RESEARCH ARTICLE

PREVALENCE AND ANTIMICROBIAL RESISTANCE PATTERN OF METHICILLIN RESISTANT STAPHYLOCOCCUS AUREUS ISOLATED FROM DIFFERENT CLINICAL SAMPLES.

Dr. Nida Khan¹, Dr. Vineeta Khare, Dr. Rahmat Farid and Dr. Shadma Yaqoob.

Background: Methicillin Resistant Staphylococcus aureus (MRSA) since its emergence, has been a challenge for the community owing to its potential to cause life threatening events like sepsis, endocarditis and osteomyelitis.

Aims and objectives: To investigate the prevalence of MRSA in various clinical samples and to find the antibiotic resistance pattern of the MRSA isolates.

Materials and Methods: Oxacillin resistance screening agar with 5.5% Nacl and Kirby–Bauer disk diffusion method (cefoxitin 30µg disk) were used for MRSA confirmation and antibiotic resistance testing was done as per CLSI 2016 guidelines. E-test was done for testing sensitivity to Vancomycin.

Result: Staphylococcus aureus was isolated from 350 specimens. Out of these 350 isolates, 220 (62.86%) were methicillin sensitive and rest 130 (37.14%) isolates were MRSA. Most of the MRSA isolates were from urine samples (43.71%) followed by pus (24.0%), and putum (11.14%). MRSA isolates showed high resistance to ciprofloxacin (53.38%), and clindamycin (42.31%). linezolid resistance was seen in only 6.15% and all isolates were sensitive to vancomycin.

Conclusion: A high prevalence of MRSA (37.14%) in our institution warrants the judicious use of antibiotics in treating infections caused by Staphylococcus aureus. Vancomycin and linezolid are good treatment options in infections caused by MRSA isolates.

Corresponding Author:- Nida Khan.
England, and since then it continues to be the most dreadful strains of S. aureus. It is one of the most common pathogens that cause nosocomial infections.

In India, a multicentric trial conducted between January 2008 to December 2009 at several healthcare centres found MRSA prevalence among specimen collected from outpatients, ward inpatients and ICU to be 28, 42 and 43 per cent, respectively in 2008 and 27, 49 and 47 per cent, respectively in 2009, thus indicating that MRSA has assumed a concerning proportion. Moreover, they stressed on the pattern of changing antibiotic susceptibility and also recommended robust antimicrobial stewardship and strengthened infection control measures to prevent spread and reduce emergence of resistance.

It must be kept in mind that current therapeutic options for MRSA are limited few expensive drugs like vancomycin, linezolid, teicoplanin, daptomycin and streptogramins. Another alarming sign is that emergence of resistance to Vancomycin, although at a low level has been reported in literature, thus underlining the observations made by the multicentric study cited above.

Hence, the present study was planned with an aim to assess the prevalence and antibiotic susceptibility of Methicillin resistant Staphylococcus aureus at Era’s Lucknow Medical College and Hospital (ELMCH)

Aims and Objectives:
The present study was carried out with an aim to evaluate the prevalence, antibiotic susceptibility of MRSA in a tertiary care centre. This aim was fulfilled with the help of following objectives:
1. To investigate the prevalence of MRSA in various samples obtained for culture.
2. To find the antibiotic resistant pattern of MRSA isolates

Material And Methods:
Place Of Study: Study was conducted at Department of Microbiology, Era’s Lucknow Medical College.

Duration: 12 months from Nov 2014 to Oct 2015

Study Population:
Part I: All patients visiting Era's Lucknow Medical College and Hospital, Lucknow.

Sampling Frame:
Sample size: 350

Inclusion criteria:
All patients including outpatients and admitted patients

Exclusion criteria:
Patients who did not show cooperative attitude or refuse to provide necessary information were excluded.

Sample Collection:
Part I: The sample collection was done at different clinical Department of Microbiology, Era’s Lucknow Medical College, Lucknow using standardized sampling techniques. The specimen obtained from different sources were labeled and tagged with information

Lab Diagnosis:
Microscopy:
Gram staining
Culture was done on following media:
Blood agar
Mac Conkey agar
**MRSA confirmation by:-**

**Oxacillin resistance screening agar with 5.5% NaCl:-**
Suspend 51.75 grams in 500 ml distilled water. Heat to boiling to dissolve the medium completely. Sterilize by autoclaving at 15 lbs pressure (121°C) for 15 minutes. Cool at 45-50°C and aseptically add rehydrated contents of 1 vial of Oxacillin Resistance Selective Supplement (FD191). Mix well and pour into sterile Petri plates.

