Review

Natural cell based biomimetic cellular transformers for targeted therapy of digestive system cancer

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Abstract

Digestive system cancer is the most common cause of cancer death in the world. Although cancer treatment options are increasingly diversified, the mortality rate of malignant cancer of the digestive system remains high. Therefore, it is necessary to explore effective cancer treatment methods. Recently, biomimetic nanoparticle delivery systems based on natural cells that organically integrate the low immunogenicity, high biocompatibility, cancer targeting, and controllable, versatile functionality of smart nanocarrier design with natural cells have been expected to break through the bottleneck of tumor targeted therapy. In this review, we focus on the dynamic changes and complex cellular communications that occur in vivo in natural cells based vehicles. Recent studies on the development of advanced targeted drug delivery systems using the dynamic behaviors such as specific surface protein affinity, morphological changes, and phenotypic polarization of natural cells are summarized. In addition to drug delivery mediated by dynamic behavior, functional “delivery” based on the natural cell themselves is also involved. Aiming to make the best use of the functions of cells, providing clues for the development of advanced drug delivery platforms.

Key words: Natural cell, Biomimetic cellular transformers, Digestive system cancer, Cancer targeting, Transformation

Introduction

Digestive system tumors mainly include esophageal cancer, gastric cancer, colorectal cancer (CRC), pancreatic cancer, hepatocellular carcinoma (HCC) and biliary tract cancer, and are the most common cause of cancer death worldwide. Its morbidity and mortality are in the forefront of malignant tumors [1]. There were nearly 6 million new cases of digestive system malignancies worldwide in 2020 [2]. At present, the main treatment methods for early and mid-stage digestive system cancer are still surgery and postoperative adjuvant chemoradiotherapy. The most common cancer treatment strategy is chemotherapy. However, chemotherapeutic drugs often have disadvantages such as poor water solubility, low tumor targeting ability, and severe adverse reactions, which greatly limit their clinical applications [3]. At the same time, because most cases are found in the late stage of the disease, the effective treatment methods are very limited, resulting in a high fatality rate [4-7]. Therefore, exploring new therapeutic methods for digestive system cancer has become a hotspot of current research.

Through decades of research, nanotechnology has shown great potential in diagnosing and treating various diseases [8,9]. However, the immune system distinguishes most synthetic nanocarriers as foreign substances and eliminates them [10,11]. Surface modification of nanoparticles can prolong their circulation time in blood and achieve specific
targeting, but this complicates their preparation [12]. As a result of their dynamic interactions or information exchanges with body systems, natural cells are emerging as the optimal drug delivery platform [13].

Additionally, the cell surface has specific signaling molecules, which can drive to the disease site, and can also dynamically alter its own morphology, secretion capacity or polarization state according to endogenous or exogenous factors [14]. The process of dynamic transformation can occur naturally, such as self-extrusion of red blood cells when passing through narrow capillaries, morphological changes of neutrophils passing through intercellular spaces and the release of extracellular traps, extracellular membrane vesicles with parent cell genetic material and function secreted by cells of different origins, as well as the interconversion and homeostasis of various phenotypes of macrophages in vivo [15]. On the other hand, from the perspective of “cargo”, in addition to specific substances, various functional components of the cell itself can likewise be delivered as “cargo”, and the resulting functional transmission is also particularly important in intercellular information communication. The transmission of these functions is usually carried out by the transport of extracellular vesicles, such as information transmission driven by molecular messengers such as eicosanoids, cytokines and various chemokines. In lipid bilayers, fluidity and cytoskeletal dynamics dictate the dynamics of processes and their role in the vast majority of physiopathological processes in the cell is essential [16-19]. Furthermore, the unique dynamic transformation properties of natural cells combined with advanced bioconjugation chemistry, nanotechnology and biotechnology can provide clues for the development of efficient drug delivery platforms [20,21].

Here, we review dynamic drug delivery systems which use cells with transformable behavior instead of conventional nano- or cellular carriers. In this review, four categories of dynamic carriers of cells are covered according to their transformation behavior (Figure 1): morphological deformation of erythrocytes and neutrophils, vesicle secretion, macrophage polarization, and semi-biological hybrid cells, to promote further progress in this field.

1. Morphology deformation-based cellular carriers

Physiological and pathological changes of the body often cause changes in the number or quality (function) of blood cells, and then lead to anemia, fever, infection, bleeding and vascular embolism and other clinical symptoms, and seriously affect the communication of information and material transfer between cells [22,23]. In the microvascular system, when red blood cells flow through narrow capillaries, due to shear stress, red blood cells will undergo compression and deformation in capillaries, thus promoting material exchange between red blood cells and surrounding tissues [24,25]. In tumor microenvironment (TME), neutrophils release cytokines...
and other special messengers to enhance the ability of tumor cells to metastasize [26]. Subsequently, neutrophils are activated by inflammation and TME to metastasize and accumulate in tumor tissue [27]. In addition, neutrophils have multiple receptors on their surfaces, thus facilitating drug enrichment at the tumor site when neutrophils’ membranes interact with chemokines and adhesion factors in the tumor microenvironment [28].

1.1. Red blood cell membrane

The red blood cells (RBCs) in our bodies circulate for a long time, lasting about 100 to 120 days [29]. The long-term circulation effect of erythrocytes is mediated by a series of membrane proteins on the surface of the cell membrane, among which integrin-related protein CD47 plays a key role [30]. CD47 is expressed on erythrocyte membrane and, as a self-protective protein, can recognize signal regulatory protein-α (SIRP-α) on macrophage membrane and send inhibitory signals to prevent phagocytosis of macrophages [31,32]. In addition, the double concave disc-shaped structure of mature red blood cells is anuclear and less rigid structure, so it is easier to deform than other cells [33]. When the osmotic pressure of the surrounding medium decreases, RBCs can become cup-shaped and finally form a sphere. This swelling characteristic is the key to combine RBCs with drugs or other chemicals [34]. The modification of traditional nanoparticles (NPs) on the surface of red blood cell carrier is a new bionic strategy to improve the half-life of drugs in vivo.

The surface area of a single RBC membrane reaches 160 μm², which can be used as a good drug loading platform, and the conjugation of drugs onto the RBCs membrane can reduce the damage to the structure and function of RBCs to some extent [35]. It can not only load various small molecular compounds, but also macromolecular compounds such as proteins and nucleic acids, and its membrane fluidity provides better flexibility when passing through narrow capillaries [36,37]. However, the high specificity of conjugating RBCs to drugs by means of membrane surface attachment leads to a lower range of potential indications. At present, the major forms of conjugation currently include the erythropoietin receptor (EPOR) mediated linkage [38], the RBCs complement receptor type I (E-CR1) mediated linkage, the avidin-biotin mediated linkage [39], and linkage via chemical bonds. Magnani et al. found that the cell activity of biotin acylation by biotin N-hydro-succinimide ester (NHS biotin) is the best, the recovery rate of RBCs can reach more than 90%, about 1000 biotins are connected to each RBCs membrane, and the activity is not affected for 24 hours in vivo [40]. Glutaraldehyde, as a commonly used cell cross-linking and fixative agent, can cross-link with proteins on the cell membrane and increase its stability, thereby avoiding RBCs hemolysis, and also slow down the drug loss from the cell, while having some targeting properties [41]. Animal experiments confirmed that RBCs carrier treated with high concentrations of glutaraldehyde could target to the liver, whereas those treated with low concentrations of glutaraldehyde target to the spleen. However, the osmotic fragility of the RBCs carrier treated by this method is increased, the cell deformability is reduced, and it is difficult to pass through the narrow capillaries. Band 3 protein (Band³) cross-links into clusters on the membrane of senescent or degenerated RBCs, promotes the connection of IgG, and is more easily phagocytosed by macrophages. Therefore, RBCs carriers treated with Band³ cross-linking reagent have macrophage targeting properties. Lotero et al. treated RBCs carriers carrying etoposide with bis(sulfosuccinimidyl)suberate (BS²) and in vitro results showed that macrophage phagocytosis increased from 3% to 11%. The in vivo results showed that the treated RBCs carriers were liver-targeted due to the strong attachment of the Band³ to macrophages [42].

In addition to using cross-linking agent to couple drugs to RBCs membrane, adsorbing drugs to RBCs membrane is also an effective drug loading strategy. Brenner et al. developed a “red blood cell hitchhiking (RH)” technology, which adsorbs drug loaded NPs onto RBCs and then infuses them to patients through intravenous injection or arterial intubation (Figure 2a). After entering the blood circulation, the NPs are desorbed from the RBCs membrane under the influence of blood flow shear stress or in direct contact with endothelial cells (Figure 2b). The results show that RH can deliver NPs to various types of cells on the surface of capillary lumen, and the type of cells to which NPs are delivered depends on the pathophysiological environment. This new drug delivery technology can accurately deliver nano drug carriers to selected organs by placing intravascular catheters in animal or human lungs, while avoiding RBCs hemolysis or toxicity to target organs, and has good biocompatibility [43].

In addition to oxygen transport, RBCs have a variety of additional immunological related functions. Researchers have found that RBCs can bring immune complexes and bacteria from the circulatory system to liver macrophages and professional antigen presenting cells (APCs) in the spleen by capturing them on their surface. Through exploiting RBC’s unique ability to present antigens in the spleen, Anvay Ukidve et al. developed strategies to generate
cellular and humoral immune responses against antigens. Researchers designed a RBCs hitchhiking system to mainly deliver the attached NPs to the spleen but not the lungs, thereby activating cellular immunity and humoral immunity, and they termed this process as erythrocyte-driven immune targeting (EDIT). Specifically, the researchers coat surfaces of polystyrene carboxylate (PS-COOH) with ovalbumin (OVA) as a model antigen using 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (EDC) chemistry to generate protein-capped NPs attached to RBCs. Dendritic cells found all particles to be monodisperse and easily internalized and activated by them. As mentioned earlier, particulate matter can be removed from the lungs with RH technology because shear stress is experienced by RBCs undergoing deformation in the lung capillaries. Increasing the NPs to RBCs dosing ratio helps to improve the shear resistance in the lungs, thus allowing more NPs to be targeted to the spleen or other sites [44].

