Predominant D<sub>1</sub> Receptors Involvement in the Over-expression of CART Peptides after Repeated Cocaine Administration

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The aim of this study was to investigate the involvement of dopaminergic receptors (DR) in behavioral sensitization, as measured by locomotor activity, and the over-expression of cocaine- and amphetamine-regulated transcript (CART) peptides after repeated administration of cocaine in mice. Repeated administrations of cocaine induced behavioral sensitization and CART over-expression in mice. The levels of striatal CART mRNA were significantly increased on the 3<sup>rd</sup> day. CART peptides were over-expressed on the 5<sup>th</sup> day in the striata of behaviorally sensitized mice. A higher proportion of CART<sup>+</sup> cells in the cocaine-treated mice were present in the nucleus accumbens (NAc) shell than in the dorsolateral (DL) part of caudate putamen (CP). The concomitant administration of both D<sub>1</sub>R and D<sub>2</sub>R antagonists, SCH 23390 (D<sub>1</sub>R selective) and raclopride (D<sub>2</sub>R selective), blocked cocaine-induced behavioral sensitization, CART over-expression, and cyclic adenosine 5'-monophosphate (cAMP)/protein kinase A (PKA)/phospho-cAMP response element-binding protein (pCREB) signal pathways. SCH 23390 more predominantly inhibited the locomotor activity, CART over-expression, pCREB and PKA activity than raclopride. Cocaine-induced behavioral sensitization was also attenuated in the both D<sub>1</sub>R and D<sub>2</sub>R knockout (KO) mice, respectively. CART over-expression and activated cAMP/PKA/pCREB signal pathways were inhibited in the D<sub>1</sub>R-KO mice, but not in the D<sub>2</sub>R-KO mice. It is suggested that behavioral sensitization, CART over-expression and activated cAMP/PKA/pCREB signal pathways induced by repeated administration of cocaine could be more predominantly mediated by D<sub>1</sub>R.

Key Words: cAMP, cAMP/PKA/pCREB signal pathways, CART, Cocaine, Dopamine receptors

INTRODUCTION

Repeated administration of cocaine elicits specific behaviors referred as sensitization, reward, reinforcement, and drug induced psychosis in animals and humans [1-5]. The striatofugal pathway (direct pathway) contains neurons with dynorphin, substance P, and dopaminergic D<sub>1</sub> receptors (D<sub>1</sub>R), whereas striatopallidal pathway (indirect pathway) preferentially contains neurons with enkephalin and dopaminergic D<sub>2</sub> receptors (D<sub>2</sub>R) [6]. D<sub>1</sub>R and D<sub>2</sub>R antagonists inhibited locomotor activity induced by psychostimulants [7,8]. However, the concomitant administration of D<sub>1</sub>R and D<sub>2</sub>R agonists enhances behavioral sensitization (BS) [9]. Cocaine did not increase locomotor activity in D<sub>1</sub>R knockout (KO) mice [10,11]. The locomotor response to cocaine was attenuated in D<sub>2</sub>R KO mice [12].

Cocaine- and amphetamine-regulated transcript (CART) peptides that are highly expressed within the nucleus accumbens (NAc), hypothalamus, and ventral tegmental area (VTA) may contribute to drug reinforcement, rewarding-seeking behaviors, and addiction liability of psychostimulants [13,14]. CART mRNA was originally shown to be up-regulated in the rat striatum after acute cocaine or amphetamine administration [15]. Repeated administration of cocaine or amphetamine gradually reduced the level of over-expressed CART [16,17]. Direct injections of CART peptide into the NC attenuate both cocaine- and dopamine-induced locomotor activity [18]. D<sub>1</sub> and D<sub>2</sub>/D<sub>3</sub>R antagonists inhibit the ethanol-induced over-expression of CART in the rat NC [19]. CART expression is regulated via adenylyl cyclase and cyclic adenosine 5'-monophosphate (cAMP)/protein kinase A (PKA)-mediating signaling, and likely through the activation of cAMP response element-binding protein (CREB) [20-22]. While both DRs and CART peptides contribute substantially to the development and/or expression of cocaine-induced behavioral sensitization, it remains unclear how the expression of CART peptides is mediated via both D<sub>1</sub>R and D<sub>2</sub>R.

