The effects of *Michelia alba* oil against mould on brown rice and assessing the brain response using electroencephalogram (EEG)

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Abstract The objective of this study was to develop antifungal fragrant brown rice using the vapour phase of *Michelia alba* oil and to understand the effects of fragrant brown rice on consumer reactions using an electroencephalography (EEG) technique. The effect of *M. alba* oil vapour (300–900 μL/L) on the growth of moulds was studied in brown rice. Then, optimisation of the *M. alba* oil vapour (300–900 μL/L) was studied through sensory evaluation. Next, EEG was used to investigate the effect of fragrant cooked brown rice on human brain activity. In addition, the key components of *M. alba* oil on the sensory effects were determined. The results indicated that *M. alba* oil vapour ≥ 450 μL/L provided effective antifungal activity against natural moulds on brown rice for at least 90 days of storage at 25 °C at 100% RH. Furthermore, the optimal concentration of *M. alba* oil vapour for enhancing consumer preference and acceptance of cooked brown rice was 300–600 μL/L with a rejection threshold of 2,052 μL/L. Moreover, it was found that linalool was the main key component and caryophyllene and β-elemene were the minor components affecting the sensory quality enhancement. Interestingly, the EEG results showed that fragrant cooked brown rice could increase the power of alpha and beta waves in the human brain, indicating anti-stress effects and a relaxed mood. Therefore, *M. alba* oil vapour demonstrated good potential to enhance consumer acceptance and preference for cooked brown rice while controlling the significant growth of moulds in brown rice.

Keywords Antifungal · Fragrant brown rice · *Michelia alba* oil vapour · Sensory · Electroencephalography

Introduction

Brown rice not only contains high nutritional values such as high protein, fibre, vitamins and mineral contents (Kalpanadevi et al. 2018) but also contains salutary health components, namely, tocopherol, phytosterol, γ-oryzanol, and γ-aminobutyric acid as well as phenolics and anthocyanins (Sapna et al. 2019). However, brown rice and its products are responsive to postharvest diseases caused by various moulds, especially, *Aspergillus* spp. *A. flavus* are a major cause of postharvest losses and diseases in brown rice and its products with a high frequency worldwide (Suhem et al. 2015). Recently, has been discovered that the application of essential oils such as that of *Litsea cubeba* (Suhem et al. 2015), bergamot (Songsamoe et al. 2016) and *Michelia alba* (Songsamoe et al. 2017) can suppress the growth of *A. flavus* in brown rice.

The new era of COVID-19 pandemic gives insights into the need for bioactive ingredients from food and herbs which could be used to increase the human immune system against infections (Galanakis 2020). The oil of *Michelia alba* contains aromatic constituents that exhibit various bioactivities. For example, it can be used as an anti-inflammatory agent to treat cramps, abdominal pain (Lee et al. 2005), fever, syphilis and malaria (Asaruddin et al. 2003), and It can also be used to protect against the growth of*.
of A. niger, A. flavus, Penicillium sp., Rhizopus sp., Fusarium sp., and Cladosporium sp. (Songsamoe et al. 2017). In addition, it has also been reported that the oil of M. alba has long been used in healthy food recipes because of its pleasant flavour and because its name sounds like a flower (Rout et al. 2006). Therefore, essential oil could be used as a functional additive in food products and in the development of new products with higher market value (Galanakis 2012). In order to avoid negative sensory effects and reduce the cost of using essential oil on a large scale, the use of M. alba oil in the vapour phase. Flavour and taste impact as well as consumer acceptance should be addressed in order to take the emerging technologies.

In consumer and sensory research of food products, consumers’ physiological and emotional responses are emphasised because they are fully correlated with consumer product acceptability. Flavour and taste impact, as well as consumer acceptance should be addressed so as to utilize the emerging technologies (Galanakis 2013). Therefore, good understanding of consumers’ physiological and emotional responses to food products is essential for success in food product design and food services (Desmet and Hekkert 2009). The samples and test situations could elicit positive or negative emotions which may generate different emotions to overall product performance (Worch et al. 2020). Many traditional explicit measurements, namely, preference, acceptance, liking and hedonic valuation, have been used for the evaluation of consumers’ feelings and preferences (King and Meiselman 2010). Nowadays, brain activity, facial expression and heart rate could be used to study a physiological response of consumers (Lagast et al. 2017). To understand of consumers’ emotional responses, non-verbal methods could be done such as look at food and smelling for cognitive by consumers (Kaneda et al. 2004). The measurement of brain activity by electroencephalography (EEG) is one interesting technique that has recently been applied in the consumer and sensory research of food products. It can provide useful and detailed data to support the understanding of consumer responses to food (Lagast et al. 2017). EEG data obtained from consumers after eating food have been considered in terms of brain function and consumers’ emotional responses. A change in the spectral power of EEG waveforms can be interpreted as the feelings or brain functions of the consumer after consuming food. In addition, the asymmetry of EEG power from the right and left hemispheres of the brain can be interpreted as motivational tendency or food acceptability. Therefore, the objective of this work was to study the effect of M. alba oil on mould growth and to find a suitable volume of M. alba oil for application in brown rice to promote a positive reception from the consumer.

