Optimal imaging time for Tc-99m phytate lymphoscintigraphy for sentinel lymph node mapping in patients with breast cancer

Ching-Chun Ho*, Yu-Hung Chen**, Shu-Hsin Liu***, Hwa-Tsung Chen†, Ming-Che Lee∗

ABSTRACT

Objectives: Sentinel lymph node (SLN) sampling has become a standard practice in managing early-stage breast cancer. Lymphoscintigraphy is one of the major methods used. The radioactive tracer used in Taiwan is Tc-99m phytate. However, this agent is not commonly used around the world and the optimal imaging time has not been studied. Thus, we investigated the optimal imaging time of Tc-99m phytate lymphoscintigraphy for SLN mapping in patients with breast cancer. Materials and Methods: We retrospectively reviewed SLN Tc-99m phytate lymphoscintigraphies in 135 patients with breast cancer between August 2013 and November 2017. The time for the first SLN to be visualized after radiotracer injection was recorded to determine the optimal imaging time. If no SLN was identified on imaging, the scan was continued to 60 min. We also recorded the presurgical technical and clinical factors to analyze the risk factors for nonvisualization of SLN. Each patient’s postoperative axillary lymph node status was also recorded. Results: Axillary SLNs were identified on imaging in 94.8% of the patients. All first SLNs presented within 30 min. In 6 of 7 patients with negative imaging, SLNs were identified during surgery using either blue dye or a hand-held gamma probe. Nonvisualization of SLNs on lymphoscintigraphy was significantly associated with a lower injection dose (1.0 mCi vs. 2.0 mCi), 4-injection protocol (compared to 2-injection), and injection around an outer upper quadrant tumor. In addition, patients with axillary lymph node metastasis had a higher percentage of SLN image mapping failure, with a marginally significant difference. Conclusion: Based on our study, 30 min after Tc-99m phytate injection is the optimal time for lymphoscintigraphy and delayed imaging beyond 30 min is not necessary. In addition, a lower injection dose, the 4-injection method, and an injection near the outer upper quadrant tumor should be avoided to minimize nonvisualization of SLNs.

KEYWORDS: Breast cancer; Lymphoscintigraphy, Phytate, Sentinel lymph node

INTRODUCTION

Breast cancer is one of the most prevalent cancers and is the leading cause of cancer death in women worldwide [1]. The treatment of breast cancer is multidisciplinary and requires detailed risk stratification. Lymph node status is one of the most important factors guiding the subsequent management and prognosis [2-5]. Therefore, assessing lymph node status is of paramount importance in treating patients with breast cancer.

In the era of advanced molecular imaging, many state-of-the-art noninvasive imaging modalities have emerged; yet, sentinel lymph node (SLN) sampling remains the procedure of choice in detecting regional lymph node metastasis in early-stage breast cancers [6,7]. Owning to its high diagnostic performance and prognostic significance, SLN sampling has become standard practice [8,9].

Colloid materials in the tissue can be absorbed and transported by the lymphatic system, and introducing a radiolabel to the colloid material makes it suitable for SLN mapping. Ample studies have demonstrated that radioactive colloids are highly reliable and capable of finding SLN, and radioactive colloids are even superior to their similar counterpart, blue dye [10,11]. Besides these advantages, radioactive colloids allow preoperative SLN imaging (lymphoscintigraphy), which is noninvasive and useful in guiding intraoperative nodal localization with a hand-held gamma probe [12]. Thus, preoperative radiotracer lymphoscintigraphic mapping is also highly...
recommended [13]. Despite the promising features of preoperative lymphoscintigraphy, the imaging procedure might interfere with operating room scheduling, and prolonged scanning time might also intensify patient discomfort. Unfortunately, the imaging time for preoperative lymphoscintigraphy has been less addressed in the literature. There has been only one study evaluating the optimal imaging time for SLN mapping in patients with breast cancer [14]. Furthermore, the most commonly used radioactive colloid in Taiwan is radioactive phytate (Tc-99m phytate), which is not commonly used around the world, and is different from the radiotracer used in the aforementioned study. Because the physical properties of each radioactive colloid are quite different, it might not be possible to generalize the study results of one radioactive colloid to other radioactive colloids [13,15]. Therefore, the ideal preoperative imaging time for Tc-99m phytate lymphoscintigraphy for SLN mapping in patients with breast cancer is yet to be thoroughly studied.

