Advances in the chemistry, pharmacological diversity, and metabolism of 20(R)-ginseng saponins

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ABSTRACT

Ginseng has been used as a popular herbal medicine in East Asia for at least two millennia. However, 20(R)-ginseng saponins, one class of important rare ginsenosides, are rare in natural products. 20(R)-ginseng saponins are generally prepared by chemical epimerization and microbial transformation from 20(S)-isomers. The C20 configuration of 20(R)-ginseng saponins are usually determined by 13C NMR and X-ray single-crystal diffraction. 20(R)-ginseng saponins have anticancer, antioxidative, antifatigue, neuroprotective, and osteoclastogenesis inhibitory effects, among others. Owing to the chemical structure and pharmacological and stereoselective properties, 20(R)-ginseng saponins have attracted a great deal of attention in recent years. In this study, the discovery, identification, chemical epimerization, microbial transformation, pharmacological activities, and metabolism of 20(R)-ginseng saponins are summarized.

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1. Introduction

Ginseng, a functional food and health-enhancing supplement, is a global herb and has been shown to have extensive range of pharmacological effects on cognition and blood circulation, as well as antitumor, antioxidative, and antifatigue effects, among others [1]. Many of its components, such as ginsenosides, polysaccharides, peptides, polyacetylenic alcohols, and fatty acids, have been separated and characterized. Ginsenosides are common pharmacological components in ginseng herbs, named after the originating countries, for example Panax ginseng, P. japonicas, P. notoginseng, P. quinquefolius, and P. vietnamensis [2]. In 1854, Garriques [3] conducted a chemical study on ginseng and separated a saponin fraction from P. quinquefolium L. for the first time. Despite these findings, the saponin fraction, which contained so-called ginsenosides, was not isolated and identified again until 1963 [4].

So far, more than 100 different ginsenosides with different pharmacological activities have been separated and identified from the root of red ginseng (P. ginseng). Red ginseng is prepared by steaming fresh ginseng at 90–100 °C for a reasonable time and then dried until the moisture content is less than 15% [5]. The amounts of some ginsenosides, such as Rb1, Rc, Rb2 and Rd, in red ginseng are higher than those in fresh ginseng [6], and they have biological functions including the amelioration of various disease symptoms via antioxidant mechanisms in cells and animals (especially in rodents) [7]. Meanwhile, 20(R)-ginseng saponins have been found in P. ginseng. Kitagawa et al [9] isolated 20(R)-Rh1 (4) and 20(R)-Rg2 (5) as characteristic constituents of Ginseng Radix Rubra in 1983. 20(R)-Rh2 (1) was first isolated from P. ginseng leaves and identified in 1987 by Chen et al [10].

The C20 configuration confirmation of ginsenosides had become the focus of this research field because they might play a vital role in pharmacological activities. Compared with other ginsenosides, 20(R)-ginseng saponins have better pharmacological activities [8], such as antitumor [11,12], antioxidative [13], antifatigue [14], neuroprotective [15], and osteoclastogenesis inhibitory effects [16]. Li et al [13] found that 20(R)-Rg3 (2) exhibited better antioxidative activity in vitro than 20(S)-Rg3. In addition, 20(R)-Rh2 (1), but not...
20(S)-Rh2, is a selective osteoclastogenesis inhibitor without any cytotoxicity [16]. Thus, chemical structure, pharmacological action, and metabolism of 20(R)-ginseng saponins have aroused interest. In this article, the discovery, structure identification, chemical epimerization, microbial transformation, pharmacological activities, and metabolism of 20(R)-ginseng saponins are summarized.

2. Discovery of 20(R)-ginseng saponins

20(R)-dammarane-type saponins are generally classified into protopanaxadiol (PPD) type and protopanaxatriol (PPT) type. The PPD-type saponin involves the attachment of saccharide(s) to C3 and/or C20, and the PPT-type saponin involves the attachment of saccharide(s) to C3, C6, and/or C20. The ocottillol-type saponin is a tetracyclic triterpenoid, containing a tetrahydrofuran ring [17]. The structures of all 20(R)-ginseng saponins are shown in Fig. 1.

2.1. 20(R)-PPD-type saponins

20(R)-Rh2 (1, 40 mg) was first isolated from the stems and leaves of P. ginseng Meyer (100 kg) by Chen et al. [10]. It [mp. 288–290°C; δ1H 4.11, δ13C (MeOH)] was identified by 1H NMR, 13C NMR, and MS and compared with the reported 20(S)-Rh2 13C NMR data [18–20].

20(R)-Rg3 (2, 1.5 g, mp. 298–300°C) and 20(R)-PPD (3, 8 mg, mp. 243–245°C) were isolated from the root, stems, and leaves of P. ginseng (5 kg) by incision during the steam process [21,22]. They found that Compound 3 was generated by steaming fresh ginseng, and the content was higher than that in red ginseng. The content was also higher in fine roots than that in main roots. These structures were identified by 1H NMR, 13C NMR, and electrospray ionization mass spectrometry (ESI-MS) [18–20,23].

