An Amish founder variant consolidates disruption of CEP55 as a cause of hydranencephaly and renal dysplasia

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Abstract

The centrosomal protein 55 kDa (CEP55 (OMIM 610000)) plays a fundamental role in cell cycle regulation and cytokinesis. However, the precise role of CEP55 in human embryonic growth and development is yet to be fully defined. Here we identified a novel homozygous founder frameshift variant in CEP55, present at low frequency in the Amish community, in two siblings presenting with a lethal foetal disorder. The features of the condition are reminiscent of a Meckel-like syndrome comprising of Potter sequence, hydranencephaly, and cystic dysplastic kidneys. These findings, considered alongside two recent studies of single families reporting loss of function candidate variants in CEP55, confirm disruption of CEP55 function as a cause of this clinical spectrum and enable us to delineate the cardinal clinical features of this disorder, providing important new insights into early human development.

Introduction

The centrosomal protein 55 kDa (CEP55) is a centrosome- and midbody-associated protein that has been shown to play a central role in cell cycle regulation and is recognised as a key protein involved in the abscission process, the final stage of cytokinesis [1–3]. CEP55 facilitates abscission through the recruitment of two ESCRT (endosomal sorting complex required for transport)-I subunit associated proteins to the midbody: tumour susceptibility gene 101 (TSG101) and apoptosis-linked gene 2 interacting protein X (ALIX) [4]. The transcribed CEP55 centrosomal protein has three central coiled-coil domains and is expressed at the perinuclear membrane, cytoplasm, and nucleus [2, 5].

Recently candidate homozygous nonsense variants in CEP55 have been identified in five foetuses from two families in association with a lethal disorder, the features of which include dysplastic kidneys and complex brain malformations [6, 7]. Homozygous cep55l knockout mutant zebrafish display a significant reduction in the size of brain structures, and a decreased number of renal tubules, consistent with the human phenotype [6].

Here we describe a novel Amish homozygous founder frameshift CEP55 variant in two affected Amish foetuses presenting with hydranencephaly and Potter sequence secondary to cystic renal dysplasia and anhydramnios.

Materials and methods

Samples were taken with informed consent (University of Arizona protocol 10-0050-01 and Akron Children’s Hospital project number986876-3) for DNA extraction. Single-nucleotide polymorphism (SNP) genotyping was
performed using the HumanCytoSNP-12 v2.1 beadchip array (Illumina). Whole-exome sequencing (WES) analysis (NextSeq500: Illumina) involved: Agilent Sureselect Whole Exome v6 targeting, read alignment (BWA-MEM (v0.7.12), mate-pairs fixed and duplicates removed (Picard v1.129), InDel realignment/ base quality recalibration (GATK v3.4-46), single-nucleotide variant (SNV)/InDel detection (GATK HaploymeCaller), annotation (Alamut v1.4.4), and read depth (GATK DepthOfCoverage). Dideoxy sequencing was undertaken using standard techniques. The CEP55 variant (NM_018131.4: c.514dup; p.(Ile172Asnfs*17) and associated phenotype data were submitted to ClinVar (www.ncbi.nlm.nih.gov/clinvar, accession SCV000808984).

**Fig. 1** CEP55 variants, family pedigree, genotype, and images of affected individuals. a Schematic representation indicating the position of the disease-associated CEP55 gene variants in relation to genomic organisation. The CEP55 c.514dup; p.(Ile172Asnfs*17) variant identified in the current study is highlighted by red text (exons; coloured boxes, introns; black lines). The correlation between coding regions and CEP55 polypeptide functional domains is also indicated; tumour suppressor gene 101 (TSG101) and apoptosis-linked gene 2 interacting protein X (ALIX) binding domains (green), and terminal region of the protein involved in localisation during cytokinesis (yellow). b Simplified pedigree of the Amish family investigated, with electropherograms showing the DNA sequence at the position of the CEP55 variant (c.514dup) confirmed as homozygous in affected twin B and the deceased male sibling, and heterozygous in both parents and unaffected twin A. CEP55 genotype is shown in red under the affected individuals in generations V and VI (+, c.514dup; –, WT). c–h Clinical features of individuals homozygous for the CEP55 variant (c.514dup). c–e VI:1 showing features of Potter sequence (also known as oligohydramnios sequence and used to describe a combination of distinctive facial and other associated phenotypic features that are a result of too little amniotic fluid. Features include epicanthic folds, retrognathia, a flattened nose, low-set ears, pulmonary hypoplasia, and limb contractures including talipes) [15]. Additional features observed included bilateral 2–5 toe syndactyly with a widened first web space and a bulbous nasal tip. f–h VI:3 (twin B) showing features of Potter sequence. The features of both affected foetuses that can be attributed as secondary to oligohydramnios include redundant skin folds, short neck, flattened face, small palpebral fissures and low-set ears, brachydactyly, tapering fingers, and short fifth fingers with clinodactyly.

