The cyclization reaction of CBD seems to occur following an acid-catalyzed activation of a specific double bond. The main products of this cyclization are Δ9-tetrahydrocannabinol (Δ9-THC) and Δ9-tetrahydrocannabinolic acid (Δ9-THCA). Based on the electrospray positive ion mode, the detection of cannabidiolic acid (CBDA; m/z 359 > 219), cannabigerolic acid (CBGA; m/z 361 > 343), cannabigerol (CBG; m/z 317 > 193), CBD (m/z 315 > 193), THC (m/z 315 > 193) and cannabiol (CBN; m/z 311 > 223) was performed by satisfying separation with high density of C18 column. Oil samples (50 mg) were diluted with isopropanol (5 mL), to which stable isotope internal standards were added by dilution with methanol/water (50/50), and accuracy rates ranged from 97.8 to 102.2%. This method was used to evaluate the CBD oil products (5 kinds) from the Japanese market. Our survey found obvious counterfeit (non-detectable CBD) CBD oil from Japanese market. Following that, we investigated the conversion of THC in CBD oil samples in simple conditions such as 10% acetic acid and 70 °C for 6 h and discovered that converts THC proportions are approximately 5% ((THC content/CBD content)×100) and <1.0%. Thus, our developed LC-MS/MS assay could be applied to monitor the CBD concentration and convertible THC from CBD oil.

Introduction
Recently, the fashioning trends of cannabis (Cannabis sativa L.), specifically cannabidiol (CBD), containing natural health cosmetics and supplements in Asia-Pacific (Japan, Australia and New Zealand) markets from increasing import growth. On the other hand, in Japanese markets, CBD-containing products are not widely available in general stores. However, specific/internet retailers selling natural health cosmetics also do business in various aspects of CBD products, such as hemp seed oil and related items advertised to help with anxiety, relaxation, restful sleeping, and anti-aging. CBD products have been appeared to be imported from the United States and other countries. In addition, the presence of the psychoactive/illegal tetrahydrocannabinol (THC) and impurity from these products have been assessed before being available to the market in Japan. However, in terms of important conditions, ingredients, and storage period from common markets, it has remained insufficient that this quality evaluation of THC and/or impurity is recognized in circulated CBD products. As a result, we should investigate the possibility of impurity-containing products under a variety of conditions, including additive interaction, acid-catalyzed reaction, and unknown phenomenon. Thus, it is needed to develop the useful and simple assay for the evaluation of CBD products in Japanese markets.

The cyclization reaction of CBD seems to occur following an acid-catalyzed activation of a specific double bond. The main products of this cyclization are Δ9-THC (trans-Δ9-tetrahydrocannabinol) and Δ9-THCA (trans-Δ9-tetrahydrocannabinolic acid). Recently, Marzullo et al. investigated the susceptibility and selectivity of CBD cyclization, different reactions, including the use of Lewis and protic acids in different solvents and varying the temperature and reaction time, concluded that CBD is a difficult substrate to exploit the chemical reactivity of natural alkenes and phenols. Pratap Singh et al. reported that acidic cannabinoids decarboxylation kinetics in hemp seed oil based on various antioxidants. Moreover, Kiselak et al. investigated to utilize a weaker acid, such as vinegar (5.4% acetic acid) to isomerize CBD to Δ8-THC, Δ8-THCA, and other impurity chemicals from original simulated-foods. Unfortunately, it is impossible to obtain the useful, veridical, and reliable information about impurity-containing CBD oil from potential conditions such as coexistence in these conditions regarding to Japanese markets. Thus, we examined a LC-tandem mass spectrometry (LC-MS/MS) assay to assess the quality evaluation of CBD and cannabinoids profiles in hemp seed oil products from Japanese markets, as well as apply convertible THC in acetic acid conditions. The most frequently reported generic methods are applied with chromatographic separation such as LC-MS/MS for screening assay of CBD and other cannabinoids in various products. In this study, we used the previous LC-MS/MS assay to measure CBD, THC, and other cannabinoids in hemp seed oil products.

