A functional SNP in miR-625-5p binding site of AKT2 3'UTR is associated with noise-induced hearing loss susceptibility in the Chinese population

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Abstract

This study aimed to explore the association of several single nucleotide polymorphisms (SNPs) within the AKT2 gene and noise-induced hearing loss (NIHL) susceptibility and explore the potential mechanism underlying NIHL. Three SNPs (rs2304186, rs41275750 and rs76524493) were genotyped in a Chinese population which consists of 690 NIHL patients and 650 normal hearing controls. Bioinformatic analysis was conducted to predict the potential miRNA-binding site of SNPs. Cell transfection and dual-luciferase reporter assay were performed to investigate the potential molecular mechanism of SNPs involved in NIHL. The results revealed rs2304186 GT genotype (OR = 1.41; 95% CI = 1.09–1.83) and TT genotype (OR = 1.51; 95% CI = 1.08–2.10) imparted increased risk of NIHL, and the increased risk could also be found in a dominant model (OR = 1.44; 95% CI = 1.12–1.84). The stratification analysis showed that rs2304186 GT/TT conferred a higher risk for NIHL, especially in subgroups of male, age (35–45 and > 45 years), noise exposure time (> 16 years), and noise exposure level (≤ 85 and ≥ 92 dB), compared with GG genotype. In addition, the haplotype TCCTACT (rs2304186-rs41275750-rs76524493) was associated with NIHL risk (OR = 1.19; 95% CI = 1.02–1.40). Rs2304186 G allele combined with hsa-miR-625-5p mimics could significantly decrease the luciferase activity compared with T allele, indicating rs2304186 altered the binding affinity of hsa-miR-625-5p to SNP rs2304186 mutation region, thus directly targeting AKT2. In conclusion, our study provides evidence for the first time that SNP rs2304186 of AKT2 3’UTR affects NIHL susceptibility by affecting the binding affinity of has-miR-625-5p in an allele-specific manner and it may act as a potential biomarker of NIHL susceptibility.

Introduction

Occupational noise is known as a common harmful factor affecting workers' health in occupational health field. Noise-induced hearing loss (NIHL) has become the second leading form of progressive sensorineural hearing defect caused by occupational noise exposure, after age-related hearing impairment (Miao et al. 2019). In addition, NIHL is the most common occupation-related hearing defect and is an urgent public health issue around the world. The World Health Organization (WHO) previously reported that approximately 10% of population worldwide are exposed to high-intensity noise level environment and may develop NIHL (Basner et al. 2014). The previous study found that 16% of adult disabling hearing loss worldwide can be attributed to occupational noise, accounting for 4.1 million disability-adjusted life years (DALYs) (Nelson et al. 2005). Nowadays, in China, NIHL has become the third largest occupational health hazard, accounting for about one-sixth of the annual increase in occupational related-illnesses (Yu 2016).

NIHL is considered to be a complex hearing disorder caused by environmental factors and genetic effects. Previous studies have found individuals show different degrees of NIHL hazard even though they are in the similar noise environment, indicating that genetic susceptibility might involve in the process of NIHL (Sliwinska-Kowalska & Pawelczyk 2013). Recently, numerous studies intending to identify NIHL-related pathogenic genes along with single nucleotide polymorphisms (SNPs) contributing to NIHL involves in several vital pathways including potassium recycling, oxidative stress, heat shock protein, apoptosis signaling and notch signaling pathways (Miao et al. 2019). However, the susceptibility genes responsible for NIHL have not been adequately explored.
AKT, also called protein kinase B (PKB), is one of key molecules in the PI3K-AKT signaling pathway. There are three human isoforms of AKT, including AKT1, AKT2 and AKT3, encoding by AKT1/PKBa, AKT2/PKBb and AKT3/ PKBc, separately (Zhu et al. 2016). Human AKT2 gene is located in chromosome 19q13.1-q13.2 and is consist of 14 exons (Cheng et al. 1992). AKT2 is highly expressed in adipocytes and muscle tissue and is involved in insulin-mediated regulation of glucose homeostasis (Zdychova & Komers 2005). AKT2 as key component of PI3K-AKT pathway, which participates in an extensive cellular biological processes, covering cell metabolism, survival, growth, and proliferation (Pereira et al. 2015). Dysregulation of AKT2 is concerned with several human diseases comprising cardiovascular, diabetes, cancer, and nerve system disorders (Hers et al. 2011). It has been indicated SNPs act an important type of genetic variations and affect individual's susceptibility to disease (Cargill et al. 1999). Previously, it was reported SNPs in AKT2 were related to gastric cancer (Zhang et al. 2014). Hildebrandt found AKT2 rs892119 was related to the survival of esophageal cancer (Hildebrandt et al. 2009). Moreover, a missense mutation R274H in AKT2 contributing to autosomal dominant inheritance of severe diabetes was observed in a previous study (George et al. 2004).

