Antibiotic Innovation May Contribute to Slowing the Dissemination of Multiresistant Streptococcus pneumoniae: The Example of Ketolides

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Abstract

Background: Despite increasingly frequent bacterial resistance to antibiotics, antibacterial innovation is rare. Ketolides constitute one of the very few new antibiotic classes active against Streptococcus pneumoniae developed during the last 25 years. Their mechanism of action resembles that of macrolides, but they are unaffected by common resistance mechanisms. However, cross-resistance to ketolides has been observed in some macrolide-resistant strains. We examined how new antibiotic exposure may affect overall pneumococcal resistance patterns in the population. The aims of this study were to assess the potential dissemination of newly emerged resistances and to control the selection of strains already multiresistant to existing antimicrobials.

Methodology/Principal Findings: We developed an age-structured population model for S. pneumoniae transmission in a human community exposed to heptavalent vaccine, and β-lactams, macrolides and ketolides. The dynamics of intra-individual selection of resistant strains under antibiotic exposure and interindividual transmission were simulated, with antibiotic-specific resistance mechanisms defining the path to co-resistances and cross-resistances, and parameters concerning the French situation. Results of this simulation study suggest that new antibiotic consumption could markedly slow the diffusion of multiresistant strains. Wider use was associated with slower progression of multiresistance. When ketolides were prescribed to all ages, resistance to them reached 10% after >15 years, while it took >40 years when they were prescribed only to adults. In the scenario according to which new antibiotics totally replaced former antimicrobials, the β-lactam resistance rate was limited at 70%.

Conclusions: In a context of widespread vaccination and rational use of antibiotics, innovative antibiotic, prescribed to all age groups, may have an added impact on multiresistant-strain dissemination in the population.

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Introduction

Bacterial resistance to antibiotics has become a major public health problem worldwide and direct consequences are now being documented in terms of increased morbidity, mortality and hospitalization costs [1–4]. Minimizing selection of pathogenic bacteria that have acquired mechanisms of resistance to antibiotics has become a top priority of international public health authorities. Streptococcus pneumoniae is a major cause of morbidity and mortality worldwide [5], and the main cause of community-acquired respiratory tract infections, e.g., otitis media, sinusitis and pneumonia. It is also a major pathogen responsible for invasive disease, like septicemia and meningitis.

In recent decades, antibiotic-resistant and multiresistant S. pneumoniae strains have spread throughout the community. In some European countries, such as France or Spain, penicillin non-susceptible strains represent >50% of all pneumococci, as do macrolide-resistant strains [6]. Multiresistance is also being observed more and more frequently. Despite this increased antibiotic resistance, very few new antibacterial classes have been developed by pharmaceutical companies over the last few decades [7].

The 7-valent pneumococcal conjugate vaccine (PCV7) markedly decreased resistant S. pneumoniae infections [8,9], but recent observations suggest the emergence of non-vaccine serotypes with high levels of resistance to all approved antibiotics [10].
Since antibiotic exposure is the main driving force behind the selection of strains harboring resistance to current drugs, a new class of antibiotic, without preexisting resistant strains, could help control the selection of resistance to older drugs, but emergence of new resistances to these new antibiotics is to be expected. The aims of this study were to examine that hypothesis using mathematical modeling, and to optimize strategies of new compound use by maximizing the time until emergence of new resistances and controlling the selection of resistant strains to preexisting antimicrobials.

This investigation was specifically conducted on ketolides, one of the newest antibiotic classes marketed for community-acquired respiratory tract infections.

**Materials and Methods**

We built a compartmental model of the transmission of *S. pneumoniae* in the French community under multiple antibiotic exposures. The model was specific to different mechanisms of resistance and the natural history of pneumococcal colonization. Using this model, we explored the diffusion of colonizing resistant and multiresistant pneumococci, according to antibiotic exposure. The objectives were to anticipate trends in new ketolide-resistance selection and multiresistance selection. First, we describe the model structure, then detail the parameters and, finally, present the scenarios that were simulated.

**Model Description**

The model developed herein takes into account 3 different antimicrobial classes (P: penicillin as an example of β-lactams, M: macrolides and K: ketolides), PCV7, and pertinent parameters concerning the French situation taken from the literature (Table 1).

To reproduce the selection and spread of resistant bacteria in the community through interindividual transmission of *S. pneumoniae* strains, the population was divided into several groups or compartments. Compartments were selected with respect to age, vaccination status, colonization and antibiotic exposure (Figure 1). To take into account differences according to the hosts’ ages, the population was divided into 3 age classes corresponding to young children (0–2 years), older children (2–15 years) and adults (>15 years). Hosts entered the population at birth as young children non-carriers. Mortality rates were defined for each class. We assumed that a fraction of young children was vaccinated each year. Vaccinated individuals were protected against carriage of serotypes included in PCV7, but not against carriage of serotypes not included in it.