**Experimental**

**Chemicals and Reagents** Δ9-THC (1.0 mg/mL) was obtained from Sigma-Aldrich Co (St Louis, MO, U.S.A.). CBD (1.0 mg/mL), cannabiol (CBN) (1.0 mg/mL), cannabigerol (CBG) (1.0 mg/mL), cannabidiolic acid (CBDA) (1.0 mg/mL), cannabigerolic acid (CBGA) (1.0 mg/mL), CBD-d1 (100 μg/mL), and Δ9-THCA-d1 (100 μg/mL) were obtained...
from Cerilliant (St. Texas, TX, U.S.A.). FUJIFILM Wako Pure Chemical Corporation (Osaka, Japan) provided methanol, acetonitrile, isopropanol, acetic acid, and formic acid. The mobile phase was purified water, and sample preparation was accomplished using a PURELAB Flex 5 system (ELGA, U.K.). Methanol was used to adjust a stock solution of analytes (50 µg/mL), which was then stored at −80 °C. In 2021, CBD products were purchased from the Japanese market (via online shopping).

**LC-MS/MS Instrument and Condition** The LC system was a Waters Acquity H Class Plus (Waters Co., Milford, MA, U.S.A.). An RP analysis was performed via an Inertsil ODS-HL column (3 µm, 2.1 × 150 mm, GL Sciences, Inc., Tokyo, Japan) at 40 °C. For the optimal separation, the TSKgel ODS-100V (3 µm, 2.0 × 150 mm, Tosoh Co., Tokyo, Japan), InertSustain AQ-C18 (1.9 µm, 2.1 × 150 mm, GL Sciences, Inc.), TSKgel ODS-120H (3 µm, 2.0 × 150 mm, Tosoh Co.), Acquity UPLC BEH C18 (1.7 µm, 2.1 × 150 mm, Waters, MA, U.S.A.), TSKgel ODS-100Z (3 µm, 2.0 × 150 mm, Tosoh Co.), and Atlantis T3 (3 µm, 2.1 × 150 mm, Waters) were used. The injection volume was set at 5 µL. The mobile phase was delivered at a flow rate of 0.2 mL/min and consisted of solvent A (0.1% formic acid in water) and solvent B (0.1% formic acid in acetonitrile). The linear gradient elution was as follows: 75% solvent B at 0 min, 75% solvent B at 9 min, 98% solvent B at 9 min, 98% solvent B at 19 min, 75% solvent B at 19.1 min, and 75% solvent B at 25 min. A Waters Xevo TQD triple quadrupole mass spectrometer was operated with an electrospray ionization (ESI) source in the positive mode. The following ionization source conditions were used: capillary voltage of 2.0 kV, extractor voltage of 3 V, RF lens voltage of 2.5 V, cone voltage of 15–40 V, collision energy of 15–30 eV, source temperature of 150 °C and desolvation temperature of 400 °C. The cone and desolvation gas flows were 50 and 800 L/h, respectively, which were obtained using a nitrogen source (N. Supplier Model 24S, Anest Iwata Co., Yokohama, Japan).

**Accuracy and Precision** Olive oil (0.05 g) samples were diluted by adding 5 mL of isopropanol. Then, 1.0 mL was diluted by adding water/methanol (1000 times dilution), added with 100 µL of a mixed internal standard (detected level of 10 ng/mL) and 1 mL of mixed standard solution (detected level of 500 ng/mL). The accuracy value was defined as follows: \( F/F_0 + A \times 100 \% \), where \( F \) and \( F_0 \) are the concentrations of the analytes in the spiked and unspiked samples, respectively, and \( A \) is the spiked concentration. Using olive oil, it is the \( F_0 = 0 \). For the precision, the same procedure was repeated for intra-day (three replicates for one day, \( n = 3 \) for standard deviation, ±S.D., %).

