Review Article

Protective Effect and Mechanism of Traditional Chinese Medicine on Myocardial Ischemia Reperfusion Injury

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Received 2 February 2022; Revised 24 March 2022; Accepted 25 March 2022; Published 31 March 2022

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After acute myocardial infarction, early restoration of myocardial perfusion by thrombolysis or percutaneous coronary intervention is the most effective way to reduce the size of myocardial infarction and improve clinical outcomes. However, recovery of blood flow to the ischemic myocardium may cause ischemia-reperfusion (I/R) injury, a phenomenon that instead reduces the efficacy of myocardial reperfusion. Traditional Chinese medicine (TCM) has long been used for the treatment of cardiovascular diseases and has shown remarkable efficacy. Many studies have shown that some TCMs and their active components can exert protective effects against myocardial I/R injury through different mechanisms. This review summarized the protective mechanisms and current research advances of TCMs in myocardial I/R injury.

1. Introduction

Ischemia-reperfusion (I/R) injury refers to the oxygen imbalance, functional disorder, and local tissue damage of nonblood supply tissue after tissue ischemia caused by various reasons (trauma, surgery, vascular obstruction, and so on). Although the blood perfusion at the ischemic site can be restored in time and effectively, it can cause further tissue damage. This may be related to mechanisms such as a burst of oxygen free radicals, rapid restoration of physiological pH, insulin resistance, intracellular calcium overload, mitochondrial damage, and inflammatory response. In clinical practice, this phenomenon is also common after thrombolysis or percutaneous coronary intervention. This may be related to oxygen free radical burst, rapid recovery of physiological pH, insulin resistance, intracellular calcium overload, mitochondrial damage, or inflammatory response [1, 2]. The mechanism is shown in Figure 1.

Insufficient oxygen supply during myocardial ischemia leads to the destruction of the balance between the intracellular oxidation and antioxidant system [3]. When blood is refluxed, the antioxidant system is difficult to resist the generation of a large number of oxygen free radicals, resulting in the damage of intracellular protein, lipid, and genetic material [4]. The cells then develop energy metabolism disorder and stress state.

Due to the increase of glycolysis during ischemia and hypoxia, the energy supply of cardiomyocytes is insufficient, resulting in the reduction of calcium ions transported outside the cells [5]. In addition, reactive oxygen species (ROS) causes the destruction of cell membrane and the influx of extracellular calcium ions, leading to calcium overload in cells [6]. Studies have shown that there is a synergistic effect between the excessive production of ROS and intracellular calcium overload [7], both of which can induce the excessive opening of mitochondrial permeability transition pore (MPTP), resulting in apoptosis via the mitochondrial pathway [8].

Besides that, the myocardial I/R process is accompanied by the production of inflammatory substances on the myocardial cell membrane [9]. Large amount of tumor necrosis factor-α (TNF-α) and inflammatory factors such as interleukin-1 (IL-1) mediate neutrophil migration and adhesion to vascular endothelium and myocardial tissue [10, 11], which damage cardiomyocytes and vital organelles (mitochondria), and cause autophagy of endothelial cells and microvascular plugging [12, 13], leading to cardiac dysfunction and hemodynamic impairment.
Oxidative stress, calcium overload, mitochondrial damage, inflammatory reaction, and other mechanisms affect the normal metabolism, division and appreciation, biosynthesis, absorption and excretion, information transmission, and other functions of cells, thus opening the journey of cell death. Apoptosis and autophagy are already turned on when chromatin condensation, nuclear fragmentation, and excessive degradation of organelles occurring within cardiomyocytes and dead cells are cleared [14]. The two not only share some signal pathways but also regulate each other through induction/inhibition factors [15], which eventually leads to irreversible myocardial injury.

Traditional Chinese medicine (TCM) treatment for myocardial I/R injury has not only a long history but also remarkable efficacy and is able to improve the tissue structure and function damage of cardiomyocytes formed by ischemia and hypoxia. Many TCM monomers, components, and prescriptions have been used to prevent and treat myocardial I/R injury. Table 1 provides different protective effects and mechanisms, such as inhibiting oxidative stress (Panax notoginseng saponins, total glucosides of paeony, and Shexian Shengmai oral liquid, 5 (S)-5-carboxystrictosidine), promoting the recovery of calcium homeostasis (Elatoside C and Aralosides), inhibiting inflammatory reaction (Diosgenin, celestrol, and ophiopogonin D), reducing mitochondrial damage (calendula E and Asiatic acid), improving the energy metabolism (ginsenoside Rb1 and Panax notoginseng saponins), inhibiting apoptosis (Cryptotanshinone, Ginsenoside Rb1, ophiopogonin D, withaferin A, and salidroside), regulating autophagy (Formononetin and Paeonol), and improving microvascular function (Tongxinluo).

This review mainly discussed the TCM or its active components used to treat myocardial I/R injury and summarized their main roles and potential mechanisms in myocardial I/R protection.

