Research Article
Influence of Classical Massage on Biochemical Markers of Oxidative Stress in Humans: Pilot Study

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Classical massage is one of the most popular forms of conservative treatment in various diseases. Despite the wide scope of research, the mechanisms of massage are not fully known and understood. Apart from the well-described effects on individual body systems, there are few scientific reports on the effects of massage on the human body at the subcellular level. The study was designed to assess changes in oxidative stress parameters in healthy volunteers after a single session of classical massage. 29 healthy volunteers aged 22.4 ± 3.64 participated in the study. Before and 30 minutes after the massage procedures, blood samples were taken by experienced personnel. Biochemical markers of oxidative homeostasis were assessed with highly specific methods for each parameter: oxidase ceruloplasmin, glutathione, malondialdehyde, glutathione peroxidase, glutathione S-transferase, and superoxide dismutase. The study demonstrates that massage therapy caused statistically significant decrease in the concentration of glutathione peroxidase (red blood cells) and increase in the level of glutathione peroxidase (plasma), superoxide dismutase, and malondialdehyde. In contrast, statistically significant changes in the hematocrit, glutathione, NO2-/NO3-, and oxidase ceruloplasmin were not observed. The results show that complex influence of classical massage therapy on human organism may be reflected in parameters of the oxidative stress. To understand this mechanism clearly, further research is needed.

1. Introduction
Oxygen is involved in many metabolic processes. It plays a role of an oxidant. During the reaction with organic compounds, it collects electrons and undergoes a process of reduction. When reduction process of oxygen molecules is incomplete, reactive oxygen species (ROS) may be formed [1–3]. Reactive oxygen species include neutral molecules or ions and oxygen free radicals, i.e., atoms or molecules that have at least one or more unpaired electrons [4, 5]. Among ROS, the following should be named: alkoxy radical RO, radical ROO peroxide, singlet oxygen O2, ozone O3, hydrogen peroxide H2O2, superoxide anion O2-, and hydroxyl radical OH [6]. ROS easily react with cell elements which may lead to changes in the cell structure and numerous damages. ROS play an important role in maintaining homeostasis: they influence signaling cells or relay cells, providing proper cell metabolism [4, 7]. ROS also participate in functioning of the immune system, excretion of hormones, contraction of muscles, and muscle tone regulation.

There are defense mechanisms within the human organism which guard against ROS activity [4], including,
e.g., antioxidative enzymes: superoxide dismutase (SOD-1), catalase (CAT), glutathione peroxidase (GPx), glutathione reductase (GR), glutathione S-transferase (GST), and glutathione (GSH), vitamin E, vitamin C, bilirubin, ceruloplasmin (CP), uric acid, ferritin, and transferrin. Extensive production of ROS and inefficient action of the antioxidant defense mechanisms cause negative results. Oxidation reaction may be unbalanced which leads to a phenomenon called oxidative stress [8, 9]. The human body may be exposed to numerous environmental factors, such as unhealthy food, alcohol, cigarette smoke, medications, stimulants, air pollution, and pathogenic microorganisms, which may provoke generation of various oxygen forms in the organism [10]. Every prohealth initiative such as physical activity, optimal diet, and physiotherapy may decrease the risk of free radical-related diseases. Numerous studies show the influence of physical activity on oxidative stress parameters but there is a lack of research into the effect of therapeutic massage on oxidative stress parameters [11–13].

Massage therapy still remains a popular and natural way of treatment, constituting part of health prevention. During years of evolution of soft tissue therapy, a lot of different types of massage were developed all over the world. Classical massage, also called Swedish massage, is very a complex and standardized activity which consists in using successive techniques of effleurage, friction, petrissage, tapotement, rolling, shaking, and vibration [14].

