Genome-Wide Association Study of Susceptibility Loci for Radiation-Induced Brain Injury

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Abstract

**Background:** Radiation-induced brain injury is a nonnegligible issue in the management of cancer patients treated by partial or whole brain irradiation. In particular, temporal lobe injury (TLI), a deleterious late complication in nasopharyngeal carcinoma, greatly affects the long-term life quality of these patients. Although genome-wide association studies (GWASs) have successfully identified single nucleotide polymorphisms (SNPs) associated with radiation toxicity, genetic variants contributing to the radiation-induced brain injury have not yet been assessed.

**Methods:** We recruited and performed follow-up for a prospective observational cohort, Genetic Architecture of Radiotherapy Toxicity and Prognosis, using magnetic resonance imaging for TLI diagnosis. We conducted genome-wide association analysis in 1082 patients and validated the top associations in two independent cohorts of 1119 and 741 patients, respectively. All statistical tests were two-sided.

**Results:** We identified a promoter variant rs17111237 (A > G, minor allele frequency [MAF] = 0.14) in CEP128 associated with TLI risk (hazard ratio = 1.45, 95% confidence interval = 1.26 to 1.66, $P_{\text{combined}} = 3.18 \times 10^{-7}$) which is in moderate linkage disequilibrium (LD) with rs162171 (MAF = 0.18, $R^2 = 0.69$), the top signal in CEP128 (hazard ratio = 1.46, 95% confidence interval = 1.29–1.66, $P_{\text{combined}} = 6.17 \times 10^{-9}$). Combining the clinical variables with the top SNP, we divided the patients into different subgroups with varying risk with 5-year TLI-free rates ranging from 33.7% to 95.5%.

**Conclusions:** This three-stage study is the first GWAS of radiation-induced brain injury that implicates the genetic susceptibility gene CEP128 involved in TLI development and provides the novel insight into the underlying mechanisms of radiation-induced brain injury.
challenges for the discovery of reliable biomarkers to predict radiation-susceptible patients and for the disclosure of the mechanism of radiation-induced brain injury. Temporal lobe injury (TLI), a deleterious late radiotherapy toxicity of nasopharyngeal carcinoma (NPC) (2,3) resulting from unavoidable radiation exposure of the inferior and medial aspects of temporal lobes, however, provides several unique conditions for such study, including no direct effects of brain tumors, longer survival of patients, and distinct phenotypic characterization. Although intensity-modulated radiotherapy (IMRT) with more controlled dose deposition in normal tissue has reduced the rate of TLI, there are still 3.2–12.9% of NPC survivors suffering TLI in months to years postradiotherapy (4–9). The TLI symptoms including dizziness, headache, memory impairment, and neurocognitive dysfunctions (10), varied among the patients, often irreversible and that negatively influence the patients’ quality of life.

Some clinical parameters including treatment protocols (11) and tumor stages (4) are risk factors of TLI development in NPC patients. However, even after these factors are adjusted, interpatient variability of TLI could not be fully explained, indicating a possible role of a genetic contribution at the individual-level radiosensitivity. Many genetic studies of radiation-induced adverse responses focused on candidate genes involved in DNA repair, DNA damage signaling, cell cycle control, and inflammatory response but many of them had limited sample sizes and suffered a lack of independent replication (12–15). Although several recent genome-wide association studies (GWAS) (16–21) have successfully identified single nucleotide polymorphisms (SNPs) associated with radiation toxicity in patients with prostate and/or breast cancer, the genetic impact of radiation-induced TLI is yet to be reported. Therefore, we initiated a prospective observational clinical trial, GARTP (Genetic Architecture of the Radiotherapy Toxicity and Prognosis) to identify the susceptibility genes contributing to TLI development.