A previous correlation study found noise exposure could lead to energy and hypoxia metabolism disturbances of hair cells in inner ear, thus leading to hair cell death (Kurabi et al. 2017). However, associations between AKT2 SNPs and the risk of NIHL has not yet been investigated. Considering the important functional roles of AKT2 in cellular growth process, we speculated that potentially functional SNPs in AKT2 gene and their interactions might be responsible for NIHL susceptibility. Therefore, we executed a case-control study to confirm the possible relationships of the AKT2 SNPs (rs2304186, rs41275750 and rs76524493) and susceptibility to NIHL in a Chinese population and deduce the potential mechanism.

Materials And Methods

Study subjects

All recruited study subjects were the frontline workers chronically exposed to occupational noise in the workplaces between 2013 and 2018 in Jiangsu province, China. All workers were required to accept the annual occupational health check-up, including routine physical examination, peripheral venous blood collection and pure-tone audiometry (PTA). Additionally, professionals carried out a questionnaire survey to inquiry subjects’ family history, personal medical history, smoking and drinking habits, and drug use. However, workers with familial hereditary deafness and blast deafness history, some diseases could impair normal hearing (e.g. nervous tinnitus, pyogenic tympanitis, skull trauma, etc.) or have recently taken harmful drugs (e.g. aminoglycosides, aspirin, quinolones, etc.) that can damage the inner ear were excluded.

The present study was approved by the Ethics Committee of Zhongda Hospital, Affiliated Hospital of Southeast University. Informed consents were also obtained from all participants.

Pure-tone audiometric (PTA) examination
After all subjects were asked to stop occupational noise exposure at least 12 hours or more, each subject received PTA examination performed by a certified audiologist in an acoustic room with Madsen Voyager 522 audiometer (Madsen, Taastrup, Denmark) at the frequencies of 0.5, 1, 2, 3, 4, and 6 kHz, respectively. According to GB/T7582-2004, the raw data of hearing thresholds of both ears were revised by age and sex. The diagnosis criteria of NIHL and normal hearing are based on the Chinese Diagnostic Criteria of Occupational NIHL (GBZ 49-2014).

**Definitions of NIHL cases and normal hearing controls**

In the current study, we defined occupational noise exposure as equivalent continuous dB(A)-weighted sound pressure levels of at least 85 dB(A) during a nominal 8-hour working day. NIHL was defined as subjects with average binaural hearing threshold level at high frequencies (3, 4 and 6 kHz) higher than 25 dB. However, normal hearing was defined as subjects with average binaural hearing threshold level less than 25 dB at high frequencies. All subjects were divided into NIHL group and control group. The control subjects were frequency-matched with NIHL cases for sex and age. Eventually, the current study included 690 NIHL patients and 650 controls.

**DNA extraction**

Peripheral venous blood used for DNA isolation was collected from each subject. Genomic DNA was collected with the RelaxGene Blood DNA Kit (Tiangen Biotech, Beijing, China) in accordance with the instructions and was stored at -80°C until use.

**SNPs selection and genotyping**

 Candidate SNPs in AKT2 gene were selected based on the HapMap database and previous results from the literature. (Genomes Project et al. 2015) In this study, the selection of candidate SNPs based on the following criteria: (1) identified by Haploview software; (2) minor allele frequency (MAF) ≥ 0.05 for Chinese Han in Beijing (CHB) population; (3) r² for linkage disequilibrium (LD) value > 0.8. In the end, three SNPs (rs2304186, rs41275750 and rs76524493) were selected for subsequent association studies.

Next generation sequencing was implemented for SNPs genotyping. Briefly, the economical and efficient method consisted of designing of chimeric specific primers containing the target and universal sequences used for three-round multiplex PCR, and the PCR products sequencing (Chen et al. 2016). In our study, the next generation sequencing was accomplished by Biowing Applied Biotechnology Company (Shanghai, China).

Bioinformatics analysis

For the interested and functional SNP rs2304186, which is in 3’UTR region of the AKT2 gene, therefore, we performed bioinformatic analysis to determine if it is located at the miRNA-binding site. Three tools, PicTar (Krek et al. 2005) TargetScan (Friedman et al. 2009) and MirSNP (Liu et al. 2012) were applied for this objective.

Previous studies have found the secondary structure of mRNAs could affect its approachability to miRNA (Mahen et al. 2010). In order to determine the possible effects of major allele G or minor allele T on the
secondary structure of 3'UTR of AKT2 gene, the AKT2 3'UTR fragments along with different length containing allele G or allele T of rs2304186 were confirmed using RNAfold online tool of ViennaRNA web services (http://rna.tbi.univie.ac.at) together with the minimum free energy (MFE) algorithm (Gruber et al. 2008).

