Introduction

Intellectual disability (ID), with close to 3% prevalence, is one of the most common neurodevelopmental disorders.1 The important risk factor in most of Iran is consanguineous marriage2 that leads to high frequency of recessive disorders.3,4 The prevalence of ID has increased three- to five-folds in children born to first-cousin marriages.5 According to the results of some studies, autosomal recessive ID (ARID) is not rare and 13%–24% of ID cases may be caused by AR genes.6 Usually, ARID occurs by housekeeping gene mutations such as protein degradation, metabolism, DNA transcription and translation and cell division genes.6

ID and developmental delay (DD) is a heterogenous disorder with complex etiology. Whole exome sequencing (WES) facilitates the diagnosis of affected individuals with no diagnosis.7 Here, we report WES in a consanguineous family with two ID/DD affected individuals. In this study, we identified a homozygous mutation in the peptidyl-tRNA hydrolyase domain containing 1 (PTRHD1) gene in the patients. In two previous independent studies, a homozygous missense mutation in PTH2 domain (p.Cys52Tyr, p.His53Tyr) of PTRHD1 has been reported in two Iranian families with ID and Parkinsonism.8,9 Also, homozygous frameshift in the PTH2 domain (p.Ala57Argfs*26) has been recently identified in a South African family with ID and juvenile-onset Parkinsonism.10 However, in our study, the PTRHD1 variant was identified in individuals with ID/DD with first-cousin parents and without clinical symptoms of Parkinsonism. Iran has a high frequency of relative marriage (approximately 40%).11,12 Our molecular investigation highlighted the autosomal recessive of ID as the genetic cause in this family.

Materials and Methods

Patients

The study was performed on two siblings affected with ID/DD from healthy first-cousin parents (Figure 1A) originating from central Iran (Qazvin province). The study was approved by the Ethics Committee of Genetic Research Center (GRC) at the University of Social Welfare and Rehabilitation Sciences, Tehran, Iran. Informed consent was obtained from parents, and patients had been screened for fragile X syndrome.

Patient V:1

The proband is an 18-year-old male, who was born full-term with a birth weight of 3400 g (-0.26 SD), occipitofrontal circumference (OFC) of 35 cm (-0.40 SD) and length of 53 cm (+1.08 SD). He had psychomotor and speech delay. Seizure started at the age of 9 months and is being controlled by aripiprazole and MemoPlus. On examination (18 years old), his OFC was 58 cm (+2.02 SD), height was 178 cm (+0.43 SD), and weight was 100 kg (+2.20 SD). The patient was characterized by moderate ID, learning disability, self-talking but no evidence of tremor gait disturbances, psychiatric phenotypes and postural instability. There was no family history of mental retardation (Table 1).
**Patient V:3**

The second patient of the family was an 11-year-old female born full-term with an unremarkable pregnancy and neonatal period. Neonatal birth weight was 3,400 g (-0.05 SD), OFC was 35 cm (+0.11 SD) and length was 53 cm (+1.29 SD). She had a slight psychomotor and speech delay. On examination (11 years old), her OFC was 56 cm (+2.46 SD), height was 156 cm (+1.64 SD), and weight was 40 kg (+0.25 SD). She had moderate ID, hyperactivity, self-talking, aggressive behavior and learning disability; she never developed tremor gait disturbances, psychiatric phenotypes and postural instability (Table 1).

**Whole Exome Sequencing**

Genomic DNA was extracted from the peripheral blood sample of the patients and close members of the family using the salting out protocol. WES was performed for patient V:1 (Figure 1A). Exome capture and prepare library were executed using the Agilent SureSelect<sup>XT</sup> kit (Version 6) (Agilent Technologies, Lake forest, CA, USA). The Illumina NextSeq 500 system (Illumina, San Diego, CA, USA) provided paired-ends sequencing. WES analysis was carried out according to previous publication<sup>6</sup>; briefly, the sequencing quality was detected by FastQC 11.5 software.<sup>14</sup> Burrows–Wheeler Aligner (BWA) version 0.7.12-r1039 was used to align the row reads with reference genome (GRCh37/hg19); variant calling and annotation were then implemented using GATK toolkit version 3.6 and Annovar32, respectively. Variant filtering was performed by exclusion of all non-coding regions and synonymous variants. At first, the known ID/DD-causing genes were considered.<sup>6,15</sup> Variant prioritization was categorized by mineral allele frequency ≤5% in population-scale resources such as 1000 Genome project,<sup>16</sup> genomAD, ExAC Browser<sup>17</sup> and ESP6500,<sup>18</sup> Table 1.

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**Table 1. Clinical Features in Patients with PTRHD1 Variants**

| Clinical Features             | This Study (n=2) | Jaberi et al, 2016 (n=2) | Khodadadi et al, 2017 (n=2) | Kuipers et al, 2018 (n=3) |
|------------------------------|------------------|--------------------------|----------------------------|--------------------------|
| Gender                       | F/M              | M                        | M                          | F                        |
| Origin                       | Iranian          | South African (Xhosa)    |                            |                          |
| Parkinsonism                 | -                | +                        | +                          | +                        |
| Intellectual disability      | +                | +                        | +                          | +                        |
| Developmental delay          | +                | +                        | +                          | +                        |
| Learning disability          | +                | nm                       | +/-                        | +/-                      |
| Seizure                      | +/-              | +/-                      | nm                         | +/-                      |
| Abnormal behavior            | +                | +/-                      | -                          | nm                       |
| PTRHD1 variant               | p.Arg122Trp      | p.Cys52Tyr               | p.His533Tyr                | p.Ala57Argfs*26          |
| Zygosity                     | HOM              | HOM                      | HOM                        | HOM                      |

