ABSTRACT: Nucleoside analogues are among the most successful bioactive classes of druglike compounds in pharmaceutical chemistry as they are well-known for their numerous effective bioactivities in humans, especially as antiviral and anticancer agents. Coronavirus disease 2019 (COVID-19) is still untreatable, with its causing virus, the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), continuing to wreak havoc on the ground everywhere. This complicated international situation urged all concerned scientists, including medicinal chemists and drug discoverers, to search for a potent anti-COVID-19 drug. Cordycepin (3′-deoxyadenosine) is a known natural adenosine analogue of fungal origin, which could also be synthetically produced. This bioactive phytochemical compound is characterized by several proven strong pharmacological actions that may effectively contribute to the comprehensive treatment of COVID-19, with the antiviral activities being the leading ones. Some new studies predicted the possible inhibitory affinities of cordycepin against the principal SARS-CoV-2 protein targets (e.g., SARS-CoV-2 spike (S) protein, main protease (M(pro) enzyme, and RNA-dependent RNA polymerase (RdRp) enzyme) based on the computational approach. Interestingly, the current research showed, for the first time, that cordycepin is able to potently inhibit the multiplication of the new resistant strains of SARS-CoV-2 with a very minute in vitro anti-SARS-CoV-2 EC₅₀ of about 2 μM, edging over both remdesivir and its active metabolite GS-441524. The ideal pharmacophoric features of the cordycepin molecule render it a typical inhibitor of SARS-CoV-2 replication, with its flexible structure open for most types of derivatization in the future. Briefly, the current findings further support and suggest the repurposing possibility of cordycepin against COVID-19 and greatly encourage us to confidently and rapidly begin its preclinical/clinical evaluations for the comprehensive treatment of COVID-19.

1. INTRODUCTION

About 2 years now and the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) still represents a major global threat and concern to human beings since its first emergence in the famous Chinese city Wuhan. The acute disease caused by this virus, the coronavirus disease 2019 (COVID-19), is also still ongoing with confirmed infections and deaths to more than 260 and 5.18 million, respectively, people worldwide due to this universal pandemic. The increasing evolution of very resistant and virulent novel variants/strains of the COVID-19 virus, especially in the current year 2021, puts a very heavy load on the concerned scientific communities in all of the countries to become much more active in searching for and finding successful medicines and vaccines effective in inhibiting and fighting this irritating virus, along with finding drugs having the abilities to counteract all, or most of, the very severe effects of the COVID-19 on human bodies; thus, searching for efficient comprehensive or dual-action anti-SARS-CoV-2/anti-COVID-19 therapeutic agents will be extremely advantageous as an excellent solution for this untreatable infection and its health sequelae. Tens of new and repurposed promising compounds are nowadays under broad international/multinational investigations (including in vitro, in vivo, and clinical trials) to be biologically evaluated as effective candidate anti-COVID-19 drugs, e.g., remdesivir (GS-5734; FDA-approved administration for emergent use), GS-441524, GS-443902, favipiravir, cyanorona-20, hydroxychloroquine, chloroquine, CoViTris2020, taroxaz-104, Chlo-ViD2020, teriflunomide, leflunomide, ivermectin, arbidol, and colchicine, but none of them has proved its successful eventual broad-spectrum effectiveness until now (i.e., the final results of almost all of these worldwide investigations are not disclosed to date).

Nucleoside and nucleotide analogism is among the top therapeutic options in drug designers’ and medicinal chemists’ minds to combat and stop coronavirus replication inside the human body. In this COVID-19 therapeutic option or
strategy, the natural or synthetic nucleoside/nucleotide analogue uses its resemblance to the normal nucleosides and nucleotides to mislead and trick the SARS-CoV-2 RNA-dependent RNA polymerase (RdRp) through incorporation and embedding in the viral genetic strands instead (i.e., in place) of the normal naturally occurring nucleosides and nucleotides, leading to repeated excessive ambiguous coding and premature termination of mRNA synthesis with the generation of vague RNA strands at the end; these pseudostrands, in turn, form abnormal noninfectious and inactive viral particles, and hence no further replication of the virus occurs (Figure 1). Some of the aforementioned investigational anti-COVID-19 agents, such as remdesivir and cyanorona-20 and their active metabolites GS-441524 and favipiravir, respectively, depend on this tactical mechanism in their inhibitory bioactivities on the SARS-CoV-2 (Figure 2).4−9 Unfortunately, all the four compounds are synthetic and did not give satisfactory results in the in vitro/in vivo anti-SARS-CoV-2 activity assays (except cyanorona-20, which has a significant anti-SARS-CoV-2 EC50 value of 0.45 μM); therefore, searching for more potent natural anti-COVID-19 drugs is very demanding.4−9

Cordycepin is a known unique natural adenosine analogue found and extracted from various species in the fungal kingdom, mainly those from the two genera Cordyceps (e.g., the species Cordyceps militaris) and Ophiocordyceps (e.g., the species Ophiocordyceps sinensis).22−25 Chemically, the compound cordycepin is 3′-deoxyadenosine (3-dA), which is IUPACally named as (2R,3R,5S)-2-(6-amino-9H-purin-9-yl)-5-(hydroxymethyl)oxolan-3-ol or 9-(3-deoxy-β-D-ribofuranosyl)adenine (Figure 3).26 Recently, the natural cordycepin content was biologically increased to more than twofold in the entomopathogenic medicinal fungus C. militaris using newly designed techniques of optimized cultivation and fermentation, which significantly enhanced the biosynthesis of this precious bioactive metabolite.27 Cordycepin could also be easily synthesized from many readily available simple starting materials, such as D-glucose, D-xylose, and adenosine, using different chemical synthetic routes.28,29 Cordycepin molecule is specifically a nucleosidic adenosine molecule that lacks only one

---

**Figure 1.** Representation of the nucleoside/nucleotide analogism approach used for the potent blockade of the SARS-CoV-2 replication in COVID-19 therapy.

