Parameters of Glucose and Lipid Metabolism Affect the Occurrence of Colorectal Adenomas Detected by Surveillance Colonoscopies

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**Purpose:** Limited data are available regarding the associations between parameters of glucose and lipid metabolism and the occurrence of metachronous adenomas. We investigated whether these parameters affect the occurrence of adenomas detected on surveillance colonoscopy.

**Materials and Methods:** This longitudinal study was performed on 5289 subjects who underwent follow-up colonoscopy between 2012 and 2013 among 62171 asymptomatic subjects who underwent an initial colonoscopy for a health check-up between 2010 and 2011. The risk of adenoma occurrence was assessed using Cox proportional hazards modeling.

**Results:** The mean interval between the initial and follow-up colonoscopy was 2.2±0.6 years. The occurrence of adenomas detected by the follow-up colonoscopy increased linearly with the increasing quartiles of fasting glucose, hemoglobin A1c (HbA1c), insulin, homeostasis model assessment of insulin resistance (HOMA-IR), and triglycerides measured at the initial colonoscopy. These associations persisted after adjusting for confounding factors. The adjusted hazard ratios for adenoma occurrence comparing the fourth with the first quartiles of fasting glucose, HbA1c, insulin, HOMA-IR, and triglycerides were 1.50 [95% confidence interval (CI), 1.26–1.77; \( p_{\text{trend}}<0.001 \)], 1.22 (95% CI, 1.04–1.43; \( p_{\text{trend}}=0.024 \)), 1.22 (95% CI, 1.02–1.46; \( p_{\text{trend}}=0.046 \)), 1.36 (95% CI, 1.14–1.63; \( p_{\text{trend}}=0.004 \)), and 1.19 (95% CI, 0.99–1.42; \( p_{\text{trend}}=0.041 \)), respectively. In addition, increasing quartiles of low-density lipoprotein-cholesterol and apolipoprotein B were associated with an increasing occurrence of adenomas.

**Conclusion:** The levels of parameters of glucose and lipid metabolism were significantly associated with the occurrence of adenomas detected on surveillance colonoscopy. Improving the parameters of glucose and lipid metabolism through lifestyle changes or medications may be helpful in preventing metachronous adenomas.

**Key Words:** Colorectal adenoma, surveillance, glucose metabolism, dyslipidemia

**INTRODUCTION**

The incidence of colorectal cancer (CRC) and mortality associated with it are effectively reduced by removing precursor lesions through colonoscopy and polypectomy. The likelihood of developing a metachronous colorectal neoplasm (CRN) during surveillance differs according to the baseline adenoma characteristics, and thus the surveillance colonoscopy intervals depend on the baseline adenoma characteristics. In addition to determining baseline adenoma characteristics, identifying other risk factors for metachronous CRN may help to establish guidelines for more individualized and optimal surveillance intervals.
Diabetes and dyslipidemia have been linked to an increased risk of CRC precursor lesions, adenomas, as well as CRC.\textsuperscript{4-7} Moreover, some studies have reported that high levels of parameters of glucose metabolism such as fasting glucose, hemoglobin A1c (HbA1c), insulin, and homeostasis model assessment of insulin resistance (HOMA-IR) are significantly associated with the risk of CRN.\textsuperscript{8-11} In addition, several studies have demonstrated that high levels of triglyceride, total cholesterol, and low-density lipoprotein-cholesterol (LDL-C), and low levels of high-density lipoprotein-cholesterol (HDL-C) were associated with an increased risk of CRN.\textsuperscript{12-16} Our previous study also found that the prevalence of CRN increased with increasing levels of glucose, HbA1c, insulin, HOMA-IR, triglycerides, total cholesterol, LDL-C, and apolipoprotein B (ApoB), and with decreasing levels of HDL-C and apolipoprotein A-1 (ApoA-1).\textsuperscript{17} However, all of these previous studies, including our own, were cross-sectional investigations that evaluated the associations between the prevalence of CRN and the levels of parameters of glucose and lipid metabolism.

Recently, a few studies have reported that metabolic factors such as obesity, impaired glucose tolerance, hypertension, and high waist circumference are risk factors for developing metachronous adenomas detected by surveillance colonoscopy.\textsuperscript{18-24} Based on these results, the parameters of glucose and lipid metabolism may also affect adenoma occurrence detected by surveillance colonoscopy. To date, there have been no studies conducted on the associations between the parameters of glucose and lipid metabolism and the occurrence of metachronous adenomas. Therefore, we conducted a longitudinal study to determine whether the parameters of glucose and lipid metabolism (including lipoproteins as well as lipids) influence the occurrence of adenomas detected by surveillance colonoscopy.

**MATERIALS AND METHODS**

**Study population**
The study population consisted of asymptomatic subjects who had undergone a colonoscopy as part of a comprehensive health screening program at Kangbuk Samsung Hospital in Seoul and Suwon, Korea, between 2010 and 2011 (defined as “initial colonoscopy”) (n=62171). Of these participants, 7318 subjects underwent repeat colonoscopy as part of a health checkup at Kangbuk Samsung Hospital from 2010 to 2011 (n=62171). The exclusion criteria were as follows: poor bowel preparation (n=1744), lack of an adequate biopsy (n=43), missing anthropometric data (n=47), a history of CRC or colorectal surgery (n=17), a history of inflammatory bowel disease (n=70), detection of a colorectal carcinoid tumor during this study (n=36), detection of a colorectal adenocarcinoma during this study (n=3), and age <30 years old (n=69). The total number of eligible subjects for the study was 5289 (Fig. 1). No subject had undergone a colonoscopy in which the cecum was not reached.

