Adenosine kinase and adenosine receptors A₁R and A₂A R in temporal lobe epilepsy and hippocampal sclerosis and association with risk factors for SUDEP

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Abstract

Objective: The “adenosine hypothesis of SUDEP” (sudden unexpected death in epilepsy) predicts that a seizure-induced adenosine surge combined with impaired metabolic clearance can foster lethal apnea or cardiac arrest. Changes in adenosine receptor density and adenosine kinase (ADK) occur in surgical epilepsy patients. Our aim was to correlate the distribution of ADK and adenosine A₂AR and A₁R receptors (A₂AR and A₁R) in surgical tissue from patients with temporal lobe epilepsy and hippocampal sclerosis (TLE/HS) with SUDEP risk factors.

Methods: In 75 cases, patients were stratified into high-risk (n = 16), medium-risk (n = 11) and low-risk (n = 48) categories according to the frequency of generalized seizures before surgery. Using whole-slide scanning Definiens image analysis we quantified the labeling index (LI) for ADK, A₂AR, and A₁R in seven regions of interest: temporal cortex, temporal lobe white matter, CA1, CA4, dentate gyrus, subiculum, and amygdala and relative to glial and neuronal densities with glial fibrillary acidic protein (GFAP) and neuronal nuclear antigen (NeuN).

Results: A₁R showed predominant neuronal, A₂AR astroglial, and ADK nuclear labeling in all regions but with significant variation. Compared with the low-risk group, the high-risk group had significantly lower A₂AR LI in the temporal cortex. In HS cases with severe neuronal cell loss and gliosis predominantly in the CA1 and CA4 regions, significantly higher A₁R was present in the amygdala in high-risk than in low-risk cases. There was no significant difference in neuronal loss or gliosis between the risk groups or differences for ADK labeling.

Significance: Reduced cortical A₂AR suggests glial dysfunction and impaired adenosine modulation in response to seizures in patients at higher risk for SUDEP. Increased neuronal A₁R in the high-risk group could contribute to periictal amygdala dysfunction in SUDEP.
1 | INTRODUCTION

Adenosine has varied physiological functions in the central nervous system (CNS) including endogenous anticonvulsant actions. Although not a conventional neurotransmitter, its binding with high affinity G protein–coupled presynaptic and postsynaptic receptors (A₁R and A₂AR) modulates synapses with predominantly inhibitory and neuroprotective effects. Adenosine levels rise before seizure termination; activation of hyperpolarizing presynaptic A₁R suppresses seizures and their spread, whereas A₁R-selective antagonists reverse this effect. By contrast, A₂AR neuronal activation has mainly proconvulsive actions.

Alteration in adenosine receptors densities and their cellular distribution has been shown in animal models of epilepsy and human tissues. An imbalance between the inhibitory and excitatory effects of adenosine through differential activation of A₁R and A₂AR receptors may contribute to temporal lobe epilepsy (TLE). In addition to seizure termination, postictal depression, refractoriness, and respiratory depression are considered manifestations of overactivation of A₁R coupled with underactivation of A₂AR. This has led to the “adenosine hypothesis of SUDEP” (sudden unexpected death in epilepsy) as common clinical phenomena, respiratory depression, and impaired arousal could be mediated through overstimulation of these receptors in autonomic and brainstem centers in the postictal period.

Experimental evidence also supports the adenosine SUDEP hypothesis; in a kainic acid model, with pharmacologically impaired adenosine clearance, induced seizures triggered sudden death while the adenosine receptor antagonist caffeine administered after seizure onset was associated with increased survival times. Similar effects were observed in seizure-induced death in DBA/2 mice, where administration of an adenosine antagonist protected from seizure-induced respiratory arrest; in genetically epilepsy-prone rats inhibitors of adenosine metabolism, were associated with increased duration of postictal respiratory depression.

Adenosine receptors have not been assessed in human tissues in relation to SUDEP. We used a large TLE surgical series, risk stratified for SUDEP similar to prior magnetic resonance imaging (MRI) studies based on the greatest risk factor of three or more generalized seizures (GS) occurring in the last year prior to surgery. We evaluated several regions, including the hippocampus, amygdala, and temporal cortex, to explore the differential expression of adenosine receptors and its major regulator, ADK, in relation to regional neuronal and glial populations.

