Exocrine pancreatic function is preserved in systemic sclerosis

Gracijela Bozovic1, Rille Pullerits2,3†, Arne Ståhl3, Kristina Ydström4, Daniel Wenger5, Jan Marsal6, Pontus Thulin3 and Kristofer Andréasson7*

Abstract

Background: Systemic sclerosis (SSc) has been suggested to cause exocrine pancreatic dysfunction. However, a case-control-based autopsy study failed to associate systemic sclerosis with any pancreatic histopathology. The primary objective of this study was to examine the exocrine pancreatic function in consecutive SSc patients in relation to an age- and sex-matched control group. A secondary objective was to relate exocrine pancreatic function to radiological, laboratory, and clinical SSc characteristics.

Methods: One hundred twelve consecutive patients fulfilling the 2013 American Congress of Rheumatology/European League Against Rheumatism criteria for SSc and 52 control subjects were matched for sex and age. Exocrine pancreatic function was assessed by ELISA-based measurement of fecal elastase, and levels ≤ 200 μg/g were considered pathological, i.e., representing exocrine pancreatic insufficiency. Patients were characterized regarding SSc manifestations including gastrointestinal and hepatobiliary function, by use of laboratory and clinical examinations. Pancreas parenchyma characteristics were evaluated by high-resolution computer tomography (HRCT).

Results: A similar proportion of subjects exhibited pathological levels of fecal elastase among SSc patients (6/112; 5.4%) and control subjects (3/52; 5.8%). Patients with fecal elastase ≤ 200 μg/g did not differ from other SSc patients with respect to laboratory and clinical characteristics, including malnutrition. SSc subjects with low levels of fecal elastase displayed significantly lower pancreas attenuation on HRCT examinations compared to the control subjects.

Conclusions: In this study encompassing 112 consecutive SSc patients and 52 matched control subjects, we were unable to associate systemic sclerosis with clinically significant exocrine pancreatic dysfunction.

Keywords: Pancreas, Systemic sclerosis, Fecal elastase, Malnutrition

Introduction

Systemic sclerosis (SSc) is a heterogeneous systemic disease characterized by the development of autoimmunity, vasculopathy, and multiorgan fibrosis. Involvement of the gastrointestinal (GI) tract is common, affecting up to 90% of patients, and is a significant contributor to both morbidity and mortality [1]. Malnutrition is common, and its etiology is multifactorial and incompletely understood. Factors that may play a role include reduced appetite, poor functional status of the hands and fingers, esophageal and GI dysmotility, and small intestinal bacterial overgrowth [2]. SSc has been linked to exocrine pancreatic insufficiency (EPI), and EPI has been suggested to contribute to malnutrition in SSc [1, 3–6].

EPI is easily and efficiently treated with pancreatic enzyme replacement therapy [7]. Consequently, it is important to identify SSc patients suffering from this disorder. During the last decades, the measurement of fecal elastase (FE) has been established as a reliable method to screen for clinically significant EPI with a reported sensitivity above 90% [8]. Assessment of exocrine pancreatic function by FE measurement has also been recommended in the evaluation of SSc-associated malnutrition and steatorrhea [9, 10].

SSc-related pancreatic tissue pathologies are characterized by both inflammation and the conversion of
functional parenchyma to a fibrous stroma, sometimes with the replacement of parenchyma with fat. Pancreatic fat can be quantified by non-contrast-enhanced computed tomography (CT), resulting in lower attenuation measured in Hounsfield units (HU) [11].

The purpose of this study was to investigate the prevalence of EPI in a consecutively assembled cohort of SSc patients in relation to an age- and sex-matched control group. As a secondary aim, we wanted to investigate if EPI in SSc was associated with any specific SSc characteristic or radiological alterations of the pancreas parenchyma.

Methods
Study population
Consecutive SSc patients at scheduled routine visits to our clinic were invited to participate in this study from April 2014 to June 2015. Age- and sex-matched control subjects were recruited from the staff of our clinics (n = 40), from the spouses of patients (n = 4), and from a neighboring orthopedic ward (n = 8). Patients and controls with concomitant pancreatic disease (including chronic pancreatitis and pancreatic cysts), a history of pancreatic surgery, or alcohol abuse were excluded from the study in order not to include patients with non-SSc-related EPI. Among the controls, subjects with rheumatic disease were also excluded. A separate control group was used for the radiological study, see below.

