Experimental studies to evaluate the immunomodulatory and anti-inflammatory potential of optimized polyherbal preparations in experimental model of asthma

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Abstract

The study assessed the effect of standardized optimized formulation of UNIM-352, a polyherbal preparation used in the traditional Unani system of medicine in experimental model of bronchial asthma. Wistar Rats (200-250g) were sensitized with ovalbumin adsorbed to aluminium hydroxide. After sensitization, rats were treated orally for 14 days with either of UNIM-352, optimized preparation of UNIM -352 (OP-1, OP-2, OP-3, OP-4) or positive control, and prednisolone in separate groups. The rats were challenged with OVA on 14th day and after 24 h of challenge, they were anesthetized and bronchoalveolar lavage (BAL) were collected for the assays of IL-4, IgE, eosinophils and neutrophils cell counts. The results showed that both doses of UNIM-352 significantly reduced the levels of IgE and IL-4 in BAL fluid as compared to OVA sensitized and challenged control group. Similarly, both doses OP 1 and OP 3 treated rat significantly reduced levels of IL-4 which was comparable to UNIM-352 in dose dependent manner. Further, optimized preparations of UNIM-352 (OP-1, OP-2, OP-3 and OP-4) at 200 and 400 mg/kg showed reduced number of eosinophils and neutrophils in BAL fluid vs control group. The results were comparable with the standard drug; prednisolone. UNIM-352 and its optimized preparations differentially reduced the eosinophils and neutrophils counts, the effector inflammatory cells in asthma, which was accompanied with the reduction in levels of IgE and IL-4 in BAL fluid suggesting their anti-inflammatory and immunomodulatory effects. This was comparable to the positive control, prednisolone in experimental model of asthma. This relative potential of various optimized preparations of this polyherbal drug in reducing/controlling the airway inflammation associated with bronchial asthma has been discussed.

Introduction

Asthma is a chronic inflammatory disease defined by airway inflammation, reversible airway obstruction and airway hyper-responsiveness [1]. The inflammation of airway involves the activation of inflammatory and structural cells responsible for synthesis and release of inflammatory mediators which are responsible for the pathophysiology of asthma. Activation of mast cells, macrophages, eosinophil and neutrophil leads to release of pro-inflammatory cytokines (TNF-α), Th2 type cytokines (IL-4) and IgE antibody production. Activated inflammatory cells like macrophages and eosinophil also produce reactive oxygen species (ROS) leading to oxidative stress [2-4].

Bronchodilators (β2 agonist) and anti-inflammatory agents (corticosteroids) are the drug of choice for the treatment of asthma. Corticosteroids have long been used as the main therapeutic drugs for asthma [5]. However, the limitation of available pharmacotherapy is that these drugs provide only symptomatic treatment which is usually accompanied with adverse effects. Additionally, prolonged use of corticosteroids may result in steroid resistance [6]. Hence, phytopharmaceuticals and traditional herbal preparation could be a choice which possesses potential to treat such conditions with minimal side effects.

Traditional herbal medicine has a history of thousands of years and is still widely used. The evolution of traditional herbal medicine is based on observations and practical experience accumulated from millions of practitioners for thousands of years. Since traditional herbal medicine does not have its own history of scientific development, there is little scientific research conducted on the safety of herbal formulation. Herbal medicines have often been provided to humans without scientifically rigorous toxicity testing [7-12]. The herbal drugs used in Indian traditional system of medicine viz. Ayurveda, Unani, Homeopathy, Siddha possess a great potential to cure various disease. These herbal drugs are considered to be relatively safe because of natural origin. UNIM-352 is a polyherbal Unani formulation, which has been clinically used in India for the treatment of bronchial asthma [13,14]. UNIM-352 contains six different herbal ingredients and all are well known traditional medicines, viz. Linum usitatissimum, Trigonella foenum-graecum, Allium sativum, Strychnos potatorum, Caesalpinia bonducella, and Pongamia glabra. Previous studies showed that UNIM-352 has anti-inflammatory, immunomodulatory effects in the experimental model of bronchial asthma [15].

