Molecular Prevalence of Cryptosporidium spp. among Companion Birds Kept in Pet Shops in Japan

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Abstract: Cryptosporidium is the most common protozoan that can infect a wide range of animals, including mammals and birds. Avian Cryptosporidium spp. can cause enteric and respiratory diseases which can be fatal in birds and some species are zoonotic. Companion birds have the potential as reservoir due to their close contact with humans. Pet shops are the major source of companion birds. However, few reports are available regarding Cryptosporidium spp. infection among companion birds kept in pet shops. The present study reports the prevalence and molecular characteristics of Cryptosporidium spp. among companion birds kept in pet shops in Japan. A total of 265 fresh fecal samples were obtained from birds kept in 4 pet shops; these birds belonged to 41 species in 3 bird orders. A nested polymerase chain reaction (PCR) assay targeting the small subunit rRNA gene was employed for the detection of Cryptosporidium spp. A total of 24 samples (9.1%) were positive, and Cryptosporidium spp. were detected from all pet shops. The prevalence of Cryptosporidium spp. in each of the bird orders was 6.5% (10/153) in Psittaciformes, 14.4% (13/90) in Passeriformes, and 4.5% (1/22) in Galliformes. Based on sequence analysis, 13 (54.2%) isolates were classified to C. galli, whereas 8 (33.3%) were avian genotype III, and the remaining 3 (12.5%) were C. baileyi. No infection with zoonotic C. meleagridis and no coinfection with multiple Cryptosporidium spp. and/or genotypes were observed. The zoonotic potential of Cryptosporidium spp. infecting companion birds kept in pet shops in Japan is likely to be low.

Key words: Cryptosporidium, C. baileyi, C. galli, bird, pet shop, avian genotype III

The parasite Cryptosporidium is one of the most common protozoans that can infect a wide range of animals, including mammals and birds worldwide [1]. In birds, cryptosporidiosis, which is often fatal, is mainly induced by the 3-dominant species: C. meleagridis, C. galli and C. baileyi. C. meleagridis and C. galli infect the gastrointestinal tract and cause enteritis [2,3], whereas C. baileyi can infect many organs and mainly causes respiratory disorders [3]. In addition, Cryptosporidium avian genotypes III and V have the pathogenic potential to cause mortality, weight loss, chronic vomiting and diarrhea in birds [4-6]. Many other species and genotypes were also identified in birds [3,5]. Among the avian isolates of Cryptosporidium spp., C. meleagridis can infect humans and cause digestive tract obstruction in immunocompromised adults and children [7-10]. In some locations, approximately 10% of human cryptosporidiosis is caused by C. meleagridis [8]. Therefore, Cryptosporidium spp. infection in birds has pathogenic significance for birds and zoonotic risk for humans. In particular, companion birds have a considerable potential to serve as a reservoir due to their close contact with humans. However, the knowledge about Cryptosporidium spp. infection in companion birds is limited, and no report exists regarding Cryptosporidium spp. infection among companion birds kept in pet shops in Japan, although pet shops are the major source of companion birds for private owners. The present study reports the recent prevalence and molecular characteristics of Cryptosporidium spp. among companion birds kept in pet shops in Japan.

Between March 2015 and May 2016, a total of 265 fresh voided fecal samples were collected on a single occasion from each birdcage (birds of a single species were kept in each cage) in 4 pet shops (Shop 1-4) located in the Kanto region, which includes the prefectures of Tokyo (Shop 1; n = 78), Chiba (Shop 2; n = 53), Saitama (Shop 3; n = 59), and Gunma (Shop 4; n = 75), in Japan. All pet shop managers granted permission to include their birds in the examination. The birds belonged to 41 species in 3 orders (Psittaciformes, Passeriformes, and Galli-
formes). The fecal samples were collected immediately after natural defecation and were stored at 4˚C prior to DNA extraction (within 3 days). The extraction of *Cryptosporidium* spp. DNA was performed using a QIAamp DNA Mini Kit (QIAGEN GmbH, Hilden, Germany) according to the manufacturer’s instructions, and the extracted DNA samples were stored at -20˚C prior to analysis.

