Neisseria macacae bacteremia: report of two cases with a literature review

Yasumasa Yamamoto 1*, Norihiko Terada 1, Tomoyo Sugiyama 1, Hanako Kurai 1 and Kiyofumi Ohkusu 2

Abstract

Background: Neisseria macacae was discovered in the oral cavity of monkeys in 1983. In humans, it has been isolated from the upper respiratory tract of neutropenic patients. However, only two cases of N. macacae bacteremia have been reported in a 65-year-old man with infective endocarditis and a 5-month-old child with fever and petechiae. There are no reports of infections in cancer patients. Here, we present two cases of N. macacae bacteremia in cancer patients.

Case presentation: In the first case, a 42-year-old woman who underwent ovarian cancer surgery presented with duodenal invasion associated with multiple lymph node metastasis. N. macacae was isolated from her blood culture and identified using matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry (MS). In the second case, a 69-year-old woman with a long-standing history of esophagogastric junction cancer presented with fever. She had stage IVB cancer with lung, bone, and multiple lymph node metastasis. The last chemotherapy was administered 5 weeks before N. macacae was detected using MALDI-TOF MS and nitrate test negative. In both cases, transthoracic echography showed no vegetation. Antibiotics were administered for 14 and 13 days in the first and second cases, respectively. In both cases, fever alleviated on day 4 of antibiotic administration. Both patients were discharged after their conditions improved.

Conclusions: This, to our knowledge, is the first report of N. macacae bacteremia in cancer patients. Both patients, mucosal damage was observed in the upper gastrointestinal tract. Therefore, exclusion diagnosis suggested that bacteremia invasion was caused by mucosal rupture in both cases. Both cases responded well to treatment with β-lactam antibiotics and improved after 2 weeks. Modifying the treatment based on the source of the infection may shorten the treatment period. Therefore, further research on N. macacae bacteremia is necessary.

Keywords: Neisseria macacae, Neisseria species, Bacteremia, Cancer
(MALDI-TOF) mass spectrometry (MS) and 16S RNA analysis.

Case presentation

Case 1

A 42-year-old woman with a long-standing history of ovarian cancer presented with fever. She had stage IVB cancer with multiple lymph node metastasis and peritoneal dissemination. Abdominal total hysterectomy, bilateral salpingo-oophorectomy, and omentectomy were performed 4 weeks ago. The patient denied any contact with monkeys. On examination, she appeared unwell and was experiencing intense rigors. Her temperature was 38.3 °C, heart rate was 95 beats/min and regular, blood pressure was 102/61 mmHg, respiratory rate 18 breaths/min, and oxygen saturation was 96% in room air. Laboratory investigations revealed an elevated white blood cell count of 8770/μL (neutrophil count 6840/μL), microcytic anemia (hemoglobin level 9.3 g/dL), C-reactive protein (CRP) 19.99 mg/dL (normal < 0.30 mg/dL), and persistently elevated cholestatic liver enzyme levels (gamma glutamyltransferase 78 U/L [normal < 30 U/L] and alkaline phosphatase 575 U/L [normal 115–359 U/L]). Mild tenderness in the upper abdomen was observed. Blood and urine cultures were performed. No significant bacteria were detected in the urine cultures, including Neisseria. A computed tomography scan of the patient revealed duodenal invasion with enlarged lymph nodes. Empirical antimicrobial therapy comprising intravenous piperacillin-tazobactam at 4.5 g was administered every 6 h. Only the aerobic culture showed bacterial growth 34 h after starting culture using BD resin bottles (Becton Dickinson, Tokyo, Japan) and the BD Bactec™ FX blood culture system (Becton Dickinson, Tokyo, Japan). Subsequently, gram-negative diplococcus was identified and confirmed as N. macacae using MALDI-TOF MS (Biotyper:BRUKER Score Value 2.10) (Table 1). No other organisms were detected in the blood culture.

