**Monoporella projecta** (Cheilostomata: Bryozoa), a New Uniserial Species from the Continental Shelf and Slope of Japan

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*Monoporella projecta* sp. nov. from the continental shelf and slope east of the Boso Peninsula, Japan adds to the bryozoan fauna along the northwestern Pacific coast, where more than 10 species of this genus have been recorded. Colonies of this species are uniserial encrusting in growth form, and autozooids are large and much inflated with a projecting orificial rim. These colony and zooidal characteristics may be related to the deep-water environment.

**Key Words:** Bryozoa, *Monoporella*, uniserial, projected orificial rim, Japan.

**Introduction**

*Monoporella* Hincks, 1881 is a long-duration genus of cheilostomatous bryozoans, recorded from the late Cretaceous to the Recent. Thirteen living species have been assigned to this genus (Bock 2016), including six from the Aleutian Islands (Dick 2008) and three from Philippine region (Canu and Bassler 1929; Tilbrook 2006). Dick (2008) noted that the high diversity of *Monoporella* in the Aleutian archipelago might represent either a relict of past high diversity occurring broadly around the North Pacific rim, or a local radiation restricted to the western Aleutians.

A number of different *Monoporella* species also occur in Japan. Fossil material from Miocene to Pleistocene deposits in Japan have been reported as *Monoporella* *fimbriata* Canu and Bassler, 1929 (Sakakura 1935; Kataoka 1961; Hayami 1970, 1975, 1976; Arakawa 1995), although these identifications need reexamination (Dick 2008). *Monoporella* specimens from Japan studied by Harmer (1926) were re-examined by Tilbrook (2006), who commented that they represent two species distinct from *M. nodulifera* (Hincks, 1881) and *M. fimbriata*. Finally, Ostrovsky and Taylor (2005) described one more unnamed Recent species from Okinawa, Japan (as *Monoporella* sp. 1).

In this study, I report from Japan a species of *Monoporella* that forms uniserial encrusting colonies. It may contribute to the study of evolution in this genus around the North Pacific.

**Materials and methods**

All of the study material came from the research expedition *Hakurei-Maru* cruise GH80-2 (Geological Survey of Japan) in 1980. Three sampling stations ranged from 144 to 350 m in depth on the continental shelf and slope east of the Boso Peninsula, Japan (Table 1). A Smith-McIntyre grab sampler was used at two stations (St. 1709 and 1734), and a chain-bag dredge was employed at one station (St. 1715).

Colonies and fragments of *Monoporella* species were picked from dried seafloor deposits under a stereoscopic microscope, and were cleaned in water or a hydrogen peroxide solution. For SEM, specimens were mounted on aluminum stubs with adhesive tape. Some were coated with Pt-Pd in a sputter coater (Model E-102, Hitachi) and observed with a Hitachi Model S-2400 scanning electron microscope at the Department of Earth Science, Faculty of Sciences, Chiba University. Other specimens were coated with Au in an ion coater (model SC-701AT, Sanyu Denshi) and observed with a JEOL Model JSM-5310 microscope at the National Museum of Nature and Science, Tsukuba.

Zooidal characters were measured from SEM images.

**Table 1.** Location and bottom sediments of sampling stations (Honza and Miyazaki 1984; Nishimura 1984).

| Station No. | Latitude    | Longitude    | Depth (m) | Bottom sediments       |
|-------------|-------------|--------------|-----------|------------------------|
| 1709        | 35°14.02′N  | 140°45.20′E  | 144       | sand and gravel        |
| 1715        | 35°54.80′-55.35′N | 140°22.80′-23.08′E | 303-350  | medium sand and rock   |
| 1734        | 35°03.13′N  | 140°26.99′E  | 155       | medium sand            |

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taken at about 360×, 480×, or 1700× magnification. Measurements (in millimeters) are presented as the range, followed by the arithmetic mean and standard deviation. Sample sizes (n) are given as the number of zooids from which a character was measured, followed by the number of colony fragment from which measurements were taken. Abbreviations used for characters measured are as follows: ZL, autozooid length; ZW, autozooid width; OrL, orifice length; OrW, orifice width.

Type material and other specimens examined in this study are deposited in the National Museum of Nature and Science, Tsukuba (collection code NMNS PA). Additional specimens examined are deposited in the collection of the science laboratory of Seishin-Gakuen, Kashima City, Ibaraki, Japan (collection code SGBC).

