Docking predictions based *Plasmodium falciparum* phosphoethanolamine methyltransferase inhibitor identification and in-vitro antimalarial activity analysis

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**Abstract**

The increased multidrug resistance among antimalarial drugs produces the urgency of potent anti malarial to combat resistant malaria and the malaria burden worldwide. The protein which may prevent the growth or transmission of malaria parasite may be the great target for rational drug designing. *Plasmodium falciparum* phosphoethanolamine methyltransferase (Pfpmt) absent in human catalyzes triple methylation of ethanolamine into phosphocholine for phosphatidylcholine biosynthesis from serine decarboxylation phosphoethanolamine methyltransferase pathway for the membrane development at asexual as well as sexual stages of parasite. The *Plasmodium* requires production of membrane rapidly for growth and multiplication. Hence, the phosphoethanolamine methyltransferase of *Plasmodium falciparum* was selected as drug target for rational drug designing. Using Discovery studio 3.5 software the library of zinc compounds was screened against target and analyzed. The compounds with better druglike properties and docking affinity and binding interaction for target protein were procured for in vitro analysis against *Plasmodium falciparum* culture (IC₅₀). Compounds ZINC02103914 and ZINC12882412 were found to have good druglike properties and affinity for Pfpmt also inhibited *P. falciparum* growth at very low µM IC₅₀ concentration 3.0 µM and 2.1 µM respectively also found nontoxic in vitro against HEK-293 cells. Simulation study of best inhibitor revealed the specificity for the target protein. Hence, the compounds possessed the immense probability of being inhibitors of Pfpmt and may be optimized as antimalarial agent for combinational therapy to overcome the multidrug resistance and may also be used as template for optimization and rational drug designing.

**Keywords:** Phosphatidylethanolamine, Pfpmt, ADMET, Docking, Invitro, HEK-293

**Background**

Malaria is a mosquito borne disease in humans caused by unicellular microorganism called parasitic *Plasmodium* protozoa. According to the WHO, million new malaria cases were found and people were killed by malaria globally [1].

Malaria treatment recommended by WHO, must be followed by government as well private health organizations. Once the suspected cases are diagnosed with malaria, the treatment should be started according to the WHO guidelines for *Plasmodium falciparum*, *vivax* and others. Uncomplicated *P. falciparum* malaria treatment should be started with artemisinin combination therapy (Table 1). Since *Plasmodium falciparum* malaria can more be complicated than *P. vivax* infection, so extra attention is needed while observing the patient and should be treated as per drug policy. Patients with severe malaria should be given artemesunate through intravenous or intramuscular route for at least 24 h or until patient become in condition to take oral medication. After 24 h
and patient can tolerate medication orally, artemisinin combination therapy (ACT) treatment should be given for 3 days, (primaquine dose can be added in case of low transmission). Artemether should be preferred over quinine if artesunate (intravenous and intramuscular) is not available. In case of pre referral treatment when complete treatment of severe malaria is not available, in that case patient (children/adult) should be given Intramuscular artesunate and refer for complete treatment. But when both injection of artesunate and complete treatment are not available, in that case rectal dose of artesunate should be given to children < 6 years (avoid in older children and adults) and refer for complete treatment. *Plasmodium ovale* and *malariae* are rarely found in India, however *P. ovale* and *P. malariae* infection should be treated as *P. vivax* and *P. falciparum* [2, 3] respectively as given in Table 1. Since, the resistance against chloroquine and artemisinin has increased so *Plasmodium falciparum* infection cases should be treated with ACT because it is combination of drugs with different mechanism of action to block parasite development.

As the world is facing problem of antimalarial drug resistance is a greatest challenge today to stop the spread of malaria to new areas and re-emergence of malaria [4]. Hence, there is need for new drug target to develop potential inhibitors against the disease. So, a novel drug target *Plasmodium falciparum* phosphoethanolamine methyltransferase gene that is devoid in humans was selected to screen potent antimalarial drugs. Phosphatidylcholine is most frequently needed phospholipid for the survival of the parasites. *Plasmodium* synthesizes phosphatidylcholine abundantly from serine decarboxylation phosphoethanolamine methyltransferase pathway (SDPM) for production of new membranes at very fast rate for rapid multiplication during not only intraerythrocytic cycle (with maximum expression at trophozoite stage) but also during gametocyte development [5].