### Materials and methods

#### Essential oil

*M. alba* oil extracted by steam distillation was obtained from Thai China Flavours and Fragrances Co., Ltd. of Thailand. The components of *M. alba* oil were determined using gas chromatography–mass spectrometry analysis. This analysis was carried out on a gas chromatograph–tandem mass spectrometer model 7890 B GC-700D MS from Agilent, USA, equipped with an Agilent HP-5 ms Ultra Inert column (30 m × 250 μm × 0.25 μm). The average helium carrier gas flow rate was 1 mL min⁻¹, the injected volume was 1 μL, the split ratio of the column was 10:1, and the injector temperature was set at 250 °C. The initial column oven temperature was set at 40 °C, then increased to 180 °C at a rate of 10 °C/min and held for 1 min and then raised to 300 °C at a rate of 5 °C/min. The detector temperature was set at 300 °C. The components of the oil were identified by comparison of their mass spectra fragmentation and computer matching using the Wiley 10 and NIST 14 libraries (Database/ChemStation data system).

#### Effect of *M. alba* oil vapour against *A. flavus* on MEA and natural mould on brown rice

**Culture**

*Aspergillus flavus* (10⁶ CFU/mL) was isolated from brown rice and obtained from the Research Center of Excellence in Innovation of Essential Oil, Walailak University, Nakhon Si Thammarat, Thailand. *A. flavus* was inoculated onto malt extract agar (MEA) before incubation at 25 °C for 7 days. The spore suspension was prepared by pouring 9 mL of sterile water onto the agar slant and mixing. The number of viable spores was counted on the MEA.

**Antifungal activity testing on MEA**

The antifungal activity of *M. alba* oil vapour (extracted from the flowering part of the plant) was observed. For this, *A. flavus* (diameter ~ 5 mm) was added onto the centre of the MEA plate. *M. alba* oil of different volumes (150–900 μL) was added into the container (1 L). Then, the plate (without a lid) was placed inside the container. All containers were incubated at 25 °C for 5 days. The control sample was treated in the same way, but without the essential oil. After incubation, the colony diameter (mm) of *A. flavus* on each plate was checked. The percent inhibition of mould growth was computed based on the following Eq. 1:
Growth inhibition (\%) = \frac{(\text{Control} - \text{Treatment}) \times 100}{\text{Control}} \quad (1)

where Control = the colony diameter (mm) of A. flavus on the control plate; Treatment = the colony diameter (mm) of A. flavus on the sample plate.

**Brown rice grain storage with M. alba oil vapour**

The effect of *M. alba* oil vapour (0 \( \text{control} \), 300, 450, 600, 750 and 900 \( \mu \)L/L) against natural mould on brown rice grains (100 g) was examined in a polyethylene (PE) plastic bag (available volume = 250 mL, thickness = 220 \( \mu \)m). The bag was stored at 25 \( ^\circ \)C and 100% RH for 90 days. During storage, the mould growth on the brown rice surface was observed, and the inhibitory period (day) was reported.

**Sensory assessment of *M. alba* oil vapour on cooked brown rice**

**Preference and acceptance testing**

Different volumes of *M. alba* oil vapour (300–900 \( \mu \)L) were applied to brown rice (400 g) in a container (1 L). The container was closed and stored at 25 \( ^\circ \)C for 30 days. Then, the brown rice (150 g) was soaked in a pot for 5 min with 400 mL of deionised water. Next, the rice was cooked in an automatic rice cooker (Otto Kingglass Co., Ltd. Thailand). After cooking, the brown rice samples were subjected to sensory analysis by an untrained panel (89 panellists) ranging in age from 20 to 45 years.

In order to find the preferred concentration of *M. alba* vapour in brown rice, five sessions of paired preference tests were conducted for each panellist. Each pair of samples consisted of a cooked brown rice control sample (without *M. alba* oil vapour, Sample A) and a stimulus sample (treated with *M. alba* oil vapour at concentrations of 300 [B], 450 [C], 600 [D], 750 [E] and 900 \( \mu \)L [F]). Panellists were asked to taste the samples in each pair (A–B, A–C, A–D, A–E and A–F at random) and to indicate their preference (Lima Filho et al. 2015). In each test, after the panellists rinse their mouths with water, they received a new pair of samples every 5 min. The preferred concentration of essential oil in cooked brown rice compared with the control was analysed using the chi-squared (\( \chi^2 \)) (\( p = 0.05 \)).

In addition, a nine-point hedonic scale (9 = extremely like, 1 = extremely dislike) was used to determine the degree of acceptance of the specimens in terms of odour, flavour (a combination sensation perceived by the oral and nasal cavities), taste (the sensation perceived by the tongue) and overall liking (Meilgaard et al. 1999).

**Rejection threshold**

The rejection threshold (RT) was also determined. A graph was constructed of the average hedonic value (Y-axis) as a function of the *M. alba* oil vapour concentration in the brown rice sample under investigation (X-axis). The cut-off point of the Y-axis was denoted on the graph by a dashed line identifying the hedonic score of 5 (the hedonic term “indifferent”), demonstrating the transition point between sensory acceptance and rejection of the cooked brown rice. The “indifferent” term of the hedonic scale was considered a rejection point since a consumer indifferent to a product is not likely to buy it (Della Lucia et al. 2014). To determine the *M. alba* oil vapour concentration at which sensory rejection of the cooked brown rice begins to occur, a regression model was adjusted to the points of the Y-axis. To select the model that was the best fit of the data, the significance of the regression coefficients and the coefficient of determination \( R^2 \) was evaluated using Statistica software (StatSoft, USA).