Diagnosis. Colony encrusting or erect. Frontal wall cryptocystal, granular, perforated, raised medially in general, with a pair of opesiules at the distal corners of cryptocyst. Lateral and proximal gymnocyct absent. Oral and lateral rim raised. Opesia reduced, coincident with the operculum. Oral spines present or absent. Avicularia absent. Ovicell large, spinocystal, with longitudinal costae, slit-like foraminia, small pores and lateral windows. Basal pore-chambers present.

Type species. Haploporella nodulifera Hincks, 1881.
Uniserial species of *Monoporella*

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*Monoporella projecta* sp. nov. (Figs 1–4)

*Monoporella* sp.: Arakawa 1999: 57, pl. 2, fig. D.

**Materials examined.** *Holotype*: NMNS PA 16855 (on a bivalve, with separated zooids from the same colony), Station 1734, *Hakurei-Maru* cruise GH-80-2. *Paratypes*: NMNS PA 16857 (on a fragment of molluscan shell), and 18451A (on a stone, not coated with metal), Station 1734, *Hakurei-Maru* cruise GH-80-2; NMNS PA 18451B (opercula from 18451A), Station 1734, *Hakurei-Maru* cruise GH-80-2; NMNS PA 18452, SGBC-0379 and -0380 (all on fragments of molluscs), Station 1734, *Hakurei-Maru* cruise GH-80-2.

**Diagnosis.** Colony encrusting, uniserial. Frontal cryptocyst inflated, higher than low mural rim, showing tessellate surface, with scattered pores and a pair of small opesiules. Orificial rim highly projecting. Oral spines absent.

**Etymology.** The specific name comes from the Latin *pro-*.  

![Image](image-url)  

Fig. 2. Autozooids. A, holotype, NMNS PA 16855, ancestrulae "a" and subsequent zooids; B, paratype, NMNS PA 16857, ancestrulae "0" and subsequent zooids; C, zooid "L4" in A, separated from the substrate by cleaning; D, zooid "3" in B, moved from the original position. Scale bars: 1 mm in A, B, and 200 µm in C, D.
jectus, referring to the highly projecting orificial rim.

**Measurements (in mm).** NMNS PA 16855, 16857 and 18452; SGBC-0379 and -0380. Autozooids (24, 4): ZL, 0.99–1.52 (1.300±0.166); ZW, 0.63–0.91 (0.710±0.061); OrL, 0.11–0.16 (0.141±0.013); OrW, 0.15–0.21 (0.175±0.015).

**Description.** Colony encrusting, uniserial; zooids generally budding one or two zooids, rarely three (Figs 1, 2A, B). Zooids elliptical, much inflated. Lateral wall very low, with pair of large communication pores laterally and one distally. Frontal wall cryptocrystal, inflated above low mural rim, finely granulated; granules aligned to form reticulum of narrow ridges (Fig. 3A, B), and perforated with scattered pores; median carina sometimes observed (Fig. 2C, D); interior of frontal wall nearly smooth, with tube structure connecting small opesiules (Fig. 3C). Orifice semicircular, with straight or slightly concave proximal border, bearing small shelf in each proximo-lateral corner, sometimes with a pair of round condyles; vestibular arch well-developed, crenulated, often boomerang-shaped inside (Fig. 4A, B). Orificial rim highly projecting (Fig. 2D), about 0.3 mm tall; its surface finely granulated, not tessellate (Fig. 3D). Oral spines absent. Operculum black; outer surface smooth, not pitted; inner surface with thin central area and proximolateral crests for attachment of occlusor muscles (Fig. 4C, D). Ovicell not found. Avicularia absent. Ancestrulae resembling subsequent autozooids, not so small (zooid “a” in Fig. 2A, zooid “0” in Fig. 2B).

**Distribution.** Presently known only from the eastern continental shelf and slope of the Boso Peninsula, Japan. The bathymetrical range is from 144 m to 350 m in depth.

**Remarks.** This species is characterized by the uniserial colony form, the highly inflated cryptocrystal and the projecting orificial rim.

