Nitrogen Dynamics Associated with Organic and Inorganic Inputs to Substrate Commonly Used on Rooftop Farms

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Abstract. Employing rooftops for the cultivation of crops in limited urban space has garnered interest in densely populated cities in the United States, where there is a growing demand for locally sourced vegetable products. Fertility management recommendations for rooftop farming, however, are scant. With insufficient research on nutrient cycling within rooftop farming systems, which tend to use soilless substrates with low organic matter content, the potential tradeoffs between the negative impacts (e.g., nutrient runoff) and the benefits (e.g., increased locally produced vegetables, stormwater retention, etc.) associated with rooftop farms are unclear. The objective of this study was to evaluate the effects of organic and inorganic nitrogen (N) inputs on the N dynamics within substrate typically used on rooftop farms. Substrate without added N inputs (control) was compared with substrates receiving N sources that are both realistic for and/or reflective of amendments currently applied on urban rooftop farms: a synthetic fertilizer (Osmocote® 14N–4.2P–11.6K), and three organic N inputs—composted poultry manure, municipal green waste (MGW) compost, and vermicompost. Aboveground crop biomass and yields of Beta vulgaris (swiss chard), along with inorganic N availability (ammonium: NH4+ and nitrate: NO3–), potential mineralizable nitrogen (PMN), leachate-inorganic N concentrations, and pH and electrical conductivity (EC) levels were measured during an 8-week greenhouse experiment. Despite differences in carbon-to-nitrogen ratios (C:N), few differences in N cycling and yields were found among the treatments receiving organic N inputs. Crop yields from the synthetic fertilizer and MGW compost treatments were higher than the other organic N input treatments. Inorganic N levels in the synthetic fertilizer treatment decreased from 129 mg N/L at the start of the season to 113 mg N/L at the end of the season, while nearly 10-fold decreases of inorganic N concentrations in the substrate of the control and organic N input treatments from week 0 (79.5–117.8 mg N/L) to week 8 (12.8–16.6 mg N/L) were observed. Greater N availability at critical periods during the season may have promoted greater crop N uptake efficiency and, therefore, higher yields in the system receiving synthetic fertilizer. However, the greatest losses of NH4+ and NO3– via leachate were also measured from this treatment. Our results show that the type of N input influenced plant-available N and yields and that the MGW compost treatment best achieved the balance between higher yields and reduced N losses to potential roof runoff. Furthermore, additional N inputs to these systems, particularly to the treatments receiving organic composts, will likely be necessary if a high N-demanding crop (such as swiss chard) is to be grown in the same substrates for more than 8 weeks. Rooftop farming is an emergent component of urban agriculture; regulations and guidelines for nutrient management of rooftop farms are necessary to optimize productivity and long-term benefits and to minimize negative environmental impacts.

Expanding agricultural capacity within or close to urban centers has the potential to supplement the growing demand for locally grown produce, while also reducing food transportation costs, lowering greenhouse gas emissions associated with food transportation, increasing food and economic security, and making a positive change in our food systems (Norberg-Hodge et al., 2002). Because land of suitable agricultural quality is often in high demand, and other forms of development are usually more economically viable, land availability is a major obstacle for urban farmers (Whittinghill and Rowe, 2012). Employing rooftops for the cultivation of crops in limited urban space has garnered interest and popularity in densely populated cities in the United States (e.g., Brooklyn Grange Farm in Queens and Brooklyn, NY; Eagle Street Rooftop Farm in Brooklyn, NY; Uncommon Ground, Chicago, IL). Experience with fertility management (e.g., basic N recommendations) and understanding of the environmental impacts associated with rooftop farms, however, is lacking.

Studies have shown that green roofs can provide multiple environmental benefits to urban areas, including attenuating surface temperatures, mitigating the urban heat island (UHI) effect, managing stormwater retention and flow, and restoring ecological habitat and biodiversity (Brenneisen, 2006; Oberndorfer et al., 2007; Solecki et al., 2005; Teemusk and Mander, 2007; VanWoert et al., 2005). While a rooftop farm essentially applies green roof technology for food production on a building roof, the aforementioned benefits associated with landscaped green roofs may not necessarily be associated with rooftop farms. A key question regarding the environmental impacts of rooftop farms is how does the fertility management of rooftop farms impact stormwater retention, detention, and pollution attenuation? One reason to expect a reduction in environmental benefits is rooftop farms operate with seasonal vegetation, i.e., biomass is low at planting and is then harvested at the end of the season; whereas, landscaped green roofs have perennial biomass cover. It has been suggested that plant establishment and the reduction of labile organic matter in the substrate via decomposition and assimilation would decrease the NO3– leaching from the vegetated green roofs (Berghage et al., 2009). Reduced environmental benefits can also be expected because rooftop farm crops will likely require more nutrient additions (e.g., N fertilizers) to maintain crop productivity compared with traditional green roof vegetation; this may lead to higher levels of nutrient leaching and unsafe pollutant contamination of stormwater runoff, a major concern for urbanized areas (Gobel et al., 2007; Groffman et al., 2004).

