Positron emission tomography of tumour $[^{18}\text{F}]$fluoroestradiol uptake in patients with acquired hormone-resistant metastatic breast cancer prior to oestradiol therapy

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Abstract

Purpose  Whereas anti-oestrogen therapy is widely applied to treat oestrogen receptor (ER) positive breast cancer, paradoxically, oestrogens can also induce tumour regression. Up-regulation of ER expression is a marker for oestrogen hypersensitivity. We, therefore, performed an exploratory study to evaluate positron emission tomography (PET) with the tracer $16\alpha-[^{18}\text{F}]$fluoro-17$\beta$-oestradiol ($^{18}\text{F}$-FES) as potential marker to select breast cancer patients for oestradiol therapy.

Methods  Eligible patients had acquired endocrine-resistant metastatic breast cancer that progressed after $\geq 2$ lines of endocrine therapy. All patients had prior ER-positive histology. Treatment consisted of oestradiol 2 mg, three times daily, orally. Patients underwent $^{18}\text{F}$-FES-PET/CT imaging at baseline. Tumour $^{18}\text{F}$-FES-uptake was quantified for a maximum of 20 lesions and expressed as maximum standardised uptake value ($\text{SUV}_{\text{max}}$). CT-scan was repeated every 3 months to evaluate treatment response. Clinical benefit was defined as time to radiologic or clinical progression $\geq 24$ weeks.

Results  $^{18}\text{F}$-FES uptake, quantified for 255 lesions in 19 patients, varied greatly between lesions (median 2.8; range 0.6–24.3) and between patients (median 2.5; range 1.1–15.5). Seven (37 %) patients experienced clinical benefit of oestrogen therapy, eight progressed (PD), and four were non-evaluable due to side effects. The positive and negative predictive value (PPV/NPV) of $^{18}\text{F}$-FES-PET for response to treatment were 60 % (95 % CI: 31–83 %) and 80 % (95 % CI: 38–96 %), respectively, using $\text{SUV}_{\text{max}} > 1.5$.

Conclusion  $^{18}\text{F}$-FES-PET may aid identification of patients with acquired antihormone resistant breast cancer that are unlikely to benefit from oestradiol therapy.

Keywords  Oestrogen receptor · Imaging · Breast cancer · Oestradiol therapy · $^{18}\text{F}$-FES-PET

Introduction

Until the introduction of tamoxifen, additive oestrogens such as the synthetic diethylstilbestrol (DES) were considered the hormone treatment of choice in postmenopausal women. In a randomised study in 143 postmenopausal patients with metastatic breast cancer, first-line endocrine therapy with DES was equally effective as tamoxifen with a response rate of 41 % vs. 33 %. Yet, tamoxifen became the preferred agent because it showed fewer side effects [1]. An emerging number of anti-oestrogen therapies have become available since. Recently, however, additive oestrogen therapy has regained interest by showing efficacy in $\approx 35$ % of patients that are extensively pretreated with anti-oestrogens [2]. Interestingly, an update of the randomised study showed a superior 5-year survival for DES compared to tamoxifen (35 % vs. 16 %) after 14 years of follow-up [3]. Moreover, in a recent study a lower dose of only 6 mg oestradiol rendered similar clinical benefit rates as 30 mg oestradiol with fewer side effects. Finally, clinical results suggest that oestrogens can restore the sensitivity to anti-oestrogens [4]. As the majority of patients will not benefit from additive oestrogen therapy, a biomarker for patient selection would be helpful.
In preclinical studies, long-term oestrogen deprivation triggered hypersensitivity to oestrogens, which is accompanied by a five- to 10-fold increase in ER expression [5, 6]. Thus, patients that have been treated with anti-oestrogens for a long time may likewise have become hypersensitive to oestrogens. If so, patients that are most likely to benefit from oestradiol therapy could potentially be identified by high tumour ER expression. Positron emission tomography (PET) with 16α-[18F]fluoro-17β-oestradiol (18F-FES) can visualise and quantify ER expression in breast cancer lesions [7]. The aim of this exploratory study was, therefore, to evaluate 18F-FES-PET as a potential marker to select breast cancer patients for oestradiol therapy.