**Determination of a key component of *M. alba* oil vapour in sensory effects**

The components of *M. alba* oil vapour absorbed into brown rice grains and remaining in cooked brown rice were investigated using GC–MS analysis. Each sample (10 g) was extracted with ethyl acetate (10 mL). Then, it was mixed gently by shaking overnight. Next, it was filtered and centrifuged at 12,000 rpm for 5 min. Afterward, the supernatant was reduced to dryness under a stream of nitrogen at room temperature. Finally, the residue of each sample was dissolved in 1 mL ethyl acetate before 1 mL aliquots of each solution were subjected to GC–MS analysis under the same conditions used in the previous section.

In addition, the components of *M. alba* oil vapour which had the main effects in enhancing the sensory qualities of cook brown rice were determined using sensory analysis. Differences from the control were used to investigate matches between the odour of treated cooked brown rice and the odours of components of *M. alba* oil. The control (cooked brown rice treated with *M. alba* oil vapour at 600 \( \mu \)L/L) was prepared by the same procedure used in the previous section. The pure components of *M. alba* oil (li-nalool, \( \beta \)-elemene and caryophyllene) were prepared in a smelling box. Afterward, the specimens were subjected to sensory analysis by an untrained panel (50 panellists, 35 females and 15 males) ranging from 18 to 47 years of age. The specimens were presented to panellists seated separately in control booths. A 10-point scale of difference...
from the control ranging from “not different” to “extremely different” was used to determine the degree of difference from the control of the specimens in terms of odour.

**Physical and chemical properties of brown rice after treatment with *M. alba* oil vapour**

Brown rice treated with *M. alba* oil vapour at 600 µL/L at 25 °C for 30 days and the control (without essential oil) evaluated for physical and chemical properties. In brief, optimal cooking time was determined using the glass plate-white center method. The colour of cooked brown rice was determined by a colorimeter (ColorFlex, Hunter Associates Laboratory, USA). The texture profile analysis (TPA) of brown rice was tested using a texture analyzer (LR 5K MK4, Lloyd Instrument Co., Ltd., England). A cylinder probe of 45 diameters was used to compress the sample to 75% deformation at the test speed of 5 mm/s. The chemical compositions of brown rice were determined by the standard methods of AOAC (2019).

**Effect of cooked brown rice containing *M. alba* oil vapour on human brain activity**

**Participants**

Ten healthy volunteers (male = 5, female = 5) between the ages of 20 and 31 were recruited from the staff and students at Walailak University. They were determined as healthy by a medical questionnaire. The testing protocol was approved by the Human Research Ethics Committee of Walailak University (Protocol Number WUEC-16-115-01).

**Cooked brown rice samples**

Brown rice specimens were treated with *M. alba* oil vapour at 600 µL/L at 25 °C for 30 days. After cooking in an automatic rice cooker, the brown rice specimens were used to test olfactory responses by the EEG technique.

**EEG procedure**

The effect of cooked brown rice containing *M. alba* oil vapour on human brain activity was investigated using electroencephalography (EEG) (Research Institute for Health Science, Health Sciences Research Center Building, Walailak University, Nakhon Si Thammarat, Thailand). The internationally standardised 10–20 system (FP1, FP2, F7, F3, FZ, F4, F8, T7, C3, CZ, C4, T8, M1, M2, P7, P3, PZ, P4, P8, O1, O2) was employed to record the spontaneous EEGs obtained from participants during smelling and mouthing and after swallowing of cooked brown rice.

Power spectral analysis was performed using Curry 7 (Compumedics, Australia), and results were compared between rice with and without *M. alba* oil vapour.

The testing sessions began at 10:00 a.m. Participants were instructed to refrain from alcohol consumption for 12 h (overnight), to sleep more than 8 h prior to the test sessions and to arrive at the EEG laboratory at 8:30 a.m. Participants were then informed that they should not drink or eat until the test session began at 10:00 a.m. The test conditions and procedures were explained to the participants prior to completion of the questionnaire. Then, the participants’ heart rates, temperatures, weights and heights were recorded. Next, electrodes were placed on the scalp of participants while they were seated in a recliner with minimal movement. The EEG recording was carried out following the procedure shown in Fig. 1. The baseline EEG was recorded while the participants closed their eyes for 2 min. Then, EEGs were recorded during smelling, mouthing, and swallowing and after swallowing (5 min) the cooked brown rice samples. Each step took place in 2-min intervals. After all the steps were completed, the electrode cap was removed.

**Statistical analysis**

All results were expressed as the mean ± standard deviation. One-way ANOVA and Duncan’s post hoc test, with *p* < 0.05 being considered statistically significant, were employed. Kruskal–Wallis and Wilcoxon matched-pairs signed rank tests were used with non-parametric data. Statistical analysis was performed using Statistica software (StatSoft, USA).

**Results and discussion**

**Effect of *M. alba* oil vapour on mould growth**

The effects of *M. alba* oil in the vapour phase on the growth of *A. flavus* on MEA found that the vapour phase of *M. alba* oil at a concentration of 450 µL/L could inhibit the growth of *A. flavus* on MEA (by 100%), followed by vapour at concentrations of ≥ 150–400 µL/L (by > 90%). These findings demonstrated that *M. alba* oil vapour possessed good antifungal activity. In addition, the results from the sensory test demonstrated that the appropriate volume of *M. alba* oil vapour was 300–600 µL/L in cooked brown rice, which showed the like slightly to like moderately in the sensory test. Therefore, using *M. alba* oil vapour from 300 to 600 µL/L on brown rice showed positive effects on antifungal activity and consumer acceptance. In addition, the effects of *M. alba* oil in the vapour phase on the growth of natural mould on brown rice clearly
showed that M. alba oil vapour (≥ 450 µL/L) on brown rice under accelerated storage conditions could prolong the shelf-life of brown rice by more than 6 times (90 days) compared with that of the control sample (15 days).