2. Protective Effects and Mechanisms of TCM in Myocardial I/R Injury

Traditional Chinese medicine believes that the pathological mechanism of myocardial I/R injury lies in Qi deficiency, blood stasis, and phlegm dampness, and the treatment concept of supplementing Qi and nourishing blood, promoting blood circulation, and removing blood stasis can just contribute to the improvement of cardiac function after blood reflow [39]. Many studies have shown that treatment of myocardial I/R injury with TCM reduces infarct size, improves hemodynamic index, and attenuates pathological damage to myocardial tissue [28, 32], and the main mechanisms are as follows.

2.1. Inhibit Oxidative Stress. Reperfusion can generate a large number of ROS, which are involved in cardiomyocyte I/R injury through many pathways, such as damaging cell membranes, organelles, and ion channels. Not only that, ROS consumes a large amount of intracellular antioxidant substances thereby aggravating the cellular stress response, leading to myocardial dysfunction [40]. Some TCMs can increase the antioxidant capacity of cardiomyocytes, thereby ameliorating the changes in ventricular pressure after injury and regulating coronary blood flow. It is reported that Panax notoginseng saponins can promote the expression of miR-30c-5p, downregulate the expression of lactate dehydrogenase (LDH) and p53 protein in cells, inhibit the release of malondialdehyde (MDA) in injured cells [17], and then reduce the level of oxidative stress in cells [16].

Yi et al. [18] demonstrated increased ROS generation in cardiomyocytes of reperfused mice by the DHE staining method, while ROS generation was significantly inhibited by pretreatment with asiatic acid. Total glucosides of paeyony (TGP) can inhibit the expression and activity of oxidative stress-related substances and promote levels of antioxidant
substances. After treatment with TGP, the levels of intracellular ROS and activities of LDH and MDA were decreased, while the activities of superoxide dismutase (SOD) and glutathione peroxidase (GSH-px) were increased, and the mechanism may be related to the inhibition of the PI3K/Akt signaling pathway [19].

Other TCMs have shown similar effects to TGP, such as aralosides [20], elatoside C [21], Shenxian Shengmai oral liquid [22], 5 (S)-5-carboxystrictosidine [23] and withaferin A [24]. 5 (S)-5-Carboxystrictosidine decreased intracellular LDH levels, while it increased SOD and catalase contents in I/R cardiomyocytes of rats, thus avoiding excessive elevation of left ventricular end diastolic pressure and reducing infarct size, and the mechanism may be related to the regulation of mitochondrial KATP channels. Whether hydrogen peroxide is used to induce I/R injury or not, Withaferin A can promote the expression of antioxidants in H9C2 cells and inhibit the production of ROS. This antioxidant effect can be significantly inhibited by the Akt inhibitor.

### 2.2. Promote the Recovery of Calcium Homeostasis

I/R injury leads to calcium overload in cardiomyocytes, resulting in myocardial systolic and diastolic dysfunction. Examination of hemodynamic indexes can observe a decrease in left ventricular systolic pressure (LVSP) and an increase in left ventricular end diastolic pressure (LVEDP) in mice subjected to I/R [20]. It has been shown that pretreatment with elatoside C is able to maintain a normal calcium concentration in cardiac diastole and significantly attenuate cardiac mechanical dysfunction resulting from abnormal calcium changes. The mechanism may be related to PI3K/Akt, ERK1/2, and JAK2/STAT3 pathways [21]. Wang et al. [20] pretreated rat myocardium with different doses of aralosides and found that LVEDP decreased with the increase of drug concentration, but there was no significant change in LVSP. Further studies revealed that it promoted calcium ion transport inside and outside the cell due to enhanced activities of Na⁺-K⁺-ATPase and Ca²⁺-Mg²⁺-Mg²⁺-ATPase, which in turn played a role in protecting the heart.

| TCM or its ingredients | Experiment object | Protective effects | Potential mechanisms | Ref. |
|------------------------|-------------------|-------------------|---------------------|------|
| Panax notoginseng saponins | H9C2 cells | Inhibit oxidative stress | miR-30c-5p pathway | [16] |
| Asiatic acid | Male C57BL/6 mice | Reduce mitochondrial damage | p38 MAPK pathway | [18] |
| Total glucosides of paony | H9C2 cells | Inhibit oxidative stress | PI3K/Akt pathway | [19] |
| Aralosides | SD rats | Promote the recovery of calcium homeostasis | — | [20] |
| Elatoside C | SD rats | Promote the recovery of calcium homeostasis | PI3K/Akt pathway | [21] |
| Shenxian Shengmai oral liquid 5 (S)-5-carboxystrictosidine | SD rats | Inhibit oxidative stress | — | [22] |
| Withaferin A | H9C2 cells | Inhibit oxidative stress | Mitochondrial KATP pathway | [23] |
| Diosgenin | SD rats | Inhibit inflammatory reaction | p38 MAPK pathway | [25] |
| Celastrol | H9C2 cells | Inhibit inflammatory reaction | NF-κB pathway | [26] |
| Ophiopogonin D | SD rats | Inhibit inflammatory reaction | NF-κB pathway | [27] |
| Calendula E | SD rats | Reduce mitochondrial damage | AMPK pathway | [28] |
| Ginsenoside Rb1 | SD rats | Improve energy metabolism | RhoA pathway | [29] |
| Ginsenoside Rb1 | SD rats | Inhibit apoptosis | mTOR pathway | [30] |
| Cryptotanshinone | — | Inhibit apoptosis | PI3K/Akt pathway | [24] |
| Salidroside | H9C2 cells | Inhibit apoptosis | PERK pathway | [32] |
| Araloside C | H9C2 cells | Inhibit apoptosis | PERK/eIF2α pathway | [33] |
| Formononetin | Aged male mice | Regulate autophagy | — | [34] |
| Paenol | H9C2 cells | Regulate autophagy | — | [35] |
| Tongxinluo | Human cardiac microvascular endothelial cells | Improve microvascular function | PPARα/ANGPTL4 pathway | [36] |
| Tongxinluo | SD rats | Improve microvascular function | PKA pathway | [37, 38] |
2.3. Inhibit Inflammatory.

