Research Article

Macrophage migration inhibitory factor -173 G>C single nucleotide polymorphism and its association with active pulmonary tuberculosis

Mirela Gehlen¹, Elis Regina Dalla Costa², Maria Lucia Rosa Rossetti³, Denise Rossato Silva¹,4*

¹ Programa de Pós-Graduação em Ciências Pneumológicas, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, RS, Brazil, ² Centro de Desenvolvimento Científico e Tecnológico, Secretaria Estadual da Saúde do Rio Grande do Sul (CDCT/SES), Porto Alegre, RS, Brazil, ³ Programa de Pós-Graduação em Biologia Molecular e Celular Aplicada à Saúde ( Biosaúde), Universidade Luterana do Brasil (ULBRA), Canoas, RS, Brazil, ⁴ Faculdade de Medicina, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, RS, Brazil

* denise.rossato@terra.com.br

Abstract

Purpose

The establishment of candidate genes associated with susceptibility to TB is a challenge especially due to divergent frequencies among different populations. The objective of this study was to evaluate the association between macrophage migration inhibitory factor (MIF) -173 G>C single nucleotide polymorphism (SNP) and susceptibility to pulmonary TB in a population of southern Brazil.

Methods

Case-control study. Patients > 18 years old, diagnosed with pulmonary TB were included. The control group consisted of blood donors and household contacts, not relatives, healthy and > 18 years old. MIF -173 G>C SNPs were genotyped using real-time PCR using a Taq-Man SNP Genotyping assay.

Results

174 patients and 166 controls were included. There were no statistically significant differences between cases and controls regarding genotype prevalence (p>0.05). Comparing patients with normal genotype (GG) with those with at least one C allele, there was also no statistically significant difference (p = 0.135). Also, there was no statistically significant difference comparing the homozygous for the mutation (CC) with the other patients (GG and CG) (p = 0.864).
Conclusions
We did not find association between MIF -173 G>C polymorphism and susceptibility to pulmonary TB.

Introduction
The natural history of tuberculosis (TB) follows a variable course after the initial infection, with only 10% of infected individuals actually developing clinical disease. Genetic, environmental and bacterial virulence factors can influence the clinical presentation of TB [1]. Many studies indicate that genetic factors play a major role in determining the susceptibility and resistance to TB [2–5].

Polymorphisms of genes play an important role in the occurrence and development of TB. In recent years, many TB susceptibility genes have been detected [6–13]. The macrophage migration inhibitory factor (MIF) is a cytokine with proinflammatory chemokine-like functions that have been recognized to play a central role in mediating a wide variety of immune responses against invading pathogens, and may be associated with the onset and / or progression of TB. In fact, a metaanalysis revealed a strong association of a MIF polymorphism with autoimmune and infectious diseases [14]. In TB, MIF is probably the first cytokine to appear in inflammatory response, inhibits macrophage migration, and promotes macrophage accumulation and T lymphocytes activation in inflamed TB lesions [15,16]. Four polymorphisms were identified in the MIF gene; however, only the MIF-173 and MIF (CATT5-8) are related to changes on MIF levels [17]. A few studies had evaluated these polymorphisms in TB, in different populations, with controversial results [16,18–22].

The establishment of candidate genes associated with susceptibility to TB is a challenge especially due to divergent frequencies among different populations [23]. Therefore, studies on the relationship between the susceptibility to TB and genetic polymorphisms in various populations and ethnic groups are of great importance. The objective of this study was to evaluate, in a case-control study, the association between MIF-173 G>C single nucleotide polymorphism (SNP) and susceptibility to pulmonary TB in a population of southern Brazil.

Materials and methods
Study design and location
This study was a case-control study done on 174 patients and 166 controls and conducted in two hospitals in a city in Southern Brazil with high incidence of TB (80.4 cases/100,000 inhabitants) [24]: Hospital de Clínicas de Porto Alegre, a tertiary hospital, university-affiliated, with 750 beds and about 300 cases of TB treated per year, and Hospital Sanatório Partenon, a referral hospital for TB treatment. The study was conducted between 2016 and 2019. The study was approved by the Research Ethics Committee of Hospital de Clínicas de Porto Alegre (number 16–0599), and all research was in accordance with regulations. All cases and controls signed Informed Consent Form prior to inclusion in the study.