Independently of their carriage status, individuals could be exposed to β-lactams (penicillin) and macrolides, as they are the main antibiotic classes prescribed in France [11]; and ketolides as the antibiotic innovation. We assumed that individuals were exposed to only 1 antibiotic class at a time. Because prescription frequencies depend on age and antibiotic classes, the matrix $a_{ij}$ describes the rates of exposure as a function of age $i$ and antibiotic class $j$. Under antibiotic exposure, strains with higher levels of resistance can emerge through genetic events and be selected, thereby replacing the strains originally harbored in the host in whom colonization had not been eliminated.

**Parameters**

Parameter values are listed in table 1. We used demographic data from the French National Institute of Statistics and Economical Studies (INSEE) [12] for the birth rate ($\mu_0$) and mortality rates of each age class ($\mu_1$, $\mu_2$, $\mu_3$). Pneumococcal colonization was defined as decreasing with age [13,14]: 50% in young children (<2 years old), 30% in older children (2–15 years old) and 10% in adults (>15 years old). The contact matrix was considered to be symmetric. Values of the age-specific contact frequencies depend on age and antibiotic classes, the matrix $a_{ij}$ were numerically calibrated so that the prevalence of each antibiotic class was observed in the French population.

In the absence of antibiotic exposure, natural immunity was defined as inducing carriage clearance in colonized individuals, according to their age: on average, 2 months for young children...
(1/λ₁), 1 month for older children (1/λ₂) and 2 weeks for adults (1/λ₃) [15–18].

**Pneumococcal conjugate vaccine.** PCV7 protects against both invasive disease and pharyngeal colonization [8], and has been recommended for young children in several countries [19–21]. In France, about 30% of children <2 years old, corresponding to all children in day care, were vaccinated each year before 2006. In 2006, recommendations were extended to systematic vaccination of all children <2 years old [22]. Previously developed models, describing the effect of vaccination on pneumococcal carriage and resistance, predicted replacement of serotypes included in the vaccine by non-vaccine serotypes [23,24]. In our model, young children were vaccinated at rate v. Vaccinated children were assumed to have received 4 doses of PCV7 (including 1 booster dose), and thus to be fully immunized against serotypes included in PCV7. We assumed progressively increasing vaccination coverage between 2002 and 2004 from no-vaccination to vaccination of 30% of young children. After 2007, date of systematic vaccination of young children, we considered young children’s vaccine coverage to be 90%. Vaccine immunity (d/v) was supposed to last throughout childhood.

**Antibiotic-exposure parameters.** We considered the global antibiotic consumption regardless of indication of the drugs. We used annual global data published by the French National Health Insurance Agencies (CNAM–TS, CANAM) [25], covering >95% of the population, to evaluate prescription frequencies for the 3 antibiotic classes of interest for each age class. Consumption varied with age: children <5 years old had high rates of exposure, while prescriptions were quite low for adults. To be consistent with the objective of overall antibiotic-use reduction in France, a progressive and homogeneous decrease of global antibiotic exposure was assumed for all age groups (reaching 25%, 20% and 10% reductions over a 5-year period in young children, older children and adults, respectively). We investigated scenarios of ketolide exposure from the current status to their total replacement of old antibiotic classes, as developed below in detail.

**Treatment duration (1/γ) was assumed to be the same regardless of the antibiotic prescribed (8 days). We considered that the antibiotic effect on colonization (σ) started after an average of 4 days of exposure (1/ν) and that it depended on the colonizing strain’s level of susceptibility (see below).**

**Antibiotic resistance.** Three stages of *S. pneumoniae* penicillin-susceptibility, corresponding to increasing minimum inhibitory concentrations (MIC), were modeled [26]: susceptible (S₀), intermediate (I₀), and resistant (R₀), for strains with respective MIC <0.125, 0.125 ≤ MIC <2 and MIC ≥2 mg/mL. Furthermore,

![Figure 1: Schematized model structure.](image-url)
since S. pneumoniae’s macrolide-resistance acquisition by ribosomal structure modification can be viewed as an on/off mechanism [27], corresponding to resistant/susceptible bacteria, macrolide-susceptible and -resistant (SM and RM) strains were considered. Ketolides have a mechanism of action similar to that of macrolides [29], but RM strains are generally susceptible to ketolides (SK).

Importantly, the few documented examples of ketolide-resistant (RK) strains concerned those already resistant to macrolides. That observation suggests that ketolide resistance is conditioned by previous resistance to macrolides (cross-resistance) [29–31]. Therefore, macrolide resistance was considered to be a prerequisite for the later emergence of ketolide resistance, which was ascribed to all strains already RM. Consequently, all RK strains colonizing individuals in the model were also RM.