In Korea, the Industrial Safety and Health Law requires employees to participate in annual or biennial health examinations. About 60% of the participants were employees of various companies and local governmental organizations and their spouses, and the remaining participants registered individually for the program. In most Korean companies, the mandatory retirement age is approximately 55 years old. As part of the welfare policy, companies often subsidize annual or biennial comprehensive health examinations including colonoscopies, regardless of the current guidelines. Such programs are popular in Korea.\textsuperscript{25} Therefore, our database had a relatively large group of subjects aged <50 years who underwent screening colonoscopy. In addition, although clinicians recommend surveillance colonoscopy intervals according to the current guideline,\textsuperscript{4} some subjects opt to undergo a colonoscopy biennially regardless of the recommendation.

Before the patients underwent the colonoscopies, general practitioners conducted interviews to ensure that the subjects had no gastrointestinal symptoms, such as visible rectal bleeding or abdominal pain. Persons with these symptoms were urged to seek medical care, and only asymptomatic participants participated in this screening program.

![Fig. 1. Flow diagram illustrating the selection of study subjects.](https://doi.org/10.3349/ymj.2017.58.2.347)
This study was approved by the Institutional Review Board of Kangbuk Samsung Hospital, which exempted the requirement for informed consent because we retrospectively accessed only de-identified data.

Measurements and definitions
The patient data on medical history and smoking were collected through a self-administered questionnaire, whereas the physical and serum biochemical parameters were measured by trained staff. Self-reported use of nonsteroidal anti-inflammatory drugs (NSAIDs) (regular use over the previous month) was also assessed. A family history of CRC was defined as CRC that occurred in one or more first-degree relatives at any age. A history of diabetes was defined as a self-reported diagnosis of diabetes or the use of an antidiabetic medication. A history of dyslipidemia was defined as a self-reported diagnosis of dyslipidemia or the use of a medication for dyslipidemia. Body mass index (BMI) was calculated by dividing measured weight (kg) by height squared (m$^2$). Obesity was defined as BMI ≥30 kg/m$^2$, based on the proposed cutoff for obesity of the World Health Organization.$^{26}$

Blood samples were collected from the antecubital vein after at least a 10-hour fast. The fasting glucose level was measured using the hexokinase method (Hitachi Modulator D2400; Roche, Tokyo, Japan). HbA1c was measured using an immunoturbidimetric assay with a Cobra Integra 800 automatic analyzer (Roche Diagnostics, Basel, Switzerland) at a reference value of 4.4–6.4%. The fasting serum insulin was measured by electrochemiluminescence immunoassay (Hitachi Modular E 170; Roche). Insulin resistance was assessed with HOMA-IR according to the following equation: fasting blood insulin (mU/mL)×fasting blood glucose (mmol/L)/22.5. The serum levels of total cholesterol and triglycerides were determined using an enzymatic colorimetric assay; LDL-C and HDL-C levels were determined using a homogeneous enzymatic colorimetric assay; and serum ApoA-1 and ApoB levels were determined using an immunoturbidimetric assay.

### Table 1. Baseline Characteristics between Subjects with vs. without Occurrence of Colorectal Adenoma

| Parameter                                      | All subjects (n=5289) | Occurrence (n=1038) | No occurrence (n=4251) | p value |
|------------------------------------------------|-----------------------|---------------------|-------------------------|---------|
| Age, yrs                                       | 42.0±6.8              | 44.6±7.4            | 41.4±6.6                | <0.001  |
| Male sex                                       | 4596 (86.9)           | 947 (91.2)          | 3649 (85.8)             | <0.001  |
| Current or ex-smoker                           | 3440 (65.0)           | 724 (69.7)          | 2716 (63.9)             | <0.001  |
| Family history of CRC                         | 420 (7.9)             | 90 (8.7)            | 330 (7.8)               | 0.332   |
| Use of NSAIDs                                  | 249 (4.7)             | 41 (3.9)            | 208 (4.9)               | 0.198   |
| Fatty liver                                    | 2253 (42.6)           | 492 (47.4)          | 1761 (41.1)             | <0.001  |
| Obesity (BMI ≥30 kg/m$^2$)                     | 250 (4.7)             | 58 (5.6)            | 192 (4.5)               | 0.145   |
| History of diabetes                            | 229 (4.3)             | 68 (6.6)            | 161 (3.8)               | <0.001  |
| History of dyslipidemia                        | 842 (15.9)            | 204 (19.7)          | 638 (15.0)              | <0.001  |
| Parameters of glucose metabolism               |                       |                     |                         |         |
| Fasting glucose (mg/dL)                        | 95.5±15.0             | 96.9±15.6           | 95.2±14.9               | 0.001   |
| HbA1c (%)                                      | 5.7±0.5               | 5.8±0.6             | 5.7±0.5                 | <0.001  |
| Fasting insulin (mg/dL)                        | 4.0 (2.7–5.8)         | 4.1 (2.8–6.0)       | 4.0 (2.6–5.8)           | 0.031   |
| HOMA-IR                                        | 0.9 (0.6–1.4)         | 1.0 (0.6–1.5)       | 0.9 (0.6–1.4)           | 0.005   |
| Parameters of lipid metabolism                 |                       |                     |                         |         |
| Triglycerides                                  | 111 (78–160)          | 119 (83–171)        | 109 (77–158)            | <0.001  |
| Total cholesterol (mg/dL)                      | 204±35                | 206±35              | 203±34                  | 0.025   |
| LDL cholesterol (mg/dL)                        | 129±32                | 130±32              | 128±31                  | 0.057   |
| HDL cholesterol (mg/dL)                        | 53±13                 | 52±12               | 53±13                   | 0.009   |
| Apolipoprotein B (mg/dL)                       | 95±23                 | 98±23               | 94±22                   | <0.001  |
| Apolipoprotein A1 (mg/dL)                      | 135±22                | 135±23              | 135±22                  | 0.742   |
| Initial colonoscopy finding                    |                       |                     |                         | <0.001  |
| Normal group                                   | 3108 (58.8)           | 444 (42.8)          | 2664 (62.7)             |         |
| Low-risk group                                 | 1724 (32.6)           | 437 (42.1)          | 1287 (30.3)             |         |
| High-risk group                                | 457 (8.6)             | 157 (15.1)          | 300 (7.1)               |         |
| Follow-up interval, yrs*                       | 2.18±0.61             | 2.23±0.64           | 2.17±0.61               | 0.009   |

