Deciphering Genomic Underpinnings of Quantitative MRI-based Radiomic Phenotypes of Invasive Breast Carcinoma

Yitan Zhu1,*, Hui Li2,*, Wentian Guo3, Karen Drukker2, Li Lan2, Maryellen L. Giger2 & Yuan Ji1,4

Magnetic Resonance Imaging (MRI) has been routinely used for the diagnosis and treatment of breast cancer. However, the relationship between the MRI tumor phenotypes and the underlying genetic mechanisms remains under-explored. We integrated multi-omics molecular data from The Cancer Genome Atlas (TCGA) with MRI data from The Cancer Imaging Archive (TCIA) for 91 breast invasive carcinomas. Quantitative MRI phenotypes of tumors (such as tumor size, shape, margin, and blood flow kinetics) were associated with their corresponding molecular profiles (including DNA mutation, miRNA expression, protein expression, pathway gene expression and copy number variation). We found that transcriptional activities of various genetic pathways were positively associated with tumor size, blurred tumor margin, and irregular tumor shape and that miRNA expressions were associated with the tumor size and enhancement texture, but not with other types of radiomic phenotypes. We provide all the association findings as a resource for the research community (available at http://compgenome.org/Radiogenomics/). These findings pave potential paths for the discovery of genetic mechanisms regulating specific tumor phenotypes and for improving MRI techniques as potential non-invasive approaches to probe the cancer molecular status.

Precise cancer diagnosis and treatment rely on the integration of information from various sources, such as the phenotypic and genotypic profilings of tumors. Radiogenomics aims to integrate computer-extracted phenotypes from radiological imaging data with genomic data, providing an opportunity to investigate the association between the radiomic tumor phenotypes and the genomic measurements of the same tumors. Such a study may identify the genetic mechanisms that regulate the development of specific tumor phenotypes. Radiomic phenotypes that are highly correlated with important genomic biomarkers can potentially serves as diagnosis and prognosis tools for patient monitoring, and therefore augment the utility of radiological imaging as a non-invasive technology for cancer care.

Radiogenomics is a new scientific field with scarce applications. This is mainly due to the lack of data consisting of both imaging and genomic measurements on the same set of tumors. Nevertheless, a few recent studies have pioneered early endeavor. Studies in lung cancer1–2, head & neck cancer2, glioblastoma multiforme3, and clear cell renal cell carcinoma4 attempted to correlate tumor radiomic phenotypes with DNA mutations4, mRNA expressions1–3 and copy number variations3. For breast cancer, Yamamoto et al.5 collected Magnetic Resonance Imaging (MRI) data and gene expression data from 10 human tumors and correlated 26 imaging phenotypes defined by radiologists with the expressions of individual genes and gene sets. Maurowski et al.6 extracted radiomic phenotypes based on 48 patients and...
discovered the phenotypes associated with the luminal B subtype of breast cancer. Based on 56 Estrogen Receptor Positive (ER⁺) breast cancers, Ashraf et al.⁷ used computationally derived radiomic phenotypes to predict the recurrence likelihood score defined by Oncotype DX, a validated gene expression assay including 21 selected genes. Agner et al.⁸ extracted quantitative radiomic features of 76 breast lesions and used them to differentiate the triple-negative breast cancer from other subtypes. In addition, BRCA1/2 and UGT2B variations have been associated with computer-extracted radiomic phenotypes⁹–¹¹.

Here, we report a comprehensive radiogenomic study of breast invasive carcinoma based on the integration of The Cancer Imaging Archive (TCIA)¹² and The Cancer Genome Atlas (TCGA)¹³, two leading cancer research projects supported by the U. S. National Institutes of Health. We integrated the Dynamic Contrast Enhanced-MRI (DCE-MRI) data with the multi-platform genomic data for 91 primary breast tumors. Through an extensive investigation, we identified statistically significant associations between various genomic features and radiomic phenotypes in breast invasive carcinoma that have yet to be reported. Among the many novel findings, we discovered highly specific associations of radiogenomic features, which are potentially useful for (1) the imaging based diagnosis that can inform the genetic progress of tumor and (2) the discovery of genetic mechanisms that regulate the development of tumor phenotype. We believe that our study is the first of its kind that investigates the relationships between the multi-layer tumor molecular system and the various quantitative radiomic phenotypes of breast cancer.

