INTRODUCTION
Copper is one of the pollutions. Ironically, the human body required a lower number of coppers for maintaining homeostasis, but the higher intake of copper can cause various organ damages, impaired regulation of lipid metabolism, impaired antimicrobial defense, neuronal activity, and resistance chemotherapies. Furthermore, the high levels of copper in the body can also stimulate reactive oxygen species (ROS) production and modify low-density lipoprotein to promote atherogenesis by enhancing the transformation of macrophages and developing vasoconstrictor and prothrombotic characteristics [1].

Some studies have been performed to investigate the environment pollution in some countries. One of these studies was reported that Some Rainwater Tanks in Adelaide Region, Australia, have been polluted by several heavy metals. Most of the pollution was lead. Other heavy metals such as zinc, cadmium, and copper were detected in the fewer samples, especially in the Adelaide Hills and Foothills [2].

The higher number of the copper may be eliminated by Reticular Endothelial System. One of these systems, Spleen, is located in the left upper quadrant of the abdomen. This organ functions included filtration of blood, production of white blood cells, and regulating blood. The overload copper may cause dysfunction of the spleen, reducing red blood cells, white blood cells, and platelet in the circulation. Therefore, the immunity will decrease and vulnerable to be an infection [3].

Some studies have been performed to look for natural resources that can be used to protect the human body against the impact of copper pollution. One of the natural resources is white turmeric rhizome (Curcuma zedoaria). It contains curcumin with some pharmacological properties, including anti-inflammatory, antioxidant, and anti-cancer activity [4, 5]. Due to the presence of antioxidant activity from the white turmeric, it potentially protects various organs against copper. Some studies have been performed to look for white turmeric’s protective effect against kidney and systemic protection (indicated by hematologic study) [6-8]. However, none of the studies investigates the protective effect of white turmeric against copper in spleen tissue. For these reasons, this study was designed to investigate the spleen protective effect of white turmeric rhizome (C. zedoaria) against copper using male Wistar rats as the animal trial.

METHODS
This was an experimental study with post only group control design. This study was performed on July 2020–October 2020 at Pharmacology Laboratory, Universitas Prima Indonesia. This study’s procedure has been approved by the Health Research Ethics Committee from Universitas Prima Indonesia with registration no. 052/KEPK/UNPRI/V/2020.

Materials
The material used in this study included white turmeric, copper sulfate pentahydrate, 96% ethanol, distilled water, sodium carboxymethylcellulose, alcohol, buffer phosphate, xylok, paraffin, and HE Staining.

Identification and preparation of white turmeric
The White turmeric (C. zedoaria) was obtained from UPT Materia Medica Batu, Jawa Timur. The white turmeric was cleaned, dried, and meshed to get the simplicia powder. Moreover, the simplicia powder was soaked into 96% ethanol as the solvent, every 24 h it was filtered, and the filtrate was collected, and the residue was soaked again as before, these processes were repeated twice. Finally, the filtrate was evaporated by rotary evaporator at 50°C until the filtrate becomes concentrated [8, 9].

Formulation of oral suspension
The oral suspension was made of sodium carboxymethylcellulose. Amount of 0.5 g sodium carboxymethylcellulose was suspended into 30 ml hot distilled water in the mortar for 15 min until it formed a clear suspension. Moreover, this suspension was ground until it becomes homogeneous. After that, it was dissolved until 100 ml by distilled water.

The obtained suspension was used as vehicle for extract and copper sulfate pentahydrate. Amount of 45 mg, 67.5 mg, and 90 mg of
extract was mixed into 5 ml of 0.5% sodium carboxymethyl cellulose suspension to form extract suspension dosage 10 mg/200 g BW, 20 mg/200 g BW, and 40 mg/200 g BW, respectively. Last, the amount of 0.36 mg copper sulfate pentahydrate was mixed into 5 ml of 0.5% sodium carboxymethyl cellulose to form copper sulfate suspension [10].

