Title
Correction: Thermodynamic Driving Force of Hydrogen on Rumen Microbial Metabolism: A Theoretical Investigation.

Permalink
https://escholarship.org/uc/item/7rr0k9jz

Journal
PloS one, 11(12)

ISSN
1932-6203

Authors
van Lingen, Henk J
Plugge, Caroline M
Fadel, James G
et al.

Publication Date
2016

DOI
10.1371/journal.pone.0168052

Peer reviewed
Thermodynamic Driving Force of Hydrogen on Rumen Microbial Metabolism: A Theoretical Investigation

Henk J. van Lingen1,2 *, Caroline M. Plugge3, James G. Fadel4, Ermias Kebreab3, André Bannink5, Jan Dijkstra2

1 TI Food and Nutrition, Wageningen, The Netherlands, 2 Animal Nutrition Group, Wageningen University, Wageningen, The Netherlands, 3 Laboratory of Microbiology, Wageningen University, Wageningen, The Netherlands, 4 Department of Animal Sciences, University of California, Davis, Davis, California, United States of America, 5 Animal Nutrition, Wageningen UR Livestock Research, Wageningen, the Netherlands

* henk.vanlingen@wur.nl

Abstract

Hydrogen is a key product of rumen fermentation and has been suggested to thermodynamically control the production of the various volatile fatty acids (VFA). Previous studies, however, have not accounted for the fact that only thermodynamic near-equilibrium conditions control the magnitude of reaction rate. Furthermore, the role of NAD, which is affected by hydrogen partial pressure ($P_{H_2}$), has often not been considered. The aim of this study was to quantify the control of $P_{H_2}$ on reaction rates of specific fermentation pathways, methanogenesis and NADH oxidation in rumen microbes. The control of $P_{H_2}$ was quantified using the thermodynamic potential factor ($F_T$), which is a dimensionless factor that corrects a predicted kinetic reaction rate for the thermodynamic control exerted. Unity $F_T$ was calculated for all glucose fermentation pathways considered, indicating no inhibition of $P_{H_2}$ on the production of a specific type of VFA (e.g., acetate, propionate and butyrate) in the rumen. For NADH oxidation without ferredoxin oxidation, increasing $P_{H_2}$ within the rumen physiological range decreased $F_T$ from unity to zero for different NAD+ to NADH ratios and pH of 6.2 and 7.0, which indicates thermodynamic control of $P_{H_2}$. For NADH oxidation with ferredoxin oxidation, increasing $P_{H_2}$ within the rumen physiological range decreased $F_T$ from unity at pH of 7.0 only. For the acetate to propionate conversion, $F_T$ increased from 0.65 to unity with increasing $P_{H_2}$, which indicates thermodynamic control. For propionate to acetate and butyrate to acetate conversions, $F_T$ decreased to zero below the rumen range of $P_{H_2}$, indicating full thermodynamic suppression. For methanogenesis by archaea without cytochromes, $F_T$ differed from unity only below the rumen range of $P_{H_2}$, indicating no thermodynamic control. This theoretical investigation shows that thermodynamic control of $P_{H_2}$ on individual VFA produced and associated yield of hydrogen and methane cannot be explained without considering NADH oxidation.
Introduction

Carbohydrates ingested by ruminants are degraded into monomers by action of rumen microbial enzymes and subsequently fermented to products such as volatile fatty acids (VFA) and alcohols. The most common pathway of hexose metabolism in rumen microbes is glycolysis, which yields two equivalents of pyruvate, ATP and NADH. The NADH, a cofactor carrying electrons, needs to be oxidized back to NAD\(^+\) to keep the glycolysis possible and to maintain further metabolic steps of the overall microbial metabolism that depend on pyruvate [1, 2]. The oxidation of NADH to NAD\(^+\) may be directly coupled to the product formation from pyruvate that follows glycolysis. Production of butyrate couples the oxidation of NADH to the reduction of acetooacetyl-CoA as well as crotonyl-CoA [3]. Various fermentative micro-organisms are also able to convert pyruvate into ethanol, lactate or succinate [4], which results in direct oxidation of NADH. Acetate is quantitatively the main VFA in the rumen, but its production from pyruvate is not directly coupled to the oxidation of NADH. In this case, NADH is oxidized via \(\text{H}_2\) production, which is thermodynamically inhibited at elevated hydrogen partial pressure \((P_{\text{H}_2})\). Oxidation of NADH may be thermodynamically feasible by coupling it to the oxidation of reduced ferredoxin [5]. Many methanogenic archaea utilize \(\text{H}_2\) to reduce \(\text{CO}_2\) to \(\text{CH}_4\). This keeps \(P_{\text{H}_2}\) at a low level, which enables NADH oxidation in bacteria that are not able to directly couple NADH oxidation to reduction of metabolites [4].

Multiple estimates of rumen VFA (e.g., acetate, propionate, butyrate and other) production from feed substrate have been reported in literature based on factors including type of organic matter fermented and type of diet [6]. Such estimates are required in rumen models to predict the amount and type of VFA entering the intermediary metabolism of ruminants. Another application of these estimates is the prediction of enteric CH\(_4\) production, which is of interest in terms of the environment. Accuracy of predicted CH\(_4\) emission by the model used by Bannink et al. [7] appeared to be mostly affected by the error in the representation of the molar proportion at which individual VFA are produced. Reducing this error contributes to more adequate prediction of enteric CH\(_4\) emission [8]. A recent metabolic model of mixed culture fermentation [9] represents how incorporation of thermodynamically controlled cofactor dynamics may improve the prediction of end products such as VFA from glucose fermentation.

Thermodynamic control of rumen fermentation pathways by \(P_{\text{H}_2}\) has been investigated to explain variation in observed VFA concentrations [10]. Thermodynamic control is often evaluated by Gibbs energy change \((\Delta G)\). Negative values of \(\Delta G\) indicate a reaction to proceed in the forward direction, positive values in the reverse direction, and \(\Delta G = 0\) indicates equilibrium. Using \(\Delta G\), it has been explained that increased concentrations of \(\text{H}_2\) result in a shift to pathways forming propionate at the expense of acetate as an alternative way of accepting electrons to \(\text{H}_2\)-forming pathways because the latter become thermodynamically less favorable [11]. Reaction rates of fermentation pathways have been prescribed by setting the quotient of kinetic rate constants for the forward and reverse reaction equal to the thermodynamic equilibrium constant [10, 12]. However, the quotient of the rate laws for reverse and forward reaction does not necessarily reflect the stoichiometry of a reaction and is not in general similar to the thermodynamic equilibrium constant. Besides, classical thermodynamic functions such as \(\Delta G\) have no implications for the magnitude of reaction rate, except for near-equilibrium situations [13], and may not rigorously account for the thermodynamic driving force on reaction [14]. Furthermore, various investigations on the control of \(P_{\text{H}_2}\) on rumen fermentation have ignored the role of NAD, or have mentioned it without quantifying the redox state as affected by varying \(P_{\text{H}_2}\) (e.g., [1, 2, 11, 12]). The aim of the present study is to quantify the thermodynamic effect of
on the reaction rate of specific fermentation pathways, NADH oxidation and methanogenesis in the rumen.

