Sixteen New Prenylated Flavonoids from the Fruit of Sinopodophyllum hexandrum

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Abstract: Sixteen new prenylated flavonoids, sinoflavonoids P–Z (1–11) and sinoflavonoids NA–NE (12–16), were isolated from the fruit of Sinopodophyllum hexandrum, along with eight known analogues (17–24). Their structures were elucidated on the basis of extensive spectroscopic data (HR-ESI-MS, 1H-NMR, 13C-NMR, HSQC, HMBC). The cytotoxic activities of compounds 1–18, 20, and 22 were evaluated by MTT assay. Compound 6 showed the most potent cytotoxicity in MCF-7, and HepG2 cell lines, with IC50 values of 6.25 and 3.83 μM, respectively.

Keywords: Sinopodophyllum hexandrum; prenylated flavonoid; cytotoxic activity

1. Introduction

Sinopodophyllum hexandrum, belonging to the family of Berberidaceae, are widely distributed in the Southwest of China [1]. As an important medicinal plant, it was described in the Chinese Pharmacopoeia and is used in Tibetan medicine. The fruit of S. hexandrum is edible and popular with Tibetan people. It has been widely used in the treatment of amenorrhea, dead fetus, and placental retaining [2]. Previous phytochemical investigations on S. hexandrum allowed the isolation and identification of aryltetralin [1,3–6] and tetrahydrofuranoid lignans [7], flavonoids [2,8–11], labdane diterpenes [12], steroids [13], and phenolics [14]. As particularly rich in aryltetralin lactone lignans, the roots and rhizomes of the plant are mainly used for extracting podophyllotoxin, which is raw medicinal material for production of etoposide and teniposide [15]. However, its fruit is a rich source of prenylated flavonoids [2,10,11]. In our previous reports, 15 new prenylated flavonoids were found in the fruit of S. hexandrum [10,11]. As part of our continuous efforts toward discovering new cytotoxic natural products, 16 new prenylated flavonoids (1–16), together with eight known analogues (17–24), were isolated from the fruit of S. hexandrum. Details of the isolation, structure elucidation of all isolated compounds, as well as cytotoxicity of compounds 1–18, 20, and 22 against MCF-7 and HepG2 cell lines are described here (Figure 1).
The EtOH extract of the fruit of *S. hexandrum* was partitioned between petroleum ether (PE), CH$_2$Cl$_2$, EtOAc, *n*-BuOH and water, respectively. The EtOAc layer was fractionated and purified by repeated column chromatography, allowing the isolation of 24 flavonoids (1–24), including 16 new prenylated flavonoids, sinoflavonoids P–Z (1–11) and sinoflavonoids NA–NE (12–16), along with eight known analogues (17–21). By comparing their physical and spectroscopic data with literature values, the known metabolites were identified as 8-prenylkaempferol (17) [2], topazolins (18) [16], nor-β-anhydrocaitin (19) [17], citrusinol (20) [18], dysosmaflavone B (21) [19], 4’-methykaempferol (22) [20], rhamnetin (23) [21], kaempferol (24) [8].

Compound 1 was obtained as a yellow, amorphous powder and possessed a molecular formula C$_{25}$H$_{26}$O$_7$, as revealed from its HR-ESI-MS analysis (m/z 439.1760 [M + H]$^+$, calc 439.1757). The $^1$H-NMR spectrum (Table 1, see Figure S1) showed two aromatic systems including one 1,2,3,4-tetra-substituted benzene ring δ 6.74 (1H, d, $J=8.2$ Hz) and 6.70 (1H, d, $J=8.2$ Hz), one penta-substituted benzene ring δ 6.27 (1H, s), two 3-methyl-2-butenyls for two olefinic protons δ 4.94 (1H, t, $J=6.8$ Hz) and 5.01 (1H, t, $J=6.8$ Hz), four methyl groups δ 1.27 (3H, s), 1.39 (3H, s), 1.45 (3H, s) and 1.53 (3H, s), and two methylene groups δ 3.25 (2H, d, $J=6.8$ Hz) and 3.21 (2H, d, $J=6.8$Hz), and five phenolic hydroxyl groups δ 12.46 (1H, s), 10.64 (1H, s), 9.73 (1H, s), 8.77 (1H, s), and 8.42 (1H, s). The $^{13}$C-NMR spectrum (Table 2, see Figure S2) revealed a flavon skeleton including one carbonyl group δ 176.5, two benzene rings, two oxygen-bearing olefinic carbons δ 150.7, 136.2, besides two 3-methyl-2-butenyls δ 20.9, 121.99, 130.6, 17.3, 25.4, 25.7, 122.04, 129.8, 17.2, 25.2. These spectroscopic data indicated that compound 1 was a prenylated flavonol derivative. The HMBC correlations (Figure 2) of methylene group δ 3.21 (2H, d, $J=6.8$ Hz, H-1") with C-7 (δ 160.8), C-8 (δ 105.4), and C-9 (δ 154.2), and δ 3.25 (2H, d, $J=6.8$ Hz, H-1’”) with C-1’ (δ123.0), C-2’ (δ128.1), and C-3’
(δ143.0), indicated that two 3-methyl-2-butenyls were located at C-8 and C-2', respectively. Thus, the structure of compound 1 was elucidated as 8,2'-di(3-methyl-2-butenyl)-5,7,3',4'-tetrahydroxyflavone, and named sinoflavonoid P.

Compound 2 was obtained as a yellow, amorphous powder. The 1H-NMR (Table 1, see Figure S5) and 13C-NMR data (Table 2, see Figure S6) of compound 2 were closely correlated with those of 1, but differed in the appearance of one methoxy group [δ 3.56 (3H, s), δ 60.3], and one olefinic carbon δ 93.3 at upper field instead of C-8 (δ 97.7) in 1. The above data suggested that compound 2 was 6,2'-diprenyl-3-methoxyquercetin, which was also supported by HR-ESI-MS and the HMBC spectrum. The HR-ESI-MS gave an [M + H]+ ion peak at m/z 453.1910 (calcd 453.1913), being 14 mass units more than that of 1. Two 3-methyl-2-butenyls were linked to C-6 and C-2', respectively, due to the long range correlations of methylene group 3.21 (2H, d, J = 7.1 Hz, H-1") with C-5 (δ 158.5), C-6 (δ 111.1), and C-7 (δ 162.3), and of δ 3.24 (2H, d, J = 7.3 Hz, H-1") with C-1' (δ 123.2), C-2' (δ 128.2), and C-3' (δ 143.7) in the HMBC spectrum (Figure 2). The methoxy group was located at C-3 by the HMBC correlation of the methylene group δ 3.56 (3H, s) with C-3 (δ 139.0). Thus, the structure of compound 2 was elucidated as 6,2'-di(3-methyl-2-butenyl)-5,7,3',4'-tetrahydroxy-3-methoxyflavone, and named sinoflavonoid Q.

