Circulating Angiopoietin-2 Is a Marker for Early Cardiovascular Disease in Children on Chronic Dialysis

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Abstract

Cardiovascular disease (CVD) is increasingly recognised as a complication of childhood chronic kidney disease (CKD) even in the absence of diabetes and hypertension. We hypothesized that an alteration in angiopoietin-1 and -2, growth factors which regulate endothelial and vascular function could be involved. We report that the endothelial survival factor, angiopoietin-1 is low in children with pre-dialysis CKD whereas the pro-inflammatory angiopoietin-2 is elevated in children on dialysis. In dialysis patients, angiopoietin-2 positively correlated with time on dialysis, systolic blood pressure, and carotid artery intima media thickness. Elevated angiopoietin-2 levels in dialysis versus pre-dialysis CKD patients were also associated with an anti-angiogenic (high soluble VEGFR-1 and low VEGF-A) and pro-inflammatory (high urate, E-selectin, P-selectin and VCAM-1) milieu. Ang-2 was immunodetected in arterial biopsy samples whilst the expression of VEGF-A was significantly downregulated in dialysis patients. Serum urate correlated with angiopoietin-2 levels in dialysis patients and addition of uric acid was able to induce rapid release of angiopoietin-2 from cultured endothelial cells. Thus, angiopoietin-2 is a marker for cardiovascular disease in children on chronic dialysis and may act as an anti-angiogenic and pro-inflammatory effector in this context. The possibility that the release of angiopoietin-2 from endothelia is mediated by urate should be explored further.

Introduction

Children with chronic kidney disease (CKD) develop early onset cardiovascular disease (CVD). [1] Manifestations of CVD in childhood CKD include arterial stiffening [2] and calcification, [3] premature atherosclerosis, [4] and left ventricular hypertrophy. [5] Over time, CKD developing in children is associated with increased cardiovascular mortality that markedly accelerates once dialysis is initiated. [6,7]

One of the earliest signs of CVD in individuals with CKD is endothelial damage and dysfunction, [8] and this has been shown even in children with pre-dialysis CKD. [9] In this context, potential causes of endothelial damage and aberrant repair are disturbances in growth factors involved in the formation of vascular networks. [10] Angiopoietin-1 (Ang-1) binds and activates the Tie-2 receptor on endothelia where it promotes cell survival and decreases vascular permeability. [11] As such, Ang-1 is usually considered beneficial for endothelial cell function. In contrast, Ang-2 is released from Weibel-Palade bodies by various stimuli [12,13] and acts as an antagonist of Ang-1. [14] Ang-2 has pro-inflammatory actions [15,16] and can also promote or retard angiogenesis dependent on the ambient levels of vascular endothelial growth factor-A (VEGF-A). [14] Other evidence exists that, in certain circumstances, Ang-2 may have biological effects, independent of the antagonism of Ang-1. [17,18]

Elevated circulating Ang-2 has been reported in adults with CKD. David and colleagues [19] found an inverse relationship between circulating Ang-2 levels and glomerular filtration rate in adults with CKD. Two other studies reported that Ang-2 levels were elevated in adults on hemodialysis (HD) or peritoneal dialysis (PD) compared with healthy controls. [20,21] In one of these studies Ang-2 correlated with scoring for coronary and peripheral arterial disease. [20] In the other study, Ang-2 correlated with cholesterol, high-sensitive C-reactive protein and osteoprotegerin and was an independent predictor of mortality. [21]

To date, no clinical studies have examined angiopoietins in childhood CKD, despite the latter having similar cardiovascular complications as adults with CKD, but at a proportionately earlier age. [1] These effects are more likely to be directly attributed to the uremic milieu because children seldom have diabetes or dyslipidaemia, uncontrolled hypertension or are smokers which themselves predispose to CVD. We hypothesized that an imbalance of angiopoietin vascular growth factors, which would be detrimental to endothelial structure and function, might be
present in children with CKD. Specifically, we predicted that childhood CKD would be associated with elevated Ang-2 and that it would correlate with inflammatory markers.

Materials and Methods

Patient cohort

Informed written consent was obtained from the next of kin, caretakers, or guardians on the behalf of the minors/children participants, and children also gave their assent where appropriate. The study was approved by the Great Ormond Street Hospital and UCL Institute of Child Health research ethics committee. From January to December 2010, 20 children in predialysis CKD stages 4-5 and 30 on dialysis (14 PD, 16 on HD) were recruited from Great Ormond Street Hospital. Primary diagnoses included renal dysplasia (n = 20), posterior urethral valves (n = 9), focal segmental glomerulosclerosis (n = 6), nephroblastic (n = 4), cortical necrosis (n = 3), and 2 each with autosomal recessive polycystic kidney disease, congenital nephrotic syndrome, bilateral Wilms’ tumors and unknown causes. None of the children had diabetes and none were smokers. Children with underlying inflammatory disorders, such as glomerulonephritis and vasculitides were excluded. Patients were compared with healthy age- and gender-matched children who formed part of a contemporaneous study and are previously described. [22]

