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Live imaging of wound angiogenesis reveals macrophage orchestrated vessel sprouting and regression

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Abstract

Wound angiogenesis is an integral part of tissue repair and is impaired in many pathologies of healing. Here, we investigate the cellular interactions between innate immune cells and endothelial cells at wounds that drive neoangiogenic sprouting in real time and in vivo. Our studies in mouse and zebrafish wounds indicate that macrophages are drawn to wound blood vessels soon after injury and are intimately associated throughout the repair process and that macrophage ablation results in impaired neoangiogenesis. Macrophages also positively influence wound angiogenesis by driving resolution of anti-angiogenic wound neutrophils. Experimental manipulation of the wound environment to specifically alter macrophage activation state dramatically influences subsequent blood vessel sprouting, with premature dampening of tumour necrosis factor-α expression leading to impaired neoangiogenesis. Complementary human tissue culture studies indicate that inflammatory macrophages associate with endothelial cells and are sufficient to drive vessel sprouting via vascular endothelial growth factor signalling. Subsequently, macrophages also play a role in blood vessel regression during the resolution phase of wound repair, and their absence, or shifted activation state, impairs appropriate vessel clearance.

Keywords angiogenesis; inflammation; macrophages; neutrophils; wound

Subject Categories Development & Differentiation; Vascular Biology & Angiogenesis

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Introduction

Healthy tissues rely on a plentiful vascular supply in order to receive the appropriate amount of oxygen and nutrients and to efficiently remove waste products. This reliance increases in instances of tissue damage where the healing process leads to intensive metabolic demands due to increased local cell signalling, migration and proliferation. Most tissue damage involves the disruption or destruction of the local vasculature, which must then be restored and increased in density to facilitate tissue repair. This process of new blood vessel growth from existing vessels, termed neoangiogenesis, must be tightly controlled; delayed or reduced neoangiogenesis may result in chronic non-healing wounds (Nunan et al., 2014), while overexuberant neoangiogenesis is associated with the formation of keloid scars and other forms of pathological fibrosis (Johnson & Wilgus, 2014). These two extreme outcomes of aberrant and dysregulated wound healing represent a substantial social and financial health burden (Sen et al., 2009). Therefore, understanding mechanisms of wound repair that might lead to improvements in wound care generally and management of wound neoangiogenesis in particular are highly relevant and desirable.

The process of wound healing progresses through a series of stereotypical, overlapping phases (Eming et al., 2014; Johnson & Wilgus, 2014; Cash & Martin, 2016). Innate immune cells play significant roles throughout these wound healing phases, mediating host defence, wound debridement, inflammatory status and signalling to coordinate cellular activities leading to appropriate migration, proliferation and tissue restoration (Eming et al., 2014; Martin & Nunan, 2015). Extensive studies of macrophages during developmental morphogenetic episodes have revealed that they associate with and support blood vessel sprouting and anastomosis in the hindbrain and retina of embryonic and neonatal mice (Fantin et al., 2010; Rymo et al., 2011), as well as the trunk vasculature in zebrafish embryos (Fantin et al., 2010). Some vessel regression during post-natal retinal
Macrophages drive wound angiogenesis

Macrophages are thought to play key roles in developmental angiogenesis in both mouse and fish and appear to interact with blood vessel sprouts and anastomoses during the establishment of the embryonic vascular network (Fantin et al., 2010; Stefater et al., 2011). We reasoned that a similar association might be important in the context of wound angiogenesis since wound healing frequently recapitulates developmental processes (Martin & Parkhurst, 2004). Firstly, we characterised the wound neoangiogenic response in mice following a simple dorsal skin punch biopsy (Fig 1A) by immunostaining for the endothelial cell marker CD31 in wounds at time-points throughout the repair process (Fig 1B). Our studies show that the first new blood vessels appear at 3 days post-injury (DPI) and are highly disorganised and poorly interconnected (Fig 1B and C). Wound blood vessels subsequently increase in number and branch pattern until they peak at 7 DPI, after which they resolve away from the wound area, becoming more organised and beginning to return to a distribution and alignment pattern resembling unwounded skin by 14 DPI (Fig 1B–D). This time course of events complements and expands on previous studies examining sprouting and regression of wound blood vessels using completely different technologies such as MicroCT angiography imaging (Urao et al., 2016).

Results

Macrophages are associated with wound angiogenesis in mammals

Macrophages are associated with blood vessels in mouse wounds.

Figure 1. Macrophages are associated with blood vessels in mouse wounds.

A  En face schematic illustration of a mouse wound, showing macrophage–blood vessel sprouts entering a full-thickness skin wound in relation to the overlying scab and associated inflammatory cells during wound healing.

B  Representative multiphoton projection images of mouse wounds stained for endothelial cells (CD31) over the full duration of blood vessel infiltration and resolution.

C  Quantification of total wound area occupied by blood vessels, based on pixel count (see Materials and Methods) and measured from time course represented in (B). Extent of wound vasculature increases to a maximum attained at 7 DPI, after which wound vasculature resolves towards uninjured levels by 14 DPI. N = 6 independent mice per timepoint.

D  Quantification of blood vessel alignment, based on pixel orientation (see Materials and Methods) and measured from time course represented in (B). Vessels begin to restore their coherence and orientation by 14 DPI. N = 6 independent mice per timepoint.

E  Representative multiphoton projection images of wound tissue co-stained for endothelial cells (CD31, red) and macrophages (CD68, green), showing stereotypical association of macrophages with blood vessel sprouts during vessel infiltration into wound site and their subsequent regression from the wound. Asterisks indicate macrophage–sprout interaction; arrows indicate phagocytosed vessel material within macrophages. These are matched with representative images of electron micrographs showing macrophage–endothelial cell interactions, taken at the repair time course of the repair process, measured from the time course represented in (E). N = 5 independent mice per timepoint.

F  Quantification of blood vessel tip–macrophage association during the repair time course of the repair process, measured from the time course represented in (E).

Data information: Error bars indicate mean ± SD. Scale bars: (B) = 200 μm, (E) = 10 μm.
Figure 1.
We next examined for potential interactions between macrophages and blood vessels at various stages throughout the repair period. Co-immunostaining of excised wounds for macrophage and endothelial cell markers (CD68 and CD31, respectively) throughout wound repair and resolution phases revealed several typical interactions. Initially, during early and mid stages of wound repair from 3 to 7 DPI, macrophages appeared to be associated with the tips of newly formed neoangiogenic sprouts (Fig 1E and F). During later stages of wound repair, from 7 to 10 DPI, macrophages exhibited a shift in their associations with vessels, with less sprouting tip contact and a “hugging” behaviour with vessels that no longer appeared to be actively sprouting (Fig 1E). At the latest stages of repair, 10–14 DPI, some macrophages appeared to contain engulfed endothelial cell material (19.3 ± 3.8% of total macrophages) and this was confirmed by use of VE-Cadherin antibody as an alternative vessel marker (Figs 1E and EV1A and B, Movie EV1). We also observed each of these various types of macrophage–endothelial cell contacts at the ultrastructural level by transmission electron microscopy, which reveals details of such interactions. For example, in the period when vessel resolution is occurring, we see macrophages in the process of phagocytosing cell debris in close proximity to vessels (Fig 1E). Together, these observations suggest that the functions of macrophages in regulating angiogenesis may change throughout the wound repair time course, from nurturing and supporting endothelial cell sprouts during early stages of repair through to clearing of endothelial cells during later stages of wound resolution.

