Characterisation of vascular changes in different stages of Stargardt disease using double swept-source optical coherence tomography angiography

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ABSTRACT

Objective To describe vascular changes in different stages of Stargardt disease (STGD) via double swept-source optical coherence tomography angiography.

Methods and analysis Prospective, cross-sectional case–control study. Twenty-three patients (45 eyes) with ABCA4 mutations graded according to the Fishman STGD classification and 23 controls (23 eyes) were included. Two independent investigators quantified the foveal avascular zone (FAZ) in the superficial and deep capillary plexus (SCP/DCP) and the areas presenting rarefied flow and complete vascular atrophy in the outer retina to choriocapillaris (ORCC) and choriocapillaris (CC) slab.

Results The mean age at first diagnosis of STGD was 24.0 years (range 9–50) and 37.9 years (range 18–74) at the time of examination. Eleven patients were assigned to the Fishman STGD classification stage (S) 1, three to S2, eight to S3 and one to S4. The FAZ in SCP and DCP was increased in all stages compared with controls (p<0.01). Areas with rarefied flow signal and vascular atrophy were detected in the ORCC and the CC layer and grew with increasing stage of disease (p<0.01). The duration of disease correlated with the extent of the enlarged FAZ in the SCP/DCP and with the area of reduced flow in the ORCC and CC layer (p<0.01). Best corrected visual acuity correlated negatively with the extent of the enlarged FAZ in the SCP/DCP (p<0.0001), as well as with enlarged atrophic area in the ORCC and CC layer (p=0.026 and p=0.074).

Conclusions Patients with STGD reveal vascular changes in the retina and CC in all disease stages. The avascular zone in the SCP/DCP and areas with rarefied flow signal in the ORCC/CC increase with the duration and stage of disease, indicating progressive vascular decay most likely secondary to retinal pigment epithelium and neuronal loss. Furthermore, increased vascular damage is associated with decreased vision.

INTRODUCTION

Stargardt disease (STGD) is the most common cause of hereditary juvenile macular dystrophy with a prevalence of about 1:10,000. The disease mainly affects young patients (6–45 years) and causes a progressive, irreversible bilateral loss of central vision. Fundus examination reveals a beaten bronze or a bull’s-eye macular appearance and characteristic deep yellowish-white fish-shaped flecks in the macular and perimacular region. The end-stage fundus aspect is characterised...
by extensive retinal pigment epithelium (RPE) and choriocapillaris (CC) atrophy, with resorbed flecks and sparse pigmentation.3

The underlying pathophysiology of STGD is not yet fully understood. It is generally accepted that mutations in the ABCA4 gene cause ATP-binding cassette (ABC) protein dysfunction, leading to malfunctioning photoreceptor cells and the deposition of lipofuscin in RPE cells.4 The initial step of RPE damage is detectable in fundus autofluorescence (FAF) images, currently the standard diagnostic modality for STGD.4 Other diagnostic tools such as optical coherence tomography (OCT), fluorescein angiography and indocyanine green angiography exhibit concomitant thinning and loss of the CC in STGD.5

Optical coherence tomography angiography (OCTA) is a non-invasive technique for imaging the microvasculature of the retina and choroid.6 Few studies to date have analysed the retinal and choroidal vasculature in STGD using OCTA.3 7 8 To the best of our knowledge, none of those studies analysed OCTA findings in different disease stages, or examined vascular changes in relation to the disease’s duration.

The aim of this STGD cohort study is therefore to describe the vascular characteristics in the retina and choroid in different stages of STGD using double swept-source OCTA (dSS-OCTA) and to correlate them to the duration and stage of disease.

MATERIALS AND METHODS

Study design
This was a prospective, cross-sectional, monocentric, observational case–control study. All patients consulted our eye centre for the first time between 2000 and 2018 and were examined in the study between 2017 and 2018.

