Review Article

Artemisia spp.: An Update on Its Chemical Composition, Pharmacological and Toxicological Profiles

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_Artemisia_ plants are traditional and ethnopharmacologically used to treat several diseases and in addition in food, spices, and beverages. The genus is widely distributed in all continents except the Antarctica, and traditional medicine has been used as antimalarial, antioxidant, anticancer, antiinocceptive, anti-inflammatory, and antiviral agents. This review is aimed at systematizing scientific data on the geographical distribution, chemical composition, and pharmacological and toxicological profiles of the _Artemisia_ genus. Data from the literature on _Artemisia_ plants were taken using electronic databases such as PubMed/MEDLINE, Scopus, and Web of Science. Selected papers for this updated study included data about phytochemicals, preclinical pharmacological experimental studies with molecular mechanisms included, clinical studies, and toxicological and safety data. In addition, ancient texts and books were consulted. The essential oils and phytochemicals of the _Artemisia_ genus have reported important biological activities, among them the artemisinin, a sesquiterpene lactone, with antimalarial activity. _Artemisia absinthium_ L. is one of the most famous _Artemisia_ spp. due to its use in the production of the absinthe drink which is restricted in most countries because of neurotoxicity. The analyzed studies confirmed that _Artemisia_ plants have many traditional and pharmacological applications. However, scientific data are limited to clinical and toxicological research. Therefore, further research is needed on these aspects to understand the full therapeutic potential and molecular pharmacological mechanisms of this medicinally species.

1. Introduction

The search and development of medicines from plant raw materials have been one of the important areas of human science for centuries [1]. A new breath of this scientific direction was given by the discovery of unique antimalarial agent artemisinin by the Chinese scientist Youyou Tu, for which she received the Nobel Prize in Physiology or Medicine in 2015. The source of this medicine was the _Artemisia annua_ L., which has long been known in Chinese folk medicine [2]. This plant is not the only one of the well-known representatives of the _Artemisia_ L. genus. _Artemisia_ genus (Asteraceae), named after the Greek goddess of the hunt and fertility Artemis, is considered one of the most widely distributed genera all over the world [3] and unites more than 400 species of plants of various life forms (grasses, shrubs, and less often trees) [4]. Many of them are weeds, some are invasive in certain regions of the planet, and at the same time, some species are listed in the Red Book. Wide distribution, a variety of component composition, and the resulting wide range of pharmacological effects made plants of the _Artemisia_ genus popular remedies of traditional medicine and ensured their study and subsequent introduction into official medicine [5–7].

The medicinal and aromatic applications of _Artemisia_ are well known for a long time as it produces volatile oil which has applications in medicine, cosmetics, and food production [8]. Some species of _Artemisia_ are edible [9], and others especially those grown in Korea have been traditionally applied in treating inflammations and ulcers. The most famous species around the world are _A. annua_ and _A. absinthium_ L. which are known for their uses in traditional medicine [10, 11]. In South Africa, _Artemisia afra_ Jacq. ex Willd. is commonly used for treating inflammation, coughs, colds, malaria, fever, influenza, and diabetes [12], while, in North America, _Artemisia dracunculus_ L. is widely used for the treatment of wounds and possesses antioxidant and antidiabetic activities [13, 14]. Furthermore, _Artemisia vulgaris_ L. is used in traditional medicine and has several activities such as anticancer, hepatoprotective, antiinflammatory, antimalarial, and insecticidal properties [15–19]. Other various species such as _Artemisia nilagirica_ (C.B.Clarke) Pamp., _A. dracunculus_, _Artemisia herba-alba_ Asso, _A. armeniaca_ Lam., and _Artemisia scoparia_ Waldst. & Kitam. possess significant therapeutic properties [20]. _A. scoparia_ also has a long history in medicine, and it has been in clinics as a diuretic, choleretic, and hepatoprotective [21]. _A. scoparia_ was used to treat hepatitis, jaundice, sores, pruritus, asthma, gastritis, and expel parasites and to treat spiders’ bites. A combination of _A. scoparia_, _Gardenia jasminoides_ J. Ellis, and rhubarb ( _Rheum rhabarbarum_ L.) was reported to be a classical prescription for curing jaundice [21]. Currently, several biopharmaceutical products containing _Artemisia_ extracts are available nowadays in the local markets to treat various diseases [22]. The purpose of the review is to systematize updated scientific data on the chemical composition and new insights in the pharmacological mechanisms of action and discusses the toxicological profiles of the _Artemisia_ genus.

2. Methodology

Two biomedical literature databases were searched for this review: PubMed/MEDLINE and Web of Science using the following MeSH search terms: “Artemisia/chemistry,” “Phytochemicals,” “Artemisinins/pharmacology,” “Artemisinins/therapeutic use,” “Medicine, Traditional,” “Phytotherapy/methods,” “Plant Extracts/pharmacology,” “Plant Extracts/therapeutic use,” “Plant Oils/pharmacology,” “Plant Oils/therapeutic use,” “Structure-Activity Relationship,” “Animals,” and “Humans.” Inclusion criteria are as follows: preclinical and clinical studies on the sources, acquisition approaches, experimental pharmacology, toxicology, and safety data regarding _Artemisia_ spp. were included. Both in vivo and in vitro pharmacological studies which underlie the molecular mechanism of action were included. Exclusion criteria are as follows: studies with data not relevant for the aim of this updated review, or poor quality of studies, duplicate studies. The taxonomy of plant species has been validated using the World Flora Online [23].

3. Geographical Distribution of _Artemisia_ spp.

The genus _Artemisia_ is widely cosmopolitan and distributed worldwide except the Antarctica [24–26]. The genus is heterogeneous and inhabits from the sea level to high altitudes
of around 4000 masl (meters above sea level) [27]. The species of *Artemisia* grows abundant in the Northern Hemisphere, and a low degree of colonization has been reported in the Southern Hemisphere [27, 28]. The main centre of species diversity of *Artemisia* is located in Central Asia consisting the region of Uzbekistan, Tadzhikistan, Turkmenistan, Kazakhstan, Kyrgyzstan, parts of Russia, China, and Mongolia. Other relevant centres of diversity include the territory of Iran-Turanian and Mediterranean regions and in western North America [29–33]. *Artemisia* has been spread beyond its native origin and successfully distributed and colonized in most of the arctic-alpine, temperate, and subtropical zones of the Northern Hemisphere. The distribution of the genus from Northern Asia primarily follows the main three routes: (1) in the West, it migrates into Europe, Western Asia, Mediterranean Basin, and Africa; (2) Siberia and into western North America; and (3) further south into Asia [34, 35]. Only a few number of species, not exceeding 25 taxa, have been reported from the Southern Hemisphere although a small diversity centre occurs in South America and it is found in Oceania as allochthonous taxa [32] (Figure 1).

4. Phytochemical Composition

4.1. Essential Oils. The essential oils (EOs) present in botanicals have been used for centuries in the form of spices, medicines, and their pleasant odour [36]. It has been possible only due to the development of distillation techniques in the middle ages and is used in their ancient applications in food, drugs, or cosmetics [37]. While, in the last decades, the EO industry entered different sectors with new dimensions and targets due to its various therapeutic applications. The chemical composition of *Artemisia* genus EOs has been reported by several authors around the world. The composition of EOs varies depending on several factors including the plant part, growing season, age of the plant, location, extraction techniques, solvent, and timing [38]. The detailed investigations on the EO composition of the *Artemisia* genus from different geographical regions have been presented in Table 1.