ABBREVIATIONS: DR, dopaminergic receptors; CART, cocaine- and amphetamine-regulated transcript; NAc, nucleus accumbens; DL, dorsolateral; CP, caudate putamen; cAMP, cyclic adenosine 5'-monophosphate; PK, protein kinase A; pCREB, phospho-cAMP response element-binding protein; NO, knockout.
D1R in the development of cocaine-induced behavioral sensitization. In the present study, the mRNA expression of CART mRNA and distribution in the striatum of male C57BL/6 mice were assessed after repeated administration of cocaine. We investigated the involvement of D1R and D2R in the overexpression of CART peptides and cAMP/PKA/CREB signaling pathways in the striata of behaviorally sensitized mice. The levels of CART peptides after repeated administration of cocaine were compared with those of both D1R- and D2R- knockout (D1R−/− and D2R−/−) mice, and wild type (WT) mice.

METHODS

Materials

Cocaine, D1R antagonist [R(+)-SCH-23390 hydrochloride] and D2R antagonist [(S)-raclopride(+)-tartrate salt] were purchased from the Sigma Chemical Company (St. Louis, MO, USA). All chemicals were dissolved immediately in physiological saline before use. The CART and GAPDH primers, M-MLV reverse transcriptase and AccuPower PCR Premix were purchased from Bioneer Co. (Seoul, Korea). The 2X SYBR Green Master Mix was purchased from Applied Biosystems (Carlsbad, CA, USA). The anti-CART and anti-GAPDH antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). The anti-phospho[Ser-133] CREB, anti-CREB, and anti-PKA antibodies were purchased from Abcam (Cambridge, MA, USA). The Vectastain ABC reagent was purchased from Vector Laboratories Co. (Burlingame, CA, USA). The Cyclic AMP XP Assay Kit was purchased from Cell Signaling Technology Inc. (Danvers, MA, USA). Protein G Sepharose 4 Fast Flow was purchased from GE Healthcare Bio-Sciences AB (Uppsala, Sweden). The PKA Assay Kit was purchased from Millipore Co. (Bedford, MA, USA). The [γ-32P] ATP was purchased from Perkin Elmer, Inc. (Covina, CA, USA). All other materials were of the highest grade available and were obtained from normal commercial sources.

Animals

Adult male C57BL/6 mice weighing 25–30 g were purchased from the Samtako Company (Osan, Korea). The adult heterozygous D1R-KO mice (D1R+/-) were purchased from the RIKEN BioResource Center (Tsukuba, Japan). The mice were genotyped by PCR analysis of genomic DNA using standard PCR protocols (http://www2.brc.riken.jp/cache_all/RBRC01080, data not shown). The homozygous D1R-KO mice (D1R−/−) were purchased from the Jackson Laboratory (Bar Harbor, ME, USA). The mice were genotyped by PCR analysis of genomic DNA using the JAX® genotyping protocols (http://jaxmice.jax.org/strain/003190.html, data not shown). All of the experiments using animals were performed in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals (NIH publication No. 85-23, revised 1985), and the Institutional Animal Care and Use Committee of Chungbuk National University approved the protocol.

Measurement of locomotor activity

All of the experiments were done in a randomized, balanced repeated-measures design, such that each mice received all of the treatments. A separate group of at least 8 mice were used for each experiment. The locomotor activity was measured using a tilting-type ambulometer (AMB-10, O’Hara, Tokyo, Japan). Each mouse was placed in an activity cage (20-cm diameter, 18-cm height). The control mice were given saline subcutaneously under the same conditions. Cocaine was administered to mice once per day for 7 days. The mice were first allowed to perambulate for 10 min in the activity cages followed by a 1-h test period immediately after saline or cocaine administration. The development of behavioral sensitization after 5 or 7 days was evidenced by increased locomotor activity in response to cocaine, and compared with activity on the 1st day [23]. In addition, to measure inhibitory effects of cocaine-induced hyperactivity, SCH 23390 (0.25 mg/kg) or raclopride (0.4 mg/kg) was pretreated intraperitoneally (i.p.) to mice, 25 min prior to cocaine administration once per day for 5 days [24]. Cocaine (15 mg/kg, s.c.) was also administered to the D1R- and D2R-KO mice for once a day for 5 days.

