SYNTHESIS OF SILVER NANOPARTICLES FROM THE MEDICINAL PLANT BAUHINIA ACUMINATA AND BIOPHYTUM SENSITIVUM–A COMPARATIVE STUDY OF ITS BIOLOGICAL ACTIVITIES WITH PLANT EXTRACT

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INTRODUCTION

Nanoparticle synthesis from bio routed method is the growing field of bionanotechnology due to their low expensive, non-toxic nature. So this method of synthesis can be considered as economic and valuable alternative source for the synthesis of silver nanoparticles. Medicinal plants have been in use from ancient times, and their efficacy has been increasing day by day in the current world. Naturally, acquired compounds are considered as environmentally friendly and also more effective than the synthetic drug. These medicinal Plants represent a foundation for many pharmaceutical compounds since it is comprised of many secondary metabolites which have been used in the treatment of many human diseases [1]. The phytochemical constituents present in plant materials serve as both reducing and capping agent in silver nanoparticle synthesis. The green method of nanoparticle synthesis targets mostly on silver nanoparticles as they are striving towards the edge level utilities in every aspect of science and technology including the medical fields. Colloidal silver is a current area of interest due to its distinctive properties viz. good conductivity, catalytic activity, chemical stability and antimicrobial activity [2]. The major benefit of using plant extracts for the synthesis of silver nanoparticles is that they are readily available, safe, and nontoxic and have a wide range of phytochemicals that can assist in the reduction of silver ions [3]. Silver compounds and its salts have been used as antimicrobial agents and wound care products since time immemorial [4]. Silver nanoparticles (AgNPs) have been widely studied and proved as an effective antimicrobial agent. The mechanism is similar as the metallic silver, but it is more effective as an antimicrobial agent than other silver compounds. Silver nanoparticles and silver ions released by silver nanoparticles interacts with the biomolecules contain sulphur (proteins) and phosphorus (DNA) which interrupts cellular metabolism. Recent studies highlighted that silver nanoparticles are able to generate reactive oxygen species (ROS) results in microbial membrane damage [5]. The current work emphasis on the green routed silver nanoparticle synthesis using two medicinal plants Bauhinia acuminata, Biophytm sensitivum and comparing it's biological activities with plant extracts.

MATERIALS AND METHODS

Chemicals

Ferric chloride, Hydrochloric acid, Sodium hydroxide, Sulphuric acid, Chloroform, Fehling’s A, Fehling’s B, Million’s reagent, 1,1-Diphenyl-2-picryl hydrosul (DPPH), DMSO, Nutrient agar, Nutrient broth, Brain heart infusion agar, Potato dextrose agar, Muller hinton agar were obtained from Hi-Media Pvt Ltd. All the chemicals purchased were of analytical grade.

Collection of plant material

The leaves of Bauhinia acuminata and Biophytm sensitivum were collected from Kannur (11.8698°N, 75.3555°E) and Alappuzha (9.49N, 76.33 E) districts of kerala, India.

Preparation of leaf extracts

The leaves of Bauhinia acuminata and Biophytm sensitivum were collected and washed thoroughly using distilled water and cut into small pieces. The leaves were shade dried for 1 w. Dried leaves were uniformly ground using a mechanical grinder to make a fine powder. The powder was serially extracted in methanol and distilled water (10%w/v) using a soxhlet apparatus. These extracts were concentrated at 40 C under reduced pressure (72mbar) with a rotary evaporator and dried using lyophilizer. Dried extracts was collected and stored in airtight container at 4 C for further use [6].
Nanoparticle synthesis
Preparation of plant extracts for synthesis of nanomaterial
Finely ground powder (10 mg) of plant material was mixed with 100 ml of solvents (milliQ water and methanol) and kept for incubation in a rotary shaker for 24-48 h [7].

Biosynthesis of silver nanoparticles
Biosynthesis of silver nanoparticles was followed by the method described by Pannerselvam et al. [8] with some modifications. 95 ml of 0.1 mmol of aqueous Silver nitrate (AgNO₃) was prepared in a conical flask, to which 5 ml of each plant extracts were added, and the flakes were covered with aluminium foil to prevent photoreaction of silver nitrate. The reaction mix showed colour change after 30 min, still for better synthesis the mixture was kept for incubation at room temperature for 24 h. After incubation, nanoparticle synthesis was confirmed using UV-Visible spectroscopy and the mixture was centrifuged at 5000 rpm for 20 min. The resultant pellets were collected and washed thrice in distilled water and centrifuged to remove unbound biomass residues for better Ag nanoparticles for further characterization studies. The silver nanoparticles were dried and stored.

