Multicenter evaluation of EIAs and CLIAs in the detection of hepatitis C virus antibody among blood donors in China

Le Chang  
Beijing Hospital

Huimin Ji  
Beijing Hospital

Fei Guo  
Beijing Hospital

Xinyi Jiang  
Beijing Hospital

Lunan Wang (✉ lunan99@163.com)  
Beijing Hospital, Graduate School, Peking Union Medical College, Chinese Academy of Medical Sciences  
https://orcid.org/0000-0002-8208-1718

Research article

Keywords: Hepatitis C virus antibodies, enzyme immunoassay, chemiluminescent immunoassay, blood donations

Posted Date: January 13th, 2021

DOI: https://doi.org/10.21203/rs.3.rs-47853/v3

License: ☺️ ☀️ This work is licensed under a Creative Commons Attribution 4.0 International License.  
Read Full License
Abstract

**Background.** Recent data on anti-HCV screening assays among large population of blood donors is limited. The present study aimed to perform a multicenter evaluation of EIAs and CLIAs for detection of anti-HCV among blood donors in 16 different Chinese blood establishments.

**Methods.** A total of 1,309 samples including 582 anti-HCV screening negatives and 727 positives collected from 15 blood establishments all over China. Ten different anti-HCV assays (eight EIAs and two CLIAs) were evaluated in 16 different blood centers/banks. Confirmatory testing was performed using recombinant immunoblot assay and HCV RNA tests.

**Results.** The plasma panel contained 963 negative samples, 261 positives, and 85 indeterminate samples, based on the results of confirmatory test. False positive rate of screening tests was 39.67% (382/963) and the positive prediction value was only 35.76% (260/727). Among ten anti-HCV assays, Roche and InTec had the highest sensitivity (98.47%), while KHB and Wantai (indirect) had the highest specificity (99.23%). Then we analyzed the combined performance of these assays with two assays’ strategy widely used in China: Ortho or Abbott together with InTec could find all the true positives and Wantai (indirect) with Livzon (sandwich) got a highest specificity of 97.80%. Indeterminate samples showed quite different signal to cutoff (S/CO) ratios tested by Roche compared with confirmed positives (4.84 vs 19.36, p<0.0001), and higher S/CO ratios than confirmed negatives (4.84 vs 2.94, p=0.020).

**Conclusion.** False reactivity in anti-HCV screening should be treated as urgent issue. RIBA indeterminate donations may be a special group, while it still worth to be further studied.

Background

Hepatitis C virus (HCV) infection is of special concern due to a rising burden of its related cirrhosis, hepatocellular carcinoma (HCC), and liver related death worldwide [1-3]. In 2015, the global prevalence of viremia HCV is estimated to be 1.0%, which means approximately 71.1 million population worldwide are suffering from Hepatitis C and HCV-related diseases [4]. Especially in China, the proportion is about 0.7% (more than 9 million people are infected).

HCV is also a blood-borne virus. Laboratory diagnosis of HCV infection is usually made on the basis of the detection of circulating antibodies and/or HCV RNA [5]. Serological tests are of great importance to preventing transfusion transmitted infections, especially because large majority of HCV infection are anti-HCV reactive and HCV RNA negative [6]. Enzyme immunoassay (EIA) and chemiluminescent immunoassay (CLIA) are two kinds of widely used serological screening methods. In China, serological screening strategy of HCV infection involves two different anti-HCV EIA assays that are approved by National Medical Products Administration (NMPA) and if they show inconsistent results, twice repeated tests using the reactive one are performed. When any of the two EIAs is repeatedly reactive, the donation is disqualified regardless of the NAT results and the donor is deferred for safety [7]. CLIA, which has been
widely used in recent years, can automatically and quickly test anti-HCV antibodies from serum/plasma samples with higher sensitivity and specificity but haven’t been used in Chinese blood establishments yet.

Recent data on anti-HCV screening assays among large population of blood donors is limited. This information is of critical value to analyze and find difference among different assays and improve assays’ performance. The present study aimed to perform a multicenter evaluation of enzyme immunoassays and chemiluminescent immunoassays for detection of antibodies against hepatitis C virus among blood donors in 16 different Chinese blood establishments.

**Methods**

**Plasma panel**

A total of 1,309 plasma samples collected by 15 different blood establishments during Apr 2015 to Sep 2015, including Changchun (abbreviated as CC), Chongqing (CQ), Hebei (HB), Heilongjiang (HLJ), Henan (HN), Heze (HZ), Jining (JN), Jiangsu (JS), Kuming (KM), Liaoning (LN), Shandong (SD), Shenzhen (SZ), Tianjin (TJ), Tongzhou (TZ), Xiangyang (XY) blood center/blood bank. Among them, there were 582 anti-HCV negative randomly selected donations (HBsAg, anti-HCV, anti-HIV, and anti-TP antibodies were all negative with unqualified ALT level) and all the 727 anti-HCV positive donations detected in blood establishments during the period were included. All the plasma bags were sent to the Chinese National Center for Clinical Laboratories (NCCL), confirmed, split, numbered, stored at -40°C, and then sent back to all the participants. The transportation of the plasma panel was at 2-8°C and within 3 days. Sample source were listed in Table 1. All these samples were from 1,309 blood donors, no consecutive samples from one donor included.

