High Sensitive Troponin as Biomarker for Coronary Artery Disease

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Abstract

There are various cardiac biomarkers available for diagnosis and stratification of patients with acute coronary syndrome but only few are cardiospecific and sensitive. Troponins are protein molecules that are part of cardiac and skeletal muscle. Smooth muscle cells do not contain troponins. There are three subunits of troponin T, I and C subunits. Cardiac troponins are known to increase on myocardial cell damage leading to myocardial necrosis. The kinetic pattern of release of troponin is known. The technological advancements in Immunoassay has led to improvement in the limit of detection, (LoD) Limit of quantitation (LoQ) and cut off detection limits. There are decreases in false positive cases due to antibody antibodies use of sandwich fifth generation Troponin T immunoassay. Recently the troponin T fifth generation by (Roche diagnostics) is approved by US-FDA. The cutoffs detections and positive 99th percentile of reference population cutoffs has to be established by verification of manufacturers reference cutoff s limits. The delta change value after serial measurements of suspected patients with symptoms of AMI leads to diagnosis.

Introduction

Cardiac troponins are highly sensitive and specific biomarkers of myocardial necrosis and are used for the diagnosis of acute myocardial infarction. Troponin is a specific cardiac marker for stratification of diagnosis of coronary artery disease. Troponin assay combined with clinical symptoms of chest pain and other diagnostic finding helps in diagnosis of STEMI and NSTEMI.

Troponin consists of three subunits-troponin I, troponin T, and troponin C. Each subunit has a function: Troponin T binds the troponin components to tropomyosin, troponin I inhibits the interaction of myosin with actin, and troponin C contains the binding sites for Ca2+ that helps initiate contraction. Cardiac troponins are the products of different genes consisting of immunological distinct entities. The total quantity of troponin is very less compared to myofibrillar protein actin and myosin. Cardiac isoforms of troponin T and Troponin I are known to be cardiospecific and are increased in myocardial damage. Release kinetics following MI show a first peak due to loosely bound troponin pool and a second prolonged elevation due to degradation of the contractile apparatus [1]. cTnT have characteristic higher molecular weight, higher fraction of unbound cTnI, less degradation, whereas cTnI is more frequently found as binary or tertiary complex in blood. Previous studies proves that early appearing troponin pool may give information on the quality of microvascular reperfusion, while the concentration of cTn on Day 3 or 4 reflects myocardial infarct size [2].

The National Academy of Clinical Biochemistry issued a guideline in 2007 that stated that “in the presence of a clinical history suggestive of ACS, the following is considered indicative of myocardial necrosis consistent with myocardial infarction an abnormal value is that above the 99th percentile of the healthy population as a cutoff using an assay with acceptable precision.

The 99th percentile cutoff point for cardiac troponin T (cTnT) is well-known at 0.01 ng/mL (with 10% coefficient of variance value at the 99th percentile of 0.03 ng/mL), at least one occasion during the first 24 h after the clinical event” [3].

Different manufactures assay are commercially available for cardiac troponin I (cTnI), so the 99th percentile cut point varies based on the assay being used. It's recommended for each local laboratory to verify the reference cut off limits of the 99th percentile of a reference decision limit (medical decision cutoff) for cardiac troponin (cTn) assays.

hs-cTn Assays

Rapid advancements in immunological assays have led to manufacturers to provide a traceable troponin calibration standards leading to improvement in analytical sensitivity and precision. Roche's Elecsys TrT Gen 5 STAT test was cleared by FDA recently is the first high sensitive troponin T assay. The fifth-generation blood test uses two monoclonal antibodies against cardiac troponin T to pick up the marker of myocardial damage with a turn-around time of 9 minutes were performed on Cobase 601 module of Roche fully automatic analyzer. The FDA clearance was based in part on a study of more than 1,000 patients with suspected acute MI in more than a dozen U.S. centers [4].

The hs-cTnT (fifth generation) assay cTnT assay uses fragment antigen-binding (FAB) of two cTnT-specific mouse monoclonal antibodies in a sandwich format. The capture antibody (M7) is biotinylated and directed against an epitope at amino acid residues 125–131 located in the central part of the cTnT molecule. The detection antibody is directed against an epitope at amino acid residues 136–147. Detection of cTnT is based on an electrochemiluminescence immunoassay using a Tris (bipyridyl)-ruthenium (II) complex as a label. The original antibody (M11.7) has been genetically reengineered, with the constant C1 region of the FAB...
being replaced by a human IgG C1 region to produce a mouse–human chimeric detection antibody [5]. The rationale for this replacement was to further reduce the susceptibility to interference by heterophilic antibodies. The analytical sensitivity was improved by increasing the sample volume from 15 µL to 50 µL, increasing the ruthenium concentration of the detection antibody, and lowering the background signal via buffer optimization. The fifth generation assay is calibrated against recombinant human cTnT produced in Escherichia coli cell culture. Assay calibration is different from that of the fourth-generation assay, so identical samples measured with the fourth-generation and hs-cTnT assays will give different results. As a result of these modifications in label and antibody, the analytic performance of the hs-cTnT assay was significantly improved; specifically, the LoD was 0.003 ng/mL (3 ng/L), the 99th percentile cut-off point was 0.014 ng/mL (14 ng/l), and the CV was 10% at 0.013 ng/mL (13 ng/L). Due to a lower LoD and a increased precision, the hs-cTnT assay is able to detect more subtle elevations indicative of cardiac injury [6].

