Invasive grass negatively affects growth and survival of an imperiled butterfly

Cale S. Nordmeyer1,2,*, Erik Runquist1, Seth Stapleton1,2

1Conservation Department, Minnesota Zoo, Apple Valley, MN 55124, USA
2Department of Fisheries, Wildlife and Conservation Biology, University of Minnesota, St. Paul, MN 55108, USA

ABSTRACT: With only ~1% of native prairie remaining in North America, populations of many prairie-obligate species, including the imperiled Dakota skipper butterfly, have drastically declined in recent decades. Unfortunately, population recovery is impeded by an insufficient understanding of Dakota skipper biology. Because larvae have never been naturally observed in the wild, even basic life history elements including preferred host plant(s) are not well understood, and potential hosts have been inferred from grasses inhabiting remnant sites rather than direct observations. To improve our understanding of Dakota skipper biology and habitat needs and inform recovery efforts, we conducted a no-choice performance experiment offering larvae 1 of 5 commonly occurring native grasses and 2 pervasive invasive grass species found across their historic range. We monitored larvae during key life history intervals and evaluated host plant quality by measuring larval and pupal mass, time to pupation, and survivorship. Larvae fed on all offered host grasses, but mass, phenology, and survivorship varied among treatments. Larvae reared on prairie dropseed and porcupine grass had the highest survival, the shortest time to adulthood, and the greatest mass, whereas larvae provided smooth brome and Kentucky bluegrass fared poorly for all observed metrics. All other grasses offered during the study were deemed 'medium' quality. Our results suggest that although larvae can feed on a variety of potential host plants, these hosts vary in quality. Invasive grasses across prairies in North America may pose an ecological trap to the conservation of Dakota skipper and other prairie-obligate Lepidoptera.

KEY WORDS: Ecological trap · Hesperia dacotae · Host quality · Invasive species · Larval development · Lepidoptera · Prairie · Skipper

1. INTRODUCTION

Grasslands are among the most imperiled ecosystems in North America (White et al. 2000), with less than 1% of native tallgrass prairies remaining (Samson & Knopf 1994). The widespread loss of grasslands over the last century has largely resulted from agricultural conversion and other development (White et al. 2000), markedly reducing habitat available to prairie-dependent species and yielding an increasingly fragmented landscape which promotes colonization by non-native plant species (Cully et al. 2003).

This reduction and degradation of habitat has resulted in the decline of many prairie-obligate species (Leach & Givnish 1996, Brennan & Kuvlesky 2005, Miles & Knops 2009).

The Dakota skipper Hesperia dacotae (Skinner, 1911) is a habitat specialist of northern tallgrass and upland dry mixed prairies (Royer & Marrone 1992) that has been severely impacted by habitat loss. Once predictably common from southeastern Saskatchewan and southern Manitoba through North Dakota, South Dakota, Minnesota, Iowa, and northeastern Illinois (McCabe 1981, Dana 1991, Cochrane &
Dakota skipper populations have declined precipitously in recent decades. The species is now extirpated from more than 76% of its known historic localities and is listed at national COSEWIC 2014, USFWS 2014 and international levels (Royer 2019). Even in protected remnant grassland parcels, populations of some prairie endemic butterflies continue to decline (Schlicht et al. 2009, Swengel & Swengel 2015). Habitat degradation may now pose a more significant threat to the long-term viability of Dakota skipper populations than complete habitat destruction, although the mechanism(s) underlying the declines remain(s) unclear (e.g. displacement of necessary nectar sources, preferred larval hosts, or something else entirely) (USFWS 2011, 2018). Indeed, non-native plants may pose additional novel threats to imperiled Lepidoptera by creating population sinks and even ecological traps if use of those novel plants as oviposition sites and/or larval hosts is mal-adaptive (Schlaepfer et al. 2005, Keeler & Chew 2008, Yoon & Read 2016).

Dakota skipper life history under in situ conditions is poorly understood (Dana 1991, Cochrane & Delph ey 2002, USFWS 2014). As with other grass skippers, Dakota skipper larvae feed exclusively on graminoids like grasses and sedges. Larvae have never been observed in the wild, however, so it is unclear if Dakota skipper larvae are specialists (e.g. feeding on species in a single genus, or even a single species) or polyphagous (perhaps due to variability in availability). Host plants are assumed for many grass skippers based on oviposition observations, adult proximity, or unsubstantiated reports from field guides or other anecdotal sources (MacNeill 1964, Narem & Meyer 2017). To date, research has focused on developing environmental and vegetative associations of Dakota skippers in remnant populations (Royer et al. 2008, Rigney 2013, Seidle 2018), but these efforts have not been paired with observed interactions. A better understanding of how Dakota skippers use prairie vegetation at a fine spatial scale (e.g. host foraging and shelter construction) is needed to inform conservation actions, including in situ habitat management (Rigney 2013) and ex situ breeding and reintroduction programs (Delph ey et al. 2016, USFWS 2019). Indeed, despite the increasing use of ex situ breeding and reintroduction programs as conservation tools for imperiled butterflies (Crone et al. 2007, Thomas et al. 2011), their successes historically have been limited and difficult to quantify (Oates & Warren 1990); many likely are unsuccessful due to poor species-specific knowledge (Schultz et al. 2008, Bierzychudek & Warner 2015) and/or scientific rigor (Daniels et al. 2018).

