Novel Single Nucleotide Polymorphism Markers for Low Dose Aspirin-Associated Small Bowel Bleeding

Akiko Shiotani1*, Takahisa Murao1, Yoshihiko Fujita2, Yoshinori Fujimura3, Takashi Sakakibara3, Kazuto Nishio2, Ken Haruma1

1 Division of Gastroenterology, Department of Internal Medicine, Kawasaki Medical School, Kurashiki City, Okayama, Japan, 2 Department of Genome Biology, Kinki University Faculty of Medicine, Sayama City, Osaka, Japan, 3 Department of Gastroenterology, Sakakibara Heart Institute of Okayama, Okayama City, Okayama, Japan

Abstract

Background: Aspirin-induced enteropathy is now increasingly being recognized although the pathogenesis of small intestinal damage induced by aspirin is not well understood and related risk factors have not been established.

Aim: To investigate pharmacogenomic profile of low dose aspirin (LDA)-induced small bowel bleeding.

Methods: Genome-wide analysis of single nucleotide polymorphisms (SNPs) was performed using the Affymetrix DMET™ Plus Premier Pack. Genotypes of candidate genes associated with small bowel bleeding were determined using TaqMan SNP Genotyping Assay kits and direct sequencing.

Results: In the validation study in overall 37 patients with small bowel bleeding and 400 controls, 4 of 27 identified SNPs: CYP4F11 (rs1060463) GG (p=0.003), CYP2D6 (rs28360521) GG (p=0.02), CYP24A1 (rs4809957) T allele (p=0.04), and GSTP1 (rs1695) G allele (p=0.04) were significantly more frequent in the small bowel bleeding group compared to the controls. After adjustment for significant factors, CYP2D6 (rs28360521) GG (OR 4.11, 95% CI. 1.62 -10.4) was associated with small bowel bleeding.

Conclusions: CYP4F11 and CYP2D6 SNPs may identify patients at increased risk for aspirin-induced small bowel bleeding.

Citation: Shiotani A, Murao T, Fujita Y, Fujimura Y, Sakakibara T, et al. (2013) Novel Single Nucleotide Polymorphism Markers for Low Dose Aspirin-Associated Small Bowel Bleeding. PLoS ONE 8(12): e84244. doi:10.1371/journal.pone.0084244

Introduction

Aspirin had long been regarded as safe beyond the duodenum because of its rapid absorption and lack of an enterohepatic recirculation [1]. However, in recent studies, small bowel injury and enteropathy associated with low dose aspirin (LDA) are increasingly being recognized [2-5]. In a recent Japanese retrospective cohort study of patients taking LDA, the incidence of overt GI bleeding and suspected of small bowel blood loss was 3.9% (upper GI 3.1%, colon 0.7%, and small bowel 0%) and 1.4%, respectively [6]. In another retrospective study of patients undergoing LDA therapy following percutaneous coronary intervention (PCI), the incidence of GI bleeding within 30 days after PCI was 4.3% including occult bleeding (1.9%), small bowel ulcer (0.5%), and overt not upper GI bleeding(0.5%) indicating that attention must be paid to small bowel and lower GI bleeding as well as upper GI bleeding [7].

LDA is now widely used for primary and secondary prevention of cardiovascular events [8-10]. The thienopyridine derivatives, ticlopidine or clopidogrel are also regarded as mandatory following PCI at least until the coronary stents are fully endothelialized which requires approximately 3 months for bare metal stents and up to 1 year for drug-eluting stents [11]. One concern regarding prolonged antiplatelet or anticoagulant (antithrombotic) therapy is the risk of gastrointestinal (GI) bleeding including bleeding from the small intestine [10-13]. However, the pathogenesis and risk factors for small bowel damage induced by aspirin are not also well understood making prevention difficult.

