Vitamin B12 and folate levels in healthy Swiss senior citizens: a prospective study evaluating reference intervals and decision limits

Martin Risch¹*, Dominik W. Meier², Benjamin Sakem², Pedro Medina Escobar², Corina Risch², Urs Nydegger² and Lorenz Risch³

Abstract

Background: The vitamin B12 and folate status in nonanaemic healthy older persons needs attention the more so as decrease in levels may be anticipated from reduced haematinic provision and/or impaired intestinal uptake.

Methods: A total of 1143 subjectively healthy Swiss midlands participants (637 females and 506 males), ≥60 years of age were included in this study. Levels of vitamin B12, holotranscobalamin (holoTC), methylmalonic acid (MMA), homocysteine (Hcy), serum folate, red blood cell (RBC) folate were measured. Further, Fedosov’s wellness score was determined. Associations of age, gender, and cystatin C/creatinine-based estimated kidney function, with the investigated parameters were assessed. Reference intervals were calculated. Further, ROC analysis was done to assess accuracy of the individual parameters in recognizing a deficient vitamin B12 status. Finally, decision limits for sensitive, specific and optimal recognition of vitamin B12 status with individual parameters were derived.

Results: Three age groups: 60–69, 70–79 and ≥80 had median B12 (pmol/L) levels of 237, 228 and 231 respectively (p = 0.22), holoTC (pmol/L) of 52, 546 and 52 (p = 0.60) but Hcy (μmol/L) 12, 15 and 16 (p < 0.001), MMA (nmol/L) 207, 221 and 244 (p < 0.001). Hcy and MMA (both p < 0.001), but not holoTC (p = 0.12) and vitamin B12 (p = 0.44) were found to be affected by kidney function. In a linear regression model Fedosov’s wellness score was independently associated with kidney function (p < 0.001) but not with age. Total serum folate and red blood cell (RBC) folate drift apart with increasing age: whereas the former decreases (p = 0.01) RBC folate remains in the same bandwidth across all age groups (p = 0.12). A common reference interval combining age and gender strata can be obtained for vitamin B12 and holoTC, whereas a more differentiated approach seems warranted for serum folate and RBC folate.

Conclusion: Whereas the vitamin B12 and holoTC levels remain steady after 60 years of age, we observed a significant increment in MMA levels accompanied by increments in Hcy; this is better explained by age-related reduced kidney function than by vitamin B12 insufficiency. Total serum folate levels but not RBC folate levels decreased with progressing age.

Keywords: Age, Folate, Holotranscobalamin, Homocysteine, Glomerular filtration rate, Kidney function, Longitudinal study methylmalonic acid, Senescence, Reference interval, Vitamin B12
Background
The clinical significance of water soluble vitamin B12 (cobalamin) and of folate, and the importance of their routine analysis, is supported by recent findings in physiology regarding their function [1–8]. The simple yet biologically important role of support by these vitamins in catalysis of methyl group transfer is essential for life maintenance [9]. The function of folate overlaps with vitamin B12 both of them being essential in methylation reactions. With telomere shortening emerging as a research topic of senescence, it has been shown that the one-carbon metabolism pathway may affect telomere length through DNA-methylation [10].

Vitamin B12 deficiency later in life is estimated to affect 10 % of people over the age of 60 [11, 12]. Recent metaanalytic surveys [13, 14] confirmed the finding of vitamin B12-, and folate- deficiencies in elderly patients [15]. Optional but frequently practiced fortification of grain-based food supply is done with folic acid and vitamin B12 (www.blv.admin.ch).

Next to vitamin B12, folates are essential in eukaryotic cells for single carbon transfer reactions. Folate receptors transport their ligand via endocytosis [16] and jejunal folate resorption is of no clinical concern unless viral infection interferes [17]. In contrast the vitamin B12/gastric intrinsic factor cargo undergoes a risky trip down to ileal uptake involving vitamin B12 detachment from food proteins, and pancreatic protease scission of the vitamin B12-haptocorrin complex. There has recently been an upsurge of interest in the proper functioning of these interactions because of the extension of life expectancy and a concomitant increase in comorbidity, requiring updates in medical-geriatric check-ups [18]; early diagnosis in the elderly of vitamin deficiency is mandatory [19]. Some sources propose RBC folate a more stable indicator of folate sufficiency because of its intracellular confinement.

It is not enough to quantitatively estimate total vitamin B12 in order to test for vitamin B12 deficiency. It is true that MMA and Hcy are surrogate markers for vitamin B12 deficiency, and correctly indicate vitamin B12 failure to produce succinic acid and methionine. However, the effectiveness of these markers also depends on kidney function, which is now assessed by measuring the age-related impairment of glomerular filtration rate (eGFR: estimated equations) [20, 21]. In fact, kidney clearance of both MMA and Hcy is compromised in reduced kidney function [21].

Reference intervals (RI) for most laboratory analysis are now included by the providers of automatic instrument systems; their establishment is part of the brand and often calculated with serum banks of healthy blood donors in their work force age groups. In addition sex differences are issued only for relating analytes, mostly endocrinological.

It remains the responsibility of a laboratory to translate the RIs into good medical practice. RIs established with healthy sample donors from the region served by the medical laboratory need to be compared to those provided by the manufacturer of automated platforms and adjustments [22–24] made as required. The Swiss SENIORLAB study (www.seniorlabor.ch) is a study with over 1400 elderly participants allowing establishment of reference intervals of different analytes including vitamin D [25] and, analyzed here, vitamin B12/folate status.

