**TNF-α and TGF-β Contributes in Recurrent Otorrhea of Active Mucosal Chronic Otitis Media**

Dewi Pratiwi1,*, Marisa Rizqiana1, Adisetya Wicaksono1, Defitaria Permatasari1, Ratna Dwi Restuti2, Tri Nugraha Susilawati3, Sutarno4

1Department of Otorhinolaryngology Head and Neck Surgery, Faculty of Medicine, Universitas Sebelas Maret – Moewardi Hospital, Jl. Kolonel Sutarto No.132, Surakarta, Indonesia
2Department of Otorhinolaryngology Head and Neck Surgery, Faculty of Medicine, Universitas Indonesia – Cipto Mangunkusumo Hospital, Jl. Diponegoro No.71, Jakarta, Indonesia
3Department of Microbiology, Faculty of Medicine, Universitas Sebelas Maret, Jl. Ir. Sutami No.36, Surakarta, Indonesia
4Faculty of Mathematics and Natural Sciences, Universitas Sebelas Maret, Jl. Ir. Sutami No.36, Surakarta, Indonesia

*Corresponding author. E-mail: pratiwidewi81@staff.uns.ac.id

Received date: Nov 17, 2021; Revised date: Jan 13, 2022; Accepted date: Jan 17, 2022

**Abstract**

**BACKGROUND:** Active mucosal chronic otitis media (COM) is prevalent in lower-income countries and is associated with recurrent episodes of otorrhea due to chronic inflammation of the middle ear. Cytokines, which are well-known for their effects on the immune system, play an important role in the inflammatory response and tissue remodeling. The specific contributions of proinflammatory and immunoregulatory cytokines in the pathophysiology of active mucosal COM remain unclear. This study aimed to compare the levels of serum tumor necrosis factor (TNF)-α and transforming growth factor (TGF)-β in patients with active mucosal COM vs. healthy subjects.

**METHODS:** Total 20 subjects with active mucosal COM and 20 healthy subjects participated in this study. The levels of serum TNF-α and TGF-β were measured using enzyme-linked immunosorbent assay (ELISA).

**RESULTS:** The average level of serum TNF-α in subjects with active mucosal COM was significantly higher than the healthy subjects (46.37±41.76 pg/mL vs. 15.02±7.16 pg/mL; \(p=0.004\)). In contrast, the average level of serum TGF-β in subjects with active mucosal COM was lower compared to the healthy subjects, although the difference is not statistically significant (9.96±3.2 ng/mL vs. 11.78±8.48 ng/mL; \(p=0.552\)). Further analysis showed that in subjects with active mucosal COM, the levels of serum TNF-α had a medium positive correlation with the level of TGF-β (\(r=0.525\); \(p=0.018\)).

**CONCLUSION:** TNF-α and TGF-β, which are proinflammatory and immunoregulatory cytokines, may contribute to the pathogenesis of recurrent episodes of otorrhea in an active mucosal COM.

**KEYWORDS:** tumor necrosis factor-α, transforming growth factor-β, active mucosal chronic otitis media

**Indones Biomed J. 2022; 14(1): 59-65**

---

**Introduction**

Active mucosal chronic otitis media (COM) is characterized by intermittent or constant discharge of pus from the ear through the tympanic membrane perforation (otorrhea). Ear discharge is notoriously difficult to treat, prone to recur, and even capable of causing death in some cases.(1) The World Health Organization (WHO) considers active mucosal COM to as neglected tropical disease. It affects 65–330 million individuals globally, mostly in lower-income nations. High-income nations have a modest prevalence of active mucosal COM (<1%), but low and middle-income countries have a much higher frequency (>2%).(1-3) The prevalence of...
active mucosal COM in Indonesia is 3.9% of the general population. There is evidence that active mucosal COM is more common in children, but other research shows that it is more common in adults.(4,5)

Many disorders, including active mucosal COM, have immunological and inflammatory responses mediated by a group of glycoproteins called cytokines. The inflammatory response and tissue remodeling in the middle ear mucosa are both mediated by cytokines. Cytokines can be released from numerous cell types found within the middle ear cleft, including all types of epithelial, endothelial, and immune cells.(6) It is possible to utilize these cells as biomarkers to detect or track disease progression, and they may also be employed as treatment-specific criteria in the clinic.(7) Middle ear mucosa contains a variety of cytokines, including proinflammatory cytokines such as tumor necrosis factor (TNF)-α, interleukin (IL)-1β, interferon (IFN)-γ and IL-6; immunoregulatory cytokines such as IL-2, IL-4, IL-5, IL-10 and transforming growth factor (TGF)-β and granulocyte-macrophage colony-stimulating factor (GM-CSF); and other mediators such as receptor activator of nuclear factor kappa-B ligand (RANKL).(8) However, the presence and functions of proinflammatory and immunoregulatory cytokines in the pathogenesis of active mucosal COM remain unclear.

