Promising Approach in the Treatment of Glaucoma Using Nanotechnology and Nanomedicine-Based Systems

Fidinaina Rina Juliana 1, Samuel Kesse 2,3✉, Kofi Oti Boakye-Yiadom 2,3, Hanitrarimalala Veroniaina 4, Huihui Wang 1 and Meihao Sun 1,*

1 College of Chemistry and Life Sciences, Zhejiang Normal University, Jinhua 321004, China; younitra@gmail.com (F.R.J.); sanket.acharya4@gmail.com (H.W.)
2 School of Pharmacy, Shanghai Jiao Tong University, 800 Dongchuan Road, Shanghai 200240, China; kessejr@yahoo.com (S.K.); otiboakye1000@gmail.com (K.O.B.-Y.)
3 Department of Pharmaceutics, School of Pharmacy, China Pharmaceutical University, Nanjing 211198, China
4 State Key Laboratory of Modern Chinese Medicine, China Pharmaceutical University, Nanjing 210009, China; scholarha@yahoo.com
* Correspondence: mhsun@zjnu.cn

Academic Editors: Luigi Paduano and Paola Luciani
Received: 20 August 2019; Accepted: 19 October 2019; Published: 22 October 2019

Abstract: Glaucoma is considered a leading cause of blindness with the human eye being one of the body’s most delicate organs. Ocular diseases encompass diverse diseases affecting the anterior and posterior ocular sections, respectively. The human eye’s peculiar and exclusive anatomy and physiology continue to pose a significant obstacle to researchers and pharmacologists in the provision of efficient drug delivery. Though several traditional invasive and noninvasive eye therapies exist, including implants, eye drops, and injections, there are still significant complications that arise which may either be their low bioavailability or the grave ocular adverse effects experienced thereafter. On the other hand, new nanoscience technology and nanotechnology serve as a novel approach in ocular disease treatment. In order to interact specifically with ocular tissues and overcome ocular challenges, numerous active molecules have been modified to react with nanocarriers. In the general population of glaucoma patients, disease growth and advancement cannot be contained by decreasing intraocular pressure (IOP), hence a spiking in future research for novel drug delivery systems and target therapeutics. This review focuses on nanotechnology and its therapeutic and diagnostic prospects in ophthalmology, specifically glaucoma. Nanotechnology and nanomedicine history, the human eye anatomy, research frontiers in nanomedicine and nanotechnology, its imaging modal quality, diagnostic and surgical approach, and its possible application in glaucoma will all be further explored below. Particular focus will be on the efficiency and safety of this new therapy and its advances.

Keywords: nanotechnology; nanomedicine; drug delivery; MEMs; glaucoma; intraocular pressure; imaging

1. Introduction

Nanotechnology is a scientific nanoscale engineering technology conducted between the scales of nanometers [1]. In 1959, Richard Feynman, who is perceived as the father of nanotechnology, proposed the use of nanoscale machines in molecular and atomic modifications [1]. This submission was not properly examined until 1981 when the first scientist observed the nanoscale on a scan tunneling microscope [2]. Thereon after, our understanding and expertise of molecular and atomic
scale modification became vastly revamped. Between the years 2000–2019, there has been a worldwide investment of approximately US$150.5 billion in the continuous research and development of nanotechnology. There has also been an increase in annual publications on nanotechnology since 1999 all the way to 2019, and this was mainly focused on drug delivery [2–4]. Nanotechnology is defined in two ways by Wagner et al. [5]: as technology using already existing knowledge on the human body and providing molecular aid for treatment and diagnostics, and it is further described as using nanostructure materials with divergent therapeutic effects. Kim et al. [6] further elaborated the description of nanomedicine in 2014 as using the aid of engineered nano-devices and nanostructures that operate extensively side by side at a single cell level for thorough examination, defense, control, repair, structuring, and enhancement at a molecular level of human biological systems, thus championing its medical goal.

The size of the particles used is seen as the main characteristic of nanotechnology [7]. Although using the traditional physics approach may not elaborately define matter interactions on a macroscopic level, nanoscale interfaces remain, however, primarily defined using quantum mechanics [8], therefore, nanomaterials have distinct chemical and physical characteristics diverse from materials at a macroscopic scale. One result of smaller particles is the expanded surface-to-volume proportion. Considering the per-unit size of a given material, a nanomaterial will offer a larger number of locales for synthetic responses than a macro scale material [8]. There are various nanomaterials that could be of extraordinary interest to the medicinal network because of their one-of-a-kind physical and compound attributes. Many nanomaterials give a gamut of useful options that embrace electrical conductivity, magnetic properties, biocompatibility, and biodegradability [5,9,10].

Glaucoma, a disease characterized by neurodegeneration which affects the retinal nerve fiber layers and the optic nerve head, is considered as one of the world’s leading causes of visual impairment. It is projected that by 2040, 111.8 million people worldwide will have the disease [11,12]. Amidst numerous subtypes, primary open-angle glaucoma (POAG) is the most typical type [13]. An important pathological risk for primary open-angle glaucoma is the degree of the intraocular pressure (IOP) [14]. The IOP is generated within the eye’s anterior chamber by resisting aqueous humor evacuation from the trabecular meshwork (TM) and the inner wall of the Schlemm’s canal (SC) (Figure 1) [15,16].

Ocular diseases have a direct effect on the sense of vision and value of human life. An assessment from thirty-nine countries found visual impairment affects 285 million people. Among these, 82% of the visually impaired are patients over 50 years [17]. A lot of improvements have been made in research on the management of ocular diseases and its pathogenesis [18–21]. However, because of the unique physiological obstacles and anatomic structures of the eye, the diagnostics and therapy of these diseases may lead to low potency and lack of specificity. Presently, therapy may only sometimes act to complete the restoration of eyesight or be able to detect serious eye diseases at an early stage [22]. Hence, there is a rapid increase in the attention shown on improving medical specificity and diagnostics and the medication for eye diseases. This review is centered on the advances in nanotechnology systems used in medical aid, therapy, and imaging in glaucoma therapy. Foremost, the precise anatomy and also the related limitations in ocular drug delivery systems are introduced. A number of the standard, as well as other drug delivery systems, are also explored. Next, in order to provide a more complex insight into the mechanism of nanosystems, many examples of nanotechnology and nanomedicine in the control of glaucoma are reviewed. Lastly, we present a summary of the nanotechnology angle and current obstacles in glaucoma diagnosis and therapy. Our review will not only provide incentive for superior design of glaucoma managements but also inspiration for further research.
2. Anatomy of the Human Eye and its Constraints

The human eye is an organ for vision and reacts to pressure and light [23]. It provides a three-dimensional mobile image usually colored in daylight. Cone cells and rod cells located in the retina help in sensing light and vision perception. Colors can be distinctively differentiated and depth perception is observable. The eye can distinctly tell apart about 10 million [24] colors and may be capable of detecting a single photon [25]. Akin to other mammalian eyes, the eyes of humans have non-image-forming photosensitive ganglion cells located in the retina, which allow light signal reception and regulate pupil size and repress the presence of the melatonin hormone and body-clock training [26]. The human eye has more of a coalesced two-piece unit shape as opposed to the conventional spherical shape proposition. There are two segments: the anterior, which encompasses the cornea, lens and iris, and; the posterior (Figure 2).

The cornea, which is about 0.5 mm (500 μm) thick and 11.5 mm (0.3 in) in diameter, with a clear curvature connected to the larger posterior section comprised of the retina, vitreous humor, choroid, and the outermost white shell known as the sclera [23]. The posterior fragment makes up its remaining five sixths with a diameter of about 24 mm. The sclera and cornea are linked together by the limbus. The pigmented framework around the eye center and the darkly pigmented pupil is called the iris. The pupil size regulates the quantity of light penetrating the eyes, this is controlled by the iris dilator and muscles in the sphincter [23].

The human eye consists of three layers surrounding many anatomical structures [27,28]. The furthermost part is the fibrous tunic, containing the sclera and cornea. The middle part, which is called the uvea or vascular tunic, is composed of the iris, ciliary body, pigmented epithelium, and choroid. The inner part of the eye is the retina, which uses retinal vessels (interiorly) and choroid blood vessels for oxygenation [23].
The environment in the human eye has two major walls—the blood-aqueous and blood-retina walls. The former wall encloses the ciliary body’s pigment-less epithelium, which contains the Schlemm’s canal endothelium, iris epithelium, and the iris vessel endothelium. Cellular transports are controlled by the cell junction encompassing both the para and active types [29–31]. The latter barrier is fragmented into two, i.e., the outer layer and inner layer. The inner blood-retinal barrier carries the retinal vascular endothelium with tight junctions, while the outer blood-retinal barriers have a single layer of retinal pigment epithelium (RPE) with tight junctions [30,32]. The mentioned fragments constrict molecular penetrations into the intraocular chamber, leading to a deficiency in intraocular tissue treatment. Additionally, site-based drug delivery on the anterior ocular section is usually restricted by a corneal surface clearance mechanism and other pre-corneal elements, which could be the blinking of the eye, solution discharge, tear turnover, and film eye tearing [33]. The tear film of the human eye has a quick restoration period of about 2–3 minutes. Therefore, many site-based delivery drugs will be cleaned out in a short time after installation, e.g., precisely 2-3 minutes. If the drug solution which is topically applied is greater than 30 µL (the volume of the upper limit quartered in the cul-de-sac), a vast majority of drugs used may either be emptied by gravity-induced discharge or nasolacrimal drainage. Due to these obstacles and factors, the total drug efficiency of the medicine given is considerably less than 5%, thus rendering the bioavailability low [34,35].

![Figure 2. Anatomy of the human eye and administration routes for both nano based medicines and traditional drugs: Blue arrows indicates various eye structures and the green arrows indicate the various administration routes.](image)

### 3. Advancements of Nanomedicine in Glaucoma

IOP-lowering agents are the most common form of anti-glaucoma treatment which is usually administered as eye drops [36]. Production of aqueous humor is suppressed by beta-blockers [37], carbonic anhydrase inhibitors [38], and alpha agonists which will reduce ocular pressure [39]. In order to enhance outflow, prostaglandin-like substances and other compounds can be used [40]. Patient compliance is very low as the medicine must be administered several times. There are other factors to be considered, like cornea permeability, tear turnover, the bioavailability of the medication, and short drug action duration. In order to overcome these limitations associated with this treatment, a more effective drug delivery system must be employed. It is in this case that nanotechnology-based delivery systems can be useful for newly developed drugs, which will provide sustained release, better bioavailability, an accurate dosing regimen, negligible irritation to tissues, lengthier shelf-life, targeted delivery, and enhanced solubility properties (Figure 3) [41,42].
Figure 3. Major challenges in ocular disease treatment with eye drop formulations.

The nano delivery systems can make use of nanoparticles (NPs) [43], nanoemulsions [44], nanodiamonds (NDs) [45], nanocrystals [46], liposomes [47], dendrimers [48], cyclodextrins [49], and other devices [50] together with contact lenses (Figure 4). The physicochemical properties (hydrophobicity, length, and stability), target tissue and the administration route determine which device and materials to use. Nonetheless, in order to get the desired effect, nontoxicity of the delivery system, its biocompatibility, balance, and several other factors must be considered to suit the therapy.