A nested polymerase chain reaction (PCR) assay targeting the small subunit (SSU) rRNA gene was employed for the detection of *Cryptosporidium* spp. In the primary reaction, the forward primer (5'-TTCTAGAGCTAATACATGCG-3') and the reverse primer (5'-CCCATTTCCTTCGAAACAGGA-3') were used to amplify a DNA fragment of approximately 1,325 bp, and in the secondary reaction, the forward primer (5'-GGAAGGGTTATTATAGAAAAG-3') and the reverse primer (5'-AAGGATAAAGGAACAACCTCCA-3') were used to amplify a fragment of approximately 826 bp [11]. For the primary reaction, the PCR mixture comprised a 1× buffer containing 1.5 mM MgCl₂, 200 μM aliquots of each dNTP, 0.5 μM aliquots of each primer, 1.25 units of GoTaq DNA polymerase (Promega Corporation, Madison, Wisconsin, USA), and 3.0 μl of template DNA in a total reaction volume of 25 μl. For the secondary reaction, the PCR mixture was the same as that for the primary reaction, except the amplicons from the primary PCR reaction were used as the template. The following cycling parameters were used for the primary reaction: after an initial denaturation of 3 min at 95˚C, 35 cycles were performed, each consisting of 45 sec at 95˚C for denaturation, 45 sec at 59˚C for annealing, and 60 sec at 72˚C for extension, followed by a final extension step of 5 min at 72˚C. The cycling parameters for the secondary reaction were as follows: an initial denaturation step of 3 min at 95˚C, 35 cycles of 30 sec at 95˚C, 60 sec at 58˚C, and 1 min at 72˚C; and a final extension step of 5 min at 72˚C.

All secondary PCR products were identified by electrophoresis on 1.5% agarose gels. The specific DNA fragments (approximately 826 bp in length) were confirmed by alternative ethidium bromide staining and visualization under UV light using a transilluminator. Secondary PCR amplicons of the predicted size were purified using a QIAquick Gel Extraction kit (QIAGEN GmbH, Hilden, Germany) and sequenced with the primer set used in the secondary PCR reaction. Sequences were analyzed by a commercial laboratory (Takara Bio Inc., Kusatsu, Japan).

![Fig. 1. PCR products on 1.5% agarose gel. Lane 1: Cryptosporidium positive control (C. canis), lane 2: C. baileyi from domestic canary, lane 3: C. galli from barred parakeet, lane 4: Cryptosporidium avian genotype III from Lilian’s lovebird, lane 5: Cryptosporidium negative control, lane 6: 100 bp DNA ladder. ← indicates approximately 826 bp.](image-url)

### Table 1. Molecular prevalence and characterization of *Cryptosporidium* spp. among companion birds kept in pet shops

| Order of host | Shop 1 (Tokyo) | Shop 2 (Chiba) | Shop 3 (Saitama) | Shop 4 (Gunma) | Overall |
|--------------|---------------|---------------|-----------------|---------------|---------|
| Psittaciformes | 5/53 (9.4%) Avian genotype III (5)* | 0/28 (0%) | 4/37 (10.8%) C. galli (2) Avian genotype III (2) | 1/35 (2.8%) | 10/153 (6.5%) |
| Passeriformes | 3/25 (12.0%) C. galli (3) | 7/25 (28.0%) C. galli (6) C. baileyi (1) | 1/14 (7.1%) C. baileyi (1) | 2/26 (7.6%) C. galli (2) | 13/90 (14.4%) |
| Galliformes | - | - | 0/8 (0%) | 1/14 (7.1%) C. baileyi (1) | 1/22 (4.5%) |
| Total | 8/78 (10.3%) | 7/53 (13.2%) | 5/59 (8.5%) | 4/75 (5.3%) | 24/265 (9.1%) |

*Numbers of isolates.
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Sequence alignment and compilation were performed using the MEGA 6.06 (www.megasoftware.net) program. To determine the species and/or genotypes of Cryptosporidium, the DNA sequences were compared to GenBank references by BLAST searches (http://www.ncbi.nlm.nih.gov/), and the similarity between the isolated and reference sequences was determined based on the degree of sequence identity.

