Background: Optimal management of androgen excess in 21-hydroxylase deficiency (21OHD) remains challenging. 11-oxygenated C19 steroids (11-oxyandrogens) have emerged as promising biomarkers of disease control, but data regarding their response to treatment are lacking.

Objective: To compare the dynamic response of a broad set of steroids to both conventional oral glucocorticoids (OG) and circadian cortisol replacement via continuous subcutaneous hydrocortisone infusion (CSHI) in patients with 21OHD based on 24-hour serial sampling.

Participants and Methods: We studied 8 adults (5 women), ages 19-43 years, with poorly controlled classic 21OHD who participated in a single-center open-label phase I–II study comparing OG with CSHI. We used mass spectrometry to measure 15 steroids (including 11-oxyandrogens and Δ5 steroid sulfates) in serum samples obtained every 2 h for 24 h after 3 months of stable OG, and 6 months into ongoing CSHI.

Results: In response to OG therapy, androstenedione, testosterone (T), and their four 11-oxyandrogen metabolites:11β-hydroxyandrostenedione, 11-ketoandrostenedione, 11β-hydroxytestosterone and 11-ketotestosterone (11KT) demonstrated a delayed decline in serum concentrations, and they achieved a nadir between 0100-0300. Unlike DHEAS, which had little diurnal variation, pregnenolone sulfate (PregS) and 17-hydroxypregnenolone sulfate peaked in early morning and declined progressively throughout the day. CSHI dampened the early ACTH and androgen rise, allowing the ACTH-driven adrenal steroids to return closer to baseline before mid-day. 11KT concentrations displayed the most consistent difference between OG and CSHI across
INTRODUCTION

Congenital adrenal hyperplasia (CAH) is a set of autosomal recessive defects in cortisol biosynthesis, and of these, 21-hydroxylase deficiency (21OHD) represents the most common form (1, 2). The molecular spectrum of 21OHD is wide, ranging from absent or minimal 21-hydroxylase activity, to fully compensated enzymatic defects (3). Conventionally, 21OHD forms associated with clinically overt adrenal insufficiency are collectively grouped into “classic” 21OHD, while cases with normal or near normal cortisol production are termed “non-classic” (3). The traditional biomarkers of 21OHD control are the androgen precursors 17α-hydroxyprogesterone (17OHP) and androstenedione (A4).

All forms of 21OHD are characterized by variable excess in adrenal androgen synthesis, which is driven by ACTH. Clinical manifestations of hyperandrogenism range from severe virilization, premature adrenarche or puberty, to hirsutism, acne, and infertility. Glucocorticoids have been the mainstay of treatment for patients with classic 21OHD, not only as replacement therapy for adrenal insufficiency, but also to lower ACTH and the resulting androgen surplus. With conventional oral glucocorticoid (OG) therapy, however, supra-physiological doses and nighttime administration are typically needed to reduce the early morning ACTH rise. Supra-physiologic glucocorticoid use contributes to higher rates of obesity, metabolic syndrome, and bone mass decline in adults with 21OHD (4, 5). Efforts towards optimizing the management of hyperandrogenism in patients with 21OHD while minimizing glucocorticoid daily dose include the development of modified-release OG (6) and continuous glucocorticoid delivery systems (7, 8), as well as a variety of non-glucocorticoid therapies (9).

Continuous subcutaneous hydrocortisone infusion (CSIH) that approximates circadian cortisol secretion was previously shown to improve disease control in patients with 21OHD poorly controlled with OG therapy based on the traditional biomarkers 17OHP and A4 (7).

Additional challenges in the management of 21OHD derive from the uncertainties surrounding the optimal use of biomarkers to assess disease control (10, 11). The conventional steroid panel of patients with 21OHD has been expanded in the recent years, to include previously neglected androgens (12–14). In particular, a set of 11-oxygenated−C19 steroids (11-oxyandrogens), including 11β-hydroxyandrostenedione (11OHA4) and 11-ketotestosterone (11KT), have been shown to be elevated in patients with classic (12, 15) and non-classic 21OHD (16). 11-oxyandrogens were found to correlate with parameters of poor 21OHD control (11, 17). The dynamic responses of 11-oxyandrogens to glucocorticoid therapy have not been characterized, and this aspect will be key for clinical interpretation. Herein, we compared the 24-hour response of serum 11-oxyandrogens and conventional androgens to standard OG vs. CSHI in patients with classic 21OHD.