Inclusion and exclusion criteria
Patients with STGD and controls with relevant systemic disease like diabetes mellitus or cardiovascular diseases were excluded. Only patients with STGD with proven ABCA4 mutation and no other retinal diseases were included. Controls were only included when exhibiting no retinal or other ophthalmological diseases. Eyes revealing media opacities, such as an advanced cataract, were also excluded, due to reduced OCTA signal strength. Only images with signal strength equal to or greater than 7 were included. Furthermore, images with other artefacts like motion artefacts were excluded.

Ophthalmological examination
Patients underwent a comprehensive ophthalmological examination including measurement of best corrected visual acuity (BCVA in logMAR), assessment of intraocular pressure, fundus examination and FAF imaging (Spectralis HRA, Heidelberg, Germany). OCTA examination was performed using the Plex Elite OCTA system (Zeiss PLEX Elite 9000, Germany), which employs a full spectrum dSS-OCT (wavelength 1050 nm, A-scan rate 100,000 A-scans/s, A-scan depth 3.0 nm in tissue). 6×6 mm² and 9×9 mm² volume scans were performed.

Used OCTA slabs
The automated segmentation of the superficial capillary plexuses (SCP, slab from internal limiting membrane to inner plexiform layer), deep capillary plexuses (DCP, slab from inner plexiform layer to outer plexiform layer), the outer retina to choriocapillaris layer (ORCC, slab from outer plexiform layer to RPE) and the CC (slab 29–49 μm under the RPE) were obtained using the manufacturer’s built-in software. Automatic segmentation was controlled and corrected by the investigators for misalignments.

Phenotypical classifications
Patients were clinically graded according to the Fishman STGD classification.9 Stage (S) 1 is characterised by macular pigmentary changes and irregular pigmentary spots (flecks) located within one disc diameter of the fovea. S2 is identified when the pigmentary changes and pisciform flecks are located beyond the vascular arcades temporally and often extend nasally to the optic disc. S3 is defined by the resorption of previously diagnosed flecks resulting in focal CC and RPE atrophy of the macula. S4 is characterised by diffusely absorbed flecks and extensive CC and RPE atrophy throughout the entire central fundus.

Image analysis
To analyse vascular changes in different STGD stages, OCTA images were exported and analysed using a self-designed analysis tool programmed in language R (www.r-project.org). Two independent and masked investigators (MR, AG) quantified the foveal avascular zone (FAZ) in the SCP and DCP using a freehand selection tool (figure 1). The FAZ in the SCP and DCP in all STGD stages and controls were compared with each other. In
the ORCC and CC the area of rarefied flow and the area of complete vascular atrophy enabling a direct view of the choroid were determined and compared among the four disease stages (figure 2). If the area to be measured accommodated into the 6×6mm² volume scan, that scan was preferred due to its higher resolution. As soon as the area to be measured exceeded the 6×6mm² volume scan, the 9×9mm² volume scan was used.

Patient and public involvement
This research was done without patient involvement. Patients were not invited to comment on the study design and were not consulted to develop patient-relevant outcomes or interpret the results. Patients were not invited to contribute to the writing or editing of this document for readability or accuracy.

Statistical analysis
SPSS V.20.0 was used for statistical analysis. For the linear mixed models R-package lmerTest was used. A probability (p) value of α<0.05 was considered statistically significant. For descriptive data analysis, the mean and SD were calculated. The Mann-Whitney U test was used to compare patient characteristics. Bland-Altman plots and intraclass correlation coefficient (ICC) were used to determine the reproducibility and variability of the manual quantification in pixels of the FAZ in the SCP and DCP, as well as the area of rarefied flow and the area of complete vascular atrophy in the ORCC and CC layer between the two examiners. To reveal any association between the patient’s age, disease duration and BCVA with vascular changes, we calculated the Spearman’s r.

RESULTS
Study population
Twenty-three patients with STGD met all inclusion criteria. Twenty-three age-matched, healthy participants with a BCVA of 0.0 logMAR or better served as controls.