Phytochemical screening of plant extracts
Phytochemical screening of the leaf extracts of Bauhinia acuminata and Biophytum sensitivum was carried out by following the standard protocol described by JB Harborne [9].

Antioxidant assay-DPPH radical scavenging activity of plant extracts and silver nanoparticles
The ability of the leaf extracts to obliterate the DPPH radical ([1, 1-diphenyl-2-picryl hydrazyl] was performed by the method described by Blois et al. 1958. A stock solution of leaf extracts was maintained at a concentration of 1 mg/ml. From the stock solution different concentrations viz; 100, 200, 300 of plant extracts were prepared and made it to 1 ml. To this solution, 1 ml of DPPH solution dissolved in DMSO was mixed, and the reaction mixture was incubated in the dark for 30 mins at room temperature and the absorbance was recorded at 517 nm. Ascorbic acid was used as a standard. DMSO was used as blank. DMSO and DPPH mixture was maintained as control [10]. The same procedure was followed for the radical scavenging activity of silver nanoparticles.

The percentage DPPH, the radical scavenging activity of each extract, was done using the following formula:

\[
\text{DPPH radical scavenging activity} = \left( \frac{Ac-At}{Ac} \right) \times 100
\]

Where, Ac is the absorbance of blank (distilled water) and At is the absorbance of test sample.

Hemolytic assay of plant extract and silver nanoparticles
Preparation of erythrocytes suspension
Five millilitres of blood was collected from a healthy individual in a centrifuge tube containing EDTA (Ethylene diamine tetra acetic acid). The collected blood was then centrifuged at 1500 rpm for 3-5 min. Plasma (supernatant) was discarded, and the resultant pellet was washed thrice with sterile phosphate buffer saline of pH 7.2 by centrifugation at 1500 rpm for 5 min. The cells were then resuspended in 0.5% of normal saline [11].

Hemolytic activity
In vitro hemolytic activity was performed by spectrophotometer method [11]. Plant extracts were maintained in a stock solution of 1 mg/ml in sterile phosphate buffer. Different concentrations (25, 50, 75 and 100 µg/ml) of extracts were made from the stock solution. A volume of 1 ml of the previously treated cell suspension was added into 1 ml of different concentration of plant extracts. The reaction mixtures were incubated at 37°C for 30 min. The incubated mixtures were again subjected to centrifugation at 1500 rpm for 10 min in a laboratory centrifuge. The free haemoglobin in the supernatant was measured in a UV-Visible spectrometer at 540 nm. Phosphate buffer and distilled water were maintained as a positive control and blank. The percentage hemolysis of each extract was calculated according to the following formula:

\[
\text{Hemolysis}\% = \left( \frac{At - An}{Ac - An} \right) \times 100
\]

Where, At is the absorbance of test sample
Ac is the absorbance of blank (distilled water) and An is the absorbance of the positive control (phosphate buffer saline).

Antimicrobial activity of plant extracts and silver nanoparticles
Bacterial strains such as Bacillus cereus (MTCC 1272), Listeria monocytogenes (MTCC 657) were obtained from Microbial Type Culture Collection, Chandigarh, India. Staphylococcus aureus and Escherichia coli were obtained from Biosciences laboratory of VIT University, Tamilnadu, India. The fungal culture includes Rhizopus spp. which was isolated from spoiled food. All the cultures were maintained on their specific growth medium.

Agar well diffusion assay
Antimicrobial activity was performed by agar well diffusion method [11]. The 24 h fresh cultures of microorganisms were standardised using Mc-Farland turbidity assay. The culture organisms were swabbed on Muller Hinton agar (for bacteria) and potato dextrose agar (for fungal) plates. Wells were bored using sterile cork borer and 20 µl of each plant extracts of 1 mg/ml concentration was pipetted out into each well. Distilled water was used as negative control. The plates were incubated at 37°C for 24 h. After incubation plates were checked for the zone of inhibition. This assay was done for plant extracts as well as silver nanoparticles.