Figure 4. A scheme showing the structures of some nano-formulations used in glaucoma treatment: Nanoparticles with the ability to pass through biological barriers, nanoemulsions with increased drug solubility and membrane penetration, nanodiamonds with accessible surface area and tailorable surface chemistry, hydrogels, and nanocrystals for the conveyance of poorly water-soluble medicines, dendrimers, and liposomes capable of carrying hydrophobic and hydrophilic medicines, contact lens capable enzyme triggered release and cyclodextrins with the ability to form inclusion complexes enclose the lipophilic drug without changing its molecular structure.
Pilocarpine (a muscarinic receptor agonist), brimonidine (α2-adrenergic agonist), timolol and carteolol (beta-blockers), latanoprost (prostaglandin analog), and brinzolamide (carbonic anhydrase inhibitor) are modern anti-glaucoma medicines that have been employed in formulations as nanomedicines in glaucoma therapy [41,51,52]. In vivo rabbit studies used nano-drug preparations of timolol in the form of nanofiber patches and contact lenses and a significant reduction in the intraocular pressure was observed as compared with using drugs solutions [53]. In other studies, sustained drug release was observed when subconjunctival administration of anti-glaucoma microspheres was administered [54,55]. This was observed for three continuous months in in vitro experiments [56]. In addition, under in vitro conditions, 5% 1:1 timolol: propoxylated glyceryl triacrylate nanoparticles loaded onto a contact lens exhibited sustained release properties for up to one month [57].

**Triggered, Controlled and Sustained Drug Release in Glaucoma**

During initial animal research in beagles, an effective reduction in the intraocular pressure was observed. A different and new contact lens layout containing vitamin E and timolol, known as ACUVUE TruEye, elicited a drastic reduction in the intraocular pressure [41,58]. Not very long ago, a device for the delivery of timolol was engineered and its lens was made to be enzyme-triggered [45]. Nanodiamonds were coupled to polyethyleneimine then chitosan was incorporated. A poly-HEMA matrix was fused to the nanodiamonds and fixed onto the lens. No drug release was seen when lysozyme was absent. It can be concluded, that the nanodiamonds ensured a controlled release of timolol when lenses were embedded with enzyme-cleavable polymers [45]. Alternatively, a nanofiber patch gadget containing polymers (polycaprolactone and polyvinyl alcohol) were also examined for the delivery of timolol after insertion of the cul-de-sac [59]. A 24-hour sustained drug release and intraocular pressure management for a period of 72 hours were observed in NZ (albino) rabbits using this new approach.

Natarajan et al. reported on a long-acting nanocarrier-based design for use in glaucoma therapy [47,60]. They reported a significant and successful improvement of a drug-encapsulated nanocarrier for decreasing intraocular pressure. A diseased nonhuman primate model was used and an explanation was given as to why in vivo sustained drug release was possible. Latanoprost, which is a prostaglandin analog, was used in this experiment and the carrier chosen was a unilamellar vesicle that was nanosized. Isothermal titration calorimetry explained the mechanism of action of this particular drug-nanocarrier interaction. Dynamic light scattering (DLS) and a Cryo-TEM were used to demonstrate that the shape of the liposomes was not altered and there was maximum drug loading of Latanoprost in the nanocarrier. As the first nanomedicine in glaucoma therapy, in vivo results clearly indicated good outcomes and confirmed the ability of this formulation to lower intraocular pressure (Figure 5) [47]. Considering the results, further work must be done to ensure nanocarriers are employed in glaucoma therapy using higher primates. A reduction in ocular pressure was observed in other experiments [61–64].

Recently, brinzolamide nanocrystal was synthesized using stable crystals [46,65]. A preliminary fast dissolution rate was observed indicating better bioavailability. Nonetheless, all the drug-loaded nanocrystals dissolved completely after one-hour. In comparing with the marketed product (49%), the intraocular pressure-lowering effect of the nanocrystals (79%) was very high in the animal models [46]. After a glaucoma surgical operation, a heat-sensitive injectable hydrogel for the administration of bevacizumab has been recently developed [66]. Polyethylene glycol–poly-ε-caprolactone–polyethylene glycol hydrogel after being injected into glaucomatous rabbit models exhibited no signs of increased intraocular pressure and infection. Comparing postoperative and preoperative values, the drug-loaded hydrogels showed a vast decrease in intraocular pressure within a period of three weeks.
Liao et al. [67] in recent research, engineered pilocarpine-loaded gelatin-covered mesoporous silica nanoparticles (p/GM). These silica nanoparticles were administered via the intracameral route into the anterior chamber of the rabbit eye. This was done in order to reduce intraocular pressure (Figure 6A). Considering the in vitro release data, a long-lasting (36 days) and high percentage release (50%) of p/GM were observed. Intraocular pressure was maintained in the eye suffering from ocular hypertension for a period of 21 days (Figure 6B). For a sustained release behavior, the gelatin coat was controlled by varying the thickness on mesoporous silica nanoparticle, thereby enhancing the drug delivery of the nanoparticle. Looking at the in vivo and in vitro data, the synthesized p/GM-0.05 sample showed an extended-release profile and the potency to reduce intraocular pressure.

Hassan et al. [68] in new research, studied the use of cationic nanoparticle (leciplex) in carvedilol delivery to ocular surfaces as a glaucoma therapy since other research [69,70] illustrated that carvedilol had an effect on intraocular pressure. As such, the leciplex formulae filled with carvedilol were made with cationic surfactant (CTAB/DDAB), soy phosphatidylcholine (SPC), and characterizations were done. After the formula used was evaluated and compared with carvedilol solution in vivo-studies, nanoparticles in leciplex were observed to be spherical in shape with an entrapment efficiency of 95% in all formulae. The leciplex preparation of DDAB and SPC in a molar proportion of 1:1 had the tiniest particle size (16.04 ± 1.2 nm), coefficient of the highest apparent permeability of the cornea at 0.1157 cm/h, and a good zeta potential (53.9 ± 0.91 mv). Carvedilol-leciplex complex was capable of decreasing intraocular pressure in rabbits with hypertension to a regular range 30 mins after administration over a period of 24 hours. Nonetheless, the solution of carvedilol decreased intraocular pressure 60 mins after administration for a period of 6 h. Previous examinations of carvedilol-leciplex administered to rabbit eyeballs with glaucomatous eyes improved their retinal atrophy. Proving that leciplex had a noticeable improvement in carvedilol bioavailability and transcorneal permeation.

Figure 5. Comparative analysis between Xalatan eye drops and Latanoprost-loaded liposomes administered via the subconjunctival route in nonhuman primates. A great reduction of intraocular pressure was observed when a single injection of Latanoprost-loaded liposomes was done via the subconjunctival route for a period of 120 days in comparison with Xalatan eye drops. After a second injection, the intraocular pressure reduced further over another period of 180 days. Reproduced from [47] with permission from the American Chemical Society.
Figure 6. (A) Pilocarpine-loaded gelatin-covered mesoporous silica nanoparticles (p/GM) designed to reduce intraocular pressure. (B) Time-course slit-lamp biomicroscopy images of rabbit eyes injected with p/MSN, p/GM-0.05, and p/GM-0.5. After pilocarpine injection, the observation times are indicated as H12, D2, and W3 on a scale bar of 4 mm. Reproduced from [67] with permission from The Royal Society of Chemistry.

Li et al. [71] engineered novel nanoparticles to be used topically in eye-controlled drug delivery systems by insertion of betaxolol hydrochloride (BH) into the inner sheet of Na-montmorillonite (Na+Mt) and in addition, enashing chitosan nanoparticles (Figure 7). The obtained nanoparticles had a diameter of about 460 ± 0.6 nm on average and a positive charge (+29 ± 0.18 mV).

The in vitro research on the drug release profile showed regulated drug release patterns. The experimental studies of human immortalized cornea epithelial cells (iHCEC) and CAM-TBS found satisfactory ocular tissue tolerance. Additionally, nanoparticles were surprisingly found to have the ability to penetrate the iHCEC through cellular uptake observation by confocal layer scan microscopy (CLSM) measurements. Furthermore, iHCEC with many layers were used in in vivo precocular capacity of retention studies by corneal epithelial cells barrier stimulation showing that BH-Mt/CS nanoparticles compared with betaxolol hydrochloride solution had the ability to increase retention time. The pharmacokinetic studies of the eye through the technique of microdialysis sampling reported that MRT0−t of BH-Mt/CS nanoparticles and AUC0−t remained approximately 1.99-times and 1.75-times more than the betaxolol hydrochloride solution, reporting greater bioavailability. However, blood drug concentration studies by some researchers pinpointed that drugs with low levels could enter the blood, highlighting less system side-effects. Notably, the studies in pharmacodynamics disclosed that BH-Mt/CS nanoparticles could significantly drop glaucomatous rabbit’s intraocular pressure. Through this finding of chitosan/montmorillonite nanoparticles, it can be envisaged that
BH-Mt/CS will be a potential carrier for betaxolol hydrochloride, thus making it an ideal candidate for glaucoma therapy.

Figure 7. Schematic representation of the BH-Mt/CS nanoparticles preparation process. Reproduced from [71] with permission from Dovepress.

4. Nanosystems for Posterior and Anterior Glaucoma Therapy

Over the last decade, there has been development of a lot of traditional nanosystems for use in glaucoma anterior disease (Table 1). Contrary to anterior glaucoma diseases, diseases also affect the posterior section. Nanosystems currently used in the treatment of posterior glaucoma disease therapy are provided in Table 2. Many strategies based on nanotechnology in diagnosis of glaucoma diseases are further listed in Table 3.