TNF-α, IL-6, IL-1β, IFN-γ mRNA and protein levels were higher in the middle ear mucosa of patients with active mucosal COM than in healthy persons.(9) Tissue damage and the shift from acute to chronic otitis media are also possible outcomes of increased levels of these proinflammatory cytokines.(5) TNF-α is one of the most significant inflammatory mediators in otitis media and was detected in human middle ear effusion. Animal models of otitis media induced by S. pneumoniae and lipopolysaccharide (LPS) have demonstrated elevated levels of TNF-α in the early stage of otitis media. TNF-α overexpression in otitis media increases the chance of recurrence, longer courses, and chronicity of otitis media. Using TNF-α inhibitors, such as TNF-α -binding protein or inhibitor of TNF-α -soluble receptor, is recommended in clinical practice to prevent chronicity and recurrence of otitis media.(10)

TGF-β is an immunoregulatory cytokine with proinflammatory and pro-fibrotic effects. It can control the initiation and regression of inflammation by regulating chemotaxis and activity of inflammatory cells, such as macrophages and granulocytes. In the event of tissue damage, TGF-β can help to promote matrix deposition and tissue healing. It can also cause abnormal collagen deposition and fibrosis when present in inflammatory settings.(11) Consequently, fibroblast proliferation and middle-ear-cleft thickening are two side effects. In the chronic stage of otitis media, fibroblast proliferation continues, and extracellular matrix (ECM) proteins are continually deposited in the middle ear submucosa layer, eventually leading to fibrosis. In the acute stage of otitis media, they contribute to the production of granulation tissue, which continues and intensifies during the chronic stage. TGF-β expression is high in secretory otitis media, according to a prior study.(12,13)

The etiology of active mucosal COM included proinflammatory and immunoregulatory cytokines, although their presence and function are as yet unclear. This study aimed to compare the levels of serum TNF-α and TGF-β in patients with active mucosal COM vs. healthy subjects and describe the microorganisms in the ear discharge of patients with active mucosal COM.

## Methods

### Study Design and Subjects

A cross-sectional study was conducted from March to May 2021 at the Otorhinolaryngology Clinic, Dr. Moewardi General Hospital, and its affiliated institutions in Surakarta, Indonesia. A total of 40 adult subjects (20 subjects with active mucosal COM and 20 healthy subjects) were recruited into this study.

Subjects were diagnosed with active mucosal COM if they had episodes of recurrent otorrrhea within 3 months prior to recruitment. Subjects were excluded from the study if they had medical and/or surgical procedures or illnesses, such as infection and/or inflammation either in an acute or chronic form that may affect the outcome of the study. All subjects signed informed consent and the study protocol was approved by the Research Ethics Committee at Dr. Moewardi General Hospital (No. 372/IV/HREC/2021).

Recurrent otorrrhea and hearing loss were the two most prevalent complaints that had been reported by all active mucosal COM subjects. Clinical presentation and otoscopic examination were employed to diagnose active mucosal COM. The decibel level of hearing (dB HL) was assessed using an audiological test. A diagnostic audiometer was used to measure the hearing threshold in each ear at frequencies of 1 kHz, 2 kHz, and 4 kHz.

### Collection and Culture of Bacteria from Exudate

Middle ear exudate was collected under the direction of an otomicroscope using sterile small cotton-tip swabs (Copan
Venturi Transystem 108C use Amies) to avoid contamination by microorganisms from the external auditory canal. Swabs of purulent exudate were cultured overnight at 37°C on blood and MacConkey agar plates. Routine procedures for bacterial organism identification were used to identify the microorganisms.