Depending on the technique, massage may energize or relax tissues and systems. It acts both locally and, through the central nervous system, globally. Classical massage has an effect on functioning of the human body on various levels, from cells to whole systems, including nervous, circulatory, pulmonary, digestive, urinary, and endocrine system [15, 16]. Thus, the influence of massage on the body can be described as complex and diverse. Literature also frequently describes a decrease in cortisol levels, changes in blood parameters, or an increase in dopamine levels, which has a positive effect on the patient’s well-being [16, 17]. This is explained by the fact that the mechanical stimulus changes the activity of the vagus nerve, putting the patient in a parasympathetic state leading to a regeneration reaction. An increase in the presence of NK cells as well as CD4 is also observed. Similarly, the immunity is positively influenced by chromogranin A which was described by Noto et al. [18]. Hypothesis stating that such complex influence may be reflected in parameters of the oxidative stress can be true. Massage as a mechanical stimulus can potentially cause microtrauma in connective tissue, which may correspond to some of the effects of physical activity and, as a consequence, trigger an antioxidant reaction.

Until now, the evidence for such impact is limited and more concentrated on physical activity or combination of physical activity and massage [19, 20]. In the available literature, no publications that deal exclusively with the topic of the influence of massage on the antioxidative response were found. Therefore, the given study is aimed at assessing the human early antioxidative response to the single classical massage session [11, 19].

2. Material and Methods

2.1. Material. The study group consisted of 29 healthy people (15 females and 14 males) mean age 22.24 ± 3.64 years, with no health conditions (fever, contagious diseases, blood clots, pregnancy, kidney and liver conditions, cancer, inflammations, and hypertension were excluded as a contraindication during general medical examination). The assessed group did not have regular massages previously. Participants were not practicing sport professionally and have no regular medications, addictions, obesity (BMI between 19 and 23), or diabetes. Every person was fasted till the last blood sample collection.

This study was conducted in accordance with the Declaration of Helsinki and approved by the local institutional review board. Freely given written informed consent was obtained from every patient prior to the study.

2.2. Methods. Participants were treated using classical massage therapy in the Department of Physiotherapy, Ludwik Rydygier Collegium Medicum in Bydgoszcz, Poland. Every patient had one session of the massage for at least 30 minutes. The classic massage covered the dorsal part of the torso, including the spinal and dorsal area (lateral part of abdominal oblique muscles, trapezius, rhomboids, latissimus dorsi, and erectors of the spine). The procedure was carried out under repeatable and unified conditions for the whole group, by one experienced physiotherapist with intermediate intensity, without crossing pain barrier. The massage consisted in the use of successive techniques of effleurage, friction, petrissage, tapotement, rolling, shaking, and vibration, in accordance with the standard of their performance. Blood samples were collected twice: before massage and after the massage.

In the experiment, activity of following biochemical markers of oxidative homeostasis was assessed: oxidase ceruloplasmin (CP), glutathione (GSH), malondialdehyde (MDA), glutathione peroxidase (GPx), glutathione S-transferase (GST), and superoxide dismutase (SOD-1).

The material for analysis was venous blood collected in an amount of approx. 8 ml of the antecubital vein into lithium heparin tubes and tubes without anticoagulant. First blood samples were collected in the morning from 8.00 to 9.00 before massage, and second were 30 minutes after procedure. Then, collected material was transported to laboratory. Tests were carried out on the same day in the Department of Biochemistry of Nicolaus Copernicus University Collegium Medicum in Bydgoszcz, within one hour of material collection. From the blood drawn into tubes without anticoagulant (approx. 3 ml) serum was obtained by centrifugation of the material over 5 min at 5000 × g; then it was transferred to Eppendorf tubes and frozen at −80°C.

The prepared serum was stored to determine the activity of the oxidase ceruloplasmin (CP). Before preparing the hemolysate, 500 μl blood was collected to determine the levels of glutathione (GSH) in the erythrocytes, and the remaining aliquot of blood (approx. 5 ml) was centrifuged to obtain plasma, wherein the concentration of nitrate/nitrite was determined. The remaining cells were used for the
preparation of the hemolysate, wherein the dialdehyde malonic concentration (MDA) and the activity of the enzymes glutathione peroxidase (cGPx), glutathione S-transferase (GST), and superoxide dismutase (SOD-1) were determined.

