Developmental pathology, accidents, and tumor resection frequently cause bone loss,\(^1,^2\) which is still a problem in orthopedic and reconstructive surgery. To resolve these problems, autografts and allografts have long been used to fill bone defects caused by surgery, trauma or disease.\(^3\) Autografts have the advantage of being biocompatible, osteoconductive and osteoinductive. These tissue grafts require additional surgery, which can cause trauma and donor site morbidity that leads to an increase in surgical time and hospital costs.\(^4\) Allografts overcome these problems, but can potentially introduce the risk of transmission infection or provoke an immunogenic response. To overcome these problems, a variety of osteoconductive biomaterials such as ceramics and titanium alloys have been considered as bone graft substitutes.\(^5\) Titanium (Ti), which is biointert, does not bond directly but has close contact with bone. Other synthetic bone grafts are made from biphasic calcium phosphates, and beta-tricalcium phosphate, calcium polylphosphate, hydroxyapatite (HA),\(^6\) and carbonated hydroxyapatite (CHA). Recently, osteoconductive biomaterials suggest that they can be made in greater quantities and present no immunogenic concerns.\(^7\) These biomaterials have gained much attention, not only due to its ionic composition similar to bones but also due to its functional properties favorable for bone growth such as resorbability by living cells.\(^8\) Moreover, some investigators have

**Biologic Response to Carbonated Hydroxyapatite Associated with Orthopedic Device: Experimental Study in a Rabbit Model**

Samira Jebahi\(^1,^2,^3,^4,^5\) · Mongi Saoudi\(^3\)

Riad Badraoui\(^1\) · Tarek Rebai\(^1\)

Hassane Oudadesse\(^1\) · Zoubaier Ellouz\(^5\)

Hassib Keskese\(^5\) · Abdelfattah El Feki\(^1\)

Hafed El Feki\(^1\)

\(^1\)Laboratory of Histology, Medicine Faculty of Sfax; \(^2\)Laboratory of Science of Materials and Environment, Sciences Faculty of Sfax; \(^3\)Animal Ecophysiology Laboratory, Department of Life Sciences, Sciences Faculty of Sfax, Sfax, Tunisia; \(^4\)University of Rennes 1, UMR CNRS 6226, Campus of Beaulieu, 263 av. General Leclerc, 35042 Rennes, France; \(^5\)Laboratory of Orthopaedic and Traumatology, Medicine Faculty of Sfax, Sfax, Tunisia

**Background:** Carbonated hydroxyapatite (CHA) and related calcium phosphates have been studied for many years as implant materials due to their similarity with the mineral phase of bone. The main limitation of CHA ceramics as well as other bioactive materials is that they have poor mechanical proprieties. It is thought that the mechanical device can cause an increase in metabolic activity and bone healing. In this study we investigated the reactivity and tissue behaviour of implanted CHA biomaterial reinforced by mini external fixator. **Methods:** The evaluation of biomaterial biocompatibility and osteogenesis was performed on a rabbit model over a period of 6 weeks by radiological, histological and scanning electron microscopy (SEM) coupled with energy dispersive X-ray SEM-energy-dispersive X-ray (EDX) analysis. **Results:** While rabbits treated with CHA exhibited more bone formation, and fibrous tissue was observed when empty bone defects were observed. EDX analysis detected little calcium and phosphorus on the surface of the bone that was not implanted, while high content of calcium (62.7%) and phosphorus (38%) was found on the interface bone cement. **Conclusions:** Bone repairing showed that the mini external fixator stimulated the ossification which was pushed when grafted by CHA. This effect may play an important role in the prevention of implant loosening.

**Key Words:** Biocompatible materials; Testing bone substitute
Biologic Bone Response to Carbonated Hydroxyapatite

reported that HA is useful as an osteoporosis model. The present study was undertaken to evaluate the regenerative effects of both CHA and mini external fixator in large cortical defect. The utilization of these materials as bone grafts may reduce the need for autogenous bone graft, increase stability of the implant, lead to faster bone regeneration and facilitate the healing process.