A number of factors (e.g., substrate composition, type of vegetation, and fertilizer additions) can potentially influence the quantity and quality of runoff from vegetated roofs (Czemiel Berndtsson, 2010; Hathaway et al., 2008). Studies evaluating the properties of the substrate material (Moran et al., 2005) and fertilizer application rate (Emilsson et al., 2007) have found that green roof systems can act as sources of NO3– and NH4+. Rooftop farms use soilless substrate, engineered to be lightweight while providing good moisture holding and aeration properties. However, these characteristics also mean that the substrate has limited nutrient holding and retention capacities. After the nutrients in the

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organic component (e.g., compost) of the substrate are depleted, rooftop farms rely on external inputs to maintain their productivity. Nitrogen, a key nutrient, can be introduced through the sole or combined additions of synthetic fertilizers and organic amendments, such as composted animal manure, composted plant and/or food materials, and green manures (e.g., cover crop residues). Unless N availability is well synchronized with plant need and uptake, there is a significant potential for the loss of added N from vegetated roofs via leaching after irrigation and/or precipitation events. Excess N from agricultural sources (e.g., fertilizer leaching, runoff from fields, etc.) and urban activities contribute to nutrient pollution in waterways and the degradation of neighboring riparian and marine ecosystems (Carpenter et al., 1998; Michael Beman et al., 2005). In turn, N leaching from agricultural systems translates to the loss of a principal—often limiting—nutrient in plant growth (Chapin III et al., 1986).

The type and form of N inputs play a major role in determining the fate and the quantity of N that will be available for plant uptake as well as N loss from an agricultural system (e.g., Goh and Haynes, 1986) and from green roofs (e.g., Clark and Zheng, 2013). Compared with synthetic fertilizers, which readily release plant-available N, organic amendments must be mineralized and therefore release mineral N to field crops slower and more gradually throughout the growing season (e.g., Burger and Jackson, 2003; Sikora and Szmidt, 2001). Mineralization of added organic matter is of particular interest to systems such as rooftop farms that apply organic amendments as the primary source of plant-available N. Composts, particularly those made from food scraps and yard trimmings, will likely be more accessible for urban agriculture as cities and countries recognize the impact of diverting organic waste from landfills on greenhouse gas emissions (Materials Management & Product Stewardship Workgroup–West Coast Climate and Materials Management Forum, 2011). While the gradual release of inorganic N from composts could also reduce N losses and substantial evidence indicates that compost applications can improve soil conditions (e.g., Roe, 2001; Smith, 1996), N availability from composts needs to be synchronized with crop demand to optimize crop yield. A better understanding of N dynamics and cycling in rooftop farm substrate would elucidate how to best manage rooftop farm systems for optimal productivity and environmental benefits, while minimizing N losses via runoff.

This study is one of the first investigations of N dynamics in substrate used on rooftop farms. The composts used in this experiment represent feasible N input options for both conventionally and organically managed urban rooftop farms in New York City, where this study took place. Three hypotheses were tested: 1) synthetic fertilizer will provide the highest N availability for crop growth, thereby leading to the highest yields, but the high levels of available N in excess of plant demand will also lead to the greatest loss of N in leachates compared with the other systems, 2) the organic N input with the lowest C:N ratio will release plant available N at a rate most closely synchronized with crop N demand, thereby leading to the smallest N loss among the systems and the highest yields among the organic N treatments, and 3) N release from the organic amendment with the highest C:N will not meet plant N demand and, therefore, lead to reduced yields and the highest leachate-N among the organic N treatments.

Materials and Methods

Experimental design. Starting in June 2012, an 8-week container experiment was conducted in a climate-controlled aluminum and glass rooftop greenhouse at Barnard College, Columbia University (New York, NY). Beta vulgaris (swiss chard) was selected as the test crop because of its high economic value to local rooftop farm businesses (B. Flanner, personal communication). rooftop® intensive ag (Skyland USA LLC, Avondale, PA), a soilless substrate designed for rooftop farms, consisting of lightweight mineral aggregates and an organic composted component [HydroRocks® (Big River Industries, Inc., Livingston, AL and Evrinenville, LA) and mushroom compost, respectively], was used as the base substrate (select characteristics shown in Table 1). rooftop® intensive ag, alone, served as the control to be evaluated against blends consisting of rooftop® intensive ag amended with four different N inputs (Table 2): “composted poultry manure” from the Stone Barns Center for Food and Agriculture (Pocantico Hills, NY); MGW compost from the New York City Department of Sanitation, which is available to urban farmers in New York City at no cost; “vermicompost” (compost produced by red wiggler worms, Eisenia fetida) from the Lower East Side Ecology Center (New York, NY), and “synthetic fertilizer” (Osmocote® Smart-Release 14N–4.2P–11.6K; The Scotts Company, Marysville, OH; a resin-coated, granular slow-release fertilizer (The Scott Company, 2009)). An average of 125, 110, and 170 g of composted poultry manure, vermicompost, and MGW compost, respectively, were added to an average of 730 g base substrate per container; these application rates were based on the 1 compost:4 substrate ratio (by volume) used at the Brooklyn Grange RoofTop Farm (B. Flanner, personal communication). The synthetic fertilizer was added to the substrate according to the manufacturer’s recommendation for vegetable plantings [2.5 g L–1 substrate (The Scott Company, 2009)]. Estimated rates of N application on an area basis are reported for each N addition in Table 2. The containers for both the control and fertilizer addition treatments contained 775 g substrate, which was equivalent in volume to the treatments receiving the organic N inputs (e.g., compost plus base substrate; ≈11 cm total substrate depth).

