Article abstract—Objective: To analyze the MRI and CSF oligoclonal bands (OB) changes in patients with MS who underwent an autologous hematopoietic stem cell transplantation (AHSCT). Background: AHSCT is evaluated as an alternative therapy in severe MS. In previous series of AHSCT for MS, data on MRI or OB outcome were limited or not provided. Methods: Five patients with a median Kurtzke’s EDSS score of 6.5, more than two attacks, and confirmed worsening of the EDSS in the previous year received an AHSCT. Hematopoietic stem cells were mobilized with cyclophosphamide (3 g/m²) and granulocyte colony-stimulating factor (5 µg/kg/d). The graft was T cell depleted by positive CD 34+ selection. Conditioning regimen included BCNU (300 mg/m²), cyclophosphamide (150 mg/kg in 3 days), and antithymocyte globulin (60 mg/kg in 4 days). MRI scans were scheduled at baseline and 1, 3, 6, and 12 months and OB analysis at baseline and 3 and 12 months post-AHSCT. Results: Four patients had a stable or improved EDSS after a median follow-up of 18 months (range, 12 to 24 months). The fifth patient’s condition deteriorated during AHSCT. She partially improved and remained stable after month 3 after AHSCT. The baseline CSF OB persisted 1 year after AHSCT. MRI studies after AHSCT showed no enhanced T1 lesions and no new or enlarging T2 lesions. The median percentage change of T2 lesion load was −11.8% (range, −26.6 to −4.0%). All patients had a decrease of corpus callosum area at 1 year (median, 12.4%; range, 7.8% to 20.5%) that did not progress in the two patients evaluated at 2 years after AHSCT. Conclusions: Although the persistence of CSF OB suggests the lymphocytes were not eliminated from the CNS, the follow-up MRI studies showed no enhanced T1 brain lesions and a reduction in the T2 lesion load that correlated with the clinical stabilization of MS after AHSCT.

Immune ablation with autologous hematopoietic stem cell transplantation (AHSCT) is evaluated as a potential treatment for severe cases of MS and other systemic autoimmune diseases.1,2 The rationale for using AHSCT to treat MS is based on isolated MS case reports of patients who underwent this treatment for a concomitant hematologic malignancy and the positive effect of syngeneic or autologous bone marrow transplantation on the prevention or remission of experimental allergic encephalomyelitis.3 The aim of AHSCT is to produce a profound T-cell depletion and to reconstitute an immune system with a new immune tolerance. Although this objective would be better accomplished by use of an allogeneic hematopoietic stem cell transplantation, the morbidity and mortality of this type of transplantation prevents its use in patients with MS.

Few studies of AHSCT in MS have been published.4–7 They addressed the feasibility of the procedure, but no detailed information was provided on the evolution of the MRI lesions or the CSF abnormalities that would help in the understanding of the effects of the AHSCT. We describe the clinical, MRI, and CSF evolution of the first five patients with MS receiving an AHSCT at our institution, after a median follow-up of 18 months.

Methods. Patients. The patients reported are the first five included in a prospective protocol to evaluate the safety of T cell–depleted AHSCT for patients with severe MS. The treatment protocol is described in the Appendix. Eligibility criteria include: 1) age between 18 and 60 years; 2) clinically definite secondary progressive (SPMS) or relapsing remitting (RRMS) with a Kurtzke’s Expanded Disability Status Scale (EDSS) of 4.0 to 6.5; 3) an increase in the EDSS by 1.0 point with an EDSS ≥5.5 or less or 0.5 with an EDSS >5.5 over the previous year in spite of treatment with interferon or other immunotherapies that are stopped at least 1 month before the AHSCT. Patients with RRMS must have at least two relapses in the last year. Patients are excluded if there is significant history of medical illness precluding transplantation, cognitive deterioration, or severe atrophy seen in the brain MRI. The protocol is approved and monitored by the ethical and re-

From the Services of Neurology (Drs. Saiz and Graus), Bone Marrow Transplantation Unit (Drs. Carreras, Martinez, and Rovira), Hematology, Radiology (Drs. Berenguer and Pujol), Immunology (Dr. Yague), and Blood Bank (Dr. Marin), Hospital Clinic, Institut d’Investigació Biomèdica August Pi i Sunyer (IDIBAPS), University of Barcelona; and Service of Neurology (Dr. Arbizu), Unidad de Esclerosis Múltiple, C.S.U. de Bellvitge, L’Hospitalet, Spain.

