SHORT COMMUNICATION

Dose efficacy study of two schedules of high-dose bolus administration of interleukin 2 and interferon alpha in metastatic melanoma

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Summary Forty-three patients with metastatic melanoma were treated with a 5 day (18 patients) and a 3 day (25 patients) schedule of high-dose IL-2 11.7 MIU m2 and IFN-α 3 MIU m2 i.v. by bolus administration every 8 h, repeated every 21 days for a total of three courses. The 5 day schedule resulted in a high response rate of 41% (CI 18–67%), but was accompanied by severe cardiotoxicity (41%) and central nervous system toxicity (28%). The 3 day schedule was associated with manageable toxicity, but yielded a moderate response rate of 20% (CI 7–45%).

Keywords: interleukin 2; interferon alpha; metastatic melanoma

The biological agents, interferon alpha (IFN-α) and interleukin 2 (IL-2) have shown evidence of activity against metastatic melanoma. IFN-α produced objective tumor regression in approximately 15% of patients (Creagan et al., 1990; Kirkwood, 1991). IL-2, with or without lymphokine-activated killer cells, yielded response rates from 10–25% (Rosenberg et al., 1989a; Parkinson et al., 1990; Whitehead et al., 1991). With the combination of IL-2 and IFN-α the reported response rates varied between 0–44% (Rosenberg et al., 1989b; Dillman et al., 1993; Sparano et al., 1993; Keilholz et al., 1993; Kruit et al., 1995). A phase I–II study with increasing dose levels of bolus IL-2 and IFN-α produced an objective response in 33% of patients (Rosenberg et al., 1989b). The highest response rate of 44% was reached with a schedule of IL-2 11.7 MIU m2 and IFN-α 3 MIU m2 three times a day by intravenous bolus administration, 5 days per cycle. We performed a phase II study with this schedule in order to confirm these results.

Patients and methods

Patient population

The study was divided into two parts. Owing to the occurrence of severe toxicity the 5-day regimen was replaced by a 3-day schedule. All patients had histologically confirmed metastatic melanoma. Eligibility criteria included: Karnofsky performance status of at least 80, age between 18 and 70 years, no prior immunotherapy with IL-2 or IFN-α, no metastases in the central nervous system, no cardiovascular history, normal pulmonary function, serum creatinine ≤ 1.25 times the upper limit of normal, or creatinine clearance ≥ 50 ml min⁻¹, leucocyte count ≥ 4.0 x 10⁹ l⁻¹, platelet count ≥ 100 x 10⁹ l⁻¹, haemoglobin ≥ 9.5 g 100 ml⁻¹ and normal liver function with the exception of liver function disturbances owing to metastatic disease. All patients gave informed consent and the study was carried out with ethical committee approval.

Treatment

In part I of the study patients received recombinant IL-2 at a dose of 11.7 MIU m⁻² (Teeleukin; Hoffman La Roche, Nutley, NJ, USA) and recombinant IFN-α at a dose of 3 MIU m⁻² (Roferon-A, Hoffman La Roche, Basle, Switzerland), each administered as an intravenous injection over 15 min, every 8 h on days 1–5. Treatment was repeated every 21 days up to a total of three cycles. With this schedule we were confronted with severe cardiotoxicity, albeit the first 3 days of treatment were relatively well tolerated. Therefore, we decided to modify the treatment schedule, using the same daily dosages of IL-2 and IFN-α, but for 3 instead of 5 days.

The World Health Organization (WHO) criteria were used to grade toxicity (WHO, 1979). Treatment was permanently discontinued if grade 3 neurotoxicity or cardiovascular toxicity occurred. In case of grade 3 hypotension, therapy was continued while giving symptomatic treatment with colloids. If volume expansion gave no improvement dopamine was added. Treatment was discontinued if the patient remained hypotensive and/or oliguric. In all other cases of grade 3 toxicity with the exception of fever, nausea/vomiting and diarrhoea, immunotherapy was discontinued until toxicity improved to grade 1 or resolved. Resumption of treatment at 50% of the previous dose was allowed in the next cycle if the grade 3 toxicity decreased to grade 1 or less. Corticosteroid administration was not allowed, except in the event of life-threatening toxicity.

The first tumour assessment was performed at 8 weeks after the start of therapy. Response evaluations were repeated every 4 weeks. Response categories were defined according to WHO criteria (WHO, 1979). The treatment results were analysed on an intent-to-treat basis. The median survival was estimated using the Kaplan–Meier method.

