INTRODUCTION

Paragonimus species are dispersed globally, but are most frequently found in Asia, Africa, and the Americas, where nine of about 50 nominal species are known to infect humans. In the USA, Paragonimus kellicotti is known to cause human illness, while Paragonimus mexicanus is accepted as a pathogenic species in Central and South American countries [1-3]. The metacercariae of P. mexicanus are unique in lacking a cyst wall, thus being very motile inside the body of crab hosts [4]. Recent research on the scanning electron microscope-based morphological and molecular features of Paragonimus metacercariae from Mexico has evidenced P. mexicanus as a species complex, and P. ecuadoriensis was recognized as possible cryptic/sibling species [5].

In Ecuador, P. ecuadoriensis was described as a new species based on the distinct morphology of its ovary and testis in relation to that of P. mexicanus, Paragonimus inca, and Paragonimus peruvianus [6]. However, further comparative studies revealed that P. ecuadoriensis is a junior synonym of P. mexicanus [4,7]. In Manabí Province, adult parasites obtained from experimentally infected animals showed similar morphological characteristics to P. mexicanus [3]. In Esmeraldas Province, bordering Manabi, isolated metacercariae were also identified as P. mexicanus by sequencing the second internal transcribed spacer (ITS2) region of ribosomal RNA gene [2].

Two siblings showing pulmonary symptoms such as productive coughs with bloody sputa were seen as outpatients in a hospital in Quito. They came from the Pedernales area in Manabí Province and had a history of consuming undercooked freshwater crabs captured in tributaries of the Cheve River. Accordingly, they were both diagnosed with pulmonary...
paragonimiasis and then successfully treated. The discovery of these cases prompted us to carry out an intensive field survey around the Cheve River to determine (1) the presence of active transmission of *Paragonimus* in the area, (2) the *Paragonimus* sp. involved, and (3) the identification of crab hosts and their respective infection rate, which are presented in this study.

**MATERIALS AND METHODS**

**Study site and crab collection**

Freshwater crabs, *Hypolobacera guayaquilensis*, were collected by local people in August 2011 in 2 tributary streams of the Cheve River flowing through the Pedernales area of Manabí Province, northwestern Pacific Coast of Ecuador (Fig. 1). This region, located between 120 and 500 m above sea level, and at latitude 0.138633° N and longitude 79.874267° W, is a seasonal green forest of the Equatorial Pacific Coast Range in Chongon-Colonche Mountains. Selected geographical areas are dedicated to agriculture and cattle ranching [8].

To collect crabs, an active search was performed in advance in the 2 tributaries of the Cheve River, which descends from the mountains. Shallow and slowly flowing streams with rocky and muddy areas are the typical habitats where crab burrows are found. Collected crabs were transferred to the laboratories in Quito and maintained alive in 20-L plastic containers with 2 L of river water at about 25°C (room temperature) until examined.

**Examination of crabs for *Paragonimus* metacercariae**

The crabs were sacrificed using thermal shock by immersing them in water at about 2°C for 10 min [9]. Then, the genitalia of each crab was examined for gender and species identification. Next, the maximum width of the carapace was measured using a caliper. After that, the carapace was removed from the posterior end of the body and only the hepatopancreas was removed as previously described [10]. The hepatopancreas was finally compressed between 2 glass plates (6×10×0.2 cm), which were examined under a dissecting microscope (CRX-32, Olympus, Tokyo, Japan). When *Paragonimus* metacercariae were detected, the glass plate was carefully removed to recover the metacercariae using teasing needles or tweezers. The isolated metacercariae were transferred into a Petri dish containing physiological saline solution using a capillary pipet. After taking photographs under a microscope (DP71, Olympus), *Paragonimus* metacercariae were preserved in 70% ethanol for molecular analysis.

**Molecular identification of metacercariae**

The molecular characterization of the metacercariae was performed by DNA isolation, amplification of the ITS2 region by PCR, and PCR-restriction fragment length polymorphism (PCR-RFLP); all processed material was sequenced as previously described [9]. Specifically, the primers used for PCR amplification and sequencing were 3S (forward, 5'-GGTACCGGTGGATCACTCGGCTCGTG-3' [11]) and A28 (reverse, 5'-GGGATCTGGTTAGTTTCTTTTCGCTGG-3' [12]). The

![Fig. 1. Map of the study area in the Pedernales area, Manabi Province, Ecuador. The Cheve River, collection site, and Cheve Medio community are indicated in the left panel. Cheve Medio is located ~20 km inland from nearby Pedernales on the coast of the Pacific Ocean.](image-url)
PCR amplification was conducted using 0.25 µM of each primer and 2.5 U of Phusion High-Fidelity DNA Polymerase (Thermo Fisher Scientific, Waltham, Massachusetts, USA). The PCR was carried out on a thermal cycler (TaKaRa PCR Thermal Cycler Dice Gradient, Takara Bio, Shiga, Japan), with 30 cycles of 98°C for 10 sec, 55°C for 10 sec, and 72°C for 15 sec. An initial denaturation and final extension were performed at 98°C for 30 sec and at 72°C for 7 min, respectively. For the PCR-RFLP, 10-µl portions of the amplified products were treated with 5 U of the restriction enzyme *Hinc* II (New England Biolabs, Ipswich, Massachusetts, USA) at 37°C for 1 hr. Then, the amplicons with or without enzymatic treatment were separated by electrophoresis on 2% (w/v) agarose gels. All amplified products were sequenced using the corresponding primers and the BigDye Terminator v3.1 Cycle Sequencing Kit (Thermo Fisher Scientific) on an automated sequencer (3730xl DNA Analyzer, Thermo Fisher Scientific). Sequences were aligned and compared using GENETYX-Win software (Ver. 13, Genetyx Co., Tokyo, Japan). Following the procedures described above, we also processed metacercariae of *P. kellicotti* isolated from crayfish collected in Missouri, USA [13], for comparison purposes.

