The effects of synbiotics on indoxyl sulphate level, constipation, and quality of life associated with constipation in chronic haemodialysis patients: a randomized controlled trial

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Abstract

Background: Gut microbiota dysbiosis in patients with chronic kidney disease on haemodialysis (CKD-HD) creates an increase in proteolytic bacteria activity, leading to an increase in the production of uraemic toxins, such as indoxyl sulphate, worsening of constipation symptoms and reducing patients' quality of life. Improving gut microbiota dysbiosis is expected to improve this condition. This study aimed to evaluate the effect of synbiotics on indoxyl sulphate levels, constipation symptoms, and constipation-related quality of life in haemodialysis patients.

Methods: This was a double-blinded randomized controlled clinical trial with a parallel design involving haemodialysis patients. We included chronic haemodialysis patients with gastrointestinal complaints, difficulty defecating, faeces with hard consistency, or a bowel movement frequency of fewer than three times per week. Patients were randomly divided into two groups (synbiotics (\textit{Lactobacillus acidophilus} and \textit{Bifidobacterium longum} 5x10^9 CFU) and placebo) for 60 days of oral intervention. All participants, caregivers, and outcome assessors were blinded to group assignment. The primary outcome was a decrease in indoxyl sulphate toxin levels. Meanwhile, improvement in constipation symptoms (measured using the Patient Assessment of Constipation: Symptoms (PAC-SYM) questionnaire) and improvement in constipation-related quality of life (measured using the Patient Assessment of Constipation Quality of Life (PAC-QOL) questionnaire) were assessed as secondary outcomes.

Results: We included 60 patients (30 intervention; median age of 51.23 (13.57) years, 33.3% male; 30 control; median age of 52.33 (11.29) years, 36.7% male). There was no significant difference in terms of pre- and postintervention indoxyl sulphate toxin levels in the synbiotics group compared to the placebo group (\(p=0.438\)). This study found an improvement in constipation symptoms (\(p = 0.006\)) and constipation-related quality of life (\(p=0.001\)) after synbiotic administration.

Conclusion: Two months of synbiotic supplementation did not lower indoxyl sulphate toxin levels. Nevertheless, it had a major effect in improving constipation and quality of life affected by constipation in patients undergoing chronic haemodialysis.

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Introduction

Dysbiosis is an imbalance in the gut microbiota that causes changes in the number and activities of the microbiota in the gastrointestinal tract. Conditions in chronic kidney disease (CKD) that serve as risk factors for gut microbiota dysbiosis include a low-fibre diet, uraemia, prolonged colonic transit time, disturbance of protein assimilation, use of drugs such as antibiotics, phosphate-binding agents, and iron supplements, as well as other comorbidities [1]. Dysbiosis in CKD is characterized by an increase in the activity of proteolytic bacteria, such as the family Enterobacteriaceae (especially Enterobacter, Klebsiella and Escherichia), Enterococci, Clostridium perfringens and Pseudomonas, accompanied by a decrease in the activity of saccharolytic bacteria, such as the family Bifidobacteriaceae (especially Bifidobacterium, Lactobacillaceae and Prevotellaceae). This leads to an increase in the production of uraemic toxins, such as indoxyl sulphate (IS), p-cresyl sulphate (p-CS) and indole acetic acid (IAA), as well as a decrease in the production of short-chain fatty acids (SCFAs), such as butyrate and propionate [2].

Indoxyl Sulphate (IS) is a result of tryptophan metabolism by bacteria with the tryptophanase enzyme in the colon. In normal kidneys, this toxin is excreted through the secretion process in the tubules. This toxin is 90% bound to albumin in the circulation and, therefore, cannot be eliminated through the haemodialysis process [3]. As kidney function decreases, the accumulation of IS increases in the body of haemodialysis patients. IS activates nuclear factor-κB (NF-κB) and the expression of plasminogen activator inhibitor (PAI) type 1 in proximal tubular cells, mediating tubulointerstitial fibrosis, and is strongly associated with the progression of kidney damage. Additionally, IS is associated with aortic calcification, vascular stiffness, and increased oxidative stress in vascular endothelial cells, leading to increased cardiovascular risks in patients with CKD [4].

In addition to being associated with an increase in uraemic toxins, dysbiosis of the gut microbiota is also related to constipation, as it is one of the most frequent gastrointestinal symptoms experienced by haemodialysis patients [5]. Inflammation, increased uraemic toxin, and decreased butyrate acid are suspected to affect intestinal motility [6]. With inadequate treatment, persistent constipation affects patients’ mental and physical quality of life. Zhang et al. [7] reported that haemodialysis patients with constipation had lower scores on physical and mental health.