**Collection of Blood Serum**

Blood serum samples were collected from the venous blood of all patients and healthy subjects into SST containers. The fresh blood was allowed to stand at 25°C for 30 min to clot, the blood then centrifuged at 3500 rpm for 15 min. The clarified upper light-yellow liquid layer is the serum. It was preserved on -80°C and moved into -20°C a day before detection.

**Enzyme-linked Immunosorbent Assay (ELISA)**

The levels of serum TNF-α and TGF-β were measured using ELISA method. The Elabscience Human TNF-α and TGF-β ELISA Kit (Elabscience, Hubei, China) utilizes an antibody specific for human TNF-α and TGF-β coated on a 96-well plate. Recombinant human TNF-α and TGF-β dilutions were used to produce the standard curves. The wavelength at which absorbance may be detected is 450 nm. The overall procedures were conducted in the Biomedical Laboratory, Faculty of Medicine, Universitas Sebelas Maret.

**Statistical Analysis**

Data were analyzed using Statistical Package for the Social Sciences (SPSS) Statistic version 25 (IBM Corporation, Armonk, NY, USA). Mean±standard deviation of cytokine levels were used in calculations. The significance level for the Kolmogorov–Smirnov and Shapiro–Wilk normality tests was <0.05. The Mann–Whitney test was used to assess if there were significant differences in the blood levels of TNF-α and TGF-β between the active-mucosal-COM and healthy subjects. TNF-α and TGF-β levels were compared between the active-mucosal-COM and healthy subjects using Pearson rank correlation. Statistical significance was defined as a $p$-value of 0.05.

### Results

#### Characteristics of Subjects

Table 1 showed the characteristics of subjects of included in this study. Participants’s ages ranged from 19 to 64 years old and the majority of them were in their productive ages; i.e., 36-45 years old (active mucosal COM subjects) and 26-35 years old (healthy subjects). Most subjects were female (55%) and those with active mucosal COM often had a mild degree of hearing loss (30%). In all cases, the frequency of otorrhea was more than 6 times annually.

**Bacterial Pathogen Detected by Culture**

Eighteen samples that were collected from subjects with active mucosal COM yielded positive cultures on blood and MacConkey agar plates, with *Pseudomonas aeruginosa* being the most prevalent organism (90%). The other two samples yielded yeast and these were considered as contaminants originated from skin commensals of the external auditory canal (Table 2).

**Cytokine Levels in Relation to Bacterial Pathogens**

The levels of serum TNF-α and TGF-β were measured and analysed further in regards to their relationship with bacterial pathogens (Table 3). The average level of serum TNF-α was higher in samples yielding gram-positive bacteria compared to the infected by gram-negative bacteria, although the difference was not statistically significant. However, the average level of serum TGF-β was equal among two groups.
Serum Cytokines Level
Serum TNF-α and TGF-β were detected in all serum samples. The average level of serum TNF-α was significantly higher in subjects with active mucosal COM compare to the healthy subjects ($p=0.004, \text{95\%CI: 19.96-41.42}$). In contrast, the average level of serum TGF-β level in subjects with active mucosal COM subjects was lower than the healthy subjects, although the difference was not statistically significant (Table 4).

Results of Pearson Correlation Test
We performed Pearson’s rank correlation test to assess the correlation between the levels of a proinflammatory cytokines (i.e., TNF-α) and an immunoregulatory cytokine (i.e., TGF-β) in the serum samples. Figure 1 demonstrated that the level of TNF-α had a medium positive correlation with the level of TGF-β in subjects with active mucosal COM subjects ($r=0.525; p=0.018$), whilst they had a weak negative correlation in healthy subjects ($r=-0.341; p=0.141$).

Discussion
COM is the most frequent inflammatory diseases of the middle ear among children and adults and is especially problematic in lower-income nations. The condition causes chronic inflammation of the middle ear cleft and the clinical manifestations include otorrhea, permanent perforation, and hearing loss. COM can be divided into active, inactive or healed COM depending on the clinical disease condition. Based on the histologic features of the middle ear mucosa and the edges of the perforated tympanic membrane, COM is divided into mucosal and squamous COM. Active mucosal COM refers to a permanent defect of the pars tensa with an inflamed middle ear mucosa that produces ear discharge (otorrhea). (14,15)

