Semisynthetic fluorescent pH sensors for imaging exocytosis and endocytosis

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The GFP-based superecliptic pHluorin (SEP) enables detection of exocytosis and endocytosis, but its performance has not been duplicated in red fluorescent protein scaffolds. Here we describe “semisynthetic” pH-sensitive protein conjugates with organic fluorophores, carbocyanine, and Virginia Orange that match the properties of SEP. Conjugation to genetically encoded self-labeling tags or antibodies allows visualization of both exocytosis and endocytosis, constituting new bright sensors for these key steps of synaptic transmission.

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Synaptic transmission is mediated by the rapid fusion of synaptic vesicles (SVs) with the plasma membrane. Precise monitoring of exocytosis is important for elucidating fundamental mechanisms of cell–cell communication, and investigating the underlying causes of neurological disorders. The lumen of synaptic vesicles are typically acidified (pH 5.6) by the action of vesicle-resident V-ATPases, which creates the driving force for neurotransmitter uptake. Upon fusion with the plasma membrane, the contents of the vesicle rapidly equilibrate with the extracellular environment (pH 7.4). This large change in pH allows for the visualization of exocytosis using a pH-sensitive variant of green fluorescent protein (GFP) that is expressed as a fusion with a vesicular membrane protein. This “supercleptic pHluorin” (SEP) exhibits ideal properties for detecting the change in pH upon vesicle fusion, with near-ideal pKa, cooperative protonation, and low background fluorescence in the protonated state, making it an excellent tool for monitoring exocytosis in living cells.

A useful extension of this technology has been the creation of pH sensors based on red fluorescent proteins (RFPs) such as mOrange2, pHTomato3, pHOran4, and pHuji5. Longer excitation wavelengths are less phototoxic, elicit lower levels of autofluorescence, facilitate multicolor imaging experiments, and allow concomitant use of optogenetics. Nevertheless, it has proven difficult to engineer red-shifted pHluorins that match the optimal pKa, cooperativity, and dynamic range of SEP, perhaps due to inherent limitations in RFP scaffolds. More generally, these techniques rely on overexpression of reporter proteins in SVs and the effect of overexpression is a confounding factor in interpreting experimental results.

To circumvent the problems with genetically encoded red fluorophores, we develop a "semisynthetic" sensor platform using red-shifted chemical fluorophores derived from fluorescein—carbofluorescein (CFI) and Virginia Orange (VO)—with pH sensitivities similar to SEP. We have adapted these fluorophores to serve as probes for exo- and endocytosis in two ways. First, we synthesize benzylguanine derivatives, which bind specifically to SNAP-tag ligands, to label exogenous vesicular proteins fused with the SNAP-tag. Second, we label an antibody targeted to the extracellular epitope of a vesicular protein, synaptotagmin1, which allow the imaging of the exo-/endocytosis cycle of an endogenous protein.

Results
In vitro characterization of CFI and VO as pH sensors. Given the limitations of pH-sensitive RFPs and the potential problems with overexpression of sensor proteins, we pursued an alternative strategy: creation of semisynthetic pH indicators using organic pH-sensitive dyes attached to either expressed self-labeling tags such as the SNAP tag or antibodies that recognize native vesicular proteins (Fig. 1a). To match the performance of SEP, we required a pH-sensitive organic dye that can undergo a cooperative transition from a bright, fluorescent form at neutral pH to a nonfluorescent form at low pH. Unfortunately, the majority of pH-sensitive dyes do not meet these requirements. The

![Figure 1 Design and characterization of semisynthetic fluorescent reporters for exocytosis.](image)
Fig. 2 Detection of single exocytosis events in PC12 cells. **a** Experimental scheme: prior to imaging, cell-permeable VO/CFI-SNAP-tag ligand is incubated with transfected neuronal cultures to specifically label VAMP2-SNAP. Both VAMP2-SEP and VAMP2-SNAP-VO/CFI are quenched at acidic pH and exhibit maximal fluorescence at the surface. **b, d, f** TIRF microscopic images of PC12 cells expressing VAChT-SEP (left) and VAChT-red shifted indicator (right) undergoing fusion. Images of whole cell with a fusion event indicated with arrow (top); time-lapse of the marked exocytic event (middle, 3.5 × 3.5 µm); normalized intensity traces for the marked event in the green and red channels (bottom); scale bars: 5 µm. **b** VAChT-SNAP-CFl. **d** VAChT-SNAP-VO. **f** VAChT-pHuji. **c, e, g** Plots of normalized fluorescence vs. time averaged across single-vesicle fusion events for vesicles labeled with VAChT-SEP (green) or VAChT-red-shifted indicator (magenta); error bars show ± s.e.m. **c** VAChT-SNAP-CFl, n = 113 events from 5 cells. **e** VAChT-SNAP-VO, n = 116 events from 10 cells. **g** VAChT-pHuji, n = 126 events from 3 cells.
archetypical small-molecule pH sensor is fluorescein (Fl, 1, Fig. 1b), which transitions between a highly fluorescent dianion (12−) and a less fluorescent monoanion (1−) with a relatively low pKₐ value of 6.3 (Fig. 1c)8. Other unsuitable synthetic pH probes include the ratiometric seminaphthorofluor (SNARF) dyes9 that exhibit high background, as well as cyanine and rhodamine-based pH sensors that show the opposite pH sensitivity profile10,11.

