Molecular Epidemiology of Hepatitis C Virus and Predominant Genotype in India

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Abstract: Analysis of the HCV genome has demonstrated extremely high heterogeneity in both structural and nonstructural coding regions and there are at least six different genotypes that have generally been divided into several subtypes. Of the 6 different Hepatitis genotypes, genotypes 1-3 is common worldwide, type 1a and 1b are the most common, accounting for about 60% of global infections. They predominate in Northern Europe, Southern and Eastern Europe, North America, and Japan respectively. Type 2 is less frequently represented than type 1. Type 3 is endemic in south-east Asia and is variably distributed in different countries. The determination of the infecting genotype is important for the prediction of response to antiviral treatment; genotype 1 is generally associated with a poor response to interferon alone, unlike genotypes 2 and 3 which are associated with better responses. A total of 238 plasma samples were received from patients attending gastroenterology department across India for treatment from March 2008 – Aug 2010. The samples were analyzed for viral load by real time PCR by Taqman principle. HCV genotyping was carried out on the samples whose viral load was more than 300IU/ml (limit of detection as per the kit). Qiagen RNA columns were used for RNA extraction, followed by reverse transcription (Promega) and genotyping was performed by conventional PCR. Out of 238 samples, 117 were positive for 3a, 44 samples were load negative, 43 samples were non-typable due to less viral load i.e. less than 1000 IU/ml. 26 were type 1a, 107 were 3a, and 11 were 2a.

Keywords: HCV, PCR, Reverse Transcription, Genotyping

1. Introduction

The genome of HCV is highly mutable. HCV being an RNA virus, lacks efficient proofreading ability as it replicates, virions infecting humans undergo evolution with time, giving rise to the notion that HCV persists as a collection of virus quasispecies (1).

The mutation occurs in hyper variable region of the genome coding for the envelope proteins and escapes immunity by the host and at the same time knocks host’s innate immunity resulting in HCV chronic infection. Analysis of the HCV genome has demonstrated extremely high heterogeneity in both structural and nonstructural coding regions and has identified at least six different genotypes that have generally been divided into several subtypes. The development of vaccine for multiple serotypes is challenging for global use. The different genotypes show varied response to interferon/ribavirin combination therapy (2).

Hepatitis C virus (HCV) remains the major causative agent for parenterally transmitted non-A, non-B hepatitis. The detection of HCV RNA provides evidence of early and active infection. HCV qualitative and quantitative molecular tests and HCV genotyping tests have proved useful in making management decisions (3).

The development of vaccine for multiple serotypes is challenging for global use. The different genotypes show varied response to interferon/ribavirin combination therapy. Hepatitis C virus (HCV) remains the major causative agent for parenterally transmitted non-A, non-B hepatitis. The detection of HCV RNA provides evidence of early and active HCV infection. Improvements in therapy have
resulted in better virologic response rates, and HCV qualitative and quantitative molecular tests and HCV genotyping tests have proved useful in making management decisions (4).

HCV genotype is the strongest foretelling factor for sustained virological response since patients with different HCV genotypes act in response differently to alpha interferon therapy. Solid evidence has been established that HCV genotype-2 and genotype-3 infected patients are more likely to have a sustained virological response (SVR) to anti-viral therapy than patients infected with genotype-1 HCV infections (5).

The global prevalence of HCV infection is (approximately 3%) 170 million people. In the United States, approximately 3.9 million people (1.8% of the population) are positive for HCV antibodies.

The primary mode of HCV transmission is exposure to infected human blood via intravenous drug use or unscreened transfusions. The practice of screening donors for HCV antibodies in developed countries since 1990 has substantially lowered the risk of acquiring HCV infection from a transfusion to approximately 1 in 263,000. This is based on results from nucleic amplification testing (NAT) of pooled donor samples that has been performed in the USA.

On the basis of its extensive genetic heterogeneity, HCV has been divided into six major genotypes (represented by Arabic numerals) and at least 80 subtypes (represented by lowercase letters). Different genotypes share approximately 65% sequence homology. Genotypes 1, 2, and 3 are found throughout the world; but the other genotypes are common in particular geographic regions (genotype 4 is common in North Africa and the Middle East, genotype 5 is common in South Africa, and genotype 6 is common in Southeast Asia). The predominant genotype in patients with chronic HCV infection in the United States is genotype 1 (72% of patients), followed by genotype 2 (16%) and genotype 3 (10%) (6).