SUBJECTS AND METHODS

Patients and Study Design

Eight adult patients (five women) with difficult to control classic 21OHD seen at the National Institutes of Health (NIH), Bethesda, Maryland were included in an open-label phase II study (NCT01859312) comparing CSHI with conventional OG, as previously described (7). The diagnosis of classic 21OHD was confirmed by hormonal and genetic analyses in all participants.

All patients were treated with stable OG doses for at least 3 months prior to study entry. For the CSHI, the total daily hydrocortisone dose was calculated based on the patient’s estimated cortisol clearance, which was conducted in each patient at baseline (7). Patients were admitted for frequent hormonal sampling (every 2 hours) on two separate occasions: at study entrance, while receiving conventional OG, and 6 months after initiation of CSHI. The OG equivalent dose (mg/m2/day) was calculated using a factor of 1 for hydrocortisone, 5 for prednisone and prednisolone, and 80 for dexamethasone (5).

Hormonal Assays

We quantified 15 steroids by liquid chromatography-tandem mass spectrometry (LC-MS/MS), including: cortisol, 17OHP, 16α-hydroxyprogesterone (16OHP), 21-deoxy cortisol (21dF), progesterone, A4, testosterone (T), 11β-hydroxyandrostenedione (11OHA4), 11-ketoandrosterone (11KA4), 11β-hydroxytestosterone (11OHT), 11-ketotestosterone (11KT), pregnenolone sulfate (PregS), 17-hydroxyprogrenolone sulfate (17OH-PregS), dehydroepiandrosterone sulfate (DHEAS) and androst-5-ene-3β,17β-diol sulfate (AdiolS). Steroid extraction and quantitation were carried out as previously described (12, 13). Plasma ACTH was analyzed at the NIH Clinical Center, Bethesda, Maryland by chemiluminescent immunoassay on Siemens Immulite 200 XPi analyzer (7).
Statistical Analyses
Logarithmic transformation was used to achieve normal data distribution where needed. The linear-up log-down trapezoidal rule was used to calculate the areas under the curve (AUC) for the 24 h periods studied, as well as for the morning (0500-1100h) and evening (1700-2300h) segments. Wilcoxon signed-rank test was used to compare AUCs between the two therapies. We assessed correlations between steroids and ACTH with the Spearman correlation test. Data analysis was conducted with R 3.6.2 (R Foundation for Statistical Computing). Two-tailed p values <0.05 were deemed statistically significant.

RESULTS
Of the eight participants, five were women, age 19–42 years, and three men, age 25–43 years. The median BMI was 40.1 kg/m² (range: 27.7–54.1 kg/m²). While receiving standard OG therapy, the glucocorticoid equivalent doses ranged from 8.8 to 26.5 mg/m²/day.

During conventional OG therapy, ACTH demonstrated a sharp peak around 0700h with a nadir 2300-0300h, which was mirrored by 17OHP, 16OHP and 21dF (Figure 1). A4, T, and their four 11-oxyandrogen metabolites demonstrated a delayed decline in serum concentrations, and they achieved a nadir between 0100-0300h (Figure 1). Not surprisingly, DHEAS and AdiolS displayed relatively flat circadian levels, with no significant differences between morning and nocturnal concentrations (Figure 1 and Table 1); conversely, PregS and 17OHPregS peaked in early morning and declined progressively throughout the day.

Overall, most steroid concentrations were lower during CSHI as compared with OG therapy (Figure 1 and Table 2). Following a more subtle morning ACTH peak during CSHI, steroids with diurnal fluctuations returned to baseline levels around 1100 (Figure 1) and remained stable throughout the rest of the day. Notably, while T was lowered by CSHI as compared with OG in women, T increased in men, suggesting an improvement of the testicular function in parallel with 21OHD control in men (Figure 2).