Clinical, biochemical and vascular parameters

All measures were taken at the same clinical visit; pre a midweek session of HD or at clinic review for pre-dialysis CKD and PD patients. All children had their height, weight, body mass index (BMI) and Doppler blood pressure measured; these were expressed as standard deviation score (SDS) for age and gender. [23] Routine blood tests (including creatinine, calcium, ionized calcium, phosphate, parathyroid hormone and serum urate) were performed. All children above 5 years of age (n = 24 children on dialysis [11 on PD, 13 on HD]; 14 children in pre-dialysis CKD and 25 healthy controls) underwent vascular scans to assess carotid artery intima media thickness (cIMT) and aortic pulse wave velocity (PWV) using methods previously described [24] and expressed as SDS for age. [25,26] Serum was obtained and ELISA used to assess circulating levels of human Ang-1, Ang-2, VEGF-A, Fli-1, E-selectin, P-selectin, intracellular adhesion molecule 1 (ICAM-1) and vascular cell adhesion molecule 1 (VCAM-1) (R & D Systems). In some cases, serum samples were taken both preand post-HD.

Immunolocalisation of Ang-1, Ang-2 and VEGF-A in intact arteries

Medium-sized muscular arteries routinely removed at omentectomy during a peritoneal dialysis catheter insertion or at renal transplantation were obtained from some of the pre-dialysis CKD and dialysis patients (n = 4 in each group). [3] Tissues were fixed in formalin, embedded, then sections cut for immunohistochemistry as described [27] for the following antibodies: rabbit anti-mouse Ang-1 (ADI); rabbit anti-mouse Ang-2 (ADI); rabbit anti-human VEGF-A (Santa Cruz) and rabbit anti-human von Willebrand factor (DAKO). Intensity of staining was quantified by a blinded observed and scored between 0 (no reactivity) to 3 (strong staining); at least four images were obtained from each vessel and a mean value obtained for each specimen.

Uric acid stimulation of human umbilical vein endothelial (HUVEC) and aortic smooth muscle cells (HAoSMC)

HUVEC and HAoSMC (Lonza) were cultured in either EGM-MV or DMEM supplemented with 20% FBS, 25 mM HEPES, 100 U/ml penicillin and 100 mg/ml streptomycin respectively. Cells from passage 2-4 were grown to 70% confluence, placed in low-serum media for 24 hours and challenged with varying concentrations of uric acid (3-12 mg/dl) [28] for 15 minutes, 24 hours and 72 hours. Conditioned media was collected at all time-points to assess Ang-2 levels and cell lysates extracted for protein measurements. In other experiments, RNA was extracted from cells stimulated with uric acid for 6 hours and used for RT-PCR for Ang1, Ang2, organic anion transporters 1-4 (Oat1-4) Tie1, Tie2, Toll-like receptor 4 (Tlr4) and human uric acid transporter 1 (Urat1) using previously described methods. [29] Quantitative RT-PCR was also performed for Ang2 on HUVEC exposed to uric acid (n = 3 for each dose) with hypoxanthine-guanine phosphoribosyltransferase (HPRT) used as a house-keeping gene. Primer details available on request.

Statistics

Results

Circulating Ang levels in pre-dialysis CKD and dialysis patients

Demographic and clinical parameters of the groups studied are summarized in Table 1. The pre-dialysis CKD and dialysis patients were similar in all demographic, clinical and biochemical markers except that 25-hydroxyvitamin D was lower and serum cholesterol and urinary albumin/creatinine ratio higher in dialysis patients (Table 1). The healthy controls had significantly higher BMI SDS and lower blood pressure SDS and urate levels versus the patients.

Ang-1 levels were modestly but significantly (p = 0.02) lower in pre-dialysis CKD patients compared with healthy controls (respectively means±SD being 2.9±1.8 and 4.3±1.8 ng/ml). In dialysis patients, Ang-1 levels (mean±SD 5.0±3.5 ng/ml) were similar to values found in healthy controls. Circulating Ang-2 levels were not significantly different between healthy children and those with pre-dialysis CKD, but were markedly and significantly increased in the dialysis group (means±SD in controls 2.7±1.2, pre-dialysis CKD 2.7±0.9, dialysis 10.5±6.9 ng/ml, p<0.0005 in comparisons between dialysis patients and both the other groups). As explained in the Introduction, Ang-2 acts an endogenous antagonist to Ang-1, such that comparative levels may be relevant; hence we evaluated the Ang-2/Ang-1 ratio. There was no difference in the Ang-2/Ang-1 ratios (Figure 1C) between control
and pre-dialysis CKD individuals but it was significantly higher in dialysis patients compared with the other groups (means ± SD in controls 0.8 ± 0.7, pre-dialysis CKD patients 1.2 ± 0.7 and dialysis patients 2.5 ± 1.4 ng/ml, p < 0.0005 in comparisons between dialysis and both the other groups).

