Factors triggering queen executions in the Argentine ant

Silvia Abril & Crisanto Gómez

Competition among queens in polygynous societies may result in queen executions or conflicts over personal reproduction. Understanding the factors that mediate the executions of ant queens should provide insight into how queen numbers are regulated in polygynous insect societies. The Argentine ant is a widespread invasive species that displays secondary polygyny, and workers execute 90% of their nestmate queens each spring. In this study, we investigated: (1) whether ambient temperature, queen number, and protein deprivation have an effect on queen executions and (2) whether workers select the queens slated for execution based on their cuticular hydrocarbon (CHC) profiles. We found that the percentage of queens executed was positively correlated with temperature and queen number but that protein deprivation did not play a role. As for queen fate, the levels of some CHCs were higher in surviving queens. One of these CHCs is associated with queen productivity (i.e. egg-laying rate and ovarian index) suggesting that workers execute the least productive queens. Our findings suggest that chemical cues related to fertility signaling may mediate queen executions in Argentine ants.

Among social insects, ants stand out as a group in which polygyny (i.e., the presence of several queens in the same colony) may be the predominant social structure. Polygyny in ants can be primary or secondary. Primary polygyny, the less frequent of the two, occurs when multiple queens without workers collaboratively found a new colony (pleometrosis), which means that the foundresses co-exist during the colony’s initial stages. Primary polygyny can also occur when multiple queens accompanied by workers found a new colony after budding from larger colonies. Secondary polygyny is more common and occurs when monogynous colonies—established either via claustral colony founding or via budding from larger polygynous colonies—become polygynous after gyne acceptance and/or after colony fusion. Executions may then be used to reduce queen numbers and thus maximise egg production.

Polygyny can lead to reproductive competition among queens, which can result in queen executions or conflicts over personal reproduction. Queen-queen competition occurs in both primary and secondary polygyny but manifests itself differently in the two systems. In primary polygyny, there are fights among queens or between queens and workers after which a single queen remains; all the others are executed. In secondary polygyny, in contrast, there is a decrease in queen fecundity as queen numbers increase. Among ants, this negative effect of queen number on the fecundity of individual queens has frequently been observed in several species. According to Vargo, there are two likely mechanisms by which increased queen numbers could diminish individual fecundity. First, if there are more queens sharing the same given amount of resources, each queen might receive less food and thus create fewer eggs. Second, there might be pheromone-mediated reproductive competition among queens.

The Argentine ant, (Linepithema humile, Mayr), an invasive species found across the globe, displays secondary polygyny and carries out a dramatic number of queen executions. Every spring, workers execute about 90% of their nestmate queens, resulting in a 7% loss in the colony’s total biomass. Previous studies examined if physiological factors such as mass, egg-laying rate, and the quantity of sperm stored differed between surviving and executed queens, but no relationship was found. Reuter et al. also explored whether relatedness played a role in queen executions; they found that workers were, on average, equally related to surviving and executed queens. Queen executions therefore do not seem to constitute a mechanism for increasing the inclusive fitness of workers. That said, Inoue et al. showed that genetic diversity in Argentine ant workers significantly decreased from May to September and suggested that queen executions are a contributing factor.

In the Argentine ant, as in other ant species, queen pheromones can inhibit gyne development. The spring execution of queens seems to respond to the workers’ need to lower queen pheromone levels so that larvae can undergo sexualisation. Consequently, the concentration of queen pheromones in colonies is thought to play a
Effect of temperature and queen number on queen executions. A significantly higher percentage of queens were killed in the colonies kept at 28°C and 32°C than in the ones kept at 24°C (GLMM: t = 2.47, p < 0.01; results of paired t-tests: 28°C and 32°C vs. 24°C—p < 0.05) (Fig. 1). Also, a greater percentage of queens were killed in polygynous nests with eight queens than in monogynous nests (GLMM: t = 1.96, p < 0.05; results of paired t-tests: monogynous vs. polygynous with eight queens—p < 0.05) (Fig. 1).