Methods
Study population
Subjects were recruited from February 2009 to December 2011 in the context of the Swiss SENIORLAB study, which is an ongoing investigation in the canton of Berne (Switzerland) aimed at creating appropriate reference intervals (RI’s) of several analytes in the senior citizens (http://www.seniorlabor.ch). Subjectively healthy senior Caucasian volunteers aged 60 years and older were recruited. The study participants were contacted through newspaper advertisements, clubs and associations where there was a high probability that the membership would include healthy senior citizens (e.g., mountaineering clubs, sports clubs) and through personal contacts of those involved in organizing the study. A personal history of the patients was taken including specific information on medication and presence of several common diseases such as known cardiovascular disease, known diabetes mellitus, known thyroid disease, or known cognitive impairment. Anthropometric measurements were performed, and venous blood was drawn into S-Monovettes® (Sarstedt, Sevelen, Switzerland) after an overnight fasting period. In a longitudinal follow-up after baseline examination, all study participants were contacted and questioned on several issues regarding quality of life and survival. In the determination of reference intervals in elderly persons, survival can be employed to demonstrate a certain degree of health [26].

Of the 1467 participants, those with high C-reactive protein levels (>10 mg/L), vitamin B12-outliers (vitamin B12 >1100 pmol/L), holoTC-outliers (holoTC > 128 pmol/L), serum folic acid outliers (>45.3 nmol/L) or a hemoglobin suggesting anemia in the elderly (<110 g/L) [27] were excluded. Further exclusion criteria comprised the intake of proton pump inhibitors, antidiabetic medication, and vitamin B12 or folic acid supplementation as well as a clinical history suggesting the presence of cognitive impairment or a survival of less than one year (Fig. 1). Of the included study participants, 515 were between 60 and 69 years old, 435 between 70 and 79 years and 193 were 80 years old or older (Table 1). This study was in accordance with the Declaration of Helsinki and was approved by the cantonal institutional review board.
(KEK-Bern 166/08). All of the participants provided written informed consent.

**Biochemical measurements**

Analyses were performed on whole blood anticoagulated with EDTA, separated plasma therefrom or serum and kept at room temperature, on the same day of morning blood drawing. For CBC we use a Sysmex XE-5000 automated hematology analyzer (Sysmex, Horgen, Switzerland) providing 38 haematology values.

Total vitamin B12 and serum folic acid were assayed using the Abbott Architect i2000 analyzer (Abbott Diagnostics, Baar, Switzerland). Holotranscobalamin (holoTC) was assayed using an Axsym platform (Abbott Diagnostics, Baar, Switzerland). Because of restricted funds, only a partial sample (n = 1063) was analyzed on a SCIEX API 4000 LC-MS/MS system (AB Sciex, Brugg, Switzerland) for methyl malonic acid (MMA) [22, 28–32]. RBC folate was estimated using the relationship between red blood cell folate level, as measured with Beckman Unicel DxI 800 (Beckman Coulter, Nyon, Switzerland) and hematocrit. Creatinine was tested using an IDMS-standardized kinetic Jaffe method (Roche Diagnostics, Rotkreuz, Switzerland). The cystatin C levels and high sensitive CRP were determined using the nephelometric method with a Siemens Prospecc.

**Table 1** Baseline characteristics. The parameters given are associated with markers of vitamin B12 and folate acid status and are classified according to three different age groups. P-values are given for differences among the three different groups.

| Age group | Female/Male (n) | Hb (g/L) | MCV (fl) | Fedosov's ω | B12 (pmol/L) | holoTC (pmol/L) | MMA (nmol/L) | Hcy (μmol/L) | Serum folate (nmol/L) | Red cell folate (nmol/L) | Folate/B12 | eGFR (ml/min/1.73 m²) | Mentzer index |
|-----------|----------------|---------|----------|-------------|--------------|----------------|-------------|------------|-------------------|------------------------|-------------|----------------------|---------------|
| 60-69 yrs | 278/237        | 143.00  | 89.00    | 0.13        | 237          | 52.3           | 207         | 12.5       | 19.5              | 899                    | 83.4        | 90 (82, 98)          | 18.70         |
| 70-79 yrs | 247/188        | 142 (134, 149.00) | 90.00 (87.00, 92.00) | 0.02 (−0.30, 0.33) | 228 (180.00, 292) | 54.1 (39.3, 69.0) | 221 (179, 294) | 13.9 (11.6, 16.7) | 17.7 (12.9, 25.3) | 17.2 (13.0, 25.3) | 870 (669,1157) | 81.5 (55.9,112.0) | 82 (69, 89); n = 435 |
| ≥80 yrs   | 278/237        | 139 (131.00, 146.00) | 91.00 (88.00, 93.00) | −0.10 (−0.39, 0.22) | 231 (185, 301) | 51.8 (38.2, 67.3) | 244 (190, 326) | 15.5 (12.8, 19.9) | 17.2 (13.0, 25.3) | 17.2 (13.0, 25.3) | 837 (646, 1136) | 76.8 (57.3, 104.5) | 67 (54, 78); n = 193 |

P values for continuous variables:
- Table 1: **<0.001**
- Table 2: **<0.001**
- Table 3: **<0.001**

