ABSTRACT

When epithelial cells are exposed to potentially threatening external stimuli such as allergens, bacteria, viruses, and helminths, they instantly produce “alarmin” cytokines, namely, IL-33, IL-25, and TSLP. These alarmins alert the immune system about these threats, thereby mobilizing host immune defense mechanisms. Specifically, the alarmins strongly stimulate type-2 immune cells, including eosinophils, mast cells, dendritic cells, type-2 helper T (T\textsubscript{H2}) cells, and type-2 innate lymphoid cells (ILC2s). Given that the alarm-raising role of IL-33, IL-25, and TSLP was first detected in allergic and infectious diseases, most studies on alarmins focus on their role in these diseases. However, recent studies suggest that alarmins also have a broad range of effector functions in other pathological conditions, including psoriasis, multiple sclerosis, and cancer. Therefore, this review provides an update on the epithelium-derived cytokines in both allergic and non-allergic diseases. We also review the progress of clinical trials on biological agents that target the alarmins and discuss the therapeutic potential of these agents in non-allergic diseases.

Keywords: Alarmins; Hypersensitivity; Autoimmune disease; Biological therapy

INTRODUCTION

The body’s first line of defense against allergens, viruses, bacteria, and helminths consists of the epithelial lining of the skin, intestines, and lungs, which acts as physical barriers. However, recent studies have shown that epithelial cells also play a hitherto poorly understood but apparently vital role in immunological host defense by sensing potential physical/chemical or infectious threats and then initiating immune responses (1). This crucial early role is largely mediated by the epithelial-cell release of IL-33, IL-25, and TSLP, which have been designated “alarmins” (2,3). Once released in response to allergen or infection, these cytokines prime the immune system to produce type-2 immune responses characterized by upregulated eosinophils, mast cells, dendritic cells (DCs), type-2 helper T (T\textsubscript{H2}) cells, and type-2 innate lymphoid cells (ILC2s) (Fig. 1). It is now well known that
### Abbreviations
AD, atopic dermatitis; AHR, airway hyperresponsiveness; CNS, central nerve system; COPD, chronic obstructive pulmonary disease; CRS, chronic rhinosinusitis; CTCL, cutaneous T-cell lymphoma; DC, dendritic cell; EAE, experimental autoimmune encephalomyelitis; IL-17RA, IL-17 receptor A; IL-17RB, IL-17 receptor B; IL-25R, IL-25 receptor; ILC, innate lymphoid cell; IMQ, imiquimod; lTSLP, long-form TSLP; MMP, metalloproteinase; MS, multiple sclerosis; sTSLP, short-form TSLP; Th2, type 2 helper T; TME, tumor microenvironment; TSLPR, TSLR receptor.

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this pathway plays a critical role in the initiation of allergic diseases such as asthma and atopic dermatitis (AD) \(^2\). However, recent studies have discovered that alarmins can also directly regulate many innate and adaptive immune cells and several non-immune cells \(^3\) and in fact are important in various inflammatory diseases that do not involve type-2 responses, including psoriasis, multiple sclerosis (MS), and cancer \(^4\-10\). Therefore, here we will discuss these findings. We will also present the current status of the clinical trials on biologics that target the epithelium-derived alarmins.

### MATURATION OF EPITHELIUM-DERIVED CYTOKINES

#### IL-33
IL-33 is a member of the IL-1 family and binds to heterodimeric receptors composed of IL-1 receptor accessory protein and the transmembrane isoform of ST-2 \(^2\). Once IL-33 binds to its receptor ST2, the cytoplasmic domain of ST2 recruits the adaptor protein MyD88, which ignites NF-κB and AP-1 signaling pathways and induce the expression of pro-inflammatory molecules \(^11\). In homeostatic conditions, the 270-amino acid IL-33 protein is constitutively expressed by epithelial and endothelial cells as a full-length immature form that is sequestered in the nucleus \(^12\). However, once epithelial cells are exposed to an allergen, bacterium, or virus, IL-33 is cleaved into mature forms by either an exogenous or endogenous proteolytic mechanism. The exogenous proteolysis cleaves the central domain of IL-33 and results in the IL-33\(^{96-270}\), IL-33\(^{107-270}\), and/or IL-33\(^{109-270}\) fragments. This cleavage is mediated by serine and cysteine proteases from either neutrophils and mast cells or allergens (e.g. proteases in fungi, house dust mite, pollen, bacteria, and cockroach) \(^13\-15\). By contrast, the endogenous proteolysis is triggered in epithelial cells when they directly detect...
an allergen: this activates the ripoptosome, an intracellular signaling platform that consists of pro-caspase 8, FADD, and RIP1. Once triggered, caspase 8 activates caspase 3 and 7. These caspases then cleave full-length IL-33 in the C-terminal domain into the IL-33 fragments (16). These exogenous and endogenous cleavage events all result in mature IL-33 fragments in the extracellular environment that bind to and activate a plethora of immune cells, including macrophages, eosinophils, mast cells, T\textsubscript{H}2 cells, Tregs, and ILC2s (17-20).

The exogenous cleavage event significantly increases the biological activity of IL-33: for example, it is 10–30-fold more potent than full-length IL-33 in terms of inducing ILC2s to produce type-2 cytokines (13,14,21). Travers et al. (22) suggested that this is because the mature IL-33 is released in a complex with histones; they observed that the histones synergized with IL-33 when activating the pro-inflammatory cytokine production of mast cells. However, the relative role and significance of cleaved IL-33 endogenously and exogenously remain unclear. Nonetheless, these findings suggest that therapies for diseases that are triggered by IL-33 should target the mature form of IL-33.

