Why does a mitochondrion need its individual cristae to be functionally autonomous?

Marc Liesa

Department of Medicine, Endocrinology, David Geffen School of Medicine at UCLA, Los Angeles, CA, USA; Molecular Biology Institute at UCLA, Los Angeles, CA, USA

ABSTRACT

We demonstrated that individual cristae in one mitochondrion function as autonomous electrochemical units, with this autonomy being maintained by cristae junction modulators. Here, I will summarize our novel findings, discuss the advantages of cristae insulation and reconcile this newly characterized cristae autonomy with previous studies showing coordinated cristae function.

The difference in electric potential generated by the proton gradient across the mitochondrial inner membrane (ΔΨm) is used to transform the energy released from nutrient catabolism to ATP. However, ΔΨm is also required for influx/efflux of metabolites feeding anabolic reactions and activating stress-responses, as well as for the import of nuclear-encoded mitochondrial proteins. The dependency on ΔΨm shared by these processes and the organization of the inner membrane (IM) in cristae raise these questions:

(i) How does a single mitochondrion organize the use of the energy contained in the ΔΨm to fuel and/or prioritize different competing processes?

(ii) Why does the abrogation of cristae junction (CJ) modulators decreases ΔΨm and impairs electron transport chain (ETC) function?

ΔΨm was demonstrated to be continuous along the IM, as photodamaging a fraction of one mitochondrion causes concurrent ΔΨm dissipation in proximal and distal areas from the site of photodamage. Consequently, it seemed unlikely that cristae formation allowed compartmentalizing the IM to regions with distinct ΔΨm. In contrast, topological studies of the ETC components and ATP synthase challenged the absence of ΔΨm heterogeneity along the IM. ETC complexes are located in the lateral surfaces of cristae and ATP synthase is located at cristae edges. Furthermore, the inner boundary membrane (IBM) is the preferred location of protein import (Figure 1). With this heterogeneous distribution of ΔΨm-generating and consuming processes, it was possible that ΔΨm could be different in distinct areas of the IM.

By using super-resolution microscopy and four mitochondrial dyes, Nonyl Acridine Orange (NAO), Tetramethylrhodamine ethyl ester (TMRE), Mitotracker-green (MTG) and Rhodamine-123, we visualized individual cristae in respiring cells. These dyes require ΔΨm to accumulate in mitochondria. However, NAO and MTG do not properly report on ΔΨm, as when they reach mitochondria, they stay bound in a ΔΨm-independent manner. Only Rhodamine-123 and TMRE report on ΔΨm real-time. We confirmed that TMRE reported ΔΨm in cristae, as cristae TMRE was dissipated by photodamage, mitochondrial uncoupling, and spontaneous depolarization. TMRE dissipation occurred without changes of cristae MTG signal, further proving ΔΨm-sensitivity of cristae decoration by TMRE. Then, why diffusible TMRE preferentially labeled cristae? We concluded that this preference was reflecting a close juxtaposition of TMRE to the matrix-side of the IM, caused by the highest ΔΨm values expected in closer apposition to the IM.

A challenge using TMRE to compare ΔΨm in cristae versus the IBM is that cristae membrane composition is different. Therefore, a higher fraction of TMRE molecules could be decorating membranes in a non-Nernstian manner (binding) in cristae, when compared to the IBM. To address this, we measured the response of the difference in TMRE fluorescence between the IBM and cristae to compounds acutely changing ΔΨm, such as the ATP synthase inhibitor oligomycin. If elevated TMRE signal in cristae was exclusively caused by increased binding to membranes, acute treatment with oligomycin should not change the difference in TMRE signal between cristae and IBM. Supporting this preference was reflecting a close juxtaposition of TMRE to the matrix-side of the IM, caused by the highest ΔΨm values expected in closer apposition to the IM.

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CONTACT Marc Liesa; mliesa@mednet.ucla.edu; Center for Health Sciences, 650 Charles E. Young Dr., CHS 27-200F, Los Angeles, CA 90095, USA

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cristae and IBM. Furthermore, the abrogation of mitochondrial dynamics-dependent quality control, through co-deletion of Opa1 & Dynamin 1 like (Dnm1l, aka Drp1), induced the formation of cristae vesicles detached from the IBM.\(^5\) These results demonstrated that cristae can become even physically independent from the IBM. We speculate that the formation of cristae vesicles could be revealing an uncharacterized recycling process of mitochondrial components controlled by CJ modulators. A defect in this recycling process, and thus in mitochondrial quality control, could help explaining decreased ETC activity induced by the abrogation of CJ modulators and cristae structures.

How cristae autonomy is reconciled with \(\Delta \Psi_m\) continuity along the IM?\(^2\) Studies demonstrating concurrent \(\Delta \Psi_m\) depolarization in proximal and distal areas from the photodamage site had a low temporal resolution, obtaining the first \(\Delta \Psi_m\) measurements 5 s after photodamage.\(^1\) Thus, it was possible that signaling events leading to depolarization could be completed in less than 5 s. To address this, we measured \(\Delta \Psi_m\) every 0.15 s after photodamage and found that \(\Delta \Psi_m\) depolarization showed a wave-like pattern, starting at the site of photodamage and spreading to distal areas.\(^5\) This wave-pattern supports that photodamage generates a diffusible and/or self-propagating signal that induces distal depolarization. We speculate that this signal could be reactive oxygen species (ROS) and/or self-propagating lipid peroxides (Figure 1). The existence of such a signal would allow sensing unsolvable damage and communicate the need to eliminate the injured mitochondrion, as depolarization tags mitochondria for removal by mitophagy.\(^10\) If such damage occurred in a long mitochondrion, fragmentation into smaller mitochondria would allow to segregate damaged areas and selectively tag them for mitophagy. Undamaged areas would form units that could recover \(\Delta \Psi_m\) and escape from mitophagy. Consequently, the connection of \(\Delta \Psi_m\) cristae by diffusible signals inducing depolarization is explained by i) the need to remove an entire mitochondrion by mitophagy to ensure quality control and ii) depolarization being the signal selected to eliminate mitochondria by mitophagy.

Cristae autonomy could also allow compartmentalizing the energy within \(\Delta \Psi_m\) in the IM, to avoid competition for \(\Delta \Psi_m\) consumption of concurrent mitochondrial processes. IBM insulation permits dedicating \(\Delta \Psi_m\) in IBM mostly to support protein import. Then, the electrical insulation of one crista from the IBM and from other cristae would let one crista to retain the hyperpolarization needed to produce ROS for signaling, while other cristae could concurrently consume \(\Delta \Psi_m\) to fuel ATP production (Figure 1).

We believe our work has provided important evidence explaining why cristae morphology and junctions are key determinants of mitochondrial function in health and disease.

**ORCID**

Marc Liesa  [http://orcid.org/0000-0002-5909-8570](http://orcid.org/0000-0002-5909-8570)

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