Novel Drug Delivery Systems for Loading of Natural Plant Extracts and Their Biomedical Applications

Heshu Sulaiman Rahman1,2
Henn Hassan Othman3
Nahidah Ibrahim Hammadi4
Swee Keong Yap5
Kawa Mohammad Amin6
Nozlena Abdul Samad7
Noorjahan Banu Alitheen8

1Department of Physiology, College of Medicine, University of Sulaimani, Sulaymaniyah 46001, Republic of Iraq;
2Department of Medical Laboratory Sciences, College of Health Sciences, Komar University of Science and Technology, Sulaymaniyah, Republic of Iraq;
3Department of Pharmacology and Toxicology, College of Pharmacy, University of Sulaimani, Sulaymaniyah 46001, Republic of Iraq;
4Department of Histology, College of Veterinary Medicine, University of Al-Anbar, Ramadi, Republic of Iraq;
5China-ASEAN College of Marine Sciences, Xiamen University Malaysia, Sepang, Malaysia;
6Department of Microbiology, College of Medicine, University of Sulaimani, Sulaymaniyah 46001, Republic of Iraq;
7Integrative Medicine Cluster, Institut Perubatan dan Pergigian T ermaju (IPPT), Sains@BERTAM, Universiti Sains Malaysia, Kepala Batas 13200, Pulau Pinang, Malaysia;
8Department of Cell and Molecular Biology, Faculty of Biotechnology and Bio-Molecular Sciences, Universiti Putra Malaysia, Selangor, Malaysia

Correspondence: Noorjahan Banu Alitheen
Department of Cell and Molecular Biology, Faculty of Biotechnology and Bio-Molecular Sciences, Universiti Putra Malaysia, 43400 UPM Serdang, Selangor, Malaysia
Tel +60 126866947
Email noorjahan@upm.edu.my

Heshu Sulaiman Rahman
Department of Physiology, College of Medicine, University of Sulaimani, Sulaymaniyah 46001, Republic of Iraq
Tel +964 772615958
Email heshu.rhaman@univsul.edu.iq

Abstract: Many types of research have distinctly addressed the efficacy of natural plant metabolites used for human consumption both in cell culture and preclinical animal model systems. However, these in vitro and in vivo effects have not been able to be translated for clinical use because of several factors such as inefficient systemic delivery and bioavailability of promising agents that significantly contribute to this disconnection. Over the past decades, extraordinary advances have been made successfully on the development of novel drug delivery systems for encapsulation of plant active metabolites including organic, inorganic and hybrid nanoparticles. The advanced formulations are confirmed to have extraordinary benefits over conventional and previously used systems in the manner of solubility, bioavailability, toxicity, pharmacological activity, stability, distribution, sustained delivery, and both physical and chemical degradation. The current review highlights the development of novel nanocarrier for plant active compounds, their method of preparation, type of active ingredients, and their biomedical applications.

Keywords: nanomedicine, natural plant metabolite, biomedical application, carrier formulation, drug delivery

Introduction

Based on the recently reported data, more than 70% of new drugs formulated are showing poor water solubility, which becomes the limiting factor in the absorption drug after oral admission.1 The limitation in the bioavailability of natural products active components includes poor solubility of the ingredient, poor stability due to gastric and colonic acidity, poor metabolism by the effect of gut microflora, poor absorption across the intestinal wall, poor active efflux mechanism and first-pass metabolic effects are among the factors that make the failure of clinical trials.2,3

In this respect, developed novel drug delivery system and carriers for herbal drugs should ideally accomplish some prerequisites such as proper delivering of the drug at a rate oriented by the needs of the body, over the period of treatment and it should pass the active entity of herbal drug to the site of action.4 Many approaches have been adopted to increase drug solubility, sustainability, bioavailability and gastrointestinal permeability.5 Nanocarrier has gained tremendous attention in the development of new pharmaceutical carrier and delivery systems. One of the strategies to thwart this problem is to encapsulate natural plant metabolites into the biodegradable and biocompatible nanoparticle.6

Employment of innovative drug delivery systems including utilization of nanocarrier delivery to overcome the physicochemical and pharmacokinetic limitation of phytochemicals enhanced the controlled release and even efficacy of the
bioactivities. This innovation shows the promising future of nanomedicine as a potential solution for impressive hindrance and handling of various chronic diseases.  

Additionally, altering the main features of nanocarriers such as their constituents (organic, inorganic or hybrid), sizes (small, medium or large), shapes (sphere, rod or cube) and surface properties (charge, functional groups, PEGylation or attachment of targeting moieties) are considered as a leading cause for tuning the physicochemical properties of nanocarriers. The overall aim of employing nanocarriers in drug delivery is to treat an unwellness effectively with the lowest side effects and potential outcomes.  

Nanomedicine has recently earned enhanced attraction for its ability to efficaciously diagnose and treat various ailments. Therefore, the aim of this review is to display the types of nanocarrier loaded natural plant products and focus on their role in various disease therapies with the promising use of nanomedicine.

Design of the Review

In this review, nanocarriers were being classified based on the types of nanocarrier, i.e. i) organic nanocarriers; ii) inorganic nanocarriers; iii) hybrid nanocarriers; and iv) biological nanocarriers. References were searched in Scopus data based using each class of nanocarriers as the keyword. Articles after the year 2010 were selected (unless the significant references for a particular type of nanocarrier, which were downloaded separately) and sorted based on the specific type of carrier for each of the above classes.

Nanocarrier

Nanocarrier is hopefully utilized to overcome the difficulty and issues related to conventional drug delivery systems such as their nonspecificity, side effects, burst release and detrimental destroying of large populations of the normal cells. Nanocarrier improves the bioavailability and therapeutic efficiency of drugs, as well as providing a preferential accumulation at the target site. Nowadays, a large number of nanocarriers have been produced but only some of them are clinically authorized for the delivery of materials because of their motivated actions at the targeted sites, especially antitumor agents.

The particles of a nanocarrier vary in size, and those ranged from 10 to 100 nm give the most acceptable physicochemical characteristics. The main advantages of nanonization are improving solubility, reducing medicinal doses and side effects, and increasing the absorbency of medicinal herbs compared with the respective crude extract preparations.

Types of Nanocarrier

Organic Nanocarrier

Lipid and Polymer-Based Nanocarrier

Solid Lipid Nanoparticle (SLN)

SLN is a colloidal drug carrier that developed in the early 1990s in which the particle size ranges from 50 to 1000 nm (Figure 1A). SLN is processed by using emulsifier(s) to stabilize the dispersion that composed of melted solid lipid(s) in water. The high-pressure homogenization (HPH) technique and microemulsification are the most commonly used methods for preparing SLN.

The main advantages of SLN are providing a highly lipophilic lipid matrix for drugs to be dispersed in, allowing the encapsulation, embedding with a wide range of molecules (such as drugs, antigens, proteins, and nucleotides) and also promoting the delivery of therapeutic loading into specific tissues and cells. Improving the in vitro and in vivo stability and reducing the adverse effects are also among the acceptable features of SLN. SLN is quite similar to nanoemulsions except that both solid and liquid lipids (oils) are used in the formulation of SLN whereas only liquid lipids are used in nanoemulsions.

The most extensively employed SLN is puerarin-loaded SLN in rats that characterized by rapid absorption, relatively improved bioavailability and increased tissue concentrations in targeted organs (heart and brain). Another group developed triptolide-loaded SLN as an antioxidant and anti-inflammatory product that showed a significant reduction in glutathione (GSH) and myeloperoxidase (MPO) activities. The aim of this development was to improve solubility, reduce toxicity, hyperemia, and irritation to the gastrointestinal tract (GIT) through minimizing direct contact with the mucosal surface, gradual drug-releasing, and avoiding high local drug concentrations. More examples in this respect are addressed in Table 1.
Nanostructured Lipid Carrier (NLC)

It is considered as a second-generation lipid nanoparticle that contains a mixture of solid and liquid lipids (Figure 1B) and was originally developed from SLN but with more lipid matrix imperfections. A wide variety of solid lipids have been utilized such as hydrogenated palm oil (HPO), glyceryl monostearate, stearic acid, and cetyl alcohol whereas the most commonly used liquid lipids are olive oil, mustard oil, castor oil, and cod liver oil. The preferable stabilizer in this system is thimerosal.

Generally, NLC preferred over the SLN because of better controlling of the drug release, more stability, enhanced drug-loading capacity, and minimized drug ejection during deposition. Thus, various active ingredients have been incorporated into NLC in studies focused on modifying water solubility, enhancing gastrointestinal absorption and oral bioavailability, controlling release, lengthening circulation time by reducing identification by the reticuloendothelial system (RES), and co-delivery. Therefore, it is realized that NLC is a better carrier for oral delivery of several natural and chemically synthesized compounds.

In this respect, silymarin loaded NLC is the best example that has been used clinically to overcome many hepatic diseases as its low solubility, permeability, and bioavailability often occur with its therapy. NLC loaded tripterine, curcumin, and triptolide are also other successful examples of corroborated absorption enhancement by this system which may be due to their small particle size, lipid components, and surfactant contents.

Cardomom essential oil (CEO) loaded NLCs have successfully been synthesized using food-grade lipids including cocoa butter and olive oil. The CEO loaded NLCs had a small size (90%), loading capacity (>25%) and provides good physical and chemical stability. This work overcame the limitation of applying the CEO to aqueous-based foods. Currently, various novel and innovative NLC have been produced as a carrier to target anticancer functions such as zerumbone, thymoquinone and citral and as a worthy drug observably increased antitumor activity in leukemia and breast tumor cells in vitro and in vivo. More examples of compounds loaded NLC are presented in Table 1.

Nanoemulsion (NE)

It refers to an optically single isotropic and thermodynamic stable transparent (translucent) nonhomogeneous colloidal dispersion system (Figure 2) with a droplet size...
| Nanocarrier | Example | Feature | Reference |
|-------------|---------|---------|-----------|
| **I. Organic Nanocarrier** | | | |
| **1. Lipid and Polymer-Based Nanocarrier** | | | |
| SLN | Puerarin (from *Pueraria lobata* (Wild) Howe)-loaded SLN | Rapidly and well absorbed, and its relative bioavailability was improved more than 3-fold as compared with that of the puerarin suspensions with increased tissue concentrations in targeted organs, particularly the heart and brain. | 15,17 |
| | Triptolide (from *Tripterygium wilfordii* Hook F)- incorporated SLN | Improved solubility, reduced toxicity, hyperemia, and irritation to the gastrointestinal tract (GIT) through minimizing direct contact with the mucosal surface, gradual drug-releasing, and avoiding high local drug concentrations. | 18 |
| | Cantharidin (from *Mylabris phalerata* Pakas or *Mylabris cichori Lineaeus*)-loaded SLN | Sustained release profile (half-life in circulation) without a burst effect, higher bioavailability after oral administration in rats induced with gastric mucus membrane irritation. | 19 |
| | Noscapine (from Papaveraceae family) PEG conjugated SLN | Improved biological half-life, drug delivery and higher anticancer efficacy in glioblastoma in vitro (U87 cells) and Swiss male albino mice induced with brain cancer. | 20 |
| | Tetrandrine (from *Stephania tetrandra* S. Moore)-loaded SLN | Prolonged the in vitro drug release, significantly enhanced the bioavailability in rabbit, and showed more efficient cellular uptake into the human lens epithelial cell line (SRA 01.04). | 21 |
| NLC | Cardamom essential oil (from *Elettaria cardamomum* Maton)-loaded NLC | Confirmed the encapsulation was able to protect the antimicrobial activity by Broth Macrodilution method. The results showed that Cardamom-loaded NLC could be used as food supplement. | 26 |
| | β-Elemene (from *Nigella damascena* L.)-loaded SLN | Showed significantly higher bioavailability in male Wistar rats and anti-tumor efficacy in H22 hepatoma bearing Kunming mice than Elemene, as well as less venous irritation and less toxicity after intravenous injection in New Zealand White rabbits. | 37 |
| | Thymoquinone (from *Nigella sativa*)-loaded NLC | Showed bioavailability and oral delivery enhancement in 4T1 bearing Balb/C mice together with the improvement of most liver biomarkers and anti-oxidant power, and correction of most liver injuries caused by a toxic dose of paracetamol in male albino rats. | 29 |
| | Citral (from *Cymbopogan citratus*)-loaded NLC | Enhanced the water solubility of the pure citral and sustained release, as well as exhibited no toxic effects on the proliferation of mice splenocytes and 3T3 cells. | 30 |
| | Zerumbone (from *Zingiber zerumbet* L. Smith)-loaded NLC | Enhanced water solubility, bioavailability, sustained result with better anticancer effects in vitro (Jurkat, MDA-MB231, 4T1, WEHI-3B, Caco-2, CMT-stylo cell lines) and in vivo (Balb/C mice model of leukemia and 4T1 challenged mice) with no toxicity. | 24,27,31–35 |
| NE | Hydroxy-safflor yellow A (from *Carthamus tinctorius*) NE | Improved bioavailability, enhanced systemic absorption along with produced higher cumulative transport of digested ME via lipid digestion by pancreatic lipase. | 322 |
| | Elemene oil (from *Curcuma species*) NE | Showed good stability, and improved oral bioavailability in Sprague Dawley rats than a commercial elemene emulsion. | 50 |
| | Oregano oil (from *Origanum vulgare*) NE | Controlled and reduced the growth of food-borne bacteria (L. monocytogenes, S. Typhimurium, and E. coli) on fresh lettuce. | 323 |
| | Basil oil (from *Ocimum basilicum*) NE | Demonstrated antibacterial activity against pure *E. coli* culture. | 51 |
| | Quercetin (from many plant parts such as nuts) NE | Stable O/W formula showed a remarkable increased in cutaneous permeability (reached the systemic circulation) with lower skin retention. | 53 |
| Compound Name | Source or Details | Improved Properties | References |
|---------------|-------------------|----------------------|------------|
| Tetrandrine | from the root tuber of *Stephania tetrandra* S. Moore | Improved liposolubility and controlled drug release in vitro, and oral absorption and bioavailability in Male Sprague-Dawley rat. | 57 |
| Cucurbitacin I | from different plants of family Cucurbitaceae Juss. | Avoid challenging the relative polarity of cucurbitacin I along with its hydrophobicity. | 58 |
| Zanthoxylum rhoifolium EO | | Improved drug release, water solubility, stability, and reduced toxicity, degradation under the action of oxygen, light and moderate temperatures and generating EO with insecticidal (Remia tabaci) action to optimize the pest control system. | 59 |
| Adhrocline satenroides EO | | AS-NC treatment protected the Wister's rat cardiac tissue damage from oxidative stress caused by *Trypanosoma evansi*. | 60,61 |
| Quercetin | from many plant parts such as nuts | Improved liposolubility and controlled drug release, more skin penetration with lower skin retention using ex vivo study and less toxicity on the other organs. | 62 |
| Oridonin (from *Rabdosia rubescens*)-conjugated PEG | | Improved bioavailability, hydrophilicity, prevented premature drug release, and improved pharmacokinetic behavior. | 62 |
| Curcumin (from *Curcuma longa* Linn)-phospholipid complex | | Offered an advantage to enhance the anticancer potential of cisplatin in A549 (lung carcinoma), HeLa (cervical carcinoma), and THP-1 (acute monocytes leukemia) cells. Induced more effectiveness in inhibiting NF-κB activation and in suppressing the expression of cyclin D1, MMR-9, and VEGF involved in cell proliferation, metastasis, and angiogenesis. Also increased lifespan in mice bearing Ehrlich's ascites carcinoma cells, with apparent regression of tumor volume. | 68, 69 |
| Trans-resveratrol (RSV) (from plants of berry family)-conjugated PCL and PLGA-PEG-COOH | | Controlled the RSV release, mimicking the acidic prostate cancer (DU-145, PC-3, and LNCaP) cell lines microenvironment, improved cytotoxicity and maximized uptake between NPs and cells, resulting in enhanced accumulation through endocytosis, and proving a consistent sensitivity toward both the androgen-independent DU-145 and hormone-sensitive LNCaP cells. Thus, offering the possibility of the administration of nanosystems via the parenteral and oral route. | 70 |
| Plumbagin (from *Plumbago indica* L.)-loaded aptamer (PLGA-PEG-COOH) | | Enhanced solubility, bioavailability, intracellular uptake, drug release, and biodistribution and thus enhanced to inhibit the growth, metastasis, and invasion of prostate cancer cells (LNCaP) in vitro. | 71 |
| PTX (from the bark of *Taxus brevifolia* or pacific yew)-loaded/PEGylated/saturated PC-based liposome | | Produced fine, homogeneous, and membrane filterable drug nanocarrier suitable for intravenous dosing. Enhanced solubility, bioavailability, intracellular uptake, and biodistribution. Improved solubility, sustained-release behavior, higher distribution with enhanced the drug-concentration in the brain tissues after intravenous administration in rats with middle cerebral artery occlusion model with highest targeting in striatum and cerebellum. Improved solubility with the sustained release in vitro. Prolonged the drug circulation time and increased the oral bioavailability of the drug in the male Sprague-Dawley rat by reducing the effect of adriamycin-injured myocardial ultrastructure and cardiomyocytes that showed an evident protective action. Improved water solubility, enhanced oral bioavailability and tissue distribution in liver tumor-bearing Kunming mice. Enhanced stability, solubility, sustained release in vitro and in vivo (Male Sprague-Dawley). Improved bioavailability and tissue (liver) distribution in Kunming mice after oral administration. | 78,79, 80, 81, 82, 83 |

(Continued)
Table 1 (Continued).

| Nanocarrier | Example | Feature | Reference |
|-------------|---------|---------|-----------|
| Transferosome | Paeonol (From peonies such as Paeonia moutan Slims)-loaded transferosome | Improved solubility, stability, transdermal delivery, skin retention and permeation in vitro using rat skin. Reduced skin irritation and inflammation in an Ex vivo using a male rat. | 89 |
| Transferosome | Capsaicin (from Capsicum plants)-loaded transferosome | Improved solubility, stability, flexibility and skin penetration and permeation in vitro using abdominal skin of Wistar rats. The product also improved anti-inflammatory effects, as well as it shows acceptable skin tolerability and anti-arthritis in the rat. | 90 |
| Transferosome | Apigenin (from fruits and vegetables such as parsley)-loaded transferosome | Showed good stability and a promising approach to improve the permeability of apigenin in sustained release for a prolonged period of time. | 91 |
| Transferosome | Epigallocatechin-3-gallate (from Camellia sinensis or green tea) and hyaluronic acid- loaded transferosome | Improved solubility and stability in vitro and skin permeation activity in ex vivo. Increased cell viability, reduced lipid peroxidation, intracellular ROS levels and expression of MMPs (2 and 9) in human keratinocyte cell line (HaCaT) that underline the potential application of the developed transfersomes in sunscreen crem/lotions for improvement of UV radiation-protection along with deriving antioxidant and anti-aging effects. | 92 |
| Transferosome | Emulgin (exudate from the aloe plant)-loaded transfersome | Improved solubility and stability in vitro. Showed anti-obesity in vitro using male rat by significantly reducing the body weight, wet weight of visceral fat, PBF and mRNA expression of G0S2 from perirenal fat tissue. As well as it improved insulin sensitivity. | 93 |
| Niosome | Lawsone (from Persian Henna, Lawsonia inermis)-loaded niosome | Improved stability, sustained release, bioavailability, and permeability in vitro. Significantly increased the antitumor activity in MCF-7 cells in vitro (enhanced therapeutic efficiency). | 102 |
| Niosome | Spermacoce hajedra-loaded niosome | Improved stability, sustained release, bioavailability, and permeability in vitro. Enhanced anti-tuberculosi in vitro. | 103 |
| Niosome | Embelin (from Embelia ribes Burm.)-loaded niosome | Improved stability, sustained release, bioavailability, and biocompatibility in vitro. Ameliorated streptozotocin-induced diabetes in Albino Wistar rats with potential antioxidant activity. | 107 |
| Niosome | Nerium oleander-loaded niosome | Improved cell effectiveness and improved tolerability of active substances. Enhanced in vitro cytotoxicity toward cervical and alveolar cancer cells (HeLa and A549) respectively, using MTT assay. | 105 |
| Niosome | Rosemarinic acid (from Rosmarinus officinalis)-loaded niosome | Improved niosomal gel of rosmarinic acid for sustained delivery to bacteria (Propionibacterium acne and Staphylococcus aureus) infected cells (anti-acne vulgaris) in vitro. Enhanced delivery of naturally occurring antimicrobial and anti-inflammatory agents, in deeper tissues of skin in vivo using Swiss albino mice. | 106 |
| Ethosome | Description | Improved Properties | References |
|----------|-------------|---------------------|------------|
| Colchicine (from dried coms and seeds of the genus Colchicum)-transethosomal gel | Improved stability, solubility, sustained release, bioavailability and skin diffusion in vitro. Enhanced drug accretion, tissue biodistribution and skin permeation in an ex vivo using Sprague Dawley rats’ back skin. | 109 |
| Apigenin (from many fruits and vegetables such as chamomile)-loaded ethosome | Produced strong anti-inflammatory activity caused by ultraviolet B light exposure after topical application. | 108 |
| Ginsenoside RII (from the root of Panax ginseng Mayer)-loaded ethosome | Enhanced skin permeation, retention and deposition in vitro using human cadaver skin. Also, the gel improved skin delivery of the compound on rat dorsal skin. | 88 |
| Cryptotanshinone (from Salvia miltiorrhiza)-loaded ethosomal gel | Enhanced more transdermal flux, skin permeation and deposition on pigskin in vitro. Also, the gel improved anti-acne activity with reduced skin irritation in the ear of rabbit model. | 111 |
| Berberis aristata extract loaded ethosomal gel | Enhanced permeation profile, as well as the transdermal delivery of the extract through ethosomal system, may be a better approach for dermatological disorders. | 112 |

