Beyond GLMs: A Generative Mixture Modeling Approach to Neural System Identification

Lucas Theis1,2, André Maia Chagas1,3,4, Daniel Arnstein3,4, Cornelius Schwarz1,3,5, Matthias Bethge1,5,6

1 Werner Reichardt Centre for Integrative Neuroscience, Tübingen, Germany, 2 Graduate School of Neural Information Processing, University of Tübingen, Tübingen, Germany, 3 Hertie Institute for Clinical Brain Research, Tübingen, Germany, 4 Graduate School of Neural and Behavioural Sciences, University of Tubingen, Tübingen, Germany, 5 Bernstein Center for Computational Neuroscience, Tübingen, Germany, 6 Max Planck Institute for Biological Cybernetics, Tübingen, Germany

Abstract

Generalized linear models (GLMs) represent a popular choice for the probabilistic characterization of neural spike responses. While GLMs are attractive for their computational tractability, they also impose strong assumptions and thus only allow for a limited range of stimulus-response relationships to be discovered. Alternative approaches exist that make only very weak assumptions but scale poorly to high-dimensional stimulus spaces. Here we seek an approach which can gracefully interpolate between the two extremes. We extend two frequently used special cases of the GLM—a linear and a quadratic model—by assuming that the spike-triggered and non-spike-triggered distributions can be adequately represented using Gaussian mixtures. Because we derive the model from a generative perspective, its components are easy to interpret as they correspond to, for example, the spike-triggered distribution and the interspike interval distribution. The model is able to capture complex dependencies on high-dimensional stimuli with far fewer parameters than other approaches such as histogram-based methods. The added flexibility comes at the cost of a non-concave log-likelihood. We show that in practice this does not have to be an issue and the mixture-based model is able to outperform generalized linear and quadratic models.

Introduction

To account for the stochasticity inherent to neural responses, single cells as well as populations of cells are often characterized in terms of a probabilistic model. A popular choice for this task are generalized linear models (GLMs) and related approaches [1–6]. These models can often be seen as a convex optimization problem where a global optimum can be found. This guarantee comes at a price, as GLMs tightly constrain the computations which can be performed on the input. More complex computations can nevertheless be implemented by choosing a nonlinear feature representation of the input which is then fed into the linear model. In practice, however, it is typically very challenging to select the appropriate feature space because it presupposes a deeper understanding of the cell’s nonlinear behavior or unfeasibly large amounts of data.

Several approaches have been suggested to overcome the limitations of the generalized linear model. A natural extension is given by generalized quadratic models [7–9]. While a quadratic model represents a true generalization of a linear model, it can also be viewed as a linear model with a quadratic extension of the feature space (and, depending on the parametrization, some additional constraints on the parameters). Consequently, it shares similar limitations. A linear combination of quadratic features might still fail to represent the kind of stimulus properties a neuron responds to, but going to higher-dimensional general-purpose feature spaces quickly leads to overfitting. The number of parameters which need to be estimated grows linearly with the stimulus dimensionality in a linear model, quadratically in a quadratic model, and correspondingly faster if one uses a feature space of higher order.

An alternative approach is offered by nonparametric methods such as maximally informative dimensions (MID) [10]. Here, one first seeks a projection of the stimulus onto a lower-dimensional subspace such that as much information as possible is retained about the presence or absence of a spike. Afterwards, histograms are used to map out the nonlinear dependence of the neuron on the projected stimulus. This approach has the advantage that it can, at least in principle, capture arbitrary dependencies on the stimulus. However, the number of parameters that need to be estimated grows exponentially with the dimensionality of the stimulus subspace. This limits the approach to cells which are selective for only a few stimulus dimensions, although nonlinear extensions of this approach exist [11].

Here, we explore a different tradeoff. We derive a much more flexible neuron model for single cells which can, at least in principle, approximate arbitrary dependencies on the stimulus. The model can be viewed as generalizing generalized linear and quadratic models, but in contrast to quadratic models cannot easily be reduced to a GLM by choosing a different representation.
Author Summary

An essential goal of sensory systems neuroscience is to characterize the functional relationship between neural responses and external stimuli. Of particular interest are the nonlinear response properties of single cells. Inherently linear approaches such as generalized linear modeling can nevertheless be used to fit nonlinear behavior by choosing an appropriate feature space for the stimulus. This requires, however, that one has already obtained a good understanding of a cell's nonlinear properties, whereas more flexible approaches are necessary for the characterization of unexpected nonlinear behavior. In this work, we present a generalization of some frequently used generalized linear models which enables us to automatically extract complex stimulus-response relationships from recorded data. We show that our model can lead to substantial quantitative and qualitative improvements over generalized linear and quadratic models, which we illustrate on the example of primary afferents of the rat whisker system.

