Biomarker Discovery in Human Prostate Cancer: an Update in Metabolomics Studies

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Abstract

Prostate cancer (PCa) is the most frequently diagnosed cancer and the second leading cause of cancer death among men in Western countries. Current screening techniques are based on the measurement of serum prostate specific antigen (PSA) levels and digital rectal examination. A decisive diagnosis of PCa is based on prostate biopsies; however, this approach can lead to false-positive and false-negative results. Therefore, it is important to discover new biomarkers for the diagnosis of PCa, preferably noninvasive ones. Metabolomics is an approach that allows the analysis of the entire metabolic profile of a biological system. As neoplastic cells have a unique metabolic phenotype related to cancer development and progression, the identification of dysfunctional metabolic pathways using metabolomics can be used to discover cancer biomarkers and therapeutic targets. In this study, we review several metabolomics studies performed in prostatic fluid, blood plasma/serum, urine, tissues and immortalized cultured cell lines with the objective of discovering alterations in the metabolic phenotype of PCa and thus discovering new biomarkers for the diagnosis of PCa. Encouraging results using metabolomics have been reported for PCa, with sarcosine being one of the most promising biomarkers identified to date. However, the use of sarcosine as a PCa biomarker in the clinic remains a controversial issue within the scientific community. Beyond sarcosine, other metabolites are considered to be biomarkers for PCa, but they still need clinical validation. Despite the lack of metabolomics biomarkers reaching clinical practice, metabolomics proved to be a powerful tool in the discovery of new biomarkers for PCa detection.

Introduction

Systems biology applied to cancer research encompasses the “omics” tools, including genomics, transcriptomics, proteomics, and metabolomics, which complement each other and are capable of measuring changes in several entities (genes, transcripts, proteins, or metabolites, respectively) simultaneously, providing an overview of various physiological or pathological conditions [1–3].

Metabolomics can provide an idea of the physiological status of a biological system, and therefore, alterations in the “normal” metabolome may be indicative of disease. These alterations in the “normal” metabolome have the potential to deliver new diagnostic markers for the detection and prognosis of diseases and to monitor the response to therapeutic interventions [4]. Metabolomics also has the potential to give new understanding of the phenotypic changes resultant from genetic alterations, environmental influence, and toxicological influence [5].

The term metabolomics includes the assessment of all the endogenous metabolites produced by the organism including small molecule intermediates and end products of biochemical reactions in
a cell (approximately ≤1500 Da), as well as exogenous metabolites, such as drugs, products from the body flora, and food. Instead of genes and proteins that participate in these biological processes, the metabolites produced are indicators of what is happening in the metabolism of a cell in physiological or pathophysiological conditions. Thus, metabolites can be altered in such diseases as cancer [1,6–9].

As neoplastic cells have a unique metabolic phenotype related to cancer development and progression, the identification of dysfunctional metabolic pathways through metabolomics can be used to identify cancer biomarkers and discover therapeutic targets [5,6,10].

**Prostate Cancer**

Prostate cancer (PCa) is the second most diagnosed cancer in men [11], principally affecting men over 50 years old [12], and is the fifth leading cause of cancer-related deaths in men worldwide [11]. Statistically, in 25% of men worldwide with PCa that develop metastatic disease [13], the bones are the principal targets of PCa metastasis [14]. Given that PCa has a long latency period and is potentially curable, it is essential to develop efficient and precise screening methods for its early detection and characterization [12].

The quantification of prostate serum antigen (PSA) and the digital rectal examination are the most common screening techniques used for PCa diagnosis. However, performing a prostate biopsy is mandatory for a final diagnosis [12]. Serum PSA levels higher than 4.0 ng/ml are a sign of PCa [14], although PSA is not able to differentiate patients with aggressive PCa from those with indolent disease [15]. The value of PSA screening is also controversial because of its limited sensitivity and specificity [1,16]. Recent studies suggested that certain PCa patients may present with PSA levels below 4.0 ng/ml [14]. This fact leads to false negatives, as no reliable cutoff values exist to demonstrate the unequivocal presence of PCa [16,17]. Furthermore, PSA levels may be affected by several other factors, such as age, prostatitis, urinary tract infection, and benign prostate hyperplasia, leading inevitably to false-positive results [14,16,18,19].

The biopsy analysis can also provide false-negative results when the tumor is small; when the cancer cells are distributed heterogeneously; and in early PCa stage when, histologically, the tumor appears benign [20–22]. Thus, samples obtained during the biopsy for histopathologic analysis may not be representative of the cancer [23].

The lack of a consistent biomarker for PCa diagnosis and monitoring highlights the need for novel, specific, sensitive, and cost-effective biomarkers to implement the best treatment approach in a precocious state of the disease [14].

**Altered Metabolism of PCa Cells**

In 1920, Otto Warburg discovered that cancer cells, unlike nonmalignant cells, preferentially produce ATP through the glycolytic pathway (anaerobic pathway) instead of the Krebs cycle, even in the presence of oxygen. This capability of cancer cells to sustain high rates of glycolysis for ATP generation is known as the Warburg effect [24–26].

Despite the relevance of the Warburg effect in cancer cells, the Krebs cycle and oxidative phosphorylation also play an important role in many types of cancer, including PCa. Recent evidence suggests that increased citrate oxidation is an important metabolic characteristic in PCa that supports the high cellular energy demand [27]. One of the major functions of prostate cells is the production of citrate, PSA, and polyamines, such as spermine, which are the major components of prostatic fluid. Therefore, prostate cells have a distinct metabolic profile as they produce specific compounds [1,16]. The production of citrate by prostate cells is very high in comparison with other organs [28]. Unlike other cells in the organism, prostate cell metabolism significantly favors citrate synthesis over citrate utilization, which makes the prostate peripheral zone epithelium unique among human cells [29]. Usually, cells degrade citrate in aerobic ATP production, with citrate being oxidized during the Krebs cycle as part of the intermediary metabolism of glucose. However, nonmalignant prostate cells accumulate and secrete citrate. The oxidation of citrate is catalyzed by mitochondrial aconitase (m-aconitase).

In normal prostate cells, m-aconitase is inhibited by the high intracellular concentrations of zinc, leading to citrate accumulation (Figure 1) [28,29].

