The frequency of genes encoding three putative group B streptococcal virulence factors among invasive and colonizing isolates

Shannon D Manning¹, Moran Ki², Carl F Marrs³, Kiersten J Kugeler⁴, Stephanie M Borchardt⁵, Carol J Baker⁶ and Betsy Foxman*³

Address: ¹National Food Safety and Toxicology Center and Department of Pediatrics and Human Development, Michigan State University, East Lansing, Michigan, USA, ²Department of Preventive Medicine, Eulji University School of Medicine, Daejeon, Korea, ³Department of Epidemiology, University of Michigan School of Public Health, Ann Arbor, Michigan, USA, ⁴Centers for Disease Control and Prevention, Division of Vector-Borne Infectious Diseases, Bacterial Zoonoses Branch, Fort Collins, Colorado, USA, ⁵Fargo Veterans Administration Medical Center, Fargo, North Dakota, USA and ⁶Department of Pediatrics, Molecular Virology and Microbiology, Baylor College of Medicine, Houston, Texas, USA

Email: Shannon D Manning - Shannon.Manning@ht.msu.edu; Moran Ki - kimoran@eulji.ac.kr; Carl F Marrs - cmarrs@umich.edu; Kiersten J Kugeler - bio1@cdc.gov; Stephanie M Borchardt - Stephanie.Borchardt@va.gov; Carol J Baker - cbaker@bcm.tmc.edu; Betsy Foxman* - bfoxman@umich.edu

* Corresponding author

Abstract

Background: Group B Streptococcus (GBS) causes severe infections in very young infants and invasive disease in pregnant women and adults with underlying medical conditions. GBS pathogenicity varies between and within serotypes, with considerable variation in genetic content between strains. Three proteins, Rib encoded by rib, and alpha and beta C proteins encoded by bca and bac, respectively, have been suggested as potential vaccine candidates for GBS. It is not known, however, whether these genes occur more frequently in invasive versus colonizing GBS strains.

Methods: We screened 162 invasive and 338 colonizing GBS strains from different collections using dot blot hybridization to assess the frequency of bca, bac and rib. All strains were defined by serotyping for capsular type, and frequency differences were tested using the Chi square test.

Results: Genes encoding the beta C protein (bac) and Rib (rib) occurred at similar frequencies among invasive and colonizing isolates, bac (20% vs. 23%), and rib (28% vs. 20%), while the alpha (bca) C protein was more frequently found in colonizing strains (46%) vs. invasive (29%). Invasive strains were associated with specific serotype/gene combinations.

Conclusion: Novel virulence factors must be identified to better understand GBS disease.

Background

Group B Streptococcus (GBS) causes sepsis and meningitis in young infants, febrile complications in pregnant women and invasive disease in adults with underlying medical conditions [1]. Capsular polysaccharide, which defines GBS serotype, is the primary virulence factor found in most GBS strains, and different serotypes contribute to disease in different populations. For example, 30% of GBS disease in non-pregnant adults is caused by serotype V [2], while serotype III causes more than 70% of
infant meningitis and most late-onset (7–89 days of age) disease [3]. Vaccines currently under development target the most prevalent GBS serotypes [4].

Other than the polysaccharide capsule, little is known about other GBS components important in pathogenesis. Many putative virulence factors and genes have been identified recently (for a review see [5]), though most are either present in all GBS strains, or are lacking sufficient data to pinpoint their role in the pathogenic process. Three proteins, however, have been studied extensively and were recommended as potential GBS vaccine candidates [6-8]. These include the protein Rib [7] encoded by rib [9], and the alpha [10] and beta [10] C proteins encoded by bca [11] and bac [12], respectively. All three proteins trigger antibody production that offers protection from GBS infection in animal models [7,8,13], though the frequency of these proteins and the genes that encode them varies by disease status [14-19] as well as serotype. For example, Rib has been found predominantly in serotype III strains [7]. To date, large, population-based studies comparing the frequencies of genes encoding the Rib, alpha and beta C proteins among invasive and colonizing isolates have been limited.