Real-time polymerase chain reaction (qRT-PCR) analysis

The separate groups of 4 mice were sacrificed by decapitation 22 h after the administration of cocaine (5, 15 and 30 mg/kg, s.c.) for 3, 5, 7 and 14 days, respectively. The total number of animals is 48 mice. The striata including the NAc, were extracted at the coronal level at + 1.6 and + 1.0 mm from the bregma, according to the stereotaxic atlas [25]. The CART mRNA levels in the striata were quantified using qRT-PCR. The sequences for the primers and an internal control for CART were designed in accordance with a previously published paper (CART: forward primer: CGAGAAGAGTACGCGCAAGC; reverse primer: GGAATATGGGAAACCGAAGGT; GAPDH: forward primer: AAATTCAACGGCCACGTCCA; reverse primer: GAACGGACGGAGAATGGACGGTGATGAC) [26]. The total RNA was extracted using TRIzol reagent. The reverse transcription reaction using M-MLV reverse transcriptase was performed according to the manufacturer’s instructions. The qRT-PCR was performed using a 7,500 detection system (Applied Biosystems, Carlsbad, CA, USA). The reaction was conducted in a 20-μl reaction mixture containing 2X SYBR Green Master Mix, 250 nM primers, and 1.0 μg of RNA per sample. The thermal cycling conditions were programmed as follows: preheating for 10 min at 95°C, followed by 45 cycles of two-step PCR consisting of 15 s at 95°C and 1 min at 60°C (the extension temperature). The accumulation of PCR products was monitored through the increase in fluorescence. A standard curve was used for relative quantification.

Immunohistochemistry analysis

After completion of the experiments, each group of 4 mice was anesthetized with pentobarbital sodium (42 mg/kg, i.p.; Sigma Co., St. Louis, MO) and fixed by perfusion using saline and cold 4% paraformaldehyde (PFA). Their brains were removed using the same experimental protocol used for qRT-PCR analysis and post-fixed in fresh 4% PFA overnight at 4°C, dehydrated in 30% sucrose at 4°C for 24–48 h, and stored at −80°C. Coronal sections (15-μm thick) were cut using a cryostat microtome (−25°C), and immunohistochemistry was subsequently performed. The tissue sections were post-fixed in 4% PFA for 10 min. The sections were incubated with the CART antibody (1:50) di-
luted in TBS overnight at 4°C following a blocking step in diluted normal serum for 30 min. After three washes with TBS-T, the sections were bound with a diluted biotinylated secondary antibody solution and the Vectastain ABC reagent. The sections were incubated in a diamino-benzidine (DAB) solution until the desired stain intensity developed. It was then counterstained with a hematoxylin. Finally, sections were dehydrated in ethanol, cleared in xylene, mounted with permount (Millipore Co., Bedford, MA, USA), and evaluated using light microscopy (Carl Zeiss Co., Jena, Germany). To determine the expression of CART, the stained cells were counted. The twelve coronal sections with four different animal brains in each group were analyzed. Four striatal regions, including the dorsolateral (DL) part of caudate putamen (CP), ventrolateral (VL) part of CP, NAc shell, and NAc core in each section were selected. Cells at three randomly selected areas (100 × 100 μm) in each region were assessed. The immunoreactive cells by CART antibody were counted and expressed as percentage of stained cells. The quantity of CART-positive cells was expressed as the average number of reactive cells per high-power field.

