Impact of Temperature Changes to the Adhesion Strength of Molar Tubes: an *in Vitro* Study

Benedikta Palesik (✉ benedikta.palesik@gmail.com)
Lithuanian University of Health Sciences

Kotryna Šileikytė
Vilnius Gediminas Technical University

Julius Grįškevičius
Vilnius Gediminas Technical University

Rimantas Stonkus
Vilnius Gediminas Technical University

Antanas Šidlauskas
Lithuanian University of Health Sciences

Kristina Lopatienė
Lithuanian University of Health Sciences

---

**Research Article**

**Keywords:** Orthodontic, adhesive, molar tubes, temperature, thermal cycle

**Posted Date:** February 4th, 2022

**DOI:** https://doi.org/10.21203/rs.3.rs-1307282/v1

**License:** ☺️ ☑️ This work is licensed under a Creative Commons Attribution 4.0 International License.

Read Full License
Abstract

**Background:** The main purpose of this was to determine study adhesion strength of molar tubes bonding with a composite adhesive after exposure to a sudden change in temperature (thermal cycles).

**Methods:** The study sample consisted of 40 recently extracted human first permanent molars, which were randomly divided into two groups of 20: group 1 was the experimental group (affected by thermal cycles), and group 2 was the control group. Molar tubes were bonded with a light-cure tube adhesive. The experimental group teeth were dipped 2,000 times in saline at 5°C and at 55°C. The control group were immersed in 37°C saline. Molar tubes for both groups were removed with an adapted Mecmesim Multitesters 2.5 - I, and the data were recorded with EMPEROR software. ANOVA was used to calculate and compare the results.

**Results:** In the experimental group of the teeth, the maximum force was obtained at 94.2 N and the lowest force was 19.69 N. In the control group of the teeth, the maximum force was obtained at 159.1N and the lowest force was 28.1N. In the experimental group, the mean debonding force (59.12 N) was statically significantly smaller than in the control group (79.88N), p=0.0345. The forces in the control group were by 1.35 times greater than those in the experimental group.

**Conclusions:** The forces of the adhesion of molar tubes to the tooth surface were reduced after exposure to a sudden change in temperature (thermal cycles). The results were significantly different between the experimental group and the control group.

Background

Orthodontic treatment with fixed appliances improves facial aesthetics and oral function, which have a significant impact on both dental health and human psychological well-being [1]

During orthodontic treatment with fixed appliances, one of the main problems is bracket/molar tube detachment, as it can reduce the success of orthodontic procedures while increasing the duration and the cost of the treatment and damaging tooth enamel [2-3] Studies of bracket/molar tube detachment have shown that the tubes detached from the molars more often than the brackets from the anterior teeth did [4-8].

In order to optimise orthodontic treatment, various in vitro and in vivo studies have been performed to improve the quality of the bonding technique (direct and indirect) and to develop new adhesives based on the need to increase the shear bond strength, to shorten bonding time, to effectively reduce clinical bonding steps, and to preserve enamel. Orthodontic adhesives are divided into three major groups: chemical-curing (glass ionomer cement, which sets by an acid-base reaction), light-curing (polyacid-modified composite resin (compomer)), and a tri-cure mechanism (resin-modified glass ionomer cement) [5,9-13].
Studies have reported that clinical bonding should be considered to be successful when the minimum shear bond strength is 5.9-10MPa, and the debonding of brackets/molar tubes should be performed easily and without damage to the enamel surface at the end of the treatment [14-16]. According to bonding protocols, the tooth surface is etched with 35 -40% phosphoric acid before fixing the bracket/molar tube to ensure the desired dissolution of the surface enamel, which results in micro-porosity on the surface allowing the resin monomers to penetrate and mechanically bond. The bond strength obtained using this bracket/molar tube bonding protocol is generally high, ranging from 9 to 35MPa [17]. Studies have shown that the bond strength in vivo was significantly lower than in vitro [18-19].