The results from this experiment confirmed that M. alba in the vapour phase (≥ 450 µL/L) could completely inhibit the growth of A. flavus. By performing a comparison with the M. alba oil from the leafy part of the plant, it was found that oil from the flowering part is less effective against A. flavus than that from the leafy part (Songsamoe et al. 2016). Linalool (the major component of M. alba oil), a monoterpenic alcohol, is widely known as a highly effective antimicrobial compound (Suppakul et al. 2011). It can disrupt the cell walls and membranes of moulds (Zeng et al. 2011). Linalool has been found to have a safety profile in studies showing that oil from the flowering part is less effective against A. flavus than that from the leafy part (Songsamoe et al. 2016). Linalool (the major component of M. alba oil), a monoterpenic alcohol, is widely known as a highly effective antimicrobial compound (Suppakul et al. 2011). It can disrupt the cell walls and membranes of moulds (Zeng et al. 2011). Linalool has been found to have a safety profile in studies showing that oil from the flowering part is less effective against A. flavus than that from the leafy part (Songsamoe et al. 2016).

Effect of M. alba oil vapour on the sensory qualities of cooked brown rice

The results of consumer acceptability of cooked brown rice treated with M. alba oil vapour in Table 1 show that brown rice treated with M. alba oil vapour at 300–900 µL/L could increase the consumer acceptability of cooked brown rice. Concerning the effects of different concentrations of M. alba oil vapour, treating brown rice with M. alba oil vapour at low concentrations (300–600 µL/L) could significantly improve all sensory attributes of the cooked brown rice (odour, flavour, taste, colour and overall liking). Increasing the concentration of M. alba oil vapour to higher than 600 µL/L initially resulted in a decrease in the liking scores for taste, followed by flavour and overall liking, but the participants’ liking scores were still higher than those for normal cooked brown rice (untreated with M. alba vapour). While there were lower effects from higher vapour concentrations on the liking scores of the odour of cooked brown rice, the liking scores were still high.

The rejection threshold (RT) is best predicted by the model Eq. 2. The coefficient of determination ($R^2$) was 0.75, which indicated that the model equations adequately fit the data. The RT was determined as 2052 µL/L, above which concentration the cooked brown rice will be rejected by consumers. Taken together, these findings indicated that treating brown rice grains with M. alba oil vapour at 300–600 µL/L was optimal for improving the sensory qualities of cooked brown rice.

$$Y = -0.0011X + 7.2573 \quad (2)$$

where Y is the predicted hedonic score and X is the M. alba vapour concentration.

It is well known that the high volatility, reactivity, odour and flavour of essential oils might cause severe interference with food product sensory qualities, resulting in positive or negative characteristics (Espina et al. 2014; Kim et al. 2013). The strong flavour of many essential oils is primarily limiting when used in products at high concentrations when applied to food, while the appropriate matching of essential oil flavours to food products can provide higher acceptance at higher concentrations. For example, lemon

### Table 1 The effect of cooked brown rice treated with M. alba oil vapour on consumer acceptance

| Treatments          | Odour   | Taste    | Flavour  | Overall   |
|---------------------|---------|----------|----------|-----------|
| M. alba flower oil 300 µL/L | 6.8 ± 1.5<sup>a</sup> | 6.9 ± 1.4<sup>a</sup> | 6.8 ± 1.3<sup>a</sup> | 7.1 ± 1.3<sup>a</sup> |
| M. alba flower oil 450 µL/L | 6.6 ± 1.4<sup>a</sup> | 6.5 ± 1.6<sup>b</sup> | 6.5 ± 1.4<sup>ab</sup> | 6.7 ± 1.5<sup>b</sup> |
| M. alba flower oil 600 µL/L | 6.5 ± 1.5<sup>a</sup> | 6.6 ± 1.5<sup>ab</sup> | 6.5 ± 1.6<sup>a</sup> | 6.9 ± 1.4<sup>a</sup> |
| M. alba flower oil 750 µL/L | 6.4 ± 1.8<sup>a</sup> | 6.2 ± 1.8<sup>b</sup> | 6.0 ± 1.8<sup>bc</sup> | 6.3 ± 1.7<sup>ab</sup> |
| M. alba flower oil 900 µL/L | 6.3 ± 1.7<sup>a</sup> | 6.1 ± 1.8<sup>b</sup> | 6.0 ± 1.9<sup>c</sup> | 6.3 ± 1.7<sup>ab</sup> |
| Control             | 5.9 ± 1.6<sup>b</sup> | 6.0 ± 1.6<sup>b</sup> | 5.9 ± 1.5<sup>c</sup> | 6.1 ± 1.4<sup>c</sup> |

<sup>a,b</sup> Different superscripts are significantly different (p < 0.05)
oil was acceptable at higher concentrations than were other essential oils when applied in tomato juice, vegetable soup, or poultry burgers, because of the higher appreciation of its sour, citrus-like flavour (Espina et al. 2014). Therefore, food compatibility and the matching of essential oil components with the physicochemical characteristics of food products have a significant impact on essential oil applications concerning food product sensory qualities. In addition, avoiding using essential oils in the liquid phase could reduce the concentration and the effects on food properties (Tyagi et al. 2012). The results of this work illustrated that appropriate matching of the *M. alba* properties (Tyagi et al. 2012). The results of this work illustrated that appropriate matching of the *M. alba* oil and cooked brown rice flavour by using a small volume of *M. alba* oil in the vapour phase could improve the sensory quality of the cooked brown rice in terms of odour, flavour, taste and colour and increase consumer preference and acceptance. Remarkably, this finding can apply to *M. alba* oil vapour in brown rice at a wide range of concentrations. This is because we can use *M. alba* oil vapour at concentrations up to approximately 1200 µL/L (predicted) without negative effects on sensory properties, and the concentration can be increased to < 2052 µL/L (RT) and remain within the consumer acceptance level.

