Influence of structural reinforcements on the twist-to-bend ratio of plant stems

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ABSTRACT

During biological evolution, plants have developed a wide variety of body plans and concepts that enable them to react or adapt to changing environmental conditions. Their morphological-anatomical and mechanical adaptations to conflicting conditions are especially interesting. A good example is the trade-off between flexural and torsional rigidity, as represented by the dimensionless twist-to-bend ratio. We have developed geometric models of a plant tissue reflecting the 2D situation of triangular cross-sections comprising of a parenchymatous matrix with vascular bundles surrounded by an epidermis and analysed them by using mathematical models (finite element analysis) to measure the effect of either reinforcements of the epidermal tissue or fibre reinforcements such as collenchyma and sclerenchyma on the twist-to-bend ratio. The change from an epidermis to a covering tissue of corky periderm increases both the flexural and the torsional rigidity and decreases the twist-to-bend ratio. Furthermore, additional fibre reinforcement strands in a parenchymatous ground tissue lead to a strong increase of the flexural and a weaker increase of the torsional rigidity and thus resulting in a marked increase of the twist-to-bend ratio. Within the developed model, a reinforcement by 49 sclerenchyma fibre strands or 24 collenchyma fibre strands is optimal in order to achieve high twist-to-bend ratios. Dependent on the mechanical quality of the fibres, the twist-to-bend ratio of collenchyma-reinforced axes is noticeably smaller, with collenchyma having an elastic modulus that is approximately 20 times smaller than that of sclerenchyma.

Keywords: flexural rigidity, torsional rigidity, Carex pendula, elastic modulus, torsional modulus, epidermis, periderm, collenchyma fibres, sclerenchyma fibres, vascular bundles, fossil plants, thin rods, finite element method

Introduction

Morphology, Anatomy and Biomechanics of Plants

During biological evolution, plants have developed a variety of tissues for which they had to invest different amounts of energy. The fibre reinforcements of the plant axes are especially costly, because of the thickness of their cell walls and/or the impregnation of their cell walls with lignin. Mechanical tests on whole biological plant axes can determine their overall properties, but not the individual in situ contribution of particular tissues such as fibres, vascular bundles, epidermis or parenchyma to the general mechanical performance of the whole axes. Within the framework of this interdisciplinary collaboration between scientists from the fields of plant biomechanics and applied mathematics, we have developed models and simulations that enable experiments in which the influence of in situ (fibre)reinforcements can be analysed. From the results of our experiments, in which either the geometrical arrangement or the mechanical properties of the individual tissues have been changed, we can draw conclusions about the in situ influence of individual tissues on the overall mechanical performance of the respective plant axis. For example, the number of fibre bundles can be increased or reduced, even tough the amount of material remains the same. Furthermore, in the simulation, lignified sclerenchyma fibres can be replaced by non-lignified collenchyma fibres, whereby the elastic modulus of the sclerenchyma fibres is one order of magnitude higher than that of the collenchyma fibres. In contrast, the non-lignified parenchyma, which often takes up the largest part of the plant cross-section, and the epidermis, a dermal tissue comprising a single layer of cells, have very low elastic moduli, which are on average...
one to three orders of magnitude smaller than those of the collenchyma fibres\textsuperscript{1–5}. The simulation also allows us to change a non-lignified single-layered epidermis into a corky multi-layered peridermal tissue, which is at least 10-times as stiff as the epidermis\textsuperscript{6} (see mechanical values in Tables 1 and 2).

With a focus on the topic of this publication, some straightforward comments can be given here with regard to functional morphology of plant axes including their morphology, anatomy and biomechanics. Flexural rigidity and torsional rigidity are composed variables that combine material properties and geometrical data: flexural rigidity ($EI$) is determined by the bending elastic modulus ($E$) and the axial second moment of area ($I$), whereas torsional rigidity ($GJ$) is determined by the torsional modulus ($G$) and the polar second moment of area ($J$). The bending and torsional moduli ($SI$ unit: Nm$^{-2}$ or Pascal) specify the stress-strain relationship as a measure for rigidity in the linear-elastic range, whereby mechanical stress is defined as the applied force per unit area and strain is understood as the displacement relating to a reference condition. The geometrical data, namely the axial and polar second moments of area ($SI$ unit: m$^4$), reflect the way in which the points of a cross-section are distributed in relation to a bending or torsional neutral plane or torsional axis, respectively\textsuperscript{7}. In this context, the dimensionless twist-to-bend ratio ($EI/GJ$) is particularly useful, as it provides information about the trade-off between rigidity in bending and in torsion. Bending and torsion are two different types of loading to which plants are exposed by their own weight and additional loads such as wind, flowers, fruits, snow, and animals sitting on the plants. A high twist-to-bend ratio reflects high bending rigidity combined with low torsional stiffness. High bending stiffness guarantees that leaves or flower stalks stand mostly upright, even when carrying large top loads (e.g. leaf blade, flowers and fruit). A high torsional flexibility allows streamlining in the wind, so that even plant parts with a large surface area (e.g. leaf blades) turn out of the wind, thereby reducing drag and enabling the plant to withstand higher wind loads without damage. During biological evolution, morphological-anatomical characteristics and biomechanical properties have been developed that, although not optimal for the individual type of loading, are optimised in terms of the sometimes conflicting demands of bending and torsion.