Improvement of dysbiosis by synbiotic administration is expected to lower IS toxin levels, improve constipation symptoms, and enhance constipation-related quality of life. However, currently available studies were conducted with small sample sizes, and not all of them were designed as double-blind randomized clinical trials. The results of these studies vary and cannot provide full evidence of the role of synbiotics in the improvement of gut dysbiosis, decreased uraemic toxins, and gastrointestinal symptoms [8–11].

This study aimed to show the benefit of synbiotic administration in lowering uraemic toxins, specifically IS, and to investigate its benefits on constipation symptoms and the quality of life of haemodialysis patients in Indonesia.

Methods

Trial design

This study was a double-blinded randomized controlled clinical trial with a parallel design organized in Dr. Cipto Mangunkusumo Hospital, a national referral hospital in Jakarta, Indonesia, from August through December 2020.

Participants

Subjects included CKD patients on haemodialysis who met the inclusion criteria and were recruited using a consecutive sampling method.

The inclusion criteria were as follows: 1) patients over 18 years old who underwent standard haemodialysis treatment twice a week for five hours for at least three months and 2) patients with gastrointestinal complaints (i.e., difficulty defecating, faeces with hard consistency, or a bowel movement frequency of fewer than three times a week). Patients with a history of malignancy, chemotherapy or radiotherapy, patients with autoimmune disorders or receiving immunosuppressants, patients who underwent gut resection, patients with Crohn's disease or ulcerative colitis, patients whose haemodialysis schedule was altered, patients consuming prebiotics/probiotics/synbiotics, and patients suffering from infection or consuming antibiotics were excluded from this study.

Intervention

All included subjects underwent history taking, physical examination, and laboratory examination (blood) and provided consent to participate in the study. The subjects then underwent history taking for demographic data, comorbidities and medications as well as physical examination for vital signs, body weight, and body height. We
tested the blood sample to evaluate haemoglobin, leucocytes, thrombocytes, urea, creatinine, and albumin to determine baseline characteristics.

After randomization, each subject received 60 capsules containing synbiotics (Lactobacillus acidophilus and Bifidobacterium longum 5x10⁹ CFU and 60 mg of fructooligosaccharides (FOS)) or 60 capsules containing placebo (Saccharum lactis). The daily dosage was two capsules per day taken every morning before a meal. All subjects were given a compliance card that had to be completed every day after they took the drug. Medication adherence was assessed every 30 days, and the subjects returned any medicine left and showed the compliance card at the subsequent follow-up.

Food intake was assessed before and after the intervention was administered using 24-hour food recall and was carried out by a nutritionist. To evaluate caloric intake and carbohydrates, protein, fat, and fibres consumed by the patient throughout the study, a programme (nutriSurvey) was used. In addition, all participants were asked to continue their dietary habits and previous lifestyle habits and were prohibited from consuming any motility agents during the study period.

After 30 days, a follow-up was performed to evaluate side effects and compliance with the drug regimen. Each subject was then provided with another 60 synbiotic capsules or 60 placebo capsules based on their previous grouping.

The exclusion criteria included subjects who withdrew from the study, those who missed their dose of synbiotics or placebo for more than three consecutive days, those with infections that required antibiotics, those whose haemodialysis schedule changed from twice a week to three times a week or whose treatment modality changed from dialysis to peritoneal dialysis or to renal transplant and those who experienced gastrointestinal symptoms, such as diarrhoea or profuse vomiting requiring hospital admission. Patient adherence to the medication under investigation was expected to reach over 90%.

Outcome
The primary outcome of this study was a decrease in IS levels. The examination was conducted twice, before and after the intervention was administered. Blood samples were taken predialysis and after each subject fasted for ±8-10 hours. Approximately 100 μL of each subject’s serum was added to 900 L of acetonitrile to precipitate the protein. The supernatant was then added to 500 μL of 5 M NaCl for salting-out-assisted liquid–liquid extraction (SALLE) and centrifuged at 14000 rpm for 10 minutes. As a result, two phases were formed: the organic phase (IS in acetonitrile and internal standard) and the aqueous, NaCl and matrix constituent phases. The organic phase was then separated, and as much as 0.2 L was injected into the HPLC system with a fluorescence detector (Agilent Technologies with MassHunterChemStation Software version B04.03.E). IS was measured quantitatively using seven calibration levels with a calibration range of 0.02-100 mg/L. The concentration of IS is expressed in units of mg/L.

