3-D imaging reveals four extraordinary cases of convergent evolution of acoustic communication in crickets and allies (Insecta)

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When the same complex trait is exhibited by closely related species, a single evolutionary origin is frequently invoked. The complex stridulatory apparatus present in the forewings of extant crickets, mole crickets, katydids, and prophalangopsids, is currently interpreted as sharing a single common origin due to their similarity and unique function. An alternative hypothesis of convergent evolution in these ensiferan groups has challenged this common view, but remained controversial because of competing interpretations of wing venation. Here we propose another hypothesis for the widely and long debated homology of ensiferan stridulatory apparatus, performing the first 3D reconstruction of hidden structures at the wing bases. This approach allowed defining the homology of each vein from its very origin rather than after its more distal characteristics, which may be subjected to environmental pressure of selection. The stridulatory apparatus involves different veins in these four singing clades. In light of the most recent phylogenetic evidence, this apparatus developed four times in Ensifera, illustrating extraordinary convergent evolutions between closely related clades, by far exceeding the number of evolutionary steps ever proposed for calling ability in this group.

The Orthoptera Ensifera are well known for their capacity to emit and perceive sounds, having developed a high diversity of morphological structures and behaviors not commensurate with other animal groups¹. Their sounds are produced by the friction between two sclerotized parts of the body (=stridulation), in two main modalities: a) wing stridulation, i.e., the friction between the two forewings (or tegmina), in crickets, propha팡opsids, katydids, and mole crickets; b) femoro-abdominal stridulation, i.e., the friction of a hind femora against the adjacent abdominal tergites²,³. Vibratory or acoustic communication can be found in representatives of all the currently recognized ensiferan families. Yet efficient sound production by wing stridulation is amazingly complex needing several crucial biophysical and neurological couplings, this fact being counter-intuitive for convergence.

Ensiferan wing stridulation is based on intricate anatomical structures located on the veins of the basal part of the tegmina (venation terminology in Supporting Information). When the tegmina come to overlap by a lateral movement of opening - closing, the ventral face of one tegmen, bearing a row of teeth called the file, rubs the thickened rim (scraper) of the other tegmen¹. The generated vibrations are amplified by a resonator located very close to the file and the scraper: a circular area (mirror) in katydids (Tettigonioidea), a triangular harp sustained by parallel and generally oblique veins in mole crickets (Gryllotalpoidea, Gryllotalpidae) and crickets (Gryllidea), or a variation of large wing cells in prophalangopsids (Hagloidea, Prophalangopsidae). The
resonator multiplies the frequency and amplifies the intensity of the signal, and the vibrations propagated along the main veins. In crickets, the resonator properties fit the closing speed of the wings, and both fit the dominant frequency of the emitted song, resulting in musical signals. *Cyphoderris* emits a highly-tuned signal, which contrasts with the signal most often broad-banded with a large ultrasonic part emitted by most katydids. In mole crickets, the efficiency of sound production increases when the male sings from inside its burrow. The main controversy regarding acoustic origins concerns the homology of acoustic apparatus, which can be approximate through the nature of the file, carried by veins whose homology is discussed. These studies lead to two alternative hypotheses for file location, on posterior cubital or first anal veins. The last study concluded in favor of a single origin of acoustic communication on the basis of the hypothesis that the file is located on CuP and homologous in all singing Ensifera.

Most recent attempts of establishment of the homology of these structures were based on the relative positions and relative convexity vs. concavity of the concerned cubital and anal veins, without exams of the extreme bases of the veins and their departures from the basivenal sclerites. The identity of each vein can be more accurately determined by examining from where it begins than from its relative position and relative convexity in distal parts, which are more subject to environmental pressure of selection. Traditionally the long and thin organs that constitute insect wings are intuitively simplified as a 2D structure and examined using optical microscopy. This technique is unable to separate strongly approximate or touching veins at the very base of the wings, as are the CuP and A1. Here we examine the relative positions of the veins in 3D space using X-ray microtomography (XMT): 3D modeling of venation, from the extreme bases of the veins to the functional acoustic apparatus, helps following each vein individually from its emergence from a basivenal sclerite, but also potential vein anastomoses, fusions and separations. We explored the diversity of wing venations in singing Ensifera (see Suppl. Table 1), to establish the homology of the stridulatory structures in crickets, mole crickets, prophantopods, and katydids.