UV-visible spectroscopy characterization of silver nanoparticles
The reduction of pure AgNPs was recorded under UV-Visible spectroscopy using Shimadzu UV1800 UV-Visible spectrometer between an absorbance ranges from 300-800 nm. The UV-Visible Spectra of the plant leaf extract and silver nitrate solution were also recorded.

Characterization of plant extracts and silver nanoparticles
GC-MS analysis of plant extracts
Bioactive methanol extracts of Bauhinia acuminata and Biophytum sensitivum was analysed for the presence of various compounds using the Gas chromatography-mass spectroscopy instrument. Helium was used as a carrier gas at 0.1 ml/min flow rate. The temperature of the column was programmed at 600 °C for 2 min ramp 106 °C/min to 3000 °C with a holding of 6 min. The injector temperature was maintained at 2500 °C for the analysis. The resulting peaks were identified on the basis of existing mass spectra data correlation provided from the Wiley 8. LIB and NIST08. LIB library spectrum provided by the software in GC-MS system [11].

FTIR analysis of plant extracts and silver nanoparticles
The dried plant extracts, as well as nanoparticles, were analysed by attenuated total reflectance FTIR (ATR-FTIR) method using Shimadzu at an infrared range of 400-4000 cm⁻¹.

SEM analysis of silver nanoparticles
The resulted dried powder of silver nanoparticles by green synthesis was subjected for SEM analysis, to determine the particle size of formed silver nanoparticles. The samples were sputter coated using 60% of gold and 40% of palladium and the images of nanoparticles were studied under SEM using Zeiss (EVO18) [12].

RESULTS
Phytochemical screening of plant extracts
The phytochemical screening of the four plant extracts showed the presence of carbohydrates, flavonoids, saponins, tannins, terpenoids, phenols and proteins. The result of the phytochemical analysis is shown in table 1.
Table 1: Qualitative analysis of phytochemical components of B. acuminata and B. sensitivum leaf extracts

| Tests for  | B. acuminata (Methanol extract) | B. acuminata (Aqueous extract) | B. sensitivum (Methanol extract) | B. sensitivum (Aqueous extract) |
|------------|---------------------------------|---------------------------------|---------------------------------|---------------------------------|
| Carbohydrates | +ve                             | +ve                             | +ve                             | +ve                             |
| Flavonoids  | +ve                             | +ve                             | +ve                             | +ve                             |
| Phenols     | -ve                             | +ve                             | -ve                             | +ve                             |
| Proteins    | -ve                             | +ve                             | -ve                             | -ve                             |
| Saponins    | +ve                             | +ve                             | +ve                             | +ve                             |
| Tannins     | +ve                             | +ve                             | -ve                             | -ve                             |
| Terpenoids  | +ve                             | +ve                             | +ve                             | -ve                             |

Nanoparticle synthesis
The primary confirmation of silver nanoparticle formation was observed by the colour change in silver nitrate solution (0.1 mmol) on the addition of plant extracts. Test samples were compared with the standard, 0.1 mmol silver nitrate solution i.e, white in colour. The aqueous extract and methanol extract of Bauhinia acuminata and Biophytum sensitivum showed the excellent result. Colour change was observed in the range of gold colour to light brown colour (methanol extract) and gold colour to dark brown colour (aqueous extract).

Antioxidant assay-DPPH radical scavenging activity plant extracts and silver nanoparticles
The DPPH scavenging activity of antioxidant compounds are due to their hydrogen donating ability. The DPPH quenching activity of aqueous and methanol extracts of the plant and as well as nanoparticles was evaluated and compared with that of the standard ascorbic acid. The methanolic leaf extract of Bauhinia acuminata showed high DPPH scavenging activity of 90% (IC50 value 4.45 µg/ml). The methanolic leaf extract of Bauhinia acuminata (BAMP) exhibited maximum radical scavenging activity.

![Fig. 1: DPPH assay of Bauhinia acuminata and Biophytum sensitivum extract and NP extracts](image1)

Hemolytic assay of plant extracts and silver nanoparticles
Haemolytic activity of leaf extracts of Bauhinia acuminata and Biophytum sensitivum and the nanoparticle (NP) extracts were performed to observe its toxic effects on human erythrocytes. The extracts showed low haemolytic activity. The percentage hemolysis was found in the range of 3%-6.7%. Thus, the extracts are found to be safe for human use.