**Western blot analysis**

WT and KO mice were respectively sacrificed by decapitation 22 h after the last administration of cocaine or saline. The striata, including the NAc, were extracted at the coronal level at +1.6 and +1.0 mm from the bregma, according to the stereotaxic atlas [25]. The striata were dissected in ice-cold saline and homogenized in the protein extraction solution (Millipore Co., Bedford, MA, USA). The supernatant was collected and stored frozen (-20°C). The protein concentration of the supernatant was determined using the Bradford method with bovine serum albumin as the standard. Equal amounts of the proteins were separated on a SDS/16%-polyacrylamide gel and subsequently transferred to a polyvinylidene difluoride (PVDF) membrane. The membrane was blocked in 0.5% non-fat milk in TBS-T, followed by three washes with TBS-T. The membranes were bound with specific antibodies using the SNAP id system (Millipore Co., Bedford, MA, USA). Anti-CART (1:67 dilution) antibodies were used in this study. The membrane was incubated with the corresponding conjugated immunoglobulin G-horseradish peroxidase. The immunoreactive cells by CART antibody were counted and expressed as percentage of stained cells. The quantity of CART-positive cells was expressed as the average number of reactive cells per high-power field.

**CAMP analysis**

WT and KO mice were respectively sacrificed by decapitation 22 h after the last administration of cocaine or saline. The striata, including the NAc, were extracted at the coronal level at +1.6 and +1.0 mm from the bregma, according to the stereotaxic atlas [25]. The striata were dissected in ice-cold saline and homogenized in the protein extraction solution. The cAMP levels were determined using a commercial enzyme-linked immunoassay (ELISA). The absorbance for this developed color was measured by the spectrofluorometer (BMG Co., Ortenberg, Germany) for emission at 450 nm. Measurement of the absorbance using the cAMP standard allows us to calculate the absolute amount of cAMP in a sample of interest. The amount of tissue was 5.0 mg, and the results were expressed as nmol (nM) cAMP/mg tissue.

**PKA radiometric analysis**

WT and KO mice were respectively sacrificed by decapitation 22 h after the last administration of the chemicals or saline. The striata, including the NAc, were extracted at the coronal level at +1.6 and +1.0 mm from the bregma, according to the stereotaxic atlas [25]. The striata were dissected in ice-cold saline and homogenized in protein extraction solution. The PKA protein was purified from the total protein (300 μg) by immunoprecipitation using the anti-PKA antibody. The PKA immunoprecipitate was dissolved in assay dilution buffer for radiometric analysis. Equal amounts of the PKA immunoprecipitate were added to incubation tubes containing 1.67 μM CAMP, 0.08 mM kemptide, 0.33 μM PKC/CaMK inhibitor cocktail, and 1.67 μCi of the [γ-32P]ATP/magnesium/ATP cocktail brought to a total volume of 60 μl with PBS. The complex solution was incubated with shaking at 30°C for 10 min. A square piece of numbered P81 phosphocellulose paper was blotted with a 25-μl aliquot and then washed with 0.75% phosphoric acid 3 times and with acetone once. The result was read in a Wallac 1450 Microbeta Trilux liquid scintillation counter (Cardinal Health Co., Dublin, OH, USA) and calculated by comparing the counts per minute (CPM) for PKA with the CPM for control samples that did not contain the enzyme. The results are expressed as pmol phosphate incorporated into Kemptide/min/μg protein.

**Statistical analysis**

The results are presented as the means±SEM. The significance of the effects was assessed between treatment groups by Student’s t test and within groups by one-way analysis of variance (ANOVA) followed by Dunnett’s post

**Fig. 1. Effects of cocaine (5, 15 and 30 mg/kg) on the locomotor activity of mice for 7 days.** The data are given as the means±SEM of at least 8 mice. *p<0.01 and ***p<0.001, compared with that of the saline group. *p<0.05, **p<0.01 and ***p<0.001, compared with that of the cocaine group on the 1st day.
RESULTS

Repeated cocaine induced behavioral sensitization in mice

Repeated administrations of cocaine (5 and 15 mg/kg) induced behavioral sensitization (n=8~10, p<0.001 and p<0.05, Fig. 1), but not at 30 mg/kg used in this experiment. Cocaine-induced behavioral sensitization was initiated on the 2nd day [F(3, 34) = 8.89, p<0.001, Fig. 1], peaked on the 5th day [F(3, 31)=28.89, p<0.001, Fig. 1] and continued to the 7th day.

