Isoforms, structures, and functions of versatile spectraplakin MACF1

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Spectraplakins are crucially important communicators, linking cytoskeletal components to each other and cellular junctions. Microtubule actin crosslinking factor 1 (MACF1), also known as actin crosslinking family 7 (ACF7), is a member of the spectraplakin family. It is expressed in numerous tissues and cells as one extensively studied spectraplakin. MACF1 has several isoforms with unique structures and well-known function to be able to crosslink F-actin and microtubules. MACF1 is one versatile spectraplakin with various functions in cell processes, embryo development, tissue-specific functions, and human diseases. The importance of MACF1 has become more apparent in recent years. Here, we summarize the current knowledge on the presence and function of MACF1 and provide perspectives on future research of MACF1 based on our studies and others. [BMB Reports 2016; 49(1): 37-44]

INTRODUCTION

Cytoskeleton in most multicellular organisms is a highly organized and interconnected network of filaments composing of microfilaments (F-actin), microtubules (MTs), and intermediate filaments (IFs). Cytoskeletal network acts as a cytoplasmic scaffold, defining the cell shape, cell mechanical properties, and functions of many other cellular events, including cell polarization, migration, division, adhesion, intracellular trafficking, and organelle locomotion. Thus, cytoskeleton is both static and adaptive, providing a dynamic cellular architecture that can remodel itself to respond to cellular changes. The tight organization and dynamics of these filaments strongly depend on their associated proteins. It has been demonstrated that spectraplakins are cytoskeletal crosslinkers with ability of interacting with all three types of cytoskeletal filaments, i.e., F-actin, MTs, and IFs (1). Spectraplakins are exceptionally long and gigantic with molecular weight of >500 kD. They are multi-domain cytoskeletal proteins and master orchestrators for coordinating cytoskeletal elements by binding to F-actin, MTs, and IFs (1, 2). Spectraplakins are evolutionarily conserved. They belong to both spectrin and plakin superfamilies (3). “Spectraplakins” are named as combinations of “spectrin” and “plakin” because they share features of both spectrins and plakins (3, 4). So far, mammalian spectraplakins have two members: microtubule actin crosslinking factor 1 (MACF1) and bullous pemphigoid antigen 1 (BPAG1). The spectraplakin family also consists of Shot/Kakapo (in Drosophila), Magellan (in zebrafish), and Vab-10 (in Caenorhabditis elegans).

MACF1, also known as actin crosslinking family 7 (ACF7), MACF, macrophin, and trabeculin-α, is a widely expressed critical spectraplakin (1). By crosslinking both MTs and F-actin and by controlling the cytoskeletal dynamics, MACF1 plays key roles in the regulation of cell migration (5-8) and cell proliferation and the maintenance of tissue integrity (9, 10). In addition, MACF1 mediates signal transduction pivotal for embryo development (11). More recently, a human disease caused by MACF1 mutation has been reported (12).

In this paper, basic molecular characteristics of MACF1 are introduced and current knowledge on MACF1 is reviewed by highlighting its versatile functions. In addition, perspectives on future research of MACF1 are provided based on studies by us and others to provide researchers the current status of MACF1 research.

