The first described turtle beetles from Eocene Baltic amber, with notes on fossil Chelonariidae (Coleoptera: Byrrhoidea)

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Abstract. Chelonariidae, or turtle beetles, are rarely represented in the fossil record. Two new extinct species of this thermophilous coleopteran family, Chelonarium and-abata Alekseev and Bukejs sp. nov. and Ch. dingansich Alekseev and Bukejs sp. nov., are described and illustrated from Eocene Baltic amber using X-ray micro-computed tomography (micro-CT). They are the first formally described species of turtle beetles from Eocene Baltic amber and the first known European representatives of this family. Based on modern habitats of the group, the presence of the plants with which their larvae are associated (epiphytic orchids) is proposed in the Eocene amber forest. The Eocene Florissant Formation fossil Chelonarium montanum Wickham, 1914, which was originally placed within Chelonariidae, is discussed based on its original description, and placement as incertae sedis within Byrrhoidea is proposed for this compression fossil (http://zoobank.org/References/C2EE164D-59DD-42FE-937D-B01C78DCD228, last access: 8 February 2021).

1 Introduction

The coleopteran superfamily Byrrhoidea consists of 12 extant families (Ślipiński et al., 2011), and half of them have been reported from Eocene Baltic amber (Klebs, 1910; Larson, 1978; Kirejtshuk and Azar, 2013; Alekseev, 2019). Six fossil species were described from this Lagerstätte within Elmidae, Limnichidae and Ptilodactylidae (Bollow, 1940; Pütz et al., 2004; Bukejs et al., 2015; Alekseev and Jäch, 2016; Hernando et al., 2018), but the three remaining families (Byrrhidae, Heteroceridae and Chelonariidae) are still awaiting further attention from researchers to complete formal taxonomic descriptions from the amber deposit.

Chelonariidae are rarely represented in the fossil record. Until now, only two fossil species have been described as belonging to this family: (1) a dubious chelonariid fossil (see critical note in the Discussion) – Chelonarium montanum Wickham, 1914 – from the Eocene Florissant Formation (Wickham, 1914) and (2) Eochelonarium belle Kirejtshuk in Kirejtshuk and Azar (2013), a monotypic chelonariid beetle genus from Lower Cretaceous Lebanese amber (Kirejtshuk and Azar, 2013). Reports of chelonariid beetles from Eocene Baltic amber have been made in the recent literature (Kirejtshuk and Azar, 2013; Alekseev, 2019), but even generic assignment of the specimens is never mentioned in these works. In the present paper, the first turtle beetle species from Eocene Baltic amber of the Kaliningrad Region (western Russia) is described and illustrated using X-ray micro-computed tomography (micro-CT).
2 Material and methods

The material examined is deposited in the following collections:

- the Palaeontology Collection of the Royal Saskatchewan Museum (Regina, Saskatchewan, Canada) [RSKM specimen number prefix]
- the collection of the Museum of Amber Inclusions, University of Gdańsk (Poland) [MAIG].

The X-ray micro-CT observations of specimen RSKM_P3000.141 were conducted at the McGill University Integrated Quantitative Biology Initiative in Montreal, Canada, using a Zeiss Xradia 520 Versa system. Scans used an X-ray beam with an energy level of 60 kV and 83 µA, an LE6 filter, a source-to-sample distance of 38 mm, and sample to detector distance of 333 mm for phase contrast. Tomographic slices were generated from 1601 rotational steps through 180° of rotation, with a 0.4× objective and 5 s exposure times. Images were binned (2 × 2 × 2), with a resolution of 7.0 µm. The specimen fit within a single field of view, and a 40 min warmup scan was conducted. Dragonfly Pro (ver. 2020.1) software was used for segmentation and 3D visualization, as well as for producing videos of scan data.