**IL-25**

IL-25 is also known as IL-17E and belongs to the 6-member (A–F) IL-17 family. All members bind to a homodimeric or heterodimeric receptor composed of 2 subunits of the 5-member (A–E) IL-17 receptor family (23). IL-25 binds to a heterodimeric receptor (IL-25R) composed of IL-17 receptor A (IL-17RA) and IL-17 receptor B (IL-17RB) (Fig. 2). Ligation of IL-25R recruits the adaptor proteins, such as ACT1 and TRAF6, and then activates NF-\textkappaB, MAPK/ERK, and JNK signaling to upregulate genes involved in proliferation and differentiation (24). Unlike the other IL-17 family members, which promote type-3 inflammation, IL-25 amplifies type-2 immunity in multiple tissues (23,25). Given this unique function, IL-25 has been designated a distinct IL nomenclature.
IL-25 can be secreted from a variety of cells, including lung epithelial cells, brain capillary endothelial cells, and synovial fibroblasts in rheumatoid arthritis patients. This diversity of sources suggests that IL-25 may participate in many diseases (26-28). When IL-25-expressing cells are exposed to external stimuli such as helminths or allergens, IL-25 is secreted as a disulfide-linked homodimer (29,30). Although very little is known about early IL-25 processing and its effect on IL-25 bioactivity, Goswami et al. (31) have shown that IL-25 is cleaved by matrix metalloproteinase (MMP) 7. They also showed that this extracellular cleavage event is important for IL-25 function: the MMP7-cleaved form of IL-25 elevated anti-CD3/28-induced splenocyte expression of IL-4, IL-5, and IL-13 significantly better than full-length IL-25. Moreover, when MMP7 was knocked out, Aspergillus-induced asthma model mice developed less airway hyperresponsiveness (AHR), lower type-2 cytokine levels, and lower eosinophil counts in the bronchoalveolar lavage (31). Thus, IL-25 may resemble IL-33 in the sense that post-translational maturation is needed for its bioactivity. This supports the notion that therapies should target mature alarmins.

**TSLP**

TSLP is a member of the IL-2 cytokine family and is mainly produced by epithelial and stromal cells. It exerts its effector functions by binding to the heterodimeric receptor composed of the TSLP receptor (TSLPR) and IL-7Rα (32,33) (Fig. 2). When TSLP binds to its receptor, JAK1 and 2 are recruited and phosphorylated, and downstream STAT5 is activated to initiate expression of pro-inflammatory genes (34).

TSLP has 2 isoforms with opposing immunomodulatory effects, namely, the anti-inflammatory short-form TSLP (sTSLP) and the pro-inflammatory long-form TSLP (lTSLP). sTSLP is constitutively expressed in the skin, intestine, and lung under homeostatic conditions whereas lTSLP is upregulated in inflammatory environments such as the asthmatic lung and the intestine in ulcerative colitis patients (35,36). Dong et al. (35) reported that treatment with synthetic sTSLP improved house dust mite-induced asthma and airway epithelial cell damage in mice whereas treatment with lTSLP impaired and worsened barrier function. However, to date, the sTSLP receptor has not been fully identified and the regulatory mechanism by which it acts has not been reported in non-human species (37). For this reason, most studies on TSLP have been conducted on lTSLP. Therefore, we will not discuss sTSLP further in this review. All further references to TSLP allude to the lTSLP isoform.

The primary sources of TSLP are the epithelial cells in the lungs, skin, and gastrointestinal tract, although immune cells, such as DCs, mast cells and mucosal T cells can also express it (35,38,39). Like the other alarmins, TSLP also appears to be activated by extracellular proteolytic activity. Thus, Nagarkar et al. (40) reported that proteases in nasal polyp extract degrade full-length TSLP and that the resulting cleaved TSLP enhances the IL-5 produced by IL-1β-stimulated mast cells. Furthermore, Poposki et al. (41) showed that proprotein convertases in nasal polyp extracts generate truncated TSLP products that potently upregulate the ability of DCs and ILC2s to produce CCL17 and IL-5, respectively. However, the exact cleavage sites of TSLP have not yet been identified. Notably, TSLP cleavage has not been observed in non-primates, which suggests that the murine model is inappropriate for examining the effects of truncated TSLP (41).
ROLE OF EPITHELIUM-DERIVED CYTOKINES IN ALLERGIC DISEASES

Asthma

Asthma is an obstructive airway disease that is characterized by sporadically recurring symptoms, including dyspnea, coughing, and wheezing (42). It is underpinned by both genetic and environmental risk factors (Fig. 1), many of which associate strongly with epithelium-derived cytokines (26,43-47). Thus, several genome-wide association studies have shown that single nucleotide polymorphisms in IL33, IL1RL1 (the gene expressing the IL-33 receptor ST2), and TSLP associate with asthma severity and blood eosinophil counts (45-47).

While associations between IL25 polymorphisms and asthma have not been found to date, an analysis of bronchial biopsies from asthma patients has shown that allergen inhalation increases IL-25-and IL-25R-expressing cell frequencies and that these frequencies correlate negatively with maximum change in forced expiratory volume in 1 second (FEV1) % (48). Moreover, bronchial epithelial biopsies from asthma patients have higher IL-33, IL-25, and TSLP protein levels than healthy controls (49-51). When murine models of allergic asthma are treated with anti-IL-33, anti-IL-25, or anti-TSLP mAbs, they reduce AHR and the type-2 cytokine levels and eosinophil counts in the lung (52-54).

It appears that these alarmins can act indirectly and directly on asthma-associated immune cells, including eosinophils, mast cells ILC2s, effector T\textsubscript{H}2 cells, and Tregs (Fig. 1). Regarding eosinophils, Cherry et al. (18) showed with in vitro experiments that IL-33 promotes eosinophil superoxide production, degranulation, and survival as potently as the key eosinophil regulator IL-5. This prosurvival effect of IL-33 appears to relate to its ability to induce the autocrine secretion of GM-CSF in eosinophils (55). Moreover, Salter et al. (56) found that the eosinophilic potential of supernatants from cultured bronchial epithelial cells (as measured by adding the supernatants to non-adherent mononuclear cell cultures) was much higher when the epithelial cells were from asthma patients rather than from healthy controls; in addition, this eosinophilic potential was extinguished by antibodies against TSLP. Thus, IL-33 and TSLP can directly influence eosinophil formation and activity. For mast cells, IL-33 induces mast-cell maturation and degranulation and enhances their adherence to fibronectin in vitro (57,58). TSLP induces mast-cell production of cytokines (IL-5, IL-6, and IL-13) and chemokines (CXCL8 and CCL1) in vitro but only when IL-1β and TNF are also present (43).

