Development and Internal Validation of a Preoperative Prediction Model for Sentinel Lymph Node Status in Breast Cancer: Combining Radiomics Signature and Clinical Factors

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Purpose: To develop and internally validate a nomogram combining radiomics signature of primary tumor and fibroglandular tissue (FGT) based on pharmacokinetic dynamic contrast-enhanced magnetic resonance imaging (DCE-MRI) and clinical factors for preoperative prediction of sentinel lymph node (SLN) status in breast cancer patients.

Methods: This study retrospectively enrolled 186 breast cancer patients who underwent pretreatment pharmacokinetic DCE-MRI with positive (n = 93) and negative (n = 93) SLN. Logistic regression models and radiomics signatures of tumor and FGT were constructed after feature extraction and selection. The radiomics signatures were further combined with independent predictors of clinical factors for constructing a combined model. Prediction performance was assessed by receiver operating characteristic (ROC), calibration, and decision curve analysis. The areas under the ROC curve (AUCs) of models were corrected by 1,000-times bootstrapping method and compared by Delong’s test. The added value of each independent model or their combinations was also assessed by net reclassification improvement (NRI) and integrated discrimination improvement (IDI) indices. This report referred to the “Transparent Reporting of a multivariable prediction model for Individual Prognosis Or Diagnosis” (TRIPOD) statement.

Results: The AUCs of the tumor radiomic model (eight features) and the FGT radiomic model (three features) were 0.783 (95% confidence interval [CI], 0.717–0.849) and 0.680 (95% CI, 0.604–0.757), respectively. A higher AUC of 0.799 (95% CI, 0.737–0.862) was obtained by combining tumor and FGT radiomics signatures. By further combining tumor and FGT radiomics signatures with progesterone receptor (PR) status, a nomogram was developed and showed better discriminative ability for SLN status [AUC 0.839 (95% CI,
INTRODUCTION

Breast cancer is the most common cancer with heterogeneous features and leading cause of cancer death in females worldwide (1). Sentinel lymph node (SLN), as the first site for tumor spreading in breast cancer patient, plays an important role in treatment decision-making. There is no need to carry out axillary lymph node (ALN) dissection in breast cancer patient with negative SLN, and one to two positive SLNs for breast conserving surgery (2, 3). SLN status was determined by invasive SLN biopsy with potentially high false-negative rate (4), possible complications of lymphedema, axillary numbness, and brachial plexus injury (5), and inconsistency. Therefore, preoperative biomarkers for SLN status is indeed needed for avoiding additional unnecessary surgical procedures.

Magnetic resonance imaging (MRI) as a noninvasive modality with different sequences is usually used to visualize morphological, diffusion, and pharmacokinetic characteristics of tumor before surgery in breast cancer patients. MRI features mainly derived from dynamic contrast-enhanced (DCE) and diffusion-weighted imaging (DWI) allow for independently predicting lymph node status. The low apparent diffusion coefficient (ADC) value and rim enhancement of tumor in patients with breast cancer were associated with lymph node metastasis (6, 7). Moreover, heterogeneous and rim enhancement, and peritumoral–tumor ADC ratio were independent predictors for SLN metastasis in breast cancer (8).

In addition to conventional feature analysis, radiomics as a novel tool applied in medicine is able to extract high-throughput quantitative features from medical images obtained by noninvasive technique using mathematical algorithm (9–11), which has been used to predict malignancy (12), molecular subtypes (13), pathological complete response to neoadjuvant chemotherapy (14), and ALN or SLN metastasis in breast cancer (15, 16).

Based on MRI, previous studies demonstrated that radiomics signature of tumor with different sequences could predict ALN metastasis, and a combined model obtained a better performance (15). SLN status in breast cancer can also be predicted by radiomic features of primary tumor and peritumor based on different sequences including T2WI, DWI, and DCE (16, 17). However, most studies only focused on the tumor and peritumor, which ignored the fibrogranular tissue (FGT). Previous studies have demonstrated that the amount of FGT was associated with breast cancer risk (18), and the rate of FGT enhancement may predict response to neoadjuvant chemotherapy in breast cancer patients (19). The usefulness of FGT radiomic features and combined features from tumor and FGT in predicting SLN status for breast cancer remains unclear. Furthermore, for prediction of SLN status, it is valuable to evaluate whether the simultaneous addition of FGT radiomic and clinical factors into tumor radiomic features could improve the prediction performance (16).

