R-High-CHOP/CHASER/LEED with autologous stem cell transplantation in newly diagnosed mantle cell lymphoma: JCOG0406 STUDY

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Abbreviations: ASCT, autologous hematopoietic stem cell transplantation; CHASER, cyclophosphamide, high-dose cytarabine, dexamethasone, etoposide, and rituximab; CHOP, cyclophosphamide, doxorubicin, vincristine, and prednisone; CI, confidence interval; CPA, cyclophosphamide; CR, complete response; GELA, Groupe d’Etude des Lymphomes de l’Adulte; HDAC, high-dose cytarabine; HDAC, high-dose chemotherapy; JCOG-LSG, Japan Clinical Oncology Group- Lymphoma Study Group; LEED, melphalan, CPA, etoposide and dexamethasone; LYSA, Lymphoma Study Association; MCL, mantle cell lymphoma; MIPI, mantle cell lymphoma international prognostic index; MIPI-c, modified combination of the Ki-67 index and MIPI; NHL, non-Hodgkin lymphoma; OS, overall survival; ORR, overall response rate; PBSC, peripheral blood stem cell; PFS, progression-free survival; PS, performance status; R, rituximab; R-DHAP, rituximab, dexamethasone, HDAC and cisplatin; WHO, World Health Organization.

Clinical Trial registration: UMIN Clinical Trials Registration number: UMIN000001220

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Although induction immunochemotherapy including high-dose cytarabine and rituximab followed by high-dose chemotherapy (HDC) with autologous stem cell transplantation (ASCT) is recommended for younger patients (<65 years old) with untreated mantle cell lymphoma (MCL), no standard induction and HDC regimen has been established. We conducted a phase II study of induction immunochemotherapy of R-High-CHOP/CHASER followed by HDC of LEED with ASCT in younger patients with untreated advanced MCL. Eligibility criteria included untreated MCL, stage II bulky to IV, and age 20-65 years. Patients received 1 cycle of R-High-CHOP followed by 3 cycles of CHASER every 3 weeks. Peripheral blood stem cells (PBSC) were harvested during CHASER. LEED with ASCT was delivered to patients who responded to R-High-CHOP/CHASER. Primary endpoint was 2-year progression-free survival (PFS). From June 2008 to June 2012, 45 patients (median age 59 years; range 38-65 years) were enrolled. PBSC were successfully harvested from 36 of 43 patients. Thirty-five patients completed ASCT. Two-year PFS was 77% (80% CI 68-84), which met the primary endpoint. Five-year PFS and overall survival were 52% (95% CI 34-68%) and 71% (95% CI 51-84%), respectively. Overall response and complete response rates after induction immunochemotherapy were 96% and 82%, respectively. The most common grade 4 toxicities were hematological. In younger patients with untreated MCL, R-High-CHOP/CHASER/LEED with ASCT showed high efficacy and acceptable toxicity, and it can now be considered a standard treatment option.

KEYWORDS
autologous stem cell transplantation, cytarabine, high-dose chemotherapy, mantle cell lymphoma, rituximab

1 | INTRODUCTION

Mantle cell lymphoma (MCL) is a well-recognized B-cell lymphoma subtype that accounts for approximately 5% of all patients with NHL. The clinical course of MCL ranges from indolent to aggressive, with a poor prognosis and a median OS of about 3-5 years with conventional chemotherapy.2,3 The prognosis when using conventional chemoinmunotherapy remains poor. Two-year PFS of 30% was reported in a phase II study of MCL patients treated with 6 cycles of rituximab, an anti-CD20 antibody, and CHOP chemotherapy.4 However, a randomized phase III study by the European MCL Network that compared myeloablative radiochemotherapy followed
by ASCT with interferon-α (IFN-α) maintenance during the first remission after a CHOP-like regimen demonstrated significant superiority of ASCT, with PFS of 45% in younger patients aged 65 years or less in patients with advanced-stage MCL.5 Promising approaches to improving CR rate before ASCT, as well as PFS and OS, include modified induction therapy with HDAC-based chemotherapy regimens and rituximab. These strategies are based on clinical studies in which the addition of rituximab to an HDAC-containing regimen was reported to ensure tumor depletion in vivo while allowing the collection of PBSC with conserved engraftment capability that was devoid of tumor cells.6,7

