In silico evaluation of the compounds of the ayurvedic drug, AYUSH-64, for the action against the SARS-CoV-2 main protease

Thrigulla Saketh Rama, Manne Munikumar, Vankudavath Naik Raju, Parasannanavar Devara, Naveen Kumar Boiroju, Rajkumar Hemalatha, P.V.V. Prasad, Manohar Gundeti, Brijesh S. Sisodia, Sharad Pawar, G.P. Prasad, Mukesh Chincholikar, Sumeet Goel, Anupam Mangal, Sudesh Gaidhani, N. Srikanth, K.S. Dhiman

CCRAS-National Institute of Indian Medical Heritage, Revenue Board Colony, Gaddiannaram, Hyderabad, 500036, Telangana State, India
NIN-TATA Centre for Excellence in Public Health Nutrition, ICMR-National Institute of Nutrition, Hyderabad, 500007, Telangana State, India
Nutrition Information, Communication & Health Education (NICHE), ICMR-National Institute of Nutrition, Hyderabad, 500007, Telangana State, India
ICMR-National Institute of Nutrition, Hyderabad, 500007, Telangana State, India
CCRAS-Raja Ramdeo Anandilal Podar (RRAP) Central Ayurveda Research Institute for Cancer, Mumbai, India
ICMR-National Institute of Nutrition, Hyderabad, 500007, Telangana State, India
CCRAS-Regional Ayurveda Research Institute for Drug Development, Gwalior, India
CCRAS-Regional Ayurveda Institute for Fundamental Research, Pune, India
Central Council for Research in Ayurvedic Sciences, New Delhi, India

Abstract

Background: Outbreak of Corona Virus Disease in late 2019 (COVID-19) has become a pandemic global Public health emergency. Since there is no approved anti-viral drug or vaccine declared for the disease and investigating existing drugs against the COVID-19.

Objective: AYUSH-64 is an Ayurvedic formulation, developed and patented by Central Council of Research in Ayurvedic Sciences, India, has been in clinical use as anti-malarial, anti-inflammatory, anti-pyretic drug for few decades. Thus, the present study was undertaken to evaluate AYUSH-64 compounds available in this drug against Severe Acute Respiratory Syndrome-Corona Virus (SARS-CoV-2) Main Protease (Mpro; PDB ID: 6LU7) via in silico techniques.

Materials and methods: Different molecular docking software’s of Discovery studio and Auto Dock Vina were used for drugs from selected AYUSH-64 compounds against SARS-CoV-2. We also conducted 100 ns period of molecular dynamics simulations with Desmond and further MM/GBSA for the best complex of AYUSH-64 with Mpro of SARS-CoV-2.

Results: Among 36 compounds of four ingredients of AYUSH-64 screened, 35 observed to exhibit good binding energies than the published positive co-crystal compound of N3 peptide. The best affinity and interactions of Akuammicine N-Oxide (from Alstonia scholaris) towards the Mpro with binding energy (AutoDock Vina) of –8.4 kcal/mol and Discovery studio of Libdock score of 147.92 kcal/mol. Further, molecular dynamics simulations with MM/GBSA were also performed for Akuammicine N-Oxide docked complex to identify the stability, specific interaction between the enzyme and the ligand. Akuammicine N-Oxide is strongly formed h-bonds with crucial Mpro residues, Cys145, and His164.

Conclusion: The results provide lead that, the presence of Mpro-Akuammicine N-Oxide with highest Mpro binding energy along with other 34 chemical compounds having similar activity as part of AYUSH-64 make it a suitable candidate for repurposing to management of COVID-19 by further validating through experimental, clinical studies.

© 2021 The Authors. Published by Elsevier B.V. on behalf of Institute of Transdisciplinary Health Sciences and Technology and World Ayurveda Foundation. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).
1. Introduction

The outbreak of corona virus disease (now known as COVID-19) in late 2019 in Wuhan, China led to a global public health emergency that currently affects more than 200 countries worldwide. Coronavirus, crown-like spikes positive-sense ribonucleic acid (RNA) from Coronaviridae family. Over the past two decades, severe acute respiratory syndrome coronavirus (SARS-CoV) and Middle East respiratory syndrome coronavirus (MERS-CoV) lead to pandemic in 2002 and 2012 resulted in a 10% and 37% mortality rate respectively [1]. In December 2019, a new coronavirus of probable bat origin caused an outbreak of human pulmonary disease in Wuhan, China, that quickly spread throughout the world [2,3]. The causative agent initially named as 2019-novel coronavirus (2019-nCoV) by world health organisation (WHO). As per name recommendation from International Committee of Corona virus Study Group (ICCSG), the virus named as Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2), its RNA genome shares 82.30% and similar pathogenesis (host) of its identity with SARS coronavirus (SARS-CoV) [1,4]. Ultimately, the disease became known as corona virus disease 2019 (COVID-19) (WHO, 2020). On January 30, 2020, WHO officially declared the COVID-19 epidemic a public health emergency of international concern and on March 11, 2020, the outbreak was confirmed a pandemic (WHO, 2020). The SARS-CoV and SARS-CoV-2 shares the similar pathogenesis (host) due to similarities in their genome [1].

Though several drug targets of coronaviruses have been identified, the main protease (Mpro), also known as 3-chymotrypsin-like protease (3CLpro), has emerged as the best-described drug target [5]. The Mpro processes the large polypeptide, translated from the viral RNA, specifically at 11 splicing regions mostly Leu to Gln (Ser, Ala and Gly). These significant splicing regions non-homologous to human. Inhibiting enzyme activity might results in blocking viral pathogenesis, making it an attractive drug target for SARS-CoV-2. Further, a mechanism-based peptide-like inhibitor (N3) has also been identified via computer-aided drug design, and then crystal structure of the Mpro of SARS-CoV-2 in complex with this compound has been determined [6].

Given the lack of any validated, approved therapeutic intervention, such as a drug or vaccine, the COVID-19 pandemic continues to spread around the world. In such circumstances, the concept of drug repositioning might be a cost-effective, time-efficient, and less labour-intensive option for development of possible therapeutic and/or prophylactic lead candidate(s) from existing traditional and/or approved drugs [7]. In comparison to synthetic inhibitors, plant based-drugs are known to have low toxicity and are much safer to use. Natural products, such as traditional medicines and plant-derived compounds (phytochemicals) are considered rich sources of promising antiviral drugs [8].

Natural products were the sources of approximately 44% of the approved antiviral drugs from 1981 to 2006 [9]. Recent studies have suggested that drug poisoning and severe to moderate adverse effects associated with hydroxychloroquine have been reported in diabetic and hypersensitive patients, in whom COVID-19 tends to affect severely. Further, hydroxychloroquine is also known for inhibition of pro-inflammatory cytokines which leads to Acute Respiratory Distress Syndrome (ARDS) [10]. Due to the severe adverse effects shown by these synthetic compounds, natural, non-synthetic, target specific drugs with minimal side effects are urgently required for prophylaxis and treatment of COVID-19 [11].

