MyD88 ADAPTER-LIKE (MAL)/TIRAP INTERACTION WITH TRAF6 IS CRITICAL FOR TLR2- AND TLR4-MEDIATED NF-κB PROINFLAMMATORY RESPONSES.

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ABSTRACT:
TIR-containing adapters are critical in orchestrating the different signal transduction pathways following Toll-like receptor (TLR) activation. MyD88 adapter-like (Mal), also termed TIRAP, is involved in bridging MyD88 to the receptor complex for TLR-2 and TLR4 signaling in response to bacterial infection. We have previously reported an interaction between Mal and TNF receptor associated factor (TRAF)-6 via a TRAF6-binding motif, the disruption of which inhibited TLR-mediated NF-κB-luciferase reporter activity. Given the recent report of intracellular TRIF-related adapter molecule (TRAM) localization promoting sequential signaling in TLR4 responses, we further characterized Mal interaction with TRAF6, the cellular localization and the outcomes of disrupting this association on TLR inflammatory responses. We found that Mal and TRAF6 directly interact in response to TLR2 and TLR4 stimulation, although membrane localization is not necessary to facilitate interaction. Critically, reconstitution of murine Mal-deficient macrophages with MalE190A, containing a mutation within the TRAF6-binding motif, fails to reconstitute the proinflammatory response to TLR2 and TLR4 ligands compared to wild type Mal. Furthermore, Mal interaction with TRAF6 mediates Ser phosphorylation of the p65 subunit of NF-κB and thus controls transcriptional activation, but not nuclear translocation of NF-κB.

This study characterizes the novel role for Mal in facilitating the direct recruitment of TRAF6 to the plasma membrane, which is necessary for TLR2- and TLR4-induced transactivation of NF-κB and regulation of the subsequent pro-inflammatory response.

INTRODUCTION
Toll-like receptors (TLRs) recognize and respond to both pathogen-associated molecular patterns (PAMPs) and endogenous signals associated with danger (1). Upon ligand-induced dimerization the TLRs activate, via their cytosolic Toll/IL-1 receptor (TIR) domains, the homotypic recruitment of one or more proteins of a family of five cytosolic TIR-containing adapter proteins (2). All TLRs, with the exception of TLR3, recruit MyD88 to their receptor complex; as do members of the IL-1 receptor family. MyD88 recruits Interleukin-1 receptor-associated kinase (IRAK)-1, IRAK4 and then TNF receptor associated factor 6 (TRAF6) that results in the nuclear translocation of the prototypic inflammatory transcription factor Nuclear Factor (NF)-κB, termed the canonical pathway (3,4) which encodes inflammatory genes such as TNF-α and interleukin (IL)-6. Although TLRs induce common signaling pathways, there is specificity in recruitment of TIR-containing adapter proteins. MyD88-adapter like (Mal)/TIRAP was the second described adapter...
capable of mediating NF-κB activation and responsible for signaling selectively via TLR4 (5,6) and TLR2 signaling (7,8). TIR-domain containing adapter protein inducing IFN-β (TRIF, also known as TICAM1) was subsequently found to mediate the MyD88-independent pathway leading to TLR4-mediated activation of the transcription factor IRF3, which regulates Type I IFN production (9). Importantly, TRIF also mediates downstream signaling from TLR3, independent of MyD88. TRIF-related adapter molecule (TRAM, also known as TICAM2) specifically acts to bridge TLR4 with TRIF, where TRAM-deficient macrophages are ablated in their responses to TLR4 activation, but not TLR3 (10,11).

Mal and TRAM have been described as bridging adapters, responsible for specific recruitment of MyD88 and TRIF proximal to the surface localized TLR2 and TLR4 receptor complexes (10,12,13). Membrane localization of these bridging adapters has further demonstrated the spatial coordination required for transmission of MyD88 (12) and TRAM (14) signals allowing specificity of downstream signaling mediators that induce cytokine production. Mal acts to bridge MyD88 to TLR2 and TLR4 specifically via its TIR domain. The recent description of a Mal functional variant associated with protection against pneumococcal disease, bacteremia, malaria and tuberculosis (15) has highlighted the potential importance of Mal in human disease. Furthermore, the recent depiction of the rare TIR domain variant, Mal D96N, which is unable to interact with MyD88 and impairs downstream TLR2/TLR4-mediated responses (16) has emphasized the importance of protein:protein interactions in TLR-mediated signaling and highlighted the need for a greater understanding of how Mal mediates TLR-induced inflammation.

We have previously demonstrated that Mal interacts with TRAF6 (17) via a putative TRAF6-binding motif (18) which when disrupted, inhibited TLR2- and TLR4-induced NF-κB activation. However the specificity and biological consequences of this interaction were not clearly defined. As disruption of Mal function appears to provide a protective phenotype from disease, we endeavored to investigate the specificity of Mal interaction with TRAF6, cellular localization and what the biological outcomes of disrupting this interaction were on TLR-mediated pro-inflammatory responses.

In this study, we demonstrate the direct interaction of Mal with TRAF6 and the localization of this association to the plasma membrane. Critically, reconstitution of Mal-deficient macrophages with wild type Mal reconstituted transactivation of the p65 subunit of NF-κB, which was ablated with disruption of the TRAF6-binding motif. Importantly, NF-κB translocation to the nucleus was not affected in these cells. Significantly, while Mal-deficient macrophages reconstituted with wild type Mal express the expected pro-inflammatory cytokines TNF-α and IL-6 in response to TLR2 and TLR4 stimulation, cells reconstituted with a single point mutation within the TRAF6-binding motif (MalE190A) display a significantly disrupted cytokine response to TLR stimulation. Together these studies characterize the novel role for Mal in facilitating the direct recruitment of TRAF6 to the plasma membrane, which is necessary for TLR2- and TLR4-induced transactivation of NF-κB and regulation of the subsequent pro-inflammatory response.

MATERIALS AND METHODS

Cell culture and reagents.

Cell culture - Human embryonic kidney (HEK) 293, HEK293T cells and murine immortalized Mal-deficient macrophages (16) were cultured in DMEM medium supplemented with 10% FCS (Invitrogen, Carlsbad, CA), 2 mM glutamine, 100 U/ml penicillin and maintained in a 37°C humidified atmosphere. Human monocyte THP-1 cells were maintained in RPMI 1640 medium 2 mM glutamine, 100 U/ml penicillin and maintained in a 37°C humidified atmosphere. LPS K235 (Sigma, St Louis, MO) was re-purified as described previously (19), Pam3Cys was obtained from EMC microcollections (Tuebingen, Germany), CL-75 (InvivoGen, San Diego, CA), CpG DNA oligonucleotide (GeneWorks, Adelaide, Australia), anti-Flag M2-agarose beads (Sigma) and Glutathione-Agarose beads (Amersham Biosciences). GST, TRAF6 and IκBα antibodies were sourced from Santa Cruz.

