Detection of Heteromers Formed by Cannabinoid CB\textsubscript{1}, Dopamine D\textsubscript{2}, and Adenosine A\textsubscript{2A} G-Protein-Coupled Receptors by Combining Bimolecular Fluorescence Complementation and Bioluminescence Energy Transfer

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Received July 23, 2008; Revised October 1, 2008; Accepted October 1, 2008; Published October 11, 2008

Functional interactions in signaling occur between dopamine D\textsubscript{2} (D\textsubscript{2}R) and cannabinoid CB\textsubscript{1} (CB\textsubscript{1}R) receptors, between CB\textsubscript{1}R and adenosine A\textsubscript{2A} (A\textsubscript{2A}R) receptors, and between D\textsubscript{2}R and A\textsubscript{2A}R. Furthermore, direct molecular interactions have been reported for the pairs CB\textsubscript{1}R-D\textsubscript{2}R, A\textsubscript{2A}R-D\textsubscript{2}R, and CB\textsubscript{1}R-A\textsubscript{2A}R. Here a combination of bimolecular fluorescence complementation and bioluminescence energy transfer techniques was used to identify the occurrence of D\textsubscript{2}R-CB\textsubscript{1}R-A\textsubscript{2A}R hetero-oligomers in living cells.

KEYWORDS: G-protein-coupled receptors, adenosine A\textsubscript{2A} receptor, cannabinoid CB\textsubscript{1} receptor, dopamine D\textsubscript{2} receptor, heterotrimers, oligomers of three different protomers, bimolecular fluorescence complementation, bioluminescence energy transfer, BRET

INTRODUCTION

The mechanism of action responsible for the motor-depressant effects of cannabinoids, which operate through centrally expressed cannabinoid CB\textsubscript{1} receptors (CB\textsubscript{1}R), is still a matter of debate. The cellular and subcellular localization of CB\textsubscript{1}R and D\textsubscript{2} receptors (D\textsubscript{2}R) in the basal ganglia have been described in
The evidence suggests a colocalization of striatal CB₁R and D₂R predominantly in the soma and dendrites of the GABA enkephalinergic neurons and also in corticostriatal glutamate terminals where A₂A receptors (A₂A R) are also present.[5,6,7,8,9,10]. Functional interactions between CB₁R and A₂A R or D₂R receptors that are relevant for striatal function have been reported. Thus, CB₁R and A₂A R form heteromers in cotransfected HEK-293T cells and in rat striatum[11]. In a human neuroblastoma cell line, CB₁R signaling was found to be completely dependent on A₂A R activation. Accordingly, blockade of A₂A R counteracted the motor-depressant effects produced by the intrastratral administration of a CB₁R agonist[11]. The existence of CB₁R-D₂R heteromers has been demonstrated in transfected cell lines by coimmunoprecipitation[12] and by fluorescence resonance energy transfer (FRET) experiments[13]. Antagonistic CB₁R-D₂R interactions have been discovered at the behavioral level[13,14,15,16]. In rats, the CB₁R receptor agonist CP 55,940 at a dose that did not change basal locomotion was able to block quinpirole-induced increases in locomotor activity. In addition, not only the CB₁R antagonist rimonabant, but also the specific A₂A R antagonist MSX-3, blocked the inhibitory effect of CB₁R agonist on D₂-like receptor agonist-induced hyperlocomotion[13]. Taken together, these results give evidence for the existence of antagonistic CB₁-D₂ receptor-receptor interactions within CB₁R-D₂R heteromers in which A₂A R may also participate.

A₂A R-D₂R was one of the first reported heteromers[17]. A close physical interaction between both receptors has been shown using coimmunoprecipitation and colocalization assays[17], and FRET and bioluminescence resonance energy transfer (BRET) techniques[18,19,20]. At the biochemical level, two kinds of antagonistic A₂A R-D₂R interactions have been discovered that can explain the A₂A R-D₂R interaction observed at both the functional and behavioral levels[9,21,22,23]. First, by means of an intramembrane interaction, i.e., by intramolecular cross-talk within the heterodimer, stimulation of A₂A R decreases the affinity of D₂R for their agonists[24]. Second, the stimulation of D₂R, a Gi/o protein–coupled receptor, inhibits the cAMP accumulation induced by the stimulation of the Gs/olf protein–coupled A₂A R[17]. Therefore, it has been suggested that the A₂A R-D₂R interaction cross-talk in the central nervous system may provide new therapeutic approaches for Parkinson’s disease, schizophrenia, and drug addiction[23,25].

