Effect of one-lung ventilation on end-tidal carbon dioxide during cardiopulmonary resuscitation in a pig model of cardiac arrest

Dong Hyun Ryu, Yong Hun Jung, Kyung Woon Jeung, Byung Kook Lee, Young Won Jeong, Jong Geun Yun, Dong Hun Lee, Sung Min Lee, Tag Heo, Yong Il Min

1 Department of Emergency Medicine, Chonnam National University Hospital, Gwangju, Republic of Korea, 2 Department of Emergency Medical Services, Honam University, Gwangju, Republic of Korea

☯ These authors contributed equally to this work.

* neoneti@hanmail.net

Abstract

Unrecognized endobronchial intubation frequently occurs after emergency intubation. However, no study has evaluated the effect of one-lung ventilation on end-tidal carbon dioxide (ETCO₂) during cardiopulmonary resuscitation (CPR). We compared the hemodynamic parameters, blood gases, and ETCO₂ during one-lung ventilation with those during conventional two-lung ventilation in a pig model of CPR, to determine the effect of the former on ETCO₂. A randomized crossover study was conducted in 12 pigs intubated with double-lumen endobronchial tube to achieve lung separation. During CPR, the animals underwent three 5-min ventilation trials based on a randomized crossover design: left-lung, right-lung, or two-lung ventilation. Arterial blood gases were measured at the end of each ventilation trial. Ventilation was provided using the same tidal volume throughout the ventilation trials. Comparison using generalized linear mixed model revealed no significant group effects with respect to aortic pressure, coronary perfusion pressure, and carotid blood flow; however, significant group effect in terms of ETCO₂ was found (P < 0.001). In the post hoc analyses, ETCO₂ was lower during the right-lung ventilation than during the two-lung (P = 0.006) or left-lung ventilation (P < 0.001). However, no difference in ETCO₂ was detected between the left-lung and two-lung ventilations. The partial pressure of arterial carbon dioxide (PaCO₂), partial pressure of arterial oxygen (PaO₂), and oxygen saturation (SaO₂) differed among the three types of ventilation (P = 0.003, P = 0.001, and P = 0.001, respectively). The post hoc analyses revealed a higher PaCO₂, lower PaO₂, and lower SaO₂ during right-lung ventilation than during two-lung or left-lung ventilation. However, the levels of these blood gases did not differ between the left-lung and two-lung ventilations. In a pig model of CPR, ETCO₂ was significantly lower during right-lung ventilation than during two-lung ventilation. However, interestingly, ETCO₂ during left-lung ventilation was comparable to that during two-lung ventilation.
Introduction

Unrecognized endobronchial intubation frequently occurs following emergency intubation [1–3]. Multiple studies have suggested that clinical assessments, including chest auscultation, are unreliable in detecting endobronchial intubation [1,4,5]. Chest radiograph remains one of the definitive means to detect endobronchial intubation. However, this modality is not generally used during cardiopulmonary resuscitation (CPR) because obtaining chest radiograph inevitably requires chest compression interruption. Thus, inadvertent endobronchial intubation is likely to remain undetected during CPR.

End-tidal carbon dioxide (ETCO₂) is a non-invasive measure of pulmonary perfusion during cardiac arrest. It has been shown to reflect changes in cardiac output during CPR and predict resuscitation outcomes, making it a useful guide for resuscitation efforts [6–12]. Currently, resuscitation guidelines recommend the use of physiological parameters, such as ETCO₂, during CPR [13,14]. Under normal physiological conditions, alveolar ventilation is the principal determinant of alveolar carbon dioxide concentration in the lungs. The partial pressure of carbon dioxide within the alveoli is inversely related to the amount of alveolar ventilation. In the case of unrecognized endobronchial intubation, the total tidal volume is delivered to the intubated lung, resulting in doubling of ventilation/perfusion ratio in the intubated lung. Thus, theoretically, ventilation through an endotracheal tube inadvertently placed into the main stem bronchus would cause a decrease and an increase in alveolar carbon dioxide in the intubated and contralateral lungs, respectively. ETCO₂ would decrease following endobronchial intubation because capnography measures the partial pressure of carbon dioxide only in the intubated lung. Several studies performed in a non-arrest animal model also reported a decrease in ETCO₂ following one-lung ventilation achieved through endobronchial intubation or bronchial occlusion [15,16]. Meanwhile, the relationship between ETCO₂ and alveolar ventilation becomes more complicated during CPR because chest compressions, as well as ventilations, influence the pulmonary gas exchange [17]. Thus, the effect of one-lung ventilation on ETCO₂ during CPR may differ from that during the normal cardiac output state. However, to our knowledge, no study has evaluated the effect of one-lung ventilation on ETCO₂ during CPR.

