Diagnostic Value of Optical Coherence Tomography Image Features for Diagnosis of Basal Cell Carcinoma

Fieke ADAN1,2, Klara MOSTERD1,2, Nicole W. J. KELLENERS-SMEETS1,2 and Patty J. NELEMANS3
1Department of Dermatology, Maastricht University Medical Centre, 2GROW Research Institute for Oncology and Developmental Biology, Maastricht University and 3Department of Epidemiology, Maastricht University, Maastricht, The Netherlands

Optical coherence tomography (OCT) is a non-invasive diagnostic method. Numerous morphological OCT features have been described for diagnosis of basal cell carcinoma (BCC). The aim of this study is to evaluate the diagnostic value of established OCT features and to explore whether the use of a small set of OCT features enables accurate discrimination between BCC and non-BCC lesions and between BCC subtypes. For each lesion, the presence or absence of specific OCT features was recorded. Histopathology was used as a gold standard. Diagnostic parameters were calculated for each OCT feature, and multivariate logistic regression analyses were performed to evaluate the loss in discriminative ability when using a small subset of OCT features instead of all features that are characteristic for BCC according to the literature. The results show that the use of a limited number of OCT features allows for good discrimination of superficial BCC from non-superficial BCC and non-BCC lesions. The prevalence of BCC was 75.3% (225/299) and the proposed diagnostic algorithm enabled detection of 97.8% of BCC lesions (220/225). Subtyping without the need for biopsy was possible in 132 of 299 patients (44%), with a predictive value for presence of superficial BCC of 84.3% vs 98.8% for presence of non-superficial BCC.

Key words: basal cell carcinoma; optical coherence tomography; diagnostic; non-invasive.

Accepted Nov 1, 2021; Epub ahead of print Nov 1, 2021
Acta Derm Venereol 2021; 101: adv00607.

Corr: Fieke Adan, Department of Dermatology, Maastricht University Medical Centre, P. Debyelaan 25, NL-6229 HX Maastricht, The Netherlands. E-mail: fieke.adan@mumc.nl

The incidence of non-melanoma skin cancer (NMSC) has increased over the past decades, with basal cell carcinoma (BCC) being the most prevalent cancer diagnosed in the Caucasian population worldwide (1–4). Although there are many histopathological subtypes, a simplified classification by Rippey roughly groups all BCCs into 3 subtypes: superficial, nodular, and aggressive (5). Currently, histopathological examination of a punch biopsy is the gold standard to discriminate BCC from alternative diagnoses and to determine the histopathological subtype (6).

In recent years, optical coherence tomography (OCT) has emerged as a promising non-invasive diagnostic method for diagnosis of BCC, generating real-time in vivo cross-sectional images of tissue microarchitecture with a depth of approximately 1.5 mm (7). OCT is based on light interferometry; the interference of 2 optical beams reflected by tissue produces different shades in the black and white spectrum. Despite the high lateral resolution of <7.5 mm and axial resolution of <5 mm, imaging of individual cells is not possible. OCT is therefore suitable for pattern recognition in tissue, similar to, for example, ultrasound, allowing for the identification of morphological features of BCC, which have been established in recent years (8–12). In 2015, Hussain et al. systematically reviewed diagnostic criteria for BCC. The authors evaluated 17 studies and found that, in 100% of these studies, rounded dark (hyporeflective) structures in the upper dermis, surrounded by a bright (hyper-reflective) halo, sometimes surrounded by a hyporeflective border and disruption of epidermal layering, were described as characteristic for BCC (8). The rounded dark structures resemble the basaloid cell nests seen in histology, the hyper-reflective halo surrounding the rounded structures corresponds to the surrounding tumour stroma and a hyporeflective border at the periphery resembles the peripheral palisading at the margins of basaloid cell nests (13). Other features that have been described for BCC are shown in Table I, some of which are illustrated in Fig. 1 (9–12).

It is not known which OCT features are most discriminative for BCC diagnosis. Therefore, the primary aim of
this study is to evaluate the diagnostic value of established morphological OCT features that can be used for diagnosis and subtyping of BCC. The second objective is to explore whether the use of a small set of features with the highest predictive value enables accurate discrimination between BCC and non-BCC lesions and between BCC subtypes.