2 | METHODS

2.1 | Case selection

Cases were selected from the UCL Epilepsy Society brain and tissue bank from adult patients who had undergone temporal lobectomy between 2009 and 2017 with a clinical and radiological diagnosis of suspected hippocampal sclerosis (HS). The project has ethical approval and patients provided written informed consent for participation in research. In 75 cases, tissue samples were available of the temporal lobe, hippocampal body, and amygdala. The neuropathology diagnosis was reviewed and the subtype of HS revised according to International League Against Epilepsy (ILAE) criteria.

2.2 | SUDEP risk stratification

Clinical records were reviewed to extract data regarding seizure history, including age at onset, seizure type, and recent frequency (Table 1). Risk stratification for SUDEP was based on the number of GS in the previous 12 months at the last preoperative assessment: Patients with ≥3 GS were high risk, no GS were low risk, and 1-2 GS were intermediate risk.

2.3 | Tissue preparation and immunohistochemistry

From each case, three tissue samples from the temporal lobe (taken approximately 2.5 cm posterior to the temporal...
pole), the hippocampal body, and amygdala were selected. Serial sections were cut at 5 μm thickness and an immunohistochemistry staining panel carried out using standard techniques: A1R (ab124780, Abcam monoclonal, dilution 1:200), A2AR (ab3461, Abcam polyclonal, dilution 1:1000), adenosine kinase (ADK) (LSB6272, Lifescience bio monoclonal dilution 1:500), glial fibrillary acidic protein (GFAP) (Z0334, DAKO polyclonal, 1:2500), and neuronal nuclear antigen (NeuN) (Millipore, 1:2000) using standard methods (detailed in Supplementary Files).

Additional double-labeling immunofluorescence was carried out on select cases representing each risk group for A1R, ADK, and A2AR combined with GFAP and neuronal markers NeuN or MAP2 using standard methods (see Supplementary Files) and visualized with Zeiss Axio Imager Z2 fluorescent microscope (Zeiss).

The slides were scanned at 40× magnification using a Leica SCN400 (Leica Microsystems) and Zeiss AxioScan Zen 2.3 slide scanners. Seven regions of interest (ROIs) were defined using Definiens Tissue Studio (Definiens AG) image analysis: ROI 1 Temporal cortex to include the entire cortical thickness from the sulcus, adjacent to the superior temporal gyrus, to both gyral crowns, ROI 2 Temporal cortex white matter region of subcortical white matter underlying ROI 1 (Figure S1A), ROI 3 dentate gyrus to include the granule cell layer and molecular layer, ROI 4 CA4 subfield of hippocampus to include the region between the blades of the dentate gyrus, ROI 5 CA1 subfield of hippocampus to include the maximal representation of pyramidal cell layer but excluding CA2, ROI 6 Subiculum to include its maximal representation on section (Figure S1B), ROI 7 Amygdala regions outlined with cytoarchitecture compatible with amygdala as identified on NeuN (Figure S1C); due to the fragmented nature of amygdala samples, it was not possible to delineate subnuclei. With Definiens Tissue Studio software, the defined seven ROIs were transposed onto the serial immunostained sections to ensure similar ROI capture across each sample; each individual section was manually edited for any slight misalignment of ROI. An intensity threshold was set for the chromogen detection for each antibody, which was applied across all cases (Figure S1D,E) and the total labeling index (LI) (percentage area of cellular immunostaining with high-intensity threshold) for each ROI was measured.

Statistical analysis was carried out with SPSS (version 22, IBM) for nonparametric tests to compare data between cause of groups (Kruskal-Wallis and Mann-Whitney tests), regions within groups (Friedman test), and clinical correlations (Spearman’s rho test); P values of <.05 were regarded as significant and corrected for multiple comparisons as detailed in results. For graphical representation of data, GraphPad Prism 7 (University of California, San Diego) was used.

3 | RESULTS

Our 75 cases were classified for SUDEP risk as high (16), intermediate (11), and low (48). The three risk groups had