![Fig. 2: Hemolysis assay of Bauhinia acuminata and Biophytum sensitivum leaf extract and NP extract](image2)
Antimicrobial activity of plant extracts and silver nanoparticles

The anti-microbial activity was assessed using Kirby-Bauer method. The antimicrobial activity of methanolic as well as aqueous extracts of plant and silver nanoparticles were checked against some of the food pathogens like Bacillus cereus, Escherichia coli, Listeria monocytogenes, Staphylococcus aureus and Rhizopusps. Extract dose of 1 mg/ml of Bauhinia acuminata methanolic leaf extract showed excellent activity towards Bacillus cereus (inhibition zone 13 mm). The methanolic leaf extract, as well as the silver nanoparticle of Biophytum sensitivum, showed the best activity towards Listeria monocytogenes (inhibition zone 13 mm, 12 mm). Other extracts showed moderate antagonistic activity. The leaf extract, as well as the silver nanoparticle, showed very weak activity towards the fungal strain.

UV-visible spectroscopy characterization of silver nanoparticles

UV-Visible absorption spectra of the silver nanoparticles in solution were recorded. The UV-Visible spectra of plant-mediated (Bauhinia acuminata and Biophytum sensitivum) silver nanoparticles showed the excellent result. High peak of absorbance was observed between wavelengths of 400-460 nm. The uv-visible absorption spectrum of the silver nanoparticles in solution is shown in fig. 4.

Structural analysis of plant extracts and silver nanoparticles

GC-MS analysis

The methanolic leaf extracts of the plants, Bauhinia acuminata and Biophytum sensitivum were undergone GC-MS analysis and showed the presence of DL-Alpha-Tocopherol and Alpha-Tocopherol-Beta-D-Mannoside. All these compounds are proven to have excellent antioxidant, anti-cancer activities. The results are depicted in fig. 5.

FTIR (Fourier Transform Infrared Spectroscopy)

FTIR analysis was done for the characterization of the resulted leaf extracts and nanoparticle extracts of Biophytum sensitivum as it showed high activity. The FTIR absorption spectra of Biophytum sensitivum extracts of the plant as well as nanoparticles were found to be in a range of 981.77 cm⁻¹–3317.56⁻¹. These absorption bands correspond to various functional groups of amides, aromatic rings, nitro groups, esters, alkenes, aldehydes, amines, etc. The amide, carboxyl, hydroxyl, aromatic groups i.e, the phenols, saponins, terpenoids found in the plant extracts are responsible for the reduction and capping during the silver nanoparticle synthesis. The FTIR results of the plant and nanoparticle extracts are shown in table 2.

SEM analysis

The SEM image is employed to predict the size and morphology of resultant silver nanoparticles. It was observed that most of the nanoparticles are spherical in shape with an average diameter of 2 micrometers (µm).
Fig. 5: GC-MS analysis of *Bauhinia acuminata* and *Biophytum sensitivum* methanol leaf extract

Table 2: FTIR spectral peaks and its functional groups and vibrations

| Extracts | Absorption   | Functional group | Type of vibration |
|----------|--------------|------------------|-------------------|
| BSWP     | 3304.06      | Amides           | N-H stretch       |
|          | 1581.63      | Aromatic rings   | C=C=C-C stretch  |
|          | 1382.96      | Nitro groups     | N=O bend         |
|          | 1020.34      | Esters           | C-O stretch      |
|          | 3234.62      | Carboxylic acids | O-H stretch      |
| BSMP     | 3186.40      | Alkenes          | Triple bond C-H  |
|          | 2924.09      | Alkanes          | C-H stretch      |
|          | 2852.72      | Carboxylic acid  | O-H stretch      |
|          | 1604.77      | Alkenes          | C=C=C stretch    |
|          | 1442.75      | Alkanes          | H-C-H bend       |
|          | 1377.17      | Alkanes          | CH-CH bend       |
|          | 1265.30      | Alkyl halides    | C-F stretch      |
|          | 1043.49      | Alcohol          | C-O stretch      |
| BSW      | 1722.43      | Aldehydes        | C-O stretch      |
|          | 1620.21      | Alkenes          | C=C=C stretch    |
|          | 1028.06      | Ethers           | =C-O=C-C-O stretch|
| BSM      | 3317.56      | Alkynes          | Triple bond C-H bend |
|          | 2916.37      | Alkanes          | H-C-H symmetric  |
|          | 2860.79      | Carboxylic acid  | And assymmetric stretch |
|          | 1722.43      | Aldehydes        | O-H stretch      |
|          | 1598.99      | Amides           | C=O stretch      |
|          | 1371.39      | Alkane           | N-H bend         |
|          | 1253.73      | Alkyl halides    | CH-CH bend       |
|          | 981.77       | Alkenes          | C-F stretch      |