**Plasmid construction, cell culture and transfection**

To evaluate the role of AKT2 3'UTR segment containing either wild-type G allele or mutant T allele of SNP rs2304186, the plasmid containing the sequence of rs2304186 was synthesized by Hanbio, Biotechnology Co., Ltd (Shanghai, China) and was cloned into the psiCHECK-2 vector (Promega). The constructed vector was inoculated on the Luria-Bertani (LB) medium and then cultured on a shaking table at 37°C for 14 h. The plasmids were isolated with high-purity plasmid extraction kit (QIAGEN) and subsequently sequenced.

Human embryonic kidney 293T cells (HEK293T) were purchased from Shanghai Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences (Shanghai, China). The HEK293T cells were maintained in Dulbecco's modified Eagle medium (DMEM) (HyClone) with 10% fetal bovine serum (FBS) (Sigma) in a humidified situation with 5% CO$_2$ at 37°C. Besides, has-miR-625-5p and negative control of miRNA mimics were acquired from Hanbio (Shanghai, China).

1.5 × 10$^5$ cells were cultured per well in 96-well plates. When the cells reached 60%-70% confluence, 100 ng of wild-type and mutant plasmids were co-transfected with 5 pmol has-miR-625-5p mimics or negative control of miRNA mimics into the cells using the Lipofectamine™ 3000 reagent (Invitrogen, Carlsbad, CA, USA).

**Dual-luciferase reporter assays**

After 48 h of transfection, cells were harvested and washed three times using the phosphate-buffered saline (PBS). The cells were adequately dissociated via adding 100 μl of 1 × passive lysis buffer (Promega, Madison, WI, USA). Dual luciferase activities were measured using the Dual-Luciferase Reporter Assay System (Promega, Madison, WI, USA). Relative luciferase activity was determined as the ratio of firefly luciferase activity to renilla luciferase activity. Three independent assays were implemented, and each assay was executed in triplicate.

**Statistical analysis**

The categorical and continuous variables were computed as the frequencies and mean ± standard deviation (SD). Goodness-of-fit χ$^2$ test was utilized to decide the Hardy-Weinberg equilibrium (HWE) of selected SNPs of the control subjects. Differences in age, noise exposure year, exposure level and binaural hearing threshold shifts were calculated by Student’s t-test. Pearson’s χ$^2$ test were applied to determine the differences in allele and genotype frequencies of the SNPs between the NIHL cases and controls. Besides, logistic regression model was applied to adjust for the potential confounders (sex, age, smoking status and drinking status), and to determine the possible relationships between the allele and genotype, and NIHL risk, as well as the strength of association. The associations between the genotypes of each SNP and the high frequency hearing threshold shifts were calculated using one-way ANOVA. Moreover, haplotype analysis of the studied SNPs was carried out by using the SHEsis tool.(Shi & He 2005) The interactions among the
studied SNPs was analyzed applying multifactor dimensionality reduction (MDR) analysis. The statistical differences were regarded significant at a $p < 0.05$. Statistical analyses were accomplished via employing SPSS software (SPSS, Chicago, Illinois, USA).

Results

Characteristics of study subjects and HWE tests of selected SNPs

Totally, 1,340 subjects were recruited in our present study, consisting of 690 NIHL cases and 650 controls. Demographic characteristics of the NIHL and control individuals are demonstrated in Table 1. There were no significant differences between the two groups in terms of age, sex, noise exposure time, smoking status, drinking status, and noise exposure level ($P > 0.05$). However, there was a significant difference in the high frequency hearing threshold between NIHL and control groups. The average high frequency hearing threshold for NIHL patients was significantly higher (37.02 ± 12.17) than that of controls (16.88 ± 4.92; $P < 0.001$).

General information for the three studied SNPs and the corresponding results of HWE tests are listed in Table S1. Three SNPs including rs2304186, rs41275750 and rs76524493 are located in 3′UTR region of AKT2. The genotype distributions of the three studied SNPs in the controls was in accord with HWE ($P > 0.05$).

Association between AKT2 SNPs and risk of NIHL

As shown in Table 2, there were significant different in genotypes frequencies of AKT2 rs2304186 between the NIHL patients and controls. When the rs2304186 GG genotype was used as a reference, the GT and TT genotypes imparted increased risk of NIHL (OR = 1.41; 95% CI = 1.09-1.83 for GT genotype; 1.51; 1.08-2.10 for TT genotype), and the increased risk could also be observed in a dominant model (OR = 1.44; 95% CI = 1.12-1.84). Moreover, in the allelic model, rs2304186 T allele conferred a significantly increased risk for NIHL (OR = 1.23; 95% CI = 1.05-1.44) compared with the G allele. Interestingly, rs41275750 heterozygous CG genotype had marginal effect association with NIHL (OR = 0.70; 95% CI = 0.49-1.01; $P = 0.052$). However, the distribution of genotypes frequencies of rs76524493 were not significant difference between two groups.