In the following, we will derive this linear model from a generative modeling point of view. This will help to motivate and see the connections to the extension presented later.

Let us consider the distribution over the stimulus $x$ conditioned on $y$. If $y$ equals 1, this distribution corresponds to the spike-triggered distribution. If $y$ equals 0, we will call it the non-spike-triggered distribution. At least for the moment, let us assume that both distributions are Gaussian, that is,

$$p(x|y) = N(\mu_y, \Sigma_y)$$

with means $\mu_y, \mu_1$ and covariances $\Sigma_y, \Sigma_1$. Bayes’ rule allows us to turn these assumptions into a probabilistic model of the neuron’s behavior,

$$p(y|x) = \frac{p(x|y)p(y)}{p(x)}$$

Using a few simple calculations, the probability of observing a spike, or firing rate, can be seen to be

$$p(y=1|x) = \sigma(f(x)),$$  \hspace{1cm} (2)

where

$$f(x) = x^T K x + w^T x + a,$$  \hspace{1cm} (4)

where we have performed the reparametrization

$$K = \frac{1}{2} (\Sigma_0^{-1} - \Sigma_1^{-1}),$$

$$w = \Sigma_1^{-1} \mu_1 - \Sigma_0^{-1} \mu_0,$$

and the bias term is given by

$$a = \frac{1}{2} \mu_0^T \Sigma_0^{-1} \mu_0 - \frac{1}{2} \mu_1^T \Sigma_1^{-1} \mu_1 + \frac{1}{2} \log \frac{\Sigma_1}{\Sigma_0} + \log \frac{p(y=1)}{p(y=0)}.$$  

If the spike-triggered and non-spike-triggered covariances are assumed identical, the quadratic term vanishes and we obtain the linear-nonlinear-Bernoulli model from above. Without this assumption, we are left with a quadratic model [7–9].

The unconstrained quadratic model is equivalent to a GLM with a quadratic extension of the feature space, since

$$x^T K x = \sum_{ij} K_{ij} x_i x_j$$  \hspace{1cm} (5)

is linear in the parameters $K_{ij}$. In practice, $K$ is often replaced by a low-rank approximation $\sum_{m=1}^M \beta_m u_m v_m^T$ [7–9,12], where $M$ controls the rank. The quadratic form in this case is given by

$$f(x) = \sum_{m} \beta_m (u_m x)^2 + w^T x + a.$$  \hspace{1cm} (6)
When choosing this parametrization, the optimization is no longer a convex problem [9] and the model no longer a GLM. In the following, we will use “quadratic model” only to refer to the unconstrained version—a GLM with a quadratic feature space—and “linear model” to refer to the GLM without quadratic features.

Spike-triggered mixture model

The generative point of view immediately suggests generalizations by loosening the assumptions of Gaussian distributed spike-triggered and non-spike-triggered stimuli. In the following, we consider mixtures of Gaussians as possible candidates,

\[ p(x|y) = \sum_k z_k \mathcal{N}(x; \mu_k, \Sigma_k). \]

Mixture models provide a good compromise between the assumptions of the tightly constrained generalized linear models and nonparametric approaches such as histograms. By plugging the mixture distributions into Equation 3, we obtain a new neuron model whose complexity can be controlled by adjusting the number of mixture components. We dub this model the spike-triggered mixture model (STM).

In the same manner that we have derived a model for the neuron’s dependency on the stimulus, we can incorporate dependence on other features as well. Let \( \tau \) be the time past since the neuron fired its last spike. Using Bayes’ rule, we obtain

\[ p(y|x, \tau) \propto p(x|y)p(\tau|y)p(y), \]

where here we have made the additional assumption that \( x \) and \( \tau \) are independent given \( y \). This assumption is also known as the naive Bayes assumption and is often employed in classification. It has empirically been observed that naive Bayes classifiers often perform well even when the assumption of independence is not met [13,14].