The concentration of reduced glutathione (GSH) was assayed using the Beutler method [21]. The principle of this method is based on the reaction of reduction of the disulfide compound—dithio-bis-2-nitrobenzoic acid (DTNB) by compounds containing sulfhydryl groups. Blood free sulfhydryl groups unrelated to proteins were derived almost only from GSH. The product of the described reaction is a compound of yellow color. Color density was measured at 412 nm. The calculations used the molar absorption coefficient, which, when attached to the mentioned wavelength, is equal to 13.6 [mol⁻¹ × 1 × cm⁻¹]. The results were expressed in mmol/LRBC. The coefficient of variation for this method was 2.4%.

The activity of glutathione peroxidase (GPx) in erythrocytes was assayed by a two-stage Paglia and Valentine method [22]. In the first stage, GPx reacts with tert-butyl peroxide and reduced glutathione (GSH). The product of this reaction is glutathione disulfide (GSSG). The second stage involves the action of glutathione reductase (GR) reducing GSSG to GSH with the participation of NADPH in one minute at 25°C. The length of 545 nm. It is directly proportional to the concentration of GSH. The concentration of malondialdehyde (MDA) in the preparation of the hemolysate, wherein the dialdehyde malonic concentration (MDA) and the activity of the enzymes glutathione peroxidase (cGPx), glutathione S-transferase (GST), and superoxide dismutase (SOD-1) were determined.

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cGPx activity was calculated based on the loss of the reduced form of coenzyme in time (test Wartburg). In the calculations, millimolar absorption coefficient for NADPH at 340 nm, equal to 6.22 [mmol⁻¹ × 1 × cm⁻¹], was used. The results were expressed in U/g Hb, where 1 µmol oxidation of NADPH in one minute at T = 25°C was adopted as a unit of enzyme activity. The coefficient of variation for this method was 2.9%.

Determination of glutathione S-transferase (GST) activity in RBCs was performed according to the method of Habig and Jakob [23]. In this method, there is a decrease in absorbance (which is measured at a wavelength of 340 nm) due to the formation of a conjugate of glutathione (GSH) with 1-chloro-2,4-dinitrobenzene (CDNB). The decrease in absorbance is proportional to the glutathione S-transferase activity. GST activity assay was carried out in the presence of phosphate buffer and CDNB. The results were expressed in nmol/CDNB-GSH/mg Hb/min.

Superoxide dismutase (SOD-1) activity in RBCs was determined using the Misra and Fridovich method, which is based on the inhibition of adrenaline oxidation reaction by superoxide dismutase at pH 10.2 [24]. The increase in absorbance was measured at a wavelength of 480 nm. It is proportional to the increase in the concentration of oxidation products of adrenaline. The activity of SOD-1 was expressed in U/g Hb. The amount of enzyme which inhibits the oxidation of adrenaline 50% was adopted as a U unit. The coefficient of variation for this method is 6.3%.

The concentration of malondialdehyde (MDA) in the erythrocytes was determined by the Placer et al. method, which is based on the reaction of thiobarbituric acid and cer-

3. Discussion

The internal balance of the organism is perceived systemically, but also at the cellular and subcellular level. Lifestyle, leisure activities, improper diet, and application of
stimulants may provoke generation of various oxygen forms in the organism. On the other hand, every pro-health initiative such as physical activity, optimal diet, and physiotherapy (including massage) may decrease the risk of free radical-related diseases. Systemic and local antioxidant systems depend also on age—they are less efficient in children and adolescents than in middle-aged people, which indicates also age-related immaturity of antioxidant mechanisms [29]. Our results confirmed the hypothesis concerning influence of classical massage on selected parameters of oxidative stress, including GPxRBC, GPxp, SOD-1, and MDA.

Integrated and effective human defense system is needed to control the production of free radicals as well as counteract their biological consequences. Extensive production of ROS may be limited/balanced. Aforementioned defense system should consist of both enzymatic and nonenzymatic mechanisms that dispose ROS.