We recently synthesized new derivatives of fluorescein (1) where the xanthene oxygen was replaced with a gem-dimethylcarbon moiety. This work resulted in "carbofluorescein" (CFI, 2, Fig. 1b)12, and the difluorinated derivative "Virginia Orange" (VO, 3, Fig. 1b)13. We discovered that this oxygen→carbon substitution elicited significant changes in photophysical and chemical properties of the fluorescein scaffold. Fl (1) exhibits λₑm/λᵦₘᵢₜ = 491 nm/510 nm at high pH, whereas CFI (2) and VO (3) are red-shifted with λₑm/λᵦₘᵢₜ = 544 nm/567 nm and 555 nm/581 nm, respectively. In addition to this bathochromic shift, the pH sensitivity of the dyes was markedly different. Fluorescein exhibits strong visible absorption at both pH 5.6 (vesicle pH) where the monoanion 1− dominates, and pH 7.4 (extracellular pH) where the dianion form 12− is prevalent—this can be observed by eye (Fig. 1c, d). In contrast, CFI (2) undergoes a cooperative transition between a highly colored dianion species (22−) and a colorless lactone form (2⁰-lactone; Fig. 1c). This is also evident visually as a solution of CFI (2) is colorless at pH 5.6, but shows robust visible absorption at pH 7.4 (Fig. 1d). Fluorescence-based titrations (Fig. 1e) gave pKₐ values of 6.3 and Hill coefficient (n_H) value of 0.97 for fluorescein (1), consistent with previous reports8. CFI (2) and VO (3) displayed pKₐ values of 7.5 and 6.7, and n_H values of 1.6 and 1.5, respectively. This cooperative transition likely stems from the altered lactone–quinoid equilibrium observed in the carbon-containing analogs of fluorescein and rhodamine dyes12. These pH sensitive red fluorophores have extinction coefficients and quantum yields similar to the pH sensitive red fluorescent proteins pHoran4 and pHuji5 (Supplementary Table 1). Moreover, we tested the resistance of these fluorophores to photobleaching (Supplementary Fig. 1a). While CFI showed similar photobleaching rate as Fl (τ = 5.33 s and τ = 5.35 s), VO was two to three times more photostable (τ = 14.54 s).

Fig. 3 Detection of synaptic vesicle exocytosis and recycling in hippocampal neurons. a Fluorescence images of hippocampal axons expressing VAMP2-SEP and VAMP2-SEP-CFI with the locations of vesicle fusion indicated with arrows (top); scale bar: 5 µm. Time-lapse images of the exocytosis events (bottom). Scale bar: 5 µm. b Average VAMP2-SEP (green) and VAMP2-SNAP-CFI (magenta) fluorescence signals in response to field stimulation for 10 s at 20 Hz. Fluorescence decay constant after stimulation was not different between the two probes (VAMP2-SEP 17.8 ± 1.5 s, VAMP2-SNAP-CFI 15.6 ± 1.0 s, p = 0.27, n = 25, Student’s t-test). c Fluorescence images of hippocampal axons expressing VAMP2-SEP (left) and VAMP2-SNAP-VO (right) with the insets indicated with arrows (top). Time-lapse of the exocytosis events (bottom). Scale bar: 5 µm. d Average VAMP2-SEP (green) and VAMP2-SNAP-VO (magenta) fluorescence signals in response to field stimulation for 10 s at 20 Hz. Fluorescence decay after stimulation was not different between the two probes (VAMP2-SEP 13.2 ± 1.7 s; VAMP2-SNAP-VO 13.0 ± 1.5 s, p = 0.51, n = 13, Student’s t-test). e Fluorescence images of hippocampal axons expressing VAMP2-SEP (left) and VAMP2-pHuji (right) with the insets indicated with arrows (top). Time-lapse of the exocytosis events (bottom). Scale bar: 5 µm. f Average VAMP2-SEP (green) and VAMP2-pHuji (magenta) fluorescence signals in response to field stimulation for 10 s at 20 Hz.
Therefore, the longer absorption and emission wavelengths, higher $pK_a$, resistance to photobleaching, and the cooperative colorless $\rightarrow$ colored transition upon increasing pH make both CFl and VO attractive scaffolds for building indicators to monitor synaptic vesicle fusion events.

To allow for specific labeling of expressed proteins, we prepared the SNAP-tag ligands attached to CFl (4) or VO (5) (Fig. 1f, Supplementary Fig. 1b). We tested the effects of protein conjugation on the properties of the dye by labeling SNAP-tag protein in vitro with CFl–SNAP-tag ligand 4 (Fig. 1g). We observed a shift in $pK_a$ to 7.3, and a decreased Hill coefficient ($\eta_H = 1.2$; Fig. 1h). The active site of the SNAP-tag enzyme is flanked with two Lys, one Arg, and one His (PDB structure 3KZZ, DOI: 10.2210/pdb3kzz/pdb). The resulting Coulombic interaction between these positively charged amino acid residues and the CFl label most likely explains the decrease in $pK_a$ upon conjugation. This polar surface might also stabilize the open form of the dye, resulting in the decreased cooperativity of the colored–colorless transition. Despite this lower $pK_a$ value and Hill coefficient, the fluorescence of the SNAP-tag-CFl conjugate is still completely suppressed at pH 5.6 (Fig. 1i).