Emergence of resistance or an increase of its level was possible in colonized individuals exposed to antibiotics. Resistance levels evolved independently for penicillin and macrolides. Their rates of increased resistance (SP→IP, S→RP, I→→RP, and SM→RM) were numerically estimated so that simulation results agreed with the observed historical trends (before 1992) of emergence and diffusion of RP and RM strains (Φ). For ketolides, the same rates as those estimated for macrolides were used.

Multiresistance. Because pneumococcal strains can combine resistances to different antibiotics, 2 types of multiresistant strains were included and evaluated in the model: penicillin-macrolide-resistant strains (denoted: (IP/RP) and penicillin-macrolide-ketolide-resistant strains (IP/RP+RM+RK). Macrolide-ketolide-resistant (RM+RK) but penicillin-susceptible (SP) strains were not considered multiresistant strains, because of the macrolide-ketolide cross-resistance.

Model Fitting and Simulations

To validate model predictions, we used historical data on antibiotic consumption in France [32], and compared simulation results with pneumococcal resistance and multiresistance rates from 1992 to 1997 reported by the French National Center for Pneumococci (CNRP), which coordinates the national surveillance system of pneumococcal infections [33].

Initialization. We initialized the model with a population in which resistant pneumococcal rates were equivalent to the 2002 French situation [33] with >50% of IP/RP strains, >55% of RM strains and 0.1% preexisting RK strains [34–37].

Scenarios. We investigated the impact of ketolide exposure by simulating 3 scenarios (Figure 2). The first, Baseline, corresponded to no ketolide-use, i.e. the French situation before 2005. The second scenario, K-Adults, consisted of replacing 25%, 50%, 75% or 100% of IP/RP strains, >55% of RM strains and 0.1% preexisting RK strains [34–37].

To study RK emergence and selection, we estimated the times required for 1% or 10% of strains colonizing the population to become RK. Furthermore, we simulated the expected dynamics of multiresistance diffusion over the next 50 years, defined as resistance to at least penicillin and macrolides (IP/RP+RM), according to the scenarios.

Sensitivity analysis. A sensitivity analysis was undertaken to evaluate the dependence of model predictions (percentages of RK and multiresistant strains in the colonized population) on small variations of all input parameters (such as the contact matrix, antibiotic exposure levels, etc.). The parameters were allowed to vary within a range of their possible values. Latin Hypercube Sampling (LHS) was used and partial rank correlation coefficients (PRCC) were computed for all parameters [38].

Results

The historical time course of macrolide resistance, penicillin resistance and penicillin-macrolide multiresistance from 1992 to 1997 were well reproduced by model simulations (Figure 3). In a previous study using data from the United States, we showed that the simulated decrease over the first 3 years of mass vaccination was consistent in magnitude with the observed data, in terms of invasive disease incidence and in terms of RP [24].

Emergence of ketolide-resistant strains. In scenario K-Adults, ketolides were prescribed only to adults. Times to emergence of 1% and 10% RK pneumococcal strains in the population are presented in figure 4A for different levels of ketolide exposure. For up to 50% of ketolide use among adults, the mean emergence time of 10% RK exceeded 50 years. Under the hypothesis that ketolides effectively replaced high rate of the preexisting antimicrobials in total adult antibiotic consumption, it still took >35 years to reach 10% RK.

When ketolides were prescribed to all ages (scenario K-All), times before RK emergence were shorter (Figure 4B). When ketolides replaced at least 20% of all preexisting antibiotic consumption, it took <40 years to reach 10% RK, whereas for quasi-total (90%) ketolide replacement of former antibiotics, simulations predicted that 10% RK could be reached within a dozen years.

Dynamics of multiresistant strains. When the new drugs were prescribed only to adults (scenario K-Adults, figure 5A), the dissemination of (IP/RP+RM) multiresistant strains was barely slowed by ketolide use over the first few years. However, after RK emergence and selection, the diffusion of strains resistant to P, M and K (IP/RP+RM+RK) was accelerated with increased ketolide use, as a consequence of the co-selection of RM strains under ketolide exposure, owing to macrolide-ketolide cross-resistance. Twenty years after RK emergence and its selection in the population, no significant differences in the frequencies of carriage of multiresistant pneumococcal strains remained between the scenario using ketolides (K-Adults) and the one not using them (Baseline).