| SUDEP | Hippocampal | Gender, | Side L: | Age at onset | Focal | Nocturnal |
|-------|-------------|---------|---------|--------------|-------|-----------|
| risk group, | sclerosis | F: M (%) | R (%) | Age at surgery | seizures with LOA (weekly seizures) (% of cases) | Sz, Episodes (% of cases) |
| N = number | subtypes | | | Duration (years, (SD)) | | |
| High risk (HR), | Type 1 62.5% | 56.3:43.8 | 62.5:37.5 | 13.7 (11.3) | 100% | 100% (73.3%) |
| N = 16 | Type 2 18.8% | | | 34.9 (9.4) | | 31.3% |
| | Other 18.8% | | | 21.1 (12) | | 20% |
| Intermediate risk (IR), | Type 1 82% | 63.6:36.4 | 63.6:36.4 | 12.7 (7.1) | 100% | 100% (55.5%) |
| N = 11 | Type 2 9% | | | 40 (9) | | 36.4% |
| | Other 9% | | | 26.7 (11.7) | | 36.4% |
| Low risk (LR), | Type 1 75% | 50:50 | 58.3:41.7 | 13.5 (11.8) | 62.5% | 80.6% |
| N = 48 | Type 2 8.3% | | | 42.2 (11.1) | | (63%) |
| | Other 16.7% | | | 28.7 (14.8) | | 8.3% |

Significance between Groups: ~

Note: ~Kruskal-Wallis test was used to compare groups.

Abbreviations: GS, generalized seizures; IPI, initial precipitating injury; LOA, loss of awareness; SE, status epilepticus; Sz, seizures.

*The frequency of GS in last 12 months was used as the criterion to stratify groups. Hippocampal sclerosis subtypes other = type 3, no HS and cases with uncertain HS types (incomplete subfields represented in the resected specimen).
no significant differences in the ILAE HS subtypes (type 1 comprised 62.5% to 82%), gender, age at onset, age at surgery, frequency of focal seizures with impaired awareness, or history of initial precipitating injury (IPI). There was a lower frequency of nocturnal seizures ($P = .02$) and episodes of status epilepticus ($P = .002$) in low-risk compared with intermediate and high-risk groups (Table 1).

### 3.1 Distribution of $A_1R$, $A_2AR$, and ADK in temporal lobe regions

**$A_1R$. Temporal neocortex:** Pancortical neuronal cytoplasmic labeling was observed with $A_1R$ and membranous or nuclear labeling was not observed (Figure 1A-D). Reduced intensity of superficial compared with deep cortical neurons was seen in some cases, particularly in cases with superficial neuronal loss (Figure 1B, C). Scattered neurons in layer I were also $A_1R$ positive (Figure 1A) and in the interstitial white matter (Figure 1E); scattered rare glial cells showed weak labeling with $A_1R$ (Figure 1E, arrow). **Hippocampus:** The granule cells of the dentate gyrus showed variable $A_1R$ labeling, particularly in the basal aspect (Figure 1F,G); labeling appeared less intense in dispersed granule cells (Figure 1E). Cytoplasmic labeling of residual CA4 and labeling appeared less intense in dispersed granule cells particularly in the basal aspect (Figure 1F,G); labeling appeared less intense in dispersed granule cells (Figure 1E). Cytoplasmic labeling of residual CA4 and CA1 pyramidal neurons was confirmed with double labeling for MAP2 and NeuN (Figure 1E, inset) and labeling of horizontal interneurons in CA1 and subiculum pyramidal neurons but no labeling of astroglia (Figure 1H, inset). **Amygdala:** Intense neuronal cytoplasmic $A_1R$ (Figure 1F) and occasional labeling of glial cells was noted in the periamygdala white matter but mainly little coexpression noted with GFAP.

**$A_2AR$. Temporal neocortex:** A stereotypical labeling pattern between cases was seen with $A_2AR$ labeling of astroglial cell types in the subpial layer and cortical layer I (Figure 2A), with white matter astrocytes in a perivascular distribution and subcortical location in addition to endothelial labeling (Figure 2B). No neuronal labeling was evident. **Hippocampus:** There was overall marked increase in intensity of $A_2AR$ labeling, particularly in the sclerotic hippocampus (Figure 2C) compared with the temporal lobe, predominantly in astroglial cell types and some endothelial (Figure 2E). The dentate granule cell layer and subgranular zone showed striking labeling with dense meshworks of glial cells and processes (Figure 2C,D) confirmed with double labeling for GFAP (Figure 2D); in some cases $A_2AR$ expression dominated in basal cell layer astroglia. CA1 showed marked labeling of astroglial cells (Figure 2C), and $A_2AR$ labeling was also seen in the hippocampal fissure and streams of glial cells in the white matter. In the few cases with no sclerosis, there was less intense $A_2AR$ labeling of glia in CA4, CA1, and the dentate gyrus; also reduced labeling for $A_2AR$ in the dentate gyrus was apparent in type 2 compared to type 1 HS. **Amygdala:** Infrequent $A_2AR$ glial cells and processes were scattered with some labeling of capillary endothelium but no neuronal labeling (Figure 2F).

