Glutamine, fish oil and antioxidants in critical illness: MetaPlus trial post hoc safety analysis

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Abstract

Background: The role of plasma glutamine, fish oil and antioxidants concentrations in the treatment effect of immune-modulating high-protein versus high-protein enteral nutrition on 6-month mortality in critically ill patients is explored, as unexpected negative outcomes of recent large randomized controlled trials on immune-modulating nutrients have raised questions about safety of these interventions.

Methods: Post hoc analysis of the MetaPlus randomized controlled trial which was performed in a total of 301 medical, surgical and trauma critically ill patients in fourteen European intensive care units. Patients received either immune-modulating (glutamine, fish oil and antioxidant enriched) high-protein (IMHP) or isocaloric high-protein (HP) enteral nutrition. Six-month mortality and baseline, day 4 and day 8 plasma concentrations of glutamine, (eicosapentaenoic acid + decosahexaenoic acid)/long-chain fatty acid plasma level ratio ((epa + dha)/lcf ratio), selenium, vitamin c, vitamin e and zinc were measured.

Results: The harmful treatment effect of the IMHP versus HP enteral nutrition on 6-month mortality was only demonstrated in the medical subgroup (HR 2.52, 95% CI 1.36–4.78, \( P = 0.004 \)). Among medical patients, when corrected for age groups and APACHE-II scores, there were no statistically significant associations between baseline plasma levels and 6-month mortality, except for zinc (HR 1.06, 95% CI 1.00–1.12, \( P = 0.026 \)). IMHP feeding resulted in statistically significant increase in plasma levels of glutamine, vitamin e, vitamin c and (epa + dha)/lcf ratio from baseline to day 4, while only the change from baseline to day 4 of (epa + dha)/lcf ratio was statistically significant associated with 6-month mortality (HR 1.18, 95% CI 1.02–1.35, \( P = 0.021 \)) and identified as mediator for the harmful treatment effect of IMHP enteral nutrition among medical ICU patients.

Conclusion: We hypothesize that the harmful effect of IMHP compared to HP enteral nutrition in a heterogeneous group of critically ill patients is limited to the medical critically ill patients and mediated by an early increase in (epa + dha)/lcf ratio.

Trial Registration Dutch Trial Register 26 January 2010 (NTR2181 http://www.trialregister.nl/trialreg/admin/rctview.asp?TC=2181).

Keywords: Critically ill, Mortality, Glutamine, Fish oil, Antioxidants, Nutritional support, Immune-modulating nutrients, Enteral nutrition, Clinical outcome

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Safety of immune-modulating nutrients like glutamine, fish oils and antioxidants to modulate infectious morbidity and enhance recovery from critical illness is under debate [1–6]. Recent large randomized trials in ICU patients failed to demonstrate benefits of immune-modulating nutrients and even demonstrated increased mortality [7–11].

Unexpected negative study results raised concern about safety of immune-modulating nutrients. It has been suggested that supplementation with immune-modulating nutrients should be reserved for specifically identified patients with compromised availability and plasma levels should be measured before supplementation [5]. However, low plasma levels could reflect adaptive and beneficial stress responses rather than conditional deficiency. The conditional deficiency hypothesis of glutamine has been challenged [12], and it has been suggested that interfering with a potential adaption could be deleterious [3].

This could suggest that supplementation with immune-modulating nutrients in critically ill patients does not improve outcome [3] and for safety reasons any patient with multi-organ failure in the ICU should not receive immune-modulating nutrients [2].

Studying mortality in critical illness and plasma status of immune-modulating nutrients after supplementation may provide pathophysiological insight and potentially facilitate redefinition of practice guidelines recommendations.

In the MetaPlus study, immune-modulating high-protein (IMHP) enteral feeding enriched with glutamine, fish oil and antioxidants was compared with standard high-protein (HP) enteral feeding in critically ill patients, and a hazard ratio (HR) of 1.57 (95% CI, 1.03–2.39; P = 0.04) was shown for 6-month mortality adjusted for age and APACHE-II score comparing IMHP with HP [7]. Univariate analysis suggested that this harmful effect was mainly observed among medical patients, although interaction between “type of patient” and “treatment” was not tested. The MetaPlus study is unique as plasma concentrations of glutamine, eicosapentaenoic acid + docosahexaenoic acid)/long-chain fatty acid plasma level ratio ((epa + dha)/lcf ratio), selenium, vitamin c, vitamin e and zinc were obtained at baseline (before treatment) and at days 4 and 8.

In this post hoc analysis, we address the interaction between IMHP versus HP and medical versus non-medical patients and 6-month mortality and subsequently associations of baseline plasma levels and changes in immune-modulating nutrient plasma levels and 6-month mortality.
and “Acute Physiology and Chronic Health Evaluation-II (APACHE-II) scores” as covariates within each subgroup. Associations of baseline plasma concentrations and 6-month mortality were tested using univariate Cox proportional hazard regression and multivariate Cox proportional hazard regression with “age groups” and “APACHE-II score” as covariates.