Since trimers formed by cannabinoid, adenosine, and dopamine receptors were suspected, strategies to detect them were developed in our laboratory. In one of them, trimers were detected by sequential application of BRET and FRET[26]. In this paper, the formation of hetero-oligomer complexes formed by cannabinoid CB₁, dopamine D₂, and adenosine A₂A G-protein-coupled receptors (GPCRs) is confirmed by another technique[27], which consists of combining bimolecular fluorescence complementation and bioluminescence energy transfer.

**MATERIALS AND METHODS**

**Cell Culture**

HEK-293T cells were grown in Dulbecco’s modified Eagle’s medium (DMEM) (Gibco) supplemented with 2 mM L-glutamine, 100 U/ml penicillin/streptomycin, and 5% (v/v) heat inactivated fetal bovine serum (FBS) (all supplements were from Invitrogen, Paisley, Scotland, U.K.). Cells were maintained at 37°C in an atmosphere of 5% CO₂, and were passaged when they were 80–90% confluent, i.e., approximately twice a week.

**Fusion Proteins and Expression Vectors**

Full-length YFP was subcloned in the XhoI site of pcDNA3.1 vector (Invitrogen). The N-terminal truncated version of YFP, named nYFP (amino acids 1 to 155), was made by PCR amplification and cloning into the XhoI site of pcDNA3.1 using the following primers: FnYFP (5’-
CCGCTCGAGACCATGGTAGCAAGGCGAGGAGC-3') and RnYFP (5'-CCGTCTAGATCAGGCCATGATATAGACGTTG-3'). Also, a C-terminal truncated version of YFP, named cYFP (amino acids 155 to 231), was made using the same strategy and the following primers: FcYFP (5'-CCGCTCGAGACCATGGACAGAAGAACGGC-3') and RcYFP (5'-CCGTCTAGATTACTTGTACAGCTCGTCCAT-3'). G\(\alpha_s\) cloned in SFV1 vector (generously given by H. Vogel, Ecole Politecnique Federal de Laussane, Switzerland) or G\(\gamma\) and G\(\beta\) cloned in pEYFP-C1 vector (generously provided by S. Cotecchia, Department of Pharmacology and Toxicology, University of Lausanne, Switzerland) were amplified to miss their stop codons using sense and antisense primers harboring unique NheI and BamHI sites to clone G\(\alpha_s\) and G\(\beta\) in pcDNA3.1-nYFP and pcDNA3.1-cYFP, respectively, and HindIII and BamHI sites to clone G\(\gamma\) in Rluc vector (pRluc-N1 PerkinElmer, Wellesley, MA). The amplified fragments were subcloned to be in-frame with the multiple cloning site of the vectors to give the plasmids G\(\alpha_s\)-nYFP, G\(\beta\)-cYFP, and G\(\gamma\)-Rluc, respectively. All plasmids express luminescent or part of fluorescent proteins on the C-terminal ends of proteins. The human cDNAs for A\(2A\)R, CB\(1\)R, D\(2\)R, or D\(4\)R cloned in pcDNA3.1 were amplified without their stop codons using sense and antisense primers harboring unique NheI and BamHI sites to clone A\(2A\)R, CB\(1\)R, and D\(2\)R in pcDNA3.1-cYFP, pcDNA3.1-nYFP, and pRluc-N1, respectively, and XhoI and BamHI sites to clone D\(4\)R in pRluc-N1 vector. The amplified fragments were subcloned to be in-frame with the multiple cloning site of the vectors to give the plasmids A\(2A\)-cYFP, CB\(1\)-nYFP, D\(2\)-Rluc, or D\(4\)-Rluc.

**Transient Transfection and Protein Determination**

HEK-293T cells growing in six-well dishes were transiently transfected with the corresponding fusion protein cDNAs by PEI (PolyEthylenImine, Sigma, Steinheim, Germany) method. Cells were incubated with the corresponding cDNA, 5.47 mM (in nitrogen residues) PEI, and 150 mM NaCl in a serum-free medium. After 4 h, cells were placed in a fresh complete culture medium. Forty-eight hours after transfection, cells were rapidly washed twice in HBSS containing 10 mM glucose, detached, and resuspended in the same buffer. To control for cell number, sample protein concentration was determined using a Bradford assay kit (Bio-Rad, Munich, Germany) using bovine serum albumin as reference. Cell suspension (20 µg of protein) was distributed into 96-well black plates with a transparent bottom for fluorescence determinations or white plates with white bottom for BRET experiments.