**Disc diffusion method** [Kirby bauer (cefoxitin 30 mg disk)]

**Procedure (As per manufacturer’s instructions):-**
Bacterial Suspension was prepared according to 0.5 McFarland standard and results were interpreted as Resistant, Intermediate or Sensitive for each antimicrobial using CLSI guidelines.

E-TEST:Vancomycin EzyMIC strips (HiMedia) were used to test for vancomycin sensitivity over a range of MIC from 0.016-256 mcg/ml. The E Test was done and interpreted as per manufacturer’s instructions. Apply the Etest strip to the agar surface with the MIC scale facing upwards. MICs were read where the edge of inhibition ellipse intersected the strips.

**Statistical Tools Employed:-**
The statistical analysis was done using SPSS (Statistical Package for Social Sciences) Version 15.0 statistical Analysis Software. The values of mean, standard deviation and p value were calculated and a p value of p<0.05 was considered significant. Chi square test was done to differentiate between MRSA and MSSA.

**Results:-**
A total of 350 specimens from the patients were collected which were subjected to MRSA sensitivity.

**Table 1:-** Distribution of Cases according to MRSA status (n=350)

|          | Number of cases | Percentage |
|----------|-----------------|------------|
| MRSA     | 130             | 37.14      |
| MSSA     | 220             | 62.86      |

Out of 350 specimens, 220 (62.86%) were found to be MSSA and only 130 (37.14%) specimens were found to be MRSA. Prevalence of MRSA in our tertiary care centre was found to be 37.14.

**Table 2:-** Comparison of Type of specimen between MRSA and MSSA cases

|          | Total | MRSA (n=130) | MSSA (n=220) | Statistical significance |
|----------|-------|--------------|--------------|-------------------------|
|          | No.   | %            | No.          | %                        | χ²       | p          |
| BC       | 5     | 3.85         | 0            | 0.00                     | 47.289   | <0.001     |
| Blood    | 15    | 2            | 13           | 5.91                     |          |            |
| Body fluid | 30    | 2            | 28           | 12.73                    |          |            |
| Catheter tip | 24    | 10           | 14           | 6.36                     |          |            |
| Pus      | 84    | 51           | 33           | 15.00                    |          |            |
| Sputum   | 39    | 11           | 28           | 12.73                    |          |            |
| Urine    | 153   | 49           | 104          | 47.27                    |          |            |
Out of 350 specimens, most common specimen was Urine (n=153; 43.71%), followed by Pus (n=84; 24.0%), Sputum (n=39; 11.14%), Body fluid (n=30; 8.57%). 15 (4.29%) were blood specimens and 5 (1.43%) were from blood culture (BC).

Proportion of MRSA specimens was higher than MSSA for Blood components – Blood culture (3.85% vs. 0.0%), Catheter tip (7.69% vs. 6.36%), Pus (39.23% vs. 15.00%).

Table 3: Distribution of MRSA positive specimen according to Method of Identification (n=130)

| SN  | Method                                                   | No. of specimen identified | Percentage |
|-----|----------------------------------------------------------|-----------------------------|------------|
| 1.  | Disc diffusion method : Kirby Bauer (cefoxitin 30mg disk) | 130                         | 100        |
| 2.  | Oxacillin resistance screening agar with 5.5% NaCl       | 90                          | 69.2       |
| 3.  | Both                                                     | 90                          | 69.2       |

All the MRSA samples were identified by Kirby-Baur disc diffusion method (cefoxitin 30 mg). Using Oxacillin resistance screening agar with 5.5% NaCl helped in identification of 90 (69.2%) specimen. Both the methods identified MRSA in 90 (69.2%) specimens.

Difference in type of specimens between MRSA and MSSA was found to be statistically significant (p<0.001).