Compounds 3 and 4 were obtained as yellow, amorphous powders. Their HR-ESI-MS showed the same molecular formula of C28H28O7, according to an [M + H]+ quasi-molecular ion peak (m/z 453.1910 (calcd 453.1913) in 3; m/z 453.1912 (calcd 453.1913) in 4). Their 1H (Table 1, see Figures S9 and S13) and 13C-NMR (Table 2, see Figures S10 and S14) spectra were similar to each other, and closely correlated with those of 2, differing by the presence of one 2,2-dimethylidihydropyrano group instead of one 3-methyl-2-butenyl in 2. One 2,2-dimethylidihydropyrano group was based on a series of signals consisting of two methylene groups (δ 2.54 (2H, t, J = 6.8 Hz), 1.74 (2H, t, J = 6.8 Hz)), δ 16.7, 30.8 in 3; δ 2.62 (2H, t, J = 6.7 Hz), 1.81 (2H, t, J = 6.7 Hz), δ 15.7, 30.9 in 4), two tertiary-methyl groups (δ 1.30 (6H, s), δ 26.4 (×2) in 3; δ 1.30 (6H, s), δ 26.3 (×2) in 4), and one oxygen-bearing aliphatic quaternary carbon (δ 74.7 in 3; δ 76.2 in 4). The methylene group (δ 2.54 (2H, t, J = 6.8 Hz, H-1") in 3; δ 2.62 (2H, t, J = 6.7 Hz, H-1") in 4) showed the long range correlations (Figure 2) with C-5 (δ 154.6 in 3; δ 158.2 in 4), C-6 (δ 105.0 in 3; δ 104.4 in 4), and C-7 (δ 159.8 in 3; δ 159.8 in 4), indicating that the 2,2-dimethylidihydropyrano group was attached to C-6 and C-7, or C-5 and C-6. The presence of the phenolic hydroxy group (δ 10.60 (1H, s, 7-OH) in 3; δ 13.00 (1H, s, 5-OH) in 4) supported the linking position of the 2,2-dimethylidihydropyrano group (C-5 and C-6 in 3; C-6 and C-7 in 4). Thus, the structures of compounds 3 and 4 were deduced respectively as 5,6-(2,2-dimethylidihydropyrano)-2'-(3-methyl-2-butenyl)-7,3',4'-tri hydroxy-3-methoxyflavone (3) and 6,7-(2,2-dimethylidihydropyrano)-2'- (3-methyl-2-butenyl)-5,3',4'-tri hydroxy-3-methoxyflavone (4), and named sinoflavonoids R and S.

Compound 5 was obtained as a yellow, amorphous powder. Its 1H (Table 1, see Figure S17) and 13C-NMR spectra (Table 2, see Figure S18) was quite similar to those of 3, except for the appearance of another aromatic proton at δ 7.46 (1H, d, J = 2.2 Hz) instead of the 3-methyl-2-butenyl in 3, suggesting the absence of 3-methyl-2-butenyl at C-2' in 5. This was further supported by the HR-ESI-MS of 5 which gave an [M + H]+ ion peak at m/z 385.1262 (calcd 385.1287), being 68 mass units less than that of 3. Thus, compound 5 was deduced as 5,6-(2,2-dimethylidihydropyrano)-7,3',4'-trihydroxy-3-methoxyflavone, and named sinoflavonoid T.
Table 1. $^1$H-NMR Spectroscopic Data (500 MHz, DMSO-$d_6$) of 1–16.

| No. | 1     | 2     | 3     | 4     | 5     | 6     | 7     | 8     |
|-----|-------|-------|-------|-------|-------|-------|-------|-------|
| 6   | 6.27 s| 6.35 s| 6.28 s| 6.34 s| 6.40 s| 6.53 s| 6.31 s| 6.29 s|
| 8   |       | 7.46 d (2.2) | 7.54 d (2.2) | 6.86 d (8.5) | 6.90 d (8.5) | 6.90 d (8.4) | 6.79 d (8.4) | 6.79 d (8.4) |
| 9   | 6.70 d (8.2) | 6.76 d (8.3) | 6.68 d (8.2) | 6.73 d (8.4) | 6.86 d (8.5) | 6.90 d (8.5) | 6.90 d (8.4) | 6.79 d (8.4) |
| 2   | 6.74 d (8.2) | 6.74 d (8.3) | 6.72 d (8.2) | 6.76 d (8.4) | 7.35 dd (8.5, 2.2) | 7.43 dd (8.5, 2.2) | 7.59 d (8.4) | 7.09 d (8.4) |
| 1   | 3.21 d (6.8) | 3.21 d (7.1) | 2.54 d (6.8) | 2.62 t (6.7) | 2.54 t (6.7) | 3.06 d (8.4) | 3.30 d (6.8) | 3.30 d (6.8) |
| 2   | 5.01 t (6.8) | 5.16 t (7.1) | 1.74 t (6.8) | 1.81 t (6.7) | 1.72 t (6.7) | 4.75 t (8.4) | 5.06 t (6.8) | 5.05 t (6.8) |
| 4   | 1.45 s | 1.61 s | 1.30 s | 1.30 s | 1.30 s | 1.13 s | 1.49 s | 1.58 s |
| 5   | 1.53 s | 1.71 s | 1.30 s | 1.30 s | 1.30 s | 1.14 s | 1.56 s | 1.60 s |
| 1′′′| 3.25 d (6.8) | 3.24 d (7.3) | 3.21 d (6.9) | 3.25 d (6.9) | 7.05 d (2.1) | 3.44 dd (17.1, 6.8) | 2.95 dd (17.1, 2.4) | 6.03 br.s |
| 9   | 6.17 s | 6.30 s | 6.30 s | 6.18 s | 6.12 s | 6.17 s | 6.16 s | 6.30 s |
| 10  | 6.43 s | 7.80 d (2.0) | 7.73 d (2.1) | 6.87 d (8.5) | 6.89 d (8.5) | 6.75 d (8.5) | 6.76 d (8.2) | 6.75 d (8.2) |
| 11  | 3.23 d (6.9) | 2.67 m | 2.83 d (6.6) | 2.84 t (6.6) | 2.83 dd (16.5, 5.1) | 4.20 d (2.7) | 3.13 d (8.5) | 4.74 t (8.5) |
| 12  | 1.20 s | 1.48 s | 0.98 s | 1.32 s | 1.32 s | 1.23 s | 1.32 s | 1.32 s |
| 13  | 1.27 s | 1.54 s | 0.98 s | 1.32 s | 1.32 s | 1.27 s | 1.36 s | 1.14 s |
| 14  | 3.23 d (6.9) | 2.75 dd (17.0, 5.4) | 7.01 s | 2.68 t (6.6) | 2.63 m | 2.66 m | 2.66 m | 2.66 m |
| 15  | 4.99 t (6.9) | 3.60 dd (8.2, 5.4) | 1.73 t (6.6) | 1.71 m | 1.73 m | 1.73 m | 1.73 m | 1.73 m |
| 16  | 1.27 s | 1.17 s | 5.77 s 5.28 s | 1.32 s | 1.32 s | 1.32 s | 1.32 s | 1.32 s |
| 5″  | 1.44 s | 1.32 s | 2.10 s | 3.78 s | 3.58 s | 3.59 s | 3.57 s | 3.57 s |
| 5‴  | 3.57 s | 3.57 s | 3.65 s | 3.78 s | 3.58 s | 3.59 s | 3.57 s | 3.57 s |
| 7-OH| 12.45 s | 12.57 s | 12.67 s | 12.65 s | 12.24 s | 12.44 s | 12.65 s | 12.87 s |
Table 2. $^{13}$C-NMR Spectroscopic Data (100 MHz, DMSO-$d_6$) of 1–16.