Differences between IMHP and HP in plasma concentration changes from baseline to day 4 and from baseline to day 8 were analyzed as continuous variables using two-sample t tests. The criterion was set at $P$ value <0.05.

The relation between plasma nutrient status changes and 6-month mortality was evaluated with multivariate Cox proportional hazard models for 6-month mortality including “age groups”, “APACHE-II score” and “baseline plasma nutrient concentrations” as covariates.

To assess which plasma nutrient changes mediated the effects on 6-month mortality, the causal steps approach was used [13]. This approach requires estimating each path depicted in Fig. 1.

Model I in Fig. 1 represents treatment effects of IMHP versus HP on 6-month mortality as analyzed using Cox proportional hazard regression analysis with “age groups” and “APACHE-II score” as covariates. In Model II, changes in plasma concentrations are suggested to mediate the relationship between treatment (IMHP vs. HP) and outcome (6-month mortality) when both the A and B association paths are statistically significant present and the C′ path association coefficient is closer to zero than in the C path in model I.

In Model II, the A path depicts the relation between the intervention (IMHP vs. HP) and changes in plasma nutrient status. This was analyzed using two-sample t tests. The B path describes the relation between changes in plasma nutrient status and 6-month mortality and analyzed using multivariate Cox proportional hazard models for 6-month mortality including “age groups”, “APACHE-II score” and “baseline plasma nutrient concentrations” as covariates. The Cox proportional hazard regression model with “age groups” and “APACHE-II score” as covariates was used to evaluate the C and C′ path, only for potential mediators having significant ($P < 0.05$) associations in both the A and B paths. The C′ path is evaluated in a Cox proportional hazard regression model including changes in plasma nutrient status variables as possible mediators.

Statistical analyses were performed on an intention-to-treat basis with SAS software, version 9.2 (SAS Institute Inc, Cary, USA).

**Results**

“Type of patient (medical vs. non-medical)” was the only factor that interacted with the effect of “treatment group (IMHP vs. HP)” (interaction term $P = 0.051$). Table 1 a/b depicts the IMHP versus HP treatment effects among medical and non-medical patients, showing a statistically significant treatment effect among medical patients with a HR of 2.52 (95% CI 1.36–4.78, $P = 0.004$) and no treatment effect in the non-medical group with an HR of 0.97 (95% CI 0.54–1.74, $P = 0.909$).

Associations between baseline plasma concentrations and 6-month mortality in all patients and in medical and non-medical subgroups are shown in Table 2. Univariate analyses showed statistically significant associations between baseline glutamine and 6-month mortality and between (epa + dha)/lcf ratio and 6-month mortality. Baseline zinc plasma concentrations were statistically significant associated with 6-month mortality in non-medical patients. However, when corrected for “age groups” and “APACHE-II scores,” there were no statistically significant associations, except for baseline zinc concentration and 6-month mortality among medical patients.

Baseline plasma concentrations and plasma concentrations on day 4 and day 8 for IMHP and HP are shown in Fig. 2 for the medical and non-medical patients. There were no statistical significant differences in baseline concentrations between IMHP and HP groups. In medical and non-medical patients, the changes in plasma levels
from baseline to day 4 and from baseline to day 8 were statistically significant larger in IMHP-treated patients compared with HP patients for (epa + dha)/lcf ratio, vitamin e and vitamin c, while plasma glutamine change from baseline to day 4 was statistically significant in the medical patients (Table 3).

The associations between changes in plasma concentrations from baseline to day 4 and day 8 and 6-month mortality are shown in Table 4. There is a statistically significant positive association of changes in (epa + dha)/lcf ratios from baseline to day 4 with 6-month mortality in the medical patients, and a statistically significant negative association of changes in zinc from baseline to day 8 with 6-month mortality in the non-medical patients.

The change in (epa + dha)/lcf ratios from baseline to day 4 is a potential candidate for mediator analysis as for this parameter IMHP versus HP conferred a statistically significant increase in plasma concentrations (Table 3; Fig. 1 association path A) and a statistically significant association between plasma concentration changes and 6-month mortality (Table 4; Fig. 1 association path B). Mediator analysis showed that a change in (epa + dha)/lcf ratio from baseline to day 4 was found to be a mediator for the treatment effect of IMHP versus HP concerning 6-month mortality, as including change in (epa + dha)/lcf ratio from baseline to day 4 into the statistical model decreased the coefficient of the statistical treatment effect on 6-month mortality (coefficient 0.823; P value: 0.016) to a non-statistical significant treatment effect (coefficient 0.618; P value: 0.291, Fig. 1).

