An application of topological graph clustering to protein function prediction

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Abstract

We use a semisupervised learning algorithm based on a topological data analysis approach to assign functional categories to yeast proteins using similarity graphs. This new approach to analyzing biological networks yields results that are as good as or better than state of the art existing approaches.

1 Introduction

Determining protein function is an integral part of understanding biological mechanisms. Until recently, proteins were characterized one at a time, slowly building our knowledge of a given pathway or mechanism. The yeast *Saccharomyces cerevisiae* was one of the first organisms to have its genome sequenced. As a model organism it has been thoroughly studied. However, at the time of sequencing only about a third of the genes had been characterized. With the increasing ease of DNA sequencing, databases were made of proteins with known function from different organisms. Sequences could be compared with those in the database, and those with sequence similarity would likely correspond to a similar protein in a different organism. For *S. cerevisiae*, about one third showed sequence similarity to genes in other organisms, leaving the last third, approximately 2000 genes, completely unknown. The number of unknown genes and difficulty in understanding their
function called for a new approach.

The shift to large scale biology has brought about both necessity for large scale protein function identification and many new avenues for large scale protein characterization. There are now hundreds of genomes sequenced. Before, being able to sequence a gene was a major obstacle. Now, the difficulty has shifted to determining the function of a large number of genes. There are numerous new high throughput methods producing information about all different attributes of a protein, like the Pfam domain, protein-protein interaction, protein expression, etc. This gives us a more complete picture of these complex systems, and also gives us vast amounts of information for which there are not always obvious conclusions. For more background information, see [14] and [6].

We can represent information about different proteins in the form of a graph. For example, with protein-protein interaction, the proteins are represented by vertices and edges between vertices represent interactions. Given such a graph, it is natural to apply approaches from machine learning to find clusters of vertices. The idea is that proteins that form a tightly grouped cluster in such a graph will have similar functions. Thus we can predict the function of an unknown protein when there are proteins of known function in its cluster. This strategy has been successfully implemented by many groups. In this paper, we apply a novel semisupervised topological data analysis approach to the same problem. This algorithm is based on the TILO/PRC (Topologically Intrinsic Lexicographic Ordering/Pinch Cluster Ratio) algorithm developed by the second and third author [4,5] and described in the next section.

Many approaches to functional annotation given a protein interaction graph have been proposed [9]. Some have focused on techniques from statistics and machine learning such as Markov random field models [2] and Support Vector Machines (SVM) [7]. Clustering using TILO/PRC bears some similarity to Spectral clustering [10] in that it attempts to find a partition of a graph with small boundary but large interior weight. Our approach is therefore similar in spirit to methods that make use of the graph Laplacian in clustering or propagation of labels [8][11][13]. Note that several of these methods use a combined graph obtained as a weighted average of several other graphs. The weights are obtained as part of the procedure using, for example, semidefinite programming in [7]. We have not attempted to do this in the present work, and use a simple average to form a combined graph. As we will see, our
results are on par with methods that attempt to select optimal weights with regard to some objective function.

The contributions of this paper are as follows:

1. We describe the TILO algorithm for semisupervised learning.
2. We demonstrate that this algorithm is effective at predicting protein function.
3. We demonstrate that this algorithm performs well in comparison to existing semisupervised methods, including the Markov random field model of Deng, Chen, and Sun [2], and the kernel methods of Lanckriet et al. [7].

Code for the project is located at http://seanbowman.me/yeast-protein

2 The TILO algorithm

First we describe the TILO/PRC algorithm, which clusters the vertices of a graph by finding a linear order on the vertices with nice properties. See [4,5] for more details. Let $G = (V, E)$ be a graph, where $V$ is the set of vertices and $E$ is a set of weighted edges. Given a set of vertices $A \subseteq V$, define the boundary $\partial A \subseteq E$ to be the set of edges with one endpoint in $A$ and the other in $V \setminus A$. The size of the boundary, $|\partial A|$, is defined to be the sum of weights of $\partial A$.