**Intervention**

This study was used 30 rats which were divided into six groups. The intervention was shown as the following.

a. Normal: Rats did not receive any interventions.

b. Negative: Rats received 1 ml of copper sulfate pentahydrate by intragastric oral tube once a day in the 12th, 13th, and 14th.

c. Positive: Rats received 1 ml of extract suspension dosage 10 mg/200 g BW by intragastric oral tube once a day every day for 14 days and 1 ml of copper sulfate pentahydrate by intragastric oral tube once a day in the 12th, 13th, and 14th.

d. Ethanol extract of white turmeric-I: Rats received 1 ml of extract suspension dosage 10 mg/200 g BW by intragastric oral tube once a day every day for 14 days and 1 ml of copper sulfate pentahydrate by intragastric oral tube once a day every day for 14 days and 1 ml of copper sulfate pentahydrate by intragastric oral tube once a day in the 12th, 13th, and 14th.

e. Ethanol extract of white turmeric-II: Rats received 1 ml of extract suspension dosage 20 mg/200 g BW by intragastric oral tube once a day every day for 14 days and 1 ml of copper sulfate pentahydrate by intragastric oral tube once a day every day for 14 days and 1 ml of copper sulfate pentahydrate by intragastric oral tube once a day in the 12th, 13th, and 14th.

f. Ethanol extract of white turmeric-III: Rats received 1 ml of extract suspension dosage 40 mg/200 g BW by intragastric oral tube once a day every day for 14 days and 1 ml of copper sulfate pentahydrate by intragastric oral tube once a day in the 12th, 13th, and 14th.

All rats can freely access food and drink. After 14 days, all rats were sacrificed to obtain the spleen for histology evaluation.

**Sacrificed of rats**

All rats were sacrificed by inhalation of chloroform in the close room. After that, the rat abdomen was incised horizontally. The spleen was collected from the chest cavity and washed into the normal saline. The washed spleen was soaked into 10% formalin buffer saline, and it was reserved until the tissue processing [9,10].

**Tissue processing**

The spleen was sliced to thickness 4–6 mm. After that, it was dehydrated with several alcohol concentrations (70%, 80%, 90%, and 95%) for 24 h and followed by 100% alcohol for an hour for 3 times. After that, it was purifyed by xylol for an hour for 3 times. Moreover, the tissue was infiltrated into the paraffin, and it was incised to thickness 4–5 microns. Finally, the incision was attached to the slide and stained by hematoxylin-eosin (HE) staining [9,10].

**Evaluation of spleen tissue**

The spleen tissue was observed under the light microscope at the ×100 and ×400 magnified. Four parameters which were used to evaluated spleen tissue included congestion or vasodilation of blood vessels, neutrophile infiltration. The scoring system was used to evaluate that the spleen histology is shown in Table 1 [11,12].

**Data analysis**

Each of the parameters was analyzed descriptively. Moreover, these data were analyzed by Chi-square for congestion/vasodilatation of blood vessels and neutrophile infiltration. Meanwhile, lymphoid necrosis and macrophages were analyzed by either one-way ANOVA or Kruskal-Wallis, depending on the normality of data. The Shapiro-Wilk analyzed the normality of that.

**RESULTS**

The white turmeric extract significantly affects the severity of lymphoid necrosis, the number of macrophage cells, the presence of congestion/vasodilatation of blood vessels, and neutrophile infiltration. These results are shown in Table 2.

| Group                  | Congestion/vasodilation blood vessel | Total | p-value |
|------------------------|-------------------------------------|-------|---------|
| Yes                    | 4 (16.7)                            | 0 (0) | 4 (16.7)     |
| No                     | 0 (0)                               | 4 (16.7) | 0.002    |
| Normal                 | 4 (16.7)                            | 0 (0) | 4 (16.7)     |
| Negative               | 0 (0)                               | 4 (16.7) | 4 (16.7)   |
| Positive               | 2 (8.3)                             | 2 (8.3) | 4 (16.7)   |
| Ethanol extract of     | 2 (8.3)                             | 2 (8.3) | 4 (16.7)   |
| white turmeric-I       | 0 (0)                               | 4 (16.7) | 4 (16.7)   |
| Ethanol extract of     | 0 (0)                               | 4 (16.7) | 4 (16.7)   |
| white turmeric-II      | 0 (0)                               | 4 (16.7) | 4 (16.7)   |
| Ethanol extract of     | 0 (0)                               | 4 (16.7) | 4 (16.7)   |
| white turmeric-III     | 0 (0)                               | 4 (16.7) | 4 (16.7)   |
| Total                  | 8 (33.3)                            | 16 (66.7) | 24 (100.0) |

p-value was obtained by Chi-square test

The white turmeric extract significantly affects the severity of lymphoid necrosis, the number of macrophage cells, the presence of congestion/vasodilatation of blood vessels, and neutrophile infiltration. These results are shown in Table 3.

