کارگاه‌های آموزشی مرکز اطلاعات علمی

مقاله نویسی علوم انسانی

اصول تنظیم قراردادها

آموزش مهارت های کاربردی در تدوین و چاپ مقاله
Anti-Aging Effects of Some Selected Iranian Folk Medicinal Herbs-Biochemical Evidences

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Introduction

Aging as a complex of natural circumstance is exhibited by an augmentation in the chance of illness and finally death. Although some theories have been proposed as the mechanisms of aging but the one relating aging and cellular oxidative stress have received more supports. Therefore, it can be said that reduced sex hormones and augmented quantity of oxidative stress parameters or inflammatory cytokines are main biochemical manifestations of aging (1-2). In accord with this theory, the production of reactive oxygen species (ROS) and/or free radicals can injure cells and tissues paralleled by malfunction of many systems. The eventual consequence of these actions is aging and finally premature cell death (3). During aging process, various pro-inflammatory molecules are generated to strengthen inflammation cascade associated with different age-related pathologies (4).

One of the problems in testing anti-aging compounds is lack of suitable animal models. Although several models have been used so far but among them, typical mouse D-galactose-induced model of aging is the best one that gives closer results to clinical studies. D-galactose is a sugar that at higher levels converts to aldose and hydroperoxide during the catalysis of galactose oxidase, culminated in the generation of free radicals (6). These modifications are substantially similar to the normal aging process demonstrated as neurological deterioration, diminished activity of antioxidant enzymes, and miserable immune responses (7-8).

Many scientists and pharmaceutical companies try to develop a drug to reduce speed of human aging
but no effective drug has been discovered yet. In the last decade the importance of folk medicine and herbal medicines have been revisited that resulted in developing many effective drugs for many human diseases. For instance, in the recent years, efficacy of herbal medicines in diseases like inflammatory bowel diseases (9-10), obesity (11), diabetes (12), pancreatitis (13), osteoporosis (14), hyperlipidemia (15), and so on has been proved. Our recent systematic review specified anti-aging herbs and their characteristics in different clinical or experimental models (16). Most of anti-aging herbs have antioxidant components and reduces free radicals which are by-product of abnormal body metabolism in the elderly.

We recently proved anti-aging potential of naturally-based drugs like IMOD and Angipars which have strong antioxidant power (2). On the basis of our systematic review, among various species we could select nine herbs with the strongest antioxidant effects such as *Z. officinale*, *G. glabra*, *R. officinalis*, *P. harmala*, *A. vera*, *S. hortensis*, *T. scordium*, *H. perforatum* and *S. marianum* to test in D-galactose-induced model of mouse aging.

**Materials and Methods**

**Chemicals**

Thiobarbituric acid (TBA), trichloroacetic acid (TCA), n-butanol, hexadecyltrimethyl ammonium bromide (HETAB), tri (2-pyridyl)-s-triazine (TPTZ), HCl, malondialdehyde (MDA), ferric chloride (FeCl3-6H2O), D-galactose, and vitamin E (Trolox) were purchased from Merck (Germany). Rat specific tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β), interleukine-6 (IL-6), NF-kappa B (NF-kb) ELISA kits were purchased from BenderMed Systems (Austria). Testosterone and dehydroepiandrosterone ELISA kits were purchased from Dia Metra (Italy).

**Preparation of herbs, extraction, and lyophilization**

Herbs were provided from the Research Institute of Medicinal Plants Karaj during June 2009 and were air-dried at room temperature. Samples were authenticated by a botanist (Y. Ajani), and voucher specimens were preserved in the central herbarium of medicinal plants (RIMP). The scientific names and tested parts of the herbal materials are detailed in Table 1. The dried plants powder (40 g) was extracted using percolation method by methanol at room temperature. Solvents were completely removed by drying under reduced pressure at 40°C in a rotary evaporator. The samples were stored at 4°C until use. Specifically, the *A. vera* leaves (1000 g) were washed in a suitable bactericide (chlorhexidine). The files were grounded to a liquid, and the pulp was removed by filtering. The resultant gel was then freeze dried.