**Results**

**Subjects**

An Ohio Amish couple, distantly related through a fourth-generation common ancestor (Fig. 1b), presented in their second pregnancy with dichorionic diamniotic twins, one male (twin A) and one female (twin B). Antenatal ultrasound scanning (USS), undertaken at 21 + 2 weeks of gestation, revealed twin B to have hydranencephaly, a multicystic dysplastic right kidney (the left kidney was not visualised), shortened bowed long bones and anhydramnios. Twin A was found to have bilateral renal pelvis dilatation, but no further abnormalities were detected. A subsequent USS undertaken at 28 weeks of gestation to further examine...
**Table 1** Comparison of cases with biallelic CEP55 variants

| CASE                  | Bondesen et al. [7] | Bondesen et al. [7] | Frosk et al. [6] | Frosk et al. [6] | Frosk et al. [6] | This study | This study |
|-----------------------|---------------------|---------------------|------------------|------------------|------------------|------------|------------|
| **Patient**           |                     |                     |                  |                  |                  |            |            |
| CEP55                 |                     |                     |                  |                  |                  |            |            |
| (NM_018131.4)         |                    |                    |                  |                  |                  |            |            |
| **Genotype**          | c.256C>T p. Arg86*  | c.256C>T p. Arg86* | c.1274C>A p. S425* | c.1274C>A p. S425* | c.1274C>A p. S425* |            |            |
| **Sex**               | M                   | M                   | M                | M                | M                | M          | F          |
| **Pregnancy outcome** | TOP (20)            | IUFD (14 + 6)       | SB (30)          | ND (35)          | SB (32)          | SB (42)    | ND (42)    |
| (gestation)           |                     |                     |                  |                  |                  |            |            |
| **Gestation anomalies** | 19+4               | 10+4               | 20               | n/k              | n/k              | 19         | 21+2       |
| identified            |                     |                     |                  |                  |                  |            |            |
| **POTTER SEQUENCE**   | n/a                 | n/a                 | +                | +                | +                | +          | +          |
| **RENS FEATURES**     |                     |                     |                  |                  |                  |            |            |
| Renal aplasia/        | +                   | n/k                 | +                | +                | n/k              |            |            |
| dysplasia             |                     |                     |                  |                  |                  |            |            |
| Ureteral agenesis     | n/k                 | n/k                 | +                | +                | n/k              | n/k        | n/k        |
| Renal cysts           | +                   | n/k                 | -                | -                | n/k              | +          |            |
| Bilateral             | +/-                 | n/k                 | -                | -                | -                |            |            |
| hydronephrosis        |                     |                     |                  |                  |                  |            |            |
| Oligohydramnios       | +                   | n/a                 | +                | +                | +                | +          | +          |
| Pulmonary hypoplasia  | n/k                 | n/k                 | +                | +                | n/k              | n/k        | n/k        |
| Contractures          | -                   | n/a                 | +                | +                | +                | -          | +          |
| Talipes               | +                   | n/a                 | +                | +                | +                |            |            |
| **CNS FEATURES**      |                     |                     |                  |                  |                  |            |            |
| Hydranencephaly       | +                   | -                   | +                | +                | +                |            | +          |
| Cerebral cysts        | -                   | +                   | -                | -                | -                |            |            |
| Cerebellar            | +                   | n/k                 | +                | +                | n/k              | n/k        | n/k        |
| hypoplasia            |                     |                     |                  |                  |                  |            |            |
| Encephalocoele        | -                   | +                   | -                | -                | -                |            |            |
| Multinucleated neurons| n/k                 | n/k                 | +                | +                | n/k              | n/k        | n/k        |
| Skull asymmetry       | +                   | -                   | -                | -                | -                |            |            |
| **GROWTH**            |                     |                     |                  |                  |                  |            |            |
| IUUGR                 | +                   | n/k                 | n/a              | n/a              | n/a              | +          |            |
| **CARDIOAN FEATURES** |                     |                     |                  |                  |                  |            |            |
| Dysplastic aortic     | n/k                 | n/k                 | +                | n/k              | +                | n/k        | n/k        |
| valve                 |                     |                     |                  |                  |                  |            |            |
| Dilated left ventricle| n/k                 | n/k                 | +                | n/k              | +                | n/k        | n/k        |
| Pericardial effusion  | n/k                 | n/k                 | -                | -                | -                |            |            |
| **SKELETAL FEATURES** |                     |                     |                  |                  |                  |            |            |
| Shortened/bowed       | n/k                 | n/a                 | -                | -                | -                |            | +          |
| long bones            |                     |                     |                  |                  |                  |            |            |
| Vertebral abnormalities| n/k                | n/a                 | +                | -                | n/k              | n/k        |            |
| **OTHER FEATURES**    |                     |                     |                  |                  |                  |            |            |
| Cystic hygroma        | +                   | +                   | +                | +                | +                | -          | -          |
twin B revealed intrauterine growth restriction (IUGR) and pericardial effusion, and the bladder and stomach were not identified. The twins were born following an uncomplicated spontaneous vaginal delivery at 42 weeks of gestation. Twin B survived for 90 min after birth; the clinical features of Potter Sequence with cutaneous syndactyly were documented (Fig. 1c). The couple had previously experienced a stillbirth of a male foetus at 42 weeks of gestation. The foetus presented with multiple foetal anomalies at 19 weeks of gestation. Anomalies included: IUGR, anhydramnios, hydranencephaly, described as ‘hydrocephalus seen throughout the cranium’, and bilateral hydronephrosis. The foetal birth weight was 3 lb 14 oz (1.76 kg) and features of Potter sequence, bilateral lower limb bowing, talipes, and syndactyly were identified. (Fig. 1c). Karyotyping of placental samples was normal and post-mortem (PM) examination was declined for both foetuses. Assuming that a founder variant was responsible for the condition, we used a combination of autozygosity mapping and WES to study this novel syndrome and identify the underlying molecular cause.