Methods

We describe the frequency of genes encoding three virulence factors among five GBS strain collections including invasive (n = 162) and colonizing (n = 338) isolates (Table 1); invasive disease status was not known for 29 strains. All isolates tested were obtained with the approval of an appropriate institutional ethics committee. Invasive isolates originated from the blood or cerebrospinal fluid (CSF) of newborns <7 days of age (n = 100), the urine of college students (n = 4) [20,21] and pregnant women presenting to the University of Michigan Medical Center (UMMC) for prenatal care associated with GBS isolation from the urine (n = 58), and the placenta following delivery (n = 5) [22]. Newborn isolates, described by Zaleznik et al. [23] (n = 65), were collected between 1993 and 1996, while the remainder (n = 35) came from the same Houston hospitals between 1997 and 2000. Colonizing GBS isolates were from the anal orifice or urine of healthy male (n = 58) college students, the anal orifice, vagina, cervix or urine or healthy female (n = 86) college students [21], pregnant women from UMMC (n = 49) [22], and sexually active college women with a urinary tract infection not caused by GBS (n = 102) and their most recent male sex partner (n = 43) [20]. Among colonizing isolates, 17 (6.9%) were from individuals colonized with multiple isolates in multiple sites; the dot blot profile was determined only for those isolates that were unique by pulsed-field gel electrophoresis (PFGE) as described previously [20,21,24].

Serotyping using hyperimmune rabbit antisera to GBS polysaccharide types Ia, Ib, and II-VIII was performed as described previously [20,21]. We amplified DNA for genes encoding bca, bac and rib using PCR (Table 2). Control strain A909 was used to amplify bca and bac [25], while BM110 was used for rib [9]. PCR DNA was purified and fluorescein-labeled as described previously [26].

DNA was isolated using a modified E. coli protocol [27] in which cells were lysed overnight. Dot blot hybridization and subsequent analyses were performed as described previously [26,28] with two negative and positive controls per membrane. The signal intensity of each dot was reported as a percentage of the positive control present on each membrane in ImageQuant (Molecular Dynamics, CA). Percentages were corrected for the background signal of the negative controls and graphed. The x-axis represented values from one membrane and the y-axis consisted of values from the duplicate membrane. A cutoff was established based on each graph distribution [26]. Isolates within the intermediate range were repeated. Sixty-eight hybridizations yielded equivocal results despite repeated probing, and thus, were confirmed for the presence or absence of each gene by PCR and sequencing using the same primers described in Table 2. Eleven remained equivocal following PCR and were excluded from the analyses.

Chi square tests were used to assess differences in gene frequencies by collection and serotype. SAS was used for all statistical analyses [29].

Results

Across GBS strain collections, the bca gene occurred most frequently, followed by rib and bac (Table 3). bca and bac occurred most frequently among colonizing isolates from college students (Collections 1, 2, and 4, Table 1), while the frequency of rib was similar across collections. Because gene frequency varies by capsular serotype, we described the frequency of each gene by serotype (Figure 1). When assessing the frequencies among invasive versus colonizing isolates, only isolates from newborns with GBS disease (n = 100) were considered invasive, while colonizing isolates consisted of those isolates known to not cause a UTI and those that were isolated during pregnancy as part of routine GBS screening (n = 360). In this analysis, rib occurred slightly more frequently among invasive versus colonizing isolates (p = .09) (Table 4), while both bac (p = .55) and bca (p = .002) occurred less frequently in the invasive strains.

After stratifying by serotype, invasive versus colonizing capsular serotype Ia strains were significantly less likely to have bca (p = .002), while Ib invasive strains were more likely to have bca (p = .03) (Table 4). Invasive versus col-
onizing capsular serotype III strains, however, were more likely to have both rib (p = .09) and bac (p = .06), and less likely to have bca (p = .09).

Because a previous study also indicated that rib occurs more frequently in invasive isolates [7], we further examined its frequency by colonization site. Among invasive

Table 1: Number of group B streptococcal isolates (n = 529) screened via dot blot hybridization and characteristics of each collection.*

