Leaf Nitrogen Traits in Response to Plant Density and Nitrogen Supply in Oilseed Rape

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Abstract: Understanding the response of plant nitrogen (N) and carbon (C) economies in oilseed rape, as well as their role in defining phenotypic plasticity, is necessary for designing new strategies to optimize plant and canopy C assimilation to improve potential yield. This paper aims to elucidate the extent to which the interaction between N supply and plant population density alters N distribution in oilseed rape plant (Brassica napus L.) and whether this interaction changes plant investment in leaf area or leaf mass per area. Spring oilseed rape was grown at two rates of N supply (50 and 150 kg N·ha⁻¹) and two plant population densities (50 and 150 plants·m⁻²). Photosynthesis, leaf area, leaf biomass, and N content of selected leaves were measured at 20% of flowers on main raceme open. The interaction between N supply and plant population density altered leaf N content per area, which is the main determinant of photosynthesis. This interaction also affected leaf mass per area, while N supply determined N content per unit leaf mass. These results suggest that the interaction between N supply and population density affects both nitrogen distribution and leaf mass per area, which could have important implications for light distribution and, therefore, for C assimilation at the plant level.

Keywords: Brassica napus L.; leaf area; leaf mass per area; light interception; nitrogen distribution; nitrogen supply

1. Introduction

During the last 50 years, oil crops have been the commodities with the greatest relative increase in terms of contribution to the world’s food supply [1]. Oil from oilseed rape (Brassica napus L.) is the third most consumed vegetable oil after soybean and palm oil, while it is one of the healthiest edible oils for human consumption [2]. Additionally, the rising demand of biofuels [3] has resulted in an exponential growth of oilseed rape production since 1980, reaching 71 million tons in 2017 [4]. Future increases in crop production should be accompanied by improved sustainability of crop production systems; therefore, there is a need for developing and improving crop nitrogen (N) management which can maintain high levels of production, while minimizing N input [5], and more insight into regulatory mechanisms controlling plant N economy is vital for improving N-use efficiency [6,7]. Since the N economy is closely associated with the carbon (C) economy [8], the interaction between both should be considered.

Oilseed rape has the potential of maintaining a similar seed yield across a wide range of plant population densities [9,10], opening the opportunity to reduce sowing rates. As the light environment...
at every leaf and leaf traits are modified by plant population density, the plant acclimation and photosynthetic performance would be different as well. However, little is known on the effects of the interaction between N supply and light interception on plant and leaf architectural traits in oilseed rape.

Leaf photosynthetic capacity is strongly related to N content per unit of leaf area ($N_{\text{area}}$) [11], but wide variation in the photosynthetic capacity for a given $N_{\text{area}}$ has been observed in different plant species [12,13]. $N_{\text{area}}$ is associated with leaf mass per area ($LMA$ mg mass cm$^{-2}$), which is an indicator of the plant investment in light-capturing and photosynthetic organs [14]. This sensitivity to photosynthetically active radiation (PAR) contributes to explain the sensitivity of $LMA$ to different plant population densities observed [15]. However, the response of $LMA$ to N supply is less clear; this trait has been found to be negatively correlated with N supply [16] or not correlated at all [17], depending on the plant or crop species. The other physiological determinant of $N_{\text{area}}$ is N content per leaf mass ($N_{\text{mass}}$), but the environmental factors affecting $N_{\text{mass}}$ are poorly understood. Neither LAI nor $LMA$ was affected in response to N supply and plant population density in maize (Zea mays L.) [18].

The N demand of plants is not completely matched by the N supplied as fertilizer; available N is variable at a short time scale due to environmental constraints that affect the N cycle [19]. Under N deficiency, due to low availability in the soil, N distribution in plants is more dependent on N soil status and subsequent N uptake by the plant, rather than on leaf light interception [20]. Ipomoea tricolor [21], grown in an hydroponic solution at 0.12 and 12 mmol NO$_3^-$·L$^{-1}$, showed a decrease in $N_{\text{area}}$ associated with aging of leaves, while leaf $N_{\text{mass}}$ of Xanthium canadense was strongly affected by leaf position on the plant, independent of the relative photon flux density, in an experiment under low N availability, where the plants did not receive N fertilizer supply [22].

A substantial body of literature about N distribution stems from studies performed on woody, perennial crops or wild plant species [17,23,24], while only a limited number of studies have been performed on annual crops [25,26]. Since wild plants and crops differ in their preferences for N forms (i.e., NO$_3^-$, NH$_4^+$), mycorrhizal associations, amounts of N storage, breeding pressure, and adaptive responses to low soil N availability [27], the conclusions from ecological studies cannot be directly extrapolated to crop species such as oilseed rape, especially when grown under different plant population densities.