**Figure 2.** Chemical structures of remdesivir, GS-441524, cyanorona-20, and favipiravir.
hydroxyl group at the 3′ position of the five-membered ring of its ribose moiety (Figure 3). This extreme structural similarity with the cellular nucleoside adenosine renders cordycepin biologically acting on adenosine receptors and very similar to this pivotal nucleoside in all its biological activities in living organisms. Additionally, this structural analogism also provides camouflage abilities to the cordycepin molecule to be able to antagonist/block and impair/disrupt many of the biological actions of the adenosine molecule itself. Cordycepin has a very broad spectrum of diverse important pharmacological activities since it effectively acts as, for example, antiviral (also in viral myocarditis), antifungal, antibacterial, antitubercular, antimalarial, antiprotozoal, antimicrobial, insecticidal, larvicidal, antinfiammatory, antioxident (and in several oxidative stress statuses of many different diseases), immunomodulatory/immunoregulatory, antileukemic (since it has exhibited promising in vitro cytotoxicities against various leukemic cell lines), antitumor/anticancer (antineoplastic), antiproliferative, antifibrotic, antinflammatory, antitum generic (in cancer therapy), apoptosis inducer (in cancer cells), antihyperglycemic/antidiabetic, antihyperlipidemic (as in cardiovascular diseases and vascular disorders), antihypercholesterolemic, antiarhythmic, antitachy cardiac, coronary vasodilator, antihypertensive, angiogenic, antithrombotic, anticoagulant aggregation, antithrombotic/thrombolytic/fibrinolytic, anti-ischemic (as in myocardial infarction/ischemia and cerebral ischemia injuries), reperfusion therapy, antistroke, hepatoprotective (as in hepatitis and liver cirrhosis), renal functions improver/nephroprotective (renal, fibrosis and chronic kidney disease (CKD)), chondrogenesis promoter, antiarthritis (as in osteo arthritis), antosteoporotic (in bone loss), antihyperuricemic, antirheumatic (as in multiple sclerosis (MS) and rheumatoid arthritis (RA)), intervertebral disc regenerator, bronchial asthma reliever, acute lung inflammation/injury healer (as in acute respiratory distress syndrome (ARDS), cystic fibrosis (CF), and chronic obstructive pulmonary disease (COPD)), pneumoprotective, cough/common cold suppressant, antihypoxic, antidepressant, nootropic, energizer, tonic, stimulant, mood modulator, natural endurance booster, pain killer/analgics, antifatigue, antinociceptive, dysthrophic, sleep depres sor (as in acute colitis conditions), some toxins antidote, and cosmeceutical (in skin photoaging and other cosmeceutical skin/hair problems, also in inflammatory skin conditions/diseases like atopic dermatitis). All of the aforementioned interesting biological activities make cordycepin one of the most important promising phytotherapeutic agents. Cordycepin is already being clinically investigated as a potential effective antileukemic/anticancer chemotherapeutic agent of the anti metabolites class since 1997 until now in several clinical settings worldwide to pass the clinical phases 1 and 2 (e.g., clinical trials NCT00003005 and NCT00709215).  

Most of the previously mentioned diverse biological activities of cordycepin are extremely needed in the comprehensive treatment of COVID-19; this makes cordycepin a very promising potential comprehensive anti-COVID-19 drug rather than being only a candidate SARS-CoV-2 inhibitor (the complementary part that was proven in the current study). The antiviral activities of cordycepin are very potent and broad since they cover several species of human viruses (including almost all virulent RNA viruses, especially those belonging to the genus Flavivirus), e.g., adenoviruses (AVs), dengue virus (DENV), Epstein-Barr virus (EBV), hepatitis B virus (HBV), hepatitis C virus (HCV), Herpes simplex virus 1 and 2 (HSV-1 and HSV-2), human immunodeficiency virus (HIV), human poliovirus (HPV), human rhinovirus (HRV), influenza viruses (IVs), Kyasunar Forest disease virus (KFDV), murine leukemia viruses (MLVs or MuLVs), murine sarcoma viruses (MSVs or MuSVs), Newcastle disease virus (NDV), Omsk hemorrhagic fever virus (OHFV), Powassan virus (POWV), rotaviruses (RV), vaccinia virus (VACC or VV), West Nile virus (WNV), yellow fever virus (YFV), and Zika virus (ZIKV). These strong inhibitory activities extend to some plant viruses, such as cowpea chlorotic mottle virus (CCMV) and tobacco mosaic virus (TMV). The strong antiviral actions of cordycepin are mediated through specific molecular mechanisms that mainly originate due to the extreme structural analogism with the nucleoside adenosine, as this analogism renders the human biological system unable to recognize this molecule, e.g., many enzymes fail to discriminate between it and the endogenous adenosine. Through this camouflage, administered cordycepin could easily be involved in successfully inhibiting/blocking certain several biochemical pathways and reactions in the human body and viral particles; for example, it could cause potent poly(A) polymerase inhibition (i.e., acts as a polyadenylation inhibitor), severe shortening of poly(A) tails, continuous destabilization of mRNAs, strong purine biosynthesis inhibition, and also premature termination of protein synthesis.  

A recently published study sheds light on the possible potent inhibitory effects of cordycepin against SARS-CoV-2 RdRp using an in silico computational molecular docking approach. The evaluation results of this validated simulation protocol clearly revealed the very strong intermolecular interactions with the most vital amino acid residues of the principal active site of the SARS-CoV-2 RdRp, proposing that cordycepin molecule actively interacts with the giant molecule of RdRp (Figure 4). These strong inhibitory interactions were reflected in the resultant relatively low binding energy, which reaches about 8.2 kcal/mol at the best pose of docking, with the formation of a very stable cordycepin−RdRp complex. Interestingly, it was found that the cordycepin molecule strikes and binds to the two pivotal catalytic amino acid residues, Asp760 and Asp761, of the active site of the SARS-CoV-2 RdRp via both strong hydrogen bonds and hydrophobic interactions. Additionally, the cordycepin molecule was found to bind also to some residues that are very close to the catalytic ones in the active site, e.g., Trp617, Asp618, Trp800, and Glu811 residues. These additional intermolecular interactions are mediated via diverse bonds/interactions, such as nonbonding interactions, hydrogen bonds, and hydrophobic interactions.
Herein, we suggest that cordycepin could effectively inhibit the coronaviral replication (i.e., act as a SARS-CoV-2 replication inhibitor) by significantly reducing the number of replicated coronaviral copies, which is expected to result from locking mainly the genomic RNA synthesis occurred via the SARS-CoV-2 RdRp, using the nucleoside analogism strategy (as previously mentioned). In this effective mechanism of anti-RNA action, the nucleoside-like cordycepin molecule is first readily phosphorylated to its mono-, di-, and triphosphate forms (i.e., its nucleotide analogues) intracellularly, then the superactive nucleotide analogue cordycepin triphosphate (Cor-TP) can be easily incorporated into RNA in place of the endogenous chemicosimilar bionucleotide adenosine triphosphate (ATP); this inhibits and terminates transcription elongation and synthesis of viral RNA strands in all stages (i.e., acts as an RNA elongation inhibitor due to the hydroxyl moiety absence at the 3′ position of...
the molecule, this one-hydroxyl group deficiency significantly antagonizes the SARS-CoV-2 RdRp activity as previously mentioned, giving incomplete disrupted premature RNAs in the developing mRNA strands and viral genomes, and finally, this ambiguous coding leads to strong inhibition of SARS-CoV-2 copying/replication and generation of inactive, noninfectious, mutated, useless, nonpathogenic, and nonviral/non-SARS-CoV-2 particles instead of the active, parent, correct, original, infectious, pathogenic, and virulent SARS-CoV-2 particles (Figure 5). This currently presented logical hypothesis, which proposes the potential strong anti-SARS-CoV-2/anti-COVID-19 properties of the cordycepin molecule, was actually tested and evaluated in this current work through a validated anti-SARS-CoV-2 bioassay (along with a full toxicological evaluation). Based on all of the previous promising literature data together with the very interesting in silico and in vitro (i.e., biological) findings of the current research, cordycepin can be repurposed to in vivo evaluate its protective and inhibitory activities specifically against SARS-CoV-2 particles’ invasion (as an anti-RNA virus agent) and to clinically evaluate its protective and inhibitory activities comprehensively against COVID-19 status as a whole (as an anti-COVID-19 condition agent) in COVID-19 patients.

