Arylpyrrole and fipronil analogues that inhibit the motility and/or development of *Haemonchus contortus* in vitro

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**ABSTRACT**

Due to widespread drug resistance in parasitic nematodes, there is a need to develop new anthelmintics. Given the cost and time involved in developing a new drug, the repurposing of known chemicals can be a promising, alternative approach. In this context, we tested a library (n = 600) of natural product-inspired pesticide analogues against exsheathed third-stage-larvae (xL3s) of *Haemonchus contortus* (barber's pole worm) using a whole-organism, phenotypic screening technique that measures the inhibition of motility and development in treated larvae. In the primary screen, we identified 32 active analogues derived from chemical scaffolds of arylpyrrole or fipronil. The seven most promising compounds, selected based on their anthelmintic activity and/or limited cytotoxicity, are arylpyroles that reduced the motility of fourth-stage larvae (L4s) with significant potency (IC\textsubscript{50} values ranged from 0.04 ± 0.01 \(\mu\)M to 4.25 ± 0.82 \(\mu\)M, and selectivity indices ranged from 10.6 to 412.5). Since the parent structures of the active compounds are uncouplers of oxidative phosphorylation, we tested the effect of selected analogues on oxygen consumption in xL3s using the Seahorse XF24 flux analyser. Larvae treated with the test compounds showed a significant increase in oxygen consumption compared with the untreated control, demonstrating their uncoupling activity. Overall, the results of the present study have identified natural product-derived molecules that are worth considering for chemical optimisation as anthelmintic drug leads.

1. Introduction

The control of gastrointestinal nematodes of livestock has relied largely on the use of a limited number of anti-parasitic drugs (Besier et al., 2016; Harder, 2016). However, drug resistance is now very widespread (Kaplan and Vidyasagar, 2012; Kotze and Prichard, 2016) and no vaccines are available for the vast majority of these worms, such that the development of new drugs is crucial to ensure effective and sustained nematode control into the future. Although the development of the compound monepantel (Kaminsky et al., 2008; Prichard and Geary, 2008) provided hope for the design of new classes of nematocides, there has been relatively little success in discovering new drugs using conventional and high throughput screening approaches (Geary et al., 2015).

Given the massive cost and time required to develop and commercialise a new anthelmintic (Geary et al., 2015), the repurposing of known drugs or bioactive chemicals (Corsello et al., 2017) offers a pragmatic and sound option. The repurposing of chemicals with known safety profiles and modes of action has the potential to proceed to clinical trials more rapidly than newly discovered chemotypes (Oprea and Mestres, 2012; Andrews et al., 2014; Panic et al., 2014; Corsello et al., 2017).

Recently, in partnership with philanthropic, academic and industry partners, we screened compound libraries of bioactive compounds for...
inhibitory activity against *Haemonchus contortus* (barber’s pole worm), in an attempt to repurpose synthetic and natural compounds against parasitic nematodes (Preston et al., 2016, 2017; Herath et al., 2017). For instance, we have shown that pesticides such as tolfenpyrad (Preston et al., 2016) and other pyrazole-5-carboxamides (e.g., a-15 and a-17; Jiao et al., 2017) have potent inhibitory activity on the motility and/or development of exsheathed third-stage (xL3s) and/or fourth-stage (L4s) larvae of *H. contortus* in vitro and that these chemicals are relatively selective for the parasite compared with a mammalian cell line (see Preston et al., 2016; Jiao et al., 2017). Using medicinal chemical methods, the potency and selectivity of some of these chemotypes have been significantly increased (unpublished findings). Taken together, this information indicates that there is merit in taking a repurposing route, provided that medicinal chemistry follows the screening effort, in order to optimise potency and safety in vitro and in vivo, pharmacokinetics and efficacy.