**Methods**

**Metabolic pathways**

Glucose can be fermented via various pathways depending on the microbial diversity and the conditions in the rumen environment. To quantify the effect of $P_{H_2}$ on reaction rates, five rumen glucose fermentation pathways each yielding different VFA, three $H_2$-dependent interconversions of VFA (viz. acetate to propionate, propionate to acetate and butyrate to acetate), oxidation of NADH with and without reduced ferredoxin oxidation, and methanogenesis were considered (Table 1). Selected reactions focus on formation of VFA and have been taken from ref. [3] for reactions b, j and k; ref. [4] for reactions a, g, h and i; ref. [15] for reaction f; ref. [16] for reaction c when butyrate is produced via the kinase route; and ref. [17] for reactions c when butyrate is produced via the CoA-transferase route, and reactions d and e. Conversions of acetate to butyrate, butyrate to propionate and propionate to butyrate are discussed, but the effect of $P_{H_2}$ on reaction rate is not shown because these conversions do not yield any $H_2$ or have limited physiological significance. Glucose fermentation reactions in Table 1 are ordered following the stoichiometry of $H_2$ formation. The number of NADH oxidized with $H_2$ formation for the interconversion reactions were obtained considering reactions f and g as linear combinations of reactions a and d, and reaction h as a linear combination of reactions a and c. Various other cofactors are involved in the microbial degradation of glucose as well, but only NAD is involved in both the glycolysis and in further metabolic pathways of pyruvate to VFA or other fermentation products. The redox state of this cofactor explains the shift in pathways of glucose fermentation and therefore the focus is on oxidation of NADH. Besides being involved in NADH oxidation via confurcation, ferredoxin is involved in the production of acetate and butyrate, which explains why the $H_2$ yield reported for metabolic pathways in Table 1 may not be equal to the number of NADH oxidized with $H_2$ formation.

Moreover, as has been compared to the formation of propionate at the expense of acetate, reductive acetogenesis may be a potential alternative $H_2$ sink to methanogenesis in the rumen [18], but will not be considered in the present investigation. Although this conversion is associated with carbon turnover and is common in environments such as the human colon [17] and foregut of kangaroos and wallabies [19], acetogenic bacteria in the rumen have been hypothesized to be unable to compete for $H_2$ with the methanogens (e.g., [20]). Unless mentioned otherwise, respiration was assumed not to be occurring within the rumen microbiome.

**Thermodynamic potential factor**

The thermodynamic control on rates of rumen fermentation pathways was quantified using the thermodynamic potential factor ($F_T$) as derived by Jin and Bethke [14]. This factor modifies commonly used rate laws and makes them thermodynamically consistent by accounting for the difference between the energy available through fermentation and the energy conserved. The energy available through fermentation is calculated from the ratio of reactants and products, which is associated with the progress of the forward and reverse direction of a reaction. A rate law that accounts for the forward as well as the reverse direction of a reaction is thermodynamically consistent and may be represented as:

$$ r = k[X] \frac{[S]}{[S] + K_s F_T},$$

(1)
with the kinetic rate constant \( k \), the microbial biomass concentration \( [X] \), the substrate concentration \( [S] \) and the half-saturation constant \( K_S \). Kinetic rate laws, however, are often developed assuming that a large thermodynamic force drives a metabolic reaction forward. Under this condition, kinetic rate laws do not need to be corrected with any factor like \( F_T \). This assumption is reasonable when the environment is rich in chemical energy, that is where the metabolic reaction is far from equilibrium. The \( F_T \) is mathematically represented as:

\[
F_T = 1 - \exp\left(-\frac{\Delta G_A - \Delta G_C}{\chi RT}\right),
\]

where \( \Delta G_C \) is the energy conserved (J mol\(^{-1}\)), which is commonly determined from the number of ATP produced times the Gibb's energy of phosphorylation (\( Y_{ATP} \Delta G_F \)). \( \Delta G_F \) is approximated by 44 kJ (mol ATP)\(^{-1}\) for rumen microbes in the present study; \( \Delta G_A \) is the energy available through fermentation (J mol\(^{-1}\)); \( \chi \) is the average stoichiometric number representing the number of times elementary steps of product formation occurs relative to the main reactant; \( R \) the gas constant (8.31 J mol\(^{-1}\) K\(^{-1}\)); \( T \) the temperature (312K in the rumen); this makes \( F_T \) dimensionless by definition. For \( \Delta G_A > \Delta G_C \) and common values of \( T \) and \( \chi \), \( F_T \) approaches 1 also designated as unity, and the net reaction rate is 100% of the forward rate, and \( F_T \) can be neglected in determining rates of reaction in microbial metabolism. When \( \Delta G_A \) approaches \( \Delta G_C \) the forward and reverse reaction approach equilibrium, which is reflected in \( F_T \) approaching zero. For \( \Delta G_A < \Delta G_C \), \( F_T \) becomes negative, suggesting that a reaction net proceeds in the reverse direction; for \( \Delta G_A \ll \Delta G_C \) and common values of \( T \) and \( \chi \), \( F_T \) approaches \(-\infty\).

### Table 1. Possible glucose fermentation pathways to VFA (Ac\(^\text{−}\), Pr\(^\text{−}\) and Bu\(^\text{−}\) for acetate, propionate and butyrate, respectively), volatile fatty acid (VFA) interconversions, hydrogenase-catalyzed NADH oxidation and methanogenesis in the rumen and their yield of ATP (\( Y_{ATP} \)), number of NADH to be oxidized with \( H_2 \) formation (\( Y_{NADH} \)), the standard reaction Gibbs energy (\( \Delta G^o \)) in kJ mol\(^{-1}\), standardized to concentrations of 1 M, pH of 0, gas pressure of 1 bar) adjusted to 312K, and the average stoichiometric number \( \chi \).