In the setting of ER-positive metastatic breast cancer, response assessment is notoriously difficult due to the high incidence of bone metastases. Bone is the most common site affected in breast cancer [8]. However, bone metastases are regarded non-measurable by the response evaluation criteria in solid tumours (RECIST) [9]. This underlines the need for objective measures to predict and evaluate response, for example by molecular imaging techniques. Recent studies have shown that 18F-FES-PET can predict response to various forms of anti-oestrogen therapy [10–13]. Its value as a biomarker for additive oestrogen therapy is, however, unknown.

In addition to molecular imaging techniques, also serum markers may be valuable to assess response in patients with bone-dominant disease in which response assessment is difficult. The ASCO recommendations for the use of tumour markers in breast cancer indicate that CA 15.3 and CEA can be regarded non-measurable by the response evaluation criteria in solid tumours (RECIST) [9]. This underlines the need for objective measures to predict and evaluate response, for example by molecular imaging techniques. Recent studies have shown that 18F-FES-PET can predict response to various forms of anti-oestrogen therapy [10–13]. Its value as a biomarker for additive oestrogen therapy is, however, unknown.

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approximately 200 MBq $^{18}$F-FES intravenously. $^{18}$F-FES-PET/CT to evaluate tumour ER-expression was performed at baseline on a hybrid PET/CT camera with a 64-slice CT and high definition and time-of-flight PET (Siemens Medical Systems). Low-dose CT-scan was used for attenuation correction in all patients. Patients were scanned from skull to mid-thigh, 3 min per bed position (usually 7–8 bed positions per patient). In all patients, baseline $^{18}$F-FES-PET was combined with a contrast-enhanced diagnostic CT scan. For representative $^{18}$F-FES-PET, CT and $^{18}$F-FES-PET/CT images see Fig. 1.

Tumour $^{18}$F-FES uptake was quantified by a nuclear medicine physician according to the guidelines of the European Association of Nuclear Medicine (EANM) for an arbitrary maximum of 20 randomly chosen lesions per patient [21]. Whole-body CT-scan was evaluated by a radiologist and used to allocate tumour lesions and identify possible $^{18}$F-FES-negative lesions. Lesion $^{18}$F-FES-uptake was expressed as maximum standardised uptake value ($SUV_{max}$). To calculate the predictive value of $^{18}$F-FES-PET for treatment response in individual patients, the median $SUV_{max}$ of all lesions quantified was used. Quantification of tumour $^{18}$F-FES-uptake was performed while blinded for treatment outcome. Patients and treating physician were held blinded for $^{18}$F-FES-PET results.

Tumour markers (CA-15.3, CEA) and bone turnover markers (PINP, CTx, and BALP) were also determined at baseline and repeated every 3 months or at the time of progression. Patients were considered evaluable for tumour marker response if one or both tumour markers were increased at baseline (CA15.3 > 33 kU/L, CEA > 5 μg/L). A 10 % increase in tumour marker was defined as biochemical progression. Patients were considered evaluable for bone turnover markers when they had evidence of bone metastases on imaging. Serum PINP > 95 ng/mL was considered the threshold for increased bone turnover based on literature [17]. For CTx and BALP, the optimum thresholds were determined by receiver operating characteristic (ROC) analysis.

Statistical analysis

The expected study time frame was 3 years for inclusion of 50 patients to evaluate the positive predictive value (PPV) and negative predictive value (NPV) of $^{18}$F-FES-PET/CT for response to oestradiol therapy by ROC analysis. After 3 years and 21 patients included, the study was terminated. We here report the PPV and NPV for $^{18}$F-FES-PET/CT, which was the predefined primary end point of the study. PPV and NPV were calculated using a ROC analysis for the median tumour $SUV_{max}$ in patients. The association between biochemical tumour markers and bone turnover markers and benefit from oestradiol was calculated by a Mann–Whitney U test. Analyses were performed in SPSS Statistics version 20.0.