| No. | 1    | 2    | 3    | 4    | 5    | 6    | 7    | 8    |
|-----|------|------|------|------|------|------|------|------|
| 2   | 150.7 s | 159.6 s | 155.1 s | 159.7 s | 151.4 s | 155.7 s | 156.6 s | 156.0 s |
| 3   | 136.2 s | 139.0 s | 140.6 s | 138.5 s | 139.7 s | 137.7 s | 137.7 s | 138.0 s |
| 4   | 176.5 s | 178.5 s | 172.0 s | 178.2 s | 172.1 s | 178.0 s | 178.1 s | 178.1 s |
| 5   | 158.3 s | 158.5 s | 154.6 s | 158.2 s | 154.5 s | 155.3 s | 160.0 s | 159.0 s |
| 6   | 97.7 d  | 111.1 s | 105.0 s | 104.4 s | 105.0 s | 108.8 s | 98.2 d  | 98.2 d  |
| 7   | 160.8 s | 162.3 s | 159.8 s | 159.8 s | 159.8 s | 166.2 s | 161.6 s | 161.6 s |
| 8   | 105.4 s | 93.3 d  | 93.2 d  | 94.4 d  | 93.1 d  | 88.5 d  | 106.0 s | 105.9 s |
| 9   | 154.2 s | 154.9 s | 156.4 s | 154.4 s | 156.0 s | 156.2 s | 153.9 s | 153.9 s |
| 10  | 103.5 s | 104.7 s | 107.6 s | 104.3 s | 107.2 s | 105.1 s | 104.3 s | 104.4 s |
| 1’  | 123.0 s | 123.2 s | 121.8 s | 121.1 s | 121.4 s | 120.7 s | 113.4 s | 117.8 s |
| 2’  | 128.1 s | 128.2 s | 127.6 s | 127.8 s | 115.0 d | 115.7 d | 128.1 s | 127.0 s |
| 3’  | 143.0 s | 143.7 s | 143.2 s | 143.3 s | 145.1 s | 145.2 s | 143.1 s | 145.4 s |
| 4’  | 146.5 s | 147.4 s | 146.5 s | 147.1 s | 147.7 s | 148.1 s | 145.5 s | 143.3 s |
| 5’  | 112.4 d | 112.9 d | 112.4 d | 112.4 d | 115.6 d | 115.4 d | 110.4 d | 115.2 d |
| 6’  | 121.0 d | 121.5 d | 120.9 d | 121.0 d | 119.8 d | 120.5 d | 125.9 d | 122.3 d |
| 1”  | 20.9 t  | 21.4 t  | 16.7 t  | 15.7 t  | 16.7 t  | 25.7 t  | 21.1 t  | 21.1 t  |
| 2”  | 121.99 d| 122.6 d | 30.8 t  | 30.9 t  | 30.8 t  | 91.5 d  | 122.4 d | 122.4 d |
| 3”  | 130.6 s | 131.1 s | 74.7 s  | 76.2 s  | 74.8 s  | 70.0 s  | 131.0 s | 131.3 s |
| 4”  | 17.3 q  | 18.1 q  | 26.4 q  | 26.3 q  | 26.4 q  | 24.8 q  | 17.6 q  | 17.7 q  |
| 5”  | 25.4 q  | 25.9 q  | 26.4 q  | 26.3 q  | 26.4 q  | 25.9 q  | 25.3 q  | 25.4 q  |
| 1”” | 25.7 t  | 26.2 t  | 26.0 t  | 25.7 t  | 25.7 t  | 107.3 d | 38.2 t  | 107.3 d |
| 2”” | 122.04 d| 122.3 d | 122.9 d | 122.8 d | 146.6 d | 101.0 d |        |      |
| 3”” | 129.8 s | 130.6 s | 129.9 s | 130.2 s |        |        |        |      |
| 4”” | 17.2 q  | 17.8 q  | 17.4 q  | 17.4 q  |        |        |        |      |
| 5”” | 25.2 q  | 25.7 q  | 25.4 q  | 25.4 q  |        |        |        |      |
| OCH$_3$ | 60.3 q  | 59.4 q  | 59.9 q  | 59.2 q  | 59.6 q  | 60.6 q  | 60.2 q  |      |
Table 2. Cont.

| No. | 9    | 10    | 11    | 12    | 13    | 14    | 15    | 16    |
|-----|------|-------|-------|-------|-------|-------|-------|-------|
| 2   | 158.7 s | 158.2 s | 156.7 s | 156.6 s | 147.7 s | 158.3 s | 158.0 s | 158.0 s |
| 3   | 139.1 s | 138.7 s | 137.7 s | 138.2 s | 136.1 s | 139.2 s | 139.5 s | 138.8 s |
| 4   | 178.2 s | 178.2 s | 178.9 s | 178.3 s | 175.9 s | 178.1 s | 178.1 s | 178.1 s |
| 5   | 159.4 s | 159.0 s | 158.7 s | 161.6 s | 158.0 s | 158.75 s | 160.3 s | 161.8 s |
| 6   | 98.9 d  | 98.2 d  | 98.2 d  | 99.0 d  | 98.5 d  | 98.8 d  | 98.7 d  | 93.3 d  |
| 7   | 158.7 s | 161.5 s | 161.7 s | 164.7 s | 159.1 s | 158.82 s | 158.8 s | 166.2 s |
| 8   | 99.0 s  | 105.9 s | 107.2 s | 94.1 d  | 99.7 s  | 99.0 s  | 100.3 s | 104.4 s |
| 9   | 154.1 s | 154.1 s | 154.1 s | 155.8 s | 153.0 s | 154.0 s | 156.0 s | 151.3 s |
| 10  | 105.4 s | 104.5 s | 104.4 s | 103.6 s | 105.5 s | 105.8 s | 105.4 s | 105.4 s |
| 1'  | 121.2 s | 120.5 s | 113.5 s | 121.7 s | 122.2 s | 120.3 s | 120.2 s | 120.2 s |
| 2'  | 127.7 s | 120.6 s | 129.4 s | 130.2 d | 114.8 d | 121.3 s | 121.0 s | 121.1 s |
| 3'  | 143.3 s | 141.0 s | 142.7 s | 121.5 s | 145.1 s | 142.0 s | 141.9 s | 142.0 s |
| 4'  | 147.1 s | 147.9 s | 144.9 s | 156.8 s | 146.7 s | 148.3 s | 148.2 s | 148.3 s |
| 5'  | 112.5 d | 112.9 d | 111.0 d | 117.5 d | 115.7 d | 112.3 d | 112.8 d | 112.7 d |
| 6'  | 121.0 d | 121.5 d | 125.9 d | 128.1 d | 120.2 d | 121.1 d | 121.4 d | 121.2 d |
| 1"  | 24.9 t  | 21.0 t  | 17.5 t  | 22.2 t  | 15.7 t  | 24.7 t  | 73.9 d  | 25.2 t  |
| 2"  | 66.8 d  | 122.0 d | 47.8 t  | 32.5 t  | 30.9 t  | 66.6 d  | 67.3 d  | 91.5 d  |
| 3"  | 78.9 s  | 130.9 s | 68.7 s  | 75.8 s  | 76.2 s  | 78.9 s  | 78.8 s  | 69.9 s  |
| 4"  | 20.8 q  | 17.4 q  | 28.8 q  | 27.0 q  | 20.2 q  | 23.1 q  | 24.8 q  | 24.8 q  |
| 5"  | 25.1 q  | 25.5 q  | 28.8 q  | 27.0 q  | 26.3 q  | 25.2 q  | 24.5 q  | 25.7 q  |
| 1'''| 25.7 t  | 29.5 t  | 104.2 d | 21.1 t  | 19.9 t  | 20.1 t  |        |        |
| 2'''| 122.9 d | 67.8 d  | 156.5 s | 31.8 t  | 31.8 t  | 31.7 t  |        |        |
| 3'''| 130.2 s | 76.8 s  | 132.5 s | 73.9 s  | 73.5 s  | 73.9 s  |        |        |
| 4'''| 17.2 q  | 19.8 q  | 114.1 t | 26.0 q  | 26.4 q  | 26.2 q  |        |        |
| 5'''| 25.2 q  | 25.3 q  | 18.9 q  | 26.0 q  | 26.4 q  | 26.2 q  |        |        |
| OCH₃| 59.8 q  | 60.2 q  | 60.1 q  | 60.1 q  | 59.2 q  | 60.2 q  | 60.2 q  | 60.2 q  |
| OCH₃|        |        |        |        |        |        |        | 56.9 q  |
Compound 6 was obtained as a yellow, amorphous powder. The molecular formula was found to be C_{21}H_{20}O_{8}, as deduced by analysis of the [M + Na]^+ molecular ion peak at m/z 423.1056 (calcd 423.1056) in the HR-ESI-MS. Its ^1H (Table 1, see Figure S21) and ^13C-NMR spectra (Table 2, see Figure S22) were similar to those of 5, except that one 2-(1-hydroxy-1-methylthyl)dihydrofurano group was observed instead of one 2,2-dimethylidihydropyran group in 5. 2-(1-Hydroxy-1-methylthyl)dihydrofurano group was deduced by the HMBC correlations (Figure 2) from two tertiary methyls δ1.13 (3H, s, H-4”), 1.14 (3H, s, H-5”) to C-3” (δ 70.0), C-2” (δ 91.5), from oxymethine group δ 4.75 (1H, t, J = 8.4 Hz, H-2”) to C-1” (δ 25.7), C-3” (δ 70.0), C-4” (δ 24.8), and C-5” (δ 25.9), from methylene group δ 3.06 (2H, d, J = 8.4 Hz, H-1”) to C-2” (δ 91.5). 2-(1-Hydroxy-1-methylthyl)dihydrofurano group was located at C-6 and C-7 from the HMBC correlations of oxymethine group δ 4.75 (1H, t, J = 8.4 Hz, H-2”) with C-7 (δ 166.2), of methylene group δ 3.06 (2H, d, J = 8.4 Hz, H-1”) with C-5 (δ 155.3), C-6 (δ 108.8), C-7 (δ 166.2), and in combination with the chelated phenolic hydroxyl group δ 12.92 (1H, s, 5-OH). Thus, compound 6 was deduced as 6,7-[2-(1-hydroxy-1-methylthyl)dihydrofurano]-5,3’,4’-trihydroxy-3-methoxy flavone, and named sinoflavonoid U.