| Microbial conversion | \( Y_{ATP} \) | \( Y_{NADH} \) | \( \Delta G^o \) | \( \chi \) |
|----------------------|--------------|--------------|--------------|--------|
| Glucose fermentation |              |              |              |        |
| a) \( C_6H_{12}O_6 + 4H_2O \rightarrow 2 Ac^− + 2HCO_3^− + 4H_2 + 4H^+ \) | 4 | 2 | -52 | 4 |
| b) \( C_6H_{12}O_6 + 2.67H_2O \rightarrow 0.67 Ac^− + 0.67 Bu^− + 2HCO_3^− + 3.33H^+ + 6.76H_2 \) | 3.33 | 0.67 | -11 | 3.33 |
| c) \( C_6H_{12}O_6 + 2H_2O \rightarrow Bu^− + 2HCO_3^− + 2H_2 + 3H^+ \) | 3 | 0 | -138 | 3 |
| d) \( C_6H_{12}O_6 + H_2O \rightarrow Ac^− + Pr^− + HCO_3^− + H_2 + 3H^+ \) | 3.67 | 0 | -159 | 3 |
| e) \( C_6H_{12}O_6 \rightarrow 0.67 Ac^− + 1.33 Pr^− + 0.67HCO_3^− + 2.67H^+ \) | 2.67 | -0.67 | -196 | 2.67 |
| VFA interconversion |              |              |              |        |
| f) \( Ac^− + HCO_3^− + H^+ + 3H_2 \rightarrow Pr^− + 3H_2O \) | 0 | -2 | -113 | 1 |
| g) \( Pr^− + 3H_2O \rightarrow Ac^− + HCO_3^− + H^+ + 3H_2 \) | 0.33 | 2 | 113 | 2 |
| h) \( Bu^− + 2H_2O \rightarrow 2Ac^− + H^+ + 2H_2 \) | 0.33 | 2 | 86 | 2 |
| Cofactor oxidation |              |              |              |        |
| i) \( NADH + H^+ \rightarrow NAD^+ + H_2 \) | 0 | NA | -25 | 1 |
| j) \( NADH + Fd_{RED} + 3H^+ \rightarrow NAD^+ + Fd_{OX} + 2H_2 \) | 0 | NA | -102 | 2 |
| Methanogenesis |              |              |              |        |
| k) \( HCO_3^− + H^+ + 4H_2 \rightarrow CH_4 + 3H_2O \) | 1.5 or 0.5\(^a\) | 0 | -172 | 2 |

\(^a\) Butyrate production via the kinase route
\(^b\) Either for butyrate production via the kinase route or a linear combination of reaction a) and 2Ac\(^−\) + 2C\(_6\)H\(_{12}\)O\(_6\) + 2H\(_2\)O \rightarrow 3Bu\(^−\) + 4HCO\(_3\)\(^−\) + 2H\(_2\) + 5H\(^+\) for butyrate production via the CoA-transferase route
\(^c\) Propionate production via succinate
\(^d\) Propionate production via lactate
\(^e\) for archaeal species with and without cytochromes

doi:10.1371/journal.pone.0161362.t001

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PLOS ONE | DOI:10.1371/journal.pone.0161362 October 26, 2016 4/18
suggesting that the forward reaction is even negligibly small compared to the reverse reaction. Negative \( F_T \) may not be useful for prediction of reaction rate since common rate laws of such as the Monod equation are not used for reactions that overall proceed in the reverse direction. At a microbial level, a reverse reaction would consume energy rather than contribute to a cell’s energy budget, which is not enzymatically supported and the metabolism may stop.

The \( \Delta G_A \) is further specified as:

\[
\Delta G_A = -\Delta G^o - RT \ln Q,
\]

with \( \Delta G^o \) the standard reaction Gibbs energy, and \( Q \) the reaction quotient, which is:

\[
Q = \prod \alpha_j^{v_j},
\]

where \( \alpha_j \) denotes the concentration of substance J, and \( v_j \) its corresponding stoichiometric number in the chemical equation, which is positive for products and negative for reactants. Substituting Eqs 3 and 4 into Eq 2 yields:

\[
F_T = 1 - Q^{-1} \exp \left( \frac{\Delta G^o + \Delta G_C}{\zeta RT} \right).
\]

Substances that are in the gaseous state under rumen conditions are represented in partial pressure instead of aqueous concentrations; water activity is assumed to be 1 and omitted from the reaction quotient in any case. To illustrate, \( F_T \) for the glucose to acetate conversion (reaction a, Table 1) by substituting into Eq 5 gives:

\[
F_T = 1 - [\text{Ac}^-]^{0.5} [\text{HCO}_3^-]^{0.5} P_{\text{H}_2}[\text{H}^+] [\text{C}_6\text{H}_{12}\text{O}_6]^{-0.25} \exp \left( \frac{-52 \cdot 10^3 + 4 \cdot 44 \cdot 10^3}{4 \cdot 8.31 \cdot 312} \right).
\]

**Reaction specific energy conservation and elementary reaction steps**

In anaerobic fermentation, ATP is mostly produced by substrate level phosphorylation, but some electron transport phosphorylation may take place during fermentations [16, 21]. Reaction steps associated with electron transport phosphorylation include fumarate reduction in the pathways of pyruvate to propionate, crotonyl-CoA reduction in the pathway of acetyl-CoA to butyrate, and the oxidations of succinate and butyryl-CoA in the syntrophic conversions of propionate to acetate and butyrate to acetate. Yield of ATP (shown in Table 1 for every reaction considered) was assumed to be 2 for the common pathway of glucose to 2 pyruvate, and 2, 1.33 and 1 for the conversion of 2 pyruvate into 2 acetate, 2 propionate and 1 butyrate, respectively [4, 9]; 0.33 for the oxidations of propionate and butyrate to acetate [4]; 0 for the reduction of acetate and \( \text{HCO}_3^- \) to propionate [22]; and 1.5 or 0.5 per equivalent of \( \text{CH}_4 \) produced by archaean species with and without cytochromes [3, 23]. Since ATP was described to be generated by substrate level phosphorylation only for *Clostridium pasteurianum* [3], and uncertain ATP yield from electron transport phosphorylation was predicted for rumen *Butyrivibrios* [16], reference values of ATP yield used in the present study may be subject to revision.

