Visualization of Phosphoinositides That Bind Pleckstrin Homology Domains: Calcium- and Agonist-induced Dynamic Changes and Relationship to Myo-[3H]inositol-labeled Phosphoinositide Pools

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Abstract. Phosphatidylinositol 4,5-bisphosphate (PtdIns[4,5]P_2) pools that bind pleckstrin homology (PH) domains were visualized by cellular expression of a phospholipase C (PLC) δ PH domain–green fluorescent protein fusion construct and analysis of confocal images in living cells. Plasma membrane localization of the fluorescent probe required the presence of three basic residues within the PLC δ PH domain known to form critical contacts with PtdIns(4,5)P_2. Activation of endogenous PLCs by ionophores or by receptor stimulation produced rapid redistribution of the fluorescent signal from the membrane to cytosol, which was reversed after Ca^{2+} chelation. In both ionomycin- and agonist-stimulated cells, fluorescent probe distribution closely correlated with changes in absolute mass of PtdIns(4,5)P_2. Inhibition of PtdIns(4,5)P_2 synthesis by quercetin or phenylarsine oxide prevented the relocalization of the fluorescent probe to the membranes after Ca^{2+} chelation in ionomycin-treated cells or during agonist stimulation. In contrast, the synthesis of the PtdIns(4,5)P_2 imaged by the PH domain was not sensitive to concentrations of wortmannin that had been found inhibitory of the synthesis of myo-[3H]inositol–labeled PtdIns(4,5)P_2. Identification and dynamic imaging of phosphoinositides that interact with PH domains will further our understanding of the regulation of such proteins by inositol phospholipids.

Key words: phosphoinositides • calcium • green fluorescent protein • pleckstrin-homology domain • phospholipase C
Inositol phospholipids were analyzed from either COS-7 cells transfected with the PH domain–phosphoinositide interaction was how the binding of PH domains to the inositol headgroup affects the primary signaling functions of PtdIns(4,5)P2, particularly its availability to PLC enzymes, or to PtdIns 3 kinases. It is known that other proteins that bind PtdIns(4,5)P2, such as profilin, greatly impair the ability of PLC enzymes to hydrolyze this lipid.

The present studies were designed to examine whether specific association of the PLCβ PH domain (PHβCLδ) with PtdIns(4,5)P2 can be exploited to visualize the cellular distribution of phosphoinositides, that are capable of binding PH domains in intact cells, and to follow their changes in real time after stimulation. Our results, obtained using a fusion protein of the PHβCLδ coupled to the green fluorescence protein (GFP), provide imaging of the dynamics of membrane PtdIns(4,5)P2 pools that interact with PH domains. They also reveal a prominent difference between the regulation of these pools compared with those labeled with myo-[3H]inositol.

Materials and Methods

Materials

Angiogenins I and II; (human) was obtained from New Life Health Technologies, Inc. (San Carlos, CA) and PDGF (recombinant, AB) from Life Technologies, Inc. (Gaithersburg, MD). Thapsigargin, ionomycin, and 1,2-bis(2-aminophenoxy)ethane N,N,N’,N’-tetraacetic acid (BAPTA) were purchased from Calbiochem-Novabiochem, Corp. (La Jolla, CA), and Wurtmann was a gift from Kyowa Hakko Laboratories (Tokyo, Japan). 2,3-Dimercaptopropanol (BAL), phenylarsine oxide, and quercetin were purchased from Calbiochem-Novabiochem, Corp. (La Jolla, CA) from human cDNAs (marathon cDNA from brain and K562 leukemia cells). Another question that is raised by studies on PH domain–phosphoinositide interactions is how the binding of PH domains to membranes. A small pool was introduced into the pH domain sequence which changed S34 to a T but this substitution did not change any characteristic compared with the wild-type protein. All mutations were confirmed by dye-exchange sequencing and the expression of the fusion protein by Western blot analysis.