| Dendrimer | Description | Improved Properties | References |
|-----------|-------------|---------------------|------------|
| Curcumin (from Curcuma longa Linn) encapsulated in PAMAM | Improved solubility, releaseability and delivery, thus the loaded dendrimer showed higher anti-proliferative activity against lung cancer A549 cell lines and had the better effect on the generation of intracellular reactive oxygen species (ROS), the mitochondrial membrane potential and cell apoptosis. | 122,123 |
| Puerarin (from the root of the Pueraria lobata (Wild) Howe)-loaded PAMAM | Promoted solubility, sustained release and improved oral bioavailability in the rat. In vitro hemolytic toxicity study revealed that this dendrimer did not cause hemolysis of fresh rat erythrocytes. | 124,125 |
| Silybin (from milk thistle, Silymarin)-loaded PAMAM | Improved aqueous solubility, stability, afforded the highest complex stoichiometry, and more extended release time. Additionally, it reduced the inherent dendrimer cytotoxicity using Alamar Blue cell viability assay on the human embryonal kidney 293 (HEK 293) cell line in vitro. | 126 |
| Anthocyanin (from Daucus carota L.) encapsulated silica-PAMAM dendrimer | Improved stability, solubility, sustained release, and cytotoxicity to neuroblastoma (A2) cell line with no toxicity to Yero (African green monkey kidney) normal cell lines in vitro. | 127 |
| Liquiritin (From Glycyrrhiza uralensis)-loaded PAMAM | Improved biocompatibility, solubility, permeability, and stability. No cytotoxicity of PAMAM dendrimers on human colon cancer (Caco-2) cells by MTT was observed in vitro. | 128 |

| Micelle | Description | Improved Properties | References |
|---------|-------------|---------------------|------------|
| Curcumin (from Curcuma longa Linn)-loaded polymeric micelle (MPEG-PCL) | Improved solubility, stability, and slow-released in vitro and improved pharmacokinetics in vivo (Sprague Dawley rat), as well as efficiently inhibited the angiogenesis on transgenic zebrafish model. Produced stronger cytotoxicity on C-26 colon carcinoma cells in vitro and in vivo (Balb/c mice) after intravenous injection. | 136 |
| Berberine (from Berberis plants) and diosmin (from citrus fruits)-loaded casein micelle | Improved solubility, delivery, and premature drug release. Enhanced superior cytotoxicity and higher cellular uptake against HepG2 liver cancer cells bearing mice revealed by down-regulation of cell necrosis markers (NF-κB and TNF-α), inflammatory marker COX2, inhibition of angiogenesis and induction of apoptosis. | 137 |
| 10-Hydroxy camptothecin (from Camptotheca acuminata) -loaded polymeric micelles | Enhanced liver targeting and pharmacokinetic (absorption) behavior in vivo using male Sprague Dawley rat model. Also showed a strong inhibitory effect on the activity of glutathione S-transferase after oral coadministration with vinegar baked Radix Bupleuri in the rat. | 138 |
| Shikonin (from the root of Lithospermum erythrorhizon and some other plants)-loaded thermosensitive micelle | Improved solubility, biodegradability, cellular internalization, and tumor accumulation. Promoted temperature-regulated passive targeting in vitro against breast cancer cells (MCF-7) and in vivo after intravenous administration to the BALB/c nude mice bearing breast cancer (MCF-7). | 139 |
| Sesbania grandiflora-loaded polymeric micelles | Enhanced solubility, stability, and sustained release. Improved antibacterial activity against S. aureus in vitro with no toxicity in vivo using healthy adult silkworm model. | 140 |

(Continued)
| Nanocarrier | Example | Feature | Reference |
|-------------|---------|---------|-----------|
| Nanosphere  | Silymarin (from *Silybum marianum*)-loaded PLGA | Improved encapsulation efficiency, sustained release, high internalization by cells and preferential toxicity to prostate cancer cells. | 147 |
|             | Nerolidol (from ginger and some other plants)-loaded nanosphere | Improved solubility, tissue targeting, therapeutic efficacy, and enabled the transporting of active principle through the blood-brain barrier (BBB). Produced effective elimination of *Trypanosoma evansi* from the central nervous system (CNS) in female mice after oral gavage. | 141 |
|             | Ginkgo biloba-loaded starch nanosphere | Improved solubility, sustained release, bioavailability, biocompatibility, biodistribution in vitro to enhance therapeutic efficacy. | 143 |
|             | Zanthoxyllum riedelianum fruit essential oil-loaded poly(-caprolactone) nanosphere | Improved solubility, stability, controlled release, and protection against photodegradation. Gained better insecticidal and deterrent activities against whitefly (*Bemisia tabaci*). | 148 |
|             | Menthol (essential oils of some plants)-loaded PLGA nanosphere | Enhanced control release, and biodegradability. The degradation of menthol-loaded PLGA nanoparticles in artificial saliva significantly affected the particles morphology and appears to be an effective medium for releasing menthol. | 149 |
|             | Nabilone (Cesamet®/Lilly) nanocrystal (from cannabis or marijuana) | Improved water solubility, stability, biodistribution, drug loading, biodegradation as an antiemetic agent for oral delivery. | 156–158 |
|             | Apigenin (from fruits and vegetables such as parsley) nanocrystal | Improved solubility, stability, bioavailability, biodistribution, and drug loading in vitro. Doubled antioxidant capacity that makes it available for dermal application. | 159, 160 |
|             | Curcumin (from *Curcuma longa* Linn)- nanocrystal | Improved solubility, stability, bioavailability and biodistribution. Enhanced skin penetration and uptake, and targeting hair follicles are also seen in vitro using the porcine skin. | 161 |
|             | Ursolic acid (from many plants such as *Mirabilis jalapa*) nanocrystal | Improved solubility, permeability, dissolution rate and oral bioavailability with prolonged retention. | 162 |
|             | Quercetin (from many plant parts such as nuts) nanocrystal | Improved solubility, permeability, dissolution rate, oral bioavailability with prolonged retention and enhanced antioxidant activity in vitro. | 163 |
| Phytosome or | Epigallocatechin gallate (from *Camellia sinensis*)-loaded phytosome | Improved solubility and bioavailability. Additionally, showed physicochemical stability through organoleptic, water content, and physicochemical properties during 6 weeks at various temperatures. Solved the problem of poor absorption, instability, insolubility and fast releasing, in vivo study using male albino rats showed a marked reduction in body weight, adipose tissue weight, and lipid profile. | 170 |
| Herbosome    | Soybean seed (from *Glycine max* L.) phytosome-based thermogel | Improved solubility, stability, releasing dynamics and bioavailability in vitro. Also fortified to be a good candidate as an antioxidant agent. | 171 |
|             | Rutin (from citrus fruits)-loaded phytosome | Improved solubility, stability, releasing dynamics and bioavailability in vitro. Also fortified to be a good candidate as an antioxidant agent. | 172, 173 |
|             | Butea monosperma flower extract-loaded phytosome | Improved solubility, stability, bioavailability, release dissolution pattern and free radical scavenging activity in vitro using DPPH model. | 174 |
|             | Gingerol (from *Zingiber officinale*)-loaded phytosome | Improved stability, bioavailability, sustained release and showed potent antioxidant, antibacterial (against *Staphylococcus aureus* and *E. coli*), and anti-inflammatory activities in vitro. Also, improved drug release and better oral absorption, showed no toxicity of blood parameters (pharmacodynamics study) in vivo using a New Zealand rabbit. Additionally, in vivo study of antioxidant, antibacterial and anti-inflammatory experiments were consistent and correlated with that of in vitro outcomes. | 175 |
| SNEDDS                        | Improved stability, solubility, oral bioavailability, absorption and sustained release in vitro with increased permeation in an ex vivo model using stomach and small intestine of the sacrificed male Sprague–Dawley rat. |
|-----------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Ellagic acid (from berries)-loaded SNEDDS |                                                                                                                                                                                                 |
| Quercetin (from many plant parts such as nuts)-loaded SNEDDS | Improved stability, solubility, sustained release and absorption in vitro using human Caco-2 cell monolayers. Also improved oral bioavailability and intestinal absorption in vivo using male Sprague–Dawley rat. |
| Akebia saponin (from the rhizome of Dipsacus asper Wall)-loaded SNEDDS | Improved liposolubility, stability, also the oral bioavailability and absorption in Sprague Dawley rats was enhanced significantly. |
| Curcumin (from Indian saffron)-loaded SNEDDS | Improved solubility, stability and releasing pattern in vitro and in vivo. Also improved cellular uptake in vitro using MDA-MB-231 breast cancer cell line. Enhanced oral absorption and bioavailability; intestinal perfusion in Sprague–Dawley rats with enhancement of cytotoxic action in metastatic breast carcinoma cell line in vitro and in 4T1 tumor-bearing BALB/c mice. Additionally reduced oxidative stress in treated animals. |
| Naringenin (from fruits such as grapes)-loaded SNEDDS | Improved solubility, stability, bioavailability and drug release in vitro and in vivo using male Albino Wistar rats. |

| SMEDDS                        | Improved solubility, stability, bioavailability, drug release and absorption in vitro using human intestinal cancer cell, the Caco-2 monolayer with fewer toxicity effects towards this cancer cell (due to the reduced toxic effect of the surfactant in the formula). Additionally, plasma concentration-time profiles from the oral absorption studies in male New Zealand white rabbits dosed with the system showed absorption of curcumin. |
|-----------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Curcumin (from Curcuma longa Linn)-loaded SMEDDS |                                                                                                                                                                                                 |
| Pueraria Flavone (from Radix puerariae)-loaded SMEDDS | Improved solubility, stability, and drug release in vitro, whereas enhanced oral bioavailability and absorption in the rat via the lymphatic uptake pathway. |
| Silymarin (from the fruit of milk thistle extraction (Silybum marianum or Carduus marianus))-loaded SMEDDS | Soft single capsule administration showed rapid absorption and high oral bioavailability in volunteer patients. |
| Camptothecin (from Camptotheca acuminata) -loaded SMEDDS | Improved solubility, and showed long-term stability with equipotent as compared to doxorubicin and had low toxicity in cervical cancer cells (HeLa), breast cancer cells (MCF-7), and leukemia (HL-60) cell line. |
| Lutein (from dark green leafy vegetables such as spinach and kale)-loaded SMEDDS | Improved small intestine absorption, transferring of into lymph and tissue distribution after oral administration in vivo using thoracic lymph-cannulated rats. |

(Continued)
| Nanocarrier               | Example                                                                 | Feature                                                                                                                                                                                                 | Reference |
|--------------------------|-------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------|
| Nanofiber                | Aloe vera gel-loaded nanofiber                                          | Increased hydrophilicity of fabricated nanofiber. Enhanced in vitro biocompatibility of gel-coated scaffold on fibroblast cells using MTT assay. Accelerated the in vivo wound-healing process and skin regeneration using male Balb/C mice model of the induced wound. | 205       |
| Sophora flavescens-loaded nanofiber | Improved fiber size distribution, thermal stability and increased the electrical conductivity of the polymer. Enhanced in vitro antimicrobial activity against a bacterial bioaerosol (Staphylococcus epidermidis) and filtration characteristics, such as the particle filtration efficiency and pressure drop. | 203       |
| Copaiba (from Copaifera plant species) oil-loaded nanofiber | Demonstrated a controlled drug release, increased hydrophilicity with greater antimicrobial action against Staphylococcus aureus in vitro. | 204       |
| Lycium barbarum-loaded nanofiber | The in vitro sustained release over several weeks through diffusion path and slow degradation enhanced both proliferation and neuronal differentiation of rat pheochromocytoma (PC12) cells induced by nerve growth factor (NGF), as well as peripheral nerve regeneration and neuroprotection. The promotion of rat Schwann cells myelination and neurite outgrowth of DRG neurons were also observed on loaded scaffolds by LSCM with immunostaining. | 207       |
| Gissus quadrangularis-loaded nanofiber | Increased in the adhesion, proliferation and osteogenic differentiation of mesenchymal stem cells (MSCs), as well as enhanced osteogenic differentiation of MSCs in vitro. Promoted mineralization when immersed in a simulated body fluid for 14 days. | 206       |
| Polymersome              | Doxorubicin-loaded polyphosphazene (PEP) polymersome                   | Provided the potential encapsulation of hydrophobic/hydrophilic drugs. The in vivo investigation in growth inhibition of MCF-7 xenograft tumors in nude mice demonstrated that PEP polymersomes could enhance life safety without compromise of therapeutic efficacy. Demonstrates similar efficacy but less toxicity compared to standard delivery methods. Overcome some limitations of oxymatrin such as short elimination half-life and poor distribution in the liver that resulting in low biological availability and some side-effects in male Sprague–Dawley rat model after intravenous injection. | 214       |
| Oxymatrin (from the root of Sophora flavescens)-loaded polymersomes | Overcome some limitations of oxymatrin such as short elimination half-life and poor distribution in the liver that resulting in low biological availability and some side-effects in male Sprague–Dawley rat model after intravenous injection. | 215       |
| Bacosides (from Brahmi, Bacopa monniera)-loaded polymerosome | Improved brain targeting and significant memory loss reversal in the chemically induced memory deficit mouse model using MRI technique. As well as to support a pharmacokinetic study of oxymatrin. | 216       |
| Lactoferrin-loaded polymerosome holding doxorubicin | Improved cytotoxicity, and increased cellular uptake and distribution in glioma cells (C6) in vitro. Enhanced a significant reduction in the tumor volume, and elongated the median survival time in the glioma model rat. | 217       |
| Paclitaxel (from the bark of the Pacific yew tree, Taxus brevifolia)-loaded polymerosome | Improved stability, bioavailability, and drug release. In vitro cytotoxicity showed the reduced viability of cultured SKBR3 breast cancer cells. | 208       |
| Substance                              | Description                                                                                                                                                                                                 | Page |
|---------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------|
| Cubosome Piperine (from fruits of family Piperaceae)-loaded cubosome | Improved suffering from hydrophobicity and first-pass metabolism with sustained release and more stability. Toxicological studies using male Wistar rats contended safety of the system on kidneys, liver, and even brain. In vivo studies using sporadic dementia of Alzheimer’s type (SDAT) model also revealed that this system has a potential to significantly enhance piperine cognitive effect and even restore cognitive function to the normal level. Additionally, exhibited potential anti-inflammatory, antioxidant, and anti-apoptotic activities, indicating the potential to stop Alzheimer’s disease (AD) progression and treatment. | 225  |
| Hinokitiol (HKL) (from the wood of trees in the family Cupressaceae)-loaded cubosome | In vitro, skin permeation experiments using the dorsal skin of female hairless mice revealed that the flux of this formula was much higher than in the case of HKL dissolved in water. Thus, the formula is thought to be one of the potent carriers in a hair tonic claiming for hair growth promotion. | 226  |
| Curcumin (from Curcuma longa L.)-loaded cubosome | Produced more stable and nano-sized vesicles that able to improve curcumin antibacterial (Escherichia coli) activity in topical drug delivery. No signs of erythema or redness were observed until 7 days on skin irritation study using healthy Wistar rats. Enhanced skin permeability in vitro using goat ear skin. | 227  |
| Adyranthes bidentata polysaccharides- loaded cubosome | Improved stability, feasible immunomodulatory and less cytotoxicity to splenic lymphocytes in vitro. Promoting a potential lymphocyte proliferation and triggering the transformation of T-lymphocytes into Th-cells using flow cytometry study (improved immune response). | 228  |
| Ulva fasciata polysaccharides-loaded cubosome | Showed significant decreases in total cholesterol (TC), triglycerides (TG) and total lipid (TL) in vivo using a rat model of hyperlipidaemia. Displayed significant reduction in malondialdehyde (MDA), whereas insignificant changes were detected in nitric oxide (NO), glutathione (GSH) levels and total antioxidant capacity (TAC). Inhibited the expression of VCAM-1 and ICAM-1, which is known to have a protective effect on the progression of atherosclerosis. | 229  |

(Continued)
Table 1 (Continued).

| Nanocarrier | Feature | Reference |
|-------------|---------|-----------|
| 1.2 Biopolymer Based Nanocarrier | | |
| Pure Biopolymer Nanocarrier | Enhanced antitumor efficacy of Chit-nanoEGCG in subcutaneously implanted 22Rv1 tumor xenografts in athymic nude mice through significant inhibition of tumor growth and secretion of prostate-specific antigens. Additionally, there was the significant induction of poly (ADP-ribose) polymerases cleavage, increase in the protein expression of Bax with a concomitant decrease in Bcl-2, activation of caspases and reduction in Ki-67 and proliferating cell nuclear antigen. | 237 |
| Curcumin (from Curcuma longa L.)-loaded chitosan | Decreased cell viability and induced apoptosis of B16F10 melanoma cells as well as significantly decreased the expression of metalloproteinases, a key biomarker for migration and proliferation of cancer cells in vitro. Enhanced attenuation of melanoma in the lungs by decreasing pulmonary tumor formation in a female C57BL/6 mice model of experimental metastasis. | 238 |
| Curcumin (from Curcuma longa L.) loaded chitosan nanoparticles impregnated into collagen-alginate scaffolds | Improved in vitro biodegradability, biocompatibility, and drug release. Enhanced significant in vivo wound contraction and closure with complete epithelialization and thick granulation tissue formation using adult male Wistar rats. | 239 |
| Rotenone (from the seeds and stems of Jicama vine, and the roots of several members of Fabaceae) -loaded chitosan | Enhanced the production of nontoxic drug carriers that can be used to solubilize, stabilize, and control the release of lipid-soluble rotenone in water that contributes to the development of green and efficient nano-botanical pesticide preparing methods for water-based formulations. | 240 |
| Zingiber cassumunar Roxb-loaded chitosan | Confirmed the hydrophilicity of the herbal blended patches through moisture uptake, swelling ratio, erosion and porosity. Enhanced the in vitro controlled release and skin permeation behavior using newborn pigskin. | 241 |
| Biopolymer Hydrogels | | |
| Yerba mate (Ilex paraguariensis A. St.-Hil.)-loaded calcium alginate hydrogels containing cornstarch | Increased the in vitro entrapment capacity of yerba mate polyphenols, modulated the antioxidants release rate and diminished the contribution of matrix erosion to the whole release mechanism. Improved the classic calcium alginate system, leading to a promising strategy to protect and deliver yerba mate antioxidants into food products. | 245 |
| β-carotene (from many fruits and vegetables)-loaded rice starch-based hydrogels | Increased β-carotene bioaccessibility that leads to faster digestion of trapped lipid droplets inside starch filled hydrogels in simulated gastrointestinal tract conditions. Encouraged the development of rice-based functional food products fortified with lipophilic bioactive components to improve human health and wellness. | 246 |
| The hydrogel containing Achyrocline satureioides extract loaded nanoemulsion | Improved in vitro protection of the porcine ear skin against oxidative stress generated by UVA/UVB light using TBARS, protein carbonylation, and protein thiol content assays. | 247 |
| Cashew gum (from Anacardium occidentale L.) modified with phthalic anhydride-carboxymethyl Cellulose hydrogel | Showed potent antibacterial activity against Staphylococcus aureus and Pseudomonas aeruginosa strains in vitro which confirmed by reduction in MIC and MBC, respectively. Improved wound healing profile in vivo using male Wister rat model with cervical dorsum wounds using manual trichotomy method. | 248 |
| Salix alba-loaded chitosan-based hydrogel | Showed no toxicity to human embryonic kidney (HEK 293) cells (wound healing model cell) in vitro using MTT assay. Showed highest inhibitory rate (antimicrobial activity) towards Salmonella typhi and Candida guilliermondii microorganisms using disc diffusion method. Suggested using such thin film coatings in medically relevant applications for the wound treatment. | 249 |
| Biopolymer Drug Conjugate                                                                 | Improvement                                                                                       | References |
|--------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------|------------|
| Resveratrol (from berry family)-loaded whey protein-dextran colloidal complex for delivery of β-carotene (from many fruits and vegetables) | Significant improvement of environmental stress (ionic strength, heat, and pH) and storage stability. Chemical stability was also remarkably enhanced when exposed to UV light and thermal treatment. Provided a better alternative to effectively protect and deliver hydrophobic nutraceuticals. | 254        |
| Chitin-glucan-aldehyde-quercetin (from many plant parts such as nuts) conjugate               | Grafting of quercetin depicted several changes on its surface with observing more crystalline nature. Displayed stronger antioxidant activity in vitro using DPPH* and ABTS*+ scavenging assays. Showed excellent cytotoxicity in vitro against macrophage cancer cells (J774) but no cytotoxicity towards peripheral blood mononuclear cells (PBMCs). | 255, 256   |
| Curcumin (from Curcuma longa L)/cyclodextrin polymer inclusion complex                       | Improved physio-chemical characterization compared to free curcumin using XRD, FTIR, DSC and UV assays. Exhibited novel antioxidant activity by scavenging the ABTS and DPPH free radicals. Displayed higher antiproliferative activity with apoptosis induction on A375 cells. | 257        |
| Redox-responsive PEGylated periplocymarin (PPM)-vitamin E conjugate                         | Improved in vitro stability, controlled release, and cytotoxicity towards liver and breast cancer cells of HepG2 and MCF-7, respectively using MTT assay. Prolonged in vivo circulation time with an elimination phase half-life. Induced in vivo suppression of tumor growth without obvious systemic toxicity in malignant H22 (hepatocarcinoma)-bearing Kunming mice. Suggested that this inclusion complex could be developed as a novel natural antioxidant with potential applications in cancer chemoprevention. | 258        |
| Camptothecin (from Camptotheca acuminata) conjugates                                         | Improved stability, solubility and biocompatibility in vitro. Enhanced permeability and retention effect. Empowered nanoparticles to be selectively targeting cervical (HeLa) and liver (HepG2) cancer cells in vitro. | 259        |