For microbial catabolism, likely rate-determining steps may be substrate level phosphorylation during fermentation, proton translocation, substrate activation or electron transfer to extracellular electron acceptors [14]. For rumen glucose fermentation, the rate-determining step was chosen to be equal to the ATP yield from substrate level phosphorylation, which results in \( \chi \) equal to 4, 3.33, 3, 3 and 2.67 for reactions a to e (Table 1). When assuming reactivity of NADH oxidation to be dominated by hydride transfer [24], the rate-determining step
occurs only once per equivalent of NADH oxidized, indicating $\chi = 1$ for NADH oxidation without ferredoxin oxidation. Although various aspects of hydrogenase-catalyzed cofactor oxidation require further clarification, a hydride intermediate may also be formed in the oxidation of reduced ferredoxin [25]). NADH oxidation via electron confurcation (i.e., combining electrons from two dissimilar donors to generate a single product such as H$_2$) would then be associated with two hydrides intermediates, indicating $\chi = 2$ for NADH oxidation with ferredoxin oxidation. The rate-determining step for the reduction of acetate and HCO$_3^-$ (reaction f, Table 1) was assumed to be the activation of acetate to acetyl phosphate. This activation occurs once per equivalent of acetate, which makes $\chi = 1$. The butyryl-CoA and succinate oxidations are the energetically most demanding steps in the overall pathways of butyrate and propionate fermentation [4], where electron transfer was taken as the rate-determining step. Two electrons are transferred for both the oxidation of butyryl-CoA and succinate, which indicates $\chi = 2$ for both conversions. The rate-determining steps in methanogenesis, with and without the involvement of cytochromes, were assumed to be the methyltetrahydromethanopterin–coenzyme M methyltransferase and the reduction of the disulfide of coenzymes B and M, respectively. Both steps are coupled to the translocation of two sodium ions [23], which occurs once per equivalent of CH$_4$ produced, indicating $\chi = 2$.

Continuous input variables and uncertainty of $F_T$

For reactions a to h and k, concentrations were 1 mM hexose, 60 mM acetate, 20 mM propionate, 12.5 mM butyrate and 40 mM bicarbonate, 0.25 bar partial pressure of CH$_4$ and pH was equal to 6.45; for reaction j, $F_{\text{RED}}^{2}/F_{\text{OX}}$ was equal to 9. Values for $\Delta G^\circ$ of fermentation pathways and standard redox potentials of cofactors were taken from refs. [3, 10]. Values of $\Delta G^\circ$ of metabolite formation were adjusted to rumen temperature using the Van’t Hoff equation (e.g., [26]).

The uncertainty of $F_T$ to variation in inputs other than $P_{\text{Hi}}$ was assessed for the five glucose fermentation pathways, the three VFA interconversions and methanogenesis (Table 1). Ten thousand different samples were drawn randomly from uniform distributions for glucose, acetate, propionate, butyrate concentrations, pH, $P_{\text{CO}_2}$, $P_{\text{CH}_4}$ and $\Delta G_P$ ranging from 0.1 to 2.0 mM, 35 to 90 mM, 7 to 30 mM, 5 to 21 mM, 5.7 to 7.2, 0.35 to 0.80 bar, 0.15 to 0.35 bar and 35 to 50 kJ·mol$^{-1}$, respectively. For completeness, proton concentrations were calculated from pH and HCO$_3^-$ concentrations were calculated using the Henderson-Hasselbalch equation (e.g., [27]).

Uncertainty of $F_T$ approaches zero when $F_T$ approaches unity. If $F_T$ of a specific reaction deviated from unity for the range of $P_{\text{Hi}}$, considered, a 95% confidence interval of $F_T$ was calculated for 10 values of $P_{\text{Hi}}$, for which $F_T$ was close to zero at the previously mentioned fixed concentrations. Values of $P_{\text{Hi}}$ increased exponentially in steps according to $P_{\text{Hi},n} = a \cdot b^{n-1}$, where $a$ is the start value, $b$ is the factor by which $P_{\text{Hi},n}$ increases per step, and $n$ runs from 1 to 10 for the number of steps. The exact values of $a$ and $b$ were chosen based on the visual representation of the uncertainty by the error bar. Applying this, the uncertainty of $F_T$ was assessed for $P_{\text{Hi}}$ at $\{2.00 \times 10^{-5}, 2.60 \times 10^{-5}, \ldots, 2.12 \times 10^{-4}\}$ bar for methanogenesis yielding 0.5 ATP, $\{6.00 \times 10^{-4}, 8.10 \times 10^{-4}, \ldots, 8.94 \times 10^{-3}\}$ bar for methanogenesis yielding 1.5 ATP, $\{7.70 \times 10^{-5}, 9.63 \times 10^{-5}, \ldots, 5.74 \times 10^{-4}\}$ bar for acetate to propionate conversion, $\{5.00 \times 10^{-6}, 6.50 \times 10^{-6}, \ldots, 5.30 \times 10^{-5}\}$ bar for propionate to acetate conversion, and $\{1.95 \times 10^{-6}, 2.93 \times 10^{-6}, \ldots, 7.50 \times 10^{-5}\}$ bar for butyrate to acetate conversion. The actual ranges of the 95% confidence intervals of $F_T$ depends on metabolite concentrations and values of $\Delta G^\circ$, $\Delta G_C$ and $\chi$, explicitly shown for the particular conversion of glucose into two equivalents of acetate (Eq 6). Eq 6 also shows the nonlinearity of $F_T$ to its input, which makes the 95% confidence intervals asymmetric.
Calculation of the 95% confidence intervals of $F_T$ at discrete values of $P_{H_2}$ and plotting of $F_T$ as a function of $P_{H_2}$ was performed in R statistical software. Code is provided as supporting information (S1–S3 Files).

Results and Discussion

Glucose fermentation and NADH oxidation

The $F_T$ for the fermentation pathways a to d (Table 1) did not deviate from unity for $P_{H_2}$ between $2 \times 10^{-5}$ and $5 \times 10^{-2}$ bar and had zero uncertainty (Fig 1), which is inherent to $F_T$ approaching unity. This indicates these fermentation reactions proceed far from thermodynamic equilibrium and implies no inhibition on reaction rates since $P_{H_2}$ in the rumen varies between $2 \times 10^{-4}$ and $1 \times 10^{-2}$ bar [2]. No $F_T$ curve is shown for the conversion of glucose into 0.67 equivalents of acetate and 1.33 equivalents of propionate because it does not involve $H_2$. The actual value of $F_T$ for this conversion also yielded unity (result not shown) and indicates no thermodynamic inhibition of this fermentation pathway under the conditions assumed and range of $P_{H_2}$ considered. In this investigation, we assumed an ATP yield of 3 per equivalent of

![Fig 1. Thermodynamic potential factor ($F_T$) as a function of $P_{H_2}$ for glucose fermentation pathways and methanogenesis.](https://example.com/figure1.png)

The black line for glucose to VFA is valid for the reactions a to d (yielding acetate, propionate or butyrate), the solid and dotted gray lines represent methanogenesis with 0.5 and 1.5 mol of ATP per mol of $CH_4$, respectively; a more detailed description of the glucose fermentation pathways to VFA and methanogenesis is given in Table 1. Confidence intervals represent uncertainty of $F_T$ to variation in inputs other than $P_{H_2}$. Vertical lines demarcate the rumen physiological range of $P_{H_2}$. A log scale is used to plot the x-axis.

doi:10.1371/journal.pone.0161362.g001
butyrate if only substrate level phosphorylation takes place. Accounting for electron transport phosphorylation as well would predict an ATP yield of ~ 4.5 per equivalent of glucose [16]. Production of propionate via succinate has also been mentioned to yield 4 ATP per equivalent of glucose [28]. Adjusting $\Delta G_C$ of reactions associated with propionate and butyrate to these higher yields of ATP still did not make $F_T$ deviate from unity for the considered range of $P_{H_2}$ (result not shown).