**Study design**

16 different blood centers or blood banks over China participated into this multicenter evaluation, including CC, CQ, Cangzhou (CZ), HB, HLJ, HN, JN, JS, KM, LN, SD, Shanghai (SH), SZ, TJ, TZ, XY blood center/bank. The moment all these establishments above received the panel, they used one or two different EIAs to test it within 3 days by automated detection systems. Each sample was only tested once by one assay. All the results were sent back to NCCL within 10 days. Besides, NCCL tested all the samples once by two different CLIA assays simultaneously. All results were performed according to the manufacturers’ instructions.

**Confirmatory tests and evaluated assays**

All the donation samples were subjected to confirmatory testing with anti-HCV recombinant immunoblot assay (Mikrogen Diagnostic, Martinsried, Germany), which showed the results as positive, negative, and indeterminate (IND). All the indeterminate samples were further tested HCV RNA by Cobas Taqscreen MPX Test, Version 2 (Roche Molecular systems, Branchburg, NJ). All results were performed according to the manufacturers’ instructions. Technical specifications of confirmatory assay see Table 2. There were
10 different commercially available anti-HCV assays in this study. Technical specifications of evaluated assays see Table 2.

Data and Statistical analysis

Since plasma panel was tested by different blood establishments and different evaluated assays in the study, majority results on one sample would be treated as a “true value”, if not all results from different blood establishments were consistent.

The sensitivity, specificity, Youden’s index (The formula was below) and Kappa coefficient of each assay and combined assays were calculated by SPSS software (v.21.0). Results were analyzed with One-Way ANOVA analysis and Chi-square test. \( p<0.05 \) was considered as statistically significance.

Sensitivity = number of positive results/ number of confirmed positive in %;

Specificity = number of negative results/ number of confirmed negative in %;

Youden's index = Sensitivity+Specificity-1

Results

Confirmatory results of plasma panel

In 1,309 plasma samples, 963 were true negative, 261 were true positive by confirmatory testing. 85 samples were indeterminate by immunoblot and HCV RNA were also undetectable. The final results and validation algorithm see Figure 1. In 727 anti-HCV initial reactive samples only 260 were true positive, indicating the average positive prediction value (PPV) of anti-HCV screening tests was only 35.76% (260/727) and the average false positive rate was as high as 39.67% (382/963). Besides, we also found one true positive samples from Liaoning blood center were undetected by screening assays. The confirmatory results from each establishments was listed in Table 1.

Performance of evaluated assays

Among all the true negatives, 724 (724/963, 75.18%) were detected negative by all 10 assays. 209 (209/261, 80.08%) positives were detected positive by all assays. 376 samples showed discrepant results, among which there were 52 confirmed positive samples could not be detected by all evaluated assays (Table S1). InTec and Roche detected 48 samples, while Abbott only detected 29 ones. All samples had positive results of antibody against HCV protein Core 1, while other antibodies were weaker than confirmed positive samples with consistent results by all evaluated assays. On the basis of assay's results of 963 true negatives and 261 true positives, we calculated the sensitivity, specificity, Youden's index and Kappa Coefficient. The results showed assays’ sensitivity varied from 91.19% to 98.47%, InTec and Roche owned the highest sensitivity (98.47%), while sensitivity of Abbott was the lowest (91.19%). As a routine strategy in China, two different kinds of EIAs are used simultaneously. Thus we calculated the
combined performance index between two different assays. The combined sensitivity showed that when we use InTec and Ortho, 100% positives could be detected. Similarly, different assays showed different specificity (92.94%-98.23%), KHB was the highest, while InTec was the lowest. When Wantai (sandwich) and Wantai (indirect) were used together, we could get the highest combined specificity (97.92%). Youden's index, a way of summarizing the performance of a diagnostic test, equals sensitivity plus specificity minus 1. Roche got the highest index (96.08%) and the two EIAs, KHB and Livzon (indirect), owned the highest combined Youden's index. Results of Kappa coefficient, a statistic which measures inter-rater agreement, indicated that Wantai (indirect) had perfect agreement with confirmatory results, and Wantai (indirect) and Livzon (sandwich) together owned the highest kappa coefficient. As a whole, CLIA didn't perform better than EIAs on the detection of anti-HCV in this study. All the detail results see Table 3.

