Modulation of immunity in young-adult and aged squirrel, *Funambulus pennanti* by melatonin and p-chlorophenylalanine

Seema Rai, Chandana Haldar and Rajesh Singh

Address: Department of Zoology, Pineal Research Lab, Banaras Hindu University, Varanasi – 221005, India

E-mail: Seema Rai - raiseema_pk@rediffmail.com; Chandana Haldar* - chaldar2001@yahoo.com; Rajesh Singh - rajeshsingh@rediffmail.com

*Corresponding author

Published: 23 April 2009 Received: 23 December 2008

*Immunity & Ageing* 2009, 6:5 doi: 10.1186/1742-4933-6-5 Accepted: 23 April 2009

This article is available from: http://www.immunityageing.com/content/6/1/5

© 2009 Rai et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License ([http://creativecommons.org/licenses/by/2.0](http://creativecommons.org/licenses/by/2.0)), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

**Background:** Our interest was to find out whether pineal gland and their by melatonin act as modulator of immunosenescence. Parachlorophenylalanine (PCPA) – a β adrenergic blocker, is known to perform chemical pinealectomy (Px) by suppressing indirectly the substrate 5-hydroxytryptamine (5-HT) for melatonin synthesis and thereby melatonin itself. The role of PCPA, alone and in combination with melatonin was recorded in immunomodulation and free radical load in spleen of young adult and aged seasonal breeder Indian palm squirrel *Funambulus pennanti*.

**Results:** Aged squirrel presented reduced immune parameters (i.e. total leukocyte count (TLC), Lymphocytes Count (LC), % stimulation ratio of splenocytes (% SR) against T cell mitogen concanavalin A (Con A), delayed type hypersensitivity (DTH) to oxazolone) when compared to young adult group. Melatonin administration (25 μg/100 g body mass/day) significantly increased the immune parameters in aged more than the young adult squirrel while PCPA administration (4.5 mg/100 g body mass/day) reduced all the immune parameters more significantly in young than aged. Combination of PCPA and melatonin significantly increased the immune parameters to the normal control level of both the age groups. TBARS level was significantly high in aged than the young group. PCPA treatment increased TBARS level of young and aged squirrels both while melatonin treatment decreased it even than the controls. Nighttime peripheral melatonin level was low in control aged group than the young group. Melatonin injection at evening hours significantly increased the peripheral level of nighttime melatonin, while combined injection of PCPA and melatonin brought it to control level in both aged and young adult squirrels.

**Conclusion:** PCPA suppressed immune status more in aged than in adult by reducing melatonin level as it did chemical Px. Melatonin level decreased in control aged squirrels and so there was a decrease in immune parameters with a concomitant increase in free radical load of spleen. Decreased immune status can be restored following melatonin injection which decreased free radical load of spleen and suggest that immune organs of aged squirrels were sensitive to melatonin. Increased free radical load and decreased peripheral melatonin could be one of the reasons of immunosenescence.
Background
Melatonin (N-acetyl-5-methoxytryptamine) is synthesized mainly in pineal gland. It plays a major role in regulating sexual maturity, reproductive cycle, stress and the immune responses [1-3]. On the other hand, it has also been observed that melatonin declines with age [4] and may act as a key regulator in ageing and senescence [5,6]. Aging is associated with a decline in immune function known as immunosenescence. This situation implies increased susceptibility to infectious disease due to decreased capacity of the immune system to respond to antigenic stimulation [7]. Many hormones also decline with advancing age (growth hormone (GH), estrogen, dehydroepiandrosterone and melatonin) playing a significant role in immunosenescence [8]. Among those, melatonin has been demonstrated to bear a general immune-enhancing effect in many animal species including humans [9]. Whether this immune-enhancing property of melatonin exists in aged animals needs to be verified. Pharmacological and surgical pinealectomy also modulate various immune parameters including plaque-forming cells and blastogenic responses of splenocytes and thymocytes to various mitogens [10]. Further, disruption of the night-time peak of melatonin completely abolished Colony Forming Unit-Granulocyte Macrophages (CFU-GM) proliferation of bone marrow cells [11,12].