Understanding the combined effects of N supply and plant population density on leaf N distribution, as well as their impact on plant N and C partitioning, is of major importance for the design of new strategies to improve seed yield of oilseed rape. For that reason, the current study addresses the following research questions: (1) to what extent does the interaction between N supply and plant population density affect N distribution in oilseed rape? (2) does the plant respond to this interaction by modifying assimilate investment in leaf area or by modifying leaf mass per area (LMA)?

To answer these questions, we focused on the plant developmental stage Biologische Bundesanstalt, Bundessortenamt and Chemical industry (BBCH) 62 (20% of flowers on main raceme open) because the plant carbon status during the period bracketing flowering determines branch appearance [28] and seed yield in oilseed rape [29,30]. Moreover, while exploring this period, N remobilization from leaves to reproductive organs is largely avoided.

2. Materials and Methods

2.1. Experimental Set-Up and Treatments

On 17 April 2013, spring oilseed rape (B. napus “Solar CL”) was sown under an open-sided shelter with transparent roof in Wageningen, the Netherlands (51°58’ north (N), 5°40’ east (E)). The treatments consisted of combinations of two rates of N fertilization, 50 (N50) and 150 (N150) kg ha$^{-1}$, and two plant population densities, 50 (D50) and 150 (D150) plants m$^{-2}$, with the goal to obtain contrasting N supply and light environments and, thus, different N distribution at the plant level. The plant population density of 150 plants m$^{-2}$ was chosen according to similar experiments performed in spring oilseed rape [31]. On the other hand, we assessed a contrasting low population density (50 plants-m$^{-2}$) taking
into account that this density increases branching and modifies carbon partitioning within the plants. The N supplies of 150 and 50 kg N·ha\(^{-1}\) applied in our experiment were also contrasting and similar to the highest and lowest N availabilities used in a study performed under similar conditions [31].

Treatment effects were tested on plants growing in containers (0.7 × 0.9 m), which contained from bottom to top a layer of 35 cm of sandy soil, upon which fertilizers (Ca\((H_2PO_4)\_2\) and K\(_2SO_4\cdotMgSO_4\)) were applied and covered with 1 cm of soil. On this layer, seeds were placed in a hexagonal spatial pattern and covered with a layer of 2 cm soil. The containers were arranged closely together to ensure canopy homogeneity, and 12 guard containers were placed around the experimental containers aimed at avoiding border effects on the plants to be measured. Treatments were assessed in a single container and we used individual plants as pseudo-replicates as in previous studies [32–34], which, in agreement with the homogeneity of the soil (coefficient of variation of N present in the soil at sowing was 3.7%; data not shown) and plants in this well-managed experiment, allowed very high similarity across plants at each experimental unit. In addition, the containers were rearranged twice to reduce position effects in the open-sided shelter. Phenological growth stages in oilseed rape were recorded according to the coding of developmental growth stages of mono- and dicotyledonous plant species designed by the Biologische Bundesanstalt, Bundessortenamt and Chemical industry (also known as BBCH phenological scale) [35].

The N fertilizer KAS-27 (13.5% NO\(_3^-\) and 13.5% NH\(_4^+\)) was applied to the soil by hand (containing 30 kg N·ha\(^{-1}\) and 3.1% organic material) to reach 50 and 150 kg N·ha\(^{-1}\). In the lower N treatment, 50 kg N·ha\(^{-1}\) was applied 2 days after emergence, while, in the higher N treatment, 150 kg N·ha\(^{-1}\) was split into two applications: 100 kg·ha\(^{-1}\) 2 days after emergence (BBCH 10) and 50 kg·ha\(^{-1}\) when the fifth internode was expanded (BBCH 35) (Figure 1). The containers were irrigated daily or twice per day dependent on the environmental temperature, while fungal diseases such as *Phoma lingam* and *Sclerotinia sclerotiorum* were chemically controlled by using the fungicides prothioconazole (Proline®, Bayer CropScience, Monheim, Germany) and iprodione (Rovral®, BASF, Ludwigshafen, Germany), respectively. Weeds were manually removed.

\[\text{Figure 1. Schematic diagram of oilseed rape developmental stages, leaf appearance, moment of N fertilization, and measurements.}\]
2.2. Temperature Conditions

Temperature was recorded (Data taker DT600, Datataker Data Loggers, Cambridgeshire, UK) with shielded thermocouples (type T, TempControl Industrial Electronic Products, Voorburg, The Netherlands) at 15 min intervals from sowing to flowering. Thermocouples were placed at mid-canopy and below the soil surface at 7 cm depth. Plant development and leaf appearance were expressed in thermal time ($TT$) after emergence and calculated by summing daily mean temperatures ($T_{\text{mean}}$) using a base temperature ($T_{\text{base}}$) of 0 °C for spring oilseed rape; $n$ is the variable being summed up to and $i$ is the starting term [36].

$$TT = \sum_{i}^{n} (T_{\text{mean}} - T_{\text{base}}), \quad (1)$$

For the first 15 days after emergence, thermal time was estimated on the basis of soil temperature, since the apex was below the soil surface; canopy temperature was used thereafter.