Patient characteristics
A total of 45 eyes of 23 patients with genetically secured STGD (13/10 females/males) were included. One eye was excluded due to artefacts and resulting partially reduced signal strength because of significant posterior polar cataract. The average age at initial diagnosis of the disease was 24.0 years (range 9–50) and 37.9 years (range 18–74) at the time of the study examination. Table 1 presents detailed information of our study group. Twenty-two eyes of 11 patients were assigned to Fishman STGD classification S1, 6 eyes of 3 patients to S2, 15 eyes of 8 patients to S3 and 2 eyes of 1 patient to S4. The age at disease onset was similar in all stages (S1: 24.8±12.8 years; S2: 14.7±6.0 years; S3: 27.5±9.5 years; S4: 14.0 years, p=0.32). However, patients in S3 and S4 were older than those in S1 and S2 at the time of the OCTA examination (S1: 33.1±10.8 years; S2: 21.3±2.9 years; S3: 48.9±15.5 years; S4: 52.0 years, p=0.01). Twenty-three controls (11/12 females/males, 23 eyes) with an average age of 37.1 years (range 20–69) were included. There was no significant difference in age between the STGD group and controls (p=0.97).

Qualitative analysis of the OCTA images in STGD
dSS-OCTA imaging yielded detailed images of the retinal microcirculation and a homogeneous texture of the ORCC and CC flow signal with a dense and fine granular appearance in all control patients (figure 2). All patients with STGD, in contrast, exhibited structural vascular changes with rarefied or even totally absent flow signal in SCP, DCP, ORCC and CC (figure 2). These structural changes were subtle in S1 and S2 and most pronounced in S3 and S4. Rarefied flow in the ORCC and CC was observed in all 45 eyes (100%) while ORCC and CC atrophy was detected in 25 of 45 eyes (53%) of patients with STGD. Concomitant rarefaction and atrophy of the CC was detected in 4 of 22 eyes (18%) in S1, in 4 of 6 eyes (67%) in S2, in all 15 eyes (100%) in S3 and in both eyes (100%) in S4.

Quantitative OCTA analysis of OCTA images in STGD
To quantitatively assess the described vascular alterations, OCTA images of the SCP, DCP, ORCC and CC were analysed by two independent and masked examiners as described above. Bland-Altman plots (figure 3) reveal...
Table 1  Clinical and molecular data on the patients with STGD in our study group