Genetic studies

Whole-genome SNP genotyping of both twins and the affected brother identified a number of homozygous genomic regions particular to the affected individuals, the largest of which was a 18.3 Mb region of 10q22.3-q24.1 (rs1769756-rs7081796, chr10:g.79,164,647-97,444,769 [hg38]). Subsequent WES of DNA from the affected twin excluded previously described Amish founder variants associated with cystic kidney disease, including NPHP3 [8]. Rare variants predicted to have a functional consequence were cross-referenced with SNP genotyping data, identifying a single candidate homozygous variant of relevance to the phenotype. This variant (NM_018131.4: c.514dup; p. (Ile172Asns*17), chr10:g.93507042dup, [hg38]) in CEP55, predicted to result in a premature stop, is within the chromosome 10 locus. It is present in a single heterozygote in gnomAD and was not listed in ClinVar, NCBI (National Center for Biotechnology Information), or HGMDpro (Human Gene Mutation Database Professional) databases. Dideoxy sequencing confirmed the presence and cosegregation of the variant (Fig. 1b). A total of 179 samples from healthy Amish adults were analysed and 7 heterozygous carriers were identified, corresponding to an estimated allele frequency of 0.02 in this population.

Discussion

This is the third reported family with likely homozygous loss of function variants in CEP55 identified in association with a lethal foetal disorder (comparison of cases; Table 1). Frosk et al. [6] reported a family with Dutch–German Mennonite ancestry and three affected foetuses homozygous for a CEP55 nonsense variant (NM_018131.4) c.1274C>A; p.Ser425* presenting with dysplastic kidneys hydranencephaly, cerebellar hypoplasia, and multinucleated neurones at PM. The authors termed this disorder MARCH syndrome (multinucleated neurons, anhydramnios, renal dysplasia, cerebellar hypoplasia and hydranencephaly syndrome (Online Mendelian Inheritance in Man (OMIM) 236500)) and highlighted an additional nine cases in the literature with phenotypic overlap including features of hydranencephaly, renal dysplasia, and syndactyly [9–13]. However, as far as we are aware, CEP55 genetic analysis has not been undertaken in these individuals. Interestingly, in two cases neuropathological PM findings identified multinucleated neurons [9, 10]. Bondeson et al. [7] reported a Swedish couple with two affected foetuses homozygous for CEP55 c.256C>T; p.Arg86* (NM_018131.4) with features in one including: hydranencephaly, enlarged cystic kidneys, oligohydramnios, and cystic hygroma [7]. The second foetus had a slightly different phenotype comprising