Table 1 Case 1: MALDI-TOF MS results

| Rank | Matched Pattern            | Score Value |
|------|----------------------------|-------------|
| 1    | Neisseria macacae DSM1917ST | 2.05        |
| 2    | Neisseria flavescens C12PGM | 1.89        |
| 3    | Neisseria mucosa DSM17611T  | 1.74        |
| 4    | Neisseria mucosa DSM1591PGM | 1.70        |
| 5    | Neisseria lactamica VA33589 | 1.67        |
| 6    | Neisseria sp 0807 M15057901 IBS | 1.65       |
| 7    | Neisseria flavescens DSM17633T | 1.58       |
| 8    | Neisseria sicca DSM17713T   | 1.56        |
| 9    | Neisseria perflava DSM18009T | 1.55        |
| 10   | Neisseria subflava DSM17610T | 1.53        |

Broth microdilution method (DP34, Eiken, Japan) showed that the bacterial culture was susceptible to ceftriaxone, ampicillin, meropenem, and minocycline but resistant to penicillin, trimethoprim-sulfamethoxazole, and levofloxacin (Clinical Laboratory Standards Institute [CLSI] testing methods and MIC break point for N. meningitidis was substituted). Transthoracic echography showed no vegetations. Fever alleviated by day 4 of antimicrobial administration. On day 6, her treatment was changed to intravenous ampicillin-sulbactam 3 g every 6 h. The total antibiotic treatment period was 14 days. Her condition improved, and she was discharged. After discharge, she was followed up for 1 month in an outpatient setting, and there was no recurrence.

Case 2

A 69-year-old woman with a long-standing history of esophagogastric junction cancer presented with fever. She had stage IVB cancer with lung, bone, and multiple lymph node metastasis. The last chemotherapy was performed 4 weeks ago. The patient denied any contact with monkeys. On examination, she appeared unwell and was experiencing intense rigors. Her temperature was 37.8 °C, heart rate was 100 beats/min and regular, blood pressure was 137/82 mmHg, respiratory rate was 18 breaths/min, and oxygen saturation 99% in room air. Laboratory investigations revealed an elevated white blood cell count of 5450/μL (neutrophil count 4180/μL), microcytic anemia (hemoglobin level 9.8 g/dL), CRP 0.64 mg/dL [normal < 0.30 mg/dL], and persistently elevated gamma glutamyltransferase 40 U/L [normal < 30 U/L]. Mild tenderness in the upper abdomen was observed. Blood and urine cultures were performed. No significant bacteria were detected in the urine cultures, including Neisseria. A computed tomography scan of her abdomen was performed which revealed esophagogastric junction cancer and multiple lymph node metastatic lesions which were enlarged. Empirical antimicrobial therapy comprising intravenous piperacillin-tazobactam was administered at 4.5 g every 6 h. Only the aerobic culture showed bacterial growth 38 h after starting culture. 16S rDNA sequence of this organism was difficult to determine because there were three strains with more than 99.9% homology with N. macacae, N. sicca, and N. mucosa with a difference in one, two, and three base values. Subsequently, the gram-negative diplococcus was identified as N. macacae based on MALDI-TOF MS analysis (Table 2) and biochemical characterization (nitrate test negative).

No other organisms were isolated from the blood culture. Broth microdilution method (DP34, Eiken, Japan) showed that the bacterial culture was susceptible to ceftriaxone, ampicillin, meropenem, and minocycline but resistant to penicillin, trimethoprim-sulfamethoxazole,
and levofloxacin (CLSI testing methods and MIC break point for \textit{N. meningitidis} was substituted). Transthoracic echography showed no vegetation. Fever had alleviated by day 4 of antimicrobial administration. Her treatment was changed to intravenous ceftriaxone at 2 g every 24 h. The total antibiotic treatment period was 13 days. Her condition improved, and she was discharged and followed up for 1 month in an outpatient setting, with no recurrence.

### Discussion and conclusions

\textit{N. macacae} bacteremia is a rare disease, and its entry point and mechanism are often unknown. To our knowledge, there have been 10 specimens of \textit{N. macacae} isolated in human samples, including these two cases (Table 3). There have only been two reports of \textit{Neisseria} bacteremia in humans including a 5-month-old child and a healthy adult [1, 2]. This is the first report of two cases of \textit{Neisseria} bacteremia in patients with cancer, one of whom had recovered from cancer. \textit{N. macacae} is known to reside in the mouth of monkeys [3]; however, neither of our patients had a history of contact with monkeys. Only one case of infectious endocarditis in an adult with \textit{N. macacae} bacteremia was reported [2].

