Control of resistance against bacteriophage killing by a metabolic regulator in meningitis-associated Escherichia coli

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Ecoologically beneficial traits in bacteria are encoded by intrinsic and horizontally acquired genes. However, such traits are not universal, and the highly mosaic nature of bacterial genomes requires control at the transcriptional level to drive these processes. It has emerged that regulatory flexibility is widespread in the Escherichia coli species, whereby preexisting transcription factors can acquire new and unrelated roles in regulating beneficial traits. DsdC is the regulator of D-serine tolerance in E. coli, is essential for D-serine catabolism, and is often encoded by two copies in neonatal meningitis-associated E. coli (NMEC). Here, we reveal that DsdC is a global regulator of transcription in NMEC and does not require D-serine for the control of novel beneficial traits. We show that DsdC binds the chromosome in an unusual manner, with many binding sites arranged in clusters spanning entire operons and within gene coding sequences, such as neuO. Importantly, we identify neuO as the most significantly down-regulated gene in a strain deleted for both dsdC copies, in both the presence and absence of D-serine. NeuO is prophage encoded in several NMEC K1 isolates and mediates capsule O-acetylation but has no effect on attachment to or invasion of human brain endothelial cells. Instead, we demonstrate that NeuO provides resistance against K1 bacteriophage attack and that this critical function is regulated by DsdC. This work highlights how a horizontally acquired enzyme that functions in cell-surface modulation can be controlled by an intrinsic regulator to provide a key ecological benefit to an E. coli pathotype.

Significance

Bacterial genomes are often subject to extensive horizontal gene transfer. Genes must be controlled at several levels of regulation to ensure their encoding functions are expressed appropriately and offer beneficial traits. Here, we reveal that an Escherichia coli transcription factor, historically linked to a narrow-spectrum metabolic pathway, interacts with the chromosome globally and by binding to DNA in an unusual fashion. These interactions affect regulation of several distinct processes, including biogenesis and modification of capsular polysaccharide. Importantly, the latter function occurs independently of the canonical metabolic inducer of this regulator and results in enhanced resistance to killing by bacteriophages that target the capsule. This work highlights that specific regulators can be reprogrammed to perform far-removed, yet critical, functions in bacteria.
showed high conservation, with 98% homology across 98% of the coding sequence. There were four amino acid substitutions occurring in a cluster within the substrate binding domain, as well as five substitutions in the C-terminal coding sequence (SI Appendix, Fig. S1). To test the functionality of NMEC DsdC in D-ser catabolism, we deleted both dsdC1 and dsdC2 individually and together in the same background (ΔdsdC1Δ2). Growth of both ΔdsdC1 and ΔdsdC2 on MOPS minimal medium plates containing 10 mM D-ser as a sole carbon source was comparable to that of the wild type (WT) (Fig. 1B). This result was mirrored in an experiment using a dsdXA promoter–green fluorescent protein fusion as a reporter for activation of D-ser catabolism. WT NMEC, ΔdsdC1, and ΔdsdC2 displayed identical activation levels of dsdXA transcription in response to D-ser, indicating equal functionality of both regulator copies (Fig. 1C). Conversely, a ΔdsdC1Δ2 double mutant was unable to grow on D-ser as a sole carbon source (Fig. 1B), but expression of either dsdC1 or dsdC2 in trans was able to fully restore the growth defect of ΔdsdC1Δ2. Collectively, these data indicate that DsdC1 and DsdC2 have functional redundancy in the context of D-ser catabolism.