Several factors affect the bond strength of orthodontic brackets/molar tubes, including contamination, type of the composite resin, viscosity of the adhesive, etching type of the enamel, storage conditions, size and shape of the bracket/molar tube base, temperature of the composite during bracket/molar tube fixation, enamel surface damage by caries or fluorosis, and restorations of the tooth [20]. Ingestion of food or beverages often causes sudden changes in temperature in the oral cavity, which also affects fixed orthodontic appliances, and thus it is necessary to study whether these temperature changes may affect the bond strength of brackets/molar tubes and to revise the orthodontic recommendations before treatment. To simulate the temperature changes in the oral cavity, thermally controlled water baths are used in in vitro studies [21].

However, a little research has been done on the effect of the thermal cycles on the adhesive after bracket/molar tube fixation and no studies have examined the correlation between bracket/molar tubes displacement and the force required during debonding. The formula of composites used in orthodontic for bonding brackets/molar tubes is improved every year. The aim of this in vitro study was to evaluate the effect of the thermal cycles on tube bonding with composite adhesion. Following this, a null hypothesis was formulated: there is no difference between the adhesives affected by thermal cycles and the control group in shear bond strength.

**Materials And Methods**

This *in vitro* study was performed in the Lithuanian University of Health Sciences at the Department of Orthodontic and the Laboratory of the Department of Biomechanical Engineering of VILNIUS TECH. The study was performed following an individual protocol, the methods of which were planned based on the previous studies [1, 22-26]. Bioethical approval was obtained from the University's Bioethical Committee, No. BEC-OF-05

Informed consent was obtained for experimentation with human teeth. The privacy rights of human subjects must always be observed.

The sample size of the trial was calculated according to the formula 2.1:
Formula 2.1. n – the minimum sample size for each group; \( z_{1-\frac{\alpha}{2}} = 1.96 \) and \( z_{1-\beta} = 0.84 \), when \( \alpha = 0.05 \) and \( \beta = 0.2 \); \( s_1, s_2 \) – standard deviation of the first samples; and \( \Delta \) – minimal clinically important difference.

Calculations were performed using G * Power (Version 3.1.9.2) statistical software. The following parameters were adjusted accordingly: significance level, 5%, strength test, 80%, standard deviation, 29.514N and 20.755N for pilot tests, and the least significant effect applied, 2.

The estimated minimum sample size required is 15 teeth for each group.

Tooth inclusion criteria were the following: molars with intact buccal enamel surface recently extracted for periodontal purposes, removed after jaw fractures, unsealed, not damaged by caries, not damaged by fluorosis, and without endodontic treatment. The teeth were collected over a month’s period. The extracted teeth were stored in a disinfectant (Gigasept Istru AF) for 15 minutes, then washed under running water for 1 minute and kept in room-temperature (of 22°C) saline according to the protocol of the previous tests in order to avoid a significant effect of the storage medium on the bond strength. The isotonic solution was changed daily to avoid bacterial growth.

After applying the selection criteria, 40 permanent teeth from a sample of 57 were included into our study. The teeth were randomly divided into two groups of 20, group 1 being the experimental group (E) (affected by thermal cycles), and group 2 – the control group (C).

According to the protocol, the buccal surface of each tooth was polished for 30 seconds with non-fluoridated polishing paste and a rubber brush hand piece set at low speed, washed with water for 30 seconds, and blow-dried with compressed air for 10 seconds. the prepared enamel area was etched with 37% phosphoric acid for 40 seconds, washed with water for 30 seconds, and dried with compressed air until the tooth surface became non-glossy. Using a micro brush, the etched enamel surface was coated with a thin, even layer of binder resin (HIGH-Q-BOND BRACKET. PRIMER), and the air was blown until the binder became non-flowable. Tubes (American Orthodontic, ifit) were bonded to the centre of the clinical crown with a light-curing tube adhesive (HIGH-Q-BOND BRACKET), pressed with a 100g weight on the buccal tooth surface and cured with a polymerisation lamp (Translux Wave, Heraeus Kulzer, Germany, 1000 mW/cm²) for 40 seconds, keeping the light source at 1mm to the surface of the tube. The bonding was performed by one person to ensure accuracy.

Each tooth was centralised and fixed up to the neck area in iron rectangular boxes of the same shape filled with epoxy resin (Faserverbung) to ensure the stability of the samples. The molars were attached to the loom so that the buccal tooth surface would be parallel to the “Mechemsi” tension crushing device.
All samples were numbered: the experimental group (affected by thermal cycles), E1-E20, and the control group of teeth, C1-C20.