The X-ray micro-CT observations of specimen 6696 [MAIG] were conducted at Daugavpils University Integrated Quantitative Biology Initiative in Latvia, using a Zeiss Xradia 510 Versa system. Scans were performed with a polychromatic X-ray beam with 40 kV of energy and 3 W of power. Sample–detector distance was set to 24 mm, and source-to-sample distance to 44.5 mm. Tomographic slices were generated from 1601 rotational steps through a 360° rotation using a 4× objective, and exposure time during each projection was set to 7 s. Variable exposure was set to two times at the thickest part of the amber to achieve similar amounts of photon throughput over the whole sample. Acquired images were binned (2 × 2 × 2), giving a voxel size of 4.4 µm. Since specimen length was bigger than the field of view for selected parameters, we carried out image acquisition using an automated vertical stitch function for three consecutive scans with identical scanning parameters. Between those scans field of view was set to overlap 41 % of data between adjacent fields of view. Prior to the full scan a 14 min warmup scan was conducted in which source–sample and sample–source distances were changed to accommodate the whole specimen in the field of view. For the warmup scan the rotational steps were reduced to 201, binning was set to 4 and exposure time was set to 1 s. Images were imported into the Dragonfly Pro (ver. 2020.1) software platform for interactive segmentation and 3D visualization.

Photographs of specimen RSKM_P3000.141 were taken using a Visionary Digital imaging system, consisting of a Canon MP-E 65 mm macrophotography lens, as well as a Dun Inc. microphotography system, consisting of Mitutoyo 5× and 10× Plan Apo objective lenses on a modified Canon EF 200 mm lens, attached to a Canon EOS 5D camera. The camera was on an automated camera lift with studio flash lighting. Supplementary images were taken with a Leica MZ12.5 stereomicroscope equipped with a Canon T6i camera. Extended depth of field at high magnifications was achieved by combining multiple images from a range of focal planes using Helicon Focus 6.8.0 software, and the resulting images were edited to create figures using Adobe Photoshop CS5.

Photographs of specimen 6696 [MAIG] were taken using a Canon 70D camera with a macro lens (Canon MP-E 65 mm). Extended depth of field at high magnifications was achieved by combining multiple images from a range of focal planes using Helicon Focus v. 6.0.18 software, and the resulting images were edited to create figures using Adobe Photoshop CS5.

The following references were used for the generic attribution and comparison with recent and fossil taxa: Méquignon (1932), Mandl (1967), Paulus (1969), Satô (2001), Ivie (2002), Leschen and Early (2004), Kirejtshuk and Azar (2013), and Beutel and Leschen (2016).

3 Systematic palaeontology

Superfamily Byrrhoidea Latreille, 1804

Family Chelonariidae Blanchard, 1845

Genus Chelonarium Fabricius, 1801

Remarks

The studied amber specimens possess the combination of characteristics unequivocally corresponding to the family Chelonariidae: biconvex body shape (i.e. fusiform in lateral view), head declined under semicircular pronotum and not visible in dorsal view, tarsomere 3 of the pentameric tarsi with long membranous lobe, pronotum and scutellum distinctly crenulate basally, pro- and mesosternum with deep groove-like excavation for reception of the basal antennomeres, ventral side of body with depressions for reception of legs, and anterior edge of pronotum curving downwards and posteriad.

Based on the presence of the above-mentioned characteristics and the following set of characteristics, both chelonarid specimens under consideration from Baltic amber are similar to Chelonarium representatives from the Recent fauna and can be considered as congeneric: (1) pronotum margined anteriorly and laterally with complete sharp carina (in contrast to pronotum without such completely surrounding sharp carina in Brounia Sharp or Pseudochelonarium Pic), (2) hypomera with depressions for reception of forelegs (absent in Brounia, only weakly developed in Eochelonarium Kirejtshuk), and (3) protibiae flattened (narrow in Eochelonarium and Brounia) and with spinose outer margin.
Chelonarium andabata Alekseev and Bukejs sp. nov.

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Figs. 1–5, 8b

Derivatio nominis

The specific epithet “andabata” is a Latin word meaning “a type of gladiator who fought blind because of a restrictive helmet” and refers to the strongly declined head of the beetle, which is concealed from above. The specific epithet is used as a noun in apposition.