Thus, this study aimed to develop noninvasive radiomics signatures from primary tumor and FGT based on pharmacokinetic DCE-MRI to predict SLN status in patients with breast cancer. The radiomic–clinical nomogram was also built and evaluated by integrating the radiomics signatures and independent clinical predictors. The study workflow is shown in Figure 1. The manuscript was prepared according to the TRIPOD checklist (20).

METHODS

Source of Data and Participants

The single-center data from our hospital was collected and analyzed in the current study. Under approval from our Institutional Review Board (IRB), we retrospectively reviewed female patients with breast cancer proven by histopathology between August 2015 and December 2018 and identified 497 patients who underwent dedicated breast MRI. The inclusion criteria were as follows: (1) patients underwent preoperative DCE-MRI; (2) patients were pathologically confirmed unilateral breast cancer; (3) patients received SLN biopsy/ALN dissection; and (4) patients did not receive neoadjuvant chemotherapy before preoperative DCE-MRI. The exclusion criteria were as follows: (1) the volume of interest (VOI) was small (longest diameter ≤ 5 mm); (2) patients with two or more lesions; (3) patients with nonmass-like breast cancer; and (4) MRI images with poor quality (non-diagnostic or non-analytic for tumor and FGT because of artifacts). Finally, a total of 186 patients were enrolled. Among the enrolled patients, 93 positive SLN and 93 negative SLN were balanced and matched based on “Age” of patients (at a ratio of 1:1) to help improve the robustness of the selected radiomics features and model performance (21, 22). The requirement of informed consent
for this study was waived by our IRB. All patient data were anonymized and de-identified before analysis.

### Outcome
The primary outcome was the SLN status. Blue dye tracer was used to identify SLN during operation. After the induction of general anesthesia, 1 ml of 1% methylene blue (Jiangsu Jichuan Pharmaceutical Co. Ltd., China) was injected subcutaneously around the areolar periphery at 12, 3, 6, and 9 o’clock positions before breast surgery. A SLN was defined as the first blue-stained node, which was along the blue-stained lymphatic channel. All the blue dye-stained SLNs were removed by following the blue lymphatic channels. Then, all SLNs were sent for immediate frozen sectioning. The SLN was defined as positive when cancer cells were identified by hematoxylin & eosin (H&E) staining. According to the American Society of Clinical Oncology (ASCO), in T1–2 breast cancer patients with one to two metastatic SLN, SLNB alone was performed; otherwise, ALN dissection was performed when SLN was confirmed with metastasis (3, 23).

### Predictors
Our database consists of 8 clinical factors, 198 tumor radiomic features, and 198 FGT radiomic features.

Clinical data were collected from patient medical records including age, sex, menstrual status, estrogen receptor (ER) status, progesterone receptor (PR) status, human epidermal growth factor receptor-2 (HER2) status, and Ki-67 proliferation index. The expression levels of ER, PR, HER2, and Ki-67 were determined by immunohistochemistry (IHC) after surgery. The ER or PR status was considered positive when at least 1% of tumor cell showed nuclear staining by IHC. The HER2 status was confirmed as positive when the staining score was 3. Fluorescence in situ hybridization (FISH) was further performed when HER2 score was 2. Ki-67 was considered positive when the expression was more than 14%. Tumor size was manually measured as the maximal transverse diameter in the enhanced phase with best contrast of DCE-MRI.

### Sample Size and Missing Data
Referring to the rule to have at least 10 outcome events per variable (EPV) (20, 24), we ensured less than nine features retained for each radiomics model in order to avoid overfitting, with respect to 93 SLN outcome events in each balanced class in the current study.

There were no missing data among clinical factors. For radiomic features, if missing data existed in one feature, the...
missing value was replaced by the median value of such radiomic feature values of 186 patients.