Plasmids and protein purification.
Full-length pDC304 Mal-HA, pDC304 Mal-HA N-terminal (1–74), pDC304 Mal-HA TIR domain (74–235) have been previously described (5). Previously described Mal variants: MalE190A-HA (17), Mal-GFP, Mal 4KK-GFP (PIP2 binding domain) and Mal (1-20aa)-GFP (first 20 amino acids) (12). MalS180L-HA, MalE190A-GFP and MalΔx3-GFP mutants were generated using the QuikChange II site-directed mutagenesis kit with pfu-Turbo (Stratagene, La Jolla, CA) using the pDC304 Mal-HA or Mal-GFP template respectively. Lentiviral vectors encoding the various Mal-GFP gene were generated by subcloning the GFP-tagged cDNA from pEGFP-N1 into pLV-CMV. Gal4-p65(1–551) plasmid encoding the full-length p65 subunit of NF-κB fused to Gal4 DNA-binding domain was a kind gift from Lienhard Schmitz (German Cancer Research Centre, Heidelberg, Germany). The Gal4-luciferase reporter gene pFA-Jun and pFA-Elk-1 fusion vectors for analysis of JNK and Erk1/2, respectively, were obtained from Stratagene.

Mal-GST and MyD88-GST purification have been described previously (20). MalE190A-GST purification was prepared the same as for Mal-GST. GST-TRAF6-Flag protein was transformed into BL21-CodonPlus™ -RIL series E. coli (Stratagene) and grown in Luria broth. A bacterial cell culture was grown to OD600 = 0.5 under ampicillin selection. Protein expression was induced with 50 μM IPTG and incubated O/N at 18ºC. Bacterial cells were harvested and resuspended in low salt soluble KalB buffer (50 mM Hpes, pH 7.6, 50 mM NaCl, 0.5 mM EDTA, 0.1 mM PMSF and 1 tablet of protease inhibitor cocktail (Roche, Mannheim, Germany). Cells were lysed by sonication, centrifuged and cell pellet resuspended in high salt soluble buffer (50 mM Hpes, pH 7.6, 500 mM NaCl, 0.5% Triton-X100, 0.5 mM EDTA, 0.1 mM PMSF and protease inhibitor cocktail (Roche)). Cell debris were separated by centrifugation (18,000 rpm, 4°C). Protein purification was facilitated by affinity purification on glutathione-Sepharose and removal of GST-fusion tag by PreScission™ (Amersham Biosciences) enzyme cleavage according to the manufacturer’s instructions.

**Immunoprecipitation, and immunoblot analysis.**

HEK293T cells (2 × 10^6 cells/10 cm dish) were transfected using Fugene 6 (Roche) with the indicated plasmids where the total amount of DNA (2.5 μg per dish) was kept constant. Twenty-four h later, cells were lysed in KalB buffer as described (21). The indicated antibodies (2 μg) or anti-Flag sepharose beads (20 μl, 50% slurry) were incubated with the cell lysates for 2 h, followed by the addition of 40 μl of 50% protein-G slurry for 1 h. The immune complexes were precipitated, washed, eluted by addition of sample buffer, followed by SDS-PAGE and immunoblotted using the indicated antibodies. For GST pull-down experiments, lysates prepared from HEK293T cells transfected with indicated vectors were used in a GST pull-down assay whereby cell lysates were incubated for 2 h at 4°C with recombinant GST fusion protein coupled to glutathione-Sepharose. Complexes were washed three times in lysis buffer, separated by SDS-PAGE, and immunoblotted as indicated in the figure legend. For endogenous immunoprecipitation, confluent THP-1 cells (2x10^6) were resuspended in 1ml conditioned media, centrifuged and harvested as per immunoprecipitation method.

Protein samples were separated by SDS-PAGE, transferred to nitrocellulose and immunoblotted with the indicated antibodies. Immunocomplexes were visualized by using SuperSignal West Pico chemiluminescent substrate solution (Pierce, Rockford, IL) followed by exposure to X-ray film (Hyperlum, Amersham Pharmacia) to detect chemiluminescence or visualized using the Odyssey fluorimager (LI-COR) and probed with AlexaFluor 680 (Molecular Probes) and IRDye800CW (Rockland, Gilbertsville, PA)) fluorescent labeled secondary antibodies.

**Confocal imaging analysis.**

HEK293 cells were co-transfected with TRAF6-Flag and fluorophore-conjugated proteins at the concentrations indicated for 48 h at 32°C. Cells were washed in PBS, fixed in 4% paraformaldehyde, and permeabilized in 0.1% Triton-X100. Cells were immunohistostained with anti-Flag antibody (Sigma) at 1:200 O/N at 4°C and probed with Alexa Fluor 594 (Texas Red) or Alexa Fluor 488 (Green)-conjugated IgG secondary antibody (Invitrogen). Mounted cells on coverslips were examined with 40X oil lens objective on Leica TCS NT upright confocal microscope (Leica Microsystems GmbH, Welzlau, Germany) using Leica TCS NT LASAF software.
Deconvolution and co-localization was performed using the IMARIS software (Bit-plane AG, Zurich Switzerland).

**Luciferase reporter assay.**

HEK293 cells were seeded at 2x10⁴ cells/well in a 96-well plate 24 h prior to transfection with Fugene 6 (Roche). NF-κB-dependent gene expression was determined using the 5x κB-luciferase reporter construct (Stratagene) concomitantly with indicated vectors. Using the PathDetect transient transfection kit (Stratagene), co-transfection of pFR-luciferase in combination with Gal4-fusions p65, pFA-Jun, pFA-CHOP or pFA-Elk-1, respectively, were used to analyze activation of p65 transactivation and MAP kinase respectively. The Renilla luciferase-thymidine kinase encoding plasmid (pRL-TK) (Promega, Madison, WI) was used to normalize for transfection efficiency, and pEF-BOS empty vector was used to maintain constant DNA. Transfected cells were lysed using Passive lysis buffer (Promega) and assayed for luciferase and Renilla activity using luciferase assay reagent (Promega). Luminescence readings were corrected for Renilla activity and expressed as fold increase over non-stimulated control values.