Polyherbal formulations without any upper limit of ingredients are allowed only in some countries, whereas, in Europe, US etc. formulations with up to 3-4 plants only can be used. So, this effective and validated formulation can achieve global usage only if its ingredients are brought down without affecting its efficacy and safety. Therefore, in the present study an attempt has been made to optimize UNIM 352 by comparing the efficacy and safety of the original six ingredients with the new optimized preparations (OP) with successive elimination of 1 to 3 ingredients deemed less important in light of available information.

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Materials and methods

Experimental animals

Wistar rats, weighing 180-220g were used in the present study. They were housed in polyacrylic cages and kept in standard controlled room with natural light and dark cycle. Rats were fed with standard food pellets and water ad libitum. The experimental protocol was approved by Institutional Animal Ethics Committee (IAEC/7/2015) and the guidelines of Indian National Science Academy, New Delhi. Rats were followed for care and safety of animals in scientific research.

Drugs and chemicals

UNIM-352 and optimised preparations of UNIM-352 used in the present study, are standardized formulations prepared and supplied by Central Research Institute of Unani Medicine (CRIUM), Hyderabad, under the auspices of Central Council for Research in Unani Medicine (CCRUM), Ministry of AYUSH, Government of India, New Delhi.

UNIM-352 contains 6 ingredients: Linum usitatissimum L. [16] (1g), Trigonella foenum-graecum L. (2g), [17] Allium sativum L (3.5g), [18] Strychnos potatorum L (7g), [19] Caesalpinia bonducella Fleming (1g), [20] Pongamia glabra Vent (1g), [21] and Honey (q.s.) in 20g of preparation.

Optimised preparation-1(OP-1) contains 3 ingredients: Strychnos potatorum L, Trigonella foenum-graecum L, Pongamia glabra Vent, Honey.

Optimised preparation-2(OP-2) contains 4 ingredients: Strychnos potatorum L, Trigonella foenum-graecum L, Pongamia glabra Vent, Allium sativum L, and Honey.

Optimised preparation-3(OP-3) contains 4 ingredients: Strychnos potatorum L, Trigonella foenum-graecum L, Pongamia glabra Vent, Linum usitatissimum L, and Honey.

Optimised preparation-4(OP-4) contains 4 ingredients: Strychnos potatorum L, Trigonella foenum-graecum L, Pongamia glabra Vent, Caesalpinia bonducella Fleming, and Honey. Prednisolone, and Ovalbumin (OVA) were purchased from M/s. Sigma Aldrich. Rat IgE and IL-4 ELISA kit were purchased from QAYEE-BIO (Cat no-QY-E11192) and Diaclone (Cat no-8650.020.096) respectively.

Experimental groups and treatment

Wistar rats were divided randomly into four groups each containing six rats: (i) Normal group: Rat were treated only with distilled water (vehicle) orally for 14 days; (ii) Control group: rats were immunized and challenged with OVA and treated on day 14 and treated with distilled water orally; (iii) and (iv) UNIM-352(200) and (400) groups: rats were immunized and challenged with OVA and treated with UNIM-352 at the dose of 200 or 400 mg/kg from day 1 to 14, orally (v) and (vi) Optimised preparation-1 (OP-1): rats were immunized and challenged with OVA and treated orally with OP-1 at the dose of 200 or 400 mg/kg; (vii) and (viii) Optimised preparation-2 (OP-2): rats were immunized and challenged with OVA and treated orally with OP-2 at the dose of 200 or 400 mg/kg; (ix) and (x) Optimised preparation-3, (OP-3): rats were immunized and challenged with OVA and treated orally with OP-3 at the dose of 200 or 400 mg/kg; (xi) and (xii) Optimised preparation-4, (OP-4): rats were immunized and challenged with OVA and treated orally with OP-4 at the dose of 200 or 400 mg/kg; (xiii) Prednisolone group: rats were immunized and challenged with OVA and treated orally with prednisolone at the dose of 10mg/kg.

Immunization and challenge protocol

All rats in each group except Normal group were immunized and challenged with OVA according to the procedure described by Kwasniewski, et al. (1998) [22]. Briefly, rats were sensitized by intraperitoneal injection of OVA (10 mg per rat) emulsified with 10µg of aluminium hydroxide in 0.5 ml of normal saline on the first day of the study. After 14 days of sensitization, animals were challenged with 1 mg of ovalbumin.