Results

Patient characteristics

Between May 2010 and May 2013, 30 patients were screened for participation in the trial, out of whom 21 were included and 19 started oestradiol therapy, one man and 18 women. Mean age was 57 years (range 36–76). Seventeen patients had bone metastases; 14 patients also had visceral or nodal metastases. Two patients had only visceral lesions. All patients had postmenopausal status, which was in two patients achieved by the use of LHRH agonists, while others were truly postmenopausal. Tumour histology was positive for ER in all patients, 12 (63 %) were also PR positive, and none were HER2 positive. All patients were heavily pre-treated; 11 patients had already received 3–4 lines of systemic therapy, and seven patients ≥ 5 lines. Patient characteristics are summarised in Table 1. For an overview of screening, inclusion, and exclusion see the CONSORT diagram (Fig. 2).
Tumour response

Twelve patients had measurable lesions on baseline CT according to RECIST, four patients had non-measurable visceral lesions, and three patients had only bone metastases. Four of 19 (21%) discontinued oestradiol because of side effects and were, therefore, not evaluable for treatment response.

Seven of the remaining 15 patients experienced clinical benefit from oestradiol therapy as indicated by stable disease ≥24 weeks. Four had radiological measurable stable disease, and three patients had no new lesions detected on radiological examination, no progression of non-measurable lesions, improvement or stabilization of symptoms, and no evidence of biochemical progression ≥24 weeks. They eventually experienced PD according to RECIST criteria at 26, 28, and 48 weeks, respectively.

Finally, eight patients had PD; in five there was radiologic PD and in three patients there was substantial clinical deterioration, meriting discontinuation of therapy. One had laboratory signs of bone marrow invasion, confirmed with a biopsy; one had rising liver function test values, a threefold increase in tumour marker CA15.3, and clinical deterioration; and one patient had deterioration of pain symptoms from bone lesions, rising alkaline phosphatase, and worsening of performance score. Overall clinical benefit rate was 37% in all treated patients (intention-to-treat; n=19 patients) and 47% in all patients available for response assessment (n=15 patients). Mean progression-free-survival was 4.7 months (range 0.4–15.3 months). Oestradiol therapy induced an increase in serum oestradiol levels from 89±15 pmol/L at baseline to 1241±225 pmol/L after 1 month of therapy. The increase in oestradiol levels was equal among responders and non-responders.

Toxicity

In four patients (21%), oestradiol therapy was terminated prematurely due to adverse events. These side effects were progressive thrombocytopenia (n=1), transient ischemic accident with atrial fibrillation (n=1), mood disorders (n=1), and signs of congestive heart failure (n=1). Other grade 3 serious adverse events requiring hospital admission were tumour flare (n=1), hypercalcaemia (n=2), pneumonia (n=1), and atrial fibrillation (n=1). The patient who experienced a tumour flare had rapid increase of pain symptoms at known metastatic sites, starting as early as the day after initiation of oestrogen therapy. Laboratory results were suggestive of tumour flare with increased lactate dehydrogenase and other liver enzymes. Symptoms and laboratory findings resolved after oestradiol discontinuation. Interestingly, this patient with grade 3 clinical flare reaction showed highest tumour uptake of 18F-FES (median SUV_{max} 15.5, maximum SUV_{max} 24.3). Common,
but manageable grade 1–2 adverse events were tumour flare, fatigue, nausea, and vaginal bleeding.

**Predictive value of $^{18}$F-FES-PET for response to oestradiol therapy**

$^{18}$F-FES uptake in tumour lesions was quantified for a total of 255 lesions (214 bone, 24 lung, 12 lymph nodes, one breast, one soft-tissue, and one brain lesion) out of which 42 (16 %) were $^{18}$F-FES-negative ($\text{SUV}_{\text{max}}$ < 1.5). Twelve out of 19 patients (63 %) had only $^{18}$F-FES-positive lesions, six (32 %) had both FES-positive and FES-negative lesions, and one had only FES-negative lesions. Absolute $^{18}$F-FES-uptake ($\text{SUV}_{\text{max}}$) varied widely between lesions (median 2.8; range 0.6–24.3) and patients (median 2.5; range 1.1–15.5), as is depicted in Fig. 3a and b. ROC analysis indicated that the most optimum threshold to differentiate between patients with clinical benefit and patients with PD was a median $\text{SUV}_{\text{max}}$ of >1.5. For patients with response assessment available ($n=15$ patients), this threshold produced a PPV of 60 % (95 % CI: 31–83 %) and an NPV of 80 % (95 % CI: 38–96 %) (Fig. 4a), with an area under curve of 0.62.

