Review

Chromone and Flavonoid Alkaloids: Occurrence and Bioactivity

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Abstract: The chromone and flavonoid alkaloids represent an unusual group of structurally diverse secondary metabolites, derived from the convergence of multiple biosynthetic pathways that are widely distributed through the plant and animal kingdoms. Many of them have been discovered through bioassay-guided chemical investigations of traditional medicines, suggesting potential therapeutic significance. Their unique structures and varied pharmacological activities may provide important new leads for the discovery of drugs with novel mechanisms of action. Potential therapeutic indications are as diverse as cancer and viral infections, inflammation and immunomodulation, neurological and psychiatric conditions, and diabetes.

Keywords: flavonoids; alkaloids; flavonoid alkaloids; chromone alkaloids; natural occurrence

1. Introduction

There is absolutely no doubt that natural products have provided key leads for drug discovery [1]. The search for novel natural products with interesting biological properties is an ongoing exercise. Produced by many organisms including humans, alkaloids are a large group of secondary metabolites containing usually basic (in some cases neutral or quaternary) nitrogen derived from an amino acid (or a purine, pyrimidine or other source such as transamination in the case of pseudoalkaloids), in a heterocycle (or aliphatic in the case of protoalkaloids). Most alkaloids are classified chemically according to the nitrogen-containing ring system [2]. However, in terms of chromone and flavonoid alkaloids, this classification is based on the part of the molecule to which the nitrogenous moiety is
attached. A structure consisting of a nitrogen system (such as pyridine, piperidine, pyrrolidine) linked to the ‘A’ ring of chromone (Figure 1) is referred to as a chromone alkaloid [3]. This group of compounds can be sub-divided into two types, namely those in which the chromone nucleus exists as noreugenin (5,7-dihydroxy-2-methylchromone) (chromone alkaloids) and those which bear an aryl substituent at C-2 (flavonoid alkaloids or ‘flavoalkaloids’) [4]. The flavonoids appearing in the flavoalkaloid structures include flavans, flavons, flavonols, flavanones, flavanones, and flavan-3-ols (catechins and epicatechins). Chromone and flavonoid alkaloids are of interest not only due to their amphoteric (being both bases and phenols) nature, but because of the pronounced biological activity of some of the natural sources which contain them. In our continuing effort to review less well-known classes of natural products and their bioactivities [5], this article will discuss the natural occurrence and known biological activities of chromone and flavonoid alkaloids. However, those flavoalkaloids with more than two rings (except one tricycle) have not been discussed here for the sake of brevity.

**Figure 1.** Flavonoid core and chromone structure.

![Flavonoids, Chromone, Noreugenin](image)

2. Results and Discussion

2.1. Flavonoid Alkaloids

The first known flavoalkaloids ficine [5,7-dihydroxy-8-(1-methyl-2-pyrrolidinyl)-2-phenyl-4H-1-benzopyran-4-one] (1, Figure 2) and isoficine [5,7-dihydroxy-6-(1-methyl-2-pyrrolidinyl)-2-phenyl-4H-1-benzopyran-4-one] (2) were isolated from the wild fig, *Ficus pantoniana* King, Moraceae [6]. The alleged proteolytic and vermicidal properties of ficine reported in the literature (for example see [2]) are, in fact, related to the enzyme ficin and not the flavoalkaloid ficine.

**Figure 2.** Ficine (1) and isoficine (2).

![Ficine and isoficine](image)

Capitavine [5,7-dihydroxy-6-(1-methylpiperidin-2-yl)flavone] (3, Figure 3) is a flavoalkaloid from the seeds of *Buchenavia capitata* Eichler, Combretaceae [7]. Two other capitavine derivatives, namely 4’-hydroxycapitavine [5,7,4’-trihydroxy-6-(N-methyl-2”-piperidinyl)flavone] and 2,3-dihydro-4’-hydroxycapitavine [5,7,4’-trihydroxy-6-(N-methyl-2”-piperidinyl)flavanone] (5 and 6, respectively)
were also isolated from the same plant. However, N-demethylcapitavine and 2,3-dihydrocapitavine [5,7-dihydroxy-6-(N-methyl-2″-piperidinyl)flavanone] (4 and 7, respectively) were found only in the fruits of *Buchenavia macrophylla* Eichler.

