Association between cagA negative *Helicobacter pylori* status and nonalcoholic fatty liver disease among adults in the United States

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Abstract

We investigated the relationship of *H. pylori* stratified by cytotoxin-associated gene A (cagA) status with nonalcoholic fatty liver disease (NAFLD) in the general population of the United States (US). We utilized the Third National Health and Nutrition Examination Survey from 1988 to 1994 in this study. NAFLD was defined by ultrasonographic detection of hepatic steatosis in the absence of other known causes of liver diseases and significant alcohol consumption. Hepatic steatosis was assessed by parenchymal brightness, liver to kidney contrast, deep beam attenuation, bright vessel walls and gallbladder wall definition. Antibodies to *H. pylori* and cagA of participants were measured using *H. pylori* IgG and anti-cagA IgG enzyme-linked immunosorbent assays. Among 5,404 participants, the prevalence of NAFLD was higher in *H. pylori* positive subjects (33.5±1.8%) compared to *H. pylori* negative subjects (26.1±1.7%, p<0.001). In terms of cagA protein status stratification, while cagA positive *H. pylori* group did not demonstrate an association with NAFLD (OR: 1.05; 95% CI: 0.81–1.37), cagA negative *H. pylori* group was noted to have a significant association with NAFLD in a multivariable analysis (OR: 1.30; 95% CI: 1.01–1.67). In conclusion, our study demonstrated that cagA negative *H. pylori* infection was an independent predictor of NAFLD in the US general population.

Introduction

Nonalcoholic fatty liver disease (NAFLD) is the most prevalent liver disease in the United States (US) and in many parts of the world [1,2]. NAFLD is a clinical and pathological entity that encompasses nonalcoholic fatty liver and nonalcoholic steatohepatitis complicated by progressive stages of fibrosis, cirrhosis, and hepatocellular carcinoma [3]. Because the mechanism underlying the development of NAFLD has been linked to insulin resistance and metabolic...
syndrome, NAFLD is associated with many risk factors of cardiovascular disease, such as obesity, diabetes, and dyslipidemia [4–7].

*Helicobacter pylori* (*H. pylori*) infection has been associated with a variety of extra-digestive conditions, including cardiovascular, lung, hematologic, neurologic and hepatobiliary diseases [8,9]. In the Finnish Alpha-Tocopherol, Beta-Carotene Cancer Prevention (ATBC) study, *H. pylori* seropositivity was associated with an increased risk of biliary tract cancer [10]. Bacterial infection of intestinal tract may compromise mucosal integrity, allowing passive or facilitated entry of harmful bacteria or their products into the circulation [11]. Notably, cytotoxin-associated gene A (cagA) positive strains can introduce *H. pylori* proteins into the epithelial cells and induce an antibody response to the cagA protein. These strains was found to be associated with an increased risk of gastric cancer and peptic ulcer disease and a decreased risk of esophageal reflux disease [12]. Until now, several small-scale epidemiological studies have reported a relationship between NAFLD and *H. pylori* infection [8,13]. However, association between NAFLD and *H. pylori* infection with regard to cagA status has not been studied in a large, general population.

Therefore, in this study, we used the National Health and Nutrition Examination Survey (NHANES) data to investigate the impact of *H. pylori* infection with regard to cagA status on NAFLD.

**Methods**

**Study population and study design**

This study represents an analysis of the third NHANES data (1988–1994, the National Center for Health Statistics, the Centers for Disease Control and Prevention [CDC]). NHANES employed a stratified, multistage, clustered probability sampling design to obtain a representative sample of the non-institutionalized civilian population in the US. In total, 7,275 adults from 20 to 74 years of age, who participated in the NHANES III survey Phase I, underwent laboratory tests at a mobile examination center (Fig 1). Subjects with significant alcohol consumption (>21 drinks/week in men and >14 drinks/week in women) [14], viral hepatitis (positive serum hepatitis B surface antigen and/or positive serum hepatitis C antibody), iron overload (transferrin saturation ≥ 50%), and pregnant women were excluded (n = 816). Among the remaining 6,459 participants, hepatic steatosis could be evaluated in 5,910 (91.5%). We further excluded participants whose tests did not examine *H. pylori* status. Finally, 5,404 subjects were included and analyzed in this study. The original NHANES survey was approved by the CDC’s Institutional Review Board, and all participants provided written informed consent to participate. This retrospective study was exempted by the Institutional Review Board of the Seoul National University Hospital because the dataset used in this analysis was completely deidentified. This study was conducted according to the Guideline for Good Clinical Practice (GCP) and the provisions of the Helsinki declaration.

