Bacteremia among Jordanian children at Princess Rahmah Hospital: Pathogens and antimicrobial susceptibility patterns

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ABSTRACT

Objective: To investigate microorganisms causing bacteremia in Jordanian children and to assess their sensitivity to various groups of antimicrobials.

Methods: A retrospective study was conducted on positive blood cultures taken from 378 children aged below 15 year, who sought medical attention at Princess Rahmah Hospital between January and December 2008.

Results: Out of 4475 tested blood samples, 378 isolates were recovered from blood cultures. The male to female isolate ratio was (1.26:1.0). The most frequent pathogen found was Staphylococcus aureus (86.2%), followed by Klebsiella spp. (9%), Escherichia coli (1.9%), Streptococcus spp. (1.9%), Pseudomonas spp. (0.8%), and Acinetobacter sp. was found in only one culture (0.3%). The susceptibility rate of S. aureus was recorded the highest (99.6%) for vancomycin, and the lowest susceptibility rate (3.2%) was recorded for aztreonam.

Conclusions: Staphylococcus aureus was the main isolate in bacteremic children, with all isolates demonstrating susceptibility to vancomycin. Overall, aztreonam resistance was near 97%, and this rate was not affected by sex and blood isolate type. This information should be considered when empirical therapy is recommended or prescribed for children with bacteremia.

Keywords: Bacteremia, Child Fever, Antimicrobials, Drug resistance.

INTRODUCTION

Bacteremia is a common cause of morbidity and mortality in children (1) worldwide (2–5). Therefore, bacteremia continues to be a serious problem that needs immediate attention and treatment. For an accurate diagnosis and an appropriate choice of antimicrobials, blood culture, which usually takes a few days, is required. The empirical choice of antimicrobials for the treatment of bacteremia is guided by an awareness of previous culture reports. Up-to-date information on the local etiologic patterns and antimicrobial sensitivities is also important. Different factors contribute to the prognosis of the infection such as the type of microorganism, age, underlying disease and microorganism entry (6). However, bloodstream infections in hospitalized patients are usually attributable to the use of central venous lines.

In a prospective five-year study on 344 clinically significant episodes of pediatric septicemia the most common organisms found were Salmonella spp. (15.0%) followed by methicillin-resistant S. aureus. They reported that Haemophilus influenzae accounted for 2.0% of all episodes (7). Another study also reported that of 408 bacterial strains, Salmonella spp. were the most commonly isolated (23%), followed by S. aureus and Acinetobacter-Moraxella (8). The most frequent etiologic agents of bacteraemia cases include Staphylococcus spp., Streptococcus spp., Enterobacter spp., Escherichia coli, Klebsiella pneumoniae and Pseudomonas spp. (1, 6). However, etiologies of bacteremia and sensitivities have been changing over the past years (9, 10).

The rapid emergence of multidrug resistant
bacteremia in developing countries is a new potential threat to the survival of newborn babies, who are in a poor health condition. Therefore, this study was conducted to assess the causative organisms and susceptibility pattern of bacteremia pathogens isolated from children in 2008 (from Jan to Dec. 2008) at Princess Rahmah Hospital in Irbid, Jordan. The importance of this study is to aid clinicians to facilitate the empiric treatment and management of children with symptoms of bacteremia. Moreover, the data would also help authorities to formulate antibacterial prescription policies.

MATERIALS AND METHODS

This retrospective study was conducted on 4475 blood specimens obtained from sick children (< 15 years of age) who attended the Princess Rahmah Hospital as outpatients or inpatients and were diagnosed with bacteremia between January and December, 2008. A total of 378 isolates were recovered from blood cultures, and the repeated positive blood cultures were not considered.

The microorganisms and antibacterial susceptibility data were obtained from the clinical microbiology laboratory records which filled in a prepared data sheet. Sampling process, culturing, bacterial identification and susceptibility testing for antimicrobials were as follows:

Blood specimens were collected in a blood culture bottle that contained 50 ml of tryptose phosphate broth and 0.02% polyanethol sulfonate (liquid). Following standard aseptic procedure, culture was incubated at 37°C for 24 hours prior to the isolation and identification of the bacteria.