Our study showed that there is no sex preference for the active mucosal COM with male-to-female ratio was almost equal (45\% vs. 55\%). This is similar to a previous study in Tanzania that found 54.4\% of the study subject with active mucosal COM were male. (16) Another study found that males were more likely to have active mucosal COM (53.5\% vs. 46.5\%). (17) This is related to male’s tendency of working outside the house making them susceptible to infection. (18)

Otitis media is typically thought to be a disease of children and the proportion of active mucosal COM in older subjects tend to decline. Children are more susceptible to otitis media than adults because their eustachian tubes are shorter, more horizontal, and more susceptible to obstruction by enlarge adenoids. Furthermore, in young children, viral infections and allergies are frequent, and both can induce eustachian tube inflammation. It can, however, affect adults and the elderly. (10,19) According to the WHO, the highest prevalence of active mucosal COM occurs in the first year of life (15.40 per thousand), with the lowest incidence

### Table 2. Bacterial culture from subjects with active mucosal COM subjects (n=18).

| Microorganisms              | n (\%) | TNF-α (pg/mL) | TGF-β (ng/mL) |
|-----------------------------|--------|---------------|---------------|
| Gram-positive               |        |               |               |
| Corynebacterium striatum    | 6 (33.3)| 58.63±42.03   | 9.76±2.08     |
| Staphylococcus haemolyticus | 1 (5.5)| 33.87         | 7.79          |
| Staphylococcus xylosus      | 1 (5.5)| 54.88         | 13.44         |
| Gram-negative               |        |               |               |
| Pseudomonas aeruginosa      | 8 (44.4)| 40.89±43.70   | 10.11±4.67    |
| Acinetobacter spp.          | 2 (11.1)| 16.75±7.95    | 9.94±1.12     |

### Table 3. Inflammatory mediators in middle ear culture positive for gram-positive and -negative bacteria.

| Serum Cytokines | Gram-positive (n=8) | Gram-negative (n=10) | $p$-value |
|-----------------|---------------------|----------------------|-----------|
| TNF-α (pg/mL)   | 55.06±39.87         | 36.06±40.00          | 0.33      |
| TGF-β (ng/mL)   | 9.97±2.36           | 10.08±4.13           | 0.94      |
occurring beyond 65 years of age (2.51 per thousand). This is confirmed by our present study that found the proportion of active mucosal COM in the 56-65 age group was being the lowest incident (10%).

Active mucosal COM is often followed by hearing loss. We found a mild degree of hearing loss (30%) to profound hearing loss (5%) among the cases. Suppurative chronic otitis media can induce both conductive (CHL) and sensorineural hearing loss (SNHL). A pus-filled middle ear is the characteristic of chronic suppurative otitis media and can interfere with sound transmission to the inner ear. The use of topical antibiotics for an extended period or the absorption of chemicals or toxins from active mucosal COM through the round window membrane might also cause sensorineural hearing loss.

Our study found P. aeruginosa as the most prevalent pathogen responsible for the development of active mucosa COM which is similar to the results of previous studies. Other studies from China reported Staphylococcus aureus as the most common pathogen in active mucosal COM, followed by P. aeruginosa. Different patient populations, regional variations, and antibiotic use before culture might all account for the discrepancies across studies. As fungi live well in humid ears, fungal infections of the middle ear have been found in the cultures of patients with active mucosal COM and who had used topical or systemic antibiotics for an extended period.

Gram-negative bacteria accounted for 55.5% of the discharged ear isolates, and gram-positive bacteria accounted for the remaining 40.5%. Another study had similar findings, which found 59.6% gram-negative bacteria in their isolates. The chronic nature of infection, in which gram-negative bacteria from external sources gain access to the auditory canal, suppressing the gram-positive normal flora and eventually becoming dominant, could explain the higher prevalence.

Previous studies showed that leukocytes produce different patterns of proinflammatory cytokines and chemokines when stimulated with gram-positive or gram-negative bacteria. Monocyte/macrophage cytokine release and TLR expression are both triggered by bacteria. TNF-α production is higher in gram-positive bacteria than in gram-negative bacteria. TNF-α, IFN-γ, and IL-12 are secreted as a result, whereas only IL-6 and IL-10 are secreted in the presence of gram-negative bacteria. Extravasation

### Table 4. Serum cytokines level in active-mucosal-COM and healthy subjects

| Serum Cytokines | Active Mucosal COM Subjects (n=20) | Healthy Subjects (n=20) | p-value |
|-----------------|----------------------------------|------------------------|---------|
| TNF-α (pg/ml)   | 46.37±41.76                     | 15.02±7.16             | 0.004   |
| TGF-β (ng/ml)   | 9.963±3.2                       | 11.78±8.48             | 0.552   |

