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COVID-19 Vaccination Induced Lymphadenopathy in a Specialized Breast Imaging Clinic in Israel: Analysis of 163 cases

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Introduction: Following vaccination of Israeli population with Pfizer-BioNTech COVID-19 Vaccine, an unusual increase in axillary-lymphadenopathy was noted. This study assesses the rate and magnitude of this trend from breast-imaging standpoint.

Materials and Methods: Participants undergoing breast-imaging, in whom isolated axillary-lymphadenopathy was detected were questioned regarding SARS-CoV-2 vaccine to the ipsilateral arm. Patients’ and imaging characteristics were statistically compared. In order to perform a very short-term follow-up, twelve healthy vaccinated medical staff-members, underwent axillary-ultrasound shortly after the second dose, and follow-up.

Results: Axillary-lymphadenopathy attributed to vaccination was found in 163 women undergoing breast-imaging, including BRCA-carriers. During the study, number of detected lymphadenopathies increased by 394% (p = 0.00001) in comparison with previous 2 consecutive years. Mean cortical-thickness of abnormal lymph-nodes after second dose vaccination was 5 ± 2 mm. Longer lymph-node diameter after second vaccination was noted (from 15 ± 5 mm, to 18 ± 6 mm, p = 0.005). In the subgroup of medical staff members, following trends were observed: in patients with positive antibodies, lymph-node cortical-thickness was larger than patients with negative serology (p = 0.03); lymph-node cortical-thickness decreased in 4-5 weeks follow-up (p = 0.007). Lymphadenopathy was evident on mammography in only 49% of cases.

Discussion: Vaccine-associated lymphadenopathy is an important phenomenon with great impact on breast-imaging clinic workload. Results suggest the appearance of cortical thickening shortly after both doses. Positive serology is associated with increased lymph-node cortical-thickness. In asymptomatic vaccinated women with ipsilateral axillary-lymphadenopathy as the only abnormal finding, radiological follow-up is probably not indicated. BRCA-carriers, although at higher risk for breast-cancer, should probably receive the same management as average-risk patients.

Key Words: COVID-19 vaccine; SARS-Cov-2; axillary lymphadenopathy; breast imaging; BRCA-carriers.
**BRCA2** mutation carriers. Most of the cases with lymphadenopathies were detected by ultrasound and MRI, with a part of them shown also by mammography.

Ipsilateral axillary lymphadenopathy has been previously reported shortly after smallpox, Bacille Calmette-Guérin (BCG), human papillomavirus HPV and H1N1 influenza A virus vaccines (1–3). Yet these vaccination campaigns have never been so massive and deployed on a nation-wide basis as the current SARS-CoV-2 vaccine campaign in Israel. Recently, several case reports on the occurrence of post-COVID 19 vaccination axillary lymphadenopathy were published (4–8). In February 2021, a scientific expert panel published recommendations regarding post vaccination lymphadenopathy for patients undergoing imaging that includes the axillae (9,10).

Herein, we performed an observational study to assess the rate and magnitude of post SARS-CoV-2 vaccination axillary lymphadenopathy on ultrasound and MRI, and the change in appearance and size of lymph nodes over time. Since our breast imaging unit serves the largest Israeli **BRCA** carrier surveillance clinic, information on lymphadenopathy in that high risk group was also collected. Furthermore, data on antibody levels against SARS CoV- 2 and comparison to the magnitude of the lymph node cortical thickening was performed.

**MATERIAL AND METHODS**

**Patients**

A retrospective observational study was carried out: the study population included all women who underwent breast imaging during the study period (between January 11, 2021 and February 04, 2021) at our breast imaging center, who presented newly detected unilateral axillary lymphadenopathy on imaging, and had a history of recent COVID-19 vaccination to the ipsilateral arm. In the regular anamnesis, if the radiologist observed axillary lymphadenopathy, the patients were asked if they were vaccinated for COVID-19 in the ipsilateral arm, in order to decide if the lymphadenopathy would be attributable to the vaccine or if further investigation was necessary. This was noted in the report, which was afterwards retrieved from the computerized hospital information system using the OpisoftCare software. The study population was composed of women undergoing routine average-risk screening for breast cancer (n = 67), women with past history of breast cancer undergoing annual imaging surveillance (n = 28), high-risk population undergoing screening for breast cancer due to family history of breast cancer (n = 5), women with newly-diagnosed breast cancer (n = 3), women with a clinically palpable mass in the axilla and lymphadenopathy detected on breast mammography or ultrasound (n = 4). Additionally, women who are carriers of pathogenic sequence variants in the **BRCA1** or **BRCA2** genes that underwent regular breast imaging surveillance between January 11, 2021 and March 31, 2021 were also included (n = 44). For each participant the data collected included age, relevant clinical data on patient’s personal history of breast cancer (if diagnosed), vaccinated arm side, dates of first and second vaccination, postvaccination symptoms such as pain, fever, were recorded. In addition, serology dates and results and biopsy results were also collected, when these were performed and available on the patients’ medical files. Furthermore, in order to perform a very short term follow-up on the imaging course of the axillary lymphadenopathy, twelve healthy medical staff members who were vaccinated, underwent baseline axillary US shortly after the second dose (range: 1–7 days) and a follow-up axillary US (average 4–5 weeks, range: 3–6 weeks post vaccination).

