Capsule Formulation Essay of Herbal Extracts of Trunk Bark of Anogeissus leiocarpus (DC) Guill. Et Perr. (Combretaceae) for the Treatment of Hypertension

Ouedraogo Salfo¹, Sombie C. Bavouma², Diawara Zime Hermine², Yameogo B.G. Josias²,³, Traore Tata Kadiatu²,³*, Nitiema Mathieu¹, Belemnaba Lazare¹, Ouedraogo Sylvain², Smde Rasmané²

INTRODUCTION

The African continent is at a crossroads as it faces an unprecedented epidemic of cardiovascular disease. Indeed, there are millions of hypertensives in Africa, and the proportion of people affected is expected to increase by 2025.¹

The burden of cardiovascular disease is overwhelming in Africa, that it has become a public health problem throughout the African Region.²

Given increasing prevalence of hypertension and the complexity of this disease, modern therapy offers a wide range of antihypertensive drugs.³ There are emergency forms, usually administered in injectable form, and ambulatory forms. Unfortunately, the costs of these drugs are often beyond the reach of Third World populations.³ This leads a large population segment to treat themselves with plant and animal-based medicines transmitted from generation to generation.⁴,⁵ In recent years, there has been an increase in research on antihypertensive medicinal plants used by traditional practitioners that show some effectiveness.⁶ Among these plants, we note Anogeissus leiocarpus which has been the subject of numerous studies (ethnobotanical, phytochemical, pharmacological, toxicological).⁷,⁸,⁹,10,11,12 Despite these studies, no adapted galenic form that would facilitate its use is available. The literature indicates that herbal medicines range from simple forms, herbal teas, capsules containing plant powders, to more elaborate arrangements forms in which a purified plant extract or a pure molecule isolated from a plant enters as the "active" of the medicine, then designated as a phyto-drug.¹³ The present aim to develop a pharmaceutical grade capsule formulation from the freeze-dried aqueous extracts of the trunk bark of Anogeissus leiocarpus.

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1. Département de médecine et pharmacopée traditionnelles-pharmacie (MEPHATRA-PH), Institut de recherche en science de la santé (IRSS/CNRST), 03 BP 7047 Ouaga 03, Burkina Faso.
2. Laboratoire du Développement du Médicament (LADME), Ecole doctorale de la santé, Université Joseph Ki-Zerbo, 03 BP 7021 Ouaga 03, Burkina Faso.
3. Laboratoire National de Santé Publique, Ministère de la santé, 09 BP 24 Ouagadougou 09, Burkina Faso.
MATERIALS AND METHODS

Materials

Plant’s materials

Fresh trunks barks of Anogeissus leiocarpus (Combretaceae) were harvested at Loumbila commune (Burkina Faso) and identified by a botanist from the ecology laboratory of the University of Joseph Ki-ZERBO in reference to the herbarium N° 1544. The barks have been dried and crushed to powder.

Excipients and packaging materials

The galenic form studied here was a dry oral formulation with conventional release. The active ingredient was some lyophilised aqueous extracts of Anogeissus leiocarpus trunks bark powders. Different types of excipients and packaging materials have been used. Corn starches (Batch 03406) were provided by CERESTAR GL, Germany. Magnesium stearate (Batch 14.153.874), Polyvinylpyrrolidone K30 (Batch 14G04-B05-290549) were purchased from FAGRON (Belgium). Empty capsules (Batch 51532812) were provided by CAPSUGEL, (Belgium). Food bag (Batch 03550100) was obtained from BUERKLE GMBH, (Germany). Ethanol was obtained from CAMEG (Burkina Faso). Distilled water was prepared in the galenic laboratory (IRSS).