2. RESULTS AND DISCUSSION

2.1. In Silico Predictive Toxicological and Stability Properties of Cordycepin. The prediction of toxicities and adverse/side effects of any new potential medicine is a very critical issue of the drug discovery journey. Interestingly, modern virtual toxicity predictions have several favorable merits over the practical classic toxic dose determinations in living creatures, as they afford, e.g., more rapid data, lower research costs, and a lower number of diverse animal experiments. The data obtained from using the ProTox-II (Prediction of Toxicity of Chemicals) Virtual Laboratory disclosed the predicted relative biosafety of the compound cordycepin (over many investigational potential anti-COVID-19 drugs).46 The results revealed that cordycepin lies in the second toxicity class (toxicity class II) with a predicted lethal dose 50 (LD50) of 8 mg/kg of the body weight (BWt) (with an excellent average similarity of 97.27% and a high prediction accuracy of 72.9%). With a relatively small molecular weight of about 251.24 (which is approaching the mean molecular weight of the dataset, 319.67), cordycepin resides in the molecular weight region of the safest druglike compounds in the dataset of this virtual laboratory (Figure S1). The resulting data also predicted that cordycepin is cytotoxicologically inactive (with a probability percentage of more than 58%) (Table 1 and Figure S2). Interestingly, analysis of all diverse toxicity endpoints using the different ProTox-II toxicity model tests showed that cordycepin has high levels of safety with respect to the diverse organ toxicities and adverse outcome pathways since it exhibits very high percentages of inactivities, reaching 100% in some examples, against almost all of them (e.g., cordycepin is predicted to be extremely immunotoxically inactive with a probability percentage of more than 99%) (Table 1and Figure S2). The only exception is the mutagenicity since cordycepin is predicted to be mutagenic with a moderate probability percentage of 69% (Table 1 and Figure S2). According to the newly established PredMS Application methodology, cordycepin is supposed to be metabolically stable in human liver microsomes with a probability percentage of more than 91%.48 All of the previous predictive toxicological/stability speculations are in line by a significant degree with the previous respective literature data about cordycepin.

2.2. In Vitro Anti-SARS-CoV-2 and Cytotoxic Bioactivities of Cordycepin. Table 2 shows the resulting values from both the in vitro anti-SARS-CoV-2 and cytotoxic bioassays in detail. The SARS-CoV-2 strain used in the anticoronaviral assay is one of the newest strains, the first Variant of Concern from December 2020 (VOC-202012/01), which is a very virulent and resistant strain of the virus. The data demonstrated in the table interestingly disclosed the considerably higher antiviral effectiveness of cordycepin against the newly appeared variants of SARS-CoV-2 in comparison to that of each of the two positive control reference drugs remdesivir and GS-441524 (the placebo drug dimethyl suloxide “DMSO”) showed extremely weak activities, i.e., negligible results). Cordycepin was found to successfully impair and block the entire coronaviral replication/transcription in Vero E6 cells with EC50 considerably smaller

| classification                  | target                                      | prediction | probability |
|---------------------------------|---------------------------------------------|------------|-------------|
| organ toxicity                  | hepatotoxicity                              | inactive   | 0.63        |
| toxicity endpoints              | carcinogenicity                              | inactive   | 0.68        |
| toxicity endpoints              | immunotoxicity                              | inactive   | 0.99        |
| toxicity endpoints              | mutagenicity                                | active     | 0.69        |
| toxicity endpoints              | cytotoxicity                                | inactive   | 0.99        |
| Tox21-nuclear receptor signaling pathways | aryl hydrocarbon receptor (AhR) | inactive | 0.89        |
| Tox21-nuclear receptor signaling pathways | androgen receptor (AR)                    | inactive   | 0.99        |
| Tox21-nuclear receptor signaling pathways | androgen receptor ligand-binding domain (AR-LBD) | inactive | 1.0         |
| Tox21-nuclear receptor signaling pathways | aromatase                                   | inactive   | 0.91        |
| Tox21-nuclear receptor signaling pathways | estrogen receptor (ER)                     | inactive   | 0.98        |
| Tox21-nuclear receptor signaling pathways | estrogen receptor ligand-binding domain (ER-LBD) | inactive | 1.0         |
| Tox21-nuclear receptor signaling pathways | peroxisome proliferator-activated receptor γ (PPAR-γ) | inactive | 0.80        |
| Tox21-stress response pathways | nuclear factor (erythroid-derived 2)-like 2/antioxidant responsive element (nrf2/ARE) | inactive | 0.99        |
| Tox21-stress response pathways | heat shock factor response element (HSE)   | inactive   | 0.99        |
| Tox21-stress response pathways | mitochondrial membrane potential (MMP)     | inactive   | 0.97        |
| Tox21-stress response pathways | phosphoprotein (tumor suppressor) p53       | inactive   | 0.81        |
| Tox21-stress response pathways | ATPase family AAA domain-containing protein 5 (ATAD5) | inactive | 0.71        |
Table 2. Anti-SARS-CoV-2/Anti-COVID-19 Activities (along with Cytotoxicities) of the Target Drug Cordycepin (Using Both Remdesivir and GS-441524 as the Positive Control/Reference Drugs, and DMSO as the Negative Control/Placebo Drug) against SARS-CoV-2 (VOC-202012/01 strain) in Vero E6 Cells

| classification | compound name | CC_{50} (µM) | 100% CPE inhibitory concentration (CPEIC_{100}) | 50% reduction in infectious virus (EC_{50}) | 50% reduction in viral RNA copy (EC_{50}) |
|----------------|----------------|--------------|-----------------------------------------------|------------------------------------|-------------------------------------|
| target compound | cordycepin | >100 | 5.98 ± 0.41 | 2.01 ± 0.12 | 2.35 ± 0.15 |
| reference drugs | remdesivir | >100 | 25.17 ± 2.51 | 21.00 ± 1.97 | 22.92 ± 1.99 |
| | GS-441524 | >100 | 17.40 ± 1.83 | 15.60 ± 0.76 | 16.04 ± 0.81 |
| placebo solvent | DMSO | >100 | >100 | >100 | >100 |