When ketolide prescriptions were extended to the entire population (scenario K-All, figure 5B), the dissemination of (IP/RP+RM) strains differed markedly according to the prescription level. The higher the ketolide-prescription rate, the lower the initial diffusion of (IP/RP+RM) strains, but the emergence of (IP/RP+RM+RK) multiresistant strains no longer slowed the resistance rise and even enhanced it. K-All (IP/RP+RM+RK) multiresistance appeared more rapidly than in K-Adults, and quickly encompassed all resistant strains. Importantly, when ketolides replaced >50% of current antibiotics, the simulations showed 2-step dynamics: first, a strong deceleration of (IP/RP+RM) strain dissemination; second, an acceleration, increasing with the rise of the replacement rate. In the extreme case in which ketolides totally replaced current antibiotics (100%), the maximum multiresistance level was reached after only 20 years. It should be noted that, when ketolides totally replaced preexisting antibiotics, the model predicted that multiresistant-strain diffusion would cease at ~70%, maintaining a reservoir of SP strains.

Sensitivity analysis. The most significantly sensitive parameters are listed in table 2. Concerning RK and multiresistance rate outcomes, the model was particularly sensitive to the frequencies and duration of antibiotic exposure: when either increased, the proportion of resistant strains increased. Moreover, the model was sensitive to the average time until treatment acts: when it decreased, resistance increased (PRCC ~0.6). Furthermore, the multiresistance rate was positively
sensitive to PCV7 use and negatively to the frequencies of antibiotic exposure in all age groups. Transmission rates between and within age-classes did not significantly influence the global resistance rates.

Discussion

The results of this simulation study suggest that new antimicrobial use could markedly slow the diffusion of \( (I_P/R_P)^+R_M \) multiresistant strains. During the initial period after ketolide introduction, their wider use was associated with a slower progression of multiresistance. The model predicted it would take 15 years to reach 10% of RK in the population when the new drug was prescribed to all ages, and 40 years, when it was prescribed only to adults. In all cases, except when ketolides replaced all other drugs in the community, bacterial resistance increased, eventually reaching high rates of strains multiresistant to the 3 drugs \( [(I_P/R_P)^+R_M^+R_K] \), due to co-selection of \( R_M^+R_K \) strains under ketolide exposure. When all drugs but ketolides were discontinued, the resistance to \( \beta \)-lactams was contained at ~70%.

Obviously, this unrealistic strategy raises contradictory economic questions: should new drugs be developed in order to preserve old drug efficacy?

Current trends in \textit{Streptococcus pneumoniae} epidemiology are difficult to anticipate: first, the epidemiological situation is complex because of multiresistant strains and multi-exposures to antibiotics; second, important changes have occurred recently with the availability and wide use of PCV7 and the measures undertaken to reduce antibiotic prescriptions. In the past, models helped to analyze probable changes, predicting changes in serotypes after vaccine use [23,39], investigating the associations between antibiotic use and RP [40] or explaining why \( \beta \)-lactam resistance presented a bimodal shape [41]. Here, using a model taking these recent changes into account, we investigated how the use of a new antimicrobial can modify the resistance patterns in the community.

As non-vaccine strains are currently less resistant to macrolides [9,42], immunization may delay the occurrence of RK. For that reason, and also because wide vaccine coverage might select for non-vaccine-serotype multiresistant stains [10], PCV7 was included in this study. No precise estimate exists for the duration

Figure 2: Description of epidemiological pertinent facts and scenarios, showing how parameters of antibiotic-exposure rates (P, penicillin; M, macrolides; K, ketolides) and PCV7 immunization rates evolved in the simulations. (A) Scenarios of ketolide introduction. Baseline: K are never prescribed; K-Adults: K are prescribed only to adults, to replace P and M; K-All: K are prescribed to all individuals, to replace P and M. In the two K-scenarios, the level of replacement varies from 0% to 100%. (B) Timeline of relevant epidemiological changes included in the model. doi:10.1371/journal.pone.0002089.g002
of PCV7-induced immunity as the vaccination program started only a few years ago. In our model, we assumed that PCV7-induced immunity would last throughout childhood. A shorter (longer) duration of immunity would be nearly equivalent to predictions of immunizing a smaller (greater) portion of young children.

The model and results presented here are specific to the introduction of ketolides into a population exposed to high rates of penicillin and macrolide use. First, the study was restricted to penicillin, macrolides and ketolides, excluding other antibiotics, such as quinolones or cyclines. In most countries, as in France, β-lactams and macrolides are the most frequently prescribed antimicrobial classes. Moreover, *S. pneumoniae* resistance to them has reached high percentages in several Western countries and become a major public health concern [11]. In contrast, fluoroquinolone and cycline antibiotic resistances remain quite low [35] and are not, at this time, a worrisome problem in terms of clinical failure in Europe and the US. For those reasons, we focused on the control of RM and RK and multiresistance. Considering other antibiotics and *S. pneumoniae* resistance to them in the model would have had little to no effect on the predicted changes in the main resistance patterns in the community in the near future.