A phase II MCL-2 study by the Nordic Lymphoma Group including HDAC and rituximab prior to stem cell mobilization, followed by HDC and ASCT, demonstrated an excellent ORR (96%), with a CR rate of 56%, and PFS of 70% and OS of 70% after 6 years.8 Similar promising results were also reported in another phase II study of an induction regimen with R-CHOP and R-DHAP followed by ASCT, which was conducted by GELA.9 This regimen of 6 cycles of alternating R-CHOP/R-DHAP followed by consolidative HDC with ASCT resulted in a superior PFS compared with the regimen of 6 cycles of R-CHOP followed by consolidative HDC with ASCT reported for a randomized phase III study by the European Mantle Cell Network (European MCL Network).10 Recently, rituximab maintenance after ASCT was shown to improve event-free survival, PFS, and OS in younger patients with MCL in a randomized phase III study by LYSAl.11 Thus, HDC with ASCT after intensive immunochemotherapy using rituximab and HDAC as first-line treatment, followed by rituximab maintenance, appears to be the only current therapy that might improve the outcomes of younger patients with untreated advanced MCL.10-12 However, there was no established regimen for induction and HDC when the present study was planned.

We developed the R-High-CHOP/cyclophosphamide, high-dose cytarabine, dexamethasone, etoposide, and rituximab (CHASER) regimen as an induction therapy, and LEED therapy as HDC. One cycle of the R-High-CHOP regimen was incorporated to enhance MCL tumor reduction. CHASER and CHASE were originally developed for both salvage therapy without a platinum agent to avoid renal toxicity and for in vivo efficacy of purging of B-cell lymphoma cells during harvesting of autologous PBSC.13,14 Based on promising data from a small pilot study of R-High-CHOP/CHASER/LEED in newly diagnosed MCL in a single institute of the Aichi Cancer Center (data not shown), we conducted a phase II study (JCOG0406) of this regimen using a new induction procedure comprising R-High-CHOP/CHASER13,14 with rituximab and HDAC followed by HDC (LEED therapy) with ASCT.

Written, informed consent was obtained from each patient before enrolment in accordance with the Declaration of Helsinki. The trial is registered with the UMIN Clinical Trials Registry (UMIN000001220).

2.2 | Eligibility criteria

Eligibility criteria included the following: newly pathologically diagnosed with MCL according to the WHO classification 2001,15 positive staining for cyclin D1 in the nucleus of lymphoma cells; positive for CD5 and CD20 on lymphoma cells by flow cytometry or by immunohistochemistry; age between 20 and 65 years; ECOG performance status (PS) of 0, 1, or 2; clinical stage II with bulky disease, stages III or IV according to the Ann Arbor staging system; lymphoma cells in peripheral blood ≤10 000/μL; no involvement of the central nervous system; at least 1 measurable lymphomatous lesion; no previous history of chemotherapy, radiation therapy, IFN, and/or antibody therapy; adequate organ function; and written, informed consent. Patients were excluded if they had a history of glaucoma, uncontrollable diabetes mellitus, uncontrollable hypertension, hepatitis B virus surface antigens or antibodies to hepatitis C virus, interstitial pneumonia, severe bacterial infection, or another active neoplasm.

2.3 | Treatment

The R-High-CHOP/CHASER regimen comprised 1 cycle of R-High CHOP consisting of 1500 mg/m² CPA and 75 mg/m² doxorubicin followed by 3 cycles of CHASER consisting of 1200 mg/m² CPA on day 3, 2 g/m² cytarabine on days 4 and 5, 100 mg/m² etoposide on days 3-5, 40 mg/body weight dexamethasone on days 1, 3-5 and 15, and 375 mg/m² rituximab on days 1 and 15 (Figure 1). HDC in LEED therapy consisting of 130 mg/m² melphalan, 60 mg/kg CPA, 500 mg/m² etoposide, and 40 mg/body dexamethasone was started on day 36-49 of the last CHASER therapy. PBSC were harvested after the 2nd cycle of CHASER, and further harvesting was added after the 3rd CHASER cycle if the number of PBSC was less than 4 × 10⁶ cells/kg. Although the recommended number of harvested CD34-positive cells was ≥4 × 10⁶ cells/kg, patients obtaining ≥2 × 10⁶ cells/kg could proceed to HDT with ASCT. Patients received sulfamethoxazole-trimethoprim from induction therapy until 6 months after ASCT to prevent opportunistic infections caused by Pneumocystis jirovecii.

2.4 | Central pathology review

A central pathology review was carried out as previously reported.16 Antigens routinely examined by immunohistochemistry included CD3, CD5, CD10, CD20, cyclin D1, SOX-11, Ki-67, and cyclin D1. Four hematopathologists reviewed the pathology specimens and classified them according to the WHO classification system 2001.15 The diagnosis by the central pathology review committee was used in this article.
Mantle cell lymphoma international prognostic index (MIPI) and modified combination of the Ki-67 index and MIPI (MIPI-c)

Patients were classified into low-risk, intermediate-risk, and high-risk groups based on the 4 prognostic factors (age, PS, lactate dehydrogenase [LDH], and leukocyte count) according to MIPI. Patients were classified into low-risk, low-intermediate risk, high-intermediate risk, and high-risk groups according to MIPI-c.