Results

Summary of Associations between Genomic Features and Radiomic Phenotypes. Through the Gene-Set Enrichment Analysis (GSEA)¹⁴–¹⁵ (Supplementary Information Section 5) and the linear regression analysis (Supplementary Information Sections 6–8), we performed a quantitative study to associate genomic features, including miRNA expressions, protein expressions, and gene somatic mutations, and transcriptional activities and gene CNVs of all genetic pathways in the Kyoto Encyclopedia of Genes and Genomes (KEGG)¹⁶ database, with six categories of radiomic phenotypes, including tumor size, shape, morphology, enhancement textures, kinetic curve assessments, and enhancement-variance kinetics. The study schema is presented in Fig. 1 and the main results are presented in Fig. 2. Specifically, Fig. 2a shows the statistically significant associations and Fig. 2b summarizes the numbers of associations between different categories of genomic features and radiomic phenotypes. Fisher’s exact test¹⁷–¹⁸ was applied to the numbers reported in Fig. 2b and concluded that the frequencies of statistically significant associations are dependent on the categories of genomic features and radiomic phenotypes (p-value ≤ 1.0 × 10⁻⁸). In other words, some types of genomic features and radiomic phenotypes are more likely to be associated than others.

The most intriguing findings in Fig. 2 are related to the associations of two types of genomic features, 1) transcriptional activities of pathways and 2) miRNA expressions. Specifically, pathway transcriptional activities are associated with all six types of radiomic phenotypes with statistical significance (Table S3), indicating that they can regulate various aspects of the tumor phenotype. Strikingly, statistically significant associations between pathway transcriptional activities and all four tumor size phenotypes (including lesion volume, effective diameter, surface area, and maximum linear size) are extremely specific in that more than 97.7% of the associations are positive (adjusted p-values ≤ 6.21 × 10⁻⁹ by the Chi-squared proportion tests with equal proportions of positive and negative associations), indicating that many pathways are up-regulated during tumor growth since larger tumors are associated with mostly higher pathway activities. We find that pathway transcriptional activities are mostly negatively associated with two tumor morphological features including margin sharpness and variance of radial gradient histogram (with adjusted p-values ≤ 0.043 from the proportion tests). This suggests a positive correlation between...
the transcriptional activities of genetic pathways and a blurred tumor margin, which is potentially a sign of tumor invasion into the surrounding tissue. Also, the transcriptional activities of pathways are mostly positively associated with the irregularity of tumor shape, another sign of aggressive tumor, characterized by two radiomic phenotypes \textit{irregularity} and \textit{surface to volume ratio} (with adjusted p-values ≤ 0.00285 from the proportion tests).

The associations between miRNA expressions and radiomic phenotypes are highly specific in that miRNA expressions are only associated with primarily two types of radiomic phenotypes, tumor size and enhancement texture (Fig. 2 and Table S3). Statistically significant associations between miRNA expressions and three out of the four tumor size phenotypes are dominantly positive with proportions ≥ 92.3% and adjusted p-values ≤ 0.00118 from the proportion tests. This suggests that miRNAs may mainly mediate the growth of tumor and the heterogeneity of blood vessel system in tumor. Such insights on the role of miRNA may facilitate the cancer mechanism study and the design of miRNA targeted treatment. Conversely, due to the specificity in the associated phenotypes, it is possible to use radiomic phenotypes characterizing the tumor size and enhancement texture to predict miRNA activities without the need for tumor biopsy and miRNA profiling.

Compared to the transcriptional activities of genetic pathways, the CNVs of pathways have much fewer statistically significant associations (Fig. 2), which are enriched with only the tumor size...
phenotypes (adjusted $p$-value $= 7.95 \times 10^{-8}$, Table S3) and the enhancement-variance kinetics (adjusted $p$-value $= 4.46 \times 10^{-6}$, Table S3). TCGA uses the Reverse Phase Protein Array (RPPA) to measure the expression levels of 142 proteins and phospho-proteins related to breast cancer. Protein expressions show enriched associations with the tumor size phenotypes (adjusted $p$-value $= 0.0486$, Table S3) and the morphological phenotypes (adjusted $p$-value $= 0.0244$, Table S3), but not with any other phenotype category.

We discuss the identified associations of different types of genomic features in detail in the next four sub-sections.

**Associations between Genetic Pathways and Radiomic Phenotypes.** The associations between the transcriptional activities of KEGG pathways and the radiomic phenotypes were studied using GSEA\textsuperscript{14–15}. A total of 1,103 statistically significant (adjusted $p$-values $\leq 0.05$) associations have been identified (Fig. 2 and Table S4). Fig. 3 provides some examples involving cancer-related pathways, which we elaborate below.

**Cell Cycle, DNA Replication and Ribosome.** Tumor growth requires excessive cell proliferation, for which 1) DNA replication, 2) protein synthesis, and 3) cell cycle are essential. Genes involved in cell cycle and DNA replication are positively associated with all four tumor size phenotypes (Fig. 3), indicating their activations during tumor growth. All these three gene modules are also positively associated with enhancement at the first post-contrast time point, normalized total rate variation, and maximum variance of enhancement, which characterize the blood flow dynamics and the contrast uptake heterogeneity in tumor.