One of the most basic information needs for the conservation of an herbivorous insect is an understanding of the relationships between the target species and their prospective host plants, particularly as novel potential host plants are invading suitable habitat. We designed and implemented a no-choice performance experiment to better understand the relationship between larval host plants and Dakota skipper ecology and fitness. We leveraged the existing Dakota skipper ex situ conservation program at the Minnesota Zoo (Apple Valley, Minnesota, USA) and measured key life history parameters of larvae reared on different potential host grasses. This study improves our understanding of Dakota skipper host plant associations and promotes their conservation by informing in situ prairie management and enabling ex situ programs to optimize husbandry protocols.

2. MATERIALS AND METHODS

2.1. Study design

We implemented a no-choice performance study design in which Dakota skipper larvae were offered a single host plant species during their entire larval period. Dakota skippers are univoltine, with just a single generation annually. We conducted the experiment with the 2016–2017 and the 2017–2018 Dakota skipper generations (Gen 1 and Gen 2, respectively) reared at the Minnesota Zoo. For each cohort, we randomly assigned larvae annually to 1 of 7 treatment species and their prospective host plants, particularly as novel potential host plants are invading suitable habitat. We designed and implemented a no-choice performance experiment to better understand the relationship between larval host plants and Dakota skipper ecology and fitness. We leveraged the existing Dakota skipper ex situ conservation program at the Minnesota Zoo (Apple Valley, Minnesota, USA) and measured key life history parameters of larvae reared on different potential host grasses. This study improves our understanding of Dakota skipper host plant associations and promotes their conservation by informing in situ prairie management and enabling ex situ programs to optimize husbandry protocols.

We assessed survival at multiple life history stages and transitions and collected biometric data includ-
Table 1. The different host plant treatment species offered to Dakota skipper larvae at the Minnesota Zoo during 2016 to 2018. For analyses, host plants were grouped as native or non-native and as cool season (C₃) or warm season (C₄). The experiment was conducted for 2 generations (Gen 1 and Gen 2) with differing numbers of larvae being randomly assigned to each host treatment}

| Host species                  | Origin   | Seasonality | n (Gen 1) | n (Gen 2) |
|-------------------------------|----------|-------------|-----------|-----------|
| Big bluestem Andropogon gerardii | Native   | C₄          | 27        | 29        |
| Side-oats grama Bouteloua curtipendula | Native   | C₄          | 27        | 33        |
| Porcupine grass Hesperostipa spartea | Native   | C₃          | 26        | 29        |
| Little bluestem Schizachyrium scoparium | Native   | C₄          | 27        | 28        |
| Prairie dropseed Sporobolus heterolepis | Native   | C₄          | 25        | 31        |
| Smooth brome Bromus inermis | Non-native | C₃          | 27        | 27        |
| Kentucky bluegrass Poa pratensis | Non-native | C₃          | 27        | 35        |

Table 2. Graminoid species offered to Dakota skipper larvae at the Minnesota Zoo ex situ. All of the graminoid species have solicited a feeding response from neonate larvae. Non-native species that have been introduced to North America are indicated in bold

| Scientific name                  | Common name       |
|-----------------------------------|-------------------|
| Andropogon gerardii⁠¹,²,³          | Big bluestem      |
| Bouteloua curtipendula⁠¹,²          | Side-oats grama   |
| Bouteloua dactyloides              | Buffalo grass     |
| Bouteloua gracilis                | Blue grama        |
| Bromus inermis                    | Smooth brome      |
| Bromus kalmia                      | Prairie brome     |
| Elymus canadensis                  | Canada wild-rye   |
| Hesperostipa comata               | Needle-and-thread grass |
| Hesperostipa spartea⁠¹,²,³          | Porcupine grass   |
| Koeleria macrantha⁠²,³              | Junegrass          |
| Muhlenbergia racemosa              | Marsh muhly       |
| Muhlenbergia richardsonis          | Mat muhly         |
| Nassella viridula                  | Green needlegrass |
| Panicum virgatum                  | Switchgrass       |
| Phleum pretense⁠²,³                 | Timothy grass     |
| Poa pratensis⁠¹,²,³                 | Kentucky bluegrass|
| Schizachyrium scoparium⁠¹           | Little bluestem   |
| Sorghastrum nutans                 | Indian grass      |
| Sporobolus cryptandrus            | Sand dropseed     |
| Sporobolus heterolepis⁠¹,²          | Prairie dropseed  |
| Carex brevior                      | Plains oval sedge |
| Carex pensylvanica                 | Pennsylvania sedge|