“CC_{50} or 50% cytotoxic concentration is the concentration of the tested compound that kills half of the cells in an uninfected cell culture. CC_{50} was determined with serially diluted compounds in Vero E6 cells at 48 h post incubation using the CellTiter-Glo Luminescent Cell Viability Assay (Promega).”

50% CPE inhibitory concentration is the lowest concentration of the tested compound that causes 50% inhibition of the cytopathic effects (CPE) of SARS-CoV-2 VOC-202012/01 virus in Vero E6 cells under increasing concentrations of the tested compound at 48 h post infection. Compounds were serially diluted from 100 µM concentration. “EC_{50} or 50% effective concentration is the concentration of the tested compound that is required for 50% reduction in infectious SARS-CoV-2 VOC-202012/01 virus particles in vitro. EC_{50} is determined by infectious virus yield in culture supernatant at 48 h post infection (log_{10} TCID_{50}/mL).”

The nucleoside analogue to a more biocompatible form of nucleotide analogue (the more required form) without any metabolic hydroxylation of the 3′ position of the molecule (as previously mentioned, the nucleotide analogue Cor-TP could be successfully merged into viral RNA and strongly inhibits the transcription elongation and RNA generation because of the absence of a hydroxyl moiety at the 3′ position). Clinically, when cordycepin enters the blood circulation after absorption from oral administration, a portion of its amount may be broken down to other structurally related nucleosides, mainly 3′-deoxyinosine by the enzymes adenosine deaminases (which are present in blood plasma, erythrocytes, and vessel walls). It was previously thought that 3′-deoxyinosine is a biologically inactive metabolite; thus, it was clinically suggested to coadminister a potent adenosine deaminase inhibitor, e.g., the antimebolite pentostatin (PTN), with cordycepin to protect it from deamination by the adenosine deaminase since this inhibitor mimics the nucleoside analogue cordycepin and therefore inhibits the enzyme adenosine deaminase and interferes with the ability of the human biological system to break down cordycepin molecules. Surprisingly, some recent studies refuted the need for adenosine deaminase inhibitor coadministration to achieve the ideal benefits of cordycepin since these studies proved that 3′-deoxyinosine could be intracellularly reconverted to cordycepin again in the form of the very active Cor-TP; thus, the metabolite 3′-deoxyinosine previously considered inactive is now considered as one of the biological metabolites that are well-responsible for the therapeutic actions of cordycepin after its oral administration, and the use of an adenosine deaminase inhibitor is no more a must (Figure 6). Another suggestion (in addition to the optional use of a potent adenosine deaminase inhibitor) to increase the blood level of cordycepin and resist any rapid metabolism is the use of a nonspecific inhibitor of nucleoside transporters (including adenosine nucleoside transporters, like dipyridamole (DPM), to solve this issue and allow cordycepin accumulation in the human blood, since this nucleoside transport inhibitor will block the cellular reuptake of cordycepin into platelets, erythrocytes, and endothelial cells, resulting in elevated concentrations of cordycepin extracellularly (i.e., extended clinical half-life of cordycepin). This, on the other hand, causes an increase in coronary vasodilatation, which is an extra beneficial effect needed in the comprehensive treatment of COVID-19. It should be taken into consideration
that DPM has very useful biological effects that can help to prevent or suppress the SARS-CoV-2 exacerbation of respiratory/cardiovascular status and combat the complications of the comprehensive COVID-19 condition. Additionally, the two adjuvant drugs DPM and PTN can also have some inhibitory activities against the coronaviral RNA synthesis and replication.

Cordycepin is an ideal druglike compound since it fully obeys Lipinski’s rule of five (Ro5) with no violations. Cordycepin is also a highly biocompatible molecule with enhanced abilities to pass through biologic membranes when compared with the adenosine molecule due to the relatively higher lipophilic characters resulting from the shortage of a hydroxyl group (i.e., the cordycepin molecule is less hydrophilic than the adenosine molecule). Metabolic similarity with the biosimilar adenosine molecule greatly helps cordycepin in tricking the human biological system and doing its intended therapeutic roles. The experimental findings of the present research are in full agreement with the previously proven effects of cordycepin to partially inhibit and impair mRNA translation. These current findings are also in agreement with almost all of the computational predictions previously presented and discussed in Section 1.

3. CONCLUSIONS AND FUTURE RECOMMENDATIONS

COVID-19 treatment is still a critical challenge to all respective scientists. Cordycepin is a promising potential comprehensive anti-COVID-19 agent that should be put under the microscope in the coming days. Some new studies reported the in silico inhibitory activities of cordycepin on SARS-CoV-2 spike (S) protein and main protease (MPro) enzyme, and, recently, another computational study showed the in silico inhibitory activities of cordycepin on the SARS-CoV-2 RdRp enzyme. In light of the findings of the previous in silico studies and the current in vitro study, it is proven that the cordycepin molecule has interesting superiority and advantage over most of the other investigational anti-SARS-CoV-2 molecules, especially those acting only on the S protein and/or MPro enzyme, as, first, it acts via the expected synergistic triple anti-SARS-CoV-2 mode of action (i.e., it acts on the three different target proteins, thus inhibiting all the three major stages of the SARS-CoV-2 infection in humans, which are the viral entry, replication/multiplication, and pathogenic phases) and, second, it acts as nonspecific/nonselective anti-SARS-CoV-2, i.e., it is capable of acting on all SARS-CoV-2 strains and variants since its anti-SARS-CoV-2 bioactivity does not depend only on the predicted action on the changeable and mutated S protein (cordycepin has general broad-spectrum anti-SARS-CoV-2 activities effective on all SARS-CoV-2 strains and variants with different mutations).