Patients
We included patients > 18 years old, diagnosed with pulmonary TB, during any time between the diagnosis and completion of treatment. Patients with concomitant extrapulmonary TB were excluded. The control group consisted of blood donors and household contacts, not
relatives, healthy and > 18 years old. Cases and controls were negative for human immunodeficiency virus (HIV), and hepatitis B and C virus (HBV and HCV).

Data collection

Patients were interviewed using a standardized questionnaire. The following data were recorded: demographic data (gender, age, race, years of schooling); presence of symptoms; smoking history; alcohol and drug abuse, and comorbidities. The racial and ethnic composition of Brazilian society is the result of a confluence of people from many different ethnic backgrounds: the original indigenous people, black Africans, Portuguese settlers, and later, European, Arab and Japanese immigrants, as well as other Asian people and from South American countries [25]. We collected data on race and classified patients in white and non-white. The diagnosis of pulmonary TB was accomplished through chest radiography, sputum smear and culture for mycobacteria, and based on consensus criteria [26]. All patients used the same treatment (the standard treatment in Brazil: rifampicin, isoniazid, ethambutol, and pyrazinamide).

DNA extraction

Whole blood samples were collected in EDTA tubes. Total genomic DNA was isolated from peripheral blood leukocytes by the salting out method [27] and stored at -20°C until analysis.

MIF Genotyping-173 G>C

The MIF -173 G>C polymorphism (rs755622) was genotyped by real-time polymerase chain reaction (RT-PCR) using a TaqMan SNP Genotyping assay (part number C_2213785_10; Applied Biosystems, Foster City, CA, USA), according to a protocol of Gomez et al [16], using StepOne ™ Real-Time PCR Systems. The PCR was carried out with mixes consisting of 8 ng of genomic DNA, 2.5 ml of Taqman master mix, 0.125 μl of 20 X assay mix and ddH₂O up to 5 μl of final volume. The amplification protocol used was 50˚C for 2 min and initial denaturation at 95˚C for 10 min followed by 50 cycles of denaturation at 92˚C for 15 s and annealing/extension at 60˚C for 1 min [16].

Statistical analysis

Data analysis was performed using SPSS 18.0 (Statistical Package for the Social Sciences, Chicago, Illinois). Genotypic frequencies were tested for Hardy-Weinberg using the chi-square test. Frequencies of genotypes and alleles were compared between cases and controls by logistic regression with adjustment for gender and age. Odds ratio (OR) was used as the point estimates of risk and was calculated along with their 95% confidence intervals (95% CI). P value less than 0.05 was considered as statistically significant. Considering data of a previous study [20], with confidence interval of 95% and a power of 80%, at least 83 cases and 83 controls will be needed.

Results

During the study period, 174 patients and 166 controls met the inclusion criteria and were included in the analysis. The age range was 18–78 years (cases) and 18–61 years (controls). There were 120 (69.0%) males and 54 (31.0%) females among cases and 105 (63.3%) males and 61 (36.7%) females among controls. One hundred and fifteen patients were white (66.1%) and 59 (33.9%) were non-white among cases and 114 (68.7%) were white and 52 (31.3%) were non-white among controls. The prevalence of genotypes GG (normal), GC (heterozygous),
and CC (homozygous for mutation) among cases was 94 (54.0%), 72 (41.4%), and 8 (4.6%), respectively. In controls, the frequencies of genotypes GG (normal), GC (heterozygous), and CC (homozygous for mutation) among cases were 103 (62.1%), 56 (33.7%), and 7 (4.2%), respectively. The genotype frequencies were not found to be significantly different from those predicted by the Hardy–Weinberg equilibrium. There were no statistically significant differences between cases and controls regarding genotype prevalence, controlled for age and sex (Table 1).

Comparing patients with normal genotype (GG) with those with at least one C allele, there was also no statistically significant difference (p = 0.135). In addition, there was no statistically significant difference comparing the homozygous for the mutation (CC) with the other patients (GG and CG) (p = 0.864).