**Indeterminate samples**

There were 85 samples indeterminate by confirmatory testing and HCV RNA undetected. We further explored the antigen distribution and found there were six different formats: core1 single, helicase single, helicase/NS3, NS3/NS4, NS4/NS5 and NS4/NS5, among which core1 single was the majority (53/85, 62.35%), next was helicase single (17/85, 20%). Only three samples belong to last three formats. Different assays showed quite different results on these indeterminate samples: of all the indeterminate samples, 45 were detected by Abbott, while Wantai (sandwich) and Murex got only 25 positives. In the aspect of antigen distribution among different assays, most assays showed similar results as the total, except Murex. Helicase single in Murex positive indeterminate samples accounted for a main proportion (40%), next is core1 single (32%) and Helicase/NS3 format (28%).

Besides, we also analyzed the S/CO ratios of Roche and Abbott among 367 samples with discrepant results. There was statistically significant between the indeterminate and positive (4.84 vs 19.36, \( p<0.0001 \)), between negative and positive (2.94 vs 19.36, \( p<0.0001 \)) and between determinate and negative (4.84 vs 2.94, \( p=0.020 \)) detected by Roche. However, we didn't find any difference among the three groups when detected by Abbott (positive 1.40 vs indeterminate 1.74 vs negative 1.55, \( p=0.567 \)) (Figure 2)

**PPV of CLIA on different S/CO ratios**

In order to determine the relationship between PPV and S/CO ratio on two CLIA, we analyzed the assays results when different S/CO ratios were as the cutoff values of the two CLIA. When S/CO ratio went to 1.0, Roche's PPV on detection of anti-HCV was 92.09% (256/278), and Abbott was only 78.55% (238/303). When PPV was above 95% (246/258 on Roche, 196/205 on Abbott), cutoff S/CO ratios need to be more than 8.2 on Roche, and more than 4.2 on Abbott. When PPV was 100% (65/65 on Roche, 177/177 on Abbott), cutoff S/CO ratios need to be 46.8 on Roche, and 8.0 on Abbott. Results of ROC and AUC indicated that the two assays both showed a good AUC: Roche was 0.994 and Abbott was 0.984. However, the best S/CO was quite different on the two: S/CO ratio=1.515 was the best cutoff for Roche,
on this S/CO the assay’s sensitivity was 98.08% and the specificity was 97.82%. Since the relatively low sensitivity of Abbott, the best S/CO was only 0.385 (sensitivity=99.23%, specificity=86.50%).

Discussion

False positive reactions in anti-HCV screening assays is a well-known concern and an obstacle to improve assays’ performance all along. In the present study, we found that the average false positive rate of anti-HCV screening assays was as high as 52.54%, which meant large amount of donations were discarded and qualified donors were deferred unfortunately. It is reported in Uganda, 7.6% (76/1000) were serologically reactive by Ortho, but none were confirmed [8]. Similarly, Schroter M, et al [9] also found false positive results by widely used anti-HCV EIAs were at least 10%, which, compared to other virus screening assays, was unacceptable. In another study in Africa, only 13% of initial anti-HCV positive samples were confirmed by NAT or immunoblot and 2.46% (409/16,613) of blood donations were wrongly discarded [10].

Whereas the false positive rate in this study might partly result from the serological screening strategy in China. Use of two EIAs no doubt would relatively increase the sensitivity but sacrifice the specificity to a certain extent. In addition, among all screened positive samples in the panel, 40% were repeated reactive by single assay, which contributing only 1% confirmed positivity and leading to the high false positive rate. In the next place, samples used in screening tests were directly from blood collection tubes, while samples in the plasma panel were from plasma bag, which contained about 20% blood preserving fluid, diluting the samples and resulting in missing detection of some weak positives. Besides, researchers also found in low risk populations, 40–50% of screening reactives were negative in supplemental immunoblot tests, which was similar to our results [11].

Secondly, performance analysis of ten different assays indicated that generally all assays’ performance was worse than reported [12-20] or claimed by their instructions, which may be mainly due to sample source. Large amount of false positive samples increased the difficulty of testing a lot and false positive results of an assay during the first screening are likely to be false positives again. Specifically, Roche got the highest sensitivity (98.47%) and Youden’s index (0.961) but specificity (97.61%) was lower than four EIAs (KHB 98.23%, Wantai (indirect) 98.23%, Wantai (sandwich) 98.03% and Livzon (sandwich) 98.13%). The other CLIA, Abbott didn't perform well: it had the lowest sensitivity, but its results were highly comparable to those of Ortho, which also has been shown in other studies [18-21]. Therefore, Two CLIAs especially in the respect of specificity didn't show much better performance than EIAs, no matter imported assays or domestic ones.

