Population pharmacokinetics of intravenous colistin sulfate and dosage optimization in critically ill patients

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Aims: To explore the population pharmacokinetics of colistin sulfate and to optimize the dosing strategy for critically ill patients.

Methods: The study enrolled critically ill adult patients who received colistin sulfate intravenously for more than 72 h with at least one measurement of plasma concentration. Colistin concentrations in plasma or urine samples were measured by ultraperformance liquid chromatography tandem mass spectrometry (LC-MS/MS). The population pharmacokinetics (PPK) model for colistin sulfate was developed using the Phoenix NLME program. Monte Carlo simulation was conducted to evaluate the probability of target attainment (PTA) for optimizing dosing regimens.

Results: A total of 98 plasma concentrations from 20 patients were recorded for PPK modeling. The data were adequately described by a two-compartment model with linear elimination. During modeling, creatinine clearance (CrCL) and alanine aminotransferase (ALT) were identified as covariates of the clearance (CL) and volume of peripheral compartment distribution (V2), respectively. In addition, colistin sulfate was predominantly cleared by the nonrenal pathway with a median urinary recovery of 10.05% with large inter-individual variability. Monte Carlo simulations revealed a greater creatinine clearance associated with a higher risk of sub-therapeutic exposure to colistin sulfate. The target PTA (≥90%) of dosage regimens recommended by the label sheet was achievable only in patients infected by pathogens with MIC ≤0.5 mg/L or with renal impairments.

Conclusion: Our study showed that the dose of intravenous colistin sulfate was best adjusted by CrCL and ALT. Importantly, the recommended dosing regimen of 1.0–1.5 million units daily was insufficient for patients with normal renal functions (CrCL ≥80 ml/min) or those infected by pathogens with high MIC values.
with MIC $\geq 1.0$ mg/L. The dosage of colistin sulfate should be adjusted according to renal function and drug exposure.

**KEYWORDS**
colistin sulfate, population pharmacokinetics, urinary recovery, dosing regimens, critically ill patients

**Introduction**

In recent years, the global prevalence of nosocomial infections caused by multidrug-resistant Gram-negative bacteria has led to the resurgence of the use of polymyxins (Nang et al., 2021). At present, there are three kinds of polymyxins used clinically, namely polymyxin B, colistin sulfate and colistin methanesulfonate sodium (CMS) (You-ning et al., 2021). Over the past decade, numerous studies have verified significant individual variability in plasma concentrations of polymyxins, especially among critically ill adult patients (Nation et al., 2016; Kubin et al., 2018; Wang et al., 2020; Li et al., 2021b). Because the pathophysiological changes of critically ill patients resulted in extreme inter- and intra-individual pharmacokinetic (PK) variability of most drugs (Abdul-Aziz et al., 2020), it was recommended that pharmacokinetics/pharmacodynamics (PK/PD) principles of antibiotics should be followed for optimizing their dosing regimens to maximize efficacy and minimize toxicity and resistance (Evans et al., 2021). Accordingly, the study on the pharmacokinetics of polymyxins is crucial for optimizing therapy regimens in critically ill patients.

Colistin sulfate, the active form of colistin, has been widely used in China since 2019 (Yu et al., 2022; You-ning et al., 2021). To date, there are limited data on pharmacokinetic properties of colistin sulfate in humans. In animal studies, the pharmacokinetic properties of colistin sulfate are closer to those of polymyxin B than those of CMS. CMS is the form of inactive prodrug which needs to be converted into the active moiety (colistin) to exert its bactericidal effect (Al-Khayyat and Aronson, 1973; Sivanesan et al., 2016). The current understanding of colistin sulfate pharmacokinetics in patients is based on one study (Yu et al., 2022), in which the PK parameters of colistin sulfate were reported to be similar to those of polymyxin B. In that study, plasma samples were used to describe the pharmacokinetic characteristics of colistin sulfate and creatinine clearance (CrCl) was identified as a covariate for the clearance of colistin sulfate. Because urine samples were not collected, it remained unclear whether colistin sulfate could be eliminated by kidney and whether a dosage adjustment was required based on renal function. Therefore, it is worth further investigating the pharmacokinetic characteristics of colistin sulfate in patients and how dosing recommendation might vary across the range of renal functions.

