An adult patient with suspected of monkeypox infection differential diagnosed to chickenpox

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Abstract

Background: Monkeypox is a zoonosis. The disease has a similar appearance to chickenpox caused by the varicella-zoster virus (VZV). On May 9th 2019, there was one laboratory-confirmed case of monkeypox reported in Singapore. A man was also suspected of having monkeypox on June 1st 2019 in Surabaya, Indonesia, a neighboring country.

Objective: To report on a suspected case of monkeypox with differential diagnosis to chickenpox.

Case: A 51-year-old male was suspected of having monkeypox after a differential diagnosis of chickenpox. His chief complaint was multiple blisters on his body. From the dermatological status on his facial, trunk and extremity regions, there were multiple pleomorphic vesicles, some with umbilication, with a centripetal distribution, and crusts.

Methods and Results: A PCR using VZV specific primers, followed by genome sequencing showed homologies of more than 99% to other VZVs and less than 50% to Monkeypox sequences.

Conclusion: Molecular laboratory techniques have confirmed the case as chickenpox.

Introduction

Monkeypox is a rare zoonotic disease caused by the monkeypox virus which belong to Orthopoxvirus genus in the family of Poxviridae. It is transmitted when a person comes into contact with virus from an animal, a human, or any material contaminated with the disease. The virus enters the body through broken skin (even if not visible), the respiratory tract, or the mucous membranes (eyes, nose, or mouth). Animal-to-human transmission may occur by bites or scratches, bush meat preparation, direct contact with body fluids or lesion material, or indirect contact with lesion material, such as through contaminated bedding. The first human case of monkeypox was recorded in 1970 in the Democratic Republic of Congo during a period of intensified effort to eliminate smallpox. An outbreak happened in the USA in 2003 when the Centers for Disease Control and Prevention (CDC) received reports of patients with a febrile rash illness who had close contact with pet prairie dogs and other animals. Of the 87 monkeypox cases, 20 (23%) were laboratory confirmed at the CDC. In 2017-2018 a monkeypox outbreak happened in Nigeria. While in May 2019, the Ministry of Health in Singapore notified the WHO of one laboratory-confirmed case of monkeypox. The case-patient was a 38 years old Nigerian man who arrived in Singapore on 28th April 2019. Prior to his travel to Singapore, he had worked in Nigeria in the Delta state, and had attended a wedding on 21st April 2019 in a village in Ebenyi State. The patient developed fevers, muscle aches, chills and a skin rash on 30th April. The patient reported that he had remained in his hotel room most of the time between 1st and 7th May. He was transferred to a public hospital by ambulance on 7th May and referred to the National Centre for Infectious Diseases (NCID) on the same day, where he was isolated for further management. Skin lesion samples were taken on 8th May and showed positive for monkeypox virus by the National Public Health Laboratory on the same day. The Complications of monkeypox include secondary bacterial infections, respiratory distress, bronchopneumonia, gastrointestinal involvement, dehydration, sepsis, encephalitis, and corneal infections with ensuing loss of vision. The case fatality rate for Monkeypox is 10%, lying between the case fatality rate of variola (or smallpox) major (30%) and variola minor (1%).

Chickenpox caused by VHZ has become the primary differential diagnosis because it has a similar clinical appearance. It is distributed worldwide, but it seems that temperate and tropical climates are also affect age-specific incidences. It is typically seen in children from 1 to 9 years of age. Primary infection in adults is usually more severe. A study in the Democratic Republic of Congo (formerly Zaire) showed that among 730 patients diagnosed as cases of chickenpox, 3.3% had monkeypox by laboratory diagnostic testing and amongst these 7.3% were diagnosed as “atypical chickenpox”. Laboratory confirmation of infection is critical, because human monkeypox closely resembles several other febrile rash illnesses including smallpox and chickenpox.
spread to his back, chest, and upper arms. This was accompanied by an itchy sensation. The patient had a contact history of feeding a monkey that belonged to the patient’s neighbour, but the owner of the monkey did not have any symptoms or abnormalities. A history of travelling prior to these complaints was denied. A history of chickenpox or vaccination before was also denied. Two weeks before, the patient’s son was diagnosed with chickenpox.