The experiment consisted of the control and four N input treatments, replicated five times per 2-week destructive sampling interval [5 treatments × 5 replicates × 5 sampling events = 125 total containers (experimental units)]. The containers (round, white, polypropylene pots; 15 cm diameter; 182 cm2 area; 14 cm height; ≈2 L volume; ITM–Meyers Industries Inc., Canada) were randomly arranged ±15 cm apart on greenhouse benches. Two B. vulgaris seeds per container were seeded on 19 June 2012 (week 0). Germination was observed on 22 June 2012 and swiss chard plants were subsequently thinned to one shoot per container on 3 July 2012 (week 2). The crop was watered via drip irrigation as needed to prevent wilting and grown to maturity (≈56 d).

Measurements

Crop and substrate sampling and analyses. Samples of the crop aboveground biomass and substrate were collected biweekly: at the start of the experiment (week 0) and weeks 2, 4, 6, and 8. Crop aboveground biomass (referred to as “yield” for the final harvest during week 8) was collected, weighed fresh for weight, dried at 60 °C until the mass no longer changed, weighed for dry weight, and then ground. From each experimental unit, one subsample of the moist substrate was collected (≈30 g), weighed, passed through a 2-mm sieve, dried at 60 °C, and then ground [in accordance with the standard methodology used for determination of chemical characteristics of green roof substrates (FLL, 2002)]; another subsample of the moist substrate was collected for inorganic N measurements via the Saturated Media Extract (SME) method (described below). Both aboveground crop biomass and substrate samples were analyzed for C and N content using a FLASH EA 1112 elemental analyzer (Thermo Scientific, Cambridge, U.K.).

Plant-available inorganic N (NH4+ and NO3–), pH, and EC values associated with the substrate were measured via the SME method, as described in Warncke (2011). The SME method is the currently established...
method of testing soilless greenhouse media and green roof substrate, both in the United States and internationally. Its goal is to extract the most soluble nutrients that are immediately available to plants. Briefly, deionized water was added to approximately 60 g moist substrate to create a saturated paste, and then allowed 90 min to equilibrate. The pH of the saturated paste was measured, then the saturated paste was 0.22-µm-vacuum filtered, and then EC was measured on the extracts before being stored at approximately -20 °C until colorimetric analysis. Extracts were thawed before ammonium (NH₄⁺) and nitrate (NO₃⁻) analyses using segmental flow analysis (SEAL AutoAnalyzer 3; SEAL Analytical GmbH, Norderstedt, Germany). Measurements of the substrates taken immediately after amending the rooflite® intensive ag with the four different N inputs are represented in Table 3 under sampling week 0.

Leachate collection and analyses. At the onset of the experiment (week 0) and each following week (weeks 1, 2, 3, 4, 5, 6, 7, and 8), leachate was collected following the PourThru program, adapted from Whipker et al., (2001). Before collecting leachate, all containers were irrigated to saturation. After draining for 1 h, deionized water was added to designated containers (e.g., during weeks 1 and 2, leachate was collected from pots assigned for destructive sampling in week 2) to collect approximately 70 mL leachate from each container (approximately 50% of total volume of deionized water applied). The leachate was subsequently filtered through Whatman no. 1 filter paper and EC and pH were measured. About 30 mL of the leachate was subsampled, 0.22-µm-filtered, and then stored at approximately -20 °C until colorimetric analysis for inorganic N content (in the same manner as that described above for the SME extracts).