Not only granulocytes but also tissue-resident innate lymphocytes, particularly ILCs, can mediate type 2 inflammation in an immediate response to alarmins (59-61). Kim et al. (62) found with in vitro experiments that ILCs are directly activated by IL-33 produced by alveolar macrophages, DCs, and type-2 pneumocytes. Their in vivo experiments using ST2-deficient mice then showed that activated ILCs are needed to induce the AHR and airway inflammation in glycolipid-induced asthma (62). Two studies have also shown that when ovalbumin-induced asthma model mice are treated with neutralizing IL-25 Ab during the antigen challenge period, it markedly abrogates the development of AHR and the IL-5 and IL-13 levels and ILC2 counts in the lung (52,63). In addition, TSLP receptor-deficient mice showed reduced BAL eosinophilia and type 2 cytokine production from ILC2 after Alternaria alternata challenge (64). Interestingly, Liu et al. (65) found that the ILC2s from the TSLP-rich bronchoalveolar lavage of asthma patients are more resistant to steroids than the circulating ILC2s and that this resistance can be reversed in vitro by clinically available inhibitors of MEK and STAT5. Kabata et al. (66) also found that TSLP, but not IL-25 or IL-33, induces the steroid...
resistance of ILC2s in the lung of asthma model mice, and that this is mediated via STAT5 phosphorylation and anti-apoptotic Bcl-xL expression. Thus, targeting TSLP could be an alternative therapeutic strategy for uncontrolled steroid-resistant asthma.

Although receiving less attention than innate lymphocytes, T lymphocytes are also affected by epithelium-derived alarmins at sites of inflammation. It has been shown that while mice that are deficient in all 3 alarmins have normal T-cell priming in the lymph node, effector T_{H2} cells cannot undergo terminal differentiation in peripheral tissues (67). In an in vitro T cell differentiation system, each alarmin directly enhances T_{H2} differentiation (68-70). Besides T helper cells, Treg cells, which are pivotal in maintaining immune tolerance against innocuous external agents (71,72), are also affected by alarmins in allergic conditions: Chen et al. (73) showed that lung-resident Tregs express ST-2 and respond to intranasal IL-33 administration by upregulating GATA3, IL5, and IL13 expression; consequently, these cells come to resemble “T_{H2}-like” Tregs and associate with impaired airway tolerance and increased airway inflammation. Thus, the epithelial-derived alarmins may play multiple roles in the development of asthma. Further studies on these roles are warranted.

AD
AD is an allergic type of eczema characterized by epidermal hyperplasia and crusted, inflamed, highly pruritic, erythematous, and oozing vesicular lesions (74). AD shares many of the pathological features of asthma, including T_{H2}-dominant immune responses and elevated plasma IgE levels (75). Several clinical and pre-clinical studies suggest that targeting alarmins in AD may have therapeutic potential because they appear to promote the disruption of the barrier epithelium, as follows (76-81). First, several studies show that AD associates with overexpression of IL-33 and TSLP by local keratinocytes and IL-25 overexpression by dermal DCs (38,76-79). Second, studies have found that 1) IL-33, IL-25, or TSLP treatment directly reduces the AD keratinocyte expression of skin barrier proteins (claudin-1 and/or filaggrin) in vitro (82-85); 2) IL-25R-deficient mice have better barrier integrity and thinner dermal epithelium than wild-type mice after AD induction (86); 3) keratinocyte-specific overexpression of TSLP and IL-33 elicit type-2 inflammation and AD-like phenotypes in mice (87,88); and 4) disruption of the barrier epithelium can promote AD by inducing trans-epithelial water loss, allergen penetration, and bacterial colonization in the skin (89). These findings together suggest that alarmins may act directly on keratinocytes in an autocrine manner and that targeting alarmins therapeutically could ameliorate AD formation and progression. The latter notion is supported by several genome-wide association studies that suggest the susceptibility loci for AD-associated with epithelial barrier integrity (90-93).

Several studies also show that ILC2s are key cellular players in AD pathogenesis. First, ILC2s appeared to be enriched in skin lesions from AD patients (80,81,94). Second, depletion of skin ILC2s reduces the skin thickness and type-2 cytokine production that is caused by topical application of MC903 (calcipotriol, a vitamin D analog), which induces AD-like inflammation (81). Thus, alarmins may contribute to AD via their well-known ability to stimulate ILCs. This is supported by Kim et al., (80) who reported that knocking out TSLP (but not IL-33 or IL-25) blocks MC903-induced AD. Interestingly, a very similar study showed that knocking out IL-25 or IL-33 attenuates the skin inflammation and ILC2 infiltration in MC903-induced AD better than knocking out TSLP (81). The reason for this discrepancy between studies is unclear. Nonetheless, these studies suggest that targeting alarmins may improve AD.

Two studies have also suggested that blocking alarmins could improve pruritis in AD.
Pruritus is mediated by somatosensory neurons whose bodies are located in the dorsal root ganglia. Liu et al. (8) and Wilson et al. (95) showed that dorsal root ganglion neurons can express the alarmin receptor molecules ST-2 and TSLPR, and that IL-33 and TSLP bind directly to these cells. Thus, alarmins may promote pruritus by acting as signaling mediators between the epithelium and cutaneous sensory neurons. Given that scratching itself can exacerbate skin inflammation (96), these studies suggest that targeting alarmins may improve not only pruritis but also AD inflammation.

**ROLE OF EPITHELium-DERIVED CYTOKINES IN AUTOIMMUNE DISEASES**

**Psoriasis**
Psoriasis is a chronic autoimmune skin disease that is characterized by epidermal hyperplasia, dermal infiltration with immune cells, and increased dermal capillary density; this leads to scaly red skin plaques (97). Unlike asthma and AD, psoriasis is considered to be a type-17 disease, meaning that it is mediated by IL-17, IFN-γ, and IL-23 produced by Th17 cells and neutrophils (10,98-103). Thus, these molecular and cellular entities are the key therapeutic targets in psoriasis. Nonetheless, multiple clinical studies have hinted that alarmins may also participate in the pathophysiology of psoriasis. We will discuss the evidence for each alarmin in turn.