* n, Number; F, Female; M, Male; nm, not mentioned; HOM, Homozygous.
The PTRHD1 Mutation in ID

**Results**

WES was reached with the mean depth of coverage 57X with 95.4% and 91.9% coverage at 10X and 20X, respectively. A novel missense mutation, NM_001013663: c.364C>T; p.Arg122Trp was identified in the PTRHD1 gene with autosomal recessive inheritance. Of note, no potential causative homozygous and compound heterozygous/homozygous variants of known disease genes were identified. Nonetheless, the homozygous mutation in PTRHD1 was detected by an exome-wide analysis of V:1 individual. Sanger sequencing in the affected siblings confirmed the homozygous potential causative allele and also heterozygous status of the variant region was detected in the healthy family members (Figures 1B and C).

The variant is not present in different population resources (1000 Genome Project, ExAc, ESP, genomAD and our in-house database (Iranian database: http://www.iranome.com) or in Clinvar and HGMD databases. More than five in silico prediction tools have revealed the variant to be disease-causing (Table 2). Multiple sequence alignment was widely conserved across species (Figure 2).

**Discussion**

Here, we report a novel autosomal recessive variant in the C-terminal of the PTH2 domain (located in 25-139 residue positions) of the PTRHD1 gene (C2orf79) in an Iranian consanguineous family with ID. The conserved C-terminal of the PTH2 domain has a role in catalytic activity, the active site region is amino acids 72 to end residues. Variants in PTRHD1 were recently reported in three studies as combined phenotypes of ID and Parkinsonism, but in this study, we found a PTRHD1 variant in two siblings with ID and without any parkinsonism manifestations (Figure 3). For the first time, in an Iranian related family, Jaberi et al detected a homozygous missense variant in PTRHD1 (p.Cys52Tyr) in two male siblings with early-onset Parkinsonism and healthy parents. They also found a homozygous variant in ADORA1 to mention the stronger disease potential causative variant. Rest and jaw tremor, postural instability, psychomotor retardation, Bradykinesia and mental retardation features in the younger affected manifested in their patients. Early after the first report, Khodadadi et al identified a homozygous missense mutation (p.His53Tyr) in two Iranian male siblings with the same features such as movement disorder, muscle stiffness, postural instability, rest and postural tremor, speech disorder, gait disturbances, psychiatric phenotypes and mild ID phenotypes. The PTRHD1 gene was suggested as the novel causative gene for combined clinical features of Parkinsonism and ID. In the investigation by Kuipers et al, a homozygous frameshift variant in the PTRHD1 (p.Ala57Argfs*26) was detected in three affected individuals with juvenile-onset Parkinsonism and ID in a

| Tools          | Prediction Value |
|----------------|------------------|
| CADD           | 29.2             |
| SIFT           | 0.001            |
| Polyphen2      | 1                |
| LRT            | 0 (D)            |
| MutationTaster | 0.999            |
| PROVEAN        | -6               |
| DANN           | 0.999            |
| Fathmm-MKL_coding | 0.647         |

A variant is considered to be damaging and deleterious if the predicted values are as follows: CADD: higher values are more deleterious, SIFT > 0.05 (damaging), Polyphen > 0.8 (probably damaging), LRT: D: Deleterious, Mutation Taster > 0.5 (disease-causing), Provean> − 2.5 (deleterious), DANN: higher values are more deleterious, Fathmm-MKL_coding ≥ 0.5 (deleterious).

Figure 2. Multiple sequence alignment shows the region (highlighted residue) which is conserved across species.

Table 2. In Silico Analysis Tools Used for the PTRHD1 Mutation (p.Arg122Trp) Pathogenicity Prediction
South African Xhosa family (Table 1). PTHRHD1 has a ubiquitous activity that involves indirectly in ubiquitin proteasome pathway by the ubiquitin-like (UBL) protein, the PTH2 (peptidyl-tRNA hydrolase 2) domain is a UBL binding protein. PTHRHD1 was first mentioned as a peptidyl-tRNA hydrolase; however, peptidyl-tRNA was not cleaved in functional study, but in yeast PTH activity is independent of its ubiquitin proteasome pathway. The UBL acts as a shuttle protein in ubiquitin proteasome systems (UPS) that binds to Rad23 and Dsk2 to link ubiquitinated substrates, as well as Rad23 and Dsk2 acting as guides to bear UBL-substrates to the proteasome. Rad23 and Dsk2 are ubiquitin like-ubiquitin associated (UBL-UBA) proteins which interact with C-terminal of the PTH2 domain. Protein turnovers were provided by UPS whose dysfunction is well-known in the pathogenesis of neurodegenerative disorders such as ID. Neurogenesis, neurite formation, neuronal migration and synapse function are affected by the UPS which reveals its involvement in neurodevelopmental disorders.

In conclusion, in line with previous studies in ID families with consanguineous marriage, our study also confirms the elevation of the incidence of homozygous ID risks. In addition, with our genetic finding, further evidence is provided for ID-causing by PTHRHD1. The novel homozygous variant in our study probably affects substrate binding to the PTH2 active site. Functional analysis could confirm the pathogenicity of the gene in ID/DD.

Authors’ Contribution
SCh involved in analysis and interpretation of data, HN, KK and RN collaborated in conception and design of study, SM and SCh helped in proposal approval process and data collection, all authors reviewed and approved the final draft of manuscript.

Conflict of Interest Disclosures
The authors declared no conflict of interest.

Ethical Statement
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