In the field, the number of queens per two litres of nest soil in May was markedly different between the contact zone and the invaded zone (mean queen number in early May: contact zone = 3.50 ± 10.8; invaded zone = 1.8; GLMM: t = −3.40, p < 0.01). It also differed significantly over time—it grew smaller as the weeks went by (GLMM: t = −3.50, p < 0.01). In addition, there was a significant interaction between these two factors (area and week). This result indicated that the decrease in the number of queens differed between the two areas across time and that, more specifically, it was more dramatic in the contact zone (GLMM: t-value = −3.17; p < 0.05) (Fig. 2a). The slopes of the regression lines for the mean number of queens present per week in May were markedly different between the two areas (contact zone: y = −3.17x + 12.42; invaded zone: y = −0.4x + 3.08), which underscored that the decline was more dramatic in the contact zone. Furthermore, although the two areas displayed differences in the number of queens present in early May, these differences had disappeared by the end of May (mean ± SD number of queens in two litres of nest soil: contact zone = 1.5 ± 1.38; invaded zone = 1.6 ± 1.21).
Given that the decrease in queen numbers was most likely due to queen executions, we estimated that 86.15% of queens were eliminated in the contact zone, as compared to 43.53% of queens in the invaded zone (Fig. 2b).

Effect of protein deprivation on queen executions. Protein deprivation did not play a significant role in queen executions because there was no difference in the percentage of queens executed by colonies given a protein-rich diet versus by colonies given a protein-free diet (GLMM: $t = -0.012$, $p = 0.989$). Interestingly, larvae in the latter colonies did not die. Instead, they pupated earlier than larvae from the former colonies. Consequently, the workers that emerged from these pupae were smaller than average (pers. obs.).

Relationship between CHC profiles and queen fate. There were significant differences in the CHC profiles of executed versus surviving queens (Figs 3 and 4). Two discriminant functions explained most of the variance (global model: Wilk’s $\lambda = 0.031$, $F = 5.9$, $p < 0.0001$; 85.45% correct classification; first function: $R = 0.93$, Wilk’s $\lambda = 0.031$, $\chi^2 = 176.50$, $p < 0.0001$; second function: $R = 0.841$, Wilk’s $\lambda = 0.238$, $\chi^2 = 73.10$, $p < 0.0001$). The second discriminant function fully differentiated between them: executed queens were clustered at one end of the spectrum and surviving queens at the other (Fig. 4). The main CHCs responsible for these differences were two di-methyl alkanes (5,11-diMeC$_{29}$ and 5,11-diMeC$_{33}$), a single alkene (C$_{31}$:1), and one mono-methyl alkane (5-MeC$_{31}$). Given that the relative quantities of these four compounds differed between executed and surviving queens, we examined their absolute quantities. Overall, surviving queens had higher quantities of 5,11-diMeC$_{29}$ and 5,11-diMeC$_{33}$ than executed queens (5,11-diMeC$_{29}$: GLMM: $t = -4.09$, $p < 0.005$; 5,11-diMeC$_{33}$: GLMM: $t = -3.18$, $p < 0.01$) (Fig. 5; Table 1).

Discussion
This study has revealed new information about how queen numbers are regulated in the Argentine ant, a globally invasive species that displays secondary polygyny. We found that higher queen numbers and higher temperatures resulted in a greater percentage of queen executions. These results support that queen pheromone concentration plays a key role in regulating queen numbers, and, therefore, in precipitating queen executions$^{27}$. According to
Vargo and Passera²⁷, in natural populations of the Argentine ant, queen numbers are regulated by controlling the levels of inhibitory pheromone present in the colony. In this way, the development of gyne-potent larvae—which are present throughout the year—is inhibited for most of the year because more queens means more queen
In the spring, the stimulatory effect of overwintering increases the tendency of workers to rear gynes from gyne-potent larvae; it also enhances the developmental potential of larvae. The massive execution of queens has been observed in invasive populations of the Argentine ant across the globe. The execution period coincides with the period of larva sexualisation, suggesting that queens must be executed to decrease levels of the inhibitory queen pheromone and thus allow gyne production. Gyne40

The hypothesis is that temperature influences the emission of queen pheromones in ants. However, based on previous studies, it seems reasonable to hypothesise that temperature influences the emission of queen pheromones in the same way. Therefore, we posit that the increase in temperature led to an increase in queen pheromone levels, which led, in turn, to an increase in queen executions.

In contrast, protein deprivation does not seem to be an important factor in queen executions. In the Argentine ant, proteins provide amino acids that are predominately used to enhance larval growth and queen egg laying. Protein deprivation does not seem to be an important factor in queen executions. In the Argentine ant, proteins provide amino acids that are predominately used to enhance larval growth and queen egg laying.