| GBS Collection | Isolation source | Culture date | Age range | Race/ethnicity | Number of strains |
|----------------|------------------|--------------|-----------|----------------|------------------|
| 1a. Sexually active college women with UTI receiving care from a Student Health Services at the University of Michigan (UM) [22]. | urine, anal orifice, vaginal | Sept. 1996 to April 1999 | 18–30 | White (76%), Non-White (24%) | Colonizing (n = 102), Invasive† (n = 2) |
| 1b. Most recent male sex partner of women with UTI receiving care from the Student Health Services at UM [22]. | urine, anal orifice | Sept. 1996 to April 1999 | 18–35 | White (71%), Non-White (29%) | Colonizing (n = 43), Invasive† (n = 0) |
| 2a. Sexually active college women without UTI presenting to the Student Health Services at UM [22]. | urine, anal orifice, vaginal | Sept. 1996 to April 1999 | 18–28 | White (80%), Non-White (20%) | Colonizing (n = 57), Invasive† (n = 0) |
| 2b. Most recent male sex partner of women without UTI presenting to the Student Health Services at UM [22]. | urine, anal orifice | Sept. 1996 to April 1999 | 19–33 | White (73%), Non-White (27%) | Colonizing (n = 35), Invasive† (n = 0) |
| 3. Newborns with early onset disease from hospitals affiliated with Baylor College of Medicine [19]. | blood, CSF | 1993 to 2000 | < 7 days | Hispanic (56%), African American (24%), Caucasian (16%), Asian (4%) | Colonizing (n = 0), Invasive† (n = 100) |
| 4a. Random sample of college aged women from the UM community [21] | urine, anal orifice, vaginal | Sept. to Nov. 1998 | 17–49 | Caucasian (65%), Asian (16%), African American (10%), Hispanic (5%), Other (5%) | Colonizing (n = 29), Invasive† (n = 1) |
| 4b. Random sample of college aged men from the UM community [21] | urine, anal orifice | Sept. to Nov. 1998 | 19–45 | Caucasian (60%), Asian (28%), African American (4%), Hispanic (3%), Other (4%) | Colonizing (n = 23), Invasive† (n = 1) |
| 5. Pregnant women presenting at the UM Medical Center for prenatal care [34]. | urine, rectal, vaginal, placental | Aug. 1999 to Mar. 2000 | 16–42 | Caucasian (67%), African American (18%), Other (7%), Unknown (9%) | Colonizing (n = 49), Invasive† (n = 53), Unknown (n = 29) |

* Seventeen individuals (7 from collection 1a, 4 from collection 2a, 3 from collection 2b, and 3 from collection 4b) were colonized with two genetically distinct strains, as determined by pulsed-field gel electrophoresis.
†Invasive isolates originated from the blood or cerebrospinal fluid of newborns <7 days of age, the urine of adults at ≥100,000 cfu/ml, pregnant women presenting for prenatal care associated with GBS isolation from the urine, and the placenta following delivery. Colonizing GBS were from the anal orifice, lower vagina, cervix or urine of healthy individuals, and sexually active college women with a urinary tract infection not caused by GBS.

Table 2: PCR primers used to amplify DNA regions specific to the genes encoding the alpha (bca) and beta (bac) C proteins, and the protein Rib (rib).*

| Gene | Forward primer | Reverse primer | Reference | Size | Annealing temperature | Extension time |
|------|----------------|----------------|-----------|------|-----------------------|----------------|
| bca  | 5'-TAACAGTTATGATACCTTCAGAC-3' | 5'-ACGACTTTCTTCCGCTCACTAGG-3' | [11] | 535 bp | 68°C | 33 sec |
| bac  | 5'-CTTTTTTGTGATTGACATGCAA-3' | 5'-GTCGTTACTTCCGAGATGTAAC-3' | [12] | 592 bp | 60°C | 36 sec |
| rib  | 5'-CAGGAAGTGCTGTTACGTAAAAC-3' | 5'-CGTCCCATTTAGGTTCC-3' | [9] | 369 bp | 58°C | 22 sec |

* PCR conditions for each reaction included a five minute denaturation step at 95°C, and 30 cycles of the following: 35 second denaturation, 40 second annealing and varying extension times. The extension temperature was 73°C for all reactions.
isolates from newborns, the odds of isolation from the cerebrospinal fluid (CSF) compared to blood was 3.6 higher when rib [95% CI: (0.86, 15.44), p = .04], and 4.1 times higher when bac [95% CI: (0.92, 17.91), p = .03] were present. There was no association with bca. Because rib occurred in 92% of capsular serotype III isolates and was found infrequently in other serotypes, and type III occurred more frequently among infants with invasive disease, it is likely that the association with CSF is attributable to confounding. When we examined the colonization site by serotype among rib positive strains, 26% of serotype III and no serotype II strains (the only other serotype with rib) were isolated from the CSF. By contrast, among isolates without rib, 6%, 10% and 17% of serotype Ia, II and Ib strains, respectively, were isolated from the CSF. In a similar analysis of among bac positive strains, 50% of serotype III and 18% of serotype Ib, but no serotype II or V strains were isolated from the CSF. When the analysis was restricted to serotype III strains, rib was not associated with CSF isolation, but bac was (OR: 4.7, 95% CI: 0.43, 60.74), although the sample size was too small to achieve statistical significance (p = .12).

The bca and bac genes frequently occurred together; 74 strains contained both genes among 324 strains with at least one gene (p < .0001). rib was significantly less likely to occur with either bca (11/333, p < .0001) or bac (14/224, p = .01). These relationships were similar when stratified by isolate type with a few exceptions. Among the 14 strains with rib and bac, 8 (57%) were invasive (p = .002); 7 of these 8 were from newborns and 6 of the 7 newborn strains were serotype III. Strains with both bca and rib together were more frequent in colonizing versus invasive strains (p = .03) as were strains with both bca and bac (p = .10).