P values for proportion female/male:
- Table 1: **0.53**
- Table 2: **0.09**

Footnotes:
- Values are median and interquartile range (IQR) for continuous variables. n = number
- *Kruskal-Wallis test for continuous variables; chi square test for proportion female/male
instrument (Siemens Zurich, Switzerland). The α-amino acid homocysteine (Hcy) was measured using chemiluminescence on a Immulite 2000 instrument (Siemens, Zurich, Switzerland). The coefficient of variation in measuring the parameters of vitamin B12 and folic acid metabolism in our hands was 6.6 % for vitamin B12 (at a mean concentration of 162 pmol/L) 8.7 % for folic acid in serum (18.9 nmol/L), 6.8 % for holoTC (46 pmol/L), 9.3 % for Hcy (12.9 μmol/L), and for 1.8 % for MMA (390 nmol/mL). Estimated glomerular filtration rate (eGFR) was computed using the combined CKD-EPI version involving creatinine and corrected cystatin C level (Chronic Kidney Disease Epidemiology Collaboration) [20, 33, 34]. The Mentzer index was calculated by dividing the RBC count into the MCV [35]. The Fedosov wellness quotient (ω, combined indicator of vitamin B12 status) was calculated according the formula: ω = log_{10}(holoTC_{n}) + log_{10}(B12_{n}) - log_{10}(MMA_{n}) - log_{10}(Hcy_{n}), where variables correspond to the combination of the variables into normalized concentrations; for example, holoTC_{n} = holoTC/ holoTC_normal [36, 37]. The normal concentrations were obtained from reference [36]. Accuracy and precision of our assays comply with external and internal quality control requirements of the Swiss commission for quality assurance in the medical laboratory (QUALAB).

Data analysis
We used descriptive statistical methods to define reference intervals and to cross-compare blood concentrations of different analyte values. To find a non-parametric association between selected analyte concentrations we sought for significance of Spearman rank order correlation. Reference intervals for total B12, holoTC, folate and RBC folate were calculated by means of the non-parametric method, as recommended by the CLSI guideline C28-A3c [38, 39]. The prevalence of vitamin B12 deficiency was assessed by Fedosov’s quotient [36]. The Kruskal-Wallis test was employed to evaluate differences in analyte concentrations among different age-groups (60–69, 70–79 and ≥80). Proportions were compared by chi-square test and chi-square test for trend. To characterize the diagnostic accuracy of the different analytes, receiver operating characteristic (ROC) curve analysis was performed; cutoff thresholds and ROC analysis of 4 separate markers of vitamin B12 deficiency (vitamin B12, holoTC, MMA, Hcy) was calculated assuming negative values ω < 0.5 point to vitamin B12 deficiency [36]. Analogously to Fedosov [35], we used ROC curve analysis to determine the optimum decision threshold for the individual markers as lower cut-off for evaluation of a vitamin deficiency. We also determined the cut-offs, where an individual marker possesses a sensitivity or specificity of at least 99 %. As a marker of diagnostic accuracy, the areas under the curve of individual markers were compared according to DeLong et al. [40] Linear regression analysis was also done, in order to investigate a relationship between the metabolites Hcy and MMA with kidney function, independent of vitamin concentrations. The computer program Medcalc version 14.12.0 (Mariakerke, Belgium) was used for statistical calculations. Graphs were constructed by Medcalc and Graphpad Prism version 5.04 (GraphPad Software, La Jolla, USA).

Results

Vitamin B12 and surrogate markers
Spearman rank correlations revealed statistically insignificant associations between vitamin B12 or holoTC and progressing years of age (p = 0.56 and p = 0.68). Total vitamin B12 serum levels and holoTC in the entire cohort thus remained stable over three arbitrarily defined age groups of 60–69, 70–79 and ≥80 year (Table 1). However, there were significant sex differences of holoTC in the age groups of 60–69 (p < 0.001), 70–79 years (p < 0.001), whereas no such differences could be detected in persons ≥80 years (p = 0.77) (Fig. 2 and Fig. 3). Therefore, a common reference interval for elderly persons could be determined for persons aged ≥80 year (21.3 (90 % confidence interval, CI, 19.3-24.0) – 118.2 (90 % CI 97–125.3) pmol/L), whereas...
showed a significant trend (p < 0.001 for both) as follows: 8.0 % (37/426), 13.4 % (55/356) and 19.0 % (35/194).

There was a statistically significant increase in serum concentrations of Hcy and MMA with advancing age (p < 0.001 for both analytes); because it is known that these analytes pass the glomerular barrier, their senescent increment depends on eGFR, which significantly deteriorated over the three age groups (p < 0.001) (Table 1). Additional file shows this in more detail [Additional file 1]. Using a linear regression model employing age, serum folate, vitamin B12, gender and eGFR as independent and Hcy or MMA as dependent variables revealed a highly significant (p < 0.001) inverse relationship of both Hcy and MMA with eGFR. It should be noted that both vitamin B12 (p = 0.44) and holoTC (p = 0.12) were unrelated to eGFR in univariate analysis but also in a regression model adjusting for age and gender [Additional file 2].