PMN measurements. Potentially mineralizable N estimates have been widely used to assess the effects of various management practices, such as tillage, crop rotations, and fertilization, on N availability (e.g., Campbell and Souster, 1982; Doran, 1987; Franzluebbers et al., 1995). Potentially mineralizable N was measured using a 28-day aerobic incubation study, following the method outlined in Curtin and Campbell for mineral soils (Curtin and Campbell, 2006), which was similar to the method used to measure PMN in a soilless potting mix (Boydston et al., 2008). About 48 h before the start of the incubation, 10 g of 4-mm-sized moist substrate from each of the treatments (five replicates per treatment) was brought to approximately 20% water holding capacity (WHC) with deionized water. This equilibration or “priming” step is necessary to avoid the period of stimulated microbial activity upon rewetting the substrate. After priming, samples were brought to 60% WHC and sealed within canning jars (946 mL). On day 0 of the incubation, 3 h after being placed in the canning jars, one set of samples was extracted with 100 mL 0.0125 M CaCl₂ at a rate of 1 sample:10 extractant (w/v) [according to the guidelines for soil analyses of green roof substrates (FLL, 2002)] and then shaken on a reciprocating shaker at approximately 150 rpm for 1 h. The slurries were filtered through a Whatman no. 1 filter, and EC and pH were measured on the extracts. Extracts were 0.22-µm-syringe filtered and stored at approximately -20 °C. The second set of samples was incubated at approximately 25 °C for 28 d, and aerated every three days for approximately 10 min. On the final day of the incubation (day 28), the samples were removed from the jars and extracted in the same manner as the first set of samples. Frozen extracts were thawed and then analyzed for NH₄⁺ and NO₃⁻ as per the methods described above for the SME extracts.

Statistical analyses
Analysis of variance (ANOVA) was used to compare the sensitivity of substrate C and N content, yields and crop-N, inorganic N in leachate, pH and EC values, and PMN concentrations with regard to experimental factors (sampling week and N input) using the function aov() of the R software package (R, 2013). Means separation was found using Tukey’s honestly significant difference, and significance was determined at α = 0.05 unless otherwise stated. Because treatments were destructively sampled at different sampling periods, results were interpreted with caution and greater scrutiny.

Results and Discussion
Nitrogen dynamics
Plant-available nitrogen. With the exception of week 0, substrate NO₃⁻-N concentrations were consistently higher (by at least 30%) in the synthetic fertilizer treatment than the treatments receiving organic N inputs (Table 3). The higher availability of NO₃⁻-N in the synthetic fertilizer treatment was likely due to the formulation of this N input, i.e., readily available ammonium nitrate and ammonium phosphate. Despite week-to-week variability within the treatments (e.g., substrate NO₃⁻-N concentrations in the vermicompost treatment increased from week 0 to 2 and then decreased in the subsequent weeks), plant available NO₃⁻-N in the control and all four N input treatments decreased linearly over the course of the experiment (rates not shown). The NO₃⁻-N concentrations in the substrate of the control and organic N input treatments showed nearly a 10-fold decrease from week 0 to week 8 (Table 3); in contrast, NO₃⁻-N concentrations in the substrate of synthetic fertilizer treatment

Table 1. Select specifications (particle size distribution and water measurements) of the rooftop substrate (rooflite® intensive ag, Skyland USA, LLC) used in this study. These details were provided by B. Flanner (personal communication).

| Particle size analysis (diam, mm) (Wt %) |       |
|----------------------------------------|-------|
| <0.002                                 | 2.0   |
| 0.002–0.05                             | 4.6   |
| 0.05–0.25                              | 4.1   |
| 0.25–1.0                               | 10.0  |
| 1.0–2.0                                | 15.7  |
| 2.0–3.2                                | 16.9  |
| 3.2–6.3                                | 36.2  |
| 6.3–9.5                                | 10.4  |
| 9.5–12.5                               | 0.1   |
| >12.50                                 | 0     |

Physical measurements
Bulk density (dry, g·cm⁻³) 0.65
Bulk density (at max. water holding capacity, g·cm⁻³) 1.08
Total pore volume (%) 63.7
Maximum water holding capacity (%) 44.7
Air-filled porosity (at max. water holding capacity, %) 19.0
Saturated hydraulic conductivity (cm·s⁻¹) 0.058

Table 2. Total nitrogen (N) content and carbon-to-nitrogen (C:N) ratio of the organic N input additions and the substrate (control), along with the amount of N applied via the N additions to the different treatments. Mean separation in the same column, among organic N inputs only, by Tukey’s honestly significant difference test at α = 0.05.

| Treatment                     | N input | Total N (g N/kg) | C:N | N application (kg N/ha) |
|-------------------------------|---------|-----------------|-----|------------------------|
| Municipal green waste        | Composted municipal green waste | 11.0 b | 22.9 a | 189 |
| Composted poultry manure     | Composted poultry manure | 10.7 b | 23.0 a | 136 |
| Vermicompost                 | Vermicompost | 12.3 a | 13.0 b | 170 |
| Synthetic fertilizer         | Osmocote® Smart-Release | 14–14–14 | 6.57 | 19.5 | 126 |
| Control                      |         |                 |     |                        |

