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Regulation of acute reflectory hyperinflammation in viral and other diseases by means of stellate ganglion block. A conceptual view with a focus on Covid-19

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ABSTRACT

Whereas the autonomic nervous system (ANS) and the immune system used to be assigned separate functions, it has now become clear that the ANS and the immune system (and thereby inflammatory cascades) work closely together. During an acute immune response (e.g., in viral infection like Covid-19) the ANS and the immune system establish a fast interaction resulting in “physiological” inflammation. Based on our knowledge of the modulation of inflammation by the ANS we propose that a reflectory malfunction of the ANS with hyperactivity of the sympathetic nervous system (SNS) may be involved in the generation of acute hyperinflammation. We believe that sympathetic hyperactivity triggers a hyperresponsiveness of the immune system (“cytokine storm”) with consecutive tissue damage. These reflectory neuroimmunological and inflammatory cascades constitute a general reaction principle of the organism under the leadership of the ANS and does not only occur in viral infections, although Covid-19 is a typical current example therefore. Within the overreaction several independent pathological positive feedback loops can be detected in which the SNS plays an important part. Consequently, there is a chance to regulate the hyperinflammation by influencing the SNS. This can be achieved by a stellate ganglion block (SGB) with local anesthetics, temporarily disrupting the pathological positive feedback loops. Thereafter, the complex neuroimmune system has the chance to reorganize itself. Previous clinical and experimental data have confirmed a favorable outcome in hyperinflammation (including pneumonia) after SGB (measurable e.g. by a reduction in proinflammatory cytokines).

1. Introduction

In viral infections, currently seen in the Covid-19 pandemic caused by the coronavirus SARS-CoV-2, the course is often mild, but some patients experience severe disease due to hyperresponsiveness of the immune system with excessive acute inflammation (hyperinflammation, cytokine storm) and consecutive tissue damage. This can lead to acute respiratory distress syndrome (ARDS), multi-organ involvement, microcirculatory disturbances, and microthrombosis (Dreher et al., 2020; Merad and Martin, 2020; Varga et al., 2020). In lab tests the so-called “cytokine storm”, a strong increase of cytokines (including TNF-α, IL-1, IL-6 and chemokines) is found (Clark and Vissel, 2017; Coperchini et al., 2020; Huang et al., 2020; Liu et al., 2020; Merad and Martin, 2020; Qin et al., 2020; Ruan et al., 2020; Velavan and Meyer, 2020; Zhu et al., 2020). Therefore, tissue damages are not solely due to direct virus-induced cytopathic effects.

The immune and inflammatory processes are – also in the case of a virus infection – reflectory controlled by the autonomic nervous system (ANS) (Elenkov et al., 2000; Tracey, 2002; Tracey, 2009; Grebe et al., 2010; Jäning, 2014a; Steenblock et al., 2020) which may thus be also...
The black arrows symbolize that the systems are connected to each other via infection, but an expression of a malfunction of the ANS with sympathetic hyperactivity, inflammation, and coagulopathy are not different entities in a viral infection – the disease is then determined by the organ or system. (For literature, see text). Interestingly, the reactions in this system take place in principle in the same way, regardless of whether the initial stimulus was a viral infection, mechanical trauma, psychological stress, or other cause. The clinical picture – the disease – is then determined by the organ or system mainly affected (see text). In our opinion, the consequences of acute hyperinflammation such as ARDS, endothelial dysfunction, microcirculation disturbance, and coagulopathy are not different entities in a viral infection, but an expression of a malfunction of the ANS with sympathetic hyperactivity.

2. Autonomic nervous system and immune system

2.1. Anatomy and physiology of the autonomic nervous system

The ANS (sympathetic and parasympathetic nervous system) consists of a central (brain and spinal cord) and peripheral part. It maintains an internal environment adapted to various factors. In addition, it significantly contributes to the control of inflammatory and immune processes as well as to the regulation of (micro)circulation (Elenkov et al., 2000; Tracey, 2002; Jäning, 2006; Jäning, 2014a).

The ANS, by virtue of its largely ubiquitous distribution as a feedback autoregulatory system, influences the functions of cells and organs, including the vascular system, endocrine system, and immune system. The SNS is distributed efferently from the sympathetic centers in the brain, including the hypothalamus, rostral ventral lateral medulla (RVLM) (Ogundele et al., 2017; Paton and Sper, 2013), through the spinal nucleus of the spinal cord (nucleus intermediolateralis), via the para- and prevertebral ganglia. The postganglionic nerve fibers run into the interstitium, with influence to mast cells, macrophages, fibrocytes, leukocytes (van der Zypen, 1967; Fischinger, 2016; Heine, 2011; Benias et al., 2018), and to the organs and organ systems. In addition, the hypothalamic-pituitary-adrenal axis (HPA) stimulates the adrenal cortex, and the sympatho-adrenal (SA) system influences the adrenal medulla (Jäning, 2014a). Sensory nerve fibers in the truncus sympathetic project from the periphery (including the interstitium) via pre- and para-vertebral ganglia and dorsal root ganglia to the spinal cord and from there to the centers in the brain. A second way into the brain is via cervical ganglia and the vascular plexus.

The efferent parasympathetic fibers have their origin in the core areas of the vagus nerve (nucleus ambiguus and nucleus dorsalis n. vagi) and form various plexuses together with the SNS (plexus pulmonary, plexus cardiacus, and plexus coeliacus). The sensory fibers in the vagus nerve project into areas in the brain stem (nucleus tractus solitarius [NTS]). There, the NTS establishes connections to efferent parasympathetic and sympathetic core areas and to integration centers such as the hypothalamus (Ross et al., 1985; Affleck et al., 2012) (Figs. 2 and 3).

2.2. Neuroimmune interactions

Whereas the nervous and the immune system were traditionally thought to be involved in separate functions, it has become clear since the works of Elenkov et al. in 2000 and Tracey in 2002 that the ANS and the immune system (and thereby inflammatory cascades) are in fact inseparable. Central networks of the ANS are informed about the peripheral inflammation and immune status (Tracey, 2002) by sensory nerve fibers (nociceptive neurons, sensory nerve fibers in the truncus sympathetic and in the vagal nerve) as well as by the humoral pathway – especially cytokines, which (among others) originate from the inflamed areas and lymphoid organs (Jäning, 2014a) (Figs. 2 and 3). Humoral and afferent neuronal signals co-determine the strength of the efferent “response” of the vagus and especially of the SNS from the autonomous centers of the brain (Kenney and Ganta, 2014). The ANS can control not only a general inflammatory and immune response, but also local processes in the damaged tissue (with sensory feedback to the brain) (Figs. 2 and 3). Thus, immunological homeostasis is a continuous, homodynamic process controlled by the ANS which allows throttling or stimulation in effector tissue including microcirculation and immune cells (Jäning, 2011; Jäning, 2014a; Kenney and Ganta, 2014).

