Circulating levels of growth hormone in postural orthostatic tachycardia syndrome

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Postural orthostatic tachycardia syndrome (POTS) is a cardiovascular autonomic disorder occurring predominantly in young women. POTS is characterized by an excessive heart rate increase when assuming upright posture accompanied by symptoms of orthostatic intolerance1. However, many POTS patients suffer from a wide array of non-specific ailments such as deconditioning, headache, dyspnea, muscle fatigue and cognitive impairment1.

Different phenotypes of POTS have been identified, e.g. neuropathic POTS and hyperadrenergic POTS2. Likewise, POTS can be physiologically classified based on patterns of peripheral blood flow and peripheral arterial resistance into low-flow POTS, normal-flow POTS and high-flow POTS3. This variety of phenotypes suggests that POTS is a heterogeneous syndrome with many manifestations sharing common pathophysiological mechanisms.

Human growth hormone (GH) is a peptide hormone secreted from the anterior hypophysis. GH plays a vital role in stimulating growth and development and maintenance of the cardiovascular system. Both up- and down-regulation of GH have been linked to increased cardiovascular morbidity and mortality. Upregulation of GH may cause hypertension, arrhythmias, diabetes mellitus, concentric cardiac hypertrophy, coronary artery disease, chronic heart failure, and stroke4, whereas downregulation of GH is characterized by dyslipidemia, insulin resistance, endothelial dysfunction, vascular atherosclerosis, a decrease in cardiac mass together with a post-exertional impairment of systolic function5. A possible explanation may be that subjects demonstrating GH level alterations have higher prevalence of hypertension, diabetes and cardiomyopathy6,7.

Previous evidence based on high-throughput proteomics analysis by proximity extension assay (PEA) technique suggested that levels of circulating GH might also differ between patients with POTS and other possible autonomic disturbances with a history of syncpe but normal response to tilt testing8. Although targeted proteomics is an ideal approach for establishing biomarker signatures in various clinical settings, dedicated detection methods and adequately matched control populations are key for external validation of PEA-guided biomarker discovery.

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Accordingly, we aimed to investigate circulating GH levels in POTS versus age- and sex-matched healthy control group from the same geographical location by high-sensitivity chemiluminescence sandwich immunoassay for plasma GH detection.

**Methods**

The data that support the findings of this study are available from the authors upon reasonable request.

**Study design and population.** The present study is based on the Syncope Study of Unselected Population in Malmö (SYSTEMA) cohort, with over 2200 patients investigated for syncope and severe orthostatic intolerance at the Skåne University Hospital in Malmö, Sweden between 2008 and 2019. Details of the SYSTEMA cohort are described elsewhere. The SYSTEMA study protocol consists of cardiovascular autonomic testing including head-up tilt (HUT) testing with continuous hemodynamic monitoring, as well as other cardiac tests including ambulatory electrocardiogram (ECG) or 24-h ambulatory BP monitoring, when appropriate.

Forty-two consecutive POTS patients with heart rate increase of ≥30 bpm during HUT and chronic symptoms for ≥6 months were selected from SYSTEMA cohort. Forty-six controls were recruited through personal invitation, among healthy medical students, Skåne University Hospital staff and younger participants of parallel population-based epidemiological programs in Malmö, Sweden. Controls had no history of syncope, orthostatic intolerance or endocrine disease. This part of the SYSTEMA project was approved by The Regional Ethical Review Board of Lund University, Sweden in a separate decision (2017/295). All participants, both POTS and controls, were examined at the Clinical Research Unit of Dept. of Internal Medicine, Skåne University Hospital, Malmö, Sweden.