MATERIALS AND METHODS

Animal model

New Zealand White Male rabbits weighing 1.6-1.9 kg and bred in the Central Animal House were used in this study. The animals were fed on a pellet diet (Sico, Sfax, Tunisia) and water ad libitum. The animals were maintained in a controlled environment under standard conditions of temperature (22±2°C) and humidity (55±5%) with an alternating light-and-dark (12/12 hours) cycle. All rabbits were acclimatized for one week prior to the start of the experimentation. They were divided into 3 groups, each one containing 5 rabbits: control rabbits (T), intact tibias without the surgical creation of bone defects; operated rabbits with implant (IP), bone defects filled with CHA to be tested; operated rabbits without implant (NIP), bone defects without any filler.

Surgical and postoperative protocol

CHA powder (particle ≤ 20 μm), was sterilized by γ-irradiation from a 60Co Source gamma irradiation at a dose of 25 Gy (The-ratron external beam teletherapy, Equinox, Ottawa, ON, Canada), and then implanted and stabilized by mini external fixator (Fig. 1A). Anesthesia was induced with 10 mg/kg of ketamine (Ketaminol, Intervet International GmbH, Unterschleisheim, Germany) and 0.1 mg/kg of Xylazine (Rompun, Bayer Healthcare, Puteaux, France). Supplemented local anesthesia was applied after 15 to 20 minutes using 4 mg/kg carprofen (Rimadyl, Pfizer, Paris, France). Cutaneous and under cutaneous incisions on the inner face of the tibia followed by an opening of the muscular aponeurose were carried out. A gap (1 cm of diameter) in the mid-diaphyseal level of the tibia was created aseptically. CHA filled the loss of osseous substance only for the first group (IP) (Fig. 1B). On the other hand, no filling was made for the second group (NIP) (Fig. 1C). During a period of 6 weeks, the subjects were checked daily for clinical lameness or other complications. The handling of the animals was approved by the Tunisian Ethical Committee for the care and use of laboratory animals.

Scanning electron microscopy (SEM) characterization

Implanted bones were analyzed by SEM, using a JEOL JSM 6301F (Tokyo, Japan). Polymethylmethacrylate embedded specimens obtained from the histological preparations were mounted on the SEM. Quantification for calcium (Ca, mol%), phosphorus (P, mol%) and magnesium (Mg, mol%) was determined by wavelength dispersive X-ray spectroscopic, and named CHA (A), bone (B) and interface between the implant and bone (C), the ratio of calcium to phosphorus (Ca/P) was calculated by dividing the values obtained from calcium (mol%) and phosphorous (mol%) at each point.

Fig. 1. A mini external fixator (A), gap with (B) and without biomaterial (C).
Radiographic analysis

To assess bone and CHA density, conventional X-ray radiographs (Polymobil III, Siemens AG Wittelsbacherplatz 2 D-80333, Muenchen, Germany) were taken following craniocaudal views. Radiography evaluated the quality of CHA osteointegration and the levels of resorption.

Microscopic evaluation

The callus formation of each animal was carefully dissected out. The sections were taken on both sides of the implants (CHA-bone specimens were cut sagittally or transversally). The undecalcified CHA-bone specimens were defleshed and fixed in 10% formaldehyde for 24 hours at 4°C. The next day, the bone samples were embedded in methylmetacrylate without previous decalcification, as previously reported. Seven micrometer-thick coronal sections were cut dry in parallel to the long axis of the tibia bone samples, using a heavy-duty microtome (Polycut S Reichert Jung, Heidelberg, Germany) equipped with tungsten carbide knives. The sections were stained with a modified Goldner trichrome. Other sections were stained with Giemsa fast (10%). In addition, the decalcified samples of CHA-bone specimens were examined with hematoxylin and eosin staining. The histological observations in the rabbit tibias after 6 weeks permitted us to study the repairing potency of the osseous substitute. The histological features, such as intensity of inflammatory reaction, cellular activity, necrosis, hemorrhage, presence of connective tissue, apposition of bone tissue and presence of particles of biomaterial were recorded in a descriptive way.

Statistical analysis

The statistical analysis of the data was made using the Student’s t-test. All values are expressed as means±standard error. Differences are considered significant at the 95% confidence level (p<0.05).