During an immune response (e. g. in a viral infection) the brain and the immune system “talk to each other” (Elenkov et al., 2000; Tracey, 2002; O’Connor et al., 2004; Jäning, 2014a; Ji et al., 2016) via afferent and efferent pathways and know about their mutual state (Figs. 2 and 3). Immune cells express adrenergceptors (Bellinger and Lorton, 2014), and, conversely, nerve endings of sensory neurons register changes in the local concentration of immune mediators (cytokines, chemokines) and transmit this information to the brain (Tracey, 2002; Schaible, 2014). Autonomous centers in the brain localize the tissue damage at a specific location in the organism, which would not be possible via the bloodstream. Inflammatory mediators such as cytokines including chemokines are actually neuromodulators (Schaible et al., 2011; Chiu et al., 2013; Ji et al., 2016). In this localized neuroinflammation, also glial cells in the dorsal root ganglia (DRG), the spinal cord and the brain are

Fig. 1. Overview of affected systems, e. g. following a virus infection like Covid-19. The grey area symbolizes that the autonomic nervous system (ANS) is integrated in the communication of all systems and has a coordinating function. The black arrows symbolize that the systems are connected to each other via neuronal and non-neuronal feedback loops and are therefore interdependent. The same neuronal and non-neuronal substances can have both activating and inhibiting effects, depending on the current state of the system as well as on the point in time (For literature, see text). Interestingly, the reactions in this system take place in principle in the same way, regardless of whether the initial stimulus was a viral infection, mechanical trauma, psychological stress, or other cause. The clinical picture – the disease – is then determined by the organ or organ system mainly affected (see text). In our opinion, the consequences of acute hyperinflammation such as ARDS, endothelial dysfunction, microcirculation disturbance, and coagulopathy are not different entities in a viral infection, but an expression of a malfunction of the ANS with sympathetic hyperactivity.
Interestingly, not only immune cells (macrophages, monocytes, lymphocytes) can produce cytokines, but also neurons of the central and peripheral nervous system, microglia and astrocytes, and vascular endothelial cells (Galic et al., 2012). Peripheral nerves have partly the same molecular recognition pathways (such as toll-like receptors) as the immune cells (Chiu et al., 2012; Chiu et al., 2013), allowing a very direct and fast communication (interaction). During noxious stimulation (trauma, toxins, viruses, bacteria, etc.) nerve fibers themselves release proinflammatory neuropeptides such as substance P (SP) and calcitonin gene-related peptides (CGRP). These are also released by immune cells such as macrophages, lymphocytes, and mast cells (Stanisz et al., 1986). This results in neurogenic inflammation with plasma extravasation and edema (Chiu et al., 2012; Chiu et al., 2013; Jänig, 2014a; Ji et al., 2016; Matsuda et al., 2019). Neuropeptides secreted by nerve fibers can in turn induce cytokine production in immune cells (Ansel et al., 1993; Hosoi et al., 1993; Stanisz, 2001; Veres et al., 2007) and generate in this way a positive feedback loop. Neurogenic inflammation also occurs in virus-associated respiratory infections (King et al., 2001), and viral infections can also activate toll-like receptors in nociceptors thus allowing a modulation of pain and local neuroinflammation (Ji et al., 2016).

3. Why a stellate ganglion block (SGB)?

Surprisingly, the basic neuroimmune interactions in heterogenous clinical pictures (“diagnosis”) with overreaction of the immune and
inflammatory system do not seem to differ (see Section 3.5), and thereby allow principally to be influenced (peripherally and centrally) by SGB (Fig. 4) (see Section 3.5). In the past, we have gained experience with SGB in heterogeneous clinical pictures which were accompanied by inflammations, immune disorders, circulatory disorders, and pain (Barop, 2017; Fischer, 2020). Therefore we thought of a common pathogenetic mechanism and a “cooperation” between the ANS and the immune system in inflammations, and that their central mechanisms are also influenced by SGB.

3.1. Anatomy and physiology of the stellate ganglion

The sympathetic SG is located at the level of the 1st rib head in the lamina prevertebralis fasciae cervicalis. The neurons of the ganglion receive their impulses via the rami communicantes albi from the spinal core areas of the nucleus intermediolateralis from C8 to Th5. After switching via divergence principle (McLachlan, 2003) to postganglionic neurons in the SG, the latter supply the upper body quadrant.

The SG modulates the functions in its immediate area of supply: brain, neck, lung, heart, upper extremity, vessels including microcirculation and endothelial function, interstitium, and immune system. The SG does not only influence the immune processing in the periphery but also brain areas involved in the control of the immune system. Therefore, the influence of the SG is not limited to pathophysiological processes in its immediate peripheral supply area (e.g. viral pneumonia, myocarditis) of the upper body quadrant, but also modifies inflammation remote from the SG, e.g. of the intestine. The reason for this is that neuronal and non-neuronal (cytokines, etc.) afferent signals from the intestine are received and integrated in the vagal and sympathetic brain centers which in turn influence the intestine (and its immune system) by feedback (Fig. 3). The autonomous centers in the brain are also under the influence of the SG (Westerhaus and Loewy, 2001), explaining why the brain (supplied by the SG) can provide ubiquitous immune responses and influence inflammation via the sympathetic and parasympathetic system (Elenkov et al., 2000; Tracey, 2002; Tracey, 2009; Janig, 2014b; Lipov and Candido, 2017). In a review of the literature of the anatomy of SG we noted that the sympathetic and vagal fibers are intertwined by a continuous commingling and via rami communicantes (Puente de la Vega Costa et al., 2016). Therefore, SGB is not a purely sympathetic block; vagal fibers are always influenced by the injection, partly because of their anatomical proximity, partly through the rami communicantes. This is desirable because both sympathetic and vagal efferents and afferents are important for the temporary interruption of the immune and inflammatory reflex paths (Puente de la Vega Costa et al., 2016).

3.2. Regulation of the acute neurogenic hyperinflammation (cytokine storm) by SGB

In the literature, the terms “proinflammatory”, “hyperinflammation” and “cytokine storm” are not exactly defined. We use the terms analogous to the following descriptions.

Proinflammatory: by this we mean substances or processes that have a proinflammatory effect, such as substance P, proinflammatory cytokines, etc. Here, the proinflammatory effect may well be physiological for defense against infectious agents, in trauma, etc. (Esch and Stefano, 2002).

Hyperinflammation: this goes beyond the physiological inflammatory response and causes tissue damage. Scientific publications and the media often use the terms “hyperinflammation” and “cytokine storm” analogously, especially in connection with Covid-19. This is practicable but not very precise. We think the term “acute hyperinflammation” is more precise.

Cytokine storm has no precise definition (Sinha et al., 2020) and “no single definition of cytokine storm as an umbrella term is widely accepted” (Fajgenbaum and June, 2020). Cytokine storm is a hyperactive immune response with excessive increased cytokines and other mediators. This is more than a physiological innate immune answer of an infection; cytokine storm causes via hyperinflammation cell and organ damage and often death (Burke et al., 2020; Sinha et al., 2020; Kim et al., 2021).

In the following, we postulate that in many tissue damages, e.g. after virus infection, the overreaction of the immune system (e.g. cytokine storm) with hyperinflammation is the consequence of an SNS overreaction, and that SGB is a logical possibility to counteract it (Figs. 2 and 3). The physiological balance between the SNS and the vagus nerve is biased towards SNS dominance (Alston et al., 2011; Puente de la Vega Costa et al., 2016; Koopman et al., 2017). Speranski, a student of Pavlov, was able to show in 1950 in animal experiments that after experimental preloading of the SNS (“first strike”) (e.g. chronic artificial hypothalamus and pituitary gland irrigation), a subsequent, additional stimulus (“second strike”) caused various overshooting, sympathetically triggered, reflexoric inflammations (such as arthritis). For Speranski (1950), excessive inflammation was the result of an overreaction of the preloaded SNS.