Despite improving the targeting and safety of drug delivery, RH strategy still has some problems: 1) NPs are not firmly connected to RBCs and easy to undergo desorption. 2) At present, the use of RH strategy to achieve the targeting of organs such as heart and brain requires the aid of arterial vascular cannulation, which is not suitable for chronic, outpatient treatment of diseases [45]. 3) Construction of hitchhiking RBCs drug loading systems requires multiple steps such as RBCs isolation, ex vivo loading and reinfusion of RBCs, or infusion of donor RBCs, each of which can have an impact on the final yield. The adhesion of NPs to RBCs is the basis of the RH strategy, but due to the small surface area to volume ratio of RBCs, there is also the problem that the drug loading is difficult to increase [46]. 4) Currently, there is a lack of studies on the distribution of NPs transferred from RBCs carrier to the vascular cells in the vasculature of addressee organs [47]. 5) In the RH strategy, NPs nonspecifically attach to random components of the RBCs membrane, which may affect the normal function of RBCs. However, a spectrum of affinities to the RBCs may also arise. Understanding this aspect and designing selective coupling of NPs to carrier RBCs may enable specific transfer to intended vascular areas and cells [47,48]. Although the application of RH strategy in clinic is still immature, its advantages still cannot be ignored. By adjusting the ratio of NPs to RBCs, a drug delivery system targeting different organs may be successfully constructed. Besides, using the innate immune function of RBCs to design NPs for transfer to the spleen provides an alternative format for delivering NPs to the spleen, avoiding excessive modification of NPs. The immune memory induced by EDIT can continuously drive therapeutic responses. As a general strategy, combining RH strategy with tumor immunotherapy may provide new hope for treating solid tumors such as digestive system cancers.

1.2 Neutrophils cell membrane

Neutrophils account for about 50% ~ 70% of the total number of leukocytes and are the most abundant leukocytes in human body [49]. As the first responder of inflammatory signals, neutrophils will be “captured” by selectin (SEL) synthesized by vascular endothelial cells when flowing rapidly in the blood, making it roll slowly along the inner surface of blood vessels, detecting inflammatory signals such as complement fragment C5a and lipopolysaccharide (LPS), and then rapidly expressing integrin (INT). INT interacts with intercellular cell adhesion molecule-1 (ICAM-1) expressed on the surface of endothelial cells. Then a chemokine signal causes neutrophils to stop rolling and migrate toward the site of inflammation by passing through the vascular endothelium [50]. Although the spacing between endothelial cells is not enough for a cell to pass through, neutrophils can be deformed by squeezing itself through the cell gap. This is also known as

Figure 2. Clinically translatable nanocarriers adsorb onto red blood cells. (a) Procedural steps of RBC hitchhiking. NCs are first adsorbed onto the RBCs ex vivo. (b) Scanning electron micrographs of PS-NPs and nanogels attached to the surface of murine RBCs. Adapted with permission from [43]. Copyright Year 2018, Springer Nature.
leukocyte extravasation. However, excessive neutrophils infiltration is also the main reason for the continuous deterioration of some diseases. Researchers have found neutrophils infiltration in a variety of tumor models [51-55]. Chemokines and cytokines produced by tumor cells and the surrounding microenvironment will actively recruit neutrophils. Neutrophils express chemokine receptors CXCR1 and CXCR2 [56], and tumor cells express the ligands of these receptors, thereby promoting the recruitment of tumor associated neutrophils. Neutrophils can also interact with circulating tumor cells (CTCs) through adhesion molecules on cells and neutrophil extracellular traps (NETs) to promote tumor metastasis [57,58]. Therefore, neutrophils can be used as a good provider of tumor targeted cell membrane biomimetic materials [59].

The use of neutrophils to deliver drugs or NPs generally takes two forms. The first is to assemble NPs into neutrophils in vitro. However, in practical applications, due to the short lifespan (~ 7 h) of neutrophils, intracellular degradation of drugs, insufficient number of obtained cells, and the potential for in vitro contamination, these methods are not effective [60]. The second is to use NPs to “hitchhiking” neutrophils in situ (i.e., during blood circulation). Although this approach is more beneficial to clinical practice, it is crucial that NPs are designed to have high affinity for neutrophils. Li et al. constructed a strategy based on pathogen mimicking nano-pathogenoids (NPNs) to “hitchhiking” circulating neutrophils, and combined photothermal therapy to enhance the recruitment of neutrophils by NPNs from the blood circulation and the ability to eliminate residual tumor cells [61]. The researchers first constructed NPNs by coating bacteria-secreted outer membrane vesicles (OMVs) collected from E. coli on NPs by extrusion method. During photothermal therapy (PTT), the chemokines produced stimulate the neutrophils and inform them of the tumor’s location. Pathogen-associated molecular patterns (PAMPs) on NPNs are sensed and recognized by pattern recognition receptors (PRR) on neutrophils to “hitchhiking” circulating neutrophils, on the basis of which biological barriers are overcome and actively accumulate at the tumor site. After NPNs enter tumor tissue, in the process of forming neutrophil extracellular traps (NETs), NPNs are released from neutrophils when plasma membranes are destroyed and internalized by tumor cells to exert anti-tumor effect (Figure 3). Real-time intravital microscopy was used to examine the migration patterns of neutrophils in PTT-treated tumors, thus confirming that chemotaxis of neutrophils to tumors can overcome certain biological drug delivery barriers. Within 2 hours after PTT, a large number of neutrophils aggregated into tumors. In enlarged region I, the shape of neutrophils changed from oval to irregular as it moved slowly along blood vessels, sometimes adhering to vessel walls. In enlarged region II, when neutrophils are elongated, they can move more freely in small capillaries, allowing them to be evenly distributed throughout the tissue. In addition, neutrophils on blood vessels also enter the interstitium by elongating and squeezed through endothelial cells. The above results all indicated that the recruited neutrophils could cross the vascular barrier in a dynamic deformation manner and increase the distribution in the tumor tissue interstitium. The researchers further induced an inflammatory environment with PMA medium and verified that the encapsulated drugs could be released from neutrophils. The mechanism of its release lies in the dynamic deformation of activated neutrophils and eventually form NETs, which indicates the plasma membrane of neutrophils has disintegrated, thereby leading to the final release of the drug. Further results from in vivo experiments confirmed that PTT-induced inflammation led to the release of encapsulated NPs by NETs, followed by its eventual endocytosis by tumor cells. Usually, as a result of the limited penetration of light through tumor tissue, the uneven distribution of heat simply retards tumor growth but does not eliminate it completely, the result was a recurrence and metastasis of the tumor. By combining PTT and chemotherapeutic drugs, NPNs dislodge circulating tumor inducing neutrophils in situ, thereby eradicating tumors in all treated mice. This method also provides new ideas for improving nanomedicine delivery and tumor therapy [61].

1.3 Stem cell membrane

Stem cells are a type of pluripotent cells with self-replication ability. Studies have shown that cytokines and chemokines secreted by tumor cells can induce the enrichment of stem cells into tumor tissue [62]. Therefore, many types of stem cells, such as bone marrow-derived mesenchymal stem cells (BMSCs), can be used to construct tumor targeted nanomedicines. MSCs have a strong ability to orientate malignant lesions, that is, the homing characteristics of stem cells. When ischemic, hypoxic, or injured, body endogenous or exogenous stem cells have the trait of directional migration toward the site of injury and distribution [63]. The homing mechanism of stem cells is a multi-step and coordinated process, which mainly involves chemokine receptor interactions, adhesion of endothelial cells, and so on [64]. Chemokines released by tumor and tumor associated stromal cells attract BMSCs with corresponding
receptors against a concentration gradient to reach the tumor site [65]. After stem cells are mobilized and migrated, their vascular cell adhesion molecule (VCAM) and intercellular adhesion molecule (ICAM-1) interact with endothelial cells to allow stem cells to adhere to the capillary wall for targeting across the endothelial cell layer to the tumor site by extrusion deformation [66]. MSCs are easy to isolate and culture in vitro, and encapsulating nanoparticles with stem cell membrane can endow the nanocarrier with active targeting property, which can specifically accumulate in the tumor site, reduce the damage to the body’s normal tissues and improve the therapeutic efficacy.

Figure 3. Schematic illustration showing the chemotaxis-driven delivery of NPNs for complete eradication of tumors post-phototherapy. 

#1a The released G-CSF and GM-CSF increased neutrophil production from bone marrow. 

#1b The released CXCL1 and MIP-2 broadcasted the location of the inflamed tumor. 

#2 Neutrophils entered the blood circulation and encountered the injected NPNs. 

#3 Neutrophils sensed NPNs with the recognition of LPS and lipoprotein by TLRs and subsequently engulfed them. 

#4 Neutrophils laden with NPNs were recruited into the tumor site in response to the chemokine gradient through the following cascade: adhesion, crawling and transmigration. 