Response and toxicity criteria

Tumor assessments were carried out on all target lesions identified at baseline by PET and CT scans after R-High-CHOP/CHASER and after LEED therapy completion (day 50-63 after ASCT). In patients in CR after LEED therapy, tumor assessment was done every 6 months for 2 years and, thereafter, every year for the next 3 years. Tumor response was determined by the Revised Response Criteria for Malignant Lymphoma 2007. Toxicities were evaluated according to the NCI Common Terminology Criteria for Adverse Events version 3.0.

Statistical analysis and endpoints

Primary endpoint was 2-year PFS. Planned sample size was 45, with an expected 2-year PFS of 50%, a threshold of 30%, 1-sided $\alpha$ of 10%, and power of 90% based on the results of conventional chemotherapy and toxicities of R-High-CHOP/CHASER/LEED (M. Ogura, unpublished data, 2000).

Secondary endpoints were ORR, CR rate, proportion of CR and ORR after induction therapy, PFS, 5-year PFS, OS, 2-year OS, 5-year OS, and toxicity. OS was calculated from the date of registration until death as a result of any cause or censored at the last follow-up date. PFS was calculated from the date of registration to the date of relapse or progression, death as a result of any cause, or censored at the date of the last follow-up. CR rate and ORR (CR + PR) and 95% CI were estimated by an exact binomial method. OS and PFS were estimated according to the Kaplan-Meier method, and CI were estimated by Greenwood's formula. These analyses were carried out using SAS release 9.2 (SAS Institute, Cary, NC, USA).
TABLE 1 Characteristics of patients in the present study (N = 45)

|                     | N   | %    |
|---------------------|-----|------|
| Gender              | 4/41| 9/91 |
| Age (years)         | 59  |      |
| Range               | 38-65|      |
| PS                  | 0/1/2| 38/6/1| 85/13/2|
| Clinical stage      | II bulky/III/IV | 0/5/40 | 0/11/89 |
| Central pathological review | MCL | 45 | 100 |
| International prognostic index (IPI) | L/LI/HI/H | 8/17/17/2 | 18/39/39/4 |
| Mantle cell lymphoma IPI (MIPI) | L/Int/H | 28/14/2 | 64/32/4 |
| MIPI-c              | L/LI/HI/H | 21/13/8/2 | 48/30/18/4 |
| SOX-11              | Positive/negative | 43/1 | 98/2 |
| Bulky mass, tumor size (cm) | Size <5 cm | 29 | 64 |
|                     | 5 cm ≤ Size < 10 cm | 7 | 16 |
|                     | Size ≥10 cm | 9 | 20 |
| Extranodal lesions  | 0-1 | 19 | 42 |
|                     | ≥2  | 26 | 58 |
| Bone marrow involvement | 36 | 82 |
| PB involvement      | 19 | 43 |

\*In IPI, MIPI, MIPI-c, SOX11, and involvement of lymphoma cells in bone marrow and peripheral blood, the total number of patients was 44. H, high-risk; HI, high-intermediate risk; Int, intermediate-risk; L, low-risk; LI, low-intermediate risk; MCL, mantle cell lymphoma; PB, peripheral blood; PS, performance status.

45) disease. Distribution of risk according to MIPI and MIPI-c is shown in Table 1. Supplementary analyses of 44 patients whose formalin-fixed paraffin-embedded tissue blocks were available at the institution were conducted in June 2016 (data cut-off date was June 2015).

3.2 | Feasibility

Flowchart of the clinical course of enrolled patients is shown in Figure 2. All 45 enrolled patients received R-High-CHOP, among whom 40 patients completed R-High-CHOP followed by 3 cycles of CHASER, and 5 patients did not (Figure 2). Although all 40 patients who completed the R-High-CHOP/CHASER induction therapy achieved CR or PR, 5 of these 40 patients did not receive high-dose LEED therapy with ASCT because of insufficient harvest of auto-PBSC in 4 patients and an adverse event in 1 patient. Therefore, 35 patients received high-dose LEED therapy with ASCT. PBSC were harvested in 43 of 45 patients because induction therapy was discontinued at less than 2 cycles as a result of disease progression or patient's refusal in 1 case each. In 43 patients, median number of harvested PBSC was 3.80 (range 0.40-38.40) × 10^9/kg cells. In 36 patients, ≥2 × 10^9/kg cells PBSC were harvested, whereas in 7 patients, less than 2 × 10^9/kg cells PBSC were harvested. Percentage of successfully collected PBSC was 84%. In 2 of 7 patients with insufficient harvest of auto-PBSC, additional harvests were carried out outside the protocol, and they received high-dose LEED therapy with ASCT. These 2 patients were finally judged as having protocol treatment.