The imbalance in the ANS can also promote inflammation when the SNS is dominant (Alston et al., 2011; Puente de la Vega Costa et al., 2016; Koopman et al., 2017). Generally (with exceptions), the hyperactivated SNS has a proinflammatory effect (Schaible and Straub, 2014) while the activated parasympathetic fibers of the vagus nerve have an anti-inflammatory effect. Nevertheless, the sympathetic and parasympathetic nervous systems cannot be considered in isolation; they are more than the sum of their parts (Fischer, 1998; Tracey, 2002), since they are not only centrally but also peripherally more closely intertwined than previously assumed (Puente de la Vega Costa et al., 2016) (see above). Although the pathophysiology was not known in detail at the time of Speranski’s experiments, later research has confirmed the “leadership” of the ANS, especially of the SNS, in (excessive) immune reactions and inflammation (Piedimonte et al., 1999; King et al., 2001). Tracey, one of the pioneers of seeing inflammation and immune processes as part of a neuronal reflex involving the SNS and the vagus (Tracey, 2002; Tracey, 2009), proposed neuronal different “setpoints” for the immune response to a stimulus such as infection: A normal “setpoint” results in a protective response (normal inflammation and tissue repair), a decreased “setpoint” in immunosuppression with...
increased infection, and an increased “setpoint” in hyperinflammation and tissue damage. Similar as in Speranski’s (1950) experiments, a direct (hypothesis [Kumagai et al., 2012], obesity, or stress) or indirect (organ damages) preload on the SNS may shift Tracey’s “setpoint” towards an increase (corresponding to Speranski’s “first strike”), so that the additional stimulus – such as an infection (Speranski’s “second strike”) – leads to an overreaction of the SNS which disturbs the balance in the ANS (Speranski, 1950; Fischer, 2002; Tracey, 2009; Barop, 2017; Fischer, 2019). This hyperactivity would then be measurable by a cytokine storm and other laboratory parameters as well as tissue damage. In such situations, various authors have proposed to stimulate the vagus nerve in order to improve the balance between the sympathetic system and the vagus nerve (Tracey, 2002; Tracey, 2009; Pavlov and Tracey, 2015; Breit et al., 2018; Pavlov et al., 2020; Tsavara et al., 2020; Caravaca et al., 2019). Effects of vagus nerve stimulation (reduction of inflammation, decreased cytokine production) correspond in principle to SGB which we are promoting here (see Section 5.3). Even though the Tracey model focuses predominantly on systemic autoimmune diseases and not on viral infections, there is striking evidence that the pathogenetic reflex mechanisms might be the same (uniform pathogenesis) with different initial (also viral) stimuli (see Section 3.5.2).

The severity of inflammatory disease is also closely related to the activity of sensory TRPV1-expressing neurons (transient receptor potential channel, formerly known as vanilloid receptor 1 or capsaicin receptor) in the truncus sympathetic and in the vagus nerve, as well as to the activity of sympathetic and vagal efferents in the lungs which modulate inflammation and immune system (Baral et al., 2018; Nahama et al., 2020). These neurons are in cross talk with the immune system (Tränkner et al., 2014; Chavan et al., 2017) and can release more proinflammatory molecules such as SP and cytokines once activated. In order to reduce overproduction of cytokines, the authors suggest to block TRPV1-positive fibers which produce SP. Nahama et al. (2020) propose a periganglionic block corresponding to SGB. Baral et al. (2018) confirmed this approach in animal studies (ablation of TRPV1).

Numerous authors stress the importance of the nervous system in excessive inflammation: Neuronal reflex pathways can increase the secretion of proinflammatory cytokines and inflammation (Heijnen et al., 2002; Tracey, 2002; Perez et al., 2009; Jänig and Baron, 2011; Bellinger and Lorton, 2014). A virus-induced exaggerated immune response (cytokine storm) is considered a consequence of excessive neurogenic mechanisms (prolonged airway hyperresponsiveness) (Perianin et al., 1989; Mejías et al., 2005). Cytokine overproduction seems to be important in tissue damage, e. g. in the lung (Wang et al., 2020). In order to regulate the cytokine storm, we believe it is crucial to regulate the overshooting neurogenic mechanisms, e. g. by SGB, to achieve a different “setpoint” (Tracey, 2009).

Ligand (virus)-receptor interactions, too, activate the secretion of proinflammatory cytokines. The latter activate sensory nerve fibers (Fig. 2) and evoke a reflex response via the SNS and vagus (Tracey, 2009). If this response is overshooting and sympathetic-dominant, electrical vagus nerve stimulation can be applied therapeutically (Tracey, 2002; Tracey, 2009), which corresponds functionally to SGB in the case of a shift towards sympathetic nerve activity. SGB reduces the production of TNF-α, IL-1, IL6, and IL-8 (Liu et al., 2013; Yang et al., 2015; Chen et al., 2018) and increases the production of IL-10, which is anti-inflammatory (Table 1) (Liu et al., 2013; Yang et al., 2015; Chen et al., 2018). Chemical sympathectomy showed basically the same results (Pierach and Stotz, 1952). Stimulation of the vagus nerve also reduces the production of proinflammatory cytokines in the spleen by unknown mechanisms (Tracey, 2009), stressing the communication (interaction) between the nervous and the immune system (see above). If even the nucleus tractus solitarius is affected by virus invasion, as described, e. g. in some cases of Covid-19 (Lj et al., 2020), it can lead to a sympathetic (catecholamine) storm, which then induces a cytokine storm leading to neurogenic pneumonia (Ur and Verma, 2020a; Ur and Verma, 2020b). Early treatment of the nervous system is – as the authors stress – crucial in this situation (Ur and Verma, 2020b). We believe that SGB is the right choice because it reduces hyperinflammatory mechanisms in the CNS as well as in lung and heart area etc.

As we could show in our previous works, the spinal cord is also strongly involved in the development of peripheral inflammation, with indications that the effects are mediated via the SNS (Boettger et al., 2010a, 2010b). SGB – as early as possible (Strong et al., 2018) – is a logical intervention to interrupt feedback loops.

We assume that a neuroplastically altered SG intensifies pneumonia towards ARDS, consistent with the neural hypothesis of lung edema (Ur and Verma, 2020b). Neuroimmunologically controlled hyper-inflammation also involves functional and structural changes in the endothelium and microcirculation and possibly also coagulopathy (see Sections 3.3 and 3.4). This results in further feedback loops (Fig. 1).

Since SGB shuts down the mentioned feedback loops simultaneously (because they are interdependent), the neuroimmune system has now the chance to reorganize itself (autoregulation). This reorganization is therefore not tied to the duration of action of the LA (procaine); rather, it far outlasts the short-term SGB (Puente de la Vega Costa et al., 2016; Barop, 2017; Fischer, 2019).

### 3.3. Regulation of the endothelial dysfunction by SGB

The ANS influences endothelial functions from outside the vessels (Amiya et al., 2014). Blood vessels are innervated by sympathetic and – only in a few places in the body – parasympathetic nerve fibers. Endothelial dysfunction and inflammation are the consequence of pathologically increased sympathetic nerve activity (Sheng and Zhu, 2018). On the other hand, the endothelium can influence the SNS via vasoactive factors. This results in complex interactions involving positive feedback loops. Inflammation impairs the endothelial function (Sinisalo et al., 2000). Even anxiety or sympathetic hyperactivity, e. g. with Takotsubo syndrome (Naegele et al., 2016) is associated with endothelial dysfunction (Narita et al., 2007) further stressing the connection between ANS, immune system, and endothelium (Mousa et al., 2010).