Materials and Methods

The GARTP Study

The GARTP study (registered with www.chictr.org.cn/, ChiCTR-ROC-17012658) is a prospective observational clinical trial that aims to identify the susceptibility genes contributing to recurrence, metastasis, and radiation-induced normal tissue injury in NPC patients. This study was approved by the Human Ethics Approval Committee of Sun Yat-sen University Cancer Center (SYSUCC) on June 15, 2005 (No. YB2005001). The recruitment criteria are described in the Supplementary Methods (available online). Human glioblastoma U87 cell line and 293 T cell lines were purchased from the American Type Culture Collection (Manassas, VA) and were cultured in RPMI-1640 or Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10.0% fetal bovine serum (FBS, GibCO, Carlsbad, CA). All cell lines were maintained in a humidified incubator containing 5.0% CO2 at 37.0°C. For the radiation treatments, cells were irradiated with 6-MV X-rays from a Primus linear accelerator (Siemens, Malvern, PA) with a dose rate of 198 cGy/min.

Follow-up and Diagnosis for TLI

After completing therapy, the patients were followed up by MRI of the nasopharynx and/or neck regularly (maximum follow-up time of 15.0 years). We defined the time of TLI development as the time between the date of commencement of the primary radiotherapy and the time when TLI occurred. For patients who had not developed TLI before the end of the study (December 31, 2016), the end times were defined as the last MRI follow-up. For the patients who were treated with a second course of radiotherapy due to local recurrence, their times were defined as the duration between the beginning of the primary radiotherapy and the beginning of the second radiotherapy.

MRI was used for the diagnosis of TLI as previously described (3,4,23) (Supplementary Figure 1, available online). The radiologists and clinical radiation oncologists specializing in head-and-neck cancers evaluated the MR images independently. Any disagreement was resolved by consensus.

DNA Extraction, Genotyping, Imputation, and Quality Control

Detailed description of DNA extraction, genotyping, imputation, and quality control is shown in the Supplementary Methods (available online) and Supplementary Table 1 (available online).

Cell Lines, Constructs, and Functional Assays

For a full description of the laboratory experiments, please see the Supplementary Methods (available online). Human glioblastoma U87 cell line and 293 T cell lines were purchased from the American Type Culture Collection (Manassas, VA) and were cultured in RPMI-1640 or Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10.0% fetal bovine serum (FBS, GibCO, Carlsbad, CA). All cell lines were maintained in a humidified incubator containing 5.0% CO2 at 37.0°C. For the radiation treatments, cells were irradiated with 6-MV X-rays from a Primus linear accelerator (Siemens, Malvern, PA) with a dose rate of 198 cGy/min.
### Statistical Analysis

All the analysis procedures are shown in Supplementary Figure 2 (available online). GWAS was performed using Cox proportional hazard regression model (24) assuming additive SNP effect. The proportional hazard assumption was verified using Schoenfeld residuals. Two reported risk factors, tumor stage (T stage) and the radiation technique (IMRT or 2D-CRT) (4), as well as age at NPC diagnosis, were adjusted as covariates. For allele stage, SNPs with two-sided \( P \) curves were estimated by the R package “survminer.” In the dissertation using the R package “survival” and the adjusted survival hazard ratios and their 95% confidence intervals [CIs] were estimated using the R package “survminer,” as age at NPC diagnosis, were adjusted as covariates. Per allele effect. The proportional hazard assumption was verified using Schoenfeld residuals. Two reported risk factors, tumor stage (T stage) and the radiation technique (IMRT or 2D-CRT) (4), as well as age at NPC diagnosis, were adjusted as covariates. Per allele stage, SNPs with two-sided \( P \) curves were estimated by the R package “survminer.” In the discovery stage, SNPs with two-sided \( P \) less than \( 1.0 \times 10^{-5} \) were selected for validation. A quantile–quantile plot was used to evaluate the overall statistical significance of the GWAS and the deviation of observed vs expected distribution of \( P \) values was represented by inflation factor \( \hat{\lambda}_{QC} \) (25). Linkage disequilibrium (LD) between SNPs was calculated with the Haploview program (v. 4.1). The survival tree was constructed by using recursive partitioning under conditional inference framework using the R package “party.” For the stopping criterion, Bonferroni correction was used and the level of statistical significance was chosen as .01. We also restricted the maximum depth of the tree to three. A two-sided \( t \)-test was used to test the statistical significance between cell survival fractions of control and different shRNAs of CEPI28. Statistical significance was defined as \( P \) less than .05. Pathway/gene set analysis and expression quantitative trait locus (eQTL) analysis is described in the Supplementary Methods (available online). All statistical tests were two-sided.

