Analysis of KIT gene mutations in patients with melanoma of the head and neck mucosa: a retrospective clinical report

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INTRODUCTION

The development of melanoma is a classic example of a neoplasm that progresses through different known stages. However, the key molecular event that triggers the progression of this neoplasm has still not been clarified, which explains why there is no specific therapy and why there have been so few advances in the multimodal therapy of this disease [1].

Somatic mutations in the KIT gene, which were identified in a small number of skin melanomas, appear to have a higher incidence in mucosal melanomas. Curtin et al. found mutations or an increase in the number of copies of the KIT in 39% of mucosal melanomas. Moreover, there was an increase in KIT protein expression in mucosal melanomas, which supports their role in the progression of this melanoma subtype [2]. Given the evidence of a possible pathogenic role of the KIT gene in a number of mucosal melanomas, including those of the head and neck, screening for KIT aberrations may have diagnostic value, and the gene may represent a therapeutic target in these patients [3].

The identification of activating mutations in the KIT gene in patients with mucosal melanoma is important to improve knowledge of tumor biology and the design of clinical research protocols with imatinib. Because mucosal melanoma of the head and neck is a rare condition, the frequency of KIT mutations has not been characterized in these tumors. The objective of the present study was to evaluate the frequency of KIT mutations and their prognostic value in a significant number of head and neck mucosal melanomas [4].
RESULTS

Twenty-eight cases were included in the study. Summary information on each case is given in Table 1. Patient age ranged from 27 to 88 years, with a median of 59.5 years; 16 patients (57.1%) were women; 24 patients (85.7%) were white, and 4 patients were black. The majority of the tumors were classified as T4 (75%) and the majority of the cases were N0 or stage IV patients. Seven patients (25%) had undifferentiated primary tumors, 17 patients (60.5%) had tumors with a mitotic index higher than 10 mitoses/mm², and only 8 patients (28.6%) exhibited amelanotic tumors.

Analysis of KIT gene mutations

Analysis of KIT gene mutations was possible in all of the 28 cases studied; 7 patients had KIT mutations and 21 patients had wild-type KIT. KIT mutations were most frequently detected in exon 11 (42.8%) and exon 9 (42.9%). Table 2 describes all the mutations found. Figure 1 shows a chromatogram of a mutation in exon 11 of the KIT gene, and loss of heterozygosity is represented in Figure 2. There was no significant relationship between the clinical and demographic variables and the presence of the KIT mutation ($p > 0.05$; Table 3).

The DFS (disease-free-survival) rate for the cancer was 53.6% in 24 months and 37.5% in 60 months. The absence of adjuvant treatment and a mitotic index higher than 10 mitoses/mm² were associated with a higher probability of death ($p = 0.05$, log-rank test). The presence of vascular invasion and angiolymphatic dissemination were also statistically significant ($p = 0.04$ and $p = 0.02$, respectively, log-rank test). In addition, there was a statistically significant relationship between recurrence and a mitotic index higher than 10 mitoses/mm², vascular invasion, angiolymphatic dissemination and perineural dissemination ($p = 0.05$, $p = 0.043$, $p = 0.008$ and $p = 0.034$, respectively, log-rank test). There were no statistical differences between the groups, and the presence of mutation did not play a role, either in protecting or promoting relapse or death.

DISCUSSION

Because of the rarity of this disease, only 28 cases were enrolled in the study. Kanda (2003) presented a study with a total of 54 cases from 3 different institutions, which demonstrates the rarity of the condition in the national context [5]. Therefore, much of the literature that exists on the subject addresses isolated cases and consists of retrospective analyses of series with relatively small sample sizes [6, 7].

Some peculiarities of the biological behavior of these tumors are extremely important: the anatomical location of the lesion, including its staging. Thus, some early-stage tumors may exhibit an aggressive behavior due to their location.

The historical results of HNMM treatment are disappointing. This soon led researchers to test new treatment strategies, such as the addition of chemotherapy or targeted therapy.

Defining prognostic parameters for HNMM is a much more complex task because the depth of invasion—the most important prognostic factor in skin melanomas—cannot be used due to the lack of histological points of reference similar to the papillary and reticular dermis.