Here, we extend previous studies (Preston et al., 2016; Jiao et al., 2017) to screen a library of compounds (*n* = 600) obtained from the Research Institute of Elemento-Organic Chemistry, Nankai University, China. This library contains novel analogues of various pesticides, including antofine (Gao et al., 2012), arlypyroles (Kuhn, 1997), diflubenzuron (Post and Vincent, 1973; Cohen and Casida, 1980), etoxazole (Nauen and Smagghe, 2006), fipronil (Cheng et al., 2009) and spirodiclofen (Van Pottelberge et al., 2009). Most of these analogues had been tested previously at Nankai University, and exhibited insecticidal and/or acaricidal properties (Li et al., 2012; Liu et al., 2014; Ma et al., 2014), suggesting that these compounds might act on other edysoxoans, including nematodes of veterinary importance. Therefore, we screened the library against larval stages of *H. contortus* - one of the most important parasitic nematodes of livestock animals (Gasser and von Samson-Himmelstjerna, 2016). We employed a semi-automated phenotypic screening technique (Preston et al., 2015) to identify active (‘hit’) compounds and to assess their potency against larval stages of *H. contortus*, followed by an evaluation of the cytotoxicity of these hits on a mammary epithelial cell line in vitro, with the aim of identifying candidate compounds.

2. Materials and methods

2.1. Parasite production and maintenance

*Haemonchus contortus* (Haecon-5 strain) was maintained in experimental sheep as described previously (Schwarz et al., 2013; Preston et al., 2015), in accord with institutional animal ethics guidelines (permit no. 1413429; The University of Melbourne, Australia). Third-stage larvae (L3s) were cultured and exsheathed using established methods (Preston et al., 2015). In brief, L3s were exsheathed by incubation in 0.15% v/v sodium hypochlorite (NaClO) for 20 min at 37 °C and then washed five times in sterile, physiological saline (pH 7.0, 37 °C). The exsheathed L3s (xL3s) were then suspended in Luria Bertani (LB) medium supplemented with 100 IU/ml of penicillin, 100 mg/ml of streptomycin and 2.5 mg/ml of amphotericin (Fungizone, antibiotic-antimycotic; cat. no. 15240-062; Gibco, USA) (Preston et al., 2015). The supplemented LB was designated as LB*. L4s were produced by incubating xL3s in LB* at 10% v/v CO2 and 38 °C for 7 days (Preston et al., 2015).

2.2. Screening for anthelmintic activity

The library of chemicals from the Research Institute of Elemento-Organic Chemistry, Nankai University, China, contained pure chemicals derived from natural (120) and synthetic (480) products with known activities against arthropods (flies and/or mites) (Supplementary file 1). All 600 compounds were individually screened (in triplicate) in 96-well plates using a previously described protocol (Preston et al., 2015; cf. Fig. 1). In brief, the chemicals were each dissolved in 100% DMSO to prepare 20 mM stocks, diluted into 40 μM in 50 μl of LB* (containing 1% DMSO) and arrayed in triplicate in flat-bottom 96-well microplates (cat. no. 3635; Corning 3650, Life Sciences, USA). The xL3s (∼300 in 50 μl per well) were added to wells and exposed to a final compound concentration of 20 μM (with 0.5% DMSO). The two commercially available drugs monepantel (Zolvix, Novartis Animal Health, Switzerland) and moxidectin (Cydectin, Virbac, France) were included as positive controls, and LB* + 0.5% DMSO as the negative control (six wells). Plates were placed in an incubator for 72 h (10% v/v CO2; 38 °C). For data acquisition, plates were agitated for 20 min using an orbital shaker (126 rotations per min) (model KOM5, Ratek, Australia), and a 5 s video was recorded from each well. Digital recordings were processed using a customised script in Image J (imageJ. nih.gov/ij/), and changes in light intensity transformed into a motility index (Mi; Preston et al., 2015). The raw data were normalised with respect to positive and negative controls, in order to remove plate-to-plate variation, and the percentage of motility reduction was calculated for each well using the program Prism (v.7.02 GraphPad Software). A compound was recorded as being active if it reduced xL3 motility by ≥ 70% and/or induced phenotypic alterations in comparison with the untreated wild-type control worms (at 72 h).

2.3. Dose-response curves for active compounds (xL3 motility, L4 motility and L4 development)

All active compounds were tested as a two-fold dilution series (18 points; starting at 100 μM) to assess a reduction in xL3 motility and an inhibition of L4 development (Preston et al., 2015, Fig. 1). To measure motility, video recordings of individual wells were taken at 24 h, 48 h and 72 h. To assess development, plates were incubated further under the same conditions for four more days; worms in each well were then fixed with 50 μl of 1% iodine and the development to L4 determined by examining 30 worms from each well under a microscope at 20× magnification. L4s were differentiated from xL3s based on the presence of well-developed mouth and pharynx.

