Near and Mid Infrared Spectroscopy to Assess Milk Products Quality: A Review of Recent Applications

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Abstract

The requirement of food safety and the recently increase in the consumption of dairy products in worldwide have drawn great attention for potential researches in these areas. Dairy products as milk, cheese, milk powder, whey milk powder, whey concentrated and isolated powder have a considerable demand and so are vulnerable to economic adulteration. Techniques of infrared spectroscopy have been used not only for the authenticity of dairy products, but also to their determination of quality, assisting issues as cheese ripening, composition and contamination. Based upon these applications and considering the importance of monitoring food quality this review presents some of the latest researches using infrared spectroscopy techniques for assessing intrinsic quality of dairy products.

Keywords: Adulteration; Dairy products; Food quality; Spectroscopic techniques

Introduction

Milk and dairy products are important sources of nutrients for human health [1]. In the last years, it has been observed an increase in their consumption. More than 6 billion people worldwide consume milk and milk products, which make them of paramount commercial importance within the food industry and plays an important role in the country’s economy [2,3]. Besides, great attention has been paid to the development of innovative and high-value dairy products that better fit consumers’ desires, which are mainly related to functional aspects important for human nutrition and health [1]. At the same time, over the past decade, the authentication of milk products has become a crucial issue once several major adulteration events were discovered [4]. The replacement of original substance partially or completely with more easily available and cheap compound is the most common procedure performed by defrauders. Those products presenting high-value and, which undergo a number of processing steps before they appear on the market are the most vulnerable for adulteration [5,6]. Fraud not only generates a product with food quality problems, but can also pose serious threats to consumers’ health [4,7]. Moreover, there are other problems, such as the contamination of milk by residues of veterinary drugs that may be present when the cow is milked in grace period. The most common drugs are antimicrobials and anti-inflammatoryities [2,8,9]. Therefore, the determination of food quality and authentication is an important issue for final consumer’s satisfaction, food processors and regulatory authorities [10]. For this, it is important to ensure the ingredient authenticity by monitoring continuously their quality in food industry, through the development of sophisticated and accurate standardized new techniques [7,11].

Infrared spectroscopy is increasingly the most used method for determining authenticity and adulteration in foods [12] and one of the most relevant technologies in the analysis of raw materials, process control and specifications of final products in the dairy industry [1]. Techniques like Near-Infrared (NIR) and Mid-Infrared (MIR) have been successfully applied in the assessment of milk and dairy products quality, including milk powder, whey, WPCs and cheeses [13-17]. Figure 1 furnishes some analysis using MIR infrared for some types of cheeses and whey protein concentrate.

The infrared spectroscopy techniques are based on the analysis of the interaction between electromagnetic waves and matter [19] and the figure 2 shows a representation of this interaction.
The basic assumption behind the application of optical spectroscopic techniques to the quality evaluation of foods relies on the generation of a “fingerprint”. An individual dairy product with a given chemical composition exposed to a light source will produce a characteristic spectrum, which is the result of the absorption of the electromagnetic energy by the various chemical constituents of the food [6].

Infrared spectroscopy has the advantage of being non-destructive and is suitable for use in a research laboratory or in an industrial environment (in online and online applications at different stages).

Laboratory techniques are generally developed and used mainly in research before the technology is applied in the industrial field. During this period, experiments carried out determine basic principles and obstacles and propose new methodologies to face problems and improve results [21]. Regarding the industrial setting, overall, the online applications are limited, but they refer to process control through the monitoring of crucial traits (e.g., recording of real-time production data) that allow rapid action and cost saving. The on-line applications are mainly related to end product labeling specification based on the development of prediction models using reference methods [1]. Both applications can increase efficiency, improve profitability, and enhance quality of the products. Thus, the infrared spectroscopy techniques provide an alternative to the traditional analytical strategies that normally are relied on wet chemistry to determine the amount of a marker compound or compounds in a suspect material [6]. Besides, most of them make use of hazardous chemicals, are labour intensive and time-consuming with requirements of extensive sample preparation, as well as skilled and experienced staff [7].