Among the 265 fecal samples from the companion birds kept in pet shops, 9.1% (24 samples) were positive for Cryptosporidium spp., as determined by conventional PCR (Fig. 1). Cryptosporidium spp. were isolated from birds in each pet shop, with a prevalence of 5.3-13.2% (Table 1). Of the 41 examined bird species in the 3 orders, 10 of the bird species belonging to one of the 3 orders were positive for Cryptosporidium spp. (Table 2). The prevalence of Cryptosporidium spp. in each of the 3 orders of birds was 6.5% (10/153) in Psittaciformes, 14.4% (13/90) in Passeriformes, and 4.5% (1/22) in Galliformes. In the order Psittaciformes, birds of 5 bird species were positive for Cryptosporidium spp.: the Rosy-faced lovebird (17.2%, 5/29), the Lilian’s lovebird (20.0%, 1/5), the barred parakeet (100%, 2/2), the Pacific parrotlet (33.3%, 1/3), and the cockatiel (10.0%, 1/10). In the order Passeriformes, birds of 4 species were determined to be infected with Cryptosporidium spp.: the society finch (33.3%, 5/15), the Java sparrow (4.0%, 1/25), the zebra finch (20.0%, 4/20), and the domestic canary (13.6%, 3/22). The only species in the order Galliformes that tested positive was the chicken (6.7%, 1/15).

Based on sequence analysis, 2 species and one genotype of Cryptosporidium were identified. The identity of the 24 positive samples was determined as follows (sequence similarity 99.2-100%): 13 (54.2%) isolates were C. galli (accession number KY409554), 8 (33.3%) isolates were avian genotype III (accession number AB694729), and the remaining 3 (12.5%) isolates were C. baileyi (accession number KU744846) (Fig. 2). No infection with C. meleagridis and no coinfection with multiple Cryptosporidium spp. and/or genotypes were observed. C. galli was found in birds from all pet shops (Shops 1, 2, 3, and 4). Avian genotype III was detected in 3 pet shops (Shops 1, 3, and 4), and C. baileyi was also detected in 3 pet shops (Shops 2, 3, and 4). Thirteen C. galli isolates were found in 5 bird species, one of these was the barred parakeet (2 isolates).

Table 2. Bird species determined to be infected with Cryptosporidium spp.

| Host                          | Positive/examined | Cryptosporidium species/genotypes |
|-------------------------------|------------------|----------------------------------|
| **Psittaciformes**             |                  |                                  |
| Rosy-faced lovebird (Agapornis roseicollis) | 5/29 (17.2%)     | Avian genotype III (5)*          |
| Lilian’s lovebird (Agapornis lilianae) | 1/5 (20.0%)     | Avian genotype III (1)           |
| Barred parakeet (Bolborhynchus lineola) | 2/2 (100%)      | C. galli (2)                     |
| Pacific parrotlet (Forpus coelestis) | 1/3 (33.3%)     | Avian genotype III (1)           |
| Cockatiel (Nymphicus hollandicus) | 1/10 (10.0%)    | Avian genotype III (1)           |
| **Passeriformes**             |                  |                                  |
| Society finch (Lonchura striata domestica) | 5/15 (33.3%)    | C. galli (5)                     |
| Java sparrow (Padda oryzivora) | 1/25 (4.0%)     | C. galli (1)                     |
| Zebra finch (Taeniopygia guttata) | 4/20 (20.0%)    | C. galli (4)                     |
| Domestic canary (Sturnus canaria) | 3/22 (13.6%)   | C. baileyi (2), C. galli (1)     |
| **Galliformes**               |                  |                                  |
| Chicken (Gallus gallus domesticus) | 1/15 (6.7%)    | C. baileyi (1)                   |

*Numbers of isolates.

Fig. 2. Phylogenetic analysis of the small subunit rRNA gene sequences from Cryptosporidium spp. isolates among companion birds kept in pet shops in Japan.
in the order Psittaciformes. The other 4 bird species were the society finch (5 isolates), the Java sparrow (1 isolate), the zebra finch (4 isolates), and the domestic canary (1 isolate), all of which belong to the order Passeriformes. Eight positive isolates of avian genotype III were found in 4 bird species; the Rosy-faced lovebird (5 isolates), the Lilian’s lovebird (1 isolate), the Pacific parrotlet (1 isolate), and the cockatiel (1 isolate), all of which belong to the order Psittaciformes. Three *C. baileyi* isolates were identified in 2 bird species. One of these species was the domestic canary (2 isolates), in the order Passeriformes; the other was the chicken (1 isolate), in the order Galliformes.