**Animals**

Male BALB/c mice (12 weeks old, 18–22 g) were provided from Tehran University of Medical Sciences (TUMS) animal house. The animals were housed in standard polypropylene cages with wired-net top in a controlled room (temperature 23±1°C, humidity 55±10%, 12 hr light–dark cycle) and were allowed free access to standard laboratory pellet diet and water during the experiments. All ethical issues on the use of animals were carefully considered and the study protocol was approved by TUMS review board with code number of 90-03-33-15668.

**Experimental design**

Before starting the main study, a pilot was designed to set up aging model and to get proper doses of herbal materials. In the main study, 120 mice were randomly divided into 12 groups, each consisting of 10 animals. D-galactose was dissolved in a measured quantity of mice drinking water. D-galactose was given to 11 out of 12 groups of animals at 500 mg/kg D-galactose per 1 ml drinking water for 6 weeks by gavage (2, 17). The 12th group of animals was the sham group which was not given D-galactose. After 2 weeks, the 11 groups which had been given D-galactose were randomly divided into aging control group (500 mg/kg D-galactose per 1 ml drinking water, for 6 weeks), positive control group (500 mg/kg D-galactose per 1 ml drinking water plus vitamin E 200 mg/kg/day by gavage for 4 weeks) and herb-treated groups including 9 groups that each received 500 mg/kg D-galactose per 1 ml drinking water plus vitamin E 200 mg/kg/day by gavage for 4 weeks) and herb-treated groups including 9 groups that each received 500 mg/kg D-galactose per 1 ml drinking water plus vitamin E 200 mg/kg/day by gavage for 4 weeks) and herb-treated groups including 9 groups that each received 500 mg/kg D-galactose per 1 ml drinking water plus vitamin E 200 mg/kg/day by gavage for 4 weeks) and herb-treated groups including 9 groups that each received 500 mg/kg D-galactose per 1 ml drinking water plus vitamin E 200 mg/kg/day by gavage for 4 weeks).

**Table 1. The scientific names and tested parts of the plant materials**

| Scientific name          | Tested parts | Extraction yield (mg/g) | Used Dose (mg/kg) | References |
|--------------------------|--------------|-------------------------|-------------------|------------|
| Zingiber officinalis     | Rhizome      | 140.57                  | 250               | 18         |
| Glycyrrhiza glabra       | Root         | 129.52                  | 150               | 19         |
| Rosmarinus officinalis   | Aerial parts | 236.51                  | 300               | 20         |
| Peganum harmala          | Seed         | 169.25                  | 50                | 21         |
| Aloe vera                | Gel          | 4.87                    | 150               | 22         |
| Satureja hortensis       | Aerial parts | 13.4                    | 200               | 23         |
| Teucrium scordium        | Aerial parts | 205                     | 200               | 24         |
| Hypericum perforatum     | Aerial parts | 100.58                  | 135               | 25         |
| Silybum marianum         | Seed         | 123.49                  | 150               | 26         |

*Iran J Basic Med Sci, Vol. 16, No. 11, Nov 2013*
mg/kg/day) and S. marianum (150 mg/kg/day), respectively by gavage for 4 weeks (18-26).

Twenty-four hours after the last treatment, blood samples were taken of each animal under anesthesia through the tail vein. Serum samples were obtained by centrifuging the whole blood at 1000 x g at 4°C for 10 min and the supernatants were transferred into several microtubes for separate biochemical assays and maintained at -80°C until the analyses were performed. Biochemical markers including TNF-α, IL-β, IL-6, NF-kb, ferric reducing reducing (TAP), lipid peroxidation (LPO) and male sex hormones including testosterone and dehydroepiandrosterone-sulfate (DHEA-S) were measured in the serum.

Measurement of LPO
LPO was measured by the reaction of thiobarbituric acid (TBA) with lipid peroxides. Samples were mixed with TCA (20%) and the precipitate was dispersed in H₂SO₄ (0.05 M). After addition of TBA (0.2% in sodium sulfate), the sample was heated for 30 min in a boiling water bath. Then, TBA reactive substances (TBARS) as LPO marker adducts were extracted by n-butanol and absorbance was measured at 532 nm as described in details in our previous work (27). Data were expressed as nM.