Endothelial cells can also produce vasoactive substances such as nitric oxide (NO) and endothelin. Endothelin maintains local and generalized processes. It can stimulate the peripheral and central SNS and can also be released by postganglionic sympathetic nerves and control catecholamine release and vascular tone (Yamaguchi, 1997; Schiffrin, 2012). In cooperation with the SNS, endothelin has a pathophysiological role in arterial hypertension including pulmonary hypertension, atherosclerosis, vascular inflammation with fibrosis, and pulmonary fibrosis (Schiffrin, 2012). Increased sympathetic tone combined with infiltration of the blood vessel wall can lead to hypertension (Zhang et al., 2020). Chen et al. (2012) observed a decrease in endothelin (and hypertension) in rats after applying SGB. Like endothelin, NO is involved in the complex regulatory circuits of endothelium–inflammation–ANS

| Table 1 | Effects of the stellate ganglion block. |
|---|---|
| **Regulation of the immune system/hyperinflammation** | **Reduction of natural killer cell activity** (Sugimoto et al., 1995; Yokoyama et al., 2005) |
| **Reduction of inflammatory cytokines IL-1, IL-4, IL-6, IL-8, TNF-α** (Liu et al., 2013; Yang et al., 2015; Chen et al., 2018) | **Increase of anti-inflammatory cytokine IL-10 and of CGRP** (Liu et al., 2013; Yang et al., 2015; Chen et al., 2018) |
| **Regulation of the endothelial dysfunction** (Chen et al., 2012) | **Regulation of microcirculation and coagulopathy** (Levy and Lorch, 1996; Hanamatsu et al., 2002; Pfister and Fischer, 2009; Yamazaki et al., 2012; Zhou et al., 2014; Punj, 2019) |
| **Reduction of (neurogenic) pulmonary edema** (Pierach and Stotz, 1952; Zhang et al., 2013; Liu et al., 2017; Chen et al., 2018) | **Reduction of pulmonary arterial hypertension** (Meyers et al., 1996) |
| **Reduction of pathological positive feedback loops** (Fischer, 2003; Jänig, 2011; Fischer, 2019) | |
and has been favorably influenced by SGB in animal experiments (Chen et al., 2012). Leptin, released from adipocytes, also plays a role in arterial hypertension in collaboration with the SNS, and it causes endothelial dysfunction and increased blood pressure (Wang et al., 2013). Both pathologies are mediated by SNS activation (Wang et al., 2013). These effects of leptin were neutralized by sympathetic denervation (ganglionectomy) (Wang et al., 2013). That obesity and hypertension are risk factors for severe courses of Covid-19 is also understandable from this point of view (see also Section 5.2).

3.4. Regulation of microcirculation – disturbance and coagulopathy by SGB

The severity of an infectious disease is not only determined by pathogenic damage and reflectory (hyper-)inflammation of the organs (e.g. in pneumonia), but also by microcirculatory disturbance with hypercoagulopathy, as with Covid-19. Inflammation is the predominantly sympathetically controlled vascular response to irritants such as viruses and others (Ricker, 1924; Marvar and Harrison, 2012). The response includes vasodilation with disturbed blood flow in irritated tissue, increased vascular permeability, production of cytokines and chemokines, and mobilization of immune cells. In a feedback loop, the brain modulates the intensity of inflammation after being informed by sensory nerve fibers from the irritated tissue (Figs. 2 and 3) (Sternberg, 2006; Marvar and Harrison, 2012). In 1994 already, Greiff et al. reported a microvascular exudative hyperresponsiveness in human coronavirus-induced common cold (Salgado et al., 2010) described disturbed microcirculatory blood flow in patients with severe influenza A (H1N1). The importance (and the link) of microvascular alterations and coagulopathy with microthrombi are also discussed in the pathogenesis of Covid-19 (Damiani et al., 2020; Magro et al., 2020; Varga et al., 2020; Spezia et al., 2020). In our opinion, the SNS should be more considered. Overactivity of the SNS including the hypothalamic-pituitary-adrenal axis can lead to hypercoagulability with thrombus formation (von Känel andDimsdale, 2000; Pagano et al., 2014). Microthrombi of the lungs and other organs are mainly mediated by cytokines (Levi et al., 2008; Merad and Martin, 2020), and therefore can be triggered by a pathologic ANS hyperactivity. Wang et al. (2019) also described the role of sympathetic activity on the pathogenesis of pulmonary embolism. Crucial is sympathetic hyperactivity which influences coagulation and fibrinolysis pathways and causes microthrombi with increased plasma D-dimers (Ur andVerma, 2020b). SGB also has an effect on pulmonary arterial hypertension and thus on microcirculation (Myers et al., 1998; Zhao et al., 2019) and is therefore a logical intervention for microcirculation disorders and coagulopathy (Murakawa, 1993; Levy andLorch, 1996; Hanamatsu et al., 2002; Pfister and Fischer, 2009; Yamazaki et al., 2012; Zhou et al., 2014; Punj, 2019) (Fig. 4).

3.5. Further clinical and experimental data

3.5.1. SGB in pneumonias

The SNS increases proinflammatory cytokines and exacerbates influenza A virus pathogenesis (Grebe et al., 2010). The authors showed that peripheral chemical sympathectomy in mice reduced morbidity and mortality from influenza A virus-induced pneumonia. After sympathectomy they observed a reduced cytokine response and a reduced inflammatory influx of monocytes, neutrophils, and natural killer (NK) cells. In the old works by the surgeon Leriche, it is suggested that sympathectomy for inflammation corresponds in effect to repeated SGB with procaine (Leriche and Fontaine, 1934). The theoretical background of hyperinflammation due to disequilibrium of the ANS with a dominance of the sympathetic part in acute lung injury stimulated the animal experiments of Liu et al. (2017). They showed that the regulation of the ANS, especially the sympathetic part, by SGB reduced inflammation and improved lung function in rabbits. In principle, vagus nerve stimulation produces a better balance in the ANS in case of overstimulated SNS. Vagus nerve stimulation also had an anti-inflammatory effect, as did the administration of cholinesterase inhibitors, but both were less potent than SGB (Liu et al., 2017). SGB not only reduced proinflammatory cytokines, but also the swelling of microvascular endothelial cells and epithelial type I cells in the lung (Liu et al., 2017).

Chen et al. (2018) showed in septicaemia with acute lung injury that SGB reduces TNF-α and IL-6, increases the anti-inflammatory IL-10, and inhibits the infiltration of inflammatory cells into lung tissue. Baral et al. (2018) produced a methicillin-resistant staphylococcus aureus-pneumonia in mice: TRPV1-expressing sensory neurons in the trunci sympathicus and the vagus nerve were chemically ablated (this corresponds to our opinion to the repeated SGBs in this circuit). Survival was improved and cytokine storm significantly reduced. Thus reduced activity of TRPV1-positive nerve fibers therefore probably prevents the transition of mild pneumonia into ARDS. In our opinion, this is a further plausible reason for SGB in such situations, supported by the fact that the TRPV1 channels can be influenced by LA such as procaine (Yanagidate and Strichartz, 2007). This even raises the question of the additional benefit of an intravenous procaine infusion (Papathanasiou, 2020), especially since procaine is anti-inflammatory and antithrombotic (Cassuto et al., 2006; Konrad et al., 2006).

Okada et al. (1996) found a reduction in the activity of NK-cells in the spleen after resection of the cervical border strand, and Sugimoto et al. (1999) found that SGB causes a short-term reduction in cytotoxic T-cell activation, i.e. in NK-cells. Yokoyama et al. (2000), also found a modified distribution of lymphocyte subsets and NK cell activity following SGB. SGB also regulates the increased hormone release of the HPA axis, especially the norepinephrine levels (Yokoyama et al., 2000).