All participants, i.e. POTS patients and healthy controls, were examined in a dedicated research facility. The study participants performed an active standing test, with 10-min rest in the supine position prior to standing. Blood pressure and heart rate were measured twice in the supine position by a validated automated oscillometer device (Omron, Kyoto, Japan), and then after 1, 3, 5, and 10 min of standing. An average of two measurements in the supine position was used for group comparisons. For orthostatic heart rate increase, difference between standing heart rate after 3 min and supine heart rate was calculated. All cardiovascular pharmacological agents such as beta-blockers, ivabradine, midodrine and droxidopa, were discontinued 72 h prior to examination. Standing heart rate after 3 min and supine heart rate was calculated. All cardiovascular pharmacological agents such as beta-blockers, ivabradine, midodrine and droxidopa, were discontinued 72 h prior to examination. POTS diagnosis established during previous tilt-testing and characteristic symptoms were considered valid for POTS diagnosis in patients with pronounced orthostatic intolerance symptoms who were unable to abstain from medication or non-pharmacological measures (such as increased fluid intake or compression garments) that lead to improved hemodynamic responses on orthostatic stress and active standing.

**Orthostatic hypotension questionnaire (OHQ).** All study participants completed an adapted questionnaire based on the validated Orthostatic Hypotension Questionnaire (OHQ) consisting of two components: the six-item symptoms assessment scale (OHSA), and a four-item daily activity scale (OHDAS) to assess the burden of symptoms.

Following symptoms are included in the OHSA: dizziness, problems with vision, generalized weakness, fatigue, trouble concentrating and head/neck discomfort. The participants were asked to rate each item from zero (none) to ten (worst possible) based on the severity of their symptoms during the past week.

The OHDAS includes questions related to how participants’ orthostatic intolerance affects their daily life, e.g. activities that require standing/walking for a short/long time. Participants were asked to rate each item from zero (no interference) to ten (total interference), based on how much the activity has been interfered with during the past week.

**Growth hormone analysis.** Study participants were asked to fast overnight and refrain from smoking, since both factors are known to influence the GH levels. Ethylenediaminetetraacetic acid (EDTA) plasma samples were collected in the supine position and frozen at −80 °C. GH levels were assessed using high-sensitivity chemiluminescence sandwich immunoassay for plasma GH detection (SphingoTec GmbH, Hennigsdorf, Germany).

The assay consists of two antigen-specific monoclonal antibodies, derived against recombinant 22 kDa growth hormone (somatropin) (NIBSC code 98/574, National Institute for Biological Standards and Control, Hertfordshire, UK) in a microtiter plate format. Both monoclonal antibodies bind to the antigen Vascular-Risk Human Growth Hormone (vr-hGH) at two individual epitopes. During incubation the vr-hGH interacts with the luminescent tracer antibody and capture antibody in the plasma. Upon indirect binding of the tracer antibody to the microtiter plate, the tracer remnant is washed away, and the level of captured tracer antibody can be analyzed using a luminometer.

The obtained luminescence signal is directly proportional to the level of vr-hGH in the plasma sample. The lower detection limit is 0.001 ng/mL and the functional assay sensitivity is 0.004 ng/mL, with an inter-assay coefficient of variation of 20%. Growth hormone is stable for 7 days in EDTA plasma at 2–35 °C and up to five freeze thaw cycles. Further details on the method are described elsewhere.

**Statistical analysis.** Between-groups comparison was performed using standard statistical tests, ANOVA for continuous variables and Pearson’s Chi-square test for categorical variables. Baseline characteristics of the study population were reported as mean and standard deviation or median and interquartile range, as appropriate, for continuous variables, and as counts and percentages for categorical variables. Growth hormone values were log-transformed and standardized before the group comparison to transform skewed data to approximately conform to normality. The relationships among GH levels, hemodynamic data and symptoms of orthostatic
intolerance by OHQ were explored by linear regression analysis. Data were analyzed using SPSS software version 26 (SPSS, Chicago, IL, USA). P-value of $< 0.05$ was considered statistically significant.

**Ethical approval.** The SYSTEMA project was approved by the Regional Ethical Committee in Lund, Sweden (82/2008), and all study participants provided informed written consent. All procedures were carried out in line with relevant current guidelines and regulations.

