Visible Human Project® female surface based computational phantom (Nelly) for radio-frequency safety evaluation in MRI coils

Gregory M. Noetscher1,2*, Peter Serano3,4*, William A. Wartman1*, Kyoko Fujimoto5*, Sergey N. Makarov1,2,4*

1 Department of Electrical and Computer Engineering, Worcester Polytechnic Institute, Worcester, Massachusetts, United States of America, 2 NEVA Electromagnetics, LLC, Yarmouth Port, Massachusetts, United States of America, 3 Ansys, Inc., Canonsburg, Pennsylvania, United States of America, 4 Athinoula A. Martinos Center for Biomedical Imaging, Massachusetts General Hospital, Charlestown, Massachusetts, United States of America, 5 Center for Devices and Radiological Health, US Food and Drug Administration, Silver Spring, Maryland, United States of America

These authors contributed equally to this work.

* gregn@wpi.edu

Abstract

Quantitative modeling of specific absorption rate and temperature rise within the human body during 1.5 T and 3 T MRI scans is of clinical significance to ensure patient safety. This work presents justification, via validation and comparison, of the potential use of the Visible Human Project (VHP) derived Computer Aided Design (CAD) female full body computational human model for non-clinical assessment of female patients of age 50–65 years with a BMI of 30–36 during 1.5 T and 3 T based MRI procedures. The initial segmentation validation and four different application examples have been identified and used to compare to numerical simulation results obtained using VHP Female computational human model under the same or similar conditions. The first application example provides a simulation-to-simulation validation while the latter three application examples compare with measured experimental data. Given the same or similar coil settings, the computational human model generates meaningful results for SAR, B1 field, and temperature rise when used in conjunction with the 1.5 T birdcage MRI coils or at higher frequencies corresponding to 3 T MRI. Notably, the deviation in temperature rise from experiment did not exceed 2.75˚C for three different heating scenarios considered in the study with relative deviations of 10%, 25%, and 20%. This study provides a reasonably systematic validation and comparison of the VHP-Female CAD v.3.0–5.0 surface-based computational human model starting with the segmentation validation and following four different application examples.

Introduction

Quantitative assessment of radio frequency (RF) absorption experienced by a patient undergoing a Magnetic Resonance Imaging (MRI) procedure is prohibitively difficult to obtain due to...
the invasive nature of such measurements. Numerical simulations with computational human models are used to estimate electric field strength, specific absorption rate (SAR), current densities in and around tissues, and expected temperature rise [1–9] and to comply with relevant international and domestic standards [10–14]. While well over 30 whole-body human models created from patients of various age, body composition, sex and race exist, (cf. for example, [15]), documented validation of these models (which involves verification against independent measurements and simulations under the same or similar settings) is not always available.

The Visible Human Project (VHP) Female v.3.0–5.0 computational human model (also known under the nickname 'Nelly' to users of Dassault Systèmes SIMULIA software CST), shown at left in Fig 1, is an anatomically accurate heterogeneous female (~60 years old, ~ 88 kg, BMI of ~36, obese, with heart pathology) surface-based human body model. It was constructed from the photographic cryosection data of the VHP conducted by the US National Library of Medicine. Its construction has been well documented in the literature [15–21], but several points are worth emphasizing here:

1. There are 249 distinct components or triangular 2-manifold surface meshes (with an additional 40 characterizing embedded implants). No intersections or joint faces between discrete meshes are allowed, enabling unique assignment of electromagnetic, thermal, or other material properties.

2. The source data for the model are freely and publicly available [22,23]. The complete co-registration data for all model cross-sections are also made publicly available [24].

One advantage of the model is its topological and computational simplicity. The model size is purposely limited to approximately 0.4 M triangular facets in total with an attempt to keep the anatomical accuracy within the body within 2–7 mm. This makes it possible to apply virtually any commercial or custom-tailored finite-element or boundary element computational solver in a very reasonable amount of time. Nearly identical results are obtained when using different software packages and numerical methods [25].

Yet another advantage is the separation of the muscular system of the body into individual muscle objects (about 50 in total).

Some recent uses of the VHP Female v3.0–5.0 computational human model have been documented in the literature—cf. for example [25–42]—and compared with experimental data on radio frequency propagation. A number of studies have focused on the characteristics and behaviors of antennas near the human body [26–34]. Others have examined through simulation various biomedical applications including transmission channel modeling [35], transcranial direct current stimulation [36], estimation of bone density [37], gastroenterology [38,39], SAR simulations in MRI coils at 3 T [40], and safety of active implants under MRI procedures [41,42]. A simplified version of the model, VHP-Female v. 2.2, has over 500 registered users and is available for download online [43].

However, a systematic model validation and comparison have not been performed. The present study attempts to fulfill this task using published experimental [44–49] and modeling [72] data, and starting with the initial segmentation validation. The most relevant context of use is a non-clinical assessment model for female patients of age 50–70 years with a BMI of 30–36 during 1.5 T and 3 T based MRI procedures. Note that, compared to validation and comparison methodologies for models based on patients that are still living and available for direct field measurements [50], the present study has to employ different human subjects with the exception of the simulation-to-simulation validation in the first application example.
Fig 1. a)–Full-body Visible Human Project Female v5.0 CAD based computational phantom. The phantom is composed of 249 individual structures. Some individual muscles are removed for clarity. b-e) Examples of coregistration maps for surface meshes in different transverse planes with tissue labeling. In case b), non-anatomical separation between scalp and skull was corrected. In case d), the tissue labeling list is not shown. The complete full-body coregistration maps with the vertical resolution of 1 mm and with tissue labeling in every cross-sectional plane are available online in ‘.mp4’ format [24] for independent inspection and verification purposes.

https://doi.org/10.1371/journal.pone.0260922.g001
Materials and methods
Validation of model segmentation

The co-registration method has been used, which implies direct superposition of transverse cross-sections of all surface tissue meshes onto the original cryosection images. An in-house MATLAB module (Fig 1B–1E) was written that performs such superposition with the resolution of 1 mm and simultaneously labels all tissue meshes which are present for a given cross-section. Its output is a scanning sequence in ‘.mp4 format [24].

Further visual assessment was performed by a number of anatomical experts in their respective areas with the participation of Profs A. Nazarian (orthopaedics), A. R. Opotowsky, (cardiovascular systems), V. Poylin, (gastroenterology), E. K. Rodriguez (musculoskeletal tissue components), A. Pascual-Leone (cranial and intracranial anatomy) from Beth Israel Deaconess Med. Center, and Massachusetts General Hospital, Boston MA, and Prof. G. Haleblian (urology) from Saint Vincent Hospital, Worcester MA. The results will be summarized in the results section.

Numerical simulation method used in application examples

In each reported case presented herein, Ansys Electronics Desktop (HFSS) 2019 R1 finite-element solver with adaptive mesh refinement has been employed to solve Maxwell’s equations in three-dimensional space. In this way, the model is considered as an arbitrary (inhomogeneous) isotropic medium with piecewise-constant electric permittivity \( \varepsilon \) having the units of F/m and with constant magnetic permeability \( \mu \) having the units of H/m. After Maxwell’s equations are solved for an electric field (or the electric field intensity) \( E(r, t) \) [V/m] and for a magnetic field (or the magnetic field intensity) \( H(r, t) \) [A/m], volumetric electric current is directly related to the electric field by a local form of Ohm’s law,

\[
J(r, t) = \sigma(r)E(r, t)
\]  

where \( \sigma(r) \) is (generally piece-wise constant) medium conductivity with the units of S/m, \( r \) is the position vector (or location in 3D space) and \( t \) is time.