4.2. Other Bioactive Compounds. The phytochemical diversity assessment of the *Artemisia* genus exhibited the presence of different types of secondary metabolites reported by several authors around the world (Table 2).

5. Pharmacological Effects of *Artemisia* spp. Extracts and Its Bioactives: Underlying Molecular Mechanisms

*Artemisia* spp. have a broad range of pharmacological activities such as antiulcer, anticancer, hepatoprotective, antidiabetic, antioxidant, and antimicrobial. Some of the preclinical studies of these species’ activities are summarized in Table 3 and Figure 2.

One of the available famous drugs derived from *Artemisia* species is artemisinin which exists in the leaves and flowers of *A. annua*. Other species which are known free of artemisinin were found to be active against malaria as *A. vulgaris*, *A. absinthium*, *A. dracunculus*, and *A. scoparia*; this activity was attributed to EOs and other sesquiterpenes [89]. Moreover, other studies also mentioned that artemisinin was not the only antimalarial substance in *A. annua* extracts [90–92].

5.1. Antioxidant. Antioxidants are a group of compounds that can help support the integrity of cells in the face of free radicals, unstable molecules that our body inevitably produces [93–95]. Natural antioxidants are thus essential for the proper functioning of the body [96–98]. Several studies have been reported the antioxidant activity of *A. absinthium*. Phenolic compounds (gallic acid, coumaric acid, vanillic acid, syringic acid, and chlorogenic salicyclic acid) and flavonoids (quercetin and rutin) present in *A. absinthium* showed the potential of this plant against diseases related to oxidative stress [99–101]. These compounds reduce lipid peroxidation (thiobarbituric acid-reactive substances (TBARS) and recover endogenous antioxidant (e.g., superoxide dismutase (SOD) and glutathione (GSH)).

5.2. Anti-inflammatory. Inflammation is the body’s natural response to protecting itself and recovering from an injury [102, 103]. It has the function of protecting the body from harmful substances and regenerating the damaged tissue [1, 104, 105]. *A. absinthium* extracts exhibit anti-inflammatory properties which may be linked to its secondary metabolites including flavonoids and sesquiterpene-type compounds and their role in inflammatory regulator inhibition such as bradykinins, histamine, prostaglandins, and serotonin [106] and through suppression of proinflammatory mediator expression such as inducible nitric oxide synthase (iNOS), prostaglandin E-2 (PGE2), cyclooxygenase-2 (COX-2), factor nuclear factor-kappa-B (NF-κB), and tumor necrosis factor-α (TNF-α) [11].

5.3. Anticancer. Cancer is a disease in which the body’s cells grow uncontrollably, forming a tumor that can spread to different parts of the body [107–111]. The mechanism of the anticancer effect *A. absinthium* extract was due to the activation of the mitogen-activated protein kinase/extracellular signal-regulated kinase MEK/ERK signaling pathway, which in turn stimulates the mitochondrial pathway of caspase activation and regulates Bad and Bcl-2 family proteins, resulting in the apoptotic death of MCF-7 and MDA-MB231 human cancer cells [108].

5.4. Neuroprotective. Neurocerebral disorders, especially neurodegenerative ones, refer to several progressive brain syndromes that affect memory, thinking ability, behavior, and emotions [112–115]. *A. absinthium* has been shown to have neuroprotective effects on cerebral damage caused by reperfusion through its nicotinic and muscarinic action. The protective mechanism of ethanolic extract of *A. absinthium* may be due to its anticholinesterase activity as well as the ability to change the behavior of rats by restoring acetylcholinesterase (AChE) and monoamine oxidase (MAO) enzymes to near-normal activity [11]. The sesquiterpenoid dimer—caruifolin D—found in *A. absinthium* may
be used for the treatment of neurodegenerative diseases such as Alzheimer’s or Parkinson’s due to its inhibitory action on the production of neuroinflammatory mediators in BV2 microglial cells and the reactive oxygen species (ROS) production; leading to inhibitory effects on the activations of protein kinase C (PKC) and c-Jun N-terminal kinase (JNK) [116].

5.5. Hepatoprotective. *A. absinthium* hydroalcoholic extract improves hepatic function and lowers oxidative stress indicators and consistently stimulates and preserves the structural morphology of the hepatocellular membrane, resulting in lower serum aspartate (ASAT) and alanine aminotransferase (ALAT) activity. The proposed hepatoprotective mechanisms include liver microsomal drug-metabolizing enzyme suppression, free radical scavenging activity, and calcium channel blockage [117].

5.6. Antidiabetic. Diabetes is a metabolic disease that causes excess blood glucose (hyperglycemia) [118]. This disease is incurable and once diagnosed requires lifelong treatment [119]. *A. absinthium* extracts showed an insulin-sensitizing action due to their role in adenosine monophosphate-activated protein kinase (AMPK) stimulation and glucose transporter type 4 (GLUT4) activity. The proposed hepatoprotective mechanisms include liver microsomal drug-metabolizing enzyme suppression, free radical scavenging activity, and calcium channel blockage [117].

5.7. Antimalarial. A sesquiterpene lactone, artemisinin, which is the main active ingredient in *A. annua* is used for the treatment of *Plasmodium* parasites; these parasites are characterized by their substantial hemoglobin uptake and digestion. This produces large quantities of free redox-active heme and free ferrous iron (Fe²⁺), which are assumed to be responsible for artemisinin’s parasite specificity. Infected erythrocytes convert excess heme to hematin, which is toxic to the parasite due to oxidative damage and direct cell membrane rupture; but malarial parasites have evolved a detoxification mechanism that uses a biocrystallization process to convert hematin to the less toxic and inert crystallized hemozoin. Activated artemisinin has been shown to inhibit the development of hemozoin by alkylating heme. As a result, artemisinin’s activator and target are both free heme from hemoglobin breakdown [122].

### 6. Clinical Studies

Long traditional usage and functional preclinical studies of different *Artemisia* species for the treatment of several diseases encouraged their clinical evaluation to support the evidence of their potential as antimalarial, antioxidant, anticancer, antinociceptive, anti-inflammatory, and antiviral agents [4].