**ADK:** All regions showed mainly nuclear labeling of small glial cells and occasional neurons with some cytoplasmic labeling, confirmed with co-localization with NeuN (Figure 2G). The superficial temporal cortex showed more prominent labeling compared with the deeper cortex, and nuclei were also prominent in the granule cell layer.

### 3.2 Quantitative analysis

#### 3.2.1 Regional differences in expression

The highest LI in all three risk groups for $A_1R$ was in the dentate gyrus, with significant variation across the seven ROIs ($P < .0001$) (Table 2). For $A_2AR$, the highest LI in the high- and intermediate-risk groups was in the dentate gyrus but for the low-risk group in CA1 (Table 2); significant variation across ROIs was seen for the low- and high-risk groups only ($P < .0001$). For ADK, the highest LI was in CA1 for the high- and low-risk groups and in temporal cortex for the intermediate-risk group (Table 2); significant variation between ROIs was present in the low-risk group only ($P < .0001$).

#### 3.2.2 Comparison of high- and low-risk SUEDP groups

There was no significant difference in $A_1R$ or ADK LI for any region between the three risk groups. $A_2AR$ LI was lower in high- than low-risk group for all regions except the amygdala, reaching significance in the temporal cortex ($P = .035$, corrected for multiple comparisons, Figure 3A).

When considering type 1 HS cases only, $A_1R$ in the amygdala was significantly different across the risk groups ($P = .034$, corrected for multiple comparisons), with mean values higher in both medium- and high-risk group compared with the low-risk group, reaching significance for the medium-risk group ($P = .006$, corrected for multiple comparison, Figure 3B). There was also significantly lower $A_2AR$ in the temporal cortex in the high- than in low-risk groups ($P = .032$, corrected for multiple comparisons, Figure 3A) but no differences were noted for ADK.

Because $A_1R$ is expressed mainly in neurons and $A_2AR$ in astroglia, we stained for NeuN and GFAP to evaluate differences in neuronal loss or gliosis between regions and risk groups. There was significant variation across ROIs for both GFAP and NeuN LI ($P < .0001$), but there was no
significant difference in NeuN or GFAP LI for any ROIs between the three risk groups; this was also the case when considering type 1 HS cases alone. Across all cases, there was a positive correlation with NeuN and $A_1R$ LI in all ROIs ($P < .01$ to $P < .000$) apart from the dentate gyrus and CA1. There was a positive correlation with GFAP LI and $A_2A_R$ for all hippocampal and amygdala ROIs ($P < .05$ to .001) but not for temporal cortex ROIs. There were no significant correlations between ADK and GFAP labeling in any region.

### 3.2.3 Clinical correlations

There was a negative correlation between duration of epilepsy and $A_1R$ LI (ROI1, $P = .04$ and ROI2, $P = .03$) and
ADK LI (ROI1, \(P = .03\)) but a positive correlation with \(A_2\alpha R\) LI (ROI2, \(P = .04\)) in temporal lobe ROIs. In patients with a history of nocturnal seizures, lower \(A_2\alpha R\) labeling occurred in CA4 (\(P = .02\)), CA1 (\(P = .001\)), and amygdala ROIs (\(P = .019\)) (Figure 3C); it was also significantly lower in CA1 in patients with a history of status epilepticus compared with those without this history (\(P = .02\)).

4 | DISCUSSION

In our TLE patients, we identified dominant neuronal expression of adenosine \(A_1 R\), expression of \(A_2\alpha R\) in regions of astrocytosis, and predominant nuclear ADK expression. In patients at high risk for SUDEP, we observed a significant reduction in \(A_2\alpha R\) in the temporal cortex and increased amygdala \(A_1 R\); this could implicate regional cellular imbalances in adenosine signaling as a vulnerability factor for SUDEP. The role of the purine nucleotide adenosine in seizure modulation is supported by extensive animal and human studies. Experimental evidence suggests that adenosine may precipitate or contribute to the progression from seizure to SUDEP postictally, although this has not been investigated using human tissues.