Macrophages are associated with neoangiogenesis throughout the repair of zebrafish wounds

Our next objective was to directly observe these dynamic interactions between macrophages and blood vessels in living tissue. To this end, we developed several wounding protocols for zebrafish larvae, which are translucent and therefore more amenable to live imaging than the opaque tissues of mice. We characterised the time course of wound angiogenesis in zebrafish by utilising the Tg(fli1:GFP) transgenic line, which marks all blood vessels. We first performed a relatively large mechanical needle-stick flank wound with a 30-gauge needle to the dorsal somite area opposite the cloaca and monitored the subsequent wound angiogenic response (Fig 2A). Zebrafish wound neoangiogenesis is temporally very similar to that of mammalian wounds, with the first blood vessel sprouts appearing within the wound at 1–3 DPI (Fig 2B). Unlike mature, undamaged vessels, these sprouts, and their resulting new vessels, are initially leaky, as evidenced by loading of blood vessels with high molecular weight rhodamine–dextran, which under unwounded conditions is too large to travel across the vessel wall (Fig EV2A and B). These new sprouts proceed to anastomose and form tortuous and densely branched vessels at 4–6 DPI (Fig 2B), by which stage they cease to leak loaded dextran (Fig EV2A and B). After reaching a post-wounding peak of complexity at about 6 DPI as measured by quantification of individual sprout and vessel node numbers, as well as total vessel length, these vessels subsequently resolve back to a pattern resembling that of uninjured tissue by 10 DPI (Fig 2B and C). Utilising a double-transgenic fish labelling arteries in green, Tg(dll4:GFP), and all vessels in red, Tg(kdrl:mCherry-CAAX), we were able to determine the nature of vessels involved in repair. In contrast to some studies of embryonic development where, for example, vessel sprouting and vascularisation of the spinal cord appear to proceed primarily from venous endothelium (Wild et al., 2017), we observed that both arteries and veins readily contribute to wound angiogenesis, resulting in chimeric wound vessels (contribution towards artery length ranged from 55.6% to 84.7%, while contribution towards vein length ranged from 15.3% to 44.4%, n = 10 independent fish at 4 DPI, Fig 2B). To determine whether physical vessel damage is required to stimulate
Figure 2.
wound neoangiogenesis, we performed smaller scale mechanical injuries using a fine tungsten needle, targeting somitic tissue but without direct damage to any vessel. Even without vessel damage, these wounds attracted new vessel sprouts, indicating that signalling from a non-endothelial exogenous source such as immune cells is responsible for driving wound neoangiogenesis (Fig EV2C).

Utilising transgenic fish lines with labelled neutrophils Tg(mpox:GFP) and macrophages Tg(mpox:mCherry), we next examined the response of innate immune cells to a 30-gauge needle-stick injury (Fig 2D and E). Neutrophil recruitment to the wound peaks at 12 HPI and then resolves back to levels observed in uninjured tissue by 4 DPI (Fig 2D and E). Macrophages take longer to reach their peak in the wound area, at about 2 DPI, and also longer to resolve, with some cells still present up to 10 DPI and beyond (Fig 2D and E). These invading waves of neutrophils and later macrophages at the site of injury are similar to those reported in previous wound studies (Gray et al., 2011). In combination with the Tg(fli:GFP) transgenic line, we examined interactions between blood vessels and both of these recruited immune lineages at the wound site. Neutrophils are rapidly drawn to the vicinity of wound blood vessels, but are only ever transiently observed in direct contact with sprouting tips and soon leave the immediate vicinity of wound angiogenesis. By contrast, macrophages form direct intimate associations with newly formed sprouting tips at early repair timepoints (Fig 2F and G), similar to what we observe in mouse wounds. These results indicate that the stereotypical interactions between macrophages and blood vessels observed during mammalian wound angiogenesis are largely conserved in zebrafish wounds.

To better analyse these cellular interactions over the full time course of vessel repair and resolution, we performed a second, smaller type of lesion by microlaser ablation (Fig 2H); these resolve in a shorter time window, thus enabling imaging for the full duration of repair. We performed microlaser ablations of trunk blood vessels of Tg(fli:GFP); Tg(mpox:GFP); Tg(mpox:mCherry) triple transgenic fish and timelapse imaged the subsequent repair process. Small, partial intersegmental vessel (ISV) ablations allowed for imaging of the entire repair process through to vessel patency, while larger total ISV ablations allowed for a more extensive examination of immune cell–vessel interaction during sprouting. We observed that vessel sprouting at the wound site appears to follow a similar process to that reported for embryonic development of the ISVs (Isogai et al., 2001, 2003), with rapid filopodial extension and sprout growth. If the extent of vessel loss is less than 30 μm (small ablation), then vessel outgrowth generally occurs in a ventral to dorsal direction and leads to anastomosis and luminaisation by approximately 600 min post-injury (Fig 2I, Movie EV2). For full ISV ablations where vessel damage is greater than 60 μm, filopodial extension and sprout formation occur from both the dorsal aorta (DA) and dorsal longitudinal anastomotic vessel (DLAV); vessel tips extend filopodia in a highly dynamic fashion apparently sensing directionality before meeting and undergoing fusion at a midpoint between the two major vessels (Fig EV2D, Movie EV3). Movies of partial ISV ablations confirmed that neutrophil interactions with damaged blood vessels occur early and are generally transient (between 30 and 120 min post-injury), after which neutrophils generally had no further association with the vessel repair process (Fig 2I–K, Movie EV2). By contrast, interactions between macrophages and damaged blood vessels persist for the full duration of vessel repair (Fig 2I–K, Movies EV2 and EV4). Macrophages initially localise to the gap between the two damaged vessel tips, after which they often wrap entirely around and "hug" the repairing blood vessel during the sprouting and early anastomosis phases (approx. 300 min post-injury); these intimate contacts are generally maintained until after vascular patency is restored (approx. 750 min post-injury; Fig 2I and J, Movies EV2 and EV4). To confirm these observations, we performed complementary experiments on Tg(kdr:mCherry-CAAX); Tg(mpox:GFP); Tg(mpox:mCherry) fish (Fig EV2E, Movie EV5). These live-imaging experiments strongly indicate important roles for macrophages that change during wound angiogenesis and its subsequent regression.

Macrophages drive wound angiogenesis

To examine whether interactions between macrophages and blood vessels at the site of damage were directly facilitating wound neoangiogenesis, we utilised the metronidazole–nitroreductase system to specifically ablate macrophages. The double-transgenic line Tg(mpox:Gal4FF); Tg(UAS:NfsB-mCherry), which drives nitroreductase enzyme via the Gal4-UAS system under the control of the macrophage mpox promoter and enables efficient ablation of macrophages after treatment with metronidazole (Palha et al., 2013). We observe > 90% macrophage ablation within 48 h of exposure to metronidazole, with no detectable associated developmental defects (Fig EV3A). Recovery of metronidazole-treated fish by transfer into fresh water post-injury led to a partial restoration of macrophage numbers at the wound site at 4 DPI, by comparison to continually treated fish (Fig EV3C–E).

In the context of wound neoangiogenesis, ablation of macrophages prior to injury and throughout the entire first four days of repair lead to a complete failure of wound neoangiogenesis (Fig 3A and B). To complement this chemical-genetic approach, we used a pharmacological method for depleting macrophages involving the injection of toxic clodronate liposomes, an ablation regime previously utilised in mice (Van Rooijen & Sanders, 1994) and zebrafish regenerative studies (Carrillo et al., 2016; Figs 3A and B, and EV3B). This alternative macrophage ablation method resulted in similar retardation of wound angiogenesis (Fig 3A and B). Short-term metronidazole treatment up to the moment of injury followed by recovering the fish in fresh water during the injury repair period resulted in a partial blockade of wound angiogenesis, with some new vessels present at 4 DPI (Fig EV3C–E). Together, these results confirm that macrophages are required for neoangiogenesis at the wound and indicate that their impact on the angiogenic process is proportional to the extent of the macrophage response.