Statistical analyses

A Poisson regression was performed to analyze the relationship between carapace size and the number of *Paragonimus* metacercariae per crab. We tested for a non-zero-slope term (the measure of effect) using a likelihood ratio test. Differences between gender and infection and between carapace size (quartiles) and infection were determined using a Chi-square test. All analyses were performed in R software (https://www.r-project.org/) considering $\alpha=0.05$.

### RESULTS

Of a total of 75 crabs examined, 50 were males (66.7%) and 25 were females (33.3%). All male crabs had gonopods with morphological features characteristic *H. guayaquilensis*. The average carapace size of the 75 crabs was 3.5 cm ($\pm 1.18$ cm, SD), ranging from 1.3 to 5.7 cm. Of the 75 crabs, 39 (52.0%) were infected with *Paragonimus* metacercariae. Of these crabs, 24 (32.0%) were males and 15 (20.0%) were females; no statistically significant relationship was detected between crab gender and metacercarial presence ($P=0.46$; Table 1).

All metacercariae were excysted, dorso-ventrally flattened, and very motile under the microscope. The excretory bladder was clearly observed in the midline of each metacecarial body, between the right and left intestines, which was yellow in color and connected to the oral sucker through the pharynx. These morphological features were in accordance with those of *P. mexicanus* (Fig. 2).

Using PCR amplification, ITS2 products of about 520 base pairs (bp) were generated from the DNA samples prepared from the excysted metacercariae (Fig. 3; lane 1). The sequence analysis of the PCR amplicons revealed that the amplified product was 461 bp (without primer sequences). The sequences obtained were identical to those previously obtained *P. mexicanus* metacercariae (DDBJ/EMBL/GenBank accession

### Table 1. Prevalence of *Paragonimus mexicanus* metacercariae according to crab gender and carapace size group

| Crab gender | Total (No.) | Positive (No.) | Negative (No.) | Chi$^2$ | Degrees of freedom | P-value |
|-------------|-------------|----------------|----------------|--------|--------------------|---------|
| Male        | 50 (66.7)   | 24 (32.0)      | 26 (34.7)      | 0.54   | 1                  | 0.46    |
| Female      | 25 (33.3)   | 15 (20.0)      | 10 (13.3)      |        |                    |         |
| Overall     | 75 (100)    | 39 (52.0)      | 36 (48.0)      |        |                    |         |

| Carapace size (cm) | No. of crabs | Positive (No.) | Negative (No.) | Chi$^2$ | Degrees of freedom | P-value |
|--------------------|--------------|----------------|----------------|--------|--------------------|---------|
| 1.3-2.9            | 19 (25.3)    | 5 (6.7)        | 14 (18.7)      | 8.96   | 3                  | 0.03*   |
| 3-3.5              | 20 (26.7)    | 10 (13.3)      | 10 (13.3)      |        |                    |         |
| 3.6-4.4            | 20 (26.7)    | 12 (16.0)      | 8 (10.7)       |        |                    |         |
| >4.4               | 16 (21.3)    | 12 (16.0)      | 4 (5.3)        |        |                    |         |
| Overall            | 75 (100)     | 39 (52.0)      | 36 (48.0)      |        |                    |         |

No difference in the infection rate was found between genders, but significant difference was found among carapace size groups.

*Significance = $P<0.05$. Ethical approval of the study was given by the Ethics Committee of the Universidad Central del Ecuador (License number LEC IORG 0001932, FWA 2482, IRB 2483.CObI-AM-PHI-0064-11).
number: AB308377) and eggs (AB308378) occurring in Ecuador [14]. Therefore, the nucleotide sequence obtained in this study has been deposited as the metacercarial stage of *P. mexicanus* in the DDBJ/EMBL/GenBank database under the accession number LC317061.

Species discrimination by RFLP of the PCR amplicons obtained from the excysted metacercariae of *Paragonimus mexicanus* (lane 1 and lane 2) and *Paragonimus kellicotti* (lane 3 and lane 4). The restriction endonuclease *Hinc* II used for RFLP digested the amplicons from *P. mexicanus* (lane 2, showing 2 fragments, 380 bp and 140 bp, arrow heads), but not the amplicons from *P. kellicotti* (lane 4, showing an undigested single fragment of 520 bp).