Table 1. The current and traditional strategies based on nanotechnology used in the therapy of anterior glaucoma disease.

| Formulation                  | Role/Function                                                                 | Cargo    | Material Type | Size (nm) | Stage            | Ref.       |
|------------------------------|------------------------------------------------------------------------------|----------|---------------|-----------|------------------|------------|
| Nanoparticle                 | Exhibition of superior transfection potency at the anterior region of the eye. | Gene     | Chitosan      | ~200      | Preclinical stage | [72]       |
| Nanowafer                    | High level of nontoxicity with a low dose and ability to efficiently treat corneal neovascularization as compared with eye drops. | Axitinib | Polymer       | 500       | Preclinical stage | [73]       |
| Nanosuspension               | Increasing retention time and penetration in corneal tissues.                | Diclofenac | Polymer       | 105       | Preclinical stage | [74]       |
| Hydrogel (Virgan)            | Treatment of herpes simplex infection in the eye.                           | Ganciclovir | Polymer       | Data not found | On the market | [75–78]   |
| Nanoparticle                 | Increased anti-inflammatory effect via the topical route.                    | Flurbiprofen | Polymer       | 200–300    | Preclinical stage | [79]       |
| Nanoparticle                 | Improved the efficacy of the drug to aid in corneal graft rejection.         | Dexamethasone sodium phosphate | Polymer | 100–500 | Preclinical stage | [80]       |
| Nanoscale dispersed ointment | Improved efficacy of corneal surface restoration and tear film repairing.   | Data not found | Polymer | 100 | Preclinical Stage | [81]       |
| Nanoparticle                 | Inhibitory effect in rabbit models with a mild response using a low drug concentration. A higher volume of the drug penetrated the aqueous humor in comparison with eye drops. | Flurbiprofen | Polymer | 100 | Preclinical stage | [82]       |
Table 1. Cont.

| Formulation | Role/Function | Cargo | Material Type | Size (nm) | Stage         | Ref.   |
|-------------|---------------|-------|---------------|-----------|---------------|--------|
| Hydrogel    | Improvement of drug bioavailability in the eye as well as retention time on the surface of the cornea. | Diclofenac | Polymer | Data not found | Preclinical stage | [83]  |
| Liposome    | High anti-cataract effect with enhanced superoxide dismutase activity and glutathione reduction. | Coenzyme-Q10 | Polymer | 100–200 | Preclinical stage | [84]  |
| Nanoparticle | Increase miotic response to 40%. | Pilocarpine | Polymer | 83 | Preclinical stage | [85]  |

Table 2. The current and traditional strategies based on nanotechnology used in the therapy of glaucoma posterior diseases.

| Formulation | Role/Function | Cargo | Material Type | Size (nm) | Stage         | Ref.   |
|-------------|---------------|-------|---------------|-----------|---------------|--------|
| Nanoparticle | Provide sustained drug release via the subconjunctival route. | Latanoprost acid | Polymer | 80 | Preclinical stage | [47]  |
| Hydrogel    | Sustained drug release of Bevacizumab in SD rats for a period of four months. | Bevacizumab | Polymer | Data not found | Preclinical stage | [86]  |
| Hydrogel (Timoptic-XE) | Treatment of glaucoma. | Timolol maleate | Polymer | Data not found | On the market | [87]  |
| Liposome    | Ability to pass through biological barriers after topical administration via annexin-A5 mediated endocytosis | Bevacizumab | Polymer | 100–200 | Preclinical stage | [73]  |
| Hydrogel    | Sustained drug release and decrease intraocular pressure as well as good compatibility with polymer. | Mitomycin C | Polymer | Data not found | Preclinical stage | [88]  |
| Liposome    | Entrapment of RPE cells and increased siRNA delivery by four-fold. | Gene | Polymer | 130–230 | Preclinical stage | [89,90] |
| Micelle     | Sustain drug release for a period of one year after intravitreal injection in rat eyes. | Triamcinolone acetonide | Polymer | 200–350 | Preclinical stage | [72]  |
| Dendrimer   | Effective gene transfection in RPE cell lines. | Gene | Polymer | ~50 | Preclinical stage | [91]  |
| Nanoparticle | Ability to prevent retinal degeneration and increase histological properties. | Gene | Peptide/polymer | ~180 | Preclinical stage | [92]  |
| Nanoparticle | Promotion of gene expression in RPE cells. | Gene | Polymer | ~250 | Preclinical stage | [93]  |

5. Diagnostic and Therapeutic Glaucoma Nanodevices

5.1. Therapeutic Nanodevices

Devices are mostly used in the drainage of glaucoma when trabeculectomy and IOP-lowering medication prove inefficient [50,94]. The mentioned devices give a different route for the aqueous humor to the collection plate beneath the conjunctiva from the anterior section of the eye. Those made by Ahmed, Molteno, Krupin and Baerveldt are the major devices commercially available for glaucoma draining [95–97]. Despite the fact that some differences exist between them, for example, their shape, size of plates, composition, and rigidity, they have successfully been able to contain and regulate IOP (within a period of five years) and there were no noticeable differences in vision preservation. Additionally, no noticeable differences were spotted in the total incidence of suprachoroidal hemorrhage, postoperative hypotony, plate surrounding fibrosis, and inflammation [98]. Fibrosis is the main reason for the overall success decrease in five years from 40% to 50% in drainage implants. Even
if antimetabolite injections like mitomycin C (MMC) and 5-fluorouracil (5-FU) have been found to both post and intraoperatively reduce complications [99], administration of injections to the bleb still causes a risk of infection and discomfort in patients. However, if the drainage device can be coated and used as a delivery vehicle, it could potentially have a greater effect on drug delivery systems (Figure 8B). Ponnusamy et al. [100] successfully manufactured a film of polylactic-co-glycolic acid (PLGA) with two layers which were loaded with 5-FU and MMC for drug release at a continuous rate. The studies conducted in vitro reported MMC was only stable on the surface, so 5-FU was loaded at the bottom. 5-Fluorouracil indicated to be released after three to five days continuously until day 28. After five days, the information obtained showed cell proliferation was inhibited by a COS-I cell culture model. This gives a promising use of glaucoma devices as an inflammation and fibrosis prevention method.

Pan et al. [101] engineered an artificial nano-drainage system that can be implanted in the sclera of the eye and aid flow of aqueous humor. The nano-drainage implant known as ANDI (Figure 8A) is composed of a polymeric shaft and a nanoporous membrane. There was clogging of proteins in the pores of the nanofiltration membrane, though the membrane was able to give the designed flow resistance. This promising nano-drainage system may be further improved by altering the surface chemistry.

A glaucoma valve that contains ferrofluid nanoparticles was developed by Paschalis et al. [102]. The ferrofluid was 10–100 nm in size and was magnetic in nature, had inert properties, and showed supramagnetic behavior. Until the magnetic pressure was lower than the liquid flow pressure, the ferrofluid served as a valve. Using x-ray diffraction, the stability of the device ascertained elicited no oxidation post air and water exposure. The intraocular pressure (11.8 ± 2 mm Hg) reduced after two weeks in comparison with contralateral control (14 ± 3 mm Hg) in in vivo rabbit studies. A plug filter was then engineered by Maleki et al. [103] to address postoperative hypotony (Figure 8C). Intraocular pressure is reduced when this filter is used but only for the short-term. Nonetheless, the hurdle of postoperative hypotony was overcome. To curtail this, the filter was engineered with polylactic acid (PLA) or PLGA. The plug was capable of reducing the intraocular pressure for a period of 15 days.

Another work worth looking at was reported by Harake et al. [104]. They fabricated and engineered an antifouling micro glaucoma apparatus employing microelectromechanical methods. The device was composed of PEG-4000 and PEG-214. This polymer composition prevents swelling and clogging, thus making it an appropriate tool in preventing biofouling. The design was effective and resistant to E.coli and in vitro studies showed a lower specificity for protein absorption in comparison with polypropylene, glass, polymethyl-methacrylate, and polydimethylsiloxane. Further studies are being worked on to fully validate the efficiency of the device.

**Figure 8.** Examples of drainage devices engineered using nanotechnology approaches. (A) The ANDi is implanted in the sclera. Reproduced from [101] with permission from IEEE. (B) Drug-loaded drainage devices are manufactured with a film layer loaded with drug for continuous release. Reproduced from [99] with permission from Elsevier. (C) The plug filter is manufactured with a biodegradable polymer to eliminate postoperative hypotony. Reproduced from [103] with permission from IEEE.
5.2. Measurement of Intraocular Pressure

The extent of intraocular pressure plays a vital role in glaucoma therapy [105]. Leo et al. [106] developed a non-invasive, wireless, and soft silicone contact lens capable of detecting changes in intraocular pressure. A gauge sensor of platinum-titanium which was 170 nm thick and embedded in two layers was used. In order to power the sensor, a gold antenna, microprocessor, and an integrated circuit were fixed in the lens, this also aided in wireless communication via an external recording unit [106]. As the stress in the gauge increased, other mechanical forces compressed and altered the electrical forces present in the gauge. Leo et al. observed that the contact lens exhibited linear and reproducible intraocular pressure in the range (15–30 mm Hg) in enucleated pig eyes. Nonetheless, the contact lens was tested on 11 patients by Mossbock and Faschinger and had limited success [106,107].

Aside from developing nanomaterials in drug delivery, nanotechnology has been able to be applied in the manufacturing of monitors capable of tracking intraocular pressure non-invasively [108–112].

A piezoresistive sensor capable of measuring intraocular pressure was engineered by Rizq et al. [113]. The sensor is capable of detecting changes in electrical resistance when stress is applied and had a radio frequency powering reverse telemetry as well as an interface circuit. In addition, Dresher et al. developed a compact circuit with low power which can be fixed in a wireless intraocular pressure monitoring system [114]. Applying the Bourdon tube technique, i.e., using a thin-walled, curved and hollow tube, other devices have been produced and are capable of measuring intraocular pressure [115–117].

Chen et al. used the Bourdon tube technique in developing a sensor that had a protective parylene membrane [115]. Extraordinarily, the device is capable of measuring intraocular pressure without depending on an external energy source. Though these sensors have an advantage, it should be noted that using the Bourbon tube is an invasive procedure and may uncomfortable for some patients [115,118]. All this research and work done serves as a good platform for wireless and implantable devices to be employed in patients with glaucoma and ocular hypertension in the future.

5.3. Imaging

Disc photography initiated the role of imaging in glaucoma, thus enabling comparison and recording of qualitative changes in the optic nerve over a period of time [119]. Clinicians can have oversight in early changes in the optic nerve head (ONH) if they solely rely on photography, as that will be subjective analysis. For this reason, improvements have been made in fabricating stereoscopic images by means of flicker photography, which when employed, improves interobserver agreement on the neuroretinal rim width [120]. For diagnosis and management of glaucoma, the most widely used model is digital evaluation in the form of optical coherence tomography (OCT)[121–125]. Examination of the retinal nerve fiber layer (RNFL) using spectral-domain OCT (SD-OCT) is a good means of differentiating a glaucomatous eye from a healthy eye [126].

For a better understanding of biological pathways, nanoparticles have been employed in fluorescent imaging in glaucoma therapy [73,127,128]. Tam et al. used CdSe/ZnS core/shell nanoparticles which are quantum dots to trace lymphatic drainage in the eye of mice [129]. Unlike other conventional techniques, quantum dots are ideal for non-invasive in vivo imaging as a result of their tunable emission spectra, narrow emission sequence, and broad spectrum of excitation as well as a high photobleaching threshold [130]. The emission spectra may vary accordingly by altering the diameter of the quantum dots, thus allowing good adaption in several instances [129]. The quantum dots showed effective drainage via the skin and peritoneal spaces, and also enabled visualization of a freshly labeled lymphatic drainage path for fluid exiting the eye [129].

5.4. Surgical Implants

Surgical implants are capable of delivering drugs to the eye over a very long period of time. There are already implants for the long-term delivery of steroids. Examples include Retisert [131], an intravitreal implant that conveys fluocinolone acetonide for chronic uveitis over a period of
30 months [132], and Ozurdex (Allergan), a dexamethasone implant capable of conveying the drug over a period of six months via the intravitreal route of administration [133]. Surmodics I-vation implant, a triamcinolone acetonide coated implant with helical screws, is already available for delivering drugs intravitreal and has already undergone a Phase I clinical trial [132]. The use of surgical implants is not totally safe or cost-effective due to: the high cost; invasiveness of initial surgery; and should any adverse effect occur after the surgery, the complexity of removing the surgical implant. A vast number of glaucoma patients are mostly drawn back from undergoing surgery as a result of its associated risks [134].