**Fluorescence Measurements**

To quantify fluorescence, cells (20 µg protein) were distributed in 96-well microplates (black plates with a transparent bottom) and fluorescence was read in a Mithras LB 940 (Berthold Technologies, DLReady, Germany) using a 10-nm bandwidth excitation and emission filters at 485 and 530 nm, respectively. Protein fluorescence expression was determined as fluorescence of the sample minus the fluorescence of cells that were not transfected.

**BRET Assays with Bimolecular Fluorescence Complemented Proteins**

HEK-293T cells were transiently cotransfected with a constant amount of cDNA encoding for the protein fused to Rluc and with increasingly equal amounts of cDNA corresponding to proteins fused to one of the two complementary parts of the YFP protein (nYFP and cYFP). Fluorescence was measured as indicated above. The equivalent to 20 µg of cell suspension was distributed in 96-well microplates (Corning 3600, white plates with white bottom) and 5 µM coelenterazine H (Molecular Probes, Eugene, OR) was added. After 1-min delay, collection of readings started using a Mithras LB 940, which allows the integration of the signals detected in the short-wavelength filter at 485 nm (440–500 nm) and the long-wavelength filter.
at 530 nm (510–590 nm). To quantify for protein-Rluc expression, luminescence readings were performed after 10 min of adding 5 μM coelenterazine H. The net BRET is defined as [(long-wavelength emission)/(short-wavelength emission)]-Cf, where Cf corresponds to [(long-wavelength emission)/(short-wavelength emission)] for the Rluc construct expressed alone in the same experiment.

Immunostaining

For immunocytochemistry, transiently transfected HEK-293T cells were fixed in 4% paraformaldehyde for 15 min and washed with PBS containing 20 mM glycine (buffer A) to quench the aldehyde groups. Then, after permeabilization with buffer A containing 0.2% Triton X-100 for 5 min, cells were treated with PBS containing 1% bovine serum albumin. After 1 h at room temperature, cells expressing D2R-Rluc were labeled with the primary mouse monoclonal anti-Rluc antibody (1/100, Chemicon) for 1 h, washed, and stained with the secondary antibody Cy3 Donkey antimouse (1/100, Jackson Immunoresearch Laboratories, Baltimore, PA). Heterodimers of receptors fused to complementary fragments of YFP were detected by their fluorescence properties. Samples were rinsed and observed in a Leica SP2 confocal microscope (Leica Microsystems, Mannheim, Germany).

RESULTS AND DISCUSSION

In the early 1980s, and based on indirect functional evidence, it was proposed that GPCRs could interact at the level of the neuronal plasma membrane. In the early 1990s, electrophoretic mobility and coimmunoprecipitation assays gave the first indication of GPCR homomerization. More recently, the development of the biophysical techniques BRET and FRET allowed the demonstration of GPCR homodimerization and heteromerization of two GPCRs in living cells[23,28,29,30,31,32,33,34,35,36]. Nevertheless, the lack of assays monitoring interactions between more than two proteins simultaneously makes it very difficult to draw a map of molecular networks involving protein-protein interactions. By the approach previously reported by Héroux et al.[27], we here show that the combination of bimolecular fluorescence complementation and bioluminescence energy transfer is useful to detect heteromerization of three different GPCRs.

For bimolecular fluorescence complementation assay, the reconstitution of a reporter fluorescence protein (YFP) from its two fragments attached to the potential interacting protein partners under study[37,38] is taken as evidence for the molecular interaction between the partners[39]. The usefulness of the bimolecular fluorescence complementation technique to detect protein heterodimers was proved using cells transfected with the following subunits of heterotrimeric G proteins: GαnYFP and GβcYFP. Positive complementation was detected by the increase of fluorescence at 530 nm upon increasing the amount of transfected proteins (Fig. 1). As negative control, no signal was detected when either GαnYFP and cYFP, or GβcYFP and nYFP, were cotransfected (Fig. 1). These results prove the ability of the bimolecular complementation technique to detect heterodimers. The bimolecular fluorescence complementation technique was used to detect CB1R-A2AR heterodimers in HEK-293 cells. Fusion of nYFP and cYFP fragments to CB1R or to A2AR did not prevent the receptor functionality determined as ERK1/2 phosphorylation (results not shown). Cells transfected with CB1RnYFP and A2ArYFP (see Methods) showed fluorescence at the membrane level (Fig. 2A). No fluorescence was detected when cells were cotransfected with A2ArYFP and nYFP (Fig. 2A) or with CB1RnYFP and cYFP (results not shown), showing the specificity of the signal. In agreement, positive complementation was detected by the increase of fluorescence at 530 nm upon increasing the amount of transfected receptors (Fig. 2B). As a negative control, no signal was detected when either A2ArYFP and nYFP, or CB1RnYFP and cYFP, were cotransfected (Fig. 2B). Taken together, these results validate the usefulness of the bimolecular fluorescence complementation technique to monitor the formation of CB1R-A2AR heteromers.
FIGURE 1. Molecular interaction between Gα and Gβ subunits of heterotrimeric G proteins detected by bimolecular fluorescence complementation. HEK-293 cells were cotransfected with equal amounts of cDNAs corresponding to the fusion proteins GαnYFP and GβcYFP (black), GαnYFP and cYFP (white), or GβcYFP and nYFP (grey). Forty-eight hours post-transfection, fluorescence was determined at 530 nm. Values are mean ± SEM of four independent experiments. One-way ANOVA followed by Newman-Keuls test showed significant differences respective to both negative controls. ***p < 0.001.

