The Current Practice of Skin Testing for Antibiotics in Korean Hospitals

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Background/Aims: Antibiotic skin testing is a useful procedure for identifying patients with IgE-mediated hypersensitivity to antibiotics. The procedures, however, have not been standardized, and the testing is performed with diverse protocols in Korean hospitals wards. Thus, we examined the current practice of antibiotic skin testing in Korea.

Methods: We sent questionnaires to 12 allergists working in secondary or tertiary referral hospitals and collected them by e-mail or fax. The questionnaire included items such as the types and concentrations of the tested antibiotics, the methods of antibiotic skin testing, and the interpretation of the results.

Results: All hospitals responded to the questionnaire. The antibiotic skin testing protocols were variable, inconsistent, and differed with regard to the type and concentrations of antibiotics, the volume injected, and the interpretation of the results. Moreover, the protocols differed from the commonly recommended procedures in the medical literature.

Conclusions: Standardized guidelines for antibiotic skin testing are needed for the safe and effective use of antibiotics in Korea. (Korean J Intern Med 2010;25:207-212)

Keywords: Anti-bacterial agents; Drug hypersensitivity; Skin tests

INTRODUCTION

Adverse reactions to antibiotic drugs are common, and up to 25% of patients who are treated with antimicrobial agents report allergic reactions to antimicrobials [1]. Penicillin is the most frequently implicated agent; approximately 10% to 20% of hospitalized patients have a history of penicillin allergy [1,2]. As the use of semi-synthetic penicillins (amoxicillin and ampicillin) and cephalosporins has increased, these agents are becoming a major cause of allergic responses [3]. Generally, non-beta-lactam antibiotics such as vancomycin and fluoroquinolones are commonly used in patients with a history of beta-lactam allergy to avoid allergic reactions. The use of alternative antibiotics is associated with an increased number of infections caused by multidrug-resistant bacterial strains, which is associated with higher costs for care, morbidity, and mortality [4-7]. To solve this problem, identifying patients in the clinical setting who are at high risk for immediate IgE-mediated reactions is essential. Antibiotic skin testing (AST) is well-known to be a safe and reliable method for detecting immediate allergic reactions to beta-lactams. Although skin testing is a routine antibiotic test in Korean hospital wards, methods vary from hospital to hospital. The purpose of this study was to examine the current practice of AST in Korea.

Received: November 5, 2009
Accepted: December 14, 2009

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METHODS

We selected 12 hospitals that employed allergy specialists. To represent general practices in Korea, we tried to avoid deviations in grade and location of the hospitals, so three secondary hospitals (J to L, Table 1) and nine tertiary hospitals (A to I, Table 1) were enrolled. Among them, one secondary hospital (J, Table 1) and five tertiary hospitals (C to G, Table 1) were located in Seoul, and two secondary hospitals (K and L, Table 1) and four tertiary hospitals (A, B, H, and I, Table 1) were located in the province. Questionnaires were sent to the allergists, and the responses were collected either by e-mail or fax. The questionnaire included items such as the type and concentration of the antibiotics tested, the testing methods, and the interpretation of the test results. The questions were as follows: 1) Do you routinely conduct AST before using antibiotics in your wards? If so, how many and which antibiotics do you test? 2) Clarify the details of the methods for AST, e.g., testing method (skin prick testing vs. intradermal testing), concentrations and doses of antibiotics tested, test site, and whether a control substance is tested. 3) Clarify the method for AST interpretation, e.g., time lag between testing and interpretation and criteria for positivity.

RESULTS

The response rate was 100%. In all hospitals, the AST was a routine procedure before starting certain antibiotics in general wards and was usually conducted by a nurse without regard to allergic reaction history. However, the number of antibiotics tested varied from three to five. Among 12 hospitals, the ASTs for penicillins and cephalosporins were performed in 10 and 11 hospitals, respectively. In one hospital (E, Table 1), ASTs for cephalosporins were not routinely performed based on the idea that no study has demonstrated the clinical utility of cephalosporin skin testing for screening in subjects without a drug allergy history. Moreover, in two hospitals (H and I, Table 1), ASTs for penicillins were not performed because penicillins were never prescribed in these hospitals for fear of an adverse reaction. For other beta-lactam antibiotics, semisynthetic penicillins such as ampicillin and amoxicillin were tested in eight hospitals, carbapenems such as imipenem and meropenem in nine hospitals, and monobactams such as aztreonam in one hospital. Additionally, ASTs for vancomycin were conducted in four hospitals, and ASTs for aminoglycosides such as streptomycin were carried out in three hospitals. The details are shown in Table 1. Antibiotics other than those listed above were used without prior skin tests in all hospitals.