Note: BAMP—*Bauhinia acuminata* methanolic leaf extract, BSMP—*Biophytum sensitivum* methanolic leaf extract, BAM—*Bauhinia acuminata* derived silver nanoparticles, BSM—*Biophytumsensitivum* derived silver nanoparticles.

Fig. 6: FTIR spectra of *Biophytum sensitivum* plant extract (aqueous extract) and nanoparticles
Fig. 7: FTIR spectra of Biophytum sensitivum plant extract (methanol extract) and nanoparticles

Fig. 8: SEM image of Bauhinia acuminata aqueous extract

Fig. 9: SEM image of Bauhinia acuminata methanol extract

Fig. 10: SEM image of Biophytum sensitivum aqueous extract
DISCUSSION

Plant-based compounds have a pharmacological activity for the prevention and treatment of complex diseases. Therefore, the researcher provides special attention to the plant derived metabolites. The antimicrobial, anticancer and antioxidant properties of the medicinal plants depend on the presence of its secondary metabolites viz., phenolic acids, flavonoids, tannins, coumarins, lignins, quinones, stilbenes and curcuminoïds [13].

The secondary metabolites of plants are used as antioxidants, anti-blood clotted agents, food additives, in the preparation of cosmetics, etc. The result of the present study showed that the aqueous extracts of Biophytum sensitivum have showed the presence of flavonoids and carbohydrates. The results of the present study coincided with previous observations and supported the pharmacological evidence. There is a contradictory in the case of Bauhinia acuminata, the present study shows the presence of flavonoids in methanolic leaf extract and presence of terpenoids in both aqueous as well as in methanolic extract [14].

In the present study, the methanol extract of Bauhinia acuminata showed high DPPH scavenging activity of 90% whereas the DPPH scavenging percentage in a previous study was found to be 84% [15]. Kalia et al. observed the maximum DPPH scavenging activity of 43.96% in Biophytum leaf extract, while in the present study we observed the inhibition percentage of 76% [16]. Hemolytic activity of plant extracts in the present study showed less haemolytic activity.

The structural analysis of plant extracts using GC-MS revealed the presence of pharmacologically important compounds like tocopherols, phenols, diethyl halite, phytol, etc. which has proven presence of pharmacologically important compounds like Di-alpha tocopherol, which is proven to have a wide range of antioxidant, anti-cancer, anti-inflammatory activities. They are also used in food industry, cosmetic industry, etc. The most important significance of the present work is the synthesis of silver nanoparticles from these less explored medicinal plants. This current investigation is the first reported case of silver nanoparticles from Bauhinia acuminata and Biophytum sensitivum.

CONCLUSION

The present investigations suggest that the plant extracts, as well as the silver nanoparticles, showed a good result of antioxidant activity, haemolytic activity. It was found that the high rates of phytochemicals are widely distributed in Bauhinia acuminata and Biophytum sensitivum. The plant extracts exhibited the presence of flavonoids, saponins, carbohydrates, tannins, terpenoids, etc. These compounds are the sources that render pharmacological activities to the plant. The structural analysis yielded pharmacologically important compounds like Di-alpha tocopherol, which is proven to have a wide range of antioxidant, anti-cancer, anti-inflammatory activities. They are also used in food industry, cosmetic industry, etc. The most important significance of the present work is the synthesis of silver nanoparticles from these less explored medicinal plants. This current investigation is the first reported case of silver nanoparticles from Bauhinia acuminata and Biophytum sensitivum.

These plants can be exploited further to undergo more research. The nanoparticle synthesises using Bauhinia acuminata and Biophytum sensitivum can be widened to get better yield in synthesis and activity. In future, the silver nanoparticle from these plants can be used as potent therapeutics and can be further exploited for their other beneficial effects in the pharmacological studies. Thus from this current study, we can conclude that the plant extracts can be used as potent therapeutics than silver nanoparticles.