Based on the Gram-staining characteristics of the bacteria growth in the blood culture bottle was subcultured onto MacConkey agar, Salmonella-Shigella agar, and Nutrient agar and/or blood agar plates. Bacteria isolated from colonies were further characterized by special biochemical and serological methods (11).

All isolates were tested for their susceptibilities to at least 8 out of 15 antimicrobials using antimicrobial diffusion discs (12). Bacterial sensitivity was tested for the following antimicrobials: Amikacin, Amoxicillin-Clavulanic acid, Ampicillin, Aztreonam, Cefaclor, Cefotaxime, Ceftazidime, Ceftriaxone, Cephalexin, Ciprofloxacin, Gentamicin, Imipenem, Pipracillin, Tobramycin and Vancomycin.

Data were analyzed statistically using SPSS (version 15 for Windows) program calculating the frequency and cross tabulations.

This protocol was approved by the Ethics Committee of the Ministry of Health in Jordan (MOH, REC, 08, 0057).

RESULTS

In a 12-month duration (January to December 2008), a total of 378 out of 4475 blood samples of children below 15 years of age (55.8% male and 44.2% female) that gave a positive blood culture reaction were studied. Results showed that the majority of pathogens isolated were Staphylococcus aureus (86.2%), followed by Klebsiella spp. (9%), Escherichia coli (1.9%), Streptococcus spp. (1.9%), Pseudomonas spp. (0.8%) and Acinetobacter spp. (0.3%) (Table 1). The antimicrobial susceptibility of bacteremia isolates for 15 selected antimicrobial agents used in this study are summarized in Table 2.

The highest susceptibility rate of S. aureus was to vancomycin (99.6%), whereas the lowest susceptibility rate was to aztreonam (3.2%). The highest susceptibility rate of other isolates i.e. Klebsiella spp. were to ciprofloxacin 91.3%, E. coli and Pseudomonas spp demonstrated 100% susceptibility to amikacin. Streptococcus spp. and Acinetobacter spp demonstrated 100% susceptibility to both aztreonam and ciprofloxacin. Whereas, the lowest susceptibility rate for all other isolates, Klebsiella spp. E. coli, Pseudomonas, Streptococcus spp. and Acinetobacter spp was to ampicillin (0%). However, vancomycin was detected to give the highest susceptibility rate (91.5%) to a variety of bacteremia isolates, aztreonam exhibited the lowest susceptibility rate of 5.8% (Table 3).

| Table 1. Microbiological characteristics blood bacterial isolates. |
|-----------------|---|---|---|---|
| **Bacteria**     | **Sex** | **Total** | (%) |
|                 | **M** | **F** |    |    |
| 1 S. aureus      | 176  | 150  | 326 | 86.2 |
| 2 Klebsiella spp.| 22   | 12   | 34  | 9    |
| 3 E. coli        | 5    | 2    | 7   | 1.9  |
| 4 Streptococcus spp. | 5  | 2    | 7   | 19   |
| 5 P. aeruginosa  | 2    | 1    | 3   | 0.8  |
| 6 Acinetobacter spp. | 1  | 0    | 1   | 0.3  |
| **Total**       | 211  | 167  | 378 | 100.0 |

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Table 2. Susceptibility rates of blood bacterial isolates to antimicrobial agents.