**Application for Convertible THC in Acetic Acid Condition** The 0.1 mL of acetic acid was added into CBD oil (1.0 g) samples, and the sample was heated to 70 °C on a heat block for 6 h. After that, 50 mg of samples were diluted with 5 mL of isopropanol. The 1.0 mL was diluted 1000 times with water/methanol before being mixed with 20 µL of a mixed internal standard (detected level of 100 ng/mL) for LC-MS/MS assay.

**Quantitative Procedure** Fixed concentrations of the stan-
Standard solutions (LOQ – 1000 ng/mL) were prepared by sequential dilutions of the stock solutions. The LOD and LOQ values were evaluated based on the signal-to-noise ratio (S/N) obtained while detecting the concentration of analytes and indicated $S/N = 3$ and $S/N > 10$. The calibration curves were built from eleven different concentrations to assess the linearity at each concentration level by plotting the peak-area ratio of the standard solutions: internal standard (y) vs. each concentration of the adjusted standard solution (x) (curve typesetting: linear, origin: exclude, weighting: $1/x$, axis: none).

Fig. 2. Typical Selected Reaction Monitoring (SRM) Chromatograms of Targeted CBD and Other Cannabinoids

a)–f) Standard solution (125 ng/mL). g)–l) CBD oil (No. 1). m)–r) CBD oil (No. 5).

Table 1. Accuracy Test of Targeted CBD and Other Cannabinoids in Olive Oil

| Analytes | Accuracy ± S.D. (%) |
|----------|---------------------|
| CBDA     | 99.7 ± 2.4          |
| CBGA     | 99.6 ± 2.3          |
| CBG      | 100 ± 1.4           |
| CBD      | 99.9 ± 7.1          |
| THC      | 99.7 ± 7.7          |
| CBN      | 99.1 ± 5.8          |

For the precision, the same procedure was repeated for intra-day (three replicates for one day, $n = 3$ for standard deviation, ± S.D., %).

Table 2. CBD Concentration Levels of CBD Oil from the Japanese Market

| Sample No. | Labeled value of CBD (mg/g) | Detectable value of CBD (mg/g) |
|------------|----------------------------|-------------------------------|
| 1          | 72.1                       | 60.6                          |
| 2          | 55.6                       | 43.9                          |
| 3          | 76.5                       | 66.0                          |
| 4          | 65.6                       | 57.8                          |
| 5          | 32.8                       | N.D. (<2.0)                   |
Results and Discussion

LC-MS/MS Assay for CBD Profiling  In this study, the targeted CBD profiling was shown in Fig. 1 and selected 6 chemicals. In this profiling, the psychoactive/illegal cannabinoids are THC and THCA in Japan. According to drug law, the THC standard solution was legally obtained from the imported U.S.A. However, due to a procedural issue that is monitored to the SRM with the same molecule weight of CBDA ($m/z$ 359 > 219), we are unable to obtain the THCA standard in the study.11) In addition, the minor oxidized CBD (CBND) standard was not obtained from common route and monitored to the SRM with the same molecule weight of CBN ($m/z$ 311 > 223).

![Fig. 3. Investigation of Convertible THC in Acetic Acid Condition](image)

a) Investigation of heating time for convertible THC (three replicates for one day). b) Investigation of heating temperature for convertible THC (three replicates for one day). c) Investigation of acetic acid for convertible THC (three replicates for one day). d) SRM chromatograms (upper $m/z$ 315 > 193 for analytes and lower $m/z$ 318 > 196 for IS) of CBD oil (No. 2) with 10% acetic acid at room temperature for 6h. e) SRM chromatograms (upper $m/z$ 315 > 193 for analytes and lower $m/z$ 318 > 196 for IS) of CBD oil (No. 2) with 10% acetic acid at high temperature (70°C) for 6h.
Supplementary Table S1 shows the MS conditions such as monitoring ion, cone voltage, and collision energy. Following that, it was determined that the resolution in SRM chromatograms of CBG (m/z 317 > 193), CBD (m/z 315 > 193), and THC (m/z 313 > 193) should be examined using RP columns. Specifically, these resolution values (Rs) of CBG/CBD are performed such as TSKgel ODS-100V (Rs = 0.40), Inertsustain AQ-C18 (Rs = 0.60), TSKgel ODS-120H (Rs = 0.70), Acquity UPLC BEH C18 (Rs = 0.86), TSKgel ODS-100Z (Rs = 1.33), Atlantis T3 (Rs = 1.49) and Inertis ODS-HL (Rs = 1.56), respectively. As a result of this study, the Inertsil ODS-HL column was found to be a valid and feasible column for LC-MS/MS analysis of CBD profiling. Figures 2-a)–f) and Supplementary Table S1 shows the SRM chromatograms and quantitative performance.