Second, we described the pathway to RK based on current observations: RK can be developed only in RM strains. This assumption effectively limits the selection for RK to strains already RM, so that progression to resistance is slower than in a setting in which resistance to the new drug would appear independently of others. The evolution of resistance to such a drug could easily be analyzed in the same setting: it is expected that, in the same context, the emergence of a new resistance would be faster but the selection slower.

Third, the efflux-pump resistance mechanism, which endows *S. pneumoniae* with low-level RM, was not considered in the model. Indeed, for macrolides, the model developed herein was based on the French situation. In France, the vast majority of RM isolates have acquired a ribosomal modification resistance mechanism [35]. Moreover, the efflux-pump mechanism does not impair ketolide activity, while the ribosomal modification mechanism has marked impact in terms of selective pressure on both RM and RK. Consequently, in the French context, adding the possibility of 3
different levels of macrolide-susceptibility (susceptible, low-level resistant, and highly resistant) would have limited influence on the predictions.

Fourth, the model could also be used in a setting with lower exposure to penicillins and macrolides. In that case, initial \( R_P \) and \( R_M \) rates would also be lower, and the emergence and selection of new resistances would be expected to slow, and the impact on multiresistance trends of a new antibiotic to be delayed.

In the scenarios, we assumed a sudden replacement of part of current antibiotics by the innovative agent, while it could be more progressive over time in real life. The simulated situation constitutes a worst-case scenario for the selection of \( R_K \).

Figure 5: Impact of ketolide prescriptions on the emergence and selection of multiresistant \( S. \) pneumoniae. Evolutions of multiresistant-strain carriage as a function of time are shown. These simulations were carried out according to the 2 scenarios of K prescriptions: (A) K-Adults: ketolides were only used by adults after 2007, with 4 different replacement rates of adult antibiotic prescriptions: 0% (+); 25% (o); 75% (+); and 100% (-). (B) K-All: K prescriptions were extended to the entire population (including children) with the same replacement rates as previously.

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Higher level of details could be achieved, for instance using to their inclusion in the PCV7 rather than individually modeled. A considered in this study. Lastly, serotypes were grouped according with several strains having different resistance patterns was not (figure 4). Second, the possibility of colonization (and transmission) resistant strains depending on the different drug-use scenarios we focused here on the time needed to select and spread these stronger selective advantage. However, once resistance has emerged, resistant strains have a nity for sensitive strains to become resistant is decreased; antibiotics eliminate sensitive strains more quickly, the opportu- took longer and resistance became more prevalent). When acts (when was shorter, the mean time for resistance selection resistant strains increased); and the mean time until treatment would not be affected by these latter choices. At present, few RK strains exist in the community [37]; therefore, an epidemiologically meaningful resistance level in the population. The model uses several simplifications regarding the diversity of S. pneumoniae ecology and the results presented should be interpreted in the context of the following 3 limitations. First, our simulation did not examine the time required for pneumococci to acquire a new RK mechanism, but rather that needed to reach an epidemiologically meaningful resistance level in the population. At present, few RK strains exist in the community [37]; therefore, we focused here on the time needed to select and spread these resistant strains depending on the different drug-use scenarios (figure 4). Second, the possibility of colonization (and transmission) with several strains having different resistance patterns was not considered in this study. Lastly, serotypes were grouped according to their inclusion in the PCV7 rather than individually modeled. A higher level of details could be achieved, for instance using serotype-specific duration of carriage or immunity to S. pneumoniae [43]. However, the main conclusions regarding the impact of ketolides on the emergence and progression of multiresistance would not be affected by these latter choices.

In our sensitivity analysis, the parameters that caused the largest changes in results were: the frequencies and duration of antibiotic exposure (when they increased, the proportion of resistant strains increased); and the mean time until treatment acts (when was shorter, the mean time for resistance selection took longer and resistance became more prevalent). When antibiotics eliminate sensitive strains more quickly, the opportu- nity for sensitive strains to become resistant is decreased; however, once resistance has emerged, resistant strains have a stronger selective advantage.

It has been suggested [44,45] that the transmissibility of resistant strains could be lower than that of susceptible strains because resistance acquisition could imply a fitness cost. We did not include any fitness difference in our model. In terms of prediction, we would expect that introducing fitness cost for resistant strains, which increases with multiresistance, would delay the second-wind rise of multiresistance in non-vaccine strains. To date, however, biological evidence on this possibility at the human level is sorely lacking [46].

Recently, the US Food and Drug Administration approved a new label for ketolides, removing specific indications (acute bacterial sinusitis and acute exacerbation of chronic bronchitis) and adding warnings and contraindications [47]. This new labeling was incited by a rare hepatitis linked to ketolide use [48,49]. Consequently, the ketolide-use benefit predicted by the model should be balanced by the incidence of severe adverse effects. At this point, precise quantification of the costs, morbidity and mortality associated with pneumococcal antibiotic resistance becomes urgent [50–52]. Modeling, complemented with such data, should enable estimation of benefits and risks and should become a real support for public health decision-making.