Inflammatory reaction is an important pathophysiological mechanism in myocardial I/R injury [41]. Studies have shown that activation of neutrophils mediates myocardial injury in I/R injury, whereas NF-κB played a key role in the production of inflammatory factors and the regulation of leukocytes. H&E staining showed that the cardiomyocytes pretreated with diosgenin had almost no inflammatory cell infiltration after blood reflow and the infarct area decreased significantly. Compared with the I/R group, NF-κB p65 phosphorylation in cardiomyocytes was inhibited, and tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), and the level of myocardial markers in rat serum decreased significantly [25]. During reperfusion, cardiac microvessels are the main damaged targets. A large number of inflammatory cell infiltration not only leads to microembolism and circulatory disorders [42] but also the release of inflammatory factors that leads to apoptosis, necrosis, and gene mutation of microvascular endothelial cells [12], which further aggravates I/R injury. Celastrol is a kind of traditional Chinese medicine, which has the effect of dispelling wind and activating blood circulation. It has been proved to have a certain therapeutic effect on cardiovascular diseases. Li et al. [26] demonstrated that low-dose celastrol inhibited NF-κB activation, prevented inflammatory factor-related gene transcription, and protected injured vascular endothelium in vitro hypoxia/reoxygenation models.

Epoxycosatetraenoic acid (EET) is a metabolite of arachidonic acid (AA) [43], which is involved in the processes of cellular inflammatory reaction, oxidative stress, and apoptosis. It acts on the same signaling pathway as celastrol to inhibit angiotensin II-induced endothelial injury [44]. Ophiopogonin D can promote the expression of cytochrome P450 cyclooxygenase in cardiovascular tissue, which is the main substance involved in the AA metabolism [45], so as to enhance the expression of EETs and inhibit inflammatory response. This mechanism is mediated by the PI3K/Akt/eNOS signaling pathway [27].

2.4. Reduce Mitochondrial Damage.

Mitochondrion, as the hub of the cell energy metabolism, is an important part of maintaining cell activity and function. In the stage of myocardial ischemia-reperfusion, the increase of reactive oxygen species, calcium overload, and inflammatory reaction caused by blood reflow lead to mitochondrial damage, activate mitochondrial autophagy, inhibit cardiomyocyte viability, and damage cardiac function. As the most vulnerable target organelle during endogenous oxidative stress, mitochondria will appear vacuoles, even lysis or autophagy during reperfusion [18].

Drp1, OPAL, and Mfn1/2 are important proteins that regulate mitochondrial dynamics. The former promotes membrane fission, and the latter two mediate inner and outer membrane fusions [46, 47]. In the study of Wang et al. [28], calendula E can effectively increase the expression of Mfn1/2 and OPAL protein in cardiomyocytes and meanwhile reduce the expression of Drp1 and reverse mitochondrial division. Asiatic acid can stabilize mitochondrial morphology and protect membrane structure from damage by regulating endogenous oxidative stress [18].

2.5. Improve Energy Metabolism. The impairment of the energy metabolism in cardiomyocytes is the result of multiple mechanisms of I/R injury and is also key to driving the further progression of injury. During ischemia-reperfusion, mitochondrial damage and insufficient oxygen supply lead to increased glycolysis and reduced ATP production. Although the enhanced fatty acid metabolism can produce more ATP, it aggravates cell hypoxia [48, 49]. ATP 5D is a subunit of ATP synthase. Studies have shown that its expression decreased in I/R-injured cardiomyocytes. Ginsenoside Rb1 treatment is able to promote the transcription of this subunit, allowing the activity of ATP synthase to increase, which in turn improves the cardiomyocyte metabolism [29]. There are studies showing that this effect is associated with the regulation of the RhoA signaling pathway [50].

Panax notoginseng saponins can not only reduce the damage caused by oxidative stress but also regulate the energy metabolism of cardiomyocytes. Through protein spectrum analysis, Zhao et al. [17] observed that Panax notoginseng saponins changed the expression of proteins related to tricarboxylic acid cycle. Because these proteins participate in the formation of important coenzymes in the process of aerobic oxidation, they effectively improve the energy supply of cells and strive for more time for cardiomyocyte repair.