**H. pylori status**

Antibodies to *H. pylori* were measured in participants 20 years and older using a *H. pylori* IgG enzyme-linked immunosorbent assay (ELISA) (Wampole Laboratories, Cranbury, New Jersey) in serum surplus samples [15]. Additionally, anti-cagA IgG was also measured in surplus sera among participants positive for *H. pylori*, using a method developed and standardized by Vanderbilt University, as described previously [16]. On the basis of the *H. pylori* and cagA status, we classified participants into three groups: *H. pylori* positive and cagA positive, *H. pylori* positive and cagA negative, and *H. pylori* negative [17].
Assessment of NAFLD

We used a previously described method to assess NAFLD [2]. Briefly, the original NHANES III survey included gallbladder ultrasonography as part of the assessment for digestive disease in adults aged 20 and above. Three board-certified radiologists reviewed the archived gallbladder ultrasonography images to assess fatty liver [18]. Assessment of fatty liver was performed using the following criteria: (1) parenchymal brightness; (2) liver to kidney contrast; (3) deep beam attenuation; (4) bright vessel walls; and (5) gallbladder wall definition. Overall assessment, made using an algorithm based on these five criteria, defined normal versus mild, moderate, or severe hepatic steatosis [18]. For our study, NAFLD was diagnosed as the presence of any degree of fatty liver (mild to severe).

Advanced fibrosis by noninvasive panels

The noninvasive panels used for the evaluation of advanced fibrosis have been previously described [2, 19]. Briefly, advanced fibrosis was assessed by NAFLD fibrosis score, FIB-4 score, and aspartate aminotransferase to platelet ratio index. Due to the small sample size of subjects with high probability of advanced fibrosis, ‘suspected advanced fibrosis’ was evaluated and defined as presence of high probability for advanced fibrosis on at least one of the three non-invasive fibrosis panels.

Clinical and laboratory evaluation

A vast array of demographic, lifestyle, dietary, anthropometric and comprehensive laboratory data were available in this NHANES dataset. Hypertension was defined as systolic blood pressure $\geq 140$ mmHg or diastolic blood pressure $\geq 90$ mmHg and/or previous use of antihypertensive medication. Diabetes mellitus was defined as having a history of diabetes and/or treatment with a hypoglycemic agent or insulin. Current smokers were participants who reported ongoing smoking or those who had smoked at least 100 cigarettes in the preceding 5 years.
Statistical analyses

The outcome variable in this study was the presence of NAFLD. Analyses were conducted using the SAS-callable SUDAAN 10.0.1 (Research Triangle Institute, Research Triangle Park, NC), which allows for the stratified sampling scheme by NHANES to project these data to the US population [20]. We analyzed proportions of categorical variables and means ± standard error of continuous variables (PROC CROSSTAB, PROC DESCRIPT). We used chi-squared tests for the categorical variables and Student's t-test and one-way analysis of variance (ANOVA) for the continuous variables (PROC CROSSTAB, PROC REGRESS). A multivariable logistic regression model was used to investigate the independent association between NAFLD and *H. pylori* status (PROC LOGISTIC). All subjects in NHANES III aged 17 years and over were followed for mortality through December 2006. We utilized Cox proportional hazards regression testing for survival analysis of overall mortality.

Results

Baseline characteristics according to NAFLD

The prevalence of NAFLD among the eligible subjects was 31.9%. Demographic and clinical characteristics of subjects with NAFLD are shown in Table 1. Subjects with NAFLD were more likely to be older, male, hypertensive, and diabetic than those without NAFLD. Similarly, body mass index (BMI), waist circumference, lipid panels, liver enzymes, and fasting glucose were higher in subjects with NAFLD than those without NAFLD.

Baseline characteristics according to *H. pylori* status

Table 2 shows the distribution of participants based on *H. pylori* status by demographic and clinical risk factors in the overall study population, including those with data on *H. pylori* cagA status. Subjects with positive *H. pylori* serology results were more likely to be older, male and with a higher BMI and waist circumference than that of *H. pylori*-negative participants. Subjects of a race/ethnicity other than non-Hispanic white and those who had a history of diabetes or hypertension were more likely to be *H. pylori*-positive. Similarly, lipid panels, liver enzymes, and fasting glucose were higher in subjects with positive *H. pylori* serology than without. There was no apparent association of *H. pylori* positivity with current smoking status. Among subjects with positive *H. pylori* serology, those without cagA were likely to be older with higher prevalence of hypertensive versus those with cagA. Subjects of a race/ethnicity other than non-Hispanic white were more likely to be cagA positive.

NAFLD prevalence according to *H. pylori* status

The prevalence of NAFLD was significantly higher in the *H. pylori* positive subjects than that in the *H. pylori* negative subjects (33.5 ± 1.8% vs. 26.1 ± 1.7%, p <0.001). Among *H. pylori* positive subjects, the prevalence of NAFLD was significantly higher in the cagA negative subjects than in the cagA positive subjects (36.4 ± 2.4% vs. 31.1 ± 2.3%, p <0.001) (Fig 2). Subjects with NAFLD showed positive relationships with *H. pylori* positivity in the univariate analysis (OR: 1.43, 95% CI: 1.23–1.66). However, after adjustment for known metabolic risk factors, multivariate regression analysis showed that this association was attenuated (OR: 1.17, 95% CI: 0.95–1.43) with marginal significance. To identify the influence of cagA on NAFLD, we performed an analysis based on cagA status. NAFLD was significantly associated with the cagA positive group (OR: 1.28, 95% CI: 1.07–1.53) and cagA negative group (OR: 1.62, 95% CI: 1.30–2.03) in univariate analysis. CagA positivity was not significantly associated with NAFLD after adjustment of multiple risk factors (OR: 1.05, 95% CI: 0.81–1.37). However, the cagA
negative *Helicobacter pylori* positive group was significantly associated with NAFLD in multivariable regression analysis (OR: 1.30, 95% CI: 1.01–1.67) after adjustment (Table 3). The effect size of cagA negative *Helicobacter pylori* positivity was similar to that of Mexican Americans (OR 1.29; 95% CI 1.02–1.63), however, much lower than that of diabetes (OR 2.03; 95% CI 1.37–3.01) and hypertension (OR 1.67; 95% CI 1.26–2.19) (S1 Table).