Of the steroids measured, 17OHP and 16OHP correlated best with ACTH (r=0.81 and 0.77, respectively, p<0.0001), followed by A4 (r=0.68), PregS (r=0.62), and 11KT (r=0.61, p<0.0001 for

![FIGURE 1](# | Serum concentrations of ACTH and steroids during conventional oral glucocorticoid therapy (OG, circles), and into 6 months of continuous subcutaneous hydrocortisone infusion (CSHI, triangles). Central data represent the geometrical means of all participants, and the vertical lines represent the standard errors of the geometrical means. 21dF, 21-deoxycorticisol; 17OHProg, 17α-hydroxyprogesterone; 16OHProg, 16α-hydroxyprogesterone; A4, androstenedione; 11OHA4, 11β-hydroxyandrostenedione; 11KT, 11-ketotestosterone; T, testosterone PregS, pregnenolone sulfate; 17OHPregS, 17α-hydroxypregnenolone sulfate; DHEAS, dehydroepiandrosterone sulfate; AdiolS, Androstenediol-3-sulfate.)
all). A4 and its 11-oxygenated derivatives correlated tightly among each other, as well as with 11OHT and 11KT (Table 3). Conversely, 11KT correlated directly with T only in women, while in men, 11KT and T displayed an inverse correlation, similar to our previous studies (12, 17), supporting their origin from independent sources in well-controlled men.

### DISCUSSION

The circadian rhythmicity of cortisol is regulated via ACTH (18) as dictated by the hypothalamic suprachiasmatic nucleus (19). Physiologic cortisol synthesis has a characteristic circadian pattern, with an early morning peak and a late-night nadir (20–22). In patients with classic 21OHD, the early morning rise in ACTH is further amplified by the reduced negative feedback from cortisol. Like cortisol, 11-oxyandrogens are synthesized via the adrenal cortical enzyme 11β-hydroxylase (CYP11B1), which is regulated by ACTH (23). Although 11KT is produced primarily from adrenal-derived 11OHA4 via peripheral metabolism, we found that 11KT still correlates highly with ACTH, in agreement with an earlier study (17). In healthy individuals, 11-oxyandrogens display

---

**Table 1** | Comparison of steroid concentrations between 0700h and 0100h during oral glucocorticoid treatment.

| Hormone | Time segment | OG AUC (ng/dL x h) | CSHI AUC (ng/dL x h) | p value |
|---------|--------------|-------------------|---------------------|--------|
| 21dF    | 0100 h       | 204.7 [71.7, 1079.4] | 864.6 [530.6, 2889] | 0.008  |
| 16OHProg| 0100 h       | 16.3 [14.7, 147.7]  | 252.8 [185, 1313.7] | 0.008  |
| 17OHProg| 0100 h       | 243.5 [151.4, 1592] | 3445.3 [2080.1, 11789.9] | 0.006  |
| T       | 0100 h       | 58.4 [24.8, 91.7]   | 74 [32, 126.9]      | 0.023  |
| A4      | 0100 h       | 104.3 [66.1, 214.8] | 189.3 [111.1, 459.7] | 0.016  |
| Prog    | 0100 h       | 24.5 [7.5, 52.3]    | 138.5 [28.8, 204.1] | 0.016  |
| 110HT   | 0100 h       | 24.2 [11.9, 52.1]   | 42.5 [32.5, 91]     | 0.039  |
| 11KT    | 0100 h       | 83.7 [49.6, 197.2]  | 145.8 [125.8, 292.6] | 0.008  |
| 11OHA4  | 0100 h       | 97.4 [64.3, 507.5]  | 301.5 [122.4, 955.8] | 0.039  |
| 11KA4   | 0100 h       | 16.4 [12.3, 77.8]   | 46.7 [29.2, 133.3]  | 0.008  |
| PregS   | 0100 h       | 820.9 [661.8, 1490.1] | 3104.8 [745.7, 4940] | 0.039  |
| 17OHProg| 0100 h       | 292.9 [220, 1343.5] | 3919.3 [973.9, 9506.8] | 0.055  |
| DHEAS   | 0100 h       | 17974.4 [3627.5, 32826.8] | 1740.4 [4453.5, 34320.6] | 0.742  |
| AdiolS  | 0100 h       | 3919.3 [973.9, 9506.8] | 3935.2 [1124.5, 7723.7] | 1      |