In view of the above, there is an urgent need to investigate the PK/PD of colistin sulfate in critically ill patients and to optimize its clinical application. In this study, plasma and urine samples were collected to build a population PK model as well as to investigate the urinary excretion of colistin sulfate following intravenous administration. The objectives of the current study were to identify patient factors influencing PK, to explore the correlation of PK/PD index with therapeutic efficacy and then to propose dosage regimens of colistin sulfate using Monte Carlo simulation. The findings of this study will provide important new information on the renal handling of colistin sulfate and dosage regimens for clinical use.

**Materials and methods**

**Study design and patient population**

This study was conducted at the Third Xiangya Hospital of Central South University between May 2021 and April 2022. The design of this research fully conformed to the principles of the Helsinki Accords, and the study protocol was approved by the Research Ethics Committee of the Third Xiangya Hospital of Central South University (No: 2021-S396). Written informed consent was obtained from all patients or their legal representatives. Adult patients (aged $\geq 18$ years) who had received intravenous colistin sulfate (Colistin, Asia Pioneer Pharmaceutical Co., Ltd., Shanghai, China) for $\geq 72$ h with at least one colistin concentration measurement were eligible for inclusion.

Colistin sulfate treatment was at the discretion of their medical teams. In clinical practice, colistin sulfate was administered at 1.0–1.5 million IU (10,000 IU = 0.44 mg) per day divided into 2 or three doses of infusions over 60–120 min. Colistin sulfate was routinely subjected to therapeutic drug monitoring (TDM) in our hospital. Blood samples (2 ml each) for colistin sulfate TDM samples were acquired immediately before starting and upon completing the infusion after at least 3 days of therapy. Other blood samples were collected randomly from residual blood of blood gas analysis or routine blood tests. In addition, urine samples from 6 patients with catheters in the intensive care unit were collected and recorded across a dosage interval (0–12 h after dosing) after 72 h of therapy. All samples were centrifuged for 10 min (4,000 ×g), and the supernatant was immediately collected and stored at $-80^\circ$C pending analysis. The therapeutic regimen of colistin sulfate, including dosage, dosing interval and infusion duration and sampling time, was recorded in detail for each patient.

The demographic information and clinical variables were retrieved from the computerized medical records, including age,
sex, weight, infection diagnosis, comorbidities, acute physiology and chronic health evaluation (APACHE II), liver function indices (such as alanine aminotransferase (ALT), aspartate aminotransferase (AST), total bilirubin (TBIL), direct bilirubin (DBIL), total bile acid (TBA), total protein (TP) and albumin (ALB)), renal function indices (such as uric acid (UA), blood urea nitrogen (BUN) and creatinine clearance (CrCL) estimated by the Cockcroft-Gault equation (Cockcroft and Gault, 1976)), routine blood tests (e.g., white blood cell (WBC), neutrophilic granulocyte percentage (NEUT), hemoglobin (HGB), red blood cell (RBC), hematocrit (HCT), platelet count (PLT)), coagulation indices (such as prothrombin time (PT), thrombin time (TT), activated partial thromboplastin time (APTT), prothrombin activity (PA), fibrinogen (FIB), C-reactive protein (CRP), procalcitonin (PCT), lactic acid (LAC) value and 24-h urine volume.

Quantification of colistin sulfate concentrations

Colistin A and colistin B in plasma and urinary samples were quantified using a validated ultra-performance liquid chromatography tandem mass spectrometry (LC-MS/MS) assay. Briefly, samples were pretreated with acetonitrile, and 1 μL supernatant was injected into the LC-MS/MS instrument for analysis. A Waters XBridge BEH HILIC (4.6 × 150 mm, 2.5 μm) column was used. The mobile phase, consisting of eluents A (0.1% formic acid in water, vol/vol) and B (methanol), was delivered at a flow of 0.5 ml/min. Analysis of independently prepared quality control samples indicated good reproducibility (coefficients of variation ≤12.5%), and accuracy ranged from 88.52% to 109.43%. The limit of quantification was 0.034 mg/L for colistin A and 0.059 mg/L for colistin B. Since colistin A and B had similar structures, molecular weights and pharmacokinetic characteristics, the concentrations of colistin A and B were added together and referred to as colistin (Li et al., 2003; Couet et al., 2011).