**Fig. 4** Detection of synaptic vesicle exocytosis with labeling of endogenous proteins. 

- **a** Chemical structure of VO-NHS ester (6).
- **b** Protocol for staining with anti-Syt1-VO.
- **c** Fluorescence images of hippocampal axons expressing Syt1-SEP and stained with anti-Syt1-VO with the locations of vesicle fusion indicated with arrows (top); scale bar: 5 µm. Bottom, time-lapse images of the exocytosis events in boutons containing only anti-Syt1-VO (i) or containing both Syt1-SEP and anti-Syt1-VO (ii).
- **d** Average anti-Syt1-VO (magenta) and Syt1-SEP (green) fluorescence signals colocalized in the same boutons, and average anti-Syt1-VO fluorescence signals present in the complete neuronal population (orange) in response to field stimulation for 10 s at 20 Hz; $n = 41$.
- **e** Same as **d** for neurons transfected with VAMP2-SEP. Average anti-Syt1-VO (magenta) and VAMP2-SEP (green) fluorescence signals colocalized in the same boutons, and average anti-Syt1-VO fluorescence signals present in the complete neuronal population (orange); $n = 10$.
- **f** Average normalized traces corresponding to the recordings in **e**. Fluorescence decay after stimulation was $10.6 \pm 1.9$ s for VAMP2-SEP, $7.9 \pm 0.4$ s for anti-Syt1-VO co-localized with VAMP2-SEP, and $6.5 \pm 0.2$ s for all boutons ($n = 10$ fields). Data are represented as mean ± s.e.m.
Detection of single exocytosis events in PC12 cells. Next, we tested these SNAP-tag-based probes in living cells. Building on existing SEP-based constructs, we designed several SNAP-tag fusion proteins: (i) SNAP-tag inserted within an intra-luminal loop of the vesicular acetylcholine transporter VACHT (VACHT-SNAP), and glutamate transporter VGluT1 (VGluT1-SNAP), and (ii) SNAP-tag protein attached to the luminal C-terminal side of the vesicle protein VAMP2 (VAMP2-SNAP). We first expressed VACHT-SNAP and VAMP2-SNAP in neuroendocrine PC12 cells. VACHT is targeted to small synaptic-like vesicles (SSLV) while VAMP2 is found in both SSLV and large dense core vesicles. We found that the propensity of the CFI and VO fluorophores to adopt the neutral lactone form (Fig. 1c, d) allows for efficient intracellular labeling (Fig. 2a) with SNAP-tag ligands 4 or 5 without the use of other masking groups (e.g., acetate esters), which are typically required for fluorescent-based compounds. To monitor exocytosis, we depolarized cells with stimulation buffer containing high [K+] and imaged single small vesicles as they fused with the plasma membrane using total internal reflection fluorescence (TIRF) microscopy. Cells expressing VACHT constructs displayed events at high frequency. Events detected in cells co-expressing VACHT-SEP and VACHT-SNAP (labeled with CFI ligand 4) showed comparable fold increase in fluorescence at exocytosis (2.19 ± 0.07 vs. 2.40 ± 0.12, mean ± s.e.m.) with similar decay kinetics in both the green and red channels (Fig. 2b, c). We also compared VACHT-SEP to VACHT-SNAP-VO (Fig. 2d, e) and VACHT-pHuji (Fig. 2f, g). Like the semisynthetic indicator from CFI ligand 4, the VACHT-SNAP-VO derived from compound 5 also showed comparable performance to the SEP sensor (2.65 ± 0.10 vs. 2.60 ± 0.16-fold increase; Fig. 2d, e). However, in PC12 cells the RFP-based VACHT-pHuji sensor showed lower relative performance when compared with VACHT-SEP under the same conditions (2.01 ± 0.05 vs. 1.32 ± 0.02-fold increase; Fig. 2f, g) making events harder to detect with pHuji than with the other pH-sensitive proteins. We also observed individual fusion events using VAMP2-SEP or VAMP2-SNAP-CFI (Supplementary Fig. 2), albeit at low frequency, perhaps due to poor incorporation of this construct in PC12 cells.