Today, the only available ketolide drug, telithromycin, is approved for use only in adults, although children represent a large portion of antibiotic users. However, other new ketolide-class antibiotics, with the same activity as telithromycin, like cethro-myacin, are being developed [53,54] and could be prescribed to children in the future. In that context, the model predictions could show how the use of such an antibiotic innovation could impact the global pattern of resistance, at the population level.

We want to underline that the main pathways for resistance control remain the global reduction of antibiotic consumption, generalized PCV7 immunization of young children, and cautious and appropriate use of antibiotics at the individual level. While the results of this study emphasize the potential benefits of a partial replacement of currently used antibiotics by a new antibiotic class, our conclusion is not to encourage the wide use of ketolides but to demonstrate that innovation could be a complementary way to reduce antibacterial resistance at the community level.

According to our model, restriction of ketolide prescriptions to adults, even at a high replacement rate, would have a limited effect on the global resistance pattern and would mainly lead to clinical success. When extended to children, ketolide use would be associated with only a short-term positive effect. Using a model that included age-structured population and specific resistance and multiresistance mechanisms (cross-resistance and co-resistances), our findings are in agreement with the situation described by Wang and Lipsitch [55]: upgrading antibiotic use within a class (fluoroquinolones vs macrolides/ketolides here) would lead to a trade-off between high prevalence of highly multiresistant strains and successful treatment. Facing the choice between enhancing therapeutic success and controlling global resistance, these models can be useful for decision-makers.

In conclusion, our simulation results predict that in a population with widespread use of PCV7 and rational use of antibiotics, antibiotic innovation, prescribed to all age groups, particularly children, could have an added impact on multiresistance rates: the more the new drug is prescribed, the slower multiresistance would diffuse. However, at the same time, the more this new drug is used, the faster new multiresistant pneumococcal strains (l/Rp+Rk strains here) would be selected.

Paradoxical as it may seem, to prevent community-wide dissemination of strains simultaneously resistant to all available antibiotics, the optimal solution for the next 20-years, would be to

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**Table 2. Sensitivity analysis of the frequency of ketolide-resistant or multiresistant strains in the colonized population.**

| Parameter                                                                 | Variable | PRCC* |
|---------------------------------------------------------------------------|----------|-------|
| Frequency of ketolide-resistant strains as output                          |          |       |
| Frequency of ketolide exposure in adults (>15 years)                       | 4.22     | 0.3   |
| (Duration of antibiotic exposure)−1                                       | γ        | −0.7  |
| (Time until treatment acts)−1                                             | γ        | 0.6   |
| Probability of S→R incremental resistance of macrolide/ketolide          | p4       | 0.22  |
| Frequency of multiresistant strains as output                              |          |       |
| Frequency of exposure                                                     |          |       |
| β-Lactam in children (2–15 years)                                         | 12       | 0.1   |
| β-Lactam in adults (>15 years)                                            | 13       | 0.2   |
| Ketolide in young children (<2 years)                                     | 3.1      | 0.1   |
| Macrolide in adults (>15 years)                                           | 2.2      | 0.1   |
| (Duration of antibiotic exposure)−1                                       | γ        | −0.6  |
| (Duration of colonization in adults)−1                                     | γ        | 0.1   |
| (Time until treatment acts)−1                                             | γ        | 0.6   |
| Vaccination rate                                                          |          | −0.2  |

*Partial rank correlation coefficient (PRCC) indicates the degree of monotonicity between a specific input variable and a particular outcome variable. The sign of the PRCC indicates the qualitative relationship between input and output variables. The magnitude indicates the importance of uncertainty in estimating the value of the input variable in contributing to the imprecision in predicting the value of the outcome variable.

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switch antibiotics totally, preserving one or more of the older drugs for which non-resistant strains still exist.