The stratification analysis showed rs2304186 GT/TT conferred a higher risk for NIHL, especially in subgroups of male, age (35-45 and > 45 years), and work time with noise (> 16 years) (OR = 1.52; 95% CI = 1.17-1.96 for male; 1.53; 1.06-2.20 and 1.75; 1.07-2.88 for age 35-45 and > 45 years; 1.55; 1.09-2.21 for noise exposure time > 16 years), compared with rs2304186 GG genotype (Table 3). For individuals exposed to occupational noise level $\leq 85$ dB, those carrying the rs2304186 GT/TT had a higher risk for NIHL (OR = 1.71; 95% CI = 1.16-2.54). Moreover, individuals exposed to $\geq 92$ dB carrying rs2304186 GT/TT also showed an increased risk for NIHL (OR = 1.55; 95% CI = 1.05-2.30). We further conducted a stratified analysis of rs41275750 and NIHL risk in dominant model, the corresponding results are demonstrated in Table S2. Our data showed that individuals aged 35-45 years (OR = 0.53; 95% CI = 0.31-0.89) and exposed to noise 86-91 dB (OR = 0.36; 95% CI = 0.15-0.85) carrying CG/GG genotypes exhibited a significantly decreased risk for NIHL, suggesting rs41275750 CG/GG genotypes were protective factors for NIHL.
Association between AKT2 haplotypes and risk of NIHL

Table 4 summarizes the haplotype frequencies of selected three SNPs between NIHL group and control group. Three common haplotypes (frequency > 2%) originated from the three SNPs were identified, and the rest of the haplotypes (frequency < 2%) were coalesced into the mixed group. We found that the haplotype TCCTACT (rs2304186-rs41275750-rs76524493) was associated with an increased risk of NIHL (OR = 1.19; 95% CI = 1.02-1.40).

Association between the high frequency hearing threshold shift and SNP genotypes

The association analysis results between the high frequency hearing threshold shift and genotypes of rs2304186, rs41275750 and rs76524493 in all subjects were demonstrated in Fig. 1. For rs2304186, the range of high frequency hearing threshold shift was 25.61 ± 13.18 dB at GG genotype, but the GT and TT genotypes were 28.28 ± 13.26 dB and 30.11 ± 13.87 dB. Subjects carrying rs2304186 GT and TT genotypes were observed to possess higher high frequency hearing threshold shifts than the individuals carrying GG genotype ($P = 0.007$ and $P < 0.001$, respectively). However, no significant statistical differences in terms of the high frequency hearing threshold shift between the genotypes of rs41275750 and rs76524493 were found ($P > 0.05$).

Multifactor dimensionality reduction (MDR) analysis of interactions between SNPs

The corresponding results of MDR analysis on the interactions between the three SNPs are displayed in Table S3 and Fig. S1. Fig. S1 shows that the distributions of high-risk and low-risk genotypes in the best locus model. The interaction results indicated that the interactions among rs2304186, rs41275750 and rs76524493 were associated with an increased risk of NIHL ($P = 0.0107$).

Prediction of RNA secondary structures of AKT2 3′UTR harboring rs2304186

The secondary structures of AKT2 3′UTR segment containing G allele or T allele of rs2304186 were explored by RNAfold. In this study, four different lengths including 85, 105, 125, and 205 bp were used for prediction. Four predictions accordantly indicated rs2304186 G allele mutates to T allele could alter the MFE, suggesting SNP rs2304186 might be associated with the alteration of mRNA secondary structure of AKT2 3′UTR and might influence the accessibility of miRNA target site (Fig. 2A-D).

Prediction of miRNAs that potentially bind to AKT2 rs2304186

Three online tools were applied to determine if functional SNP rs2304186 is located at miRNA target site. We found has-miR-625-5p could target AKT2 rs2304186 (Fig. 2E). has-miR-625-5p was confirmed by the three tools that it could perfectly match with the G allele of rs2304186 in the miRNA seed region.

Analysis of SNP rs2304186 interfered with the interaction between hsa-miR-625-5p and AKT2

AKT2 SNP rs2304186 is located in the 3′UTR of AKT2, to assess whether or not the rs2304186 G > T variant would affect the binding of the predicted has-miR-625-5p to the 3′UTR of AKT2 mRNA, two recombinant constructs containing the G allele or T allele were transiently co-transfected with has-miR-625-5p or
corresponding negative control mimics into HEK293T cells. As indicated in Fig. 3, a notable decrease of luciferase activity was found in the transfected group of AKT2 3’UTR wild G allele together with hsa-miR-625-5p, whereas the luciferase activity containing 3’UTR of the mutant T allele was not visibly changed compared with the corresponding control. Moreover, we found the luciferase activity of the reporter containing G allele combined with hsa-miR-625-5p was decrease by about 50% compared with the T allele in the presence of hsa-miR-625-5p ($P < 0.001$). Therefore, these results indicated that the G allele of SNP rs2304186 had stronger binding with hsa-miR-625-5p, that is, hsa-miR-625-5p might play regulatory role in directly targeting AKT2 with the rs2304186 G allele. Meanwhile, it is indicated that the SNP rs2304186 may modify miRNA-AKT2 gene regulation by affecting the binding of has-miR-625-5p in an allele-specific manner, eventually causing NIHL susceptibility.