Type material

Holotype: collection number “RSKM_P3000.141”, “Holotype/Chelonarium andabata sp. nov./Alekseev and Bukejs des. 2021” [red printed label] [RSKM]; adult, male. Almost complete beetle (right antennomeres 10–11 lacking) included in a transparent, yellow amber piece with approximate dimensions of 8 mm × 16 mm and a maximum thickness of 8 mm; preserved without supplementary fixation. The dorsal part of the specimen is partially obscured by milky amber. Syninclusions are absent.

Type stratum

Baltic amber from Eocene amber-bearing Blue Earth layers; a predominantly Bartonian age has been interpreted for the extinct central European resin-producing forests (Bukejs et al., 2019).

Type locality

Yantarny settlement (formerly Palmnicken), Sambian (Sambia) Peninsula, Kaliningrad Region, Russia.

Description

Measurements: body length 5.6 mm, maximum body width 3.2 mm; pronotum length 1.2 mm, maximum pronotum width 2.5 mm; elytra length 4.3 mm, maximum combined elytra width 3.2 mm.

Body ovoid, widely oval, biconvex; total body length / maximum body width equal to 1.75; integument unicolorous dark brown (as preserved). Pubescence unicolorous and dark: dorsum covered with short, semierect setae in moderate density; elytral vestiture uniform, simple, without any trichome-like structures, setae almost straight to slightly curved; setae at elytral lateral sides slightly stouter; venter and legs with dense, short recumbent setae.

Head declined, not visible from above, rounded pentagonal, slightly convex; densely punctate; punctures round and small, distance between punctures equal to 0.2–0.5× diameter of one puncture. Clypeus not distinct, frontoclypeal su-
ture absent. Eyes small, entire, almost round, slightly convex, finely faceted, without interfacetal setation; interocular frontal distance equal to about 2× diameter of one eye. Antennal insertions hidden. Antennae rather long; scape hidden, rounded and short; pedicel and antennomeres 3–11 flattened; antennomere 3 longest, antennomere 4 shortest; antennomeres 2–3 and 11 elongate, antennomeres 5–10 triangular in outline. Relative length ratios of antennomeres 1–11 equal to 1.5:3:7:0.7:1.8:2:2:2:2:2:3.4.

Pronotum transverse, 2.1× as wide as long, widest at base, with fine and dense punctuation. Posterior angles of pronotum acute. Anterior edge of pronotum almost semicircular in dorsal view; posterior edge bisinuate and crenulate. Pronotum margined anteriorly and laterally with sharp, raised carina dividing pronotum into upper (moderately convex dorsally) and lower (inclined posteriad) parts. Lower pronotal area (between prominent anterior carina and posterior edge of concealed head) wider than protibial width. Pro- and mesosternum with deep, elongate, intercoxal median excavation for reception of basal antennomeres. Hypomeron excavated to receive profemora; meso- and metaventrite with excavations for receiving meso- and metafemora as well as tibiae.

Scutellum subpentagonal, almost as long as wide. Elytra moderately convex, widely oval, about 1.4× as long as combined width; distinctly wider than pronotal posterior margin, about 3.6× as long as pronotum. Elytral punctuation irregular, dense and round along sides, sparser and finer on disc, distance between punctures equal to 0.3–3.0× diameter of one puncture, interspaces flat. Epipleura narrow, reaching abdominal ventrite 1. Metepisternum wide, about 2× as wide as maximum epipleural width, with dense, small punctuation. Metaventrite slightly convex laterally, with almost flat disc; densely covered with small punctuation; distance between punctures smaller than diameter of one puncture; interspaces slightly convex; discrimen distinct in anterior half of metaventrite.