DsdC1 and DsdC2 Bind Globally to the NMEC Chromosome in Two Distinct Patterns. E. coli TFs often adapt their functionality to regulate diverse gene sets beyond their primary role (2). However, it was unknown if a role for DsdC existed beyond D-ser catabolism in NMEC, which formed a key question of this study. We first validated the ability of both DsdC1 and DsdC2 to directly bind their respective dsdCX promoter regions in vitro. Electrophoretic mobility shift assays (EMSAs) using purified DsdC1 and DsdC2 confirmed that both TF copies were able to bind to their respective promoter regions, but not a control fragment of the kanamycin gene (SI Appendix, Fig. S2A). To increase the resolution of this interaction, DNase I footprinting revealed a region of protection within the dsdC and dsdX4 intergenic regions for both copies of DsdC (SI Appendix, Fig. S2B). The protected site was 47 bp in length and is identical in sequence between both the dsdC1 and dsdC2 promoter regions (SI Appendix, Fig. S2C). LysR-type transcriptional regulators typically bind as dimers to two 13-bp sites in close proximity upstream of regulated genes, which may explain the size of this protected region (13). Addition of D-ser to the reaction resulted in a reduction of binding to the latter protected site, suggesting D-ser can influence the strength of DsdC binding to target promoters. These data confirmed that both copies of DsdC directly and specifically bind to target DNA, regulating their respective dsdCX4 loci in a similar fashion in NMEC.

Given the broad transcriptional responses of distinct E. coli pathotypes to D-ser and the fact that NMEC encodes two functional copies of DsdC, we hypothesized that this TF may play a wider role in gene regulation (16, 18). To address this, we used chromatin immunoprecipitation sequencing (ChIP-seq) to map all binding sites of DsdC1 and DsdC2 along the NMEC genome in vivo. We first genetically engineered two NMEC strains, where either DsdC1 or DsdC2 was tagged with a 3X FLAG epitope at the C terminus (ΔdsdC1Δ2FLAG and ΔdsdC2Δ1FLAG). These strains were then grown in M9 minimal media spiked with 1 mM D-ser at mid exponential phase (hours 3–5 in the growth curve) to assess if the presence of the FLAG-tag fusions had an impact on gross physiology. Growth was identical to that of the WT under these conditions (SI Appendix, Fig. S3A and B). In parallel, we analyzed samples from these growth assays by whole-cell Western blot to confirm that the DsdC1Δ1FLAG and DsdC2Δ2FLAG fusions were indeed being expressed. Note that addition of D-ser only resulted in a modest increase in DsdC1Δ1FLAG and DsdC2Δ2FLAG expression (SI Appendix, Fig. S3C). ChIP-seq analysis of these
strains, grown in the presence and absence of a D-ser spike-in, revealed an unexpected pattern of DsdC binding on a genomewide scale (Fig. 2A). DsdC1FLAG and DsdC2FLAG bound to 95 and 140 sites, respectively, in the absence of D-ser, and 217 and 177 sites, respectively, in the presence of D-ser (P ≤ 0.01) (SI Appendix; Tables S1–S4). This suggests that D-ser likely influences which genes are targeted by DsdC, but also that DsdC may act as a global regulator of diverse NMEC processes, regardless of whether D-ser is present or not. To validate the interactions, we tested binding of DsdC to one of the novel identified regions (the waaV promoter region) by EMSA, confirming the interaction in vitro (SI Appendix; Fig. S4). A key question from this global analysis was whether or not DsdC1 or DsdC2 were capable of regulating unique gene sets. Importantly, closer inspection of the read-mapping data revealed that the DsdC1/2-specific ChIP-seq peaks identified in the peak calling pipeline were indeed present in both strains but that some peaks did not meet statistical cutoff during the data processing. This indicated that the function of both DsdC copies is likely redundant, and therefore, for the remainder of the study, we focused on regulation by DsdC1 and DsdC2 as a common process. Gene ontology (GO) analysis of open reading frames located near DsdC binding sites revealed a diverse functional range of potential targets for DsdC regulation (SI Appendix; Fig. S5A). The main functional groups of genes associated with DsdC binding sites were metabolism, transport and membrane associated, regulation, and virulence associated. The latter group is of potentially critical importance for understanding NMEC virulence regulation and included genes involved in lipopolysaccharide (LPS) and capsule biosynthesis, fimbral adhesion, and a putative T3SS (SI Appendix; Fig. S5B) (18, 19, 20). This result suggested that DsdC is potentially a key regulator of many cellular processes in NMEC and not exclusively a regulator of D-ser catabolism.

