Light Emitting Diodes as Alternative Light Sources: Effects of Ultraviolet Frequencies on Microbial Replication

Cardaci Rosa¹, Sandra Burgassi¹, Spataro Giuseppe², Tani Marco³, Cevenini Gabriele⁴ and Messina Gabriele¹,2*

¹Department of Molecular and Developmental Medicine, Italy
²Post Graduate School of Public Health, University of Siena, Italy
³FeT Elettronica Snc, via A. Volta 28, Poggibonsi (Siena), Italy
⁴Department of Medical biotechnology, Italy

*Corresponding author: Messina Gabriele, Department of Molecular and Developmental Medicine, via Aldo Moro 2 – 53100 Siena, Italy.

ARTICLE INFO

Received: March 27, 2019
Published: April 09, 2019

Citation: Cardaci R, Burgassi S, Spataro G, Tani M, Cevenini G, et al. Light Emitting Diodes as Alternative Light Sources: Effects of Ultraviolet Frequencies on Microbial Replication. Biomed J Sci & Tech Res 16(5)-2019. BJSTR. MS.ID.002925.

ABSTRACT

Background: In the biomedical field the UV-C rays are known for altering the effect of replication on different microbial species. Most of the studies reported in bibliography use the UV-C lamps, as light sources, that have a broad spectral emission, on the contrary Light Emitting Diodes have specific wavelengths. The aim of our research is to test these innovative sources of light at different wavelengths on microorganism’s replication in different time.

Methods: The present pilot study in which pre and post exposure phases were performed at different LED sources. It was conducted in the months of January and February 2018 in the Environmental Hygiene Laboratory of the University of Siena. The tested LEDs had a power and the following wavelengths 276 and 279 nm (UV-C); 305,306 and 308 nm (UV-B); 343 and 345 nm (UV-A). For each of them, tests were carried out using: Enterococcus faecalis, Pseudomonas aeruginosa, Staphylococcus aureus, Escherichia coli and Bacillus subtilis. For each of these strains, bacterial suspensions were prepared. Aliquots of these were seeded in double in Petri dishes with selective medium for each microorganism chosen. Treatment involved illuminating exposed Petri dishes with a UV LED for 10-30 and 60 minutes and then incubated at 36°C for 24 hours. For each microorganism, four Petri dishes contaminated but not exposed to UV radiation were used as controls. They were also incubated in a thermostat at 36°C and then read at 24 hours.

Results: We found differences in the replication of microorganisms between the different wavelengths. For UV-C there was a reduction between 97% and 100% already at 10 minutes. For UV-B we noted that the replication for all microorganisms was greater than the UV-C and time-related. For UV-A although with some uncertainties, in general, we still found a time-related reduction.

Conclusion: The results show that the efficacy of the LED depends on exposure times, wavelengths and microbial species. There is evidence that monochromatic UV LED source, influence the microbial replication in different way. Such an eventuality opens innumerable hypotheses of study and possible innovative applications.

Introduction

Hospital acquired infections (HAIs) are a major global problem as they increase morbidity, mortality, hospitalization length and the cost of care for hospitalized patients [1,2]. Several pathogenic microorganisms including Pseudomonas aeruginosa, E. coli, Enterococcus spy (including VRE) and methicillin-resistant
**Staphylococcus aureus** are able to remain on medical surfaces and devices for long periods [3,4]. Studies carried out by Weber et al. [5] show that about 20-40% of HIAs are due to the lack of hygienic attention of healthcare workers who may have become contaminated through direct infection with the patient in hospital or indirectly by touching the contaminated environmental surfaces. In fact, the environment is increasingly recognized as a significant element for microbial cross-contamination [6]. For this reason, a report by the CDC highlights the need to not underestimate the standards of good hygiene practice, which, although obvious, remain a reference element [7]. Among the methods of disinfection, the use of disinfectants is another element to be considered in order to implement a proper sanitization of environments and objects. In addition, the use of ultraviolet radiation is also becoming more and more emerging [8].

The radiation with the most germicidal effect is represented by UV-C (200-280 nm). It has long been known that the damage that these determine is given by the formation of bonds between the pyrimidine nitrogenous bases present in the DNA of microorganisms that combine to form dimers of thymine that if they remain prevent microorganisms from being able to replicate [9]. Such radiations are artificially produced by the classic germicidal lamps. Recent researches have shown that other electronic frequencies can have a biocidal effect. Also, the appearance of alternative light sources: Light Emitting Diodes (LEDs) allows you to select the frequencies of interest to capture the effects of these on various species of microbes [8,10]. In view of this, it is necessary to examine the issue in greater depth in order to be able to consider its application in the health environment. The purpose of this work is to verify the biocidal effect of the various UV frequencies (A-B-C) emitted by LEDs on some microbial species, relevant in HIAs, at different exposure times (10-30-60 minutes).