Data are expressed as medians and interquartile ranges. Comparisons of baseline 0700h vs. 0100h were done with the Wilcoxon signed-rank test.

**Table 2** | Comparison of steroid concentrations during conventional oral glucocorticoid therapy (OG) vs. continuous subcutaneous hydrocortisone infusion (CSHI).

| Hormone | Time segment | OG AUC (ng/dL x h) | CSHI AUC (ng/dL x h) | p value |
|---------|--------------|-------------------|---------------------|--------|
| 21dF    | 24 h         | 12582.6 [7762.4, 36130.4] | 4513.6 [2944.8, 17916.7] | 0.023  |
| 16OHProg| 24 h         | 3077.2 [1554.8, 12975.3] | 1568.5 [545.2, 2378.7] | 0.055  |
| 17OHProg| 24 h         | 40649.6 [18889.8, 112674.8] | 12074 [6734.9, 18718.8] | 0.023  |
| T       | 24 h         | 1416.5 [725.8, 2379.3] | 725 [562, 2344.8] | 0.641  |
| A4      | 24 h         | 439.1 [187.8, 779.9] | 190.5 [140.2, 278.6] | 0.641  |
| Prog    | 24 h         | 2055.5 [989.1, 10706.6] | 353.7 [94.7, 12189.2] | 0.641  |
| 110HT   | 24 h         | 11567.1 [3900.8, 16714] | 1778 [1226.8, 831] | 0.844  |
| 11KT    | 24 h         | 1100 h            | 105 [14.7, 2372.9] | 0.008  |
| 11OHA4  | 24 h         | 5439.8 [1819.4, 3864.8] | 2794 [1719.2, 1314.9] | 0.109  |
| 11KA4   | 24 h         | 5072.9 [1620.8, 8198.3] | 593.3 [404.5, 675.2] | 0.055  |
| PregS   | 24 h         | 1502.9 [469.4, 27020.5] | 27883.1 [5685, 30451.4] | 0.023  |
| 17OHProg| 24 h         | 13783.5 [7435.8, 52818.6] | 8054.5 [4519.1, 13381.3] | 0.008  |
TABLE 2 | Continued

| Hormone | Time segment | OG AUC (ng/dL x h) | CSHI AUC (ng/dL x h) | p value |
|---------|--------------|--------------------|----------------------|---------|
| DHEAS   | 0500 – 1100 h | 4260.6 [1401.1, 17786.5] | 2548.3 [1447.6, 4540.2] | 0.023 |
|         | 1700 – 2300 h | 3148.5 [1428.9, 11432.8] | 1739.8 [752.7, 2173] | 0.008 |
|         | 24 h          | 278225.1 [110016.9, 921487] | 177721.3 [53305.4, 425090.4] | 0.008 |
| AdiolS  | 0500 – 1100 h | 56092.5 [29166.4, 210158.1] | 47525.4 [12323.5, 117190.3] | 0.039 |
|         | 1700 – 2300 h | 62665.7 [27821.9, 263578.4] | 43929 [14807.3, 81494.3] | 0.008 |
| ACTH    | 0500 – 1100 h | 89933.2 [25443.2, 181370.8] | 62435.3 [13767.9, 129204.2] | 0.039 |
|         | 1700 – 2300 h | 19499.9 [5331.1, 44425.2] | 10673.3 [5963.9, 29584.9] | 0.055 |

Data are expressed as medians and interquartile ranges of areas under the curves (AUC).