NIR infrared spectroscopy uses the near-infrared region of the electromagnetic spectrum (from 750 - 2500 nm or 14290 - 4000 cm⁻¹) and is based on molecular overtone and combination vibrations [8,22]. The spectra can be recorded in reflection, transmission, or interaction modes providing complex chemical and physical information. In particular, overtones and combinations of the vibrations such as C-H, O-H, N-H, and C=O make it possible the measurements of various structures [23].

MID infrared spectroscopy (from 2500 - 25000 nm or 4000 - 400 cm⁻¹) monitors the fundamental vibrational and rotational stretching modes of molecules, which reflect the chemical profile of the sample [8,22,24]. Once the technique measures the fundamental vibrations instead of the overtones and combination bands measured in the NIR region, it provides a greater amount of chemical information regarding the scanned sample. The main bands obtained in the MIR spectra belong to two distinctive regions: the one called “functional group” (4000 to 1500 cm⁻¹) and the one known as “fingerprint region” (1500 to 500 cm⁻¹) [4]. However, most of the relevant information that is used to interpret MIR spectra is usually obtained from the functional group region, where as the fingerprint region is normally complex. It contains a lot of structural rather than functional group information, and many overlapped bands because each different compound produces a different and unique absorption pattern in this region [25].

Once NIR and MIR spectra contain large quantities of data that require processing, infrared methodologies are generally supported by chemometrics, defined as an inter-disciplinary field that uses methods such as multivariate statistics, mathematics and computer science for extracting information from chemical systems [26,27]. It can be based on unsupervised pattern recognition (principal component analysis - PCA, hierarchical cluster analysis - HCA), supervised classification (partial least squares-discriminant analysis - PLS-DA, soft independent modeling of class analog - SIMCA, linear discriminant analysis - LDA, canonical variate analysis - CVA, artificial neural network - ANN) and multivariate calibration (partial least squares - PLS, principal component regression - PCR) [22]. The chemometric approaches can enhance significantly the potential applications of infrared spectroscopy techniques [23].

Considering the importance of monitoring food quality this review presents some of the latest researches using infrared spectroscopy techniques for assessing intrinsic quality of dairy products.

Recent Applications of Infrared Spectroscopy for Dairy Products Quality Evaluation

Since the 1960s, many studies have been carried out using infrared spectroscopy applied to the food industry, mainly dairy products [2,28,29]. In 1961, Goulden performed a quantitative analysis of milk and other emulsions using infrared absorption [30]. FT-MIR and FT-NIR are spectral complementary techniques, so they have advantages and disadvantages. FT-MIR is the worldwide method of choice for composition and quality controls during routine liquid milk testing. It allows a fast, nondestructive quantification of milk chemical properties to avoid reference methods, which are usually tedious, expensive, and time consuming [31]. MIR is good for qualitative and quantitative identification and presents well-defined bands for organic functional groups [32], fat, protein, lactose that are part of the milk composition. Medium infrared devices generally use a minimum sample volume for milk analysis compared to NIR equipment. However, its main disadvantages are presence of the huge band of absorption of water, since milk consists of about ~87 % water [2]. The need for sample preparation (in the absence of ATR) is another disadvantage of FT-MIR [32] and for milk analysis there is still the experimental complexity. Expensive transmitting materials [32], the MID-infrared devices generally are much more expensive. In this respect, the NIR stands out. The NIR has advantages: simple and cheaper transmission instruments with glass optics and cheaper and less expensive light source compared to the MIR. For dairy analyzes, the samples are checked daily, therefore, for this purpose, the NIR is economically advantageous. Weak absorption due to water over to NES enables analysis of high moisture product. The NIR is not influenced by CO2 eliminating instrument purging and extremely high signal-to-noise in spectral data [32].
Many studies indicate that NIR can predict the chemical composition of milk and dairy products and can monitor the cutting-point during cheese manufacturing. Other studies have demonstrated the potential of NIR to predict sensory characteristics (for example, hardness and tenderness) of dairy products [33]. Liu et al., [31] reports different approaches to detect some potential biomarkers in milk. However, they furnish accurate results after a complex sample preparation and professional instrument operation. The advantages of NIR spectroscopy also include fast and a simultaneous, non-destructive measurement of a number of milk constituents and potential for online analysis [17]. The information of spectra relates to overtones and combinations of vibrations of some characteristic bonds, such as C-H, N-H, O-H and S-H, which typically exist in all organic molecules [31]. The disadvantage of NIR is that the spectra are less pronounced making the signal differences more subtle. According Quet et al., [23] NIR has limited capacity for estimating heterogeneous samples. Generally, it cannot identify components with concentration less than 1% [32]. Moreover, low precision and subjectivity of the reference methods are also constraint for its applications. Consequently, robust calibrations are needed with regard to better sampling procedures and improvement of reference methods. For example, relies on statistical methodology - PCA, PLS, SIMCA [32].

