Three new freshwater species of the genus *Achnanthidium* (Bacillariophyta, Achnanthidiaceae) from Taiping Lake, China

Pan Yu¹, Qingmin You¹, J. Patrick Kociolek²,³ & Quanxi Wang¹*

¹ College of Life Sciences, Shanghai Normal University, Shanghai 200234, P. R. China; *Corresponding author e–mail: wangqx@shnu.edu.cn
² Museum of Natural History, University of Colorado, Boulder, CO 80309, USA
³ School of Life Science, Shanxi University, Taiyuan, P. R. China

**Abstract:** We describe three new *Achnanthidium* species, *A. lacustre* sp. nov., *A. sublanceolatum* sp. nov., and *A. taipingensis* sp. nov., from Taiping Lake, Anhui Province (China) based on light and scanning electron microscopy. *A. lacustre* sp. nov. belongs to the “*A. minutissimum* complex” of the genus, based on it having straight external distal raphe fissures and round to elliptical areolae. Both *A. sublanceolatum* sp. nov., and *A. taipingensis* sp. nov. belong to the “*A. pyrenaicum* complex” of the genus, based on them having transpically–elongated areolae and deflected external distal raphe fissures. All three species are sufficiently different from other similar species based on valve outline, shape of the axial and center areas, and striae density. These three new species are all observed in benthic collections from Taiping Lake.

**Key words:** *Achnanthidium*, diatom, morphology, new species, Taiping Lake, China

**INTRODUCTION**

The genus *Achnanthidium* is known from freshwaters, and species of this genus are widely distributed in various types freshwater habitats and can be common and abundant in those habitats (Karthick et al. 2017; Liu et al. 2016; Pinseel et al. 2015; Novais et al. 2011; Potapova & Ponader 2004; Kobayasi et al. 2006). *Achnanthidium* species can occur across a broad range of trophic conditions, from oligotrophic to eutrophic waters (Karthick et al. 2017; Ponader & Potapova 2007). This genus was initially described by Kützing (1844) as a subgenus of *Achnanthes*, and the species *Achnanthes microcephalum* Kützing was the type species (Pérès et al. 2014; Kingston 2003). Later, *Achnanthidium* was elevated to the level of genus by Round & Potapova (1996). Because the size of many *Achnanthidium* is small, and there may be few identification characteristics evident as viewed with the light microscopy (LM), and the variability of diagnostic features often overlap, this genus may be challenging to study in terms of species recognition (Karthick et al. 2017; Marquardt et al. 2017; Ponader & Potapova 2007).

The genus *Achnanthidium* has been subdivided into three subgroups, the “*Achnanthidium minutissimum* complex”, “*A. pyrenaicum* complex” and the “*A. exiguum* complex” (Karthick et al. 2017; Pérès et al. 2014; Wojtal et al. 2011; Wojtal et al. 2010). The *A. minutissimum* complex has straight external distal raphe ends and, usually, linear to linear–lanceolate valve shapes, striae density increasing towards the apices and round external areolar openings (Pinseel et al. 2015; Compère & Van De Vijver 2011). Members of *A. pyrenaicum* complex have external distal raphe ends that are deflected or hooked to one side of the valve and slit–like areolar openings (Pinseel et al. 2015; Jüttner et al. 2011; Round & Bukhtiyarova 1996). Members of the *A. exiguum* complex have external distal raphe ends curved in opposite directions.

Presently, the number of species of the genus *Achnanthidium* is greater than 200 (Marquardt et al. 2017; Kociolek et al. 2018). Only a limited number of new *Achnanthidium* species have been described from China (e.g. Liu et al. 2016; Yu et al. 2018, accepted). During a survey of the freshwater diatoms from Taiping Lake, in the framework of water quality monitoring, three unknown *Achnanthidium* species were encountered. The purpose of this present is to document and formally describe those species with light microscopy (LM) and scanning electron microscopy (SEM), and to compare the new species with morphologically similar taxa.
**Material and Methods**

For this study we used samples that were collected from Taiping Lake (30°14’–30°28’N, 117°55’–118°12’E), located in the District of Huangshan Mountain, Southern Anhui Province, China, in May 2018. Taiping Lake is the largest artificial reservoir lake in Anhui province. In the field, several water chemistry characteristics were recorded, including: pH, temperature, dissolved oxygen, salinity, total dissolved solids (TDS), and conductivity. These were all measured using a YSIPro Plus multiparameter meter (YSI, Ohio, USA). Diatom samples were collected from natural substrates, including stones, or from navigation buoys, by clean toothbrushes, and the samples were placed in a bottle and preserved with formalin (4% final concentration).

In the laboratory, diatom samples were cleaned with concentrated nitric acid using the Microwave Accelerated Reaction System (Model MARS, CEM Corporation, Charlotte, USA) (Parr et al. 2004), with a pre–programmed digestion scheme (temperature, 180°C) (You et al. 2015; Yu et al. 2017). Next, samples were alternately centrifuged for 5 min at 3500 rpm (TDZS–WS, Luyi Corporation, Shanghai, China) and washed six times using distilled water until the pH of the sample was close to neutral. Finally, the cleaned samples were kept in 95% ethanol. Cleaned diatom frustules were made mounted in Naphrax for light microscopy (LM) or air–dried onto cover slips and mounted onto alloy stubs for observation with the scanning electron microscope (SEM). LM studies were made with an Olympus BX–53 microscope fitted with DIC optics and a 100× oil immersion objectives (1.4 numerical aperture). SEM examination was made using a Hitachi SU8010 (1–2 kV, WD less than 6 mm) (Tokyo, Japan). Images were compiled with Adobe Photoshop CS6. Morphological terminology follows Round et al. (1990). All of the diatom samples and permanent slides are housed in the Biology Department Diatom Herbarium, Shanghai Normal University (SHTU). Isotype material and slides has been deposited in the Kociolek Collection at the University of Colorado, Boulder (COLO).