Repeated cocaine increased the levels of CART mRNA and peptides in the mouse striatum

In the qRT-PCR experiments, repeated cocaine (5, 15 and 30 mg/kg) administration increased the level of CART mRNA by 3.1-, 3.5-, and 2.8-fold, respectively, on the 5th day (Fig. 2A). Mice given 15 mg/kg of cocaine had significantly elevated the levels of CART mRNA on the 3rd and 5th days (n=4, p<0.01, Fig. 2B), which peaked on the 3rd day (n=4, p<0.01, Fig. 2B) and continued to the 7th day.

Western blot analysis revealed that cocaine increased the levels of the CART peptides in the striata in an inverted...
Over-Expression of CART Peptides by Cocaine

"U" manner. Cocaine significantly elevated the expression of CART peptides on the 5th day (n=4, p<0.01, Fig. 2C). A significant increase in levels of the CART peptides was observed on the 5th day (n=4, p<0.01, Fig. 2D) after mice were given 15 mg/kg of cocaine when compared with the control (saline, n=4) group, but returned to the control levels on the 7th and 14th days.

Distribution of CART+ cells were found in the cocaine-treated mice striatum

Within the NAc shell of the striata in mice treated with saline (n=4), 21±2% of cells were CART+, whereas 40±5% of CART+ were found in mice treated with cocaine (15 mg/kg, n=4) on the 5th day (Fig. 3A). The proportion of CART+ cells within DL part of CP in the cocaine-treated mice was 32±4% (Fig. 3C). These results indicated a higher proportion of CART+ cells in the cocaine-treated mice were present within the NAc shell than DL part of CP. However, the proportion of CART+ cells in the VL part of CP and NAc core was not significantly different in cocaine-treated mice compared with saline-treated mice (Fig. 3B).

D1R and D2R antagonists inhibited cocaine-induced behavioral sensitization and CART peptides over-expression

Both SCH 23390 (D1R antagonist, 0.25 mg/kg) and raclopride (D2R antagonist, 0.4 mg/kg) inhibited behavioral sensitization induced by cocaine (15 mg/kg) on the 5th day of treatment (F(4,41)=25.51, p<0.001, n=8, Fig. 4A). Cocaine-induced behavioral sensitization was also significantly decreased by the concomitant administration of SCH 23390 and raclopride (n=8, p<0.001, Fig. 4A). Subsequently, each SCH 23390 and raclopride treatment inhibited CART peptides over-expression (n=4, p<0.001, Fig. 4D), whereas concomitant administration of two antagonists inhibited CART peptides over-expression (n=4, p<0.001, Fig. 4D), cAMP (n=4, p<0.001, Fig. 4B), and PKA activity (n=4, p<0.01 and p<0.001, Fig. 4C), and pCREB (n=4, p<0.01 and p<0.001, Fig. 4E) more than SCH 23390 and raclopride alone. It is suggested that cocaine-induced overexpression of CART peptides were directly mediated simultaneously by both D1R and D2R-mediated signaling pathways. However, the level of CREB in the striata did not change significantly after

Fig. 4. Effects of SCH 23390 (0.25 mg/kg) and raclopride (0.4 mg/kg) on cocaine (15 mg/kg)-induced locomotor, CART expression, cAMP levels, PKA activity, and pCREB levels in the mice striatum. (A) Cocaine was administered in mice one time per day for 5 days. The locomotor activity was measured using the tilting-type ambulometer. (B) The cAMP levels were measured using the competition enzyme-linked immunosassay (ELISA). (C) After PKA was purified by immunoprecipitation, the PKA activity was measured using a radiometric assay. The CART levels (D), CREB phosphorylation (E), and (F) CREB levels were observed using western blotting. A quantitative analysis was conducted by measuring the relative OD of the immunoreactive signal for preproCART, pCREB, CREB, and GAPDH. Data represent the means±SEM of 4 mice. The total twenty mice were used. **p<0.01 and ***p<0.001, compared with that of the control (saline) group; +p<0.01 and +++p<0.001, compared with that of the cocaine group; *p<0.01 and ****p<0.001, compared with that of the raclopride group.
administration of cocaine alone or in combination with SCH 23390 and raclopride, and it appeared to be almost the same as the control level (n=4, Fig. 4F).