3.5.2. SGB in other situations of inflammation or acute hyperinflammation (cytokine storm)

The following studies and experiments come to uniform basic statements about the effect of SGB – although different organ systems such as the lungs, vessels, etc. or a trauma, a postoperative state, an autoimmune disease are affected. In principle, the mechanisms of predominantly sympathetic-triggered inflammation or hyperinflammation are always the same since the body has just a few reaction patterns to respond to stimuli such as infection, trauma, etc. The neuro-immunological and inflammatory cascade seem to be in motion regardless of the initial stimulus. The affected end organ determines the specific clinical appearance. Some examples: Analogous to the overreaction (cytokine storm) of the neuro-endocrine-immune network in Covid-19, the same processes are found in the early phase of severe trauma. It is called systemic inflammatory response syndrome (SIRS) (Liu et al., 2013) and is one of the major factors in complications, organ failure and death. SIRS, too, is associated with an overproduction of proinflammatory cytokines such as IL-1β, IL-6 and TNF-α. SGB regulates the early systemic inflammatory response in severe trauma patients (Liu et al., 2013). The authors found that the concentrations of IL-1β, IL-6 and TNF-α (but not IL-4 and IL-10) are significantly lower after SGB than in the control group without SGB. SGB regulates the hyperactive neuro-endocrine-immune response to allow homeostasis and is recommended as an effective therapy for impending SIRS.

Yang et al. (2015) found that SGB can regulate cellular and humoral immune functions in patients with traumatic brain injury (TBI); likewise, it can improve an overreacting nerve-endocrine-immune system, excessive vasoconstriction, abnormal blood rheology, and thus restore the balance between the sympathetic (excessive sympathetic tone) and parasympathetic nervous systems. In 50 patients with TBI, Yang et al. (2015) showed that the excessive inflammatory response after TBI is an important reason that aggravates the secondary brain injury. IL-1β, IL-6 and TNF-α were significantly reduced after SGB compared to the control group. The authors also suspect a positive effect of SGB on central and peripheral vasoconstriction.

Thus, severe trauma can cause an overreactive stress response involving the complex feedback in the neuro-endocrine-immune system
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This technique allows a very shallow depth of the injection, so that
rather suggest regulation of the neuroimmune system by short-lasting
SGB than blockade of immune functions by selective drugs, since a
“selective” drug-induced blockage of parts of the neuroimmunological
processes may be associated with serious side effects including deteri-
oration of immunity and subsequent secondary infection or increase in
viral load (Sanders et al., 2020; Velavan and Meyer, 2020).

Since (despite the term SGB) we do not block but regulate, we do not
interfere with the body’s natural ability to develop an immunological
memory (adaptive immune system) and therefore there is no greater risk
of subsequent re-exposure. It is true that the SGB modifies the distri-
bution of the lymphocyte subsets and the NK cell activity (increase in the
proportion of B- and T-cells, and a decrease in NK-cells) (see Section
3.5.1), but these effects were transient (Yokoyama et al., 2000). The
transient SGB serves mainly to regulate an excessive immune response
to a physiological immune response.

3.7. Properties of procaine

In contrast to amide-structured LA (lidocaine, bupivacaine), procain-
e has a direct vasodilating (regulating) effect in addition to the
sympatholytic effect. Furthermore, procaine has an antiinflammatory
effect (Hollmann and Durieux, 2000; Cassuto et al., 2006). LA also act
on non-neuronal cells such as mast cells, neutrophils and monocytes as well
as on cytokine production and also have antibacterial and antiviral ef-
ects (Hollmann et al., 2001; Cassuto et al., 2006). LA can also modulate
an excessive inflammatory response that is no longer protecting, but
destroying tissue (Brown et al., 1995; Groeben et al., 1996; Groeben
et al., 1999; Hollmann and Durieux, 2000). In addition, LA can reduce
inflammation in the microglia (Zheng et al., 2019).

Procaine has no significant side effects (Fischer et al., 2005; Fischer
et al., 2010; Barop, 2017) and no interactions with other drugs because
it is not broken down by the liver but by pseudocholinesterase, which is
ubiquitously present. This is also an advantage in Covid-19, since most
of the time, severely ill patients are elderly, receiving poly-
pharmaceutical treatment.

4. Applying SGB using a minimally invasive technique

In 2016 we presented a modified technique (Puente de la Vega Costa
et al., 2016) based on Leriche and Fontaine (1934), Dosch (1986),
Fischer (1998), and Barop (2017). Technical details were described in
“Autonomic Neuroscience” (Puente de la Vega Costa et al., 2016). We
recommend the bilateral SGB with a time interval of ca 1 h (disappearing
Horner complex) twice daily for 2 or 3 days.

4.1. Material

Procaine 1% 5 ml each side without any additives. Needle: 20 × 0.4 mm
or 12 × 0.3 mm, depending on body type (short needles are possible
because of the remaining finger on Chassagnac’s tubercle).

This technique allows a very shallow depth of the injection, so that
very thin needles can be used, which allows the stellate injection even in
anticoagulated patients. However, with extremely thin needles, aspira-
tion is more difficult and must be performed slowly and continuously.
For safety reasons, anticoagulated patients must be given several mi-
utes of compression at the injection site.
4.2. Possible complications

Paralysis of the recurrent or phrenic nerve on the opposite side can lead to breathing problems. Accidental injection of a LA into a brain-bound blood vessel or into the cranial area of the cerebrospinal fluid space (cave: cervical cerebrospinal fluid cyst) can cause cramps, unconsciousness, cardiac and respiratory arrest.

5. Discussion

5.1. The neuroimmune system – complex and nonlinear?

There is evidence that an imbalance in the ANS (in particular an excessive sympathetic tone) due to pre-existing conditions is at the beginning of the overshooting reactions in Covid-19 and other diseases. Even small changes in non-physiologically balanced complex systems can therefore have a major effect. This is typical in a system with interdepending positive feedback loops (iterations) as described in the previous chapters. This gives the neuroimmune system a complex nonlinear character, analogous the mathematical chaos theory as it was described for nature and biology (Mandelbrot, 1967; Lorenz, 1972; Nicolis and Prigogine, 1977; Feigenbaum, 1978; Kluge and Neugebauer, 1994). We postulate several positive feedback loops dependent on the ANS which play a role in the overreaction of the neuroimmune system, in which neural and humoral factors reinforce each other. In positive feedback loops it is often difficult to find the “starting points”. We will try to illustrate this with some examples: Since neurons express receptors for inflammatory mediators (Schaible, 2014; Ji et al., 2016), SP or cytokines activate sensory nerve fibers to increase the synthesis (White et al., 2005; Zelenka et al., 2005; Xu et al., 2018) and release of even more SP (O'Connor et al., 2004; Matsuda et al., 2019) and cytokines (Galic et al., 2012). This positive feedback loop is a very fast way to reach high concentrations of SP and cytokines, but there is also the danger of overreaction when the system is already preloaded. This could be one of the mechanisms of a cytokine storm. SP can additionally stimulate the production of cytokines by macrophages (e.g. IL-1, IL-6 and TNF-α) (Bill et al., 1979) and induce chemotaxis and degranulation of human neutrophils (Serra et al., 1988; O'Connor et al., 2004). Furthermore, immune cells express adrenocorticotropin (Bellinger and Lorton, 2014), induced by proinflammatory cytokines (Heijnen et al., 2002; Perez et al., 2009). Thus, the SNS can induce their cytokine production – and cytokines are also produced by nerve fibers themselves (White et al., 2005; Zelenka et al., 2005). Furthermore, certain cytokines (IL-1) increase the synthesis of SP in sympathetic ganglia (Freidin and Kessler, 1991) and stimulate their secretion in afferent neurons (Inoue et al., 1999). The production and secretion of SP by lymphocytes and macrophages (De Giorgio et al., 1998; Lai et al., 1998) causes further positive feedback with expansion and intensification of neurogenic inflammation. Positive feedback loops are strongly interconnected, typical for a complex, iterative, chaotic system.