The expected comprehensive nature of cordycepin in COVID-19 treatment mainly comes from two practically proven pathways. First, it can inhibit the replication and survival of the COVID-19-causing virus itself with potent broad-spectrum activities (including activities against the most recent strains of SARS-CoV-2), which reach an EC50 of about 2 μM. This anti-SARS-CoV-2 action of cordycepin is mainly due to its chemical similarity/analogism with the human adenosine. During viral replication, the SARS-CoV-2 RdRp incorporates Cor-TP into the newly synthesized RNA strands instead of using real ATP. Consequently, when the SARS-CoV-2 RdRp attempts to copy the destabilized RNA containing Cor-TP, it either falsely interprets it or fails to interpret it. This interrupted and hindered interpretation and the consequent errors in the viral genetic code cause a very significant number of mutations in all downstream coronaviral-2 copies that much exceeds the threshold the virus can survive, an effect known in virology as lethal mutagenesis or viral error catastrophe. Second, it can mitigate and relieve COVID-19 characteristic severe symptoms and sequelae that are mainly related to the SARS-CoV-2 invasion of the respiratory and cardiovascular systems of the patients since this attack results primarily in acute biological disruptions of immunologic/inflammatory origin (see the numerous diverse and broad pharmacological activities of cordycepin in Section 1).
Promisingly, the net distinction, in almost all anti-COVID-19 parameters/properties/activities, of cordycepin over the potent antiviral agents remdesivir and GS-441524 supports cordycepin candidacy to be a superior potential comprehensive therapeutic agent against COVID-19. It is also worth mentioning that the cordycepin molecule has more than 10 sites that are considered as very active positions for chemical reactions; thus, hundreds of possible derivatives with required enhanced pharmacokinetic and/or pharmacodynamic properties can be designed and synthesized based on the biocompatible scaffold of this golden multitask molecule, which belongs to the nucleoside analogues’ class of antiviral/antineoplastic therapeutic agents. Last but not the least, it is highly suggested to the scientific community to start the cordycepin repurposing journey against COVID-19 by conducting extensive worldwide preclinical and clinical studies to seriously evaluate the efficacy and safety of cordycepin (as a monotherapy or, optionally, as either a double-combination therapy with DPM or a triple-combination therapy with both DPM and PTN) to be used for the comprehensive treatment and prevention of all types of COVID-19 infections.

4. MATERIALS AND METHODS

4.1. In Silico Predictive Toxicological and Stability Properties of Cordycepin. There is an increasing need for such type of in silico toxicological and metabolic studies for cordycepin due to their absence in the previous literature. Therefore, a complete simulative toxicological study of cordycepin was implemented in the current work using the ProTox-II web server, which is a speculative web-based laboratory for the prediction of toxicities/adverse effects of small chemical molecules.\(^{48}\) ProTox-II web server computationally integrates the principles of molecular similarity, fragment propensities, fragment similarity-based cluster cross-validation machine learning, and most frequent features of artificial intelligence (AI) with each other, establishing a diverse collection of more than 33 models for the analysis and prediction of diverse toxicity endpoints, e.g., cytotoxicity, general acute toxicity, organ toxicity “mainly hepatotoxicity”, carcinogenicity, immunotoxicity, mutagenicity, adverse outcomes (Tox21) pathways, and several toxicity targets.\(^{48,58}\) Oral toxic doses in this validated virtual laboratory are given as LD50 values in mg/kg BWt.\(^{38,58–60}\) Practically, the LD50 can be defined as the experimental dose at which 50% of test subjects die upon exposure to a compound.\(^{48,58–60}\) This interesting virtual web server classifies the different chemicals into six classes of decreasing order of toxicity (according to the Globally Harmonized System of Classification and Labeling of Chemicals “GHS”) as follows: toxicity class I “fatal if swallowed” (LD50 ≤ 5 mg/kg BWt), toxicity class II “fatal if swallowed” (5 mg/kg BWt < LD50 ≤ 50 mg/kg BWt), toxicity class III “toxic if swallowed” (50 mg/kg BWt < LD50 ≤ 300 mg/kg BWt), toxicity class IV “harmful if swallowed” (300 mg/kg BWt < LD50 ≤ 2000 mg/kg BWt), toxicity class V “may be harmful if swallowed” (2000 mg/kg BWt < LD50 ≤ 5000 mg/kg BWt), and toxicity class VI “nontoxic” (LD50 > 5000 mg/kg BWt).\(^{48,58–60}\) A small complementary simulative/spectulative study was also done to determine the expected degree of metabolic stability of cordycepin in its free form, using the PredMS Application.\(^{49}\) This application is a new web server utilized to predict the metabolic stability of a certain chemical compound, where the compound is said to be either stable (if ≥50% was predicted to remain intact after 30 min) or unstable (if <50% was predicted to remain intact after 30 min) in human liver microsomes.\(^{49}\)