**MRI Acquisition**
All the MRI sequences were performed on a 3.0-T scanner with a 16-channel breast coil (Skyra, Siemens Healthcare, Erlangen, Germany). T2WI, T1 mapping, and DCE were conducted with patients in a prone position as our previous study (13). T1 mapping and DCE images were collected for radiomic analysis. T1 mapping was performed with parameters: repetition time (TR) = 5.64 ms; echo time (TE) = 2.46/3.69 ms; field of view (FOV) = 360 × 360 mm; slice thickness = 2.5 mm; matrix = 269 × 384; flip angles = 2°/15°. Gadodiamide (Omniscan, 0.1 mmol/kg; GE Healthcare, Milwaukee, USA) was intravenously administered at a rate of 2.5 ml/s after T1 mapping, followed by 20 ml of saline flush with the same injection rate. Twenty-six consecutive DCE phases were acquired using the CAIPIRINHA-Dixon-TWIST-VIBE sequence with parameters: TR = 5.64 ms; TE = 2.46/3.69 ms; FOV = 360 × 360 mm; slice thickness = 2.5 mm; matrix = 269 × 384; flip angle = 10°.

**Radiomics Workflow**
The radiomics procedure included VOI segmentation, radiomic feature extraction, feature selection, and prediction model construction.

**VOI Segmentation and Feature Extraction**
T1 mapping and DCE-MRI data were imported to Omni-Kinetics (GE Healthcare, Milwaukee, USA). Reference Region mode with a reference region of interest in the contralateral pectoralis major was used to generate voxel-wise perfusion maps. Two radiologists (more than 8 years of experience) blinded to the pathological diagnosis participated in the interpretation of breast MRI. VOIs of tumor and ipsilateral FGT were manually outlined slice by slice in the enhanced phase with best contrast between lesion and FGT, respectively. In order to guarantee the reproducibility of the extracted radiomic features, 20 patients’ images were randomly chosen for intra- and inter-observer agreement analysis. The tumor and FGT VOIs were independently segmented in the subset of 20 patients twice by one radiologist at 1-month interval to assess the intra-observer agreement and once by another radiologist at the same period for inter-observer agreement analysis.

Perfusion and enhanced features were calculated based on the VOIs of tumor and FGT. A total of 121 pharmacokinetic parameters were obtained using histogram analysis, including the maximum, minimum, median, mean, area, 10%, 25%, 75%, and 90% of volume transfer constant (Ktrans), rate contrast (Kep), volume fraction of plasma (Vp), time to peak (TTP), maximum concentration (MAX Conc), area under curve (AUC), maximal slope (MAXSlope), blood flow (BF), blood volume (BV), and mean transit time (MTT). Seventy-seven features based on the images in the first post-contrast phase were extracted, consisting of 29 first-order features, 13 gray-level co-occurrence matrix (GLCM), 10 Haralick features derived from GLCM, 16 gray-level run length matrix (GLRLM), and 9 morphology metrics features.

**Radiomic Feature Selection and Radiomics Model Construction**
All data were used for feature selection and model construction, and the same feature selection method was applied for tumor and FGT features. The SLN status was labeled by nominal number, “1” for negative while “0” for positive status. The negative SLN was the main predicting event.

Radiomic features with low reproducibility were excluded firstly. Intra- and inter-observer reproducibility of all extracted radiomic features were assessed using intraclass correlation coefficients (ICCs), and features with ICC less than 0.7 for intra- or inter-observation were excluded (25). Furthermore, following the exclusion of features with variance ≤ 1, the Z-score standardization and normalization were performed.

Next, minimal redundancy maximum relevancy (mRMR) was used to select the optimal 20 features, which were non-redundant and highly informative (26, 27). The least absolute shrinkage selection operator (LASSO) with 10-fold cross-validation (the criteria as maximum area under the ROC curve) was further used to select most candidate radiomic features. Finally, the backward stepwise logistic regression with minimum Akaike Information Criterion (AIC) was conducted and the retained features were involved to construct logistic regression model. Therefore, two independent radiomics “Radscore” for tumor and FGT were calculated according to Equation (1), which were further used as the radiomics signature of each patient:

\[
\text{Radscore} = \beta_0 + \beta_1x_1 + \beta_2x_2 + \ldots + \beta_nx_n
\]  

(1)

where \(\beta_0\) is the constant, \(\beta_i\) is logistic regression coefficient, and \(x_i\) is the value of selected features.

**Construction of Clinical and Combined Radiomics–Clinic Models**
The independent predictors among clinical factors were selected sequentially by univariate (\(p < 0.1\)) and multivariate backward stepwise logistic regression with minimum AIC criteria. The retained clinical factors were used to construct independent clinic model using logistic regression method.