The question arises if positive feedback loops make sense in physiology. In our opinion these iterations are the quickest way to generate a physiological inflammation. In case of an infection, cytokines and SP can be provided very quickly. However, there is also the disadvantage that such systems are unstable, and we hypothesize that if the SNS is pre-loaded (such as hypertension, obesity, stress, negative emotions) or the regulatory capacity reduced, the (positive feedback) system can over-react and lead to a cytokine storm with a consecutive acute hyperinflammation in case of viral infections such as SARS-CoV-2. Thus, we believe, that the severity of e. g. viral pneumonia or even ARDS does not depend linearly on the initial stimulus (amount of virus). Depending on time and the current state of the system, the same parts of the ANS can have either an activating or inhibiting effect. For example, the SNS can have a pro- or anti-inflammatory (late phase) effect (Pongratz and Strauh, 2014; Strong et al., 2018). Therefore, neuronal pathways can use the same neurotransmitters either synergistically or antagonistically (Tracey, 2009). These observations also show the complexity of the system. In our opinion, the therapeutic manipulation of only one cell type (e. g. by receptor blockers) as a linear, reductionist intervention can only be associated with disadvantages and side effects (as experience has shown), since the cell type in question is protective under physiological conditions and only causes tissue damage in the case of dysregulation (hyperresponsiveness) (Ji et al., 2016). By contrast, the regulation of the ANS by means of short term SGB has an influence on cytokines and chemokines, for example, and – since regulation is used instead of suppression – has no adverse effects on the “physiological” performance of the inflammatory and immune system.

For this reason, the temporary shutdown by means of SGB is a logical intervention, since SGB has an influence at several places at the same time, and the system has the chance to reorganize itself afterwards (autoregulation) which is typical for a complex, nonlinear system.

5.2. Pre-existing conditions strain the neuroimmune system

We put forward the idea that with the hyperresponsiveness of the immune and inflammatory system, an overreaction of the already pre-loaded ANS plays an important role, specifically, by activity-enhancing positive feedback loops (see Section 3.2). So, the “cytokine storm” is caused by a “sympathetic storm” (Lemke, 2004) in the pre-loaded ANS; the “setpoint” (Tracey, 2002, 2009) is shifted, corresponding to Speranski’s (1950) “first strike” (see Section 3.2).

Thus, pre-existing sympathetic hyperactivity (including hypertension, obesity, psychological stress, multimorbidity, polymedication, metabolic syndrome, diabetes, intestinal diseases) and the associated central and peripheral neuroimmunological imbalance can lead to severe, sometimes fatal processes in case of a “second strike” (see Section 3.2) as in a viral infection with Covid-19 (Belliniva et al., 2020) especially in older people, in whom the neuroimmunological system appears to be less adaptable.

Hypertension and obesity are two major recognized risk factors for a severe course of Covid-19, both of which are characterized by sympathetic hyperactivity and imbalance of the SNS and vagus, respectively. As a result, the release of leptins from adipocytes is increased (Wang et al., 2013). Leptins cause endothelial dysfunction with increased release of endothelin, and further increase in blood pressure via the SNS (Wang et al., 2013) (see Section 3.3). Also, central mechanisms play a role: Kumagai et al. (2012) describe the importance of RVLM (rostral ventrolateral medulla) in determining efferent sympathetic nerve activity: this is modulated by proinflammatory cytokines (Heijnen et al., 2002; Tracey, 2002; Tracey, 2009; Ji et al., 2016), induced by proinflammatory cytokines (Heijnen et al., 2002; Perez et al., 2009). Thus, the SNS can induce their cytokine production – and cytokines are also produced by nerve fibers themselves (White et al., 2005; Zelenka et al., 2005). Furthermore, certain cytokines (IL-1) increase the synthesis of SP in sympathetic ganglia (Freidin and Kessler, 1991) and stimulate their secretion in afferent neurons (Inoue et al., 1999). The production and secretion of SP by lymphocytes and macrophages (De Giorgio et al., 1998; Lai et al., 1998) causes further positive feedback with expansion and intensification of neurogenic inflammation. Positive feedback loops are strongly interconnected, typical for a complex, iterative, chaotic system.

The question arises if positive feedback loops make sense in physiology. In our opinion these iterations are the quickest way to generate a physiological inflammation. In case of an infection, cytokines and SP can be provided very quickly. However, there is also the disadvantage that such systems are unstable, and we hypothesize that if the SNS is pre-loaded (such as hypertension, obesity, stress, negative emotions) or the regulatory capacity reduced, the (positive feedback) system can over-react and lead to a cytokine storm with a consecutive acute hyperinflammation in case of viral infections such as SARS-CoV-2. Thus, we believe, that the severity of e. g. viral pneumonia or even ARDS does not depend linearly on the initial stimulus (amount of virus). Depending on time and the current state of the system, the same parts of the ANS can have either an activating or inhibiting effect. For example, the SNS can have a pro- or anti-inflammatory (late phase) effect (Pongratz and Strauh, 2014; Strong et al., 2018). Therefore, neuronal pathways can use the same neurotransmitters either synergistically or antagonistically (Tracey, 2009). These observations also show the complexity of the system. In our opinion, the therapeutic manipulation of only one cell type (e. g. by receptor blockers) as a linear, reductionist intervention can only be associated with disadvantages and side effects (as experience has shown), since the cell type in question is protective under physiological conditions and only causes tissue damage in the case of dysregulation (hyperresponsiveness) (Ji et al., 2016). By contrast, the regulation of the ANS by means of short term SGB has an influence on cytokines and chemokines, for example, and – since regulation is used instead of suppression – has no adverse effects on the “physiological” performance of the inflammatory and immune system.

For this reason, the temporary shutdown by means of SGB is a logical intervention, since SGB has an influence at several places at the same time, and the system has the chance to reorganize itself afterwards (autoregulation) which is typical for a complex, nonlinear system.

5.3. Vagus nerve stimulation or stellate ganglion block?

The imbalance between the SNS and the vagus with sympathetic hyperactivity may be an important factor in hyperinflammation with cytokine storm. The balance can in principle be improved by vagus nerve stimulation (VNS) or by SGB. Piedimonte et al. (1999) and King et al. (2001) considered early on in their therapeutic considerations the neuronal regulation of neurogenic inflammation in viral infections. Neuroromodulation by VNS in reflex inflammation is proposed by various authors (Borovikova et al., 2006; Tracey, 2002; Tracey, 2009; Ji et al., 2016; Breit et al., 2018; Caravaca et al., 2019; Pavlov and Tracey, 2015; Pavlov et al., 2020; Tsaava et al., 2020). Tracey (2002, 2009) suspects that in the future, further neuroimmunological reflexes always will be discovered which can produce homeostasis in the innate and the adaptive immune system through therapy of the ANS. In general, according to these authors, it should be possible in the future to treat the immune response and the progression of the inflammation therapeutically via the neuronal networks. Caravaca et al. (2019) see the therapeutic potential
of the VNS also for autoimmune diseases with hyperinflammation such as chronic inflammatory intestinal diseases, rheumatoid arthritis, etc.

