New advances on Au–magnetic organic hybrid core–shells in MRI, CT imaging, and drug delivery

Fatemeh Mohajer, Ghodsi Mohammadi Ziarani and Alireza Badiei

Magnetic nanoparticles have been widely studied for various scientific and technological applications such as magnetic storage media, contrast agents for magnetic resonance imaging (MRI), biolabelling, separation of biomolecules, and magnetic-targeted drug delivery. A new strategy on Au–magnetic nano-hybrid core–shells was applied in MRI, CT imaging, and drug delivery, which has been received much attention nowadays. Herein, the designing of different magnetic core–shells with Au in MRI and cancer treatment is studied.

1. Introduction

There are various nanoparticles for medical applications, including carbon dots and organic core/shell nanoparticles. Among all these materials, Fe₃O₄ (ref. 3) and MnFe₂O₄ (ref. 4) gain the first step in terms of usage. However, TiO₂, SnO₂, Ag, Au, S, BaTiO₃, and ZnO are known for their electrical applications. Through the coating process, various materials such as polyaniline (PA), polystyrene (PS), graphene, oleic acid, polyvinylpyrrolidone (PVP), hyperbranched aromatic polyanime (HBP), poly(methylmethacrylate) (PMMA), are differently used, and the provided materials are used extensively for batteries, optical sensors, and magnetic imaging. The metal–organic frameworks (MOFs) have received much attention in terms of bioimaging in living cells, as porous functional materials, which have large surface areas, high porosity, fluorescence functionalities, and good biocompatibility. Organic or inorganic fluorescent materials such as fluorescent dyes, quantum dots, and metal nanoclusters can also be applied in medicine for bioimaging.

The importance of magnetite nanoparticles (Fe₃O₄ NPS) was extensively studied due to their many biomedical uses in cancer cells, magnetic resonance imaging (MRI) as a contrast agent, drug delivery, and hyperthermia treatment. In terms of toxicity and susceptibility, magnetite NPs are safe and have superparamagnetic applications. For improving the stability and dispersing the NPs in water, Fe₃O₄ NP was coated with Au NPs to improve the stability and dispersion of the NPs in aqueous media as efficient materials due to their biocompatibility, stability, resistance, which provided the potential medical requests. Gold nanoparticles (AuNPs) have received much attention because of their unique physicochemical activities such as biological, chemical, and biomedical implementations. AuNPs have been used for sensing, catalysis, imaging and diagnosis, and therapy (Fig. 1). Core–shell (CS) nanostructures have received much attention due to the known constituents and configuration. In this area, the immunization of Au on Fe₃O₄@ promoted the optical properties of computed tomography (CT) and MRI, which increased the resolution in 3D visual images in this process. Through the importance of bioimaging in living cells, metal–organic frameworks (MOFs) were used which organic or inorganic fluorescent compounds like quantum dots, fluorescent dyes, nanosheets, and metal nanoclusters can provide modified MOFs to give fluorescent nano-composites for an imaging agent. In continuing our previous work, we decided to introduce another application of core shells, including Au–magnetic organic hybrid in MRI, CT imaging, and drug delivery.

Fig. 1 The application of the different nanoparticles as core–shells.
2. Au–magnetic organic hybrid core–shells

2.1. Fe$_3$O$_4$@SiO$_2$@Au NPs

Fe$_3$O$_4$ NP was provided by ferrous (Fe$^{2+}$) and ferric (Fe$^{3+}$) through the co-precipitation method with sodium hydroxide (NaOH) under inert nitrogen gas at room temperature. To obtain the Fe$_3$O$_4$@Au CSNPs, the Fe$_3$O$_4$ was added to the HAuCl$_4$ solution in Na$_3$C$_6$H$_5$O$_7$ and sodium citrate (CS) under sonicated conditions. According to the result, Fe$_3$O$_4$@Au CSNPs are suitable for CT and MRI imaging. The importance of this method is to give Fe$_3$O$_4$@Au CSNPs through the rapid sonochemical synthesis (Scheme 1).^{28}