In this era of personalized medicine, the technology allows one to identify genetic risk factors in relation to side effects of medical therapy. To date, there are few studies of the
association between genetic polymorphisms and the risks of aspirin-induced ulcer or its complications[14-17]. Individuals with two single nucleotide polymorphisms (SNPs) of cyclooxygenase-1 (COX-1), A-842G and C50T, exhibit increased sensitivity to aspirin and lower prostaglandin synthesis capacity but the polymorphism lacked statistical significance in relation to an association with bleeding peptic ulcer [18]. Our recent Japanese study suggested that the SLC01B1 521TT genotype and the SLC01B1 *1b haplotype were significantly associated with the risk of peptic ulcer and ulcer bleeding in patients taking low dose enteric-coated aspirin [17]. However, data on the related factors with small intestinal or lower GI events among the patients taking LDA are lacking. Particularly, to our knowledge the association between genetic markers and risk of small bowel bleeding induced by NSAIDs including LDA are still lacking. Therefore, the aim of the present study was to identify the genetic markers related with small bowel bleeding among the patients taking LDA.

Materials and Methods

Study subjects consisted of patients taking 100 mg of enteric coated aspirin (Bayer Health Care, Osaka, Japan) and suspected of bleeding from the small intestine and controls. All patients with at least a one year history of aspirin and or at least a 3 month history of anti-thrombotics use were entered. The study was conducted in accordance with the Declaration of Helsinki from 2007 January to 2012 April at the hospital of Kawasaki Medical School and Sakakibara Heart Institute of Okayama, Japan. The permission was granted by the Ethics Committees of both hospitals, and written informed consent was obtained from each patient.

Subjects

Patients who had complaints of fresh GI bleeding or exacerbating anemia with positive fecal occult blood test had undergone abdominal ultrasonography, upper GI endoscopy, and total colonoscopy. If the patients had no identified source for GI bleeding, bleeding from the small intestine was suspected. All patients with suspected bleeding from the small intestine underwent video capsule endoscopy (VCE) within one month and the diagnosis of aspirin induced enteropathy was made by VCE findings such as multiple erosions and/ or ulcers. Outpatients taking 100 mg of enteric-coated aspirin who had no complaint of GI bleeding or exacerbating anemia and had no identified source for bleeding or peptic ulcer by upper GI endoscopy were included as controls. Patients were excluded if they had lesions identified as causing small bowel bleeding such as malignant, tumorous, inflammatory, or vascular lesions. Patients were also excluded if they had gastric cancer or other malignant lesions.

Demographic data were collected at entry included age, gender, alcohol and smoking consumption, and drug treatments including doses and internal use periods. These data were collected by interview using structured questionnaires and from the patients' clinical records. The most evaluated medicines were continued more than 3 years including aspirin, and the all evaluated medicines were confirmed to be unchanged from others within 2 months. The medicines which had been prescribed just before endoscopy or VCE were not evaluated.

Genotyping

Genomic DNA was extracted from 200 μL of peripheral blood using QiAamp DNA Blood Mini kits (QiAGEN GmbH, Hilden, Germany).

 Genome-wide analysis of SNPs. Seventeen patients taking enteric coated LDA and suspected bleeding from small intestine and 18 patients taking aspirin without bleeding and peptic ulcer who were matched with age, sex, medicine taken and diseases were enrolled. Genome-wide analysis of SNPs was performed using Affymetrix DMET™ Plus Premier Pack. The DMET™ Plus GeneChip array (Affymetrix Inc, Santa Clara, CA) contains 1931 SNPs and five Copy and number Variants (CNVs) distributed on 225 drug metabolizing enzymes and transporters genes. Amplified and non-amplified DNA samples were combined for the annealing and amplification steps, in which molecular inversion probes (MIP) technology was exploited to genotype all the genomic sites of interest in a single reaction. DNA samples were subsequently purified, fragmented, labeled and hybridized to the array to be scanned with the Gene Chip Scanner 3000 (Affymetrix Inc, Santa Clara, CA).