Extensive metabolic alterations occur when prostate cells experience neoplastic transformation. One of the most relevant alterations is citrate oxidation, because cancer cells are unable to accumulate zinc, and without elevated levels of zinc, m-aconitase is no longer inhibited and can catalyze citrate oxidation [2,4,27,28,30]. This transformation of citrate accumulation in healthy prostate cells to oxidized citrate in malignant prostate cells results in more efficient energy production. This is probably an early event in the progression to malignancy and precedes the histopathological identification of malignant cells [27,31,32].

For citrate synthesis, oxaloacetate and acetyl-coenzyme A (acetyl-CoA) are essential, but whereas oxaloacetate is regenerated in the Krebs cycle, acetyl-CoA is consumed. To ensure that cancer cells have the needed energy for rapid proliferation, it is necessary to maintain elevated rates of citrate oxidation, and thus, the availability of acetyl-CoA is required. Some studies suggested that to maintain this accelerated citrate oxidation, alterations in fatty acid metabolism are needed to provide both ATP and acetyl-CoA [27,33,34] (Figure 1).

Beyond the Krebs cycle and glycolysis, glucose also can be degraded by the pentose phosphate pathway. This metabolic pathway provides NADPH and ribose-5-phosphate (important for the synthesis of nucleic acids and nucleotides), thus promoting anabolic reactions and redox homeostasis. In a recent study, Tsouko et al. (2014) demonstrated that androgen receptor (AR) signaling augmented the levels of glucose-6-phosphate dehydrogenase (G6PD) (key enzyme for pentose phosphate pathway). NADPH, and ribose synthesis in hormone-sensitive PCa cells and castrate-resistant PCa (CRPC) cells. After inhibition of mammalian target of rapamycin with rapamycin, the upregulation of G6PD is abolished. Hence, these studies revealed a relationship between the upregulation of G6PD via AR and mammalian target of rapamycin. These results suggested the importance of pentose phosphate pathway for PCa growth [35].

Cell proliferation and intercellular signaling are dependent on increased lipid biosynthesis. Acetyl-CoA also plays an important role in this metabolic alteration because it is a precursor for lipogenesis and cholesterol biosynthesis and can be produced by transformation of citrate in the cytosol [1]. Sterol regulatory element-binding protein–1, an essential transcription factor for lipogenesis, is also implicated in AR transcriptional regulation. Beyond increased lipogenesis, sterol regulatory element-binding protein–1 also increased reactive oxygen species production and the expression of NAPDH oxidase, which leads to proliferation, migration, and invasion of PCa cells [36–38]. In PCa cells, the levels of choline and creatine are increased because there is an augmentation of membrane synthesis for cell proliferation [16].

Glutamine also has an important role in the maintenance of lipogenesis, as well as to provide intermediates for the Krebs cycle through glutaminolysis (where glutamine is transformed into glutamate by glutaminase and then glutamate is transformed into
α-ketoglutarate). The observation that the glutamine transporter and glutaminase are both overexpressed in PCa cells was proof of the importance of this mechanism in PCa [38–40]. The α-ketoglutarate derivative from glutamine can contribute to the formation of citrate when incorporated into the Krebs cycle (oxidation pathway); however, α-ketoglutarate can also be transformed into citrate by the reversal of the tricarboxylic acid cycle through reductive carboxylation. This alteration of oxidation to reductive carboxylation is promoted by hypoxia and leads to lipid synthesis and tumor growth [41]. The tumor-stromal interactions also have an important role in PCa development. The myofibroblastic microenvironment, formed from the interaction of cancer cells with “cancer-associated fibroblasts”, is important for the reverse Warburg effect. Cancer-associated fibroblasts in the myofibroblastic microenvironment undergo the Warburg effect, induced by epithelial cancer cells, and secrete lactate and pyruvate. The lactate and pyruvate are taken up by the PCa cells and used for the Krebs cycle, anabolic metabolism, and cell proliferation [38,42,43].

The use of urine as a sample for metabolomics studies has many advantages compared with serum: urine is easier to obtain and handle, needs less sample preparation, and has higher amounts of metabolites and a lower protein content [12,44,45]. Blood plasma/serum has some advantages compared with urine, as the diurnal variation and the intra- and intervariability are lower. However, serum and plasma are more complex matrices than urine, having a higher concentration of proteins, and sample collection is more invasive [5].

Seminal fluids, obtained by ejaculation, come from the seminal vesicles, prostate, and epididymis. Prostatic fluid is collected after prostate massage, and the composition of this biofluid is simpler than seminal fluid [46,47]. The use of seminal/prostatic fluids has some advantages compared with the use of other biofluids, as these samples are richer in prostatic metabolites because the metabolites do not need to cross blood-tissue barriers once they are naturally secreted into the seminal/prostatic fluid. Thus, seminal/prostatic fluids are less affected by confounding factors. However, these biofluids may be difficult to collect in men with erectile dysfunction, and a portion of men may have personal or ethical problems with giving these types of samples [48].

The collection of tissue samples is more invasive than the collection of other matrices; however, the use of matched malignant and normal

Metabolomics Studies in PCa Model Systems and Biological Fluids

The most common models and biological fluids used to perform metabolomics studies are tissue and cultured cell lines and human urine, serum/plasma, prostatic fluid/seminal fluid, respectively.

Figure 1. Schematic illustration of the most significantly altered metabolic pathways in PCa cells. Dashed lines = downregulated pathway; continuous line = upregulated pathway. Metabolites overexpressed in PCa cells are shown in bold. TCA, tricarboxylic acid (cycle); AAs, amino acids; DNA, deoxyribonucleic acid; GNMT, glycine N-methyltransferase; SARDH, sarcosine dehydrogenase; G6P, glucose-6-phosphate; 6P, 6-phosphate; 3PG, 3-phosphoglycerate; CoA, coenzyme A.
adjacent prostate tissue is a good strategy to reduce intraindividual variability in metabolomics studies [49].

In vitro models are increasingly used because of interindivdual variability, and difficulties enrolling patients in these models are nonexistent. In addition, cell lines have a perfectly defined cell state which allows the analysis of a targeted metabolic status [2,50,51]. However, they do not efficiently simulate the complex cell-cell and cell-matrix interactions occurring within an organism [2,52].