Following that, 20 teeth of the experimental group were dipped 2,000 times in cold saline at 5°C and hot saline at 55°C. The immersion or dwell time in each bath was 30 seconds with a transfer time of 2-3 seconds. The saline temperature was maintained by a baby food heater/sterilizer. Meanwhile, 20 teeth of the control group were immersed in 37°C saline. These prepared teeth were kept in saline at room temperature of 22°C until the start of the test (i.e., for 5 hours).

The teeth were fixed on a loom. An adapted Mecmesim Multitesters 2.5-I (Mecmesim Limited, United Kingdom) material tester was used to remove the tubes. A tightening tube gripping mechanism and an epoxy-fixed tooth holder were fabricated for the attachment (Figure 1). A constant speed of 0.1 mm/s was used to remove the tubes. This system was fully connected to the computer, thus the force and the displacement of debonding of the orthodontic tubes from the tooth surface was fixed. The adhesion of the molar tubes to the tooth surface was assessed by the debonding force and the displacement of the molar tubes during removal.

**Statistical analysis**

The data were recorded with EMPEROR software. ANOVA (statistically used MATLAB program) was used to calculate and compare the results. One-way analysis of variance (ANOVA) was used to analyse the effects of temperature on the adherence of the tubes. The null hypothesis tested was whether the mean maximum debonding force of the tubes would be the same in the control and the experimental groups. Multiple comparisons using the Tukey-Kramer test were used to assess the significance of the mean difference. Statistical analysis was performed with the MATLAB (Mathworks Inc, USA) software package.

The difference in data between the control and the experimental groups was regarded as statistically significant when (p < α, α = 0.05).

**Results**

In this study, we evaluated molar tube adhesion forces with tooth surfaces; The comparison of the debonding force and its distribution in the experimental and the control groups is presented in Figure 2. In the experimental group, the mean debonding force (59.12N) was statically significantly smaller than in the control group (79.88N), (p=0.0345), and thus the zero hypothesis was rejected.

The results of force and displacement dependencies in the experimental are presented in Figure 3 and the results of control groups are presented in Figure 4. The purpose of this specific test is to determine the correlation between the displacement distance of the molar tubes and the force required during debonding.
In the experimental group of the teeth, the maximum force was obtained at 94.2 N with a displacement of 0.75 mm, and the lowest force was 19.69 N with a displacement of 0.08 mm. In the experimental group the maximum displacement (1.15 mm) was obtained at 62 N debonding force.

In the control group of the teeth, the maximum force was obtained at 159.1 N with a displacement of 0.69 mm, and the lowest force was 28.1 N with a displacement of 0.14 mm. In the control group the maximum displacement (1.6 mm) was obtained at 121 N debonding force.

The comparison of force dependence on the displacement in the experimental and the control groups is presented in Figure 5. The forces in the control group were by 1.35 times greater than in the experimental group, and the displacement was by 1.75 times greater in the experimental group than in the control group.

**Discussion**

To improve orthodontic treatment, evaluations of the factors that may impair bracket/molar tube adhesion are carried out. Laboratory tests are often used to evaluate the performance of adhesive systems before long-term clinical trials to determine the clinical efficacy of improved adhesive systems in the oral cavity [24]. The adhesion between the enamel and the bracket/molar tubes weakens due to the three different phenomena: mechanical, chemical, and thermal changes. Although in vitro studies cannot accurately reproduce in vivo conditions, when properly prepared and exposed, we can simulate different phenomena that occur in the oral cavity [28]. Thus, thermal cycling is fully accepted and widely used by the scientific community in experimental studies to create conditions similar to those in the oral cavity [12]. Even though some differences in temperature changes measured in the mouth and different tolerances for extreme temperatures have been reported, researchers agree that in laboratory tests on thermal cycle samples, the temperature should be from 5°C to 55°C [12]. These temperatures also comply with the technical specification of ISO TS 11405 for testing the adhesion to the tooth structure [22]. The number of cycles in laboratory testing has not been based on scientific data [28].