**Conserved Regulations in Cancer.** KEGG does not provide a genetic pathway dedicated to breast cancer, but provides a comprehensive regulation map called “pathways in cancer”\textsuperscript{19} that includes the conserved regulation mechanisms across cancer types. The transcription activity of this large molecular regulation system is positively associated with 14 radiomic phenotypes and negatively associated with 9 radiomic phenotypes (Fig. 3). It is positively associated with the tumor size phenotypes. Its activity is also associated with an increased tumor shape irregularity characterized by the irregularity and sphericity phenotypes, which is usually a sign of malignant and aggressive tumor\textsuperscript{20}.

**JAK-STAT Signaling Pathway.** The JAK-STAT signaling cascade forms the principal signaling transduction mechanism in response to a variety of cytokines and growth factors\textsuperscript{21}. Over-activation of the JAK-STAT pathway can cause cancer by evading apoptosis and forming self-sufficient growth signals\textsuperscript{22}. According to our analysis, the transcriptional activity of JAK-STAT signaling pathway is positively associated with the tumor size phenotypes (Fig. 3). Also, we found a statistically significant association between...
its activity and the tumor shape irregularity measured by two radiomic phenotypes, *irregularity* and *sphericity*. We also see that the pathway has statistically significant positive associations with *maximum enhancement* and *enhancement at first post-contrast time point*, which implies that tumors with a higher JAK-STAT pathway activity have more leaky microvessels to support its growth.

**Cell Adhesion Molecules.** The cell adhesion molecules are in direct or indirect control of cellular activities such as adhesion, proliferation, migration and differentiation. Aberrant activities of cell adhesion molecules disrupt normal cell-cell and cell-matrix interactions and can facilitate tumor formation and metastasis. In Fig. 3, we find that expressions of cell adhesion genes are correlated with the signs of tumor malignancy and aggressiveness, such as a large tumor size, an increased tumor shape irregularity, and increased blood flow dynamics (measured by *maximum enhancement*, *enhancement at first post-contrast time point*, and *uptake rate*).

**TGF-beta Signaling Pathway.** The role of TGF-beta signaling pathway in breast cancer has been intensively studied. *TGF-beta* serves as a tumor suppressor at the initial stage of tumorigenesis, but loses its growth inhibition function during cancer progression and diverts towards promoting motility, invasion and metastasis at late stages. We find that a strong activity of TGF-beta signaling pathway is positively associated with irregular tumor shape characterized by *sphericity* (Fig. 3). Its statistically significant positive association with the *maximum variance of enhancement* implies a correlation between the activity of TGF-beta signaling pathway and the heterogeneous blood distribution in tumor.

Section 5 in the Supplementary Information provides more explanations on the associations involving other cancer-related pathways shown in Fig. 3. We also identified statistically significant associations between the CNVs of pathways and radiomic phenotypes (Fig. S2 in the Supplementary Information).
Associations between miRNA Expressions and Radiomic Phenotypes. Using the linear regression analysis (see Section 6 in the Supplementary Information), we found statistically significant (adjusted p-value ≤ 0.05) associations between miRNA expressions and primarily two types of radiomic phenotypes, tumor size and enhancement texture (Fig. 2). Table S5 in the Supplementary Information shows all the identified associations. We curated a list of miRNAs related to cancer development, especially breast cancer formation, by literature survey26–29, and present in Fig. 4a the statistically significant associations involving these cancer-related miRNAs. MiR-128-1 plays an oncogenic role in drug-resistant breast cancer cell by interfering with TGF-beta signaling27. Its expression is found to be positively associated with the tumor size. MiR-18a has been reported to induce tumor growth and tumor vascularization26. Its expression is positively associated with lesion volume and difference variance. Both miR-19a and miR-18a belong to the miR-17-92 cluster. The expression of miR-19a is statistically significantly associated with enhancement texture phenotypes, including contrast, correlation, difference variance, entropy, and maximum correlation coefficient. These associations indicate that the expression of miR-19a correlates with the heterogeneity of tumor enhancement texture, which is also a sign of aggressive and malignant lesion26. Let-7b shows an opposite association pattern with the enhancement texture phenotypes compared to miR-19a, probably due to the tumor suppressive function of the let-7 family26. MiR-10b has been reported as a modulator of tumor invasion and metastasis26,27. Its expression is associated with tumor effective diameter.