By applying SGB we regulate not only the SNS, but also the vagus nerve because, on the one hand, of their close connections within the periphery (anatomical and rami communicantes), as we described in our review (Puente de la Vega Costa et al., 2016), on the other hand, indirectly, because of the involvement of the sensory vagus “receiving site” in the brain (nucleus tractus solitarius). In addition, SGB enables us to reach the central sympathetic “output” centers such as the hypothalamus, pituitary gland, RVLM, etc. as well as peripheral sympathetic efferents and sensory fibers in the truncus sympathicus. Therefore, from a pathophysiological point of view, we consider SGB more advantageous than VNS, also confirmed experimentally by Liu et al. (2017).

5.4. Why has the SGB not been used more frequently in medicine?

The most common uses of SGB are in CRPS, circulatory disorders, and sympathetically maintained pain. The fear of hemodynamic problems has been a topic for decades. However, we could show that the hemodynamic parameters hardly change (Puente de la Vega Costa et al., 2016). There is a respect for other complications. They mainly occur when applying old techniques with significantly larger amounts of long-acting LA with a high diffusion capacity (undesired diffusion to neighboring structures). With the short-acting, only slightly diffusing procaine in lower amounts, we have not seen any complications; only a few have been noted in the literature (Fischer et al., 2005; Fischer et al., 2010) – with the reservation that certainly not all complications have been reported and published.

The decades-long fear of the allergic potential of procaine has not been confirmed in recent years (Fischer et al., 2005; Barop, 2017). In the past, mainly the additives triggered allergies. Procaine even has an anti-allergic potency (Barop, 2017; Fischer, 2019).

Another reason for the limited breadth of its application is that SGB is not considered guideline-compliant for most indications – and usually only specially trained anesthetists perform SGBs.

In addition, the range of indications is not known to many clinically active physicians due to decades of neglect of the ANS in pain, immune and inflammatory processes. Oyama and Node (2014) put it aptly: “Most of the previous studies lacked careful consideration of the impact of the ANS. The main reason is the difficulty to evaluate sympathetic nerve activity directly and easily.”

5.5. Limitations and factors that may impair the efficacy of a stellate ganglion block

Whether the injection can be performed during anticoagulation has to be decided individually (risk-benefit assessment, lack of other therapeutic options, or these have been exhausted). However, the area of injection can well be compressed manually after the (atraumatic) injection, preventing hematoma. Nevertheless, we principally advise against SGB in anti-coagulated patients. This is another reason why early recognition of a threatening course of Covid-19 is important; thus, SGB can be performed shortly before anticoagulation is started. Therefore, clinical and laboratory monitoring of high-risk patients should be close-meshed.

The timing of SGB plays a major role: it should be as early as possible, if laboratory or clinical evidence (e.g. beginning respiratory insufficiency) shows that the neuroimmunological networks are overreacting (Burke et al., 2020; Caricchio et al., 2021). In general, we have seen the best results in acute or acutely exacerbated inflammation (Pfister and Fischer, 2009; Schaible and Straub, 2014; Strong et al., 2018) for “timing” (see also Section 3.5.2).

In our experience, corticosteroids or other inhibitory drugs as well as old age can impede the self-organization expected after SGB (Barop, 2017; Fischer, 2019).

The correct technique plays an important role: not only the equilateral Horner symptom complex should be positively checked, but also a temperature increase of the equilateral upper extremity of >0.5 °C; because if the SNS is blocked cranially the SG, a Horner symptom complex is though created, but the postganglionic fibers for the lungs and heart are not directly affected by the block.

In previous studies and experiments, SGB was usually only conducted unilaterally. However, we propose the bilateral SGB with a time interval (see Section 4).

Another limiting factor could be old age (reduced capacity for self-organization) – we got this impression during our clinical work – but this would still have to be verified by means of studies.

6. Conclusions

The sympathetically maintained interdependent positive feedback loops give the neuroimmune system an iterative, nonlinear character (compatible with the mathematical chaos theory). Such systems have the fundamental ability to reorganize themselves (autoregulation) after changes of state e.g. by means of SGB. With this regulation the nonlinearity of the system is respected.

Previous clinical and experimental data show that the repeated, temporary SGB with LA procaine, is capable to regulate the sympathetically triggered neurogenic inflammation processes.

Because of the very low material costs, this therapy is also suitable for countries with low financial resources in the health system.

Despite the existence of clinical and experimental data on the effect of SGB and its logical mechanisms of action, larger studies are necessary. We hope that this work will serve as a basis for applications to the ethics committees.

Because of the lack of direct measurements of the ANS especially in pain, immune and inflammatory processes, the ANS was not considered enough in the past in our opinion. We hope that our conceptual view with partially new pathophysiological inputs may help to show how important the careful consideration of the ANS is. We also think that the therapy by the ANS is just at the beginning and that there is still a great potential here.

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Declaration of competing interest

None.

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References

Aiflick, V.S., Coote, J.H., Pyner, S., 2012. The projection and synaptic organisation of NTS afferent connections with presympathetic neurons, GABA and nNOS neurons in the paraventricular nucleus of the hypothalamus. Neuroscience 219 (1–2), 48–61. https://doi.org/10.1016/j.neuroscience.2012.05.070.

Alston, E.N., Farrish, D.C., Hasan, W., Tharp, K., Pahlmeyer, L., Habecker, B.A., 2011. Cardiac ischemia-reperfusion regulates sympathetic neuropeptide expression through gp130-dependent and independent mechanisms. Neuroptides 45 (1), 33–42. https://doi.org/10.1016/j.npep.2010.10.002.

Amino, M., Yoshioka, K., Morita, S., et al., 2007. Is the combination therapy of Ikr-channel blocker and left stellate ganglion block effective for intractable ventricular arrhythmia in a cardiopulmonary arrest patient? CardioI. J. 14 (4), 355–365.

Amiya, E., Watanabe, M., Komuro, I., 2014. The relationship between vascular function and the autonomic nervous system. Ann. Vasc. Dis. 7 (2), 109–119. https://doi.org/10.3400/avd.ra.14-00048.
Piedimonte, G., Rodriguez, M.M., King, K.A., McLean, S., Jiang, X. 1999. Respiratory syncytial virus upregulates expression of the substance P receptor in rat lungs. Am. J. Respir. Cell Mol. Biol. 27 (4), 483–491. https://doi.org/10.1165/racb.1999.27.4.483.

Pierach, A., Stottz, K. 1952. Therapy of pulmonary edema with procaine block of the right stellate ganglion. Desch. Med. Wochenschr. 77 (44), 1344–1346. https://doi.org/10.1002/dmww.2620774417.

Pichichero, A., Spagnolli, A. 2010. Das System der Grundregulation, 11. Aufl. Haug, Stuttgart.

Pongratz, G., Straub, R.H. 2014. The sympathetic nervous response in inflammation. Arthritis Res. Ther. 16 (6), 504. https://doi.org/10.1186/s13075-014-0054-2.

Puente de la Vega Costa, K., G€anke, N., Kiyono, H., Yoshiya, I. 1999. Lymphocyte activation is attenuated by stellate ganglion block. Reg. Anesth. Pain Med. 24 (3), 30–35. https://doi.org/10.1096/raj.99.3.9001621.

Tan, T., Khoon, B., Mills, E.G. 2020. Association between high serum total cortisol concentrations and mortality from COVID-19. Lancet Diabetes Endocrinol. https://doi.org/10.1016/S2213-8578(20)30126-3.