| Group                  | Neutrophil infiltration | Total | p-value |
|------------------------|-------------------------|-------|---------|
| Yes                    | 4 (16.7)                | 0 (0) | 4 (16.7)     |
| No                     | 0 (0)                   | 4 (16.7) | <0.05    |
| Normal                 | 4 (16.7)                | 0 (0) | 4 (16.7)     |
| Negative               | 0 (0)                   | 4 (16.7) | 4 (16.7)   |
| Positive               | 3 (14.3)                | 1 (4.8) | 4 (16.7)   |
| Ethanol extract of     | 3 (14.3)                | 1 (4.8) | 4 (16.7)   |
| white turmeric-I       | 0 (0)                   | 4 (16.7) | 4 (16.7)   |
| Ethanol extract of     | 0 (0)                   | 4 (16.7) | 4 (16.7)   |
| white turmeric-II      | 0 (0)                   | 4 (16.7) | 4 (16.7)   |
| Ethanol extract of     | 0 (0)                   | 4 (16.7) | 4 (16.7)   |
| white turmeric-III     | 0 (0)                   | 4 (16.7) | 4 (16.7)   |
| Total                  | 15 (62.5)               | 9 (37.5) | 24 (100.0) |

p-value was obtained by Chi-square test
neutrophile infiltrations, while the normal and negative groups showed the opposite. Furthermore, all-white turmeric groups (I-III) showed reduced rats with neutrophil infiltration, depending on the extract’s dosage. Fig. 1 below shown the microscopic view of neutrophile infiltration.

This study was also analyzed the severity of lymphoid necrosis by the Kruskal–Wallis test and followed by the Mann–Whitney test. Table 4 below showed the result of this analysis. Table 4 above showed that each group had significant differences in lymphoid necrosis score due to p<0.05 (p=0.002). The normal and positive group did not show any degrees of lymphoid necrosis. However, the highest score of lymphoid necrosis was shown by the negative group. Nevertheless, the median score of lymphoid necrosis showed a decrease followed by the group’s increase, but this reduction was not significant compared to the negative group, instead of the normal or positive group.

Finally, this study was also analyzed the number of macrophages cells in the spleen tissue in the same way used to analyze the severity of lymphoid necrosis. Table 5 below showed the result of the analysis. According to Table 5 above, the negative group has the highest number and narrowest range of macrophage cells. As the opposite, the normal, positive, and white turmeric-I group were the lowest number of macrophage cells. Meanwhile, the increasing white turmeric dosage was followed by the increasing number of macrophage cells. However, the highest dosage of white turmeric has a significant difference in the number of macrophage cells against white turmeric’s lowest dosage. Fig. 2 below showed microscopic view of the macrophages cell in medium - (∗100 magnified) and high-power field (∗400 magnified).

**Table 4: Analysis of lymphoid necrosis in spleen of the intervention groups**

| Group          | Lymphoid necrosis [Median (Min–Max)] | p-value* |
|----------------|--------------------------------------|----------|
| Normal         | 0.00 [0.00–0.00]                     | 0.002    |
| Negative       | 2.00 [1.00–3.00]                     |          |
| Positive       | 0.00 [0.00–0.00]                     |          |
| Ethanol extract of white turmeric-I | 1.50 [1.00–2.00] |          |
| Ethanol extract of white turmeric-II | 1.50 [1.00–2.00] |          |
| Ethanol extract of white turmeric-III | 1.00 [1.00–1.00] |          |

*p-value was obtained from Kruskal–Wallis Test, **There is significant differences against the negative group according to Mann–Whitney Test, *There is significant difference against both normal and positive group according to Mann–Whitney test