4.2. In Vitro Anti-SARS-CoV-2 and Cytotoxic Bioactivities of Cordycepin. This credible and robust in vitro anti-COVID-19 test (including the cytotoxicity assay) is based mainly upon the validated procedures of Rabie.\(^{8,9,12–15}\) The complete procedures were carried out in a specialized biosafety level 3 (BSL-3) laboratory. The assayed new strain of SARS-CoV-2 virus, VOC-202012/01, was isolated from the fresh nasopharynx aspirate and throat swab from a 50-year-old man with confirmed COVID-19 infection using Vero E6 cells (ATCC CRL-1586) on September 21, 2021. Stock virus (10^{7.25} TCID_{50}/mL) was prepared after three serial passages in Vero E6 cells in infection media (Dulbecco’s modified Eagle medium (DMEM) supplemented with 4.5 g/L d-glucose, 100 mg/L sodium pyruvate, 2% fetal bovine serum (FBS), 100 000 U/L penicillin-streptomycin, and 25 mM N-(2-hydroxyethyl)-piperazine-N’-ethanesulfonic acid (HEPES)). Cordycepin (3’-deoxyadenosine, CAS registry number: 73-03-0) and remdesivir (GS-5734, CAS registry number: 1809249-37-3) were purchased from Biosyn Carbosynth (Carbosynth Ltd., Berkshire, U.K.) (for cordycepin, product code: N02930, purity: ≥98%; for remdesivir, product code: AG170167, purity: ≥98%), while the other reference compound GS-441524 (CAS registry number: 1191237-69-0) was purchased from MedChemExpress (MCE, MedChemExpress LLC, New Jersey, U.S.A.) (catalog number: HY-103586, purity: 99.77%). The ultrapure solvent DMSO (CAS registry number: 67-68-5) was purchased from a local distributor, El-Gomhouria Company For Drugs (El-Gomhouria Co. For Trading Drugs, Chemicals & Medical Supplies, Mansoura Branch, Egypt) (purity: ≥99.9% “anhydrous”). Preliminary pilot assays were performed mainly to determine the best concentration of cordycepin, remdesivir, and GS-441524 to start the in vitro anti-COVID-19 and cytotoxicity tests with. Accordingly, the stocks of the tested compounds were accurately prepared by dissolving each of the three compounds in DMSO to obtain a 100 μM concentration of each compound. Additionally, DMSO was used for the purpose of a negative control comparison to make the study placebo-controlled. To evaluate the in vitro anti-SARS-CoV-2 activity of the target compound, cordycepin, in comparison to that of each of the two positive control drugs, remdesivir and GS-441524, along with that of the negative control solvent, DMSO, Vero E6 cells were pretreated with the four compounds diluted in infection media for 1 h prior to infection by the new variant of the SARS-CoV-2 virus at MOI = 0.02. The four tested compounds were maintained with the virus inoculum during the 2 h incubation period. The inoculum was removed after incubation, and the cells were overlaid with infection media containing the diluted test compounds. After 48 h of incubation at 37 °C, supernatants were immediately collected to quantify viral loads by TCID_{50} assay or quantitative real-time reverse transcription polymerase chain reaction (RT-PCR) “qRT-PCR” (TaqMan Fast Virus 1-Step Master Mix). Viral loads in this assay were fitted in a logarithmic scale (log_{10} TCID_{50}/mL and log_{10} viral RNA copies/mL), not in a linear scale, under increasing concentrations of the tested compounds. Four-parameter logistic regression (GraphPad Prism) was used to fit the dose–response curves and determine the EC_{50} of the tested compounds that inhibit SARS-CoV-2 viral replication (CPEIC_{50} was also determined for each compound). Cytotoxicity of each of the four tested compounds was also evaluated in Vero E6 cells using the CellTiter-Glo Luminescent Cell Viability Assay (Promega). The final results were represented as the mean (μ) ± the standard deviation (SD) from the triplicate biological experi-
ments. Statistical analysis was performed using SkanIt 4.0 Research Edition software (Thermo Fisher Scientific) and Prism V5 software (GraphPad). All reported data were significant at p < 0.05.

**ASSOCIATED CONTENT**

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.1c05998.

Graphic comparison of the molecular weight of cordycepin with the mean value of molecular weights of all the dataset compounds (Figure S1); screenshot of the output table for the ProTox-II Toxicity Model Report (Figure S2) (PDF)

**AUTHOR INFORMATION**

Corresponding Author

Amgad M. Rabie — Dr. Amgad Rabie’s Research Lab, for Drug Discovery (DARLD), Mansoura 35511, Egypt; Pharmaceutical Organic Chemistry Department, Faculty of Pharmacy, Mansoura University, Mansoura 35516, Egypt; orcid.org/0000-0003-3681-114X; Phone: 002-01019733188; Email: amgadpharmacist1@yahoo.com, dr.amgadrabie@gmail.com

Complete contact information is available at: https://pubs.acs.org/10.1021/acsomega.1c05998

**Notes**

The author declares no competing financial interest.

**ACKNOWLEDGMENTS**

This new discovery did not receive any external funding. The author gratefully thanks and deeply acknowledges everyone who helped to make this new discovery and work coming out to light.

**REFERENCES**

(1) Hui, D. S.; Azhar, E. I.; Madani, T. A.; Ntoumi, F.; Kock, R.; Dar, O.; Ippolito, G.; Mughith, T. D.; Memish, Z. A.; Drosten, C.; Zulma, A.; Peterse, E. The continuing 2019-nCoV epidemic threat of novel coronaviruses to global health — The latest 2019 novel coronavirus outbreak in Wuhan, China. Int. J. Infect. Dis. 2020, 91, 264–266.

(2) COVID-19 Map. Available from Johns Hopkins Coronavirus Research Center, https://coronavirus.jhu.edu/map.html (accessed Nov 25, 2021).

(3) Jiang, S.; Du, L.; Shi, Z. An emerging coronavirus causing pneumonia outbreak in Wuhan, China: calling for developing therapeutic and prophylactic strategies. Emerging Microbes Infect. 2020, 9, 275–277.

(4) Moirangthem, D. S.; Surbala, L. Remdesivir (GS-5734) in COVID-19 Therapy: The Fourth Chance. Curr. Drug Targets 2021, 22, 1346–1356.

(5) Yan, V. C.; Muller, F. L. Advantages of the Parent Nucleoside GS-441524 over Remdesivir for Covid-19 Treatment. ACS Med. Chem. Lett. 2020, 11, 1361–1366.

(6) Brunotte, L.; Zheng, S.; Mecate-Zambrano, A.; Tang, J.; Ludwig, S.; Rescher, U.; Schloer, S. Combination Therapy with Fluoxetine and the Nucleoside Analog GS-441524 Exerts Synergistic Antiviral Effects against Different SARS-CoV-2 Variants In Vitro. Pharmacuticals 2021, 13, No. 1400.

(7) Cai, Q.; Yang, M.; Liu, D.; Chen, J.; Shu, D.; Xia, J.; Liao, X.; Gu, Y.; Cai, Q.; Yang, Y.; Chen, C.; Li, X.; Peng, L.; Huang, D.; Zhang, J.; Zhang, S.; Wang, F.; Liu, J.; Chen, L.; Chen, S.; Wang, Z.; Zhang, Z.; Cao, R.; Zhong, W.; Liu, Y.; Liu, L. Experimental Treatment with Favipiravir for COVID-19: An Open-Label Control Study. Engineering 2020, 6, 1192–1198.

(8) Rabie, A. M. Discovery of (E)-N-(4-cyanobenzylidene)-6-fluoro-3-hydroxyppyrazine-2-carboxamide (cyanorona-20): the first potent and specific anti-COVID-19 drug. Chem. Pap. 2021, 75, 4669–4685.

(9) Rabie, A. M. Cyanorona-20: The first potent anti-SARS-CoV-2 agent. Int. Immunopharmacol. 2021, 98, No. 107831.

(10) Ip, A.; Ahn, J.; Zhou, Y.; Goy, A. H.; Hansen, E.; Pecora, A. L.; Sinclair, B. A.; Bednarz, U.; Marafelis, M.; Sawczuk, I. S.; Underwood, J. P.; III; Walker, D. M.; Prasad, R.; Sweeney, R. L.; Ponce, M. G.; La Capra, S.; Cunningham, F. J.; Calise, A. G.; Pulver, B. L.; Ruocco, D.; Mojares, G. E.; Eagan, M. P.; Zientz, K. L.; Mastroyiakios, P.; Goldberg, S. L. Hydroxychloroquine in the treatment of outpatients with mildly symptomatic COVID-19: a multi-center observational study. BMC Infect. Dis. 2021, 21, No. 72.