6.1. Anti-inflammatory Activity. In a randomized double-blind clinical trial, oral treatments of 42 patients by Arthrem (supercritical CO₂-extracted *A. annua*) at doses 150 mg and 300 mg or placebo twice daily for 12 weeks were tested for
| Plant species                  | Parts used | Chemical composition                                                                 | Region/country | References |
|-------------------------------|------------|--------------------------------------------------------------------------------------|----------------|------------|
| Artemisia absinthium L.       | L          | Camphor; E-caryophyllene; eucalyptol; germacrene D; α-cadinol                       | Brazil         | [39]       |
| Artemisia anethoides Mattf.   | AP         | 1,8-Cineole; terpinen-4-ol; 2-isopropyltoluene; pinocarvone                           | China          | [40]       |
| Artemisia annua L.            | AP, F, L   | Artemisia ketone; α-caryophyllene; germacrene D; (E)-β-Farnesene; germacranol B     | Russia         | [41]       |
| Artemisia arborescens (Vail.) L. | F, L       | β-Thujone, camphor, terpinen-4-ol, germacrene D, chamazulene                        | Italy          | [42]       |
| Artemisia argyi H. Lév. & Vaniot | L          | α-Thujone; bornanone; terpinen-4-ol; cis-2-methylheptol; bornol; cis-sabinol; α-terpineol; β-caryophyllene; germacryllyl oxide; neointermedeol | China          | [43]       |
| Artemisia campestris L.       | L, S       | β-Pinene; germacrene D; 1,8-cineole; 1,4-cyclohexadiene; β-Myrcene; 2-naphthalanemethanol; 2-decanone | Algeria        | [44]       |
| Artemisia dracunculus L.      | AP         | p-Allylinalisole; ocime (e)-β; ocime (z)-β; limonene                                | Portugal       | [45]       |
| Artemisia gmelinii Weber ex Stechm. | AP        | Cyclobutane etanol; endo-borneol; germacrene D; eucalyptol; selin-6-en-4α-ol; bisabolone oxide A; caryophyllene; terpinen-4-ol | China          | [46]       |
| Artemisia herba-alba Asso      | AP         | 3-Thujanone a; 3-thujanone b; camphor                                               | Saudi Arabia   | [47]       |
| Artemisia jordanica Danin     | L          | 2,3-Dehydro-1,8-Cineole; camphene; endo-borneol; bornyl acetate; geranyl isovalerate | Palestine      | [48]       |
| Artemisia judaica L.           | AP         | Butanoic acid; β-linalool; 2-cyclohexen-1-one, 3-methyl-6-(1-methylheyl); acenaphthen; davana ether | Jordan         | [49]       |
| Artemisia monosperma Delile    | AP         | γ-Costol; (Z)-en-yn-dicycloether; α-selinene; selina-4,11-diene (eudesma-4,11-diene); (E)-β-farnesene; 2-methylbutyl 2-methylbutyrate; (Z)-β-ocimene | Argentina      | [50]       |
| Artemisia nilagirica (C.B.Clarke) Pamp. | AP      | γ-Costol; (Z)-en-yn-dicycloether; α-selinene; selina-4,11-diene (eudesma-4,11-diene); (E)-β-farnesene; 2-methylbutyl 2-methylbutyrate; (Z)-β-ocimene | Saudi Arabia   | [51]       |
| Artemisia pedemontana Balb.    | F, L       | 1,8-Cineole; terpinen-4-ol; 2-isopropyltoluene; pinocarvone; trans-pinocarveol; α-pinene; 1,8-cineole; artemisaglutaric acid B and D | China          | [52]       |
| Artemisia persica Boiss.       | AP         | Santolina triene; α-thujone; eucalyptol; α-sabinene; trans-2-methylhept-1-ol; α-selinene, caryophyllene epoxide | Iran           | [53]       |
their efficacy on stiffness, pain, and functional limitations in osteoarthritis of the hip and knees. Results showed a significant decrease in visual analogue scale (VAS) score and improvement in WOMAC (Western Ontario and McMaster Universities Osteoarthritis) total score only at the low dose of 150 mg [154]. Afterwards, an open-label 6 month extension trial was proceeded to examine the safety of Artrem in the long run (6 months). Results showed that Artrem could be a safe and effective agent for osteoarthritis management [155]. In a similar study, 90 patients diagnosed with osteoarthritis applied 3% *A. absinthium* ointment, 3% *A. absinthium* liniment, or piroxicam gel (PG) on their knees for 4 weeks. Results showed that *A. absinthium* could be a safe and effective agent in the long run (6 months). Results showed that Arthrem ointment, 3% *A. absinthium* powder (750 mg three times daily) along with its basic therapy for 6 weeks, where another ten patients were selected for investigating the efficacy of anti-histamines [158].

Furthermore, in a randomized controlled clinical trial, 10 patients with Crohn’s disease administrated dried *A. absinthium* powder (750 mg three times daily) along with their basic therapy for 6 weeks, where another ten patients served as the control group. Results showed a significant reduction in serum TNF-α level and Crohn’s disease activity index (CDAI) scores as well as remission of symptoms in eight patients [157].

In a further clinical study, a nasal spray containing an extract of *A. abrotanum* mainly composed of EO (4 mg/mL) and flavonoids (2.5 μg/mL) was established for treating 12 patients with allergic rhinitis. The EO fraction is composed of 1,8-cineole, linalool, and davanone, while the flavonol fraction contained centauredin, casticin, and quercetin dimethyl-ethers, which are well known for their anti-inflammatory effect. Most of the patients exhibited a significant reduction in nasal congestion, sneezing, and rhinorrhea as well as relief of eye symptoms when compared to the effect of anti-histamines [158].

In the sight of cosmetics, twenty-five sensitive skin patients were selected for investigating the efficacy of *A. annua* extract on skin reliving. Results showed that consuming cosmetics having *A. annua* extract for 4 weeks could improve the hydration degree of the cheek cuticle by 63.90%, reduce the transepidermal water loss by 21.51%, reduce the sensitized area by 77.47%, repair skin damage, and reduce redness [159].

### 6.2. Anticancer Activity

Interestingly, artemisinin and its derivatives were reported as potent antitumor compounds with high selectivity on cancer cells without any side effects on normal cells [160]. Their mechanism of action is based on the cleavage of their endoperoxide bridge by Fe²⁺ in cancerous cells and the production of ROS involved in apoptosis, DNA damage, autophagy, and cell cycle arrest G0/G1 of the cancerous cells. Additionally, they can suppress angiogenesis by inhibiting the secretion of vascular endothelial growth factor (VEGF), vascular endothelial growth factor receptor 2 (VEGFR2), and kinase insert domain receptor (KDR)/flk-1 in tumors, as well as affecting different signaling pathways and transcription factors related to tumor growth [161, 162]. Besides the antitumor effect of artemisinin and its derivatives against human cancer cell lines *in vitro* [160, 163, 164], different clinical studies were established to ensure their potency. For instance, in a pilot clinical trial at phase II, the anti-tumor effect of dihydroartemisinin (200 mg/day) was tested against advanced cervical carcinoma for 3 weeks in ten women. Results showed a significant reduction in vaginal discharge and pain, with no sign of severe toxicity, as well as improvement in overall signs and symptoms. These patients also exhibited a lower expression of epidermal growth factor receptor (EGFR) and Ki-67 oncopgenes [165].

Further, in a randomized, controlled clinical trial, artesunate (120 mg, once daily, IV) was combined with vinorelbine (25 mg/m², once daily, IV) and cisplatin (25 mg/m², once daily, IV) to treat patients with the advanced lung cancer stage. Results showed significant improvement in the survival rate and hindering of the progression time of the cancerous cell, devoid of extra side effects [166]. In a case report conducted by Singh and Verma [167], artesunate (50 mg) proved to reduce the tumor size of the larynx of a patient with stage II cancer by 70% following 2 weeks of treatment [167]. As well in another case report conducted by Berger et al. [168], the combination of artesunate (50 mg twice/day) with standard chemotherapy showed a significant reduction in death risk and stabilization of the disease case in 2 patients with metastatic uveal melanoma stage IV, when compared to the chemotherapy alone [168].