Loss of *Pf*pmt could severely distort the multiplication and gametocytes formation but reintroduction of *Pf*pmt effects were restored to wild type. *Pf*pmt has one catalytic domain for three methylation steps this implies that same catalytic domain is used for all three methylations (Fig. 1). Therefore, the inhibition of catalytic domain of *Pf*pmt may lead to complete inhibition of SDPM pathway [6]. Hence, the *Pf*pmt is a crucial target for structure based drug designing.

Few compounds such as sinefugin (AdoMet analog), miltefosine (Choline analog) amodiaquine (4-aminquinoline) had been identified to inhibit the PMTs of *Plasmodium* and nematode [7–10]. Miltefosine with

| Table 1 Current treatment of *Plasmodium* malaria infection |
|--------------------------------------------------------------|
| **P. falciparum** Uncomplicated | **First-line treatment** |
| *Uncomplicated* | Artesunate + sulfadoxine pyrimethamine |
| 50 mg or 100 mg of artesunate and tablets containing 500 mg of sulfadoxine + 25 mg of pyrimethamine |
| Artesunate + mefloquine/amodiaquine |
| Recommended if the use of sulfadoxine–pyrimethamine (SP) is contraindicated. Artesunate 15–20 mg/kg and mefloquine 8 (5–11) mg/kg/dose daily for 3 days |
| Primaquine (0.25 mg base/kg body weight, maximum dose 15 mg) is recommended for uncomplicated *P. falciparum* malaria as gametocytocidal |
| Second-line treatment |
| Artemether–lumefantrine |
| Available as tablets coartem (20 mg artemether and 120 mg lumefantrine) twice daily for 3 days |
| Primaquine (0.25 mg base/kg body weight, maximum dose 15 mg) is recommended for uncomplicated falciparum malaria as gametocytocidal |
| Third line treatment |
| In case of contraindication with first line and second line the third line treatment is recommended as Atovaquone 250 mg/proguanil 100 mg (malarone) tablets daily for 3 days |
| Quinine + doxycycline for 7 days |
| Quinine and clindamycin is the preferred for pregnant women during first trimester |
| **P. falciparum** (Special risk groups) Infants (< 5 kg body weight): ACT (mg/kg) as given to the children of 5 kg body weight |
| Malaria co-infected with HIV: avoid artesunate + sulfadoxine pyrimethamine if patient is on clotrimazole |
| Avoid artesunate + amodiaquine if patient is on efavirenz or zidovudine |
| *P. falciparum* Severe malaria | *Artesunate through intravenous (IV) or intramuscular (IM) route for at least 24 h or until patient become in condition to take oral medication. After 24 h ACT treatment for 3 days (primaquine can be added) In absence of IV/IM artesunate, artemether should be preferred* |
| *P. vivax* | *P. vivax*: chloroquine 25 mg for 3 days, in combination with primaquine 0.25 mg/kg and 15 mg for adults for 14 days in case of chloroquine-sensitive *P. vivax* malaria. Other ACTs should be combined with primaquine for chloroquine-resistant |
| *P. ovale* | *P. ovale*: treatment recommended for *P. ovale* malaria infection is same as for *P. vivax* malaria |
| *P. malariae and knowlesi* | *P. malariae* and *P. knowlesi*: infection is recommended for chloroquine like *P. vivax* malaria. No radical cure is required since, no hypnozoites are formed in infection |
Singh et al. BMC Chemistry (2019) 13:43