**Results**

*Achnanthidium lacustre* P. Yu, Q.–M. You et Kociolek sp. nov. (Figs 1–20, 90–105)

**Description:** In LM (Figs 1–20), frustules are bent in girdle view with a concave raphe valve and a convex raphelose valve. Frustules are heterovalvar, monoraphid. Valves are irregularly linear–lanceolate in shape, with bluntly rounded apices. Valve length 20–23 µm, breadth 3.0–3.5 µm (n=200). Both valves possess a linear axial area which widens slightly towards the central area. On the raphe valve, central area is a small oval in shape forming an elliptical fascia. Striae are uniseriate, comprised of 2–3 round to transapically elongate areolae in the middle part of the valve (Figs 98, 102), and 1–3 irregularly round or transapically elongate areolae at the ends (Fig 100). Some striae have one slit–like areola on the valve mantle (Figs 92–93). Areolar occlusions are positioned within the opening and can be seen from the exterior (Fig 96). Internally, the raphe terminates distally as elevated helictoglossae (Figs 91, 94), and the proximal raphe endings are short, deflected in opposite directions (Figs 91, 95). Areolae are large round to oblong in shape and the openings are occluded with fine hymenate structures that include small openings around the periphery (Fig 97).

On the external of the raphelose valve, the axial area is linear, widening slightly towards the central area (Fig 98). Striae are uniseriate, comprised of 2–3 round to transapically elongate areolae in the middle part of the valve (Figs 98, 102), and 1–3 irregularly round or transapically elongate areolae at the ends (Fig 100). Some striae have one slit–like areola on the valve mantle (Figs 100, 102). Areolae are occluded with fine hymenate structure that can be seen externally (Fig 104). Internally, areolae are round to transapically–elongate, occluded on valve face and mantle, and the occlusions are comprised of a fine hymenate structure (Figs 99, 101, 103, 105).

**Holotype:** SHTU!, slide TPH–1805009!, holotype illustrated in Figs 2, 11.

**Isotypes:** COLO! slide 628097, Kociolek Collection, University of Colorado, Museum of Natural History Diatom Herbarium, Boulder, USA.

**Type locality:** CHINA. Taiping Lake, Anhui Province, 30°23'07"N, 118°02'26"E, altitude: 210 m, collected by Q.X. Wang et al., 21th May 2018.

**Etymology:** The species is named for it being found in a lake.

**Ecology:** Collected in two samples (TPH–20180509; TPH–201805011) on stones and navigation buoys. pH 8.5, water temperature 25.5 °C, Salinity 0.05‰, TDS 69.6 mg.l⁻¹, Conductivity 108 µs.cm⁻¹).

**Distribution:** So far, the new species is known only from the two sampling localities.

*Achnanthidium sublanceolatum* P. Yu, Q.–M. You et Kociolek sp. nov. (Figs 21–55, 106–123)

**Description:** In LM (Figs 21–55), Frustules are heterovalvar, monoraphid. Valves are linear–lanceolate in shape, with rounded or weakly protracted apices. Valve length 18–35 µm, breadth 4.0–4.5 µm (n=300). The raphe valve is concave, and the axial area is narrow,
linear–lanceolate, widening slightly towards the central area. Striae nearly parallel, 20–23 in 10 µm at the middle portion, 36–42 in 10 µm near the apices, on the raphe valve. The rapheless valve is convex, with narrow linear–lanceolate axial area weakly expanded at the middle portion of the valve. Striae are nearly parallel, 21–24 in 10 µm in the center, and 30–36 in 10 µm near the apices. Areolae are not visible in LM.

In the SEM, both valves have a narrow hyaline area at the valve face: mantle junction (Figs 106, 115). Raphe valve: Externally, the raphe is filiform and straight (Fig 106), distal raphe ends are deflected to the same side at an angle of nearly 90°, and the proximal raphe ends are straight and teardrop-shaped (Figs 106, 108, 110). Striae are uniseriate, containing 3–4 transapically–oriented areolae in the middle portion of the valve, and 1–2 long narrow areolae at the apices (Figs 108, 110, 112–113). Valve mantle with a single row of linear areolae extended around the apices with small interruption at the ends (Fig. 108). Internally, the raphe terminates in raised helictoglossae close to apices (Figs 107, 109), and proximal raphe ends are weakly deflected in opposite directions (Figs 107, 111). Areolae are transapically elongate in the central portion of the valve and becoming larger and oblong at the apices (Figs 109, 111). Areolar openings have fine, complex hymenate occlusions (Fig 114).

Rapheless valve: Externally, the axial area is linear–lanceolate, being weakly expanded in the central area and the apices (Figs 115, 117–118, 122). Striae are uniseriate, composed of 2–5 narrow and linear areolae in the central area (Fig. 122), and 1–2 long narrow areolae at the apices (Fig. 117–118). Valve mantle with a single row of linear areolae extended around apices with small interruption at the ends (Fig. 108). Internally, the raphe terminates in raised helictoglossae close to apices (Figs 107, 109), and proximal raphe ends are weakly deflected in opposite directions (Figs 107, 111). Areolae are transapically elongate in the central portion of the valve and becoming larger and oblong at the apices (Figs 109, 111). Areolar openings have fine, complex hymenate occlusions (Fig 114).