In validation examples 2 and 4, the solutions of these simulations have been coupled to Ansys Mechanical 2019 R1 via Ansys Workbench. Pennes’ bioheat equation, based on the heat diffusion equation, is a standard approximation for heat transfer in biological tissues [51–53] partially implemented in the Ansys Mechanical software. It has the form

\[
\rho(r)C(r)\frac{\partial T(r, t)}{\partial t} = \nabla \cdot (k(r)\nabla T(r, t)) = \rho(r)Q(r) + \rho(r)\text{SAR}(r) - \rho_sC_sB(r)(T(r, t) - T_b) = 0 \tag{2}
\]

where \( T(r, t) \) is the local temperature, \( \rho(r) \) is density, \( C(r) \) is specific heat capacity (at constant pressure), \( Q(r) \) is the metabolic heat generation rate, \( B(r) \) is the perfusion rate [1/s]; index \( b \) is related to blood.

All electromagnetic, mechanical, and thermal tissue properties used in the present study are catalogued in the IT’IS database [54]. The electromagnetic material properties are given as a function of frequency, which is suitable for the present analysis.

The local SAR (W/kg) is defined through averaging the dissipated power per unit mass over a small (ideally infinitesimally small) volume \( V \), that is

\[
\text{SAR}(r) = \frac{1}{V} \int_V \frac{\sigma(r)}{2\rho(r)} |E(r)|^2 dV \tag{3A}
\]

Here, \( |E(r)| \) is the electric field magnitude at the observation point. The body-averaged or the
whole-body SAR is given by averaging over the entire body volume, as
\[
\text{SAR}_{\text{body}} = \frac{1}{V_{\text{body}}} \int_{V_{\text{body}}} \frac{\sigma(r)}{2\rho(r)} |E(r)|^2 dV \tag{3B}
\]
Similarly, SAR_{1g} is given by averaging over a cubic volume with a mass of 1 g
\[
\text{SAR}_{1g}(r) = \frac{1}{V_{1g}} \int_{V_{1g}} \frac{\sigma(r)}{2\rho(r)} |E(r)|^2 dV \tag{3C}
\]
\(\text{SAR}_{10g}(r)\) is found in a similar fashion.

To solve Eq (3), the electric field is required, which is difficult to measure directly during an MRI procedure. However, it is possible to obtain the \(B_{1+}\) field, which can be used to predict and estimate the electric field. According to [55], various \(B_{1+}\)-mapping methods have been proposed to measure the magnitude of \(B_{1+}\) components (especially for the transmit \(B_{1+}\) component), such as

1. using multiple acquisitions with different flip angles of spins [56,57];
2. applying identical RF pulses followed by two delays of different repetition times (TRs) [58];
3. utilizing phase-sensitive means based on composite RF pulses [59] or on the Bloch–Siegert phase shift [60],
4. employing stimulated echoes acquisition mode (STEAM) in multipulse sequences [61,62] or in a single sequence followed by a tailored gradient echo train [63].

On the other hand, there is no direct measurement to quantitatively obtain the absolute phase distribution of the transmit or receive \(B_{1+}\) field [55].

Local SAR values given by Eq (3) can now be derived from the \(B_{1+}\) field mapping [64]. In summary, we express the electric field form Ampere’s law in frequency domain,
\[
E(r) = \frac{\nabla \times H_{1+}(r)}{j\omega k(r)}, \quad k(r) = \varepsilon(r) - j\sigma(r)/\omega \tag{4}
\]
Once \(E(r)\) is known from Eq (4), the local SAR in Eq (3),
\[
\text{SAR}(r) = \frac{\sigma(r)}{2\rho(r)} |E(r)|^2 \tag{5}
\]
can be computed. Variations of this method are available [55].

Modeling setup for validation Example 1: Normalized SAR predicted by two different modeling techniques in 1.5 T birdcage whole body MRI coil
In the first example, a generic, whole-body high-pass birdcage coil with 16 rungs and 32 matching capacitors, loaded with the VHP Female model, is considered. The coil has a diameter of 64 cm and length of 69 cm consistent with [65]. The simulation geometry is shown in Fig 2A. Simulations have been conducted at shoulder/heart and abdominal landmarks. For the former, the coil center is oriented to coincide with the top of the T7 vertebra; the latter has the coil center located at the top of the L1 vertebra. The coil was tuned to the desired frequency of 64 MHz \((B_0 = 1.5 \text{ T})\) when loaded with the subject at each landmark as described in S1 Appendix (cf. [72]). Similar to [72], an ideal excitation was applied with 32 sources placed in the two
end-rings to perform the function of the capacitors. This excitation provides results which are very similar to the conventional two-port or four-port excitations [72].

A comparison was further made with the results of Ref. [72] which was using a nearly identical high-pass birdcage coil (diameter of 63 cm and length of 70 cm), the nearly identical heart landmark, and the identical coil excitation type. However, an in-house voxel model for the same VHP-Female dataset was employed in [72] followed by the FDTD simulation method with the resolution of 5 mm.

To compare the solution variation as a function of FEM mesh density and the solution convergence trends, solutions were generated first with 1 and then with 8 adaptive mesh refinement passes, created approximately 0.5 M and 2.0 M tetrahedra, respectively. As an example, Fig 2 shows the corresponding local SAR distributions at two different FEM resolutions in the coronal plane for the coil loaded with the VHP-Female v3.0 computational human model given $B_1^+$ amplitude of 1 $\mu$T at the coil center. Fields solutions from Ref. [72] were also normalized given the desired magnitude of $B_1^+$ at the coil center of 1 $\mu$T. The normalization is done in the form

$$\text{SAR} \rightarrow \frac{\text{SAR}}{(B_1^+/1\mu T)^2}$$ (6)

The local SAR was computed in Ansys HFSS and then exported to MATLAB over a uniform 3D grid of 2 mm in size. Whole-body SAR$_{body}$ was computed from this data directly in MATLAB. SAR$_{1g}$ was also calculated by finding a volume $V_{1g}$ surrounding the observation point having the mass of exactly 1 g, and then performing averaging according to Eq (3C). This averaging volume contains approximately $5\times5\times5$ individual voxels ($2\times2\times2$ mm each) closest to the observation point. The observation points form a 3D sub-grid spaced of 20 mm and 10 mm, respectively. SAR over any 1g was approximated as the SAR over any 1cm$^3$. SAR$_{10g}$ was computed in the same way. In this case, the averaging volume contained approximately 1250 individual voxels ($2\times2\times2$ mm each) closest to the observation point.
Modeling setup for comparison Example 2: Metal nail implant heating in 1.5 T birdcage whole body MRI coil

ASTM phantoms have been used as the standard method for testing compliance of implants within MRI environment [73]. The phantom is not an equivalent of intricate human body structures. Therefore, we consider this example, which employs the experimental phantom data, as a comparison, not as the validation.