During the analysis, we noticed that only some binding sites corresponded to the characteristic peak shape typically expected of TF binding events (an intergenic peak upstream of a gene 5′ end with a bimodal read distribution) (21). This canonical TF binding is exemplified by DsdC at the dsaA promoter region, which displays a discrete peak upstream of the target genes (Fig. 2B). Analysis of the peak location with respect to the nearest 5′ gene boundary for all DsdC binding sites revealed that ~50% of these sites were located close to genes, with the remainder located much farther up- or downstream of genes (Fig. 2C). By interrogating the distribution of signal enrichment at the latter regions, we observed an unusual pattern of DsdC binding defined by clusters of intergenic and intragenic peaks appearing in close succession and spanning entire operons (Fig. 2D). Importantly, these regions include operons encoding genes for key NMEC virulence–associated processes such as O-antigen, core LPS, and K1 capsular polysaccharide biosynthesis. Collectively, these data reveal that DsdC is a global regulator in NMEC, encoded by two functionally redundant copies, and binds target genes in two highly distinct manners.

### DsdC Regulates the Expression of Capsular and LPS Biosynthesis Genes

To determine if the unusual global binding pattern of DsdC identified by ChIP-seq resulted in functional changes to NMEC gene expression, we used comparative RNA-seq analysis. The experimental design consisted of three pairwise transcriptomic comparisons (SI Appendix; Fig. S6) to determine DsdC-associated differentially expressed genes (DEGs; false-discovery rate P ≤ 0.05 and absolute fold change ≥1.5). We began by reanalyzing previously published data from our group (that documented the response of NMEC and ΔdsdC1/2 to D-ser) comparing WT NMEC with ΔdsdC1/2 grown in M9 minimal media alone to identify DsdC-regulated genes in the absence of D-ser (16). Next, we tested whether D-ser could alter the regulatory function of DsdC by comparing RNA from WT NMEC and ΔdsdC1/2 cells exposed to D-ser during the exponential phase of growth (culture in M9 for 3 h followed by addition of 1 mM D-ser for 2 h). Eighty-two significant DEGs were identified comparing the WT and ΔdsdC1/2 grown without D-ser, whereas in the presence of D-ser, the ΔdsdC1/2 mutant displayed 436 DEGs (Fig. 3A and SI Appendix; Tables S5 and S6). Because the cellular accumulation of D-ser in the ΔdsdC1/2 background would be expected to cause shifts in global gene expression associated with D-ser toxicity and not necessarily a lack of DsdC regulatory effects, a third comparison was performed to mitigate this (8). This comprised WT NMEC and ΔdsdC1/2 complemented with a plasmid expressing DsaA in trans (pDsdA) under the same conditions of D-ser exposure. The logic behind this experimental design was to determine the DEGs associated with DsdC that were dependent on the presence of D-ser (i.e., sensing) but, in principle, not caused by the toxic effects of D-ser accumulation in ΔdsdC1/2 because of the expression of D-ser deaminase. This
comparison revealed 552 DEGs in the ΔdsdCΔ2 pDsdA strain after D-ser exposure (SI Appendix, Table S7). To narrow down the specificity of DEGs to DsdC and not the effects of D-ser accumulation, we identified 30 common DEGs regulated in the same direction (i.e., displayed up- or down-regulation) in the mutant and complement comparisons (Fig. 3B). This allowed us to identify a core set of DEGs that were specific to DsdC, regardless of whether D-ser was catabolized or accumulated within cells (SI Appendix, Table S8). The most striking result from this analysis was the identification of newO, encoding the O-acetyltransferase of the K1 capsule, as the most significantly down-regulated gene in all three comparisons (−7.9-fold, \( P = 1.66 \times 10^{-20} \); −4.57-fold, \( P = 8.56 \times 10^{-16} \); and −8.15-fold, \( P = 5.07 \times 10^{-71} \), respectively) (17). This finding suggests that there exist key DsdC-regulated genes in NMEC that are not dependent on a physiological response to D-ser.