Measurement of TNF-α, IL-1β, IL-6 and NF-kb
Quantitative detection of TNF-α, IL-1β, IL-6 and NF-kb levels in serum were performed using an enzyme-linked immunosorbent assay rat specific ELISA kit according to each specific protocol. The absorbance of the final colored product was measured in 450 nm as the primary wave length and 620 nm as the reference wave length. TNF-α, IL-1β, IL-6 and NF-kb levels were expressed as pg/ml.

Measurement of TAP
Serum TAP was evaluated by measuring the ability to reduce Fe³⁺ to Fe²⁺. Interaction of TPTZ with Fe²⁺ results in formation of a blue color with a maximum absorbance at 593 nm. The whole procedure has been described in our previous study (27). Data were expressed as nM.

Measurement of testosterone and DHEA-S
For determination of testosterone and DHEA-S, specific ELISA kits were used and the instruction of their brochure was followed. Testosterone and DHEA-S were expressed as ng/ml.

Statistical analysis
Results are expressed as mean±standard error of the mean (SEM). Data were analyzed by one-way ANOVA followed by Tukey post-hoc test for multiple comparisons to ensure the variances of the data are distributed properly. A P-value less than 0.05 were considered significant. The Stats Direct version 2.7.9 was used.

Results
A significant increase in TBARS (Figure 1, 11.9±0.2 vs. 20.66±0.88, P<0.05) and a significant decrease in TAP (Figure 2, 218±8 vs. 120±7.5, P<0.05) were observed when sham group was compared with D-galactose-received aged group. Figures 3-6 show the effects of aging on the levels of TNF-α, IL-6, IL-1β, and NF-kb, respectively in comparison to sham (32±2.3 vs. 59±15, P<0.05; 1.2±0.5 vs. 2.5±0.33, P<0.05; 27±3.9 vs. 49.66±3.4, P<0.05; 45.7±2.4 vs. 97±21.2, P<0.05). As shown in Figures 7 and 8, testosterone and DHEA-S (0.6±0.05 vs. 0.25±0.03, P<0.05; 1.2±0.2 vs. 0.6±0.08, P<0.05) in aged mice was lower than that in the sham.

Effects of Z. officinale in aged mice
Z. officinale treatment recovered D-galactose-induced rats by reducing TBARS (14.5±1.6 vs. 20.66±0.88, P<0.05), and increasing TAP (169±3.5 vs. 120±7.5, P<0.05), (Figures 1, 2). Figures 3-6 show that administration of Z. officinale recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kb (39±2.6 vs. 59±15, P<0.05; 1.3±0.3 vs. 2.5±0.33, P<0.05; 32.3±0.54 vs. 49.66±3.4, P<0.05; 68±1±5.7 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, Z. officinale increased testosterone and DHEA-S (4.6±0.04 vs. 2.5±0.03, P<0.05; 1.28±0.17 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of G. glabra in aged mice
D-galactose-induced elevation of TBARS and reduction of TAP (Figures 1, 2) were significantly recovered following treatment with G. glabra (13.1±1.01 vs. 20.66±0.88, P<0.05; 203±17 vs. 120±7.5, P<0.05). Figures 3-6 show that administration of G. glabra recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β and NF-kb (33±9.5 vs. 59±15, P<0.05; 1.2±0.14 vs. 2.5±0.33, P<0.05; 30±8.31 vs. 49.66±3.4, P<0.05; 57±2±8.7 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, G. glabra increased testosterone and DHEA-S levels (0.49±0.05 vs. 0.25±0.03, P<0.05; 1.3±0.34 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of R. officinalis in aged mice
R. officinalis treatment recovered D-galactose-induced rats by reducing TBARS (13.2±0.63 vs. 20.66±0.88, P<0.05), and increasing TAP (167±5.3 vs. 120±7.5, P<0.05) (Figures 1, 2). Figures 3-6 show that administration of R. officinalis recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kb (42±12 vs. 59±15, P<0.05; 1.2±0.2 vs. 2.5±0.33, P<0.05; 33.5±4.1 vs. 49.66±3.4, P<0.05; 58±1±3.8 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, R. officinalis increased testosterone, (0.46±0.09 vs. 0.25±0.03, P<0.05) and DHEA-S (1.17±0.19 vs. 0.6±0.08, P<0.05) in aged mice.
Figure 1. Effects of herbs on serum thiobarbituric acid reactive substances (TBARS) as lipid peroxidation (LPO) marker in aged mice. Data are mean±SEM of ten animals.