Prior to conducting simulations using the VHP Female v3.0 model, a necessary workflow and RF power calibration have been established. Fig 3 depicts to scale simulation of the original experiment performed in [44–46], which was used for calibration purposes. At left in Fig 3, a homogenous experimental AGAR gel phantom is positioned within a 1.5 T birdcage MRI coil operating at 64 MHz. All examinations [44–46] were performed with a 1.5T MR scanner (MAGNETOM Symphony, SIEMENS) with the phantom at the center of the coil. A birdcage shaped transmit/receive body coil was used there with the inner diameter of 60 cm and the length of 70 cm; these dimensions have been reproduced in the modeling setup. Coil matching and excitation was performed as described in the previous example. We again use ideal excitation which is very similar [72] to the two-port excitation used in [44–46].

A 24 cm long metallic orthopaedic nail implant of Zimmer, Inc. made of stainless steel (originally very slightly bent but modeled a straight rod of the same diameter) has been

![Fig 3. Simulations to establish calibration for experiments conducted in [43–45]. a)–The metallic nail implant placed within a homogeneous loop-like Agar phantom [43–45] at a depth of 2 cm. b)–The current density produced with a whole-phantom SAR of 4.0 W/kg. c)–The simulated temperature given a total volume power loss of 120 W exactly corresponding to experiment [43–45]. Simulation results produced a temperature rise of 10.02°C, slightly less than the 12.6°C experimentally observed in [43–45].](https://doi.org/10.1371/journal.pone.0260922.g003)
embedded in this phantom such that it is 2 cm away from the top and side edges of the phantom. Both the phantom and metallic rod have been assigned dimensions and material properties consistent with experiment [44–46].

**Fig 4** at left shows the VHP Female computational human model in a 1.5T birdcage MRI coil. The computational model has been oriented so that the center of the femur bone is aligned with the center of the coil. The left quadriceps muscle within the VHP model is not shown in Fig 4 so that the position of the femur can clearly be seen. Within the left femur, the same 24 cm long cylindrical metallic implant has been inserted. This rod is assigned material properties consistent with [44–46]. The coil is driven at 64 MHz and at a power such that the volumetric power loss within the human model is again 120 W, also consistent with the published experimental results.

A length-based mesh constraint of no edge larger than 2.5 mm was enforced for the metallic rod mesh and a total of about 636,000 tetrahedral elements were used in the Ansys HFSS simulation.

Once the electromagnetic simulation was complete, the results were passed to Ansys Mechanical software by linking the two simulations in Ansys Workbench. A mesh refinement was again employed on the faces of the rod to ensure that a dense enough mesh was created to capture the local temperature changes. Approximately 629,000 tetrahedral elements were used in the transient thermal simulation. Volumetric losses (SAR) produced by the Ansys HFSS simulation were imported into Ansys Mechanical and used as the internal heat generation source density. The RF coil and the heat sources were active for 900 s and the model was allowed to cool for another 600 s. The values for the implant temperature observed from 0 to 900 s are shown in **Fig 5**; they will be discussed in the Results section below.
Modeling setup for validation example 3 $B_1^+$ maps for 3 T birdcage MRI whole body coil

The experiment [47,48] uses an 8-channel volume coil, which means each rung is independently driven. This is needed for body imaging at 3 T as the wavelength in the body is fairly short, resulting in an inhomogeneous $B_1^+$ field. This is not a birdcage design. It is a TEM coil (where each rung terminates to the shield) with a decoupling ring (not the same as a birdcage end ring). A co-simulation with a circuit solver is therefore needed to carefully tune, match and decouple all the channels (by adjusting capacitors) and then drive each channel with its own power source with the desired amplitude/phase differences.

Fortunately, in the experimental study [47], the parallel transmit body coil was driven in the so-called quadrature mode with equal current amplitude in all TX elements at 45 degrees phase differences between neighboring elements. This mimics the behavior of a birdcage resonator [47] and therefore makes it possible to use the present birdcage coil model with the nearly identical diameter of both coils (64 cm vs 63 cm [48]) and the same diameter of both RF shields, but with a somewhat longer length of our birdcage (69 cm vs ~42 cm [48]).

![Fig 5. Comparison of the simulated numerical values (red stars) obtained using the VHP Female computational phantom with published experimental data (two black curves) [43–45] for the maximum temperature rise near the implant. The depth of the implant within the VHP model is approximately 4 cm and the maximum temperature value after 900 seconds of coil operation time is represented by the top red star. The VHP model predicts a slightly higher (by 2.75˚) maximum temperature rise than what was measured in [43–45]–see Table 2. This is likely due to the different (non-homogeneous) material properties employed in the present study and the slightly angled orientation of the nail.](https://doi.org/10.1371/journal.pone.0260922.g005)
From experimental data for two male subjects [47], we choose healthy volunteer #2 since he has body mass of 68 kg [48] being closer to the VHP-Female human model which is important for calibration.

The VHP Female computational human model shown in Fig 6A is then positioned in the 3 T birdcage MRI coil at the abdominal landmark to match the physical setup in [47,48]. Adaptive mesh refinement has again been employed to produce a tetrahedral mesh with about 1.7 M tetrahedra throughout the volume. Simulations have been conducted at 128 MHz ($B_0 = 3$ T). Coil matching and excitation was performed as described in the previous examples.

We compare the $B_1^+$ distribution within the body with that obtained for healthy volunteer #2 (male, age 29 to 43 years) [47]. The power provided to the coil has been manually calibrated to produce a whole-body SAR of 0.51 W/kg within the VHP Female human model to exactly comply with the experimental data [47,48].

Modeling setup for validation example 4: Tissue heating due to single-loop coil close to skin surface at 165 MHz

The final validation case models experimental setup [49]; it is shown in the top portion of Fig 7. The forearm of the VHP Female computational human model has been isolated to speed up the simulation. Internal structures, including the humerus, ulna and radius bones, extensor, flexor, triceps and biceps muscles, radial nerve, and various arteries and veins, are encapsulated within concentric layers of muscle, fat and skin tissue. The single loop antenna [49] is modeled as a 80 mm diameter copper torus with a minor diameter of 2 mm driven at 165 MHz by a 50 Ohm lumped antenna port and a thin sheet of Teflon separates the antenna from the forearm. These settings exactly correspond to experiment. All thermal and electromagnetic material properties associated with the internal body structures correspond to those supplied in [49].

In experiment [49], the forearm size and origin were not specified. However, it seen in Fig 3 of Ref. [49] that it is a male arm fully covering the coil ring. The arm of the VHP-Female model in Fig 7A and 7B is also covering the entire ring. In both models, the coil ring is underneath the arm (cf. Fig 3 of Ref. [49] and Fig 7A and 7B of the present study). However, the arm of the VHP-Female model is more round in shape since it was not supported by anything during the cryogenic freezing.

According to [49], the antenna is provided sufficient power to dissipate approximately 31 W within the forearm and produce peak local SAR values in the lower corners of the forearm of approximately 450 W/kg. These power conditions have been replicated in simulations with the forearm of the VHP Female model. Adaptive mesh refinement produced a volumetric mesh consisting of approximately 0.1 M tetrahedra.