Methods

Preparation of extracts and preliminary phytochemical caractérisation of the extract

Hundred (100) grams of vegetable powder have been boiled with 1L sterile distilled water while 30 minutes. The extracts obtained were frozen for 24 hours and then lyophilized at 96 hours. These extracts were analysed using different methods, such as macroscopic examination of botanical characteristics. The determination of the organoleptic characteristics consisted of powders observation (touching, smelling and tasting). The residual moisture content of the extracts were determined using the thermogravimetric method. One (01) g is weighed in triplicate and placed in a previously tared watch glass in an infrared moisture Analyser (MF-50, US).

Chemical screening with TLC

Phytochemical screening was performed on chromatoplates (60 F254, 20 x 20 cm glass support, Fluka -Silica gel) following methods described in the literature. The main chemical groups were searched by thin layer chromatography (TLC) such as steroidal compounds, terpene compounds, phenolic compounds and alkaloidal compounds. The dry sample was solubilised in its extraction solvent at the concentration of 10 mg/mL (10 mg in 1 mL of solvent) and 5 µL were deposited on the TLC plate for chromatogram development.

Table 1: The composition of granules (amounts of the substances are given in parts)

| Formule/ gram            | F1   | F2   | F3   |
|--------------------------|------|------|------|
| Polyvinylpyrrolidone K25 (PVP) | 0    | 0.28 | 0.12 |
| Magnesium stearate       | 0.12 | 0.12 | 0.12 |
| Extract                  | qs   | qs   | qs   |
| Corn starch              | 11.12| 10.9 | 11   |

qs*: Sufficient quantity to make

Determination of phenolic compounds by UV-spectrophotometric method

The amount of total phenolics in the drug retained from the plant was estimated by the Singleton method, using the Folin-Ciocalteu reagent (FCR). FCR is a mixture of phosphotungstic acid and phosphomolydbic acid, which is reduced during the oxidation of phenols in alkaline medium to a mixture of blue oxides of tungsten and molybdenum. The latter show an absorption maximum at the wavelength of 760 nm, with an intensity proportional to the quantity of polyphenols present in the sample.

The reaction mixture consisted of 25µL of the 0.1 mg/mL sample, 105 µL of 0.2N FCR which was left to incubate for five minutes in the dark. To this mixture was added 100 µL of a sodium carbonate solution (75 g/L in distilled water). The mixture was then left to incubate for one (1) hour in the dark and then the absorbance was measured at 760 nm wavelength in a spectrophotometer against a gallic acid standard curve.

The tests were performed four times, and the results expressed as grams of gallic acid equivalent per 100 g of dry sample (mg EAT/100 g).

The total phenolic content of the extract was obtained by the formula:

\[ T = \frac{c \times D}{Ci} \times 100 \]

Where:
- \( T \) = Content in mg Gallic Acid Equivalent in 100 g extract
- \( c \) = sample concentration read (µg EAT/mL) on the standard curve
- \( D \) = Dilution factor of the sample under assay
- \( Ci \) = initial concentration of the sample solution to be determined

Preparation of granules

The granules were prepared by the wet granulation method. It was carried out using a GLA-ORY FREWITT type granulator. The mass to be granulated was weighed, mixed and wetted with ethanol until a crumbly paste was obtained. The extract was mixed with corn starch and Polyvinylpyrrolidone K25 (PVP) as binding agents are added till it produces coherent mass a then required quantity of magnesium stearate using ethanol as wetting liquid was added. The set was passed through an oscillating granulator with a 125 mesh sieve. Granules obtained were gently spread and dried in a MIGHTERT oven at a temperature of 45 °C for 12 hours. Dry granules were weighed and their weight was recorded. The dried granules were mixed with magnesium stearate were added in required quantities.
Evaluation of granules

The macroscopic and organoleptic characteristics and the residual moisture content were carried out according to the methods mentioned above.

Prepared granules were subjected for determination of bulk density, tapped density, Hausner’s ratio, Carr’s index, angle of repose and bulkiness to assess granule’s the flow property.\textsuperscript{20,21}

Fabrication and quality control of capsules

Prepared granules were packed into hard gelatin capsule (size 3) using semi-automatic capsule filler machine.