#5 NPNs were released from neutrophils to kill tumor cells along with the formation of NETs in the inflamed tumor. Adapted with permission from [61]. Copyright Year 2020, Springer Nature.
To reduce the systemic toxicity of doxorubicin (DOX), Liu et al. constructed a MSCs membrane encapsulated DOX superparamagnetic iron oxide (SPIO) NPs for colorectal cancer treatment. DOX-SPIO@MSCs with the biological function of MSCs membrane, thus achieving highly efficient delivery of intracellular drugs, prolonging serum half-life, and actively targeting to the tumor site. Compared with free DOX and DOX-SPIO, DOX-SPIO@MSCs can not only avoid the premature clearance of drugs by the mononuclear phagocyte system, but also have stronger targeting to the MC38 xenograft tumor. The anti-tumor treatment efficiency was significantly improved without obvious systemic toxicity [67]. Compared with BMSCs, human umbilical cord derived MSCs are popular sources because of their stemness, ease of acquisition, high rate of proliferation, and provision of abundant plasma membranes. Yang et al. noted that PLGA NPs encapsulated by umbilical cord derived MSCs could enhance the uptake of DOX, and the results showed that the uptake efficiency of membrane coated NPs by tumor cells was 3 times higher than that of membrane uncoated NPs. Moreover, encapsulation of umbilical cord derived MSCs improved the release efficiency of DOX at acidic microenvironment and inhibited MHCC97H subcutaneously tumor growth in nude mice with liver cancer [68]. Notably, stem cells exhibit different growth impacts for different tumor models, suggesting that the targeting ability of stem cell membrane biomimetic NPs may be tumor-specific and not applicable to all tumor types [69]. At the same time, it is still necessary to further confirm whether the stem cell carrier has the risk of promoting tumor growth or even metastasis, so its clinical application still needs further exploration.

2 Cellular dynamic release-based delivery

Under physiological and pathological conditions, almost all kinds of microorganisms or mammalian cells will dynamically release extracellular vesicles (EVs). EVs are diverse membranous vesicles derived from endosomes and cell membranes released by cells into the extracellular environment [70]. As early as 1976, Allan et al. found that RBCs release EVs from their surface [71]. EVs can not only participate in the transport of biomolecules such as lipids, proteins and RNA between cells, but also mediate the transmission of information between cells in biological system [72-74]. EVs can be divided into exosomes, microvesicles (MVs), apoptotic bodies, synaptoneurosomes and other types [75,76]. The diversity of EVs stems from its different production mechanisms [77]. Because EVs mediate the communication between cells, have the ability of material loading and certain targeted homing ability, and have high biocompatibility and can exist stably in vivo, research on EVs and drug delivery has increased gradually in the recent years [78]. Therapeutic drugs can be loaded into EVs in various ways. A comparison of the advantages and disadvantages of each drug loading method is shown in Table 1. In addition, EVs have lipid bilayers, so lipid-soluble molecules can be anchored and inserted into the membrane. Combined with cell membrane surface modification technology, functionalized EVs can be obtained, which further expands the application of EVs as drug carrier [79]. This part will focus on the bionic drug delivery system based on the dynamic release of these cells.

| Drug loading methods | Advantages | Disadvantages | Ref |
|----------------------|------------|---------------|-----|
| Freeze thaw          | Medium drug loading efficiency and low cost of materials | Protein degradation, irreversible change of EVs membrane | [80] |
| Co-incubation        | Simple operation, no large equipment required, no damage to membrane integrity | Affects the size of EVs, results in low yield, low entrapment and uncontrollable drug loading | [81,82] |
| Electroformation     | Load macromolecular substances or hydrophilic molecules into EVs | Molecular aggregation, EVs structural instability | [83] |
| Extrusion            | High drug loading efficiency | Destroy membrane integrity and alter membrane properties | [84] |
| Saponin assisted encapsulation | Highest drug loading efficiency | Presence of toxicity | [85,86] |
| Sonication           | Suitable for biological molecules like small RNAs, high drug loading efficiency | Membrane dysregulation, with incomplete drug release, non-suitability for hydrophobic drug delivery | [82] |
| Click chemistry      | Fast and efficient, with strong binding site controllability | Potential Toxicity of Chemical Crosslinkers | [82] |
| Genetic engineering of parental cells | Allows the loading of RNA, DNA, and peptides of choice into the EVs | High cost and low efficiency, choice of specific EVs derived cells is an associated limitation | [87,88] |
| Noncovalent binding and Hydrophobic inlay | Can dissociate by changes in chemical shift, temperature, or solvent | Too costly to be synthesized in bulk | [89,90] |
| Microfluidics technology | Preservation of EVs integrity, stability maintenance, high throughput precision, and low sample volume | Sometimes polymeric material or non-EVs ingredient causes co-precipitation along with EVs | [82] |

2.1 Extracellular vesicles of animal origin as dynamic carriers

EVs is a highly heterogeneous vesicle [91]. The release of EVs is considered to be an important regulator of cell-cell interaction and has a wide range of physiological functions [92]. It can be used as antigen to bind to receptor cells, and can also be used
as carrier to deliver functional substances to receptor cells, such as protein, nucleic acid, DNA, mRNA and non-coding RNA. Receptor cells can interact with EVs through several active uptake modes, including membrane fusion, receptor-mediated endocytosis, endocytosis and phagocytosis [93]. The characteristics of EVs are different depending on the source cells [94]. The EVs derived from mesenchymal stem cells (MSCs) can transfer many components such as genetic material and drugs. Their homing ability to inflammatory sites can also have a variety of effects on tumor growth [95]. Embryonic stem cells (ESCs) can remain undifferentiated for a long time in the process of in vitro culture, so as to generate EVs with stable characteristics, which can be used as drug carriers for the development of new cancer therapeutic drugs. Macrophage derived EVs can penetrate the blood-brain barrier, interact with and accumulate in cancer cells [96]. Tumor derived EVs have also attracted the attention of many researchers. Its tumor targeting homing property enables drugs to reach cancer cells effectively, which not only protects therapeutic drugs from degradation, but also has low immunogenicity [97]. Dendritic cell-derived EVs contain a variety of antigen-presenting molecules and costimulatory molecules, which can activate T cells, enhance the function of natural killer cells, and promote the eradication of tumors [98]. In addition to the above characteristics, the main advantage of EVs as drug carriers is that they can effectively load therapeutic drugs without significantly changing membrane structure and surface proteins, and have stability, biocompatibility, targeting and the ability to overcome natural barriers in blood circulation. They also have immune tolerance and can overcome the limitations of synthesizing NPs. Based on the above advantages, extracellular vesicles have become a promising tool for drug delivery [92,99-101].

Inspired by EVs mediated intercellular communication, Silva et al. constructed a hybrid carrier consisting of macrophage derived EVs encapsulating different therapeutic agents and iron oxide NPs to achieve spatial control of drug delivery. Studies have found that under a magnetic field, the uptake of EVs by tumor cells can be dynamically regulated and spatially controlled, and that magnetic targeting enhances tumor cell death with attenuated toxic side effects [102].

With endogenous exosomes, immune reactions can be avoided. Nevertheless, the efficiency of exosome delivery depends on both the parent and the recipient cells, and native exosomes have difficulties locating their target cells [103]. To meet the experimental needs, the technology of constructing engineered exosomes by surface modification has emerged [104]. To precisely target therapeutic drugs to lesions, chemical attachment of targeting peptides, genetic engineering of exosome membranes, magnetic nanoparticle technology, electrostatic interactions and post-insertion have become a common method in recent years [105,106]. In order to produce the precise and improved therapeutic effect, Tamura R et al. modified exosomes with cationic amylopectin to target asialoglycoprotein receptors on hepatocytes [107]. As an alternative method, Lee J et al. treated parental cells with azide membrane-fused liposomes (MFL) to create azide exosomes that were then conjugated with a tumor targeting peptide conjugate DBCO-CGKPK via bioorthogonal reaction [108]. MFL exogenously incorporated into exosomes for surface modification does not damage membrane proteins critical to the exosomes’ biological function (Figure 4).

### 2.2 Extraglandular vesicles of plant or microbial origin as dynamic carriers

The majority of studies at present focus on EVs derived from animal cells. However, the production efficiency of EVs derived from animal cells is low, and only a few micrograms can be obtained in billions of cells [109]. Therefore, it is usually necessary to culture...
a large number of cells to produce enough EVs for in vitro and in vivo experiments [109], resulting in expensive experiments. Compared with EVs of animal origin, a major benefit of EVs derived from plants or microorganisms is that they can be produced in larger quantities as well as having a higher economic benefit. Not only that, plant-derived exosome-like nanoparticles (PDENs), which are often enriched in various bioactive lipid, protein, RNA, and other components, are natural nano-formulations that are proven to have significant regulatory roles in cancer, immunomodulation, and antiviral versus antioxidant effects [110]. For example, Zhu et al. found that after oral administration of ginger derived EVs in mice, they mainly accumulated in the liver and mesenteric lymph nodes, and could inhibit ROS production when acting on hepatocytes [111]. Citrus derived EVs can participate in regulating the gene expression of tight junction proteins, thereby restoring intestinal barrier function [112]. Grape derived EVs accelerate the repair of intestinal mucosa epithelium by regulating the expression of genes that promote the growth and proliferation of intestinal stem cells through Wnt signaling pathway [113]. Broccoli derived EVs can target dendritic cells (DCs) to activate AMP-activated protein kinase (AMPK) in DCs, thereby reducing the release of IFN-γ and TNF-α. It can prevent the activation of intestinal DCs and improve the symptoms of DSS induced colitis in mice [114]. Lemon derived EVs can inhibit the proliferation of cancer cells by promoting tumor necrosis factor related apoptosis inducing ligand (TRAIL) - mediated apoptosis and inhibiting the secretion of vascular endothelial growth factor A (VEGF-A), IL-6 and IL-8 [115]. Lentinus edodes derived EVs can alleviate D-galactosamine/lipopolysaccharide induced liver injury in mice by inhibiting the activation of NLRP3 inflammasome [116]. At the same time, in addition to their inherent therapeutic potential, source plants possess low toxicity and safety characteristics, making them ideal vehicles for delivering drugs and genes, with broad application prospects [117].