Table 9: Comparison of Resistance of MRSA and MSSA cases

| Resistant | MRSA (n=130) | MSSA (n=220) | Statistical significance |
|-----------|--------------|--------------|-------------------------|
|           | No. | %    | No. | %    | \(\chi^2\) | P      |
| CEFOXITIN | 130 | 100.00 | 0   | 0.00 | 350.00    | <0.001 |
| CIPROFLOXACIN | 70 | 53.38 | 63  | 28.6 | 22.04    | <0.001 |
| DOXYCYCLINE | 20 | 15.38 | 17  | 7.7  | 5.068    | 0.024  |
| NORFLOXACIN | 18 | 13.85 | 15  | 6.8  | 4.726    | 0.030  |
| LINEZOLID | 8  | 6.15  | 8   | 3.6  | 1.187    | 0.276  |
| CLINDAMYCIN | 55 | 42.31 | 47  | 21.4 | 17.37    | <0.001 |
| VANCOMYCN | 0  | 0.00  | 0   | 0.00 | -        | -      |
| AMIKACIN | 13 | 10.00 | 11  | 5.0  | 3.198    | 0.074  |
All the MRSA specimen were resistant against Cefoxitin. The resistance rate in decreasing order was Ciprofloxacin (53.38%), Clindamycin (42.31%), Doxycycline (15.38%), Norfloxacin (13.85%), Amikacin (10%) and Linezolid (6.15%). Vancomycin was sensitive against 100% of MRSA isolates.

All MSSA specimen were sensitive against Cefoxitin. The resistance rate in decreasing order was Ciprofloxacin (28.6%), Clindamycin (21.4%), Doxycycline (7.7%), Norfloxacin (6.8%), Amikacin (5%) and Linezolid (3.6%). Vancomycin was sensitive against 100% of MRSA isolates.

**Discussion:**

Methicillin resistant *Staphylococcus aureus* since its emergence has been a challenge for the healthcare workers owing to its potential to cause life-threatening events like sepsis, endocarditis, and osteomyelitis. Major outbreaks of MRSA and its different phage types have also been recorded and reported in healthcare facilities. Since resistance to multiple antibiotics among MRSA isolates is very common, there is a possibility of extensive outbreaks, which may be difficult to control. MRSA is now one of the commonest nosocomial pathogens, and asymptotically colonized healthcare workers are the major sources of MRSA in the hospital environment. Early detection of MRSA and formulation of effective antibiotic policy in tertiary care hospitals is of paramount importance from the epidemiological point.

In our study a total of 350 clinical specimen positive for *Staphylococcus aureus* were obtained and assessed for methicillin resistance. Out of these 130 (37.14%) were found to be Methicillin resistant. Thus prevalence of Methicillin resistant *S. aureus* was found to be 37.14% in present study. Prevalence of MRSA has been shown to vary substantially in some contemporary clinical series from the region. Table D1 shows the prevalence of MRSA in some contemporary clinical series:

| SN | Author (Year) | Sample size and characteristic | MRSA prevalence rate |
|----|---------------|--------------------------------|----------------------|
| 1. | Saikia *et al.* (2009)12 Dibrugarh, Assam | 276 Clinical specimen | 34.78 |
| 2. | Ahmad *et al.* (2009)13, Armed Forces Hospital, Saudi Arabia | 106 Specimen collected from different hospital | 22.3% |
| 3. | Tiwari *et al.* (2011)14, Bhubaneswar, India | 204 Clinical samples | 55.8% |
| 4. | Sharma *et al.* (2013)15, Mangalore, India | 685 | 23.25% |

An overview of Table D1 above shows a wide variability in clinical prevalence rate of MRSA in different studies. In present study, Kirby Bauer disc diffusion method (Cefoxitin 30 mg) proved to be more sensitive than Oxacillin resistance agar method for detection of MRSA. This finding is in accordance with the observations made by Datta *et al.* who also showed that Cefoxitin disc diffusion is more sensitive than Oxacillin resistance agar method in the detection of MRSA16.