antitumor and antileishmanial activities also found to arrest the function of Pfpm at higher concentration in radioactivity-based assay [11–13]. It was also observed that aminoquinolines and amino alcohols such as quinacrine, quinidine and quinine had no Pfpm inhibitory activity at concentrations up to 200 μM [14]. Among the antimalarials chloroquine and primaquine were also found to inhibit Pfpmt in millimolar [7]. Eleven compounds from National Cancer Institute Open Chemical Repository were screened as Pfpm inhibitor based on Pfpm-specific enzyme-coupled spectrophotometric assay. These 11 compounds inhibited both Pfpmt and gametocytes. Compounds NSC-323241 (3-azabicyclo[3.2.2.][3.2.2.]nonane-3-carboxenic acid, [1-(2-pyridinyl) ethylidene]hydrazide) and NSC-158011 (N-naphthalen-1-yl-2-phenylsulfanylethanolthioamid) found effective against early-stage gametocyte development where NSC-158011 found to inhibit the protein competitively [13] (Table 2). In this paper work we used the binding site of NSC-158011 (phosphoethanolamine/phosphocholine binding site) for screening the compound library to find the compounds with novel scaffold with better antimalarial activity.

We carried out study for finding specific inhibitors of Pfpmt based on docking studies and ADMET analysis. The Natural compounds of ZINC database were shortlisted based on the Lipinski rule of five, ADMET and docking score. The final seven compounds, which were not only interacting with crucial amino acids but

Table 2 Reported inhibitors of Pfpmt

| S. no. | Compound | (µM) PIPMT activity IC50 | (µM) asexual proliferation IC50 |
|-------|----------|--------------------------|--------------------------------|
| 1     | NC 39225 | 2.4±1.5                  | 5.6                            |
| 2     | NC 22225 | 3.1±2.0                  | 0.8±1.1                        |
| 3     | NC 109268| 3.3±1.3                  | 2.7                            |
| 4     | NC 310551| 3.5±1.6                  | 3.8±1.8                        |
| 5     | NC 348401| 3.9±0.3                  | 5.2±0.2                        |
| 6     | NC 641296| 3.8±0.7                  | 3.6±1.2                        |
| 7     | NC 125034| 4.3±2.7                  | 4.7                            |
| 8     | NC 668394| 7.7±4.8                  | 1.3±0.4                        |
| 9     | NC 150080| 8.5±4.9                  | 4.8                            |
| 10    | NC 158001| 9.2±3.5                  | 5.2                            |
| 11    | NC 323241| 9.5±3.4                  | 1.1±0.2                        |

Fig. 1 SDPM pathway a different route for PC in P. falciparum in comparison with human predominant PC synthesis. Plant-like reactions are indicated as in dotted circle. DAG diacylglycerol, CDP cytidine diphosphate, PC phosphatidylcholine.
also occupying the catalytic dyad important for triple methylation were procured for invitro analysis. Two compounds ZINC02103914 and ZINC12882412 showed good parasite inhibition at 3.0 µM and 2.1 µM IC₅₀ respectively. The Molecular dynamics (MD) simulation of best inhibitor ZINC12882412 did using Desmond v4.2 also confirmed affinity and specificity of the inhibitor towards the Pfpm. These compounds were also found nontoxic to the HEK-293 cells. Hence these compounds may further be optimized as antimalarial and may be used as template for further structure and ligand based drug designing.

Results and discussion

ADMET analysis of compound library

The ZINC natural compounds library before docking was subjected for computational (absorption, distribution, metabolism, excretion and toxicity) ADMET analysis. Compounds procured were fulfilling the rule of five for oral absorption. Solubility of compounds was analyzed based on solubility where levels 3 and 4 stated the good and optimum solubility. The compounds given in Table 3 were having the good solubility profile.