Physiotherapy may play a significant role in this process [30]. Current evidence concerning influence of classical massage on oxidative stress is limited. Study by Karabulut et al. [19] showed that regular physical activities combined with postexercise massage increase SOD-1 activity and do not change adenosine deaminase (ADA). Such outcome

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**Table 1**: Mean values of activity or concentration of the selected parameters of oxidative stress before and after classical massage.

| Parameter          | Before (n=29) | After (n=29) | ±95% CI of change | p value | Cohen’s d |
|--------------------|--------------|--------------|-------------------|---------|-----------|
|                    | Mean (SD)    | Mean (SD)    |                   |         |           |
| HT                 | 43.845 (3.303)| 43.862 (3.359)| 0.112; -0.078     | 0.713   | 0.005     |
| GSH (mmol)         | 2.441 (0.258)| 2.441 (0.261)| 0.020; -0.019     | 0.971   | 0.001     |
| GPx (U)            | 237.797 (38.932)| 282.010 (58.625)| 60.443; 27.985   | 0.001   | 0.888     |
| GPxRBC (U)         | 18.431 (2.358)| 16.945 (2.323)| -0.922; -2.050   | 0.001   | -0.635    |
| GST (nmol)         | 3.007 (0.674)| 2.910 (0.627)| 0.067; -0.260    | 0.236   | -0.149    |
| SOD-1 (U)          | 2648.620 (334.480)| 2844.140 (374.743)| 258.960; 132.074| 0.001   | 0.550     |
| MDA (mmol)         | 0.291 (0.029196)| 0.253 (0.514)| -0.021; -0.053   | 0.001   | -0.102    |
| NO2/NO3 (μmol/L)   | 3.117 (3.267)| 3.240 (3.139)| 0.801; -0.555    | 0.713   | 0.038     |
| CP (IU)            | 1141.676 (391.677)| 1148.334 (375.960)| 44.287; -30.970| 0.720   | 0.017     |

HT: hematocrit; GSH: glutathione; GPx: glutathione peroxidase (plasma); GPxRBC: glutathione peroxidase (red blood cells); GST: glutathione S-transferase; SOD-1: superoxide dismutase; MDA: malondialdehyde; NO2/NO3: nitrate/nitrite; CP: ceruloplasmin oxidase.
supports the assumption that regular physical activity and massage has favorable health effects. Our results are consistent with aforementioned results in SOD-1. However, it should be noted that Karabulut et al. [19] investigated effects of physical exercise alone and in combination with massage, but not massage alone. Moreover, the final outcome was similar regardless of the massage intervention. Analogous results are presented by Heydari et al. who showed that the use of various forms of massage during recovery and rest after exhausting physical exertion may also influence SOD level. [13]

Study by Amano et al. showed that overall health care program, including body massage (cupping and vibration), may significantly improve health status (including oxidative stress) of the preobese or mildly obese middle-aged women. It is difficult to point to the key factor that had impact on health of the examined women. The influence of massage alone was not investigated in the aforementioned findings, yet it may play an important role in comprehensive approach [31].

Presented outcome may support evidence that classical massage may prove helpful in neutralizing radical superoxide by enzymatic defense system [29], which is also confirmed in the given study by the fact that an increased activity of GPx was observed after massage, as compared to the values before the massage. This result is consistent with the results presented by Heydari et al. [13]. Another important parameter of oxidative stress is GST, taking part i.a. in regulation of GPx [32]. Our studies show lack of differences in the GST activity before/after the massage. Similar outcome is observed by Ciechomski et al. [33] who assess GPx and GSH in patients relaxing after training (passively or undergoing massage). The effect of the damages caused by oxidative stress may be also measured indirectly using MDA [34]. MDA concentration increases with extensive production of ROS in the organism [35]. Our study showed a decrease in MDA level after massage compared to the value before the massage. Similar results were presented by Heydari et al. and Karabulut et al. [13, 19]. NOx also play an important role as an element of antioxidant barrier of the organism [36, 37]. Our study has failed to show a significant impact of the massage. The outcome is similar to the one showed by Karabulut et al. [19]. Another important parameter of oxidative stress is CP [38]. We have not observed significant differences in CP level before/after massage.

The mechanism of the observed reaction may be due to the microdamage of the muscles after the massage. Activation of monocytes, increasing the expression of inflammatory cytokines leads to the activation of endothelial leukocyte adhesion molecules P-selectin and E-selectin. Neutrophils intensify the breakdown of tissue, leading to the release of free radicals in the environment and, consequently, a change in the studied parameters [39].