There are two different metabolomics approaches to discover biomarkers for cancer: the top-down approach and the bottom-up approach. Both approaches have advantages and disadvantages. The top-down approach has the advantage of starting the metabolome

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**Table 1. Metabolomic Studies Performed in Urines from PCa Patients**

| PCa Subject Group | Control Group | Analytical Platform | Statistical Methods | Total Metabolites Found/Discriminative Metabolites Found | Discriminatory Metabolites/Biomarkers | Metabolic Pathways Dysregulated | Ref. |
|-------------------|---------------|---------------------|---------------------|----------------------------------------------------------|-------------------------------------|---------------------------------|------|
| n=13 n=24 | GS-MS | Binary strings, Similarity coefficients | 91/21 | Buthanolactone, methyl vinyl ketone, N-methylformamide, N,N-dimethylalanine, N,N-dimethylpyrrolidine, N-acetylformamide, acetonitrile, acetaldehyde, acetamide, 1-methyl-piperidinedione, 1-piperidinedione, dimethylamine, pyrrole, methacrolein, N,N-dimethylalanine, 3-methyl-pyrrolidine, 2-methyl-1H-pyrrrole, 2-octanone, 3-methyl-1H-pyrrrole, 2-n-butylacrolein and methyl propyl disulfide | Sarcosine (+) | NS | [65] |
| n=59 n=51 | GS-MS | GS-MS Wilcoxon P test, hierarchical clustering, nonparametric tests | 583/34 | Sarcosine (+) | Alterations in glycine synthesis and degradation | [53] |
| n=59 n=57 | GC-MS | Nonparametric statistical tests and ROC | NS/0 | No relevant differences in sarcosine levels between patients with and without PCa | | | [61] |
| n=3 | LC-MS | NS | NS/5 | 1. Sarcosine (+) 2. Proline 3. Kynurenine (+) 4. Uric acid (+) 5. Glycerol 3-phosphate (+) | | | [55] |
| n=25 PCa patients developing biochemical recurrence n=29 PCa patients who remained recurrence-free | GC-MS | ROC | 8/2 | Sarcosine (+) Cysteine (+) (in the group developing biochemical recurrence) | | | [56] |
| n=33 n=23 | GC-MS | Nonparametric statistical tests and ROC | NS/1 | Sarcosine (+) | | | [57] |
| n=86 n=45 | GC-MS | ROC | NS/1 | Diagnostic value of sarcosine was modest; relationship with clinicopathologic parameters was not found | | | [58] |
| n=20 n=28 | GC-MS | PCA ROC | 81/5 | Dihydroxybutaric acid (+), xylonic acid (+), pyrimidine (+), ribofuranoside(+), xylonic acid (+) | | | [60] |
| n=211 n=134 | GC-MS | ROC | NS/1 | Sarcosine (+) | | | [59] |
| n=32 n=32 | LC-MS | PCA PLS-DA | 1132/15 | 1. Glycine (+), serine (+), threonine (+), alanine (+) 2. Glutamine (+), citrate (+), aconitate (+), succinate (+) 3. Sarcosine (+), sorbose (+), arabinose (+), arabinose (-), inositol (-), galactaric acid (+) 4. Carnitines (-) | | | [63] |
| n=59 n=43 | GC-MS | RF LDA | 196/4 | 1. 2,6-dimethyl-7-octen-2-ol (+), 3-octanone (-), 2-octanone (-) 2. Pentanal (+) | | | [64] |

BPH: benign prostatic hypertrophy; GS-MS: Gas chromatography–mass spectrometry; HC: healthy controls; LDA: linear discriminant analysis; LC-MS: Liquid chromatography–mass spectrometry; NS: not specified; PCA: Principal component analysis; PLS-DA: Partial least squares discriminant analysis RF: random forest; ROC: receiver-operator characteristic.