We then analyzed the combined performance of any two EIAs assays. Interestingly, not two assays both with good sensitivity had the best combined sensitivity, but Ortho or Abbott together with InTec had a 100% sensitivity indicating these two assays just made up for each other's missing detection. In the terms of specificity, though use of two assays would definitely decrease detection specificity, we found that Wantai (indirect) together with Livzon (sandwich) got a 97.80% specificity, even better than only using
one assay (InTec 92.94%, Ortho 90.13%, Livzon(indirect) 93.15%, Murex 97.20%, Roche 97.61%, Abbott 93.25%).

Based on the performance of ten evaluated assays in the study, we recommended Roche for screening anti-HCV in blood donations and to those could only use EIA assays, Wantai (indirect), Wantai (sandwich), and Livzon (sandwich) also performed well. If blood establishments use two assays’ strategy when testing anti-HCV antibody, Wantai (indirect) with KHB, Wantai (indirect) with Livzon (sandwich), or Wantai (sandwich) with Livzon (sandwich) would be a better choice. In addition, a confirmatory testing is necessary for the initial positive samples.

Usually, an immunoblot test and/or NAT are served as a way to confirm screening reactive samples. However, the recombinant immunoblot assay as a confirmatory method for serological screening test of HCV, is of high cost and laborious to be used in every laboratory in China and these confirmatory tests often give many indeterminate and/or HCV RNA negative results, indicating the donor may be a resolved HCV infection, infection with a different genotype, early seroconversion, occult HCV infection or just non-specific reactivity [22], which makes us much difficult to estimate the donors’ true status. Like our results, we got 85 (6.49%, 85/1,309) indeterminate/HCV RNA negative samples, and most (53/85, 62.35%) were due to isolated reactivity to antigens from the core1 region of the HCV genome. We also found these indeterminate samples showed quite different average S/CO ratios detected by Roche compared with confirmed positives (4.84 vs 19.36, \(p<0.0001\)), and higher S/CO ratios than confirmed negatives (4.84 vs 2.94, \(p=0.020\)). Similarly to Kiely P’s results [22], the antigen distribution and band intensity were different between indeterminate samples and confirmed positives. All these results indicated indeterminate samples were a special group different from negative and positives.

Finally, we evaluated the corelationship between PPV and S/CO ratio on two CLIAas. When S/CO was 8.2 on Roche and 4.2 on Abbott, the PPV could achieve more than 95%. The S/CO ratio of ARCHITECT anti-HCV established for FDA–approved is more than 5.0 [5], similar to our results. But there is no such approved ratio on Elecsys Anti-HCV II. Some studies suggested that the value predictive of a true positive \(\geq95\%\) of the time with the assay could be set at an S/CO ratio of 20.0, much higher than our study [14, 16], which meant it was required to verify the value in further study.

There were some limitations in the study. First, not all the information of anti-HCV positive donors were available in this study, therefore we didn't know the true infection status of these donors. Secondly, different automated sampling and testing equipment were used in the 16 different blood establishments involved in the evaluation study, and all these difference may influence the detection accuracy.

**Conclusion**

Based on the multicenter evaluation of 10 common used anti-HCV screening assays, false reactive problem of anti-HCV screening should be solved urgently. Assays showed different performance on sensitivity and specificity and CLIAas didn’t performed better than EIAs, especially on specificity. Indeterminate samples may be a special group, should be further studied.
Abbreviations
HCV, Hepatitis C virus; EIA, Enzyme immunoassay; CLIA, Chemiluminescent immunoassay; NCCL, National Center for Clinical Laboratories.

Declarations

Ethics approval and consent to participate: The ethics committee of Beijing Hospital has approved the study. The methods in the study were in accordance with the guidelines of the Declaration of Helsinki. Written informed consent at the time of blood donation was obtained from all blood donors participating in this research.

Consent for publication: Not applicable

Availability of data and materials: The data generated and analyzed during the current study are available from the corresponding author on reasonable request.

Competing interests: None declared.

Funding: This work was supported by a grant from National Science and Technology Major Project of the Ministry of Science and Technology of China (Grant number 2018ZX10306412-004).

Authors’ contributions: All the authors have accepted responsibility for the entire content of this submitted manuscript and approved submission. L.N.W. and L.C. designed the study. L.C., H.M.J. and F.G. conducted the laboratory tests. H.M.J., F.G. and X.Y.J. collected and L.C. analyzed data and prepared the manuscript. All authors reviewed the manuscript and approved the final version of the manuscript and take responsibility for the integrity of the data and accuracy of data analysis.

Acknowledgements: We thank involved laboratory staff from blood establishments for participating in sample collection and testing.