(11) Wang, M.; Cao, R.; Zhang, L.; Yang, X.; Liu, J.; Xu, M.; Shi, Z.; Hu, Z.; Zhong, W.; Xiao, G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. Cell Res. 2020, 30, 269–271.

(12) Rabie, A. M. Two antioxidant 2,5-disubstituted-1,3,4-oxadiazoles (CoViTris2020 and ChloViD2020): successful repurposing against COVID-19 as the first potent multitarget anti-SARS-CoV-2 drugs. New J. Chem. 2021, 45, 761–771.

(13) Rabie, A. M. CoViTris2020 and ChloViD2020: a striking new hope in COVID-19 therapy. Mol. Diversity 2021, 25, 1839–1854.

(14) Rabie, A. M. Potent toxic effects of Taroxaz-104 on the replication of SARS-CoV-2 particles. Chem.- Biol. Interact. 2021, 343, No. 109480.

(15) Rabie, A. M. Discovery of Taroxaz-104: The first potent antidote of SARS-CoV-2 VOC-202012/01 strain. J. Mol. Med. 2021, 124, No. 131106.

(16) Kaur, H.; Saras, P.; Bhattacharya, A.; Sharma, S.; Chhimpa, N.; Prajapat, M.; Prakash, A.; Kumar, S.; Singh, A.; Singh, R.; Avti, P.; Thota, P.; Medhi, B. Efficacy and safety of dihydroorotate dehydrogenase (DHODH) inhibitors “leflunomide” and “teriflunomide” in Covid-19: A narrative review. Eur. J. Pharmacol. 2021, 906, No. 174233.

(17) Rabie, A. M. Teriflunomide: A possible effective drug for the comprehensive treatment of COVID-19. Curr. Res. Pharmacol. Drug Discovery 2021, 2, No. 100055.

(18) Caly, L.; Druce, J.; D.; Catton, M. G.; Jans, D. A.; Wagstaff, K. M. The FDA-approved drug ivmecin inhibits the replication of SARS-CoV-2 in vitro. Antiviral Res. 2020, 178, No. 104778.

(19) Wang, X.; Cao, R.; Zhang, H.; Hu, J.; Xu, M.; Hu, L.; Li, Y.; Zhao, L.; Li, W.; Sun, X.; Yang, X.; Shi, Z.; Deng, F.; Hu, Z.; Zhong, W.; Wang, M. The anti-influenza virus drug, arbidol is an efficient inhibitor of SARS-CoV-2 in vitro. Cell Discovery 2020, 6, No. 28.

(20) Tardif, J.-C.; Bouabdallahoui, N.; L’Allier, P. L.; Gaudet, D.; Shah, B.; Pillinger, M. H.; Lopez-Sendon, J.; Da Luz, P.; Verret, L.; Audet, S.; Dupuis, J.; Denault, A.; Pelletier, M.; Tessier, P. A.; Samson, S.; Fortin, D.; Tardif, J.-D.; Busseul, D.; Goulet, E.; Lacoste, C.; Dubois, A.; Joshi, A. Y.; Waters, D. D.; Hsue, P.; Lepor, N. E.; Lesage, F.; Santuret, N.; Roy-Cavel, E.; Bassevit, Z.; Orfano, A.; Stamatescu, G.; Grégoire, J. C.; Busque, L.; Lavallée, C.; Hétu, P.-O.; Paquette, J.-S.; Deftereos, S. G.; Levesque, S.; Cossette, M.; Nozza, A.; Chabot-Blanchet, M.; Dubé, M.-P.; Guertin, M.-C.; Boivin, G.; for the COLCORCORAN Investigators. Colchicine for community-treated patients with COVID-19 (COLCORON): a phase 3, randomised, double-blinded, adaptive, placebo-controlled, multicentre trial. Lancet Respir. Med. 2021, 9, 924–933.

(21) Chien, M.; Anderson, T. K.; Jochusch, S.; Tao, C.; Li, X.; Kumar, S.; Russo, J. J.; Kirchdoerfer, R. N.; Ju, J. Nucleotide Analogues as Inhibitors of SARS-CoV-2 Polymerase, a Key Drug Target for COVID-19: A narrative review. J. Mol. Med. 2020, No. 100055.

(22) Cunningham, K. G.; Manson, W.; Spring, F. S.; Hutchinson, S. A. Cordycepin, a Metabolic Product isolated from Cultures of Cordyceps militaris (Linn.) Link. Nature 1950, 156, 949.

(23) Zhou, X.; Luo, L.; Dressel, W.; Shadier, G.; Krumbeig, D.; Schmidtke, P.; Zepf, F.; Meyer, C. U. Cordycepin is an Immunoregulatory Active Ingredient of Cordyceps sinensis. Am. J. Chin. Med. 2008, 36, 967–980.
Entomopathogenic Fungus

(24) Bibi, S.; Wang, Y.-B.; Tang, D.-X.; Kamal, M. A.; Yu, H. Prospects for Discovering the Secondary Metabolites of Cordyceps Senus Lato by the Integrated Strategy. Med. Chem. 2021, 17, 97–120.

(25) Yue, K.; Ye, M.; Zou, Z.; Sun, W.; Lin, X. The genus Cordyces: a chemical and pharmacological review. J. Pharm. Pharmacol. 2013, 65, 474–493.

(26) Cordycepin. PubChem CID: 6303, 2021. Available from PubChem, https://pubchem.ncbi.nlm.nih.gov/compound/6303 (accessed Oct 6, 2021).

(27) Wang, Y.; Yang, Z.; Bao, D.; Li, B.; Yin, X.; Wu, Y.; Chen, H.; Tang, G.; Li, N.; Zou, G. Improving Hypoxia Adaption Causes Distinct Effects on Growth and Bioactive Compounds Synthesis in an Entomopathogenic Fungus Cordyceps militaris. Front. Microbiol. 2020, 11, No. 998436.

(28) Li, Q.; Yang, R.; Ruan, Z.; Hu, T.; Ding, H.; Xiao, Q. Total Synthesis of Cordycepin. Chin. J. Org. Chem. 2013, 33, 1340–1344.

(29) Huang, S.; Liu, H.; Sun, Y.; Chen, J.; Li, X.; Xu, J.; Hu, Y.; Li, Y.; Deng, Z.; Zhong, S. An effective and convenient synthesis of cordycepin from adenine. Chem. Pap. 2018, 72, 149–160.