Plants differ in their so-called general body plan, which is a set of morphological features common to many members of a phylum\textsuperscript{8}. This includes, for example, the outer shape of their plant axes. Thus, the cross-sections of the petioles of the elephant ear (\textit{Caladium bicolor} (Aiton.) Vent.; hereafter \textit{C. bicolor}) are massive and round\textsuperscript{9}, the stems of the giant reed (\textit{Arundo donax} L.) are hollow and round\textsuperscript{10}, the branches of the prickly pear (\textit{Opuntia ficus-indica} (L.) Mill.; hereafter \textit{O. ficus-indica}) are elliptical\textsuperscript{11}, the flower stalks of the motherwort (\textit{Leonurus cardiaca} L.; hereafter \textit{L. cardiaca}) are square\textsuperscript{5} and the flower stalks of the drooping sedge (\textit{Carex pendula} Huds.; hereafter \textit{C. pendula}) are triangular\textsuperscript{12}. Looking at the ratio of the axial and polar second moments of area ($I/J$) of the various shapes, one generally finds values of 0.25 for elliptical cross-sections if the minor axis is twice the major axis, 0.2 to 0.5 for U-shaped cross-sections\textsuperscript{7}, 0.5 for circular cross-sections, 0.81 for squared cross-sections, 0.83 for isosceles triangles, and 1.25 for elliptical cross-sections if the major axis is twice the minor axis\textsuperscript{11}. Therefore, if the $I/J$ ratios are usually below 1, high twist-to-bend ratios ($EI/GJ$) must be attributed to high ratios of elastic and torsional modulus ($E/G$)\textsuperscript{11}. Since plants can be regarded as fibre-reinforced materials systems defined by a number of material properties, the body plan of plants also includes the three-dimensional arrangement of various tissues with their differing mechanical properties, which ultimately also determine the ratio of $E/G$. Additionally, natural structures are not only anatomically inhomogeneous and mechanically anisotropic, but also possess a spatial and temporal heterogeneity because of their growth and reaction capacity\textsuperscript{5}.

**Modelling of Biological Materials Systems**

Plants are multifunctional structures whose diverse functions are anchored to five hierarchical levels (molecule, organelle, cell, tissue, organ, plant). We focus here on the hierarchical level of tissues and the modelling of the various tissues in the cross-section of plant axes. A cornerstone of plant biomechanics is the performance of mechanical tests under a variety of applied loads such as tension, bending, compression and torsion, both in the linear–elastic range and up to ultimate strength, i.e. at failure. These mechanical data and the corresponding geometrical data on size, shape and tissue arrangement are necessary to develop analytical and numerical models. The models not only enable a deeper understanding of the functional principles of the plant model, but are also an indispensable precondition for the transfer of knowledge to technical developments, because models represent a common language for natural scientists, mathematicians, materials scientists and engineers\textsuperscript{12}. According to the biomimetic approaches of the “biology push process” or “bottom-up approach” and the “technology pull process” or “top-down approach”, every successful biomimetic product has to go through a step of abstraction\textsuperscript{13}.

Both analytical and numerical modelling have their intrinsic advantages and shortcomings when they are used to improve our understanding of the form-structure-function relationship of the biological models or to facilitate the transfer into biomimetic applications. Analytical models, for example, allow fast predictions about the influence of a variety of structural, i.e., morphological and/or anatomical, changes in a plant organ on its mechanical properties and also enable the inclusion of predictions about non-existing intermediate or extreme forms in the analysis. Similar to the requirements of a “closed” mathematical description, as desired in analytical models, the number of variables and the boundary conditions have to be
limited, and a careful simplification in the description of the biological model is typically necessary. The reduction to a few characterising parameters allow, on the one hand, a (much) faster analysis of the biological models. On the other hand, this procedure unfortunately includes dangers of oversimplification and of decisive parameters being overlooked. Numerical models (often) permit the precise description of form and structure of a biological model, but frequently need a plethora of parameters. The measurement of all of these parameters to the accuracy needed for numerical models is mostly extremely time-consuming (and sometimes even impossible). Moreover, numerical models are only of limited value for the inclusion and prediction of non-existing intermediate or extreme forms in the analysis. The latter is especially true as far as changes in form and structure are involved, whereas variations in mechanical parameters are typically easy to include in numerical models. Now that the advantages and shortcomings of analytical and numerical modelling have been pointed out, it becomes obvious that a combination of both approaches can be considered as the “royal road” of modelling, combining the advantages and avoiding the disadvantages of both approaches. In this work, we have therefore used simplified materials models and have made some geometrical assumptions to be able to focus on the essential parameters of the model. The resulting description is too complex to provide a closed form solution but, is simple enough that, by using a finite element approach, we can efficiently analyse a whole range of parameter values.

Aim of the Study

The aim of this project has been to find answers to the following scientific question: “To what extent do individual tissues such as fibres, vascular bundles, epidermis and parenchyma contribute to the flexural rigidity, the torsional rigidity and thus to the twist-to-bend ratio of a plant axis?”. Mathematical calculations have therefore been carried out, based on mechanical and geometrical data from the literature and from investigations of the triangular flower stalk of *C. pendula*. In this context, the effect of additional reinforcements on the flexural rigidity, the torsional rigidity and the twist-to-bend ratio have been examined with regard to: (i) the formation of a periderm instead of an epidermis, (ii) an increasing number of fibre strands up to an optimum and (iii) the replacement of sclerenchyma fibres by collenchyma fibres, the two fibre types differing notably in their elastic moduli.

Plant Data

Selected Model Plant

*C. pendula* is a suitable model plant from which the necessary geometrical and mechanical data for the mathematical calculations presented below have been collected, as in previous studies of the Plant Biomechanics Group Freiburg. Figure 1 presents a stained section and schematic drawing of its flower stalks showing the cross-sectional distribution of various tissues. The triangular cross-section shows an outer epidermis (e). Individual lignified sclerenchyma fibre strands (sc) in the periphery and vascular bundles (vb) scattered within the outer half of the cross-section are embedded in a non-lignified ground tissue consisting largely of parenchyma (pa) with interspaced smaller regions of aerenchyma (ae) close to the periphery and a layer of chlorenchyma (ch) directly underneath the epidermis.

Table 1 presents experimental data of the flower stalk of *C. pendula*. Mechanical properties from bending and torsional tests are provided, as are geometrical data of the entire internode and of individual tissues.

