Genome analysis

JEPEGMIX: gene-level joint analysis of functional SNPs in cosmopolitan cohorts

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Abstract

Motivation: To increase detection power, gene-level analysis methods are used to aggregate weak signals. To greatly increase computational efficiency, most methods use as input summary statistics from genome-wide association studies (GWAS). Subsequently, gene statistics are constructed using linkage disequilibrium (LD) patterns from a relevant reference panel. However, all methods, including our own Joint Effect on Phenotype of eQTL/functional single nucleotide polymorphisms (SNPs) associated with a Gene (JEPEG), assume homogeneous panels, e.g. European. However, this renders these tools unsuitable for the analysis of large cosmopolitan cohorts.

Results: We propose a JEPEG extension, JEPEGMIX, which similar to one of our software tools, Direct Imputation of summary STatistics of unmeasured SNPs from MIXed ethnicity cohorts, is capable of estimating accurate LD patterns for cosmopolitan cohorts. JEPEGMIX uses this accurate LD estimates to (i) impute the summary statistics at unmeasured functional variants and (ii) test for the joint effect of all measured and imputed functional variants which are associated with a gene. We illustrate the performance of our tool by analyzing the GWAS meta-analysis summary statistics from the multi-ethnic Psychiatric Genomics Consortium Schizophrenia stage 2 cohort. This practical application supports the immune system being one of the main drivers of the process leading to schizophrenia.

Availability and implementation: Software, annotation database and examples are available at http://dleelab.github.io/jepegmix/.

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Supplementary information: Supplementary material is available at Bioinformatics online.

1 Introduction

Univariate analysis of genome-wide association studies (GWAS) has emerged as the main tool for identifying trait/disease-associated genetic variants (Burton et al., 2007). However, most variants reported by complex trait GWAS are common single nucleotide polymorphisms (SNPs) with weak or moderate effect sizes, which account for only a small fraction of the overall phenotypic variation (Manolio et al., 2009). This is due to the fact that, due to their small effect sizes, most common causal variants are unlikely to be detected in GWAS (Yang et al., 2010).

A reasonable approach to increase the power to detect true association signals with small effect sizes is to aggregate them by jointly analyzing multiple SNPs. To leverage information from multiple SNPs, multivariate association tests (Ehret et al., 2012; Wood et al., 2011; Yang et al., 2012) have been also proposed. However, these methods typically test all SNPs, regardless of their functionality.

Given that functional SNPs are likely to jointly impact on gene expression, to increase detection power, our group proposed JEPEG (Joint Effect on Phenotype of eQTL/functional SNPs associated with
a Gene; Lee et al., 2013b), which (i) uses only summary association statistics, (ii) imputes summary statistics of unmeasured functional SNPs and (iii) boosts detection power by jointly analyzing measured and imputed functional variants. However, similar to direct imputation methods based on summary statistics, e.g. DIST (Lee et al., 2013) and ImpG (Pasaniuc et al., 2014), it is only applicable to homogeneous cohorts. To overcome this limitation, concurrently with Adapt-Mix (Park et al., 2015) and DISSCO (Xu et al., 2015), our group developed DISTMIX (Direct Imputation of summary STratistics of unmeasured SNPs from MIXed ethnicity; Lee et al., 2015a). It extends DIST capabilities to the analysis of mixed ethnicity cohorts by estimating their linkage disequilibrium (LD) patterns as a mixture of the LD patterns from the constituent ethnicities of large reference panels, e.g. 1000 Genomes data (1KG) (Altshuler et al., 2010). Here, for the gene level analysis of the ever more common (and well powered) mixed ethnicity cohorts, we propose JEPEG for MIXed ethnicity cohorts (JEPEGMIX), which adapts the LD estimation strategy used by DISTMIX, while retaining all JEPEG advantages.