**MATERIALS AND METHODS**

Data for this study were derived from 1 arm of a randomized multi-centre non-inferiority trial in 1 academic and 2 general Dutch hospitals: the dermatology outpatient clinic of Maastricht University Medical Centre+ (Maastricht), Catharina Hospital (Eindhoven) and Zuyderland Medical Centre (Heerlen). Included were consecutive patients (18 years or older) with an indication for skin biopsy of a lesion clinically suspected for BCC. Excluded were patients with a lesion located in the “H-zone” of the face, patients with a large BCC referred to our head and neck tumour working group, and patients who were unable to sign informed consent. The marked biopsy area of the clinically most aggressive part was scanned with OCT without any preparation of the skin in advance (Vivosight Multi-beam Swept-Source Frequency Domain OCT, Michelson Diagnostics, Maidstone, Kent, UK).

OCT scanning and interpretations were conducted by a single experienced investigator (FA), who used morphological OCT features that are characteristic for identifying BCC lesions (8–10, 14). For each lesion, the presence or absence of specific morphological OCT features was recorded (Table I). Features relating to the epidermis included: superficial scaling/crust/ulceration, atrophy of the epidermis, protrusions into the upper dermis with dark rim, superficial basaloid nests with a firm connection to the epidermis, dermal-epidermal junction interrupted/poorly defined. Features relating to the dermis included: signal-poor ovoid structures, ovoid structures with bright centre, dark rim/hyperreflective border, bright (hyper-reflective) peritumoural stroma/hyper-reflective halo, prominent vessels, small ovoid signal-poor structures “shoal of fish” or “bunch of grapes”, dark/black (areflective) areas/cysts, basaloid nests in the dermis, basaloid nests with necrotic cell debris in centre, peripheral palisading, collagen compression between adjacent nests, dilated capillaries, nests in morpheaform and micronodular basal cell carcinoma, respectively, area of liquefactive necrosis.

**Table I. Optical coherence tomography (OCT) image features and corresponding histopathology features (8–12, 19)**

| OCT Feature                                | Histopathology Feature                                |
|--------------------------------------------|-------------------------------------------------------|
| Superficial scaling/crust/ulceration       | Superficial scaling/crust/ulceration                  |
| Atrophy of the epidermis                   | Atrophy of the epidermis                              |
| Thickening of the epidermis                | Thickening of the epidermis                           |
| Protrusions into the upper dermis with dark rim | Superficial basaloid nests with a firm connection to the epidermis |
| Dermo-epidermal junction                   | Interrupted/poorly defined                            |
| Epidermis                                  |                                                       |
| Superficial scaling/crust/ulceration       | Superficial scaling/crust/ulceration                  |
| Atrophy of the epidermis                   | Atrophy of the epidermis                              |
| Thickening of the epidermis                | Thickening of the epidermis                           |
| Superficial basaloid nests with a firm connection to the epidermis | Superficial basaloid nests with a firm connection to the epidermis |
| Dermo-epidermal junction                   | Interrupted/poorly defined                            |
| Dermis                                     |                                                       |
| Signal-poor ovoid structures/rounded dark structures | Basaloid nests in the dermis                          |
| Ovoid structures with bright centre         | Basaloid nests with necrotic cell debris in centre     |
| Dark rim/hyperreflective border            | Peripheral palisading                                 |
| Bright (hyper-reflective) peritumoural stroma/hyper-reflective halo | Collagen compression between adjacent nests           |
| Prominent vessels                          | Dilated capillaries                                   |
| Small ovoid signal-poor structures “Shoal of fish” or “Bunch of grapes” | Nests in morpheaform and micronodular basal cell carcinoma, respectively |
| Dark/black (areflective) areas/cysts       | Area of liquefactive necrosis                         |

Fig. 1. Optical coherence tomography (OCT) images of different basal cell carcinoma (BCC) subtypes. (A) OCT image of a superficial BCC. Red arrow points towards a protrusion into the upper dermis with a dark rim, corresponding to a basaloid cell nest with a firm connection to the epidermis (10). Vessels in the upper dermis are dilated (blue arrow) and the dermal epidermal junction (DEJ) is disrupted (yellow arrow). (B) OCT image of a nodular BCC. Red arrows point towards a fully encompassing signal-poor ovoid structure located in the dermis with a dark rim and bright peritumoural stroma. Inside the nest, well-circumscribed black areas are observed, representing signs of liquefactive necrosis (19). Vessels in the upper dermis are dilated and directed towards the tumour nests (blue arrow). The epidermis above the nest is atrophic, the DEJ is disrupted (yellow arrow). (C) OCT image of an aggressive BCC. Inside the red rectangle, smaller signal-poor ovoid structures with surrounding bright peritumoural stroma are visible, also described as “shoal of fish” or “bunch of grapes”, indicating an aggressive BCC subtype (9).
The study was approved by the Medical Ethical Committee of MUMC+(METC 18-043). All patients provided written informed consent.

