Quinalphos Induced Alterations in the Levels of Ions and Whole Animal Oxygen Consumption of Freshwater Fish, *Cyprinus Carpio* (Linnaeus, 1758)

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Abstract

The fingerlings of *Cyprinus carpio* were exposed to the lethal concentration (7.5 μg/l) of quinalphos (Emulsified Concentrate 25 percent) for 1, 2, 3 and 4 days respectively to study the levels and whole animal oxygen consumption. Exposed fish showed a significant decrease in the whole animal oxygen consumption due to the respiratory distress as a consequence of the impairment of oxidative metabolism. The ionic content of Na+, K+ and Ca²⁺ also decreased significantly in gill, liver and muscle tissue of exposed fish, signifying the altered cellular proliferation and deranged Na⁺, K⁺ and Ca²⁺ ionic pump due to probable consequence of tissue damage.

Keywords: Ions; Toxicity; Gill; Liver; Muscle; Calcium; Sodium; Potassium; Quinalphos

Introduction

The use of insecticides is being increase in the recent years to control the pest in which only 1% of the pesticide applied hits the target pest while, the remaining 99% of the pesticide drifts into the environment contaminating soil, water and biota [1]. This poses a constant threat to the non-target organisms especially to fishes; because pesticides are known to alter their behaviour pattern, growth, nutritional value and physiology. One of the early symptoms of acute pesticide poisoning is failure of respiratory metabolism. Some pesticides decrease the oxygen uptake of fish. The rate of oxygen consumption can be used as a bio-detector in monitoring the physiological effects of pesticides and the oxygen consumption pattern will indicate the possible mapping of metabolic pathways influenced by the pesticide stress [2]. Freshwater fishes are hyperosmotic to their medium. They gain water osmotically and tend to loosed solute by diffusion. In the regulation of osmolarity of system sodium, potassium and calcium ions play a significant role to keep the hyperosmotic properties of these animals [3].

Quinalphos is one of the organophosphate insecticide extensively used in agriculture in our area. The Environment Centre of National Toxicology declared that, there are a dozen highly dangerous chemicals including quinalphos. Quinalphos is a hard insecticide, which has become a matter of concern because of its potentiality and hazardous effect. The fish *Cyprinus carpio* was selected as experimental model because of its wide availability in local tanks and ponds of our area. It is also serves as a cheap protein rich source of food for vast population of India.

Hence in the present study, an attempt has been made to determine the relation between tissue ion content and oxygen consumption in fish, *Cyprinus carpio* exposed to quinalphos for different exposure periods.

Materials and Methods

*Cyprinus carpio* weighing 5±2 g and measuring an average length of 5 cm were collected from the State Fisheries Department, Dharwad, Karnataka, India and kept in large cement tank previously treated with potassium permanganate to clean it from microbial infection if any. All the experimental fish were acclimatized to laboratory conditions for 15 days with dechlorinated water whose physico-chemical character were analysed by following the APHA [4]. Organophosphate insecticide quinalphos (EC 25%) was obtained from Nagarjuna Chemical Ltd. Hyderabad, Andra Pradesh, India. The LC₅₀ value for 96 hr was determined by following the Finney [5] method and was found to be (7.5 μg/l). The fish were exposed to lethal concentration for 1, 2, 3, and 4 days.

Required quantity of pesticide quinalphos was added to the exposure container using variable micropipette. The oxygen consumption was measured for lethal concentration for 1, 2, 3, and 4 days by following the method of Welsh and Smith [6] and the apparatus set up was the same as described by Saroja [7]. After the stipulated time gill, muscle and liver isolated and weighted. They were wet ashed in 50:50 (v/v) concentrated perchloric acid and nitric acid [8] for half an hour until the organs were evaporated at 100°C to 200°C temperature. The residue obtained was dissolved in glass-distilled water and made up to 10ml. It was filtered through Whatman No.1 filter paper and appropriate dilutions were made prior to estimations. Na⁺, K⁺ and Ca²⁺ were estimated with the help of Atomic Absorbance Spectrophotometer (GBC 932 plus) and the values expressed in μg/g wet weight of the organ. The date were subjected to analysis of variance and the means were compared by Duncan’s new multiple range test at 0.05% level [9] to draw the mean comparison among the results.