The number of samples, the bracket/molar tube model, the time spent in the water baths, the transfer time, the number of cycles, and the removal techniques are chosen by the researchers, and the result of the adhesion of the brackets/molar tubes to the tooth surface can vary greatly depending on the method used. For this reason, Fritz with other authors proposed a separate control for each study [23].

It is important to mention that previous studies have shown that molar tubes debonded more often than brackets of premolars and anterior teeth (canines and incisors) [4-7]. In a year-long study of bracket/molar tube debonding frequency, Maijer and Smith showed that debonding rates of incisor, canine, premolar, and molar brackets/molar tubes were, respectively, 3.6%, 1.6%, 4.8%, and 11.6%, and it was reported that there was no significant difference of molar tube debonding rates between the first and the second molars [8]. For this reason, first molar teeth were used in this study.
In our study, while taking into account other studies [1, 22-26] the teeth of the experimental group were dipped 2,000 times in 5°C and 55°C saline. The obtained results showed that the debonding force in the experimental group was, on average, by 20.75N, or by 1.35 times lower than that in the control group. The statistically significant difference in forces of the composite (HIGH-Q-BOND BRACKET) between the control and the experimental groups was confirmed (p>alpha, p=0.0345, alpha=0.05).

In studies where the conventional bracket/molar tube bonding method was used [24], the mean values of bracket/molar tube adhesion to the tooth surface after 2,000 and 5,000 thermal cycles decreased slightly. These reductions were not statistically significant. However, Daub et al. [25] reported that the average adhesion of the brackets/molar tubes, using the conventional method of attaching the bracket/molar tube to human premolars and using Transbond XT adhesive, was significantly reduced by 16.7% after 500 thermal cycles. Saito et al. [26] observed a significant decrease in bracket/molar tube adhesion after 2,000 and 5,000 thermal cycles using the conventional bracket/molar tube attachment method.

Also in our study, using computer equipment, by debonding the molar tubes in the vertical direction, minimal displacement of the molar tubes until its complete detachment from the tooth surface was recorded. Such research has not been done in other studies. Comparing the correlation between the molar tubes displacement and force during debonding in both groups, we found that in the experimental group to achieve the same force as in the control group, the molar tubes displacement during debonding is 1.75 greater.

Based on the results of our and other studies [25-26], an additional recommendation of orthodontist to the patient before orthodontic treatment should be included: to change dietary habits and not to cause sudden changes in the temperature in the oral cavity as this weakens the adhesion and increases the chance of debonding of the brackets/molar tubes.

Regarding the causes of the deterioration of the adhesion of the brackets/molar tubes, it is important to mention that during the thermal cycle test, the samples undergo sudden temperature changes and are exposed to water. Differences in the coefficient of the thermal expansion of metal brackets/molar tubes, the adhesive, and the tooth result in repetitive shrinkage/expansion stresses [12, 29]. The resin expands and contracts because its coefficient of thermal expansion is higher than that of the teeth; the higher the coefficient of thermal expansion of the resin, the worse it would be for the adhesive bond, as the volumetric changes in the resin will be greater. In addition, the water in this procedure causes hygroscopic expansion as well as chemical decomposition of the resinous components, which is called plastification [30-31].

It has been suggested that the minimum bracket/molar tube adhesion force should be 6–10MPa to achieve an acceptable clinical result [15]. From 1975, a standard procedure in the orthodontic practice is to etch the tooth surface with 35-40% phosphoric acid before bonding the brackets/molar tubes to ensure the micro-porosity on the tooth surface to allow the resin monomers to penetrate and mechanically bond and to increase the adhesion strength of the brackets/molar tubes from 9 to 35 MPa [1]. Our results far
exceeded these limits. The difference could be due to the choice of different protocols and molar tube removal methods [17].

In vitro studies such as this have some limitations. Saliva and patients’ oral hygiene, diseases, and habits may affect the results, but in vitro, we cannot accurately simulate a multifactorial oral environment, thus the results of our study show the effect of only one factor on the debonding force of orthodontic brackets/molar tubes. In addition, some other factors, such as the type of the adhesive used, the mechanical and chemical surface preparations, and the number of thermal cycles may also affect the adhesion of the brackets/molar tubes. Thus, in order to obtain more accurate results and to evaluate and compare the effect of the thermal cycles on different types of adhesives, a greater number of subjects and studies are required. The effect of the thermal cycles on the adhesion strength of ceramic brackets/molar tubes to the tooth surface should also be evaluated.