Statistical analysis

The distribution of baseline characteristics in patients who underwent OCT examination was summarized by absolute numbers and percentages for categorical variables and by mean values with standard deviations (SD) or median with range for continuous variables. Diagnostic parameters were calculated for each morphological OCT feature. Diagnostic odds ratios (DOR) with 95% confidence intervals (95% CI) were calculated using univariate logistic regression analyses.

Subsequently, multivariate logistic regression analyses were performed to evaluate the loss in discriminative ability when using a small subset of morphological features instead of all morphological features that are characteristic for BCC according to the literature (8–10). Histologically verified presence or absence of BCC was the dependent variable. Discriminative ability was expressed as the area under the receiver operating characteristic curve (AUC) and differences in AUCs between models were tested for significance using an algorithm for paired comparison of AUCs from DeLong et al. (15).

Similar analyses were performed to identify subsets of features that can be used for accurate subtyping. These analyses were restricted to the subgroup of patients with histologically confirmed BCC. Discrimination between superficial BCC and non-superficial BCC subtypes was used as dependent variable. A DOR > 1 indicates that superficial BCC can be treated non-invasively. Therefore, histologically verified superficial BCC vs non-superficial BCC subtypes was used as dependent variable. A DOR = 1 indicates that the presence of a feature is indicative for the presence of superficial BCC vs non-superficial BCC:

BCC: basal cell carcinoma; SCC: squamous cell carcinoma.

| Characteristic            | Presence in BCC (%)* (Sensitivity) | Absence in non-BCC (%) (Specificity) | Probability of BCC if present (PPV) | Probability of non-BCC if absent (NPV) | Odds ratio (95% CI) | p-value |
|---------------------------|------------------------------------|--------------------------------------|-------------------------------------|---------------------------------------|---------------------|---------|
| **Epidermis**             |                                    |                                      |                                     |                                       |                     |         |
| Superficial scaling/crust/ ulceration | 57.3 (129/225)                   | 40.5 (30/74)                        | 74.6 (129/173)                     | 23.8 (30/126)                         | 0.92 (0.54–1.56)   | 0.79    |
| Atrophy of the epidermis  | 40.8 (92/225)                     | 94.6 (70/74)                        | 95.8 (92/96)                       | 34.5 (70/203)                        | 12.11 (4.27–34.32) | <0.0001 |
| Thickening of the epidermis | 16.0 (36/226)                  | 31.1 (23/74)                        | 41.4 (36/87)                       | 10.8 (23/212)                        | 0.09 (0.05–0.16)   | <0.0001 |
| Protrusions into upper dermis with dark rim | 61.3 (138/225)            | 90.5 (67/74)                        | 95.2 (138/145)                     | 43.5 (67/154)                        | 15.18 (6.66–34.59) | <0.0001 |
| **Dermal–epidermal junction** |                                    |                                      |                                     |                                       |                     |         |
| Dermal–epidermal junction poorly defined/interrupted Dermis | 74.7 (168/225)            | 75.7 (56/74)                        | 90.3 (168/186)                     | 49.6 (56/113)                        | 9.17 (4.98–16.88) | <0.0001 |
| **Dermis**                |                                    |                                      |                                     |                                       |                     |         |
| Signal-poor ovoid structures | 69.8 (157/225)                | 79.7 (59/74)                        | 91.3 (157/172)                     | 46.5 (59/127)                        | 9.08 (4.82–17.12) | <0.0001 |
| Ovoid structures with bright centre | 28.9 (65/225)                | 93.2 (69/74)                        | 92.9 (65/70)                       | 30.1 (69/229)                        | 5.61 (2.16–14.53) | <0.0001 |
| Dark rim                  | 96.4 (217/225)                   | 70.3 (52/74)                        | 90.8 (217/239)                     | 86.7 (52/60)                         | 64.11 (27.02–152.11) | <0.0001 |
| Bright peritumoral stroma | 81.3 (183/225)                   | 91.9 (68/74)                        | 96.8 (183/189)                     | 61.8 (68/110)                        | 49.38 (20.09–121.40) | <0.0001 |
| Prominent vessels         | 56.9 (128/226)                   | 44.6 (33/74)                        | 75.7 (128/169)                     | 25.4 (33/130)                        | 1.06 (0.63–1.80)   | 0.89    |
| Shoil of fish/bunch of grapes | 22.2 (50/225)                 | 97.3 (72/74)                        | 96.2 (50/52)                       | 29.1 (72/247)                        | 10.29 (2.44–43.40) | <0.0001 |
| Black areas, cysts        | 27.6 (62/225)                    | 95.9 (71/74)                        | 95.4 (62/65)                       | 30.3 (71/234)                        | 9.00 (2.73–29.64)  | <0.0001 |