①Species incorporated into this study; ②Species offered to neonate Dakota skippers as part of a similar no-choice performance experiment by Dana (1991); ③Species offered to neonate Dakota skippers as part of a similar no-choice performance experiment by McCabe (1981)

Larvae were reared following protocols developed by the Minnesota Zoo. Within a few hours of hatching, we placed neonates individually in a 50 ml, transparent plastic tube with living leaf blades of the randomly assigned host grass fed through a foam stopper in the top. Grasses and vials with larvae were kept indoors in a temperature-controlled laboratory, at a constant 25°C under full spectrum, T5 fluorescent grow lights, and timers were set to mimic natural day–night cycles in Apple Valley, Minnesota (44.765° N, 93.201° W) in mid-July. All larvae showed signs of feeding (e.g. notches in the leaves, frass accumulation) within 72 h of placement.

After 1 to 2 wk, at the second instar developmental stage (i.e. early August), we transitioned larvae to a host grass planted in Vigoro® All-Purpose Garden Soil, in 4 inch diameter, 4 inch tall, round plastic pots. All plants were in their second growing season and approximately 20 to 25 cm tall; culms comprised approximately 25% of the growing surface. We used plants that had enough biomass to potentially sustain a Dakota skipper throughout its entire larval development, but were not so massive as to not fit in a 4 inch pot. We sealed the pots with an upright, fine-mesh screen cage to prevent the escape of larvae and invasion by larval predators while providing the larvae full access to the host plant (Fig. 1). We selected this life stage to transition larvae to full plants to accommodate an apparent shift in feeding strategies for Dakota skipper larvae, from chewing notches mid-way on the grass leaf, to shelter building at the base of host grass (Minnesota Zoo unpubl. data). Because the amount of dead material and duff was inconsistent among plants, all dead material was removed from each plant prior to placing the larvae. We added segments of dried leaf blades of each respective assigned host, 5 cm in length and weighing a total of 2 g, to each pot to standardize the amount of material available for shelter construction.
Larvae and their associated potted plants were then housed in an outdoor, screened hoop-house at the Minnesota Zoo.

Host plants were bottom-watered and exteriors were inspected daily for mesh damage, host plant health and potential predators. If a host grass died or was nearly entirely consumed, the larva was removed and placed on a new host plant of the same species. Because Dakota skipper larvae spend most of the time concealed in their shelters and minor disturbance can lead to shelter abandonment (Minnesota Zoo unpubl. data), we only censused them during key life stage transitions (i.e. when all individuals could be censused simultaneously), or opportunistically to change a failing host plant.

Entering winter diapause, we located and removed larvae from their shelters and measured head capsule widths and mass. Dakota skippers enter diapause as either third or fourth instar larvae (Dana 1991). In keeping with the Minnesota Zoo Dakota skipper husbandry protocols to maximize larval survivorship, we overwintered larvae in artificial diapause conditions by placing each individual in a 5 ml plastic vial lined with paper towel. Vials were positioned in plastic cups filled one-third with water-saturated HydroStone® gypsum cement (United States Gypsum Company). Cups were then maintained in an environmental growth chamber at −5°C to mimic natural winter hibernation conditions until the following spring. We recorded the mass of all individuals again post-diapause (early May).

We transitioned post-diapause larvae back to their respective host plants in early May and housed them again in the outdoor, screened hoop-house. Larvae were passively observed daily until they pupated. We determined pupation date by the appearance of the water-repellant wax which encases their shelters near the time of pupation (McCabe 1981, Dana 1991). We extracted pupae from their shelters, and weighed and sexed each individual.

Research specimens that survived to pupation were incorporated into the Minnesota Zoo’s formal reintroduction program to a site in southwestern Minnesota from which the species had been recently extirpated (Delphey et al. 2017). For this reintroduction program, individuals were brought out to the release site as pupae, kept in a sealed enclosure at the site, and released daily as they eclosed as adults. We monitored eclosion date and status. Adult morphometrics (wing size, shape, and total dry mass) and reproductive output were not recorded in order to minimize handling of adults and maximize health and survival post-release.

2.3. Analysis

We evaluated stage-specific and cumulative survival of Dakota skippers using known fates analyses in the program MARK, version 9.0 (White & Burnham 1999). After initial placement of neonates on host plants, we censused all individuals during 5 sampling occasions: second instar (~1–2 wk after hatching), pre-diapause (~12 wk after the previous sampling occasion), post-winter diapause emergence (~26 wk), pupation (~8–10 wk), and adult emergence (1.5 wk). During each sampling occasion, we recorded an individual as alive, dead, or unrecovered. We censored unrecovered individuals from survival analyses for that sampling occasion. Although these stages varied in length, they were the most biologically relevant for evaluating survival
of Dakota skippers and minimized disturbance to larvae.