Population pharmacokinetic modeling

Population PK analysis was performed using a non-linear mixed-effect modeling with the Phoenix® NLME software (version 8.3.4, Certara L.P. United States). The first-order conditional estimation-extended least-squares (FOCE-ELS) method was used to develop the PPK model. One- and two-compartment models with linear elimination were compared based on Akaike information criteria (AIC) and Bayesian information criterion (BIC). The inter-individual variability of PK parameters was assumed to follow a log-normal distribution and described using an exponential model, and residual variability was described by an additive, proportional, or combined error model. The coefficient of variation (CV%), goodness-of-fit plots, and the likelihood ratio test (-2 loglikelihood; -2LL) were used to evaluate the model.

Then, a stepwise covariate modeling (SCM) method was used to explore candidate covariates on PK parameters, including age, sex, weight, ALT, AST, TBIL, DBIL, TBA, TP, ALB, UA, BUN, CrCL, WBC, NEUT, HGB, RBC, HCT, PLT, PT, TT, APTT, PA, FIB, CRP, PCT, LAC, and 24-h urine volume, where continuous covariates (e.g., age) were modeled by using the power function and categorical covariates (e.g., sex) were described by an exponential function. By comparing with the base model, a drop in objective function value (OFV) > 3.84 (p < 0.05, df = 1) during forward selection steps and an increase of OFV >6.63 (p < 0.01, df = 1) in backward elimination steps were the inclusion criteria for covariates.

Finally, the reliability of the final model was evaluated by goodness-of-fit plots. Additionally, the performance of the final model was assessed by a prediction-corrected visual predictive check (pcVPC) of 1,000 replicates using Monte Carlo simulations, and a bootstrap analysis with 1,000 samples was conducted to evaluate the model stability.

The renal clearance (CLR) of colistin was calculated as the ratio of the amount recovered in urine to the area under the plasma concentration-time curve (AUC) during the same period (Sandri et al., 2013). The percentage of colistin sulfate excreted in urine was defined as the amount of colistin recovered in urine divided by the dose administered. The fraction of renal clearance was estimated by CLR divided by total body clearance (CL) (Li et al., 2003).

Monte carlo simulation

Based on the final population model, Monte Carlo simulations with 1,000 replicates were performed for 12 clinically relevant dosing regimens at different MICs ranging from 0.5 to 2 mg/L with various renal functions (CrCL: 10, 50, 80, 120 mL/min). The 12 fixed dosing regimens were a loading dose of 1 million units, 1.5 million units or no loading dose, followed by 500, 000 units q12h, 500, 000 units q8h, 750, 000 units q12h or 1.0 million units q12h. The infusion time for a dose was set as 2 h.

The AUC for each dosing regimen was calculated at 24 or 72 h after therapy to evaluate the probability of target attainment (PTA) of fAUC/MIC ≥20 (Dudhani et al., 2010a; Cheah et al., 2015), where f is the unbound fraction of colistin defined as 0.49 (Nation et al., 2016). Dosing regimens were considered appropriate if PTA ≥90% was achieved. In addition, the average steady-state plasma colistin concentration (C\text{ss,avg}) was used to evaluate the probabilities of efficacy and toxicity, and the desired C\text{ss,avg} were designed to achieve target attainment rates of >80% for C\text{ss,avg} ≥ 2 and <30% for C\text{ss,avg} ≥ 4 mg/L (Nation et al., 2016).
Outcomes

The primary endpoint was the clinical outcome, and the secondary endpoint was the microbiological outcome. Clinical outcomes were defined as valid, if signs and symptoms of infection disappeared or improved on Day 7 after the initiation of colistin therapy; or invalid, defined as any of the following criteria: death related to infection, hemodynamic instability, deteriorated Sequential Organ Failure Assessment (SOFA) score, constant fever >38.5°C, repeat isolation of bacterial phenotypically identical to the index isolate. Microbiological outcomes were defined as clearance, if bacterial culture did not find the original bacteria from the same infection site within the full course of colistin sulfate, or uncleared, defined as repeat isolation of bacteria on or after Day 7 after administration of colistin sulfate. Colistin-induced nephrotoxicity was defined using RIFLE (risk, injury, failure, loss of kidney function, and end-stage renal disease) criteria during treatment (Bellomo et al., 2004).