Legs rather short, flattened; densely covered with small punctures; distance between punctures smaller than diameter of one puncture. All coxae distinctly separated; pro- and mesocoxae widely oval, slightly transverse, about 1.3–1.4× as wide as long; metacoxae narrow, strongly transverse, excavate, with short metacoxal plates and with triangular outer edge. Femora and tibiae subequal in length; pro- and mesofemur comparatively wider than metafemur. Tibiae narrower than femora, protibia about 0.5× as wide as profemur, mesotibia about 0.5× as wide mesofemur, metatibia about 0.6× as wide as metafemur; protibia spinose along inner margin. Tarsi 5 segmented, tarsomere 3 with long membranous lobe ventrally, tarsomere 4 minute. Relative length ratios of mesotarsomeres 1–5 equal to 15:10:7:4:25. Pretarsal claws with denticle basally.

Abdomen with five visible ventrites; abdominal sutures entire, slightly concave; ventrite 5 simple, with widely rounded, semicircular apical margin; finely and densely punctate; distance between punctures equal to 0.5–1.0× di-
ameter of one puncture. Relative length ratios of abdominal ventrites 1–5 equal to 7:4.5:4:3.5:6.5 (medially).

Aedeagus robust, with widely rounded base and gradually narrowed apically (Fig. 5).

Differential diagnosis

The genus *Chelonarium* is very species-rich but poorly documented: it is in need of revision for many of its inadequately illustrated and briefly described species. The Eocene taxon *Ch. andabata* sp. nov. is similar to one Recent North American chelonariid species, *Ch. lecontei* Thomson, in its general antennal structure, but it differs in having smaller eyes (inte-
ocular distance in *Ch. lecontei* is slightly less than one eye diameter), denser head punctation and uniform elytral vestiture (without maculae). The newly described fossil species can be distinguished from the westernmost Palaearctic *Ch. vartianae* Mandl (Afghanistan) based on having a pronotum without transverse impression. *Chelonarium andabata* sp. nov. clearly differs from *Ch. ornatum* Klug in its ratio of four basal antennomere lengths; it differs from *Ch. kurosawai* Satō in possessing unicolorous elytra; and it differs from *Ch. beauvoisi* Latreille, *Ch. convexum* Méquignon and *Ch. cupreum* Méquignon in its smaller body size. The combination of characteristics mentioned in the species description, additional characteristics of body part ratios, and details of vestiture and punctation should distinguish the new fossil species from all Recent congeners.
Figure 3. X-ray micro-CT renderings of *Chelonarium andabata* Alekseev and Bukejs sp. nov., holotype, RSKM_P3000.141 [RSKM], habitus: (a) ventral view without legs, showing depressions for legs reception; (b) ventral view with antennae and legs in different colours; (c) frontal view; and (d) caudal view. Scale bar = 1.0 mm.

*Chelonarium dingansich* Alekseev and Bukejs sp. nov.
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Figs. 6–7, 8a

*Derivatio nominis*

The specific epithet “*dingansich*” is derived from the German word combination “Ding an sich” (in English: “thing in itself”), a well-known philosophic concept introduced by Immanuel Kant. The species name is dedicated to this famous Königsberg (now Kaliningrad)-native Prussian philosopher and thinker, in the run-up to the 300-year anniversary of his birth in 2024. The name is used as a noun in apposition.

*Type material*

Holotype: collection number “6696” [MAIG] (ex. coll. Jonas Damzen JDC 7210), “Holotype/Chelonarium dingansich sp. 
Figure 4. Antennae of *Chelonarium andabata* Alekseev and Bukejs sp. nov., holotype, RSKM_P3000.141 [RSKM]: (a) X-ray micro-CT rendering and (b) reconstruction. Abbreviations: a1–a11 – antennomeres 1–11 respectively.

Figure 5. X-ray micro-CT renderings of *Chelonarium andabata* Alekseev and Bukejs sp. nov., holotype, RSKM_P3000.141 [RSKM], aedeagus: (a) dorsal view, (b) lateral view and (c) ventral view. Scale bar = 0.1 mm.
Figure 6. Photomicrographs of *Chelonarium dingansich* Alekseev and Bukejs sp. nov., holotype, 6696 [MAIG], habitus: (a) dorsal view, (b) ventral view and (c) left lateral view. Scale bar = 1.0 mm.

nov./Alekseev and Bukejs des. 2021” [red printed label] [MAIG]; adult, sex apparently female. Almost complete beetle (antennomeres 4–11 of both antennae lacking) included in a transparent, yellow amber piece with approximate dimensions of 40 mm × 20 mm and a maximum thickness of 10 mm; preserved without supplementary fixation. Syninclusions: four very small stellate trichomes and a few small gas vesicles.