Transfection of Cells for Confocal Microscopy

Cells were plated onto poly-L-lysine–coated 30-mm-diam circular cover slips at a density of 5 × 10^4 cells/dish and cultured for 3 d before transfection with plasmid DNAs (1 μg/ml) using the Lipofectamine reagent (10 μg/ml). Life Technologies, Inc.) and OPTI-MEM (Life Technologies, Inc.). 48 h after transfection cells were washed twice with a modified Krebs-Ringer solution, containing (mM): NaCl 120, KCl 4.7, CaCl2 1.2, MgSO4 0.7, glucose 10, Na-Hepes 10, pH 7.4, and the coverslip was placed into a chamber that was mounted on a heated stage with the medium temperature kept at 33°C. Cells were incubated in 1 ml of the Krebs-Ringer buffer and the stimuli were added in 0.5 ml prewarmed buffer after removing 0.5 ml medium from the cells. Cells were examined in an inverted microscope under a 40× oil-immersion objective (Nikon, Inc., Melville, NY) and a BioRad laser confocal microscope system (MRC-1024) with the LaserSharp acquisition software (Bio-Rad Laboratories, Hercules, CA). All pictures presented are original recordings without postacquisition enhancing.

Analysis of Inositol Lipids

Inositol phospholipids were analyzed from either COS-7 cells transfected with the AT1a Ang II receptor together with selected GFP–PH domain fusion constructs, or from untransfected NIH-3T3 cells after labeling with myo-[3H]inositol for 24 h (COS-7 cells) or 48 h (NIH-3T3 cells) as previously described (4, 17). [3H]Inositol phosphates were analyzed by HPLC (17) or, in the COS cell experiments, by Dowex micromulds. PtdIns(4,5)P2 mass was determined from NIH-3T3 cells and bovine adrenal glomerulosa (BAG) cells that were cultured on 12-well plates. After stimulation at 33°C, reactions were terminated with 400 μl ice-cold perchloric acid (5% final) followed by an acidic lipid extraction (3). Samples were taken to dryness with N2 stream and were subjected to alkaline hydrolysis to liberate Ins(1,4,5)P3, from PtdIns(4,5)P2 for quantitation in a radioreceptor assay, essentially as described in Chailly (see reference 6) using bovine adrenal cortical membranes (15) and [3H]Ins(1,4,5)P3 (Amerham Corp.).

Results

Plasma Membrane Localization of the PLCβ PH Domain Requires Interaction with PtdIns(4,5)P2

Among the known PH domains reported to interact with PtdIns(4,5)P2, the PH domain of PLCδ has the highest affinity (12). Therefore, we chose this PH domain to create a fluorescent probe for imaging purposes and fused it to the NH2 terminus of GFP. Expression of GFP alone showed no specific cellular localization and was present in the cytosol, as expected from a protein of its size (27 kD) lacking a localization signal. Addition of the PH domain of PLCδ (1–170) to the GFP (PHδ–GFP) was sufficient to localize the construct to the plasma membrane when transiently expressed in various cell types (Fig. 1). A small amount of fluorescence was always present in the cytosol and the nucleus, but very little in internal membranes apart from some vesicular structures that could be membrane invaginations. Although this result was consistent with binding of the construct to membrane PtdIns(4,5)P2,
PtdIns(4,5)P2 pools, then hydrolysis of these phospholipids by endogenous PLCs should change the distribution of the fusion protein to report those phosphoinositide changes. PLC was activated in intact NIH-3T3 cells by treatment with ionomycin (10 μM) in the presence of external Ca\(^{2+}\) (1.2 mM) to allow a large increase in cytosolic Ca\(^{2+}\) concentration ([Ca\(^{2+}\)]) with sequential activation of PLC isoforms (30). As shown in Fig. 2, this manipulation, indeed, caused rapid disappearance of fluorescence from the plasma membrane and its simultaneous appearance in the cytosol, consistent with PLC-mediated hydrolysis of the membrane PtdIns(4,5)P2 at high [Ca\(^{2+}\)]. Once released into the cytosol, this 40-kD fusion construct only slowly appeared in the nucleus. Preincubation of the cells with neomycin (10 mM) for 10 min to inhibit the hydrolysis of PtdIns(4,5)P2 (8), completely prevented the release of the fluorescent signal from the membranes in the majority (>90%) of cells (not shown).