**Figure 3.** Capitavine (3) and capitavine derivatives (4–7).

Buchenavianine (8, Figure 4) is the major alkaloid from the leaves of *B. macrophylla* and is also found in *B. capitata* [7]. O-Demethylbuchenavianine (9), which is found in both species, showed anti-HIV activity [8]. N-Demethylbuchenavianine (10) is another alkaloid from the leaves and fruits of *B. macrophylla*. Finally, N,O-bisdemethylbuchenavianine [5,7-dihydroxy-8-(2-piperidinyl)flavone] (11) has been isolated from the fruits of *B. macrophylla* [7].

**Figure 4.** Buchenavianine (8) and Buchenavianine derivatives (9–11).

Extensive and ongoing research has been conducted and funded by the U.S. National Center for Complementary and Alternative Medicine and the U.S. National Cancer Institute in support of the discovery of selective anti-HIV and anti-cancer drugs from natural sources. Therefore, the identification of novel molecules and their congeners such as these flavoalkaloids with relevant
bioactivities provides important leads for further drug development including structure-activity relationship analysis.

*Aquilegia ecalcarata* Maxim., Ranunculaceae, Chinese Columbine, is a Chinese medicinal plant used for the treatment of pustulosis, necrotic boils, and other infections [9]. Aquiledine [(2S)-6-(1,4-ureylenebutyl)-5,7-dihydroxyflavanone] (12, Figure 5) and isoaquiledine [(2S)-8-(1,4-ureylenebutyl)-5,7-dihydroxyflavanone] (13) were isolated from *A. ecalcarata* [10,11]. Their structures contain a saturated 1,3-diazepin-2-one ring attached to C-6 and C-8 positions of the flavonone.

**Figure 5.** Aquiledine (12) and isoaquiledine (13).

Cheliensisine (14, Figure 6) was isolated from *Goniothalamus cheliensis* Hu, Annonaceae, a plant growing in China. This plant has shown antineoplastic activity [12]. Cheliensisine belongs to the oxepinochromone family of natural products, for which syntheses are being developed due to their unusual structures and potential for drug development (for example, see [13]).

**Figure 6.** Cheliensisine (14).

*Lilium candidum* L., Liliaceae, Madonna lily, is a plant found in eastern Europe and western Asia. A pyrrolidinoflavonol called lilaline [3,4′,5,7-tetrahydroxy-8-(4-methyl-5-oxo-2-pyrrolidinyl)-flavone] (15, Figure 7) was isolated from the aerial parts (flowers) [14].

**Figure 7.** Lilaline (15).
In traditional medicine, it is believed that the flower of *L. candidum* has anti-inflammatory properties [15]. The relevance of the flavonoid constituents of lily flowers to anti-inflammatory activities such as cyclooxygenase inhibition has been demonstrated (for example, see [16]).

The only known pyrrolidinoflavan, vochysine [3,4-dihydro-2-(4-hydroxyphenyl)-7-methoxy-8-(2-pyrrolidinyl)-2H-1-benzopyran-5-ol] (16, Figure 8) was isolated from the fruits of the Amazonian tree *Vochysia guianensis* Aubl., Vochysiaceae [17].

**Figure 8.** Vochysine (16).

![Vochysine](image)

Sea-grass, *Phyllospadix iwatensis* Makino, Zosteraceae, grows in the coastal region of Japan and Russia. The flavoalkaloid, phyllospadine [4′,5,7-trihydroxy-6-methoxy-8-(1-methyl-2-pyrrolidinyl) flavone] (17, Figure 9) was isolated from this plant [18]. Phyllospadine is similar to two other flavoalkaloids, ficine and isoficine in terms of possessing an *N*-methylpyrrolidinyl substitution (Figure 2).

**Figure 9.** Phyllospadine (17).

![Phyllospadine](image)

Lotthanongine [afzelechin-(4β→2″)-N-(p-coumaroyl)-6″-hydroxytryptamine] (18, Figure 10) is a flavoalkaloid isolated from the roots of *Trigonostemon reidioides* Craib, (a synonym for *Baltispermum reidioides* Kurz), Euphorbiaceae [19]. In Thai traditional medicine, *T. reidioides* is used as an antidote for poisonous mushrooms, as an expectorant, laxative, and topical antiseptic. Lotthanongine contains an indole moiety attached to the C-4 position of an afzelechin molecule. There are many scientific publications on possible mechanisms by which flavonoid derivatives may be hepatoprotective, which would be relevant to a putative mechanism of the traditional antidote for poisonous mushrooms and as a valuable direction for future research on these flavoalkaloids.