PCPA has been proposed as an indirect antagonist of melatonin as it inhibits the melatonin biosynthesis pathway by acting on tryptophane hydroxylase the enzyme responsible for synthesis of serotonin (serotonin is the substrate for melatonin). PCPA, being a specific depletor of serotonin (5-HT) contributes also to the prenatal development of the central nervous system, acting as a morphogen in the young embryo and later as a neurotransmitter [18,19]. If endogenous melatonin synthesis is blocked by administration of propanolol (PRO) or p-chlorophenylalanine (PCPA) during evening hrs, it, significantly depresses the antibody production in human and mice [20,21].

Further, the basic problem of aging is the declined efficiency in all organisms. This will lead to dysfunction and degeneration, increase in free radical load and reactive oxygen species (ROS) in particular. Further, direct free radical scavenging [22] and indirect antioxidant stimulatory property of melatonin was used in number of studies for improving metabolic function [23,24]. Hardly any report exist explaining the age related changes in immune function and free radical load along with protective effect of melatonin on them in any wild mammals. Therefore, in the present study we recorded the immune status and free radical load of aged and compared it with young adult male squirrel, Funambulus pennanti on one hand and effect of melatonin, PCPA (a specific depletor of serotonin level) and PCPA plus melatonin on the improvement of immune functions on the other. We selected Indian palm squirrel, Funambulus pennanti as animal model because most of the parameters and doses for melatonin and PCPA related to present study has already been established which eventually helped in planning this study.

Materials and methods
All the experiments on the animals were conducted in accordance with Institutional practice and with the framework of revised Animals (Specific Procedure) Act of 2002 of Govt. of India on animal welfare.

Animal Maintenance
Squirrels were collected from the vicinity of Varanasi (Latitude 25°, 18 ' N, Longitude 83°, 01' E) as they were available in plenty of numbers and were maintained in laboratory under ambient conditions adapted for several weeks for young adult squirrels and years for aged squirrels. Young adults (weighing 70 ± 10 g, ~5 months old) and aged (weighing 140 ± 10 g, ~2.0 years old) squirrels as judged by incisor length and cranium diameter, [25] were used for present study. The animals were maintained in an open air fenced area, where they can move freely. During experiment they were housed in wire net cages (25" × 25" × 30" in size) in the same housing area. The squirrels were fed with soaked gram, Cicer arietinum along with other seasonal food grains, nuts and water ad libitum. Squirrels were then randomized and divided into following groups containing 12 squirrels as mentioned in Table 1.

Drug and treatment protocols
Para-chlorophenylalanine (PCPA) and melatonin were purchased from (SIGMA, St. Louis, Missouri, USA). The control squirrels were injected (subcutaneously) with the normal ethanolic (20 μl) saline (0.9%NaCl) 0.1 ml/day.

Table 1:

| Groups               | Treatment               | No. of young/aged squirrels |
|----------------------|-------------------------|-----------------------------|
| Control              | 0.9% saline/day         | 12/12                       |
| PCPA treated         | 4.5 mg/squirrel/day     | 12/12                       |
| PCPA + Melatonin     | 4.5 mg/squirrel/day + 25 μg/squirrel/day | 12/12 |
| Melatonin            | 25 μg/squirrel/day      | 12/12                       |
Melatonin solution was made by dissolving it in few drops (20 μl) of ethanol and then diluted with normal saline (0.9% NaCl) up to the desired concentration. PCPA was dissolved in 1 N NaOH and diluted with PBS. Melatonin 25 μg/squirrel/day (subcutaneously; S.C.) and PCPA, 4.5 mg/squirrel/day (intra peritoneal; i. p.) at evening hrs. (4.30–5.30 pm, i.e. 1.30 hrs before lights off) were injected daily for 60 consecutive days.