Taken together, the input to the sigmoid nonlinearity (Equation 2) is given by

\[ f(x, \tau) = \log \frac{\sum_k z_k \mathcal{N}(x; \mu_k, \Sigma_k)}{\sum_k z_k \mathcal{N}(x; \mu_k, \Sigma_k)} + \log h_1 + \log \frac{\pi}{1 - \pi}, \tag{7} \]

where \( \pi \) represents the prior probability of observing a spike and we have used histograms \( h_0 \) and \( h_1 \) to represent the interval distributions, \( p(\tau|y) = h_\tau \) (Figure 1). Note that if we do not constrain the parameters, there are several redundancies in this parametrization. For example, we can multiply both \( h_1 \) and \( h_0 \) by a common factor without changing the model’s predictions. If we reparametrize the model to get rid of redundancies and in addition assume that one mixture component is enough to represent the non-spike-triggered distribution, the input to the sigmoid takes the much simpler form

\[ f(x, \tau) = \log \sum_k \exp \left[ x' K_k x + w_k' x + a_k \right] + \log h_1. \tag{8} \]

The assumption of Gaussian distributed non-spike-triggered stimuli is sensible, for instance, if an \textit{a priori} Gaussian distributed stimulus is used to drive the neuron and the width of each bin of the spike train is small such that the posterior probability of observing a spike is generally also small, since in this case

\[ p(x|y = 0) \propto p(y = 0|x)p(x) \approx p(x). \]

The spike history dependent term on the right-hand side of Equation 8 can also be written in terms of a linear filter,

\[ \log h_1 = v \phi(z), \]

where \( z \in \{0,1\}^T \) represents the spike history, and \( \phi(z) = e_1 \) is the unit vector with zeros everywhere except at the position of the most recent spike. That is, the only difference to a linear model with history dependent term \( v^T z \) is that here all but one spike are suppressed by \( \phi \). In our experiments, we found that the two forms of spike history dependency worked equally well for most cells.

It is instructive to compare Equation 8 with Equation 4. While the quadratic model can be cast into the form of a linear model with a quadratic feature space, this is in general not possible for the STM. The function \( \log \sum_k \exp f_k \) is also known as soft maximum, since it can be viewed as a smooth approximation to the maximum of the \( f_k \). Our model is thus effectively taking the maximum of the responses of a number of quadratic models. Also note that the number of parameters is only a constant times the number of parameters of the quadratic model, which means it still grows only quadratically in the number of stimulus dimensions. But the number of parameters can be reduced further, as discussed in the next section.

Reducing the number of parameters

Assuming a single non-spike-triggered mixture component as in Equation 8 and ignoring the spike history for the moment, the number of parameters of the STM grows as \( O(KN^2) \), where \( N \) is the stimulus dimensionality and \( K \) is the number of mixture components. This growth might still be impractical where \( N \) is

![Figure 1. Illustration of the spike-triggered mixture model (STM). A. A sigmoidal nonlinearity is applied to a log-likelihood ratio of two mixtures of Gaussians to determine the firing rate of the model, which is then used to generate spikes. B. By making a naive Bayes assumption, additional information and measurements such as inter-spike interval distributions can easily be incorporated into the model in the form of additional log-likelihood ratios.

doi:10.1371/journal.pcbi.1003356.g001](https://journals.plos.org/ploscompbiol/article/file?jid=pcbi&doi=10.1371/journal.pcbi.1003356&fileId=10.1371/journal.pcbi.1003356.g001)
large or the amount of available data is small, as is often the case with neural data.

To reduce the number of parameters, we can employ the same trick as for the quadratic model and replace the matrices \( K_k \) by low-rank approximations (Equation 6). If we additionally share features \( u_m \) between the different components, we obtain

\[
f(x) = \log \sum_k \exp \left( \sum_m \beta_m (u_m^T x)^2 + w_k^T x + a_k \right). \tag{9}
\]

The number of parameters now grows as \( O(K \cdot M + M \cdot N + K \cdot N) \), where \( M \) is the number of quadratic features \( u_m \) contributing \( M \cdot N \) parameters, \( K \cdot M \) is the number of coefficients \( \beta_{\text{lin}} \), and \( K \cdot N \) is the number of parameters added by the linear features \( w_k \).

That is, for fixed \( M \) and \( K \), the number of parameters is linear in the number of stimulus dimensions. We will refer to this variant of the model as the factorized STM.

**Experimental setup**

We tested our model on spike trains obtained from 18 whisker-sensitive trigeminal ganglion cells of adult Sprague-Dawley rats. Recordings were made with a single electrode (sampling frequency: 20 kHz, bandpass filter: 300–5000 Hz). Manual stimulation was used to identify which whisker the neuron innervated as well as the approximate preferred direction of the whisker, after which the whisker was placed inside a plastic tube driven by a metal stimulator arm. The stimulator arm was programmed to deliver low-pass filtered (100 Hz) Gaussian white noise stimulation along the neuron’s preferred movement direction. Stimulation was divided into 50 unfrozen trials in which the stimulation sequence varied between trials, and 50 frozen trials in which a Gaussian white noise sequence was generated for the first trial only and then repeated for each subsequent trial. Spikes were extracted offline on the basis of waveform shape and all cells were categorized as either slowly adapting (SA) or rapidly adapting (RA). Example spike trains of two cells for frozen stimuli are shown in Figure 2.