Clinical characteristics
SSc was defined according to the 2013 American Congress of Rheumatology/European League Against Rheumatism (ACR/EULAR) criteria, and patients were subdivided into diffuse cutaneous SSc (dcSSc) and limited cutaneous SSc (lcSSc) [12, 13]. SSc disease duration was defined as years since the first non-Raynaud symptom of disease. Body mass index was recorded, and malnutrition was assessed according to the validated Malnutrition Universal Screening Tool (MUST) [14]. All patients were systematically questioned regarding the following GI symptoms: heartburn (dyspepsia), dysphagia, diarrhea, and/or constipation. These were recorded as present or not. Lung fibrosis was defined as the presence of fibrosis on high-resolution computed tomography (HRCT) as specified in the ACR/EULAR criteria [12]. Esophageal function was assessed by barium cineradiography and graded 0 (normal) to 2 (aperistalsis) since this investigation has been suggested to be a marker of GI manifestations of SSc [15].

Laboratory analyses
FE was measured with ELISA utilizing a monoclonal antibody towards pancreatic elastase 1 (Schebo Biotech, Giessen, Germany). Samples from control subjects and patients were analyzed in duplicates on the same ELISA plates. According to the manufacturer and the literature [7, 8], levels below 15 μg/g were considered indicative of severe pancreatic dysfunction while levels between 15 and 200 μg/g were categorized as possible insufficiency. With a suggested cutoff of 200 μg/g, this analysis has been shown to identify EPI with high sensitivity and specificity [8]. F-calprotectin, a marker of GI inflammation has previously been associated with GI manifestations of SSc, was measured with ELISA (Calpro, Lysaker, Norway) [16]. Markers of hepatobiliary function (aspartate aminotransferase, alaninaminotransferase, alkaline phosphatase, gamma-glutamyltransferase) were assessed as well as pancreatic amylase. Prealbumin (transthyretin) and albumin were measured as markers of malnutrition. Prealbumin levels below 200 mg/l were considered indicative of malnutrition [17]. Vitamin D was measured since low levels of this vitamin have been associated with both SSc and EPI [18, 19].

Radiological assessment of the pancreas
All patients had been examined with a mandatory HRCT in search for pulmonary involvement of SSc. Although focused on the lungs, it includes all or most of the pancreas since the basal parts of the lungs extend over the upper abdomen. We included a separate control group for the radiology analysis. These control subjects were randomly selected from the radiologist’s workflow during a 6-month period and matched for sex and age ± 6 years apart, with the exception of two patients. They had been referred to the radiologist for HRCT and did not have any concomitant pancreatic disease, diabetes mellitus, a history of pancreatic surgery, or alcohol abuse. Radiology subjects were divided into three groups, each pre-specified to encompass at least seven subjects:

1) SSc patients with low FE levels (n = 7; FE ≤ 210 μg/g);
2) Age- and sex-matched SSc patients with normal levels of FE (n = 21);
3) Age- and sex-matched control subjects without SSc (n = 21).

The HRCT examinations were performed with routine scan settings and reconstruction parameters on CT scanners. HRCT parameters are summarized in Additional file 1: Table S1. All scans were done with 120 kV using dose modulation to optimize the image quality according to patient size. Reconstructions were done with 3 mm slice thickness and with kernel/filter according to Additional file 1: Table S1.

Retrospectively, the CT number (HU value) was measured by one radiologist blinded for the clinical data. A
circular region of interest (ROI) up to 1 cm in diameter was drawn in the pancreatic body and tail respectively, avoiding vascular structures and cysts if present (Fig. 1). Furthermore, a similar ROI was placed in the spleen as a reference for normalization of attenuation to compensate for the variability between different CT machines [20].

Statistics
Non-parametric statistics and Fischer’s exact test were consistently used in this study when describing and comparing FE in the patient and control group, as well as when analyzing FE in relation to patient data. Spearman’s rank-order correlation was used when relating FE to F-calprotectin and esophageal function. Parametric statistics were used when analyzing the radiological examinations. \( p < 0.05 \) was considered significant.