The Fedosov wellness quotient calculated from the four vitamin B12 dependent analytes exhibited a statistically weak inverse correlation with progressing age in univariate analysis (r = −0.153, p < 0.001) [41]. However, a linear regression model (R² = 0.95) incorporating MMA, Hcy, holoTC, vitamin B12, eGFR and gender as independent and Fedosov’s wellness quotient as dependent variable reveals the four parameters of vitamin B12 status as well as kidney function to be independent predictors (p < 0.001 for all) of Fedosov’s wellness score. Age (p = 0.97) and gender (p = 0.59) did not have a significant association in this model.

To scrutinize the diagnostic accuracy of vitamin B12 dependent analyses, we performed a ROC analysis (Fig. 2). It appears that the largest area under the curve (AUC) is seen with holoTC: 0.923 (95 % CI 0.905-0.938), followed by MMA: 0.916 (95 % CI 0.897-0.932), followed by vitamin B12: 0.884 (95 % CI 0.863-0.903); Hcy reached an AUC value of 0.807 (95 % CI 0.782-0.831). A comparison of the AUC revealed holoTC to display significantly better diagnostic characteristics than vitamin B12 (p = 0.03) and homocysteine (p < 0.001). HoloTc (p < 0.001), MMA (p < 0.001) and vitamin B12 (p = 0.009) had significantly better diagnostic characteristics than Hcy. There were no differences in AUC between holoTC and MMA (p = 0.67) as well as between MMA and vitamin B12 (p = 0.14). As obtained by ROC curve analysis, the three decisive cut-offs (cut-off with 99 % specificity, cut-off with 99 % sensitivity, optimum decision threshold) for the different markers were: >25.8 pmol/L (specificity 99.04 %; sensitivity 32.81 %), ≤57 pmol/L (specificity 99.22 %; sensitivity 46.06 %), ≤342 pmol/L (specificity 93.75 %; sensitivity 75.85 %) for holoTC; >485 nmol/L (specificity 99.04 %; sensitivity 32.81 %), ≥216 nmol/L (specificity 99.22 %; sensitivity 55.85 %), ≥284 nmol/L (specificity 85.94 %; sensitivity 82.55 %) for MMA; ≤31 pmol/L (specificity 99.04 %; sensitivity 29.69 %), ≥316 pmol/L (specificity 99.22 %; sensitivity 20.21 %), ≤194 pmol/L (specificity
for vitamin B12; >9.5 µmol/L (specificity 11.17 %; sensitivity 99.22 %), ≥26.8 µmol/L (sensitivity 9.38 %; specificity 99.04 %), ≥15.1 µmol/L (sensitivity 77.34 %; specificity 72.87 %) for Hcy. As an alternative to reference intervals, a 2 cut-off model indicating probable vitamin B12 deficiency, probable sufficient vitamin B12 status as well as a grey zone can be derived from these cut-offs. For the 4 investigated parameters, these cut-offs are given in a separate table (Table 2).

Together, the specific cut-offs of holoTC (25.8 pmol/L) and vitamin B12 (131 pmol/L) as determined by biochemical vitamin B12 status obtained by Fedosov’s wellness quotient and ROC curve analysis fall close to the 90 % CI of the lower limit of the combined reference interval for holoTC (21.9-24.0 pmol/L) and vitamin B12 (114-126 pmol/L).

Folate
The total serum folate decreased with increasing age (r = −0.08; p = 0.01). Further, significant sex-specific differences can be seen in the age groups from 60–79 (p < 0.0001) but not in individuals ≥80 years (p = 0.07). The sex-specific reference intervals in the age group 60–69 years are: 8.1 (90 % CI 6.3-8.8) – 40.8 (90 % CI 36.8-45.1) nmol/L for males and 9.6 (90 % CI 8.4-10.7) – 43.3 (90 % CI 42.3-44.6) nmol/L for females. In the age group 70–79 years, the sex-specific reference intervals are 6.7 (90 % CI 3.7-8.1) – 37.1 (90 % CI 34.5-44.1) nmol/L in males and 9.1 (90 % CI 7.4-9.8) – 43.9 (90 % CI 40.4-45.1) nmol/L in females. A combined reference interval for the age group ≥80 years can be determined at 8.0 (90 % CI 6.5 – 8.7) – 43.0 (90 % CI 39.2-45.1) nmol/L. A reference interval combining all gender and age strata for practical purposes reveals a lower limit of 8.2 (90 % CI 7.6-8.5) and an upper limit of 42.8 (90 % CI 41.4-43.8) nmol/L.

Interestingly, RBC folate remained stable over increasing age (r = −0.045; p = 0.12). There was a significant difference in RBC folate levels in females and males (p = 0.02). Accordingly, gender specific reference intervals for all age groups were calculated. The reference interval was 452 (90 % CI 396–503) – 1885 (90 % CI 1735–2045) nmol/L for males and 435 (90 % CI 399–485) – 2067 (1903–2331) nmol/L for females. Finally, a reference interval combining all gender and age strata for practical purposes reveals a lower limit of 444 (90 % CI 408–480) and an upper limit of 1934 (90 % CI 1869–2066) nmol/L.

As age increased, the folate/vitamin B12 ratio tended to decrease (r = −0.06; p = 0.049), an observation in line with the observation that serum levels of folic acid recede with age by contrast with vitamin B12. Serum folate and Hcy were inversely correlated (r = −0.485, <0.0001). As a matter of fact, such correlation was better than the one resulting from comparison of vitamin B12 and Hcy (r = −0.253, p < 0.0001) [Additional file 2]. Such an association was absent with RBC folate (r = 0.01; p = 0.58) [Additional file 2]. In a linear regression model incorporating age, sex and eGFR as independent and serum folate as dependent variable maintained the independent relationship of eGFR and serum folate (p = 0.03).