**Methods**

This research is a pilot study in which pre and post exposure phases were performed at different LED sources, with the following frequencies: 276 and 279 nm (UV-C) by CUD7GF1A; 306 and 308 nm (UV-B) by LEUVA66G; 305 and 306 nm (UV-B) by CUD1AF4C; 343 and 354 nm (UV-A) by CUD4AF1B. The survey was conducted in January and February 2018 in the Laboratory of Environmental Hygiene (microbiological section) of the University of Siena.

**Experimental Setting**

In order to conduct and standardize the experiments it was necessary to build a setting that would allow a uniform distribution between the light source and the Petri dishes. The setting to conduct the study was designed with the help of the 3D modeling program, Solid works, and then realized with a 3D lithographic printer, Formlab Form 2. Our setting had two supports, shaped as an inverted cone, in which the vertex had a hole specifically created to allow the insertion of the various LEDs. Those LEDs were soldered on PCBs and powered in order to generate 3 mW. To standardize this power and measure the different wavelengths emitted by the LEDs, an integrating sphere connected to a spectrophotometer was used.

**Selection of Microorganisms and Preparation Protocol**

The microorganisms were chosen on the basis of the following features

a. Morphological characteristics

b. Ability to produce spores (forms of resistance)

c. Causation of HIAs.

They were: *Staphylococcus aureus* ATCC13150; *Pseudomonas aeruginosa* ATCC27853; *Escherichia coli* (not ATCC); *Bacillus subtilis* (not ATCC); *Enterococcus faecalis* ATCC51299. In the case of *Bacillus subtilis* the inoculation was on Nutrient Agar with the addition of a Manganese chloride solution. The incubation of the Petri dishes took place at a temperature of 30°C and lasted for about 10 days until the complete transformation of the microorganisms in sporogenous form. For the spores of *Bacillus subtilis* and for all other microorganisms in the study, bacterial suspensions were set up in 10 ml of sterile Phosphate Buffered Saline (PBS) in order to obtain a concentration of 0.3 McFarland. For each bacterial and sporogenous suspension dilutions were made and for the present work a dilution of 10-2 was used.

Rates of 40μL of these dilutions were sown in duplicate by spatula in 55 nm diameter Petri dishes in the agar specific medium. For *Staphylococcus aureus* (Mannitol Salt Agar); *Pseudomonas aeruginosa* (Cetrimide agar); *Escherichia coli* (BriBiance E. coli Coliform Selective Medium); *Bacillus subtilis* (Nutrient Agar); *Enterococcus faecalis* (Slanet And Bartley Medium). The Petri dishes thus contaminated were exposed to the radiation of the various LEDs. Each Petri dish was individually mounted in the cone inverted setting, and exposed to a specific UV wavelength at 10, 30 and 60 minutes. Subsequently, the irradiated plates were incubated in a thermostat at 36°C, for each of them reading was made at 24 hours. For each microorganism, four Petri dishes contaminated but not exposed to UV radiation were used as controls. They were also incubated in a thermostat at 36°C and then read at 24 hours.

**Results**

Differences in the effectiveness of LEDs in reducing the microbial load have been observed, from UV-C frequencies to UV-B and eventually to UV-A. The average reduction percentage linked to the frequency of UV-C LEDs was 100% already at 10’ for *Staphylococcus aureus*, *Escherichia coli* and *Pseudomonas aeruginosa*, while for *Enterococcus faecalis* there was a reduction of 97% and 99% for *Bacillus subtilis*. After 30’ of exposure for *Staphylococcus aureus* and *Bacillus subtilis* there was a reduction of 98.2%, for *Escherichia coli* of 99.7%, for *Enterococcus faecalis* and *Pseudomonas aeruginosa* of 100%. All the microorganisms tested after 60’ of exposure showed, instead, a reduction of 100%. The average percentage of reduction related to the frequency of UV-B LEDs was lower than the results obtained by testing UV-C rays, the microbial charge decreased as the exposure time increased. In particular, it emerges that *Pseudomonas aeruginosa* was most affected by their effectiveness, and at 10’ it was already...
reduced by 70.8% and 95.5% at 60', while the microorganism that was less sensitive was Escherichia coli, with a reduction of 22.1% at 10' and 95% at 60'. The average percentage of reduction related to the frequency of UV-A LEDs is the lowest compared to the others mentioned above, but still gave good results compared to the controls, although with certain critical issues, especially for Staphylococcus aureus, Enterococcus faecalis and Bacillus subtilis.