### Results

We included 1082 NPC patients in the discovery stage (Figure 1) and the detailed clinical characteristics are shown in Table 1. The average MRI follow-up was 4.10 times and the median follow-up months were 30.7. Among 1082 patients, 243 (22.5%) developed TLI after radiotherapy. Tumor stage and radiation techniques have been reported to be associated with TLI risk in a previous study (4) and in this clinical trial we confirmed the results. The patients treated with IMRT had a lower TLI incidence, with a hazard ratio (HR) of 0.42 (95% CI = 0.28 to 0.64) compared with those treated by 2D-CRT. Patients with advanced tumor stage exhibited elevated TLI risks with HRs of 2.86 (T2 vs T1, 95% CI = 1.13 to 7.19), 4.30 (T3 vs T1, 95% CI = 1.74 to 10.63), and 7.65 (T4 vs T1, 95% CI = 3.05 to 19.21), respectively (Table 1). Additionally, we found that older people were more likely to develop TLI, with HRs of 1.54 (95% CI = 0.84 to 2.83), 2.08 (95% CI = 1.16 to 3.74), 2.31 (95% CI = 1.28 to 4.18), and 1.67 (95% CI = 0.77 to 3.65) for those in age groups of 31–40 years, 41–50 years, 51–60 years, and 61 years and older, compared with those age 30 years or younger, respectively (Table 1).

After quality controls, 445 078 autosomal SNPs were included in the genome-wide association analysis under an additive assumption using a Cox proportional hazard model (24,26), adjusting age at NPC diagnosis, radiation techniques, and tumor stages, following radiogenomics reporting guidelines (27). The distribution of observed vs expected \( P \) values are shown in the Supplementary Figure 2 (available online). We identified seven potentially statistically significant association signals with \( P \) values smaller than \( 1.0 \times 10^{-5} \) (Figure 2A and Supplementary Table 2, available online). Three of them were located at the introns of CEPI28 with high linkage disequilibrium (LD) (\( R^2 = 0.97–1.00 \)) and the signal could be represented by rs162171 with HR of the minor allele of 1.66 (minor allele frequency [MAF] = 0.17; 95% CI = 1.33 to 2.07) and \( P = 7.75 \times 10^{-6} \) (Table 2). Additionally, two intron SNPs of KCTD1 and one at DISC1FP1 were associated with TLI risk at a similar statistical significance level.

We then fine mapped 100 kb regions upstream/downstream of CEPI28, KCTD1, and DISC1FP1 by imputation (28) analysis. We searched the potential functional SNPs with \( P \) less than .001 in these regions and found three variants located at the promoter.

**Figure 1.** The flow diagram of the study design. NPC = nasopharyngeal carcinoma; GARTP = Genetic Architecture of the Radiotherapy Toxicity and Prognosis; SNP = single nucleotide polymorphism; GWAS = genome-wide association study; TLI = temporal lobe injury; MRI = magnetic resonance imaging; RT = radiotherapy.