Ethics
The study was approved by the Regional Ethics Review Board, Lund, Sweden, reference number 2011/596. Informed written consent was obtained from all subjects before study inclusion, and the study conformed to the ethical guidelines of the Declaration of Helsinki.

Results
F-elastase
The characteristics of patients and control subjects are described in Table 1. A minority of the SSc patients (6/112; 5.4%) exhibited FE levels \( \leq 200 \mu g/g \), of which no one presented levels indicative of severe dysfunction (<15 \( \mu g/g \)). In the control group, 3/52 (5.8%) had FE levels \( \leq 200 \mu g/g \), of which no one had levels below 15 \( \mu g/g \), which was not statistically different from patients with SSc (\( p = 1.00 \)). Median (interquartile range [IQR]) levels of FE were similar between patients (800 [515–1475] \( \mu g/g \)) and control subjects (1200 [435–1700] \( \mu g/g \); \( p = 0.189 \)), as shown in Fig. 2.

Clinical characteristics
Three of the six patients with low FE levels had dcSSc. This disease subtype was not statistically overrepresented compared to the lcSSc (\( p = 0.137 \)).

None of the 13 patients with a body mass index < 20 and none of the 18 patients with a MUST score ≥ 1 (indicating malnutrition) had FE \( \leq 200 \mu g/g \). Median FE levels did not differ between patients with and without pathological MUST scores (940 [590–1850] and 800 [510–1400] \( \mu g/g \), respectively; \( p = 0.428 \)). Also, median FE levels did not differ between those with and without pathological prealbumin levels (675 [423–1065] and 780 [565–1400] \( \mu g/g \), respectively; \( p = 0.284 \), \( n = 68 \)). Only 1 of 16 patients with pathological prealbumin levels had FE \( \leq 200 \mu g/g \). The presence of GI symptoms was not associated with pathological FE testing (\( p = 0.377 \), \( p = 0.648 \), \( p = 0.562 \), and \( p = 0.691 \) for heartburn, dysphagia, diarrhea, and constipation, respectively). Anti-mitochondrial
antibodies were present in 7 patients, 5 patients had a diagnosis of primary biliary cirrhosis, and 3 had diabetes mellitus; none of these had pathological FE testing.

FE levels were not associated with intestinal inflammation as assessed by F-calprotectin ($r_s = 0.00$, $p = 0.952$) or intestinal dysmotility as assessed by cineradiography ($r_s = -0.08$, $p = 0.422$). FE $\leq 200 \mu g/g$ did not associate with laboratory markers of hepatobiliary function, pancreatic amylase, SSc disease duration, or antibody profile (Table 2).

### Radiological assessment

In this analysis, 28 patients and 21 control subjects were analyzed. Controls were patients who underwent a HRCT for chronic obstructive pulmonary disease, pulmonary fibrosis, or bronchiectasis. The HRCT examination was done within 1 year of the FE sampling in 22 of the 28 subjects with SSc. The median (IQR) age in the control group ($n = 21$) was 63 (47–72) years, similar to the median age of the SSc subjects with low FE ($n = 7$, median age 67 [62–74] years) and normal FE ($n = 21$, median age 71 [58–73] years) who were subject to radiological analysis ($p = 0.046$). We identified an age-dependent variation in pancreas attenuation both in the SSc subjects ($r = -0.39$, $p = 0.041$) and the control subjects ($r = -0.45$, $p = 0.044$). Pancreas attenuation, normalized in reference to the spleen, was significantly lower in SSc patients with low levels of FE compared to control subjects (0.798 vs. 0.932; $p = 0.024$), as shown in Fig. 3. However, SSc patients with normal levels of FE did not express significantly different attenuation compared to control subjects (0.910 vs. 0.932, $p = 0.201$).