α = 0.05 (significant difference test at α = 0.05).
Table 3. Measurements of pH, electrical conductivity (EC), total nitrogen (N), carbon-to-nitrogen ratio (C:N), and inorganic N levels (NO$_3^-$-N and NH$_4^+$-N) in the substrate at sampling events during an 8-week greenhouse experiment. Means (n = 5) among treatments within a sampling week not followed by the same letter are significantly different at P $\leq$ 0.05.

| Sampling week | N input treatment | pH   | EC (dS·m$^{-1}$) | Substrate N (g N/kg) | C:N | NO$_3^-$-N (mg·L$^{-1}$) | NH$_4^+$-N (mg·L$^{-1}$) |
|---------------|------------------|------|------------------|----------------------|-----|------------------------|-------------------------|
| 0             | Control          | 7.39 | 5.61 a           | 6.37 a               | 19.8 a | 117.8 a                | 1.22 b                  |
|               | MGW compost      | 7.48 | 6.76 a           | 7.06 b               | 17.9 b | 99.3 b                 | 0.30 b                  |
|               | Synthetic fertilizer | 7.22 | 6.99 a           | 5.99 a               | 19.1 ab | 113.9 a                | 15.2 a                  |
|               | Composted poultry manure | 7.39 | 5.30 a           | 8.03 b               | 17.2 bc | 94.5 a                 | 0.88 b                  |
|               | Vermicompost     | 7.51 | 4.33 a           | 8.52 a               | 16.0 b | 79.5 a                 | 0.97 b                  |
| 2             | Control          | 7.51 | 4.38 b           | 8.07 ab              | 19.6 a | 70.2 c                 | 0.24 b                  |
|               | MGW compost      | 7.43 | 4.66 b           | 9.05 a               | 17.4 ab | 61.4 c                 | 0.39 b                  |
|               | Synthetic fertilizer | 6.97 | 7.11 a           | 6.63 b               | 19.1 a | 165.8 a                | 30.8 a                  |
|               | Composted poultry manure | 7.38 | 4.28 b           | 9.52 a               | 17.0 b | 88.8 bc                | 0.53 b                  |
|               | Vermicompost     | 7.51 | 4.56 b           | 9.86 a               | 16.0 b | 116.3 b                | 0.39 b                  |
| 4             | Control          | 7.65 | 3.14 a           | 8.82 b               | 18.4 a | 50.0 b                 | 0.50 b                  |
|               | MGW compost      | 7.70 | 4.83 a           | 8.88 b               | 16.5 b | 48.1 b                 | 1.03 b                  |
|               | Synthetic fertilizer | 7.39 | 4.78 a           | 8.69 b               | 16.9 ab | 131.8 a                | 28.7 a                  |
|               | Composted poultry manure | 7.61 | 3.30 a           | 11.6 a               | 16.2 bc | 61.3 b                 | 0.34 b                  |
|               | Vermicompost     | 7.72 | 3.31 a           | 9.51 ab              | 14.7 c | 64.7 b                 | 1.02 b                  |
| 6             | Control          | 7.55 | 4.26 ab          | 8.32 a               | 18.8 a | 60.0 a                 | 1.28 b                  |
|               | MGW compost      | 7.77 | 5.26 a           | 9.56 a               | 16.2 b | 26.9 b                 | 0.67 b                  |
|               | Synthetic fertilizer | 7.45 | 4.67 ab          | 8.85 a               | 16.9 ab | 68.3 a                 | 28.8 a                  |
|               | Composted poultry manure | 7.68 | 2.75 ab          | 9.94 a               | 17.4 ab | 39.4 a                 | 0.61 b                  |
|               | Vermicompost     | 7.71 | 2.45 b           | 9.55 a               | 15.4 bc | 33.5 b                 | 0.61 b                  |
| 8             | Control          | 7.46 | 2.53 a           | 7.30 b               | 19.0 a | 16.6 b                 | 0.49 b                  |
|               | MGW compost      | 7.38 | 3.33 a           | 9.50 ab              | 17.1 ab | 13.7 b                 | 0.64 b                  |
|               | Synthetic fertilizer | 7.00 | 4.41 a           | 7.50 b               | 19.1 a | 84.0 a                 | 29.0 a                  |
|               | Composted poultry manure | 7.39 | 2.73 a           | 10.4 a               | 16.8 b | 14.9 b                 | 0.84 b                  |
|               | Vermicompost     | 7.39 | 2.46 a           | 10.0 a               | 15.4 bc | 12.8 b                 | 0.67 b                  |

*Treatments include control, no added N input; composted poultry manure; composted municipal green waste (MGW compost); vermicompost; and synthetic fertilizer, Osmocote® Smart-Release 14N–4.2P–11.6K.