In present study, pus was the most common source of MRSA (39.23%) followed by urine (37.69%), sputum (8.46%) and catheter tip (7.69%) respectively. Saikia *et al.* (2009)12 also showed maximum isolation of MRSA from pus/wound swabs (46.67%) followed by sputum/throat swab (42.86%) while Ahmad *et al.*13 found source of MRSA to be 22.2% from pus, 23.8% from wound swabs, 33.3% from aspirates and 13.3% from sputum. The findings of present study are in accordance with the observations of Tiwari *et al.*14 (2011) who reported pus (45%) followed by urine (20.5%) to be the major source of MRSA in their study. With slight difference in proportions, these two sources comprise the major source of MRSA in our study too. In present study, MRSA positivity rate in pus samples was significantly higher (51/84; 60.7%) as compared to other specimen (79/266; 29.7%). This finding is in accordance with some other studies too that have also reported pus to have a higher MRSA positivity rate as compared to other specimen12,14,15.

MRSA specimens were most sensitive against Vancomycin (100.0%) and Linezolid (93.85%). Cefoxitin, Ciprofloxacin and Clindamycin showed maximum resistance (100%, 53.38% and 42.31% respectively). The antibiotic susceptibility has been reported to vary substantially in different studies. Goyal et al(2013)17 showed from a study conducted in a teaching hospital at Agra that MRSA specimens were most sensitive against
Vancomycin, Teicoplanin and Linezolid (100%), Ampicillin, Erythromycin and Chloramphenicol showed maximum resistance (100%, 88%, 66.7% respectively). Another study conducted by Sharma et al. (2013) from Mangalore, India showed most of the MRSA strains were sensitive against Linezolid (98.27%) and Ciprofloxacin (64.7%), Tetracycline (58.38%) showed maximum resistance. The findings of the present study are in accordance with Tiwari et al. (2011) from Bhubaneswar, India who showed all the isolated MRSA strains were sensitive against Vancomycin and Linezolid (100%). Vancomycin and Linezolid are most sensitive drugs with most of the studies reporting their sensitivity rates between 90% to 100%. The findings of present study also emulated the same and showed vancomycin and linezolid to be 100% and 93.85% sensitive. For other conventional antibiotics different studies have shown a high resistance pattern. A number of studies have also reported multiresistance. In present study too for conventional antibiotics the resistance rates were ranged from 42.31% to 100%. A number of specimen were polyresistant too.

These findings in turn indicate the need to develop an antibiotic protocol dependent on MRSA profile of the pathogen in order to reduce the unnecessary burden of antibiotics.

Conclusions:-

The present study was carried out in the Department of Microbiology, Era's Lucknow Medical College & Hospitals to evaluate the prevalence, antibiotic susceptibility and carriage rate of MRSA in a tertiary care centre.

1. Out of 350 specimens during the study period, 220 (62.86%) specimens were found to be MSSA and 130 (37.14%) were MRSA. Prevalence of MRSA in our institution was 37.14%.
2. Proportion of MRSA specimens was higher than MSSA for Blood culture (3.85% vs. 0.0%), Catheter tip (7.69% vs. 6.36%), Pus (39.23% vs. 15.00%) while proportion of MSSA was higher than MRSA for body fluid (12.73% vs. 1.54%), Sputum (12.73% vs. 8.46%) and Urine (47.27% vs. 37.69%).
3. MRSA specimens were most sensitive against Vancomycin (100.0%) and Linezolid (93.85%) while least sensitive for Cefoxitin (0.0%).
4. 90 isolates were detected by oxacillin resistance screening agar and 130 isolates were detected by Kirby Bauer disk diffusion method. Thus Kirby Bauer method provided an addition of 30.8% in overall MRSA detection.

The findings of present study thus showed that MRSA was prevalent in our hospital. There is a progressive increase in MRSA positivity and multi-drug resistance in strains of *Staphylococcus*, vancomycin and linezolid were found to have absolute sensitivity.