**Results**

Characteristics of the study population are displayed in Table 1. Supine heart rate and supine diastolic BP were significantly higher in POTS patients compared with controls ($69.0 \pm 11.1$ bpm vs. $63.3 \pm 10.8$ bpm, $p = 0.02$; and $72.9 \pm 9.1$ mmHg vs. $69.0 \pm 8.5$ mmHg, $p = 0.04$). Heart rate after 3 min of standing was also significantly increased in POTS patients ($96.5 \pm 16.0$ vs. $83.1 \pm 13.6$, $p < 0.001$). However, we did not observe any significant difference in supine systolic BP ($116.6 \pm 13.3$ mmHg vs. $115.2 \pm 10.0$ mmHg, $p = 0.60$) between the groups.

**POTS and OHQ score.** POTS patients had significantly higher composite OHQ score than controls ($60.0 \pm 18.6$ vs. $4.2 \pm 7.5$, $p < 0.001$), as well as OHSA ($36.2 \pm 10.0$ vs. $3.6 \pm 6.4$, $p < 0.001$) and OHDAS scores ($23.8 \pm 9.7$ vs. $0.6 \pm 1.3$, $p < 0.001$).

**Plasma levels of growth hormone in POTS.** Plasma levels of GH were significantly lower in POTS patients compared with controls (median $0.53$ ng/mL, IQR, $0.10–2.83$ vs. $2.33$ ng/mL, IQR, $0.26–7.2$, $p = 0.04$, Fig. 1), although the GH levels were still within the normal range. Moreover, we found that levels of GH were inversely related to OHDAS ($p = 0.049$) among POTS patients (Fig. 2). However, OHQ and OHSA scores were not significantly associated with GH levels in POTS group ($p = 0.37$ and $p = 0.84$, respectively (not illustrated). In addition, post-hoc analysis showed no relationship between POTS symptom duration (median 4 years, IQR: 2–11) and levels of GH ($p = 0.134$) (not illustrated).

Further, levels of GH were inversely associated with supine systolic BP ($B = -0.07$, SE = $0.02$; $p < 0.001$) in all participants, and in POTS ($B = -0.06$, SE = $0.02$, $p = 0.004$) but there was no association with heart rate, neither supine ($p = 0.33$) nor standing ($p = 0.18$), even when restricting the analysis to POTS subgroups ($p = 0.53$ and 0.25, respectively) (not illustrated). The association between lower GH and higher SBP demonstrated a distinct threshold level whereby SBP levels were significantly higher only in the lowest tertile of GH, both in all participants and POTS subgroup (Fig. 3).

|                  | POTS (n = 42) | Controls (n = 46) | P-value |
|------------------|---------------|------------------|---------|
| Mean age, years  | $31 \pm 9$ (19–62) | $32 \pm 9$ (18–59) | 0.54    |
| Female, n (%)    | 36 (86)       | 35 (76)          | 0.25    |
| SBP supine, mmHg | $116.6 \pm 13.3$ | $115.2 \pm 10.0$ | 0.60    |
| DBP supine, mmHg | $72.9 \pm 9.1$  | $69.0 \pm 8.5$   | 0.04    |
| HR supine, bpm   | $69.0 \pm 11.1$ | $63.3 \pm 10.8$  | 0.02    |
| HR 3 min standing, bpm | $96.5 \pm 16.0$ | $83.1 \pm 13.6$  | $< 0.001$ |
| OHQ score        | $60.0 \pm 18.6$ | $4.2 \pm 7.5$    | $< 0.001$ |
| OHSA score       | $36.2 \pm 10.0$ | $3.6 \pm 6.4$    | $< 0.001$ |
| OHDAS score      | $23.8 \pm 9.7$  | $0.6 \pm 1.3$    | $< 0.001$ |

Table 1. Baseline characteristics and intergroup comparison between POTS and healthy controls. DBP: diastolic blood pressure; HR: heart rate; OHDAS: orthostatic hypotension daily activities; OHQ: Orthostatic Hypotension Questionnaire composite score; OHSA: Orthostatic Hypotension Symptom Assessment; POTS: postural orthostatic tachycardia syndrome; SBP: systolic blood pressure.
Discussion

In this study, we have demonstrated that patients with postural orthostatic tachycardia syndrome have lower levels of growth hormone. The lower levels of GH were associated with the overall severity of daily activity impairment among POTS patients, but not with the general and less specific symptoms such as weakness or problems with concentration.