We extracted 10 ms windows from the stimulus and reduced their dimensionality by keeping the first 10 principal components (>99.99% explained variance). We also extracted 25 ms of the spike history preceding each bin of the spike train. The dimensionality of the spike history was reduced to 100 by binning spikes into 100 equally sized bins of 250 µs width (no bin contained more than 1 spike). We then removed all but the most recent spike from the spike history and used this as input to all models. A linear projection of this vector is equivalent to the form of spike history dependency in Equation 8.

Filters of generalized linear models were first trained assuming a sigmoid nonlinearity. Together with a Bernoulli output distribution, this guarantees a concave log-likelihood such that an optimal solution can be found. Afterwards, we replaced the sigmoid nonlinearity with a more flexible nonlinearity consisting of a sum of Gaussian blobs,

\[
g(x) = \tanh \left( \sum_i \gamma_i \exp \left( -\frac{\lambda_i}{2} (x - \mu_i)^2 \right) \right),
\]

where the hyperbolic tangent ensures that the predicted probability of a spike does not exceed 1. We jointly optimized the parameters of this nonlinearity and the linear filter by alternately maximizing the average log-likelihood of the linear-nonlinear model using limited-memory BFGS [15], a standard quasi-Newton method (see Text S1 of the supporting information for gradients of the parameters). In a final step, we used a nonparametric histogram estimate (150 bins) to map out the nonlinearity. Through this multi-step procedure we tried to maximize the chances of finding a linear-nonlinear description of a neuron’s behavior where one exists. Note that strictly speaking, this model is no longer a generalized linear model (since the nonlinearities used are not invertible and the nonlinearities’ parameters are optimized). Quadratic models were optimized using the same procedure after extending the input by quadratic features.

The parameters of the STM (Equation 7) were initialized by estimating the spike-triggered, non-spike-triggered, and interspike interval distributions. Mixtures of Gaussians were fitted using standard expectation maximization [14,16] and interval distributions were estimated using histograms. While naive Bayes classifiers often already work well, it can be beneficial to directly optimize the conditional log-likelihood [17]. After initializing the parameters, we thus discriminatively finetuned the parameters using BFGS [18]. We found that this indeed helped to substantially improve the performance where the model depended on both the stimulus and the spike history.

We used between three and five components for the spike-triggered distribution and one and two components for the non-spike-triggered distribution, which was found to work well in preliminary runs on a different but related dataset with similar stimuli. Using two non-spike-triggered components increased the stability of the optimization for some cells. Finally, factorized STMs were trained discriminatively using limited-memory BFGS with randomly initialized parameters.

All models were trained on the 50 unfrozen trials and performance was evaluated based on the 50 frozen trials.

**Results**

We qualitatively and quantitatively compare the performance of the generalized linear, quadratic and spike-triggered mixture model (STM) for different cells and find in both cases that the STM can lead to substantial improvements.

**Qualitative comparison**

Figure 2 shows spike trains generated by the different models when fitted to one SA cell and one RA cell. The trial-to-trial variability of the responses of most cells in the dataset is quite low. This behavior is well captured by the STM, while the responses of the generalized linear and quadratic models generally seem to be noisier. This difference is more pronounced for SA cells than for RA cells, where all models appear to give a reasonably good fit. Corresponding peristimulus time histograms (PSTHs) can be seen in Figure 3 (details on how the PSTHs were computed are given in the next section).

Similar conclusions can be drawn by looking at spike-triggered distributions (Figure 4). Ensembles of spike-triggered positions \( x_t \) and velocities \( \dot{x}_t \) of the time-varying stimulus suggest a complex dependency of the responses on the stimulus for at least some cells. Note, however, that even a linear neuron can produce non-Gaussian spike-triggered distributions when the stimulus is correlated over time and the cell's firing depends on its history of generated spikes. Also note that while here we show 2-dimensional spike-triggered distributions, the input to the models was a 10-dimensional stimulus (and a 100-dimensional spike history), as described above.

To get a better intuition for the degree of nonlinearity of a cell, we can compare the cell’s spike-triggered distribution with the spike-triggered distribution of the best matching linear model. In
The given examples, the linear model is unable to reproduce the spike-triggered distributions of the cells displayed in Figure 4. For the SA cell, even the quadratic model fails to reproduce many of the features of the neuron's spike-triggered distribution, while the STM's behavior much more closely resembles that of the real cell.