In this study, we compared the hemodynamic parameters, blood gases, and ETCO₂ during one-lung ventilation with those during conventional two-lung ventilation in a pig model of CPR, to determine the effect of the former on ETCO₂ during CPR. We hypothesized that ETCO₂ would decrease during one-lung ventilation compared with two-lung ventilation.

Materials and methods

This prospective, randomized, crossover study was conducted in 12 Yorkshire/Landrace cross pigs weighing 22.8 ± 3.2 kg, which were intubated with double-lumen endobronchial tube to achieve lung separation. Additionally, ETCO₂ was assessed in one animal after tracheal and endobronchial intubations using a standard endotracheal tube. Thus, 13 pigs were used in this study. The Animal Care and Use Committee of Chonnam National University approved the protocol (CNU IACUC-H-2017-1). Animal care and experiments were conducted based on the author’s Institutional Animal Care and Use Committee Guidelines.

Animal preparation

Following the administration of premedications (20 mg/kg ketamine and 2.2 mg/kg xylazine intramuscularly), the animals were placed in a supine position in a U-shaped trough and were orally intubated. Anesthesia was provided using 70%:30% N₂O:O₂ and 0.5–2% sevoflurane, which was titrated to prevent signs of pain (reactive wide pupils, tachycardia, and
hypertension). A double-lumen catheter was inserted via the right femoral artery for blood pressure monitoring and blood sampling. The right external jugular vein was cannulated with an 8 F introducer sheath to monitor the right atrial (RA) pressure and insert a right ventricle (RV) pacing catheter. The right common carotid artery was surgically exposed, and an ultrasonic flow probe (Transonic Inc., Ithaca, NY, USA) was placed around it for carotid blood flow (CBF) measurement. An ETCO₂ sample line (B40 Patient Monitor; GE Healthcare, Chalfont St Giles, UK) was connected to the ventilator circuit. In large pigs, the tip of the orally intubated tracheal tube does not reach the carina although the tube’s rostral end is positioned deep into the oral cavity. Furthermore, in our preliminary experience, placement of a tracheal tube into the desired main stem bronchus was extremely difficult during CPR because of the tracheal compression caused by chest compression and presence of oedema in the airways, which resulted in CPR interruptions for up to several minutes. For these reasons, a 35 F double-lumen endobronchial tube (Broncho-Cath™, Covidien, Mansfield, MA, USA) was placed under bronchoscopic guidance (Ambu® aScope™; Ambu A/S, Ballerup, Denmark) after tracheal tube removal, with the tip of the longer bronchial lumen placed in the left main stem bronchus while the tip of the shorter tracheal lumen remained in the trachea, to achieve lung separation during CPR. Immediately after the insertion, a fiberoptic bronchoscope was introduced into the tracheal lumen. While observing the carina, the tube position was adjusted until the bronchial cuff was just below the carina, and both the bronchial and tracheal cuffs were inflated to seal the left bronchus and trachea, respectively. We chose to place the bronchial lumen in the left main stem bronchus because the right upper lobe bronchus arose from the trachea immediately above the carina in our pigs. Throughout the rest of the preparation period, both lungs were ventilated using a Y-adapter for the proximal ends. After a 20 min stabilization period, baseline measurements were obtained, and vecuronium (0.05 mg/kg) was administered intravenously to inhibit the potential confounding effect of gasping.

Experimental protocol

Ventricular fibrillation (VF) was induced by applying an electrical current (60 Hz, 30 mA alternating current) via an RV pacing catheter. After 5 min of untreated VF, the animals underwent a 2 min CPR period to wash out the accumulated carbon dioxide in the trachea while adjusting the chest compression depth. Chest compressions were delivered at a rate of 100 /min using a piston-driven chest compression device (Life-Stat; Michigan Instruments, Grand Rapids, MI, USA). The compression depth was adjusted to decrease the anterior–posterior diameter of the chest by 20%. Following the 2 min CPR period, the animals underwent three 5 min ventilation trials during CPR, with each animal receiving the following three types of ventilation based on a randomized crossover design: left-lung, right-lung, or two-lung ventilation (which refers to ventilation through the bronchial lumen, tracheal lumen, or both lumens using a Y-adapter, respectively) (Fig 1). The order of the ventilation types was counterbalanced and randomized using closed envelope method. Throughout this period, asynchronous positive-pressure ventilations with high-flow oxygen (14 l/min) were provided with a tidal volume of 10 ml/kg and rate of 10 /min using a volume-marked bag devised by Cho et al [18]. A tidal volume identical to that used for two-lung ventilation was utilized during right- and left-lung ventilations to simulate one-lung ventilation in unrecognized endobronchial intubation. During the right- or left-lung ventilation, the non-ventilated lumen was left open to the atmosphere. The investigator ventilating the animal was blinded to the ETCO₂ level, but not to the ventilation type. The animals were not resuscitated after completion of the experiment, and thus an additional euthanasia procedure was not required in our study. Autopsy
was routinely performed to ensure that each lung was adequately ventilated during one-lung ventilation.