*Significantly different between sham group and other groups at P<0.05.

*Significantly different between D-galactose group and other groups at P<0.05.

Figure 2. Effects of herbs on serum total antioxidant power (TAP) of aged mice. Data are mean±SEM of ten animals.

*Significantly different between sham group and other groups at P<0.05.

*Significantly different between D-galactose group and other groups at P<0.05.

Figure 3. Effects of herbs on serum interleukin-1 beta (IL-1β) in aged mice. Data are mean±SEM of ten animals.

*Significantly different between Sham group and other groups at P<0.05.

*Significantly different between D-Galactose group and other groups at P<0.05.

Figure 4. Effects of herbs on serum tumor necrosis factor-alpha (TNF-α) in aged mice. Data are mean±SEM of ten animals.

*Significantly different between Sham group and other groups at P<0.05.

*Significantly different between D-Galactose group and other groups at P<0.05.
**Figure 5.** Effects of herbs on serum interleukin 6 (IL-6) in aged mice. Data are mean±SEM of ten animals.

- Significantly different between Sham group and other groups at \( P < 0.05 \)
- Significantly different between D-Galactose group and other groups at \( P < 0.05 \)

**Figure 6.** Effects of herbs on serum NF-kappaB (NF-kB) in aged mice. Data are mean±SEM of ten animals.

- Significantly different between Sham group and other groups at \( P < 0.05 \)
- Significantly different between D-Galactose group and other groups at \( P < 0.05 \)

**Figure 7.** Effects of herbs on serum testosterone in aged mice. Data are mean±SEM of ten animals.

- Significantly different between Sham group and other groups at \( P < 0.05 \)
- Significantly different between D-Galactose group and other groups at \( P < 0.05 \)

**Figure 8.** Effects of herb on serum dehydroepiandrosterone-sulfate (DHEA-S) in aged mice. Data are mean±SEM of ten animals.

- Significantly different between Sham group and other groups at \( P < 0.05 \)
- Significantly different between D-Galactose group and other groups at \( P < 0.05 \)
Effects of P. harmala in aged mice

D-galactose-induced elevation of TBARS and reduction of TAP (Figure 1, 2) were significantly recovered following treatment with P. harmala (15.5±1.73 vs. 20.66±0.88, P<0.05; 172±13.9 vs. 120±7.5, P<0.05). Figures 3-6 show that administration of P. harmala recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kB (41.8±6.8 vs. 59±15, P<0.05; 1.7±0.18 vs. 2.5±0.33, P<0.05; 35.1±1.42 vs. 49.66±3.4, P<0.05; 69±7.2 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, P. harmala increased testosterone and DHEA-S (0.45±0.08 vs. 0.25±0.03, P<0.05; 0.9±0.7 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of A. vera in aged mice

A. vera treatment recovered D-galactose-induced elevation of TBARS (Figure 1, 13.5±1.7 vs. 20.66±0.88, P<0.05), and improved TAP (Figure 2, 171±4.03 vs. 120±7.5, P<0.05). Figures 3-6 show that administration of A. vera recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kB (42±6.94 vs. 59±15, P<0.05; 1.5±0.09 vs. 2.5±0.33, P<0.05; 35±0.77 vs. 49.66±3.4, P<0.05; 59±14 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, A. vera increased testosterone and DHEA-S levels (0.47±0.09 vs. 0.25±0.03, P<0.05; 1.12±0.19 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of S. hortensis in aged mice