In contrast, protein deprivation does not seem to be an important factor in queen executions. In the Argentine ant, proteins provide amino acids that are predominately used to enhance larval growth and queen egg laying. In contrast, protein deprivation does not seem to be an important factor in queen executions. In the Argentine ant, proteins provide amino acids that are predominately used to enhance larval growth and queen egg laying. In contrast, protein deprivation does not seem to be an important factor in queen executions.
protein is in high demand by queens and larvae—such as during the execution period—workers would eliminate supernumerary queens to enhance larval development. Interestingly, we found that protein deprivation did not provoke queen executions in the Argentine ant, which suggests that factors unrelated to diet are in operation.

With regards to queen fate, our results are consistent with those of a recent study, which suggested that Argentine ant workers cull queens based on their CHC profiles. We found that surviving queens bore different CHC profiles than executed queens, and it appears that two di-methyl alkanes were the main compounds responsible for these differences. Compared to executed queens, surviving queens bore significantly greater amounts of 5,11-diMeC₂₉ and 5,11-diMeC₃₃. Interestingly, Abril et al. also observed large amounts of these two di-methyl alkanes in the CHC profiles of queens sampled at the end of the execution period (i.e., surviving queens). This result indicates that, in the Argentine ant, it is likely that a queen's CHC profile determines her fate and that workers use the di-methyl alkanes 5,11-diMeC₂₉ and 5,11-diMeC₃₃ as signals in the culling process. Greater amounts of di-methyl alkane 5,11-diMeC₂₉ are associated with higher egg-laying rates and ovarian indices in Argentine ant queens, which suggests that workers could use this CHC to selectively kill the least productive queens, as occurs in Lasius niger. This latter species founds colonies via pleometrosis, and levels of the methylalkane 3MeC₃₁ and of the alkene C₃₁, compounds that are both correlated with queen productivity and maturity, were found to be higher in surviving queens than in executed queens. Later, it was discovered that 3MeC₃₁ was both a primer pheromone (i.e., affecting worker reproduction) and a releaser pheromone (i.e., affecting worker behavior, notably by increasing aggressiveness towards queens). Consequently, the di-methyl alkanes identified here could serve as releaser queen pheromones in the Argentine ant (i.e., queen pheromones that affect the behaviour of other nestmates) and thus increase worker aggressiveness towards queens.

In the Argentine ant, queens experiencing the same set of experimental conditions, whether monogynous or polygynous, displayed marked variation in their egg-laying ability. Similarly, when experimental conditions were polygynous as opposed to monogynous, the oviposition rates of queens shifted: there were always one or two main queens who laid the eggs, and the contribution of other queens was small to null. These results underscore that there might be some form of reproductive inhibition among queens. This hypothesis is supported by the fact that Argentine ant queens can regulate colony social organization using queen pheromones. It is thus possible that, in this species, reproductive competition occurs among a colony's queens and leads to the execution of the least productive individuals during the queen execution period.

Interestingly, Abril et al. found greater amounts of di-methyl alkanes 5,11-diMeC₂₉ and 5,11-diMeC₃₃ in young queens than in the executed queens reported in the present study (i.e. old queens). This finding suggests that the elimination of old queens to allow gyne production in the Argentine ant could serve as a mechanism to increase queen productivity in the colony. A previous study on the fecundity of Argentine ants found that the egg-laying of young queens was similar to that of the old ones. However, young queens in this study underwent all of their maturation in the laboratory and this may have influenced the process of fat accumulation, and, therefore, their fecundity. As the CHC profile reflects the hormonal state more than the current egg-laying rate, the results obtained in the present study provide new information about the biological reasons beyond the annual execution period in the Argentine ant.

Further research is necessary to assess whether the compounds mentioned above are, in fact, queen pheromones that help regulate queen numbers and, ultimately, whether they have a role in suppressing egg laying activity by fellow queens in polygynous situations. It is also important to identify the inhibitory queen pheromone present in this highly polygynous ant species. This information will allow us to gain a broader understanding of the social organization of polygynous ant species and inform strategies for managing Argentine ant invasions, potentially through the use of pheromones that induce colony self-destruction.