Evaluation of the presence of A and B association paths in model II Fig. 1 was based on the criterion P value < 0.05. Relaxing this criterion to a P value < 0.15 did result in more parameters being investigated as possible mediators. However, none of those showed any suggestion for a mediating effect (data not shown).

Additional analysis on the association between the mediator (epa + dha)/lcf ratio and 6-month mortality showed that patients with an increase (≥0) in (epa + dha)/lcf plasma level ratios compared with those with a decrease (<0) from baseline to day 4 demonstrated higher 6-month mortality risk (P = 0.007, HR = 2.8) (Table 5).

### Discussion

Based on this post hoc analysis of the MetaPlus prospective randomized double-blind multicenter trial in which a high-protein enteral nutrition enriched with immune-modulating nutrients was compared with a standard isocaloric isonitrogenous high-protein enteral nutrition in a heterogeneous mechanically ventilated ICU population, we hypothesize that the harmful treatment effect of IMHP versus HP on 6-month mortality is specific for medical patients and that this harmful effect is mediated by an acute increase from baseline to day 4 in plasma (epa + dha)/lcf plasma level ratios compared with those with a decrease from baseline to day 4 demonstrated higher 6-month mortality risk (P = 0.007, HR = 2.8) (Table 5).

### Table 1 Multivariate Cox proportional hazard regression analysis on treatment effects among medical patients (a) and non-medical patients (b)

| Parameter | Parameter estimate | Standard error | Hazard ratio | 95% CI of the hazard ratio | P value |
|-----------|--------------------|----------------|--------------|--------------------------|---------|
| IMHP versus HP | 0.922 | 0.319 | 2.515 | [1.360, 4.783] | 0.004 |
| Age group 1 versus 4 | −2.651 | 0.788 | 0.071 | [0.011, 0.275] | <0.001 |
| Age group 2 versus 4 | −1.439 | 0.393 | 0.237 | [0.110, 0.522] | <0.001 |
| Age group 3 versus 4 | −1.120 | 0.409 | 0.326 | [0.145, 0.733] | 0.006 |
| APACHE-II score | 0.017 | 0.019 | 1.017 | [0.980, 1.057] | 0.376 |
| IMHP versus HP | −0.034 | 0.296 | 0.967 | [0.540, 1.736] | 0.909 |
| Age group 1 versus 4 | −1.595 | 0.533 | 0.203 | [0.068, 0.562] | 0.003 |
| Age group 2 versus 4 | −1.489 | 0.434 | 0.226 | [0.095, 0.531] | <0.001 |
| Age group 3 versus 4 | −0.467 | 0.381 | 0.627 | [0.299, 1.353] | 0.219 |
| APACHE-II score | 0.101 | 0.026 | 1.106 | [1.052, 1.165] | <0.001 |

IMHP immune-modulating high-protein enteral nutrition, HP high-protein enteral nutrition, age group 1 age ≤50 years, age group 2 age 51–70 years, age group 3 age 71–80 years, age group 4 age >80 years, APACHE-II acute physiology and chronic health evaluation-II
Table 2  Associations of baseline plasma concentrations of immune-modulating ingredients and 6-month mortality in all, medical and non-medical critically ill patients