Determination of uniformity of weight

This test was performed according to European Pharmacopoeia 2.9.5.\textsuperscript{22} Twenty (20) capsules were randomly selected and individually weighed using a precision balance (Sartorius, France). The full capsules were individually weighed and then carefully emptied of their contents with a swab. The empty capsules were then individually weighed to determine the difference between the contents of the capsules. The average mass (M), mass deviations, in percent \[100 \text{ (mi - M)/M}\] from the average mass and coefficient of variation (CV) were calculated. These deviations were compared to the tolerated limits specified in the pharmacopoeia.\textsuperscript{22}

Disintegration time of capsules

This test involved 6 capsules taken at random and introduced individually into the tubes of a disintegrating device. A disc was placed on each capsule, and the disintegrating medium used was distilled water maintained at 37°C. The time of complete disintegration of the first capsule and that of the last one was noted.\textsuperscript{22}

Determination of total phenolics as a tracer

The amount of total phenolics in the plant extracts was estimated by the Singleton method using the Folin-Ciocalteu reagent (FCR).\textsuperscript{19} The reaction mixture consisted of 25µL of extract at 0.1mg /mL, 105µL of 0.2N RCF, which was incubated for 5 min in the dark. To this mixture was added 100µL of a sodium carbonate solution (75g/L in distilled water). It was incubated for 01 h in the dark and then the absorbance was measured at the wavelength of 760 nm with a spectrophotometer against a gallic acid standard curve. The tests were carried out four times on each sample, and the results are expressed in micrograms of gallic acid equivalent per milligram of dry extract (µg GAE / mg extract).

Uniformity of total phenolic content of capsules

A sample of 10 capsules was taken at random. Each capsule was weighed and emptied of its contents and macerated individually in water for 10 min by magnetic agitation in a beaker containing 50 ml of distilled water at 25°C. The determination was carried out according to method described by Singleton et al.\textsuperscript{19} and interpretation according to the European Pharmacopoeia 10th edition 2.9.6.\textsuperscript{22}

RESULTS AND DISCUSSION

Preliminary phytochemical analysis of extract

The freeze-dried extract of \textit{Anogeissus leiocarpus} (Combretaceae) was brown (Figure 1) with astrigent odor and slightly bitter flavor. The extraction yield was 26.77 ± 2.86, and the residual moisture content (THR) was 4.43±0.49%.

Figure 1: Freeze-dried extract of \textit{Anogeissus leiocarpus}

The chromatographic analysis revealed the presence of several chemical groups represented by figure 2. This figure could be used as a chromatographic fingerprint. The blue fluorescence after spraying with NEU reagent characterises the presence of phenolic compounds in the extracts. These compounds could be used as tracers for the quality control of the extract and finished products. Indeed, chromatographic techniques allow a quantitative and qualitative analysis. To facilitate the quality control of raw materials and plant-based medicines, the European Medicines Agency (EMA) proposes an approach based on the determination of the chemical fingerprint of these phytomedicines and the monitoring of chemical tracers.\textsuperscript{23,24}

Figure 2: TLC of the hexane fractions of the freeze-dried extracts after daylight (A) and UV observation at 254 nm (B).

The determination of phenolic compounds chosen as tracer to give a value of 1.71899± 0.0426 µg EAG/mg extract. The calibration curve of the gallic acid (Figure 3) obtained has the equation: \[ y = 29.07x + 0.0202 \quad (R^2 = 0.999) \] This curve was used for the determination of gallic acid in the samples.