At the end of the experiment (60th days), six animals from each group of young adult and aged were subjected for the Delayed Type Hypersensitivity (DTH) response test for ear swelling tests according to Phanuphak et al. (1974) and Disis et al. (2000) [26,27]. Remaining six squirrels following their sacrifice at evening hrs (5.00 – 6.00 PM) after complete anesthesia (anesthetic ether) were subjected for spleen weight analysis, MDA assay [28] and blastogenic response of splenocytes. Blood plasma was separated by centrifugation and was stored at -20° for radioimmunoassay (RIA) of melatonin. Blastogenic response of splenocytes was noted in terms of tritiated thymidine uptake (3H-TdR; Specific activity 8.9 ci mM; BARC Mumbai, India) against T cell mitogen concanavalin A [29] and expressed in terms of percent stimulation ratio (% SR) of splenocytes.

\[
\text{% Stimulation Ratio} = \frac{\text{cpm with mitogen} - \text{cpm without mitogen}}{\text{cpm without mitogen}} \times 100
\]

**Lipid peroxidation (LPO) assay**

Free radicals have a very short half-life, which makes them very hard to measure in the laboratory. A commonly used alternate approach measures markers of free radicals rather than the actual radical. Thiobarbituric acid reactive substances (TBARS) are produced during oxidative damage to cell membrane. Malondialdehyde (MDA), one of the major lipid breakdown product and commonly used parameter will be assessed by the method of Ohkawa et al. 1978 [28].

All spleens were excised and weighed for the preparation of 10% tissue homogenates in 20 mM Tris Hydrochloride (HCl) buffer (pH 7.4). The homogenates were centrifuged at 3000 g for 15 min at 4°C and supernatant was subjected to thiobarbituric acid (TBA) assay by mixing it with 8.1% SDS, 20% acetic acid, 0.8% TBA and boiling for 1 h at 95°C. The reaction mixture was immediately cooled in running water and vigorously shaken with n-butanol and pyridine reagent (15:1) and centrifuged for 10 min at 1500 g. The absorbance of the upper phase was measured at 534 nm. LPO was expressed as TBARS in nmol/g tissue wt., by taking 1,1,3,3 tetraethoxy propane (TEP) as standard. The standard curve was calibrated using 10 nM concentration of TEP.

**Statistics**

All the data were expressed as means ± SEM of at least 6 animals per point. Data comparisons were statistically analyzed by using ANOVA followed by Student Newman-Keuls’ multiple range tests among the groups while, student t test was performed to compare the aged group with young one. The differences were considered statistically significant when \( p < 0.05 \).
than fifty percent decrease in spleen weight of aged squirrel was recorded when compared with young group.

PCPA treatment decreased spleen weight of aged and young both groups of squirrels when compared with saline treated control group. Further, a combined treatment of melatonin along with PCPA significantly \((P < 0.01)\) increased the spleen weight of aged squirrels only without affecting the spleen weight of young adult squirrel when compared with their respective controls. Moreover, melatonin alone given to the aged squirrels showed a significant \((P < 0.01)\) increase in spleen weight when compared with control, PCPA and PCPA plus melatonin treated groups (Figure 1).

**Total leukocyte and lymphocyte count (TLC and LC)**

There was a decrease in no of TLC and LC in control aged group when compared with control young adult group. PCPA treatment significantly \((P < 0.01)\) decreased TLC and LC of young adult and aged squirrels when compared with their respective control. Further, melatonin and PCPA treatment to young adult and aged squirrels significantly \((P < 0.01)\) enhanced the TLC and LC when compared with the control and PCPA treated groups. Melatonin alone to the aged squirrels significantly \((p < 0.01)\) increased TLC and LC when compared with the control and PCPA treated aged squirrels and had similar value as the combined treatment of PCPA plus melatonin (Figure 2a, b). Melatonin alone or in combination with PCPA had no significant effect on TLC and LC of young adult group when compared with young adult control group.

**Delayed Type Hypersensitivity (DTH) response to oxazolone**

Aged squirrels presented a general decline in DTH response than the young adult squirrels. A significant decrease of DTH response was noted following the PCPA treatment in

![Figure 1](image1.png)

**Figure 1**

**Effect of p-chlorophenylalanine and melatonin treatment on spleen weight of young adult and aged squirrels.** Histogram represents Mean ± SE; \(n = 12\) for each group. Con = Control, PCPA = para-chlorophenylalanine, PCPA + Mel = para-chlorophenylalanine + Melatonin, Mel = Melatonin Significance of difference aged vs young adult; \(* = P < 0.05; ** = P < 0.01\) Significance of difference treated aged vs treated young adult; \(a = P < 0.05; b = P < 0.01\).