Discussion

In the present study, we did not find association between MIF -173 G>C polymorphism and susceptibility to pulmonary TB. There were no statistically significant differences between cases and controls regarding genotype prevalence. In addition, there was no statistically significant difference comparing patients with normal genotype (GG) with those with at least one C allele, and comparing the homozygous for the mutation (CC) with the other patients (GG and CG).

There is increasing evidence that host genetic factors are implicated in susceptibility to TB, corroborated by monozygotic and dizygotic twin studies [5,28], genome-wide linkage studies [29–31], and genome-wide association studies [32,33]. Polymorphisms in candidate genes have shown influence on this TB susceptibility. MIF -173 G>C gene polymorphism has been shown to play a role in TB risk in several studies [16,18–22]. MIF gene is located at chromosome22q11.2 and encodes a T-cell-derived cytokine that plays an important role activating macrophage functions, regulating Th1/Th2 balance, and also is involved in delayed-type hypersensitivity reaction [34–36]. MIF is expressed by epithelial cells of the bronchi and alveolar macrophages, which have substantial influence on defense against tubercle bacilli [37]. In a Ugandan cohort, it was showed that genetic low expressers of MIF were 2.4-times more frequently identified among patients with Mycobacterium tuberculosis bacteremia than those without. Also, MIF deficient mice have an inadequate innate immune response and, consequently, are more susceptible to mycobacterial pathology [38].

Table 1. Genotype frequency of MIF -137 G/C polymorphism in cases and controls.

| Genetic model | Cases, n (%) | Controls, n (%) | OR (95% CI) | P value |
|---------------|-------------|----------------|-------------|---------|
| Codominant    |             |                |             |         |
| GG            | 94 (54.0)   | 103 (62.1)     | 1.00 (reference) |         |
| GC            | 72 (41.4)   | 56 (33.7)      | 0.71 (0.45–1.11) | 0.133   |
| CC            | 8 (4.6)     | 7 (4.2)        | 0.79 (0.28–2.29) | 0.675   |
| Dominant      |             |                |             |         |
| GG            | 94 (54.0)   | 103 (62.1)     | 1.00 (reference) |         |
| GC + CC       | 80 (46.0)   | 63 (37.9)      | 0.72 (0.47–1.11) | 0.135   |
| Recessive     |             |                |             |         |
| GG + GC       | 166 (95.4)  | 159 (95.8)     | 1.00 (reference) |         |
| CC            | 8 (4.6)     | 7 (4.2)        | 1.09 (0.39–3.09) | 0.864   |
| Allele        |             |                |             |         |
| C             | 88 (25.3)   | 70 (21.1)      | -           | 0.195   |
| G             | 260 (74.7)  | 262 (78.9)     | -           | 0.195   |

https://doi.org/10.1371/journal.pone.0234565.t001
We did not find differences in MIF -173 G>C genotype prevalence comparing patients with TB and the control group. Gomez et al [16] studied 230 northwestern Colombian patients with pulmonary TB, negative for HIV, and 235 matched healthy individuals, and found on multivariate analysis that MIF -173C allele was associated with disease (odds ratio = 1.64) in a dominant pattern. In a Moroccan population, the authors demonstrated a statistically significant increase of the MIF –173CC homozygote genotype and MIF –173°C allele frequencies in PTB patients compared with healthy controls [21]. Hashemi et al [18], showed in an Iranian population that the MIF -173 G/C polymorphism increased the risk of PTB in codominant (GC vs GG, OR = 1.76) and dominant (GC+CC vs GG, OR = 1.78) tested models. Three studies were conducted in Chinese population. One of them [19] evidenced that distribution of MIF -173 genotypes (GC + CC) was significantly higher in TB cases than in controls, and the frequencies for MIF -173 (GG vs. GC+CC) were statistically significant different comparing total cases of TB, new cases of TB, and retreatment cases of TB to controls, respectively. Yanlin et al [22] demonstrated that the frequency of MIF -173 (GC + CC) was higher in TB patients than in controls (OR = 2.12). Last, Kuai et al [20] found a significant association of MIF -173 C alleles with susceptibility to active TB. Since polymorphisms occur at different frequencies in populations of different ethnicities, the high prevalence of miscegenation in our country may have contributed to these results [25]. In addition, one should also consider the fact that the population of southern Brazil (study setting) has Italian and German descent, and that these polymorphisms have not been demonstrated in these populations, only in Chinese [19,20,22], Moroccan [21], Colombian [16], and Iranian [18] populations. In addition, the sample size could be small to come to a conclusion at this point; however, we calculated a priori sample size, and it is bigger than 3 out of 6 studies conducted on this topic.