**Methods**

**Effect of temperature and queen number on queen executions.** We collected 156 queens from 36 different Argentine ant nests before the execution period (in early March and April) along the southern edge of the Gavarres Massif, near the village of Castell d’Aro (NE Iberian Peninsula) (41°49’N3°00’E). In this area, there are invaded and uninvaded cork oak secondary forests. In the invaded forests, Argentine ant colonies can cover up to 323 ha and mainly occur under stones. Here, we use Heller et al.’s definition of an Argentine ant colony: a group of interacting nests. From October to June, Argentine ant nests within a radius of 3–4 m aggregate in large clusters. Therefore, the nests sampled were at least 10 meters apart to ensure that they truly represented different colonies. In the laboratory, we created 12 monogynous colonies: each contained one queen and approximately 300 workers. We also created 24 polygynous colonies: 12 contained four queens, and 12 contained eight queens. Each polygynous colony had the same ratio of workers to queens—approximately 300 workers per queen. In these artificial colonies, the queen(s) and workers were from the same nest of origin. The colonies were housed in plaster structures (180 × 115 × 25 mm high) and were fed the Bhatkar diet and mealworms on a daily basis. To prevent the ants from escaping, the inner sides of the structure were coated with liquid PTFE (Fluon). The colonies were kept under conditions of 80% relative humidity.

To test the effect of temperature and queen number on queen executions, four monogynous colonies, four polygynous colonies with four queens, and four polygynous colonies with eight queens were placed in environmental chambers kept at one of three temperatures (±0.1°C): 24°C ± 0.1, 28°C ± 0.1, and 32°C ± 0.1. We chose these temperatures because we observed in previous studies that workers executed queens with varying frequency within this range. We checked the colonies daily for a period of three weeks to count the number of executed queens.

**Pattern of queen executions in the field.** We characterised queen execution patterns in the field, by comparing queen numbers during May 2013 between an area along the invasion front where the Argentine ant was still in contact with the native ant community (hereafter, contact zone) and an area almost exclusively
occupied by the Argentine ant (hereafter, invaded zone). These two areas were located in cork oak secondary forests in the NE Iberian Peninsula; the first was at the southern edge of the Gavarres Massif, near Santa Cristina d’Aro (41°48’51.71”N; 3°01’50.57”E), and the second was in the Cadiretes Massif, near Pedralba (41°47’31.53”N; 2°58’52.79”E). Colony queen numbers are known to differ between these two areas: they were found to be higher in the contact zone than in the invaded zone\(^\text{30}\).

In the Argentine ant, new colonies are founded via fission in mid-February or early March\(^\text{51–53}\). Consequently, the drop in queen numbers reported during the month of May could be caused by queen executions, as reported by Keller et al.\(^\text{20}\), since no other ecological or biological processes have been observed during this period that could decrease queen numbers\(^\text{31–33}\). For this reason, we assumed in this study that the decline in queen numbers in May was the result of queen executions.

To compare queen numbers during the execution period in these two areas, we collected 2-litre soil samples from a total of 24 nests in the contact zone and 24 nests in the invaded zone. To ensure sample independence, the nests sampled were at least 10 m apart. Samples were taken weekly during the whole month of May (six nests were sampled in each area every week). We assumed that the execution period had ended when queen numbers remained stable for two consecutive weeks, which had occurred by the last week of May. All the queens present in these samples were counted \textit{in situ} in the field by manual extraction of the queens present in each sample.

The contact zone and the invaded zone were ±1 km apart and had similar environmental characteristics, which meant that differences in queen numbers were unlikely to be related to abiotic differences between the areas.

**Effect of protein deprivation on queen executions.** To investigate the effect of protein deprivation on queen executions, we collected 12 nests before the execution period (in early April) from the same location as for the temperature/queen number experiment. We created 12 experimental colonies, each containing 15 queens and approximately 100 workers per queen. They also all contained larvae from several developmental stages (mainly from earlier stages so that larvae require more food from workers). Specifically, 0.1 g of larvae were weighed to the nearest 0.01 mg using a digital analytical balance. They were then carefully introduced into the experimental colonies. In these artificial colonies, the queen(s), workers and larvae were from the same nest of origin. Each experimental colony was housed in a rectangular plastic box (17 × 11 × 3 mm high) with a water source (a plastic tube filled with water that was wicked through a cotton ball). To prevent the ants from escaping, the inner sides of the box were coated with liquid PTFE (Fluon). The colonies were kept at 26 °C ± 0.1 and under conditions of 80% relative humidity; they were exposed to approximately 1 h of light per day (i.e., the time needed for feeding and cleaning). The twelve colonies were fed on a daily basis, but their diet differed: six received the Bhaktar diet, a sugar solution, and \textit{ad libitum} mealworms, and six received only the sugar solution. We checked the nests daily for three weeks to count the number of executed queens.