To more precisely live image the differences in vessel sprouting in the presence or absence of macrophages, we generated microlaser ablations of entire individual blood vessels in either Tg(fli:GFP); Tg(mpox:GFP); Tg(mpox:mCherry) transgenic fish or Tg(fli:GFP); Tg(mpox:Gal4FF); Tg(UAS:NfsB-mCherry) transgenic fish, treated with metronidazole as a control or to ablate macrophages throughout the repair process, respectively (Fig 3C, Movies EV6 and EV7). Macrophage ablation resulted in poorly directed and unsupported filopodial sprouting from leading tip cells, and a failure...
of sprout extension and anastomosis. We also observed that in the absence of macrophages, neutrophils are retained at the wound site and maintain long-term associations with damaged blood vessel tips rather than the transient interactions that occur in wounds with an intact macrophage response (Fig 3C–E, Movie EV7). Conversely, we observed contact-dependent neutrophil displacement from vessel tips driven by macrophages in the unablated situation, similar to that seen previously in other wound scenarios (Tauzin et al., 2014). To test whether these neutrophils might be inhibitory to wound angiogenesis, we performed quantitative reverse transcription–polymerase...
chain reaction (quantitative PCR) on FAC-sorted neutrophils from early stab wounds; these neutrophils exhibit increased expression of anti-angiogenic genes relative to those from unwounded fish (Fig 3F). Specifically, early wound neutrophils express a soluble, truncated version of vegf receptor 1, sflt1, which has a high affinity for vegf but lacks a membrane domain (Kendall & Thomas, 1993). We find that it is only this truncated version of vegf receptor 1 that is upregulated and that primers designed specifically against the membrane version of sflt1 indicated a lack of expression in neutrophils (Fig EV3F). Expression of sflt1 has previously been implicated in fine-tuning vegf signalling during developmental angiogenesis, being expressed by vessel stalk cells and thereby acting as a localised vegf sink, resulting in maximised vegf signalling and guidance to vessel tip cells (Chappell et al., 2009). We further show that early wound macrophages express increased levels of cyba and tyrk, a pair of redox-regulated Src family kinase genes normally associated with appropriate macrophage recruitment and subsequent neutrophil reverse migration (Tauzin et al., 2014; Fig 3G). These results, together with our in vitro data (see later), indicate that early wound neutrophils may provide an initial angiogenic brake and that macrophages are involved in dispersing neutrophils to remove this brake, as well as positively directing vascular sprouts at the wound site by supporting and stabilising their initial outgrowth as they progress to vessel anastomosis.

The activation state of macrophages at the wound site influences the extent of neoangiogenesis in zebrafish wounds

Tissue inflammation at the site of injury is a typical early step in wound repair. Since macrophages are known to switch phenotypic state during the repair period (Novak & Koh, 2013), we reasoned that this may influence their capacity to regulate various aspects of wound neoangiogenesis and subsequent vessel resolution throughout the wound repair process. To observe the number of pro-inflammatory, activated macrophages at the wound site over time, we utilised the Tg(tnfα:GFP); Tg(mpeg1: mCherry) double-transgenic fish, since tumour necrosis factor-α (tnfa) is a known marker of the pro-inflammatory phenotypic state (Parameswaran & Patial, 2010; Marjoram et al., 2015). After tissue wounding, we observed a wave of pro-inflammatory macrophages constituting more than 50% of the total macrophages at the wound at 1 DPI but decreasing to almost none by 4 DPI (Fig 4B–E). In order to manipulate the inflammatory state of wound macrophages, fish were treated immediately after wounding with either lipopolysaccharide (LPS) as a non-specific immunostimulant (Fig EV4A) or with recombinant zebrafish interferon gamma (Ifng-γ) as a more specific immunostimulant (Fig 4A and B). In another series of wounded fish, we suppressed wound inflammation using hydrocortisone (Fig EV4A) or recombinant zebrafish interleukin-10 (Il-10) (Fig 4A and B). These contrasting treatments allowed us to manipulate macrophage phenotype either towards or away from a tnfα-positive, pro-inflammatory phenotype (Fig 4A). As expected, we observed an increase in the proportion of tnfα-positive, inflammatory wound macrophages when treated with either immunostimulant, versus a decrease in proportion of inflammatory wound macrophages when treated with either immunosuppressive agent (Figs 4B, D and E, and EV4A–C). We next examined how the modulated wound inflammatory response impacted wound neoangiogenesis. Our experiments revealed a substantial decrease in the level of blood vessel sprouting and a general impairment of wound neoangiogenesis following treatment with either of the immunosuppressive agents for needle-stick wounds (Figs 4B and F, and EV4A and D). Conversely, the degree of wound neoangiogenesis was generally increased when treated with either immunostimulatory agent (Figs 4B and F, and EV4A and D). However, injection of high molecular weight dextran into LPS-treated fish, and to a lesser extent Ifng-γ-treated fish, indicated that the newly established blood vessels at these wounds were leakier than those found at equivalent stages in untreated wounds at the same stage post-wounding (4 DPI; Fig EV4E).

To manipulate wound macrophage phenotype using complementary genetic approaches, we also wounded fish mutant for colony-stimulating factor 1 receptor a (csf1ra). csf1ra mutant zebrafish are

Figure 3. Macrophages are necessary for wound angiogenesis and act to stabilise wound blood vessel sprouting tips.

A Representative confocal projection images of needle-stick injured Tg(fli1:GFP); Tg(mpeg1:mCherry) or Tg(fli1:GFP); Tg(mpeg1:KalTA4); Tg(UAS: nbs8-mCherry) zebrafish in combination with various treatments (as indicated), for ablating macrophages (red) during the peak stage of vessel (green) sprouting (4 DPI). Boxed area denotes wound site. In control (DMSO vehicle) and metronidazole-treated fish lacking nitroreductase expressing macrophages, a normal inflammatory response and wound angiogenesis is observed. Upon ablation of macrophages using the nitroreductase–metronidazole system, wound angiogenesis is almost completely blocked. The same is true when macrophages are ablated using clodronate liposome injections.

B Quantification of total blood vessel length in the presence/absence of macrophages at the wound site, measured from images represented in (A), using Angioanalyser. N = 16 independent fish per condition. Statistical significance, as determined by one-way ANOVA, is P ≤ 0.0001. Subsequent Bonferroni multiple comparison test, determines level of significance, as indicated. Significance values:** P ≤ 0.0001.

C Images taken from timelapse movies of 4 DPI laser wounded Tg(fli1:GFP); Tg(mpeg1:mCherry), control or Tg(fli1:GFP); Tg(mpx:GFP); Tg(mpeg1:KalTA4); Tg(UAS: nbs8-mCherry) macrophage ablation transgenic zebrafish, treated with metronidazole and imaged 30–930 min post-injury (MPI). Boxed area denotes wound site. Arrowheads indicate position of damaged, sprouting vessel tips.

D Quantification of proportion of vessel repaired within 930 MPI, measured from movies represented in (C). N = 8 independent fish per condition. Statistical significance is indicated, as determined by two-tailed t-test.

E Graphical representation of typical cellular interaction events and their time course, measured from movies represented in (C). N = 8 independent fish per condition.

F Gene expression of FAC-sorted neutrophils extracted from Tg(mpx:GFP), Tg(mpeg1:mCherry) needle-stick wounded fish at 12 HPI, showing relative gene expression for sflt2 compared to unwounded fish at 4.5 DPI. N = 50 independent fish per condition per replicate, 3 replicates. Statistical significance is indicated, as determined by two-tailed t-test.

G Gene expression of FAC-sorted macrophages from Tg(mpx:GFP), Tg(mpeg1:mCherry) transgenic zebrafish at 12 HPI post-needle-stick injury, showing relative gene expression for cyba2 and tyrk compared to unwounded fish at 4.5 DPI. N = 50 independent fish per condition per replicate, 3 replicates. Statistical significance is indicated, as determined by two-tailed t-test.

Data information: For all graphs, error bars indicate mean ± SD. Scale bars: (A) 100 μm; (C) 40 μm.
Figure 4.
Figure 4. Manipulation of wound inflammation and macrophage phenotypic state affects the extent of wound angiogenesis.

A. Schematic showing the factors used to skew macrophage phenotype towards either pro-inflammatory or anti-inflammatory states.

B. Representative confocal projection images of needle-stick injured zebrafish, taken from either 1 DPI Tg(mpeg:mCherry); Tg(tnfα:GFP) transgenic zebrafish (top panel) or 4 DPI Tg(fli:CFP) transgenic zebrafish (bottom panel), treated, as indicated, with DMSO (control), Ifn-γ or II-10. Boxed area denotes wound site.

C. Representative confocal projection images of needle-stick injured zebrafish taken from either 1 DPI Tg(mpeg:mCherry); Tg(tnfα:GFP) transgenic fish (top panel; middle panel for GFP only) or 4 DPI Tg(fli:CFP) transgenic fish (bottom panel), csf1ra−/− mutant and treated with Ifn-γ. Boxed area denotes wound site.

D. Quantification of number of macrophages at the wound site between 1 and 4 DPI, measured from images represented in (B and C). N = 14 (from B) and N = 16 (from C) independent fish per timepoint per condition.