![Figure 1. Pearson correlation between TNF-α and TGF-β in the patients with active mucosal COM (A) and healthy subjects (B).](image-url)
of polymorphonuclear leukocytes into infected tissue is dependent on TNF-α, which also contributes to prostaglandin and cytokine production, neutrophil activation, and eosinophil and macrophage activation; thus, TNF-α is essential. Patients with active mucosal COM had higher levels of the cytokines TNF-α and TGF-β, which were then linked to the kind of bacteria found by culture in the middle ear. We found that the levels of serum TNF-α were greater in cases infected with gram-positive bacteria. This result is similar to a previous study which also demonstrated that gram-positive bacteria is related to the higher levels of TNF-α in the serum.(30)

TNF-α is a cytokine with the ability to promote bone osteolysis. Our study showed that the levels of serum TNF-α levels in patients with active mucosal COM were 3.08-fold greater than in healthy people, while other study found that TNF-α levels in COM with cholesteatome were 3.8-fold greater than in normal skin. This may imply that the increasing level of proinflammatory cytokines such as TNF-α in active mucosal COM causes chronic inflammation to exacerbate, resulting in recurrent otorrhea.(8,31)

TGF-β is an immunoregulatory cytokine that plays an important role in the control of inflammatory processes as well as tissue repair. TGF-β is thought to be responsible for granulation tissue formation and ossicular fixation in the middle ear.(12) A high expression of TGF-β in the serum of patients with active mucosal COM showed that TGF-β has an important role in the chronic inflammatory course of active mucosal COM. As TGF-β can permeate into middle ear mucosa, it causes higher expression in middle ear discharge in patients with active mucosal COM and is not only produced by the middle ear mucosa. A previous study had suggested that perturbation of the TGF-β signaling pathways can lead to chronic otitis media.(32) Prolonged TGF-β activation has been linked to the development of chronic inflammation and fibrosis in other parts of the body.(33)

Our study showed no statistically significant difference in the expression of TGF-β in between active-mucosal-COM and healthy subjects. However, TGF-β might limit the production of mucin elicited by bacteria by downregulating MUC5AC in the middle ear cavity, which may decrease the host’s innate immunity to a pathogen, delay the removal of microorganisms, and eventually cause the condition to become chronic.(34)

Nonetheless, our study found a positive correlation between the levels of serum TNF-α and TGF-β; i.e., the higher the level of TNF-α, the higher the level of TGF-β. It demonstrates that these two types of cytokines contribute to the middle ear inflammatory responses, particularly otorrhea in patients with active mucosal COM. Further research into the presence and interplay of proinflammatory and immunoregulatory cytokines in active mucosal COM is needed to offer a more comprehensive picture of the inflammatory process in the middle ear.

Further research is required to determine the association between middle ear secretion TNF-α and TGF-β levels in conjunction with active mucosal COM to explain the involvement of cytokines in the pathophysiology of active mucosal COM. Our study is limited to the absence of measurements of cytokine levels in middle ear discharge. Furthermore, due to the lack of pediatric patients, a direct comparison of differences between pediatric and adult patients was impossible.

### Conclusion

The level of TNF-α serum was significantly higher in patients with active mucosal COM than in healthy subjects, but the level of TGF-β serum was not. The level of TNF-α had a medium positive correlation with the level of TGF-β in subjects with active mucosal COM. These two cytokines may contribute to the pathogenesis of recurrent episodes of otorrhea in an active mucosal otitis media. Further study is needed to explain the roles of other cytokines in the middle ear inflammatory responses.

### Acknowledgments

The Faculty of Medicine at Universitas Sebelas Maret in Surakarta, Indonesia fully financed this initiative. Thanks to Ahmad Yasin, our research assistant, for his help with the manuscript's editing.

### Authors Contribution

DP, RDR, TNS, and S were involved in the planning and supervision of the work, as well as performing the measurements. MR, AW, and DP processed the experimental data, performed the analysis, drafted the manuscript, and designed the figures. DP performed the computations and statistical analysis and also supported MR and AW in interpreting the results and writing the manuscript. Each author discussed the findings and provided feedback on the manuscript.
References

1. Orji F. A survey of the burden of management of chronic suppurrative otitis media in a developing Country. Ann Med Health Sci Res. 2013; 3(4): 598–601.