**Type stratum**

Baltic amber from Eocene amber-bearing Blue Earth layers; a predominantly Bartonian age has been interpreted for the extinct central European resin-producing forests (Bukejs et al., 2019).
Figure 7. X-ray micro-CT renderings of *Chelonarium dingansich* Alekseev and Bukejs sp. nov., holotype, 6696 [MAIG], habitus: (a) dorsal view, (b) ventral view and (c) right lateral view. Scale bar = 1.0 mm.

**Type locality**

Yantarny settlement (formerly Palmnicken), Sambian (Samland) Peninsula, Kaliningrad Region, Russia.

**Description**

Measurements: body length 6.2 mm, maximum body width 2.7 mm; pronotum length 1.4 mm, maximum pronotum width 2.3 mm; elytra length 4.9 mm, maximum combined elytra width 2.7 mm.

Body elongate oval, biconvex; total body length/maximum body width equal to 2.3; integument unicolorous dark brown (as preserved). Pubescence dark, with small patches of denser and paler setae on elytral disc: dorsum covered with short, semierect setae in moderate density; lateral sides of elytra and pronotum with slightly
longer, distinctly curved setae; elytral vestiture simple, without any trichome-like structures; venter and legs with dense, recumbent, almost straight setation, setae shorter than setae of dorsum.

Head declined, not visible from above, slightly convex; densely covered with small punctuation; distance between punctures smaller than diameter of one puncture. Clypeus not distinct; frontoclypeal suture absent. Eyes small, entire, almost round, slightly convex, finely faceted, without interfacetal setation; interocular frontal distance equal to about $2 \times$ diameter of one eye. Antennal insertions hidden. Antennae rather long; scape hidden; pedicel short, about $0.4 \times$ as long as antennomere 3; antennomere 3 longest, flattened.

Pronotum transverse, $1.6 \times$ as wide as long, widest at base; densely covered with small punctuation; distance between punctures less than diameter of one puncture. Posterior angles of pronotum acute. Anterior edge of pronotum almost semicircular in dorsal view; posterior edge bisinuate and crenulate. Pronotum margined anteriorly and laterally, with sharp, raised carina dividing pronotum into upper (moderately convex dorsally) and lower (inclined posterior) parts. Lower pronotal area, between prominent anterior carina and posterior edge of concealed head, wider than protibial width. Pro- and mesosternum with deep, elongate, intercoxal median excavation for reception of basal antennomeres. Hypomeron excavated to receive profemora; meso- and metaventrite with excavations for receiving meso- and metafemora and tibiae.

Scutellum subpentagonal, almost as long as wide, densely covered with semierect, pale setae. Elytra moderately convex, forming elongate oval about $1.9 \times$ as long as combined width; slightly wider than pronotal posterior margin, about $3.6 \times$ as long as pronotum. Elytral punctuation irregular, small, dense and round, with distance between punctures less than
diameter of one puncture; interspaces almost flat. Epipleura narrow, reaching abdominal ventrite 1. Metepisternum wide, about \(2 \times\) as wide as epipleural maximum width; with dense, small punctation. Metaventrite slightly convex laterally, with almost flat disc; densely covered with small punctation; distance between punctures smaller than diameter of one puncture; interspaces slightly convex; discriment distinct in anterior half of metaventrite.