Nine patients terminated treatment with ER-antagonists 5 weeks before initiating oestrogen therapy. Three of these patients had an earlier $^{18}$F-FES-PET obtained in another study (NCT01377324). These patients had much lower $^{18}$F-FES-uptake than on the earlier scans and several lesions could no longer be observed. For example, one patient had, on earlier $^{18}$F-FES-PET, a median tumour $^{18}$F-FES-uptake of 6.5, while in the current study, $\text{SUV}_{\text{max}}$ was only 1.1. This patient benefited from oestradiol despite the relatively low tumour $^{18}$F-FES-uptake. The remaining six patients had no earlier scans available, but also had relatively low tumour $^{18}$F-FES-uptake compared to patients without recent use of drugs that can bind ER. Thus, ER-antagonists may reduce tumour $^{18}$F-FES-uptake beyond the currently used 5-week drug withdrawal period [18]. In an explorative analysis, using the results of the previous $^{18}$F-FES-PET scans instead of the current PET scans, the PPV and NPV increased to 64 and 100 %, respectively (Fig. 4b).

**Tumour markers and bone turnover markers**

Seventeen patients had increased tumour markers. A tumour marker response was seen in seven patients. The PPV and NPV for tumour marker response were 67 and 71 %, respectively.

Bone turnover markers were explored as potential effect sensors in patients with bone metastases. Mean levels were 175 ng/mL for P1NP (range 17–613 ng/mL), 90 U/L for BALP (range 27–298 U/L), and 467 pg/mL for sCTx (range 26–1369 pg/mL). Change in bone turnover markers was not associated with treatment response. Baseline P1NP levels were 85 vs. 234 ng/mL ($P=0.032$), and sCTx levels 195 pg/mL vs. 623 pg/mL ($P=0.032$) in patients with clinical benefit and PD, respectively. The PPV of baseline P1NP was 100 % (five out of five patients with P1NP $\leq$ 95 ng/mL responded) and the NPV 88 % (seven out of eight patients with P1NP $>95$ ng/mL progressed) [17]. All four patients with sCTx levels $<200$ pg/mL responded, and seven of nine patients with sCTX $>200$ pg/mL progressed. BALP levels were non-informative in this exploratory study. The PPV and NPV for $^{18}$F-FES-PET/CT, tumour markers, and the bone turnover marker P1NP are shown in Table 2.
This is the first exploratory study evaluating \(^{18}\text{F}-\text{FES}\)-PET/CT as predictive marker for oestradiol therapy in patients with metastatic endocrine resistant breast cancer. While the mechanism of anti-oestrogen therapy is well known, this is not the case for the addition of oestrogens. Based on preclinical data, we hypothesised that very high \(^{18}\text{F}-\text{FES}\) uptake would predict response to oestradiol therapy. The value of \(^{18}\text{F}-\text{FES}\)-PET, however, turned out to be especially its ability to identify patients that are unlikely to benefit from oestradiol therapy as a result of low or absent \(^{18}\text{F}-\text{FES}\) uptake in metastases.

There are currently no good upfront predictive biomarkers to select patients for oestradiol therapy. Assessing ER status by a biopsy is the current gold standard, but is sometimes unreliable due to heterogeneous ER expression within and among lesions, and detection of non-functional ER. \(^2\text{'-[18F]fluoro-2'-deoxyglucose (18F-FDG) PET imaging has been tested to predict response to oestriol therapy. A study randomised 66 patients to 6 or 30 mg oestriol daily, 43 patients underwent 18F-FDG-PET imaging before and 24 h after the initiation of oestriol therapy [4]. A metabolic flare reaction upon oestriol therapy, predefined as a \(\geq 12\%\) increase in tumour \(^{18}\text{F}-\text{FDG}\)-uptake, had a PPV of 80\% (12 of 15 patients), and an NPV of 87\% (27/31 patients) for response to oestriol therapy. Metabolic flare on \(^{18}\text{F}-\text{FDG}\)-PET in 51 patients subsequently treated with an aromatase inhibitor or fulvestrant had an even higher PPV and NPV of 100 and 94\% [12]. \(^{18}\text{F}-\text{FES}\)-PET was evaluated earlier in three studies as a predictive biomarker before the initiation of aromatase inhibitors, tamoxifen, or fulvestrant. In these studies, the PPV of \(^{18}\text{F}-\text{FES}\)-PET ranged between 32 and 79\% and the NPV between 82 and 100\% [10–12], which is comparable with our findings. It is hypothesised that a negative \(^{18}\text{F}-\text{FES}\)-PET can identify tumours that have lost ER-expression during the course of disease [7, 10–12]. Recently, \(ESRI\) (ER) gene mutations have been described, some of which strongly reduce ligand binding affinity and induce endocrine resistance [22]. These phenomena might explain the good NPV of low tumour uptake of \(^{18}\text{F}-\text{FES}\).