### Discussion

Malnutrition afflicts approximately 18% of patients with SSc, is hard to manage, and is associated with both

---

**Table 1** Patient and control subjects characteristics

|                      | SSc subjects ($n = 112$) | Control subjects ($n = 52$) |
|----------------------|--------------------------|-----------------------------|
| Age (years)          | 62 (50, 69)              | 62 (51, 66)                 |
| Sex (female/male)    | 89/23 (3.9:1)            | 41/11 (3.7:1)               |
| Disease duration (years) | 7 (3, 15)           |                             |
| Disease subtype (dcSSc/lcSSc) | 26/86 (1:3.3)    |                             |
| ANA positive ($n \%$) | 105 (94\%)               |                             |
| ACA positive ($n \%$) | 39 (35\%)                |                             |
| ATA positive ($n \%$) | 20 (18\%)                |                             |
| ARA positive ($n \%$) | 10 (9\%)                 |                             |
| Lung fibrosis ($n \%$) | 37 (33\%)              |                             |
| Cineradiography (normal; mild to moderate pathology; aperistalsis) ($n = 110$) | 22; 80; 8 |                             |
| MUST score (0; 1; 2) | 94; 15; 3               |                             |
| Prealbumin $< 200 \text{mg/l} (n \%)^*$ | 16 (24\%) |                             |
| Heartburn$^\dagger$ | 59 (53\%)               |                             |
| Dysphagia$^\dagger$  | 47 (42\%)               |                             |
| Diarrhea$^\dagger$   | 12 (11\%)               |                             |
| Constipation$^\dagger$ | 14 (13\%)            |                             |

Values are expressed as median (interquartile range) if not otherwise stated
dcSSc diffuse cutaneous systemic sclerosis, lcSSc limited cutaneous systemic sclerosis, ACA anti-centromere antibodies, ATA anti-topoisomerase antibodies, ARA anti-RNA polymerase 3 antibodies, MUST Malnutrition Universal Screening Tool [13]

*Prealbumin analyzed in 68 patients
$^\dagger$Data available on 111 patients

---

**Fig. 2** Fecal elastase levels in systemic sclerosis and control subjects. Box plot indicating fecal elastase levels in patients with systemic sclerosis and age- and sex-matched controls
### Table 2: Laboratory and clinical characteristics of patients with and without pathological levels of fecal elastase

|                  | ALT (U/L) | AST (U/L) | GGT (U/L) | ALP (U/L) | Pancreatic amylase (U/L) | Calcium (mmol/l) | Magnesium (mmol/l) | Albumin (g/l) | Prealbumin (g/l) | Vitamin D3 (nmol/l) | Disease duration (years) | Age (years) | ACA (n) | ATA (n) | ARA (n) |
|------------------|-----------|-----------|-----------|-----------|--------------------------|------------------|-------------------|---------------|-----------------|----------------------|--------------------------|-------------|---------|---------|---------|
| FE ≤ 200 µg/g    | 25 (16–41) | 29 (22–40) | 55 (22–156) | 71 (22–32) | 24 (22–32) | 2.4 (2.3–2.5) | 0.93 (0.77–1.1) | 40 (37–42) | 0.33 (0.19, 0.36) | 48 (29, 65) | 5 (1, 15) | 70 (57, 76) | 1 | 1 | 1 |
| (n = 6)          |           |           |           |           |             |                  |                  |               |                 |                      |                         |             |         |         |         |
| FE > 200 µg/g    | 19 (14–24) | 24 (21–29) | 71 (52–81) | 25 (18–25) | 2.3 (2.3–2.4) | 0.82 (0.77–0.86) | 39 (36–41) | 0.25 (0.2, 0.3) | 70 (45, 78) | 7(3, 15) | 60 (61, 69) | 38 | 19 | 9 |
| (n = 106)        |           |           |           |           |             |                  |                  |               |                 |                      |                         |             |         |         |         |

Systemic sclerosis patients with pathological FE testing did not differ compared to other patients with regard to laboratory markers of liver function and malnutrition, disease duration, age, and antibody profile. *p > 0.05* for all variables when comparing patients with FE ≤ 200 µg/g to patients with FE > 200 µg/g. Values are given as median (interquartile range).

FE: fecal elastase, ALT: alanin aminotransferase, AST: aspartate aminotransferase, GGT: gamma-glutamyltransferase, ALP: alkaline phosphatase, ACA: anti-centromere antibodies, ATA: anti-topoisomerase antibodies, ARA: anti-RNA polymerase III antibodies.
significant morbidity and mortality [2]. EPI is an important potential cause of malnutrition since it can be easily and efficiently treated with specific replacement therapy. In order to assess the prevalence of clinically significant EPI in SSc, we have examined 112 consecutive SSc patients by measurements of FE and related these findings to 52 age- and sex-matched controls. Low levels of FE were uncommon in both the SSc and the control groups. These data indicate that EPI is not a major cause of SSc-related malnutrition.