Mitsui et al. (102) showed that psoriasis patients have higher serum levels of IL-33 than healthy controls. Moreover, Zeng et al. (10) found with a murine model of imiquimod (IMQ)-induced psoriasis that keratinocytes in the inflamed skin predominantly produce IL-33 and also express the IL-33 receptor ST-2. This study then showed that mice with IL-33-deficient keratinocytes are less susceptible to IMQ-induced psoriasis (10). In addition, Duan et al. (104) reported that treating HaCaT human epidermal cells with IL-33 increases their proliferation. Besides, IL-33 administration on IMQ-induced psoriasis mice inhibits the expression of autophagy-related proteins from skin inflamed lesions (104): the latter finding is relevant because downregulation of autophagy associates with the progression of several inflammatory and autoimmune diseases, including psoriasis and AD. Thus, similar to the findings in AD, these studies suggest that IL-33 from keratinocytes may induce or exacerbate the epithelial hyperplasia in psoriasis (and AD) in an autocrine fashion. Immune cells may also participate in the IL-33-mediated exacerbation of psoriasis. Hueber et al. (105) showed that intradermal injection of IL-33 in murine ears not only induced an inflammatory psoriasis-like lesion that increased ear thickness, it also elevated the local levels of the neutrophil chemokine CXCL1 and neutrophil counts. Local mast cell counts also rose and were at least partly responsible for the inflammation since mast-cell deficient mice (KitW-sh/W-sh mice) showed a significant delay in the evolution of the psoriatic phenotype (105).

Like the IL-33, both IL-25 and TSLP were also involved in the pathogenesis of psoriasis. Papp et al. (9) showed that treating psoriasis patients with an IL-17RA (1 of the 2 IL-25 receptor subunits) blockade had strong therapeutic effects on psoriasis. Moreover, Suto et al. (106) showed that IL-25 stimulates DCs to produce IL-1β, and this is essential for Th17 cell-mediated contact hypersensitivity since Il25−/− mice are protected from this inflammatory response. Thus, the IL-25/IL-17R axis may be an effective target for psoriasis therapeutics.

Regarding TSLP, Suwarsa et al. (103) found that psoriasis patients have higher serum levels
of TSLP than healthy controls while Volpe et al. (107) reported that the keratinocytes in untreated skin lesions from psoriasis patients strongly expressed TSLP. Notably, the latter study also found that TSLP works synergistically with CD40L to activate skin and blood DCs and that once activated in this manner, these DCs express high levels of IL-23 in vitro (107). This is significant because IL-23 (a heterodimeric cytokine composed of IL-12p40 and IL-23p19) induces epidermal acanthosis in mice when injected intradermally. Moreover, blocking IL-23 with a mAb against IL-24p40 reduces psoriasis symptoms in patients. Since IL-23 induces and activates both type-17 T cells and ILC3s, these findings suggest that although TSLP is known to promote type-2 immune responses, it can also aid type-17 inflammation. Additional studies suggest that keratinocytes are not only a key source of skin TSLP (38), they also express TSLPR (108). This suggests that keratinocyte-produced TSLP not only has paracrine immune effects, it also affects keratinocyte behavior in an autocrine manner. This is supported by an in vitro study that showed that treatment of primary keratinocytes with recombinant TSLP induces their proliferation in a dose-dependent fashion (109). Thus, these studies together suggest that IL-33, IL-25, and TSLP also contribute to psoriasis pathogenesis and that blocking alarmin signaling may be a therapeutic target in psoriasis.

**MS**

MS is a neurodegenerative disease that is characterized by demyelination and neurodegeneration in the central nerve system (CNS) (110). Analyses of IL-33 reporter mice showed that IL-33 is constitutively expressed in the corpus callosum, hippocampus, thalamus, and cerebellum (111); moreover, the astrocytes in both the brain and spinal cord express high levels of this alarmin (112). The involvement of IL-33 in the pathogenesis of MS is supported by increased IL-33 and/or ST2 expression in CNS lesions of MS patients (113,114) and experimental autoimmune encephalomyelitis (EAE), a mouse model for human MS (7,115). IL-33 was elevated in plasma (116,117), cerebrospinal fluid (117), and brain tissue (113,114) from MS patients. Several studies suggest that blocking IL-33 signalings can protect against MS. Li et al. (7) reported that IL-33 blockade reduces the development of EAE and that this effect is mediated by inhibition of T,1-like and/or T,17-related immune responses. Similarly, when myelinating rat neurospheres are treated in vitro with recombinant IL-33, their axon myelination is inhibited (113). As the defect on axon myelination is the major feature of the MS, accumulated IL-33 may be involved in the initiation and aggravation of MS. However, other studies have found the opposite. Jiang et al. (115) showed that in vivo administration of IL-33 attenuates the development of EAE by promoting T,2 cell and M2 macrophage polarization. Moreover, genetic deletion of IL33 in oligodendrocyte precursor cells impairs oligodendrocyte maturation and differentiation (118). IL-33 receptor-deficient (St2−/−) mice also develop more severe EAE and exhibit enhanced inflammatory T cell accumulation in the brain and spinal cord (119). Thus, the therapeutic potential of targeting the IL-33/ST-2 axis in MS remains unclear.

Unlike the controversial roles of IL-33, IL-25 may play a protective role in MS since MS patients have very low serum IL-25 levels (120). Moreover, IL-25-deficient (Il25−/−) mice are highly susceptible to EAE, and this susceptibility is blocked when the IL-25 receptor IL-17A is neutralized (6). In addition, when T cells are activated in vitro by anti-CD3/28 in the presence of IL-25, they lose their ability to kill fetal neurons. This effect is mediated by IL-25-induced inhibition of LFA-1 on the T cells, which generates the cytotoxic immune synapse between T cells and their target cells (121). In addition, lentiviral-mediated administration of IL-25 into the CNS reduces the neuroinflammation in both EAE and the entorhinal cortex lesion model of neuroinflammation; this effect is due to IL-25-induced shifting of the microglia from the pro-inflammatory M1 to the anti-inflammatory M2 phenotype (122). Thus, IL-25 could
potentially be a therapeutic target for MS.