2.2. 20(R)-PPT-type saponins

20(R)-Rh1 (4, 900 mg, mp. 217–219°C) was first separated and purified from P. ginseng Meyer (5 kg) by silica gel column chromatography (CHCl3/MeOH/H2O, 6:1:0.1) and solid-phase extraction high performance liquid chromatography (SP-HPLC, MeOH/H2O, 70:30) [21]. Wang et al. [24] obtained Compound 4 from the root of P. notoginseng (Burk.) F. H. Chen (Araliaceae) by steaming and baking. Compound 4 (35 mg) and 20(R)-Rg2 (5, 16 mg) from Gynostemma yixingense (100 g) were subjected to silica gel column chromatography (CHCl3/MeOH, 20:1 to 1:1) [20]. Their structures were determined by spectroscopic analyses, including 1D-NMR, 2D-NMR, and ESI-MS [18,19].

20(R)-RZ (6, 6 mg) and 20(R)-PPT (7, 120 mg, mp. 259–261°C) were isolated from the stems and leaves of P. ginseng (5 kg) and purified by silica gel column chromatography (CHCl3/MeOH/H2O, 3:1:0.1) by Xu et al. [25] and Yang et al. [21]. Their structures were identified by IR, field desorption-mass spectrometry (FD-MS), and comparison with the 20(S)-isomer's 13C NMR data [26,27].

2.3. Ocottillol-type saponins

20(R)-PF11 (32, 11 mg) was obtained from American red ginseng (2.5 kg), and novel structures will likely continue to be reported with the aid of modern and more sensitive characterization techniques. Its structure was elucidated as (20R,24R)-dammar-20, 24-epoxy-3α, 6α,12β, 25-tetraol [28,29].

3. Identification of 20(R)-ginseng saponins

3.1. 13C NMR analysis

NMR plays a vital role in the structural elucidation of 20(R/S)-ginseng saponins. The obvious identifiers are the chemical shift values of C20 and its contiguous C17, C21, and C22.

Asakawa et al. [30] found that the chemical shift values of C17, C21, and C22 of 20(R)-ginseng saponins were ca. 55, 27, and 35 ppm, respectively, distinctively different from those of the 20(R)-isomers (ca. 50, 22, and 43 ppm). In addition, changes in chemical shifts between the 5 form and R form at C17, C21, and C22 in 13C NMR spectra were approximately △δ (δR–δS) = –1.1 ± 0.4, –4.3 ± 0.1, and +7.4 ± 0.1 ppm, respectively [31] (Table 1).

The observed chemical shift values of each compound are within the ranges mentioned previously, which permits the assignment of the C20 configuration as 5 form or R form.

3.2. Single-crystal X-ray diffraction

The C20 configuration of 20(R/S)-ginseng saponins could also be determined by single-crystal X-ray diffraction analysis. In our previous study, Compounds 3 and 7 were prepared from P. quinquefolium L. with citric acid and sodium hydrate in glycerol, successively. (20R,24S)-epoxy-3-acetyldammar-3β,12β,25-triol (33), (20R,24R)-epoxy-3-acetyldammar-3β,12β,25-triol (34), (20R,24S)-20,24-epoxy-3-oxo-12β,25-dihydrodammarane (39), and (20R,24R)-20,24-epoxy-3-oxo-12β,25-dihydroxy-dammarane (40) have been synthesized from Compound 3. Their structures were elucidated by spectral studies, and the configurations were confirmed by single-crystal X-ray diffraction [32,33] (Fig. 2).

4. Chemical epimerization and microbial transformations of 20(S)-ginseng saponins

20(R)-ginseng saponins are rare in natural products. They are mainly obtained by chemical epimerization and microbial transformation from their corresponding 20(S)-isomers.

4.1. Chemical epimerization with acid

The C20 configuration of 20(S)-ginseng saponins can be epimerized with acid via an S41 mechanism (Fig. 3). The common acids mainly include dl-tartaric acid, citric acid, lactic acid, acetic acid, etc.

Since Compound 2 was extracted from red ginseng in 1980, researchers have found that it is more effective than 20(S)-Rg3 in exhibiting antitumor [8], antioxidative [62], antifatigue [14], neuroprotective [58], and osteoclastogenesis inhibitory effects [16], among others. Therefore, the extraction, synthesis, and evaluation of Compound 2 were prioritized, and the yield was gradually increasing in different optimized conditions. Sun et al. [35] obtained Compound 2 at 50% yield by hydrolyzing PPD-type saponins in optimized conditions (dl-tartaric acid, 10 mol/L, 110°C and 2.5 h). Yao et al. [36] also acquired Compound 2 from major ginsenosides with a confined microwave–promoted degradation method. The actual Compound 2 yield was predicted to be 94.52% in specific conditions (dl-tartaric acid, 1.19 mol/L, 107.9°C and 2.79 h) through canonical analysis with maximum responses [37].