MACF1: GENE, ISOFORM, AND TISSUE DISTRIBUTION

Gene and isoforms of MACF1

MACF1, a large protein with molecular weight of ~600 kD, was first discovered by Byers et al. in the effort to screen for additional members of the actin crosslinker superfamily (13). They isolated a partial cDNA of MACF1 using degenerate primer-mediated PCR and named it ACF7. Subsequently, murine ACF7 was further characterized (14) and its full-length cDNA encoding a 608-kD protein was cloned (15). Due to the associ-
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MACF1/ACF7 is encoded by MACF1 gene located on human chromosome 1p32 or mouse chromosome 4 (13, 14, 18). Human MACF1 gene contains at least 102 exons and spans over 270 kb. Genomic organization analysis of human MACF1 gene has demonstrated that MACF1 and ACF7 are the same gene (18). MACF1 gene is a hybrid of genes encoding plakins and spectrins/dystrophins (15, 18). MACF1 shares similar gene sequences with plectin, a well-characterized plakin. It has the same exon-intron boundaries for N-terminal actin-binding domain (ABD), one large exon encoding plectin repeats, and a similar serine/glycine-rich C-terminus containing GSR repeats. In addition, the spectrin repeats in MACF1 are similar to those of dystrophin gene. This hybrid gene structure of MACF1 reveals that it is related to Drosophila gene shot and Caenorhabditis elegans gene vab-10. Due to the hybrid structure of these genes and their combined features of both spectrin and plakin family members, they were assigned to the spectraplakin family (3).

Like dystonin/BPAG1 (19, 20), MACF1 displays isoform diversity. Several MACF1 isoforms (Fig. 1) produced by alternative splicing and different promoter usage have been reported. Three murine ACF7/MACF1 isoforms with a common 3' sequence but a unique 5' region were first identified by Bernier et al. (14). Isoforms ACF7-1 and ACF7-2 contain identical actin-binding domains (ABDs) but different 5'UTRs, while isoform ACF7-3 contains a unique 5'UTR and a longer N-terminal sequence with just the second half of the ABD (14). Subsequently, a fourth isoform MACF1-4 was cloned (18). Different from the three MACF1 isoforms mentioned above, MACF1-4 has no ABD at the N-terminus. Instead, it has plectin repeats (Fig. 1). Later, another alternatively spliced murine isoform of MACF1 encoding a gigantic protein of approximately 800-kD was revealed (21). It was named MACF1b with the original isoform renamed as MACF1a. MACF1b harbors extra plakin repeats between the plakin domain and the spectrin repeats of MACF1a (Fig. 1). With different N-terminal domains, the first three isoforms ACF7-1, ACF7-2, and ACF7-3 were renamed as MACF1a1, MACF1a2, and MACF1a3, respectively (21). While studying the role of MACF1 in the nervous system by adopting MACF1a conditional knockout (cKO) mice, a novel isoform MACF1c was found by Goryunov et al. (22). MACF1c is identical to MACF1a except that it has no N-terminal ABD domain (Fig. 1).

**Fig. 1.** Isoforms of MACF1 with a combination of different unique domain structure. There are seven main types of functional domains, including an actin binding domain (ABD) with CH1 and CH2 fragments, a plakin domain, a spectrin repeats rod domain with 23 α-helical spectrin repeats, two EF hand motifs, a GAR domain, plectin repeats domain, and plakin repeats domain (PRD). A, B, C represents different types.
In addition, a putative MACF/MACF1 isoform MACF2 has been reported by Sun et al. (23). Based on sequence homology analysis, a partial human cDNA clone (KIAA0728) encoding a polypeptide sharing 68% amino acid identities with human MACF was found and called MACF2. However, they found differences in chromosomal locations and nucleotide sequences between MACF and MACF2 transcripts based on unigene database, suggesting that human MACF and MACF2 represent two distinct protein products from two different genes (23). Consequently, MACF2 was demonstrated to be an isoform of BPAG1 and renamed as BPAG1-a (20). Therefore, up to date, a total of six isoforms of MACF1 (MACF1a1, MACF1a2, MACF1a3, MACF1-4, MACF1b, and MACF1c, Fig. 1) have been reported.

MACF1’s tissue distribution
As a cytoskeletal linker protein, MACF1 is ubiquitously expressed in different tissues with some isoforms having different distributions (Table 1).