We created a set of 8 candidate models based on *a priori* hypotheses to explain variability in larval survivorship (see Table 3). We explored models incorporating the 7 host plant species as well as time-varying survival and time-constant survival. We considered additive and interactive relationships between the host plant species and time steps (e.g. to test the hypothesis that survival varied among host plant treatments differently over time). We also hypothesized that some aspects of host plant biology may affect larval survivorship. We first pooled the host plants into natives (i.e. big bluestem, side-oats grama, porcupine grass, little bluestem, prairie dropseed) and exotics (i.e. smooth brome, Kentucky bluegrass), hypothesizing that native grasses (those grass species Dakota skipper had most recently evolved with) would support higher survivorship than the non-native species. We also pooled host plants into C3 (porcupine grass, smooth brome, Kentucky bluegrass) and C4 (big bluestem, side-oats grama, little bluestem, prairie dropseed) subsets for analyses. It has been hypothesized that C3 (cool-season) grasses are nutritionally superior to C4 (warm-season) grasses (Caswell et al. 1973), and because C4 grasses are often tougher than C3 grasses, they are more difficult for insect herbivores to consume (Caswell & Reed 1976). To control for potential variation in survival between study years, we incorporated a ‘generation’ covariate in some models. We assessed support for candidate models using Akaike’s information criteria (AIC; Akaike 1974).

We completed all analyses for mass metrics and phenology using R Statistical Software, version 3.5.3 (R Core Team 2019). We evaluated variability in pre-diapause larval and pupal mass among host plant treatments with ANOVAs and, where appropriate, completed post-hoc Tukey tests. For the analyses in which pupal mass was assessed, we conducted separate analyses by sex because Dakota skipper adults are sexually dimorphic (females are typically about 30% larger than males; Minnesota Zoo unpubl. data). Preliminary analyses suggested that results were consistent across generations, so we did not incorporate variability by generation in further analyses.

We determined timing of individuals to pupation after winter diapause using a standard degree-day (DD) model (i.e. (maximum daily temperature + minimum daily temperature)/2 − lower temperature threshold), modified from Dearborn & Westwood (2014). We used 10°C as the lower temperature threshold rather than 6°C based on previous Dakota skipper models developed by Gerald Selby for southern Minnesota (G. Selby unpubl. data). We used this approach for controlling for phenology to accommodate differences in temperature conditions between the 2 years of the experiment and because larval removal from diapause varied by up to 5 calendar days within each generation. We calculated DD to pupation (rather than adult emergence; cf. Dearborn & Westwood 2014) because the exact dates of eclosion were not known for some individuals that were included in an ongoing reintroduction project.

We conducted ANOVAs to evaluate variability in accumulated DD from post-diapause to pupation by host plant treatments and used post-hoc Tukey tests where appropriate. Sexes were analyzed separately because Dakota skipper are known to exhibit pro-tandry, in which males eclose before females (Dana 1991), similar to many other butterflies (Fagerström & Wiklund 1982). For all mass and phenology analyses, we examined Q-Q plots and residuals, and conducted Shapiro-Wilks tests to confirm that assumptions of normality were met.

### 3. RESULTS

We randomly assigned 398 Dakota skipper larvae to 1 of the 7 host plant treatments. A total of 121 individuals survived to pupation, and we documented variability in the target life history metrics: survival, mass of immature stages, and phenology.

#### 3.1. Survival

The most highly supported models explaining variation in survival included terms for all host plant species with full time variation (i.e. independent estimates of survival for each host plant species at each time step, or life history stage); the 2 most-supported models included >99% of all model weight (Table 3). Because the second most-highly supported model only differed from the top model in its inclusion of a term for ‘generation’, we considered ‘generation’ an uninformative parameter and based our inference on the most highly supported model (Arnold 2010). Although the host plant × time model required estimation of many parameters relative to the treatment sample sizes, its support in this modeling framework suggests that there is significant variation in Dakota skipper survival among different host grasses, which is affected by life history stage. Modeling provided no support for more simplistic models (e.g. time varying including an additive relationship with host plant
species; time-constant) or models that pooled host plant species by biological characteristics.