Receiver operating characteristic (ROC) curves were generated to assess the drug exposure parameters AUC<sub>0-24h</sub>/MIC, the trough, peak and average concentrations at steady state (C<sub>ss</sub>,min, C<sub>ss</sub>,max and C<sub>ss</sub>,avg) as predictors of clinical outcome. Then the area under the ROC curve (AUC<sub>ROC</sub>) was calculated to evaluate the correlation between the above parameters and clinical outcome.

Results

Patients

A total of 20 patients were enrolled in the study. The demographic data of the patients are summarized in Table 1. Renal functions varied considerably among the patients, and the CrCL ranged from 6.5 to 193.8 ml/min. Two patients received continuous renal replacement therapy (CRRT). Treatment was initiated with a loading dose in four patients, and the most common daily maintenance dose was 1.5 million units divided into 2 or three infusions lasting 1 or 2 h. Carbapenem-resistant A. baumannii (50%) and K. pneumoniae (30%) were the two most common bacteria isolated from bronchoalveolar lavage fluid, blood, puncture fluid or urine.

Population pharmacokinetic modeling

Overall, 98 plasma concentration measurements collected after 72 h of therapy were included in the analysis. Among them, 38.8% (n = 38) were samples with troughs, 27.6% (n = 27) were samples with peaks, and 33.7% (n = 33) were random samples. A two-compartment model with linear elimination performed better than a one-compartment model (AIC: 160.2 vs. 174.2, BIC: 183.5 vs. 187.1, respectively). Thus, a two-compartment model with first-order elimination was selected as the base model, and a proportional error model was used to evaluate the residual variability. Next, covariate model building identified CrCL as the effective covariate of central compartment clearance.
TABLE 2 Parameter estimates and bootstraps results of the final population pharmacokinetic model.

| Parameter             | Final model | Bootstrap |
|-----------------------|-------------|-----------|
|                       | Estimate  | CV (%) | 95% CI        | Shrinkage (%) | Median | 95% CI       |
| tvV(L)                | 16.1      | 8.70   | 13.3–18.9     | 43.6           | 16.2   | 12.9–19.0    |
| tvV2(L)               | 50.5      | 22.0   | 28.4–72.6     | NA             | 53.8   | 24.8–200     |
| tvCL (L/h)            | 1.50      | 11.7   | 1.15–1.84     | 5.06           | 1.46   | 0.69–1.89    |
| tvCL2 (L/h)           | 1.71      | 22.0   | 0.96–2.45     | NA             | 1.78   | 1.29–3.77    |
| dV2dALT               | 0.635     | 16.6   | 0.426–0.844   | NA             | 0.633  | 0.146–0.954  |
| dCLdCrCL              | 0.353     | 29.8   | 0.144–0.563   | NA             | 0.373  | 0.0766–1.12  |
| Interindividual variability |          |        |                |                |        |              |
| u2V                   | 0.0267    | 66.4   | NA             | NA             | 0.0174 | 0.000–0.113  |
| u2CL                  | 0.197     | 29.2   | NA             | NA             | 0.175  | 0.0656–0.803 |
| Residual variability (σ) | 0.228   | 15.4   | 0.158–0.297   | NA             | 0.221  | 0.147–0.292  |
| stdev0                |           |        |                |                |        |              |

CV%, percent confidence of variation; CI, confidence interval; tvV, typical value of volume of central compartment distribution (V); tvV2, typical value of volume of central compartment distribution (V2); tvCL, typical value of central compartment clearance (CL); tvCL2, typical value of inter-compartmental clearance (CL2); dV2dALT, exponential parameter coefficient of alanine aminotransferase (ALT) to V2; dCLdCrCL, exponential parameter coefficient of creatinine clearance (CrCL) to CL; u2V, variance of inter-individual variability for V; u2CL, variance of inter-individual variability for CL; stdev0, standard deviation.