Associations between Protein Expressions and Radiomic Phenotypes. All statistically significant (adjusted p-value ≤ 0.05) associations between protein expressions and radiomic phenotypes are shown in Fig. 4b. P-cadherin is a calcium-dependent cell-cell adhesion glycoprotein encoded by CDH3 in human. Its expression has been shown to be correlated with high histologic grade, increased proliferation, and poor patient survival in breast cancer30–33. We find a statistically significant positive association between the expression of P-cadherin and the tumor size, measured by effective diameter, surface area, and lesion volume. JNK2 is a mitogen-activated protein kinase encoded by MAPK9 in the MAPK signaling pathway. It is considered as a negative regulator of cellular proliferation30, and cooperates with JNK1 in activating the p53 singling pathway to induce apoptosis30. The anti-tumorigenic role of JNK2 is demonstrated in our analysis as its expression is negatively associated with the tumor size phenotypes and positively associated with tumor margin sharpness, a phenotype signaling the absence of tumor invasion into the surrounding tissue.

Associations between Somatic Gene Mutations and Radiomic Phenotypes. We compared the measurements of a radiomic phenotype for patients harboring somatic mutations in a gene versus those not (Section 8 in the Supplementary Information). Table S7 in the Supplementary Information shows all the associations in which a gene mutated in at least five patients and the obtained p-value ≤ 0.05.

PIK3CA is an oncogene participating in the signaling cascades of cell growth, survival, proliferation, motility and morphology. Four radiomic phenotypes, including one kinetic curve assessment and three enhancement texture phenotypes, are associated with PIK3CA mutations. GATA3 has been observed by TCGA as the third most frequently mutated gene in breast invasive carcinoma after TP53 and PIK3CA, with an overall mutation rate larger than 10%13. GATA3 encodes a transcription factor that regulates luminal epithelial cell differentiation in the mammary gland34. Its expression is progressively lost during luminal breast cancer progression as cancer cells acquire a stem cell-like phenotype35. Our analysis shows that mutations in GATA3 are negatively associated with tumor size (measured by three tumor size phenotypes), tumor shape irregularity (measured by irregularity), and sum entropy that measures the randomness of enhancement texture. Such an observation leads to the hypothesis that mutations in GATA3, although frequent, might not be driver mutations causing tumor progression, because a large tumor size, an irregular tumor shape, and random enhancement texture are usually signs of malignant and aggressive tumors. MAP2K4 encodes a kinase in the MAPK signaling pathway and is considered as a tumor suppressor36. Its mutations are positively associated with time to peak and negatively associated with uptake rate, indicating tumors with MAP2K4 mutations have a relatively slow blood flow and fewer leaky microvessels.

We also studied the associations of somatic gene mutations at the pathway level, by comparing the measurement of a radiomic phenotype for patients with gene mutations in a KEGG pathway versus those without (see Section 8 in the Supplementary Information). Table S8 in the Supplementary Information shows all the identified statistically significant associations. An interesting observation is that somatic mutations in the p53 signaling pathway was found to be positively associated with tumor effective diameter, indicating potentially that DNA mutations can damage the tumor suppressive function of the p53 signaling pathway and thus induce tumor growth.

Discussion
Based on the integrated data from TCIA and TCGA, we have conducted a comprehensive radiogenomic study to explore the association between multi-platform genomic profiles and MRI-based tumor
phenotypes for breast invasive carcinoma. Our study generated two major findings that have not been previously reported. First, we identified statistically significant associations between six types of radiomic tumor phenotypes and various genomic features involved in multiple molecular regulation layers. Our large-scale study produced a new resource (available at http://compgenome.org/Radiogenomics/) for exploring the genetic mechanisms that potentially regulate the formation of various tumor phenotypes. We expect our findings will facilitate future radiogenomic research and provide a template for the type of analyses that could be carried out. Second, we observed highly specific patterns for the identified associations. Many genetic pathways are more active in tumors with a large size, irregular shape, and blurred margin. MiRNA expressions are associated with only tumor size and enhancement texture, but not other types of radiomic phenotypes. These patterns provide new insights on the genetic mechanisms that regulate tumor development. They are also potentially useful in the clinical diagnosis of cancer by suggesting the candidate radiomic phenotypes for predicting genomic features, although further validations are needed. If validated, these findings could augment the use of MRI as a non-invasive technology, not only for examining tumor phenotypes, but also for probing the underlying molecular status of tumor, which is crucial for personalized treatment.