*Data subset includes 225 BCCs and 74 non-BCCs.

PPV: positive predictive value; NPV: negative predictive value; 95% CI: 95% confidence interval.

225 (75.3%) patients had a BCC and 74 had an alternative diagnosis. Of these 225 BCCs, 66 were superficial, 79 nodular and 12 aggressive. The remaining 68 BCCs were of a mixed subtype (Table II).

**RESULTS**

A total of 598 patients with 598 skin lesions clinically suspicious for BCC were included in the randomized trial between March 2019 and September 2020. Data for 299 patients, who were randomized to the OCT group, were used for this study. According to histological diagnosis,
peritumoural stroma (OR 49.38, 95% CI 20.09–121.40). Protrusions into the upper dermis with a dark rim (OR 15.18, 95% CI 6.66–34.59), poorly defined/interrupted dermal epidermal junction (DEJ) (OR 9.17, 95% CI 4.98–16.88) and signal-poor ovoid structures (OR 9.08, 95% CI 4.82–17.12) were also strongly associated with presence of BCC. The ORs were statistically significant \((p<0.0001)\) for all features, except superficial scaling/crust/ulceration \((p=0.79)\) and prominent vessels \((p=0.89)\).

The AUC of the full multivariate logistic model including all 12 morphological features was 95.2% (95% CI 91.8–98.7%). The AUC of a model including the 5 features with the highest DORs was 94.7% (95% CI 91.4–98.1%). A final model included the 4 features that were most discriminative, based on clinical experience of the investigator: signal-poor ovoid structures, dark rim, bright peritumoural stroma and protrusions into the upper dermis with a dark rim. For this model, the AUC decreased to 94.1% (95% CI 90.8–97.3%). However, the decreases in AUC when using 4 or 5 features instead of all features were minor and non-significant \((p=0.49\) and \(p=0.24\), respectively). Based on these results we conclude that 4 features can be used to discriminate between BCC and non-BCC without significant loss of diagnostic performance.

### Discriminating between superficial basal cell carcinoma and non-superficial basal cell carcinoma subtypes

Table IV shows the frequency of each morphological feature, as well as diagnostic parameters for discrimination between superficial BCC and non-superficial BCC subtypes. The diagnostic odds ratio (DOR) was significant for 8 features. The high DOR for “protrusions into the upper dermis with a dark rim” and an “interrupted DEJ” indicates that the presence of these features is strongly indicative for presence of superficial BCC (sBCC). The other 6 features: (i) atrophy of the epidermis, (ii) signal-poor ovoid structures, (iii) ovoid structures with bright centre, (iv) bright peritumoural stroma, (v) shoal of fish/bunch of grapes, and (vi) black areas, cysts are associated with DORs that are significantly lower than 1, indicating that these features are highly predictive for non-superficial BCC.

The AUC of the full multivariate logistic model including all 12 features was 95.5% (95% CI 93.0–98.0). The use of a model including the 8 features that were associated with a significantly increased or decreased DOR resulted in a comparable AUC \((p=0.88)\) of 95.6% (95% CI 93.2–98.0%). Use of a subset of 6 of these 8 features (excluding “bright peritumoural stroma” and “interrupted DEJ”) led to a significant \((p=0.01)\) decrease in the AUC (94.0% with 95% CI 91.1–97.0%).

