Double primary malignant fibrous histiocytoma and squamous cell carcinoma of the larynx treated with laser laryngeal conservation surgery

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Abstract

Background: Synchronous multiple malignancies of the larynx are rare. We present a case here of synchronous primary laryngeal squamous cell carcinoma (SCC) and malignant fibrous histiocytoma (MFH) in a patient with hoarseness though with no history of exposure to radiation. Clinical, intraoperative, and histopathological findings in this patient are discussed.

Methods: Wide laser excision of the left supraglottic lesion and laser cordectomy of the right true vocal cord were performed.

Results: The patient presented with a recurrence of the MFH alone (with no recurrence of the SCC) two months after the first operation and was managed with an extended second look laser cordectomy. The patient is under regular follow-up and remained disease-free nine months from diagnosis.

Conclusions: Our results show that early-stage simultaneous tumours of the larynx and particularly MFH and SCC can be treated efficiently with endoscopic laryngeal surgery alone. Close follow-up is of paramount importance because of the aggressive nature of MFH.

Keywords: fibrous histiocytoma, squamous cell carcinoma, larynx, laser
Introduction

The first case of multiple primary tumours was reported in 1889 by Billroth [1]. Synchronous multiple primary malignancies of the larynx are very rare. Only very few simultaneous multiple malignancies of the larynx have been reported to date [2]. We report here a case of synchronous laryngeal SCC and primary MFH.

Case report

A 69-year-old man with a history of smoking and alcohol consumption was referred to otolaryngology complaining of hoarseness for almost one year. Full head and neck examination was performed including flexible laryngoscopy revealing a mass emanating from the left supraglottic part of larynx without erosion or haemorrhage and a lesion of the true right vocal cord presenting as leukoplakia (Figure 1). Bilateral vocal cord mobility was normal. Computerised Tomography (CT) of the neck and chest were normal without any lymph nodes or distant metastases. Furthermore laboratory tests, chest x-ray, and abdominal ultrasonography were also normal. There was no previous history of radiotherapy to the neck. Microlaryngoscopy and biopsy were performed (Figure 1). Histopathological analysis demonstrated a double malignancy in the left supraglottic lesion, i.e. a moderately differentiated SCC and MFH. Sections stained with haematoxylin and eosin from the mass of the left supraglottic area showed a biphasic neoplasm composed of two populations in close relation, which covered at least focally by stratified squamous epithelium, with mild to moderate dysplasia (Figures 2 and 3)

Figure 1. A mass emanating from the supraglottic part of larynx without erosion or haemorrhage and a lesion of the true right vocal cold presenting as leukoplakia.

Figure 2. Low-power view of the mass shows two neoplastic populations (sarcomatoid and epithelial component) and the overlying dysplastic epithelium (H&E, X40).
A B

Figure 3. (A) Low-power view demonstrates the neoplastic populations to intermingle (H&E, X100). (B) Low-power view shows the sarcomatoid component and the overlying epithelium (H&E, X100).

The prominent neoplastic population consisted mainly of plump spindle cells arranged in short fascicles in a storiform pattern. A portion of these cells were arranged in a diffuse growth pattern. In these areas, the neoplastic cells had hyperchromatic, irregular nuclei with prominent one or more nucleoli. Giant, bizarre, or multinucleated cells were commonly along with typical and atypical mitotic figures. Aggregates composed predominantly of lymphocytes and plasma cells and strands of collagen were seen in the stroma. The immunophenotype (Table 1) and the morphological features of this neoplastic population were compatible with a pleomorphic undifferentiated sarcoma/malignant fibrous histiocytoma (PUS/MFH).

The second neoplastic population consisted of epithelial component and corresponded to a moderately differentiated SCC. The neoplastic cells were arranged in solid masses or cords and had eosinophilic or clear cytoplasm and big irregular nuclei with prominent nucleoli. Intercellular bridges were present. The immunophenotype of this population was typical to that of any SCC (Table 1). The cells of the two populations intermingled (Figure 4).

A wide laser cordectomy of the left supraglottic mass and the right vocal cord with CO2 laser were performed. The results of the right true vocal cord biopsy were compatible with moderately differentiated SCC. During follow-up period, two months after the operation a very small granulomatous-type exophytic mass was found just above the anterior commissure (Figure 5). An extended second look laser cordectomy was performed and final histopathological analysis confirmed once again MFH. The patient is under regular follow-up and nine months later remained disease-free.

Discussion

SCC represents over 95% of all laryngeal malignant tumours. MFH is a non-epithelial tumour of mesenchymal origin that is composed of a variety of cell types, such as fibroblasts, atypical giant cells, and histiocyte-like cells. MFH is usually localised in the limbs, the abdomen, and the retroperitoneum. Almost 3–10% of all MFH are localised in the head and neck region. The most frequent localisations are sinonasal cavities (30%), craniofacial bones (15%), and larynx (10–15%) [3, 4]. MFH represents approximately 2% of all malignancies in the larynx. The number of cases found in the literature is limited [5].