References

1. Dore GJ, Hatzakis A, Negro F, Waked I. Estimating HCV disease burden-volume 4 (editorial). J Viral Hepat 2017;24 Suppl 2:4-7.

2. Chen DS, Hamoudi W, Mustapha B, Layden J, Nersesov A, Reic T, et al. Strategies to manage hepatitis C virus infection disease burden-Volume 4. J Viral Hepat 2017;24 Suppl 2:44-63.

3. Chan HLY, Chen CJ, Omede O, Al Qamish J, Al Naamani K, Bane A, et al. The present and future disease burden of hepatitis C virus infections with today’s treatment paradigm: Volume 4. J Viral Hepat 2017;24 Suppl 2:25-43.

4. Polaris Observatory HCVC. Global prevalence and genotype distribution of hepatitis C virus infection in 2015: a modelling study. Lancet Gastroenterol Hepatol 2017;2(3):161-76.
5. Kamili S, Drobeniuc J, Araujo AC, Hayden TM. Laboratory diagnostics for hepatitis C virus infection. Clin Infect Dis 2012;55 Suppl 1:S43-8.

6. Kuroishi A, Yasui K, Matsukura H, Tani Y, Furuta RA. Comparison of neutralization profiles for anti-HCV reactive donor samples with or without detectable HCV RNA. Vox Sang 2015;109(4):319-26.

7. Li L, Li KY, Yan K, Ou G, Li W, Wang J, et al. The History and Challenges of Blood Donor Screening in China. Transfus Med Rev 2017;31(2):89-93.

8. Mullis CE, Laeyendecker O, Reynolds SJ, Ocama P, Quinn J, Boaz I, et al. High frequency of false-positive hepatitis C virus enzyme-linked immunosorbent assay in Rakai, Uganda. Clin Infect Dis 2013;57(12):1747-50.

9. Schroter M, Feucht HH, Schafer P, Zollner B, Polywka S, Laufs R. Definition of false-positive reactions in screening for hepatitis C virus antibodies. J Clin Microbiol 1999;37(1):233-4.

10. Candotti D, Sauvage V, Cappy P, Boullahi MA, Bizimana P, Mbensa GO, et al. High rate of hepatitis C virus and human immunodeficiency virus false-positive results in serologic screening in sub-Saharan Africa: adverse impact on the blood supply. Transfusion 2020, 60(1):106-16.

11. Majid AM, Gretch DR. Current and future hepatitis C virus diagnostic testing: problems and advancements. Microbes Infect 2002;4(12):1227-36.

12. Seigneres B, Descamps F, Croise R, Barlet V, Bouvier-Alias M, Chevaliez S, et al. Multicenter clinical evaluation of the new 3rd generation assay for detection of antibodies against hepatitis C virus on the VIDAS((R)) system. J Clin Virol 2016;78:20-6.

13. Hyun J, Ko DH, Kang HJ, Whang DH, Cha YJ, Kim HS. Evaluation of the VIDAS Anti-HCV Assay for Detection of Hepatitis C Virus Infection. Ann Lab Med 2016;36(6):550-4.

14. Yoo SJ, Wang LL, Ning HC, Tao CM, Hirankarn N, Kuakarn S, et al. Evaluation of the Elecsys((R)) Anti-HCV II assay for routine hepatitis C virus screening of different Asian Pacific populations and detection of early infection. J Clin Virol 2015;64:20-7.

15. Sommese L, Sabia C, Paolillo R, Parente D, Capuano M, Iannone C, et al. Screening tests for hepatitis B virus, hepatitis C virus, and human immunodeficiency virus in blood donors: evaluation of two chemiluminescent immunoassay systems. Scand J Infect Dis 2014;46(9):660-4.

16. Yang R, Guan W, Wang Q, Liu Y, Wei L. Performance evaluation and comparison of the newly developed Elecsys anti-HCV II assay with other widely used assays. Clin Chim Acta 2013;426:95-101.

17. Esteban JI, van Helden J, Alborino F, Burgisser P, Cellera C, Pantaleo G, et al. Multicenter evaluation of the Elecsys(R) anti-HCV II assay for the diagnosis of hepatitis C virus infection. J Med Virol 2013;85(8):1362-8.

18. Berger A, Rabenau H, Allwinn R, Doerr HW. Evaluation of the new ARCHITECT anti-HCV screening test under routine laboratory conditions. J Clin Virol 2008;43(2):158-61.

19. Watterson JM, Stallcup P, Escamilla D, Chernay P, Reyes A, Trevino SC. Evaluation of the Ortho-Clinical Diagnostics Vitros ECi Anti-HCV test: comparison with three other methods. J Clin Lab Anal 2007;21(3):162-6.
20. Zachary P, Ullmann M, Djeddi S, Meyer N, Wendling MJ, Schvoerer E, et al. Evaluation of three commercially available hepatitis C virus antibody detection assays under the conditions of a clinical virology laboratory. J Clin Virol 2005;34(3):207-10; discussion 16-8.