Supported by grant 97/001 Fundació La Marató TV3.

Received July 25, 2000. Accepted in final form December 30, 2000.

Address correspondence and reprint requests to Dr. Francesc Graus, Service of Neurology, Hospital Clinic. Villarroel 170, 08036 Barcelona, Spain; e-mail: graus@medicina.ub.es

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search committees of the institution and the Spanish Ministry of Health.

Clinical evaluation. The neurologic status of the patients was evaluated (neurologic examination, EDSS, Ambulatory Index [AI] score) at baseline, at the end of stem cell mobilization, at months 1, 3, 6, 9, and 12 after AHSCT, and then every 6 months. Hematologic follow-up after hospital discharge and schedule of immunizations after treatment were done according to guidelines previously reported. Systemic adverse effects were scored by using the Bearman’s scale.

MRI evaluation. Brain MRI scans were obtained before the stem cell mobilization (baseline) and months 1, 3, 6, and 12 after AHSCT with a 1.5-T unit Siemens Magnetom SP (Erlangen, Germany) with a quadrature head coil. Two-dimensional gradient-echo scout images, acquired in three planes, were used for positioning slices in the MRI procedure. Sections with a thickness of 5 mm were acquired in the axial plane (parallel to the body of the corpus callosum) by using the following parameters: minimal interslice gap (<1 mm) and 192 × 256 matrix.

The MRI protocol consisted of sagittal spin-echo T1-weighted images (repetition time [TR], 608 msec; echo time [TE], 14 msec), axial spin-echo T1-weighted images (TR/TE 608/14), and sagittal fast spin-echo T2-weighted images (TR/TE 4600/90). Axial spin-echo T2-weighted images included proton density (TR/TE 2500/15) and T2 (TR/TE 2500/90). All T1-weighted images also were acquired 5 minutes after IV injection of 0.1 mmol/kg gadodiamide (Omniscan; Nycomed, Inc, Princeton, NJ).

A single experienced neuroradiologist (J.B.) visually identified and counted the number of hypointense and enhanced lesions in precontrast and postcontrast T1-weighted images and hyperintense lesions in T2-weighted images. The measurement of lesion volume by using T2 sequences of the baseline and 1 year post-AHSCT MRI was calculated with a workstation Sienet DRC 104 (Siemens, Erlangen, Germany). Lesion volume calculations were obtained by using manual tracking, based on the lesional volume obtained by multiplying the total hyperintense lesion area by the thickness of the section and summation of the value of each section. The atrophy measures were based on the precontrast axial (lateral ventricle and brain widths) and sagittal (corpus callosum area) T1-weighted images and calculated in the workstation monitor. The mean coefficients of variation, obtained as described, were 1.5%, 0.3%, and 5% for ventricle width, brain width, and corpus callosum area. All studies were done blind to the clinical evolution of the patients.

CSF analysis. Paired serum/CSF samples were obtained at baseline and 3 and 12 months after AHSCT and stored at −80 °C. Integrity of the blood–brain barrier (BBB) was estimated by the albumin index (upper normal value, 9.0). The identification of IgG-specific oligoclonal bands (OB) in CSF and serum was performed with an IgG-IIF Kit (Helena BioScience, Sunderland, UK) according to manufacturer's instructions. Basically, paired serum/CSF samples were run in an agarose gel isoelectric focusing, pH 3–10. The separated proteins were transferred to a nitrocellulose membrane, immunofixed with sheep anti-human IgG peroxidase conjugated, and developed with 3-amino-9 ethylcarbazole (Sigma, St. Louis, MO). The patterns were interpreted qualitatively by comparing the presence or absence of oligoclonal bands in CSF or serum.