BMI, body mass index; HbA1c, hemoglobin A1c; HOMA-IR, homeostasis model assessment of insulin resistance; LDL, low-density lipoprotein; HDL, high-density lipoprotein; CRC, colorectal cancer; NSAIDs, nonsteroidal anti-inflammatory drugs.

Data are presented as mean±SD, median (interquartile range) or number (%). p value by t-test or Mann-Whitney U-test for continuous variables and chi-square test for categorical variables.

*Interval between the initial and follow-up colonoscopy.
Colonoscopy and histologic examination
The colonoscopies were performed by 1 of 13 experienced gastroenterologists (>1000 cases) by using an EVIS LUCERA CV-260 colonoscope (Olympus Medical Systems, Tokyo, Japan). All the participants took 4 L of polyethylene glycol solution for bowel preparation.

All of the detected polyoid lesions were biopsied or removed and histologically assessed by experienced pathologists. The polyps were classified by number, size, and histologic characteristics (tubular, tubulovillous, or villous adenoma; hyperplastic polyp; inflammatory polyp; sessile serrated adenoma or traditional serrated adenoma). Pathologic results that indicated hyperplastic polyps, inflammatory polyps, or lipomas were considered normal findings. The grade of dysplasia was classified as low or high. An advanced adenoma was defined as the presence of one of the following features: >10 mm diameter, tubulovillous or villous structure, and high-grade dysplasia (HGD). 3 The study subjects were categorized into normal, low-risk, and high-risk groups according to their initial colonoscopy findings. The normal group was defined as subjects with no adenoma; the low-risk group, as subjects with 1–2 tubular adenomas <10 mm; and the high-risk group, as subjects who had adenomas with villous histology, HGD, size ≥10 mm, or 3 or more adenomas detected during the initial colonoscopy.3

Statistical analysis
The data were expressed as mean±SD, median (interquartile range), or number (%). Baseline characteristics were compared between subjects with and without adenoma detected on follow-up colonoscopy by using the chi-square test for categorical variables and t-test, or the Mann-Whitney U-test for continuous variables.

The associations of the individual glucose and lipid markers with the risks of occurrence of any adenoma and advanced adenoma were assessed using Cox proportional hazards modeling, adjusted for potential confounding variables such as age, gender, smoking status, family history of CRC, use of NSAIDs, obesity, and initial colonoscopy findings (normal, low-risk, and high-risk groups). We estimated the hazard ratios (HRs) of adenoma occurrence with 95% confidence intervals (CIs) by comparing quartiles 2 to 4 of markers of glucose and lipid metabolism with the first quartile. Person-years were calculated as the sum of the follow-up duration from visit 1 to visit 2. All of the reported p values are two-tailed, and statistical significance was set at p<0.05. SPSS Version 18 (IBM Corp., Armonk, NY, USA) was used for statistical analyses.

RESULTS
Baseline characteristics of study population
A total of 5289 participants were eligible for the analysis (Fig. 1). At baseline, the mean age of the 5289 subjects was 42.0±6.8 years, and 86.9% of the subjects were men. The mean interval between the initial and follow-up colonoscopy was 2.18±0.61 years, with a total of 11532 person-years. According to the initial colonoscopy findings (baseline adenoma characteristics), 3108 (58.8%) subjects were assigned to the normal group, 1724 (32.6%) to the low-risk group, and 457 (8.6%) to the high-risk group.

The baseline characteristics at initial colonoscopy of the subjects who had an adenoma detected during the follow-up surveillance colonoscopy and the subjects who did not are compared in Table 1. Subjects who had an adenoma detected had a higher mean age compared to subjects without an adenoma (44.6±7.4 vs. 41.4±6.6, p<0.001). The occurrence of an adenoma was seen more frequently in males (91.2% vs. 85.8%, p<0.001), current or ex-smokers (69.7% vs. 63.9%, p<0.001), and patients with fatty livers, (47.4% vs. 41.1%, p<0.001), history of diabetes (6.6% vs. 3.8%, p<0.001), and history of dyslipidemia (19.7% vs. 15.0%, p<0.001). There were no significant differences in the proportion of subjects with a family history of CRC or the use of NSAIDs between subjects with and without adenoma occurrence.