Cumulative survival rates derived from the most highly supported model varied significantly among larvae reared on different host plants (Fig. 2), with survival rates exceeding the pooled treatment average for individuals reared on porcupine grass, prairie dropseed, and side-oats grama across nearly all life history stages (Fig. 3). Additionally, stage-specific survival rates varied among host plant species (Fig. 4). All host plant treatments supported relatively high survivorship of larvae (73 to 89%) over winter diapause (i.e. the life history stage between pre- and post-diapause). For the pupae to adult eclosion life history interval, survival exceeded 93% for larvae reared on all treatments other than smooth brome; by contrast, only approximately 43% of pupae reared on smooth brome successfully eclosed. Although 7 individuals reared on smooth brome survived to pupation, the only 2 males died as pupae, and 3 of the surviving females failed to eclose properly and were unable to fly as adults. Notably, these were the only 3 individuals during the course of the experiment (on any host plant) that survived to adulthood but were unable to expand their wings.

### 3.2. Immature stage masses

Larval mass entering winter diapause varied among host treatments ($F_{5,212} = 3.620; p = 0.0019$). Larvae reared on porcupine grass were significantly larger than those grown on big bluestem, although larval mass did not significantly differ among other treatments (Table 4). Preliminary analyses suggested that

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**Table 3. Model selection results of candidate models explaining variability in survival of Dakota skippers reared on different host plants at the Minnesota Zoo during 2016 to 2018. AICc: corrected Akaike’s information criterion**

| Model description                                                                 | ΔAIC  | AICc weight | Model likelihood | Number of parameters | Deviance     |
|----------------------------------------------------------------------------------|-------|-------------|------------------|----------------------|--------------|
| Survival varies by host plant treatment type, with an interactive effect with life history stages | 0.00  | 0.59        | 1.00             | 35                   | 1163.94      |
| Survival varies by host plant treatment type, with an interactive effect with life history type and an additive effect of generation | 0.71  | 0.41        | 0.70             | 36                   | 1162.53      |
| Survival varies by host plant treatment, with an additive effect of life history stage | 8.72  | 0.01        | 0.01             | 11                   | 1222.56      |
| Survival varies by native and introduced grass types, with an additive effect of life history stage | 20.71 | 0.00        | 0.00             | 10                   | 1236.60      |
| Survival varies by cool- (C3) and warm-season (C4) grass types, with an additive effect of life history stage | 34.49 | 0.00        | 0.00             | 10                   | 1250.38      |
| Survival varies by life history stage only | 36.50 | 0.00        | 0.00             | 5                    | 1262.52      |
| Survival varies by host plant treatment only | 44.12 | 0.00        | 0.00             | 7                    | 1266.09      |
| Survival constant across life history stages and host plant treatments | 75.46 | 0.00        | 0.00             | 1                    | 1309.52      |

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![Fig. 2. Cumulative survival estimates for Dakota skippers integrated in an experiment evaluating different host plant species at the Minnesota Zoo during 2016 to 2018. *Invasive grass species*](image-url)
although generation (i.e. Gen 1 or Gen 2) did not affect size, pupal mass varied by sex, so we conducted individual ANOVAs by sex to evaluate the relationship between host plant and pupal mass. Females reared on porcupine grass as larvae were significantly larger as pupae than females reared on smooth brome and Kentucky bluegrass (Table 4). Males reared on prairie dropseed were significantly larger than males reared on big bluestem and Kentucky bluegrass (Table 4). Because only 2 males reared on smooth brome survived to pupation, this treatment was excluded from the analysis.

3.3. Phenology

DD did not significantly differ between generations ($F_{1,123} = 0.833; p = 0.364$), but did vary by sex ($F_{1,117} =$

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**Fig. 3.** Total survivorship of Dakota skipper by offered host plant species at defined life-history intervals. (solid line) Cumulative survival at each time step for each host plant treatment; (dotted reference line) mean cumulative survival across all treatments and years; *invasive grass species
31.732; p < 0.001), so male and female DD were analyzed separately. DD accumulation from winter diapause to pupation varied among host plant treatments for female pupae ($F_{6,52} = 8.561; p < 0.001$), with individuals reared on smooth brome taking significantly longer to reach pupation than those reared on little bluestem, side-oats grama, porcupine grass, or prairie dropseed (Fig. 5). Male pupae did not differ by DD to pupation ($F_{5,60} = 0.771; p = 0.599$; Fig. 5), but as noted earlier, males reared on smooth brome were excluded from this analysis due to poor survivorship.

4. DISCUSSION

Determining what a caterpillar eats seems like an easily answered question, but in the case of Dakota skipper and many other graminoid-feeding grass-
land specialist insects, the answer can be obscure and complex. We documented that Dakota skipper larvae will feed on a wide range of commonly available graminoid hosts that can solely sustain them through all their developmental stages, suggesting they are fairly polyphagous with respect to graminoids. However, variation in pupal mass, timing to pupation, and total survivorship to adulthood highlights that there are differences in host plant quality.