With the advent of hs-assays and the emphasis on imprecision (% CV) of assays at the 99th percentile URL and the limit of quantification (LoQ), along with the increasing role of using the limit of blank (LoB) and the limit of detection (LoD) as cutoffs for early rule-out of AMI, understanding what these values mean is important all are analytical parameters used to describe the low concentrations of cardiac troponin measurements.

**Total Imprecision (% CV) at 99th Percentile**

Day-to-day imprecision of cardiac troponin assays is defined by the % CV and is determined using multiple lots of both reagents and calibrators over multiple days [7]. The CLSI EP5-A2 document details the evaluation protocol that spans 20 days, 2 repeats a day, with 2 different lots used for reagents and calibration materials. For clinical use, cardiac troponin assays have been deemed “guideline acceptable” if they have a % CV of ≤ 10% at the 99th percentile, "clinically usable" if the % CV is >10% to ≤ 20%, and "not clinically acceptable" if the % CV is ≥ 20% [6,7]. The hs-assays meet the highest standards of clinical practice guideline precision recommendations (% CV<10%) at the 99th percentile, whereas contemporary and POC cardiac troponin assays have a % CV between 10% and 20% at the 99th percentile. Using hs-cTnI assays decreases analytical noise, allowing reporting of real cardiac troponin increases above the 99th percentile indicative of myocardial injury, rather than increases in cardiac troponin resulting from analytical imprecision, thereby improving diagnostic accuracy.

**Statistical Approach to Define 99th Percentile**

Uniform, standardized statistical approach to calculate cardiac troponin 99th percentiles are required. The nonparametric method is a distribution-free method using ranks of observed values to determine percentiles for a given reference interval/cutoff. It is simple to calculate and easy to determine a sample size. For a 99th percentile and 90% CI, n ≥ 299 study participants are required, as recommended by the IFCC TF-CIB.

**Implementing Δ Cardiac Troponin Values**

There is no universal Δ value for cTnI or cTnT values to best optimize clinical specificity [8]. Deltas will be assay dependent and vary between a percentage change for contemporary and POC assays as compared to absolute concentration changes for hs-assays. Further, the calculation of Δ values will vary depending on the timing between serial samples, i.e., 0–1, 2 or 3 h (hs assays) vs 0 to 6 h (contemporary assays), as well as whether the initial cardiac troponin concentration is within the reference range below the 99th percentile (at the time of the disease evolution) or above the 99th percentile. Small concentration changes may also result from poor analytical imprecision for contemporary and POC assays that tend to have poor imprecision at low cardiac troponin concentrations, particularly those near the 99th percentile. A recommendation has been made for hs-cTnT to use a 50% change near the 99th percentile URL and a 20% change when the baseline value is increased above the 99th percentile within a 3 h interval [9]. Shorter time intervals (≤ 3 h) will be required with hs-cTn assays to assist in ruling out AMI.

**False Positive Cardiac Troponin Results**

**Analytical issues**

Falsely elevated troponin concentration due to analytical interference caused by various interferents. Fibrin clots in serum as a result of incompletely clotted specimen, e.g. in patients with coagulopathy or on anticoagulant therapy so Plasma is commonly used as specimen for analysis, heterophile antibodies, human anti-animal antibodies decreased due to use of higher generation immunonassey with improved specificity, rheumatoid factor, and autoantibodies, interference from other endogenous components in the blood such as bilirubin and hemoglobin interfere with absorbance, immunocomplex formation, microparticles in specimen, high concentration of alkaline phosphatase, analyzer malfunction are some of the commonly known (Table 1) [10-12].