From a physical examination, his general condition appeared to be sick. The temperature was 36.6°C. There was no lymph node enlargement on the cervical, axillar, or inguinal regions. From the dermatological status (Figure 1), on the facial region, there were multiple pleiomorphic vesicles (multiple stages of development) and multiple crusts. On the trunk, extremities (superior and inferior regions), there were multiple pleiomorphic vesicles (multiple stages of development), some of them with umbilication and centripetal distribution. A Laboratory examination revealed a hemoglobin level at 12.5 g/dL, a leucocytes level at 6,870/μL, with a neutrophils level at 65.4%. Vesicle fluid was collected for PCR amplification with specific primers for VZV. Patient sample DNA was extracted using Qiagen. The extracted DNA was used as a template for the amplification of the respective Open Reading Frame (ORF) 14.

PCRs were performed with the PCR master mix (Promega, GoTaq Green Master Mix), 200 μL primers for VZV detection (10 picomoles/microliter) and 20 μL positive control (Herpesvirus genome 45 ng/mL). The thermocycling condition included a 5-min denaturation step of 94°C, followed by 40 cycles of 30 sec at 94°C, 30 sec at 53°C, and 45 sec at 72°C. ORF 14 was amplified using primers VZV R2 Fw (5’-CGGTAAATCTGGCATGCG-3’) and VZV R2 Rv (5’- AAGGG GAGCGTG-GATGTGTC–3’). Nucleotide sequences of the amplified fragments were determined using the BigDye Deoxy Terminator cycle sequencing kit with an AB 3500XL Genetic Analyzer (Applied Biosystems, Foster City, CA). The result was positive and matching.

Table 1. Nucleotide sequence homology of the patient to other viruses from the GenBank database.

| Patient Nucleotide Sequence (813 bp) | Reported isolates from the GenBank | Percentage homology, % |
|-------------------------------------|----------------------------------|------------------------|
| Sby-1.gnu 813                       | KM355703.1 *                     | 99.385                 |
|                                    | MH709324 *                       | 99.262                 |
|                                    | KJ642616.1 **                    | 46.885                 |
|                                    | NC_003310.1 **                   | 46.885                 |
|                                    | AY243312 ***                    | 45.395                 |
|                                    | KC813493 ****                   | 44.737                 |

* Sequences of VZV, ** of Monkeypox, *** of Vaccinia, and **** of Cowpox Viruses, indicated by the accession numbers.
with positive control in the range of 810 bp (Figure 2). It is confirmed at more than 98% sequence homology to several published VZVs from the GenBank. The patient treated with acyclovir. After 10 days of treatment, all the skin lesions became crusted and the patient was discharged. A written informed consent was obtained.

Discussion

Monkeypox presents challenges for public health officials and healthcare personnel in terms of surveillance and laboratory capacities, and management and treatment of disease. On May 9th 2019, there was one laboratory-confirmed case of monkeypox reported in Singapore. This situation causes an awareness of the outbreak of monkeypox in Indonesia considering that Singapore is located near to Indonesia and the mobility of travelers between these countries is high. However, the authority had warned that patients with suspected monkeypox should be provided with more careful and accurate laboratory examinations to establish a diagnosis. The patient suspected with monkeypox had been admitted to Dr. Soetomo General Academic Hospital. Outbreak investigations commenced immediately: isolation of the suspected case-patient, a laboratory examination, and contact tracing were conducted. A PCR examination, followed by genome sequencing, was completed. PCR has a very high sensitivity and specificity, ready availability, and a relatively quick (one day or less) turnaround time. For detection of VZV, vesicle fluid is the best specimen for PCR analysis, but lesion scrapings, crusts, VZV, vesicle fluid is the best specimen for PCR examination, followed by genome sequencing and the results obtained were compared with reported Nucleotide Sequence Data from the Genbank. From several nucleotide sequences that were compared, results showed high homology to KM355703.1-HHV3-Bandim as much as 99.38% and to MH709324-HHV3-USA as much as 99.26%. Both of them are VZV strains. Whereas the homology to KJ642616.1-Monkeypox-Liberia nucleotides was only 46.88%, and to NC_003310.1-MonkeyPox nucleotides was as low as 46.88% (Table 1).

Based on this data, the possibility of monkeypox infection in this patient could be ruled out and the diagnosis of chickenpox could be established.

Conclusions

A VZV of an adult case in Surabaya, Indonesia, which was clinically similar to a more pathogenic Monkeypox, was confirmed by a molecular diagnostic laboratory test. We are relieved, since a confirmed monkeypox was recently found in an African citizen who came into Singapore, a neighbouring country.

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