Subsequent chelation of external Ca\(^{2+}\) by either BAPTA or EGTA (2 mM) caused the slow reappearance of the fluorescence at the plasma membrane and, most notably, in some intracellular membrane structures (Fig. 2 B). In many cells, the first signs of relocation were the appearance of bright foci at perinuclear membranes and in other intracellular structures that eventually translocated to the plasma membrane (Fig. 2 B). These changes were considered to reflect the resynthesis of the phosphoinositide pools as PLC activity declined in concert with the falling [Ca\(^{2+}\)] (see below).

To analyze whether Ca\(^{2+}\) release from intracellular stores without Ca\(^{2+}\) influx was sufficient to cause the breakdown of membrane phosphoinositides that bind PH domains, we applied ionomycin to the transfected cells in Ca\(^{2+}\)-free medium. Under these conditions ionomycin failed to evoke a lipid signal that could be detected by the redistribution of the fluorescent construct (not shown). To calculate the concentration of Ca\(^{2+}\) that was required to activate PtdIns(4,5)P2 breakdown in the presence of ionomycin, we titrated extracellular Ca\(^{2+}\) in the presence of 2 mM BAPTA and measured [Ca\(^{2+}\)] under identical conditions in Fura-2-loaded NIH-3T3 cells. Based on these measurements, the concentration of Ca\(^{2+}\) that was required to activate PtdIns(4,5)P2 hydrolysis (assessed by the release of fluorescence from the membrane into the cytosol) was around 10 μM. The need for relatively high Ca\(^{2+}\) to trigger this process was also indicated by the finding that lower concentrations of ionomycin (1 μM), that evoke rapid Ca\(^{2+}\) release with consequential activation of the capacitive Ca\(^{2+}\) entry pathway, did not induce translocation of the fluorescent construct (not shown). Similarly, thapsigargin (100 nM) or PDGF (25–100 ng/ml), both of which evoked moderate [Ca\(^{2+}\)] increases in NIH-3T3 cells, had no effect on the distribution of the fusion protein (not shown).

**Ca\(^{2+}\)-induced Changes in Cellular PtdIns(4,5)P2 Mass Closely Correlate with Plasma Membrane Localization of PLC\(\delta\) PH Domain**

To examine how the cellular PtdIns(4,5)P2 pool is affected by the same manipulations that caused the redistribution of the fusion construct, we analyzed the total mass of this phospholipid from NIH-3T3 cells after ionomycin treat-

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**Figure 1.** Membrane localization of PLC\(\delta\) PH domain–GFP fusion protein in transfected NIH-3T3 cells. The PH domain of PLC\(\delta\) (1-170) was fused to the NH\(_2\) terminus of the GFP protein and this construct or its mutants were transiently expressed in NIH-3T3 cells. Confocal images were obtained in living cells at 33°C. For technical reasons, the K30L, K32L, and R40L mutants also carried the S34T mutation but the latter alone was without effect on localization. A partial sequence of the PLC\(\delta\) PH domain containing the first three β strands is shown with the mutated residues indicated by arrows. Residues that form critical contacts with Ins(1,4,5)P3 (12), are labeled by small squares. Bar, 10 μm.

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ment and subsequent Ca\(^{2+}\) chelation. The total mass of PtdIns(4,5)P\(_2\) in cultured cells was measured by lipid extraction followed by alkaline hydrolysis to liberate Ins(1,4,5)P\(_3\), which was then quantitated by a radioreceptor assay (6). As shown in Fig. 2C, ionomycin (10 \(\mu M\)) caused rapid breakdown of PtdIns(4,5)P\(_2\), and upon Ca\(^{2+}\) chelation the level of this phospholipid rapidly returned nearly to its original value within 5 min.