Hematologic and immune evaluation. Peripheral blood was examined at baseline and months 3, 9, and 12 after transplantation, and then every 6 months. T cells (CD3+), helper/inducer T cells (CD3+CD4+), naive (CD4+CD45RA+) and memory (CD4+CD45RO) helper/inducer T cells, suppressor/cytotoxic T cells (CD3+CD8+), and B cells (CD19+) were analyzed on a FACSCan (Becton Dickinson Immunocytometry Systems [BDIS], San Jose, CA).

Results. Baseline patient characteristics. The five patients had a mean disease duration of 9 years (range, 6 to 14 years). Two patients had RRMS and three had SPMS. The median scores of EDSS was 6.5 (range, 5.0 to 6.5), and that of AI, 4 (range, 2 to 6). Both patients with RRMS had six relapses the previous year despite treatment with interferon β and repeated boluses of IV methylprednisolone (table 1).

Toxicity. There were no major systemic complications (table 1). Patient 5 presented an exacerbation of her leg weakness while receiving granulocyte colony-stimulating factor (G-CSF), which returned to baseline in a few days. Patient 2 developed a severe paraparesis concomitant with high fever associated with the administration of antithymocyte globulin (ATG), and her EDSS score increased from 6.5 to 8.0. She slowly improved to an EDSS of 7.5 over the ensuing 3 months. Four patients had uncomplicated urinary tract infections during the first 3 months after hospital discharge. Menstruation did not return in the oldest woman.

Neurologic outcome. At the last evaluation (median follow-up, 18 months; range, 12 to 24 months), the EDSS improved in Patients 4 (from 6.5 to 5.0) and 5 (from 6.5 to 5.5), was unchanged in Patients 1 and 3, and remained stable after month 3 after AHSCT in Patient 2, whose condition worsened during the transplantation. Patient 3 had two relapses and Patient 4 one after AHSCT. All the episodes were subjective sensory symptoms that lasted a few days and did not require treatment. The neurologic examination during the relapse did not disclose any new sign, but the distribution of the sensory symptoms suggested an involvement of the spinal cord. Fatigue (although not formally measured) and urinary symptoms (once urinary infections were controlled) improved along the follow-up period in the five patients. None of the patients needed additional immunotherapy after AHSCT.

MRI evaluation. The baseline MRI characteristics are shown in table 2. Enhanced T1 lesions were presented in Patients 1, 3, and 5 and disappeared in the MRI done 1 month after AHSCT. The follow-up MRI studies did not show enhanced T1 lesions and no new hypointense T1 lesions, except in one patient who had a new hypointense lesion that corresponded to an enhancing lesion at baseline MRI. No new or enlarging hyperintense lesions on T2-weighted images were observed at the follow-up MRI studies in the five patients. The median percentage change of T2 lesion volume was −11.8% (range, −26.6 to −4.0%) (table 2).

A decrease in the corpus callosum area was observed in the five patients at 1 year (median decrease, 12.4%; range, 7.8% to 20.5%). The atrophy was independently confirmed by another neuroradiologist, with interobserver median
variation in the percentage of the atrophy of the corpus callosum of 1.2% (range, 0.1% to 3.9%). More than 50% of the reduction of the corpus callosum area occurred in the first 3 months after AHSCT in all of the patients. The percentage of loss during this period was 82% in Patient 2 and 80% in Patient 5. The MRI 2 years after AHSCT done in Patients 1 and 2 showed no new enhanced or hypointense T1 lesions, a further reduction of the T2 lesion load, and the arrest in the decrease of the corpus callosum area seen during the first year after transplantation.