(CL) and ALT as a significant covariate for volume of peripheral compartment distribution (V2). Age, sex, weight, and other clinical variables had no statistically significant relationship with PK parameters. The final PK model equations are shown in Eqs 1–3, where the value of inter-compartmental clearance (CL2) was estimated as 1.71 L/h with no inter-individual variability. The population PK estimate parameters and bootstrap replicates are presented in Table 2, which indicated that the final model had qualified stability.

\[
V(L) = 16.1 \times \exp(\eta V) \tag{1}
\]

\[
V2(L) = 50.5 \times \left(\frac{ALT}{37}\right)^{0.635} \tag{2}
\]

\[
CL(L/h) = 1.50 \times \left(\frac{CrCL}{57.5}\right)0.353 \times \exp(\eta CL) \tag{3}
\]

Goodness-of-fit plots for the final model adequately described the observed data, as shown in Figure 1. The scatterplots of observed concentrations versus PRED and IPRED revealed a perfect correlation. The CWRES versus time or PRED of the final model showed a randomly normal distribution, and most residuals were centered on zero within an acceptable range (−2 to 2). The pcVPC showed that the model reliably predicted the observed data presented in Figure 2, where the most observed plots were within the 90% CIs of the predicted corresponding quantiles. Overall, the results supported the predictive performance of the final model and its use for subsequent dosing simulations.

A total of 41 urine samples from six patients were used to describe urinary excretion of colistin sulfate. The median percentage of colistin dose that was excreted in urine was 10.05% (range, 2.24%–32.09%), and the median renal clearance (CLcr) was 0.209 L/h (range, 0.053–0.405 L/h). The CLcr of colistin sulfate was a small percentage of CL with large variability (median, 12.37%; range, 2.52%–63.91%) (Figure 3 and Supplementary Table S1).

Monte carlo simulations

The PTAs of colistin with various dosing regimens when CrCL values ranged from 10 to 120 ml/min, predicted from the Monte Carlo simulations, are presented in Figure 4 and Supplementary Table S2. For an MIC of 0.5 mg/L, the PTAs of almost all simulated regimens were greater than 90% on Day 3, except for the regimen with no loading dose followed by 0.5 MU q12h. When the MIC was less than 1 mg/L, administration of the doses recommended by the label instruction (1.0 MU or 1.5 MU) achieved the PTA over 90% only in patients with CrCL ≤10 ml/min. Yet, when the MIC value was at the current EUCAST susceptibility breakpoint of 2 mg/L, most simulated regimens failed to achieve the target PTA; the only regimen that reached the target PTA was in patients with CrCL ≤10 ml/min treated with 1.0 MU loading dose and maintained on 0.75 MU q8h. Furthermore, we found that a loading dose made it possible to achieve the target PTA on Day 1, which indicated that the loading dose was essential for colistin sulfate treatments.

In addition, the Cavg, avg for various dosing regimens with CrCL ranging from 10 ml/L to 120 ml/min are presented in Figure 5 and Table 3. Based on the simulation results, we conclude that the dose of colistin sulfate should be adjusted according to CrCL. For patients with severe renal insufficiency (≤10 ml/min), a 1.0 MU daily maintenance dose should be selected. For patients with CrCL in 50 ml/min, 80 ml/min and
120 ml/min, the recommended regimen was a loading dose of 1.5 MU followed by daily maintenance doses 1.5, 2.0 or 2.25 MU divided into 2 or three infusions, respectively.

**Correlation of drug exposure to outcome**

In this study, we evaluated the efficacy of colistin sulfate among 16 patients who received colistin sulfate as definitive therapy; the other four patients who received colistin as empirical therapy could not be assessed in accordance with our evaluation criteria. Overall, clinical outcomes were successful in 12 of 16 cases (75%), and microbiological eradication was observed in 13 of 16 subjects (81.25%) with a causative pathogen MIC range of 0.5–2 mg/L (Supplementary Table S3). None of the patients experienced colistin-induced nephrotoxicity during the treatment. The AUC$_{RUC}$ (0.854, $p = 0.039$) for AUC$_{ss, 0–24h}$/MIC was the largest, indicating that AUC$_{ss, 0–24h}$/MIC had the strongest correlation with efficacy (Supplementary Figure S1). The mean ratio of AUC$_{ss,0–24h}$/MIC was 85.16 h (range 27.49–211.18) and 40.99 h (range 13.83–63.91) in the valid group and invalid group, respectively ($p = 0.104$).

