Serum Biomarkers of Endothelial Dysfunction in Fabry Associated Cardiomyopathy

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Background: Fabry disease (FD) is characterized by early development of vasculopathy and endothelial dysfunction. However, it is unclear whether these findings also play a pivotal role in cardiac manifestation. As Fabry cardiomyopathy (FC) is the leading cause of death in FD, we aimed to gather a better insight in pathological mechanisms of the disease.

Methods: Serum samples were obtained from 17 healthy controls, 15 FD patients with and 7 without FC. FC was defined by LV wall thickening of >12 mm in cardiac magnetic resonance imaging and serum level of proBNP, high sensitive Troponin T (hsT), and globotriaosylsphingosine (lyso-GB3) were obtained. A multiplex ELISA-Assay for 23 different angiogenesis markers was performed in pooled samples. Markers showing significant differences among groups were further analyzed in single samples using specific Elisa antibody assays. L-homoarginine (hArg), L-arginine, asymmetric (ADMA), and symmetric Dimethylarginine (SDMA) were quantified by liquid chromatography—mass spectrometry.

Results: Angiostatin and matrix metalloproteinase 9 (MMP-9) were elevated in FD patients compared to controls independently of the presence of FC (angiostatin: 98 ± 25 vs. 75 ± 15 ng/mL; p = 0.001; MMP-9: 8.0 ± 3.4 vs. 5.0 ± 2.4 µg/mL; p = 0.002). SDMA concentrations were highest in patients with FC (0.90 ± 0.64 µmol/l) compared to patients without (0.57 ± 0.10 µmol/l; p = 0.027) and vs. controls (0.58 ± 0.12 µmol/l; p = 0.006) and was positively correlated with indexed LV-mass (r = 0.61; p = 0.003), hsT (r = 0.56, p = 0.008), and lyso-Gb3 (r = 0.53, p = 0.013). Accordingly, the ratio of L-homoarginine to SDMA (hArg/SDMA) was lowest in patients with FC (2.63 ± 1.78) compared to controls (4.16 ± 1.44; p = 0.005). For L-arginine, hArg and ADMA no significant differences among groups could be detected, although a trend toward higher ADMA and lower hArg levels could be observed in the FC group. Furthermore, a significant relationship between kidney and cardiac function could be revealed (p = 0.045).
INTRODUCTION

Fabry disease (FD) is an X-linked recessive multi-systemic storage disorder caused by a decreased activity of the lysosomal enzyme alpha-Galactosidase A (GLA) (1, 2). Due to the deposition of globotriaosylceramide (Gb3) in vascular lysosomes neutral glycosphingolipids accumulate in plasma and various tissues throughout the body (3). Typical manifestations of FD are cardiac, neurological, renal, ocular, dermatological, and gastrointestinal (4–6) with cardiovascular disease as the leading cause of death in FD patients (7). Prognosis of cardiomyopathy is particularly poor (8), thus, appropriate diagnosis and treatment of Fabry associated cardiomyopathy (FC) is crucial. Enzyme Replacement Therapy (ERT) has proven to significantly reduce accumulation of Gb3, especially intracellular deposits in the coronary endothelium (9, 10) and to halt or even partially reverse FC. However, in advanced stages of FD with a severe cardiac phenotype the effectiveness of ERT is profoundly diminished and the disease can even progress (11–13). Therefore, a better understanding of the underlying mechanisms contributing to the development of FC is urgently needed to improve treatment and outcome of FD patients.

Clinical studies provide evidence of increased intima-media thickness (IMT) and impaired artery flow-mediated dilatation in FD (14, 15) indicating an early onset of atherosclerosis in these patients. Moreover, different studies suggest that myocardial fibrosis, detected by cardiac magnetic resonance imaging (cMRI), may contribute to left ventricular remodeling in FD (16, 17). Myocardial fibrosis and subsequent remodeling are caused by an altered extracellular matrix turnover, which is catalyzed by Matrix Metalloproteinases (MMP) including MMP-9 (18). This is in line with the detection of increased serum MMP-9 level in Fabry patients compared to controls (19). Matsunaga et al showed that the inhibition of NO synthase resulted in increased MMP-9 and MMP-2 activities suggesting a link between oxidative stress and extracellular matrix turnover. Several clinical and experimental studies demonstrated inflammatory activity and endothelial nitric oxide synthase (eNOS) alterations in vascular cells of FD patients (20–23). These findings support the hypothesis of early occurring vasculopathy and endothelial dysfunction in FD. Whether these findings play a pivotal role in cardiac manifestation has not yet been sufficiently investigated. Accordingly, the aim of this study was to gather an insight in underlying pathological mechanisms by determining serum markers of endothelial dysfunction, angiogenesis and cardiac function in FD patients with and without FC.