Absence of thermodynamic inhibition for any of the glucose fermentation pathways is not in line with conclusions drawn previously [11], where the conversion of glucose into VFA was considered to be directly affected by the level of $H_2$. For common values of $T, F_T$ approaches unity when $\Delta G_A \gg \Delta G_C$, representing the far-from-equilibrium situation. This applies to the glucose fermentation pathways considered indicating that $\Delta G$ cannot be used as a measure of reaction rate for these reactions. This is in accordance with the fact that classical thermodynamic functions such as $\Delta G$ have no implications for magnitude of reaction rate, except for near-equilibrium situations [13]. Only a difference between $\Delta G_A$ and $\Delta G_C$ closer to zero than approximately $-20 \text{ kJ mol}^{-1}$, which may be the cutoff for near-equilibrium, makes $F_T$ deviate from unity. Additional evidence for $\Delta G \approx -20\text{kJ mol}^{-1}$ as a cutoff value for inhibited progress of microbial metabolism is given by Schink [29] who assumed a heat loss of about $20 \text{ kJ mol}^{-1}$ for irreversible metabolic processes that generate ATP. However, it was experimentally shown that syntrophic bacteria metabolize up to a zero difference between $\Delta G_A$ and $\Delta G_C$ [30], which corresponds to $F_T = 0$.

The $F_T$ for NADH oxidation without reduced ferredoxin oxidation decreased to zero upon an increase of $P_{H_2}$ from $2 \cdot 10^{-4}$ to $1 \cdot 10^{-2} \text{ bar}$, whereas $F_T < 1$ may already be obtained at $P_{H_2} < 5 \cdot 10^{-3} \text{ bar}$ for a high $\text{NAD}^+$ to NADH ratio and $pH = 7.0$ (Fig 2a). The $F_T$ for NADH oxidation with reduced ferredoxin oxidation decreased to zero at $P_{H_2} > 1 \cdot 10^{-2} \text{ bar}$, whereas $F_T < 1$ may already be obtained at $P_{H_2} > 2 \cdot 10^{-4} \text{ bar}$ when $pH = 7.0$ (Fig 2b). The actual value of $F_T$ depends on pH and $\text{NAD}^+$ to NADH ratio. Partial pressure of $H_2$ and intracellular pH of microbes in the rumen are assumed to vary between $2 \cdot 10^{-4}$ and $1 \cdot 10^{-2} \text{ bar}$ [2], and 6.2 and 7.0 [31], respectively. Shortly after new feed enters the rumen, the rate of fermentation will increase, which results in a high $P_{H_2}$ [32] and a low pH; whereas during fasting, $P_{H_2}$ will be low and pH high. In an experimental study in which the effects of starch type and level on rumen fermentation were evaluated [33], the lowest acetate to propionate ratio was observed at 2 h after feeding, whereas the lowest pH was observed at 4 h after feeding. Achieving the lowest acetate to propionate ratio before the lowest pH may suggest that after feed consumption the increase in pH and $\text{NAD}^+$ to NADH ratio. Partial pressure of $H_2$ and intracellular pH of microbes in the rumen are assumed to vary between $2 \cdot 10^{-4}$ and $1 \cdot 10^{-2} \text{ bar}$ [2], and 6.2 and 7.0 [31], respectively. Shortly after new feed enters the rumen, the rate of fermentation will increase, which results in a high $P_{H_2}$ [32] and a low pH; whereas during fasting, $P_{H_2}$ will be low and pH high. In an experimental study in which the effects of starch type and level on rumen fermentation were evaluated [33], the lowest acetate to propionate ratio was observed at 2 h after feeding, whereas the lowest pH was observed at 4 h after feeding. Achieving the lowest acetate to propionate ratio before the lowest pH may suggest that after feed consumption the increase in $P_{H_2}$ occurs faster than the decrease in pH. This indicates that elevated $P_{H_2}$ thermodynamically inhibits NADH oxidation shortly after feeding, but this is compensated by decreased pH later.

Although effects of the redox state of ferredoxin on the thermodynamic inhibition of NADH oxidation are not explicitly shown, ferredoxin is reduced during fermentation and the $\text{Fd}_{\text{RED}}^{2-}$ to $\text{Fd}_{\text{OX}}$ ratio, which was assumed to be 9, may increase in response to increased metabolism shortly after ingestion of feed. If an increased $\text{Fd}_{\text{RED}}^{2-}$ to $\text{Fd}_{\text{OX}}$ ratio applies to rumen bacteria, the inhibition of NADH oxidation is potentially alleviated. To evaluate this alleviation, the solid, dashed and dot-dashed lines in Fig 2b may, alternative to keeping the $\text{Fd}_{\text{RED}}^{2-}$ to $\text{Fd}_{\text{OX}}$ constant at 9 and $\text{NAD}^+$ to NADH ratios of 9, 3 and 1, correspond to keeping the $\text{NAD}^+$ to NADH ratio constant at 9 and $\text{Fd}_{\text{RED}}^{2-}$ to $\text{Fd}_{\text{OX}}$ ratios of 9, 27 and 81, respectively. This implies that the value of $F_T$ is closer to 1 for more reduced ferredoxin, which weakens the thermodynamic force that inhibits NADH oxidation. Since ferredoxin is involved in the pathway from pyruvate to acetate and butyrate only and not in the glycolysis, whereas NAD may be involved in both pathways, the $\text{NAD}^+$ to NADH ratio may change more rapidly after feeding than the $\text{Fd}_{\text{RED}}^{2-}$ to $\text{Fd}_{\text{OX}}$ ratio. Inhibition of NADH oxidation may therefore occur shortly after feeding, but may be compensated later. Nonetheless, the present study demonstrates that
the mechanism of NADH oxidation is critical for the magnitude of its inhibition; the inhibition of NADH oxidation is also determined by the thermodynamic state of the rumen with $P_{H_2}$ and intracellular pH both being important determinants.