Monitoring exocytosis and recycling of synaptic vesicles. We then tested these sensors in living neurons, focusing first on VAMP2-based constructs, which have been used extensively to follow SV exocytosis in neurons. We co-transfected hippocampal neurons with VAMP2-SEP and either VAMP2-pHuji or VAMP2-SNAP incubated with CFI ligand 4 or Virginia Orange ligand 5. For all the sensors, we observed a robust increase in fluorescence following electrical stimulation in fields covered with transfected axons, signaling SV exocytosis. The relative increase in fluorescence upon SV exocytosis was slightly higher for the SEP channel relative to the red-shifted fluorescent indicators, VAMP2-SNAP-CFI (Fig. 3a, b), VAMP2-SNAP-VO (Fig. 3c, d), and VAMP2-pHuji (Fig. 3e, f), which behaved similarly. The kinetics of decay, which tracks endocytosis and re-acidification of the vesicle, were similar for all four labels (Fig. 3b, d, f). We then tested whether the added CFI or VO could label the whole SV population efficiently. To do so, we normalized the fluorescence increase induced by stimulation with an application of a buffer containing 50 mM NH₄Cl, which quickly increases the intravesicular pH to extracellular pH. We found that the proportion of SVs undergoing exocytosis estimated with this method was similar for all three red fluorescent probes (SNAP-CFI, SNAP-VO, and pHuji) and similar to SEP (Supplementary Fig. 3a–f). We also tested whether the semisynthetic pH sensor system could be used in multicolor imaging experiments with GFP-based indicators. We co-transfected neurons with GCaMP6f and VAMP2-SNAP, which we labeled with CFI ligand 4. This allowed simultaneous imaging of both calcium ion transients and vesicle fusion in the same cell (Supplementary Fig. 3g–j). Finally, we tested the ability to monitor exocytosis by labeling another SV protein, VGluT1, which has been used previously with SEP to monitor SV exocytosis. The three probes (VGluT1-SNAP-CFI, VGluT1-SNAP-VO, and VGluT1-pHuji) were able to report SV exocytosis (Supplementary Fig. 3k–m).

Imaging of endogenous protein exo-/endocytosis cycling. One interesting feature of small organic fluorophores is their ability to label not only genetically encoded domains, such as SNAP-tag, but also ligands and antibodies and hence endogenous proteins. To enable imaging of endogenous vesicular proteins, we labeled a monoclonal antibody that recognizes a luminal epitope of synaptotagmin 1 (Syt1), a SV protein, with VOAc₂-NHS ester (Supplementary Fig. 1b) followed by mild deprotection of the acetate esters using hydroxylamine (Fig. 4a). This antibody has previously been used to detect endogenous Syt1 present on the plasma membrane after exocytosis in active synapses. To mark vesicular Syt1, we incubated neurons with this antibody-VO conjugate (10 nM) for 3 h in stimulation buffer, followed by extensive washing to remove the unbound antibodies (Fig. 4b). The antibody labeling was done in neurons transfected with Syt1-SEP or VAMP2-SEP to compare the performance of this labeling technique with the overexpressed, genetically encoded GFP-based pH sensors. Electrical stimulation evoked a robust increase in fluorescence in axons transfected with Syt1-SEP (Fig. 4c, d) or VAMP2-SEP (Fig. 4e), and in axons of untransfected neurons without detectable SEP. We found that the decay kinetics after stimulation was faster for the VO-antibody conjugate (8.8 ± 0.5 s) than the Syt1-SEP (12.2 ± 0.9 s, n = 41; paired t-test p < 0.0001, Fig. 4d) and VAMP2-SEP (Fig. 4f), suggesting a difference in overexpressed vs. endogenous protein behavior after SV exocytosis. Remarkably, the fluorescence transients were substantially higher in untransfected than in transfected neurons (Fig. 4d, e), perhaps stemming from steric hindrance of the overexpressed SV proteins or through quenching of the two fluorophores.

Discussion

We have developed new “semisynthetic” pH-sensitive proteins that allow for the imaging of synaptic vesicle fusion events in living cells. This sensor system combines the highly tunable properties of small-molecule fluorophores with the specificity of self-labeling tags or antibodies. The SNAP-tag-based system constitutes the first genetically encoded long-wavelength pH sensor with similar or better performance than SEP in different cell types. The antibody-based pH sensor allows for imaging of vesicle fusion events without the need for overexpression of sensor proteins. Addition of other self-labeling or epitope tags by genome editing methods could allow cell- and protein-specific labeling without the need for overexpression. Future improvements of both the protein and the dye within this semi-synthetic scaffold should further enable imaging of this key biological process in increasingly complex systems.

Methods

General organic synthesis methods. Commercial reagents were obtained from reputable suppliers and used as received. All solvents were purchased in septum-sealed bottles stored under an inert atmosphere. All reactions were sealed with septa through which a nitrogen atmosphere was introduced unless otherwise noted. Reactions were conducted in round-bottomed flasks or septum-capped crimp-top vials containing Teflon-coated magnetic stir bars. Heating of reactions was accomplished with a silicon oil bath or an aluminum reaction block on top of a stirring hotplate equipped with an electronic contact thermometer to maintain the indicated temperatures.
Reactions were monitored by thin layer chromatography (TLC) on precoated TLC glass plates (silica gel 60 F254, 250 μm thickness) or by LC/MS (Phenomenex Kinetic 2.1 mm x 30 mm 2.6 μm C18 column; 5 μL injection; 5–98% MeCN/H2O, linear gradient, with constant 0.1% v/v HCO2H additive; 6 min run; 0.5 mL/min flow; ESI; positive ion mode). TLC chromatograms were visualized by UV illumination or developed with anisaldehyde, ceric ammonium molybdate, or KMnO4 stain. Reactions were purified by flash chromatography on an automated purification system using pre-packed silica gel columns or by preparative HPLC (Phenomenex Gemini–MXT 30 x 150 mm 5 μm C18 column). Analytical HPLC analysis was performed with an Agilent Eclipse XDB 4.6 x 150 mm 5 μm C18 column under the indicated conditions. High-resolution mass spectra were acquired at the Mass Spectrometry Facility in the Department of Medicinal Chemistry at the University of Washington and the High Resolution Mass Spectrometry Facility at the University of Iowa.

