Introduction

The treatment of wound in the past was carried out by allowing the wound to dry and form a hard layer, which would then peel off by itself. However, over the advancement in science and technology, the treatment of the wound has changed. It is known that the wound heals more rapidly if the moist is covering the wound as the wound dressing. Traditionally, gauzes made of cotton were used as wound dressings. There are some types of wound dressings such as hydrocolloid, hydrogel, foam, and semi-permeable adhesive membranes [1,2]. The membrane is a preferable one because it is transparent thus easy to observe the level of healing, comfortable to use on the elbows and knees because it is elastic and not easily torn. In addition, membranes are also permeable to oxygen and water vapor to allow skin respiration [3,4].

Membranes are pharmaceutical preparations that have thicknesses between micrometers to millimeters and made by various methods using one or more polymers. Polymers are the main ingredient of the membrane. Meanwhile, plasticizer is usually added to form elastic and flexible membranes. The plasticizer can reduce intermolecular forces and increase the flexibility of membranes by widening the space of molecules and weakening the hydrogen bonds of polymer chains. The amount of plasticizer used can affect the mechanical properties of the membrane. In a previous study [4], the honey membrane was formulated with glycerol, polyethylene glycol, and propylene glycol as plasticizers, which are from polyalcohol groups [5,6].

Membranes with a homogeneous polymeric bonding structure are used to treat damaged skin areas and generally protect injured areas from external factors [7,8]. Aside from being wound dressing, the membrane can also function to help wound healing with the presence of active ingredients contained in the membrane. Several previous studies showed that natural ingredients were added as active substances into the membrane formula such as chitosan [9], haruan (Channa striatus) extract [10], and honey [4].

Eels are Pisces class animals that contain many ingredients useful for humans. These include omega-3 and omega-6 fatty acids. The content of arachidonic acid and DHA in an eel body is 8.25 and 6.21 g per 100 g fat, respectively [11]. Based on a previous study by Febriyenti et al. [12], the fatty acid composition of eel extracts was dominated by oleic acid (19.7%), palmitic acid (18.7%), pentadecanoic acid (15.81%) and octadecanoic acid (4.87%). Omega-3 and omega-6 fatty acids play an important role in the wound healing process. Omega-3 fatty acids play an important role in the wound healing process.
Fatty acids act as anti-inflammatory agents that work to inhibit eicosanoid production [13]. Therefore, eels can be used as additional nutrients to accelerate the wound healing process. The present study aimed to formulate the eel extract membrane with a variety of plasticizers in order to evaluate the effect of the type of plasticizers on thickness, permeability, and mechanical properties of the membranes.

Materials and Methods

Materials

Eels were purchased from Bukittinggi, West Sumatera, Indonesia. Eel extract was prepared, according to Febriyenti et al. [12]. PVA 146,000 was bought from VWR International, Belgium. Glycerol, propylene glycol, PEG 400, methylparaben, and propylparaben were acquired from R&M Chemicals, UK. Distilled water was purchased from Bratachem, Indonesia. All chemicals were used without further purification.

Preparations of Eels Extract Membranes

PVA was prepared by the hot mechanical method [4,14,15]. PVA was dissolved in hot distilled water. Eel extract, methylparaben, propylparaben were dispersed in glycerol, propylene glycol or PEG 400, and then mixed with PVA solution using Ultra turrax at 10,000 rpm for 15 minutes [4,16,17]. The formulae can be seen in Table 1. Furthermore, the membrane was prepared using casting techniques. Some of the liquid membrane formulae were poured into the glass mold (size 20 cm x 20 cm), then allowed to dry at room temperature for approximately 24 hours [4,10,18].

Membrane Thickness

The membrane thicknesses were measured based on the method in previous studies [4,10,19,20] by using a micrometer (Digimatic micrometer, Mitutoyo, Tokyo, Japan).