Compared to the associations at the transcriptional level, we found much fewer statistically significant associations for pathway CNVs and gene somatic mutations. There could be two reasons for this observation. Firstly, DNA mutation events, such as CNVs and somatic mutations are rarely shared across many patients (Table S6). Thus, there lacks sufficient statistical power for identifying a potential association, especially given the small sample size in almost all radiogenomic studies. We believe that our initial results can trigger the motivation for future large-scale radiogenomic studies that include a large number of samples. Secondly, compared to DNA mutations, gene expressions are more directly related to phenotype in the process of genetic events influencing phenotype development. Genetic mutations are more upstream in the functional activities of the cellular system. Therefore, gene expressions present more associations with tumor phenotypes.

An interesting finding is that the transcriptional activity of basal cell carcinoma pathway is statistically significantly associated with 12 radiomic phenotypes of breast invasive carcinoma (Table S9). Actually, among all the cancer-type-specific KEGG pathways, the basal cell carcinoma pathway is associated with the largest number of breast cancer radiomic phenotypes (Section 9 and Table S9 in the Supplementary Information). Such associations between breast and skin cancer are consistent with their correlation in terms of disease prevalence. Patients with basal cell carcinoma, especially those diagnosed at a young age, have been reported to have an increased risk for noncutaneous cancers including breast cancer.

Our study is based on genomic data generated by a single tissue sample from each primary tumor. Tumor contains spatially heterogeneous cell populations. A single biopsy sample of a tumor usually contains multiple cell subpopulations, but typically cannot encompass all the subclones of a tumor. Therefore, the genomic profile of a single tumor sample may be incomplete and only partially reflect the overall genomic landscape of the entire tumor. All these can affect the results obtained through the radiogenomic analysis. A comprehensive study would require multiple biopsy samples from the same tumor, which is often costly and labor-intensive in practice. In return, such a study will be more informative and accurate.

In addition to the association analysis, we did clustering analysis on the tumor samples to provide an overview of the radiomic and expression data used in the analysis (Fig. S1 and Section 3 in the Supplementary Information). The clustering partitions of tumors were associated with the clinical subtypes of tumors defined by their pathological state and molecular receptor status (Table S2). Tumor clustering partitions based on gene expressions, miRNA expressions, and protein expressions are all statistically significantly (adjusted p-values ≤ 0.05) associated with the statuses of Estrogen Receptor (ER) and Progesterone Receptor (PR), which means that ER+ patients and PR+ patients show different expression patterns from ER− patients and PR− patients, respectively, at multiple molecular levels (Table S2 and Fig. S1).

Survival analysis is not included in our study, because of the short overall follow up (median is 870 days) and the small number of mortality events (1 out of 91). In this work, we are focusing on understanding the relationship between MRI tumor phenotypes and underlying genetic mechanisms, if there are any. We attempt to address a critical aspect of patient care – the use of noninvasive technology. Genomic features reflect the molecular characteristics of a tumor, but are obtained through invasive procedures such as surgery or biopsy. Through the study of radiogenomics, we aim to identify good surrogate radiomic features that can reveal genetic changes of tumors, thereby establishing noninvasive means for monitoring tumor progression.

More analyses have been planned for future radiogenomic study of breast cancer. Integration of multiple genomic, epigenomic, proteomic features simultaneously with radiomic phenotypes can provide a better understanding of how the multiple molecular regulation layers generate the observed tumor phenotypes. Graphical models can be a powerful tool to study the complex relationships between radiomic and genomic features, which takes into account the potential competitive regulations and conditional dependence between them. Another interesting topic is to use predictive modeling to predict the status of a genomic feature (especially the biomakers important for diagnosis, prognosis, and response to therapy) based on imaging phenotypes that have been shown to associate with genomic features.
Methods

The preparation of data and the organization of the analysis in this study are illustrated in Fig. 1. The DCE-MRIs of 91 breast cancers were downloaded from TCIA. These cases were contributed by four institutions, including Memorial Sloan Kettering Cancer Center, Mayo Clinic, University of Pittsburg Medical Center, and Roswell Park Cancer Institute. Section 1 in the Supplementary Information introduces the imaging cases, including the patient populations and the MRI pulse sequences used. Using a quantitative MRI radiomics workstation, i.e. the Quantitative Image Analysis (QIA) workstation\textsuperscript{20,40–45} that had been initially developed for computer-aided diagnosis research, we computationally extracted 38 radiomic phenotypes from the DCE-MRIs for each of the 91 primary tumors to characterize size, shape, margin, enhancement texture, kinetics, and variance kinetics. See Table S1 in the Supplementary Information for a summary of the radiomic phenotypes by category. Section 1 in the Supplementary Information also provides the mathematical descriptions of radiomic phenotypes. Using TCGA-Assembler\textsuperscript{46}, we retrieved and processed the multi-layer genomic data of these tumors from TCGA, including gene expressions, copy number variations (CNV), protein expressions, miRNA expressions, and somatic mutations. Section 2 in the Supplementary Information introduces the sample inclusion criteria of TCGA and the assays, platforms, and algorithms used for generating the genomic data.