For a better comparison with the last studies on the topic, we use the same nomenclature of wing venation elaborated by Béthoux & Ne (adapted with slight modifications by Béthoux in subsequent papers). Corresponding abbreviations are: CuPa = anterior branch of CuP; CuPα = anterior branch of CuPα; CuPβ = posterior branch of CuP; CuPb = posterior branch of CuP; first anal vein = A1. Abbreviation for the other veins and color codes are listed in the Supporting Information.

XMT reconstructions show how the stridulatory file of the Gryllidea is on A1 (Fig. 1a–e, Supporting Information), while it is on CuPb in the Gryllotalpidae (Fig. 1f–k, Supporting Information). Consequently the triangular harp is an area between A1 and the most anterior branch CuP of CuP in crickets, unlike the hypothesis of Béthoux, where it is an area between the two branches CuPα and CuPβ in mole crickets, in accordance with Béthoux. Thus, the stridulatory files and harps of crickets and mole crickets are not homologous (Ref. 2, contra 5,12), while they are currently considered as sister groups. Béthoux proposed that in mole crickets CuA is separating from M very basally and fused with CuPα into a very long CuA + CuPα, unlike in Gryllidea. We recover a strong transverse vein between M + CuA and CuPα in the mole cricket that could correspond to the CuA.

The Gryllidea are characterized by a very weak (sometimes completely reduced, especially in Gryllinae. s. str.) CuPb vanishing in area between CuPα and A1. Béthoux considered this vein as a sclerotization located between CuPα and CuPb, but this vein is clearly emerging from the cubital basivenal sclerite in all the Gryllidea we examined (see for example *Oecanthus* sp., where it is especially strong: Fig. 1a,b,d, suppl. movies 1,2). This vein is not homologous to the secondary structure present in *Gryllotalpa* between CuA + CuP and CuPb, which is not emerging from a basivenal sclerite (Fig. 1f–k, suppl. movies 3,4). In the same line of evidence, the side wing of the Jurassic *Liassophyllum caii* Gu & Ren, 2012 (currently in Haglidae: Cyrtophyllitinae) shows a short but ending in a node of veins situated at the posterior end of the file.

The Gryllidea are interpreted as of composite origin (CuPα, column, and handle) by Béthoux and Chivers et al. Furthermore in the katydids, CuPb is absent while the basal part of CuPα is weak and concave, and runs between the strongly convex A1 and the convex M + CuA. Therefore, even if the katydid and gryllid stridulatory files are on the vein A1, their whole stridulatory apparatus are not homologous.

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Figure 1. Wing venation interpretations. (a–e) Gryllidae (Oecanthus sp.). (a) 3D modeling of wing venation using XMT, view from above. (b) 3D modeling of wing venation using XMT, view from below. (c) General habitus (copyright S. Hugel). (d) Complete tegmen (copyright L. Desutter-Grandcolas). (e) Cut xy (1 colored; 2 without color; orientation of tomogram plane YZ, tomogram number 1035). (f–k) Gryllotalpidae (Scapteriscus sp.). (f) 3D modeling of wing venation using XMT, view from above. (g) 3D modeling of wing venation using XMT, view from below. (h) General habitus (copyright S. Hugel). (i) Complete tegmen (copyright L. Desutter-Grandcolas). (j) Cut uv (1 colored; 2 without color; orientation of tomogram plane XZ, tomogram number 1043). (k) Cut st (1 colored; 2 without color; orientation of tomogram plane XZ, tomogram number 876). (3D XMT L. Jacquelin).
From a functional point of view, the file has to be situated on a strong relief of the ventral side of the tegmen to rub the other tegmen. Prophalangopsids and gryllotalpids have easily ‘solved’ the problem as the file is located on the ventral side of the concave vein CuPb (thus highly protruding on the ventral side of the wing and in a position to hit the scraper). In the grylloids, the file is on the distal part of vein A1, which is concave at the file level but convex basally (and thus changes its convexity, rendering its homology difficult to interpret on the sole criteria of the position and convexity of the vein). In the katydids, A1 is convex but strongly flattened so that its ventral part bearing the file is ventrally protruding.