FIGURE 2 | Serum concentrations of testosterone (T) and 11-ketotestosterone (11KT) during conventional oral glucocorticoid therapy (OG, circles), and into 6 months of continuous subcutaneous hydrocortisone infusion (CSHI, triangles), in women (N=5) and men (N=3). Central data represent the geometrical means, and the vertical lines represent the standard errors of the geometrical means.
morning peaks and nocturnal nadirs, similar to cortisol (22). In a recent study of patients with classic 21OHD, salivary 11OHA4 and 11KT were highest in the morning, and a recent study of patients with classic 21OHD, salivary after 1100h, reflecting the improved disease control achieved with circadian cortisol replacement.

11KT is a potent androgen, with bioactivity similar to that of T, which also derives from the gonads, 11KT reflecting the adrenal component of androgen production alone. 11KT is particularly useful in men with 21OHD, as both steroids are produced primarily by the adrenal glands. The serum concentrations of 11-oxyandrogens return to baseline levels after 1100h, reflecting the improved disease control achieved with circadian cortisol replacement.

11KT is a potent androgen, with bioactivity similar to that of T, which also derives from the gonads. 11KT reflecting the adrenal component of androgen production alone. 11KT is particularly useful in men with 21OHD, as both steroids are produced primarily by the adrenal glands. The serum concentrations of 11-oxyandrogens return to baseline levels after 1100h, reflecting the improved disease control achieved with circadian cortisol replacement.

11KT is a potent androgen, with bioactivity similar to that of T, which also derives from the gonads. 11KT reflecting the adrenal component of androgen production alone. 11KT is particularly useful in men with 21OHD, as both steroids are produced primarily by the adrenal glands. The serum concentrations of 11-oxyandrogens return to baseline levels after 1100h, reflecting the improved disease control achieved with circadian cortisol replacement.

11KT is a potent androgen, with bioactivity similar to that of T, which also derives from the gonads. 11KT reflecting the adrenal component of androgen production alone. 11KT is particularly useful in men with 21OHD, as both steroids are produced primarily by the adrenal glands. The serum concentrations of 11-oxyandrogens return to baseline levels after 1100h, reflecting the improved disease control achieved with circadian cortisol replacement.

Although quantitatively abundant, DHEA and DHEAS have virtually no androgenic activity (25). Moreover, DHEAS is a poor biomarker of 21OHD control, which can be paradoxically low in poorly controlled classic 21OHD patients (12, 29). This enigma could be partly explained by the relative shift in sulfotransferase type 2A (SULT2A1) substrates, as suggested by the higher PregS serum concentrations in patients with 21OHD vs. age and BMI-matched controls (12). We have previously shown both in vivo and in vitro, that in contrast with DHEAS and AdiolS, PregS responds acutely to ACTH stimulation (30). PregS has also been found to be associated with clinical surrogates of poor 21OHD control (17). Thus, PregS could be more relevant than DHEAS in the clinical evaluation of patients with 21OHD.
Practices vary regarding the laboratory evaluation of patients with 21OHD, and 24-hour sampling is only done in a research setting. Monitoring treatment by consistently timed hormone measurements relative to the time of glucocorticoid administration is recommended (31), as glucocorticoid acutely suppress adrenal steroid production. Although 17OHP and A4 show good correlation, we previously found that 17% of laboratory evaluations done in the early morning resulted in discrepant findings between these two traditional biomarkers (11). 11-oxyandrogens are the major androgens and androgen precursors found in classic and nonclassic 21OHD (12, 16) and likely provide more direct evidence of adrenal androgen excess. However, their use in the medical management and monitoring of 21OHD has yet to be determined. Our study of 24-hour steroid profiles of these newly defined biomarkers in addition to the traditional biomarkers provides insight into the 24-hour rhythms of adrenal androgens in patients with classic 21OHD. Our findings suggest that 11-oxyandrogens remain elevated for a longer period of time than 17OHP in patients with poor disease control and demonstrate a circadian rhythm.

