Contribution of oxytocin receptor polymorphisms to amygdala activation in schizophrenia spectrum disorders

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Background
Oxytocin has been proposed to mediate amygdala dysfunction associated with altered emotion processing in schizophrenia, but the contribution of oxytocin pathway genes is yet to be investigated.

Aims
To identify potential differential contributions of three oxytocin receptor polymorphisms (rs53576, rs237902 and rs2254298) between patients with schizophrenia spectrum disorders (SCZ), affective spectrum disorders (AD) and healthy controls (HC).

Method
In a total of 346 participants (104 with SCZ, 100 with AD, and 142 HC) underwent genotyping and functional magnetic resonance imaging (fMRI) during an emotional faces matching paradigm. Genetic association analyses were performed to test the possible effects on task-induced BOLD amygdala response to fearful/angry faces.

Results
In participants with SCZ, the rs237902 G allele was associated with low amygdala activation (left hemisphere: $b = -4.99$, Bonferroni corrected $P = 0.04$) and interaction analyses showed that this association was disorder specific (left hemisphere: Bonferroni corrected $P = 0.03$; right hemisphere: Bonferroni corrected $P = 0.03$). There were no associations between oxytocin polymorphisms and amygdala activation in the total sample, among AD patients or HC.

Conclusions
Rs237902 was associated with amygdala activation in response to fearful/angry faces only in patients with SCZ. Our findings indicate that the endogenous oxytocin system could serve as a contributing factor in biological underpinnings of emotion processing and that this contribution is disorder specific.

Declaration of interest
O.A.A. received speaker’s honoraria from GSK, Otsuka, Lundbeck.

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Social cognitive deficits, including deficits in emotion processing, precede onset of illness, are stable impairments over time and important predictors for functional disabilities in patients with schizophrenia.1,2 Add-on trials with oxytocin administered intra-nasally in this patient group suggest a treatment effect on these deficits as well as a modulating effect on amygdala response to facial emotion processing.3,4 Trials from healthy subjects show an effect on amygdala activation in a possible sexual dimorphic pattern, but the diverse direction of effects seen are not yet established.5-7 The results from oxytocin trials are inconsistent in general,3,4,8,9 and this inconsistency has been attributed to individual differences, including genetically based differences in the oxytocin system,8,9,10 and a poor understanding of drug targets and mechanisms.11 The field is also troubled by small sample sizes with an average sample size in schizophrenia and oxytocin studies to date, consisting of patients with schizophrenia spectrum (SCZ) and affective spectrum disorders (AD) as well as a community representative sample of healthy participants (HC), specifically targeting the SNPs: rs53576, rs2254298 and rs237902. As emotion stimuli processing is particularly altered in patients with schizophrenia, we here hypothesised that the contribution of the three SNPs would be more pronounced in the SCZ group. Because of the reported sex differences in the response to exogenous oxytocin, we additionally investigated interaction effects between SNPs and sex in the case of significant associations between SNPs and amygdala response.

Method

Sample characteristics
Participants, all of Caucasian origin, were included in the Norwegian multicentre Thematically Organized Psychosis (TOP) research study recruiting patients from in- and out-patient clinics in the greater Oslo area in south-eastern Norway. None of the participants had a history of head injury, neurological disorder, autoimmune or infectious disorders or malignancies, or an IQ below 70. Patient assessments included clinical and physical examinations by a physician, neuropsychological testing by a psychologist, collection of blood samples for somatic screening...
The Structured Clinical Interview for DSM-IV Axis I Disorders (SCID-I) was used for diagnostic purposes. The sample of 104 patients with SCZ included the following diagnoses: schizophrenia (n=62), schizophreniform disorder (n=6), schizoaffective disorder (n=12) and psychotic disorder not otherwise specified (NOS) (n=24). The AD group (n=100) consisted of patients with bipolar disorder I (BD I, n=50), bipolar disorder II (BD II, n=38), bipolar disorder NOS (BD NOS, n=6) and depressive psychosis (n=6) (Table 1). The healthy controls (HC) (n=142) were invited via stratified random selection from statistical records of persons from the same catchment area as the patient groups and excluded in case of the presence of severe mental disorders, including schizophrenia and AD in the participant and in first-degree relatives, or current illegal substance use. The study received approval from the Norwegian Scientific-Ethical Committees and the Norwegian Data Protection Agency, and all participants provided written informed consent.