Recent studies have evaluated the oncogenic role of KIT mutations in HNMM, as well as the benefits of therapy with tyrosine kinase inhibitors in these tumors. The results appear to be encouraging, showing significant benefits in survival time over chemotherapy and targeted therapy [8].

HNMM most frequently affects patients between the fifth and seventh decades of life, with more than 60% of patients belonging to this age group [9]. In this study, 16 (57%) patients were aged over 60. When we compared these 2 age groups, i.e., patients aged over 60 and those aged under 60, with regard to disease recurrence and mortality, there was no significant difference ($p = 0.38$ and $p = 0.648$).

According to some authors there are no differences in the incidence of the location of HNMM, i.e., the incidence of tumors of the sinuses of the face and nose is equivalent to that of tumors of the oral cavity [9, 10]. However, we found a higher incidence of tumors of the sinuses of the face (75%) in our sample. This is due to factors related to the treatment; considering that in INCa more complex tumors are treated, we believe this is due to the sample selected.

Retrospective series did not demonstrate a relationship between the primary site of the tumor and both survival and local control [11, 12]. This observation was confirmed in the present study. However, there are some reports of a poorer prognosis related to tumors originating in the sinuses of the face, probably because of the presence of locally advanced disease at diagnosis [13].

We used the TNM classification in the present study. The majority of tumors were classified as T4 (75%) and the majority of patients had N0 neck and were considered stage IV. We did not find any differences in OS and DFS ($p = 0.899$ and $p = 0.523$, respectively). In the present study, we observed a mitotic index higher than 10 mitoses per field in 60.7% of the patients. This is an important result as it suggests a greater aggressiveness in these tumors. It is interesting to highlight that after analysis of DFS and OS, the mitotic index was found to be an independent prognostic factor for both end-points. OS in patients with a mitotic index higher than 10 mitoses per field was 36.4% in 48 months versus 51.5% in the other group ($p = 0.05$). Thompson et al. (2003) studied 115 patients with HNMM and obtained similar results, i.e., a poorer prognosis in...
Table 1: Summary of cases included in the study

| Case | Gender | Age | Ethnicity | Treatment | Relapse | SG (months) | KIT |
|------|--------|-----|-----------|-----------|---------|-------------|-----|
| 1    | Female | 69  | White     | CX + RXT  | Yes     | 117         | Wild |
| 2    | Female | 66  | White     | CX + RXT  | Yes     | 54          | Wild |
| 3    | Female | 38  | White     | CX        | Yes     | 11          | Wild |
| 4    | Female | 88  | White     | CX + RXT  | Yes     | Still Alive | Wild |
| 5    | Female | 64  | White     | CX + RXT  | No      | Still Alive | Mutated |
| 6    | Female | 61  | White     | CX + RXT  | No      | Still Alive | Wild |
| 7    | Female | 62  | White     | CX + RXT  | Yes     | 27          | Wild |
| 8    | Female | 62  | White     | CX        | Yes     | 29          | Wild |
| 9    | Female | 39  | White     | CX + RXT  | Yes     | Still Alive | Wild |
| 10   | Female | 62  | White     | CX        | Yes     | 10          | Wild |
| 11   | Female | 27  | White     | CX        | Yes     | 8           | Wild |
| 12   | Female | 59  | White     | CX        | Yes     | Still Alive | Wild |
| 13   | Female | 60  | White     | CX        | Yes     | 11          | Wild |
| 14   | Female | 76  | White     | CX        | Yes     | 14          | Wild |
| 15   | Female | 63  | White     | CX        | Yes     | 19          | Wild |
| 16   | Female | 69  | White     | CX        | Yes     | 43          | Wild |
| 17   | Male   | 32  | White     | CX + RXT  | Yes     | 99          | Mutated |
| 18   | Male   | 85  | White     | CX + RXT  | Yes     | 60          | Mutated |
| 19   | Male   | 47  | White     | CX        | Yes     | Still Alive | Wild |
| 20   | Male   | 39  | White     | CX + RXT  | Yes     | 27          | Wild |
| 21   | Male   | 53  | White     | CX + RXT  | Yes     | 15          | Mutated |
| 22   | Male   | 52  | White     | CX + RXT  | Yes     | 21          | Mutated |
| 23   | Male   | 62  | White     | CX + RXT  | Yes     | 64          | Wild |
| 24   | Male   | 51  | White     | CX + RXT  | Yes     | Still Alive | Wild |
| 25   | Male   | 40  | White     | CX + RXT  | Yes     | 11          | Mutated |
| 26   | Male   | 81  | Black     | CX        | Yes     | 30          | Mutated |
| 27   | Male   | 69  | Black     | CX + RXT  | Yes     | 21          | Mutated |
| 28   | Male   | 76  | Black     | CX + RXT  | Yes     | 11          | Mutated |