(+): levels increased in PCa; (-): levels decreased in PCa.
| PCa Subject Group | Control Group | Analytical Platform | Statistical Methods | Total Metabolites Found | Discriminative Metabolites Found | Discriminatory Metabolites/Biomarkers | Metabolic Pathways Dysregulated | Ref. |
|-------------------|---------------|---------------------|---------------------|-------------------------|-------------------------------|-------------------------------------|----------------------------------|------|
| 962 n | 1061 HC | GC-MS | Conditional logistic regression | NS/5 | Palmitic acid (+), stearic acid (−), myristic acid (+), linolenic acid (+), eicosapentaenoic acid (+) | Alteration in lipid metabolism | [70] |
| 85 HG plus 120 | 114 HC | F I A - M S / M S LC-MS/MS | ROC and logistic regression model | 112/5 | 1. Lyposphatidyl-choline (C16:0 and C18:0) (-) 2. Serotonin (−) 3. Aspartic acid (+) 4. Ornithine (−) | 1. Alteration in lipid metabolism 2. Alteration in growth inhibition and induction of apoptosis 3. Alteration in protein biosynthesis 4. Ornithine decarboxylase overexpression | [75] |
| 561 n | 1034 HC | GC-MS LC-MS | Wilcoxon signed rank tests and χ2 tests | 7/3 | Choline (+), vitamin B2 (+), methylmalonic acid (−) | 1. Alteration in lipid metabolism 2. Alteration in growth inhibition and induction of apoptosis 3. Alteration in protein biosynthesis 4. Ornithine decarboxylase overexpression | [71] |
| 134 n | 666 HC | LC-MS | ROC | 9/3 | Cystathionine (+), homocysteine (−), cysteine (−) | Alteration in membrane phospholipidic metabolism | [73] |
| 28 Serum from patients developing biochemical recurrence | 30 Serum from patients with recurrence free 5 years after prostatectomy | LC-MS GC-MS | ROC | 19/7 | Glutamine (−), alanine (−), valine (−), isoleucine (±), alanine (−), ornithine (±), lysine (−) | Alteration in free amino acid metabolism | [56] |
| 36 Fasting plasma from PCa patients 3 months after the therapy initiation | 36 Fasting plasma from PCa patients 3 months after the therapy initiation | LC-MS GC-MS | t tests | 504/56 | 1. DHEAS (−), epiandrosterone sulfate (−), androsterone sulfate (−), cortisol (−), 4-androsten-3β (−), 17β-diol diol sulfate 1 & 2 (−), 5α-androstan-3β (−) 17β-diol diol sulfate (−), pregnen-diol diol sulfate (−), pregn steroid monosulfate (−) and andro steroid monosulfate 1 & 2 (−), 2. Cholate (−), glycocholate (±), taurocholate (±), chenodeoxycholate (±), taurochenodeoxycholate (±), ursochenodeoxycholate (±), hyodeoxycholate (±), deoxycholate (±), taurochenodeoxycholate glycodeoxycholate (±), glycochenodeoxycholate (±), 7-ketochenodeoxycholate (±) glycochenodeoxycholate (±), glycolithocholate sulfate (±), glycocholate sulfate (−), and taurocholate sulfate (−), 3. Carnitines (−), ketone bodies (−), dicarboxylic acids 4. 2-hydroxybutyrate (−) and branched-chain ketone acid dehydrogenase complex products (−) | 1. Steroids metabolism 2. Bile acids and intermediates of bile acid metabolism 3. Lipid oxidation 4. Markers of insulin resistance | [78] |
| 290 n | 312 Fluorometric assay | ROC | 1/1 | Sarcosine (+) | Alterations in glucose synthesis and degradation | [66] |
| 105 n | 36 ESI-MS/MS | PCA and HCA | 390/35 | Phosphatidylcholines-lamine (−), ether-linked phosphatidylcholines-lamine (−), ether-linked phosphatidylcholines-lamine (−) | Alteration in lipid metabolism | [72] |
| 1122 n | 1112 LC-MS | ROC | NS/1 | Sarcosine (+) | Alterations in glucose synthesis and degradation | [67] |
| 25 n | 100 HC | Immunoassay | NS | 4/1 | Insulin (±) | Alteration in energetic metabolism | [76] |
| 64 n | 50 HC | LC-MS PCA | 480/49 | Azelaic acid, uric acid, tryptophan, lysoPC | Alteration in fatty acids | [69] |
evaluation using a real sample (urine or plasma) that could be used in a clinical practice. However, urine and plasma are complex biological matrices and have metabolites from different origins, and the metabolites may be more diluted. The advantage of the bottom-up approach (starts with the metabolic analyses of cell lines) is that cultured cell lines are a simpler biological system with less interference factors. In fact, studies with immortalized cultured cells are important to eliminate confounding factors, such as the age of patients, smoking habits, diet, and other diseases that influence the intervariability in plasma and urine. Nevertheless, the findings in cultured cell lines may not be directly extrapolated to the real disease as it is practically impossible to simulate complex cell–cell and cell–matrix interactions in cell cultures of PCa. Both approaches used together can be useful tools to obtain metabolic information that can discriminate the metabolic pathways in PCa cells.

Studies with Human Fluids and Model Systems

Urine Studies. Table 1 summarizes the major metabolites and metabolic pathways that have been found to be dysregulated in metabolomics studies performed on urine samples from PCa patients.

One of the most relevant metabolomics studies was performed by Seekum et al. (2009), where sarcosine (N-methylglycine) was discovered as possible biomarker in urine for PCa. Sarcosine, an intermediate product in the synthesis and degradation of glycine, was found to be highly elevated during PCa progression to metastasis and was not detected or was presented at very low concentrations in the urine of healthy individuals [53]. Carcinogenesis alters the biosynthesis of sarcosine, although the importance of sarcosine in carcinogenesis remains unknown. It is known that glycine N-methyltransferase (GNMT) has a significant role in the metabolism of PCa tissues. This enzyme catalyzes the conversion of glycine to sarcosine and also participates both in the metabolism of amino acids metabolism, lysophospholipids metabolism, and bile acids metabolism and alteration in steroid hormone biosynthesis pathway

| PCa Subject Group | Control Group | Analytical Platform | Statistical Methods | Total Metabolites Found | Discriminatory Metabolites/Biomarkers | Metabolic Pathways Dysregulated | Ref. |
|------------------|---------------|---------------------|---------------------|------------------------|-------------------------------------|---------------------------------|------|
| HG               | 32 HC         | 
| HG               | 29 BPH        | 
| LG                | 30 HG PCa plus | 
| HG                | 29            | NMR                 | PCA, OPLS-DA and ROC   | NS/4                                 | 1. Alanine (+), pyruvate (+)      | 1. Alteration in energetic metabolism and lipogenesis | [68] |
| HG                | 29            | LC-MS               | NMR                 | PCA and ROC            | 348/53                              | 2. Sarcosine (+) glycine (-)      | 2. Alterations in glycine synthesis and degradation | [74] |
| HG                | 29            | GC-MS               | NMR                 | PCA and ROC            | 348/53                              | 2. Sarcosine (+) glycine (-)      | 2. Alteration in fatty acids metabolism | |
| HG                | 29            | GC-MS               | NMR                 | PCA and ROC            | 348/53                              | 3. Arginine                        | 3. Alteration in amino acids metabolism | |

BPH, Benign prostatic hypertrophy; DHEAS, dehydroepiandrosterone sulfate; HC, Healthy Controls; HCA, hierarchical clustering analysis; GS-MS, Gas chromatography–mass spectrometry; HG, high grade; LC-MS, Liquid chromatography–mass spectrometry; LG, low grade; NS, Not specified; OPLS-DA, orthogonal partial least squares discriminant analysis; PCA, principal component analysis; PLS, Partial least squares; ROC, Receiver-Operator Characteristic.

(+): levels increased in PCa; (-): levels decreased in PCa.

TABLE 2 (continued)
cells for energy production. Alterations in carnitine profiles were also detected; carnitines and their derivatives are important for conservation of regular mitochondrial function as well as the transport of activated long-chain fatty acids from the cytoplasm to the mitochondrial compartment. The results also suggested disturbances in energy metabolism, including the Krebs cycle, which were expected given the Warburg effect and the alteration of the activity of m-aconitase, as previously explained (Table 1) [60,63].

Potentially of urinary volatile organic compounds to discriminate between PCa samples and controls was also evaluated in several different studies. In these studies, volatile organic compounds were able to differentiate urine from PCa patients and from control individuals (Table 1) [64,65].