Mechanical Properties of Plant Tissues

Plant tissues differ in their anatomy, biomechanics and functions. Many of their properties are related to the presence of a cell wall and the large vacuole within the cell. The epidermis is the outermost tissue of the plant and consists of a single layer of cells covered by a cuticle. The epidermal tissue (including the cuticle) protects against water loss and regulates gas change. The periderm is a secondary covering composed of multiple layers containing cork cells. It sometimes replaces the epidermis and protects the stem from desiccation and pathogen attack. The parenchyma consists of living thin-walled cells with various functions. As a result of a high turgor pressure inside the vacuoles of these cells, the parenchyma can contribute to flexural and torsional rigidity and holds the specialised strengthening tissues in place. Chlorenchyma cells contain many chloroplasts for photosynthesis. The aerenchyma is a spongy tissue that allows gas exchange. Sclerenchyma fibres are dead cells characterised by thick and lignified cell walls. Collenchyma fibres are living cells with thick and non-lignified cell walls. The sclerenchyma and collenchyma belong to the strengthening tissues that provide load-bearing support for the plant and its organs. The vascular bundles are part of the transport system for water (xylem) and sugars (phloem). Tracheids and vessel elements of the xylem are dead and possess thick-walled and lignified cell walls, which also provide mechanical support.

Table 2 shows the elastic modulus of individual tissues such as the epidermis, parenchyma, sclerenchyma fibres, collenchyma fibres and vascular bundles. Elastic moduli were estimated for the respective tissues based on values from the literature and
Figure 1. Internodal cross-section of the biological model *Carex pendula*. (a) Schematic drawing highlighting various tissues. (b) Thin-section stained with acridine orange revealing lignified tissues in bright yellow-green. Abbreviations and colour code of the schematic drawing: ae: aerenchyma (white), ch: chlorenchyma (grey), e: epidermis (blue), pa: parenchyma (grey), sc: sclerenchyma fibre strands (black), vb: vascular bundles (red)

| data                        | *Carex pendula*                                      |
|-----------------------------|------------------------------------------------------|
| plant organ                 | internode of flower stalk                            |
| cross-sectional shape       | triangular                                           |
| flexural rigidity $EI$ (N-mm$^2$) | 141 873 ± 61 896                                  |
| bending elastic modulus $E$ (N/mm$^2$) | 16 132 ± 3305                                   |
| axial second moment of area $I$ (mm$^4$) | 9.03 ± 3.74                                      |
| torsional rigidity $GJ$ (N-mm$^2$) | 825.60 ± 412.98                                   |
| torsional modulus $G$ (N/mm$^2$) | 37.90 ± 11.87                                     |
| polar second moment of area $J$ (mm$^4$) | 22.09 ± 9.43 (see Remark 1)                        |
| twist-to-bend ratio $EI/GJ$ (/) | 192.42 ± 82.53                                     |
| 4-point bending test / number $n$ (/) | 15                                               |
| torsional test / number $n$ (/) | 15                                               |
| reinforcement fibres        | sclerenchyma                                        |
| total area of cross-section (mm$^2$) | 7.07 ± 1.34                                      |
| total area of sclerenchyma (mm$^2$) | 1.19 ± 0.21                                       |
| total area of collenchyma (mm$^2$) | –                                                 |
| total area of vascular tissues (mm$^2$) | 2.09 ± 0.41                                      |
| thickness of epidermis (µm) | 14.11 ± 4.04                                       |
| number of sclerenchyma strands (/) | 56.00 ± 3.87                                     |
| number of vascular bundles (/) | 77.67 ± 4.78                                      |
| references                  | basal internode no. 2 from May 7                    |

Table 1. Geometrical and mechanical data of the flower stalk of *Carex pendula*. Mean values ± one standard deviation of the mechanical and geometrical properties of the entire internodes and the geometrical data of individual tissues with reference to the cross-section are presented.
were additionally normalised in relation to the sclerenchyma having the highest value of these elastic moduli. The normalised values given in Table 2 are also included in the mathematical analyses of the influence of fibre reinforcement of plant axes on the twist-to-bend ratio presented below.

Remark 1. We are fully aware that the torsional second moment of area only for circular cross-sections equals the polar second moment of area: \( I_{\text{torsional}} = I_{\text{pol}} = 1.57 \cdot r^4 \), with \( r \) being the radius. For an isosceles triangle with side length \( s \), the torsional moment is calculated to be \( I_{\text{torsional}} = s^4/46.2 \), i.e. is, by a factor of 1.66, smaller than its polar second moment of area \( I_{\text{pol}} = s^4/27.7 \) as used in our calculations. Since the cross-sectional shape of the flower stalk of \( C. \text{pendula} \) has strongly rounded corners and slightly outwardly curved flanks and thus its torsional behaviour probably resembles more closely a circular cross-section than a triangle with sharp corners and straight flanks, we have decided to use the polar second moment of area in our calculation.

### Mathematical Models

In order to describe the influence of fibre reinforcement to the twist-to-bend ratio of a plant, we can consider a 2D-model determining the flexural and torsional rigidity of a beam using only its cross-section. The composite properties generally described in the introductory section, such as flexural rigidity \( (EI) \), torsional rigidity \( (GJ) \) and twist-to-bend ratio \((EI/GJ)\), are now defined mathematically in the following section and are given correspondingly more detailed abbreviations.

As in our previous work, in which we assumed a constant elastic modulus \( E \), we use methods from linear elasticity, which we repeat here for the readers convenience. As we are interested in investigating mechanical properties of the long and narrow flower stalk of \( C. \text{pendula} \) we describe a plant stem as a long thin elastic rod with domain \( B = \Omega \times (0, L) \) of length \( L \) and simply connected cross-section \( \Omega \) remaining constant along the longitudinal axis. We can now consider the domain \( \Omega \) as a composite of different materials bounded by a sufficiently regular boundary curve \( \partial \Omega \). Further, we assume \( L \gg \text{diam} \Omega \) and isotropy for the materials involved. Anisotropic effects, viscosity and other time-dependent processes are neglected here since we are only interested in the influence of the cross-sectional shape and the contained distribution of various materials on the mechanical properties of the stem. For our modelling, we consider \( B \) fixed at \( z = 0 \) and bending of \( B \) to be due to an outer normal force on \( \Omega \) at \( z = L \).