We next combined functional grouping of DEGs with the ChIP-seq data set to identify potentially important processes regulated by DsdC. GO analysis of the common DsdC-associated DEGs in response to D-ser revealed that more than half of these were involved in capsular (7/30) and LPS (9/30) biosynthesis (Fig. 3C). Given the large number of DEGs identified by RNA-seq, we decided to focus on genes directly regulated by DsdC, irrespective of D-ser exposure, because these likely represent adapted functions that may be specific to NMEC biology. Intriguingly, there was very little overlap between the ChIP-seq and
RNA-seq data sets. Indeed, in the absence of D-ser, only three genes were found to show differential expression in the \( \Delta \text{dsdC1/2} \) mutant relative to WT NMEC in the absence of D-ser (Fig. 3A; Supplementary Table S1). From the 30 common DEGs that overlapped between \( \Delta \text{dsdC1/2} \) and \( \Delta \text{dsdC1/2} \) plus pDsDA strains in response to D-ser, thirty of these overlapping genes showed the same direction (either up- or down-regulated) of regulation. (C) GO analysis of DsdC-specific DEGs that were identified as being related to D-ser accumulation (\( \Delta \text{dsdC1/2} \)) or dependent on D-ser exposure (\( \Delta \text{dsdC1/2} \) and \( \Delta \text{dsdC1/2} \) plus pDsDA).

**O-acetyltransferase NeuO Mediates Protection Against K1 Bacteriocidal Attack.** The combined transcriptomic analyses revealed that neuO was the most significantly down-regulated gene as a result of \( \Delta \text{dsdC1} \) and \( \Delta \text{dsdC2} \) deletion, regardless of exposure to D-ser. NeuO is an O-acetyltransferase responsible for O-acetylation of the K1 capsule and is regulated by a spontaneous phase-variation mechanism, involving slipped-strand mispairing of a heptanucleotide repeat region within the gene termed polyP (17). Unusually, ChIP-seq identified highly significant, noncanonical binding of DsdC1\(^{\text{FLAG}}\) (\( P = 1.98 \times 10^{-109} \)) and DsdC2\(^{\text{FLAG}}\) (\( P = 2.5 \times 10^{-74} \)) both within the coding sequence and directly downstream of the neuO gene, with only a very modest peak being identified within its promoter region (Fig. 4A). To verify these interactions, we confirmed that DsdC was capable of binding to the promoter region and a fragment corresponding to the neuO 3’ coding region in vitro, while in vivo binding seemed to be enriched at the intragenic sites (Fig. 4B). As another negative control (in addition to the \( \text{kanamycin} \) gene), we used a fragment of the native \( \text{adhes} \) gene, which was not associated with a ChIP-seq peak, to verify that this was a specific DsdC-neuO interaction (Supplementary Fig. S7). Additionally, we used qRT-PCR to verify that neuO was indeed down-regulated (more than sevenfold; \( P < 0.05 \)) in the \( \Delta \text{dsdC1/2} \) mutant background (Fig. 4C). These data suggest that the noncanonical binding of DsdC within the neuO gene region somehow regulates its expression at the transcriptional level.

We next turned our attention to what the biological role of this regulatory event might be. Adhesion and invasion of the blood–brain barrier are key virulence mechanisms of NMEC, and the K1 capsule is believed to play a role in this process (7, 19, 20, 22, 23). We therefore tested the ability of WT NMEC and a \( \Delta \text{neuO} \) mutant to adhere to and invade the human brain microvascular endothelial cell line, hCDMEC/D3. We first confirmed that deletion of neuO had no observable growth defect that would have an impact on interpretation of the cell infection assays (Supplementary Fig. S8). We then determined that \( \Delta \text{neuO} \) did not have a decreased ability to adhere to hCDMEC/D3 cells (\( \sim 20–30\% \) adherence efficiency comparable to that of WT NMEC). Furthermore, the invasion efficiency was comparable for both WT and \( \Delta \text{neuO} \) (\( \sim 0.002\% \)), also displaying no statistically significant difference. In addition, we tested the ability of \( \Delta \text{dsdC1/2} \) to adhere to and invade hCDMEC/D3 cells (Supplementary Fig. S9). As expected, we...
observed no phenotype in the ΔdsdC1/2 background, suggesting that neither NeuO nor its regulator, DsdC, was involved in the ability of NMEC to attach to or invade human brain endothelial cells in vitro.