Quality control. Before proceeding to the analysis, we performed quality control checks on the data. First, we tested the concordance between the genetic and reported sex to check for errors in labeling the samples. Second, all subjects showing a genotype call rate <95% were removed. Third, SNPs mapping on the regions of interest (i.e. containing the drug metabolism genes, about 6000 SNPs) were removed if there Hardy-Weinberg p-value was less than 0.00001.

Database submission of microarray data. The microarray data were deposited in the Gene Expression Omnibus (GEO) database: http://www.ncbi.nlm.nih.gov/geo/. The GEO accession number for the platform is GSE52155.

Validation of the candidate SNPs. Genotypes of candidate genes associated with small bowel bleeding were determined by using TaqMan SNP Genotyping Assay kits (Life Technologies, Carlsbad, CA) by following the manufacturer’s instructions and were confirmed by direct sequencing.

Video capsule endoscopy(VCE)

Subjects fasted for 12 h before swallowing the video capsule and data were collected for 8 h-16h after ingestion. Water and lunch of choice were permitted 2 h and 4 h after capsule ingestion, respectively. Two well-trained physicians who remained blinded to the patients’ groups separately reviewed each of the procedures for intestinal injury and identified all suspected lesions by recording as thumb-nail photographs using RAPID® Access 5 or 6.5 software (Given Diagnostic Imaging System, Given Imaging, Tokyo). Each procedure was separately reviewed by recording as thumb-nail photographs. Adjudication was performed by at least four investigators simultaneously reviewing the thumb-nail photographs and by knowledge of the results of other diagnostic imaging such as double-balloon enteroscopy, ultrasonography, computed
tomography, etc. The patients were asked to repeat VCE, if procedure was incomplete because of a large quantity of residue or blood and persistence of the capsule in the upper GI.

Analysis
Values were expressed as the mean ± standard deviation (SD). The differences of age and body mass index were analyzed by unpaired t test, and Mantel-Haenszel statistics were used to assess the differences in the other demographic and clinical characteristics. The odds ratio (OR) and 95% confidence interval (CI) were obtained by Mantel-Haenszel statistics and multiple logistic regression analysis to identify the risk or preventive factors after adjustment for the other significant factors determined by univariate analysis. Differences in the genotype frequencies between the two groups and Hardy-Weinberg equilibrium of allele frequencies at individual loci by comparing the observed and expected genotype frequencies were assessed using the chi-square test or the Fisher’s exact probability test. A two-sided p value of less than 0.05 was considered statistically significant. All statistical computations were performed using SPSS (version 11.0 for Windows, SPSS Inc, Chicago, IL).

Results
Patients’ characteristics
The genome-wide analysis group consists of 17 of the 37 patients with bleeding and 18 of the 400 controls of the study, and a total of 437 Japanese patients (287 men and 150 women; 42-91 years old; average age 71 years) were enrolled. The study groups consisted of 37 patients with suspected bleeding from small intestine (the bleeding group) and 400 controls. Demographic and clinical characteristics are shown in Table 1. Age, Sex, drinking, or body mass index, complication of diabetes mellitus or renal failure was not significantly different between the small bowel bleeding group and controls (Table 1). The prevalence of active smokers was significantly higher (35.1% vs. 11%, p<0.001) in the bleeding group compared to controls. The percentage of the patients with ischemic heart disease treated with aspirin was significantly lower (56.8% vs. 73.5%, p=0.03) and those with non-cardiac vascular diseases including cerebrovascular diseases (24.3% vs. 6.8%, p<0.001) were significantly higher in the bleeding group compared to controls. The percentages of the patients taking warfarin (43.2% vs. 26%, p=0.03) and NSAIDs (16.2% vs. 3.0%, p<0.001) in the bleeding group were significantly higher than those in controls, but the other medicines including proton pump inhibitor (PPI) were not associated with small bowel bleeding (Table 1).

