Full Length Article

Isolation, structure characterization and prediction of antioxidant activity of two new compounds from the leaves of *Dodonaea viscosa* native to the Sultanate of Oman

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**Abstract**

Traditionally, *Dodonaea viscosa* (*D. viscosa*) is used worldwide for the treatment of different chronic diseases. The current investigation involves the isolation, structure characterization and prediction of antioxidant compounds from the leaves of *D. viscosa*, which was collected from AL-Hamra, Sultanate of Oman. The leaves samples were used for the extraction of plant constituents with methanol by using hot extraction method. The methanol solvent was evaporated by usual method and the crude extract was dissolved in water and fractionation successively with different polarity solvents to give hexane, chloroform, ethyl acetate, butanol and water extracts, respectively. The antioxidant activity of different polarity crude extracts and isolated pure compounds was determined by modified 1,1-diphenyl-2-picryl-hydrazyl (DPPH) method. The highest antioxidant activity was found in hexane extract and the lowest antioxidant activity was found in butanol extract and followed by hexane > chloroform > ethyl acetate > methanol > water > butanol extract. The hexane and ethyl acetate crude extracts were used for the isolation and separation of antioxidant compounds by using different chromatographic techniques. The isolated pure compounds were characterized as 3,3',4,5,7-pentahydroxyflavane (1) and 4-methoxylstigmasterol (2) by MS, $^1$H NMR, $^{13}$C NMR, and 2D NMR. Both the compounds, 3,3',4,5,7-penta hydroxyflavane (1) and 4-methoxylstigmasterol (2) were isolated for the first time from the selected Omani plant species. The pure compounds obtained from hexane and ethyl acetate extracts of the selected plant species could be a good source of natural antioxidant.

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**1. Introduction**

Plants are considered as essential source of medicines for humans. Plant parts like leaves, flowers, roots, stems, seeds, and fruit are used as food resources for human as well as safe medicine for the treatment of different diseases. Plant-derived herbal medicine is used as main sources for the treatment of diseases since the ancient time [1]. Antioxidants are the most important chemical components that may prevent or delay different types of cell damage. Foods, fruits, and vegetables are the main sources of natural antioxidants. The prevention or recovery of cell damage in the human body could be used foods, fruits, and vegetables as natural antioxidants. The main mechanism of antioxidants is to improve the immune system in the human body by producing the free radical. Some of natural antioxidants are also available as dietary supplements. Recently, vegetables and fruits are considered as rich sources of natural antioxidants. Several recent scientific evidences are available that eating lots of vegetables and fruits are very good sources for human health and to prevent the cell damage. Therefore, the risks of certain chronic or incurable diseases are lowered due to natural antioxidants [2].

*Dodonaea viscosa* (*D. viscosa*) is an evergreen woody plant belongs to the family Sapindaceae [3]. The selected plant species originates from Australia and recently it is available throughout the tropical and subtropical countries. The selected plant is mostly available in certain parts of Australia, Africa, Mexico, New Zealand, India, Northern Mariana Islands, Virgin Islands, Florida, Arizona, South America, and elsewhere. *D. viscosa* is a dioecious or monoecious multi stems shrub small tree up to 7 m tall (Fig. 1). The bark is blackish, thin and exfoliating in long thin strips. Several varieties are available all over the world, including Oman. It is one of the rarest medicinal plant species indigenous to Al Jabal Al Akhdar, Northern Oman, Al Hamra and Salalah [1]. Its Arabic name is Zaitoon Alramal and locally known as Shahs [4]. It possesses...
medicinal values promoted by the plant being easy to harvest and blend into a variety of therapeutic preparations [1,2].

D. viscosa is usually administered by local communities of several regions for the administration as poultice for treating a variety of diseases [2,5]. The traditional use of this plant covers a significant range of conditions, including pain relief in dental cases and headache, chronic inflammatory disorders, rheumatoid arthritis, fever, cold, gastrointestinal system disorders and female reproductive conditions [6–8]. In Saudi Arabia, the mixture of leaves, steam and seed with honey is used to treat malaria [7,9].