**Cocaine-induced behavioral sensitization and CART peptides expressions were found in D1R- and D2R-KO mice**

The repeated cocaine administration induced BS on the 5th day compared with the control (saline, n=4) group in D1R- (F21=27.24, p<0.001, Fig. 5A) and D2R-KO mice (F21=15.97, p<0.01, Fig. 5A). Furthermore, repeated administration of cocaine increased the CART peptide levels in the D1R-KO (cAMP, p<0.001; n=5, Fig. 5B) and D2R-KO mice (cAMP, p<0.01; saline, n=4, Fig. 5B) when compared with the control (saline, n=4) group. Accordingly, the increases in cAMP level, PKA activity, and pCREB levels after cocaine treatment were observed in the D1R-KO (cAMP: p<0.01, n=5, Fig. 5B; PKA: p<0.001, n=5, Fig. 5C; pCREB: p<0.01, n=5, Fig. 5E) and D2R-KO mice (cAMP: p<0.05, n=4, Fig. 6B; PKA: p<0.01, n=4, Fig. 6C; pCREB: p<0.05, n=4, Fig. 6E).

Cocaine administration did not significantly increase locomotor activity (saline, p<0.05, n=4; cocaine, p<0.01, n=5; Fig. 5A). However, CART expression was decreased in cocaine treated D1R-KO mice (saline, p<0.01, n=4; cocaine, p<0.05, n=5; Fig. 5D). cAMP/PKA/pCREB signaling pathways were reduced in D2R-KO mice (cAMP: p<0.01, n=4~5, Fig. 5B; PKA: p<0.01, n=4~5, Fig. 5C; pCREB: p<0.05, n=4~5, Fig. 5E) on the 5th day. In contrast, CART peptides were increased in cocaine treated D2R-KO mice (p<0.05, n=4, Fig. 6D) and pCREB (p<0.05, n=4, Fig. 6E). However, cAMP levels and PKA activity did not increase significantly increase repeated administrations of saline or cocaine (n=3~5, Fig. 5F and Fig. 6F).

**DISCUSSION**

It has been well known that repeated intermittent administration of cocaine produces progressive enhancement in locomotor activity, a phenomenon known as behavioral sensitization. The close correlations between behavioral sensitization and the overexpression of CART peptides by cocaine were suggested. After peaking on the 5th day, both behavioral sensitization and overexpression of CART proteins gradually decreased on the 7th day [27,28]. It was also confirmed in the present study. Repeated administration of cocaine started to simultaneously induce behavioral sensitization and CART over-expression on the 3rd day. These results are consistent with the previous findings that the level of CART peptides elevates at the onset, peaks during the process, and returns to biophysiological levels at the end in response to various repeated stimuli [29-31]. CART peptides are peptidergic neurotransmitter that is expressed.
Over-Expression of CART Peptides by Cocaine

95

Fig. 6. Effects in WT mice and D1R-KO mice treated with cocaine (15 mg/kg) on the 5th day on locomotor activity (A), cAMP levels (B), PKA activity (C), CART levels (D), pCREB levels (E), CREB levels (F). Quantitative analysis was conducted by measuring the relative OD of the immunoreactive signal for prepro CART, pCREB, CREB, and GAPDH. Data represent the means±SEM of 3–4 mice. The total sixteen mice were used. *p<0.05, **p<0.01 and ***p<0.001, compared with that of the control (saline) group; #p<0.05, compared with that of the KO or WT mice.

in regions of the brain regions involved in critical biological processes such as feeding, stress, and in areas associated with drug reward and abuse, including the dopamine-rich NAc, which can be considered part of the basal ganglia [32]. Both CART over-expression and behavioral sensitization are closely related to DR. The DR-mediated cAMP/PKA signaling pathway activation underlies the over-expression of CART peptides induced by cocaine in the striatum [16,33]. Dopaminergic neurons in the VTA and substantia nigra (SN) receive a CART input. There is also sufficient evidence for CART systems being affected by dopamine. CART was observed to particularly regulated by dopamine in the NAc, at least partly by D1R [18]. However, it remains unclear how CART peptides over-expression is mediated via D1R and D2R in the development of cocaine-induced behavioral sensitization.