The original report describing the discovery of NeuO in NMEC strain RS218 documented that neuO was encoded on the CUS-3 prophage (17, 24–26). Interestingly, CUS-3 is integrated into the genome at a known hypervariable region between dsdC and the argW transfer RNA (tRNA) gene in RS218. Many K1-specific bacteriophages use the polysialic capsule as their receptor. Prophages often encode enzymes that alter their host receptor, thereby modulating superinfection by additional bacteriophages. Given the role of NeuO in O-acetylation of the K1 capsule, we hypothesized that deletion of neuO would therefore affect the ability of a K1 bacteriophage to infect and lyse NMEC CE10 (17, 27). To test this, we assayed the formation of plaques on bacterial agar plates overlaid with a mixture of NMEC-containing agar and spotted with a K1 bacteriophage-containing buffer. Experiments were performed in biological triplicate, with representative images from each assay indicated to the Left of the graph. (B) K1 bacteriophage killing assays performed in liquid media for NMEC, ΔneuO, or ΔneuO plus pNeuO. Data points were determined by measuring optical density (600 nm) after 1 h of phage exposure. *** and **** indicate P < 0.001 and 0.0001 respectively from three biological replicates (Student's t test) with SD. *P < 0.05, ***P < 0.001, and ****P < 0.0001 derived from three biological replicates (Student t test) with SD.

DsdC Regulates NeuO-Associated Resistance to K1 Bacteriophage Killing. To assess if DsdC played a role in resistance against bacteriophage attack via its regulatory effect on neuO, we tested the ΔdsdC1/2 mutant in K1 killing assays. The ΔdsdC1/2 mutant was significantly (P < 0.0001) more susceptible to bacteriophage
killing than WT NMEC, mirroring the phenotype observed for ΔnenO (Fig. 5A). To confirm the functional redundancy of both copies of DsdC in regulation of this process, we successfully complemented the double mutant with both dsdC1 and dsdC2. Plasmid-borne expression of DsdC1 and DsdC2 significantly (P = 0.0109 and P = 0.0045, respectively) reduced the lytic burden of K1 bacteriophage in ΔdsdC1/2 by ~50% (Fig. 5A and SI Appendix, Fig. S10). Additionally, the ΔdsdC1/2 double mutant formed significantly larger plaques on bacterial soft agar plates (average 3.65 mm; P = 0.0117) than WT NMEC after K1 bacteriophage exposure (Fig. 5B). Finally, to confirm that the K1 resistance phenotype was attributable to DsdC-mediated regulation of NeuO, we also complemented the ΔdsdC1/2 mutant indirectly by constitutive expression of nenO in trans. This bypassed the requirement for DsdC1/2 and completely restored the WT plaque size, alleviating the lytic burden of K1 phage on the ΔdsdC1/2 mutant (Fig. 5B and C). Collectively, these results confirm that NeuO is a key enzyme that plays a role in defense against bacteriophage killing through modulating the K1 capsule and that maximal protection efficiency is facilitated through regulation by the core genome–encoded TF DsdC.

**Discussion**

The regulation of gene expression dictates how bacteria control cellular processes central to their survival in specific environments. Bacteria are capable of responding to their surroundings in a precise manner to express the right genes at the right time (1–3). Furthermore, the process of metabolism is often intrinsically linked to other functions, including virulence, to maximize the competitiveness of pathogens at distinct host niches (6). This is particularly important within organisms that have highly mosaic genomes, composed of both core and horizontally acquired genetic elements. Bacteria therefore require TFs to control multiple, overlapping roles in the cell through genetic regulatory adaptation (5). Here, we identify a distinct role for the regulator of D-ser catabolism in the NMEC strain CE10. DsdC controls resistance against bacteriophage attack through direct regulation of the O-acetylttransferase encoding gene neuO, representing a key regulatory adaptation contributing to the ecological success of NMEC.