3.3 | Responses

Responses of the 45 enrolled patients are shown in Table 2. ORR and CR rates after R-High-CHOP/CHASER induction therapy were 95.6% (95% CI 84.9-99.5%) and 82.2% (95% CI 68.0-92.0%), respectively. ORR and CR rates in all 45 patients after induction and LEED therapy were 77.8% (95% CI 62.9-88.8%) and 71.1% (95% CI 55.7-83.6%), respectively.

3.4 | Progression-free survival

Median follow-up time for all enrolled patients was 3.7 years in the primary analysis. Two-year PFS as the primary endpoint of all enrolled patients was estimated to be 77.3% (lower boundary of 80% CI 68.0%, which exceeded the threshold of 30%, and 95% CI 61.9-87.1%), which met the primary endpoint (Figure 3A). PFS at 5 years was estimated to be 52.2% (95% CI 33.8-67.8). PFS at 5 years for patients with low-risk (N = 28), intermediate-risk (N = 14), and high-risk (N = 2) disease according to MIPI was 59.9% (95% CI 37.4-76.6%), 51.6% (95% CI 21.6-75.1%), and 0%, respectively (Figure 3B). PFS at 5 years for patients with low-risk (N = 21), low-intermediate risk (N = 13), high-intermediate risk (N = 8), and high-risk (N = 2) disease according to MIPI-c was 66.2% (95% CI 38.4-83.8%), 44.0% (95% CI 16.8-68.4%), 58.3% (95% CI 18.0-84.4%), and 0%, respectively (Figure 3C). PFS rates at 5 years for patients who received LEED therapy followed by ASCT (n = 35) and for patients who did not receive LEED therapy followed by ASCT for any reason including harvesting failure of PBSC (n = 10) were 54.8% (95% CI 33.0-72.1%) and 42.2% (95% CI 11.1-71.3%), respectively.

3.5 | Overall survival

Overall survival of all 45 enrolled patients is shown in Figure 4A. Eleven patients died (7 died of MCL, 3 died of secondary malignancy [1 acute myelogenous leukemia (AML), 1 adult T-cell leukemia/lymphoma (ATL), and 1 diffuse large B-cell lymphoma (DLBCL)], and 1 died after postprotocol treatment). OS at 2 years and 5 years was estimated to be 91% (95% CI 77.7-96.5%) and 71.0% (95% CI 50.9-84.1%), respectively. Subgroup analysis of OS according to MIPI and
MIPI-c is shown in Figure 4B,C, respectively. OS rates at 5 years for patients who received LEED therapy followed by ASCT (n = 35) and for patients who did not receive LEED therapy followed by ASCT for any reason including harvesting failure of PBSC (n = 10) were 71.9% (95% CI 47.4-86.4%) and 67.5% (95% CI 29.1-88.3%), respectively.

3.6 | Toxicity

All 45 treated patients were evaluated for toxicity (Table 3). The most common grade 4 toxicities were hematological; the percentage of patients with grade 4 neutropenia and thrombocytopenia was 80% and 0% in R-High-CHOP, 91% and 89% in CHASER, and 94% and 77% in LEED, respectively. There were no grade 4 nonhematological or nonlaboratory toxicities during the entire protocol treatment. In R-High-CHOP therapy, 1 grade 4 increased alanine aminotransferase (ALT) was observed, and grade 3 increased aspartate aminotransferase (AST) and hyponatremia were observed in 1 patient each. All of these adverse events were reversible.

In CHASER therapy, grade 4 hypokalemia was observed in 1 patient, and grade 3 hyponatremia and hypokalemia were observed in 1 and 3 patients, respectively. All of these adverse events were reversible.

In LEED therapy, grade 4 hypokalemia was observed in 1 patient, and grade 3 increased AST, increased ALT, hyponatremia, and hypokalemia were observed in 2, 3, 1, and 5 patients, respectively. All of these adverse events were reversible. In the present study, 3 types of opportunistic infection were observed in 5 patients, including grade 3 Pneumocystis pneumonia in 2 patients, grade 3 cytomegalovirus infection in 2 patients, and grade 3 adenovirus cystitis in 2 patients; one patient had cytomegalovirus infection followed by adenovirus cystitis, sequentially. Treatment-related death occurred in 1 patient as a result of brain DLBCL after the development of Epstein Barr (EB) virus-positive post-transplantation lymphoproliferative disorder (PTLD) on day 159 after ASCT. Secondary malignancies including AML, prostate cancer, DLBCL, and ATL developed in 1 patient each. Incidence of secondary malignancies was 8.9% (95% CI 2.5-21.2%).