Plant compounds and natural phyto-molecules are most economical and ideal for exploration to develop drugs through the drug discovery process [12]. Recent studies via integrated computational approaches have highlighted the repurposing of approved and known drugs against SARS-CoV-2 drug targets, either individually or in combination, to combat the virulence of COVID-19. Several plant compounds have been reported, using in silico approaches, as potential inhibitors of the Mpro of SARS-CoV-2. Mimicked drugs of bicitravir, dovitegravir, paritavir, and raltegravir have been studied against 3CLpro and 2’-OMTase through in silico approaches [13,14]. Another study reported darunavir, remdesivir, and saquinavir, the natural compounds derivatives from coumarine and flavone which inhibits 3CLpro [13–15]. In another in silico study also shown that, Andrographis paniculata’s compound Andrographolide, inhibits Mpro of SARS-CoV-2. Its good solubility, pharmacodynamic properties, target accuracy, and adherence to Lipinski’s rule of five were also predicted using computational tools [11].

The potent plant compounds nelfinavir, lopinavir, kaempferol, quercetin, luteolin–7-glucoside, demethoxycurcumin, naringenin, apigenin–7-glucoside, oleuropein, curcumin, catechin, and epicatechin–gallate, have been shown to inhibit the Mpro [12]. At the same time, several potential anti-SARS-CoV-2 lead compounds have been revealed in the medicinal plant library of traditional Chinese medicine compounds [16]. Another study identified oolonghomobisflavan-A from the tea plant as a potential inhibitor against Mpro of SARS-CoV-2 [17]. In addition, a promising drug candidate from plant sources, bonducellin D, has been reported to exhibit broad-spectrum inhibition potential against both the SARS-CoV Mpro and the MERS-CoV Mpro [18].

AYUSH-64 is a poly-herbal compound developed by the Central Council for Research in Ayurvedic Sciences, the main organization for research and development in Ayurveda in India under the Ministry of AYUSH, Government of India. It consists of four ingredients viz., Saptaparna (Alstonia scholaris R. Br.) bark aqueous extract 100 mg, Ketuca (Picrorhiza kurroa Royle ex. Benth) root extract 100 mg, Kiratatikta (Swertia chirata Pexbex. Karst) whole-plant extract 100 mg, and Kuberaaksha (Caesalpinia crista L.) seed powder 200 mg [19].

To summarize the information provided in Table 1, it is understood that all four ingredients of AYUSH-64 are characterized by tikta rasa (bitter taste) and therefore amapachk (able to digest the Ama or undigested form of food) and hence act as Jvaraghna (anti-pyretic). The combined effects of these herbs are jvararahara (able to relieve fever), sannipata jvararahara (able to relieve intermittent fevers), krimihara (wormicidal), jantuvara (anthelmintic/antimicrobial/antiviral), and shoathara (anti-inflammatory), rendering this compound a potent combination against conditions such as Influenza Like Illnesses having the symptoms of cough, cold, headache and fever. Several compounds found in the drug have been reported to exhibit several pharmacological activities including anti-malarial, anti-viral, anti-inflammatory, immunomodulatory etc. (Table 2).

Various pre-clinical studies were performed on AYUSH-64 to study its safety and efficacy aspects (Table 3). The major studies, namely anti-malarial property in albino mice, found safe and non-toxic in a dose of 500 mg/kg of body weight for 12 weeks (Anonymous, 1987) and potential antiviral activity of Chirakin (marketed name of AYUSH-64 by Zandu) against Chikungunya virus were investigated at the department of Molecular Virology Laboratory, Rajiv Gandhi Centre for Biotechnology, Thiruvananthapuram, by protection and plaque reduction assay. The efficacy of the compound was expressed in terms of activity index and selectivity index in protection assay and Vero cells were infected with Chikungunya Virus (CHIKV) and treated with an optimum concentration of Chirakin (25 μg/mL). Chirakin treated cells and the reduction in virus yield was determined by plaque assay in Vero cells. Chirakin shows antiviral activity against Chikungunya virus in this study. Its activity was found better than ribavirin in protection assays, where as in plaque reduction assays, both perform almost
equally with a 2 log_{10} reduction in virus number (CK Katiyar. Technical note on Chirakin Tab – Unpublished report).

Attempts have also been made to clinically evaluate the compound for management of Microfilaria [20, 21] effectiveness against *Plasmodium vivax* in ring stage than in gamete stage and also the mixed infection cases of *P. vivax* [22] decline in infectivity rate of malaria, double-blind study for comparative efficacy with chloroquine/primaquine in *P. vivax* malaria and lead candidate in the management of influenza like illnesses [23] (Table 4). Overall, the compound is considered to be safe and is prescribed widely by Ayurvedic experts for the effective management of malaria, joint pains, fever, and influenza like illnesses. In this *in silico* study, 36 major compounds were analysed which are four ingredients of AYUSH-64 drug might be potent inhibitors against M_{pro} of SARS-CoV-2 observed via two different docking strategies of AutoDock Vina and Discovery studio molecular docking tools.

2. Materials and methods

In the drug discovery process for COVID-19 crisis, the development of novel drugs with potential interactions with therapeutic targets is of primal importance. Conventionally, promising-lead or drug identification is achieved by traditional or experimental high-throughput screening (HTS), which is time-consuming and cost-effective [24]. In contrast to the traditional drug discovery method (classical or forward pharmacology), the rational drug design is noted as efficient and economical [25]. In these contrasts, approach to identifying therapeutics is to repurpose approved compounds developed for other uses, by taking advantage of existing detailed information on human pharmacology and toxicology to enable rapid clinical trials and regulatory quick review. Furthermore, *in silico* drug design method is also known as reverse pharmacology, as the initial step is to identify promising drug targets (proteins/enzymes), after which they are used for screening of novel small-molecule inhibitors for the candidates [14, 26–28]. In the present study of *in silico* drug design method, molecular docking approach was used for SARS-CoV-2 M_{pro} and 36 AYUSH-64 compounds, to screen for the novel inhibitors by two different docking strategies/ algorithms, LibDock protocol of BIOVIA Discovery Studio and AutoDock Vina.

2.1. M_{pro} structure preparation

The crystal structure of COVID-19 M_{pro} in complex with an peptide inhibitor N3 (n-[5-methylisoxazol-3-yl]carbonyl)alaninyl-l-valyl-n-1-[(1r,2z)-4-{benzoyloxy}-4-oxo-1-[[3r-2oxopyrrolidin-3-yl]methyl]but-2-enyl]-l-leucinamide) peptide obtained from Protein Data Bank (PDB) [6]with the PDB ID of 6LU7 with crystal resolution 1.5 Å for the molecular docking studies. Foremost, for the preparation of the protein AutoDock Tool (ADT) used for removal of all HOH molecule, adding Kollman charges and polar hydrogen atoms, assigning hydrogen polarities. Further, Gasteiger charges were applied on prepared protein and protein structures using BIOVIA Discovery Studio and AutoDock Vina.