NMEC strains typically encode two copies of the *dsdC* locus (14). This suggests that metabolism of D-ser represents an ecological advantage to NMEC, presumably offering a fitness advantage during infection of the brain. Accordingly, we demonstrate here that both loci are functionally redundant in D-ser use. However, this represents an unusual scenario, given that homologous genes with identical functions are often not stably maintained within bacterial genomes, unless there is an advantage to the organism (28). This therefore presents three possible reasons for stably maintaining both copies of the locus: double the capacity to uptake and catabolise D-ser, discrete control in two chromosomal contexts, or regulatory functions for DsdC that exist beyond the D-ser catabolism locus. In agreement with the latter hypothesis, our global analysis of DsdC in NMEC identified that both copies of the regulator bound to hundreds of sites along the chromosome, irrespective of the presence of D-ser in the environment. Close inspection of the data revealed that DsdC1 and DsdC2 do not bind to distinct regions, suggesting they are functionally redundant in general as global regulators. Importantly, we identified binding of regions that are reported to be involved in NMEC virulence, such as LPS and capsular biosynthesis (7). Transcriptional analyses identified that these processes are differentially regulated by DsdC in a D-ser–dependent manner. This finding is in line with our previous work describing the transcriptional effects of D-ser on EHEC and UPEC virulence (16). The regulatory effects of D-ser on virulence also extend beyond the *E. coli* species. For instance, D-ser has been reported to induce expression of the *sp lipase* involved in *Staphylococcus saprophyticus* virulence, and in *Proteus mirabilis*, catabolism of D-ser caused increased fitness during polymicrobial catheter-associated urinary tract infection (29, 30). Furthermore, D-ser was shown to inhibit attachment and biofilm formation in *Staphylococcus aureus*, down-regulating key genes, including *agrA, sarS*, and *dld* (31). These studies, and ours, enforce the concept that D-ser modulates virulence regulation in multiple bacterial species. Despite these findings, deletion of both *dsdC* copies resulted in no observable virulence-related phenotype in NMEC, suggesting other roles for this regulator must exist.

Another striking observation from our ChIP-seq analyses was the pattern of binding to distinct chromosomal regions. Approximately half of the DsdC-bound regions appeared to

*Fig. 5.* DsdC regulates NeuO-associated resistance to K1 bacteriophage killing. (A) K1 bacteriophage killing assays performed in liquid media for NMEC, ΔdsdC1/2, ΔdsdC1/2 plus pDsdC1, or ΔdsdC1/2 plus pDsdC2. Data points were determined by measuring optical density (OD; 600 nm) after 1 h of phage exposure. (B) Phage plaque assays showing the diameter (mm) of plaques formed on NMEC-, ΔdsdC1/2-, and ΔdsdC1/2 plus pNeuO-containing agar plates spotted with K1 bacteriophage-containing buffer. Experiments were performed in biological triplicate, with representative images from each assay indicated to the left of the graph. (C) K1 bacteriophage killing assays performed for NMEC, ΔdsdC1/2, and ΔdsdC1/2 plus pNeuO. ns, not significant. *P < 0.05 (Student’s t test) with SD (A) from 10 biological replicates, ****P < 0.0001 (Student’s t test) with SD from (A) 10 or (C) three biological replicates.
form clusters spanning entire operons, as opposed to more traditional discrete binding sites located upstream of regulated genes. While this pattern was highly unusual, noncanonical TF binding has been observed in bacteria. For example, the RutR TF in *E. coli* K-12 binds to mostly intragenic sites, with no apparent effects on transcription (32). Indeed, most of the DsdC-bound regions in our study were not associated with transcriptional shifts. Nonregulatory binding events could therefore represent evolutionary relics, where the TF no longer functions at this site (32). However, we did observe differential expression at the LPS and capsular biosynthetic loci, which were associated with the unusual clustered binding pattern of DsdC across operons. This may represent a mechanism of transcriptional regulation that mirrors the activity of certain nucleoid-associated proteins, such as H-NS, to bind along large spans of DNA to silence transcription (33, 34). Indeed, the line between what constitutes a TF and a nucleoid-associated protein has become increasingly blurred, with the existence of seemingly overlapping functions in transcriptional regulation (35, 36). It is therefore plausible that DsdC falls into this category, acting upon its targets both by traditional promoter interactions (e.g., at *dscCXA*) and clustered interactions that may result in effects on local DNA structure and therefore the accessibility of other factors such as RNA polymerase.