**Electrophoretic Mobility Shift Assay (EMSA).**

Nuclear extracts (4 μg) were incubated with 10,000 c.p.m. of a 22 bp DNA fragment oligonucleotide containing the NF-κB consensus sequence (Promega) that had previously been labelled with [γ-32P]-ATP (10 mCi mmol) (Perkin Elmer, Waltham, MA, USA) by T4 polynucleotide kinase (Promega). Incubations were performed for 30 min at room temperature in the presence of 2 μg of poly-(dL.dC) as non-specific competitor and 10 mM Tris-HCl, pH 7.5, containing 100 mM NaCl, 1 mM EDTA, 5 mM DTT, 4% glycerol and 100 mg/ml nuclease-free BSA. Samples were subjected to electrophoresis on native 5% (w/v) polyacrylamide gels, which were subsequently dried and autoradiographed.

**ELISA and RT-PCR.**

Macrophage supernatants from virally transduced cells were used for ELISA to detect expression of TNF-α or IL-6 with reagents from BD PharMingen (St Jose, CA); according to the manufacturer’s instructions. Total cellular RNA was prepared using RNeasy mini kit according to manufacturer’s instructions (QIAGEN, Hilden, Germany). Two μg of total RNA was treated with DNase (Promega) before cDNA synthesis using Superscript III (Invitrogen) and random primers in a volume of 40 μl. cDNA was diluted 1/3 and triplicate wells run for each sample on the 7900HT Fast Real-Time PCR system (ABI, Foster City, CA). Taqman® Gene Expression assays were conducted using premade kits from ABI and all samples were multiplexed with the endogenous control 18S ribosomal RNA. Relative gene expression was determined using the ΔΔCT cycle threshold method.

**Statistical analysis.**

For statistical analysis, the data obtained from independent experiments are presented as the means ±SD. All statistical analysis was performed using GraphPad PRISM software (GraphPad Software, Inc., San Diego, CA). Mean differences were calculated using Student’s t-test. Analysis of two independent variables was calculated using a two-way ANOVA. The levels of significance are denoted as follows: * P<0.05, ** P<0.01, *** P<0.001.

**RESULTS**

*Mal interaction with TRAF6 is induced by TLR2 and TLR4 stimulation.*

We have previously demonstrated that Mal interacted with TRAF6 in ectopic co-immunoprecipitation experiments; therefore we first wished to establish whether TLR stimulation induced association between the two proteins. As can be observed in Figure 1A, TRAF6 immunoprecipitated Mal in a time-dependent manner within 10-15 mins of TLR2 (Pam3Cys) stimulation of human THP-1 monocytic cells. TLR4 (LPS) activation also induced the same effect (Fig 1B), although with slightly delayed kinetics, peaking at 20 mins following stimulation. Importantly, TLR9 (CpG DNA) stimulation failed to induce Mal:TRAF6 interaction (Fig 1C) consistent with Mal having a specific role in TLR2 and TLR4 signaling. Degradation of IκBα was also observed in cells following stimulation confirming that the respective ligands had activated their relevant TLR. These results
demonstrate a specific interaction between Mal and TRAF6 following stimulation of TLR2 and TLR4, but not TLR9, consistent with the described role for Mal in TLR2 and TLR4 signal transduction.

**Mal Interaction with TRAF6 Occurs at the Plasma Membrane.**

Recently, Kagan et al demonstrated the critical role of Mal localization to the plasma membrane via an N-terminal PIP2 binding domain (12) in TLR2 and TLR4 signaling. Localization to the plasma membrane was required to facilitate MyD88 recruitment to the plasma membrane TLR receptor complex leading to initiation of the canonical signaling pathway and subsequent activation and nuclear translocation of NF-κB. As ectopic expression of Mal is able to localize to the membrane (12) and activate NF-κB (5,17) mimicking TLR stimulation, we wished to determine if localization Mal and TRAF6 occurs at the plasma membrane or the cytosol in HEK293 cells by fluorescence microscopy.

We found that GFP-tagged Mal localized to the plasma membrane by co-staining with Cholera toxin subunit B (CTxB) (Fig S1A). We next confirmed that Mal 4KK-GFP (kind gift Ruslan Medzhitov, Yale University) which contains point mutations of the PIP2-binding region (K15,16,32,33A) and therefore disrupts Mal membrane localization and Mal 1-20-GFP, consisting of the first 20 amino acids of Mal lacking the PIP2 membrane binding motif and functionality both displayed diffuse staining throughout the cell consistent with earlier reports (12) (Fig S1B and S1C respectively). Ectopic expression of Flag-tagged TRAF6 alone was found in discrete foci scattered throughout the cytosol and did not label the cell surface when compared to CTxB staining (Fig S1D).

To investigate if membrane localization was required to facilitate interaction between Mal and TRAF6 we compared wild type Mal association with TRAF6 to Mal 4KK-GFP, as this variant still contains an intact TRAF6-binding motif (Mal 188-193), but fails to localize to the membrane and Mal 1-20-GFP which lacks the PIP-, TIR- and TRAF6-binding domains. Initial 3D view image analysis of co-expressed wild type Mal, Mal 4KK and Mal 1-20 TRAF6 (Fig. 2- first panel) appears that all three Mal proteins are co-localizing with TRAF6 either at the plasma membrane (wild type Mal- Fig 2A) or in the cytosol (Mal 4KK and Mal 1-20- Fig 2B and 2C). However when 3D iso-surface rendition analysis is applied, where the intensities of the fluorescence of each tag are standardized to remove variance in fluorescence intensity between fluorophores, it becomes apparent that while Mal and Mal 4KK co-localize in the same plane as TRAF6 and therefore appear intensely co-localized (Fig 2A and 2B- final panel), Mal 1-20 is clearly not co-localizing in the same plane and therefore is not co-localized with TRAF6 (Fig 2C, middle and final panel). Similar co-localization of Mal and TRAF6 was observed of Mal tagged with DsRed and TRAF6-GFP (Fig S2A) or Mal-GFP and TRAF6-DsRed (Fig S2B) indicating that the co-localization at the plasma membrane did not result from the tag and represents a feature of Mal and TRAF6 association itself.