| Immunonutrient (unit) | Univariate analysis | Multivariate analysis |
|-----------------------|---------------------|----------------------|
|                       | Coef. | SE  | Hazard ratio | 95% CI of the hazard ratio | P value | Coef. | SE  | Hazard ratio | 95% CI of the hazard ratio | P value |
| 2a: All patients (n = 301) |       |     |              |                            |         |       |     |              |                            |         |
| Glutamine (μmol/L)    | 0.00119 | 0.00059 | 1.001 | [1.000, 1.002] | 0.046 | 0.00034 | 0.00065 | 1.000 | [0.999, 1.001] | 0.599 |
| (epa + dha)/lcf ratio (x 10⁻²) | 0.21174 | 0.09876 | 1.236 | [1.012, 1.491] | 0.032 | -0.02082 | 0.10415 | 0.979 | [0.794, 1.195] | 0.842 |
| Selenium (μmol/L)     | 0.06623 | 0.16564 | 1.068 | [0.737, 1.416] | 0.689 | 0.11961 | 0.14465 | 1.127 | [0.814, 1.448] | 0.408 |
| Vit E (μmol/L)        | -0.00416 | 0.01402 | 0.996 | [0.968, 1.023] | 0.766 | -0.00750 | 0.01458 | 0.993 | [0.964, 1.021] | 0.607 |
| Vit C (μmol/L)        | 0.00297 | 0.00754 | 1.003 | [0.986, 1.015] | 0.694 | -0.00507 | 0.00817 | 0.995 | [0.977, 1.009] | 0.535 |
| Zinc (μmol/L)         | 0.01189 | 0.02532 | 1.012 | [0.961, 1.061] | 0.639 | 0.02327 | 0.02437 | 1.024 | [0.974, 1.071] | 0.340 |
| 2b: Medical patients (n = 109) |       |     |              |                            |         |       |     |              |                            |         |
| Glutamine (μmol/L)    | -0.00011 | 0.00090 | 1.000 | [0.998, 1.002] | 0.901 | -0.00025 | 0.00095 | 1.000 | [0.998, 1.002] | 0.792 |
| (epa + dha)/lcf ratio (x 10⁻²) | 0.16155 | 0.12798 | 1.175 | [0.905, 1.496] | 0.207 | -0.04175 | 0.13716 | 0.959 | [0.727, 1.246] | 0.761 |
| Selenium (μmol/L)     | 0.06647 | 0.17729 | 1.069 | [0.689, 1.420] | 0.708 | 0.06778 | 0.18363 | 1.070 | [0.685, 1.457] | 0.712 |
| Vit E (μmol/L)        | -0.00370 | 0.01981 | 0.996 | [0.957, 1.035] | 0.852 | -0.00299 | 0.02107 | 0.997 | [0.955, 1.038] | 0.887 |
| Vit C (μmol/L)        | -0.00611 | 0.00966 | 0.994 | [0.972, 1.009] | 0.527 | -0.00773 | 0.00995 | 0.992 | [0.970, 1.008] | 0.437 |
| Zinc (μmol/L)         | 0.04600 | 0.02592 | 1.047 | [0.992, 1.098] | 0.076 | 0.06078 | 0.02729 | 1.063 | [1.004, 1.118] | 0.026 |
| 2c: Non-medical patients |       |     |              |                            |         |       |     |              |                            |         |
| Glutamine (μmol/L)    | 0.00169 | 0.00074 | 1.002 | [1.000, 1.003] | 0.023 | 0.00067 | 0.00076 | 1.001 | [0.999, 1.002] | 0.377 |
| (epa + dha)/lcf ratio (x 10⁻²) | 0.29408 | 0.14469 | 1.342 | [0.996, 1.757] | 0.042 | 0.03667 | 0.16425 | 1.037 | [0.740, 1.410] | 0.823 |
| Selenium (μmol/L)     | 0.06196 | 0.29028 | 1.064 | [0.570, 1.785] | 0.831 | 0.11013 | 0.27021 | 1.116 | [0.625, 1.816] | 0.684 |
| Vit E (μmol/L)        | -0.00153 | 0.02043 | 0.990 | [0.948, 1.027] | 0.606 | -0.01162 | 0.02052 | 0.988 | [0.949, 1.028] | 0.571 |
| Vit C (μmol/L)        | 0.00203 | 0.01849 | 1.020 | [0.982, 1.056] | 0.274 | 0.01667 | 0.01899 | 1.017 | [0.978, 1.054] | 0.380 |
| Zinc (μmol/L)         | -0.10003 | 0.04612 | 0.905 | [0.824, 0.988] | 0.030 | -0.06168 | 0.04475 | 0.940 | [0.857, 1.022] | 0.168 |

The multivariate statistical model includes the variables “age group” and “Apache-II score” as covariates. Coef. = coefficient. The coefficient is the Cox proportional hazard regression parameter estimate; a positive coefficient indicates a worse prognosis, and a negative coefficient indicates a protective effect of the variable on 6-month mortality. Coefficient values represent changes per unit of nutrient for the different immunonutrients and change per percentage for (epa + dha)/lcf ratio. SE is the parameter estimate standard regression. P values represent Chi-square statistic testing the null hypothesis that the estimate is zero.
Fig. 2 Boxplot figures representing the plasma concentration values and variations of the immune-modulating nutrients at baseline, day 4 and day 8 among medical and non-medical patients. a Glutamine. b (epa + dha)/lcf ratio. c Selenium. d Vitamin e. e Vitamin c. f Zinc. IMHP immune-modulating high-protein enteral nutrition, HP high-protein enteral nutrition. Boxplot interpretation: 0 or +: average value, −: median, rectangle bottom: quartile 1 cutpoint (25th percentile), rectangle upper: quartile 3 cutpoint (75th percentile). 0 or +: outliers more than 1.5 times inter quartile range above quartile 3 or below quartile 1, T: highest or lowest level not being an outlier.
Table 3 Baseline plasma concentrations of immune-modulating ingredients and changes from baseline on day 4 and 8 in medical and non-medical critically ill patients