METHODS

Study Population

Serum samples from 15 FD patients with FC, 7 without FC, and 17 healthy controls were collected between September 2014 and December 2016. FD was confirmed by molecular genetic analysis revealing the following mutations: 4x p.N215S, 2x p.E341K, 2x c.1277_1278delAA, 2x c.718_719delAA, p.N320l, p.A143T, p.A230_I232del, p.Q327L, p.A389V, c.717delAA, p.I384N, p.P205T, p.S247P, p.Q327L, p.R227Q. FC was defined by LV wall thickening of >12 mm assessed in cMRI. Furthermore, 11 FD patients with FC and one patient without FC received ERT at the time of blood sampling. The study was in line with the principles outlined in the Declaration of Helsinki and approved by the local ethics committee. All participants gave their written informed consent for participation in the study.

Enzyme-Linked Immunosorbent Assays

Blood samples were centrifuged at 4000 × G for 10 min at room temperature and obtained serum was aliquoted and stored at −80°C until use. For the multiplex Enzyme-Linked Immunosorbent Assay (ELISA) a Human Angiogenesis Antibody Array Membrane (Abcam PLC, Cambridge, UK, ab169808) was used. Aliquots of serum samples were pooled into three groups: FD with FC (n = 15), FD without FC (n = 7), and controls (n = 17). The assay allowed a simultaneous and semi-quantitative analysis of 23 targets (Angiopoietin 1, Angiopoietin 2, Angiostatin, Endostatin, G-CSF, GM-CSF, I-309, IL-10, IL-1 alpha, IL-1 beta, IL-2, IL-4, I-TAC, MCP-3, MCP-4, MMP-1, MMP-9, PECAM-1, Tie-2, TNF alpha, suPAR, VEGF, VEGF-R3) and was performed in duplicates. Visualization of membrane signals was performed by ChemiDoc™ MP Imaging System and the densitometry software Image Lab™.

Markers showing significant differences among groups in the multiplex Elisa-Assay were further analyzed in single samples. Specific ELISAs from Abcam were used for MMP-9, angiostatin, soluble urokinase-type plasminogen activator receptor (suPAR), and vascular endothelial growth factor (VEGF) quantification according to the manufacturers’ instructions.

Liquid Chromatography—Tandem Mass Spectrometry (LC-MS/MS) Measurements

L-Arginine, L-homoarginine (hArg), asymmetric (ADMA), and symmetric Dimethylarginine (SDMA) were quantified as described previously (24, 25). In brief, 25 µL of EDTA plasma were diluted with 100 µL 2H7-arginine, 13C2D2N4-hArg, and 2H7-ADMA solved in methanol. Proteins were

Conclusion: Elevated MMP-9 and angiostatin levels suggest an increased extracellular matrix turnover in FD patients. Furthermore, endothelial dysfunction may also be involved in FC, as SDMA and hArg/SDMA are altered in these patients.

Keywords: matrix metalloproteinase 9, angiostatins, SDMA, homoarginine, Fabry disease, Fabry cardiomyopathy, vasculopathy, endothelial dysfunction
Cardiac MRI

Cardiac MRI was performed with a 1.5 Tesla MRI scanner (Achieva, Philips Medical Systems, Philips, Best, The Netherlands). The examination contained a retrospectively gated cine-MRI in cardiac short and long axis orientations using a steady-state free precession (SSFP) sequence to quantify regional and global left ventricular (LV) function and the LV-myocardial mass. Ten minutes after bolus injection of 0.075 mmol/kg Gd-BOPTA (MultiHance®), end-diastolic late gadolinium enhanced (LGE) images were acquired by phase-sensitive inversion recovery (PSIR) sequences to quantify areas of myocardial fibrosis. LGE images were obtained in the LV short-axis as well as in two-, three-, and four-chamber views and quantified using cvi42® software (Circle Cardiovascular Imaging Inc., Calgary, Alberta, Canada). All quantified parameters included LV function, end-diastolic and end-systolic volumes, stroke volume, ejection fraction, left ventricular mass, regional wall thickness, and LGE. Left ventricular mass was indexed to the body surface area calculated by DuBois & DuBois formula.

