROS levels (Figure 6). The levels of ROS were significantly increased (by up to 187%) in RB355 cells in comparison to the untreated cells.

Figure 7. 8-deoxylactucin triggers G2/M arrest of the RB355 retinoblastoma cells as evident by flow cytometry. The experiments were performed in triplicate.

8-Deoxylactucin arrests RB355 cells at G2/M phase

To investigate if 8-deoxylactucin affects the distribution of B355 cells in different cell cycle phases, flow cytometry was performed at 0, 12.5, 25, and 50 µM concentration of 8-deoxylactucin. It was found that the percentage of G2/M phase...
cells increased significantly, with 5.27% in control to 83.27% at 50 µM concentration of 8-deoxylactucin (Figure 7).

**8-Deoxylactucin inhibits the expression and phosphorylation of cell cycle regulatory proteins**

Because 8-deoxylactucin arrested the RB355 retinoblastoma cells at the G2/M phase, the effects of 8-deoxylactucin were also examined on the cell cycle regulatory proteins. We found that the protein expressions of cyclin A and cyclin B1 were decreased by 8-deoxylactucin treatment. Moreover, the phosphorylation of p-Cdc2 was also decreased, with slight effects on the expression of Cdc2 (Figure 8).

**Discussion**

Retinoblastoma accounts for approximately 4% of all pediatric malignancies and is associated with significant morbidity and mortality among children. Effective treatment of cancer is impeded by the scarcity of safe and effective drug regimens [12]. This study was designed to examine the growth-inhibitory effects of a natural SQL, 8-deoxylactucin, on RB355 retinoblastoma cells, and we found that 8-deoxylactucin exerts growth-inhibitory effects on these cells. The IC50 of 25 µM was determined for 8-deoxylactucin against RB355 cells. We found that the growth-inhibitory effects of 8-deoxylactucin were minimal against normal cells, indicative of the cancer cell-specific activity of 8-deoxylactucin. The anticancer activities of 8-deoxylactucin have not been reported previously, but the anticancer effects of natural sesquiterpene lactones are widely reported in the literature. Costunolide, a natural SQL, has been shown to cause significant inhibition of growth of cancer cells [13]. Consistently, costunolide was found to suppress the growth of MCF-7 cells [14]. Accumulating evidence suggests that sesquiterpene lactones exert their anticancer effects through induction of apoptosis. For instance, parthenolide has been showed to inhibit the growth of human acute myelogenous leukemia cells through induction of apoptotic cell death [15]. In the present study, we found that 8-deoxylactucin also triggered apoptosis in the RB355 cells, as shown by DAPI, PI, and annexin V/PI staining. 8-Deoxylactucin induced apoptosis and was involved in modulation of the expression of several apoptosis-related proteins. The expression of cleaved caspase 3 was considerably increased upon treatment of the RB355 cells with 8-deoxylactucin. Moreover, 8-deoxylactucin suppressed the expression of Bcl-2 and increased the expression of Bax in the RB355 cells. Bax and Bcl-2 are considered to be important biomarkers of apoptosis, and an increase in the Bax/Bcl-2 ratio favors apoptosis [16]. A number of anticancer molecules have been shown to induce apoptosis via modulation of Bax and Bcl-2 expression; for example, zerumbone induces apoptosis of liver cancer cells via increase in the Bax/Bcl-2 ratio [17]. Resveratrol has been reported to induce apoptosis in prostate cancer cells via modulation of Bax and Bcl-2 expression [18]. Cell cycle arrest caused by several anticancer agents is an essential mechanism in the growth inhibition of cancer cells [19] and our present results show that 8-deoxylactucin caused the arrest of the RB355 retinoblastoma cells at the G2/M checkpoint, which was associated with suppression of cyclin A and cyclin B. Previous studies have also shown that suppression of cyclins, especially cyclin A and B, triggers cell cycle arrest [20]. 8-Deoxylactucin also caused inhibition of the Cdc2 phosphorylation, with only slight effects on the expression of total Cdc2. Additionally, plant-derived molecules such as thymoquinone induce arrest of cancer cells via alteration of the expression of cyclins and Cdc2, further supporting our results [21].
**Conclusions**

8-Deoxylactucin inhibits the growth of retinoblastoma cells by triggering apoptosis and G2/M cell cycle arrest. Therefore, 8-deoxylactucin may prove beneficial in the management of retinoblastoma and warrants further investigation.

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**Conflict of interest**

None.