In summary, this is the first study to show 24-hour profiles of 11-oxyandrogens and related Δ4- and Δ5-steroids in patients with 21OHD. We directly compared the dynamic responses of 21OHD biomarkers, including adrenal 11-oxyandrogens and Δ4 steroid sulfates, to both conventional OG therapy and CSHI. We found that the high amplitude ACTH morning rise that occurs in patients treated with OG leads to sharp elevations of several adrenal steroids, particularly those immediately preceding the enzymatic blockade. Following such marked ACTH stimulation, 11-oxyandrogen elevations decline rather slowly throughout the day. This finding has clinical implications, especially since 11KT has androgenic activity equivalent to T. Conversely, mimicking physiologic cortisol concentrations via CSHI dampens the early ACTH rise, allowing ACTH-driven adrenal steroids to return closer to baseline before mid-day. In addition to ACTH, these dynamic steroid excursions are also impacted by protein binding and metabolism, which remain understudied. Additionally, the small number of patients and lack of nonclassic 21OHD and healthy individuals as comparators limit the extrapolation of the absolute values for broader use. Nevertheless, our data reinforce the potential utility of 11-oxyandrogens as biomarkers of disease control. The development of normative data and the establishment of 21OHD monitoring recommendations for the promising biomarkers – both 11-oxyandrogens and PregS – needs to consider their diurnal variability.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by National Institutes of Health Institutional Review Board. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

AM, AAN, and DM were investigators in the clinical study. AT proposed the current study. ATN performed mass spectrometry steroid assays. XC, LZ, JB, and AT analyzed the data. AT, DM, and RA interpreted the data. AT drafted the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported, in part, by grants 1K08DK109116 (to AT), K23HL128909 (to JB), and R01GM086596 (to RA) from the National Institutes of Health, by Michigan Institute for Clinical and Health Research (MICHRI Translational Science Award/ U046500 to AT and, in part, by the Intramural Research Program of the National Institutes of Health.

ACKNOWLEDGMENTS

We thank the patients for their participation in this study and the nursing staff of the 5SWN Metabolic Unit for their assistance in conducting the study. We also thank Amira Rabbah, for assistance with constructing the figures.

REFERENCES

1. Claahsen-van der Grinten HL, Speiser PW, Ahmed SF, Arlt W, Aucius RJ, Falhammar H, et al. Congenital Adrenal Hyperplasia - Current Insights in Pathophysiology, Diagnostics and Management. Endocr Rev (2021) bnb016. doi: 10.1210/endrev/bnb016
2. Turcu AF, Aucius RJ. Adrenal Steroidogenesis and Congenital Adrenal Hyperplasia. Endocrinol Metab Clin North Am (2015) 44(2):275–96. doi: 10.1016/j.ccm.2015.02.002
3. Merke DP, Aucius RJ. Congenital Adrenal Hyperplasia Due to 21-Hydroxylase Deficiency. N Engl J Med (2020) 383(13):1248–61. doi: 10.1056/NEJMc1909786
4. Arlt W, Willis DS, Wild SH, Krone N, Doherty EJ, Hahner S, et al. Health Status of Adults With Congenital Adrenal Hyperplasia: A Cohort Study of 203 Patients. J Clin Endocrinol Metab (2010) 95(11):5110–21. doi: 10.1210/jc.2010-0917
5. Finkielstain GP, Kim MS, Sinaei N, Nishitani M, Van Ryzin C, Hill SC, et al. Clinical Characteristics of a Cohort of 244 Patients With Congenital Adrenal Hyperplasia. J Clin Endocrinol Metab (2012) 97(12):4429–38. doi: 10.1210/jc.2012-2102
6. Merke DP, Mallappa A, Arlt W, Brac de la Perriere A, Linden Hirschberg A, Juul A, et al. Modified-Release Hydrocortisone in Congenital Adrenal Hyperplasia. J Clin Endocrinol Metab (2021) 106(5):e2063–e277. doi: 10.1210clinem/dgab051