**Measurements**

The primary outcome was ETCO₂ values, which were determined every 30 s by averaging the ETCO₂ values for the preceding 30 s interval. Aortic and RA pressures were continuously monitored (CS/3 CCM; Datex-Ohmeda, Helsinki, Finland), and the data were transferred to a personal computer using S/5 Collect (Datex-Ohmeda, Helsinki, Finland). Coronary perfusion pressure (CPP) was calculated by subtracting the end-diastolic RA pressure from the simultaneous end-diastolic aortic pressure. Aortic systolic pressure, aortic diastolic pressure, and CPP were sampled at 30 s intervals by averaging pressures from five consecutive compressions. CBF was continuously monitored, and its value was sampled every 30 s. Arterial blood gases (Rapidlab 865; Bayer Health Care, Fernwald, Germany) were measured at the pre-arrest baseline and end of each ventilation type.

**Statistical analysis**

Sample size was calculated based on the ETCO₂ data (mean ± standard deviation [SD], 14.13 ± 5.70 mmHg; variance, 32.50) from a pilot study, where the ETCO₂ values were 17.03 ± 2.77, 18.29 ± 3.12, and 7.08 ± 2.06 mmHg for two-lung, left-lung, and right-lung ventilations, respectively, and the calculated within-group variance was 25.17. We calculated that nine animals would be required to achieve a power of 80% at an α of 0.05. Considering that each animal received all three interventions in a randomized counterbalanced order based on the study design, 12 animals were used for this study. Normally distributed variables were presented as mean ± SD, and a repeated-measure analysis of variance was performed. In contrast, non-normally distributed variables were presented as medians with interquartile ranges, and a Friedman test was conducted. Generalized linear mixed model was used to compare aortic and RA pressures, CPP, CBF, and ETCO₂ during CPR. Pairwise comparison with Bonferroni adjustment was performed for post-hoc analysis. A P value of < 0.05 was considered significant.

**ETCO₂ observation in one animal undergoing one- and two-lung ventilations using a standard endotracheal tube**

The preparation and VF induction procedures were identical to those described above, except that tracheostomy was performed and a 6.5 mm internal diameter endotracheal tube (Hi-Lo; Mallinckrodt Medical, Athlone, Ireland) was placed into the tracheostomy stoma, instead of the double-lumen endobronchial tube. During 5 min of untreated VF, the endotracheal tube was advanced into the right main stem bronchus under bronchoscopic guidance. After 5 min
of untreated VF, CPR was performed as described above, during which ventilation was provided through the endotracheal tube (right-lung ventilation). Subsequently, 5 min after the start of CPR, the endotracheal tube was pulled back until the cuff was just below the tracheostomy stoma so that both lungs were ventilated. Five minutes thereafter, the endotracheal tube was advanced into the left main stem bronchus while withholding CPR, and then chest compressions and ventilations (left-lung ventilation) were resumed. Five minutes thereafter, the endotracheal tube was pulled back again so that both lungs were ventilated. Five minutes thereafter, the endotracheal tube was advanced again into the right main stem bronchus while withholding CPR, and then chest compressions and ventilations (right-lung ventilation) were provided for 5 min.