PDENs have been successfully reported for oral colon-targeted drug delivery [118]. In addition, studies have also shown that PDENs have good tolerance to digestive enzymes like pepsin, intestinal pancreatin and bile. It allows the entrapped drugs to be delivered intact to the colonic site and exert their effect, thus avoiding decomposition of drugs by the gastric or intestinal environment [110]. However, researchers have not yet elucidated how PDENs can still maintain drug loaded integrity while defending against the harsh gastrointestinal environment. A study constructed methotrexate loaded PDENs (GDNs) for targeting intestinal macrophages. After oral administration of the drug loaded PDENs, the results showed that in the intestinal lamina propria, these PDENs are highly efficient in targeting F4/80+ macrophages via micropinocytosis and clathrin-dependent cellular uptake pathways. After being internalized by macrophages, encapsulated methotrexate was dynamically released. The results showed that pretreatment with GDNs could effectively prevent weight loss and intestinal segment shortening caused by DSS induced colitis (Figure 5a, b). After treatment with GDNs, the severity and pathological score of colitis in mice decreased (Figure 5c, d). The expression of E-cadherin, IL-6 and IL-1β in colon epithelial cells were significantly reduced (Figure 5e, f). The mRNA levels of chemokines MCP-1, CXCL9 and CXCL10 were also reduced (Figure 5g), and the inflammatory neutrophil infiltration of CD11b+Ly6Chigh was significantly reduced (Figure 5h, i), suggesting that GDNs could enhance the anti-inflammatory capacity of intestinal immune cells by playing the role of immune regulators [119]. It has been reported that curcumin can be delivered to colon tissue using PDENs, and the PDENs enhanced both the stability and blood level of the drug, resulting in its specific drug delivery to inflammatory cells [120], offering new treatment options for colon cancer. In response to relevant studies, the James Graham Brown Cancer Center initiated two phase I clinical trials to test the therapeutic efficacy of PDENs in head and neck cancer and colon cancer (NCT01668849, NCT01294072). In addition, clinical studies on PDENs, multiple clinical trials on cell-free immunotherapy for cancer are also underway, with a view to developing effective cancer vaccines in the future [121].

Outer membrane vesicles (OMVs) are a type of membrane-shaped structure originated from the bacteria’s outer membrane and mainly produced by Gram negative bacteria [122,123]. OMVs contain a variety of components of bacterial origin, including enzymes, virulence factors, bacterial specific antigens, and various PAMPs, such as lipopolysaccharide (LPS), making them especially useful for the development of bacterial vaccines and adjuvants [124,125]. In addition, OMVs may also have antitumor properties that have gradually gained attention [126]. OMVs can not only induce powerful tumor immune effects through bacterial-derived proteins, but also encapsulate therapeutic drugs using their own structures to achieve PAMPs-mediated cell specific targeting and improve drug accumulation at target sites [127].
In order to relieve the state of tumor immunosuppression, combining the anti-tumor immune response mediated by OMVs with chemotherapeutic drugs is a more effective approach. By combining immunotherapy and OMVs, additional potential for tumor immunotherapy can be enhanced to prevent tumor recurrence and metastasis and complete eradication of tumors. The detoxified OMVs were modified by Ping et al. by adding polyethylene glycol (PEG) and the Arg-Gly-Asp (RGD) peptide to increase blood circulation and improve tumor targeting. On top of the tegafur-loaded NPs, modified OMVs were coated, and the resulting NPs had three antitumor functions (Figure 6). As a first step following systemic injection, innate immune cells are activated, followed by an enhanced tumor accumulation in response to the EPR effect and peptide targeting by RGD. After being taken up by tumor cells, OMVs dynamically released tegafur, which enhanced the sensitivity of cancer cells to cytotoxic CD8⁺ T cells and eliminated myeloid-derived suppressor cells (MDSCs), thereby synergistically enhancing the immunotherapy ability of OMVs [128]. Notably, OMVs-coated NPs were not harmful to major organs. Serum cytokine levels increased rapidly within a few hours of injection and rapidly decreased within 24 hours, demonstrating the safety of this drug delivery vehicles.

Figure 5. Grapefruit-derived nanoparticles (GDN) pretreatment ameliorates DSS-induced colitis in mice. C57/B6 mice were treated with either PBS/DSS or GDN/DSS. Adapted with permission from [119]. Copyright Year 2014, Elsevier.
The safety of using OMVs for cancer treatment is a major concern. Although many studies have demonstrated that the use of attenuated or detoxified OMVs as drug delivery carriers are safe in mice [129], humans are several orders of magnitude more sensitive to endotoxin than other mammals, such as mice [130]. There is still no relevant report on whether the human body can tolerate OMVs. The LPS of the cell wall of gram-negative bacteria is not only highly carcinogenic but can also cause fatal septic shock [129]. In order to further improve the safety of OMVs as drug carriers, current studies usually use the OMVs of attenuated or avirulent bacteria, and suppresses the expression of bacterial LPS through gene mutation or makes the cytoplasmic A in LPS insufficiently acylated, followed by genetic and surface modification to target the cells of interest. There have been studies to engineer avirulent *Staphylococcus aureus* (*S. aureus*) mutants by genetic engineering that produce avirulent cytolysins, and OMVs containing avirulent cytolysins that are immunogenic and nontoxic are able to protect mice against *S. aureus* [131]. Gujrati *et al.* used genetic engineering to modify the parental *Escherichia coli* (*E.coli*) to reduce the cytotoxicity of OMVs, expressed the tumor targeting ligand HER2 on their surface, and loaded siRNA to inhibit the activity of overexpressed spindle kinesin within tumor cells. Consequently, cell proliferation is...
inhibited and apoptosis is induced. The experiments showed that AffiHER2OMV-siRNA was not significantly cytoxic and could aggregate at the tumor site with high specificity, effectively inhibiting tumor growth [132].

2.3 Platelets as dynamic carriers

Megakaryocytes produce small, anucleate cells called platelets, whose lifespan is about 8 ~ 10 d, and their main function is hemostasis, regulating inflammation, and promoting thrombosis [133]. Aside from their traditional functions, platelets can also play other roles such as regulating immunity and communicating with other cells and tissues in the blood vessels. Key to the role played by platelets is their activation, which upon activation enables the dynamic release of granules (platelets contain 3 types of granules: α particles, δ particles and lysosomal particles) and EVs (microparticles 100 nm ~ 1 mm and exosomes 40 ~ 100 nm in diameter). It is possible that platelet communication with other cells in the blood vessels is achieved through microparticles. Genetic material (miRNA, mRNA, and so on), enzymes, proteins, and small molecules within these microparticles can affect cells in the vasculature and alter their functions [134,135]. Because platelet microparticles and exosomes differ in their source and specific protein composition, they may have different functions, with exosomes play the function of extracellular communication, platelet microparticles play the role of procoagulant activity and intracellular communication that can transfer their cargo (cytoplasm or membrane proteins, mRNA, and non-coding RNA) to target cells. This explains part of the role of platelet EVs in inflammation, thrombosis, immune regulation and biological information transmission [136,137].

Studies have shown that platelets are involved in tumor development, including tumor growth, tumor cell extravasation, and metastasis, by protecting tumor cells from the host immune system, whereas platelet effects on local and distant tumor host interactions may be mediated through the secretion of large amounts of platelet microparticles and exosomes [138,139]. In vivo studies have shown that platelet microparticles can promote the growth of primary tumors, stimulate vascularization, and promote the formation of distant metastases [140]. Levels of platelet microparticles are markedly elevated in a large subset of malignancies. Colon cancer patients are often accompanied by altered coagulation activity, and stage III/IV colon cancer patients have significantly increased levels of platelet microparticles in the systemic circulation, greatly shortened clotting time, and increased fibrin production, resulting in a hypercoagulable state in the blood of colon cancer patients, so platelet microparticles are potential targets for preventing coagulation in colon cancer [141].

A great deal of attention has been paid in recent years to the interactions between platelets and CTCs in the blood [142], and the aggregation of platelets surrounding CTCs helps them survive in the bloodstream and invade new tissues [143]. The high affinity between P-selectin on platelets and CD44 receptors on tumor cells explains this specific aggregation. Therefore, platelets are more likely to target CTCs when P-selectin binds specifically to CD44 [144,145]. To deliver functional small molecules with precise targeting, Hu et al. created platelet-mimicking nanovehicles (PM-NV) [146]. PM-NV consists of a small molecule drug loaded nanogel inner core and a platelet membrane based outer shell. Meanwhile, TRAIL was modified on the surface of platelet membrane, and DOX was loaded into PM-NV to prepare TRAIL-DOX-PM-NV. During in vivo experiments, PM-NV was found to contain a large amount of platelet-derived “self-recognition” proteins, which minimizes in vivo immunogenicity and prolongs circulation time. CD44 molecules are present on tumor cell membranes and can be bound by P-selectin carried on platelet membranes. This allows TRAIL-DOX-PM-NV to target tumor cells and accumulate at the tumor site. Meanwhile, TRAIL-DOX-PM-NV aggregation on tumor cell surfaces encourages TRAIL interaction with the membrane and signals exogenous apoptosis. More importantly, when TRAIL-DOX-PM-NV was internalized by cells, the acidic environment of lysosomes would mediate TRAIL-DOX-PM-NV to dynamically release DOX, allowing DOX to accumulate in tumor cell nuclei and synergistically induce tumor cell apoptosis (Figure 7). In addition, the binding of P-selectin to CD44 can eliminate CTCs, preventing tumor metastasis. Based on platelet and tumor cell specific affinity, this study successfully developed platelet membrane modified nanomedicines that delivered TRAIL and DOX to tumor cells.