To further support these findings we found that both wild type Mal-GFP and Mal 4KK-GFP, but not Mal 1-20-GFP, were able to interact with TRAF6 in co-expression immunoprecipitation experiments (Fig 2D) confirming that the GFP tag did not affect the ability of Mal and TRAF6 to interact and that association occurs independent of cellular localization.

Having established that interaction was not dependent on localization, we next assessed the effects on downstream signaling. We have previously found that Mal interaction with TRAF6 mediated the activation of the MAP kinase pathway and importantly, transactivation of the p65 subunit of NF-κB (17). We therefore tested the ability of these GFP-tagged proteins to drive NF-κB activation (Fig 2E) compared to our previously reported results using HA-tagged Mal. Consistent with our localization and immunoprecipitation data, both wild type Mal and Mal 4KK were able to induce activation of NF-κB luciferase expression, while Mal 1-20 failed to induce a similar responses. Furthermore, consistent with our earlier report (17), wild type Mal and Mal 4KK were able to induce activation of NF-κB p65 transactivation, Erk1/2 and JNK activation (Fig 1F) via reporter assays but critically not p38. As expected, Mal 1-20 was unable to induce activation of any pathway due to its lack of PIP2 and TIR domains.
Taken together these results demonstrate that Mal and TRAF6 interact either at the plasma membrane or in the cytosol independent of localization to the plasma membrane suggesting the geographical constraint of Mal to the plasma membrane by PIP2 modification may be important for spatial signaling or kinetics. Alternatively, TRAF6 may have a higher preference, but not an absolute requirement for, association for the membrane bound Mal.

Disrupting the TRAF6-binding motif in Mal inhibits Mal:TRAF6 interaction.

The requirement of an intact Mal TRAF6-binding motif for interaction with TRAF6 was next determined. In our previous study we found that a single amino acid change of E190A in Mal inhibited TLR2 and TLR4 signaling, however, in ectopic immunoprecipitation studies, this mutation was still able to interact with TRAF6. Therefore we wished to determine a more specific assessment of the role of the critical glutamic acid residue in the TRAF6-binding motif in mediating Mal interaction with TRAF6.

To begin with we assessed Mal and TRAF6 association by immunomicroscopy. As shown in Fig. 3, while it appears that both Mal (Fig 3A) and MalE190A (Fig 3B) are both co-localizing with TRAF6 at the plasma membrane, 3D iso-surface rendition analysis shows that once the fluorescence intensities are standardized, it is apparent that Mal and TRAF6 are co-localizing in the same plane (Fig 3A- middle panel) and as such appear intensely co-localized at the plasma membrane (final panel). Conversely, 3D iso-surface rendition shows that MalE190A appears as discrete foci along the plasma membrane while TRAF6 remained diffuse in the cytosol and are not in the same plane (Fig 3B- middle panel) and as such do not co-localize (final panel). To further assess this interaction, we generated a mutation of all three key residues of the TRAF6-binding motif (PxExAr/Ac) (18) from PPELRF→QPAERA, termed MalΔx3, which interestingly, does not appear to co-localize with TRAF6 both 3D view or iso-surface rendition and as such do not co-localize demonstrating a lack of association between the proteins (Fig 3C). This protein also failed to display any discernable co-localization with TRAF6 (Fig 3C) as rendered by 3D iso-surface rendition co-localization, appearing primarily throughout the cytosol. Identical results were observed using GFP-tagged TRAF6 and DsRed-Mal constructs (data not shown) indicating that the localization and the importance of an intact TRAF6-binding motif for Mal interaction with TRAF6 were indicative of a function of Mal and not the result of the fusion tag to the proteins.

Having demonstrated an interaction between Mal and TRAF6 by microscopy, we next wished to further support the findings above and investigated the specificity of interaction and whether the association was dependent of an intact TRAF6-binding motif within Mal and independent of bridging proteins. As shown in Figure 4A, recombinant GST-Mal was able to specifically immunoprecipitate ectopically expressed recombinant TRAF6 from cellular lysates while recombinant GST alone was unable to recognize TRAF6. Conversely, recombinant GST-TRAF6 was able to precipitate ectopically expressed Mal from cellular lysates, but was unable to interact with MalE190A (Fig. 4B) clearly demonstrating the requirement of an intact TRAF6-binding motif to facilitate association. In order to demonstrate that the Mal interaction with TRAF6 was direct, we next co-incubated purified recombinant fusion proteins of TRAF6 tagged with Flag, in conjunction with the indicated recombinant TIR-containing proteins. As can be observed in Figure 4C, neither recombinant GST-MalE190A, GST-MyD88 nor GST alone, were able to be immunoprecipitated with TRAF6 while GST-Mal and TRAF6 markedly formed a complex.

Taken together, these results demonstrate that Mal and TRAF6 directly interact at the plasma membrane, the interaction dependent on an intact TRAF6-binding motif within Mal. Importantly, these findings also suggest the interaction between Mal and TRAF6 are independent of post-translational modification of either Mal or TRAF6 due to formation of a specific complex of recombinant proteins.

*Mal association with TRAF6 is critical for TLR2- and TLR4-mediated inflammatory responses.*
To establish the biological importance of Mal and TRAF6 interaction in regulating the immune response to TLR2 and TLR4 activation, we examined the effect of disruption of the Mal TRAF6-binding motif on TLR inflammatory responses. To perform these experiments we utilized immortalized Mal-deficient macrophages. We first established that this cell line was unresponsive to TLR2 (Pam3Cys) and TLR4 (LPS) stimulation, but did respond to TLR9 (CpG DNA), TLR7 (CL75) and TLR3 (poly I:C) ligands (Fig S3A) as assessed by TNF-α production. Immunoblotting also demonstrated the lack of Mal in these cells (Fig S3B). Mal-deficient macrophages were then reconstituted with GFP-tagged Mal, MalE190A or MalΔx3 by lentiviral transduction. GFP+ve cells were sorted to give a homogenous population of cells expressing GFP-tagged proteins (Fig S3C). Fluorescence microscopy of cells with GFP+ve expression for each tagged protein was used to verify protein expression (Fig S4D) and supported by immunoblot analysis of Mal protein expression in reconstituted cells (Fig 5A) demonstrating comparable expression levels of wild type Mal, MalE190A, Mal Δx3 and GFP alone.