2 Methods

Similar to DISTMIX, to accurately estimate LD patterns for mixed ethnicity cohorts, JEPEGMIX first estimates the ethnic proportions of study cohorts using study allelic frequency (AF) information [see Supplementary Text S1 in supplementary data (SD) for details]. (Alternatively, when AF information is not available, user can precisely specify the proportions based on the ethnic composition information typically provided by published studies.) Next, using the estimated/user-specified ethnic proportions, the software estimates LD patterns of the study cohort as a weighted mixture of the LD matrices of all ethnic groups in a reference panel (Supplementary Text S2 of SD). Finally, it uses these estimated mixture LD patterns and association summary statistics to (i) when necessary, rapidly and accurately impute summary statistics of unmeasured functional SNPs (Supplementary Text S3 of SD) and (ii) jointly test the effect of measured and imputed functional SNPs associated with each gene (Supplementary Text S4 of SD).

3 Results

To estimate false positive rates, null hypothesis cosmopolitan cohorts were simulated using haplotype patterns from 1KG (see Supplementary Text S5 of SD). When compared with JEPEG, JEPEGMIX maintains the false positive rates at or below nominal thresholds (Supplementary Fig. S1 in SD). We obtained gene-level statistics by applying the method to association summary statistics from the large-scale cosmopolitan Psychiatric Genomics Consortium Schizophrenia stage 2 (PGC SCZ2) cohort (Schizophrenia Working Group of the Psychiatric Genomics Consortium, 2014). A subsequent Ingenuity Pathway Analysis (www.ingenuity.com) of the 61 significant JEPEGMIX genes (Supplementary Table S1 in SD), i.e. those with false discovery rate q-values < 0.05, yields a large number of immune pathways and only one (in italics) which is neurologically related (Table 1). The pattern is maintained even when excluding the 21 genes located in the immune related Major Histocompatibility (MHC) region from chromosome 6p (Supplementary Table S2).

4 Conclusions

For multi-ethnic cohorts, unlike existing methods, JEPEGMIX controls the Type I error rates at or below nominal levels. Due to ridge adjustment being inversely related to the size of 1KG relevant subpopulations (Supplementary Text S2 of SD), at present the method is rather conservative. However, the conservativeness is expected to become negligible with the advent of extremely large reference panels (http://www.haplo-type-reference-consortium.org). Thus, to the capabilities of JEPEG, JEPEGMIX adds the much needed applicability to the analysis of large cosmopolitan cohorts, which are the state-of-the-art in detecting genetic signals. For such cohorts, (i) it imputes unmeasured functional SNPs, (ii) pools in a synthetic variable the information of measured and imputed SNPs from the same functional category and (iii) combines these synthetic variables in a gene-level Mahalanobis test. JEPEGMIX application to PGC SCZ2 cohort suggests that, in the etiology of SCZ, the immune system might play a more substantial role than currently accepted.

| Pathway                                              | P-value |
|------------------------------------------------------|---------|
| Antigen presentation pathway                          | 0.0002  |
| Craft-versus-host disease signaling                   | 0.0004  |
| Autoimmune thyroid disease signaling                  | 0.0005  |
| Granulocyte A signaling                               | 0.002   |
| Dendritic cell maturation                             | 0.002   |
| Allograft rejection signaling                         | 0.002   |
| OX40 signaling pathway                                | 0.003   |
| Crosstalk between dendritic cells and natural killer cells | 0.003   |
| Communication between innate and adaptive immune cells | 0.003   |
| Cytotoxic T lymphocyte-mediated apoptosis              | 0.004   |
| Type 1 diabetes mellitus signaling                    | 0.005   |
| Role of RIG1-like receptors in antiviral innate immunity| 0.008   |
| Neuroprotective Role of THOP1 in Alzheimer’s Disease  | 0.009   |
| Nur77 signaling in T lymphocytes                      | 0.01    |
| Cdc42 signaling                                       | 0.01    |
| Calcium-induced T Lymphocyte apoptosis                | 0.02    |
| Caveolar-mediated endocytosis signaling               | 0.02    |
| CTLA4 signaling in cytotoxic T lymphocytes            | 0.03    |
| Virus entry via endocytic pathways                    | 0.03    |
| p53 Signaling                                         | 0.04    |
| G-protein coupled receptor signaling                  | 0.05    |

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