**Quantitative comparison**

To quantify the performance of the different models, we estimate the cross-entropy or negative log-likelihood,

\[ E[-\log p(y|x)] \]  

where the expectation is taken over stimuli \( x \) and spikes \( y \) generated by the real neuron. We estimate this quantity using 50 frozen trials not used during training of the model. The cross-entropy is a natural measure for comparing different models, as it is the measure which is optimized during maximum likelihood
estimation of the parameters, and many other system-identification approaches such as spike-triggered averaging can often be viewed as performing maximum likelihood or penalized maximum likelihood learning [19].

The cross-entropy can be used to lower-bound the mutual information between stimulis and spikes,

\[ I[y,x] = H[y] - H[y|x] \geq H[y] - H[y|x] \]

The better a model distribution \( p(y|x) \) approximates a cell’s behavior, the smaller the difference will be between the lower bound and the true information transmitted by the cell. Note that this mutual information only quantifies the information carried by one bin of the spike train while we are generally more interested in the information carried by an entire spike train, \( y \in \{0,1\}^N \).

The spike train’s mutual information with the stimulus can be decomposed as follows

\[ I[y,x] = \sum_t I[y_t,x[y_{<t}]] \]

where \( y_{<t} \) denotes the history of spikes preceding time \( t \). To correctly quantify the mutual information between the spike train and the stimulus, it is thus important to take spike history effects into account. If we also use the fact that a neuron’s firing will only be affected by the stimulus preceding a spike, \( x_{<t} \), we get

\[ I[y_t,x[y_{<t}]] = H[y_t|x[y_{<t}]] - H[y_t|x_{<t},y_{<t}] \]

for the mutual information of the spike train per time bin. Dividing by the bin width yields an information rate (measured in bits per second or similar). Estimating this quantity requires two models: one for approximating the distribution \( p(y_t|y_{<t}) \) and one for approximating \( p(y_t|x_{<t},y_{<t}) \). A model for the former can take the form of Equation 8 but with the stimulus-dependent terms dropped.

Results averaged over all recorded SA cells (\( N = 8 \)) and all RA cells (\( N = 10 \)) are given in Figure 5. The average improvement of the STM over the quadratic model is 45.40 bit/s for SA cells and 15.48 bit/s for RA cells (for models taking into account spike history). The improvement for the cell with the largest difference to the quadratic model is 95.15 bit/s for SA cells and 43.05 bit/s for RA cells (the cells displayed in Figures 2 to 4). The firing rates of these two neurons were 117 Hz and 52.6 Hz, respectively, so that both numbers roughly correspond to 0.8 bit/spike improvement.

In addition to comparing different models, we can also compute and compare our model’s performance to the cross-entropy of a PSTH, which has also been called oracle model [20]. We computed PSTHs by convolving the average response to the frozen stimulus with a Gaussian kernel. We took all but one trial to compute the PSTH, which has also been called oracle model [20]. We computed PSTHs by convolving the average response to the frozen stimulus with a Gaussian kernel. We took all but one trial to compute the PSTH, and compare our model’s performance to the cross-entropy of the PSTH. That is, the probability of a spike at time \( t \) in trial \( i \) was predicted to be

\[ p(y_i = 1|x) = \frac{1}{J-1} \sum_j \sum_t y_j k_o(t,s) \]

where \( J \) is the number of trials and \( k_o \) is a normalized Gaussian kernel of width \( \sigma \),

\[ k_o(t,s) \propto \exp\left(-\frac{(t-s)^2}{2\sigma^2}\right), \quad \sum_s k_o(t,s) = 1. \]

For spike counts larger than 1, the same approach could be taken by using the right-hand side of Equation 11 as the rate parameter of a Poisson distribution. We found it was necessary to add a small offset to the PSTH to achieve good results. Both the offset and the
kernel width were automatically chosen from a prespecified set of parameters to minimize the cross-entropy averaged over all trials. That is, for each individual cell, we chose the kernel width with the best prediction performance. The optimal kernel widths were found to be around 0.12 ms and 0.09 ms (full width at half maximum, FWHM) for the SA and the RA cell displayed in Figure 3, respectively.

While the performance of the PSTH does not give us a guaranteed lower bound on the achievable cross-entropy, it gives us a very optimistic estimate of the performance that can be achieved by a model which does not take spike history into account. We found that the PSTH yielded a significantly lower cross-entropy than an STM without history dependency ($p < .01$), but not significantly lower than an STM which takes spike history into account ($p = .32$ and $p = .58$, respectively; Figure 3C and D).