**Milk**

Dairy products such as milk, consumed worldwide, have high nutritional value. However, some suppliers often add adulterants such as starch, melamine, sodium, citrate, sucrose, urea, water, whey and other components into milk and thus get a higher profit [34,35]. The adulterated milk will not only cause the risk of illnesses among people, but also lead to considerable losses for the dairy industry, once the milk quality plays a very important role in the production of all types of dairy products, as cheeses for example, affecting both cheese yield and characteristics of the cheese [36].

Thus, from both the commercial and the health perspectives, milk adulterations have become a serious problem in the food industry and therefore techniques such as infrared spectroscopy can assist against this type of practice [34].

NIR infrared was successfully applied to discriminate raw cow milk from those adulterated with different pseudo proteins and thickeners [37] and melamine, which is a nitrogen-rich substance noxious to human health [38]. Also melamine addition was investigated in powdered milk products [39-41].

Mid-infrared spectroscopy is widely used in raw milk by processors for standardization purposes before milk transformation, by dairy industry for milk payment, and by farmers, technicians, and companies, which used several milk phenotypes [42]. Bassbasiet al., [43] have employed the technique to the fast and non-destructive quantitative determination of Solid Nonfat (SNF) content in raw milk. SNF is an important nutritional parameter of raw milk, once it can be determined with the appearance of several peaks in the fingerprint region (near 1000 cm⁻¹) and differentiating from the original milk spectrum. Regarding formaldehyde adulteration, a distinguishable peak around 1000 cm⁻¹ could be noted. In sucrose adulteration, the differentiation was found associated with the appearance of several peaks in the fingerprint region (near 1200 - 1000 cm⁻¹). At the same line of research, Gondimet al., [44] studied a sequential strategy to detect common adulterants in milk, including water, thickeners, preservatives and neutralizing agents. It was used the Soft Independent Modelling of Class Analogy (SIMCA) classification technique applied to Mid-Infrared (MIR) data, using the cross-validation method for the construction of the models.

Luiz et al., [2] analyzed the veterinary drugs tetracycline, enrofloxacin, penicillin, cefitiofur hydrochloride and diclofenac sodium by means of PCA-associated Fourier Transform Near-Infrared Spectroscopy (FT-NIR) to fast and accurately detect residues of these drugs in milk samples. With their results, they were able to discriminate different types of antimicrobials dissolved in milk within the maximum residue limits allowed by the European Medicines Agency and the Ministry of Agriculture, Livestock and Supply of Brazil. In addition, their methodology was able to detect real samples milked on different days after the injection of cefitiofur hydrochloride, which is, in principle, a zero antimicrobial grace period.

Recently Teixeira et al., [9] established spectroscopic patterns of antibiotics isolated and in the presence of milk, in order to contribute with another tool for the detection of minimum amounts of these compounds quickly and accurately. For that, they used vibrational spectroscopy and theoretical calculations based on Density Functional Theory in bovine milk.

**Whey and its Products**

Nowadays there is an increased demand for products with healthy and high nutritional properties; therefore, the dairy industry has recognized the value of whey proteins [45]. Whey constitutes about 85-90% of the volume of the milk used in the cheese production and it retains about 55% of the milk nutrients [46]. Even though it’s high concentration of good nutrients, whey was historically considered as a waste product and environmental pollutant, with no commercial use [47]. However, nowadays this by-product of cheese-making is getting new technical and nutritional applications [47-49]. The whey proteins can be incorporated in several food products to maintain the functional and nutritional value and also can be used in the sports nutrition market, as nutritional supplements [45,47,49]. In this area Whey Protein Concentrate (WPC), Whey Protein Isolate (WPI) and Whey Protein Hydrolysates (WPH) are the main examples. They contain 65-80% and above 90% of protein on a dry basis, respectively, being rich in essential amino acids, bioactive peptides and antioxidants [50,51]. However due the fact of being high added value products, WPC and WPI have been target for adulteration with cheaper components [52], as carbohydrates, amino acids derivatives and thermogenic substances [15,53].