Conclusion

The adhesion force of molar tubes to the tooth surface is influenced by sudden changes in temperature (thermal cycles), which reduces it statistically significantly. For this reason, when discussing the recommendations of orthodontic treatment with fixed appliances for patients, the influence of temperature changes on the adhesion of the brackets should be discussed as well.

Declarations

Ethics approval

This study was performed according to the Declaration of Helsinki, and was approved by the ethics committee of the University of Lithuanian University of Health Sciences (No.: BEC-OF-05). All patients gave their written informed consent to participate in the study.

Consent for publication

Not applicable.

Availability of data and materials

All data generated or analysed during this study are included in this published article.

Competing interests

The authors declare that they have no competing interests.

Funding

Not applicable.
Authors’ contributions

Benedikta Palesik: interpretation of data, design of the work.

Kotryna Šileikytė: the acquisition and analysis of data.

Julius Griškevičius: the acquisition and analysis of data.

Rimantas Stonkus: the acquisition and analysis of data.

Antanas Šidlauskas: the research consultant.

Kristina Lopatienė: the research supervisor.

All authors reviewed the manuscript.

Acknowledgements

Not applicable.

References

1. Ibrahim AI, Al-Hasani NR, Thompson VP, Deb S. In vitro bond strengths post thermal and fatigue load cycling of sapphire brackets bonded with self-etch primer and evaluation of enamel damage. J Clin Exp Dent. 2020 Jan 1;12(1):e22-e30. doi: 10.4317/medoral.56444. PMID: 31976040; PMCID: PMC6969965.

2. Powers JM, Kim HB, Turner DS. Orthodontic adhesives and bond strength testing. Semin Orthod. 1997 Sep;3(3):147-56. doi: 10.1016/s1073-8746(97)80065-5. PMID: 9573876. Powers, J. M., Kim, H. B., & Turner, D. S. (1997). Orthodontic adhesives and bond strength testing. Seminars in orthodontics, 3(3), 147–156. https://doi.org/10.1016/s1073-8746(97)80065-5

3. Almosa N, Zafar H. Incidence of orthodontic brackets detachment during orthodontic treatment: A systematic review. Pak J Med Sci. 2018 May-Jun;34(3):744-750. doi: 10.12669/pjms.343.15012. PMID: 30034451; PMCID: PMC6041531.

4. Roelofs T, Merkens N, Roelofs J, Bronkhorst E, Breuning H. A retrospective survey of the causes of bracket- and tube-bonding failures. Angle Orthod. 2017 Jan;87(1):111-117. doi: 10.2319/021616-136.1. Epub 2016 Jun 15. PMID: 27304230; PMCID: PMC8388589.

5. Millett DT, Mandall NA, Mattick RC, Hickman J, Glenny AM. Adhesives for bonded molar tubes during fixed brace treatment. Cochrane Database Syst Rev. 2017 Feb 23;2(2):CD008236. doi: 10.1002/14651858.CD008236.pub3. PMID: 28230910; PMCID: PMC6464028.

6. Flores-Mir C. Bonded molar tubes associated with higher failure rate than molar bands. Evid Based Dent. 2011;12(3):84. doi: 10.1038/sj.ebd.6400813. PMID: 21979772.
7. Fricker JP, McLachlan MD. Clinical studies of glass ionomer cements. Part I–A twelve month clinical study comparing zinc phosphate cement to glass ionomer. Aust Orthod J. 1985 Mar;9(1):179-80. PMID: 3911940.

8. Maijer R, Smith DC. A comparison between zinc phosphate and glass ionomer cement in orthodontics. Am J Orthod Dentofacial Orthop. 1988 Apr;93(4):273-9. doi: 10.1016/0889-5406(88)90156-4. PMID: 3281438.