Primary amine or thiol residues on the surface of platelets enable chemical attachment to NPs or biomolecules, followed by dynamic release of drug loaded microparticles via activation [147,148]. Kailashiya et al. [12] constructed platelet-transformed drug loaded platelet-derived microparticles (PMPs). This biomimetic particle has good biocompatibility and natural targeting, can carry a variety of drugs, has good stability, and can be obtained in large quantities in a short period of time. Compared with free drugs, PMPs are more toxic to cancer cells and less prone to extravascular escape [149].
3 Macrophages transformation

The body’s immune system functions as an immune surveillance that can recognize and clear mutant cells from the body [150]. However, tumor cells evade immune surveillance and immune killing by reducing immunogenicity and enhancing immune tolerance, leading to a loss of balance in the interaction of the tumor with the immune system [151]. Tumor cells establish an immunosuppressive TME by secreting immunosuppressive factors and recruiting immunosuppressive cells, which further suppress antitumor immune responses, thus leading to uncontrolled tumor growth and ultimately life-threatening health. Tumor associated macrophages (TAMs) are important components of the immunosuppressive TME [152]. The classical phenotype of macrophages is divided into M1, which is pro-inflammatory and anti-cancer, and M2, which is pro-cancer. TAMs are generally M2 type, therefore, polarizing M2 type TAMs to M1 type is a feasible scheme to restore antitumor immune responses [153].

3.1 Macrophage polarization

Peptides targeting M2 macrophages can be used to modify carriers capable of delivering drugs that promote macrophage transformation to achieve TAMs-specific regulation. Signal transducer and activator of transcription 6 (STAT6) and nuclear factor kappa-B (NF-κB) promote M2 polarization of macrophages through different mechanisms. Therefore, inhibition of these two signaling pathways may lead to a phenotypic transformation in macrophages. Xiao et al. [154] constructed a nanomedicine with dual characteristics of TME response and active targeting. The M2 targeting peptide was first masked with pH-sensitive PEG, and then the targeting peptide was used to modify the STAT6 inhibitor AS1517499 (AS) and the inhibitor of nuclear factor kappa-B kinase β (IKKβ) siRNA nanomedicines to obtain nanocomplexes. After the nanocomplex reaches the tumor, the PEG protective shell falls off under the weak acid condition of TME, exposing the M2 targeting peptide, and the loaded drug is specifically delivered to M2 macrophages, which induces the phenotypic polarization of M2 macrophages at the same time, reduce the occurrence of adverse reactions, and provide safe and effective immunotherapy. In addition, based on specific proteins on the surface of macrophages, Kulkarni et al.
designed a protein that can specifically bind to SIRP-α and inhibit the colony stimulating factor-1 receptor (CSF-1R) through computational biology. The supramolecules can self-assemble with specific compounds into nanoparticles. After targeting macrophages, the supramolecules can block the mutual binding of macrophage SIRP-α receptors and CD47, and block the CSF-1R signaling pathway, so that TAMs are polarized to the M1 type and induce an innate immune response against tumors [155].

Several classes of microbial agents and microbe derived molecules (e.g., bacterial muramyl dipeptide, IFN-γ, CD40 antibody antagonist, CP-870, 893) were shown to drive the antitumor cytotoxic effects of macrophages, related clinical research has also been carried out, but their clinical efficacy has been modest [156,157]. While the continuous extension of nanotechnology in the field of tumor immunotherapy has allowed many novel nanomedicines to be mined for their potential in tumor immunotherapy, iron oxide nanoparticles are one of them (Figure 8). For a long time, iron oxide nanoparticles have been used for imaging liver tumors due to the trait of specific aggregation in liver tissue, a property that highlights their advantages for liver tumor therapy [158,159]. Li et al. used hyaluronic acid (HA) encapsulated superparamagnetic iron oxide nanoparticles (IONs) to construct reprogrammed macrophages (HION@Macs). In vitro and in vivo studies demonstrated that HION@Macs were able to educate in situ M2 macrophages toward M1 transition and restore their immune effects via a paracrine manner. HION@Macs also exhibits strong resistance to the immunosuppressive tumor environment, enabling sustained production of inflammatory factors (i.e., NO, H2O2, and TNF-α) to inhibit the growth of cancer cells and promote their apoptosis, exerting a synergistic tumor suppressive effect [160].

Although there have been many studies that successfully induced phenotypic transformation of macrophages in vivo, this strategy is still very far from clinical translation. Because in most cases, M1 type macrophages induced to undergo polarization by in vitro stimulation readily revert to the M2 phenotype after reaching the tumor immune microenvironment [161], thereby failing to generate sustained immune killing. Therefore, to truly exert powerful therapeutic effects in the clinic, macrophage-based tumor immunotherapy must develop effective strategies to control the phenotype of adoptively transferred macrophages in vivo. Based on this purpose, a cellular “backpack” strategy that can maintain the duration of M1 type polarization of macrophages has emerged. Several drug delivery demonstrations using the “backpack” strategy have been carried out over the past few years [162-166].

![Figure 8. Reprogramming of tumor-associated macrophages by iron oxide nanoparticles as an anti-tumor therapeutic strategy [159]. Copyright Year 2021, the Authors.](https://www.thno.org)
Wyatt Shields IV et al. integrated interferon gamma (IFN-γ) into a “backpack”, and IFN-γ as a pro-inflammatory macrophage stimulator can sustain macrophages in a tumor-killing M1 type state. In mice with cancer, the maintenance effect slowed tumor growth and reduced metastasis for up to five days. Specifically, the team fabricated two layers of the biocompatible polymer poly (lactic-co-glycolic acid, PLGA) sandwiched with polyvinyl alcohol (PVA) and the cytokine IFN-γ. To complete the “backpack”, a layer of cellular adhesive was also added to help the “backpack” fit snugly on the macrophages. In addition, they attach easily and do not become phagocytosed or digested by macrophages due to their disc-shape, allowing them to survive for longer periods of time. The researchers injected “backpack”-carrying M1 macrophages into the tumors of mice and assessed them seven days later. Based on the findings of the in vivo study, macrophages carrying an IFN-γ “backpack” exhibited M1 indicators for more than 48 hours. As compared to cells injected with free IFN-γ or cells with a blank “backpack”, there was a significant difference in expression level of M1 indicators. Additionally, treatment with IFN-γ “backpack” therapy resulted in fewer metastatic nodules, smaller tumors, and longer lifespans than control group. Interestingly, not only do “backpack”-wearing macrophages stay in the M1 state, but it can also repolarize other macrophages including TAMs. Also repolarize other macrophages including TAMs -wearing macrophages stay in the M1 state, but it can also repolarize other macrophages including TAMs. In mice. In contrast to other studies, this effect was achieved at 100 times less than the maximum total dose of IFN-γ, and the mice did not show any signs of treatment toxicity [167].

As a result of the “backpack” strategy, macrophages adoptively transferred into solid tumors will retain their phenotype in an immunosuppressive environment. To improve its efficacy against solid tumors, future research can continue to delve into the optimal way of packing the stimulator into the “backpack” and its release kinetics. In addition, “backpack” can be combined with adjunctive therapies to increase the therapeutic effect. These strategies should also not be restricted to macrophages and can also engage other circulating cells with increased chemotactic sensitivity to mount a more robust tumor immune response [168,169].

4 Semi-biological hybrid cellular transformers

Abundant chemical groups on cell membranes are able to create interactions with the interfaces of nanomaterials or carriers. Based on this, coupling synthetic materials to the cell surface or encapsulating nano-drugs inside the cells can promote the exposure of tumor-associated antigens and induce tumor immune responses on the one hand, on the other hand can realize multimodal therapy of tumors by combining PDT or PTT, and most importantly, the constructed biomimetic nanocarriers have the property of actively targeting tumor sites. Such biomimetic nano-drug delivery systems constructed or modified artificially we call them semi-biological hybrid cellular transformers. By active targeting effect, it can reduce the non-specific distribution of drugs in the body, and the safety and efficacy of treatment can be improved [170]. Through endocytosis mediated by the interaction of surface conjugating ligands with membrane receptors of target cells, semi-bio hybrid cellular transformer can increase the concentration of drugs inside the target cells [171]. A series of nanocarriers modified with ligands, antibodies, antibody fragments or short peptides with high specificity and affinity have been developed so far according to the receptors or protein antigens that are highly expressed specifically on the tumor cell membrane [172]. For example, transferrin can bind to transferrin receptors on the membranes of metastatic tumors and drug-resistant tumor cells to mediate endocytosis of drugs [173]. It has been shown that Arginine glycine aspartic acid (RGD) tripeptides target tumor vessel and cell integrins αvβ3 that are upregulated on the surface of tumor cells and increase the delivery efficiency of drugs [174]. In addition to protein ligands, carbohydrate ligands (e.g., hyaluronic acid, HA), folate receptors that are highly expressed in a variety of tumor cells, and nucleic acid aptamers can also bind receptors specific to tumor cell surfaces, conjugating them onto the nano-biomimetic drug delivery system not only mediates efficient uptake of drugs, but also enables enhanced delivery efficiency and safety of nanomedicines [175,176].

In addition to artificially modified ligands, certain specific types of cells have been regarded as potential drug delivery vehicles due to their abilities to homing to tumors and long circulation in the blood. For example, tumor cell membranes can be exploited for surface functionalization of nanoparticles, thereby endowing them with the ability for homologous targeting to achieve tumor specific drug delivery [177]. Engineered CCMs prepared by fusing CCMs with other types of CCMs or making CCMs express specific proteins provide a new direction for overcoming the defects of CCMs and improving the biomedical properties of CCMs [178]. The latest research points out that the nano-drug coated with glioblastoma (GBM) cell membrane has good blood-brain barrier (BBB) permeability and no obvious side effects [179]. Besides, since a variety of immune cells are recruited to the tumor site during cancer development, this inspired researchers to use
the cell membrane coated nanomedicine of immune cells to obtain the dual roles of tumor targeting and immunotherapy. It has been confirmed that nano-biomimetic drug delivery systems based on monocyte/macrophage membrane, neutrophil membrane and immune cell membrane can improve the delivery efficiency of antitumor drugs [180-183].