Mora & Müller have rigorously derived the flexural (or bending) rigidity by considering the limit of a very slender and long rod. Following classical theory, see Crandall et.al., flexural rigidity can be deduced from the moment curvature relation

\[
\begin{bmatrix}
M_x \\
M_y
\end{bmatrix} =
\begin{bmatrix}
D_x & D_{xy} \\
D_{xy} & D_y
\end{bmatrix}
\begin{bmatrix}
\kappa_x \\
\kappa_y
\end{bmatrix}
\]

(1)

where \( M_x, M_y \) denote the bending moments on the end of the beam, \( \kappa_x, \kappa_y \) denote the curvature in the direction of \( x \) and \( y \) respectively and the moments of inertia \( D_x, D_y \) and the product of inertia \( D_{xy} \) are given by

\[
D_x = \int_{\Omega} E(x,y) \hat{x}^2 \, dx \, dy, \quad D_y = \int_{\Omega} E(x,y) \hat{y}^2 \, dx \, dy, \quad D_{xy} = \int_{\Omega} E(x,y) \hat{x} \hat{y} \, dx \, dy,
\]

where we have

\[
\hat{y} = y - \frac{\int_{\Omega} E(x,y) y \, dx \, dy}{\int_{\Omega} E(x,y) \, dx \, dy}, \quad \hat{x} = x - \frac{\int_{\Omega} E(x,y) x \, dx \, dy}{\int_{\Omega} E(x,y) \, dx \, dy}.
\]
Because of the heterogeneity of $\Omega$ the elastic modulus $E(x,y)$ depends on the cross-sectional coordinates, being piece-wise constant. The maximal and minimal flexural rigidities $D_{\text{max}}$ and $D_{\text{min}}$ along the principal axes are then given by the maximal and minimal eigenvalue of the matrix in (1) leading to

$$D_{\text{max/min}} = \left( D_{\text{mean}} \pm \sqrt{\frac{(D_x - D_y)^2}{4} + D_{xy}^2} \right)$$

with $D_{\text{mean}} = \frac{D_x + D_y}{2}$. In the following we are concerned with the problem of generating cross-sections with high flexural rigidity. Therefore we will confine ourselves to the computation of the minimal flexural rigidity $D_{\text{min}}$. High values of $D_{\text{min}}$ then lead to high resistance against bending forces along all axes. As for the flexural rigidity, the torsional rigidity for an elastic slender rod with domain $B$ has been rigorously derived by Mora & Müller.

In order to describe the torsional rigidity, we use St.Venant’s theory of pure torsion of nonhomogeneous elastic beams, which has been used, among others, by Ecsedi. As torsion is only given by a moment at the top of $B$ the torsional rigidity can be expressed by Prandtl’s stress function $\phi(x,y)$ satisfying

$$\nabla \cdot \left( \frac{1}{G(x,y)} \nabla \phi \right) = -2, \text{ in } \Omega,$$

$$\phi = 0 \text{ on } \partial \Omega$$

with the shear modulus $G(x,y)$ depending on the cross-sectional coordinates, being piece-wise constant. By using the stress function $\phi$, the torsional rigidity is then given by

$$D_z = 2 \int_{\Omega} \phi \, dx \, dy.$$

For the numerical computation of the rigidities, we have employed a P1 triangular finite element discretisation of an equilateral triangular cross-section with approximately $5 \cdot 10^6$ triangular elements in order both to solve the equation in (2) numerically and finally to compute the rigidities. The implementation of this standard finite element method (C++-code) is available in the supplementary material.

**Effect of Reinforcements on the Twist-to-bend Ratio**

During their ontogeny, plants react to increasing bending forces triggered by continuous stem growth and the formation of top loads such as flowers, seeds and fruits. An increase of their resistance against bending forces can be achieved by an increase of their flexural rigidity along the principal axes, i.e. increasing $D_{\text{min}}$. For a better understanding of this effect, we introduce a simplified model that is related to $C. \text{pendula}$ and that measures flexural and torsional rigidity. As a first approach, we describe the tissue arrangement in the cross-section of $C. \text{pendula}$ in terms of a distribution of circles (vascular bundles) in an equilateral triangular reference domain $\Omega$ additionally filled with parenchyma and surrounded by an epidermis (see Fig. 2a). The contribution to the cross-sectional area of the vascular bundles ($\approx 29\%$) and the parenchyma ($\approx 52\%$) and the thickness of the epidermis is selected to match the distribution of mass in the total cross-sectional area of $C. \text{pendula}$.

For the computation of the rigidities, we normalise the elastic moduli ($E_{\text{est}}$) of all contained materials with respect to the elastic modulus of the sclerenchyma. This means that we set $E_{\text{norm}} = 1$ for the sclerenchyma and obtain the normalised elastic moduli of the other materials by scaling accordingly (see Table 1). The ratio between the elastic modulus of the mechanically decisive materials involved is denoted by $\mu$ in the following and is referred to as the ratio between material values.

We assume a constant Poisson’s ratio $\nu$ for the materials involved and compute the normalised torsional modulus $G_{\text{norm}}$ as

$$G_{\text{norm}} = \frac{E_{\text{norm}}}{2(1 + \nu)}$$

for a given elastic modulus $E_{\text{norm}}$. The assumption of a constant Poisson’s ratio is reasonable as the value range $\nu \in [0.2, 0.5]$ is typical for many plant axes and, thus, a change in $\nu$ among the materials is negligible for our model. In the following, we set $\nu = 0.35$, which is an appropriate choice for $C. \text{pendula}$, see Hejnowicz.
Figure 2. Initial configuration. Parenchyma and vascular bundles surrounded by (a) an epidermis ($E_{\text{norm}} = 0.00111$) and (b) a peridermal covering tissue ($E_{\text{norm}} = 0.011$), respectively. For a non-lignified epidermis, both the torsional and flexural rigidity are comparatively low. If a corky periderm is formed, the flexural rigidity is increased ($\approx 14\%$) and so is the torsional rigidity ($\approx 46\%$). This leads to a decrease in the twist-to-bend ratio ($\approx 24\%$). Colour code: epidermis non-lignified: blue, periderm: green, parenchyma: grey, vascular bundles: red.