Table 1

| Name of the plant | Rasa (Taste) | Guna (Property/Quality) | Virya (Potency) | Vipaka (biotransformed rasa) | Karma (Pharmacological actions) |
|-------------------|-------------|-------------------------|----------------|---------------------------|--------------------------------|
| Saptaaparna [40–42] (Alstonia scholaris, K. Br.) | Tikta (bitter), Kashaya (astringent) | Sara, Ushna (hot), Snigdha (oleous), Deepan (kindling), Laghu (light) | Ushna (hot) | Katu (pungent) | Shoolahara (relieves pain), Gulumahara (relieves bloating), Krinmahara (wurmicidal), Hrudya (cardiac tonic), Shwasahara (useful in asthma), vronahara (wound healer), Arsdoshahara (useful in blood related diseases), jantahara (antihelmintic), Trisodhahara (pacific kapha, pitta and vata), Kasahara (useful in bronchitis), Kushtahara (useful in skin disorders), Jvaraghna (Antipyretic), Saraika Pravahkahar (useful in bloody dysentery), Vatarkahara (useful in Gout), Grahahara (useful in IBS like conditions) Udaprapaharam (relieves urticaria), Dantakrimihara (useful in dental caries), Vishaghna (Antitoxic) |
| Katuki [41, 42] (Picrorhiza kurroa, Royle ex. Benth) | Tikta (bitter) | Laghu (light), Ruksha (dry) | Shita (cold) | Katu (pungent) | Bhedana (causes purgation), Deepana (improves digestion), Hrudya (cardiac tonic), Jvaraghna (useful in fevers), Vishamjvaranashini (useful in recurrent fevers) Kaphapittahara (pacifics Kapha and pitta), Prameghahara (useful in urinary disorders/diabetes), Shwasas-Rasaghna (useful in asthma, cough), Dahaguna (relieves burning sensation), Kushtahara (useful in skin disorders), Lekhana (therapeutic scraping), Krinmahara (wurmicidal), Arochakaguna (useful in tastelessness) Arajut (useful in blood disorders), Stanyashodhan (improves quality of breast milk) Hikkanigrahana (relieves hiccups)

Shoolahara (relieves pain), Gulumahara (relieves bloating), Krinmahara (wurmicidal), Hrudya (cardiac tonic), Shwasahara (useful in asthma), vronahara (wound healer), Arsdoshahara (useful in blood related diseases), jantahara (antihelmintic), Trisodhahara (pacific kapha, pitta and vata), Kasahara (useful in bronchitis), Kushtahara (useful in skin disorders), Jvaraghna (Antipyretic), Saraika Pravahkahar (useful in bloody dysentery), Vatarkahara (useful in Gout), Grahahara (useful in IBS like conditions) Udaprapaharam (relieves urticaria), Dantakrimihara (useful in dental caries), Vishaghna (Antitoxic) |
Table 2
The pharmacological activity of AYUSH-64 by ingredient.

| S.No | Name of the plant                     | Chemical Compound(s) & pharmacological activities                                                                 |
|------|--------------------------------------|---------------------------------------------------------------------------------------------------------------|
| 1    | Saptaparni (Alstonia scholaris) R. Br.) | Methanol extracted from bark of *Alstonia scholaris*<br>Found to be more promising. Nevertheless, the observed antimalarial activity of *Alstonia scholaris* was less pronounced as compared to *Alstonia macrophylla* [44].<br>*Alstonia macrophylla*, which is seen in India, generally adulterated for *Alstonia scholaris* [44]. Four new alkaloids, alstiphyllanines A-D (1–4), were isolated from *Alstonia macrophylla* and their structures determined by MS and 2D NMR analyses. Alkaloids 1–4 showed moderate anti-plasmodial activity against *Plasmodium falciparum* and vasorelaxant activity against phenylephrine-induced contraction of isolated rat aorta. Other Studies: 1. Inhibited the carrageenan-induced inflammation in the rat paw oedema study model [45]. 2. Controls Malarial fever by virtue of its strong schizontocidal activity [46]. 3. Anti-HSV and anti-adenovirus activity of indole alkaloids from leaves of *Alstonia scholaris*, with significant inhibitory activity against herpes simplex virus (HSV) and adenovirus [47]. 4. Enhances DNA repair capacity [48]. 5. In vitro tests, alkaloids exhibited inhibition of inflammatory mediators (COX-1, COX-2 and 5-LOX), which is accordant with results on animal model [49]. 6. Potent antimalarial activity against *P. falciparum* [50,51]. |
| 2    | Katuki (Picrorhiza kurroa Royle ex. Benth) | 1. Anti-inflammatory effect by 13-adrenergic blockade [52].<br>2. Stimulates the cell-mediated and humoral components of the immune system as well as phagocytosis in experimental animals [53,54].<br>3. Improves the immune system by increasing the proliferation of lymphocytes and cytokine levels (IL-4 and IFN-gamma) in serum, in HA titre, DTH, PFC, phagocytic index and CD4/CD8 population [55].<br>4. Another study demonstrates the antioxidant and free radical scavenging activity of the leaf extract of same plant [56].<br>5. Inhibits the growth of *Plasmodium falciparum* Significantly [57–59].<br>The drug shows promising effect due to its antiparasitic activity and antimalarial activity [58].<br>1. Anti-protozoal activity: A MEOH extract of *Swertia chirata* found to inhibit the catalytic activity of topoisomerase I of *Leishmania donovani* was subjected to fractionation to yield three secoiridoid glycosides: amarogentin (a), amaroswerin (b), and sweroside (c). Amarogentin is a potent inhibitor of type I DNA topoisomerase from *Leishmania* and exerts its effect by interaction with the enzyme, preventing binary complex formation. Other Studies: 1. Inhibits the expression of viral protein R (an attractive target for HIV disease) in HeLa cells harbouring the TREx plasmid encoding full-length Vpr (TREx-HeLa-Vpr cells) [60].<br>2. Inhibited HSV-1, plaque formation at more than 70% level and viral dissemination [61].<br>3. Suppressive effects on inflammatory mediators by blocking the expression of COX-2 and phosphorylation of Akt, IKK-β, MAPK and NF-κB, activation in LPS-stimulated macrophages [62].<br>4. A study on experimental arthritis in rats suggested amelioration of oxidative and inflammatory stress, which implies the immunomodulatory effect of leaves of the above said plant. Post treatment with the leaves of the said plant, the animal subjects showed marked reduction in inflammation as well as arthritic changes [63].<br>5. Protects against red blood cell (RBC) haemolysis and DNA damage [66].<br>4. Immuno-stimulatory: increase in hemagglutinating antibody titre and a change in delayed-type hypersensitivity [26].<br>5. Two weeks after challenge with *Pseudomonas aeruginosa*, the Caesalpinia treated animals showed a significant bacterial clearance from the lungs, with less severe incidence of lung abscess [67].<br>6. Exhibited activity against the vaccinia virus [68].<br>7. In vivo experimental study, neutrophil adhesion test, hemagglutinating antibody (HA) titre, delayed-type hypersensitivity (DTH) response, phagocytic activity and cyclopophamide-induced myelosuppression were demonstrated to be positively activated pointing towards a promise in immunomodulation [26]. |
| 3    | Kiratatikta (Swertia Chirata Pexbex. Karst.) | **1.** Inhibited the carrageenan-induced inflammation in the rat paw oedema model [45].<br>2. Stimulates the cell-mediated and humoral components of the immune system as well as phagocytosis in experimental animals [53,54].<br>3. Improves the immune system by increasing the proliferation of lymphocytes and cytokine levels (IL-4 and IFN-gamma) in serum, in HA titre, DTH, PFC, phagocytic index and CD4/CD8 population [55].<br>4. Another study demonstrates the antioxidant and free radical scavenging activity of the leaf extract of same plant [56].<br>5. Inhibits the growth of *Plasmodium falciparum* Significantly [57–59].<br>The drug shows promising effect due to its antiparasitic activity and antimalarial activity [58].<br>1. Anti-protozoal activity: A MEOH extract of *Swertia chirata* found to inhibit the catalytic activity of topoisomerase I of *Leishmania donovani* was subjected to fractionation to yield three secoiridoid glycosides: amarogentin (a), amaroswerin (b), and sweroside (c). Amarogentin is a potent inhibitor of type I DNA topoisomerase from *Leishmania* and exerts its effect by interaction with the enzyme, preventing binary complex formation. Other Studies: 1. Inhibits the expression of viral protein R (an attractive target for HIV disease) in HeLa cells harbouring the TREx plasmid encoding full-length Vpr (TREx-HeLa-Vpr cells) [60].<br>2. Inhibited HSV-1, plaque formation at more than 70% level and viral dissemination [61].<br>3. Suppressive effects on inflammatory mediators by blocking the expression of COX-2 and phosphorylation of Akt, IKK-β, MAPK and NF-κB, activation in LPS-stimulated macrophages [62].<br>4. A study on experimental arthritis in rats suggested amelioration of oxidative and inflammatory stress, which implies the immunomodulatory effect of leaves of the above said plant. Post treatment with the leaves of the said plant, the animal subjects showed marked reduction in inflammation as well as arthritic changes [63].<br>5. Protects against red blood cell (RBC) haemolysis and DNA damage [66].<br>4. Immuno-stimulatory: increase in hemagglutinating antibody titre and a change in delayed-type hypersensitivity [26].<br>5. Two weeks after challenge with *Pseudomonas aeruginosa*, the Caesalpinia treated animals showed a significant bacterial clearance from the lungs, with less severe incidence of lung abscess [67].<br>6. Exhibited activity against the vaccinia virus [68].<br>7. In vivo experimental study, neutrophil adhesion test, hemagglutinating antibody (HA) titre, delayed-type hypersensitivity (DTH) response, phagocytic activity and cyclopophamide-induced myelosuppression were demonstrated to be positively activated pointing towards a promise in immunomodulation [26]. |
| 4    | Kuberaaksha (Caesalpinia crista L.) | The CH2Cl2 extract from the seed kernels of *Caesalpinia crista*, which exhibited a promising antimalarial activity against *Plasmodium berghei* infected mice in vivo, which was examined and resulted in the isolation of seven new furanocassane-type diterpenes (*caesalpins A-C, 2-acetoxy-3-deacetoxycaesaldekarin e, caesalmin B, caesaldekarin e, caesalpin F, 14(17)-dehydrocaesalpin F, 2-acetoxycaesaldekarin e, 7-acetoxybouduelcin P, C, and caesalmin G)*. Their structures were determined on the basis of spectroscopic analysis. The isolated diterpenes showed significant dose-dependent inhibitory effects on Plasmodium falciparum FCR-3/72 growth in vitro. Their IC50 values ranged from 90 nM to 6.5 μM, and norcaesalpinin E (7) showed the most potent inhibitory activity (IC50, 90 nM).<br>2. Exhibits antimalarial [64] and hepatoprotective [65] activity. Other Studies: 3. Protection against red blood cell (RBC) haemolysis and DNA damage [66].<br>4. Immuno-stimulatory: increase in hemagglutinating antibody titre and a change in delayed-type hypersensitivity [26].<br>5. Two weeks after challenge with *Pseudomonas aeruginosa*, the Caesalpinia treated animals showed a significant bacterial clearance from the lungs, with less severe incidence of lung abscess [67].<br>6. Exhibited activity against the vaccinia virus [68].<br>7. In vivo experimental study, neutrophil adhesion test, hemagglutinating antibody (HA) titre, delayed-type hypersensitivity (DTH) response, phagocytic activity and cyclopophamide-induced myelosuppression were demonstrated to be positively activated pointing towards a promise in immunomodulation [26]. |