Our study has some limitations. First, the number of patients included was lower than expected. A possible explanation is that, despite the fact that previous studies have shown the safety and benefits of oestriol therapy, physicians may be reluctant to refer patients for oestrogen therapy given that anti-oestrogen therapy is the key method to treat patients with ER-positive disease. Secondly, given the low number of measurable lesions in this population, our study was not powered.

**Table 2** Association between \(^{18}\text{F}-\text{FES}\)-PET, tumour markers, bone marker, and treatment outcome

| Marker          | Result          | n  | Response | Predictive value | PPV | NPV |
|-----------------|-----------------|----|----------|------------------|-----|-----|
| \(^{18}\text{F}-\text{FES}\)-PET | SUV \(\geq 1.5\) | 10 | 6 4 (4) | 60\% |      |     |
|                 | SUV <1.5        | 5  | 1 4 (0) | 80\% |      |     |
| Tumour marker   | R               | 6  | 4 2 (1) | 67\% |      |     |
|                 | NR              | 7  | 2 5 (3) | 71\% |      |     |
|                 | NE              | 2  | 1 1 (0) |     |      |     |
| Bone marker     | <95 pg/mL       | 5  | 5 0 (1) | 100\% |     |     |
|                 | \(\geq 95\) pg/mL | 8  | 1 7 (3) | 88\% |      |     |

CB clinical benefit, PD progressive disease, NE non-evaluable, PPV positive predictive value, NPV negative predictive value, R response, NR non-response
to evaluate the correlation between $^{18}$F-FES uptake and response in individual lesions. Also, we used CT to identify $^{18}$F-FES-negative lesions. It is possible that $^{18}$F-FDG-PET together with $^{18}$F-FES-PET increases the sensitivity for $^{18}$F-FES-negative lesions, since bone lesions are especially difficult to characterise on CT. Finally, when evaluating the predictive value of $^{18}$F-FES-PET, it is important to take concomitant and recent therapies into account. We observed low $^{18}$F-FES-uptake in several previously $^{18}$F-FES-avid lesions in patients that had used ER-antagonists up to 5 weeks before $^{18}$F-FES-PET. It is possible that the long half-lives ($t_{1/2}$) of fulvestrant ($t_{1/2}$=40 days) and of tamoxifen and metabolites ($t_{1/2}$=9 and 13 days, respectively) are responsible for the $^{18}$F-FES-negative results [23–25]. However, in the absence of biopsies, we are unable to dissect fully whether the observed effects can be attributed to altered ER-expression or to spill-over effects of recent therapies.

In preclinical studies, long-term oestrogen-deprived ER-positive breast cancer cells are used to study oestrogen-induced apoptosis. After several months of culturing in oestrogen-deprived conditions, breast cancer cells adapt to the low levels of oestrogens by increasing ER expression [26, 27]. Paradoxically, therapeutic doses of oestrogens now no longer induce growth proliferation, but induce apoptosis. More recently, $ESR1$ gene amplification was described in patient-derived mouse xenografts as a possible marker for hypersensitivity to oestradiol [28]. It would, therefore, be of interest in future studies to combine $^{18}$F-FES-PET/CT with analysis of $ESR1$ gene amplification and mutation in tumour biopsies, in order potentially to improve the selection of patients for oestrogen therapy. Although ER expression is required for response to endocrine agents, ER-positive tumours may still fail to respond, e.g., due to cross talk with other pathways. We, therefore, evaluated whether the addition of tumour and bone turnover markers could improve response prediction. The association of tumour marker response alone with the patient response classification was modest and did not add significantly to $^{18}$F-FES-PET/CT. For bone turnover markers, not the changes in bone markers during treatment, but the pre-treatment values were associated with time-to-progression. These markers are known to correlate with the number and size of bone metastases in breast and prostate cancer [15–17]. Therefore, high serum bone markers may be useful to identify patients that have a poor prognosis independent of the therapy given. Whether bone markers are of prognostic or predictive value needs to be addressed in larger studies, adhering to REMARK criteria [29].