Recently, D. viscosa extends to infectious agents where it was found to be antiviral and antimicrobial activities. In addition to its special use as a spasmolytic and hypotensive agent. Regarding medicinal effects of the plant parts, the leaves were reported to possess bioactive elements resembling the activity of local anesthetic, smooth muscle relaxant, antimicrobial, anti-inflammatory and anti-ulcerogenic agents [1,10]. However, lack of literature on medicinal properties of the selected plant indicates that no work has been done on the Omani species. On the basis of medical importance of the selected plant, now, it is commercially cultivated all over the world including Sultanate of Oman. Therefore, it is necessary to evaluate the biological activities of D. viscosa, which is abundantly available in Oman mainly in Al Jabal Al Akhdar and Al Hamra. The present investigation involves to prepare different crude extracts from the leaves of Omani D. viscosa species, and to isolate and characterize antioxidant compounds from crude extract having the highest antioxidant activity and to determine their antioxidant activity.

2. Materials and methods

2.1. Chemicals and reagents

The solvents like hexane, ethyl acetate, butanol, chloroform, and acetone were used in this experiment obtained from Sigma Aldrich Company, Germany. Methanol, silica gel, preparative TLC, gallic acid and 1,1-diphenyl-2-picrylhydrazyl (DPPH) were obtained from BDH, UK. In addition, other chemicals were used analytical grade. All glassware used in this experiment was from borosil, India.

2.2. Instruments

The absorbance of various polarities extracts and isolated pure compounds at different concentration was measured by UV–visible spectroscopy (Shimadzu, Model 1800, Japan) for the determination of antioxidant activity. The MS spectra were performed using a Varian GC–MS/MS (Model Varian CP 3800) equipped with a VF-5 fused silica capillary column (30 m × 0.25 i. d., film thickness 0.25 μm). Injector and mass transfer line temperature were set at 250 and 300 °C, respectively. 1H and 13C NMR spectra were measured on a Brucker (600 MHz) spectrometer with TMS as the internal standard. DMSO was used as solvent and chemical shifts values were given in δ (ppm).

2.3. Sample collections

The leaves of D. viscosa sample were collected from AL-Hamra in the month of September 2016 at 10 am and identified at the Biological Science Division, University of Nizwa, Oman. The leaves of the selected plant were identified by local people and confirm by the website. The collected leaves sample was transported to the Research Laboratory, University of Nizwa, Oman for cleaning and the healthy clean leaves were separated from the stems and dried under the shade for 7 days.

2.4. Extraction

The dried leaf samples (500 g) were coarse into powder by blender machine and the coarse powder samples (200 g) were extracted with methanol (300 ml) by using a hot extraction method for 3 days. After 3 days extraction, the solvent was evaporated from the extract by using rotary evaporator at under reduced pressure at ambient temperature. Then, the solvent free extract (10 g) was dissolved in water (200 ml) and fractionated with n-hexane, chloroform, ethyl acetate and n-butanol [3] with increasing polarity. The four fractions such as n-hexane, chloroform, ethyl acetate, n-butanol and the remaining water fraction were dried separately by using a rotary evaporator to give n-hexane (1.30 g), chloroform (2.81 g), ethyl acetate (1.82 g), n-butanol (1.09 g) and water extract (1.64 g), respectively. All prepared crude extracts from the leaves powder of D. viscosa were used for the determination of their antioxidant activity.

