Effects of Salicylic Acid on Fatty Acid Gene Expression in *Carthamus tinctorious* L. cv. Dinçer under Pendimethalin Stress

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Abstract

Pendimethalin is a member of the dinitroaniline class herbicide. It used to control most annual grasses and many annual broad-leaved weeds. Salicylic acid acts as an endogenous signal molecule in charge of inducing environmental stress tolerance in plants. Omega-3 fatty acid desaturase is a key enzyme for α-linolenic acid biosynthesis. Here, we searched to understand the beneficial impacts of salicylic acid on fatty acid desaturase gene (*FAD3* and *FAD7*) expression during pendimethalin stress in safflower (*Carthamus tinctorious* cv. “Dinçer”). In this study, 0.004 and 0.01 M pendimethalin was applied to safflower plants grown under controlled climatic conditions in 36 pots. 0.05 mM salicylic acid was applied to the samples where pendimethalin was applied. After application, gene expression analyze were performed using quantitative polymerase chain reaction. The floor change between the test and control groups was calculated with the formula $2^{\Delta\Delta CT}$. The change between gene expression levels was evaluated by t test ($p < 0.05$). *FAD3* and *FAD7* expression levels decreased at low pendimethalin concentration. A down-regulation in *FAD3* expression was observed in high pendimethalin stress, while an up-regulation in *FAD7* expression was detected. Salicylic acid had a ameliorative effect on the negative effect of pendimethalin stress on *FAD3* and *FAD7* gene expression.

Keywords: *Carthamus tinctorious*, omega-3 fatty acid, pendimethalin, salicylic acid

INTRODUCTION

Pesticides are used to prevent or inhibit the growth of various agricultural pests that can damage crops and reduce farm productivity (Kaur, 2019). The use of herbicides can generate stress conditions, evidenced by the increase in phytotoxicity, which affects growth, development and productivity (Agostinetto et al., 2016). Pendimethalin (PEN) is a pre-emergence herbicide used for control of annual grasses and annual broad-leaved weeds (Tomlin, 2001). It is the third largest selective herbicide in the world and has been classified as a possible human
carnogen (Hou et al., 2004; Osman et al., 2016). Vegetable oils are significant economically because they are renewable resources of highly reduced carbon and used in diets and industrial applications (Heppard et al., 1996; Yang and Xu, 2007). Fatty acid desaturases (FADs) are a key role in the maintenance of the proper structure and functioning of biological membranes (Los and Murata, 1998). In plants, ω-3 fatty acid desaturases (FAD3 and FAD7) are involved in the production of α-linolenic acid (ALA) (Xue et al., 2018).

Safflower (Carthamus tinctorius L.) is an annual herbaceous plant that is adapted to hot and dry environments (Pahlavani et al., 2004; Dajue and Mündel, 1996). The plant has been grown as a source of vegetable oil used for food and industrial purposes (Dajue and Mündel, 1996; Majidi et al., 2011). Systemic acquired resistance is an immune response of plants that provides protection to infection by pests and pathogens (Fu and Dong, 2013; Andersen et al., 2018). Salicylic acid (SA) is a plant hormone that plays a role in the induction and regulation of metabolic responses to biotic and abiotic stresses (Maruri-Lopez et al., 2019).

This research was aimed to evaluate the potential role of SA on FAD3 and FAD7 gene expression of Carthamus tinctorius cv. “Dinçer” under pendimethalin stress.

MATERIAL AND METHOD

Stress Treatments and Growth Conditions

In the present study, commercial form of PEN herbicide was used. Dinçer seeds were provided from Transitional Zone Agricultural Research Institute, Eskişehir. The seeds were grown at 23 ± 2°C in plant growth chamber with 60-65% humidity. Plastic pots (10.5 × 8 cm diameter and volume 0.46 L) containing soil were used. A total of 36 samples (pots) with 3 replicates in 6 groups were created for each gene. Samples were irrigated every 3 days with Hoagland culture solution (Hoagland and Arnon, 1938). PEN was applied after germination at the doses determined (0.004 and 0.01 M) following toxicity tests to the leaves of plants of suitable size (21 days). PEN was applied by spraying method. The application samples were divided into six groups as follows (Group 1) Control, (Group 2) SA Control; (Group 3) 0.004 M PEN, (Group 4) 0.004 M PEN+0.5 mM SA; (Group 5) 0.01 M PEN; (Group 6) 0.01 M PEN+0.5 mM SA.