FIGURE 2. CB₁R-A₂AR heterodimers in HEK-293 cells. HEK-293 cells were cotransfected with (A) 2 µg of cDNA corresponding to the fusion proteins CB₁RnYFP and A₂ARcYFP (left panel), or A₂ARcYFP and nYFP (right panel) or (B) equal amounts of cDNAs corresponding to CB₁RnYFP and A₂ARcYFP (black), A₂ARcYFP and nYFP (grey), or CB₁RnYFP and cYFP (white). In (A), confocal microscopy images obtained 48 h post-transfection are shown. In (B), fluorescence at 530 nm was determined 48 h post-transfection. Values are mean ± SEM of four independent experiments. One-way ANOVA followed by Newman-Keuls test showed significant differences respective to both negative controls. ***p < 0.001.
The positive results on the formation of CB₁R-A₂Aᵣ heteromers, obtained by the bimolecular fluorescence complementation technique, opened the possibility of combining this technology with BRET to investigate the existence of D₂R-CB₁R-A₂Aᵣ hetero-oligomers. The usefulness of the combination of the bimolecular fluorescence complementation technique and BRET to detect oligomers formed by three different proteins was first tested by transfecting the following fusion proteins of the three subunits of heterotrimeric G proteins: Gγ-Rluc, GαnYFP, and GβcYFP. Positive BRET was detected between Gγ-Rluc and complemented GαnYFP-GβcYFP. A hyperbolic BRET saturation curve was obtained upon increasing the GαnYFP-GβcYFP expression (Fig. 3A). This result proves the ability of the combination of the two techniques to detect trimolecular protein complexes as those detected for functional calcitonin gene-related peptide receptors, which are formed by the asymmetric assembly of a calcitonin receptor-like receptor homo-oligomer and a monomer of receptor activity-modifying protein-1[27].