Although semi-biological hybrid cellular transformers can also be surface modified or loaded with drugs, there are some differences with EVs drug delivery systems. The difference between the two is that EVs, as part of the cell, only inherit some of the cell’s properties. In order to develop dynamic drug delivery systems using EVs, it is usually necessary to introduce functional components into cells and release them in the form of EVs when the parent cells are exposed to specific stimuli [184]. The preparation process is complex and the yield is difficult to control. However, the direct loading of drugs into the isolated EVs may lead to vesicle aggregation and membrane protein damage [185]. In addition, the large-scale extraction and purification of EVs has been a major problem restricting their clinical application [186]. In contrast, semi-biological hybrid cellular transformers mostly use cell membranes as carriers, so the types of surface proteins and markers are more abundant, and there are more possibilities in preparing targeted carriers. In addition, semi-biological hybrid cellular transformers can be prepared by using single-cell biofilms to encapsulate drugs, or by preparing hybrid membranes to impart more properties to the carriers. The more accessible membrane materials and higher carrier yield also make it more universal than EVs in the future. It is worth noting that compared with synthetic NPs, drug carriers constructed by natural cell membranes or EVs still have incomparable advantages. They can effectively load therapeutic drugs without significantly changing the membrane structure and surface protein, and have stability, biocompatibility, extremely low immunogenicity, targeting and the ability to overcome natural barriers in the blood circulation, which can overcome the limitations of synthetic NPs [187-189].

4.1 Cargo loaded cellular transformers

Loss of tumor antigen expression is one of the important mechanisms of immune escape. Tumor cells evade killing by the immune system by downregulating the expression of strong rejection antigens, downregulating major histocompatibility antigen I (MHC-I) levels, and impairing antigen-presenting function [190]. Utilizing semi-biological hybrid cellular transformers to deliver drugs specifically to tumor cells, and induce specific types of cell death, would lead to the release of tumor associated antigens (TAAs), which in turn be recognized by immune cells and activate antitumor immune responses [191].

Recently, a new form of cell death known as ferroptosis has been discovered that could improve immunogenicity of tumors. The existing agents that induce tumor cells to undergo ferroptosis usually face two major problems of being easily cleared by the body’s immune system and being difficult to target. Jiang et al. coated sulfasalazine (SAS) - loaded magnetic iron oxide nanoparticles (Fe3O4 NP) with platelet membrane by extrusion method to obtain biomimetic nanoparticles Fe3O4-SAS@PLT with better stability. P-selectin on the surface of platelets works in conjunction with CD44 on the surface of 4T1 tumor cells to increase the uptake of nanoparticles and promote the accumulation of nanoparticles in metastatic tumors. The ingested magnetic iron oxide and sulfasalazine synergistically induce ferroptosis, expose TAAs, and activate antitumor immune responses (Figure 9). The rate of DC maturation induced by biomimetic nanocarriers was increased to 69.9%, which was significantly higher than that of nanoparticles without platelet membrane coating (28.4%). Programmed death (PD-1) inhibitors and this biomimetic nanocarrier were combined to treat tumor bearing mice, achieving 76% of mice survival up to 80 days, while the mice in other control groups all died within 50 days [192]. This approach of utilizing natural ligands of cell membranes to deliver drugs specifically to tumor sites provides a feasible strategy to enhance antitumor immunity, meanwhile, the outstanding biocompatibility from cell membranes improves the possibility of clinical translation.

There are many tumors stromal cells in the TME, including cancer-associated fibroblasts (AF), and this occupies a major position as a barrier to the development of cancer treatments. On the one hand, AF secretes many growth factors and cytokines to activate tumor related signaling pathways to promote angiogenesis, tumorigenesis, progression, metastasis, and resistance generation [193]. On the other hand, AF prevents drug entry into tumor cells by surrounding tumor cells and producing extracellular matrix (ECM) [194]. It is illustrated that AF constitutes a physical barrier to protect tumor cells and impedes the access of anticancer drugs to the tumor to exert their effects. Many studies have therefore turned to the treatment of tumors based on AF. Li et al. reported an activated AF coated semiconductor polymer nanoparticle (SPN) for enhanced tumor phototherapy [195]. This nanocomplex, called AF-SPN, is constructed from a semiconductor polymer (SP) containing NIR absorption and the AF cell membrane. As a theranostic agent, SP can be used for both
imaging and photothermal therapy by producing NIR fluorescence and singlet oxygen. By coating the AF cell membrane with NPs, homologous targeting was enabled, and promoting NPs accumulation in the vicinity of cancer cells and thus enhancing the efficacy of photodiagnostic therapy. Furthermore, this biomimetic nanocomposite can also be loaded with other drugs and imaging agents to provide a multimodal theranostic platform, providing new ideas for tumor theranostics.

4.2 Engineered modified cellular transformers

Recently, as a representative of adoptive cell therapy, CAR-T has achieved a large breakthrough in the treatment of hematological tumors. But due to the limitation of many factors, such as the lack of tumor specific antigen and the suppressive tumor immune microenvironment, CAR-T therapy still faces a great challenge in the treatment of solid tumors [196]. In view of this, to improve CAR-T cell targeting and antitumor efficacy, we must start with optimization of CAR-T target selection, reduction of antigen escape and breaking immunosuppression in the TME [197]. Owing to the high mutability of tumor cells and the changing immune microenvironment, the phenomenon of tumor immune escape is difficult to eliminate, and the dual targeting strategy can provide a more reliable and effective way for nanocarriers to target tumors. Researchers have used bioorthogonal technology to construct artificial targets BCN groups (bicyclic [6.1.0] nonyne) at the tumor site. Meanwhile, azide groups (N3) were embedded into T cell

Figure 9. Schematic illustration of platelet membrane-camouflaged magnetic nanoparticles for ferroptosis-enhanced cancer immunotherapy. Adapted with permission from [192]. Copyright Year 2020, Wiley-VCH GmbH.
membranes by glycometabolism technology, and then the extracted N₃ functionalized T cell membranes were encapsulated on the surface of indocyanine green (ICG) - loaded polymeric nanocores to construct biomimetic nanoparticles (N₃-TINPs). A special protein (T-cell antigen receptor, TCR) on the T-cell membrane endowed N₃-TINPs with immune recognition ability to tumor cells, while functionalized N₃ groups enabled efficient and specific bioorthogonal reaction with BCN groups of artificial targets for glycometabolic labeling on tumor cells (Figure 10). N₃-TINPs greatly enhance the enrichment of ICG at the tumor site to effectively overcome the tumor “off target effect”, thus improving the antitumor efficacy of nanocarriers based on their tumor targeting with immunomodulatory drugs using artificial targeting and natural target based dual guide therapy [198].

In recent years, aptamers as novel targeting ligands can also modulate the body’s antitumor immune response by binding immune related receptors, which could be applied to tumor immunotherapy. In 2019, an osteosclerotic protein aptamer drug developed by the research team of Hong Kong Baptist University has been granted orphan drug certification by FDA for the treatment of osteogenesis imperfecta [199]. Previous studies have shown that aptamers can be directly used in tumor immunotherapy as novel targeting ligands for tumor associated antigens, immune checkpoint molecules, costimulatory receptors, cytokines, and can also be coupled with drugs, siRNA, nanomaterials [200-202]. Or modify the aptamer on the cell membrane surface to exert anti-tumor effect [203]. The interaction between immune and tumor cells is an important link to achieve highly effective immunotherapy. By recognizing tumor cells on the surface of immune cell membranes, anchoring nucleic acid aptamers can directly induce the immune cells to attack the tumor cells and maximize the therapeutic effect. For example, the aptamer S4F targeting SGC-7901 gastric cancer cells were modified on the surface of CD3⁺T cells by metabolic labeling and click chemistry, giving them stronger tumor targeting and killing [204]. NK cells modified with TLS11a aptamer and programmed cell death ligand 1 (PD-L1) aptamer can both target attacking liver cancer cells and resist the inhibition of PD-1/PD-L1 immune checkpoint, enhancing the
immunotherapeutic efficacy of NK cells in solid tumors [205]. Recently, researchers used DNA aptamers as a substitute for antibodies to construct a polyvalent antibody mimic (PAM), which utilizes hydrophobic insertion to insert the PAM into the cell membrane of NK cells to make NK cells identify and capture the corresponding tumor cells effectively. This method has a broad spectrum, and any aptamer of interest can be modified on various immune cells [206]. Compared with CAR-T/NK immunotherapy, which has a large therapeutic risk and complicated operation, the aptamer modified immune cells are more easily available and have less toxic and side effects, making them highly potential for clinical applications.

Since tumor surface markers are different, the selection of suitable cell membrane surface modifiers is crucial for the targeting of biomimetic nanocarriers. Low density lipoprotein receptor related protein (LRP) receptors are overexpressed in endothelial cells of the BBB and in U87MG GBM cells. Based on this property, Shi et al. constructed an angiopep-2 modified biomimetic nanocarrier of RBCs membrane loaded with polymer coloaded DOX and lexiscan (Lex). Lex can transiently open the BBB and increase the permeability of the nanomedicine to the brain, while angiopep-2 can target the highly expressed LRP in the GBM, promoting the passage of the nanomedicine through BBB and achieving GBM targeting while prolonging the circulation time of the NPs [207]. After further research, Shi et al. constructed ApoE peptide modified RBCs membrane encapsulated temozolomide (TMZ) and the epigenetic bromodomain inhibitor (OTX015) biomimetic nanocarriers to achieve synergistic chemotherapy and immune therapy for GL261 GBM. In contrast to previous studies, ApoE peptides can target multiple overexpressed low-density lipoprotein family receptors (LDLRs) (e.g., LDLR, LRP1, and LRP2) in BBB endothelial cells and GBM cells, thus providing superior BBB penetration and tumor targeting capabilities [208]. This biomimetic nanocarrier enhances the sensitivity of TMZ to tumor cells while targeting GBM. The anti-tumor immune response was also enhanced by enhanced expression of CD4⁺ and CD8⁺ T cells through induction of immunogenic cell death and suppression of PD-1/PD-L1 coupling [209]. Compared with brain tumors, digestive system tumors exist in a large variety and complex characteristics of biomarkers. Therefore, finding suitable targets is particularly important for the targeting, efficacy and safety of biomimetic nanocarriers.