Finally, in order to evaluate the vaccination-associated excessive incidence of cases of ipsilateral lymphadenopathy, a comparison was performed with the rate of cases showing unilateral lymphadenopathy detected during the same time frame in 2020 and 2019. Using the OpisoftCare software, we searched our institutional radiological information system on a prospectively maintained database for women who underwent US examination during the same period last year. Case–by–case confirmation was then performed to identify cases with documented unilateral lymphadenopathy.

**Imaging Technique**

For the non-**BRCA** cohort, which accounts for the majority of participants 119/163 (73%), ultrasonography (US) was the imaging modality mostly applied to detect, qualify, and perform analysis of the axillary lymphadenopathy. For **BRCA** carriers’ cohort, MRI was the main imaging modality. The flowchart with the exact distribution of subgroups and imaging modalities is shown in Figure 1.

Ultrasound: Whole breast and bilateral axillae US examinations were performed by certified fellowship–trained breast radiologists, using Siemens Acuson S2000 ultrasound system with a linear transducer 18–6Mhz, (Siemens Medical Solutions USA, Inc., Mountain View, CA, USA). Image parameters were as follows: frame rate, 60 frames/second; gain, 0 dB; dynamic range, 65 Db.

MRI: Scan was performed on 1.5-T MRI (Signa Excite HDX, GE Healthcare) using a dedicated double breast coil equipped with 8 channels. A standard dynamic contrast enhanced MRI protocol was via axial vibrant multiphase 3D DCE T1-weighted sequence with the following parameters: echo time (TE)/ repetition time (TR) of 2.6/5.4 ms, flip angle 15°, bandwidth 83.3 kHz, matrix 512 × 364; field of view 310 mm × 310 mm.

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**Figure 1.** Flowchart showing patients subgroups distribution and imaging modalities.
view (FOV) 340 mm and slice thickness of 2 mm. DCE was acquired prior to and 4 times after an automated injection of contrast agent bolus (0.1 ml/kg at 2 ml/sec Dotarem, (gadoterate meglumine, Guerbet)) followed by a 20-ml saline flush. Post-contrast images were acquired, with the first acquisition centered at 1:25 minutes after injection and the delayed acquisition centered at 7:35 minutes after injection, as was previously described (11).

In our breast imaging center, we screen average risk patients with mammography, and we add handheld radiologist performed ultrasound when the breasts are heterogeneously dense or extremely dense (BI-RADS Category C or D by BI-RADS 5th ed), or if there is a clinical or mammographic finding. High-risk patients (BRCA carriers or patients with patients with ≥20% lifetime risk of having breast cancer) will undergo MRI and mammography every year.

Image Analysis

Ultrasound imaging features of the axillary lymphadenopathy included: maximal dimension on long axis, and cortex width of the largest lymph node on the vaccinated arm side. In this regard, it should be mentioned though, that a lack of consensus exists for axillary node size assessment on ultrasound (12), while focal or diffuse thickening greater than 3 mm cortical thickness are considered proper parameters (13−15). Measurements were obtained by a single reader (RF, certified fellowship-trained breast radiologist, with 8 years of experience in breast imaging). The same measurements were also performed on MRI, and were obtained on the second post contrast T1-weighted image with fat-suppression.

For non-BRCA women, 81 mammograms and 107 screening ultrasounds were performed. For BRCA carriers, 44 MRIs were performed.

Serology

In 9 participants, serology results for SARS-CoV-2 antibodies were available, as part of a service offered by the hospital to the staff. Serology was measured in units, when results equal or higher than 1 unit were considered positive. Results equal or under 0.8 unit were considered negative. Results between 0.81 and 0.99 unit were considered borderline. This results were correlated with axillary lymphadenopathy.