A widely distributed member of the mint family in western China, *Dracocephalum rupestre* Hance, Lamiaceae, has been used in the treatment of cold, cough, icterohepatitis, laryngalgia, and various other diseases [10]. Sixteen flavoalkaloids called dracocephins [e.g., 4′,5,7-trihydroxy-6-(5-oxo-2-pyrrolidinyl)flavanone and 3,4′,5,7-tetrahydroxy-6-(5-oxo-2-pyrrolidinyl)flavanone] were found in the
aerial parts of this plant [18]. Dracocephins A–D (19–34, Figure 11) were determined to be mixtures of two diastereoisomeric pairs of enantiomers. Two chiral centers, C-2 and C‘-5, are responsible for this stereoisomerism. Dracocephins are the conjugates of a flavanone (naringenin for A and B or eriodictyol for C and D) with pyrrolidin-2-one (pyrrolidinone). As shown in Figure 11, the pyrrolidinone ring is attached to the C-6 position of the flavanone in dracocephins A and C and to the C-8 position in dracocephins B and D [21]. As suggested by the traditional uses above, in addition to the extensive body of literature on their anti-inflammatory and immunomodulatory activities, recent research has also shown flavonoid derivatives’ analgesic activity to be another field worthy of further investigation (for example, see [22]).

Figure 10. Lotthanongine (18).

Figure 11. Dracocephins A (19–22), B (23–26), C (27–30), and D (31–34).

Hyperglycemia affects the pathogenesis of diabetic complications by increasing protein glycation and the build-up of the advanced glycation end products (AGEs) in body tissues [23]. Two pyrrolidinone epicathechins, 6-(2-pyrrolidinone-5-yl)-(−)-epicatechin (35, Figure 12) and 8-(2-pyrrolidinone-5-yl)-(−)-epicatechin (36), isolated from the roots of the tara vine, Actinidia arguta (Siebold & Zucc.) Planch. ex Miq., Actinidiaceae, exhibited significant in vivo inhibitory activity against AGEs formation [24]. Even under a physician’s care, the American and Canadian Diabetes Associations report a very high prevalence of potentially life-threatening complications of diabetes including heart disease and stroke (cause of death in up to 80% of diabetics), blindness, kidney disease, nerve damage, erectile dysfunction and depression. For that reason any new leads, such as epicatechin-
alkaloid derivatives, for pharmacological interventions to reduce or prevent diabetic complications are extremely important.

Figure 12. Pyrrolidinone epicatechins (35 and 36).

Prolinalin A [6-C-[(2S,5S)-prolin-5-yl] quercetin] (37, Figure 13) and its 5″-epimer prolinalin B [6-C-[(2S,5R)-prolin-5-yl] quercetin] (38) are two C-prolinylquercetins isolated from the yellow cocoon shell of the silkworm, Bombyx mori L., Bombycidae [25]. Prolinalins are the first examples of a flavonoid (flavonol) conjugated with an amino acid (i.e., proline). The fact that these flavoalkaloids are only found in the silkworm and not in its host plant (i.e., mulberry) indicates that prolinalins are produced from dietary quercetin within the insect. This suggests that prolinalins from the diet are modified by a glucosyltransferase within the silkworm that can transfer a glucose residue to the C-5 hydroxy position of quercetin.

Figure 13. Prolinan A (37) and B (38).

Senecio argunensis Turcz., Asteraceae, is a plant growing in China. It is often used as folk medicine displaying antipyretic effects and detoxicant properties against dysentery [26]. Two pyrrolidinone flavonols namely ‘8-(2″-pyrrolidinone-5″-yl)-quercetin’ (39, Figure 14) and ‘8-(2″-pyrrolidinone-5″-yl)-isorhamnetin’ (40) were isolated from S. argunensis [27].

Kopsirachin [2-(3,4-dihydroxyphenyl)-3,4-dihydro-6,8-bis(octahydro-2,4,7-trimethyl-1H-2-pyridin-1-yl)-2H-1-benzopyran-3,5,7-triol] (41, Figure 15) is a flavoalkaloid isolated from the leaves of Kopsia dasyrachis Ridl., Apocynaceae [28]. It consists of two skytanthine moieties linked to the C-6 and C-8 positions of a catechin molecule.
As a folk medicine, the fern, *Davallia mariesii* T. Moore ex Baker, Davalliaceae (also placed in the Dryopteridaceae), is used in Korea to treat common cold, neuralgia, and stomach cancer; in China it is used to treat lumbago, rheumatalgia, toothache and tinnitus [29]. Davallioside A (42, Figure 16) and its 1″-epimer Davallioside B (43) are constituents of the rhizomes [30].