Areolae at the apices (Fig. 117–118). Valve mantle with a single row of linear areolae extended around apices with small interruption at the ends (Fig. 108). Internally, the raphe terminates in raised helictoglossae close to apices (Figs 107, 109), and proximal raphe ends are weakly deflected in opposite directions (Figs 107, 111). Areolae are transapically elongate in the central portion of the valve and becoming larger and oblong at the apices (Figs 109, 111). Areolar openings have fine, complex hymenate occlusions (Fig 114).

Rapheless valve: Externally, the axial area is linear–lanceolate, being weakly expanded in the central area and the apices (Figs 115, 117–118, 122). Striae are uniseriate, containing 3–4 transapically–oriented areolae in the middle portion of the valve, and 1–2 long narrow areolae at the apices (Figs 110, 112–113). Valve mantle with a single row of linear areolae extended around the apices with small interruption at the ends (Fig. 108). Internally, the raphe terminates in raised helictoglossae close to apices (Figs 107, 109), and proximal raphe ends are weakly deflected in opposite directions (Figs 107, 111). Areolae are transapically elongate in the central portion of the valve and becoming larger and oblong at the apices (Figs 109, 111). Areolar openings have fine, complex hymenate occlusions (Fig 114).

Areolae at the apices (Fig. 117–118). Valve mantle with a single row of linear areolae extended around apices with small interruption at the ends (Fig. 108). Internally, the raphe terminates in raised helictoglossae close to apices (Figs 107, 109), and proximal raphe ends are weakly deflected in opposite directions (Figs 107, 111). Areolae are transapically elongate in the central portion of the valve and becoming larger and oblong at the apices (Figs 109, 111). Areolar openings have fine, complex hymenate occlusions (Fig 114).

Description: In LM (Figs 56–89), frustules are heterovalvar, monoraphid. The raphe valve is concave and the rapheless valve is convex. Valves linear to linear–elliptic in shape, with broadly rounded ends. Valve length 12–24 µm, breadth 3.5–4.0 µm (n=300). Raphe valve with a narrow, linear–lanceolate axial area which is weakly expanded at the middle portion of the valve. Striae are nearly parallel, becoming denser towards apices, 21–25 in 10 µm at center, 28–32 in 10 µm near the apices. Rapheless valve with narrow, linear axial area which is weakly expanded at the middle portion of the valve. Striae density is 20–24 in 10 µm at the center, up to 26–30 in 10 µm at the apices.

In the SEM, the frustule is slightly bent ventrally in girdle view (Fig.139). On the both valves, the valve mantle has a single row of linear areolae (Figs 139, 141), but there is a small depression on one side of the raphe (Figs 126–127) that is absent at the apices of rapheless valveS (Figs 142–143). Externally, the raphe is filiform, straight (Fig 124), and has distal raphe ends deflected to the same side. On the side of the deflection there is a depression near the distal raphe ends (Figs 126–127). The proximal raphe ends are straight and teardrop-shaped (Figs 128–130). Areolae are small, round to transapically–oriented, the uniserate striae are composed of 1–5, usually 4–5, areolae in the middle portion of the valve (Figs 128–130), and 1–4 areolae at the apex (Figs 126–127). Internally, distal raphe ends terminate in raised helictoglossae (Figs 131, 133), while the proximal raphe ends are very weakly deflected in opposite directions (Figs 131, 135).

On the exterior of the rapheless valve, the areolae are small, transapically–oriented, and the uniserate striae contain 2–6, usually 4–5, areolae at the center (Figs 145–146), and 1–3 areolae at the apices (Figs 142–143). On the both interior of both valves, areolae are occluded by hymenes perforated by delicate slits, and each hymene joins with the adjacent hymene (Fig. 137–38).

Ecology: Collected in two samples (TPH–201805008; TPH–201805011) on navigation mark. pH 8.5, water temperature 25.5 °C, Salinity 0.05‰, TDS 69.6 mg.l⁻¹, Conductivity 108 µs.cm⁻¹).

Distribution: So far, the new species is known only from the two sample localities.
Table 1. Comparison of morphological characteristics of *Achnanthidium lacustre* sp. nov. and closely related taxa.

| Species/Feature       | *A. lacustre* | *A. ennediense* | *A. standeri* | *A. taiaense* | *A. exile* | *A. affine* |
|-----------------------|---------------|-----------------|---------------|---------------|------------|-------------|
| Valve length (μm)     | 20–23         | 15.6–26.5       | 7–38          | 10–35         | 21–33      | 11.0–25.8   |
| Valve width (μm)      | 3.0–3.5       | 2.3–3.3         | 2.8–4.4       | 3–5           | 4–6        | 2.6–3.7     |
| Valve outline         | Irregularly linear–lanseolate | Lanceolate to rhombic–lanceolate | Linear to linear–lanceolate | Lanceolate | Rhombic–lanceolate to narrowly lanceolate | Lanceolate |
| Valve apices          | Bluntly rounded | Broadly rounded | Bluntly rounded | Acutely rounded | Acute to bluntly rounded | Broadly rounded |
| **Raphe valve**       |               |                 |               |               |            |             |
| Axial area            | Linear        | Linear          | Narrow lanceolate | Narrow linear | Lanceolate | Narrow linear |
| Central area          | Small oval    | Small, variable | Acute–angled fascia | Acute–angled fascia | Rhombic to elliptical | Wedge–shaped fascia |
| Raphe                 | Straight, distal raphe ends are expended the valve mantle | Distal raphe ends slightly elongated straight | Distal and enlarged proximal raphe endings | Distal ends are droplet-shaped | Straight proximal and distal endings | Straight, distal raphe ends are expended the valve mantle |
| Density of striae (10 μm) | 27–29 (M), 28–32 (A) | 30–32 (M), 38 (A) | 28–30 (M), 30–60 (A) | 36–40 | 25–30 (M), 36 (A) | 27–29 (M), 35–40 (A) |
| Number of areolae per striae | 2–3 (M), 1–3 (A) | 2–4 | 2–3 | 3–4 (M), 1–3 (A) | 3–6 | 3–4 (M), 1–3 (A) |
| **Rapheless valve**   |               |                 |               |               |            |             |
| Axial area            | Linear        | Narrow linear   | Narrow linear | Narrow linear | Narrowly linear–lanceolate | Linear |
| Central area          | Absent        | Absent or very small, apically elongated | Almost absent | Absent | Apically elongated, elliptical to oval | Absent |
| Density of striae (10 μm) | 26–29 (M), 28–30 (A) | 32–34 (M), 38 (A) | 24–26 (M), 30–32 (A) | 40 | 25–30 | 29–32 (M), 37 (A) |
| Number of areolae per striae | 4–5 (M), 1–3 (A) | 3–5 | 4–5 (M), 2–4 (A) | 1–3 | 2–4 | 3–4 |
| References            | Current study | Compère & Van De Veyer (2011) | Taylor et al. (2011) | Taylor et al. (2011) | Krammer & Lange–Bertalot (1991) | Czarnecki (1994) |