### Table 1: Continued.

| Plant species                  | Parts used | Chemical composition                                                                 | Region/country | References |
|-------------------------------|------------|--------------------------------------------------------------------------------------|----------------|------------|
| *Artemisia tournefortiana* Rchb. | —          | cis-Spiroether; Z-β-farnesene; trans-nerolidol; camphor                            | India          | [66]       |
| *Artemisia scoparia* Waldst. & Kitam. | AP         | Spathulenol; acenaphthene; davana ether; 2-propenoic acid, 3-phenyl-ethyl ester     | Saudi Arabia   | [59]       |
| *Artemisia sieberi* Besser   | AP         | Acenaphthene; 2-cyclohexen-1-one, 3-methyl-6-(1-methylethyl)                         | Saudi Arabia   | [59]       |
| *Artemisia vulgaris* L.      | AP         | Caryophyllene; humulene; germacrene D; borneol; Caryophyllene oxide                 | Brazil         | [67]       |
|                              |            | Germacrene D; 1,8-cineole; β-pinene; sabinene; cis-thujone; β-caryophyllene;         | Lithuania      | [68]       |
|                              |            | Caryophyllene oxide; α-humulene; davanone                                          |                |            |

AP: areal part; L: leaves; F: flowers; —: not reported.
### Table 2: Chemical composition of *Artemisia* genus from different geographical regions.

| Plant species          | Chemical composition                                                                 | Region/country | References |
|------------------------|---------------------------------------------------------------------------------------|----------------|------------|
| *Artemisia abrotanum* L. | Caffeic acid; chlorogenic acid; isochlorogenic acid; protocatechuic acid; rosmarinic acid; quercitrin | Saudi Arabia   | [69]       |
| *Artemisia absinthium* L. | Artemisinin; α-thujone; β-thujone; bornyl acetate; 4-terpineol; camphene; camazulenol; cadinene; myrcene; guaiazulene; linalool; γ-terpinene | -              | [11]       |
| *Artemisia anuva* L.    | Armeanuin B; artemisinin; artemisinic acid; scopoletin                               | China          | [70]       |
| *Artemisia campestris* L. | Catechin; vanillic acid; syringic acid; p-coumaric acid; gallic acid; methoxyluteolin | Morocco        | [71]       |
| *Artemisia capillaris* Thunb. | Neohesperidin; naringin; quercetin; quercitrin; luteolin; kaempferol; genkwanin; astragalin | China          | [72]       |
| *Artemisia dubia* L. ex B.D.Jacks. | Calotroponeanl ester; α-aminin; nonacosanoic acid; docosanoic acid; tetracosanoic acid; 1-(O-tricosanoyl) glycerol; 1-(O-pentacosanoyl) glycerol; β-sitosterol | Pakistan       | [73]       |
| *Artemisia gmelinii* Weber ex Stechm. | Coumarins; phenolics; flavonoids; caffeoylquinic acids; diterpene glycosides | Korea          | [74]       |
| *Artemisia herba-alba* Asso | Coumarins; phenolics; flavonoids; diterpene glycosides | Morocco        | [75]       |
| *Artemisia indica* Wild. | 5-Hydroxy-3,7,4′-trimethoxyflavone; ludartin; maackiain; lupenol; cis-matricaria ester; trans-matricaria ester; 6-methoxy-7,8-methyleneoxy coumarin | China          | [76]       |
| *Artemisia lactiflora* Wall. ex DC. | β-Sitosterol; daucosterol; umbelliferone; isofraxidin; scopoletin; fraxidin; mandshurin; fraxin-8-0-β-D-glucopyranoside; eumonoside; scopol; 5,8-dihydroxy-7,4′-dimethoxy-flavone; syringing; chrysoeriol; tricin; luteolin; acacetin; apigenin; 5,7-dihydroxy-3,6,4′-trimethoxy-flavone; tectorigenin; eic soy/locysol-p-coumarate; isoferic acid; ferulaldehyde; ethyl caffeate; caffeic acid; (-)-syringaresinol; (1)-diisyringaresinol; p-hydroxybenzoic acid; α-methylbenzaldehyde; cleomiscosin C; cleomiscosin A or B; bisofraxidin | China          | [77]       |
| *Artemisia mongolica* (Fisch. ex Besser) Fisch. ex Nakai | 1-(3-Hydroxyphenyl)-2-(5-hydroxy-3-methoxyphenyl)ethane; 1-(3-hydroxyphenyl)-2-(3,5-dihydroxyphenyl)ethane | China          | [78]       |
| *Artemisia myriantha* Wall. ex Besser | Blumenol A; (+)-dehydrovomifol; (+)-3-hydroxy-β-ionone; (3R, 6R, 7E)-3-hydroxy-4, 7-megastigmen-9-one; (-)-10-oxo-isodauc-3-en-15-oic acid; isorivinan; eudesmafraglaucolide; artanomalide A; 13-acetoxy-3β-hydroxygermacra-1(10)-E, 4E, 7(11)-triene-12, 6α-olide; 13-acetoxy-3β-tigloylgermacra-1(10)-E, 4E, 7(11)-triene-12, 6α-olide; 13-acetoxy-3β-(3,9-δ-methylbutanoyl)germacra-1(10)-E, 4E, 7(11)-triene-12, 6α-olide (11), 3,9-diaceetoxy-13-hydroxy-1(10)-E, 4E, 7(11)-germacatrien-12, 6α-olide; 8α-angeloyloxyxostunolide | China          | [79]       |
| *Artemisia pontica* L. | n-Hexadecanoic acid; 9,12,15-(Z,Z,Z)-octadecatrienoic acid; 2-(4a,8,12-trimethyl-7-oxo-1,2,3,4,5,6,7,8-octahydrophenanthren-2-yl)-acetic acid; 3-oxo-(1H)quinolin-4-ol-2-one; neophytadiene | Ukraine        | [80]       |
| *Artemisia rupestris* L. | Citrusin A; alaschanioside A; coniferin; citrusin B; syringaresinol-β-D-glucoside; (6R,9S)-3-carbonyl-α-ionol glucopyranoside; 3,4-dihydroxy-5-methyl-7-isofraxidin | China          | [81]       |
| *Artemisia scoparia* Waldst. & Kitam. | Eugenol; capillene; sphenathenol; capillin; scoparone; tricosane; heptacosane; nonacosane; stigmastranol; tritriacontane | Serbia         | [82]       |
| *Artemisia splendens* Willd. | Quercetin-3-O-β-D-glucoside; 3,4-dihydroxy-5-methoxyxycinnamic acid; caffeic acid; 6,7-dimethoxy coumarin | Iran           | [83]       |
| *Artemisia turanica* Krasch. | Narcisin; quercitin; luteolin; kaempferol; genkwanin; astragalin; isorhamnetin-3-O-β-D-glucoside | Iran           | [84]       |
| *Artemisia vulgaris* L. | 3,5-Dicaffeoylquinic acid; 4,5-dicaffeoylquinic acid; 3,5,3′,4′-tetrahydroxy-7,5′-methoxy flavones | Iran           | [85, 87]   |
| *Artemisia manghili* (Bess.) B disagree. | Artanoic acid; luteolin; 6-methoxy luteolin; eupatilin; o-coumaric acid; vanillic acid; protocatechuic acid; 4-hydroxyphenyl acetate; vulgarin | Vietnam        | [88]       |
| Extract/compound                  | Doses                      | In vitro/in vivo | Route of administration/ assay | Model/cells                        | Activity                      | Potential effect                                      | Reference |
|----------------------------------|----------------------------|------------------|--------------------------------|------------------------------------|-------------------------------|-------------------------------------------------------|-----------|
| *A. nilagirica*/ethanolic extracts | 500 mg/kg                  | *In vivo*        | Orally                         | Rats                               | Antiulcer                     | Gastroprotective, ↑ proteins of mucus content         | [123, 124]|
| *A. nilagirica*/methanolic extract | 150–250 mg/kg              | *In vivo*        | Orally                         | Swiss albino mice                  |                               | Gastroprotective compared to standard drug vincristine | [125]    |
| *A. absinthium*, *A. vulgaris*/flowers/methanolic extract | 62.5, 125, 250, 500 μg/mL | *In vitro*      | MTT                            | MCF7 cells                         |                               | ↑ cytotoxicity IC<sub>50</sub> = 221–500 μg/mL         | [126]    |
| *A. nilagirica*/ethyl acetate, hexane fractions | 100 μg/mL                  | *In vitro*      | SRB                            | Hepatocellular carcinoma cells     | Anticancer                    | ↑ apoptosis IC<sub>50</sub> = 0.1 mg/mL                  | [15]     |
| *A. vulgaris*/leaves/methanolic extract | 0.01–1.0 mg/mL             | *In vitro*      | MTT                            | Hepatocellular carcinoma cells     | Anticancer                    | ↑ apoptosis IC<sub>50</sub> = 0.1 mg/mL                  | [15]     |
| *A. absinthium*/methanolic extract | 20, 25 g/mL                | *In vitro*      | MTT                            | MCF-7 MDA-MB231                   |                               | ↑ cancer cells suppression                             | [108, 128]|
| *A. armeniaca*/CH<sub>2</sub>Cl<sub>2</sub> fraction | 6.25–200 μg/mL             | *In vitro*      | MTS                            | Apoptosis-proficient HL60 apoptosis-resistant K562 |                               | HL-60: IC<sub>50</sub> = 75 μg/mL, K562: IC<sub>50</sub> = 130 μg/mL | [129]    |
| *A. dracunculus*/aerial parts, roots/ethanol, aqueous extracts | 250 mg/kg                  | *In vivo*        | Orally                         | STZ-induced diabetic rats          |                               | ↑ sensitivity of insulin, ↑ insulin receptor signaling | [130, 131]|
| *A. dracunculus L.* (PMI 5011)/ethanolic extract | PMI 5011 (1%)              | *In vivo*        | Diet                            | KK-A' mice                        | Antidiabetic                   | ↑ sensitivity of insulin, ↑ insulin receptor signaling | [130, 131]|
| *A. sieberi* (A. herba-alba)/aqueous extracts | 0.39 g/kg                  | *In vivo*        | Orally                         | Alloxan-induced diabetic rats      |                               | ↑ blood glucose, ↑ RBC, ↑ WBC, ↑ PCV, ↑ ESR, ↑ neutrophils, ↑ heart rate | [132]    |
| *A. persica*/aqueous, methanolic extracts | 300, 400, 500 mg/kg       | *In vivo*        | Orally                         | Sprague-Dawley rats               | Antihypertensive               | ↓ systolic blood pressure in normotensive/hypertensive rats | [133]    |
| *A. absinthium*/aqueous extract | 50, 100, 200 mg/kg         | *In vivo*        | Orally                         | Kunming mice, NIH mice            | Hepatoprotective              | ↓ inflammatory cells, ↓ liver lipid peroxidation, ↑ SOD, ↑ GPx | [134]    |
| *A. vulgaris*/aerial parts/crude extract | 150, 300, 600 mg/kg       | *In vivo*        | i.p.                           | Balb-C mice                       |                               | ↑ liver structure, ↓ parenchyma congestion, ↓ cellular swelling, ↓ apoptotic cells | [16]     |
| *A. nilagirica*/leaf extracts | 32–512 μg/mL               | *In vitro*      | Agar disk diffusion method      | 15 bacterial strains              | Antimicrobial                  | Methanol, hexane extracts, ↑ inhibition against phytopathogens | [135]    |
| *A. herba-alba*, *A. judaica*, *A. monosperma*/EO | 10.0, 5.0, 2.5, 1.0, 0.5 μL/disc | *In vitro*      | Agar disc diffusion method      | *Staphylococcus aureus* ATCC29213, *Escherichia coli* ATCC 25922 | Antibacterial | IC<sub>50</sub> = 0.5 – 2.5 μL. *A. judaica*, *A. monosperma* plants had the highest MIC | [136]    |
Table 3: Continued.