| ID | Age (years) | Onset (years) | Sex | BCVA OD logMAR | BCVA OS logMAR | Fishman STGD classification | Mutation 1 | Mutation 2 | Mutation 3 |
|----|-------------|---------------|-----|----------------|----------------|----------------------------|-------------|-------------|-------------|
| 1* | 40          | 10            | F   | 2              | 2              | 1                          | c.5196+2T>C  | c.5882G>A   | (Splice)    |
|    |             |               |     |                |                |                            | p.Gly1961Glu | (p.Gly1961Glu) |             |
| 2  | 23          | 15            | F   | 2              | 2              | 1                          | c.2041C>T    | c.5882G>A   | (p.Arg681Xaa) (het.) |
|    |             |               |     |                |                |                            | p.Gly1961Glu | (p.Gly1961Glu) |             |
| 3  | 23          | 17            | F   | 1.3            | 1              | 1                          | c.1622T>C    | c.3113C>T   | (p.Leu541Pro) (p.Ala1038Val) |
|    |             |               |     |                |                |                            | p.Gly1961Glu | c.5882G>A   | (p.Gly1961Glu) |
| 4  | 27          | 23            | M   | 1              | 1              | 1                          | c.4462T>C    | c.5882G>A   | (p.Cys1488Arg) |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 5  | 28          | 24            | F   | 1              | 1              | 1                          | c.2588G>C    | c.2828G>A   | (p.Gly863Ala, Splice) |
|    |             |               |     |                |                |                            | p.Arg943Gln  | c.5603A>T   | (p.Asn1868Ile) |
| 6  | 39          | 31            | M   | 1.3            | 1              | 1                          | c.5461-10T>C | c.5882G>A   | (Unknown) |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 7  | 37          | 36            | F   | 0.3            | 0.1            | 1                          | c.3322C>T    | c.5882G>A   | (p.Arg1108Cys) |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 8  | 52          | 42            | F   | 1              | 1              | 1                          | c.2894A>G    | c.5882G>A   | (p.Asn965Ser) |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 9  | 49          | 48            | M   | 0.3            | 0.1            | 1                          | c.2588G>C    | c.2588G>C   | (p.Gly863Ala) |
|    |             |               |     |                |                |                            | p.Gly863Ala |             |             |
| 10 | 22          | 13            | M   | 1              | 1              | 1                          | c.3322C>T    | c.5882G>A   | (p.Arg1108Cys) |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 11 | 25          | 14            | M   | 1              | 1              | 1                          | c.5882G>A    | c.6238_6239delTC | p.Ser2080HfsX16 |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 12†| 18          | 9             | M   | 1.3            | 1              | 2                          | c.2588G>C    | c.6238_6239delTC | p.Ser2080HfsX16 |
|    |             |               |     |                |                |                            | p.Gly863Ala, Splice |             |             |
| 13†| 23          | 14            | F   | 1.3            | 1.3            | 2                          | c.2588G>C    | c.6238_6239delTC | p.Ser2080HfsX16 |
|    |             |               |     |                |                |                            | p.Gly863Ala, Splice |             |             |
| 14 | 23          | 21            | M   | 0.6            | 0.6            | 2                          | c.2588G>C    | c.858+2T>A  | (Unknown) |
|    |             |               |     |                |                |                            | (p.Gly863Ala, Splice) |             |             |
| 15 | 28          | 19            | F   | 2              | 2              | 3                          | c.1622T>C    | c.3113C>T   | (p.Leu541Pro) (p.Ala1038Val) |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 16*| 51          | 22            | M   | 1.2            | 1.1            | 3                          | c.5196+2T>C  | c.5882G>A   | (Splice)    |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 17 | 40          | 23            | F   | 0.9            | 0.9            | 3                          | c.5413A>G    | c.5714+5G>A | (p.Asn1805Asp) |
|    |             |               |     |                |                |                            | (Unknown) |             |             |
| 18*| 57          | 27            | F   | 0.9            | 0.8            | 3                          | c.5196+2T>C  | c.5882G>A   | (Splice)    |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 19 | 74          | 27            | M   | 1.3            | 1.3            | 3                          | c.5882G>A    | Deletion exon 18–19 |            |
|    |             |               |     |                |                |                            | (p.Gly1961Glu) |             |             |
| 20 | 30          | 27            | M   | 1.3            | 1.3            | 3                          | c.1622T>C    | c.3113C>T   | (p.Leu541Pro) (p.Ala1038Val) |
|    |             |               |     |                |                |                            | p.Gly1961Glu |             |             |
| 21 | 52          | 25            | F   | 1.3            | 1.3            | 3                          | c.2692G>T    | c.5606C>T   | (p.Glu698Xaa) |
|    |             |               |     |                |                |                            | (p.Pro1869Leu) |             |             |
| 22 | 59          | 50            | F   | -0.1           | 0.1            | 3                          | c.4610C>T    | c.5461-10T>C | (Unknown) |
|    |             |               |     |                |                |                            | (Unknown) |             |             |
| 23 | 52          | 14            | F   | 2.3            | 2.3            | 4                          | c.2931G>A    | c.5714+5G>A | (Unknown) |
|    |             |               |     |                |                |                            | (p.Gly978Ser) |             |             |

*Siblings.  †Siblings.