Statistical Analysis

Comparison of the rate of unilateral lymphadenopathy detected during the same periods of the study with those reported in 2019 and 2020 was carried out using chi-square test. In addition, imaging measurements were compared between cases that were examined after the first and second dose (Student’s paired 2-tailed t-test). Finally, intra-individual comparison between the staff members with lymphadenopathy that were examined twice, at baseline and follow-up US was performed (Student’s paired 2-tailed t-test). All statistical tests were calculated using SPSS V27. P values were considered significant at $p = 0.05$.

Our institutional review board approved this retrospective study and waived the requirement for informed consent.

RESULTS

Overall, the study encompassed 107 participants (non-BRCA carriers) undergoing breast and axillary ultrasound, 12 medical staff members, and 44 BRCA carriers who underwent breast ultrasound, mammography or breast MRI, for a total of 163 participants. In all participants, post vaccination lymphadenopathy was not present on previous breast imaging of the same participant and was limited to the ipsilateral axilla of the vaccinated arm.

Comparing the rate of newly diagnosed ipsilateral lymphadenopathies of all causes detected by US in the current year, with those detected in the previous 2 consecutive years, during the same period of January 11, 2020 until February 04, 2020, and January 11, 2019 until February 04, 2019 the following results were noted: 2019 − 31/484 (6.4%); 2020 − 31/547 (5.7%) and the current study 153/573 (26.7%), an increase of 394% ($p = 0.00001$) (Fig 2). Vaccination associated lymphadenopathy was attributed to 77.8% of lymphadenopathy cases of all causes in the current period (119 lymphadenopathies were attributed to the vaccine, while 34 were due to other causes, including metastatic cancer and reactive lymphadenopathies due to other causes). Illustration of the rise in the lymphadenopathy cases as compared to previous years is presented in Figure 2.

Non-BRCA Carriers

Thirty participants had ultrasound after the first vaccine dose and 77 patients after the second vaccine dose, and 12 staff members after the second dose. The mean time in days of the

Yearly-based distribution of positive axillary lymphadenopathy cases detected on US

Figure 2. Rise in lymphadenopathy cases as compared to previous years. (Color version of figure is available online.)
ultrasound after the first vaccine dose (mean ± SD) was 9 ± 6 days (range 1–20 days), and after the second vaccine dose imaging occurred after 9 ± 8 days (range 1–38 days). Mean cortical thickening of the abnormal lymph nodes after the first and second dose of vaccination, in the group of non-BRCA4 carriers was (mean ± SD), respectively: 5 ± 1.5 mm, range 3–9 mm (after the first dose) and 5 ± 2 mm, range of 3–15 mm (after the second dose) (p = 0.70). Mean ± SD length of abnormal lymph nodes after the first vaccination dose was 15 ± 5 mm (range 7–30 mm), and increased to 18 ± 6 mm (range 9–48 mm) after the second vaccination dose (p = 0.005) Table 1.

In the subgroup of patients having breast imaging after the first dose, 17 underwent mammography, and ipsilateral lymphadenopathy was shown in 9 of them 9/17 (52%). In the subgroup of patients having breast imaging after the second dose, 64 underwent mammography, and ipsilateral lymphadenopathy was shown in 31 of them 31/64 (48%). Altogether, lymphadenopathy was seen in only 40/81 (49%) of ultrasound positive cases.

In the non-BRCA4 participant group, 33/107 (30%) had a personal history of breast cancer, of whom 8 were vaccinated in the ipsilateral arm to the side that breast cancer was diagnosed. In 5/8 of these cases, the imaging score was BI-RADS 2 with no recommendation for a repeat interval breast imaging, and mild lymphadenopathy was attributed to the vaccination (the assessment was BI-RADS 2 since breast imaging was normal and there was a known clinical event which explained ipsilateral mild lymphadenopathy – the vaccination); 2 cases (1 with a palpable axillary mass) were instructed to undergo a short term (2 month) repeated US, and another one was recommended to undergo biopsy since the cortex was very thickened up to 15 mm, with a benign pathological result. Both follow up ultrasounds demonstrated normal sonographic appearing lymph node with cortex width up to 0.2 mm.