The levels of all the parameters of glucose metabolism at initial colonoscopy were significantly higher in subjects with a detected adenoma than in subjects without an adenoma. The levels of triglycerides, total cholesterol, and ApoB were significantly higher in subjects with a detected adenoma than in subjects without adenomas, while the HDL-C levels were significantly lower in subjects with a detected adenoma than in subjects without adenomas. The LDL-C levels tended to be higher in subjects with detected adenoma; however, the difference was not statistically significant. There was no significant difference in ApoA-1 levels between the two groups. The mean interval between the initial and follow-up colonoscopy was higher in subjects with a detected adenoma than in subjects without an adenoma (2.23±0.64 years vs. 2.17±0.61 years, p=0.009).

Associations between parameters of glucose metabolism and adenoma occurrence
Table 2 shows the risk for occurrence of any adenoma and advanced adenoma at the time of follow-up colonoscopy among the 5289 subjects, according to the quartiles of parameters of glucose metabolism at the time of the initial colonoscopy. The occurrence of any adenoma increased linearly with increasing quartiles of fasting glucose, HbA1c, insulin, and HOMA-IR. In the multivariable-adjusted model, the associations between markers of glucose metabolism and the occurrence of any adenoma remained significant for all markers. The adjusted HRs for any adenoma occurrence when we compared the fourth with the first quartiles of fasting glucose, HbA1c, insulin, and HOMA-IR were 1.50 (95% CI, 1.26–1.77; pmult=0.001), 1.22 (95% CI, 1.04–1.43; pmult=0.024), 1.22 (95% CI, 1.02–1.46; pmult=0.046), and 1.36 (95% CI, 1.14–1.63; pmult=0.004), respectively.

The occurrence of an advanced adenoma increased linearly

https://doi.org/10.3349/ymj.2017.58.2.347
with increasing quartiles of fasting glucose in the univariate analysis (Q2, Q3, and Q4 vs. Q1; HRs=1.07, 1.14, and 1.97; \(p_{\text{trend}}=0.036\)), but the association was not statistically significant in the multivariable-adjusted model (Q2, Q3, and Q4 vs. Q1; adjusted HRs=1.01, 1.14, and 1.28; \(p_{\text{trend}}=0.398\)). The levels of HbA1c, insulin, and HOMA-IR did not show a significant relationship with the risk of the occurrence of an advanced adenoma (\(p_{\text{trend}}=0.963, 0.822, \) and 0.822, respectively).

**Associations between parameters of lipid metabolism and adenoma occurrence**

Table 3 shows the risk for occurrence of any adenoma and advanced adenoma at the time of follow-up colonoscopy, according to quartiles of the parameters of lipid metabolism. The occurrence of any adenoma increased linearly with increasing quartiles of triglycerides, LDL-C, and ApoB. In the univariate analysis, the HRs for any adenoma occurrence when we compared the fourth with the first quartiles of triglycerides, LDL-C, and ApoB were 1.48 (95% CI, 1.25–1.76; \(p_{\text{trend}}=0.001\)), 1.26 (95% CI, 1.06–1.50; \(p_{\text{trend}}=0.013\)), and 1.35 (95% CI, 1.12–1.63; \(p_{\text{trend}}=0.004\)), respectively. However, in the multivariable-adjusted model, the associations between the markers of lipid metabolism and the occurrence of any adenoma remained significant only for triglycerides (Q2, Q3, and Q4 vs. Q1; adjusted HRs=1.02, 1.13, and 1.19; \(p_{\text{trend}}=0.041\)).

In contrast, the risk of the occurrence of an advanced adenoma did not show a significant relationship with any of the markers of lipid metabolism.

**DISCUSSION**

This large-scale, longitudinal study evaluated the associations between the parameters of glucose and lipid metabolism and adenoma occurrence detected during surveillance colonoscopy. We found that the levels of parameters of glucose and lipid metabolism at the time of the initial colonoscopy were significantly associated with adenoma occurrence detected during the surveillance colonoscopy. The occurrence of adenomas increased with increasing levels of all the parameters of glucose metabolism (including fasting glucose, HbA1c, insulin, and HOMA-IR) and triglycerides. These associations persisted after adjusting for confounding risk factors for adenomas. In addition, increasing levels of LDL-C and ApoB were significantly as-

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**Table 2. Risks of Adenoma and Advanced Adenoma Occurrence by Quartiles of Parameters of Glucose Metabolism**