Tanevaya, C., Goto, H. 2009. Fractal cardiovascular dynamics and baroreflex sensitivity after stellate ganglion block. Anesth. Analg. 109 (4), 1325–1340. https://doi.org/10.1224/01.ANE.0b013e3181b10d8.

Tracey, K.J. 2002. The inflammatory reflex. Nature 420 (6917), 853–859. https://doi.org/10.1038/913121. PMID: 12490958.

Tran, K., 2019. Multiple bilateral ultrasound-guided stellate ganglion blocks to treat acute vasculitis in a recently diagnosed patient of systemic lupus erythematosus. Indian J. Anaesth. 63 (10), 851–855. https://doi.org/10.4103/ija.IJA_331_19.

Qin, C., Zhou, L., Hu, Z., et al. 2020. Dysregulation of immune response in patients with COVID-19 in Wuhan, China. Clin. Infect. Dis. ciaa248. https://doi.org/10.1093/cid/ciaa248.

Rahimzadeh, P., Mahmoudi, K., Khodaverdi, M., Faiz, S.H.R. 2020. Effects of ultrasound guided stellate ganglion block on treatment of viral infection. J. Infect. 80 (1), 157–162. https://doi.org/10.1016/j.jinf.2020.01.023.

Rahimzadeh, P., Mahmoudi, K., Khodaverdi, M., Faiz, S.H.R. 2020. Effects of ultrasound guided stellate ganglion block on treatment of viral infection. J. Infect. 80 (1), 157–162. https://doi.org/10.1016/j.jinf.2020.01.023.

Schaible, H.G., Ebersberger, A., Natura, G. 2011. Update on peripheral mechanisms of immune responses in laparoscopic gynecologic surgery. VideoSurgeroy Miniminv 15 (2), 351–357.

Ricker, G. 1924. Pathologie als Naturwissenschaft - Relatiopathologie. Springer, Berlin.

Ross, C.A., Ruggiero, D.A., Reis, D.J. 1985. Projections from the nucleus tractus solitarii to the rostral ventrolateral medulla. J. Comp. Neurol. 242 (4), 511–534. https://doi.org/10.1002/cne.902420405.

Ruan, Q., Yang, K., Wang, W., Wang, J., Song, J. 2020. Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. Intensive Care Med. 46 (5), 846–848. https://doi.org/10.1007/s00134-020-05991-x.

Salgado, D.R., Ortiz, J.A., Favorov, R., Creteur, J., Vincent, J.L., De Backer, D. 2010. Microparticulate abnormalities in patients with severe influenza A (H1N1) infection. Can. J. Anaesth. 57 (10), 940–946. https://doi.org/10.1007/s12630-009-9565-9.

Sander, J.M., Monogue, M.L., Jodlowski, T.Z., Cutrell, J.B. 2020. Pharmacologic treatments for coronavirus disease 2019 (COVID-19): a review. JAMA. https://doi.org/10.1001/jama.2020.0844.

Schaible, H.G., 2014. Noniceptive neurons detect cytokines in arthritis. Arthritis Res. Ther. 16 (5), 470. https://doi.org/10.1186/s13075-014-0470-8.

Schaible, H.G., Straub, R.H. 2014. Function of the sympathetic supply in acute and chronic experimental joint inflammation. Auton. Neurosci. 182, 55–64. https://doi.org/10.1016/j.autneu.2013.12.004. May.

Schaible, H.G., Ebersberger, A., Natura, G. 2011. Update on peripheral mechanisms of pain: beyond prostaglandins and cytokines. Arthritis Res. Ther. 13 (10). https://doi.org/10.1186/ar3805. Apr 28.

Schirfner, E.L. 2012. The endothelium system. In: Robertson, D., Biggioli, I., Burnstock, G., Low, P.A., Paton, J.F.R. (Eds.), Primer on the Autonomic Nervous System, 3rd ed. Academic Press, San Diego, pp. 125–139. https://doi.org/10.1016/B978-0-12386525-5.00028-7.

Serra, M.C., Bazzoni, F., Della Bianca, V., Grekowiski, M., Ross, F. 1988. Activation of human neutrophils by substance P. Effect on oxidative metabolism, exocytosis, cytosolic Ca2+ concentration and inositol phosphate formation. J. Immunol. 141 (2), 139. https://doi.org/10.4049/jimmunol.141.2.139.

Sugimoto, M., Shimaoka, M., Taenaka, N., Kiyono, H., Yoshiya, I. 1999. Lymphocyte proliferations by lymphocytes from Peyer’s patches, mesenteric lymph nodes, and spleen. J. Immunol. 136 (1), 152–159. https://doi.org/10.4049/jimmunol.136.1.152.

Sternberg, E.M., 2006. Neural regulation of innate immunity: a coordinated nonspecific host response to pathogens. Nat. Rev. Immunol. 6 (4), 318–328. https://doi.org/10.1038/nri1787.

Strong, J.A., Zhang, J.M., Schaeble, H.G. 2018. The sympathetic nervous system and pain. In: Wood, J.N. (Ed.), The Oxford Handbook of the Neurobiology of Pain. https://doi.org/10.1093/oxfordhb/9780190660239.013.0041.

Sugimoto, M., Ishii, M., Shimada, M., Todorov, V., Kanczewska, W., et al. 2020. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and the neuroendocrine stress axis. Mol. Psychiatry 1–7. https://doi.org/10.1038/s41386-020-0758-9.
Zhang, L., Yao, J., Zhang, T., Jin, J., Zeng, X., Yue, Z., 2013. Stellate ganglion block may prevent the development of neurogenic pulmonary edema and improve the outcome. Med. Hypotheses 80 (2), 158–161. https://doi.org/10.1016/j.mehy.2012.11.017.

Zhao, H.Y., Yang, G.T., San, N.N., Kong, Y., Liu, Y.F., 2017. Efficacy and safety of stellate ganglion block in chronic ulcerative colitis. World J. Gastroenterol. 23 (3), 533–539. https://doi.org/10.3748/wjg.v23.i3.533.

Zhao, Y., Xiang, R., Peng, X., et al., 2019. Transection of the cervical sympathetic trunk inhibits the progression of pulmonary arterial hypertension via ERK-1/2 signalling. Respir. Res. 20 (1), 121. https://doi.org/10.1186/s12931-019-1090-2.

Zheng, Y., Hou, X., Yang, S., 2019. Lidocaine potentiates SOCS3 to attenuate inflammation in microglia and suppress neuropathic pain. Cell. Mol. Neurobiol. 39 (8), 1081–1092. https://doi.org/10.1007/s10571-019-00753-6.

Zhou, Y., Yi, X., Xing, W., Hu, S., Maslov, K.I., Wang, L.V., 2014. Microcirculatory changes identified by photoacoustic microscopy in patients with complex regional pain syndrome type I after stellate ganglion blocks. J. Biomed. Opt. 19 (8), 086017. https://doi.org/10.1117/1.JBO.19.8.086017.

Zhu, Z., Cai, T., Fan, L., et al., 2020. Clinical value of immune-inflammatory parameters to assess the severity of coronavirus disease 2019. Int. J. Infect. Dis. 95, 332–339. https://doi.org/10.1016/j.ijid.2020.04.041.

van der Zypen, E., 1967. Elektronenmikroskopische befunde an der endausbreitung des vegetativen nervensystems und ihre deutung. Acta Anat. (Basel) 67 (4), 481–515.