Bernier et al. were the first to show that MACF1 was widely expressed in postnatal mouse tissues, including brain, spinal cord, spleen, liver, heart, skeletal muscle, stomach, lung, kidney, and skin, with the strongest expression in the lung, followed by brain, spinal cord, cardiac/skeletal muscle, and skin (14). They further demonstrated a predominant expression of MACF1 in neural, muscle, and lung tissue at the beginning of embryonic development which was continued into adulthood (24). A striking difference in tissue distribution for different MACF1 transcripts has been observed. Relatively higher levels of MACF1a2a have been detected in the brain, spinal cord, and lung, while relatively lower levels of MACF1a2 have been found in the kidney, heart, and skeletal muscles without any detection in the skin, liver, stomach, or the spleen. In contrast, MACF1a1 is predominantly found in the skin, kidney, and stomach. Similarly, MACF1a1 mRNA is detected in embryos from day 7.5 to day 10.5, whereas MACF1a2 mRNA becomes detectable only at day 10.5. MACF1a3 is predominantly detected in the brain and spinal cord. Moderate levels of MACF1a3 have been found in the skin, lung, and kidney without expression in the heart, skeletal muscle, or the liver (24). MACF1b is expressed in all tissues and throughout the development of mouse embryo (21).

MACF1 is widely expressed in human tissues, including pituitary, adrenal, thyroid, salivary gland, mammary glands, pancreas, heart, and skeletal muscle at different levels (13, 16, 17). MACF1a2 is highly expressed in the brain, heart, lung placenta, liver, kidney, and pancreas (16), while the hybridization signals of MACF1-4 are visible in all tissues, with the strongest signals in the heart, lung, pituitary gland, and placenta (18).

**Table 1. The isoforms of MACF1**

| Isoform     | Tissue distribution                                   | Domains                                                                 | References |
|-------------|-------------------------------------------------------|-------------------------------------------------------------------------|------------|
| ACF7-1/MACF1a1 | Broadly expressed with predominance in skin, kidney, and stomach | ABD (CH1, CH2), Plakin, Spectrin repeats, EF hand, and GAR.             | (14, 21, 24) |
| ACF7-2/MACF1a2 | Broadly expressed with high level in brain, spinal cord, and lung | ABD (CH1, CH2), Plakin, Spectrin repeats, EF hand, and GAR. Different 5’ UTR with MACF1a1. | (14, 16, 21, 24) |
| ACF7-3/MACF1a3 | Predominant in brain and spinal cord                   | Half ABD (CH2), Plakin, Spectrin repeats, EF hand, and GAR.             | (14, 21)   |
| MACF1a4      | Broadly expressed with high level in heart, lung, pituitary gland and placenta | Plectin repeats, Plakin, Spectrin repeats, EF hand, and GAR.             | (18)       |
| MACF1b       | Broadly expressed                                      | ABD (CH1, CH2), Plakin, Plakin repeats, Spectrin repeats, EF hand, and GAR. | (21)       |
| MACF1c       | Nervous system                                         | Plakin, Spectrin repeats, EF hand, and GAR.                              | (22)       |
the structural stability of tandem CH domains (28).

**Plakin domain**
Following the ABD at the N-terminal of MACF1, there is a plakin domain (23). The plakin domain is also a feature of members of the plakin superfamily (3). It is characterized by a high α-helical content (29). Structure analysis of BPAG1 and plectin has revealed a number of spectrin repeats in the plakin domain (30, 31), suggesting that the plakin domain is derived from spectrin repeats. The plakin domain is responsible for binding to various adhesion and signaling molecules (e.g. β4 integrin, BPAG2, and Erbin) (32-35). It plays a role in the mediation of the binding to cell junctions (29). However, the structure and the function of the plakin domain in MACF1 require further investigation.

**Spectrin repeats**
MACF1 contains both the spectrin repeats and the plakin domain. It is classified as a spectraplakin. Spectrin repeats are the typical architecture of the spectrin family. MACF1 has 23 dystrophin-like spectrin repeats (15). Each repeat contains 110~120 residues that are folded into three α helices. These helices then form an antiparallel three-helical coiled coil (36, 37). These α-helical spectrin repeats then form an extended rod-like structure and act as a spacer region that separates different functional domains at the N- and C-termini. In addition, these spectrin repeats endow the protein MACF1 with flexibility (1, 3).