Main limitations of our study included a relatively small group of volunteers and no placebo group; thus, we regard our study as preliminary. Only immediate results of the massage therapy on selected biochemical parameters of the oxidative stress were assessed during the study. Long-term results will be assessed during further studies. Moreover, we assessed only the influence of the single session of the therapeutic massage. Cumulative effect of the subsequent massage therapy sessions may be stronger or longer-lasting.

Further studies require randomized controlled trials on a bigger number of volunteers, as well as taking into consideration such factors as sex, age, previous lifestyle, diet, and body mass index. Such detailed approach would allow assessment of possible prognostic signs and provide scenarios for prohealth prevention in various groups of patients.

Other parameters of oxidative stress may be used in treating disease-related complications as well as in the
interdisciplinary treatment. Intensity of the massage (and associated physical exercises), its duration, and used techniques may also influence development of the oxidative stress and its long-term consequences. The ultimate goal is optimization of classical massage in prevention and treatment of oxidative stress in both healthy and ill people, including elderly patients [40–43].

**Abbreviations**

- CAT: Catalase
- CP: Oxidase ceruloplasmin
- Gpx: Glutathione peroxidase
- Gpxpl: Glutathione peroxidase (plasma)
- GpxRBC: Glutathione peroxidase (red blood cells)
- GR: Glutathione reductase
- GSH: Glutathione
- GST: Glutathione S-transferase
- H2O2: Hydrogen peroxide
- HT: Hematocrit
- MDA: Malondialdehyde
- NO2: Nitrate
- NO3: Nitrite
- O2−: Superoxide anion
- O3: Ozone
- OH: Hydroxyl radical
- RO: Alkoxy radical
- ROO: Radical peroxide
- ROS: Reactive oxygen species
- SD: Standard deviation
- SOD-1: Superoxide dismutase

**Data Availability**

The biochemistry data used to support the findings of this study are included within the article.

**Conflicts of Interest**

The authors declare that they have no conflicts of interest.

**References**

[1] T. Rahman, I. Hosen, M. M. T. Islam, and H. U. Shekhar, “Oxidative stress and human health,” *Advances in Bioscience and Biotechnology*, vol. 3, no. 7, pp. 997–1019, 2012.

[2] A. J. Gow, D. Duran, S. Malcolm, and H. Ischiropoulos, “Effects of peroxynitrite-induced protein modifications on tyrosine phosphorylation and degradation,” *FEBS Letters*, vol. 385, no. 1–2, pp. 63–66, 1996.

[3] E. R. Stadtman and R. L. Levine, “Protein oxidation,” *Annals of the New York Academy of Sciences*, vol. 899, pp. 191–208, 2000.

[4] S. H. Abtahi-Eivari, M. Moghimian, M. Soltani et al., “The effect of Galega officinalis on hormonal and metabolic profile in a rat model of polycystic ovary syndrome,” *International Journal of Women’s Health and Reproduction Sciences*, vol. 6, no. 3, pp. 276–282, 2018.

[5] G. Ray and S. A. Husain, “Oxidants, antioxidants and carcinogenesis,” *Indian Journal of Experimental Biology*, vol. 40, no. 11, pp. 1213–1232, 2002.

[6] M. Bonnefoj, J. Drai, and T. Kostka, “Antioxidants to slow aging, facts and perspectives,” *Presse Médicale*, vol. 31, no. 25, pp. 1174–1184, 2002.

[7] P. D. Ray, B. W. Huang, and Y. Tsuji, “Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling,” *Cellular Signalling*, vol. 24, no. 5, pp. 981–990, 2012.

[8] M. Shokoohi, E. Olad Saheb Madarek, A. Khaki et al., “Investigating the effects of onion juice on male fertility factors and pregnancy rate after testicular torsion/detorsion by intrauterine insemination method,” *International Journal of Women’s Health and Reproduction Sciences*, vol. 6, no. 4, pp. 499–505, 2018.

[9] S. Vertuani, A. Angusti, and S. Manfredini, “The antioxidants and pro-antioxidants network: an overview,” *Current pharmaceutical design*, vol. 10, no. 14, pp. 1677–1694, 2004.

[10] A. Bokov, A. Chaudhuri, and A. Richardson, “The role of oxidative damage and stress in aging,” *Mechanisms of Ageing and Development*, vol. 125, no. 10–11, pp. 811–826, 2004.