Following simulation in Ansys HFSS, the geometry and resulting fields were passed via Ansys Workbench to Ansys Mechanical. Here, the human model was again re-meshed, producing just over 0.06 M elements.

The volumetric power losses (SAR) determined in Ansys HFSS were imported and evaluated as the heat generation source density. Heat generation sources were active for 120 s and the model then cooled via convection for 19.2 s; temperatures were recorded throughout the duration of the simulation. The corresponding temperature rise after 139 seconds in total was computed and compared with experiment.

Results

Segmentation validation via co-registration and anatomical expertise

After resolving multiple mesh intersections, the validation of the model segmentation shown in Fig 1A was performed by co-registration of surface meshes superimposed onto the original
cryosection images (Fig 1B–1E) and correcting obvious flaws. Further visual anatomical assessment was performed by the number of experts from Beth Israel Deaconess Med. Ctr. and Massachusetts General Hospital, Boston MA. The questionable surface meshes (~12 in total, mostly bones but also soft tissues including scalp, bladder and uterus) were corrected. An example of a non-anatomical flaw is shown in Fig 1B where the scalp was non-anatomically separated from the skull during the cryogenic process, which required proper mesh adjustment. The surface deviation for all meshes was found not to exceed 2–7 mm.

Fig 6. The VHP-Female computational phantom positioned with a 3T MRI birdcage coil at the abdominal landmark. a)–The model geometry is shown, providing a view of the phantom located in the coil. b)–The simulated $B_1^+$ map generated in the axial plane about the model mid-section. c)–The experimental $B_1^+$ map reported in [46,47]. d)–The simulated $B_1^+$ map generated in the coronal plane about the VHP Female model mid-section. e)–The measured $B_1^+$ map generated in a coronal plane at the approximate middle of the model. These values are also reviewed in Table 2. https://doi.org/10.1371/journal.pone.0260922.g006
The complete full-body co-registration maps with the vertical resolution of 1 mm and with tissue labeling in every cross-sectional plane are available online in .mp4 format [24] for independent inspection and verification purposes.

Example 1: Normalized SAR predicted by two different modeling techniques in 1.5 T birdcage whole body MRI coil

Fig 2B and 2C show the local SAR within the various tissues of the VHP-Female v3.0 model after 1 pass and 8 passes, respectively, for the shoulder/heart landmark. All results are normalized to the $B_1^+$ amplitude of 1 $\mu$T at the coil center. By analyzing Fig 2 we can see that
The accurate solution with eight adaptive mesh refinement passes generates a more realistic SAR distribution, especially with regard to the local SAR—see Table 1. In particular, two non-physical maxima of the SAR observed at the top of the head are no longer present.

The accurate solution with eight adaptive mesh refinement passes and the coarse solution with one adaptive pass generate approximately the same whole-body SAR and SAR distribution maps, but considerably different peak local SAR values—see Table 1. Note that the coil has been retuned separately in both cases.

The maximum SAR for the present landmark is observed in the upper shoulder/neck area and in the arms area.

Table 1 compares the computed SAR values with the values obtained in Ref. [72] under nearly identical conditions. For both models and both methods, the whole body SAR and the maximum SAR_{max}(r) differ by 8% and 10%, respectively.

A comparison with a variety of other modeling reference sources [66–76] has also been made both at 1.5 T and 3 T. It was found that the SAR results calculated when using the VHP Female v3.0 model are strictly within the bounds of all other reported values.

Comparison Example 2: Metal nail implant heating in 1.5 T birdcage whole body MRI coil

The computed volumetric current density within the phantom is shown at top right in Fig 3. This density matches well with the published result; it was produced by adjusting coil power to exactly replicate the RF exposure given in [44–46]—a whole-phantom SAR of 4.0 W/kg and a volumetric power loss density of 120 W in the phantom. The temperature simulation is shown at bottom right of Fig 3. Given the above power loss density as an internal heat generation source, the total temperature rise within the human model was 10.02˚C. This value is slightly less than the 12.6˚C observed in [44–46]. This small disparity may be due to differences in material properties, given that the experiments conducted in [44–46] did not explicitly provide permittivity and conductivity values, but did state that values were ‘equivalent to that of muscular tissue,’ which were replicated for the present work. However, it is close enough to give confidence in the adequate simulation setup, enabling the extension of this methodology to the case involving the VHP female human model.
Peak simulated temperature values for the implant within the human model are shown in the bottom right of Fig 4. A maximum temperature rise of approximately 11˚C is observed at the very ends of the metallic implant.

Fig 5 shows temperature dynamics: it compares simulated numerical values (red stars) obtained using the VHP Female computational phantom with published experimental data (two black curves) [44–46] for the maximum temperature rise near the implant. The depth of the implant within the VHP model is approximately 3–4 cm and the maximum temperature value after 900 seconds of coil operation time is represented by the top red star. The VHP model predicts a slightly higher (by 2.75˚) maximum temperature rise than what was measured in [44–46]–see Table 2. This is likely due to the different (non-homogeneous) material properties employed in the present study and the slightly angled orientation of the nail. In the VHP Female model, the rod is aligned with the femur and represents a more realistic position that would be encountered in a clinical setting.

Though not shown below, the electromagnetic simulation produced surface currents along the metallic implant in excess of 14 A/m current amplitude. As anticipated, the simulated SAR and temperature had high values along the end tips of the implant.

**Example 3: $B_1^+$ maps for 3T birdcage MRI whole body coil**

Experimental [47,48] $B_1^+$ field maps for the problem geometry of Fig 6A are show in two planes in Fig 6B–6E. Fig 6B and 6C depict the $B_1^+$ map in an axial plane at the abdominal region of the VHP Female mode and the corresponding measured data obtained for healthy volunteer 2 [47]. The two remaining plots in Fig 6 depict the same $B_1^+$ field map in a coronal plane throughout the center of the model. The field profiles depicted in Fig 6 are topologically similar to the measured profiles provided in [47,48]. Emphasize that the peak field values are approaching exactly 2 $\mu$T in either case when whole-body SAR of 0.51 W/kg is enforced. In both instances, hot spots are noted around the inner arm edges and the intensity of the field is shown to dissipate as it moves into the center of the model/patient. Peak values in the central region are slightly under 1 $\mu$T for both measured and simulated results.

Variation between the experimentally captured data and the results using the VHP Female v3.0 model are due to the fact that the body composition, arm positions, and the coil type are not entirely equivalent. However, the general pattern and the absolute peak values correspond very well between the experimental and simulated sources and demonstrate that the VHP Female v3.0 human model will generate representative field distributions under these conditions.

### Table 2. Comparison of simulated numerical results using the VHP Female computational model with experimental results for examples 2, 3, and 4.

| Validation/Comparison Example | Reference (s) | Quantity | Simulation results of this study | Published experimental data |
|-----------------------------|-------------|---------|---------------------------------|-----------------------------|
| 2 Full body, orthopaedic nail implant | [43–45] | Maximum Temperature Increase after 900 Seconds | −11˚C Observed at implant ends (3–4 cm depth) in VHP-Female model–see Fig 5. | −8.25˚C measured for Gel Phantom (4 cm depth) for the same implant–see Fig 3 |
| 3 Full body | [46,47] | B1+ Field Maps in Body Planes | Between 0–2 $\mu$T depending on body location–see Fig 3. Agreement with experimental field topology | Between 0–2 $\mu$T depending on body location–see Fig 4. |
| 4 Upper Extremity | [50] | Maximum Temperature Increase after 139 seconds | 6.0˚C | 6.6˚C measured 8˚C simulated–see Fig 5 |

https://doi.org/10.1371/journal.pone.0260922.t002
SAR values for this case have also been simulated and recorded. Though not shown here, they also match the data in [47,48] with the peak values approaching 4 W/kg in ether case, and with higher values at the outer portions of the arms.