NMR spectra were recorded on a 400 MHz spectrometer. 1H and 13C chemical shifts (δ) were referenced to CDCl3 (7.26 ppm) for 1H spectra and CD3OD (49.0 ppm) for 13C NMR spectra. Data for 1H NMR spectra are reported as follows: chemical shift (δ ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, m = multiplet), coupling constant (Hz), and integration. Data for 13C NMR spectra are reported by chemical shift (δ ppm) with multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, dq = doublet of quartets, dq = doublet of doublet of quartets). Hydrogen coupling constants are given in Hz.

Synthesis of TBS2-CFl-6-CO2H (10). A vial was charged with di-tert-butyl 2-bromopentanoate (9; 1.48 g, 4.14 mmol, 2 eq), sealed, and flushed with nitrogen. After dissolving the bromide in THF (7 mL) and cooling the reaction to −15 °C, iPrMgCl2Li (1.3 M in THF, 3.19 mL, 4.14 mmol, 2 eq) was added. The reaction was warmed to −10 °C and stirred for 10 h. A solution of 3-bis-(tert-butyldimethylsilyl)-oxy)-10-10-dimethylnaphthalen-9(10-H)one (11; 1.7 g, 2.07 mmol) in THF (4 mL) was then added dropwise. The reaction mixture was warmed to room temperature and stirred for 2 h. It was subsequently quenched with saturated NH4Cl, diluted with water, and extracted with EtOAc (2x). Combined organics were washed with brine, dried over anhydrous MgSO4, filtered, and evaporated. Silica gel chromatography (0–10% EtOAc/hexanes, linear gradient) provided 245 mg (17%) of 10 as a colorless solid. 1H NMR (CDCl3, 400 MHz) δ 8.16 (dd, J = 1.8, 3.1 Hz, 1H), 8.02 (dd, J = 0.8, 0.6 Hz, 1H), 7.67–7.59 (m, 1H), 7.09–7.05 (m, 2H), 6.64–6.57 (m, 4H), 1.81 (s, 3H), 1.72 (s, 3H), 1.34 (s, 3H), 0.99 (s, 18H). 13C NMR (CDCl3, 100 MHz) δ 164.5 (CH), 155.5 (C), 147.0 (C), 138.1 (C), 130.3 (CH), 129.7 (C), 129.3 (CH), 125.1 (CH), 125.0 (CH), 124.0 (CH), 119.2 (CH), 117.8 (CH), 87.0 (C), 82.5 (C), 38.2 (C), 35.0 (CH3), 33.2 (CH2), 28.2 (C), 25.8 (C), 18.4 (C), 14.7 (C), 4.19 (C); HRMS (ESI) calc’d for C28H40F2O5S [M + H]+ 567.3357, found 567.3352.

Synthesis of TBS2-VO-6-CO2H (11). A vial was charged with di-tert-butyl 2-bromopentanoate (9; 1.03 g, 2.89 mmol, 1.5 eq), sealed, and flushed with nitrogen. After dissolving the bromide in THF (5 mL) and cooling the reaction to −55 °C, VO(15) in THF (2.25 mL, 2.25 mmol, 1.5 eq) was added. The reaction was stirred at −40 °C for 4 h. A solution of 3-bis-(tert-butyldimethylsilyl)-oxy)-2,7-difuoro-10,10-dimethylnaphthalen-9(10-H)one (12; 1.00 g, 1.93 mmol) in THF (5 mL) was then added dropwise. The reaction mixture was warmed to room temperature and stirred for 2 h. It was subsequently quenched with saturated NH4Cl, diluted with water, and extracted with EtOAc (2x). The combined organics were washed with brine, dried over anhydrous MgSO4, filtered, and evaporated. Silica gel chromatography (0–10% EtOAc/hexanes, linear gradient) provided 439 mg (31%) of 11 as a white solid. 1H NMR (CDCl3, 400 MHz) δ 8.20 (dd, J = 0.8, 1.3 Hz, 1H), 8.05 (dd, J = 0.8, 0.6 Hz, 1H), 7.65–7.60 (m, 1H), 7.12 (d, J = 8.3 Hz, 2H), 6.36 (d, J = 11.3 Hz, 2H), 1.77 (s, 3H), 1.68 (s, 3H), 1.56 (s, 3H), 1.01 (s, 18H), 0.21 (s, 12H); 13C NMR (CDCl3, 100 MHz) δ 169.9 (CH), 163.5 (C), 155.5 (CH), 147.0 (C), 138.1 (C), 130.3 (CH), 129.7 (C), 129.3 (CH), 125.1 (CH), 125.0 (CH), 124.0 (CH), 119.2 (CH), 117.8 (CH), 87.0 (C), 82.5 (C), 38.2 (C), 35.0 (CH3), 33.2 (CH2), 28.2 (C), 25.8 (C), 18.4 (C), 14.7 (C), 4.19 (C); HRMS (ESI) calc’d for C28H40F2O5S [M + H]+ 567.3357, found 567.3352.
2\textsubscript{jL} = 19.7 Hz, CH\textsubscript{3}), 37.8 (C), 33.9 (CH\textsubscript{2}), 33.4 (CH\textsubscript{3}), 20.2 (CH\textsubscript{3}); HRMS (ESI) calcd for C\textsubscript{28}H\textsubscript{19}F\textsubscript{2}O\textsubscript{8} [M + H]+ 620.1363, found 620.1370.