One limitation of this study is that we recruited patients from a single region of Brazil, and, as stated above, it is well known that Brazil is an extensive country with a varied ethnic and racial composition. Therefore, this may have prevented us from finding differences in genotype frequency.

This study has been conducted in mixed population, as both cases and control study subjects consists of peoples from various ethnic origin. An elaborated study with a larger sample size is required to bring enough power to analyse the data from different ethnic groups. In addition, we didn’t collect data on body mass index (BMI), and it is well known that low BMI could contribute to TB disease establishment. Also, the patients were not followed-up after the treatment for relapse. Furthermore, we know that a cohort study including patients with MIF polymorphism and evaluating the development of TB would bring better scientific evidence; however, this kind of study is expensive and take a lot of time to get results. Moreover, MIF levels were not quantified; a correlation study between different genotypes and MIF levels in patients and controls might help to explain the association between MIF -173 G>C gene polymorphism and susceptibility/resistance to tuberculosis. In spite of these concerns, this is the first study to evaluate the association between MIF -173 G>C polymorphism and susceptibility to pulmonary TB in Brazil.

In conclusion, in the present study, there were no differences in MIF -173 G>C genotype prevalence when comparing TB cases and controls. Further studies, including patients from various regions of Brazil are necessary to better evaluate the association between MIF -173 G>C polymorphism and susceptibility to pulmonary TB.

**Author Contributions**

**Conceptualization:** Mirela Gehlen, Elis Regina Dalla Costa, Maria Lucia Rosa Rossetti, Denise Rossato Silva.
Formal analysis: Mirela Gehlen, Elis Regina Dalla Costa, Maria Lucia Rosa Rossetti, Denise Rossato Silva.

Funding acquisition: Denise Rossato Silva.

Investigation: Mirela Gehlen, Elis Regina Dalla Costa, Denise Rossato Silva.

Methodology: Mirela Gehlen, Maria Lucia Rosa Rossetti, Denise Rossato Silva.

Project administration: Mirela Gehlen, Elis Regina Dalla Costa, Maria Lucia Rosa Rossetti, Denise Rossato Silva.

Supervision: Denise Rossato Silva.

Writing – original draft: Mirela Gehlen, Denise Rossato Silva.

Writing – review & editing: Mirela Gehlen, Elis Regina Dalla Costa, Maria Lucia Rosa Rossetti.

References

1. Raviglione MC, Kochi A, Snider DE. Global Epidemiology of Tuberculosis: Morbidity and Mortality of a Worldwide Epidemic. JAMA J Am Med Assoc. 1995; 273:220–6.

2. van de Vosse E, Hoeve MA, Ottenhoff THM. Human genetics of intracellular infectious diseases: molecular and cellular immunity against mycobacteria and salmonellae. Lancet Infect Dis [Internet]. 2004 [cited 2019 Oct 21]; 4:739–49. Available from: http://www.ncbi.nlm.nih.gov/pubmed/15567123 https://doi.org/10.1016/S1473-3099(04)01203-4 PMID: 15567123

3. Hill A. The genomics and genetics of human infectious disease susceptibility.—PubMed—NCBI [Internet]. Annu Rev Genomics Hum Genet. 2001 [cited 2019 Oct 21]. p. 373–400. Available from: https://www.ncbi.nlm.nih.gov/pubmed/11701655 https://doi.org/10.1146/annurev.genom.2.1.373 PMID: 11701655