### Diagnostic algorithm

Fig. 2 proposes a diagnostic algorithm for diagnosis and subtyping of BCC. First, to discriminate between BCC and non-BCC lesions, one can use the 4 features: (i) dark rim, (ii) bright peritumoural stroma, (iii) protrusions, and (iv) signal-poor ovoid structures. If 1 or more of these features are present, the probability of BCC is 89.4% (PPV), whereas if all 4 features are absent the probability of a non-BCC lesion is 90.6% (NPV).

With respect to subtyping, absence of the feature “protrusions into the upper dermis with a dark rim” indicates a high probability of non-superficial BCC. However, when this feature is present, both superficial and non-superficial BCC can be present. For further discrimination, additional features that increase the probability of non-superficial BCC can be used: (i) atrophy of the epidermis, (ii) signal-poor ovoid structures, (iii) ovoid structures with bright centre, (iv) shoal of fish/bunch of grapes, and (vi) black areas, cysts are associated with DORs that are significantly lower than 1, indicating that these features are highly predictive for non-superficial BCC.

### Table IV. Prevalence of morphological features in superficial basal cell carcinoma (BCC) and non-superficial BCC subtypes in patients with histologically verified BCC. Odds ratios (OR) of univariate logistic regression analyses and \(p\)-values

| Characteristic | Presence in sBCC (%) \(n=66\) (Sensitivity) | Absence in non-superficial BCC subtype (%) \(n=159\) (Specificity) | Probability of sBCC if present (PPV) | Probability of non-superficial BCC subtype if absent (NPV) | Odds ratio\(^*\) (95% CI) | \(p\)-value |
|----------------|---------------------------------------------|---------------------------------------------|-------------------------------------|-------------------------------------|------------------------|----------|
| Epidermis      |                                             |                                             |                                     |                                     |                        |          |
| Superficial scaling/crust/ulceration | 56.1 (37/66) | 42.1 (67/159) | 28.7 (37/129) | 69.8 (67/96) | 0.93 (0.52–1.66) | 0.88            |
| Atrophy of the epidermis | 9.1 (6/66) | 45.9 (73/159) | 6.5 (6/92) | 54.9 (73/133) | 0.09 (0.04–0.21) | <0.001   |
| Thickening of the epidermis | 22.7 (15/66) | 86.8 (138/159) | 41.7 (15/36) | 73.0 (138/189) | 1.93 (0.93–4.04) | 0.11       |
| Protrusions into upper dermis with dark rim | 95.5 (63/66) | 52.8 (84/159) | 45.7 (63/138) | 96.6 (84/87) | 23.52 (7.09–78.04) | <0.001   |
| Dermal–epidermal junction poorly defined | 98.5 (65/65) | 35.2 (56/159) | 38.7 (65/168) | 98.2 (56/57) | 35.34 (4.78–261.55) | <0.001   |
| Dermal–epidermal junction poorly defined | 98.5 (65/65) | 35.2 (56/159) | 38.7 (65/168) | 98.2 (56/57) | 35.34 (4.78–261.55) | <0.001   |
| Dermal–epidermal junction poorly defined | 98.5 (65/65) | 35.2 (56/159) | 38.7 (65/168) | 98.2 (56/57) | 35.34 (4.78–261.55) | <0.001   |
| Dermis         |                                             |                                             |                                     |                                     |                        |          |
| Signal-poor ovoid structures | 19.7 (13/66) | 9.4 (15/159) | 8.3 (13/157) | 22.1 (15/68) | 0.03 (0.01–0.06) | <0.001   |
| Ovoid structures with bright centre | 9.1 (6/66) | 62.9 (100/159) | 9.2 (6/65) | 62.5 (100/160) | 0.17 (0.07–0.42) | <0.001   |
| Dark rim       | 95.5 (63/66) | 3.1 (5/159) | 29.0 (63/217) | 62.5 (5/8) | 0.68 (0.16–2.94) | 0.70       |
| Bright peritumoural stroma | 65.2 (43/66) | 11.9 (19/159) | 23.5 (43/183) | 45.2 (19/42) | 0.25 (0.13–0.51) | <0.001   |
| Prominent vessels | 43.9 (29/66) | 37.7 (60/159) | 22.7 (29/128) | 61.9 (60/97) | 0.48 (0.27–0.85) | 0.01       |
| Shoal of fish/bunch of grapes | 0.0 (0/66) | 68.6 (109/159) | 0.0 (0/50) | 62.3 (109/175) | 0\(a\) | <0.001   |
| Black areas, cysts | 9.1 (6/66) | 64.8 (102/159) | 9.7 (6/62) | 63.2 (102/163) | 0.18 (0.08–0.45) | <0.001   |

\(a\)Cannot be calculated.