To analyze the correlation between PtdIns(4,5)P\(_2\) changes and the redistribution of fluorescence, we needed to quantify membrane localization. Dividing the fluorescence of the membrane with that of the adjacent cytosol (values were taken from line intensity histograms on series of images) proved to be a useful index of membrane-association of the construct. As shown in Fig. 2D, PtdIns(4,5)P\(_2\) changes and membrane localization of the fluorescent construct correlated remarkably closely during manipulation of intracellular Ca\(^{2+}\), except for a clear delay observed in the plasma membrane reappearance of the fluorescence. This, however, was preceded by localization of the construct at intracellular sites (Fig. 2D), indicating that the earliest sites of PtdIns(4,5)P\(_2\) synthesis are at intracellular membrane compartments.

Ca\(^{2+}\)-mobilizing Hormones Can Also Initiate the Breakdown of the PtdIns(4,5)P\(_2\) Pool That Binds PLC\(\delta\) PH Domain

To investigate whether the PtdIns(4,5)P\(_2\) pools that are visualized by PH\(_{PLC\delta}\)-GFP show any change after agonist stimulation, we examined the effect of the calcium mobilizing hormone, Ang II, on the redistribution of the fluorescent probe in various cells. The fluorescent construct was transfected together with the AT\(_{1a}\) Ang II receptor into COS-7 or NIH-3T3 cells, or alone into primary cultures of adrenal glomerulosa cells that express endogenous AT\(_{1}\) receptors. Stimulation of the cells with Ang II caused a rapid release of the fluorescent signal from the plasma membrane to the cytosol in each of the three cell types (Figs. 3 and 4). This process was very rapid and, in contrast to ionomycin stimulation, was only transient with the construct partially relocalizing to the plasma membrane in spite of the sustained stimulation. Simultaneous changes in PtdIns(4,5)P\(_2\) mass measured in BAG cells showed again a close correlation between changes in membrane-associated fluorescence and the absolute amounts of this lipid (Fig. 4, B and C). In contrast to the effects of ionomycin, the effect of Ang II on PH\(_{PLC\delta}\)-GFP distribution was still observed in a medium nominally free of Ca\(^{2+}\), although this effect was very short-lived, the fluorescence being relocalized to the membrane within 2 min (not shown). These results indicate that stimulation of a G protein-coupled receptor can cause the breakdown of the PtdIns(4,5)P\(_2\) pool(s) that bind PH domains.

Inhibition of the Resynthesis of PtdIns(4,5)P\(_2\) Prevents Relocalization of the PLC\(\delta\) PH Domain to the Plasma Membrane

To further examine the interaction between the PLC\(\delta\) PH domain and the plasma membrane phosphoinositides, we used inhibitors that have been reported to inhibit the formation of PtdIns(4,5)P\(_2\) from PtdIns by inhibiting PtdIns 4 and PtdIns(4)P 5 kinases (34). These inhibitors were tested...
for their ability to prevent the resynthesis of PtdIns(4,5)P$_2$ after ionomycin treatment and subsequent Ca$^{2+}$ chelation. As shown in Fig. 5, both quercetin (100 µM) and phenylarsine oxide (PAO; 100 µM) were each able to prevent the resynthesis of PtdIns(4,5)P$_2$ as measured by the total mass of this phospholipid. The inhibitory effect of PAO was completely reversed by DTT, only partially by BAL and not by β-mercaptoethanol (1 mM each). These same treatments equally affected the relocalization of the fluorescent probe to the plasma membrane upon Ca$^{2+}$ chelation after ionomycin treatment: both quercetin and PAO prevented the reappearance of the fluorescent signal in membranes, and DTT but not β-mercaptoethanol, were each able to antagonize the inhibitory effect of PAO (Fig. 5B). Interestingly, although BAL only partially restored PAO-inhibited PtdIns(4,5)P$_2$ resynthesis, it fully restored the translocation of PH$^{PLC	ext{-}d}$–GFP. In bovine glomerulosa cells that were pretreated with quercetin (or with PAO), Ang II induced a more permanent release of the fluorescent construct into the cytosol instead of the transient translocation observed in control cells, indicating that once the resynthesis of the PtdIns(4,5)P$_2$ is inhibited, the Ca$^{2+}$-mobilizing agonist causes a more complete degradation of membrane PtdIns(4,5)P$_2$ (Fig. 4, B and C).