Compounds 3 and 4 were isolated from the acid hydrolysis product of P. quinquefolium L. for the first time by Ma and Yang [38] and Ma et al. [39]. Compound 3 (40 g, mp. 244–245°C) had also been obtained from P. quinquefolium L. (120 g) with 50% citric acid and deglycosylated with sodium hydrate in glycerol in our laboratory. These structures were identified by 1H NMR and 13C NMR [32,40].
Fig. 1. The structures of 20(R)-ginseng saponins. (A) Protopanaxadiol (PPD)- and protopanaxatriol (PPT)-type saponins; (B) Panaxadiol (PD)-type, panaxatriol (PT)-type, and modified saponins; (C) Modified PPD- and PPT-type saponins; (D) Ocotillol-type and modified saponins.
20(R)-RF (9) was epimerized from ginsenoside RF by citric acid (pH = 3.5, 90°C, 45 min) and identified by ultra high performance liquid chromatography/time of flight mass spectrometry (UPLC/TOFMS) in red ginseng [30,34]. The ginsenoside RF has higher chemical stability than other ginsenosides in acid environments.

### Table 1
Chemical shift of C-17, C-20, C-21, and C-22 in 20(R)-ginseng saponins (in pyridine- d6).

| Ingredient name | C-17 | C-20 | C-21 | C-22 |Refs |
|-----------------|------|------|------|------|-----|
| 20(R)-Rh2       | 52.2 | 73.4 | 23.0 | 43.7 | [9] |
| 20(S)-Rh2       | 54.8 | 72.9 | 26.9 | 35.2 |     |
| Δ320(R)-20(S)   | -2.6 | +0.5 | -3.9 | +8.5 |     |
| 20(R)-Rg3       | 50.7 | 73.0 | 22.8 | 43.3 | [9] |
| 20(S)-Rg3       | 54.8 | 73.0 | 27.1 | 35.9 |     |
| Δ320(R)-20(S)   | -4.1 | +0.0 | -4.3 | +2.6 |     |
| 20(R)-PPD       | 50.7 | 73.0 | 22.6 | 43.2 | [9] |
| 20(S)-PPD       | 54.9 | 73.0 | 25.8 | 35.3 |     |
| Δ320(R)-20(S)   | -4.2 | +0.0 | -3.2 | +7.9 |     |
| 20(R)-Rh1       | 50.6 | 73.0 | 23.0 | 43.3 | [9] |
| 20(S)-Rh1       | 54.6 | 73.0 | 26.9 | 35.9 |     |
| Δ320(R)-20(S)   | -4.0 | +0.0 | -3.9 | +7.4 |     |
| 20(R)-Rg2       | 40.7 | 73.0 | 22.7 | 43.2 | [15,16] |
| 20(S)-Rg2       | 46.5 | 72.9 | 26.9 | 35.7 |     |
| Δ320(R)-20(S)   | -4.9 | +0.1 | -4.2 | +7.5 |     |
| 20(R)-RZ2       | 51.7 | 73.4 | 22.5 | 43.6 | [9] |
| 20(S)-RZ2       | 54.7 | 72.5 | 27.2 | 36.4 |     |
| Δ320(R)-20(S)   | -3.0 | +0.9 | -4.7 | +7.2 |     |
| 20(R)-PPT       | 50.7 | 73.0 | 22.6 | 43.3 | [9] |
| 20(S)-PPT       | 54.8 | 73.0 | 27.1 | 35.9 |     |
| Δ320(R)-20(S)   | -4.1 | +0.0 | -4.5 | +7.4 |     |
| 20(R)-PF1       | 50.5 | 86.3 | 19.3 | 38.2 | [25,26] |
| 20(S)-PF1       | 48.8 | 87.3 | 28.8 | 31.5 |     |
| Δ320(R)-20(S)   | +1.7 | -1.0 | -9.5 | +6.7 |     |

PDD, protopanaxadiol; PPT, protopanaxatriol.