2.5. Antioxidant activity

The antioxidant activity of different crude extracts and isolated pure compound of the selected plant was estimated by 1,1-diphenyl-2-picrylhydrazyl (DPPH) method with modification [11]. Various concentrations (12.5, 25, 50, 100 and 200 μg/ml) of each crude extract and pure compound were used for the determination of antioxidant activity. Each concentration of each crude extract (4 ml) was placed in a clean test tube and DPPH solution (1 ml) was added to the same test tube. The mixture was shaken vigorously by hand and kept in a dark place at ambient temperature for 45 min. DPPH and methanol solvent were used as blank samples. The absorbance of all working samples was measured at a fixed wavelength 517 nm by using a UV–visible spectrophotometer [11]. The percentage of inhibition of each concentration plant crude extract was calculated by using the following formula:

\[
\%\text{Inhibition} = \frac{A_{\text{control}} - A_{\text{extract}}}{A_{\text{control}}} \times 100
\]

2.6. Isolation and characterization antioxidant compounds

The highest antioxidant hexane extract (1.20 g) and ethyl acetate crude extract (1.50 g) was used for the separation of antioxidant compounds by using column chromatography with different organic solvents as a mobile phase. According to the same Rf values, the isolated fractions from the column chromatography were combined together and evaporated the mother solvent at ambient temperature. Based on the TLC, these similar TLC pattern eluents were combined together to give Fraction 1, Fraction 2, Fraction 3, Fraction 4, Fraction 5 and Fraction 6. Similarly, the same
chromatographic techniques were used to separate the ethyl acetate crude extract by using different polar solvents. According to the TLC, four fractions (Fraction 1, Fraction 2, Fraction 3, and Fraction 4) were obtained from the ethyl acetate extract. All fractions were evaporated at ambient temperature inside the fume hood. The isolated fraction was again purified by preparative thin layer chromatography (PTLC) to give the pure compounds. The structures of purified compounds from both crude extracts were elucidated by using MS and 1D and 2D NMR.

2.7. Fraction 1

The obtained fraction 1 from ethyl acetate crude was further purified by PTLC over silica gel 60 G using hexane-ethyl acetate (1:1) as developing solvent. Fraction 1 gave one major compound with several minor compounds. The separated major compound was crystallized from ethyl acetate-chloroform to give yellowish power (5.4 mg), Rf 0.44 (hexane-ethyl acetate: 1:1); (M+, 291); 1H NMR (600 MHz, DMSO-d6): 2.44 (m, 1H, H-4b), 2.64 (dd, 1H, J = 4.38 and 4.5 Hz, H-4a), 3.98 (d, 1H, J = 2.76 Hz, H-3), 4.63 (d, 1H, J = 4.38 Hz, H-2), 4.71 (s, 1H, -OH), 5.69 (d, 1H, J = 2.16 Hz, H-6), 5.87 (d, 1H, J = 2.22 Hz, H-8), 6.87 (s, 1H, H-2'), 6.62 (m, 2H, H-5' and H-6'), 8.70 (s, 1H, -OH), 8.77 (s, 1H, -OH), 8.87 (s, 1H, -OH), 9.08 (s, 1H, -OH); 13C NMR (600 MHz, DMSO-d6): 28.18 (C-4), 64.88 (C-3), 78.03 (C-2), 94.04 (C-8), 95.02 (C-6), 98.45 (C-10), 114.72 (C-2'), 114.86 (C-5'), 117.91 (C-6'), 130.58 (C-1'), 144.41 (C-4'), 144.47 (C-3'), 155.75 (C-5), 156.20 (C-9), 156.50 (C-7). On the basis of 1H and 13C and MS spectral data it was characterized as 3,3,4,5,7-pentahydroxyflavane (1).