9. Buonocore MG. A simple method of increasing the adhesion of acrylic filling materials to enamel surfaces. J Dent Res. 1955 Dec;34(6):849-53. doi: 10.1177/00220345550340060801. PMID: 13271655.

10. Mandall NA, Hickman J, Macfarlane TV, Mattick RC, Millett DT, Worthington HV. Adhesives for fixed orthodontic brackets. Cochrane Database Syst Rev. 2018 Apr 9;4(4):CD002282. doi: 10.1002/14651858.CD002282.pub2. PMID: 29630138; PMCID: PMC6494429.

11. Newman GV. Epoxy adhesives for orthodontic attachments: progress report. Am J Orthod. 1965 Dec;51(12):901-12. doi: 10.1016/0002-9416(65)90203-4. PMID: 5214895.

12. Gale MS, Darvell BW. Thermal cycling procedures for laboratory testing of dental restorations. J Dent. 1999 Feb;27(2):89-99. doi: 10.1016/s0300-5712(98)00037-2. PMID: 10071465.

13. Buyukyilmaz T, Usumez S, Karaman Al. Effect of self-etching primers on bond strength–are they reliable? Angle Orthod. 2003 Feb;73(1):64-70. doi: 10.1043/0003-3219(2003)073<0064:EOSEPO>2.0.CO;2. PMID: 12607857.

14. Mansour AY, Drummond JL, Evans CA, Bakhsh Z. In vitro evaluation of self-etch bonding in orthodontics using cyclic fatigue. Angle Orthod. 2011 Sep;81(5):783-7. doi: 10.2319/012811-59.1. Epub 2011 May 5. PMID: 21545301.

15. Lamper T, Ilie N, Huth KC, Rudzki I, Michelhaus A, Paschos E. Self-etch adhesives for the bonding of orthodontic brackets: faster, stronger, safer? Clin Oral Investig. 2014 Jan;18(1):313-9. doi: 10.1007/s00784-013-0942-2. Epub 2013 Feb 14. PMID: 23408192.

16. Reynolds IR, von Fraunhofer JA. Direct bonding of orthodontic brackets–a comparative study of adhesives. Br J Orthod. 1976 Jul;3(3):143-6. doi: 10.1179/bjo.3.3.143. PMID: 788775.

17. Øgaard, B., & Fjeld, M. (2010, March). The enamel surface and bonding in orthodontics. In Seminars in orthodontics (Vol. 16, No. 1, pp. 37-48). WB Saunders.

18. Murray SD, Hobson RS. Comparison of in vivo and in vitro shear bond strength. Am J Orthod Dentofacial Orthop. 2003 Jan;123(1):2-9. doi: 10.1067/mod.2003.49. PMID: 12532055.

19. Hajrassie MK, Khier SE. In-vivo and in-vitro comparison of bond strengths of orthodontic brackets bonded to enamel and debonded at various times. Am J Orthod Dentofacial Orthop. 2007 Mar;131(3):384-90. doi: 10.1016/j.ajodo.2005.06.025. PMID: 17346595.

20. Akarsu S, Buyuk SK, Kucukekenci AS. Effects of adhesive systems at different temperatures on the shear bond strength of orthodontic brackets. J Dent Res Dent Clin Dent Prospects. 2019 Spring;13(2):103-108. doi: 10.15171/joddd.2019.016. Epub 2019 Aug 14. PMID: 31592305; PMCID: PMC6773916.
21. Gad MM, Rahoma A, Abualsaud R, Al-Thobity AM, Akhtar S, Siddiqui IA, Al-Harbi FA. Influence of artificial aging and ZrO2 nanoparticle-reinforced repair resin on the denture repair strength. J Clin Exp Dent. 2020 Apr 1;12(4):e354-e362. doi: 10.4317/jced.56610. PMID: 32382385; PMCID: PMC7195688.

22. ISO/TS 11405. 2015. Dentistry – testing of adhesion to tooth structure, 3rd ed. Geneva, Switzerland: International Standard Organization.

23. Fritz UB, Diedrich P, Finger WJ. Self-etching primers–an alternative to the conventional acid etch technique? J Orofac Orthop. 2001 May;62(3):238-45. English, German. doi: 10.1007/pl00001931. PMID: 11417207.