Genotyping

DNA was extracted from blood and rs237902 was genotyped using the Affymetrix Human SNP Array 6.0 (Affymetrix Inc., Santa Clara, CA, USA), as previously reported. All chips were subjected to the Birdseed-v2 algorithm developed by Affymetrix Inc. and Broad Institute and implemented in the software Affymetrix to the Birdseed-v2 algorithm developed by Affymetrix Inc. and European. The markers did not show deviation from Hardy-Weinberg equilibrium (P>0.001), low-yield (<95%) or minor allele frequencies below 0.01. Following the above-mentioned quality control, rs33576 and rs2254298 were imputed with MaCH (http://www.sph.umich.edu/csg/abecasis/MACH/download/1000G-Phase1-Interim.html) using the European samples in the Phase I release of the 1000 Genomes Project.

Experimental paradigm

A widely used and validated paradigm was employed to elicit amygdala activation. For DSM-IV Axis I Disorders (SCID-1) was used for diagnostic purposes. The sample of 104 patients with SCZ included the following diagnoses: schizophrenia (n=62), schizophreniform disorder (n=6), schizoaffective disorder (n=12) and psychotic disorder not otherwise specified (NOS) (n=24). The AD group (n=100) consisted of patients with bipolar disorder I (BD I, n=50), bipolar disorder II (BD II, n=38), bipolar disorder NOS (BD NOS, n=6) and depressive psychosis (n=6) (Table 1). The healthy controls (HC) (n=142) were invited via stratified random selection from statistical records of persons from the same catchment area as the patient groups and excluded in case of the presence of severe mental disorders, including schizophrenia and AD in the participant and in first-degree relatives, or current illegal substance use. The study received approval from the Norwegian Scientific-Ethical Committees and the Norwegian Data Protection Agency, and all participants provided written informed consent.

Genotyping

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Experimental paradigm

A widely used and validated paradigm was employed to elicit amygdala activation. In this task, participants selected which of two stimuli (displayed at the bottom of the screen) matched a target stimulus (displayed at the top). In the faces-matching task, the images displayed were human faces expressing either anger or fear (‘negative faces’), or happiness (‘positive faces’). In the sensorimotor control task, geometrical shapes were displayed. The negative and positive faces were presented in two separate runs, each with four blocks of faces interleaved by five blocks of geometrical shapes. The order of the runs was counterbalanced among participants. Within blocks of negative faces, fearful and angry faces were intermixed. Each block of faces consisted of six emotion-specific face trios derived from a standard set of facial-affect pictures. Each trial (face/shape stimulus) was presented for 5.4 s with no inter-stimulus interval, for a total block length of 32.6 s. Each run lasted 294 s. The E-prime software (version 1.0 Psychology Software Tools, Inc, Pittsburgh, PA, USA) controlled the presentations of the stimuli using VisualSystem (NordicNeuroLab, Bergen, Norway). Measures of reaction times and task accuracy were recorded through MRI-compatible Response Grips (NordicNeuroLab, Bergen, Norway).

BOLD fMRI data acquisition

fMRI was performed on a 1.5 T Siemens Magnetom Sonata scanner (Siemens Medical Solutions, Erlangen, Germany) supplied with a standard head coil. We collected 152 functional volumes per run using a T2*-weighted echo-planar imaging (EPI) sequence. Each volume consisted of 24 axial slices with a pixel size of 3 mm in the axial plane and a slice thickness of 4 mm with 1 mm gap between slices (TR=2040 ms, TE=50 ms, FA=90°, matrix 64×64, FOV 224 mm). The first seven volumes and the last one were discarded, leaving 144 volumes for analyses. Prior to fMRI, a sagittal T1-weighted 3D Magnetization Prepared Rapid Gradient Echo (MPRAGE) scan was collected (160 slices, TR=2730 ms, TE=3.93 ms, FA=7°, matrix 192×256, FOV 240 mm, voxel size 1.33×0.94×1 mm³), here used for co-registration purposes.

fMRI data analysis

fMRI data analyses were performed on a 1.5 T Siemens Magnetom Sonata scanner (Siemens Medical Solutions, Erlangen, Germany) supplied with a standard head coil. We collected 152 functional volumes per run using a T2*-weighted echo-planar imaging (EPI) sequence. Each volume consisted of 24 axial slices with a pixel size of 3 mm in the axial plane and a slice thickness of 4 mm with 1 mm gap between slices (TR=2040 ms, TE=50 ms, FA=90°, matrix 64×64, FOV 224 mm). The first seven volumes and the last one were discarded, leaving 144 volumes for analyses. Prior to fMRI, a sagittal T1-weighted 3D Magnetization Prepared Rapid Gradient Echo (MPRAGE) scan was collected (160 slices, TR=2730 ms, TE=3.93 ms, FA=7°, matrix 192×256, FOV 240 mm, voxel size 1.33×0.94×1 mm³), here used for co-registration purposes.