The first case of laryngeal MFH was described in 1972. Laryngeal MFH is more prevalent in men than in women with a ratio of 3:1, and it usually appears in adults of average age of 43 with a wide age range [4]. In men MFH is found typically in glottis, whereas in women in the subglottis [3, 4]. The majority of cases with laryngeal MFH are associated with radiation [6]. However, in our patient there was no previous exposure to radiation. This suggests that the aetiology of MFH needs further analysis. Prognostic factors of MFH are tumour size, gender, age, histological grade, metastasis, and vascular invasion [2, 7]. The most common symptoms of laryngeal MFH are hoarseness, difficulty in swallowing, and dyspnea. Symptomatology depends on the size and location of the lesion. MFH presents in the larynx as a polyp or a nodule. Radiological findings characteristic of MFH do not exist. Although diagnosis is difficult with conventional histopathology, immunohistochemistry helps to discriminate MFH among the tumours of mesenchymal origin. Histological diagnosis includes proliferation of neoplastic cells that resemble fibroblast and histiocytes with abnormal atypias and mitosis. These are organised in bundles. The most common type is the storiform one which produces a collagen matrix.
Table 1. Results of immunohistochemical analysis.

| Antibodies          | Results |
|---------------------|---------|
|                     | SCC     | PUS/MFH |
| Cytokeratin AE1/AE3 | +       | --/+    |
| Cytokeratin 5/6     | +       | -       |
| Cytokeratin 34βE12  | +       | -       |
| p63                 | +       | --/+ (second biopsy) |
| EMA                 | -       | -       |
| Vimentin            | -       | +       |
| CD31                | -       | --/+    |
| CD34                | -       | -       |
| Factor VIII         | -       | -       |
| SMA                 | -       | Rare positive cells |
| Desmin              | -       | Rare positive cells |
| Sarcomeric actin    | -       | Rare positive cells |
| Myoglobin           | Not done| Not done|
| Neurofilaments      | -       | Rare positive cells |
| CD57                | -       | -       |
| S-100               | -       | -       |
| CD1a                | -       | -       |
| CD68                | -       | --/+    |
| Lysozyme            | -       | ++/-    |
| Fascin              | -       | +       |
| CD10                | -       | ++/-    |
| BCL-2               | -       | --/+    |
| ALK1                | -       | -       |
| CD99                | -       | +       |
| β-Catenin           | -       | -       |
| Ki67/MIB1           | 25%     | 90%     |

The immunohistochemical analysis in this case was crucial for the final diagnosis. In the case of the spindle/pleomorphic population the absence of any immunoreactivity for high molecular weight cytokeratin 34βE12 (which consist of cytokeratin 1, 5, 10, and 14) and cytokeratin 5/6, along with the at least focal positivity for markers of mesenchymal differentiation (vimentin, smooth muscle actin [SMA], desmin, sarcomeric actin, neurofilaments) excluded the diagnosis of a SCC. On the other hand, the absence of specific myogenic, vascular or neuromatous differentiation (only focal immunoreactivity for these markers) lead to the diagnosis of PUS/MFH.

Focal or weak immunoreactivity for mesenchymal markers such as vimentin [8, 9], SMA [10], CD10 [11, 12], CD99 [13], CD68 [14, 15], lysozyme [16, 17], fascin [18], and other intermediated filaments like desmin [10, 19, 20] and neurofilaments [9, 19] has already observed in cases of PUS/MFH. Additionally, cases of PUS/MFH positive for cytokeratin and EMA have been reported in the literature [9, 10, 19, 20, 21, 22, 23]. Positivity for protein p63 is extremely rare [24]. Although there are no previous reports for immunoreactivity for the antigen CD31 in PUS/MFH, however, CD31 positivity highlighting cutaneous histiocytomas and histiocytoma mimics [25] CD31 immunoreactivity. On the other hand, only sarcomatoid SCC shows positivity for markers such as vimentin, SMA, CD10, CD99, CD68, lysozyme, fascin, desmin, and neurofilaments [26, 27, 28, 29, 30].
Figure 4. Diffuse immunoreactivity of squamous cell carcinoma and the overlying epithelium for cytokeratin AE1/AE3 and focal/weak immunoreactivity of the sarcomatoid component (Immunoperoxidase with haematoxylin counterstain, X100).

Figure 5. Small supraglottic mass just above the anterior commissure two months after initial treatment was excised with CO2 laser and proved to be a recurrence of histiocytoma. The patient remained disease-free nine months after the second look cordectomy.