2.6. Inhibit Apoptosis. It is well known that apoptosis is an important pathophysiological mechanism of myocardial I/R injury [51]. Many TCMs exert cardiomyocyte protective effects by inhibiting apoptosis. Salvia miltiorrhiza has the effect of promoting blood circulation and removing blood stasis. It is often used as medicine for cardiovascular diseases and has a certain therapeutic effect on myocardial I/R injury. Cryptotanshinone is a compound with pharmacological activity extracted from Salvia miltiorrhiza. Wang et al. [31] confirmed that the myocardial protective effect of cryptotanshinone was achieved by promoting the expression of MAKP and reversing cardiomyocyte apoptosis through biological experiments in vivo and in vitro. The rapamycin target protein (mTOR) pathway is an important pathway regulating growth and metabolism. It can inhibit apoptosis after being activated. Ginsenoside Rb1 was able to protect cardiomyocytes by activating the PI3K/Akt/mTOR pathway via phosphorylation as shown by Western blot analysis, which was abolished after the application of rapamycin [30]. Ophiopogonin D has multiple biological functions, and studies have shown its ability to inhibit apoptosis by regulating the caspase pathway through arachidonic acid metabolites [27]. Similar to ophiopogonin D, withaferin A improves I/R-injured H9C2 cells viability by inhibiting caspase activity [24].

Some TCMs can reduce the expression of apoptotic protein and increase the expression of antiapoptotic protein. Shen et al. [19] found that total glucosides of paony (TGP)
downregulated the expression of apoptosis-related factors by the apoptosis detection kit, while increasing the levels of procaspase-3 and antiapoptotic protein Bcl-2. Flow cytometry showed that TGP could reduce the apoptosis rate by 13.73%. Salidroside was also shown to have the similar efficacy in a study by Sun et al. [32]. It acts through PERK and IRE1α pathway that regulates apoptosis-related proteins, thereby reversing endoplasmic reticulum stress to achieve the effects of reducing I/R-induced cytotoxicity, alleviating morphological changes of apoptosis, and increasing cell viability.

Apoptosis is related to the opening of mitochondrial permeability transition pore (mPTP). When a large number of ROS are produced in cells, calcium ions are overloaded, ATP synthesis is insufficient, and the latter is largely opened [52], followed by activation of the glycolgen synthesis kinase 3β (GSK-3β) pathway and protein kinase C (PKC) pathway [49, 53], resulting in mitochondrial dependent apoptosis [52]. Asiatic acid, elatoside C, is able to improve the imbalance of Bcl-2/Bax balance, reduce the expression of apoptosis-related proteins (caspase-9 and caspase-3), and block the mitochondria-dependent apoptotic pathway [18, 21].

Endoplasmic reticulum stress was involved in the pathological process of I/R injury through phosphorylation of PERK and eIF2α. Du et al. [33] showed that I/R injury increased the expression of stress markers in cardiomyocytes, and the same effect could be produced by inducing endoplasmic reticulum stress with tunicamycin (TM). After araloside C intervention, the effect of TM was significantly inhibited, and the expression of endoplasmic reticulum stress-dependent apoptotic proteins (CHOP and caspase-12) decreased. Further studies showed that the above effects were achieved through the increased expression of heat shock protein 90.

2.7. Regulate Autophagy. Autophagy contributes to the digestion and degradation of abnormal organelles and cell contents. Many evidences show that increased autophagy can inhibit the increase of necrotic organelles and improve the quality of intracellular protein in cells, so as to protect cells from I/R injury [54–57]. Huang et al. [34] showed that although I/R could increase the number of lysosomes in cells, it could not play a protective role because lysosomes were in an immature state. Formononetin was able to significantly reduce I/R infarct size and postinjury cardiac function by promoting the lysosomal switch from immature to mature phenotype via acidification. However, excessive autophagy will also lead to excessive protein consumption, burden cells, and aggravate myocardial injury caused by I/R.

Beclin-1, as a key protein in autophagy regulation, participates in a variety of pathophysiological processes of myocardial I/R injury. It is able to accelerate autophagy progress; it can also be lost by caspase breakdown instead to promote apoptosis. When Bcl-2 binds to it, autophagy and apoptosis are both inhibited [58, 59]. Tsai et al. [35] found that paeonol significantly reduced the incidence and time of arrhythmia after I/R injury. The level of myocardial markers in serum and the myocardial infarction area decreased in the paeonol pretreatment group. The specific mechanism may be related to that paeonol can regulate the level of Bcl-2 in cardiomyocytes and inhibit excessive autophagy and apoptosis at the same time.

2.8. Improve Microvascular Function. As an important structure of myocardial perfusion, cardiac microvascular endothelial cells are closely arranged around cardiomyocytes to form an endothelial barrier and participate in a variety of pathophysiological processes of cardiomyocytes [60]. The destruction of the integrity of microvascular endothelial barrier and the increase of microvascular permeability will aggravate I/R injury [61, 62]. The increase of angiopoietin-like protein 4 (ANGPTL4) can stabilize the function of vascular endothelial barrier [63]. Peroxisome proliferator-activated receptors-α (PPAR-α) is able to promote the generation of ANGPTL4 [64]. Tongxinluo and PPAR-α have similar bioactive effects which can promote ANGPTL4 expression in microvascular endothelial cells, thereby blocking the increase in endothelial permeability. This is related to the regulation of Tongxinluo on intercellular junction structure (JAM-A, VE-cadherin, and integrin-α5) [36].