### Table 2
Summary of acid transformation to 20(R)-ginseng saponins

| Entry | Ingredient name | Substrates | Transformation condition | Refs |
|-------|-----------------|------------|--------------------------|-----|
| 2     | 20(R)-Rg3       | PDD saponins | D, L-tartaric acid       | [35,36] |
| 3     | 20(R)-PPD       | P. ginseng and P. quinquefolium | Sulfuric acid in alcoholic solution (5%) and citric acid (50%) | [32,38] |
| 4     | 20(R)-Rh1       | P. quinquefolium L. | Sulfuric acid in alcoholic solution (5%) | [38,39] |
| 7     | 20(R)-PPT       | P. ginseng and P. quinquefolium | Sulfuric acid in alcoholic solution (5%) and concentrated hydrochloric acid | [38,41] |
| 9     | 20(R)-RF        | Ginsenoside RF | Citric acid (pH 3.5) | [30,34] |
| 10    | 20(R)-PD        | P. ginseng | Sulfuric acid in alcoholic solution (5%) and hydrochloric acid (7%) | [39] |
| 11    | 20(R)-PT        | P. ginseng | Sulfuric acid in alcoholic solution (5%) and hydrochloric acid (7%) | [39] |
| 12    | 20(R)-25-epoxy-3β-acetoxy-12β-hydroxydammarane | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 13    | 20(R)-25-epoxy-3β,6α-diacetoxy-12β-hydroxydammarane | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 14    | 20(R)-25-epoxy-3β-acetoxy-6α,12β-dihydroxydammarane | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 15    | 20(R)-25-epoxy-3α,20-acetoxy-3β,12β-dihydroxydammarane | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 16    | 20(R)-25-epoxy-3β-acetoxy-6α,12β-dihydroxydammarane | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 17    | 20(R)-25-epoxy-3α,6β,12β-trihydroxydammar-6-0,6-β-D-glucopyranoside | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 18    | 20(R)-25-epoxy-3-0,6β,12β-dihydroxydammarane | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 26    | 20(R)-dammar-3β,12β,20,25-tetrol | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 27    | 20(R)-dammar-3α,6β,12β,20,25-tetrol | P. ginseng | Sulfuric acid in alcoholic solution (5%) | [39] |
| 28    | 20(R)-25-methoxydammar-3β,12β,20,25-tetrol | P. ginseng | Hydrochloric acid (18%) | [48] |
| 29    | 20(R)-25-methoxydammar-3β,6β,12β,20,25-tetrol | P. ginseng | Hydrochloric acid (18%) | [48] |
| 30    | 20(R)-20-methoxydammar-3β,12β,20,25-triol | P. ginseng | Hydrochloric acid (18%) | [48] |
| 31    | 20(R)-25-acetyldammar-3β,12β,20,25-triol | P. notoginseng | Concentrated hydrochloric acid | [50] |
| 32    | 20(R)-25-acetyldammar-3β,6α,12β,20,25-triol | P. quinquefolium L. | Citric acid (50%) | [32] |
| 33    | 20(R)-25-acetyldammar-3β,6β,12β,20,25-triol | P. quinquefolium L. | Citric acid (50%) | [32] |
| 34    | 20(R)-24β-epoxy-3-acetyldammar-3β,12β,20,25-triol | P. ginseng | Alcoholic solution of sulfuric acid (5%) and concentrated sulfuric acid (pH 3–5) | [39,51] |
| 35    | 20(R)-24β-epoxydammar-3β,6α,12β,20,25-tetrol | P. ginseng | Concentrated sulfuric acid (pH 3–5) | [51] |
| 36    | 20(R)-24β-epoxydammar-3β,6β,12β,20,25-tetrol-6-0,6-β-D-glucopyranoside | P. ginseng | Concentrated sulfuric acid (pH 3–5) | [51] |
| 37    | 20(R)-24β-epoxydammar-3β,6α,12β,20,25-tetrol-6-0,6-β-D-glucopyranoside | P. ginseng | Concentrated sulfuric acid (pH 3–5) | [51] |

PDD, protopanaxadiol.
| Entry | Ingredient name                                      | Substrates                        | Transformation condition                  | Refs         |
|-------|-----------------------------------------------------|-----------------------------------|------------------------------------------|--------------|
| 1     | 20(R)-Rh2                                           | 20(R)-Rg3                         | Aspergillus niger                        | [53]         |
| 3     | 20(R)-PPD                                           | Fermented ginseng, 20(R)-Rg3      | Lactobacillus paracasei A221             | [52,53]      |
| 4     | 20(R)-Rh1                                           | Fermented ginseng, 20(S)-Rg1      | Lactobacillus paracasei A221             | [52,54]      |
| 8     | 20(R)-R2                                            | 20(S)-R2                          | Absidia coerulea (AS 3.3389)             | [54]         |
| 19    | 20(R),25-epoxy-3-oxo12β-hydroxydammarane            | 20(R)-PD                          | Absidia corymbifera (AS 3.3387)          | [57]         |
| 20    | 20(R),25-epoxy-3-oxo7β,12β,24α-trihydroxydammarane | 20(R)-PD                          | Absidia corymbifera (AS 3.3387)          | [57]         |
| 21    | 20(R),25-epoxy-3-oxo7β,12β,24β-trihydroxydammarane | 20(R)-PD                          | Absidia corymbifera (AS 3.3387)          | [57]         |
| 22    | 20(R),25-epoxy-3-oxo12β,15α,24α-trihydroxydammaran | 20(R)-PD                          | Absidia corymbifera (AS 3.3387)          | [57]         |
| 23    | 20(R),25-epoxy-3-oxo12β,15α,24β-trihydroxydammaran | 20(R)-PD                          | Absidia corymbifera (AS 3.3387)          | [57]         |
| 24    | 20(R),25-epoxy-3-oxo7β,12β,24β-trihydroxy-15-endammarane | 20(R)-PD                          | Absidia corymbifera (AS 3.3387)          | [57]         |
| 25    | 20(R),25-epoxy-3-oxo-7β,12β,24β-trihydroxy-15,30-cyclodammarane | 20(R)-PD                          | Absidia corymbifera (AS 3.3387)          | [57]         |

PD, panaxadiol.