Water Vapor Permeability of The Membrane

The rates of water vapor permeability of the membrane were determined using the method described in USP XXIV [21]. The method was also used in previous studies [4,10].

Mechanical Properties Measurements

The mechanical properties of the membrane were measured using the same method of previous studies [4,10,22]. A texture analyzer (TA.XT2, Stable Micro System, Haslemere, Surrey, UK) was used to determine

Table 1. Formulae of eel (Monopterus albus) extract membrane

| Ingredients                  | F1  | F2  | F3  |
|------------------------------|-----|-----|-----|
| Eels extract                 | 10  | 10  | 10  |
| PVA                          | 10  | 10  | 10  |
| Glycerol                     | 3   | -   | -   |
| Propylene glycol             | -   | 3   | -   |
| PEG 400                      | -   | -   | 3   |
| Methylparaben                | 0.1 | 0.1 | 0.1 |
| Propylparaben                | 0.02| 0.02| 0.02|
| Distilled water up to        | 100 | 100 | 100 |

Table 2. Thickness and water vapor permeability of eel extract membranes

| Formula | Thickness (mm) | Water vapor permeability (mg/L/day) |
|---------|----------------|-----------------------------------|
| F1      | 0.047 ± 0.005  | 1981 ± 205                        |
| F2      | 0.043 ± 0.001  | 995 ± 328                         |
| F3      | 0.043 ± 0.002  | 631 ± 109                         |
tensile strength value and elongation at the break of the membrane. Young's Modulus (E) was calculated using the equation in Martin et al. [23].

Statistical Analysis

The data were presented as mean ± SD and analyzed statistically by one-way analysis of variance (ANOVA) to compare the results using SPSS software (version 15, USA). Post-hoc Tukey's Honestly Significant Difference (Tukey-HSD) test was applied when there was a statistically significant difference (P<0.05) [9,24].

Results and Discussion

The eel extract membranes formulated in this study function as a wound dressing to keep the wound moist and prevent secondary infection by microorganisms. Membranes have several advantages, including being transparent and permeable to water vapor, allowing the skin tissue respiration. Membranes are also comfortable to use on elbows and knees because they are elastic and not easily torn [1-3]. Based on the previous study [4], PVA is the best polymer that could produce a transparent membrane. Herein, PVA was used as a polymer to form the eel extract membrane. Meanwhile, glycerol, propylene glycol, and PEG 400 were used as plasticizers.

Results of membrane thickness and water vapor permeability tests are tabulated in Table 2. The formula that used glycerol as the plasticizer had a significantly different membrane thickness than the other formula. Different types of plasticizers are reported to affect the thickness and mechanical properties of the membrane [4,10]. The ideal wound dressing has to be permeable to water vapor that could control the moisture on the surface of the wound [1-3]. According to USP XXIV [21], materials or membranes with water vapor permeability more than 2000 mg/L/day are categorized as permeable. The results in Table 2 show that F1 was the only formula that had water vapor permeability close to 2000 mg/L/day.

The ideal wound dressing should be sturdy but pliable and elastic [9,25,26]. When a wound dressing is applied to the wound, it should not be easily separated and damaged. The results of mechanical properties in Table 3 show that all formulae have good tensile strength and elongation, but F1 has a better Young's modulus value.

Conclusion

This study concludes that the formula F1 that uses glycerol as a plasticizer produced membranes with good water vapor permeability and mechanical properties.

Table 3. Formulae of eel (Monopterus albus) extract membrane

| Ingredients | Formula (%) | Tensile strength (N/mm²) | Elongation at break (%) | Young's modulus (N/mm²) |
|-------------|-------------|--------------------------|-------------------------|-------------------------|
| F1          | 25.96 ± 0.91 | 256 ± 23.2               | 10.20 ± 1.31            |
| F2          | 26.02 ± 0.37 | 315 ± 9.2                | 8.26 ± 0.14             |
| F3          | 24.70 ± 1.02 | 313 ± 15.9               | 7.89 ± 0.59             |

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