Result

The rate of whole animal oxygen consumption (ml oxygen consumed per gram wet weight of fish per hour) of control and exposed fingerlings of *Cyprinus carpio* were studied for periods of 1, 2, 3 and 4 days. The rate of oxygen consumption due to respiratory distress was found to be the maximum in first day and remained significantly low in subsequent days in fish exposed to quinalphos. The oxygen consumption in gill, liver and muscle tissue of control fingerlings were significantly higher than that of exposed fish.

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quinalphos treated fishes are presented in (Table 1). However the data on the levels of sodium, potassium and calcium ions (μg/g wet weight of the organ) of the freshwater fish Cyprinus carpio to lethal concentration presented in (Table 2, Table 3 and 4).

Oxygen consumption

The data indicates that, the fish exposed to lethal concentrations of quinalphos oxygen consumption was reduced about 28.36% on day one and reached maximum reduction of 72.28% on day 4. The decrement was a sudden leap from day one to day two, from then it was gradual reduction up to day 4 (Table 1).

Sodium

In gill sodium ion shows a gradual decrease in lethal concentration from day 1 to day 4 ranging from 14.50% to 44.16%. In muscle similar fluctuation with 9.17% to 42.20% from day 1 to day 4 and in liver followed by 7.35% to 34.75% from day 1 to day 4 (Table 2, Figure 1).

Potassium

The potassium ion content in gill exhibited gradual decrement from day 1 to 4 ranging from 9.17% to 25.22%. In muscle a maximum decrease of about 32.33% observed on 4th day of the exposure period. However in liver the steady decrease of about 9.16% to 26.16% from day 1 to day 4 (Table 3, Figure 2).

Calcium

The calcium ion content also exhibited a steady decrement at lethal concentration. In gill, muscle and liver the maximum decrease was 18.45%, 27.66% and 22.36% respectively on day was recorded (Table 4, Figure 3). All the alterations were significant at 0.05% levels.

Discussion

Since most fish breathe in the water in which they live, changes in the chemical properties thereof may be reflected in the animal's

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Estimation   | 0.5722±a         | 0.4998±b         | 0.4062±c         | 0.2299±d         |
| SD±          | 0.0003           | 0.0004           | 0.0003           | 0.0004           |
| % change     | -28.36           | -33.11           | -59.82           | -72.28           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Gill         | 0.5405±a         | 0.4884±b         | 0.4240±c         | 0.3365±d         |
| SD±          | 0.0003           | 0.0001           | 0.0002           | 0.0001           |
| % change     | -14.52           | -22.30           | -38.33           | -44.16           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Gill         | 0.9937±a         | 0.9025±b         | 0.8737±c         | 0.8152±d         |
| SD±          | 0.0002           | 0.0002           | 0.0003           | 0.0003           |
| % change     | -13.90           | -15.89           | -18.48           | -22.36           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Gill         | 0.8627±a         | 0.7932±b         | 0.7457±c         | 0.7217±d         |
| SD±          | 0.0002           | 0.0002           | 0.0002           | 0.0002           |
| % change     | -14.33           | -22.36           | -38.36           | -44.16           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Gill         | 0.9732±a         | 0.9483±b         | 0.9033±c         | 0.8775±d         |
| SD±          | 0.0002           | 0.0002           | 0.0002           | 0.0002           |
| % change     | -11.68           | -13.59           | -16.34           | -18.95           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Gill         | 1.2352±a         | 1.0538±b         | 0.9228±c         | 0.8847±d         |
| SD±          | 0.0003           | 0.0002           | 0.0002           | 0.0002           |
| % change     | -13.30           | -22.36           | -38.36           | -44.16           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Gill         | 0.9138±a         | 0.8833±b         | 0.8663±c         | 0.7986±d         |
| SD±          | 0.0002           | 0.0002           | 0.0002           | 0.0002           |
| % change     | -3.20            | -5.08            | -12.70           | -22.63           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.

| Tissue       | Control          | Lethal Exposure Period in Days |
|--------------|------------------|-------------------------------|
|              | Day 1            | Day 2            | Day 3            | Day 4            |
| Gill         | 0.9872±a         | 0.9475±b         | 0.9033±c         | 0.8775±d         |
| SD±          | 0.0002           | 0.0002           | 0.0002           | 0.0002           |
| % change     | -11.68           | -13.59           | -16.34           | -18.95           |