Candidate SNPs discriminated by DMET
Among the 1,936 SNPs included in the DMET system, we obtained the genotyping data of 1,771 SNPs with 100% call rate; 1,215 SNPs were identical in all patients tested. As a result, we used genotyping data of the remaining 556 SNPs for statistical analysis. The SNPs detected to be significantly associated with small bowel bleeding using DMET included 27 SNPs of 23 genes and are listed in Table 2.

Validation study
The candidate SNPs were successfully evaluated in the total of 437 patients, and 55SNPs of 4 genes were significantly associated with small bowel bleeding (Tables 3, 4). The CYP4F11 20043G>A (D446N) and 4927T>C(T106I) alleles were in almost complete linkage disequilibrium. The frequencies of the CYP4F11 20043GG (70.3% vs. 43.5%, p=0.003), CYP2D6 -2178GG (45.9% vs. 25.2%, p=0.02), CYP24A1 18948 T allele (86.5% vs. 68%, p=0.04), and GSTP1*Bc.313 G allele (37.9% vs. 23.3%, p=0.04) were significantly higher in the bleeding subjects than in controls (Table 4).

Factors associated with small bowel bleeding
Smoking (adjusted OR 8.46, 95% CI 3.09-23.2), non-cardiac vascular diseases (6.59, 2.48-17.5), co-treatment of warfarin (3.59, 1.41-9.14), and GG homo-genotypes of CYP2D6 -2178 rs28360521 (4.11, 1.62-10.4) were significantly associated with small bowel bleeding in multivariate analysis after adjustment of the significant factors in univariate analysis (Table 5). NSAIDs, GG homo-genotypes of CYP4F11 20043 rs1060463, and GSTP1*Bc.313 G allele rs1695 were not significantly associated with small bowel bleeding.

Discussion
Our study attempted to identify genetic risk factors associated with small bowel bleeding. Although 27 candidate SNPs of 23 genes were identified by DMET™ Plus GeneChip array, in our validation study, LDA associated small bowel bleeding occurred more often in the patients carrying the GG homo-genotypes of CYP4F11 (rs1060463) or CYP2D6 (rs28360521), the patients carrying T allele of CYP24A1 (rs4809957), or G allele of GSTP1 (rs1695). The role of the other SNPs must await additional studies with larger numbers of samples than available from our array study.

Human CYP4F11 is one of the orphan cytochrome P450 with limited information regarding heterologous expression and functional characterization. It has been reported to be located in a cluster with the other five members of the Cytochrome P450 4F (CYP4F) subfamily on chromosome 19p13.1-2; its mRNA is expressed mainly in human liver [19]. Cytochrome P450 4F (CYP4F) enzymes are involved in cellular protection and metabolism of numerous small molecules, including drugs, toxins, and eicosanoids [20]. CYP4F11 appears to be involved in arachidonic acid and fatty-acid metabolism and catalyzes ω-hydroxylation of leukotriene B4, lipoxins A4 and B4, and hydroxyeicosatetraenoic acids (HETEs) (ie, metabolites with roles in many biological processes including platelet aggregation). Although CYP4F11 activities are much lower than those of CYP4F3 [21], CYP4F11 may have a role in platelet aggregation which would provide a theoretically basis for it being related to small bowel bleeding induced by LDA. However, its function is still not well characterized and there was no study investigating the biologic basis of CYP4F11.
SNPs in this study. The SNP of CYP4F11 (rs1060463) was the most significantly associated with small bowel bleeding among the identified SNPs. However, in multivariate analysis it lost significance as well as taking NSAIDs which is well known risk factor for small bowel injury and bleeding. There may be some interactional association between NSAIDs use and the CYP2D6 SNPs with LDA induced small bowel bleeding, and there is no report indicating the association of the SNPs with not only small bowel bleeding, but also upper GI bleeding. Moreover, we failed to confirm the significant association of the previously identified SNPs related with LDA induced peptic ulcer with small bowel bleeding (Data is not shown). The pathogenesis of small intestinal damage induced by NSAIDs including aspirin is not well understood. Based on the animal studies and in vitro studies, the same mechanisms in the stomach such as topical irritant properties of NSAIDS inducing direct mucosal toxicity, mitochondrial damage, breakdown of intercellular integrity has been assumed [25]. However, enteric bacteria is thought to cause and exacerbate the intestinal damage elevating intestinal permeability by activating neutrophils, generating induction of inducible nitric oxide synthesis [28-30]. The further studies how