**BRCA Carriers**

44 BRCA carriers with lymphadenopathy detected by breast imaging were included: 27 BRCA1, 16 BRCA2 carriers, and 1 patient that is carrier of both a BRCA1 and BRCA2 mutation. Six participants had breast ultrasound (1 due to palpable lymphadenopathy and 5 as part of the routine protocol surveillance scheme), 2 had a mammogram and 36 had a breast MRI. Figure 3 shows 2 examples of axillary lymphadenopathy on MRI. Mean ± SD cortical thickening was 6 ± 2 mm (range 3–13 mm) and mean lymph node length was 14 ± 4 mm (range 8–24 mm). All mild lymphadenopathy were also considered secondary to vaccination and these women also received a BI-RADS 2 categorization. Fourteen BRCA4 carriers had past history of breast cancer (32%), 3 of them bilateral, 6 in the contralateral vaccinated arm and 2 in the ipsilateral vaccinated arm. One BRCA4 carrier with past history of bilateral breast cancer had an ultrasound guided biopsy, in which pathology showed reactive lymph node. Both patients vaccinated in the ipsilateral arm to the previous breast cancer received a BI-RADS 3 categorization with recommendation of short term ultrasound follow up. One of the women with a personal history of left breast cancer that was vaccinated in the right arm had a new focus in the right breast – she received recommendation of biopsy of the focus, which was not seen in the day of the biopsy – this patient received a recommendation of sonographic short term follow up, that was performed 11 weeks after the second dose – which showed complete resolution of the lymphadenopathy, with a normal cortical thickness of 1 mm. Participants vaccinated in the contralateral arm and without history of breast cancer received a BI-RADS 2 with recommendation of continued routine protocol approved breast imaging surveillance.

**Biopsy of Ipsilateral Axillary Lymphadenopathy**

In 3 patients with a newly diagnosed breast cancer and ipsilateral axillary lymphadenopathy, ultrasound guided core needle biopsy was performed (an example is shown in Fig 4). Additionally, 3 cancer free women with clinically suspicious lymphadenopathy (1 with family history, 1 with previous ipsilateral breast cancer and 1 with no history of breast cancer), and an additional BRCA carrier with past history of bilateral breast cancer also had an ultrasound guided biopsy. In all 7 participants pathology results showed benign reactive lymph node. A summary of outcomes for both groups is shown in Table 2.

**Palpable Versus Subclinical Lymphadenopathy**

Overall, in 159/163 (97.5%) participants lymphadenopathy was detected only by imaging, whereas in 4 cases a palpable lymphadenopathy was noted (all of them in the non-BRCA4 carriers’ group, 1 of them with a personal history of breast cancer in the contralateral breast). A short-term ultrasound follow-up after 6–12 weeks was recommended to all the patients with palpable lymphadenopathy. One already performed repeat US after 6 weeks and adenopathy had resolved.

**Serology**

In 9 participants, serology results for SARS-CoV-2 antibodies were available. In 7 participants, serology levels were positive (higher than 1 unit). All participants with positive serology
presented with enlarged lymph nodes (mean ± SD lymph node length of 18 ± 5.2 mm, range 9-25 mm; mean ± SD cortical thickness 5 ± 2.5 mm, range 4-10 mm). Two additional participants who performed serology testing had negative serology (0.03 serology units), both had only slight cortical thickness of 3 mm which was statistically significant smaller compared to the size of the 7 serology positive participants (p = 0.03).

**Short-Term Follow-Up**

Short term ultrasonographic follow up in the medical staff cohort demonstrated that the lymphadenopathy started to recede within 4 weeks from the second dose, with a mean cortical thickness of 3 mm. Subsequent US follow up in 4 of these medical staff after 6 weeks showed continued reduction of the cortical thickness, being less than 3 mm (p = 0.007).

Summary of the staff-members lymph node data is provided in table 3.

**DISCUSSION**

In the current study we show an almost 4-fold increase in axillary lymphadenopathy detected on breast imaging in 2021 compared to 2019 and 2020 in the same medical center, an increase attributed to the massive COVID-19 vaccination campaign in Israel. Vaccination with the Pfizer-BioNTech vaccine was associated with a mostly radiologically detected, clinically subtle, ipsilateral lymphadenopathy in women who underwent breast imaging for a variety of clinical reasons: average and high risk women and in cancer free and breast cancer affected women.