| Discovery Stage | Replication Stage I | Replication Stage II |
|-----------------|----------------------|----------------------|
| 1166 eligible NPC patients were recruited for GARTP in 2005-2007 | 1164 eligible NPC patients were recruited for GARTP in 2008-2010 | 919 eligible NPC patients were collected in 2002-2004 & GARTP 2005-2007 |
| | Pre-treatment evaluation: | Collection of clinical records: |
| | (i) MRI of neck and nasopharynx; (ii) Medical history; (iii) Physical examination; | (i) MRI of neck and nasopharynx; (ii) Medical history; (iii) Physical examination; |
| | Regularly followed up the MRI of nasopharynx/brain | Regularly reviewed the MRI of nasopharynx/brain |
| | last follow-up date: 2016/12/31 | last follow-up date: 2016/12/31 |
| 1082 patients for GWAS study of TLI development | No MRI after RT (n=84) | No MRI after RT (n=45) |
| Median follow-up months: 30.7 | | |
| 1119 patients for validation of the candidate SNPs | No MRI after RT (n=45) | No MRI after RT (n=178) |
| Median follow-up months: 35.2 | | |
| 741 patients for validation of the candidate SNPs | No MRI after RT (n=178) | |
| Median follow-up months: 26.6 | | |
of CEP128 associated with TLI development (Supplementary Table 3, available online). Two SNPs, rs17111237 and rs17111246, located within 2 kb upstream of CEP128 (Figure 2B) were in nearly complete LD ($R^2 = 0.99$, Supplementary Figure 4, available online). The per-allele HR of the minor allele at rs17111237 was 1.63 (MAF = 0.13; 95% CI = 1.27 to 2.08; $P = 1.02 \times 10^{-4}$ (Table 2) and it showed moderate LD with the top signal rs162171 ($R^2 = 0.69$).

To confirm the above findings, we genotyped the identified SNPs in two stages, including 1119 NPC patients from GARTP study (2008–2010) and 741 NPC patients in 2002–2004 and in 2005–2007 (GARTP) (Figure 1 and Table 1). The average MRI follow-ups for patients in the first and the second replication stages are 4.98 and 3.01 months, respectively. All the SNPs in CEP128 were associated with TLI risk with $P$ less than .05. When combining the patients from three stages, we found that patients carrying the minor alleles at rs162171 or rs17111237 tend to have higher risks to develop TLI with per allele HRs of 1.46 (MAF=0.18, 95% CI = 1.29 to 1.66, $P_{\text{combined}}=6.17 \times 10^{-5}$) and 1.45 (MAF=0.14, 95% CI = 1.26 to 1.66, $P_{\text{combined}}=3.18 \times 10^{-5}$), respectively (Table 2, Figure 2C and Supplementary Figure 5, available online). None in KCTD1 or DSC1FP1 reached the statistical significance level of $P$ less than .05 in the replication analysis.

We further performed expression quantitative trait locus (eQTL) analysis and found that with the increasing number of risk alleles at rs162171 or rs17111237, CEP128 exhibited a lower mRNA expression level in human lymphoblastoid cell line (Hapmap database: http://www.hapmap.org/) and intralobular white matter of human brain tissue (BRAINEAC database, http://www.braineac.org/) (Figure 3A,B). Furthermore, the results of dual-luciferase assay showed that the transcription activities of promoter constructs harboring the risk allele of rs17111237 and rs17111246 were lower than that of the wildtype constructs (Figure 3C), indicating that individuals harboring risk alleles might present an impaired expression level and insufficient function of CEP128.

Radiation-induced brain injury has been speculated as a multistep process, during which direct damage and vascular abnormality may disturb the balance of oxidative metabolism, which could activate the release of reactive oxygen species and generate deleterious oxidative stress. To further investigate the role of CEP128 involved in radiation-induced brain injury, we imitated the direct radiation effect and oxidative stress effect on gliocytes by treating the glioblastoma cell line U87 with X-ray and H2O2. Our results of the clonogenic assay showed that RNAi inhibition of endogenous CEP128 expression reduced the survival fraction of U87 cell lines under the treatment of X-ray (Figure 3D,E). Additionally, we found that after being treated with X-ray or H2O2, a higher level of cell death and apoptosis was observed in U87 cell lines with RNAi inhibition of endogenous CEP128 expression, implicating a potential biological involvement of CEP128 in protecting normal temporal lobes against radiation-induced damage (Supplementary Figures 6 and 7, available online).