| Markers of glucose metabolism | Any adenoma occurrence | Advanced adenoma occurrence |
|------------------------------|------------------------|----------------------------|
|                              | Crude HR (95% CI)      | Adjusted HR (95% CI)*      |
|                              |                        |                            |
| Fasting glucose (mg/dL)      |                        |                            |
| Q1 (57–88)                   | 1                      | 1                          |
| Q2 (89–94)                   | 1.27 (1.07–1.51)       | 1.20 (1.01–1.43)           |
| Q3 (95–100)                  | 1.41 (1.18–1.69)       | 1.33 (1.12–1.60)           |
| Q4 (101–271)                 | 1.84 (1.56–2.18)       | 1.50 (1.26–1.77)           |
| \(p_{\text{trend}}\)        | <0.001                 | <0.001                     |
| HbA1c (%)                    |                        |                            |
| Q1 (4.80–5.50)               | 1                      | 1                          |
| Q2 (5.51–5.60)               | 0.96 (0.79–1.17)       | 0.94 (0.77–1.15)           |
| Q3 (5.61–5.80)               | 1.05 (0.90–1.24)       | 1.00 (0.85–1.18)           |
| Q4 (5.81–12.70)              | 1.36 (1.16–1.59)       | 1.22 (1.04–1.43)           |
| \(p_{\text{trend}}\)        | <0.001                 | 0.024                      |
| Fasting insulin (mg/dL)      |                        |                            |
| Q1 (0.20–2.67)               | 1                      | 1                          |
| Q2 (2.68–4.01)               | 1.12 (0.94–1.34)       | 1.11 (0.93–1.32)           |
| Q3 (4.02–5.81)               | 1.15 (0.96–1.37)       | 1.08 (0.90–1.29)           |
| Q4 (5.82–37.88)              | 1.32 (1.11–1.57)       | 1.22 (1.02–1.46)           |
| \(p_{\text{trend}}\)        | <0.001                 | 0.046                      |
| HOMA-IR                      |                        |                            |
| Q1 (0.03–0.61)               | 1                      | 1                          |
| Q2 (0.62–0.94)               | 1.26 (1.05–1.50)       | 1.26 (1.05–1.50)           |
| Q3 (0.95–1.40)               | 1.17 (0.98–1.41)       | 1.13 (0.94–1.36)           |
| Q4 (1.41–18.34)              | 1.53 (1.28–1.82)       | 1.36 (1.14–1.63)           |
| \(p_{\text{trend}}\)        | <0.001                 | 0.004                      |

CI, confidence interval; HR, hazard ratio; HbA1c, hemoglobin A1c; HOMA-IR, homeostasis model assessment of insulin resistance.

*Estimated from Cox proportional hazards model adjusted for age, sex, smoking status, family history of colorectal cancer, use of nonsteroidal anti-inflammatory drugs, obesity, and initial colonoscopy finding (normal, low-risk, and high-risk groups).
associated with an increasing occurrence of adenomas, although the associations were not statistically significant in the multivariate analysis.

The incidence of CRC is rapidly increasing because of the spread of a Westernized diet and lifestyle, especially in Asian countries. Interest in identifying potentially modifiable risk factors has kept pace with the increase in the incidence of CRC. As part of these efforts, many recent studies have reported that diabetes, metabolic syndrome, dyslipidemia, and obesity are associated with an increased risk of CRC. Furthermore, several studies have evaluated the associations between the parameters of glucose and lipid metabolism and CRC risk in more detail, and researchers have reported that a high fasting glucose level, HbA1c, insulin level, HOMA-IR, triglyceride level, total cholesterol level, and LDL-C level, and a low HDL-C level are associated with an increased risk of colorectal adenomas. Our previous study showed similar results.

### Table 3. Risks of Adenoma and Advanced Adenoma Occurrence by Quartiles of Parameters of Lipid Metabolism

| Markers of lipid metabolism (mg/dL) | Any adenoma occurrence | Advanced adenoma occurrence |
|------------------------------------|------------------------|-----------------------------|
|                                    | Crude HR (95% CI)      | Adjusted HR (95% CI)*       | Crude HR (95% CI)      | Adjusted HR (95% CI)*       |
| **Triglycerides**                  |                        |                             |                        |                             |
| Q1 (28–78)                         | 1                      | 1                           | 1                      | 1                           |
| Q2 (79–111)                        | 1.17 (0.98–1.40)        | 1.02 (0.88–1.26)             | 1.62 (0.83–3.18)        | 1.35 (0.68–2.66)             |
| Q3 (112–160)                       | 1.31 (1.10–1.57)        | 1.13 (0.94–1.35)             | 1.30 (0.54–2.64)        | 1.01 (0.49–2.06)             |
| Q4 (161–1967)                      | 1.48 (1.25–1.76)        | 1.19 (0.99–1.42)             | 1.52 (0.76–3.03)        | 1.01 (0.50–2.07)             |
| \( p \) for trend                  | \(<0.001\)             | 0.041                       | 0.382                  | 0.758                       |
| **Total cholesterol**              |                        |                             |                        |                             |
| Q1 (98–180)                        | 1                      | 1                           | 1                      | 1                           |
| Q2 (181–201)                       | 0.99 (0.83–1.18)        | 0.96 (0.81–1.15)             | 2.15 (1.08–4.29)        | 2.11 (1.06–4.21)             |
| Q3 (202–226)                       | 1.04 (0.88–1.24)        | 0.95 (0.80–1.12)             | 1.39 (0.67–2.90)        | 1.16 (0.55–2.43)             |
| Q4 (227–433)                       | 1.18 (0.99–1.40)        | 1.11 (0.93–1.31)             | 1.48 (0.71–3.11)        | 1.44 (0.69–3.02)             |
| \( p \) for trend                  | 0.053                  | 0.307                       | 0.651                  | 0.842                       |
| **LDL cholesterol**                |                        |                             |                        |                             |
| Q1 (27–106)                        | 1                      | 1                           | 1                      | 1                           |
| Q2 (107–127)                       | 1.06 (0.89–1.26)        | 1.00 (0.84–1.20)             | 1.20 (0.59–2.44)        | 1.12 (0.55–2.28)             |
| Q3 (128–149)                       | 1.06 (0.89–1.27)        | 0.97 (0.81–1.16)             | 1.66 (0.86–3.22)        | 1.52 (0.78–2.95)             |
| Q4 (150–277)                       | 1.26 (1.06–1.50)        | 1.17 (0.98–1.39)             | 1.26 (0.62–2.59)        | 1.23 (0.60–2.53)             |
| \( p \) for trend                  | 0.013                  | 0.123                       | 0.338                  | 0.389                       |
| **HDL cholesterol**                |                        |                             |                        |                             |
| Q1 (20–44)                         | 1                      | 1                           | 1                      | 1                           |
| Q2 (45–51)                         | 0.98 (0.83–1.16)        | 1.02 (0.86–1.20)             | 0.83 (0.44–1.58)        | 0.90 (0.47–1.70)             |
| Q3 (52–60)                         | 0.90 (0.76–1.07)        | 0.93 (0.78–1.10)             | 0.82 (0.43–1.53)        | 0.83 (0.43–1.57)             |
| Q4 (61–136)                        | 0.91 (0.76–1.07)        | 1.04 (0.88–1.24)             | 0.81 (0.43–1.53)        | 1.03 (0.54–1.96)             |
| \( p \) for trend                  | 0.160                  | 0.964                       | 0.488                  | 0.930                       |
| **Apolipoprotein B**               |                        |                             |                        |                             |
| Q1 (23–79)                         | 1                      | 1                           | 1                      | 1                           |
| Q2 (80–94)                         | 1.18 (0.98–1.43)        | 1.10 (0.90–1.33)             | 1.37 (0.63–2.99)        | 1.25 (0.57–2.73)             |
| Q3 (95–109)                        | 1.10 (0.91–1.34)        | 0.97 (0.80–1.18)             | 1.41 (0.85–3.04)        | 1.16 (0.54–2.51)             |
| Q4 (110–215)                       | 1.35 (1.12–1.63)        | 1.18 (0.97–1.42)             | 1.60 (0.76–3.37)        | 1.26 (0.59–2.69)             |
| \( p \) for trend                  | 0.004                  | 0.199                       | 0.240                  | 0.622                       |
| **Apolipoprotein A1**              |                        |                             |                        |                             |
| Q1 (62–120)                        | 1                      | 1                           | 1                      | 1                           |
| Q2 (121–133)                       | 0.89 (0.74–1.07)        | 0.91 (0.76–1.09)             | 1.41 (0.71–2.80)        | 1.42 (0.71–2.83)             |
| Q3 (134–148)                       | 0.85 (0.71–1.02)        | 0.89 (0.74–1.07)             | 0.81 (0.38–1.76)        | 0.86 (0.40–1.88)             |
| Q4 (149–285)                       | 0.84 (0.70–1.01)        | 0.89 (0.74–1.07)             | 0.90 (0.43–1.89)        | 0.97 (0.46–2.06)             |
| \( p \) for trend                  | 0.055                  | 0.188                       | 0.445                  | 0.610                       |