**Table 5: Analysis of number of macrophages cells in spleen of the intervention groups**

| Group          | Number of macrophages cells [median (IQR)] | p-value* |
|----------------|-------------------------------------------|----------|
| Normal         | 2.50 [2.00–3.00]                          | 0.003    |
| Negative       | 5.00 [5.00–5.00]                          |          |
| Positive       | 2.50 [2.00–3.00]                          |          |
| Ethanol extract of white turmeric-I | 2.50 [2.00–3.00] |          |
| Ethanol extract of white turmeric-II | 4.50 [3.00–5.00] |          |
| Ethanol extract of white turmeric-III | 5.00 [4.00–5.00] |          |

*p-value was obtained from Kruskal–Wallis Test, **There is significant differences against the negative group according to Mann–Whitney Test, *There is significant difference against both normal and positive group according to Mann–Whitney test

**DISCUSSION**

It becomes obvious that the ethanol extract of white turmeric may protect spleen from the impact of copper. The increase of ethanol extract of white turmeric gave better protection of spleen against copper. It was shown by the improvement of acute inflammatory changes in the spleen such as congestion/vasodilatation of the blood vessel, neutrophile infiltration, and lymphoid necrosis. However, prolonged consumption of white turmeric showed increasing number of macrophage cells in the spleen tissue.

Copper is an essential nutrient and cofactor for various enzymes. The regulation of this nutrient is closely regulated. The excess level of copper in the body can induce oxidative stress which leads to an inflammatory response [13,14]. White turmeric contains more than 20 types of phytochemicals can be found in the white turmeric, include furano diene, furanodienone, zederone, curzerenone, curcumeone, germacronone, 13-hydroxygermacronone, dehydrocudrine, curcumene, zedoarynediol, 13-hydroxycurzerenone, 1oxocurzerenone, curcolone, procucumenol, ermanin, curcumin, the mixture of stigmast-4-en-3,6-dione, and stigmata-4,22-dien-3,6. More than these phytochemicals are responsible for white turmeric’s pharmacology properties such as antioxidant and anti-inflammatory effect. These phytochemicals reduce the toxicity of copper by donor proton (hydrogen ion) to neutralize the free radical, and it reduces the formation of ROS [15,16].

Acute inflammation is characterized by the accumulation of fluid and plasma at the affected site (due to vasodilation), intravascular activation of platelets, and polymorph nuclear neutrophils as inflammatory cells. Meanwhile, chronic inflammation is the presence of chronic inflammatory cells such as lymphocyte, plasma cells, and macrophages, granulation tissue formation, and in a specific situation as granulomatous inflammation. When the causative agent successfully eliminated by the inflammation, the inflamed area was resolution instead of necrosis [17]. This study showed that increasing white turmeric extract dosage improved the acute inflammation, but it induced chronic inflammation by the increasing number of macrophage cells [17].

The result of this study was supported by the study performed by Fiska et al. They reported that white turmeric protected the kidney from the impact of copper by reducing the level of ureum and creatinine serum and improving the histology changes in the kidney. Another study performed by Chiuman et al. also showed that prolonged consumption of white turmeric increased granulocyte level based on a hematologic study among copper-induced rats. Meanwhile, Ongko et al. reported that the white turmeric has a potential genotoxic property [6-9].

**Fig. 1:** Neutrophile infiltration in White Pulp at ∗100 magnified (left) and ∗400 magnified (right). The black pointer pointed the neutrophile cell. Staining: HE Staining

**Fig. 2:** Macrophage cell in two different magnified: ∗100 (Left) and ∗400 (Right). The black pointer pointed the macrophage cell. Stain: HE Staining
CONCLUSION
It can be concluded that white turmeric may acutely protect spleen from the impact of copper. The increasing of white turmeric dosage shows better splenic protection against copper. However, the higher and prolonged consumption of white turmeric may induce chronic inflammation.

ACKNOWLEDGMENTS
The author(s) disclosed receipts of the financial support for the research and/or publication of this article.

AUTHORS’ CONTRIBUTION
Linda Chiuman – Writing manuscript, evaluation of microscopic view, and final approval. Fahrul Azmi Tanjung – Data collection, data analysis, interpretation of data, and final approval. Djamin – Writing manuscript, Concept and designing the study, data collection, and final approval.

CONFLICT OF INTEREST
The authors declare that there was no conflict of interest.

AUTHORS FUNDING
This study was fully funded by the Ministry of Research, Technology, and Higher Education by Funding Contact No. 282/LLI/PG/2020.

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