Deciphering the triad of infection, immunity and pathology

The factors which drive and control disease progression can be inferred from mathematical models that integrate measures of immune responses, data from tissue sampling and markers of infection dynamics.

FREDERIK GRAW

A fever, a cough, a splitting headache... Being sick often comes with tell-tale signs which worsen as the disease progresses and tissues become damaged. These symptoms result from complex interactions between the infecting pathogen, the inflammation process, and the response from the immune system. Tracking these mechanisms and how they interact, as well as identifying which factors determine when the disease recedes or progresses, is essential for establishing better treatment strategies.

In this effort, a more refined understanding of infection and immune responses has emerged from combining experimental and clinical measurements with mathematical models (Perelson, 2002). However, it is still difficult to link tissue pathology and disease severity with viral load or immune cell counts, which respectively measure the amount of virus and of certain immune actors in the body. Now, in eLife, Amber Smith and colleagues at St. Jude Children’s Research Hospital, the University of Tennessee Health Science Center and the Washington University School of Medicine – including Margaret Myers and Amanda Smith as joining first authors – report how viral infection, counteracting immune responses and lung pathology interact as mice fight off influenza A (Myers et al., 2021).

First, the team tracked how viral load and the number of CD8+ T cells, an important immune actor that helps to clear infected cells, progressed over time. In combination with mathematical models, these measurements allowed Myers et al. to estimate several parameters that reflect the pace at which the virus replicates, the strength of the immune response, and the interactions between these processes. While this had already been achieved in previous studies (e.g. Baccam et al., 2006), Myers et al. also analyzed the anatomy of the lung tissue over time, assessing the damage caused by infection and inflammation as well as how much the organ eventually regenerates.

Then, the team compared these data to values from their mathematical model that described viral load and CD8+ T cells counts, thereby linking viral load dynamics and specific immune responses to disease pathology and severity (Figure 1). In particular, the analysis shed light on how the relative number of...
immune cells correlates with the level of lung tissue cleared from the virus and, thus, the mice’s ability to recover from infection. These quantitative relationships could help to assess how well the virus is controlled within tissues simply by relying on easily accessible markers that are, for example, present in the blood. This would reduce the need for invasive tissue samples.

Individual molecular processes and specific aspects of viral replication can be studied extensively within in vitro cell culture systems. However, the full triad of infection, immunity and especially tissue pathology can only be reliably assessed within conditions that are physiologically relevant (Fackler et al., 2014). Indeed, simple cell culture systems insufficiently address the impact tissue structure can have on infection dynamics, immune activation and clearing mechanisms (Fackler et al., 2014; Imle et al., 2019).

Myers et al. used frequent samples and histological analyses to infer how infected tissues change over time. Yet, imaging technologies may continue to improve so that it becomes possible to observe the interactions between host and pathogen within tissues in real-time (Coombes and Robey, 2010). These approaches could help to investigate whether quantitative relationships as highlighted by Myers et al. also play a role in other infections and in other tissues. The expanding field of organoids – whereby simple, miniature organs are grown in the laboratory – also represents a promising step towards understanding how cells interact within structured, tissue-related environments (Gosselin et al., 2018; Bar-Ephraim et al., 2020). Combined with new technologies such as single-cell sequencing methods (Triana et al., 2021; Youk et al., 2020), these approaches will help to determine the molecular processes that govern disease progression, and how these might differ between patients.

Despite these new experimental and diagnostic technologies, data-driven mathematical modeling and analytical methods will continue to fulfil a key role for deciphering the interplay between infection, tissue pathology and disease severity. Using these models makes it possible to integrate different types of measurements from various places and times, and to disentangle the contributions of individual processes to the infection dynamics. It is only by understanding exactly how individual processes interact over time that scientists will be able to find and validate prognostic markers which predict disease progression.

Frederik Graw is in the BioQuant (Center for Quantitative Biology) at Heidelberg University, Heidelberg, Germany.

Competing interests: The author declares that no competing interests exist.

Published 01 September 2021
Bar-Ephraim YE, Kretzschmar K, Clevers H. 2020. Organoids in immunological research. Nature Reviews Immunology 20:279–293. DOI: https://doi.org/10.1038/s41577-019-0248-y, PMID: 31853049

Coombes JL, Robey EA. 2010. Dynamic imaging of host-pathogen interactions in vivo. Nature Reviews Immunology 10:353–364. DOI: https://doi.org/10.1038/nnr2746, PMID: 20395980

Fackler OT, Murooka TT, Imle A, Mempel TR. 2014. Adding new dimensions: towards an integrative understanding of HIV-1 spread. Nature Reviews Microbiology 12:563–574. DOI: https://doi.org/10.1038/nrmicro3309, PMID: 25029025

Gosselin EA, Eppler HB, Bromberg JS, Jewell CM. 2018. Designing natural and synthetic immune tissues. Nature Materials 17:484–498. DOI: https://doi.org/10.1038/s41563-018-0077-6, PMID: 29784994

Imle A, Kumberger P, Schnellbächer ND, Fehr J, Carrillo-Bustamante P, Ales J, Schmidt P, Ritter C, Godinez WJ, Müller B, Rohr K, Hamprecht FA, Schwarz US, Graw F, Fackler OT. 2019. Experimental and computational analyses reveal that environmental restrictions shape HIV-1 spread in 3D cultures. Nature Communications 10:2144. DOI: https://doi.org/10.1038/s41467-019-09879-3, PMID: 31086185

Myers MA, Smith AP, Lane LC, Moquin DJ, Aogo R, Woolard S, Thomas P, Vogel P, Smith AM. 2021. Dynamically linking influenza virus infection kinetics, lung injury, inflammation, and disease severity. eLife 10:e68864. DOI: https://doi.org/10.7554/eLife.68864, PMID: 34282728

Perelson AS. 2002. Modelling viral and immune system dynamics. Nature Reviews Immunology 2:28–36. DOI: https://doi.org/10.1038/nri700, PMID: 11905835

Triana S, Metz-Zumaran C, Ramirez C, Kee C, Doldan P, Shahraz M, Schraivogel D, Gschwind AR, Sharma AK, Steinmetz LM, Herrmann C, Alexandrov T, Boulang S, Stanifer ML. 2021. Single-cell analyses reveal SARS-CoV-2 interference with intrinsic immune response in the human gut. Molecular Systems Biology 17:e10232. DOI: https://doi.org/10.15252/msb.202110232

Youk J, Kim T, Evans KV, Jeong YI, Hur Y, Hong SP, Kim JH, Yi K, Kim SY, Na KJ, Bleazard T, Kim HM, Fellows M, Mahbubani KT, Saeb-Parsy K, Kim SY, Kim YT, Koh GY, Choi BS, Ju YS, et al. 2020. Three-dimensional human alveolar stem cell culture models reveal infection response to SARS-CoV-2. Cell Stem Cell 27:905–919. DOI: https://doi.org/10.1016/j.stem.2020.10.004, PMID: 33142113