4. Bellamy R. Genetic susceptibility to tuberculosis. Clin Chest Med [Internet]. 2005 [cited 2019 Oct 21]; 26:233–46, vi. Available from: http://www.ncbi.nlm.nih.gov/pubmed/15837108 https://doi.org/10.1016/j.ccm.2005.02.006 PMID: 15837108

5. Comstock GW. Tuberculosis in twins: a re-analysis of the Prophit survey. Am Rev Respir Dis [Internet]. 1978 [cited 2019 Oct 21]; 117:621–4. Available from: http://www.ncbi.nlm.nih.gov/pubmed/565607 https://doi.org/10.1164/arrd.1978.117.4.621 PMID: 565607

6. Vasilca V, Oana R, Munteanu D, Zugun F, Constantinescu D, Carasevici E. HLA-A and -B phenotypes associated with tuberculosis in population from north-eastern Romania. Roum Arch Microbiol Immunol [Internet]. [cited 2019 Oct 21]; 63:209–21. Available from: http://www.ncbi.nlm.nih.gov/pubmed/17240790

7. Yim J-J, Selvaraj P. Genetic susceptibility in tuberculosis. Respirology [Internet]. 2010 [cited 2019 Oct 21]; 15:241–56. Available from: http://www.ncbi.nlm.nih.gov/pubmed/20199642 https://doi.org/10.1111/j.1440-1843.2009.01690.x PMID: 20199642

8. Malik S, Abel L, Tooker H, Poon A, Simkin L, Girard M, et al. Alleles of the NRAMP1 gene are risk factors for pediatric tuberculosis disease. Proc Natl Acad Sci U S A [Internet]. 2005 [cited 2019 Oct 21]; 102:12183–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/16103355 https://doi.org/10.1073/pnas.0503368102 PMID: 16103355

9. Dissanayeke SR, Levin S, Pienaar S, Wood K, Eley B, Beatty D, et al. Polymorphic variation in TIRAP is not associated with susceptibility to childhood TB but may determine susceptibility to TBM in some ethnic groups. PLoS One [Internet]. 2009 [cited 2019 Oct 21]; 4:e6698. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19693265 https://doi.org/10.1371/journal.pone.0006698 PMID: 19693265

10. Liu W, Cao WC, Zhang CY, Tian L, Wu XM, Habbema JDF, et al. VDR and NRAMP1 gene polymorphisms in susceptibility to pulmonary tuberculosis among the Chinese Han population: a case-control study. Int J Tuberc Lung Dis [Internet]. 2004 [cited 2019 Oct 21]; 8:428–34. Available from: http://www.ncbi.nlm.nih.gov/pubmed/15141734 PMID: 15141734

11. El Sahly HM, Reich RA, Dou SJ, Musser JM, Graiviss EA. The effect of mannose binding lectin gene polymorphisms on susceptibility to tuberculosis in different ethnic groups. Scand J Infect Dis [Internet]. 2004 [cited 2019 Oct 21]; 36:106–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/15061663 https://doi.org/10.1080/00365540310018860 PMID: 15061663
12. Tosh K, Campbell SJ, Fielding K, Sillah J, Bah B, Gustafson P, et al. Variants in the SP110 gene are associated with genetic susceptibility to tuberculosis in West Africa. Proc Natl Acad Sci U S A [Internet]. 2006 [cited 2019 Oct 21]; 103:10364–8. Available from: http://www.ncbi.nlm.nih.gov/pubmed/16803959 PMID: 16803959

13. Velez DR, Hulme WF, Myers JL, Weinberg JB, Levesque ME, et al. NOS2A, TLR4, and IFNGR1 interactions influence pulmonary tuberculosis susceptibility in African-Americans. Hum Genet [Internet]. 2009 [cited 2019 Oct 21]; 126:643–53. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19575238 PMID: 19575238

14. Illescas O, Gomez-Verjan JC, Garcia-Velazquez L, Govezensky T, Rodriguez-Sosa M. Macrophage migration inhibitory factor -173 G/C polymorphism: A global meta-analysis across the disease spectrum. Front Genet. 2018; 9:1–14. https://doi.org/10.3389/fgene.2018.00001

15. Nishihira J. Macrophage migration inhibitory factor (MIF): Its essential role in the immune system and cell growth. J Interf. Cytokine Res. 2000. p. 751–62.