2. Inorganic Nanocarrier

| Metal NP                                                                 | Improved stability, solubility, biocompatibility, biodegradability and release of the compound. Suggested to be a suitable formula for natural product anticancer drug delivery. | 265        |
| Licochalcone A (from Glycyrrhiza inflata)-loaded hollow Au-NP              | Significant improvements obtained for phenolic and flavonoid contents of the plant when grown with AuNPs. Improved antioxidant activity in vitro using DPPH assay. | 266        |
| Amaranthus caudatus Linn. Pod extract-loaded Ag-NP                        | Improved in vitro cytotoxicity against murine myeloid leukemia (WEHI-3B) cells with no effect on normal mouse fibroblast (3T3) cells. Showed distinct morphological changes using fluorescent dyes; the apoptotic population was increased via flow cytometry, and the caspase activations contributed to ZnO-NPs triggered apoptotic death in WEHI-3 cells. | 267        |
| Brown seaweed (Sargassum muticum)-loaded ZnO-NP                           | Improved stability, solubility, biodegradability, and drug releasing capacity in vitro. Enhanced in vitro anti-tumor activity on human breast adenocarcinoma cells that confirmed by apoptosis and necrosis induction as well as inhibiting of migration in MCF-7 cell line. | 268        |
| Curcumin (from Curcuma longa Linn) and sulforaphane (from cruciferous vegetables)-loaded iron oxide-gold core-shell NP | Improved in vitro antioxidant, antibacterial (Sophyobacous epidermidis and Escherichia col), and antiproliferative activities toward human leukemia (MOLT-4) cell line. | 269        |
| White tea (Camela sinensa) -loaded palladium NP                           | Protected the quality and quantity of total flavonoid and phenolic content. Improved in vitro antioxidant, antibacterial (Sophyobacous epidermidis and Escherichia col), and antiproliferative activities toward human leukemia (MOLT-4) cell line. | 269        |

(Continued)
Table 1 (Continued).

| Nanocarrier                      | Example                                                                 | Feature                                                                                                                                                                                                 | Reference |
|----------------------------------|-------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------|
| Mesoporous Silica NP             | Curcumin (from *Curcuma longa* Linn)-loaded folic acid-conjugated MSNP | Displayed higher cellular uptake and sustained intracellular release. Apoptosis was induced in vitro human hepatocellular carcinoma cells (HepG2) and cervical carcinoma (HeLa) cells through specific signaling molecular pathways (Caspase-3, H2O2, c-MET, and MCL-1). Enhanced in vitro antioxidant activity using DPHH+ and ABTS+ scavenging assays. | 275       |
| Paclitaxel (from yew tree, *Taxus brevifolia*) and curcumin (from *Curcuma longa* Linn)-loaded PEGylated lipid bilayer coated MSNP | Enhanced in vitro antioxidant activity using DPHH+ and ABTS+ scavenging assays. Improved stability, dissolution ability and sustained release in vitro. Exhibited high degree dispersity that enables the intravenous administration of hydrophobic drugs. Manifested definite and persistently promoted cytotoxic effect against canine breast cancer cells (7364) in vitro using CCK8 essay. | 276       |
| Ursolic acid (from many plants such as *Mirabilis jalapa*)-loaded MSNP | Characterized by high loading capacity, high cellular uptake and sustained release with improved bioavailability. Showed in vitro cytotoxicity, proliferation inhibition, G2/M cell cycle arrest and apoptotic effects against human hepatocellular carcinoma HepG2 cells. | 277       |
| Guar gum (from leguminous plants)-capped MSNP (GG-MSN) | Showed enzymatic biodegradation of guar gum by colonic enzymes in the simulated colonic microenvironment that specifically triggered the release of 5-FU from GG-MSN. Manifested anticancer activity in colon cancer (HT-29) cells in vitro confirmed by flow cytometry and biochemical assay. GG-MSN system also demonstrated near perfect “zero release” property in absence of enzymes in different simulated conditions of the gastrointestinal tract. | 278       |
| Axitinib and celastrol (from the root extracts of *Tripterygium wilfordii* and *Celastrus regelii*)-loaded PEGylated lipid bilayer coated MSNP (ACML) | Effectively internalized and showed cytotoxicity in human breast cancer (BT-474), murine squamous cell carcinoma (SCC-7), and neuroblastoma-derived cell line (SH-SY5Y) in vitro using MTS assay. ACML-treated mice showed remarkably higher tumor inhibition in tumor xenograft models. Tumor xenograft immunohistochemistry revealed elevated caspase-3 and poly (ADP-ribose) polymerase and reduced CD31 and Ki-67 expression, suggesting tumor apoptosis through mitochondrial and antiangiogenic effects. | 279       |
The in vitro release of formulated nanocomposites showed an initial burst release in the first 18 hrs, followed by a more gradual and sustained release for 120 hrs. Disk diffusion antibacterial test showed that the loading of CPX on the Fe₃O₄@BSM nanocarrier does not have any negative effect on the structure and performance of the drug and increases the antibacterial properties of CPX.

Sargassum muticum-loaded Fe₃O₄ magnetic NP

Considered as a simple, rapid, efficient, one-step green method involving reduction of ferric chloride solution using brown seaweed aqueous extract containing hydroxyl, carboxyl and amino functional groups. Enhanced in vitro anticancer activity in human cell lines for leukemia (Jurkat cells), breast cancer (MCF-7 cells), cervical cancer (HeLa cells) and liver cancer (HepG2 cells) using MTT, Annexin V, Cell cycle and protease inhibition assays.

Argemone mexicana L. (Mexican prickly poppy) leaf extract-loaded Fe₃O₄ magnetic NP

Showed a noteworthy inhibition on Escherichia coli MTCC 443 and Proteus mirabilis MTCC 425 growth in vitro using disc diffusion method. Proved that the immobilized nanomaterials of magnetite can effectively improve the drug loading and has wide scope in opening an excellent drug delivery system.

Ocimum basilicum mucilage-loaded magnetic cobalt ferrite NP

The magnetic properties of the produced system ensure an easy separation of the nanocomposites from the aqueous medium by means of an external magnetic field. Antibacterial disk diffusion test results demonstrated that the synthesized nanocomposite exhibits excellent antibacterial activity against bacteria (Staphylococcus aureus, Bacillus cereus, Escherichia coli and Salmonella typhimurium).

Gallic acid (from a variety of fruits and plants)-loaded Fe₃O₄ magnetic NP

Enhanced the thermal stability and controlled release of the active drug from the nanocarrier. Showed no toxicity in a normal human fibroblast (3T3) line, and anticancer activity was higher in human colorectal adenocarcinoma (HT29) cells than human breast cancer cells (MCF7) in vitro.
| Nanocarrier                  | Example                                                                 | Feature                                                                                                                                                                                                 | Reference |
|-----------------------------|--------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------|
| **Nanotube**                |                                                                          |                                                                                                                                                                                                        |           |
| Halloysite Clay Nanotube    | Thyme oil (from *Thymus vulgaris*)-encapsulated HNT                       | Improved in vitro sustained release for more than 3 weeks at 4 and 25 °C using the Korsmeyer–Peppas model and Arrhenius model. showed in vitro antimicrobial effects toward *Escherichia coli* O157:H7, total mesophilic aerobic bacteria, molds and yeasts using vapor phase assay. It is suggested to be a good candidate for antimicrobial food packaging system. | 298       |
|                             | Curcumin (from *Curcuma longa* Linn)-loaded HNT                          | Improved in vitro colloidal stability and wettability that might be triggered by temperature stimuli. In vitro tests simulating the gastrointestinal transit demonstrated that the proposed delivery system allows a targeted release of curcumin, preventing its degradation in an acidic medium. | 299       |
|                             | Silibinin (from *Silbury marianum*) and quercitin (from many plants and foods such as berries)-grafted covalent cyclodextrin HNT | Improved stability, sustainability, and drug uptake in vitro. Exhibited an antiproliferative activity against the human anaplastic thyroid cancer cell lines (8505C) in vitro using MTT assay. These results demonstrated the suitability of the complex as nano-co-delivery vehicles of multi drugs to enhance synergic effects in anticancer therapy and as efficient tools for transport of drugs into living cells | 300       |
|                             | Peppermint (from *Menta piperita*) essential oil (PO)-loaded HNT          | Evidenced the successful functionalization of halloysite surfaces. Showed that the temperature increase induces an enhancement of the total PO release highlighting that the mechanical action of the bionanocomposite is thermo-sensitive. Enhanced the in vitro antioxidant activity using the DPPH method. showed strong in vitro antibacterial activity against *Escherichia coli* and *Staphylococcus aureus*. Suggested an easy strategy to prepare a functional sustainable edible film with thermo-sensitive antioxidant/antimicrobial activity that can be useful for a multi-step food conservation. | 301       |
|                             | Rosemary (from *Rosmarinus officinalis*) essential oil-loaded HNT         | showed that the in vitro kinetics of release of the rosmarinic acid is via controlled release using UV spectrometry. Antimicrobial assay indicated no mold growth after 3 months on the film stored at room temperature. Proved the potential of the systems in the active packaging field. | 302       |
| **3. Hybrid Nanocarrier**   |                                                                          |                                                                                                                                                                                                        |           |
| *Panax* notoginsenoside-loaded core-shell HN (PNS-HLV) | Improved bioavailability, stability and in vitro controlled drug release. Significantly inhibited the edema of the brain, reduced the infarct volume, markedly inhibited H2O2, modified Dixon agar, and serum lactate dehydrogenase, and increased superoxide dismutase in vivo using male Sprague Dawley rats induced with global cerebral ischemia/reperfusion and acute myocardial ischemia. Provided promising prospects for improving free drug bioactivity on oral administration. | 312       |
### 4. Biological Nanoparticles

| Name of Nanoparticle | Description |
|----------------------|-------------|
| **Potato virus X (PVX) nanoparticle** | Using a combination of ex vivo whole-organ imaging, quantitative fluorescence assays and immunofluorescence microscopy, the biodistribution and clearance of PVX was up to 30% from the colon, mammary and brain tumor tissues, remaining particles were cleared by the reticuloendothelial system organs (the spleen and liver), followed by slower processing and clearance through the kidneys and bile. Increased tumor homing and tissue penetration as well. |
| **Tobacco mosaic virus (TMV) nanoparticle** | Used in platform technology characterized by monodispersity, biocompatibility, capability for scale-up production, and amenability to multiple functionalization strategies. Enhanced biodistribution using confocal microscopy as well as improved fluorescent imaging of tissues ex vivo using Maestro Imaging System and fluorescence quantification in homogenized tissues of mouse xenograft tumor model. |
| **Black-eyed pea cowpea mosaic virus (CPMV) nanoparticle** | Suted for long-term intravital vascular imaging due to its biocompatibility and retention in the endothelium with minimal side effects. Enhanced imaging of vascular structure and intravital vascular mapping in developmental and tumor angiogenesis models using CD1 mouse embryos. |
| **Potato virus X (PVX) incorporating doxorubicin (DOX) nanoparticle** | Used for immunotherapeutic for in situ vaccine monotherapy through increased survival that represented in the enhanced antitumor cytokine/chemokine profile. Elicit delayed tumor progression in B16F10 melanoma. |
| **Recombinant plant virus-based nanoparticles (PVNs)** | Provided platforms for the induction of humoral and cellular immune responses to genetically fused antigens from pathogenic viruses, bacteria, tumors and toxins in man and animals. Developed as a prophylactic and/or therapeutic vaccines for the prevention or treatment of several microbial diseases, pathologies and toxin poisoning. |
of less than 100 nm. Generally, the NE is composed of stabilized oil and water with the aid of surfactant and cosurfactant an interfacial film molecule.9

After the lipophilic drugs loaded into either oil/water or oil/water/oil suspension, the oil droplets are engulfed by the macrophage and find in a high concentration in the spleen, liver, and kidneys since the quantity of the liquefied medicate is too big. Whereas the hydrophilic drug is encapsulated into water/oil or water/oil/water nanoemulsion, it can be well condensed in the lymphatic system through subcutaneous or intramuscular insertion due to the higher internal membrane permeability.

This system is characterized by targeted sustained release, the stability of the solubilized components, enhanced the permeability of materials to the mucous and skin, solubilized components of varied lipophilicity, improved drug absorption, lowered viscosity with inducing less pain or allergic reactions and simpleness of production and decontamination as well.39 Moreover, the intestinal absorption of NE is attributed to the lymphatic conveyance processes that ameliorate the oral bioavailability of encapsulated materials.40,41

NE serves as an attractive vehicle for the delivery of drugs and essential oils (especially as repellent and antimicrobial agents, nucleic acids as well as imaging agents).39,42,43 In the last few years ago, a modern system has upgraded the transdermal remedial use of NE, such as Transcutol®P, phospholipid, alkyl polyglycosides, PEGylated fatty acid ester, and fatty alcohol.54–46

Herbal drugs including camptothecin, rutin, genistein, resveratrol, and oils of Brueca javanica, coixenolide, and zedoary have been loaded into NE for various applications.47,48 With great application prospects of NE, Syagrus romanzzoffiana fruit pulp extracts were incorporated into O/W NE using the phase inversion method to evaluate antioxidiant activity.49 More examples of herbal loaded NE are presented in Table 1.51–53

Nanocapsule (NC)

It is a nanovesicular colloidal dispersion system (Figure 3) that exhibits a typical core-shell structure in which the drug is confined to a reservoir or within a cavity surrounded by a polymer membrane or coating.52 The cavity can contain the active substance in liquid (an oily or an aqueous core) or solid form or as a molecular in which the core-shell structure and composition are the main features of NC especially controlling the drug release.54 Likewise, this formula can be lipophilic or hydrophobic according to the preparation method and raw materials used. The main aim in developing this formula is to alter the oral bioavailability of ailing hydrophilic active components.55

Additionally, as asserted by different authors, other advantages of NC as a carrier system include high drug encapsulation efficiency due to optimized drug solubility in the core, low polymer content compared to other systems, drug polymeric shell protection against degradation factors like pH and light and the reduction of tissue irritation due to the polymeric shell.56 Recently, a ligand-modified or multifunctional NC that carries the active substance on their surfaces or imbibed in the polymeric membrane has been developed to attain higher delivery of therapies to the targeted site more actively.54 Different preparation methods such as nanoprecipitation, emulsion–diffusion, double emulsification, emulsion-coneavration, polymer-coating, and layer-by-layer were employed to develop various types of this carrier.55

In this area, the well-known anticancer natural herbal product, artemisinin (ART) (from Artemisia annua) crystals were encapsulated with polyelectrolytes (chitosan,
be used for improving age-related diseases such as cancers of various organs and Alzheimer’s disease. Resveratrol efficacy was halted significantly due to its instability, and solubility especially in vivo model. Thus, resveratrol conjugated transferrin (Tf)-modified polyethylene glycol-polylactic acid (PEG-PLA) nanoparticle (Tf-PEG-PLA-RSV) was developed to target transferrin receptor overexpression in C6 glioma cells in vitro and to inhibit tumor maturation in rats induced with C6 glioma. More examples of herbal loaded NE are presented in Table 1.

Liposome

A liposome is a spherical shaped polar lipid nanoparticle that encapsulates an aqueous core by single or multiple natural or synthetic lipid bilayer membranes, in which it freely diffuses into its interior (Figure 5A). A liposome is known to have both hydrophilic and lipophilic groups on the same molecules and thus it can load both hydrophilic and lipophilic materials and can have single or multiple homocentric membranes as well. The pharmacokinetic profiles of drugs, herbs, vitamins, and enzymes can be modified extraordinarily by encapsulating them with liposomes for the purpose of preparing vaccines, cosmetics, and nutraceuticals. Because of liposome’s unique feature of having phospholipid bilayers as well as accommodating both water-soluble and lipid-soluble agents, it is able to enhance the solubility, bioavailability, delivery, intracellular uptake and biodistribution performance of the products both in vitro and in vivo. Additionally, defending of active drug from environmental factors, overwhelming primal destruction of the loaded material, less costly and prompt treatment with minimum systemic morbidity that has magnified their use in bio-medicine formulations.

The most commonly used polymers to elongate their half-life, as well as stability, are PEG and poly(lactic-co-glycolic acid) (PLGA). On the other hand, antibodies or ligands can be conjugated to liposome in order to enhance their target specificity, such as incorporating of curcumin into liposomes coated with PSMA antibodies by Thangapazham et al to enhance targeted delivery of curcumin for prostate cancer. They used LNCaP and C4-2B human prostate cancer cell lines in their study and realized that treatment of cells with liposomal curcumin leading to at least 70–80% inhibition of cellular proliferation without affecting their viability, with a 10-fold dose advantage over free curcumin. More examples of herbal loaded NE are presented in Table 1.

Lipid Drug Conjugate (LDC) or Polymer Drug Conjugate (PDC)

The union of agents with polymers is a new approach to modify drug property and its pharmacokinetics. LDCs are lipidic drugs that covalently or noncovalently coupled to a lipid moiety, such as diglyceride, phosphoglyceride and fatty acid (Figure 4). In several instances, LDC may also be known as Pharmacosomes especially when the drug is conjugated with a phospholipid. LDC is the most accepted lipid-based nanoparticle, especially when considered drug is hydrophilic in nature in which it is converted into water-insoluble lipid-drug conjugate by conjugating it with a lipid component.

LDC is characterized by possessing controlled drug release, drug targeting, an increase in gastrointestinal permeability, an increment in bioavailability. Additionally, adding targeted motifs to the polymer to produce functionalized polymer–drug conjugate can also be constructed. One of the appreciable natural products with high edible polyphenolic content is resveratrol that is widely known to gelatin, and alginate) for the purpose of controlled release through self-assembly of polyelectrolytes on drug crystals, and improved hydrophilicity of the crystal using the layer-by-layer technique. More examples of herbal loaded NE are presented in Table 1.

