Unusual manifestation of *Helicobacter cinaedi* infection: a case report of intracranial subdural empyema and bacteremia

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**Abstract**

**Background:** There have been various reports concerning *Helicobacter cinaedi* infections. However, few reports have examined central nervous system infections.

**Case presentation:** A 52-year-old man was transferred from the local hospital because of a persistent headache and suspected intracranial subdural empyema. Neurosurgical drainage was performed via burr holes. Gram staining and results from abscess cultures were negative. The blood culture yielded *H. cinaedi*. He was given an antibiotic regimen consisting of 2 g of ceftriaxone twice a day, but the size of the abscess was not reduced in size at all after 3 weeks of treatment. Neurosurgical drainage was performed again, and the antimicrobial regimen was switched to 2 g of meropenem 3 times a day. The size of the abscess was reduced after 2 weeks of the second drainage and antimicrobial drug change to meropenem. After 4 weeks treatment with meropenem, the patient was discharged, and his symptoms had completely resolved.

**Conclusions:** *H. cinaedi* infection should be considered in the differential diagnosis of subdural empyema cases for which Gram staining and abscess culture results are negative. Meropenem can be a first-line drug of choice or an effective alternative treatment for *H. cinaedi* central nervous system infections.

**Keywords:** *Helicobacter cinaedi*, Intracranial subdural empyema, Antimicrobial susceptibility testing, Case report

**Background**

The first report of a *Helicobacter cinaedi* infection involved a man with proctitis in 1984 [1]. Since then, various foci of *H. cinaedi* infection have been reported. However, few reports have examined central nervous system (CNS) infections, and the optimal therapy for CNS infection is unknown. Here, we report a case of intracranial subdural empyema and bacteremia due to *H. cinaedi*, in which the patient experienced treatment failure after a maximum dose of ceftriaxone. Finally, he was treated successfully with adequate drainage and meropenem.

**Case presentation**

A 52-year-old man with a history of epilepsy and drug eruption due to an amoxicillin/clavulanate was transferred from the local hospital because of a persistent headache and a suspicion of chronic subdural hematoma. There were no other symptoms before the persistent headache occurred, and he could perform his work normally. He had no history of head trauma and no meningeal irritation symptoms, such as neck stiffness. He kept an outdoor dog for years, but he had no contact with other animals, such as rats, hamsters, dogs, cats, birds, or monkeys, during the past year.

Computed tomography and nuclear magnetic resonance imaging of the head showed a right subdural mass with high and mixed density/intensity (Fig. 1a).

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Because of these findings, we suspected intracranial subdural empyema (SDE).

Neurosurgical drainage was performed via burr holes. Gram staining of the purulent material showed no bacteria, and he had no predisposition for sinusitis or periodontal disease. Therefore, we thought the possibility of anaerobic and aerobic gram-negative bacteria involvement was low. Two sets of blood cultures were drawn, and empirical antibiotic therapy was started with intravenous vancomycin targeting only aerobic streptococci and staphylococci. However, the results from abscess cultures were negative. After 7 days of incubation, the blood culture (BAC-TEC FX system, Nippon BD, Tokyo, Japan) yielded Gram-negative, long spiral-shaped bacillus. Microaerobic subculture on blood agar plates with hydrogen gas showed thin transparent colonies. Because of these findings, we suspected *Helicobacter cinaedi* intracranial subdural empyema. The human immunodeficiency virus antibody screening test administered just after surgery showed negative results. An additional two sets of blood culture specimens were drawn, and he was given an antibiotic regimen consisting of 2 g of ceftriaxone twice a day. The blood culture results were negative, but the size of the abscess was not reduced in size at all after 3 weeks of treatment (Fig. 1b).

Neurosurgical drainage was performed again, and the antimicrobial regimen was switched to 2 g of meropenem 3 times a day. Although the microaerobic culture of the abscess was negative, *H. cinaedi* was identified from the abscess and blood culture by cdtB virulence factor gene-based PCR. The 16S rRNA sequence analysis revealed with 100% similarity between the abscess and blood culture (Fig. 2). Antimicrobial susceptibility testing for *H. cinaedi* using the broth microdilution method revealed minimum inhibitory concentrations (MICs) of 4 μg/mL for ceftriaxone and 0.06 μg/mL for meropenem (Table 1). The size of the abscess was reduced after 2 weeks of the second drainage and antimicrobial drug change to meropenem (Fig. 1c). After 4 weeks treatment with meropenem, the patient was discharged, and his symptoms had completely resolved. The patient has continued visiting the hospital for more than 1 year after discharge, and there has been no recurrence to date (Fig. 1d).

**Conclusion**

This case revealed two important clinical issues. First, *H. cinaedi* can cause an intracranial SDE. Secondly, treatment failure with ceftriaxone can occur when the MIC value for ceftriaxone is 4 μg/mL or higher in an *H. cinaedi* intracranial SDE case.

To our knowledge, this is the first published case of *H. cinaedi* intracranial SDE. There have been four reported cases of *H. cinaedi* central nervous system infections. Three cases concerned meningitis in adults [2–4], and the final case was a case of meningitis and bacteremia in a neonate [5]. None of those reported cases were associated with abscess formation. We must consider *H. cinaedi* as a causative organism of culture-negative intracranial SDE.

Treatment failure with ceftriaxone in cases of *H. cinaedi* intracranial SDE can occur if the MIC value for ceftriaxone is 4 μg/mL or higher. We initially chose ceftriaxone as definitive therapy because antibiotic regimens including ceftriaxone effectively treated *H. cinaedi* meningitis in the previous four
case reports. Antimicrobial susceptibility testing (AST) for *H. cinaedi* isolates was not performed in these cases, probably because AST for *H. cinaedi* is too cumbersome to perform routinely in hospital laboratories [6].