A pinch cluster in $G$ is a set of vertices $S \subseteq V$ such that for any sequence of vertices $v_1, \ldots, v_m$, if adding these vertices to $S$ or removing them from $S$ creates a set with smaller boundary, then for some $k < m$, adding/removing $v_1, \ldots, v_k$ to/from creates a set with strictly larger boundary. Informally, a pinch cluster has small boundary relative to the edges it contains, and vertices in a pinch cluster are more strongly connected to each other than to outside vertices.

Consider an ordering $O = (v_1, v_2, \ldots, v_n)$ of $V$. Let $A_i = \{v_1, \ldots, v_i\}$ and $b_i = |\partial A_i|$. The width of $O$, $w(O)$ is the tuple $(w_1, \ldots, w_{n-1})$, where $\{w_1, \ldots, w_{n-1}\} = \{b_1, \ldots, b_{n-1}\}$ and $w_{i+1} \leq w_i$ for each $1 \leq i < n - 1$. We order widths lexicographically. In [6] a property called weak reducibility is
defined for an ordering $O$, and [5] Lemma 3] shows that given a weakly reducible ordering $O$, we can find a new ordering $O'$ so that $w(O') < w(O)$. After applying this procedure a finite number of times, we arrive at a strongly irreducible ordering.

The goal of the TILO algorithm is to find a strongly irreducible ordering of the vertices of $G$. Suppose that $O$ is a strongly irreducible ordering and $i$ is the index of a local minimum of $(b_1, \ldots, b_{n-1})$. Then [5, Theorem 4] shows that $A_i$ and its complement are pinch clusters. Note that we are free to pick the index of any local minimum of the boundary vector in order to partition the data. The pinch ratio cut, defined in [4], is one heuristic for choosing such an index which has been shown to work well in practice. However, in the present work we will always make cuts at every index corresponding to a local minimum.

2.1 Semisupervised learning with TILO

In the case at hand, we are given a weighted graph $(V, E)$ which we think of as a similarity graph. The vertex set $V = \{v_1, v_2, \ldots, v_n\}$ is divided into two parts: $m$ vertices $v_1, \ldots, v_m$ are labeled $y_1, \ldots, y_m$, and the rest are unlabeled. We wish to use the structure of the graph to assign labels to $v_{m+1}, \ldots, v_n$. In the present application $y_i \in \{0, 1\}$ for $1 \leq i \leq m$, and so we may also assign probabilities $y_i \in [0, 1]$, $m < i \leq n$, representing the likelihood that vertex $v_i$ has the label 1.

For each connected component of the similarity graph, we use the TILO/PRC algorithm to divide the vertices into clusters. Given a strongly reducible ordering $O$, we make a cut at every local minimum of the boundary vector, thus cutting the graph into the maximal number of pinch clusters. If $v_{i_1}, v_{i_2}, \ldots, v_{i_k}$ are labeled vertices in a resulting cluster, we assign the probability

$$\frac{1}{k} \sum_{j=1}^{k} y_{i_j}$$

to each unlabeled vertex in the cluster. If there are no labeled vertices in the cluster, we assign the most common label in the data set to each unlabeled vertex.
The TILO algorithm can get stuck in local minima, which correspond to orderings that may realize suboptimal clusters in a given graph. We use a version of bagging \cite{1} in order to generate more accurate predictions from our labeled data. Bagging also ensures that the probability that a given vertex always receives the most common label in the data set is extremely low. After generating $N$ subsets of the unlabeled data by sampling a proportion $\lambda$ uniformly without replacement, the algorithm described above is run on each sample. The probabilities obtained are averaged over all runs, and the averaged probabilities $y_{m+1}, \ldots, y_n$ are the output of the algorithm. In the data below, we use $N = 25$ and $\lambda = 0.5$.