| Extract/compound | Doses | In vitro/in vivo | Route of administration/assay | Model/cells | Activity | Potential effect | Reference |
|------------------|-------|------------------|-------------------------------|-------------|----------|------------------|-----------|
| A. judaica/ethanol extract | 250, 500, 1000, 2000, 4000 μg/mL | In vitro | (mic90) growth inhibition | Protozoan parasite (blastocystis) | Antiprotozoal | IC<sub>50</sub> = 4000 μg/mL, ↓growth, ↓destruction of blastocystis | [137] |
| A. nilagirica/EO | 0.33 μL/mL | In vitro | Inverted petri plate technique | A. flavus, A. niger, A. ochraceus | Antifungal | ↑immobility time in the FST, ↓other activities in the OFT depressors of SNC | [138] |
| A. annua/leaves/EO ethanolic extract | EO = 470 mg/kg ethanol extract = 450 mg/kg | In vivo | i.p. | Wistar rats | Antidepressant | ↓immobility period in the FST and TST, dose-dependent antidepressant activity | [139] |
| A. vulgare/leaves/methanolic extract | 125, 250, 500, 1000 mg/kg | In vivo | i.p. | Swiss albino mice | Antiepileptic | Anticonvulsant activities were noticed using EPM and MBT | [140, 141] |
| A. capillaris/herba/ethanolic extract | 50, 100, 200, 400 mg/kg | In vivo | Orally | Mice | | | |
| A. nilagirica/leaves/ethanolic, aqueous extracts | | | | | | | |
| A. vulgaris/leaves/methanolic extract | 50, 100, 300 mg/kg | In vivo | i.p. | Swiss albino mice | Anti-Alzheimer | | |
| A. annua/aqueous, ethanolic extracts | 2 g/L | In vitro | ABTS, ORAC, FRAP | | | | |
| A. dracunculus L./leaves/methanolic extract | 20 μL | In vitro | DPPH | | | | |
| A. nilagirica/leaves/ethanolic, aqueous extracts | 100, 200 mg/kg | In vivo | i.p. | Swiss albino mice | Anti-Parkinson | | |
| A. scoparia, A. spicigera/methanolic extracts | 0.25; 0.125; 0.0625; 0.0312; 0.0156; 0.0078; 0.0039; 0.0019; 0.0009; 0.00048 mg/mL | In vitro | DPPH | | Antioxidant | Antioxidant activity of ethanolic extract > aqueous extract | [143] |
| A. nilagirica/EO, chloroform, petroleum ether methanolic extracts | 10; 8.6; 6.5; 6.5; 3.3; 2.5; 2 μg/mL | In vitro | Method recommended by WHO | Aedes albopictus mosquito | Insecticidal | LC<sub>50</sub> = 5 μg/mL | [146] |
| A. nilagirica/EO | — | In vitro | Method recommended by WHO | Aedes aegypti, Anopheles stephensi, Culex quinquefasciatus mosquito larvae | | | [146] |
| | | | | | | | |