There is some limited evidence that suggests TSLP signaling pathways contribute to MS pathogenesis. Eckhardt et al. (123) showed that when EAE is induced in TSLP-deficient (Tslp−/−) mice by myelin oligodendrocyte glycoprotein injections, the disease is both delayed and attenuated. Moreover, these mice display impaired differentiation of T cells into myelin oligodendrocyte glycoprotein-specific effector and memory T cells (123). Yu et al. (124) showed that TSLP receptor-deficient (Tslpr−/−) mice also exhibit resistance to EAE. Moreover, they found that TSLPR signaling induces neuroinflammation by activating Janus kinase, which then hyperactivates the NLR family pyrin domain containing 3 inflammasome (124), which in turn drives systemic T helper 1 and T helper 17 inflammation and chemotaxis of immune cells to the CNS. Thus, blocking TSLP signaling may be a prospective target for MS.

These studies suggest that epithelial-derived alarmins could participate in MS. However, the limited research on IL-25 and TSLP and the discrepant research on IL-33 mean that further studies are needed to determine the therapeutic potential of these cytokines in MS.

Cancer
Cancer is characterized by the development of abnormal cells that divide uncontrollably and can penetrate and destroy normal body tissues. These features of cancer cells are intimately controlled by the tumor microenvironment (TME), which participates in all stages of tumorigenesis from initiation to metastasis (125). Cytokines that mediate cell-cell communications, including the alarmins, are central to this role of the TME.

IL-33 levels are elevated in the serum and tumor tissues of patients with a wide range of cancers, including glioma (126), head and neck squamous cell carcinoma (127), gastric cancer (128), colorectal cancer (129), hepatocellular carcinoma (130), uterine leiomyoma (131), non-small cell lung cancer (132), and breast cancer (133). Additional studies on IL-33 have shown that this alarmin can both promote and suppress tumorigenesis. Both effects are mediated by the ability of IL-33 to shape innate and adaptive immunity, thereby converting TME to a pro-tumoral or anti-tumoral environment. Below, we will first discuss the studies on the pro-tumoral activities of IL-33.

Yang et al. (133) showed that the serum IL-33 levels in breast cancer patients correlate positively with their serum concentrations of molecules that are well known to associate with tumor-related angiogenesis (vascular endothelial growth factor), matrix remodeling (MMP-11), and growth and survival (platelet-derived growth factor-C). Similarly, Jovanovic et al. (134) showed that treating IL-33 on a murine breast cancer model accelerates tumor growth and metastasis and that this is mediated by increased intratumoral accumulation of myeloid-derived suppressor cells and Treg cells. Moreover, Liu et al. (129) reported that colorectal cancer cells express higher IL-33 and ST-2 levels when they are derived from patients with a poor tumor, nodes, metastasis stage, which signifies the extent to which the tumor has spread. In addition, Fang et al. (4) showed that when IL-33 transgenic mice are inoculated with murine colon cancer cells, they exhibit increased tumor growth, which is induced by macrophage recruitment to the TME and stimulating prostaglandin E2 production from recruited macrophages.

Contrary to the pro-tumor effects of IL-33, this cytokine can also have a tumor-suppressive function. The studies on the tumor-suppressor functions of IL-33 show that this suppression
is due to IL-33-modulated activation of either cytotoxic immune cells, the pro-inflammatory T\(_9\) cell subset, or ILC2s. With regard to the studies on IL-33-induced cytotoxicity, Gao et al. (135) showed that artificially upregulating IL-33 expression in cancer cells inhibits tumor growth by increasing CD8\(^+\) T and NK cell numbers in tumor tissues. Similarly, IL-33 transgenic mice display significantly increased tumor infiltration of CD8\(^+\) T and NK cells, and recombinant IL-33 treatment increases CD8\(^+\) T- and NK-cell cytotoxicity \textit{in vitro} (136). Several studies also show that IL-33 can increase anti-tumor immune activity indirectly by increasing the ability of DCs to either cross-present leukemia or melanoma Ags to CD8\(^+\) T cells (137,138) or express OX40L, which induces T\(_9\) cells that promote anti-tumor immune responses (139). Moreover, with regard to the studies showing that IL-33 fosters the anti-tumoral functions of ILC2s, Moral et al. (140) found that 1) human and mouse pancreatic ductal adenocarcinomas bear increased intratumoral ILC2 frequencies, 2) IL-33 deletion increases the growth of the tumors \textit{in vivo}, and 3) IL-33 acts by activating ILC2s, which recruit CD103\(^+\) DCs, which in turn prime tumor-specific CD8\(^+\) T cells and recruit them into the tumor. In addition, Ikutani et al. (141) reported that ILC2s promote tumor recruitment of eosinophils, which have anti-tumoral activity. Notably, a recent study that used the B16.F10 melanoma model showed that the acidification of the TME by lactic acid produced by tumor cells facilitates cancer growth by impairing ILC2 survival and function. This mechanism explains why the expression of lactate dehydrogenase A (the enzyme that produces lactic acid) in human cutaneous melanoma samples correlates negatively with ILC2 marker expression (142).

In line with IL-33, IL-25 also has a Janus face in tumor immunity. Its pro-tumoral role is mainly mediated by inducing CD4\(^+\) T cells to differentiate into T\(_{H2}\) cells, which can promote tumor progression. Thus, Jiang et al. (143) showed first that IL-25 is produced in the TME of breast cancer by CD4\(^+\) T cells and F4/80\(^+\) macrophages. Subsequently, they found that IL-25 neutralization reduces these cell frequencies, T\(_{H2}\) responses and M2 polarization in the TME. These changes enhance tumor progression (143). Nakajima et al. (144) observed a similar relationship between IL-25, T\(_{H2}\) responses, and cancer progression: they found that the keratinocytes in cutaneous T-cell lymphoma (CTCL) skin lesions express high levels of IL-25 and that PBMCs in patients with an aggressive form of this cancer express IL-25 receptors and produce high levels of the type-2 cytokine IL-13.

In contrast, the tumor-suppressor functions of IL-25 are demonstrated by Ferretti et al., (145) who showed that human germinal center-derived non-Hodgkin B cell lymphomas express the IL-25 receptor, IL-17RB, and that treating model mice with these tumors with IL-25 inhibits tumor growth by suppressing pro-angiogenic molecules. Moreover, Furuta et al. (146) showed that IL-25 treatment of breast cancer cells can induce their apoptosis \textit{in vitro}, and that this is mediated by IL-25 binding to IL-25 receptor. Interestingly, they found that the intracellular region of the IL-25 receptor bears a death domain-like segment that is recognized by death domain adaptor proteins, which subsequently activate apoptosis (146).