The NAD$^+$ to NADH ratio is sometimes assumed to be in thermodynamic equilibrium with $P_{H_2}$ [34], or in other words, $F_T$ is assumed zero for any value of $P_{H_2}$. For rumen bacteria incapable of confusion this implies the NAD$^+$ to NADH ratio is $\geq 9$ at $P_{H_2} = 2 \times 10^{-4}$ and $\leq 1$ at $P_{H_2} = 1 \times 10^{-2}$ bar (Fig 2a); for rumen bacteria in which confusion does take place this implies the NAD$^+$ to NADH ratio is $\geq 9$ for $P_{H_2} \leq 1 \times 10^{-2}$ bar (Fig 2b). The NAD$^+$ to NADH ratio was reported to be 1.4 to 2.6 in rumen microbes [35], 1.1 to 2.7 for Escherichia coli [36], and was reported to be $< 9$ in living cells [3]. These ratios largely fall within the range of our prediction but tend to be at the edge of physiological feasibility and the NAD$^+$ to NADH ratio in bacteria...
incapable of confurcation may underestimated at elevated $P_{H_2}$. Although many anaerobic and syntrophic bacteria contain enzymes that catalyze electron confurcation, it is not evident whether many of the bacteria belonging to the core community in the rumen (e.g., *Prevotella, Fibrobacter, Ruminococcaceae, Bacteroidales*; [37]) employ this mechanism. *Ruminococcus albus* 7 that is part of the rumen core community employs this mechanism [38]. In this strain, genes encoding for the hydrogenase enzyme involved in electron confurcation had a similar transcript abundance in mono- and biculture. In contrast, genes encoding for a different hydrogenase that reduces protons to molecular hydrogen using reduced ferredoxin only was 90-fold upregulated in mono- compared to biculture [39]. This suggests that the confurcating hydrogenase functions in central metabolism regardless of external $P_{H_2}$. Nonetheless, increased propionate to acetate ratios [33] and production of lactate being reported in response to feeding [40] may indicate these latter two ways of NADH oxidation are important alternatives for ferredoxin dependent oxidation of NADH. Direct evidence of how these mechanisms are applied by rumen bacteria is lacking however.

Given that the NAD$^+$ to NADH ratio becomes less than or equal to 1 (Fig 2a), glycolytic reactions may be downregulated. Glycolytic activity of *Caldicellulosiruptor saccharolyticus* was found not to be completely inhibited at a NAD$^+$ to NADH ratio equal to 1 [41], which may allow metabolic activity at ratios $< 1$. Nonetheless, highly reduced NAD is reconditioned to more oxidized NAD by the upregulation of the production of metabolites such as lactate and ethanol, as explained for gut microbiota [42]. This upregulation may take place in addition to increased proportions of propionate production. However, the production of lactate and ethanol is less favorable for microbial growth because conversion of pyruvate to either lactate or ethanol does not yield any ATP, unlike the conversion of pyruvate to acetate or butyrate, and to propionate via succinate. Another way in which bacteria may control $P_{H_2}$ and the redox state of NAD in the rumen environment is the production of formate. Formate may be produced when pyruvate is converted to acetyl-CoA as an alternative for the oxidation of reduced ferredoxin [4]. Formate can be converted to $H_2$ and $CO_2$, but may also be directly used for $CH_4$-production by methanogens [43]. In the latter case, no $H_2$ is produced and the synthesis of formate serves as a potential mechanism to maintain low $P_{H_2}$ [44].

The present theoretical exercise indicates that, in the rumen, $P_{H_2}$ does not directly control the glucose fermentation pathways. However, depending on mechanism and pH, $P_{H_2}$ does thermodynamically control NADH oxidation, which influences VFA production. NAD$^+$ to NADH ratio as a key controller of fermentation end product formation is widely recognized in literature (e.g., [45, 46]). When the NAD$^+$ to NADH ratio is low, the metabolism needs to yield more reduced products to oxidize NADH [41, 42]. Production of butyrate and propionate from reactions c and d both oxidize all NADH obtained from glycolysis back to NAD$^+$ (Table 1) but does not explain why elevated propionate but no elevated butyrate is found at increased $P_{H_2}$. A difference between these pathways is the $H_2$ yield of 2 and 1 equivalents per equivalent of glucose from reaction c and d, respectively. The higher $H_2$ yield associated with butyrate production (reaction c) will inhibit NADH oxidation more than propionate production (reaction d), which explains why propionate production is more upregulated than butyrate production at increased $P_{H_2}$. Furthermore, production of butyrate yields only one VFA per equivalent of glucose (reaction c), whereas production of acetate and propionate (reaction d) yields two VFA per equivalent of glucose, which makes the rumen environment more acidic. Shortly after a meal, propionate may be produced via lactate production, via reaction e. Lactate is a stronger acid than propionate and makes the rumen environment even more acidic. In addition to the net 0.67 NADH oxidized back to NAD$^+$, the acidic environment promotes the oxidation of NADH. Less inhibition of NADH oxidation at lower pH (Fig 2) explains why, at
neutral or alkaline pH, propionate production is more effective in maintaining the NAD\(^+\) to NADH ratio than butyrate production [9].

Thermodynamic control of \(P_{H2}\) on NADH oxidation but not on the glucose fermentation pathways, is also in line with the statement that the NAD\(^+\) to NADH ratio determines the profile of VFA produced with rumen fermentation [2]. One may designate this as the dynamic control of \(P_{H2}\) on rumen fermentation pathways. Ghimire et al. [12], building on the Molly cow model, which includes a representation of rumen fermentation processes, attempted to account for the effect of the thermodynamic state of the rumen environment on the interconversion between acetate and propionate. Besides keeping \(P_{H2}\) constant in the calculation of these rate constants, they did not consider the NAD\(^+\) to NADH ratio, which might have caused their model not to perform well in predicting observed variation in ruminal VFA production. Future modeling attempts might benefit from a representation of the NAD\(^+\) to NADH ratio.

Even though an empirical relationship between \(P_{H2}\) and proportion at which individual VFA are produced may appear from experimental data, the validity of a NAD-driven mechanistic prediction of metabolic end products is supported by the work of Salem et al. [47]. They used the NAD\(^+\) to NADH ratio as a key controller of the type of glucose degradation products to be formed. Although their modeling effort deals with the myocardial energy metabolism, which partly differs from the energy metabolism of anaerobic bacteria, a similar approach may be applied for estimating rumen fermentation products. Oxygen concentration in blood, like \(P_{H2}\) in anaerobic environments, dictates redox conditions and consequently the NAD\(^+\) to NADH ratio. Therefore, predicting the production of individual VFA in the rumen might benefit from using the NAD\(^+\) to NADH ratio as a controlling factor as was suggested from an evaluation of various VFA prediction models [48]. Future modeling attempts might benefit from a representation of the NAD\(^+\) to NADH ratio.