Previously, reports of \textit{N. macacae} infection in humans include peritonitis in patients on peritoneal dialysis [5]. Additionally, \textit{N. macacae} has been isolated from the upper respiratory tract of neutropenic patients [4]. No neutropenia or no respiratory infections such as pneumonia and infective endocarditis were observed in our patients. In addition, echoendoscope colonization was reported, suggesting that it may have been established in the upper digestive tract of humans [6]. Among patients who used antibiotics within a month, \textit{N. macacae} accounted for 4.5% of \textit{Neisseria} species that colonized in the oral cavity [4]. Neither of our patients had used antibiotics within 1 month of admission, but both had previously been treated with antibiotics and chemotherapy. In both cases, mucosal damage was observed in the upper gastrointestinal tract. Therefore, exclusion diagnosis suggested that bacteremia invasion was caused by mucosal rupture in our cases.

There are no clear susceptibility testing methods or criteria for \textit{N. macacae} detection, and the treatment for \textit{Neisseria} bacteremia is unknown. Therefore, susceptibility testing was performed using the CLSI meningococcal criterion; however, it is unknown whether these results are appropriate. Both patients responded well to treatment with \(\beta\)-lactam antibiotics and improved after 2 weeks. The duration of treatment is unknown. Modifying the treatment based on the source of the infection may shorten the treatment period. Therefore, further research on \textit{N. macacae} bacteremia is necessary.

Common \textit{Neisseria} species that are pathogenic to humans include \textit{N. gonorrhoeae} and \textit{N. meningitidis}. 16S sequence analysis may not be sufficient to identify \textit{Neisseria} species due to high intraspecies sequence variation in the 16S rRNA gene [4]. \textit{Neisseria} species can be differentiated based on their morphological, physiological, and biochemical properties and MALDI-TOF MS results. Commensal \textit{Neisseria} species that colonize oral and nasal cavity sites have only been rarely associated with disease [7]. Donati et al. have shown that commensal species have tropism for different sites in the oral cavity and oropharynx. \textit{Neisseria meningitidis} is enriched for colonization in the throat, \textit{N. flavescens} and \textit{N. subflava} populate the tongue dorsum, and \textit{N. sicca}, \textit{N. mucosa}, and \textit{N. elongata} the gingival plaque [8].

Interestingly, Bennett et al. have demonstrated that \textit{N. subflava} and \textit{N. flavescens} are phylogenetically the same species. Similarly, researchers have proposed that both \textit{N. sicca} and \textit{N. macacae} be reclassified as \textit{N. mucosa} [9, 10].

The limitation of this report was that \textit{Neisseria} spp. is difficult to fully identify with only 16sRNA. In 34% (29/85) cases of identification of the \textit{Neisseria} spp., 16S rRNA sequence analysis indicated the possibility of more

### Table 2 Case 2: MALDI-TOF MS results

| Rank | Matched Pattern                  | Score Value |
|------|----------------------------------|-------------|
| 1    | \textit{Neisseria macacae} DSM1917ST | 2.10        |
| 2    | \textit{Neisseria lactamica} CCUG26481 | 1.83        |
| 3    | \textit{Neisseria subflava} DSM17610T | 1.81        |
| 4    | \textit{Neisseria mucosa} DSM17611T | 1.78        |
| 5    | \textit{Neisseria lactamica} CCUG44970 | 1.77        |
| 6    | \textit{Neisseria elongata} spp glycolytica DSM23337T | 1.77        |
| 7    | \textit{Neisseria lactamica} CCUG17628 | 1.77        |
| 8    | \textit{Neisseria subflava} CCUG29402 | 1.70        |
| 9    | \textit{Neisseria lactamica} CCUG796 | 1.70        |
| 10   | \textit{Neisseria lactamica} CCUG17016 | 1.70        |

### Table 3 \textit{N. macacae} isolated in human samples review

| Case | Sex | Age (years) | Samples                  | References  |
|------|-----|-------------|--------------------------|-------------|
| 1    | F   | 42          | Blood culture            | Present case 1 |
| 2    | F   | 69          | Blood culture            | Present case 2 |
| 3    | M   | 0.66 (8 months) | Blood culture          | [1]         |
| 4    | M   | 65          | Blood culture            | [2]         |
| 5    | F   | 52          | Peritoneal fluid culture | [5]         |
| 6    | NA  | NA          | Mouth                    | [4]         |
| 7    | NA  | NA          | Throats swab             | [4]         |
| 8    | NA  | NA          | Respiratory tract        | [4]         |
| 9    | NA  | NA          | Respiratory tract        | [4]         |
| 10   | NA  | NA          | Respiratory tract        | [4]         |