From these experiments, both D1R- and D2R-antagonists blocked cocaine-induced behavioral sensitization. Moreover, the combined treatment of D1R and D2R antagonists produced less locomotor activity than D1R and D2R antagonists alone. In terms of mediation of reward induced by cocaine, the stimulation of D1R results in fast neuronal activation, and the stimulation of D2R results in progressive neuronal deactivation [7]. The weaker behavioral sensitization by cocaine was developed in the D1R- and D2R-KO mice compared to the WT mice, although repeated cocaine administration still induced behavioral sensitization in both D1R- and D2R-KO mice. Behavioral effects of psychostimulants were also reduced in the mutant CART KO mice [34].

Co-administration of CART peptides with D1R and D2R agonists into the NAc reduces locomotor activity, whereas co-administration of CART peptides enhances the increase in locomotor activity induced by D1R agonists [16]. Both behavioral sensitization and CART peptides may be closely correlated with simultaneous D1R and D2R activation (is that correct?). The minimal estimated proportion of neurons expressing D1R is 64% in the NAc shell, and 57% and 59% in the CP and the NAc core, respectively, although various regions in the striatum comprise a higher percentage of neurons expressing D1R than D2R according to previous reports [6]. We found that there are also differences in the proportion of CART+ cells in various regions of the striatum. The proportion of CART+ cells are significantly increased by cocaine within the NAc shell and the CP-DL, but not within the NAc core and CP-VL part. CART cells in the NAc of rats is co-localized with substance P, dynorphin, and D1R [32]. It has been suggested that behavioral sensitization and the expression of CART peptides is predominantly mediated by D1R than D2R.

Psychostimulants-induced over-expression of CART peptides and phosphorylation of ERK and CREB was mediated by D1R which stimulated the cAMP/PKA signaling in striatogiral (direct pathway) neurons [6,35]. Because the over-expression of CART peptides after repeated cocaine treatment is shown to be activated by cAMP, PKA and pCREB [36], our present study suggest that inhibiting overexpression of CART peptides could be responsible for attenuation of cAMP, PKA and pCREB levels in mice lacking D1R using a phar-
activation in mice treated with repeated cocaine treatments by Bertran-Gonzalez et al. [6] who reported that early ERK preclude the contribution of CART over-expression induced demonstrated that raclopride partially decreased cocaine-induced over-expression of CART peptides by ethanol in the rat NAc [19]. Valjent et al. demonstrated that raclopride partially decreased cocaine-induced ERK activation in the DL striatum of mice [35]. Thus, gene expression is mediated, in part, by activation of D2R in striatopallidal neurons. Interestingly, CART expression after repeated cocaine treatment in D2R-KO mice was increased, compared to WT mice which in contrast appeared to lose their IEG activation response to cocaine treatment [39]. The underlying mechanism of the higher sensitivity of CART peptides to cocaine in D2R-KO mice is presently unclear. But, it may correlate with increased stimulation of D2R due to leading to the increase of DA release in D2R-KO mice, which may delete presynaptic autoreceptors [40]. It may also depend on the activation of this gene by the release of pituitary hormone in the absence of D1R [41,42].

In summary, D1R- and D2R-mediated signal pathways simultaneously modulated the expression of CART mRNA and peptides during the development of behavioral sensitization after repeated cocaine administration, which is consistent with previous findings [9,18,36]. CART + cells in the NAc shell and DL caudate putamen were significantly increased in the cocaine-treated mouse. D1R- and D2R-antagonists blocked CART over-expression induced by cocaine. Cocaine induced CART over-expression was weaker in the D1R-KO mice and stronger in the D2R-KO mice when compared to the WT mice. These results suggest that CART over-expression induced by repeated administration of cocaine could be more predominantly involved in D1R.

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