When we evaluated the association of *H. pylori* seropositivity and/or cagA presence with advanced fibrosis among subjects with NAFLD (S2 Table), no statistically significant
| Characteristic | H. pylori- (n = 2,749) | H. pylori+ (n = 2,655) | H. pylori+ (n = 1,617) | CagA- (n = 1,038) |
|---------------|------------------------|------------------------|------------------------|-------------------|
| Age (years)   | 40.2 ± 0.12            | 48.4 ± 0.26            | 47.1 ± 0.34            | 49.9 ± 0.24       |
| Gender (male, %) | 46.1 ± 0.36            | 48.5 ± 0.52            | 49.7 ± 0.98            | 47.0 ± 0.82       |
| Body mass index (kg/m²) | 26.0 ± 0.07            | 27.2 ± 0.06            | 27.1 ± 0.07            | 27.3 ± 0.10       |
| Waist circumference (cm) | 90.3 ± 0.19            | 94.1 ± 0.20            | 93.4 ± 0.25            | 94.8 ± 0.26       |
| Race-ethnicity (%) |                       |                        |                        |                   |
| Non-Hispanic white | 85.1 ± 0.48            | 62.7 ± 0.87            | 51.3 ± 1.57            | 76.2 ± 0.43       |
| Non-Hispanic black | 7.0 ± 0.28             | 16.4 ± 0.36            | 23.0 ± 0.39            | 8.6 ± 0.29        |
| Mexican American | 2.9 ± 0.11             | 10.1 ± 0.40            | 11.3 ± 0.64            | 8.6 ± 0.38        |
| Other Race     | 4.9 ± 0.38             | 10.8 ± 0.62            | 14.4 ± 1.15            | 6.6 ± 0.25        |
| Smoking (%)    | 3.0 ± 0.42             | 29.6 ± 0.72            | 29.7 ± 1.00            | 29.5 ± 0.66       |
| History of cardiovascular disease (%) | 2.9 ± 0.15             | 6.5 ± 0.18             | 6.8 ± 0.33             | 6.1 ± 0.12        |
| Lipid-lowering medication (%) | 2.6 ± 0.14             | 3.5 ± 0.12             | 3.1 ± 0.14             | 4.0 ± 0.21       |
| Systolic blood pressure (mm Hg) | 119.2 ± 0.16           | 125.5 ± 0.20           | 124.7 ± 0.31           | 126.4 ± 0.24     |
| Diastolic blood pressure (mm Hg) | 73.2 ± 0.13            | 75.6 ± 0.11            | 75.8 ± 0.18            | 75.4 ± 0.10      |
| Total cholesterol (mg/dL) | 201.8 ± 0.42           | 211.4 ± 0.57           | 210.4 ± 0.67           | 212.5 ± 0.71     |
| HDL cholesterol (mg/dL) | 51.6 ± 0.17            | 49.8 ± 0.24            | 49.7 ± 0.31            | 49.9 ± 0.24      |
| Triglycerides (mg/dL) | 133.6 ± 1.15           | 147.2 ± 1.42           | 142.4 ± 2.21           | 153.0 ± 1.39     |
| ALT (IU/L)     | 15.5 ± 0.12            | 16.1 ± 0.16            | 16.0 ± 0.24            | 16.2 ± 0.16      |
| AST (IU/L)     | 20.0 ± 0.05            | 21.4 ± 0.12            | 21.4 ± 0.19            | 21.4 ± 0.11      |
| GGT (IU/L)     | 24.6 ± 0.14            | 28.3 ± 0.44            | 29.3 ± 0.87            | 27.1 ± 0.25       |
| Albumin (g/dL) | 4.33 ± 0.00            | 4.26 ± 0.01            | 4.2 ± 0.01             | 4.3 ± 0.01       |
| Platelet (x10³/L) | 282.9 ± 0.48           | 280.9 ± 0.72           | 280.5 ± 1.25           | 281.3 ± 0.65     |
| Glucose (mg/dL) | 96.5 ± 0.27            | 102.2 ± 0.20           | 102.9 ± 0.40           | 101.4 ± 0.21     |
| Vitamin D     | 30.3 ± 0.07            | 27.6 ± 0.18            | 27.1 ± 0.18            | 28.1 ± 0.25      |
| HbA1c (%)      | 5.2 ± 0.01             | 5.5 ± 0.01             | 5.5 ± 0.01             | 5.5 ± 0.01       |