### Table 1. Demographic and clinical characteristics of the patients.

| Variable                        | Controls n=400 (%) | Bleeding n=37 (%) | P     |
|---------------------------------|--------------------|-------------------|-------|
| Mean age yr (SD)                | 70(8.4)            | 72(10.3)          | 0.43  |
| Over 80 yr of age (%)           | 67(15.7)           | 8(23.5)           | 0.23  |
| Sex Male (%)                    | 263(65.8)          | 24(73)            | 0.47  |
| Active alcohol drinking (%)     | 118(29.5)          | 9(24.3)           | 0.83  |
| Active smoking (%)              | 44(11)             | 13(35.1)          | <0.001|
| Body mass index (SD)            | 23.5(3.1)          | 23.0 (4.0)        | 0.40  |
| Ischemic heart diseases (%)     | 294(73.5)          | 21(56.8)          | 0.03  |
| Other cardiac diseases (%)      | 88(22)             | 6(16.2)           | 0.67  |
| Cerebrovascular diseases (%)    | 27(6.8)            | 9(24.3)           | <0.001|
| Other vascular disease (%)      | 14(3.5)            | 6(16.2)           | <0.001|
| Diabetes mellitus (%)           | 115(28.8)          | 11(29.7)          | 0.84  |
| Chronic renal failure (%)       | 6 (1.5)            | 2(5.4)            | 0.13  |
| Warfarin                        | 104(26)            | 16(43.2)          | 0.03  |
| Thienopyridine                  | 111(27.8)          | 13(35.1)          | 0.34  |
| PPI                             | 145(36.3)          | 13(35.1)          | 0.90  |
| H2-RA                           | 133(33.3)          | 9(24.3)           | 0.27  |
| Ca-blocker                      | 158(39.5)          | 11(29.7)          | 0.25  |
| ARB or ACE inhibitor            | 231(57.8)          | 20(54.1)          | 0.66  |
| Statin                          | 209(52.3)          | 15(40.5)          | 0.17  |
| Nitrile                         | 113(28.3)          | 13(35.1)          | 0.37  |
| NSAID                           | 12(3.0)            | 6(16.2)           | <0.001|

*p Values; age and body mass index by unpaired t test and others by Mantel-Haenszel Chi square analyses. p Values by Mantel-Haenszel Chi square analyses; Thienopyridine, ticlopidine 200 or 300 mg/day or clopidogrel 50 or 75 mg/day; PPI, proton pompor inhibitor, omeprazole 10 mg/day, lansoprazole 15mg/day; H2-RA, H2-receptor antagonist, famotidine 10 or 20mg/day, Ca-blocker, nifedipidine 20 or 40mg/day, amiodipine 2.5mg or 5mg/day; ACE inhibitor, angiotensin-converting enzyme inhibitor, imidapril 5mg/day; ARB, angiotensin type 1 receptor blocker, candesartan 4 or 8 mg/day, telmisartan 40mg/day, olmesartan 20mg/day, Statin (HMG-Co A reductase inhibitor), pravastatin 10mg/day, atorvastatin 10mg/day, Rosuvastatin 5mg/day; Nitrile, carvedilol 20 mg/day, metoprolol 40mg/day; NSAID, Non-steroidal anti-inflammatory drugs internal use only.

doi:10.1371/journal.pone.0084244.t001
the SNPs are involved in LDA induced enteropathy or bleeding are required.