The secondary outcome of this study was improvement in constipation-related symptoms and quality of life. They were assessed using the Indonesian-validated Patient Assessment of Constipation: Symptoms (PAC-SYM) and Patient Assessment of Constipation Quality of Life (PAC-QOL) questionnaires [12, 13]. The examination was conducted twice, at the beginning and the end of the study. The PAC-SYM questionnaire includes a 0-4 scale and consists of three parts: abdominal symptoms (questions 1-4), rectal symptoms (questions 5-7), and stool symptoms (questions 8-12). Symptom improvement was defined as a decrease in the total PAC-SYM score > 1. The PAC-QOL questionnaire includes a 0-4 scale and consists of four parts: physical discomfort (questions 1-4), psychosocial discomfort (questions 5-12), worries/concern (questions 13-23), and satisfaction (question 24-28). Quality of life improvement was defined as a decrease in the total PAC-QOL score > 1.

Sample size
To estimate the sample size, a formula for comparing two means was used. With α of 0.05, power of 0.80, and dropout of 20%, the minimum sample size for each group was 30.

Randomization
We only enrolled subjects who gave consent. A third party (pharmacist) randomized the subjects into two study groups using a computer randomizer. Synbiotics and placebo were both packed on an identical clear gastroenteric coated capsule. Each subject was given one plastic pot containing either intervention or placebo drugs; each identical pot contained 60 capsules. A white paper with information on drug use and administration was placed on the cover of the pot. The drugs were distributed by a third party. None of the patients, researchers, or physicians in charge were aware of the treatment groups.

Statistical methods
We performed the statistical analysis using SPSS version 20.0. Mean and standard deviation (SD) analyses were performed for numeric data with normal distributions, whereas medians and interquartile ranges (IQRs) were calculated for data with nonnormal distributions. This study utilized intention-to-treat analysis. Bivariate
analysis using an independent t test was performed to analyse data with a normal distribution, and the Mann–Whitney U test was performed to analyse data with a nonnormal distribution. A p value of <0.05 was considered statistically significant.

**Results**

Out of 100 screened patients, 60 met the inclusion criteria. These patients were randomized into either the synbiotics or the placebo group. Twenty-seven subjects in the synbiotics group and 30 subjects from the placebo group completed the study. In the synbiotics group, two subjects refused to continue the study, whereas one dropped out due to receiving antibiotics for 15 days during the course of the study. The research algorithm is shown in Fig. 1. Baseline demographic characteristics as well as IS levels and PAC-SYM and PAC-QOL scores are provided in Table 1.

**Effects of synbiotic supplementation on indoxyl sulphate toxin levels**

In the synbiotics group, the median IS toxin levels pre- and postintervention were 26.98 mg/L (22.78-34.77 mg/L) and 27.94 mg/L (23.25-34.05 mg/L), respectively.

The initial and posttreatment median IS toxin levels of the placebo group were 20.95 mg/L (17.25-27.07 mg/L) and 22.88 mg/L (18.20-29.38 mg/L), respectively (Table 2). The synbiotics group showed an increased level of IS of 0.17 mg/L (-2.67-4.89 mg/L), whereas the placebo group showed an increased level of 0.67 mg/L (-3.01-2.14 mg/L). A nonparametric Mann–Whitney bivariate analysis test was used to compare the difference in IS levels pre- and postintervention in both groups. In Table 2, compared to placebo, no significant changes in IS levels were observed after synbiotic administration (p=0.438).

**The effect of synbiotic administration on constipation symptoms according to the PAC-SYM questionnaire**

Table 3 shows a significant difference in abdominal symptoms (questions 1-4) between the synbiotics and placebo groups (p = 0.023). We also observed significant improvement in rectal symptoms (questions 5-7) and stool symptoms (questions 8-12) after administering synbiotics. There was an overall improvement in constipation in the group that received synbiotics compared to the placebo group (p = 0.006). This study also analysed dietary intake before and after intervention and found no difference in dietary pattern in terms of caloric intake.

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![Research algorithm according to Consolidated Standards of Reporting Trials (CONSORT)](image-url)
Table 1  Initial characteristics of study subjects

