Data Article

Whole-genome sequence data of the proteolytic and bacteriocin producing strain Enterococcus faecalis PK23 isolated from the traditional Halitzia cheese produced in Cyprus

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A B S T R A C T

Halitzia is a traditional white-brined cheese produced by a limited number of producers in Cyprus. During a survey of the microbiome of a number of different Halitzia samples, we identified a bacterial strain that exhibited enhanced proteolytic activity compared to the other isolates. The strain was further studied, and it was assigned as Enterococcus faecalis PK23. We proceeded with sequencing of its whole genome using Illumina technology. Initial sequencing and assembly produced 116 scaffolds with a length of 3,149,036 bp. Comparison with the available E. faecalis genomes revealed that the strain PK23 exhibited high levels of identity to the genome sequence of E. faecalis isolate 26975_2#180 deposited in GenBank as a single complete contig. From the 116 scaffolds 106 could be aligned to the genome of isolate 26975_2#180 leading to a chromosomal length of 3,132,784

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Specifications Table

| Subject                          | Food Science: Food Microbiology                                      |
|---------------------------------|------------------------------------------------------------------------|
| Specific subject area           | Genome sequencing and in silico analysis                               |
| Type of data                    | Genome sequence; Tables and Figures                                   |
| How data were acquired          | Genome sequencing: Illumina HiSeq 2000 platform, Trimming of adaptors: | |
|                                 | BBduk, De novo sequence assembly: SPAdes-3.12.0, Chromosomal alignments: | |
|                                 | r2cat and MAUVE, Sequence annotation: Rapid Annotation using Subsystem | |
|                                 | Technology (RAST) version 2.0 server indicated the presence of seven putative | |
|                                 | genes which were related to the subsystem of Protein Degradation. This | |
|                                 | dataset provides a first overview of the proteolytic and bacteriocin producing | |
|                                 | properties of E. faecalis PK23. The dataset may also be used in future | |
|                                 | experiments which could shed light on the adaptation of the strain in the | |
|                                 | dairy environment and its role in cheese production.                  |
| Data format                     | Raw Illumina paired-end sequence reads in fastq files, fasta and genbank | |
| Parameters for data collection  | files formats of annotated sequence scaffolds                           |
| Description of data collection  | Enterococcus faecalis strain PK23 was isolated from traditional Halitza cheese | |
|                                 | produced in Cyprus and the genomic DNA sequenced was isolated from the pure | |
|                                 | culture                                                                |
| Data source location            | Laboratory of Food Quality Control and Hygiene, Department of Food Science | |
|                                 | and Human Nutrition, Agricultural University of Athens, 11855 Athens, Greece | |

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Data accessibility

Data are deposited in the respective databases and are publicly available. Raw sequence data of strain Enterococcus faecalis PK23 was deposited in the Sequence Read Archive (SRA) under accession number SRX11406112 (https://trace.ncbi.nlm.nih.gov/Traces/sra/?run=SRR15096257).

The whole-genome sequence of Enterococcus faecalis PK23 has been deposited in GenBank under accession number JAHBBU000000000 (https://www.ncbi.nlm.nih.gov/nuccore/JAHBBU000000000). In GenBank the sequence can be found annotated by PGAT.

The BioProject ID in GenBank is: PRJNA729111

The annotated genome is also available through the Rast version 2.0 server for anyone logging in with the guest account under the genome ID 1351.5033.

Value of the Data

- *Enterococcus faecalis* has been identified very frequently as part of the microbiome of artisanal cheeses. The genome sequence of strain PK23 may aid understanding the adaptation mechanisms which underpin its growth in the dairy environment.
- Data included in this manuscript may be useful for researchers in the field of Dairy Microbiology.
- Data presented here may be useful to understand the differences and similarities between commensal/pathogenic and dairy *E. faecalis* strains through comparative and evolutionary genomics.
- *E. faecalis* PK23 presents potentially important technological properties which warranty further investigation, including its proteolytic potential and bacteriocin production.

1. Data Description

Halitza is a rare white-brined cheese produced in Cyprus. It is named after its shape which resembles stones or pebbles [1]. During a recent screening of the microbial ecosystem of a number of Halitza cheese samples, we isolated a strain exhibiting enhanced proteolytic and bacteriocin producing activity compared to other isolates (our unpublished results). The strain was identified and assigned as *Enterococcus faecalis* PK23. The proteolytic activity of enterococci has been suggested to be more pronounced than other lactic acid bacteria (LAB) [2]. It is an important technological property since it contributes to the cheese ripening process, the generation of bioactive peptides or even the reduction of the allergenicity of bovine milk proteins [2–4]. Nevertheless, enterococcal proteolytic enzymes have also been related to the pathogenicity of clinical strains [5]. Furthermore, the ability of enterococci to produce bacteriocins against foodborne pathogens and food spoilage microorganisms is also an important technological trait [6].