PSTHs for model cells were estimated from 1000 spike trains (sampling spike trains was necessary since the models depend on the spike history) using the same kernel as for the real cell. The variance explained ($R^2$) by the generalized linear model, quadratic model and STM was 0.15, 0.26, and 0.47 for the SA cell, and 0.19, 0.41, and 0.5 for the RA cell (Figure 3), respectively. Note that the explained variance depends heavily on the chosen kernel width and wider kernels would yield larger coefficients.

How much data is enough?

The high firing rate of the cells and the resulting abundance of data allowed us to neglect regularization and overfitting issues. The training set contained on average about 25,000 spikes for SA cells and 6,700 spikes for RA cells. However, typically much less data is available.

To counter overfitting, different approaches to regularization can be taken. We already suggested reducing the number of parameters of the STM via factorization and parameter sharing (Equation 9). To get an idea of how the factored STM’s performance behaves as a function of the available data, we artificially reduced the amount of data by randomly picking a subset of the 50 training trials. Of that subset, we used 50% for validation and 50% for optimization. During optimization, the performance on the validation set was tested every 5 iterations. If it decreased 50 times in a row, training was stopped and the parameters with the lowest validation error until then were kept. Other than early stopping, no other form of regularization was used. The test set was the same as the one used in Figure 5.

Figure 6 shows the performance of the factored STM for different amounts of spikes in the training set. The factored STM used 6 components and 5 quadratic features (246 parameters in total) for the SA cell, and 3 components and 5 quadratic features...
(198 parameters) for the RA cell. For comparison, we also plot the performance of a generalized linear model (111 parameters) trained with early stopping on a subset of the training data, as well as the performance of non-factored STMs (532 parameters and 400 parameters, respectively) and quadratic (156 parameters) models trained on the entire training set.

For the SA cell, the performance of the factored STM started to decrease more rapidly as soon as less than 5,000 spikes were present in the training set. However, even with 2,500 spikes the average performance was still much better than the performance of a quadratic model trained on the entire dataset. For the RA cell, the performance started to deteriorate at about 2,000 spikes. Note that the performance of the linear model worsened at a similar rate. Reducing the number of parameters further by using half the spike history or six instead of ten principal components to represent the stimulus did not help to improve performance in the regime of few data points. The performance might however be improved by choosing suitable priors for the parameters, which we did not explore here.

Training with half the dataset of the RA cell (about \(2.5 \times 10^6\) data points) on average took 9.4 minutes for the factored STM and 2.7 minutes for the linear model with parallelized implementations written in C++ when run on a machine with 12 Intel Xeon E5-2630 cores (2.3 GHz).

Discussion

We have shown that a spike-triggered mixture model can lead to better performance than either linear or quadratic models, which we illustrated on the example of rat primary afferents. A possible explanation for the improved performance might be that our model can better cope with a cell's adaptation to the stimulus. Because the firing rate of our model is effectively a maximum over a number of quadratic models, the model is able to respond differently in different regions of the stimulus space. Our model may yield even bigger improvements when applied to cells higher up the hierarchy—such as cortical cells—where highly nonlinear dependencies on the stimulus are to be expected [21]. In particular, an interesting empirical question is whether STMs will be able to improve upon quadratic models in modeling complex cells [22]. As a generalization, the STM can capture the same kind of invariances that the quadratic model can capture, but in addition allows us to spend parameters in different ways by adding components instead of quadratic feature dimensions.

Here, we chose to give up on the constraint of convexity to be able to build a more flexible neuron model. In practice, non-convex or even multimodal likelihoods do not have to be an issue. Many local optima of the STM likelihood are created simply by permutations of the parameters of the different mixture components and are therefore unproblematic. We found that initializing mixture models with EM and fine-tuning with an off-the-shelf optimizer worked well for our data and the performance of the resulting model was stable across different initializations. The parameters of the factored variant of the STM (Equation 9) were randomly initialized and gave comparable results (Figure 6).

Alternatively, we could have used support vector machines, kernel logistic regression (KLR) [23] or other kernel based approaches [24] for gaining flexibility while retaining convexity. In KLR, the input to the sigmoid (Equation 2) determining the firing rate takes the form

\[
f(x) = \sum_i w_i k(x_i, x) + b,
\]

where \(i\) indexes training points \(x_i\) and \(k\) is a kernel measuring the similarity between stimuli or, more generally, inputs to the neuron. If a Gaussian RBF kernel is used, KLR becomes similar to an STM with all covariance matrices constrained to a multiple of the identity matrix and one mixture component placed on top of each data point (cf. Equation 8).