CI, confidence interval; HR, hazard ratio; LDL, low-density lipoprotein; HDL, high-density lipoprotein.

*Estimated from Cox proportional hazards model adjusted for age, sex, smoking status, family history of colorectal cancer, use of nonsteroidal anti-inflammatory drugs, obesity, and initial colonoscopy finding (normal, low-risk, and high-risk groups).
date, there have been no studies evaluating the associations between these parameters and the occurrence of metachronous adenomas, which led us to investigate whether these parameters influence the occurrence of adenomas detected during surveillance colonoscopy.

We speculated that the parameters of glucose and lipid metabolism may affect the occurrence of adenomas detected during surveillance colonoscopy based on the results of recent studies carried out on the associations between metabolic factors and the occurrence of metachronous adenomas. Some studies demonstrated that the metabolic syndrome was a risk factor for developing metachronous adenomas at surveillance colonoscopy, while other studies showed that obesity, increased fasting glucose, and hypertension were associated with an increased risk of adenoma recurrence. As expected, our study showed that all of the parameters of glucose metabolism and some parameters of lipid metabolism influenced the occurrence of metachronous adenomas.

The pathophysiological mechanisms linking altered glucose metabolism, dyslipidemia, and CRN risk have several possible explanations: hyperinsulinemia can promote carcinogenesis through the effect of insulin-like growth factor-1, and dyslipidemia can increase the production of inflammatory cytokines such as interleukin-6 and tumor necrosis factor-α, creating a protumorigenic environment. Hyperglycemia and dyslipidemia can also promote the formation of reactive oxygen species, which can damage DNA and trigger cancer progression. Based on these biologic mechanisms, it is reasonable to conclude that altered glucose and lipid metabolism affects the occurrence of adenomas detected during surveillance colonoscopy, as well as adenoma prevalence.

Recently, meta-analyses have demonstrated that the use of metformin and statins is associated with a reduced risk of CRC and several studies also revealed that metformin and statins reduced the risk of adenoma. Furthermore, a previous study reported a potential benefit of metformin use for lowering the risk of subsequent adenomas after polypectomy. These results suggest that improving glucose and lipid levels through the administration of metformin and statins might be helpful in reducing the risk of CRN. In this study, we tried to analyze the risks of adenoma occurrence according to the changes in the levels of the parameters of glucose and lipid metabolism to identify whether worsening levels increased the occurrence of metachronous adenomas or improving levels reduced the occurrence of metachronous adenoma. However, we found no association between changes in the levels of all these parameters and the occurrence of adenoma. One of the reasons may be that the follow-up interval (mean 2.2 years) was too short to evaluate the impact of changes in the parameters on the occurrence of adenomas. Future longitudinal studies are warranted to clarify whether interventions to improve the parameters of glucose and lipid metabolism (such as diet control, exercise, weight reduction, or medication use) can reduce the development of metachronous adenomas.