Spectroscopy methodologies have been used in studies with whey and its products [13,45,54-58]. O'Loughlin et al., [55] investigated thermal denaturation of whey protein solutions, characterizing the changes occurring in whey protein dispersions upon heat treatment through physical analysis of solutions and intra- and intermolecular level analysis using infrared spectroscopy. According to the authors, the use of Infrared (IR) spectroscopy for discerning protein structure...
Several authors have attested the cheese quality using infrared spectroscopy, once usual methods are normally time consuming and expensive. Most of the works are in relation to cheese ripening, composition and authenticity or authentication.

In 2005, AOAC international has approved the Fourier Transform Infrared (FTIR) method for determination of fat, protein, lactose and total solid contents in some dairy products [63]. Since then, studies on cheese ripening using spectroscopic methods have been published [64-70]. A recent and important work in this field was carried out [14]. They evaluated the ripening changes over time of special cheeses (Pecorino, ewes' ripe, and Gouda) made with ewes' milk using FTIR/ATR spectroscopy during approximately one year. The study was able to detect the proteolysis and lipolysis, which are primary reactions that define cheese ripening. Overall it was observed a downward trend of the absorbance intensity of the amide group peaks (1700 to 1500 cm\(^{-1}\)), which are linked to the breakdown of peptide bonds. Similar behavior was observed for the lipidic region (3000 to 2800 cm\(^{-1}\) and 1765 to 1730 cm\(^{-1}\)). The proteolysis occurred in a fast pace during the first trimester of the ripening process, and the lipids were converted to smaller species as the time goes by. Hierarchical cluster analysis and principal component analysis allowed the evaluation of the physico-chemical changes of the cheeses.

Regarding cheese adulteration and authenticity it can be cited [10,18,71]. Hruzikova et al., [10] developed a method for authenticity control of cheese products, which is based on the measurement of infrared spectra of the gas phase obtained by heating selected cheese under controlled conditions. The method was tested for the authenticity control of two cheese families. Cuibuset al., [71] have done a study to compare some traditional Romanian cheeses and adulterated ones using ATR-FTIR coupled to chemometrics. It was carried out a Partial Least Squares Integrated Fourier Transform Infrared (FTIR-PLS) strategy to reveal the adulteration of Dalia cheeses with fats bellowing to Palm Oil. The results proved that FTIR-PLS is a reliable technique for discriminate the adulterated cheeses with palm oil.

A recent work with the application of MIR infrared spectroscopy for the detection of adulteration was performed with butter cheeses adulterated with soybean oil in levels of 0 to 100% of replacement [18]. The main bands obtained in the spectra were related to the presence of water, lipids and proteins, which belong to the "functional group" region (4000 to 1500 cm\(^{-1}\)) [4]. Overall, regarding the lipids related bands there was an increase in the absorbance values up to 70% of adulteration. However, the band near 3007 cm\(^{-1}\) which represents stretches of groups -C=CH (cis-) of double bonds in unsaturated fatty acids [72,73] was the only one in which there was a regular increase in the absorbance values because of the increase of soybean oil content in the cheeses, even in those with adulterations of 80%, 90% and 100%. Also, another aspect observed in this peak and which was used to characterize the adulterations was the band shift. Therefore it was found that the proportion and composition of unsaturated fatty acids (highly present in soybean oil, but in low concentrations in butter oil) affect the position and intensity of this band, proving fraud. Major mid-infrared bands associated with milk are presented in table 1.