In the present study, active smoking, non-cardiac vascular diseases including cerebrovascular diseases warfarin, and NSAIIDs were significantly associated with small bowel bleeding among the patients taking LDA. Drug combinations involving antiplatelets and anticoagulants are known to lead to a greatly increased risk of GI bleeding, and the prescribing of

### Table 2. Lists of discriminating polymorphisms associated with enteropathy using DMET.

| Common Name | Chromosome | dbSNPRs ID | Alleles | p value |
|-------------|------------|------------|---------|---------|
| CYP2A7_c.821A>G(H274R) | 19 | rs4079366 | A // G | 0.000919 |
| CYP2B6*27_15708T>C(M198T) | 19 | rs36079186 | C // T | 0.003325 |
| XDH_c.3030C>T(F1010F) | 2 | rs1884725 | C // T | 0.003889 |
| CYP2B6*16_15631G>T(Q172H) | 19 | rs3745274 | T // G | 0.004337 |
| CYP2B6*4_18053A>G(K262R) | 19 | rs2279343 | A // G | 0.006724 |
| SLC10A2_c.315G>T(A171S) | 13 | rs188096 | T // G | 0.009141 |
| GSTA2_c.335G>C(S112T) | 6 | rs2180314 | T // G | 0.010715 |
| SLC10A2_c.*315G>T | 13 | rs279941 |  |  |
| CYP2B6*6_1_15631G>T(Q172H) | 19 | rs3745274 | T // G | 0.004337 |
| GSTA2_c.335G>C(S112T) | 6 | rs2180314 | T // G | 0.010715 |
| SLC10A2_c.511G>T(A171S) | 13 | rs188096 | G // T | 0.009141 |
| CYP2B6*4_18053A>G(K262R) | 19 | rs2279343 | A // G | 0.006724 |
| CYP2B6*6_15631G>T(Q172H) | 19 | rs3745274 | T // G | 0.004337 |
| SLC10A2_c.315G>T(A171S) | 13 | rs188096 | T // G | 0.009141 |
| GSTA2_c.335G>C(S112T) | 6 | rs2180314 | T // G | 0.010715 |
| SLC10A2_c.511G>T(A171S) | 13 | rs188096 | G // T | 0.009141 |
| CYP2B6*4_18053A>G(K262R) | 19 | rs2279343 | A // G | 0.006724 |
| CYP2B6*6_15631G>T(Q172H) | 19 | rs3745274 | T // G | 0.004337 |
| SLC10A2_c.315G>T(A171S) | 13 | rs188096 | T // G | 0.009141 |

### Table 3. Allele and genotype frequencies of the candidate genes.

| Genotype | Allele frequencies | Controls | Bleeding | p |
|----------|--------------------|----------|----------|---|
| CYP2A7 | GG | G=0.66 | 174(43.5%) | 26 (70.3%) | 0.007 |
| CYP2B6*16_15631G>T(Q172H) | GA | A=0.34 | 178(44.5%) | 9 (24.3%) |
| rs1060463 | AA | p_a =0.82 | 48 (12%) | 2 (5.4%) |
| CYP2D | GG | G=0.52 | 101(25.2%) | 17 (45.9%) |
| CYP4F11 | TT | C=0.43 | 65 (16.2%) | 7(20%) |
| rs1799931 | AA | p_a =0.85 | 94(23.5%) | 5 (13.5%) |
| CYP24A1 | CC | p_a =0.61 | 128 (32.8%) | 5(13.5%) |
| GSTP1*B | AG | G=0.14 | 86 (21.5%) | 12 (32.4%) |