A. nilagirica/EO, chloroform, petroleum ether methanolic extracts The EO of A. nilagirica was the most effective larvicide against A. stephensi larvae | [147] |
| Extract/compound | Doses | In vitro/in vivo | Route of administration/assay | Model/cells | Activity | Potential effect | Reference |
|------------------|-------|-----------------|-------------------------------|-------------|----------|-----------------|-----------|
| A. aucheri/methanolic extract | 25, 50, 100 mg/mL | In vitro | Scolicidal tests | Echinococcus granulosus | ↓ effect on the protoscolices of hydatid cysts | [148] |
| A. vulgaris/ethanolic extract | 1, 5, 10, 50, 100, 1000 ppm | In vitro | Method recommended by WHO | Aedes aegypti | LC50 = 65.8 ppm in 1 h and 18.6 ppm in 24 h, ↓ A. aegypti in various stages of its lifecycle | [19] |
| A. scoparia, A. spicigera/n-hexane, DCM, MeOH extracts | 20, 40, 80 mg/mL | In vitro | Toxicity bioassay | Tribolium castaneum (red flour beetle) | Insecticidal properties, ↓ activity of DCM extract | [145] |
| A. scoparia/butanol fraction | 20 mg/site | In vivo | Topically | BALB/C mice | Antiatopic dermatitis | ↓ clinical symptoms in a DNFB mouse model that induced lesions, ↓ inflammatory cytokines | [149] |
| A. scoparia/aerial parts/methanolic extract | 150, 300 mg/kg | In vivo | — | Sprague-Dawley rats | Nephroprotective | ↓ DNA damages, dose = 300 mg/kg, ↓ oxidative stress | [150] |
| A. capillaris Thunb/extract | 10 mg/mouse/day | In vivo | Topically | Dermatophagoides farinae-sensitized NC/NGA mice | Anti-inflammatory, anti-atopic dermatitis | ↓ dermatitis scores, ↓ bleeding, ↓ hyperkeratosis, ↓ hypertrophy in the dorsal skin and ear of the epidermis, ↓ histamine | [151] |
| A. pallens/aerial parts/methanolic extract | 200, 400 mg/kg | In vivo | Orally | Wistar rats | Anti-inflammatory, antioxidative | ↓ level of hepatic enzymes, ↑ renal antioxidant enzymes | [152] |
| A. vulgaris/leaf/ethanolic extract | 250, 500, 750, 1000 mg/kg | In vivo | Orally | ICR mice infected with P. berghei | ↓ P. berghei, nontoxic | IC50 = 0.778 mg/mL, IC50 = 0.998 mg/mL | [17] |
| A. scoparia, A. spicigera/dichloromethane extracts | 0–2 mg/mL 10% DMSO | In vitro | Heme biocrystallization and inhibition assay | | | IC50 = 3.27 nM, IC50 = 4.95 nM | [145] |
| A. annua/aqueous, hydro alcoholic extracts | — | In vitro | Parasite lactate dehydrogenase (pLDH) | Plasmodium falciparum | Antimalarial | | [153] |
| A. annua/aqueous hydro alcoholic extracts | Aqueous extract 1000 mg/kg/day, hydro alcoholic extract 500 mg/kg/day | In vivo | - | Plasmodium berghei NK173-infected mice | ↑ activity on malaria of artemisinin, both extracts of A. annua are effective on malaria | [153] |

↑: increase; ↓: decrease; MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide; DCM: dichloromethane; DNFB: 2,4-dinitrofluorobenzene; EPM: elevated plus maze; ESR: erythrocyte sedimentation rate; EO: essential oil; FST: forced swimming test; HDL: high-density lipoprotein; LDL: low-density lipoprotein; MBT: marble-burying test; MIC: minimum inhibitory concentration; OFT: open-field test; PCV: packed cell volume; RBC: red blood cell; SRB: sulforhodamine B; TC: total cholesterol; TGL: triglycerides; WBC: white blood cell; WHO: World Health Organization; i.p.: intraperitoneally; FRAP: ferric-reducing ability of plasma; ABTS: 2,2’-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid; ORAC: oxygen radical absorbance capacity.
Furthermore, artemether (40 mg once/day) proved to exhibit a significant improvement in the computed tomography scan of a 75-year-old patient with pituitary macroadenoma. Results showed a decrease in the tumor’s density as well as an improvement in the overall clinical signs and symptoms [169].

6.3. Antidiabetic Activity. In a randomized double-blind clinical study, 24 patients with impaired glucose tolerance (IGT) administrated A. dracunculus (1000 mg) or placebo before breakfast and dinner for 90 days. Results showed a significant decrease in systolic blood pressure, glycated hemoglobin, and total insulin secretion as well as a significant increase in the high-density lipoprotein cholesterol level [170].

In a dose-response clinical study, ethanolic extract of Artemisia princeps Pamp. containing eupatilin and jaseocidin was investigated for its antidiabetic effect in 81 patients with hyperglycemia. Patients were randomized into four groups: negative control (lactose 2000 mg/day), low-dose extract (2000 mg/day), and high-dose extract (4000 mg/day). Both doses significantly reduced glycated hemoglobin level, where the free fatty acid level in plasma was only lowered in patients administrated the high dose of the extract [171]. In a similar clinical trial, A. absinthium capsules (1 g/twice daily for 30 days) or placebo were administrated to 16 patients with type II diabetes. Results showed that A. absinthium reduced the glucose level by 32%, triglycerides by 10%, total cholesterol by 5%, and LDL level by 6% [172].

6.4. Antimalarial Activity. Malaria is considered the most common tropical disease that is provoked by certain parasites of the genus Plasmodium such as P. malariae, P. vivax, P. falciparum, and P. ovale [173]. Artemisia species are famous for their content of sesquiterpene lactones that are responsible for the high therapeutic potential of the genus [173, 174]. For instance, artemisinin and its derivatives are the most common sesquiterpene lactones among the genus. Dihydroartemisinin (the active form inside the biological systems) is produced by reducing the lactone of artemisinin. While, alkylation of the hemiacetal group yields arteether systems (all these derivatives are converted to dihydroartemisinin, where it possesses the highest activity, oral bioavailability, and tolerability with minimal side effects. These compounds are well known for their powerful activity against different species of Plasmodium as they contain the 1,2,4-trioxane moiety that may be responsible for the mechanism of action of the drugs [122]. The antimalarial activity of these compounds outcomes from the presence of Fe$^{2+}$ after the Plasmodium hemolysis. This Fe$^{2+}$ is utilized as a catalyst to open the peroxide bridge of the compound, leading to the formation of free radicals, alkylation Plasmodium proteins, and finally causing parasite death. Artemisinin also can perform its antimalarial activity by inhibiting PfATP6, an enzyme for the delivery of Ca$^{2+}$ into vesicles of the parasite, which is critical for its development [122]. In a cluster-randomized clinical trial performed in different African countries, it was determined that rectal artemesunate could take
from 4 to 6 h to reduce parasite load, progression of the disease, and risk of death, so it can be a good choice for patients who cannot be treated orally [176]. Moreover, the WHO reviewed different clinical trials performed by the African Quinine Artesunate Malaria Trial multicentre on 5400 children under the age of 15 years with multidrug-resistant severe malaria and suggested IV artesunate (2.4 mg/kg once daily) as a choice to treat malaria [177].

A meta-analysis study using single-patient data from different randomized, controlled trials was conducted to compare artemether and quinine in treating severe *P. falciparum* infection. Results showed that the death rate was significantly reduced with patients treated with artemether (14%) when compared to patients treated with quinine (17%). On the other hand, there was no difference between the 2 treatments in coma recovery, fever clearance, or the progress of neurological toxicity. However, the overall adverse outcomes of either death or neurological toxicity were significantly fewer in the artemether-treated group [178].