According to XMT results, a stridulatory file has evolved convergently in Ensifera, on the cubitus posterior vein of the mole crickets and prophalangopsids, and on the anal vein in crickets and katydids (Fig. 4), with different venation patterns each time. Following the phylogeny proposed by Song et al., with the crickets and mole crickets within Gryllidea, and the katydids and modern prophalangopsids within Tettigoniidea, the acoustic communication should have evolved independently at least four times in Ensifera. Our inference of convergence is strong enough compared to other homoplastic patterns with difficult probabilistic inference: we demonstrate differences between wings that cannot be re-interpreted as a common ancestral pattern depending on their situation on a tree. Interestingly, the Permian Permostridulidae (Palaeozoic orthopteroid order Caloneurodea) have a stridulatory file on a supplementary longitudinal veinlet between CuPa and CuPb (Supplementary Fig. 2). It represents a fifth type of acoustic apparatus developed on the forewings of an Archaeorthoptera. It is commonly accepted that acoustic communication is counter-selected by strong predation. It can then be supposed that each emergence of acoustic communication could be related to a ‘low-predation window’, i.e. a period with less intense predation pressure.

From an evolutionary and functional point of view, because of the general pattern of convergences for stridulation put in evidence here in Ensifera, the functional parameters that govern the properties of emitted signals cannot be analyzed between ensiferan clades as deriving from supposedly ancestral groups to supposedly more apical ones, which would remind a gradist approach: each clade evolved on its own under relatively similar functional constraints. This pattern invalidates the concept of ‘Grylloptera’ sensu, a ‘formal taxon encompassing ensiferans possessing a ‘file’ and a paraphyletic assemblage proposed without taking into account the actual diversity of the Ensifera. The only way to hypothesize a plesiomorphic condition for the whole Ensifera will be to perform a phylogenetic analysis including both extant and fossil species, both mute and acoustic, with well-attested homology hypotheses.