Abbreviation: H. pylori, Helicobacter pylori; NAFLD, nonalcoholic fatty liver disease; HDL, high-density lipoprotein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; GGT, gamma-glutamyltransferase. Data are expressed as the mean ± standard error or proportion ± standard error

associations were noted in the univariate and multivariable models. As shown in S3 Table, H. pylori positivity was associated with significantly higher hazards of all-cause mortality, and the adjusted hazard ratio (HR) and 95% confidence interval (CI) was 1.95 (1.49–2.55) for univariate model and 1.30 (1.01–1.68) for multivariate model 1. With further adjustment for traditional risk factors, this association was attenuated and demonstrated only marginal significance (P for trend = 0.138). The cagA negative or positive H. pylori positive group with NAFLD demonstrated significant association with increased risk of all-cause mortality in univariate model. However, this association remained insignificant in multivariate model.

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Helicobacter pylori infection was associated with an increased risk of NAFLD in all ethnicities. However, *H. pylori* infection was significantly associated with NAFLD in non-Hispanic black participants. Regarding cagA status, cagA negative *H. pylori* was associated with an increased risk.
for NAFLD in the non-Hispanic white and non-Hispanic black populations. However, cagA negative \textit{H. pylori} status was not associated with NAFLD in Mexican-Americans (Fig 3).

### Discussion

The main finding in this nationally representative, population-based study was that the prevalence of NAFLD was higher in subjects with \textit{H. pylori} than in those without. In subjects with \textit{H. pylori}, the prevalence of NAFLD was greater in cagA negative subjects compared to cagA positive subjects. Additionally, cagA negative \textit{H. pylori} positivity was significantly associated with NAFLD after adjustment for multiple classical risk factors, confirming that a relevant clinical relationship exists between these two conditions.

Several studies have investigated the association between \textit{H. pylori} and NAFLD. Polyzos et al. reported that higher rates of anti-\textit{H. pylori} IgG were observed in the NAFLD group \((n = 28)\) compared to that of the healthy control group \((n = 25)\) [8]. They also showed that biopsy-proven NAFLD patients had higher insulin and tumor necrosis factor (TNF)-\(\alpha\) levels compared to those of controls. However, there were no significant differences in steatosis grade, fibrosis stage, lobular or portal inflammation, or balloon degeneration when NAFLD patients were divided based on \textit{H. pylori} IgG seropositivity or \(^{13}\)C urea breath test positivity [8]. This study was also limited by a small sample size. Dogan et al. showed that fatty liver determined by ultrasonographic score was observed more frequently in \textit{H. pylori} positive patients, but they did not consider other metabolic confounders [13]. NAFLD is considered as the hepatic manifestation of metabolic syndrome and is strongly associated with cardiovascular disease. [21]. Several studies reported that an association between \textit{H. pylori} and metabolic syndrome is significant as shown in case of NAFLD. In a large cross-sectional study from Japan, \textit{H. pylori} seropositivity was significantly associated with metabolic syndrome [22]. On the contrary, studies from highly endemic areas of \textit{H. pylori} revealed no association between positive \textit{H. pylori} serology and NAFLD [23,24]. In a recent meta-analysis, \textit{H. pylori} infection and NAFLD demonstrated a modest, significant association (pooled OR 1.21 (95% CI 1.07–1.37) [25]. Recently, Fan et al. found that prevalence of NAFLD was significantly higher in subjects with \textit{H. pylori} infection (36.0% vs. 33.3%, \(p < 0.05\)) [26]. However, after adjusting for confounding factors, \textit{H. pylori} infection was not significantly associated with NAFLD. In our study using representative US population, we found \textit{H. pylori} and cagA negativity was associated with NAFLD, which may partly explain the inconsistent results in the literature.

### Table 3. Univariate and multivariable analyses of the risk for NAFLD according to \textit{H. pylori} status.

|                      | Univariate |                | Multivariable-adjusted |                |
|----------------------|------------|----------------|------------------------|----------------|
|                      | OR (95% CI)| \(P\) Value   | OR (95% CI)            | \(P\) Value    |
| HP positivity        |            |                |                        |                |
| Negative             | Reference  |                | Reference              |                |
| Positive             | 1.43 (1.23–1.66) | <0.001     | 1.17 (0.95–1.43) | 0.139          |
| HP and Cag A positivity |            |                |                        |                |
| Negative             | Reference  |                | Reference              |                |
| CagA positive        | 1.28 (1.07–1.53) | 0.008       | 1.05 (0.81–1.37) | 0.715          |
| CagA negative        | 1.62 (1.30–2.03) | <0.001     | 1.30 (1.01–1.67) | 0.044          |

Abbreviation: NAFLD, nonalcoholic fatty liver disease; \textit{H. pylori}, \textit{Helicobacter pylori}; OR, odds ratio; CI, confidence interval. Multivariable models adjusted for age, sex, race-ethnicity, income, diabetes, hypertension, smoking status, waist circumference, alcohol consumption, caffeine consumption, total cholesterol, high-density lipoprotein-cholesterol, and transferrin saturation.