2.3. Plant Measurements

One week after emergence, 10 representative plants of similar plant height, leaf size, and color grown in the center of the evaluated containers were randomly selected and labeled for each treatment (five plants for destructive measurements and five for photosynthesis measurements), and the leaves were coded according to their position on the stem (the phytomer rank). A leaf was considered to have appeared halfway between the last observation when it was absent and the first observation when the tip was visible. For each plant selected, the time of leaf tip appearance of leaf 9 and 11 on the main stem, as well as leaf 9.1 (i.e., the first leaf on the primary branch outgrowing from phytomer 9 on the main stem), was recorded twice a week from emergence (BBCH 10) to 20% of flowers on main raceme open (BBCH 62) in order to estimate the time of leaf appearance. Because, in oilseed rape, leaves placed at lower phytomers differ in their size, shape, and presence of petiole in comparison with middle and upper leaves [37], and because the beginning of flowering assimilate remobilization and/or senescence was evidenced in leaves positioned below phytomer rank 3, only sessile leaves positioned at higher phytomer ranks were evaluated. Plant height was measured at the beginning of flowering, as the distance from the base and the top of the main stem.

Light interception ($IPAR$) was measured twice per week at the canopy level in all the containers evaluated; thus, light interception by leaves located in a given layer was estimated by assuming $IPAR$ ($\mu$mol·m$^{-2}$·s$^{-1}$) to attenuate through the leaf canopy following the Beer–Lambert law [38].

$$IPAR = PAR_{\text{inc}} \times (1 - e^{-k \times LAI}), \quad (2)$$

where $k$ is the light extinction coefficient and depends upon the architecture of a canopy and transmission of radiation through individual leaves [39]; it was estimated by the following function:

$$k = -\ln (PAR_{\text{trans}}/PAR_{\text{inc}})/LAI, \quad (3)$$

Incident light ($PAR_{\text{inc}}$, $\mu$mol·m$^{-2}$·s$^{-1}$) was recorded by the BF5 Sunshine Sensor (Delta-T Devices Ltd., Cambridge, United Kingdom), which was placed on a horizontal level 1.5 m close to the container evaluated, and transmitted light ($PAR_{\text{trans}}$, $\mu$mol·m$^{-2}$·s$^{-1}$) was measured at ground level with a 1 m sensor probe of the SunScan Canopy Analysis System (Delta-T Devices Ltd., Cambridge, UK).

Leaf area index ($LAI$) is the total green leaf area by ground surface area. Cumulative $LAI$ was estimated for each rank phytomer and, in this way, $IPAR$ of individual leaves was estimated using Equation (2), assuming that each leaf layer has a similar $k$.

Net assimilation rate ($A_n$) measurements were done 5–7 days before flowering on leaves 9, 9.1, and-11 for treatment N150-D50 and on leaves 9 and 11 for treatments N50-D50, N150-D150,
and N50-D150 because these treatments did not produce branches or leaf 9.1. At that moment, the evaluated leaves were fully expanded. The photosynthesis rate was determined with a LI-COR 6400XT Portable Photosynthesis System (LI-COR, Lincoln, NE, USA) at saturated incident light on surface area \(I_{\text{inc}}\) \(= 1500 \text{ mmol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}\), \(\text{CO}_2 = 390 \text{ mmol} \cdot \text{mol}^{-1}\) and at a leaf temperature of 25 °C.

At BBCH 62, a destructive sampling was performed, and the leaf area of each individual leaf at each phytomer from the top of the canopy was measured. For this purpose, five plants per treatment (previously tagged) were harvested, and the leaf area of individual leaves 9, 9.1, and 11 was assessed (LI 3100; LI-COR, Lincoln, NE, USA). After oven-drying (at 70 °C for 48 h), the dry matter of leaves was also recorded. The remaining leaves and stems were also collected, to estimate dry matter and N uptake, but detached leaves were not considered for the analysis. N contents (%) of individual leaves, remaining leaves, and stems were determined in an N elemental analyzer using the combustion method of Dumas (Flash 2000, Thermo Fisher Scientific Inc., Delft, The Netherlands).