Data are means ± SD (n = 6) for an organ in a row followed by the same letter are not significantly different (p < 0.05) from each other according to Duncan’s multiple range test.
ventilatory, activity, particularly if the environment factors affect respiratory gas exchanges [10]. The fluctuated response in respiration may be attributed to reduction in gill permeability causing a drop in oxygen consumption for which the fish compensates by increasing the ventilation volume observed by Kalavathy et al. [11]. Several authors suggested that the gills are the major respiratory organs and all metabolic pathways depend upon the efficiency of the gill for their energy supply and any damage to this vital organ causes a chain of destructive events, which ultimately lead to respiratory distress [12, 13]. According to Natarajan et al. [14] the decrease in oxygen consumption uptake from pesticide water is mainly due to shrinkage of the respiratory epithelium since there is swelling of the secondary lamellar tips. The gill opercular movements increased initially to support enhanced physiological activities in stressful habitat and later decrease, possibly due to mucus accumulation on gills. This been reported by David et al. [15] and Devaprakasa Raju, [16]. The coagulation of mucus on the gill caused demolition of various important processes such as gas exchange, nitrogen excretion, salt balance and circulation of blood [17]. Decrease in oxygen up take efficiently was noted in rainbow trout exposed to fenvalerate and cypermetrin by Bradbury et al. [18]. Similar situation was observed by in Cirrhinus mrigal and Labeo rohita, exposed to fenvalerate [2]. The decrease in oxygen consumption appears to be a protective measure to ensure that there is low intake of the toxic substance.

In fishes, gills form major site for ion transport and osmotic water movements, hence also for pesticide entry. They are in constant touch with the water. This affects the permeability characteristics and osmoregulatory function of the gills there by resulting in the decrease of these ions in gill tissue upon exposure to quinalphos. An appraisal of result in the present study suggests that the sodium, potassium and calcium ions in gill, muscle and liver exposed to lethal concentration of quinalphos indicates change in the permeable properties of the cell membrane of these organs and of deranged Na\(^+\), K\(^+\) and Ca\(^{2+}\) ionic pump due to the probable consequences of tissue damage [19]. It is known that Na\(^+\) pump which regulates the ionic content of the tissues. The level of Na\(^+\) signifies the importance of tissue in the mobilization of the water transport, since sodium content in the membrane facilitates the water movement among the tissues [20]. In this investigation, it is evident that Na\(^+\) loss is higher in the case of gill indicating the derangement in Na\(^+\) transport and rupture in the respiratory epithelium of gill tissue [21].

It is known that any remarkable decrease in K\(^+\) ion level might be accompanied by serious disturbance in muscular irritability, myocardial function and respiration [22]. The creases in K\(^+\) ion content in the tissues of Cyprinus carpio exposed to quinalphos might attribute to the derangement in whole animal oxygen consumption and ionic content at tissue levels as observed in the present investigation. The decrease in Ca\(^{2+}\) ion level indicates increased decalcification. Ca\(^{2+}\) is concerned with neuromuscular excitability, cell membrane permeability and regulation of protein binding capacity [23]. In the present study, the restlessness in fish during organophosphate stress corresponds to structural change in mitochondrial integrity. Since mitochondria acts as “sink” for intracellular Ca\(^{2+}\) and as principle store house of Ca\(^{2+}\), it appears that Ca\(^{2+}\) decrease may be attributed to the disturbance in mitochondrial integrity and subsequent respiratory distress [24].

Decrease in Ca\(^{2+}\) content may also be due to deranged neural transmission and acetylcholinesterase activity during quinalphos toxicity. Since it was reported that the inhibition of acetylcholinesterase activity and subsequent accumulation of acetylcholine at the nerve
ending accounts for decrease Ca\(^{2+}\) content [25]. Hoar [26] suggested that the levels of amino acids and metabolites like pyruvate and lactate will be increase under stress conditions to compensate the loss of inorganic ions. In support to this, amino acids and lactate were found increased in tissues of *Catla catla* exposed to endosulfan [10].

Thus, is quite apparent that the fish, *Cyprinus carpio* under quinalphos stress seldom under goes total abolition of functional regulation of the ion transport and water permeability system and the imbalance in osmoregulation is compensated in harmonic manner through the production of biochemically changed components like amino acids which go to rescue and compensate the imbalance ionic composition. Thus, an intrinsic osmoregulatory ability of fish is viewed in a nutshell but clear understanding on the basis of water permeability versus Na\(^{+}\) transport and the position of Ca\(^{2+}\) in water transport mechanisms still awaits direct analysis and will certainly throw light on the principles behind this.

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