Immunological monitoring

Absolute numbers of lymphocyte subsets and cytotoxic activities of peripheral blood mononuclear cells (PBMCs) were assessed in each treatment cycle, immediately before the first administration and 1 week later. The lymphocyte subsets defined by double-staining with CD3 and CD56, CD4 and CD8, CD16 and CD19 monoclonal antibodies were assessed by multicolour immunofluorescence and flow
cytometry as described elsewhere (Gratama et al., 1996). Cytotoxic activities of lymphocytes were determined in standard 3 h $^{51}$Cr-release assays. The K562 erythromyeloid leukaemia cell line and the Daudi Burkitt’s lymphoma cell line were used as target cells for the assessment of natural killer (NK) and lymphokine-activated killer (LAK) activities respectively.

**Results**

**Patient characteristics**

Forty-three patients were entered in the study, 18 in part I and 25 in part II. The characteristics of the eligible patients are summarised in Table I. All eligible patients were evaluable for response and toxicity. In the 5 day regimen, one patient was ineligible because of unmeasurable disease. Two patients had previously been treated with isolated limb perfusion and one patient received radiotherapy before study entry. In the 3 day regimen, two patients were pretreated with adjuvant BCG or poly A–poly U.

**Table I** Patient characteristics

| Part I | Part II |
|--------|--------|
| Evaluable patients | 17 | 25 |
| **Age** | | |
| Median | 48 | 41 |
| Range | 29–61 | 20–69 |
| **Sex** | | |
| Male | 9 (53%) | 16 (64%) |
| Female | 8 (47%) | 9 (36%) |
| **Performance status (Karnofsky)** | | |
| Median | 90 | 90 |
| Range | 80–100 | 80–100 |
| **Prior therapy** | | |
| Surgery | 17 (100%) | 25 (100%) |
| Immunotherapy | 2 | 2 (8%) |
| Chemotherapy | 2 (12%) | 0 |
| Radiotherapy | 1 (6%) | 0 |
| **Distribution of metastatic sites** | | |
| Lung | 9 (53%) | 10 (40%) |
| Lymph nodes | 7 (41%) | 17 (68%) |
| Subcutaneous | 6 (35%) | 12 (48%) |
| Liver | 6 (35%) | 9 (36%) |
| Other (adrenal, pancreas, bone) | 8 (47%) | 9 (36%) |
| **Number of metastatic sites** | | |
| 1 | 6 (35%) | 7 (28%) |
| 2 | 4 (24%) | 7 (28%) |
| 3 | 4 (24%) | 7 (28%) |
| 4 | 2 (12%) | 4 (16%) |
| 5 | 1 (6%) | |

**Treatment characteristics**

Of the 17 patients in part I, nine (53%) received three cycles, three (18%) two cycles and five (29%) only one cycle. Eight patients were taken off study early; six owing to grade 3–4 toxicity, one due to rapidly progressive disease and another patient refused further treatment. The actual lymphokine doses given during the first, second and third cycle, expressed as a percentage of the planned dose, were 85%, 42% and 30% respectively.

Of the 25 patients in part II, 14 (56%), received three cycles, five (20%) two cycles and six (24%) one cycle. The lymphokine doses administered were 95%, 71% and 54% of the planned dosages respectively. Treatment was discontinued in one patient after only one single cytokine infusion owing to grade 3 toxicity. He was considered a treatment failure.

**Treatment results**

Table II shows the results at response and survival. With the 5 day schedule the overall response rate was 41% (95%
confidence interval 18–67%), including two complete responses (CRs). The overall survival was 10.2 months (range 2.6–37.5 months). With the 3 day schedule the overall response rate was 20% (95% confidence interval 7–43%). No CRs occurred. The overall survival was 6.8 months (range 0.9–24+ months).

**Toxicity**

The adverse effects are shown in Table III. With the 5 day schedule a high incidence of severe cardiac toxicity occurred. Seven patients (41%) experienced cardiac adverse events; cardiomyopathy in four, acute cardiac arrest, myocardial

| Table III Adverse events |
|--------------------------|
| **Adverse event** | **Part I** | **Part II** |
| | Grade 1/2 (%) | Grade 3/4 (%) | Grade 1/2 (%) | Grade 3/4 (%) |
| Fever | 12 | 88 | 52 | 48 |
| Skin rash | 76 | 0 | 64 | 0 |
| Nausea/vomiting | 76 | 18 | 40 | 52 |
| Diarrhoea | 53 | 35 | 48 | 44 |
| Malaise | 24 | 76 | 40 | 56 |
| Weight gain | 35 | 0 | 16 | 4 |
| Hypotension | 41 | 53 | 60 | 32 |
| Cardiac | 6 | 35 | 0 | 0 |
| Dyspnoea | 53 | 24 | 28 | 8 |
| Neuropsychiatric | 41 | 29 | 32 | 4 |
| Oliguria | 18 | 18 | 20 | 12 |
| Creatinine | 47 | 0 | 12 | 0 |
| Alkaline phosphatase | 76 | 6 | 56 | 4 |
| Bilirubin | 47 | 12 | 16 | 12 |
| Transaminases | 47 | 53 | 60 | 24 |
| Anaemia | 53 | 0 | 36 | 8 |
| Thrombocytopenia | 41 | 6 | 48 | 0 |