**EF hand**
MACF1 contains two EF-hand calcium-binding motifs located at the C-terminus that are conserved within the spectrin superfamily (23). Sun et al. have indicated that the EF-hand motifs of MACF1 have no effect on the ability of MACF1 to interact with MTs (23).

**GAS2-related protein (GAR) domain**
Following the EF-hand motifs, there is a GAR domain located at the C-terminal of MACF1 (Fig. 1). The GAR domain is restricted to spectraplakins. It can be used to distinguish spectraplakins from spectrins and plakins (3). The GAR domain is the homology region shared between Gas2 (growth arrest-specific protein 2) and GAR22 (Gas2 related on chromosome 22) proteins. It has the function to bind and stabilize MTs (15, 23).

**Plakin repeat domain (PRD)**
PRD is unique to members of the plakin family. It only exists in the MACF1b isoform (21). Based on well-characterized A, B, and C type PRDs in desmoplakin (38, 39), Lin et al. have revealed five PRDs in MACF1b, including two A type, one C type, one incomplete B type, and one incomplete A type (21). They have further demonstrated that PRD targets MACF1b to the Golgi complex via its N-terminal portion of two consecutive A-type subdomains.

**Plectin repeats**
Plectin repeats are unique to the MACF1-4 isoform (18). MACF1-4 contains eight plectin repeats instead of ABD at the N-terminus (Fig. 1). Studies from the plakins that contain plectin repeats have demonstrated the essential role of plectin repeats in binding IFs. Therefore, the same function of plectin repeats may exist in MACF1-4. This requires further study.

**MACF1 IN CELL MIGRATION**
MACF1 has a well-known function in cell migration by coordinating F-actin and MTs dynamics (5-7). MACF1 binds to both F-actin and MTs with its ABD and MT-binding domains. In endothelial cells, MACF1 can bind along MTs, especially at the plus-ends of those MTs toward the edges of migrating cells (5, 25). Loss of MACF1 causes perturbations in MTs trajectories, dynamic stabilities, and tracking ability along actin cables, which can affect cell migration in response to wounding (5). MACF1 deficiency compromises the targeting of MTs along F-actin to focal adhesions (FAs), stabilizes FA-actin networks, and impairs epidermal migration (6). The underlying mechanisms involve the F-actin binding domains in MACF1 and an intrinsic actin-regulated ATPase domain of MACF1 for the regulation of cell migration. Therefore, there is a new functional domain in MACF1. MACF1 also sustains directional cell migration in stem cells responsible for homeostasis and wound healing (7). It has been demonstrated that GSK3β can phosphorylate and uncouple MACF1 from MTs during the regulation of cell migration by MACF1 (7). Moreover, MACF1 can regulate cell migration in neurons (8) with several partners, including ErbB2 receptor and ELMO (40, 41).

**MACF1 IN CELL SIGNALING**
Given the implication that MACF1-/- mice and Wnt3-/- and Lrp5/6 double-knockout mice have similar phenotype, the role of MACF1 in Wnt/β-catenin signal transduction has been determined by Chen et al. (11). Their results have indicated that MACF1 can interact with the Axin complex, including Axin, GSK-3β, β-catenin, and APC. They have also revealed that MACF1 is responsible for the translocation of the Axin complex from the cytoplasm to the cell membrane where MACF1 also binds to co-receptor LRPS6. Reduction of MACF1 will impair the translocation of the Axin complex, thus inhibiting Wnt-induced TCF/β-catenin-dependent transcriptional activation. In addition, MACF1 has a phosphorylation site for GSK-3β (7). It can also mediate GSK-3 signaling (8). These findings have demonstrated a role of MACF1 in cell signaling. In addition, MACF1 functions in vesicles transport. In axonal vesicle transportation, MACF1 acts as a medium to transport vesicles from the trans-Golgi network to Kif5A. Malfunction of MACF1 can cause vesicles incapable of being mobilized to the cell periphery (42). Interaction between Trans-Golgi protein p230 and MACF1 can mediate the transport of mAtg9 from the trans-
Golgi network to peripheral phagophores in the early step of autophagy (43).