2.2. Fe$_3$O$_4$@SiO$_2$@Au NPs

Cisplatin (cPt) was doped on Fe$_3$O$_4$@SiO$_2$@Au to cure cancer, which was detected through MRI images. Hydrophobic Fe$_3$O$_4$ NPs were synthesized by a thermal decomposition method using FeCl$_3$·6H$_2$O and sodium oleate in ethanol/hexane. In the next step, Fe$_3$O$_4$@SiO$_2$ was obtained through the reverse microemulsion method. In this method, the mixture of Igepal CO-520 in cyclohexane and ammonium hydroxide was added to...
Fe₃O₄ NPs, followed by the addition of tetraethoxysilane (TEOS) and (3-aminopropyl)triethoxysilane (APTES) to yield Fe₃O₄@SiO₂ core–shell nanostructures, which were added to Au NPs to obtain Fe₃O₄@SiO₂@Au NPs. In the next step, 16-mercaptohexadecanoic acid (16-MHDA) as a linker with 16 carbon atoms containing thiol and carboxyl group was linked to the Au NPs surface and cPt, respectively (Scheme 2). The functionalized Fe₃O₄@SiO₂@Au NPs are assessed in photothermal cancer therapy by the irradiation of two colon cancer cell lines (SW480 and SW620) with a laser (λ = 808 nm, W = 100 mW cm⁻²). It is found that laser irradiation showed the mortality of cancer cells. Because of the synergic effect of cPt and Au NPs and laser irradiation, the functionalized Fe₃O₄@SiO₂@Au NPs are used for potential (MRI)-guided stimulated chemo-photothermal treatment of cancer.

2.3. Core–shell iron–gold (Fe@Au)
Core–shell iron–gold (Fe@Au) structures were used for MRI imaging and targeted drug delivery. They were obtained through the reverse micelle method. In this method, the iron–gold nanoparticles were coated with polyglycerol, thiol, and polymerized glycidol. Spherical core particles of iron with a thin layer of gold shell were decorated with 2- mercaptoethanol, which was linked to Au from S head to yield the Au–S on the core–shell surfaces (Scheme 3). The functionalized Fe₃O₄@SiO₂@Au by magnetic iron oxide nanoparticles through the co-precipitation process, followed by silica according to the Stöber method. Consequently, Fe₃O₄ nanoparticles were functionalized by (3-aminopropyl)triethoxysilane (APTES) to provide Fe₃O₄@SiO₂PrNH₂, which was added to the suspension of gold nanoparticles in H₂O to produce Fe₃O₄@SiO₂PrNH₂@Au. Then, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide as human hepatocellular carcinoma was immobilized on the surface of Fe₃O₄@SiO₂-PrNH₂@Au to give Fe₃O₄@SiO₂PrNH₂@Au + drugs, which was applied as MRI and CT agent (Scheme 4). Fe₃O₄@SiO₂-PrNH₂@Au was provided through the synthetic strategy according to the laser ablation in liquid (LAL) as a green route to achieve NPs in one step (Scheme 5).

2.4. Fe₃O₄@SiO₂PrNH₂@Au
Keshtkar et al. designed Fe₃O₄@Au by magnetic iron oxide nanoparticles through the co-precipitation process, followed by silica according to the Stöber method. Consequently, Fe₃O₄ nanoparticles were functionalized by (3-aminopropyl)triethoxysilane (APTES) to provide Fe₃O₄@SiO₂PrNH₂, which was added to the suspension of gold nanoparticles in H₂O to produce Fe₃O₄@SiO₂PrNH₂@Au. Then, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide as human hepatocellular carcinoma was immobilized on the surface of Fe₃O₄@SiO₂-PrNH₂@Au to give Fe₃O₄@SiO₂PrNH₂@Au + drugs, which was applied as MRI and CT agent (Scheme 4). Fe₃O₄@SiO₂-PrNH₂@Au was provided through the synthetic strategy according to the laser ablation in liquid (LAL) as a green route to achieve NPs in one step (Scheme 5).

2.5. Fe/FexOy@Fe₃O₄PrNH₂@Au
Iron/iron oxide nanoparticles Fe/FexOy@Au were provided through the electric arc discharge method. Then, particles were mixed with (3-aminopropyl)trimethoxysilane (APTMS) under sonicated conditions to provide Fe/FexOy@Fe₃O₄PrNH₂, which was functionalized by gold nanoparticles to provide Fe/FexOy@Fe₃O₄PrNH₂@Au.
Fe$_3$O$_4$PrNH$_2$@Au. The designed Fe$_3$/Fe$_3$O$_4$PrNH$_2$@Au nanoparticles as bifunctional magnetic plasmonic nanostructures were used in magnetic resonance imaging and magneto-optical thermal therapies (Scheme 6).  

2.6. Au@Bi$_2$S$_3$ NBs

Au@Bi$_2$S$_3$–PVP nano bone, which can be used as a multimodal imaging agent for PT/PA/CT image, was reported by Ouyang et al.  