\(^*\)An odds ratio > 1 indicates higher probability of superficial BCC and an odds ratio < 1 indicates higher probability of non-superficial subtype.

PPV: positive predictive value; NPV: negative predictive value; 95% CI: 95% confidence interval.
bunch of grapes, or (v) black areas, cysts. If all 5 features are absent and protrusions are present \((n=51)\), the probability of superficial BCC is 84.3%. If 1 or more of the 5 features are present and protrusions are absent \((n=81)\), there is a high probability of non-superficial BCC subtype (98.8%). In case of other combinations, there remains too much doubt and a punch biopsy is still necessary to determine the histological subtype. For instance, when both protrusions and 1 or more of the other 5 features are present, the probability of superficial BCC and non-superficial BCC is 23% and 77%, respectively.

Summarizing, the use of the diagnostic algorithm enabled detection of 97.8% of BCC lesions (220/225). In 132 of 299 patients (44%) a diagnosis of subtype could be made with high predictive value for presence of superficial BCC (84.3%) vs presence of non-superficial BCC (98.8%) without the need for biopsy.

**DISCUSSION**

This study evaluated the diagnostic value of morphological OCT features for BCC diagnosis. With the features: (i) dark rim, (ii) bright peritumoural stroma, (iii) protrusions into the upper dermis with a dark rim, and (iv) signal-poor ovoid structures (all with PPV >90%) a good discrimination between BCC and non-BCC lesions is possible. With regard to subtyping, “protrusions into the upper dermis with a dark rim” are visible in the vast majority of superficial BCCs and absence of this feature is highly predictive of non-superficial BCC when other BCC features are present. However, if “protrusions” are present, a conclusive diagnosis cannot be made. In this situation, 5 other features that are highly predictive of non-superficial BCC have to be used.

Accurate diagnosis of BCC with OCT, or other non-invasive methods, would enable the use of a one-stop shop approach in patients with a lesion clinically suspected for BCC. This approach requires that superficial BCCs can be distinguished from non-superficial BCCs in a substantial proportion of patients. Patients with superficial BCC can be treated non-invasively (i.e. imiquimod cream, photodynamic therapy). Therefore, these patients would benefit most from accurate non-invasive diagnosis.

In case of high suspicion of sBCC, non-invasive treatment can be discussed with the patient and initiated in the same visit, and an invasive procedure with risk of pain, bleeding and scar formation can be prevented. In case of high suspicion of non-superficial BCC, a surgical excision can be planned immediately. This one-stop shop approach could result in a reduction in the number of biopsies and is expected to be more efficient, patient friendly and cost-effective than regular care, where diagnosis and treatment is based on the histopathological result of a punch biopsy (16, 17).

The proposed diagnostic algorithm offers a systematic approach towards discrimination of superficial BCC from non-superficial BCC and non-BCC lesions using OCT. The first step, discrimination of BCC from non-BCC lesions, is possible with 4 features (PPV of 89.4) There is a risk (of approximately 10%) that a lesion that is diagnosed as BCC by OCT is not a BCC. The prevention of misclassification of another cutaneous malignancy as BCC is the biggest challenge. Cheng et al. evaluated the diagnostic accuracy of OCT in lesions for which sBCC was considered in the differential diagnosis. In their series, one amelanotic melanoma was misclassified as sBCC. In the current study cohort, one patient had a
superficial spreading malignant melanoma, clinically highly suspected for BCC. However, upon OCT examination, no BCC features were present, so this case was diagnosed as a non-BCC lesion with an indication for biopsy. The risk of misclassification of an amelanotic melanoma as BCC is small, but future research on morphological OCT features that can help to distinguish melanoma from BCC is needed.

The proposed diagnostic algorithm also enables discrimination of sBCC from non-superficial BCCs and non-BCC lesions, but there remains a 15.7% chance that non-superficial BCC is misclassified as sBCC, if protrusions are present and 1 or more of the other 5 features are absent. Such misclassifications may result in a higher risk of residual or recurrent BCC if non-invasive treatment is chosen. To verify that misclassifications by OCT do not compromise patient safety, a randomized controlled trial, which compares effectiveness of an OCT-guided diagnosis with regular care (biopsy) is necessary.