BCVA, best corrected visual acuity; F/M, female/male; OD, right eye (oculus dexter); OS, left eye (oculus sinister); STGD, Stargardt disease.

Inter-rater reliability between the two examiners concerning their analyses of the avascular area in the SCP and DCP, the area of rarefied and absent flow in the ORCC and CC. The ICC for the avascular area in the SCP was 0.99 (p<0.0001), respectively 0.99 (p<0.0001) in the DCP, 0.99 (p<0.0001) and 0.99 (p<0.0001) regarding the area of rarefied flow and 1.0 (p<0.0001) and 1.0 (p<0.0001) regarding the area of absent flow in the ORCC and the CC layer. Relying on such minimal interobserver variability, we employed one grader’s (MR) quantification for the analyses below.

**Analysis of the retinal vasculature**

All patients with STGD revealed an enlarged FAZ in the SCP compared with controls which expanded as the disease stage increased (S1: 0.21±0.10mm²; S2:...
Figure 3  Bland-Altman analysis of inter-rater reliability. Two independent and masked investigators (MR, AG) quantified the foveal avascular zone (FAZ) in the superficial capillary plexus (SCP, A) and deep capillary plexus (DCP, B) using a freehand selection tool. Furthermore, the area of rarefied flow and complete atrophy was quantified in the outer retina to choriocapillaris (ORCC, C and E) and the choriocapillaris (CC, D and F) layer.

Similarly, the FAZ in the DCP was enlarged in S1, S2, S3 and S4 compared with controls (S1: 0.16±0.09 mm²; S2: 0.17±0.04 mm²; S3: 0.21±0.11 mm²; S4: 0.40±0.08 mm²; controls: 0.11±0.03 mm², over all p<0.01, figure 4A2). Furthermore, patients with STGD revealed a larger FAZ in the SCP compared with DCP (0.26±0.19 mm² vs 0.19±0.10 mm², figure 4A3). In contrast, our controls exhibited a smaller FAZ in the SCP compared with DCP (0.03±0.01 mm² vs 0.11±0.03 mm², figure 4A3).

Analysis of the choroidal vasculature

Similar to the retinal vasculature findings, altered OCTA signals in ORCC and CC increased with advancing STGD stages. The area of rarefied OCTA signal in the ORCC was increased in S2, S3 and S4 when compared with S1 (S1: 1.07±0.46 mm²; S2: 8.46±0.86 mm²; S3: 7.05±2.88 mm²; S4: 9.00±0 mm²; over all p<0.01, figure 4B1). Similarly, the area of rarefied flow in the CC was increased in S2, S3 and S4 when compared with S1 (S1: 6.90±0.58 mm²; S2: 0.03±0.07 mm²; S3: 0.99±0.88 mm², over all p<0.01, figure 4B2). Interestingly, the area of rarefied flow in the ORCC (2.38±0.90 mm²) was larger than the one in the CC layer (1.86±1.80 mm²) in all patients with STGD independent of disease stage (figure 4B3).

Finally, the area of vascular atrophy in the ORCC also grew with increasing stage and was larger in S4 than in S1, S2 and S3 (S4: 6.69±0.79 mm²; S1: 0.03±0.07 mm²; S2: 0.05±0.04 mm²; S3: 1.01±0.90 mm², over all p<0.05, figure 4C1). Similarly, the area of vascular atrophy in the CC grew with increasing stage and was increased in S4 compared with S1, S2 and S3 (S4: 6.69±0.79 mm²; S1: 0.03±0.07 mm²; S2: 0.05±0.04 mm²; S3: 1.01±0.90 mm², over all p<0.05, figure 4C2). The area of atrophy in the ORCC (1.31±1.89 mm²) was similar to the one in the CC layer (1.18±1.82 mm²) in all patients with STGD independent of disease stage (figure 4C3).
Figure 4  Quantitative evaluation of vascular alterations in Stargardt disease (STGD). Box and whiskers plot demonstrating the area of vascular alterations (in mm²) of patients in different stages of STGD (S1–S4) and controls (Ctr). Data on the foveal avascular zone in the superficial and deep capillary plexuses (SCP and DCP) are depicted in A1–A3, data on the area of rarefication of flow signal/atrophy in the outer retina to choriocapillaris (ORCC) layer and choriocapillaris (CC) are shown in B1–B3 and C1–C3, respectively. To assess any correlation between the areas of rarefied flow in the ORCC compared with the CC layer, we excluded data of 17 eyes as the area of rarefied flow exceeded the maximum available image size of 9×9 mm². A linear mixed model, using the diagnosis and stage of disease as a fixed factor and the patient ID as a random factor, also including signal strength values for each image, was used to analyse differences between the groups. All analyses (A1, A2, B1, B2, C1, C2) show a significance value of p<0.01.