Docking analysis of compounds and schizonticidal activity

Compounds with good ADMET properties were selected for docking analysis. In order to find better and more thermodynamically stable compounds, the compounds with docking score and binding energy more than the pCholine docking score (51.8 kcal/mol) and binding energy (−89.5 kcal/mol) were selected. The compounds were docked with Pfpm and the top twenty compounds with dock score and binding energy more than 60 kcal/mol and −90.2 kcal/mol respectively were selected for

Table 3 In silico ADME and physicochemical parameters analysis of selected compounds are shown

| S. no. | Structure and zinc id         | RV | S   | S   | A   | Al | P    |
|--------|------------------------------|----|-----|-----|-----|----|------|
| 1      | ![Structure 1](image1)       | 0  | −2.6| 3   | 1.5 | 0   | 114.7|
| 2      | ![Structure 2](image2)       | 0  | −3.7| 3   | 2.7 | 0   | 99.8 |
| 3      | ![Structure 3](image3)       | 0  | −3.6| 3   | 2.8 | 0   | 107.1|
| 4      | ![Structure 4](image4)       | 0  | −3.3| 3   | 2.0 | 0   | 104.8|
| 5      | ![Structure 5](image5)       | 0  | −3.0| 3   | 2.5 | 0   | 94.7 |
interaction analysis. Selection of compounds was done based on better interaction with \textit{Pf}pmt. Compounds showed interaction with crucial amino acids like residues Tyr19 and His132 which are important for formation of catalytic dyad and other tyrosine residues important for transmethylation of phosphoethanolamine.

All selected compounds formed hydrogen bonds with crucial amino acids of pCholine binding pocket importantly, with Tyrosine residues. The significantly compounds inhibitors ZINC08792474, ZINC12882412 and ZINC02103914 occupied dyad between Tyr19, His132 crucial for triple methylation of pEth. Top Glide XP scoring compound ZINC08792082 formed H-bond interaction with Tyr160 important residue of pCholine site as shown in Fig. 2. All the seven compounds selected based on interaction with the important amino acids which may be preventing the formation of catalytic dyad and binding (Table 5) of pEth for transmethylation were procured and tested against trophozoites (Schizont) stage of \textit{Plasmodium falciparum}. Compound showed schizonticidal activity at less than five micromolar concentrations were selected as potent antimalarial compounds.

The compound ZINC12882412 also showed good docking score (108.80 kcal/mol) than pCholine. Compound ZINC12882412 provided four hydrogen bonds with Tyr19, Tyr181, Tyr160 and Ala129 where binding with these three tyrosine residues showed the better affinity for \textit{Pf}pmt. Both the amino acids (Tyr160 and Tyr181) acted as H-bond donor and provided side chain hydrogens to the compound’s hydroxyl oxygen. Compound also participated in the Van der Waals interaction with amino acids like His132 and Lys247 crucial for catalytic dyad formation and protein function. The binding with tyrosine residues increased the probability of inhibition of transmethylation. Compound ZINC12882412 possessed pyrrole and pyrrolidine heterocyclic rings fused with benzene rings at both ends of the it which may be crucial for the stability and activity of the compound which could inhibit the formation of schizonts at IC$_{50}$ 2.1 µM which showed the potency of the compound (Fig. 4). Very low micromolar concentration of these compounds implied the good potency and specificity of compounds for \textit{Pf}pmt as inhibitors.

Hence, these two compounds may act as potent inhibitors for \textit{Pf}pmt. Since \textit{Pf}pmt has significant identity with other pmt \textit{Plasmodium} orthologs so, the inhibitors may also be effective antimalarial worldwide.

**Cytotoxicity testing**

Five thousand HEK-293 cells were embedded with both the selected compounds diluted in concentrations (5 µM, 10 µM, 20 µM, 40 µM, 80 µM, 1600 µM)
for testing Fig. 5. Both the compounds ZINC02103914 and ZINC12882412 were found safer with LD₅₀ concentration 77.12 µM and 68.54 µM respectively. Even the higher concentration than the IC₅₀ was found nontoxic for the HEK-293 cells. Both the compounds ZINC02103914 and ZINC12882412 attained the better safety index 25.83 µM and 32.63 µM respectively Table 6.