Figure 4 A schematic illustration of Polyethylene glycate (PEG)-aptamer-liposome-doxorubicin (DOX); a type of lipid drug-conjugate. Notes: Reproduced from Dou XQ, Wang H, Zhang J, et al. Aptamer–drug conjugate: targeted delivery of doxorubicin in a HER3 aptamer-functionalized liposomal delivery system reduces cardiotoxicity. Int J Nanomed. 2018;13:763-776.
Transferosome

The conventional liposomes do not deeply penetrate the skin and remain confined to the outer layer of stratum corneum. Thus, new classes of lipid vesicles such as transfersomes have been developed as an enhanced type of liposomes,\textsuperscript{11} which is an ultra-flexible lipid-based elastic vehicle with highly deformable membranes that enhance the sending of materials to deeper skin tissues through a nonoccluded method which penetrates the intercellular lipid lamellar regions of stratum corneum due to the hydration or osmotic force of the skin.\textsuperscript{84}

It is composed of phospholipid and a single chain surfactant that provides elasticity and deformability to the vesicles (Figure 5B) that can be used topically for the purpose of supplying the nutrients locally to maintain the skin.\textsuperscript{85} This unique infrastructure endows transfersome to entrap hydrophilic, lipophilic, and amphiphilic drugs and thereof it can be utilized as drug carriers for small molecules, peptides, proteins, and herbal components as well as it can accommodate drug molecules with a wide range of solubility.\textsuperscript{9} On the other hand, chemically, transfersomes are not stable because of their predisposition to oxidative degradation and the purity of natural phospholipids is another criterion militating against the adoption of transfersomes as drug delivery vehicles.\textsuperscript{86}

Additionally, transfersomes are not difficult to scale up, as the process is simple, easy to scale up without using pharmaceutically unsatisfactory additives.\textsuperscript{87} In this respect, ginsenoside Rh1 from Red ginseng (the steamed root of Panax ginseng C. A. Mayer) transfersome has been developed for skin maintenance that provided significantly higher skin penetration and higher topical absorption in comparison to ethosome and conventional liposome using rat dorsal skin in vitro.\textsuperscript{88} More examples

Figure 5 A schematic illustration of liposome (A), transfersome (B), niosome (C) and ethosome (D).

Notes: Adapted with permission from Frontier in Pharmacology. Sercombe L, Veerati T, Moheimani F, Wu SY, Sood AK, Hua S. Advances and challenges of liposome assisted drug delivery. Front. Pharmacol. 2015;6:286.\textsuperscript{324}
of herbal loaded transfersomes are presented in Table 1.89–93

Niosome
Niosome is a nonionic nanosphere vesicle with a diameter of 100 nm to 2 μm, in which its center is watery that surrounded by layers of nonionic amphiphilic lipids in lamellar phase (Figure 5C).94 It is prepared by thin-film hydration method, sonication, microfluidization, multiple membrane extrusion, reverse phase evaporation technique, remote loading, bubble method and proniosome pre-formulation technique.95

Niosome is almost similar to liposome in structure but with more penetrating capability, more stability and therapeutic index of a drug, and less toxicity, thus it could offer more advantages over liposome.96 The advantages of niosome include cost-effectiveness, high solubility and flexibility and controlled release of its content. Therefore, they have been utilized widely as a targeting vehicle for neoplasia or as peptide carrier, hemoglobin carrier, and transdermal delivery.97

In tropical application, niosomes were also showed prolonged circulation, sustained release and retention in the skin and facilitated the permeation of the drug into the skin.98 Niosomes were reported to be more stable without significant toxicity than liposomes especially when used topically for treatment of skin diseases. In this regard, niosome loaded resveratrol for topical treatment of skin cancers is one of the potential candidate.99,100 Similarly, the topical gel from Zingiber cassumunar Roxb. extract loaded niosome for anti-inflammatory activity-enhanced skin permeation and stability of compound D was developed using croton oil-induced ear edema model in male ICR mice.101 More examples of herbal loaded niosome are shown in Table 1.102–106

Ethosome
It is a novel liposome that defined as a soft, non–invasive lipid-based elastic vesicles (Figure 5D) developed for topical, transdermal and systemic applications with the high efficient ability of both hydrophilic and lipophilic drugs and active ingredient delivery to deeper skin layers and blood circulation.8,10

Ethosome is composed of water, certain phospholipids (phosphatidylcholine, phosphatidylserine, phosphatidylethanolamine, and phosphatidylglycerol), and a relatively high concentration of alcohol (30–45%) (ethanol and isopropyl alcohol).109,110 This composition provides higher deformability and entrapment efficiency to ethosome that enhances topical drug delivery of highly concentrated active ingredients and transdermal transport efficiency and prolongs the physical stability of ethosomes via flexibility of the lecithin bilayer when compared to liposome.109

The disadvantage of ethosome is size growing from tens nanometers to micrometers due to its poor stability that caused by alcohol evaporation and then loaded compounds leaks out after a while. To control this shortcoming, alcohol can be situated with a combination of trehalose and propylene glycol.12

In this connection, curcumin-encapsulated PEGlycated and traditional liposomes and ethosomes were developed and tested for their potency as a transporter for the carrying of products to the skin. PEGlycated liposomes presented the most accepted ex vivo transdermal drug delivery system in rat skin and showed a higher suppression of paw edema in the rat model of induced inflammation.110 More examples of herbal loaded NE are presented in Table 1.88,111,112

Dendrimer
A dendrimer is a tree-like synthesized polymer that was characterized as having a single central core that gives frequent branches of variously armed macromolecules (external capping and multifunctional groups) (Figure 6) to achieve better targeting to specific sites. Generally, they are made up of natural or synthetic components such as sugars, nucleotides and amino acids.113,114

The unique feature of this polymer is that its structure and hydrophilicity are easily controllable during formation to get higher solubility, permeability, biocompatibility, biodistribution, clearance and consequently reducing side effects.115

Instantly, the polyamidoamine (PAMAM) dendrimers have recently been studied as carriers as they can be developed in various shapes, sizes and surfaces, in order to get functionalized nanoscale formulas.116 Thus, this dendrimer can offer to target ligands to promote particular binding to cellular receptors.117 Additionally, the small size of this dendrimer renders it to be promptly cleared from the body through the renal and escape from the reticuloendothelial system.118 Furthermore, broad internal cavities of PAMAM dendrimers allow them to complex hydrophobic drugs either by a covalent or non-covalent conjunction.119,120
In this regard, a group of researchers investigated the effectiveness of quercetin-loaded PAMAM dendrimers after oral administration as a Biopharmaceutical Classification System (BCS) class II molecule. They assessed the water solubility of quercetin in 4 generated dendrimers with 5 different concentrations. Consequently, they found that all generations with respective concentrations of PAMAM dendrimers showed potential positive effects on solubility enhancement and in vitro quercetin dual releasing pattern of an initial quicker release then sustained release. Furthermore, the efficacy of this dendrimer on a carrageenan-induced paw edema model to evaluate the acute activity of this nanocarrier in response to inflammation was also evaluated. More examples of herbal loaded NE are presented in Table 1. However, many other dendrimers such as polyamidoamine organosilicon (PAMAMOS), polypropyleneimine (PPI), and glycodendrimers have been developed and studied but with less common use.

Micelle
It is a nanosized (10–100 nm) polymer particles or colloidal dispersion (Figure 7) that consists of a single core-shell with narrow and small-sized self-assembly of synthetic amphiphilic di- or tri-block copolymers with both hydrophobic and hydrophilic segments in aqueous media. Solubilization enhancement, intracellular drug accumulation, and protection against degradation are provided by the inner hydrophobic core, while the hydrophilic layer providing improved biocompatibility and active site-specific cell targeting, as well as thermal, pH, and photosensitivity properties.

Figure 6 A schematic illustration of dendrimer.
Notes: Reproduced from ud Din F, Aman W, Ullah I, et al. Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors. Int J Nanomed. 2017;12:7291-7309

Figure 7 A schematic illustration of micelle.
Notes: Reproduced from ud Din F, Aman W, Ullah I, et al. Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors. Int J Nanomed. 2017;12:7291-7309
In order to achieve a higher accumulation of drugs at the tumor site and achieve a prolonged circulation in blood, micelles must maintain good stability in the body. This proper stability can be achieved in two paths that are dynamic stability in which the micelle being decomposed into the single polymer chain, and the other is thermodynamic stability in which depends on the anti-dilution capacity of polymeric micelles.\(^\text{131}\)

The best-studied block copolymers for use in micelle construction are polyethylene glycol (PEG)-block-poly3-caprolactone (PCL), PEG-b-polylactide-coglycolide, and PEG-b-poly-c-benzyl L-glutamate. Among them, PEG-PCL is the most preferable one due to its acceptable features such as biodegradability, safety, and high loading for lipid-soluble biomaterials.\(^\text{132}\)

In this respect, artemisinin encapsulated PEG-PCL micelle introduced with LyP-1 (cyclizine-amino acid peptide) has been developed that recognizes and binds to the p32/gC1qR receptor and consequently expressed highly in specific cancer cells of tissues and lymph vessels. This polymeric micelle modification enhances the artemisinin delivery to extremely metastasized mammary adenocarcinoma and its surrounding lymphatic tissues both in vitro and in vivo successfully.\(^\text{133}\)

Generally, polymeric micelles are penetrating the tumors via active or passive targeting mechanisms in which the latter enhanced permeability and retention effects produced after intravenous administration of particles, while active targeting depends on the basic receptor-mediated interaction between the ligand-modified on the surface of micelles and the molecular markers specifically over-expressed in the cancer cells, such as folate receptors, integrins, and epidermal growth factor receptors.\(^\text{134,135}\)

More examples of herbal loaded NE are presented in Table 1.\(^\text{136–140}\)

### Nanosphere (NS)

Nanosphere is a colloidal aqueous solution with amorphous or crystalline nature having a size range between 10 and 200 nm (Figure 8) that composed of a polymeric core encapsulating active ingredients and/or adsorbing them onto the nanoparticles.\(^\text{141,142}\)

The main virtue of this system includes delayed drug release, regular plasma drug concentrations, more stability in biological fluids, high protection from enzymatic and chemical degradation, improved bioavailability, potential antitumor efficiency, enhance complete entrapment of the drug, and reduced toxicity.\(^\text{143}\) These most outstanding features of NS are directly due to hydrophobic surfaces of these particles that are highly susceptible to opsonization and clearance by the reticuloendothelial system.\(^\text{12}\)

Biodegradable NS includes albumin NS, modified starch NS, gelatin NS, polypropylene dextran NS and polylactic acid NS. In addition, there are 2 more types of NS, immune NS and magnetic NS. Immunomagnetic NS can be prepared by combining the above two kinds of NS, which could significantly improve its targeting.\(^\text{144}\)

Most nanospheres are prepared with biodegradable, biocompatible, and synthetic polymers such as polylactic acid (PLA), polyglycolic acid (PGA), and their co-polymer polylactide-coglycolide (PLGA) using emulsion evaporation technique.\(^\text{144}\) Moreover, NS can be also prepared using pre-formed polymers by nanoprecipitation (omitting the oil in the formulation) or interfacial deposition of polymer (containing the oil). The type of polymer is also important for evaluating the rate of release by NS. Because of the small size of NS, they can be administered orally, locally, and systemically.\(^\text{12,145}\)

Interestingly, oridonin (ORI)-loaded poly(D, L-lactic acid) (PLA) modified with a functionalized polymer [RGD (Arg-Gly-Asp peptides)] to improve antitumor activity is generated that comes with tissue targeting, and better in vivo tumor inhibitory effects than oridonin alone.

![Figure 8 A schematic illustration of nanosphere.](image)

**Notes:** Reproduced from Harper 3D.\(^\text{142}\)
or ORI loaded PLA nanoparticles. More examples of herbal loaded NE are presented in Table 1.

Nanocrystals

They are pure drug crystals with nanosized particles (Figure 9) in which toxic side effects resulting from the encapsulating/solubilizing excipients may be eliminated. The best known features of nanocrystals are high drug-loading capacity and platform stability that render them to be widely used to deliver poorly hydrophilic materials in the form of a colloidal dispersion. Cells of the mononuclear phagocytic system can recognize the nanocrystal particles in the bloodstream as an exogenous material which results in passive accumulation in liver, spleen, and lung.

Nanocrystals are generally developed by the “bottom-up” (such as Nanomorph™, Soliqs/Abbott), “top-down” technologies (such as Dissocubes®, SkyePharma) or combination technologies (such as NANOEDGE, Baxter). However, the top-down technique is the procedure of choice for nanocrystals in products developed in the pharmaceutical, cosmetic or clinical trials mainly due to the simple and easy scale-up which better serves the industry.

**Figure 9** A schematic illustration of fenofibrate nanocrystals (FNB-NCs).

**Notes:** Reproduced from Kevadiya BD, Chen L, Zhang L, Thomas MB, Davé RN. Fenofibrate Nanocrystal Composite Microparticles for Intestine-Specific Oral Drug Delivery System. Pharmaceuticals. 2019;12(3):109-124

**Figure 10** A schematic illustration of phytosome.

**Notes:** Reproduced Karthivashan G, Masarudin MJ, Kura AU, Abas F, Fakurazi S. Optimization, formulation, and characterization of multiflavonoids-loaded flavanosome by bulk or sequential technique. Int J Nanomed. 2016;11:3417-3434.
The nanocrystals are made by wet milling methods, such as bead milling or high-pressure homogenization technique that improves the oral bioavailability and enhanced the transdermal efficacy of poorly soluble drugs.\textsuperscript{153} Physically, the biodistribution of nanocrystals is affected by particle size, morphology and surface modification. Additionally, in order to target nanocrystals to specific pathogenic sites, ligand conjugation and stimuli-responsive polymers can be used.\textsuperscript{154}

In 2011, a group of researchers prepared a natural product derived nanocrystal focused on Camptothecin (CPT) active compound that was isolated from the bark and stem of \textit{Camptotheca acuminata}, a tree native to China used as a cancer treatment in Traditional Chinese Medicine.\textsuperscript{155} In their study, they examined the particle characteristics, cellular cytotoxicity, and animal anticancer effect. Finally, they realized that CPT nanocrystals were more potent to MCF-7 human breast cancer cells than CPT alone in vitro. Additionally, CPT nanocrystals exhibited significant suppression of tumor growth in MCF-7 xenografted BALB/c mice model and the drug concentration in the tumor site was 5 times more at 24 hrs by using the nanocrystal treatment than by using the CPT solution. Storage stability study indicated that the nanocrystals were stable for at least 6 months.\textsuperscript{155} More examples of herbal loaded NE are presented in Table 1.\textsuperscript{156–163}

**Phytosome or Herbosome**

Water-soluble phytochemicals such as flavonoids and polyphenols are poorly absorbed in the body due to their large

---

**Figure 11** A schematic illustration of APC-SNEDDS dissolved in distilled water. APC: Akebia saponin D-phospholipid complex.

**Notes:** Reproduced from Shen J, Bi J, Tian H, et al. Preparation and evaluation of a self-nanoemulsifying drug delivery system loaded with akebia saponin D–phospholipid complex. \textit{Int J Nanomed}. 2016;11:4919-4929.\textsuperscript{183}
molecular size which did not allow them to be absorbed by passive diffusion, as well as their poor lipid solubility makes a serious limiting to their pass across the lipid-rich biological membranes, subsequent poor bioavailability.

Phytosome is a patented formula developed to incorporate medicinal plant active ingredients and water-soluble phytochemicals into phospholipids to create lipid-compatible molecular complexes in order to immensely modify their absorption and bioavailability. The novelty of phytosome formulation is that there is a molecular complex and chemical bonding between phosphatidylcholine and the plant materials at a ratio of either 1:1 or a 2:1 relying on the substance(s) conjugated, whereas no chemical bonds are formed in liposome and thousands of phosphatidylcholine molecules enclosing the water-soluble compound can be observed freely.

Phytolipid delivery system is a specifically modified phytosome for delivering of herbal drugs that made by incorporation of standardized plant extracts or water-soluble phytochemicals into phospholipids to produce lipid-compatible complexes to enhance better absorption and bioavailability without resorting the pharmacological or structural changes of the ingredients.

The phytosomes are small-sized particles (Figure 10) that produce better transiting from a water-soluble condition into the lipid-soluble condition of the enterocyte cell membrane and then into the cell, lastly arriving the blood and protecting the valuable ingredients of the herbal drug from gastric enzyme destruction and gut bacteria.

The recently produced phytosome-loaded herbal content is optimizing a sequential technique by a group of researchers from Malaysia to encapsulate several flavonoids in a single phytosome that named flavonosome. Three widely constituted and therapeutically valuable flavonoids named quercetin (Q), kaempferol (K), and apigenin (A) were tested in the ethyl acetate fraction of *Moringa oleifera* leaf extract and encapsulated in a single flavonosome (QKA–phosphatidylcholine) via 4 various techniques. After checking for many physicochemical properties, they suggested that this three-in-one flavonosome with sustained activity is a good candidate as an antioxidant, hepatoprotective, and heat supplement agent. More examples of herbal loaded NE are presented in Table 1.

Self Nanoemulsifying Drug Delivery System (SNEDDS) SNEDDS is a lipid-based anhydrous isotropic mixture of oil, surfactant(s) and cosurfactant(s) with a particle size of 20–200 nm (Figure 11). It produces fine oil-in-water nanoemulsions upon gentle agitation after dilution in aqueous media, such as gastrointestinal fluids; thus, it can be given orally in soft or hard gelatin capsules. This leads to in situ solubilization of drug that can subsequently be absorbed by lymphatic pathways, bypassing the hepatic first-pass effect. On the other hand, efforts were made to overcome the limited aqueous solubility, low ocular bioavailability and short pre-ocular retention and absorption of drugs by introducing SNEDD in the form of eye drop. Physically stability upon storage, easy to produce, improved dissolution rates and absorption that results in more reproducible blood–time profiles are among the most accepted features of SNEDDS. Additional advantages of SNEDDS over conventional emulsions and other lipid carriers are the significantly reduced energy requirement for their preparation and easier to manufacture in a large scale.

Among the successful example of previously prepared crude plant extract-loaded SNEDDS is a persimmon (*Diospyros kaki*) leaf extract (PLE) loaded SNEDDS that was characterized to compare its in vitro dissolution and relative bioavailability with a commercially available agent (Naoxingting tablets). They indicated that this developed formula shows better stability, solubility and sustained release than the commercial drug, as well as it is a promising drug delivery system for increasing the oral bioavailability following oral administration in fasting beagle dogs. Recently, several novel herbal loaded SNEDDSs with desirable properties have been reported and are presented in Table 1.
Self Microemulsifying Drug Delivery System (SMEDDS)

SMEDDS is a lipid-based nanoparticle that is composed of oil, surfactant (Cremophor RH40, Cremophor EL, or Polysorbate 80) and co-surfactant (Figure 12). The role of surfactant in SMEDDS is to improve intestinal permeability by lowering surface tension and hence facilitating touch with gastrointestinal mucosa and additionally inhibiting drug efflux by P-glycoprotein.186,187

Among the most preferred property of SMEDDS is bioavailability improvement due to its small particles and wide surface area, which ameliorate absorption, solubilization, and releasing capacity. Additionally, SMEDDS decreases the first-pass metabolism by facilitating drug absorption via the lymphatic system of the intestine, and thus it provides a promising way to raise bioavailability for poorly hydrophilic products. Moreover, SMEDDS is very stable, easy to administer, and easy to construct at industrial scale especially solid SMEDDS.188

SMEDDS can produce microemulsions, nanoemulsions, or emulsions followed by injection into aqueous media with mild agitation that may develop drug
precipitation. In order to overcome this phenomenon, a super-saturable self-micro emulsifying drug delivery system (S-SMEDDS) is developed that contains precipitation inhibitor with good crystallization-inhibiting capacity such as Polyvinylpyrrolidone (PVP) K17.\textsuperscript{189} Instantly, ligand-modified SMEDDS was reported with targeted delivery of active compounds to specific absorption sites such as targeting of folate receptor over-expression in colorectal carcinoma by producing folate-modified SMEDDS.\textsuperscript{190}

In this connection, resveratrol, a poorly hydrophilic component isolated from some commonly desired fruits loaded SMEDDS is generated for oral delivery. This novel SMEDDS is also characterized by maintaining high drug solubilization for long period to improve drug absorption, improved bioavailability and provided more stability due to its small particle size (approximately 50 nm) and high zeta potential in a neutral environment. The antioxidant capacity and cytotoxicity of the formulation were also detected using DCFH-DA and CCK-8 assays. The formulation exhibited a greater antioxidant capacity with less toxicity than a free compound.\textsuperscript{191} More examples of herbal loaded NE are presented in Table 1.\textsuperscript{192–196}

**Nanofiber**

It is composed of solid polymer fibers with diameters of 10–1000 nm that have a large surface area with a small pore size (Figure 13) and is prepared by the electrospinning method.\textsuperscript{12} This novel nanocarrier has a limited role in delivering active components but with potential improvements in the therapeutic treatments and support the using of active compounds in several biomedical areas such as tissue regeneration.\textsuperscript{197,198}

The nanofibers are most likely carbon-based as they are extracted from various plants and thus they can be generated from different polymers and hence have different physical properties and application potentials.\textsuperscript{199} Among the potential benefits of the nanofibers is to modify wound healing and preventing infection. Also, it is suggested that nanofibers have very strong adhesive features such as that is found with a gecko that allows it to easily climb surfaces using bundles of nanofibers on the surface of its feet. Moreover, the scaffolding of nanofibers to initiate the repair of damaged tissue is among the pronounced features.\textsuperscript{200}

Regarding the anticancer potential of nanofiber, curcumin-loaded self-assembling peptide nanofiber as a novel tumor-targeting carrier was developed that showed high cellular uptake in αvβ3 integrin-positive HepG2 liver carcinoma cells, thereby leading to significantly higher cytotoxicity than nonloaded one. Additionally, ex vivo studies further demonstrated that curcumin could accumulate markedly in mouse tumors after administration via the tail vein.\textsuperscript{201,202} More examples of herbal loaded nanofibers are displayed in Table 1.\textsuperscript{203–207}

**Polymersome (PS)**

Polymersome is a self-assembled polymeric nanosphere vesicle that may have relatively thick membranes (up to
40 nm), which are formed by synthetic amphiphilic block copolymers (Figure 14). They are able of incorporating hydrophilic and nonhydrophilic drugs, proteins, peptides, DNA and RNA fragments in their membrane which acts as a barrier to protect them from the biological environment. Additionally, the membrane flexibility of PS makes them applicable in targeting and control release.