In order to detect possible formation of hetero-oligomers composed of CB₁, D₂, and A₂Aᵣ receptors, HEK-293 cells were transfected with D₂R-Rluc, A₂AᵣcYFP, and CB₁RnYFP. Fusion of Rluc to D₂R did not prevent the receptor functionality determined as ERK1/2 phosphorylation (results not shown). Fusion proteins did not affect the normal subcellular distribution of receptors (Fig. 3B). In fact, these receptors are predominantly colocalized in the plasma membrane of cotransfected cells. In conditions to give a BRET₅₀ fusion protein expression levels, measured as described previously[26] by radioligand binding in different experimental sessions, were between 0.5 and 0.7 pmols/mg protein for A₂AᵣcYFP, between 0.9 and 1.1 pmols/mg protein for D₂R-Rluc, and between 0.6 and 0.8 pmols/mg protein for CB₁RnYFP. Triggering with coelenterazin H, these transfected cells gave a significant BRET signal. The BRET signal was specific as assessed by the saturation hyperbola obtained upon increasing the complemented A₂AᵣcYFP-CB₁RnYFP expression and by the lack of the signal using D₂RRluc instead of D₂R-Rluc as a negative control (Fig. 3C). These data indicate that D₂R, CB₁R, and A₂Aᵣ form, at least, trimolecular oligomers in cotransfected living cells. This technique is validated by the identification of D₂-CB₁- and A₂Aᵣ receptor heteromers by sequential resonance energy transfer (SRET)[26]. Apart from transmembrane regions, basic and acidic residues are involved in the epitope-epitope electrostatic interactions existing in D₂-A₂Aᵣ receptor heteromers. In fact, mass spectrometry and pull-down assays have been instrumental to show that the Arg-rich D₂R epitope may bind to two different epitopes in the C-terminal part of the A₂Aᵣ, one containing two adjacent Asp residues and another containing a phosphorylated Ser residue[18,20]. Then, it might be possible that one of the epitopes is involved in the interaction with D₂ and another in the interaction with CB₁. Further experimental work is necessary, however, to elucidate the amino acids constituting the interfaces in the D₂-CB₁-A₂Aᵣ hetero-oligomer.
FIGURE 3. D₂R-CB₁R-A₂A R heteromers detected by a combination of bimolecular fluorescence complementation and BRET. In (A), as a positive control, BRET saturation curve was performed using HEK-293 cells cotransfected with 0.75 µg of cDNA corresponding to the fusion protein G₃-RLuc (100,000 bioluminescence units) and increasing equal amounts of cDNAs corresponding to Gα₌YFP and GβcYFP (1000–6000 fluorescence units). In (B), confocal microscopy image of a cell after 48 h of transfection with 1 µg of cDNA corresponding to D₂RRLuc, 2 µg of cDNA corresponding to CB₁RnYFP, and 2 µg of cDNA corresponding to A₂ARcYFP. Proteins were identified by fluorescence (green image) or by immunocytochemistry (red image) using a monoclonal anti-RLuc primary antibody and a cyanine-3-conjugated secondary antibody. Colocalization is shown in yellow in the right image. In (C), BRET saturation curve (red) was obtained using HEK-293 cells cotransfected with 1.5 µg of cDNA corresponding to D₂RRLuc (100,000 bioluminescence units) and increasing equal amounts of cDNAs corresponding to CB₁RnYFP and A₂ArcYFP (1000–10,000 fluorescence units). As negative controls, cells were transfected with 1.5 µg of either the cDNA for D₄RLuc (black line) or for GABAB₁RLuc (green line) (100,000 bioluminescence units in each case).
There has been reported a coexpression of D₂R and A₂A R in GABAergic striatal neurons (see [9]) and of CB₁R and A₂A R in rat striatal fibrillar structures[11,13]. The demonstration of D₂R-CB₁R-A₂A R heteromers in transfected cells, together with such striatal codistribution of the three receptors in the plasma membrane of striatal neurons, strongly suggests that these three receptors are forming part of a molecular network. The function of these neurons is particularly compromised in Parkinson’s disease and in the early stages of Huntington’s disease[21]. Furthermore, neuroadaptations of glutamatergic synapses of GABAergic enkephalinergic neurons localized in the nucleus accumbens (the ventral part of the striatum) seem to be involved in compulsive drug seeking and relapse[40]. Based on the existence of antagonistic interactions between A₂A R and D₂R in the A₂A R-D₂R heteromer[21,25], A₂A R antagonists are giving successful results in clinical trials in patients with Parkinson’s disease[41]. Furthermore, A₂A R antagonists are being considered as possible therapeutic agents in end-stage drug addiction[42]. Their clinical efficacy might be related to the recently demonstrated dependence of A₂A R activation for CB₁R receptor signaling within the striatal A₂A R-CB₁R heteromers[11]. Thus, A₂A R antagonists behave as CB₁R antagonists, known to counteract cue-induced reinstatement of different addictive drugs in the experimental animal, a model for human relapse[43]. Although the intramembrane and intracellular cross-talk established by complexes formed by receptor heterodimers is already important to understand better the function of striatal enkephalinergic neurons, the occurrence of oligomers formed by three different receptors indicate a more diverse interplay between receptors for neurotransmitters. Taken together, the results already reported in the literature suggest that the A₂A R-CB₁R-D₂R receptor heteromer may act as a processor mediating the neuronal computation needed to modulate striatal dopamine neurotransmission. The demonstration of D₂R-CB₁R-A₂A R heteromers in transfected cells, together with their striatal codistribution, opens new perspectives to understand the interplay between different neurotransmitter-neuromodulator systems. Pharmacological and functional diversification expand in a macromolecular complex containing three receptors by the same simple events as described for dimers, i.e., by (1) a change in the pharmacological profile of a receptor when another receptor in the complex is activated and (2) a change in the associated signaling response-pathways depending on the receptors present in the complex, their degree of activation, and the nature of the G proteins expressed in the horizontal molecular network involved[44].