Table 1 Demographic and clinical background of the study sample

|                  | SCZ n=104 | AD n=100 | HC n=142 | Total n=346 | F or χ²  | P     |
|------------------|-----------|----------|----------|-------------|---------|-------|
| Gender (m/f)     | 63/41     | 43/57    | 84/58    | 190/156     | χ²=8.1  | 0.02  |
| Age, years (mean ± s.d.) | 31.5 (8.9) | 33.2 (11.2) | 33.4 (8.4) | 32.8 (9.5) | F=1.44  | 0.24  |
| Left amygdala activation (COPE) (mean ± s.d.) | 11.6 (13.7) | 12.3 (13.1) | 11.0 (12.9) | 11.6 (12.9) | F=0.73  | 0.69  |
| Right amygdala activation (COPE) (mean ± s.d.) | 12.0 (15.4) | 12.6 (13.3) | 12.3 (11.7) | 12.3 (13.4) | F=0.19  | 0.91  |
| Medicated (%)    | 91 (87.5) | 80 (78)  | –        | –           | –       | –     |
| ALTE/AD/AD/HYP (%) | 88/1/23/12 | 47/10/37/13 | –        | –           | –       | –     |
| Rs2254298 (ma/f) | 0.11      | 0.19     | 0.10     | 0.10        | F=0.15  | 0.70  |
| Rs35376 (ma/f)  | 0.29      | 0.28     | 0.32     | 0.30        | F=0.03  | 0.87  |
| Rs2237902 (ma/f) | 0.31      | 0.42     | 0.35     | 0.36        | F=0.03  | 0.87  |
| Behavioural data | –        | –        | –        | –           | –       | –     |
| Response time (ms) (mean ± s.d.) | 1192.8 (314.9) | 1197.6 (339) | 1009.9 (230.9) | 1141 (295.3) | F=5.83  | 0.004 |
| Accuracy rate (%) (mean ± s.d.) | 98.7 (0.06) | 98.5 (0.09) | 99.2 (0.03) | 98.9 (0.06) | F=0.29  | 0.87  |

SCZ, schizophrenia spectrum disorders; AD, affective spectrum disorders; HC, healthy controls; COPE, contrast parameter estimates; ma/f, male/female ratio; fMRI, functional magnetic resonance imaging; MPRAGE, magnetization prepared rapid gradient echo; ALTE, alcohol and other psychoactive substances; AD, affective disorders; HYP, hypomania; F, Fisher’s exact test; χ², chi-square test; COPE, contrast parameter estimates; ma/f, male/female ratio.

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and a high-pass filter with a 90-s window. A first-level general linear model (GLM) analysis was performed for each run, where the onset and duration of the on-blocks (positive faces and negative faces, respectively) was modelled with that of the off-blocks (geometrical shapes) as implicit baseline. The design matrix was filtered and convolved with a haemodynamic response function before the model fit. This analysis resulted in individual contrast parameter estimates (COPEs) reflecting the faces versus shapes contrast within runs. In the current study, we focused on participants’ amygdala responses to negative stimuli (the negative faces > shapes contrast), because of the implication of altered threat-related facial affect perception in psychotic disorders, and since this contrast has been reported to reliably engage the amygdala.27,35,36 Amygdala regions of interest (ROIs) were defined in accordance with the probabilistic Harvard–Oxford subcortical atlas provided with FSL, and were thresholded at 25% probability. Mean BOLD signal changes (parameter estimates) across all ROI voxels were extracted from FSL and used in statistical analyses.