BAL fluid collection

After 24 hrs of OVA challenge, all animals were anesthetized, and BAL fluid was collected. BAL fluid was collected by lavaging the lung through tracheal cannula with 0.9% sodium chloride solution and centrifuged at 1500 rpm at 4°C for 10 min and supernatant recovered and stored at -80°C for the assay of IL-4 cytokine [22]. The precipitated pellets were resuspended in 100 µl of normal saline. Eosinophil and neutrophil counts in BAL fluid were carried out using Neubauer chamber.

Assay for IL-4 Levels

BAL fluid samples were assayed for the levels of IL-4 using commercially available ELISA Kits. Briefly the cytokine assays were performed using solid phase sandwich ELISA. Antigen and biotinylated polyclonal antibody specific for IL-4 were added to the microtitre plate wells whose walls were precoated with polyclonal antibody specific for IL-4 and incubated for specific period. The HRP conjugated streptavidin was added and incubated. Finally, TMB substrate solution was added to induce a coloured reaction product. Then again incubate in the dark for 10-20 min at room temperature, avoid direct exposure to light. The enzyme substrate solution was stopped by adding H2So4. The absorbance of the coloured product was measured using microplate reader (ECIL) at 450nm and results were expressed in pg/ml.

Assay for IgE levels

BAL fluid samples were assayed for levels of IgE using commercially available enzyme-linked immunosorbent assay (ELISA) kits. These cytokine assays were performed using double antibody sandwich ELISA. The microtitre plate provided is precoated with monoclonal antibody specific to Immunoglobulin E (IgE). Samples were added to wells and after incubation OVA s IgE secondary antibody labelled with biotin was added. Streptavidin-HRP was added to wells in order to form immune complex and was then incubated. Chromogenic solution A and B were added to develop colour and the reaction was stopped by using a stop solution. The absorbance of colored product was measured using microplate reader at 450 nm and results were expressed in ng/ml.

Statistical analysis

Data are expressed as mean ± SEM and analyzed by using one-way ANNOVA followed by tukey's test. A P value of <0.05 was used as the level of significance in all statistical tests.

Results

Effects of UNIM-352 and optimized preparations of UNIM-352 on IgE in BAL fluid

Assay for IgE showed that sensitization and challenge with OVA resulted in significant increase in the level of IgE in BAL fluid, as compared to the Normal group. UNIM-352(200and 400mg/kg) and
both the doses of optimized formulations of UNIM-352 (OP-1, OP-2, OP-3 and OP-4) decreased the level of IgE in BAL fluid, when compared with vehicle treated Control group. UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 200 mg/kg reduced IgE levels by 60%, 81%, 63%, 73% and 80% respectively, as compared to Control. UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 400 mg/kg reduced IgE levels by 56%, 85%, 61%, 79% and 82% respectively, as compared to Control. The reductions were comparable with the standard drug Prednisolone. OP-1 at both the doses significantly reduced the IgE levels in BAL fluid as compared to UNIM-352 (400 mg/kg). Further, OP-4 at the dose of 400 mg/kg also significantly reduced the IgE levels in BAL fluid vs UNIM-352 (400 mg/kg). Interestingly, the reductions were greater in magnitude as compared to UNIM-352. Overall analysis of the data revealed that the changes in the levels of IgE after various treatment were significantly different across all groups (p<0.01) and comparisons of individual groups with Control group are shown in Figure 1.

**Effects of UNIM-352 and optimized preparations of UNIM-352 on Eosinophils and Neutrophils in BAL fluid**

Effects of optimized formulations of UNIM-352 were assessed on eosinophil cell counts in BAL fluid in OVA sensitized and challenged rats. The results showed that sensitization and challenge with OVA resulted in significant increase in the number of eosinophil cells in BAL fluid, as compared to the Normal group (not sensitized and challenged with OVA). UNIM-352 and Optimized preparations of UNIM-352 differentially reduced the number of eosinophil cells at both the dose levels (200 and 400mg) in BAL fluid, when compared with vehicle treated Control group (sensitized and challenged with OVA). UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 200 mg/kg reduced eosinophil counts by 55%, 42%, 39%, 34% and 33% respectively, as compared to Control. UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 400 mg/kg reduced eosinophil counts by 70%, 55%, 60%, 57% and 51% respectively, as compared to Control. Pre-treatment with the standard drug Prednisolone, markedly suppressed the eosinophil counts in BAL fluid. Analysis of the data revealed that the changes in the number of eosinophil cells after various treatment were significantly different across all groups (p<0.01) and comparisons of individual groups with Control group are shown in Figure 2.