**Discussion**

NIHL is a complicated sensorineural hearing loss that caused by the combination of environmental factors and genetic effects (Miao et al. 2019). The incidence of NIHL is gradually increasing around the world and severely endangers human health. SNP is known as the most common forms of genetic variations in the humans and affect individual’s susceptibility to disease (Cargill et al. 1999). Therefore, the identification of key SNPs associated with NIHL risk is so essential for understanding this disease. Nowadays, some NIHL-related susceptibility genes and SNPs have been identified, involving in several pathways including oxidative stress, potassium recycling, heat shock protein, etc (Miao et al. 2019).

In our study, the genetic association study on three polymorphisms (rs2304186, rs41275750 and rs76524493) in AKT2 gene in 690 NIHL patients and 650 controls was investigated. The results showed the rs2304186 GT/TT genotypes are associated with a significantly increased risk of NIHL. Besides, the haplotype TCCTACT (rs2304186-rs41275750-rs76524493) imparts an increased risk of NIHL. Our findings support the hypothesis that AKT2 polymorphism rs2304186 might involve in the susceptibility to NIHL in the Chinese population.

AKT2 as a key component of PI3K-AKT pathway, which implicates in a wide range of cellular processes, such as cell growth, proliferation and survival (Pereira et al. 2015). To date, several polymorphisms in AKT2 gene have been confirmed to be implicated in several human diseases, such as gastric cancer, esophageal cancer and diabetes (George et al. 2004, Hildebrandt et al. 2009, Zhang et al. 2014) indicating that some AKT2 polymorphisms might play a crucial role in human diseases. Noise exposure can cause energy and hypoxia metabolism disorders of hair cells in inner ear, leading to hair cell death (Kurabi et al. 2017). Previous studies found that during noise exposure, the level of 8-iso-prostaglandin F(2alpha) in the inner ear rises and causes cochlear vasoconstriction and blood circulation disorder (Miller et al. 2003). After noise exposure, the blood circulation of the cochlea gradually restores, leading to the cochlear ischemia-reperfusion injury, thus causing mitochondrial dysfunction and the increased productions of ROS triggering oxidative stress and inflammatory response (Honkura et al. 2016). PI3K-AKT signaling pathway has been identified to play a major part in regulating inflammatory responses (Sun et al. 2019). Currently, related studies have found inflammatory response is the essential response and involves in the pathogenesis of cochlear damage induced by noise (Yang et al. 2016). In addition, studies on animals also revealed noise-
induced differentially expressed genes of cochlear are associated with multiple inflammation-related pathways, covering the cytokine-cytokine receptor and chemokine signaling pathway, and p38/MAPK and JNK signaling pathways (Jamesdaniel et al. 2011, Murai et al. 2008, Patel et al. 2013). In our study, mutation from G allele to T allele at rs2304186 of AKT2 gene was significantly associated with increased NIHL risk. In addition, our findings further suggested AKT2-mediated inflammation may be a potential molecular mechanism for NIHL.

In the stratification analysis, we found the rs2304186 GT/TT conferred a higher risk for NIHL in noise exposure time > 16 years group and noise exposure level > 92 dB group, suggesting long period and high-level noise exposure combined with SNP rs2304186 of AKT2 could contribute to a higher NIHL risk. The findings of the interactions of gene-by-environment are consistent with the correlational studies on NIHL (Guo et al. 2017, Liu et al. 2010). Besides, we observed that the male carrying rs2304186 GT/TT genotypes had a significantly higher risk for NIHL compared with the subjects carrying GG genotype. The results are in accord with the preceding studies, showing the association between polymorphisms and NIHL is likely more common in males (Wang et al. 2017). Individuals aged > 35 years with rs2304186 GT/TT exhibited a higher risk for NIHL, suggesting age is always a risk factor affecting NIHL susceptibility. Haplotype analysis indicated the haplotype TCCTACT (rs2304186-rs41275750-rs76524493) was associated with an increased risk of NIHL. Most importantly, the individuals harboring rs2304186 GT/TT genotypes had a significantly higher level of high frequency hearing threshold shift than those with GG genotype. Taken together, our results have indicated the rs2304186 GT/TT genotypes are associated with the NIHL susceptibility. Furthermore, we found that individuals aged 35–45 years and exposed to noise level 86–91 dB with rs41275750 CG/GG genotypes exhibited a decreased risk for NIHL by stratification analysis, suggesting rs41275750 CG/GG genotypes may be protective factors against for NIHL. Meanwhile, the results suggest that the general individuals without rs41275750 mutant G allele as a protective factor, the NIHL prevention should pay more attentions to preventing age-related hearing loss and reducing noise exposure time and intensity, in order to accomplish individualized prevention.