The gold standard for assessing EPI is the secretin-cerulein test. This test is invasive and cumbersome and not recommended for screening purposes [7]. FE is a pancreas-specific enzyme that is not degraded during intestinal transport. The enzyme is inert to degradation also when stored in room temperature. It can be successfully measured with ELISA and has proven to have limited intraindividual variability [8, 21]. FE measurement is superior to other indirect tests of pancreatic function including the fecal chymotrypsin test and the $^{13}$C-mixed triglyceride breath test. Measurement of FE with the monoclonal ELISA used in this study has a sensitivity and specificity above 90% in identifying EPI. Consequently, FE measurement has been established as an alternative to secretin-cerulein test when screening for EPI in both research and clinical settings [8, 22]. Even if FE has been suggested to be of value in the evaluation of SSc-related malnutrition [9], to our knowledge, FE has not previously been studied in SSc.

Conflicting data exists regarding the prevalence of EPI in SSc. A case-control autopsy study from 1969 failed to identify SSc-specific pancreas pathology in 58 SSc patients [23]. Smaller functional studies have suggested that EPI may be prevalent in SSc [3–6, 24], although none of them included a control group. The clinical relevance of the alterations identified in these studies is unclear since mild EPI is often asymptomatic [7]. Also, the generalizability of the larger of these studies ($n = 31$) is questionable since it comprised “a highly selective group of patients deliberately sequestered by virtue of the gastrointestinal complaints” [3].

We observed that 77% of our patients had lcSSc. Although not statistically significant, we noted that EPI was more common among patients with dcSSc. In the study by Shawis et al, encompassing five dcSSc and six lcSSc subjects, two out of three patients with EPI had lcSSc [6]. Earlier works on EPI in SSc were published before these classification subtypes were established [13].

In our cohort, a pathological MUST score was present in 16% of the subjects and pathological prealbumin levels observed in 24% of the patients studied. These figures are similar to what has previously been reported in SSc and indicate that malnutrition was indeed prevalent in our cohort [14, 17]. Still, in this study, we were unable to associate malnutrition to EPI.

It was beyond the scope of this study to investigate other, non-pancreatic causes of malnutrition. Previous studies have suggested a complex mixture of SSc-related complications including GI dysmotility and small intestinal bacterial overgrowth, systemic inflammation, and extraintestinal manifestations to cause malnutrition in SSc [2]. Our results indicate that exocrine pancreatic dysfunction is not an important factor behind SSc-related malnutrition compared to the ones presented above.

In order to further investigate the pancreas in relation to SSc, patients with and without pathological FE were retrospectively studied using CT. The HRCT examinations identified an age-dependent decline in pancreatic attenuation in keeping with previous studies [11], but also a statistically significant difference in pancreatic attenuation in the subgroup of patients with low levels of
FE compared to control subjects. The lower attenuation is likely caused by the replacement of exocrine pancreatic tissue with fat (pancreatic lipomatosis), possibly reflecting a destructive inflammatory process with increased parenchymal turnover [11]. Whether these data reflect an SSc-specific process remains to be elucidated. We were unable to find any similar differences when comparing SSc patients with normal FE levels to control subjects. HRCT is not the optimal imaging modality to assess the pancreas. Our patients were examined with a mandatory chest HRCT because of suspected lung fibrosis, an examination that usually includes all or most of the pancreas due to the anatomy of the lung. With respect to the radiation risks associated with CT examinations, we chose not to assess additional radiological examination but benefit from already existing ones. Further studies using magnetic resonance imaging and endoscopy, as well as autopsy studies, are warranted to understand if and how the pancreas may be affected in SSc.

Conclusions
Our knowledge on any SSc-specific pathobiology of the pancreas is limited. In order to determine the prevalence of EPI in SSc, we have investigated a fairly large number of consecutive SSc patients and age- and sex-matched control subjects with a validated and sensitive marker of EPI and by HRCT. The radiological analyses might suggest that SSc in some cases may manifest itself in the pancreas but on the whole; our study indicates that exocrine pancreatic function is usually preserved in SSc.