![Figure 2](image2.png)

**Figure 2**

**Effect of p-chlorophenylalanine and melatonin treatment on total leukocyte (TLC; Figure 2a.) and lymphocyte count (LC; Figure 2b) of young adult and aged squirrels.** Histogram represents Mean ± SE; \(n = 12\) for each group. Con = Control, PCPA = para-chlorophenylalanine, PCPA + Mel = para-chlorophenylalanine + Melatonin, Mel = Melatonin Significance of difference aged vs young adult; \(* = P < 0.05; ** = P < 0.01\) Significance of difference treated aged vs treated young adult; \(a = P < 0.05; b = P < 0.01\).
young squirrel when compared with the control group while this decline was less evident in aged group. Combined treatment of melatonin and PCPA had no effect on DTH response of young adult but aged squirrels showed significant increased when compared with their respective saline treated control group of squirrels. Melatonin treatment alone increased significantly (P < 0.01) DTH response in both aged but not in young squirrels when compared with saline treated control group (Figure 3).

Blastogenic response of splenocytes in terms of percent stimulation ratio (% SR)
Control aged squirrel had significantly less %SR than young adult. PCPA treatment significantly (P < 0.01) decreased % SR of splenocytes in young adult and aged squirrels when compared with saline treated control group. Further, combined treatment of PCPA and melatonin to young adult and aged squirrels had no recovery effect on % SR of splenocytes when compared with saline treated control group. In addition melatonin treatment alone to the young adult squirrel could not show any effect while a significant (P < 0.01) increase was noted in % SR of splenocytes of aged squirrel when compared with their respective saline treated control, PCPA and PCPA plus melatonin treated groups (Figure 4).

Radioimmunoassay of Melatonin
Peripheral melatonin level was significantly less in aged group of squirrel than young adult group. Significant (P < 0.01) decrease in plasma melatonin was noted in young as well as aged squirrels following PCPA treatment when compared with their saline treated control group. Combined treatment of PCPA and melatonin in aged squirrels had no effect on melatonin level but it decreased melatonin level in young adult squirrels when compared with saline treated control group. However, melatonin treatment alone significantly increased its level in aged and young adult squirrels (Figure 5).

TBARS level
Significant increase was noted in TBARS level of aged squirrels as compared with the young group. PCPA treatment caused significant increase in lipid peroxidation in both the age groups while melatonin in combination with PCPA reduced it to the level of control group. However, injection of melatonin alone significantly decreased the TBARS level in young as well as in aged squirrels when compared with saline treated control group (Figure 6).

Figure 3
Effect of p-chlorophenylalanine and melatonin treatment on Delayed Type Hypersensitivity (DTH) response to oxazolone (in terms of ear thickness) of young adult and aged squirrels. Histogram represents Mean ± SE; n = 12 for each group. Con = Control, PCPA = para-chlorophenylalanine, PCPA + Mel = para-chlorophenylalanine + Melatonin, Mel = Melatonin. Significance of difference aged vs young adult; *= P < 0.05; **= P, < 0.01. Significance of difference treated aged vs treated young adult; a = P < 0.05; b = P, < 0.01.

Figure 4
Effect of para-chlorophenylalanine and melatonin treatment on blastogenic response against T cell mitogen Concanavalin (Con A) in terms of percent stimulation ratio (%SR) of young adult and aged squirrels. Histogram represents Mean Mean ± SE; n = 12 for each group. Con = Control, PCPA = para-chlorophenylalanine, PCPA + Mel = para-chlorophenylalanine + Melatonin, Mel = Melatonin. Significance of difference aged vs young adult; *= P < 0.05; **= P, < 0.01. Significance of difference treated aged vs treated young adult; a = P < 0.05; b = P, < 0.01.
Discussion