**Immune reconstitution after transplantation.** All patients had a total number of peripheral blood lymphocytes >500/μL and normal values of CD8+ T cells at 3 months after transplantation. CD4+ T cells achieved cell counts higher than 200/μL at 12 months after transplant in four of five patients. Throughout the period of the study, almost all CD4+ lymphocytes expressed CD45RO antigen (memory T cells), and only low numbers of cells expressed CD45RA antigen (naive T cells). All of the patients had normal values of B-lymphocytes at 9 months after treatment.

**Table 1 Baseline clinical characteristics, number of reinfused cells, hematologic recovery, and toxicity of the treatment**

| Characteristics                  | Patient 1          | Patient 2          | Patient 3          | Patient 4          | Patient 5          |
|----------------------------------|--------------------|--------------------|--------------------|--------------------|--------------------|
| Age, y/sex                        | 30/F               | 43/F               | 24/F               | 44/M               | 27/F               |
| MS type                           | SP                 | SP                 | RR                 | SP                 | RR                 |
| Baseline evaluation               |                    |                    |                    |                    |                    |
| EDSS                             | 6.0                | 6.5                | 5.0                | 6.5                | 6.5                |
| Ambulation index                  | 4                  | 6                  | 2                  | 4                  | 4                  |
| Number of relapses                | 3                  | 2                  | 6                  | 2                  | 6                  |
| Number of reinfused cells         |                    |                    |                    |                    |                    |
| CD34+ (× 10⁶/kg)                  | 7.6                | 3.4                | 4.7                | 2.5                | 4.8                |
| CD3+ (× 10⁴/kg)                   | 30                 | 7                  | 0.27               | 0.64               | 0.29               |
| Engraftment, d                    |                    |                    |                    |                    |                    |
| Neutrophils >500/μL               | +9                 | +11                | +11                | +12                | +9                 |
| Platelets >20,000/μL              | +11                | +14                | +12                | +15                | +12                |
| Toxicity                          |                    |                    |                    |                    |                    |
| Mucositis*                        | Yes                | Yes                | Yes                | Yes                | Yes                |
| Fever                             | Yes                | Yes                | Yes                | Yes                | Yes                |
| Documented infection              |                    | S aureus           | —                  | S epidermidis      | S epidermidis      |
| —                                |                    | E coli             | —                  |                    |                    |
| Neurologic deterioration          | No                 | Yes‡               | No                 | No                 | Yes§               |
| Other toxicities (grade)          | No                 | No                 | No                 | No                 | Hepatic (II)¶      |
| Days in hospital                  | 18                 | 29                 | 35                 | 21                 | 46                 |

* Grade I = mucositis not requiring continuous IV analgesia.
† Asymptomatic reinfection.
‡ During conditioning, incomplete recovery.
§ During mobilization, complete recovery.
¶ Grade II = AST/ALT >5 times of normal values.

SP = secondary progressive MS; RR = relapsing-remitting MS; EDSS = Expanded Disability Status Scale score.

**Discussion.** AHSCT is an experimental treatment option for severe forms of MS. The efficacy of AHSCT remains to be demonstrated, and this treatment should be considered only in the setting of approved protocols. This study confirms previous work that patients with severe MS tolerate AHSCT with acceptable toxicity4-7 and shows for the first time three features that may be relevant for future studies. First, IgG OB in the CSF did not disappear after AHSCT; second, follow-up MRI showed a decrease in T2-weighted lesion load that persisted in absence of additional immunotherapy; and third, there was a decrease in corpus callosum area in spite of an ab-
sence of inflammatory disease activity during the first year after AHSCT.

AHSCT was not associated with severe systemic toxicity. However, one of the patients presented a severe neurologic deterioration during the treatment that did not recover to the baseline level. Transient neurologic worsening is described in up to 42% of patients during the procedure, but our patient emphasizes that this complication may be only partially reversible. Another patient had a transient neurologic deterioration while receiving granulocyte colony-stimulating factor (G-CSF) for stem cell mobilization. This patient had multiple relapses in the year previous to the transplantation and the possibility of a coincidence with the G-CSF treatment cannot be excluded. The mechanism of this complication is uncertain but has been reported, particularly when G-CSF alone is used to mobilize stem cell to the periphery. The current protocol uses half of the recommended dose of G-CSF with a priming dose of cyclophosphamide that may counteract possible immunostimulatory effects of G-CSF. We have not observed this complication in the subsequent six patients who have been treated with this protocol (unpublished).