They are epicatechin glycosides carrying a γ-lactam (pyrrolidinone) substituent [8-(2-pyrrolidinone-5-yl)epicatechin-3-O-β-D-allopyranoside diastereomers].
Ethylpyrrolidinonyl theasinensin A (8′-ethylpyrrolidinonyltheasinensin A) (44, Figure 17) is a dimeric polyphenol having an N-ethyl-2-pyrrolidinone moiety. It was isolated from black tea, *Camellia sinensis* (L.) Kuntze, Theaceae [31]. It has been hypothesized that the Strecker aldehydes of tea amino acids, such as L-theanine, may be formed during the drying and enzyme deactivation stages of black tea production. These aldehydes can attack the A-rings of existed catechins, e.g., theasinensin, to form compounds such as 44 [32].

**Figure 17.** Ethylpyrrolidinonyl theasinensin A (44).

![Chemical structure of Ethylpyrrolidinonyl theasinensin A (44).](image)

2.2. Chromone Alkaloids

Rohitukine [5,7-dihydroxy-2-methyl-8-[4-(3-hydroxy-1-methyl)piperidinyl]-4H-1-benzopyran-4-one] (45, Figure 18) is a chromone alkaloid from the leaves and stems of *Amoora rohituka* (Roxb.) Wight & Arn. (a synonym of *Aphanamixis polystachya* (Wall.) R. Parker), Meliaceae [33], the stem bark of *Dysoxylum bincatariferum* (Roxb.) Hook. F. ex Hiern (a synonym of *D. gotadhora* (Buch.-Ham.) Mabb., Meliaceae [34], the bark of *Schumanniophyton magnificum* Harms (a synonym of *Tetrastigma magnificum* K. Schum.), Rubiaceae [35], and the bark of *Schumanniophyton problematicum* (A. Chev.) Aubrév. (a synonym of *Assidora problematica* A. Chev.), Rubiaceae [36]. Rohitukine showed antiinflammatory and immunomodulatory activities [37]. It also showed moderate cytotoxicity against human HL-60 promyelocytic leukemia and HCT-116 colon cancer cells [38]. Two other chromone alkaloids, rohitukine N-oxide (46, Figure 18) and N-demethylrohitukine-3′-acetate (47), were also isolated from the stem bark of *D. bincatariferum* [34] and *S. magnificum* [35], respectively. N-Demethylrohitukine-3′-acetate is found as a mixture of the 3′α- and 3′β-isomers in a 2:1 ratio. According to the U.S. National Cancer Institute, chemotherapy is often given as a combination of drugs. Combinations usually work better than single drugs because different drugs kill cancer cells in different ways. Thus, new leads such as the chromone alkaloids may prove to be valuable complements to the existing armamentarium of antineoplastic agents.

The piperidinone noreugenin [4-(5,7-dihydroxy-2-methyl-4-oxo-4H-chromen-6-yl)-piperidin-2-one] (48, Figure 19) and its N-methyl derivative [4-(5,7-dihydroxy-2-methyl-4-oxo-4H-chromen-6-yl)-1-methyl-piperidin-2-one] (49) were isolated from the root bark of *S. problematicum*, a west-African
plant [39]. In traditional medicine, the alcoholic extract from root of *S. problematicum* is used for treatment of agitated psychotic patients. In fact, the root extract of the plant showed interesting neuropsychopharmacological activities [40]. In addition to the above mentioned compounds, several complex chromone alkaloids, such as schumannniophytine, have also been isolated from *Schumannniophyton* species.

**Figure 18.** Rohitukine (45) and rohitukine derivatives (46 and 47).

![Rohitukine](image)

45. Rohitukine 46. Rohitukine N-oxide 47. N-Demethylrohitukine-3'-acetate

**Figure 19.** Piperidin-2-one-noreugenin (48) and *N*-methylpiperidin-2-one-noreugenin (49).

![Piperidin-2-one-noreugenin](image)

48. Piperidin-2-one-noreugenin 49. *N*-Methylpiperidin-2-one-noreugenin

Tubastraine [(3R,4S)-4-[7-[(4-bromobenzoyl)oxy]-5-hydroxy-2-methyl-4-oxo-4H-1-benzopyran-8-yl]-1-methyl-3-piperidinylester, 4-bromobenzoic acid] (50, Figure 20) is the first example of a chromone-containing metabolite of a marine invertebrate. This organobromine compound was isolated from the Pacific stony coral *Tubastrea micrantha* Ehrenberg, Dendrophylliidae (black turret coral) [41]. Tubastraine is believed to be responsible for the deterrent response to the crown-of-thorn seastar, *Acanthaster planci* L., Acanthasteridae, the major predator of stony corals. These predator deterrent substances from marine organisms are significant not only from an ecological perspective but also from a therapeutic perspective. The Developmental Therapeutics Program, Division of Cancer Treatment and Diagnosis, U.S. National Cancer Institute, is currently screening natural product materials derived from marine macro-organisms and micro-organisms as potential sources of novel anticancer drugs.