Previous studies have shown that POTS patients have impaired quality of life. Although it has been demonstrated that individuals with GH deficiency and compromised quality of life may benefit from GH replacement therapy, the relationship between quality of life and GH deficiency in POTS is complex and warrants in-depth research.

Promising results have been reported about the beneficial effects of GH replacement therapy on the cardiovascular system in individuals with GH deficiency. It has been shown that GH replacement significantly improves various cardiovascular risk parameters, e.g. lipid-profile and glucose metabolism, with other possible pleiotropic cardioprotective effects. However, potential clinical utility and efficacy of GH replacement therapy on symptom burden of POTS patients and its impact on cardiovascular risk profile is largely unknown.

The hypothalamus secretes growth hormone-releasing hormone (GHRH), which stimulates GH synthesis and secretion to a large extent, while somatostatin inhibits the GH secretion from the anterior hypophysis. Previous studies have shown that low levels of GH are associated with an excessive autonomic sympathetic activity. In GH deficiency, the levels of GHRH are increased, prompting the authors to assume that the effect of low GH may be attributed to a positive feedback loop between GHRH and norepinephrine. Since GH release is blocked in GH deficiency, a rise in norepinephrine occurs. This efflux of norepinephrine may in turn enhance production of GHRH and, thus, release of GH. However, this may not be the case in POTS.

POTS is associated with abnormal sympathetic hyperactivity. Norepinephrine metabolism is described as abnormal in hyperadrenergic POTS with increased levels of plasma norepinephrine. The possible effect of GH on the autonomic nervous system may be more evident in some individuals, thus, potentially leading to a greater predisposition to POTS-related symptoms. Although at the current stage of understanding, the relative place of GH and norepinephrine in the pathophysiological cascade of POTS is not yet established.

Moreover, exercise is a strong stimulus of GH secretion. Reduced exercise capacity and physical deconditioning occurs in both patients with POTS and GH deficiency, however, whether GH levels are related to mechanism of POTS is yet unknown. It may be speculated that the observed lower levels of GH in the present study may be a consequence of POTS-induced physical inactivity. Interestingly, evidence has demonstrated that deconditioning in POTS may be associated with reduced left ventricular mass (LVM), and after starting physical training the LVM increased in individuals with POTS to the same size as in controls. GH deficiency is also associated with reduced LVM, and studies have demonstrated favorable effects of physical exercise, such as aerobic training in patients with either POTS or GH deficiency. Although the advantages of exercise in GH deficiency may resemble the effects of GH replacement therapy, previous results indicate that the positive effects of physical exercise are not additive to GH therapy.

GH exerts also diabetogenic and anti-insulin properties, partly due to its indirect effects via insulin-like growth factor 1 (IGF-1), which has glucose-lowering effects similar to insulin. Consequently, it may be hypothesized that reduced GH levels may result in enhanced insulin action, conferring a higher risk of prediabetes in a subset of patients with POTS.

A previous study has reported elevated plasma levels of GH in POTS, by using a proteomics approach and analyzing simultaneously over 90 cardiovascular plasma biomarkers in one plasma sample. In our study, we observed lower levels of GH in POTS patients, by using a highly specific immunoassay method. These contradictory findings may be partly explained by the techniques used to analyze GH, since the proteomics technique targeted a wide spectrum of different biomarkers, thus obtaining relative GH levels, whereas our method was targeted at absolute quantification of GH levels. However, the contradictory findings may also be partly explained.
by partly different study groups. The previous study included patients with a normal hemodynamic test as the control group, unlike including healthy subjects from the population, which was the case in our present study. Also, the timing of the measurements in the stage of disease, as well as the clinically insignificant variations in GH level that were within normal ranges may further explain the conflicting results.