To further evaluate the underlying disease mechanisms responsible for the genetic signals, we applied gProfiler (29) and Gene-set Enrichment Analysis (GSEA) (30) to identify overrepresented pathways using SNPs located within 20 kb upstream/downstream of 212 candidate genes with $P < .001$ in the GWAS analysis. The top 20 enriched gene sets identified by gProfiler

Table 1. Characteristics and multivariable regression analysis of temporal lobe injury development in the discovery and replication stages

| Characteristics | Discovery stage | Replication stage I | Replication stage II | Combined |
|-----------------|----------------|---------------------|---------------------|----------|
| Number of patients | 243/839 | 261/858 | 177/564 | 681/2261 |
| Age, y | | | | |
| <30 | 13/68 | 16/74 | 8/49 | 37/191 |
| 31-40 | 56/241 | 69/240 | 55/166 | 180/647 |
| 41-50 | 85/256 | 84/278 | 70/172 | 239/706 |
| 51-60 | 76/181 | 71/189 | 34/113 | 181/483 |
| ≥61 | 13/93 | 27/111 | 10/64 | 44/234 |
| Sex | | | | |
| Male | 186/606 | 200/660 | 132/436 | 518/1702 |
| Female | 57/233 | 61/198 | 45/128 | 163/559 |
| Tumor stage† | | | | |
| T1 | 5/60 | 5/94 | 13/49 | 23/203 |
| T2 | 47/216 | 57/223 | 35/166 | 139/605 |
| T3 | 110/386 | 109/375 | 79/233 | 296/994 |
| T4 | 81/177 | 90/166 | 50/116 | 221/459 |
| Radiation technique | | | | |
| 2D-CRT | 215/679 | 214/541 | 173/525 | 602/1745 |
| IMRT | 28/160 | 47/317 | 4/39 | 79/516 |
| Treatment | | | | |
| RT alone | 58/248 | 41/148 | 55/215 | 154/611 |
| RT+IC/AC | 67/232 | 68/159 | 60/142 | 195/533 |
| CCRT | 118/359 | 152/551 | 62/207 | 332/1117 |

*Adjusted hazard ratios (HRs) and 95% CIs for TLI development were calculated by Cox proportional hazard model using variables including sex, age group, tumor stage, radiation technique, and treatment modality. CI = confidence interval, 2D-CRT = two-dimensional conformal radiotherapy; IMRT = intensity-modulated radiotherapy; RT = radiotherapy alone; RT+IC/AC = radiotherapy with induction and/or adjuvant chemotherapy; CCRT = concurrent chemoradiotherapy; TLI = temporal lobe injury.
†The patients were staged according to the 2002 6th USCC/AJCC staging system (22).
and GSEA are listed in Supplementary Tables 4 and 5 (available online). “GO neurogenesis” (27 of 212 genes, 12.7%) was identified by both softwares (Supplementary Table 6, available online). In addition, the candidate genes were also clustered in the gene sets such as “immune system process”, “programmed cell death”, “generation of neurons”, and “apoptotic process”.

Based on our genetic findings, we further divided the patients into six subgroups by using the genetic and clinical variables (Figure 2D) (31). For patients in the T1–T3 classification, the radiation technique was chosen as the next variable, where patients in the IMRT group carrying CC genotype at rs162171 have the lowest TLI risk (5-year TLI-free rate of 95.5%, 95% CI = 92.5 to 98.6%) and the risk elevated among those carrying the risk genotypes of rs162171 (5-year TLI-free rate of 82.1%, 95% CI = 73.2 to 92.0%). Unsurprisingly the patients in the T4 classification had the higher TLI risk. Among these patients, the ones who carried risk genotypes at rs162171 had the highest risk (5-year TLI-free rate of 33.7%, 95% CI = 25.6 to 44.4%). The resulting classification further indicated that the patients who have the same clinical characteristics could have varying TLI risk due to a different genetic background.
Table 2. Associations of SNPs with development of temporal lobe injury after radiation in nasopharyngeal carcinoma patients