Several chromone alkaloids have been isolated from leaves and bark of *Dysoxylum acutangulum* Miq., Meliaceae [38,42], including chrotacumine B (51, Figure 21), C (52), D (53), and F (54). Other chrotacumines, e.g., A and E, are not presented here due to their more complex structures (tetracycles). The chrotacumines shown here share an *N*-methyl piperidine ring attached to a noreugenin molecule. Chrotacumines B, C, and D have additional ester side chains.
The flowers of the common Indian tree, Siamese senna, *Cassia siamea* Lam. (a synonym of *Senna siamea* (Lam.) H.S. Irwin & Barneby), Fabaceae, are used to treat insomnia and asthma and their extract has strong antioxidant activity [43]. A chromone alkaloid with unique structural features called cassiadinine \([5\text{-}((1Z)-1\text{-}(2\text{-amino-}4H\text{-imidazol-}4\text{-ylidene})\text{-}2\text{-oxopropyl})\text{-}7\text{-hydroxy-}2\text{-methyl-}4H\text{-benzopyran-}4\text{-one})\) (55, Figure 22) was isolated from the flowers [44]. Cassiadinine contains an acetonylchromone attached to a guanidine moiety. Through their effects to reduce levels of free radicals, antioxidants are intimately involved in the prevention of cellular damage, a common pathway for a variety of diseases, including cancer, and aging. That is another reason why polyphenols such as the flavonoid and chromone alkaloids are potentially valuable drug development leads.
Figure 22. Cassiadinine (55).

3. Conclusions

The chromone and flavonoid alkaloids are of particular interest for several reasons. They are not common but are widely distributed in at least 17 families of plants, including ferns, monocots, and from the most primitive to the most advanced orders of dicots. They have also been reported from two very widely separated types of animals, the insects and the corals, with respect to which evidence suggests endogenous biosynthesis rather than bioaccumulation from a dietary source. This genetic distance between source organisms suggests convergent evolution to similar approaches of chromone and flavonoid alkaloid chemical defenses against herbivores/predators. Such cases of convergent evolution imply a highly successful strategy that confers a survival advantage on the host. They also imply a strong potential for the discovery of further novel natural products with interesting bioactivities from organisms producing chromone and flavonoid alkaloids.

From a biochemical perspective, chromone and flavonoid alkaloids represent a very interesting convergence of multiple biosynthetic pathways, e.g., the shikimic acid pathway to the flavonoids, or the acetate to pentaketide pathway to the chromones, with addition of a nitrogenous moiety coming from L-ornithine to pyrrolidine, L-lysine to piperidine, or from other sources. The result is a group of secondary metabolites with diverse and unique structures.

With respect to bioactivities, many of these flavonoid and chromone alkaloids have been discovered through bioassay-guided chemical investigations of traditional medicines, suggesting that they have significant potential for drug discovery. As could be anticipated, the literature reports ecologically important activities of these substances against predators and pathogens, which often correlate with cytotoxic and antiviral activities. From a drug development perspective, the key challenge will be to determine, and attempt through derivatization to enhance the selectivity of these compounds, for example as targeted antineoplastic rather than generally cytotoxic agents, since natural selection tends to favour a broad spectrum of toxicity to predators and pathogens.

Since many alkaloids exhibit toxic effects through mimicking structurally and biosynthetically related neurotransmitters, perhaps the reported potential neuropsychiatric indications for these compounds could also have been predicted. However, other therapeutically important activities that have been discovered for these flavonoid and chromone alkaloids include potential applications against diabetes, inflammation, and immunological disorders.

Despite the tremendous success of certain naturally occurring substances as drugs, their limited accessibility and sometimes challenging synthesis often make them only leads and not candidate drugs for scientists and pharmaceutical companies in the quest for new therapeutic hits. Biochemists still
need preferably small molecules to demonstrate some degree of perturbation effects on biological processes. One of the most reliable sources of guidance is Mother Nature, through synthesis of natural product-inspired totally synthetic or semi-synthetic compounds [45]. Not surprisingly, some of the flavoalkaloid semi-synthetic compounds have shown interesting bioactivities (Figure 23). For example, flavopiridol (also known as alvocidib) (56) is a semi-synthetic flavoalkaloid which inhibited cyclin-dependent kinases CDK1 and CDK2 by alteration of tyrosine phosphorylation of CDK1/CDK2 and competitive inhibition with ATP [46]. As a CDK inhibitor, flavopiridol is a promising agent that induces p53-independent apoptosis in Chronic Lymphocytic Leukemia (CLL) [47]. L-868276 (57, Figure 23) is another semi-synthetic compound derived from rohitukine with specific and potent inhibitory effects on CDK2 [48].