### 4.3 Engineered cellular bioreactors

Currently, although surgery, chemotherapy, and radiotherapy are all effective methods of suppressing primary tumors, nearly 90% of cancer-related deaths are still caused by cancer cell metastasis [210]. To improve cancer treatment outcomes, it may be beneficial to modulate the TME rather than directly destroy cancer cells [211]. Studies have confirmed that the immunosuppressive TME of solid tumors is characterized by severe hypoxia, elevated H₂O₂ content, and overexpression of glutathione, while a cascade bioreactor constructed by combining natural cell membranes with nanomedicines is able to modulate the tumor microenvironment, improve radiotherapy, and support immunotherapy.

Liu et al. used Ti₃C₂ nanosheets as carriers to chemically conjugated with glucose oxidase (GOX) and chloroperoxidase (CPO) and load deoxygenation-activated prodrug tirapazamine (TPZ) to prepare a cascaded-enzyme nanoreactor Ti₃C₂-GOX + CPO / TPZ (TGCT), which was finally encapsulated into tumor cell membrane carriers with high expression of CD47 (m₄TGCT). Because the encapsulation of highly expressed CD47 biomimetic membrane, m₄TGCT exhibits superior immune escape ability as well as homologous targeting ability, which can be preferentially targeted to tumor sites and effectively enhanced tumor cell uptake. After entering tumor cells, GOX and CPO can produce a large amount of HClO through a cascade reaction, achieving highly efficient enzyme dynamic therapy (EDT). In addition, A further laser irradiation can increase singlet oxygen (¹O₂) production by accelerating enzyme catalyzed reactions. Meanwhile, the oxygen consumption of phototherapy and EDT could aggravate the hypoxic state inside the tumor, and then activate the deoxygenation-activated prodrug TPZ to realize chemotherapy. Experimental results showed that m₄TGCT can effectively inhibit tumor growth because of the amplified synergistic therapeutic effect of tumor phototherapy, enzyme dynamic therapy, and chemotherapy (Figure 11). A cascaded enzyme nanoreactor proves to be a promising way of achieving synchronized and remarkable antitumor therapy [212]. At present, bacteria based synthetic biology has also been widely used in tumor therapy and achieved good results. Zhang et al. created a bacterium, Escherichia coli (MG1655), that overexpresses the NDH-2 enzyme (respiratory chain enzyme II) and colonizes tumors and increase the generation of H₂O₂. Subsequently, the experiments covalently linked magnetic Fe₃O₄ nanoparticles with bacteria and acted as a catalyst for Fenton-like reaction to transform H₂O₂ into toxic hydroxyl radical (•OH) for tumor therapy. The structures proved that
the constructed bioreactor sustainably produced H$_2$O$_2$ with engineered bacteria for performing Fenton-like reaction, while the generated toxic •OH could effectively induce tumor cell apoptosis. With this bioreactor, it is possible to achieve aggressive tumor colonization as well as self-sufficient Fenton-like reactions without the need for additional H$_2$O$_2$ [213].

**Conclusions and future perspectives**

Worldwide, digestive system cancer is still the leading cause of cancer death. Digestive system cancer is still the most common cause of cancer death worldwide. The clinical treatments for digestive system tumors are still conventional surgery, chemotherapy and radiotherapy. Surgical treatment, which is more effective for early-stage tumors that have not spread, but with the development and metastasis of malignant tumors, the curative effect and prognosis of advanced tumors are poor, and often cannot completely clear all cancer cells in the human body. Although chemotherapy can play a certain role, due to its lack of specificity, there is a large damage to the normal cells and immune system of the body, and it is easy to increase the body’s drug resistance and cause adverse reactions. Radiotherapy has the problem of long cycle length and significant toxic side effects to human body. Some novel therapies such as immune checkpoint inhibitors, targeted agents, and CAR-T have played an important role in the treatment of digestive system cancer, but they also suffer from defects such as drug resistance and severe adverse effects. Under this background, the new idea of biomimetic nano-drug delivery system based on natural cells may change the situation of digestive system malignant tumor treatment (Table 2). The promotion effects of biomimetic nano-drug delivery systems for digestive system tumor therapy are mainly reflected in the following aspects. 1) The optimal multi-drug co-delivery system can be designed according to the inherent characteristics of various cell membranes, and the mixing ratio of cell membrane and nanoparticles can be adjusted to optimize the fusion process and achieve higher encapsulation efficiency and good controlled release capability. 2) Precise modification of nanocarriers can be achieved by recognition, removal or addition manipulation of specific antigenic proteins on the cell membrane surface to improve the tumor targeting ability. 3) By modifying nanocarriers with hybrid membranes, or incorporating ligands consisting of antibodies, nucleic acids or enzymes into the cell membrane for surface functionalization, a variety of

![Figure 11. Schematic diagram of the construction of bionic cascaded-enzyme nanoreactor and proposed mechanism in vivo. Adapted with permission from [212]. Copyright Year 2021, Springer Nature.](https://www.thno.org)
biological functions beyond the single cell membrane are introduced to enhance the synergistic effect in tumor immunotherapy. 4) Cell membrane modified nanocarriers prepared from patients’ own cells can be utilized for individualized drug delivery and treatment to achieve the effect of precision treatment of tumors. 5) It can promote the extensive combination of chemotherapy, radiotherapy, photothermal therapy and other treatment methods, and realize the integration of various tumor treatment methods.

Different from drug delivery for brain tumors, drug delivery systems targeting digestive system tumors can achieve targeted release of drugs without crossing complex biological barriers such as blood brain barrier. However, it should be noted that drug carriers targeting digestive system tumors still need to solve the problems such as short internal circulation time, off-target effect caused by being captured and cleared by immune system. In view of the low immunogenicity and high heterogeneity of digestive system tumors, the inherent targeting ability of immune cell membrane and the homologous targeting of tumor cell membrane can be used to achieve targeted drug release when designing natural cells based biomimetic cellular transformers. Compared with hematological tumors, the TME of the digestive system is relatively stable, and the infiltrating tumor associated macrophages are highly heterogeneous and plastic. Therefore, targeting the TME can relieve the immunosuppression of the tumor site, and transforming “cold tumors” into “hot tumors”. This is an effective strategy for designing biomimetic cellular transformers as well. Our review focuses on the dynamic properties of native cells and reviews their transformation characteristics. Targeting digestive system tumors by transferring specific functional and biological properties of native cells (such as dynamic deformation, phenotypic polarization, enhanced immunogenicity, immune escape, tumor-specific targeting) into nanocarriers treatment offers a wider range of possibilities.

Nonetheless, research on natural cell-based biomimetic nanoparticle drug delivery platforms in the diagnosis and treatment of digestive system tumors is still in its infancy, and many challenges need to be overcome for the transition from experimental research to clinical applications. One of the most important points is how to guarantee the safety of biomimetic NPs. Current production technologies of biomimetic NPs are not mature enough, and the resulting complex and varied production processes, inconsistent standards and poor reproducibility will affect the efficacy and safety of the final products; Second, existing characterization methods are limited, and the success of membrane coating is verified only by particle size detection and morphological observation. Western blot analysis can only demonstrate whether the surface composition of biomimetic nanocarriers is similar to that of the source cell membrane, but cannot verify whether the membrane is partially disrupted after membrane coating and whether biochemical reactions occur during the coating process or introduce unknown components that pose a hidden danger to drug safety. Moreover, biomimetic NPs with improper storage are prone to denaturation of membrane proteins, resulting in potential immune responses to endogenous antigens, which in turn cause tissue damage to the body. Additionally, its safety issue should be well considered when using certain special cell membranes such as tumor cell membrane or bacterial membrane. Inappropriate immunogenic exposure may induce detrimental immune responses, even against normal cell types, and long-term safety should be considered. Overall, to further improve the safety and efficacy of biomimetic NPs, the problem of poor reproducibility and inconsistent standards in the preparation process should first be addressed. Besides, innovative methods and infrastructure for natural cell acquisition require optimization, along with high-quality storage of natural cells and efficient characterization methods. During the production preparation process, it is imperative for the surface markers and specific functions of natural cells to not be destroyed. Yet, this remains an urgent problem in the current natural cell biomimetic nanoparticle drug delivery system towards clinical application. In addition, the tumor permeability, systemic cumulative toxicity, self-degradability and body excretion pathway after targeting biomimetic nanoparticle drug delivery to the tumor site still need to be explored.

The ultimate goal of discovering and developing biomimetic nano-drug delivery system is to reach the clinical stage and achieve large-scale industrial production. Most of the studies on the drug-loaded biomimetic NPs are still in the preclinical stage. Some formulations have undergone clinical research and may be used to serve patients soon (Table 3). In conclusion, despite the great success of natural cell biomimetic nanoparticle drug delivery for treating tumors, this innovative strategy still stays at the laboratory stage and cannot meet the requirements for clinical applications because there are currently no suitable methods to ensure the controllability of the preparation process and the safety and efficacy of the use process. However, challenges always coexist with opportunities. From the laboratory to the clinic, there are still many issues that must be addressed urgently.
But the unique advantages and application potential of the natural cell biomimetic nanoparticle drug delivery system are still undeniable, which provides the design and application of nano-drugs. The new thinking mode will have a place in the field of precision medicine.