The compounds were also found nontoxic noncarcinogenic and nonmutagenic in computational toxicity predictions which correlated with invitro. Hence, the better safety index of the compounds implied about therapeutic safety that these may be safer within the body.

**Dynamic simulation of protein–ligand complex**

In order to confirm binding modes and interaction of best Pfpmt inhibitor (ZINC12882412), the molecular dynamic simulation (MDS) was run using Desmond. The behavior of the protein and inhibitor was studied in
terms of root mean square deviation (RMSD) and root mean square fluctuation between protein and ligand and protein–ligand interactions throughout the simulations.

**Root mean square deviation**
To explore the dynamic stability of systems, MD properties were observed up to 20 ns for protein–ligand complex and RMSD values over time from the starting structure was analyzed as depicted in Fig. 4. The MD analysis of protein–ligand complex showed that the RMSD trajectories within the allowed range (1–3 Å) for small, globular proteins. From 0 to 5 ns system showed slight fluctuation to equilibrate with the external environment and after 5 ns, the system remained equilibrated at the end of simulation that give insight into its stable structural conformation. The backbone of protein showed not as much of deviation as side chain, this might be due to interaction of ligand with the flexible side chain residues (Fig. 6).

The less RMSD of Lig fit Prot’ as compared to the RMSD of protein, demonstrated the stability of inhibitor interactions with protein as the movement of inhibitors remained within the binding pocket throughout the simulation. Throughout the dynamic simulation the stability of protein was remained constant but there little RMSD fluctuations were observed in the inhibitors due to conformational adjustment and interaction with new amino acids within the active site.

**Root mean structure fluctuation**
Root mean structure fluctuation (RMSF) of protein–ligand complex was monitored throughout the simulation period that indicated that the secondary structure elements like alpha helices and beta strands are usually more rigid than the unstructured part of the protein.

Fig. 3 Interaction of compounds ZINC02103914. a Orientation in the hydrophobic surface view, b 2D interaction within the binding pocket, c schizonticidal activity (IC_{50}) of the ZINC02103914 compound.
Minor fluctuations were seen in the active site pocket (residue 80–100) and (residue 150–250) that indicate that the active site residues may be fluctuating in order to form interaction with the ligand (Fig. 7).

**Protein ligand contacts**
Interaction of protein with the ligand throughout the simulation has been explained in protein ligand contacts. The compound ZINC12882412 had ten rotatable bonds which undergone continuous conformational adjustments throughout the simulation. Protein–ligand interactions (or ‘contacts’) categorized into four types: hydrogen bonds, hydrophobic, ionic and water bridges. We found, that the inhibitors had stable H-bond interaction with the active site residues that had initial interaction (Tyr 19, Ile 86, Ala 129, Tyr 160, Tyr 181, Ser 37, and Asp 85) as H-bond interactions are important because of their strong influence on drug specificity, metabolism and adsorption. We also observed interaction of other residues within the binding pocket that may be due to flexibility of ligand in presence of water shown in Fig. 8.

**Experimental**

**Ethics approval and consent to participate**
This study was conducted under the approval of the Human Ethical Committee of National Institute of Malaria Research (NIMR), New Delhi (Approval No. ECR/NIMR/EC/2015/206) with proper written documentation of informed consent. The consent was also explained verbal to the patient. The minors were excluded from study.

**Compound library and ADMET analysis**
Compound library was made from Natural compound of ZINC database. The ADMET that is computational analysis of pharmacokinetic properties of a compound was calculated from ADMET and toxicity prediction (TOP-KAT) modules of discovery Studio 3.5 (DS3.5) [15]. The
compounds, devoid of ADME descriptors and Lipinski criteria were removed at very start to save experimental cost and time [16, 17]. Carcinogenicity, mutagenicity and affinity for Cytochrome P450 2D6 (Cyp2d6) were also analyzed. DS3.5 is robust software for insilico ADME and toxicity analysis [18].