Fig. 2. The oak ridge thermal ellipsoid plot (ORTEP) figures of 3, 7, 34, 39, and 40, showing the atom-labeling scheme. Displacement ellipsoids are drawn at the 30% probability level, and H atoms are shown as spheres of arbitrary radii.
Fig. 3. Epimerization of 20(S)-ginseng saponins to 20(R)-isomers in acid. The dehydration reaction produced a carbocation intermediate at C-20 of 20(S)-ginseng saponins. Upon rehydration, an oxonium ion was generated, resulting in the conversion of the configuration at C-20 and the creation of 20(R)-ginseng saponins.

Inhibition than 20(R)-Rg3 [anticancer API in Shenyi capsule (YBZ00842003) in China]. Zhang and Zhao [49] found that unsaturated solution and low temperature were beneficial to the separation of 20(R)-25-methoxyl-PPD. They found that under the optimized conditions (acetone, 26 mL, 4 °C), the yield and purity of AD-1 crystals were 38.76% and 97.83% from a nonracemic enantiomeric mixture of compound 28 (R: 59.34%; S: 38.35%), respectively.

20(R)-25-acetyldammar-3β,12β,20-triol (31) was first isolated from hydrolysates of stems and leaves of P. notoginseng (Burk) F. H. Chen with hydrochloric acid. It was purified by RP-HPLC (CH₃OH/H₂O, 88:12) and identified by comparison with the 1H NMR and 13C NMR data of Compounds 25 and 26 [50]. The contents of Compounds 3, 26, and 31 were highest in the stems and leaves of P. notoginseng.

In our previous study, two C-24 epimeric 3-acetylated ocotillol-type saponins, (20R,24S)-epoxy-3-acetyldammar-3β,12β,20-triol (33, m.p. 241–243°C) and (20R,24R)-epoxy-3-acetyldammar-3β,12β,20-triol (34, m.p. 242–244°C), were prepared and isolated from Compound 3 with acetic anhydride in pyridine at room temperature using silica gel column chromatography (ethyl acetate/petroleum ether, 10:1) and crystallized from ethyl acetate. Their structures were confirmed by 1H NMR, 13C NMR, HR-MS, and single-crystal X-ray diffraction [32,58].

(20R,24R)-epoxydammar-3β,6α,12β,25-tetraol (35, m.p. 289–291°C) and (20R,24S)-epoxydammar-3β,6α,12β,25-tetraol (36, m.p. 225–226°C) were systematically semisynthesized from the residue of Compound 7 by Yang et al [51]. (20R,24R)-epoxydammar-3β,6α,12β,25-tetraol-6-O-β-D-glucopyranoside (37, m.p. 187–189°C) and (20R,24S)-epoxydammar-3β,6α,12β,25-tetraol-6-O-β-D-glucopyranoside (38, m.p. 185–187°C) were separated from the residue of Compound 4. Structures were elucidated based on comprehensive 1H NMR, 13C NMR, heteronuclear singular quantum correlation (HSQC), heteronuclear multiple bond correlation (HMBC), rotating frame overhauser enhancement spectroscopy (ROESY), and MS. The configurations at C20 or C24 and the number of glycosyl at C-3 were shown to have an important influence on the compounds’ cytotoxicity.

Recently, (20R,24S)-20,24-epoxy-3-oxo-12β,25-dihydroxymarane (39, m.p. 213–215°C), and (20R,24R)-20,24-epoxy-3-oxo-12β,25-dihydroxymarane (40, m.p. 223–224°C), (20R,24S)-20,24-epoxy-12-oxo-3β,25-dihydroxymarane (41, m.p. 251–253°C) and (20R,24R)-20,24-epoxy-12-oxo-3β,25-dihydroxymarane (42, m.p. 90.5–92.5°C) were synthesized by oxidation and hydrolysis from Compound 3. Their structures were confirmed by hr-ms, 1H NMR, and 13C NMR. Simultaneously, the configurations of Compounds 39 and 40 were authenticated by single-crystal X-ray diffraction [33].

The C20 configuration of ginsenosides is unchanged during alkaline degradation. Consequently, 20(R)-ginseng saponins are primarily obtained by acid epimerization, and some of them possess good pharmacological activities [8] (Table 2). However, some side reactions such as epimerization and hydration were easily induced. The optimization of chemically catalyzed and scaled-up preparation of 20(R)-ginseng saponins still faces tremendous difficulties.

4.2. Microbial transformation

Compounds 3 (328 mg) and 4 (43 mg) could be isolated from fermented ginseng (1 kg) and crystallized by Lactobacillus paracasei A221 containing minor ginsenosides and metabolites of
fermentation [52]. Compound 3, the main metabolite of enzymatic reactions of Compound 2 from black ginseng (pH 5.0, 55°C), with 100% conversion, possessed potential antimutant properties [53].

