**Case Report**

**Campylobacter fetus** subsp. *fetus* bacteremia in a patient with myelodysplastic syndrome: A case report

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**ABSTRACT**

*Campylobacter fetus* is an important pathogen of extra intestinal infections in patients with immunodeficiency. A case of *Campylobacter fetus* subsp. *fetus* bacteremia in a patient with myelodysplastic syndrome was presented in this study. A 78-year-old woman with myelodysplastic syndrome was admitted to the emergency due to chest pain and dyspnea. A diagnosis of pneumonia was made, and blood, throat, and urine samples were obtained for microbiological tests. Trimethoprim-sulfamethoxazole and piperacillin-tazobactam were empirically initiated. *C. fetus subsp. fetus* was isolated from blood cultures and clarithromycin was added. The patient responded an overall 21-day antimicrobial treatment. Because repeated blood cultures were negative, and her general condition improved, upon that she was discharged from the hospital.

In conclusion, the incidence of *C. fetus* infections was low, and case reports are rare in current literature in Turkey. The infections can occur with these bacteria should not be overlooked in the clinical laboratory.

**Key words:** *Campylobacter fetus* subsp. *fetus*, bacteremia, myelodysplastic syndrome

**INTRODUCTION**

The *Campylobacter* genus is a Gram-negative, curved microorganism including 143 species within the NCBI Taxonomy.¹ *Campylobacter* is a significant pathogen of intestinal infections in human being. This microorganism sporadically causes bacteremia or extra intestinal infections. Although intestinal infections are usually caused by *Campylobacter jejuni*, *Campylobacter fetus* is the important causative microorganism of extra intestinal infections.²–⁴ *C. fetus subsp. fetus* causes infertility and miscarriage in sheep, goats, and bovine. Additionally it is an opportunistic pathogen in humans, mostly immunocompromised and elderly patients. At first the bacterium causes gastroenteritis, and then dissemi...
nates throughout the blood and causes meningitis and other systemic infections via bacteremia.\(^2\)

We report a case of \textit{C. fetus subsp. fetus} bacteremia in a patient with myelodysplastic syndrome.

**CASE PRESENTATION**

A 78-year-old female patient with myelodysplastic syndrome was admitted to the emergency room due to chest pain intensifying with inspiration and difficulty in breathing. Physical examination was unremarkable except for diminished breath sounds and bilateral crackles over the lower lung fields by auscultation. Vital signs revealed a blood pressure of 110/70 mmHg, an apical heart rate of 78/minute, a respiratory rate of 16/minute, and a body temperature of 38.6 °C. Her leukocyte count was 30.7 K/µL (94% polymorphonuclear cells), erythrocytes 1.68×10\(^6\) K/µL, hemoglobin concentration 5.14 g/dL and platelet count 708 K/µL. Her C-reactive protein level was 379 mg/L, procalcitonin 8.5 ng/mL. Chest x-ray revealed bilateral patchy infiltrates intensifying at the right lower lobe segment. Oxygen saturation was 90%.

After a diagnosis of pneumonia had been made, blood, throat, and urine samples were obtained for microbiological tests. Sputum specimen could not be obtained. Then trimethoprim-sulfamethoxazole (800/160 mg) t.i.d, po and piperacillin-tazobactam (4.5 g) t.i.d, iv treatments were promptly initiated as empirical therapy. Four units of packed red cells were transfused. Recombinant erythropoietin was administered for treatment of MDS. Cytotoxic chemotherapy was not administered to the patient.

The patient recovered clinically, and her respiratory findings improved but she still had fever. While urine culture and throat swab yielded no pathogen, blood cultures obtained on admission were positive for spiral-shaped and thin Gram-negative bacilli with a typical view of \textit{Campylobacter} species. Upon receiving culture results of growth of \textit{C. fetus subsp. fetus} in two consecutive blood samples, clarithromycin (500 mg, b.i.d, p.o.) was added on the 8\(^{th}\) day of the present regimen. She denied any recent gastrointestinal disturbances like diarrhea but described an abdominal pain before her admission. The patient had a history of contact with bovine and sheep products. Her stool culture didn’t yield growth of \textit{C. fetus}. The patient responded the overall 21-day antimicrobial treatment and her fever, the C-reactive protein and procalcitonin levels declined day by day. Repeated blood cultures were negative. Her general condition improved and upon that she was discharged from the hospital.