E. Quantification of proportion of macrophages that are tnfα positive at the wound site from 1 to 4 DPI, measured from images represented in (B and C). N = 14 (from B) and N = 16 (from C) independent fish per timepoint per condition.

F. Quantification of total blood vessel length at 4 DPI, measured from images represented in (B and C), using Angioanalyser. N = 14 (from B) and N = 16 (from C) independent fish per timepoint per condition. Statistical significance, as determined by one-way ANOVA, is P ≤ 0.0001. Subsequent Bonferroni multiple comparison test, determines level of significance, as indicated. Significance values: *P ≤ 0.05, ***P ≤ 0.0001.

G. Representative confocal projection images taken from laser wounded Tg(fli:GFP), Tg(mpeg:mCherry); Tg(tnfα:GFP) triple transgenic zebrafish, imaged 30-930 DPI. Boxed area denotes wound site. tnfα-positive, inflammatory macrophages (arrows) associate with damaged, sprouting blood vessels earlier and in larger numbers than tnfα-negative, non-inflammatory macrophages (arrowheads).

H. Quantification of numbers of inflammatory versus non-inflammatory macrophages associated with damaged, sprouting blood vessels, measured from images represented in (C). N = 8 independent fish. Statistical significance is indicated, as determined by two-tailed t-test.

I. Graphical representation of common cellular interaction events and their time course, measured from movies represented in (G) and compared to events observed in Fig 3C. N = 8 independent fish.

Data information: For all graphs, error bars indicate mean ± SD. Scale bars: (B, C) 100 μm; (G) 40 μm.

known to have deficiencies in raising an appropriate immune response to bacterial infection (Pagan et al., 2015). While this mutant line exhibited macrophage recruitment to the wound site, it displayed a deficiency in its pro-inflammatory response, with considerably reduced macrophage tnfα expression (Figs 4C–E, and EV4A–C). We further observed that wound angiogenesis was inhibited in these pro-inflammation deficient fish (Figs 4C and F, and EV4A and D). Treating csf1ra mutants with immunostimulants LPS or Ifn-γ throughout the wound healing process rescued both the wound inflammation tnfα response as well as the wound angiogenesis deficit (Figs 4C and F, and EV4A and D). Taken together, these data indicate that wound inflammation status in general, and the switching of macrophages to a pro-inflammatory phenotype in particular, is critical in promoting wound angiogenesis during the early stages of wound repair.

Finally, laser ablation of single vessels in Tg(fli:GFP); Tg(tnfα:GFP); Tg(mpeg:mCherry) triple transgenic fish revealed that tnfα-positive macrophages interacted earlier and more frequently with laser damaged blood vessel tips than did tnfα-negative, non-inflammatory macrophages (Fig 4G–I, Movie EV8). Complementary movies of vessel repair following injuries performed on Tg(kdr:mCherry-CAAX); Tg(tnfα:GFP); Tg(mpeg:mCherry) triple transgenic fish further demonstrated the rapid responsiveness of tnfα-positive macrophages to damaged vessel tips (Fig EV4F, Movie EV9). Additionally, observing multiple such movies revealed that some of these early responding tnfα-positive macrophages exhibited reduced GFP over time in the vicinity of others that remained bright, thus excluding photobleaching as an explanation; this suggests that tnfα expression by macrophages is transient and that at least some cells are switching from tnfα positive to tnfα negative during the repair period (Fig EV4F, Movie EV9). In the context of the overall vessel repair response, the presence of these tnfα-positive inflammatory macrophages temporarily coincides with the early sprouting events, whereas non-inflammatory macrophages dominate later, as vessels undergo anastomosis (Fig 4G–I). These observations suggested that different macrophage phenotypes might be regulating different aspects of the wound angiogenesis process.

Inflammatory and non-inflammatory macrophages play different roles in driving wound angiogenesis

The concept of a pro-inflammatory environment potentially acting as a trigger for vessel sprouting has been previously suggested (Sainson et al., 2008). To examine the cellular interactions underlying this observation more closely and to better understand what role they may be playing in driving neoangiogenesis, we established an in vitro co-culture assay using human umbilical vein endothelial cells (HUVECs), which when cultured together with feeder human fibroblasts spontaneously generated vessels in culture (Bishop et al., 1999; Richards & Mellor, 2016). To this assay, we added primary human macrophages isolated from peripheral blood mononuclear cells that had been treated with cytokines to direct them towards pro-inflammatory (interferon γ, IFN-γ) or anti-inflammatory (interleukin-4, IL-4) activation states (Fig 5A). We confirmed that macrophages had indeed reached differential activation states by testing for levels of gene expression of well-established pro-inflammatory (tumour necrosis factor-α, TNF-α) or anti-inflammatory (mannose receptor C-type 1, MR1C1) genes (Roszer, 2015; Fig EV5A). To complement these co-culture cell-based approaches, we also exposed HUVECs to conditioned media extracted from the in vitro macrophage cultures, to dissect any differential contributions of cell-to-cell contact versus secreted factors produced by macrophages in these altered states (Fig 5A). The resultant macrophage–endothelial cell interactions were imaged and quantified after 4 days of co-culture. We observed three different types of macrophage–vessel interactions in vitro—macrophages can interact with blood vessel tips, or localise to the sides of blood vessels, or to vessel intersections and anastomoses. Intriguingly, we observed that pro-inflammatory macrophages were far more likely to associate with vessel tips than any other part of the vessel, whereas non-inflammatory cells (treated with IL-4) associated randomly with different areas of vessels (Fig 5B and C).

We next quantified the extent of vessel sprouting in these in vitro conditions. Our results showed that, just as in vivo in the zebrafish, the presence of pro-inflammatory macrophages (or their conditioned media) led to a significant increase in the amount and complexity of
Figure 5.
blood vessels in the co-culture (Figs 5D and E, and EV5B). By contrast, the presence of non-inflammatory macrophages (or their conditioned media) trended to an overall decrease in resultant blood vessels (Figs 5D and E, and EV5B). Similar increases and decreases in terms of vessel sprouting were observed when using LPS as a pro-inflammatory mediator, or dexamethasone as an anti-inflammatory mediator, respectively (unpublished observations). Quantitative PCR on macrophages cultured in immunostimulating conditions revealed high levels of VEGFA, suggesting that inflammatory macrophages might exert their function as angiogenic stimulators by providing a source of vascular growth factor (Fig 5F), complementing previous studies describing similar macrophage-derived VEGF as a driver for angiogenesis in mice (Xiong et al., 1998; Ramanathan et al., 2003; Cattin et al., 2015). Indeed, addition of VEGFA blocking antibody to cultures with pro-inflammatory macrophages reduced the amount of vessels formed to levels observed in control macrophage co-cultures (Fig 5G and H). These results suggest that pro-inflammatory macrophages play a specific role in initiating and driving vessel sprouting, at least in part, via release of VEGFA.

Since our earlier in vivo studies suggested that another pro-angiogenic role of macrophages might be to dislodge inhibitory neutrophils from vessel tips, we sought to examine this neutrophil anti-angiogenic activity using our co-culture in vitro assay. We isolated primary human neutrophils and cultured them, with or without pro-inflammatory interferon-α (IFN-α; Pylaeva et al., 2016; Shaul et al., 2016), together with HUVECs. Quantifying the extent of in vitro vessel sprouting indicated that neutrophils suppressed vessel growth, with IFN-α-treated neutrophils significantly more inhibitory (Fig 5I and J). These results suggest that neutrophils can indeed inhibit angiogenic influences and thus may require dislodging at sites of in vivo angiogenesis.

To investigate further whether tnfa-positive, pro-inflammatory macrophages were indeed delivering pro-angiogenic cues at the wound site, we FAC-sorted tnfa-positive versus tnfa-negative macrophages from zebrafish wounds and performed quantitative PCR for a range of angiogenic genes. Our results confirmed that tnfa-positive macrophages express higher levels of vegfaa (Fig 6A). Quantitative PCR performed on whole zebrafish wounds in the context of macrophage ablation showed greatly reduced levels of vegfaa expression, suggesting that wound macrophages are indeed necessary for vegfaa expression at the wound site (Fig 6B). These observations were supported by whole-mount in situ hybridisation (WISH) for vegfaa performed on wounded Tg(tnfaa:GFPl); Tg(mpeg:mCherry) transgenic fish, which showed a marked increase in vegfaa levels at the wound site, particularly by tnfa-positive macrophages (Fig 6C). Furthermore, performing WISH for vegfaa on metronidazole-treated Tg(mpeg:Gal4FF); Tg(UAS:In5B-mCherry) transgenic fish revealed a decrease in vegfaa staining at wounds where macrophages were absent (Fig 6D).