**Table 1.** Demographic, clinical, anthropometric, and biochemical characteristics of patients and control subjects.

| Characteristics                      | Pre-dialysis CKD (n = 20) | Dialysis (n = 30) | Healthy Controls (n = 25) | p    |
|--------------------------------------|---------------------------|-------------------|--------------------------|------|
| Age (yr)                             | 10.7 ± 4.1                | 14.2 ± 3.9        | 13.1 ± 2.8               | 0.68 |
| Sex (males/females)                  | 12/8                      | 17/13             | 14/11                    | 0.82 |
| Race (White/Asian/Black/other)       | 14/5/1/0                  | 19/7/2/2          | 16/7/2/0                 | 0.85 |
| eGFR (ml/min per 1.73 m²)            | 18.3 ± 6.0                | -                 | 113 ± 9.8                | -    |
| Time in CKD4-5 pre-dialysis (yr median [IQR]) | 4.5 (1.1–9.2)          | 3.9 (0.2–7.9)     | -                        | 0.70 |
| Time on dialysis (yr median [IQR])   | -                         | 1.4 (0.2–3.9)     | -                        | -    |
| Dialysis modality (PD/HD)            | -                         | 14/16             | -                        | -    |
| BMI SDS                              | −0.6 ± 1.1                | −0.7 ± 0.3        | 1.1 ± 0.7                | 0.42 |
| Systolic BP index*                   | 1.9 ± 0.8                 | 1.5 ± 2.5         | 0.8 ± 0.2                | 0.21 |
| Numbers on antihypertensive medications | 11                        | 4                 | 0                        | 0.6  |
| Numbers on ACEi or ARB               | 4                         | 0                 | 0                        | 0.1  |
| Hemoglobin (g/dl)                    | 12.3 ± 1.9                | 11.4 ± 0.8        | 12.1 ± 0.9               | 0.51 |
| Albumin (g/L)                        | 39.0 ± 4.1                | 41 ± 4.8          | 40 ± 0.6                 | 0.34 |
| Total cholesterol (mmol/L)           | 3.5 ± 1.3                 | 4.1 ± 0.9         | 3.1 ± 0.7                | 0.07 |
| Triglycerides (mmol/L)               | 1.1 ± 0.7                 | 1.4 ± 2.1         | 0.9 ± 0.6                | 0.11 |
| No. on statins                       | 0                         | 1                 | 0                        | -    |
| Albumin-adjusted calcium (mmol/L)    | 2.4 ± 0.2                 | 2.4 ± 0.1         | 2.4 ± 0.2                | 0.9  |
| Serum phosphate levels (mmol/L)      | 1.4 ± 0.6                 | 1.6 ± 0.8         | 1.2 ± 0.2                | 0.9  |
| Parathyroid hormone (pmol/L)         | 5.2 ± 1.1                 | 8.9 ± 3.7         | -                        | 0.06 |
| Serum urate level (μmol/L)           | 260 ± 20.8                | 278 ± 29.3        | 184 ± 33.0               | 0.88 |
| 25-hydroxyvitamin D (nmol/L)         | 40.1 ± 16.2               | 12.9 ± 9.8        | -                        | 0.04 |
| Urinary albumin / creatinine ratio (mg/mmol) | 122.8 ± 18.6             | 260.0 ± 64.3 (n = 21) | -                        | 0.04 |

All values are presented as mean ± SD; p value indicates comparisons between the pre-dialysis CKD and dialysis groups. Parathyroid hormone, 25-hydroxyvitamin D and urinary albumin / creatinine ratio were not measured in healthy controls due to small volumes of serum and lack of urine samples. * Systolic BP index = measured BP / 95th centile BP for age, gender, and height. ARB, angiotensin II receptor blocker; ACEi, angiotensin-converting enzyme inhibitor; BMI, body mass index; SBP, systolic BP; SDS, SD score.

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Correlation of Ang levels with clinical and vascular measures

Ang-2 levels had no significant relation to age or gender, but increased linearly with time on dialysis (r = 0.37, p = 0.002) whereas there was no association of Ang-2 levels with the time spent in pre-dialysis CKD (p = 0.8, Figure 2A). There was no
difference in Ang-2 levels between HD and PD patients. Circulating Ang-2 levels was also not significantly related to the presence of residual renal function. To determine whether Ang-2 was cleared by HD, we obtained serum samples pre- and post-HD from 5 individuals. There was no significant differences in Ang-2 levels (means ± SD 5.0 ± 1.1 and 4.6 ± 0.9 ng/ml, p = 0.7).

Serum urate levels were significantly increased in both predialysis CKD and dialysis patients compared with controls (Table 1) and showed a weak positive correlation with systolic blood pressure SDS in these patients (r = 0.12, p = 0.048). Urate levels positively correlated with Ang-2 levels in the dialysis group (r = 0.52, p = 0.004, Figure 2B). There was a strong positive correlation between Ang-2 levels and systolic blood pressure SDS in the dialysis patients (r = 0.64, p = 0.003), but not in the predialysis CKD group. No significant correlations were found between Ang-1 or Ang-2/Ang-1 ratio with any clinical, biochemical or vascular parameters.