The present study is the first report to demonstrate the prevalence of *Cryptosporidium* spp. among companion birds kept in pet shops in Japan. Information regarding the prevalence of infection by *Cryptosporidium* spp. among birds kept in pet shops and/or bird kept in markets is limited [12-16]. Previously, the prevalence has been reported as 6.8% (7/103) in birds kept in bird markets and pet shops in Brazil [13] and 3.2-13.4% from birds kept in bird markets or pet shops in China [12,14-16]. Due to differences in research methodology, population and scale, comparison between the present study and previous reports is difficult. However, the present results suggest that infection with *Cryptosporidium* spp. is infrequent but widespread among companion birds kept in pet shops in Japan because isolates of *Cryptosporidium* spp. were obtained from companion birds in all pet shops and bird orders examined. In the present study, 10 bird species belonging to 3 orders were found to be infected with *Cryptosporidium* spp. Since these species are common as companion birds, *Cryptosporidium* spp. infections have previously been detected in the same or closely related bird species [12-20].

In the present study, *C. galli* was found in the orders Psittaciformes and Passeriformes, and most of the isolates were from the order Passeriformes. Avian genotype III was limited to the order Psittaciformes. *C. baileyi* was found in the orders Passeriformes and Galliformes. In contrast, 2 or 3 species of *Cryptosporidium* were confirmed in all shops. Previous reports demonstrated that *C. galli* has been isolated from the bird orders Psittaciformes, Passeriformes, and Galliformes, and others [5,14,15]. Avian genotype III was isolated from the bird orders Psittaciformes and Passeriformes [5,14]. *C. baileyi* was infrequently isolated in the present study, but was determined to infect quite a wide range bird orders, including Psittaciformes, Passeriformes, and Galliformes [3,5,14,15]. Avian genotypes III, V, and *C. galli* have been detected in companion birds of the order Psittaciformes kept in private households in Japan [4,6,17]. Additionally, *C. baileyi* has been isolated from Passeriformes [6]. Therefore, the present study is the first to identify *C. galli* in Passeriformes and *C. baileyi* in Galliformes among pet birds in Japan. Although *C. meleagridis* was not detected here, this zoonotic species was isolated from companion birds kept in private households in Japan [6,17]. Thus, potential for zoonotic transmission of *C. meleagridis* from companion birds kept in pet shops to humans is not negligible.

Although the pathogenic potential of the species and genotypes of *Cryptosporidium* isolates in the present study was indicated [2-6], the infected birds exhibited no clinical signs, with the exception of one Pacific parrotlet (infected with avian genotype III) that displayed anorexia. These results suggested that many companion birds kept in pet shops infected with *Cryptosporidium* spp. display subclinical infection, and those carrier birds have a risk of developing to clinical cryptosporidiosis in future.

**CONFLICT OF INTEREST**

On behalf of all authors, the corresponding author states that there is no conflict of interest.

**REFERENCES**

1. Ryan U, Fayer R, Xiao L. *Cryptosporidium* species in humans and animals: current understanding and research needs. Parasitology 2014; 141: 1667-1685.
2. Gharagozlou MJ, Dezfooli O, Rahbari S, Bokaei S, Jahanzad I, Razavi AN. Intestinal cryptosporidiosis in turkeys in Iran. J Vet Med A Physiol Pathol Clin Med 2006; 53: 282-285.
3. Ryan U. *Cryptosporidium* in birds, fish and amphibians. Exp Parasitol 2010; 124: 113-120.
4. Makino I, Abe N, Reavill DR. *Cryptosporidium* avian genotype III as a possible causative agent of chronic vomiting in peach-faced lovebirds (*Agapornis roseicollis*). Avian Dis 2010; 54: 1102-1107.
5. Nakamura AA, Meireles MV. *Cryptosporidium* infections in birds—a review. Rev Bras Parasitol Vet 2015; 24: 253-267.
6. Abe N, Makino I, Kojima A. Molecular identification of *Cryptosporidium* isolates from pet birds in Japan. Jpn J Vet Parasitol 2016; 15: 19-23.
7. Kurniawan A, Dwintasari SW, Connelly L, Nichols RA, Yuniasztuti E, Karyadi T, Djauzi S. *Cryptosporidium* species from human immunodeficiency-infected patients with chronic diarrhea in Jakarta, Indonesia. Ann Epidemiol 2013; 23: 720-723.
8. Wang Y, Yang W, Cama V, Wang L, Cabrera L, Ortega Y, Bern C, Feng Y, Gilman R, Xiao L. Population genetics of *Cryptosporidium* species in humans and animals: current understanding and research needs. Parasitology 2014; 141: 1667-1685.
meleagridis in humans and birds: evidence for cross-species transmission. Int J Parasitol 2014; 44: 515-521.