Reinforcement without Fibres: from Epidermis to Periderm

In this section, we compare the effect of a single-layered epidermis and a multi-layered corky periderm on the twist-to-bend ratio of the triangular cross-section (see Fig. 2). In this configuration and with respect to the normalised elastic modulus of the epidermis ($E_{\text{norm}} = 0.00111$), the torsional rigidity ($D_z \approx 0.00013$) and the flexural rigidity ($D_{\text{min}} \approx 0.00118$) are comparatively low and so is the twist-to-bend ratio ($D_{\text{min}}D_z \approx 9.37$) (see Fig. 2a). The resistance against bending forces and, hence, the flexural rigidity increases by the formation of a ring of corky periderm ($E_{\text{norm}} = 0.011$). This formation of a closed ring of strengthening tissue, however, simultaneously increases the flexural rigidity ($D_{\text{min}} \approx 0.00134$) and the torsional rigidity ($D_z \approx 0.00019$) impeding the plant’s property of being able to twist easily and, moreover, decreasing its twist-to-bend ratio to ($D_{\text{min}}D_z \approx 7.05$).

Reinforcement by Numerous Sclerenchyma Fibre Strands

In addition to the above-mentioned reinforcement of the epidermal tissue, fibre reinforcement is extremely common in plant axes. As one can see in the biological model (Fig. 1), individual sclerenchyma strands in the periphery of the triangular cross-section of $C. \text{pendula}$ are striking. Therefore, we can additionally incorporate sclerenchyma fibre strands ($E_{\text{norm}} = 1$) into the parenchyma ($E_{\text{norm}} = 0.00044$) of the cross-section from Fig. 2a. Using this idea, we have developed a model to describe the effect of fibre reinforcement. The mechanically decisive materials for this experiment are the parenchyma and sclerenchyma having the ratio $\mu$ between their elastic moduli of $\mu = 0.00044$. We fix the proportion of the sclerenchyma ($\approx 17\%$; see Table 1) in the total cross-sectional area. Further, we consider a distribution of fibre bundles (sclerenchyma) around the inner boundary $\partial \Omega$ of $\Omega$ with circular cross-sections centred at a fixed distance to $\partial \Omega$. Starting with 6 fibre strands for each side of the triangle we refine the distribution, such that a higher number of fibre bundles is used in each step of the refinement, see Fig. 3. The proportion of the fibre bundles in the total cross-sectional area is fixed during the whole process. The arrangement and the total cross-sectional area of the fibre strands are now determined by the structure of the Carex ground tissue, see Fig. 1, and, hence, the number of fibre bundles is the only free variable in this model. The procedure is stopped before single fibre strands become connected, as such a closed ring of sclerenchyma would immediately (markedly) increase the torsional rigidity and thus decrease the twist-to-bend ratio.

For simplicity, circular shapes of the fibre bundles are taken, as circular cross-sections exhibit high torsional rigidity and, therefore, any other cross-sectional shape of the fibre strands is likely to amplify the effect of decreasing torsional rigidity while increasing the number of fibre bundles.

The numerical experiments show that, by increasing the number of fibre strands, the torsional rigidity $D_z$ initially decreases when the flexural rigidity remains nearly bounded and, thus, the twist-to-bend ratio increases (see Fig. 4). A regression analysis for torsional rigidity up to 51 fibre strands shows that $D_z$ is given as a function of the number of fibre bundles $N$, with

$$D_z(N) = \frac{a}{N} + C$$

for constants $a, C \in \mathbb{R}$. Therefore, increasing the number of fibre bundles initially decreases the torsional rigidity scaling as
Figure 3. Reinforcement by fibre strands. From (a) to (d), the number of reinforcing (sclerenchyma) fibre strands increases whereas their total cross-sectional area remains constant. If the number is too high, single fibre strands become connected (d), thereby changing the torsional rigidity drastically and, thus, the procedure is stopped before this occurs. Colour code: epidermis non-lignified: blue, parenchyma: grey, sclerenchyma fibres: black, vascular bundles: red.

\( N^{-1} \) in dependence on the number of fibre bundles with asymptote \( C \). Because of the boundedness of the flexural rigidity, the twist-to-bend ratio increases showing a similar asymptotic behaviour and scaling as \( N \) in dependence on the number of fibre strands. However, when the distance between the single fibre strands becomes very small, even when the fibre strands are not yet connected, see (a) in Fig. 6, the cycle-like structure of the fibre arrangement causes the torsional rigidity to increase again leading to a decrease in the twist-to-bend ratio. This occurs because the gradient of \( \phi \) in the space between the fibre strands increases as the distance between fibre strands decreases, similar to the behaviour of a Neumann sieve. Thus, the value of \( \phi \) in the inner part of the cross-section is raised, see (e) and (f) in Fig. 6 resulting in a substantial increase of the torsional rigidity, since this increase is integrated over a large domain.

As the proportion of the cross-sectional area filled with parenchyma is comparatively high, the effect becomes dominant as the distance of single fibre strands decreases and therefore leads to an increase in the torsional rigidity, see Fig 4a. Therefore, in the setting described above, torsional rigidity and the twist-to-bend ratio reach an optimum for 49 fibre strands, see Fig. 4a and c. This characterises the number of fibre bundles \( N \) as a design variable in an optimisation problem in order to maximise the twist-to-bend ratio and simultaneously minimise the torsional rigidity. The results of the experiments, see Fig. 4, then justify the conjecture of the existence of an optimal number of fibre bundles.

Reinforcement by Numerous Collenchyma Fibre Strands

As stated in the previous section, an optimal number of sclerenchyma strands achieves a high twist-to-bend ratio within the configuration of scattered vascular bundles embedded in a parenchymatous matrix that is surrounded by an epidermis (see Figure 3(a)). However, we can also reasonably investigate the dependency of the optimal number of fibre strands needed to increase the twist-to-bend ratio on the ratio \( \mu \) between the elastic moduli of the mechanically decisive materials involved.