S. hortensis treatment recovered D-galactose-induced elevation of TBARS (Figure 1, 13.9±2.2 vs. 20.66±0.88, P<0.05), and increased TAP (Figure 2, 176±14.16 vs. 120±7.5, P<0.05). Figures 3-6 show that administration of S. hortensis recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kB (37±9.9 vs. 59±15, P<0.05; 1.2±0.05 vs. 2.5±0.33, P<0.05; 34.7±1.8 vs. 49.66±3.4, P<0.05; 56.2±13.9 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, S. hortensis increased testosterone and DHEA-S (0.45±0.05 vs. 0.25±0.03, P<0.05; 1.02±0.15 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of T. scordium in aged mice

D-galactose-induced elevation of TBARS and reduction of TAP (Figure 1, 2) were significantly recovered following treatment with T. scordium (14±0.76 vs. 20.66±0.88, P<0.05; 170±6.74 vs. 120±7.5, P<0.05). Figures 3-6 show that administration of T. scordium recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kB (39±10.26 vs. 59±15, P<0.05; 1.4±0.28 vs. 2.5±0.33, P<0.05; 33±1.1 vs. 49.66±3.4, P<0.05; NF-kB: 59±24.42 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, T. scordium increased testosterone and DHEA-S (0.44±0.05 vs. 0.25±0.03, P<0.05; 1.08±0.24 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of H. perforatum in aged mice

H. perforatum treatment recovered D-galactose-induced rats by reducing TBARS (13±1.9 vs. 20.66±0.88, P<0.05) and increasing TAP (190±23 vs. 120±7.5, P<0.05) (Figures 1, 2). Figures 3-6 show that administration of H. perforatum recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β and NF-kB (35.5±2.12 vs. 59±15, P<0.05; 1.1±0.09 vs. 2.5±0.33, P<0.05; 26±11±1 vs. 49.66±3.4, P<0.05; 45±4.2 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, H. perforatum increased testosterone and DHEA-S (0.51±0.06 vs. 0.25±0.03, P<0.05; 1.2±0.18 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of S. marianum in aged mice

D-galactose-induced elevation of TBARS and reduction of TAP (Figure 1, 2) were significantly recovered following treatment with S. marianum (12.58±0.64 vs. 20.66±0.88, P<0.05; 210±12.14 vs. 120±7.5, P<0.05). Figures 3-6 show that administration of S. marianum recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kB (36±5.4 vs. 59±15, P<0.05; 1.1±0.09 vs. 2.5±0.33, P<0.05; 26±11 vs. 49.66±3.4, P<0.05; 45±4.2 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, the S. marianum recovered D-galactose-induced reduction of testosterone and DHEA-S (0.57±0.09 vs. 0.25±0.03, P<0.05; 1.3±0.22 vs. 0.6±0.08, P<0.05) in aged mice.

Effects of vitamin E in aged mice

D-galactose-induced elevation of TBARS and reduction in TAP (Figure 1, 2) were significantly recovered following treatment with Vitamin E (13±0.83 vs. 20.66±0.88, P<0.05; 190±13.1 vs. 120±7.5, P<0.05). Figures 3-6 show that administration of Vitamin E recovered D-galactose-induced increase in TNF-α, IL-6, IL-1β, and NF-kB (35.5±2.12 vs. 59±15, P<0.05; 1.4±0.26 vs. 2.5±0.33, P<0.05; 30.1±2.2 vs. 49.66±3.4, P<0.05; 57±3.9 vs. 97±21.2, P<0.05), respectively. As shown in Figures 7 and 8, Vitamin E increased testosterone and DHEA-S (0.44±0.02 vs. 0.2±0.03, P<0.05; 1.0±0.16 vs. 0.6±0.08, P<0.05) in aged mice.