Surgical implants may nonetheless provide a good platform for the delivery of neuroprotective medicines, as they are capable of delivering neuroprotective medicines to the retina effectively over a long period. For instance, the delivery of ciliary neurotrophic factor (CNTF) [135] can be done from a rice-sized implant through encapsulated cell technology for a period of one year [136]. The ciliary neurotrophic implant has undergone Phase I clinical trials for retinitis pigmentosa. All the patients used in the study showed high tolerance and enhanced visual acuity [137]. For a drug delivery system to be termed as ideal for glaucoma, it must be capable of ensuring a sustained release of the drug over a period of 3–4 months [138]. The 3–4 months drug release period will be ideal to ensure good results are obtained for follow-up evaluations.

Refined Surgical Implants

Surgical implants are capable of delivering ocular medications over a long period of time. Several refined implants are coming up aside of the ones already available. If possible, one would prefer self-medication of ocular drugs to lower intraocular pressure and expect the drug to last for about 3–4 months just as that delivered by the surgical implants [139]. One innovative method is to implant a reservoir system in the subconjunctival space. Electrolysis that creates bubbles and pushes the medicine pot of the reservoir of the device is possible using a microelectromechanical system (MEMS) permitting multiple drug loading [140,141]. There are some similarities between the surgical steps and available glaucoma drainage devices, as the implant can be reloaded a lot of times and showed high tolerance in rabbit studies [142]. This is promising for the delivery of both small and large neuroprotective molecules, for example, growth factors [143,144].

An additional benefit of the microelectromechanical system is that by controlling electrolysis, the rate of drug release from the device can be regulated. Via an active delivery system, any clinician can alter the delivery rate upon assessment [145,146]. With very little modification, multiple drugs can also be administered via the intravitreal route using this device. In order to evaluate the sustained release function of the device and its stability, long term studies must be done. A major demerit and risk is that this implant must be surgically implanted in the eye, which poses some long term challenges [147].

Table 3. Prospective nanotechnology-based approaches for glaucoma diagnostics.

| Formulation | Role/Functions | Target | Material Type | Size (nm) | Stage | Ref. |
|-------------|----------------|--------|---------------|-----------|-------|------|
| Nanoparticle | Nanoparticles coated with calcium elicit slight damage and can be used in retina imaging | Retina | Silver | 80 | Preclinical stage | [148] |
| Nanoparticle | Improvement of angiogenic vessels in a rabbit corneal neovascularization model | Corneal neovascularization | Gold | ~260 | Preclinical stage | [149] |
| Nano-cage | Good optical resonance | Retina | Gold | 35 | Preclinical stage | [150] |
| Nanoparticle | Enhanced and clear fluorophores in eye imaging | Intraocular cancer | Quantum dots | 3–6 | Preclinical stage | [129, 151] |
| Nanoparticle | Diffusion of nanoparticles present in in vitro model of human vitreous humor | Retinal detachment | Magnetic Nanoparticles (Fe3O4) | 10 | Preclinical stage | [152] |
6. Replacement and Regeneration of the Trabecular Meshwork (TM)

The primary drainage system for aqueous humor is the trabecular meshwork (TM), and it works as the major structure for controlling intraocular pressure [153]. This tissue has a sheet-like extracellular matrix (ECM) covered with endothelial-like and beams. Present are irregular inter-trabecular spaces making up the juxtacanalicular region, adjacent to the Schlemm’s canal (SC), and in between the sheets and beams. Prior to aqueous humor passing through the Schlemm’s canal (SC) and juxtacanalicular, the outer layer of the trabecular meshwork, which is phagocytic in nature, filters cell debris. Numerous fluctuations have been observed in the structure and function of the trabecular meshwork for a glaucomatous eye. This includes herniation in the extracellular matrix, variations in the levels of protein in the extracellular matrix, and improved action of calcification markers [154,155].

One of the recommended approaches for TM-focused treatments is gene therapy. Over the years, gene transduction has been made possible via the generation of recombinant adenoviral (Ad) vectors [156]. A dominant-negative RhoA gene carried by an adenoviral vector was effective in inactivating RhoA and lowering intraocular pressure in organ cultures. Actin cytoskeleton reorganization ceased as a result of inactive RhoA incapable of activating Rho kinase (ROCK). In this regard, intraocular pressure was reduced due to cellular relaxation in the trabecular meshwork [157], thus making ROCK inhibitors potentially beneficial in glaucoma therapy. It is possible to transduce genes using adenoviral vectors. To do this, an adenoviral vector carrying the matrix metalloproteinase 163 gene [158] controlled by glucocorticoid response elements was engineered, and it was observed that it could effectively lower intraocular pressure in a glaucomatous steroid sheep model [159].

Cell-based restoration of the trabecular meshwork tissue is a good alternate strategy because of its promising results in decreasing disease-related trabecular meshwork cells. To this effect, TM cells have been studied and were shown to localize in trabecular tissues after injection into the anterior capsule of mice for up to four months. This was done to ascertain its localization and differentiation into functional trabecular meshwork cells [160]. The presence of the expressed TM marker protein CHI3L1 and cell differentiation was detected in week one. This research served as a promising outcome for cell-based therapies and acts as a platform for TM tissue-based treatment.

Likewise, a biomimetic inner wall of the Schlemm’s canal that can serve as a tool for studying outflow physiology and drug screening was engineered with microfabrication techniques [161]. Furthermore, to test biomechanical roles and the effect of various therapeutic agents, artificial trabecular meshwork tissues are a good candidate. Micropatterned porous SU-8 scaffolds coated with poly-L-lysine or gelatin to enhance trabecular meshwork attachment and cultured with human TM-cells for a 14-day period was the first attempt in this regard [162] and others followed [163,164]. A gelatin-coated 12-µm SU-8 microstructure was concluded to be the best design for meshwork formation and cell growth using perfusion studies and immunohistochemistry [162].

Snider et al. in 2018 [165] used magnetic steering to direct magnetic nanoparticles (Prussian blue nanocubes [PBNCs]) to the trabecular meshwork after injection of the nanoparticles into the eye’s anterior chamber. Comparing unlabeled cells and TM cells, PBNC-labeled stem cells displayed improved delivery after only a 15-minute exposure to a magnetic field (Figure 9). Further, the delivery of PBNC-labeled mesenchymal stem cells (MSCs) to the entire circumference of the trabecular meshwork was not possible without steering with a magnet. The multi-potency and viability of the mesenchymal cell were not affected by the PBNC (Figure 10). In conclusion, a targeted and high-efficiency delivery of stem cells to the trabecular meshwork was achieved using this approach, which are essential steps concerning regenerative medicine therapies in regulating ocular hypertension in glaucoma patients.

Dillinger et al. [16] proposed a novel, causative therapeutic idea which involves delivering small interfering RNA against CTGF via the intracameral route. A final layer of hyaluronan (HA) with layer-by-layer coated nanoparticles of 200–260 nm were synthesized. It was observed that the HA-coating allowed binding to TM and SC cells via CD44 and provided the nanoparticles enough mobility in ECM. The strength of the model was established by screening primary SC cells and TM ex vivo, in vivo, and in vitro. There was elevation of CD44 expression in healthy versus glaucomatous cells.
by about 2–6 times. CD44 is considerably involved in the cellular uptake of HA-coated nanoparticles. Compared to control nanoparticles, ex vivo organ culture of murine, human eyes and porcine validates up to threefold higher accumulation of HA and far enhanced permeation into the target tissue. In conclusion, a significant reduction of CTGF expression is brought about by gene silencing in primary human TM cells. Thus, RNA interference combined with HA-coated nanoparticles has great prospects in glaucoma therapy.

Figure 9. Outcome of Magnetic Field Exposure Time on directing 200 nm PBNC-MSCs to the trabecular meshwork. Figure (A–D) Shows anterior segment of the eye post mesenchymal stem cell delivery. There were bar magnets around the limbal region with *C* indicating the center of the human cornea. Figure (E–H) shows the intensity of total fluorescence within the trabecular meshwork area. P-values were calculated by means of Kuiper’s V test to evaluate if distribution was non-uniformly skewed in the direction of the focus of the magnet (0°). (A,E) 15 mins (n = 4 eyes), (B,F) 30 mins (n = 5 eyes), (C,G) 60 mins (n = 3 eyes), and (D,H) overnight (n = 4 eyes) with its exposure to the magnet are presented. Figure (I) indicates the percentage of fluorescent cell signal in magnetic quadrant in the trabecular meshwork, defined as the section encompassing 45° to −45°. The dotted line designates the expected value should there have been uniform cell delivery to the entire circumference of the TM (25%). Figure (J) Trabecular region fluorescence signal in comparison with non-trabecular fluorescent signal within the zone encompassing −45° to +45°. All evaluations between groups were not significant (p > 0.05) using one-way ANOVA with post hoc Tukey survey. Adapted from [165] with permission from Scientific Reports.
7. Major Challenges, Expected Breakthrough, and Conclusions

7.1. Major Challenges

Effective glaucoma drug treatment has numerous setbacks, all of which come under the headline of building a better eye drop [166,167]. Issues of how safe nano-based medicines are pose a major challenge in nanotechnology, though there have been great advancements in nano-based drug delivery [168]. The presence of particles, contaminants, and inflammatory variants in nanoparticle systems is a major
As a result of Van der Waals forces, induced dipole force of attraction and hydrophobic interactions, numerous nanoscale materials show different characteristics. This allows interactions of proteins, biological membranes, and bodily fluids with nanoparticles possible. The shape and size of nanoparticles play a vital function in the effect on health. The size of the nanoparticles is inversely proportional to the surface area-to-mass ratio. This yields several reactive sites capable of undergoing bio interactions and uptake mechanisms [170,171].

The coating and surface charge of the nanoparticle determines its degree of effect on health, specifically with respect to toxicity. There is a better chance of positively charged particles undergoing maximum cellular uptake and interacting with intercellular membranes that are negatively charged. There is a higher probability of charged particles interacting with membranes and being cleared faster in comparison with neutral particles [170]. However, if released from the surface, the composition of the surface coating can cause adverse toxicity [172]. Particularly, the basis of biological interactions depends on the chemistry of the nanoparticles involved. Several nano-based systems have been used in the area of ophthalmology ranging from synthetic polymers to natural polysaccharides with their toxicity profiles very low [169]. Among these nano-based systems, PLGA, which is a biodegradable polymer, is considered the least toxic and approved by the US Food and Drug Administration (FDA). Likewise, at the cell penetration stage, polycaprolactone nano-capsules are also considered safe by the FDA [173]. Nano-micelles have also shown slight toxicity in biological systems [174] as well as acrylic copolymer Eudragit nanoparticles [169].