It is widely considered that 3′UTR plays a critical role in potentially regulating the stability, translation and localization of mRNA. Previous studies observed when SNPs are located at the miRNA-binding site of 3′UTR and they can affect the biological function of miRNA by means of generating or removing binding site of miRNA in the target gene mRNA, thus causing the cellular dysfunction and resulting in the occurrence of diseases (Zhu et al. 2017). Recent studies have found functional SNP rs11077 in the 3′UTR of XPO5 involved in NIHL is disruptive to the miRNAs including miR-4763-5p, miR-5002-3p and miR-617 and affects NIHL susceptibility by disturbing XPO5 expression (Wang et al. 2020). Besides, studies have suggested that several SNPs in 3′UTR may affect the corresponding gene expression through influencing the binding affinity for specific miRNAs, causing the occurrence of diseases (Knox et al. 2018; Pirooz et al. 2018). Considering SNP rs2304186 is located at the 3′UTR of AKT2, we examined whether rs2304186 associated with NIHL influenced AKT2 expression. First, we used three tools (PicTar, TargetScan and MirSNP) to determine if SNP rs2304186 is located at miRNA target site. We found that has-miR-625-5p was confirmed by the three databases that it could perfectly match with the G allele of SNP rs2304186 in the miRNA seed region. Next, we further performed vitro experiments to investigate its potential molecular mechanisms. The luciferase reporter assay results suggested the activity of the reporter containing 3′UTR of rs2304186 G allele of AKT2
gene was decreased to 50% when co-transfected with has-miR-625-5p, whereas the reporter containing 3'UTR of T allele showed no significant alteration of luciferase activity compared with negative control. The findings showed the G allele of rs2304186 could enhance a binding site for has-miR-625-5p and affect miRNA-AKT2 gene regulation.

To our knowledge, has-miR-625-5p belongs to human miR-625 family. It has reported that miR-625 family members play a major part in the development and progression of several types of cancers (Lou et al. 2013; Wang et al. 2014). Zheng et al. (Zheng et al. 2015) confirmed that miR-625-3p exhibits oncogenic functions in colorectal carcinoma by regulating SCA1/E-cadherin/MMP9 pathway. Roth et al. (Roth et al. 2012) detected the expression of miR-625 in non-small cell lung cancer and found miR-625 may have a protective effect on the development of non-small cell lung cancer. In this study, we found has-miR-625-5p might be involved in NIHL through targeting AKT2 to regulate its expression, which could be modified by SNP rs2304186.

**Strengths And Limitations**

Nevertheless, there are some potential limitations to our study. First, all the subjects in the current study were only restricted to the Chinese population. The findings appeared to be more favorable to the Chinese population but not on behalf of other ethnic population. Second, out of the three selected SNPs in AKT2 gene, only rs2304186 was significantly associated with NIHL. It may not be sufficiently to clarify such a complex disease with only single SNP. Third, the personal hearing protection measure of study participants was not taken into account, which might decrease the confidence level of the current results.

**Conclusions**

In conclusion, our current study provides evidence that SNP rs2304186 within miRNA-binding site of AKT2 3'UTR is associated with NIHL susceptibility in the Chinese population and might act by affecting the binding affinity of has-miR-625-5p. Therefore, functional SNP rs2304186 might be a potential biomarker of NIHL susceptibility for Chinese workers exposed to occupational noise. Moreover, our study indicates that the SNPs alter gene expression through miRNAs might be a crucial underlying molecular mechanism NIHL susceptibility.

**Declarations**

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**Authors' contributions**

YP designed the research study. LM and BW performed the research and analyzed the data. LM drafted the manuscript. JZ, LY critically reviewed and revised the manuscript. All authors read and approved the final manuscript.
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Data availability

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Compliance with ethical standards

Conflict of interest

The authors declare that they have no conflicts of interest.

Ethical approval

The present study was approved by the Ethics Committee of Zhongda Hospital, Affiliated Hospital of Southeast University.

Consent to participate

Informed consent was obtained from all individual participants included in the study.

Consent to publish

The participant has consented to the submission of the case report to the journal.