To assess overall host plant quality, we developed a post-hoc scoring system summarizing the parameters evaluated in this study (Table 5). Our system was based on how results from individual metrics likely reflected host plant quality, such that we categorized each metric as ‘Low’ (1), ‘Medium’ (2), or ‘High’ (3) quality. For female pupal mass and timing to pupation, we assigned scores based on the results of post-hoc Tukey tests. Host plants that supported statistically larger individuals and individuals that pupated more rapidly were assigned ‘High’ rankings for pupal mass and timing to pupation, respectively, whereas host plants that yielded statistically smaller individuals and larvae that required longer to reach pupation were assigned ‘Low’ scores for the respective categories. For cumulative survival, host plant treatments were categorized as ‘Low’ if larvae reared

| Host Plant          | Pre-diapause larvae | Female pupae | Male pupae |
|---------------------|---------------------|--------------|------------|
| Big bluestem        | 0.021 (0.018–0.024) | 0.213 (0.182–0.244) | 0.197 (0.169–0.225) |
| Side-oats grama     | 0.026 (0.024–0.028) | 0.233 (0.214–0.253) | 0.226 (0.221–0.242) |
| Porcupine grass     | 0.029 (0.027–0.031) | 0.273 (0.252–0.294) | 0.230 (0.212–0.248) |
| Little bluestem     | 0.026 (0.024–0.028) | 0.219 (0.194–0.243) | 0.204 (0.180–0.225) |
| Prairie dropseed    | 0.026 (0.025–0.028) | 0.260 (0.238–0.283) | 0.245 (0.229–0.262) |
| Smooth brome        | 0.024 (0.021–0.027) | 0.201 (0.170–0.231) | na         |
| Kentucky bluegrass  | 0.024 (0.022–0.027) | 0.204 (0.177–0.230) | 0.197 (0.176–0.182) |

Table 4. Dakota skipper mean pre-diapause larval and pupal mass (g) by offered host plant species (95% confidence intervals). N: individuals surviving to that sampling event. Cells within columns that do not share a letter are significantly different from each other (at \( \alpha = 0.05 \)) with Tukey’s post-hoc test. na: not applicable (as only 2 males reared on smooth brome survived to pupation, this treatment was excluded from the analysis)

Fig. 5. Accumulated degree-days (DD) to pupation after winter diapause for female (left) and male (right) Dakota skippers. Within each box, black lines denote median values; boxes extend from the 25th to the 75th percentile of each group’s distribution of values; whiskers denote adjacent values; x-marks denote outliers. No male Dakota skippers survived to adulthood on smooth brome. Different letters indicate results that are significantly different from each other as analyzed with a post hoc Tukey test. *Invasive grass species
on that species exhibited survival rates below the pooled treatment mean or ‘High’ if larvae survived at a rate above the combined mean (Fig. 3). This post-hoc assessment identified porcupine grass and prairie dropseed as the most optimal hosts of the provided graminoid species with the highest survival, the largest pupae, and the shortest time to pupation, and these ranked highest in our post-hoc assessment of overall host plant quality. We classified big bluestem, side-oats grama, and little bluestem as moderate hosts, whereas the 2 invasive species, smooth brome and Kentucky bluegrass, ranked poorly and are considered inferior hosts.

Many herbivorous insects have tight host plant niches that may be brought about by a co-evolutionary arms race with their respective hosts, resulting in specialization on just a few plant species or genera (Ehrlich & Raven 1964). This is often not the case for graminoid feeding insects, which are characterized as having a broader taxonomic host range (Tscharntke & Greiler 1993). Although grasses do defend themselves from herbivory (Vicari & Bazely 1993), this is often achieved less by means of secondary metabolites than through mechanisms such as silicates (Massey et al. 2006), fungal endosymbionts (Ball et al. 2006), and leaf toughness (Nakasuji 1987, Moore & Johnson 2017). Despite this battery of defense mechanisms, many grass skippers overcome these barriers. Grass skipper larvae hatch with large head-capsules relative to other Lepidoptera, allowing them to feed on some of the toughest grass leaves (Nakasuji 1987, Seko & Nakasuji 2004). Additionally, fungal endosymbionts, although found to be toxic to many livestock, do not impact the development and survivorship of the tawny-edged skipper Polites themistocles (Jokela et al. 2016), another Hesperiinae grassland specialist butterfly that historically occurs sympatrically with the Dakota skipper.