We herein conduct a retrospective review of breast cancer patients who underwent Tc-99m phytate lymphoscintigraphy for SLN mapping. The preoperative imaging time will be analyzed, and the factors affecting the identification rate on lymphoscintigraphy will be investigated as well.

**MATERIALS AND METHODS**

**Ethics statements**

The study was conducted in accordance with the Declaration of Helsinki and was approved by the local ethics committee of the institution (IRB106-182-B). Informed written consent was waived because the study was a retrospective data analysis.

**Clinical and imaging data**

We retrospectively reviewed the medical charts and lymphoscintigraphic images of adult patients referred for SLN mapping from August 2013 to November 2017 in Buddhist Tzu Chi General Hospital, Hualien, Taiwan. Only patients with a diagnosis of breast cancer were included for analysis. We included only early-stage (Stage I and II) breast cancer patients, and no neoadjuvant chemotherapy or radiotherapy had been administered before the lymphoscintigraphy.

To perform SLN lymphoscintigraphy, Tc-99m phytate was injected intradermally around the tumor or around the areola. The total volume of the radiotracer was 0.4 mL and the total activity was 1.0 or 2.0 mCi. The radiotracer was divided into two or four doses for injection. Gentle massage was applied to the injection sites for all patients. Immediately after radiotracer injection and massage, image acquisition commenced using a dual-head gamma camera (Infinia Hawkeye 4, GE Healthcare, Milwaukee, Wisconsin, USA) equipped with a parallel-hole low-energy high-resolution collimator. The image matrices were 128 × 128 and the energy window was 15% centered over 140 keV. An initial set of dynamic images was acquired at 1 min/frame for 10 min. Subsequently, static images were acquired at 5 min intervals. If there was no SLN on the image, image acquisition was continued until 60 min was reached. All lymphoscintigraphic images were displayed on a GE Xeleris 2.1753 Workstation (GE Healthcare).

All patients then underwent SLN biopsy using the dual-tracer method (using both blue dye and an intraoperative hand-held gamma probe). Blue or hot nodes were regarded as SLNs and were sent to the pathologist for frozen section examination. Lymph node dissection over Level I and Level II of the axilla was performed if the frozen section was positive for metastasis; otherwise, lymph node dissection was not performed. If no SLN was identified during surgery, the surgeon performed lymph node dissection [Figure 1].

**Statistical analysis**

The data were expressed as frequencies and mean or standard deviation depending on the characteristics of each item. Fisher’s exact test was used for bivariate analysis of the association between different variables and the rate of failure to identify SLNs on lymphoscintigraphy. To fit into the bivariate analysis, tumor sizes were separated into two groups with a cutoff of 1.3 cm using the receiver operating curve method [Figure 2]. To eliminate possible confounders, the effect of each presurgical technical and clinical variable on the failure rate was further examined by multivariate logistic regression analysis. We used stepwise analysis to identify the

![Figure 1: Flowchart showing the sentinel lymph node biopsy procedure in our study](image-url)
most predictive presurgical variables. Data were expressed as β coefficients and 95% confidence intervals of β and associated P values. Statistically significant differences were defined as P < 0.05. All statistical analyses were performed using SPSS software (version 19.0; SPSS Inc., Chicago, IL, USA).

RESULTS

A total of 135 patients were included for analysis and their baseline characteristics are summarized in Table 1. SLNs were detected by lymphoscintigraphy in the majority of patients (94.8%). In addition, all SLNs were detected within 30 min of imaging. These SLNs were then detected by the dual-tracer method and harvested intraoperatively. Only seven patients did not show SLNs on lymphoscintigraphy. SLNs were identified in six of these seven patients during the surgery using the dual-tracer method. Only one patient had negative image findings and had no SLNs detected during the surgery.

Tumor size, injection dose, and injection site showed statistical significance with the failure rate of SLN mapping on imaging [Table 2]. The technical and clinical variables were further examined by stepwise multivariate logistic regression analysis [Table 3]. The initial analysis disclosed that injection dose (1.0 mCi or 2.0 mCi) was the only independent variable that affected the SLN identification rate. In addition, the number of injections (4 vs. 2) and tumor location (UOQ vs. others) showed marginally significant associations. After eliminating nonsignificant variables and combining the tumor location with injection site, the most significant variables that were associated with nonvisualization of SLN on imaging were a lower injection dose, 4 injections (compared with 2 injections), and radiotracer injection around an upper outer quadrant tumor.