The radiomic data and genomic data were then combined as described in Fig. 1. See Table 1 for a summary of the integrated data. ER, PR and HER2 refer to Estrogen Receptor, Progesterone Receptor, and Human Epidermal growth factor Receptor 2, respectively.

| Data Platform         | Number of Features | Number of tumors |
|-----------------------|--------------------|------------------|
| Radiomics             | 38                 | 91               |
| Gene expressions      | 20531 genes (186 pathways) | 91               |
| Copy number variations| 19950 genes (186 pathways) | 91               |
| miRNA expressions     | 1046               | 91               |
| Protein expressions   | 142                | 62               |
| Mutated genes         | 3734               | 91               |

| Number of tumors with different pathological stages |
|-----------------------------------------------------|
| Pathological Stage | T | M | N | Overall |
|---------------------|---|---|---|---------|
| 0                   | 91 | 46 |
| I                   | 38 | 34 | 22 |
| II                  | 50 | 6  | 58 |
| III                 | 3  | 4  | 11 |
| X                   |    |    | 1  |

| Number of tumors with different molecular receptor statuses |
|-------------------------------------------------------------|
| Receptor Status | ER | PR | HER2 |
|-----------------|----|----|------|
| Negative        | 14 | 19 | 72   |
| Positive        | 77 | 72 | 19   |

Table 1. Summary of integrated data. ER, PR and HER2 refer to Estrogen Receptor, Progesterone Receptor, and Human Epidermal growth factor Receptor 2, respectively.
Sections 6, 7 and 8 of the Supplementary Information introduce the details of linear regression analysis for identifying associations between radiomic phenotypes and three types of genomic features, including miRNA expression, protein expression, and gene somatic mutation, respectively. Data preprocessing steps, mathematical formulas, and the analysis procedure are included in those sections. Table S5 shows the analysis results of all identified associations involving miRNA expressions. Table S6 summarizes the frequencies of somatic gene mutations among patients. Table S7 and Table S8 present the analysis results of all identified associations involving somatic gene mutations at the single gene level and at the pathway level, respectively.

Section 9 and Table S9 in the Supplementary Information summarize the numbers of associations between the radiomic phenotypes of invasive breast carcinoma and the transcriptional activities of KEGG pathways dedicated to other types of cancer. This information is provided for the discussion on the relationship between breast cancer and basal cell carcinoma.