With this information, \(N_{\text{area}}, LMA, N_{\text{mass}},\) and N uptake were calculated (see Table 1 for an explanation of all abbreviations used). Leaf N content per unit of leaf area \((N_{\text{area}})\) of leaves 9, 9.1, and 11 was determined as follows:

\[
N_{\text{area}} = \frac{LMA \times N_{\text{mass}}}{N_{\text{total}}} \tag{4}
\]

where \(LMA\) is the leaf mass per unit of area, and \(N_{\text{mass}}\) is the amount of leaf N per unit of leaf dry weight \([22]\).

| Symbol | Variable Name | Definition | Units |
|--------|--------------|------------|-------|
| \(A_{\text{in}}\) | Rate of photosynthesis | Net CO\(_2\) uptake per leaf area per time | \(\mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}\) |
| \(\text{IPAR}\) | PAR intercepted | PAR intercepted by the canopy | \(\mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}\) |
| \(k\) | Light extinction coefficient | Extinction coefficient for light in a stand | - |
| \(\text{LAI}\) | Leaf area index | Green leaf area/ground surface area | - |
| \(\text{LMA}\) | Leaf mass per area | Leaf dry matter/leaf area | mg mass cm\(^{-2}\) |
| \(\text{PAR}\) | Photosynthetically active radiation | Spectral range of solar radiation from 400 to 700 nm | nm |
| \(\text{PAR}_{\text{inc}}\) | PAR incoming | PAR incoming above the canopy | \(\mu\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}\) |
| \(N_{\text{area}}\) | N content per green leaf area | \(LMA/N_{\text{mass}}\) | mg N cm\(^{-2}\) |
| \(N_{\text{mass}}\) | N content per unit of mass | Amount of N/leaf dry weight | g N g\(^{-1}\) leaf |
| \(\text{NO}_3^-\) | Nitrate | Nitrate content | g m\(^{-2}\) |
| \(N_{\text{total}}\) | Total mineral nitrogen | \(\text{NO}_3^- + \text{NH}_4^+\) | g m\(^{-2}\) |
| \(\text{N uptake}\) | Nitrogen uptake | N contained in the plant | g plant\(^{-1}\) |
| \(TT\) | Thermal time | Accumulation of daily mean temperature above a base temperature | °C-day |

### 2.4. Soil Sampling

Nitrogen is present in the soil as a mixture of organic and inorganic forms \([40]\), from which nitrate (\(\text{NO}_3^-\)) is the most absorbed form of N \([41]\), and the most associated with developmental and growing processes in oilseed rape \([42,43]\). In addition, due to N fertilization, mineralization, biological fixation, and losses by volatilization, the status of this nutrient varies over time; for that reason, the time course of \(\text{NO}_3^-\) availability is a more accurate way of analyzing the effects of N supply on oilseed rape than total mineral N availability. For this purpose, soil samples were collected at four different stages between sowing and beginning of flowering (BBCH 62), and total mineral N (\(\text{NO}_3^- + \text{NH}_4^+\)) and nitrate were determined on the basis of extraction with the 0.01 M calcium chloride method \([44]\).

To obtain the dynamics of N availability and then the \(\text{NO}_3^-\) available at the moment that each leaf appeared, a three-parameter power function was fitted to the \(\text{NO}_3^-\) measured.

\[
Y = A + B \times R^{TT}, \tag{5}
\]

where \(Y\) is \(\text{NO}_3^-\) availability, \(A\) is the intercept, \(B\) determines the slope of the curve, \(R\) is the asymptotic maximum \(\text{NO}_3^-\) value, and \(TT\) is thermal time.
2.5. Statistical Analysis

Experimental data were subjected to analysis of variance using GENSTAT 17th edition [45], and differences were considered to be statistically significant at a probability level of 5%. To evaluate the effects of N supply, plant population density, and their interaction at plant level (i.e., plant height, leaf appearance, leaf area, N content per leaf, Narea, LMA, Nmass, and N uptake m−2 and per plant), a two-way ANOVA was used and, when significant effects were found, differences between high and low N level or between high and low plant population level were evaluated by means of a two-sample t-test. When leaf position was also included as a factor, evaluated variables (Yijk; Yijk being leaf appearance, leaf area, N content per leaf, Narea, LMA, or Nmass) were analyzed using the following model:

\[ Y_{ijk} = \mu + N_i + D_j + ND_{ij} + NDL_{ijk} + \varepsilon_{ijk}, \]

where \( \mu \) represents the mean of the observations, and the effects of the treatments are represented as follows: \( N_i \) is the fixed effect of N supply, \( D_j \) is the fixed effect of plant population density, \( ND_{ij} \) represents the interaction between N supply and plant population density, and \( NDL_{ijk} \) represents the effect of leaf position within N supply and population density, plus a residual \( \varepsilon_{ijk} \), which represents the random variation. Since not all the treatments produced leaf 9.1, the effect of leaf position \( (Lk) \) was not evaluated. When significant effects were found, differences among treatments were evaluated by means of the Fischer least significant difference (LSD) test, and, when only two N level or densities were compared, a t-test was performed (see Supplementary Materials). Linear and exponential models were used to assess the associations between variables, and adjusted r-square (adjusted \( r^2 \)) was used to evaluate goodness of fit of these models, which is the difference between residual and total mean squares of the total mean square. The adjustment takes into account the number of parameters in the model compared to the number of observations [45].