Postsurgical pathologic results showed that 25 patients had metastatic lymphadenopathy in the axillary region. Lymphoscintigraphy failed to identify SLNs in three patients. SLNs were successfully identified during surgery in two of these three patients, while no SLN was detected in the other patients even during the surgery. The pathological lymph node status was marginally associated with the failure rate of SLN mapping on imaging [Table 2, P = 0.099]. The actual number of metastatic lymph nodes, on the other hand, did not correlate with the imaging or clinical variables, including size of the primary tumor, number of SLNs on imaging, and the time to first SLN on imaging.

DISCUSSION

SLN mapping has been applied to many different cancers, including breast cancer, melanoma, head and neck cancers, and even cancer of the genitalia [16,17]. Due to the high prevalence of breast cancer and the advent of screening mammography, more patients can be diagnosed early and sentinel lymphoscintigraphy is increasingly being used. The imaging time for sentinel lymphoscintigraphy is thus an important issue, especially for surgical departments with a tight schedule. Sadeghi et al. studied the imaging time using Tc-99m antimony sulfide colloid and concluded that a 30-min scan is sufficient [14]. However, the particle migration speed in the lymphatic tracts and the time to visualize SLN after radiotracer injection varies according to the particle size [13,18,19]. The particle size of Tc-99m phytate

![ROC Curve](image)

**Table 1: Baseline characteristics of patients (n=135)**

| Biopsy method | Value |
|---------------|-------|
| Excisional biopsy | 68    |
| CNB | 67    |

| SLN mapping failure rate (in %) | 92.3 |

**Table 2:** The receiver operating characteristic curve for different tumor sizes in predicting sentinel lymph node visualization. A cutoff of 1.3 cm obtained from the ROC curve analysis has an acceptable sensitivity of 74.2% and an acceptable specificity of 71.4%. The area under the curve is 0.739.
Table 2: Analysis of the association of variables with failed sentinel lymph node identification on lymphoscintigraphy (Fisher’s exact test)

| Variables                          | Image mapping failure (%) | P     |
|------------------------------------|---------------------------|-------|
| Age (years)                        |                           |       |
| <55                                | 4.5                       | 1.00  |
| ≥55                                | 5.9                       |       |
| Tumor size (cm)                    |                           |       |
| <1.3                               | 14.3                      | 0.0128*|
| ≥1.3                               | 2.0                       |       |
| Biopsy method                      |                           |       |
| Excisional biopsy                  | 8.8                       | 0.1153|
| Core-needle biopsy                 | 1.5                       |       |
| Injection dose (mCi)               |                           |       |
| 1.0                                | 25.0                      | 0.0008*|
| 2.0                                | 1.7                       |       |
| Number of injections               |                           |       |
| 2 injections                       | 1.6                       | 0.124 |
| 4 injections                       | 8.2                       |       |
| Injection site                     |                           |       |
| Around the tumor                   | 8.4                       | 0.0432*|
| Around the areola                  | 0                         |       |
| Tumor location                     |                           |       |
| UOQ                                | 10.4                      | 0.0967|
| Others                             | 2.3                       |       |
| Pathological lymph node metastasis |                           |       |
| Presence                           | 12                        | 0.099 |
| Absence                            | 3.6                       |       |

*Statistically significant. UOQ: Upper outer quadrant.

An injection dose of 1.0 mCi (vs. 2.0 mCi), the 4-injection method, and radiotracer injection around an upper outer quadrant tumor were the only three independent risk factors with statistical significance for nonvisualization of SLN on lymphoscintigraphy. These three technical factors have not been reported to be associated with nonvisualization of SLN on lymphoscintigraphy in the literature. The injection dose, nonetheless, is reasonably regarded as a predictive variable because higher injected radioactivity theoretically produces a more intense signal from the SLN and hence is easier to detect by a gamma camera. Although most studies used a radioactivity dose <1.0 mCi, our study results imply that a 2.0 mCi total radioactivity dose might achieve a better detection rate when Tc-99m phytate is used [14,22-26]. This phenomenon can be endorsed by the hypothesis we propose. The smaller Tc-99m phytate particles migrate to the SLN faster. Higher injected total radioactivity, with the same distribution of Tc-99m phytate particle sizes, results in an overall higher migrated radioactivity to the SLN. Quantitative analysis of filtered Tc-99m phytate might be able to verify this theory in the future. Another possible explanation for lower doses with similar high SLN visualization rates in other studies is that those studies had longer imaging times. For example, Koizumi et al. performed lymphoscintigraphy with 0.4–1.5 mCi of Tc-99m phytate and achieved an SLN visualization rate of 100% on imaging [22]. However, their imaging time was 1–2 h, which was considerably longer than ours (within 60 min). Although no SLN was found between 30 and 60 min in our study, we still cannot exclude the possibility of an improved SLN visualization rate when the imaging time is extended beyond 60 min.