To do so, we can incorporate collenchyma fibres \( (E_{\text{norm}} = 0.05555) \) instead of sclerenchyma fibres \( (E_{\text{norm}} = 1) \), whose elastic modulus is lower compared with the elastic modulus of sclerenchyma fibres, see Table 2. Since parenchyma \( (E_{\text{norm}} = 0.00044) \) and collenchyma are the decisive materials, this then leads to a ratio of material values given by \( \mu \approx 0.008 \). We can again carry out the experiment shown in Fig. 3 but with collenchyma fibres instead of sclerenchyma fibres and with the normalised elastic modulus \( E_{\text{norm}} = 0.05555 \) for the collenchyma fibres. The elastic moduli of the other materials involved (epidermis, parenchyma, vascular bundles) remain the same.

Again, we find that up to a certain number of collenchyma strands, the torsional rigidity decreases linearly, whereas the flexural rigidity remains nearly constant, see Fig. 5a and b. However, now that the ratio \( \mu \) is lower, the number of fibre strands that is optimal in order to decrease torsional rigidity and to increase the twist-to-bend ratio, is reduced noticeably to 24 and,—after a slight drop—the twist-to-bend ratio remains nearly constant up to 36. For more than 24 collenchyma strands, we can see the same effect as above, namely the torsional rigidity increases again and, hence, the twist-to-bend ratio decreases, see Fig. 5a and c.

This experiment in which the sclerenchyma fibres are replaced by collenchyma fibres illustrates the effect of the ratio \( \mu \) between the material values of the mechanically decisive materials on the twist-to-bend ratio and on the optimal number of fibre strands included in fibre reinforcement. As a preview to the discussion, we can conjecture that the ratio \( \mu \) plays a role in the sense that,
Figure 4. Reinforcement by sclerenchyma fibre strands. Trend of (a) torsional rigidity, (b) flexural rigidity and (c) twist-to-bend ratio. The ratio $\mu$ between the elastic moduli of the parenchyma and sclerenchyma is $\mu = 0.00044$. The sclerenchyma and parenchyma are the mechanically decisive materials for this experiment. By increasing the number of fibre strands used for reinforcement, torsional rigidity first decreases linearly, see (a), whereas the flexural rigidity remains nearly constant during the procedure, see (b). The twist-to-bend ratio first increases nearly linearly reaching a maximum for $N = 49$. For more than 49 fibre bundles, the distance of single fibre bundles is too small and the torsional rigidity is caused to increase again, whereas the twist-to-bend ratio is reduced by this effect, see (a) and (c). The numerically computed values of the torsional rigidity are further interpolated using a first order polynomial regression (orange line). For up to 51 fibre strands, the torsional rigidity behaves as $f(N) = \frac{a}{N} + C$, see (a), where $C = 2.1 \cdot 10^{-4}$ and $a = 0.0031$. The R-squared value for the polynomial regression is 0.99.

Discussion

When embryophytic plants colonised the land and lost the buoyancy of surrounding water around 470 million year ago during the mid-Ordovician, they faced entirely different mechanical constraints. These concerned, in particular, their anchorage (root system) and the mechanical loads on their upright aerial parts (stems with branches and leaves). Stems, roots and leaves had evolved as early as the mid Devonian (ca. 390 million years BP) and the first forests of tall trees existed by the late Devonian (ca. 370 million years BP)\textsuperscript{22, 23}. Since then, evolutionary processes have shaped stem and root form and internal structure enabling plants to cope with the various mechanical loads that act upon them in their diverse environments. A plethora of evolutionary adaptations can be observed in aerial stems, especially with regard to bending and torsional loading, which represent the predominant load cases in plants with self-supporting upright stems. In addition to experimental analyses, theoretical considerations including analytical and numerical simulations can help to decipher the complex interplay between form, structure and mechanical properties of the involved plant tissues\textsuperscript{16, 24–27}.

In this article, we present our finite element analysis involving a triangular cross-section consisting of parenchyma with scattered embedded vascular bundles surrounded by an epidermis (see Fig. 2a). In contrast to real experiments, computer experiments allow one particular variable to be altered and the resulting effect to be quantified precisely. By replacing the epidermis with a periderm or by including additional fibre strands with variable numbers and/or mechanical quality, we can quantify the mechanical effect of structural reinforcement on the flexural and torsional rigidity and thus the twist-to-bend ratio of plant axes (see Table 3). The biological model that we have used for our simulation is the isosceles-triangular cross-section of the flower stalk of \textit{C. pendula}. This long, drooping stem can attain 2.5 metres in length and can apically carry pendulous inflorescences. In summer, the inflorescences develop into a multiple fruit with many relatively heavy fruits.

From Epidermis to Periderm

The outermost single cell layer of plants is called the epidermis, the main function of which is to protect the plant. This primary covering with an elastic modulus of $E \approx 50$ MPa can support tensile loads. Woody stems can also produce a secondary
Figure 5. Reinforcement by hypothetically occurring collenchyma fibre strands. Trend of (a) torsional rigidity, (b) flexural rigidity and (c) twist-to-bend ratio. The ratio $\mu$ between the elastic moduli of the parenchyma and collenchyma is $\mu = 0.008$. The collenchyma and parenchyma are the mechanically decisive materials. By increasing the number of fibre strands used for reinforcement, the torsional rigidity first decreases linearly, see (a), whereas the flexural rigidity remains nearly constant during the procedure, see (b). The twist-to-bend ratio first increases nearly linearly reaching a maximum for $N = 24$. For more than 24 strands, the torsional rigidity is caused to increase again, whereas the twist-to-bend ratio is reduced by this effect, see (a) and (c).