Legs rather short, flattened; densely covered with small punctures; distance between punctures smaller than diameter of one puncture. All coxae distinctly separated; procoxa nearly round; mesocoxa oval; metacoxa narrow, strongly transverse, excavate, with short metacoxal plates and with triangular outer edge. Femora and tibiae subequal in length; pro- and mesofemora comparatively wider than metafemur. Tibiae narrower than femora, protibia about 0.6\(\times\) as wide as profemur, mesotibia about 0.3\(\times\) as wide mesofemur, metatibia about 0.6\(\times\) as wide as metafemur; protibia spinose along inner margin. Tarsi 5 segmented, tarsomere 3 apparently with membranous lobe ventrally, tarsomere 4 minute. Tarsal claws with denticle basally.

Abdomen with five visible ventrites; abdominal sutures entire, slightly concave to almost straight; ventrite 5 simple, with widely triangular apical margin; finely and densely punctate; distance between punctures equal to 0.5–1.0\(\times\) diameter of one puncture. Relative length ratios of abdominal ventrites 1–5 equal to 6:4:4:3.5:7 (medially).

### Differential diagnosis

*Chelonarium dingansich* sp. nov. differs from *Ch. andabata* sp. nov. in possessing longer and distinctly curved setae on elytral and pronotal lateral sides (setae comparatively shorter and almost straight in *Ch. andabata* sp. nov.); an elytral disc with small patches of paler setae (without small patches of paler setae in *Ch. andabata* sp. nov.); a widely rounded, semicircular apical margin of abdominal ventrite 5 (widely triangular in *Ch. andabata* sp. nov.); and a distinctly more elongate oval body: less transverse pronotum, 1.6\(\times\) as wide as long (2.1\(\times\) as wide as long in *Ch. andabata* sp. nov.), and more elongate elytra, 1.9\(\times\) as long as combined width (1.4\(\times\) as long as combined width in *Ch. andabata* sp. nov.).

### Remarks

The sex of the examined specimen is determined based on micro-CT results. There is no sclerotized aedeagus-like structure present inside the abdomen; therefore, the specimen appears to be female.

### 4 Discussion

#### 4.1 Zoogeographical and ecological notes

Chelonariidae are a small coleopteran family, which comprises three Recent genera with about 300 described species that are predominantly distributed in tropical regions (Beutel and Leschen, 2016). *Brounia* Sharp is restricted to New Zealand; *Pseudoche lonarium* Pic (consisting of two subgenera, *Pseudoche lonarium* and *Neochelonarium* Méquignon) occurs in New Guinea, India, eastern and southeastern Asia; and *Chelonarium* Fabricius is mainly found in the neotropics but also occurs in Asia and Australia (Leschen and Early, 2004; Beutel and Leschen, 2016). In the present-day Palaeartic region, Chelonariidae are restricted to the southernmost periphery of eastern Asia (Afghanistan; Bhutan; China; Hainan; India; Japan; Taiwan) and represented by four species of *Chelonarium* and seven species of *Pseudocha lenarium* (Satô, 2016). No representatives of the family are known from modern Europe or Africa.

Chelonariid larvae are probably detritus feeders in moist-to-dry litter on the ground. Their known habitats include the bases of trees, orchids and other plants; the refuse heaps of ants or termite galleries in branches; and under the bark of dead trees (Ivie, 2002). A preference for the roots of orchids and tree epiphytes, as well as myrmecophilous or termitophilous tendencies, has been suggested (Beutel and Leschen, 2016); however, detailed observations of larval feeding habits are lacking. Interestingly, European Eocene amher-producing forests were thermophilous, humid mixed forest communities that grew in a climate thought to have had low seasonality (Alekseev, 2017; Bogri et al., 2018, 2020).

Both of the larval habitats that have been recorded for Recent congeners of the newly described fossil species have also been documented for this Lagerstätte: (1) the oldest orchid fossil known at present was recently described from Baltic amber on the basis of orchid pollinaria (Poinar and Rasmussen, 2017), and (2) the fauna of termites and ants in the Baltic amber forest was diverse and species-rich (e.g. Wheeler, 1915; Engel et al., 2007). A similar biology is tentatively suggested for *Chelonarium andabata* sp. nov. and *Ch. dingansich* sp. nov., with the fossil taxa having been subcortical or inquiline in moist forest habitats with numerous overmatured trees and epiphytic orchids, and with an abundance of different termite and ant species.