In 1889, the first case of multiple primary tumours was reported. Primary multiple malignant tumours of the larynx are extremely rare. On reviewing the literature we only found seven cases, and to our knowledge our case is the second report of synchronous laryngeal SCC and MFH [2]. The supraglottis is a rare location. Surgery is the treatment of choice for MFH, and it depends on location and size of the tumour, as happens with SCC. Radiotherapy or chemotherapy is not recommended as it does not offer any benefits to surgery. Wide removal with free clear margins is desirable. MFH is a very aggressive tumour [31]. The most common metastases are in the lung, liver, and spleen [32]. Close and frequent follow-ups are obligatory so that recurrences can be detected early and treated efficiently [31, 33]. In our case the extended supraglottic laser cordectomy which was performed, ensured a complication-free postoperative course and at nine-month follow-up the patient remained disease-free. In general, survival for MFH is approximately 60% after five years and 40% after ten years.

Conclusion

In conclusion synchronous multiple malignancies of the larynx are uncommon. We report here a case of a laryngeal SCC, moderately differentiated, with simultaneous primary MFH which was confirmed with histopathological and immunohistochemical examination. Our results show that early-stage of simultaneous tumours in the larynx and in particular the MFH and SCC can be removed via laser laryngeal conservation surgery. Close follow-up is mandatory in order to detect early recurrences or a distal metastasis, considering the fact that MFH is an aggressive tumour.
References

1. T Billroth Die allgemeine chirurgische pathologie und therapie 51 Vorlesungen In Reimer G (ed) Handbuch Fur Studirende und Arzte14th edition Berlin 908 1889

2. Cao X et al (2012) Simultaneous squamous cell carcinoma with primary malignant fibrous histiocytoma of the larynx: a case report Mol Med Rep 5(4) 971–3 PMID: 2307748 PMCID: 3493047

3. Barnes L and Ferlito A (1993) Soft tissue neoplasms En: Ferlito A (ed) Neo- plasms of the larynx Churchill Livingstone, Edinburgh 265–304

4. Barnes L and Kambour (1988) A Malignant fibrous histiocytoma of the head and neck. A report of 12 cases Arch Otolaryngol Head Neck Surg 114(10) 1149–56 DOI: 10.1001/archotol.1988.01860220083030 PMID: 2843204

5. Saleem M et al (1998) Fibrous histiocyto ma of the larynx J Laryngol Otol 112(12) 1205–7

6. Guney E et al (2002) Postirradiation malignant fibrous histiocytoma of the larynx: a case report Am J Otolaryngol 23(5) 293–6 DOI: 10.1053/ajot.2002.124554 PMID: 12239696

7. Ortiz Bish F et al (2004) [Malignant laryngeal fibrous histiocytoma (MLFH). Report of two unusual cases] Acta Otorrinolaringol Esp 55(8) 390–4 DOI: 10.1016/S0001-6519(04)78541-6 PMID: 15552216

8. Lawson CW, Fisher C and Gatter KC (1987) An immunohistochemical study of differentiation in malignant fibrous histiocytoma Histopathology 11(4) 375–83 DOI: 10.1111/j.1365-2559.1987.tb02642.x PMID: 2439432

9. Metten M and Soini Y (1989) Malignant fibrous histiocytoma. Heterogeneous patterns of intermediate filament proteins by immunohistochemistry Arch Pathol Lab Med 113(12) 1363–6 PMID: 2556088

10. Hasegawa T et al (2003) Expression of smooth muscle markers in so called malignant fibrous histiocytomas J Clin Pathol 56(9) 666–71 DOI: 10.1136/jcp.56.9.666 PMID: 12944549 PMCID: 1770045

11. Deniz K, Çoban G and Okten T (2012) Anti-CD10 (56C6) expression in soft tissue sarcomas Pathol Res Pract 208(5) 281–5 DOI: 10.1016/j.prp.2012.02.002 PMID: 22464152

12. Clarke LE et al (2010) CD10-positive myxofibrosarcomas: a pitfall in the differential diagnosis of atypical fibroxanthoma J Cutan Pathol 37(7) 737–43 DOI: 10.1111/j.1600-0560.2010.01532.x PMID: 20175824

13. Hartel PH et al (2006) CD99 immunoreactivity in atypical fibroxanthoma and pleomorphic malignant fibrous histiocytoma: a useful diagnostic marker J Cutan Pathol 33 Suppl 2 24–8 DOI: 10.1111/j.1600-0560.2006.00492.x PMID: 16972949

14. Guo H et al (2008) Reassessment of the pathological diagnosis in 33 cases of malignant fibrous histiocytoma Beijing Da Xue Xue Bao 40(4) 374–9 PMID: 18677383

15. Anghelina F et al (2009) Malignant fibrous histiocyto ma of larynx with giant cell: case report and histological-clinical considera- tions Rom J Morphol Embryol 50(3) 481–5 PMID: 19690778