**Relationship between CHC profiles and queen fate.** We extracted the CHCs of the queens from the artificial colonies used in the temperature/queen number experiment. First, we collected 19 executed queens that had remained in the colony for 0–24 hours until their corpses were found. Second, we collected 19 queens that had survived the three weeks of observations. As a control, we also sampled 24 queens before the execution period (in early April) from the same location as for the temperature/queen number experiment. All the samples were frozen (−20 °C) until CHC extraction could occur. The queens’ gasters were eliminated to prevent any contamination by the Dufour gland. The samples were thus composed of just their heads, thoraces, and legs. We immersed the corpses in 50 μl of dichloromethane (GC grade) for 10 min. They were then removed, and the resulting extract was stored at −20 °C until further analysis. The extracts were evaporated and then redissolved in 50 μl of dichloromethane containing eicosane (8 ng/μl) as an internal standard. Next, 1 μl of each extract was injected into a gas chromatography (GC; Agilent 7820 A Series, Agilent Technologies, USA) equipped with a HP-5 capillary column (30 m × 0.32 mm × 0.25 μm) and a flame ionization detector. Sample analysis was performed using helium as a carrier gas; the flow rate was set to 2 ml/min (34.9 cm/sec); an injection volume of 1 μl with a split ratio of 1:5 was used; and the inlet temperature was set to 275 °C. The following temperature program was used: 75 °C held for 0 min; an increase from 75°C to 200°C at a rate of 20 °C min⁻¹; then another increase from 200°C to 315°C at a rate of 5 °C min⁻¹; and 315°C held for 15 min. The FID detector was set to a temperature of 300 °C. CHCs were identified using standard alkanes, compound libraries, and Kovats retention indices. CHC quantities were determined using the eicosane area as the internal standard (ng per head and thorax [i.e., one queen]).

Because we were analysing the CHCs of executed queens that may have remained in their nests for as long as 24 hours before being collected, we conducted an experiment to determine if queen CHC profiles degraded over time. We collected 6 nests containing 24 queens from the same location as for the temperature/queen number experiment. We randomly assigned 12 queens to the experimental group and 12 queens to the control group. In the experimental group, queens were freeze killed, warmed back up to ambient temperature, and returned to their artificial colonies of origin for 24 hours. Their corpses were then frozen again until the CHC analysis could be conducted. In the control group, queens were left unharmed in their artificial colonies for 24 hours. They were then freeze killed, thawed, and frozen again until the CHC analysis could be conducted. We found no significant differences between the CHC profiles of experimental queens and those of control queens (R = 0.43, Wilks’ λ = 0.815, χ²₁₂ = 2.10, p = 0.73) (Fig. 4). Furthermore, the two groups did not differ in their total quantities of CHCs (GLMM t₁₄ = −1.54, p = 0.19) or the quantities of the two main CHCs responsible for the differences in the CHC profile between surviving and executed queens (5,11-diMeC₃₃: GLMM: t = 0.27, p = 0.79; 5,11-diMeC₃₅: GLMM: t = −0.54, p = 0.61). These results indicate that the CHC profiles of Argentine ant queens did not significantly degrade even if dead queens remained in the nest for 24 hours prior to being collected.
Statistical analyses. Generalised linear mixed models (GLMMs) were performed to compare the percentage of executed queens (binomial error distribution and logit link function) based on the ambient temperature, the number of queens present, and protein availability (presence/absence). We included colony as a random factor. When overall significant differences were detected, pairwise comparisons were performed using t-tests employing pooled standard deviation.

The numbers of queens in the contact zone and the invaded zone were analysed using GLMMs (Poisson error distribution and log-link function). The response variable was queen number. The area (contact zone or invaded zone) and time (specific week of May) were fixed factors, and location (Santa Cristina d'Aró or Pedralta) was a random factor. We used the function glmmPQL in the MASS package to carry out the GLMMs.

To examine the queens’ CHC profiles, we obtained the relative contribution of each CHC peak to the total. Only identified peaks whose mean relative quantities were above 2% in at least one of the groups of queens were used. Discriminant analyses were then performed to investigate how CHC profiles differed between executed and surviving queens. To identify the CHCs that characterised each group, principal component analysis (PCA) was used. We focused on the CHCs demonstrating the greatest differences—those whose factor loadings on the first axis had an absolute value > 0.70. We further investigated these data using GLMMs in which the response variable was the mass of a given peak in nanograms (calculated by comparing the area of the peak with that of the internal standard) and colony was a random factor. To ensure normality, all the variables were log-transformed prior to being used in the discriminant analyses and the PCA. In addition, Levene’s test was used to determine whether the variables displayed homogeneity of variance; only those that did were retained. We used the function line in the nlm package to carry out the GLMMs.