Initially we wished to determine if an intact TRAF6-binding domain was required for NF-κB nuclear translocation to the nucleus following TLR stimulation. As can be observed in Figure 5, macrophages reconstituted with either wild type Mal (lanes 1, 4-6), MalE190A (lanes 2, 7-9) or MalΔx3 (lanes 3, 10-12) respectively display comparable NF-κB translocation to the nucleus in response to TLR2 (Fig 5A), TLR4 (Fig 5B) and TLR7 (Fig 5C) stimulation which suggests that interaction between Mal and TRAF6 is not required for efficient NF-κB translocation via the MyD88 canonical pathway.

We have previously reported that Mal interaction with TRAF6 is required for transactivation of NF-κB (17) and that Mal is crucial for Ser536 phosphorylation of the p65 subunit of NF-κB (21). As transactivation of NF-κB is a critical process in the transcriptional regulation of the inflammatory response (22), we next investigated if Mal interaction with TRAF6 was critical to this response. As can be observed in Figure 6A, only cells reconstituted with Mal were able to demonstrate robust phospho-Ser536 p65 in response to TLR2 stimulation with Pam3Cys (lanes 2, 7-9) compared to uninfected cells (lanes 1, 4-6) in a time-dependent manner. Critically, reconstitution of Mal-deficient macrophages with MalE190A (lanes 3, 10-12) failed to recapitulate TLR2-induced phosphorylation of the p65 subunit, thereby ablating transactivation of NF-κB. Notably, we observed that cells reconstituted with Mal demonstrate TLR-induced Ser276 phosphorylation of p65, which is absent in MalE190A reconstituted cells (compare lanes 7-9 to lanes 10-12). Mal has not been implicated previously in this transactivation event. Interestingly, in response to TLR4 stimulation, MalE190A reconstituted cells display partially restored p65 Ser536 or Ser276 phosphorylation (Fig. 6B), perhaps due to compensation by the MyD88-independent pathway.

To determine if Mal:TRAF6-mediated NF-κB transactivation plays a role in the inflammatory cytokine response to TLR stimulation, we next investigated cytokine expression in reconstituted Mal-deficient macrophages. Mal-deficient macrophages were reconstituted with wild type Mal, MalE190A or MalΔx3 and stimulated with LPS, Pam3Cys or the TLR7/8 agonist CL-75 (23) to induce IL-6 or TNF-α mRNA and protein gene expression. While reconstitution with Mal was able to mediate potent IL-6 or TNF-α mRNA and protein gene expression. While reconstitution with Mal was able to mediate potent IL-6 (Fig 7A and 7D) and TNF-α (Fig. 7B and 7E) expression compared to the GFP reconstituted or non-infected cells alone, both MalE190A and MalΔx3 displayed a significantly inhibited TLR2 and TLR4 proinflammatory responses. Importantly, as expected, TLR7/8 stimulation induced a robust response with all constructs and was independent of the absence or presence of Mal. Significantly, TLR4 induction of IFNβ mRNA was also not affected either in the presence or absence of Mal (Fig 7C), consistent with Mal-independent TLR4-induced IFNβ expression (8).

Taken together, these results clearly demonstrate the critical role Mal interaction with TRAF6 plays in generating the pro-inflammatory response as a consequence of TLR2 and TLR4 activation and that the mechanism is via a pathway of NF-κB transactivation, independent of the canonical MyD88-dependent NF-κB nuclear translocation.
DISCUSSION

In this study we have extended and characterized our earlier finding that the novel interaction of Mal with TRAF6 has a critical role in TLR2- and TLR4-induced inflammatory responses. Localization studies demonstrate that the geographical constraint of Mal to the plasma membrane may be important in the spatial control of Mal-mediated signaling. Significantly, the data presented characterizes the direct interaction between Mal and TRAF6, which regulates transactivation of the p65 subunit of NF-κB but does not affect NF-κB nuclear translocation. The association of Mal and TRAF6 therefore appears to control the transcriptional expression of NF-κB-dependent pro-inflammatory cytokines. Reconstitution of Mal-deficient macrophages with MalE190A containing a point mutation within the TRAF6-binding motif, significantly inhibits both TLR2- and TLR4-mediated inflammatory responses compared to wild type Mal. These results describe a specific role for Mal in TLR signaling and identify a direct interaction between Mal and the key signaling mediator TRAF6, that is separate to that of acting simply as a bridging adapter for MyD88 signaling.

The importance of Mal in TLR-mediated signal transduction in human disease was suggested by the recent description of the S180L functional variant and its association with protection against pneumococcal disease, tuberculosis, malaria, bacteremia and systemic lupus erythematosus (15,24). We found no affect of this variant on the ability of Mal to recruit TRAF6 (data not shown) however, our studies with MalE190A ,those by Hill and co-workers, combined with the inability of the rare Mal D96N SNP to interact with MyD88 (16) highlight the fact that structural modifications to Mal can have important implications in the formation of the plasma membrane proximal signaling complex. Interestingly, while Mal has been reported to undergo Tyr phosphorylation (25,26), caspase-1-mediated cleavage (27) and now the physiological impact of the S180L variant, none of these events altered the ability of Mal to interact with TRAF6 in immunoprecipitation experiments as ectopically expressed single amino substitution variants were still recognized by recombinant TRAF6 (data not shown) which combined with the direct interaction of the recombinant Mal and TRAF6 proteins strongly suggests that post-translational modification is not required for the interaction to occur. This suggests therefore there is still more to be understood as to the signaling capabilities of Mal. Critically, this study illustrates the importance of a single amino acid substitution within the critical TRAF6-binding motif of Mal and demonstrates how the immune response to TLR2 and TLR4 is profoundly affected. Consequently, Mal does not act solely as a bridging adapter, facilitating MyD88 recruitment to the membrane proximal receptor complex, but directly recruits TRAF6 to the plasma membrane to regulate NF-κB transactivation and ultimately transcriptional activation of the inflammatory response, independent of NF-κB nuclear translocation.

Interestingly, the TRAF6-binding motif in Mal is situated within its TIR domain raising the possibility of MyD88 and TRAF6 competing for binding sites. A recent paper by Ohnishi et al. demonstrated through complex structural modeling 2 Mal proteins bound MyD88 based on residues responsible for TIR-TIR interaction (28). Their model suggested that MyD88-TIR binding would not interfere with interactions between Mal-TIR and TLR4, TLR2, or TRAF-6 as the Glu190 in Mal was distal to the Mal:MyD88 interaction surface. This model is therefore suggestive of the formation of a signaling complex comprising TLR4, Mal, TRAF6 and MyD88 to initiate the signaling cascade.