Similarly, effects of optimized formulations of UNIM-352 were assessed on neutrophil cell counts in BAL fluid in OVA sensitized rats. The results showed that sensitization and challenge with OVA resulted in significant increase in the number of neutrophil cells in BAL fluid, as compared to the Normal group. UNIM-352 and Optimized preparations of UNIM-352 reduced the number of neutrophil cells at both the dose levels (200 and 400mg) in BAL fluid, vs Control group. UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 200 mg/kg reduced neutrophil counts by 42%, 39%, 31%, 64% and 56% respectively, as compared to Control. UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 400 mg/kg reduced neutrophil counts by 56%, 60%, 50%, 68% and 65% respectively, as compared to Control. The reductions were comparable with the standard drug Prednisolone. Overall analysis of the data revealed that the changes in the number of neutrophil cells after various treatment were significantly different across all groups (p<0.01) and comparisons of individual groups with Control group are shown in Figure 3.

**Effects of UNIM-352 and optimized preparations of UNIM-352 on IL-4 in BAL fluid**

Assay for IL-4 showed that sensitization and challenge with OVA resulted in significant increase in the level of IL-4 in BAL fluid, as compared to the Normal group. UNIM-352(200and 400mg/kg) and both the doses of optimized formulations of UNIM-352 (OP-1, OP-2, OP-3 and OP-4) decreased the level of IL-4 in BAL fluid, when compared with vehicle treated Control group. UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 200 mg/kg reduced IL-4 levels by 25%, 33%, 21%, 28% and 22% respectively, as compared to Control. UNIM-352, OP-1, OP-2, OP-3 and OP-4 at dose 400 mg/kg reduced IL-4 levels by 39%, 50%, 30%, 49% and 30% respectively, as compared to Control. The reductions were comparable with the standard drug Prednisolone. OP-1 and OP-3 at the dose of 400 mg/kg significantly reduced the IL-4 levels in BAL fluid as compared to UNIM-352 (200 mg/kg). Overall analysis of the data revealed that the changes in the levels of IL-4 after various treatment were significantly different across all groups (p<0.01) and comparisons of individual groups with Control group are shown in Figure 4.

**Discussion**

Asthma is a chronic inflammatory disease which is characterized by eosinophilic airway inflammation, mucus hyper-secretion, and bronchial hyper-responsiveness. Its pathophysiology is regulated by the complex interactions between cellular and humoral components. The pharmacotherapy depends on controllers (corticosteroids) and relievers (bronchodilators) agents and life-long therapy is required for management of bronchial asthma, once the patient is diagnosed. Though significant incidence of adverse effects related to these drugs has been a key area of concern, the increasing incidence of refractoriness to conventional forms of therapy has further complicated the problem [5,6]. Therefore, it is the need of time to focus extensively on alternative

![Figure 1](image1.png)  
**Figure 1.** Effects of UNIM-352 and optimized preparations of UNIM-352 on IgE levels in BAL fluid of OVA immunized and challenged rats. Data are expressed as Mean ± SEM. *p< 0.01 vs normal group; **p < 0.05 and ***p < 0.01 vs control group; †p < 0.05 vs UNIM-352 (200 mg/kg) and ‡p < 0.05 vs UNIM-352 (400 mg/kg).

![Figure 2](image2.png)  
**Figure 2.** Effects of UNIM-352 and optimized preparations of UNIM-352 on eosinophil counts in BAL fluid of OVA immunized and challenged rats. Data are expressed as Mean ± SEM. *p< 0.01 vs Normal group; †p < 0.05 and ‡‡p < 0.01 vs control group; ††p < 0.05 vs UNIM-352 (200 mg/kg) and ‡‡†p < 0.01 vs Control group.
strategies for rational management of the disorder. Medicinal plants and plant derived products have long been recognized as major resources for alternative therapy for several disease states. The World Health Organization also encourages, recommends and promotes traditional or herbal drugs in National Health Care Programmes because of their easy availability, low cost, safety and the faith of people in such remedies. Although, Indian traditional medicine like Ayurveda and Unani have several formulations to cure bronchial asthma based on clinical experience but, there is need for scientific validation of such claims before recommendation for complimentary therapy.