As per Gower et al., Genotype occurrence was available for ninety-eight countries. Globally, genotype 1 was the most common (46%), followed by G3 (22%), G2 (13%), and G4 (13%), (13)

The prevalence and number of HCV sero-positives have increased from 2.3% to 2.8% which is 122 million to >185 million between 1990 and 2005. Central and East Asia and North Africa/Middle East are estimated to have high prevalence. South and Southeast Asia, sub-Saharan Africa, Andean, Central, and Southern Latin America, Caribbean, Oceania, Australasia, and Central, Eastern, and Western Europe have moderate prevalence. Asia Pacific, Tropical Latin America, and North America have low prevalence (<1.5%). (14)

HCV genotypes display significant differences in their global distribution and prevalence, making genotyping a useful method for determining the source of HCV transmission in an infected localized population. Quick and accurate genotyping of hepatitis C virus (HCV) is becoming increasingly important for clinical management of chronic infection and as an epidemiological marker (7).

The prevalence and distribution of HCV genotypes depend on geographical location. Three broad patterns of genotype distribution have been identified to date. One pattern, characterized by high genetic diversity, involves geographically discrete areas of West Africa with types 1 and 2, Central Africa with type 4 and Asia with types 3 and 6. This pattern is suggestive of a long period of endemic infection. Another pattern involves areas with a few subtypes circulating in specific risk groups, e.g., subtype 3a in drug addicts. The third pattern involves areas where a single subtype is present, such as in Egypt with subtype 4a and South Africa with subtype 5a (3).

Hepatitis C virus is one of the most important causes of chronic liver disease in the United States. It accounts for about 20% of acute viral hepatitis cases; 60% to 70% of chronic hepatitis cases; and 30% of cirrhosis, end-stage liver disease, and liver cancer cases. Seventy-five percent of HCV-infected individuals become persistently infected and many develop chronic hepatitis, which progresses eventually to liver cirrhosis and hepatocellular carcinoma (8).

Americans and Asians with HCV have a 2-fold and 4-fold increased risk, respectively, of developing HCC when compared to Caucasians with HCV. The rate of chronic HCV infection is lower in patients who present with clinical symptoms such as jaundice during the acute onset of HCV infection as compared to those who do not have such symptoms. There have been extensive studies focusing on the natural course of disease progression from chronic hepatitis C to cirrhosis, hepatocellular carcinoma (HCC) and death (8).

2. Materials and Methods

A total of 238 plasma samples were analyzed from patients who had HCV infection referred to Manipal acunova central laboratory by Gastroenterologists across India from Jan 2007 to October 2010 from different states in India. Out of 238 patients 54 were females in the age group ranging from 5 to 70 years with a mean age of 43.90 and std of 15.94 and 184 were males in the age group ranging from 5 to 70 years with a mean age of 46.78 and std of 14.34. 10 patients were from Karnataka accounting to 4.2%, followed by Andhra Pradesh 30 (12.6%), New Delhi 156 (65.5%) Punjab 26(10.9) West Bengal 6 (2.5%) Chatisgarh 9 (3.8%) and Tamil Nadu 1(0.4%). Diagnosis of HCV was made by detection anti HCV antibodies by ELISA method and rapid method.

The EDTA blood samples were shipped at −20°C, DNA extraction was done using Qiagen DNA columns. HCV quantification was performed prior to genotyping using Taqman assay, reagents were from Qiagen. Samples whose viral load was less than 300 IU/ ml were not included for genotyping. The procedure was followed as per TOMOYOSHI OHNO et al. 1997(12). Reverse transcription kits were from Promega and the primers were from Sigma.
Out of 238 samples, 16 samples were negative for viral load i.e. less than the detection limit of the kit. Genotyping required for successful genotyping is 1000 IU/ml, kit detection limit being 300IU/ml. 

Of the total 238, (156, 65.5%) were from New Delhi followed by (30, 12.6%) from Andhra Pradesh, (26, 10.9%) from Punjab, (10, 4.2%) from Karnataka, (9, 3.8%) from Chhattisgarh, (6, 2.5%) from West Bengal and (1, 0.4%) from Tamil Nadu.