3.7 | Pathological characteristics

A central review of the pathological diagnosis was carried out for 45 enrolled patients, and all were confirmed as having cyclin D1-positive MCL, although 1 patient was diagnosed as having CD5-negative MCL. Although CD5 positivity was defined as an eligibility criterion in this study, the Central Pathology Review Committee decided that this patient was pathologically eligible, as CD5 negativity was not unequivocally confirmed by other techniques, including flow cytometry.
This phase II study showed that treatment of untreated younger MCL patients with R-High-CHOP/CHASER followed by LEED HDC with ASCT resulted in high ORR and CR rates with durable PFS and OS and acceptable toxicity profiles. These results show that this regimen of R-High-CHOP/CHASER/LEED with ASCT can now be considered a standard treatment option in this population.

In the present study, PFS with the R-High-CHOP/CHASER/LEED with ASCT regimen was comparable with that reported for other regimens containing rituximab and HDAC followed by consolidative HDC with ASCT. Addition of both HDAC and rituximab (MCL2
study) dramatically improved the PFS (4-year PFS of 73% and 6-year PFS of 66%) compared to the PFS (4-year PFS of 37%) with previous MCL-1 protocol treatments.\(^8,20\) The 5-year event-free survival in the GELA phase II study of 3 cycles of R-CHOP and 3 cycles of R-DHAP followed by ASCT was 64%.\(^7\) In the randomized phase III study by the European MCL Network, the 5-year PFS in the experimental arm of 6 cycles of the alternating R-CHOP/R-DHAP regimen followed by consolidative HDC with ASCT and the control arm of 6 cycles of R-CHOP followed by consolidative HDC with ASCT was 65% and 44%, respectively.\(^10\) The present results provide further data to support the treatment approach used in these prior studies. Combined with the present data, the present findings strongly suggest that HDAC-based high-dose consolidation therapy followed by ASCT has dramatically improved the prognosis of MCL compared with the

**FIGURE 4** Kaplan-Meier curves of overall survival (OS) (A) of all 45 enrolled patients, and Kaplan-Meier curves of OS according to risk stratified by MIPI (n = 44) (B) and MIPI-c (n = 44) (C). MIPI, mantle cell lymphoma international prognostic index; MIPI-c, modified combination of the Ki-67 index and MIPI.
Recently, rituximab maintenance after ASCT was reported to improve event-free survival, PFS, and OS in younger patients with MCL.\(^5\) PFS at 4 years was 83% in the rituximab maintenance group versus 64% in the observation group (\(P < .001\)), and OS was 89% vs 80% (\(P = 0.04\)), respectively. In this study, no patients received rituximab maintenance, and PFS and OS at 4 years were almost the same as in the observation group. It is likely that adding rituximab maintenance to our regimen will improve outcomes further.

In order that as many patients as possible can proceed to ASCT, it is important to achieve CR or PR after the induction chemotherapy before ASCT. In the present study, ORR and CR rates after R-High/CHOP/CHASER induction therapy were 95.6% (95% CI 84.9-99.5%) and 82.2% (95% CI 68.0-92.0%), respectively. In the Nordic MCL-2 study, ORR and CR rates including uncertain CR were 96.3% and 54.4%, respectively.\(^8\) In the GELA phase II study, ORR and CR rates after induction therapy were 95% and 57%, respectively.\(^7\) In the European MCL Network phase III study, ORR and the CR including unconfirmed CR rate after alternating R-CHOP/R-DHAP were 94% and 55%, respectively.\(^10\) In a phase III LYSA study to evaluate the usefulness of rituximab maintenance treatment, induction therapy was 4 cycles of R-DHAP (6.7% of patients received an additional 3 cycles of R-CHOP), and ORR and CR rates were 94% and 74% (including 34% with unconfirmed CR), respectively.\(^11\) Thus, the high CR rate of 82.2% in the present study is notable. Recently, retrospective data of a nationwide study of MCL in Japan were reported.\(^22\) In that study, 501 patients with newly diagnosed MCL with a median age of 67 (range 22-90) years treated with rituximab-containing therapy between 1992 and 2012 were analyzed. PFS and OS at 5 years were 25% and 60%, respectively. These results are lower than in the present study (PFS of 52% and OS of 71% at 5 years). Although comparison is difficult, the most probable reason for these differences might be differences in the study populations: The median age was 67 years in the nationwide retrospective study and 59 years in the present study, and 48% of patients received R-CHOP without ASCT in the retrospective nationwide study, whereas all patients in the present study received treatment with the intent of ASCT.