All the statistical analyses were performed using R

Data Availability

The datasets generated and analyzed during the current study are available from the corresponding author on reasonable request.

References
1. Keller, L. Queen Number and Sociality in Insects. (Oxford University Press, 1993).
2. Hölldobler, B. & Wilson, E. O. The number of queens: an important trait in ant evolution. Naturwissenschaften 64(1), 8–15 (1977).
3. Herbers, J. M. Ecological determinants of queen number in ants. In Queen Number and Sociality in Insects (ed. Keller, L.) 16–44 (Oxford University Press, 1993).
4. Fletcher, D. J. C. & Blum, M. S. Regulation of queen number by workers in colonies of social insects. Science 219, 312–314 (1983).
5. Fortelius, W., Rosengren, R., CheriX, D. & Chautems, D. Queen recruitment in a highly polygynous supercolony of Formica lobicriss (Hymenoptera, Formicidae). Oikos 67, 197–200 (1993).
6. Sundström, L. Queen acceptance and nestmate recognition in monogyne and polygyne colonies of the ant Formica truncorum. Anim. Behav. 53, 499–510 (1997).
7. Vásquez, G. M. & Silverman, J. Queen acceptance and the complexity of nestmate discrimination in the Argentine ant. Behav. Ecol. Sociobiol. 62(4), 537–548 (2008).
8. Elmes G. W. & Keller, L. Distribution and ecology of queen number in ants of the genus Myrmica. In Queen Number and Sociality in Insects (ed. Keller, L.) 16–44 (Oxford University Press, 1993).
9. Bartz, S. H. & Hölldobler, B. Colony founding in Myrmecocystus mimicus Wheeler (Hymenoptera: Formicidae) and the evolution of foundress associations. Behav. Ecol. Sociobiol. 10, 137–147 (1982).
10. Rissing, S. W. & Pollock, G. B. Social interaction among pleometrotic queens of Veromessor pergandei (Hymenoptera: Formicidae) during colony foundation. Anim. Behav. 34, 226–233 (1986).
11. Sommer, K. & Hölldobler, B. Pleometrosis in Lasius niger. In Biology and Evolution of Social Insects (ed. Billen, J.) 47–50 (Leuven University Press, 1992).
12. Sommer, K. & Hölldobler, B. Colony founding by queen associations and determinants of reduction in queen number in the ant Lasius niger. Anim. Behav. 50, 287–294 (1995).
13. Mercier, B., Passera, L. & Suzzoni, J. P. Etude de la polygynie chez la fourmi Plagiolepis pygmaea Latr. (Hym. Formicidae). I. La fécondité des reines en condition expérimentale monogyne. Insects Soc. 32, 335–348 (1985).
14. Keller, L. Evolutionary implications of polygyny in the Argentine ant, Iridomyrmex humilis (Mayr) (Hymenoptera: Formicidae): an experimental study. Annu. Behav. 36, 159–165 (1988).
15. Aribé, S., Oliveras, J. & Gómez, C. Effect of temperature on the oviposition rate of Argentine ant queens (Linepithema humile Mayr) under monogygous and polygynous experimental conditions. J. Insect Physiol. 54, 265–272 (2008).
16. Vander Meer, R. K., Morel, L. & Lofgren, C. S. A comparison of queen oviposition rates from monogyne and polygyne fire ant, Solenopsis invicta, colonies. Physiol. Entomol. 17, 384–390, https://doi.org/10.1111/j.1365-3022.1992.tb01036.x (1992).
17. Vargo, E. L. Mutual pheromonal inhibition among queens in polygyne colonies of the fire ant Solenopsis invicta. Behav. Ecol. Sociobiol. 31, 205–210 (1992).
18. Passera, L., Vargo, E. L. & Keller, L. Le nombre de reines chez les fourmis et sa conséquence sur l'organisation sociale. Ann. Biol. 30(3), 137–173 (1991).
19. Markin, G. P. The seasonal life cycle of the Argentine ant, Iridomyrmex humilis (Hymenoptera: Formicidae), in southern California. Ann. Entomol. Soc. Am. 63(5), 1238–1242 (1970).
20. Keller, L. & Passera, L. & Suzzoni, J. P. Queen execution in the Argentine ant, Iridomyrmex humilis. Physiol. Entomol. 14, 157–163 (1989).
21. Reuter, M., Balloux, F., Lehmann, L. & Keller, L. Kin structure and queen execution in the Argentine ant Linepithema humile. J. Evolution. Biol. 14, 954–958 (2001).
22. Inoue, M. N., Ito, E. & Golke, K. Queen execution increases relatedness among workers of the invasive Argentine ant, Linepithema humile. Ecol. Evol. 5(18), 4098–4107 (2015).
23. Villalta, I., Abril, S., Cerdá, X. & Boulay, R. Queen control or queen signal in ants: what remains of the controversy 25 years after Keller and Nonacs’ seminal paper? J. Chem. Ecol. 44(9), 805–817 (2018).
24. Bach, D., Passera, L. & Aron, S. Le rôle des reines dans l'agression du couvain sexué par les ouvrières de la fourmi d'Argentine Iridomyrmex humilis. Actes Coll. Insects Sociaux 8, 47–54 (1993).
25. Passera, L., Aron, S. & Bach, D. Elimination of sexual brood in the Argentine ant Linepithema humile: queen effect and brood recognition. Entomol. Exp. Appl. 73(2), 203–212 (1995).
26. Vargo, E. L. & Passera, L. Pheromonal and behavioral queen control over the production of gynes in the Argentine ant Iridomyrmex humilis (Mayr). Behav. Ecol. Sociobiol. 28(3), 161–169 (1991).
27. Vargo, E. L. & Passera, L. Gyne development in the Argentine ant Iridomyrmex humilis: role of overwintering and queen control. *Physiol. Entomol.* 17, 193–201 (1992).