3. Results

3.1. Plant and Leaf Characteristics

The times of emergence (BBCH 10), rosette formation (BBCH 25), and beginning of flowering (BBCH 62) were not affected \( (p > 0.05) \) by the treatments. Plants grown under N150 were 12 cm taller than plants grown under N50, while plants grown at low plant population density (D50) were 15 cm taller than those at high density (D150) (Figure 2a). Plant height was not affected \( (p > 0.05) \) by the interaction between N supply and plant population density.

![Figure 2](image-url)

**Figure 2.** (a) Plant height in response to N supply and plant density. Vertical bars indicate the standard error of means. (b) Leaf area for each phytomer rank (includes main stem and branch leaf area) at Biologische Bundesanstalt, Bundessortenamt and Chemical industry (BBCH) 62 in response to N supply and plant population density. Horizontal bars indicate the standard error of means.
Remarkable differences in leaf area at phytomer rank in response to N supply and, to a lesser degree, to plant population density were observed (Figure 2b), while yellowing and senescence were evident from phytomer 3 downward. In all the treatments, leaves positioned on phytomers 1 and 2 had already dropped at that time. These processes of leaf senescence and dropping are associated with reduced PAR and a decreased red/far-red ratio in leaves positioned at the bottom of the plant canopy [46,47]. The area of leaves positioned in phytomer ranks 3 and 4 was slightly higher in N150-D150 than in N150-D50, while these differences were not evident at higher phytomer ranks. This means that, at early stages, when plant competition was less of a limiting factor, plants grown at higher densities expanded slightly larger leaves than at low plant population density.

A significant interaction among N supply, plant population density, and leaf position was observed in leaf appearance, individual leaf area, and N content per leaf of the evaluated leaves (Table 2). Among the three positions measured, the leaf positioned at phytomer 9 was the first to appear, followed by leaves 11 and 9.1 in second and third place, respectively. While the timing of appearance was not affected in leaf 9 by the treatments and their interaction, leaf 11 appeared earlier in treatments N50-D150 and N50-D50 than in treatments N150-D150 and N150-D50. Across treatments, the leaf area of leaves 9 and 11 was larger in N150-D50 than in treatments with low N supply (N50-D50 and N50-D150), and the leaf area of leaf 9.1 was not different from the leaf area of leaf 11 ($p > 0.05$). The total N content of leaf 9 and 11 was higher in N150-D50 than in the other treatments.

### Table 2. Time of appearance, leaf area, and N content of leaves at phytomers 9, 9.1, and 11.

| Treatment   | Leaf Position | Leaf Appearance (°C·Day) | Individual Leaf Area (cm$^2$ per Leaf) | N content Per Leaf (mg N Per Leaf) |
|-------------|---------------|--------------------------|----------------------------------------|-----------------------------------|
| N150-D50    | Leaf 11       | 539.1 d                  | 14.86 cd                               | 2.868 b                           |
|             | Leaf 9        | 473.1 a                  | 60.74 a                                | 9.916 a                           |
|             | Leaf 9.1      | 690.9 e                  | 11.83 de                               | 1.180 cd                          |
| 150-D150    | Leaf 11       | 535.2 cd                 | 9.44 def                               | 0.892 cd                          |
|             | Leaf 9        | 470.4 a                  | 19.26 c                                | 2.184 bc                          |
|             | Leaf 9.1      | 690.9 e                  | 11.83 de                               | 1.180 cd                          |
| N50-D50     | Leaf 11       | 527.6 bc                 | 6.27 ef                                | 0.756 d                           |
|             | Leaf 9        | 466.9 a                  | 26.59 b                                | 2.729 b                           |
|             | Leaf 9.1      | 623.7 b                  | 3.44 f                                 | 0.136 d                           |
| N50-D150    | Leaf 11       | 523.7 b                  | 3.44 f                                 | 0.136 d                           |
|             | Leaf 9        | 469.3 a                  | 9.19 def                               | 0.722 d                           |
|             | Leaf 9.1      | 623.7 b                  | 3.44 f                                 | 0.136 d                           |

**N supply (N)** *** *** ***
**Plant population density (D)** *** *** ***
**N × D × Leaf position** *** NS **

Different letters indicate significant differences between leaf positions in response to treatments according to ANOVA and Fischer least significant difference (LSD) test, where letter a indicates the highest value. Asterisks indicate differences between the treatments evaluated (** significant at $p < 0.01$; *** significant at $p < 0.001$; NS = not significant).