References:-

1. Ma X, Ito T, Tiensasitorn C, Jamklang M, Chongtrakool P, Boyle-Vavra S et al. Novel type of staphylococcal cassette chromosome mec identified in community-acquired methicillin resistant *Staphylococcus aureus* strains. Antimicrobial Agents Chemotherapy, 2002; 46(4): 1147-1152. Link
2. DeLeo FR, Otto M, Kreiswirth BN, Chambers HF. Community-associated methicillin-resistant *Staphylococcus aureus*. Lancet. 2010; 375: 1557-1568. Link
3. Kennedy AD, Deleo FR. Epidemiology and Virulence of Community-Associated MRSA. Clinical Microbiology Newsletter. 2009; 31: 153-160. Link
4. Kennedy AD, Deleo FR. Epidemiology and Virulence of Community-Associated MRSA. Clinical Microbiology Newsletter. 2009; 31: 153-160. Link
5. Chambers HF, Deleo FR. Waves of resistance: *Staphylococcus aureus* in the antibiotic era. Nature Reviews Microbiology. 2009; 7: 629-41. Link
6. DeLeo FR, Chambers HF. Re-emergence of antibiotic-resistant *Staphylococcus aureus* in the genomics era. J Clin Invest., 2009; 119(9): 2464–2474. Link
7. Jevons MP. "Celbenin"-resistant staphylococci. Br Med J 1961; 124:124–125. Link
8. 8 Jevons MP, Coe AW, Parker MT. Methicillin resistance in staphylococci. Lancet 1963;1:904-907. Link
9. Assadullah S, Kakru DK, Thoker MA, Bhat FA, Hussain N, Shah A. Emergence of low level Vancomycin resistance in MRSA. Indian J Med Microbiol 2003;21:196-198. Link
10. National Committee for Clinical Laboratory Standards. 2003. Approved standard: M2-A8. Performance standards for antimicrobial disk susceptibility tests, 8th ed. National Committee for Clinical Laboratory Standards, Wayne, Pa.
11. Cox RA, Conquest C, Mallaghan C, Marple RR. A major outbreak of methicillin resistant *Staphylococcus aureus* caused by a new phage type (EMRSA-16). J Hosp Infect. 1995;29:87–106. Link
13. Saikia L, Nath R, Choudhury B, Sarkar M. Prevalence and antimicrobial susceptibility pattern of methicillin-resistant *Staphylococcus aureus* in Assam. Indian Journal of Critical Care Medicine : Peer-reviewed, Official Publication of Indian Society of Critical Care Medicine. 2009;13(3):156-158. Link

14. Ahmad S, Alenzi FQ, Al-Juaid NF, Ahmed S. Prevalence and antibiotic susceptibility pattern of methicillin resistant *Staphylococcus aureus* at Armed Forces Hospital in Saudi Arabia. Bangladesh Med Res Counc Bull. 2009;35(1):28-30. Link

15. Tiwari S, Sahu M, Rautaraya B, Karuna T, Mishra SR, Bhattacharya S. Prevalence of methicillin-resistant *Staphylococcus aureus* and its antibiotic susceptibility pattern in a tertiary care hospital. J Indian Med Assoc. 2011;109(11):800-1. Link

16. Sharma NK, Garg R, Baliga S, Bhat K. G. Nosocomial Infections and Drug Susceptibility Patterns in Methicillin Sensitive and Methicillin Resistant *Staphylococcus aureus*. Journal of Clinical and Diagnostic Research : JCDR. 2013;7(10):2178-2180. Link

17. Datta P, Gulati N, Singla N, Vasdeva HR, Bala K, Chander J, Gupta V. Evaluation of various methods for the detection of meticillin-resistant *Staphylococcus aureus* strains and susceptibility patterns. J. Med. Microbiol. 2011;60:1613-1616. Link

18. Goyal A, Diwakar MK, Bhooshan S, Goyal S, Agrawal A. Prevalence and Antimicrobial Susceptibility Pattern of Methicillin-resistant *Staphylococcus aureus* [MRSA] isolates at a Tertiary Care Hospital in Agra, North India – A systemic annual review. IOSR Journal of Dental and Medical Sciences (IOSR-JDMS) 2013;11(6):80-84. Link.

19. Sharma NK, Garg R, Baliga S, Bhat K. G. Nosocomial Infections and Drug Susceptibility Patterns in Methicillin Sensitive and Methicillin Resistant *Staphylococcus aureus*. Journal of Clinical and Diagnostic Research : JCDR. 2013;7(10):2178-2180. Link

20. Tiwari S, Sahu M, Rautaraya B, Karuna T, Mishra SR, Bhattacharya S. Prevalence of methicillin-resistant *Staphylococcus aureus* and its antibiotic susceptibility pattern in a tertiary care hospital. J Indian Med Assoc. 2011;109(11):800-1. Link.