Larval host plants are often determined by observing oviposition events. For many Lepidoptera, it is often assumed that adults will oviposit on their optimal host plant and that doing so will optimize their fitness (Ehrlich & Raven 1964, Levins & MacArthur 1969, Jaenike 1978). However, observations of oviposition alone do not necessarily indicate that the recipient plant is an optimal, or even a palatable, host (Casagrande & Dacey 2007). Although many herbivorous insects will oviposit on optimal host plants, this trend appears less pronounced in polyphagous species with abundant access to edible species, and additional information is needed to improve our understanding of the response of polyphagous species (Gripenberg et al. 2010). It is often the case that graminoid-feeding Lepidoptera in grassland habitats will oviposit nearly randomly on available substrates, presumably because potential host plants are in close enough proximity for the neonate larvae to find on their own (Wiklund 1984). Dakota skipper oviposition has been rarely observed in the wild (Selby 2006), but includes native bunchgrasses and invasive grasses, as well as other unsuitable live and dead non-graminoid plant material and even inert surfaces (McCabe 1981, Dana 1991, Minnesota Zoo unpubl. data). Similar oviposition ‘mistakes’ have been recorded in other Hesperia (MacNeill 1964). Thus, female Dakota skipper oviposition behavior may not be mediated by optimal hosts, though more research is needed.

McCabe (1981) first documented that Dakota skipper neonate larvae accepted big bluestem, porcupine grass, Junegrass, Timothy grass Phleum pretense, Kentucky bluegrass, prairie dropseed, and an unspecified Carex species. Twenty-two graminoid species have been offered to neonate Dakota skipper larvae over the course of the ex situ program at the Minnesota Zoo (Table 2), and neonates have fed on all of them within 72 h of hatching. Although triggering a feeding response by a neonate larva is a good indicator of the plant being a candidate host, it does not necessarily indicate that it is a quality, or sustainable food source (Gómez Jiménez et al. 2014). Thus, it is clear that an understanding of host plant effects throughout development is essential when evaluating the importance of different hosts for a particular insect herbivore and detecting potential sub-lethal effects.

Table 5. Host plant quality rankings. Metrics to assess Dakota skipper performance on different host plants were categorized as ‘Low’ (score = 1), ‘Medium’ (2), or ‘High’ (3). Scores were added to produce a final rank quality of 3 to 9, with 9 the most optimal host plant

| Pupal mass (female) | Timing to pupation | Total survivorship | Rank quality |
|---------------------|--------------------|--------------------|--------------|
| Big bluestem        | Medium (2)         | Low (1)            | Medium (5)   |
| Side-oats grama     | Medium (2)         | Medium (2)         | Low (1)      |
| Porcupine grass     | High (3)           | High (3)           | Medium (9)   |
| Little bluestem     | Medium (2)         | High (3)           | Medium (8)   |
| Prairie dropseed    | Medium (2)         | High (3)           | Low (3)      |
| Smooth brome*       | Low (1)            | Low (1)            | Low (3)      |
| Kentucky bluegrass* | Low (1)            | Low (1)            | Low (3)      |

*Invasive grass species

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Our results suggest that Dakota skipper performance does vary among different host plants. We documented variability in all performance metrics for larvae provided different host grass treatments. Dana (1991) reported the high quality of prairie dropseed, but suggested that porcupine grass was likely an unsuitable host given its leaf toughness. Although we did not measure leaf toughness in this experiment, even neonate larvae were capable of feeding on porcupine grass, and our results suggest it is a high-quality host. Importantly, Dana (1991) concluded his studies prior to winter diapause, where larvae had only gone through half their developmental stadia. Our research indicates that performance differences among host plants become amplified in later stages after winter diapause.

4.1. Future work

Our controlled, no-choice performance experimental design may not account for variation in host utilization under wild conditions (Forister 2008). As the native prairie habitat of the Dakota skipper is made up of a mosaic of grasses, a next step in understanding their host plant dynamics is to implement a choice study in which larvae have access to multiple species. Although some Lepidoptera larvae may be unable to discern between suitable and unsuitable host plants (Gómez Jiménez et al. 2014), some polyphagous species have demonstrated selection (Nylin et al. 2000, Gamberale-Stille et al. 2014) and are even able to modify their feeding preference in the presence of predators (Bernays & Graham 1988) or differentially choose alternative hosts if parasitized (Smilanich et al. 2011). We were unable to determine if Dakota skippers can discern between suitable and unsuitable hosts, or if foraging on multiple hosts would promote a fitness advantage (e.g. larger, more fecund adults). Similarly, observations of neonate behavior and mobility (Thompson 1983) in situ or in controlled areas similar to those conducted by Bierzychudek & Warner (2015) would improve our understanding of Dakota skipper ecology and the threat posed by smooth brome and other invasive species. Unfortunately, traditional experimental host choice studies are less feasible with obligate shelter-building butterflies like Dakota skippers than they can be with other butterflies that will initiate feeding without first building a complex shelter.