Note: “M” means middle; “A” means apices.
Table 2. Comparison of morphological characteristics of *Achnanthidium sublanceolatum* sp. nov. and closely related taxa.

| Species/Feature | *A. sublanceolatum* | *A. linannulum* | *A. sinense* | *A. gracillimum* | *A. chitrakootense* | *A. deflexum* |
|----------------|---------------------|----------------|--------------|----------------|------------------|---------------|
| Valve length (μm) | 18–35               | 15.5–32.5       | 17.5–31.7   | 19–31.5         | 13–42            | 7.4–27.1      |
| Valve width (μm)  | 4–4.5               | 2.5–4.5         | 4.1–6.0     | 3–4             | 3.4–4.2          | 3.5–5.2       |
| Valve outline     | Linear–lanceolate   | Linear elliptical to lanceolate | Narrow lanceolate | Elliptical to lanceolate | Linear to linear–elliptical | Linear to elliptical |
| Valve apices      | Rounded or weakly protracted | Rounded or slightly protracted | Not protracted, acute round | Narrowly rostrate to subcapitate | Subcapitate to rounded | Subrostrate |
| **Raphe valve**   |                      |                |              |                 |                  |               |
| Axial area        | Narrow, linear–lanceolate | Lanceolate     | Narrow lanceolate | Linear          | Linear–lanceolate | Linear         |
| Central area      | Absent              | Indistinct to weakly expanded | Rhombic–lanceolate | Asymmetrical    | Indistinct to weakly expanded | Absent        |
| Raphe             | Distal raphe fissures deflected to the same side | Distal fissures deflected to the same side at an angle of 80°–90° | Distal raphe ends curved to the same side | Distal raphe fissures are sharply bent | Distal raphe ends strongly curved to the same side | Distal raphe fissures are hooked towards the same side |
| Density of striae (10 μm) | 20–23 (M), 34–42 (A) | 24–27 (M), 32–34 (A) | 21–28 (M), 40 (A) | 22 (M), 36 (A) | 26–30 | 15–28 (M), 35–50 (A) |
| Number of areolae per stria | 3–4 (M), 1–2 (A) | 2–3 | 5–6 (M), 1–3 (A) | 4–5 (M), 1–2 (A) | 2–4 (M), 1–3 (A) | 3–5 (M), 1–3 (A) |
| **Rapheless valve** |                      |                |              |                 |                  |               |
| Axial area        | Narrow linear–lanceolate | Narrow lanceolate | Linear       | Linear          | Linear           | Linear         |
| Central area      | Absent              | Weakly expanded to absent | Absent       | Weakly expanded to absent | Absent          | Absent        |
| Density of striae (10 μm) | 21–24 (M), 30–36 (A) | 24–26 (M), 28–30 (A) | 21–27 (M), 34 (A) | 22 (M), 36 (A) | 26–30 | 15–26 (M), 30–35 (A) |
| Number of areolae per stria | 2–5 (M), 1–2 (A) | 2–4 | 6–7 (M), 1–3 (apices) | 4–5 (M), 1–3 (apices) | 4–5 (M), 2–4 (apices) | 3–4 |
| References        | Current study        | LIU et al. (2016) | KOYAYASI et al. (2006) | WOSTAL et al. (2010) | POTAPA V & PONADER (2004) |

Note: “M” means middle; “A” means apices.
Table 3. Comparison of morphological characteristics of *Achnanthidium taipingensis* sp. nov. and closely related taxa.

| Species/Feature | *A. taipingensis* | *A. pseudoconspicuum* var. *yomensis* | *A. convergens* Kobayasi | *A. rivulare* | *A. crassum* | *A. japonica* |
|----------------|------------------|-------------------------------|-----------------|-----------------|---------------|--------------|
| Valve length (μm) | 12.0–24.0 | 17.7–25.2 | 10.0–25.0 | 5.4–21.3 | 7.3–19.6 | 13.0–19.5 |
| Valve width (μm) | 3.5–4.0 | 4.3–5.7 | 4.0–4.5 | 2.6–4.4 | 3.1–4.5 | 3.2–5.0 |
| Valve outline | Linear to linear–elliptic | Linear to elliptical | Linear lanceolate | Linear–elliptical | Elliptic to elliptic–lanceolate | Linear–elliptical |
| Valve apices | Broadly rounded | Round to broadly rounded | Round to subrostrate | Round or protracted | Broadly rounded | Rounded |