The diagnostic algorithm uses 6 features for subtyping, although the discriminative ability of a multivariate diagnostic model using 6 features was significantly worse than that of a model with 8 features, which also included “bright peritumoural stroma” and “interrupted DEJ”. Still, the latter 2 features were not used in the algorithm. The reason is that the multivariate model evaluates the probability of sBCC given all combinations of the presence and/or absence of features, whereas the diagnostic algorithm considers only a limited number of possible combinations. Absence of “protrusions” in combination with 1 or more of features indicative of non-superficial BCC results in high probability of non-superficial BCC being present. However, in this respect, addition of the feature “bright peritumoural stroma”, which is indicative of non-superficial BCC, was not helpful, because its prevalence is also high in superficial BCC. The feature “interrupted DEJ” was also left out. It had little added value, because it nearly always occurred in combination with the feature “protrusions into the upper dermis with a dark rim” (except for 2 lesions).

In this study, the selection of features that are useful for diagnosis and subtyping of BCC was based on reports from the literature. A limitation is that there is high variance in the use of terminology in the literature. Describing the 4 features that discriminate well between BCC and non-BCC lesions, Hussain et al. (8) referred to: (i) rounded dark structures in the upper dermis (corresponding to signal-poor ovoid structures); (ii) surrounded by a hyper-reflective halo (bright peritumoural stroma); (iii) possibly surrounded by a hyporeflective border (dark rim); and (iv) disruption of epidermal layering (poorly defined/interrupted DEJ). Cheng et al. (10) concluded that “protrusions into the upper dermis with a dark rim” are highly predictive for superficial BCC, and described this feature as “hyporeflective ovoid structures with firm attachment to the DEJ and a clefting region focused or solely visible at the inferior margin”. The high prevalence of an atrophic epidermis and ovoid structures with bright centre (referred to as intranodular small bright dots) in nodular BCC has also been reported by other studies (11, 12). Since the evidence for the use of OCT for BCC diagnosis is increasing, more uniformity in terminology is desirable for future implementation of OCT for BCC diagnosis. This study used conventional OCT, whereas the use of dynamic OCT provides additional information by visualizing the vascular patterns and thus allows for better differentiation between BCC subtypes. Therefore, with use of dynamic OCT the proposed diagnostic algorithm may lead to even better diagnostic performance, which can be evaluated in future studies (18).

A limitation of this study is that predictive values, such as PPV and NPV, are highly dependent on the prevalence of BCC and sBCC subtypes in a study population. In this study, patients were included with lesions suspected for BCC, based on clinical and dermoscopic examination by a dermatologist, leading to a study population with a high prevalence of BCC (75.3%). In other study populations, where for BCC suspected lesions would have been selected by physicians with less experience in clinical and dermoscopic examination, the prevalence of BCC might be lower, which could result in different predictive values of the morphological features. Furthermore, the proposed diagnostic algorithm is intended as support for OCT users who do not yet have much experience with interpretation of OCT images. The trial aimed to evaluate whether OCT-guided diagnosis and treatment does not compromise patient safety compared with regular care (always biopsy). For ethical reasons, it was decided to not yet include high-risk patients with large lesions or lesions in the “H-zone”. Furthermore, the H-zone surface areas are often convex or concave, making it more difficult to obtain an OCT image of sufficient quality. More studies are therefore required to determine whether OCT is suitable in this subpopulation. Also, the diagnostic algorithm needs to be validated in future studies.

In conclusion, the results of this study show that the use of a limited number of morphological features allows for good discrimination of superficial BCC from non-superficial BCC and non-BCC lesions, with the potential to obviate the need for punch biopsy in 44% of patients. Hence, novice OCT assessors commencing OCT training could focus initially on recognizing these selected features.

ACKNOWLEDGEMENTS

The study was financed by a grant from the Netherlands Organisation for Health Research and Development (ZonMw; 852001928). ZonMw is a governmental institution financing research to improve healthcare in the Netherlands. The funding source had no role in the design or conduct of the study; collection, management, analysis or interpretation of the data; or preparation, review or approval of the manuscript.
The authors have no conflicts of interest to declare.

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