**Discussion**

This population PK study has made a significant contribution to understanding the clinical application of
Colistin sulfate administered intravenously in critically ill patients. To date, there have been limited population pharmacokinetic studies of colistin sulfate in patients (Yu et al., 2022). Our results showed that a two-compartment model with first-order elimination best fitted the PK of colistin sulfate in critically ill patients, which was inconsistent with a previous study in which a one-compartment model with linear-elimination was used to describe the pharmacokinetic characteristics of colistin sulfate (Yu et al., 2022). This could be due to the intensive combined scattered sampling strategy used in the current study (Chen et al., 2022). We collected 6-8 blood samples from six patients across the distribution and elimination phases of colistin in dosing intervals, which fits to better describe the characteristics of distribution for colistin sulfate in humans (Li et al., 2021a). We also collected urine samples from six patients with catheters among 20 subjects to explore urinary excretion. The urinary recovery of colistin sulfate was low with significant inter-individual variability (median, 10.05%; range, 2.24%–32.09%) in humans, which was higher than those reported in animal studies, in which 0.18 ± 0.14% or 0.13 ± 0.09% of the dose was recovered from urine of Sprague-Dawley rats (Li et al., 2003) or dogs (Al-Khayyat and Aronson, 1973), respectively. Renal clearance of an unchanged drug was a relatively small fraction of total clearance in our study (median, 12.37%), which indicated that non-renal clearance may be the major elimination pathway of colistin sulfate in humans as demonstrated in rats (Li et al., 2003), and was similar to polymyxin B sulfate in patients (Sandri et al., 2013). However, renal excretion of colistin sulfate varied widely among patients. The CLR of patient eight was 63.91%, which was far higher than that of the other patients (Supplementary Table S1). This implied that colistin sulfate was subject to very extensive net reabsorption from tubular urine back into blood, as previously reported in rats (Li et al., 2003) and for polymyxin B in patients (Sandri et al., 2013).

In the present study, the estimated typical value of CL was 1.50 L/h for colistin sulfate, which was lower than the CL of colistin converted from CMS (2.92 L/h) (Couet et al., 2011), and similar to the CL of polymyxin B reported previously (range 1.59–2.86 L/h) (Wang et al., 2021; Chen et al., 2022). CrCL was identified to have a significant effect on the CL of colistin sulfate, which was in agreement with a previous study (Yu et al., 2022) and the disposition characteristic of polymyxin B (Wang et al., 2020; Li et al., 2021b; Yu et al., 2021). Monte Carlo simulations demonstrated that the colistin sulfate exposure showed significant negative correlation with renal function. Accordingly, in patients with renal dysfunction (CrCL ≤ 50 ml/min), 1.0–1.5 million IU per day divided into 2-3 doses in accordance with the label recommended regimen, would achieve the target therapeutic window. However, in patients with normal renal function (CrCL > 80 ml/min), a regimen of no less than 2.0 million IU/day would be a better option.

In the current study, we used $\text{AUC/MIC} \geq 20$ to evaluate the PTA (Dudhani et al., 2010b), which is widely used to optimize the regimen for polymyxin B (Li et al., 2021b; Yu et al., 2021). The regimen of a 1.0–1.5 million IU daily dose recommended by the current label of colistin sulfate reach PTA ≥90% only for MIC values ≤0.5 mg/L, which was in line with a previous study (Yu et al., 2022). High-dose regimens should be considered for the patients infected by organisms with an MIC of ≥1 mg/L. In our limited cases with causative pathogens, all valid patients were infected by organisms with MIC values of 0.5 mg/L, and 50% of invalid patients had a causative pathogen with MIC ≥1 mg/L (Supplementary Table S1).
In addition, we confirmed that AUC_{ss, 0-24h}/MIC showed a stronger correlation with the clinical efficacy than Css_{avg}, Css_{min}, or Css_{max} (Supplementary Figure S1).