In order to build combined radiomics–clinic model, the constructed radiomics signature (tumor or FGT or Tumor + FGT) were mixed with the clinical factors to sequentially receive univariate (\(p < 0.1\)) and the backward stepwise logistic regression with minimum AIC criteria. The combined model could be constructed if the radiomics Radscore and any clinical factor were retained simultaneously. The nomogram based on the logistic regression coefficients was constructed for the final combined model.

**Evaluation of Model Performance**
The discrimination ability of each model was evaluated by receiver operating characteristic (ROC) curve analysis, from which the AUC, sensitivity, specificity, accuracy, negative predictive value (NPV), and positive predictive value (PPV) could be derived. The continuous net reclassification improvement (NRI) and integrated discrimination improvement (IDI) indices were also calculated to assess the added value of each independent model and their
combinations (28). The model calibration was evaluated by calibration curve analysis and Hosmer and Lemeshow test. The decision curve analysis (DCA) was used for assessing clinical usefulness and net benefit resulted from the predicting model. The Delong’s test was used for comparing each pair of model’s AUC.

Considering the robustness of the independent tumor and FGT radiomics model, the internal validations were performed using 1,000-times bootstrapping (20). Furthermore, the performance of radiomics models was tested in the subsets of different molecular subtypes, i.e., luminal A, luminal B, HER2 enriched, and triple-negative breast cancer.

**Statistical Analysis**

Statistical analysis was implemented by IBM SPSS software (version 22.0, NY, USA) and R software (version 3.5.3; http://www.r-project.org). The continuous variables with normal distribution were evaluated by Student’s t-test and illustrated as mean ± standard deviation, while the continuous variables with non-normal distribution were evaluated by Mann–Whitney U test and represented as median (interquartile range [IQR]). The categorical variables were evaluated by Chi-square to compare the clinical factors between groups with positive and negative SLN. The main R packages were as follows: "icc" for ICC calculation by setting as "twoway" and type of "agreement", "findCorrelation" in "caret" package for correlation analysis, "mRMR" package for mRMR analysis, "glmnet" package for logistic regression and LASSO logistic regression analysis, "pROC" package for ROC analysis, "PredictABEL" package for continuous NRI and IDI indices calculation, and "rmda" package for DCA. The statistical significance was set as two-sided p < 0.05.

**RESULTS**

**Participants**

The data were collected from our hospital for more than 3 years from August 2015 to December 2018. A total of 404 variables were collected from 186 patients. The patients with positive SLN (n = 93) and negative SLN (n = 93) were matched. There were no missing data for clinical factors.

In the positive SLN group, the median number of node was 1 (IQR: 1, 2). As shown in Table 1, the clinical and histopathological variables including age, menopause, tumor size, HER2 status, and ER status between patients with and without SLN metastasis had no significant difference (all p > 0.05), while there were significant differences in ER and PR status between these two groups (both p < 0.05).

| Group           | Patients with positive SLN (n = 93) | Patients with negative SLN (n = 93) | p-value |
|-----------------|-----------------------------------|-----------------------------------|---------|
| Age (mean ± SD) | 49.7 ± 10.2                       | 47.9 ± 9.5                        | 0.208   |
| Menopause       | 45                                | 41                                | 0.556   |
| Median tumor size (IQR) | 2.1 (1.8, 2.6)           | 2.0 (1.5, 2.5)          | 0.083   |
| ER              |                                   |                                   | 0.023   |
| Positive        | 82                                | 70                                |         |
| Negative        | 11                                | 23                                |         |
| PR              |                                   |                                   | 0.003   |
| Positive        | 80                                | 63                                |         |
| Negative        | 13                                | 30                                |         |
| HER2            |                                   |                                   | 0.275   |
| Positive        | 22                                | 16                                |         |
| Negative        | 71                                | 77                                |         |
| Ki-67           |                                   |                                   | 0.869   |
| Positive        | 68                                | 67                                |         |
| Negative        | 25                                | 26                                |         |
| Molecular subtypes |                                   |                                   |         |
| Luminal A       | 21                                | 21                                |         |
| Luminal B       | 61                                | 48                                |         |
| HER2 enriched   | 6                                 | 7                                 |         |
| Triple negative | 5                                 | 17                                |         |