The half-maximum inhibitory concentration (IC50) of each compound was determined by transforming the compound concentrations to log10, and fitting the data using a variable slope four-parameter equation (v.7.02 GraphPad Software) (cf. Preston et al., 2015). IC50 values were obtained from results from two independent biological assays with three technical replicates for each compound. Compounds were ranked based on the level of xL3 motility reduction, L4 development inhibition and minimal cytotoxicity. A subset of compounds was tested for their ability to inhibit L4 motility, again using a two-fold dilution series and in two biological assays with three technical replicates.

2.4. Cytotoxicity assay

The cytotoxicity of individual compounds was tested as described previously (Kumarasingha et al., 2016; Fig. 1) at the Victorian Centre for Functional Genomics in Cancer, Melbourne, Australia, with minor modifications. In brief, normal breast epithelial (MCF10A) cells were cultured in Dulbecco’s Modified Eagle Medium: Nutrient Mixture F-12 (DMEM/F12; Thermofisher, USA) containing 5% v/v horse serum (Life Technologies, Australia), 100 ng/ml of cholera toxin (Sigma, Australia),
2.5. Larval oxygen consumption assay

The primary screen of the library (Fig. 1) identified 21 compounds that reduced the motility of *H. contortus* xL3s by ≥ 70% and induced a non-wildtype ("circular") phenotype as well as 11 compounds that induced a "circular" phenotype but did not reduce the xL3 motility by ≥ 70% (Supplementary file 1). All of these 32 compounds were categorised as 'hits' (Fig. 1); 30 of them were pyrrole analogues (the
The effects of seven active analogues on xL3 and L4 motility (24 h, 48 h and 72 h) and L4 development (7 days) of *Haemonchus contortus* in vitro. A comparison of half of the maximum inhibitory concentration (IC50) values of compounds with those of monepantel or moxidectin, expressed as mean IC50 ± standard error of mean or a selectivity index (SI) of the compounds on motility (xL3 and L4) and development (L4) of *H. contortus* at different time points compared with monepantel.

| Compound | IC50 (in μM) | Selectivity index (SI) for *H. contortus* |
|----------|--------------|---------------------------------|
|          | xL3 motility | L4 motility | L4 development |
|          | 24 h | 48 h | 72 h | 24 h | 48 h | 72 h | 7 days |
| Zpx019   | nd | 11.81 ± 0.19 | 7.40 ± 1.22 | 13.45 ± 3.66 | 3.13 to 6.25 | 3.7 b | 4.28 ± 0.38 |
| Zpx020   | nd | nd | 3.86 ± 0.28 | 3.47 ± 2.36 | 3.22 ± 1.64 | 2.28 ± 0.67 | 7.24 ± 0.97 |
| Zpx022   | nd | 10.62 ± 4.07 | 9.14 ± 3.32 | 17.68 ± 0.11 | 4.3 b | 4.25 ± 0.82 | 13.98 ± 0.98 |
| Zpx024   | 25.26 ± 8.50 | 9.17 ± 2.60 | 3.2 b | 2.45 ± 1.22 | 1.07 ± 0.15 | 1.30 ± 0.15 | 1.99 ± 0.02 |
| Zpx027   | 9.79 ± 2.54 | 3.10 ± 0.01 | 1.22 ± 0.09 | 0.51 ± 0.29 | 0.14 ± 0.12 | 0.09 ± 0.04 | 1.18 ± 0.13 |
| Zpx028   | 8.63 ± 0.90 | 2.9 b | 1.25 ± 0.36 | 0.16 ± 0.13 | 0.10 ± 0.03 | 0.04 ± 0.01 | 0.57 ± 0.04 |
| Zpx040   | nd | 3.1 b | 1.68 ± 0.66 | 12.92 ± 4.00 | 2.6 b | 3.36 ± 0.86 | 1.58 ± 0.09 |
| Monepantel | 2.12 ± 0.55 | 0.76 ± 0.32 | 0.59 ± 0.01 | 6.52 ± 2.18 | 0.7 b | 0.1 b | 0.43 ± 0.01 |
| Moxidectin | 0.50 ± 0.42 | 0.19 ± 0.04 | 0.08 ± 0.02 | 0.92 ± 0.90 | 0.002 ± 0.04 | 0.003 ± 0.01 | not applicable |