Like the other 2 alarmins, TSLP both promotes and suppresses tumors. Ragonnaud et al. (147) demonstrated that TSLP is secreted from breast cancer cell lines and supports pre-B cell proliferation in the bone marrow, ultimately leading to cancer metastasis. Xie et al. (148) reported that cervical cancer cells also secrete TSLP and that stimulates cancer cells’ \textit{in vitro} growth in an autocrine manner. The same group also showed with \textit{in vitro} experiments that cervical cancer-derived TSLP may induce the recruitment of eosinophils which produces anti-inflammatory cytokines, further promotes cervical cancer cell proliferation, and reduces their apoptosis (149). Moreover, Takahashi et al. (150) reported that TSLP is expressed by CTCL
lesional skin TSLP enhances CTCL progression by directly stimulating cancer cell growth and by inducing a T_{\text{H}2}-dominant TME that is hostile to T_{\text{H}1} anti-tumor immune responses. This was supported by a study on human pancreatic cancer cells, which found that the source of TSLP in the cancer lesions is cancer-associated fibroblasts; these cells were shown to activate TSLP^+ DCs, which then induce T_{\text{H}2} differentiation (151).

The studies showing that TSLP can also have anti-tumor effects include that by Demehri et al., (152) who observed that the keratinocyte-specific overexpression of TSLP in K14-TSLP-transgenic mice leads to T_{\text{H}2} cell-mediated inflammation that blocks the spontaneous development of metastatic breast cancer in MMVT-pyMT mice. This suggests that the TSLP can prevent the early stages of tumorigenesis (152). Moreover, an analysis of NCBI GEO datasets by Yue et al. (153) showed that human colon adenomas express lower levels of TSLP than normal mucosa. This was confirmed by analysis of 40 additional colon adenomas, which also showed that the clinical severity of the cases correlated with lower TSLP expression. They then reported that TSLP treatment induces primary human colonic cancer cells to undergo caspase-8-initiated apoptosis in vitro and that peritumoral injection of TSLP suppresses colon cancer growth in vivo (153).

Given these mixed pro- and anti-tumoral roles of all 3 epithelial cell-derived cytokines, it remains unclear whether targeting these molecules could have therapeutic value. Nonetheless, the research in this area has greatly extended our view of alarmins: they are no longer seen as mere inducers of an immune response, rather, it is clear that they can modulate cancer cell growth both directly via receptor ligation and indirectly by shaping the immune status in the TME. Therefore, further research that untangles these roles is warranted.

DEVELOPMENT OF BIOLOGICS AND ON-GOING CLINICAL TRIALS TARGETING EPITHELIUM-DERIVED CYTOKINES

Although the roles of epithelium-derived cytokines in some diseases remain controversial, many mouse studies and clinical findings in allergic diseases have confirmed their therapeutic possibilities. In particular, these studies have shown that the alarmins can induce more extensive and potent allergic responses than downstream mediators such as IgE or type-2 cytokines. This suggests that targeting the alarmins may result in better immunomodulatory effects than the existing therapeutics. This has led to multiple clinical trials on alarmins. Most have been or are being conducted on allergic diseases due to the well-established roles of IL-33, IL-25, and TSLP in these diseases. The trial drugs are Astegolimab (154-159), Etokimab (160-163), GS-K3772847 (164,165), Itepekimab (166-173), and MEDI3506 (174-177), which target the IL-33/ST-2 pathway (Table 1), and Tezepelumab (178-194) and CSJ117 (195-198), which target the TSLP pathway (Table 2). Although blocking the IL-25 pathway is also a possible target, there are no clinical trials on agents against IL-25 at present. While biologics that target IL-33 and TSLP have not yet received FDA approval, some have promising results, as detailed below, and are now in phase-3 clinical trials.

IL-33/ST-2 signaling pathway blockade

Astegolimab (MSTT1041A/RG 6149) is a human IgG2 mAb that blocks IL-33 signaling by targeting its receptor ST-2. Of the 6 trials on Astegolimab, 2 are in phase-1 studies for healthy subjects, patients with mild asthmatics (154), or chronic rhinosinusitis (CRS) with nasal polyps (155). The remainders are phase-2 studies on patients with AD (157), chronic
Table 1. Clinical trials that target IL-33/ST-2