**MACF1 IN EMBRYO**

As a ubiquitously-expressed protein, MACF1 is present at embryonic day 7.5 (E7.5) in mouse (24), which is quite early, suggesting that MACF1 may act as a critical molecule during embryo development. Chen et al. have confirmed the essential role of MACF1 in embryo development using MACF1−/− mice (11). The MACF1−/− mice died at the gastrulation stage with a “head-without-trunk”. They had developmental retardation at E7.5, lacking primitive streak, node, and mesoderm, similar to “head-without-trunk”. They had developmental retardation at E7.5, lacking primitive streak, node, and mesoderm, similar to the phenotypes of Wnt3−/− embryos (44) and LRP5/6 double-knockout embryos (45). Moreover, a deletion mutation in the magelan gene encoding MACF1 in zebrafish caused oocyte polarization failure (46). Taken together, these data demonstrate the importance of MACF1 in embryo development.

**MACF1 IN THE NERVE**

High levels of MACF1 and significant physiological function of the homolog of MACF1 Kakapo/Shot in the nervous system (15, 47) indicate that MACF1 might play important role in the nervous system. By generating a nervous system-specific Macf1 knockout (cKO) mouse model, Goryunov et al. (22) have underscored the importance of MACF1 in nervous system development. MACF1 cKO brains have been found to have multiple neuronal developmental defects (22). Defects in neuronal migration in the cKO cortex are also observed, suggesting a critical role of MACF1 in neuronal migration (22). In addition, the MACF1c isoform that lacks an ABD domain to MACF1a cannot compensate for the loss of MACF1a (22). Their findings indicate the necessity to interact with actin for MACF1’s function in the nervous system. Other studies have confirmed the role of MACF1 in the regulation of neuronal growth, survival, and migration (8, 48, 49). These evidences indicate the importance of MACF1 in the nerve.

**MACF1 IN SKIN**

The physiological function of ACF7/MACF1 in mammalian skin has been uncovered mainly by Elaine Fuchs’ group. Karakesisoglou et al. first showed the expression of MACF1 in the epidermis of mice and identified the functional binding sites of MACF1 for both actin and MTs (25). Subsequently, Wu et al. elucidated the function of MACF1 in the skin by conditionally knocking out (cKO) MACF1 in epidermal cells (6, 7). No gross morphological change in the skin or hair coat was observed in MACF1 cKO mice. However, a significant delay in repairing full-thickness wounds due to defects in epidermal cell migration was detected in the skin of cKO mice in response to injury. Further studies revealed that cell migration defects were rooted in perturbations in epidermal-ECM adhesion caused by a malfunction in focal adhesion dynamics due to MACF1 absence (6). Moreover, a link between MACF1 and GSK3β controlling skin stem cell migration and polarized locomotion upon injury was identified (7). Taken together, these results suggest a key role of MACF1 in maintaining skin integrity and regulating epidermal cell migration through coordinating with F-actin and MTs dynamics. Recently, MACF1 has been linked to cellular processes involved in human epidermal cells in response to extremely-low-frequency electric fields (50).

**MACF1 IN BONE**

MACF1 is a widely expressed protein. However, its role in bone tissue is not well understood. Our group have focused on the role of MACF1 in bone and reported some primary findings in osteoblasts. MACF1 is widely expressed in osteoblastic cells. It partially co-localizes with F-actin and MTs. It is scattered in the cytoplasm and cell periphery. MACF1 also participates in the osteoblast in response to environmental stimuli (51). Under diamagnetic levitation conditions, the distribution of MACF1 in osteoblasts was altered and the co-localization of MACF1 with F-actin and its co-alignment with MTs were also changed (51). Our further studies have demonstrated that MACF1 is crucial for maintaining cell shape and cell proliferation in osteoblasts (52). Knockdown of MACF1 can induce large cells with a binuclear or multinuclear structure, disrupt the organization of F-actin and MTs, and inhibit osteoblast proliferation (52). These data indicate that MACF1 is important in osteoblasts and bone.