Compound 7 was obtained as a yellow, amorphous powder and possessed a molecular formula of C_{23}H_{20}O_{7}, derived from its HR-ESI-MS analysis (m/z 407.1111 [M−H]^−, calcd C_{23}H_{19}O_{7}, 407.1131). The ^1H (Table 1, see Figure S25) and ^13C-NMR spectra (Table 2, see Figure S26) of compound 7 was quite similar to those of 1, except for the appearance of one methoxy group and one disubstituted furan ring for two olefinic protons δ 7.05 (1H, d, J = 2.1 Hz, H-3”) and 8.09 (1H, d, J = 2.1 Hz, H-2”) instead of one 3-methyl-2-butenyl in 1. This was also further supported by HMBC spectrum (Figure 2). The olefinic protons δ 7.05 (1H, d, J = 2.1 Hz, H-1”) and 8.09 (1H, d, J = 2.1 Hz, H-2”) showed the HMBC correlations with C-1’ (δ 113.4), C-2’ (δ 128.1), C-3’ (δ 143.1), and C-2’ (δ 128.1), C-3’ (δ 143.1), respectively, indicating the disubstituted furan ring was attached to C-2’ and C-3’. The methoxy group was located at C-3 by the HMBC correlation of the methoxy group δ 3.65 (3H, s) with C-3 (δ 137.7). Thus, compound 7 was deduced as 2’,3’-furano-5,3’,4’-trihydroxy-3-methoxy flavone, and named sinoflavonoid V.

Compound 8 was obtained as a yellow, amorphous powder. Its ^1H (Table 1, see Figure S29) and ^13C-NMR spectra (Table 2, see Figure S30) were quite similar to those of 7, except that one 2-hydroxydihydrofuran ring was observed instead of one furan ring in 7. The 2-hydroxydihydrofuran ring was determined by one methylene group δ 3.44 (1H, dd, J = 17.1, 6.8 Hz), 2.95 (1H, dd,
δ 132.5. The furan ring was inferred from one olefinic proton δ 38.2, one dioxymethine group δ 6.03 (1H, brs), δ 101.0. Those were further supported by its HR-ESI-MS, which gave an [M + H]⁺ quasi-molecular ion peak m/z 427.1390 (calcd 427.1393), being 18 mass units more than that of 7. Thus, compound 8 was identified as 8-(3-methyl-2-butenyl)-2,3'-(2-hydroxydihydrofurano)-5,7,4'-trihydroxy-3-methoxyflavone, and named as sinoflavonoid W.

Compounds 9 and 10 were obtained as yellow, amorphous powders. Their molecular formulae were assigned as C_{26}H_{26}O_{8} by HR-ESI-MS (m/z 491.1683 [M + Na]⁺ (calcd 491.1682) in 9; m/z 491.1669 [M + Na]⁺ (calcd 491.1682) in 10). Their \(^1\)H (Table 1, see Figures S33 and S37) and \(^{13}\)C-NMR spectra (Table 2, see Figures S34 and S38) were similar to those of 8. Two tertiary-methyl signals (δ 1.27 (3H, s), 1.20 (3H, s), 1.27 (3H, s), 1.17 (3H, s), δ 25.3, 19.8 in 10), one methylene group (δ 2.79 (1H, dd, J = 16.5, 5.2 Hz), 2.47 (1H, dd, J = 16.5, 7.1 Hz), δ 24.9 in 9; δ 2.75 (1H, dd, J = 17.0, 5.4 Hz), 2.55 (1H, dd, J = 17.0, 8.2 Hz), δ 29.5 in 10), and one oxymethine group (δ 3.67 (1H, dd, J = 7.1, 5.2 Hz), δ 66.8 in 9; δ 3.60 (1H, dd, J = 8.2, 5.4 Hz), δ 67.8 in 10) were observed, implying the presence of one 2,2-dimethyl-3-hydroxydihydropyrano group in 9 and 10 instead of 2-hydroxydihydrofurano ring in 8, respectively. The 2,2-dimethyl-3-hydroxydihydropyrano group and 3-methyl-2-butenyl in 9 were attached to C-7 and C-8, and C-2’ and C-3’, respectively, by the HMBC correlations (Figure 2) from H-1” (δ 2.79, 2.47) to C-7 (δ 158.7), C-8 (δ 99.0), and C-9 (δ 154.1), and from H-1”” (δ 3.23) to C-1’ (δ 121.2), C-2’ (δ 127.7), and C-3’ (δ 143.3). In contrast, the 3-methyl-2-butenyl and 2,2-dimethyldihydropyrano group in 10 was located at C-8, and C-2’, C-3’, respectively, by the HMBC correlations of H-1” (δ 3.23) with C-7 (δ 161.5), C-8 (δ 105.9) and C-9 (δ 154.1), and of H-1”” (δ 2.75, 2.55) with C-1’ (δ 120.5), C-2’ (δ 120.6) and C-3’ (δ 141.0). Thus, compounds 9 and 10 were elucidated respectively as 7,8-(2,2-dimethyldihydropyrano)-2’-(3-methyl-2-butenyl)-5,3’-tri hydroxy-3-methoxyflavone (9), 8-(3-methyl-2-butenyl)-2,3’-(2,2-dimethyldihydropyrano)-5,7,4’-tri hydroxy-3-methoxyflavone (10), and named sinoflavonoids X and Y.