The combination of bimolecular fluorescence complementation and bioluminescence energy transfer techniques constitutes a powerful approach to detect the protein-protein interactions localized in the plane of the membrane, and thus allows identification of the horizontal molecular networks like the receptor networks in local circuits. This new knowledge will hopefully provide novel therapeutic approaches for neurodegenerative diseases, mental disorders, and drug addiction.

ACKNOWLEDGMENTS

This research was supported by grants SAF2006–00170 and SAF2005-00170 from the Spanish Ministerio de Ciencia y Tecnologia and grant 060110 from Fundació La Marató de TV3.

REFERENCES

1. Egertova, M. and Elphick, M.R. (2000) Localisation of cannabinoid receptors in the rat brain using antibodies to the intracellular C-terminal tail of CB. J. Comp. Neurol. 422, 159–171.
2. Khan, Z.U., Gutierrez, A., Martin, R., Penafiel, A., Rivera, A., and De La Calle, A. (1998) Differential regional and cellular distribution of dopamine D2-like receptors: an immunocytochemical study of subtype-specific antibodies in rat and human brain. J. Comp. Neurol. 402, 353–371.
3. Matyas, F., Yanovsky, Y., Mackie, K., Kelsch, W., Misgeld, U.A., and Freund, T.F. (2006) Subcellular localization of type 1 cannabinoid receptors in the rat basal ganglia. Neuroscience 137, 337–361.
4. Missale, C., Nash, S.R., Robinson, S.W., Jaber, M., and Caron, M.G. (1998) Dopamine receptors: from structure to function. Physiol. Rev. 78, 189–225.
5. Pickel, V.M., Chan, J., Kearns, C.S., and Mackie, K. (2006) Targeting dopamine D2 and cannabinoid-1 (CB1)
6. Köfalvi, A., Rodrigues, R.J., Ledent, C., Mackie, K., Vizi, E.S., Cunha. R.A., and Sperlágh, B. (2005) Involvement of cannabinoid receptors in the regulation of neurotransmitter release in the rodent striatum: a combined immunochemical and pharmacological analysis. J. Neurosci. 25, 2874–2884.

7. Katona, I., Urban, G.M., Wallace, M., Ledent, C., Jung, K.M., Piomelli, D., Mackie, K., and Freund, T.F. (2006) Molecular composition of the endocannabinoid system at glutamatergic synapses. J. Neurosci. 26, 5628–5637.

8. Yin, H.H. and Lovinger, D.M. (2006) Frequency-specific and D2 receptor-mediated inhibition of glutamate release by striatal endocannabinoid signaling. Proc. Natl. Acad. Sci. U. S. A. 103, 8251–8256.

9. Ferré, S., Agnati, L.F., Ciruela, F., Lluis, C., Woods, A.S., Fuxe, K., and Franco, R. (2007) Neurotransmitter receptor heteromers and their integrative role in 'local modules': the striatal spine module. Brain Res. Rev. 55, 55–67.

10. Uchigashima, M., Narushima, M., Fukaya, M., Katona, I., Kano, M., and Watanabe, M. (2007) Subcellular arrangement of molecules for 2-arachidonoylethanolamine-mediated retrograde signaling and its physiological contribution to synaptic modulation in the striatum. J. Neurosci. 27, 3663–3676.

11. Carriba, P., Ortiz, O., Patkar, K., Justinova, Z., Stroik, J., Themann, A., Muller, C., Woods, A.S., Hope, B.T., Ciruela, F., Casado, V., Canela, E.I., Lluis, C., Goldberg, S.R., Moratalla, R., Franco, R., and Ferré, S. (2007) Striatal adenosine A1A and cannabinoid CB1 receptors form functional heteromeric complexes that mediate the motor effects of cannabinoids. Neuropsychopharmacology 32, 2249–2259.

12. Kearn, C.S., Blake-Palmer, K., Daniel, E., Mackie, K., and Glass, M. (2005) Concurrent stimulation of cannabinoid CB1 and dopamine D2 receptors enhances heterodimer formation: a mechanism for receptor cross-talk? Mol. Pharmacol. 67, 1697–1704.

13. Marcellino, D., Carriba, P., Filip, M., Borgkvist, A., Frankowska, M., Bellido, I., Tanganelli, S., Muller, C., Fisone, G., Lluis, C., Agnati, L.F., Franco, R., and Fuxe, K. (2007) Antagonistic cannabinoid CB1/dopamine D2 receptor interactions in striatal CB1/D2 heteromers. A combined neurochemical and behavioral analysis. Neuropsychopharmacology 54, 815–823.