ER, estrogen receptor; PR, progesterone receptor; HER2, human epidermal growth factor receptor-2; SLN, sentinel lymph node.
Radscore\textsubscript{Tumor} = -0.338 - 0.369 \times BF \_ 10percent + 0.677 \\
\times TTP \_ 25percent - 1.194 \times TTP \_ Max \\
- 0.809 \times MTT \_ Max - 0.602 \\
\times MAXSlope \_ Max + 0.722 \times Variance \\
- 0.643 \times Quantile5 - 3.773 \\
\times MeanDeviation \tag{2}

For the FGT model, 172 radiomic features remained with ICC > 0.7 after intra- and inter-observer agreement analysis. Among the 20 optimal radiomic features selected by mRMR, 5 features were selected using LASSO and 3 features were finally retained after backward multivariate logistic regression for logistic model construction (Table 2). The radiomics score for FGT model was expressed as Eq. (3).

Radscore\textsubscript{FGT} = 0.390 + 3.455 \times BF \_ Min - 0.905 \\
\times TTP \_ Max + 0.450 \times Variance \tag{3}

The statistical differences for the selected radiomic features and the constructed Radscores in the tumor model and FGT model between the positive and negative SLN groups are summarized in Tables S1, S2. The Radscore in the negative SLN group was significantly higher than the positive SLN group in both tumor and FGT models.

The clinical factor selection results after univariate and multivariate logistic regression are summarized in Table S3 and the tumor size and PR status were retained to construct the independent clinic model. The clinic model scores were calculated based on the following equation:

Score\textsubscript{Clinic} = 1.930 - 0.476 \times Tumor \text{ size} - 1.178 \times PR

The logistic regression method was also applied to construct the combined models based on the radiomics signature of tumor, FGT, and clinical factors. The univariate and multivariate backward logistic regression analysis results are summarized in Tables S4–S6. The model scores for each combined model were expressed as follows:

\begin{align*}
\text{Score}^{\text{Tumor + FGT}} &= -0.018 + 0.889 \times \text{Radscore}_\text{Tumor} + 0.590 \times \text{Radscore}_\text{FGT} \\
\text{Score}^{\text{Tumor + PR}} &= 1.407 + 1.139 \times \text{Radscore}_\text{Tumor} - 1.776 \times PR \\
\text{Score}^{\text{Tumor + Clinic}} &= 2.401 + 1.163 \times \text{Radscore}_\text{Tumor} - 0.692 \times Tumor \text{ size} - 1.198 \times PR \\
\text{Score}^{\text{Tumor + FGT + PR}} &= 1.379 + 1.032 \times \text{Radscore}_\text{Tumor} + 0.569 \times \text{Radscore}_\text{FGT} - 1.769 \times PR
\end{align*}

**Prediction Performance of Different Models**

The AUC, specificity, sensitivity, accuracy, NPV, and PPV of different predictive models for negative SLN are summarized in Table 3 and Figure 2.

For the tumor model, the AUC and NPV were 0.783 (95% CI, 0.717–0.849) and 0.728 (95% CI, 0.629–0.809), respectively. The AUC of averaged model performance in the resampled test set after 1,000-times bootstrap was 0.710 (95% CI, 0.706–0.713) and the optimism-corrected AUC was 0.700, as shown in Table S7. In addition, during 1,000-times bootstrap, all the radiomic features involved in the tumor model appeared more than 600 times (Figure S1), which indicated the robustness and reliability of the selected features. The AUCs of the tumor model in the subset of luminal A, luminal B, HER2 enriched, and triple-negative breast cancer were 0.728 (95% CI, 0.556–0.899), 0.812 (95% CI, 0.734–0.890), 0.976 (95% CI, 0.910–1.0), and 0.741 (95% CI, 0.489–0.993), respectively.