| CASE | Bondesen et al. [7] | Frosk et al. [6] | Frosk et al. [6] | Frosk et al. [6] | This study | This study |
|---|---|---|---|---|---|---|
| Redundant neck skin | n/a | n/a | + | + | + | + |
| Bulbous nasal tip | n/a | n/a | + | + | + | + |
| Syndactyly | n/a | n/a | + | + | + | + |
| Brachydactyly | n/a | n/a | + | + | + | + |
| Widened first web space | n/a | n/a | + | + | + | + |
| Single umbilical artery | + | n/a | – | – | – | – |

The symbol ‘+’ indicates presence of a feature in an affected subject and ‘−’ indicates absence of a feature in an affected subject.

**Table 1** (continued)
occipital encephalocele, cerebral cyst, and cystic hygroma, and the kidneys were severely degraded due to foetal autolysis. PM examination was only possible for one foetus, and although neither were identified to have polydactyly or liver abnormalities, the authors classified the combination of clinical features as a Meckel-like syndrome.

All seven cases described with biallelic CEP55 variants display phenotypical similarity, with renal dysplasia or cysts resulting in oligohydramnios and Potter sequence, and central nervous system (CNS) abnormalities comprising hydranencephaly or cerebral cysts, as the cardinal clinical features (Table 1). Interestingly, as noted by Bondeson et al. [7], the presence of these two congenital anomalies fulfills two of the characteristic Meckel syndrome (MKS) clinical triad [7]. However, in the absence of polydactyly, none of the seven cases described to date fulfills the triad. Although PM examination was undertaken in only three cases, hepatic ductal plate malformation, a frequent finding in classical MKS, was not identified. Occipital encephalocele, the most frequent CNS abnormality in MKS (83.8% of cases) [14], was documented in one foetus [7]. A spectrum of other CNS abnormalities have been described in MKS cases, including cerebral cysts and hydrocephalus, of which hydranencephaly can be considered a severe form. Several other features reported in MKS were identified in this patient cohort, including syndactyly, hydrenephrosis, and short bowed limbs. In view of the wide variability seen in MKS, the lack of diagnostic criteria, and the phenotypic overlap with CEP55 patients, this disorder should be considered a MKS-like condition [7].

The Amish siblings described here enable us to more precisely delineate the clinical consequences of CEP55 loss of function, with hydranencephaly and cystic renal dysplasia as the predominant features. The identification of seven additional Amish control samples heterozygous for the CEP55 frameshift variant highlights the importance of testing when an Amish affected foetus presents with Potter sequence or an MKS-like phenotype. Antenatal USS and PM investigations following stillbirth or neonatal death are infrequently undertaken by the Amish and, despite no previous reports, the allele frequency suggests that this condition is underrecognised.

Unfortunately, while no tissue was available for cerebral histological analysis from the Amish and Bondeson cases [7], all three Frosk cases [6] displayed multinucleated neurons in cerebral tissue, and in one case multinucleated hepatocytes. Disruption of the CEP55 binding site for TSG101 and ALIX likely results from both the p.Arg86* and p.(Ile172Asnfs*17) variants (Fig. 1a), which may prevent the cytokinesis abscission process resulting in incomplete cell division, providing a plausible explanation for the multinucleated neurons. The c.256C>T; p.Arg86* variant, located in exon 3, would be predicted to result in nonsense mediated mRNA decay; however, studies on heterozygous parents identified equal levels of wild-type and truncated transcript [7]. The p.Ser425* variant is predicted to delete the terminal 40 amino acids critical for localisation during cytokinesis. Consistent with this, subcellular localisation studies showed the variant disrupts localisation to the midbody during cell division [6].

The lack of development of cerebral structures suggests that loss of CEP55 function may also play a role in cell migration during embryogenesis and development. Impaired cytokinesis may contribute to impaired neuronal migration in cells with aberrant cellular division, potentially explaining the MKS-like phenotype as both impaired cilia function and cell division processes may cause abnormal neuronal migration. However, further studies of CEP55 function are required to fully determine the precise patho-molecular basis of this lethal multisytem congenital anomaly disorder. Taken together, our findings consolidate CEP55 as a molecule fundamental to normal human development, with homozygous loss of function associated with a Meckel-like lethal foetal disorder profoundly affecting brain and kidney development.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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