| Characteristics                        | Symbiotics (N=30) | Placebo (N=30) | p value |
|----------------------------------------|-------------------|----------------|---------|
| Gender, n(%)                           |                   |                |         |
| Male                                   | 10 (33.3)         | 11 (36.7)      | 0.787a  |
| Female                                 | 20 (66.7)         | 19 (63.3)      |         |
| Age (years), mean (SD)                 | 51.23 (13.57)     | 52.33 (11.29)  | 0.734b  |
| HD duration (months), median (IQR)     | 70.50 (46.25-118.75) | 57.50 (29.5-99.7) | 0.329c  |
| Comorbidities, n(%)                    |                   |                |         |
| - Hypertension                         | 25 (83.3)         | 25 (83.3)      | 0.635d  |
| - Diabetes                             | 11 (36.7)         | 8 (26.7)       | 0.405a  |
| - Heart failure                        | 5 (16.7)          | 5 (16.7)       | 0.635d  |
| - Coronary heart disease               | 4 (14.3)          | 2 (6.7)        | 0.335d  |
| - Stroke                               | 1 (3.3)           | 3 (10)         | 0.306d  |
| Medication taken, n(%)                 |                   |                |         |
| - ACEi/ARB                             | 12 (40.0)         | 14 (46.7)      | 0.602a  |
| - CCB                                  | 20 (66.7)         | 18 (60.0)      | 0.592a  |
| - Beta blocker                         | 3 (10.0)          | 9 (30)         | 0.053a  |
| - Alpha blocker                        | 14 (46.7)         | 9 (30)         | 0.184a  |
| - Insulin                              | 8 (26.7)          | 3 (10)         | 0.095a  |
| - Oral hypoglycaemic drugs             | 3 (10.0)          | 4 (13.3)       | 0.500d  |
| - Phosphate binder                     | 28 (93.3)         | 27 (90.0)      | 0.640d  |
| - Oral iron supplements                | 1 (3.3)           | 3 (10.0)       | 0.306d  |
| HD adequacy (Kr/V), mean (SD)          | 1.92 (0.35)       | 1.83 (0.30)    | 0.284b  |
| Body mass index (kg/m²), mean (SD)     | 22.56 (4.80)      | 24.22 (4.64)   | 0.180b  |
| Laboratory                             |                   |                |         |
| - Haemoglobin (g/dL), median (IQR)     | 9.4 (7.9-10.9)    | 9.4 (8.80-10.42) | 0.487c  |
| - Leucocyte (/uL), mean (SD)           | 8142 (2380.9)     | 7978 (1783.9)  | 0.764b  |
| - Thrombocyte (/uL), mean (SD)         | 257933.3 (61952.11) | 244333.3 (63318.39) | 0.407b  |
| - Albumin (mg/dL), mean (SD)           | 3.90 (0.28)       | 3.87 (0.4)     | 0.789b  |
| - Urea (mg/dL), mean (SD)              | 144.77 (44.83)    | 131.94 (38.45) | 0.148b  |
| - Creatinine (mg/dL), mean (SD)        | 11.86 (3.46)      | 12.35 (3.7)    | 0.547b  |
| Dietary intake                         |                   |                |         |
| - Calories (kcal/kg/day), median (IQR) | 28.82 (22.32-36.34) | 26.84 (18.41-36.21) | 0.220c  |
| - Protein (gram/kg/day), mean (SD)     | 0.91 (0.38)       | 0.79 (0.30)    | 0.178b  |
| - Carbohydrate (gram/day), mean (SD)   | 183.98 (71.05)    | 206.99 (94.99) | 0.293b  |
| - Fat (gram/day), mean (SD)            | 70.02 (20.9)      | 64.49 (20.51)  | 0.306b  |
| - Fibre (gram/day), mean (SD)          | 6.09 (3.76)       | 6.11 (3.33)    | 0.980b  |
| Indoxyl sulphate (mg/L), median (IQR)  | 26.98 (22.78-34.77) | 20.95 (17.25-27.07) | 0.062b  |
| PAC-SYM score, median (IQR)            | 8 (5-11.25)       | 6 (4-9.5)      | 0.137c  |
| PAC-QOL score, median (IQR)            | 17.5 (14-24)      | 18 (13-26.25)  | 0.970c  |

ACEi angiotensin-converting enzyme inhibitor, ARB angiotensin receptor blocker, CCB calcium channel blocker, HD haemodialysis, PAC-SYM Patient Assessment of Constipation Symptoms, PAC-QOL Patient Assessment of Constipation Quality of Life

Data were analysed using the *chi-square test,* *t* test, *Mann–Whitney U test,* and *Fisher test*

Table 2  The effects of synbiotics and placebo supplementation on indoxyl sulphate levels

| Indoxyl sulphate | SYNBIOtICS (N=30) | PLACEBO (N=30) | P   |
|------------------|-------------------|----------------|-----|
|                  | Pre               | Post           | Δ   | P   |
|                  | ( median (IQR))   | (median (IQR)) | (Δ) |   |
| Median (IQR) (mg/L) | 26.98 (22.78-34.77) | 27.94 (23.25-34.05) | -0.17 | 0.728a  |
|                  | 20.95 (17.25-27.07) | 22.88 (18.20-29.38) | -0.67 | 0.586 a  |

* Pre- and postintervention were analysed using the Wilcoxon test

* Delta IS between groups was analysed using the Mann–Whitney test
and protein, carbohydrate, fat, and fibre consumption, which can affect defecation patterns.

Table 4 shows significant improvement in physical discomfort (questions 1-4), psychosocial discomfort (questions 5-12), and worries/concerns (questions 13-23) after synbiotic administration compared to placebo. However, synbiotic administration did not show significant results in terms of satisfaction (questions 24-28). Synbiotic administration showed an overall significant difference in quality of life caused by constipation ($p=0.001$).