were $\approx 114$ mg·L$^{-1}$ at week 0, increased to $\approx 166$ mg·L$^{-1}$ at week 2, and did not fall below 68 mg·L$^{-1}$ for the remainder of the experiment (Table 3). The concentration of plant-available NH$_4^+$-N in the synthetic fertilizer treatment after week 0 was consistently $\approx 30$ mg·L$^{-1}$. This was an order of magnitude higher than NH$_2$-N concentrations in the treatments amended with compost ($<1$ mg·L$^{-1}$, Table 3). Plant available NH$_2$-N in neither the control nor the N input treatments showed significant changes across the season, though. The pool of plant-available NO$_3^-$-N in the synthetic fertilizer treatment was only $\approx 10$% larger than that of NH$_2$-N across the season, whereas, substantially more N was available as NO$_3^-$ than as NH$_2$-N in the control and organic N input treatments ($\approx 10$–100 times more). By the end of the season, the proportion of N available as NO$_3^-$ vs. NH$_2$ declined more in the control and organic N input treatments than in the substrate receiving synthetic fertilizer (data not shown). The greater decrease in the ratio of NO$_3^-$-N:NH$_2$-N suggests that N mineralization and subsequent release of NO$_3^-$-N and NH$_2$-N from the compost treatments and the control were less consistent than mineral N release in the synthetic fertilizer treatment (at the rates applied).

Substrate depths on rooftop farms are similar to intensive green roof depths ($>17$ cm). To the best of the authors’ knowledge, nutrient application recommendations are, however, not easily obtainable for intensive green roofs planted with vegetable crops. The common practice regarding intensive green roof fertilization is one that is endorsed by green roof expert, Charlie Miller, P.E., and that is to apply care and maintenance as prescribed to a similar garden situation (Roofmeadow, 2015). The total N application via the synthetic fertilizer treatment was similar to an N fertilization recommendation of 136 N kg·ha$^{-1}$ for field-grown swiss chard (Masabni, 2011); whereas, the total N applied from the composted poultry manure, MGW compost, and vermicompost were 13, 58, and 42% higher than the recommendation for field-grown swiss chard. Despite the lower N application rate, the synthetic fertilizer treatment provided a more consistent supply of both NO$_3^-$- and NH$_2$-N throughout the period of the study, which might have been better synchronized with swiss chard N need and uptake than the organic N input treatments with higher N application rates.

**Potentially mineralizable nitrogen.** Although the C:N ratios of the composts were expected to be strong predictors of the size of the PMN pools [i.e., the lower the C:N, the larger the PMN pool (Nicolardot et al., 2001)] and there were differences in C:N among the N input treatments at week 0 (Table 3), no significant differences in total PMN were found among treatments (Fig. 1). This suggests that the contribution of mineralization to plant-available N is not different among the treatments. While the lack of differences in PMN among the N input treatments corresponds with the similarities in substrate and inorganic N availability (NO$_3^-$ and NH$_4^+$) at week 0, it does not reflect the differences in inorganic and total N in the substrates at the end of the experiment (Table 3). Also, the higher total substrate N content of the compost amended systems at the end of the experiment suggested that these systems had a larger pool of total N that could be
higher concentration of NO– synthetic fertilizer addition had a significantly at weeks 0, 1, 4, 5, and 8, leachate from the differences in NH + concentrations of NO– from the organic N input treatments and the composites may have contributed to lower substrate as a result of the increase in C from the immobilization of existing NO – negative implications from the proliferation in the synthetic fertilizer treatment at all organic N input treatments.

Ammonium-N in the leachate. Rooftop farmers face the challenge of applying N in sufficient amounts to maintain crop yields, while not contributing to nutrient loading of stormwater runoff, which would diminish the benefits associated with a vegetated roof. Concentrations of NO3-N in the leachate from the synthetic fertilizer and organic N input treatments at week 0 ranged from 49.5 to 129.9 mg NO3-N/L (Fig. 2) and decreased to a range between 2.99 and 23.9 mg NO3-N/L by week 8. At weeks 0, 1, 4, 5, and 8, leachate from the synthetic fertilizer addition had a significantly higher concentration of NO3-N than leachate from the organic N input treatments and the control (Fig. 2). The relatively higher concentration of NO3-N in the leachate from the synthetic fertilizer treatment suggests greater negative implications from the proliferation of this type of N input than the compost amendments to rooftop farms. Reduced rates of mineralization of the composts and immobilization of existing NO3-N in the substrate as a result of the increase in C from the composts may have contributed to lower NO3-N concentrations in the leachate of the organic N input treatments.

Ammonium-N in the leachate was highest in the synthetic fertilizer treatment at all sampling weeks (Fig. 3). Concentrations of NH4-N in the leachate of the control and treatments receiving organic N inputs did not significantly change over the crop growth period (0.2–0.45 mg NH4-N/L) and no differences in NH4-N were found among the leachate of the organic N input treatments. The overall low levels of NH4-N in both the leachate and in the substrate may have been

Fig. 2. Total nitrate concentration (mg NO3-N/L) in leachate water from rooftop farm substrate during an 8-week greenhouse experiment. Treatments include control, no added N input to rooftop farm substrate; poultry manure, composted poultry manure; vermicompost; municipal green waste (MGW) compost, composted municipal green waste; and synthetic fertilizer, Osmocote® Smart-Release 14N–4.2P–11.6K. Error bars represent standard error of the mean (n = 5).
the substrate will change, and the porosity of the substrate may be altered by crop roots and compaction. On-farm research is needed to better understand the impacts of rooftop farm fertility management on stormwater runoff quality.