p Values by using the chi-square test; a, Hardy-Weinberg equilibrium (HWE) of allele frequencies at individual loci was assessed by comparing the observed and expected genotype frequencies.
aspirin with warfarin was reported to increase risk of GI bleeding than that observed with each drug alone [31]. There is no report indicating association of small bowel bleeding with smoking. Moreover, there is no evidence indicating that the prevalence of GI bleeding was more frequent in aspirin users with non-cardiac vascular diseases compared to those with cardiac disease, and the risk of GI bleeding seems to be similar according to the previous studies although there is no data comparing the risk of small bowel bleeding [32]. The significant results of underlying disease treated by LDA possibly were caused by selection bias. Wallace JL et al [30] recently reported PPIs significantly exacerbated NSAIDs-induced intestinal ulceration and bleeding in the rat. However, in the present study there was no association between PPI use and small bowel bleeding. The limitation of our study is case control and the small number of patients with small bowel bleeding, and the large scale clinical cohort studies are absolutely required to investigate the possibility of this important issue.

Although our data needs to be validated and extended in a larger cohort, this exploratory study suggests that CYP4F11 and CYP2D6 SNPs may identify patients at increased risk for aspirin-induced small bowel bleeding.

**Acknowledgements**

We thank Ms. Maki Nomura and Ms. Tomoko Yobimoto (Kawasaki Medical School, Okayama, Japan) for assistance of laboratory work.

**Author Contributions**

Conceived and designed the experiments: AS KN KH. Performed the experiments: AS TM Y. Fujita. Analyzed the data: AS Y. Fujita KN. Contributed reagents/materials/analysis tools: AS TM Y. Fujimura TS. Wrote the manuscript: AS Y. Fujita.

**References**

1. Shiotani A, Kamada T, Haruma K (2008) Low-dose aspirin-induced gastrointestinal diseases: past, present, and future. J Gastroenterol 43: 581-588. doi:10.1007/s00535-008-2206-5. PubMed: 18709479.
2. Endo H, Hosono K, Inamori M, Kato S, Nozaki Y et al. (2009) Incidence of small bowel injury induced by low-dose aspirin: a crossover study using capsule endoscopy in healthy volunteers. Digestion 79: 44-51. doi:10.1159/000204465. PubMed: 19246922.
3. Endo H, Hosono K, Inamori M, Nozaki Y, Yoneda K et al. (2009) Characteristics of small bowel injury in symptomatic chronic low-dose aspirin users: the experience of two medical centers in capsule endoscopy. J Gastroenterol, 44: 544-549. doi:10.1007/s00535-009-0040-z. PubMed: 19373431.
4. Smecuol E, Pinto Sanchez MJ, Suarez A, Argonz JE, Sugai E et al. (2009) Low-dose aspirin affects the small bowel mucosa: results of a
16. Shiotani A, Sakakibara T, Nomura M, Yamanaka Y, Nishi R et al. (2010) Aspirin-induced peptic ulcer and genetic polymorphisms. J Gastroenterol Hepatol 25 Suppl 1: S31-S34. doi:10.1111/j.1440-1673.2009.05637.x. PubMed: 1997.v112.pm9041264. PubMed: 9041264.

17. Whittle BJ, László F, Evans SM, Moncada S (1995) Induction of nitric oxide synthase and microvascular injury in the rat jejunum provoked by indomethacin. Br J Pharmacol 116: 51-61. Available online at: doi:10.1039/gast.1997.v112.pm9041264. PubMed: 9041264.

18. Shiotani A, Murao T, Sakakibara T, Tarumi K, Manabe N et al. (2012) Association of SLC01B1 1b with peptic ulcer amongst Japanese patients taking low-dose aspirin. Dig Liver Dis 44: 201-205. doi:10.1016/j.dld.2011.06.057. PubMed: 22098426.

19. van Oijen MG, Laheij RJ, Koetsier M, de Kleine E, Te Morsche RH et al. (2006) Effect of a specific cytochrome P450 polymorphism (A-842G/C50T) on the occurrence of peptic ulcer hemorrhage. Dig Dis Sci. 51: 2348-2352. doi:10.1007/s10620-006-9475-8. PubMed: 17078001.