Figure 3: Standard line obtained with gallic acid

Preparation and evaluation of granules

The pellets were prepared from the extract using various excipients at different concentrations by the wet granulation method. The pellets were brown (Figure 4) with an astrigent odor and slightly bitter flavor. This indicates retention of the extract characteristics. The residual moisture content (RMC) was 6.21±0.32%.
Table I: Pharmacotechnical characteristics of granules

| Designations | Bulk density (g/ml) | Tapped density (g/ml) | Hausner ratio | Carr’s index (%) | Angle of repose (°) | Bulkiness |
|--------------|-------------------|----------------------|---------------|-----------------|-------------------|-----------|
| F1           | 0.8±0.02          | 0.96±0.05            | 1.14±0.01     | 12.50±0.05      | 27.1±0.03         | 1.37±0.04 |
| F2           | 0.89±0.01         | 0.99±0.01            | 1.11±0.04     | 10.10±0.03      | 23.7±0.05         | 1.43±0.02 |
| F3           | 0.81±0.03         | 0.91±0.03            | 1.12±0.02     | 10.99±0.02      | 24.4±0.07         | 1.32±0.05 |

Formulation and evaluation of capsules

All capsule formulations were subjected to various evaluations, and the results are shown in Table 2.

The capsules manufactured were ivory in color and size 3. (Figure 5).

Concerning mass variation, the capsules complied with the standards accepted by the 10th edition of the European Pharmacopoeia, which stipulates that the mass of the capsules may vary up to 10% for capsules of less than 300 mg and 7.5% for weights above 300 mg.22 The capsules analysed had mean masses of 123.5±0.004 mg to 128.4±0.0053 mg, with coefficients of variation ranging from 2.17 to 3.64 (Table II). Thus, none of the analysed capsules containing powder mixtures exceeded the acceptable range.

This test of uniformity of mass of the capsules conforming with the pharmacopoeia standards presumes that the quantity of active extract by capsule would be thus homogeneous and would make it possible to avoid any variability of the administered dose. As for the disintegration time, the formulations without binding agent disintegrated faster than the others. This finding confirmed a correlation between capsule qualities and the physicochemical properties of the excipients. However, the F2 formulation had the highest disintegration time. It was above the standards accepted by the 10th edition of the European Pharmacopoeia, stipulating that this value should be less than 15 minutes.

The disintegration test of the capsules showed an acceptable disintegration for the capsules of formulations F1 and F3 according to the 10th edition of the European Pharmacopoeia. The capsules disintegrated in 11 minutes for F1, 15 minutes for F2 and 13 minutes for F3. Formulations F1 and F3 were then used for further studies concerning the uniformity of tracer content (phenolic compounds).

Table II: Mass uniformity and disintegration time

| Designations | Weight variation (mg) | Disintegration Test (mins) |
|--------------|-----------------------|---------------------------|
| F1           | 123.5±0.004           | 11.18±0.8                 |
| F2           | 128.4±0.0053          | 15.36±0.3                 |
| F3           | 127±0.0045            | 13.21±0.7                 |

The uniformity of phenolic compounds content retained as the tracer is represented in Tables III and IV. The phenolic compound content of 10 capsules randomly taken from each formulation was 0.039±0.0097 mg GSE/capsule for F1 and 0.059±0.0063 for F3. From these results, both formulations are within the standards of the European Pharmacopoeia 10 ed. The latter states that the preparation passes the test if the individual content of up to one unit is outside the limits of 85 per cent to 115 per cent of the mean content and if it is not outside the boundaries of 75 per cent to 125 per cent of the mean content.22
**CONCLUSIONS**

This study produced capsule formulations based on freeze-dried aqueous extract of *Anogeissus leiocarpus* for the treatment of hypertension. These formulations were made through the addition of excipients. The use of wet granulation of the mixtures in the form of granules followed by a filling of the capsule made it possible to obtain a galenic form meeting the recommendations of the pharmaceutical standards. The pharmaceutics characteristics follow the European Pharmacopeia 10th ed specifications for the F1 and F3 formulations. These forms could serve as alternatives for the administration of the extracts of trunks barks of the plant. They could also improve the quality of treatment and compliance through ease of administration and storage. Further stability studies should be considered.

**CONFLICT OF INTEREST**

The author has no conflict of interest.

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