The five patients’ conditions remained neurologically stable or improved after discharge from the hospital. The absence of T1-enhancing lesions and the lack of new or enlarging hyperintense T2 lesions in the follow-up MRI suggests that AHSCT had a positive impact on active inflammation and seems to agree with the clinical stabilization of our patients in the first year after AHSCT. However, these MRI features are not a good predictor of long-term disability, and only the follow-up will unambiguously show whether the treatment has been able to modify the clinical course of these patients.

All patients showed an increase in the atrophy of the corpus callosum in spite of improvement of the other MRI variables. The relationship between inflammatory activity and the development of brain atrophy is unclear. In longitudinal studies of brain

### Table 2 MRI outcomes after autologous hematopoietic stem cell transplantation

| Outcomes                                    | Patient 1 | Patient 2 | Patient 3 | Patient 4 | Patient 5 |
|----------------------------------------------|-----------|-----------|-----------|-----------|-----------|
| Baseline Mean T2 lesion volume, cm³         | 19.32     | 31.48     | 37.57     | 2.89      | 3.08      |
| Enhanced T1 lesions                         | 12        | 0         | 5         | 0         | 6         |
| Hypointense T1 lesions⁹                     | 22        | 18        | 22        | 5         | 2         |
| Atrophy variables                           |           |           |           |           |           |
| Lateral ventricle width, mm                  | 29        | 24        | 24        | 33        | 16        |
| Brain width, mm                              | 86        | 92        | 87        | 94        | 95        |
| Corpus callosum area, mm²                    | 685       | 523       | 461       | 594       | 515       |
| 1 Year after transplantation                 |           |           |           |           |           |
| Percentage change of mean lesional volume from baseline | -19.7     | -26.6     | -10.3     | -4.0      | -11.8     |
| Enhanced T1 lesions                         | 0         | 0         | 0         | 0         | 0         |
| Hypointense T1 lesions                       | 22        | 18        | 22        | 5         | 3         |
| Percentage change of atrophy from baseline   |           |           |           |           |           |
| Lateral ventricle width                       | 0         | 0         | +17.2     | +13.5     | 0         |
| Brain width                                  | -4.6      | 0         | -3.4      | 0         | 0         |
| Corpus callosum area                         | -7.8      | -14.9     | -11.7     | -12.4     | -20.5     |

*All lesions less than 1.3 mm.*

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Figure. Identification of immunoglobulin G–specific oligoclonal bands (OB) from Patient 5. The baseline CSF had at least three OB (open triangles). These bands persisted at month 3 after transplantation, and they were also seen in the serum but with a weaker intensity. At month 12, the three bands were only observed in the CSF. Serum at month 3 showed multiple OB (closed triangles) that were also present in the CSF. Some of these bands were still present in the serum and CSF 12 months after treatment. SO/CO = Baseline serum/CSF; S3/C3 = serum/CSF 3 months after transplantation; S12/C12 = serum/CSF 12 months after transplantation.
atrophy in untreated patients with RRMS, the decrease in corpus callosum area appears to be influenced by the baseline number of gadolinium-enhancing lesions. However, similar studies that used volumetric techniques failed to show a correlation of brain atrophy with enhanced lesion load.