Three out of 14 (21%) pre-dialysis CKD patients had increased cIMT compared with age-matched controls (0.37 ± 0.03 versus 0.38 ± 0.02 mm respectively), but there was no significant correlation between Ang-2 and cIMT in this group (p = 0.82, Figure 2D). In contrast, cIMT was increased in 16 of 24 (66%) dialysis patients (0.46 ± 0.05 mm) and showed a strong positive correlation with Ang-2 (r = 0.62, p = 0.0005, Figure 2D). PWV was increased in two out of 14 children with predialysis CKD (5.1 ± 0.2 m/sec in pre-dialysis CKD versus 5.0 ± 0.3 m/sec in controls) and 7 out of 24 (5.6 ± 0.5 m/sec) on dialysis but did not show any correlation with Ang-2 in either group. Ang-2 levels were not significantly correlated with blood cholesterol, triglyceride, albumin, calcium, phosphate, parathyroid hormone, 25-hydroxyvitamin D or urinary albumin/creatinine levels in predialysis CKD or dialysis patients. On multiple regression analysis the significant determinants of Ang-2 levels were systolic blood pressure and serum urate levels (Table 2). Carotid IMT was significantly and independently influenced by the time on dialysis, calcium x phosphate product and Ang-2 levels (Table 2).

Circulating levels of VEGF-A and sFlt-1

The biological actions of Ang-2 on blood vessels are dependent on VEGF-A availability; [14] so we measured circulating levels of this growth factor and the endogenous VEGF-A inhibitor, sFlt-1. Ang-2 levels were similar in healthy controls and pre-dialysis CKD patients and therefore VEGF-A and sFlt-1 levels were only measured in pre-dialysis CKD and dialysis patients. VEGF-A levels were significantly lower in individuals on dialysis compared with pre-dialysis CKD patients (respective medians being 6.9 and 33.5 pg/ml, p = 0.003, Figure 3A). In contrast, sFlt-1 levels were significantly higher in dialysis patients compared with pre-dialysis CKD (respective means ± SD of 222 ± 78 and 121 ± 54 pg/ml respectively, p<0.0001, Figure 3B). There were no significant...
correlations of either VEGF-A or sFlt-1 with time or mode of dialysis, urate levels, blood pressure SDS or any other vascular measures.

Circulating levels of soluble cell adhesion molecules

Ang-2 has been shown to have pro-inflammatory actions. Therefore, circulating levels of cell adhesion molecules which attract inflammatory cells were measured. Compared with pre-dialysis CKD individuals, patients treated with dialysis had significantly elevated levels of soluble E-selectin (respective means ± SD being 72 ± 31 and 54 ± 21 ng/ml, p = 0.03, Figure 4A), soluble P-selectin (75 ± 27 versus 52 ± 17 ng/ml, p = 0.001, Figure 4B) and soluble VCAM-1 (1.9 ± 0.5 versus 1.3 ± 0.4 μg/ml, p < 0.0001, Figure 4C) but there was no difference in ICAM-1 (354 ± 99 versus 341 ± 91 ng/ml, Figure 4D). In the dialysis population, circulating levels of Ang-2 positively correlated with soluble VCAM-1 (r = 0.41, p = 0.02), but there were no significant correlations with E-selectin, ICAM-1 or P-selectin.

Immunolocalisation of vascular growth factors in arteries

To seek potential source(s) of Ang-1, Ang-2 and VEGF-A immunohistochemistry was undertaken on intact arteries obtained from a subset of the pre-dialysis CKD and dialysis patients. Ang-1 protein was detected in the media of vessels from pre-dialysis CKD (Figure 5A) and dialysis patients (Figure 5B). As scored by an observer blinded to the source of the samples, there was no difference in staining intensity between the two groups (Figure 5C). Ang-2 was also immunodetected in the media of both pre-dialysis CKD (Figure 5D) and dialysis (Figure 5E) vessels with a similar intensity in each group (Figure 5F). Ang-2 expression was also detected in the endothelial layer which was also positive for von Willebrand factor (Figure 5G, 5H). VEGF-A immunostaining was prominent in the media of pre-dialysis CKD vessels (Figure 5I), but was significantly decreased in dialysis patients (Figure 5J and K).

Effect of uric acid exposure on Ang-2 release in-vitro

As demonstrated above, Ang-2 levels strongly and positively correlated with urate levels in dialysis patients. We hypothesized that elevated urate might increase Ang-2 expression by, and/or release from, endothelial and/or vascular smooth muscle cells. There has been a previous report that uric acid can stimulate release of contents from Weibel-Palade bodies including Ang-2. [13] We first examined how urate may enter HUVEC and detected the urate transporter, Urat1, but not Oat1-4; the mRNA levels of Urat1 showed a tendency to decrease with increasing doses of uric acid stimulation (Figure 6A). HUVEC also expressed Ang-1, Ang-2, Tie-1 and Tie-2 (Figure 6A).