### The Resynthesis of PtdIns(4,5)P$_2$ That Binds PH Domains Is Not Sensitive to WT Inhibition

Interestingly, high concentrations of wortmannin (WT; 10 µM) shown to inhibit type III PtdIns 4-kinases (27) did not affect relocalization of the fluorescent construct after Ca$^{2+}$ chelation in ionomycin-treated cells, although it partially inhibited PtdIns(4,5)P$_2$ resynthesis measured by the mass assay (Fig. 5). Similarly, 10 µM WT had no effect on the Ang II–induced transient translocation of the fluorescent signal (not shown). This result contrasted our previous finding that the metabolically labeled hormone-sensitive PtdIns(4,5)P$_2$ pools are maintained by WT-sensitive PtdIns 4-kinase(s) in several agonist-stimulated cells and that stimulation of these cells in the presence of 10 µM WT leads to the depletion of these phosphoinositide pools in myo-[3H]inositol–labeled cells (27).

These findings suggest that although the myo-[3H]inositol–labeled, agonist-sensitive PtdIns(4,5)P$_2$ pools are formed by WT-sensitive PtdIns 4-kinase(s), additional PtdIns(4,5)P$_2$ pools that bind PH domains are synthesized by WT-insensitive mechanisms that are only inhibited by less-specific PI kinase inhibitors, such as quercetin or PAO (34).
PtdIns(4,5)P2 pools that bind PH domains are largely overlapping and that the resynthesized PtdIns(4,5)P2 is still available for agonist-induced hydrolysis.

**PH Domains Can Inhibit the Agonist-induced Hydrolysis of Myo-[3H]inositol–labeled PtdIns(4,5)P2**

Since PH domains bind to and therefore cover the inositol phosphate headgroup of phosphoinositides, they can hinder its accessibility and hydrolysis by the agonist-regulated PLC enzymes. Alternatively, PH domains may interfere with the membrane localization of the PLC enzymes that are activated by an agonist. Although all of the known PLC isoenzymes contain a PH domain, its affinity and specificity toward inositides has not been analyzed in every case (10). To test the possibility that PH domains interfere with agonist-induced Ins(1,4,5)P3 formation from [3H]inositol-labeled PtdIns(4,5)P2, we overexpressed various PH domain–GFP constructs in COS-7 cells together with the AT1a Ang II receptor and measured Ang II–stimulated [3H]inositol phosphate formation in cells prelabeled by myo-[3H]inositol. Fig. 7A shows that PLCδ PH domain greatly inhibited Ang II–stimulated formation of [3H]inositol phosphates and that other PH domains with low affinity for PtdIns(4,5)P2, such as that of the Bruton’s tyrosine kinase or the Akt protein kinase, as well as that of dynamin (13, 18, 31), showed no similar inhibitory effect. Fluorescent constructs containing these PH domains did not show the same membrane localization as those with the PLCδ PH domain (not shown). Also, mutations within the PH domain of PLCδ that prevented its interaction with PtdIns(4,5)P2, and hence its membrane localization, failed to inhibit Ang II–induced inositol phosphate production (Fig. 7B). The corollary of this finding is that fluorescent PH domain constructs with high enough affinity to “label” PtdIns(4,5)P2 pools are most likely to also interfere with the agonist-sensitive phosphoinositide pools, since their binding to PtdIns(4,5)P2 impedes their access to the relevant PLC enzymes.