Compound 11 was obtained as a yellow, amorphous power and possessed a molecular formula of C_{26}H_{26}O_{8}, derived from its HR-ESI-MS analysis (m/z 465.1544 [M − H]−, calcd C_{26}H_{25}O_{8}, 465.1549). Its \(^1\)H-NMR (Table 1, see Figure S41) and \(^{13}\)C NMR data (Table 2, see Figure S42) were closely correlated with those of 7, but differed in the appearance of an isopropenyl and 3-hydroxy-3-methylbutyl in 11 instead of an olefinic proton δ 8.09 (1H, d, J = 2.1 Hz) and 3-methyl-2-butenyl in 7, respectively. The isopropenyl was deduced by two olefinic protons δ 5.77 (1H, s), 5.28 (1H, s), one tertiary-methyl group δ 2.10 (3H, s), δ 18.9, two olefinic carbons δ 114.1, 132.5. The furan ring was inferred from one olefinic proton δ 7.01 (1H, s), and two characteristic olefinic carbons δ 104.2, 156.5. In the HMBC spectrum (Figure 2), the long range correlations of the olefinic protons δ 5.77 (1H, s), 5.28 (1H, s), the methyl group 2.10 (3H, s) with the olefinic carbon δ 156.5 (C-2’”) indicated the isopropenyl was linked to C-2’”. One 3-hydroxy-3-methylbutyl was based on a series of signals consisting of two olef inic groups δ 2.67 (2H, m), 1.47 (2H, m), δ 17.5, 47.8, two tertiary-methyl groups δ 0.98 (6H, s), δ 28.8 (×2), and one oxygen-bearing aliphatic quaternary carbon δ 68.7. The HMBC spectrum also showed the long range correlations of the methane protons δ 2.67 (2H, m, H-1”) with C-7 (δ 161.7), C-8 (δ 107.2), and C-9 (δ 154.1), indicating that 3-hydroxy-3-methylbutyl was attached to C-8. Thus, compound 11 was elucidated as 8-(3-hydroxy-3-methylbutyl)-2,3’-(2-isopropenylfurano)-5,7,4’-trihydroxy-3-methoxyflavone, and named sinoflavonoid Z.

Compound 12 was obtained as a yellow, amorphous powder and possessed a molecular formula of C_{21}H_{20}O_{6}, derived from its HR-ESI-MS analysis (m/z 369.1336 [M + H]⁺, calcd C_{21}H_{20}O_{6}, 369.1338). Its \(^1\)H and \(^{13}\)C-NMR spectra were similar to kaempferol [8], except for the appearance of 2,2-dimethyldihydropyrano group and one methoxy group in 12. 2,2-Dimethyldihydropyrano group was proved by two tertiary-methyl signals δ1.32 (6H, s), δ 27.0 (×2), two methylene groups δ 1.82 (2H, t, J = 6.5 Hz), 2.83 (2H, t, J = 6.5 Hz), δ 32.5, 22.2, and one oxygen-bearing aliphatic quaternary carbon δ 75.8. By the HMBC correlations (Figure 2) of δ 2.83 (2H, t, J = 6.5Hz, H-1”) with C-2’ (δ 130.2), C-3’ (δ 121.5) and C-4’ (δ 156.8), 2,2-dimethyldihydropyrano group was linked
to C-3′ and C-4′. The methoxy group was located at C-3, based on the HMBC correlation between the methoxy group δ 3.78 (3H, s) and C-3 (δ 138.2). Thus, compound 12 was elucidated as 3′,4′-(2,2-dimethyldihydropyrano)-5,7-dihydroxy-3-methoxyflavone, and named sinoflavonoid NA.

Compound 13 was obtained as a yellow, amorphous powder and possessed the molecular formula C_{20}H_{18}O_{7}, as revealed from its HR-ESI-MS analysis (m/z 371.0989 [M + H]^+, calcld C_{20}H_{16}O_{7}, 371.1131). Its 1^H (Table 1, see Figure S49) and 13C-NMR spectra (Table 2, see Figure S50) were similar to quercetin [8], except for the appearance of 2,2-dimethyldihydropyrano group in 13. 2,2-Dimethyldihydropyrano group was proved by two tertiary-methyl signals δ 1.32 (6H, s), δ 26.3 (2×2), two methylene groups δ 1.87 (2H, t, J = 6.6 Hz), 2.84 (2H, t, J = 6.6 Hz), δ 15.7, 30.9, and one oxygen-bearing aliphatic quaternary carbon δ 76.2. By the HMBC correlations (Figure 2) of δ 2.84 (2H, t, J = 6.6 Hz, H-1′′) with C-7 (δ 159.1), C-8 (δ 99.7) and C-9 (δ 153.0), 2,2-dimethyldihydropyrano group was linked to C-2′ and C-3′, by the HMBC correlations (Figure 2) of the methylene protons δ 2.68 (2H, t, J = 6.6 Hz, H-1′′′) with C-1′ (δ 120.3), C-2′ (δ 121.3), and C-3′ (δ 142.0). Thus, compound 13 was elucidated as 7,8-(2,2-dimethyldihydropyrano)-5,3′,4′-tetrahydroxy-3-methoxyflavone, named sinoflavonoid NB.

Compound 14 was obtained as a yellow, amorphous powder. Its 1^H (Table 1, see Figure S53) and 13C-NMR spectra (Table 2, see Figure S54) were quite similar to those of 9, respectively, except for the observation of 2,2-dimethyldihydropyrano group in 14 instead of 3-methyl-2-butenyl in 9. This was further confirmed by their HR-ESI-MS, which gave the same molecular formula C_{26}H_{28}O_{8} by the quasi-molecular ion peak (m/z 469.1853 [M + H]^+ (calcld 469.1862) in 14, m/z 491.1682 [M + Na]^+ in 9). The 2,2-dimethyldihydropyrano group was attached to C-2′ and C-3′, by the HMBC correlations (Figure 2) of the methylene protons δ 2.68 (2H, t, J = 6.6 Hz, H-1′′′) with C-1′ (δ 120.3), C-2′ (δ 121.3), and C-3′ (δ 142.0). Thus, compound 14 was elucidated as 7,8-bis-2′,3′-(2,2-dimethyldihydropyrano)-5,4′,2″-trihydroxy-3-methoxyflavone, and named sinoflavonoid NC.

Compound 15 was obtained as a yellow, amorphous powder. Its 1^H (Table 1, see Figure S57) and 13C-NMR spectra (Table 2, see Figure S58) were quite similar to those of 14, respectively, except for the observation of one methoxy group δ 3.31 (3H, s), δ 56.9 in 15, suggesting 15 to be a further methyl ether derivative of 14. This was further confirmed by the HR-ESI-MS, which gave the molecular formula C_{27}H_{30}O_{9} by the quasi-molecular ion peak m/z 521.1792 [M + Na]^+ (calcld 521.1788), being 14 mass units more than that of 14. In the HMBC spectrum (Figure 2), the additional methoxy group was located at C-1′′, based on the long range correlation between the methoxy group δ 3.31 (3H, s) and C-1′′ (δ 73.9). Thus, compound 15 was elucidated as 7,8-bis-2′,3′-(2,2-dimethyldihydropyrano)-5,4′,2″-trihydroxy-3,1″-dimethoxyflavone, and named sinoflavonoid ND.

Compound 16 was obtained as a yellow, amorphous powder. Its 1^H (Table 1, see Figure S61) and 13C-NMR spectra (Table 2, see Figure S62) were quite similar to those of 14, except for the appearance of one 2-(1-hydroxy-1-methylethyl)dihydrofurano group in 16 instead of 3-hydroxy-2,2-dimethyldihydropyrano group in 14. 2-(1-Hydroxy-1-methylethyl)dihydrofurano group was deduced by the HMBC correlations (Figure 2) from two tertiary-methyls δ1.13 (3H, s, H-4′′′), 1.14 (3H, s, H-5′′′) to C-3″ (δ 69.9), C-2″ (δ 91.5), from oxymethine group δ 4.74 (1H, t, J = 8.5 Hz, H-2″′) to C-1″ (δ 25.2), C-3″ (δ 69.9), C-4″ (δ 24.8), and C-5″ (δ 25.7), from methylene group δ 3.13 (2H, d, J = 8.5 Hz, H-1′′′) to C-2″ (δ 91.5). 2-(1-Hydroxy-1-methylethyl)dihydrofurano group was located at C-7 and C-8, based on the HMBC correlation of methylene group δ 3.13 (2H, d, J = 8.5 Hz) with C-7 (δ 166.2), C-8 (δ 104.4), C-9 (δ 151.3). Thus, compound 16 was deduced as 7,8-[2-(1-hydroxy-1-methylethyl)dihydrofurano]-2′,3′-(2,2-dimethyldihydropyrano)-5,4′-dihydroxy-3-methoxyflavone, and named sinoflavonoid NE.