NA Not available
than one species [4]. Therefore, the usefulness of MALDI-TOF for the identification of Neisseria spp. is being examined. In CASE 2, the 16sRNA analysis results showed that N. macacae, N. sicca and N. mucosa were the three possible species. A combination of biochemical tests ruled out N. mucosa but did not differentiate N. macacae from N. sicca. However, N. sicca was not within rank 10 on the MALDI-TOF, and therefore N. macacae was identified.

This case report suggests that N. macacae bacteremia may be caused by gastrointestinal mucosal damage. Scrutiny of the upper gastrointestinal tract should be considered in patients with N. macacae bacteremia caused by an unknown source.

Abbreviations
MALDI-TOF: Matrix-assisted laser desorption/ionization time-of-flight; MS: Mass spectrometry; CRP: C-reactive protein; CLSI: Clinical Laboratory Standards Institute

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Authors’ contributions
YY designed the study; YY, and HK assessed the patient's documentation and evaluated study inclusion criteria; YY and NT were involved in data collection and interpretation; TS and KO conducted the laboratory analyses. All of the authors interpreted the data, drafted and critically revised the manuscript, and approved the final version.

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Consent for publication
Written informed consent was obtained from both the patients for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor of this journal.

Competing interests
The authors declare that they have no competing interests.

Author details
1Division of Infectious Diseases, Shizuoka Cancer Center Hospital, 1007 Shimonagakubo, Suntou-gun, Shizuoka, Nagaiizumi 411-8777, Japan.
2Department of Microbiology Tokyo Medical University, Shinjuku, Tokyo, Japan.

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References
1. Schouten MA, Suvandy F, Quaak M. Neisseria macacae bacteraemia in a child with fever and petechiae. Ned Tijdschr Med Microbiol. 2014;22(Suppl): S83–4.
2. Vecten M, Martel H, Casalta JP, Hubert S, Lepidi H, Habib G, et al. Fatal Neisseria macacae infective endocarditis: first report. Infection. 2017;45:369–71.
3. Vedros NA, Hoke C, Chun P. Neisseria macacae sp. nov., a new Neisseria species isolated from the oropharynges of rhesus monkeys (Macaca mulatta). Int J Syst Evol Microbiol. 1983;33:515–20.
4. Arj M, Achour W, Ben HA. Intraspecific 16s RNA gene diversity among clinical isolates of Neisseria species. APMIS. 2013;122:437–42.
5. Iyama T, Hamada S, Takata T, Hoi S, Fukuda S, Iromoto H. Refractory peritoneal dialysis peritonitis due to Neisseria macacae: a case report and review of the literature. Intern Med. 2020;4832–20.
6. Coton T, Sbai W, Beaussc M, Luciano L, Gay G, Garnotel E. New flexible endoscopes: surprising bacterial colonization post-disinfection. Clin Res Hepatol Gastroenterol. 2017;41:e63–4.
7. Dong PN, Pham LQ, Nguyen HT, Nguyen MXB, Nguyen TV, May F, et al. Decreased cephalosporin susceptibility of oropharyngeal Neisseria species in antibiotic-using men who have sex with men in Hanoi, Vietnam. Clin Infect Dis. 2020;70:1169–75.
8. Donati C, Zollo M, Albanese D, Truong DT, Asnicar F, lebba V, et al. Uncovering oral Neisseria tropism and persistence using metagenomic sequencing. Nat Microbiol. 2016;1:16070.
9. Bennet JS, Jolley KA, Earle SG, Coton C, Bentley SD, Parkhill J, et al. A genomic approach to bacterial taxonomy: an examination and proposed reclassification of species within the genus Neisseria. Microbiology. 2012;158:1570–80.
10. Weyand NJ. Neisseria models of infection and persistence in the upper respiratory tract. Pathogen Dis. 2017;75:fx031.

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