CX + RXT = Surgical resection followed by adjuvante radiotherapy, CX = Exclusive surgical resection, OS = Overall survival.

Table 2: Distribution of the KIT mutations found

| Patient | Mutational status | Type of mutation | Note |
|---------|-------------------|------------------|------|
| 5       | Exon 11           | V551I            | Heterozygous |
| 21      | Exon 11           | V551I            | Heterozygous |
| 22      | Exon 13           | L657F            | Homozygous  |
| 25      | Exon 9            | L455M            | Heterozygous |
| 26      | Exon 11           | L576P            | Heterozygous |
| 27      | Exon 9            | S480F            | Heterozygous |
| 28      | Exon 9            | G499S            | Heterozygous |
the group with a high mitotic index \((p = 0.026)\) [14]. In a recent work published by Moreno et al. (2010), 95% of patients with a high mitotic index died from the HNMM, whereas patients with a mitotic index of less than 10 had a better response to systemic therapy [15]. In our sample, we found 8 cases (28.6%) of amelanotic tumors, all confirmed by immunohistochemistry. Data from the literature suggest greater aggressiveness in amelanotic tumors, this being the only histopathological factor related to survival; no patient was alive after 48 months, compared with 47.6% survival in patients with pigmented tumors [15]. Our results did not confirm these data, but we had an OS of 55% in 48 months in patients with pigmented tumors, compared with 18.8% in those with amelanotic tumors. The explanation accepted for a poorer prognosis in amelanotic tumors is that they are associated with a delayed diagnosis.

In the sample, it was not possible to compare surgical treatment with other therapy modalities, because all the patients were submitted to resection. We obtained free margins in 82.15% of the patients; however, the finding of free margins had no impact on survival. Patients with positive surgical margins were 21 times more likely to die due to therapeutic failure, but we did not find this relation in the sample [16].

In local recurrent disease, and in the absence of metastatic dissemination, surgical removal should be considered, but the extent of the resection should be carefully planned. The chance of a successful surgical removal is less than 25%, and the chances of distant metastases are increased [17].

The main motivation for performing this study was to investigate a marker with therapeutic potential for a

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**Figure 1:** Chromatogram showing mutation in exon 11 of the gene KIT/L576P of a representative case (case 26).

**Figure 2:** Electropherogram showing loss of heterozygosity of the KIT gene (HK 8810 Marker) (analysis using an Agilent 2100 bioanalyzer).
disease that, although rare, is highly lethal. Moreover, there are few studies involving the research of KIT in HNMM, possibly as a result of the rarity of the disease.

The presence of KIT mutations in malignant melanomas has already been demonstrated in the literature, in particular in melanomas of the mucosa (21% of KIT mutations), acral melanomas (11% of KIT mutations), and skin melanomas caused by chronic exposure to sunlight (17% of KIT mutations) [2]. This suggests that KIT mutations have a role in the physiopathogeny and, therefore, are a potential therapeutic target in these subtypes of melanoma. In contrast, KIT mutations are rarely found in the largest subgroup of malignant melanomas, SM not associated with chronic exposure to sunlight: 1 in 100 cases studied by Willmore-Payne et al. (2005) [18]. Furthermore, phase II studies with imatinib in patients with SM, without KIT mutation analysis, were disappointing [19–21].