16. Gómez LM, Sánchez E, Ruiz-Narvaez EA, López-Nevo T, Anaya JM, Martín J. Macrophage migration inhibitory factor gene influences the risk of developing tuberculosis in northwestern Colombian population. Tissue Antigens. 2007; 70:28–33. https://doi.org/10.1111/j.1399-0039.2007.00843.x PMID: 17559578

17. Donn R, Alourfi Z, De Benedetti F, Meazza C, Zeggini E, Lunt M, et al. Mutation screening of the macrophage migration inhibitory factor gene: positive association of a functional polymorphism of macrophage migration inhibitory factor with juvenile idiopathic arthritis. Arthritis Rheum [Internet]. 2002 [cited 2019 Oct 21]; 46:2402–9. Available from: http://www.ncbi.nlm.nih.gov/pubmed/1235488 https://doi.org/10.1002/art.10492 PMID: 12355488

18. Hashemi M, Sharifi-Mood B, Rasooli A, Amininia S, Naderi M, Taheri M. Macrophage migration inhibitory factor -173 G/C polymorphism is associated with an increased risk of pulmonary tuberculosis in Zahedan, Southeast Iran. EXCLI J. 2015; 14:117–22. https://doi.org/10.17179/excli2014-636 PMID: 27065766

19. Liu A, Li J, Bao F, Zhu Z, Feng S, Yang J, et al. Single nucleotide polymorphisms in cytokine MIF gene promoter region are closely associated with human susceptibility to tuberculosis in a southwestern province of China. Infect Genet Evol. Elsevier B.V.; 2016; 39:219–24. https://doi.org/10.1016/j.mgeid.2015.12.003 PMID: 26656832

20. Kuai SG, Ou QF, You DH, Shang ZB, Wang J, Liu J, et al. Functional polymorphisms in the gene encoding macrophage migration inhibitory factor (MIF) are associated with active pulmonary tuberculosis. Infect Dis (Auckl). 2016; 48:222–8.

21. Sadki K, Lamsyah H, Rueda B, Ali El, Sadak A, Martin J, et al. Analysis of MIF, FCGR2A and FCGR3A gene polymorphisms with susceptibility to pulmonary tuberculosis in Moroccan population. J Genet Genomics [Internet]. Institute of Genetics and Developmental Biology and the Genetics Society of China; 2010; 37:257–64. Available from: https://doi.org/10.1016/S1673-8527(09)60044-8 PMID: 20439102

22. Li Y, Yuan T, Lu W, Chen M, Cheng X, Deng S. Association of tuberculosis and polymorphisms in the promoter region of macrophage migration inhibitory factor (MIF) in a Southwestern China Han population. Cytokine [Internet]. Elsevier Ltd; 2012; 60:64–7. Available from: https://doi.org/10.1016/j.cyto.2012.06.010 PMID: 22742858

23. Vannberg FO, Chapman SJ, Hill AVS. Human genetic susceptibility to intracellular pathogens. Immunol Rev [Internet]. 2011 [cited 2019 Oct 21]; 240:105–16. Available from: http://www.ncbi.nlm.nih.gov/pubmed/21349089 https://doi.org/10.1111/j.1600-065X.2010.00996.x PMID: 21349089

24. Ministério da Saúde. Boletim Epidemiológico [Internet]. 2018. Available from: www.saude.gov.br

25. Instituto Brasileiro de Geografia e Estatística (IBGE). Censo 2010 [Internet]. 2010. Available from: www.ibge.gov.br

26. Conde MB, Melo FAF de, Marques AMC, Cardoso NC, Pinheiro VGF, Dalcín P de TR, et al. III Brazilian Thoracic Association Guidelines on tuberculosis. J Bras Pneumol [Internet]. 2009 [cited 2019 Oct 21]; 35:1018–48. Available from: http://www.ncbi.nlm.nih.gov/pubmed/19918635 https://doi.org/10.1590/s1806-37132009001000011 PMID: 19918635

27. Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res. 1988; 16:1215. https://doi.org/10.1093/nar/16.3.1215 PMID: 3344216