Example 4: Tissue heating due to single-loop coil close to skin surface at 165 MHz

Fig 7A and 7B replicate the experimental problem geometry [49]. The simulated SAR generated in a plane through the VHP Female forearm directly above the center of the coil is shown in Fig 7C and 7D as compared with the experiment [49]. The peak SAR value experienced in the outermost model layer is approximately 450 W/kg, consistent with the value reported in [49] to within 5%. There is a lower value of SAR within the second layer (fat tissue) and it again rises in the muscle tissue. As expected, due to their material properties, higher SAR values are displayed in the nerve and cardiovascular tissues.

The simulated and measured [49] thermal maps within the forearm in a plane directly above the antenna center are given in Fig 7E and 7F after 139 seconds of coil operation. The peak simulated temperature change is about 8˚C, which is slightly higher than the measured 6˚C change reported in [49]. It should be noted that the peak simulated value of 8 deg reported here is in almost a perfect agreement with the 8.8˚C simulated temperature change reported in [49].

The spatial temperature map is also consistent with the measured map despite the different wrist geometry and tissue composition. There is some difference in the observed depth of temperature change. This is likely mostly due to the fat layer of the VHP Female model which is thicker than the subject reported in [49].

Discussion

Initial model validation by co-registration

Since the original VHP-Female image dataset with the isotropic resolution of 0.33 mm is freely and publicly available, the co-registration validation performed in this study is also made public [24]. It is pertinent to VHP-Female model v5.0, which has a larger number of individual muscles than the model v3.0.

Review of subjects, settings, and results used in validation and comparison examples

During a validation study, simulated or measured data should be compared to an independent modeling or measuring technique. Ideally, when performing such a validation study, the setup of the validation should closely resemble the simulation or measurement that is being validated. As an example, in our study it would be ideal to have the same human body model in both the simulation/measurement and the validation setup. However, since the human body model that we want to validate in our study is deceased, this is not a possibility. Therefore, comparison with the results for different human body models and/or subjects is used in this study. When possible, the attempt is made to use data obtained from human body models that are comparable to the VHP-Female model.

Validation example #1 (whole body SAR and the maximum SAR$_{1g}(r)$ evaluation) is a purely modeling study at 64 MHz. The same original dataset from the US National Library of Medicine was used for human model construction in both the cases, along with the nearly identical coil dimensions and settings. As a result, an excellent agreement in whole body SAR and in the
maximum $SAR_r(r)$ was obtained in Table 1 despite the different model type (voxel vs CAD) and the different solver (FDTD vs FEM).

Comparison example #2 (implant heating in the ASTM phantom at 64 MHz) also used identical coil construction, dimensions, and settings, but the implant temperature was compared with that simulated within the femur of the VHP-Female model. Despite the obvious field differences and a slightly different implant position caused by human anatomy, the implant temperature dynamics (Figs 4 and 5 and Table 2) are rather similar in both cases, for this particular anatomical location and the particular patient landmark. These are particularly relevant results given that assessment of RF safety for medical implants generally consists of obtaining measurements from inside a saline gel phantom and conducting simulations using computational human body models. It is therefore critical that these body models are anatomically correct and have been validated for this purpose. Ideally, a suite of human body models, characterizing a range of age, sex, BMI, and other pertinent characteristics, is employed given that a single model may not provide enough evidence to guarantee the safety of a medical implant. Subsequent models could follow a similar validation procedure as presented here.

Validation example #3 ($B_1^+$ map at 128 MHz in vivo) employs a younger male subject with the closest weight and a coil of a different length (but with the same diameter and mimicking the same birdcage resonator), which results in a good agreement, both qualitative and quantitative, in the $B_1^+$ maps in the transverse plane (Fig 6B and 6C). Some disagreement in the coronal plane (Fig 6D and 6E) could likely be due to different arm positions (beside body in [47]).

Validation example #4 (in vivo heating of a human arm at 165 MHz by a single loop antenna) employs the identical antenna dimensions, settings and position as well as nearly the same arm width with respect to the antenna size. However, the arm cross-sections are quite different (cf. Fig 7) since the arm of the VHP-Female model is not supported while the experiment assumes some arm support. Despite this fact, a decent agreement in the measured $SAR$ and especially in temperature rise is obtained (cf. Fig 7).

Other comparison results
A comparison with a variety of other modeling reference sources [66–76] has also been performed both at 1.5 T and 3 T. It was found that the SAR results calculated when using the VHP Female v3.0–5.0 model are strictly within the bounds of all other reported values.

Conclusions
This study provided a reasonably systematic validation and comparison of the VHP-Female CAD v.3.0–5.0 surface-based computational human model starting with the segmentation validation and following four different application examples. The target context of use is a non-clinical assessment model for female patients of age 50–70 years with a BMI of 30–36 during 1.5 and 3.0 T based MRI procedures.

It was found that given the same or similar coil settings, the computational human model generates meaningful results for SAR, $B_1^+$ field, and temperature rise when used in conjunction with the 1.5 T birdcage MRI coils or at higher frequencies corresponding to 3 T MRI.

Notably, the temperature rise deviation from experiment did not exceed 2.75°C for three different heating scenarios considered in the study. Two of them included the long nail orthopaedic implant in the phantom and in the body, respectively. The relative temperature rise difference was 10% and 25%, respectively. The higher error for the implant in the body is to be expected. In the last example for in vivo heating of the human arm, the relative temperature rise difference was 20%.
Therefore, the tested human model could likely be applied to the safety evaluation of implantable medical devices within this temperature error margin.

The approach taken in this study might also be used as an example for validation of similarly developed computational human models from no longer living subjects.

Supporting information
S1 Appendix. Tuning procedure for the MRI coil used in this study.

(DOCX)

Acknowledgments
The authors thank Profs A. Nazarian, A. R. Opotowsky, V. Poylin, E. K. Rodriguez, A. Pascual-Leone from Beth Israel Deaconess Med. Center, and Massachusetts General Hospital, Boston MA, and Prof. G. Haleblian (urology) from Saint Vincent Hospital, Worcester MA for their feedback on multiple anatomical model details. Directions of Prof. N. Weiskopf, Drs. S. Geyer, and M. Kozlov (Max Planck Institute for Human Cognitive and Brain Sciences, Leipzig, Germany) are acknowledged.

The authors wish to express their sincerest thanks to Dr. G. Bogdanov of the Electrical and Computer Engineering Department at Worcester Polytechnic Institute for his critical advice on matching and exciting the MRI coils used in this study.