KIT mutations were evaluated in a selected group of patients with HNMM. Molecular analysis of KIT mutations was possible in 28 patients and showed mutations in 7 of them (25%). This is consistent with the discoveries of Antonescu et al. (2007), and Rivera et al. (2008), who detected mutations in the KIT gene in 3 out of 20 patients (15%) and in 4 out of 18 patients (22%) with mucosal melanomas of the anal region and of the oral cavity, respectively [22–24]. Thus, KIT mutations occur in up to 20% of mucosal melanomas, regardless of the location of the primary tumor. Three mutations were detected in exon 11, 3 mutations in exon 9, and 1 mutation in exon 13; this distribution with a higher frequency of mutations in exons 11 and 13 is in line with the literature [25, 26]. Table 4 shows the prevalence of the mutation in various studies.

The role of the location of the KIT mutation in HNMM has been little studied. However, its prognostic value in GIST tumors has been shown in the literature: exon 11, which codifies the juxtamembrane domain, is involved in autoinhibition of the receptor; the mutations in this juxtamembrane domain impede this inhibitory function, increasing the dimerization of the receptor, independently of its activation. This location, in turn, is the most sensitive (responds better) to treatment with tyrosine kinase inhibitors. This indicates that HNMM is potentially sensitive to tyrosine kinase inhibitors [31].

In our results, there was not one mutation more frequent than the others: 2 cases V551I, 1 case L657F, 1 case L455M, 1 case L576P, 1 case S480F, and 1 case G499S. Although the available studies comprise a small number of patients and several studies have demonstrated the wide variety of mutations (Table 5). We attempted to

| Variables     | Wild N = 21 (75%) | Mutation N = 7 (25%) | p’   |
|---------------|-------------------|----------------------|------|
| Age           |                   |                      |      |
| >60           | 12 (42.9%)        | 4 (14.3%)            | 0.666|
| <60           | 9 (32.1%)         | 3 (10.7%)            |      |
| Sex           |                   |                      |      |
| Female        | 12 (42.9%)        | 9 (32.1)             | 0.672|
| Male          | 4 (14.3%)         | 3 (10.7%)            |      |
| Race          |                   |                      |      |
| Whites        | 18 (64.3%)        | 6 (21.4%)            | 0.747|
| Blacks        | 3 (10.7%)         | 1 (3.6%)             |      |
| Smoking       |                   |                      |      |
| Yes           | 9 (32.1%)         | 4 (14.3%)            | 0.412|
| No            | 12 (42.9%)        | 3 (10.7%)            |      |
| Alcohol use   |                   |                      |      |
| Yes           | 5 (17.9%)         | 2 (7.1%)             | 0.581|
| No            | 16 (57.1%)        | 5 (17.9%)            |      |
| Location      |                   |                      |      |
| Nasosinusal   | 4 (14.3%)         | 4 (14.3%)            | 0.220|
| Oral cavity   | 17 (60.7%)        | 3 (10.7%)            |      |

* Fischer’s exact test.
evaluate clinical-pathological characteristics and link them to KIT mutations. The results were discouraging, as none of the characteristics analyzed presented higher prevalence of mutation, despite the number of mutations identified being highly significant (8/28 patients, 25%).

An interesting result was the distribution of mutations in terms of anatomical sites. In nasosinusual tumors, mutations were found in 4 cases (19% of patients), whereas in the group of patients with mouth tumors mutations were found in 3 patients (75% of patients).

Our results suggest a higher incidence of mutations in melanomas of the oral cavity. This finding is not consistent with the literature data, in which mutations are more common in tumors of the sinuses [30]. No prognostic impact caused by the presence of the KIT mutation was found in the present study. We evaluated the relationship of the mutation with OS and DFS. DFS was 28.6% in 48 months in both groups, i.e., in patients with the mutation and in those who presented the wild-type KIT ($p = 0.771$). SG was higher in patients with the wild-type gene, with a 5-year OS of 47.6%, but without statistical significance ($p = 0.935$).

KIT mutations have been evaluated in various other tumors, and in some neoplasms they are an important prognostic factor. In gastrointestinal stromal tumors (GIST), the KIT mutation is an independent risk factor for OS and DFS; this relationship is so important that exon 11 is related to a better therapeutic response. GIST tumors are an example of the importance of KIT in the selection for treatment and prognosis. In patients with leukemia, KIT mutations can suggest a greater risk of relapse [35].