Taken these data together, one cannot rule out that the AH SCT could be effective in arresting the inflammatory activity but fail to reverse the pathologic process responsible for the brain atrophy. A second explanation is that the atrophy could be caused, at least in part, by the AH SCT. More than 50% of the atrophy occurred in the 3 months after AH SCT, particularly in the two patients who had a more complicated AH SCT. This possibility would be further supported by the observation that the atrophy of the corpus callosum did not increase during the second year after the treatment. Brain atrophy was observed in patients undergoing autologous bone marrow transplantation for chronic myeloid leukemia, but specific measurements of the corpus callosum area were not provided. If the brain atrophy found in our patients is related to the treatment, this effect could be more evident in transplantation protocols that include total body radiation because of the potential synergistic effect of chemotherapy and radiation therapy in causing neurotoxicity.

The CSF analysis 3 months after AH SCT showed a disruption of the BBB and the presence of many OB that were also identified in the serum. Serum OB was reported in up to 87% of patients who had an AH SCT for multiple myeloma and persisted 7 months on the average. The significance of these serum OB is unclear, but they could reflect recapitulation of early B-cell ontogeny. Persistence of CSF OB at 1 year after AH SCT was previously observed in two patients who were treated with busulfan, cyclophosphamide, and antithymocyte globulin. The serial analysis of the CSF OB at 3 and 12 months after AH SCT in our patients supports the idea that the B cells responsible for the IgG synthesis in the CNS survived the conditioning regimen. Unlike other CSF OB detected at 3 months after AH SCT, the OB persisted 1 year after AH SCT in our patients supports the idea that the atrophy of the corpus callosum could be caused, at least in part, by the AH SCT. More than 50% of the atrophy occurred in the 3 months after AH SCT, particularly in the two patients who had a more complicated AH SCT. This possibility would be further supported by the observation that the atrophy of the corpus callosum did not increase during the second year after the treatment. Brain atrophy was observed in patients undergoing autologous bone marrow transplantation for chronic myeloid leukemia, but specific measurements of the corpus callosum area were not provided. If the brain atrophy found in our patients is related to the treatment, this effect could be more evident in transplantation protocols that include total body radiation because of the potential synergistic effect of chemotherapy and radiation therapy in causing neurotoxicity.

The clinical impact of the presumable persistence of the lymphocytes in the CNS is unclear because the MRIs did not show features compatible with disease activity, and the EDSS did not increase after hospital discharge. However, the evolution of CSF OB and the brain atrophy should be recorded in transplantation protocols to better understand the effects of AH SCT in MS.

Appendix

Treatment protocol

**Mobilization of hematopoietic stem cells:** Cyclophosphamide 3 g/m² + granulocyte colony-stimulating factor (G-CSF) (5 µg/kg/d)

**Peripheral blood stem-cell collection:** Fenwal CS3000 (Baxter)

**Leukapheresis:** One T-cell–depleted + one non–T-cell–depleted (backup) (target in both: 3 × 10⁶ CD34⁺/kg)

**Positive CD34⁺ selection:** Isolex 300 (Baxter) (Patients 1, 2)

**ClimiMACS (Miltenyi) (Patients 3–5)**

**Conditioning regimen:**

- Day -6—BCNU (300 mg/m²)
- Day -5—Antithymocyte globulin (Merieux) (ATG) (15 mg/kg), cyclophosphamide (50 mg/kg)
- Day -4—ATG (15 mg/kg), cyclophosphamide (50 mg/kg)
- Day -3—ATG (15 mg/kg), cyclophosphamide (50 mg/kg)
- Day -2—ATG (15 mg/kg)
- Day -1—Rest

**Day 0—Hematopoietic stem cell transplantation**

**Supportive care:** Lamian airfow room; low microbial diet; oral ciprofloxacin, fluconazole, and acyclovir; inhaled pentamidine; IV Ig, G-CSF (day +1), and ganciclovir if positive cytomegalovirus antigenemia

**Acknowledgment**

The authors thank Dr. B. Casanova, Hospital La Fe, Valencia, for referring Patient 4.