[11] C. V. De Sousa, M. M. Sales, T. S. Rosa, J. E. Lewis, R. V. de Andrade, and H. G. Simões, “The antioxidant effect of exercise: a systematic review and meta-analysis,” *Sports Medicine*, vol. 47, no. 2, pp. 277–293, 2017.

[12] A. Thirupathi, R. A. Pinho, and Y. Z. Chang, “Physical exercise: an inducer of positive oxidative stress in skeletal muscle aging,” *Life Sciences*, vol. 252, 2020.

[13] B. Heydari, M. Ghofrani, and M. E. Bahram, “Effects of three types of massage on serum levels of malondialdehyde, superoxide dismutase and glutathione peroxidase after one session of exhaustive exercise in female futsal players,” *Complementary Medicine Journal*, vol. 10, no. 4, pp. 328–339, 2021.

[14] K. Kassolik, W. Andrzejewski, M. Brzozowski et al., “Comparison of massage based on the tensegrity principle and classical massage in treating chronic shoulder pain,” *Annals of Complementary and Alternative Medicine*, vol. 7, no. 10, pp. 418–427, 2013.

[15] M. Cassar, *Handbook of Clinical Massage: A Complete Guide for Students and Professionals*, Elsevier, Edinburgh, 2004.

[16] T. Field, “Massage therapy research review,” *Complementary Therapies in Clinical Practice*, vol. 24, pp. 19–31, 2016.

[17] A. Moraska, R. A. Pollini, K. Boulanger, M. Z. Brooks, and L. Teitlebaum, “Physiological adjustments to stress measures following massage therapy: a review of the literature,” *Evidence-based Complementary and Alternative Medicine*, vol. 7, no. 4, 418 pages, 2010.

[18] Y. Noto, M. Kudo, and K. Hirota, “Back massage therapy promotes psychological relaxation and an increase in salivary chromogranin A release,” *Journal of Anesthesia*, vol. 24, no. 6, pp. 955–958, 2010.

[19] A. B. Karabulut, M. E. Kafkas, A. S. Kafkas, Y. Onal, and T. R. Kiran, “The effect of regular exercise and massage on oxidant and antioxidant parameters,” *Indian Journal of Physiology and Pharmacology*, vol. 57, no. 4, pp. 378–383, 2013.

[20] M. C. Polidori, P. Mecucci, A. Cherubini, and U. Senin, “Physiological activity and oxidative stress during aging,” *International Journal of Sports Medicine*, vol. 21, no. 3, pp. 154–157, 2000.

[21] E. Beutler, “Red Cell Metabolism,” in *A Manual of Biochemical Methods*, E. Beutler, Ed., pp. 11-12, Grune-Stratton, New York, NY, USA, 1971.
[22] U. Reiss and D. Gershon, “Comparison of cytoplasmic superoxide dismutase in liver, heart and brain of aging rats and mice,” *Biochemical and Biophysical Research Communications*, vol. 73, no. 2, pp. 255–262, 1976.

[23] W. H. Habig and W. B. Jakoby, “[51] Assays for differentiation of glutathione S-Transferases,” *Methods in Enzymology*, vol. 77, pp. 398–405, 1981.

[24] H. Misra and P. J. Fridovich, “The role of superoxide anion in the autoxidation of epinephrine and a simple assay for superoxide dismutase,” *Journal of Biological Chemistry*, vol. 247, no. 10, pp. 3170–3175, 1972.

[25] J. W. Rowe and R. L. Kahn, *Successful Aging. Healthy Aging. Challenges and Solutions*, K. Dychtwald, Ed., Aspen Publishers, Inc., Gaithersburg, MD, USA, 1999.

[26] P. Griess, “Bemerkungen zu der abhandlung der HH. Weselky und Benedikt. Ueber einige azoverbindungen,” *Berichte der Deutschen Chemischen Gesellschaft*, vol. 12, no. 1, pp. 426–428, 1879.

[27] H. A. Ravin, “An improved colorimetric enzymatic ceruloplasmin assay,” *Journal of Laboratory and Clinical Medicine*, vol. 58, pp. 161–168, 1961.