| Drug                          | S. aureus (n = 326) | Klebsiella spp. (n = 34) | E. coli (n = 7) | Streptococcus spp. (n = 7) | Pseudomonas spp. (n = 3) | Acinetobacter spp. (n = 1) |
|-------------------------------|---------------------|--------------------------|----------------|---------------------------|------------------------|---------------------------|
|                               | No      | S%      | No      | S%      | No      | S%      | No      | S%      | No      | S%      | No      | S%      |
| Amoxicillin-Clavulanic acid   | 34      | 76.4    | 5       | 20      | 0       | 0       | 0       | 0       | 1       | 0       | 0       | 0       |
| Amikacin                      | 319     | 81.8    | 31      | 38.7    | 7       | 100.0   | 7       | 28.5    | 3       | 100.0   | 1       | 0       |
| Ampicillin                    | 106     | 32      | 13      | 0       | 2       | 0       | 2       | 0       | 1       | 0       | 1       | 0       |
| Aztreonam                     | 311     | 3.2     | 30      | 16.6    | 7       | 71.4    | 5       | 100.0   | 3       | 33.3    | 1       | 100.0   |
| Cefazidime                    | 276     | 25      | 29      | 41.3    | 7       | 85.7    | 6       | 16.6    | 3       | 33.3    | 1       | 100.0   |
| Cefaclor                      | 7       | 85.7    | 2       | 0       | 1       | 100.0   | 0       | 0       | 0       | 0       | 0       | 0       |
| Cephalxin                     | 83      | 13.2    | 9       | 33.3    | 1       | 100.0   | 0       | 0       | 0       | 0       | 0       | 0       |
| Ciprofloxacin                 | 262     | 79.2    | 23      | 91.3    | 7       | 85.7    | 5       | 100.0   | 2       | 100.0   | 1       | 100.0   |
| Ceftriaxone                   | 289     | 44.9    | 31      | 16.1    | 7       | 71.4    | 6       | 83.3    | 3       | 0       | 1       | 0       |
| Cefotaxime                    | 262     | 69.4    | 32      | 12.5    | 6       | 66.6    | 7       | 85.7    | 3       | 66.6    | 0       | 0       |
| Gentamicin                    | 282     | 60.6    | 31      | 193     | 7       | 42.8    | 7       | 42.8    | 3       | 66.6    | 1       | 0       |
| Imipenem                      | 272     | 4.7     | 22      | 9.0     | 7       | 42.8    | 3       | 0       | 3       | 0       | 1       | 100.0   |
| Pipracillin                   | 303     | 52.1    | 30      | 13.3    | 7       | 47.1    | 5       | 100.0   | 3       | 66.6    | 1       | 0       |
| Tobramycin                    | 317     | 83.5    | 30      | 20.0    | 7       | 85.7    | 7       | 57.1    | 3       | 66.6    | 1       | 0       |
| Vancomycin                    | 295     | 99.6    | 24      | 20.0    | 7       | 14.2    | 6       | 83.3    | 0       | 0       | 0       | 0       |

n = Number of total isolates
No = Number of tested isolates
S% = Percentage of Sensitive isolates

DISCUSSION

This current study provides information regarding the main etiological agent S. aureus that causes bacteremia in children of both inpatients and outpatient and its antimicrobial susceptibility patterns. These results are in agreement with other studies that reported S. aureus as the most common bacteria isolated from blood of children (13-15).

An increase in the occurrence of Staphylococcal bacteremia is likely to be related to the increased use of intravascular catheters in medical care centers and puncture wounds (16, 17). The second most common organism causing bacteremia in this study was klebsiella spp. (9%). Similar results were reported by Rahman et al 2002 (14). However Klebsiella was the most common cause of neonatal sepsis in Karachi, Pakistan (18). However, in another studies conducted elsewhere, Pseudomonas aeruginosa was the most common organism (38.3%) followed by Klebsiella (30.4%) and E. coli (15.6%) (19).