Our study has a few limitations. First, the great majority of the study subjects were male and younger than 50 years old. Moreover, this was a retrospective study that included participants who underwent a colonoscopy at two centers in Korea. As a result, there was likely some degree of selection bias. Therefore, interpretation of our findings requires careful consideration when applied to other setting or populations. Second, the follow-up interval is too short to evaluate the risk for occurrence of adenomas, especially advanced adenomas. The reason why the parameters of glucose and lipid metabolism did not have a significant impact on the occurrence of advanced adenomas might be the short follow-up interval. In addition, some percentage of the adenomas found at the time of surveillance colonoscopy might have been missed during the initial colonoscopy. Third, we did not investigate withdrawal time that is important in complete colonoscopy although our medical examination center makes it a rule to withdrawal 6 minutes or more. Finally, dietary factors, physical activity, and medications such as metformin, which could all affect the occurrence of adenomas, were not considered.

In conclusion, higher levels of fasting glucose, HbA1c, insulin, HOMA-IR, and triglycerides at initial colonoscopy were associated with an increased occurrence of adenomas detected by surveillance colonoscopy. Improving the parameters of glucose and lipid metabolism through lifestyle changes or medications may be helpful in decreasing the development of metachronous adenomas.

REFERENCES

1. Lee HS, Jeon SW. Is retroflexion helpful in detecting adenomas in the right colon?: a single center interim analysis. Intest Res 2015; 13:326–31.
2. Lin OS, Kozarek RA, Cha JM. Impact of sigmoidoscopy and colonoscopy on colorectal cancer incidence and mortality: an evidence-based review of published prospective and retrospective studies. Intest Res 2014;12:268-74.
3. Lieberman DA, Rex DK, Winawer SJ, Giardiello FM, Johnson DA, Levin TR; United States Multi-Society Task Force on Colorectal Cancer. Guidelines for colonoscopy surveillance after screening and polypectomy: a consensus update by the US Multi-Society Task Force on Colorectal Cancer. Gastroenterology 2012;143:844-57.
4. Luo S, Li JY, Zhao LN, Yu T; Zhong W, Xia ZS, et al. Diabetes mellitus increases the risk of colorectal neoplasia: an updated meta-analysis. Clin Res Hepatol Gastroenterol 2016;40:110-23.
5. Kim BC, Shin A, Hong CW, Sohn DK, Han KS, Ryu KH, et al. Association of colorectal adenoma with components of metabolic syndrome. Cancer Causes Control 2012;23:727-35.
6. Liu CS, Hsu HS, Li CJ, Jan CJ, Li TC, Lin WY, et al. Central obesity and atherogenic dyslipidemia in metabolic syndrome are associated with increased risk for colorectal adenoma in a Chinese population. BMC Gastroenterol 2010;10:51.
7. Park H, Ko SH, Lee JM, Park JH, Choi YH. Troglitzazone enhances the apoptotic response of DLD-1 colon cancer cells to photodynamic therapy. Yonsei Med J 2016;57:1494-9.
8. Koku TO, Lund PK, Galanko J, Simmons JG, Woosley JT, Sandler
Insulin resistance, apoptosis, and colorectal adenoma risk. Cancer Epidemiol Biomarkers Prev 2005;14:2076-81.
9. Rampal S, Yang MH, Sung J, Son HJ, Choi YH, Lee JH, et al. Association between markers of glucose metabolism and risk of colorectal adenoma. Gastroenterology 2014;147:78-87.
10. Vidal AC, Lund PK, Hoyo C, Galanko J, Burcal L, Holst RN, et al. Elevated C-peptide and insulin predict increased risk of colorectal adenomas in normal mucosa. BMC Cancer 2012;12:389.
11. Ortiz AP, Thompson CL, Chak A, Berger NA, Li L. Insulin resistance, central obesity, and risk of colorectal adenomas. Cancer 2012;118:1774-81.
12. Misciagna G, De Michele G, Guerra V, Cistermino AM, Di Leo A, Freudenberg HJ; INTEROEP Group. Serum fructosamine and colorectal adenomas. Eur J Epidemiol 2004;19:425-32.
13. Yamada K, Araki S, Tamura M, Sakai I, Takahashi Y, Kashiwara H, et al. Relation of serum total cholesterol, serum triglycerides and fasting plasma glucose to colorectal carcinoma in situ. Int J Epidemiol 1998;27:794-8.
14. Yang MH, Rampal S, Sung I, Choi YH, Son HJ, Lee JH, et al. The association of serum lipids with colorectal adenomas. Am J Gastroenterol 2013;108:833-41.
15. Bayerdörffer E, Mannes GA, Richter WO, Ochsenkühn T, Sehölzer G, Kößpe W, et al. Decreased high-density lipoprotein cholesterol and increased low-density cholesterol levels in patients with colorectal adenomas. Ann Intern Med 1993;118:481-7.
16. van Duijnhoven FJ, Bueno-De-Mesquita HB, Calligaro M, Jenab M, Pischon T, Jansen EH, et al. Association between body size and colorectal adenoma recurrence. J Clin Gastroenterol 2013;47:174-8.