### Table 2. The application of natural cell based biomimetic cellular transformers in the digestive system cancer

| Cancer type      | Nanomaterials | Sources of biomimetic materials | Cargo                  | Surface modifier | Target                  | Outcome/properties                                               | Biosecurity                          | Ref          |
|------------------|---------------|---------------------------------|------------------------|------------------|-------------------------|------------------------------------------------------------------|--------------------------------------|-------------|
| Esophageal cancer| Poly (lactic-co-glycolic acid) (PLGA) | RBC membrane                  | TE10 cells membrane   | Doxorubicin/curcumin | Desp-PEG 15000         | TE10/Dox xenografted tumor                                        | Homologous targeting effect, synergistic anti-tumor effect | High biosafety | [214]       |
| Gastric cancer   | Zeolitic imidazole frameworks-8 | PLGA                            | RBC membrane           | Paclitaxel       | Anti-EGFR-RGD          | EGFR extracellular domain, αvβ3 and neuropilin-1 receptors     | Dual tumor-targeting strategy, improve the radioresponsitization | Relatively safe concerning both manufacture and clinical use | [215]       |
| Gastric cancer   | AGS cell      | PLGA                            | RBC membrane           | Paclitaxel/triptolide | none                     | AGS tumor                                                        | Synergistic therapy of SDT and chemotherapy, induce pyroptosis of cancer cells and play anti-tumor effects | Excellent biosafety and biosafety | [216]       |
| Gastric cancer   | SGC7901 cell membrane | PLGA                            | RBC membrane           | Paclitaxel       | Injectable hydrogel    | SGC7901 tumor                                                    | Enhanced retention effect of nanoparticles in tumors | Low systemic toxicity | [217]       |
| Gastric cancer   | KPC cells membrane | PLGA                            | RBC membrane           | Paclitaxel       | Anti-EGF-R-PEG M2ep peptide | Pancreatic orthotopic tumor                                      | Targeting both tumor cells and M2-like TAMs, achieve effective combination of chemotherapy and immune checkpoint inhibitor therapy | Combinational therapy is biosafe | [218]       |
| Pancreatic cancer| KPC cells membrane | PLGA                            | RBC membrane           | Gemcitabine      | M2ep peptide           | pancreatic orthotopic tumor                                      | Targeting both tumor cells and M2-like TAMs, achieve effective combination of chemotherapy and immune checkpoint inhibitor therapy | Combinational therapy is biosafe | [219]       |
| Pancreatic cancer| β-cyclodextrin | PLGA                            | RBC membrane           | Gemcitabine/erlotinib/IRAK4 siRNA | none                     | PANC-1/ SW1990 tumor                                             | In vitro synergistic anti-proliferation, anti-migration and pro-apoptosis effects | Excellent biosafety and biosafety | [220]       |
| Pancreatic cancer| none | PATU-8988 cells membrane | Paclitaxel/triptolide | none | PATU-8988/PL-45 cells | Induced cellular apoptosis and suppressed cell proliferation via the inhibition of RAS signaling | none | [221]       |
| Pancreatic cancer| Gold nanoparticles | PLGA                            | RBC membrane           | Cyclopamine/oral administration | none                     | Capan-2 tumor xenografts                                         | Obtained excellent PTT efficacy | Combinational therapy is biosafe | [222]       |
| Pancreatic cancer| PLGA | RBC membrane                  | Cyclopamine/paclitaxel | none | Capan-2 tumor xenografts | Improved drug delivery to tumor, significantly enhance the anti-tumour efficacy | Improved drug delivery to tumor, significantly enhance the anti-tumour efficacy | Combinational therapy is biosafe | [223]       |
| Colorectal cancer| human platelet membrane | none | none | R848 | none | MC38 tumor                                                        | Active tumour-specific T cell immune responses and long-term protective immunity | No acute toxicity | [224]       |
| Colorectal cancer| DC-MC38 fusion cell membrane | PLGA                            | CpG ODN                | none | MC38 tumor                                                        | A vaccine to prevent tumor development, a therapeutic agent to regress established tumors | Safe for in vivo use | [225]       |
| Colorectal cancer| Zine gallogenanter persistence luminescence nanomaterials | PLGA                            | RBC membrane           | Erythrocyte/20ST cell hybrid membrane | none | CT26 tumor                                                        | Excellent immune escapability and cancer active targeting ability | Excellent in vitro and in vivo biosafety | [226]       |
| Colorectal cancer| Leukocyte-deriv ed membrane | none | none | Doxorubicin | none | HCT-116 organoids                                                  | Target the inflamed vasculature associated with colorectal cancer | none | [227]       |
| Colorectal cancer| Bovine serum albumin/Fe304 | none | none | Doxorubicin | none | CT26 tumor                                                        | Homologous targeting, combinational SDT and CDT could accelerate cell apoptosis | Excellent in vivo biosafety | [228]       |
| Colorectal cancer| Mesoporous silica and zinc gallogenante | none | none | Lactobacillus reuteri biofilm | 5-FU | none | HT-29/MC-38 cells                                                  | Withstanding the digestion of gastric acid and targeted release 5-FU to colorectum | Good biosafety and no toxic effects on the primary organs | [229]       |
| Liver cancer     | Pectin | RBC membrane                  | Doxorubicin | none | BEL-7402 tumor                                                    | Prolong blood circulation, enhance active targeting              | Poorly effective targeting | Low systemic toxicity, no significant in vivo toxicity | [230]       |
| Liver cancer     | PLGA | RBC membrane                  | Doxorubicin | none | Nanosone | Huh-7/Hepa-1 tumor                                               | Reverse the immunosuppressive TME, potentiate the systemic antitumor immunity | None | [231]       |
| Liver cancer     | PLGA | HUVEC membrane                | Oxaliplatin/hydroxychloro- quine | none | TRAIL tumor/HEK293 cells | Orthotopic HCC/LM3 tumor/HepG2 and Huh-7 or HCCLM3 metastatic tumor | Execute targeted autophagy inhibition and antitumor metastatic effect | Low systemic toxicity and excellent biosafety | [232]       |
| Liver cancer     | PLGA | RBC membrane                  | Doxorubicin | none | H22 tumor                                                        | Exert combinatorial antitumor effects of chemotherapy and thermotherapy | Excellent biosafety and no obvious cardiotoxicity | [233]       |
**Table 3. Clinical status of natural cell based biomimetic cellular transformers in the digestive system cancer**

| Conditions                          | Interventions/treatment                                      | Carrier source | NCT number          | Phase | Clinical Status      |
|------------------------------------|---------------------------------------------------------------|----------------|---------------------|-------|----------------------|
| Advanced Esophageal Cancer         | Anti-MUC1 CAR-T combined with PD-1                           | CAR-T cell     | NCT03736326         | Phase 1/2 | Recruiting          |
| Gastric Cancer                     | Pharmacological: activated DCs                              | DCs membrane   | NCT03410732         | Phase 2 | Recruiting          |
| Pancreatic Cancer                  | Drug: GRASPA                                                 | RBCs           | NCT01523080         | Phase 1 | Completed           |
| Rectal Cancer                      | None                                                          | Tumor EVs      | NCT04552633         | None   | Recruiting          |
| colon cancer                       | Curcumin                                                     | Plant exosomes | NCT01294072         | Phase 1 | Recruiting          |
| Colorectal Cancer                  | NK22D CAR-NK                                                | CAR-NK cell    | NCT01293159         | Phase 1 | Recruiting          |
| Gastrointestinal Junction Cancer   | N-803, Pembrolizumab, PD-L1 t-haNK                           | CAR-NK cell    | NCT01547466         | Phase 2 | Recruiting          |
| Gastrointestinal Cancer            | Cyclophosphamide, Fludarabine, Aldesleukin, Pembrolizumab    | CAR-T cell     | NCT03412877         | Phase 2 | Recruiting          |
| Solid Tumor                        | Cyclophosphamide                                             | CAR-T cell     | NCT02498912         | Phase 1 | Active, not recruiting |
| Solid Tumor                        | KITE-718, Cyclophosphamide, Fludarabine                      | CAR-T cell     | NCT03139370         | Phase 1 | Active, not recruiting |
| Solid Tumor                        | ROBO1 CAR-NK cells                                           | CAR-NK cell    | NCT03948020         | Phase 1/2 | unknown recruiting |
| Advanced Solid Tumor               | Anti-CAR-NK Cells                                            | CAR-NK cell    | NCT0194709          | Early Phase 1 | Recruiting |
| Metastatic Malignant Solid Neoplasm| Atezolizumab, Cyclophosphamide, Fludarabine, PDI Inhibitor | CAR-T cell     | NCT04639245         | Phase 2 | Suspended            |
| Metastatic Melanoma                | Cyclophosphamide, Fludarabine, Aldesleukin                   | Tumor infiltrating lymphocytes | NCT03169875 | Phase 2 | Terminated          |

**Abbreviations**

APCs: antigen-presenting cells; CTCs: circulating tumor cells; EVs: extracellular vesicles; MDSCs: myeloid-derived suppressor cells; NPs: nanoparticles; NETs: neutrophil extracellular traps; OMVs: outer membrane vesicles; PAMPs: pathogen-associated molecular patterns; PTT: photothermal therapy; PDEMs: plant-derived exosome-like nanoparticles; PEG: polyethylene glycol; PM-NV: platelet-mimicking nanovehicles; PDT: photodynamic therapy; TAAs: tumor associated antigens; PD-1: programmed death-1; PD-L1: programmed death ligand-1; RBCs: red blood cells; SIRP-α: signal regulatory protein-α; TME: tumor microenvironment; TAMs: tumor associated macrophages.

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**Competing Interests**

The authors have declared that no competing interest exists.

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