It has previously been demonstrated that the myristoylation sequence in the N-terminal of TRAM is critical for its plasma localization and the ability to mediate TLR4 signaling (29). Kagan and Medzhitov demonstrated that TRAM also requires further phosphoinositide modification that facilitates recruitment of TRIF and subsequently, TRAF3 to the complex to promote other signaling pathways (14). This modification or geographical constraint appears to provide the TRAM/TRIF complex with the opportunity to specifically recruit TRAF3, an adapter required for induction of IFN. This description of ‘spatial’ signaling control of TRAM (14,30) and its geographical restriction to the endosomal membrane as a means of organizing and coordinating the two TLR4 adapter systems may have similarities to what we have observed in this
study for Mal. Consequently, selective recruitment of TRAF family members to TIR adapter proteins due to localization may provide spatial control over signaling, which integrates signals and allows coordination of NF-κB, MAP kinase and IRF signaling pathways. Indeed our study found that the Mal 4KK variant that was unable to localize to the plasma membrane could still activate MAP kinases and NF-κB transactivation commensurate with wild type Mal suggesting membrane localization was not critical for mediating this aspect of Mal function. Analogous to that observed with TRAM, Mal localization to the membrane via PIP2 activation may provide an optimal platform for Mal and TRAF6 to associate which would be distinct from the MyD88-IRAK-TRAF6 complex utilized by other TIR-signaling pathways. The experiments described herein suggest that Mal interaction with TRAF6 does not affect MyD88-dependent pathway signal transduction leading to NF-κB nuclear translocation. However, the nuclear translocated NF-κB would not appear to undergo transactivation without the Mal/TRAF6 interaction. It would appear that Mal interaction with TRAF6 is implicit in transactivation of the p65 subunit of NF-κB, seemingly via the MAP kinase pathway (17). Thus the plasma membrane localization of Mal and TRAM appears to allow the spatial recruitment of specific signaling components and as such, coordinating downstream signaling events (30) analogous to that described with TRAM and TRAF3 to regulate IFN induction.

Using complementation experiments in Mal-deficient macrophages, we were able to clearly demonstrate the importance of interaction between Mal and TRAF6 for a robust inflammatory response to TLR2 and TLR4 ligands. Disruption of the TRAF6-binding motif in Mal inhibited both NF-κB transactivation (phospho-p65 Ser276 and Ser536) (22) and significantly ablated cytokine gene expression and production in response to TLR2 and TLR4 stimulation. Interestingly, while the TRAF6-binding motif is important for some of the functionality of Mal, all three critical amino acids of the motif required mutation to significantly inhibit LPS-mediated TNF-α production. This result suggests other pathways may play a role in TLR4-induced TNF-α secretion. This disparity in effect may be a combination of the complexity of TLR4 signaling (ie. MyD88-dependent and MyD88-independent signaling) combined with the intricacies of TNF secretion requiring expression, trafficking and transmembrane cleavage, which may explain the discrepancy between these results. Interestingly, Baltimore and co-workers have suggested that TNF-α is expressed by the MyD88-independent pathway via IRF3, which was time dependent (31).

Additionally, p38 has been shown to play a role in regulating both TNF-α message stability and protein expression (32,33). We have previously described that Mal interaction with TRAF6 mediates Erk1/2 and JNK activation (17), but not p38, which may explain the discrepancy that the MalE190A mutant is still able to induce TNF-α expression. Moreover, we also demonstrated that reconstitution of Mal-deficient cells with MalE190A did not affect the ability of TLR4 to drive the MyD88-independent pathway (as witnessed by a lack of an effect on IFNβ induction). Therefore, it may be possible that inhibition of TNF-α mRNA expression by MalE190A represents ‘early’ activation, but the ‘late’ MyD88-independent response has compensated for this inhibition.

Overall, this study clearly demonstrates a critical role for Mal in TLR2- and TLR4-mediated inflammatory responses that are novel to its role as a bridging adapter for MyD88. Mal regulation of the transcriptional control of NF-κB via TRAF6 interaction and the subsequent expression of inflammatory cytokines is a key regulatory step in TLR2 and TLR4 inflammatory responses. This interaction allows activation of both the MyD88-dependent canonical and Mal/TRAF6-dependent transactivation pathways, culminating in the coordinated translocation and subsequent transactivation of NF-κB; both of which are critical to production of pro-inflammatory cytokines.

Significantly, a single point mutation within the TRAF6-binding motif of Mal is able to potently inhibit the pro-inflammatory response to TLR2 and TLR4 ligands, the primary response mechanism for Gram-positive and Gram-negative bacteria. This study identifies Mal as a key regulator of TLR2 and TLR4 signal transduction via a novel interaction with TRAF6, the possible disruption of which may provide a putative specific therapeutic intervention for TLR2- and TLR4-mediated...
inflammatory diseases without impacting on MyD88-dependent or TRIF/TRAM-dependent signaling pathways.

ABBREVIATIONS USED:
TLR, Toll-like receptor; TIR, Toll/IL-1 receptor; GST, glutathione S-transferase; IRAK, IFN, Interferon; Interleukin, IL; Interleukin-1 receptor-associated kinase; LPS, lipopolysaccharide, Mal, MyD88-adaptor like; NF-κB, Nuclear Factor-κB; PIP2, phosphatidylinositol 4,5-biphosphate; TRAF, TNF receptor associated factor; TRAM; TRIF-related adapter molecule; TRIF, TIR-domain containing adapter protein inducing IFNβ

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Figure Legends.

Figure 1. **Mal immunoprecipitates with TRAF6 following TLR2 and TLR4 stimulation.** Human monocyteic THP1 cells (2x10^7) were stimulated with (A) Pam3Cys (100 ng/ml), (B) LPS (100 ng/ml) or (C) CpG-DNA (500 nM) for indicated times. Cells were lysed and cellular debris removed by centrifugation (14,000 rpm, 10 mins, 4°C). Lysates were pre-cleared with Protein A-Sepharose beads and the lysates probed with α-TRAF6 mAb (1μg) bound to α-mouse IgG beads. Detection of immunoprecipitated endogenous Mal was analyzed by immunoblotting with α-Mal antibody (Pearl-1) and IκBα detected with α-IκBα mAb. Results are representative of three independent experiments.