Statistical Analysis

Obtained data were analyzed using SPSS, version 23. Not normally distributed variables were log transformed if necessary. Assessment of group differences was performed by analysis of variance (ANOVA) or analysis of covariance (ANCOVA) when additionally adjusted for age, sex and eGFR. Post-hoc group comparisons were analyzed if the global differences were significant. Correlations were investigated by Pearson’s correlation tests. Logistic regression was performed to investigate the relationship between renal and cardiac function. Concentrations are presented as mean ± standard deviation and a p-value of <0.05 was set as statistically significant.

RESULTS

Baseline Characteristics

Cardiac MRI measurements revealed significantly higher indexed left-ventricular masses of FD patients with FC compared to controls and FD patients without FC as shown in Table 1. Furthermore, similar results could be shown for septal thickness and mean LGE size, which were significantly thicker/higher in FD patients with FC. No differences among groups could be detected for left ventricular ejection fraction and stroke volume. Laboratory values revealed significantly higher levels of proBNP, hsT, and lyso-Gb3 in the cardiomyopathy group. Moreover, patients with FC showed elevated renal function parameters.

Quantification of Angiogenesis Markers by Enzyme-Linked Immunosorbent Assays

The multiplex ELISA indicated differences among the pooled groups for angiostatin, MMP-9 and suPAR. In Figure 1 the MMP-9 and angiostatin levels from the ensuing ELISA assays are demonstrated. MMP-9 concentrations were 1.51 times higher in FD patients with FC and 2.07 times higher in FD patients without FC compared to controls. The mean MMP-9 level of all FD patients was 1.67 times higher compared to controls (Table 2). Accordingly, angiostatin ELISA provided 1.33 times higher angiostatin levels in FD patients with FC and 1.23 times higher concentrations in FD patients without FC compared to controls. The mean angiostatin level of all FD patients was 1.3 times higher compared to controls (Table 2). However, no significant differences between the two FD groups could be detected for angiostatin and MMP-9. Moreover, neither suPAR nor VEGF concentrations revealed a significant difference between FD patients and healthy controls.

Correlations of Biomarkers to Anthropometric and Clinical Phenotypes

No statistically relevant age or sex dependencies for MMP-9, angiostatin, SDMA, or hArg/SDMA could be observed. As shown in Figure 3A, SDMA was positively correlated with LV-mass, lyso-Gb3, and hsT and negatively correlated with eGFR. Accordingly, as shown in Figure 3B, hArg/SDMA showed a negative correlation to LV-mass, lyso-Gb3, and a positive correlation with eGFR. Moreover, correlations of indexed LV-mass with ADMA, hArg/ADMA, suPAR, lyso-Gb3, cardiac, and renal parameters could be revealed (Table 3). For MMP-9 and angiostatin no significant correlations to any of these variables could be observed (data not shown).

Relationship of SDMA and HArg/SDMA to Renal Function

Analysis of covariance with adjustment for eGFR diminished the significant group differences. Therefore, SDMA and hArg/SDMA
TABLE 1 | Baseline characteristics, cMRI measurements, and laboratory values.