Moreover, ALT was identified as a significant covariate for volume of peripheral compartment distribution (V2) which explained inter-individual variability; ALT has also been used to describe the pharmacokinetic characteristics of tacrolimus (Lu et al., 2015) and atorvastatin lactone (Dostalek et al., 2012). In our study, because ALT values varied widely from 7 U/L to 495 U/L, the effect of the liver on the disposal of colistin sulfate was considered. Biliary excretion has been verified as one of the pathways for the elimination of polymyxin B (Manchandani et al., 2016) and CMS (Li, 2004) in animal models. We speculated that the liver may

![Figure 4](image)

**FIGURE 4**
PTAs for dosing regimens performed based on the final PPK model on Day 3. The dotted line represents the target PTA of 90%. A to C show MICs of 0.5 mg/L, 1.0 mg/L, and 2.0 mg/L, respectively.

| Dosing regimens          | Probability (%) of Css_{avg} > 2 mg/L | Probability (%) ofCss_{avg} > 4 mg/L |
|--------------------------|--------------------------------------|-------------------------------------|
|                          | 10 ml/min | 50 ml/min | 80 ml/min | 120 ml/min | 10 ml/min | 50 ml/min | 80 ml/min | 120 ml/min |
| 0.5MU q12h               | 74        | 29        | 21        | 15         | 4         | 0         | 0         | 0         |
| 1.0MU + 0.5MU q12h       | 85.5      | 37.5      | 28        | 17         | 7.5       | 0.5       | 0         | 0         |
| 1.5MU + 0.5MU q12h       | 94        | 53        | 42        | 33         | 28.5      | 3         | 1.5       | 1.5       |
| 0.5MU q8h                | 96.5      | 75        | 62.5      | 49         | 46        | 7         | 2.5       | 1         |
| 0.75MU q12h              | 96.5      | 73.5      | 58        | 45         | 41        | 7         | 2.5       | 0.5       |
| 1.0MU + 0.5MU q8h        | 98        | 77        | 69.5      | 56.5       | 56        | 16.5      | 12.5      | 7.5       |
| 1.5MU + 0.5MU q12h       | 98        | 70        | 59        | 43.5       | 44.5      | 12        | 6         | 3         |
| 0.5MU q12h               | 98.5      | 82        | 68.5      | 57         | 66.5      | 20        | 12        | 7.5       |
| 1.0MU + 0.5MU q8h        | 98.5      | 81        | 65.5      | 52.5       | 59.5      | 14.5      | 7.5       | 4.5       |
| 1.5MU + 0.75MU q12h      | 99.5      | 88.5      | 78        | 69         | 76        | 33        | 23        | 13        |
| 1.0 MU q12h              | 99.5      | 88.5      | 78        | 69         | 79        | 39        | 26.5      | 16.5      |
| 1.5MU + 1.0 MU q12h      | 100       | 93        | 87        | 80.5       | 87        | 55.5      | 36.5      | 28.5      |

10,000 U = 0.44mg; q12h: Every 12 h; q8h: every 8 h; Css_{avg}: Average steady-state plasma concentration.
also be involved in the disposal of colistin sulfate, which remains to be further explored.

Colistin sulfate and polymyxin B are administered by an active moiety and show similar pharmacokinetic properties. Weight-based dosing strategies for polymyxin B are recommended by international consensus guidelines (Tsuji et al., 2019). However, similar to the study of polymyxin B in Chinese patients (Li et al., 2021b; Wang et al., 2021; Yu et al., 2021), no effect of total body weight on the colistin sulfate PK parameter was observed during modeling in the present study. This may be due to the narrow range of weight values among the included subjects (range, 45–65 kg) and the lack of weight-based doses administered in the clinic, affording low power to detect an effect if present. More research needs to be carried out to explore the relationship between weight and colistin sulfate exposure and whether either weight-based dosing scheme should be used.