Table 3
Preclinical pharmacological and toxicological/safety studies of AYUSH-64 and its ingredients.

| No  | Preclinical pharmacological/safety studies                             |
|-----|---------------------------------------------------------------------|
| 1   | In albino mice, oral administration of AYUSH-64 at doses of 250–750 mg/kg for five days exhibited significant anti-malarial property [67]. The experimental studies of AYUSH-64 have shown that it was safe and non-toxic in a dose of 500 mg/kg of body weight for 12 weeks [57]. |
Table 4
Clinical studies of AYUSH-64.

| S.No | Clinical studies of AYUSH-64                                                                 |
|------|------------------------------------------------------------------------------------------|
| 1    | Clinically effective in cases of microfilariaus [20,21].                                  |
| 2    | 81% curative effect on Plasmodium vivax, drug is more effective in ring stage than in Gamete stage. In the cases of mixed infection of P. vivax and P. falciparum the curative effect was found to be 75% after longer therapy [69]. |
| 3    | Good effect in the management of microfilaraemia [70].                                    |
| 4    | In a study group of 4500 participants, AYUSH-64 was observed to be safe and nontoxic with good anti-malarial activity and also a major decline in the infectivity rate was noted by administering AYUSH-64 along with anti-mosquito measures, in various types of fevers [70]. |
| 5    | A double-blind study demonstrated comparative efficacy of AYUSH-64 to the chloroquine/primaquine as the standard modern control in sixty cases of P. vivax malaria [70]. |
| 6    | In a prospective, open-label, nonrandomized, single group, single-centre pilot study with pre-test and post-test design one-week intervention of ‘AYUSH-64’ in a dose of 3 g/day effectively helped to recover from Influenza Like Illnesses, and also the quicker return to normal life with reduced frequency of usage of acetaminophen/antihistaminic. No adverse effects were found during the study [71]. |
| 7    | A pilot study done on 112 participants produced 97.14% result with prophylactic treatment in P.F. Malaria [72]. |

lengths of 2.87, 2.37, 2.8, 2.98, 2.83, 2.89, and 2.85 respectively. In both the docking strategies, the grid box was generated around the 4Å region of N3 peptide in the Mpro of SARS-CoV-2 [15].

2.2. Ligand preparation

Selected 36 compounds of AYUSH-64 were obtained from the PubChem database, as 3D coordinates of structure-data file (SDF) format, and each compound was converted to PDBQT file format with the help of PyMol and ADT tool, generating an input compound/ligand file for docking study in AutoDock Vina. In BIOVIA Discovery studio, 225 maximum conformations were generated for selected 36 compounds, and separate isomer conformations were also created within the threshold of 20.0 kcal/mol relative energy. The CHARMM force field (fast and accurate) was applied to minimize the compounds with 1000 steps of steepest descent (SD) algorithm. The root-mean-square deviation (RMSD) with <1.0 Å were considered as duplicates, and the higher specified RMSD value will reduce the number of output ligand poses and RMS gradient 0.001 [15].