| decisive materials         | $\mu$ | flexural rigidity | torsional rigidity | twist-to-bend ratio |
|---------------------------|-------|-------------------|--------------------|---------------------|
| parenchyma/epidermis      | 0.4   | reference value   | reference value    | reference value     |
| parenchyma/periderm       | 0.04  | increase $\approx 14\%$ | increase $\approx 46\%$ | decrease $\approx 24\%$ |
| parenchyma/sclerenchyma*  | 0.00044 | increase $\approx 5600\%$ | increase $\approx 112\%$ | increase $\approx 2536\%$ |
| parenchyma/collenchyma**  | 0.008 | increase $\approx 315\%$ | increase $\approx 34\%$ | increase $\approx 198\%$ |

Table 3. Summary of the simulation results. The mechanical effect of various changes of the structural reinforcement is given as a percentage increase or decrease with reference to the initial configuration (= reference value). Figure 2a shows the initial configuration in terms of a triangular cross-section comprising of parenchyma with embedded vascular bundles surrounded by an epidermis. $\mu$ is the ratio of estimated elastic moduli ($E_{est}$) of the decisive materials (see Table 2). The percentage values inherent to the fibre reinforcements are computed with respect to the optimal number of fibre strands. namely *49 sclerenchyma strands and **24 collenchyma strands (see Figs. 4 and 5).
(a) Gap between fibre strands with inscribed triangulation and a total of 51 circular fibre strands inside a triangular cross-section.

(b) Prandtl’s stress function $\phi$ in fibre-reinforced triangular cross-section with horizontal and vertical lines $L_1$ and $L_2$.

(c) Representation of Prandtl’s stress function $\phi$ along the horizontal line $L_1$ for a total of 36 fibre strands. The integral of $\phi$ over $L_1$ is $\int \phi \approx 5.0 \cdot 10^{-4}$.

(d) Representation of Prandtl’s stress function $\phi$ along the horizontal line $L_1$ for a total of 51 fibre strands. The integral of $\phi$ over $L_1$ is $\int \phi \approx 4.5 \cdot 10^{-4}$.

(e) Representation of Prandtl’s stress function $\phi$ along the vertical line $L_2$ for a total of 36 fibre strands. The dashed line shows the increase of $\phi$ between the two fibre strands.

(f) Representation of Prandtl’s stress function $\phi$ along the vertical line $L_2$ for a total of 51 fibre strands. The dashed line shows the increase of $\phi$ between the two fibre strands.

**Figure 6. Local representation of Prandtl’s stress function.** For different numbers of fibre strands, the stress function $\phi$ is plotted along horizontal and vertical lines, $L_1$ and $L_2$, in (b) and thereby two opposing effects are illustrated. By increasing the number of fibre strands but by fixing their total area, $\phi$ decreases within the fibre strands, see (e) and (d), but increases in the inner part of the cross-section, see (e) and (f). This because, as the distance between the fibre strands becomes smaller, the gradient of $\phi$ between two fibre strands is increased and, therefore, the value of $\phi$ in the inner part of the cross-section is raised. As the proportion of area filled with parenchyma is comparatively high, this effect becomes dominant from a certain number of fibre strands and, thus, increases the torsional rigidity. Further, the impact of these effects on the torsional rigidity depends on the ratio $\mu$ between the elastic moduli of the two mechanically decisive materials. For high ratios $\mu$, the impact of the effect in (e) and (f) is more pronounced, whereas for low ratios, the impact of the effect in (e) and (d) is more intense. The fineness of the triangulation is sufficiently high, so that even small gaps between fibre strands can be resolved, see (a).
Whereas, in extant perennial arborescent growth forms, the mechanical properties and stability are governed by secondary wood (= secondary xylem), this was not the case in a group of important forest-forming plants during the late Devonian/early Carboniferous members of the progymnosperms, lycopsids and pteridosperms\(^23, 26, 27, 29\). Periderm formation not only allows secondary stem growth increasing girth with growth along the outer protection layer, but also may take over mechanical functions\(^29–31\) and ensure repair after injuries\(^28\).

### Reinforcement by Sclerenchyma Fibre Strands

As documented in Table 2, sclerenchyma fibres are the stiffest material in our simulation with an elastic modulus of $E \approx 45,000$ MPa. Thus, we selected it as our reference material ($E_{\text{norm}} = 1$). We added fibre strands to the initial configuration, with the total area of the strands remaining constant. In this simulation, we investigated the effect of an increasing number of individual peripheral sclerenchyma strands in a parenchymatous matrix with vascular bundles being surrounded by an epidermis (see Fig. 3). The flexural rigidity remained almost constant independent of the number of sclerenchyma strands, namely between 18 and 63 strands (see Fig. 4b). In contrast, the torsional rigidity decreased nearly linearly from 18 to 49 strands. From 50 to 63 strands, it increased again to $\approx 140\%$ of the initial value (see Fig. 4a). Since the flexural rigidity remained almost constant, the twist-to-bend ratio initially increased almost linearly up to an optimum at 49 strands and then decreased to $\approx 2200\%$ of the initial value (see Fig. 4c).

This optimum of $N = 49$ strands found in the simulation corresponds well with the average value of $49.40 \pm 7.83$, which is calculated from the numbers of strands of the apical internode ($42.80 \pm 4.31$) and the more basal internode ($56.00 \pm 3.87$, see Table 1) of $C. \text{pendula}^7$. The clear optimum of the U-shaped curve (see Fig. 4a) is of interest in so far that sclerenchyma fibres are energetically highly costly for plants because of their extremely thick secondary cell walls that are impregnated with the macromolecule lignin. For annual plants, in particular, this is an expensive investment in a strengthening tissue that is, however, highly rigid. Since sclerenchyma fibres are dead cells, no further investment is required once they have been formed. This is especially advantageous for perennial plants.

With twist-to-bend ratios of up to 403, the internodes of $C. \text{pendula}$ show the highest values ever measured in plant axes\(^7\). The high flexural rigidity and low torsional rigidity of the flower stalk are particularly advantageous under dynamic wind loads. The high torsional flexibility allows the streamlining of the stalks together with the apical pendulous inflorescences or multiple fruits. In summer, the flower stalks have to bear these additional heavy fruits without bending down to the ground. Our simulations show that flexural rigidity cannot be achieved by the formation of additional sclerenchyma strands, as the number is already in the range of the optimum. However, a further increase in flexural rigidity is ensured by the rigid leaf sheaths that enclose large parts of the flower stalk. Since the leaf sheaths are not firmly attached to the stem, they are unlikely to have a strong negative effect on torsional flexibility.