We focused on advances in nanotechnology and nanomedicine in glaucoma therapy in this review. Nanotechnology has demonstrated to be an effective tool in glaucoma therapy. In vitro and in vivo delivery of nanoparticles to treat glaucomatous eyes have been seen in several nanosystems with diverse cargos. Nonetheless, a lot of challenges exist and must be addressed in future research. Some include: (1) The rabbit’s eye possesses low tear production, high mucus production, high surface sensitivity and has a size comparable with the human eye [175]. These qualities are not precisely as the human eye, thus making mimicking difficult. (2) A number of the research is done for in vitro studies and has not focused much on in vivo studies. (3) Expansion and increase in the size of some nanoparticles after injection via the intravitreal route [176–178]. This can affect drug distribution and delivery efficiency. (4) Ocular disease-related biomarkers, molecular and cellular mechanisms, are not fully understood for targeted delivery [179]. All these must be addressed to fully curb and ensure more effective glaucoma therapy.

7.2. Expected Breakthrough

Evidence is building that nanoparticles will serve as a promising drug delivery system, and it will not take much time until this technology reaches patients in the mainstream [180,181]. The quest for tailored medicine approaches in fighting glaucoma as a disease may be promising only through the development of nanotechnology platforms which encompasses molecular-level engineering. Nanoparticle engineering is a common thread for several imaging paradigms and drug delivery systems [182,183]. Nanotechnology has served, is still serving, and has promising prospects in glaucoma therapy and diagnosis. The eye is considered as one of the body’s most delicate organs and a perfect platform for drug and gene delivery because it bypasses systemic circulation. More than 1600 gene therapy clinical trials for ophthalmology are ongoing according to data from the Wiley Library [184]. Non-invasive routes of administration should be focused on using various nanomaterials and devices. Lastly, a system that combines therapeutic functions and diagnostics should be built to aid in visual tracking in glaucoma therapy.

7.3. Conclusions

Several effective anti-glaucoma medicines and devices have been developed over the years. On the other hand, poor patient adherence, side effects, poor bioavailability, and inefficient delivery systems limit their clinical efficacy. Innovative and more competent delivery systems are being developed
to improve patient adherence, reduce toxicity profiles, and side effects. Finally, these novel delivery systems for potential neuroprotective drugs and lowering intraocular pressure can initiate greater treatment possibilities and preservation of vision in glaucoma. Further investigations and research need to advance our understanding of the fundamentals of nanoparticles and encourage development of proper delivery routes for glaucoma therapy. Considerable clinical and basic research needs to be done to ensure progress in the direction of personalized medicine for refining treatment outcomes for patients with glaucoma [185].

Funding: This paper received funding from the Chinese Scholarship Council (CSC).

Acknowledgments: The authors are appreciative of the Chinese Scholarship Council (CSC) grant number-2016288010 for their generous financial support to this Paper.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Initiative, U.S.N.N. What It Is and How It Works. Available online: https://www.nano.gov/nanotech-101/what (accessed on 1 May 2019).
2. Li, X.; Chen, H.; Dang, Y.; Lin, Y.; Larson, C.A.; Roco, M.C. A longitudinal analysis of nanotechnology literature: 1976–2004. J. Nanoparticle Res. 2008, 10, 3–22. [CrossRef]
3. Gao, W.; Chan, J.M.; Farokhzad, O.C. pH-responsive nanoparticles for drug delivery. Mol. Pharm. 2010, 7, 1913–1920. [CrossRef] [PubMed]
4. Wang, B.; Tang, Y.; Oh, Y.; Lamb, N.W.; Xia, S.; Ding, Z.; Chen, B.; Suarez, M.J.; Meng, T.; Kulkarni, V. Controlled release of dexamethasone sodium phosphate with biodegradable nanoparticles for preventing experimental corneal neovascularization. Nanomed. Nanotechnol. Biol. Med. 2019, 17, 119–123. [CrossRef] [PubMed]
5. Wagner, V.; Dullaart, A.; Bock, A.-K.; Zweck, A. The emerging nanomedicine landscape. Nat. Biotechnol. 2006, 24, 1211. [CrossRef] [PubMed]
6. Kim, N.J.; Harris, A.; Gerber, A.; Tobe, L.A.; Amireskandari, A.; Huck, A.; Siesky, B. Nanotechnology and glaucoma: A review of the potential implications of glaucoma nanomedicine. Br. J. Ophthalmol. 2014, 98, 427–431. [CrossRef]
7. Bucolo, C.; Drago, F.; Salomone, S. Ocular drug delivery: A clue from nanotechnology. Front. Pharmacol. 2012, 3, 188. [CrossRef]
8. Zarbin, M.A.; Montemagno, C.; Leary, J.F.; Ritch, R. Nanomedicine in ophthalmology: The new frontier. Am. J. Ophthalmol. 2010, 150, 144–162.e142. [CrossRef]
9. Initiative, U.S.N.N. What’s So Special about the Nanoscale? Available online: https://www.nano.gov/nanotech-101/special (accessed on 1 May 2019).
10. Ibrahim, M.M.; Jablonski, M.M. The Impact of R-801 Nanoparticles as a Long Acting Topical Glaucoma Therapy. J. Biomed. Nanotechnol. 2019, 15, 1968–1981. [CrossRef]
11. Tham, Y.-C.; Li, X.; Wong, T.Y.; Quigley, H.A.; Aung, T.; Cheng, C.-Y. Global prevalence of glaucoma and projections of glaucoma burden through 2040: A systematic review and meta-analysis. Ophthalmology 2014, 121, 2081–2090. [CrossRef]
12. Gross, R.L.; McMillian, B. Current medical management of glaucoma. In Ophthalmol., 5th ed.; Elsevier: Philadelphia, PA, USA, 2019.
13. Braunger, B.M.; Fuchshofer, R.; Tamm, E.R. The aqueous humor outflow pathways in glaucoma: A unifying concept of disease mechanisms and causative treatment. Eur. J. Pharm. Biopharm. 2015, 95, 173–181. [CrossRef]
14. Overby, D.R.; Stamer, W.D.; Johnson, M. The changing paradigm of outflow resistance generation: Towards synergistic models of the JCT and inner wall endothelium. Exp. Eye Res. 2009, 88, 656–670. [CrossRef]
15. Tamm, E.R. The trabecular meshwork outflow pathways: Structural and functional aspects. Exp. Eye Res. 2009, 88, 648–655. [CrossRef]
16. Dillinger, A.E.; Guter, M.; Friemel, F.; Weber, G.R.; Perkumas, K.; Stamer, W.D.; Ohlmann, A.; Fuchshofer, R.; Breunig, M. Intracameral Delivery of Layer-by-Layer Coated siRNA Nanoparticles for Glaucoma Therapy. Small 2018, 14, 1803239. [CrossRef]
17. Pascolini, D.; Mariotti, S.P. Global estimates of visual impairment: 2010. Br. J. Ophthalmol. 2012, 96, 614–618. [CrossRef] [PubMed]

18. Baughman, R.; Lower, E.; Ingledue, R. Management of ocular sarcoidosis. Sarcoidosis Vasc. Diffus. Lung Dis. 2012, 29, 26–33.

19. Weng, Y.; Liu, J.; Jin, S.; Guo, W.; Liang, X.; Hu, Z. Nanotechnology-based strategies for treatment of ocular disease. Acta Pharm. Sin. B 2017, 7, 281–291. [CrossRef] [PubMed]

20. Xue, C.; Rosen, R.; Jordan, A.; Hu, D.-N. Management of ocular diseases using Lutein and Zeaxanthin: What have we learned from experimental animal studies? J. Ophthalmal. 2015. [CrossRef] [PubMed]

21. Sacchetti, M.; Lambiase, A. Diagnosis and management of neurotrophic keratitis. Clin. Ophthalmal. (Auckl. Nz) 2014, 8, 571.

22. Schoenfeld, E.R.; Greene, J.M.; Wu, S.Y.; Leske, M.C. Patterns of adherence to diabetes vision care guidelines: Baseline findings from the Diabetic Retinopathy Awareness Program. Ophthalmology 2001, 108, 563–571. [CrossRef]

23. Atchison, D.A.; Smith, G.; Smith, G. Optics of the Human Eye; Butterworth-Heinemann Oxford: Oxford, UK, 2000.

24. Deane, B.; Wyszecki, G. Color in Business, Science and Industry; Wiley: Hoboken, NJ, USA, 1963.

25. Palczewska, G.; Vinberg, F.; Stremplewski, P.; Bircher, M.P.; Komar, K.; Zhang, J.; Cascella, M.; Wojtkowski, M.; Kefalov, V.J. Human infrared vision is triggered by two-photon chromophore isomerization. Proc. Natl. Acad. Sci. 2014, 111, E5445–E5454. [CrossRef] [PubMed]

26. Zimmer, C. Our strange, important, subconscious light detectors. Discov. Mag.. Retrieved 2012, 02–18.

27. Mann, I. The Development of the Human Eye; CUP Archive: Cambridge, UK, 1949.

28. Treuting, P.M.; Wong, R.; Tu, D.C.; Phan, I. Special senses: Eye. In Comparative Anatomy and Histology; Elsevier: Amsterdam, The Netherlands, 2012; pp. 395–418.

29. Yi, X.-j.; Wang, Y.; Fu-Shin, X.Y. Corneal epithelial tight junctions and their response to lipopolysaccharide challenge. Investig. Ophthalmol. Vis. Sci. 2000, 41, 4093–4100.

30. Cunha-Vaz, J.G. The blood-ocular barriers: Past, present, and future. Doc. Ophthalmal. 1997, 93, 149–157. [CrossRef]

31. Gipson, I.K.; Argueso, P. Role of mucins in the function of the corneal and conjunctival epithelia. Int Rev Cytol 2003, 231, 1–49. [CrossRef]

32. Gaudana, R.; Jwala, J.; Boddu, S.H.; Mitra, A.K. Recent perspectives in ocular drug delivery. Pharm. Res. 2009, 26, 1197. [CrossRef]

33. Masini, E.; Carta, F.; Scozzafava, A.; Supuran, C.T. Antiglaucoma carbonic anhydrase inhibitors: A patent review. Expert Opin. Ther. Pat. 2013, 23, 705–716. [CrossRef] [PubMed]

34. Burke, J.A. Combinations of selective alpha-adrenergic agonists and antagonists useful in lowering intraocular pressure. US Patent 5,281,591, 1994. [CrossRef]

35. Medeiros, F.A.; Weinreb, R.N. Medical backgrounders: Glaucoma. Drugs Today (Barc) 2002, 38, 563–570. [CrossRef] [PubMed]

36. Goyal, G.; Garg, T.; Rath, G.; Goyal, A.K. Current nanotechnological strategies for treating glaucoma. Crit. Rev. Ther. Drug Carr. Syst. 2014, 31. [CrossRef]

37. Pita-Thomas, D.W.; Goldberg, J.L. Nanotechnology and glaucoma: Little particles for a big disease. Curr. Opin. Ophthalmal. 2013, 24, 130–135. [CrossRef]

38. Salama, H.A.; Ghobab, M.; Mahmoud, A.A.; Hady, M.A. PLGA nanoparticles as subconjunctival injection for management of glaucoma. Aaps Pharmscitech 2017, 18, 2517–2528. [CrossRef]
44. Morsi, N.; Ibrahim, M.; Refai, H.; El Sorogy, H. Nanoemulsion-based electrolyte triggered in situ gel for ocular delivery of acetazolamide. *Eur. J. Pharm. Sci.* 2017, 104, 302–314. [CrossRef]