Last but not least, the authors would like to thank Drs. M. I. Iacono and L. Angelone of the US Food and Drug Administration for their constructive guidance and assistance in selecting the experimental cases against which the numerical results produced during this effort were compared.

Author Contributions
Conceptualization: Gregory M. Noetscher, Peter Serano, William A. Wartman, Kyoko Fujimoto.

Formal analysis: Gregory M. Noetscher.

Investigation: Gregory M. Noetscher, Peter Serano, William A. Wartman, Kyoko Fujimoto, Sergey N. Makarov.

Methodology: Gregory M. Noetscher, Peter Serano, William A. Wartman, Kyoko Fujimoto, Sergey N. Makarov.

Software: Gregory M. Noetscher, Peter Serano, William A. Wartman, Sergey N. Makarov.

Supervision: Kyoko Fujimoto.

Writing – original draft: Gregory M. Noetscher, Sergey N. Makarov.

Writing – review & editing: Gregory M. Noetscher, Peter Serano, William A. Wartman, Kyoko Fujimoto, Sergey N. Makarov.

References
1. Jian-Ming J. Electromagnetic Analysis and Design in Magnetic Resonance Imaging. 1st ed. Boca Raton: CRC Press; 1999. https://doi.org/10.1016/s0730-725x(99)00064-8 PMID: 10576717
2. Abadi E, Segars WP, Tsui BMW, Kinahan PE, Bottenus N, Frangi AF, et al. Virtual clinical trials in medical imaging: a review. J Med Imaging. 2020; 7: 042805. https://doi.org/10.1117/1.JMI.7.4.042805 PMID: 32318817
3. Hand JW, Li Y, Thomas EL, Rutherford MA, Hajnal JV. Prediction of specific absorption rate in mother and fetus associated with MRI examinations during pregnancy. Magn Reson Med. 2006; 55: 883–93. https://doi.org/10.1002/mrm.20824 PMID: 16508913

4. Kozlov M, Turner R. A comparison of Ansoft HFSS and CST Microwave Studio simulation software for multi-channel coil design and SAR estimation at 7 T MRI. Piers Online. 2010; 6: 395–399.

5. Serano P, Angelone LM, Katnani H, Eksandar E, Bonmassar G. A Novel Brain Stimulation Technology Provides Compatibility with MRI. Sci Rep. 2015; 5: 9805 https://doi.org/10.1038/srep09805 PMID: 25924189

6. Kabil J, Belguerras L, Trattnig S, Pasquier C, Feiblinger J, Missoufe A. A Review of Numerical Simulation and Analytical Modeling for Medical Devices Safety in MRI. Yearb Med Inform. 2016; 10: 152–158. https://doi.org/10.15265/IY-2016-016 PMID: 27830244

7. Giovannetti G, Tiberi G. Radiofrequency magnetic resonance coils and communication antennas: Simulation and design strategies. Magn Reson Imaging. 2017; 44: 1–7. https://doi.org/10.1016/j.mri.2017.07.021 PMID: 28757459

8. Kozlov M, Bazin PL, Möller HE, Weiskopf N. Influence of Cerebrospinal Fluid on Specific Absorption Rate Generated by 300 MHz MRI Transmit Array. Proceedings of the 10th European Conference on Antennas and Propagation; 2016 Apr 10–15; Davos, CH.

9. Stocker T, Vahedipour K, Pfugfelder D, Shah NJ. High Performance Computing MRI Simulations. Magn Reson Med. 2010; 64:186–93. https://doi.org/10.1002/mrm.22406 PMID: 20577987

10. U.S. Food and Drug Administration. Guidance for the submission of premarket notifications for magnetic resonance diagnostic devices. Rockville, MD: Center for Devices and Radiological Health, U.S. Food and Drug Administration; 1998.

11. European Committee for Electrotechnical Standardization. EN 60601–2–33:2002. Medical electrical equipment: Part 2–33: Particular requirements for the safety of magnetic resonance equipment for medical diagnosis. Brussels: CENELEC; 2002.

12. Medical Devices Agency. Guidelines for magnetic resonance equipment in clinical use with particular reference to safety. London: Medical Devices Agency; 2002.

13. U.S. Food and Drug Administration. Criteria for significant risk investigations of magnetic resonance diagnostic devices. Rockville, MD: Center for Devices and Radiological Health, U.S. Food and Drug Administration; 2003.

14. International Committee on Non-Ionizing Radiation Protection (ICNIRP). Medical MR procedures: protection of patients. Health Physics 2004; 87: 197–216. https://doi.org/10.1097/00004032-200408000-00008 PMID: 15257220

15. Makarov SN, Noetscher GM, Yanamadala J, Piazza MW, Louie S, Prokop A, et al., Virtual Human Models for Electromagnetic Studies and Their Applications. IEEE Reviews in Biomedical Engineering. 2017; 10:95–121. https://doi.org/10.1109/RRBME.2017.2722420 PMID: 28682265

16. Noetscher GM, Yanamadala J, Kozlov M, Louie S, Nazarian A, Makarov S. VHP-Female v3.0 FEM/BEM Computational Human Phantom. Proceedings of the 24th Int. Meshing Roundtable; 2015 Oct 12–14; Austin, USA.

17. Yanamadala J, Noetscher GM, Louie S, Prokop A, Kozlov M, Nazarian A, et al. Multi-Purpose VHP-Female Version 3.0 Cross-Platform Computational Human Model. Proceedings of the 10th European Conference on Antennas and Propagation; 2016 Apr 10–15; Davos, CH.

18. Tankaria H, Jackson XJ, Borwankar R, Srichandhru GNK, Tran AL, Yanamadala J, et al. VHP-Female Full-Body Human CAD Model for Cross-Platform FEM Simulations—Recent Development and Validations. Proceedings of the 38th Annual Int. Conf. of the IEEE Engineering in Medicine and Biology Society; 2016 Aug 16–20; Orlando, USA. https://doi.org/10.1109/EMBC.2016.7591173 PMID: 28268773

19. Noetscher G, Yanamadala J, Louie S, Prokop A, Nazarian A, Makarov S. Creating a Computational Human Model. IEEE Pulse. 2016; Available from: http://pulse.embs.org/march-2016/creating-a-computational-human-model/

20. Noetscher GM, Yanamadala J, Tankaria H, Louie S, Prokop A, Nazarian A, et al. Computational Human Model VHP-FEMALE Derived from Datasets of the National Library of Medicine. Proceedings of the 38th Annual Int. Conf. of the IEEE Engineering in Medicine and Biology Society; 2016 Aug 16–20; Orlando, USA.

21. Noetscher GM, Yanamadala J, Tankaria HV, Jackson XJ, Makarov SN, Louie S, et al. VHP-Female CAD Human Model Family for Antenna Modeling. Proceedings of the 2016 IEEE Int. Sym. on Antennas and Propagation/USNC-URSI National Radio Science Meeting. 2016 Jun; Puerto Rico.

22. The Visible Human Project, US National Library of Medicine. Online: https://www.nlm.nih.gov/research/visible/visible_human.html.
23. Ackerman MJ. The Visible Human Project: From Body to Bits. IEEE Pulse. 2017, vol. 8(4), pp. 39–41. https://doi.org/10.1109/MPUL.2017.2701221 PMID: 28715313

24. VHP Female v.3.0–5.0 Co-Registration Standard. 08/23/2017. Online: https://www.dropbox.com/sh/f1unwyang9e0h2v/AACczcq7EpR8wYx0wbnHC-eAe?dl=0.