**Evaluation of CBD Oils** For the validation test, the standard solutions were spiked in an olive oil sample (n = 3). The spiked olive oils were pretreated and subjected to internal standard and absolute calibrations (Table 1). These accuracy and precision values were also presented in our previous report. In addition, a similar preparation using isopropanol and precision values were also presented in our previous report considering import of CBD products such as CBD oil. The authors declare no conflict of interest.

**Conflict of Interest** The authors declare no conflict of interest.

**Supplementary Materials** This article contains supplementary materials.

**References**

1. McGregor I. S., Cairns E. A., Abelev S., Cohen R., Henderson M., Couch D., Arnold J. C., Gauld N., Int. J. Drug Policy, 85, 103935 (2020).
2. The narcotics control department of Japan, “For those who are considering import of CBD products such as CBD oil.” https://www.nccd.mhlw.go.jp/pdl_data/cbd/guidecbd_en.pdf, cited May, 2020.
3. Gaoni Y., Mechoulam R., Tetrahedron, 22, 1481-1482 (1966).
4. Mechoulam R., Hanus L., Chem. Phys. Lipids, 121, 35-43 (2002).
5. Marzullo P., Foschi F., Coppini D. A., Fanchini F., Maynarri L., Rusconi S., Luzzani M., Passarella D., J. Nat. Prod., 83, 2894-2901 (2020).
6. Pratap Singh A., Fathordoobady F., Guo Y., Singh A., Kats D. D., Sci. Rep., 10, 10567 (2020).
7. Kiselak T. D., Koerbier R., Verbeck G. F., Forensic Sci. Int., 308, 110173 (2020).
8. Pacifici R., Marchei E., Salvatore F., Guandalini L., Busardo F. P., Pichini S., Clin. Chem. Lab. Med., 55, 1555-1563 (2017).
9. Brighenti V., Licata M., Pedrazzi T., Maran D., Bertelli D., Pelliati F., Benedetti S., J. Chromatogr. A., 1597, 179-185 (2019).
10. Meng Q., Buchanan B., Zuccolo J., Poulin M. M., Gabriell J., Be-
ranowski D. C., PLOS ONE, 13, e0196396 (2018).

1) Nemeškalová A., Hájková K., Mikulů L., Sýkora D., Kuchař M., Talanta, 219, 121250 (2020).
2) McRae G., Melanson J. E., Anal. Bioanal. Chem., 412, 7381–7393 (2020).
3) Takashina S., Igarashi Y., Takahashi M., Kondo Y., Inoue K., Anal. Sci., 36, 1427–1430 (2020).
4) Jang E., Kim H., Jang S., Lee J., Baeck S., In S., Kim E., Kim Y. U., Han E., Forensic Sci. Int., 306, 110064 (2020).

15) Gorley B. J., Murphy T. P., Gul W., Walker L. A., ElSohly M., J. Diet, 17 (Suppl.), 599–607 (2020).
16) Ministry of Health, Labour and Welfare, Japan, “About THC included product.”: https://www.mhlw.go.jp/stf/seisakunitsuite/bunya/kenkou_iryou/iyakuhin/yakubuturanyou/other/torishimari_00004.html, cited February, 2021.
17) Adams R., Pease D., Cain C., Baker B., Clark J., Wolff H., Wearn R., J. Am. Chem. Soc., 62, 2245–2246 (1940).