The association between tumor location and the rate of imaging mapping failure might be explained by the obscuring effect of the intense radioactivity in the injection site around the tumor, which hampers detection of the much weaker radioactivity in the SLN. No significant association with SLN visualization was found on multivariate analysis when only the injection site was considered (injection around the tumor vs. injection around the areola). These results suggest that a lower SLN detection rate on imaging is not only related to the site of injection or tumor location alone but also related to a combination of these two factors.

Currently, there is no consensus on the number of tracer injections, and the technique varies across different nuclear medicine departments. In our study group, the “2-injection method” and “4-injection method” differs in the volume per injection. A greater volume will be delivered to an injection site when the “2-injection method” is used, which can generate higher tissue pressure to facilitate lymphatic drainage. This is concordant with the findings reported by other groups [27-29]. However, the results are conflicting and the effect of injection volume on the nodal detection rate might be multifactorial [25]. Because the 4-injection method is surely less tolerable for patients, the 2-injection method is thus recommended for routine practice, since the detection rate is not inferior.

Finally, our study results showed that the rate of no SLN on imaging is higher in patients with pathological lymph node metastasis. Although only a statistical trend was reached, this
finding is congruent with other authors’ work investigating the association of lymphoscintigraphic patterns with axillary nodal metastasis [20,30]. The increased axillary metastatic burden obstructs lymphatic drainage and hampers radiotracer transition to the SLN. Thus, no SLN can be visualized on imaging.

There were several limitations in our study. First, not all of the study participants received axillary dissection. Thus, it is not possible to determine the false-negative rate. Second, the study population was relatively small and the number of patients without successful lymphoscintigraphic mapping was very limited. A larger study population is required to minimize statistical noise. Third, the study was retrospective and thus susceptible to selection bias. And finally, this retrospective study reviewed lymphoscintigraphies across 4 years. The confounding effect of technical maturity on the performance of SLN mapping cannot be excluded completely.

**Conclusion**

SLNs were visualized on lymphoscintigraphy in the majority of breast cancer patients, and most nodes could be shown within 30 min after tracer injection. To ensure a high surgical throughput and to minimize patient discomfort, imaging should not be longer than 30 min. To maximize the rate of successful mapping on imaging, we should avoid using lower doses of Tc-99m phytate (1.0 mCi), multiple injections (4-injection method), and injection around a UOQ tumor.

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| Table 3: Stepwise multivariate logistic regression analysis of presurgical variables with failed sentinel lymph node mapping on imaging (n=135) |
|---------------------------------------------------------------|
| Variables | β | CI | P |
| All variables | | | |
| Age (years) | | | |
| ≥55 versus <55 | 1.456 | -1.155-4.067 | 0.274 |
| Tumor size (cm) | | | |
| ≥1.3 versus <1.3 | -1.285 | -3.782-1.212 | 0.313 |
| Biopsy method | | | |
| Excisional versus CNB | 0.740 | -2.104-3.584 | 0.610 |
| Injection dose (mCi) | | | |
| 2.0 versus 1.0 | -3.763 | -6.869--0.837 | 0.012* |
| Number of injections | | | |
| 4 versus 2 | 3.268 | -0.209-6.745 | 0.065 |
| Injection site | | | |
| Around the tumor versus around the areola | 17.226 | -9279-9313 | 0.997 |
| Tumor location | | | |
| UOQ versus others | 2.578 | -0.268-5.424 | 0.076 |
| Selected variables | | | |
| Injection dose (mCi) | | | |
| 2.0 versus 1.0 | -3.744 | -6.129--1.359 | 0.002* |
| Number of injections | | | |
| 4 versus 2 | 3.299 | 0.367-6.231 | 0.027* |
| Tumor location and injection site | | | |
| Injection around UOQ tumor versus others | 2.823 | 0.465-5.181 | 0.019* |

*Statistically significant. CNB: Core needle biopsy, UOQ: Upper outer quadrant, CI: Confidence interval

**Conflicts of interest**

There are no conflicts of interest.

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