Results

Table 1 shows the pre-arrest baseline measurements. Fig 2 displays the aortic and RA pressures, CPP, CBF, and ETCO₂ during each ventilation type. Comparison using generalized linear mixed model revealed no significant group effects with respect to aortic and RA pressures, CPP, and CBF; however, significant group effect in terms of ETCO₂ was noted (P < 0.001). In the post hoc analyses, a significant difference in ETCO₂ was found between the right-lung and two-lung ventilations (P = 0.006) and between the left-lung and right-lung ventilations (P < 0.001). However, no difference in ETCO₂ was noted between the left-lung and two-lung ventilations (P = 0.288). Table 2 shows the arterial blood gases obtained at the end of each ventilation type. A significant difference in the partial pressure of arterial carbon dioxide (PaCO₂), partial pressure of arterial oxygen (PaO₂), and oxygen saturation (SaO₂) was observed between the three types of ventilation (P = 0.003, P = 0.001, and P = 0.001, respectively). The post hoc analyses revealed a higher PaCO₂, lower PaO₂, and lower SaO₂ during right-lung ventilation than during two-lung or left-lung ventilation. However, the post hoc analyses also showed that the levels of these blood gases did not differ between the left-lung and two-lung ventilations. A significant difference in the PaCO₂-ETCO₂ gradient was observed between the three types of ventilation (P = 0.020). The post hoc analyses showed a higher PaCO₂-ETCO₂ gradient during right-lung ventilation than during two-lung or left-lung ventilation. However, the PaCO₂-ETCO₂ gradient did not differ between the left-lung and two-lung ventilations. Fig 3 displays the ETCO₂ values of one animal undergoing two-lung, left-lung, and right-lung ventilations via a standard endotracheal tube during CPR.

Table 1. Pre-arrest baseline measurements.

| Parameter                              | Value            |
|----------------------------------------|------------------|
| Aortic systolic pressure (mmHg)        | 129.0 (113.8–136.0) |
| Aortic diastolic pressure (mmHg)       | 86.1 ± 13.8      |
| Right atrial systolic pressure (mmHg)  | 11.5 (9.5–14.3)  |
| Right atrial diastolic pressure (mmHg) | 7.0 (4.5–9.8)    |
| Heart rate (/min)                      | 98.1 ± 15.0      |
| pH                                     | 7.450 ± 0.061    |
| PaCO₂ (mmHg)                           | 42.1 ± 6.0       |
| PaO₂ (mmHg)                            | 102.3 ± 28.0     |
| HCO₃⁻ (mmol/l)                         | 28.6 ± 2.9       |
| SaO₂ (%)                               | 97.5 (93.1–98.8) |
| End-tidal carbon dioxide (mmHg)        | 36.0 ± 3.7       |
| Carotid blood flow (ml/min)            | 262.3 ± 77.4     |

https://doi.org/10.1371/journal.pone.0195826.t001
Fig 2. Hemodynamic parameters during two-lung, left-lung, and right-lung ventilations. (A) Aortic systolic pressure, (B) aortic diastolic pressure, (C) right atrial systolic pressure, (D) right atrial diastolic pressure, (E) coronary perfusion pressure, (F) carotid blood flow, (G) end-tidal carbon dioxide. RA, right atrial.

https://doi.org/10.1371/journal.pone.0195826.g002
The present study evaluated the effect of one-lung ventilation occurring in the case of unrecognized endobronchial intubation in a pig model of CPR, and the results showed that ETCO$_2$ and PaO$_2$ were significantly lower but PaCO$_2$ was significantly higher during right-lung ventilation than during two-lung ventilation. However, interestingly, ETCO$_2$ and arterial blood gas values during left-lung ventilation were comparable to those during two-lung ventilation. A consistent finding was also observed when one-lung and two-lung ventilations were simulated using a standard endotracheal tube.

### Table 2. Arterial blood gases and PaCO$_2$-ETCO$_2$ gradient obtained at the end of each ventilation type.

|                          | Two-lung ventilation (n = 12) | Left-lung ventilation (n = 12) | Right-lung ventilation (n = 12) | P   |
|--------------------------|-----------------------------|-------------------------------|-------------------------------|-----|
| pH                       | 7.162 ± 0.118               | 7.157 ± 0.117                 | 7.146 ± 0.084                 | 0.749|
| PaCO$_2$ (mmHg)          | 64.7 ± 14.1$^a$             | 64.8 ± 12.5$^a$               | 73.6 ± 14.0$^b$               | 0.003|
| PaO$_2$ (mmHg)           | 59.5 (53.1–75.8)$^a$        | 70.0 (55.9–92.5)$^a$          | 41.5 (38.7–48.1)$^b$          | 0.001|
| HCO$_3^-$ (mmol/l)       | 22.4 ± 4.3                  | 22.2 ± 3.9                    | 24.5 ± 3.8                    | 0.087|
| SaO$_2$(%)               | 76.0 (65.6–85.1)$^a$        | 81.3 (60.0–93.6)$^a$          | 42.3 (39.3–55.0)$^b$          | 0.001|
| PaCO$_2$-ETCO$_2$ gradient | 42.8 ± 12.4$^a$             | 39.0 ± 12.8$^a$               | 57.8 ± 12.4$^b$               | 0.020|