In this study, the occurrence of E. coli, Streptococcus spp, Pseudomonas spp and Acinetobacter spp were 1.9%, 1.9% 0.8% and 0.3% respectively. Higher occurrence of these blood isolates was reported in different literature (13, 19, 20).
The most effective antimicrobial agent against *Staphylococcus aureus* demonstrated in this study was vancomycin (99.6%), followed by cefaclor (85.7%), tobramycin (83.5%), ciprofloxacin (79.3%), amoxicillin-clavulanate (76.4%) and gentamicin (60.6%). Whereas low susceptibility rates were observed with ceftazidime (25%), cephlexin (13.2%), imipenem (4.7%) and aztreonam (3.2%). Similar results for vancomycin, ciprofloxacin and amoxicillin-clavulanate were reported in Jordan (21) and elsewhere (22). However, same authors reported high susceptibility rates of *S. aureus*, reaching 85-100% to both ceftazidime and gentamicin respectively.

In this study, *Staphylococcus aureus* showed resistant rate to ampicillin accounted for 67% which was lower than documented resistance rate of 85% conducted elsewhere (23).

However, the highest resistance rate (100%) to ampicillin observed in this study was for *Klebsiella*, *E.coli*, *Streptococcus spp*, *Pseudomonas spp* and *Acinetobacter spp*. Similarly low susceptibility rate (0%) of *Acinetobacter spp* to ampicillin has been reported in the literature (22).

In this study, the susceptibility rate of *Staphylococcus aureus* to cefaclor, amikacin, gentamicin and ceftriaxone was 85.7%, 81.8%, 60.6% and 44.9% respectively, which was lower than that reported by Shwe et al. 2002 (24).

The increasing rates of resistance in *Staphylococcus aureus* may be due to changes in the pathogen over the past years (10, 24). Ceftriaxone shown to be the most effective drug for *Klebsiella spp* with susceptibility rate of 91.3% which was higher than that reported previously in Jordan (25). However, the higher susceptibility rate of 100% to ciprofloxacin was reported in Tanzania (22).

At the same time, all blood isolates from selected children of this study showed highest susceptibility rate to vancomycin (91.5%) followed by ciprofloxacin (81.0%), amikacin (77.5%), tobramycin (77.5%), cefaclor (70.0%), amoxicillin-clavulanic acid (67.5%), cefotaxime (63.8%) and gentamicin (55.8%). Comparatively lower susceptibility rate to pipracillin (49.5%), ceftriaxone (37.1%), ampicillin (28%), ceftazidime (27.6%), cephlexin (16.1%), imipenem (7.5%) and aztreonam (5.8%) was demonstrated. These results are correspondent with the reported data (22, 26).

To conclude, the finding of this study showed that bacteremia in children is mainly caused by *Staphylococcus aureus* organisms, which develop resistance to commonly used antimicrobials. This emergence of multiple drug resistance calls for continuous monitoring and reviewing of antimicrobial policy in hospitals and the country at large. Therefore, this study is important for clinicians in order to facilitate the empiric treatment of children with symptoms of bacteremia. Moreover, the data would also help authorities to formulate antimicrobial prescription policies.