Researchers are actively engaged in solving the age related decline in various physiological functions. Evidences supporting the properties of melatonin that affect the ageing process comes from the studies in mice where pathological changes resembling senescence occurred when pineal gland was destroyed. When pineal gland of young mice was grafted in older animals they exhibited a prolonged life span \[32,33\]. We have already proposed that melatonin act as an immunostimulator in *Funambulus pennanti* when tested in *in vivo* as well as *in vitro* \[8\]. Whether the immunostimulatory effect of melatonin is age dependent in this seasonal breeder, *F. pennanti* is being reported here in order to specify the role of melatonin in immunosenescence. We used an indirect blocker of melatonin synthesis i.e. PCPA to support it. Our data clearly suggest an age related decline of the immune system (almost all the parameters) associated with the decrease in plasma melatonin level \[34\]. Further, a significant suppression of spleen weight following treatment of PCPA in young squirrels was noted while, aged squirrels showed no such reduction as they had already reduced spleen weight when compared with the young adult squirrels. Combined treatment of melatonin and PCPA showed a significant increase in splenic mass when compared with control and PCPA treated group. This indicates that melatonin is able to compromise the PCPA induced suppression of spleen mass and thereby immunity. The magnitudes of the PCPA effect in reducing immune activity were more prominent in young squirrels than the aged ones. Our above observation support the earlier report of Maestrini and his coworkers (1986) \[35\] that deprivation of nocturnal circulating melatonin by constant light exposure (physiological pinealectomy) and administration of propanolol or PCPA (chemical pinealectomy) reduced the splenic and thymic cellularity and that pinealectomy caused thymic involution \[36\] pointing towards the significance of the melatonin in the maintenance of the lymphoid organ activity. This could be the reason behind observation when we noted that PCPA significantly suppressed the total leukocyte and lymphocyte count in young adult squirrels, while no suppression was noted in the aged squirrels as their TLC and LC number were already quite low. However, melatonin treatment to the aged squirrel increased the peripheral TLC and LC once more supporting the immunostimulatory action of it which is independent of age.

The Delayed Type Hypersensitivity (DTH) response to oxazolone as a measure of T-cell mediated immune function \[26,27\] and splenocytes proliferative response to the mitogen Con A which reflected that PCPA treatment significantly suppressed the cell mediated immunity and immune status while melatonin
treatment improved the suppressed immune functions, but the magnitude of the response was more prominent in aged squirrels than the young squirrels. Decrease in plasma melatonin level following PCPA signifies its action as chemical pinealectomy in young adult more prominently.

The present result of increased lipid peroxidation (LPO) production in aged squirrels suggest involvement of free radicals and the role of redox imbalance could be a result of immunosenescence which was restored following the melatonin treatment. The data of the present study is also in support of the previous literatures suggesting the role of PCPA on increased lipid peroxidation products and reduced glutathione content in renal and brain tissue [37]. Our results also support the additive pro-oxidant action of PCPA in spleen to exacerbate the oxidative stress during ageing when the LPO was noted very high which might have reduced immune functions as well as proved the possible antioxidant property of melatonin. Melatonin stimulates the production of glutathione [38] and therefore immunoenhancing role of melatonin following its administration may be partly due to its influence on the maintenance of intracellular glutathione level.

PCPA treatment was more effective in reducing the immune parameter via reducing melatonin level in young group, whereas in aged squirrels it did not suppress the immune parameters significantly because of (i) low level of melatonin and (ii) already low level of immunity. This is the reason that melatonin treatment in young squirrels could not affect the studied immune parameters whereas in aged squirrels it was more effective in inducing the immune status.

Conclusion
The result reported here indicates that the immunomodulatory role of melatonin is age independent and the immunosenescence is due to low level of melatonin production with age. The mechanism by which melatonin exerts its immuno-potentiating action could be partially explained via its action in reducing the free radical load [39].

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
RS participated in execution of the experiments. SR was involved in estimations (RIA and Free radicals etc.), statistical analysis, drafting of manuscript etc. CH made contribution in conception of experimental project, planning, interpretation of data critical evaluation of MS for intellectual content and has given final touch in preparation of MS for publication.

Acknowledgements
Authors are thankful to Department of Science and Technology, New Delhi, India for financial assistance. Instrument gift of Alexander von Humboldt Foundation, Germany is gratefully acknowledged.