Endothelial nitric oxide synthase (eNOS) is an enzyme mainly responsible for NO production in vascular endothelium [65]. The increase of eNOS activity can promote the utilization of NO by cells, thus to protect myocardium from blood reperfusion injury. Studies have shown that Tongxinluo is able to indirectly stimulate eNOS phosphorylation with the aid of extracellular vesicles, and the specific mechanism is related to the PI3K/Akt signaling pathway [37, 38].

3. Protective Effects and Mechanisms of Chinese Medicinal Prescriptions in Myocardial I/R Injury

Doctors of traditional Chinese medicine are good at using different kinds of medicinal materials together to make Chinese medicine prescriptions to achieve the efficacy of one plus one greater than two, which is the traditional Chinese medicine theory of “monarch, miner, assistant, and guide.” Based on this theoretical foundation and combined with the analysis of network pharmacology, Liu et al. [66] constructed a traditional Chinese medicine prescription “Tanyutongzhi” for the treatment of myocardial I/R injury.

The main active components of dried ginger-aconite decoction include aconitine, 6-ginger, and mesaconitine. Feng et al. [67] confirmed that dried ginger-aconite decoction can reduce cardiomyocyte apoptosis through the hypoxia/reperfusion model in vitro and coronary artery ligation model in vivo. In vitro, it increased the antioxidants in H9C2 cells and significantly restored the cell viability. In vivo, it improved the swelling, fiber breakage, and nuclear damage of cardiomyocytes caused by I/R injury.

Huang Qi Tong Bi decoction has the functions of warming Yang and supplementing Qi, nourishing Yin and...
blood, removing blood stasis, and dredging collaterals. It has been used in the treatment of coronary heart disease for a long time. It has been found to inhibit the HMGB1/TLR/NF-κB pathway, thereby effectively reducing expressions of TNF-α, IL-6, and IL-1β in cardiomyocytes, avoiding myocardial damage from neutrophil infiltration [68].

4. Summary

With characteristics of multitarget and multipathway, TCMs show unique advantages in the prevention and treatment of cardiovascular diseases. This study reviews the protective effects of TCMs and their active components on myocardial I/R injury. A variety of TCM components and prescriptions can ameliorate I/R-induced myocardial injury by inhibiting oxidative stress, promoting the recovery of calcium homeostasis, inhibiting inflammatory reaction, reducing mitochondrial damage, improving energy metabolism, inhibiting apoptosis, regulating autophagy, and improving microvascular function. The specific mechanisms involve different signal pathways, such as the MAPK pathway, PKA pathway, AMPK pathway, PI3K/Akt pathway, NF-κB pathway, PERK pathway, and IRE1α pathway. However, the active components and protective mechanism of some TCMs are still unclear, so more experiments and further research studies are needed. Besides these, the use and dosage of TCMs for clinical use also need to be explored by a large number of clinical experiments, in order to obtain better TCM prescriptions and preparations, and consequently expand the application scope of TCM in the treatment of myocardial I/R injury.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Kuo Liu and Demin Liu contributed equally to this study. They are mainly responsible for writing this article. Wei Cui contributed to the design of this study and made important corrections.