2.3. ADME and toxicity prediction

To predict the drug-likeness properties of selected 36 AYUSH-64 compounds, in silico absorption, distribution, metabolism, excretion and toxicity (ADME/T) prediction was carried out. We explored the ADME/T properties of the 36 AYUSH-64 selected compounds using the ADME/T Protocol in the Discovery Studio software package (Accelrys, San Diego, CA, USA). These examinations were exclusively founded on the substance structure of the particle. Some of the parameters that were calculated included, ADME 2 Dimensional Fast Polar Surface Area (ADME 2D FPSA), Atom-based Log P98 (ALogP98), Blood Brain Barrier (BBB), Cytochrome P4502D6 (CYP2D6), and Hepatotoxicity (HEPATOX).

2.4. Molecular docking

The prepared 36 AYUSH-64 compounds (ligands) were docked on an individual basis to the Mpro of SARS-CoV-2 with grid coordinates (grid center). The selected compounds were set flexible with conformation condition while interacting with the macromolecule of Mpro under rigid conditions of the protein. The prepared configuration file was run through ADT command line with PDBQT file as input with grid size of X; 25 × Y; 25 × Z; 25 points, grid centre set at X: 11.74, Y: 13.63 and Z: 70.63 dimensions having 1000 Å grid radius. By using AutoDock Vina scoring algorithm to measure Gibbs Free Energy (ΔG kcal/mol) for selected AYUSHES-64 compound binding affinities [29].

The LibDock protocol module of BIOVIA Discovery Studio was used for molecular docking studies of the 6LU7 and 36 AYUSH-64 compounds, the binding site is created with the same dimensions of the AutoDock Vina i.e. 4 Å radius of N3 peptide binding site. The LibDock protocol is a high-throughput algorithm developed by Diller and Merz, in 2001 [29], consisting the polar and a polar feature as “Hotspots.” Among the 36 compounds of AYUSH-64, A)

Fig. 1. Superimposition of the docked Mpro—N3 peptide with its X-ray crystal structure. A) Blue and yellow color indicates experimentally derived structure of Mpro—N3 peptide, the docked complex of orange — brown and green — yellow orange is derived from Discovery studio and Autodock Vina docking approaches respectively. B) 2D interaction of experimental and docked Mpro of SARS- CoV-2.
which were passed the applied filters while ligand preparation process, subjected to dock with the prepared 6LU7 crystal structure. The prepared compounds were docked to analyses and compare interacting amino acids. The post-docking analyses of two different docking strategies were visualized for H-bond, hydrophobic molecular interactions and bond distances with 4 Å radii in the Discovery Studio v2020 in the Mpro of SARS-CoV-2. The top ranked molecular docked complex selected for molecular dynamics simulations using Desmond v2019-4 for 100 nanoseconds (ns).

2.5. Molecular dynamics simulations

The first ranked docked molecular complex, AYUSH-64 with Mpro selected for carrying out the dynamic’s simulations using Desmond v2019-4 protocol. Where initially the selected molecular docked complex atoms calibrated by the “Optimized Potentials for Liquid Simulation (OPLS-2005)” force field. Further, geometric chemical structure corrections in molecular docked complex are performed using “Protein Preparation Wizard (macro model)”. To put the molecular docked complex in solvent environment, Simple Point charge-HOH model was used with the cubic box dimensions 10 Å × 10 Å × 10 Å, and for maintaining the acid-base balance for the molecular docked complex solvent, electrical ions NaCl were added with 0.15 M [15,30–32]. To model the maximum possible atomic bond energy (covalent and non-covalent) flexibility of molecular docked complex, hybrid algorithm “Steepest Descent and the limited-memory Broyden-Fletcher-Goldfarb-Shanno” is used. To create the initial minimized condition model of dynamic particles, pressure, and temperature of the molecular docked complex, “Berendsen thermostat and barostat NPT ensemble” method is used for 5 ns.

To assert constant temperature at 300 K and pressure of 1 atm throughout the molecular dynamics’ simulation process, “Nose-Hoover thermostat algorithm [33]” and “Martyna-Tobias-Klein Barostat algorithm [34]” is used respectively. To affirm coulombic long-range atomic interactions in the molecular docked complex throughout the molecular dynamic simulations process “Smooth Particle Mesh Ewald” method is used. To maintain the accuracy and tolerance of long-range interactions with the smaller value of 1e-9 is set for accurate computational implemented by SHAKE algorithm. The final production run was carried out for 100 ns, and the

![Docking Score](image)

**Fig. 2.** Molecular docking score of AYUSH-64 compounds. Among Akuammicine N-Oxide (green) has least docking score of -8.4 kcal/mol and N3 peptide (red) has docking score -4.1 kcal/mol.
trajectory sampling was done at an interval of 1.0 ps [15,30,31]. Subsequently, in the MD simulations study, the binding free energy ($\Delta G_{\text{bind}}$) for all simulated protein-ligand complexes were estimated by Desmond and Prime-molecular mechanics/generalized born surface area (MM-GBSA) method using thermal_mmgbbsa.py script embedded in Schrödinger suite.

2.6. Prime MM-GBSA

The MM-GBSA is a suite of protocols that combines OPLS-2005 molecular mechanics energies (EMM), a Surface Generalized Born (SGB) solvation model for polar solvation (GSGB), and a nonpolar solvation term (GNP), composed of the nonpolar solvent accessible surface area and vdW interactions. The binding energy calculations are much accurate than the docking score [35]. The binding energy is calculated by the following equations [36,37].

$$\Delta G_{\text{bind}} = \Delta E + \Delta G_{\text{solv}} + \Delta G_{\text{SA}}$$

Where

$$\Delta E = E_{\text{complex}} - E_{\text{protein}} - E_{\text{ligand}}$$

Where $E_{\text{complex}}, E_{\text{protein}}$, and $E_{\text{ligand}}$ are the minimized energies of the receptor–lead complex, receptor, and leads respectively

$$\Delta G_{\text{solv}} = G_{\text{solv}}(\text{complex}) - G_{\text{solv}}(\text{protein}) - G_{\text{solv}}(\text{ligand})$$

Where $G_{\text{solv}}$(complex), $G_{\text{solv}}$(protein), and $G_{\text{solv}}$(ligand) are the solvation free energies of the complex, protein, and inhibitor, respectively

$$\Delta G_{\text{SA}} = G_{\text{SA}}(\text{complex}) - G_{\text{SA}}(\text{protein}) - G_{\text{SA}}(\text{ligand})$$

Where $G_{\text{SA}}$(complex), $G_{\text{SA}}$(protein), and $G_{\text{SA}}$(ligand) are the surface area energies for the complex, protein and inhibitor, respectively.

The simulations were carried out using the GBSA continuum model in Prime [38]. The Prime uses a Surface Generalized Born (SGB) model employing a Gaussian surface instead of a vdW surface for better representation of the solvent-accessible surface area [35,39]. The binding free energy can be computed for all the trajectories or a set of frames within generated frames from MD simulations. Considering all frames, perfect (or imperfect) poses might have a crucial contribution in the binding free energy calculation, hence, each and every frame of the entire MD simulated trajectories were used, that is highly rigorous and more accurate approach to validate the docking and dynamics approaches in the study for calculating the binding free energy. Therefore, in the present study, all MD trajectory frames from start to end (~1000 frames) were selected to estimate the $\Delta G_{\text{bind}}$ for proposed and native MD simulated complex.