21. Oethinger M, Mayo DR, Falcone J, Barua PK, Griffith BP. Efficiency of the ortho VITROS assay for detection of hepatitis C virus-specific antibodies increased by elimination of supplemental testing of samples with very low sample-to-cutoff ratios. J Clin Microbiol 2005;43(5):2477-80.

22. Kiely P, Styles C. Anti-HCV immunoblot indeterminate results in blood donors: non-specific reactivity or past exposure to HCV? Vox Sang 2017;112(6):542-8.

Tables

Table 1. Sample source and confirmatory results

| Samples source (blood establishments) | screening assays | anti-HCV- by screening assays | anti-HCV+ by screening assays | False negative % | False positive % |
|--------------------------------------|------------------|-----------------------------|-----------------------------|-----------------|-----------------|
|                                      |                  | total | - | + | total | - | + | +/- |                  |
| NCCL                                 | a-h              | 0     | 0 | 0 | 129   | 87  | 33 | 9   | 0.00             | 67.44           |
| CC                                   | a, e             | 11    | 11| 0 | 35    | 9   | 23 | 3   | 0.00             | 25.71           |
| CQ                                   | b, h             | 22    | 22| 0 | 48    | 24  | 16 | 8   | 0.00             | 50.00           |
| HB                                   | b, f             | 18    | 18| 0 | 48    | 31  | 12 | 5   | 0.00             | 64.58           |
| HLJ                                  | a, e             | 76    | 76| 0 | 30    | 12  | 16 | 2   | 0.00             | 40.00           |
| HN                                   | d, e             | 111   | 111| 0 | 66    | 31  | 23 | 12  | 0.00             | 46.97           |
| HZ                                   | a, e             | 21    | 21| 0 | 5     | 2   | 3  | 0   | 0.00             | 40.00           |
| JN                                   | a, b             | 0     | 0 | 0 | 4     | 4   | 0  | 0   | 0.00             | 100.00          |
| JS                                   | b, f             | 50    | 50| 0 | 50    | 33  | 14 | 3   | 0.00             | 66.00           |
| KM                                   | f, h             | 15    | 15| 0 | 11    | 3   | 6  | 2   | 0.00             | 27.27           |
| LN                                   | a, b             | 50    | 49| 1 | 37    | 12  | 19 | 6   | 2.00             | 32.43           |
| SD                                   | d, f             | 50    | 50| 0 | 49    | 23  | 21 | 5   | 0.00             | 46.94           |
| SZ                                   | c, d             | 50    | 50| 0 | 50    | 30  | 13 | 7   | 0.00             | 60.00           |
| TJ                                   | c, e             | 49    | 49| 0 | 101   | 43  | 42 | 16  | 0.00             | 42.57           |
| TZ                                   | c, h             | 50    | 50| 0 | 35    | 20  | 11 | 4   | 0.00             | 57.14           |
| XY                                   | a, b             | 9     | 9 | 0 | 29    | 18  | 8  | 3   | 0.00             | 62.07           |
| Total                                |                  | 582   | 581|1 | 727   | 382 | 260| 85  | 0.17             | 52.54           |

Note: a, KHB Diagnostic Kit for Antibody to Hepatitis C virus (Shanghai Kehua Bioengineering co., Shanghai, China); b, InTec Diagnostic Kit for Antibody to Hepatitis C virus (InTec Products, Inc., Xiamen, China); c, ORTHO HCV Version 3.0 ELISA (Ortho-Clinical Diagnostics, Raritan, New Jersey); d, Livzon Diagnostic Kit for Antibody to Hepatitis C virus (indirect) (Zhuhai Livzon Diagnostics Inc., Zhuhai, China); e, Wantai Diagnostic Kit for Antibody to Hepatitis C virus (indirect) (Beijing Wantai Biological Pharmacy, Beijing, China); f, Wantai Diagnostic Kit for Antibody to Hepatitis C virus (sandwich) (Beijing Wantai Biological Pharmacy, Beijing, China); g, Livzon Diagnostic Kit for Antibody to Hepatitis C virus (sandwich) Zhuhai Livzon Diagnostics Inc., Zhuhai, China); h, Murex anti-HCV (version 4.0) (Diasorin, Saluggia, Italy). -: true negative; +: true positive; +/-: indeterminate.
Table 2. Technical specifications of evaluated assays and confirmatory assay