The majority of the Mesozoic singing Ensifera are currently attributed to fossil families (e.g., Mesoedischiidae Gorochov, 1987, Hagldae Handlirsch, 1906) or to the Prophalangopsidae, without strong supporting synapomorphies. Our new hypothesis for the homologies of the singing apparatus provides a new frame to reconsider their relationships. For instance, the Triassic Mesoedischia obliqua Gorochov, 1987 or Termitidium ignotum Westwood, 1854 could have a file patterns of tettigonioid type (see Fig. 5a). The recent description of a leaf-mimicking Tettigonioidea in the Middle Permian demonstrates the antiquity of this clade contr. The grylloid type is present in an undescribed Liassic fossil from Grand-Duché de Luxembourg (see Fig. 5b, Suppl. Fig. 3).
Figure 3. Wing venation interpretations. (a–i) Prophalangopsidae (*Cyphoderris monstrosa*). (a) 3D modeling of wing venation using XMT, view from above. (b) 3D modeling of wing venation using XMT, view from below. (c) General habitus (copyright and thanks to D. Gwynne for authorization to use). (d) Complete tegmen (copyright L. Desutter-Grandcolas). (e) Cut xy (1 colored; 2 without color; orientation of tomogram plane XY, tomogram number 1159). (f) Cut uv (1 colored; 2 without color; orientation of tomogram plane XY, tomogram number 1131). (g) Cut rs (1 colored; 2 without color; orientation of tomogram plane XY, tomogram number 1134). (h) Cut mm (1 colored; 2 without color; orientation of tomogram plane XY, tomogram number 1138). (i) Cut op (1 colored; 2 without color; orientation of tomogram plane XY, tomogram number 1112). (j–o) Tettigonioidae (*Quiva* sp.). (j) 3D modeling of wing venation using XMT, view from above. (k) 3D modeling of wing venation using XMT, view from below. (l) General habitus (copyright and thanks to P.S. Padron for authorization to use). (m) Complete tegmen (copyright L. Desutter-Grandcolas). (n) Cut xy (1 colored; 2 without color; orientation of tomogram plane XZ, tomogram number 130). (o) Cut uv (1 colored; 2 without color; orientation of tomogram plane XZ, tomogram number 207). (p) Cut rs (1 colored; 2 without color; orientation of tomogram plane XZ, tomogram number 223). (q) Cut mm (1 colored; 2 without color; orientation of tomogram plane XZ, tomogram number 232). (r) Cut op (1 colored; 2 without color; orientation of tomogram plane XZ, tomogram number 235). (3D XMT L. Jacquelin).
**Figure 4.** Molecular phylogeny of Ensifera (after Song et al.12). The four singing groups are indicated by photographs of representatives and the reconstructions of the cubito-anal fields (arrows indicate the fields) (copyrights of photographs S. Hugel, D. Gwynne, P.S. Padron).

**Figure 5.** Wing venation interpretations. (a) *Mesodeschius obliqua* Gorochov, 1987, holotype PIN 2240/4074, Triassic, Kyrgyzstan, photograph (copyright A. Rasnitsyn); for the reconstruction of tegmen see ref. 28. (b) Reconstruction of tegmen of a Grylloidea indet., MNHN.F.A57514, Torcian, Bascharage, Grand-Duché du Luxembourg (copyright A. Nel).
Beyond the necessary reanalysis of the venation of the other modern and fossil Ensifera in the light of the paradigm we validate here, several questions need to be addressed in the next future.

First, what are the molecular mechanisms underlying diversified wing venation in Ensifera? Second, are the same genomic regions responsible for more than one wing venation pattern? Third, are the same genes responsible for the same wing venation pattern for all the Ensifera? Turning on and off a key set of genes may have resulted in different wing patterns, by alteration of their expression regulatory pathway25, supporting the hypothesis that repeated reversals or convergences of characters are likely in the evolution of Ensifera. In this context, the exploration of genes and their expression by means of high throughput sequencing methods would enlighten us on the reversible and irreversible genetic changes driving morphogenesis in insects, over long periods of evolutionary time26, 27. In responding to questions such as these, the new framework we present here for understanding Ensifera acoustics paves the way for a more detailed understanding of the processes giving rise to convergent evolutionary outcomes.

Methods

Materials. The studied specimens are housed at the Entomology Department of the Muséum national d’Histoire naturelle, Paris, France. The inventory numbers are detailed in Suppl. Table 2, see MNHN collection data base at [https://science.mnhn.fr/institution/mnhn/collection/eo/]

Imaging. The four extant specimens were imaged under X-ray, with phase contrast, at the microtomograph of the University of Poitiers.

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Author Contributions
L.D.-G. and L.J. are first authors with equal ranks, L.J., R.G., L.D.-G., and A.N. performed research. R.G., L.J., and R.B. contributed new analytic tools. L.J., S.H., M.H., J.I.C.-M., L.D.-G., P.N. and A.N. analyzed data. L.J., B.H.W., L.D.-G., P.G., and A.N. wrote the paper with inputs from all coauthors. L.D.-G. and A.N. designed the program. P.G. and A.N. are last authors with equal rank.

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