For the FGT model, the AUC and NPV were 0.680 (95% CI, 0.604–0.757) and 0.660 (95% CI, 0.561–0.747), respectively. The AUC of averaged model performance in the resampled test set after 1,000-times bootstrap was 0.651 (95% CI, 0.646–0.654) and the optimism-corrected AUC was 0.661, as shown in Table S7. Meanwhile, the features in the FGT model also showed reliability with more than 800 times appearing frequency during 1,000-
times bootstrap (Figure S1). The AUCs of the FGT model tested in the subset of luminal A, luminal B, HER2 enriched, and triple-negative breast cancer were 0.632 (95% CI, 0.459–0.806), 0.682 (95% CI, 0.579–0.785), 0.500 (95% CI, 0.151–0.848), and 0.635 (95% CI, 0.301–0.969).

After combining the radiomics signature of tumor and FGT, the AUC and NPV were 0.799 (95% CI, 0.737–0.862) and 0.773 (95% CI, 0.674–0.849), respectively. The AUC of the combined radiomics model was significantly higher than the independent FGT model (Delong’s test, \( p = 0.001 \)), while not significantly different from the independent tumor model (Delong’s test, \( p = 0.145 \)). The AUCs of the combined radiomics model tested in the subset of luminal A, luminal B, HER2 enriched, and triple-negative breast cancer were 0.753 (95% CI, 0.593–0.912), 0.821 (95% CI, 0.744–0.897), 0.976 (95% CI, 0.910–1.0), and 0.753 (95% CI, 0.507–0.999), respectively.

By involving the hormone factor PR expression status into the radiomics model, the AUC of Tumor + PR was significantly higher than that of the independent tumor model (Delong test, \( p = 0.041 \)). For Tumor + PR model, the AUC and NPV were 0.824 (95% CI, 0.765–0.883) and 0.838 (95% CI, 0.736–0.906), respectively. For FGT + Clinic model, the Radscore \( FGT \), tumor size, and PR were simultaneously involved. The AUC and NPV were 0.757 (95% CI, 0.687–0.827) and 0.747 (95% CI, 0.647–0.827), respectively. The introduction of tumor size and PR had a statistically significant effect on AUC improvement for the FGT radiomics model (Delong test, \( p = 0.029 \)).

For the Tumor + FGT + PR model, the AUC and NPV were 0.839 (95% CI, 0.783–0.895) and 0.853 (95% CI, 0.754–0.918), respectively. The AUC of the Tumor + FGT + PR model was significantly higher than any independent or other combined models (Delong test, all \( p < 0.05 \)), except for the Tumor + PR model (Delong test, \( p = 0.098 \)). The nomogram for the combined model is illustrated in Figure 3.

The continuous NRI and IDI indices for the different models are summarized in Table S8. Compared with the independent tumor or FGT radiomics model, the Tumor + FGT model had significant improvement in discrimination and reclassification (both \( p < 0.05 \)). Meanwhile, the combination of tumor or FGT with clinical factors also showed significant improvement by IDI and NRI indices (all \( p < 0.05 \)), compared with each independent model. In addition, the combination of tumor, FGT, and PR simultaneously could obtain further improvement compared with the model combining any two models (all \( p < 0.05 \)).

The calibration curve (Figure 2) and decision curve (Figure 4) also demonstrated that the independent tumor radiomics model and combined model involving tumor signature had a good fit to actual observations (Hosmer and Lemeshow test, \( p > 0.05 \), as shown in Table S9) and had net benefit in a wide threshold probability of 0.35–0.8.

**DISCUSSION**

Lymph node status is an important prognostic factor in breast cancer. This study is the first attempt to predict negative SLN by combining tumor and FGT radiomic features, and further
combining radiomics signature with clinical feature, which is helpful for treatment regimen in breast cancer. The prediction achieved a high AUC of 0.799 for combined radiomics model by combining tumor and FGT radiomics signatures. Furthermore, we developed a nomogram by combining tumor and FGT signatures and histopathological PR expression status to predict SLN status and achieved a higher AUC of 0.839, with a high NPV of 0.853, suggesting benefits for identification of patients with positive SLN, which was consistent with a previous study that combined radiomic features and clinical factors, which were predictive for positive and negative SLN (16) and may help breast cancer patients to avoid unnecessary SLNB and the corresponding complications.