### 3.2. Leaf N Content ($N_{area}$) and Photosynthesis

There was a significant interaction effect ($p < 0.05$) between N supply and plant population density on $N_{area}$ in leaves 9, 9.1, and 11 (Figure 3a). The highest $N_{area}$ value was reached in leaves 9 and 11 (0.16 and 0.20 mg·cm$^{-2}$, respectively) grown under N150-D50, while leaf rank 9.1, the youngest leaf, recorded the lowest $N_{area}$ (0.10 mg·cm$^{-2}$). $N_{area}$ of leaves 9 and 11 grown under N150-D150, N50-D50, and N50-D150 did not differ significantly ($p > 0.05$). Light-saturated photosynthesis rate per area ($A_n$) was asymptotically related to $N_{area}$ across treatments and leaf positions (Figure 3b), whereas, at low $N_{area}$, $A_n$ increased markedly until 0.12 mg N·cm$^{-2}$ reaching a plateau at around 0.20 mg N·cm$^{-2}$. This trend in oilseed rape has also been observed but at different critical values in a wide diversity of other crops including sunflower (*Helianthus annuus* L.), kenaf (*Hibiscus cannabinus* L.), and cynara (*Cynara cardunculus* L.) [48,49].
with the value for plants grown at low N supply (N50), while the high plant population density (D150) was analyzed considering both N and plant population density as independent factors because there was an interaction between them (Figure 4b). High N supply (N150) increased leaf N mass by 65% compared with the value for plants grown at low N supply (N50), while the high plant population density (D150) showed Nmass 11% lower than the low density (D50). These last results suggest that Nmass was affected by N availability per plant.

Narea was closer associated with LMA (Figure 5a) than with Nmass (Figure 5b) and showed a linear relationship depending on N supply. The intercepts of those relationships were lower at N150 than at N50; therefore, the minimum Narea was reached with smaller LMA at N150 than at N50.

**Figure 3.** (a) Leaf N content per area (Narea) of leaves at different phytomer ranks in response to N supply and plant density. (b) Light-saturated photosynthesis (An) (linc = 1500 µmol·m−2·s−1; CO2 = 390 µmol·mol−1) in relation to Narea at 7 days before flowering (BBCH 62). Different letters indicate significant differences across treatments (p < 0.05), and vertical bars indicate the standard error of means.

### 3.3. Physiological and Structural Determinants of Narea

LMA was affected (p < 0.01) by the interaction among N supply, plant population density, and leaf position. Within a treatment, differences in LMA (Figure 4a) among the evaluated leaves were significant (p < 0.05) in N150-D50 where leaves 9 and 11 showed higher LMA than leaf 9.1, while the other treatments did not show differences among leaves.

**Figure 4.** (a) Leaf mass per unit of area (LMA) and (b) N content per unit of mass (Nmass), of leaves at different phytomer ranks in response to N supply and plant population density at 7 days before flowering (BBCH 62). Different letters indicate significant differences across treatments (p < 0.05), and vertical bars indicate the standard error of means.

Among treatments, LMA of leaf 11 was higher in treatments N150-D50 and N50-D50 than in N150-D150 and N50-D150. On the other hand, Nmass, the other physiological determinant of Narea, was analyzed considering both N and plant population density as independent factors because there was no interaction between them (Figure 4b). High N supply (N150) increased Nmass by 65% compared with the value for plants grown at low N supply (N50), while the high plant population density (D150) showed Nmass 11% lower than the low density (D50). These last results suggest that Nmass was affected by N availability per plant.

Narea was closer associated with LMA (Figure 5a) than with Nmass (Figure 5b) and showed a linear relationship depending on N supply. The intercepts of those relationships were lower at N150 than at N50; therefore, the minimum Narea was reached with smaller LMA at N150 than at N50.
N50-D150 recorded the minimum N uptake with only 0.04 g N per plant. Population density was not significant (p > 0.05) N uptake per plant with 0.29 g N per plant followed by N150-D150 and N50-D50, while N50-D150 recorded the minimum N uptake with only 0.04 g N per plant.

A positive relationship was observed between LMA and light interception by leaves located in a given layer for treatments grown at low plant population density (N150-D50 and N50-D50) irrespective of N supply (Figure 6) (p < 0.05). Thus, LMA was more sensitive to plant population density than to N supply. However, this trend was not valid for the high-density treatments (N150-D150 and N50-D150).