45. Kim, H.-J.; Zhang, K.; Moore, L.; Ho, D. Diamond nanogel-embedded contact lenses mediate lysosome-dependent therapeutic release. *Acx Nano* 2014, 8, 2998–3005. [CrossRef]

46. Tuomela, A.; Liu, P.; Puranen, J.; Rönkkö, S.; Laaksonen, T.; Kalesnykas, G.; Oksala, O.; Ilkka, J.; Laru, J.; Järvinen, K. Brinzolamide nanocrystal formulations for ophthalmic delivery: Reduction of elevated intraocular pressure in vivo. *Int. J. Pharm.* 2014, 467, 34–41. [CrossRef]

47. Natarajan, J.V.; Darwitan, A.; Barathi, V.A.; Ang, M.; Htoo, H.M.; Boey, F.; Tam, K.C.; Wong, T.T.; Venkatraman, S.S. Sustained drug release in nanomedicine: A long-acting nanocarrier-based formulation for glaucoma. *Acx Nano* 2014, 8, 419–429. [CrossRef]

48. Kambhampati, S.P.; Kannan, R.M. Dendrimer nanoparticles for ocular drug delivery. *J. Ocul. Pharmacol. Ther.* 2013, 29, 151–165. [CrossRef] [PubMed]

49. Rodrigues-Aller, M.; Guinchard, S.; Guillarme, D.; Pupier, M.; Jeannerat, D.; Rivara-Minten, E.; Veuthey, J.-L.; Gurny, R. New prostaglandin analog formulation for glaucoma treatment containing cyclodextrins for improved stability, solubility and ocular tolerance. *Eur. J. Pharm. Biopharm.* 2015, 95, 203–214. [CrossRef] [PubMed]

50. Cardigos, J.; Ferreira, Q.; Crisóstomo, S.; Moura-Coelho, N.; Cunha, J.P.; Pinto, L.A.; Ferreira, J.T. Nanotechnology-Ocular Devices for Glaucoma Treatment: A Literature Review. *Curr. Eye Res.* 2019, 44, 111–117. [CrossRef] [PubMed]

51. Liu, S.; Jones, L.; Gu, F.X. Nanomaterials for ocular drug delivery. *Macromol. Biosci.* 2012, 12, 608–620. [CrossRef] [PubMed]

52. Jin, Q.; Li, H.; Jin, Z.; Huang, L.; Wang, F.; Zhou, Y.; Liu, Y.; Jiang, C.; Oswald, J.; Wu, J. TPGS modified nanoliposomes as an effective ocular delivery system to treat glaucoma. *Int. J. Pharm.* 2018, 553, 21–28. [CrossRef] [PubMed]

53. Wadhwa, S.; Paliwal, R.; Paliwal, S.R.; Vyas, S. Hyaluronic acid modified chitosan nanoparticles for effective management of glaucoma: Development, characterization, and evaluation. *J. Drug Target.* 2010, 18, 292–302. [CrossRef]

54. Fedorchak, M.V.; Conner, I.P.; Medina, C.A.; Wingard, J.B.; Schuman, J.S.; Little, S.R. 28-day intraocular pressure reduction with a single dose of brimonidine tartrate-loaded microspheres. *Exp. Eye Res.* 2014, 125, 210–216. [CrossRef]

55. Liu, H.; Wan, J.; Hughes, P.M. Microsphere drug delivery system for sustained intraocular release. US Patent 9,504,653, 2016.

56. Bertram, J.P.; Saluja, S.S.; McKain, J.; Lavik, E.B. Sustained delivery of timolol maleate from poly(lactic-co-glycolic acid)/poly (lactic acid) microspheres for over 3 months. *J. Microencapsul.* 2009, 26, 18–26. [CrossRef]

57. Jung, H.J.; Abou-Jaoude, M.; Caribia, B.E.; Plummer, C.; Chauhan, A. Glaucoma therapy by extended release of timolol from nanoparticle loaded silicone-hydrogel contact lenses. *J. Control. Release* 2013, 165, 82–89. [CrossRef]

58. Peng, C.-C.; Burke, M.T.; Caribia, B.E.; Plummer, C.; Chauhan, A. Extended drug delivery by contact lenses for glaucoma therapy. *J. Control. Release* 2012, 162, 152–158. [CrossRef]

59. Garg, T.; Malik, B.; Rath, G.; Goyal, A.K. Development and characterization of nano-fiber patch for the treatment of glaucoma. *Eur. J. Pharm. Sci.* 2014, 53, 10–16.

60. Natarajan, J.V.; Ang, M.; Darwitan, A.; Chattopadhyay, S.; Wong, T.T.; Venkatraman, S.S. Nanomedicine for glaucoma: Liposomes provide sustained release of latanoprost in the eye. *Int. J. Nanomed.* 2012, 7, 123.

61. Wong, T.T.; Novack, G.D.; Natarajan, J.V.; Ho, C.L.; Htoo, H.M.; Venkatraman, S.S. Nanomedicine for glaucoma: Sustained release latanoprost offers a new therapeutic option with substantial benefits over eye drops. *Drug Deliv. Transl. Res.* 2014, 4, 303–309. [CrossRef] [PubMed]

62. Vold, S.; Ahmed, I.I.K.; Craven, E.R.; Mattox, C.; Stamper, R.; Packer, M.; Brown, R.H.; Ianchulev, T.; Group, C.S. Two-year COMPASS trial results: Supraciliary microstenting with phacoemulsification in patients with open-angle glaucoma and cataracts. *Ophthalmology* 2016, 123, 2103–2112. [CrossRef] [PubMed]

63. Sheybani, A.; Dick, H.B.; Ahmed, I.I. Early clinical results of a novel ab interno gel stent for the surgical treatment of open-angle glaucoma. *J. Glaucoma* 2016, 25, e691–e696. [CrossRef] [PubMed]
64. Loewen, R.T.; Roy, P.; Parikh, H.A.; Dang, Y.; Schuman, J.S.; Loewen, N.A. Impact of a glaucoma severity index on results of trabeculectomy surgery: Larger pressure reduction in more severe glaucoma. Plos One 2016, 11, e0151926. [CrossRef] [PubMed]

65. Wu, W.; Li, J.; Wu, L.; Wang, B.; Wang, Z.; Xu, Q.; Xin, H. Ophthalmic delivery of brinzolamide by liquid crystalline nanoparticles: In vitro and in vivo evaluation. Aaps PharmSciTech 2015, 14, 1063–1071. [CrossRef]

66. Han, Q.; Wang, Y.; Li, X.; Peng, R.; Li, A.; Qian, Z.; Yu, L. Effects of bevacizumab loaded PEG-PCL-PEG hydrogel intracameral application on intraocular pressure after glaucoma filtration surgery. J. Mater. Sci. Mater. Med. 2015, 26, 225. [CrossRef]

67. Liao, Y.-T.; Lee, C.-H.; Chen, S.-T.; Lai, J.-Y.; Wu, K.C.-W. Gelatin-functionalized mesoporous silica nanoparticles as a novel controlled-release topical ophthalmic delivery system for the treatment of glaucoma. Int. J. Nanomed. 2018, 13, 3975. [CrossRef] [PubMed]

68. Woodward, D.F.; Gil, D.W. The inflow and outflow of anti-glaucoma drugs. Trends Pharmacol. Sci. 2004, 25, 238–241. [CrossRef] [PubMed]

69. Szumny, D.; Szelag, A. The influence of new beta-adrenolytics nebivolol and carvedilol on intraocular pressure and iris blood flow in rabbits. Graefe’s Arch. Clin. Exp. Ophthalmol. 2014, 252, 917–923. [CrossRef] [PubMed]

70. Jiang, M.; Gan, L.; Zhu, C.; Dong, Y.; Liu, J.; Gan, Y. Cationic core–shell liponanoparticles for ocular gene delivery. Biomaterials 2012, 33, 7621–7630. [CrossRef] [PubMed]

71. Li, J.; Tian, S.; Tao, Q.; Zhao, Y.; Gui, R.; Yang, F.; Zang, L.; Chen, Y.; Ping, Q.; Hou, D. Montmorillonite nanoparticles with sustained release properties for intracameral pharmacotherapy of glaucoma. J. Mater. Chem. B 2017, 5, 7008–7013. [CrossRef]

72. Hassan, D.; Abdelmonem, R.; Abdellatif, M. Formulation and Characterization of Carvedilol Leciplex for Glaucoma Treatment: In-Vitro, Ex-Vivo and In-Vivo Study. Pharmaceutics 2018, 10, 197. [CrossRef]

73. Yuan, X.; Marcano, D.C.; Shin, C.S.; Hua, X.; Isenhart, L.C.; Pflugfelder, S.C.; Acharya, G. Ocular drug delivery nanowafer with enhanced therapeutic efficacy. Adv. Healthc. Mater. 2015, 4, 714–718. [CrossRef] [PubMed]

74. Shi, S.; Zhang, Z.; Luo, Z.; Yu, J.; Liang, R.; Li, X.; Chen, H. Chitosan grafted methoxy poly (ε-caprolactone) nanosuspension for ocular delivery of hydrophobic diclofenac. Sci. Rep. 2015, 5, 11337. [CrossRef]

75. Colin, J. Ganciclovir ophthalmic gel, 0.15%: A valuable tool for treating ocular herpes. Clin. Ophthalmol. (Auckl. Nz) 2007, 1, 441.

76. Foster, C.S. Ganciclovir gel—a new topical treatment for herpetic keratitis. Ganciclovir Gel—A New Top. Treat. Herpetic Keratitis 2013. [CrossRef]

77. Kaufman, H.E.; Haw, W.H. Ganciclovir ophthalmic gel 0.15%: Safety and efficacy of a new treatment for herpes simplex keratitis. Curr. Eye Res. 2012, 37, 654–660. [CrossRef]

78. Chou, T.Y.; Hong, B.Y. Ganciclovir ophthalmic gel 0.15% for the treatment of acute herpetic keratitis: Background, effectiveness, tolerability, safety, and future applications. Ther. Clin. Risk Manag. 2014, 10, 665. [CrossRef]

79. Vega, E.; Egea, M.; Valls, O.; Espina, M.; Garcia, M. Flurbiprofen loaded biodegradable nanoparticles for ophthalmic administration. J. Pharm. Sci. 2006, 95, 2393–2405. [CrossRef]

80. Pan, Q.; Xu, Q.; Boylan, N.J.; Lamb, N.W.; Emmert, D.G.; Yang, J.-C.; Tang, L.; Hefflin, T.; Alwadani, S.; Eberhart, C.G. Corticosteroid-loaded biodegradable nanoparticles for prevention of corneal allograft rejection in rats. J. Control. Release 2015, 201, 32–40. [CrossRef]

81. Zhang, W.; Wang, Y.; Lee, B.T.K.; Liu, C.; Wei, G.; Lu, W. A novel nanoscale-dispersed eye ointment for the treatment of dry eye disease. Nanotechnology 2014, 25, 125101. [CrossRef]

82. Pignatello, R.; Buocolo, C.; Spedaleri, G.; Maltese, A.; Puglisi, G. Flurbiprofen-loaded acrylic polymer nanosuspensions for ophthalmic application. Biomaterials 2002, 23, 3247–3255. [CrossRef]