Moreover, we found that lower levels of GH were associated with higher supine systolic blood pressure in both POTS and healthy individuals. Interestingly, alterations in heart rate were not associated with GH levels. Thus, lower GH levels may paradoxically indicate higher supine BP but reduced orthostatic intolerance, suggesting that alternative mechanisms other than hemodynamic, may be involved in POTS-symptom generation mediated by lower GH levels.

Another previous study reported that patients with GH deficiency also have increased supine systolic blood pressure in comparison with healthy controls, whereas no difference was observed in heart rate. Notably, the study found that individuals with GH deficiency have an abnormal blood pressure response during orthostatic stress, possibly attributed to structural changes of the aorta and carotid artery, which are responsible for the baroreflex control of BP.

Another possible explanation of our observations may be an upsurge in norepinephrine. Since secretion of GH is regulated by norepinephrine, low levels of GH may contribute to elevated SBP by stimulating the sympathetic

**Figure 3.** Supine systolic blood pressure by tertiles of circulating growth hormone levels in all participants (A, n = 84) and in POTS patients (B, n = 42). GH, growth hormone; POTS, postural orthostatic tachycardia syndrome.
nervous system. Ultimately, the underlying pathophysiological link needs further investigation in larger and independent populations.

**Strengths and limitations.** With the biology of POTS incompletely explored, our study supports the association between GH levels and POTS. To the best of our knowledge, this is the first study aimed at directly measuring the levels of GH in POTS. The discordant findings observed in this study with regard to previous proteomics evidence with PEA technology needs further verification in independent cohorts. A biomarker-driven approach for phenotyping POTS may improve the understanding of its pathophysiology.

Although GH levels were associated with impairment of daily life activities and higher supine systolic blood pressure, however, whether there exists a causal relationship between the two events remains to be investigated. It is also unclear if exercise and growth hormone supplementation may potentially influence our observed findings. Moreover, the study sample size of our study was relatively small. Likewise, the observed levels of growth hormone in POTS and controls were in fact within normal ranges, e.g. in adult males 0.4 to 10 ng/mL, and in adult females 1 to 14 ng/mL.

As far as possible clinical implications of this explorative study are concerned, the present results do not yet have any direct therapeutic or diagnostic implications in subjects with POTS. In the long term and for possible clinical relevance, the role of GH should be tested not only among POTS, but also in similar diseases of cardiovascular autonomic function. It may be speculated that measurement of GH in plasma may be useful in POTS to determine the underlying neuroendocrine alterations and aid in clinical decision-making regarding a possible pharmacological intervention or substitution. Considering that patients with POTS have pronounced central nervous system symptomatology, it would be of interest to determine whether the hormonal dysregulation is at the hypothalamic level rather than the pituitary. Future studies should be aimed at assessing the hormonal profile in POTS, including the hypothalamic–pituitary–adrenal axis, which is involved in fluid and salt conservation, as well as measurement of insulin-like growth factor 1 (IGF-1).

**Conclusions**

The multifaceted nature of postural orthostatic tachycardia syndrome poses many diagnostic and therapeutic challenges. Our study adds to the understanding of POTS and shows that patients with POTS have significantly lower plasma levels of circulating growth hormone. Impairment of daily life activities is inversely related with growth hormone levels in POTS. A higher supine diastolic blood pressure is inversely associated with growth hormone levels in both POTS and healthy individuals. Further research is needed to confirm these findings in larger and independent populations.

**Data availability**

The datasets analyzed during the current study are available from the corresponding author on reasonable request.

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**References**

1. Fedorowski, A. Postural orthostatic tachycardia syndrome: Clinical presentation, aetiology and management. *J. Intern. Med.* 285, 352–366. https://doi.org/10.1111/joim.12852 (2019).

2. Ross, A. J., Oconn, A. J., Medow, M. S. & Stewart, J. M. A double-blind placebo-controlled cross-over study of the vascular effects of midodrine in neuroatropic compared with hydropneumonic postural tachycardia syndrome. *Clin. Sci.* 126, 289–296. https://doi.org/10.1042/CS20130222 (2014).