Bulk aqueous solutions were autoclaved at 150 °C for 30 minutes. The solution was concentrated and purified using a SepPak C\textsubscript{18} cartridge (Waters) eluted with 100% water and then dried over anhydrous MgSO\textsubscript{4}, filtered through a 0.22 mm PTFE syringe filter, diluted with 0.1% TFA, and lyophilized.

UV-vis and fluorescence spectroscopy. Spectroscopy was performed using 1-cm path length, 3.5-ml quartz cuvettes or 100-ml quartz microcuvettes. All measurements were taken at ambient temperature (~22 °C). Absorption spectra were recorded on a Cary Model 100 spectrometer (Varian), and fluorescence spectra were recorded on a Cary Eclipse fluorimeter (Varian). The pK\textsubscript{a} values for compounds 1, 2, and 3 were determined in buffers containing 150 mM NaCl and 10 mM buffer. The following buffer systems were used: (pH 4.0–6.8) glycine/tris (pH 7.8–9.0) carbonate (pH 9.2–10.8). Fluorescence values were read on 500 mM samples (n = 3) and fitted to a sigmoidal dose response curve (variable slope) using GraphPad Prism software. Samples for visual inspection contained 5 mM 1 or 2 in citrate (pH 5.6) or phosphate (pH 7.4) buffer.

Measurement of phosphoros bleaching. Solutions of 1 mM fluorescein (1), carboxyfluorescein (2), or Virginia Orange (3) were prepared in 50 mM sodium borate, pH 9.7. To create aqueous microdroplets, a sample of these solutions was added to 1-octanol in 1:9 ratio and this mixture was vortexed briefly. The resulting emulsion was placed on a glass slide and slide with a coverslip. Fluorophor bleaching was accomplished by illuminating an entire isolated microdroplet of aqueous dye solution using an upright microscope (Zeiss Axio Observer Z1) and a 20× NA 0.8 objective. Fluorescein (1) was bleached under 488 nm (Sapphire 488, Coherent) laser excitation at 4.3 mW power (Intensity 3.25 W/cm\textsuperscript{2}) and the emission was collected using a 550BP38 filter. Carboxyfluorescein (2) and Virginia Orange (3) fluorescence were measured using a mercury lamp (Xite, Series 120-Q) with 550BP38 filter and the emission intensity 4.76 W/cm\textsuperscript{2}; emission was collected using 625BP90 filter. Fluorescence was detected by a fiber coupled Avalanche photodiode (SPDM-AQRH14, Pacer).

Protein chemistry. SNAP-tag protein (NEB) was labeled with excess CFI SNAP-tag ligand (4) in PBS containing 1 mM DTT. The protein concentration was purified and concentrated using a Bio spin desalting column (7 K MWCO; Thermofisher). The protein sample was analyzed by gel electrophoresis using a NuPAGE 4–12% Bis-Tris polyacrylamide gel (Thermofisher) and imaged using a Typhoon Trio + scanner (GE Healthcare); the gel was also stained with Coomassie Plus (ThermoFisher) and compared to a Mark 12 protein standard ladder (ThermoFisher). Absorbance measurements were taken in citrate (pH 5.6) or phosphate (pH 7.4) buffers described above.

Plasmid constructs. VAMP2-SEP and synaptotagmin1-SEP were kindly gifts from Jörn Kluenga. GCaMP6f was obtained from Addgene (#40755). Vcath-SNAP and Vcath-T-Huji were created by replacing the open reading frame of SEP for either SNAP-tag (New England Biolabs) or pHuji5 in the VACHT-SEP construct15. VGlut1-SNAP and VGlut1-pHuji were created by replacing the open reading frame of SEP for either SNAP-tag (New England Biolabs) or pHuji5 in the VGlut1-SEP construct17. Vp2-huji was created by swapping the SEP gene for pHuji5 in VAMP2-SEP sensor construct2.

Cell culture and transfections. PC12 cells (obtained from Dr. Rae Nishi, The Molecular Biology Labs) were grown in DMEM containing 40 mM L-glutamine, 1 mM sodium pyruvate, 100 units/ml penicillin, 100 units/ml streptomycin, at 37 °C in 5% CO\textsubscript{2}. Cells were plated onto 20-mm round poly-D-lysine-coated glass cover-slips and transfected ~24 h later with 1 μg of plasmid DNA and the appropriate concentrations of each reporter construct using calcium phosphate transfection procedure22. Cells were imaged 24 h after transfection.