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In the present study, we found only infection with cagA negative H. pylori was significantly associated with NAFLD. CagA plays an important role not only in gastric inflammation but also in the development of gastric cancer. Regarding the effect of cagA on cardiovascular and metabolic risk factors, a German population-based cohort study showed decreased incidence of cardiovascular mortality in the cagA positive population compared with that of *H. pylori*.
negative subjects. The difference of incidence between the two groups was 33% - 38%, but the cardiovascular mortality of the cagA negative population was similar to that of the H. pylori-negative group [27]. Analysis of the association between H. pylori and mortality in the NHANES III study showed an inverse relationship between mortality from stroke and cagA positive H. pylori infection [28]. The inverse relation between cagA and cardiovascular mortality can be explained by regulatory T cells (T-reg). In mice model, cagA plays an important role in the migration of CD4+ T cells in the gastric mucosa and cagA-dependent T-cell priming induces T-reg differentiation [29]. The presence of T-reg in the gastric mucosa of H. pylori-infected subjects suggested their involvement in suppressing mucosal immune responses, contributing to the infection persistence and modulation of the H. pylori-induced gastritis [30]. Reduced risk of asthma and allergies in H. pylori-infected populations appears to be related to the induction of T-reg responses [31]. T-reg is also known to be important in regulating inflammatory processes in NAFLD [31]. The highly stimulated regulatory immune system in the setting of cagA may be the reason why only cagA negative H. pylori infection is related with NAFLD as shown in this study.

Insulin resistance can be another possible explanation for the association between H. pylori and NAFLD. Insulin resistance is a key factor in both metabolic syndrome and NAFLD. One study showed that insulin resistance, which is measured as HOMA-IR, was significantly higher in the H. pylori positive group than in the H. pylori negative counterparts [32]. A systematic review of nine studies investigating the association between H. pylori infection and insulin resistance indicated that H. pylori infection was positively associated with insulin resistance [33]. It is well known that even chronic subclinical inflammation is associated with cardiovascular disease [34]. Long-term H. pylori infection induces a ‘state of inflammation’ by stimulating excessive release of pro-inflammatory cytokine and vasoactive substances, such as interleukin (IL)-6, IL-8, IL-1β, and TNF-α [35,36]. TNF-α is a key mediator of both direct and indirect effects of H. pylori infection on NAFLD. TNF-α interferes with insulin signaling, thereby favoring steatosis, and may play a pro-inflammatory role in the pathogenesis of NAFLD [37,38]. In addition, TNF-α can accelerate lipolysis, leading to an increase in free fatty acids. This can result in hepatocyte dysfunction, including increased oxidative stress, induction of endoplasmic reticulum dysregulation and subsequent expression of pro-inflammatory cytokines [39,40]. We noted that H. pylori positivity was associated with increased risk for all-cause mortality among subjects with NAFLD, though after adjusting for traditional risk factors, the association remained at marginal significance and thus needs further confirmation. This result perhaps suggests that the effect of H. pylori positivity in NAFLD on all-cause mortality may be mediated in part by diabetes, hypertension, and smoking.

In case of a disruption of the intestinal barrier, lipopolysaccharide from gram-negative bacteria increases in the portal circulation and is accompanied by increased levels of endotoxin-mediated cytokines in the liver. So bacterial constituents and cytokines enhance hepatic inflammation and fibrosis [41]. Miele et al. also found that in NAFLD patients, increased gut permeability and the prevalence of small intestine bacterial overgrowth correlated with the severity of steatosis but not with the presence of steatohepatitis [42]. H. pylori is believed to colonize not only the stomach and duodenal epithelium but also the biliary epithelium [43]. So H. pylori colonization in stomach and duodenum can increase in lipopolysaccharides and endotoxin in portal circulation and can promote hepatic inflammation and fibrosis.

The prevalence of H. pylori infection showed racial disparities. Both non-Hispanic blacks and Mexican-Americans had a significantly higher prevalence of H. pylori than non-Hispanic whites [44]. The prevalence of cagA positive H. pylori was higher in non-Hispanic blacks and Mexican-Americans versus non-Hispanic whites [44]. However, in this study, a significant association between cagA negative H. pylori strains and NAFLD was found in non-Hispanic
whites and non-Hispanic blacks but not in Mexican-Americans. Further research is needed to confirm these ethnic differences.

Currently, only two studies have reported the effect of H. pylori eradication on metabolic profile or NAFLD. One prospective study revealed that in the group with successful eradication, levels of total adiponectin and each multimer form were significantly increased 12 weeks after completion of treatment [45]. This study suggested that eradication could have a beneficial effect on metabolic risk by increasing adiponectin levels. A recent small-scale randomized open label study showed that lifestyle modification plus H. pylori eradication did not result in significant differences in liver fat content by ultrasonography and in insulin resistance compared with that of lifestyle modification alone [46]. However, the primary outcome of this study is relatively subjective, and the open label design may be a pitfall. Therefore, further large prospective studies are needed to confirm the effect of H. pylori eradication on NAFLD.