Compounds 1–18, 20, 22 were tested for their in vitro cytotoxic activities against MCF-7 and HepG2 cell lines using the MTT assay [11], with 5-fluorouracil as a positive control, and IC_{50} values were summarized in Table 3. Among the tested compounds, only compound 6 exhibited the most potent cytotoxic activities against MCF-7 and HepG2 cell lines, with an IC_{50} value of 6.25 and 3.83 μM, respectively. Compound 6 was more cytotoxic than 5-fluorouracil, whereas compounds
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5 displayed no cytotoxicity against MCF-7 and HepG2 cell lines. Compound 5 has the same B and C rings from flavone skeleton as 6, so the variation in cytotoxicity between them indicates 2-(1-hydroxy-1-methylethyl)dihydrofurano group on ring A is structurally required for the cytotoxicity against the MCF-7 and HepG2 cell lines. Furthermore, the cytotoxic activity may be affected by the position of furano or dihydrofurano group on the ring A, which needs to be verified with more similar derivatives. With the promising cytotoxicities against two cell lines, compound 6 may be the optimal lead compound for structure optimization studies.

Table 3. Cytotoxicities of 1–18, 20, 22 against MCF-7 and HepG2 cell lines (IC50, µM).

| Compound | MCF-7 | HepG2 | Compound | MCF-7 | HepG2 |
|----------|-------|-------|----------|-------|-------|
| 1        | 33.8 ± 2.0 | 75.9 ± 5.3 | 15       | 25.5 ± 1.8 | 17.2 ± 1.3 |
| 2–5, 7–12, 17, 18 | >100  | >100  | 16       | 41.8 ± 3.5 | 55.4 ± 4.9 |
| 6        | 6.25 ± 0.49 | 3.83 ± 0.26 | 20       | 48.3 ± 3.2 | 50.6 ± 4.4 |
| 13       | 59.7 ± 4.1  | 45.3 ± 3.5  | 22       | 59.3 ± 5.7  | >100    |
| 14       | 30.4 ± 2.6  | 23.1 ± 1.7  | 5-fluorouracil | 33.4 ± 3.0 | 18.2 ± 2.5 |

3. Materials and Methods

3.1. General Experimental Procedures

The UV spectra were measured on a Shimadzu UV-1700 spectrometer (Shimadzu Corporation, Kyoto, Japan). The IR spectra were measured on a Nicolet 10 Microscope Spectrometer (Thermo Scientific, San Jose, CA, USA). The 1D and 2D-NMR spectra were recorded on Bruker-AC (E)-500 spectrometer (Bruker AM 500, Fällanden, Switzerland) using tetramethylsilane (TMS) as an internal standard. The HR-ESI-MS was determined on a Bruker microTOF-Q instrument (Bruker BioSpin, Rheinstetten, Germany). Column chromatography was performed with silica gel (200–300 mesh; Qingdao Marine Chemical Inc., Qingdao, China), sephadex LH-20 (GE Healthcare), and ODS (50 µm; YMC Co. LTD., Kyoto, Japan). Preparative high performance liquid chromatography (HPLC) separations were performed on a SEP system (Beijing Sepuruisi scientific Co., Ltd., China) equipped with a variable-wavelength UV detector, using a YMC-Pack ODS-A column (250 × 20 mm, 5 µm). Chemical reagents for isolation were of analytical grade and purchased from Tianjin Siyou Co., Ltd., China. Biological reagents were from Sigma Company. Human hepatocellular (HepG2), and breast (MCF-7) cancer cell lines were from Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, China.

3.2. Plant Material

The plant material was collected from Deqin, Yunnan Province, China, in September 2013, and identified by Prof. Chengming Dong as the fruit of *S. hexandrum*. A voucher specimen (SE 20130929) was deposited at the School of Pharmacy, Henan University of Traditional Chinese Medicine.

3.3. Extraction and Isolation

The powered fruit of *S. hexandrum* (9.1 kg) were refluxed with 95% EtOH three times (each, 2h, 20L). The filtrate was concentrated under reduced pressure to yield a dark brown residue (1.6 kg). The residue was suspended in water and partitioned with petroleum ether (PE), CH2Cl2, EtOAc, and n-BuOH, successively.

The EtOAc layer (142.71 g) was fractionated by silica gel column chromatography (CC, 100 × 10 cm) with a gradient of PE (60–90 °C)-acetone. Sixteen fractions E1–E16 were obtained on the basis of TLC monitoring. Fraction E7 (4.79 g) was chromatographed over open ODS (50 × 2 cm) eluted with a gradient of methanol–H2O (v/v: 60:40, 70:30, 80:20, 90:10) to obtain sub-fractions E7-1–E7-6. Sub-fraction E7-4 (1.62 g) was further purified by silica gel CC (20 × 1 cm) eluted with PE-acetone (100:10, 100:30, 100:50) to give 12 (4.8 mg), 17 (53.8 mg), 18 (6.4 mg), 19 (2.6 mg), 20 (1.9 mg). Fraction E8 (5.02 g) was...
subjected to sephadex LH-20 CC (90 × 2.5 cm) eluted by methanol to yield sub-fractions E8-1–E8-5. Sub-fraction E8-4 (1.03 g) was further submitted to preparative HPLC eluted with methanol-H₂O (72: 28) at a flow rate of 7 mL/min to obtain 10 (1.8 mg, tᵣ 25 min), 2 (5.9 mg, tᵣ 36 min). Sub-fraction E8-5 (1.75 g) was further applied to preparative HPLC eluted with methanol-H₂O (75:25) at a flow rate of 7 mL/min to give 24 (30.8 mg, tᵣ 16 min), 23 (1.8 mg, tᵣ 19 min), 22 (2.6 mg, tᵣ 27 min), 21 (2.2 mg, tᵣ 32 min). Fraction E9 (2.45 g) was separated by silica gel CC eluted by PE-acetone (100:30) to give 5 (6.2 mg) and 11 (4.9 mg). Fraction E10 (2.05 g) was submitted to sephadex LH-20 CC (1.5 × 50 cm) eluted by methanol to yield sub-fractions E10-1 to E10-3. Sub-fraction E10-2 was purified by preparative HPLC eluted with MeOH-H₂O (69:21) at 7 mL/min to yield 6 (8.5 mg, tᵣ 42 min), 16 (7.6 mg, tᵣ 62 min), 14 (5.3 mg, tᵣ 66 min). Fraction E11 (1.95 g) was applied to silica gel CC (45 × 2 cm) eluted by PE-acetone (100:7, 100:10, 100:15, 100:20) to yield sub-fractions E11-1 to E11-4. Sub-fraction E11-4 was further purified by preparative HPLC eluted with methanol-H₂O (80:20) at a flow rate of 7 mL/min to give 15 (4.5 mg, tᵣ 21 min), 4 (3.7 mg, tᵣ 69 min). Fraction E12 (1.70 g) was chromatographed over open ODS (25 × 2 cm) eluted with a gradient of methanol–H₂O (50:50, 70:30, 80:20) to yield sub-fractions E12-1 to E12-3. Sub-Fraction E12-3 (0.5 g) was further purified by preparative HPLC eluted with MeOH–H₂O (60:40) at 7 mL/min to yield 1 (13.5 mg, tᵣ 26 min), 7 (8.5 mg, tᵣ 40 min). Fraction E13 (2.58 g) was subjected to silica gel CC (35 × 2 cm) eluted by PE-acetone (100:40) to give 3 (4.8 mg) and 13 (5.2 mg). Fraction E14 (1.73 g) was submitted to sephadex LH-20 CC (60 × 2.5 cm) eluted by methanol to yield sub-fractions E14-1 to E14-3. Sub-fraction E14-3 was subjected to preparative HPLC eluted with methanol–H₂O (75:25) at a flow rate of 7 mL/min to give 9 (2.7 mg, tᵣ 16 min), 8 (3.2 mg, tᵣ 25 min).