| SNPs       | Locus | Stage   | MAF   | N   | TLI %  | N   | TLI %  | N   | TLI %  | HR (95% CI)† | P† |
|------------|-------|---------|-------|-----|--------|-----|--------|-----|--------|----------------|-----|
| rs162171   | CEP128| Discovery | 0.171 | 748 | 19.4%  | 297 | 29.3%  | 37  | 29.7%  | 1.66 (1.33 to 2.07) | 7.75 × 10^-6 |
| C/A‡       | Intron| Replication I | 0.194 | 714 | 22.0%  | 368 | 25.3%  | 32  | 34.4%  | 1.34 (1.09 to 1.65) | 0.005 |
|            |       | Replication II | 0.180 | 491 | 21.0%  | 211 | 30.3%  | 25  | 32.0%  | 1.43 (1.11 to 1.84) | 0.006 |
|            |       | Combined    | 0.182 | 1953| 20.7%  | 876 | 27.9%  | 94  | 31.9%  | 1.46 (1.29 to 1.66) | 6.17 × 10^-9 |
| rs1711237  | CEP128| Discovery | 0.131 | 788 | 20.4%  | 235 | 31.5%  | 19  | 21.1%  | 1.34 (1.09 to 1.65) | 0.005 |
| A/G‡       | Promoter| Replication I | 0.138 | 823 | 22.8%  | 271 | 24.0%  | 18  | 38.9%  | 1.32 (1.05 to 1.66) | 0.02  |
|            |       | Replication II | 0.143 | 544 | 21.9%  | 180 | 28.9%  | 16  | 37.5%  | 1.41 (1.07 to 1.84) | 0.01  |
|            |       | Combined    | 0.137 | 1953| 20.7%  | 876 | 27.9%  | 94  | 31.9%  | 1.46 (1.29 to 1.66) | 6.17 × 10^-9 |
| rs9304497  | KCTD1 | Discovery | 0.278 | 576 | 20.1%  | 409 | 31.5%  | 96  | 31.3%  | 1.60 (1.33 to 1.94) | 1.19 × 10^-4 |
| G/A‡       | Intron| Replication I | 0.269 | 605 | 22.2%  | 416 | 26.4%  | 91  | 17.6%  | 1.03 (0.85 to 1.25) | 0.77  |
|            |       | Replication II | 0.298 | 376 | 24.5%  | 288 | 25.0%  | 77  | 16.9%  | 0.88 (0.70 to 1.11) | 0.29  |
|            |       | Combined    | 0.280 | 1557| 22.0%  | 1113| 25.1%  | 264 | 22.4%  | 1.19 (1.06 to 1.34) | 0.004 |
| rs10501719 | DISC1FP1| Discovery | 0.111 | 850 | 21.2%  | 221 | 25.8%  | 9   | 66.7%  | 1.81 (1.40 to 2.35) | 7.70 × 10^-4 |
| A/G‡       | Intron| Replication I | 0.111 | 877 | 23.5%  | 222 | 22.5%  | 12  | 33.3%  | 1.00 (0.76 to 1.31) | 0.98  |
|            |       | Replication II | 0.123 | 570 | 24.2%  | 159 | 23.9%  | 12  | 8.3%   | 0.97 (0.69 to 1.36) | 0.87  |
|            |       | Combined    | 0.114 | 2297| 22.8%  | 602 | 24.1%  | 33  | 33.3%  | 1.24 (1.05 to 1.46) | 0.01  |

*Genotypes are shown as AA for major-allele homozygotes, AB for heterozygotes and BB for minor-allele homozygotes. SNP = single-nucleotide polymorphism; HR = hazard ratio; CI = confidence interval; TLI = temporal lobe injury; MAF = minor allele frequency.

†HRs, 95% CIs and two-sided Wald test P-values were calculated using multivariate Cox proportional hazard regression models assuming additive effect of minor alleles and adjusting age at NPC diagnosis, tumor stage, and radiotherapy technique as covariates.

‡The major allele/minor allele of the SNP.