17. Jung YS, Ryu S, Chang Y, Yun KE, Park JH, Kim HJ, et al. Association between parameters of glucose and lipid metabolism and risk of colorectal neoplasm. Dig Dis Sci 2015;60:2996-3004.
18. Taniguchi L, Higurashi T, Uchiyama T, Kondo Y, Uchida E, Uchihashi Y, et al. Metabolic factors accelerate colorectal adenoma recurrence. BMC Gastroenterol 2014;14:187.
19. Jacobs ET, Martinez ME, Alberts DS, Jiang R, Lance P, Lowe KA, et al. Association between body size and colorectal adenoma recurrence. Clin Gastroenterol Hepatol 2007;5:5982-90.
20. Laiyemo AO, Doubeni C, Badurdeen DS, Murphy G, Marcus PM, Schoen RE, et al. Obesity, weight change, and risk of adenoma recurrence: a prospective trial. Endoscopy 2012;44:813-8.
21. So H, Han S, Park HW, Kim EH, Lee JY, Lee HS, et al. Metabolic factors affect the occurrence of colorectal neoplasm on surveillance colonoscopies. J Gastroenterol Hepatol 2016;31:1273-9.
22. Chiu HM, Lee YC, Tu CH, Chang LC, Hsu WF, Chou CK, et al. Effects of metabolic syndrome and findings from baseline colonoscopies on occurrence of colorectal neoplasms. Clin Gastroenterol Hepatol 2015;13:1134-42.
23. Lin CC, Huang KW, Luo JC, Wang YW, Hou MC, Lin HC, et al. Hypertension is an important predictor of recurrent colorectal adenoma after screening colonoscopy with adenoma polypectomy. J Chin Med Assoc 2014;77:508-12.
24. Kim MC, Jung SW, Kim CS, Chung TH, Yoo CI, Park NH. Metabolic syndrome is associated with increased risk of recurrent colorectal adenomas in Korean men. Int J Obes (Lond) 2012;36:1007-11.
25. Park HW, Byeon JS, Yang SK, Kim HS, Kim WH, Kim TI, et al. Colorectal neoplasm in asymptomatic average-risk Koreans: the KASID prospective multicenter colonoscopy survey. Gut Liver 2009;3:35-40.
26. Kitahara CM, Berndt SI, de González AB, Coleman HG, Schoen RE, Hayes RB, et al. Prospective investigation of body mass index, colorectal adenoma, and colorectal cancer in the prostate, lung, colorectal, and ovarian cancer screening trial. J Clin Oncol 2013;31:2450-9.
27. Ng SC, Wong SH. Colorectal cancer screening in Asia. Br Med Bull 2013;105:29-42.
28. Hsu YC, Chiu HM, Liu JY, Chang CC, Lin JT, Liu HH, et al. Glycated hemoglobin A1c is superior to fasting plasma glucose as an independent risk factor for colorectal neoplasia. Cancer Causes Control 2012;23:321-8.
29. Kim BJ, Kim YH, Sinn DH, Kang KJ, Kim JY, Chang DK, et al. Clinical usefulness of glycosylated hemoglobin as a predictor of adenomatous polyps in the colorectum of middle-aged males. Cancer Causes Control 2010;21:939-44.
30. Sasaki Y, Takeda H, Sato T, Orii T, Nishise S, Nagino K, et al. Serum Interleukin-6, insulin, and HOMA-IR in male individuals with colorectal adenoma. Clin Cancer Res 2012;18:392-9.
31. Otani T, Iwasaki M, Ikeda S, Kozu T, Saito H, Mutoh M, et al. Serum triglycerides and colorectal adenoma in a case-control study among cancer screening examinees (Japan). Cancer Causes Control 2006;17:1245-52.
32. Sun ZJ, Huang YH, Wu JS, Yang YC, Chang YE, Lu FH, et al. The association of serum lipids with the histological pattern of rectosigmoid adenoma in Taiwanese adults. BMC Gastroenterol 2011;11:54.
33. Shikata K, Ninomiya T, Kyohara Y. Diabetes mellitus and cancer risk: review of the epidemiological evidence. Cancer Sci 2013;104:9-14.
34. Esteve E, Ricart W, Fernández-Real JM. Dyslipidemia and inflammation: an evolutionary conserved mechanism. Clin Nutr 2005;24:16-31.
35. Cowlay S, Hardy R. The metabolic syndrome: a high-risk state for cancer? Am J Pathol 2006;169:1505-22.
36. Franciosi M, Lucisano G, Lapice E, Strippoli GF, Pellegrini E, Nicolicucci A. Metformin therapy and risk of cancer in patients with type 2 diabetes: systematic review. PLoS One 2013;8:e71583.
37. Liu Y, Tang W, Wang J, Xie L, Li T, He Y, et al. Association between statin use and colorectal cancer risk: a meta-analysis of 42 studies. Cancer Causes Control 2014;25:237-49.
38. Kim YH, Noh R, Cho SY, Park SJ, Jeon SM, Shin HD, et al. Inhibitory effect of metformin therapy on the incidence of colorectal advanced adenomas in patients with diabetes. Intest Res 2015;13:145-52.
39. Cho YH, Ko BM, Kim SH, Myung YS, Choi JH, Han JP, et al. Does metformin affect the incidence of colonic polyps and adenomas in patients with type 2 diabetes mellitus? Intest Res 2014;12:139-45.
40. Broughton T, Sington J, Beales IL. Statin use is associated with a reduced incidence of colorectal adenomatous polyps. Int J Colorectal Dis 2013;28:469-76.
41. Marks AR, Pietrofesa RA, Jensen CD, Zebrowski A, Corley DA, Doubeni CA. Metformin use and risk of colorectal adenoma after polypectomy in patients with type 2 diabetes mellitus. Cancer Epidemiol Biomarkers Prev 2015;24:1692-8.