Plasma and Serum Studies. A summary of metabolomics studies performed on serum and plasma samples can be found in Table 2.

Studies evaluating sarcosine as a biomarker for PCa were performed in serum samples using different analytical platforms [fluorometric assay, liquid chromatography–mass spectrometry (LC-MS), and 1H-nuclear magnetic resonance (NMR)]. The results showed that PCa samples had elevated levels of sarcosine and could distinguish low-grade from high-grade PCa, suggesting plasmatic level of sarcosine as a good biomarker for PCa (Table 2) [66–68].

Beyond the alteration in sarcosine levels, metabolomics studies of serum/plasma from PCa patients also revealed alterations in fatty acids, amino acids, lysophospholipids, bile acids, and metabolites related to the steroid hormone biosynthesis pathway. The alteration in fatty acids is related to changes in lipid β-oxidation necessary to provide energy for abnormal cell proliferation (Table 2) [69,68,70–75]. Alterations in energetic metabolism are also common [68,76] (Table 2), and increased levels of glucose in serum samples at the time of PCa diagnosis were associated with an increased risk of recurrences after therapy with radical prostatectomy or radiation therapy [77].

Serum and plasma metabolomics studies can also be used to assess alterations in the metabolic profile caused by medical treatment. The metabolic profile of fasting plasma from PCa patients before starting androgen deprivation therapy and 3 months after the therapy initiation was analyzed [78]. As expected, steroid levels decreased during androgen deprivation therapy, whereas the levels of most bile acids and their metabolites increased with therapy. Bile acids have an important role in the control of serum lipids, glycemic regulation, and energy homeostasis. Lower levels of metabolites related to lipid metabolism after 3 months of treatment were also observed. Carnitines, ketones, dicarboxylic acids, and the levels of 2-hydroxybutyrate and branched-chain keto-acid dehydrogenase complex products (bio-markers of insulin resistance) were also present in lower levels after the therapy, indicating a reduction in the catabolic state [78].

Prostatic and seminal fluids are other biofluids that may be used to perform PCa metabolomics studies to discover alterations in cancer cell metabolism and noninvasive biomarkers for PCa detection. As previously explained, normal prostate cells have the ability to accumulate zinc and consequently accumulate citrate. However, PCa cells lose this ability. Several metabolomics studies support this theory. The analysis of prostatic and seminal fluid using different analytical platforms (fluorescence technique and NMR) revealed reduced levels of zinc and citrate in PCa groups when compared with the controls (Table 3) [46,79–82]. Kline et al. (2006) also concluded that citrate level tests outperform the PSA test in PCa detection. Additionally, the analysis of citrate in semen has the same efficacy as the analysis of citrate in prostatic secretion for detecting PCa [81].

Another important function of normal prostate cells is the synthesis of polyamines, such as spermine and myo-inositol. The analysis of prostatic and seminal fluid from PCa patients showed significantly decreased levels of spermine and myo-inositol (Table 3) [80,82]. Serkova et al. (2008) also demonstrated that the reduction in citrate, spermine, and myo-inositol levels is independent of the patient’s age [82].

Ex Vivo Tissue Studies. Table 4 presents a summary of metabolomics studies performed in PCa tissues.

The value of sarcosine as a PCa biomarker was also evaluated in prostate tissue samples using different analytical platforms. The levels of sarcosine were increased in PCa samples (Table 4) [15,53,59,83]. Results also revealed that sarcosine levels were significantly elevated in metastatic PCa and clinically localized PCa tissue samples, whereas in benign samples, sarcosine was not detected. These results indicate that sarcosine may be a good biomarker for monitoring disease progression and aggressiveness [53].