Table 2
Toxicity assessment of seven arylpyrrole analogues on MCF10A cells. A comparison of half of the maximum inhibitory concentration (IC50) values of compounds with monepantel or doxorubicin; expressed as mean IC50 ± standard error of mean, and selectivity indices (SI) of the compounds on motility (xL3 and L4) and development (L4) of *Haemonchus contortus* at different time points compared with monepantel.

| Compound | IC50 (in μM) for MCF10A cells | Selectivity index (SI) for *H. contortus* |
|----------|--------------------------------|---------------------------------|
|          | xL3 motility | L4 motility | L4 development |
|          | 24 h | 48 h | 72 h | 24 h | 48 h | 72 h | 7 days |
| Zpx019   | > 50 | nd | > 4.5 | > 6.8 | > 3.7 | > 8.0 | > 13.6 | > 11.7 |
| Zpx020   | > 50 | nd | nd | > 12.9 | > 14.4 | > 15.7 | > 21.9 | > 7.0 |
| Zpx022   | > 50 | nd | > 4.7 | > 5.5 | > 2.8 | > 11.7 | > 11.7 | > 3.6 |
| Zpx024   | 26.11 ± 3.92 | 1.0 | 2.9 | 8.1 | 10.7 | 24.4 | 20.1 | 13.1 |
| Zpx027   | 6.36 ± 3.29 | 0.7 | 2.1 | 5.2 | 20.5 | 45.6 | 70.9 | 5.4 |
| Zpx028   | 16.50 ± 2.58 | 1.9 | 5.7 | 13.2 | 103.1 | 165 | 412.5 | 28.9 |
| Zpx040   | 35.7 b | n/d | 11.5 | 21.3 | 2.8 | 13.9 | 10.6 | 22.6 |
| Monepantel | 32.8 b | 15.5 | 43.3 | 55.7 | 5.0 | 47.4 | 328.4 | 76.4 |

Table 1

**a** IC50 values that could not be accurately determined using the log (inhibitor) vs. response-variable slope four parameters model are indicated as not determined (nd). **b** Estimated from the graphs in Fig. 2.

**Mqq and Zpx series** could not be tested for toxicity to MCF10A cells, as IC50 values that were not established, SI was not determined (nd).

**Mqq and Zpx series** were fipronil analogues (Zqq series) (Supplementary file 1).

In the dose response assays (cf. Fig. 1), 13 of the 32 hits induced a dose-dependent inhibition of xL3 motility, with an IC50 of ≤ 50 μM at 24 h (Supplementary file 2). The IC50 values of these 13 compounds ranged from 8.63 ± 0.90 μM to 48.13 ± 2.71 μM (Supplementary file 2). Compound Zpx028, a pyrrole analogue, had the highest potency at 24 h, with an IC50 of 8.63 ± 0.90 μM. Compound Zpx027 had the second highest potency at inhibiting xL3 motility at 24 h (IC50 = 9.79 ± 2.54 μM), and was most potent at inhibiting xL3 motility at 72 h (IC50 = 1.22 ± 0.09 μM) (Table 1; Supplementary file 2).

All 32 hits, except Zpx021 and Zpx90, elicited a dose-dependent inhibition of L4 development; six compounds achieved an IC50 of ≤ 2 μM (Supplementary file 2). The two compounds, Zpx028 and Zpx027, with the highest potency at inhibiting xL3 motility, were also the most potent inhibitors of L4 development (Fig. 2; Table 1) - the IC50 values (0.57 ± 0.04 μM for Zpx028; 1.18 ± 0.10 μM for Zpx027) are comparable to those of the commercial anthelmintic monepantel in vitro (IC50 = 0.43 ± 0.01 μM) (Fig. 2; Table 1).