Chen et al [54] produced Compound 4 and 20(R)-R2 (8) from microbial transformation of 20(S)-PPD-type saponins by Absidia coerulea (26°C, 95% ethanol). Compound 8 [m.p. 239–240°C; \( \frac{1}{2}C_{20}D + 17 \) (c 0.17, MeOH)] was identified as a new metabolite by spectroscopic data [55,56].

Chen et al [57] investigated the microbial transformation of Compound 10 by Absidia corymbifera AS 3.3387. Seven oxidized and hydroxylated products 20(R)-25-epoxy-3-oxo-12β-hydroxydammarane (19), 20(R)-25-epoxy-3-oxo-7β,12β,24α-trihydroxydammarane [20, m.p. 312–313°C; \( \frac{1}{2}C_{20}D + 47.2 \) (c 0.1, MeOH)], 20(R)-25,28-epoxy-3-oxo-7β,12β,24α-trihydroxydammarane [21, m.p. 334–336°C; \( \frac{1}{2}C_{20}D + 24.7 \) (c 0.1, MeOH)], 20(R)-25,28-epoxy-3-oxo-12β,15α,24α-trihydroxydammarane [22, m.p. 285–287°C; \( \frac{1}{2}C_{20}D + 18.7 \) (c 0.1, MeOH)], 20(R)-25-epoxy-3-oxo-12β,15α,24α-trihydroxydammarane [23, m.p. 311–313°C; \( \frac{1}{2}C_{20}D + 21.2 \) (c 0.1, MeOH)], 20(R)-25,28-epoxy-3-oxo-7β,12β,24α-trihydroxy-15-endoendammarane [24, m.p. 292–294°C; \( \frac{1}{2}C_{20}D + 26.4 \) (c 0.1, MeOH)], and 20(R)-25-epoxy-3-oxo-7β,12β,24α-trihydroxy-15,30-cyclodammarane [25, m.p. 352–353°C; \( \frac{1}{2}C_{20}D + 30.3 \) (c 0.1, MeOH)] were obtained, which are new compounds reported for the first time. Their structures were elucidated by \(^1H\) NMR, \(^13C\) NMR, HSQC, HMBC, ROESY, and HR-MS.

Compounds 23, 24, and 25 exhibited strong cytotoxic activities against four cancer cell lines (Du-145, Hela, HepG2, and MCF-7) and one normal cell line Vero.

Microbial transformation is an impactful approach to diversify natural product structures and prepare a variety of derivatives to search for new lead compounds (Table 3). However, optimizing incubation conditions, microbial transformation conditions, and extraction processes is still a huge challenge.

5. Pharmacological activities of 20(R)-ginseng saponins

5.1. Effect on nervous system

He et al [58] reported that 20(R)-Rg3 (2) could attenuate the neuronal apoptosis caused by cerebral ischemia—reperfusion injury through the downregulation of calpain 1 and caspase-3. The expression of calpain I and caspase-3 mRNA could significantly be inhibited at doses of 10 and 20 mg/kg.

Li [15] reported that 20(R)-ginseng saponins (10 and 20 μM of Rh1 or Rh2) could suppress 6-hydroxypoline (6-OHDA) toxicity in SH-SYS5 cells, induce neurite outgrowths in PC-12 cells, and reduce the 6-OHDA—induced phosphorylation of extracellular signal—regulated protein kinase (ERK) (20 μM).

5.2. Protective effects of human umbilical vein endothelial cells (HUVECs) against injury

It has been reported that 20(R)-Rg3 (2) can protect cultured HUVECs from injury by ipopolysaccharide (LPS) and tumor necrosis factor-α (TNF-α). The mechanism might be closely associated with elevating the mobilization of cytosolic calcium, attenuating the generation of plasmagenal activator inhibitor-1 (PAI-1), and elevating the tissue-type plasminogen activator (t-PA) level [59,60]. It could induce HUVEC proliferation at micromolar concentration (10 μM), cell migration, and tube formation in vitro (1–103 μM), as well as ex vivo endothelial spraying through the activation of protein kinase B/extracellular signal—regulated protein kinase (ERK)-endothelial nitric oxide synthase (AKT/ERK-eNOS) signaling pathways.

Compound 2 (150 and 600 nM) also remarkably reduced basic fibroblast growth factor—induced angiogenesis in an in vivo Matrigel plug assay [61,62].

Keung et al [11] found that Compound 2 could inhibit angiogenesis. They demonstrated that Rg3-induced angiosuppression (20 nM) could be mediated by miR-520h, which was proposed to negatively regulate angiogenesis through suppressing EphB2 and EphB4 expression.

5.3. Antiinflammatory and antioxidative effects

It had been reported that 20(R)-Rh2 (1; 10, 30, and 50 μM) has matrix metalloproteinase inhibitory, antiinflammatory, and antioxidative activities by inhibiting nitric oxide (NO), prostaglandin E2 (PG2), reactive oxygen species (ROS) and pro—matrix metalloproteinase-9 (pro-MMP-9) levels [63].