**MACF1 IN OTHER TISSUES**

MACF1 also plays important roles in the heart, colony, and other tissues (53, 54). Fassett et al. (53) have identified a role of MACF1 in regulating cardiomyocyte MTs distribution and adaptation to hemodynamic overload for the first time using inducible cardiac-specific MACF1 knockout (KO) mice. The heart size or function of MACF1 KO mice was not affected under basal conditions. However, exacerbated pressure overload induced left ventricular (LV) hypertrophy, LV dilation, and contractile dysfunction (53). This manifestation has been found to be related to altered MT distribution and several relevant signaling proteins caused by the absence of MACF1 (53). MACF1 is also critical to the maintenance of colonic paracellular permeability because cKO of MACF1 causes significant decrease in colonic mucosal permeability associated with disturbed epithelial arrangement and cytoskeleton dysregulation (54).

**MACF1 AND HUMAN DISEASE**

In 2014, a human disease caused by a mutation in MACF1 was reported (12). This disease was found in a 12-year-old boy (the proband) who suffered from a skeletal muscle problems
with mainly manifestation of generalized hypotonia and pain. Genetic analysis revealed a single duplicated region on chromosome 1p34.3 covering a large part of the MACF1 gene. This affected all 4 major MACF1 isoforms in the proband. Further studies demonstrated that the duplication resulted in a remarkable reduction in MACF1 expression, resulting in reduced motility and internal structural changes of endothelial and satellite cells in the skeletal muscle in the proband. This novel myopathy caused by the duplication in MACF1 was named spectraplakinopathy type 1 (12). The discovery of this MACF1-related genetic human disease has shed light on the importance of MACF1 in human.

Recently, it was found that MACF1 was involved in cancer. A genetic alteration of MACF1 has been detected in numerous cancers, including breast cancer, colorectal cancer, gliomas, and others cancers (55-59). Although the function of MACF1 in cancer requires further investigation, these data suggest that MACF1 might be used as a marker for cancer diagnosis and cancer therapy.

CONCLUSIONS AND FUTURE PERSPECTIVES

In this review, we summarized the current knowledge of MACF1. As an extensively studied mammalian spectraplakin, MACF1 is broadly expressed with versatility. It has different isoforms, structures, and functions. Similar to other spectrakins, MACF1 has several isoforms expressed in different tissues. MACF1 participates in many cellular processes and specific-tissue functions. MACF1 is crucial to cell polarization and migration by regulating the dynamics of F-actin and MTs. In addition, MACF1 plays a key role in mediating cellular signaling transduction. The embryonic lethality due to a lack of MACF1 has demonstrated its essential role in embryo development. In addition, MACF1’s ubiquitous expression in all tissue indicates its physiological significance. In fact, MACF1 plays critical roles in controlling the normal development of the nervous system, maintaining skin integrity and colonic paracellular permeability, and adapting cardiomyocyte to hemodynamic overload. Moreover, our findings suggest a novel role of MACF1 in bone tissue. More importantly, mutation in MACF1 has been described in a novel myopathy disease in humans and cancer development. Thus, MACF1 is attracting more attention.

Although MACF1’s functions have been extensively studied and uncovered, many aspects of MACF1 have not been well understood. For example, how is the MACF1 expression regulated? What is the role of MACF1 with its ATPase activity? What is the exact role of MACF1 in cancer development? Considering the fact that MACF1 is involved in both osteoblast function and Wnt/β-catenin signal transduction critical for the regulation of osteoblast differentiation and bone formation, it will be interesting to see whether MACF1 can control bone formation via Wnt/β-catenin signaling. Further study of MACF1 is merited to unravel interesting and exciting new functions of MACF1 in both physiological and pathological settings.

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