In the present study, 22.2% of all enrolled patients did not receive ASCT, whereas in the Nordic MCL-2 study 6.8% of all 160 eligible patients did not proceed to the high-dose regimen because of toxicity or patient refusal.\(^8\) Five of the present 10 patients did not proceed to ASCT due only to insufficient harvest. In 7 of all 43 harvested patients, collected PBSC did not reach the threshold number of stem cells per protocol (\(\geq 2 \times 10^6\) CD34-positive cells per kg body weight). Thus, the percentage of PBSC collected was 84%.

According to the protocol, auto-PBSC were harvested during the second CHASER and, if necessary, during the third CHASER, to avoid contamination of harvested auto-PBSC with MCL cells. As highly efficient harvest of auto-PBSC was observed during the first CHASER (data not shown), it is possible that a greater number of patients may proceed to ASCT if the harvest of auto-PBSC is initiated during the first CHASER. Possibility that the harvest of auto-PBSC in the early period of intensive induction chemotherapy may result in a highly efficient harvest was suggested in a recent phase II study.\(^23\) In that randomized phase II study by the Southwest Oncology Group that compared HyperCVAD/MA with rituximab and bendamustine with rituximab (S1106 study), stem cell collection was planned and carried out after the 3rd of 4 HyperCVAD/MA cycles. The R-HyperCVAD/MA arm was prematurely closed as a result of failure of stem cell collection and/or delay of therapy because of hematological toxicity.\(^23\) Percentage (84%) of collected PBSC in the present study was higher than that (66%) reported in the alternating R-CHOP/R-DHAP regimen in the European MCL Network phase III study.\(^10\) PFS and OS at 5 years for patients who received LEED therapy followed by ASCT were relatively higher than those for patients who did not receive LEED therapy followed by ASCT for any reason, although the comparison is difficult because of the small sample size and because it was a subset analysis according to outcome.

Only 5 of the 45 eligible patients in the present study did not complete induction chemotherapy (R-High CHOP/CHASER) because of toxicity (\(N = 3\)), disease progression (\(N = 1\)), or refusal (\(N = 1\)). These proportions were much lower than the reported 29% of patients who did not complete the planned 6 or 8 HyperCVAD/MA cycles,\(^23\) and the reported 63% of patients who did not complete the planned 4 HyperCVAD/MA cycles with 3 deaths during therapy in a phase II study by Gruppo Italiano Studio Linfomi,\(^24\) but was

### TABLE 3 Toxicity (\(N = 45\)) in all protocol treatments

| CTCAE 3.0 | G1 | G2 | G3 | G4 | % G3-4 |
|-----------|----|----|----|----|--------|
| Leukocytes | 0  | 0  | 1  | 44 | 100%   |
| Hemoglobin | 0  | 3  | 27 | 14 | 91.1%  |
| Platelets  | 1  | 0  | 4  | 40 | 97.8%  |
| Neutrophils | 0 | 0  | 0  | 45 | 100%   |
| Hypoaalbuminemia | 27 | 17 | 0  | –  | 0%     |
| Bilirubin   | 20 | 5  | 0  | 0  | 0%     |
| AST         | 25 | 6  | 3  | 0  | 6.7%   |
| ALT         | 23 | 12 | 4  | 1  | 11.1%  |
| GGT         | 15 | 12 | 2  | 0  | 4.7%   |
| Cholesterol | 19 | 1  | 0  | 0  | 0%     |
| Creatinine  | 10 | 0  | 0  | 0  | 0%     |
| Hypernatremia | 11 | 0 | 0  | 0  | 0%     |
| Hyponatremia | 38 | – | 2  | 0  | 4.4%   |
| Hyperkalemia | 16 | 1 | 0  | 0  | 0%     |
| Hypokalemia | 27 | – | 9  | 1  | 22.2%  |
| Hypercalcemia | 1 | 0 | 0  | 0  | 0%     |
| Hypocalcemia | 18 | 7 | 0  | 0  | 0%     |

ALT, alanine aminotransferase; AST, aspartate aminotransferase; CTCAE, common terminology criteria for adverse events; G, grade; GGT, gamma-glutamyl transpeptidase; -, not applicable.
comparable to the 3% of such patients reported in the Nordic MCL-2 study and the 6.7% of such patients reported in the GELA study.