The initial assembly of the paired-end Illumina sequencing reads resulted in 116 scaffolds. After manual BLASTn searches [7], a number of closely related *E. faecalis* genomes were identified and downloaded from GenBank. Further analysis with the Average Nucleotide Identity (ANI) calculator [8] and the Genome-to-Genome Distance Calculator (GGDC) 2.1 [9] revealed that the closest related genome currently available for strain PK23 was the chromosomal sequences of *E. faecalis* isolate 26975_2#180. The two strains exhibited an ANI value of 99.88 and a dDDH value of 99.20. Alignment of the 116 PK23 scaffolds against the sequence of isolate 26975_2#180 resulted in the ordering of 106 of them (Fig. 1A). The ordered scaffolds had a length of 3,132,784 bp and a GC content of 37.3% (Table 1). The circular map of the sequence indicated that the organization of the PK23 pseudochromosome had a typical bacterial GC skew from the replication initiation region towards the replication termination region (Fig. 1B). Five additional scaffolds of approx. 2.7 kb could be correlated to chromosomal *E. faecalis* regions according to BLASTn.
Fig. 1. (A) Alignment of the Enterococcus faecalis PK23 pseudochromosome (i.e. concatenated ordered scaffolds) against the chromosome sequence of E. faecalis isolate 26975_2#180 using MAUVE. MAUVE uses local collinear blocks to align regions of high identity. (B) Circular map of the PK23 pseudochromosome generated using DNAPlotter. Genomic features drawn from the periphery to the centre of the map: 1. Forward CDSs (red); 2. Reverse CDSs (blue); 3. %GC plot; 4. GC skew.

Table 1
Statistics of the assembled sequence of Enterococcus faecalis PK23 according to the RAST version 2.0 server.

| Sequence trait                  | Value  |
|---------------------------------|--------|
| Size (bp)                       | 3,149,036 |
| GC Content (%)                  | 37.3%  |
| Number of Scaffolds             | 116    |
| Number of Subsystems            | 253    |
| Number of Coding Sequences      | 3,161  |
| Number of RNAs                  | 62     |

searches. These scaffolds could not be ordered with the rest of the PK23 scaffolds as they were probably absent from the chromosome of isolate 26975_2#180. Finally, five scaffolds exhibited high identity to plasmid sequences. More specifically, scaffold 54 showed high identity with the entire length of plasmid pEF1071 of E. faecalis strain BFE 1071 [10] while scaffold 77 showed high identity with the entire sequence of the cryptic plasmid unnamed_5 of Enterococcus faecium strain PR05720-3. The final three scaffolds (i.e. scaffold 88, 105 and 106) represented short fragments of larger plasmids.

Annotation with Rapid Annotation using Subsystem Technology (RAST) version 2.0 server [11] of the whole genome sequence of E. faecalis PK23 identified 3161 coding sequences and 62 RNA sequences (Table 1). From the total number of proteins encoded in the genome of strain PK23 27% and 73% could or could not be assigned in subsystem categories, respectively. The most
abundant subsystem feature counts were Carbohydrates (205 counts), Amino Acids and Derivatives (157 counts) and Protein Metabolism (154 counts) (Fig. 2). Given the proteolytic activity of strain PK23, preliminary analysis identified seven putative proteolytic enzymes in subsystems counts for Protein Degradation (Table 2). Additionally, annotation of scaffold 54 showed that it carries the biosynthetic genes for enterocins 1071A and 1071B described previously for plasmid pEF1071 [10]. This observation may explain the antimicrobial activity of the strain against certain foodborne pathogens (our unpublished results).

2. Experimental Design, Materials and Methods

E. faecalis PK23 was isolated from traditional Halitza cheese. The culture was routinely grown in M17 broth (Oxoid) at 37 °C statically. High quality genomic DNA was extracted from the PK23 strain with the PureLink® Genomic DNA Mini Kit (Invitrogen, Life Technologies) according to the manufacturer’s instructions. Genome sequencing was performed by SNPzsaurus (Eugene, OR) using an Illumina HiSeq 2000 platform (Illumina, CA). Standard workflows were followed for library preparation, sequencing, read trimming and assembly. In detail, a Nextera kit (Illumina) was used for library preparation, followed by 2 × 150-bp paired-end read sequencing, trimming of adaptors with BBduk (https://sourceforge.net/projects/bbmap) and scaffold assembly with SPAdes-3.12.0 using default parameters [12]. This analysis generated total sequence corresponding to > 200x coverage of the PK23 genome. As mentioned above, the closest E. faecalis genomes to the PK23 scaffolds were identified by manual BLASTn searches [7]. These genomes were then
analysed against the PK23 scaffolds with the ANI calculator [8] and the GGDC 2.1 [9]. PK23 scaffolds were ordered against the chromosomal sequence of E. faecalis isolate 26975_2#180 using the r2cat [13] and MAUVE [14] tools. DNAPlotter was employed for drawing the circular map of the PK23 pseudochromosome [15]. The scaffolds which could not be aligned to the reference genome were analysed with BLASTn to determine whether they derived from chromosomal or plasmid sequences. The entire whole-genome sequence of strain PK23 was annotated with the RAST version 2.0 server [11] and the Prokaryotic-genome Analysis Tool (PGAT) [16].

3. Nucleotide Sequence Accession Number

Raw sequence data of strain Enterococcus faecalis PK23 was deposited in the Sequence Read Archive (SRA) under accession number SRX11406112. The Whole Genome Shotgun project has been deposited at DDBJ/ENA/GenBank under the accession JAHBBU0000000000. The version described in this paper is version JAHBBU010000000.

Ethics Statement

N/A.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships which have or could be perceived to have influenced the work reported in this article.

CRediT Author Statement

Konstantinos Papadimitriou: Supervision, Conceptualization, Methodology, Data curation, Writing – original draft; Anastasia Venieraki: Methodology, Writing – original draft; Markella Tsigkrimani: Formal analysis, Writing – original draft; Panagiotis Katinakis: Conceptualization, Methodology, Data curation, Writing – original draft; Panagiotis N. Skandamis: Supervision, Conceptualization, Methodology, Data curation, Writing – original draft.

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