Discussion

In this study, for the first time, we analyzed the anti-aging potentials of nine famous herbs in a well-setup animal aging model using chronic administration of D-galactose. Our results showed that production of free radicals is the principal reason of up-regulation of pro-inflammatory cytokines and the main determinant involved in the D-galactose-induced aging model. Furthermore, these herbs dramatically diminished oxidative stress and proinflammatory cytokines in the aged mice. Supporting the mechanism of action of these herbs and the theory of oxidative stress in aging, vitamin E was used as the standard and showed the similar effects in examined markers of aging.
Interestingly, present results indicated improvement of testosterone and DHEA-S by herbs in the aged mice. Decline of steroid hormones with aging is already known and is believed a major contributor to elevation of pro-inflammatory markers (28).

Recent studies have shown the mechanisms of action of anti-aging herbs in reducing aging process that is divided into four categories including antioxidant, anti-inflammatory, effect on memory/cognition/mood, and the sex hormones (Figure 9). This indicates that most of anti-aging herbals have antioxidant components (16) and thus supports the present findings and hypothesis of this study.

Ginger [Zingiber officinale Roscoe (Zingiberaceae)] and supplements derived from ginger like zingerone, shogaols and gingerols posses the abilities for the treatment of chronic inflammation. The protective effects of Z. officinale in lessening macromolecular damage in aged mice were shown in this study. Besides, recent study has shown that ginger extracts owns antioxidant activity (29). It has been recently shown that pre-trial administration of this herb expedites conditioned inhibitory learning in adult rats (30). Also, it has been found that Z. officinale has possibly good effects on age-related execution shortages and defends against oxidative stress in old rats, suggesting this compound as a useful factor in treating age-related disturbances (31).

G. glabra (licorice extract) or licorice is the root of G. glabra from which a sweet flavor can be extracted. The results of this study showed that G. glabra has the protective effects in declining macromolecular damage in aged mice. It has been shown that G. glabra extract is the safest pigment-lightening agent with the fewest side effects (32). Additionally, G. glabra has anti-inflammatory properties hypothetically helpful in diminishing skin reddness and postinflammatory hyperpigmentation. Interestingly, it appears to be more useful for the hyperpigmentation related to skin aging (33).

R. officinale leaves possess a variety of bioactive agents, including antioxidants and anti-inflammatories (34). The most potent antioxidant constituents are polyphenolics such as quercitrin and quercetin (35). The results of this study showed that R. officinale has the protective effects in decreasing macromolecular damage in aged mice during aging. In addition, R. officinale extract has shown free radical scavenging effect in the hippocampus (36). This is supported with a raising number of reports showing that natural extracts and phytochemicals have a constructive effect on brain aging through their action on ROS, specifically in the hippocampus (37).

P. harmala L. is known as Syrian rue, Wild rue and Harmal. The P. harmala has antibacterial, antifungal, antiviral, antioxidant, anti-diabetic, antitumor, antileishmanial, insecticidal, cytotoxic, hepatoprotective, and antinociceptive effects (38-39). In this study P. harmala showed the protective effects in improving antioxidant, anti-inflammatory and male sex hormones that were affected in aged mice during aging. In fact, flavonoids as a powerful antioxidant isolated from P. harmala, can remove the lipid peroxide radicals (40). Also, P. harmala treatment appeared to be a versatile strategy to conserve testicular uprightness and function during aging in male rats (41).

The leaves of A. vera (A. barbadensis) (Fam. Liliaceace) are the source of aloe vera gel. A. vera gel is greatly used in cosmetics and toiletries for its moisturizing and regenerating action. Also, the leaf
of A. vera could assist cellular repairing, imbibition of foods, vitamins, minerals and vital nutrients (42). In this study A. vera showed a protective effect in improving antioxidant, anti-inflammatory and male sex hormones that were affected in aged mice. The anti-inflammatory property of A. vera has been documented in inflammation through suppression of free radicals and ROS (43). It has been shown that the life-long dietary supplementation of A. vera suppresses many age-related consequences in rats (44). Also, it has been suggested A. vera could suppress oxidative damage and age-related increases in hepatic cholesterol during life-long dietary (45).