Exposure of HUVECs to uric acid for 15 minutes led to an increase in Ang-2 release versus control media as evaluated by the proportion of Ang-2 protein in the conditioned media. The most prominent response was observed with 12 mg/dl with Ang-2 levels significantly elevated compared with all other groups (Figure 6B). This was an acute effect because longer term stimulation with uric acid for 72 hours did not enhance the release of Ang-2 protein in the conditioned media (Figure 6C). Within the cells, uric acid

### Table 2. Multiple regression analyses for independent predictors of Angiopoietin-2 (Ang2) and carotid intima media thickness (cIMT).

| Variables         | β   | SE   | p       | Model R² |
|-------------------|-----|------|---------|----------|
| Ang2              | 71% |       |         |          |
| Systolic BP       | 2.54| 0.21 | <0.001  |          |
| Serum urate level | 0.14| 0.06 | 0.03    |          |
| cIMT              | 68% |       |         |          |
| Time on dialysis  | 0.50| 0.02 | 0.008   |          |
| Ca x P product    | 0.37| 0.12 | 0.02    |          |
| Ang2              | 0.26| 0.06 | 0.05    |          |

β - Unstandardized regression coefficient; indicates the difference in the outcome variable (Ang2 or cIMT) per unit change in the independent variables.
SE = standard error
Model R² - The amount of variance in the dependent variable that can be explained by the model.

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Figure 3. Circulating levels of VEGF-A and sFlt-1 in pre-dialysis CKD and dialysis patients. VEGF-A levels were significantly lower in individuals on dialysis compared with pre-dialysis CKD patients (A). In contrast, sFlt-1 were significantly higher in the dialysis patients (B).
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stimulation led to a decreased abundance of Ang-2 mRNA after 6 hours of stimulation (Figure 6D), compared with controls. It has been suggested that the acute release of Ang-2 from endothelia is mediated by Tlr4 [13] and we detected mRNA levels of Tlr4 on HUVECs (Figure 6A). Prior studies have shown that HAoSsMC express the Urat1 receptor [28] and in the current study they were also found to express transcripts for Ang-1, Ang-2 and Tie-2, but not Tie-1 (data not shown); however, we did not detect Ang-2 protein in the conditioned media with/without addition of uric acid.

Discussion

Our study demonstrated that circulating Ang-2 levels were markedly elevated in dialysis patients compared with healthy controls and pre-dialysis CKD individuals. Amongst the dialysis patients, Ang-2 positively correlated with time on dialysis, systolic blood pressure and cIMT, but not PWV. These findings may indicate that circulating Ang-2 is a marker for the early cardiovascular changes occurring in children with CKD on dialysis. Previous studies have demonstrated that in the more compliant vessels of children with CKD structural changes precede functional alterations with increases in cIMT observed before alterations in PWV. [30] Furthermore, our work examining intact vessels from children on dialysis indicated that the vessel calcium load showed a strong linear association with cIMT but not with PWV or the coronary calcification score. [3]

Our findings concur with several studies that have shown a relationship between circulating Ang-2 levels and cardiovascular complications in adults. Elevated circulating Ang-2 is associated with scores for coronary and peripheral arterial disease in adults with CKD on PD or HD [20] and positively correlated with systolic blood pressure and left ventricular hypertrophy in 4000 young to middle-aged individuals. [31]. A further study [21] demonstrated that Ang-2 was an independent predictor of mortality in CKD patients and correlated with markers of vascular disease (cholesterol, hsCRP and osteoprotegerin) but not the degree of vascular calcification or arterial stiffness. The observation that circulating Ang-2 is also elevated in children on dialysis suggests that the uraemic environment may directly influence vascular growth factor expression. This is because children do not have many of the cardiovascular comorbidities that are commonly seen in adults. In addition, the pathophysiology of CVD in children may be different to that found in adults, for example, our

Figure 4. Circulating levels of soluble cell adhesion molecules. Compared with pre-dialysis CKD individuals, patients treated with dialysis had significantly elevated levels of soluble E-selectin (A), P-selectin (B) and VCAM-1 (C); there was no difference in ICAM-1 levels (D). doi:10.1371/journal.pone.0056273.g004
previous work has shown that children on dialysis develop arteriosclerosis with exclusively medial involvement [3] whereas adults are much more likely to have both intimal lesions as well as medial damage [32]. Therefore results from adults may not be able to be directly extrapolated to the paediatric population and studies in children with CKD are necessary.