4.1 | \(A_2\alpha R\), ADK, and astrocytosis in epilepsy

Epilepsy is associated with altered cellular distribution of adenosine receptors and ADK that may occur as primary or secondary adaptive phenomena. Earlier studies utilizing human tissues found enhanced neuronal expression of \(A_1 R\) and upregulation of \(A_2\alpha R\) in Rasmussen's encephalitis, with \(A_2\alpha R\) expression in lesional astrocytes, endothelium, as
In TLE, A2AR expression was primarily co-localized with hippocampal GFAP-positive astrocytes but not neuronal markers, showing increased receptor expression compared with postmortem controls. Astrocytic A2AR regulates cellular Na+/K+ ATPase, glutamate uptake and inflammatory cytokine release, and receptor upregulation was observed following varied brain insults. In our study, A2AR expression was predominantly astroglial, supported by the correlation between A2AR and GFAP staining in mesial temporal regions; there was also a positive correlation with duration of epilepsy in the temporal lobe regions. The relative reduction of A2AR in the temporal cortex in the high SUDEP risk group, despite no quantitative differences in gliosis between the risk groups, suggests impaired adaptation and adenosine modulation as well as glial dysfunction in response to seizures as a vulnerability factor for SUDEP.

Our study was limited to the temporal cortex and it would be important to extend a similar study to other cortical regions, for example, the insular and cingulate cortex, that have been structurally and functionally implicated in recent clinical and neuroimaging studies in SUDEP. Furthermore, A2AR receptors have high density on normal cerebral microvessels in contrast to A1R, and A2AR modulates blood-brain barrier permeability. Our previous postmortem study found no evidence for acute blood-brain barrier insufficiency in SUDEP postmortem brains. Nevertheless, in the present series, intense endothelial A2AR expression was observed in some cases, although this was not separately quantified; it is of potential relevance to the significant microvascular dysfunction in focal epilepsy and warrants further investigation.

Astrocytes regulate the adenosine cycle during and after seizures: Following high levels of neuronal activity, cytoplasmic ATP is released through exocytosis or hemi-channels ("gliotransmission"); then following degradation, glial reuptake of adenosine occurs via nucleotide transporters with phosphorylation by cytoplasmic ADK. Astroglial ADK is the major metabolic clearance route for adenosine and represents a candidate treatment target in epilepsy. Glial proliferation and astrocytosis occurs in most surgical epilepsy specimens with myriad pathophysiological contributions, such as recycling neuromodulators. ADK upregulation was reported in the balloon cells of focal cortical dysplasia, reactive astrocytes in Rasmussen’s encephalitis, and in TLE. In contrast to these reports, we did not demonstrate significant cytoplasmic glial ADK in our series or a correlation with gliosis as measured by GFAP labeling. This discrepancy may be explained by our antibody, which was selected for its optimal labeling in temporal lobe fixed-tissue samples but has preferential recognition of the long isoform of ADK. ADK isoforms are developmentally regulated in the brain; the short isoform of ADK preferentially locates to the cytoplasm for adenosine clearance, whereas the long isoform is nuclear, with mainly epigenetic regulatory roles.

| Table 2 | Labeling index (LI) and standard deviation (SD) for A1R, A2AR, and ADK in seven regions of interest in the temporal lobe in the three SUDEP risk groups |
|---------|---------------------------------------------------------------|
| Group/Region | ROI1/Cortex | A1R LI | A2AR LI | ADK LI × 10−2 (SD) |
| Low risk, N = 48 | 0.039 (0.02) | 0.005 (0.01) | 0.1 (0.1) |
| Intermediate risk, N = 11 | 0.044 (0.03) | 0.08 (0.09) | 0.18 (0.24) |
| High risk, N = 16 | 0.043 (0.02) | 0.002 (0.003) | 0.025 (0.004) |

Note: Significant differences between risk groups shown in bold.
4.2 | A1R expression and the amygdala

High A1R binding densities occur in CA1,\(^36\) intermediate to high concentrations in the amygdala, and lower levels in brainstem.\(^37\) In the cortex, A1R neuronal labeling was more intense in layer V than superficial cortex,\(^38\) consistent with our observations. In quantitative studies of TLE tissues, A1R labeling in the hippocampus and dentate gyrus was reduced compared with nonepilepsy controls,\(^39\) whereas another study of the amygdala found similar A1R-binding densities in TLE patients and controls.\(^40\) Here, we observed increased A1R in the amygdala in the high-risk group when we stratified cases into type I HS alone. The extent of amygdala sclerosis associated with HS is a relevant variable,\(^41\) but we did not observe any difference in NeuN labeling in any region, as a measure of neuronal number, between the risk groups. In addition, we found lower A2A R labeling in amygdala and mesial temporal regions in patients with nocturnal seizures, a risk factor for SUDEP,\(^42\) together this supports amygdala A1R/A2A R imbalance in cases with higher SUDEP risk.