We observed a clinical benefit rate of 37 % for oestradiol 6 mg orally daily, which is comparable to the study by Ellis et al. [4]. Our study is the second evaluating this low-dose regimen, as compared to the previous standard of 30 mg orally daily. The clinical benefit rate observed in our study in patients that were extensively pre-treated (median of four prior regimens) provides further evidence for the efficacy of oestradiol 6 mg daily. The 21 % of patients that terminated treatment prematurely due to toxicity was relatively high; grade 3 adverse events were noted in 42 % of the patients. The high incidence of toxicity, however, underlines the value of upfront predictive markers for this treatment.

Conclusion

Patients with acquired endocrine-resistant metastatic breast cancer may paradoxically benefit from oestradiol therapy. The relatively low response rate and toxicity accompanying oestradiol therapy warrants exploration of potential biomarkers to predict response. $^{18}$F-FES-PET/CT may aid to identify patients that are unlikely to respond to oestradiol therapy.

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Compliance with Ethical Standards

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Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval All procedures were performed in accordance with the ethical standards of the institutional and national research committee, and the declaration of Helsinki.

Informed consent Informed consent was obtained from all individual participants included in the study.

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References

1. Ingle JN, Ahmann DL, Green SJ, Edmonson JH, Bisel HF, Kvols LK, et al. Randomized clinical trial of diethylstilbestrol versus tamoxifen in postmenopausal women with advanced breast cancer. N Engl J Med. 1981;304:16–21.
2. Lonning PE, Taylor PD, Anker G, Iddon J, Wie L, Jorgensen LM, et al. High-dose estrogen treatment in postmenopausal breast cancer
patients heavily exposed to endocrine therapy. Breast Cancer Res Treat. 2001;67:111–6.
3. Peethambaram PP, Ingle JN, Suman VJ, Hartmann LC, Loprinzi CL. Randomized trial of diethylstilbestrol vs. tamoxifen in postmenopausal women with metastatic breast cancer. An updated analysis. Breast Cancer Res Treat. 1999;54:117–22.
4. Ellis MJ, Gao F, Dehdashti F, Jeffe DB, Marcom PK, Carey LA, et al. Lower-dose vs high-dose oral estradiol therapy of hormone receptor-positive, aromatase inhibitor-resistant advanced breast cancer: a phase 2 randomized study. JAMA. 2009;302:774–80.
5. Song RX, Santen RJ. Apoptotic action of estrogen. Apoptosis. 2003;8:55–60.
6. Song RX, Zhang Z, Mor G, Santen RJ. Down-regulation of Bcl-2 enhances estrogen apoptotic action in long-term estradiol-depleted ER(+) breast cancer cells. Apoptosis. 2005;10:667–78.
7. van Kruchten M, de Vries EG, Brown M, de Vries EF, Glaudemans AW, Dierckx RA, et al. PET imaging of oestrogen receptors in patients with breast cancer. Lancet Oncol. 2013;14:e465–75.
8. Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, et al. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). Eur J Cancer. 2009;45: 228–47.
9. Mortimer JE, Dehdashti F, Siegel BA, Trinkaus K, Katzenellenbogen JA, Welch MJ. Metabolic fluore: indicator of hormone responsiveness in advanced breast cancer. J Clin Oncol. 2001;19:2977–803.
10. Linden HM, Stekhova SA, Link JM, Gralow JR, Livingston RB, Ellis GK, et al. Quantitative fluoroestradiol positron emission tomography imaging predicts response to endocrine treatment in breast cancer. J Clin Oncol. 2006;24:2793–9.
