Von Hippel-Lindau Disease

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Abstract

A germline mutation in the Von-Hippel Lindau (VHL) gene predisposes carriers to development of abundantly vascularised tumours in the retina, cerebellum, spine, kidney, adrenal gland and pancreas. Most VHL patients die from the consequences of cerebellar haemangioblastoma or renal cell carcinoma. The VHL gene is a tumour suppressor gene and is involved in angiogenesis by regulation of the activity of hypoxia-inducible factor 1-α (HIF1-α). Clinical diagnosis of VHL can be confirmed by molecular genetic analysis of the VHL gene, which is informative in virtually all VHL families. A patient with (suspicion for) VHL is an indication for genetic counselling and periodical examination.

Introduction

A germline mutation in the Von-Hippel Lindau (VHL) gene predisposes carriers to development of abundantly vascularised tumours in multiple organs. These tumours may include haemangioblastoma in the retina (also referred to as retinal angioma), cerebellum and myelum, renal cell carcinoma (clear cell type), phaeochromocytoma, islet cell tumours of the pancreas, and endolymphatic sac tumours, as well as cysts and cystadenoma in the kidney, pancreas, epididymis and broad ligament (Fig. 1) [1-3]. VHL disease is an autosomal, dominant inherited tumour syndrome with an estimated prevalence of 2-3 per 100,000 persons (OMIM #193300) [4]. At present, metastases from renal cell carcinoma and neurological complications from cerebellar haemangioblastoma are the most common causes of death [5].

Using DNA diagnostics virtually all cases of classic VHL disease are identified, enabling early and presymptomatic diagnosis in families. Subsequently, periodical clinical examination and advanced operation techniques are likely to reduce both morbidity and mortality in patients with VHL disease.

Diagnostic criteria

In the presence of a positive family history, VHL disease can be diagnosed clinically in a patient with at least one
variants of VHL tumours. Typical VHL tumours are retinal, spinal, and cerebellar haemangioblastomas, renal cell carcinoma, and phaeochromocytoma. Endolymphatic sac tumours and multiple pancreatic cysts suggest a positive carriership (in the presence of a positive VHL family history), since they are uncommon in the general population. In contrast, renal and epididymal cysts occur frequently in the general population and are, as sole manifestation, not reliable indicators for VHL disease. In patients with a negative family history of VHL-associated tumours, a diagnosis of VHL disease can also be made when they exhibit two or more haemangioblastomas, or a single haemangioblastoma in association with another typical manifestation [3].

Molecular genetics

The gene that, in mutated form, is responsible for the disease is located on chromosome 3 (3p25-26) [6]. The VHL gene is a relatively small gene that covers approximately 14,500 basepairs of genomic DNA. The VHL gene encodes a ubiquitously expressed messenger RNA of 4,700 nucleotides and the protein-coding region is contained in three exons. Germline mutation analysis has revealed two mutation hotspots that concur with important functional domains of the VHL protein (Fig. 2).

The VHL gene is a tumour suppressor gene according to Knudson’s ‘two-hit’ hypothesis [7]: for a normal cell, inactivation of both copies of the VHL gene is required to develop into a tumour cell. In carriers of a VHL gene germline mutation, tumours tend to occur multicentrically and bilaterally, and manifest at a younger age than in patients without a VHL gene germline mutation, where each of the two VHL gene alleles in a cell has to become affected by an independent hit at the somatic level.

VHL protein

The VHL gene encodes a 213 amino acid protein that does not closely resemble any other human protein. The VHL protein resides predominantly in the cytoplasm and is widely expressed in normal human tissues [8]. Strong expression is detected in target tissues of the disease, such as cerebellar Purkinje cells, proximal and distal renal tubules and exocrine pancreas. The VHL protein is also expressed in organs not at risk for the disease, such as the pituitary gland, colon and thyroid. The protein plays a vital role in embryonic development. Foetal mice deprived of both copies of the VHL gene are not viable. In human embryos, the VHL protein is expressed in all three germ layers [9].