Data are presented as mean ± standard deviation or medians with interquartile ranges. Superscripts a and b represent pairwise post hoc analyses. For a particular variable, values with different superscripts are significantly different, whereas those with common superscripts are not significantly different from each other.

https://doi.org/10.1371/journal.pone.0195826.t002

### Discussion

The present study evaluated the effect of one-lung ventilation occurring in the case of unrecognized endobronchial intubation in a pig model of CPR, and the results showed that ETCO$_2$ and PaO$_2$ were significantly lower but PaCO$_2$ was significantly higher during right-lung ventilation than during two-lung ventilation. However, interestingly, ETCO$_2$ and arterial blood gas values during left-lung ventilation were comparable to those during two-lung ventilation. A consistent finding was also observed when one-lung and two-lung ventilations were simulated using a standard endotracheal tube.

![Fig 3. ETCO$_2$ in an animal undergoing two-lung, left-lung, and right-lung ventilations via a standard endotracheal tube during cardiopulmonary resuscitation.](https://doi.org/10.1371/journal.pone.0195826.g003)
A previous study performed in a non-arrest animal model reported that one-lung ventilation resulted in a significant but transient decrease in ETCO$_2$ [16]. Johnson et al. conducted a study to assess the changes in ETCO$_2$ after left main stem bronchus occlusion in open-chested dogs, and their findings revealed that ETCO$_2$ decreased at the onset of bronchial occlusion but returned to its pre-occlusion values within 3 min as hypoxic pulmonary vasoconstriction (HPV) redistributed blood flow from the non-ventilated to the ventilated lung [16]. In the present study, right-lung ventilation resulted in a significant decrease in ETCO$_2$ compared with the two-lung ventilation. This decrease appeared to remain relatively constant throughout the 5 min ventilation trial. We speculate that the persistent decrease in ETCO$_2$ in our study could be attributed to inhibition of HPV during CPR with high-flow oxygen [19].

We initially expected that left-lung ventilation would result in decreases in ETCO$_2$ that are equivalent to those observed in the right-lung ventilation. However, in contrast to our expectation, ETCO$_2$ during left-lung ventilation was significantly higher than that during right-lung ventilation and did not differ from that during two-lung ventilation. In fact, ETCO$_2$ was higher in left-lung ventilation than in two-lung ventilation, although the difference did not reach statistical significance. ETCO$_2$ is determined by total body carbon dioxide production, alveolar ventilation, and pulmonary perfusion. The total body carbon dioxide production was unlikely to differ between the right-lung and left-lung ventilations. Thus, the observed difference in ETCO$_2$ between the right-lung and left-lung ventilations has two possible explanations: different alveolar ventilation or different pulmonary perfusion. Although ventilations were delivered with the same tidal volume and respiratory rate, the actual alveolar ventilation could have differed between the right-lung and left-lung ventilations. In the present study, the gradient between PaCO$_2$ and ETCO$_2$, which is regarded as an index of dead space ventilation (although it is also affected by pulmonary perfusion), was significantly higher in right-lung ventilation than in left-lung ventilation. Thus, the difference in ETCO$_2$ between the right-lung and left-lung ventilations might be attributable to the difference in the amount of dead space ventilation. The volume of anatomic and apparatus dead space might have differed between the right-lung and left-lung ventilations because of the structural difference of the bronchial and tracheal lumens of a double-lumen tube. However, consistent results regarding the ETCO$_2$ during right-lung and left-lung ventilations were observed in the animal that underwent ventilation trials using a standard endotracheal tube, thus making this possibility less likely.

Another, more likely explanation is that left-lung ventilation might have resulted in less atelectasis and more effective alveolar recruitment, consequently improving ventilation in the relatively better-perfused, dependent part of the lung. Studies suggest that a substantial amount of atelectasis occurs in the dependent part of the lung during CPR [20,21]. Because the left lung is smaller than the right lung, the ventilation volume relative to lung volume would be larger in left-lung ventilation than in right-lung ventilation, despite the same tidal volume delivery. Thus, the relatively higher left-lung ventilation volume might have opened the dependent, atelectatic part of the lung, which receives more perfusion than the non-dependent ventral part of the lung owing to gravity, thereby improving ventilation/perfusion matching in the dependent part of the lung.