| Cardiac Cause | Noncardiac Causes |
|---------------|-------------------|
| Cardiac contusion resulting from trauma | Pulmonary embolism |
| Cardiac surgery | Severe pulmonary hypertension |
| Cardioversion | Renal failure |
| Endomyocardial biopsy | Stroke, subarachnoid hemorrhage |
| Acute and chronic heart failure | Infiltrative diseases, eg, amyloidosis |
| Aortic dissection | Cardiotoxic drugs |
| Aortic valve disease | Critical illness |
| Hypertrophic cardiomyopathy | Sepsis |
| Tachyarrhythmia | Extensive burns |
| Bradyarrhythmia, heart block | Extreme exertion |
| Apical ballooning syndrome | |
| Post-‐percutaneous coronary intervention | |
| Rhabdomyolysis with myocyte necrosis | |
| Myocarditis or endocarditis/pericarditis | |

**Table 1:** Causes of Elevated Plasma Cardiac Troponin Other Than Acute Coronary Syndromes [13].

**Implementation of hs-Assays**

The following draft check-list, derived from the evidence based literature, will help educate laboratories currently using and prepare

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laboratories not using hs-assays for the conversion from contemporary and POC assays to hs-assays. First, education of laboratory medicine and clinical staff on relevant literature pertaining to the manufacturers’ assay being implemented will be necessary. Distribute analytical, diagnostic and risk assessment outcomes studies to your clinical colleagues, and present information on the new hs-assay at their staff meetings and clinical conferences. Second, develop understanding among users of the new hs-assay that the concentration numbers are going to change, and that they should not expect a conversion factor. Third, a URL at the 99th percentile will need to be established, following the IFCC TK-LB educational materials. Fourth, preparation will be needed for changing from a single to sex-specific 99th percentile, recognizing that this value for women will be less than for men. Fifth, conversion to reporting only whole numbers in ng/L will be needed. Sixth, define a QC material at the 99th percentile to monitor %CV. Seventh, consider using cardiac troponin values LoD of the hs-assay as a potential rule-out characteristic. Eighth, provide serial testing protocols that consider earlier measurements such as at baseline, 1.5 h and 3 h for diagnostic determinations. Ninth work on assuring good preanalytical sampling as the hs-assays are so sensitive that poor sample quality can be a problem [14,15].

References

1. Thygesen K, Mair J, Katus H, Plebani M, Venge P, et al. (2010) Recommendations for the use of cardiac troponin measurement in acute cardiac care. Eur Heart J 31: 2197-2204.
2. Giannitsis E, Steen H, Kurz K, Ivandic B, Simon AC, et al. (2008) Cardiac magnetic resonance imaging study for quantification of infarct size comparing directly serial versus single time-point measurements of cardiac troponin T. J Am Coll Cardiol 51: 307–314.
3. Wu AH, Apple FS, Gibler WB, Jesse RL, Warshaw MM, et al. (1999) National Academy of Clinical Biochemistry Standards of Laboratory Practice: recommendations for the use of cardiac markers in coronary artery diseases. Clin Chem 45: 1104-1121.
4. Apple FS, Collinson PO, IFCC Task Force on Clinical Applications of Cardiac Bio-Markers (2012) Analytical characteristics of high-sensitivity cardiac troponin assays. Clin chem 58: 54-61.
5. Apple FS, Jaffe AS (2012) Clinical implications of a recent adjustment to the high-sensitivity cardiac troponin T assay: user beware. Clin Chem 58: 1599-1600.
6. Apple FS (2009) A new season for cardiac troponin assays: it’s time to keep a scorecard. Clin Chem 55: 1303-1306.
7. Apple FS, Jaffe AS, Collinson P, Mockel M, Ordonez-Llanos J, et al. (2015) IFCC educational materials on selected analytical and clinical applications of high sensitivity cardiac troponin assays. Clin Biochem 48: 201-203.
8. Thygesen K, Mair J, Giannitsis E, Mueller C, Lindahl B, et al. (2012) How to use high-sensitivity cardiac troponins in acute cardiac care. Eur Heart J 33: 2252-2257.
9. Kricka LJ (1999) Human anti-animal antibody interferences in immunological assays. Clin Chem 45: 942-956.
10. Kricka LJ, Schmerfeld-Pruss D, Senior M, Goodman DB, Kaladas P (1990) Interference by human anti-mouse antibody in two-site immunoassays. Clin Chem 36: 892-894.
11. Mahajan VS, Jarolim P (2011) How to Interpret Elevated Cardiac Troponin Levels, Circulation 124: 2350-2354.
12. Mahajan VS, Jarolim P (2011) How to interpret elevated cardiac troponin levels. Circulation 124: 2350-2354.
13. Apple FS, Sandoval Y, Jaffe AS, Ordonez-Llanos J (2017) Cardiac Troponin Assays:Guide to Understanding Analytical Characteristics and Their Impact on Clinical Care. Clinical Chemistry 63: 173-181.