Antibody labeling. Mouse monoclonal antibodies against the luminal part of Synaptotagmin 1 (100 μg, Synaptic Systems 503 105 1) were coupled to calcium buffer (pH 9.3 to 3.4 eq VO\textsubscript{2+}-NHS ester (6); solubilized in DMSO) for 1 h. VO\textsubscript{2+} was then added and incubated with hydrazine for 48 h. VO\textsubscript{2+} was purified by size exclusion chromatography on a Superdex 2000/10 column (GE Healthcare) equilibrated with PBS; final concentration of VO\textsubscript{2+} was determined on a size exclusion column and eluted in PBS at a final concentration of 0.1 μM. The degree of labeling was estimated to be 6.8% per antibody by absorbance spectroscopy.

Cell staining with CFI and VO ligands. PC12 cells were washed with Fluorobrite media (Fluorobrite DMEM (Invitrogen) supplemented with 5% fetal bovine serum, 3% horse serum, 4 mM L-glutamine, 1% prorovate, and 1% penicillin/streptomycin), and then incubated with 6 μM CFI ligand (4) or VO ligand (5) in Fluorobrite media for 3 h at 37 °C. Cells were then washed thoroughly, incubated in media for an additional 2 h before imaging. SVs were labeled with...
CFl-SNAP-tag ligand (4), VO-SNAP-tag ligand (5), and anti-Syt1-VO antibody. Neurons were exposed to 10 µM CFl-SNAP-tag ligand (4) or VO-SNAP-tag ligand (5) for 2 h in conditioned culture medium, washed thoroughly and then placed back in conditioned culture medium for at least 30 min before imaging. Neurons were incubated with 10 nM anti-Syt1-VO in a 37 °C incubator for 3 h in a carbonate buffer containing 105 mM NaCl, 20 mM KCl, 2.5 mM CaCl₂, 1 mM MgCl₂, 10 mM glucose, 18 mM NaHCO₃. Cells were then washed three times before imaging.

Live cell imaging and analysis of PC12 cells. Cells were imaged in buffer containing 130 mM NaCl, 2.8 mM KCl, 5 mM CaCl₂, 1 mM MgCl₂, 10 mM HEPES and 10 mM glucose. pH was adjusted to 7.4 with 1 N NaOH. The stimulation buffer contained 50 mM NaCl, 105 mM KCl, 5 mM CaCl₂, 1 mM MgCl₂, 10 mM HEPES and 1 mM NaH₂PO₄, pH was adjusted to 7.4 with 5 M KOH. Experiments were carried out at 25 °C using TIRF microscopy.14, 23. Before experiments, 100 nm yellow fluorescent beads (Invitrogen) were imaged in the green and red channels, and superimposed by mapping the coordinates of the brightest pixel bead positions.

Live cell imaging and analysis of hippocampal neurons. The experiments with neurons were carried out in a buffer solution containing 120 mM NaCl, 5 mM KCl, 2 mM CaCl₂, 2 mM MgCl₂, 5 mM glucose, 10 mM HEPES adjusted to pH 7.4 and 270 mM Osmol/l. Neurons were stimulated by electric field stimulation (platinum electrodes, 10 mm spacing, 1 ms pulses of 50 µA, and alternating polarity at 20 Hz) applied by constant current stimulus isolator (SIU-102, Warner Instruments) in the presence of 10 µM 6-cyano-7-nitroquinoxaline-2,3-dione (CNQX) and 50 µM D,L-2-amino-5-phosphonovaleric acid (AP5) to prevent recurrent activity.

Experiments were performed on an inverted microscope (IX83, Olympus) equipped with an Apochromat N oil ×100 objective (NA 1.49). Images were acquired with an electron multiplying charge coupled device camera (QuantEM:512SC: Roper Scientific) controlled by MetaVue7.1 (Roper Scientific). Samples were illuminated by a 473 nm laser (Cobolt) for green imaging, as well as by a coaligned 561 nm laser (Cobolt) for red imaging. Emitted fluorescence was detected after passing filters (Chroma Technology Corp.): 595/50 nm for pHui, CFI and VO imaging, and 525/50 nm for SEP/GFP imaging. Simultaneous dual color imaging was achieved using a DualView beam splitter (Roper Scientific). To correct for x/y distortions between the two channels, images of fluorescently labeled beads (Tetraspeck, 0.2 µm, Invitrogen) were taken before each experiment and used to align the two channels. Time lapse images were acquired at 1 or 2 Hz with integration times from 50 to 150 ms.

Image analysis was performed using Metamorph (Molecular Devices) and custom scripts on MATLAB (Mathworks). The co-ordinates of the brightest pixel were imaged in the green and red channels. and green images were aligned post acquisition using projective image transformation14, 23. Before experiments, 100 nm yellow-green fluorescent beads (Invitrogen) were imaged in the green and red channels, and superimposed by mapping bead positions.

Image analysis was performed using Metamorph (Molecular Devices) and custom scripts on MATLAB (Mathworks). The co-ordinates of the brightest pixel in the first frame of each fusion event in the green channel was identified by eye, and time was normalized to 0 s. A circular ROI of 6 pixels (~0.99 mm) diameter and a square of 21 pixels (~3.5 µm) were drawn around the fusion co-ordinates. The average minimum pixel intensity in the surrounding square from five frames before fusion was subtracted from the intensity in the circular ROI, and the values were normalized to the frame before fusion in the green and red channels.