**Determination of a key component of *M. alba* oil vapour in sensory effects**

The components identified in *M. alba* oil, brown rice grain samples and cooked brown rice samples are shown in Table 2. In *M. alba* oil, 67 components were identified, representing 99.62% of the total components. Linalool (43.47%), β-elemene (8.03%), caryophyllene (6.80%), methyl 2-methylbutyrate (4.05%) and β-selinene (4.00%) were found to be major and minor components of *M. alba* oil. Only 24 components were found in brown rice grains after exposure to *M. alba* oil vapour. This illustrated that not all components of the *M. alba* oil vapour were absorbed into the brown rice grains. However, all major and minor components could be released from the absorbent material and were absorbed by brown rice grains in similar ratios to their presence in *M. alba* oil, since the vapour was present at a high ratio within the system, while other components with very low ratios could be not detected in the sample. After soaking and cooking the brown rice, some components were lost, and it was found that only 14 components remained in the cooked brown rice. The components with lower molecular weights and boiling points such as methyl 2-methylbutyrate (MW = 116.158 g mol\(^{-1}\), boiling point = 116.00 °C) were easier to lose due to heat more compared to the components with higher molecular weights and boiling points such as β-selinene (MW = 204.357 g mol\(^{-1}\), boiling point = 263.00 °C). Noticeably, among the major and minor components, linalool was lost due to heat during cooking more than were β-elemene and caryophyllene, since it has a lower molecular weight and boiling point, which affected the ratio of components in the cooked brown rice (β-elemene = 34.38%, caryophyllene = 19.58% and linalool = 11.32%). Hence, the MW and number of aromatic rings, hydroxyl, carboxylic and methylation groups constitute different solutes that were attracted into the target material (Galanakis 2015).

Although β-elemene was present at a higher ratio in cooked brown rice than were caryophyllene and linalool, it did not play the main role in terms of the odour improvement of cooked brown rice. The score of difference from the control in sensory analysis indicated that the odour of linalool resembled that of the control (cooked brown rice treated with *M. alba* oil vapour) more than did those of β-elemene and caryophyllene, respectively (Table 2). However, the scores obtained for the three components were slightly different compared with the control. This might demonstrate that not only one component of this essential oil provided the sensory effects on cooked brown rice, but instead, the combination of the major components, minor components and other components took a part in the improvement. Accordingly, essential oils are natural, complex aromatic compounds, and their odours and flavours are obtained from the collaboration of many volatile components (Xiao et al. 2017a, b).

Linalool has a strong pleasant aroma because of the acyclic monoprenene, which is described as a floral, citrus and sweet odour (Asikin et al. 2018). It is extensively used as a fragrant ingredient in many cosmetic and cleaning products. Furthermore, it is commonly used as a fragrance and flavour agent in food products (Aprotosoaie et al. 2014). Therefore, linalool is the reasonable component which played the main role in improving the sensory qualities, especially the odour, flavour and taste, of cooked brown rice in this study. β-elemene is described as having a grassy and woody odour, and caryophyllene is described as having a chewing gum and woody odour (Asikin et al. 2018). Therefore, these compounds have a lesser effect on the odour of cooked brown rice when compared with linalool. In addition to the beneficial sensory qualities, these compounds have been proven to have many beneficial biological properties. Linalool has many properties, such as antimicrobial, sedative, anxiolytic, analgesic, anticonvulsant and anti-inflammatory effects (Aprotosoaie et al. 2014), β-elemene has been used as an anticancer drug (Zhao et al. 2007). Caryophyllene has been reported to have anticancer, anti-inflammatory, antioxidant and antimicrobial activities (Basha and Sankaranarayanan 2016).
### Table 2: The components presented in *M. alba* oil, brown rice grain samples and cooked brown rice samples