Moderna reported on higher rates of axillary lymphadenopathy compared to prior vaccines (16,17); In the Moderna clinical trials, axillary swelling or tenderness was reported in 11.6% of patients (5.0% placebo) after the first dose and 16.0% (4.3% placebo) after the second, while per Pfizer trials, reports of ipsilateral axillary and supraclavicular lymphadenopathy in the first month after the second dose were imbalanced, with more cases in the vaccine group versus the placebo group, on clinical examination (18–20). In our

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**TABLE 1. Patients and Lymph Node Imaging Data as Demonstrated on Ultrasound (Excluding Medical Staff-Members)**

|                      | First Dose | Second Dose |
|----------------------|------------|-------------|
| Patients that had cortical thickening (n) | 30         | 77          |
| Mean ± SD lymph node cortical thickness | 5 ± 1.5 mm | 5 ± 2 mm    |
| Mean ± SD lymph node length | 15 ± 6 mm  | 18 ± 6 mm   |
| Previous history of breast cancer | 9 patients | 25 patients |

**TABLE 2. Summary of Outcomes in 107 Non-BRCA (excluding staff- members) and 44 BRCA Women**

|                     | Non-BRCA | BRCA |
|---------------------|----------|------|
| BIRADS 2            | 99       | 40   |
| BIRADS 3            | 2        | 3    |
| BIRADS 4            | 6        | 1    |
| Pathology           | Reactive lymph node | Reactive lymph node |
| Outcome of BIRADS 3 | Cortex up to 2 mm after 2-8 weeks | Cortex up to 2 mm after 6-8 weeks |
| Number of breast cancers detected | 3 | - |

![Figure 4. A 27 years old patient that had pain in the left arm and left breast after the first dose of the vaccine (in the ipsilateral arm), and felt a mass in the left breast. Ultrasound showed a suspicious mass in the upper outer quadrant of the left breast (A) and a pathological axillary lymph node (B). A biopsy was performed of both the mass and the axillary lymph node on pathology, the mass was a triple negative invasive ductal breast cancer and the lymph node was reactive (considered to be reactive to the vaccine). (Color version of figure is available online.)](image-url)
study, most of the adenopathy 159/163 (97.5%) was subclinical and mostly detected by imaging only. Previously vaccine associated lymphadenopathy was anecdotaly reported. Mehta et al described 3 cases of patients coming for routine screening mammography and ultrasound, and 1 patient presented due to palpable lump in the ipsilateral vaccinated arm (4). Eifer and Eshet reported a patient who underwent lumpectomy for breast cancer in whom contralateral lymphadenopathy and FDG uptake were attributed to reactive post vaccination lymphadenopathy (5). Özütémiz reported 5 cases of oncology patients with axillary lymphadenopathy that mimicked metastasis (7).

The size of enlarged lymph nodes was roughly but statistically significant correlated with vaccination schedule: 15 mm after the first dose and 18 mm after second vaccination dose (p = 0.005), and mean cortical thickness was abnormal and unchanged over time, measuring 5 mm. The reassuring feature of this radiological phenomenon is that it is self-regressing as shown by the limited follow up in the present study.

Despite the limited number of cases where serology data were available, it seems that minimal cortical thickening is associated with negative serology for SARS-CoV-2 antibodies. It seems plausible that the absence of sizable adenopathy in individuals with a limited measurable antibody levels could be due to a low local immune response and could be related to the absence of vaccine induced antibody formation. This possibility needs to be tested in larger future studies.

The study limitations include a limited number of cases in a single medical center in Israel, and the selection bias that may have resulted. Furthermore since not all vaccinated women underwent axillary sonography, the precise rate of this subclinical phenomenon cannot be accurately determined. The strength of this study is the fact that it was done in a country, where approximately 90% of the adult population have been vaccinated (Israel Ministry of Health).

The clinical impact of this study may be to defer breast imaging screening in asymptomatic individuals to 6 weeks after vaccination. In addition, it would be helpful to query all women undergoing breast imaging, if a vaccine were given, when and in which arm. In asymptomatic vaccinated women, subclinical lymphadenopathy as the only abnormal finding is in all likelihood secondary to vaccination, and radiological follow up is probably not indicated. Similarly, Keshavarz et al. have recently recommended in their review to rely on patient’s clinical context and updated resources to prevent potential disease upstaging and change in therapy, regarding post vaccination axillary lymphadenopathy (21). Patients with current and past breast cancer should be vaccinated in the contralateral arm, as recommended by Becker et al (22). Patients with newly diagnosed breast cancer and ipsilateral lymphadenopathy should have a biopsy performed to exclude malignancy, even if they have a history of vaccination. BRCA carriers, although at a higher risk for breast cancer, should probably receive the same management as patients in average risk.

In conclusion, imaging detected axillary lymphadenopathy appears shortly after vaccination, with a mean cortical thickening of 5 mm, which starts to decrease after 4–5 weeks. In asymptomatic low risk women with normal breast exam, isolated axillary lymphadenopathy and a history of recent COVID-19 vaccination to the ipsilateral arm, a further investigation or follow up is most probably not warranted, though additional studies are required in order to confirm these preliminary observations.

**DECLARATIONS OF INTEREST**

None.

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