Our R-High-CHOP/CHASER/LEED therapy was well tolerated. The nonrelapse mortality of 4.4% (2/45), which included 1 case with ATL who was a HTLV type-1 carrier and developed acute type ATL after the completion of the protocol treatment, and 1 case with DLBCL as PTLD, is comparable with that reported in the Nordic MCL-2 study (5%). Apart from these 2 secondary malignancies, 2 secondary malignancies including AML and latent prostatic carcinoma developed after postprotocol treatment. Although a patient who developed latent prostatic carcinoma completed protocol treatment including ASCT, a patient who developed AML did not receive LEED therapy followed by ASCT. The cause of these 2 secondary malignancies is difficult to determine, especially with respect to the relationship with protocol treatment, because they developed following postprotocol treatment. No grade 2 or greater increased creatinine levels occurred during any of the treatment cycles, because R-High-CHOP/CHASER/LEED therapy does not contain any platinum agents. In contrast, 8.3% (N = 5; 3 patients with grade 3 or 4) of 60 enrolled patients developed renal insufficiency in the GELA study during which they were treated with DHAP using cisplatin.

In the present study, opportunistic infections including cytomegalovirus infection, adenovirus cystitis and Pneumocystis pneumonia were observed in 5 patients. Although the patients recovered from these infections, routine monitoring of the number of peripheral CD4+ lymphocytes was essential because intensive immunotherapy caused severe lymphocytopenia. After encouraging all participating institutions to do this examination, no further opportunistic infections were reported. In the present study, 22.2% of patients developed grade 3 or 4 hypokalemia. Grade 4 hypokalemia developed in 1 patient after both induction therapy and LEED high-dose therapy. The most likely reason for hypokalemia may have been the frequent use of diuretics for increased body weight as a result of the considerable volume of infusion fluid for CHASER and LEED therapy.

Recently, Hoster et al reported that a modified MIP1 that combines the Ki-67 index with MIPI (MIPI-c) provided a refined risk stratification, reflecting their strong complementary prognostic effects while integrating the most relevant prognostic factors available in routine clinical practice. In the present study, a very good OS at 5 years of 95.2% (95%CI 70.7-99.3%) in low-risk group (N=21), and a very poor OS of 0% in high-risk group (N=2) were also observed when risk group was stratified according to MIPI-c.

Last, the incorporation of new molecular targeted drugs into the current standard regimens is an attractive approach to improving the efficacy of treatment strategies for MCL patients. It has been reported that Bruton’s tyrosine kinase inhibitors and phosphoinositide 3-kinase inhibitors showed high efficacy in relapsed or refractory MCL patients. Further improvements of our regimen, including more efficient harvest of auto-PBSC and/or incorporation of promising new molecular targeting agents, are important future goals.

In conclusion, R-High-CHOP/CHASER including HDAC and rituximab followed by LEED HDC with ASCT showed a high CR rate and durable PFS and OS in patients aged 65 years or younger with newly diagnosed advanced MCL. This regimen compares favorably with other regimens including HDAC, rituximab, and HDC with ASCT and can now be considered a standard treatment option for newly diagnosed younger patients with MCL.

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CONFLICTS OF INTEREST

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REFERENCES

1. Schmidt C, Dreyling M. Therapy of mantle cell lymphoma: current standards and future strategies. Hematol Oncol Clin North Am. 2008;22:953-963.

2. Fisher RI, Dahlberg S, Nathwani BN, et al. A clinical analysis of two indolent lymphoma entities: mantle cell lymphoma and marginal zone lymphoma (including the mucosa-associated lymphoid tissue and monocytoid B-cell subcategories): a Southwest Oncology Group study. Blood. 1995;85:1075-1082.

3. Yatabe Y, Suzuki R, Tobinai K, et al. Significance of cyclin D1 overexpression for the diagnosis of mantle cell lymphoma: a clinicopathologic comparison of cyclin D1-positive MCL and cyclin D1-negative MCL-like B-cell lymphoma. Blood. 2000;95:2253-2261.

4. Howard OM, Gribben JG, Neuberger DS, et al. Rituximab and CHOP induction therapy for newly diagnosed mantle-cell lymphoma: molecular complete responses are not predictive of progression-free survival. J Clin Oncol. 2002;20:1288-1294.

5. Dreyling M, Lenz G, Hoster E, et al. Early consolidation by myeloablative radiochemotherapy followed by autologous stem cell transplantation in first remission significantly prolongs progression-free survival in mantle-cell lymphoma: results of a prospective randomized trial of the European MCL Network. Blood. 2005;105:2677-2684.

6. Magni M, Di Nicola M, Devizzi L, et al. Successful in vivo purging of CD34-containing peripheral blood harvests in mantle cell and indolent lymphoma: evidence for a role of both chemotherapy and rituximab infusion. Blood. 2000;96:864-869.