| No. | Abbreviations of assays | Test format † | Antigens | Blood establishments |
|-----|--------------------------|--------------|----------|----------------------|
| a   | KHB                      | EIA, indirect| core, antigen, non-structure | CQ, HLJ, HN, XY |
| b   | InTec                    | EIA, indirect| N/M ‡    | CC, HB, JN, JS, XY, XY |
| c   | Ortho                    | EIA, indirect| core(c22-3), NS3+NS4(c200), NS5 | CQ, SZ, TJ |
| d   | Livzon (indirect)        | EIA, indirect| N/M      | CC, JS, SD |
| e   | Wantai (indirect)        | EIA, indirect| N/M      | CZ, LN, SD |
| f   | Wantai (sandwich)        | EIA, sandwich| N/M      | CZ, HB, HLJ, HN, LN, TJ |
| g   | Livzon (sandwich)        | EIA, sandwich| core, NS3, etc. | JN, LN |
| h   | Murex                    | EIA, indirect| core, NS3, NS4, NS5 | KM, SH, TZ |
| i   | Roche                    | ECLIA, sandwich| core, NS3, NS4 | NCCL |
| j   | Abbott                   | CMIA, indirect| core, NS3(33c), NS4(c100-3) | NCCL |
| k   | Mikrogen                 | recombinant immunoblot assay| core1, core2, helicase, NS3, NS4, NS5 | NCCL |

Note:
† EIA: enzyme immunoassay; ECLIA: electrochemiluminescence immunoassay; CMIA: chemiluminescent microparticle immunoassay.
‡ N/M: not mentioned in the package inserts.

Table 3. Performance index of different assays
Sensitivity (red), specificity (green), Youden’s index (blue), and Kappa Coefficient (purple) of single assay and two combined assays were shown. The darker the color, the better the performance the assays have. The number in the black box shows the best performance (sensitivity, specificity, Youden’s index, or Kappa Coefficient) of all the combined assays.

|                  | KHB     | InTec   | Ortho   | Livzon (indirect) | Wantai (indirect) | Wantai (sandwich) | Livzon (sandwich) | Murex | Roche  | Abbott |
|------------------|---------|---------|---------|-------------------|-------------------|-------------------|-------------------|-------|--------|--------|
| KHB              | 95.02%  |         |         |                   |                   |                   |                   |       |        |        |
| InTec            | 99.23%  | 98.47%  |         |                   |                   |                   |                   |       |        |        |
| Ortho            | 98.08%  | 100.00% | 92.72%  |                   |                   |                   |                   |       |        |        |
| Livzon (indirect)| 99.23%  | 98.85%  | 92.23%  | 91.05%            |                   |                   |                   |       |        |        |
| Wantai (indirect)| 98.85%  | 98.85%  | 99.62%  | 99.23%            | 97.70%            |                   |                   |       |        |        |
| Wantai (sandwich)| 98.47%  | 98.85%  | 92.23%  | 97.70%            | 97.32%            |                   |                   |       |        |        |
| Livzon (sandwich)| 98.47%  | 98.85%  | 92.23%  | 97.70%            | 97.32%            |                   |                   |       |        |        |
| Murex            | 96.17%  | 99.23%  | 96.55%  | 99.23%            | 98.08%            | 97.70%            | 97.70%            |       | 92.72% |        |
| Roche            | 98.85%  | 98.85%  | 100.00% | 99.23%            | 98.47%            | 98.50%            | 98.47%            | 98.85%| 98.47% |        |
| Abbott           | 96.93%  | 100.00% | 94.25%  | 98.85%            | 99.62%            | 99.23%            | 99.23%            | 95.79%| 99.62% | 91.19% |

|                  | KHB     | InTec   | Ortho   | Livzon (indirect) | Wantai (indirect) | Wantai (sandwich) | Livzon (sandwich) | Murex | Roche  | Abbott |
|------------------|---------|---------|---------|-------------------|-------------------|-------------------|-------------------|-------|--------|--------|
| KHB              | 98.23%  |         |         |                   |                   |                   |                   |       |        |        |
| InTec            | 91.80%  | 92.94%  |         |                   |                   |                   |                   |       |        |        |
| Ortho            | 89.72%  | 84.53%  | 90.13%  |                   |                   |                   |                   |       |        |        |
| Livzon (indirect)| 92.21%  | 87.23%  | 84.74%  | 93.15%            |                   |                   |                   |       |        |        |
| Wantai (indirect)| 96.88%  | 91.80%  | 88.70%  | 92.32%            | 98.23%            |                   |                   |       |        |        |
| Wantai (sandwich)| 96.68%  | 91.59%  | 88.58%  | 92.00%            | 98.03%            |                   |                   |       |        |        |
| Livzon (sandwich)| 96.79%  | 91.69%  | 88.79%  | 92.32%            | 97.72%            | 98.13%            |                   |       |        |        |
| Murex            | 95.85%  | 90.55%  | 88.60%  | 91.07%            | 95.74%            | 95.53%            | 95.64%            |       | 97.20% |        |
| Roche            | 96.26%  | 91.17%  | 88.27%  | 91.80%            | 97.40%            | 97.30%            | 97.40%            | 95.22%| 97.61% |        |
| Abbott           | 92.94%  | 86.71%  | 87.75%  | 87.54%            | 91.90%            | 91.69%            | 91.80%            | 91.38%| 91.48% | 93.25% |