The well-vascularised phenotype of VHL tumours suggests that inactivation of the VHL gene induces either upregulation of an angiogenic factor or downregulation of an inhibitor of angiogenesis. In VHL-associated haemangioblastoma and renal cell carcinoma, various proteins that are involved in angiogenesis are upregulated. These angiogenic proteins include vascular endothelial growth factor (VEGF), plasminogen activator inhibitor 1 (PAI-1) and erythropoietin [10-12]. The genes coding for these proteins have in common that they are regulated by hypoxaemia. During hypoxaemia a transcription factor, hypoxia-inducible factor (HIF)-1α, binds to the promoters of these genes and stimulates their transcription [10]. In normoxic conditions HIF-1α binds to the β-domain of the VHL protein (Fig. 3). The α-domain of the VHL protein binds to Elongin C, which is connected with Elongin B in a multi-protein complex (Fig. 3) [13]. This complex targets the HIF-1α substrate for degradation in a proteasome via a process called ubiquitination. Consequently, excessive blood vessel formation may occur when hypoxia-inducible proteins are not properly degraded. In other words, in cells lacking...
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Fig. 2. Functional domains of the VHL protein and distribution of germline mutations (from VHL database) [22]. Hotspots for mutations are readily visible at amino acids 67-117, hypoxia-inducible factor (HIF)-1α binding domain (in the β-domain) and 157-170, Elongin C (EloC) binding domain (in the α-domain). Copyright 2003, The Endocrine Society [16]

wild-type VHL protein, degradation of HIF-1α is impaired and thus those cells behave as if they were deprived of oxygen. In addition to the regulation of HIF-1α, the VHL protein has been demonstrated to interact with fibronectin and urokinase, which are required for extracellular matrix formation [11, 14].

Natural history

The relationship between the presence of a VHL germline mutation and the occurrence of one or more VHL-related tumours is a well-established observation in the literature. Only scarce evidence is provided for reduced or even non-penetrance of some VHL germline mutations [4]. The mean prevalence of VHL-associated tumours is depicted in Figure 1. Generally, VHL gene germline mutation carriers present tumours at a relatively young age. However, the age at diagnosis also depends on the intensity of pursuit. The mean age at diagnosis (with intervals) for VHL tumours are: retinal haemangioblastoma 25 years (1-68), phaeochromocytoma 30 years (5-56), cerebellar haemangioblastoma 30 years (9-78), and renal cell carcinoma 35-40 years (15-69) [1-5].

Based upon clinical expression of the disease, VHL disease has been divided into four subtypes, with a central role for phaeochromocytoma (Table 1). Patients with VHL type 1 have no phaeochromocytomas. Type 2 families have phaeochromocytomas and are divided into subtypes with a low (2A) or high risk (type 2B) of renal cell carcinoma, while type 2C families present with phaeochromocytoma only. In addition to evidence for interfamilial variability, also intrafamilial variability is a well-observed characteristic in VHL disease. This observation indicates that other genetic ('modifier' genes) and/or environmental factors are involved in the clinical manifestations of VHL germline mutations [15]. Apparently, there is no simple relationship between a germline mutation in the VHL gene and the manifestation (age of onset and type) of VHL-related tumours. Moreover, these observations do not enable individualised monitoring of VHL disease.

Pathophysiology

Whereas VHL disease manifests itself as an autosomal dominant trait, it can be considered as a recessive trait at the cellular level. Similar to other hereditary tumours caused by germline inactivation of a tumour suppressor gene, VHL tumours arise by a loss of the wild-type VHL allele, while maintaining the mutated allele. Such loss of heterozygosity (LOH, caused by deletion, non-disjunction, somatic recombination, etc.) at the VHL locus has been
The VHL protein and oxygen-dependent ubiquitination of the hypoxia-inducible factor (HIF)-1α. The VHL protein contains two functional domains: alpha (α) and beta (β). The α-domain of the VHL protein binds to Elongin C, which is connected with Elongin B in a multi-protein complex consisting of Cul2, Rbx1 and E2 ubiquitin conjugating enzyme (E2). The β-domain directly binds the substrate, HIF-1α. The VHL protein directs, depending on the amount of available oxygen, the breakdown (ubiquitination) of HIF-1α in the proteasome. In normoxic circumstances, HIF-1α is hydroxylated and binds to an intact VHL protein and is ubiquitinated in the proteasome (left). During hypoxic circumstances HIF-1α is not hydroxylated. The non-hydroxylated HIF-1α does not bind to the VHL protein and accumulates (right). In the case of a defect or absent VHL protein, HIF-1α also accumulates. Subsequently, genes that are regulated by HIF-1α, like vascular endothelial growth factor (VEGF) and erythropoietin (Epo), are upregulated, leading to (neo) angiogenesis and tumour growth.