There are strengths and limitations in our study. We studied a large representative sample from the US general population; thus, the results can be generalizable. Second, this study performed analysis according to cagA seropositivity and studied the differential impact of cagA presence (versus cagA absence) on the association between NAFLD and H. pylori. However, there may be other predictive factors that may play an influential role in impacting the association of H. pylori and NAFLD. Third, we included sufficient confounders from the NHANES III data, which increased the accuracy of this study. Finally, our study included various ethnic/racial groups, and thus, our conclusions may be applicable to the general population. This study also has some limitations. First, because of its cross-sectional design, a causal relationship between H. pylori infection and NAFLD could not be identified. Second, our diagnosis of NAFLD was based on ultrasonographic examination, which is not able to differentiate nonalcoholic steatohepatitis from nonalcoholic fatty liver. In addition, hepatic ultrasonography cannot identify fatty infiltration below 30% and has intra- and inter-observer variability in making a diagnosis. Third, our study is unable to offer a mechanistic explanation to associate cagA negative H. pylori seropositivity with the risk of NAFLD. Further studies are warranted to determine mechanism(s) explaining this association.

In conclusion, the present study suggests that H. pylori infection is related to an increased prevalence of NAFLD in the general population. CagA negative H. pylori infection was associated with an increased risk for NAFLD, confirming that a relevant clinical relationship exists between these two conditions. However, since pathogenic mechanisms have not been fully elucidated, future studies are needed to clearly understand the clinical implications of our results.

Supporting information
S1 Table. Univariate and multivariable analyses of the risk for advanced fibrosis among subjects with NAFLD according to H. pylori status.
(DOCX)

S2 Table. Univariate and multivariable cox-regression analyses of the risk for all-cause mortality among subjects with NAFLD according to H. pylori status.
(DOCX)

S3 Table. Multivariable analysis of the risk for NAFLD.
(DOCX)

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References
1. Ong JP, Younossi ZM. Epidemiology and natural history of NAFLD and NASH. Clin Liver Dis. 2007; 11:1–16, vii. https://doi.org/10.1016/j.cld.2007.02.009 PMID: 17549468
2. Kim D, Kim WR, Kim HJ, Theeau TM. Association between noninvasive fibrosis markers and mortality among adults with nonalcoholic fatty liver disease in the United States. Hepatology. 2013; 57:1357–1365. https://doi.org/10.1002/hep.26156 PMID: 23175136
3. The Korean Association for the Study of the Liver (KASL). KASL clinical practice guidelines: Management of nonalcoholic fatty liver disease. Clin Mol Hepatol. 2013; 19(4):325–348. https://doi.org/10.3350/cmh.2013.19.4.325 PMID: 24459637
4. Chung GE, Kim D, Kim W, Yim JY, Park MJ, Kim YJ, et al. Non-alcoholic fatty liver disease across the spectrum of hypothyroidism. J Hepatol. 2012; 57:150–156. https://doi.org/10.1016/j.jhep.2012.02.027 PMID: 22425701
5. Park HE, Kwak MS, Kim D, Kim MK, Cha MJ, Choi SY. Nonalcoholic Fatty Liver Disease Is Associated With Coronary Artery Calcification Development: A Longitudinal Study. J Clin Endocrinol Metab. 2016; 101(8):3134–3143. https://doi.org/10.1210/jc.2016-1525 PMID: 27253666
6. Kim D, Choi SY, Park EH, Lee W, Kang JH, Kim W, et al. Nonalcoholic fatty liver disease is associated with coronary artery calcification. Hepatology. 2012; 56(2):605–613. https://doi.org/10.1002/hep.25593 PMID: 22271511
7. Choi SY, Kim D, Kang JH, Park MJ, Kim YS, Lim SH, et al. Nonalcoholic fatty liver disease as a risk factor of cardiovascular disease: relation of non-alcoholic fatty liver disease to carotid atherosclerosis. Korean J Hepatol. 2008; 14:77–88. https://doi.org/10.3350/kjhep.2008.14.1.77 PMID: 18367860
8. Park MJ, Choi SH, Kim D, Kang SJ, Chung SJ, Choi SY, et al. Association between Helicobacter pylori Seropositivity and the Coronary Artery Calcium Score in a Screening Population. Gut Liver. 2011; 5(3):321–327. https://doi.org/10.5009/gnl.2011.5.3.321 PMID: 21927661
9. Suzuki H, Franceschi F, Nishizawa T, Gasbarrini A. Extragastric manifestations of Helicobacter pylori infection. Helicobacter. 2011; 16 Suppl 1:65–69. https://doi.org/10.1111/j.15235378.2011.00883.x
10. Murphy G, Michel A, Taylor PR, Albanes D, Weinstein SJ, Virtamo J, et al. Association of seropositivity to Helicobacter species and biliary tract cancer in the ATBC study. Hepatology, 2014; 60:1963–1971. https://doi.org/10.1002/hep.27193 PMID: 24797247
11. Fox JG, Feng Y, Theve EJ, Racynski AR, Fiala JL, Doernte AR, et al. Gut microbes define liver cancer risk in mice exposed to chemical and viral transgenic hepatocarcinogens. Gut. 2010; 59:88–97. https://doi.org/10.1136/gut.2009.183749 PMID: 19850960
12. Chen Yu, Segers Stephanie, Blaser Martin J. Association between Helicobacter pylori and mortality in the NHANES III study. Gut. 2013; 62:1262–1269. https://doi.org/10.1136/gutjnl-2012-303018 PMID: 23033440
13. Dogan Z, Filik L, Ergul B, Sarikaya M, Akbai E. Association between Helicobacter pylori and liver-to-spleen ratio: a randomized-controlled single-blind study. Eur J Gastroenterol Hepatol. 2013; 25:107–110. https://doi.org/10.1097/MEG.0b013e3283590c10 PMID: 23013624
14. Chalasani N, Younossi Z, Lavine JE, Diehl AM, Brunt EM, Cusi K, et al. The diagnosis and management of non-alcoholic fatty liver disease: practice Guideline by the American Association for the Study of Liver Diseases, American College of Gastroenterology, and the American Gastroenterological Association. Hepatology. 2012; 55:2005–2023. https://doi.org/10.1002/hep.25762 PMID: 22486764
15. Everhart JE, Kruszon-Moran D, Perez-Perez GI, Tralka TS, McQuillan G. Seroprevalence and ethnic differences in Helicobacter pylori infection among adults in the United States. J Infect Dis. 2000; 181:1359–1363. https://doi.org/10.1086/315364 PMID: 10762567
16. Blaser MJ, Perez-Perez GI, Kleanthous H, Cover TL, Peek RM, Chyou PH, et al. Infection with Helicobacter pylori strains possessing cagA is associated with an increased risk of developing adenocarcinoma of the stomach. Cancer Res. 1995; 55:2111–2115. PMID: 7743510
17. Cho I, Blaser MJ, Francois F, Mathew JP, Ye XY, Goldberg JD, et al. Helicobacter pylori and overweight status in the United States: data from the Third National Health and Nutrition Examination Survey. Am J Epidemiol. 2005; 162:579–584. https://doi.org/10.1093/aje/kwi237 PMID: 16093294