(30) Adenosine. PubChem CID: 60961, 2021. Available from PubChem, https://pubchem.ncbi.nlm.nih.gov/compound/60961 (accessed Oct 6, 2021).

(31) Tan, L.; Song, X.; Ren, Y.; Wang, M.; Guo, C.; Guo, D.; Gu, Y.; Li, Y.; Cao, Z.; Deng, Y. Anti-inflammatory effects of cordycepin: A review. Phytother. Res. 2021, 35, 1284–1297.

(32) Ashraf, S. A.; Elkalifa, A. E. O.; Siddiqui, A. J.; Patel, M.; Awadellekareem, A. M.; Snoussi, M.; Ashraf, M. S.; Adnan, M.; Hadi, S. Cordycepin for Health and Wellbeing: A Potent Bioactive Metabolite of an Entomopathogenic Medicinal Fungus Cordyceps with Its Nutraceutical and Therapeutic Potential. Molecules 2020, 25, No. 2735.

(33) Qin, P.; Li, X.; Yang, H.; Wang, Z.-Y.; Lu, D. Therapeutic Potential and Biological Applications of Cordyceps and Metabolic Mechanisms in Cordycepin-Producing Fungi. Molecules 2019, 24, No. 2231.

(34) Das, K. S.; Masuda, M.; Sakurai, A.; Sakakibara, M. Medicinal uses of the mushroom Cordyceps militaris: Current state and prospects. Fitoterapia 2010, 81, 961–968.

(35) Li, B.; Hou, Y.; Zhu, M.; Bao, H.; Nie, J.; Zhang, G. Y.; Shan, L.; Yao, Y.; Du, K.; Yang, H.; Li, M.; Zheng, B.; Xu, X.; Xiao, C.; Du, J. 5′-Deoxyadenosine (Cordycepin) Produces a Rapid and Robust Inhibitor of Cellular RNA Synthesis. Proc. Natl. Acad. Sci. U. S. A. 1974, 71, 2008–2012.

(36) White, J. L.; Dawson, W. O. Effect of Cordycepin Triphosphate on In Vitro RNA Synthesis by Plant Viral Replicases. J. Virol. 1979, 29, 811–814.

(37) Dawson, W. O. Tobacco mosaic virus protein synthesis is correlated with double-stranded RNA synthesis and not single-stranded RNA synthesis. Virology 1983, 125, 314–323.

(38) Verma, A. K. Cordycepin: a bioactive metabolite of Cordyceps militaris and polyadenylation inhibitor with therapeutic potential against COVID-19. J. Biomed. Struct. Dyn. 2020, 1–8, DOI: 10.1080/07391102.2020.1850352.

(39) Bibi, S.; Hasan, M. M.; Wang, Y.-B.; Papadakos, S. P.; Yu, H. Cordycepin as a Promising Inhibitor of SARS-CoV-2 RNA Dependent RNA Polymerase (RdRp). Curr. Med. Chem. 2022, 29, 152–162.

(40) ProTox-II Virtual Laboratory. https://tox-new.charite.de/protox_II (accessed Oct 5, 2021).

(41) PredMS Application. https://predms.netlify.app (accessed Oct 5, 2021).

(42) Lee, J. B.; Radhi, M.; Cipolla, E.; Gandhi, R. D.; Sarmad, S.; Zgair, A.; Kim, T. H.; Feng, W.; Qin, C.; Adrower, C.; Ortori, C. A.; Barrett, D. A.; Kong, L.; Fischer, P. M.; de Moor, C. H.; Gershkovitch, P. A novel nucleoside rescue metabolic pathway may be responsible for therapeutic effect of orally administered cordycepin. Sci. Rep. 2019, 9, No. 15760.

(43) Li, G.; Nakagome, I.; Hirono, S.; Itoh, T.; Fujiwara, R. Inhibition of adenosine deaminase (ADA)-mediated metabolism of cordycepin by natural substances. Pharmacol. Res. Perspect. 2015, 3, No. e00121.

(44) Rottenberg, M. E.; Masocha, W.; Ferella, M.; Pettit-Assis, F.; Goto, H.; Kristenson, K.; McCaffrey, R.; Wigzell, H. Treatment of African Trypanosomiasis with Cordycepin and Adenosine Deaminase Inhibitors in a Mouse Model. J. Infect. Dis. 2005, 192, 1658–1665.

(45) Rodman, L. E.; Farnell, D. R.; Coyne, J. M.; Allan, P. W.; Hill, D. L.; Duncan, K. L. K.; Tomaszewski, J. E.; Smith, A. C.; Page, J. G. Toxicity of Cordycepin in Combination with the Adenosine Deaminase Inhibitor 2′-Deoxycoformycin in Beagle Dogs. Toxicol. Appl. Pharmacol. 1997, 147, 39–45.

(46) Fata-Hartley, C. L.; Palmenberg, A. C. Dipyridamole Reversibly Inhibits Mengovirus RNA Replication. J. Virol. 2005, 79, 11062–11070.

(47) Wong, Y. Y.; Moon, A.; Duffin, R.; Barthet-Barateig, A.; Meijer, H. A.; Clemens, M. J.; de Moor, C. H. Cordycepin Inhibits Protein Synthesis and Cell Adhesion through Effects on Signal Transduction. J. Biol. Chem. 2010, 285, 2610–2621.

(48) Verma, A. K.; Aggarwal, R. Repurposing potential of FDA-approved and investigational drugs for COVID-19 targeting SARS-CoV-2 spike and main protease and validation by machine learning algorithm. Chem. Biol. Drug Des. 2021, 97, 836–853.

(49) Suwannarach, N.; Kumla, J.; Sujarit, K.; Pattananandecha, T.; Saenjum, C.; Lumyong, S. Natural Bioactive Compounds from Fungi as Potential Candidates for Protease Inhibitors and Immunomodulators to Apply for Coronavirus. Molecules 2020, 25, No. 1800.

(50) Banerjee, P.; Eckert, A. O.; Schrey, A. K.; Preissner, R. ProTox-II: a webserver for the prediction of toxicity of chemicals. Nucleic Acids Res. 2018, 46, W257–W263.

(51) Drwal, M. N.; Banerjee, P.; Dunkel, M.; Wettig, M. R.; Preissner, R. ProTox-II: a web server for the in silico prediction of rodent oral toxic. Nucleic Acids Res. 2014, 42, W53–W58.

(52) Balenrek, P.; Dehnbostel, F. O.; Preissner, R. Prediction Is a Balancing Act: Importance of Sampling Methods to Balance Sensitivity and Specificity of Predictive Models Based on Imbalanced Chemical Data Sets. Front. Chem. 2018, 6, No. 362.