ADC value by DWI and heterogeneous or rim enhancement by DCE-MRI were associated with ALN and SLN metastasis (6–8). Regarding the prediction of SLN status, the AUC of MRI features (heterogeneous and rim enhancement, and peritumoral–tumoral ADC ratio) was 0.80, suggesting that the DCE-MRI can play an important role in the prediction of SLN status. More than enhancement pattern, DCE-MRI also allows for revealing the perfusion and permeability of tissue characterized by multiple pharmacokinetic parameters, including Ktrans, Kep, Vp, TTP, MaxSlope, AUC, and MaxCon, which were able to predict the prognosis in breast cancer (29, 30). As an emerging tool for medical application in recent years, radiomics provides a large number of quantitative features derived from post-contrast images based on DCE-MRI. Liu et al. combined pharmacokinetic parameters and radiomic features of maximum layer in primary tumor to predict SLN status and obtained a satisfactory performance (AUC 0.80) (31). Pharmacokinetic parameters can also be further analyzed by histogram, one kind of radiomics analysis, which has the potential to be a noninvasive biomarker for preoperative differentiation of molecular subtypes (32), lymph node metastasis (33), and proliferative activities (34) of breast cancer. In our study, we analyzed pharmacokinetic parameters using histogram combined with radiomic features of post-contrast images from whole tumor, which obtained a satisfactory performance (AUC 0.783) to predict SLN status, consistent with a previous study (31). Furthermore, regardless of the small sample of subtype-specific analysis, HER2 enriched had the best predictive performance (AUC 0.976).

Although radiomic features from primary tumor and peripheral region are difficult to explain, they allow for

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**FIGURE 3** | Nomogram for model combining tumor, fibroglandular tissue (FGT), and progesterone receptor (PR) in prediction of negative sentinel lymph node (SLN). The different variable values corresponds to a point at the top of the figure, and the sum of the points for all the variables obtaining a total point corresponds to the probability of negative SLN at the bottom of the figure. FGT, fibroglandular tissue; PR, progesterone receptor; SLN, sentinel lymph node.

**FIGURE 4** | Decision curves of different prediction models. The black line represents the assumption that no patients have negative sentinel lymph node (SLN) status (NONE), and the gray line represents the assumption that all the patients have negative SLN status (ALL). The colored decision curves respectively show the net benefit per patient based on the different models: Tumor model (green); FGT model (brown); Clinic model (blue); Tumor+FGT combined model (pink); Tumor+PR combined model (purple); FGT+Clinic combined model (black); Tumor+FGT+PR combined model (magenta). The decision curve represents the clinical net benefit (y-axis) calculated by subtracting false-positive rate (weighted by relative harm: threshold probability/(1−threshold probability)) from the true-positive rate, when choosing different threshold probabilities based on the model cut-value. The closer the decision curves to the black and gray curves, the more similar the net benefit of the models as those from the assumption that “NONE” or “ALL” patients have positive labels (the negative SLN in the current study). The decision curves also could be used for comparing the net benefit of different models within a specific threshold probability point or range. The higher the decision curve of the model, the larger the net benefit. FGT, fibroglandular tissue; PR, progesterone receptor.
quantitatively expressing tissue heterogeneity. Dong et al. demonstrated that Global_Var, GLCM, GLSZM, and GLRLM from T2-weighted fat suppression or DWI were selected to predict SLN metastasis and the AUC was 0.863 (17). Liu et al. selected GLCM, NGLDM, and Laws_Skewness after LSS5 of intratumoral or peritumoral regions derived from wash-out, wash-in, or signal enhancement ratio maps based on DCE-MRI for prediction of SLN metastasis (AUC 0.836) (16). In summary, both aforementioned second-order (GLCM, GLRLM, GLSZM, and NGLDM) and first-order radiomic features were probably selected for prediction of SLN status (31, 35). Nevertheless, in our study, for intratumoral radiomic feature from post-contrast phase, first-order features (Variance, Quantile5, MeanDeviation) were mainly selected to predict negative SLN, which also implied the tumor heterogeneity. Moreover, the selected features from whole tumor in our study also included pharmacokinetic parameters (BF_10percent, TTP_25percent, TTP_Max, MTT_Max, MAXSlope_Area) after histogram analysis, which is consistent with a previous study (31), suggesting that massive angiogenesis in tumor may contribute to the perfusion change.