On the other hand, the difference in ETCO$_2$ between the right-lung and left-lung ventilations might be attributable to different pulmonary perfusion. However, the total pulmonary blood flow was unlikely to differ between the right- and left-lung ventilations. Chest compression depth remained constant among the three ventilation types. Although cardiac output was not measured in the present study, aortic pressure, CPP, and CBF did not differ among the three ventilation types. The observed difference in ETCO$_2$ between the left-lung and right-lung ventilations may be explained by the unequal distribution of pulmonary blood flow between the right and left lungs. In normal subjects, the left lung is slightly less perfused than
the right lung, as the left lung is smaller than the right lung [22]. However, the left pulmonary artery of pigs, similar to that of humans, represents a direct continuation of the pulmonary trunk and the main pulmonary artery is directed towards the left pulmonary artery as shown in S1 Movie. Thus, the leftward direction of the main pulmonary artery itself may favor flow toward the left lung. Moreover, the left pulmonary artery courses posteriorly, while the right pulmonary artery follows a longer and more horizontal course as it crosses the mediastinum. Thus, our finding can be explained by supposing that, in addition to the effect of the leftward direction of the main pulmonary artery, the hydrostatic pressure due to gravity also causes the blood to be carried preferentially into the left pulmonary artery, and also supposing that the effect of these factors is significantly operational in low-pulmonary blood flow states such as during CPR, while this effect is negligible in normal blood flow states. However, our explanations remain speculative at best. Thus, our finding should be considered as hypothesis-generating.

Our findings may have potential clinical implications. First, the changes in ETCO₂ caused by endobronchial intubation may confound the interpretation of ETCO₂ during CPR. In particular, the decrease in ETCO₂ following right endobronchial intubation may lead to inappropriate resuscitative effort termination. Second, the results of our study also suggest the importance of verifying correct endotracheal tube position in preventing hypoxemia during CPR. During one-lung anesthesia in otherwise normal subjects, adequate oxygenation is achieved despite one-lung ventilation due to a high inspired oxygen concentration. However, in the present study, PaO₂ fell to a dangerous level (41.5 mmHg [38.7–48.1]) following right-lung ventilation. This finding indicates that inadvertent right endobronchial intubation can have a catastrophic effect on oxygenation during CPR despite the use of 100% oxygen because pulmonary gas exchange is already impaired during CPR [23]. To date, the optimal PaO₂ level during CPR remains to be determined. However, a study that investigated the association between PaO₂ during CPR and survival to hospital admission among 145 adult cardiac arrest patients reported that hypoxemia, defined as PaO₂ ≤ 60 mmHg, was associated with decreased rate of survival to hospital admission [24].

Our study has several limitations. First, this study was conducted on pigs. Thus, the results should be verified in humans. Second, in the present study, care was taken to ensure that the endobronchial tube did not abut against the bronchial wall and the same tidal volume was delivered throughout the ventilation trials. However, endobronchial intubation in clinical settings can result in varying ventilation patterns depending on the tube tip position. If an endotracheal tube is inserted too deep into the main bronchus, the tube itself can cause obstruction of a lobar bronchus. In contrast, the non-intubated lung can be partially ventilated through the Murphy’s eye in cases when only the tip of endotracheal tube is positioned in the main stem bronchus. Third, the non-ventilated lumen was left open to the atmosphere during the one-lung ventilation trials, which might have facilitated the collapse of the non-ventilated lung. This might also have facilitated passive ventilation of the non-ventilated lung induced by chest compressions. Thus, our one-lung ventilation trials might not have precisely replicated the clinical setting of endobronchial intubation. Fourth, a randomized crossover design was utilized to reduce the number of animals used for this study. Previous studies suggested that changes in the non-ventilated lung, such as atelectasis, persisted even after restoration of ventilation [16,25]. Thus, the effects of one-lung ventilation might have persisted even after the completion of one-lung ventilation trial, which might have affected the results. Fifth, one-lung ventilation using a double-lumen endobronchial tube might exert different effects from that using a standard endotracheal tube. However, adequate respiratory excursions of the lungs were found during ventilation through the double-lumen tube at autopsy. Sixth, we could not determine the effect of one-lung ventilation on resuscitability.
Conclusions

In conclusion, ETCO$_2$ was significantly lower during right-lung ventilation than during two-lung ventilation in a pig model of CPR. However, interestingly, the ETCO$_2$ level during left-lung ventilation was comparable to that during two-lung ventilation. Further studies are required to confirm our findings and elucidate the underlying mechanism.