Polymersomes have some similarities to liposomes especially in the structure but are more stable and less permeable than liposomes. The PS is capable to bind with biologically active ligands, antibodies and biotinylated conjugation to their surface which enhances targeted therapy and imaging strategy.

It was documented that PS was used as anti-tumor agents for several drugs due to its controlled release, high permeation, retention and loading capacity of drugs into PS than liposome membrane. In this respect, biotin functionalized leuko-polymersome to proctor and treat inflammation, cancer, and cardiovascular disease and Tat-loaded PS as a fantabulous agent for cellular tracking were also investigated as a promising tumor-fighting agent. More examples of herbal loaded PS are found in Table 1.

Cubosome

It is a viscous isotropic vesicle that made mainly of unsaturated monoglycerides (monoolein-water) binary system with thermodynamically steric stable bi-continuous cubic liquid crystalline phase (poloxamers) (Figure 15).

Features such as high internal surface area per unit volume (approximately 400 m²/g) and a 3D structure with hydrophilic and hydrophobic domains make them entrap water-soluble and nonsoluble and amphiphilic materials successfully. Its large interfacial area can provide a complex diffusion pathway for sustained release of entrapped drug molecules, whereas lipid constituents are biocompatible, bio-adhesive, and digestible. They are usually constructed via dispersion or fragmentation of the cubic phases of gels in a liquid condition. Previous works on somatostatin, insulin, indomethacin, and rifampicin drug encapsulation within cubosomes have been done intensively. Additionally, various pharmaceutical applications of cubosomes have also been investigated such as peptides, enzymes, antimuscarnic drugs, antibiotics, and analgesic delivery.

Cubosomes easily evacuate their contents to the epidermis as they have an almost same structure to that of the stratum corneum, as well as the properties of adhesion and penetration enhancement of cubosomes suggest their potential utility in skin cancer (melanoma) treatment. On the highlight of this, very recently, a report of biocompatible polymer-free cubosomes for potential application in both photodynamic therapy and bioimaging of skin malignant melanoma has been published with very low cytotoxicity to the cutaneous formulation. More examples of herbal loaded cubosome are shown in Table 1.

Biopolymer-Based Nanocarrier (BBN)

They have derived from proteins (such as gelatin, albumin, and milk proteins), polysaccharides (such as chitosan, hyaluronan, dextran, cyclodextrins, pectin, guar gum, cellulose, sodium alginate, and starch), and/or their modified versions, derivatives or their combinations. The most interesting features of these materials that render them to be used for BBN...
production are a biological realization, bioactivity, biodegradability, less toxicity, easy modification, and simplicity of producing gels from them.\textsuperscript{230} This type of nanocarrier is well established to have plausible water solubility, stability, degradation, and biocompatibility for a wide range of utilization that earned from their variable charges, molecular weights, and compositions.\textsuperscript{9,12} Various approaches for fabrication of BBN delivery systems are well addressed which including coacervation, spray drying, electrospinning, electrospray, supercritical fluid, emulsion–diffusion, reverse micelle, emulsion-droplet coalescence, emulsification/solvent evaporation, salting-out, ultrasonication and high-pressure homogenization.\textsuperscript{231}

Pure Biopolymer Nanoparticles (PBN)

Chitosan, a cationic biocompatible and biodegradable linear polysaccharide, containing d-glucosamine and N-acetyl glucosamine units, which is extracted from the exoskeleton of crustacean arthropods such as insects, crabs, lobster and shrimps\textsuperscript{232} reported to be the best example of natural pure biopolymer to deliver plant components such as curcumin, quercetin,\textsuperscript{233} and transresveratrol with better mucoadhesion, solubility, dissolution rate, and specific targeting.\textsuperscript{234,235} Additionally, chitosan has been utilized for various biomedical applications such as in tissue engineering in the form of scaffolds, drug delivery carriers, fabricating surgical thread, bone healing, and as a wound dressing substance. On the other hand, modified chitosan molecules such as dextran sulfate-conjugated chitosan, biotinylated and galactosylated chitosan are well developed with advanced properties such as altering the surface charge, providing pH-sensitive swelling, more stability, enhancing bioactivity by modifying targeting to the specific site of action (Figure 16).\textsuperscript{236} More examples on this nanoparticle are available in Table 1.\textsuperscript{237–241}

Figure 18 A schematic illustration of biopolymeric drug conjugate.

Notes: Reproduced from Safer AM, Leporatti S, Jose J, Soliman MS. Conjugation Of EGCG And Chitosan NPs As A Novel Nano-Drug Delivery System. Int J Nanomed. 2019;14:8033-8046.\textsuperscript{333}
Biopolymer-based Hydrogels (BBH)

Hydrogels are cross-linked polymeric networks with hydrophilic functionalities that supply spaces for homing aqueous biological fluids (Figure 17) and have is known as a promising bio-compatible material in numerous therapeutic applications. This formula is characterized by adorable biocompatibility, high porosity, hydrophilicity that results in controlled drug release. Naturally available biopolymers such as chitosan, alginate, hyaluronic acid (HA), collagen, and gelatin are used to construct inherently biodegradable BBH that frequently pre-

![Citrate](image)

**Figure 19** A schematic illustration of gold nanoparticle.
**Notes:** Reproduced with permission from Luna Nanotech. 260

functionalized to integr binding sites permitting for adherence and integrated cellular responses. However, the application of these substances is somewhat restricted because significant batch-to-batch variability and potential immunogenicity within foreign models are obtained. 243

In this connection, biopolymer-based pH-sensitive hydrogels were prepared using chitosan with PEG of different molecular weights in the presence of silane crosslinker. The incorporated components remain undissolved in different swelling media as they are connected by siloxane linkage which was confirmed by FTIR spectroscopy. The swelling in water was enhanced by the addition of higher molecular weight PEG. The swelling behavior of the hydrogels against

![Images](image)

**Figure 21** A schematic illustration of magnetic nanoparticle.
**Notes:** Adapted with permission from Frontier in Microbiology. Souza AC, Amaral AC. Antifungal therapy for systemic mycosis and the nanobiotechnology era: improving efficacy, biodistribution and toxicity. Front Microbiol. 2017;8(336):1–13. 280

**Figure 22** A schematic illustration of veteran cockle shell-derived calcium carbonate nanoparticles.
**Notes:** Reproduced from Muhammad Mailafy M, Abubakar K, Danmaigoro A, et al. Cockle Shell-Derived Calcium Carbonate (Aragonite) Nanoparticles: A Dynamite to Nanomedicine. Appl Sci. 2019;9(14):2897-2922. 334

---

![Images](image)

**Figure 20** A schematic illustration of silica nanoparticle.
**Notes:** Reproduced from ud Din F, Aman W, Ullah I, et al. Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors. Int J Nanomed. 2017;12:7291-7309. 8

---

Dovepress

International Journal of Nanomedicine 2020:15

submit your manuscript | www.dovepress.com

Dovepress

2469

Dovepress

Dovepress
pH showed high swelling in acidic and basic pH, whereas, the low swelling was examined at pH 6 and 7. This characteristic pH-responsive behavior at neutral pH made them suitable for injectable controlled drug delivery. More examples on this nanoparticle are available in Table 1.

Biopolymer Drug Conjugate (BDC)
Thermally sensitive biopolymer with the potential ability to quickly form insoluble viscous co-acervate at body temperature can be used for this purpose (Figure 18). Although some of the biomedical polymer–drug conjugates are approved for clinical trials, they lack photothermal properties and multi-imaging capabilities, impeding them from imaging-guided precision cancer therapy and total cancer arrested development. Thus, researchers introduced a novel all-in-one biopolymer–drug conjugate nanotheranostics, such as that of intracellular pH-sensitive polydopamine–doxorubicin conjugate nanoparticles under a mild situation that are characterized by excellent photothermal attribute, dual stimuli-triggered drug release activity, and about elongated blood circulation time than nonconjugated doxorubicin. More examples on this nanoparticle are available in Table 1.

Inorganic Nanocarrier
Recently, different kinds of inorganic nanocarriers have been developed and investigated for their potential delivery of plant active ingredients.

Metal Nanoparticle (MN)
These are nanoparticles such as silver nanoparticles (AgNPs), gold nanoparticles (AuNPs) (Figure 19), copper nanoparticles (CuNPs), zinc oxide (ZnONPs) nanoparticles, quantum dots, cerium oxide (CeO₂) nanoparticles, iron oxide nanoparticles (Fe₂O₃), yttrium oxide (Y₂O₃) nanoparticles and titanium dioxide nanoparticles (TiO₂) possess special benefits in biomedical application due to their contents of essential mineral elements that have strong activity for human body. These nanoparticles gained their noncovalent interaction or covalent conjugation drug-loading capacity due to their surface plasmon resonance (SPR) ability, structural diversity, poor toxicity, and high biocompatibility. Thus, they can be utilized for achieving intracellular drug delivery and controlled release through a photothermal route.

Moreover, a novel metal nanoparticle with multi-functional groups is also investigated by developing much active component-loaded complex metal nanoparticle integrated multifunctional liposomes to improve intracellular drug delivery, overwhelm multi-drug resistant (MDR), fasten anti-tumor activity, and lower side effects. Very recently, it has proven that biopolymers complexed with bioactive nanoparticles endowing antimicrobial and anti-inflammatory properties have a fantastic effect in wound care to prompt the healing mechanism of wound infections caused by hyperglycemia.

In this regard, a combination of antibacterial nanoparticles such as silver, gold, or copper nanoparticles with polymeric matrix could potentially suppress bacterial propagation and similarly fastens the healing process of a wound and mitigate the diabetes mellitus-based foot ulcer. More examples on this nanoparticle are available in Table 1.

Mesoporous Silica Nanoparticle (MSN)
MSN is the most recent promising carrier for drug storage and delivery that has large surface area with high loading capacity for therapeutic agents, high pore volume and porosity (honeycomb-like architecture), adjustable pore diameter, modifiable surface potential, selective surface functionality, morphology control, desirable biocompatibility and controlled release properties (Figure 20).

Moreover, MSN has been applied in pharmaceuticals to improve drug bioavailability, reduce drug toxicity, and deliver with cellular targetability. Particularly, the exciting progress in the development of MSN-based effective delivery systems for poorly soluble drugs, anticancer agents, and therapeutic genes.

In general, MSNs are synthesized by using a silica precursor (tetraethylorthosilicate or sodium silicate) in an alcoholic solution under basic conditions and incorporating a surfactant. On the other hand, the synthesis of mesoporous silica particles in the nonalcoholic medium was conducted but the formation of spherical particles is limited by the amount of the surfactant.
The developed formula in this area is silybin from the seed of the milk thistle (*Silybum marianum*)-meglumine encapsulated MSN with high drug-loading capability, in vitro, sustained release, and in vivo high absorption ability. More examples on this nanoparticle are available in Table 1.

Magnetic Nanoparticle (MNP)

Attributing with the use of developed magnetic nanoparticle (especially iron oxide) owing to receive the highest target positioning and best trigger drug release, biocompatibility, and nontoxicity in a magnetic field (Figure 21). Magnetic nanoparticles with appropriate surface coatings are used clinically for various biomedical applications, such as magnetic resonance imaging, hyperthermia, drug delivery, tissue repair, cell, and tissue targeting and transfection. Other benefits of using magnetic nanocarriers are referred to be more rapid and effective for curing diseases even if a small amount of drug is consumed; thus, it can reduce the concentration of the drug in healthy tissues, and consequently diminished side effects. Moreover, MNP small size renders them to gain more bioaccessibility to deserted tissues and bio interact with them at molecular and cellular levels, also binding to particular tumor-suppressor antibodies and conveying these adsorbed anti-tumor materials to the site of the tumor. In this respect, gambogic acid (from the brownish or orange resin from *Garcinia hanburyi*)-loaded magnetic iron oxide (*Fe₃O₄*) nanoparticles were produced and investigated for its improvement in the water solubility of gambogic acid and halting the proliferation and migration.
of Panc-1 pancreatic cells by inactivating transcription factor ETS1 in vitro using MTT and scratch assays, respectively. More examples on this nanoparticle are available in Table 1.

Calcium Carbonate (CaCO₃) Nanoparticle

Aragonite CaCO₃ nanoparticle is less stress to achieve, environmentally pleasant, and less costly process that involved a simply automated stirring of cockle shell powder in the occurrence of BS-12 as a biomineralization catalyst (Figure 22). CaCO₃ is utilized as therapeutic agents with outstanding efficacy due to its biocompatibility, nontoxicity, changeable surface chemistry, excellent physicochemical property, simple preparatory methods in a bulk scale, controlling release, slow biodegradability, pH-sensitivity, and porous nature.

CaCO₃ potential to be functionalized with targeting agents gives it the distinctive property that can be used in targeted delivery systems for anticancer drugs, in addition to slow CaCO₃ matrix degradation, constant and organized discharge property, controlled release, at the targeted location.

The best example in this area is doxorubicin-loaded CaCO₃-NPs for cancer therapy. Generally, the physiological pH of blood and the extracellular spaces around tumors is about 6.8–7.2, while the pH of endolysosomes of cancer cells is highly acidic (pH <6). Then, CaCO₃ nanoparticles holding DOX are swiftly unprotected to the acidic environments of endosomes, loss its stability, and is believed to discharge drugs at lesser pH, which results in a rise in the cellular uptakes of drugs. Unfortunately, until this moment plant metabolite loaded CaCO₃ nanoparticles are not available in the research area.

Nanotube

Halloysite Clay Nanotubes (HNT)

They are aluminosilicate clay that constructed from 2 different dimensional structures (tetrahedral and octahedral) through surface weathering of aluminosilicate and composed of aluminum, silicon, oxygen, and hydrogen (Figure 23). These hollow tubes with several nanocavities are best known for having the high surface area, biocompatibility and loading capacity.

Halloysite nanotubes are natural green cylindrical clays that are not costly, not difficult to collect, and possessing chemical composition similar to that of kaolin. Features such as proper lumens, high aspect length–diameter ratio and low hydroxyl density on their surface make them be more adjustable to be used for many projects.

Continuing in this area, resveratrol (from berry family)-loaded halloysite nanotube coated layer-by-layer with polyelectrolytes in order to control its drug release ability and to reduce its toxicity is developed successfully. Additionally, the system showed enhanced resveratrol cytotoxicity to MCF-7, breast cancer cells and produced pronounced apoptosis. More examples on this nanoparticle are available in Table 1.

Carbon Nanotubes (CNT)

CNTs are allotropes of carbon with a cylindrical nanostructure that have unusual properties, which are valuable
for nanotechnology, electronics, and optics with some remarkable properties such as excellent thermal conductivity, mechanical strength, and electrical conductivity (Figure 24).303,304

One of the most recently developed nanodevices for biomarker detection is CNT in which a single-walled CNT as a high-resolution atomic force microscopy (AFM) for the selective detection of specific sequences of kilobase-size DNA from single-base mismatch sequences was used.305 The principle of this technique is hybridizing of targeted DNA fragments with labeled oligonucleotides to be easily detected by AFM. This method enabled the straight detection of genetic disorders causing cancers that encoded by specific haplotypes. Additionally, this model can be utilized as nanoscale carriers for bioimaging, drug delivery and used for photothermal destruction of cancer cells.306

In this connection, freshly prepared Ocimum tenuiflorum (tulsi extract) mediated photosynthesized AgNP loaded into emulsiﬁed multiwalled carbon nanotube (MWCNT) was developed that characterized by a spherical shape, 5–40 nm size and surface plasmonic resonance at 430 nm. Their targetability to the intracellular part of the sperm cell (without disrupting the sperm cell membrane) for its further application in biosensing-based infertility diagnosis was also investigated in detail and conﬁrmed that AgNP-MWCNT composite is suitable in fertility diagnosis and reproductive health care.307

Hybrid Nanocarrier (HNC)

This type of nanocarrier is composed either from the combination of 2 different organic materials or a combination of an organic with an inorganic material to emerge improved drug delivery performance. Many HNC possesses a core-shell structure that composed of different types of biomaterials (Figure 25).308,309 This composition endows the HNC with many desirable properties, such as high encapsulation and loading ability, the betterment of stability, sustained release, improvement of intracellular drug delivery, and the enhancement of conjugating with targeting ligands.295 On the other hand, speciﬁc functionalities of organic materials at the surface of inorganic materials can be promoted to improve the selectivity and efﬁciency of therapy especially those utilized as anticancer agents.310 In this respect, hybrid nanocarrier conjugated folic acid for targeted letrozole (LTZ) delivery for breast cancer treatment was produced in which physicochemical properties, in vitro in vitro drug release, cytotoxicity and ex vivo work of the formula were studied intensively. As a result, the system could overwhelm the restrictions related to the LTZ as a potent nonsteroidal drug. Finally, it was concluded that both the entrapment and therapeutic efﬁciency of LTZ in the amphiphilic carrier were enhanced using the lipid nanoparticles and the surface modification, respectively.311 More example of this nanoparticle is available in Table 1.312

Biological Nanocarriers (BNC)

These are naturally occurring highly diverse nanoparticles that are shared a common structure of a shell composed of capsid proteins surrounding the DNA or RNA viral genome. They have variable sizes (within the nanometer range) and morphologies from simple spheres to rods to icosahedrons (Figure 26).313,314 In this respect, viruses have been dedicated to targeting the most pronounced organisms and tissues. Most applications use virus nanoparticle (VNP) and virus-like particles (VLP) are native viral capsid proteins without nucleic acid in order not to cause infection.299 VNP and VLP are nanosized (approximately 100 nm), self-assembled robust protein net that possessing uniform nanostructures and distinct geometry. Recently, this system is used for many purposes such as drug delivery and gene therapy in the form of nanoreactors, filamentous or spherical scaffolds.12

Modified VNP and VLP are also utilized in vivo for vaccination either to induce immunity against the parent virus or to modify other diseases. Additionally, VLP conjugated to appropriate epitopes have been used for anti-tumor vaccines and for vaccines against chronic diseases such as hypertension.315 Regarding their use in bioimaging system, VLP has also been adapted for use as contrast agents in MRI and PET. Furthermore, the natural afﬁnity of VLP for deﬁned cell types allows targeted delivery, as well as the VLP, can be modiﬁed by conjugation to targeting molecules, such as folic acid, for cell-speciﬁc delivery.316 More examples of herbal loaded BNC are presented in Table 1.317–321

Conclusion

Poor solubility in water and bioavailability have limited the therapeutic efﬁcacy of naturally available potential natural plant products. Currently, recent studies have attempted to address these problems using nanocarriers and studies have anticipated that nanomedicine as a plausible approach for diagnosis, imaging, and therapeutics for a variety of disease treatment and management including cancer, diabetes, hyperglycemia, hypertension, and anemia. Currently, several nanocarrier-encapsulated natural plant extract formulations are in clinical or preclinical
development and some of them are already approved by the Food and Drug Administration (FDA) to be used safely in human especially those that are already confirmed that they do not have potential long-term toxicity, degradation, and incomplete metabolism after mitigated by the concept of modifications and piolot study that could be developed as an inexpensive, safe, tolerable, and an appropriate approach for disease control and management.

**Acknowledgement**

The authors appreciate a grant that has been funded by a Research University Grant Scheme (RUGS) (Project No. UPM/700-2/1/GPBI/2017/9554100) provided by Universiti Putra Malaysia (UPM), Malaysia.

**Disclosure**

The authors declared that there is no conflict of interest in this current review article.