2.8. Fraction 2

Fraction 2 obtained from hexane extract by column chromatography was repeated column chromatography to give one major and three minor fractions (A-D). The major fraction C was further purified by preparative thin layer chromatography (PTLC) to give two compounds (1 and 2). The compound 1 with high amount was collected from the preparative TLC by spatula. The compound 1 was eluted from the silica gel by methanol. The amount of purified compound 1 was 13 mg by weight. Finally, the compound 1 was crystallized from dichloromethane to give a white crystals (5 mg), Rf 0.64 (hexane-ethyl acetate: 7:2); (M+, 442); 1H NMR (600 MHz, CDCl3): δ 0.77 (d, 3H, J = 2.04 Hz, CH3-27), 0.81 (d, 3H, J = 1.5 Hz, CH3-26), 0.82 (d, 3H, J = 7.4 Hz, CH3-25), 0.84 (d, 3H, J = 4.8 Hz, CH3-21), 0.93 and 0.96 (2s, 6H, CH3-19 and CH3-18), 0.88 (m, 2H, H-1), 1.00 (m, 4H, H-15 and H-16), 1.02 (s, 2H, H-2), 1.26 (m, 4H, H-11 and H-12), 2.01 (s, 1H, H-24), 4.43 (m, 1H, H-3), 4.04 (s, 3H, -OCH3), 5.00 (s, 1H, H-8), 5.05 (s, 1H, H-24), 5.09 (m, 2H, H-22 and H-23), 5.00 (s, 1H, H-23), 5.11 (m, 1H, H-6); 13C NMR (600 MHz, CDCl3): δ 140.6 (C-5), 138.4 (C-22), 129.1 (C-23), 121.8 (C-6), 71.9 (C-3), 56.7 (C-17), 56.9 (C-14), 50.9 (C-9), 50.7 (C-24), 42.6 (C-13, 4), 39.6 (C-12), 37.4 (C-1), 40.2 (C-20), 36.7 (C-10), 31.4 (C-8, 7), 31.7 (C-2), 30.9 (C-23), 29.23 (C-OCCH3), 28.8 (C-16), 24.8 (C-15), 24.7 (C-28), 21.5 (C-11), 20.8 (C-26), 20.4 (C-19), 19.7 (C-27), 19.1 (C-21). On the basis of 1H and 13C and MS spectral data, it was characterized as 4-methoxyxstigmastanol (1).

3. Results

The methanol solvent was used for the preparation of crude extract from the leaves of D. viscosa, which was collected from Al Hamra. After extraction, the methanol solvent was evaporated by rotary evaporator at ambient temperature and then defatted with water and fractionation with different polarity of solvent to give hexane; chloroform, ethyl acetate, butanol, and water extract (Table 1).

3.1. Antioxidant activity

The antioxidant activity of different crude extracts of leaves of D. viscosa was evaluated by DPPH method with modification (Aziza et al., 2015). The highest antioxidant activity was found in hexane extract with percentage of inhibition value of 94.41 ± 0.12 and the lowest activity of found in butanol extract with percentage of inhibition value of 78.57 ± 0.08 (Table 2). The other crude extracts of the selected plant species also showed significant inhibition compare to standard gallic acid.

3.2. Isolation and characterization antioxidant compounds

The high antioxidant hexane and ethyl acetate crude extract was used for the isolation and separation of antioxidant compounds by different chromatographic system with different polar and non polar organic solvents. According to the similar Rf values, the separated fractions were combined together. The pure compounds were purified from the fraction 2 of hexane extract and fraction 1 of ethyl acetate extract which was obtained from column chromatography. The major compounds from both fractions were purified again by preparative thin layer chromatography (PTLC) to give pure compounds 1 and compound 2 (Fig. 2). The structures of the purified compound 1 and compound 2 were elucidated by using MS and NMR.

3.3. Antioxidant activity of pure compounds

The antioxidant activity of the isolated pure compound 1 and compound 2 at different concentrations obtained from ethyl acetate and hexane extracts of D. viscosa was determined by the same modified DPPH method. Both pure compounds showed significant percentage of inhibition at all applied concentrations against DPPH method and presented in Table 3. The compound 1 showed the highest percentage of inhibition compare to compound 2.

4. Discussion

Huge number of medicinal plants has been investigated worldwide by the scientists for their biological activities. Most of the scientists are interested to find out the naturally occurring new drugs with significant medicinal values from the nature without side effect for the treatment of diseases. The plant crude extract contains several chemical compounds which is responsible for biological activities. The bioactivities of crude extracts obtained from the plants are completely depends on the bioactive constituents. The selected medicinal plant species showed several biological activities like antioxidant, antimicrobial, anticandidal, antiadibetic, anti-fungal, and antiarheal activities [1,12] and the plant is used traditionally for the treatment of different diseases. The leaves of the selected medicinal plant species are used traditionally worldwide including Oman for the treatment of itching, swelling, aches,