All 32 hits were tested for toxicity to MCF10A breast epithelial cells in vitro. The IC50 values obtained ranged from 2.16 μM to 50 μM, and selectivity indices (at 24 h) varied from 0.04 to 1.91 for xL3 motility, and from 0.26 to 28.95 for L4 development (7 days; Supplementary file 3). Eight compounds, Zpx028, Zpx030, Zpx023, Zpx026, Zpx024, Zpx040, Zpx090 (pyrrole analogues) and Zqq1341 (fipronil analogue), exhibited moderate toxicity (IC50 of 16.50–44.98 μM; Supplementary file 2). Four pyrrole analogues, Zpx019, Zpx020, Zpx021 and Zpx022, had limited toxicity to the cells, with IC50 values of ≥ 50 μM (Table 2; Supplementary file 2).

On the basis of the dose-dependent inhibition of xL3 motility (at 24 h) and L4 development (7 days) and/or the degree of cytotoxicity in vitro (Supplementary file 2), seven compounds (Zpx019, Zpx020, Zpx027...
Zpx022, Zpx027, Zpx024, Zpx028 and Zpx040; all pyrrole analogues) were assessed for their ability to inhibit the motility of L4s (Fig. 1). All of these seven compounds exhibited a dose-dependent inhibition of L4 motility at 24 h, with IC50 values ranging from 17.68 ± 0.11 μM to 0.16 ± 0.13 μM (Fig. 2; Table 1). Interestingly, compounds Zpx019, Zpx20, Zpx022 and Zpx040, which did not inhibit xL3 motility in a dose-dependent manner at 24 h, did inhibit L4 motility at the same time point (Fig. 2; Table 2). The two most potent inhibitors of both xL3 motility and L4 development also exhibited the highest potency (IC50 = 0.16 ± 0.13 μM for Zpx028 and IC50 = 0.31 ± 0.29 μM for Zpx027) and selectivity (SI = 20.5 for Zpx027 and SI = 103.1 for Zpx028) for inhibiting L4 motility at 24 h (Fig. 2; Tables 1 and 2).

As pyrroles act as insecticides via respiratory uncoupling (Black et al., 1994; Liu et al., 2014), we assessed compounds Zpx027 and Zpx028 for their effects on respiration in H. contortus. In response to each of these compounds, the oxygen consumption rate (OCR) of the xL3 stage increased (Fig. 3). Neither the negative (no-compound) nor the positive (monepantel) control showed a significant difference from the basal OCR in xL3s throughout the experimental period (Fig. 3). The OCR patterns for compounds Zpx028 and Zpx027 were similar to that of FCCP, a standard mitochondrial uncoupler (Fig. 3), and there was a significant difference (P < 0.001 for Zpx028 and P < 0.01 for Zpx027) in total oxygen consumption (cf. Fig. 3) between larvae treated with each test compound and the untreated and monepantel-treated controls (Fig. 3).
4. Discussion

The screening of a series of pesticide analogues identified 32 compounds with activity against *H. contortus* xL3s. Seven compounds were chosen for further evaluation based on their potency against *H. contortus* larvae (motility and development) and mostly limited toxicity on a human cell line (MCF10A). Selected compounds (n = 7) were found to be rapid inhibitors of L4 motility, and the two most potent ones in all worm assays (Zpx027 and Zpx028) were shown to increase oxygen consumption in xL3s. This increase mirrored the behaviour of a recognised mitochondrial uncoupler, FCCP (cf. Fig. 3), inferring that heightened respiration rates stemmed from an increased electron flow through the mitochondrial electron transport chain, rather than from a non-specific removal of dissolved oxygen. Therefore, the compounds tested seem to have a mode of action that is the same or similar to that of the arylypyrrole parent structure representing 30 of the 32 compounds tested seem to have a mode of action that is the same or similar to that of the arylypyrrole parent structure representing 30 of the 32 compounds.

The founding member of insecticidal and acaridical arylypyrroles was a natural product, dioxapyrrolomycin, first identified as a metabolite from *Streptomyces fumarus* (see Carter et al., 1987). The structure of dioxapyrrolomycin was modified to yield the commercial product chlorfenapyr (Addor et al., 1992), a compound that is effective against a range of parasitic arthropods of plants, including the southern army-mite (*Empoasca abrupta*), tobacco budworm (*Heliothis virescens-Fabri-cius*), western potato leafhopper (*Empoasca fabrea*) and red spider mite (*Tetranychus urticae*) (Kuhn, 1997). Chlorfenapyr is a pro-drug that is metabolically converted to the active compound (CL303268) by N-dealkylation inside the pathogen (Black et al., 1994). The biological effects of CL303268 and dioxapyrrolomycin seem to be based on their activity as uncouplers of oxidative phosphorylation (Black et al., 1994), and it is reasonable to propose that this is also the mode of anthelmintic action of the present arylypyrrole hit compounds, given the uncoupling activity exhibited by the two members tested (i.e. Zpx027 and Zpx028). Once again, natural product-inspired s have proven to be suitable starting points to identify potential anti-infective leads.