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Tables
| Variables                        | Cases (n = 690) | Controls (n = 650) | P  |
|---------------------------------|-----------------|--------------------|----|
|                                 | n               | %                  | n  | %   |      |
| Age (years)                     | 690             | 100                | 650| 100  |      |
| Mean ± SD                       | 39.98 ± 8.01    | 39.92 ± 8.09       | 0.738<sup>a</sup> |
| < 35                             | 182             | 26.4               | 178| 27.4 |      |
| 35-45                           | 331             | 48.0               | 298| 45.9 |      |
| > 45                            | 177             | 25.7               | 174| 26.8 |      |
| Sex                             | 649             | 94.1               | 612| 94.2 |      |
| Male                            | 41              | 5.9                | 38 | 5.9  |      |
| Work time with noise (years)    | 331             | 48.0               | 335| 51.5 |      |
| Mean ± SD                       | 18.08 ± 9.07    | 17.41 ± 8.88       | 0.192<sup>a</sup> |
| ≤ 16                            | 331             | 48.0               | 335| 51.5 |      |
| > 16                            | 359             | 52.0               | 315| 48.5 |      |
| Smoking status                  | 385             | 55.8               | 337| 51.8 |      |
| No                              | 305             | 44.2               | 313| 48.2 |      |
| Drinking status                 | 225             | 32.6               | 208| 32.0 |      |
| No                              | 465             | 67.4               | 442| 68.0 |      |
| High frequency hearing threshold (dB) | 37.02 ± 12.17   | 16.88 ± 4.92       | <0.001<sup>a</sup> |
| ≤ 26                            | 78              | 11.3               | 650| 100  |      |
| > 26                            | 612             | 88.7               | 0  | 0.0  |      |
| Expose level with noise (dB)    | 87.54 ± 7.61    | 87.27 ± 7.97       | 0.586<sup>a</sup> |
| ≤ 85                            | 286             | 41.5               | 269| 41.4 |      |
|     |     |     |     |     |
|-----|-----|-----|-----|-----|
| 86-91 | 126 | 18.3 | 106 | 16.3 |
| ≥ 92 | 278 | 40.3 | 275 | 42.3 |

*aTwo-sided χ² test. bStudents’ t test.

Abbreviations: SD, standard deviation; dB, decibel.
| SNP no.     | Genetic models | Genotypes | Genotype | Cases | Controls | P   | Adjusted OR (95% CI) |
|------------|----------------|-----------|----------|-------|----------|-----|---------------------|
| rs2304186  | Co-dominant    | GG        |          | 155   | 195      | 1.00| (ref)               |
|            |                | GT        | 351      | 311   | 0.008    | 1.41| (1.09-1.83)        |
|            |                | TT        | 132      | 109   | 0.012    | 1.51| (1.08-2.10)        |
|            | Dominant       | GG        | 155      | 195   | 1.00     | (ref)|                    |
|            |                | GT/TT     | 483      | 420   | 0.003    | 1.44| (1.12-1.84)        |
|            | Recessive      | GG/GT     | 506      | 506   | 1.00     | (ref)|                    |
|            |                | TT        | 132      | 109   | 0.183    | 1.20| (0.91-1.60)        |
|            | Alleles        | G         | 661      | 701   | 1.00     | (ref)|                    |
|            |                | T         | 615      | 529   | 0.009    | 1.23| (1.05-1.44)        |
| rs41275750 | Co-dominant    | CC        | 619      | 568   | 1.00     | (ref)|                    |
|            |                | CG        | 58       | 76    | 0.052    | 0.70| (0.49-1.01)        |
|            |                | GG        | 4        | 0     | 0.158    | -   |                     |
|            | Dominant       | CC        | 619      | 568   | 1.00     | (ref)|                    |
|            |                | CG/GG     | 62       | 76    | 0.108    | 0.75| (0.53-1.04)        |
| rs76524493 | Co-dominant | CTACT/CTACT | 598 | 89.9 | 549 | 87.7 | 1.00 (ref) |
|------------|-------------|-------------|-----|------|-----|------|------------|
|            | CTACT/CT    | 63          | 9.5 | 76   | 12.1| 0.129| 0.76 (0.54-1.09) |
|            | CT/CT       | 4           | 0.6 | 1    | 0.2 | 0.426| 3.52 (0.39-31.69) |
| Dominant   | CTACT/CTACT | 598         | 89.9| 549  | 87.7| 1.00 (ref) |
|            | CTACT/CT +  | 67          | 10.1| 77   | 12.3| 0.204| 0.80 (0.57-1.13) |
|            | CT/CT       | 4           | 0.6 | 1    | 0.2 | 0.407| 3.61 (0.40-32.52) |
| Recessive  | CTACT/CTACT | 661         | 99.4| 625  | 99.8| 1.00 (ref) |
|            | CTACT/CT    | 4           | 0.6 | 1    | 0.2 | 0.407| 3.61 (0.40-32.52) |
| Alleles    | CTACT       | 1259        | 94.7| 1174 | 93.8| 1.00 (ref) |
|            | CT          | 71          | 5.3 | 78   | 6.2 | 0.332| 0.85 (0.61-1.18) |

\(^a\)Two-sided \(\chi^2\) test. \(^b\)Adjusted for age, sex, smoking and drinking status in logistic regression model.