3. Results

3.1. Validation of docking protocol

Preferably, the distinguishing proof of the ideal docking and scoring blend will diminish the number of false positives and false negatives while ensuring optimal hit rates. It has accounted for enormous strategies for validating docking approach and scoring functions. Utilized strategy through pose selection whereby docking programs are exploited to re-dock into the target’s active/binding site with a known conformation and orientation, ordinarily from known co-crystal structure. The approaches that can restore poses under a pre-selected RMSD esteem from the known compliance/orientation (1.5 or 2 Å relying upon ligand size) are considered having performed effectively (Fig. 1). Pose selection is then trailed by scoring and positioning to contemplate which of the accessible scoring capacities most precisely positions the stances regarding their RMSD esteems. In the present study, the different docking protocols of Discovery studio and AutoDock protocols are validated through, re-docked with known co-crystal structure of 6LU7 with N3 peptide (Fig. 1). The re-docked results of RMSD revealed that, the AutoDock vina determined with 1.25A and Discovery studio has 1.02Å. Also, 2D atomic coordination of the re-docked complexes was supplanted experimentally derived structure. Thus, these docking protocols was viewed good enough for replicating the docking results similar to co-crystal structure and consequently can be applied for further molecular docking analysis.

3.2. Molecular interactions of M$^{\text{pro}}$ with AYUSH-64

The SARS-CoV-2, M$^{\text{pro}}$ plays a significant role in RNA translation and is essential for viral replication. The three-dimensional structure of M$^{\text{pro}}$(6LU7) is a monomer structure, consisting three major structural domains: domain I (residues 8–101), domain II (residues 102–184), which contribute an antiparallel β-barrel structure, and domain III (residues 201–303), which contains a five-fold anti-parallel α-helix cluster and is attached to long loop (residues 185 to 200) of domain II [6]. The amino acid of cysteine–Histidine dyad plays a significant role in the maturation cleavage, which is located in the cleft between domains I and II the precursor M$^{\text{pro}}$ [6]. Moreover, M$^{\text{pro}}$ was non-homologous to human and it is an ideal anti-viral drug target for SARS-CoV-2 [15]. The co-crystal ligand of N3 peptide is located in the cleft region.

**Fig. 3.** A) Molecular interactions of Akuammicine N-Oxide with 2019-nCoV of M$^{\text{pro}}$. B) 2D interaction of docked M$^{\text{pro}}$ of SARS-CoV-2 with Akuammicine N-Oxide.
between domain I and II. The SARS-CoV-2 MPro shares highly (~99%) similarity and identity (100%) with binding site residues of SARS-CoV MPro [15].

In the present study, the selected 36 compounds of AYUSH-64 were substituted (shows a similar inhibitor-binding mode) within the cleft between domain I and II, found to be having good docking score than the native substrate of N3 peptide (Supplementary Table 1 and Fig. 1A). ADME/T properties prediction results are illustrated in Supplementary Table 2 for drug-likeness. Two docking strategies, Akuammicine N-Oxide compound has best docking score, binding affinity and good ADME/T properties than the other selected 35 AYUSH-64 compounds and reference substrate of N3 peptide (Supplementary Table 3 and Fig. 2). The side chain residues, His41 (2), Met49 (2), Tyr54, Asn 142 (2), Gly 143, Ser144, Cys145 (2), His163, Met165, and His172 formed 14 H-bonds with Akuammicine N-Oxide within 4 Å region of MPro (Fig. 3). Along the side, Cys44, Pro52, Phe140, Leu141, Asn142, Arg188, Asp187, Gln189 were formed vdW interactions to the Akuammicine N-Oxide within 4 Å region in both docking strategies (Supplementary Table 2 and Fig. 3B). To summarize, Akuammicine N-Oxide compound reveal a similar inhibitory mechanism to occupy the substrate-binding pocket and block the enzyme activity of SARS-CoV-2 MPro.

Further, the docked complex of MPro– Akuammicine N-Oxide and native MPro– N3 peptide complex was subjected to perform molecular dynamics simulations to check the stability at 100ns employing Desmond 2019-4.

3.3. Molecular dynamics simulations

For understanding the conformational deviations/fluctuations and binding free energy of docked and native complexes, 100 ns period of molecular dynamics simulations was performed for each trajectory. The MPro– Akuammicine N-Oxide and native N3 peptide complex depicted the total energy average of −9.2271.69 kcal/mol, potential energy of −114792.08 and −121645.95 kcal/mol, was relatively stable with the average temperature 298.67 K, pressure -0.822 bar, and volume ~366704.2 Å³ throughout the simulations run (Fig. 4). The post MD simulations, each trajectory was analyzed for stability conformation through RMSD, root mean square fluctuation (RMSF), H-bond monitoring and ΔG binding energy (Figs. 5–7 and Supplementary Figs. 1–3).

Fig. 4. The stable total energy, potential energy, temperature, pressure and volume of molecular dynamics simulations at 100 ns period of 2019-nCoV MPro with Akuammicine N-Oxide(A) and N3 peptide (B).
The average RMSD value observed for \( M^{\text{pro}} \) Ca atoms is 1.4 Å and for proposed Akuammicine N-Oxide is 1.17 Å, which was found stable throughout the simulation period (Fig. 5A). In the native complex of \( M^{\text{pro}} \) – N3 peptide, average RMSD values 9.5 Å and 2.05 Å were observed for the Ca atoms and maximum RMSD of \( M^{\text{pro}} \) Ca atoms is 14.12 Å, that constitutes significantly not stable during MD simulations (Fig. 5B). To measure the atomic fluctuations in \( M^{\text{pro}} \) residues, RMSF values were analysed for each residue Ca atoms bound with side-chain of docked and native complex of \( M^{\text{pro}} \) of SARS-CoV-2. The average RMSF observed for each Ca atoms: 1.4 Å and side chain: 2.5 Å in the overall simulation run docked with Akuammicine N-Oxide (green lines) (Fig. 6A). In the native N3 peptide (green lines) with \( M^{\text{pro}} \) complex observed in the 1000 frames of MD simulations where the results revealed with average RMSF values for Ca and side-chain have 3.77 Å and 4.52 Å respectively (Fig. 6B).

Notably, post dynamics simulation of the \( M^{\text{pro}} \) with Akuammicine N-Oxide and native N3 peptide, formed significant H-bonds, hydrophobic and water associated bridges correlating with co-crystal structure (Fig. 7). The Akuammicine N-Oxide formed seven H-bonds in \( M^{\text{pro}} \) with Ser46 (~7%), Asn142 (~7%), His164 (~27%), Glu166 (~3%), Val186 (~30%), Arg188 (~4%) and Gln189 (~22%); three hydrophobic interactions with Leu27 (~30%), Met49 (~31%) and Cys145 (~39%); and more than 10% of water associated bridges with Asn142 (~18%), Glu166 (~39%) and Gln189 (~12%) within 4Å region of binding site (Fig. 7A). The co-crystal N3 peptide molecular interaction was substituted with the above furnished residue interactions of Akuammicine N-Oxide with \( M^{\text{pro}} \) of SARS-COV-2.