Correlation between vascular alterations and patient’s age, disease duration and BCVA

No association between patient age and the FAZ in the SCP (r=0.06, p=0.66) or the DCP (r=0.01, p=0.95) was revealed. Similarly, no significant correlation between patient age and the area of flow rarefication in the ORCC
Figure 5  Association between best corrected visual acuity (BCVA) and vascular changes. Illustrated is the correlation between BCVA (in logMAR) and the foveal avascular zone (FAZ) in the superficial capillary plexus (SCP, A) and deep capillary plexus (DCP, B) and the areas presenting rarefied flow (C, D) and complete vascular atrophy (E, F) in the outer retina to choriocapillaris (ORCC) and choriocapillaris (CC). To assess any correlation between BCVA and the area of rarefied flow in the ORCC, respectively the CC layer, we excluded data of 17 eyes, respectively 16 eyes, as the area of rarefied flow exceeded the maximum available image size of 9×9 mm².

Furthermore, disease duration correlated with an enlarged FAZ in the SCP (r=0.42, p<0.01) and the DCP (r=0.39, p<0.01), as well as with an enlarged rarefied

(r=0.15, p=0.33) and the CC (r=0.16, p=0.30) was identified. However, patient age associated significantly with an enlarged area of atrophy in the ORCC (r=0.30, p=0.048) and the CC layer (r=0.34, p=0.02).
and atrophic area in the ORCC (r=0.37, p=0.01/r=0.66, p<0.001) and CC layer (r=0.40, p<0.01/r=0.67, p<0.001).

BCVA correlated negatively with the extent of the enlarged FAZ in the SCP (r=0.81, p<0.001, figure 5A) and DCP (r=0.63, p<0.0001, figure 5B) in patients with STGD and controls, as well as with enlarged atrophic area in the ORCC (r=0.33, p=0.026, figure 5E) and a trend of enlarged atrophic area in the CC layer (r=0.27, p=0.074, figure 5F) in patients with STGD. We detected no significant correlation between BCVA and enlarged rarefied area in the ORCC and CC layer (r=0.07, p=0.72, figure 5C/r=−0.01, p=0.96, figure 5D).

**DISCUSSION**

To identify vascular alterations in STGD is an important prerequisite for defining disease pathophysiology, individual prognosis and the optimal time point for therapeutic interventions in the future. However, little is known about the microvascular alterations associated with STGD. Former studies using fluorescein or indocyanine green angiography already detected vascular changes in STGD. These studies, however, were limited by low imaging resolutions and dye leakiness which precluded exact quantification of the microvascular network. The advent of OCTA has enabled in-depth imaging and analysis of the retinal and choroidal vasculature in chorioretinal disease. The present study aimed to characterise vascular changes in STGD using dSS-OCTA. All our patients with STGD exhibited vascular alterations in the retina and choroidea that increased with age and disease stage, indicating progressive vascular decay over the disease course. Furthermore, increased vascular damage seems to be associated with decreased vision.