**REFERENCES**

1. Reimer LG, Wilson, ML., Weinstein, MP. (1997). Update on detection of bacteria and fungemia. *Clin Microbiol Rev* 1997; 10: 444–465.
2. Dawodu A, Al-Umran K, Twum-Danso K. A case control study of neonatal sepsis: experience from Saudi Arabia. *J Trop Pediatr* 1997; 43: 84–8.
3. Stoll BJ, Holman RC, Schuchat A. Decline in sepsis associated neonatal and infant deaths in the United States, 1979 through 1994. *Pediatrics* 1998; 102: e18.
4. Bhatta ZA, Yusuf K. Neonatal sepsis in Karachi: factors determining outcome and mortality. *J Trop Pediatr* 1997; 43: 65–70.
5. Orritt FA, Shurland SM. Neonatal sepsis and mortality in a regional hospital in Trinidad: a etiology and risk factors. *Ann Trop Paediatr* 2001; 21: 20–25.
6. Cisterna R, Cabezas V, Gomez E, Busto C, Atutxa I, Erpeleta C. Community-acquired bacteremia. *Rev Esp Quimioter* 2001; 14: 369–382.
7. Cheng AFB, Fok TF, Duthie R, French GL. A five year prospective study of septicemia in hospitalized children in Hong Kong. *J Trop Med Hyg* 1991; 94: 295.
8. Gedeou M, Tassew A, Azene G. Blood culture isolates from an Addis Ababa hospital frequency and its antibiotic sensitivities. *East African Med J* 1984; 61: 190.
9. Gedeou M. Clinical sources and resistance to antimicrobial agents of *Klebsiella* isolates from Addis Ababa hospital. *Ethiopia Med J* 1982; 20: 109.
10. Shah M, Watanakunakorn C. Changing patterns of *Staphylococcus aureus* bacteremia. *Am J Med Sci* 1979; 278: 115–21.
11. Ewing WH. Edward. Ewing’s Identification of Enterobacteriaceae. 4th ed. New York: Elsevier Science Publishing Company, 1986.
12. Bauer AW, Kirby WMM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized simple disc method. *Am J Clin Pathol* 1960; 45: 493.
13. Nimri LF, Batchoun R. Community-acquired bacteremia in a rural area: predominant bacterial species and antibiotic resistance. *J Med Microbiol* 2004; 53: 1045–1049.
14. Rahman S, Hameed A, Roghani MT, Ullah Z. Multidrug resistant neonatal sepsis in Peshawar, Pakistan. *Arch Dis Child Fetal Neonatal Ed* 2002; 87: F52–F54.
15. Sabu T, Tudehope DI, Tilsie MJ. Clinical significance of quantitative blood cultures in newborn infants. *Pe-
16. Lark RI, Saint S, Chenoweth C, Zemenuck Jk, Lipsky BA, Plorde J. Four-year prospective evaluation of community-acquired bacteremia: epidemiology, microbiology, and patient outcome. *Diagn Microbiol Infect Dis* 2001; 41:15–22.

17. Miller LG, Mathisen GE, Chang S. Staphylococcus aureus meningitis in a patient with acquired immunodeficiency syndrome. *Mayo Clin Proc* 1998;73: 1083–4.

18. Bhutta ZA, Yusuf K. Neonatal sepsis in Karachi: factors determining outcome and mortality. *J Trop Pediatr* 1997; 43: 65–70.

19. Joshi SJ, Ghole VS, Niphadkar KB. Neonatal gram negative bacteremia. *Indian J Pediatr* 2000; 67: 27–32.

20. Berkley JA, Lowe BS, Mwangi I, Williams T, Bauni E, Mwarumba S, et al. Bacteremia among Children Admitted to a Rural Hospital in Kenya. *N Engl J Med* 2005; 352: 39–47.

21. El-Nasser Z and Awad F. Prevalence and pattern of antimicrobial susceptibility of methicillin-resistant and methicillin-sensetive staphylococcus aureus in north Jordan. *J M J* 2009;43: 15-20.

22. Blomberg B, Manji KP, Urassa WK, Tamim BS, Mwakagile DS, Jureen R, et al. Antimicrobial resistance predicts death in Tanzanian children with bloodstream infections: a prospective cohort study. *BMC Infect Dis.* 2007 May 22; 7: 43.

23. Orrett FA, Shurland SM. Neonatal sepsis and mortality in a regional hospital in Trinidad: aetiology and risk factors. *Ann Trop Paediatr* 2001; 21: 20–5.

24. Manges AR, Natarajan P, Solberg OD, Dietrich PS, Riley LW: The changing prevalence of drug-resistant Escherichia coli clonal groups in a community: evidence for community outbreaks of urinary tract infections. *Epidemiol Infect* 2006; 134: 425–31.

25. Bataineh. H and Alrashed K. Resistant gram-negative bacilli and antibiotic consumption in Zarqa, Jordan. *Pak J Med Sci* 2007; 1: 59-63.

26. Aurangzeb B, Hameed A. Neonatal sepsis in hospital-born babies: bacterial isolates and antibiotic susceptibility patterns. *J Coll Physicians Surg Pak.* 2003; 13: 629–32.