**Raphe valve**

| Axial area | Narrowly linear–lanceolate | Narrow and fusiform | Linear–lanceolate | Linear–lanceolate | Narrowly linear–lanceolate | Lanceolate |
| Central area | Absent | Absent | Absent | Absent | Absent | Absent |
| Raphe | Distal raphe fissures hooked towards the same side | Distal raphe fissures bend sharply towards the same side | Distal raphe ends strongly curved to the same side | Distal raphe fissures are hooked towards the same side | Distal raphe ends are unilaterally deflected | Distal raphe ends strongly curved to the same side |
| Density of striae (10 μm) | 21–25 (M), 28–32 (A) | 17–19 (M), 36–40 (A) | 18 (M), 36–40 (A) | 19–25 (M), 50 (A) | 20–24 (M), 36–40 (A) | 16–20 (M), 30 (A) |
| Number of areolae per stria | 4–5 (M), 2–4 (A) | 3–5 (M), 2–3 (A) | 5–6 (M), 2–3 (A) | 2–4 (M), 1–3 (A) | 3–4 (M), 1–3 (A) | 3–4 (M), 1–3 (A) |

**Rapheless valve**

| Axial area | Linear | Linear | Linear–lanceolate | Narrowly linear | Narrowly linear | Linear–lanceolate |
| Central area | Absent | Absent | Absent | Absent | Absent | Absent |
| Density of striae (10 μm) | 20–24 (M), 26–30 (A) | 16–20 (M), 24–28 (A) | 22–28 (M), 28–32 (A) | 19–28 (M), 43 (A) | 20–24 (M), 35–40 (A) | About 18 |
| Number of areolae per stria | 2–6 (M), 1–3 (A) | 4–5 (M), 1–3 (A) | 4–6 (M), 1–3 (A) | 5–6 (M), 1–3 (A) | 4–5 (M), 1–4 (A) | 3–6 (M), 2–4 (A) |
| References | Current study | YANA & MAYAMA (2015) | KOBAYASI et al. (1986) | POTAPOVA & PONADER (2004) | POTAPOVA & PONADER (2004) | KOBAYASI et al. (1986) |

Note: “M” means middle; “A” means apices.
**DISCUSSION**

The three new species possess characters that support their assignment to the genus *Achnanthidium*, including, having small linear–lanceolate to lanceolate elliptic valves, shallow V-shaped valve in girdle view, uniseriate striae, a fine raphe and straight or deflected external distal raphe fissures (Vijver et al. 2011; Ponder & Potapova 2007). *A. lacustre* belongs to the “*A. minutissimum* complex” based on its possession of straight external distal raphe fissures, while *A. sublanceolatum* and *A. taipingensis* belong to the “*Achnanthidium pyrenaicum* complex” based on them having deflected external distal raphe fissures. *A. lacustre* is similar to a few species, including *A. ennediense* (Compère) Compère et Van De Vijver (Compère & Van De Vijver 2011), *A. standeri* Taylor, Morales et Ector (Taylor et al. 2011), *A. taiaense* Taylor, Morales et Ector (Taylor et al. 2011), *A. exile* (Kützing) Heiberg (Krammer & Lange–Bertalot 1991) and *A. affine* (Grunow) Czarnecki (Czarnecki 1994). To facilitate comparison between *A. lacustre* sp. nov. and these similar species their morphological characteristics are summarized in Table 1. In valve shape, *A. lacustre* sp. nov. can easily be separated from other similar species with irregularly linear–lanceolate outlines. *A. ennediense*, for example, has valves with lanceolate to rhombic–lanceolate outlines while the valve of *A. standeri* are linear to linear–lanceolate, *A. taiaense* are lanceolate, *A. exile* are rhombic–lanceolate to narrowly lanceolate, and *A. affine* with a lanceolate valve. *A. lacustre* sp. nov. and *A. standeri* have bluntly–rounded apices, while *A. ennediense* and *A. affine* have valves with broadly rounded ends. *A. taiaense* and *A. exile* bear acutely rounded apices, helping to distinguish them from *A. lacustre* sp. nov. In the raphe valve, the center area is small oval of *A. lacustre* sp. nov. which different other species, *A. ennediense* with a oval to very slightly apically elongated center area, *A. standeri* and *A. taiaense* have a acute–angled fascia in middle portion, *A. exile* with rhombic to elliptical center area, and *A. affine* with a large wedge–shaped fascia in middle portion. In addition, the striae density at the apices of *A. lacustre* sp. nov. is less (28–32/10 µm) at the apices than *A. ennediense* (38/10 µm), *A. standeri* (30–60/10 µm), *A. taiaense* (36–40/10 µm), *A. exile* (36/10 µm), and *A. affine* (35–40/10 µm). Furthermore, the striae density of *A. lacustre* sp. nov. is less (26–29/10 µm at the middle, 28–30/10 µm at the apices) than *A. ennediense* (32–34/10 µm at the middle, 38/10 µm at the apices), *A. standeri* (30–32/10 µm at the apices), *A. taiaense* (40/10 µm throughout the length of the valve) and *A. affine* (29–32/10 µm at the middle, 37/10 µm at the apices), but the striae density of the middle portion of *A. lacustre* sp. nov. is higher than *A. standeri* (24–26/10 µm). On the rapheless valve, *A. lacustre* sp. nov., *A. standeri*, *A. taiaense* and *A. affine* all lack a distinguished central area, but valves of *A. ennediense* have a small apically–elongated central area, and in *A. exile* the valves have a apically–elongated, elliptical to oval central area.