The ASTs were performed on the volar forearm using the intradermal method. However, skin tests using a control solution (e.g., saline) were not routinely conducted with the AST. Skin tests with saline were performed as a negative control in only two hospitals (F and G, Table 2),
following the AST and for suspicious, false-positive results cases. The injection amounts varied among the hospitals, ranging from 0.02 to 0.3 mL (Table 2). The concentrations tested also varied among hospitals (Table 1). For example, testing concentrations for penicillin G showed a 10,000 fold difference, ranging from 5 IU/mL to 50,000 IU/mL. However, the time lag (15 to 20 minutes) between testing and interpretation was similar in all hospitals. The criteria for positive results on the AST varied and were inconsistent among the hospitals: either a wheal or erythema larger than standard in six hospitals, only a wheal regardless of erythema in four hospitals, and only erythema in one hospital. Only one hospital considered both a wheal and erythema for interpreting the results. Additionally, the wheal and erythema standards for a positive response varied among the hospitals (Table 2).

**DISCUSSION**

We examined the current practice of AST in 12 Korean general hospitals with different grades and locations. We emphasize that what we examined in this study was the routine testing performed for screening in all patients in general wards and not what allergy specialists do. Unexpectedly, the AST protocols varied from hospital to hospital with regard to the types of antibiotics tested, testing concentrations and amounts, and the interpretation method. Therefore, discussing the similarities and differences between the current practices and the commonly recommended procedures in the medical literature is warranted.

**AST methods**

Some experts recommend that the general method for the AST is an intradermal test after confirming a negative skin prick test [8-10]. Intradermal testing is performed on the volar surface of the forearm with a 0.02 to 0.05 mL injection of the reagent solution, raising a 3 mm wheal, with saline as the negative control and with histamine as the positive control [8-10]. Similarly, all hospitals enrolled in the present study conducted AST using intradermal methods. Because the AST protocol in this study was the practice performed in wards, only intradermal testing was performed. However, intradermal testing was performed without any control, which makes the interpretation of the test results more difficult and increases the likelihood of a false-negative or false-positive result. Moreover, the volumes injected varied from one to ten times more than the volume recommended in the medical literature. Although the intradermal test is thought to be relatively safe, it may induce severe reactions, such as anaphylaxis, especially when a large volume is injected. Considering this, AST should be performed using the proper volume of antibiotics.

The antibiotic concentrations tested should not cause any significant irritation in subjects with good tolerance to antibiotics. Currently, the maximum concentrations used

| Hospitals | Injection amount, mL | Reading |
|-----------|----------------------|---------|
| A         | Make a bleb, 5 - 10 mm | 15 - 20 | W > 10 mm or E > 15 mm |
| B         | 0.2 - 0.3            | 15      | W > 5 mm |
| C         | 0.02                 | 15      | W > initial |
| D         | 0.02                 | 15 - 20 | W > 10 mm or E > 15 mm |
| E         | 0.02                 | 15 - 20 | W > 4 mm and E (+) |
| F         | 0.1                  | 15 - 20 | E > 10 mm |
| G         | Make a bleb, 3 mm    | 15 - 20 | W > 1.5 times |
| H         | 0.02                 | 15 - 20 | W > initial or E (+) |
| I         | Make a bleb, 3 mm    | 15      | W > initial |
| J          | 0.1                 | 15 - 20 | W > 10 mm or E > 15 mm |
| K          | 0.1                 | 15      | W > initial or E (+) |
| L          | 0.1                 | 15 - 20 | W > 10 mm or E > 10 mm |

W, wheal; E, erythema.

* Secondary hospitals; the others are tertiary hospitals.
differ from the accepted guidelines recommended by the American Academy of Allergy, Asthma and Immunology (AAAAI) / Joint Council of Allergy Asthma and Immunology (JCAAI) [8], the European Network for Drug Allergy (ENDA) / European Academy of Allergy, and the European Academy of Allergy and Clinical Immunology (EAACI) [9,10], as shown in Table 3. Notably, the testing concentrations in the hospitals we studied varied widely from one-thousandth less to ten times more than those recommended by published guidelines [8-10]. This is an important issue because testing with too low an antibiotic concentration can result in a false-negative, and too high a concentration can cause irritation and a false-positive result. Furthermore, high concentrations of antibiotics elevate the risk for anaphylaxis.

### AST interpretation

Interpretation is recommended 15 to 20 minutes after the injection [3,8-10]. For the intradermal test, the wheal area should be marked initially and again 15 to 20 minutes later, and an increase in the diameter greater than 3 mm is considered positive [3,8-10]. As mentioned above, we found that the criteria for a positive result varied too much and were inconsistent among the hospitals, although the time lag between test and interpretation was similar (15 to 20 minutes). These inconsistencies made a comparison of test results among hospitals difficult.