83. Zhang, Z.; He, Z.; Liang, R.; Ma, Y.; Huang, W.; Jiang, R.; Shi, S.; Chen, H.; Li, X. Fabrication of a micellar supramolecular hydrogel for ocular drug delivery. Biomacromolecules 2016, 17, 798–807. [CrossRef]

84. Zhang, J.; Wang, S. Topical use of Coenzyme Q10-loaded liposomes coated with trimethyl chitosan: Tolerance, precorneal retention and anti-cataract effect. Int. J. Pharm. 2009, 372, 66–75. [CrossRef]
86. Tyagi, P.; Barros, M.; Stansbury, J.W.; Kompella, U.B. Light-activated, in situ forming gel for sustained suprachoroidal delivery of bevacizumab. Mol. Pharm. 2013, 10, 2858–2867. [CrossRef]
87. Shedden, A.H.; Laurence, J.; Barrish, A.; Olah, T.V. Plasma timolol concentrations of timolol maleate: Timolol gel-forming solution (TIMOPTIC-XE®) once daily versus timolol maleate ophthalmic solution twice daily. Doc. Ophthalmol. 2001, 103, 73–79. [CrossRef]
88. ElShaer, A.; Mustafa, S.; Kasar, M.; Thapa, S.; Ghatora, B.; Alany, R. Nanoparticle-laden contact lens for controlled ocular delivery of prednisolone: Formulation optimization using statistical experimental design. Pharmaceutics 2016, 8, 14. [CrossRef]
89. Chen, C.-W.; Yeh, M.-K.; Shiau, C.-Y.; Chiang, C.-H.; Lu, D.-W. Efficient downregulation of VEGF in retinal pigment epithelial cells by integrin ligand-labeled liposome-mediated siRNA delivery. Int. J. Nanomed. 2013, 8, 2613.
90. Chen, C.-W.; Lu, D.-W.; Yeh, M.-K.; Shiau, C.-Y.; Chiang, C.-H. Novel RGD-lipid conjugate-modified liposomes for enhancing siRNA delivery in human retinal pigment epithelial cells. Int. J. Nanomed. 2011, 6, 2567. [CrossRef]
91. Mastorakos, P.; Kambhampati, S.P.; Mishra, M.K.; Wu, T.; Song, E.; Hanes, J.; Kannan, R.M. Hydroxyl PAMAM dendrimer-based gene vectors for transgene delivery to human retinal pigment epithelial cells. Nanoscale 2015, 7, 3845–3856. [CrossRef]
92. Read, S.P.; Cashman, S.M.; Kumar-Singh, R. POD nanoparticles expressing GDNF provide structural and functional rescue of light-induced retinal degeneration in an adult mouse. Mol. Ther. 2010, 18, 1917–1926. [CrossRef]
93. Rajala, A.; Wang, Y.; Zhu, Y.; Ranjo-Bishop, M.; Ma, J.; Mao, C.; Rajala, R.V. Nanoparticle-assisted targeted delivery of eye-specific genes to eyes significantly improves the vision of blind mice in vivo. Nano Lett. 2014, 14, 5257–5263. [CrossRef]
94. Mitrev, P. Glaucoma drainage device implant. US Patent 6,589,203, 2003.
95. Coleman, A.L.; Hill, R.; Wilson, M.R.; Choplin, N.; Kotas-Neumann, R.; Tam, M.; Bacharach, J.; Panek, W.C. Initial clinical experience with the Ahmed glaucoma valve implant. Am. J. Ophthalmol. 1995, 120, 23–31. [CrossRef]
96. Syed, H.M.; Law, S.K.; Nam, S.H.; Li, G.; Caprioli, J.; Coleman, A. Baerveldt-350 implant versus Ahmed valve for refractory glaucoma: A case-controlled comparison. J. Glaucoma 2004, 13, 38–45. [CrossRef]
97. Yalvac, I.; Eksioglu, U.; Satana, B.; Duman, S. Long-term results of Ahmed glaucoma valve and Molteno implant in neovascular glaucoma. Eye 2007, 21, 65. [CrossRef]
98. Hong, C.-H.; Arosemena, A.; Zurakowski, D.; Ayyala, R.S. Glaucoma drainage devices: A systematic literature review and current controversies. Surv. Ophthalmol. 2005, 50, 48–60. [CrossRef]
99. Alvarado, J.A.; Hollander, D.A.; Juster, R.P.; Lee, L.C. Ahmed valve implantation with adjunctive mitomycin C and 5-fluorouracil: Long-term outcomes. Am. J. Ophthalmol. 2008, 146, 276–284.e272. [CrossRef]
100. Ponnusamy, T.; Yu, H.; John, V.T.; Ayyala, R.S.; Blake, D.A. A novel antiproliferative drug coating for glaucoma drainage devices. J. Glaucoma 2014, 23, 526–534. [CrossRef] [PubMed]
101. Pan, T.; Brown, J.D.; Ziaie, B. An artificial nano-drainage implant (ANDI) for glaucoma treatment. In Proceedings of the 2006 International Conference of the IEEE Engineering in Medicine and Biology Society, New York, NY, USA, 30 August–3 September 2006; pp. 3174–3177.
102. Paschalis, E.I.; Chodosh, J.; Sperling, R.A.; Salvador-Culla, B.; Dohlman, C. A novel implantable glaucoma valve using ferrofluid. Plos One 2013, 8, e67404. [CrossRef] [PubMed]
103. Maleki, T.; Chitinis, G.; Park, J.H.; Cantor, L.B.; Ziaie, B. Biodegradable microfabricated plug-filters for glaucoma drainage devices. Ieee Trans. Biomed. Eng. 2012, 59, 1507–1513. [CrossRef]
104. Harake, R.S.; Ding, Y.; Brown, J.D.; Pan, T. Design, fabrication, and in vitro testing of an anti-biofouling glaucoma micro-shunt. Ann. Biomed. Eng. 2015, 43, 2394–2405. [CrossRef]
105. Leonardi, M.; Pitchon, E.M.; Bertsch, A.; Renaud, P.; Mermoud, A. Wireless contact lens sensor for intraocular pressure monitoring: Assessment on enucleated pig eyes. Acta Ophthalmol. 2009, 87, 433–437. [CrossRef]
106. Fasching, C.; Mossböck, G. Continuous 24 h monitoring of changes in intraocular pressure with the wireless contact lens sensor Triggerfish™. First results in patients. Der Ophthalmol. Z. Der Dtsch. Ophthalmol. Ges. 2010, 107, 918–922. [CrossRef]
108. Lee, G.A.; Khaw, P.T.; Ficker, L.A.; Shah, P. The corneal thickness and intraocular pressure story: Where are we now? *Clin. Exp. Ophthalmol.* 2002, 30, 334–337. [CrossRef]

109. Graham, S.; Drance, S. Nocturnal hypotension: Role in glaucoma progression. *Surv. Ophthalmol.* 1999, 43, S10–S16. [CrossRef]

110. Irazoqui, P.P.; Chow, E.Y.; Chappell, W.J.; Yang, C.-I.; Ziaie, B. Intra-ocular pressure sensor. U.S. Patent 5,281,591, 2013.

111. Tanihara, H.; Inoue, T.; Yamamoto, T.; Kuwayama, Y.; Abe, H.; Suganami, H.; Arai, M.; Group, K.C.S. Intra-ocular pressure-lowering effects of a Rho kinase inhibitor, ripasudil (K-115), over 24 hours in primary open-angle glaucoma and ocular hypertension: A randomized, open-label, crossover study. *Acta Ophthalmol.* 2015, 93, e254–e260. [CrossRef]

112. Chen, G.-Z.; Chan, I.-S.; Leung, L.K.; Lam, D.C. Soft wearable contact lens sensor for continuous intraocular pressure monitoring. *Med Eng. Phys.* 2014, 36, 1134–1139. [CrossRef] [PubMed]

113. Rizq, R.N.; Choi, W.-H.; Eilers, D.; Wright, M.M.; Ziaie, B. Intraocular pressure measurement at the choroid surface: A feasibility study with implications for implantable microsystems. *Br. J. Ophthalmol.* 2001, 85, 868–871. [CrossRef] [PubMed]

114. Dresher, R.P.; Irazoqui, P.P. A compact nanopower low output impedance CMOS operational amplifier for wireless intraocular pressure recordings. In Proceedings of the 2007 29th Annual International Conference of the IEEE Engineering in Medicine and Biology Society, 22–26 August 2007, Lyon, France; pp. 6055–6058.

115. Chen, P.-J.; Rodger, D.C.; Saati, S.; Humayun, M.S.; Tai, Y.-C. Microfabricated implantable parylene-based wireless passive intraocular pressure sensors. *J. Micromechanical Syst.* 2008, 17, 1342–1351. [CrossRef]

116. Chen, P.-J.; Rodger, D.C.; Humayun, M.S.; Tai, Y.-C. Unpowered spiral-tube parylene pressure sensor for intraocular pressure sensing. *Sens. Actuators A Phys.* 2006, 127, 276–282. [CrossRef]

117. Chen, P.-J.; Rodger, D.C.; Agrawal, R.; Saati, S.; Meng, E.; Varma, R.; Humayun, M.S.; Tai, Y.-C. Implantable micromechanical parylene-based pressure sensors for unpowered intraocular pressure sensing. *J. Micromechanics Microengineering* 2007, 17, 1931. [CrossRef]

118. Phan, A.; Truong, P.; Camp, A.; Stewart, K.; Suen, B.; Weinreb, R.N.; Talke, F.E. A Wireless Handheld Pressure Measurement System for in vivo Monitoring of Intraocular Pressure in Rabbits. *Ieee Trans. Biomed. Eng.* 2019. [CrossRef]

119. Zilly, J.; Buhmann, J.M.; Mahapatra, D. Glaucoma detection using entropy sampling and ensemble learning for automatic optic cup and disc segmentation. *Comput. Med Imaging Graph.* 2017, 55, 28–41. [CrossRef]

120. Morgan, J.E.; Sheen, N.J.L.; North, R.V.; Choong, Y.; Ansari, E. Digital imaging of the optic nerve head: Monoscopic and stereoscopic analysis. *Br. J. Ophthalmol.* 2005, 89, 879–884. [CrossRef]

121. Fallon, M.; Valero, O.; Pazos, M.; Anton, A. Diagnostic accuracy of imaging devices in glaucoma: A meta-analysis. *Surv. Ophthalmol.* 2017, 62, 446–461. [CrossRef]

122. Jia, Y.; Wei, E.; Wang, X.; Zhang, X.; Morrison, J.C.; Parikh, M.; Lombardi, L.H.; Gatley, D.M.; Armour, R.L.; Edmunds, B. Optical coherence tomography angiography of optic disc perfusion in glaucoma. *Ophthalmology* 2014, 121, 1322–1332. [CrossRef]

123. Liu, L.; Jia, Y.; Takusagawa, H.L.; Pechauer, A.D.; Edmunds, B.; Lombardi, L.; Davis, E.; Morrison, J.C.; Huang, D. Optical coherence tomography angiography of the peripapillary retina in glaucoma. *Jama Ophthalmol.* 2015, 133, 1045–1052. [CrossRef] [PubMed]