KLR is equivalent to a linear-nonlinear-Bernoulli model with a cleverly chosen feature space whose dimensionality grows with the number of data points. Hence, one advantage of KLR is that its objective function is convex. Advantages of a parametric model like the one presented in this paper are more readily interpretable parameters and lower computational costs when the number of training points is large. Ultimately, whether kernel based methods or a generative approach should be preferred presumably depends on whether one has a better intuition of what represents a good kernel for the input space, or a better intuition of what represents a good characterization of the spike-triggered distribution.

Figure 6. Performance as a function of available data. The factored STM was trained with different random subsets of the training trials and evaluated on all test trials for one SA cell (A) and one RA cell (B). The horizontal axis represents the number of spikes in the training set. Shown are the average performances (solid blue line) along with 90% confidence intervals (5th and 95th percentile). For comparison, we also show the performance of the linear model trained with different subsets of the data, the average performances of the non-factored STM, and the quadratic model trained on the entire training set. Note that the factored STM outperforms the generalized linear model even when only a small fraction of the dataset is used.

doi:10.1371/journal.pcbi.1003356.g006
The idea of using spike-triggered distributions to construct and motivate neuron models is not new. However, most work in this direction has focused on spike-triggered averages and covariances [25–29]. Here we used mixtures of Gaussians and histograms to derive a new neuron model, but other distributions might work better in a different context and might be worth exploring.

Yet another related approach is to use feed-forward neural networks [30–32]. While standard feed-forward neural networks are in principle also able to represent arbitrarily complex stimulus-response relationships [33], one can hope to get away with fewer parameters, less data, or simpler optimization schemes when using a model tailored to the task at hand. In contrast to general nonlinear regression strategies, a generative approach can lead to much more problem-specific architectures and nonlinearities (Equations 8 and 9). Similar cascades of linear-nonlinear units have been proposed but motivated by physiological rather than statistical considerations [20,34–36].

STMs can easily be extended to model populations of neurons similar to how GLMs are extended to populations by introducing coupling filters [5,37]. Analogous to how we incorporated dependency on the spike history of a single neuron, a form for the dependency between neurons can also be motivated via a log-likelihood ratio for the distribution of crossinterspike intervals.

Code for fitting STMs is provided at http://bethgelab.org/code/theis2013a/.

Supporting Information

Text S1 Gradients for the log-likelihoods of the STM and factored STM. (PDF)

Author Contributions

Conceived and designed the experiments: LT AMC CS MB. Performed the experiments: LT AMC DA CS. Analyzed the data: LT AMC CS MB. Wrote the paper: LT MB.