Wei et al [8] found that Compound 2 (10 mg/kg) exhibited significantly higher antioxidant effects than its 20(S)-isomer (10 mg/kg) in mice. It might inhibit Cy-induced oxidative stress in mice and elevate the activities of catalase, superoxide dismutase, and lysozyme (p < 0.05).

Cheng et al [63] reported that Compound 2 (50 μg/mL) could suppress the early formation of hypertrophic scarring (HS) and later HS hyperplasia by inducing the apoptosis of fibroblasts. Yoon et al [64] found that Compound 2 (20 mg/mL) might act as a dual therapeutic regulator for the treatment of inflammatory and oxidative stress—related diseases and downregulating VEGF expression [26,66].

20(R/S)-Rg3 exhibited obvious protection against H2O2—induced oxidative stress in SK-N-SH cells, with Compound 2 and 20(S)-Rg3 decreasing ROS formation by 44.1% and 29.2%, respectively. Thus, the 20(R)-isomer displayed better antioxidant activity than 20(S)-isomer in vitro [13].

Fufang Ejiao Syrup (FES, 20(R)-Rg3 present) is an extensively used immune booster in Traditional Chinese Medicine in eastern Asian countries. Compound 2 (50 g/mL) demonstrated cytoprotective effects on bEnd.3 cells against oxidative injury and was discarded to perform the preprotective study [67].

5.4. Antitumor effects

Lv et al [67] found that 20(R)-Rh2 (1) suppressed the growth of H22 transplanted tumors in vivo, and the highest inhibition rate was 46.8% (p < 0.05). Qi et al [68,69] demonstrated that it could stimulate A549 cell apoptosis by activating the bK kinase/nuclear factor—κ-gene binding (IκB/NF—κB) signaling pathway.

Kim [70] reported that Compound 2 (CS7 BL/6 mice, 100 g intravenous (i.v.); 1,000 g per os (p.o.)) could inhibit pulmonary metastasis of B16-BL6 melanoma cells in vitro and in vivo by oral administration.

Compound 2 (100 mM) had a strong inhibitory effect on UGT1A8 and UGT1A1. Compound 2, 20(R)-Rh2, 20(R)-PPD, and 20(R)-Rh1, had no apparent inhibitory effect on UGT1A1 [71]. Kim et al [71] found that Compound 2 (25 and 50 g/mL) could suppress lung cancer migration, invasion, and anoikis resistance of A549 lung cancer cells in vitro by inhibiting the tubuloglomerular feedback—β1 (TGF—β1)—induced epithelial to mesenchymal transition (EMT).

An efficacy study showed that Compound 2 (3 mg/kg) could significantly inhibit the growth of H22 transplanted tumors in mice, and the inhibition rate of tumor growth was 40.9% [72,73].

Compound 2 notably improved the clinical therapeutic efficacy and quality of life of patients. The clinical relief rate of patients treated with antiangiogenic agent Compound 2 (36.6%) was higher than that of the patients not treated with it (16.7%, p < 0.05). It provided better therapeutic efficacy against early—stage cancer than in advanced stages (p < 0.05) [60].
Compounds 35, 36, 37, and 38 (ocotilloil-type saponins) were synthesized from 20(R)-Rh1 (4) and 20(R)-PPT (7). They possessed better cytotoxicity (200 μg/mL) against HeLa, A549, and MCF-7 cells (>90%) than their corresponding 20(S)-isomer, suggesting that the carbonyl group at C3 might improve their cytotoxicity [74].

**Shenry capsule (20(R)-Rg3, API)** inhibited the proliferation of HO-8910PM cells and the apoptosis of melanoma B16-4A5 cells [75]. Huang et al [75] found that it could treat advanced primary liver cancer, with treatment group survival and efficiency rates of 90% and 70%, respectively.

### 5.5. Other effects

20(R)-Rh2 (1) might play a large role in selective osteoclastogenesis inhibition. Liu et al [16] reported that Compound 1 showed selective osteoclastogenesis inhibitory activity without any cytotoxicity (RAW264 cells) up to 100 μM in vitro. However, the mechanism was ambiguous and needs further research.

Tang et al [14] predicted a benefit of Compound 2 (0.1 and 0.5 mg/kg) as an antifatigue treatment by intranasal administration in mice (p < 0.05). The mechanism was related to the increase of the storage of hepatic glycogen and the decrease of the accumulation of metabolites such as lactic acid and serum urea nitrogen.

Compared with other ginsenosides, 20(R)-ginsenoside possessed better pharmacological activities, such as antioxidant, cytotoxicity, and osteoclastogenesis inhibitory effects. Some studies found that Compounds 1 and 2 have better osteoclastogenesis inhibitory and antioxidative activities than the corresponding 20(S)-isomers. **Shenry capsule (API: 20(R)-Rg3)** and FES (API: 20(R)-Rg3) were approved by the China Food and Drug Administration (CFDA) in China as antitumor and immune-boosting drugs. However, the mechanisms of actions are not clear and require investigation.