Table 1 shows the results of the specific species-frequency pairings.

| LED | Staphylococcus aureus | Escherichia coli | Pseudomonas aeruginosa | Enterococcus faecalis | Bacillus subtilis |
|-----|-----------------------|-----------------|------------------------|-----------------------|------------------|
| UVC | 276                   | 100,0           | 99,4                   | 100,0                 | 97,0             |
|     | 279                   | 98,2            | 99,7                   | 100,0                 | 99,8             |
| UVB | 306                   | 57,6            | 22,1                   | 70,8                  | 32,3             |
|     | 308                   | 83,6            | 69,6                   | 93,3                  | 38,8             |
| UVA | 343                   | 47,2            | 1,9                    | 3,3                   | 25,5             |
|     | 345                   | 44,3            | 8,3                    | 24,7                  | 47,6             |

Table 1: Average percentage reduction linked to the frequency of the various LEDs tested.
Discussion

The results of testing LEDs at different wavelengths show that their effectiveness depends on the dissimilar sensitivity of microorganisms to ultraviolet light and on the time of irradiation, which results in a progressive reduction of the microbial load. This is consistent with most microorganisms [11]. Recent study shows that an effect that alters DNA replication is not only dependent on UV-C (200-280 nm), but also on UV-B (280-320 nm). [10] Our results have shown that UV-C, UV-B and UV-A have a reducing effect on the microbial load, albeit with important differences between them. The penetration capacity of UV-A and UV-C is different; the former does not induce direct damage to DNA, capable of forming thymine dimers, [12] but damage cellular proteins, induce the formation of reactive oxygen species such as singlet oxygen and hydrogen peroxide as also claimed by Hargreaves A et al. [13]. Noteworthy is the fact that while the DNA damage induced by UV-C can be repaired by a photolytic enzyme [14], those reported by UV-A cannot be healed in any way because the enzymes used for the repair are damaged. Our results have shown that UV-A, although to a lesser extent, partially inhibits bacterial replication. In this case, such a reduction could be assumed to be due to this mechanism.

The possibility of associating the effects of both UV-A and UV-C with bacterial replication is currently a field in which several studies are being conducted. Akgun M.P. and collaborators have recently conducted a study where the combination of UV-C and UV-A has allowed a greater effectiveness in reducing the microbial load [10,15]. In our study for Staphylococcus aureus, Enterococcus faecalis and Bacillus subtilis, critical issues were found. The number of colony-forming units (CFU) was higher than the longest exposure times to UV-C and UV-A, probably due to a sampling problem, possible limit of this pilot study. Moreover, almost all the Petri dishes showed a contamination on the circumference borders of them, probably due to their direct contact with the cone rested in the on the medium. This was particularly evident with the experiments conducted with UV-C, in which these contaminations were very sharp due the inability of the radiation to reach the resting surface of cone circumference. However, this limit could be useful to confirm a positive sowing of the Petri Dishes. This observation leads to a probable underestimation of the results, which could have had a greater percentage reduction. This small systematic inconvenience was controllable and allowed us to think about possible improvements in the experimental setting. In fact, assuming to use of 90 mm Petri dishes, instead of 55 mm in diameter, we could have, for each plate treated, a matched control of itself, in the external part of the Petri dishes.

Conclusion

The results obtained in testing the efficacy of LEDs at different wavelengths are consistent with what has been reported in recent literature. In particular, it is stressed that the greatest action with a germicidal effect is performed by UV-C rays (200-280 nm) [10], but similarly other frequencies of the light can be useful to reduce the microbial replication although efficacy decreases at increasing wavelength, progressively, from UV-C, UV-B (280- 320 nm) and finally UV-A (320- 400 nm). Moreover, the hypothesis of being able to combine and control wavelengths emitted by UV-A, UV-B and UVC suggests the possibility of having a greater effectiveness in inhibiting microbial replication by exploiting its different properties. Also, the use of these innovative sources, although still very “young” and have a large margin for improvement, can be found in numerous disinfection application [6,16,17]. LEDs have many advantages over UV lamps, including small size; impact resistance; no need to heat up to operate; low energy consumption; longer life than germicidal lamps; no mercury content; but most importantly, they emit multiple individual wavelengths [10,15]. Such an eventuality opens in fact innumerable hypotheses of study and possible applicative relapses.