18. NCHS. NHANES III Hepatic Steatosis Ultrasound Images Assessment Procedures Manual. Available from: https://wwwn.cdc.gov/nchs/Data/Nhanes3/Manuals/hsuiaap.pdf.

19. Kim D, Li AA, Ahmed A. Leucocyte telomere shortening is associated with nonalcoholic fatty liver disease-related advanced fibrosis. Liver Int. 2018 May 24 Epub. https://doi.org/10.1111/liv.13886 PMID: 29797393

20. Ingram DD, Makuc DM. Statistical issues in analyzing the NHANES I Epidemiologic Followup Study. Series 2: Data evaluation and methods research. Vital Health Stat 2. 1994; 121:1–30.

21. Li M, Shen Z, Li YM. Potential role of Helicobacter pylori infection in nonalcoholic fatty liver disease. World J Gastroenterol. 2013; 19:7024–7031. https://doi.org/10.3748/wjg.v19.i41.7024 PMID: 24222944

22. Gunji T, Matsuhashi N, Sato H, Fujibayashi K, Okumura M, Sasabe N, et al. Helicobacter pylori infection is significantly associated with metabolic syndrome in the Japanese population. Am J Gastroenterol. 2008; 103:3005–3010. https://doi.org/10.1111/j.1572-0241.2008.02151.x PMID: 19086952

23. Okushin K, Takahashi Y, Yamamichi N, Shimamoto T, Enooku K, Fujinaga H, et al. Helicobacter pylori infection is not associated with fatty liver disease including non-alcoholic fatty liver disease: a large-scale cross-sectional study in Japan. BMC Gastroenterol. 2015:15–25. https://doi.org/10.1186/s12876-015-0244-z

24. Baeg MK, Yoon SK, Ko SH, Noh YS, Lee IS, Choi MG. Helicobacter pylori infection is not associated with nonalcoholic liver disease. World J Gastroenterol. 2016; 22(8):2592–2600. https://doi.org/10.3748/wjg.v22.i8.2592 PMID: 26937147

25. Wijarnpreecha K, Thongprayoon C, Panjawanatan P, Manatsathit W, Jaruvongvanich V, Ungprasert P. Helicobacter pylori and Risk of Nonalcoholic Fatty Liver Disease: A Systematic Review and Meta-analysis. J Clin Gastroenterol. 2018; 52(5):386–391. https://doi.org/10.1097/MCG.0000000000000784 PMID: 28098578

26. Fan N, Peng L, Xia Z, Zhang L, Wang Y, Peng Y. Helicobacter pylori infection is not associated with non-alcoholic fatty liver disease: A cross-sectional study in China. Front Microbiol. 2018; 9:73. https://doi.org/10.3389/fmicb.2018.00073 PMID: 29445363