| Immune-modulating nutrient concentrations | IMHP (n = 54) mean (SD) | HP (n = 55) mean (SD) | P value |
|------------------------------------------|-------------------------|-----------------------|---------|
| 3a: Medical patients (n = 109)           |                         |                       |         |
| Glutamine (μmol/L)                        |                         |                       |         |
| Baseline (BL)                             | 393 (155)               | 406 (168)             | 0.687   |
| Day 4—BL                                 | 102 (141)               | 36 (141)              | 0.017   |
| Day 8—BL                                 | 77 (148)                | 10 (168)              | 0.070   |
| (epa + dha)/lcf ratio ($\times 10^{-2}$)  |                         |                       |         |
| Baseline                                 | 2.5 (1.1)               | 2.5 (1.1)             | 0.948   |
| Day 4—BL                                 | 3.3 (2.1)               | −0.4 (0.5)            | <0.001  |
| Day 8—BL                                 | 5.3 (2.2)               | −0.7 (0.7)            | <0.001  |
| Selenium (μmol/L)                        |                         |                       |         |
| Baseline                                 | 0.84 (0.37)             | 1.08 (1.00)           | 0.113   |
| Day 4—BL                                 | 0.16 (0.33)             | −0.16 (1.08)          | 0.057   |
| Day 8—BL                                 | 0.21 (0.43)             | −0.08 (1.26)          | 0.200   |
| Vit E (μmol/L)                            |                         |                       |         |
| Baseline                                 | 20.3 (7.6)              | 22.1 (7.2)            | 0.208   |
| Day 4—BL                                 | 19.5 (7.9)              | 2.5 (6.3)             | <0.001  |
| Day 8—BL                                 | 29.1 (18.8)             | 5.7 (8.9)             | <0.001  |
| Vit C (μmol/L)                            |                         |                       |         |
| Baseline                                 | 11.1 (14.0)             | 15.6 (22.0)           | 0.245   |
| Day 4—BL                                 | 13.6 (17.5)             | −4.4 (21.9)           | <0.001  |
| Day 8—BL                                 | 19.7 (18.5)             | −5.6 (26.3)           | <0.001  |
| Zinc (μmol/L)                             |                         |                       |         |
| Baseline                                 | 8.09 (6.05)             | 7.06 (4.05)           | 0.321   |
| Day 4—BL                                 | 0.73 (4.17)             | 1.56 (3.56)           | <0.001  |
| Day 8—BL                                 | 2.09 (3.90)             | 3.66 (4.34)           | 0.119   |
| 3b: Non-medical patients (n = 192)        |                         |                       |         |
| Glutamine (μmol/L)                        |                         |                       |         |
| Baseline (BL)                             | 350 (163)               | 327 (104)             | 0.252   |
| Day 4—BL                                 | 55 (180)                | 27 (118)              | 0.206   |
| Day 8—BL                                 | 16 (191)                | 30 (137)              | 0.586   |
| (epa + dha)/lcf ratio ($\times 10^{-2}$)  |                         |                       |         |
| Baseline                                 | 2.5 (1.0)               | 2.7 (0.9)             | 0.226   |
| Day 4—BL                                 | 3.5 (2.2)               | −0.2 (0.6)            | <0.001  |
| Day 8—BL                                 | 5.0 (2.7)               | −0.4 (0.6)            | <0.001  |
| Selenium (μmol/L)                        |                         |                       |         |
| Baseline                                 | 0.99 (0.46)             | 0.99 (0.55)           | 0.928   |
| Day 4—BL                                 | 0.13 (0.59)             | 0.03 (0.56)           | 0.272   |
| Day 8—BL                                 | 0.17 (0.73)             | 0.15 (0.68)           | 0.900   |
| Vit E (μmol/L)                            |                         |                       |         |
| Baseline                                 | 19.6 (6.6)              | 20.5 (8.2)            | 0.399   |
| Day 4—BL                                 | 19.7 (13.9)             | 3.3 (5.8)             | <0.001  |
| Day 8—BL                                 | 26.7 (14.4)             | 6.6 (7.5)             | <0.001  |
| Vit C (μmol/L)                            |                         |                       |         |
| Baseline                                 | 9.7 (8.2)               | 9.8 (7.7)             | 0.904   |
| Day 4—BL                                 | 7.0 (13.2)              | −2.1 (6.3)            | <0.001  |
| Day 8—BL                                 | 11.6 (14.7)             | −0.2 (9.3)            | <0.001  |
| Zinc (μmol/L)                             |                         |                       |         |
Medical versus non-medical patients

The debate regarding safety of supplementation of immune-modulating nutrients has mainly emerged from negative results of the REDOXs [9] and OMEGA [10] trials with, respectively, 80 and 85% medical ICU patients, while in the SIGNET trial [8], showing no benefit nor harm of immune-modulating nutrients, only 25% of included patients were medical. Meta-analyses of results of immune-modulating nutrient intervention studies among surgical patients have not reported any harmful effects and even show positive effects [14, 15]. Our post hoc analyses suggest harmful treatment effects of IMHP on 6-month mortality among medical patients and not among non-medical patients, and are in line with these observations. Medical patients were older, had higher APACHE-II scores and more frequently suffered from sepsis and pulmonary diseases [7]. Previously, it was recommended not to use arginine in ICU patients [16] or patients with severe sepsis [17] due to possible vasodilatory effects from NO production in patients with circulatory shock. This mechanism might have played a role as glutamine serves as precursor for de novo arginine production through the citrulline–arginine pathway [18].