Few investigations have examined the retinal and choroidal vasculature in patients with STGD using OCTA. Mastropasqua et al assessed the CC in 12 patients with STGD using OCTA and detected an extensive loss of the CC in areas of RPE atrophy. Guduru et al quantitatively assessed the CC in STGD and demonstrated that the RPE loss was more pronounced than the concomitant CC loss indicating primary RPE degeneration followed by vascular atrophy. Both studies, however, were limited by the use of spectral domain OCTA (SD-OCTA) technology, which is prone to image artefacts when assessing the choroidal vasculature. To overcome this limitation, we employed dSS-OCTA, as it delivers more accurate imaging of the CC thanks to its longer wavelength and better sensitivity roll-off. We distinguished two different CC textures in STGD: a central area of complete CC atrophy which permitted direct visualisation of deep choroidal vessels and a surrounding area of rarefied CC. These data support fluorescein angiography findings demonstrating a progressively hyperfluorescence in the borders of the area of atrophy in STGD, while the centre was isohypofluorescent. While all patients with STGD revealed areas of rarefied CC in our study, areas of CC atrophy were predominantly found in more advanced STGD stages. In line with this observation, we detected a positive correlation between the time since first diagnosis and the area of CC atrophy, indicating progressive vascular decay in the CC layer. Prospective studies entailing longitudinal follow-up are warranted to clarify whether the areas of CC rarefication eventually become atrophic, and whether they precede or follow RPE atrophy during the disease course.

To the best of our knowledge, the ORCC layer has not been examined in STGD so far. Hitherto this layer has mainly been used to visualise features of type I and type II choroidal neovascularization (CNV). The ORCC slab includes parts of the outer retina, the RPE and, to a small degree, the CC, and reveals a homogeneous architecture in healthy individuals. Patients with STGD, however, exhibit an ORCC slab emitting irregular and rarefied OCTA signals, and areas revealing a total loss of retinal signal texture with uncovered large-calippered vessels which increase with duration and stage of disease. Although recent studies indicate that SS-OCTA is capable of delivering ORCC angiograms with better image quality than SD-OCTA, the projection artefacts from the inner retinal vessels (reduced in density in STGD) cannot be completely ruled out. Furthermore, shadowing artefacts from lipofuscin-containing flecks or window defects from atrophic RPE may enhance the area of alterations in the ORCC. Since the affected area in the ORCC was significantly larger than the SCP, DCP and CC, it is likely that a combination of the aforementioned phenomena contributes to the changes observed in the ORCC, and that the ORCC represents an overall measure to visualise retinal, RPE and choroidal anomalies in STGD.

While the choroidal vasculature has attracted the most interest, considerably less is known about the retinal vasculature in STGD. Mastropasqua et al studied 12 patients with STGD and reported on a reduction in vessel density in the fovea within the SCP and DCP, a finding in line with ours. In contrast to Battaglia Parodi et al who examined 19 patients with STGD, our study revealed an enlarged FAZ in the SCP compared with the DCP, suggesting that the vascular damage is exacerbated in the inner retinal layers. This discrepancy may be attributed to the older and more advanced cohort analysed in our study which may exhibit more pronounced vascular decay in the SCP secondary to neuronal loss. However, the small sample sizes of our study and the aforementioned studies preclude definitive conclusions. Further studies are needed to determine any differences between the SCP and DCP with confidence. Nevertheless, our study supports the aforementioned OCTA investigations revealing a reduced vascular density in the inner retina, and demonstrates that these vascular changes worsen with time and advancing disease stages. It is currently unknown whether the decay of the inner retinal vessels in STGD is a primary or secondary consequence of the weakening metabolic demand in the degenerating neuroretina. The hypothesis of primary regression of the vascular tissue with secondary loss of neurons, however, is unlikely since the ABCA4 gene is predominantly expressed in
photoreceptor cells and only weakly in endothelial cells in the central nervous system. Nevertheless, it is a possibility that cannot be entirely ruled out, and future studies are necessary to examine patients with STGD at a very early disease stage to prove or disprove that hypothesis. On the other hand, the vascular decay in the retina could be attributable to a reduced metabolic demand in the retina caused by progressing retinal degeneration. This hypothesis would be in line with findings demonstrating a correlation between macular vessel density and retinal atrophy and would be supported by our study showing a correlation between the disease duration and vascular degeneration. In any case, the considerable vascular decay in the inner retina may constitute an important feature of STGD pathology and may specifically exacerbate the progressive degeneration of photoreceptors and inner retinal neurons over the disease course. This finding is particularly important in the context of promising therapeutic interventions such as gene therapy or stem cell transplantation, which need to be administered at an early disease stage or at least guided by the vascular pattern as visualised on OCTA.