To genetically test the importance of vegfaa signalling, we utilised the vegfaa+mcdn2 mutant zebrafish, which has severe developmental vascular defects but can be rescued via vegfaa RNA injection (Rossi et al., 2016). Injuring vegfaa null “rescued” larvae, indicated that, while they were able to raise a typical immune response to wounding, they failed to undergo wound angiogenesis (Fig 6D and E), suggesting that vegfaa is indeed critically important for vessel repair. To more precisely control the timing of vegf signal blockage, we treated needle-stick injured Tg(fi:GFPl); two vegfr2 inhibitors S KlB1002 and SU5416 which have previously been shown to inhibit developmental sprouting angiogenesis in zebrafish (Serbedzija et al., 1999; Zhang et al., 2011); treating larvae with either of these inhibitors completely blocked wound angiogenesis compared to vehicle-treated wounded controls (Figs 6F and G, and EV6A and B). Live imaging of laser ablations of entire vessels in Tg (fi:GFPl); Tg(mpeg:mCherry) transgenic fish treated with SKLB1002 showed that while macrophages were still attracted to damaged...
Figure 6.
Macrophages drive wound angiogenesis  
David B Gurevich et al

At later stages of wound repair, blood vessels at wounds are pruned back to uninjured levels because the reduced metabolic demand in the repaired tissues no longer necessitates such an extensive vascular network. Since our later stage murine wounds indicate that the repaired tissues no longer necessitates such an extensive vascular network, we investigated whether the reduced metabolic demand in the damaged tissue was responsible for vessel pruning. At later stages of wound repair, blood vessels at wounds are pruned to uninjured levels because the reduced metabolic demand in the repaired tissues no longer necessitates such an extensive vascular network. Since our later stage murine wounds indicate that the repaired tissues no longer necessitates such an extensive vascular network, we investigated whether the reduced metabolic demand in the damaged tissue was responsible for vessel pruning.

Blood vessel resolution at the wound site is dependent on macrophage presence and phenotypic state

At later stages of wound repair, blood vessels at wounds are pruned back to uninjured levels because the reduced metabolic demand in the repaired tissues no longer necessitates such an extensive vascular network. Since our later stage murine wounds indicate that the repaired tissues no longer necessitates such an extensive vascular network, we investigated whether the reduced metabolic demand in the damaged tissue was responsible for vessel pruning. At later stages of wound repair, blood vessels at wounds are pruned to uninjured levels because the reduced metabolic demand in the repaired tissues no longer necessitates such an extensive vascular network. Since our later stage murine wounds indicate that the repaired tissues no longer necessitates such an extensive vascular network, we investigated whether the reduced metabolic demand in the damaged tissue was responsible for vessel pruning.

Figure 6. Zebrafish wound angiogenesis is driven by wound inflammatory macrophage-mediated vegf signalling.

A Gene expression for FAC-sorted zebrafish wound macrophages from Tg(nfs/C6:GFP); Tg(mpeg:mCherry) zebrafish, extracted at 1 DPI following needle-stick injury, trsfα-positive inflammatory macrophages express higher levels of vegfas than trsfα-negative non-inflammatory macrophages. N = 50 independent fish per condition, 3 replicates. Statistical significance is indicated, as determined by two-tailed t-test.

B Gene expression for whole-dissected zebrafish needle-stick wounded from Tg(mpeg:KalTA4); Tg(UAS:nfsb-mCherry) zebrafish following treatment with metronidazole, versus control DMSO-treated wounded fish, at 2 DPI and 4 DPI. Macrophage absence from wounds results in an overall decrease in vegfas levels at the wound site. N = 30 independent fish per condition, 3 replicates. Statistical significance is indicated, as determined by two-tailed t-test.

C Representative confocal projection images taken from needle-stick wounded Tg(mpeg:KalTA4); Tg(UAS:nfsb-mCherry); Tg(nfs/C6:GFP) triple transgenic zebrafish, DMSO control or metronidazole treated indicated, upon which vegfas WISH was performed. Inset image shows high levels of vegfas expression (blue) in trsfα-positive macrophages (yellow, asterisk) versus low levels in trsfα-negative macrophages (red, arrowhead). Boxed area denotes wound site. N = 12 independent fish per condition.

D Schematic and representative confocal images showing rescue experiment and subsequent needle-stick injury performed on vegfas homozygous mutant zebrafish or siblings, transgenic for Tg(elt2:GFP). Embryos derived from an incross of vegfas-/- adults were injected with “rescue” mRNA encoding for vegfas-165 at the one-cell stage. At 4 DPI, these mutant fish had an impaired but functional complement of ISVs. Needle-stick injuries performed on these mutants and their siblings at 4 DPI indicated a failure of wound angiogenesis by 4 DPI, despite a relatively normal inflammatory response. Boxed area denotes wound site.

E Quantification of total blood vessel length at 4 DPI, measured from images represented in (D), using Angioanalyser. N = 12 independent fish per condition. Statistical significance is indicated, as determined by two-tailed t-test.

F Representative confocal projection image taken from needle-stick wounded Tg(elt2:GFP) transgenic zebrafish, imaged 4 DPI and treated with vegfr2 inhibitor SKLB4002 from the moment of injury. Boxed area denotes wound site.

G Quantification of total blood vessel length, measured from images represented in (F) using Angioanalyser. N = 14 independent fish. Statistical significance is indicated, as determined by two-tailed t-test.

H Representative confocal projection images taken from laser wounded Tg(elt2:GFP); Tg(mpeg:mCherry) double-transgenic zebrafish, imaged 30-90 MPFI and treated with SKLB4002 from moment of injury. Macrophages still associate with damaged vessels (arrowheads), but these vessels do not sprout appropriately and fail to repair. Boxed area denotes wound site.

I Graphical representation of common cellular interaction events and their time course, measured from movies represented in (H) and compared to events observed in Fig 3C. N = 8 independent fish.

Data information: For all graphs, error bars indicate mean ± SD. Scale bars: (C, D, F) 100 μm; (H) 40 μm.
altering the total number of macrophages at this timepoint (Fig 7F–H). The same treatment regime in Tg(flrl1:GFP) transgenic fish leads to a clear failure in vessel regression at the wound site (Fig 7F and I). Finally, we performed quantitative PCR for vegfa on FAC-sorted tnf-a-positive versus tnf-a-negative macrophages from these later stage (7 DPI) zebrafish wounds. Our results confirm that the inflammatory macrophages express higher levels of vegfa than non-inflammatory macrophages at these later wounds, a scenario similar to that observed in early vessel repair (Fig 7J). Taken together, these results demonstrate that appropriate vessel resolution is dependent not only on the presence of macrophages, but that these macrophages must be of a particular phenotype that no longer expresses pro-angiogenic signals.

**Discussion**

In this study, we describe the process of wound angiogenesis and its resolution in mouse and zebrafish and investigate how the innate immune cell influx into the wound impacts on these neangiogenesis events. We show that neutrophils are only transiently drawn to the tips of damaged vessels whereas macrophages exhibit long-term, intimate associations throughout the duration of vessel sprouting and anastomosis; at the later resolution stages, these wound macrophages contain phagocytosed vessel material. Macrophages are critically important for neangiogenesis in wound repair, and ablating macrophages at either early or late stages of wound healing disrupts vessel sprouting and vessel regression. The early influx of macrophages is largely pro-inflammatory and express tnf-a, but the proportion of these macrophages diminishes over time. By modulating this phenotypic switch with exogenous chemokines or genetically, we can skew macrophages towards a pro- or anti-inflammatory state at early or late stages in the zebrafish wound repair process; these in vivo experiments suggest that pro-inflammatory macrophages express vegfa and are required for early vessel sprouting, but that tnf-a must be switched off for macrophages to effectively participate in vessel clearance. Our in vitro co-culture experiments confirm these interactions and reveal that VEGFA expression is a key component of the pro-sprouting capacity of inflammatory macrophages that preferentially associate with sprouting tips.