Other metabolites, namely citrate, lactate, and alanine, were frequently altered in PCa tissue samples. These results suggest alterations in citrate synthesis (Krebs cycle) and in energetic metabolism. As previously explained, the PCa cells switch from
| PCa Subject Group | Control Group | Analytical Platform | Statistical Methods | Total Metabolites Found/Discriminative Metabolites Found | Discriminatory Metabolites/Biomarkers | Metabolic Pathways Dysregulated | Ref. |
|-------------------|--------------|---------------------|---------------------|--------------------------------------------------------|-------------------------------------|---------------------------------|------|
| n= 21             | n= 66        | MRS                 | LDA                 | NS/6                                                   | 1. Citrate (-)                      | 1. Reduced citrate synthesis    | [85] |
|                   |              |                     |                     |                                                       | 2. Taurine (+), glutamate (+)      | ( Krebs cycle)                  |      |
|                   | n= 10        |                     |                     |                                                       | 1. Citrate (-)                      | 2. Alteration in energy metabolism | [86] |
| Adenocarcinoma    |              |                     |                     |                                                       | 2. Choline (+)                      | 1. Reduced citrate synthesis    |      |
|                   | n= 15        | 1H-NMR              | Linear regression   | NS/2                                                   | 1. Citrate (-)                      | 3. Altered membrane metabolism  | [87] |
|                   |              |                     | analysis            |                                                       | 2. Spermine (-)                     |                                |      |
|                   | n= 27        | MRS                 |                     | 22/3                                                   | 1. Citrate (-)                      | 1. Reduced citrate synthesis    | [88] |
| Adenocarcinoma    |              |                     |                     |                                                       | 2. Choline (+)                      | 2. Increased membrane turnover  |      |
|                   | n= 20        | 1H-NMR              | Nonparametric test  | NS/8                                                   | 1. Choline (+), phosphocholine (+), | 1. Increased cellular proliferation | [89] |
|                   |              |                     | of Kruskal-Wallis    |                                                       | glycerophospho-choline (+)          | 1. Alteration in phospholipid   |      |
|                   | n= 15        | 1H-NMR              | Z statistics        | NS/5                                                   | 1. Citrate (-), polyamines (-)     | 2. Alteration in energy metabolism |      |
|                   |              |                     |                     |                                                       | 2. Myo-inositol (+), scyllo-inositol (+) | 3. Altered membrane metabolism |      |
|                   | n= 16        | 1H-NMR              | NS                  | NS/2                                                   | 1. Lactate (+)                      | 4. Reduced citrate and polyamines synthesis | [90] |
|                   |              |                     |                     |                                                       | 2. Alanine (+)                      | Alterations on phospholipid      |      |
|                   | n= 18        | 1H-NMR              | LR                  | NS/7                                                   | rCho/Cr (+), Cho/GC (+), GPC + PC/Cr (+), Lac/Al (+) | 1. “Warburg effect”            | [84] |
|                   | n= 12        | LC-MS               | Wilcoxon P test,    | 626/60                                                 | 1. Sarcoine (+)                     | 2. Intensification in glycolytic  |      |
| Localized PCa     | n= 14        |                     | hierarchical        |                                                       | 2. Uracil (+)                       | flux and increased protein       |      |
|                   |              |                     | clustering,         |                                                       | 3. Kynurenine (+)                   | synthesis in cancer cells        |      |
|                   |              |                     | nonparametric tests |                                                       | 4. Glycerol-3-phosphate (+),       | Alterations in citrate synthesis |      |
|                   | n= 27        | NMR                 | NS                  | NSA                                                   | 3. Proline (+)                      | (Krebs cycle), in membrane       |      |
| patients with     |              |                     |                     |                                                       | 5. Leucine (+), proline (+)        | turnover, and in energetic       |      |
| chemical failure  | n= 32        | 1H-NMR              | PCA                 | NSA                                                   |                                    | metabolism                      |      |
|                   |              |                     |                     |                                                       |                                    | 1. Alterations in glyceine       |      |
|                   | n= 41        | 1H-NMR              | Binary logistic     | 13/6                                                   | 1. Sarcosine (+)                    | synthesis and degradation        | [53] |
|                   |              |                     | regression          |                                                       | 2. Uracil (+)                       | Sarcoine is an intermediate      |      |
|                   | n= 92        | GC-MS               | Nonparametric       | NS/1                                                   | 3. Kynurenine (+)                   | compound in the metabolism of    |      |
|                   |              |                     | statistical tests   |                                                       | 4. Glycerol-3-phosphate (+),       | choline.                        |      |
|                   | n= 331       | GC-MS               | ROC                 | 469/200                                                | 3. Myo-inositol (+), scyllo-inositol (+) | 2. Alteration in pyrimidine      |      |
|                   |              |                     |                     |                                                       | 4. Choline (+), glutamate (+)      | metabolism                      |      |
|                   | n= 11        | GC-MS               | ROC                 | NSA                                                   |                                    | alterations in phospholipid      | [83] |
| Localized PCa     |              |                     |                     |                                                       |                                    | membrane synthesis and         |      |
| plus n= 10        | n= 11        |                     |                     |                                                       |                                    | hydrolysis                      |      |
| metastatic PCa    |              |                     |                     |                                                       |                                    | 4. Alteration in energy metabolism|      |
|                   |              |                     |                     |                                                       |                                    | 1. Alteration in phospholipid     | [94] |
|                   |              |                     |                     |                                                       |                                    | membrane synthesis and hydrolysis|      |
|                   |              |                     |                     |                                                       |                                    | 2. Altered membrane metabolism  | [83] |
|                   |              |                     |                     |                                                       |                                    | 3. Alterations in glyceine       |      |
|                   |              |                     |                     |                                                       |                                    | synthesis and degradation        | [15] |
|                   |              |                     |                     |                                                       |                                    | Sarcoine is an intermediate      |      |
|                   |              |                     |                     |                                                       |                                    | compound in the metabolism of    |      |
|                   |              |                     |                     |                                                       |                                    | choline.                        |      |
|                   |              |                     |                     |                                                       |                                    | 2. Alteration in kynurenine      |      |
|                   |              |                     |                     |                                                       |                                    | pathway                          |      |
|                   |              |                     |                     |                                                       |                                    | 3. Alteration in amino acids      |      |
|                   |              |                     |                     |                                                       |                                    | metabolism                       |      |
|                   |              |                     |                     |                                                       |                                    | 4. Alteration in energy metabolism|      |
|                   |              |                     |                     |                                                       |                                    | 5. Alteration in amino acids      |      |
|                   |              |                     |                     |                                                       |                                    | metabolism                       |      |
|                   |              |                     |                     |                                                       |                                    | 5. Alteration in pyrimidine      |      |
|                   |              |                     |                     |                                                       |                                    | metabolism                       |      |
|                   |              |                     |                     |                                                       |                                    | 6. Reduced polyamines synthesis  |      |
|                   |              |                     |                     |                                                       |                                    | Alterations in glyceine synthesis|      |
|                   |              |                     |                     |                                                       |                                    | and degradation                  |      |
|                   |              |                     |                     |                                                       |                                    | Sarcoine is an intermediate      |      |
|                   |              |                     |                     |                                                       |                                    | compound in the metabolism of    |      |
|                   |              |                     |                     |                                                       |                                    | choline.                        |      |

(continued on next page)
citrate accumulation to citrate oxidation when becoming malignant. PCA cells also undergo the Warburg effect, all of which explain these alterations (Table 4) [15,84–91].

It is also well established that cancer cells have elevated proliferation rates, and this is reflected in alterations in membrane metabolism. Several metabolomics studies performed in PCA tissue revealed an increase of choline levels in PCA samples, which indicates alterations in phospholipid membrane synthesis and hydrolysis (Table 4) [84,86,88,89,92–94]. Because this elevated proliferation rate also increases cell energy requirements, PCA samples also have alterations in lipid metabolism (as lipids may be used by the cells to produce energy) (Table 4) [49,95].