We acknowledge that our analysis has limitations, including subjective classification of the type of disease and the absence of longitudinal follow-up. Due to our cohort's small size, the group size of the four different STGD stages varies between 2 and 22 eyes, which makes the classes harder to compare. Furthermore, our study enrolled only one patient presenting S4, which may have led to an overestimating or underestimating the OCTA findings at that stage. Using two eyes of a single patient might result in within-subject correlation. Nevertheless, since STGD is a rare disease, and although our cohort managed to attain a good size compared with previous studies, sample size is still small. Therefore, we decided, as did other investigators, to include both eyes of the patients with STGD in our study. We performed additional analyses only including the right eye of the patients. Due to smaller sample sizes, not all analyses that resulted in a significant level when using all eyes also reach significance when only using the right eye. Nevertheless, a trend toward significance becomes obvious in all our analyses (data not shown). Although carrying out multiple statistical tests in our study, we did not correct the p values due to small sample sizes. Therefore, some of the significant p values might be spurious. Furthermore, different dimensions of the OCTA scans may lead to different quantitative values. Nevertheless, in view of the higher precision of the 6×6 mm² volume scan, we decided to prefer it, and it was only whenever the area to be measured exceeded the size of that scan that we opted to use the 9×9 mm² volume scan. Furthermore, since 9×9 mm² was the largest frame available, the areas of rarefied flow and atrophy in ORCC and CC may have been underestimated in some patients and therefore potential differences in eyes affected by large areas of rarefied flow and atrophy went undetected. Despite the existing body of evidence suggesting that the retinal plexuses merge at the edge of the FAZ, which may be thus considered a singular structure throughout the entire foveal thickness and increased variability of measurements when assessing the FAZ size at different segments, we chose to investigate the FAZ area separately in different retinal capillary plexuses (SCP and DCP) so as to compare our findings to the latest literature. As a result of choriocapillary atrophy, the large choroidal vessels are detected in the corresponding areas. Due to this limitation which prohibits a more objective measurement of, for example, perfusion density or vessel length density, we decided to manually detect the area of vascular changes/atrophy, as described before. Nevertheless, a more objective means of detecting vascular changes would be desirable. To the best of our knowledge, however, there is no such analysis procedure available that does justice to the present situation of patients with STGD. In light of these limitations, further prospective studies offering increased power, improved OCTA imaging technology and longitudinal follow-up data are warranted to determine the vascular phenotype of STGD with confidence.

In summary, to the best of our knowledge this is the first and largest cohort-based study investigating vascular changes in different stages of STGD using dSS-OCTA technology, which enables improved visualisation of the deep retinal and choriocapillary layers and thus fewer image artefacts. Our study confirms the essential findings of previous studies using angiography or SD-OCTA and illustrates with more detail the considerable vascular alterations in all layers of the retina and choroid in STGD which increase with the disease’s duration and advancing stage, indicating progressive vascular decay over the disease course. Furthermore, we demonstrated that increased vascular damage is associated with decreased vision. Further prospective and especially longitudinal OCTA studies are required to validate our results, which may have a significant impact on individual prognosis and the optimal timing for possible therapeutic interventions.

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