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References

1. Davis SR (2000) The state of antibiotic resistance surveillance: an overview of existing activities and new strategies. Mil Med 165: 35–39.
2. Tomas A (1997) Antibiotic resistance in Streptococcus pneumoniae. Clin Infect Dis 25: S85–S88.
3. Levy SB, Marshall B (2004) Antibacterial resistance worldwide: causes, challenges and responses. Nat Med 10: S122–S129.
4. Feikin DR, Schuchat A, Kolczak M, Barrett NL, Harrison LH, et al. (2000) Mortality from invasive pneumococcal pneumonia in the era of antibiotic resistance. 1993–1997. Am J Public Health 90: 223–229.
5. Obaro SK, Adegbola RA, Banya WA, Greenwood BM (1996) Carriage of pneumococci after pneumococcal vaccination. Lancet 348: 271–272.
6. European Antimicrobial Resistance Surveillance System (EARS) (2006) Annual Report 2005. Available: http://www.rivm.nl/cnms/; Home; Cnms; htm; Monitoring; reports.
7. Cassel GH, Mekalanos J (2001) Development of antimicrobial agents in the era of new and reemerging infectious diseases and increasing antibiotic resistance. JAMA 285: 601–603.
8. Whitney CG, Farley MM, Hadler J, Lum LM, Cuny GD, et al. (2003) Decline in invasive pneumococcal disease after the introduction of protein-poly saccharide conjugate vaccine. N Engl J Med 348: 1737–1746.
9. Kyaw MH, Linfield E, Hadler J, Lum LM, Cuny GD, et al. (2006) Effect of introduction of the pneumococcal conjugate vaccine on drug-resistant Streptococcus pneumoniae. N Engl J Med 354: 1453–1463.
10. Pichichero ME, Casey JR (2007) Emergence of a multiresistant serotype 19A pneumococcal strain not included in the 7-valent conjugate vaccine as an otopathogen in children. JAMA 298: 1772–1778.
11. Riedel S, Beckmann SE, Helmling KP, Richter SS, Garcia-de-Lomas J, et al. (2007) Antimicrobial use in Europe and antimicrobial resistance in Streptococcus pneumoniae. Eur J Clin Microbiol Infect Dis 26: 485–490.
12. Institut National de la Statistique et des Etudes Economiques (INSEE) (2007) Indices et series statistiques. Demographie. Available: http://www.insee.fr/fr/indexfr/.../home/home_page.asp. Accessed January 2005.
13. Reyes-Yochay G, Raz M, Dagan R, Porat N, Shainberg B, et al. (2004) Nasopharyngeal carriage of Streptococcus pneumoniae by adults and children in community and family settings. Clin Infect Dis 30: 632–639.
14. Hendley JO, Sande MA, Stewart PM, Gwaltney JM (1975) Spread of Streptococcus pneumoniae in Families. I. Carrier rates and distribution of types. J Infect Dis 132: 55–61.
15. Ekdahl K, Ahlinder I, Hansson HB, Melander E, Molstad S, et al. (1997) beta-lactam resistance in Streptococcus pneumoniae transmissible diseases) relative to vaccination by heptavalent pneumococcal conjugate vaccine. J Infect Dis 132: 55–61.
16. Decousser JW, Ovetchkine P, Collignon A, Chaplain C, Estrangin E, et al. (2004) Multicentre study of the molecular epidemiology, serotypes and serogroup-related differences in observed durations of nasopharyngeal carriage of penicillin-resistant pneumococci. J Clin Microbiol 43: 948–952.
17. (1993) Advice of the French Superior Council on Public Health (section on transmissible diseases) relative to vaccination by heptavalent pneumococcal conjugate vaccine (Prevenar). Meeting of March 18, 2002. Arch Pediatr 9: 982–984.
18. Centers for Disease Control and Prevention (CDC) (2000) Preventing pneumococcal disease among infants and young children. Recommendations of the Advisory Committee on Immunization Practices for use of PCV7 in the USA. MMWR 49: R1–R30.
19. Centers for Disease Control and Prevention (CDC) (2004) Updated recommendations for use of pneumococcal conjugate vaccine. MMWR 53: 177–178.
20. Institut de Veille Sante´ (InVS) (2006) Calendrier vaccinal 2006. Bulletin Epidemiologique Hebdomadaire 29: 30–212.
21. Temme I, Guillomet D, Boelle PY (2004) Short- and long-term effects of pneumococcal conjugate vaccination of children on penicillin resistance. Antimicrob Agents Chemother 48: 2206–2213.
22. Temme I, Guillomet D, Boelle PY (2006) Pneumococcal resistance in the postvaccine era. Pediatr Infect Dis J 25: 382–383.
23. French National Health Insurance. French Surveillance of Antimicrobial Consumption Program, CNAM-TS and CANAM. Available: http://www.anel.fr.

Author Contributions

Conceived and designed the experiments: PB LO LT DG. Performed the experiments: LO. Analyzed the data: LO. Contributed reagents/materials/analysis tools: LO LT RD. Wrote the paper: PB LO LT DG. Other: Performed the modeling and simulation parts: LO. Provided the French antibiotic resistance data: EV. Participated in identifying pertinent parameters for an adequate construction of the model: HD RL.