Beyond the study of the metabolic pathways involved in cancer development, it is also important to assess which metabolic pathways are involved in the growth of bone metastases. The results obtained from a study [78] revealed a significant increase in cholesterol levels in bone metastases tissues from PCA patients. The metabolic profile from PCA bone metastases indicates high energy metabolism, which may be related to highly proliferating cells. This conclusion was based on the elevated levels of certain metabolites, such as threonine, glutamate, phenylalanine, citrate, fumarate, glycerol-3-phosphate, and fatty acids. Another relevant metabolic alteration in PCA bone metastases tissue was the elevated levels of myo-inositol-1-phosphate, which may indicate active cell signaling involving inositol-based compounds as second messengers. Inositol-based molecules are related to the activation of protein kinase C, and the activation of this molecule is important for cell proliferation, apoptosis, differentiation, invasion, and angiogenesis. The concentration of sarcosine was increased in bone metastases from PCA and from other cancers, which reveals that sarcosine may not be specific to PCA. Metabolites such as threonine, asparagine, fumarate, and linoleic acid are present in high levels in samples from bone metastases. The levels of these metabolites were also increased in samples of primary prostate tissues from patients with confirmed bone metastases. Linoleic acid, an essential fatty acid, was associated with PCA progression. Furthermore, linoleic acid may also be associated with the inflammatory response because linoleic acid is transformed into arachidonic acid. Arachidonic acid is a precursor for prostaglandins, which have an important role in inflammation [96].

*In Vitro Studies.* Table 5 summarizes general information on in vitro metabolomics studies in PCA-derived cell lines.

The biological relevance of sarcosine was evaluated in four immortalized PCA cell lines, in primary benign prostate epithelial cells, and in an immortalized benign prostate epithelial cell line. Sarcosine levels were increased in malignant cell lines when compared with benign cell lines [53,97]. Furthermore, alterations in the expression of the enzymes involved in sarcosine metabolism influence cell proliferation, invasion, and cell death, which suggest the importance of sarcosine in PCA metabolism (Table 5) [53,59]. In an effort to understand the role of sarcosine in PCA progression, Sreekumar et al. (2009) evaluated the role of androgen signaling and the genes ERG and ETV1 (important mediators of PCA progression). The results showed that after treatment with androgens, cell lines that were ERG positive and ETV1 positive had increased GNMT expression and decreased sarcosine dehydrogenase (SARDH) expression [53].

In agreement with the results from metabolomics studies performed in other matrices (urine, plasma/serum, and tissues), presented here previously, alterations in amino acid metabolism were also observed in the studies performed in PCA cell lines. PCA cell lines revealed alterations in the levels of certain amino acids, such as leucine, valine, or isoleucine (Table 5) [97–99].

The increase of lactate and alanine levels in PCA cell lines is frequently observed in metabolomics studies. These alterations suggest changes in cellular energy metabolism (Table 5) [97–100].

As reported for other matrices, metabolomics studies in PCA also revealed that PCA cells experience alterations in membrane metabolism with changes, for example, in choline metabolite levels (Table 5) [98,101]. Androgen signaling has an important role in the development and progression of PCA. In fact, one current therapy for metastatic PCA is the use of antiandrogen agents; however, with the progression of the disease, patients normally develop resistance to this therapy, and it is currently impossible to predict if the cancer will progress into a castration-resistant state. The androgen-responsive cell lines can be characterized by increased levels of spermine, N-acetylspermine, serine, threonine, lysine, homocysteine, asparagine, alanine, and glutamic acid, as well as decreased levels of S-adenosylmethionine, with a simultaneous increase in levels of its breakdown product, homocysteine. These findings indicate that androgen-responsive cell lines have an elevated methylation activity. Androgen treatment resulted in further perturbations in amino acid metabolism and in a shift toward increased methylation.

| Table 4 (continued) |
|---------------------|
| PCA Subject Group | Control Group | Analytical Platform | Statistical Methods | Total Metabolites Found/Discriminative Metabolites/Biomarkers | Metabolic Pathways Dysregulated | Ref. |
|---|---|---|---|---|---|---|
| n-95 | n-95 | Normal adjacent prostate tissue | GC-MS | ROC, Univariate Cox regression, Kaplan-Meier analyses, and multivariate Cox regression analyses | 820/9 | 1. Gluconic acid (−), maltotriose (−) | 1. Alteration in carbohydrate metabolism [49] |
| n-30 LG PCa plus n-47 | n-81 HG PCa | Normal adjacent tissue | MRS | PCA, PLS and PLS-DA | 23/2 | 1. Citrate (−), Spermine (−) | 1. Reduced citrate synthesis [91] |

**BP**: Benign prostate hypertrophy; Cho/Cr, choline/creatinine; Cit/Cr, citrate/creatinine; GS-MS, Gas chromatography–mass spectrometry; (GPC + PC)Cr, glycerol-phosphocholine + phospholyl-choline)/creatinine; HC, Healthy Controls; HG, High grade; Lac/Al, lactate/alanine; LC-MS, Liquid chromatography–mass spectrometry; LIA, Linear discriminant analysis; LG, Low grade; LR, Linear regression MRS, magnetic resonance spectroscopy; NMR, Nuclear magnetic resonance; NS, Not specified; PCA, Principal component analysis; PLS, partial least squares; PLS-DA, partial least squares discriminant analysis; rChol/Cr, total choline/creatinine. (+): levels increased in PCa; (−): levels decreased in PCa.
Table 5. Metabolomic Studies Performed in Human PCa-Derived Cell Lines