There is accumulating clinical evidence for amygdala involvement in SUDEP; MRI studies revealed increased rightsided gray matter volumes\(^16\) and clinical studies record apnea following electrical stimulation of the amygdala.\(^43-45\) Of interest, tolerance to hypoxia is mediated through A1R,\(^46\) and patients with apnea undergoing amygdala stimulation had no symptoms of dyspnea,\(^45\) which could be an adenosine-mediated effect. Furthermore, evidence supports brainstem spreading depolarization as a final postictal event in SUDEP\(^47;\) in the CACNA1A model of SUDEP, spreading depolarization extended through subcortical regions and the amygdala before brainstem extension.\(^48\) There is evidence that adenosine may be responsible for prolonged depression of synaptic transmission after spreading depolarization via A1R receptor activation.\(^49\) Of note, the neuronal labeling we observed included cytoplasmic labeling and is likely also detecting internalized A1R nonfunctioning receptors, in addition to functional membrane receptors. A1R

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**FIGURE 3** Graphs of regional A1R and A2A R labeling in relation to SUDEP risk, hippocampal sclerosis and nocturnal seizures. A, Scatter graph of A2A R labeling index shown in high and low risk groups, in all cases and with type I HS only, both showing significantly lower A2A R in the temporal cortex (ROI1) in high risk group. B, Scatter graph of A1R labeling index in the amygdala (ROI7) which was higher in high risk and medium risk compared to low SUDEP risk group. C, A2A R regional labeling in cases with nocturnal seizures showed lower labeling in hippocampal and amygdala ROI; *significant in CA4 (P = .02), CA1 (P = .001) and amygdala ROI (P = .019).

Abbreviations: AMYG, amygdala; DG, dentate gyrus; H, high; HS, hippocampal sclerosis; L, Low; M, medium; TCX, temporal cortex; ROI, region of interest; SUBIC, subiculum; WM, white matter
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CONFLICT OF INTEREST

The authors have no conflicts of interest to declare. We confirm that we have read the Journal’s position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

REFERENCES

1. Canas PM, Porciuncula LO, Simoes AP, Augusto E, Silva HB, Machado NJ, et al. Neuronal adenosine A2A receptors are critical mediators of neurodegeneration triggered by convulsions. eNeuro. 2018;5:6.
2. Welth L, Reemmer J, Boison D. The role of adenosine in epilepsy. Brain Res Bull. 2019;151:46–54.
3. Van Gompel JJ, Bower MR, Worrell GA, Stead M, Chang SY, Goerss SJ, et al. Increased cortical extracellular adenosine correlates with seizure termination. Epilepsia. 2014;55:233–44.
4. Barros-Barbosa AR, Ferreirinha F, Oliveira A, Mendes M, Lobo MG, Santos A, et al. Adenosine A2A receptor and ecto-5’-nucleotidase/CD73 are upregulated in hippocampal astrocytes of human patients with mesial temporal lobe epilepsy (MTLE). Purinergic Signal. 2016;12:719–34.
5. He X, Chen F, Zhang Y, Gao Q, Guan Y, Wang J, et al. Upregulation of adenosine A2A receptor and downregulation of GLT1 is associated with neuronal cell death in Rasmussen’s encephalitis. Brain Pathol. 2020;30(2):246–60.
6. Luan G, Gao Q, Zhai F, Zhou J, Liu C, Chen Y, et al. Adenosine kinase expression in cortical dysplasia with balloon cells: analysis of developmental lineage of cell types. J Neuropathol Exp Neurol. 2015;74:132–47.
7. Devinsky O, Hesdorffer DC, Thurman DJ, Lhatoo S, Richerson G. Sudden unexpected death in epilepsy: epidemiology, mechanisms, and prevention. Lancet Neurol. 2016;15:1075–88.
8. Erlichman JS, Leiter JC, Gourine AV. ATP, glia and central respiratory control. Respir Physiol Neurobiol. 2010;31(173):305–11.
9. Boison D. Adenosine dysfunction in epilepsy. Glia. 2012;60:1234–43.
10. Shen HY, Li T, Boison D. A novel mouse model for sudden unexpected death in epilepsy (SUDEP): role of impaired adenosine clearance. Epilepsia. 2010;51:465–8.
11. Zhang J, Yin D, Wu F, Zhang G, Jiang C, Li Z, et al. Microinjection of adenosine into the hypothalamic ventrolateral preoptic area enhances wakefulness via the A1 receptor in rats. Neurochem Res. 2013;38:1616–23.
12. Scislo TJ, O’Leary DS. Purinergic mechanisms of the nucleus of the solitary tract and neural cardiovascular control. Neurol Res. 2005;27:182–94.
13. Faingold CL, Randall M, Komnajosyula SP. Susceptibility to seizure-induced sudden death in DBA/2 mice is altered by adenosine. Epilepsy Res. 2016;124:49–54.
14. Komnajosyula SP, Randall ME, Faingold CL. Inhibition of adenosine metabolism induces changes in post-ictal depression, respiration, and mortality in genetically epilepsy prone rats. Epilepsy Res. 2016;119:13–9.
15. Allen LA, Harper RM, Kumar R, Guye M, Ogren JA, Lhatoo SD, et al. Dysfunctional brain networking among autonomic regulatory structures in temporal lobe epilepsy patients at high risk of sudden unexpected death in epilepsy. Front Neurol. 2017;8:544.