28. Deslippe, R. J. The killing of nestmate queens in ant colonies. *Southwest. Entomol.* 25, 111–121 (2002).

29. Abril, S., Olivers, J. & Gómez, C. Effect of seasonal dynamics on queen densities of the Argentine Ant (Linepithema humile) in an invaded natural area of the NE Iberian Peninsula. *Sociobiol.* 51(2), 1–10 (2008).

30. Abril, S., Díaz, M., Fernández, M. I. & Gómez, C. More and bigger queens: a clue to the invasive success of the Argentine ant Iridomyrmex humilis in natural habitats. *Myrmecol. News* 18, 19–24 (2013).

31. Markin, G. P. Food distribution within laboratory colonies of the Argentine ant, Iridomyrmex humilis (Mayr). *Insect. Soc.* 17, 127–158 (1970).

32. Passera, L., Keller, L. & Suzzoni, J. P. Control of brood male production in the Argentine ant Iridomyrmex humilis (Mayr). *Insect. Soc.* 35(1), 19–33 (1988).

33. Abril, S., Olivers, J. & Gómez, C. Foraging activity and dietary spectrum of the Argentine ant (Hymenoptera: Formicidae) in invaded natural areas of the northeast Iberian Peninsula. *Environ. Entomol.* 36(5), 1166–1173 (2007).

34. Abril, S., Olivers, J. & Gómez, C. Effect of temperature on the development and survival of the Argentine ant, *Linepithema humile*. *J. Insect Sci.* 10, 97 available online: insectscience.org/10.97 (2010).

35. Boutilis, A., Détournay, C., Francis, F. & Verheugen, F. Will climate change affect insect pheromonal communication? *Curr. Opin. Insect. Sci.* 17, 87–91 (2016).

36. Abril, S. et al. Cuticular hydrocarbons correlate with queen reproductive status in native and invasive Argentine ants (*Linepithema humile*, Mayr). *PLoS One* 13, e0193115 (2018).

37. Passera, L., Keller, L. & Suzzoni, J. P. Queen replacement in queenless colonies of the Argentine Ant *Iridomyrmex humilis* (Mayr). *Psyche* 95, 59–65 (1988).

38. Liu, Y.-B. & Haynes, K. F. Temporal and temperature-induced changes in emission rates and blend ratios of sex pheromone components in *Trichopius ni*. *J. Insect Physiol.* 40(4), 341–346 (1994).

39. Sentis, A., Ramon-Portugal, F., Brodeur, J. & Hemiptinje, J. L. The smell of change: warming affects species interactions mediated by chemical information. *Glob. Chang. Biol.* 21(10), 3586–3594 (2015).