24. Elekdag-Turk S, Turk T, Isci D, Ozkalayci N. Thermocycling effects on shear bond strength of a self-etching primer. Angle Orthod. 2008 Mar;78(2):351-6. doi: 10.2319/122906-537.1. PMID: 18251621.

25. Daub, J., Berzins, D. W., Linn, B. J., & Bradley, T. G. (2006). Bond strength of direct and indirect bonded brackets after thermocycling. The Angle orthodontist, 76(2), 295–300. https://doi.org/10.1043/0003-3219(2006)076[0295:BSODAI]2.0.CO;2

26. Saito K, Sirirungrojying S, Meguro D, Hayakawa T, Kasai K. Bonding durability of using self-etching primer with 4-META/ MMA-TBB resin cement to bond orthodontic brackets. Angle Orthod. 2005 Mar;75(2):260-5. doi: 10.1043/0003-3219(2005)075<0256:BDOUSP>2.0.CO;2. PMID: 15825793.

27. Morresi AL, D'Amario M, Capogreco M, Gatto R, Marzo G, D'Arcangelo C, Monaco A. Thermal cycling for restorative materials: does a standardized protocol exist in laboratory testing? A literature review. J Mech Behav Biomed Mater. 2014 Jan;29:295-308. doi: 10.1016/j.jmbbm.2013.09.013. Epub 2013 Sep 27. PMID: 24135128.

28. Eliasson ST, Dahl JE. Effect of thermal cycling on temperature changes and bond strength in different test specimens. Biomater Investig Dent. 2020 Jan 29;7(1):16-24. doi: 10.1080/26415275.2019.1709470. PMID: 32128509; PMCID: PMC7033714.

29. Helvatjoglu-Antoniades M, Koliniotou-Kubia E, Dionyssopoulos P. The effect of thermal cycling on the bovine dentine shear bond strength of current adhesive systems. J Oral Rehabil. 2004 Sep;31(9):911-7. doi: 10.1111/j.1365-2842.2004.01318.x. PMID: 15369475.

30. Mohammadi E, Pishevar L, Mirzakouchaki Boroujeni P. Effect of food simulating liquids on the flexural strength of a methacrylate and silorane-based composite. PLoS One. 2017 Dec 12;12(12):e0188829. doi: 10.1371/journal.pone.0188829. PMID: 29232691; PMCID: PMC5726734.

31. González-Serrano C, Baena E, Fuentes MV, Albaladejo A, Míguez-Contreras M, Lagravère MO, Ceballos L. Shear bond strength of a flash-free orthodontic adhesive system after thermal aging procedure. J Clin Exp Dent. 2019 Feb 1;11(2):e154-e161. doi: 10.4317/jced.55540. PMID: 30805120; PMCID: PMC6383895.

32. Al-Khatieeb, M. M., Mohammed, S. A., & Al-Attar, A. M. (2015). Evaluation of a new orthodontic bonding system (Beauty Ortho Bond). Group, 22, 23.

33. Demirovic K, Slaj M, Spalj S, Slaj M, Kobasljia S. Comparison of Shear Bond Strength of Orthodontic Brackets Using Direct and Indirect Bonding Methods in Vitro and in Vivo. Acta Inform Med. 2018 Jun;26(2):125-129. doi: 10.5455/aim.2018.26.125-129. PMID: 30061785; PMCID: PMC6029916.
**Figures**

**Figure 1**

A - attaching teeth to the loom (Mecmesim Limited, United Kingdom) from the right side of the tooth. B - attaching teeth to the loom (Mecmesim Limited, United Kingdom) from the front of the tooth.
Figure 2

The results of mean, distribution, maximum, and minimum of debonding forces of E (the experimental group) and C (the control group). The mean of debonding force is indicated by red lines. The distribution of the debonding force is indicated by blue lines, and the maximum and minimum debonding forces are marked with black lines.
Figure 3

The dependencies of maximum debonding force and displacement in each tooth of the experimental group.
**Figure 4**

Dependencies of the maximum debonding force and displacement in each tooth of the control group.
Figure 5

Dependence of the debonding force on the displacement in the experimental group (red line) and the control group (blue line) tubes.