16. Wandschneider B, Koepp M, Scott C, Micallef C, Balestrini S, Sisodiya SM, et al. Structural imaging biomarkers of sudden unexpected death in epilepsy.Brain. 2015;138:2907–19.

17. DeGiorgio CM, Curtis A, Hertling D, Moseley BD. Sudden unexpected death in epilepsy: Risk factors, biomarkers, and prevention. Acta Neurol Scand. 2019;139:220–30.

18. Blumcke I, Thom M, Aronica E, Armstrong DD, Bartolomei F, Bernasconi A, et al. International consensus classification of hippocampal sclerosis in temporal lobe epilepsy: A Task Force report from the ILAE Commission on Diagnostic Methods. Epilepsia. 2013;54:1315–29.

19. Cieslak M, Wojtczak A, Komoszynski M. Role of the purinergic signaling in epilepsy. Pharmacol Rep. 2017;69:130–8.

20. Richerson GB, Boison D, Faingold CL, Ryvlin P. From unwitnessed fatality to witnessed rescue: Pharmacologic intervention in sudden unexpected death in epilepsy. Epilepsia. 2016;57(Suppl 1):35–45.

21. Luan G, Wang X, Gao Q, Guan Y, Wang J, Deng J, et al. Uptregulation of neuronal adenosine A1 receptor in human rasmussen encephalitis. J Neuropathol Exp Neurol. 2017;1(76):720–31.

22. Cunha RA. How does adenosine control neuronal dysfunction and neurodegeneration? J Neurochem. 2016;139:1019–55.

23. Lacuey N, Hampson JP, Theeranaew W, Zonjy B, Vithala A, Hupp NJ, et al. Cortical structures associated with human blood pressure control. JAMA Neurol. 2018;1(75):194–202.

24. Lacuey N, Zonjy B, Theerannaew W, Loparo KA, Tatsuoka C, Sahadevan J, et al. Left-insular damage, autonomic instability, and sudden unexpected death in epilepsy. Epilepsy Behav. 2016;55:170–3.

25. Ogren JA, Tripathi R, Macey PM, Kumar R, Stern JM, Elishash DS, et al. Regional cortical thickness changes accompanying generalized tonic-clonic seizures. Neuroimage Clin. 2018;20:205–15.

26. Allen LA, Harper RM, Lhatoo S, Lemieux L, Diehl B. Neuroimaging of Sudden Unexpected Death in Epilepsy (SUDEP): insights from structural and resting-state functional MRI studies. Front Neurol. 2019;10:185.

27. Kalaria RN, Harik SI. Adenosine receptors and the nucleoside transporter in human brain vasculature J Cereb Blood Flow Metab. 1988;8:32–9.

28. Bynoe MS, Viret C, Yan A, Kim DG. Adenosine receptor signaling: a key to opening the blood-brain door. Fluids Barriers CNS. 2015;2(12):20.

29. Michalak Z, Obari D, Ellis M, Thom M, Sisodiya SM. Neuropathology of SUDEP: Role of inflammation, blood-brain barrier impairment, and hypoxia. Neurology. 2017;7(88):551–61.

30. Gorter JA, Aronica E, van Vliet EA. The roof is leaking and a storm is raging: repairing the blood-brain barrier in the fight against epilepsy. Epilepsy Curr. 2019;19:177–81.