### Selection of antibiotics tested

AST is recommended as a routine practice for patients with suspected immediate-type allergy to penicillins (including semisynthetic penicillins) and cephalosporins but not to other antibiotics [11-16]. The proportion of patients labeled as allergic to penicillin who have a true IgE-mediated reaction to penicillin is very low [8,17,18]. Among these patients, over 80% have negative skin test results and no allergic reaction if challenged, whereas most patients with positive skin test results will react if challenged [19]. Penicillin skin testing has a high negative predictive value and is reliable for identifying individuals at risk for an immediate allergic reaction to penicillins [11,20,21]. Recent studies have shown that using the penicillin skin test helps determine an appropriate antibiotic treatment for patients with a history of penicillin allergy and reduces the use of broad-spectrum antibiotics, which may be a useful strategy for managing antibiotic resistance [11,19,22,23]. Many patients with a history of penicillin allergy and positive penicillin skin test results have a vague history of a prior reaction to penicillin [24]. In other words, the patient history may not be accurate, and penicillin skin testing should be considered for confirmation.

The increased use of beta-lactam antibiotics other than penicillin, such as amoxicillin, ampicillin, and cephalosporins, has increased the chance for allergic reactions to these antibiotics; thus, evaluating primary sensitization and cross-reactivities is important. The AST value for semisynthetic penicillins has been assessed extensively [10,25-27]. Semisynthetic penicillins may induce specific reactions and skin test positivity without cross-reacting to penicillins, which makes it necessary to include semisynthetic penicillins in ASTs. However, the negative predictive value of skin testing is unknown for cephalosporins [15,28]. Previous studies have reported a high negative predictive value when using parent cephalosporins at 2 mg/mL, but further study is needed for confirmation [16,29]. Moreover, cross-reactivity information between penicillins and cephalosporins is lacking. Considering a 4.4% rate of cross-reaction to cephalosporins in patients with a history of penicillin allergy and a positive result on a penicillin skin test [30], cephalosporin skin testing should be performed in patients with a history of penicillin allergy. The AST is not only a relatively simple and safe procedure but also a minimally required procedure to avoid medical / legal problems.

The usefulness of skin testing for carbapenems has not been determined. Moreover, the cross-reactivity between penicillins and carbapenems is unclear, as studies have reported conflicting findings [21,31-35]. Recently, Romano et al. [35] published two prospective studies on the utility of skin testing to detect immediate-type allergic reactions to imipenem-cilastatin and meropenem [36]. Because skin testing with an unmetabolized drug is

### Table 3. Beta-lactam concentrations recommended for skin testing

| Hapten                  | USA\(^a\) | Europe\(^b\) |
|-------------------------|-----------|--------------|
| Penicillin G, IU/mL     | 10,000    | 10,000       |
| PPL, mmol/L             | 6 × 10 \(^{-9}\) | 6 × 10 \(^{-6}\) |
| MDM, mmol/L             | 10 \(^{-2}\) | 2 × 10 \(^{-2}\) |
| Cephalosporin, mg/mL    | 2 - 3     | 2            |
| Semisynthetic penicillin\(^c\), mg/mL | - | 20 |

PPL, penicilloyl-polylysine; MDM, minor determinant mixture.

\(^a\)Adopted from reference [8].

\(^b\)Adopted from references [9,10].

\(^c\)Ampicillin, amoxicillin, and piperacillin.
unlikely to detect all sensitive individuals, patients with a negative reaction underwent a graded challenge, and no patients reacted. The authors concluded that this approach appeared safe, that the cross-reactivity was better defined, and that carbapenem reagents for skin testing are available. However, further confirmation is needed.

Many hospitals enrolled in the present study performed ASTs with antibiotics such as vancomycin, streptomycin, and monobactam that have not yet been validated. Up to five different antibiotics were identified in six hospitals (A, B, C, E, J, and K, Table 1). An AST should be performed to identify immediate-type allergic reactions to penicillins and cephalosporins, as discussed above, and not to other antibiotics in which the reliability of AST has not been validated.

This inconsistency in the AST procedure can result in a waste of time, money, labor, and the nation’s resources and can put patients at risk. Moreover, inconsistencies in the AST procedure make it difficult to conduct comparative studies on adverse antibiotic events among countries and hospitals. Of course, we should not accept other countries guidelines blindly but make our guidelines adequate for our circumstances. In conclusion, the practice of AST performed in hospital wards in Korea was too varied and inconsistent. The standardized guidelines for AST must be followed for the safe and effective use of antibiotics in Korea.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

Acknowledgements

We thank to Dr. Soo-Gul Lee, professor at Dong-A University Medical Center; Dr. Hae-Kyung Park, professor at Busan National University Hospital; Dr. Byung-Jae Lee, professor at Samsung Medical Center; Dr. Tae-Bum Kim, professor at the Asan Medical Center; Dr. Gyu-Young Hur, professor at the Korea University College of Medicine; Dr. Yong-Won Lee of the Yonsei University College of Medicine; Dr. In-Seon Choi, professor at Chonnam National University Hospital; Dr. Yong-Eun Kwon, professor at Chosun University Hospital; Dr. Sook-Hee Song of the Seoul Medical Center; Dr. Hee-Kyoo Kim, professor at Kook University College of Medicine; and Dr. Sang-Rok Lee of Cheongju St. Mary’s Hospital. This study was supported by a grant from the Korea Health 21 R&D Project, Ministry for Health, Welfare and Family Affairs, Korea (A030001).

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