Our studies found that the elevation in circulating Ang-2 levels were similar immediately before and after a HD session. Both Ang-1 and Ang-2 form multimeric structures composed of monomers of 55 kDa [33] and therefore unlikely to be affected by dialytic clearance. In contrast to adults with CKD [19,21] we did not detect different Ang-2 levels in children with pre-dialysis CKD compared with healthy controls. One explanation for this

Figure 5. Immunolocalisation of vascular growth factors in arteries. Ang-1 was detected in the media of vessels from both pre-dialysis CKD (A) and dialysis patients (B); no differences in staining intensity were observed between the two groups (C). Ang-2 was immunodetected in both the media and endothelia (arrows) in pre-dialysis CKD (D) and dialysis (E) vessels with similar intensity (F). The endothelial layer was also positive for von Willebrand factor (arrows, G and H). VEGF-A immunostaining was prominent in the media of pre-dialysis CKD vessels (I), but was significantly decreased in dialysis patients (J and K). All fields taken with ×40 objective. doi:10.1371/journal.pone.0056273.g005
discrepancy could be that the children under study had not been exposed to diabetes mellitus, and that dyslipidaemia and hypertension were less common than in adults with CKD. Indeed, each of these factors have been shown to be associated with elevated Ang-2. [31,34] Instead, children with pre-dialysis CKD had decreased circulating Ang-1 compared with healthy controls. This loss of Ang-1 in pre-dialysis CKD children may decrease blood vessels stability and be an early sign of the endothelial dysfunction which occurs in these patients. [9] Potential sources of Ang-1 not only include the vessel wall, but also platelets [35] and one caveat to consider when measuring circulating Ang-1 in serum samples is that ex-vivo activation may increase Ang-1 levels within serum tubes. [36] In future studies, it would be of interest to quantify both circulating Ang-1 and platelet-derived Ang-1.

Elevated circulating Ang-2 levels in dialysis versus non-dialysis CKD children were associated with an anti-angiogenic environment as demonstrated by decreased circulating VEGF-A and elevated soluble sFlt-1 (Table 3). Increased sFlt-1 [37] and reduced circulating VEGF-A [38] have been demonstrated in adult populations with CKD. In the presence of low VEGF-A, Ang-2 will destabilise blood vessels leading to vessel regression. [14] This milieu of growth factors may therefore contribute to the impaired endothelial function seen in CKD children on dialysis. [9,39]

Increased circulating Ang-2 in CKD children on dialysis was also associated with pro-inflammatory responses with high urate, E-selectin, P-selectin and VCAM-1 (Table 3). Systemic inflammation is seen in children with CKD with dialysis [3,40] and Ang-2 may play a direct role in this process. Ang-2 can sensitise the endothelium to inflammatory responses; [16] and directly affect the biology of inflammatory cells which express the Tie-2 receptor themselves. [41,42] Although, we demonstrated that Ang-2/Ang-1 was also elevated in CKD children on dialysis it did not correlate with any cardiovascular parameters. This suggests that the total Ang-2 levels are important in biological responses in dialysis patients, rather than the relative balance between Ang-2 and Ang-1.

We detected Ang-2 in the endothelium of intact arteries isolated from children with CKD and cultured HUVECs indicating this cell type is a potential source of the increased Ang-2 in dialysis patients. Our studies detected Ang-2 in the walls of intact arteries from both pre-dialysis CKD and dialysis patients and Ang-2 transcripts were detected in cultured HoASMCs. Using Ang-2/ LacZ mice positive expression in renal arterial walls during kidney development has been observed; [43] whilst Ang-2 has been detected in cultured mouse embryonic fibroblasts [44] and smooth muscle cells derived from the heart microvasculature. [45] We could not detect any Ang-2 released from HoASMCs suggesting
vascular smooth muscle cells may not contribute to the increase in Ang-2 seen in dialysis patients. We cannot rule this out completely as the cells used in these experiments were not derived from patients, nor did we reproduce the uremic milieu they will be exposed to in *in vivo*. Another potential source of Ang-2 are macrophages. [41,46] Although prior studies [3] have demonstrated that macrophages are not present in the intact arteries of children on dialysis they may be found in the circulation and increase Ang-2.

There are several potential mechanisms for the increase in circulating Ang-2 in patients with CKD. The increase in Ang-2 may be a direct consequence of elevated blood pressure. Korff and colleagues [47] demonstrated that hypertension in mice led to release of stored Ang-2 from Weibel-Palade bodies. There is also evidence that mediators of vascular tone such as angiotensin II can directly alter Ang-2 expression. [48] A lack of endothelial nitric oxide may also predispose to a release of Weibel-Palade bodies that would theoretically increase Ang-2 levels. [49]

One potential factor that could bring these various mechanisms together is uric acid. Urate is retained in CKD and found to correlate with Ang-2 levels in the dialysis patients. We showed that uric acid could directly induce the release of Ang-2 from HUVECs with a corresponding decrease in mRNA abundance within the cell, consistent with prior reports that uric acid stimulates release of Weibel-Palade bodies. [13] These effects are likely to be mediated by *Urat1* and *Tlr4* [13], both of which were found to be expressed on endothelia. Future studies using inhibitors specific for *Urat1* (probenecid) [29] and *Tlr4* (TAK-242) [50] would help to determine the specific role of these molecules in Ang-2 release from endothelia exposed to uric acid. In addition, there is increasing evidence that urate may have a role in hypertension via effects that include inducing endothelial dysfunction, oxidative stress and the production of angiotensin II. [51] These findings might account for why urate can contribute to cardiovascular complications. [32,33]

In conclusion, Ang-2 is a marker for cardiovascular disease in children on chronic dialysis. Furthermore, we suggest that Ang-2 may also be an anti-angiogenic and pro-inflammatory effector in this context.