### Adverse events observed during the course of the study

During the course of this study, six subjects in the synbiotics group and two subjects in the placebo group complained of diarrhoea (two to five times a day) during the first two weeks of intervention. Five subjects showed improvement without antidiarrheal treatment, and three subjects required antidiarrheal treatment to recover. Despite recovery in all patients suffering from diarrhoea, two subjects in the synbiotics group opted to discontinue their participation in this study and were therefore considered dropouts. Two subjects in the synbiotics group and three subjects in the placebo group experienced nausea. One subject in the placebo group needed a proton pump inhibitor for two days, whereas the rest improved without additional treatment. As shown in Table 5, one subject in the synbiotics group felt bloated, and two subjects in the placebo group complained of colicky pain, but all improved without any additional treatment. No subjects died or required hospitalization.

Six subjects (out of 27) in the synbiotics group and seven subjects (out of 30) in the placebo group had an adherence of less than 100%. The mean adherence in subjects who completed this study was 99.4% and 99.2% in the synbiotics and placebo groups, respectively.

### Discussion/conclusion

The excretion of uraemic toxins, such as IS, decreases with declining kidney function. Although haemodialysis is an advanced kidney replacement therapy, IS still cannot be completely eliminated. This is because this

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**Table 3** The effects of synbiotics and placebo supplementation on constipation symptoms based on PAC-SYM questionnaire

| PAC-SYM   | SYNBIOtICs (n=30) | PLACEBO (n=30) | P   |
|-----------|-------------------|----------------|-----|
|           | Pre | Post | Pre | Post |
| Abdominal symptoms (questions 1-4), median (IQR) | 3 (0-9) | 0 (0-3) | 2 (0-9) | 1 (0-8) | 0.023$^a$ |
| Rectal symptoms (questions 5-7), median (IQR) | 0 (0-7) | 0 (0-2) | 0.5 (0-7) | 0 (0-4) | 0.025$^a$ |
| Stool symptoms (questions 8-12), median (IQR) | 4 (0-14) | 1 (0-6) | 3 (0-12) | 2 (0-7) | 0.047$^a$ |
| Total score, median (IQR) | 8 (2-25) | 1 (0-8) | 6 (1-28) | 5.5 (0-17) | 0.006$^a$ |

$^a$ Postintervention data were analysed using the Mann–Whitney test

**Table 4** The effects of synbiotics and placebo supplementation on constipation-related quality of life based on PAC-QOL questionnaire

| PAC-QOL | SYNBIOtICs (n=30) | PLACEBO (n=30) | P   |
|---------|-------------------|----------------|-----|
|         | Pre | Post | Pre | Post |
| Physical discomfort (questions 1-4), median (IQR) | 2 (0-8) | 0 (0-2) | 2 (0-11) | 0.5 (0-10) | 0.007$^a$ |
| Psychosocial discomfort (questions 5-12), median (IQR) | 2 (0-16) | 0 (0-3) | 3 (0-12) | 2 (0-11) | 0.000$^a$ |
| Worry/concerns (questions 13-23), median (IQR) | 6 (2-29) | 0 (0-9) | 4.5 (0-24) | 3 (0-12) | 0.001$^a$ |
| Satisfaction (questions 24-28), median (IQR) | 8 (4-12) | 9 (7-13) | 9.5 (2-17) | 8.5 (4-14) | 0.609$^a$ |
| Total score, median (IQR) | 18 (10-57) | 11 (8-21) | 18 (10-44) | 16 (7-33) | 0.001$^a$ |

$^a$ Postintervention data were analysed using the Mann–Whitney test

**Table 5** Postintervention adverse events

| Components | SYNBIOtICs (n=30) N (%) | PLACEBO (n=30) N (%) | p value |
|------------|-------------------------|----------------------|---------|
| - Diarrhoea | 6 (20) | 2 (6.7) | 0.259$^a$ |
| - Nausea/vomiting | 2 (6.7) | 3 (10) |
| - Bloating | 1 (3.3) | 0 (0) |
| - Stomachache/heartburn | 0 (0) | 2 (6.7) |
| Hospitalization | 0 (0) | 0 (0) |
| Death | 0 (0) | 0 (0) |

$^a$ Data were analysed using the chi-square test
toxin is large, and 90% is bound to albumin [14]. Various strategies to improve IS removal, including (using) haemodiafiltration machines, increasing dialysate flow, increasing dialyzer membrane size, or adding a sorbent to the dialysate, have not shown promising results [15–17]. This suggests the need for an alternative way to lower the level of IS in CKD or CKD-HD patients. It was expected that the administration of synbiotics would suppress IS production in the gastrointestinal tract by improving gut microbiota dysbiosis. A previous study by Cossola C et al. found that a combination of prebiotics and probiotics restored the balance of gut microbiota and suppressed IS production [18]. However, some studies and a recent meta-analysis showed no correlation between synbiotics and IS reduction in CKD-HD subjects [8, 9, 19, 20]. Likewise, our study found nonsignificant results.