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**Fig. 3.** The VHL protein and oxygen-dependent ubiquitination of the hypoxia-inducible factor (HIF)-1α. The VHL protein contains two functional domains: alpha (α) and beta (β). The α-domain of the VHL protein binds to Elongin C, which is connected with Elongin B in a multi-protein complex consisting of Cul2, Rbx1 and E2 ubiquitin conjugating enzyme (E2). The β-domain directly binds the substrate, HIF-1α. The VHL protein directs, depending on the amount of available oxygen, the breakdown (ubiquitination) of HIF-1α in the proteasome. In normoxic circumstances, HIF-1α is hydroxylated and binds to an intact VHL protein and is ubiquitinated in the proteasome (left). During hypoxic circumstances HIF-1α is not hydroxylated. The non-hydroxylated HIF-1α does not bind to the VHL protein and accumulates (right). In the case of a defect or absent VHL protein, HIF-1α also accumulates. Subsequently, genes that are regulated by HIF-1α, like vascular endothelial growth factor (VEGF) and erythropoietin (Epo), are upregulated, leading to (neo) angiogenesis and tumour growth. Copyright 2003, The Endocrine Society [16].

So far, there is no evidence for downregulation of an inhibitor of angiogenesis in VHL tumourigenesis. The abundant vascularisation of haemangioblastoma and renal cell carcinoma can be readily explained with the HIF-1α and ubiquitination theory as described above. Loss of VHL function reduces HIF-1α degradation and increases VEGF expression, which leads in turn to angiogenesis. In general, protein truncating mutations and missense mutations in the β-domain of the VHL protein, predicting a loss of function, seem to be associated with VHL type 1 (Fig. 2 and Table 1).

In contrast, putative gain of function mutations are associated with VHL type 2C, involving non-excessively vascularised phaeochromocytoma only [14]. VHL type 2C families are predominantly associated with specific missense mutations, notably in the α-domain. In type 2A and type 2B, mutations hamper either the binding to ElonginC by the α-domain of VHL or the capture of target proteins by the β-domain of VHL. Consequently, such target proteins (e.g. HIF-1α) cannot be properly degraded.
Table 1. Genotype-phenotype correlation for von Hippel-Lindau disease and possible responsible pathophysiological mechanism

| Type VHL | Type of VHL gene germline mutation | Retinal HAB | CNS HAB | RCC | PHAEO |
|----------|-----------------------------------|------------|--------|-----|-------|
| 1        | missense                           | +          | +      | +   | -     |
|          | microdeletions                      |            |        |     |       |
|          | insertions                          |            |        |     |       |
|          | splice site                         |            |        |     |       |
|          | nonsense                             |            |        |     |       |
|          | large deletions                      |            |        |     |       |
|          |                                    |            |        |     | loss of function (i.e. HIF-1α degradation) |
| 2A       | missense                           | +          | +      | -   | +     |
|          |                                    |            |        |     | gain of function (PHAEO) |
| 2B       | missense                           | +          | +      | +   | +     |
|          |                                    |            |        |     | loss of function (HAB+RCC) |
| 2C       | missense                           | -          | -      | -   | +     |
|          |                                    |            |        |     | gain of function, fibronectin |

HAB, haemangioblastoma; CNS, central nervous system; RCC, renal cell carcinoma (clear cell type); PHAEO, phaeochromocytoma; HIF, hypoxia-inducible factor; +, tumour present; -, tumour absent

Mutation detection

VHL gene germline mutations are detected in virtually all classic families with more than one affected family member, or classic sporadic patients with multiple VHL-related tumours [17]. Moreover, VHL gene germline mutations are identified in 4% of patients with apparently sporadic haemangioblastoma, in 1.6% with renal cell carcinoma and in 3% to 9% with phaeochromocytoma [18-21]. So, VHL gene germline mutations are not only identified in patients who do meet clinical diagnostic criteria, but also in patients with one or more VHL tumour and/or without a negative family history. This may occur when there is 1) incomplete family history data; 2) non-penetrance of a germline mutation in one of the parents; 3) a ‘de novo’ mutation. De novo mutations occur in approximately 20% of the identified VHL gene germline mutation carriers [4].

The mutation spectrum is heterogeneous, with mutations scattered throughout most of the VHL gene (Fig. 2) [22]. Missense mutations are found in 40% of the families with an identified VHL gene germline mutation [17]. Microdeletions (1-18 nucleotides), insertions (1-8 nucleotides), splice site and nonsense mutations, all predicted to lead to a truncated protein, are found in 30% of the families. Large deletions (deletions encompassing the entire gene) account for the remaining 30% of the VHL gene germline mutations. Since the size of the protein-encoding region of the VHL gene is small, it seems justified to perform screening for point mutations for VHL always by direct nucleotide sequence analysis of the entire protein-encoding region and splice-junctions. Deletions are readily detected with Southern blotting and
Table 3. VHL protocol for periodic clinical surveillance