3 Experimental Results

We use the data set from \cite{12} and also used in \cite{2} and \cite{7} (among others) in order to compare our results. The goal of the algorithm is to predict functional classes of 3588 yeast proteins, with true labels given by the MIPS Comprehensive Yeast Genome Database. There are 13 classes in the top level hierarchy, and we view the problem as 13 separate binary classification tasks.

The input data consists of five symmetric matrices $W_i$, $i = 1, 2, 3, 4, 5$, whose entries describe different types of interaction between the row and column proteins. The first matrix, $W_1$, comes from the Pfam domain of the proteins. Each protein was characterized based on the presence or absence of 3950 different structural domains. This results in 3950-bit vector, and the dot products of these vectors are used to generate the matrix. The next three, $W_2$, $W_3$, and $W_4$, are from the combined data in the MIPS Comprehensive Yeast Genome Database. The matrix $W_2$ indicates if there is co-participation in a protein complex. The matrix $W_3$ indicates known protein-protein interactions. The matrix $W_4$ indicates known genetic interactions. The final matrix $W_5$ is based on comparing the proteins’ gene expression profiles.

The graphs obtained by considering the $W_i$ as adjacency matrices are highly disconnected and have many components with a small number of vertices. We do not consider isolated vertices, since it is impossible to infer a label using our graph based approach. Furthermore, $W_5$ is so sparse that we have
not used it in our tests. We also examine the integrated graph

\[ \frac{1}{5} \sum_{i=1}^{5} W_i. \]

This graph is relatively well connected. We use 5 fold cross validation 3 times and report the receiver operating characteristic (ROC) score on the test set.

Our results are shown in Table 1. Shown are the mean ROC scores, as described above, for the graphs \( W_1, W_2, W_3, W_4 \), and the integrated graph. The graph \( W_1 \) has 432 components, 2809 vertices, and 48445 edges; \( W_2 \) has 29 components, 1051 vertices, and 1872 edges; \( W_3 \) has 140 components, 1342 vertices, and 1844 edges; \( W_4 \) has 106 components, 819 vertices, and 1006 edges; the integrated graph has 96 components, 3278 vertices, and 56371 edges. The columns labeled SVM/\( W_1 \) and SDP/SVM are results from [7]. The first shows the ROC values obtained using a 1–norm SVM with \( C = 1 \) on a normalization of our matrix \( W_1 \), and the SDP/SVM column shows the performance of a semidefinite programming SVM approach which uses a weighted combination of the \( W_i \). The last column shows the performance of the Markov random field algorithm of [2] on the weighted combination used in the previous column.

4 Conclusion

Understanding protein function is integral to our understanding of pathways and mechanisms in biological systems, and the huge amount of data available today necessitates techniques which can work on a large scale. We have described a semisupervised learning algorithm based on TILO/PRC and shown that this algorithm performs well on the task of predicting protein functional classes on a data set of yeast proteins.