Yields and crop N. Swiss chard biomass (fresh weight of aboveground biomass) was similar among the treatments at week 2 (Fig. 4). In weeks 4 and 6, the synthetic fertilizer treatment showed a significant increase in Swiss chard biomass compared with the MGW compost treatment and the control. At harvest (week 8), the yield in the treatment receiving synthetic fertilizer (fresh weight = 44.4 g) was nearly 67% higher than the yield from the control (Fig. 4). Although biomass in the MGW compost treatment was lower than the synthetic fertilizer at weeks 4 and 6, yields between these two treatments were similar at harvest. Throughout the experiment, biomass and yields among the compost treatments were not different and did not differ from the control (Fig. 4).

In a greenhouse study, Echer et al. (2012) evaluated the impact of a range of urea-N fertilization doses on Swiss chard and found linear increases in total and marketable yields (5.4 t·ha⁻¹ increase for each 40 kg·ha⁻¹ N applied). Engelbrecht et al. (2010) found the fresh biomass weight of Swiss chard increased with increasing N doses and did not show a decrease even at an application of 800 kg N/ha. The N application rates in our study (126–189 kg N/ha) approached the highest N dose in the Echer et al. (2012) study (160 kg N/ha), but were less than 25% of the highest N dose used in Engelbrecht et al. (2010). Therefore, applying more N to the rooftop farm substrate might still lead to a linear increase in Swiss chard yields. However, other factors, such as phosphorus concentration, planting density, salinity, etc., could become limiting to growth at higher N input levels.

At harvest, N content of the Swiss chard yields among the four treatments were not different from the control (Table 4). Despite the generally higher inorganic N availability in the synthetic fertilizer treatment, yield N (33 g N/kg biomass) in the synthetic fertilizer treatment was similar to the yield N from the organic N input treatments and was only higher than N content in Swiss chard from the composted poultry manure treatment (21 g N/kg biomass). The higher C:N ratio in the Swiss chard from the composted poultry manure (C:N = 15.2) vs. the synthetic fertilizer addition treatment (C:N = 10.9; \( P = 0.06 \); Table 4) was consistent with the differences observed in yield N.

Fertility management

Substrate conditions: pH and EC. Along with organic matter content, green roof substrate characteristics that are important to nutrient release include salinity, pH, particle size, porosity, and WHC (FLL, 2002). We observed pH values within the range of 6.97 to 7.77 for all treatments across the season (Table 3). Values of pH were generally highest at weeks 4 and 6 and did not change significantly among the treatments.
significantly within treatments across the season, suggesting that these systems are well buffered (statistical significance among weeks not shown in Table 3). With the exception of week 6, the pH of the substrate of the synthetic fertilizer treatment was significantly lower than that of the control and organic N input treatments, which were generally not different. Although the measured pH values for the treatments fell within the range of pH 5.5–8.0 that is recommended in the FLL (2002) guidelines for actively growing intensive green roofs, these values are higher than the preferred pH range of 6.75 for swiss chard grown in mineral soils (Jett, 2005). The lower pH of the substrate in the synthetic fertilizer treatment (Table 3) may have, along with other factors, contributed to higher yields in comparison with the control and organic N input treatments.

Substrate EC values across all treatments ranged from 2.46 to 7.66 dS·m⁻¹ (Table 3), with EC values gradually decreasing from the start of the experiment toward harvest. This was to be expected as salts from the substrate and N inputs will gradually leach out with each irrigation event that exceeds substrate moisture holding capacity and plant water uptake. The substrate from the synthetic fertilizer and MGW compost treatments showed the highest EC values across the season (5.68 and 5.15 dS·m⁻¹, respectively). According to Warncke and Krauskopf (1983), substrate EC values exceeding 5.00 dS·m⁻¹ can result in plant wilting and leaf burn. Shannon and Grieve (1998) report that swiss chard growth diminishes in sand cultures with EC exceeding 11 dS·m⁻¹ (≈3.67 dS·m⁻¹ for saturated extract EC). While neither leaf burn nor wilting were observed in any of the treatments, rooftop farms using substrates similar to those in this study should monitor salinity to optimize crop productivity and substrate health.