The reasons behind these findings might be because a 60-day administration and observation period was not sufficient to demonstrate a significant IS reduction. Thus, a longer observation period and serial examination of IS might be needed. Another factor that might have played a role in the findings is the fact that a previous study used a synbiotic combination that differed from ours. The synbiotics used by previous studies contained different strains of bacteria and higher amounts of prebiotics, which might have led to various effects [8, 9, 18–20]. Furthermore, the results of these studies show the importance of determining the type, dosage and duration of administration of synbiotics to suppress IS levels [8, 9, 18–20]. Additionally, this illustrates the need for faecal microbiota analysis to determine the most suitable synbiotics to be given to CKD-HD patients. In Indonesia, intestinal microbiota profile examination has never been performed on CKD-HD patients, and to date, there are no accurate data on the most suitable synbiotics for CKD-HD patients in Indonesia.

Saccharolytic bacteria provide benefits in improving intestinal microbiota homeostasis, lowering the amount of pathogenic bacteria, improving intestinal transit time, increasing the frequency of defecation, improving faecal consistency, decreasing bloating symptoms and producing butyric acid to increase intestinal barrier functions [21]. The presence of saccharolytic bacteria is expected to lower the amount and activity of proteolytic bacteria, which lowers the level of uraemic toxins, such as IS. The presence of fructooligosaccharides (FOS) modulates the growth of bacteria, such as Bifidobacterium, and increases the ratio of Roseburia/E. rectal, which plays a role in increasing the level of butyrate acid, which in turn serves as the source of energy in the regeneration of cells in the host’s intestines. Fructooligosaccharides also serve as substrates for saccharolytic bacteria contained in the synbiotic preparation, leading to increased SCFA production and suppressed uraemic toxin production by proteolytic bacteria [22].

Patients with CKD experience constipation more often than the normal population. This is due to the restriction of fibre and water, reduced physical activity, use of medications such as oral iron supplements, and gut microbiota dysbiosis in haemodialysis patients [23]. An in vitro study reported an inflammation process due to gut-derived uraemic toxin disrupting intestinal motility. Nishiyama et al. demonstrated that rats with CKD also had gut dysbiosis, reduced intestinal motility, reduced amounts of faeces, and intestinal inflammation [24]. A study by Ramos et al. showed that CKD patients suffering from constipation tended to have higher uraemic toxin levels [25]. Constipation increases colonic transit time, leading to an increase in the activity of proteolytic bacteria in metabolizing amino acids and producing uraemic toxins. The presence of uraemic toxin itself may worsen constipation. In other words, constipation and gut dysbiosis affect each other [26]. A study by Salmean et al. showed an improvement in defecation frequency among 13 CKD patients receiving high fibre supplementation for 12 weeks [27]. In our study, we found that the administration of synbiotics capsules improved complaints related to constipation symptoms patterns, as shown in the improved score on the PAC-SYM questionnaire. This is because the administration of synbiotics increases the amount and activity of saccharolytic bacteria in producing butyric acid. Butyric acid is the source of energy for the regeneration of colonocytes; thus, the increase in its production improves intestinal motility and contractility, lowering colonic transit time. Additionally, improving gut dysbiosis decreases intestinal inflammation, which causes disturbances in intestinal motility [6, 28].

Constipation, if not well treated, may affect the quality of life in haemodialysis patients. Zhang et al. reported that haemodialysis patients with constipation symptoms tended to have poorer quality of life and were prone to depression compared to subjects without constipation [7]. Ranganathan et al. reported that the administration of probiotics for six months in stage 3-4 CKD patients increased their quality of life [29]. A study by Haghihat et al. showed that the administration of synbiotics in haemodialysis patients improved their mental status, including depression and anxiety symptoms, but was not associated with significant improvement in terms of quality of life [30]. The results of this study were consistent with those of previous studies, which showed an improvement in terms of constipation-related quality of life as assessed using the PAC-QOL questionnaire. We observed significant improvements in terms of physical discomfort, psychosocial discomfort, and worries, although we did not observe significant improvement.
in terms of patient satisfaction. To date, current studies have shown various results. The pathogenesis of how synbiotics may affect the quality of life and the possibility of a gut-brain axis role are not yet completely understood. Hence, the need for further studies remains.