Supporting information

S1 Data. Raw data.
(XLSX)

S1 File. ARRIVE guidelines checklist.
(DOC)

S1 Movie. Computed tomography (CT) showing the course of the pulmonary artery in a pig undergoing cardiopulmonary resuscitation (CPR). CT was performed immediately after withholding CPR. Note the contrast material preferentially filling the left pulmonary artery.
(GIF)

Acknowledgments

The authors wish to acknowledge Jun-A Cho for her assistance in the preparation of this paper.

Author Contributions

Conceptualization: Kyung Woon Jeung, Tag Heo, Yong Il Min.

Formal analysis: Dong Hyun Ryu, Byung Kook Lee.

Funding acquisition: Kyung Woon Jeung.

Investigation: Dong Hyun Ryu, Yong Hun Jung, Byung Kook Lee, Young Won Jeong, Jong Geun Yun, Dong Hun Lee, Sung Min Lee.

Methodology: Dong Hyun Ryu, Yong Hun Jung, Young Won Jeong, Jong Geun Yun, Dong Hun Lee, Sung Min Lee, Tag Heo.

Supervision: Kyung Woon Jeung, Yong Il Min.

Validation: Dong Hyun Ryu.

Writing – original draft: Kyung Woon Jeung.

Writing – review & editing: Dong Hyun Ryu, Yong Hun Jung, Kyung Woon Jeung, Byung Kook Lee, Young Won Jeong, Jong Geun Yun, Dong Hun Lee, Sung Min Lee, Tag Heo, Yong Il Min.

References

1. Bissinger U, Lenz G, Kuhn W. Unrecognized endobronchial intubation of emergency patients. Ann Emerg Med. 1989; 18: 853–855. PMID: 2757282

2. Timmermann A, Russo SG, Eich C, Roesssler M, Braun U, Rosenblatt WH, et al. The out-of-hospital esophageal and endobronchial intubations performed by emergency physicians. Anesth Analg. 2007; 104: 619–623. https://doi.org/10.1213/01.ane.0000253523.80050.e9 PMID: 17312220

3. Dronen S, Chadwick O, Nowak R. Endotracheal tip position in the arrested patient. Ann Emerg Med. 1982; 11: 116–117.
4. Sitzwohl C, Langheinrich A, Schober A, Kräfft P, Sessler DI, Herkner H, et al. Endobronchial intubation detected by insertion depth of endotracheal tube, bilateral auscultation, or observation of chest movements: randomised trial. BMJ. 2010; 341: c5943. https://doi.org/10.1136/bmj.c5943 PMID: 21062875

5. Brunel W, Coleman DL, Schwartz DE, Peper E, Cohen NH. Assessment of routine chest roentgenograms and the physical examination to confirm endotracheal tube position. Chest. 1989; 96: 1043–1045. PMID: 2509149

6. Ryu SJ, Lee SJ, Park CH, Lee SM, Lee DH, Cho YS, et al. Arterial pressure, end-tidal carbon dioxide, and central venous oxygen saturation in reflecting compression depth. Acta Anaesthesiol Scand. 2016; 60: 1012–1023. https://doi.org/10.1111/aas.12728 PMID: 27080141

7. Weil MH, Biserà J, Trevino RP, Rackow EC. Cardiac output and end-tidal carbon dioxide. Crit Care Med. 1985; 13: 907–909. PMID: 3931979

8. Sheak KR, Wiebe DJ, Leary M, Babaeizadeh S, Yuen TC, Zive D, et al. Quantitative relationship between end-tidal carbon dioxide and CPR quality during both in-hospital and out-of-hospital cardiac arrest. Resuscitation. 2015; 89: 149–154. https://doi.org/10.1016/j.resuscitation.2015.01.026 PMID: 25643651

9. Hamrick JL, Hamrick JT, Lee JK, Lee BH, Koehler RC, Shafnner DH. Efficacy of chest compressions directed by end-tidal CO2 feedback in a pediatric resuscitation model of basic life support. J Am Heart Assoc. 2014; 3: e000450. https://doi.org/10.1161/JAHA.113.000450 PMID: 24732917

10. Sanders AB, Kern KB, Otto CW, Milander MM, Ewy GA. End-tidal carbon dioxide monitoring during cardiopulmonary resuscitation. A prognostic indicator for survival. JAMA. 1989; 262: 1347–1351. PMID: 2761035

11. Callaham M, Barton C. Prediction of outcome of cardiopulmonary resuscitation from end-tidal carbon dioxide concentration. Crit Care Med. 1990; 18: 358–362. PMID: 2108000