It has a smooth operculum in common with all of six Monoporella species from the Aleutian Islands (Dick 2008), and four of which (Monoporella flexibila Dick, 2008, M. elongata Dick, 2008, M. ellefsoni Dick, 2008, M. aleutica Dick, 2008) also lack oral spines. Among these, M. flexibila is most similar to M. projecta in having a low mural rim and in the obscurity of the marginal boundary of cryptocrystal, especially in the initial encrusting portion of the colony.

The frontal wall of Monoporella projecta resembles the tessellate cryptocrystal in M. flexibila and M. elongata, but the concavities in the cryptocrystal of this species are shallower and broader than in the latter two species, and the bottom of each concavity is also generally granulated (Figs 2D, 3A). In addition, the surface of the orificial rim is not pitted in M. projecta.
Discussion

*Monoporella projecta* sp. nov. increases the known diversity of *Monoporella* along the northwestern Pacific coast. Including Aleutian species, some of which show similar characteristics of the frontal wall.

Several species of *Monoporella* bearing an inflated frontal cryptocyst occur in the northwestern Pacific. It is very conspicuous in *M. flexibila* and *M. projecta*. Although the phylogeny is not clear, it is noteworthy that *Monoporella sulcoecia* Kataoka, 1961 from the Pleistocene Ryukyu Limestone also has an inflated cryptocyst and low mural rim. A highly inflated cryptocyst is also evident in the Cretaceous *Monoporella convexa* (Canu, 1911) from Argentina, but the cryptocyst (except for the median carina) is lower than the mural rim in most *Monoporella* species.

In addition, the tessellate surface of cryptocyst in *Monoporella* has been also observed only in species from the northwestern Pacific Ocean, including Japan.

Scholz (1991) described an ancestrula of a *Monoporella* species (identified as *M. nodulifera*) in which the proximal part of the cryptocyst is apparently tessellate. This Philippine species differs from *M. projecta* in having oral spines and a pitted operculum. Another example is *Monoporella* sp. 1 in Ostrovsky and Taylor (2005) from Okinawa, Japan, showing a cryptocyst with somewhat deep concavities and a finely granulated orificial rim, and bearing two to six oral spines.

Tilbrook (2006) re-examined Harmer’s (1926) Siboga specimens, and referred to two *Monoporella* species from Japan. He identified one of them lacking oral spines and a pitted operculum as *Monoporella* sp. from Alaska in Taylor and McKinney (2002), i.e., *Monoporella* sp. 2 in Ostrovsky and Taylor (2005). This species was named *Monoporella elongata* by Dick (2008). If the conclusions of Tilbrook (2006) and Dick (2008) are correct, three *Monoporella* species with a tessellate cryptocyst, including *M. projecta*, have been recorded in Japan.

On the other hand, the characteristics of *Monoporella projecta* may be related to the deep water environment.

Among cheilostomes, relatively few species form uniserial encrusting colonies, except in some families such as Ae-teidae, Electridae and Hippothoidae. *Monoporella projecta* differs from species in these families in numerous characteristics, especially zooids lacking cauda. Bishop (1989) termed such non-caudate species “category 1 of uniserial runners”.

Some other species have similar uniserial colonies and zooids, e.g., *Thalamoporella linearis* Canu and Bassler, 1929 from 161 m in depth near Hong Kong; *Microporella lineata* Canu and Bassler, 1929 from 192 m in the Philippines; and *Psilopsella uniseriata* Canu and Bassler, 1927 from 37–192 m

Fig. 4. Orifice and operculum. A, orifice of zooid L4 in Fig. 2A; B, orifice of one zooid of the lower-right row in Fig. 1A; C, outer surface of operculum, NMNS PA 18451B; D, inner surface of operculum, NMNS PA 18451B. Scale bars: 50 µm.
in the Philippines. *Microporella lineata* was also recorded from the Kermadec Ridge of New Zealand (Gordon 1984), and this species was found with another uniserial species *Nimba terraenovae* (Powell, 1967) at the same sampling stations, K826, 840, 857 and 872 (165–490 m in depth). *Microporella lineata* was also recorded from the Kermadec Ridge. New Zealand Oceanographical Institute Memoir 91: 1–198.

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Fig. 5. *Hippothoa connata* Ortmann, 1890, encrusting the same substrate (a molluscan shell) as *Monoporella projecta*, NMNS PA 18452. Scale bar: 500 µm.
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