27. Schottker B, Adamu MA, Weck MN, Muller H, Brenner H. Helicobacter pylori infection, chronic atrophic gastritis and major cardiovascular events: A population-based cohort study. Atherosclerosis. 2012; 220:569–574. https://doi.org/10.1016/j.atherosclerosis.2011.11.029 PMID: 22189198

28. Stein M, Ruggiero P, Rappuoli R, Bagnoli F. Helicobacter pylori CagA: From pathogenic mechanisms to its use as an anti-cancer vaccine. Front Immunol. 2013; 4:328. https://doi.org/10.3389/fimmu.2013.00328 PMID: 24133496

29. Swiderska M, Jaroszewicz J, Stawicka A, Parfeniuk-Kowerda A, Chabowski A, Flisiak R. The interplay between Th17 and T-regulatory responses as well as adipokines in the progression of non-alcoholic fatty liver disease. Clin Exp Hepatol. 2017; 3(3):127–134. https://doi.org/10.5114/che.2017.68466 PMID: 29062902

30. Aydemir S, Bayraktaroglu T, Sert M, Sokmen C, Atmaca H, Mungan G, et al. The effect of Helicobacter pylori on insulin resistance. Dig Dis Sci. 2005; 50:2090–2093. https://doi.org/10.1007/s10620-005-3012-z PMID: 16240220

31. Polyzos SA, Kountouras J, Zavos C, Deretzis G. The association between Helicobacter pylori infection and insulin resistance: a systematic review. Helicobacter. 2011; 16(2):79–88. https://doi.org/10.1111/j.1523-5378.2011.00822.x PMID: 21435084

32. Crabtree JE, Shallcross TM, Heatley RV, Wyatt JI. Mucosal tumour necrosis factor alpha and interleukin-6 in patients with Helicobacter pylori associated gastritis. Gut. 1991; 32 (12):1473–1477. PMID: 1773951

33. Basso D, Plebani M, Kusters JG. Pathogenesis of Helicobacter pylori infection. Helicobacter. 2010; 15 Suppl 1:14–20. https://doi.org/10.1111/j.1523-5378.2010.00781.x PMID: 21054648
37. Hui JM, Hodge A, Farrell GC, Kench JG, Kriketos A, George J. Beyond insulin resistance in NASH: TNF-alpha or adiponectin? Hepatology. 2004; 40:46–54. https://doi.org/10.1002/hep.20280 PMID: 15239085

38. Feldstein AE, Werneburg NW, Canbay A, Guicciardi ME, Bronk SF, Rydzewski R, et al. Free fatty acids promote hepatic lipotoxicity by stimulating TNF-alpha expression via a lysosomal pathway. Hepatology. 2004; 40:185–194. https://doi.org/10.1002/hep.20283 PMID: 15239102

39. Rolo AP, Teodoro JS, Palmeira CM. Role of oxidative stress in the pathogenesis of nonalcoholic steatohepatitis. Free Radic Biol Med. 2012; 52:59–69. https://doi.org/10.1016/j.freeradbiomed.2011.10.003 PMID: 22064361

40. Ibrahim SH, Kohli R, Gores GJ. Mechanisms of lipotoxicity in NAFLD and clinical implications. J Pediatr Gastroenterol Nutr. 2011; 53:131–140. https://doi.org/10.1097/MPG.0b013e31822578db PMID: 21629127

41. Gabele E, Dostert K, Hofmann C, Wiest R, Scholmerich J, Hellerbrand C, et al. DSS induced colitis increases portal LPS levels and enhances hepatic inflammation and fibrogenesis in experimental NASH. J Hepatol. 2011; 55:1391–1399. https://doi.org/10.1016/j.jhep.2011.02.035 PMID: 21703208

42. Miele L, Valenza V, La Torre G, Montalto M, Cammarota G, Ricci R, et al. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. Hepatology. 2009; 49:1877–1887. https://doi.org/10.1002/hep.22848 PMID: 19291785

43. Figura N, Franceschi F, Santucci A, Bernardini G, Gasbarrini G, Gasbarrini A. Extragastric manifestations of Helicobacter pylori infection. Helicobacter. 2010; 15 Suppl 1:60–68. https://doi.org/10.1111/j.1523-5378.2010.00778.x PMID: 21054655

44. Grad YH, Lipsitch M, Aiello AE. Secular trends in Helicobacter pylori seroprevalence in adults in the United States: Evidence for sustained race/ethnic disparities. Am J Epidemiol. 2012; 175(1):54–59. https://doi.org/10.1093/aje/kwr288 PMID: 22085628

45. Ando T, Ishikawa T, Takagi T, Imamoto E, Kishimoto E, Okajima A, et al. Impact of Helicobacter pylori eradication on circulating adiponectin in humans. Helicobacter. 2013; 18:158–164. https://doi.org/10.1111/hel.12028 PMID: 23167259

46. Jamalir, Mofid A, Vahedi H, Farzaneh R, Dowlatshashi S. The effect of helicobacter pylori eradication on liver fat content in subjects with non-alcoholic Fatty liver disease: a randomized open-label clinical trial. Hepat Mon. 2013; 13:e14679 https://doi.org/10.5812/hepatmon.14679 PMID: 24358044