While checking the H-bond occupancies over the course of MD simulation trajectories, Phe140 (~28%), Leu141 (~28%), Gly143 (~92%), Ser144 (~15%), His163 (~78%), and His164 (~97%) was observed with native N3 peptide. In another instance, residue Phe140 (~90%), Glu166 (~97%) and His172 (~94%) constructed \( M^{\text{pro}} \) – N3 peptide contacts via ionic bond occupancies with 1000 frames. Moreover, \( M^{\text{pro}} \) residues has constituted the hydrophobic: Val3 (~10%), Met49 (~10%), Cys145 (~97%), and water bridges: Leu4 (~10%) Thr26 (~10%) with native N3 peptide.

From the comparative MM-GBSA plots between docked \( M^{\text{pro}} \) – Akuammicine N-Oxide and native \( M^{\text{pro}} \) – N3 peptide complexes, in total of 1000 frames obtained from 100 ns MD simulations were considered for binding free energy calculations (\( \Delta G_{\text{bind}} \)) (Supplementary Fig. 1). The docked complex illustrated the stable
average binding free energy of $-44.57$ kcal/mol (Supplementary Fig. 1A) than fluctuated native complex with average binding free energy of $-38.47$ kcal/mol (Supplementary Fig. 1B). The stable $\Delta G_{\text{bind}}$ values obtained from docked complex is self explanatory that, the proposed $\text{Mpro - Akuammicine N-Oxide}$ exhibited significantly stable and higher binding free energy in comparison with the native complex during 100 ns of MD simulations.

While $\text{Mpro}$ of secondary structure elements (SSE) contributing to the overall protein stability was examined and noticed that, the protein was maintained an average of $\sim42\%$ SSE, mostly composed of $\sim18.08\%$ helices and $\sim24.14\%$ of strand during 100ns simulation period. Remarkably, the Akuammicine N-Oxide also showed the substantial ligand properties of Radius of Gyration ($R_g$: $\sim4.0$), Molecular Surface Area ($\text{MolSA}$: $\sim363.37$), Solvent Accessible Surface Area ($\text{SASA}$: $\sim209.8$) and Polar Surface Area ($\text{PSA}$: $\sim138.47$) (Supplementary Fig. 2). Significantly, one intra-molecular interaction was observed during the simulation time of 100 ns period. Moreover, every rotatable bond of the Akuammicine N-Oxide formed H-bonds with $\text{Mpro}$ was stable throughout the simulation trajectory (Supplementary Fig. 3). To summarize from all the observed findings of molecular docking and dynamics simulations, $\text{Mpro - Akuammicine N-Oxide}$ complex was highly stable and shown significant molecular interactions when compared with native $\text{Mpro - N3 peptide}$ complex of SARS-COV-2.

4. Discussion

$\text{Mpro}$ is one of the best-characterized pan drug target for the coronavirus's family. Along with $\text{Mpro}$ an essential enzyme for the endurance of virus, processing the polyproteins which are
translated from the viral RNA [13–15]. During translation, the Mpro exclusively cleaves at all 11 cleavage sites (after a glutamine residue) of poly peptide sequences on the large poly protein named as 1 ab (replicase 1-ab, ~790 kDa); Inhibiting the activity of this enzyme would block the viral replication. The cleavage site recognition sequence at most sites is Leu-Gln (Ser, Ala, Gly). To the best of our knowledge no human proteases with a similar cleavage site specificity are known, such inhibitors are unlikely to be toxic [13–15].

In this present study, we performed different docking strategies of Discovery studio and Auto Dock Vina, Mpro of 6LU7 with 36 compounds of AYUSH-64. The results revealed that, 35 compounds have a good binding affinity and strong H-bond than the native N3 peptide bound with the Mpro of SASR-CoV-2 binding site (Supplementary Table 1 and Fig. 2). The top scored docking pose compound, Akuammicine N-Oxide formed fourteen H-bond with Mpro side chain residues along number bonds mentioned within brackets, His41 (2), Met49 (2), Tyr54, Asn142 (2), Gly143, Ser144, Cys145 (2), His163, Met165, and His172 of Mpro significant binding site residues (Fig. 3). The Akuammicine N-Oxide formed vdW interactions with the binding site residues of Cys44, Pro52, Phe140, Leu141, Asn142, Arg188, Asp187, and Gln189 within 4 Å region in both docking strategies (Supplementary Table 2 and Fig. 3B). Thus, the molecular docking approaches utile to identify the novel hits/leads against several pathogenesis. Docking aims to reproduce a conformational selection specific to target conformations that are recognized by the hit/lead to form molecular-interactions in the form of thermodynamically favored complex. The MD simulations were successfully used to achieve the major objective of substantially stabilize the Mpro quaternary structure which involves an increase of the inter-molecular interactions by strengthening the ionic contacts and structural flexibility of the Mpro binding pocket to allow flexible exploration of its conformational space, thus extracting representative frame from docked complexes.

To investigate the top scored docking pose complex (Mpro – Akuammicine N-Oxide) binding conformational stability with SARS-CoV-2 Mpro, we performed 100 ns MD simulations using Desmond. The MD simulations results revealed, Akuammicine N-Oxide formed seven stable H-bonds with Ser46, Asn142, His164, Glu166, Val186, Arg188 and Gln189; three hydrophobic interactions

Fig. 7. Post molecular dynamics simulations of (A) Akuammicine N-Oxide and (B) N3 peptide were formed interactions with Mpro of SARS-CoV-2.
with Leu27, Met49 and Cys145; and water associated bridges with Asn142, Glu166 and Gln189 within 4A region of binding site (Fig. 7A). The co-crystal N3 peptide molecular interaction was substituted with Akuammicine N-Oxide in MPro of SARS-CoV-2. Furthermore, advanced MD simulations of MM-GBSA analysis (total frames obtained from 100 ns run) also confirmed the proposed MPro—Akuammicine N-Oxide complex has consistent binding free energy (ΔGbind) score when compared to native complex. This study from sequence levels to advanced structural dynamics confirmed, Akuammicine N-Oxide of AYUSH-64 have potential inhibitor against SARS-CoV-2. Superimposition of experimentally derived X-ray crystal structure of the MPro of MERS-CoV, SARS-CoV, and SARS-CoV-2, shares structural similarity and high conservativeness of substrate binding site. These observances are enlight-ened; Akuammicine N-Oxide was a potential pan-coronavirus inhibitor against corona family. Although these results were obtained from in silico approaches, the formation of these interactions provides insight to design and repurposing of novel drugs that might enlighten scientific community and researchers for further immune-modulatory investigations in human. Further, it is pertinent to note that AYUSH-64 is a compound herbal preparation, which is already tested to some extent for its safety and efficacy for treating malaria, joint disorders and influenza like illness. The currents in silico study provide a valid basis for its repurposing in the management of COVID-19.