Discussion
Our study on subjectively healthy elderly Caucasian participants from a circumscribed geographical region shows for the first time that holoTC, but also vitamin B12 levels from 60 year onwards remain within RIs set by international expert panels with people from all ages [42, 43] whereas serum folate concentrations – though not RBC folate - recede to some extent. It could be shown that MMA, Hcy as well as Fedosov’s wellness quotient rather are associated with kidney function than with age. Finally, we were able to present reference intervals and decision limits for the different investigated markers in the elderly.

An overwhelming proportion of the literature implies that, with progressing age, vitamin B12 and folate levels in senior citizens fall short of currently validated RIs [44–48], a finding that is challenged by our data, obtained from participants apparently in good health and with a relatively uniracial background [49, 50]. Most other studies, on metaanalysis, reveal cohorts comprised of study subjects seeking treatment or already ill or even subject to multimorbidity [14, 47, 51, 52]. The recruitment of study subjects apparently devoid of cognitive impairment might further augment the proportion of those with maintained vitamin B12 and folate levels now known to impinge on cognitious performance [44, 45, 53–55]. With the recent discoveries regarding the indispensable functions of vitamin B12 and folate in living organisms [1, 2, 7], it comes as little surprise that vitamin B12 and, to a lesser extent, folate, maintain their levels throughout our life cycle or else the participating subjects would not have been able to

| Cut-offs indicating probable vitamin B12 deficiency, probable vitamin B12 sufficiency and a grey zone of diagnostic uncertainty. These cut-offs are given for four markers of vitamin B12 status and were obtained by ROC-analysis |
|-------------------------------------------------|-----------------|-----------------|
| Vitamin B12 (pmol/L) | <131 | 131 – 315 | ≥316 |
| Holotranscobalamin (pmol/L) | <25.8 | 25.8 – 56.9 | ≥57.0 |
| MMA (nmol/L) | >485 | 217 – 485 | ≤216 |
| Homocysteine (µmol/L) | >26.8 | 9.6 – 26.8 | ≤9.5 |
enter the study in advanced age under the classification 'healthy'.

This was a community-dwelling cohort and haematinic sufficiency [56] can be assumed among participants whose BMI was normal and who were omnivorous, well fed and unlikely to be vegetarians. Marginal vitamin B12 haematinic provision remains a topic in daily health care. Switzerland counts only around 20,000 lactovegetarians distributed over all age groups; inadequate dietary intake of these two vitamins cannot be regarded as a relevant issue in this study. The effect of food fortifications on the levels of vitamin B12 and folate could not be addressed. With the Canadian Health Measures Survey on healthy folate replete subjects, the prevalence of folate deficiency was close to zero while vitamin B12 deficiency (cutoff < 148 pmol/L) came up to 5 % [43] but without effect of advanced age in older adults (60–79 years). In Switzerland, intentional fortification of food with vitamin B12 and folic acid is not enforced by law unless vitamin supplementation is a recommended component of pregnancy care. We thus might assume that dietary intake by the healthy elderly studied here is adequate.

The drop in folate but not RBC folate levels observed here may be linked to an intracellular maintenance of folic acid entrapped in RBCs but no full explanation can be forthcoming until half-life studies in both body compartments, plasma and RBC, have been conducted. Some authors hold that free and RBC folate are equivalent [57] and guidelines from the UK judge serum folate measurement as sufficient for clinical purpose [58]. Some are of the opinion that estimation of RBC folate is required to appreciate general tissue folate supply; should the latter view prevail for interpretation of our observations, then increasing age at least is not connected to folate deficiency.

In line with acknowledged understanding, our study links the surrogate markers of vitamin B12 status, i.e., MMA & Hcy, more closely to waning kidney function than to age. It is widely confirmed that with advancing age and particularly over age 50, functional kidney performance declines by > 1 ml/min/year/1.73 m² eGFR [59–61]. Our cohort confirms receding kidney function with progressing age ≥60 years using state-of-the art evaluation with creatinine and corrected cystatin C levels [20]. Thus logic would dictate that there are limits to applicability of MMA and Hcy as surrogate markers in geriatrics. Unhelpfully, quite a few studies have used claims of high levels of MMA and Hcy to underscore their findings of vitamin B12/folate reductions without testing for impaired kidney function, which is now well acknowledged, to increase MMA and Hcy levels [62].

It is noteworthy that significantly lower, but still normal, vitamin B12 median levels emerge in elderly males by comparison with females in the 60–79 years age range. Although no such sex difference showed up in a Canadian-HMS cycle 1 [43], it was observed that elderly males clearly were at higher risk for Vitamin B12/folate deficiency in a NHANES survey on 1770 elderly persons [47] and in our study women maintain sufficient vitamin B12/folate levels from their reproductive life cycle onwards.

In our study we have seen an increment in MCV with increasing age. Although the index values steadily increase by merely 1 fl from the youngest to the oldest age group, the difference is statistically significant because of the narrow distribution. This observation confirms previous reports indicating a link between macrocytosis and poorer age-related cognitive performance [18]; the selection criteria prior to study entry used here preclude such an association in the cohort studied here [52]. Out of curiosity, we also estimated the Mentzer index, now undergoing a revival as a test for safety of stem cell apheresis from healthy donors [35]. The values here obtained confirm those observed with MCV and in our case served as a validity criterion for our observations.