2. Li MG, Hotez PJ, Vrbacek JT, Donovan DT. Is Chronic Suppurative Otitis Media a Neglected Tropical Disease? PLoS Negl Trop Dis. 2015; 9(3): e0003485. doi: 10.1371/journal.pntd.0003485.

3. World Health Organization. Chronic Suppurative Otitis Media: Burden of Illness and Management Options. Geneva: World Health Organization; 2004.

4. Mutthah S, Mackenzie I, Faragher B, Brabin B. Prevalence of chronic suppurrative otitis media (CSOM) and associated hearing impairment among school-aged children in Yemen. Oman Med J. 2015; 30(5): 358–65.

5. Mittal R, Lisi CV, Gerring R, Mittal J, Narasimhan G, et al. Current concepts in the pathogenesis and treatment of chronic suppurrative otitis media. J Med Microbiol. 2015; 64(10): 1103–16.

6. Massa HM, Lim DJ, Kurono Y, Cripps AW. Middle ear and eustachian tube mucosal immunology. Mucosal Immunol Fourth Ed. 2015; 2(2): 1923–42.

7. Kany S, Vollrath JT, Relja B. Cytokines in inflammatory disease. Int J Mol Sci. 2019; 20(23): 6008. doi: 10.3390/ijms20236008.

8. Kuczkwoski J, Sakowicz-Burkwicz M, Ilyka-Swieszewska E, Mikaszewski B, Pawelczyk T. Expression of tumor necrosis factor-α, interleukin-1α, interleukin-6 and interleukin-10 in chronic otitis media with bone osteolysis. ORL J Otorhinolaryngol Relat Spec. 2011; 73(2): 93–9.

9. Si Y, Zhang ZG, Chen SJ, Zheng YQ, Chen Y Bin, Liu Y, et al. Attenuated TLRs in middle ear mucosa contributes to susceptibility of chronic suppurrative otitis media. Hum Immunol. 2014; 75(8): 771–6.

10. Schilder AGM, Chonnaintree T, Cripps AW, Rosenfeld RM, Casselbrant ML, Haggard MP, et al. Otitis media. Nat Rev Dis Primer. 2016; 2(1): 16063. doi: 10.1038/nrdp.2016.63.

11. Sulaeman A, Amiruddin AR, Lawrence GS. Metabolic syndrome (MetS) and nonalcoholic steatohepatitis (NASH): study of biochemical markers free fatty acid (FFA), total antioxidant status (TAOS), adiponectin, transforming growth factor (TGF-beta1), in occurrence of NASH. Indones Biomed J. 2009; 1(1): 40–4.

12. Qiu J, Wang Y, Guo W, Xu L, Mou Y, Cui L, et al. Role of TGF-β1-mediated epithelial-mesenchymal transition in the pathogenesis of tympanosclerosis. Exp Ther Med. 2020; 20(6): 1–1.

13. Wang B, Cheng Y, Xu M. Characterization of the t-cell subpopulations in the granulation tissues of chronic suppurrative otitis media. Biomed Rep. 2016; 4(6): 694–8.

14. Gleeson M, Browning GG, Burton MJ, Clarke R, John H, Jones NS, et al. Scott-Brown’s Otorhinolaryngology Head and Neck Surgery. 7th ed. London: Hodder Arnold; 2008.

15. Ibekwe TS, Nwaorgu OGB. Classification and management challenges of otitis media in a resource-poor country. Niger J Clin Pract. 2011; 14(3): 262–9.

16. Abraham ZS, Nutanagzi D, Kahinga AA, Mapondella KB, Massawe ER, Nkwiwi EI, et al. Prevalence and etiological agents for chronic suppurrative otitis media in a tertiary hospital in Tanzania. BMC Res Notes. 2019; 12(1): 429. doi: 10.1186/s13104-019-4483-x.

17. Metri Basavaraj C, Jyothi P. Chronic suppurrative otitis media (CSOM): Etiological agents and antibiotic sensitivity pattern of the isolates. J Med Bangladesh. 2015; 16(2): 78–82.