|                  | KHB     | InTec   | Ortho   | Livzon (indirect) | Wantai (indirect) | Wantai (sandwich) | Livzon (sandwich) | Murex | Roche  | Abbott |
|------------------|---------|---------|---------|-------------------|-------------------|-------------------|-------------------|-------|--------|--------|
| KHB              | 0.933   |         |         |                   |                   |                   |                   |       |        |        |
| InTec            | 0.910   | 0.914   |         |                   |                   |                   |                   |       |        |        |
| Ortho            | 0.878   | 0.845   | 0.829   |                   |                   |                   |                   |       |        |        |
| Livzon (indirect)| 0.914   | 0.861   | 0.840   | 0.851             |                   |                   |                   |       |        |        |
| Wantai (indirect)| 0.957   | 0.906   | 0.884   | 0.915             | 0.959             |                   |                   |       |        |        |
| Wantai (sandwich)| 0.951   | 0.904   | 0.878   | 0.912             | 0.955             | 0.963             |                   |       |        |        |
| Livzon (sandwich)| 0.954   | 0.905   | 0.880   | 0.913             | 0.956             | 0.950             | 0.934             |       |        |        |
| Murex            | 0.920   | 0.898   | 0.846   | 0.903             | 0.938             | 0.932             | 0.933             | 0.899 |        |        |
| Roche            | 0.951   | 0.900   | 0.883   | 0.910             | 0.959             | 0.958             | 0.959             | 0.941 | 0.961  |        |
| Abbott           | 0.899   | 0.867   | 0.820   | 0.864             | 0.915             | 0.909             | 0.910             | 0.872 | 0.911  | 0.844  |

|                  | KHB     | InTec   | Ortho   | Livzon (indirect) | Wantai (indirect) | Wantai (sandwich) | Livzon (sandwich) | Murex | Roche  | Abbott |
|------------------|---------|---------|---------|-------------------|-------------------|-------------------|-------------------|-------|--------|--------|
| KHB              | 0.977   |         |         |                   |                   |                   |                   |       |        |        |
| InTec            | 0.822   | 0.839   |         |                   |                   |                   |                   |       |        |        |
| Ortho            | 0.776   | 0.700   | 0.749   |                   |                   |                   |                   |       |        |        |
| Livzon (indirect)| 0.830   | 0.737   | 0.698   | 0.801             |                   |                   |                   |       |        |        |
| Wantai (indirect)| 0.923   | 0.819   | 0.769   | 0.832             | 0.945             |                   |                   |       |        |        |
| Wantai (sandwich)| 0.916   | 0.816   | 0.763   | 0.826             | 0.936             | 0.938             |                   |       |        |        |
| Livzon (sandwich)| 0.918   | 0.818   | 0.767   | 0.822             | 0.938             | 0.931             | 0.940             |       |        |        |
| Murex            | 0.883   | 0.799   | 0.737   | 0.808             | 0.893             | 0.887             | 0.889             | 0.889 |        |        |
| Roche            | 0.909   | 0.808   | 0.762   | 0.822             | 0.931             | 0.929             | 0.931             | 0.887 | 0.936  |        |
| Abbott           | 0.829   | 0.736   | 0.717   | 0.743             | 0.826             | 0.820             | 0.792             | 0.792 | 0.798  |        |

Sensitivity (red), specificity (green), Youden’s index (blue), and Kappa Coefficient (purple) of single assay and two combined assays were shown. The darker the color, the better performance the assays have. The number in the black box shows the best performance (sensitivity, specificity, Youden’s index, or Kappa Coefficient) of all the combined assays.

Figures
Figure 1

Confirmatory results of plasma panel Note: +, POS: positive/reactive; -, NEG: negative/non-reactive; +/-, IND: indeterminate; R: Elecsys Anti-HCV II (Roche Diagnostics GmbH, Penzberg, Germany); A: ARCHITECT anti-HCV (Abbott, Illinois, US).
Figure 2

S/CO ratio distribution among confirmed positive, indeterminate and negative samples. Note: +: positive/reactive; -: negative/non-reactive; +/-: indeterminate; Roche: Elecsys Anti-HCV II (Roche Diagnostics GmbH, Penzberg, Germany); Abbott: ARCHITECT anti-HCV (Abbott, Illinois, US).

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- SupplementaryMaterial.docx