In an earlier report on a small subsample of the present cohort, we reported a borderline difference of eGFR in participants with low and normal holoTC [38]. In line with other investigations, we now could demonstrate that holoTC is not influenced by kidney function [63]. The fact that we found holoTC to possess the highest accuracy in recognizing an insufficient vitamin B12 status together with the independence of the parameter from reduced kidney function (a problem often occurring in the elderly) and from age seems to make holoTC an attractive first line choice for assessing vitamin B12 status in the elderly.

There are several possibilities to interpret markers of vitamin B12 and folate status. The most often used approach is the comparison with reference intervals. We were able to report reference intervals for all relevant parameters relating to the investigation of vitamin B12 and folate status in the elderly. Each of the reported stratum provides results from far more than 120 subjects, a lower limit generally accepted for evaluation of valid reference intervals [39]. Reference intervals have been reported from substantially smaller collectives [22]. The present investigation to the best of our knowledge is the largest collective of elderly persons, where reference intervals for vitamin B12 and folate status have been derived in an integral approach.

Another way of interpreting laboratory results is the comparison with pathophysiological conditions. We have taken this approach with ROC curve analysis, which allows to describe areas of relative certainty (i.e., above or below two cut-offs) together with a greyzone. Interestingly, the lower limits of the reference intervals are similar to the specific but insensitive cut-offs. Applying classically evaluated reference intervals for assessment of vitamin B12
deficiency can thus be regarded as very specific and relatively insensitive. Within the difficult context of diagnosing vitamin B12 and folate deficiency, we think that using an approach with decision limits provided in Table 2 is preferable to the use of reference intervals. Unfortunately, a functional score for detecting folate deficiency in analogy to the Fedosov wellness quotient in vitamin B12 deficiency is lacking so far.

Our study contains limitations:

(i) The medication and supplementation intake of the participants was self-reported. It might be that some patients had supplements and medications leading to erroneous inclusion of a participant in this analysis. We can therefore not rule out the possibility that a portion of our participants have normal vitamin B12 levels because supplemented. As impaired cognition was an exclusion criterion, it can be assumed that the number of affected individuals is low.

(ii) It is unfortunate that the questionnaire for study subjects did not include more details on eating habits

Conclusions
Whereas the vitamin B12 and holoTC levels remain steady after 60 years of age, we observed a significant increment in MMA levels accompanied by increments in Hcy; the latter is better explained by age-related reduced kidney function than by vitamin B12 insufficiency. Total serum folate levels but not RBC folate levels decreased with progressing age. The present work evaluated decision limits for the use of these parameters in the elderly.

Additional files
Additional file 1: Estimated glomerular filtration rate of the individuals studied. A statistically significant reduction (<0.001) in eGFR levels with increasing age was observed.

Additional file 2: Association of folate, vitamin B12 and related markers with kidney function (eGFR). The 6 panels show the serum concentrations of each of vitamin B12 (r=0.02, p=0.44), holoTC (r=-0.04, p=0.16), MMA (r=-0.29, p=0.001), Hcy (r=-0.46, p<0.001), serum folate (r=0.10, p<0.001) and RBC folate (r=0.01, p=0.58) as compared to eGFR plotted independently of age.

Abbreviations
AUC: Area under the curve; CI: confidence interval; ECL: electronic chemiluminescence; GFR: glomerular filtration rate; Hcy: homocysteine; holoTC: holotranscobalamin; MCV: mean corpuscular volume; MMA: methylmalonic acid; OTC: over the counter (to denote: nonprescription drugs); PPI: proton pump inhibitor; RBC: Red Blood Cells; Ri: Reference interval; ROC: Receiver operating characteristic.

Competing interests
The authors declare that they have no competing interests.

Authors contributions
DM, BS, MR and LR made substantial contributions in analysis of data and designing the figure material.

Acknowledgement
The work of Mrs Elisabeth Lenggenhager as study nurse is gratefully acknowledged. The authors acknowledge the linguistic and editorial improvements made by Mr. Nigel Lindup (BA Oxon.; retired UN translator), Geneva, Switzerland. The final text is the sole responsibility of the authors.