At increasing intraspecific competition, other environmental factors become limiting for leaf structure (i.e., red/far-red ratio, below-ground competition). To assess the efficiency of the N treatments, N uptake per square meter was evaluated (Figure 7a), and N uptake per square meter of N150 was two times higher than that of N50 (p < 0.01), while the interaction between N supply and plant population density was not significant (p > 0.05). At the plant level (Figure 7b), the interaction between N supply and plant population density was significant (p < 0.01) and treatment N150-D50 showed the highest (p < 0.05) N uptake per plant with 0.29 g N per plant followed by N150-D150 and N50-D50, while N50-D150 recorded the minimum N uptake with only 0.04 g N per plant.

Figure 5. Leaf N content per unit of area (Narea) versus (a) leaf mass per unit of area (LMA) and (b) N content per unit of mass (Nmass) at 7 days before flowering (BBCH 62). Each symbol corresponds to an individual leaf.

Figure 6. Relationship between leaf mass per unit of area (LMA) and light interception by leaves located in a given layer at 7 days before flowering (BBCH 62). Vertical bars indicate the standard error of means.
4. Discussion

4.1. Effects at Plant Level

Both assessed factors, N supply and plant population density, affected plant growth and architecture, while the times of plant emergence, rosette formation, and beginning of flowering...
were not affected by the treatments. Only the interaction among N supply, plant population density, and leaf position affected leaf appearance. Low N supply reduced plant height and leaf area, while, at low density, plants were taller but developed smaller leaves than at high density. Reductions in plant height and individual leaf area in response to low N supply corroborate previous work in *Triticum aestivum* L. [50], *Zea mays* L. [51], and *Gossypium hirsutum* L. [52]. According to the literature [53], the interaction between N supply and plant population density influences the leaf area index, which determines the proportion of light intercepted by the canopy. It has been observed that the higher impact of N supply on leaf area is at early stages (i.e., budding stage), while the effect of plant population density is at later stages (i.e., seed filling stages) [53]. As our study focused on the phenological stage of the beginning of flowering (BBCH 62), both effects resulted as an emergent property of the interaction as a consequence of the effective availability of resources per plant.

On the other hand, an increase in plant density induces competition for solar radiation and nutrients among individuals, leading to taller plants and thinner stems (not measured in the current experiment), resulting in an increase in lodging risk [54], one of the most important problems in oilseed rape. In our study and at high N rates, the tallest plants were found at low plant density (50 plants·m⁻²), and not at high plant density (150 plants·m⁻²). These results are supported by studies performed in oilseed rape in field experiments (when comparing sowing rates of 15 vs. 45 plants·m⁻²) [55] and flax (*Linum usitatissimum* L.) [56], where increasing population density decreased plant height. This behavior is the result of an intensive intraspecies competition for resources (i.e., light or nutrients), at developmental stages when the plant requirements are higher than the resources available to satisfy such requirements, as has been demonstrated in sunflower [57] and now in oilseed rape. Thus, at conventional plant densities (e.g., 50 plants·m⁻²), increasing plant population density (e.g., 150 plants·m⁻²) leads to taller plants, but there is a critical level above which further increases in plant population density will cause a reduction in plant height, due to a shortage in resources available for plant C assimilation, causing a reduction in photoassimilates available for plant growth. Variations in leaf area in response to N supply were significant (*p < 0.05*) in all the evaluated leaves, while a delay in leaf appearance was evident only in the most apical leaves evaluated (i.e., leaves 11 and 9.1).

When soil N status exceeds the plant N requirements, N available in the soil is enough to satisfy the plant N demand [8]. Thus, N is absorbed, assimilated, and distributed in the plant to maximize photosynthesis [38] by means of building tissues for photosynthesis, meristematic activity, and stems [8]. However, when plant N demand exceeds N availability, N and C are mostly allocated to the roots, and shoot/root and leaf/stem ratios are reduced [8]; thus, reductions in leaf area and delays in leaf appearance are expected. The lower N uptake per square meter (Figure 7a) and the lower N leaf/stem ratio (Figure 7b) observed in treatments with low N supply, added to the later appearance of the younger leaves (leaves 11 and 9.1), coincided with the moment when N availability most likely started to become a limiting factor (Figure 8), confirming that N availability played an important role in leaf development and expansion. Because N uptake per plant was significantly affected by the interaction between N supply and plant density (Figure 7b), it is not possible to establish the individual importance of N supply and plant population density in plant N uptake. Since detached leaves were not considered as part of the evaluation, it is possible that N uptake and N redistribution were slightly underestimated.