Tables 1 and 2 show that for each of the 13 functional classes studied, the semisupervised TILO method performs better than the SVM method of [7] when using \( W_1 \). The same is true for \( W_2, W_3, \) and \( W_4 \). In all, the mean ROC score improves from 0.767 to 0.842 on \( W_1 \), albeit with slightly higher standard deviations. For the combined graph, we obtain similar scores (a mean ROC of 0.844 versus 0.853 for the SDP/SVM approach of [7]) while using a simple average of the \( W_i \) instead of a weighted average.
| Class | W1    | W2    | W3    | W4    |
|-------|-------|-------|-------|-------|
| 1     | 0.880 ± 0.019 | 0.768 ± 0.046 | 0.700 ± 0.027 | 0.833 ± 0.043 |
| 2     | 0.879 ± 0.031 | 0.684 ± 0.098 | 0.722 ± 0.059 | 0.756 ± 0.165 |
| 3     | 0.825 ± 0.022 | 0.793 ± 0.040 | 0.755 ± 0.033 | 0.881 ± 0.029 |
| 4     | 0.876 ± 0.014 | 0.844 ± 0.020 | 0.842 ± 0.021 | 0.857 ± 0.034 |
| 5     | 0.896 ± 0.019 | 0.847 ± 0.032 | 0.816 ± 0.058 | 0.830 ± 0.091 |
| 6     | 0.881 ± 0.021 | 0.751 ± 0.046 | 0.767 ± 0.039 | 0.830 ± 0.023 |
| 7     | 0.878 ± 0.022 | 0.863 ± 0.042 | 0.887 ± 0.027 | 0.866 ± 0.039 |
| 8     | 0.794 ± 0.034 | 0.656 ± 0.103 | 0.651 ± 0.052 | 0.776 ± 0.067 |
| 9     | 0.823 ± 0.054 | 0.589 ± 0.149 | 0.811 ± 0.063 | 0.744 ± 0.079 |
| 10    | 0.767 ± 0.034 | 0.755 ± 0.045 | 0.776 ± 0.045 | 0.804 ± 0.025 |
| 11    | 0.629 ± 0.050 | 0.625 ± 0.056 | 0.674 ± 0.055 | 0.635 ± 0.070 |
| 12    | 0.961 ± 0.019 | 0.756 ± 0.109 | 0.796 ± 0.056 | 0.759 ± 0.126 |
| 13    | 0.854 ± 0.053 | 0.577 ± 0.159 | 0.710 ± 0.104 | 0.753 ± 0.107 |

Table 1: Mean ROC of the semisupervised TILO/PRC algorithm on the graphs $W_i$.

We believe that algorithms inspired from ideas in topology, such as the TILO/PRC algorithm described here, are uniquely suited to this problem for several reasons. They have a good theoretical grounding with clear links to both topology and existing machine learning approaches, they operate directly on protein interaction graphs, they are less sensitive to geometric properties of the data (which are not always relevant), and they are capable of meeting the challenges of large scale biology described above.

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Table 2: Mean ROC of the semisupervised TILO/PRC algorithm on the integrated graph. Columns SVN/W1 and SDP/SVM are results of [7], and the last column is results from [2]. See text for details.

| Class | Integrated | SVM/W1  | SDP/SVM | MRF   |
|-------|------------|---------|---------|-------|
| 1     | 0.866 ± 0.018 | .8373 ± .0037 | .8825 ± .0042 | .7532 ± .0042 |
| 2     | 0.871 ± 0.033 | .8107 ± .0113 | .8563 ± .0100 | .7173 ± .0102 |
| 3     | 0.839 ± 0.019 | .7547 ± .0062 | .8464 ± .0053 | .6990 ± .0045 |
| 4     | 0.882 ± 0.011 | .8085 ± .0048 | .9024 ± .0028 | .7490 ± .0049 |
| 5     | 0.908 ± 0.014 | .8349 ± .0058 | .9094 ± .0050 | .7375 ± .0102 |
| 6     | 0.868 ± 0.022 | .8069 ± .0046 | .8742 ± .0049 | .7183 ± .0059 |
| 7     | 0.880 ± 0.019 | .8010 ± .0074 | .9149 ± .0040 | .7534 ± .0085 |
| 8     | 0.811 ± 0.030 | .7023 ± .0089 | .8023 ± .0094 | .7285 ± .0120 |
| 9     | 0.823 ± 0.043 | .7309 ± .0113 | .8623 ± .0091 | .6849 ± .0107 |
| 10    | 0.790 ± 0.023 | .6906 ± .0079 | .8120 ± .0078 | .6954 ± .0060 |
| 11    | 0.658 ± 0.043 | .5952 ± .0148 | .6575 ± .0093 | .5691 ± .0092 |
| 12    | 0.954 ± 0.018 | .9331 ± .0038 | .9674 ± .0023 | .8575 ± .0076 |
| 13    | 0.841 ± 0.068 | .6678 ± .0227 | .8083 ± .0091 | .6612 ± .0169 |
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