RESULTS

General features and behavior

The animals were allowed to move freely in their cages after surgical operation. All surgical interventions were performed without complications. The postoperative healing was uneventful in all rabbits. The rabbits walked without a limp after 5 days. The operated rabbits remained healthy and showed no signs of discomfort or lameness. Any infection surrounding the fixator or the osseous-substitute was detected. No problems occurred in the bone defects during surgeries and healing, i.e., no failure of any implantation site was observed during the whole study period. All the implants appeared to be fixed securely in the tibias.

SEM findings

The results presented by energy-dispersive X-ray analysis (Fig. 2) in the IP group revealed a high distribution of Ca and P by (62.7%) and (38%), respectively (Table 1). The percentage of Ca and P determined in this group was similar to those determined in the T group. Moreover, the Ca/P ratio (1.65) in CHA-bone interface was close to that determined in the T group (1.63). However, Ca/P ratio (1.58) in the surface of the NIP bone tissue (Fig. 3) was lower than in the T group (Table 2). In addition, in the IP group the Mg quantity (0.94%) was similar to the T group (1.20%). This rate of Mg presented only about (0.01%) in NIP rabbits.

Fig. 2. (A) Calcium and (B) phosphorus mapping energy-dispersive X-ray data carbonated hydroxyapatite.
Macroscopic and radiographic evaluation

Macroscopically, all experimental bone seemed to be well osteointegrated within the bone tissue. No obvious inflammatory reaction or bone tissue formation in the adjacent soft tissue was observed. The implant was well osteointegrated in IP rabbits. Bone defects filled with CHA detected by radiology were slightly radio-opaque. The evaluations of new calli graft materials allowed the checking of the implant position in the IP group (Fig. 4A), and osseous consolidation for the NIP group (Fig. 4B).

Qualitative histological findings

Clinically and histologically, the implanted biomaterials were well tolerated. There were no signs of rejection, necrosis or infection. We noted the presence of mesenchymal stem cells surrounded by amorphous and homogeneous substances (Fig. 5A). Gradually, these cells appeared to be transformed into fibroblasts resorption which metamorphosed into active osteoblasts at 6 weeks. Furthermore, we noticed the presence of an array of randomly oriented collagen (Fig. 5B). The newly formed bone showed new blood vessels (cavities of small arteries) (Fig. 5D). Histological studies also showed the deposition of salts on collagen fibres (Fig. 5C). The ossification was set up gradually with periphery towards the center and thus in a centripetal way (Fig. 5E, F). The NIP rabbit (empty gap defects) indicated that only endochondral ossification formed (Fig. 5I). Moreover, a fibrous

| Table 1. Quantification for Ca, P, and Mg determined from EDX analysis of an area including bone, CHA bone and the interface |
|-----------------|---|---|---|---|
| Element (mol%)  | Ca    | P    | Mg    | Ca/P  |
| Interface       | 62.7 ± 5 | 38 ± 2 | 0.94 ± 0.043 | 1.65 |
| Bone            | 60.75 ± 7 | 37.26 ± 0.2 | 1.20 ± 0.043 | 1.63 |
| CHA             | 65.00 ± 36 | 38.39 ± 0.4 | 0.18 ± 0.012 | 1.69 |

EDX, energy-dispersive X-ray; CHA, carbonated hydroxyapatite.

| Table 2. Quantification for Ca, P, and Mg determined from EDX analysis of a new bone area without implant |
|-----------------|---|---|---|---|
| Element (mol%)  | Ca    | P    | Mg    | Ca/P  |
| New bone without implant | 60.02 ± 80 | 37.8 ± 5.1 | 0.010 ± 0.0023 | 1.58 |

EDX, energy-dispersive X-ray.
conjunctive tissue layer with low bone density also appeared (Fig. 5J-M). The direction of progression was always of the centripetal type. We noted that IP rabbits exhibited more intensive bone remodeling when compared to the NIP group. Despite the important bone tissue regeneration in the IP group, it’s considered as limited when compared to the T group (Fig. 5N, O). SEM analysis confirmed the biocompatibility (Fig. 5G, H) and osteointegration of CHA (Fig. 6) in the bone tissue.