The presence in Baltic amber of representatives belonging to an extant coleopteran genus often associated with epiphytic orchids may provide additional support for (1) the presence and comparative abundance of their host plants (Orchidaceae, possibly Epidendroideae, from tribes Dendrobieae and/or Epidendreae) in the Eocene amber forest and (2) the possible epiphytic nature of these orchids. Given the scarcity of data about Eocene orchids, and the indirect connection between these plants and fossil Chelonariidae, it will not be possible to establish their habitat connection with con-
clear. According to the ICZN paragraph 32.2.1 (i.e. if a name is spelled in more than one way in the work in which it was established, then, except as provided otherwise in this article, the correct original spelling is that chosen by the first reviser), and the ICZN paragraph 24.2.3 (selection of correct original spellings), we decided the possible name confusion should be discussed and prevented. To the best of our knowledge, no published papers have used the name *Eochelonarium bellum* or *Eochelonarium belle*, and an electronic database (Clapham, 2014) included only the variant “*Eochelonarium belle*” without any comments. Consequently, we can act as the “first reviser” in this dubious case.

As it is stated in the original etymology, the epithet of this new species means “beautiful”, “handsome”, “good-looking”, “fine” and “pretty”. The one variant is “*belle*”, and the second is “*bellum*”, both of which overlap this meaning to some extent. The word “*belle*” is a French adjective meaning “attractive, pretty, handsome etc.” or a Latin adverb meaning “beautifully, attractively etc.”. The word “*bellum*” in Latin as a substantive means “war”, or it functions as an adjective for “beautiful, handsome etc.” in a neutral case. Based on grammatical correctness of the species name, *Eochelonarium bellum* would be preferred, as an amendment to the grammatically incorrect “*belle*” variant. On the other hand, the name “*belle*” was used first in the paper (in the summary), was used as the name for the type species of the new genus (Kirejtshuk and Azar, 2013, p. 116) and was used by the author of the original description in an online catalogue of fossils (Kirejtshuk and Ponomarenko, 2014); meanwhile *Eochelonarium bellum* only appears in figure captions. On the basis of these arguments, we select the name *Eochelonarium belle* as correct and propose the use of the specific epithet as a noun in apposition.

**Data availability.** All material included in the paper is deposited in the Palaeontology Collection of the Royal Saskatchewan Museum (Regina, Saskatchewan, Canada) [RSKM] and in the collection of the Museum of Amber Inclusions, University of Gdańsk (Poland) [MAIG], and all data are included in the description. X-ray microtomography volume renderings of the habitus, habitus without legs, antennae and aedeagus of *Chelonarium andabata* sp. nov., holotype, RSKM_P3000.141 [RSKM], and habitus of *Ch. dingansich* sp. nov., holotype, 6696 [MAIG], are available as video supplements.

**Video supplement.** The videos are available at
- [https://doi.org/10.5446/49584](https://doi.org/10.5446/49584) (Mitchell et al., 2021a)
- [https://doi.org/10.5446/49583](https://doi.org/10.5446/49583) (Mitchell et al., 2021b)
- [https://doi.org/10.5446/49582](https://doi.org/10.5446/49582) (Mitchell et al., 2021c)
- [https://doi.org/10.5446/49581](https://doi.org/10.5446/49581) (Mitchell et al., 2021d)
- [https://doi.org/10.5446/49580](https://doi.org/10.5446/49580) (Bukejs and Kairišs, 2021).
Author contributions. VIA and AB designed the study, identified specimens, performed systematic placement and prepared new species descriptions. HCEL performed micro-CT measurements. JM reconstructed micro-CT data and created volume renderings and 3D models of Chelonarium andabata sp. nov. with help from MB. VIA and AB drafted the discussion. RCM located and prepared the specimen and prepared photomicrographs of Chelonarium andabata sp. nov. and plates. All authors drafted the manuscript and contributed to the writing and discussion.

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