Discussion

Based on the suggestion that the alpha and beta C proteins and protein Rib protein serve as potential vaccine GBS candidates either in glycoconjugates or alone [6-8], it was estimated in 1988, before the emergence of serotype V GBS, that a vaccine containing the alpha C protein and a serotype III component would prevent at least 90% of GBS cases [30]. Although we did not find these three genes significantly more frequently in invasive versus colonizing GBS strains, bac was found more frequently among isolates from CSF than blood in invasive serotype

different populations.

Table 3: The frequency of genes encoding the alpha (bca) and beta (bac) C protein, and the protein Rib (rib) among various GBS populations.

| GBS Collection | alpha antigen (bca) ‡ | beta antigen (bac) ‡ | Rib protein (rib) |
|----------------|-----------------------|----------------------|-----------------|
|                | Number screened | n (%) | Number screened | n (%) | Number screened | n (%) |
| 1. Sexually active college women with UTI and sex partner† | 145 63 (43) | | 147 26 (18) | | 146 33 (23) | |
| 2. Sexually active college women without UTI and their sex partner | 93 58 (62) | | 93 26 (28) | | 93 17 (18) | |
| 3. Infected newborns < 7 days of age | 100 29 (29) | | 100 20 (20) | | 100 28 (28) | |
| 4. Random sample of college students† | 53 25 (47) | | 53 18 (34) | | 53 13 (25) | |
| 5. Pregnant women | 135 41 (30) | | 131 18 (14) | | 134 27 (20) | |
| Total | 526 216 (41) | | 524 108 (21) | | 526 118 (22) | |

Note: n (%) represents the number of participants with the respective gene in each population; the number screened varies slightly by gene and population because a result could not be obtained for 4 strains tested for rib and bca, and 6 strains tested for bac despite repeated testing.
†There was no difference in gene frequency by gender so the results were combined for presentation.
‡Using the Chi-square test, a significant difference at the p = .05 significance level was observed between bca frequencies in collection 2 versus 3 (p < .0001), 5 (p < .0001) and 1 (p = .004); and between bac frequencies in collection 2 versus 5 (p = .008).
III isolates from newborns, suggesting it may increase disease severity.

Although we detected differences in the frequency of specific genes, it is possible that the encoded proteins are differentially expressed [31,32] and thus, differences in pathogenicity could be attributable to differences in gene expression. A prior study demonstrated that protein Rib [7] was present in more invasive versus colonizing serotype III strains. In this study, invasive strains were more likely to have rib (p = .09), but the association was only marginally significant. A similar observation was found for bac (p = .06), which is consistent with a prior report [18]. However, we cannot exclude the possibility that the differences in collection date and geographic location are responsible for this result. Further, and possibly more important, the isolates assessed may contain other unknown virulence characteristics important to invasion, as the virulence of GBS is probably attributable to multiple genes. Our collections of invasive isolates were limited to those from newborns, pregnant women and healthy young women. It is possible that these virulence genes might have different impacts in other susceptible populations, such as the elderly or those with underlying chronic disease.

**Conclusion**

We observed only a marginally significant difference in bac, bca and rib frequency between invasive and colonizing serotype III strains, thereby raising the possibility that other genes explain the association of serotype III with invasive disease. It is noteworthy, however, that both rib and bac were found more frequently in the newborn serotype III isolates, while bca was found less frequently. Because various genotyping methods, such as multilocus sequence typing (MLST), have distinguished between colonizing and invasive strains, [33] this warrants further study. Using the framework provided by MLST, for example, may allow us to assess the distribution of these genes by sequence types found to be associated with invasiveness. In addition, it is clear that GBS disease pathogenesis is complex, thus novel virulence genes need to be identified and evaluated to understand their role in the pathogenic process, and provide additional vaccine targets. Recently published GBS DNA sequences [34-36] will facilitate the identification of these novel factors.

**Competing interests**

The author(s) declare that they have no competing interests.

**Authors’ contributions**

SDM conducted the data analysis and drafted the manuscript, MK performed the PCR on strains yielding an equivocal result; CFM and BF oversaw and participated in the study design, analysis and writing; KJK and SMB performed the dot blot assays; and CJB provided strains, performed serotyping and assisted with the manuscript. All authors read and approved the final manuscript.

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References

1. Schrag SJ, Zell ER, Lynfield R, Roome A, Arnold KE, Craig AS, Harrison LH, Reingold A, Stefonek K, Smith G, Gamble M, Schuchat A: A population-based comparison of strategies to prevent early-onset group B streptococcal disease in neonates. N Engl J Med 2002, 347:233-239.