### 6. Metabolism of 20(R)-ginseng saponins

The absorption of Compound 2 (3.2 mg/kg) was rapid in the human body, and its elimination was rapid after oral administration in vivo. The pharmacokinetic results showed that it exhibited first-order kinetic characteristics in 14 healthy volunteers [76,77].

Mami [55] found that Compound 2 (100–1000 μg/mouse) could induce a significant decrease in lung metastasis of B16-BL6 melanoma by oral administration. The mechanism of the antimetastatic effect was related to inhibition of the lung and invasion of tumor cells and to antiangiogenesis activity.

Anaerobic incubation of Compound 2 quickly yielded 20(R)-Rh2 (1) or 20(R)-PPD (3) with bacteria isolated from human fecal microflora, Bacteroides sp., Eubacterium sp., and Bifidobacterium sp. However, Fusobacterium sp. could metabolize Rg3 only to Rh2. **Compound 3 (50–100 mg/mL)** potently inhibited the growth of Helicobacter pylori [78].

Peng et al [78] found that 20(R/S)-Rg3 could be deglycosylated to their corresponding chiral metabolites, Compound 1 or Compound 3, to different extents. However, Compound 3 can undergo single-direction chiral inversion to 20(S)-Rg3 in rats. The chiral inversion rate was calculated to be 7.9% after i.v. administration and was estimated to be greater than 9.7% after i.g. administration. Stereo-selective pharmacokinetic parameters, metabolic degrees, and chiral inversion extents of Rg3-epimers in rats were also discussed for the first time.

20(R)-Rh1 was identified as an Rg metabolite in rats. The microbial transformation metabolites of Re were used as standard references, and they were identified by HPLC-ESI-MS/MS [79].

Pharmacokinetic study showed that 20(R)-Rg2 (2 mg/kg) was rapidly absorbed and eliminated in plasma after i.v. administration by a simple and reproducible HPLC method [80].

Many studies reported that the absorption, distribution, metabolism, and excretion of 20(R)-ginsenosides are rapid in vivo. 20(S)-ginsenosides can be transformed to 20(R)-isomers by various microorganisms in vivo. However, the microbial transformation condition is difficult in vitro and needs further exploration.

### 7. Conclusion and perspectives

Panax ginseng, known as Koran ginseng, one of the most commonly used traditional plants, has shown a wide range of pharmacological applications. Ginseng saponins are the major active ingredients found in ginseng and are responsible for biological and pharmacological activities, such as antioxidation, antiinflammation, vasorelaxation, and anticancer actions. Our understanding of 20(R)-ginsenoside saponins has advanced tremendously in the last few years. Investigations related to pharmacological and stereo-selective activities of 20(R)-ginsenoside saponins have been helpful in verifying the functions and values of the corresponding 20(S)-isomers, such as antitumor, antioxidant, cytotoxicity and osteoclastogenesis inhibitory effects. The effects of 20(R)-ginsenoside saponins in cancer chemoprevention and therapy are one of the most popularly studied areas, and the curative effects are known in clinical terms. **Shenry capsule (YBZ00842003, 20(R)-Rg3 as API) and FES (Z37021371, API: 20(R)-Rg3),** approved by CFDA, combined with some chemotherapy agents, have remained the favorable treatment for primary lung cancer and liver cancer and improving immunity. Based on our review, 20(R)-ginsenoside saponins function as anticancer agents because of their antiproliferative, antiinflammatory, and antioxidant effects. They have not only great potential capacity for varied mechanisms for cancer treatment but also a lack of toxicity to normal cells, making 20(R)-ginsenoside saponins appealing candidates for drug development.

To date, studies have mostly focused on identifying and purifying single 20(R)-ginsenoside saponins and investigating their pharmacological activities and molecular mechanisms in cell lines and animal models. Few 20(R)-ginsenoside saponins have been tested in humans despite the fact that they are widely accepted to have therapeutic effects when used alone or in combination with other therapeutic agents in the management of a wide range of chronic diseases. Therefore, studies demonstrating the therapeutic effects of ginseng and 20(R)-ginsenoside saponins are in high demand. Moreover, further studies using single 20(R)-ginsenoside saponins need to include the molecular and cellular mechanisms of action, specificity, structure—function relationship, pharmacokinetic profile, and toxicity in animal models and humans. These studies could maximize the potential of 20(R)-ginsenoside saponins as promising herbal medicines, thereby further contributing to the promotion of global health. There are still many challenges in the optimization of chemical semisynthesis, microbial transformation, and scaled separation of optical 20(R)-ginsenoside saponins. A number of recent studies have presented evidence showing that fermentation by microorganisms is the most effective and environmental transformation mode for the separation of 20(R)-ginsenoside saponins. Some metabolites produced by the action of microbes have different structures from those of naturally occurring 20(R)-ginsenoside saponins in vivo or vitro. Microbial transformation may well become a research hot spot, opening a new avenue for ginseng development in the future. Therefore, further studies are needed to optimize microbial transformation conditions and scale up separation techniques of 20(R)-ginsenoside saponins.
Conflicts of interest

The authors declare no conflict of interest.

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Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.jgr.2019.01.005.

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