The NAD\(^+\) to NADH ratio as a key controller of the type of VFA produced explains why feeding rapidly degradable carbohydrates induces a shift from acetate to propionate production in the rumen. This shift has been confirmed by various studies, among which a regression analysis of molar proportions of VFA production [6] and a metabolic model of mixed culture fermentations [9, 46]. Different carbohydrate polymers such as cellulose and amylose are broken down to the same monomers, and can be converted into the same fermentation end products. Degradation rate of carbohydrates, however, determines the magnitude of the increase in \(P_{H2}\) and decrease in NAD\(^+\) to NADH ratio obtained via the glycolysis, which controls pathways of VFA production from pyruvate. The ability of specific microbial species to catalyze the breakdown of a certain type of carbohydrate polymer might be related to the production of specific VFA, like starch hydrolysis favors propionate production. Nonetheless, this may also be regarded as the NAD\(^+\) to NADH ratio controls fermentation pathways, where the metabolic physiology of these species has been adapted to degrade specific carbohydrate polymers in the rumen.

**VFA interconversion**

Interconversion of VFA in the rumen has been discussed various times in the literature [10, 12]. After measuring VFA production rates in the rumen of lactating dairy cows by infusion of \(^{14}\text{C}\) labeled VFA, all six possible conversions between acetate, propionate and butyrate were confirmed to occur [49]. Of these conversions, acetate to propionate, propionate to acetate and butyrate to acetate are \(H_2\)-dependent.

Acetate to propionate conversion was observed at 2.0% and 2.6% of de novo synthesized acetate being converted into propionate at normal and low-roughage diets, respectively [49]. The higher conversion rate from the low-roughage diets may be attributed to higher \(P_{H2}\) from
the more rapidly degradable carbohydrates. To the authors’ knowledge, there is only one study that has described this conversion [15]. Therein, H₂-dependent propionate production from acetate and CO₂ by a pure culture of Desulfobulbus propionicus was reported. This particular study focuses on freshwater sediments and other microbial species might be responsible for this conversion in the rumen. The $F_T$ for this reaction increased from zero to unity for $P_{H_2}$ between approximately $1.5 \times 10^{-4}$ and $5 \times 10^{-4}$ bar, and zero is no longer within the confidence interval of $F_T$ for $P_{H_2} > 2.3 \times 10^{-4}$ bar (Fig 3a), implying the conversion of acetate to propionate to be controlled by $P_{H_2}$ and thermodynamically feasible under common rumen conditions. However, Laanbroek et al. [15] also reported not having observed any propionate from acetate and CO₂ in the presence of sulfate. Traces of sulfate may enter the rumen with regular feed-stuffs and will be metabolized by the microbes [50]. Especially when diets contain co-products from grain milling industries rumen sulfate concentrations may be high. Apart from $P_{H_2}$, also the sulfate concentration might control the rate of conversion of acetate to propionate. Besides, sulfate is an electron acceptor for respiration and will also compete for electrons and lower CH₄ production [51].

The H₂-dependent conversions of propionate and butyrate into acetate yield multiple equivalents of H₂ (reactions g and h, Table 1) and require very low $P_{H_2}$ to make them exergonic and proceed. For both reactions, values of $F_T \geq 0$ are within the 95% confidence interval for $P_{H_2} < 4 \times 10^{-5}$ bar (Fig 3b and 3c). This indicates these conversions do not occur under conditions that are common in the rumen where $P_{H_2}$ is usually higher. However, propionate or butyrate degrading bacteria may aggregate with H₂-consuming methanogens in typical syntrophic associations. This association of cells enables interspecies H₂ transfer by diffusion, and its flux is enhanced when the intermicrobial distance decreases [4, 52]. If this local interspecies H₂ transfer occurs, $P_{H_2}$ is lower than in other locations of the rumen, which makes the oxidation of propionate and butyrate exergonic. Furthermore, sulfate- and nitrate-reducing conditions have been reported to thermodynamically favor the degradation of propionate and butyrate [52]. Degradation of VFA under these conditions is H₂-independent, as was reported for propionate to acetate conversion in the presence of sulfate [15]. Therefore, the inhibition of $P_{H_2}$ on the
butyrate to acetate and propionate to acetate conversions might be counteracted in the presence of external electron acceptors. These conversions, though, require microbes capable of respiration.

Another pathway involving propionate to acetate conversion was described by De Bok et al. [53]. Using $^{13}$C labeled compounds, they found Smithella propionica to convert propionate into acetate and butyrate via a six-carbon intermediate. This particular conversion of propionate also gives physiological evidence for the conversion of propionate into butyrate. Hydrogen is not directly involved in this pathway and indicates the conversion of propionate into either acetate or butyrate is not affected by $P_{H_2}$. Depending on the $P_{H_2}$, the concentrations of acetate, propionate and butyrate, and the abundance of microbial aggregates, this particular propionate conversion into acetate and butyrate may enable butyrate oxidation in methanogenic ecosystems in case the classical propionate oxidation pathway would be endergonic [54]. In other words, this makes sense for the range of $P_{H_2}$ with $F_T > 0$ for butyrate oxidation and $F_T < 0$ for propionate oxidation. This range is negligibly small and below $2 \times 10^{-4}$ bar (Fig 3b and 3c), explaining why this particular oxidation of propionate is not expected to occur under rumen conditions.

Besides the VFA interconversions discussed in the paragraphs above, the acetate to butyrate conversion is ecologically significant [55] and seems to be more substantial than the other VFA interconversions in the rumen [49]. The final metabolic step of butyrate production, butyryl-CoA to butyrate, proceeds via butyrate kinase or via butyryl-CoA:acetate CoA-transferase [55]. Acetate to butyrate conversion may be described by the latter mechanism. For this conversion, apart from acetate, another substrate such as hexose is required to yield butyryl-CoA. From human colon microbiota, genes encoding for enzymes for both pathways were detected in various Butyrivibrio fibrisolvens strains and Clostridium species that also reside in the rumen. The butyryl-CoA:acetate CoA-transferase step does not yield $H_2$ and will not be affected by $P_{H_2}$. The $F_T$ for the conversion of glucose and acetate to butyrate did not deviate from unity for $P_{H_2}$ between $2 \times 10^{-5}$ and $5 \times 10^{-2}$ bar. Furthermore, for butyrate formed via both butyryl-CoA:acetate CoA-transferase and butyrate kinase, butyryl-CoA is formed from pyruvate with the same metabolic steps. Hence, the two mechanisms of butyrate production yield the same $H_2$ balance and oxidize equal equivalents of NADH to NAD$^+$ per equivalent of glucose (reaction c, Table 1; [3, 56]. Butyrate production via butyryl-CoA:acetate CoA-transferase and via butyryl kinase are therefore not controlled differently by the NAD$^+$ to NADH ratio and $P_{H_2}$. This would make a specific $P_{H_2}$-controlled flux of acetate to butyrate conversions in rumen dynamic modeling efforts redundant.