14. Giuffrida, A., Parsons, L.H., Kerr, T.M., Rodríguez de Fonseca, F., Navarro, M., and Piomelli, D. (1999) Dopamine activation of endogenous cannabinoid signaling in dorsal striatum. Nat. Neurosci. 2, 358–363.

15. Thiemann, G., Di Marzo, V., Molleman, A., and Hasenöhrl, R.U. (2008) The CB1 cannabinoid receptor antagonist AM251 attenuates amphetamine-induced behavioural sensitization while causing monoamine changes in nucleus accumbens and hippocampus. Pharmacol. Biochem. Behav. 89, 384–391.

16. Hillion, J., Canals, M., Torvinen, M., Casado, V., Scott, R., Terasmaa, A., Hansson, A., Watson, S., Olah, M.E., Mallol, J., Canela, E.I., Zoli, M., Agnati L.F., Ibanez, C.F., Lluis, C., Franco, R., Fuxe, S., and Fuxe, K. (2002) Aggregation, cointernalization, and codesensitization of adenosine A2A receptors and dopamine D2 receptors. J. Biol. Chem. 277, 18091–18097.

17. Canals, M., Marcellino, D., Fanelli, F., Ciruela, F., de Benedetti, P., Goldberg, S.R., Neve, K., Fuxe, K., Agnati, L.F., Woods, A.S., Ferre, S., Lluis, C., Bouvier, M., and Franco, R. (2003) Adenosine A2A-dopamine D2 receptor-receptor heteromerization. Qualitative and quantitative assessment by fluorescence and bioluminescence resonance energy transfer. J. Biol. Chem. 278, 46741–46749.

18. Kamiya, T., Suitoh, O., Yoshioka, K., and Nakata, H. (2003) Oligomerization of adenosine A2A and dopamine D2 receptors in living cells. Biochem. Biophys. Res. Commun. 306, 544–549.

19. Ferré, S., Burgueño, J., Casado, V., Canals, M., Marcellino, D., Goldberg, S.R., Bader, M., Fuxe, K., Agnati, L.F., Lluis, C., Franco, R., Ferré, S., and Woods, A.S. (2004) Combining mass spectrometry and pull-down techniques for the study of receptor heteromerization. Direct epitope-epitope electrostatic interactions between adenosine A2A and dopamine D2 receptors. Anal. Chem. 76, 5354–5363.

20. Ferré, S., Fredholm, B.B., Morelli, M., Popoli, P., and Fuxe, K. (1997) Adenosine-dopamine receptor-receptor interactions as an integrative mechanism in the basal ganglia. Trends Neurosci. 20, 482–487.

21. Ferre, S., Ciruela, F., Woods, A.S., Canals, M., Burgueno, J., Marcellino, D., Karcz-Kubička, M., Hope, B.T., Morales, M., Popoli, P., Goldberg, S.R., Fuxe, K., Lluis, C., Franco, R., and Agnati, L.F. (2003) Glutamate mGlur5-adenosine A2A-dopamine D2 receptor interactions in the striatum. Implications for drug therapy in neuro-psychiatric disorders and drug abuse. Curr. Med. Chem. 3, 1–26.

22. Agnati, L.F., Ferré, S., Lluis, C., Franco, R., and Fuxe, K. (2003) Molecular mechanisms and therapeutic implications of intramembrane receptor/receptor interactions among heptahelical receptors with examples from the striatopallidal GABA neurons. Pharmacol. Rev. 55, 509–550.

23. Ferré, S., von Euler, G., Johansson, B., Fredholm, B.B., and Fuxe, K. (1991) Stimulation of high-affinity adenosine A2 receptors decreases the affinity of dopamine D2 receptors in rat striatal membranes. Proc. Natl. Acad. Sci. U. S. A. 88, 7238–7241.

24. Ferré, S., Ciruela, F., Canals, M., Marcellino, D., Burgueno, J., Casado, V., Hillion, J., Torvinen, M., Fanelli, F., Benedetti, P., Goldberg, S.R., Bouvier, M., Fuxe, K., Agnati, L.F., Lluis, C., Franco, R., and Woods, A. (2004) Adenosine A2A-dopamine D2 receptor-receptor heteromers. Targets for neuro-psychiatric disorders. Parkinsonism Relat. Disord. 10, 265–271.
26. Carriba, P., Navarro, G., Ciruela, F., Ferré, S., Casadó, V., Agnati, L., Cortés, A., Mallol, J., Fuxé, K., Canela, E.I., Lluis, C., and Franco, R. (2008) Detection of heteromerization of more than two proteins by sequential BRET-FRET. Nat. Methods 5(8), 727–733.