References

[1] M.-Y. Wu, G.-T. Yang, W.-T. Liao et al., “Current mecha-nistic concepts in ischemia and reperfusion injury,” Cellu-lar Physiology and Biochemistry, vol. 46, no. 4, pp. 1650–1667, 2018.
[2] R. O. S. Soares, D. M. Losada, M. C. Jordani, P. Évora, and O. Castro-e-Silva, "Ischemia/reperfusion injury revisited: an overview of the latest pharmacological strategies," International Journal of Molecular Sciences, vol. 20, p. 5034, 2019.
[3] S. Cadenas, "ROS and redox signaling in myocardial ischemia/reperfusion injury and cardioprotection," Free Radical Biology and Medicine, vol. 117, pp. 76–89, 2018.
[4] E. Dongó, I. Hornyák, Z. Benkó, and L. Kiss, "The cardioprotective potential of hydrogen sulfide in myocardial ischemia/reperfusion injury (Review)," Acta Physiologica Hungarica, vol. 98, no. 4, pp. 369–381, 2011.
[5] S. Zhu, T. Xu, Y. Luo et al., "Luteolin enhances sarcoplasmic reticulum Ca2+-ATPase activity through p38 MAPK signaling thus improving rat cardiac function after ischemia/reperfu-sion," Cellular Physiology and Biochemistry, vol. 41, no. 3, pp. 999–1010, 2017.
[6] M. Ohtsuka, H. Takano, M. Suzuki et al., "Role of Na+-Ca2+-exchanger in myocardial ischemia/reperfusion injury: evaluation using a heterozygous Na+-Ca2+-exchanger knockout mouse model," Biochemical and Biophysical Research Communications, vol. 314, no. 3, pp. 849–853, 2004.
[7] C. Wang, N. Liu, R. Luan et al., "Apelin protects sarcoplasmic reticulum function and cardiac performance in ischaemia-reperfusion by attenuating oxidation of sarcoplasmic reticulum Ca2+-ATPase and ryanodine receptor," Cardiovascular Research, vol. 100, no. 1, pp. 114–124, 2013.
[8] S. M. Davidson, D. M. Yellon, M. P. Murphy, and M. R. Duchen, "Slow calcium waves and redox changes precede mitochondrial permeability transition pore opening in the intact heart during hypoxia and reoxygenation," Cardiovascular Research, vol. 93, no. 3, pp. 445–453, 2012.
[9] S. E. Boag, E. Andreano, and I. Spyridopoulos, "Lymphocyte communication in myocardial ischemia/reperfusion injury," Antioxidants and Redox Signaling, vol. 26, no. 12, pp. 660–675, 2017.
[10] F. Arslan, D. de Kleijn, L. Timmers, P. Doevendans, and G. Pasterkamp, "Bridging innate immunity and myocardial ischemia/reperfusion injury: the search for therapeutic targets," Current Pharmaceutical Design, vol. 14, no. 12, pp. 1205–1216, 2008.
[11] A. K. Singhal, J. D. Symons, S. Boudina, B. Jaishy, and Y.-T. E. Shiu, "Role of endothelial cells in myocardial isch-emia-reperfusion injury," Vascular Disease Prevention, vol. 7, no. 1, pp. 1–14, 2010.
[12] N. Schanz, C. Bode, and D. Duerschmied, "Platelet contribu-tions to myocardial ischemia/reperfusion injury," Frontiers in Immunology, vol. 10, p. 1260, 2019.
[13] I. Russo, C. Penna, T. Musso et al., "Platelets, diabetes and myocardial ischemia/reperfusion injury," Cardiovascular Diabetology, vol. 16, no. 1, p. 71, 2017.
[14] P. Xia, Y. Liu, and Z. Cheng, "Signaling pathways in cardiac myocyte apoptosis," BioMed Research International, vol. 2016, Article ID 9583268, 22 pages, 2016.
[15] Y. Dong, H. Chen, J. Gao, Y. Liu, J. Li, and J. Wang, "Mo-lecular machinery and interplay of apoptosis and autophagy in coronary heart disease," Journal of Molecular and Cellular Cardiology, vol. 136, pp. 27–41, 2019.
[16] L. Wang, X. Chen, Y. Wang, L. Zhao, X. Zhao, and Y. Wang, "MiR-30c-5p mediates the effects of panax notoginseng sa-ponins in myocardial ischemia/reperfusion injury," BioMed Research International, vol. 2016, Article ID 109963, 2020.
[17] X. Zhao, F. Zhang, and Y. Wang, "Proteomic analysis reveals Xuesaitong injection attenuates myocardial ischemia/reperfu-sion injury by elevating pyruvate dehydrogenase-mediated aerobic metabolism," Molecular BioSystems, vol. 13, no. 8, pp. 1504–1511, 2017.
[18] C. Yi, M. Song, L. Sun et al., "Asiatic acid alleviates myocardial ischemia-reperfusion injury by inhibiting the ROS-mediated mitochondria-dependent apoptosis pathway," Oxidative Medicine and Cellular Longevity, vol. 2022, Article ID 3267450, 16 pages, 2022.
[19] P. Shen, J. Chen, and M. Pan, "The protective effects of total paenony glycoside on ischemia/reperfusion injury in H9C2 cells via inhibition of the PI3K/Akt signaling pathway,"
[20] R. Wang, M. Yang, M. Wang et al., "Total saponins of aralia elata (mii) seem alleviate cardiac homeostasis imbalance and endoplasmic reticulum stress-related apoptosis induced by myocardial ischemia/reperfusion injury," *Cellular Physiology and Biochemistry*, vol. 50, no. 1, pp. 28–40, 2018.

[21] M. Wang, G.-B. Sun, J.-Y. Zhang et al., "Elatoside C protects the heart from ischaemia/reperfusion injury through the modulation of oxidative stress and intracellular Ca²⁺ homeostasis," *International Journal of Cardiology*, vol. 185, pp. 167–176, 2015.

[22] Y. Zhao, X. Zhang, J. Luan et al., "Shenxian-shengmai oral Liquid reduces myocardial oxidative stress and protects myocardium from ischemia-reperfusion injury," *Cellular Physiology and Biochemistry*, vol. 48, no. 6, pp. 2503–2516, 2018.

[23] Y. Han, C. Li, P. Zhang et al., "Protective effects of 5 (S)-5-carboxystrictosidine on myocardial ischemia-reperfusion injury through activation of mitochondrial KATP channels," *European Journal of Pharmacology*, vol. 920, Article ID 174811, 2022.

[24] X. Huang, Y. Wang, Y. Wang, L. Yang, J. Wang, and Y. Gao, "Protective effect of celastrol on myocardial ischemia-reperfusion injury via upregulating CYP2J3/EETs in rats," *Cellular Physiology and Biochemistry*, vol. 49, no. 4, pp. 1646–1658, 2018.