| Study title                                                                 | Reference | Identifier   | Stage | Drug | Disease                      | Status            |
|-----------------------------------------------------------------------------|-----------|--------------|-------|------|------------------------------|-------------------|
| A first-in-human, double blind, single dose study in healthy subjects and subjects with mild atopic asthma | (154)     | NCT01928368 | I     | Astegolimab | Asthma                        | Completed         |
| A study to evaluate the safety, tolerability, pharmacokinetics, and pharmacodynamics of AMG 282 in healthy subjects and subjects with chronic rhinosinusitis with nasal polyps | (155)     | NCT02170337 | I     | Astegolimab | CRS with NP                   | Completed         |
| A study to assess the efficacy and safety of MSTT1041A in participants with uncontrolled severe asthma | (156)     | NCT02918019 | II    | Astegolimab | Asthma                        | Completed         |
| A study to assess the efficacy and safety of MSTT1041A in participants with moderate to severe atopic dermatitis | (157)     | NCT03747575 | II    | Astegolimab | AD                           | Completed         |
| Anti-ST2 (MSTT1041A) in COPD (COPD-ST2OP)                                   | (158)     | NCT03615040 | II    | Astegolimab | COPD                         | Completed         |
| A study to evaluate the efficacy and safety of astegolimab in participants with chronic obstructive pulmonary disease | (159)     | NCT05037929 | II    | Astegolimab | COPD                         | Recruiting        |
| A study investigating the efficacy, safety, and PK profile of ANB020 administered to adult subjects with moderate-to-severe AD (ATLAS) | (160)     | NCT03533751 | II    | Etokimab    | AD                           | Recruiting        |
| Proof of concept study to investigate ANB020 activity in adult patients with severe eosinophilic asthma | (161)     | NCT03469934 | II    | Etokimab    | Asthma                        | Completed         |
| Etokimab in adult patients with chronic rhinosinusitis with nasal polyps (CRS+NP) | (162)     | NCT03614923 | II    | Etokimab    | CRS with NP                   | Completed         |
| Placebo-controlled study to investigate ANB020 activity in adult patients with peanut allergy | (163)     | NCT02920021 | II    | Etokimab    | Peanut allergy                | Completed         |
| A study to evaluate the safety and tolerability, pharmacokinetics (PK) and pharmacodynamics (PD) of Melrilimab (GSK3772847) in healthy participants | (164)     | NCT04366349 | I     | GSK3772847  | Healthy/asthma                | Completed         |
| Efficacy and safety study of GSK3772847 in subjects with moderately severe asthma | (165)     | NCT03207243 | II    | GSK3772847  | Asthma                        | Completed         |
| Study of REGN3500 and dupilumab in patients with asthma                     | (166)     | NCT03112577 | I     | Itepekimab  | Asthma                        | Completed         |
| Study of safety, tolerability, and pharmacokinetics of multiple ascending doses of REGN3500 in adults with moderate asthma | (167)     | NCT02999711 | I     | Itepekimab  | Asthma                        | Completed         |
| Evaluation of SAR440340 and as combination therapy with dupilumab in moderate-to-severe asthma participants | (168)     | NCT03387852 | II    | Itepekimab  | Asthma                        | Completed         |
| Efficacy, safety, and pharmacokinetic profiles of REGN3500 administered to adult patients with moderate-to-severe atopic dermatitis | (169)     | NCT03738423 | II    | Itepekimab  | AD                           | Completed         |
| Efficacy and safety of REGN3500 monotherapy and combination of REGN3500 plus dupilumab in adult patients with moderate-to-severe atopic dermatitis | (170)     | NCT03736967 | II    | Itepekimab  | AD                           | Terminated (Lack of efficacy) |
| Proof-of-concept study to assess the efficacy, safety and tolerability of SAR440340 (anti-IL-33 mAb) in patients with moderate-to-severe chronic obstructive pulmonary disease (COPD) | (171)     | NCT03546907 | II    | Itepekimab  | COPD                         | Completed         |
| Study to assess the efficacy, safety, and tolerability of SAR440340/REGN3500/Itepekimab in chronic obstructive pulmonary disease (COPD) (FINER-COPD) | (172)     | NCT04701983 | III   | Itepekimab  | COPD                         | Recruiting        |
| Safety and tolerability of MEDI3506 in healthy participants, in patients with COPD and healthy Japanese participants | (173)     | NCT04751487 | III   | MEDI3506    | Healthy/COPD                  | Completed         |
| Study to assess the efficacy and safety of MEDI3506 in adults with uncontrolled moderate-to-severe asthma (FRONTIER-3) | (174)     | NCT0396795  | I     | MEDI3506    | Healthy/COPD                  | Completed         |
| Efficacy and safety of MEDI3506 in adult subjects with atopic dermatitis | (175)     | NCT04570657 | II    | MEDI3506    | Asthma                        | Recruiting        |
| A phase II, randomized, double-blind, placebo-controlled study to assess MEDI3506 in participants with COPD and chronic bronchitis (FRONTIER-4) | (176)     | NCT04212169 | II    | MEDI3506    | COPD/chronic bronchitis       | Recruiting        |

NP, nasal polyp.
*Stage I, II, and III represent phase 1, phase 2, and phase 3 respectively.

obstructive pulmonary disease (COPD) (158,159), or asthma (156). The latter study has shown that Astegolimab significantly reduces the asthma exacerbation rates in a broad range of adult patients with severe asthma, including those with low eosinophil counts and those with inadequately controlled severe asthma (156,199).

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https://doi.org/10.4110/in.2022.22.e11
### Table 2. Clinical trials that target TSLP

| Study title                                                                 | Reference | Identifier   | Stage | Drug    | Disease     | Status    |
|----------------------------------------------------------------------------|-----------|--------------|-------|---------|-------------|-----------|
| A study to evaluate the pharmacokinetics of MEDI9929 (AMG 157) in adolescents with mild to moderate asthma | (178)     | NCT02512900  | I     | Tezepelumab | Asthma      | Completed |
| Study to evaluate the pharmacokinetics of tezepelumab in children with asthma (TRAILHEAD) | (179)     | NCT04673630  | I     | Tezepelumab | Asthma      | Recruiting |
| Double-blind, multiple dose study in subjects with mild atopic asthma       | (180)     | NCT01405963  | I     | Tezepelumab | Asthma      | Completed |
| Safety study of AMG 157 in healthy subjects and subjects with atopic dermatitis | (181)     | NCT00757042  | I     | Tezepelumab | Healthy/AD  | Completed |
| Study to evaluate the efficacy and safety of MEDI9929 (AMG 157) in adult subjects with inadequately controlled, severe asthma | (182)     | NCT02054130  | II    | Tezepelumab | Asthma      | Completed |
| Effects of anti-TSLP in patients with asthma (UPSTREAM)                     | (183)     | NCT02698501  | II    | Tezepelumab | Asthma      | Completed |
| Study to evaluate tezepelumab on airway inflammation in adults with uncontrolled asthma (CASCADE) (CASCADE) | (184)     | NCT03688074  | II    | Tezepelumab | Asthma      | Completed |
| Phase 2a study to evaluate the efficacy and safety of MEDI9929 in adults with atopic dermatitis (ALLEVIAD) | (185)     | NCT02525094  | II    | Tezepelumab | AD          | Completed |
| Anti-TSLP (AMG 157) plus antigen-specific immunotherapy for induction of tolerance in individuals with cat allergy | (186)     | NCT02237196  | I/II  | Tezepelumab | Cat allergy/cat hypersensitivity | Completed |
| Study to evaluate tezepelumab in adults with chronic spontaneous urticaria (INCEPTION) | (187)     | NCT04833855  | II    | Tezepelumab | Chronic spontaneous urticaria | Recruiting |
| Tezepelumab COPD exacerbation study (COURSE)                               | (188)     | NCT04039113  | II    | Tezepelumab | COPD        | Recruiting |
| Study to evaluate tezepelumab in adults & adolescents with severe uncontrolled asthma (NAVIGATOR) | (189)     | NCT03347279  | III   | Tezepelumab | Asthma      | Completed |
| Study to evaluate the efficacy and safety of tezepelumab in reducing oral corticosteroid use in adults with oral corticosteroid dependent asthma (SOURCE) | (190)     | NCT03406078  | III   | Tezepelumab | Asthma      | Completed |
| Long-term safety of tezepelumab in Japanese subjects with inadequately controlled severe asthma (NOZOMI) | (191)     | NCT04048343  | III   | Tezepelumab | Asthma      | Completed |
| Extension study to evaluate the safety and tolerability of tezepelumab in adults and adolescents with severe, uncontrolled asthma (DESTINATION) | (192)     | NCT03706079  | III   | Tezepelumab | Asthma      | On going (active) |
| Study to evaluate tezepelumab in adults with severe uncontrolled asthma (DIRECTION) | (193)     | NCT03927157  | III   | Tezepelumab | Asthma      | Recruiting |
| Efficacy and safety of tezepelumab in participants with severe chronic rhinosinusitis with nasal polypysis (WAYPOINT) | (194)     | NCT04851964  | III   | Tezepelumab | CRS with NP | Recruiting |
| A bronchoprovocation study to assess the safety, tolerability, pharmacokinetics and pharmacodynamics of CSJ117 in adult subjects with mild atopic asthma | (195)     | NCT03138811  | I     | CSJ117    | Asthma      | Completed |
| Study of efficacy and safety of CSJ117 in patients with severe uncontrolled asthma | (196)     | NCT04410523  | II    | CSJ117    | Asthma      | Recruiting |
| Study of safety of CSJ117 in participants with moderate to severe uncontrolled asthma | (197)     | NCT04946318  | II    | CSJ117    | Asthma      | Recruiting |
| Study of effect of CSJ117 on symptoms, pharmacodynamics and safety in patients with COPD | (198)     | NCT04882124  | II    | CSJ117    | COPD        | Recruiting |