25. Yanamadala J, Noetscher G, Louie S, Prokop A, Kozlov M, Nazarian A, et al. Multi-Purpose VHP-Female Version 3.0 Cross-Platform Computational Human Model. Proceedings of the 10th European Conference on Antennas and Propagation; 2016 Apr 10–15; Davos, CH.

26. Lemdiakov R, Venkatasubramanian A. Transmit coil design for Wireless Power Transfer for medical implants. Proceedings of the 39th Annual International Conference of the IEEE Engineering in Medicine and Biology Society; 201 Jul 11–15; Jeju, KR. https://doi.org/10.1109/EMBC.2017.8037282 PMID: 29060324

27. Venkatasubramanian A, Gifford B. Modeling and design of antennas for implantable telemetry applications. Proceedings of the 38th Annual Int. Conf. of the IEEE Engineering in Medicine and Biology Society; 2016 Aug 16–20; Orlando, USA. https://doi.org/10.1109/EMBC.2016.7592210 PMID: 28269728

28. Nikolayev D, Zhadobov M, Karban P, Sauleau R. Conformal antennas for miniature in-body devices: The quest to improve radiation performance, in URSI Radio Science Bulletin, vol. 2017, no. 363, pp. 52–64, Dec 2017. https://doi.org/10.23919/URSIRSB.2017.8409427

29. Nikolayev D. Modeling and Characterization of in-Body Antennas, 2018 IEEE 17th International Conference on Mathematical Methods in Electromagnetic Theory (MMET), Kiev, 2018, pp. 42–46. https://doi.org/10.1109/MMET.2018.8460279

30. Barbi M, Garcia-Pardo C, Cardona N, Nevarez A, Pons V, Frasson M. Impact of Receivers Location on the Accuracy of Capsule Endoscope Localization, 2018 IEEE 29th Annual International Symposium on Personal, Indoor and Mobile Radio Communications (PIMRC), Bologna, 2018, pp. 340–344. https://doi.org/10.1109/PIMRC.2018.8580862

31. Garcia-Pardo C, Andreu C, Forges-Leal A, Castello-Palacios S, Perez-Simbor S, Barbi M, et al. Ultrawideband Technology for Medical In-Body Sensor Networks: An Overview of the Human Body as a Propagation Medium, Phantoms, and Approaches for Propagation Analysis. IEEE Antennas and Propagation Magazine. 2018; 60: 19–33.

32. Nikolayev D, Zhadobov M, Le Coq L, Karban P, Sauleau R. Robust Ultraminiature Capsule Antenna for Ingestible and Implantable Applications. IEEE Transactions on Antennas and Propagation. 2017; 85: 6107–6119.

33. Perez-Simbor S, Andreu C, Garcia-Pardo C, Frasson M, Cardona N. UWB Path Loss Models for Ingestible Devices. IEEE Transactions on Antennas and Propagation. 2019; 67: 5025–5034.

34. Chen L, Eaton G, Noetscher G, Makarov S. Radiofrequency Propagation Close to the Human Ear and Accurate Ear Canal Models," Proceedings of the 40th Annual Int. Conf. of the IEEE Engineering in Medicine and Biology Society; 2018 Jul 17–21 Honolulu, USA.

35. Eloia JM, Noetscher GM, Makarov SN, Pascual-Leone A. Continuous wave simulations on the propagation of electromagnetic fields through the human head. IEEE Trans. Biomedical Engineering. 2014; 61:1676–1683. https://doi.org/10.1109/TBME.2013.2297781 PMID: 24845277

36. Noetscher GM, Yanamadala J, Makarov SN, Pascual-Leone A. Comparison of cephalic and extracephalic montages for Transcranial Direct Current Stimulation—A numerical study. IEEE Trans. Biomed. Eng. 2014; 61: 2488–2498. https://doi.org/10.1109/TBME.2014.2322774 PMID: 25014947

37. Makarov SN, Noetscher GM, Arum S, Rabiner R, Nazarian A. Concept of a Radiofrequency Device for Osteopenia/Osteoporosis Screening. Sci Rep. 2020; 10: 3540. https://doi.org/10.1038/s41598-020-60173-5 PMID: 32103042

38. Perez-Simbor S, Andreu C, Frasson M, Cardona N. UWB Path Loss Models for Ingestible Devices. IEEE Transactions on Antennas and Propagations. 2019; 67: 5025–5034.

39. Simbor SP. In-Body to On-Body Experimental UWB Channel Characterization for the Human Gastrointestinal Area. PhD Thesis, Universitat Politecnica de Valencia. 2019. Available from: https://dialnet.unirioja.es/servlet/tesis?codigo=267752.

40. Prokop A, Wittig T, Morey A. Using Anatomical Human Body Model for FEM SAR Simulation of a 3T MRI System. In: Makarov SN, Noetscher GM, Nummenmaa A. Brain and Human Body Modeling 2020. Springer Nature: New York; 2020. pp. 273–281.

41. Brown JE, Qiang R, Stadnik PJ, Stotts LJ, Von Arx JA. Calculation of MRI RF-Induced Voltages for Implanted Medical Devices Using Computational Human Models. In: Makarov SN, Horner M, Noetscher GM. Brain and Human Body Modeling. Springer Nature: New York; 2019. pp. 283–294.

42. Brown JE, Qiang R, Stadnik PJ, Stotts LJ, Von Arx JA. RF-Induced Unintended Stimulation for Implantable Medical Devices in MRI. In: Makarov SN, Noetscher GM, Nummenmaa A. Brain and Human Body Modeling 2020. Springer Nature: New York; 2020. pp. 283–292.
43. Neva Electromagnetics LLC, Static VHP-Female model v2.2. Available from: https://www.nevaelectromagnetics.com/vhp-female-2-2.

44. Muranaka H, Horiguchi T, Usui S, Ueda Y, Nakamura O, Ikeda F, et al. Evaluation of RF heating on humerus implant in phantoms during 1.5 T MRI imaging and comparisons with electromagnetic simulation. Magn Reson Med Sci. 2006; 5: 79–88. https://doi.org/10.2463/mrms.5.79 PMID: 17008764

45. Muranaka H, Horiguchi T, Usui S, Ueda Y, Nakamura O, Ikeda F. Dependence of RF heating on SAR and implant position in a 1.5T MR system. Magn Reson Med Sci. 2007; 6: 199–209. https://doi.org/10.2463/mrms.6.199 PMID: 18239357

46. Muranaka H, Horiguchi T, Ueda Y, Tanki N. Evaluation of RF heating due to various implants during MR procedures. Magn Reson Med Sci. 2011; 10: 11–19. https://doi.org/10.2463/mrms.10.11 PMID: 21441723

47. Homann H, Börmert P, Eggers H, Nehrke K, Dössel O, Graesslin I. Towards Individualized SAR Models and in vivo validation. Magn. Reson. Med. 2011; 66: 1767–1776. https://doi.org/10.1002/mrm.22948 PMID: 21630346

48. Homann H. SAR Prediction and SAR Management for Parallel Transmit MRI. 1st ed. Karlsruhe: KIT Scientific Publishing; 2012.