A significant and unique advantage of performing our live-imaging studies in zebrafish larvae is the ease of imaging in translucent tissues. Unlike the opaque tissues of mouse that require complicated and invasive procedures to image, we can visualise blood vessels prior to injury induction and can therefore selectively injure areas of living zebrafish larval tissue with or without damage to adjacent vessels. This has enabled us to observe that wound angiogenesis occurs in response to tissue damage regardless of whether there is damage to existing vessels. Another outcome of our live-imaging studies is that wound neoangiogenesis appears to largely recapitulate earlier episodes of embryonic developmental angiogenesis. The resultant intimate macrophage–vessel association shown in our wounding models appears to mimic certain elements of what has previously been observed in developmental contexts such as retinal (Rymo et al., 2011) and hindbrain angiogenesis in embryonic mice (Fantin et al., 2010), and in the connection of intersomitic vessels in the embryonic zebrafish trunk (Fantin et al., 2010). Indeed, the dynamic nature of filopodial formation and vessel sprouting during repair of larval zebrafish wounds appears to replicate that observed during normal developmental angiogenesis (Isogai et al., 2003).

A recent study in which zebrafish vessels were damaged by hypoxia also reports that the responding macrophages are largely tnf-a positive (Gerri et al., 2017). Our live-imaging studies in fish combined with in vitro studies go beyond this and demonstrate that
this inflammatory phenotype is needed for sprouting in wounds and that the pro-inflammatory macrophage-mediated sprouting is facilitated by vegf. Clearly, the VEGF signal does not have to be locally delivered because macrophage-conditioned medium added to our in vitro co-culture vessel assay triggers increased vessel sprouting and outgrowth. However, macrophages, particularly inflammatory ones, associate with vessel tips in both human co-cultures and fish wounds, suggesting that macrophages will tend to act as point sources of vegf signalling. Similar, vegf-based macrophage-endothelial cell guidance has previously been suggested in the context of mediating repair of peripheral nerves by bridging the gap between damaged nerve ends (Cattin et al, 2015); our studies reveal that this mechanism is part of the inflammatory response to injury which must be tightly controlled in order to establish fully functional, patent vessels in a variety of wound repair scenarios.

During the later resolution stages, we observe that macrophages contain phagocytosed vessel material. Furthermore, ablation of macrophages during later phases of repair in fish wounds results in a failure to appropriately remodel the vasculature, and this failure is reproduced by global inhibition of apoptosis. The dynamics of vessel pruning and regression in our wounds also appears to be very similar to that observed in zebrafish retinas during embryo development, with apoptotic endothelial cells observed almost exclusively in thin, unicellular tubes as blood vessels progress from multicellular lumenised structures to the point of detachment (Kochhan et al, 2013). Whether wound macrophages simply clear away this vessel death, or are actively driving vessel remodelling by directing endothelial cell death as previously reported in hyaloid vessel regression in mouse retina (Lang & Bishop, 1993; Lobov et al, 2005; Rao et al, 2007), is an area of active investigation. We observe that appropriate vessel regression fails if pro-inflammatory signals persist or are re-stimulated at later stages of repair, and we show that one of the mechanisms preventing this regression is continued vegf expression by inflammatory macrophages. This experimental scenario may reflect pathologies in which the dysregulation of wound angiogenesis is associated with an altered inflammatory profile, such as atherosclerosis or chronic wounds (Frykberg & Banks, 2015; Malecic & Young, 2017). Understanding how and when to nudge the innate immune wound response towards a pro-inflammatory state to stimulate sprouting and then away from this state in a timely fashion to allow for vessel regression is therefore critical in devising new therapies to deal with a broad range of pathologies.

It is interesting that during normal wound repair we see neutrophils displaying very transient associations with vessel tips, in contrast to the much longer macrophage–vessel interactions. These wound neutrophils express soluble vegf receptor 1 (sflt1), which means they may act as an angiogenic brake during the early period following wounding; indeed, when we delete macrophages, we see that neutrophils persist at these sites for considerably longer than when macrophages are present, and, as a consequence, wound neangiogenesis fails. This presence of anti-angiogenic neutrophils could explain previous observations of increased and prolonged neutrophil numbers in human chronic wounds, which fail to heal in part due to lack of appropriate revascularisation (Martin & Nunan, 2015). Furthermore, early wound macrophages express genes involved in mediating neutrophil resolution from wounds, a function that has previously been demonstrated in other wound contexts (Tauzin et al, 2014; de Oliveira et al, 2016). These results indicate that as well as their direct roles in regulating wound angiogenesis, macrophages play an indirect role in controlling when the brakes to angiogenesis are removed by inducing anti-angiogenic neutrophils to depart the repairing wound. How neutrophils and macrophages identify damaged vessel tips that they subsequently associate with, and whether macrophages play further roles in regulating the behaviours of other important cells in the repair process that may influence endothelial cells—such as fibroblasts and pericytes—remain open questions.

In larger wounds, we see numerous macrophages associating with damaged vessels and driving vessel sprouting, whereas in smaller vessel laser lesions we observe individual macrophages straddling the gap between two vessel tips, just as has also been shown in a recent zebrafish stroke model of brain vessels microlesions (Liu et al, 2016). We presume that macrophages facilitate the repair of vessels in larger injuries in a stepwise manner, firstly encouraging sprouting angiogenesis and extension of vessel tips via vegf signalling until they reach close proximity with one another and then, just as in repair of microlesions, support ligation and anastomosis at sites of vessel fusion. Furthermore, we observe tnfα expression by the majority of early wound responding macrophages associated with spraying tips, while later macrophages appear to downregulate tnfα expression during the anastomosis and vessel maturation process. Indeed, some of our timelapse experiments directly demonstrate tnfα-positive macrophages switching to a tnfα-negative phenotype, indicating a pro- to anti-inflammatory switch in macrophage phenotype previously observed only indirectly in a variety of mouse wound contexts (Arnold et al, 2007; Ramachandran et al, 2012). These in vivo data, along with our in vitro results showing that anti-inflammatory IL-4-treated macrophages lead to fewer blood vessels, suggest that differential action in macrophage function may be driven by macrophage phenotype switching. Defining precisely what mechanisms are utilised by macrophages to facilitate vessel sprouting, stabilisation and “nurturing,” in addition to delivery of angiogenic factors, warrants further study. Moreover, whether the phenotype switching that we observe is normally triggered by exogenous factors or is a passive time-limited occurrence, and whether the manipulation of these phenotypic states will yield therapeutic benefits are all important questions for further investigation.

Materials and Methods

Mouse housing and excisional wounding

All experiments were conducted with approval from the local ethical review committee at the University of Bristol and in accordance with the UK Home Office regulations (Guidance on the Operation of Animals, Scientific Procedures Act, 1986). Male wild-type C57Bl/J6J mice (7–10 weeks old) were purchased from Charles River and kept under standard pathogen-free conditions with food (regular chow diet) and water ad libitum and 12:12-h light–dark cycle. For skin wounding experiments, mice were anaesthetised with isoflurane and four full-thickness excisional wounds made to the shaved dorsal skin (4-mm biopsy punch, Kai Industries). Wounds and 0.5 cm surrounding skin were harvested on days 0, 1, 3, 7, 10 and 14 after wounding and either prepared for cryosectioning or fixed overnight.
in 4% paraformaldehyde at 4°C overnight on a rocker for whole-mount multiphoton imaging.

Tissue preparation for multiphoton imaging

After fixation in 4% PFA, wounds were washed with PBS. Blocking and permeabilising was achieved with 1% Triton X-100, 1% BSA, 3% normal goat serum in PBS at room temperature overnight. Tissues were then exposed to antibodies diluted in block/perm solution incubated for 48–72 h at 4°C on a rocker. Thereafter, tissues were mounted in Prolong Diamond (Life Technology) for multiphoton imaging. For frozen sections, mouse wounds were fresh snap-frozen in optimal cutting temperature (OCT) compound (Tissue-Tek) on liquid nitrogen-cooled isotopentane and sectioned at 50 μm.