Shelter morphometrics and potential variation among grass hosts also require further research. Like many other grass skippers, Dakota skippers defend themselves from predators and the elements in constructed shelters at the base of their host plants (MacNeill 1964). Second instar larvae begin stitching blades of grasses together, and the larvae feed and overwinter in these shelters until finishing development the following spring. The cryptic larvae and their shelters are virtually undetectable in the landscape, and it is unknown how structure construction might vary across host grass species, though variation has been observed within dicot feeding Pyrrophygini skippers (Greeney et al. 2010a). Due to interspecific variation in leaf size and basal architecture, we expect that larval shelter construction will necessarily vary among host grasses, and that some of these variants may be disadvantageous to Dakota skipper larvae. For example, a more tightly bound shelter may be possible with prairie dropseed (numerous narrow leaf blades around a circular base) relative to little bluestem (fewer broad blades around a flattened base). These differences may result in variations in visibility to predators and parasitoids and microclimatic differences that may influence survivorship and/or growth rates (Greeney et al. 2010b). Anecdotally, over the course of this study, variation in shelter size and orientation above or below surface of the soil line was observed.

4.2. Management implications

The invasion of smooth brome into prairies appears to represent a sink that complicates Dakota skipper conservation efforts. Despite the apparent lack of female oviposition preference provided by opportunistic observations, larvae reared exclusively on this exotic grass consistently had the lowest survivorship, smallest pupal mass, and took the longest to reach pupation. Extended development time could indicate that smooth brome does not provide sufficient larval nutritional content (Bauerfeind & Fischer 2005). Notably, greater pupal mass corresponded with higher overall survivorship and shorter development time (e.g. individuals fed prairie dropseed and porcupine grass), suggesting that longer development time is indicative of a nutritional deficiency or stress.

In addition to lower survivorship and mass, delayed development on suboptimal hosts, like smooth brome, may have serious negative impacts on Dakota skipper populations. Host plant quality can impact the number of annual generations and may have an even greater impact on phenology than temperature in some Lepidoptera (Abarca et al. 2018).
For a univoltine species with a short reproductive window like the Dakota skipper, delayed development and a shift in adult flight could prevent overlap with potential mates, or asynchrony with preferred adult nectar resources (Van Dyck et al. 2015). Indeed, smooth brome-reared Dakota skippers reached adulthood up to 2 wk later than their siblings reared on other more suitable hosts. Adult Dakota skipper lifespan under natural conditions is likely less than 2 wk. Thus, individuals feeding exclusively on smooth brome may not reach adulthood until after the flight and thus be excluded from the year’s reproduction.

Egg-laying females seem to exhibit little to no oviposition selection for optimal plant hosts (McCabe 1981, Minnesota Zoo unpubl. data), and as documented here, Dakota skipper neonate larvae will consume host plants that result in poor performance in the long term. As a result, both adults and larval Dakota skippers may be unable to distinguish optimal hosts. The inability to distinguish optimal from suboptimal plants has been observed in other Lepidoptera (Gómez Jiménez et al. 2014). Indeed, female host preference appears to evolve independently from larval host performance (Forister 2005, Friberg et al. 2015). The apparent inability of Dakota skipper to identify prime host plants suggests that the presence of large patches or complete invasions of smooth brome may represent population sinks for Dakota skipper. As smooth brome has already largely invaded much of the Dakota skipper range (DeKeyser et al. 2013), it may be a significant stressor for remnant populations.

Little bluestem has been closely associated with Dakota skipper habitats (Royer & Marrone 1992, Rigney 2013, Seidle et al. 2018), and big bluestem is the dominant grass species at extant Dakota skipper sites (Rigney 2013). Indeed, native bunch grasses, specifically little bluestem, have been deemed essential for Dakota skipper conservation (USFWS 2018). However, this designation may not be indicative of Dakota skipper biological needs. Results from this experiment suggest that little bluestem and big bluestem are moderate quality host plants (Table 5). Furthermore, it is unlikely that little bluestem alone could support Dakota skipper larvae in situ. For the purposes of this study, all treatment grasses were propagated under laboratory conditions so that they would be of consistent sizes when offered. However, even compared to other C₄ grasses, little bluestem develops new growth fairly late into the season (Gillen & Ewing 1992). Assuming Dakota skipper begin foraging after winter diapause (once average daily temperatures reach about 6 to 10°C; Dearborn & Westwood 2014), anecdotal observations from field surveys suggest that little bluestem often may not have growth available for forage at this time. Though not characterized to the same extent as little bluestem, porcupine grass and prairie dropseed (Royer & Marrone 1992) also commonly occur at remnant prairie sites supporting Dakota skipper and are available earlier in the growing season.

Land management practices developed for the support of Dakota skipper should prioritize porcupine grass and prairie dropseed and emphasize the suppression of smooth brome. Given the widespread and sometimes near complete invasions of smooth brome into North American prairies, this may be a tricky prospect and novel approaches may be needed to avoid the potential ecological trap that it may present to Dakota skippers (Schlaepfer et al. 2005).

Methodologies utilized to determine Dakota skipper host plant suitability may inform important research for other imperiled graminoid-feeding specialist insects for which much of the life history is unknown (e.g. Poweshiek skippering Oarisma poweshiek).

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