3. Stewart, J. M. & Montgomery, L. D. Regional blood volume and peripheral blood flow in postural tachycardia syndrome. *Am. J. Physiol. Heart Circ. Physiol.* 287, H1319–1327. https://doi.org/10.1152/ajpheart.00086.2004 (2004).

4. Hallgren, E. et al. Fasting levels of high-sensitivity growth hormone predict cardiovascular morbidity and mortality: The Malmo Diet and Cancer study. *J. Am. Coll. Cardiol.* 64, 1452–1460. https://doi.org/10.1016/j.jacc.2014.03.063 (2014).

5. Lombardi, G. et al. The cardiovascular system in growth hormone excess and growth hormone deficiency. *J. Endocrinol. Invest.* 35, 1021–1029. https://doi.org/10.3275/8717 (2012).

6. Melmed, S. et al. A consensus on the diagnosis and treatment of acromegaly complications. *Pituitary* 16, 294–302. https://doi.org/10.1007/s11120-012-0420-x (2013).

7. Colao, A., Manni, M., Di Somma, C. & Lombardi, G. Growth hormone and the heart. *Clin. Endocrinol.* 53, 137–154. https://doi.org/10.1111/j.1365-2265.2003.01218.x (2001).

8. Medic Spahić, J. et al. Proteomic analysis reveals sex-specific biomarker signature in postural orthostatic tachycardia syndrome. *BMC Cardiovasc. Disord.* 20, 190. https://doi.org/10.1186/s12872-020-01465-6 (2020).

9. Fedorowski, A., Burri, P., Struck, J., Juel-Moller, S. & Melander, O. Novel cardiovascular biomarkers in unexplained syncopal attacks: The SYSTEMA cohort. *J. Intern. Med.* 273, 359–367. https://doi.org/10.1111/joim.12043 (2013).

10. Low, P. A. Neurogenic orthostatic hypotension: Pathophysiology and diagnosis. *Am. J. Manag. Care* 21, s248–257 (2015).

11. Wilkins, J. N. et al. Nicotine from cigarette smoking increases circulating levels of cortisol, growth hormone, and prolactin in male chronic smokers. *Psychopharmacology* 78, 305–308. https://doi.org/10.1007/BF00433730 (1982).

12. Iansz, A., Doorenbos, H. & Reitsma, W. D. Effect of food intake on growth-hormone level. *Lancet* 1, 250–251. https://doi.org/10.1016/s0140-6736(63)90955-7 (1963).

13. Biddingmaier, M. et al. High-sensitivity chemiluminescence immunoassays for detection of growth hormone doping in sports. *Clin. Chem.* 55, 445–453. https://doi.org/10.1373/clinchem.2008.112458 (2009).

14. Bagai, K. et al. Sleep disturbances and diminished quality of life in postural tachycardia syndrome. *J. Clin. Sleep Med.* 7, 204–210 (2011).

15. Benrud-Larsen, L. M. et al. Quality of life in patients with postural tachycardia syndrome. *Mayo Clin. Proc.* 77, 531–537. https://doi.org/10.4065/77.6.531 (2002).

16. Brod, M., Pohlmans, B., Hojbjerre, L., Adalsteinsson, J. E. & Rasmussen, M. H. Impact of adult growth hormone deficiency on daily functioning and well-being. *BMC Res. Notes* 7, 813. https://doi.org/10.1186/1756-0500-7-813 (2014).
17. Gola, M., Bonadonna, S., Doga, M. & Giustina, A. Clinical review: Growth hormone and cardiovascular risk factors. J. Clin. Endocrinol. Metab. 90, 1864–1870. https://doi.org/10.1210/jc.2004-0545 (2005).

18. Gazzaruso, C., Gola, M., Karamouzis, I., Giubbini, R. & Giustina, A. Cardiovascular risk in adult patients with growth hormone (GH) deficiency and following substitution with GH—an update. J. Clin. Endocrinol. Metab. 99, 18–29. https://doi.org/10.1210/jc.2013-2394 (2014).