Acknowledgement

We want to thank EBV Elektronik (Dr. Pierluigi Rossetti) and Seoul Viosys for providing us samples of LEDs.

References

1. Messina G, Giuseppe Sputaro, Daniele Rosadini, Sandra Burgassi, LorenzoMariani, et al. (2018) A novel approach to stethoscope hygiene: A coat-pocket innovation. Infection, Disease & Health 23(4): 211-216.
2. Lv Y, Chen L, Yu J, Xiang Q, Tang QS, et al. (2019) Hospitalization costs due to healthcare-associated infections: An analysis of propensity score matching. J Infect Public Health.
3. Kramer A, I Schwebeke, G Kampf (2006) How long do nosocomial pathogens persis on inanimate surfaces? A systematic review. BMC Infect Dis 6: 130.
4. Fattorini M, Bononcore G, Lenzi D, Burgassi S, Cardaci RMR, et al. (2018) Public Health since the beginning: Neonatal incubators safety in a clinical setting. J Infect Public Health 11(6): 788-792.
5. Weber DJ, Rutala WA, Miller MB, Hulseg K, Sickbert Bennett E, et al. (2010) Role of hospital surfaces in the transmission of emerging health care-associated pathogens: norovirus, Clostridium difficile, and Acinetobacter species. Am J Infect Control 38(Suppl 1): S25-33.
6. Messina G, Fattorini M, Nante N, Rosadini D, Serafini A, et al. (2016) Time Effectiveness of Ultraviolet C Light (UVC) Emitting by Light Emitting Diodes (LEDs) in Reducing Staphylococcus Contamination. Int J Environ Res Public Health 13(10).
7. William AR, David J Weber, Robert A Weinstein, Jane D Siegel, Michele L Pearson, et al. (2008) Guideline for Disinfection and Sterilization in Healthcare Facilities, CDC.
8. Pyrek KM (2015) Understanding the Essential of Germicidal UV Light- Infection Control Today.
9. Setlow RB, JK Setlow (1962) Evidence that ultraviolet-induced thymine dimers in DNA cause biological damage. Proc Natl Acad Sci U S A 48: 1250-1257.
10. Akgun MP, S Uralturuk (2017) Effects of ultraviolet light emitting diodes (LEDs) on microbial and enzyme inactivation of apple juice. Int J Food Microbiol 260: 65-74.
11. Kowalski W (2009) Ultraviolet Germicidal Irradiation handbook: UVGI for Air and Surface Disinfection.
12. Ravanat JL, T Douki, J Cadet (2001) Direct and indirect effects of UV radiation on DNA and its components. J Photochem Photobiol B 63(1-3): 88-102.
13. Hargreaves A, Taiwo FA, Duggan O, Kirk SH, Ahmad S (2007) Near-ultraviolet photolysis of beta-phenylpyruvic acid generates free radicals and results in DNA damage. J Photochem Photobiol B 89(2-3): 110-116.
14. Oguma K, H Katayama, S Ohgaki (2002) Photoreactivation of E. coli after low or medium-pressure UV disinfection determined by endonuclease sensitive site assay. Applied Environmental Microbiology 68(12): 6029-6035.

15. Chevremont AC, Fernet AM, Coulomb B, Boudenne JL (2012) Effect of coupled UV-A and UV-C LEDs on both microbiological and chemical pollution of urban wastewaters. Sci Total Environ 426: 304-310.

16. Messina G, Burgassi S, Messina D, Montagnani V, Cevenini G (2015) A new UV-LED device for automatic disinfection of stethoscope membranes. Am J Infect Control 43(10): e61-66.

17. Messina G, Rosadini D, Burgassi S, Messina D, Nante N, et al. (2017) Tanning the bugs - a pilot study of an innovative approach to stethoscope disinfection. J Hosp Infect 95(2): 228-230.

ISSN: 2574-1241
DOI: 10.26717/BJSTR.2019.16.002925

Messina Gabriele. Biomed J Sci & Tech Res

Assets of Publishing with us
- Global archiving of articles
- Immediate, unrestricted online access
- Rigorous Peer Review Process
- Authors Retain Copyrights
- Unique DOI for all articles

Submission Link: https://biomedres.us/submit-manuscript.php