Most radiomics studies in breast cancer emphasized on the features extracted from intratumoral and peritumoral regions for prediction of molecular subtypes (13), lymph node metastasis (15), and prognosis (14, 36). To our best knowledge, few studies have constructed a radiomics model using FGT features for classification and prediction in breast cancer. The predictive value of FGT in breast cancer studies is still controversial. Vreemann et al. determined that FGT was related to short-term breast cancer risk (18). However, Dontchos et al. found no difference in FGT amount between cancer and control groups (37). We tried to investigate the predictive performance of FGT for SLN status in breast cancer. In the current study, the predictive value of FGT was relatively low (AUC 0.680), as well as in all the subtype sets. However, the introduction of FGT features into the tumor model could improve the predictive performance as shown by continuous NRI and IDI indices in our study, achieving a higher AUC of 0.799 compared with the FGT or tumor model alone, consistent with previous studies that show that the combination model could improve predictive performance (16, 17, 31).

Clinical factors such as age, tumor size, tumor location, tumor histological type, and lymphovascular invasion were significant predictors for SLN metastasis in breast cancer (38, 39). Nomograms based on the clinical factors, such as the Memorial Sloan-Kettering Cancer Center (MSKCC) nomogram, have been developed to predict the likelihood of SLN metastasis in breast cancer with good prediction performance in different populations (39–41). In the current study, the clinical factors PR and tumor size were selected as independent predictors to construct an independent clinic model for predicting SLN status. The PR expression with an odds ratio of 0.308 means that the positive PR is inversely related to the negative SLN, consistent with a previous study that shows that PR was a predictor for SLN metastasis (42). The biological mechanism of PR for SLN metastasis in breast cancer is still unclear. One probable interpretation is that PR is associated with cell proliferation, migration, attachment, and invasiveness in breast cancer (43, 44). In addition, the tumor size with an odds ratio of 0.621 indicated that the smaller tumor size is related to higher probability of the negative SLN, which is in accordance with previous studies (38, 39, 42).

Furthermore, previous studies have demonstrated that the combination of clinical factors and radiomic features can improve the prediction performance of SLN status (16, 45). We found that the AUC was improved from 0.799 to 0.839 for the radiomics model combining tumor with FGT after further involving the clinical factor of PR, which was consistent with a previous study (16). However, different studies obtained different AUCs of predictive models based on radiomic features from intratumoral and peritumoral regions and clinical factors (16, 17, 31, 35). One of the possible reasons is that we predict negative SLN instead of positive SLN. The other possible reason is that the different features could be extracted by different imaging series, image processing, software, and composition of breast cancer subtypes.

There were several limitations in this study. First, the sample size of the study was relatively small for breast cancer patients and small for different molecular subtypes. Therefore, we tried to use the whole dataset to construct the model and internally validated by 1,000-times bootstrapping. Second, the segmentations of tumor and FGT for radiomic features extraction were conducted by manual method although the acceptable inter- and intra-observer reproducibility was achieved. Third, external validation of this study is absent. Multicenter validation is needed to achieve high-level evidence for further clinical application. Thus, the data should be further collected and further validation and physiological explanation for the robust features should be explored as well.

In conclusion, introducing FGT and hormone receptor PR on the basis of tumor radiomics model could help improve the prediction performance of SLN status in breast cancer. A nomogram integrating the DCE-MRI radiomics signature of tumor and FGT and PR status achieved good performance in the prediction of SLN status. This study provided a potential biomarker for predicting preoperative SLN status, which might be helpful for planning treatment strategy.

DATA AVAILABILITY STATEMENT

The datasets presented in this article are not readily available because they were also part of an ongoing study. Requests to access the datasets should be directed to PZ, penghyzhou@126.com.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by SCCHE2015029. Written informed consent for
participation was not required for this study in accordance with the national legislation and the institutional requirements.

AUTHOR CONTRIBUTIONS
GX, JR, PZ, CW, and XC contributed to conception and design of the study. XC, CW, HL, YL, RM, and MW organized the database and analyzed the data. SL performed the statistical analysis. CW wrote the study. XC, CW, HL, YL, RM, and MW organized the database and the national legislation and the institutional requirements. participation was not required for this study in accordance with national legislation and institutional requirements.

FUNDING
Chengdu Science and Technology Program; No. 2021-YF05-01507-SN.

SUPPLEMENTARY MATERIAL
The Supplementary Material for this Article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2021.754843/full#supplementary-material

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