Whereas, in extant perennial arborescent growth forms, the mechanical properties and stability are governed by secondary wood (= secondary xylem), this was not the case in a group of important forest-forming plants during the late Devonian and Carboniferous: the arborescent lycopsids. In this group with scale trees (genus: $\text{Lepidodendron}$) and seal trees (genus: $\text{Sigillaria}$), the periderm, which contributed over 90% of the stem volume, was by far the mechanically dominant tissue\(^23, 24, 29, 32\).

Peripheral stiffening structures were the first specialised stabilising tissues evolved in early land plants. After the colonisation of
We have shown here that the ratio of the elastic moduli of the materials, which are decisive for the mechanical performance of the withered leaf stalk cannot be restored to its healthy positioning, even with sufficient water support, the evolution of a redundant This work includes no experimental data. A C++-implementation including the computation of rigidities is available in the supplementary material.

Reinforcement by Collenchyma Fibre Strands
Collenchyma fibres are living cells that possess a vacuole and a thick non-lignified primary cell wall. Therefore, their elastic modulus is turgor-dependent. With an elastic modulus of \( E \approx 2,500 \text{ MPa} \), collenchyma fibres belong to the strengthening tissues \((E_{\text{norm}} = 0.05555)\) and are often found in still-growing shoots and leaves. From an energy point of view, collenchyma fibres are much cheaper to build than sclerenchyma fibres, because neither thick secondary cell walls nor the macromolecule lignin need to be formed in the former. Furthermore, collenchyma fibres have the advantage that they are still able to grow or to be stretched. On the other hand, as living cells, they always consume physiological energy and hence, are predominantly found in young growing tissues and in annual plants or plant organs.

The square-shaped flower stalks of the perennial \( L. \text{ cardiaca} \) show pronounced strands of collenchyma fibres at the four corners and small strands in the middle of the four sides. Dependent on the height above ground and increasing age, the cross-sectional percentage area of collenchyma decreases from 8.61\% to 6.18\%. With increasing age and mass attributable to the formation of flowers and heavy fruits, the peripheral lignified vascular tissues with a percentage cross-sectional area increasing from 15.55\% to 21.00\% increasingly take over the mechanical support of the stem. Analyses have demonstrated that the area sum of the vascular tissues and parenchyma exhibit moderate positive allometric scaling, whereas the collenchyma shows clear negative scaling.\(^5\).

In the circular leaf stalks of \( C. \text{ bicolor} \) ‘Candyland’, the cross-sectional percentage area of the \( \approx 40 \) peripheral collenchyma strands has a value of 3.9\% \pm 0.7\%. The cross-sectional area of the parenchyma and the lignified elements are 89.0\% \pm 3.3\% and 1.2\% \pm 0.4\%, respectively. Parenchyma and collenchyma are turgor-dependent. Together, they form a doubly secured mechanical system that is sensitive to drought stress. The decrease of flexural rigidity and, thus, the wilting of the leaf stalk are the result of a turgor-loss-induced decrease of the elastic moduli of both the collenchyma fibres and the parenchyma cells\(^9\). As a withered leaf stalk cannot be restored to its healthy positioning, even with sufficient water support, the evolution of a redundant mechanical system to maintain the flexural rigidity of the plant, in particular, is of great advantage for selection.

In the case of \( C. \text{ pendula} \), the replacement of sclerenchyma fibres by collenchyma fibres showed an increase of flexural rigidity by \( \approx 315\% \) instead of \( \approx 5600\% \) (see Table 3). Reinforcement by collenchyma fibre strands would be much too weak mechanically to support the stalk. Moreover, even the addition of an optimal number of sclerenchyma strands is probably insufficient. During autumn, in particular, when the flower stalk has a heavy top load of fruits, additional leaf sheaths increase the flexural rigidity of the overall system.

Conclusion
We have shown here that the ratio of the elastic moduli of the materials, which are decisive for the mechanical performance of the entire plant axis (\( \mu \)), plays a crucial role in plant stems. Reinforcements generally increase the flexural and torsional rigidities in a triangular cross-section composed of a parenchymatous matrix with embedded scattered vascular bundles surrounded by an epidermis. Closed ring-shaped reinforcement of the epidermal tissue (e.g. a periderm) leads to a considerable increase in torsional rigidity and a moderate increase in flexural rigidity in areas of the plants that are mechanically heavily loaded or at risk of damage. Therefore, epidermal reinforcements decrease the twist-to-bend-ratio. fibre reinforcement noticeably increases the flexural rigidity and moderately increases the torsional rigidity of the entire plant axis. The flexural rigidity is almost independent of the number of fibre strands, whereas the torsional rigidity and thus the twist-to-bend ratio is a function of strand numbers. Obviously, torsional stiffness is the key factor for changing the twist-to-bend ratio through structural reinforcements. The evolution of structural reinforcements including cells with walls strengthened by lignin was a prerequisite for land plants to be able to for colonise terrestrial habitats and successfully enabled them to face notably different mechanical constraints compared with those experienced in an aquatic environment. As outlined above, the primary (e.g. hypodermal sterome) and secondary (e.g. wood, cortex, periderm) strengthening tissues allowed plants not only to increase markedly in height, but also to colonise more and more hostile (drier) habitats leading to the plethora of plant life forms that we know from extant and fossil flora\(^{22-25,29}\).

Data availability
This work includes no experimental data. A C++-implementation including the computation of rigidities is available in the supplementary material.
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Author contributions statement

S.W.-V. conducted the mathematical experiments and wrote the first draft of the manuscript. O.S. was responsible for the description of the plants and contributed to the improvement of the first draft. T.S., O.S. and P.W.D. initiated the study. All authors contributed to the data interpretation and reviewed and improved the final draft of the manuscript.

Additional information

The authors declare no competing interests.