28. Möller M, Hoal EG. Current findings, challenges and novel approaches in human genetic susceptibility to tuberculosis. Tuberculosis (Edinb) [Internet]. 2010 [cited 2019 Oct 21]; 90:71–83. Available from: http://www.ncbi.nlm.nih.gov/pubmed/2026579

29. Cooke GS, Campbell SJ, Bennett S, Lienhardt C, McAdam KPWJ, Sirugo G, et al. Mapping of a novel susceptibility locus suggests a role for MC3R and CTSZ in human tuberculosis. Am J Respir Crit Care Med. 2008; 178:203–7. https://doi.org/10.1164/rccm.200710-1554OC PMID: 18420963
30. Bellamy R, Beyers N, McAdam KPWJ, Ruwende C, Gie R, Samaai P, et al. Genetic susceptibility to tuberculosis in Africans: A genome-wide scan. Proc Natl Acad Sci U S A. 2000; 97:8005–9. https://doi.org/10.1073/pnas.140201897 PMID: 10859364

31. Mahasirimongkol S, Yanai H, Nishida N, Ridruecha C, Matsushita I, Ohashi J, et al. Genome-wide SNP-based linkage analysis of tuberculosis in Thais. Genes Immun [Internet]. 2009 [cited 2019 Oct 21]; 10:77–83. Available from: http://www.ncbi.nlm.nih.gov/pubmed/18843276 https://doi.org/10.1038/gene.2008.81 PMID: 18843276

32. Mahasirimongkol S, Yanai H, Mushiroda T, Promphittayarat W, Wattanapokayakit S, Phromjai J, et al. Genome-wide association studies of tuberculosis in Asians identify distinct at-risk locus for young tuberculosis. J Hum Genet [Internet]. 2012 [cited 2019 Oct 21]; 57:363–7. Available from: http://www.ncbi.nlm.nih.gov/pubmed/22551897 https://doi.org/10.1038/jhg.2012.35 PMID: 22551897

33. Png E, Alisjahbana B, Sahiratmadja E, Marzuki S, Nelwan R, Balabanova Y, et al. A genome wide association study of pulmonary tuberculosis susceptibility in Indonesians. BMC Med Genet. 2012;13. https://doi.org/10.1186/1471-2350-13-13

34. Calandra T, Froidevaux C, Martin C, Roger T. Macrophage migration inhibitory factor and host innate immune defenses against bacterial sepsis. J Infect Dis [Internet]. 2003 [cited 2019 Oct 21];187 Suppl 2: S385–90. Available from: http://www.ncbi.nlm.nih.gov/pubmed/12792855

35. Bacher M, Metz CN, Calandra T, Mayer K, Chesney J, Lohoff M, et al. An essential regulatory role for macrophage migration inhibitory factor in T-cell activation. Proc Natl Acad Sci U S A. 1996; 93:7849–54. https://doi.org/10.1073/pnas.93.15.7849 PMID: 8755565

36. Martínez A, Orozco G, Varadé J, Sánchez López M, Pascual D, Balsa A, et al. Macrophage migration inhibitory factor gene: influence on rheumatoid arthritis susceptibility. Hum Immunol [Internet]. 2007 [cited 2019 Oct 21]; 68:744–7. Available from: http://www.ncbi.nlm.nih.gov/pubmed/17869648 https://doi.org/10.1016/j.humimm.2007.06.007 PMID: 17869648

37. Kibiki GS, van der Ven AJAM, Geurts-Moespot A, Shao J, Calandra T, Sweep FCGJ, et al. Serum and BAL macrophage migration inhibitory factor levels in HIV infected Tanzanians with pulmonary tuberculosis or other lung diseases. Clin Immunol. 2007; 123:60–5. https://doi.org/10.1016/j.clim.2006.12.004 PMID: 17275414

38. Das R, Koo MS, Kim BH, Jacob ST, Subbian S, Yao J, et al. Macrophage migration inhibitory factor (MIF) is a critical mediator of the innate immune response to Mycobacterium tuberculosis. Proc Natl Acad Sci U S A. 2013; 110.