| Component                                      | R.T  | Peak area (%) | *M. alba* flower oil | Brown rice grain | Cooked brown rice |
|------------------------------------------------|------|---------------|----------------------|------------------|-------------------|
| Methyl-2-butan-1-ol                             | 5.61 | 0.16          | –                    | –                | –                 |
| Methyl 2-methylbutyrate                         | 6.19 | 4.05          | 9.98                 | –                | –                 |
| Ethyl 2-methylbutyrate                          | 7.25 | 0.36          | –                    | –                | –                 |
| Ethyl 3-methylbutanoate                         | 7.30 | 0.15          | –                    | –                | –                 |
| Methyl 2-methylcrotonate                        | 7.52 | 0.23          | –                    | –                | –                 |
| Methylethylacetic acid                          | 8.23 | 1.55          | 9.21                 | –                | –                 |
| α-Pinene                                        | 8.71 | 0.24          | 0.11                 | –                | –                 |
| (-)-camphene                                    | 8.98 | 0.11          | –                    | –                | –                 |
| Hexanoic acid                                   | 9.94 | 0.06          | –                    | –                | –                 |
| β-Pinene                                        | 9.43 | 0.48          | 0.22                 | –                | –                 |
| β-Myrcene                                       | 9.52 | 0.12          | –                    | –                | –                 |
| o-Cymene                                        | 10.13| 0.07          | –                    | –                | –                 |
| 1,5-Cyclooctadiene, 1,5-dimethyl-               | 10.21| 0.19          | –                    | –                | –                 |
| β-cis-Ocimene                                   | 10.27| 2.94          | 1.61                 | 2.14             | –                 |
| 3-Carene                                        | 10.45| 2.84          | 1.42                 | 2.36             | –                 |
| γ-Terpinene                                     | 10.66| 0.07          | –                    | –                | –                 |
| Ethyl 2-(5-methyl-5-vinyltetrahydrofuran-2-yl) propan-2-yl carbonate | 10.88 | 0.26          | 0.16                 | –                | –                 |
| Cyclohexene, 1-methyl-4-(1-methylethylidene)-   | 11.12| 0.19          | –                    | –                | –                 |
| Linalool                                        | 11.46| 43.47         | 52.07                | 11.32            | –                 |
| Phenylethyl alcohol                             | 11.68| 0.69          | –                    | –                | –                 |
| endo-Borneol                                    | 12.41| 0.18          | –                    | –                | –                 |
| Terpinen-4-ol                                   | 12.54| 0.20          | –                    | –                | –                 |
| α-Terpineol                                     | 12.72| 1.29          | 1.10                 | –                | –                 |
| Anisole, p-allyl-                               | 12.77| 0.16          | 0.08                 | –                | –                 |
| Nerol                                           | 13.45| 0.56          | 0.14                 | –                | –                 |
| Linalool oxide acetate                          | 13.92| 0.20          | 0.10                 | –                | –                 |
| Safrole                                         | 14.04| 0.10          | –                    | –                | –                 |
| 1H-indole                                       | 14.11| 0.09          | –                    | –                | –                 |
| δ-Elemene                                       | 14.70| 0.15          | 0.12                 | –                | –                 |
| α-Cubebene                                      | 14.86| 0.34          | 0.30                 | –                | –                 |
| Eugenol                                         | 14.89| –             | 0.12                 | 5.66             | –                 |
| Cycloisosativene                                | 15.19| 0.09          | –                    | –                | –                 |
| Copaene                                         | 15.27| 1.40          | 0.92                 | 3.83             | –                 |
| β-Elemene                                       | 15.48| 8.03          | 9.76                 | 34.38            | –                 |
| cis-α-Bergamotene                               | 15.76| 0.21          | 0.34                 | 1.43             | –                 |
| α-Santalene                                     | 15.86| 0.23          | –                    | –                | –                 |
| Caryophyllene                                   | 16.01| 6.80          | 4.90                 | 19.58            | –                 |
| Bicyclosesquiphellandrene                       | 16.06| 0.16          | –                    | –                | –                 |
| α-Guaiene                                       | 16.11| 0.16          | –                    | –                | –                 |
| (E)-β-Famesene                                  | 16.17| 0.20          | –                    | –                | –                 |
| Cadina-3,5-diene                                | 16.36| 0.13          | –                    | –                | –                 |
| 1,4,7, -Cycloundecatriene, 1,5,9,9-tetramethyl-, Z, Z, Z- | 16.46| 1.71          | 1.23                 | 5.30             | –                 |
| γ-Muurolene                                     | 16.55| 0.10          | –                    | –                | –                 |
| Methyl isoegenol                                | 16.85| 3.30          | –                    | –                | –                 |
| β-Selene                                        | 16.95| 4.00          | 3.54                 | 5.74             | –                 |
| 1R,4R,7R,11R-1,3,4,7 Tetramethyltricyclo [5.3.1.0(4,11)] undec-2-ene | 17.05| 2.25          | –                    | –                | –                 |
Table 2 continued

| Component | R.T     | Peak area (%) | M. alba flower oil | Brown rice grain | Cooked brown rice |
|-----------|---------|---------------|--------------------|------------------|-------------------|
| Eremophila-1-(10),11-diene | 17.14   | 0.31          | –                  | –                |
| γ-Cadinene | 17.27   | 0.21          | –                  | –                |
| δ-Cadinene | 17.38   | 3.30          | 1.37              | 5.96             |
| Nerolidol | 17.79   | 0.74          | –                  | –                |
| α-Calacorene | 18.01  | 0.08          | –                  | –                |
| Caryophyllene oxide | 18.45   | 0.70          | 0.26              | 0.94             |
| Isoaromadendrene epoxide | 18.66   | 0.08          | –                  | –                |
| Isopathulenol | 18.78   | 0.09          | –                  | –                |
| Cubenol (1,10 di-epi) | 19.06   | 0.53          | –                  | –                |
| τ-Muurolol | 19.27   | 0.93          | –                  | –                |
| α-Cadinol | 19.48   | 0.27          | –                  | –                |
| Neointermedeol | 19.56   | 0.52          | –                  | –                |
| Spatulenol | 19.74   | 0.28          | –                  | –                |
| 6-(1-Hydroxymethylvinyl)-4,8a-dimethyl-3,5,6,7,8,8a-hexahydro-1H-naphthalen-2-one | 19.94   | 0.09          | –                  | –                |
| (1R,7S, E)-7-Isopropyl-4,10-dimethylenecyclodec-5-enol | 21.05   | 0.09          | –                  | –                |
| Bicyclo[4.4.0]dec-5-ene, 1,5-dimethyl-3-hydroxy-8-(1-methylene-2-hydroxethyl-1)- | 21.36   | 0.09          | –                  | –                |
| 9-Cycloheptadecen-1-one, (Z)- | 22.78   | 0.24          | –                  | –                |
| Linoleic acid ethyl ester | 22.90   | 0.25          | –                  | –                |
| Nonadecane | 23.19   | 0.10          | –                  | –                |
| Hexadecanoic acid, methyl ester | 23.65   | 0.12          | 0.16              | 0.62             |
| Hexadecanoic acid, ethyl ester | 24.81   | –             | 0.77              | 0.74             |
| 9,12-Octadecadienoic acid (Z,Z)-, methyl ester | 26.59   | 0.22          | –                  | –                |
| 9,12,15-Octadecatrienoic acid, methyl ester, (Z,Z,Z)- | 26.70   | 0.11          | –                  | –                |
| Total | 99.62   | 100           | 100               | 100              |