References
1. Kavalali, E. T. & Jorgensen, E. M. Visualizing presynaptic function. Nat. Neurosci. 17, 10–16 (2013).
2. Sankaranarayanan, S., De Angelis, D., Rothman, J. E. & Ryan, T. A. The use of pHluorins for optical measurements of presynaptic activity. Biophys. J. 79, 2199–2208 (2000).
3. Li, H. et al. Concurrent imaging of synaptic vesicle recycling and calcium dynamics. Front. Mol. Neurosci. 4, 34 (2011).
4. Li, Y. & Tsien, R. W. pHTomato, a red, genetically encoded indicator that enables multiplex interrogation of synaptic activity. Nat. Neurosci. 15, 1047–1053 (2012).
5. Shen, Y., Rosendale, M., Campbell, R. E. & Perrais, D. Phospho-sensitive red fluorescent protein for imaging of exo- and endocytosis. J. Cell. Biol. 207, 419–432 (2014).
6. Soykan, T., Maritzen, T. & Haucke, V. Modes and mechanisms of synaptic vesicle recycling. Curr. Opin. Neurobiol. 39, 17–23 (2016).
7. Mollwitz, B. et al. Directed evolution of the suicide protein O6-alkylguanine-DNA alkyltransferase for increased reactivity results in an alkylated protein with exceptional stability. Biochemistry 51, 986–994 (2012).
8. Matteoli, M., Takei, K., Perin, M. S., Südhof, T. C. & Camilli, P. D. Traf1 is required for vesicle recycling and subsequent exocytosis. Nat. Commun. 7, 833–839 (2016).
9. Grimm, J. B. et al. Carbofluorescins and carboxorhodamines as scaffolds for high-contrast fluorogenic probes. ACS Chem. Biol. 8, 1303–1310 (2013).
10. Shirota, T., Kyotani, Y., Shiozaki, K., Fujii, T. & Nishi, K. New pHosFRET probes with exceptional stability. Biotechniques 47, 769–774 (2009).
11. Asanuma, D. et al. Acidic-pH-activatable fluorescence probes for visualizing exocytosis dynamics. Angew. Chem. Int. Ed. 53, 6085–6089 (2014).
12. Hua, Y. et al. A readily retrievable pool of synaptic vesicles. Nat. Neurosci. 14, 833–839 (2011).
13. Grimm, J. B. et al. gi and carboxorhodamines as scaffolds for high-contrast fluorogenic probes. ACS Chem. Biol. 8, 1303–1310 (2013).
14. Grimm, J. B., Gruber, T. D., Ortiz, G., Brown, T. A. & Lavis, L. D. Virginia Orange: a versatile, red-shifted fluorochrome scaffold for single- and dual-input fluorogenic probes. Bioconjug. Chem. 27, 474–480 (2016).
15. Schackh, K. A. et al. Imaging the post-fusion release and capture of a vesicle membrane protein. Nat. Commun. 3, 1154 (2012).
16. Brauchi, S., Krapivinsky, G., Krapivinsky, L. & Clapham, D. E. TRPM7 facilitates cholinergic vesicle fusion with the plasma membrane. Proc. Natl. Acad. Sci. USA 105, 8304–8308 (2008).
17. Chen, T.-W. et al. Ultrasensitive fluorescent probes for imaging neuronal activity. Nature 499, 295–300 (2013).
18. Voglmaier, S. M. et al. Distinct endocytic pathways control the rate and extent of synaptic vesicle protein recycling. Neuron 51, 71–84 (2006).
19. Matteoli, M., Takei, K., Perin, M. S., Südhof, T. C. & Camilli, P. D. EXO-endocytic recycling of synaptic vesicles in developing processes of cultured hippocampal neurons. J. Cell. Biol. 117, 849–861 (1992).
20. Opatz, F. et al. Limited intermixing of synaptic vesicle components upon vesicle recycling. Traffic 11, 808–812 (2010).
21. Mikuni, T., Nishiyama, J., Sun, Y., Kamasawa, N. & Yasuda, R. High-throughput, high-resolution mapping of protein localization in mammalian brain by in vivo genome editing. Cell 165, 1803–1817 (2016).
22. Kaeche, S. & Banker, G. Culturing hippocampal neurons. Nat. Protoc. 1, 2406–2415 (2006).
23. Wienschi, M. & Klingauf, J. Vesicular proteins exocytosed and subsequently retrieved by compensatory endocytosis are nonidentical. Nat. Neurosci. 9, 1019–1027 (2006).
24. Trelles, A. J., Schackh, K. A. & Taraska, J. W. Imaging the recruitment and loss of proteins and lipids at single sites of calcium-activated exocytosis. Mol. Biol. Cell. 27, 2423–2434 (2016).

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Author contributions
M.M. performed and analyzed experiments on neurons. A.S. performed experiments on PC12 cells, and A.S. and J.W.T. analyzed the data. J.B.G. performed organic synthesis. T.D.G. prepared and analyzed protein conjugates. L.D.L. performed spectroscopy. M.M.,
L.D.L., J.W.T. and D.P. wrote the manuscript and all the other authors edited the manuscript.

**Additional information**

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**Competing interests:** L.D.L. and J.B.G. have filed patent applications whose value might be affected by this publication. The remaining authors declare no competing financial interests.

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