**References**

1. Krishnaiah YS. Pharmaceutical technologies for enhancing oral bioavailability of poorly soluble drugs. *J Bioequivalence Bioavailab.* 2010;2(2):28–36. doi:10.4172/jbb.1000027
2. Teeranachaideekul V, Müller RH, Junyaprasert VB. Encapsulation of acesrol palmitate in nanostructured carrier lipids (NLCs) effects of formulation parameters on physicochemical stability. *Int J Pharm.* 2007;340(1–2):198–206. doi:10.1016/j.ijpharm.2007.03.022
3. Siddiqui IA, Sanna V. Impact of nanotechnology on the delivery of natural products for cancer prevention and therapy. *Mut Nutr Food Res.* 2016;60(6):1330–1341. doi:10.1002/mnfr.201600035
4. Aql F, Munagala R, Jeyabalan J, Vadhanam MV. Bioavailability of phytochemicals and its enhancement by drug delivery systems. *Cancer Lett.* 2013;334:133–141. doi:10.1016/j.canlet.2013.02.032
5. Adhani VM, Mukhtar H. Human cancer chemoprevention: hurdles and challenges. *Top Curr Chem.* 2013;329:203–220.
6. Bharali DJ, Siddiqui IA, Adhani VM, et al. Nanoparticle delivery of natural products in the prevention and treatment of cancers: current status and future prospects. *Cancers.* 2011;3(4):4024–4045. doi:10.3390/cancers3044024
7. Wang S, Su R, Nie S, et al. Application of nanotechnology in improving bioavailability and bioactivity of diet-derived phytochemicals. *J Nutr Biochem.* 2014;25:363–376. doi:10.1016/j.jnutbio.2013.10.002
8. ud Din F, Aman W, Ullah I, et al. Effective use of nanocarriers as drug delivery systems for the treatment of selected tumors. *Int J Nanomed.* 2017;12:7291–7309. doi:10.2147/IJN.S146315
9. Saraf A. Applications of novel drug delivery system for herbal formulations. *Fitoterapia.* 2010;81(7):680–689. doi:10.1016/j.fitote.2010.05.001
10. Ni S. Nanoparticles carrying natural product for drug delivery. *J Drug Delivery Ther.* 2017;7(3):73–75. doi:10.22270/jdtt.v7i3.1425
11. Nagalingam A. Drug delivery aspects of herbal medicines. *Jpn Kampo Med Treat Common Dis Focus Inflammation.* 2017;17:143.
12. Liu Y, Feng N. Nanocarriers for the delivery of active ingredients and fractions extracted from natural products used in traditional Chinese medicine (TCM). *Adv Colloid Interface Sci.* 2015;221:60–76. doi:10.1016/j.cis.2014.04.006
13. Lin CH, Chen CH, Lin ZC, Fang JY. Recent advances in oral delivery of drugs and bioactive natural products using solid lipid nanoparticles as the carriers. *J Food Drug Anal.* 2017;25(2):219–234. doi:10.1016/j.jfda.2017.02.001
14. Yingchoncharoen P, Kalinowski DS, Richardson DR. Lipid-based drug delivery systems in cancer therapy: what is available and what is yet to come. *Pharmacol Rev.* 2016;68(3):701–787.
15. Luo CF, Yuan M, Chen MS, et al. Pharmacokinetics, tissue distribution and relative bioavailability of paeruquin solid lipid nanoparticles following oral administration. *Int J Pharm.* 2011;410:138–144. doi:10.1016/j.ijpharm.2011.02.064
16. Rostami E, Khashanian S, Azandaryani AH, Faramarzi H, Dolatabadi JE. Omidfar K. Drug targeting using solid lipid nanoparticles. *Chem Phys Lipids.* 2014;18:56–61. doi:10.1016/j.chemphyslip.2014.03.006
17. Luo CF, Hou N, Tian J, et al. Metabolic profile of paeruquin in rats after intragastric administration of paeruquin solid lipid nanoparticles. *Int J Nanomed.* 2013;8:933–940. doi:10.2147/IJN.S39349
18. Zhang C, Gu C, Peng F, et al. Preparation and optimization of triptolide-loaded solid lipid nanoparticles for oral delivery with reduced gastric irritation. *Molecules.* 2013;18(18):13340–13356. doi:10.3390/molecules181113340
19. Dang YJ, Zhu CY. Oral bioavailability of cantharidin-loaded solid lipid nanoparticles. *Chin Med.* 2013;8:1–6. doi:10.1186/1749-8546-8-1
20. Madan J, Pandey RS, Jain V, Kataro OP, Chandra R, Katyal A. Poly (ethylene)-glycol conjugated solid lipid nanoparticles of nosophine improve biological half-life, brain delivery and efficacy in glioblastoma cells. *Nanomedicine.* 2013;9(4):492–503. doi:10.1016/j.nano.2012.10.003
21. Li J, Guo X, Liu Z, et al. Preparation and evaluation of charged solid lipid nanoparticles of tetrancrid for ocular drug delivery system: pharmacokinetics, cytotoxicity and cellular uptake studies. *Drug Dev Ind Pharm.* 2014;40:980–987. doi:10.3109/03639045.2013.795582
22. Rahman HS, Rasheedee A, How CW, et al. Zerumbone-loaded nanostructured lipid carriers: preparation, characterization, and antileukemic effect. *Int J Nanomed.* 2013;8:2769–2781. doi:10.2147/IJN.S45313
23. Das S, Ng WK, Tan RB. Are nanostructured lipid carriers (NLCs) better than solid lipid nanoparticles (SLNs): development, characterizations and comparative evaluations of clamitraumzole-loaded SLNs and NLCs? *Eur J Pharm Sci.* 2012;47(1):139–151. doi:10.1016/j.ejps.2012.05.010
24. Muhammad HS. Anti-Leukemic Effects of Zerumbone Nanoparticle on Human Jurkat T Lymphoblastoid Cell Lines In vitro and Marine Leukemic WEHI-3B Model In vivo [Doctoral dissertation]. Universiti Putra Malaysia; 2014.
25. Shangguan M, Feng Q, Zhao J, et al. Binary lipids-based nanostructured lipid carriers for improved oral bioavailability of silymarin. *Food Chem Toxicol.* 2012;50:1460–1467.
26. Nahr FK, Ghanbarzadeh B, Hamishehkar H, Kafis HS. Food grade nanostructured lipid carrier for cardamom essential oil: preparation, characterization and antimicrobial activity. *J Funct Foods.* 2018;40:1–8. doi:10.1016/j.jff.2017.09.028
27. Rahman HS, Rasheedee A, Abdul AB, et al. Zerumbone-loaded nanostructured lipid carrier induces G2/M cell cycle arrest and apoptosis via mitochondrial pathway in a human lymphoblastic leukemia cell line. *Int J Nanomed.* 2014;9:527–538. doi:10.2147/IJN.S43436
28. Ng WK, Saiiful Yazan L, Yap LH, et al. Thymoquinone-loaded nanostructured lipid carrier showed enhanced anticancer activity in 4T1 tumor-bearing mice. *Nanomedicine.* 2018;13(13):1567–1582. doi:10.2217/nmn-2017-0322
30. Nordin N, Yeap SK, Zamberi NR, et al. Characterization and toxicity of citral incorporated with nanostructured lipid carrier. Peer J. 2018;6:e3916. doi:10.7717/peerj.3916
31. Mohamad NE, Abu N, Rahman HS, et al. Nanostructured lipid carrier improved in vivo anti-tumor and immunomodulatory effect of zerumbone in 4T1 challenged mice. RSC Adv. 2015;5(28):22066–22074. doi:10.1039/C5RA0144G
32. Hosseinpour M, Abdul AB, Rahman HS, et al. Comparison of apoptotic inducing effect of zerumbone and zerumbone-loaded nanostructured lipid carrier on human mammary adenocarcinoma MDA-MB-231 cell line. J Nanomater. 2014;2014:1–10. doi:10.1155/2014/742738
33. Rahman HS, Rasoeae A, How CW, et al. Antileukemic effect of zerumbone-loaded nanostructured lipid carrier in WEHI-3B cell-induced murine leukemia model. Int J Nanomed. 2015;10:1649–1666. doi:10.2147/SN67113
34. Rahman HS, Rasoeae A, Othman HH, et al. Acute toxicity study of zerumbone-loaded nanostructured lipid carrier on BALB/c mice model. Biomed Res Int. 2014;2014:1–15.
35. Jia Ning F, Gayathri TS, Rahman HS, et al. Zerumbone-loaded nanostructured lipid carrier induces apoptosis of canine mammary adenocarcinoma cells. Biomed Res Int. 2018;2018:1–18.
36. Nathani C, Elaine-Lee YL, Yee BC, et al. Zerumbone-loaded nanostructured lipid carrier induces apoptosis in human colorectal adenocarcinoma (Caco-2) cell line. Nanosci Nanotechnol Lett. 2016;8(4):294–302. doi:10.1016/nnl.2016.2136
37. Shi F, Yang G, Ren J, Guo T, Du Y, Feng NP. Formulation design, preparation, and in vitro and in vivo characterizations of β-ele- mene-loaded nanostructured lipid carriers. Int J Nanomed. 2013; 8:2533–2541. doi:10.2147/IJN.S46578
38. Jaiswal M, Duhde R, Sharma PK. Nanoemulsion: an advanced mode of drug delivery system. J Biotech. 2015;5(2):123–127. doi:10.1007/s13205-014-0214-0
39. Mahato R. Nanoemulsion as targeted drug delivery system for cancer therapeutics. J Pharm Sci Pharmacol. 2017;3(2):83–97. doi:10.1166/jsp.2017.1082
40. Lovelyn C, Attama AA. Current state of nanoemulsions in drug delivery. J Biomater Nanobiotechnol. 2011;2(05):626–639. doi:10. 4236/jbn.2011.225075
41. Chhabra G, Chuttani K, Mishra AK, Pathak K. Design and development of nanoemulsion drug delivery system of amloidipine besi- late for improvement of oral bioavailability. Drug Dev Ind Pharm. 2011;37(8):907–916. doi:10.3109/03639045.2011.550590
42. Topuz OK, Ozvural EB, Zhao Q, Huang Q, Chikindas M, Gölükçü M. Physical and antimicrobial properties of anise oil loaded nanoemulsions on the survival of food-borne pathogens. Food Chem. 2016;203:117–123. doi:10.1016/j.foodchem.2016.02.051
43. Zhang S, Zhang M, Fang Z, Liu Y. Preparation and characterization of blended cloves/cinnamon essential oil nanoemulsions. LWT-Food Sci Tech. 2017;75:316–322. doi:10.1016/j.lwt.2016.08.046
44. Kotta S, Khan AW, Pramod K, Ansari SH, Sharma RK, Ali J. Exploring oral nanoemulsions for bioavailability enhancement of poorly water-soluble drugs. Expert Opin Drug Deliv. 2012;9 (5):585–598. doi:10.1517/17425247.2012.668523
45. Mahajan HS, Mahajan MS, Nerkar PP, Agrawal A. Nanoemulsion-based intranasal drug delivery system of saquinavir mesylate for brain targeting. Drug Deliv. 2014;21(2):148–154. doi:10.3109/ 10717543.2014.838014
46. Khan S, Keyhanfar F, Amani A. Design and evaluation of oral nanoemulsion drug delivery system of mebudipine. Drug Deliv. 2016;23(6):2035–2043. doi:10.1080/10717544.2015.1088597
47. Patel RP, Joshi JR. An overview on nanoemulsion: a novel approach. Int J Pharm Sci Res. 2012;3(12):4640–4650.
56. Raj Tiwari, C. Evaluation of nanotechnology-based carrier for delivery of paclitaxel: in vitro evaluation. Int J Pharm. 2015;470:114–119. doi:10.1016/j.ijpharm.2014.12.033

57. Leitão PS, Loureiro AM, Medeiros N, et al. Characterization of transfersomal formulations containing saxagliptin: in vitro and in vivo evaluation. J Control Release. 2015;211:146–152. doi:10.1016/j.jconrel.2015.04.030

58. Wang Y, Wang S, Su B, et al. Enhanced solubility and oral bioavailability of naringenin via liposomal nanoformulation: preparation and in vitro and in vivo evaluations. AAPS PharmSciTech. 2017;18(3):586–594. doi:10.1007/s12249-016-0537-8

59. Prabhu RH, Patravale VB, Joshi MD. Polymeric nanoparticles for targeted treatment in oncology: current insights. Int J Nanomed. 2015;10:1001–1018. doi:10.2147/IJN.S56932

60. Chauhan P, Tyagi BK. Herbal novel drug delivery systems and transfersomes. J Drug Delivery Ther. 2018;8(3):162–168. doi:10.22270/jdtt.v8s3.1772

61. Rai S, Pande V, Rai G. Transfersomes as versatile and flexible nanovehicular carriers in skin cancer therapy: the state of the art. Nano Rev Exp. 2017;8(1):1–18. doi:10.1020/200227727.2017.1325708

62. El-Refaie WM, Elnaggar YS, El-Massik MA, Abdallah OY. Novel curcumin-loaded gel-core hyaluronates with promising burn-wound healing potential: development, in-vitro appraisal and in vivo studies. Int J Pharm. 2015;486(1–2):88–98. doi:10.1016/j.ijpharm.2015.03.052

63. Choi HJ, Cho SH, Yun JJ, Yu YB, Cho CW. Ethosomes and transfersomes for topical delivery of ginsenoside Rhi from red ginseng: characterization and in vitro evaluation. J Nanosci Nanotechnol. 2015;15(8):5660–5662. doi:10.1166/jnn.2015.10462

64. Ma H, Guo D, Fan Y, Wang J, Cheng X, Zhang X. Paconol-loaded ethosomes as transdermal delivery carriers: design, preparation and evaluation. Molecules. 2018;23(7):1756–1771. doi:10.3390/molecules23071756

65. Neupane YB, Sabir MD, Ahmad N, Ali M, Kohli K. Lipid drug conjugate nanoparticle as a novel lipid nanocarrier for the oral delivery of decitabine: ex vivo gut permeation studies. Nanotechnology. 2013;24(41):1–11. doi:10.1088/0957-4484/24/41/415102

66. Guo W, Li A, Jia Z, Yuan Y, Dai H, Li H. Transferrin-modified PEG-PLA-resveratrol conjugates: in vitro and in vivo studies for glioma. Eur J Pharmacol. 2013;718:41–47. doi:10.1016/j.ejphar.2013.09.034

67. Shen J, Zhang D, Zhao Z, et al. Synthesis, characterization, in vitro and in vivo evaluation of PEGylated oridonin conjugates. J Pharm Sci. 2013;102:80–86. doi:10.1016/j.xpharm.2013.08.014

68. Maiti K, Mukherjee K, Gantait A, Saha BP, Mukherjee PK. Curcumin-phospholipid complex: preparation, therapeutic evaluation and pharmacokinetic study in rats. Int J Pharm. 2007;330(1–2):155–163. doi:10.1016/j.ijpharm.2006.09.025

69. Singh M, Bhattacharjee P, Mishra S, et al. PLGA encapsulated tea polyphenols enhance the chemotherapeutic efficacy of cisplatin against human cancer cells and mice bearing ehrlich ascites carcinoma. Int J Nanomed. 2015;10:6789–6809. doi:10.2147/IJN.S79489

70. Sanna V, Siddiqui IA, Sechi M, Mukhtar H. Resveratrol-loaded nanoparticles based on poly(epsiloncaprolactone) and poly(D,L-lactic-co-glycolic acid)-poly(ethylene glycol) blend for prostate cancer treatment. Mol Pharm. 2013;10:3871–3881. doi:10.1021/mp400342f

71. Pan M, Li W, Yang J, et al. Plumbagin-loaded aptamer-targeted poly D, L-lactic-co-glycolic acid-b-polyethylene glycol nanoparticles for prostate cancer therapy. Medicine. 2017;96(30):e7405. doi:10.1097/MD.0000000000007405

72. Ganesan P, Narayanasamy D. Lipid nanoparticles: different preparation techniques, characterization, hurdles, and strategies for the production of solid lipid nanoparticles and nanostructured lipid carriers for oral drug delivery. Sustain Chem Pharm. 2017;7:37–56. doi:10.1016/j.scpp.2017.07.002

73. Robson AL, Dastoor PC, Flynn J, et al. Advantages and limitations of current imaging techniques for characterizing liposome morphology. Front Pharmacol. 2018;9(80):1–8. doi:10.3389/fphar.2018.00080

74. Lee WH, Loo CY, Young PM, Traini D, Rohanizadeh R. The development and achievement of polymeric nanoparticles for cancer drug treatment. Part Technol Delivery Ther. 2017;25:82.

75. Song Z, Lin Y, Zhang X, et al. Cyclic RGD peptide-modified liposomal drug delivery system for targeted oral apatinib administration: enhanced cellular uptake and improved therapeutic effects. Int J Nanomed. 2017;12:1941–1958. doi:10.2147/IJN.S125573

76. Hong SS, Kim SH, Lim SJ. Effects of triglycerides on the hydrophobic drug loading capacity of saturated phosphatidylcholine-based liposomes. Int J Pharm. 2015;483(1–2):142–150. doi:10.1016/j.ijpharm.2015.02.013

77. Thangapazham RL, Puri A, Tele S, Blumenthal R, Maheshwari RK. Evaluation of nanotechnology-based carrier for delivery of curcumin in prostate cancer cells. Int J Oncol. 2008;32:1119–1123.

78. Hong SS, Choi JY, Kim JO, Lee MK, Kim SH, Lim SJ. Development of paclitaxel-loaded liposomal nanocarrier stabilized by triglyceride incorporation. Int J Nanomed. 2016;11:4465–4477. doi:10.2147/IJN.S113723

79. Kannan V, Balabhathula P, Divi MK, Thoma LA, Wood GC. Optimization of drug loading to improve physical stability of paclitaxel-loaded long-circulating liposomes. J Liposome Res. 2015;25(4):308–315. doi:10.3109/08988210.2014.995671

80. Li N, Feng L, Tan Y, Xiang Y, Zhang R, Yang M. Preparation, characterization, pharmacokinetics and biodistribution of bicalain-loaded liposome on cerebral ischemia-reperfusion after iv administration in rats. Molecules. 2018;23(7):1747–1761. doi:10.3390/molecules23071747
98. Ambwani S, Tandon R, Ambwani TK, Malik YS. Current knowledge on nanodelivery systems and their beneficial applications in enhancing the efficacy of herbal drugs. J Exp Biol Agric Sci. 2018;6(6):87–107. doi:10.18066/j.biosci.2018.6.87107

99. Ramesh M, Sharififar F, Mehrabani M, Pardakhht A, Farsinejad A, Mehrabani M. Proliferation and in vitro wound healing effects of the microniosomes containing Narcissus tazetta L. bulb extract on primary human fibroblasts (HDFs). DARU. 2018;26(1):31–42. doi:10.1007/s40199-018-0211-7

100. Barani M, Mirzaei M, Torkzadeh-Mahani M, Nematollahi MH. Lawsone-loaded niosomes and its antioxidant activity in MCF-7 breast cancer cell line: a nano-herbal treatment for cancer. DARU. 2018;26(1):11–27. doi:10.4103/daru.daru.2018.261847

101. Anghore D, Kulkarni GT. Development of novel niosome gels as drug delivery system of spermacoce Roxb. extract for anti-inflammatory activity enhanced skin permeation and stability of compound D. AAPS PharmSciTech. 2016;17(3):631–639. doi:10.1208/s12249-015-0376-2

102. Barani M, Mizraei M, Torkzadeh-Mahani M, Nematiollahi MH. Niosome gel of Zingiber cassumunar Roxb. extract for anti-inflammatory activity enhanced skin permeation and stability of compound D. AAPS PharmSciTech. 2016;17(3):631–639. doi:10.1208/s12249-015-0376-2

103. Alpi MS, Ahdad A, Abidin L, Aqil M, Mir SR, Mujeeb M. Embelin-loaded oral niosomes ameliorate streptozotocin-induced diabetes in wistar rats. Biomed Pharmacother. 2018;97:1514–1520. doi:10.1016/j.biopha.2017.11.073

104. Gunes A, Guler E, Un RN, et al. Niosomes of nerium oleander leaves as carriers for the transdermal delivery of colchicine: statistical optimization of the microniosomes containing Narcissus tazetta L. bulb extract for anti-inflammation and mitigating toxicity for effective anticancer drug and gene delivery. Acta Biomater. 2016;43:14–29. doi:10.1016/j.actbio.2016.07.015

105. Liu F, Sun Y, Kang C, Zhu H. Pegylated drug delivery systems: in vitro assessment of bioactive nanovesicular structures. Drug Deliv. 2016;23(1):254–262. doi:10.3109/10717544.2014.910564

106. Wang L, Xu X, Zhang Y, et al. Encapsulation of curcumin within poly (amidoamine) dendrimers as drug carriers for delivery to cancer cells. J Mater Sci Mater Med. 2013;24:2137–2144. doi:10.1007/s10856-013-4969-3

107. Falconieri MC, Adamo M, Monasterolo C, Bergonzi MC, Coronello M, Bilia AR. New dendrimer-based nanoparticles enhance curcumin solubility. Planta Med. 2017;83(5):420–425. doi:10.1055/s-0042-175252

108. Gu L, Wu ZH, Qi X, et al. Polyamidoamine dendrimers: an excellent drug carrier for improving the solubility and bioavailability of puerarin. Pharm Dev Technol. 2013;18(6):1051–1057. doi:10.1080/10837450.2011.653822

109. Kambhampati SP, Kannan RM. Dendrimer nanoparticles for ocular drug delivery. J Ocul Pharmacol Ther. 2013;29(2):151–165. doi:10.1089/jop.2012.0232

110. Diaz C, Guzmán J, Jiménez VA, Alderete JB. Partially PEGylated PAMAM dendrimers as solubility enhancers of silybin. Pharm Dev Technol. 2018;23(7):689–696. doi:10.1080/10837450.2017.1315134

111. Yesil Celiktas O, Pala C, Cetin Uyanikgil EO, Sevimli Gur C. Synthesis of silica-PAMAM dendrimer nanoparticles as promising carriers in neuro blastoma cells. Anal Biochem. 2017;519:1–7. doi:10.1016/j.ab.2016.12.004

112. Qi WJ, Li HF, Su YY, Dong ZQ, Ge YR. Absorption enhancing ability of andesartan cilexetil and mitigating toxicity for effective antihypertensive drug action. Eur J Pharm Sci. 2016;336:1–15. doi:10.1016/j.ejps.2015.12.031

113. Han R, Sun Y, Kang C, Sun H, Wei W. Amphiphilic dendritic nanomicelle-mediated co-delivery of 5-fluorouracil and doxorubicin for enhanced therapeutic efficacy. J Drug Target. 2017;25:140–148. doi:10.1080/10613044.2016.1207649

114. Wang Z, Yu Y, Ma J, et al. LyP-1 modification to enhance delivery of artemisinin or fluorescent probe loaded polymer micelles to highly metastatic tumor and its lymphatics. Mol Pharm. 2012;9:2646–2657. doi:10.1021/mp3002107
134. Rahman J, Farghaly S, Tawfik R, et al. A micelle-like structure of polyoxamer—methotrexate conjugates as nanocarrier for methotrexate delivery. *Int J Pharm*. 2015;487(1–2):177–186. doi:10.1016/j.ijpharm.2015.04.014

135. Zhang Y, Zhang H, Wang X, Wang J, Zhang X, Zhang Q. The eradication of breast cancer and cancer stem cells using octreotide modified paclitaxel active targeting micelles and salinomycin passive targeting micelles. *Biomaterials*. 2012;33(2):679–691.