Nutrient management for maximizing crop yields and reducing N loss. Like most agricultural operations, rooftop farms must maximize their crop production with minimal inputs to be viable. In the case of N inputs, cropping systems with high nitrogen use efficiency (NUE) are key to optimizing the trade-offs among production, profit, and environmental impact. Cropping system NUE can be increased by achieving greater uptake efficiency of applied N inputs, by reducing the amount of N lost from organic and inorganic N pools in the growth medium, or both. Nitrogen applied via synthetic fertilizer (126 kg N/ha) and MGW compost (189 kg N/ha) were the lowest and highest, respectively, among the treatments. The partial productivity factor of N applied (PFPN), calculated as the ratio of the Swiss chard yield (g dry weight per container) to the total amount of N applied to the treatment (g N per container), was highest for the synthetic fertilizer treatment (6.37 g yield/g N applied) and did not differ among the organic N input treatments (2.97–4.39 g yield/g N applied; Table 4). The substrate NO₃-N levels in the synthetic fertilizer treatment, which showed the highest yields, were significantly greater than the other systems only at weeks 2, 4, and 8 (Table 3), which suggests that mineral N availability during these periods was critical for maximizing crop growth. The higher concentration of inorganic N in the leachates from the synthetic fertilizer treatment is a trade-off of the higher NUE. This corroborates our hypothesis that the synthetic fertilizer would provide the highest N availability for crop growth, thereby leading to the greatest yields, but the levels of available N in excess of plant demand would also lead to the greatest loss of N in leachates, which increases the potential for negative environmental impact compared with the compost systems.

The NO₃-N and NH₄-N available in the substrate, as well as in the leachate of the vermicompost treatment were different from the other organic N input treatments (except at week 2; Table 3), despite the lower C:N ratio of the vermicompost treatment. Also, the yield from the vermicompost treatment was not different from the other composts. Therefore, our hypothesis that the organic N input treatment with the lowest C:N ratio would release plant available N (NO₃-N and NH₄-N) at a rate synchronized with crop N demand, and lead to the smallest N loss among the treatments, plus the highest yield among the compost systems was not corroborated. Moreover, our third hypothesis that N released from the organic N input treatment with the highest C:N ratio would not meet plant N demand and, therefore, lead to reduced yields, was not corroborated because the MGW compost treatment (one of the highest C:N; Table 3) produced yields similar to the other organic N input treatments.

Of the four N input treatments, the treatment receiving MGW compost-N best achieved the balance between higher yields and reduced N losses to potential roof runoff. It is not evident from our measurements, however, that inorganic N availability, total substrate N, and potential N mineralization concentrations contributed to the higher yields of the MGW compost treatment and the synthetic fertilizer treatments. Phosphorus and other macronutrients and micronutrients, which were not measured in this study, are also critical to crop growth (Fageria et al., 2010) and may have played a potential role in the differences observed in the yields. While the moisture holding capacities of the MGW compost, vermicompost, and composted poultry manure treatments (12%, 11%, and 15%, respectively, including base substrate) were similar, differences in porosity (not measured) may have also been influential in creating conditions that promoted crop growth and N loss.

Nitrate- and ammonium-N availability in the organic N input treatments and the control were lower at the end of the 8-week experiment than at the beginning of the season (Table 3; significance not shown). Williams and Nelson (1992) tested several organic N sources, including sewage sludge, poultry manure sludge, bone meal, pine needles, and poultry feathers, and found that all sources ceased releasing sufficient N after 6–7 weeks. To replenish the inorganic N pools and sustain crops grown in soilless rooftop farm substrate, applying N inputs throughout the growing season (which can be longer than the 8-week period studied in this experiment) and/or supplementing organic amendments with synthetic fertilizers might improve N availability, thereby maximizing yields, providing N for subsequent crops, and promoting long-term productivity. Cover cropping with leguminous N-fixing crops (e.g., vetch, clover, and/or beans) have been shown to replenish N pools and accumulate organic matter in traditional cropping systems (Dakora and Keya, 1997; Kuo et al., 1997; Stivers and Shennan, 1991), and should be considered for fertility management on rooftop farms.

Conclusions

Despite the lower total N applied, the higher inorganic N (NO₃-N and NH₄-N) availability in the substrate of the synthetic fertilizer treatment led to higher swiss chard yields and crop-N but also to higher inorganic N in leachate compared with the treatments receiving organic N inputs. Our measurements of total and inorganic substrate-N and NH₄-N and NO₃-N in the leachate suggest that the MGW compost system (which had the highest C:N ratio of the organic N input treatments) best achieved the balance between higher yields and reduced N losses via leachate. Moreover, our data show that while the form of N input influenced plant-available N and yields, additional N inputs, particularly to the treatments receiving composts, will likely be necessary if a high N-demanding crop (such as swiss chard) is to be grown in the same substrates in the long-term. It is clear that further research, particularly involving in situ experiments, is necessary to examine the complex dynamics between abiotic and biotic factors controlling mineralization and N availability in rooftop farm substrates receiving different amendments and fertilizers. As urban and rooftop farming become more prevalent, establishing standards, and recommendations for best management practices will be pertinent to
optimize productivity and minimize negative environmental impacts.

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