**Discussion**

The present experiments demonstrate that PH domains which bind PtdIns(4,5)P2 with high affinity and specificity, such as that of PLCδ1, can be used to visualize certain PtdIns(4,5)P2 pools in single living cells. Decreasing the affin-
ity of the PH domain by mutation of any one of three basic residues known to form contacts with the inositol phosphate headgroup (12, 35) was sufficient to eliminate membrane localization of the construct. Also, PH domains with lower affinity for PtdIns(4,5)P₂ such as that of the GTP-binding protein, dynamin, or with different specificity such as those of the Bruton’s tyrosine kinase and the protein kinase, Akt, did not show the clear plasma membrane localization that was observed with the PLC₃PH domain. The high concentration of the fluorescent probe at the plasma membrane was consistent with previously established views that PtdIns(4,5)P₂ is most abundant in the plasma membrane (26). The lack of localization to intracellular membranes, including the nuclear membrane where a separate inositide system has been described (7), suggests that in quiescent cells these membranes either contain only small amounts of this lipid, or that such pools are not accessible to the expressed PH domain. Since the interaction of the PH domain is formed with the inositol phosphate headgroup of PtdIns(4,5)P₂, hydrolysis of the lipid by PLC is reflected in the release of the fluorescent probe from the plasma membranes to the cytosol. Such change in localization was dramatically demonstrated when endogenous PLC was activated by Ca²⁺ influx via Ca²⁺ ionophores. After Ca²⁺ chelation, when PLC activity subsided and PtdIns(4,5)P₂ resynthesis began to take place, the appearance of bright spots at perinuclear structures preceded the localization of the fluorescence to the plasma membrane, indicating that some PtdIns(4,5)P₂ synthesis also occurs in intracellular membranes.

This novel methodology, which allowed analysis of the regulation of cellular PtdIns(4,5)P₂ pools from the viewpoint of PH domains showed that stimulation of a G protein–coupled Ca²⁺-mobilizing receptor, the AT₃ Ang II receptor, was also able to activate the hydrolysis of PH domain–tagged PtdIns(4,5)P₂. In a recent report, Stauffer et al. (see reference 32) reported that stimulation of PAF receptors (another G protein–coupled receptor) in a basophilic leukaemia cell line also caused a transient translocation of PLC₃PH domain using a similar methodology. Interestingly, activation of PLCγ by PDGF in NIH-3T3 cells (in this study), or by FCeRI stimulation in RBL cells (33) did not cause any visible change in the distribution of a PH domain–GFP construct (the PH domain of pleckstrin was used in reference 33). Whether this reflects the inability of PLCγ to hydrolyze PtdIns(4,5)P₂ when it is covered by a PH domain remains to be determined.

Overexpression of the PLC₃PH domain was also found to inhibit agonist-induced formation of myo-[³H]inositol–labeled inositol phosphates in transfected COS-7 cells, and this effect was closely correlated with the ability of the construct to localize to the membrane. Similar finding was presented in a recent report, in which pleckstrin, via its PH domain, was shown to inhibit [³H]inositol phosphate formation in transfected COS-1 or HEK 293 cells regardless of the type of PLC that was activated (1). These results suggest that expressed PH domains also can interfere with the agonist-regulated PLC activation mechanism, either by masking the PtdIns(4,5)P₂ headgroup from the enzyme’s catalytic site or by competing with the PH domains.
of the PLC enzymes to inhibit their localization to the membranes. These results also emphasize that the PLC activity detected in cells is greatly influenced by membrane components that interact with phosphoinositides, and that the use of cell-free systems and artificial substrates cannot reveal this aspect of PLC regulation.