136. Guo M, Men K, Shi H, et al. Curcumin-loaded biodegradable polymeric micelles for colon cancer therapy in vitro and in vivo. *Nanoscale*. 2011;3(4):1558–1567. doi:10.1039/c008075g

137. Abdelmonem MA, Mahmoud M, Zaky A, et al. Dual-targeted casein micelles as green nanomedicine for synergistic phototherapy of hepatocellular carcinoma. *J Control Release*. 2018;278:73–83. doi:10.1016/j.jconrel.2018.08.026

138. Wu H, Yu T, Tian Y, Wang Y, Zhao R, Mao S. Enhanced liver-targeting via coadministration of 10-hydroxyapomorphine polymeric micelles with vitamin baked radix bupleuri. *Phytomedicine*. 2018;44:1–8. doi:10.1016/j.phymed.2018.04.022

139. Su Y, Huang N, Chen D, et al. Successful in vivo hyperthermal therapy toward breast cancer by Chinese medicine shikonin-loaded thermosensitive micelle. *Int J Nanomed*. 2017;12:4019. doi:10.2147/IJN.S132639

140. Anantaworasakul P, Okonogi S. Encapsulation of sesbania grandiflora extract in polymeric micelles to enhance its solubility, stability, and antibacterial activity. *J Microencapsul*. 2017;34(1):73–81. doi:10.1080/02652048.2017.1284277

141. Baldissera MD, Souza CF, Boligon AA, et al. Solving the challenge of cancer stem cells in breast cancer therapy through preclinical evaluation of oridonin. *Biomaterials*. 2017;154. doi:10.1016/j.biomaterials.2017.01.003

142. Pereira K, Quintela E, Da Silva D, et al. Characterization of nanoencapsulated flavonoids in green' nanomaterials: a novel nanomedical approach. *BioMed Res Int*. 2017;2017:6130278. doi:10.1155/2017/6130278

143. Wang T, Wu C, Fan G, et al. Optimization, formulation, and degradation in artificial saliva. *Mater Res*. 2018;21(2):1–9. doi:10.1590/1980-5373-mr-2017-0488

144. Wang T, Wu C, Fan G, Li, Gong H, Cao F. Ginkgo biloba flavonoids mediated nanocrystals. *Phytomedicine*. 2018;404. doi:10.1016/j.phymed.2018.04.022

145. Elzoghby A. Polymeric nanocarriers as robust platforms for cancer therapy. *Curr Pharm Des*. 2017;23(35):5211–5212. doi:10.2174/138161281766617106125801

146. Elzoghby A. Polymeric nanocarriers as robust platforms for cancer therapy. *Curr Pharm Des*. 2017;23(35):5211–5212. doi:10.2174/138161281766617106125801

147. Li C, Zhang D, Guo H, et al. Preparation and characterization of galactosylated bovine serum albumin nanoparticles for liver-targeted delivery of oridonin. *Int J Pharm*. 2013;448:79–86. doi:10.1016/j.ijpharm.2013.07.012

148. Elnaggar M, El-Shazly A, Hamdy A, et al. Paclitaxel-loaded PLGA micro and nanoparticles: synthesis, characterization and degradation in artificial saliva. *Mater Res*. 2018;21(2):1–9. doi:10.1590/1980-5373-mr-2017-0488
Anwar E, Farhana N. Formulation and evaluation of phytosome-loaded maltodextrin-gum Arabic microsphere system for delivery of Camellia sinensis extract. J Young Pharm. 2018;10(2s):S56. doi:10.5530/jyp
El-Menshawe SF, Ali AA, Rabab MA, Khalil NM. Nanosized soy phytosome-based thermogel as topical anti-obesity formulation: an approach for acceptable level of evidence of an effective novel herbal weight loss product. Int J Nanomed. 2018;13:307–318. doi:10.2147/IJNN.S135429
Hooresfand Z, Ghanbarzadeh S, Hamishehkar H. Preparation and characterization of rutin-loaded nanophytosomes. Pharm Sci. 2015;21(3):145–151. doi:10.1515/PS.2015.29
Vu HT, Hook SM, Siqueira SD, Müllertz A, Rades T, McDowell A. Are phytosomes a superior nanodelivery system for the antioxidant rutin? Int J Pharm. 2018;122:214–229.
Gahandule MB, Jadhav SJ, Gadhave MV, Gaikwad DD. Formulation and development of hepato-protective butea monosperma-phytosome. Int J Res Pharm Sci. 2016;1(4):21–27.
Singh RP, Gangadharpappa HV, Mruthunjaya K. Phytosome complexed with chitosan for gingerol delivery in the treatment of respiratory infection: in vitro and in vivo evaluation. Eur J Pharm Sci. 2018;122:214–229. doi:10.1016/j.ejps.2018.06.028
Kassem AA, Molsen AM, Ahmed RS, Essam TM. Self-nanoemulsifying drug delivery system (SNEDDS) with enhanced solubilization of nystatin for treatment of oral candidiasis: design, optimization, in vitro and in vivo evaluation. J Mol Liq. 2016;218:219–232. doi:10.1016/j.molliq.2016.02.081
Senapati PC, Sahoo SK, Sahu AN. Mixed surfactant based (SNEDDS) self-nanoemulsifying drug delivery system presenting efavirenz for enhancement of oral bioavailability. Biomed Pharmacother. 2016;80:42–51. doi:10.1016/j.biopha.2016.02.039
Chatterjee B, Hamed Almursi S, Ahmed Mahdi Dukhan A, Mandal UK, Sengupta P. Controversies with self-emulsifying drug delivery system from pharmacokinetic point of view. Drug Deliv. 2016;23(9):3639–3652. doi:10.1080/10717544.2016.1214990
Agrawal AG, Kumar A, Gide PS. Formulation of solid self-nanoemulsifying drug delivery systems using N-methyl pyrrolidone as cosolvent. Drug Dev Ind Pharm. 2015;41:594–604. doi:10.3109/03639045.2014.886695
Li W, Yi S, Wang Z, et al. Self-nanoemulsifying drug delivery system of persimmon leaf extract: optimization and bioavailability studies. Int J Pharm. 2011;420(1):161–171. doi:10.1016/j.ijpharm.2011.08.024
Avachat AM, Patel VG. Self-nanoemulsifying drug delivery system of stabilized elagic acid-phospholipid complex with improved dissolution and permeability. Saudi Pharm J. 2015;23(3):276–289. doi:10.1016/j.sjps.2014.11.001
Tran TH, Guo Y, Song D, Bruno RS, Lu X. Quercetin-containing self-nanoemulsifying drug delivery system for improving oral bioavailability. J Pharm Sci. 2013;10(3):840–852. doi:10.1002/jps.23588
Shen J, Bi J, Tian H, et al. Preparation and evaluation of a self-nanoemulsifying drug delivery system loaded with akebia saponin D-phospholipid complex. Int J Nanomed. 2016;11:4919–4929. doi:10.2147/IJNN.S108765
Shukla M, Jaiswal S, Sharma A, et al. A combination of complexion and self-nanoemulsifying drug delivery system for enhancing oral bioavailability and anticancer efficacy of curcumin. Drug Deliv. Ind Pharm. 2017;43(5):847–861. doi:10.1080/03639045.2016.1239732
Khan AW, Kotta S, Ansari SH, Sharma RK, Ali J. Self-nanoemulsifying drug delivery system (SNEDDS) of the poorly water-soluble grapefruit flavonoid naringenin: design, characterization, in vitro and in vivo evaluation. Drug Deliv. 2015;22(4):552–561. doi:10.1080/10717544.2013.878003
Yeom DW, Chae BR, Son HY, et al. Enhanced oral bioavailability of valsartan using a polymer-based supersaturable self-microemulsifying drug delivery system. Int J Nanomed. 2017;12:3533–3545. doi:10.2147/IJNN.S136599
Yan B, Wang Y, Ma Y, Zhao J, Liu Y, Wang L. In vitro and in vivo evaluation of poly (acrylic acid) modified mesoporous silica nanoparticles as pH response carrier for β-encelme self-micro emulsifying. International Journal of Pharmaceutics. 2019;572:118768–18778.
Allen JL. Basics of complicating-nonsterile: complicating self-emulsifying drug delivery systems and other self-emulsifying lipid formulations, part 1. Int J Pharm Compd. 2018;22(3):220–228.
Jabak G, Fülop V, Santha K, Szerőczei D, Balogh E, Antal L. Formulation possibilities of self-emulsifying drug delivery systems, microemulsions and nanoemulsions. Acta Pharm Hung. 2017;87(1):27–34.
Zhang L, Zhu W, Yang C, et al. A novel folate-modified self-microemulsifying drug delivery system of curcumin for colon targeting. Int J Nanomed. 2012;7:151–162. doi:10.2147/IJN.S27639
Chen Y, Zhang H, Yang J, Sun H. Improved antioxidant capacity of optimization of a self-microemulsifying drug delivery system for resveratrol. Molecules. 2015;20:21167–21177. doi:10.3390/molecules201219750
Jaisamut P, Wiwattanawongs K, Gradaist P, Sangsen Y, Wiwattanapatree R. Enhanced oral bioavailability of curcumin using a supersaturable self-microemulsifying system incorporating a hydrophilic polymer; in vitro and in vivo investigations. AAPS PharmSciTech. 2018;19(2):730–740. doi:10.1208/s12249-017-0857-3
Qiao J, Ji D, Sun S, et al. Oral bioavailability and lymphatic transport of pueraria flavione-loaded self-emulsifying drug delivery systems containing sodium taurocholate in rats. Pharmaceutics. 2018;10(3):147–160. doi:10.3390/pharmaceutics10030147
Sornsuvit C, Hongwiset D, Yotsawimonwat S, Toonkum M, Thongsawat S, Taesotikul W. The bioavailability and pharmacokinetics of silymarin SMEDDS formulation study in healthy thai volunteers. Evid Based Complement Altern Med. 2018;1:1–7. doi:10.1155/2018/1507834
Dhulam DM, Akamanchi KG. Self-microemulsifying drug delivery system for camptothecin using new bicephalous heterolipid with tertiary-amine as branching element. Int J Pharm. 2018;541(1–2):48–55. doi:10.1016/j.ijpharm.2018.02.030
Sato Y, Joumura T, Nishimoto S, et al. Enhancement of lymphatic transport of lutein by oral administration of a solid dispersion and a self-microemulsifying drug delivery system. Eur J Pharm Biopharm. 2018;127:171–176. doi:10.1016/j.ejpb.2018.02.013
Karami Z, Rezaeian I, Zahedi P, Abdollahi M. Preparation and performance evaluations of electro spun poly (ε-caprolactone), poly (lactic acid), and their hybrid (50:50) nanoﬁbrous mats containing thymol as an herbal drug for effective wound healing. J Appl Polym Sci. 2013;129(2):756–766. doi:10.1002/app.38683
Available from: http://scienceضعیفیابیمگمجردی.org/content/294/5547/1684/F1. Accessed December 5, 2019.
Wang H, Wei J, Yang C, et al. The inhibition of tumor growth and metastasis by self-assembled nanoﬁbers of taxol. Biomaterials. 2012;33(24):5848–5853. doi:10.1016/j.biomaterials.2012.04.047
Wagh A, Singh J, Qian S, Law B. A short circulating peptide nanoﬁber as a carrier for tumoral delivery. Nanomedicine. 2013;9(4):449–457. doi:10.1016/j.nano.2012.10.009
Liu J, Xu H, Zhang Y, et al. Novel tumor-targeting, self-assembling peptide nanoﬁber as a carrier for effective curcumin delivery. Int J Nanomed. 2014;9:197–207. doi:10.2147/IJNN.S55875
Ranjbar MM, Bahrani SH. Electros spun curcumin loaded poly (ε-caprolactone)/gum tragacanth nanoﬁbrers for biomedical application. Int J Biol Macromol. 2016;84:448–456. doi:10.1016/j.ijbiomac.2015.12.024
Choi J, Yang BJ, Bae GN, Jung JH. Herbal extract incorporated nanoﬁber fabricated by an electrospinning technique and its application to antimicrobial air ﬁltration. J Ocul Pharmacol Ther. 2015;7(45):25313–25320. doi:10.1021/acsmi.5b07441
204. Bonan RF, Bonan PR, Batista AU, et al. In vitro antimicrobial activity of solution blow spin poly (lactic acid)/polyvinylpyrollidone nanofibers loaded with copaiba (Copaifera sp.) oil. *Mater Sci Eng C.* 2015;48:372–377. doi:10.1016/j.msec.2014.12.021

205. Jouybar A, Seyedi Jafari A, Ardehiyajami A, et al. Enhanced skin regeneration by herbal extract-coated poly-L-lactic acid nanofibrous scaffold. *Artif Organs.* 2017;41(11):E296–E307. doi:10.1111/aor.12926

206. Parvath V, Krishnan AG, Anitha A, Jayakumar R, Nair MB. Poly (L-lactic acid) nanofibers containing cissus quadrangularis induced osteogenic differentiation in vitro. *Int J Biol Macromol.* 2018;110:514–521. doi:10.1016/j.ijbiomac.2017.11.094

207. Wang J, Tian L, He L, et al. Lycium barbarum polysaccharide encapsulated poly lactic-co-glycolic acid nanofibers: cost effective herbal medicine for potential application in peripheral nerve tissue engineering. *Sci Rep.* 2018;8(1):8669–8683. doi:10.1038/s41598-018-26837-z

208. Lee JS, Feijen J. Biodegradable polymersomes as carriers and release systems for paclitaxel. *J Control Release.* 2014;177:207–215. doi:10.1016/j.jconrel.2014.08.004

209. Jouybar A, Seyedjafari E, Ardeshirylajimi A, et al. Enhanced skin permeation across the skin. *J Drug Deliv Sci Technol.* 2018;48:75–81. doi:10.1016/j.jddst.2018.09.002

210. Bazylitska U, Kulbacka J, Schmidt J, Talmon Y, Murgia S. Polymer-free cubosomes for simultaneous bioimaging and photodynamic action of photosensitizers in melanoma skin cancer cells. *J Colloid Interface Sci.* 2018;522:163–173. doi:10.1016/j.jcis.2018.03.063

211. Gupta PK, Asthana S, Jaiswal AK, et al. Exploitation of lectinized lipo-polymerosome encapsulated poly lactic-co-glycolic acid nano. *Nanomedicine (Lond).* 2013.08.004

212. Rahman et al. 2015.3534. doi:10.1002/jps.24548

213. Hammer DA, Robbins GP, Haun JB, et al. Leukopolymersomes. *Synthesis of curcumin loaded polymeric nanoparticles from crab shell derived chitosan for drug delivery.* *Int J Biol Macromol.* 2018;109:748–760. doi:10.1016/j.ijbiomac.2017.11.064

214. Matloub AA, AbouSamra MM, Salama AH, Rizk MZ, Aly HF, Fouad GI. Cubic liquid crystalline nanoparticles containing a polysaccharide from ulva fasciata with potent antihyperlipidaemic activity. *Saudi Pharm J.* 2018;26(2):224–231. doi:10.1016/j.sjps.2017.12.007

215. Nitta SK, Numata K. Biopolymer-based nanoparticles for drug/gene delivery and tissue engineering. *Int J Mol Sci.* 2013;14(1):1629–1654. doi:10.3390/ijms14011629

216. Fathi M, Martin A, Mc Clements DJ. Nanocapsulation of food ingredients using carbohydrate based delivery systems. *Trends Food Sci Tech.* 2014;39(1):18–39. doi:10.1016/j.tifs.2014.06.007

217. Biranje SS, Madiwale PV, Patankar KC, Chhabra R, Dandekar-Jain P, Adivearekar RV. Hemostasis and anti-necrotic activity of wound healing dressing containing chitosan nanoparticles. *Int J Biol Macromol.* 2018;121:936–946. doi:10.1016/j.ijbiomac.2018.10.125

218. Tan Q, Liu W, Guo C, Zhai G. Preparation and evaluation of quercetin-loaded lecithin-chitosan nanoparticles for topical delivery. *Int J Nanomed.* 2011;6:16211630. doi:10.2147/IJN.S25646

219. Bu L, Gan LC, Guo XQ, et al. Trans-resveratrol loaded chitosan nanoparticles modified with biotin and avidin to target hepatic carcinoma. *Int J Pharm.* 2013;452(1–2):355–362. doi:10.1016/j.ijpharm.2013.05.007

220. Samrot AV, Burman U, Philip SA, Shobana N, Chandrasekar K. Synthesis of curcumin loaded polymeric nanoparticles from crab shell derived chitosan for drug delivery. *Int Med Unlocked.* 2018;10:159–182. doi:10.1016/j.imult.2017.12.010

221. Esfanjani AF, Jafari SM. Biopolymer nano-particles and natural nano-carriers for nano-encapsulation of phenolic compounds. *Colloids Surf B BioInterfaces.* 2016;146:532–543. doi:10.1016/j.colsurf.b.2016.06.053

222. Khan N, Bharali DJ, Adhami VM, et al. Oral administration of naturally occurring chitosan-based nanofORMulated green tea polyphenol EGCG effectively inhibits prostate cancer cell growth in a xenograft model. *Carcinogenesis.* 2014;35:415–423. doi:10.1093/carcin/bgt211

223. Loch-Neckel G, Santos-Bubbniak L, Mazzarino L, et al. Orally administered chitosan-coated polycaprolactone nanoparticles containing curcumin attenuate metastatic melanoma in the lungs. *J Pharm Sci.* 2015;104:3524–3534. doi:10.1002/jps.24548
239. Karri VV, Kuppasamy G, Talluri SV, et al. Curcumin loaded chitosan nanoparticles impregnated into collagen-alginic scaffolds for diabetic wound healing. Int J Biol Macromol. 2016;93:1519–1529. doi:10.1016/j.ijbiomac.2016.05.038

240. Feng B, Ashraf MA, Peng L. Characterization of particle shape, zeta potential, loading efficiency and outdoor stability for chitosan-rinicolic acid loaded with rotenone. Open Life Sci. 2016;11 (1):380–386. doi:10.1515/open-2016-0050