124. Wang, X.; Jiang, C.; Ko, T.; Kong, X.; Yu, X.; Min, W.; Shi, G.; Sun, X. Correlation between optic disc perfusion and glaucomatous severity in patients with open-angle glaucoma: An optical coherence tomography angiography study. *Graefes Arch. Clin. Exp. Ophthalmol.* 2015, 253, 1557–1564. [CrossRef] [PubMed]

125. Bussel, I.I.; Wollstein, G.; Schuman, J.S. OCT for glaucoma diagnosis, screening and detection of glaucoma progression. *Br. J. Ophthalmol.* 2014, 98, ii15–ii19. [CrossRef] [PubMed]

126. Strouthidis, N.G.; Scott, A.; Peter, N.M.; Garway-Heath, D.F. Optic disc and visual field progression in ocular hypertensive subjects: Detection rates, specificity, and agreement. *Investig. Ophthalmol. Vis. Sci.* 2006, 47, 2904–2910. [CrossRef]

127. Chiang, B.; Kim, Y.; Doty, A.; Grossniklaus, H.; Schwendeman, S.; Prausnitz, M. Sustained reduction of intraocular pressure by supraciliary delivery of brimonidine-loaded poly (lactic acid) microspheres for the treatment of glaucoma. *J. Control. Release* 2016, 228, 48–57. [CrossRef]
149. Anderson, S.A.; Rader, R.K.; Westlin, W.F.; Null, C.; Jackson, D.; Lanza, G.M.; Wickline, S.A.; Kotyk, J.J. Magnetic resonance contrast enhancement of neovasculature with αvβ3-targeted nanoparticles. Magn. Reson. Med. Off. J. Int. Soc. Magn. Reson. Med. 2000, 44, 433–439. [CrossRef]

150. Cang, H.; Sun, T.; Li, Z.-Y.; Chen, J.; Wiley, B.J.; Xia, Y.; Li, X. Gold nanocages as contrast agents for spectroscopic optical coherence tomography. Opt. Lett. 2005, 30, 3048–3050. [CrossRef]

151. Akerman, M.; Chan, W.; Laakkonen, P.; Bhatia, S.; Ruoslahti, E. Nanocrystal targeting in vivo Proc Natl Acad Sci USA 99: 12617–12621. Find This Artic. Online 2002.

152. Schachar, R.A.; Chen, W.; Woo, B.K.; Pierscionek, B.K.; Zhang, X.; Ma, L. Diffusion of nanoparticles into the capsule and cortex of a crystalline lens. Nanotechnology 2007, 19, 025102. [CrossRef]

153. Saccà, S.C.; Pascotto, A.; Camicione, P.; Capris, P.; Izzotti, A. Oxidative DNA damage in the human trabecular meshwork: Clinical correlation in patients with primary open-angle glaucoma. Arch. Ophthalmol. 2005, 123, 458–463. [CrossRef] [PubMed]

154. Acott, T.S.; Kelley, M.J. Extracellular matrix in the trabecular meshwork. Exp. Eye Res. 2008, 86, 543–561. [CrossRef] [PubMed]

155. Vranka, J.A.; Kelley, M.J.; Acott, T.S.; Keller, K.E. Extracellular matrix in the trabecular meshwork: Intraocular pressure regulation and dysregulation in glaucoma. Exp. Eye Res. 2015, 133, 112–125. [CrossRef] [PubMed]

156. Borrás, T. Advances in glaucoma treatment and management: Gene therapy. Invest. Ophthalmol. Vis. Sci. 2012, 53, 2506–2510. [CrossRef] [PubMed]

157. Nakajima, E.; Nakajima, T.; Minagawa, Y.; Shearer, T.R.; Azuma, M. Contribution of ROCK in contraction of trabecular meshwork: Proposed mechanism for regulating aqueous outflow in monkey and human eyes. J. Pharm. Sci. 2005, 94, 701–708. [CrossRef] [PubMed]

158. De Groef, L.; Van Hove, I.; Dekeyster, E.; Stalmans, I.; Moons, L. MMPs in the trabecular meshwork: Promising targets for future glaucoma therapies? Investig. Ophthalmol. Vis. Sci. 2013, 54, 7756–7763. [CrossRef] [PubMed]

159. Gerometta, R.; Spiga, M.-G.; Borrás, T.; Candia, O.A. Treatment of sheep steroid–induced ocular hypertension with a glucocorticoid-inducible MMP1 gene therapy virus. Invest. Ophthalmol. Vis. Sci. 2010, 51, 3042–3048. [CrossRef]

160. Du, Y.; Yun, H.; Yang, E.; Schuman, J.S. Stem cells from trabecular meshwork home to TM tissue in vivo. Investig. Ophthalmol. Vis. Sci. 2013, 54, 1450–1459. [CrossRef] [PubMed]

161. Dautriche, C.N.; Szymanski, D.; Kerr, M.; Torrejon, K.Y.; Bergkvist, M.; Xie, Y.; Danias, J.; Stamer, W.; Sharfstein, S.T. A biomimetic Schlemm’s canal inner wall: A model to study outflow physiology, glaucoma pathology and high-throughput drug screening. Biomaterials 2015, 65, 86–92. [CrossRef]

162. Torrejon, K.Y.; Pu, D.; Bergkvist, M.; Danias, J.; Sharfstein, S.T.; Xie, Y. Recreating a human trabecular meshwork outflow system on microfabricated porous structures. Biotechnol. Bioeng. 2013, 110, 3205–3218. [CrossRef]

163. Bergkvist, M.; Brenner, S.; Danias, J.; Sharfstein, S.; Xie, Y.; Gracias, A.; Torrejon, K. Bioengineered human trabecular meshwork for biological applications. U.S. Patent 9,506,907, 2016.

164. Xie, Y.; Torrejon, K.; Sharfstein, S.; Bergkvist, M.; Danias, J.; Dautriche, C.N. Use of vascular cells to create the conventional outflow tract. US Patent 16/044,806, 2019.

165. Snider, E.; Kubelick, K.; Tweed, K.; Kim, R.; Li, Y.; Gao, K.; Read, A.; Emelianov, S.; Ethier, C. Improving Stem Cell Delivery to the Trabecular Meshwork Using Magnetic Nanoparticles. Sci. Rep. 2018, 8, 12251. [CrossRef] [PubMed]

166. Ikuta, Y.; Aoyagi, S.; Tanaka, Y.; Sato, K.; Inada, S.; Koseki, Y.; Onodera, T.; Oikawa, H.; Kasai, H. Creation of nano eye-drops and effective drug delivery to the interior of the eye. Sci. Rep. 2017, 7, 44229. [CrossRef] [PubMed]

167. Alqawlaq, S.; Huzil, J.T.; Ivanova, M.V.; Foldvari, M. Challenges in neuroprotective nanomedicine development: Progress towards noninvasive gene therapy of glaucoma. Nanomedicine 2012, 7, 1067–1083. [CrossRef] [PubMed]

168. Yadav, K.S.; Rajpurohit, R.; Sharma, S. Glaucoma: Current treatment and impact of advanced drug delivery systems. Life Sci. 2019. [CrossRef] [PubMed]

169. Prow, T.W. Toxicity of nanomaterials to the eye. Wiley Interdisciplinary Rev. Nanomed. Nanobiotechnol. 2010, 2, 317–333. [CrossRef] [PubMed]
170. Sutariya, V.B.; Groshev, A.; Dave, V.S.; Saluja, H.; Bhatia, D.; Sadana, P.; Pathak, Y. Nanoparticles and human health: A review of epidemiological studies. *Biointeractions Nanomater.* 2014, 175–208.

171. Albanese, A.; Tang, P.S.; Chan, W.C. The effect of nanoparticle size, shape, and surface chemistry on biological systems. *Annu. Rev. Biomed. Eng.* 2012, 14, 1–16. [CrossRef]

172. del Pozo-Rodriguez, A.; Delgado, D.; Solinis, M.; Gascon, A.; Pedraz, J. Solid lipid nanoparticles for retinal gene therapy: Transfection and intracellular trafficking in RPE cells. *Int. J. Pharm.* 2008, 360, 177–183. [CrossRef]

173. Calvo, P.; Vila-Jato, J.L.; Alonso, M.J. Comparative in vitro evaluation of several colloidal systems, nanoparticles, nanocapsules, and nanoemulsions, as ocular drug carriers. *J. Pharm. Sci.* 1996, 85, 530–536. [CrossRef]

174. Vadlapudi, A.D.; Mitra, A.K. Nanomicelles: An emerging platform for drug delivery to the eye. *Ther. Deliv.* 2013, 4, 1–3. [CrossRef]

175. Araújo, J.; Gonzalez, E.; Egea, M.A.; Garcia, M.L.; Souto, E.B. Nanomedicines for ocular NSAIDs: Safety on drug delivery. *Nanomed. Nanotechnol. Biol. Med.* 2009, 5, 394–401.

176. Marmor, M.F.; Negi, A.; Maurice, D.M. Kinetics of macromolecules injected into the subretinal space. *Exp. Eye Res.* 1985, 40, 687–696. [CrossRef]

177. Amrite, A.C.; Kompella, U.B. Size-dependent disposition of nanoparticles and microparticles following subconjunctival administration. *J. Pharm. Pharmacol.* 2005, 57, 1555–1563. [CrossRef] [PubMed]

178. Cheruvu, N.P.; Amrite, A.C.; Kompella, U.B. Effect of eye pigmentation on transscleral drug delivery. *Investig. Ophthalmol. Vis. Sci.* 2008, 49, 333–341. [CrossRef] [PubMed]

179. Tezel, G. A proteomics view of the molecular mechanisms and biomarkers of glaucomatous neurodegeneration. *Prog. Retin. Eye Res.* 2013, 35, 18–43. [CrossRef]

180. Sanvicens, N.; Marco, M.P. Multifunctional nanoparticles—properties and prospects for their use in human medicine. *Trends Biotechnol.* 2008, 26, 425–433. [CrossRef]

181. Sahoo, S.K.; Misra, R.; Parveen, S. Nanoparticles: A boon to drug delivery, therapeutics, diagnostics and imaging. In *Nanomedicine in Cancer*; Pan Stanford: Stanford, CA, USA, 2017; pp. 73–124.

182. Crommelin, D.J.; Florence, A.T. Towards more effective advanced drug delivery systems. *Int. J. Pharm.* 2013, 454, 496–511. [CrossRef]

183. Smith, B.R.; Gambhir, S.S. Nanomaterials for in vivo imaging. *Chem. Rev.* 2017, 117, 901–986. [CrossRef]

184. Osborne, N.N. Recent clinical findings with memantine should not mean that the idea of neuroprotection in glaucoma is abandoned. *Acta Ophthalmol.* 2009, 87, 450–454. [CrossRef]

185. Moroi, S.E.; Raoof, D.A.; Reed, D.M.; Zöllner, S.; Qin, Z.; Richards, J.E. Progress toward personalized medicine for glaucoma. *Expert Rev. Ophthalmol.* 2009, 4, 145–161. [CrossRef]