References
1. Roman-Garcia P, Quiros-Gonzalez I, Mottsam L, Lieben L, Sharan K, Wangwivatpin A, et al. Vitamin B12-dependent taurine synthesis regulates growth and bone mass. J Clin Invest. 2014; doi:10.1172/JCI72606.
2. Fenech M. Folate (vitamin B9) and vitamin B12 and their function in the maintenance of nuclear and mitochondrial genome integrity. Mutat Res. 2012;733(1–2):21–33.
3. Sanghoon Shin D, Hee Min S, Russell L, Zhao R, Fiser A, Goldman D. Functional roles of aspartate residues of the proton-coupled folate transporter (PCFT-SLC46A1): a DIS3L1 mutation causing hereditary folate malabsorption. Blood. 2010;116(24):5162–9.
4. Nielsen MJ, Rasmussen MR, Andersen CB, Nexø E, Moestrup SK. Vitamin B12 transport from food to the body’s cells - a sophisticated, multistep pathway. Nat Rev Gastroenterol Hepatol. 2012;9(6):345–54.
5. Li Z, Ghezamin C, Lesniale NA, Baneree R. Glutathione-dependent one-electron transfer reaction catalyzed by a B12 trafficking protein. The Journal of biological chemistry. 2014;289(23):16487–97.
6. Padmanabhan M, Geary-Joo C, Wu X, Ferguson-Smith AC, Fung E, Bieda MC, et al. Mutation in folate metabolism causes epigenetic instability and transgenerational effects on development. Cell. 2013;155(1):81–93.
7. Gos M, Specht-Procka A. Genetic basis of neural tube defects. II Genes correlated with folate and methionine metabolism J Appl Genet. 2002;43(4):S11–24.
8. Quadros EV, Nakayama Y, Sequeira JM. The protein and the gene encoding the receptor for the cellular uptake of transcobalamin-bound cobalamin. Blood. 2009;113(1):186–92.
9. Kung Y, Ando N, Doukov T, Blasiak IC, Bender G, Servalli J, et al. Visualizing molecular juggling within a B12-dependent methyltransferase complex. Nature. 2012;494(7393):265–9.
10. Holohan B, Wright WE, Shaj JW. Telomeropathies: An emerging spectrum disorder. Cell biology of disease JCB. 2014;205:289–99.
11. Meziere A, Audureau E, Vairelles S, Krypck S, Docko M, Morie M, et al. B12 deficiency increases with age in hospitalized patients. A study on 14,904 samples. J Gerontol A Biol Sci Med Sci. 2014;69(12):1576–82.
12. Baik HW, Russel. Vitamin B12 deficiency in the elderly. Ann Rev Nutr. 1999;19:357–77.
13. Carmel R. Biomarkers of cobalamin (vitamin B12) status in the epidemiologic setting. a critical overview of context, applications, and performance characteristics of cobalamin, methylmalonic acid, and holotranscobalamin II, Am J Clin Nutr. 2011. p. 1–11. doi:10.3945/ ajcn.111.013441.
14. Dullemeijer C, Souverein OW, Doets EL, van der Voet H, van Wijngaarden JP, de Boer WJ, et al. Systematic review with dose-response meta-analyses between vitamin B-12 intake and European Micronutrient Recommendations Aligned’s prioritized biomarkers of vitamin B-12 including randomized controlled trials and observational studies in adults and elderly persons. Am J Clin Nutr. 2013;97(2):390–402.
15. Andres E, Luklii MH, Noel E, Kaltenbach G, Abdelgheni MB, Perrin AE, et al. Vitamin B12 (cobalamin) deficiency in elderly patients. CMAJ. 2004;171(3):251–9.

16. Wilbobo AS, Singh M, Reeder KM, Carter JJ, Kovach AR, Meng W, et al. Structures of human folate receptors reveal biological trafficking states and diversity in folate and antifolate recognition. Proceedings of the National Academy of Sciences of the United States of America. 2013;110(38):15180–8.

17. Xiao S, Tang Y, Khan R, Zhang Y, Kusumachip S, Stabler SP, et al. Inference of physiologic folate deficiency on human papilloma type 16 (HPV16)-harboring human keratinocytes in vitro and in vivo. The Journal of biological chemistry. 2012;287(12):7599–77.

18. Gamaldo AA, Ferrucci L, Rifkind J, Longo DL, Zonderman AB. The relationship between mean corpuscular volume and cognitive performance in older persons. J Am Geriatr Soc. 2013;61(1):84–9.

19. Palacios G, Solà R, Barrios L, Pietrzik K, Castillo MJ, Gonzalez M. Algorithm for the early diagnosis of vitamin B12 deficiency in elderly people. Nutrition Hospital. 2013;28(1):447–52.

20. Inker LA, Schmid CH. Estimating glomerular filtration rate from serum creatinine and cystatin C. New Engl J Med. 2012;367(1):20–9.

21. Lewerin C, Ljungman S, Nilsson-Ehle H. Glomerular filtration rate as measured by serum cystatin C is an important determinant of plasma h homocysteine and serum methylylmethacetic acid in the elderly. J Intern Med. 2007;261(1):65–73.

22. Valente E, Scott JM, Ueland PM, Cunningham C, Casey M, Mollon AM. Diagnostic accuracy of holotranscobalamin, methylylmethacetic acid, serum cobalamin, and other indicators of tissue vitamin B12 status in the elderly. Clinical chemistry. 2015;57(6):856–63.

23. Katayev A, Fleming JK. Patient results and laboratory test accuracy. J Health Care Qual Assur. 2014;27(1):165–70.

24. Katayev A, Balizca C, Seccombe DW. Establishing reference intervals for human serum vitamin B12. Am J Clin Pathol. 2010;133:180–6.

25. Sakem B, Nock C, Stanga Z, Medina P, Nydegger UE, Risch M, et al. Serum concentrations of 25-hydroxvitamin D and immunoglobulins in an older Swiss cohort: results of the Senior Labour Study. BMC medicine. 2013:11:176.

26. Yourman LC, Lee SJ, Schonberg MA, Widera EW, Smith AK. Prognostic indices for older adults: a systematic review. Jama. 2012;307(2):182–92.

27. Culleton BF, Manns BJ, Zhang J, Tonelli M, Klarenbach S, Hemmelgarn BR. Impact of anemia on hospitalization and mortality in elderly. Blood. 2006;107(10):3841–6.