However, in the present case, AST yielded useful information for changing the therapeutic strategy. With the doses normally used to treat bacterial meningitis, the concentrations of ceftriaxone in cerebrospinal fluid range from 2 to 8 μg/mL, and levels are nearly constant in children and adults [7, 8]. These concentrations are close to the MIC observed in this case (4 μg/mL), which may negate the culture results, but there is a possibility of treatment failure especially in the presence of an abscess. On the other hand, when 2 g of meropenem was administered every 8 h, the concentration in the cerebrospinal fluid was reported to be 1.29 μg/mL, even in the trough value [9], which is considerably higher than the MIC in this case (0.06 μg/mL). Therefore, meropenem can be a first-line drug of choice or an effective alternative treatment for CNS infection, especially when the MIC value is 0.06 μg/mL or lower.

In conclusion, *H. cinaedi* infection should be considered in the differential diagnosis of SDE cases for which Gram staining and abscess culture results are
negative. AST for H. cinaedi isolates must be performed for cases of CNS infections. When the MIC value to ceftriaxone is 4 μg/mL or higher, treatment failure can occur. Meropenem can be a first-line drug of choice or an effective alternative treatment for H. cinaedi CNS infections.

Abbreviations
AST: Antimicrobial susceptibility testing; CNS: Central nervous system; MIC: Minimum inhibitory concentration; SDE: Subdural empyema

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Availability of data and materials
Data sharing not applicable to this article as no datasets were generated or analyzed during the current study.

Table 1 Antimicrobial susceptibility testing results of the H. cinaedi isolate

| Antibiotic                        | MIC (μg/mL) |
|-----------------------------------|-------------|
| Penicillins                       |             |
| Amoxicillin                       | 4           |
| Carbenicillin                     | 8           |
| Piperacillin                      | 4           |
| Piperacillin/Tazobactam           | 4           |
| Cephalosporins                    |             |
| Cefepime                          | 4           |
| Ceftriaxone                       | 4           |
| Carbapenems                       |             |
| Imipenem                          | 0.06        |
| Meropenem                         | 0.06        |
| Aminoglycosides                   |             |
| Gentamicin                        | 0.25        |
| Kanamycin                         | 0.5         |
| Tetracycline                      |             |
| Tetracycline                      | 0.06        |
| Macrolides                        |             |
| Erythromycin                      | >64         |
| Quinolones                        |             |
| Ciprofloxacin                     | 16          |
| Levofoxacin                       | 4           |
| Moxifloxacin                      | 0.5         |
| Metronidazole                     |             |
| Metronidazole                     | >64         |

Authors’ contributions
MY, IY, and SK performed the isolation and identification of the organism. JT and YK performed 16S rRNA sequence analysis and antimicrobial susceptibility testing. TH treated the patient, and was a major contributor in writing the manuscript. All authors read and approved the final manuscript.

Competing interests
The authors declare that they have no competing interests.

Consent for publication
We have had a signed consent for publication from the patient.

Ethics approval and consent to participate
Not applicable.

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References
1. Pasternak J, Bolivar R, Hopfer RL, Fainstein V, Mills K, Rios A, Bodey GP, Fennell CL, Totten PA, Stamm WE. Bacteremia caused by campylobacter-like organisms in two male homosexuals. Ann Intern Med. 1984;101:339–41.
2. Sugiyama A, Mori M, Ishiwada N, Himuro K, Kuwabara S. First adult case of helicobacter cinaedi meningitis: J Neurol Sci. 2014;336:263–4.
3. Okubo H, Goto M, Sato M, Sugiyama T, Kawano M, Matsunaga T, Akaie T. Helicobacter cinaedi meningitis: a case report and review of previous cases. J Neurol Sci. 2014;347:396–7.
4. Uwamino Y, Muranaka K, Hase R, Otsuka Y, Hosokawa N. Clinical features of community-acquired helicobacter cinaedi bacteremia. Helicobacter. 2016;21:24–8.
5. Orlicek SL, Welch DF, Kuhls TL. Septicemia and meningitis caused by helicobacter cinaedi in a neonate. J Clin Microbiol. 1993;31:569–71.
6. Oyama K, Khan S, Okamoto T, Fuji S, Ono K, Matsunaga T, Yoshitake J, Sawa T, Noma K, Kawamura Y, et al. Identification of and screening for human helicobacter cinaedi infections and carriers via nested PCR. J Clin Microbiol. 2012;50:3893–900.
7. Del Rio M, McCracken Jr GH, Nelson JD, Crane D, Shelton S. Pharmacokinetics and cerebrospinal fluid bactericidal activity of ceftriaxone in the treatment of pediatric patients with bacterial meningitis. Antimicrob Agents Chemother. 1982;22:622–7.
8. Cabellos C, Viladrich PF, Verdaguer R, Pallares R, Linares J, Gudiol F. A single daily dose of ceftriaxone for bacterial meningitis in adults: experience with 84 patients and review of the literature. Clin Infect Dis. 1995;20:1164–8.
9. Blaschmann U, Roehr AC, Frey OR, Vetter-Kerkhoff C, Thon N, Hope W, Briegel J, Huse V. Cerebrospinal fluid penetration of meropenem in neurocritical care patients with proven or suspected ventriculitis: a prospective observational study. Crit Care. 2016;20:343.