The ecological significance of the conversion of butyrate to propionate is low. Because $\Delta G^\circ$ for the propionate conversion into acetate and butyrate is nearly zero [57], the reverse reaction from butyrate to propionate might occur too. Furthermore, the metabolism of threonine fermentation in Clostridium propionicum has been described to yield both propionate and butyrate via 2-oxobutyrate [58]. The conversion of butyrate into propionate might occur as a side reaction, albeit the actual occurrence via 2-oxobutyrate is questionable.

The different fluxes of rumen VFA in the three-pool model of Sutton et al. [49] suggests that accounting for $P_{H_2}$ controlled VFA interconversions in dynamic model predictions is compatible with the conversions of acetate to propionate, butyrate to acetate and propionate to acetate. Nonetheless, these VFA interconversions are still controlled by the NAD$^+$ to NADH ratio of which the dynamics, described in the present investigation, may already explain an important part of the observed variation in the proportion of individual VFA. Prediction of VFA interconversion would also require information such as intermicrobial distance in syntrophic aggregates and concentration of external electron acceptors such as nitrate and sulfate. Including this information in a model next to control by NAD$^+$ to NADH ratio increases the model...
complexity, and it needs to be further investigated whether it aids in explaining observed variation in the proportion of individual VFA. Furthermore, functions that microorganisms carry out in certain experimental settings may differ greatly, depending on the presence or absence of other community members [42]. Applying this differing of functions to VFA interconversions makes dynamic predictions of rumen VFA concentrations uncertain.

**Methanogenesis**

The $F_T$ for methanogenesis increased from zero to unity for $P_{H_2}$ at $\sim 10^{-5}$ bar for archaea without cytochromes and at $\sim 10^{-3}$ bar for archaea with cytochromes (Fig 1). This indicates a certain threshold of $P_{H_2}$ to make methanogenesis proceed, depending on the physiology of the archaea. For methanogenesis by archaea with cytochromes, $F_T = 0$ for $P_{H_2} \approx 3 \cdot 10^{-3}$ bar and and based on the 95% confidence interval $F_T \leq 0$ for $P_{H_2} < 8 \cdot 10^{-4}$ bar (Fig 1). Rumen $P_{H_2}$ may be as low as $2 \cdot 10^{-4}$ bar [2] which will yield a negative $F_T$ and may explain why archaea with cytochromes are hardly found in the methanogenic community in the rumen [23, 59]. Given that $F_T$ approaches unity with rather minor uncertainty at $P_{H_2}$ as low as $2 \cdot 10^{-4}$ bar (Fig 1), methanogenesis by archaea without cytochromes is hardly restricted by the thermodynamic state of the rumen environment.

The amount of H$_2$ present in the rumen has been expressed as dissolved H$_2$ concentration [11]. It is common to express gas contents in pressure, but the possible occurrence of supersaturation of dissolved H$_2$ (e.g., [60]) would necessitate the use of dissolved H$_2$ concentration instead of $P_{H_2}$. Supersaturation, the violation of Henry’s Law, is the non-equilibrium condition between dissolved H$_2$ concentration and $P_{H_2}$ in the rumen headspace. The fact that archaea with cytochromes hardly exist in the rumen might suggest too low dissolved H$_2$ concentrations for their survival and negligible supersaturation of H$_2$. Furthermore, rumen contractions may prevent supersaturation of H$_2$ to occur. If supersaturation does occur in the rumen, survival of archaea with cytochromes may be enabled and the NAD$^+$ to NADH ratio may become lower than indicated in the present study.

Several studies have recognized the importance of adequate coefficients of production rate of individual VFA to accurately predict CH$_4$ [7, 8, 12]. The present finding that, under common rumen conditions, VFA dynamics rather than methanogenesis is controlled by $P_{H_2}$ confirms that the thermodynamic control on the type of VFA formed is significant and should be further elaborated. This finding corresponds with conclusions in previous publications [10, 11]. In contrast to these studies, however, it is argued here that the NAD$^+$ to NADH ratio should be considered as a key controller of the type of VFA produced and the associated amount of H$_2$ being formed available for methanogenesis, as also described in ref. [2]. The present theoretical effort, indicates that taking the NAD$^+$ to NADH ratio into account in dynamic rumen models is likely to improve prediction of type of VFA formed and CH$_4$ emissions.

It is concluded that fermentation of glucose to various VFA proceeds far from thermodynamic equilibrium and is not controlled by $P_{H_2}$ under rumen physiological conditions. However, oxidation of NADH does appear to be controlled by $P_{H_2}$, where the actual control also depends on the intracellular pH of microorganisms and the involvement of ferredoxin in NADH oxidation. The conversion of acetate to propionate is thermodynamic controlled by $P_{H_2}$ and also depends on the NAD$^+$ to NADH ratio. Conversions of butyrate to acetate and propionate to acetate are thermodynamically suppressed by $P_{H_2}$ and will not proceed without aggregation of rumen microbes. Rumen methanogenesis by archaea without cytochromes, which comprise most of the methanogenic population in the rumen, appears not to be thermodynamically restricted by $P_{H_2}$, implying the thermodynamic control of $P_{H_2}$ to be negligible.
Representation of the key role of the NAD\(^+\) to NADH ratio in rumen fermentation models is required to improve the accuracy of prediction of VFA and CH\(_4\) production by these models.

Supporting Information

S1 File. R code for calculating 95% confidence intervals of the thermodynamic potential factor (\(F_T\)) at discrete values of \(P_{H_2}\) and for plotting of \(F_T\) as a function of \(P_{H_2}\), including the 95% confidence intervals, for glucose fermentation and methanogenesis (Fig 1).

S2 File. R code for plotting of the thermodynamic potential factor (\(F_T\)) as a function of \(P_{H_2}\) for NADH oxidation (Fig 2).

S3 File. R code for calculating 95% confidence intervals of the thermodynamic potential factor (\(F_T\)) at discrete values of \(P_{H_2}\) and for plotting of \(F_T\) as a function of \(P_{H_2}\), including the 95% confidence intervals, for VFA interconversions (Fig 3).

Author Contributions

Conceptualization: HJvL JGF.
Data curation: HJvL.
Formal analysis: HJvL.
Funding acquisition: JD.
Investigation: HJvL CMP JGF.
Methodology: HJvL JGF.
Project administration: HJvL.
Resources: HJvL CMP JGF.
Software: HJvL JGF.
Supervision: JGF AB JD.
Validation: CMP JGF JD.
Visualization: HJvL JGF AB JD.
Writing – original draft: HJvL.
Writing – review & editing: CMP JGF EK AB JD.

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