| Investigation                                                                 | Age, frequency                  |
|------------------------------------------------------------------------------|---------------------------------|
| patients’ history,                                                           | from 10 years old, annually     |
| physical examination, blood pressure                                         | from 10 years old, annually     |
| biochemical blood tests                                                      | from 10 years old, annually     |
| 24-h urine tests (catecholamines and metanephrines)*                         | from 10 years old, annually     |
| ophthalmological examination                                                 | from 5 years old, annually      |
| upper abdominal ultrasound                                                   | from 10 years old, annually     |
| MRI (with gadolinium) cerebellum and myelum                                 | from 15 years old, two-yearly** |
| MRI upper abdomen                                                            | when indicated***               |
| MRI inner ear                                                               | when indicated****              |
| audiogram                                                                    | when indicated****              |
| neurological examination                                                     | when indicated                  |

*Accumulating evidence suggests that measurements of plasma-free metanephrines or urinary-fractionated metanephrines (normetanephrine and metanephrine separately) are the most sensitive tests for diagnosis, and are the most suitable for reliable exclusion of phaeochromocytoma [27]. These tests are particularly indicated in VHL type 2.

**Radiosurgical techniques have been developed that enable presymptomatic treatment of solid cerebellar haemangioblastoma [26], which may justify (more frequent) periodical surveillance for these tumours.

***When an MRI of the myelum is made every two years it is recommended to image the upper abdominal organs simultaneously. In this way the upper abdomen is monitored with ultrasound and MRI in alternate years.

****When an endolymphatic sac tumour (ELST) is suspected; i.e. hearing loss/deafness, tinnitus, or vertigo [2].

Practical guidelines

Clinical monitoring (Table 3) should be primarily organized around those VHL patients who have tested positive for a VHL gene germline mutation. In addition, the following persons should be monitored: first- and second-degree family members who decline a DNA test; patients (and first-degree family members) with a typical VHL tumour and features that suggest the presence of a germline mutation (i.e. the presence of multi-centric or bilateral tumours, involvement in more than one organ, a suspected family history and young age at diagnosis) [23]. With the involvement of many organs in VHL disease, it is of utmost importance that periodic monitoring is carried out in a well-co-ordinated, multidisciplinary team of physicians. On the basis of present knowledge of genotype-phenotype correlations in VHL disease, periodic monitoring cannot yet be individualised. However, regarding phaeochromocytoma, we feel periodic monitoring could be less frequent in VHL type 1 and more frequent in VHL type 2.

So far, only a small number of guidelines for the treatment of VHL-associated tumours have been developed. Most commonly, a choice is made between conservative policy (surveillance of tumours) and surgery. A good example in this dilemma is renal cell carcinoma. Options for treatment range from bilateral nephrectomy, nephron-sparing surgery to follow-up investigations only [24]. If both kidneys are affected with multiple cysts and tumours, a difficult decision has to be made between radical nephrectomy or nephron-sparing surgery. Whenever feasible,
nephron-sparing surgery is performed in order to maintain renal function as long as possible.

The other potentially life-threatening tumour in VHL disease is the cerebellar haemangioblastoma. Haemangioblastomas are regarded as benign and slow-growing tumours that do not normally invade the surrounding brain [25]. However, complications may arise due to the tumour’s tendency to form expanding cysts. Cerebellar shift may lead to herniation of the cerebellar tonsils through the foramen magnum and subsequently elevated or even life-threatening intracranial pressure. Hydrocephalus may result in rapid decompenlation with papilloedema. Cerebellar haemangioblastomas remain a major cause of morbidity and mortality in VHL patients [5]. The standard treatment is complete microsurgical removal, aided if necessary by preoperative embolisation to reduce the tumour’s vascularity. A new technique, stereotactic radiosurgery, offers the possibility of tackling multiple cerebellar lesions in a single treatment, which is particularly important in VHL patients [26]. Radiosurgery shrinks, or stops the growth of, small- or medium-sized (i.e. smaller than 3 cm) solid haemangioblastomas. Adjoining cysts, however, do not respond to radiosurgery and require later, sometimes repeated, evacuation.

Since early detection, periodic clinical surveillance and timely treatment of VHL patients lead to a better prognosis, information on the possibility of DNA analysis has to be provided to all family members. For privacy reasons, it is not allowed for health care workers to contact family members of VHL patients directly. With information brochures (spread via the patient or informed family members) persons at risk for VHL disease can be informed and advised to seek genetic counselling. In this way they have a free choice for themselves, whether they (or their offspring) want to be tested. Patients can turn to VHL support groups for support in emotional distress, information, and advice on social issues (see: www.vhl.org).

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