Additional file

Additional file 1: Table S1. High-resolution tomography parameters of machines used in this study. (DOCX 13 kb)

Abbreviations
ACR/EULAR: American Congress of Rheumatology/European League Against Rheumatism; CT: Computer tomography; dSSc: Diffuse cutaneous systemic sclerosis; EPI: Exocrine pancreas insufficiency; FE: Fecal elastase; GI: Gastrointestinal; HRCT: High-resolution computer tomography; HU: Hounsfield units; IQR: Interquartile range; lSSc: Limited cutaneous systemic sclerosis; MUS: Malnutrition Universal Screening Tool; ROI: Region of interest; SSc: Systemic sclerosis

Acknowledgements
The authors would like to thank professor emeritus Frank Wollheim and associate professor Roger Hesselstrand for founding and developing the SSc unit at our clinic and supporting this study. We would also like to thank the patient organization Reumatikerförbundet: Riksföreningen för systemisk skleros for their cooperation.

Funding
This work was supported by grants to researchers in public health care from the Swedish government (ALF for young researchers); Anna-Greta Crafoord Foundation [grant number 20162008]; Swedish Society of Medicine [grant number SLS-595121], Magnus Bergvalls Stiftelse [grant number 2016-01820], Swedish Rheumatism Association [grant number R-478421]

Availability of data and materials
The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Authors’ contributions
GB, RP, and RA designed and organized the study, acquired and analyzed the data, and prepared the manuscript. AS, KY, and DW contributed in acquiring the data and reviewing the manuscript for intellectual content. JM and PT contributed in designing the study and analyzing the data and reviewed the manuscript for intellectual content. All authors read and approved the final manuscript.

Ethics approval and consent to participate
The study was approved by the Regional Ethics Review Board, Lund, Sweden, reference number 2011/596. Informed written consent was obtained from all subjects before study inclusion and the study conformed to the ethical guidelines of the Declaration of Helsinki.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details
1Department of Medical Imaging and Physiology, Skåne University Hospital, Lund, Sweden. 2Department of Rheumatology and Inflammation Research, Sahlgrenska Academy at University of Gothenburg, Gothenburg, Sweden. 3Department of Clinical Immunology and Transfusion Medicine, Sahlgenska University Hospital, Gothenburg, Sweden. 4Department of Medical Radiation Physics, Skåne University Hospital, Lund, Sweden. 5Section of Orthopedics, Department of Clinical Sciences, Lund University, Lund, Sweden. 6Section of Gastroenterology, Department of Clinical Sciences, Lund University, Lund, Sweden. 7Department of Medical Radiation Physics, Skåne University Hospital, Gothenburg, Sweden. 8Department of Medical Imaging and Physiology, Skåne University Hospital, Lund, Sweden. 9Department of Medical Imaging and Physiology, Skåne University Hospital, Lund, Sweden. 10Department of Medical Imaging and Physiology, Skåne University Hospital, Lund, Sweden.

Received: 29 November 2018 Accepted: 3 February 2019
Published online: 12 February 2019