The reason for the rapid and dose-dependent activity of arylypyrrole analogues Zpx019, Zpx020, Zpx022 and Zpx040 against L4s, with no apparent activity against xL3s at 24 h, is not yet clear. However, we propose that it relates to differences in oral drug uptake between the two larval stages of *H. contortus*. Uptake in L4s may be greater than in xL3s due to a well-developed pharynx in this stage. At this point, it cannot be excluded that this difference might be due to variation in target function and/or in drug metabolism between the two development stages; however, given the high structural similarities among the four test compounds, this latter explanation is considered less likely, as they are expected to share the same target/mode of action and degradation pathways.

The two fipronil derivatives (Zqq-1726 and Zqq-1341) received limited experimental priority, because of their relatively low potency on *H. contortus* and higher cytotoxities relative to the selected arylypyrrole analogues. Nonetheless, their structure-activity relationship could be explored in future work. The parent compound of these structures is a broad spectrum, highly selective phenylpyrazole insecticide that exerts its biological activity by blocking GABA-gated chloride channels (Zhao et al., 2003) and glutamate-gated chloride (GluCl) channels (Gant et al., 1998; Zhao et al., 2004), so we presume that this will also be the mode of action for the analogues studied here, although we have not yet collected the evidence to support this proposal.

In conclusion, some of the analogues with favourable activity and toxicity profiles identified in this study warrant critical evaluation as anthelmintic candidates. Medicinal chemical optimisation, supported by iterative structure-activity relationship, in vitro and in vivo toxicity and pharmacokinetic assessments, should allow the potential of lead candidates to be established. Overall, the findings of the present study encourage the screening of natural product-derived libraries for compounds with anthelmintic activity.

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Appendix A. Supplementary data

Supplementary data related to this article can be found at https://doi.org/10.1016/j.ijpddr.2018.06.002.

References

Addor, R., Babcock, T., Black, B., Brown, D., Diehl, R., Furch, J., Kameswaran, V., Kambhi, V., Kremer, K., Kuhn, D., 1992. Insecticidal pyrethroids: discovery and overview. In: Baker, D.R., Fenyes, J.G., Steffens, J.J. (Eds.), Synthesis and Chemistry of Agrochemicals III. American Chemical Society, Washington, DC, USA, 9780841224735, pp. 283–297.

Andrews, K.T., Fisher, G., Skinner-Adams, K.I., Kukel, C.D., Donovan, S., 1994. Black, B.C., Hollingworth, R.M., Amsterdam, Netherland, pp. 147–152. Future: Minimizing Chronic Exposure of Humans and the Environment. IOS press, London, UK978-0-12-810395-1.

Arias, F., 2012. Structure-activity relationships for insecticidal pyrroles. In: Hedin, P.A., Hollingworth, R.M., Masler, E.P., Miyamoto, J., Thompson, D.G. (Eds.), Pyrethmecials for Pest Control. American Chemical Society, Washington, DC, USA, pp. 195–205ISBN-9780841234888.

Kumarasingha, R., Preston, S., Yeo, T.C., Lim, D.S., Tu, C.L., Palombo, E.A., Shaw, J.M., Gasser, R.B., Boag, P.R., 2016. Anthelmintic activity of selected etho-medical plant extracts on parasitic stages of Haemonchus contortus. Parasit. Vectors 9, 1458–1459.

Li, Y., Zhang, P., Ma, Q., Song, H., Liu, Y., Wang, Q., 2012. The trifluoromethyl transformation synthesis, crystal structure and insecticidal activities of novel 2-pyrrole-carboxamide and 2-pyrrolecarboxylate. Bioorg. Med. Chem. Lett. 22, 6858–6861.