RNA Isolation

Samples were taken from safflower leaves at 24, 48 and 72 hours after chemical application and frozen in liquid nitrogen. Frozen samples were taken into homogenization tubes and 600 µl lysis buffer (RLT) was added. 100 mg sample of frozen material was homogenized for 1 minute at 7 m s⁻¹ with 2.5 mm ceramic beads in an eppendorf tube (Bioprep 24). Samples were incubated at 56°C for 30 minutes in a water bath. The samples were centrifuged at 16,000 rpm for 2 minutes and the supernatant was taken into a new eppendorf tube. RNA isolation was performed according to the manufacturer’s instructions (QIAGEN, Allprep Mini Kit Cat No:80004).

cDNA Synthesis

The quality of mRNA obtained after RNA isolation was checked by spectrophotometer. RNA isolation was repeated from the samples with total RNA concentration below 4 µg mL⁻¹. In the examples, the conversion kit (QIAGEN, RT2 HT First Strand Kit, Cat No: 330411) was used for DNA elimination and a rapid first chain cDNA synthesis. cDNA synthesis was carried out by holding the reaction components at thermal cycler for 42°C at 15 minutes and at 95°C for 5 minutes.

qPCR

Quantitative PCR was performed in a total reaction volume of 25 µl. C_T cut off value was adjusted to 35 with positive and negative controls. Distribution of reaction components are 12.5 µl of RT² qPCR master mix, 10.5 µl of ddH₂O, 1.0 µl of template cDNA and 1.0 µl of RT² qPCR primer assay. Reaction conditions are given in Table 1. The data of the genes and housekeeping gene used in the reaction are given in Table 2. Sybr green was used as fluorescent dye in the reaction (QIAGEN, RT2 SYBR Green qPCR Mastermix, Cat No:330500). In the study, 3 techniques and 3 biological repeats were applied.

Data Analysis

C_T values were analyzed with the Qiagen GeneGlobe bioinformatics tool. Fold change and fold regulation values were computed with the ΔΔC_T method (Livak and Schmittgen, 2001). The floor change between the test and control groups was calculated with the formula 2^ΔΔC_T. For the statistical significance of the expression change.
between the test and control groups, p value was analyzed by t test (p <0.05).

**Table 1.** qPCR reaction conditions

| Cycle | Temperature (°C) | Time       |
|-------|-----------------|------------|
| 1     | 95              | 10 minute  |
| 40    | 95              | 15 second  |
| 40    | 53              | 40 second  |
| 40    | 72              | 30 second  |

**RESULTS AND DISCUSSION**

Pesticide is released into the environment in a controlled manner to prevent, destroy or control undesirable plant or animal species (Zikankuba et al., 2019). Fatty acid-induced signaling is one of the important defense pathways. (Kachroo et al., 2005).

The synthesis of ALA requires the activity of ω-3 FADs which were encoded by *FAD3* and *FAD7* (Peng et al., 2020). In transcriptome analysis with perilla seeds, *FAD3* and *FAD7* genes were found to be the basic genes for ALA synthesis (Kim et al., 2016). ALA, which is the product of ω-3 FAD is precursor of jasmonic acid that play crucial roles in plant development and stress responses (Weber, 2002).

It has been shown that salicylic acid applications can positively affect the formation of some plant secondary metabolites (such as essential oils, oleoresins and triterpenes) (Elyasi ve ark., 2016; Mirzajani et al., 2015; Rodrigues and Fett-Netto, 2009).

In this study, the data analysis web portal (GeneGlobe®) calculates fold change/regulation using delta delta CT method, in which delta CT is calculated between genes (*FAD3* and *FAD7*) and a housekeeping gene (HKG). Fold Change is calculated using 2^(- delta delta CT). The statistical evaluation of the difference between the expression levels of the *FAD3* and *FAD7* genes between the test groups and control groups was done with t-test (Table 3). p <0.05 was statistically significant.

The analysis results of the samples at 24th h were given in Figure 1. *FAD3* gene expression was downregulated in all application groups, but only in 0.004 M PEN, 0.004 M PEN+0.5 mM SA and 0.01M groups were statistically significant (p <0.05) (Table 3). It was observed that the downregulate in gene expression level formed with PEN application upregulated even more with SA. In the heat map, there was decrease in *FAD3* expression compared to the control group (Figure 1). *FAD7* gene expression was downregulated in all groups. This decrease in expression level was found statistically significant except 0.01 M PEN+0.5 mM SA groups. When the effect of SA on control groups was analyzed, it was found that the decrease in *FAD7* expression was higher than *FAD3*. PEN was downregulated *FAD3* and *FAD7* expression levels. Increasing of PEN concentration caused more decrease in expression level in *FAD3* than *FAD7*.

SA caused a downregulate in *FAD3* and *FAD7* expression levels on the control group at 48th h. While *FAD3* was upregulated in the test groups, *FAD7* was downregulated (Figure 1). SA application limited the decrease in *FAD7* expression in test groups exposed to pendimethalin (Table 3).