Effect of *M. alba* oil vapour on the physical and chemical properties of brown rice

The physical and chemical properties of brown rice treated and untreated with *M. alba* oil vapour are shown in Table 3. Results indicated that there were optimistic effects of *M. alba* oil vapour on the physical and chemical properties of brown rice. Treatment of brown rice with *M. alba* oil vapour could cause a significant reduction in the cooking time and the hardness of cooked brown rice, and a small change on the colour of cooked brown rice (ΔE* = 2.54 ± 0.63). Furthermore, it could increase the total phenolic content in brown rice. Moreover, the proximate composition analysis revealed that there were no significant negative effects of *M. alba* oil vapour on the quantity of the nutritional compositions in the brown rice.

The *M. alba* oil components which were absorbed in the brown rice according to the results in “Determination of a key component of *M. alba* oil vapour in sensory effects” section (Table 2), might be responsible for a decrease in cooking time and hardness of the cooked rice. In detail, this could be explained by the ability of the absorbed *M. alba* oil components to damage the rice bran layer of brown rice, thereby promoting the water uptake into the starch granule of brown rice and results into reduction in cooking time and hardness of cooked rice. In addition, the increase in the phenolic content in brown rice may be due to the accumulation of *M. alba* oil components in the brown rice since the *M. alba* oil contains several phenolic compounds.
Effect of cooked brown rice containing *M. alba* oil vapour on human brain activity

The results of the effect of cooked brown rice on human brain activity are shown in Fig. 2 and Table 4. The results indicated that consumption of cooked brown rice both treated with *M. alba* oil vapour and untreated could increase human brain waves in the alpha band (8–13 Hz) and the beta band (13–30 Hz). The alpha band started to increase during smelling the odour of the cooked brown rice and strongly increased after participants chewed and swallowed the samples. The comparison between untreated cooked brown rice samples and those treated with *M. alba* oil vapour showed that the consumption of treated cooked brown rice could increase the power of the slow alpha waves (9 Hz) more than could the consumption of the untreated cooked brown rice sample (*p* < 0.05).

These results demonstrated that the odour and flavour of *M. alba* oil in cooked brown rice could stimulate the human brain to provide increased alpha and beta waves. As noted, this stimulation was evident during smelling the odour of the rice and after chewing and swallowing. Generally, alpha waves are related to semantic information processing and to good cognitive performance (Klimesch 1999), whereas beta waves play an important role in attention or higher cognitive functions (Sauseng et al. 2009). Moreover, the increase of the slow beta wave band (8–11 Hz) correlates with a relaxed state. The low beta wave band (13–15 Hz) and high beta wave band (20–30 Hz) are correlated with musing and complex thoughts, respectively. A high ratio of alpha to high beta indicates a very calm and relaxed state (Subha et al. 2010). Therefore, increased alpha and beta wave power could indicate anti-stress effects and a relaxed mood.

This finding is similar to the effects of other foods on human brain activity. A study by Labbe et al. (2011) reported the effect of refreshing perception from a refreshing drink on consumers’ emotional responses. The EEG wave forms obtained immediately before and 15 min

### Table 3 The physical and chemical properties of brown rice treated and untreated with *M. alba* oil vapour

| Quality factors                  | Brown rice samples                                                                 |
|----------------------------------|-----------------------------------------------------------------------------------|
|                                  | Control                                                                           |
|                                  | Treatment with *M. alba* vapour (600 µL/L air)                                   |
| Cooking time (min)               | 27.57 ± 0.29<sup>a</sup>                                                         | 25.08 ± 0.50<sup>b</sup> |
| Colour                           |                                                                                   |
| L<sup>*</sup>                    | 65.97 ± 0.55<sup>a</sup>                                                         | 63.65 ± 0.21<sup>b</sup> |
| a<sup>*</sup>                    | 3.64 ± 0.11<sup>a</sup>                                                          | 3.81 ± 0.16<sup>a</sup>  |
| b<sup>*</sup>                    | 21.72 ± 0.15<sup>a</sup>                                                          | 22.47 ± 0.15<sup>a</sup> |
| ∆E<sup>a</sup>                  | 2.54 ± 0.63                                                                      |
| Texture analysis                 |                                                                                   |
| Hardness (N)                    | 292.24 ± 12.90<sup>a</sup>                                                        | 171.21 ± 5.63<sup>b</sup>|
| Springiness (mm)                | 4.64 ± 0.04<sup>a</sup>                                                          | 4.54 ± 0.13<sup>a</sup>  |
| Cohesiveness                    | 0.18 ± 0.07<sup>a</sup>                                                          | 0.20 ± 0.07<sup>a</sup>  |
| Adhesiveness (N)                | 4.69 ± 2.67<sup>a</sup>                                                          | 3.81 ± 1.91<sup>b</sup>  |
| Gumminess (N)                   | 52.25 ± 15.94<sup>a</sup>                                                        | 34.32 ± 12.97<sup>b</sup>|
| Proximate composition (% wt)    |                                                                                   |
| Moisture (%)                    | 15.33 ± 0.05<sup>a</sup>                                                          | 15.64 ± 0.05<sup>a</sup> |
| Total protein (%)               | 7.25 ± 0.05<sup>a</sup>                                                          | 7.31 ± 0.04<sup>a</sup>  |
| Lipids content (%)              | 2.36 ± 0.05<sup>a</sup>                                                          | 2.28 ± 0.04<sup>a</sup>  |
| Ash (%)                         | 1.17 ± 0.04<sup>a</sup>                                                          | 1.18 ± 0.01<sup>a</sup>  |
| Crude fiber (%)                 | 0.40 ± 0.03<sup>a</sup>                                                          | 0.35 ± 0.01<sup>a</sup>  |
| Total carbohydrate (%)          | 73.49 ± 1.51<sup>a</sup>                                                          | 73.24 ± 1.49<sup>a</sup> |
| Total phenolic content (mg gallic acid/g) | 0.50 ± 0.01<sup>b</sup>                      | 0.82 ± 0.01<sup>a</sup>  |