[25] M. Wang, R.-Y. Wang, J.-H. Zhou, X.-H. Xie, G.-B. Sun, and X.-B. Sun, "Calenduloside E ameliorates myocardial ischemia-reperfusion injury through regulation of AMPK and mitochondrial OPA1," *Oxidative Medicine and Cellular Longevity*, vol. 2020, Article ID 2415269, 12 pages, 2020.

[26] Y.-C. Cui, C.-S. Pan, L. Yan et al., "Ginsenoside Rb1 protects against ischemia/reperfusion-induced myocardial injury via energy metabolism regulation mediated by RhoA signaling pathway," *Scientific Reports*, vol. 7, no. 1, Article ID 44579, 2017.

[27] C.-Y. Li, P. Yang, Y.-L. Jiang et al., "Ginsenoside Rb1 attenuates cardiomyocyte apoptosis induced by myocardial ischemia reperfusion injury through the mTOR signal pathway," *Biomedicine & Pharmacotherapy*, vol. 125, Article ID 109993, 2020.

[28] H. Wang, W. Pang, X. Xu, B. You, C. Zhang, and D. Li, "Cryptotanshinone attenuates ischemia/reperfusion-induced apoptosis in myocardium by upregulating MAPK3," *Journal of Cardiovascular Pharmacology*, vol. 77, no. 3, pp. 370–377, 2021.

[29] M. Y. Sun, D. S. Ma, S. Zhao, L. Wang, C. Y. Ma, and Y. Bai, "Salidroside mitigates hypoxia/reoxygenation injury by alleviating endoplasmic reticulum stress-induced apoptosis in H9c2 cardiomyocytes," *Molecular Medicine Reports*, vol. 18, no. 4, pp. 3760–3768, 2018.

[30] Y. Du, M. Wang, X. Liu et al., "Araloside C prevents hypoxia/reoxygenation-induced endoplasmic reticulum stress via increasing heat shock protein 90 in H9c2 cardiomyocytes," *Frontiers in Pharmacology*, vol. 9, p. 180, 2018.

[31] Z. Huang, Y. Liu, and X. Huang, "Formononetin may protect aged hearts from ischemia/reperfusion damage by enhancing autophagic degradation," *Molecular Medicine Reports*, vol. 18, no. 6, pp. 4821–4830, 2018.

[32] X. Huang, Y. Wang, Z. Zhang et al., "Paonol protects against myocardial ischemia/reperfusion-induced injury by mediating apoptosis and autophagy crosstalk," *Frontiers in Pharmacology*, vol. 11, Article ID 586498, 2020.

[33] K. Qi, Y. Yang, Y. Geng et al., "Tongxinluo attenuates oxygen-glucose-serum deprivation/restoration-induced endothelial barrier breakdown via peroxisome proliferator activated receptor-a/angiopoietin-like 4 pathway in high glucose- incubated human cardiac microvascular endothelial cells," *Medicine*, vol. 99, no. 34, Article ID e21821, 2020.

[34] X.-D. Li, Y.-J. Yang, Y.-J. Geng et al., "Tongxinluo reduces myocardial no-reflow and ischemia-reperfusion injury by stimulating the phosphorylation of eNOS via the PKA pathway," *American Journal of Physiology—Heart and Circulatory Physiology*, vol. 299, no. 4, pp. H1255–H1261, 2010.

[35] G. Chen, C. Xu, T. G. Gillette et al., "Cardiomyocyte-derived small extracellular vesicles can signal eNOS activation in cardiac microvascular endothelial cells to protect against Ischemia/Reperfusion injury," *Theranostics*, vol. 10, no. 25, pp. 11754–11774, 2020.

[36] Q. Liu, J. Li, J. Wang, I. Li, J. S. Janicki, and D. Fan, "Effects and mechanisms of Chinese herbal medicine in ameliorating myocardial ischemia-reperfusion injury," *Evidence-Based Complementary and Alternative Medicine*, vol. 2013, Article ID 925625, 14 pages, 2013.

[37] F. He and L. Zuo, "Redox roles of reactive oxygen species in cardiovascular diseases," *International Journal of Molecular Sciences*, vol. 16, no. 11, pp. 27770–27780, 2015.

[38] A. Vincent, B. Lattuca, N. Merlet, C. Sportouch-Dukhan, and S. Barrere-Lemaire, "New insights in research about acute ischemic myocardial injury and inflammation," *Anti-Inflammatory and Anti-Allergy Agents in Medicinal Chemistry*, vol. 12, no. 1, pp. 47–54, 2013.

[39] J. Vinten-Johansen, "Involvement of neutrophils in the pathogenesis of lethal myocardial reperfusion injury," *Cardiovascular Research*, vol. 61, no. 3, pp. 481–497, 2004.

[40] G. J. Gross, A. Hsu, A. W. Pfeiffer, and K. Nithipatikom, "Roles of endothelial nitric oxide synthase (eNOS) and mitochondrial permeability transition pore (MPTP) in epoxycasioinoidic acid (EET)-induced cardioprotection against infarction in intact rat hearts," *Journal of Molecular and Cellular Cardiology*, vol. 59, pp. 20–29, 2013.

[41] X. Huang, Y. Wang, Z. Zhang et al., "Ophiopogonin D and EETs ameliorate Ang II-induced inflammatory responses via activating PPARα in HUVECs," *Biochemical and Biophysical Research Communications*, vol. 490, no. 2, pp. 123–133, 2017.