16. Nakashishi S et al (1982) Immunohistochemical observation of intracytoplasmic lysozyme in proliferative and neoplastic fibro- histiocytic lesions Acta Pathol Jpn 32(6) 949–59 PMID: 6297236

17. Ribeiro RA et al (1988) Immunohistochemical identification of lysozyme and vimentin in an experimental malignant fibrous histiocyto ma Braz J Med Biol Res 21(5) 995–7 PMID: 2854748

18. Grogg KL et al (2005) A survey of clusterin and fascin expression in sarcomas and spindle cell neoplasms: strong clusterin immunostaining is highly specific for follicular dendritic cell tumor Mod Pathol 18(2) 260–6 DOI: 10.1038/modpathol.3800294
19. Lawson CW, Fisher C and Gatter KC (1987) An immunohistochemical study of differentiation in malignant fibrous histiocytoma Histopathology 11(4) 375–83 DOI: 10.1111/j.1365-2559.1987.tb02642.x PMID: 2439432

20. Litzky LA and Brooks JJ (1992) Cytokeratin immunoreactivity in malignant fibrous histiocytoma and spindle cell tumors: comparison between frozen and paraffin-embedded tissues Mod Pathol 5(1) 30–4 PMID: 1371872

21. Murakami I et al (1991) Recurrent malignant fibrous histiocytoma with expression of cytokeratin J Dermatol 18(6) 340–4 DOI: 10.1111/j.1346-8138.1991.tb03095.x PMID: 1719053

22. Rosenberg AE et al (1993) Expression of epithelial markers in malignant fibrous histiocytoma of the musculoskeletal system: an immunohistochemical and electron microscopic study Hum Pathol 24(3) 284–93 DOI: 10.1016/0046-8177(93)90039-J PMID: 7681032

23. Barak S, Wang Z and Miettinen M (2012) Immunoreactivity for calretinin and keratins in desmoid fibromatosis and other myofibroblastic tumors: a diagnostic pitfall Am J Surg Pathol 36(9) 1404–9 DOI: 10.1097/PAS.0b013e3182556def PMID: 22531174

24. Bishop JA, Montgomery EA and Westra WH (2014) Use of p40 and p63 immunohistochemistry and human papillomavirus testing as ancillary tools for the recognition of head and neck sarcomatoid carcinoma and its distinction from benign and malignant mesenchymal processes Am J Surg Pathol 38(2) 257–64 DOI: 10.1097/PAS.0000000000000119 PMID: 24418859 PMCID: 4313886

25. Vanchinathan V et al (2015) The vascular marker CD31 also highlights histiocytes and histiocyte-like cells within cutaneous tumors Am J Clin Pathol 143(2) 177–85 DOI: 10.1309/AJCPRHMM8CZH5EMFD PMID: 25596243

26. Monteagudo C et al (2002) CD99 immunoreactivity in atypical fibroxanthoma: a common feature of diagnostic value Am J Clin Pathol 117(1) 126–31 DOI: 10.1309/2EXB-70CW-3U6P-VQ6H PMID: 11789717

27. Kanner WA et al (2010) CD10, p63 and CD99 expression in the differential diagnosis of atypical fibroxanthoma, spindle cell squamous cell carcinoma and desmoplastic melanoma J Cutan Pathol 37(7) 744–50 DOI: 10.1111/j.1600-0560.2010.01534.x PMID: 20184665

28. Wieland CN et al (2011) The role of CD10 in distinguishing atypical fibroxanthoma from sarcomatoid (spindle cell) squamous cell carcinoma J Cutan Pathol 38(11) 884–8 DOI: 10.1111/j.1600-0560.2011.01768.x PMID: 21883368

29. Roy S, Purgina B and Seethala RR (2013) Spindle cell carcinoma of the larynx with rhabdomyoblastic heterologous element: a rare form of divergent differentiation Head Neck Pathol 7(3) 263–7 DOI: 10.1007/s12105-012-0402-3 PMCID: 3738762

30. Takata T et al (1991) Spindle cell squamous carcinoma of the oral region. An immunohistochemical and ultrastructural study on the histogenesis and differential diagnosis with a clinicopathological analysis of six cases Virchows Arch A Pathol Anat Histopathol 419(3) 177–82 DOI: 10.1007/BF01626345 PMID: 1718079

31. Weiss SW and Enzinger FM (1978) Malignant fibrous histiocytoma: an analysis of 200 cases Cancer 41(6) 2250–66 PMID: 207408

32. Ferlito A (1978) Histiocytic tumors of the larynx. A clinico-pathological study with review of the literature Cancer 42(2) 611–22 PMID: 209885

33. Resta L et al (2000) Malignant fibrous histiocytoma of the larynx after radiotherapy for squamous cell carcinoma Eur Arch Otorhinolaryngol 257(5) 260–2 DOI: 10.1007/s004050050235 PMID: 10923939