28. Schmedes A, Brandslund I. Analysis of methlymalonic acid in plasma by liquid chromatography-tandem mass spectrometry. Clinical chemistry. 2011;57(12):1915–23.

29. Fedosov SN, Brito A, Miller JW, Green R, Allen LH. Combined indicator of metabolic signs of vitamin B12 deficiency in humans: analytical aspects and clinical utility, Am J Clin Nutr. 2011. p. 1S.

30. Fedosov SN, Metabolic signs of vitamin B12 deficiency in humans: a nonparametric approach. Biometrics. 1988;44:3837–45.

31. Wang TF, Chen SH, Yang SH, Su YC, Chu SC, Li DK. Poor harvest of peripheral nerve terminal function and vitamin B12 status in the elderly by gender and ethnicity. J Natl Med Assoc. 2011;103(10):870–7.

32. Andres E, Lukili MH, Noel E, Kaltenbach G, Abdelgheni MB, Perrin AE, et al. Vitamin B12 deficiency: modification for missing biomarkers and folate status. BMC Geriatrics 2015;15:82.

33. Fedosov SN, Metabolic signs of vitamin B12 deficiency in humans: a nonparametric approach. Biometrics. 1988;44:3837–45.

34. Segarra A, DelaTorre J, Ramos N, Quiroz A, Garjau M, Torres I, et al. Low vitamin b12 status and risk of cognitive decline in older adults. J Am Clin Nutr. 2007;86:1284–91.

35. Risch C, Medina P, Nydegger UE, Bahador Z, Brinkmann T, Von Landenberg P, et al. The relationship of leucocyte anisoctosis to holotranscobalamin, a marker of cobalamin deficiency: Int J Lab Hematol. 2012;34(2):192–200.

36. CCS Defining, Establishing, and Verifying Reference Intervals in the Clinical Laboratory, Approved Guideline, Third edition CLSI document C28-A3c. Clinical and Laboratory Standards Institute: Wayne PA, 2008.

37. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. Biometrics. 1988;44:837–45.

38. Risch C, Medina P, Nydegger UE, Bahador Z, Brinkmann T, Von Landenberg P, et al. The relationship of leucocyte anisoctosis to holotranscobalamin, a marker of cobalamin deficiency: Int J Lab Hematol. 2012;34(2):192–200.

39. CCS Defining, Establishing, and Verifying Reference Intervals in the Clinical Laboratory, Approved Guideline, Third edition CLSI document C28-A3c. Clinical and Laboratory Standards Institute: Wayne PA, 2008.

40. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. Biometrics. 1988;44:837–45.

41. Xiao S, Tang Y, Khan R, Zhang Y, Kusumachip S, Stabler SP, et al. Inference of physiologic folate deficiency on human papilloma type 16 (HPV16)-harboring human keratinocytes in vitro and in vivo. The Journal of biological chemistry. 2012;287(12):7599–77.

42. Fedosov SN, Metabolic signs of vitamin B12 deficiency in humans: a nonparametric approach. Biometrics. 1988;44:3837–45.

43. Wang TF, Chen SH, Yang SH, Su YC, Chu SC, Li DK. Poor harvest of peripheral nerve terminal function and vitamin B12 status in the elderly by gender and ethnicity. J Natl Med Assoc. 2011;103(10):870–7.

44. Koenig V, Stanga Z, Zerlauth M, Bernasoni L, Risch M, Huber A, et al. Prevalence of vitamin B12 deficiency and deficiency in Liechtenstein. Public Health Nutri. 2012;17(2):241–7.

45. Benoit K, Boudreau RM, Studenok SA, Newman AB. The relationship of vitamin B12 and sensory and motor peripheral nerve function in older. J Am Geriatr Soc. 2012;60:1057–1063.

46. CLSI. Defining, Establishing, and Verifying Reference Intervals in the Clinical Laboratory. Clinical and Laboratory Standards Institute: Wayne PA; 2008.

47. Fedosov SN. Biochemical markers of vitamin B12 deficiency combined in one diagnostic parameter: the age-dependence and association with cognitive function and blood hemoglobin. Clinica chimica acta; international journal of clinical chemistry. 2013:422:47–53.

48. Fedosov SN. Biochemical markers of vitamin B12 deficiency combined in one diagnostic parameter: the age-dependence and association with cognitive function and blood hemoglobin. Clinica chimica acta; international journal of clinical chemistry. 2013:422:47–53.

49. Segarra A, DelaTorre J, Ramos N, Quiroz A, Garjau M, Torres I, et al. Combined indicator of vitamin B12 status: modification for missing biomarkers and folate status and recommendations for revised cut-points, Clinical chemistry and laboratory medicine. CCLM / FESCC. 2015.
62. Lewerin C, Ljungman S, Nilsson-Ehle H. Glomerular filtration rate as measured by serum cystatin C is an important determinant of plasma homocysteine and serum methylmalonic acid in the elderly. J Intern Med. 2007;261(1):65–73.

63. Lewerin C, Nilsson-Ehle H, Jacobsson S, Karlsson MK, Ohlsson C, Mellstrom D. Holotranscobalamin is not influenced by decreased renal function in elderly men: the MrOS Sweden study. Annals of clinical biochemistry. 2013;50(Pt 6):585–94.