Figure 2. **TRAF6 co-localizes with Mal regardless of localization.** Micrographs of HEK293 co-transfected with GFP-tagged Mal (A), Mal 4KK (B), or Mal (1-20 aa) (C) and TRAF6-Flag for 48 h at 32°C. Immunofluorescence microscopy of Z-stack images (35-50) were taken of individual cells and 3D image reconstruction was performed using Imaris software (BitPlan). Co-localization of the Z-stack images (intense yellow), illustrates clusters of TRAF6-Flag (Texas Red) co-localizing with concentrated Mal-GFP in plasma membrane ruffles (arrows, right panel, bar 8 µm) following 3D iso-surface rendition. Mal4KK-GFP and Mal (1-20aa)-GFP illustrates TRAF6 localized to the cytosol. All images are sections taken with a 40X oil objective lens and are representative of at least three independent experiments where over 500 cells were examined per condition, >90% of the cells displayed similar staining. 

(D) HEK293T cells were co-transfected with TRAF6-Flag and GFP-tagged wild-type Mal, Mal 4KK or Mal (1-20 aa) where indicated. Cells were lysed and TRAF6 immunoprecipitated with α-M2 Flag agarose beads. Detection of Mal or Mal variant-GFP proteins were analyzed by immunoblotting with α-Mal antibody. Expression of TRAF6 was verified by immunoblot analysis of cellular lysates with α-Flag M2-HRP conjugated antibody while GFP-tagged protein expression was verified by α-GFP immunostaining (n=3).

(E) HEK293 cells were co-transfected with 5x κB-luciferase promoter gene plasmid (100 ng), concomitantly with 10 ng of either GFP-tagged Mal, Mal 4KK, Mal (1-20 aa) or HA-tagged Mal and MalE190A constructs for stimulation. Shown is the mean relative luciferase activity ±SEM for a representative experiment from three separate experiments. Readings are normalized for κB-luciferase expression over TK-renilla expression. *(F) HEK293 cells were co-transfected with components of the pRF-luciferase and either c-jun (2 ng), CHOP (5 ng), Elk-1 (10 ng) or p651-551 (10 ng) Gal4 fusion vector, in conjunction with either GFP-tagged Mal, Mal 4KK or Mal (1-20aa) constructs respectively. Shown is the mean relative luciferase activity ±SEM for a representative experiment from three separate experiments. All luciferase results were comparative to constitutively expressed TK-renilla. *P<0.05, **P<0.01, ***P<0.001.

Figure 3. **Mal co-localization at the plasma membrane with TRAF6 requires an intact TRAF6 binding motif.** HEK293 cells were co-transfected with DsRed-tagged Mal (A), MalE190A (B), or MalΔx3 (C) with TRAF6-Flag for 48 h at 32°C. Immunofluorescence microscopy of Z-stack (29-45) images were taken of individual cells and 3D image reconstruction was performed using Imaris software (BitPlan). Co-localization illustrates TRAF6-Flag co-localizing with Mal-DsRed in plasma membrane ruffles (arrows, right panel, bar 8 µm). MalE190A-DsRed and MalΔx3-DsRed mutant demonstrate plasma localization while TRAF6 is diffusely distributed within the cytosol (right panel, bar 8 µm) and minimal co-localization observed (right panel, bar 10 µm). All results are representative of three independent experiments.

Figure 4. **Mal directly interacts with TRAF6 via an intact TRAF6-binding motif.** (A) HEK293T cells were transiently transfected with Flag-tagged TRAF6 (1.25 μg) for 24 h. Cells were lysed and immunoprecipitated with either 1 μg of recombinant GST-Mal or GST protein. Mal interacting complexes were visualized by immunoblotting with α-Flag antibody to represent TRAF6. Expression of TRAF6 in transiently transfected cells was confirmed by immunoblot of cell lysates with α-Flag antibody.
HEK293T cells were transfected with either HA-tagged wild-type Mal or the Mal mutated TRAF6-binding motif, MalE190A (1.25 μg respectively) where indicated. Cells were lysed and immunoprecipitated with 1 μg of recombinant TRAF6-GST. TRAF6-GST protein immunocomplexed with ectopically expressed wild-type Mal but not with MalE190A when immunoblotted with α-HA antibody. Mal protein expression was confirmed by immunoblot analysis of cell lysates using α-HA antibody to detect Mal-HA. Recombinant TRAF6-Flag protein (0.75 μg) interacts specifically with recombinant GST-Mal (1.5 μg) but not with GST-fused MyD88 or MalE190A protein in an in-vitro binding assay. TRAF6 immunocomplexes were precipitated using α-M2 Flag beads in a high salt (400 mM) PBS buffer and immunoblotted with α-GST antibody to detect interacting proteins. Recombinant proteins were confirmed by immunoblot analysis of inputs with their indicated antibodies. All results are representative of three independent experiments.

Figure 5. Mal interaction with TRAF6 does not affect NF-κB nuclear translocation in response to TLR stimulation.

(A) Virally transduced GFP-positive populations for each indicated gene were gated and sorted for >90% GFP positive population, then seeded at a concentration of 3 x 10⁴ cells/well 24 h prior to lysis in KalB buffer. Cellular debris was removed by centrifugation (13000 rpm, 10 mins, 4°C) and equal protein loaded onto SDS-PAGE, transferred to nitrocellulose and the relative expression of viral reconstituted immortalized Mal-deficient macrophages assessed for GFP-expressing proteins by immunoblot of cell lysates with α-GFP antibody. Reconstituted Mal-deficient macrophages expressing the indicated genes were seeded at 1x10⁴ cells/ml in 6 well plates 72 h prior to stimulation with (B) TLR2 ligand Pam3Cys (10 ng/ml), (C) TLR4 ligand LPS (10 ng/ml) or (D) TLR7 ligand CL-75 (0.5 μg/ml) for indicated times. Nuclear extracts (4 μg) were prepared, incubated with γ32P-labeled NF-κB DNA probe and analyzed by a 6% native gel. Binding to the NF-κB probe was observed in all stimulated virally-reconstituted nuclear extracts (lanes 1-12) but only weakly observed in non stimulated cells (lanes 1-3). Uninfected-stimulated extracts observed binding from treatment with LPS (C) and CL-75 (D) but not from Pam3Cys (B) (lanes 13-15). Unbound free NF-κB probe was shown in lower panel (B). Results are representative of two independent experiments.

Figure 6. Mal interaction with TRAF6 is required for TLR2- and TLR4-mediated transactivation of the p65 subunit of NF-κB.