### Results

#### Group analyses of behavioural data and amygdala activation

We did not detect differences in amygdala activation (left hemisphere: $\chi^2=0.73$, $P=0.69$; right hemisphere: $\chi^2=0.19$, $P=0.91$) or accuracy rate ($\chi^2=0.29$, $P=0.87$) between diagnostic subgroups, but in response time ($\chi^2=13.6$, $P=0.001$) (Table 1). There were no main effects of sex on amygdala activation in the total sample (left hemisphere: $\chi^2=2.31$, $P=0.026$), (right hemisphere: $\chi^2=1.17$, $P=0.24$), COPE values from the right and left amygdala showed a strong correlation ($\rho=0.67$, $P<0.01$).

#### Genetic associations analyses

Among patients with SCZ, rs237902 was significantly associated with left amygdala activation, whereas the rs237902 G allele displayed increased risk of low amygdala activation ($P=0.014$, Bonferroni corrected $P=0.04$) (Table 2, Fig. 1). An interaction

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### Table 2: Association analyses between amygdala activity in the Negative Faces > Shapes contrast and oxytocin receptor polymorphisms

| SNP          | Amygdala hemisphere | SCZ n=104 | AD n=100 | HC n=142 | Total N=346 |
|--------------|---------------------|-----------|----------|----------|-------------|
|              | Effect size         | Effect size | Effect size | Effect size | Effect size |
| Rs2254298 (G) | Left                | b=0.12 | 0.97     | b=−1.35 | 0.67     | b=1.92 | 0.40     | b=0.72 | 0.66 |
|              | Right               | b=−1.11 | 0.77     | b=4.24  | 0.20     | b=0.45 | 0.84     | b=1.22 | 0.47 |
| Rs53576 (G)  | Left                | b=4.73  | 0.002    | b=0.37  | 0.87     | b=−3.36 | 0.06     | b=0.35 | 0.77 |
|              | Right               | b=3.28  | 0.19     | b=0.69  | 0.77     | b=−1.14 | 0.50     | b=0.77 | 0.54 |
| Rs237902 (G) | Left                | b=−4.99 | 0.014*   | b=−0.25 | 0.90     | b=2.81 | 0.08     | b=−0.56 | 0.59 |
|              | Right               | b=3.84  | 0.097    | b=−2.66 | 0.18     | b=2.65 | 0.08     | b=−1.00 | 0.36 |

SNP, single nucleotide polymorphism; SCZ, schizophrenia spectrum disorders; AD, affective spectrum disorders; HC, healthy controls.

* Results for the G allele dosage are displayed.
* Remains significant after Bonferroni corrections (3 independent tests).

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### Statistics

#### Group analyses of amygdala activation and behavioural data

Potential differences in amygdala activation, response time and accuracy rate between subgroups (SCZ, AD, and HC) were examined. The distributions were not normal within all subgroups, hence Kruskal–Wallis tests were performed. Sex effects for amygdala activation were tested for in the total sample with Mann–Whitney tests because of values without normal distribution across gender. The correlation between right and left amygdala activation was examined using bivariate correlation analyses (Spearman’s rho). These analyses were performed in SPSS version 22 (IBM, Armonk, NY).

#### Genetic associations analyses

The additive effects of allele dosage on right and left amygdala activation in the total group were investigated for rs237902 (genotyped SNP), as well as rs53576 and rs2254298 (imputed SNPs) with age, sex and diagnosis as covariates in linear regression models. In the follow-up analyses, the genetic association tests for all the three SNPs were repeated in the three subgroups separately, with age and sex as covariates (PLINK Version 1.07). In the case of significant genetic association with amygdala activation, interaction analyses between SNP and diagnosis as well as between SNP and sex in linear regression models were performed (R Version 3.2.2). All analyses were performed with and without outliers. The significance threshold was set to $P=0.05/3=0.017$ as a result of Bonferroni corrections for three SNPs. Linkage disequilibrium (LD) analyses across the three SNPs were performed (R Version 3.2.2).