11. Dehdashti F, Mortimer JE, Trinkaus K, Naughton MJ, Ellis M, Katzenellenbogen JA, et al. PET-based estradiol challenge as a predictive biomarker of response to endocrine therapy in women with estrogen-receptor-positive breast cancer. Breast Cancer Res Treat. 2009;113:509–17.
12. van Kruchten M, de Vries EG, Glaudemans AW, van Lanschot MC, van Faassen HJR, Kema IP, et al. Measuring residual estrogen receptor availability during fulvestrant therapy in patients with metastatic breast cancer. Cancer Discov. 2015;5:72–81.
13. Harris L, Fritsche H, Mennel R, Norton L, Ravdin P, Taube S, et al. American Society of Clinical Oncology. American Society of Clinical Oncology 2007 update of recommendations for the use of tumor markers in breast cancer. J Clin Oncol. 2007;25:5287–312.
14. Leeming DJ, Koizumi M, Qvist P, Barkholt V, Zhang C, Henriksen K, et al. Serum N-terminal propeptide of collagen type I is associated with the number of bone metastases in breast and prostate cancer and correlates to other bone related markers. Cancer Biomark. 2011;3:15–23.
15. Voorzanger-Rousselot N, Juillet F, Mareau E, Zimmermann J, Kalebic T, Gamero P. Association of 12 serum biochemical markers of angiogenesis, tumour invasion and bone turnover with bone metastases from breast cancer: a crosssectional and longitudinal evaluation. Br J Cancer. 2006;95:506–14.
16. Luftner D, Jozezaur D, Schildhauer S, Geppert R, Muller C, Fiolk G, et al. PINP as serum marker of metastatic spread to the bone in breast cancer patients. Anticancer Res. 2005;25:1491–9.
17. Leeming DJ, Koizumi M, Qvist P, Barkholt V, Zhang C, Henriksen M, et al. FDG PET and PET/CT: EANM procedure guidelines for tumour PET imaging: version 1.0. Eur J Nucl Med Mol Imaging. 2010;37:181–200.
18. Roemer J, Steinbach J, Kasch H. Studies on the synthesis of 16α-[18F]fluoroestradiol. Appl Radiat Isot. 1996;47:395–9.
19. Merenbakh-Lamin K, Ben-Baruch N, Yeheskel A, Dvir A, Soussan-Gutman L, Jeselsohn R, et al. D538G mutation in estrogen receptor-alpha: a novel mechanism for acquired endocrine resistance in breast cancer. Cancer Res. 2013;73:6586–64.
20. Boellaard R, O’Doherty MJ, Weber WA, Mottaghy FM, Lonsdale MN, Stroobants SG, et al. FDG PET and PET/CT: EANM procedure guidelines for tumour PET imaging: version 1.0. Eur J Nucl Med Mol Imaging. 2010;37:181–200.
21. Jesselsohn R, Yelensky R, Buchwalter G, Frampton GM, Meric-Bernstam F, Gonzalez-Angulo AM, et al. Emergence of constitutively active estrogen receptor-α mutations in pretreated advanced estrogen receptor positive breast cancer. Clin Cancer Res. 2014;20:1757–67.
22. Robinson DR, Wu YM, Vats P, Su F, Lonigro RJ, Cao X, et al. Activating ESR1 mutations in hormone-resistant metastatic breast cancer. Nat Genet. 2013;45:1446–51.
23. Song RX, Gao F, Dehdashti F, Jeffe DB, Marcom PK, Carey LA, et al. Effect of long-term estrogen deprivation on apoptotic response of breast cancer cells to 17β-estradiol. J Natl Cancer Inst. 2001;93:1714–23.
24. Santen RJ, Song RX, Masamura S, Yue W, Fan P, Sogon T, et al. Adaptation to estradiol deprivation causes up-regulation of growth factor pathways and hypersensitivity to estradiol in breast cancer cells. Adv Exp Med Biol. 2008;630:19–34.
25. Li S, Shen D, Shao J, Crowder R, Liu W, Prat A, et al. Endocrine-therapy-resistant ESR1 variants revealed by genomic characterization of breast-cancer-derived xenografts. Cell Rep. 2013;4:1116–30.
26. McShane LM, Altman DG, Sauerbrei W, Taube SE, Gion M, Clark GM. Statistics Subcommittee of the NCI-EORTC Working Group on Cancer Diagnostics. Reporting recommendations for tumor marker prognostic studies (REMARK). J Natl Cancer Inst. 2005;97:1180–4.