In the first step, Au NPs were synthesized\textsuperscript{44} and mixed with l-ascorbic acid (AA), hexamethylenetetramine, and thiocacetamide (TAA) to give Au NR Au@Bi$_2$S$_3$ NBs as the theranostic agent in cancer therapy, which was added to bismuth acetate to give Au@Bi$_2$S$_3$ core–shell NBs, followed by mixing with PVP to produce Au@Bi$_2$S$_3$–PVP NBs nano bone as an imaging agent for applying in the tumor cells (Scheme 7). The structure can be used as a nanocarrier for anticancer drugs (DOX) to be released at a special pH.

2.7. D-Au@Gd&RGD

Gold nanoparticles was provided to be functionalized by mitochondria-targeting group (Mito-S), rhodamine B derivative (RH-S), fluorescein derivative (Flu-S), tetraaza macrocyclic compounds (TAMC-S), cyclic arginine–glycine–aspartic acid peptide (cRGD-S) to yield a mono-sensitive compound. The

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latter was treated with Gd\textsuperscript{3+} to provide d-AuGd\textsuperscript{3+}RGD as the dual-sensitive structure. The application of the synthesized compound was used in MR and fluorescence imaging for a tumor in mice for tumor imaging and penetrate the blood–brain barrier (BBB) for central nervous system (CNS) problems (Scheme 8).

2.8. APG@OVA NPs

The multifunctional gadolinium-doped Au@Prussian blue nanoparticles (Au@PB–Gd@OVA) were designed as MR/SERS bimodal agents. Prussian blue nanoparticles (PB) as cyanide (CN)-bridged coordination polymer were immobilized onto the AuNP core to give a background-free surface-enhanced Raman scattering (SERS) signal. The presence of doped Gd\textsuperscript{3+} provided a sensitive agent for MRI. Through the coating of ovalbumin (OVA) physically, APG@OVA NPs were provided.\textsuperscript{47} First step: The Au NPs were prepared through the classic sodium citrate reduction process using sodium citrate and HAuCl\textsubscript{4} solutions. Second step: K\textsubscript{4}[Fe(CN)\textsubscript{6}] and FeCl\textsubscript{3} were applied as the Prussian blue nanoparticles (PB) agent, which was reacted with GdCl\textsubscript{3}. These two solutions were mixed to provide APG NPs, which was added to the OVA solution to yield Au@Prussian blue-Gd@ovalbumin nanoparticles (APG@OVA NPs) (Scheme 9).\textsuperscript{48}

2.9. Au@MnO\textsubscript{2}

In this study, a gold@manganese dioxide (Au@MnO\textsubscript{2}) as core–shell structure was synthesized, which was functionalized by glutathione GSH as a theranostic agent in photoacoustic and magnetic resonance study. The GSH-triggered Au@MnO\textsubscript{2} was applied in photoacoustic and MRI as a smart theranostic nanostructure for cancer diagnosis and treatment (Scheme 10).\textsuperscript{49}

The nanostructure of AuPd@PVP is important due to their similar magnitudes to the biomolecules, which was used for in biotechnology and medicine. The chloroauric acid hydrated solution was added to the Au precursor; and then disodium tetrachloropalladate was mixed with ascorbic acid as a reducing agent to give AuPd (Scheme 11).\textsuperscript{50} The resulting product AuPd was mixed with polyvinyl pyrrolidone (PVP) as a surfactant to give AuPd@PVP nanoparticles for biocompatibility in vivo and in vitro study in photothermal therapy (PTT) and radiotherapy (RT). Therefore, PTT and RT could be applied for cancer therapy. The AuPd@PVP NPs have photothermal therapy (PTT) activity under NIR laser irradiation at a low power. Moreover, the NPs could be applied in radiotherapy (RT) as the sensitizer agent. Through PTT and RT, AuPd@PVP core–shell nanoparticles could be efficient for cancer therapy.