| Cancer Cell Lines                                                                 | Control Group                                      | Analytical Platform | Statistical Methods | Total Metabolites Found/ Discriminatory Metabolites Found | Discriminatory Metabolites/Biomarkers                                                                 | Metabolic Pathways Dysregulated                                                                 | Ref. |
|----------------------------------------------------------------------------------|----------------------------------------------------|---------------------|---------------------|----------------------------------------------------------|---------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------|------|
| AD prostate carcinoma LNCaP cell line                                            | Androgen-independent prostate carcinoma PC-3 cell line | MRS                 | NS                  | 3/2                                                                 | Uptake of ethanolamine and N,N′-dimethylthanolamine (+) (principally in AD cells in presence of androgens) | Alteration in membrane lipid synthesis                                                                 | [101]|
| VCaP, DU145, 22RV1, and LNCaP                                                   | PrEC and RWPE                                      | LC-MS               | Wilcoxon P test, hierarchical clustering, nonparametric tests   | 1/1                                                                 | Sarcosine (+)                                                                  | Alterations in glycine synthesis and degradation                               | [53] |
| Androgen-nonresponsive PC3 and DU145 cell lines                                    | RWPE                                               | LC-MS               | HCA                 | 1553/674                                                      | Malignant cell lines                                                                                     |                                                                                   |      |
| Androgen-responsive VCaP (treated with synthetic androgen) and LNCaP cell lines  |                                                                                   |                     |                     |                                                          | 1. Sarcosine (+)                                                                                       |                                                                                   |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | 2. Threonine (+), phenylalanine (+), alanine (+), creatine (+), creatinine (+), citrulline (+), tryptophan (+), 1-methyl tryptophan (-) and kyurenine acid (-) |                                                                                   |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | 3. Serine (+), threonine (+), lysine (+), homocysteine (+), asparagine (+), alanine (+), glutamic acid (+) |                                                                                   |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | 4. S-adenosylmethionine (-), homocysteine (+)                                                          |                                                                                   |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | Androgen treatment resulted in further perturbations in amino acid metabolism and increased methylation. |                                                                                   |      |
| PC3 and LNCaP treated with LY294002 (inhibitor of the PI3K signaling pathway) or 17AAG (inhibitor of the HSP90 protein chaperone) | PC3 and LNCaP untreated                                                            | MRS                 | PCA                 | NS/24                                                      | After both treatments: lactate (-), alanine (-), fumarate (-)                                                                 | PI3K and HSP90 inhibition                                                                 | [102]|
| Low-invasiveness WPE1-NB14 and high-invasiveness WPE1-NB11 cell lines            | WPE-1                                                                 | H-NMR               | PLS-DA              | NS/10                                                      | 1. Leucine (+), valine (-), isoleucine (-), glutamine (-), glutamate (-), β-hydroxyisovalerate (-)      | 1. Increased protein synthesis and amino acid catabolism                        | [98] |
|                                                                                   |                                                                                   |                     |                     |                                                          | 2. Glycine (-)                                                                                           | 2. Alterations in synthesis and degradation of sarcosine                        |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | 3. Lactate, alanine                                                                                      | 3. Alteration in energetic metabolism                                           |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | 4. Phosphocholine (+)                                                                                   | 4. Alteration in choline metabolism                                             |      |
| Androgen-nonresponsive PC3 and androgen-responsive LNCaP cell lines              | PNT1A                                                                 | H-NMR               | Two-way analysis of variance followed by Bonferroni post test    | NS/3                                                      | Glucose consumption (+)                                                                                   | Increased levels of oxidative stress in PC3 cells.                              | [100]|
| DU145, PC3, and LNCaP (knockdown of GNMT, SARDH, or PIPOX and overexpression of GNMT, SARDH, or PIPOX (convert sarcosine back to glycine) | RWPE                                               | GC-MS               | ROC                 | NS/1                                                      | Overexpression of GNMT: sarcosine (+) (increase in invasion)                                                                 | Alteration in glycine synthesis and degradation                                  | [59] |
|                                                                                   |                                                                                   |                     |                     |                                                          | Knockdown of GNMT: sarcosine (-) (reduction in cell proliferation, invasion, and greater percentage of cell death) |                                                                                   |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | Overexpression of SARDH or PIPOX: sarcosine (-) (reduced invasion)                                                                 |                                                                                   |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | Knockdown of SARDH: sarcosine (+) (increase proliferation and invasion)                                                                 |                                                                                   |      |
|                                                                                   |                                                                                   |                     |                     |                                                          | Knockdown of PIPOX: sarcosine (+) (increased invasion)                                                                 |                                                                                   |      |
|                                                                                   | Androgen receptor positive LNCaP and MDA-PCa-2a and MDA-PCa-2b                   | LC-MS               | HCA                 | 150/38                                                     |                                                                                                           | 1. Alteration in energy metabolism and signaling                                | [99] |
|                                                                                   |                                                                                   |                     |                     |                                                          | 2. Alteration in amino acid metabolism                                                                 |                                                                                   |      |

(continued on next page)
Androgen-non-responsive cell lines and androgen-responsive cell lines also have differences in their glycolytic metabolism profiles. Androgen-dependent PCa cells and androgen-independent PCa cells also show differences in membrane lipid synthesis (Table 5) [97,100,101].

Metabolomics studies in cell lines may also be used to evaluate the alterations that occur after a pharmacological therapy. The treatment of PCa cells appears to lead to changes in energetic metabolism and choline metabolism (Table 5) [102]. Dichloroacetate (DCA) is an inhibitor of pyruvate dehydrogenase kinase, and inhibition of this enzyme has the potential to reverse the Warburg effect due to the increased pyruvate uptake into mitochondria. After treatment with this drug, the highly metastatic cells showed significantly lower levels of lactate metabolite ratios [Lac/Cr, Lac/Cho, Lac/Al, and Lac/(Cho + Cr + Al)], whereas in poorly metastatic cells, no changes in lactate/metabolite ratios were found after the treatment. These findings suggest that highly metastatic cells are more dependent on lactate production (Table 5) [103].

Conclusions and Future Directions

The introduction of PSA testing has radically altered how PCa is diagnosed and managed. However, this test may lead to a false-positive or false-negative diagnosis. This drawback has given rise to serious efforts toward the discovery of new biomarkers, preferentially noninvasive, which have better specificity and sensitivity.

Because metabolic alterations are the last step in the cellular response to diseases, metabolomics can be successfully used to discover new biomarkers for cancer. In this regard, several studies have been conducted to characterize the metabolic profile of PCa.

One of the major obstacles in data interpretation is that the metabolic profile is influenced by various factors, such as age, diet, drugs, and chronobiological variations, among others. Additional problems include sample preparation, the analytical procedures, and the statistical platforms used. Major differences in these conditions can compromise the comparison of results among different studies and consequently compromise the discovery of new biomarkers. Another intrinsic difficulty with metabolomics studies is the massive amount of data produced that is difficult statistically to analyze. Despite these difficulties, metabolomics is a powerful tool for the discovery of new biomarkers for PCa detection, biomarkers indicative of cancer prognosis, disease progression, and therapeutic response, as well as identifying new therapeutic targets.

The metabolomics studies described in this review revealed different results, but almost all studies in urine, plasma/serum, prostatic fluids, tissues, and cell lines associated PCa with decreased levels of citrate and polyamines and increased levels of choline, lactate, and amino acids.

Further studies are still needed to confirm the results of these studies and identify an inexpensive, noninvasive, sensible, and specific biomarker for PCa.

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