Products of about 520 bp were also generated from the DNA samples prepared from *P. kellicotti* metacercariae (Fig. 3; lane 3). However, these amplicons remained undigested by *Hinc* II (Fig. 3; lane 4). We found no intraspecific variation among the RFLP patterns for any of the amplicons (n = 5).

The percentage of infection within groups of crabs with similar carapace size was significantly different (P = 0.03; Table 1). A higher infection rate was observed in crabs with a carapace width > 3.6 cm, and the highest number of infected crabs was found within 3.6-4.4 cm carapace size groups. The largest number of metacercariae was found in a single male crab measuring 4.3 cm, which had 32 metacercariae. The smallest crab of our sample was a female measuring 1.6 cm and had 2 metacercariae, whereas the largest crab was a female, measuring 5.7 cm, in which only 1 metacercaria was found. We found a statistically significant effect of carapace size on the number of metacercariae under the Poisson regression model (intercept = -2.42, standard error = 0.34; slope = 0.86, standard error = 0.074, P < 0.001; Fig. 4).

**DISCUSSION**

The results of this study confirm the presence of the lung fluke *P. mexicanus* in the Pedernales area in Manabí Province, Ecuador. Furthermore, the 2 children who were diagnosed with pulmonary paragonimiasis by sputum examination and successfully treated with praziquantel in Quito were autochtho-
nous cases infected in the Cheve community. The high rate of metacercarial infection found in crabs (52.0%) represents a potential risk for residents and visitors for acquiring *P. mexicanus* infections because it is a common practice to eat raw or undercooked crabs in this region.

The crab, *H. guayaquilensis*, which has been reported from this study area [15] is reported here as a suitable second intermediate host for *P. mexicanus*, and this is a new host record. Previous studies have described 2 freshwater crab species in the same genus, namely *H. aequatorialis* and *H. chilensis* (formerly referred to as *Stregeria eigenmanni* and then *Pseudothelphusa chilensis*), as second intermediate hosts for *P. mexicanus* in Manabi Province [7]. Although, *P. ecuadoriensis* was described as the lung fluke species in Ecuador [6], it was later considered a junior synonym of *P. mexicanus* [4,7]. However, recent evidence has proposed that the individuality of *P. ecuadoriensis* should be reconsidered based on molecular and morphological evidence [5]. Further molecular studies are necessary to obtain a reliable consensus on the taxonomy of *P. mexicanus* and *P. ecuadoriensis* in Ecuador [5]. Based on the molecular methods used here (sequencing, and PCR-RFLP using *Hinc II*), we confirmed that the metacercariae examined belonged to *P. mexicanus*. The molecular region selected for PCR-RFLP did not produce an identifiable pattern in *P. kellicotti* from the USA and *P. caliensis* from Costa Rica [13,16], suggesting that PCR-RFLP using *Hinc II* can be used as a tool to discriminate *P. mexicanus* from these 2 other American lung fluke species, although *P. caliensis* specimens were unavailable for our analysis.

The infection rate of excysted metacercariae in crabs found here differs from that reported in previous studies performed in Manabi Province. For example, an infection rate of 100% was found in the crab *H. aequatorialis* in Llipijapa [6], whereas the infection rate was 16.1% in *H. chilensis* in Nalpe (Fig. 1) [10]. This difference in infection rates might be due to the different crab species acting as hosts. In addition, environmental factors, such as the deforestation that occurred over the years, might have changed the habitats of the reservoirs and intermediate hosts for *P. mexicanus*. Furthermore, the contamination of rivers with insecticides, fungicides, and other chemicals used by cattle ranches, coffee, cocoa, and African palm plantations, can directly affect the survival of the intermediate hosts, as mentioned in studies carried out in the neighboring Esmeraldas Province [10]. The high natural infection rate of 52% found in the present study might be related to the fact that primary forests in mountains and water sources of streams have remained uncontaminated. The livelihood of the reservoir animals including the first and second intermediate hosts (a mollusk, and crustacean, respectively) might not be threatened, and thus the life cycle of *P. mexicanus* is properly maintained in the area surrounding the Cheve River.

A statistical difference in infection rate was found between crab groups of different carapace sizes (Table 1). Furthermore, the likelihood ratio test under the Poisson regression model showed a significantly positive relationship between crab size and number of metacercariae (*P < 0.001*, Fig. 4). These data suggest that large crabs are heavily infected with *P. mexicanus* in the Cheve River. This relationship was first described in Esmeraldas Province [10]. Larger crabs are more valuable as food by people because they have more meat. If the local residents maintain the habit of eating raw or undercooked crabs, the probability of finding more individuals with paragonimiasis might increase.

The high natural infection rate of crabs with *Paragonimus* metacercariae, the abundance of crabs in the area, the ease of their capture, and the habit of local residents of consuming raw or undercooked crabs represent potential risk factors for the acquisition of paragonimiasis. The dissemination and familiarization of the results of the present study by the public health government agencies is important for implementing preventive measures or programs. Physicians and health personnel must consider the diagnosis of paragonimiasis in patients with pulmonary symptoms coming from this area, and thus make an early diagnosis and a timely treatment.

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**CONFLICT OF INTEREST**

We have no conflict of interest related to this work.

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