There are five species that might be confused with *A. sublanceolatum* sp. nov., namely *A. linannulum* Karthick, Taylor et Hamilton (Karthick et al. 2017), *A. sinense* Liu et Blanco (Liu et al. 2016), *A. gracillimum* (Meister) Lange–Bertalot (Kobayasi et al. 2006), *A. chitrakootense* Wojtal, Lange–Bertalot et Nautiyal (Wojtal et al. 2010) and *A. deflexum* (Reimer) Kingston (Potapova & Ponder 2004), and we compare them morphologically in Table 2. In LM, valves of *A. sublanceolatum* sp. nov. are wider (4–4.5 µm), as compared to *A. gracillimum* (3–4 µm) and *A. chitrakootense* (3.4–4.2 µm), and narrower than either *A. sinense* (4.1–6.0 µm) or *A. deflexum* (3.5–5.2 µm). There is a difference in valve outline between *A. sublanceolatum* sp. nov. and these other species, with valves of *A. sublanceolatum* sp. nov. being linear–lanceolate with rounded apices. *A. linannulum* possesses linear elliptical to lanceolate valves that have rounded or slightly protracted apices, *A. sinense* have a narrow lanceolate valve and acutely round ends, *A. gracillimum* bears elliptical to lanceolate valve and narrowly rostrate to subcapitate ends, *A. chitrakootense* has valves with linear to linear–elliptical and subcapitate to rounded ends, and *A. deflexum* possess a linear to elliptical valve and a subrostrate apices. On the raphe valve, *A. sublanceolatum* sp. nov. and *A. deflexum* usually have no distinct central area, while the central areas of *A. linannulum* and *A. chitrakootense* are indistinct to weakly expanded. In *A. sinense* the central area is rhombic–lanceolate, while in *A. gracillimum* the central area is asymmetrical. Moreover, the striae density at the apices of *A. sublanceolatum* sp. nov. is less (34–42/10 µm) than *A. sinense* (40/10 µm) and *A. deflexum* (35–50/10 µm), but higher than *A. linannulum* (32–34/10 µm) and *A. chitrakootense* (26–30/10 µm). There are also additional features that distinguish this new species from other similar species (Table 2).

Species similar to *A. taipingensis* sp. nov. are *A. pseudoconspicuum* var. *yomensis* Yana et Mayama (Yana & Mayama 2015), *A. convergens* Kobayasi (Kobayasi et al. 1986), *A. rivulare* Potapova et Ponader (Potapova & Ponder 2004) and *A. crissum* (Hustedt) Potapova et Ponader (Potapova & Ponder 2004), *A. japonica* Kobayasi (Kobayasi et al. 1986). This group of species is compared in Table 3. Externally, on the raphe valve, *A. taipingensis* sp. nov. has a depression near the terminal raphe fissures, a feature which distinguishes it from other, similar species. In addition, the internal proximal raphe fissures of *A. taipingensis* sp. nov. are only very weakly deflected in opposite directions, but in other, similar species the internal proximal raphe ends are distinctly deflected in opposite directions. Additionally, the striae density at the middle of the valves of *A. taipingensis* sp. nov. is higher (21–25/10 µm) than *A. pseudoconspicuum* var. *yomensis* (17–19/10 µm), *A. convergens* (18/10 µm), and *A. japonica* (16–20/10 µm), and the striae density at the ends of *A. taipingensis* sp. nov. is lower (28–32/10 µm) than *A. pseudoconspicuum* var.
yomensis (36–40/10 µm), A. convergens (36–40/10 µm), A. rivulare (50/10 µm), and A. crassum (36–40/10 µm).

On the raphedless valve, the striae density at the meddle of A. taipengensis sp. nov. is higher (20–24/10 µm) than A. pseudoconspicuum var. yomensis (16–20/10 µm), and A. japonica (18/10 µm). Also, the striae density at the apices of A. taipengensis sp. nov. is less (26–30/10 µm) than A. rivulare (43/10 µm), A. crassum (35–40/10 µm), and A. convergens (28–32/10 µm), and higher than A. pseudoconspicuum var. yomensis (24–28/10 µm) and A. japonica (18/10 µm).

This paper represents the first published treatment of diatoms from Taiping Lake, Southern Anhui Province, China. The three new Achnanthidium species were abundant in the samples studied, and there are some other monoraphid species that co-occur with these new species. The co-occurring monoraphid taxa include, Achnanthidium caledonicum Lange–Bertalot, A. pyrenacicum (Hustedt) Kobayasi, A. rivulare Potapova et Ponader and A. duthei (Sreen.) Edlund. We will continue to study the diatom diversity of the Taiping Lake.

Acknowledgements

This research was funded and supported by National Natural Science Foundation of China (No. 31770222), National basic science and technology work (2013FY110400), and Shanghai Engineering Research Foundation of China (No. 31770222), National basic science and technology of North America. Ecology and Classification. – pp. 531, Uchida Rokakuho Publishing Co., Ltd, Tokyo.

Kociolek, J.P.; Balasubramanian, K.; Blanco, S.; Coste, M.; Ector, L.; Liu, Y.; Kulikovskiy, M.; Lundholm, N.; Ludwig, T.; Potapova, M.; Rimey, F.; Sabbe, K.; Sasa, S.; Sar, E.; Taylor, J.; Van De Vijver, B.; Weitzel, C.E.; Williams, D.M.; Wiktowski, A. & Wiktowski, J. (2018): – In DiatomBase. Accessed at http://www.diatombase.org on 2018–03–15.

Krammer, K. & Lange–Bertalot, H. (1991): Bacillariophyceae 4. Teil: Achnanthaceae. Kritische Ergänzungen zu Navicula (Lineolatae) und Gomphonema. Gesamtliteraturverzeichnis Teil 1–4. – In: Ettl, H.; Gerloff, J.; Heyning, H. & Mollenhauer, D. (eds): Süßwasserflora von Mitteleuropa., Vol. 2/4, – pp. 1–437, Gustav Fischer Verlage, Stuttgart.