Some potential limitations should be considered when interpreting the results of the current study. First, most blood samples were collected from residual blood in the clinic, and therefore, both intensive and sparse sample schedules were used in the study, even though the dosing and sampling time were precisely recorded. Second, the sample size was relatively small, and further evaluation and external validation are needed to confirm the conclusions. Third, we explored urinary colistin sulfate excretion, and large inter-individual variability indicated that urinary excretion should not be ignored. Additional greater sample studies are needed, and TDM is needed for individualized medication. Finally, we evaluated the efficacy and PTA derived from calculated unbound drug fraction rather than measured unbound drug fraction as the previous studies (Wang et al., 2021; Li et al., 2021b; Yu et al., 2022), which may be different from the actual values especially in patients with

FIGURE 5
Violin plots of simulation results of average steady-state plasma colistin concentration (Cs, avg) for dosing regimens with various CrCL values. The dotted lines represent the interquartile of simulated exposure; the solid lines represent the median of simulated exposure; the red dashed lines represent potential toxicity concentration (4 mg/L); the blue dashed lines represent the target concentration (2 mg/L). A to D show the CrCL values of 10 ml/min, 50 ml/min, 80 ml/min and 120 ml/min, respectively.
hypoalbuminemia. Further clinical validation of the regimens based on fAUC/MIC simulation is needed, although we evaluated the clinical outcomes and toxicity in limited cases.

Conclusion

In conclusion, we have presented a reasonable two-compartment population pharmacokinetic model of intravenous colistin sulfate in critically ill patients in China. The study demonstrated that doses of intravenous colistin sulfate are best scaled by CrCL and ALT. Dosage adjustments are recommended according to renal function and drug exposure. Our data indicated that the current dosing recommendation by the label was insufficient for patients with normal renal functions (CrCL ≥80 ml/min) or those infected by pathogens with MIC ≥1.0 mg/L. TDM was recommended to optimize the dosage of colistin sulfate, and further clinical studies on colistin sulfate PK/PD are urgently needed.

Data availability statement

The raw data supporting the conclusion of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving human participants were reviewed and approved by The design of this research was completely conformed to the principles of the Helsinki Accords, and this study was approved by the Ethics Research Committee of the Third Xiangya Hospital of Central South University (No: 2021-5396). All subjects signed the informed consent that residual blood samples from arterial blood gas or blood routine were collected for colistin concentration determination. The patients/participants provided their written informed consent to participate in this study.

Author contributions

X-YL, JX, PY and Z-XC had full access to the conception and design of the study. Y-SS, X-BX, W-CF, W-H, Y-BC, X-XF and L-ZL collected the clinical data. PQ measured colistin sulfate plasma concentrations. X-YL and LW established the PPK Model; X-YL and JX wrote the first draft of the manuscript. PY and Z-XC contributed to revising and proofreading the manuscript. All authors contributed and approved the submitted version of the manuscript.

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Conflict of interest

WL was employed by Changsha VALS Technology Co. Ltd. The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2022.967412/full#supplementary-material
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Glossary

TDM therapeutic drug monitoring
APCHE II acute physiology and chronic health evaluation
ALT alanine aminotransferase
AST aspartate aminotransferase
TBIL total bilirubin
DBIL direct bilirubin
TBA total bile acid
TP total protein
ALB albumin
UA uric acid
BUN blood urea nitrogen
CrCl creatinine clearance
WBC white blood cell
NEUT neutrophilic granulocyte percentage
HGB hemoglobin
RBC red blood cell
HCT hematocrit
PLT platelet count
PT prothrombin time
TT thrombin time
APTT activated partial thromboplastin time
PA prothrombin activity
FIB fibrinogen
CRP C-reactive protein
PCT procalcitonin

LAC lactic acid
LC-MS/MS liquid chromatography tandem mass spectrometry
FOCE-ELS first-order conditional estimation-extended least-squares;
AIC Akaike information criteria
BIC Bayesian information criterion
OFV objective function value
SCM stepwise covariate modeling
CV% coefficient of variation
PPK population pharmacokinetics
DV observed concentrations
PRED population prediction
IPRED individual population prediction
CWRES conditional weighted residuals
pcVPC prediction-corrected visual predictive check
CLR renal clearance
AUC the area under the plasma concentration-time curve
Cl clearance
PTA probability of target attainment
C_{ss,avg} average steady-state plasma colistin concentration
SOFA sequential organ failure assessment
RIFLE risk, injury, failure, loss of kidney function, and end-stage renal disease
ROC receiver operating characteristic
CRRT continuous renal replacement therapy