Abbreviations: OR, odds ratio; CI, confidence interval.
| Variables                  | GG (case/control) | GT/TT (case/control) | P \(^a\) | Adjusted OR (95% CI) \(^b\) |
|---------------------------|------------------|---------------------|---------|-----------------------------|
|                           | n    | %     | n    | %     |                    |
| Sex                       |      |       |      |       |                      |
| Male                      | 142/187 | 12.0/15.9 | 458/393 | 38.8/33.3 | 0.001 | 1.52 (1.17-1.96) |
| Female                    | 13/8  | 17.8/11.0 | 25/27  | 34.2/37.0 | 0.284 | 0.70 (0.21-2.39) |
| Age (years)               |      |       |      |       |                      |
| < 35                      | 49/48 | 14.2/13.9 | 129/119 | 37.4/34.5 | 0.802 | 1.04 (0.65-1.68) |
| 35-45                     | 71/90 | 12.3/15.6 | 228/188 | 39.5/32.6 | 0.021 | 1.53 (1.06-2.20) |
| > 45                      | 35/57 | 10.6/17.2 | 126/113 | 38.1/34.1 | 0.017 | 1.75 (1.07-2.88) |
| Work time with noise (years) |      |       |      |       |                      |
| \(\leq 16\)              | 77/99 | 12.2/15.7 | 235/220 | 37.2/34.9 | 0.075 | 1.38 (0.97-1.96) |
| \(> 16\)                 | 78/96 | 12.5/15.4 | 248/200 | 39.9/32.2 | 0.018 | 1.55 (1.09-2.21) |
| Expose level with noise (dB) |      |       |      |       |                      |
| \(\leq 85\)              | 61/85 | 11.8/16.5 | 204/165 | 39.6/32.0 | 0.006 | 1.71 (1.16-2.54) |
| 86-91                     | 34/25 | 15.0/11.1 | 87/80  | 38.5/35.4 | 0.464 | 0.78 (0.43-1.43) |
| \(\geq 92\)              | 60/85 | 11.7/16.6 | 192/175 | 37.5/34.2 | 0.026 | 1.55 (1.05-2.30) |

\(^a\) Two-sided \(\chi^2\) test. \(^b\) Adjusted for age, sex, smoking and drinking status in logistic regression model.

Abbreviations: OR, odds ratio; CI, confidence interval; dB, decibel.
Table 4 Frequencies of inferred haplotypes among the cases and controls and their association with risk of NIHL

| Haplotypes<sup>a</sup> | Cases | Controls |  ρ<sup>b</sup> | OR (95% CI) | Global ρ<sup>c</sup> |
|------------------------|-------|----------|----------------|-------------|---------------------|
|                        | n     | %        | n              | %           |                     |
| GCCTACT                | 586.26| 46.9     | 592.04         | 50.3        | 0.100               | 0.88 (0.75-1.03)    | 0.076               |
| GGCT                   | 61.19 | 4.9      | 69.96          | 5.9         | 0.257               | 0.82 (0.57-1.16)    |                     |
| TCCTACT                | 598.74| 47.9     | 512.96         | 43.6        | 0.030               | 1.19 (1.02-1.40)    |                     |
| Others<sup>d</sup>     | 3.81  | 0.3      | 1.04           | 0.1         |                     | 1.00 (ref)          |                     |

<sup>a</sup>The alleles of haplotypes were arrayed as rs2304186-rs41275750-rs76524493. <sup>b</sup>Two-sided χ² test. <sup>c</sup>Generated by permutation test with 1000 times of simulation. <sup>d</sup>Haplotypes with a frequency ≤ 0.02 (GCCT/TGCT/GGCTACT/TCCT/TGCTACT) were pooled into the mixed group.

Abbreviations: OR, odds ratio; CI, confidence interval.

Figures

Figure 1

Comparison of high frequency hearing threshold shift of rs2304186 (A), rs41275750 (B), and rs76524493 (C) genotypes in all studied subjects. Data are presented as mean ± SE analyzed by ANOVA.
Figure 2

The results of bioinformatics prediction about the secondary structure of AKT2 3'UTR containing either allele G or allele T of rs2304186 and the candidate miRNAs. (A-D) Predicted secondary structure of AKT2 3'UTR containing SNP rs2304186 using RNAfold online tool, mRNA sequences (85, 105, 125, and 205 bp, respectively) containing either allele G or allele T. SNP rs2304186 allele G mutates to allele T consistently changes the MFE. (E) Predicted candidate miRNA (hsa-miR-625-5p) could bind to the SNP rs2304186 of AKT2 3'UTR. SNP rs2304186 in the miRNA-binding site of AKT2 3'UTR is shown by red font.
The luciferase activity in HEK293T cells transfected with a plasmid containing either the wild type (G allele) or the mutant allele (T allele) of SNP rs2304186 in AKT2 3’UTR together with has-miR-625-5p mimics or miRNA-negative control. Renilla luciferase was measured as an internal control, and firefly luciferase signals were normalized with renilla luciferase signals. Data were presented as the mean ± SEM.

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