Liu, Y.X., Zhang, P.X., Li, Y.Q., Song, H.B., Wang, Q.M., 2014. Design, synthesis, and biological evaluation of 2-benzylpyrroles and 2-benzoylpyrroles based on structures of insecticidal chlorfenapyr and natural pyrolymocins. Mol. Divers. 18, 593–598.

Ma, Q., Liu, Y., Zhang, P., Li, Y., Xiong, L., Wang, Q., 2014. Design, synthesis, and biological evaluation of various α-substituted benzyllpyrroles based on the structures of insecticidal chlorfenapyr and natural pyrolymocins. J. Agric. Food Chem. 62, 6072–6081.

McGee, S.L., Sadli, N., Morrison, S., Swinton, C., Suphioglu, C., 2011. DHA protects against zinc mediated alterations in neuronal cellular bioenergetics. Cell. Physiol. Biochem. 28, 157–162.

Nauen, R., Smagge, G., 2006. Mode of action of etoxazole. Pest Manag. Sci. 62, 379–382.

Oprea, T., Mestres, J., 2012. Drug repurposing: far beyond new targets for old drugs. AAPS J. 14, 759–763.

Post, L., Vincent, W., 1973. A new insecticide inhibits chitin synthesis. Naturewissenschaften 60, 431–432.

Panic, G., Duthaler, U., Speich, B., Keiser, J., 2014. Repurposing drugs for the treatment and control of helminth infections. Int. J. Parasitol. Drugs Drug Resist. 4, 185–200.

Preston, S., Jabbar, A., Nowell, C., Joachim, A., Ruttkowsky, B., Baell, J., Cardno, T., 2015. Low cost whole-organism screening of compounds for anthelmintic activity. Int. J. Parasitol. 45, 333–343.

Preston, S., Jiao, Y., Jabbar, A., McGee, S.L., Lalue, B., Willis, P., Wells, T.N., Gasser, R.B., 2016. Screening of the ‘Pathogen Box’ identifies an approved pesticide with major anthelmintic activity against the barber’s pole worm. Int. J. Parasitol. Drugs Drug Resist. 6, 329–334.

Preston, S., Khoronen, P.K., Mouchiroud, L., Cornaglia, M., McGee, S.L., Young, N.D., Davis, R.A., Crawford, S., Nowell, C., Ansell, B.R.E., Fisher, G.M., Andrews, K.T., Chang, B.C.H., Gis, M.A.M., Sternberg, P.W., Auwercx, J., Baell, J., Hofmann, A., Jabbar, A., Gasser, R.B., 2017. Deguelin exerts potent nematocidal activity via the mitochondrial respiratory chain. FASEB J. 31, 4515–4532.

Prichard, R.K., Geary, T.G., 2008. Drug discovery: fresh hope to can the worms. Nature 452, 517–518.

Schwarz, E.M., Khoronen, P.K., Campbell, B.R.E., Young, N.D., Jiao, Y., Jabbar, A., Hall, R.S., Mondal, A., Howe, A.C., Peil, J., Hofmann, A., Boag, P.R., Zhu, X.Q., Gregory, T., Loukas, A., Williams, B.A., Antoshechkin, I., Brown, C., Sternberg, P.W., Gasser, R.B., 2013. The genome and developmental transcriptome of the strongylid nematode Haemonchus contortus. Genome Biol. 14, R99.

Terao, H., 1990. Uncouplers of oxidative phosphorylation. Environ. Health Perspect. 87, 213–218.

Van Pottelberge, S., van Leeuwen, T., Khajehali, J., Tirry, L., 2009. Genetic and biochemical analysis of a laboratory-selected spirodiclofen-resistant strain of Tetranychus urticae Koch (Acari: Tetranychidae). Pest Manag. Sci. 65, 203–218.

Zhao, X., Yeh, J.Z., Salgado, V.L., Narahashi, T., 2004. Fipronil is a potent open channel blocker of glutamate-activated chloride channels in cockroach neurons. J. Pharmacol. Exp. Therapeut. 306, 914–924.

Zhou, X., Yeh, J.Z., Salgado, V.L., Narahashi, T., 2004. Fipronil is a potent open channel blocker of glutamate-activated chloride channels in cockroach neurons. J. Pharmacol. Exp. Therapeut. 310, 192–201.
Author/s:
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