References
1. Maestroni GJM, Conti A and Pierpaoli W: Role of the pineal gland in immunity. III, Melatonin antagonizes the immunosuppressive effect of acute stress via an opiatergic mechanism. Urology 1988, 32:339–343.
2. Maestroni GJM and Conti A: Melatonin in relation to the immune system. Melatonin Biosynthesis, Physiological effect and clinical applications Boca Raton, CRC Press: Yu HS, Reiter Rj 1993, 189–210.
3. Guerrero JM and Reiter RJ: Melatonin immune system relationship. Curr Top Med Chem 2002, 2:167–179.
4. Reiter RJ, Craft CM, Johnson JE, King TS Jr, Richardson BA, Vaughan GM and Vaughan MK: Age-associated reduction in nocturnal pineal melatonin levels in female rats. Endocrinology 1981, 109:1295–1297.
5. Pierpaoli W and Regelson W: Pineal control of aging: Effect of melatonin and pineal grafting on aging mice. Proc Natl Acad Sci USA 1992, 91:787–789.
6. Liu sj and Wang ZJ: Alzheimer-like tau phosphorylation induced by wortmannin in vivo and its attenuation by melatonin. Acta Pharmacol Sin 2002, 23:183–187.
7. Pawelec G, Akbar A, Caruso C, Effros R, Grubeck-Loebenstein B and Wilkby A: Is Immunosenescence infectious?. Trends Immunol 2004, 25:406–410.
8. Arlt W and Haldar C: Hormones and immune function: implications of aging. Aging Cell 2004, 3:209–216.
9. Nelson RJ: Seasonal immune function and sickness responses. Trends Immunol 2004, 25:187–192.
10. Rai S and Haldar C: Pineal Control of immune status and hematological changes in blood and bone marrow of male squirrels (Funambulus pennantii) during their reproductively inactive phase. Comp Biochemistry and Physiology (C) 2003, 136:319–328.
11. Haldar C, Haussler D and Gupta D: Response of colony forming units for granulocyte macrophages (CFU-GM) of intact and pinealectomized rat bone marrow cells to murine recombinant interleukin-3 (rIL-3), recombinant granulocyte macrophage colony stimulating factor (rGM-CSF) and human recombinant erythropoietin (rEPO). Prog Brain Res 1991, 91:323–325.
12. Haldar C, Haussler D and Gupta D: Effect of the pineal gland on circadian rhythmicity of colony forming units for granulocytes and macrophages (CFU-GM) from rat bone marrow cell cultures. J Pineal Res 1992, 12:79–83.
13. Marco J, Roiero O, Villa I, Esquifino A and Larralde J: Effect of p-chlorophenylalanine on LH, FSH, prolactin and ovulation in the rat. Rev Esp Fisiol 1982, 38:285–290.
14. Steinman JL, Carlton SM, Haber B and Willis WD: Differential effects of p-chlorophenylalanine on indoleamines in brainstem nuclei and spinal cord of rats. J Neurochem 1987, 46:297–309.
15. Ramachandran A, Patel M and Patel C: Effects of pineal indoles and parachlorophenylalanine on seasonal reproduction in the pigeon. J Exp Biol 1996, 199:793–800.
16. Ieni JR and Thurmond JB: Maternal aggression in mice: effects of treatments with PCPA, 5-HTP and 5-HT receptor antagonists. Eur J Pharmacol 1985, 111:211–220.
17. Nakajima K, Matsuyma K and Mori S: Prenatal administration of para-chlorophenylalanine results in suppression of serotonergic system and disturbance of swimming movements in newborn rats. Neurosci Res 1998, 31:135–169.
18. Shishkina GT and Dygalo NN: Role of the serotoninergic system in the acceleration of sexual maturation in wild Norway rats selected for reduced aggressiveness toward humans. Comp Biochem Physiol C Toxicol Pharmacol 2000, 125:45–51.
19. Butkevich IP, Khozhai LI, Mikhailenkov VA and Ostellin VA: Decreased serotonin level during pregnancy alters morphological and functional characteristics of tonic nociceptive