**Figure 23**. Bioactive semi-synthetic flavoalkaloids: Flavopiridol (56) and L-868276 (57).

In conclusion, the evidence from the literature suggests that these chromone and flavonoid alkaloids have significant bioactivities that likely help their host organisms survive predation and disease. These bioactive compounds may be the basis for the host organisms’ uses in traditional medicines around the world. They have unique and diverse structures that will provide new leads for the discovery of drugs sufficiently different from previous discoveries in the alkaloid world to have perhaps different mechanisms of action and potentially to be effective in some health conditions where there is established multidrug resistance. Furthermore, the spectrum of bioactivities extends beyond those easily anticipated for alkaloids, increasing the potential for the discovery of new therapeutic agents from these unusual natural products.

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**Conflict of Interest**

The authors declare no conflict of interest.

**References and Notes**

1. Newman, D.J.; Cragg, G.M. Natural products as sources of new drugs over the last 25 years. *J. Nat. Prod.* 2007, 70, 461-477.
2. Buckingham, J.; Baggaley, K.H.; Roberts, A.D.; Szabo, L.F. *Dictionary of Alkaloids; 2nd ed.* CRC Press: Boca Raton, FL, USA, 2010.

3. Houghton, P.J.; Brossi, A. *The Alkaloids: Chemistry and Pharmacology, Chapter 3- Chromone Alkaloids*; Academic Press: San Diego, CA, USA, 1987; Volume 31.

4. Houghton, P.J. Chromatography of the chromone and flavonoid alkaloids. *J. Chromatogr. A* 2002, 967, 75-84.

5. Khadem, S.; Marles, R.J. Monocyclic phenolic acids; hydroxy- and polyhydroxybenzoic acids: Occurrence and recent bioactivity studies. *Molecules* 2010, 15, 7985-8005.

6. Johns, S.R.; Russel, J.H. Ficine, a novel flavonoidal alkaloid from *Ficus pantoniana*. *Tetrahedron Lett.* 1965, 24, 1987-1991.

7. Ahond, A.; Fournet, A.; Moretti, C.; Philogene, E.; Poupat, C.; Thoison, O.; Potier, P. Premiers alcaloïdes vrais isolés de Combretaceae: *Buchenavia macrophylla* Eichl. et *Buchenavia capitata* Eichl. *Bull. Soc. Chim. Fr. Partie I* 1984, 1-2, 41-45.

8. Beutler, J.A.; Cardellina, J.H.; McMahon, J.B.; Boyd, M.R. Anti-HIV and cytotoxic alkaloids from *Buchenavia capitata*. *J. Nat. Prod.* 1992, 55, 207-213.

9. Wu, Z.Y.; Zhou, T.Y.; Xiao, P.G. *Xin Hua Compendium of Materia Medica*; Shanghai Science & Technology Press: Shanghai, China, 1998; Volume 1.

10. Chen, S.B.; Gao, G.Y.; Leung, H.W.; Yeung, H.W.; Yang, J.S.; Xiao, P.G. Aquiledine and isoaquiledine, novel flavonoid alkaloids from *Aquilegia ecalcarata*. *J. Nat. Prod.* 2001, 64, 85-87.

11. Zhou, J.; Xie, G.; Yan, X. *Encyclopedia of Traditional Chinese Medicines: Molecular Structures, Pharmacological Activities, Natural Sources and Applications*; Springer: Heidelberg, Germany, 2011.

12. Gu, Z.; Liang, H.; Chen, H.; Xu, Y.; Yang, G.; Zhang, W. A new type of alkaloid from *Goniothalamus cheliensis*. *Acta Bot. Yunn.* 2000, 22, 499-502.

13. Bruder, M.; Haseler, P.L.; Muscarella, M.; Lewis, W.; Moody, C.J. Synthesis of the oxepinochromone natural products ptaeroxylin (desoxykarenin), ptaeroxylinol, and eranthin. *J. Org. Chem.* 2010, 75, 353-358.

14. Mašterová, I.; Uhrin, D.; Tomko, J. Lilaline—A flavonoid alkaloid from *Lilium candidum*. *Phytochemistry* 1987, 26, 1844-1845.