|                      | Control (1) | Fabry no FC (2) | Fabry FC (3) | p (ANOVA) | p (1 vs. 2) | p (2 vs. 3) | p (1 vs. 3) |
|----------------------|-------------|-----------------|-------------|-----------|------------|------------|------------|
| n                    | 17          | 7               | 15          |           |            |            |            |
| Age (years)          | 30.4 ± 6.3  | 36.3 ± 10.1     | 47.5 ± 13.0 | 0.224     | –          | –          | –          |
| Male, n (%)          | 15 (88)     | 1 (14)          | 10 (67)     | 0.063     | –          | –          | –          |
| ERT, n (%)           | 0 (0)       | 1 (14)          | 11 (73)     | 0.001     | 0.817      | <0.001     | <0.001     |
| MSSI –               | –           | 5.6 ± 3.2       | 25.7 ± 7.5  | 0.011     | 0.988      | 0.015      | 0.009      |
| LVEF (%)             | 61.1 ± 3.7  | 70.0 ± 8.8      | 68.0 ± 14.0 | 0.011     | 0.988      | 0.015      | 0.009      |
| SV (ml/m²)           | 114.2 ± 20.1| 91.4 ± 13.0     | 100.8 ± 20.0| 0.001     | 0.042      | 0.003      | 0.003      |
| LVM indexed to BSA (g/m²) | 73.1 ± 8.2  | 73.3 ± 29.9     | 107.1 ± 34.7| <0.001    | 0.321      | 0.009      | <0.001     |
| Septal thickness (mm)| 8.9 ± 1.7   | 8.6 ± 1.1       | 15.5 ± 3.7  | <0.001    | 0.817      | <0.001     | <0.001     |
| LGE positive, n      | 0           | 1               | 10          | <0.001    | 0.990      | 0.001      | <0.001     |
| LGE size LV mean (5th SD) (%) | 0.0 | 0.014 ± 0.038 | 4.014 ± 3.970 | <0.001    | 0.990      | 0.001      | <0.001     |
| proBNP (pg/ml)       | 37.7 ± 35.2 | 67.6 ± 44.3     | 492.9 ± 634.0| <0.001    | 0.248      | 0.012      | <0.001     |
| hsT (pg/ml)          | 6.9 ± 8.8   | 3.7 ± 0.8       | 28.8 ± 25.2 | 0.001     | 0.672      | 0.003      | 0.001      |
| eGFR (ml/min/1.73 m²) | 104.2 ± 15.8| 95.7 ± 19.6     | 71.9 ± 21.5 | <0.001    | 0.321      | 0.009      | <0.001     |
| Creatinine (mg/dl)   | 0.94 ± 0.14 | 0.81 ± 0.14     | 1.46 ± 1.50 | 0.042     | 0.347      | 0.020      | 0.062      |
| ACR (mg/g)           | –           | 30.2 ± 24.2     | 1534.6 ± 4314.0| –         | –          | 0.006      | –          |
| Lyso-Gb3 (ng/ml)     | –           | 3.7 ± 3.9       | 36.0 ± 32.9 | –         | –          | 0.003      | –          |

Data are presented as mean ± standard deviation. Significant p-values are marked in bold. Results of the analysis of variance (ANOVA) are shown. If ANOVA was significant post-hoc group comparisons were performed. ACR, albumin-to-creatinine ratio; cMRI, cardiac magnetic resonance imaging; eGFR, estimated glomerular filtration rate using the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) formula; ERT, enzyme replacement therapy; Fabry no FC, Fabry patients without Fabry cardiomyopathy; hsT, high sensitive cardiac Troponin T; LGE, late gadolinium enhancement; LGE size LV mean (5th SD), 5th standard deviation of left ventricular mean late gadolinium enhancement size; LVEF, left ventricular ejection fraction; LVM indexed to BSA, left ventricular end-diastolic mass indexed to body surface area; Lyso-Gb3, Globotriaosylsphingosine; MSSI, Mainz severity score index; proBNP, prohormone of brain natriuretic peptide; SV, stroke volume.

FIGURE 1 | Box-plots of MMP9 (A) and angiostatin (B) concentration in 17 healthy controls, 7 FD patients without and 15 FD patients with FC. Box plots represent median, 25th and 75th percentiles. Whiskers indicate minimum and maximum without outliers and extremes. On the Y-axis concentrations of the biomarkers are presented (log scale). Brackets indicate p-values from the analysis of variance (ANOVA) with post-hoc group comparisons. MMP9, matrix metalloproteinase 9; n.s., not significant.