## Table 1

| Crude extracts | Yield (g) |
|---------------|----------|
| Hexane        | 1.30     |
| Chloroform    | 2.81     |
| Ethyl acetate | 1.82     |
| Butanol       | 1.09     |
| Methanol      | 19.30    |
| Water         | 1.64     |
trachoma, antispasmodic agent, gout, bone fraction, rash, and fever [7,9]. In addition, in Oman, the leaves and roots are mixed together and used traditionally for the treatment of toothache, headache, indigestion, ulcer, diarrhea and constipation [7,9,13]. Recently, scientists and researcher are interested to find and isolate the bioactive compounds from natural sources which have showed the biological activities. They are trying to discover new drugs from plant sources for the treatment of different diseases to avoid the

| Crude extracts | Concentration (µg/ml) | Absorption of standard | Absorption of standard | % Inhibition |
|----------------|----------------------|------------------------|------------------------|--------------|
| Methanol       | 200                  | 1.787                  | 0.134                  | 92.78 ± 0.48 |
|                | 100                  | 0.136                  | 92.38 ± 0.10           |
|                | 50                   | 0.145                  | 91.88 ± 0.09           |
|                | 25                   | 0.150                  | 91.60 ± 0.03           |
|                | 12.5                 | 0.185                  | 89.64 ± 0.11           |
|                | 200                  | 1.787                  | 0.350                  | 94.41 ± 0.12 |
|                | 100                  | 0.366                  | 90.51 ± 0.13           |
| Hexane         | 200                  | 1.787                  | 0.392                  | 79.03 ± 0.19 |
|                | 100                  | 0.399                  | 77.62 ± 0.08           |
|                | 50                   | 0.370                  | 89.29 ± 0.20           |
|                | 25                   | 0.392                  | 79.03 ± 0.19           |
|                | 12.5                 | 0.399                  | 77.62 ± 0.08           |
|                | 200                  | 1.787                  | 0.109                  | 93.90 ± 0.03 |
|                | 100                  | 0.110                  | 93.84 ± 0.11           |
| Chloroform     | 50                   | 0.115                  | 93.56 ± 0.28           |
|                | 25                   | 0.220                  | 87.76 ± 0.10           |
|                | 12.5                 | 0.250                  | 86.09 ± 0.08           |
|                | 200                  | 1.787                  | 0.089                  | 95.01 ± 0.18 |
|                | 100                  | 0.108                  | 93.84 ± 0.18           |
| Ethyl acetate  | 50                   | 0.270                  | 84.89 ± 0.07           |
|                | 25                   | 0.330                  | 81.53 ± 0.18           |
|                | 12.5                 | 0.378                  | 78.84 ± 0.13           |
|                | 200                  | 1.787                  | 0.365                  | 78.57 ± 0.08 |
|                | 100                  | 0.390                  | 78.17 ± 0.07           |
| Butanol        | 50                   | 0.456                  | 74.48 ± 0.04           |
|                | 25                   | 0.488                  | 72.69 ± 0.13           |
|                | 12.5                 | 0.499                  | 72.07 ± 0.18           |
|                | 200                  | 1.787                  | 0.249                  | 86.06 ± 0.09 |
|                | 100                  | 0.289                  | 83.82 ± 0.18           |
| Water          | 50                   | 0.370                  | 79.29 ± 0.13           |
|                | 25                   | 0.401                  | 77.56 ± 0.17           |
|                | 12.5                 | 0.479                  | 73.19 ± 0.16           |

Each value is a mean of three biological replicates.