[28] W. G. Hopkins, “A new view of statistics. Internet Society for Sport Science,” 2020, http://www.sportsci.org/resource/stats/.

[29] M. Maciejczyk, A. Zalewska, and J. Ładny, “Salivary antioxidant barrier, redox status, and oxidative damage to proteins and lipids in healthy children, adults, and the elderly,” *Oxidative Medicine and Cellular Longevity*, vol. 20, Article ID 4393460, 12 pages, 2019.

[30] K. Fisher-Wellman and R. J. Bloomer, “Acute exercise and oxidative stress: a 30 year history,” *Dynamic Medicine*, vol. 8, no. 1, 2009.

[31] S. Amano, H. Tsubone, M. Hanafusa, T. Yamasaki, S. Nishizaka, and H. Yanagisawa, “Physical and physiological effectiveness of an overall health care program for middle-aged Japanese women with mild obesity: a pilot study,” *Journal of Traditional and Complementary Medicine*, vol. 5, no. 2, pp. 88–95, 2015.

[32] C. E. Pippenger, R. W. Browne, and D. Armstrong, “Regulatory antioxidant enzymes,” in *Free Radical and Antioxidant Protocols*, D. Armstrong, Ed., Humana Press, New Jersey, 1998.

[33] J. Ciechomski, J. Karolkiewicz, Z. Kasprzak, and L. Szczesniak, “The impact of classical massage on oxidative stress induced by extreme physical exertion,” *Fizjoterapia Polska*, vol. 4, no. 2, pp. 167–175, 2004.

[34] F. Cacciapuoti, “Oxidative stress as “mother” of many human diseases at strong clinical impact,” *Journal of Cardiovascular Medicine and Cardiology*, vol. 3, pp. 1–6, 2014.

[35] D. R. Janero, “Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury,” *Free Radical Biology and Medicine*, vol. 9, no. 6, pp. 515–540, 1990.

[36] V. B. O’Donnell, P. H. Chumley, N. Hogg, A. Bloodworth, V. M. Darley-Usmar, and B. A. Freeman, “Nitric oxide inhibition of lipid peroxidation: kinetics of reaction with lipid peroxyl radicals and comparison with alpha-tocopherol,” *Biochemistry*, vol. 36, no. 49, pp. 15216–15223, 1997.

[37] E. C. Opara, “Oxidative stress,” *Disease-a-Month*, vol. 52, no. 5, pp. 183–198, 2006.

[38] V. W. Bunker, “Free radicals, antioxidants and ageing,” *Medical Laboratory Sciences*, vol. 49, no. 4, pp. 299–312, 1992.

[39] C. Waters-Banker, E. E. Dupont-Versteegden, P. H. Kitzman, and T. A. Butterfield, “Investigating the mechanisms of massage efficacy: the role of mechanical immunomodulation,” *Journal of Athletic Training*, vol. 49, no. 2, pp. 266–273, 2014.

[40] K. Porzych, B. Augustynska, M. Porzych et al., “Change of the state of the natural antioxidant barrier of a body and psychological parameters in patients aged above 60,” *Oxidative Medicine and Cellular Longevity*, vol. 2017, Article ID 6568501, 9 pages, 2017.

[41] H. Çalış-Atabek, S. Demir, R. D. Pinarbaşı, and N. Gündüz, “Effects of different resistance training intensity on indices of oxidative stress,” *Journal of Strength and Conditioning Research*, vol. 24, no. 9, pp. 2491–2497, 2010.

[42] R. Aslan, M. R. Şekeroglu, M. Tarakçiloglu, F. Bayiroglu, and I. Meral, “Effect of acute and regular exercise on antioxidative enzymes, tissue damage markers and membrane lipid peroxidation of erythrocytes in sedentary students,” *Turkish Journal of Medical Sciences*, vol. 28, pp. 411–414, 1998.

[43] K. Pawlak-Osinska, H. Kazmierczak, M. Marzec et al., “Assessment of the state of the natural antioxidant barrier of a body in patients complaining about the presence of tinnitus,” *Oxidative Medicine and Cellular Longevity*, vol. 2018, Article ID 1439575, 8 pages, 2018.