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![Fig. 1](https://www.cambridge.org/core. 07 Jul 2021 at 21:00:39, subject to the Cambridge Core terms of use.)
In this study, we show attenuated amygdala activation in response to emotionally negative faces in rs237902G allele carriers among patients with schizophrenia spectrum disorders (SCZ), while not in patients with affective disorders or healthy control participants. Rs237902 is an OXTR synonymous variant located at exon 3 (https://genome.ucsc.edu/cgi-bin/hgTracks?db=hg19 lastVirtModeType=default&lastVirtModeExtraState=&virtModeType=default&virtMode=0&nonVirtPosition=&position=chr3:8808934-8809434&hgscid=48054839_Izi7M39PmzacwlaTbAUn3la2J6O0), thus with potential influence on protein expression, conformation and function, and relative potential functional and clinical consequences.37 Montag and colleagues found rs237902G to be associated with more negative symptoms in schizophrenia,17 a clinical characteristic found to be correlated with emotion processing and face recognition in a meta-analysis.38 Thus, the main result of the current study is in line with this previous finding. Rs237902 could also serve as a proxy for rs237915 (LD: \( r^2 = 0.82 \)), the single SNP Loth and colleagues found to be associated with activation in the ventral striatum in response to threat perception in 1445 healthy adolescents, investigating 23 tag SNPs in the OXTR.39 The nucleus accumbens in the ventral striatum plays an important role in the mesocorticolimbic system and has been hypothesised to interplay with the amygdala via dopaminergic–oxytocinergic mechanisms.5

The influence of rs53576 and rs2254298, two intronic polymorphisms, has been extensively studied across social features, psychiatric symptoms and ethnic groups; however, the results are mixed in spite of large meta-analyses.19-41 Tost and colleagues showed associations between rs53576 and rs2254298 and amygdala activation in separate studies in 228 healthy individuals using the same fMRI paradigm as the present study;43,44 findings neither we nor Loth and colleagues were able to replicate. While Loth and colleagues suggest that this may be because of their use of a different emotion processing paradigm and inclusion of adolescents instead of adults, our lack of replication might be because of a smaller sample of healthy individuals and the use of a clinical sample that may not show associations to the same OXTR polymorphisms as healthy individuals.

Another possible explanation for ambiguous genetic associations regarding oxytocin pathway genes is potential interaction effects with sex.45 The expression of central oxytocin is steroid-dependent influence on protein expression, conformation and function, thus with potential influence on protein expression, conformation and function, and relative potential functional and clinical consequences.37 Montag and colleagues found rs237902G to be associated with more negative symptoms in schizophrenia,17 a clinical characteristic found to be correlated with emotion processing and face recognition in a meta-analysis.38 Thus, the main result of the current study is in line with this previous finding. Rs237902 could also serve as a proxy for rs237915 (LD: \( r^2 = 0.82 \)), the single SNP Loth and colleagues found to be associated with activation in the ventral striatum in response to threat perception in 1445 healthy adolescents, investigating 23 tag SNPs in the OXTR.39 The nucleus accumbens in the ventral striatum plays an important role in the mesocorticolimbic system and has been hypothesised to interplay with the amygdala via dopaminergic–oxytocinergic mechanisms.5

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Another possible explanation for ambiguous genetic associations regarding oxytocin pathway genes is potential interaction effects with sex.45 The expression of central oxytocin is steroid-dependent.
dependent, the central regulation differs between men and women, and men and women respond differently to exogenous oxytocin. Additionally, several brain imaging studies investigating oxytocin pathway genes in healthy individuals have detected sex-specific effects. However, we were not able to detect any interaction effects between sex and rs237902 in patients with SCZ, and all genetic association analyses included sex as a covariate. Hence, our genetic association results presented are not because of sex differences, and the disorder-specific association with rs237902 did not differ between men and women with SCZ.

As no oxytocin-related polymorphisms have been detected as risk variants for SCZ in large genome-wide association studies (http://www.broadinstitute.org/mpg/ricopoli/), we chose a hypothesis-based approach for SNP selection. Rs53576, rs2254298 and rs237902 are the only SNPs previously found associated with disease characteristics in Caucasian schizophrenia samples, hence, we were interested in these polymorphisms in particular. However, only a few studies have focused on the oxytocin pathway genes in patients with schizophrenia, and the possibility of publication bias must be taken into account. The clinical consequences and functions of these variants are also as yet unknown, and their influence on brain function should be interpreted with caution.

To conclude, we found a disorder-specific contribution of rs237902 to amygdala activation during emotional face processing in patients with SCZ. This is, to our knowledge, the first fMRI study investigating oxytocin pathway polymorphisms in this patient group. Our findings suggest that the endogenous oxytocin system contributes to biological underpinnings of emotion processing, that this contribution differs between patients with SCZ and HC and that exonic regions in OXTR are of interest for future research. Future research should investigate additional oxytocin pathway polymorphisms as well as involving other brain circuits, to better understand the biological mechanisms underlying emotion processing. We encourage replication of our findings in further imaging studies and meta-analyses of oxytocin pathway genes in this patient group.

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