In a double-blind, randomized, placebo-controlled clinical trial, artesunate was combined with sulfadoxine-pyrimethamine to test their efficacy in reducing the timing of malaria exposure during the infancy stage. Results showed that innate cells produced a balanced level of pro-inflammatory and regulatory cytokines around 2 years of age, which was accompanied by a lower risk of clinical malaria [179]. Further, it was reported that patients with uncomplicated malaria who administrated this combination therapy exhibited an 84.1% cure rate [180].

Despite the potency of artesunate as an antimalarial drug, there are also two main antimalarial regimen options: dihydroartemisinin–piperazine (DHA–PPQ) and artemether-lumefantrine (AL), which are considered as the first option for treating uncomplicated *P. falciparum* malaria globally [181]. For instance, DHA–PPQ was reported to have high benefits in children with uncomplicated malaria in endemic countries [182]. On the other hand, the AL therapy (marketed as “Coartem”) could exert its antimalarial effect through opposing the erythrocytic stages of the parasite and so reducing the number of parasites. In addition, lumefantrine had a much-extended half-life time when combined with artemether and was supposed to clear residuals of the parasites [183].

As malaria causes severe maternal and fetal problems, the Centers for Disease Control and Prevention (CDC) suggested AL therapy for treating pregnant women with uncomplicated malaria in the United States throughout the second and third trimesters of pregnancy, at the same doses assigned for nonpregnant women [184]. In a case report implemented by Daddy et al. [185], dried leaves of *A. annua* were investigated for their efficacy as an antimalarial agent in patients (with *P. falciparum* infection) not responding to artemisinin combination therapy (ACT) or artesunate (IV). After oral administration of dried leaves of *A. annua* at a dose (0.5 g/twice daily/5 days), these patients exhibited a subside in clinical symptoms and the parasites were undetected microscopically [185]. In a large-scale double-blind, randomized clinical trial, the antimalarial effect of *A. annua* and *A. afra* infusions (1 L/day of dry leaf/twig infusion for 7 days) was compared to artesunate-amodiaquine (ASAQ) by 957 patients with malaria (with *P. falciparum* infection). Results showed that patients treated with both *Artemisia* infusions exhibited trophozoites clearance after 24 h when compared to ASAQ which took up to 14 days. Moreover, fever clearance took up to 48 h for ASAQ, but only 24 h for both *Artemisia* infusions. From days 14–28, gametocytes were undetectable for patients treated with *Artemisia* infusions, whereas on day 28, ASAQ-treated patients stayed carriers for gametocytes. These results proved that *Artemisia* infusions could break the life cycle of malaria by eliminating gametocytes with better outcomes than ASAQ [186]. In a questionnaire performed in Kenya and Uganda (2011) to study the antimalarial effect of *A. annua* teas, results showed prosperous outcomes after treating about 3000 cases including 250 children and 54 women in the first trimester of pregnancy with malaria [187].

6.5. Antiviral Activity as an Approach for Treating COVID-19 Infection. Recently, transposing of medications already in clinical use is the therapeutic strategy for controlling SARS-CoV-2 (COVID-19) infection [188, 189]. The WHO has recommended *A. annua* as a promising remedy for the treatment of COVID-19; however efficacy and side effects are not determined yet [190] (www.ClinicalTrials.gov, Identifier: NCT04530617). Moreover, *A. annua* is one of the ingredients of Jinhua Qinggan granule (one of the remedies suggested in the therapeutic regimen of COVID-19 in China) [191]. Currently, a phase II clinical study is under its way to evaluate the efficacy of *A. annua* in inhibiting the replication of the SARS-CoV-2 virus in patients with high-risk factors such as diabetes and hypertension (www.ClinicalTrials.gov Identifier: NCT04530617). As well, researchers from Saudi Arabia have established a placebo-controlled trial for evaluating the effect of artemunate in patients with mild symptoms of COVID-19 (www.ClinicalTrials.gov Identifier: NCT04387240).

Scientific evidence of this strategy might be due to the promising anti-inflammatory, immunomodulatory, and antiviral properties of the bioactive compounds in different *Artemisia* species, either among the preclinical or the clinical levels [4]. For instance, 85 patients with SARS were selected for a clinical study and 62 patients received the consigned treatment combined with the traditional Chinese medicine (TCM) (one of its components is herba *Artemisia*), while 23 patients were assigned in the control group. Results showed that patients who received the combined TCM regimen daily for 3 weeks showed a significant decrease in the total score of symptoms, as well as improvement of the lung X-rays, hepatic function, quality of life, and total score of mental sentiment factors [192].

Recently, it was reported that 1250 medical staff in Tongxu County Hospital take one or more decoctions of TCM daily as well as burn *Artemisia argyi* H.Lév. & Vaniot in the hospital corridor to cut off the route of transmitting infection, where *A. argyi* was stated as one of the herbs that can be used for contagion prevention by aromatherapy [193]. Artemisinin and its derivatives proved to have promising activities as antiviral agents. For instance, artemunate (100 mg/day) was examined for its efficacy to treat a 12-
The year patient with human cytomegalovirus (HCMV) infection who exhibited resistance against assigned antiviral drugs (foscarnet and ganciclovir) after stem cell transplantation. Results showed a significant reduction in viral load at day 7, with a virus half-life of 0.9–1.9 days, representing an effective stopover in viral replication [194].

7. Safety Issues of Artemisia Species

Various reports have been published regarding toxicity related to the overdosing in humans with extracts of Artemisia; A. absinthium (wormwood) was used in the formulation of the absinthe drink and currently has been restricted in most countries because of neurotoxicity [195, 196]; it was reported that few cups of sage tea or wormwood would be essential to reach the suitable daily intakes [195]. Preclinical studies of the toxicity of Artemisia species were examined in many reports and the most important results were summarized in Table 4. Regarding Artemisia spp. toxicity, different parts of Artemisia spicigera K. Koch and Artemisia fragrans Willd. significantly increased the number of MCF7 and HEK293 cell proliferation [126]. Moreover, the toxicological study of Artemisia judaica L. has been studied by Nofal et al. [197] and observed acute and chronic toxicity. Furthermore, Artemisia parviflora Roxb. ex D. Don showed no significant toxic effect on Swiss albino mice [198] as shown in Table 4. In addition, there is only one report on the toxicity of A. vulgaris [199] while different studies of A. annua demonstrated that it is considered safe and nontoxic up to 5000 mg/kg of the extract [200]. Artemether and the closely related compound arteether are hydrophobic derivatives of dihydroartemisinin reported to have neurotoxicity action [201].

8. Limitations, Therapeutic Perspectives, and Clinical Gaps

Although Artemisia spp. and its constituents show great potential as functional foods, dietary supplements, and safe medicines, some adverse effects have been described in the literature [206]. A. absinthium, grand wormwood, has been included traditionally as a major component of the highly anise-flavoured alcoholic spirit, “Absinthe,” which was the most popular alcoholic beverage of the late 19th century in Europe [207]. Absinthe was prohibited at the beginning of the 20th century as a consequence of adverse symptoms called absinthism [208]. Absinthism symptoms included hallucinations, blindness, mental deterioration, and convulsions. The prebanned Absinthe was probably related to chronic alcoholism [207]. Several drugs can interact with the effects produced by Artemisinin, and therefore, the specialist should be consulted before taking it. A. absinthium is permitted nowadays in foods and alcoholic beverages. The consumption of thujone from Artemisia must not exceed 10 mg/kg according to the European Food Safety Authority (EFSA) and 3 mg/day/person according to European Medicines Agency (EMA) [206].