DISCUSSION
The aim of this study was to investigate the role of vasculopathy and endothelial dysfunction in FD with special respect to Fabry associated cardiomyopathy. In blood samples of FD patients generally higher levels of MMP-9 and angiostatin could be detected independently of an existing FC, supporting the...
TABLE 2 | Markers of endothelial dysfunction.

|                | Control (1) | Fabry no FC (2) | Fabry FC (3) | \( p \) (ANOVA) | \( p \) (1 vs. 2) | \( p \) (2 vs. 3) | \( p \) (1 vs. 3) |
|----------------|-------------|-----------------|--------------|-----------------|-----------------|-----------------|-----------------|
| VEGF (pg/ml)   | 59.1 ± 51.7 | 70.1 ± 100.7    | 89.8 ± 58.6  | 0.127           | –               | –               | –               |
| suPAR (ng/ml)  | 1.6 ± 0.5   | 1.7 ± 0.7       | 3.2 ± 4.4    | 0.138           | –               | –               | –               |
| MMP-9 (µg/ml)  | 5.0 ± 2.4   | 9.6 ± 3.6       | 7.3 ± 3.1    | 0.004           | 0.002           | 0.163           | 0.021           |
| suPAR (ng/ml)  | 1.6 ± 0.5   | 1.7 ± 0.7       | 3.2 ± 4.4    | 0.138           | –               | –               | –               |
| MMP-9 (µg/ml)  | 5.0 ± 2.4   | 9.6 ± 3.6       | 7.3 ± 3.1    | 0.004           | 0.002           | 0.163           | 0.021           |
| Angiostatin (ng/ml) | 75.0 ± 15.2  | 92.9 ± 19.2    | 101.0 ± 27.3 | 0.002           | 0.038           | 0.468           | 0.001           |
| - Angiostatin FD total | 75.0 ± 15.2  | 98 ± 25         | –            | –               | –               | –               | –               |
| Arginine (µmol/l) | 111.4 ± 30.1 | 114.4 ± 29.6   | 113.3 ± 31.0 | 0.944           | –               | –               | –               |
| L-homoarginine (µmol/l) | 2.36 ± 0.82  | 2.24 ± 1.43     | 1.79 ± 0.79  | 0.133           | –               | –               | –               |
| ADMA (µmol/l)  | 0.63 ± 0.13 | 0.64 ± 0.09     | 0.77 ± 0.27  | 0.179           | –               | –               | –               |
| SDMA (µmol/l)  | 0.58 ± 0.12 | 0.57 ± 0.10     | 0.90 ± 0.64  | 0.012           | 0.959           | –               | 0.027           | 0.006           |
| hArg/ADMA      | 3.86 ± 1.54 | 3.48 ± 2.22     | 2.83 ± 2.14  | 0.061           | –               | –               | –               |
| hArg/SDMA      | 4.16 ± 1.44 | 4.17 ± 3.19     | 2.63 ± 1.78  | 0.017           | 0.629           | 0.078           | 0.005           |

Data are presented as mean ± standard deviation. Significant p-values are marked in bold. Results of the analysis of variance (ANOVA) are shown. If ANOVA was significant post-hoc group comparisons were performed. ADMA, asymmetric dimethylarginine; FD total, both Fabry disease groups combined; hArg, L-homoarginine; MMP-9, matrix metalloproteinase 9; SDMA, symmetric dimethylarginine; suPAR, soluble urokinase-type plasminogen activator receptor; VEGF, vascular endothelial growth factor. *Mean ± standard deviation of both FD groups combined (2 + 3) **p-value of t-test: 2 + 3 vs. 1.

FIGURE 2 | Box-plots of SDMA concentrations (A) and the ratio of L-homoarginine and SDMA (B) in 17 healthy controls, 7 Fabry patients without and 15 Fabry patients with FC. Box plots represent median, 25th and 75th percentiles. Whiskers indicate minimum and maximum without outliers and extremes. On the Y-axis concentrations of the biomarkers are presented (log scale). Brackets indicate p-value from the analysis of variance (ANOVA) with post-hoc group comparisons. n.s., not significant; SDMA, symmetric dimethylarginine.

hypothesis of an altered extracellular matrix (ECM) turnover in FD.

In patients with FC higher concentrations of SDMA and a decreased ratio of hArg/SDMA could be revealed compared to healthy controls as well as to FD patients without overt cardiomyopathy. These parameters correlate with the ventricular mass as well as with cardiac and renal markers suggesting a potential causal relationship of kidney function and cardiac disease progress.