4.2. Effects at Leaf Level

Leaf traits such as leaf thickness, palisade depth, and mesophyll cell density are summarized in a single concept: LMA [58]. In the current study, LMA showed a significant response to the interaction among N supply, plant population density, and leaf position, showing that oilseed rape plants respond to this interaction by modifying LMA, while the leaf area of individual leaves was affected by N supply and plant population density, but not by the interaction between these two factors. These results differ from studies performed in crops like soybean [15], in which LMA is mainly modified as a result of leaf light conditions, while interspecies studies [59] showed moderate responses of LMA to nutrient shortage. The same response was observed in *Prunus persica* [17], whereas, by contrast, Jullien et al. [60] observed
in winter oilseed rape an increase in LMA grown at 200 kg·ha$^{-1}$ of N supply in comparison with a higher fertilization rate (300 kg·ha$^{-1}$). Discrepancies among these studies could be due to the fact that the effects of N and light on LMA are hidden by their interaction. In addition, there are differences in responses of LMA to N supply among plant species and genotypes [59,60]. Furthermore, assimilate allocation to different organs is a dynamic process that varies among phenological stages [8] and the moment of the season at which these phenological stages occur. This is important when comparing winter and spring oilseed rape or different varieties. For that reason, differences in LMA observed in spring oilseed rape during a specific phenological stage (BBCH 62) are not totally comparable with studies performed in winter oilseed rape at another phenological stage [60].

It is known that N supply has a higher impact on IPAR and LAI at early stages (i.e., budding stage), while density has the most significant impact during the seedling stage. Thus, the interaction between N supply and plant population density influences the IPAR and LAI during early and mid (i.e., seedling, budding and blossom) developmental stages [53]. Another determinant of $N_{\text{area}}$ is $N_{\text{mass}}$, which was affected more by N supply than by plant population density. This is supported by a global study of relationships among leaf traits, climate, and soil nutrient fertility, where $N_{\text{mass}}$ was more dependent on soil fertility than on light or other climatic factors [24], as observed in our study.

Although LMA and $N_{\text{mass}}$ are both physiological determinants of $N_{\text{area}}$, this trait was more strongly associated with LMA as demonstrated in other plant species [61]; this phenomenon reflects a trade-off between the interaction between N supply and light interception with the potential for photosynthesis per unit of area ($A_n$). The $A_n$ per unit of leaf area depends on $N_{\text{area}}$, with variations in the photosynthetic capacity for a given $N_{\text{area}}$ among species [12]. In our work, $A_n$ was strongly correlated with $N_{\text{area}}$ in an asymptotical relationship (Figure 3b), as observed in a wide diversity of other crops including sunflower (Helianthus annuus L.), kenaf (Hibiscus cannabinus L.), and cynara (Cynara cardunculus L.) [48,49], but in contrast with a linear trend in Triticum aestivum L. grown in a hydroponic system [62].

In a scenario in which N supply is limiting, N availability is not able to satisfy plant N demand, because, in oilseed rape, N deficiency impacts leaf cell and tissue structure, with consequences for senescence [63], causing reductions in photosynthesis, plant height, N uptake, N partitioned toward the leaves, $N_{\text{mass}}$, and leaf area, as observed in the current study. On the other hand, the plant aims to maximize C assimilation maximizing $A_n$, which is highly correlated with $N_{\text{area}}$. LMA was more sensitive to the interaction between N supply and plant population density than to N supply and plant population density independently.

### 4.3. Implications of the Present Results and Further Research

It is well known that the amount of N fertilizer and plant population density regulate canopy architecture and photosynthesis, affecting C accumulation and allocation between stems and reproductive organs, further influencing stem mechanical strength and final seed yield [53,64]. Although high density leads to limited growth and smaller photosynthetic area per plant, the change in LAI, IPAR, and radiation-use efficiency before flowering is the foundation of crop yield [53]. However, studies evaluating the interaction between both agronomical practices, i.e., N supply and plant density, on N distribution and leaf traits in oilseed rape or any other crop are scarce.

We show that the interaction between N supply and plant population density played a major role in altering N per unit of area ($N_{\text{area}}$), which is the main determinant of photosynthesis ($A_n$). This interaction also affects LMA, while N supply determines N content per unit leaf mass ($N_{\text{mass}}$). These results suggest that the interaction between N supply and plant population density has significant effects on both N distribution and leaf mass per area, which, together with the effects on plant height, could have important implications for light distribution and, therefore, for C assimilation at the plant level. In addition to the effects on N distribution and leaf traits, N supply and plant population density have been related to stem branching patterns in Arabidopsis, potato, and maize, which shape overall plant architecture [65–67]. We hypothesize that N availability interacting with light interception
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