CONFLICT OF INTERESTS

Declared none

REFERENCES

1. Akhila Sravya Dantu, Shankarguru P, Ramya devi D, Vedha Hari BN. Evaluation of in vitro anticancer activity of hydroalcoholic extract of Tabernaemontana divaricata. Asian J Pharm Clin Res 2012;5:59-61.
2. Ashokkumar S, Ravi S, Kathiravan V, Velumungan S. Synthesis of silver nanoparticles using A. indicum leaf extract and their antibacterial activity. Spectrochim Acta Part A 2015;134:34-9.
3. Kanchana R, Pranjita Zantye. Plant-mediated synthesis of silver nanoparticles with diverse applications. Asian J Pharm Clin Res 2016;9:159-63.
4. Khandelwal N, Kaur G, Kumar N, Tiwari A. Application of silver nanoparticles in viral inhibition: a new hope for antivirals. Dig J Nanomater Biostruct 2014;9:175.
5. Park S, Park HH, Kim SY, Kim SJ, Woo K, Ko G. Antiviral properties of silver nanoparticles on a magnetic hybrid colloid. Appl Environ Microbiol 2014;80:2343-50.
6. Anju Krishna SR, Hafza S, Poorna Chandrika G, Lekhya Priya C, Bhaskara Rao KV. Pharmacological properties, phytochemical and GC-MS analysis of Bauhinia acuminata Linn. J Chem Pharm Res 2015;7:372-80.
7. Arokiya FAA, Arumugam R, Saravanan S, Anantharaman P. Photofabrication of gold nanoparticles assisted by leaves of *Suaedamonica* and its free radical scavenging property. J Photochem Photobiol B: Biol 2014;135:75-80.

8. Pannerselvam, Balashanmugam, Pudupalayam T, Kalaichelvan. Biosynthesis characterization of silver nanoparticles using *Cassia roxburghii* DC aqueous extract and coated on cotton cloth for effective antibacterial activity. Int J Nanomed 2015;10:87-97.

9. Harbone JB. Phytochemical methods, a guide to modern techniques of plant analysis. Chapman and Hall, London; 1973.

10. Bloos MS. Antioxidant determination by the use of a stable free radical. Nature 1958;29:1199-200.

11. Priya CL, Gaurav K, Bhaskara RKV. Antioxidant activity of *Achyranthes aspera* Linn stems extracts. Pharmacologyonline 2010;2:228-37.

12. Krishnaraj C, Jagan EC, Rajasekar S, Selvakumar P, Kalaichelvan PT, Mohan N. Synthesis of silver nanoparticles using *Acalypha indica* leaf extract and its antibacterial activity against water borne pathogens. Colloid Surf B: Biointer 2010;76:50-6.

13. Cai Y, Luo Q, Sun M, Corke H. Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. Life Sci 2004;74:2175-84.

14. Anjukrishna SR, Hafza S, Poorna CG, Lekhya PC, Bhaskara RKV. Pharmacological properties, phytochemical and GC-MS analysis of *Bauhinia acuminata* Linn. J Chem Pharm Res 2015;7:372-80.

15. Anjukrishna SR, Hafza S, Poorna CG, Lekhya PC, Bhaskara RKV. Pharmacological properties, phytochemical and GC-MS analysis of *Bauhinia acuminata* Linn. J Chem Pharm Res 2015;7:372-80.

16. Venkata NK, Tadi R, Sudhakara RP, Sandeep BV. Assessment of phytoconstituents composition and anti-haemolytic activity of *Psidium guajava* leaf extract against H2O2 induced hemolysis in chicken erythrocytes. Int J Pharm Sci Res 2016;1:27-30.

17. Charalampos P, Michael. Analysis of naturally occurring phenolic compounds in aromatic plants by RP-HPLC coupled to diode array detector (DAD) and GC-MS after Silylation. Foods. Foods 2013;2:90-9.

18. Anjukrishna SR, Hafza S, Poorna CG, Lekhya PC, Bhaskara RKV. Pharmacological properties, phytochemical and GC-MS analysis of *Bauhinia acuminata* Linn. J Chem Pharm Res 2015;7:372-80.

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