Consequences for the glutamine debate

Our results show positive associations between higher baseline plasma glutamine and increased 6-month mortality in all patients and in non-medical patients. These findings contradict the hypothesis that glutamine is a conditional essential amino acid during critical illness [5].

This hypothesis is mainly based on one study from 2001 showing that ICU patients with plasma glutamine levels below 420 µmol/L show increased hospital mortality compared with patients with higher levels (420 µmol/L) [19]. However, mean age in the low glutamine group (74 years) was statistically significant different compared to the high glutamine group (63 years). Age was not included in the hospital mortality logistic regression analysis as separate explanatory factor. Including age potentially could have changed the strength and direction of associations, as in the MetaPlus study we showed that age is a strong independent mortality predictor [7] and our present analysis demonstrates that including age group covariates markedly influences associations between baseline levels and 6-month mortality. Recently, it was shown that not only low ICU admission plasma glutamine levels (<420 µmol/L), but also high plasma glutamine levels (>930 µmol/L) were associated with increased 6-month mortality, suggesting a U-shaped relation between plasma glutamine and 6-month mortality [20]. Furthermore, in septic patients, non-survivors had statistically significant higher median plasma glutamine levels (648 µmol/L) compared to survivors (460 µmol/L) [21]. In the Meta-Plus study, only 2 patients had plasma levels >930 µmol/L and 95% confidence intervals of baseline plasma glutamine levels were 339–391 µmol/L (IMHP group) and 334–378 µmol/L (HP group)[7], suggesting the univariate positive association between baseline glutamine concentration and 6-month mortality is not influenced markedly by very high baseline concentrations.

The REDOXs trial showed increased mortality with glutamine supplementation and raised questions whether critical illness is associated with low plasma glutamine levels at all [2] and whether interfering with glutamine supplementation could be deleterious [3].

Our post hoc analysis does not show associations between changes in plasma glutamine and 6-month mortality. Moreover, changes in plasma glutamine concentrations were not found to be a mediator of harmful treatment effects of IMHP versus HP in medical patients. Therefore, our post hoc analyses do not support the hypothesis that interfering with glutamine supplementation in medical critically ill patients is deleterious. However, this should be interpreted with caution and it may not be concluded that glutamine is not harmful as the enteral dose of glutamine supplementation and the effect of IMHP on glutamine levels were very modest. Furthermore, plasma glutamine levels, independent of the treatment allocation, were very comparable in both groups as seen in Fig. 2.

Consequences for the epa and dha debate

This post hoc analysis is the first to suggest that a change from baseline to day 4 in (epa + dha)/lcf plasma level ratios has a positive association with 6-month mortality risk. Average plasma (epa + dha)/lcf ratios among
Table 4 Associations of changes in plasma concentrations from baseline to day 4 and day 8 with 6-month mortality in medical and non-medical critically ill patients

| Immunonutrient | Baseline to day 4 | Baseline to day 8 |
|----------------|-------------------|-------------------|
|                | Coef.  | SE     | Hazard ratio | 95% CI of the hazard ratio |   P value | Coef. | SE     | Hazard ratio | 95% CI of the hazard ratio |   P value |
| 4a: Medical patients |        |        |             |                           |           |        |        |             |                           |           |
| Glutamine (μmol/L) | −0.002 | 0.001 | 0.998 | [0.996, 1.000] | 0.111 | −0.001 | 0.001 | 0.999 | [0.996, 1.001] | 0.302 |
| (epa + dha)/lcf ratio (×10⁻²) | 0.162 | 0.070 | 1.176 | [1.023, 1.348] | 0.021 | 0.055 | 0.053 | 1.057 | [0.949, 1.170] | 0.294 |
| Selenium (μmol/L) | 0.487 | 0.457 | 1.176 | [0.644, 3.892] | 0.286 | −0.551 | 0.615 | 0.576 | [0.159, 1.776] | 0.370 |
| Vit E (μmol/L) | −0.005 | 0.015 | 0.995 | [0.964, 1.024] | 0.758 | 0.009 | 0.012 | 1.009 | [0.985, 1.031] | 0.446 |
| Vit C (μmol/L) | −0.006 | 0.011 | 0.994 | [0.971, 1.016] | 0.614 | −0.001 | 0.011 | 0.999 | [0.976, 1.020] | 0.944 |
| Zinc (μmol/L) | −0.013 | 0.049 | 0.988 | [0.890, 1.080] | 0.799 | −0.093 | 0.064 | 0.912 | [0.794, 1.020] | 0.145 |
| 4b: Non-medical patients |        |        |             |                           |           |        |        |             |                           |           |
| Glutamine (μmol/L) | 0.001  | 0.001 | 1.001 | [0.999, 1.003] | 0.439 | 0.002  | 0.001 | 1.002 | [0.999, 1.004] | 0.180 |
| (epa + dha)/lcf ratio (×10⁻²) | −0.015 | 0.057 | 0.985 | [0.876, 1.095] | 0.793 | −0.040 | 0.050 | 0.961 | [0.866, 1.057] | 0.431 |
| Selenium (μmol/L) | 0.353  | 0.359 | 1.424 | [0.675, 2.807] | 0.324 | −0.530 | 0.440 | 0.589 | [0.238, 1.326] | 0.228 |
| Vit E (μmol/L) | −0.010  | 0.013 | 0.990 | [0.965, 1.014] | 0.429 | −0.005 | 0.012 | 0.995 | [0.971, 1.018] | 0.667 |
| Vit C (μmol/L) | −0.005  | 0.014 | 0.995 | [0.968, 1.021] | 0.700 | −0.009 | 0.013 | 0.991 | [0.966, 1.016] | 0.499 |
| Zinc (μmol/L) | −0.028  | 0.053 | 0.973 | [0.873, 1.074] | 0.600 | −0.102 | 0.051 | 0.903 | [0.812, 0.991] | 0.045 |