Kützing, F.T. (1844): Die Kieselalgen der Batillarien oder Diatomeen. – pp. 152, Nordhausen: zu finden bei W. Kühne.

Liu, B.; Blanco, S.; Long, H.; Xu, J.J. & Jiang, X.Y. (2016): Achnanthidium sinense sp. nov. (Bacillariophyta) from the Wuling Mountains Area, China. – Phytotaxa 284: 194–202.

Marquardt, G.C.; Costa, L.F.; Bicudo, D.C.; Bicudo, C.E.D.M.; Blanco, S.; Weitzel, C.E. & Ector, L. (2017): Type analysis of Achnanthidium minitissium and A. catenatum and description of A. tropicocaterata sp. nov. (Bacillariophyta), a common species in Brazilian reservoirs. – Plant Ecology & Evolution 150: 313–330.

Novais, M.H.; Hludiková, D.; Morais, M.; Hoffmann, L. & Ector, L. (2011): Morphology and ecology of Achnanthes caravelense (Bacillariophyceae), a new species from Portuguese rivers. – Algalological Studies 136: 131–150.

Parr, J.F.; Taffs, K.H. & Lane, C.M. (2004): A microwave digestion technique for the extraction of fossil diatoms from coastal lake and swamp sediments. – Journal of Paleolimnology 31: 383–390.

Péres, F.; Cohu, R.L. & Delmont, D. (2014): Achnanthidium barbei sp. nov. and Achnanthidium costei sp. nov., two new diatom species from French rivers. – Diatom Research 29: 387–397.

Pinseel, E.; Van De Vijver, B. & Kopalova, K. (2015): Achnanthidium petuniabuktianum sp. nov. (Achnanthesaceae, Bacillariophyta), a new representative of the A. pyrenacicum group from Spitsbergen (Svalbard Archipelago, High Arctic). – Phytotaxa 226: 63–74.

Potapova, M. & Ponader, K.C. (2007): Diatoms from the genus Achnanthidium in flowing waters of the Appalachian Mountains (North America): Ecology, distribution and taxonomic notes. – Limnologica 37: 227–241.

Potapova, M. & Ponader, K.C. (2004): Two common North American diatoms, Achnanthidium rivulare sp. nov. and A. deflexum (Reimer) Kingston: morphology, ecology and comparison with related species. – Diatom Research 19: 33–57.

Round, F.E.; Crawford, R.M. & Mann, D.G. (1990): The Diatoms. Biology and morphology of the genera. – pp. 747, Cambridge University Press, Cambridge.

Round, F. & Bukhtiyarova, L. (1996): Four new genera

References

Compère, P. & Van De Vijver, B. (2011): Achnanthidium ennediensis (Compère) Compère et Van de Vijver comb. nov. (Bacillariophyceae), the true identity of Navicula ennediensis compère from the Ennedi mountains (Republic of Chad). – Algalological Studies 136/137: 5–17.

Czarnecki, D.B. (1994): The freshwater diatoms culture collection at Loras College, Dubuque, Iowa. – In: Kockolek, J.P. (ed.): Proceedings of the 11th International Diatom Symposium. – Memoirs of the California Academy of Sciences 17: 155–174.

Jüttner, I.; Chimondes, J. & Cox, J. (2011): Morphology, ecology and biogeography of diatom species related to Achnanthes pyrenacium (Hustedt) Kobayasi (Bacillariophyceae) in streams of the Indian and Nepalese Himalaya. – Algalological Studies 136/137: 45–76.

Kartheck, B.; Taylor, J.C. & Hamilton, P.B. (2017): Two new species of Achnanthidium Küting (Bacillariophyceae) from Kolli Hills, Eastern Ghats, India. – Fottea 17: 65–77.

Kingston, J.C. (2000): New combinations in the freshwater Fragilariaceae and Achnanthidaceae. – Diatom Research 15: 409–411.

Kingston, J.C. (2003): Araphid and Monoraphid Diatoms. – In: Wehr, J.D. & Sheath, R.G. (eds): Freshwater Algae of North America. Ecology and Classification. – pp. 595–636, Elsevier Science.

Kobayasi, H.; Nagumo, T. & Mayama, S. (1986): Observations on the Two Rheophilic species of the Genus Achnanthes (Bacillariophyceae), A. convergens H. Kob. and A. japonica H. Kob. – Diatom 2: 83–93.

Kobayasi, H.; Idei, M.; Mayama, S.; Nagumo, T. & Osada, K. (2006): H. Kobayasi’s Atlas of Japanese Diatoms based on electron microscopy. – pp. 531, Uchida Rokakuho Publishing Co., Ltd, Tokyo.
based on *Achnanthes* (*Achnanthidium*) together with re-definition of *Achnanthidium*. – *Diatom Research* 11: 345–361.

Taylor, J.C.; Morales, E.A. & Ector, L. (2011): *Achnanthidium standeri* (Cholnoky) comb. nov. and *Achnanthidium taiaense* (J. R. Carter et Denny) comb. nov. two new combinations of morphologically similar *Achnanthidium* species from Africa. – Algological Studies 136/137: 151–166.

Vuver, B.V.D.; Ector, L.; Beltrami, M.E.; Haan, M.D., Falasco, E.; Hlubiková, D.; Jarlman, A.; Kelly, M.; Noæs, M.H. & Wotjal, A.Z. (2011): A critical analysis of the type material of *Achnanthidium lineare* W. Sm. (Bacillariophyceae). – Algological Studies 136/137: 167–191.