An important finding of the present study was the inability of WT (at μM concentrations that inhibit type III PtdIns 4 kinases) to prevent the resynthesis of the PtdIns(4,5)P2 pools available for binding of the PLCδ PH domain in both ionomycin- and agonist-stimulated cells. The same treatment completely prevented the synthesis of myo-[3H]inositol-labeled PtdIns(4,5)P2 in Ang II-stimulated adrenal glomerulosa cells (27), and also blocks the resynthesis of [3H]PtdIns(4,5)P2 after chelation of Ca2+ in ionomycin-treated NIH-3T3 cells (Varnai, P., and T. Balla, unpublished observations). This finding raised the possibility that the agonist-sensitive myo-[3H]inositol-labeled PtdIns(4,5)P2 pools are not completely identical to those that are imaged by the fluorescent PH domain of PLCδ, although both respond to agonist stimulation. Heterogeneity of phosphoinositide pools have been described earlier in a few reports. These studies indicated that some of the metabolically labeled PtdIns and PtdIns(4,5)P2 pools are not sensitive to agonist stimulation (22) and conversely, some PtdIns(4,5)P2 that is not labeled metabolically is still subject to PLC-mediated hydrolysis (20). The existence of a metabolically hyperactive hormone-sensitive PtdIns(4,5)P2 pool, however, could not be substantiated (25). Our results indicate that, unlike the [3H]inositol-labeled PtdIns(4,5)P2 pools, the PtdIns(4,5)P2 pool(s) that bind PH domains are not synthesized by WT-sensitive type III PtdIns 4 kinases. The highly abundant, tightly membrane-bound type II PtdIns 4 kinase (5), which is not sensitive to

Figure 6. Availability of newly synthesized PtdIns(4,5)P2 to repeated stimulation of phospholipase C in BAG cells. BAG cells were transfected with plasmids encoding the PLCδ PH domain–GFP fusion protein. (A) Cells were stimulated with Ang II (10−7 M) for 5 min before addition of 10 μM ionomycin, and then 5 mM EGTA. In some experiments, losartan (10−7 M, an AT1 receptor antagonist) was added for 10 min to terminate Ang II action before the administration of ionomycin (not shown). (B) Cells were stimulated with 10 μM ionomycin and, immediately after the redistribution of fluorescence, 5 mM EGTA was added to remove Ca2+. After 10 min, cells were washed twice with incubation medium containing 5% BSA to reduce ionomycin, and then incubated in normal medium (containing 1.2 mM Ca2+) for an additional 10 min before stimulation with Ang II (10−7 M). Bar, 10 μm.
even high concentrations of WT (9) is a good candidate for synthesizing the inositides that bind PH domains. Clearly, more experiments will be required to find the explanation for the apparent discrepancy between the regulation of myo-[3H]inositol–labeled and PH domain–imaged PtdIns(4,5)P$_2$.

An additional possibility to be considered in understanding the current results is that the Ins(1,4,5)P$_3$ that is formed after agonist stimulation competes for the PH domain of PLC$_D$ (and of other proteins), and contributes to the release of these proteins from the plasma membrane, especially when PtdIns(4,5)P$_2$ levels are decreasing. Such competition by Ins(1,4,5)P$_3$ with PtdIns(4,5)P$_2$ for the PH domain of PLC$_D$ has been demonstrated and also found to inhibit the catalytic efficiency of the enzyme (2, 19). Since the only known major source of Ins(1,4,5)P$_3$ is PtdIns(4,5)P$_2$, this competing effect of Ins(1,4,5)P$_3$ would distort the imaging results if there were substantial amounts of Ins(1,4,5)P$_3$ formed from PtdIns(4,5)P$_2$ pools that did not bind PH domains in their initial steady-state before stimulation. Nevertheless, this is an open possibility and a better assessment of such a displacing effect of Ins(1,4,5)P$_3$ requires additional studies with GFP-fused PH domains.

In summary, the present studies demonstrate that iso-

lated PH domains fused to the GFP are capable of recognizing phosphoinositides in living cells with remarkable specificity. These interactions allow visualization of the spatiotemporal changes in phosphoinositides at the single cell level. Analysis of cellular PtdIns(4,5)P$_2$ with this method reveals that the pools that bind PH domains may not be identical to those that can be metabolically labeled with myo-[3H]inositol. Studies on the receptor-mediated control of the PtdIns(4,5)P$_2$ pools that bind PH domains will help to understand the manner in which phosphoinositides regulate various signaling processes via protein PH domains.

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