DISCUSSION

There is currently great interest in the development of materials capable of helping bone repair. In the present study, the osteointegration of CHA with the assistance of mini external fix-
Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

REFERENCES

1. Huh GY, Lee SK. Extraskeletal osteosarcoma: a case report. Korean J Pathol 1988; 22: 489-94.
2. Kotoura Y, Yamamuro T, Kasahara K. Application of biomaterials in the treatment of osteosarcoma. Gan To Kagaku Ryoho 1987; 14(5 Pt 2): 1423-9.
3. Friedlaender GE, Horowitz MC. Immune responses to osteochondral allografts: nature and significance. Orthopedics 1992; 15: 1171-5.
4. Einhorn TA, Lane JM, Burstein AH, Kopman CR, Vigorita VJ. The healing of segmental bone defects induced by demineralized bone matrix: a radiographic and biomechanical study. J Bone Joint Surg Am 1984; 66: 274-9.
5. Tsukeoka T, Suzuki M, Ohitsuaki C, et al. Enhanced fixation of implants by bone ingrowth to titanium fiber mesh: effect of incorporation of hydroxyapatite powder. J Biomed Mater Res B Appl Biomater 2005; 75: 168-76.
6. Tienboon P, Jaruwangsanti N, Kuptmanakul S. Efficacy of hydroxyapatite in pedicular screw fixation in canine spinal vertebra. Asian Biomed 2009; 3: 177-81.
7. Miño-Fariña N, Muñoz-Guzón F, López-Peña M, et al. Quantitative analysis of the resorption and osteoconduction of a macroporous calcium phosphate bone cement for the repair of a critical size defect in the femoral condyle. Vet J 2009; 179: 264-72.
8. Liljensten E, Adolfsson E, Strid KG, Thomsen P. Resorbable and non-resorbable hydroxyapatite granules as bone graft substitutes in rabbit cortical defects. Clin Implant Dent Relat Res 2003; 5: 95-101.
9. Badraoui R, Abdelmoula NB, Sahnoun Z, Fakhfakh Z, Rebai T. Effect of subchronic exposure to tetradifon on bone remodelling and metabolism in female rat. C R Biol 2007; 330: 897-904.
10. Doi Y, Shibutani T, Moriwaki Y, Kajimoto T, Iwayama Y. Sintered carbonate apatites as bioreosorbable bone substitutes. J Biomed Mater Res 1998; 39: 603-10.
11. Landi E, Celotti G, Logroscino G, Tampieri A. Carbonated hydroxyapatite as bone substitute. J Eur Ceram Soc 2003; 23: 2931-7.
12. Pan H, Zhao X, Darvell BW, Lu WW. Apatite-formation ability: predictor of “bioactivity”? Acta Biomater 2010; 6: 4181-8.
13. Okumura M, Ohgushi H, Dohi Y, et al. Osteoblastic phenotype expression on the surface of hydroxyapatite ceramics. J Biomed Mater Res 1997; 37: 122-9.
14. Mastrogiacomo M, Scaglione S, Martinetti R, et al. Role of scaffold internal structure on in vivo bone formation in macroporous calcium phosphate biomaterials. Biomaterials 2006; 27: 3230-7.
15. Rey C, Renugopalakrishnan V, Collins B, Glimcher MJ. Fourier transform infrared spectroscopic study of the carbonate ions in bone mineral during aging. Calcif Tissue Int 1991; 49: 251-8.
16. Ooms EM, Wolke JG, van der Waerden JP, Jansen JA. Trabecular bone response to injectable calcium phosphate (Ca-P) cement. J Biomed Mater Res 2002; 61: 9-18.
17. Yuan H, Li Y, de Bruijn JD, de Groot K, Zhang X. Tissue responses of calcium phosphate cement: a study in dogs. Biomaterials 2000; 21: 1283-90.
18. Theiss F, Apelt D, Brand B, et al. Biocompatibility and resorption of a brushite calcium phosphate cement. Biomaterials 2005; 26: 4383-94.
19. Eriksson C, Ohlson K, Richter K, Billerdahl N, Johansson M, Nygren H. Callus formation and remodeling at titanium implants. J Biomed Mater Res A 2007; 83: 1062-9.