References

1. Govaert, O. et al. Non-small cell lung cancer: identifying prognostic imaging biomarkers by leveraging public gene expression microarray data–methods and preliminary results. Radiology 264, 387–296, doi: 10.1148/radiol.12111607 (2012).
2. Aerts, H. J. W. L. et al. Decoding tumour phenotype by noninvasive imaging using a quantitative radiomics approach. Nat. Commun. 5, doi: 10.1038/ncomms5006 (2014).
3. Jamshidi, N., Diefen, M., Bredel, M. & Kuo, M. D. Illuminating radiogenomic characteristics of glioblastoma multiforme through integration of MR imaging, messenger RNA expression, and DNA copy number variation. Radiology 270, 212–222, doi: 10.1148/radiol.13130078 (2014).
4. Karlo, C. A. et al. Radiogenomics of clear cell renal cell carcinoma: associations between CT imaging features and mutations. Radiology 270, 464–471, doi: 10.1148/radiol.13130663 (2014).
5. Yamamoto, S., Maki, D. D., Korn, R. L. & Kuo, M. D. Radiogenomic analysis of breast cancer using MRI: a preliminary study to define the landscape. Am. J. Roentgenol. 199, 654–665, doi: 10.2214/AJR.11.7824 (2012).
6. Mazurowski, M. A., Zhang, J., Green, L. J., Yoon, S. C. & Silber, J. I. Radiogenomic analysis of breast cancer: luminal B molecular subtype is associated with enhanced dynamics at MR imaging. Radiology 273, 365–372, doi: 10.1148/radiol.14132641 (2014).
7. Ashraf, A. B. et al. Identification of intrinsic imaging phenotypes for breast cancer tumors: preliminary associations with gene expression profiles. Radiology 272, 374–384, doi: 10.1148/radiol.14131375 (2014).
8. Agner, S. C. et al. Computerized image analysis for identifying triple-negative breast cancers and differentiating them from other molecular subtypes of breast cancer on dynamic contrast-enhanced MR images: a feasibility study. Radiology 272, 91–99, doi: 10.1148/radiol.14121031 (2014).
9. Li, H. et al. Pilot study demonstrating potential association between breast cancer image-based risk phenotypes and genomic biomarkers. Med. Phys. 41, doi: 10.1118/1.4865811 (2014).
10. Li, H., Giger, M. L., Lan, L., Janardanan, J. & Sennett, C. A. Comparative analysis of image-based phenotypes of mammographic density and parenchymal patterns in distinguishing BRCA1/2 cases, unilateral cancer cases, and controls. J. Med. Imaging 1, doi: 10.1117/1.JMI.1.3.031009 (2014).
11. Gierach, G. L. et al. Relationships between computer-extracted mammographic texture pattern features and BRCA1/2 mutation status: a cross-sectional study. Breast Cancer Res. 16, 424, doi: 10.1186/s13058-014-0424-8 (2014).
12. Clark, K. et al. The Cancer Imaging Archive (TCIA): maintaining and operating a public information repository. J. Digit. Imaging 26, 1045–1057, doi: 10.1007/s10278-013-9622-7 (2013).
13. Network, T. C. A. Comprehensive molecular portraits of human breast tumours. Nature 490, 61–70, doi: 10.1038/nature11412 (2012).
14. Subramanian, A. et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. Proc. Natl. Acad. Sci. USA 102, 15545–15550, doi: 10.1073/pnas.0506580102 (2005).
15. Väreno, L., Nielsen, J. & Nookaew, I. Enriching the gene set analysis of genome-wide data by incorporating directionality of gene expression and combining statistical hypotheses and methods. Nucleic Acids Res. 41, 4378–4391, doi: 10.1093/nar/gkt111 (2013).
16. Kanehisa, M., Goto, S., Sato, Y., Furumichi, M. & Tanabe, M. KEGG for integration and interpretation of large-scale molecular data sets. Nucleic Acids Res. 40, D109–114, doi: 10.1093/nar/gkr988 (2012).
17. Mehta, C. R. & Patel, N. R. Algorithm 643. FEXACT: A Fortran subroutine for Fisher’s exact test on unordered r×c contingency tables. ACM Trans. Math. Softw. 12, 154–161, doi: 10.1145/6497.214326 (1986).
18. Clarkson, D. B., Fan, Y. & Joe, H. A Remark on Algorithm 643: FEXACT: An Algorithm for Performing Fisher’s Exact Test in r×c Contingency Tables. ACM Trans. Math. Softw. 19, 484–488, doi: 10.1145/168173.168412 (1993).
19. Kanehisa, M. KEGG pathways in cancer - Homo sapiens (human). <http://www.genome.jp/kegg-bin/show_pathway?hsa05200> (Date of access: 18/03/2014).
20. Chen, W. Computerized analysis and interpretation of breast MR images Ph.D. thesis, The University of Chicago, (2007).
21. Dutta, P. & Li, W. X. Role of the JAK-STAT Signalling Pathway in Cancer. In: eLS. doi: 10.1002/9780470015902.a0025214 (John Wiley & Sons, Ltd, 2013).
22. Wagner, K. & Schmidt, J. W. The two faces of Janus kinases and their respective STATs in mammary gland development and cancer. J. Cancer Res. 10, doi: 10.4103/1477-3163.96677 (2011).
23. Wong, C. W., Dye, D. E. & Coombe, D. R. The role of immunoglobulin superfamily cell adhesion molecules in cancer metastasis. Int. J. Cell Biol. 2012, doi: 10.1155/2012/340296 (2012).
24. Scollen, S. et al. TGF-β Signaling Pathway and Breast Cancer Susceptibility. Cancer Epidemiol. Biomarkers Prev. 20, 1112–1119 (2011).