[42] Y. Ding, P. Tu, Y. Chen, Y. Huang, X. Pan, and W. Chen, "CYP2J2 and EETs protect against pulmonary arterial hypertension with lung ischemia-reperfusion injury in vivo and in vitro," *Respiratory Research*, vol. 22, no. 1, p. 291, 2021.

[43] C. Manechote, S. Palee, S. Kerdpoo, T. Jaiwongkam, S. C. Chattipakorn, and N. Chattipakorn, "Balancing mitochondrial dynamics via increasing mitochondrial fusion attenuates infarct size and left ventricular dysfunction in rats with cardiac ischemia/reperfusion injury," *Clinical Science*, vol. 133, no. 3, pp. 497–513, 2019.
proteins for cardioprotection,” *Journal of Cellular and Molecular Medicine*, vol. 24, no. 12, pp. 6571–6585, 2020.

[48] P. Alegre, L. Mathias, M. A. Loureño et al., “Euterpe oleracea mart. (Açaí) reduces oxidative stress and improves energetic metabolism in myocardial ischemia-reperfusion injury in rats,” *Arquivos Brasileiros de Cardiologia*, vol. 114, no. 1, pp. 78–86, 2020.

[49] E. J. Lesnefsky, Q. Chen, B. Tandler, and C. L. Hoppel, “Mitochondrial dysfunction and myocardial ischemia-reperfusion: implications for novel therapies,” *Annual Review of Pharmacology and Toxicology*, vol. 57, no. 1, pp. 535–565, 2017.

[50] L. Li, C.-S. Pan, L. Yan et al., “ Ginsenoside Rg1 ameliorates rat myocardial ischemia-reperfusion injury by modulating energy metabolism pathways,” *Frontiers in Physiology*, vol. 9, p. 78, 2018.

[51] Y. Wei, L. Ruan, G. Zhou et al., “Local ischemic post-conditioning during primary percutaneous coronary intervention: a meta-analysis,” *Cardiology*, vol. 123, no. 4, pp. 225–233, 2012.

[52] G. Paradis, V. Paradis, F. M. Ruggiero, and G. Petrosillo, “Mitochondrial bioenergetics and cardioplpin alterations in myocardial ischemia-reperfusion injury: implications for pharmacological cardioprotection,” *American Journal of Physiology—Heart and Circulatory Physiology*, vol. 315, no. 5, pp. H1341–H1352, 2018.

[53] P. Makhdoumi, A. Roohbakhsh, and G. Karimi, “MicroRNAs regulate mitochondrial apoptotic pathway in myocardial ischemia-reperfusion-injury,” *Biomedicine and Pharmacotherapy*, vol. 84, pp. 1635–1644, 2016.

[54] J. Chen, J. Gao, W. Sun et al., “Involvement of exogenous H2S in recovery of cardioprotection from ischemic post-conditioning via increase of autophagy in the aged hearts,” *International Journal of Cardiology*, vol. 220, pp. 681–692, 2016.

[55] J. Liu, P. Wu, Y. Wang et al., “Ad-HGF improves the cardiac remodeling of rat following myocardial infarction by upregulating autophagy and necroptosis and inhibiting apoptosis,” *American Journal of Translational Research*, vol. 8, no. 11, pp. 4605–4627, 2016.

[56] C.-L. Wu, C.-H. Chen, C.-S. Hwang, S.-D. Chen, W.-C. Hwang, and D.-I. Yang, “Roles of p62 in BDNF-dependent autophagy suppression and neuroprotection against mitochondrial dysfunction in rat cortical neurons,” *Journal of Neurochemistry*, vol. 140, no. 6, pp. 845–861, 2017.

[57] J. Yi, G. He, J. Yang, Z. Luo, X. Yang, and X. Luo, “Heat acclimation regulates the autophagy-lysosome function to protect against heat stroke-induced brain injury in mice,” *Cellular Physiology and Biochemistry*, vol. 41, no. 1, pp. 101–114, 2017.

[58] R. Kang, H. J. Zeh, M. T. Lotze, and D. Tang, “The Beclin 1 network regulates autophagy and apoptosis,” *Cell Death and Differentiation*, vol. 18, no. 4, pp. 571–580, 2011.

[59] S. Pattingre, A. Tassa, X. Qu et al., “Bcl-2 antiapoptotic proteins inhibit beclin 1-dependent autophagy,” *Cell*, vol. 122, no. 6, pp. 927–939, 2005.

[60] A. Colliva, L. Braga, M. Giacca, and S. Zacchigna, “Endothelial cell-cardiomyocyte crosstalk in heart development and disease,” *The Journal of Physiology*, vol. 598, no. 14, pp. 2923–2939, 2020.

[61] S. Rangasamy, R. Srinivasan, J. Maestas, P. G. McGuire, and A. Das, “A potential role for angiopoietin 2 in the regulation of the blood-retinal barrier in diabetic retinopathy,” *Investigative Ophthalmology and Visual Science*, vol. 52, no. 6, pp. 3784–3791, 2011.