Docking analysis of compounds

The crystal structure of Pf pmr 3D7 (PDB Id: 3UJ9) was retrieved from PDB (http://www.rcsb.org/pdb/home/home.do) for docking studies [19]. The protein was prepared by applying CHARMM forcefield using DS3.5. The binding site was created around the co-crystallized phosphocholine (pCholine) using Discovery Studio 3.5. The binding site covered the important interacting amino acids with pCholine to screen the compounds. Docking of the filtered compound library was carried out using LibDock docking module of DS3.5 which works based on formation of polar and apolar HotSpot [20, 21]. The compounds were scored based on the orientation of compounds within the binding site and docking score. The binding energy was also calculated. The pCholine and pEth were used as docking control in the study so; these were also docked with Pf pmr.

Compound preparation and invitro schizonticidal activity

Parasite strain Plasmodium falciparum 3D7 available at parasite bank, National Institute of Malaria Research, India was grown under optimum conditions. RPMI1640 media was used as growth medium for parasite. Other

| S. no. | Comp. id | IC<sub>50</sub> schizonticidal (µM) | LD<sub>50</sub> cytotoxicity (µM) | Safety index (LD<sub>50</sub>/IC<sub>50</sub>) |
|--------|----------|---------------------------------|---------------------------------|---------------------------------|
| 1      | ZINC02103914 | 3.0                             | 77.12                           | 25.83                           |
| 2      | ZINC12882412 | 2.1                             | 68.54                           | 32.63                           |

Fig. 5 Showed the toxicity analysis of the different concentrations of compounds against the HEK-293 cells

Fig. 6 Protein–ligand RMSD. It showed the RMSD between the backbone, side chain and heavy atoms aligned on the ligand
required growth supplements like glucose and antibiotic were added to the RPMI1640 media at the time of media preparation. Incomplete media was prepared with 5% sodium bicarbonate and complete media was prepared by adding human serum (10% heat inactivated) to the incomplete media. The human serum was collected from Indian Red Cross Society-1, Red Cross Road New Delhi-11000, INDIA. When rings formed, parasite culture was treated with 5% sorbitol and rings were separated. Parasite 1% rings were used for schizonticidal activity of compounds in 96 well-microlitre plate [22].

For each compound the stock solution of 200 µM was prepared in 0.1% DMSO and ten serial dilutions of each were prepared. Experiments were performed in triplicates with controls and reference drug (chloroquine) by placing it in CO₂ incubator at 37 °C. After 32–36 h when schizonts formed, the blood Giemsa smear glass slides were prepared for each well of control and sample for microscopic evaluation. The schizonts were counted from 200 asexual parasites microscopically and percent parasitaemia calculated.

Cytotoxicity testing

MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) based cytotoxicity testing of two compounds was done against Human embryonic kidney cells 293 (HEK293). The HEK 293 cells of human were procured from ATCC with identifier no. 293 [HEK-293] (ATCC® CRL-1573™) and catalog no CRL-1573. The cells were grown in Dulbecco's Modified Eagle's Medium. Growth medium was supplemented with glucose, penicillin and fetal bovine serum (5%). Washed and trypsinized five thousand HEK-293 grown cells were seeded with containing growth media into 96-well plate and cell viability was checked after 24 h using trypan blue dye exclusion test. Cells showed viability 98% were incubated with different concentration of primary hits to the 96 well plate for 24 h followed by 4 h incubation with MTT 5 mg/ml. The absorbance was recorded at 580 nm using Synergy/HTX MultiScan reader (BioTek) [23]. Experiments were carried out in triplicate and the Lethal dose (LD₅₀) of each primary hits was calculated and the safety index (SI) was calculated.