25. Barcellos-Hoff, M. H. & Akhurst, R. J. Transforming growth factor-β in breast cancer: too much, too late. Breast Cancer Res. 11, doi: 10.1186/bcr2224 (2009).
26. Spizzo, R., Nicoloso, M. S., Croce, C. M. & Calin, G. A. Snapshot: microRNAs in cancer. Cell 137, 586–586, doi: 10.1016/j.cell.2009.04.040 (2009).
27. Corcoran, C., Friel, A. M., Duffy, M. J., Crown, J. & O’Driscoll, L. Intracellular and extracellular microRNAs in breast cancer. Clin. Chem. 57, 18–32 (2011).
28. O’Day, E. & Lal, A. MicroRNAs and their target gene networks in breast cancer. Breast Cancer Res. 12 (2010).
29. Chen, P. S., Su, J. L. & Hung, M. C. Dysregulation of microRNAs in cancer. J. Biomed. Sci. 19, doi: 10.1186/1423-0127-19-90 (2012).
30. Paredes, J. et al. P-cadherin overexpression is an indicator of clinical outcome in invasive breast carcinomas and is associated with CD3H promoter hypomethylation. Clin. Cancer Res. 11, 5869–5877 (2005).
31. Paredes, J. et al. P-cadherin expression in breast cancer: a review. Breast Cancer Res. 9, 214, doi: 10.1186/bcr1774 (2007).
32. Sabapathy, K. & Wagner, E. F. JNK2: a negative regulator of cellular proliferation. Cell Cycle 3, 1520–1523 (2004).
33. Oleinik, N. V., Krupenko, N. I. & Krupenko, S. A. Cooperation between JNK1 and JNK2 in activation of p53 apoptotic pathway. Oncogene 26, 7222–7230 (2007).
34. Kourosh-Mehr, H., Slorach, E. M., Sternlicht, M. D. & Werb, Z. GATA-3 maintains the differentiation of the luminal cell fate in the mammary gland. Cell 127, 1041–1055, doi: 10.1016/j.cell.2006.09.048 (2006).
35. Chou, J., Provot, S. & Werb, Z. GATA3 in development and cancer differentiation: cells GATA have it! J. Cell. Physiol. 222, 42–49, doi: 10.1002/jcp.21943 (2010).
36. Curtis, C. et al. The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups. Nature 486, 346–352, doi: 10.1038/nature10983 (2012).
37. Zhu, Y. Associations between genomic features and radiomic tumor phenotypes for invasive breast carcinoma, <http://www.compgenome.org/Radiogenomics/Associations.xlsx> (Date of access: 08/10/2015).
38. Frisch, M., Hjalgrim, H., Olsen, J. H. & Melbye, M. Risk for subsequent cancer after diagnosis of basal-cell carcinoma. A population-based, epidemiologic study. Ann. Intern. Med. 125, 815–821 (1996).
39. Zhu, Y. et al. Zodiac: A Comprehensive Depiction of Genetic Interactions in Cancer by Integrating TCGA Data. JNCI-J. Natl. Cancer Inst. 107, djv129, doi: 10.1093/jnci/djv129 (2015).
40. Shimauchi, A. et al. Evaluation of clinical breast MR imaging performed with prototype computer-aided diagnosis breast MR imaging workstation: reader study. Radiology 258, 696–704, doi: 10.1148/radiol.10100409 (2011).
41. Chen, W., Giger, M. L., Bick, U. & Newstead, G. M. Automatic identification and classification of characteristic kinetic curves of breast lesions on DCE-MRI. Med. Phys. 33, 2878–2887 (2006).
42. Chen, W., Giger, M. L., Lan, L. & Bick, U. Computerized interpretation of breast MRI: investigation of enhancement-variance dynamics. Med. Phys. 31, 1076–1082 (2004).
43. Chen, W., Giger, M. L., Li, H., Bick, U. & Newstead, G. M. Volumetric texture analysis of breast lesions on contrast-enhanced magnetic resonance images. Magn. Reson. Med. 58, 562–571 (2007).
44. Gilhuijs, K. G. A., Giger, M. L. & Bick, U. Automated analysis of breast lesions in three dimensions using dynamic magnetic resonance imaging. Med. Phys. 25, 1647–1654 (1998).
45. Chen, W., Giger, M. L. & Bick, U. A fuzzy c-means (FCM)-based approach for computerized segmentation of breast lesions in dynamic contrast-enhanced MR images. Acad. Radiol. 13, 63–72 (2006).
46. Zhu, Y., Qiu, P. & Ji, Y. TCGA-Assembler: open-source software for retrieving and processing TCGA data. Nat. Methods 11, 599–600 (2014).

Acknowledgements
Maryellen L. Giger’s research was partly supported by a University of Chicago Dean Bridge Fund. She is a stockholder in R2 Technology/Hologic and receives royalties from Hologic, GE Medical Systems, MEDIAN Technologies, Riverain Medical, Mitsubishi, and Toshiba. She is a cofounder of and a stockholder in Quantitative Insights. Yuan Ji’s research is partly supported by NIH 2R01 CA132897.

Author Contributions
Y.J., M.L.G., Y.Z. and H.L. conceived the main idea and participated in all aspects of the project. Y.Z. and W.G. prepared the genomic data and performed the integrative data analysis. H.L., K.D. and L.L. prepared the computer-extracted image-based (radiomics) data. All authors edited the manuscript.

Additional Information
Supplementary information accompanies this paper at http://www.nature.com/srep

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Zhu, Y. et al. Deciphering Genomic Underpinnings of Quantitative MRI-based Radiomic Phenotypes of Invasive Breast Carcinoma. Sci. Rep. 5, 17787; doi: 10.1038/srep17787 (2015).

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/