Some Artemisia species are used in regulating fertility and thus should be avoided in pregnancy due to the possible risk of embryotoxicity at higher doses. For instance, the consumption of A. herba-alba to pregnant mice significantly decreased the fertility ratio of offspring mice [209]. Additionally, consumption of A. kopetdaghensis “Krasch., Popov & Lincz. ex Poljakov” hydroalcoholic extract in pregnancy increases the risk of abortion [210].

Skin contact dermatitis has been reported upon exposure to different Artemisia species [211, 212]. Skin prick testing showed that the majority of patients with allergic rhinitis and asthma have positive reactions to A. vulgaris. Therefore, patients with Compositae sensitization are routinely warned against exposure to Artemisia species [213]. Artemisia-induced dermatitis is attributed to the content of sesquiterpene lactones [214].

Pollens from Artemisia species can cause serious pollinosis [206]. Nasal challenge and bronchial provocation tests verified that pollens, leaves, and stems from Artemisia species are serious allergens causing allergic rhinitis and/or asthma [215, 216]. For instance, pollens from mugwort, A. vulgaris, contained allergenic substances such as profilin as well as other crossreactive allergens with immunoglobulin E (IgE) reactivity causing immediate type I allergic reactions [217]. Type I hypersensitivity involves mast cell degranulation and the release of inflammatory mediators such as histamine, causing allergic reactions such as anaphylactic shock [218]. Additionally, A. vulgaris pollens showed the highest levels of endotoxin among other collected plants across 100 locations in Europe [219]. Pollen extracts of six different Artemisia species, A. annua, A. scoparia, A. vulgaris, A. princeps, Artemisia campestris L., and Artemisia tridentata Nutt. exhibited an extensive degree of similarity and crossreactivity [220]. This study also showed that Korean and Norwegian patient sera had the same pattern of reactivity towards A. vulgaris and A. princeps [220].

Clinically, administration of the sesquiterpene lactone artemisinin as well as its derivatives such as arteether, artesunate, and artemether in appropriate therapeutic doses for short periods did not show serious side effects [221, 222]. In the liver, artemisinin is converted to various inactive metabolites, such as deoxyartemisinin, deoxoartemisinin, crystal 7, and 9,10-dihydrodeoxyartemisinin. The reaction is catalyzed by a CYP2B6 enzyme, while another CYP3A4 enzyme acts as a secondary catalyst. These enzymes belong to the cytochrome P450 group present in the smooth endoplasmic reticulum. Artemisinin derivatives are metabolized differently. They are first converted to dihydroartemisinin (DHA). DHA itself is a powerful antimalarial molecule and is active in the bloodstream for two to three hours [223]. The antimalarial activity of artemesunate is actually only through DHA [224]. Artesunate is converted to DHA within one minute of absorption. About 90% of all DHA is normally bound to blood plasma. In the liver, the cytochrome P450 enzyme system (including CYP2A6, CYP3A4, and CYP3A5) converts DHA into inactive metabolites [225]. All metabolites are subject to glucuronidation and are excreted in the urine or faeces. UDP-glucuronosyltransferases, especially UGT1A9 and UGT2B7, are responsible for the process [226]. DHA is also excreted in the bile as minor glucuronides, such as tetrahydrofuran acetate. Due to its rapid metabolism, artemisinin is a relatively safe
| Extract/compound | Doses          | In vitro/in vivo | Route of administration | Model                        | Effect                                                                 | Ref  |
|------------------|----------------|------------------|--------------------------|------------------------------|------------------------------------------------------------------------|-----|
| A. annua/leaf/hexane extract | 1000, 2000, 2500 mg/kg | In vivo          | i.p.                     | Wistar albino rats          | ↓carbohydrate, protein, lipid metabolisms, unfavourable effect on nutritional benefits, ↓hematological parameters, ↓toxicity when used acutely in rats | [202] |
| A. annua/hydroethanolic extract | 300, 2000, 5000 mg/kg | In vivo          | Orally                   | Swiss albino mice           | No toxic or lethal reactions of all the doses                            | [200] |
| A. parviflora/aerial parts/ethanolic extract | 0.10, 0.50, 1.0 g/kg | In vivo          | Orally                   | Swiss albino mice           | No significant toxic effect LD₅₀ > 1 g/kg BW                               | [198] |
| A. abyssinica, A. inculta/aerial parts/ethanolic extracts | 500 mg/kg, 1 and 3 g/kg | In vivo          | Orally                   | Swiss albino mice           | A. inculta, dose = 3 g/kg: CNS stimulation, A. abyssinica, dose = 3 g/kg: ↓locomotor activity | [203] |
| A. vulgaris/oils | —              | In vivo          | —                        | Brine shrimp Artemia sp. (larvae) | LC₅₀ = 10.4–23.3 μg/mL germacrene D, camphor, 1,8-cineole, davanone ↑toxicity | [199] |
| A. afr/aqueous extract | 1.5–5.5 g/kg i.t., 2–24 g/kg o.p. | In vivo          | i.p., orally             | BALB/C mice, Wistar rats     | Nontoxic when given acutely, low chronic toxicity, hepatoprotective effect in high doses | [204] |
| Artemether | 0, 20, 40, 80 mg/kg i.m., 0, 50, 150, 600 mg/kg p.o. | In vivo          | i.m., orally             | Beagle dogs                  | High i.m. doses: neurological damage, dose = 20 mg/kg: minimal effects occurred | [201] |
| Artemether, artesunate | 30–100 mg/kg/day | In vivo          | i.m.                     | Swiss albino mice            | Artemether neurotoxicity is significantly more neurotoxic than i.m. artesunate | [205] |
bioactive compound [227]. Because artemisinin interactions with other drugs are not fully known, future clinical studies are needed to establish their potential interaction mechanisms.

Other limitations derive from the amounts of active ingredients in plants, depending on the area of cultivation and climate [228–230]. Also, the absorption of bioactive compounds rapidly and in variable quantities requires the future development of pharmaceutical nanoformulations that will improve the bioavailability and implicitly an increased therapeutic efficacy [231, 232].

9. Overall Conclusion

Artemisia spp. have been traditionally used for pharmacological purposes and as an edible plant used in food, spices, and beverages. A. annua and A. absinthium are the most famous Artemisia species. This genus distributed worldwide presents diverse chemical constituents mainly EOs and polyphenols. These species contain sesquiterpene lactones that are largely responsible for the therapeutical potential of Artemisia genus. The most studied biological activities of this genus are antioxidant, anti-inflammatory, antitumor, antidiabetic, antimalarial, neuroprotective, and hepatoprotective activities through preclinical and clinical evidence. Artemisia spp. and their constituents show great potential as dietary supplements, functional foods, and safe medicines as antimalarial, antioxidant, antitumor, antinociceptive, anti-inflammatory, and antiviral agents. The antiviral activity for treating COVID-19 infection is a hope for the current pandemic. However, it is really important and necessary for further research investigations for discovering safer Artemisia plant-derived drugs for curing several kinds of diseases.

Data Availability

The data supporting this review are from previously reported studies and datasets, which have been cited. The processed data are available from the corresponding author upon request.

Conflicts of Interest

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

Authors’ Contributions

All authors made a significant contribution to the work reported, whether that is in the conception, study design, execution, acquisition of data, analysis, and interpretation or in all these areas, that is, revising or critically reviewing the article, giving final approval of the version to be published, and agreeing on the journal to which the article has been submitted and confirm to be accountable for all aspects of the work.

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