**BACTERIOLOGICAL METHODS**

Blood samples were inoculated in aerobic blood culture vials (aerobic bacteriological media: BD BACTEC\textsuperscript{TM} Plus Aerobic/F Medium (Becton–Dickinson Diagnostic Systems, USA). The vials were monitored by the BACTEC\textsuperscript{TM} 9120 fully automated blood culture instrument (Becton–Dickinson Diagnostic Systems, USA). The vials with a positive signal were stained with Gram stain and aliquots of 0.1 mL were subcultured on Columbia agar with 5% sheep blood, eosin methylene blue (EMB) agar and Campylosel agar (bioMérieux Clinical Diagnostics/France). EMB and Columbia agar with 5% sheep blood were incubated aerobically, and Campylosel agar was micro-aerobically incubated (5% O\(_2\), 10% CO\(_2\) and 85% N\(_2\)) at 35°C. After 48 hours incubation, growth of tiny and sprawling gray colonies was seen. Spiral-shaped and thin Gram-negative bacilli were showed on Gram stain.

The identification of bacteria were performed by traditional methods (catalase, oxidase and motility tests; positive) VITEK® MS, an automatic microorganism identification system that uses mass spectrometry technology (Matrix-assisted laser desorption ionization time of flight, MALDI-TOF) (bioMérieux Clinical Diagnostics, France). The antibiotic susceptibility tests were performed by E-test method (PDM Epsilometer; AB Biodisc, Sweden) on Mueller-Hinton agar with 5% sheep blood. The minimum inhibitory concentrations (MICs) of the isolate were 0.38 µg/mL for ampicillin; 0.25 µg/mL for tetracycline; 0.75 µg/mL for erythromycin and gentamicin; 0.2 µg/mL for ciprofloxacin; 256 µg/mL for piperacillin-tazobactam; 0.015µg/mL for imipenem, and 24 µg/mL for ceftazidime. MICs were interpreted according to the breakpoints published by the Clinical Laboratory Standards Institute.\(^3\)

**DISCUSSION**

\textit{Campylobacter} is a significant pathogen of a wide range of diseases in humans. Most common cause of intestinal infections is \textit{C. jejuni}. Extra intestinal infections such as endocarditis, meningitis, osteomyelitis and septic arthritis are rare and caused mostly by other species.\(^3\)

\textit{Campylobacter} sp. colonizes various hosts from farm animals to humans with different degrees of virulence. Among them, \textit{C. fetus} strains are sep-
arated into two subspecies, *C. fetus subsp. venerealis* and *C. fetus subsp. fetus*. C. *fetus subsp. venerealis* causes venereal diseases in cattle and is rarely associated with human infection. On the other hand, *C. fetus subsp. fetus* infects bovine, sheep, goats, and humans. The microorganism may be delivered by an injury or mostly consumption of contaminated food or water.7

*C. fetus* is primarily related with bacteremia because of its tropism for vascular bed, seeding pre-existing lesions.8 Bacteremia caused by *C. fetus* is a severe infection frequently accompanied by recurrences and complications in spite of administration of proper antibacterial therapy without any delay.6 In immunocompromised patients, lung abscess, prosthesis hip joint infection, septic arthritis, vertebral osteomyelitis, spontaneous bacterial peritonitis, and cholecystitis are among the diverse complications of *C. fetus* bacteremia.7,9,12

*C. fetus* bacteremia has also been rarely reported in immunocompetent patients. Human immunodeficiency virus infection, primary immunodeficiencies, liver cirrhosis, alcoholism, senility, pregnancy, diabetes mellitus, haematological and solid organ malignancies or transplantation, systemic lupus erythematous, and splenectomy are significant predisposing factors for *C. fetus* infections.4,13

Our research has found a single case in Turkey. In this case, a 92-year-old male patient with secondary chronic renal failure due to pyelonephritis developed bacteremia possibly after a gastrointestinal infection caused by *C. fetus subsp. fetus* was reported.14

In our patient, myelodysplasia could have been contributing factor to the bacteremia. In our case, because any respiratory specimen could not be obtained, and stool culture didn’t yield growth of *C. fetus*, the source of primary infection side could not be identified. Because the patient’s respiratory distress responded to piperacillin-tazobactam therapy and *C. fetus* is resistant to piperacillin-tazobactam, it was not likely that the pneumonia resulted from *C. fetus* bacteremia. However, *C. fetus* bacteremia developed possibly after a gastrointestinal infection without diarrhea, because she described an abdominal pain before her admission. Unlike *C. jejuni*, *C. fetus* rarely causes diarrhea, but it is known to cause isolated bacteremia.15

A limitation of our study is the absence of the molecular analysis in the bacteriological identification. Correct identification needs molecular analysis, but we could not use molecular methods for a single case because of the high cost. Biochemical identification and MALDI-TOF is used to identify *C. fetus*.15

In conclusion, despite *C. fetus* infections are low, and case reports are rare in current literature, the actual incidence of *Campylobacter* infections has not been known due to its rare occasion and the specific growth conditions. For this reason, the possibility that infections can occur with these bacteria should not be overlooked in the clinical laboratory.

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