To the best of our knowledge, this is the first study in Indonesia that successfully showed the efficacy of synbiotic supplementation on the improvement of constipation symptoms and constipation-related quality of life in patients undergoing haemodialysis, although its efficacy in decreasing the IS level has not yet been established. This study has several limitations. We did not perform genomic analysis of gut microbiota on patients’ faecal samples to assess dysbiosis patterns in Indonesian CKD-HD patients and determine whether there was any change in dysbiosis patterns after synbiotic administration. Moreover, this study only included subjects who received haemodialysis twice a week, so attention is needed to generalize this study’s results.

In conclusion, the administration of synbiotics containing Bifidobacterium longum, Lactobacillus acidophilus (5×10⁹ CFU), and 60 grams of fructo-oligosaccharides (FOS) in two capsules per day for 60 days has not been shown to reduce levels of IS toxin but can improve constipation symptoms and quality of life associated with constipation in CKD-HD patients.

**Abbreviations**

CKD-HD: Chronic Kidney Disease on Haemodialysis; CFU: Colony-Forming Unit; FOS: Fructooligosaccharides; IS: Indoxyl Sulphate; P-CS: P-Cresyl Sulphate; IAA: Indole Acetic Acid; SCFA: Short Chain Fatty Acid; NF-Kβ: Nuclear Factor-Kβ; PAI: Plasminogen Activator Inhibitor; PAC-SYM: Patient Assessment of Constipation-Symptoms; PAC-QOL: Patient Assessment of Constipation Quality of Life; SD: Standard Deviation; IQR: Interquartile Range; SCFA: Short-Chain Fatty Acids.

**Supplementary Information**

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Not applicable.

**Authors’ contributions**

Study concept and design: AL and TAI. Acquisition of the data: TAI. Analysis and interpretation of the data: AL, TAI, and AR. Drafting of the manuscript: AL and TAI. Critical revision of the manuscript for important intellectual content: AR, MA, and AL; statistical analysis: AL, TAI, and AR. Administrative, technical, and material support: TAI. Study supervision: AL, AR, and MA. The authors read and approved the final manuscript.

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**Availability of data and materials**

The data that support the findings of this study are available from the corresponding author, AL, upon reasonable request.

**Declarations**

**Ethics approval and consent to participate**

This study was conducted according to the principles in the Helsinki Declaration and the Guideline for Good Clinical Practice from ICH Tripartite Guideline (ICH-GCP). This study received ethical clearance on July 20, 2020, according to the decision of the Permanent Ethical Committee of Medical Research in Faculty of Medicine Universitas Indonesia/RSUPN Dr. Cipto Mangunkusumo Jakarta according to letter number KET.777/UN2.F1/ETIK/PPM.00.02/2020. All subjects provided consent to participate in the study after being informed both orally and in written form of the goals and procedures of the study. Informed consent was obtained from all the participants. This study was also included in the clinical trial database on www.clinicaltrials.gov with registration number NCT04527640.

**Consent for publication**

Not applicable.

**Competing interests**

The authors have no conflicts of interest to declare.