S. hortensis L. is an annual culinary herb belonging to the family Labiatae. It is known as summer savory. Besides, this plant exhibited analgesic, antibacterial, antifungal, antioxidant and antihyperglycemic properties. In addition, the antigenotoxic effects of S. hortensis L. was shown on rat lymphocytes exposed to oxidative stress (46). The major constituents of the S. hortensis are carvacrol, gamma-terpinene, thymol and para-cymene (47). The investigation showed that carvacrol, thymol and flavonoids of Satureja species are responsible to marked reduction of serum cholesterol in diabetic patients (48-50). It has been reported that age-related alterations of fatty acid composition in liver was accompanied by intake of savory essential oil through intensification of polyunsaturated fatty acids synthesis in mice liver and reduction of lipid peroxidation products (51).

As a result, S. hortensis shows protective effects in improving antioxidant, anti-inflammatory and male sex hormones that were affected in aged mice during aging.

The genus Teucrium (Labiatae) comprises 12 species, which possess antioxidant, anti-spasmodic, antinociceptive and anti-inflammatory properties (52-54). According to the literature, b-caryophyllene and caryophyllene oxide were reported as the main sesquiterpenes in many Teucrium species. Investigation of the chemical constituents of the oil of T. scordium showed b-caryophyllene, caryophyllene oxide and also (E)-b-farnesene as the major components of T. scordium (55). In addition, b-caryophyllene is known as anti-inflammatory sesquiterpene (56); this effect may confirm the anti inflammatory activity of this plant (55). Nevertheless, no reports are valid about T. scordium in relation to its possible oxidative stress inhibitory potential in aged individuals. As much as we know this is the first study that shows antioxidant and anti-inflammatory effects of T. scordium extract in the aged rats. Our results indicated that T. scordium decreases inflammatory mediators and increases anti-oxidative power and steroid hormones in aged mice. Thus, T. scordium could be included in the diet as a nutritional supplement to increase the defenses of body against oxidative stress.

H. perforatum L. is responsible for pharmacological properties, antiseptics (57), anti-inflammatory (58), antitumoral activities (59, 60). Today, H. perforatum is known as one of the few economic plants that include great ingredients of hypericins, hyperforins, and flavonoids. The results of this study showed that H. perforatum owns the protective effects in decreasing macromolecular damage in aged mice during aging. St. John’s Wort is the dried tops or aerial parts of H. perforatum which gathered before or during flowering and is used in the therapy of anxiety related to aging (61).

Silymarin (SM), a flavonoids complex known as ‘milk thistle’ is extracted from the fruit of Silybum marianum (L.) Gaertn. (Carduus marianus L., Astera-ceae). Interestingly, the present findings confirmed that S. marianum causes the best effects in improving antioxidant, anti-inflammatory and male sex hormones in aged mice. This effect is so important and should be considered as an advantage. This can be explained with current knowledge that among many medicinal plants, S. marianum, has been greatly used for centuries as a natural popular complementary medicine for the treatment of several diseases. The main indications for the use of silymarin are related to the hepatoprotection (62, 63). Also, its efficacy in inflammatory oxidative-mediated diseases like colitis has been confirmed (64, 65). It is noteworthy that several age-related brain and neurodegenerative diseases happen due to amplified oxidative stress. The search for compounds acting on the upgrading of cognitive performance and neuroprotection through antioxidant motion is now a great interest (66). Excitingly, it has been shown that use of S. marianum for prevention and treatment of neurodegenerative diseases and processes associated with aging improves physiological responses against the ROS in the neural cells (67).

Conclusion

Taking collectively, the present results confirmed our hypothesis that the herbs with highest antioxidant power may reduce speed and rate of aging as evidenced by recovery of proinflammatory cytokines and sex hormones. Among tested herbs, S. marianum showed the best effect in improving all the D-galactose-induced aging effects. Since all of the selected and examined herbs are already found safe in human and there are good information from traditional medicine, therefore, they can be supplemented into the diet of elderly people to reduce speed of aging. Testing the mixture of these herbs together or with other anti-aging products is among the plans of future.
Acknowledgment
This study was undertaken as a project and the results described in this paper were part of student thesis. Authors also thank assistance of National Elite Foundation and the INSF.

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