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The CSF analysis 3 months after AH SCT showed a disruption of the BBB and the presence of many OB that were also identified in the serum. Serum OB was reported in up to 87% of patients who had an AH SCT for multiple myeloma and persisted 7 months on the average. The significance of these serum OB is unclear, but they could reflect recapitulation of early B-cell ontogeny. Persistence of CSF OB at 1 year after AH SCT was previously observed in two patients who were treated with busulfan, cyclophosphamide, and antithymocyte globulin. The serial analysis of the CSF OB at 3 and 12 months after AH SCT in our patients supports the idea that the B cells responsible for the IgG synthesis in the CNS survived the conditioning regimen. Unlike other CSF OB detected at 3 months after AH SCT, the OB showed a stronger intensity in the CSF than in the serum. This observation suggests the B cells responsible for its synthesis were in the CNS, and the OB diffused to the serum because of the disruption of the BBB.

Unlike other published AH SCT protocols, we did not include in the conditioning regimen total body irradiation or added other drugs, as melphalan, etoposide, or cytosine arabinoside that in theory could be more effective to eradicate the lymphocytes in the CNS. We decided not to use these approaches for the concern of an increased incidence of cancer later in life. In addition, there is clinical and experimental evidence of neurologic deterioration after use of brain irradiation on demyelinating lesions.

The clinical impact of the presumable persistence of the lymphocytes in the CNS is unclear because the MRIs did not show features compatible with disease activity, and the EDSS did not increase after hospital discharge. However, the evolution of CSF OB and the brain atrophy should be recorded in transplantation protocols to better understand the effects of AH SCT in MS.

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- Day -2—ATG (15 mg/kg)
- Day -1—Rest

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The influence of cognitive impairment on driving performance in multiple sclerosis

Maria T. Schultheis, PhD; Edward Garay, BS; and John DeLuca, PhD

Article abstract—Objective: To examine the influence of impaired cognitive processing on measures of driving skills in persons with MS. Methods: Twenty-eight subjects with documented MS were divided into two groups—with [MS(+)], n = 13 and without [MS(−)], n = 15) cognitive impairment—based on neuropsychological performance. Healthy control (HC) subjects (n = 17) matched on age and driving experience were also studied. Driving-related skills were compared between the groups based on performance on two computerized driving tests: the Useful Field of Vision (UFOV) and the Neurocognitive Driving Test (NDT). Results: The MS(+) group performed significantly worse than both the MS(−) and HC groups in the latency to perform several driving-specific functions on the NDT, but no overall group differences were observed in actual errors on the NDT. On the UFOV, when compared to MS(−) and HC subjects, the MS(+) group demonstrated poorer performance on two of the three subtests. Additionally, a significantly higher percentage of MS(+) individuals were rated within the high risk (probability of crash involvement) category, relative to the MS(−) and HC groups. Conclusions: Cognitive impairment can negatively affect driving-related skills in persons with MS and should be considered in the determination of driving ability.

NEUROLOGY 2001;56:1089–1094

The impact of cognitive impairment on driving skills and abilities has been documented in various neurologic populations, including brain injury, stroke, and dementia. These studies have identified decreased attentional and visual perceptual skills, slowed information processing speed, and executive dysfunction as related to impaired driving skills and abilities. Despite recent evidence indicating the

From the Neuropsychology and Neuroscience Laboratory (Dr. Schultheis, E. Garay, and Dr. DeLuca), Kessler Medical Rehabilitation Research and Education Corporation, West Orange, NJ; and Departments of Physical Medicine and Rehabilitation (Drs. Schultheis and DeLuca) and Neuroscience (Dr. DeLuca), University of Medicine and Dentistry of New Jersey–New Jersey Medical School, Newark. Supported by grant number PP9886 from the National Multiple Sclerosis Society.

Received July 19, 2000. Accepted in final form January 9, 2001.

Address correspondence and reprint requests to Dr. Maria T. Schultheis, Neuropsychology and Neuroscience Laboratory, Kessler Medical Rehabilitation Research & Education Corporation, 1199 Pleasant Valley Way, West Orange, NJ 07052; e-mail: mschultheis@kmrrec.org

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