12. Kolar M, Krizmaric M, Klemen P, Grmec S. Partial pressure of end-tidal carbon dioxide successful predicts cardiopulmonary resuscitation in the field: a prospective observational study. Crit Care. 2008; 12: R115. https://doi.org/10.1186/cc8409 PMID: 18786260

13. Neumar RW, Otto CW, Link MS, Kronick SL, Shuster M, Callaway CW, et al. Part 8: Adult Advanced Cardiovascular Life Support: 2010 American Heart Association Guidelines for Cardiopulmonary Resuscitation and Emergency Cardiovascular Care. Circulation. 2010; 122: S729–S767. https://doi.org/10.1161/CIRCULATIONAHA.110.970988 PMID: 20956224

14. Link MS, Berkow LC, Kudenchuk PJ, Halperin HR, Hess EP, Moitra VK, et al. Part 7: Adult Advanced Cardiovascular Life Support: 2015 American Heart Association Guidelines Update for Cardiopulmonary Resuscitation and Emergency Cardiovascular Care. Circulation. 2015; 132: S444–S464. https://doi.org/10.1161/CIR.0000000000000261 PMID: 26712995

15. Gandhi SK, Munshi CA, Coon R, Bardeen-Henschel A. Capnography for detection of endobronchial migration of an endotracheal tube. J Clin Monit. 1991; 7: 35–38. PMID: 1900323

16. Johnson DH, Chang PC, Hurst TS, Reynolds FB, Lang SA, Mayers I. Changes in PETCO2 and pulmonary blood flow after bronchial occlusion in dogs. Can J Anaesth. 1992; 39: 184–191. https://doi.org/10.1007/BF03008654 PMID: 1544203

17. Jin X, Wei MH, Tang W, Povoas H, Pernat A, Xie J, et al. End-tidal carbon dioxide as a noninvasive indicator of cardiac index during circulatory shock. Crit Care Med. 2000; 28: 2415–2419. PMID: 10921572

18. Cho YC, Cho SW, Chung SP, Yu K, Kwon OY, Kim SW. How can a single rescuer adequately deliver tidal volume with a manual resuscitator? An improved device for delivering regular tidal volume. Emerg Med J. 2011; 28: 40–43. https://doi.org/10.1136/emj.2010.099911 PMID: 21131393

19. Robertsson S, Karlsson T, Wiklund L. Systemic oxygen uptake during experimental closed-chest cardiopulmonary resuscitation using air or pure oxygen ventilation. Acta Anaesthesiol Scand. 1998; 42: 32–38. PMID: 9527741

20. Markstaller K, Karmrodt J, Doebich M, Wolcke B, Gervais H, Weiler N, et al. Dynamic computed tomography: a novel technique to study lung aeration and atelectasis formation during experimental CPR. Resuscitation. 2002; 53: 307–313. PMID: 12062847

21. Markstaller K, Rudolph A, Karmrodt J, Gervais HW, Goetz R, Becher A, et al. Effect of chest compressions only during experimental basic life support on alveolar collapse and recruitment. Resuscitation. 2008; 79: 125–132. https://doi.org/10.1016/j.resuscitation.2008.03.022 PMID: 18556110

22. Cheng CP, Taur AS, Lee GS, Goris ML, Feinstein JA. Relative lung perfusion distribution in normal lungs: observations and clinical implications. Congenit Heart Dis. 2006; 1: 210–216. https://doi.org/10.1111/j.1747-0803.2006.00037.x PMID: 18377528

23. Omato JP, Bryson BL, Donovan PJ, Farquharson RR, Jaeger C. Measurement of ventilation during cardiopulmonary resuscitation. Crit Care Med. 1983; 11: 79–82. PMID: 6822084
24. Spindelboeck W, Schindler O, Moser A, Hausler F, Wallner S, Strasser C, et al. Increasing arterial oxygen partial pressure during cardiopulmonary resuscitation is associated with improved rates of hospital admission. Resuscitation. 2013; 84: 770–775. https://doi.org/10.1016/j.resuscitation.2013.01.012 PMID: 23333452

25. Markstaller K, Rudolph A, Karmrodt J, Gervais HW, Goetz R, Becher A, et al. Effect of chest compressions only during experimental basic life support on alveolar collapse and recruitment. Resuscitation. 2008; 79: 125–132. https://doi.org/10.1016/j.resuscitation.2008.03.228 PMID: 18556110