2.10. Fe\textsubscript{3}O\textsubscript{4}@Au

The manganese dioxide-encapsulated gold nanoparticle (Au@MnO\textsubscript{2} NP) was decorated by polyethylene glycol (PEG) to provide Au@MnO\textsubscript{2}–PEG and decomposed into the naked Au NPs and Mn\textsuperscript{2+} in acidic media. The resulting structure was absorbed by biomolecules to give a stimuli-responsive surface-enhanced Raman scattering (SERS) signal. The presence of doped Gd\textsuperscript{3+} provided a sensitive agent for MRI. Through the coating of ovalbumin (OVA) physically, APG@OVA NPs were provided.\textsuperscript{47} First step: The Au NPs were prepared through the classic sodium citrate reduction process using sodium citrate and HAuCl\textsubscript{4} solutions. Second step: K\textsubscript{4}[Fe(CN)\textsubscript{6}] and FeCl\textsubscript{3} were applied as the Prussian blue nanoparticles (PB) agent, which was reacted with GdCl\textsubscript{3}. These two solutions were mixed to provide APG NPs, which was added to the OVA solution to yield Au@Prussian blue-Gd@ovalbumin nanoparticles (APG@OVA NPs) (Scheme 9).\textsuperscript{48}

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microenvironment (TME); however, in other tissues, which were highly selective to tumors, there was no activation. As a result, TME-triggered exposure with Au NPs and biomolecules could meaningfully increase; moreover, the Raman fingerprints show the complete information, compared with the traditional molecular method. The importance of the SR-SERS probes is to make a difference between tumors and healthy tissues, which might be applicable for the treatment of cancers (Scheme 12).46 In fact, Au@MnO2 nanoparticles were produced through a layer-by-layer approach. Thus, the obtained Au@MnO2-PEG nanoparticles treated with H2O2 in the tumor to provide Mn2+ for improving T1-weighted MR imaging to create O2 for removing the cancer cells to X-rays. Therefore, Au nanoparticles increase the X-ray energy in tumor cells, and MnO2 reacts with endogenous tumor H2O2 to create O2 in hypoxia-associated RT resistance. Through the combination of gold nanoparticles and O2 generation by MnO2 shells, Au@MnO2-PEG core–shell nanoparticles, there is a good tumor therapeutic effect.

![Scheme 12](image)

**Scheme 12** The preparation of SR-SERS nanoprobe.

![Scheme 13](image)

**Scheme 13** The preparation of EGaIn–Au NPs.
2.11. EGaIn–Au NPs

To give nano-composites through liquid metals (LM), a mixture of gallium indium–Au nanostructures (EGaIn–Au NPs) was used for providing radio-photothermal cancer treatment. In this process, Au nanodots were reduced onto the mixture of gallium indium (EGaIn) NPs surface to achieve EGaIn–Au NPs. This strategy might open a new door to a LM-based nano-composite. The EGaIn NPs were designed through the probe-sonicating method in xanthan gum solution after centrifuging the spherical EGaIn NPs, and HAuCl₄ solution was mixed with EGaIn NPs for growth of Au NPs by the interfacial galvanic replacement reactions to give EGaIn–Au nano-composites (Scheme 13).

It was proved that the EGaIn–Au nano-composite was used to respond the X-ray and NIR (near-infrared laser) irradiation. The nano-composite with photothermal conversion and radiosensitization ability destroy cancer cells to cure. However, healthy tissues are damaged, they used for decreasing the growth of tumor tissues by NIR, and X-ray treatment in photothermal therapy and radiotherapy.

2.12. I–Pd@Au–PEG–FA nanosheets

Pd@Au–PEG–FA nanosheets were used as CT image contrast agents. The Pd@Au nanosheets were synthesized according to previous reports. FA–NHS was obtained from folic acid (FA), N-hydroxysuccinimide (NHS), and dicyclohexylcarbodiimide (DCC) in DMSO in the dark to provide FA–NHS, which was added to the NH₂–PEG–SH to provide Pd@Au–PEG–FA nanosheets, and then it was mixed with FA–PEG–SH to provide Pd@Au–PEG–FA nanosheets. Preparation of I–Pd@Au–PEG–FA nanosheets was accomplished through the reaction of Pd@Au–PEG–FA with radioiodine at room temperature, which was used to load radioiodine by ¹²⁵I at room temperature (Schemes 14 and 15). I–Pd@Au–PEG–FA nanosheets were then applied on detecting the plaques by reforming the 2D multifunctional structure by FA on the surface and evaluate the target specificity for the activated macrophages; the targeted probes show that 2D Pd@Au nanosheets have superior pharmacokinetic to achieve the cure effect.

3. Conclusion

Au nanoparticles received much attention for cancer treatment and MRI due to their high chemical stability, biocompatibility, and affinity for binding with thiol terminal groups of various organic compounds. Moreover, these mixtures give the magnetic and plasmonic properties to nanoparticles for diagnostics and therapeutic applications. However, the currently available synthesis methods for these nanoparticles are based on organic compounds.

Conflicts of interest

The authors declare that there is no declaration of competing interest in this paper.

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