A 67-year-old woman with recurrent hypoglycemia: non-islet cell tumour hypoglycemia

**THE CASE:** A 67-year-old woman was brought to hospital by her husband, who had found her sweaty and unresponsive in the morning. She was hypoglycemic, with a plasma glucose level of 1.2 mmol/L (normal range 3.8–6.1 mmol/L). After receiving dextrose intravenously, she regained consciousness and her plasma glucose level increased to 3.8 mmol/L. Apart from a recent dry cough, she was otherwise well and taking no medications. She was a known carrier of hepatitis B virus. A physical examination was notable for reduced air entry and percussion dullness over the left side of her chest. The results of initial investigations, including electrolyte levels, liver function tests and complete blood count, were all normal. She was admitted to hospital, where she experienced 2 further episodes of spontaneous hypoglycemia; serum insulin levels were undetectable on both occasions. A short Synacthen test (whereby a plasma cortisol level is checked 30 minutes after tetracosactrin is given) showed an adequate rise in cortisol concentration, which ruled out a hypoadrenal state. Chest radiography showed a large opacity over the left hemithorax (Fig. 1). A CT scan confirmed a large solid mass in the left pleural space causing compressive collapse of the left lung, but the patient’s pancreas appeared normal (Fig. 2, white arrow). A needle biopsy of the pleural mass suggested a fibrous tumour, and the specimen stained positively for CD34, raising the suspicion of non-islet cell tumour hypoglycemia secondary to a pleural fibroma. Her insulin-like growth factor (IGF)-I and total IGF-II levels were normal at 10.6 (normal range 5.0–22.5) nmol/L and 111.9 (normal range 55–150) nmol/L respectively. An elevated level of “big” IGF-II (75 [normal < 14] nmol/L) and elevated IGF-II/IGF-I ratio (11.3 [normal < 10]) confirmed the diagnosis.

The patient declined surgical resection, and long-term steroid treatment was felt to be contraindicated in view of her hepatitis B carrier status. A 2-week trial of the somatostatin analogue octreotide resulted in a slight reduction in the frequency of hypoglycemic episodes but no improvement in her biochemical profile, with “big” IGF-II levels elevated at 100 nmol/L, IGF-II/IGF-I ratio at 16.3 and plasma glucose at 6.9 mmol/L, and insulin levels suppressed to less than 10 pmol/L and C peptide levels to less than 94 pmol/L. She agreed to undergo surgery for removal of the mass. The 2285-g encapsulated tumour was confirmed histologically to be a pleural fibroma. After surgery her biochemical IGF levels returned to normal, and, after 8 days of unremarkable recovery, the patient was discharged. She has had no further hypoglycemic episodes 5 years later.
IGF-II is produced by many different tissues. Insulin, IGF-I and IGF-II each have their own receptors, and they bind to each others’ receptors with varying affinity. IGF-I, a 70-amino acid protein, has roles in skeletal and cartilage development. It exists at about half the circulating plasma levels of IGF-II, a 67-amino acid compound whose exact physiologic role is not well defined. So-called “big” IGF-II, a higher-molecular-weight form of IGF-II, represents only 10%–15% of the total circulating IGF in normal human serum. In the normal state, 70%–80% of circulating IGF is bound to insulin-like growth factor binding protein-3 (IGFBP-3) and a protein called acid labile subunit to form an inactive 150-kD complex. The sequestration of IGF forms in these complexes prevents them from causing hypoglycemia.

Some tumours may over-produce “big” IGF-II. The initiating event appears to involve sudden overexpression of the prepro-IGF-II gene by tumour cells. Pro-IGF-II from tumour cells fails to undergo the usual glycosylation process and consequently is only partly processed, resulting in “big” IGF-II. In patients harbouring such non-islet cell tumours, “big” IGF-II represents about 77% of the total IGF. “Big” IGF-II does not bind well to IGFBP-3, so there are fewer IGF–IGFBP complexes available to associate with acid labile subunit to form the normal 150-kD complex. Instead, “big” IGF-II forms a smaller 50-kD complex, which leaves more IGF-II available. This in turn leads to hypoglycemia and the biochemical abnormalities observed among these patients (Fig. 3).1

The diagnosis of non-islet cell tumour hypoglycemia can be confirmed by a combination of suppressed serum insulin levels and suppressed C peptide and growth hormone concentrations in the setting of hypoglycemia, along with elevated “big” IGF-II levels. A total IGF-II/IGF-I ratio of greater than 10 is felt to be diagnostic.2 A description of how “big” IGF-II levels are measured is included in an online appendix (www.cmaj.ca/cgi/content/full/173/359/DC1).

Glucagon increases hepatic glucose production and can be used for the short-term management of non-islet cell tumour hypoglycemia. Long-term treatment requires a continuous intravenous infusion. Growth hormone increases the formation of IGF-II–IGFBP complexes3 but often requires supraphysiologic doses, which may lead to volume overload. In view of the large tumour mass present, which was causing significant mediastinal shift, growth hormone was felt to be relatively contraindicated in our patient. Somatostatin receptors are present in a variety of neuroendo-
crine tumours, and the use of somatostatin analogues can result in reduced secretion by these tumours. Somatostatin receptors have previously been demonstrated in a pleural fibroma causing non-islet cell tumour hypoglycaemia, and this provided the rationale for octreotide therapy for our patient. Octreotide may have failed to suppress “big” IGF-II production because somatostatin receptors on the tumour were non-functional.4 Glucocorticoids can suppress tumour secretion of IGF-II, thus reversing the hypoglycaemia.4 The treatment of choice is tumour resection, which leads to normal glucose concentrations and biochemical marker levels.

Ronald C.W. Ma
Peter C.Y. Tong
Juliana C.N. Chan
Clive S. Cockram
Department of Medicine and Therapeutics

Michael H.M. Chan
Department of Chemical Pathology
Chinese University of Hong Kong
Prince of Wales Hospital
Shatin, Hong Kong

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Box 1: Causes of recurrent hypoglycemia

- Medications
  - insulin
  - sulfonylureas (and interactions with medications such as gatifloxacin)
  - meglitinides (e.g., repaglinide, nateglinide)
  - salicylates
  - pentamidine
  - quinine
  - trimethoprim–sulfamethoxazole
- Alcohol
- Endocrine disturbances
  - adrenal insufficiency
  - hypopituitarism (e.g., postsurgical, post-radiotherapy, hypophysitis, tumours such as craniopharyngioma)
- Hepatic failure
- Renal failure
- Sepsis
- Inborn errors of metabolism (e.g., glycogen storage diseases)
- Autoimmune conditions
  - antibodies to insulin receptors (augmenting insulin’s effect) or antibodies that inhibit circulation of other anti-insulin antibodies
- Tumours
  - islet cell tumours (secreting insulin)
  - non-islet cell tumours (often acting through insulin-like growth factor-II secretion) including mesenchymal tumours, fibromas, carcinoids, myelomas, lymphomas as well as hepatocellular, colorectal, lung, adrenocortical and renal cell carcinomas

Clinical trial registration

CMAJ will consider clinical trials for publication only if they have been registered in a publicly accessible clinical trials registry before the enrolment of the first patient. This policy applies to trials that start recruiting on or after July 1, 2005. For trials that began enrolment before this date, registration is required by Sept. 13, 2005. The criteria for acceptable registration are described in CMAJ (2005;172[13]:1700-2).