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**References**

1. Armani R, Ramezani A, Yasir A, et al. Gut microbiome in chronic kidney disease. Curr Hypertens Rep. 2017;19(4):29.
2. Rukavina Mikusic NL, Kouyoumdzian NM, Choi MR. Gut microbiota and chronic kidney disease: evidences and mechanisms that mediate a new communication in the gastrointestinal-renal axis. Pflugers Arch Eur J Physiol. 2020;472:303–20.
3. Lekawanvijit S, Kompa AR, Wang BH, Kelly DJ, Krum H. Cardiorenal syndrome: the emerging role of protein-bound uremic toxins. Circ Res. 2012;111(11):1470–83.
4. Ramezani A, Massy ZA, Meijers B, et al. Role of the gut microbiome in Uremia: A potential therapeutic target. Am J Kidney Dis. 2016;67:483–98.
5. Zuvela J, Tringingham C, Le Leu R, et al. Gastrointestinal symptoms in patients receiving dialysis: A systematic review. Nephrology. 2018;23:718–27.
6. Ikei R, Yano K, Tsuru T. Constipation in chronic kidney disease: It is time to reconsider. Ren Replace Ther. 2019;5:51.
7. Zhang J, Huang C, Li Y, et al. Health-related quality of life in dialysis patients with constipation: A cross sectional study. Patient Prefer Adher. 2013;7:589–94.
8. Nakabayashi I, Nakamura M, Kawakami K, et al. Effects of synbiotic treatment on serum level of p-cresol in haemodialysis patients: A preliminary study. Nephrol Dial Transplant. 2011;26:1094–8.
9. Rossi M, Johnson DH, Morrison M, et al. Synbiotics easing renal failure by improving gut microbiology (SYNERGY): A randomized trial. Clin J Am Soc Nephrol. 2016;11:223–31.
10. Guida B, Germano R, Tiro R, et al. Effect of short-term synbiotic treatment on plasma p-cresol levels in patients with chronic renal failure: A randomized clinical trial. Nutr Metab Cardiovasc Dis. 2014;24:1043–9.
11. Cruz-Mora J, Martínez-Hernández NE, Martín del Campo-López F, et al. Effects of a Symbiotic on Gut Microbiota in Mexican Patients With End-Stage Renal Disease. J Ren Nutr. 2014;24:330–5.
12. Abdullah M, Maulahela H, Utari AP, Soebandrio A, et al. Patient assessment of constipation quality of life questionnaire: validity and reliability for Indonesian population. Med J Indones. 2019;28:345–50.
13. Abdullah M, Maulahela H, Utari AP, Soebandrio A, et al. Validity and Reliability of the Patient Assessment of Constipation: Symptoms (PAC-SYM) in the Indonesian Language. JOP Conf Ser: Earth Environ Sci. 2019;248:012060. https://iopscience.iop.org/article/10.1088/1755-1315/248/1/012060.
14. Leong SC, Sirich TL. Indoxyl sulfate-review of toxicity and therapeutic strategies. Toxins (Basel). 2016;8(12):358. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5198552/.
15. Meyer TW, Peattie JD, Miller JD, et al. Increasing the clearance of protein-bound solutes by addition of a sorbent to the dialysate. J Am Soc Nephrol. 2007;18(3):686–74.
16. Krieter DH, Hackl A, Rodriguez A, et al. Protein-bound ureaemic toxin removal in haemodialysis and post-dilution haemodiafiltration. Nephrol Dial Transplant. 2010;25(1):212–8.
17. Meert N, Watellos MA, Van Landschoot M, et al. Prospective evaluation of the change of predialysis protein-bound uremic solute concentration with postdilution online hemodiafiltration. Artif Organs. 2010;34(7):580–5.
18. Cosola C, Rocchetti MT, Di Bari I, et al. An innovative symbiotic formulation decreases free serum indoxyl sulfate, small intestine permeability and ameliorates gastrointestinal symptoms in a randomized pilot trial in stage IIIb-IV CKD patients. Toxins (Basel). 2021;13(5):334. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8147955/.
19. Nguyen TTU, Kim HW, Kim W. Effects of Probiotics, Prebiotics, and Synbiotics on Uremic Toxins, Inflammation, and Oxidative Stress in Hemodialysis Patients: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. J Clin Med. 2021;10(19):4456.
20. McFarlane C, Krishnasamy R, Stanton T, Emma S, Matthew S, Mihala G, et al. Synbiotics Easing Renal Failure by Improving Gut Microbiome II (SYNERGY II): A Feasibility Randomized Controlled Trial. Nutrients. 2021;13:4481.
21. Hewadmal N, Jangra S. A Review on Probiotic and Health Benefits of Probiotics. Int J Curr Microbiol App Sci. 2019;8:1863–80.
22. Guarino MPL, Altomare A, Emerenzi S, et al. Mechanisms of action of prebiotics and their effects on gastro-intestinal disorders in adults. Nutrients. 2012. Epub ahead of print 2020:10.3390/nu12041037.
23. Sumida K, Yamagata K, Kovesdy CP. Constipation in CKD. Kidney Int Rep. 2020;5:121–34.
24. Nishiyama K, Aono K, Fujimoto Y, et al. Chronic kidney disease after 5/6 nephrectomy disturbs the intestinal microbiota and alters intestinal motility. J Cell Physiol. 2019;234:6667–78.
25. Ramos CI, Armani RG, Canziani ME, et al. Bowel Habits and the Association With Uremic Toxins in Non-Dialysis-Dependent Chronic Kidney Disease Patients. J Ren Nutr. 2020;30:31–5.
26. Ikee R, Sasaki N, Yasuda T, et al. Chronic kidney disease, gut dysbiosis, and constipation. A burdensome triplet. Microorganisms. 2020;8:1–18.
27. Salmean YA, Segal MS, Pali SP, et al. Fiber supplementation lowers plasma p-cresol in chronic kidney disease patients. J Ren Nutr. 2015;25:316–20.
28. Zhao Y, Yu YB. Intestinal microbiota and chronic constipation. Springerplus. 2016;5:1130.
29. Ranganathan N, Ranganathan P, Friedman EA, et al. Pilot study of probiotic dietary supplementation for promoting healthy kidney function in patients with chronic kidney disease. Adv Ther. 2010;27:634–47.
30. Haghighat N, Mohammadshahi M, Shayanpour S, et al. The effect of symbiotic and probiotic supplementation on mental health parameters in patients undergoing hemodialysis: A double-blind, randomized, placebo-controlled trial. Indian J Nephrol. 2021;31:149–56.