40. Stein, M. B., Thorvilson, H. G. & Johnson, J. W. Seasonal changes in bait preference by red imported fire ant, *Solenopsis invicta* (Hymenoptera, Formicidae). *Flor. Entomol.* 73, 117–123 (1990).

41. Gaut, S., Barroso, A., Cerdá, X., Amor, F. & Boulay, R. A year in an ant's life: opportunism and seasonal variation in the foraging ecology of *Aphaenogaster senilis*. *Ecology* 20, 19–27 (2013).

42. Holman, L., Dreier, S. & d’Ettorre, P. Selfish strategies and honest signalling: reproductive conflicts in ant queen associations. *Proc. R. Soc. B.* 277, 2007–2015 (2010).

43. Holman, L., Jørgensen, C. G., Nielsen, J. & d’Ettorre, P. Identification of an ant queen pheromone regulating worker sterility. *Proc. R. Soc. B.* 277, 3793–3800 (2010).

44. Vargo, E. L. & Passera, L. The killing of nestmate queens in ant colonies determined by the caste royal of the fire ant (*Solenopsis invicta*). *J. Econ. Entomol.* 87, 101–108 (1994).

45. Passera, L. & Aron, S. Factors controlling dealation and egg laying in virgin queens of the Argentine ant (*Linepithema humile*, Mayr). *Insect. Soc.* 57, 61–63 (1999).

46. Keller, L. & Passera, L. Fecundity of ant queens in relation to their age and the mode of colony founding. *Insect. Soc.* 37, 116–130 (1990).

47. Cuvillier-Hot, V., Lenoir, A., Crewe, R., Malosse, C. & Peeters, C. Fertility signalling and reproductive skew in queenless ants. *Biol. Lett.* 2, 207–216 (2006).

48. Keller, L. & Pasquariello, L. Fecundity of ant queens in relation to their age and the mode of colony founding. *Insect. Soc.* 37, 116–130 (1990).

49. Cuvillier-Hot, V., Lenoir, A., Crewe, R., Malosse, C. & Peeters, C. Fertility signalling and reproductive skew in queenless ants. *Anim. Behav.* 68, 1209–1219, https://doi.org/10.1016/j.anbehav.2003.11.026 (2004).

50. Casellas, D. Tasa de expansión de la hormiga argentina, *Linepithema humile* (Mayr 1868), (Hymenoptera, Dolichoderinae) en un área mediterránea. *Bol. AeE.* 28, 207–216 (2004).

51. Helfer, N. E., Ingram, K. K. & Gordon, D. M. Nest connectivity and colony structure in unicolonial Argentine ants. *Insect. Soc.* 55, 379–403 (2008).

52. Bhaktar, A. P. & Whitcomb, W. H. Artificial diet for rearing various species of ants. *Flor. Entomol.* 53, 229–232 (1970).

53. Newell, W. The life history of the Argentine ant *Iridomyrmex humilis* Mayr. *J. Econ. Entomol.* 2, 174–193 (1919).

54. Newell, W. & Barber, T.C. The Argentine ant. USDA, *Bull. Entomol. Bull.* 122 (1930).

55. Benois, A. Incidence des facteurs écologiques sur le cycle annuel et l’activité saisonnière de la fourmi d’ Argentine, *Iridomyrmex humilis* (Mayr). *Myrmecol. News* 18, 19–24 (2013).

56. R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL, http://www.R-project.org/ (2013).

57. StatSoft, Inc. STATISTICA for Windows [Computer program manual]. Tulsa, OK: StatSoft, Inc., 2300 East 14th Street, Tulsa, OK 74104, phone: (918) 749–1119, fax: (918) 749–2217, email: info@statsoft.com, WEB: http://www.statsoft.com (2001).

Acknowledgements

We would like to thank L. Gómez from the Mass Spectrometry Unit of the University of Girona for identifying the CHCs and for her help during all the chemical analyses. We are also grateful to J. Pearce for her English-language editing services. This work was financially supported by the Spanish Ministry of Science and Innovation and the European Regional Development Fund (CGL2010-16451).

Author Contributions

S.A. conceived and conducted the experiments, analysed the results, prepared the figures and wrote the manuscript. C.G. acquired the funding. All authors reviewed the manuscript.

Additional Information

Competing Interests: The authors declare no competing interests.

Publisher’s note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.