26. Hakenbeck R, Grebe T, Zahnner D, Stock JB (1999) beta-lactam resistance in Streptococcus pneumoniae: penicillin-binding proteins and non-penicillin-binding proteins. Mol Microbiol 33: 673–678.
27. Wierzchowski AK, Boyd D, Mulvey MR, Hoban DJ, Zhanel GG (2005) Expression of the merE gene encoding the macrolide efflux pump protein increased in Streptococcus pneumoniae with increasing resistance to macrolides. Antimicrob Agents Chemother 49: 4635–4640.
28. Lederglo R (2002) Will resistance to ketolides develop in Streptococcus pneumoniae? J Infect 44: 11–16.
29. Goldstein F, Vidal B, Kitiz MD (2003) Telithromycin-resistant Streptococcus pneumoniae. Emerg Infect Dis 11: 1489–1490.
30. Farrell DJ, Jenkins SG, Beek JR, Patel M, Lavin BS, et al. (2005) Emergence and spread of Streptococcus pneumoniae with erm(B) and mer(A) resistance. Emerg Infect Dis 11: 831–836.
31. Lederglo R (2001) Overcoming antimicrobial resistance: profile of a new ketolide antibacterial, telithromycin. J Antimicrob Chemother 48: 9–23.
32. Sonnent A, Seremet G, Boelle PY, Tafflet M, Berne© C, et al. (2004) No significant decrease in antibiotic use from 1992 to 2000, in the French community. J Antimicrob Chemother 54: 524–528.
33. Gregoire P, Fremaux A, Sissia G, Zberrane S, Zberrane C, Zberrane S (1992–2000) Centre National de Référence des Pneumocoques. Rapports d’activité.
34. Varon E, Gutmann L (2003) Centre National de Réference des Pneumocoques. Rapport d’activité 2003: Epidémiologie 2002.
35. Varon E, Gutmann L (2005) Centre National de Référence des Pneumocoques. Rapport d’activité 2005: Epidémiologie 2004.
36. Decousser JW, Ovetchkine P, Collignon A, Chaplain C, Estrangin E, et al. (2004) Multicentre study of the molecular epidemiology, serotypes and serogroup-related differences in observed durations of nasopharyngeal carriage of penicillin-resistant pneumococci. J Clin Microbiol 42: 862–864.
37. Malley R, Srivastava A, Lipsitch M, Thompson CM, Watkins C, et al. (2006) Antibody-independent, interleukin-17A-mediated, cross-serotype immunity to pneumococci in mice immunized intranasally with the cell wall polysaccharide. Infect Immun 74: 2187–2195.
38. Anderson DJ (2006) The biological cost of mutational antibiotic resistance: any practical conclusions? Curr Opin Microbiol 9: 461–465.
39. Trzcinski K, Thompson CM, Gilby AM, Dowson CG, Lipsitch M (2006) Incremental increase in fitness cost with increased beta-lactam resistance in pneumococci evaluated by competition in an infant rat nasal colonization model. J Infect Dis 193: 1296–1302.
40. Rosen DE, McGee L, Levin BR, Klugman KP (2007) Fitness costs of fluoroquinolone resistance in Streptococcus pneumoniae. Antimicrob Agents Chemother 51: 412–416.
41. U.S. Food and Drug Administration (FDA) (2007) Updated label for Ketek (telithromycin). Available: http://www.fda.gov/cder/drug/infopage/telithromycin/Accessed May 2007.
42. Ross DB (2007) The FDA and the case of Ketek. N Engl J Med 356: 1601–1604.
43. Malley R, Gooi E, Kwaeder S, Jenkins J, Galson S (2007) Ketek the FDA perspective. N Engl J Med 356: 1675–1676.
44. Desenclos JC, Guillomet D (2004) Consequences of bacterial resistance to antimicrobial agents. Emerg Infect Dis 10: 759–760.
51. Moroney JF, Fire AE, Harrison LH, Patterson JE, Farley MM, et al. (2001) Clinical outcomes of bacteremic pneumococcal pneumonia in the era of antibiotic resistance. Clin Infect Dis 33: 797–805.
52. McGowan JE Jr (2001) Economic impact of antimicrobial resistance. Emerg Infect Dis 7: 286–292.
53. Talbot GH, Bradley J, Edwards JE Jr, Gilbert D, Scheld M, et al. (2006) Bad bugs need drugs: an update on the development pipeline from the Antimicrobial Availability Task Force of the Infectious Diseases Society of America. Clin Infect Dis 42: 657–668.
54. Jorgensen JH, Crawford SA, McElmeel ML, Whitney CG (2004) Activities of cethromycin and telithromycin against recent North American isolates of Streptococcus pneumoniae. Antimicrob Agents Chemother 48: 603–607.
55. Wang YC, Lipsitch M (2006) Upgrading antibiotic use within a class: tradeoff between resistance and treatment success. Proc Natl Acad Sci U S A 103: 9655–9660.