Multiphoton imaging

Two-photon imaging with z-compensation was performed to detect the fluorophore staining. Blood vessel endothelial cells were labelled with Alexa Fluor 594-conjugated rat anti-mouse CD31 (1 μg/ml, clone ME13.3; Biolegend) (excited @ 760 nm) or Alexa Fluor 647-conjugated rat anti-mouse CD144 (2.5 μg/ml, clone BV13, Biolegend), and macrophages were labelled with Alexa Fluor 488-conjugated anti-mouse CD68 (2.5 μg/ml, clone FA-11; Biolegend—excited @ 710 nm). Multiphoton images were generated on a Leica SP8 MP/CLSM system (Leica Microsystems).

Transmission electron microscopy

Wounds were harvested at the indicated timepoints, fixed and processed as previously described (Nunan et al., 2015). Ultrathin (0.02 μm) sections were images on a Tecnai 12-FEI 120 kV BioTwin Spirit transmission electron microscope.

Zebrafish strains and maintenance

All experiments were conducted with approval from the local ethical review committee at the University of Bristol and in accordance with the UK Home Office regulations (Guidance on the Operation of Animals, Scientific Procedures Act, 1986). Wild-type and transgenic lines Tg(mpeg1:GFP) [referred to as Tg(mpeg:GFP)] (Lawson & Weinstein, 2002), Tg(mpx:GFP)(Renshaw et al., 2006), Tg(mpeg1:mCherry) [referred to as Tg(mpeg:mCherry)] (Ellett et al., 2011), Tg(mpeg1.1:Gal4FF) [referred to as Tg(mpeg:Gal4)] (Palha et al., 2013), Tg(UAS-E1b:NfsB-mCherry) [referred to as Tg(UAS:NfsB-mCherry)] (Davison et al., 2007), TgBAC(tnf:GFP) [referred to as Tg(tnf:GFP)] (Marjoram et al., 2015), Tg(dll4:EGFP) (Sacilotto et al., 2013), Tg(kdr:mCherry-CAAX) (Fujita et al., 2011), TgBAC(etv2:EGFP) [referred to as Tg(etv2:GFP)] (Proulx et al., 2010), and Tg(tbp:Gal4) together with Tg(UAS-SEC-Hsa.ANXA5-YPF,myl7:RFp) [referred to as Tg(seeA5:YPF)] (van Ham et al., 2010) were maintained on TL wild-type background, and staging and husbandry were performed as previously described (Westerfield, 1995). Mutant strains used were csf1ra+/- (Parichy et al., 2000) and vegfaa+/- (Rossi et al., 2016), maintained on AB background or used in combination with transgenic lines as indicated. csf1ra+/- mutants were genotyped by visual inspection for absence of mature xanthophores as previously described (Parichy et al., 2000). vegfaa+/- mutants were genotyped as previously described (Rossi et al., 2016), using primers 5’-CGAGAGCTGCGTGAGACATC-3’ and 5’-GAGTGTAG TGTGCTCGATCT-3’. Metronidazole cell ablation was performed as previously described (Rossi et al., 2016), using primers 5’-CAACGGTAGCTGCGTGAGACATC-3’ and 5’-GAGTGTAGTACG TGTGCTCGATCT-3’.

Needle-stick injury

Needle-stick wound induction was performed into the dorsal somites opposite the cloaca with either a 30-gauge needle (Becton Dickinson) or fine tungsten needle (Harvard Apparatus), as previously described (Gurevich et al., 2016). Subsequently, fish were immediately transferred to fresh embryo water (E3) to recover. Sham wounded fish were used as an uninjured control. Sham and wounded fish were fed as normal from 6 days post-fertilisation, and fish were rejected from analysis for experiments dealing with vessel regression if they had not grown to at least 5 mm in length by 10 DPI, as this was taken as a sign of insufficient feeding. Fluorescent stereomicroscope images were generated on a Leica M205 FA system (Leica Microsystems). Confocal images were generated on a Leica SP8 MP/CLSM system (Leica Microsystems).

Laser injury

Laser-induced injury was performed as previously described (Gurevich et al., 2016), with some modifications. Briefly, anaesthetised fish were mounted directly into a glass-bottomed 35-mm dish (Mattek). A Micropoint laser (Spectra Physics) connected to a Zeiss Axioiplan II microscope (Zeiss Microimaging) was used for injuring as previously described (Otten & Abdelilah-Seyfried, 2013), using a 40× water immersion objective and a laser pulse at a wavelength of 435 nm for cell ablation. Time lapse recordings were generated on a Leica SP8 MP/CLSM system (Leica Microsystems).

Fluorescent angiography

Zebrafish fluorescent angiographies were performed as previously described (van Rooijen et al., 2010), with some modifications. Briefly, high molecular weight rhodamine–dextran conjugate (2,000,000 MW, Sigma) was injected into the caudal vein of fish prior to imaging at the timepoints indicated.

Macrophage ablation experiments

Metronidazole cell ablation was performed as previously described (Pisharath & Parsons, 2009; Gurevich et al., 2016), with some modifications. Briefly, the pro-drug metronidazole (Sigma) was dissolved in E3 water as a 10 mM or 2.5 mM working solution, as indicated, along with added 0.2% dimethyl sulfoxide (DMSO). Transgenic zebrafish were incubated at 28.5°C in 10 mM solution from 1.5 DPI to 4 DPI and 2.5 mM immediately after injury (4 DPI) as indicated, with solutions changed every day. Control fish were subjected to the same injury and treatment procedure without metronidazole.

Clodronate liposome (Clophosome A, FormuMax) cell ablation was performed as previously described (Bernut et al., 2014; Carrillo et al., 2016), with 5 nl of either Clodronate or PBS containing liposomes injected into the caudal vein of 3 DPI fish prior to injury at 4 DPI (10-nl injections used for 6 DPI fish).
Drug and recombinant protein treatments

Fish were treated with compounds as described. 0.2% DMSO was used for all treatments as well as vehicle control. Lipopolysaccharide (referred to as LPS) (Sigma-Aldrich) was dissolved in PBS to 5 mg/ml as a stock solution and used as a working solution of 100 μg/ml diluted in E3 water. Hydrocortisone (Sigma-Aldrich) was dissolved in DMSO as a 500 mM stock solution and used as a working solution of 100 μM diluted in E3 water. Recombinant zebrafish Ifn-γ, IL-10 and IL-4 (Kingfisher Biotech) were all dissolved and maintained as a stock solution and used at working concentrations as indicated, diluted in E3 water. SKLB1002 and SU5416 Vegfr inhibitors (both Sigma-Aldrich) were dissolved in DMSO and used at working concentrations of 2.5 and 5 μM, respectively. Q-VD-OPH pan-caspase inhibitor (referred to as QVD) (Sigma-Aldrich) was dissolved in PBS to 100 mM as a stock solution and used as a working concentration of 100 μM. Fish were treated in all reagents at 28.5°C at the timepoints indicated with solutions changed every day. Control fish were subjected to the same injury and treatment procedure without compound exposure.

Whole-mount in situ hybridisation

In situ hybridisation was performed using a commercial vegfaa probe (Dr-vegfaa; catalog no. 543241) and RNAscope® technology (ACD, USA) on 4% PFA fixed larvae following manufacturers protocols.

Peripheral blood apheresis cones

Peripheral blood apheresis cones were acquired for research use in accordance with the Declaration of Helsinki and approval from the Bristol Research Ethics Committee (REC 12/SW/0199). Blood from apheresis cones was diluted 1:1 with Hanks balanced salt solution (HBSS, Sigma) containing 0.6% acid citrate dextrose (ACD) and layered onto Histopaque (Sigma, USA) on 4% PFA fixed larvae following manufacturers protocols. Blood was allowed to layer for 1.077 g/ml density gradient leaving only the red cell and neutrophil pellet at the bottom. The red cells were lysed in ice-cold lysis buffer (155 mM NH4Cl, 0.137 mM EDTA, 1 mM KHCO3, pH 7.5), on ice for 10 min. Centrifugation steps were carried out at 400 g at 4°C; lysis and centrifugation steps were repeated as necessary. Neutrophils were washed twice in cold PBS + 0.5% human serum albumin (HSA) before being resuspended in incubation media (RPMI + 20% FBS + 1% pen/strep), with or without 20 ng/ml Interferon α (IFN-α, Biogen).