Figure 3. Expression quantitative trait locus (eQTL) analysis and functional characterizations of the protective role of CEP128 gene in radiosensitivity. A,B) Visualization of association trends for rs162171, rs1711237, and CEP128 mRNA expression in (A) HapMap CHB (Han Chinese in Beijing, China) and JPT (Japanese in Tokyo, Japan) and (B) in the intralobular white matter of human brain in the UK brain expression consortium (UKBEC). Gene expression values observed in each individual are shown by points. The correlation was represented by ρ and P values were calculated by Spearman correlation test. C) The means ± SD (N = 3) of luciferase activity detection. "CEP128 promoter WT" means luciferase vector containing a segment of CEP128 promoter with rs1711237A and rs1711246G, "CEP128 promoter variant" means luciferase vector containing a segment of CEP128 promoter with rs1711237G and rs1711246A. The numbers 1, 2, 3, 4 in the x-axis indicate four independent luciferase vectors. D) The means of relative mRNA expression levels (N = 2) of CEP128 after stable inhibition of endogenous CEP128 using shRNA in the U87 cell line. shCEP128-1, 2, and 3 indicate three shRNAs targeting different regions of CEP128 mRNA. E) The clonogenic survival curve was established on day 10 after stable transfected cells received indicated doses of radiation. Survival fractions were calculated. Data represent means ± SD (N = 3) and P values were calculated by two-sided t-test for shRNA vs sh-control.
Discussion

In this prospective observational study, we identified genetic variants in a centrosomal protein CEP128 that conferred the risk of radiation-induced TLI in NPC patients. We found that rs162171 in the intron of CEP128 was statistically significantly associated with TLI development. Additionally, the A → G transition at rs17111237 in CEP128 promoter may cause downregulation of this gene, which could further increase the radiosensitivity of glioblastoma cell lines, suggesting the protective role of CEP128 against radiation toxicity in normal brain tissue.

CEP128 is a newly characterized centrosome protein (CEP) which is a key regulator of ciliation (32) and plays important roles in cell cycle progression (33). Increasing evidence has indicated that primary cilia plays important roles in coordinating a variety of cellular signaling pathways in neuronal cell and regulate cell migration, differentiation, as well as a host of adult behaviors (34). Functions of primary cilia can be dramatically affected by either the malfunction of ciliary genes or various external factors, including radiation. Interestingly, CEP128 appears to associate with both the cilia function maintenance and radiation response of cells. Knockdown of CEP128 could lead to abnormally high levels of ciliation in proliferating cells (35), and multiple CEP128 interactors enriched by ciliogenesis induction, including CASK, CEP72, and LMO7, were associated with ionizing radiation (IR) resistance in a recent genome-wide RNAi screen study (36). Identification of genetic variants in an essential gene of primary cilia provides not only novel insights into the underlying mechanisms of radiation-induced brain injury, but also a new list of potential treatment targets to prevent or ameliorate the side effects of radiotherapy.

Cerebrovascular injury and remodeling is another suggested hypothesis on the development of radiation-induced brain injury. Evidence has shown the vascular structural and functional alteration after fractional whole-brain irradiation (RWBI) in rodents (37–39) and nonhuman primate (NHP) models (40–42). A recent NHP study reported white matter-specific transcriptional alterations of genes involving cerebrovascular remodeling, blood–brain barrier integrity, neurotransmission, and inflammation (42). Interestingly, we found 34 SNPs in these genes associated with TLI with P less than .01 in our samples (data not shown). More studies are warranted to further investigate the underlying mechanisms of vascular abnormality in the development of TLI.

Temporal lobe injury has generally been regarded as a progressive and irreversible complication in the radiotherapy of NPC (43). Although the TLI rate is decreasing in the NPC survivors in the era of IMRT, there is still 3.2–12.9% of NPC patients suffering TLI (5–9,44). Moreover, temporal lobe protection is arduous, particularly in patients with T4 disease, with higher injury rates up to 25.0% (6). Hence, pretreatment assessment and prevention of adverse responses is of critical importance. Some ongoing or completed preclinical studies (39,45,46) and clinical trials (47–50) have proven that clinical intervention could effectively treat or even reverse radiation-induced brain injury. Furthermore, the risk prediction integrating the genetic findings and clinical variables showed that those patients with high risks of TLI could be identified in advance, suggesting that applying genetic risk profiling to clinical intervention may have potential value to prevent severe complications and improve the quality of life of NPC patients. Further studies might be warranted to investigate the clinical models generalized to other cancers.