However, the prognostic importance of KIT mutations in melanoma was not evaluated in a series of adequate size. Our study confirmed the presence of 25% of mutation in HNMM, thus suggesting a route to study the pathophysiology of this tumor, with a focus on the MAPKs cascade and the inclusion of patients in clinical trials with KIT inhibitors.

In conclusion, the continual evaluation of KIT mutations is essential, in an attempt to identify biomarkers and improve the selection of patients for targeted therapy.

### MATERIALS AND METHODS

For the formation of the group of this study, patients with head and neck mucosal melanomas were selected, enrolled, and treated at the Head and Neck Surgery Department of the National Cancer Institute (Instituto Nacional de Cáncer - INCa). Cases without paraffin blocks of tumor tissue or cases with incomplete information on the treatment used were excluded. Molecular analyses of KIT mutations and of the pattern of response to the treatment used (surgical resection, radiotherapy, or a combination of the two) were considered for study in addition to clinical and pathology data. The study was approved by the Research Ethics Committee, under number 272/11.

### DNA extraction method

DNA was extracted from tissue and embedded and fixed in paraffin using the MagNA Pure LC 2.0 semi-automated system (Roche Applied Science), according to the manufacturer’s instructions. The procedure consisted of deparaffination with xylene, followed by graduated addition of ethane to eliminate any xylene residues.

Deparaffination with xylene was followed by digestion by proteinase K (BioAmerica, Inc, Homestead, FL, USA) using the Wizard® Genomi DNA purification kit (Promega, Madison, WI, USA). The extraction was confirmed by running a 5 µL aliquot in 0.8% agarose gel in electrophoresis buffer (TBE 1X). To evaluate the integrity and concentration of the extracted DNA, a 1 µL aliquot was analyzed in a ND-1000 spectrophotometer (NanoDrop Technologies Inc., Delaware, USA).

### KIT mutation analysis

Mutational analysis of the gene was performed by Polymerase chain reaction (PCR) amplification and genome sequencing. The analysis began with exon 11 in the KIT gene, in which the majority of mutations are found (70% of the described mutations), followed by exons 9, 17,

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| Author               | Year | Number of Patients | KIT Mutation |
|----------------------|------|--------------------|--------------|
| Beadling et al. [27].| 2008 | 29                 | 8.4%         |
| Carvajal et al. [28] | 2011 | 5                  | 40%          |
| Schoenewolf et al. [29].| 2012 | 12                 | 0%           |
| Turri-Zanino et al. [7].| 2013 | 32                 | 12.5%        |
| Zebary et al. [30].  | 2013 | 56                 | 3.6%         |
| Present study.       | 2015 | 28                 | 25%          |
| **Total**            |      | 162                | 9.8%         |
and 13. It should be emphasized that the mutations are exclusive.

Exons 9, 11, 13, and 17 of the KIT gene were amplified using the primer pairs described in Table 6. The purified PCR products were prepared for genome sequencing, using a final volume of 7.5 µl (solution containing primer, DNA, and milli-Q water). Sequencing was performed using the Sanger method, and the software used in the analysis of the results was the automated sequencer ABI PRISMTM 377 (Applied Biosystems Foster City, California, USA), which interprets the results by titrating each base in relation to the intensity of the fluorescent compound, denominating 4 specific colors. The software used in the analysis of sequencing uses the same 4 specific colors for each base. By convention, A (adenine) was labeled as green, C (cytosine) as blue, T (Thymine) as red, and G (guanine) as black, in the electropherogram image.

### Statistical analysis

The statistical analyses were performed using the SPSS software, version 18.0 (SPSS Inc., Chicago, IL, USA). Overall survival (OS) (defined as the time between the start date of the treatment and the date of death) and disease-free survival (DFS) (defined as the time between treatment and recurrence) were estimated by the Kaplan–Meier method, and the survival curves were compared using the log-rank test. Univariate analysis was performed to evaluate the association between the variables age, sex, primary tumor site, mutational status of the KIT gene, and DFS. A $p$ value $\leq 0.05$ was considered statistically significant.

### CONFLICTS OF INTEREST

None.
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