7. Voso MT, Pantel G, Weis M, et al. In vivo depletion of B cells using a combination of high-dose cytosine arabinoside/mitoxantrone and rituximab for autografting in patients with non-Hodgkin’s lymphoma. Br J Haematol. 2000;109:729-735.

8. Geisler CH, Kolstad A, Laurell A, et al. Long-term progression-free survival of mantle cell lymphoma after intensive front-line immunochemotherapy with in vivo-purged stem cell rescue: a non-randomized phase 2 multicenter study by the Nordic Lymphoma Group. Blood. 2008;112:2687-2693.

9. Delarue R, Haïoun C, Ribrag V, et al. CHOP and DHAP plus rituximab followed by autologous stem cell transplantation in mantle cell lymphoma: a phase 2 study from the Groupe d’Etude des Lymphomes de l’Adulte. Blood. 2013;121:48-53.

10. Hermine O, Hoster E, Walewski J, et al. Addition of high-dose cytarabine to immunochemotherapy before autologous stem-cell transplantation in patients aged 65 years or younger with mantle cell lymphoma (MCL Younger): a randomised, open-label, phase 3 trial of the European Mantle Cell Lymphoma Network. Lancet. 2016;388:565-575.

11. Le Gouill S, Thieblemont C, Oberic L, et al. Rituximab after autologous stem-cell transplantation in mantle-cell lymphoma. N Engl J Med. 2017;377:1250-1260.

12. Cheah CY, Seymour JF, Wang ML. Mantle cell lymphoma. J Clin Oncol. 2016;34:1256-1269.

13. Ogura M, Kagami Y, Taji H, et al. Pilot phase I/II study of new salvage therapy (CHASE) for refractory or relapsed malignant lymphoma. Int J Hematol. 2003;77:503-511.

14. Oki Y, Ogura M, Kato H, et al. Phase II study of a salvage regimen using cyclophosphamide, high-dose cytarabine, dexamethasone, etopo- side, and rituximab in patients with relapsed or refractory B-cell non-Hodgkin’s lymphoma. Cancer Sci. 2008;99:179-184.

15. Jaffe ES, Harris NL, Stein H, et al. World Health Organization Classification of Tumours Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues, 3rd edn. Lyon: IARC Press; 2001.

16. Ogura M, Itoh K, Ishizawa K, et al. Phase II study of ABV (doroxuribin with increased dose, bleomycin and vindristaine) therapy in newly diagnosed advanced-stage Hodgkin lymphoma: Japan Clinical Oncology Group study (JCOG9705). Leuk Lymphoma. 2013;54:46-52.

17. Hoster E, Dreyling M, Klapper W, et al. A new prognostic index (MIPI) for patients with advanced-stage mantle cell lymphoma. Blood. 2008;111:558-565.

18. Hoster E, Rosenwald A, Berger F, et al. Prognostic value of Ki-67 Index, cytology, and growth pattern in mantle-cell lymphoma: results from randomized trials of the European Mantle Cell Lymphoma Network. J Clin Oncol. 2016;34:1386-1394.

19. Cheson BD, Pfistner B, Juweid ME, et al. Revised response criteria for malignant lymphoma. J Clin Oncol. 2007;25:579-586.

20. Andersen NS, Pedersen L, Elenen E, et al. Primary treatment with autologous stem cell transplantation in mantle cell lymphoma: outcome related to remission pretransplant. Eur J Haematol. 2003;71:73-80.

21. Lenz G, Dreyling M, Hoster E, et al. Immunochemotherapy with rituximab and cyclophosphamide, doroxuribin, vincristine, and prednisone significantly improves response and time to treatment failure, but not long-term outcome in patients with previously untreated mantle cell lymphoma: results of a prospective randomized trial of the German Low Grade Lymphoma Study Group (GLSG). J Clin Oncol. 2005;23:1984-1992.

22. Chihara D, Asano N, Omachi K, et al. Prognostic model for mantle cell lymphoma in the rituximab era: a nationwide study in Japan. Br J Haematol. 2015;170:657-668.

23. Chen R, Li H, Bernstein SH, et al. RB but not R14 haplotypes are linked to outcome of patients with mantle cell lymphoma. J Clin Oncol. 2008;26:934-940.

24. Merli F, Luminari S, Ilariucci F, et al. Rituximab plus HyperCVAD alternating with high dose cytarabine and methotrexate for the initial treatment of patients with mantle cell lymphoma, a multicenter trial from Gruppo Italiano Studio Linfomi. Br J Haematol. 2012;156:346-353.

25. Wang ML, Rule S, Martin P, et al. BTK with ibrutinib in relapsed or refractory mantle-cell lymphoma. N Engl J Med. 2013;369:507-516.

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