Figure 1 Median absolute numbers of CD56⁺, 3⁻ (a), CD3⁺ (b), CD4⁺ (c) and CD8⁺ (d) lymphocytes in patients treated in the 5 day schedule (■) and in the 3 day schedule (Δ). Logarithmic scales have been used for the vertical axes in order to compress the figure. Vertical bars represent confidence limits as defined by the 5th and 95th percentile of each group. The areas between the horizontal lines represent the normal range of the different lymphocyte subsets as defined by the 5th and 95th percentiles of healthy control persons.
infarction and negative T-waves in one patient each. The details of these patients have been reported elsewhere (Krütz et al., 1994). No cardiotoxicity was observed in the 3 day regimen.

Two-thirds of patients in the 5 day regimen suffered from neuropsychiatric disturbances such as agitation, disorientation, confusion and overt psychosis. With the 3 day regimen neuropsychiatric side-effects were encountered in about one-third of patients and were mild in most cases. Neurotoxicity completely resolved in all patients.

**Immunological monitoring**

Before therapy the absolute numbers of NK lymphocytes (CD56$^+$, CD16$^-$ and CD16$^+$), T lymphocytes (CD3$^+$), helper/inducer T lymphocytes (CD4$^+$), cytotoxic/suppressor T lymphocytes (CD8$^+$) and B lymphocytes (CD19$^+$) were within the normal range. During therapy, the CD56$^+$, CD3$^+$ and CD8$^+$ lymphocyte counts gradually increased above normal, whereas the CD4$^+$ and CD19$^+$ remained within the normal range (Figure 1). The number of lymphocytes after each treatment cycle (rebound lymphocytosis) reached a higher value in the 5-day study (Figure 1, open symbols) compared with the 3-day study (closed symbols). NK and LAK cytotoxic activities of PBMCs remained within the normal range in both parts of the study, with a large variation between patients (data not shown). There was no relationship between tumour response and immune parameters.

**Discussion**

From an earlier performed dose-escalating study of bolus IL-2 and IFN-α we selected the dosage schedule with the highest response rate. This treatment schedule resulted in a 41% response rate (95% CI 18–67%), which is identical to the results reported by the NCI investigators (Rosenberg et al., 1986).

We observed a high incidence of severe cardiotoxicity. Although myocardial toxicity was not so frequently encountered in the NCI study as here, several cases of myocardial infarction and elevation of cardiac enzyme levels in 15% of treatment courses were reported (Rosenberg et al., 1989b; Marincola et al., 1995). Also other recent literature indicates that high-dose IL-2 regimens bear a high risk of severe cardiotoxicity (Sznol et al., 1992; Atkins et al., 1993; Fossa et al., 1993).

The second severe toxicity of the 5-day regimen consisted of neuropsychiatric disturbances. Evidence from earlier studies showed that the frequency and severity of these side-effects are dose dependent (Denicoff et al., 1987; Dillman et al., 1993; Sparano et al., 1993; Keilholz et al., 1993; Atkins et al., 1993). The NCI investigators observed neurotoxicity including coma in 54% of patients (Marincola et al., 1995).

Thus, the promising results on response were accompanied by unacceptable toxicities. In order to maintain the high anti-tumour activity and to reduce the side-effects, we shortened the treatment duration in the second part of the study from 5 to 3 days. Indeed, the 3 day schedule was accompanied by acceptable toxicity, however, the response rate dropped to 26%. A direct comparison between the two schedules is difficult, because it was not a randomised study and the confidence intervals showed considerable overlap. The response rate and survival duration of the 3 day schedule were in the same range as observed in other studies of IL-2 and IFN-α in melanoma (Dillman et al., 1993; Sparano et al., 1993; Krütz et al., 1995). In a recent progress report the NCI investigators observed a similar decrease in treatment results after modification of the treatment schedule (one instead of three IFN-α administrations per day), made necessary by the encountered toxicity (Marincola et al., 1995). In general the response rates and survival data of combined therapy of IL-2 and IFN-α in patients with metastatic melanoma appeared not to be superior to treatment with IL-2 alone and IL-2 in combination with lymphokine-activated killer cells (Bar et al., 1990; Dutcher et al., 1991; Rosenberg et al., 1992, 1994; Sparano et al., 1993; Marincola et al., 1995).

In summary, the combination of IL-2 and IFN-α has not meaningfully improved the clinical results that may be obtained with IL-2 alone. Based on the possibly additive or synergistic effects between cytotoxic agents and cytokines, biochemotherapy may be an important new treatment modality. Combinations of biological response modifiers and cytostatics have produced encouraging response rates (Richards et al., 1992; Khayat et al., 1993; Legha et al., 1993).

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