49. Oh S, Ryu YC, Carluccio G, Sica CT, Collins CM. Measurement of SAR-induced temperature increase in a phantom and in vivo with comparison to numerical simulation. Magn. Reson. Med. 2014; 71:1923–1931. https://doi.org/10.1002/mrm.24820 PMID: 23804188

50. Murbach M, Neufeld E, Capstick M, Kainz W, Brunner DO, Samaras T, et al. Thermal tissue damage model analyzed for different whole-body SAR and scan durations for standard MR body coils. Magn Reson Med. 2014; 71: 421–431. https://doi.org/10.1002/mrm.24671 PMID: 23413107

51. Lienhard A. Heat Transfer Textbook. 5th ed. New York: Dover Publications; 2019.

52. Wissler E, Penrose’s 1948 paper revisited. J. Appl. Phys. 1998; 85: 35–41. https://doi.org/10.1109/jappl.1998.85.1.35 PMID: 9655751

53. Arkin H. Recent developments in modeling heat transfer in blood perfused tissues. IEEE Trans. Biomed. Eng. 1994; 41: 97–107. https://doi.org/10.1109/10.294920 PMID: 8026856

54. Hasgall PA, Di Gennaro F, Baumgartner C, Neufeld E, Lloyd B, Gosselin MC, et al. IT’IS Database for thermal and electromagnetic parameters of biological tissues. Available from: www.itis.ethz.ch/database.

55. Xiao Tong Z, Jiaen L, He B. Magnetic-Resonance-Based Electrical Properties Tomography: A Review. IEEE Reviews in Biomedical Eng. 2014; 7: 87–95.

56. Insko EJ, Bolinger L. Mapping of the radiofrequency field, J. Magn. Reson. A. 1993; 103: 82–85.

57. Wang J, Qiu M, Yang QX, Smith MB, Constable RT. Measurement and correction of transmitter and receiver induced nonuniformities in vivo. Magn. Reson. Med. 2005; 53: 408–417. https://doi.org/10.1002/mrm.20354 PMID: 15678526

58. Yarnykh VL. Actual flip-angle imaging in the pulsed steady state: A method for rapid three-dimensional mapping of the transmitted radiofrequency field. Magn. Reson. Med. 2007; 57: 192–200. https://doi.org/10.1002/mrm.21120 PMID: 17191242

59. Morrell GR. A phase-sensitive method of flip angle mapping. Magn. Reson. Med. 2008; 60: 889–894. https://doi.org/10.1002/mrm.21729 PMID: 18816809

60. Sacolick LI, Wiesinger F, Hancu I, Vogel MW. B1 mapping by Bloch–Siegel shift. Magn. Reson. Med. 2010; 63: 1315–1322. https://doi.org/10.1002/mrm.22357 PMID: 20432032

61. Akoka S, Franconi F, Seguin F, Le Pape A. Radiofrequency map of an NMR coil by imaging. Magn. Reson. Imag. 1993; 11: 437–441. https://doi.org/10.1016/0730-725x(93)90078-r PMID: 8505878

62. Helms G, Finsterbusch J, Weiskopf N, Dechent P. Rapid radiofrequency field mapping in vivo using single-shot STEAM MRI. Magn. Reson. Med. 2008; 60: 739–743. https://doi.org/10.1002/mrm.21676 PMID: 18727090

63. Nehrke K, Bornert P. DREAM—A novel approach for robust, ultrafast, multislice B1 mapping. Magn. Reson. Med. 2012; 68: 1517–1526. https://doi.org/10.1002/mrm.24158 PMID: 22252850

64. Voigt T, Homann H, Katscher U, Dössel O. Patient-individual local SAR determination: In vivo measurements and numerical validation. Magn. Reson. Med. 2012; 68: 1117–1126. https://doi.org/10.1002/mrm.23322 PMID: 22213053

65. Cabot E, Christ A, Kuster N. Whole Body and Local SAR in Anatomical Phantoms Exposed to RF Fields from Birdcage Coils. Proceedings of the 29th General Assembly of the International Union of Radio Science; 2008 Aug 7–16. Chicago, USA.
66. Oh S, Webb AG, Neuberger T, Park B, Collins CM. Experimental and numerical assessment of MRI-induced temperature change and SAR distributions in phantoms and in vivo. Magn Reson Med. 2010; 63: 218–23. https://doi.org/10.1002/mrm.22174 PMID: 19785018

67. Bonmassar G, Serano P, Angelone LM. Specific absorption rate in a standard phantom containing a Deep Brain Stimulation lead at 3 Tesla MRI. 6th International IEEE/EMBS Conference on Neural Engineering (NER); 2013 San Diego, USA.

68. Collins CM, Li S, Smith MB. SAR and B1 Field Distribution in a Heterogeneous Human Head Model with a Birdcage Coil. Magnetic Resonance in Medicine. 2005; 40(6): 847–856.

69. Rennings A, Chen L, Otto S, Erni D. B1-Field Inhomogeneity Problem of MRI: Basic Investigation on a Head- Tissue-Simulating Cylinder Phantom Excited by a Birdcage-Mode. 42nd European Microwave Conference; 2012 Nov Amsterdam, NL.

70. Tomas BP, Li H, Anjum MR. Design and Simulation of a Birdcage Coil using CST Studio Suite for Application at 7T. IOP Conference Series: Materials Science and Engineering; 2013 Feb 1.

71. Yeo DTB, Wang Z, Loew W, Volgel MW, Hancu I. Local SAR in High Pass Birdcage and TEM Body Coils for Multiple Human Body Models in Clinical Landmark Positions at 3T. Journal of Magnetic Resonance Imaging. 2011; 33: 1209–1217. https://doi.org/10.1002/jmri.22544 PMID: 21509880

72. Liu W, Collins CM, Smith MB. Calculation of B1 Distribution, SAR, and SNR for a Body-size Birdcage Coil Loaded with Different Human Subjects at 64 and 128 MHz. Applied Magnetic Resonance. 2005; 29: 5–18. https://doi.org/10.1007/BF03166953 PMID: 23565039

73. Winter L, Seifert F, Zilberti L, Murbach M, Ittermann B. MRI-Related Heating of Implants and Devices: A Review. J Magn Reson Imaging. 2021; 53: 1646–1665. https://doi.org/10.1002/jmri.27194 PMID: 32458559

74. Cabot E, Christ A, Kuster N. Whole Body and Local SAR in Anatomical Phantoms Exposed to RF Fields from Birdcage Coils. Proceedings of the 29th General Assembly of the International Union of Radio Science; 2008 Aug 7–16. Chicago, USA.

75. Kozlov M, Turner R. RF Transmit Performance Comparison for Several MRI Head Arrays at 300 MHz. Progress in Electromagnetic Research Symposium Proceedings. 2013 Mar 28, Taipei: 1052–1056.

76. Kozlov M, Bazin PL, Möller HE, Weiskopf N. Influence of Cerebrospinal Fluid on Specific Absorption Rate Generated by 300 MHz MRI Transmit Array. Proceedings of the 10th European Conference on Antennas and Propagation; 2016 Apr 10–15; Davos, CH.