The main limitation of this study is that dosage parameters of the temporal lobes, which were reported as important factors for the development of TLI (51,52), were not available and these parameters were not adjusted in the analysis. We made some attempts to overcome this limitation. First, we collected the information on prescribed total dose, dose per fraction, and overall treatment time of the nasopharynx. However, no association was observed between the dosage parameters and the development of TLI. These results might be explained by the similarity of dose prescription received by all the patients. Second, we took radiation techniques and tumor stages as covariates, which may reflect the dosage–volume parameter of the temporal lobes to some extent. In the patients who received IMRT or who were in early tumor stages, a smaller volume of the temporal lobes would receive high-dose irradiation. Additionally, when the associations were stratified by tumor stages or radiation techniques, the genetic effect remained statistically significant (data not shown).

In conclusion, using genome-wide association analysis followed by two independent replications and functional studies, we identified CEP128 as a susceptibility gene of TLI development in NPC patients. Additional studies on how the CEP proteins participate in radiation-induced brain injury are required. Moreover, normal tissue sensitivity to radiation exposure has been regarded as a complex, polygenic trait resulting from interactions of numerous genes in different cellular pathways (53,54). Therefore, further experiments are needed to confirm our findings or to identify other additional susceptibility loci, which could provide the basis of individualized radiation treatment for patients.

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Notes

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The main limitation of this study is that dosage parameters of the temporal lobes, which were reported as important factors for the development of TLI (51,52), were not available and these parameters were not adjusted in the analysis. We made some attempts to overcome this limitation. First, we collected the information on prescribed total dose, dose per fraction, and overall treatment time of the nasopharynx. However, no association was observed between the dosage parameters and the development of TLI. These results might be explained by the similarity of dose prescription received by all the patients. Second, we took radiation techniques and tumor stages as covariates, which may reflect the dosage–volume parameter of the temporal lobes to some extent. In the patients who received IMRT or who were in early tumor stages, a smaller volume of the temporal lobes would receive high-dose irradiation. Additionally, when the associations were stratified by tumor stages or radiation techniques, the genetic effect remained statistically significant (data not shown).

In conclusion, using genome-wide association analysis followed by two independent replications and functional studies, we identified CEP128 as a susceptibility gene of TLI development in NPC patients. Additional studies on how the CEP proteins participate in radiation-induced brain injury are required. Moreover, normal tissue sensitivity to radiation exposure has been regarded as a complex, polygenic trait resulting from interactions of numerous genes in different cellular pathways (53,54). Therefore, further experiments are needed to confirm our findings or to identify other additional susceptibility loci, which could provide the basis of individualized radiation treatment for patients.

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Author Contributions: Dr Wei-Hua Jia supervised the study and had full access to all the data in the study. Guo-Ping Shen, Ming-Yuan Chen, and Ying Sun collected the clinical data and followed up the MRI for the diagnosis of TLI. Tong-Min Wang and Guo-Ping Shen analyzed and interpreted the data. Jiang-Bo Zhang designed and performed the functional experiments. Shao-Yi Huang performed functional experiments. Xi-Zhao Li, Wen-Qiong Xue, Xiao-Hui Zheng, Shao-Dan Zhang, and Ye-Zhu Hu collected the samples. Jing He, Hai-De Qin, and Jin-Xin Bei conducted statistical analysis. Wei-Hua Jia and Tong-Min Wang wrote the article. Jianbing Mu and Yin Yao Shugart gave critical revision of the article. Jianbing Mu, Yin Yao Shugart, and Jun Ma gave important consultation to this work.

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