18. Poluan FH, Utomo BSR, Dharmayanti J. Profile benign type of chronic suppurrative otitis media in general hospital of the christian university of Indonesia. Int J Res Granthaalayah. 2021; 9(4): 229–39.

19. Al-Sadeeq H, Algarni Z, Alobaid A, Aloyaid A, Alotaibi M, Al-Qwizani A, et al. Otitis media among elderly: incidence, complication and prevention. Int J Community Med Public Health. 2018; 5(3): 839–41.

20. Aahrus L, Tams K, Kvestad E, Engdahl B. Childhood otitis media: a cohort study with 30-year follow-up of hearing (The HUNT Study). Ear Hear. 2015; 36(3): 302–8.

21. Islam MR, Abdullah M, Kabir AL, Islam SS, Rashid MHO. Hearing loss in chronic suppurrative otitis media (CSOM). Bangladesh J Otorhinolaryngol. 2020; 23(1): 59–66.

22. Hiremath B, Mudhol RS, Vagrali MA. Bacteriological profile and antimicrobial susceptibility pattern in chronic suppurrative otitis media: a 1-year cross-sectional study. Indian J Otolaryngol Head Neck Surg. 2019; 71(S2): 1221–6.

23. Wahyono DJ, Darmawan AB, Alfason L, Simbolon R, Wiiyantii SPM, Paramaiswari WT, et al. Staphylococcus aureus and pseudomonas aeruginosa in tubotympanic chronic suppurrative otitis media patients in Purwokerto, Indonesia. Indones Biomed J. 2020; 12(4): 340–8.

24. Xu J, Du Q, Shu Y, Ji J, Dai C. Bacteriological profile of chronic suppurrative otitis media and antibiotic susceptibility in a tertiary care hospital in Shanghai, China. Ear Nose Throat J. 2021; 100(9): NP391–6.

25. Juyal D, Negi V, Sharma M, Adekhandi S, Prakash R, Sharma N. Significance of fungal flora in chronic suppurrative otitis media. Ann Trop Med Public Health. 2014; 7(2): 120–3.

26. Chirwa M, Mulwafu W, Aswani J, Masinde P, Mkakosya R, Soko D. Microbiology of chronic suppurrative otitis media at Queen Elizabeth Central Hospital, Blantyre, Malawi: A cross-sectional descriptive study. Malawi Med J. 1970; 27(4): 120–4.

27. Chandra Sahu M, Swain SK. Surveillance of antibiotic sensitivity pattern in chronic suppurrative otitis media of an Indian teaching hospital. World J Otorhinolaryngol - Head Neck Surg. 2019; 5(2): 88–94.

28. Khutan Mstr, Alam KhMdF, Naznin M, Salam MdA. Microbiology of chronic suppurrative otitis media: an update from a tertiary care hospital in Bangladesh. Pak J Med Sci. 2021; 37(3): 821–6.

29. Kaur R, Casey J, Pichichero M. Cytokine, chemokine, and toll-like receptor expression in middle ear fluids of children with acute otitis media: innate immune response in children with AOM. Laryngoscope. 2015; 125(1): E39–44.

30. Lee HY, Chung JH, Lee SK, Byun JY, Kim Y Il, Yeo SG. Toll-like receptors, cytokines & nitric oxide synthase in patients with otitis media with effusion. Indian J Med Res. 2013; 138(4): 523–30.

31. Rout MR, Mohanty D, Vijaylaxmi Y, Kamalebsh B, Chakradhar M. Prevalence of cholestetoma in chronic suppurrative otitis media with central perforation. Indian J Otol. 202; 18(1): 7–10.

32. Tateossian H, Morse S, Simon MM, Dean CH, Brown SDM. Interactions between the otitis media gene, Fbxo11, And p53 in the mouse embryonic lung. DMM Dis Models Mech. 2015; 8(12): 1531–42.

33. Bhutta MF, Thornton RB, Kirkham LAS, Kerschner JE, Cheeseman MT. Understanding the aetiology and resolution of chronic otitis media from animal and human studies. DMM Dis Models Mech. 2018; 5(3): 839–41.

34. Lee SU, Kim MO, Kang MJ, Oh ES, Ro H, Lee RW, et al. Transforming growth factor β inhibits munc5ac expression by smad3/3/4 complex formation and nf-κb deacytlation at k310 in nci-h292 cells. Mol Cells. 2021; 44(1): 38–49.