Table 3
Antioxidant activity for four pure samples different antioxidant activity for four samples.

| Crude extracts | Concentration µg/ml | Absorption of standard | Absorption of standard | % Inhibition |
|----------------|--------------------|------------------------|------------------------|--------------|
| 1              | 200                | 1.787                  | 0.596                  | 98.09 ± 0.17 |
|                | 100                | 0.301                  | 98.48 ± 0.11           |
|                | 50                 | 0.065                  | 87.20 ± 0.22           |
|                | 25                 | 0.051                  | 98.43 ± 0.10           |
|                | 12.5               | 0.039                  | 97.64 ± 0.09           |
| 2              | 200                | 1.787                  | 0.095                  | 94.68 ± 0.15 |
|                | 100                | 0.128                  | 92.83 ± 0.71           |
|                | 50                 | 0.059                  | 96.69 ± 0.15           |
|                | 25                 | 0.080                  | 95.50 ± 0.19           |
|                | 12.5               | 0.111                  | 93.70 ± 0.23           |

Each value is a mean of three biological replicates.

Fig. 2. Structure of compound 1 and compound 2.
synthetic drugs. The Sultanate of Oman is considered as mega diversity with plenty of plant sources. In Oman, the selected plant species has been used widely by the local ethnic communities as traditional healing for a variety of conditions. However, until to-date, there is no scientific study conducted to prove the presence of antioxidant compounds of the selected plant. On the basis of medical importance, we are intended to work on the leaves of *D. viscosa*, which was collected from AL-Hamra. The collected samples were used for the preparation of different crude extracts by different polarities solvents. The antioxidant activity of the crude extracts of *D. viscosa* was determined by DPPH method and the results are presented in Table 2. In our experimental results showed that the highest antioxidant activity was found in hexane and ethyl acetate extract and lowest antioxidant activity was found in the butanol extract and followed by hexane > ethyl acetate > chloroform > methanol > water > butanol extract. Due to the significant antioxidant activity, the hexane and ethyl acetate crude extracts were selected for the isolation and separation of antioxidant compounds.

Different chromatography techniques such as thin layer chromatography (TLC), column chromatography (CC) and preparative TLC were used normally for the separation of compounds from the plant crude extract. As a stationary phase silica gel particle (120–260 mesh) and as mobile phase different polarity of solvents were used for the separation of compounds. Initially, in our experiment as mobile phase hexane-ethyl acetate (7:1) was used for column chromatography then increase the polarity of mobile phase by addition of ethyl acetate. The polarity of the mobile phase plays a significant role for the isolation of pure chemical compounds from the crude extracts by different chromatographic system. Based on the TLC behavior, fraction 1 contains lots of compounds compare to another fraction of ethyl acetate extract. In addition, the chemical compounds inside the fraction 1 are separated nicely by using hexane-ethyl acetate (9:1) developing solvent. The same developing solvent was used to separate the pure compounds from the fraction 1 by using preparative TLC. Again, repeat the preparative TLC and crystallization to get the pure compound 1. Similarly, the pure compound 2 was obtained from fraction 2 of hexane extract by several chromatographic systems.

The pure compound 1 was obtained as a yellowish powder. It had the molecular ion peak at [M]+ m/z 291, which corresponds to the molecular formula C15H15O6. In 1H NMR spectrum, the isolated compound 1 showed one multiplet and one doublet of doublets at δ 2.44 and δ 2.64 indicated the presence of two protons at the same position of H-4. Two doublets at δ 3.98 and δ 4.63 indicated the presence of one proton each position at H-2 and H-3. One multiple peak at δ 6.62 indicated the presence of one proton each position at the position H-5' and H-6'on the B-ring. One sharp singlet at δ 6.87 indication the presence of one proton at H-2’. Two doublets at δ 5.69 and δ 5.87 indicated the presence of two aromatic protons at the position H-6 and H-8 on the A-ring. One upfield singlet at δ 4.71 and four downfield sharp singlets at δ 8.70, 8.77, 8.87 and 9.08 indicated the presence of five –OH at different places on the A, B and C-rings (Fig. 3). In the 13C NMR and DEPT spectrum showed that the isolated compound 1 contained total fifteen carbon signals which consisted one methylene, seven methine and seven quaternary carbons (Fig. 4). The 1H–1H COSY correlations showed that the compound 1 coupling between H-2 and H-3 protons and H-3 and H-4 protons. These correlations between the positions were confirmed by HMBC (Fig. 5). Based on the above spectral data, the structure of compound 1 was isolated by several authors from different plant species; however, the compound 1 was isolated from the selected plant species for the first time in our laboratory.