15. Picci, V. The exploitation of medicinal plants of the Mediterranean area. *J. Ethnopharmacol.* 1980, 2, 81-89.

16. Francis, J.A.; Rumbeiha, W.; Nair, M.G. Constituents in Easter lily flowers with medicinal activity. *Life Sci.* 2004, 76, 671-683.

17. Baudouin, G.; Tillequin, F.; Koch, M.; Vuilhorgne, M.; Lallemand, J.Y.; Jacquemien, H. Isolation, structure and synthesis of vochysine, a pyrrolidinoflavane from *Vochysia guianensis*. *J. Nat. Prod.* 1983, 46, 681-687.

18. Takagi, M.; Funahashi, S.; Ohta, K.; Nakabayashi, T. Phyllospadine, a new flavonoidal alkaloid from the sea-grass *Phyllospadix iwatensis*. *Agric. Biol. Chem.* 1980, 44, 3019-3020.

19. Kanchanapoom, T.; Kasai, R.; Chumsri, P.; Kraisintu, K.; Yamasaki, K. Lotthanongine, an unprecedented flavonoidal indole alkaloid from the roots of Thai medicinal plant, *Trigonostemon reidioides*. *Tetrahedron Lett.* 2002, 43, 2941-2943.
20. Ren, D.M.; Guo, H.F.; Yu, W.T.; Wang, S.Q.; Ji, M.; Lou, H.X. Stereochemistry of flavonoidal alkaloids from *Dracocephalum rupestre*. *Phytochemistry* **2008**, *69*, 1425-1433.

21. Zeng, Q.; Jin, H.Z.; Qin, J.J.; Fu, J.J.; Hu, X.J.; Liu, J.H.; Yan, L.; Chen, M.; Zhang, W.D. Chemical constituents of plants from the genus *Dracocephalum*. *Chem. Biodivers.* **2010**, *7*, 1911-1929.

22. Hagenacker, T.; Hillebrand, I.; Wissmann, A.; Büßelberg, D.; Schäfers, M. Anti-allodynic effect of the flavonoid myricetin in a rat model of neuropathic pain: Involvement of p38 and protein kinase C mediated modulation of Ca^{2+} channels. *Eur. J. Pain* **2010**, *14*, 992-998.

23. Ahmed, N. Advanced glycation endproducts—Role in pathology of diabetic complications. *Diab. Res. Clin. Prac.* **2005**, *67*, 3-21.

24. Jang, D.S.; Lee, G.Y.; Lee, Y.M.; Kim, Y.S.; Sun, H.; Kim, D.H.; Kim, J.S. Flavan-3-ols having a gamma-lactam from the roots of *Actinidia arguta* inhibit the formation of advanced glycation end products *in vitro*. *Chem. Pharm. Bull.* **2009**, *57*, 397-400.

25. Hirayama, C.; Ono, H.; Tamura, Y.; Nakamura, M. C-prolinylquercetins from the yellow cocoon shell of the silkworm, *Bombyx mori*. *Phytochemistry* **2006**, *67*, 579-583.

26. Roeder, E. Medicinal plants in China containing pyrrolizidine alkaloids. *Pharmazie* **2000**, *55*, 711-726.

27. Li, N.; Shao, L.; Zhang, C.F.; Zhang, M. Two new flavonoid alkaloids from *Senecio argunensis*. *J. Asian Nat. Prod. Res.* **2008**, *10*, 1143-1146.

28. Homberger, K.; Hesse, M. Kopsirachin, ein ungewöhnliches alkaloid aus der Apocynaceae *Kopsia dasyrachis* Ridl. *Helv. Chim. Acta* **1984**, *67*, 237-248.

29. Cui, C.B.; Tezuka, Y.; Kikuchi, T.; Nakano, H.; Tamaoki, T.; Park, J.H. Constituents of a fern, *Davallia mariesii* Moore. I. Isolation and structures of davallialactone and a new flavanone glucuronide. *Chem. Pharm. Bull. (Tokyo)* **1990**, *38*, 3218-3225.

30. Cui, C.B.; Tezuka, Y.; Kikuchi, T.; Nakano, H.; Park, J.H. Davallioside-A and Davallioside-B, novel Flavan-3-ol derivatives with a gamma-lactam, from the rhizomes of *Davallia mariesii* Moore. *Chem. Pharm. Bull.* **1990**, *38*, 2620-2622.

31. Tanaka, T.; Watarumi, S.; Fujieda, M.; Kouno, I. New black tea polyphenol having N-ethyl-2-pyrrolidinone moiety derived from tea amino acid theanine: Isolation, characterization and partial synthesis. *Food Chem.* **2005**, *93*, 81-87.