20. Guengerich FP, Wu ZL, Bartleson CJ (2005) Function of human cytochrome P450sa: characterization of the orphans. Biochem Biophys Res Commun 338: 465-469. doi:10.1016/j.bbrc.2005.08.079. PubMed: 16126164.

21. Nebert DW, Russell DW (2002) Clinical importance of the cytochromes P450. Lancet 360: 1155-1162. doi:10.1016/S0140-6736(02)11203-7. PubMed: 12379068.

22. Kalsotra A, Turman CM, Kikuta Y, Strobel HW (2004) Expression and characterization of human cytochrome P450 4F11: Putative role in the metabolism of therapeutic drugs and eicosanoids. Toxicol Appl Pharmacol 199: 295-304. doi:10.1016/j.taap.2003.12.033. PubMed: 15364546.

23. Zanger UM, Raimundo S, Eichelbaum M (2004) Cytochrome P450 2D6: overview and update on pharmacology, genetics. Biochemistry - Naunyn Schmiedebergs Arch Pharmacol, 369: 23-37. doi:10.1007/s00210-003-0862-2.

24. Dagle JM, Fisher TJ, Haynes SE, Brophy PD et al. (2011) Cytochrome P450 (CYP2D6) genotype is associated with elevated systolic blood pressure in preterm infants after discharge from the neonatal intensive care unit. J Pediatr 158: 104-109. doi:10.1016/j.jpeds.2011.01.002. PubMed: 21353244.

25. Yin S, Shen L, Zhang A, Xie J, Shen W et al. (2008) Systematic polymorphism analysis of the CYP2D6 gene in four different geographical Han populations in mainland China. Genomics, 92(3): 152-158. doi:10.1016/j.ygeno.2008.05.004. PubMed: 18632250.

26. Wallace JL (1997) Nonsteroidal anti-inflammatory drugs and gastrecteropathy: the second hundred years. Gastroenterology, 112: 1000-1015. Available online at: doi:10.1053/gast.1997.v112.pm9041264. PubMed: 9041264.

27. Reuter BK, Davies NM, Wallace JL (1997) Nonsteroidal anti-inflammatory drug enteropathy in rats: role of permeability, bacteria, and enterohepatic circulation. Gastroenterology, 112(1): 109-117. doi:10.1016/S0016-5085(97)70025-7. PubMed: 8978349.

28. Wax J, Clingen WA, Varner P, Bass P, Winder CV (1970) Relationship of the enterohepatic cycle to ulcerogenesis in the rat small bowel with inflammatory bowel disease. Gastroenterology, 58: 772-780. PubMed: 5423889.

29. Kent TH, Cardelli RM, Stamler FW (1969) Small intestinal ulcers and intestinal flora in rats given indomethacin. Am J Pathol 54: 237-249. PubMed: 5765865.

30. Whittle BJ, László F, Evans SM, Moncada S (1995) Induction of nitric oxide synthase and microvascular injury in the rat jejunum provoked by indomethacin. Br J Pharmacol 116: 2286-2290. doi:10.1111/j.1476-5381.1995.tb15066.x. PubMed: 8564261.

31. Wallace JL, Syer S, Denou E, de Palma G, Vong L, et al. (2011) Proton pump inhibitors exacerbate NSAID-induced small intestinal injury by inducing dysbiosis. Gastroenterology 141:1314-1322. 1322 e1311-1315.

32. Delaney JA, Opatrny L, Brophy JM, Suissa S (2007) Drug drug interactions between antiarthritic medications and the risk of gastrointestinal bleeding. CMAJ 177: 347-351. PubMed: 17698822.

33. Derry S, Loke YK (2000) Risk of gastrointestinal haemorrhage with long term use of aspirin: meta-analysis. BMJ 321: 1183-1187. doi:10.1136/bmj.321.7270.1183. PubMed: 11073508.