9. Moore CE, Elwin K, Phot N, Seng C, Mao S, Suy K, Kumar V, Nader J, Bousfield R, Perera S, Bailey JW, Beeching NJ, Day NP, Parry CM, Chalmers RM. Molecular Characterization of Cryptosporidium species and Giardia duodenalis from symptomatic Cambodian children. PLoS Negl Trop Dis 2016; 10: e0004822.

10. Wesołowska M, Szostakowska B, Kicia M, Sak B, Kvac M, Knysz B. Cryptosporidium meleagridis infection: the first report in Poland of its occurrence in an HIV-positive woman. Ann Parasitol 2016; 62: 239-241.

11. Xiao L, Morgan UM, Limor J, Escalante A, Arrowood M, Shulaw W, Thompson RC, Fayer R, Lal AA. Genetic diversity within Cryptosporidium parvum and related Cryptosporidium species. Appl Environ Microbiol 1999; 65: 3386-3391.

12. Qi M, Wang R, Ning C, Li X, Zhang L, Jian F, Sun Y, Xiao L. Cryptosporidium spp. in pet birds: Genetic diversity and potential public health significance. Exp Parasitol 2011; 128: 336-340.

13. Gomes RS, Huber F, da Silva S, do Bomfim TC. Cryptosporidium spp. parasitize exotic birds that are commercialized in markets, commercial aviaries, and pet shops. Parasitol Res 2012; 110: 1363-1370.

14. Zhang XX, Zhang NZ, Zhao GH, Zhao Q, Zhu XQ. Prevalence and genotyping of Cryptosporidium infection in pet parrots in north China. Biomed Res Int 2015; 2015: 549798.

15. Li Q, Li L, Tao W, Jiang Y, Wan Q, Lin Y, Li W. Molecular investigation of Cryptosporidium in small caged pets in northeast China: host specificity and zoonotic implications. Parasitol Res 2016; 115: 2905-2911.

16. Yao QX, Zhang XX, Chen K, Ma JG, Zheng WB, Xu XQ, Zhu XQ. Prevalence and genetic characterization of Cryptosporidium infection in Java Sparrows (Lonchura oryzivora) in northern China. Biomed Res Int 2017; 2017: 2318476.

17. Abe N, Makino I. Multilocus genotypic analysis of Cryptosporidium isolates from cockatiels, Japan. Parasitol Res 2010; 106: 1491-1497.

18. Sevá AP, Funada MR, Richtzenhain L, Guimarães MB, Souza Sde O, Allegretti L, Sinhorini JA, Duarte VV, Soares RM. Genotyping of Cryptosporidium spp. from free-living wild birds from Brazil. Vet Parasitol 2011; 175: 27-32.

19. Ewald MPC, Martins FDC, Caldart ET, Vieira FEG, Yamamura MH, Sasse JP, Barros LD, Freire RL, Navarro IT1, Garcia JL. The first study of molecular prevalence and species characterization of Cryptosporidium in free-range chicken (Gallus gallus domesticus) from Brazil. Rev Bras Parasitol Vet 2017; 26: 472-478.

20. Ferrari ED, Nakamura AA, Nardi ARM, Santana BN, da Silva Camargo V, Nagata WB, Bresciani KDS, Meireles MV. Cryptosporidium spp. in caged exotic psittacines from Brazil: Evaluation of diagnostic methods and molecular characterization. Exp Parasitol 2018; 184: 109-114.