<sup>a,b</sup>Different superscripts are significantly different (*p* < 0.05)
After drinking the refreshing drink were considered to improve the effect of refreshing perception on mental energy in terms of cortical activation and cognitive performance. It was demonstrated that drinking the refreshing drink could increase the alpha and beta wave activity in the brain. In addition, a study by Murao et al. (2013) showed that smelling green tea, especially shaded white tea, could also increase the alpha and beta brain wave activity in the frontal and occipital regions.

Moreover, the results of this study illustrated that the measurement of brain activity by EEG techniques can provide useful data to support the understanding of consumer responses to cooked brown rice treated with M. alba oil vapour in terms of brain function. This technique is suggested as an interesting technique that can be applied in the consumer and sensory research of food products.

**Conclusion**

*M. alba* oil vapour provided effective antifungal activity against *A. flavus* (≥ 450 µL/L) on MEA and brown rice. Furthermore, consumers accepted cooked brown rice treated with *M. alba* oil vapour at concentrations 300–600 µL/L (optimal at 600 µL/L), and a rejection threshold of 2052 µL/L was determined. EEG results suggested that the consumption of cooked brown rice treated with *M. alba* oil vapour at 600 µL/L could increase the power of alpha and beta waves in the human brain, promoting anti-stress effects and a relaxed mood. Therefore, *M. alba* oil vapour demonstrated good potential to enhance consumer preference for cooked brown rice and control significant growth of *A. flavus* in brown rice and
may prove useful in the improvement of brown rice quality.

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Table 4 Power of slow–fast alpha and low-mid-high beta waves during and after consumption of cooked brown rice

| Step                        | Frequency (Hz) | Power (µV²) | Untreated brown rice | Treated brown rice | P value |
|-----------------------------|---------------|-------------|----------------------|-------------------|---------|
|                             |               |             | Mean SEM             | Mean SEM          |         |
| Smelling odour              | 9             | 0.0903      | 0.0072               | 0.1521            | 0.0124  | < 0.0001 |
|                             | 12            | 0.1037      | 0.0066               | 0.0735            | 0.0048  | 0.4457   |
|                             | 14            | 0.0237      | 0.0020               | 0.0252            | 0.0014  | 0.5059   |
|                             | 17            | 0.0245      | 0.0014               | 0.0198            | 0.0010  | 0.9416   |
|                             | 25            | 0.0174      | 0.0014               | 0.0109            | 0.0007  | 0.8807   |
| Mouthing                    | 9             | 0.1104      | 0.0109               | 0.1135            | 0.0090  | < 0.0001 |
|                             | 12            | 0.0954      | 0.0095               | 0.0894            | 0.0070  | 0.4817   |
|                             | 14            | 0.0274      | 0.0017               | 0.0218            | 0.0016  | < 0.0001 |
|                             | 17            | 0.0218      | 0.0011               | 0.0179            | 0.0010  | < 0.0001 |
|                             | 25            | 0.0124      | 0.0006               | 0.0123            | 0.0009  | 0.0803   |
| After swallowing            | 9             | 0.1314      | 0.0100               | 0.1519            | 0.0166  | 0.2189   |
|                             | 12            | 0.1288      | 0.0102               | 0.1029            | 0.0113  | 0.0003   |
|                             | 14            | 0.0214      | 0.0013               | 0.0409            | 0.0032  | 0.0224   |
|                             | 17            | 0.0188      | 0.0010               | 0.0189            | 0.0009  | 0.7411   |
|                             | 25            | 0.0095      | 0.0005               | 0.0134            | 0.0007  | < 0.0001 |
| After swallowing 5 min      | 9             | 0.0916      | 0.0076               | 0.0940            | 0.0068  | 0.0347   |
|                             | 12            | 0.0837      | 0.0061               | 0.0713            | 0.0043  | 0.1613   |
|                             | 14            | 0.0159      | 0.0007               | 0.0211            | 0.0010  | 0.1195   |
|                             | 17            | 0.0169      | 0.0008               | 0.0150            | 0.0007  | 0.1909   |
|                             | 25            | 0.0131      | 0.0007               | 0.0127            | 0.0008  | 0.0128   |
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