References
1. Jaovisidha K, Csuka ME, Almagro UA, Soergel KH. Severe gastrointestinal involvement in systemic sclerosis: report of five cases and review of the literature. Semin Arthritis Rheum. 2005;34(4):689–702.
2. Harrison E, Herrick AL, McLaughlin JT, Lal S. Malnutrition in systemic sclerosis. Rheumatology (Oxford). 2012;51(10):1747–56.
3. Drelling DA, Soto MJ. The pancreatic involvement in disseminated "collagen" disorders. Studies of pancreatic secretion in patients with scleroderma and Sjogren’s "disease". Am J Gastroenterol. 1976;66(6):546–53.
4. Cobden I, Axon AT, Rowell NR. Pancreatic exocrine function in systemic sclerosis. Br J Dermatol. 1981;105(2):189–93.
5. Hendel L, Worning H. Exocrine pancreatic function in patients with progressive systemic sclerosis. Scand J Gastroenterol. 1989;24(4):461–6.
6. Shawis TN, Chaloner C, Herrick AL, Jayson MJ. Pancreatic function in systemic sclerosis. Br J Rheumatol. 1996;35(5):298–9.
7. Lindkvist B. Diagnosis and treatment of pancreatic exocrine insufficiency. World J Gastroenterol. 2013;19(42):7258–66.
8. Vanga RR, Tansel A, Sidiq S, El-Serag HB, Othman M. Diagnostic performance of measurement of fecal elastase-1 in detection of exocrine pancreatic insufficiency: systematic review and meta-analysis. Clin Gastroenterol Hepatol. 2018;16(8):1220–8.
9. Hansi N, Thoua N, Carulli M, et al. Consensus best practice pathway of the UK scleroderma study group: gastrointestinal manifestations of systemic sclerosis. Clin Exp Rheumatol. 2014;32(Suppl 86):S214–21.
10. Gyger G, Baron M. Systemic sclerosis: gastrointestinal disease and its management. Rheum Dis Clin N Am. 2015;41(3):459–73.
11. Kim SY, Kim H, Cho JY, et al. Quantitative assessment of pancreatic fat by using unenhanced CT: pathologic correlation and clinical implications. Radiology. 2014;271(1):104–12.
12. van den Hoogen F, Khanna D, Fransen J, et al. 2013 classification criteria for systemic sclerosis: an American College of Rheumatology/European League Against Rheumatism Collaborative Initiative. Ann Rheum Dis. 2013;72(11):1747–55.
13. LeRoy EC, Black C, Fleischmajer R, Jablonska S, Krieg T, Medsger TA, Rowell N, Wollheim F. Scleroderma (systemic sclerosis): classification, subsets and pathogenesis. J Rheum. 1988;15(2):202–5.
14. Baron M, Hudson M, Steele R. Canadian Scleroderma Research Group. Malnutrition is common in systemic sclerosis: results from the Canadian scleroderma research group database. J Rheumatol. 2003;30(12):2737–43.
15. Clements PJ, Becvar R, Drosos AA, Ghattas L, Gabrieli A. Assessment of gastrointestinal involvement. Clin Exp Rheumatol. 2003;21(Suppl 29):S15–8.
16. Andreasson K, Scheja A, Saxne T, Ohlsson B, Hesselstrand R. Faecal calprotectin: a biomarker of gastrointestinal disease in systemic sclerosis. J Intern Med. 2011;269(1):50–7.
17. Codullo V, Cereda E, Klersy C, et al. Serum prealbumin is an independent predictor of mortality in systemic sclerosis outpatients. Rheumatology (Oxford). 2016;55(2):315–9.
18. Mann ST, Stracke H, Lange U, Klör HU, Teichmann J. Vitamin D3 in patients with various grades of chronic pancreatitis, according to morphological and functional criteria of the pancreas. Dig Dis Sci. 2003;48(3):533–8.
19. Vacca A, Cornier C, Piras M, Mathieu A, Kahan A, Allanore Y. Vitamin D deficiency and insufficiency in 2 independent cohorts of patients with systemic sclerosis. J Rheumatol. 2009;36(9):1924–9.
20. Zeb I, Li D, Nasir K, Katz R, Larijani VN, Budoff MJ. Computed tomography scans in the evaluation of fatty liver disease in a population based study: the multi-ethnic study of atherosclerosis. Acad Radiol. 2012;19(7):811–8.
21. Löser C, Møllgaard A, Fölsch UR. Faecal elastase-1: a novel, highly sensitive, and specific tubeless pancreatic function test. Gut. 1996;39(4):580–5.
22. Leeds JS, Oppong K, Sanders DS. The role of fecal elastase-1 in detecting exocrine pancreatic disease. Nat Rev Gastroenterol Hepatol. 2011;8(7):405–15.
23. D’Angelo WA, Fries JF, Masi AT, Shulman LE. Pathologic observations in systemic sclerosis (scleroderma). A study of fifty-eight autopsy cases and fifty-eight matched controls. Am J Med. 1969;46(3):428–40.
24. Scudamore HH, Green PA, Hofman HH 2nd, Rosewar J, Tauze WN. Scleroderma (progressive systemic sclerosis) of the small intestine with malabsorption. Evaluation of intestinal absorption and pancreatic function. Am J Gastroenterol. 1968;49(3):193–208.