2. Harrison LH, Elliott JA, Dwyer DM, Libonati JP, Ferrieri P, Billmann L, Schuchat A: Serotype distribution of invasive group B streptococcal isolates in Maryland: implications for vaccine formulation. Maryland Emerging Infections Program. J Infect Dis 1998, 177:998-1002.

3. Davies HD, Raj S, Adair C, Robinson J, McGeer A: Population-based active surveillance for neonatal group B streptococcal infections in Alberta, Canada: implications for vaccine formulation. Pediatr Infect Dis J 2001, 20:879-884.

4. Baker CJ, Edwards MS: Group B streptococcal conjugate vaccines. Arch Dis Child 2003, 88:375-378.

5. Manning SD: Molecular epidemiology of Streptococcus agalactiae (group B streptococci). Front Pediatr 2006, 851-18.

6. Baker CJ: Immunization to prevent group B streptococcal disease: victories and vexations. J Infect Dis 1999, 177:1593-1603.

7. Stalhammar-Carlemalm M, Stenberg L, Lindahl G: Protein rib: a novel group B streptococcal cell surface protein that confers protective immunity. J Exp Med 1997, 180:2491-2496.

8. Gravekamp C, Kasper DL, Paolletti LC, Madoff LC: Alpha C protein as a carrier for type III capsular polysaccharide and as a protective protein in group B streptococcal vaccines. Infect Immun 1999, 67:2481-2486.

9. Wastfelt M, Stalhammar-Carlemalm M, Stenberg L, Lindahl G: Identification of a family of streptococcal surface protein with extremely repetitive structure. J Biol Chem 1996, 271:18892-18897.

10. Bevanger L, Maeland JA: Complete and incomplete lbc protein fraction in group B streptococci. Acta Pathol Microbiol Scand [B] 1979, 87B:51-54.

11. Michel JL, Madoff LC, Olson K, Kling DE, Kasper DL, Ausubel FM: Large, identical, tandem repeating units in the C protein alpha C protein antigen gene, bca, of group B streptococci. Proc Natl Acad Sci U S A 1992, 89:10060-10064.

12. Jerlstrom PG, Chhatwal GS, Timmis KN: The IgA-binding beta antigen of the C protein complex of Group B streptococci: sequence determination of its gene and detection of two binding regions. Mol Microbiol 1991, 5:843-849.

13. Lachnauer CS, Baker CJ, Baron MJ, Kasper DL, Gravekamp C, Madoff LC: Quantitative determination of immunoglobulin G specific for group B streptococcal beta C protein in human maternal serum. J Infect Dis 2002, 185:236-374.

14. Johnson DR, Ferrieri P: Group B streptococcal lbc protein antigen: distribution of two determinants in wild-type strains of common serotypes. J Clin Microbiol 1984, 19:506-510.

15. Madoff LC, Hori S, Michel JL, Baker CJ, Kasper DL: Phenotypic diversity in the alpha C protein of group B streptococci. Infect Immun 1995, 63:2638-2644.

16. Hickman ME, Rench MA, Ferrieri P, Baker CJ: Changing epidemiology of group B streptococcal colonization. Pediatrics 1999, 104:203-209.

17. Maeland JA, Brakstad OG, Bevanger L, Krokstad S: Distribution and expression of bca, the gene encoding the C alpha C protein, by Streptococcus agalactiae. J Med Microbiol 2000, 49:193-198.

18. James H, Bohnsack JF, Takahashi S, Ouellette C, Poyart C, Trier-Cuyp K, Kunst F, Serotype P. Rusniok C, Crook DW, Harding RM, Bisharat N, Spratt BG: Multilocus sequence typing system for group B streptococcus. J Clin Microbiol 2003, 41:2530-2536.

19. Glaser P, Rusniok C, Buchrieser C, Chevalier F, Frangeul L, Msadek T, Zouine M, Couve E, Lalouel J, Poyart C, Trier-Cuyp K, Kunst F: Genome sequence of Streptococcus agalactiae, a pathogen causing invasive neonatal disease. Mol Microbiol 2002, 45:1499-1513.

20. Tettelin H, Masiyane V, Cieslewicz MJ, Eisen JA, Peterson S, Wessels MR, Paulsen IT, Nelson KE, Margerit I, Read TD, Madoff LC, Wolf AM, Beanan MJ, Brinkac LM, Daugherty SC, DeBoy RT, Durkin AS, Kolo-
Streptococcus agalactiae: implications for the microbial "pan-genome". Proc Natl Acad Sci U S A 2005, 102:13950-13955.

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