According to TLC behavior, out of two compounds (compound 1 and compound 2) from fraction 2 of hexane extract, Compound 2 is almost pure, however, the compound 1 is not 100% pure. The
separated compound 2 was crystallized from dichloromethane to give white crystals. The pure isolated compound 2 was identified on the basis of MS and $^1$H NMR, $^{13}$C NMR and 2D NMR. It had molecular weight ($M^+$, 442) with corresponding molecular formula is C$_{29}$H$_{50}$O$_2$. In the $^1$H NMR spectrum, the isolated compound 2 showed one proton each position at C-3 and C-6 as multiples at δ 5.11 and δ 4.43 indicating the presence of steroidal nucleus. Two protons appeared at δ 5.09 as a multiplet in the $^1$H NMR spectrum which was identical with the chemical shift of H-22 and H-23 respectively of stigmasterol [14,15]. One sharp singlet at δ 4.00 indicated the presence of –OCH$_3$ at the position of C-4. Two signets at δ 0.93 and δ 0.96 indicated the presence of two –CH$_3$ at the position of C-19 and C-18 on the ring system and four doublets at δ 0.77, 0.81, 0.82 and 0.84 indicating the presence of four –CH$_3$ at the position of C-27, C-26, C-29 and C-21, respectively (Fig. 6). Two olefinic protons appeared as downfield in the $^1$H NMR spectrum as multiplet at δ 5.09 indicated the presence of one proton each position at H-22 and H-23, respectively of stigmasterol [14,15]. From the DEPT 90 and 135 experiments showed that the compound 2 was having six methyl (CH$_3$) groups, eight methylene (CH$_2$), twelve methane (CH), one methoxyl (–OCH$_3$) group and three quaternary carbons groups. The $^1$H–$^1$H COSY correlations showed that the compound 2 coupling between H-2 and H-3, H-5 and H-6, H-9 and H-11, H-13 and H-14, H-20 and H-22 and H-23 and H-24 protons. These correlations between the positions were confirmed by HMBC (Fig. 5). Based on the above spectral data structure of compound 2 was identified as 4-methoxystigmasterol (2). It was also isolated from the selected plant species for the first time in our laboratory.

4.1. Antioxidant activity of pure compounds

The evaluation of antioxidant activity of the isolated pure compound 1 and 2 from ethyl acetate and hexane extract by the same
The method described earlier [16]. The compounds 1 and 2 showed significant antioxidant activity against DPPH as compared to standard gallic acid (Table 3). However, the isolated compound 1 at all concentrations showed very high antioxidant activity compared to compound 2. According to 1D and 2D NMR, the compound 2 having –OH and –OCH3 groups. The highest antioxidant activity of pure compound 1 may be due to the –OH groups. The –OH group is free in compound 1 and it is significantly enhancing its antioxidant activity. This observation is in agreement with the earlier structural-activity study [15,17,18].

5. Conclusion

Antioxidants are an important parameter to produce of free radical in the body to improve the human immune system. Therefore, the risks of certain diseases are lowered due to antioxidants. D. viscosa is a medicinal plant, which contains high levels of antioxidant activity [19,20]. Due to the antioxidant activity, the hexane and ethyl acetate crude extract were selected for the isolation and separation of antioxidant compounds. All the isolated pure compounds showed significant antioxidant activity against DPPH as compared to standard gallic acid. From our experimental results, 3,3',4',5,7-pentahydroxyflavane (1) and 4-methoxystigmasterol (2) were isolated from the leaves of locally available D. viscosa and their chemical structures characterized based on 1D and 2D NMR. Further in vivo study is required for the isolated compound 1 and compound 2 to confirm the mode biological activities as well as structure-activity relationship.

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Fig. 6. 1H NMR spectrum of pure compound 2.
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