The expression levels of *FAD3* and *FAD7* genes were upregulated compared to the control group in PEN and PEN+SA application groups at 72th h in Dincer cultivar. (Figure 1). Increases in expression levels were also found statistically significant (Table 3).

*FAD3* expression in rapeseed under injury stress is known to be stimulated by abscisic acid, a plant growth regulator (Jitao et al., 1995). *FAD7* expression was down-regulated in low temperature stress in the leaves of birch seedlings, while *FAD3* expression was up-regulated and an increase in ALA content was detected (Martz et al., 2006). Kim et al. (2016) showed that *PfrFAD3* and *PfrFAD7* were identified as key genes for ALA synthesis in seeds and leaves of *Perilla frutescens* (L.) var. frutescens. Sen Wang et al. (2014) reported that significant changes in fatty acid composition caused by consuming of *LeFAD3* in antisense transgenic tomato (*Lycopersicon esculentum*) lines were the reduce in 18:3 and the rise in 18:2 in leaves and roots. Guan et al. (2014) reported that the low *FAD3* transcript level in developing seeds is consistent with the high linoleic acid level and linolenic acid deficiency in safflower seed oil.
Table 2. Primer sequence of genes

| Symbol | Sequence |
|--------|----------|
| ACTB (Housekeeping) Acc.KJ634809.1 | F- GCGGCTGGATCCACGAGA<br>R- TCAGCAATGCCAGGGAACATAG |
| FAD3 Acc.HQ831356.1 | F- TCGTGGTGCTCCGTTAAATGAAA<br>R- GCCACAAGTACAATGGGTATGC |
| FAD7 Acc.HQ831349.1 | F- TCCACCTCACTTCCAAGAGTTG<br>R- ACTCGCTATCTCCATCGTTTCG |

Table 3. Changing in expression levels of *FAD3* and *FAD7* genes (p <0.5)

| Group 1 0.5 mM SA Control | FAD 3 | FAD 7 |
|---------------------------|-------|-------|
| 24 h | 48 h | 72 h | 24 h | 48 h | 72 h |
| Fold Change | 0.91 | 0.39 | 1.75 | 0.17 | 0.94 | 1.66 |
| P value | 0.486 | 0.000 | 0.035 | 0.001 | 0.003 | 0.043 |
| Group 2 0.004 M PEN | FAD Change | 0.21 | 1.19 | 2.93 | 0.54 | 0.39 | 2.36 |
| P value | 0.002 | 0.022 | 0.002 | 0.013 | 0.000 | 0.002 |
| Group 3 0.004 M PEN+0.5 mM SA | FAD Change | 0.20 | 1.03 | 5.29 | 0.11 | 0.45 | 5.79 |
| P value | 0.002 | 0.595 | 0.000 | 0.001 | 0.000 | 0.000 |
| Group 4 0.01 M PEN | FAD Change | 0.07 | 2.31 | 2.43 | 0.48 | 0.51 | 13.55 |
| P value | 0.000 | 0.006 | 0.015 | 0.002 | 0.000 | 0.0001 |
| Group 5 0.01 M PEN+0.5 mM SA | FAD Change | 0.72 | 2.96 | 3.16 | 0.64 | 0.93 | 6.93 |
| P value | 0.050 | 0.012 | 0.017 | 0.076 | 0.001 | 0.000 |

Figure 1. Heatmap showing expression levels of *FAD3* and *FAD7* genes at 24, 48, 72th h
Safflower is an important industrial plant with oil seed used for edible oil production and its cultivation has been increasing throughout the world in last decades. The plant is a valuable resource in terms of nutrition with its high ALA capacity. It is an important source in nutrition with its high ALA content. It has been demonstrated with this study that herbicides, which are widely used for the purpose of increasing plant production and yield, may exhibit negative results in the oil metabolism of the plant in terms of oil yield. It was also shown that SA played a role in removing negative effects of PEN (Figure 2 and 3).

**CONCLUSION**

It is thought that the use of PEN may have a decrease in plantal oil yield in *Carthamus tinctorius* L. Dinçer cultivar and cause a decrease in ALA formation in terms of nutrition and metabolism. SA can play a role in reducing the adverse effect of PEN on fatty acid metabolism.

**Figure 2.** General evaluation of fold change in *FAD3* expression levels

**Figure 3.** General evaluation of fold change in *FAD7* expression levels
CONFLICT OF INTEREST STATEMENT

The author declares that there is no conflict of interest in this study.

RESEARCH AND PUBLICATION ETHICS

STATEMENT

The author declares that the research and publication ethics are complied with in the study.

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