3.4. Spectroscopic and Physical Data

Sinoflavonoid P (1): yellow, amorphous powder; UV (MeOH) λ_max (log ε) 260 (0.40), 364 (0.22) nm; IRν_max 3356, 2924, 2854, 1651, 1560, 1511, 1423, 1364, 1316, 1260, 1211, 1188, 1149, 1112 cm⁻¹; HR-ESI-MS (positive): m/z 439.1760 [M + H]^+ (calcd. for C₂₅H₂₇O₇, 439.1757), m/z 461.1576 [M + Na]^+ (calcd. for C₂₅H₂₆O₇Na, 461.1576); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid Q (2): yellow, amorphous powder; UV (MeOH) λ_max (log ε) 262 (1.24), 321 (0.73) nm; IRν_max 3354, 2961, 2926, 2855, 1659, 1611, 1573, 1468, 1358, 1293, 1234, 1155, 1092 cm⁻¹; HR-ESI-MS (positive): m/z 453.1910 [M + H]^+ (calcd. for C₂₆H₂₉O₇, 453.1913); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid R (3): yellow, amorphous powder; UV (MeOH) λ_max (log ε) 262 (1.24), 321 (0.73) nm; IRν_max 3374, 2959, 2926, 2854, 1650, 1597, 1451, 1359, 1293, 1228, 1162, 1092 cm⁻¹; HR-ESI-MS (positive): m/z 453.1910 [M + H]^+ (calcd. for C₂₆H₂₉O₇, 453.1913); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid S (4): yellow, amorphous powder; UV (MeOH) λ_max (log ε) 261 (1.24), 325 (0.60) nm; IRν_max 3524, 2975, 2930, 2856, 1651, 1604, 1462, 1373, 1301, 1258, 1161, 1091 cm⁻¹; HR-ESI-MS (positive): m/z 453.1912 [M + H]^+ (calcd. for C₂₆H₂₉O₇, 453.1913); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid T (5): yellow, amorphous powder; UV (MeOH) λ_max (log ε) 256 (0.66), 345 (0.75) nm; IRν_max 3421, 2974, 2941, 1655, 1617, 1571, 1454, 1287, 1241, 1159, 1088 cm⁻¹; HR-ESI-MS (positive): m/z 385.1262 [M + H]^+ (calcd. for C₂₁H₂₁O₇, 385.1287), (negative): m/z 383.1116 [M − H]^− (calcd. for C₂₁H₂₀O₇, 383.1131); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid U (6): yellow, amorphous powder; [α]_D°²⁵ = −13.7 (c 0.13, MeOH); UV (MeOH) λ_max (log ε) 258 (0.48), 354 (0.56) nm; IRν_max 3393, 2925, 1656, 1588, 1455, 1341, 1239, 1159, 1088 cm⁻¹; HR-ESI-MS (positive): m/z 423.1056 [M + Na]^+ (calcd. for C₂₁H₂₀O₈Na, 423.1056), (negative): m/z 399.1084 [M − H]^− (calcd. for C₂₁H₁₉O₈s, 399.1080); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid V (7): yellow, amorphous powder; UV (MeOH) λ_max (log ε) 269 (0.11), 359 (0.07) nm; IRν_max 3330, 2957, 2925, 2870, 2855, 1650, 1610, 1562, 1511, 1496, 1453, 1424, 1364, 1334, 1307, 1230, 1154,
1098 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 409.1282 [M + H]⁺ (calcd. for C₂₃H₂₁O₇, 409.1287), (negative): \( m/z \) 407.1111 [M – H]⁻ (calcd. for C₂₃H₁₉O₇, 407.1131); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid W (8): yellow, amorphous powder; \([\alpha]_{D}^{25}\)–11.5 (c 0.09, MeOH); UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 264 (0.39), 350 (0.15) nm; IR\( \nu_{\text{max}} \) 3350, 2966, 2926, 2855, 1651, 1610, 1506, 1448, 1362, 1306, 1228, 1069 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 427.1390 [M + H]⁺ (calcd. for C₂₃H₂₃O₈, 427.1393), \( m/z \) 449.1210 [M + Na]⁺ (calcd. for C₂₃H₂₂O₈Na, 449.1212); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid X (9): yellow, amorphous powder; \([\alpha]_{D}^{25}\)–8.9 (c 0.04, MeOH); UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 264 (0.41), 350 (0.18) nm; IR\( \nu_{\text{max}} \) 3368, 2982, 2926, 2854, 1655, 1597, 1506, 1486, 1446, 1355, 1297, 1232, 1172,1149, 1033 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 491.1683 [M + Na]⁺ (calcd. for C₂₆H₂₅O₈Na, 491.1682); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid Y (10): yellow, amorphous powder; \([\alpha]_{D}^{25}\)–7.4 (c 0.03, MeOH); UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 264 (0.38), 350 (0.13) nm; IR\( \nu_{\text{max}} \) 3340, 2982, 2930, 2854, 1651, 1584, 1490, 1449, 1359, 1302, 1227, 1171, 1145, 1067 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 491.1669 [M + Na]⁺ (calcd. for C₂₆H₂₅O₈Na, 491.1682); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid Z (11): yellow, amorphous powder; UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 263 (0.14), 332 (0.09) nm; IR\( \nu_{\text{max}} \) 3403, 2957, 2922, 2851, 1654, 1578, 1488, 1457, 1375, 1361, 1302, 1229, 1161, 1076 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 465.1544 [M – H]⁻ (calcd. for C₂₆H₂₅O₆, 465.1549); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid NA (12): yellow, amorphous powder; UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 268 (0.53), 355 (0.60) nm; IR\( \nu_{\text{max}} \) 3421, 2974, 2929, 2853, 1647, 1603, 1507, 1489, 1448, 1358, 1306, 1160, 1089 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 369.1336 [M + H]⁺ (calcd. for C₂₁H₁₉O₆, 369.1338), \( m/z \) 391.1144 [M + Na]⁺ (calcd. for C₂₁H₂₀O₆Na, 391.1158); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid NB (13): yellow, amorphous powder; UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 260 (0.23), 376 (0.18) nm; IR\( \nu_{\text{max}} \) 3419, 2955, 2924, 2852, 1651, 1596, 1556, 1456, 1363, 1323, 1159, 1084 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 371.0989 [M + H]⁺ (calcd. for C₂₀H₁₅O₇, 371.1131), (negative): \( m/z \) 369.0972 [M – H]⁻ (calcd. for C₂₀H₁₄O₇, 369.0974); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid NC (14): yellow, amorphous powder; \([\alpha]_{D}^{25}\)–9.1 (c 0.07, MeOH); UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 264 (0.15), 342 (0.06) nm; IR\( \nu_{\text{max}} \) 3279, 2975, 2925, 2854, 1654, 1588, 1487, 1452, 1356, 1260, 1159, 1117, 1088 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 469.1853 [M + H]⁺ (calcd. for C₂₆H₂₉O₆, 469.1862); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid ND (15): yellow, amorphous powder; \([\alpha]_{D}^{25}\)–13.0 (c 0.08, MeOH); UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 264 (0.15), 342 (0.06) nm; IR\( \nu_{\text{max}} \) 3387, 2950, 2878, 2832, 1654, 1599, 1540, 1490, 1377, 1315, 1255, 1137, 1106, 1070 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 499.1966 [M + H]⁺ (calcd. for C₂₇H₃₁O₉, 499.1968), \( m/z \) 521.1792 [M + Na]⁺ (calcd. for C₂₇H₂₉O₉Na, 521.1788); NMR data (DMSO-d₆), see Table 1; Table 2.