5. Conclusion

This study revealed that, several compounds from an Ayurvedic drug are acting as potent inhibitors of SARS-CoV-2 MPro enzyme. Assessing the individual compounds through two different molecular docking strategies and extensive molecular dynamics simulations, explains that the Akuammicine N-Oxide is tightly bound (H-bond) with significant residues Cys145 and His164 of MPro, also found stable during pre- and post-dynamics simulations. The results indicate that AYUSH-64, an approved and safe drug for managing joint pains, fever, influenza like illnesses make it a good candidate for repurposing against COVID-19. Therefore, AYUSH-64 might be validated through further experimental, clinical studies that entailed to confirm the anti-protease properties against COVID-19.

Source(s) of Funding

None.

Conflicts of Interest

None.

Acknowledgments

The authors are highly thankful to the Indian Council of Medical Research (ICMR), New Delhi, Government of India and Central Council for Research in Ayurvedic Sciences, Ministry of AYUSH, Government of India, for providing facilities, and resources. We are also thankful to Dr. Kanipakam Hema and Dr. Dinesh Gupta from Translational Bioinformatics Group, International Centre for Engineering and Biotechnology, New Delhi, for providing Schrodinger Suite to perform MM-GBSA analysis. The efforts of Dr G Shruthi, SRF (AYU), SNOMED CT Project, NIIMH, Hyderabad and Sribendi were highly appreciated for proof reading and editing the manuscript. MM highly thankful to TATA trust for providing facilities at the NIN-TATA Centre of Excellence in Public Health and Nutrition, ICMR-National Institute of Nutrition, Hyderabad, Telangana State.

Appendix A. Supplementary data

Supplementary material associated with this article can be found online at https://doi.org/10.1016/j.jaim.2021.02.004.

References

[1] Rabhi FA, Al Zoubi MS, Al-Nasser AD, Kasasbeh GA, Salameh DM. Sars-cov-2 and coronavirus disease 2019: what we know so far. Pathogens 2020;9. https://doi.org/10.3390/pathogens9030231.
[2] Wu F, Zhao S, Yu B, Chen YM, Wang W, Song ZG, et al. A new coronavirus associated with human respiratory disease in China. Nature 2020;579:265–9. https://doi.org/10.1038/s41586-020-02088-3.
[3] Zhou P, Lou Yang X, Wang XG, Hu B, Zhang L, Zhang W, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature 2020;579:270–3. https://doi.org/10.1038/s41586-020-0212-7.
[4] Lu R, Zhao X, Li J, Niu P, Yang B, Wu H, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. Lancet 2020;395:565–74. https://doi.org/10.1016/S0140-6736(20)30251-9.
[5] Anand K, Ziebuhr J, Wadhwani P, Menters JR, Hilgenfeld R. Coronavirus main protease (MPro) Structure: basis for design of anti-SARS drugs. Science (80- ) 2003;300:1763–7. https://doi.org/10.1126/science.1085658.
[6] Jin Z, Du X, Xu Y, Deng F, Liu M, Zhao Y, et al. Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors. Nature 2020;582:289–93. https://doi.org/10.1038/s41586-020-2223-y.
[7] Li CC, Wang XJ, Wang HCR.Repurposing host-based therapeuticsto control coronavirus and influenza virus, Drug Discov Today 2019;24:726–35. https://doi.org/10.1016/j.judr.2019.01.018.
[8] Liu AL, Du GH. Antiviral properties of phytochemicals. Diet. Phytochem. Microc. Springer Netherlands; 2012. p. 93–126. https://doi.org/10.1007/978-94-007-3926-0_3.
[9] Newman DJ, Craig GM. Natural products as sources of new drugs over the last 25 years. J Nat Prod 2007;70:461–77. https://doi.org/10.1021/np060054v.
[10] Darabian-Ganjame M, Abbaspour A. Chloroquine/hydroxychloroquine be harmful in coronavirus disease 2019 (COVID-19) treatment? Clin Infect Dis 2020;71:888–9. https://doi.org/10.1093/cid/ciaa321.
[11] Ennozhi SK, Raja K, Sebastine I, Joseph J. Andrographolide as a potential inhibitor of SARS-CoV-2 main protease: an in silico approach. J Biomol Struct Dyn 2020:1–10. https://doi.org/10.1080/07391102.2020.1760136.
[12] Khaerunnisa S, Kurnawahan H, Awaluddin R, Suhartati S. Potential inhibitor of COVID-19 main protease (Mpro) from several medicinal plant compounds by molecular docking study. Preprints 2020:1–14. https://doi.org/10.20944/preprints202003.0226v1.
[13] Khan SA, Zia K, Ashraf S, Uddin R, Ul-Haq Z. Identification of chymotrypsin-like protease inhibitors of SARS-CoV-2 via integrated computational approach. J Biomol Struct Dyn 2020;13:1–10. https://doi.org/10.1080/07391102.2020.1751298.
[14] Khan RJ, Jha RK, Amera GM, Jain M, Singh E, Pathak A, et al. Remdesivir (GS-5734) as a therapeutic option of 2019-nCOV main protease - in silico approach. J Biomol Struct Dyn 2020;14:1–14. https://doi.org/10.1080/07391102.2020.1753577.
[15] Naik VR, Munikumar M, Ramakrishna U, Srujana M, Goudar G, Naresh P, et al. Remdesivir (GS-5734) as a therapeutic option of 2019-nCOV main protease - in silico approach. J Biomol Struct Dyn 2020;14:1–14. https://doi.org/10.1080/07391102.2020.1781694.
[16] Tahir uQamar M, Alqahtani SM, Alami MA, Chen LL. Structural basis of SARS-CoV-2 3CLpro and anti-COVID-19 drug discovery from medicinal plants. J Pharm Anal 2020;10(4):313–9. https://doi.org/10.1016/j.jpha.2020.03.005.
[17] Bhardwaj VK, Singh R, Sharma J, Rajendran V, Purohit R, Kumar S. Identification of bioactive molecules from tea plant as SARS-CoV-2 main protease inhibitors. J Biomol Struct Dyn 2020;1–10. https://doi.org/10.1080/07391102.2020.1766572.
[18] Gurung AB, Ali MA, Lee J, Farah MA, Al-Anazi KM. Unravelling lead antiviral phytochemicals for the inhibition of SARS-CoV-2 Mpro enzyme through in silico approach. Life Sci 2020;255:117831. https://doi.org/10.1016/j.lfs.2020.117831.
[19] CCRAS. Ayurved-64 A new ayurvedic anti-malarial compound. 1st ed. New Delhi: Central Council for Research in Ayurveda and Siddha; 1987.
[20] Pandey PN, Kishore P. Effect of ayush-64 and saptaparnaghana vati on microfilaraemia. J Res Ayurveda Siddha 1991;12:145–50.
[21] Pandey PN, Patdi MM, PK. An epidemiological survey on microfilaraemia in the villages around hubaneswar with therapeutic effect of ayush-64. J Res Ayurveda Siddha 1991;12:33–40.
[22] Chari MV, Venkatagabavan S, Seshadri C, Ramakrishna B, Shetty G. A double blind clinical trial with ayush-64 an ayurvedic drug in p. vivax malaria. J Res Ayurveda Siddha 1985;6:105–16.