31. Beamer E, Conte G, Engel T. ATP release during seizures - a critical evaluation of the evidence. Brain Res Bull. 2019;151:65–73.

32. Boison D, Steinhauser C. Epilepsy and astrocyte energy metabolism. Glia. 2018;66:1235–43.

33. Steinhauser C, Grunnet M, Carmignoto G. Crucial role of astrocytes in temporal lobe epilepsy. Neuroscience. 2016;323:157–69.

34. Aronica E, Zurolo E, Iyer A, de Groot M, Anink J, Carbonell C, et al. Uptregulation of adenosine kinase in astrocytes in experimental and human temporal lobe epilepsy. Epilepsia. 2011;52:1645–55.

35. Kiese K, Jablonski J, Boison D, Kobow K. Dynamic regulation of the adenosine kinase gene during early postnatal brain development and maturation. Front Mol Neurosci. 2016;9:99.

36. Svenningsson P, Hall H, Sedvall G, Fredholm BB. Distribution of adenosine receptors in the postmortem human brain: an extended autoradiographic study. Synapse. 1997;27:322–35.

37. Fastbom J, Pazos A, Probst A, Palacios JM. Adenosine A1 receptors in the human brain: a quantitative autoradiographic study. Neuroscience. 1987;22:827–39.

38. Schindler M, Harris CA, Hayes B, Papotti M, Humphrey PP. Immunohistochemical localization of adenosine A1 receptors in human brain regions. Neurosci Lett. 2001;297(3):211–5.

39. Glass M, Faull RL, Walker EB, et al. Loss of A1 adenosine receptors in human temporal lobe epilepsy. Brain Res. 1996;671:56–68.

40. Graebenitz S, Kedo O, Speckmann EJ, Gorgi A, Panneck H, Hans V, et al. Intercital-like network activity and receptor expression in the epileptic human lateral amygdala. Brain. 2011;134:2929–47.

41. Nakayama Y, Masuda H, Shirozu H, Ito Y, Higashijima T, Kitaura H, et al. Features of amygdala in patients with mesial temporal lobe epilepsy and hippocampal sclerosis: an MRI volumetric and histopathological study. Epilepsy Res. 2017;135:50–5.

42. Latreille V, Abdennadher M, Dworetzky BA, Ramel J, White D, Katz E, et al. Nocturnal seizures are associated with more severe hypoxemia and increased risk of postictal generalized EEG suppression. Epilepsia. 2017;58:e127–e131.

43. Nobis WP, Schuele S, Temppler JW, Zhou G, Lane G, Rosenow JM, et al. Amygdala-stimulation-induced apnea is attention and nasal-breathing dependent. Ann Neurol. 2018;83:460–71.

44. Lacuey N, Zonjy B, Loparo KA, Tatsuoka C, Sahadevan J, et al. Left-insular damage, autonomic instability, and sudden unexpected death in epilepsy. Epilepsia. 2016;55:170–3.

45. Allen LA, Harper RM, Lhatoo S, Lemieux L, Diehl B. Neuroimaging of Sudden Unexpected Death in Epilepsy (SUDEP): insights from structural and resting-state functional MRI studies. Front Neurol. 2019;10:185.

46. Allen LA, Harper RM, Lhatoo S, Lemieux L, Diehl B. Neuroimaging of Sudden Unexpected Death in Epilepsy (SUDEP): insights from structural and resting-state functional MRI studies. Front Neurol. 2019;10:185.

47. Allen LA, Harper RM, Lhatoo S, Lemieux L, Diehl B. Neuroimaging of Sudden Unexpected Death in Epilepsy (SUDEP): insights from structural and resting-state functional MRI studies. Front Neurol. 2019;10:185.

48. Allen LA, Harper RM, Lhatoo S, Lemieux L, Diehl B. Neuroimaging of Sudden Unexpected Death in Epilepsy (SUDEP): insights from structural and resting-state functional MRI studies. Front Neurol. 2019;10:185.
after spreading depolarization in brain slices. Neuroscience. 2012;223:365–76.

50. Baines AE, Correa SA, Irving AJ, Frenguelli BG. Differential trafficking of adenosine receptors in hippocampal neurons monitored using GFP- and super-eclipic pHluorin-tagged receptors. Neuropharmacology. 2011;61:1–11.

SUPPORTING INFORMATION
Additional supporting information may be found online in the Supporting Information section.

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