The most intriguing discovery in this work was the DsdC-dependent regulation of resistance to K1 bacteriophage attack. There are >70 capsular antigens; however, K1 is overrepresented in NMEC, accounting for 84% of isolates obtained from infected neonates, as well as pathogenic strains of *Neisseria* and *Klebsiella* sp (20, 37). The K1 capsule is a homopolymer of α2,8-linked N-acetylneuraminic acid residues (Neu5Ac), a molecular mimic of sialic acid produced on neural cells (38). *E. coli* K1 strains can modify their capsular structure by O-acetylation of carbon 7 or 9 of the sialyl units (39). We identified that maximal expression of *neuO* (encoding *O*-acetyltransferase) was dependent upon DsdC, irrespective of the presence of D-ser. O-acetylation of the capsule occurs in many bacterial and fungal species, for example, playing a role in protection from the bactericidal activity of serum in *Neisseria meningitidis* (40). The genomic context of *neuO* suggests it was originally acquired on a prophage, and its regulation by DsdC therefore represents an adaptive regulatory trait in NMEC. K1 bacteriophages use the K1 capsule as an attachment site, employing tailspike proteins to cleave the Neu5Ac (27). To prevent superinfections within the host cell, prophages often hijack an ancestral TF to promote its continued function—resulting in an inability to tolerate D-ser (8). Interestingly, this uptake–release-like phage appears prone to decay, because in many isolates, including RS218, genes essential for phage replication, including those encoding tail proteins and coat proteins, are annotated as pseudogenes. Moreover, there are several examples of strains exhibiting loss of genetic content in this region such that the proximity of *neuO* to *dscD* is reduced, often to as little as 8 Kb (e.g., strain CE10 and SCU-175; accession No. CP054379.1) (*SI Appendix*, Table S9). A second, less common insertion site for CUS–3-like phage and therefore *neuO* exists between *pgl* and *biot*. The size of this insertion also varies between 20 and 50 Kb, indicating that genetic decay may occur following acquisition. Interestingly, CE10 possesses phage-like elements in both locations, with the *argW* insertion appearing extensively degraded, including truncation of the 5’ extremity (nucleotides 537–777 remain) of *neuO*. The second *neuO* allele is, however, intact and as shown here functional in protection against K1 phage attack. Therefore, although genetic decay of CUS–3-like phages is common in *E. coli*, the retention of functional *neuO* alleles by the prototype NMEC isolates RS218 and CE10 indicates that capsule *O*-acetylation may provide a pathotype-specific ecological advantage through prevention of phage infection.

**Conclusion.** Tailoring transcriptional regulatory networks by recycling TFs to control alternative and often horizontally acquired genes can lead to the emergence of ecologically beneficial traits in bacteria. However, the extent of this phenomenon and associated mechanisms driving such regulation remains a key question in bacterial genetics. We identify that the TF DsdC, responsible for tolerance to D-ser, has adapted alternative metabolism-independent roles in the regulation of NMEC gene expression. We show that DsdC unusually binds to entire operons involved in envelope biogenesis, affecting their transcription, and is critical for regulating NeuO-mediated protection against bacteriophage attack. Given its carriage on a seemingly dispensable prophage, retention and subsequent control of NeuO are crucial. As such, its regulation has been hijacked by an ancestral TF to promote its continued functionality and enhance the environmental versatility of NMEC K1 isolates in the face of bacteriophage attack.

**Materials and Methods**

A complete list of all bacterial strains, plasmids, and primers used in this study can be found in *SI Appendix*, Tables S10–S12. Details of all methodology related to the experiments and analyses are included in the *SI Appendix, Materials and Methods*. This includes growth conditions, strain generation, cloning procedures, ChiP-seq, RNA-seq, qRT-PCR, protein overexpression and purification, EMSA, DNeasel footprinting, Western blotting, cell culture, phage killing assays, and data analysis. All raw sequencing data have been deposited in the European Nucleotide Archive under the accession Nos. RNA-seq study PRJEB36547 (ERS4281326–ERS4281334, ERS4281334–ERS4281354, ERS4281354–ERS4281356, ERS4281338–ERS4281340, ERS4281344–ERS4281346, and ERS4281350–ERS4281352) and ChiP-seq study PRJEB36549 (ERS4281484, ERS4281489–ERS4281494, and ERS12154679–ERS12154680).

**Data Availability.** RNA-seq and ChiP-seq data have been deposited in the European Nucleotide Archive (PRJEB36547, PRJEB36549). No previously
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