UNIM-352 is one such polyherbal Unani preparation with documented clinical use in patients of bronchial asthma. Our pilot clinical study has shown that UNIM-352 improves the efficacy and safety of conventional treatment of bronchial asthma in modern medicine and thus, could be considered as viable alternatives or an adjunct for pharmacotherapy [23-25]. Acute and sub-acute toxicity studies with UNIM-352 and its optimized preparations have shown that it is safe up to the dose of 2000 mg/kg and showed no toxicity sign as compared to that in control rats.

The inflammatory process in asthma involves antigen interaction and sustained infiltration and activation of many inflammatory cells including eosinophils, neutrophils, lymphocytes, basophils and macrophages, followed by initiation and release of various pro-inflammatory mediators and cytokines [26-28]. Th2 cells are the key orchestrators of this inflammation, which initiates and propagate inflammation by the release of cytokines, IgE, IL-4. These are well known mediators, and a potent modulator of immune and inflammatory response resulting in influx of neutrophils and eosinophils as well as bronchial hyperreactivity.

Inhibition of inflammatory responses mediated through inflammatory cells and their cytokines, is a basic therapeutic aim in treatment of asthma. Present study showed significant suppression of IgE responses by administration of optimized preparations of UNIM-352 in BAL fluid samples as compared to control. Inflammation and infiltration of the airways with eosinophils and neutrophils is the characteristic feature of asthma and in most of the phenotype is accompanied with increased levels of these cells in tissue, blood and bronchoalveolar lavage (BAL) fluid in general and has direct correlation with disease severity [29]. The result of our experiments showed that treatment with optimized formulations of UNIM-352 (OP-1, OP-2, OP-3 and OP-4) and UNIM-352 alone in both dose levels (200 and 400mg/kg) reduced the number of eosinophil and neutrophil cells in BAL fluid of ovalbumin immunized and challenged rats. Prednisolone (10mg/kg) treatment, which acted as positive control group, also decreased the number of eosinophils and neutrophils cells in separate group of rats indicating the comparability of our data.

IL-4 plays an important role in the development of allergic airway inflammation. Patients with bronchial asthma have increased IL-4 protein levels in serum and bronchoalveolar lavage fluid [30]. The results of our study showed that IL-4 levels were suppressed in BAL fluid in ovalbumin immunized and challenged rats after treatment with optimized formulations (OP-1, OP-2, OP-3 and OP-4) and UNIM-352. Interestingly, the reductions were more marked with OP-1 and OP-3 preparations as compared to UNIM-352, which may envisage better efficacy of these optimized versions. Further, results of OP-1 on suppression of IgE levels also showed better efficacy as compared to positive control prednisolone. However, detailed studies on various cellular and molecular markers of asthma are required to confirm better efficacy of OP-1 and OP-3 as compared to UNIM-352.

Taken together, it can be concluded that UNIM-352 and its optimized preparations may act by preventing infiltration of the eosinophils and neutrophils (the effector cells in asthma) and reducing the levels of IgE and Th2 cytokine, IL-4 which are responsible for release of various inflammatory mediators, thus contributing to the therapeutic benefits observed in patients of bronchial asthma.

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References

1. Shifren A, Witt C, Christie C, Castro M (2012) Mechanisms of remodeling in asthmatic airways. J Allergy (Cairo) 2012: 316049. [Crossref]
2. Behera D, Sehgal IS (2015) Bronchial asthma – issues for the developing world. Indian J Med Res 141: 380–382. [Crossref]
3. Brightling CE, Symon FA, Birring SS, Bradding P, Pavord ID, et al. (2002) TH2 cytokine expression in bronchoalveolar lavage fluid T lymphocytes and bronchial submucosa is a feature of asthma and eosinophilic bronchitis. J Allergy Clin Immunol 110: 899-905. [Crossref]
4. Busse WW, Rosenwasser LJ (2003) Mechanisms of asthma. J Allergy Clin Immunol 111: 799-807. [Crossref]
5. Wenzel SE, Covar R (2006) Update in asthma 2005. Am J Respir Crit Care Med 173: 698-706. [Crossref]