Immortalized Mal-deficient macrophages were virally reconstituted with either GFP-tagged Mal (lanes 2, 7-9) or MalE190A (lanes 3, 10-12) as previously described in Figure 5. GFP-positive populations were challenged with either 10 ng/ml of Pam3Cys (A) or LPS (B) for indicated time points, lysed and protein separated by SDS-PAGE followed by immunoblot for p65, p65 Ser276 or Ser536 phosphorylation where indicated. Results represent three independent experiments.

Figure 7. A disrupted TRAF6-binding domain in Mal inhibits TLR2- and TLR4-mediated NF-κB proinflammatory responses. Mal-deficient macrophages were reconstituted with either no virus, GFP, GFP-tagged Mal, MalE190A or MalΔx3 as indicated were seeded at 3x10⁴ cells/well 72 h prior to stimulation with 10 ng/ml of Pam3Cys, LPS, or CL-75 (0.5 μg/ml) ligand respectively. RNA was extracted from macrophages after 3 h challenge and mRNA induction levels of (A) IL-6, (B) TNF-α and (C) IFN-β measured using quantitative real-time PCR where the relative abundance was compared to 18S levels. Supernatants were collected from macrophages 24 h after stimulation and assessed for (D) IL-6 or (E) TNF-α expression by ELISA. Mal variants lacking an intact TRAF6-binding motif demonstrate defective for TLR2 and TLR4-induced pro-inflammatory cytokine expression. Results are mean ±SEM of three experiments. * P<0.05, ** P<0.01, *** P<0.001.
Figure 2

A. 3D View 3D iso-surface rendition Colocalization

Mal-GFP TRAF6-Flag

B. Mal 4KK-GFP TRAF6-Flag

C. Mal 1-20-GFP TRAF6-Flag

D. Mal-GFP + + - -
   Mal 4KK-GFP - - + -
   Mal 1-20-GFP - - - +
   TRAF6-Flag - + + +

IP: α-Flag
IB: α-GFP

IP: α-Flag
IB: α-Flag
IB: α-GFP

E. 

F. 

Fold Stimulation

Empty Vector Mal-GFP Mal 4KK-TRAF6 Mal 1-20-TRAF6

Fold Stimulation

Empty Vector Mal-GFP Mal 4KK-TRAF6 Mal 1-20-TRAF6
Figure 3

A.

Mal-DsRed
TRAFl6-Flag

B.

Mal E190A
DsRed
TRAFl6-Flag

C.

Mal Δx3
DsRed
TRAFl6-Flag
Figure 4

A.

| GST      | + | - |
| GST-Mal  | - | + |
| TRAF6-Flag | + | + |

IP: GST
IB: α-Flag
IB: α-Flag

B.

| GST-TRAF6 | + | + |
| Mal-HA    | + | - |
| Mal E1901A-HA | - | + |

IP: GST
IB: α-HA
IB: α-HA

C.

| GST-Mal   | + | - | - | - |
| GST-Mal E190A | - | + | - | - |
| GST-MyD88  | - | - | + | - |
| GST       | - | - | - | + |
| TRAF6-Flag | + | + | + | + |

IP: α-Flag
IB: α-GST
IB: α-GST
IB: α-Flag
Figure 5

A.

B.  

| Time (mins) | Mal -GFP | Mal E190A-GFP | Mal Δx3-GFP | Uninfected |
|-------------|----------|---------------|-------------|------------|
| 0          | 0        | 0             | 0           | 0          |
| 15         | 30       | 60            | 15          | 30         |
| 30         | 60       | 15            | 30          | 60         |
| 60         |          |               | 15          | 30         |

NF-κB

Free γ32P-κB probe

Lane: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15

C.  

| Time (mins) | Mal -GFP | Mal E190A-GFP | Mal Δx3-GFP | Uninfected |
|-------------|----------|---------------|-------------|------------|
| 0          | 0        | 0             | 0           | 0          |
| 15         | 30       | 60            | 15          | 30         |
| 30         | 60       | 15            | 30          | 60         |
| 60         |          |               | 15          | 30         |

NF-κB

Lane: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15

D.  

| Time (mins) | Mal -GFP | Mal E190A-GFP | Mal Δx3-GFP | Uninfected |
|-------------|----------|---------------|-------------|------------|
| 0          | 0        | 0             | 0           | 0          |
| 15         | 30       | 60            | 15          | 30         |
| 30         | 60       | 15            | 30          | 60         |
| 60         |          |               | 15          | 30         |

NF-κB

Lane: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15
Figure 6

A.

| Time (mins): | 0  | 0  | 0  | 15 | 30 | 60 | 15 | 30 | 60 | 15 | 30 | 60 |
|-------------|----|----|----|----|----|----|----|----|----|----|----|----|
| Pam, Cys    |    |    |    |    |    |    |    |    |    |    |    |    |
| Uninfected  |    |    |    |    |    |    |    |    |    |    |    |    |
| Mal-GFP     |    |    |    |    |    |    |    |    |    |    |    |    |
| Mal E190A-GFP |   |    |    |    |    |    |    |    |    |    |    |    |
| IB: α-pSer536 p65 | | | | | | | | | | | | |
| IB: α-pSer276 p65 | | | | | | | | | | | | |
| IB: α-p65   |    |    |    |    |    |    |    |    |    |    |    |    |

Lane: 1 2 3 4 5 6 7 8 9 10 11 12

B.

| Time (mins): | 0  | 0  | 0  | 15 | 30 | 60 | 15 | 30 | 60 | 15 | 30 | 60 |
|-------------|----|----|----|----|----|----|----|----|----|----|----|----|
| LPS         |    |    |    |    |    |    |    |    |    |    |    |    |
| Uninfected  |    |    |    |    |    |    |    |    |    |    |    |    |
| Mal-GFP     |    |    |    |    |    |    |    |    |    |    |    |    |
| Mal E190A-GFP |   |    |    |    |    |    |    |    |    |    |    |    |
| IB: α-pSer536 p65 | | | | | | | | | | | | |
| IB: α-pSer276 p65 | | | | | | | | | | | | |
| IB: α-p65   |    |    |    |    |    |    |    |    |    |    |    |    |

Lane: 1 2 3 4 5 6 7 8 9 10 11 12
Figure 7

A.

B.

C.

D.

E.
MyD88 adapter-like (mal)/TIRAP interaction with TRAF6 is critical for TLR2- and TLR4-mediated NF-κB proinflammatory responses

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