NP, nasal polyp.

*Stage I, II, and III represent phase 1, phase 2, and phase 3 respectively.

Etokimab (ANB020) is a human IgG1 mAb that neutralizes IL-33. It is manufactured by AnaptysBio and has been subjected to 4 completed phase-2 trials on AD (160), asthma (161), CRS with nasal polyps (162), and peanut allergy (163). The results are still pending. An interim analysis of the phase-2 study on asthma showed that Etokimab improved the FEV1 of adults with severe eosinophilic asthma compared to placebo-treated patients (161).

GSK3772847, like Astegolimab, also blocks the IL-33 receptor. It is currently being tested for safety in a phase-1 trial in healthy participants and a phase-2 trial in patients with asthma (164,165). Itepekimab (SAR440340/REGN3500) is a human IgG4 mAb against IL-33. It is the subject of 2 phase-1 trials in asthma patients (166,167), 4 phase-2 trials in asthma (168), AD (169,170), and COPD patients (171), and 2 phase-3 trials in COPD patients (172,173). The results of the phase-2 trial in COPD patients have been reported: although Itepekimab failed to meet the primary endpoint in the overall population, it did reduce the exacerbation rate...
in former smokers with COPD. These patients also demonstrated improved lung function (171,200). MEDI3506 is a newly developed IL-33-targeting mAb. It is currently being studied in 1 phase-1 trial in healthy subjects and patients with COPD (174) and 3 phase-2 trials in asthma (175), AD (176), and COPD patients (177).

**TSLP signaling pathway blockade**

Tezepelumab (MEDI9929/AMG 157) is a human IgG2 mAb that binds TSLP, thereby blocking the TSLP/TSLPR signaling pathway. It is the subject of 17 trials: 5 phase-1 trials in asthma (178-180), AD patients (181), or cat allergy (186); 7 phase-2 trials in patients with asthma (182-184), AD (185), cat hypersensitivity (186), chronic spontaneous urticaria (187), or COPD (188); and 6 phase-3 trials in asthma patients (189-193) or patients with CRS and nasal polyps (194). The phase-2 PATHWAY study showed that Tezepelumab markedly reduced the blood eosinophil counts and serum IL-5 and IL-13 levels in patients with severe and uncontrolled asthma. This suggests that it decreased asthma severity (182,201). In addition, the phase-3 NAVIGATOR trial suggested that Tezepelumab effectively reduced asthma exacerbations and improved lung function in adult patients with severe asthma (189,202). Moreover, the phase-2 ALLEVIAD trial in adults with AD showed that while Tezepelumab did not achieve significant levels in the primary endpoint, the Tezepelumab-treated patients were significantly more likely to achieve >50% improvement in the eczema site severity index (185,203).

CSJ117 is a potent neutralizing Ab fragment that targets human TSLP. It was developed as an inhaled dry powder formulation for targeted delivery to the lungs. It is the subject of 1 phase-1 trial in asthma patients (195) and 3 phase-2 trials in asthma (196,197) or COPD patients (198). An abstract recently reported that CSJ117 attenuated allergen-induced bronchoconstriction and sputum eosinophil counts in patients with mild atopic asthma (204).

**CONCLUSIONS**

This review shows that the epithelium-derived cytokines IL-33, IL-25, and TSLP play roles in a spectrum of diseases that range broadly from allergic disorders to tumors and autoimmune diseases. The roles of these alarmins not only include their well-known ability to initiate type-2 responses, but they also include participation in type-1 and type-17 immune responses. They have a direct impact against ILCs as well as many immune cells (DC, T cells, Treg cells, brain and liver macrophages, pre-B cells, eosinophils, mast cells) and non-immune cells (keratinocytes, oligodendrocytes, and tumors). These diverse targets and functions suggest that controlling alarmin signaling may be a novel therapeutic target. Indeed, numerous clinical trials have been or are being conducted on these alarmins. All focus on IL-33 or TSLP, most are on allergic diseases, and most are still ongoing, but the existing data are promising and suggest that biologics that target the alarmins can safely and effectively treat allergy. Moreover, although the research on the roles of alarmins in non-allergic diseases such as cancer and autoimmune diseases is still in its infancy, the preliminary pre-clinical and clinical data suggest that anti-alarmin biologics could be useful in these diseases as well.

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