References

1. McCullagh P, Nelder JA (1989) Generalized linear models. Chapman & Hall, second edition.
2. Nelder JA, Wedderburn RWJ (1972) Generalized linear models. Journal of the Royal Statistical Society, Series A (General) 135: 370–384.
3. Paninski L (2004) Maximum likelihood estimation of cascade point-process neural encoding models. Network: Computation in Neural Systems 15: 243–262.
4. Truccolo W, Eden UT, Fellows MR, Donoghue JP, Brown EN (2004) A point process framework for relating neural spiking activity to spiking history, neural ensemble, and extrinsic covariate effects. Journal of Neurophysiology 93: 1074–1089.
5. Pillow JW, Shlens J, Paninski L, Sher A, Litke AM, et al. (2008) Spatio-temporal correlations and visual signaling in a complete neuronal population. Nature 454: 995–999.
6. Gerwinn S, Macke J, Bethge M (2010) Bayesian inference for generalized linear models for spiking neurons. Frontiers in Computational Neuroscience 4: 12.
7. Pillow JW, Simoncelli EP (2006) Dimensionality reduction in neural models: An information-theoretic generalization of spike-triggered average and covariance analysis. Journal of Vision 6: 414–428.
8. Park IM, Pillow JW (2011) Bayesian spike-triggered covariance. In: Advances in Neural Information Processing Systems 24. pp. 1692–1700.
9. Rajan K, Marre O, Tkačik G (2013) Learning quadratic receptive fields from neural responses to natural stimuli. Neural Computation 25: 1661–1692.
10. Sharpee T, Rust NC, Bialek W (2004) Analyzing neural responses to natural signals: maximally informative dimensions. Neural Computation 16: 223–250.
11. Rajan K, Bialek W (2012). Maximaly informative stimulus energies in the analysis of neural responses to natural signals. arXiv:1201.0321.
12. Fitzgerald JD, Roskamp RJ, Sincich LC, Sharpee TO (2011) Second order dimensionality reduction using minimum and maximum mutual information models. PLoS Computational Biology 7(10): e1002249.
13. Zhang H (2004) The optimality of naive Bayes. In: Proceedings of the Seventeenth Florida International Artificial Intelligence Research Society Conference. AAAI Press, pp. 562–567.
14. Bishop CM (2006) Pattern Recognition and Machine Learning. Springer.
15. Byrd RH, Lu P, Nocedal J (1995) A limited memory algorithm for bound constrained optimization. SIAM Journal on Scientific and Statistical Computing 16: 1190–1208.
16. Dempster AP, Laird NM, Rubin DB (1977) Maximum likelihood from incomplete data via the EM algorithm. Journal of the Royal Statistical Society, Series B 39: 1–38.
17. Ng AT, Jordan MI (2002) On discriminative vs. generative classifiers: A comparison of logistic regression and naive Bayes. In: Advances in Neural Information Processing Systems 15. pp. 841–848.
18. Nocedal J, Wright SJ (2006) Numerical Optimization. Springer, second edition. 136–143 pp.
19. Wu MCK, David SV, Gallant JL (2006) Complete functional characterization of sensory neurons by system identification. Annual Review of Neuroscience 29: 477–503.
20. Vintch B, Zaharia AD, Movshon JA, Simoncelli EP (2012) Efficient and direct estimation of a neural subunit model for sensory coding. In: Advances in Neural Information Processing Systems 25. pp. 3113–3121.
21. Carandini M, Demb JB, Mante V, Tolhurst DJ, Dan Y, et al. (2005) Do we know what the early visual system does? The Journal of Neuroscience 25: 10577–10597.
22. Hubel DH, Wiesel TN (1962) Receptive fields, binocular interaction and functional architecture in the cat’s visual cortex. Journal of Physiology 160: 101–154.
23. Zha J, Hastie T (2001) Kernel logistic regression and the import vector machine. In: Advances in Neural Information Processing Systems 14. pp. 1081–1088.
24. Truccolo W, Donoghue JP (2007) Nonparametric modeling of neural point processes via stochastic gradient boosting regression. Neural Computation 19: 672–705.
25. de Ruyter van Steveninck R, Bialek W (1998) Real-time performance of a movement-sensitive neuron in the blowfly visual system: Coding and information transfer in short spike sequences. Proceedings of the Royal Society of London Series B, Biological Sciences 254: 379–414.
26. Brenner N, Bialek W, de Ruyter van Steveninck R (2000) Adaptive rescaling maximizes information transmission. Neuron 26: 695–702.
27. Simoncelli EP, Paninski L, Pillow J, Schwartz O (2004) The Cognitive Neurosciences, MIT Press, chapter Characterization of Neural Responses with Stochastic Stimuli. third edition, pp. 327–338.
28. Schwartz O, Pillow JW, Rust NC, Simoncelli EP (2006) Spike-triggered neural characterization. Journal of Vision 6: 484–507.
29. Fairhall AL, Berlinger CA, Narasimhan R, Harris RA, Puchalla JL, et al. (2006) Selectivity for multiple stimulus features in retinal ganglion cells. Journal of Neurophysiology 96: 2724–38.
30. Lelkes SR, Sejnowski TJ, Desimone R (1992) Predicting responses of nonlinear neurons in monkey striate cortex to complex patterns. Journal of Neuroscience 12: 3568–3581.
31. Lau B, Stanley GB, Dan Y (2002) Computational subunits of visual cortical neurons revealed by artificial neural networks. Proceedings of the National Academy of Sciences 99: 8974–8979.
32. Prenger R, Wu MCK, David SV, Gallant JL (2004) Nonlinear V1 responses to natural scenes revealed by neural network analysis. Neural Networks 17: 663–679.
33. Cybenko G (1989) Approximations by superpositions of sigmoidal functions. Mathematics of Control, Signals, and Systems 2: 303–314.
34. Gollisch T, Herz AVM (2005) Disentangling sub-millisecond processes within an animal’s visual transduction chain. PLoS Biology 3: e8.
35. Butts DA, Weng C, Jin J, Alonso JM, Paninski L (2011) Temporal precision in the visual pathway through the interplay of excitation and stimulus-driven suppression. The Journal of Neuroscience 31: 11313–11327.
36. McFarland JM, Cui Y, Butts DA (2013) Inferring nonlinear neuronal computation based on physiologically plausible inputs. PLoS Computational Biology 9: e1003143.
37. Stevenson IH, Rebec J, Miller LE, Körding KP (2008) Inferring functional connections between neurons. Current Opinion in Neurobiology 18: 582–588.

Beyond GLMs: A Generative Approach