Wotjal, A.Z.; Lange-Bertalot, H.; Nautiyal, R.; Verma, J. & Nautiyal, P. (2010): *Achnanthidium chitrakootense* spec. nov. from rivers of northern and central India. – Polish Botanical Journal 55: 55–64.

Wotjal, A.Z.; Ector, L.; Vuver, B.V.D.; Morales, E.; Lanza, S.B.; Piatek, J. & Smieja, A. (2011): The *Achnanthidium minutissimum* complex (Bacillariophyceae) in southern Poland. – Algological Studies 136: 211–238.

Yana, E. & Mayama, S. (2015): Two new taxa of *Achnanthidium* and *Encyonema* (Bacillariophyceae) from the Yom River, Thailand, with special reference to the areolae occlusions implying ontogenetic relationship. – Phycological Research 63: 239–252.

You, Q.M.; Kociolek, J.P. & Wang, Q.X. (2015): The diatom genus *Hantzschia* (Bacillariophyta) in Xinjiang Province, China. – Phytotaxa 197: 1–14.

Yu, P.; You, Q.M.; Kociolek, J.P.; Lowe, R. & Wang, Q.X. (2017): *Nupela major* sp. nov. a new diatom species from Maolan nature reserve, central–south of China. – Phytotaxa 311: 245–254.

Yu, P.; Kociolek, J.P.; You, Q.M. & Wang, Q.X. (Accepted): *Achnanthidium longissima* sp. nov. (Bacillariophyta), a new diatom species from Jiuzhai Valley, Southwestern China. – *Diatom Research*. DOI: 10.5507/fot.2018.015
Figs 1–20. LM valve views of Achnanthidium lacustre sp. nov.: (21–55) LM valve views of A. sublanceolatum sp. nov., (21–37) Raphe valves, (38–55) Rapheless valves; (56–89) LM valve views of A. taipingensis sp. nov., (56–72) Raphe valves, (73–89) Rapheless valves. Scale bar 10 µm.
Figs 90–97. *Achnanthidium lacustre* sp. nov. SEM views of raphe valve: (90) External view of an entire raphe valve; (91) Internal view of an entire raphe valve; (92–93) Details of the apices and the central area on the external of the raphe valve; (93) Detail of the apices and the central area on the internal of the raphe valve; (96) Detail of the areolae on the external of the raphe valve; (97) Detail of the areolae on the internal of the raphe valve. Scale bars 5 µm (90–91), 0.5 µm (92–93, 96–97), 0.2 µm (94–95).
Figs 98–105. *Achnanthidium lacustre* sp. nov., SEM views of rapheless valve: (98) External view of an entire rapheless valve; (99) Internal view of an entire rapheless valve; (100) Detail of the apices on the external of the rapheless valve; (101) Detail of the apices on the external of the rapheless valve; (105) Detail of the apices on the internal of the rapheless valve; (102–103) Middle portion of the rapheless valve; (104) Detail of the areolae on the external of the rapheless valve. Scale bars 5 µm (98–99), 1 µm (102–103), 0.5 µm (100–101, 105), 0.2 µm (104).
Figs 106–114. *Achnanthidium sublanceolatum* sp. nov., SEM views of raphe valve: (106) External view of an entire raphe valve; (107) Internal view of an entire raphe valve; (108) Detail of the apices on the external of the raphe valve; (109) Detail of the apices on the internal of the raphe valve; (110) Detail of the middle portion on the external of the raphe valve; (111) Detail of the middle portion on the internal of the raphe valve; (112–113) External areolae; (114) Internal areolae openings with fine hymenate structures. Scale bars 5 µm (106–107), 1 µm (110–111), 0.5 µm (108–109), 0.2 µm (112–114).
Figs 115–123. *Achnanthidium sublanceolatum* sp. nov., SEM views of raphe valve: (115) External view of an entire rapheless valve; (116) Internal view of an entire rapheless valve; (117–119) Details of the apices on the external of the rapheless valve; (120) Detail of the apices on the internal of the raphe valve; (121) Internal areolae openings with fine hymenate structures; (122) Detail of the middle portion on the external of the raphe valve; (123) Detail of the middle portion on the internal of the raphe valve. Scale bars 5 µm (115–116), 1 µm (122–123), 0.5 µm (117–118,120), 0.2 µm (119, 121).
Figs 124–130. Achnanthidium taipingensis sp. nov., external views of raphe valve, SEM: (124) Entire raphe valve; (125) External areolae; (126–127) Details of distal raphe ends terminating on the valve mantle to form a drop-like areas. There is a depression near the terminal raphe fissures; (128–130) Details of the middle portion of the raphe valve. Scale bars 5 µm (124), 0.5 µm (126-130), 0.2 µm (125).
Figs 131–138. Achnanthidium taipingensis sp. nov., internal views of raphe valve and rapheless valve, SEM: (131) View of an entire raphe valve; (132) View of an entire rapheless valve; (133) Detail of the apices of the raphe valve; (134) Detail of the apices of the rapheless valve; (135) Detail of the middle portion of the raphe valve; (136) Detail of the middle portion of the rapheless valve; (137–138) Areolae occluded by hymens, which are partially merged with adjacent hymens. Scale bars 5 µm (131–132), 0.5 µm (133–136), 0.2 µm (137–138).
Figs 139–146. Achnanthidium taipingensis sp. nov., SEM: (139) Girdle view, a frustule with convex rapheless valve; (140) Entire rapheless valve; (141) Details of the apices of the girdle view; (142–144) Details of the apices on the external of the rapheless valve; (145–146) Details of the middle portion of the rapheless valve. Scale bars 5 µm (139–140), 0.5 µm (141–143, 145–146), 0.2 µm (144).