**MMP-9, Angiostatin, and SuPAR**

MMP-9 is part of a family of endogenous zinc-dependent endopeptidases. In the myocardium MMPs play an important role for structural integrity of the ECM (18). In patients with familial hypertrophic cardiomyopathy (HCM) an association of MMP-9 with gadolinium enhancement in cardiac MRI was recently described and an important role of the MMP system in cardiac remodeling and fibrosis was proposed (26). In this context, Shah et al. identified significantly higher levels of MMP-9 in 29 FD patients compared to 21 healthy controls and hypothesized that MMP-9 plays an important role in the pathogenesis of FC and might be a valuable surrogate marker for the response to ERT (19). The findings from our study confirm the higher levels of MMP-9 in FD patients. However, a correlation of MMP-9 levels and Fabry associated cardiomyopathy cannot be confirmed.

MMP-9 cleaves matrix-bound plasminogen into angiostatin (27) which is a potent inhibitor of angiogenesis and has been shown to attenuate endothelial cell proliferation and migration (28). In FD the role of angiostatin has not yet been investigated. According to the elevated MMP-9 also Angiostatin concentrations increased in FD compared to controls and
showed a trend toward higher concentrations in the FC group compared to FD patients without FC.

Interestingly, a study from Matsunaga et al. could demonstrate that reduced NO production leads to an increase of MMP-2 and MMP-9 activity and higher angiostatin concentrations concluding that NO production influences coronary angiogenesis (29). Furthermore, an experimental study from Takahashi et al. revealed that angiostatin inhibits VEGF-stimulated NO production in human umbilical vein endothelial cells (30). In this context higher MMP-9 and angiostatin levels may contribute to an alteration of NO synthesis in FD. Moreover, an experimental animal study from Givvimani et al. found a switch to higher levels
of MMP-9 and anti-angiogenic markers such as angiotatin in the transition from compensatory hypertrophy to decompensated heart failure (31). Both, the effect of MMP-9 and angiotatin on NO synthesis and on a possible transition to decompensated heart failure in FC should be addressed in future studies.

Soluble uPAR is an important regulator of ECM proteolysis and is also involved in MMP activation by plasmin generation (32). For suPAR only a trend toward higher concentrations in FC could be determined, however, a positive correlation with the LV-mass could be found, as shown in Table 3. Soluble uPAR was shown to directly correlate with proteinuria (33). In this context a strong correlation to renal parameters and especially proteinuria (p < 0.001, r = 0.74) could be revealed.

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ADMA, SDMA, and hArg are non-proteinogenic amino acids structurally related to L-arginine. hArg has been shown to serve as an alternative substrate for NOS and to inhibit arginase. Thus, it is considered to increase NO formation (34, 35). In addition, low circulating concentrations of hArg have been proposed as a cardiovascular disease risk factor (36). ADMA on the other hand is an endogenous inhibitor of NOS (37), whereas its structural isomer SDMA does not directly interfere with NOS (38). However, SDMA inhibits the tubular L-arginine absorption in kidneys (39) and the y^+ transporter, which mediates the intracellular uptake of L-arginine (40). Therefore, SDMA has an indirect effect on NOS. Both dimethylarginines, ADMA and SDMA, are involved in endothelial dysfunction (41, 42), oxidative stress (43, 44), and atherosclerosis (45). A recent meta-analysis of prospective studies from Schlesinger et al. concluded that both markers are independently associated with cardiovascular disease and all-cause mortality (46).

In this context, the present study reveals a correlation of hArg, ADMA, and the ratio hArg/ADMA with the LV mass. Although group differences of ADMA and hArg showed a trend toward altered concentrations in FC patients, the effect was not significant. Moreover, the ratio of hArg/ADMA was superior compared to the single markers, but it equally missed significance in overall F-test of the analysis of variance with a p-value of 0.06.

However, in our cohort higher SDMA level in FC patients correlated with LV-mass, hsT, and lyso-Gb3 concentrations indicating the presence of endothelial dysfunction in these patients. Furthermore, this reveals a new possible mechanism of NO alteration in FD as this is the first study demonstrating higher SDMA level in FC patients’ sera. Similar results are shown for the hArg/SDMA ratio. Although the alteration of the hArg/SDMA ratio was statistically not superior compared to the higher SDMA concentrations in FC, group differences of this ratio show that hArg might also contribute to a dysfunction of NO synthesis in FC. Due to the antagonistic effects of hArg and SDMA in arginine metabolism a further investigation of this ratio might be reasonable.