241. Suksaeree J, Monton C, Madaka F, et al. Formulation, physicochemical characterization, and in vitro study of chitosan/HPMC blends-based herbal blended patches. AAPS PharmSciTech. 2015;16(1):171–181. doi:10.1208/s12249-014-0216-6

242. Zhang H, Yu D, Guo R, Deng L, Dong A, Zhang J. Composites of polymer hydrogels and nanoparticulate systems for biomedical and pharmaceutical applications. Nanomaterials. 2015;5(4):2054–2130. doi:10.3390/nano5042054

243. Dimatteo R, Darling NJ, Segura T. In situ forming injectable hydrogels for drug delivery and wound repair. Adv Drug Deliv Rev. 2018;127:167–184. doi:10.1016/j.addr.2018.03.007

244. Atta S, Khaliq S, Islam A, et al. Injectable biopolymer based hydrogels for drug delivery applications. Int J Biol Macromol. 2015;80:240–245. doi:10.1016/j.ijbiomac.2015.06.044

245. Córdoba AL, Deladino L, Martino M. Effect of starch filler on calcium alginate hydrogels loaded with yerba mate antioxidants. J Polym Sci Part A: Polym Chem. 2013;51:315–323. doi:10.1002/pola.25303

246. Mun S, Kim YR, McClements DJ. Control of β-carotene bioaccessibility using starch-based filled hydrogels. Food Chem. 2015;173:454–461. doi:10.1016/j.foodchem.2014.10.053

247. Balestrin LA, Bidone J, Bortolin RC, Moresco K, Moreira JC, Lustosa AK, de Jesus Oliveira AC, Quelemes PV, et al. In situ synthesis of biopolymer based hydrogels to deliver of anti-HIV effects of atripla as a new nanoformulated drug. Adv Drug Deliv. 2017;24(1):1170–1178. doi:10.1080/10717544.2017.1365393

248. Vio V, Jose Marchant M, Araya E, Kogan JM. Metal nanoparticles for the treatment and diagnosis of neurodegenerative brain diseases. Curr Pharm Des. 2017;23(13):1916–1926. doi:10.2174/1381612823666170105152948

249. Vodyanoy V, Daniels Y, Pustovyy O, MacCrehan WA, Muramoto S, Stan G. Engineered metal nanoparticles in the sub-nanomolar range for the treatment and diagnosis of neurodegenerative brain diseases. Curr Pharm Des. 2017;23(13):1916–1926. doi:10.2174/1381612823666170105152948

250. Vijayakumar V, Samal SK, Mohanty S, Nayak SK. Recent advancements in biopolymer and metal nanoparticle-based materials in diabetic wound healing management. Int J Biol Macromol. 2018;122:137–148. doi:10.1016/j.ijbiomac.2018.10.120

251. Dai Y, Wang LH, Meng DL, Che X. A green and facile preparation approach, icocchalone A capped on hollow gold nanoparticles, for improving the solubility and dissolution of anticancer natural product. Oncotarget. 2017;8(62):105673–105681. doi:10.18632/oncotarget.22387

252. Azeem L, Lateef A, Adelsi SA. Silver nanoparticles (AgNPs) biosynthesized using pod extract of Cola nitida enhances antioxidant activity and phytochemical composition of Amaranthus caudatus Linn. Appl Nanosci. 2017;7(1–2):59–66. doi:10.1007/s13204-017-0546-2

253. Namvar F, Rahman HS, Mohamad R, et al. Cytotoxic effects of biosynthesized zinc oxide nanoparticles on murine cell lines. Evid Based Complement Altern Med. 2015;2015:1–11. doi:10.1155/2015/593014

254. Danafar H, Shariﬁ A, Kheiri S. Co-delivery of sulfurofan and curcumin with PE-Gylated iron oxide-gold core shell nanoparticles for delivery to breast cancer cell line. Iran J Pharm Res. 2018;17(2):480–494.

255. Singh A, Dutta PK, Kumar H, Kureel AK, Rai AK. Synthesis of chitin-glucan-aldehyde-curcumin conjugate and evaluation of anticancer and antioxidant activities. Carbohydr Polym. 2018;193:99–107. doi:10.1016/j.carbpol.2018.03.092

256. Singh A, Kureel AK, Dutta PK, Kumar S, Rai AK. Curcumin loaded chitin-glucan querctein conjugate: synthesis, characterization, antioxidant, in vitro release study, and anticancer activity. Int J Biol Macromol. 2018;110:234–244. doi:10.1016/j.ijbiomac.2017.11.002
270. Poonia N, Lither V, Pandita D. Mesoporous silica nanoparticles: a smart nanosystem for management of breast cancer. Drug Discov Today. 2017;23(2):315–332. doi:10.1016/j.drudis.2017.10.022
271. Al-Asmar A, Giosafatto CV, Sabah M, Sanchez A, Villalona Santana R, Marinello L. Effect of Mesoporous Silica Nanoparticles on The Physicochemical Properties of Pectin Packaging Material for Strawberry Wrapping. Nanomaterials. 2020;10(1):52–70.
272. Zhou Y, Quan G, Wu Q, et al. Mesoporous silica nanoparticles for drug and gene delivery. Acta Pharm Sin B. 2018;8(2):165–177. doi:10.1016/j.apsb.2018.01.007
273. Desai D, Zhang J, Sandholm J, et al. Lipid bilayer-gated mesoporous silica nanocarriers for tumor-targeted delivery of zoledronic acid in vivo. Mol Pharmacol. 2017;14(9):3218–3227. doi:10.1021/acs.molpharmaceut.7b00519
274. Cao X, Deng WW, Fu M, et al. In vitro release and in vitro–in vivo correlation for silybin meglumine incorporated into hollow-type mesoporous silica nanoparticles. Int J Nanomed. 2012;7:753–762. doi:10.2147/IJN.S28348
275. AbouAitah K, Swiderska-Sroda A, Farghali AA, et al. Folic acid–conjugated mesoporous silica particles as nanocarriers of natural produgs for cancer targeting and antioxidant action. Oncotarget. 2018;9(41):26646–26490. doi:10.18632/oncotarget.25470
276. Lin J, Cai Q, Tang Y, et al. PEGLyated lipid bilayer coated mesoporous silica nanoparticles for co-delivery of paclitaxel and curcumin: design, characterization and its cytotoxic effect. Int J Pharm. 2018;536(1):272–282. doi:10.1016/j.ijpharm.2017.10.043
277. Li T, Chen X, Liu Y, et al. pH-sensitive mesoporous silica nanoparticles anticancer produgs for sustained release of ursoic acid and the enhanced anti-cancer efficacy for hepatocellular carcinoma cancer. Eur J Pharm Sci. 2017;96:456–463. doi:10.1016/j.ejps.2016.10.019
278. Kumar B, Kulnhaivel S, Mondal A, et al. Mesoporous silica nanoparticle based enzyme responsive system for colon specific drug delivery through guar gum capping. Colloids Surf B Biointerfaces. 2017;150:352–361. doi:10.1016/j.colsurfb.2016.10.049
279. Choi JY, Ramasamy T, Kim SY, et al. PEGLyated lipid bilayer-supported mesoporous silica nanoparticle composite for synergistic co-delivery of axitinib and celastrol in multi-targeted cancer therapy. Acta Biomater. 2016;39:94–105. doi:10.1016/j.actbio.2016.05.012
280. Souza AC, Amaral AC. Antifungal therapy for systemic mycosis and the nanobiotechnology era: improving efficacy, biodistribution and toxicity. Front Microbiol. 2017;8(336):1–13. doi:10.3389/fmicb.2017.00336
281. Garcia Rubia G, Peigneux A, Jabalera Y, et al. pH-dependent adsorption rate of doxorubicin on mame-bioimimetic magnetite nanoparticles. Langmuir. 2018;34(45):13713–13724. doi:10.1021/acs.langmuir.8b03109
282. Lungu II, Radulescu M, Mogosanu GD, Grumezescu AM. pH sensitive core-shell magnetic nanoparticles for targeted drug delivery in cancer therapy. Rom J Morphol Embryol. 2016;57(1):23–32.
283. Wang C, Zhang H, Chen Y, Shi F, Chen B. Gamboagic acid-loaded magnetic Fe3O4 nanoparticles inhibit panc-1 pancreatic cancer cell proliferation and migration by inactivating transcription factor ETS1. Int J Nanomed. 2012;7:781–787. doi:10.2147/IJN.S28509
284. Rayegan A, Allafchian A, Sarsari IA, Kameli P. Synthesis and characterization of basil seed mucilage coated Fe3O4 magnetic nanoparticles as a drug carrier for the controlled delivery of cephalixin. Int J Biol Macromol. 2018;113:317–328. doi:10.1016/j.ijbiomac.2018.02.134
285. Arokriyayar S, Saravanan M, Prakash Nl, Arasu MV, Vijayakumar B, Vincent S. Enhanced antibacterial activity of iron oxide magnetic nanoparticles treated with Argemone mexicana L. leaf extract: an in vitro study. Mater Res Bull. 2013;48(9):3323–3327. doi:10.1016/j.materresbull.2013.05.059
286. Namvar F, Rahman HS, Mohamad R, et al. Cytoxic effect of magnetic iron oxide nanoparticles synthesized via seaweed aqueous extract. Int J Nanomed. 2014;9:2479–2488. doi:10.2147/IJN.S59661
287. Allafchian A, Jalali SA, Hosseini F, Massoud M. Ocimum basilicum mucilage as a new green polymer support for silver in magnetic nanocomposites: production and characterization. J Environ Chem Eng. 2017;5(6):5912–5920. doi:10.1016/j.jece.2017.11.023
288. Domnani D, Hussein MZ, Kura AU, Fakurazi S, Shaari AH, Ahmad Z. Preparation of Fe3O4 magnetic nanoparticles coated with gallic acid for drug delivery. Int J Nanomed. 2012;7:5745–5756. doi:10.2147/IJN.S35746
289. Wang C, Liu X, Chen S, Hu FQ, Sun J, Yuan H. Facile preparation of phospholipid-amorphous calcium carbonate hybrid nanoparticles: toward controllable burst drug release and enhanced tumor penetration. Chem Commun. 2018;54(93):13080–13083. doi:10.1039/CSCC07694D
290. Zhao L, Zhang Y, Miao Y, Nie L. Controlled synthesis, characterization and application of hydrophobic calcium carbonate nanoparticles in PVC. Powder Technol. 2016;288:184–190. doi:10.1016/j.powtec.2015.11.001
291. Nakić S, Wang JT, Wu J, et al. Designed inorganic porous nanovesicle with controlled release and MRI features for safe administration of doxorubicin. Int J Pharm. 2019;554:327–336. doi:10.1016/j.ijpharm.2018.10.074
292. Hammad NI, Abba Y, Hisemz MNM, et al. Formulation of a sustained release doxetaxel loaded cockle shell-derived calcium carbonate nanoparticles against breast cancer. Pharm Res. 2017;34:1193–1203. doi:10.1007/s11199-017-2135-1
293. Shi H, Li L, Zhang L, et al. Designed preparation of polyacrylic acid/calcium carbonate nanoparticles with high doxorubicin payload for liver cancer chemotherapy. Cryst Eng Comm. 2015;17:4768–4773. doi:10.1039/C5CE00708A
294. Tully J, Fakhruillin R, Lvov Y. Halloysite clay nanotube composites with sustained release of chemicals. In: Nanomaterials and Nanoarchitectures. Springer, Dordrecht; 2015:87–118.
295. Lazzara G, Cavallaro G, Panchal A, et al. An assembly of organic-inorganic composites using halloysite clay nanotubes. Curr Opin Colloid Interface Sci. 2018;35:42–50. doi:10.1016/j.cocis.2018.01.002
296. Tully J, Yenduri R, Lvov Y. Halloysite clay nanotubes for enzyme immobilization. Biomacromolecules. 2016;17(2):615–621. doi:10.1021/acs.biomac.5b01542
297. Vergaro V, Lvov YM, Leportari S. Halloysite clay nanotubes for resveratrol delivery to cancer cells. Macromol Biosci. 2012;12(9):1265–1271. doi:10.1002/mabi.v12.9
298. Lee MH, Seo HS, Park PH. Thymol encapsulated in halloysite nanotubes for anti-microbial packaging system. J Food Sci. 2017;82(4):922–932. doi:10.1111/1750-3841.13675
299. Cavallaro G, Lazzara G, Massaro M, et al. Biocompatible poly (N-isopropylacrylamide)-halloysite nanotubes for thermo responsive curcumin release. J Phys Chem C. 2015;119(16):8944–8951. doi:10.1021/acs.jpcc.5b00991
300. Massaro M, Piana S, Colletti CG, et al. Multicavity halloysite nanotubes for antimicrobial packaging system. Mater Res Bull. 2015;69:159–170. doi:10.1016/j.mrsb.2015.03.050
301. Chamarro-Garcia A, Merkoçi A. Nanobiosensors in diagnostics. Nanobiomedicine. 2016;3:1849543516663574. doi:10.1177/1849543516663574.
304. Iannazzo D, Piperno A, Pistone A, Grassi G, Galvagni S. Recent advances in cancer nanotherapeutics as delivery systems for anticancer drugs. Curr Med Chem. 2013;20(11):1333–1354. doi:10.2174/0929867311320110001

305. Ng CM, Loh H-S, Muhoosamy K, Sridevi N, Manickam S. Conjugation of insulin onto the sidewalls of single-walled carbon nanotubes through functionalization and diimide-activated amidation. Int J Nanomed. 2016;11:1607–1614. doi:10.2174/1877051X16666170942153109

306. Floyed EL, Sapag K, Oh J, Lunga CT. Photothermal desorption of single-walled carbon nanotubes and coconut shell-activated carbons using a continuous light source for application in air sampling. Ann Occup Hgy. 2014;58(7):877–888. doi:10.1093/annhyg/meu043

307. Jha PK, Jha RK, Raut D, Gnanasekar S, Rana SV, Hossain M. Potential targetability of multi-walled carbon nanotube loaded with silver nanoparticles photosynthesized from Ocmium tenuiflorum (tulsi extract) in fertility diagnosis. J Drug Target. 2017;25(7):616–625. doi:10.1080/1061186X.2017.1306534

308. Azandaryani AH, Kashanian S, Derakhshandeh K. Folate conjugated hybrid nanocarrier for targeted luteolysis delivery in breast cancer treatment. Pharm Res. 2017;34(12):2798–2808. doi:10.1007/s11195-017-2260-x

309. Date T, Nimbalkar V, Kamal A, Mittal A, Mahato RI, Chitkara D. Lipid-polymer hybrid nanocarriers for delivering cancer therapeutics. J Control Release. 2017;271:60–73. doi:10.1016/j.jconrel.2017.12.016

310. Sedki M, Khalil IA, El-Sherbiny IM. Hybrid nanocarrier system for guiding and augmenting simvastatin cytotoxic activity against prostate cancer. Artif Cells Nanomed Biotechnol. 2018;46:1–10.

311. Lim WQ, Phaa SZ, Xu HV, Sreejith S, Zhao Y. Recent advances in multi-functional silica-based hybrid nanocarriers for bioimaging and cancer therapy. Nanoscale. 2016;8(25):12510–12519. doi:10.1039/C5NR07853A

312. Zhang J, Han X, Li X, et al. Core-shell hybrid liposomal vesicles loaded with panax notoginsenoside: preparation, characterization and protective effects on global cerebral ischemia/reperfusion injury and acute myocardial ischemia in rats. Int J Nanomed. 2012;7:4299–4310. doi:10.2174/1877051X128035

313. Stanley S. Biological nanoparticles and their influence on organisms. Curr Opin Biotechnol. 2014;28(6):69–74. doi:10.1016/j.copbio.2013.11.014

314. Loredo-Tovias M, Duran-Meza AL, Villagran-Escareño MV, et al. Encapsulated ultrasmall lipopolyphosphates as novel nanocarriers for highly hydrophobic anticancer drugs. Nanoscale. 2017;9(32):11625–11631. doi:10.1039/C7NR02118F

315. Singh P, Kim YJ, Zhang D, Yang DC. Biological synthesis of nanoparticles from plants and microorganisms. Trends Biotechnol. 2016;34(7):588–599. doi:10.1016/j.tibtech.2016.02.006

316. Gregory AE, Tithall R, Williamson D. Vaccine delivery using nanoparticles. Front Cell Infect Microbiol. 2013;3(13):1–13.

317. Shukla S, Wen AM, Ayat NR, et al. Biodistribution and clearance of a filamentous plant virus in healthy and tumor-bearing mice. Nanomedicine. 2014;9(2):221–235. doi:10.2217/nmm.13.75

318. Wen AM, Lee KL, Yildiz I, Bruckman MA, Shukla S, Steinmetz NF. Viral nanoparticles for in vivo tumor imaging. J Vis Exp. 2012;69:1–12.

319. Leong HS, Steinmetz NF, Ablack A, et al. Intravital imaging of embryonic and tumor neovascularature using viral nanoparticles. Nat Protoc. 2010;5(8):1406–1417. doi:10.1038/nprot.2010.103

320. Lee KL, Murray AA, Le DH, et al. Combination of plant virus nanoparticle-based in situ vaccination with chemotherapy potentiates antitumor response. Nano Lett. 2017;17(7):4019–4028. doi:10.1021/acs.nanolett.7b00107

321. Badri Narayanan K, Sood Han S. Genetic modifications of icosahedral plant virus-based nanoparticles for vaccine and immunotherapy applications. Curr Protein Pept Sci. 2017;18(11):1141–1151. doi:10.2174/138920371866617042153109

322. Qi J, Zhuang J, Wu W, et al. Enhanced effect and mechanism of water-in-oil microemulsion as an oral delivery system of hydroxysoflavone yellow A. Int J Nanomed. 2011;6:985–991. doi:10.2147/IJN.S18821

323. Bhargava K, Conti DS, da Rocha SR, Zhang Y. Application of an oregano oil nanoemulsion to the control of food-borne bacteria on fresh lettuce. Food Microbiol. 2015;47:69–73. doi:10.1016/j.fm.2014.11.007

324. Sercombe L, Vearati T, Moheimani F, Wu SY, Sood AK, Hua S. Advances and challenges of liposome assisted drug delivery. Front. Pharmacol. 2016;7:286. doi:10.3389/fphar.2015.00286

325. Hsu CY, Wang PW, Alalaiwe A, Lin ZC, Fang JY. Use of lipid Nanocarriers to improve Oral delivery of vitamins. Nutrients 2019;11(1):68–97.

326. Agnihotri N, Soni GC, Chanchal DK, Tiwari S, Fang JY. A Scientific Review On Nanomulsion For Targeting Drug Delivery System. Int J Life Sci Res. 2019;5(2):16–29.

327. Héraut N, Wagner J, Abram SL, et al. Silver-Containing Titanium Dioxide Nanocapsules for Combating Multidrug-Resistant Bacteria. Int J Nanomed. 2020;15:1267–1281.

328. Dou XQ, Wang H, Zhang J, Aptamer–drug conjugate: targeted delivery of doxorubicin in a HER3 aptamer-functionalized liposomal delivery system reduces cardiotoxicity. Int J Nanomed. 2018;13:763–776.

329. Kevadiya BD, Chen L, Zhang L, Thomas MB, Davé RN, et al. FenoFibrate Nanocrystal Composite Microparticles for Intestine-Specific Oral Drug Delivery System. Pharmaceuticals. 2019;12(3):109–124.

330. Quan G, Niu B, Singh V, et al. Supersaturable solid self-micro-emulsifying drug delivery system: precipitation inhibition and bioavailability enhancement. Int J Nanomed. 2017;12:880–8811.

331. Lee JW, Lee HY, Park SH, et al. Preparation and evaluation of dexamethasone-loaded electroporph nanofiber sheets as a sustained drug delivery system. Materials. 2016;9(3):175–186.

332. Jin X, Zhang ZH, Sun E, et al. Enhanced oral absorption of 20 (S)-protopanaxadiol by self-assembled liquid crystalline nanoparticles containing piperine: in vitro and in vivo studies. Int J Nanomed. 2013;8:641–652.

333. Safer AM, Leporatti S, Jose J, Soliman MS, et al. Conjugation Of EGCG And Chitosan NPs As A Novel Nano-Drug Delivery System. Int J Nanomed. 2019;14:8033–8046.

334. Muhammad Maila A, Abdul-Razaq K, Dannaigoro A, et al. Cockle Shell-Derived Calcium Carbonate (Aragonite) Nanoparticles: A Dynamoite to Nanomedicine. Appl Sci. 2019;9(14):2897–2922.

335. Kamal N, Kochkodan V, Zecri A, Azhi S et al. Polysulfone Membranes Embedded with Halloysites Nanotubes: Preparation and Properties. Membranes 2020;10(1):2–29.