Some important near-infrared bands associated with major components found in milk are summarized in table 2.
| Wavenumber (cm⁻¹) | Origin       | Assignment                                      | Reference |
|------------------|--------------|-------------------------------------------------|-----------|
| 900 to 1200      | Lactose      | C-O-C stretch                                   | [74]      |
| 1045             |              | C-O stretching/vibration                        | [29]      |
| 2800 to 3000     | Carbohydrates| C-H stretch                                     | [75]      |
| 800 to 1400      |              | Skeletal stretching and bending                  | [75]      |
| 1550             | Protein      | Peaks of C-N and N-N                            | [29]      |
| 1600 to 1695     | Casein       | Amide I (C=O stretch)                           | [76][77]  |
| 1520 to 1560     |              | Amide II (N-H bending and C-N stretch)          | [76][77]  |
| 1230 to 1300     |              | Amide III                                      | [76]      |
| 1060 to 1100     |              | O-P-O stretch                                   | [78][74]  |
| 1725 to 1745     | Fat          | C=O stretch                                     | [76][75]  |
| 970              |              | C=C-H bending                                   | [75]      |
| 2800 to 3000     |              | C-H stretch                                     | [75]      |
| 1520 to 1650     | WheyProtein  | Amide I and Amide II                            | [79][74]  |
| 1632             |              | Amide I (C=O stretch)                           | [59]      |
| 1531             |              | Amide II (N-H bending and C-N stretch)          | [59]      |
| 1200 to 1400     |              | C-H groups of aliphatic hydrocarbons            |           |
| Amide III        |              | [79]                                            |           |
| 1390             |              | C=O stretch in the COO-                         | [79]      |
| 1250             |              | Amide III (C-N)                                 | [59]      |
| 1750             | Lipids       | C=O                                             | [59]      |
| 1450             |              | C-H bonds in CH2 groups                         | [59]      |
| 1392             |              | C=O in carboxylic acid                          | [59]      |
| 1100 to 1130     |              | C-OH stretch, C=C and C-O-C                    | [59]      |
| 900 to 1200      |              | C-O-C and C-O vibration of carbohydrates        | [79][74]  |
| 1730 to 1765     | Triglycerides| C=O stretch                                     | [74][80]  |
| 2925             |              | Methyleneasymmetric stretching                  | [80]      |
| 2850             |              | Methyleneasymmetric stretching                  | [80]      |
| 1475             |              | Methyleneacussoring                             | [80]      |
| 1385             |              | Methyleneasymmetric formation                   | [80]      |
| 1400 to 1477     | Lipids       | C-H stretch                                     | [74]      |
| 3680             | Water        | O-H symmetric and antisymmetric stretching      | [81]      |
| 3200 to 3600     |              | O-H stretching                                  | [75]      |
| 1650             |              | H-OH stretching                                 | [75]      |
| 1020 to 1282     | Phospholipid |                                                | [82]      |
| 1055 to 1070     |              | P-O-C                                          | [83]      |
| 970              |              | Asymmetrical C-N stretch of (CH3)N             | [84]      |
| 970 to 1200      |              | P-O-C + PO2                                     | [85][84]  |
| 1025             |              | Symmetrical C-O-P stretch                      | [84]      |
| 1243             |              | Asymmetricalphosphatediester PO2              | [84]      |
| 1090             |              | Symmetrical phosphate diester stretch of PO2   | [84]      |
| 1173             |              | Asymmetrical C-O stretch of C=O-O-C            | [84]      |
| 400 to 1000      | Melatonin    |                                                | [86]      |
| 3200 to 3400     |              |                                                | [86]      |
| 1680             |              |                                                | [87]      |
| 1400 to 1500     |              |                                                | [87]      |
| 800              |              |                                                | [87]      |
| 900              |              |                                                | [87]      |
| 980              |              |                                                | [87]      |

Table 1: Wavenumbers of mid Infrared bands and their assignments.
Final Remarks and Future Outlook

The detection of food quality and authenticity requires comprehensive monitoring of food with efficient analytical methods. In this review, recent advances in food safety evaluation by infrared spectroscopy techniques are presented and discussed. It could be confirmed that NIR and MIR spectroscopy have proven to be worthwhile analytical methods, once have the potential for assessing food safety properties, detecting adulterants, as well as performing characterization and classification of dairy products. Besides the potentiality of the infrared spectroscopy is increased by its use associated with chemometrics. Their combination makes the MIR and NIR techniques more convenient and effective for the analysis of a broad variety of products.

Even though a bunch of works have been carried out in the recent years, the increasing requirement for food quality and authenticity analyses, as well as ongoing work on the development of spectroscopic methods, it will bring several research opportunities for coming years.

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