Macrophase culture

Human peripheral blood mononuclear cells (PBMCs) were isolated as previously described using the interphase layer of the density gradient (Ramos et al, 2013). CD14⁺ cells were isolated from PBMCs using a magnetic bead CD14⁺ kit (Miltenyi Biotec), LS columns (Miltenyi Biotec) and MidiMACSTM separator (Miltenyi Biotec) as per the manufacturer’s instructions. CD14⁺ cells were frozen in 50% foetal calf serum (FCS, Gibco), 40% PBS with 10% DMSO (Sigma) and stored in liquid nitrogen until required. Thawed cells were resuspended at a density of 0.33 × 10⁶/ml in RPMI 1640 (Gibco) supplemented with 10% FBS (Gibco), 10 ng/ml macrophage-colony-stimulating factor (wt/vol, M-CSF, Miltenyi Biotec), and penicillin/streptomycin at 100 U/0.1 mg per ml of media, respectively (wt/vol; Sigma); with or without the inclusion of interleukin-4 at 20 ng/ml (wt/vol, IL-4, Biogen), interferon-γ at 2.5 ng/ml (wt/vol, IFN-γ, Biogen), for directed differentiation into pro- and anti-inflammatory macrophages. Cells were incubated at 37°C with 5% CO₂. Full media changes were performed twice throughout the 7-day culture where any cells in suspension were collection via centrifugation at 300 g and replaced in the culture. Cells were harvested from the adherent macrophage culture using a detaching buffer (10 mM EDTA, 15 mM Lidocaine in PBS) for inclusion in the co-culture assay.

HVECs co-culture

Normal human adult-derived fibroblasts (NDHF, Lonza CC-2511) were cultured in 12-well plates in HVECs medium (EBM-2, Lonza), seeded at 3 × 10⁴ cells/ml onto glass coverslips triple coated with 50 μg/ml fibronectin (F0895, Sigma-Aldrich), 30 μg/ml type I collagen (5409, Advanced BioMatrix) and 0.1% gelatin (G1393, Sigma-Aldrich) before being incubated in the bottom. The red cells were lysed in ice-cold lysis buffer (155 mM NH4Cl, 0.137 mM EDTA, 1 mM KHCO3, pH 7.5), on ice for 10 min. Centrifugation steps were carried out at 400 g at 4°C; lysis and centrifugation steps were repeated as necessary. Neutrophils were washed twice in cold PBS + 0.5% human serum albumin (HSA) before being resuspended in incubation media (RPMI + 20% FBS + 1% pen/strep), with or without 20 ng/ml Interferon α (IFN-α, Biogen).

Table 1. Primers used for QPCR and genotyping.

| Gene name               | Forward primer 5’–3’ | Reverse primer 5’–3’ |
|-------------------------|----------------------|----------------------|
| e1a (danio)             | CTCTCAAGGCTGACTGTCG  | CGGCTAGTACCCTCCC     |
| soluble flt1 (danio)    | GCAAGACGAGCCAAACAGA  | TTGGCGTCTGGGTAGTGC   |
| membrane flt1 (danio)   | AGCCCTGGAGAATCAAGGC  | GCTCAAGAGAGCTGACTTT  |
| yrk (danio)             | CATCTCATGACTAGCGCTCT | CAGCTTCCAGACCTCCTCG  |
| cyba1 (danio)           | ATGCCCAAGATTGACTGCCG  | GCTCAGAGATGTTATGCT   |
| vegfaa (danio)          | AAAGACGTGGCTCAAACACC | AGACGTCCTACTGACCTTT  |
| vegfaa (danio, genotyping) | CGACAGCTGCTGGTAGACATC | GGATCTAGCTGCTCATATCT |
| EF1a (homo)             | TGCTGCTATGGAGCCTAG   | ACCGGCTAATTTTATCC    |
| TNFa (homo)             | CGCTCCCAAGAAGACAG    | AGAGCTGAAGAACCAACAC  |
| MRC1 (homo)             | TTAGAAGCTTCTTCTGAGTG  | TCTCCATAAGCCAGTTTTCA |
| VEGFA (homo)            | AGGCGAAGATCATCACGAGA  | AGGGTGCTGATTGAGTGC   |
Aldrich) in PBS, incubated for 4 days at 37°C/5% CO₂. Media was refreshed every 2 days throughout the assay. HUVECs (Lonza pooled human donors, CC-2519) were subsequently added to confluent fibroblast monolayer on day 5 at 3 × 10⁶ cells/ml, cultured in HUVEC medium (Lonza EBM-2), incubated for a further 3 days at 37°C/5% CO₂. At this point, macrophages at 1.5 × 10⁶, conditioned media or RPMI control media, were added to the co-culture assay, as indicated, and incubated for a further 3 days at 37°C/5% CO₂. Coverslips were subsequently fixed with 4% paraformaldehyde and immunofluorescently stained as previously described (Richards & Mellor, 2016), using mouse monoclonal anti-human CD31 antibody (1:200 of 0.5 mg/ml stock, BAA7, R&D Systems) to label endothelial cells, mouse monoclonal anti-human MCSFR antibody (1:200, ab183316, Abcam) to label macrophages, DAPI (D21490, Invitrogen) and goat anti-mouse secondary antibodies Alexa Fluor 488 (1:400, A11029, Invitrogen) and Alexa Fluor 594 (1:400, A11032, Invitrogen). Human VEGF₁₆₅ antibody (AF-293, R&D Systems, referred to as VEGF blocking antibody) was used as a neutralising antibody at a working concentration of 200 ng/ml, incubated from the moment of macrophage/conditioned media addition and replaced with fresh VEGF antibody at each media change.

Analysis of gene expression

For analysis of whole wounds, injured fish were anaesthetised, placed on a 3’ × 1’ glass slide in a drop of water, manoeuvred into a lateral lying position and wounds were dissected using a surgical scalpel. Uninjured fish were similarly processed, with control unwounded tissue collected from the dorsal somites opposite the cloaca. Collected tissue was processed and RNA extracted using a RNeasy Mini Plus Kit (Qiagen) as per manufacturer’s instructions. For analysis of sorted cells, wounds were harvested from transgenic zebrafish and dissociated to generate single-cell suspensions as previously described (Trinh et al., 2017). FACS analysis was performed using a Becton Dickinson InFlux (BD Biosciences San Jose, CA USA) and RNA extracted using a RNeasy Micro Kit (Qiagen) as per manufacturer’s instructions. For analysis of cultured macrophages, cells were harvested and RNA extracted using a RNeasy Mini Plus Kit (Qiagen) as per manufacturer’s instructions. cDNA was generated using a Maxima First Strand cDNA Synthesis Kit (Thermo Fisher) as per manufacturer’s instructions. Quantitative PCR was performed using a SYBR Green PCR kit (Qiagen) and primers for genes as indicated (see Table 1) in an Agilent MX3005P QPCR cycler (La Jolla). Gene expression data were normalised against Elongation factor 1-alpha. 1% agarose gels were run at 80 V and stained with ethidium bromide at 0.5 µg/ml.

Image analysis and statistics

All image analysis was performed in Velocity (Perkin Elmer) or ImageJ. Pixel counts were performed in ImageJ. Vessel orientation was measured using the ImageJ plugin OrientationJ, which rates the coherence of neighbouring pixels between 0 (isotropic, no overall direction) and 1 (one dominant direction; Fonck et al., 2009). Vessel metrics (such as total vessel length, node and sprout/branch numbers) were calculated by using the ImageJ macro, Angioanalyzer (Gholobova et al., 2015). All statistical analysis was performed using Graphpad Prism. Data were confirmed to be normally distributed via d’Agostino–Pearson test or Shapiro–Wilks test prior to further comparisons. Student’s t-test was used except in the case of comparisons involving more than two groups; in these instances, one-way ANOVA was performed for all comparisons, and a Bonferroni multiple comparison test was subsequently performed.

Expanded View for this article is available online.

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Author contributions

DBG designed and performed zebrafish wounding, treatment and imaging experiments and mouse wound electron microscopy imaging; DBG, CES, CT, AMT and HM designed and performed human tissue co-culture experiments; AG designed and performed quantitative PCR experiments; JC designed and performed mouse wounding and multiphoton imaging experiments; PM supervised the project. DBG and PM wrote the manuscript.

Conflict of interest

The authors declare that they have no conflict of interest.

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