The coefficient is the Cox proportional hazard regression parameter estimate; a positive coefficient indicates a worse prognosis and a negative coefficient indicates a protective effect of the variable on 6-month mortality. Coefficient values represent changes per unit of nutrient for the different immunonutrients and change per percentage for (epa + dha)/lcf ratio. SE is the parameter estimate standard regression. CI confidence interval. P values represent Chi-square statistic testing the null hypothesis that the estimate is zero.
healthy persons across 16 European regions vary from 3.4 to 8.9%, due to differences in food intake [22]. In our study, average ICU admission values were below this range (2.5%), while after supplementation with IMHP for 4 or 8 days, averages were within these ranges. Among HP supplemented patients receiving no epa or dha, averages remained low, or even slightly decreased, suggesting a fast response of plasma status on epa and dha supplementation in IMHP patients. In the OMEGA study among acute lung injury ICU patients, Rice showed fast increases in plasma status within 3 days in epa, dha and gamma-linolenic-supplemented patients [10]. Rice concluded that fish oil supplementation might be harmful as illustrated by higher risk of 60-day mortality. Critics of the OMEGA trial suggested that twice daily bolus administration and higher protein intake in control patients confounded results [4]. However, our present results suggest that increased 6-month mortality in the IMHP group versus the HP group with similar amounts of protein was mediated through increases in (epa + dha)/lcf ratios from baseline to day 4 during continuous feeding. Therefore, without the perceived limitations of the OMEGA trial, we now report similar hazardous effects of omega-3 fatty acids supplementation among medical critically ill patients. Consequently, our findings do not support suggested clinical benefits of omega-3 fatty acids in enteral nutrition to preserve immune function and prevent aspects of the inflammatory response [23], as we did not observe any benefits on infectious morbidity [7]. As we did not measure biomarkers of immune function or inflammation, we cannot rule out that immune stimulation or anti-inflammatory effects have occurred. However, in the OMEGA trial no reductions in levels of inflammatory biomarkers despite marked increases in plasma omega-3 fatty acids were observed. The mechanism why increases in plasma omega-3 fatty acids are associated with increased mortality in medical critically ill patients remains unclear. We speculate that due to the suggested anti-inflammatory effect of fish oil IMHP not only reduced the systemic inflammatory response but also enhanced the compensatory anti-inflammatory response syndrome (CARS). It has become apparent that CARS is not simply a consecutive response to SIRS and hyperinflammation, but that both responses may occur simultaneously in the early phase after ICU admission. Possibly, we have induced the so-called persistent inflammatory immuno-suppressed catabolic syndrome (PICS) in these patients [24]. Another speculation is based on interesting data on omega-3 fatty acids supplementation during exercise in healthy volunteers showing reduced maximal power output by 10% and maximal heart rate by 6% within 1–3 days of supplementation [25]. These negative cardiovascular and metabolic effects of omega-3 fatty acids supplementation during exercise probably are mediated by other mechanisms than omega-3 fatty acids incorporation into plasma membranes [25].