**Author Contributions**

Conceived and designed the experiments: RCS LR ASW DAL. Performed the experiments: RCS KLP MKJ AFT DAL. Analyzed the data: RCS KLP DAL. Contributed reagents/materials/analysis tools: DW JD. Wrote the paper: RCS RJJ ASW DAL.

### Table 3. Changes in circulating angiogenic and inflammatory markers between pre-dialysis CKD and dialysis patients.

| Angiogenic factors | Levels in dialysis compared with pre-dialysis CKD patients |
|--------------------|----------------------------------------------------------|
| Ang-1              | No change                                                |
| Ang-2              | ↑                                                        |
| Ang-2/Ang-1 ratio  | ↑                                                        |
| VEGF-A             | ↓                                                        |
| sFlt-1             | ↑                                                        |
| Inflammatory markers |                                                      |
| E-selectin         | ↑                                                        |
| P-selectin         | ↑                                                        |
| VCAM-1             | ↓                                                        |
| ICAM-1             | No change                                                 |

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### References

1. Goodman WG, Goldin J, Kuizon BD, Yoon C, Gales B, et al. (2000) Coronary-artery calcification in young adults with end-stage renal disease who are undergoing dialysis. *N Engl J Med* 342: 1478–1483.
2. Covic A, Mardare N, Gusbert-Tatomir P, Brumaru O, Gavrilovic C, et al. (2006) Increased arterial stiffness in children on haemodialysis. *Nephrol Dial Transplant* 21: 729–735.
3. Shroff RC, McNair R, Figg N, Skepper JN, Schurgers L, et al. (2008) Dialysis accelerates medial vascular calcification in part by triggering smooth muscle cell apoptosis. *Circulation* 118: 1748–1757.
4. Dursun I, Poyrazoglu HM, Gunduz Z, Ulger H, Yykylmaz A, et al. (2009) The relationship between endothelial microparticles and arterial stiffness and atherosclerosis in children with chronic kidney disease. *Nephrol Dial Transplant* 24: 2511–2518.
5. Minifie-FM, Barletta GM, Dresner IG, Chand DH, Geary D, et al. (2006) Severe cardiac hypertrophy and long-term dialysis: the Midwest Pediatric Nephrology Consortium study. *Pediatr Nephrol* 21: 1167–1170.
6. McDonald SP, Craig JC, Australian and New Zealand Paediatric Nephrology Association (2004) Long-term survival of children with end-stage renal disease. *N Engl J Med* 350: 2654–2662.
7. Oh J, Wunsch R, Turzer M, Bahner M, Raggi P, et al. (2002) Advanced coronary and carotid arteriopathy in young adults with childhood-onset chronic renal failure. *Circulation* 106: 100–105.
8. Lilien MR, Geroffoth JW (2009) Cardiovascular disease in children with CKD or ESRD. *Nat Rev Nephrol* 5: 229–235.
9. Kari JA, Donald AE, Vallance IY, Bruckdorfer KR, Leone A, et al. (1997) Physiology and biochemistry of endothelial function in children with chronic renal failure. *Kidney Int* 52: 468–472.
10. Long DA, Norman JT, Fine LG (2012) Restoring the renal microvasculature to treat chronic kidney disease. *Nat Rev Nephrol* 8: 244–250.
11. Kim KT, Choi HH, Steinmetz MO, Maco B, Kammerer RA, et al. (2005) Oligomerization and multimerization are critical for angiopoietin-1 to bind and phosphorylate Tie2. *J Biol Chem* 280: 20126–20131.
12. Fiedler U, Scharpfenacker M, Koidl S, Hegen A, Grunow V, et al. (2004) The Tie-2 ligand angiopoietin-2 is stored and rapidly released upon stimulation from endothelial cell Weibel-Palade bodies. *Blood* 103: 4150–4136.
13. Kuo MC, Patschan D, Patschan S, Cohen-Gould I, Park HC, et al. (2008) Ischemia-induced eoscytosis of Weibel-Palade bodies mobilizes stem cells. *J Am Soc Nephrol* 19: 2321–2330.
14. Maisonpierre PC, Suri G, Jones PF, Bartunkova S, Wiegand SJ, et al. (1997) Angiopoietin-2, a natural antagonist for Tie2, interacts with and phosphorylates Tie2. *Nat Med* 3: 235–239.
15. Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. *Science* 277: 55–60.
16. Schoiz A, Lang V, Henschler R, Gzahanka M, Vajkoczy P, et al. (2011) Angiopoietin-2 promotes myeloid cell infiltration in a β2-integrin-dependent manner. *Blood* 118: 5050–5059.
17. Fiedler U, Reiss Y, Scharpfenacker M, Grunow V, Koidl S, et al. (2006) Angiopoietin-2 sensitizes endothelial cells to TNF-alpha and has a crucial role in the induction of inflammation. *Nat Med* 12: 235–239.
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17. Felcht M, Lack R, Schering A, Seidel P, Srivastava K, et al. (2012) Angiopoietin-2 differentially regulates angiogenesis through TIE2 and integrin signaling. J Clin Invest 122: 1991–2005.