Dynamics simulation

Molecular dynamics (MD) simulations were done to analyze more intrinsically the binding mode stability and interaction pattern of best identified inhibitor through docking studies. MD simulation was run using the Desmond v 4.2 [24]. The protein was prepared for by adding hydrogen, assigning bond orders and by generating protonation states at physiological pH (7.4) for MD run, thereafter, the missing side chain and missing loop were filled and terminal residues were capped with neutral groups such as N-terminal acetyl and C-terminal amide and protein was optimized. Then, the protein–ligand complex system was solvated with TIP3P water model inside the orthorhombic box ensuring 10 Å solvent buffers and system was neutralized by adding Na⁺ and Cl⁻ ions and simulations were assigned using the

Fig. 7 Protein RMSF: showed the fluctuations among the backbone and the sidechains during the simulation. The green lines are amino acids participated in interacting with ligand throughout the simulation
OPLS-AA/2005 force field. Then, the solvated system was relaxed using relaxation protocol as implemented in Desmond. Finally, a 20 ns production simulation was performed using the NPT ensemble using Nose–Hoover chain thermostat and Martyna–Tobias–Klein with isotropic coupling at constant temperature 300 K and pressure of 1ATM. The long-range electrostatic interactions were handled using the particle-mesh Ewald (PME) method. A distance cutoff of 9.0 Å was used for short-range electrostatics and Lennard–Jones interactions. Energy and trajectory were saved at every 10 ps and 20 ps time interval respectively. The trajectory for RMSD and interaction of ligand with protein was visualized in Maestro [25].
Conclusion
Phosphatidylcholine an important component of membrane which is synthesized abundantly from serine decarboxylation pathway during intraerythrocytic cycle as well as gametocyte development cycles of Plasmodium. The knockout of Pfppmt revealed importance of Pfppmt in the phosphatidylcholine biosynthesis from serine for survival and malaria transmission and moreover absence of phosphoethanolamine methyltransferase in human makes this enzyme an excellent drug target for development of new antimalarial. The compound library built from natural compound of zinc database was screened based on the docking, Lipinski rule of five and ADMET analysis. Compounds were selected based on the good docking scoring and ADMET physicochemical properties. Five compounds (ZINC08792082, ZINC02120366, ZINC08792474, ZINC12882412 and ZINC02103914) found better based on insilico analysis and procured for in vitro testing. These compounds formed the hydrogen interaction with crucial tyrosine residues and also crucial amino acids of pCholine binding site. The schizonticidal activity was done against the 1% rings of Plasmodium falciparum where best compounds found ZINC12882412 (IC_{50} 2.1 μM) and ZINC02103914 (IC_{50} 3.0 μM) were occupied with catalytic dyad between Tyr19 and His132. These two compounds inhibited the development of schizonts from rings at very low μM concentration (3.0 μM and 2.1 μM) implied probability of being occupied with the crucial amino acids of target protein to prevent its biological function and the non-toxicity towards the kidney cells confirmed the safety of these two compounds too. The simulation study of 3D binding mode of the best inhibitor ZINC12882412 elucidated that the binding with the important active site residues were also conserved throughout the MD simulation for 20 ns that signified the stability of interaction near the active site. Hence, these compounds may act as potent inhibitors of Pfppmt and may lead as template for structure based drug designing to overcome the problem of multi drug resistance.

Abbreviations
Pfppmt: Plasmodium falciparum phosphoethanolamine methyltransferase; HEK-293: human embryonic kidney 293 cells; ADMET: absorption distribution metabolism excretion and toxicity; ACT: artemisinin combination therapy; DAG: diacylglycerol; CDP: cytidine diphosphate; PC: phosphatidylcholine; PSA: polar surface area; Cyp2d6: cytochrome enzyme; MD: molecular dynamic; RMSD: root mean square deviation; RMSF: root mean structure fluctuation; DMSO: dimethyl sulfoxide; OPLS-AA: optimized potentials for liquid simulations; NPT: normal pressure and temperature.

Authors' contributions
JS, AS and MK performed docking simulations and in vitro studies; SV assisted in vitro studies; RM assisted in computational result analysis; GCS provided the access of computational software's and assisted in the results interpretation throughout the study. All authors read and approved the final manuscript.

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Competing interests
The authors declare that they have no conflicts of interests.

Availability of data and materials
Not applicable.

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