27. Héroux, M., Hogue, M., Lemieux, S., and Bouvier, M. (2007) Functional calcitonin gene-related receptors are formed by the asymmetric assembly of a calcitonin receptor-like receptor homo-oligomer and a monomer of receptor activity-modifying protein-1. J. Biol. Chem. 282, 31610–31620.

28. Bouvier, M. (2001) Oligomerization of G-protein-coupled transmitter receptors. Nat. Rev. Neurosci. 2, 274–286.

29. Franco, R., Canals, M., Marcellino, D., Ferré, S., Agnati, L., Mallol, J., Casadó, V., Ciruela, F., Fuxé, K., Lluis, C., and Canela, E.I. (2003) Regulation of heptaspanning-membrane-receptor function by dimerization and clustering. Trends Biochem. Sci. 28, 238–243.

30. Milligan, G. (2004) G protein-coupled receptor dimerization: function and ligand pharmacology. Mol. Pharmacol. 66, 1–7.

31. Pfleger, K.D. and Eidne, K.A. (2006) Illuminating insights into protein-protein interactions using bioluminescence resonance energy transfer. Nat. Methods 3, 165–174.

32. Rashid, A.J., So, C.H., Kong, M.M., Furtak, T., El-Ghundi, M., Cheng, R., O'Dowd, B.F., and George, S.R. (2007) D1-D2 dopamine receptor heterooligomers with unique pharmacology are coupled to rapid activation of Gq/11 in the striatum. Proc. Natl. Acad. Sci. U. S. A. 104, 654–659.

33. Vilardaga, J.P., Nikolaev, V.O., Lorenz, K., Ferrandon, S., Zhuang, Z., and Lohse, M.J. (2008) Conformational cross-talk between alpha2A-adrenergic and mu-opioid receptors controls cell signaling. Nat. Chem. Biol. 4, 126–131.

34. Juhasz, J.R., Hasbi, A., Rashid, A.J., So, C.H., George, S.R., and O'Dowd, B.F. (2008) Mu-opioid receptor heterooligomer formation with the dopamine D(1) receptor as directly visualized in living cells. Eur. J. Pharmacol. 581, 235–243.

35. Ferré, S., Ciruela, F., Quiroz, C., Luján, R., Popoli, P., Cunha, R.A., Agnati, L.F., Fuxe, K., Woods, A.S., Lluis, C., and Franco, R. (2007) Adenosine receptor heteromers and their integrative role in striatal function. TheScientificWorldJOURNAL 7, 74–85.

36. Franco, R., Casadó, V., Cortés, A., Ferrada, C., Mallol, J., Woods, A., Lluis, C., Canela, E.I., and Ferré, S. (2007) Basic concepts in G-protein-coupled receptor homo- and heterodimerization. TheScientificWorldJOURNAL 7, 48–57.

37. Michnick, S.W. (2001) Exploring protein interactions by interaction-induced folding of proteins from complementary peptide fragments. Curr. Opin. Struct. Biol. 11, 472–477.

38. Hu, C.D., Chinenov, Y., and Kerppola, T.K. (2002) Visualization of interactions among bZIP and Rel family proteins in living cells using bimolecular fluorescence complementation. Mol. Cell 9, 789–798.

39. Kerppola, T.K. (2008) Bimolecular fluorescence complementation: visualization of molecular interactions in living cells. Methods Cell Biol. 85, 431–470.

40. Kalivas, P.W. and Volkow, N.D. (2005) The neural basis of addiction: a pathology of motivation and choice. Am. J. Psychiatry 162, 1403–1413.

41. Jenner, P. (2005) Istradefylline, a novel adenosine A2A receptor antagonist, for the treatment of Parkinson's disease. Expert Opin. Investig. Drugs 14, 729–738.

42. Ferré, S., Diamond, I., Goldberg, S.R., Yao, L., Hourani, S.M., Huang, Z.L., Urade, Y., and Kitchen, I. (2007b) Adenosine A2A receptors in ventral striatum, hypothalamus and nociceptive circuitry. Implications for drug addiction, sleep and pain. Prog. Neurobiol. 83, 263–276.

43. De Vries, T.J. and Schoffelmeer, A.N. (2005) Cannabinoid CB1 receptors control conditioned drug seeking. Trends Pharmacol. Sci. 26, 420–426.

44. Terrillon, S. and Bouvier, M. (2004) Roles of G-protein-coupled receptor dimerization. EMBO Rep. 5, 30–34.