**Consequences for the antioxidant debate**

Recent guidelines on nutritional support for critically ill patients by the Society of Critical Care Medicine and American Society of Parenteral and Enteral Nutrition indicate very low evidence for routine supplementation

| Table 5 Associations between changes in (epa + dha)/lcf ratio from baseline to day 4 and 6-month mortality in medical critically ill patients |
|---------------------------------------------------------------|
| (epa + dha)/lcf ratio parameters | 6-months mortality | Cox proportional hazard model analysis |
| ---------------------------------|-------------------|--------------------------------------|
| In incidence (%) | Coef. | SE | Hazard ratio | 95% CI of the hazard ratio | P value |
| (epa + dha)/lcf ratio (x 10^{-2}), continuous | 0.162 | 0.070 | 1.176 | [1.023, 1.348] | 0.021 |
| (epa + dha)/lcf ratio (x 10^{-2}), recoded to quartiles | | | | | |
| Q1 cutpoint: <-0.44 | 33 | Q1 versus Q4 | -0.667 | 0.486 | 0.513 | [0.191, 1.324] | 0.170 |
| Q2 cutpoints: >=-0.44 to <0.11 | 35 | Q2 versus Q4 | -0.960 | 0.506 | 0.383 | [0.137, 1.023] | 0.058 |
| Q3 cutpoints: >=0.11 to <3.19 | 48 | Q3 versus Q4 | 0.076 | 0.447 | 1.079 | [0.447, 2.627] | 0.865 |
| Q4 cutpoint: >=3.19 | 46 | Overall | | | | 0.105 |
| (epa + dha)/lcf ratio (x 10^{-2}), recoded to >=0 versus <0 | | | | | |
| <0 | 34 | >= 0 versus <0 | 1.030 | 0.383 | 2.800 | [1.344, 6.098] | 0.007 |
| >=0 | 46 | | | | |

*Coef coefficient. The coefficient is the Cox proportional hazard regression parameter estimate; a positive coefficient indicates a worse prognosis, and a negative coefficient indicates a protective effect of the variable on 6-month mortality. Coefficient values represent changes per percentage for (epa + dha)/lcf ratio. SE is the parameter estimate standard regression. P values represent Chi-square statistic testing the null hypothesis that the estimate is zero.*
of antioxidants summarizing studies until 2013 [26]. In these guidelines, it is indicated that antioxidant and trace element supplementation is associated with significant reductions in overall mortality. Recent REDOXs and Signet trials did not show any benefit or harm from antioxidant supplementation, but a recent retrospective study on selenium supplementation in postoperative ICU patients with sepsis showed increased mortality in univariate analysis [11]. Our post hoc analyses show positive associations with baseline zinc concentrations and 6-month mortality in medical patients. However, there was no effect of IMHP versus HP treatment on plasma zinc levels and no association between increase in plasma zinc concentrations and 6-month mortality, with similar results for selenium, vitamin e and vitamin c. Therefore, we hypothesize that harmful effects of IMHP versus HP treatment is not due to these antioxidants.

To the best of our knowledge, the MetaPlus study provides the largest database to study effects of nutritional treatment on plasma concentrations of immune-modulating nutrients in a heterogeneous mechanically ventilated ICU population. Although our data suggest mediator effects of changes in plasma (epa + dha)/lcf ratio, we must consider that findings are based on results from unplanned post hoc analyses. Furthermore, the relationship could be associative without direct causality if an underlying unknown confounding factor is involved. The intervention was a cocktail of various immune-modulating nutrients and may confer positive or negative effects on mortality, with or without interaction. Furthermore, post hoc analyses only generate hypotheses and therefore preclude firm recommendations.

Conclusions
We hypothesize that the harmful effect of immune-modulating high-protein enteral nutrition compared to high-protein enteral nutrition in the MetaPlus trial studying a heterogeneous group of critically ill patients is limited to the medical critically ill patients and mediated by an early increase in (eicosapentaenoic acid + docosahexaenoic acid)/long-chain fatty acid plasma ratio, resulting in increased 6-month mortality.

Additional file

Additional file 1. Immune-modulating nutrient analysis methods in plasma.

Authors’ contributions
Dr Van Zanten had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis. Hofman and Van Zanten were involved in study concept and designing. Van Zanten acquired the data. Swinkels, Hofman and Van Zanten were involved in statistical analysis and interpretation of data. Hofman and Van Zanten drafted the manuscript. Hofman, Swinkels and Van Zanten critically revised the manuscript for important intellectual content. Hofman obtained funding. Hofman was involved in administrative, technical or material support. Van Zanten and Hofman supervised the study. All authors read and approved the final manuscript.

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Competing interests
All authors will complete and submit the ICMJE Form for Disclosure of Potential Conflict of Interest. ZH is employed by Nutricia Advanced Medical Nutrition, Nutricia Research, Utrecht, The Netherlands. SS is employed by Nutricia Advanced Medical Nutrition, Nutricia Research, Utrecht, The Netherlands. AVZ has received honoraria for advisory board meetings, lectures and travel expenses from Abbott, Baxter, Danone-Nutricia, Fresenius Kabi, Nestle and Novartis. Inclusion fees for patients in the MetaPlus trial from Nutricia were paid to the local ICU research foundation. The authors declare that they have no competing interests.

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Role of the sponsor
The study sponsor contributed to the design of the study, the analysis and the interpretation of the study data and drafting of the manuscript. The sponsor funded the statistical analyses.

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