Out of 238 samples 16 samples were negative for viral load, i.e. less than the detection limit of the kit. Genotyping was negative in 17 samples as the minimum viral load required for successful genotyping is 1000 IU/ml, kit detection limit being 300IU/ml.

Out of 206 in which genotyping was successful, a total of 145 were positive for 3a of which 104 males, 3a, and 36 females were positive for 3a, followed by type 1a (18), 3b(17), 2a & 1a. Genotype 3a was uniformly distributed in all the states with a p value of 0.0001. 3a was the predominant genotype in both males and females which confirms there is no gender preference.

**Table 1. HCV genotype distribution as determined by genotyping across India.**

| State  | 1a | 1b | 2a | 3a | 3b | 3a&3b | 1a&2a | 4 | 6 | NT | N | Total |
|--------|----|----|----|----|----|-------|-------|---|---|----|---|--------|
| ND     | 12 | 5  | 2  | 98 | 8  | 1     | 5     | 2 | 1 | 12 | 10 | 156    |
| AP     | 3  | 1  | 0  | 14 | 6  | 0     | 0     | 0 | 0 | 3  | 3  | 30     |
| PJ     | 2  | 1  | 0  | 13 | 2  | 2     | 1     | 0 | 0 | 2  | 3  | 26     |
| KA     | 1  | 0  | 0  | 8  | 0  | 0     | 1     | 0 | 0 | 0  | 0  | 10     |
| CHAT   | 0  | 0  | 0  | 8  | 0  | 0     | 1     | 0 | 0 | 0  | 0  | 9      |
| WB     | 0  | 0  | 0  | 4  | 1  | 0     | 1     | 0 | 0 | 0  | 0  | 6      |
| TN     | 0  | 1  | 0  | 0  | 0  | 0     | 0     | 0 | 0 | 0  | 0  | 0      |
| Total  | 18 | 8  | 2  | 145| 17 | 3     | 9     | 2 | 1 | 17 | 16 | 238    |

**4. Discussion**

The hypothesis of an Asian origin of HCV-3a relies on the large number of subtypes of HCV genotype 3 isolated in this area, suggesting several centuries of endemic presence of genotype 3 in Asian population (10). Three HCV genotypes such as HCV-1, HCV-2, and HCV-3 have worldwide distribution and their relative prevalence varies from one geographic area to another. HCV-1a and 1b subtypes are the most prevailing genotypes circulating in the United States of America and Europe. In Japan the most common circulating HCV subtype is 1b. HCV-2a and 2b subtypes are mostly common in North America, Europe, and Japan. Subtype 2c is found commonly in northern Italy. HCV-4 is the most prevalent genotype circulating in North Africa and the Middle East. HCV-5 and HCV-6 genotypes are established only in South Africa and Hong Kong, respectively (10).

The region used by Okamoto et al. for their type 3a primer might not be a suitable region for the design of primers if all the common subtypes were to be detected. As their system is based on the nucleotide sequences of genotype 1a, 1b, 1d, 2a, 2b, 3a, 3b, 4, 5a, and 6a HCV isolates, they believed that this system may have a much broader application. However, the number of samples of HCV types 3 to 6 tested was still not large enough for definitive conclusions to be drawn, which should be further tested in areas in which HCV types 3 to 6 are common to further validate this genotyping method. If the accuracy of our system is confirmed in these areas, this is a convenient and cost-effective method and will assist research workers in conducting large-scale epidemiological studies.

In developed countries, in which donated blood is routinely screened for HCV, infection control and safe-injection practices are in place, as well as in developing countries, injection drug use is an important risk factor for HCV transmission. Injection drug users have highest HCV sero -positivity rate in China. More than 80% of IDUs are sero-positive in Mexico, Pakistan, and Thailand. Egypt has the highest rate of hepatitis C in the world (10%), (15).

The high prevalence of global HCV infection necessitates continuous efforts prevent primary infection, vaccine development, new approaches to secondary and tertiary prevention to reduce the burden of chronic liver disease and to improve survival for those who already have evidence of liver disease (14).

As per the KPK study, HCV genotypes 1a, 1b, 3a and 3b...
are distributed in various parts of KPK among which the genotype 3a is the most frequent genotype, our findings are similar to the above study(5).

5. Conclusion

We conclude that HCV genotypes 1a, 1b, 3a and 3b are distributed in various parts of India, 3a being the most frequent genotype circulating in India.

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