18. Kraus S, Garcia S, Ambarus CA, de Launay D, Foster M, et al. (2012) Angiopoietin-2 promotes inflammatory activation of human macrophages and is essential for murine experimental arthritis. Ann Rheum Dis 71: 1402–1417.

19. David S, Kumpers P, Lukasz A, Fliner D, Martens-Lobenhoffer J, et al. (2010) Circulating angiopoietin-2 levels increase with progress of chronic kidney disease. Nephrol Dial Transplant 25: 2571–2576.

20. David S, Kumpers P, Hellpap J, Horn R, Leitolf H, et al. (2009) Angiopoietin-2 and cardiovascular disease in dialysis and kidney transplantation. Am J Kidney Dis 53: 770–778.

21. David S, John SG, Jefferies HJ, Sigrist MK, Kumpers P, et al. (2012) Angiopoietin-2 levels predict mortality in CKD patients. Nephrol Dial Transplant 27: 1867–1872.

22. Kracht D, Shroff R, Baig S, Doyon A, Jacobi C, et al. (2011) Validating a new oscillographic device for aortic pulse wave velocity measurements in children and adolescents. Am J Hypertens 24: 1294–1299.

23. Cole TJ, Green PJ (1992) Smoothing reference centile curves: the LMS method and penalized likelihood. Stat Med 11:1305–1319.

24. Shroff RC, Donald AE, Hiorns MP, Watson A, Feather S, et al. (2007) Mineral metabolism and vascular damage in children on dialysis. J Am Soc Nephrol 18: 2996–3003.

25. Jourdan C, Wuhl E, Litwin M, Falh K, Trelewiec J, et al. (2005) Normative values for intima-media thickness and distensibility of large arteries in healthy adolescents. J Hypertens 23: 1707–1715.

26. Reza GN, Cooperkal O, Temmar M, Kis E, Cherif AB, et al. (2010) Reference values of pulse wave velocity in healthy children and teenagers. Hypertension 56: 217–224.

27. Long DA, Woolf AS, Suda T, Yuan HT (2001) Increased renal angiopoietin-1 expression in a ficoll-induced nephrotoxicity in mice. J Am Soc Nephrol 12: 2721–2731.

28. Price KL, Sautin YY, Long DA, Zhang I, Miyazaki H, et al. (2006) Human vascular smooth muscle cells express a urate transporter. J Am Soc Nephrol 17: 217–224.

29. Mestas J, Ley K (2008) Monocyte-endothelial cell interactions in the development of atherosclerosis. Trends Cardiovasc Med 18: 228–232.

30. Litwin M, Wuhl E, Jourdan C, Trelewiez J, Nierinska A, et al. (2005) Altered morphologic properties of large arteries in children with chronic renal failure and after renal transplantation. J Am Soc Nephrol 16: 1494–1500.

31. Lieb W, Zacharia JF, Xanthakis V, Safa R, Chen MH, et al. (2010) Clinical and genetic correlates of circulating angiopoietin-2 and soluble Tie-2 in the community. Cire Cardiovasc Genet 3: 300–306.

32. London GM, Marchais SJ, Guerin AP, Marchais SJ, et al. (2002) Arterial structure and function in end-stage renal disease. Nephrol Dial Transplant 17: 1713–1724.

33. Davis S, Papadopoulos N, Aldrich TH, Maisonneuve PC, Huang T, et al. (2003) Angiopoietins have distinct modular domains essential for receptor binding dimerization and superchaseriting. Nat Struct Mol Biol 10: 36–44.

34. Lim HS, Lip GY, Blann AD (2005) Angiopoietin-1 and angiopoietin-2 in diabetes mellitus: relationship to VEGF, glycemic control, endothelial damage/dysfunction and atherosclerosis. Atherosclerosis 180: 113–118.

35. Li J, Huang YQ, Basch R, Karpatkin S (2001) Thrombin induces the release of angiopoietin-1 from platelets. Thromb Haemost 85: 204–206.

36. Lukasz A, Hellpap J, Horn R, Kielstein JF, David S, et al. (2008) Circulating angiopoietin-1 and angiopoietin-2 in critically ill patients: development and clinical application of two new immunoassays. Crit Care 12: R94.

37. Di Marco GS, Reuter S, Hillebrand U, Amler S, König M, et al. (2009) The soluble VEGF receptor sFlt contributes to endothelial dysfunction in CKD. J Am Soc Nephrol 20: 2235–2243.

38. Futakaki N, Bunthe P, Laohareungpanya N, Chaisuriya P, Ratanabangnakoon K (2008) A defective angiogenesis in chronic kidney disease. Ren Fail 30: 213–217.