The discrepancy between significantly higher SDMA levels without ADMA group differences might be explained by their disparate ways of excretion. SDMA elimination is exclusively renal, whereas ADMA is also enzymatically excreted by the dimethylarginine dimethylaminohydrolases (DDAHs). DDAH-1 for example is highly expressed in the kidney and liver (47) and as FD patients do not typically show liver dysfunction enzymatic ADMA elimination might be sufficient, whereas SDMA accumulates due to the typical early occurring renal insufficiency in FD. In this regard, adjustment for eGFR showed a significant dependency of SDMA and hArg/SDMA to kidney function. Furthermore, an association of eGFR and FC could be revealed. It is well known that diastolic dysfunction caused by impaired LV relaxation may lead to congestive heart failure and consequently to renal insufficiency. In our cohort only 4 patients with FC presented with mild (grade 1) and 1 patient with moderate (grad 2) diastolic dysfunction. Thus, based on these findings one might speculate that renal insufficiency might rather contribute to FC due to an accumulation of SDMA and its negative effect on NO synthesis in the vascular system. However, a causal relationship between renal and cardiac function in Fabry disease has to be proven in further studies.

### CONCLUSIONS

This study provides evidence for an altered ECM turnover with higher levels of MMP-9 and angiotatin in FD patients independent of an existing FC. Moreover, patients with FC showed higher SDMA and hArg/SDMA level, which correlated with LV mass, hsT, and lyso-Gb3 concentration but also with impaired renal function. Renal and cardiac function showed a relationship leading to the hypothesis that accumulation of SDMA due to renal insufficiency in FD might contribute to the development of endothelial dysfunction and subsequently lead to FC.

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**TABLE 3 | Correlation to indexed left ventricular mass measured in cMRI.**

| Marker       | p-Value | r     |
|--------------|---------|-------|
| SDMA (µmol/l) | 0.003   | 0.61  |
| ADMA (µmol/l) | 0.031   | 0.47  |
| hArg (µmol/l) | 0.120   | −0.35 |
| hArg/SDMA    | 0.011   | −0.54 |
| hArg/ADMA    | 0.032   | −0.47 |
| suPAR (ng/ml) | 0.002   | 0.64  |
| hsT (pg/ml)   | < 0.001 | 0.78  |
| proBNP (pg/ml)| 0.002   | 0.65  |
| lyso-Gb3 (ng/ml)| 0.026  | 0.50  |
| eGFR (ml/min/1.73 m^2) | 0.037 | −0.46 |
| Creatinine (mg/dl) | 0.014 | 0.53  |
| ACR (mg/g)    | 0.018   | 0.51  |

Results from the correlation tests of laboratory values and markers of endothelial dysfunction correlating with indexed left ventricular mass. ACR, albumin-to-creatinine ratio; ADMA, asymmetric dimethylarginine; eGFR, estimated glomerular filtration rate using the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) formula; hArg, L-homoarginine; hsT, high sensitive cardiac Troponin T; lyso-Gb3, Globotriaosylsphingosine; proBNP, prohormone of brain natriuretic peptide; r, Pearson’s correlation coefficient; SDMA, symmetric dimethylarginine; suPAR, soluble urokinase-type plasminogen activator receptor.

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Limitations
A major limitation of this study is the small sample size of 22 investigated FD patients with only 7 patients without FC compared to more than twice as much patients with FC. Another limitation is the unequal distribution of male and female patients in the FD groups and the fact that this study is a single center study. Moreover, whether the relationship between kidney and cardiac function is causal cannot be proven by this study design.

Perspectives
To validate these findings multicenter studies including more patients to investigate the effects of MMP-9 and angiotatin on endothelial dysfunction in FD and to clarify the pathological impact of SDMA accumulation in Fabry associated cardiomyopathy are required.

ETHICS STATEMENT
This study was carried out in accordance with the principles outlined in the Declaration of Helsinki. The protocol was approved by the local ethics committee of the Ärztekammer Hamburg, Germany (approval no: PV-4056).

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AUTHOR CONTRIBUTIONS
JL: writing of the manuscript, experimental work. NL: experimental work, writing assistance. MA: MRI data acquisition, critical review of the data. NM: acquisition of clinical data, critical manuscript review. SL: statistical analysis. KC and ES: supervision and design of experiments, critical review of data analysis and manuscript. MP: experimental design, supervision of data analysis. writing of the manuscript.

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