19. Diez, J. J., Sangiao-Alvarellos, S. & Cordido, F. Treatment with growth hormone for adults with growth hormone deficiency syndrome: Benefits and risks. Int. J. Mol. Sci. https://doi.org/10.3390/ijms19030893 (2018).

20. Cordoba-Chacon, J. et al. Somatostatin dramatically stimulates growth hormone release from primate somatotrophs acting at low doses via somatostatin receptor 5 and cyclic AMP. J. Neuroendocrinol. 24, 453–463. https://doi.org/10.1111/j.1365-2826.2011.02261.x (2012).

21. Sversidisott, Y. B., Elam, M., Herlitz, H., Bengtsson, B. A. & Johannsson, G. Intense sympathetic nerve activity in adults with hypopituitarism and untreated growth hormone deficiency. J. Clin. Endocrinol. Metab. 83, 1881–1885. https://doi.org/10.1210/jcem.83.6.4895 (1998).

22. Mar, P. L. & Raj, S. R. Postural orthostatic tachycardia syndrome: Mechanisms and new therapies. Annu. Rev. Med. 71, 235–248. https://doi.org/10.1146/annurev-med-041818-011630 (2020).

23. Casanueva, F. F., Villanueva, L., Cabranes, J. A., Cabezas-Cerrato, J. & Fernandez-Cruz, A. Cholinergic mediation of growth hormone secretion elicited by arginine, clomidine, and physical exercise in man. J. Clin. Endocrinol. Metab. 59, 526–530. https://doi.org/10.1210/jcem.59.3.526 (1984).

24. Fu, Q. et al. Cardiac origins of the postural orthostatic tachycardia syndrome. J. Am. Coll. Cardiol. 55, 2858–2868. https://doi.org/10.1016/j.jacc.2010.02.043 (2010).

25. Blithsteyn, S. & Fries, D. Postural tachycardia syndrome is not caused by deconditioning. Palmd. Circ. 6, 401. https://doi.org/10.1080/20479118.2016.687757 (2016).

26. Isgaard, J. & Cittadini, A. Growth hormone and the heart in growth hormone deficiency—what have we learned so far? Endocrine 55, 331–332. https://doi.org/10.1007/s12020-016-1206-0 (2017).

27. Thomas, S. G., Esposito, J. G. & Ezat, S. Exercise training benefits growth hormone (GH)-deficient adults in the absence or presence of GH treatment. J. Clin. Endocrinol. Metab. 88, 5734–5738. https://doi.org/10.1210/jcem.2003-030632 (2003).

28. Kim, S. H. & Park, M. J. Effects of growth hormone on glucose metabolism and insulin resistance in human. Ann. Pediatr. Endocrinol. Metab. 22, 145–152. https://doi.org/10.1006/apem.2017.22.3.145 (2017).

29. Yuen, K. C. & Dunger, D. B. Therapeutic aspects of growth hormone and insulin-like growth factor-I treatment on visceral fat and insulin sensitivity in adults. Diabetes Obes. Metab. 9, 11–22. https://doi.org/10.1111/j.1463-1326.2006.00591.x (2007).

30. Barreto-Filho, J. A. et al. Familial isolated growth hormone deficiency is associated with increased systolic blood pressure, central obesity, and dyslipidemia. J. Clin. Endocrinol. Metab. 87, 2018–2023. https://doi.org/10.1210/jcem.87.5.8474 (2002).

Author contributions
The manuscript was written by M.J. together with A.F. Figures and statistical analyses were made by E.R. and A.F. All authors contributed equally in reviewing this manuscript.

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Competing interests
AF reports personal fees from Medtronic Inc. and Biotronik outside the submitted work. RS also reports personal fees from Medtronic Inc. and from Abbott Laboratories Corp; he is a shareholder in Edwards Lifesciences Corp. JS is employed by SphingoTec GmbH, the manufacturer of the GH immunoassay. The remaining authors have no conflicts to disclose.

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