Sinoflavonoid NE (16): yellow, amorphous powder; \([\alpha]_{D}^{25}\)–12.8 (c 0.11, MeOH); UV (MeOH) \( \lambda_{\text{max}} \) (log \( \epsilon \)) 265 (0.57), 349 (0.25) nm; IR\( \nu_{\text{max}} \) 3664, 2974, 2931, 1651, 1591, 1485, 1447, 1359, 1226, 1147 cm⁻¹; HR-ESI-MS (positive): \( m/z \) 469.1860 [M + H]⁺ (calcd. for C₂₆H₂₉O₆, 469.1862); NMR data (DMSO-d₆), see Table 1; Table 2.

3.5. Cytotoxicity Assay

Tumor cells were maintained in RPMI-1640 medium containing 10% heat-inactivated fetal bovine serum, penicillin (100 units/mL), streptomycin (100 μg/mL) under humidified air with 5% CO₂ at 37 °C. Exponentially growing cells were seeded into 96-well tissue culture-treated plates and precultured for 24h. The isolates were tested against MCF-7 and HepG2 cell lines, using an established MTT assay protocol [11].
4. Conclusions

Prenylated flavonoids are characterized by the presence of lipophilic prenylated group on the parent skeleton. Their structure diversity is most attributed to the different position of prenylation, and various length, further cyclization and hydroxylation of prenyl chain. With diverse chemical structure, prenylated flavonoids exhibit extensive pharmacological actions, including antioxidant, anti-inflammatory, anticoagulant, antivirus, antimicrobial antitumor [22], antigenotoxic [23], antiplasmodial [24] and estrogen regulation activities [25]. However, currently, 80% of the approximately 1100 prenylated flavonoids exist in only three plant families (Asteraceae, Cannabinaceae, Leguminosae) [22]. Consequently, their exploitation and use is limited by the narrow distribution in the plant kingdom. Forty-six flavonoids, including 37 prenylated ones, were isolated from S. hexandrum [2,8–11]. Most of them were tested for the cytotoxic activity in tumor cell lines [2,9,11]. Further phytochemical studies on S. hexandrum resulted in the isolation of 16 new prenylated flavonoids and eight known analogues. Their cytotoxic activity was evaluated against MCF-7 and HepG2 cell lines. Compound 6 was the most valuable of all tested compounds. Further research is necessary to elucidate the antitumor mechanism. This study also enriches the chemical and pharmacological diversity of prenylated flavonoids.

Supplementary Materials: The following are available online. Figures S1–S64: NMR spectra of compounds 1–16.

Author Contributions: Y.J.S. and W.S.F. designed the research; H.J.C., J.M.W., M.L.G., C.Z., R.J.H., H.C., M.L., and G.M.X. performed the research and analyzed the data; Y.J.S. wrote the paper. All authors read and approved the final manuscript.

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References

1. Zhao, C.Q.; Cao, W.; Nagatsu, A.; Ogihara, Y. Three new glycosides from Sinopodophyllum emodi (Wall.) Ying. Chem. Pharm. Bull. 2001, 49, 1474–1476. [CrossRef] [PubMed]
2. Kong, Y.; Xiao, J.J.; Meng, S.C.; Dong, X.M.; Ge, Y.W.; Wang, R.F.; Shang, M.Y.; Cai, S.Q. A new cytotoxic flavonoid from the fruit of Sinopodophyllum hexandrum. Fitoterapia 2010, 81, 367–370. [CrossRef] [PubMed]
3. Zhao, C.Q.; Zhu, Y.Y.; Chen, S.Y.; Ogihara, Y. Lignan glucoside from Sinopodophyllum emodi and its cytotoxic activity. Chin. Chem. Lett. 2011, 22, 181–184. [CrossRef]
4. Zhao, C.Q.; Huang, J.; Nagatsu, A.; Ogihara, Y. Two new podophyllotoxin glycosides from Sinopodophyllum emodi (Wall) Ying. Chem. Pharm. Bull. 2001, 49, 773–775. [CrossRef] [PubMed]
5. Zhao, C.Q.; Nagatsu, A.; Hatano, K.; Shirai, N.; Kato, S.; Ogihara, Y. New lignan glycosides from Chinese medicinal plant, Sinopodophyllum emodi. Chem. Pharm. Bull. 2003, 51, 255–261. [CrossRef] [PubMed]
6. Sun, Y.J.; Li, Z.L.; Chen, H.; Liu, X.Q.; Zhou, W.; Hua, H.M. Three new cytotoxic aryltetralin lignans from Sinopodophyllum emodi. Bioorg. Med. Chem. Lett. 2011, 21, 3794–3797. [CrossRef] [PubMed]
7. Sun, Y.J.; Li, Z.L.; Chen, H.; Liu, X.Q.; Zhou, W.; Hua, H.M. Four new cytotoxic tetrahydrofuranoid lignans from Sinopodophyllum emodi. Planta Med. 2012, 78, 480–484. [CrossRef] [PubMed]
8. Sun, Y.J.; Zhou, W.; Chen, H.; Li, Z.L.; Hua, H.M. Isolation and identification of flavonoids from the roots and rhizomes of Sinopodophyllum emodi. J. Shenyang Pharm. Univ. 2012, 29, 185–189.
9. Sun, Y.J.; Pei, L.X.; Wang, K.B.; Sun, Y.S.; Wang, J.M.; Zhang, Y.L.; Gao, M.L.; Ji, B.Y. Preparative isolation of two prenylated biflavonoids from the roots and rhizomes of Sinopodophyllum emodi by sephadex LH-20 column and high-speed counter-current chromatography. Molecules 2016, 21, 10. [CrossRef]
10. Sun, Y.J.; Sun, Y.S.; Chen, H.; Hao, Z.Y.; Wang, J.M.; Guan, Y.B.; Feng, W.S.; Zheng, X.K. Isolation of two new prenylated flavonoids from Sinopodophyllum emodi fruit by silica gel column and high-speed counter-current chromatography. *J. Chromatogr. B* 2014, 969, 190–198. [CrossRef] [PubMed]

11. Sun, Y.J.; Hao, Z.Y.; Si, J.G.; Wang, Y.; Zhang, Y.L.; Wang, J.M.; Gao, M.L.; Chen, H. Prenylated flavonoids from the fruits of Sinopodophyllum emodi and their cytotoxic activities. *RSC Adv.* 2015, 5, 82736–82742. [CrossRef]

12. Sun, Y.J.; Li, Z.L.; Chen, H.; Zhou, W.; Hua, H.M. Study on chemical constituents from the roots and rhizomes of Sinopodophyllum emodi. *Molecules* 2016, 21, 434. [CrossRef] [PubMed]

13. Sun, Y.J.; Li, Z.L.; Chen, H.; Zhou, W.; Cheng, H.; Li, Z.L.; Hua, H.M. Phenols from roots and rhizomes of Sinopodophyllum emodi. *Chin. Tradit. Herbal Drugs* 2012, 43, 226–229.

14. Yang, X.Z.; Shao, H.; Zhang, L.Q.; Zhou, C.; Xuan, Q.; Yang, C.Y. Present situation of studies on resources of podophyllotoxin. *Chin. Tradit. Herbal Drugs* 2001, 32, 1042–1044.

15. Kirmizibekmez, H.; Uysal, G.B.; Masullo, M.; Demirci, F.; Bagci, Y.; Kan, Y.; Piacente, S. Prenylated polyphenolic compounds from Glycyrrhiza iconica and their antimicrobial and antioxidant activities. *Fitoterapia* 2015, 103, 289–293. [CrossRef] [PubMed]

16. Indran, I.R.; Zhang, S.J.; Zhang, Z.W.; Sun, F.; Gong, Y.H.; Wang, X.C.; Li, J.; Erdelmeier, C.A.J.; Koch, E.; Yong, E.L. Selective estrogen receptor modulator effects of Epimedium extracts on breast cancer and uterine growth in nude mice. *Planta Med.* 2014, 80, 22–28. [CrossRef] [PubMed]