32. Drynan, J.W.; Clifford, M.N.; Obuchowicz, J.; Kuhnert, N. The chemistry of low molecular weight black tea polyphenols. *Nat. Prod. Rep.* **2010**, *27*, 417-462.

33. Harmon, A.D.; Weiss, U.; Silverton, J.V. The structure of rohitukine, the main alkaloid of *Amoora rohituka* (syn. *Aphanamixis polystachya*) (Meliaceae). *Tetrahedron Lett.* **1979**, *8*, 721-724.

34. Yang, D.H.; Cai, S.Q.; Zhao, Y.Y.; Liang, H. A new alkaloid from *Dysoxylum binectariferum*. *J. Asian Nat. Prod. Res.* **2004**, *6*, 233-236.

35. Houghton, P.J. New Chromone alkaloids from the stem bark of *Schumanniophyton magnificum*. *Planta Med.* **1988**, *54*, 239-242.

36. Houghton, P.J.; Woldemariam, T.Z. High-performance liquid-chromatographic analysis of chromone alkaloids from *Schumanniophyton* species. *Phytochem. Anal.* **1993**, *4*, 9-13.
37. Naik, R.G.; Kattige, S.L.; Bhat, S.V.; Alreja, B.; de Souza, N.J.; Rupp, R.H. An antiinflammatory cum immunomodulatory piperidinylbenzopyranone from Dysoxylum binectariferum: Isolation, structure and total synthesis. Tetrahedron 1988, 44, 2081-2086.

38. Ismail, I.S.; Nagakura, Y.; Hirasawa, Y.; Hosoya, T.; Lazim, M.I.M.; Lajis, N.H.; Shiro, M.; Morita, H. Chrotacumines A-D, chromone alkaloids from Dysoxylum acutangulum. J. Nat. Prod. 2009, 72, 1879-1883.

39. Schlittler, E.; Spitaler, U. On the contents of Schumanniphyton problematicum (Rubiaceae). Tetrahedron Lett. 1978, 2911-2914.

40. Amadi, E.; Offiah, N.V.; Akah, P.A. Neuropsychopharmacologic properties of a Schumanniphyton problematicum root extract. J. Ethnopharmacol. 1991, 33, 73-77.

41. Alam, M.; Sanduja, R.; Wellington, G.M. Tubastraine: Isolation and structure of a novel alkaloid from the stony coral Tubastraea micrantha. Heterocycles 1988, 27, 719-723.

42. Ismail, I.S.; Lazim, M.I.M.; Mahmood, I.I.; Shaari, K.; Malek, E.A. Chromone alkaloids of Dysoxylum acutangulum and dammarane triterpenes of Walsura chrysogyne. In Proceedings of The 14th Asian Chemical Congress, Bangkok, Thailand, 2011.

43. Kaur, G.; Alam, M.S.; Jabbar, Z.; Javed, K.; Athar, M. Evaluation of antioxidant activity of Cassia siamea flowers. J. Ethnopharmacol. 2006, 108, 340-348.

44. Biswas, K.M.; Mallik, H. Cassiadinine, a chromone alkaloid and (+)-6-hydroxy-mellein, a dihydroisocoumarin from Cassia siamea. Phytochemistry 1986, 25, 1727-1730.

45. Nandy, J.P.; Prakesch, M.; Khadem, S.; Reddy, P.T.; Sharma, U.; Arya, P. Advances in solution- and solid-phase synthesis toward the generation of natural product-like libraries. Chem. Rev. 2009, 109, 1999-2060.

46. Carlson, B.A.; Dubay, M.M.; Sausville, E.A.; Brizuela, L.; Worland, P.J. Flavopiridol induces G1 arrest with inhibition of cyclin-dependent kinase (CDK) 2 and CDK4 in human breast carcinoma cells. Cancer Res. 1996, 56, 2973-2978.

47. Christian, B.A.; Grever, M.R.; Byrd, J.C.; Lin, T.S. Flavopiridol in chronic lymphocytic leukemia: A concise review. Clin. Lymphoma Myeloma 2009, 9, S179-S185.

48. Azevedo, W.F.D., Jr.; Mueller-Dieckmann, H.J.; Schulze-Gahmen, U.; Worland, P.J.; Sausville, E.; Kim, S.H. Structural basis for specificity and potency of a flavonoid inhibitor of human CDK2, a cell cycle kinase. Proc. Natl. Acad. Sci. USA 1996, 93, 2735-2740.

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