Are clarithromycin, azithromycin and their analogues effective in the treatment of COVID-19?

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ABSTRACT

BACKGROUND: SARS-CoV-2, which started in Wuhan and later affected the whole world, is the most important disease of the world today. Many ways to inhibit SARS-CoV-2 virus are sought to prevent the spread of this virus. Azithromycin and clarithromycin are considered for the treatment of the SARS-CoV-2 virus, which has a high similarity to previous colonic diseases.

AIM: We aimed to determine whether azithromycin and clarithromycin, the RNA-dependent RNA polymerase protein inhibitor used in the treatment of COVID-19, is effective against SARS-Cov-2 in silico.

RESULTS AND CONCLUSION: The 503 analogues of azithromycin and clarithromycin were studied to target SARS-CoV-2 the RNA-dependent RNA polymerase protein inhibition. Maestro program was used to compare the inhibition activities of these analogues. A detailed comparison was made using the numerical value of many parameters obtained. ADME / T properties were then examined to determine the effects and reactions of analogues on human metabolism. In this study, the SARS-CoV2 virus is 6NUR and 6NUS, which is the RNA-dependent RNA polymerase protein. Among these proteins, the best inhibitor among the 503 analogues according to the docking score parameter was 9851445 with a great difference. This analogue was an analogue of azithromycin (Tab. 3, Fig. 6, Ref. 58).

KEY WORDS: SARS-CoV-2, RNA-dependent RNA polymerase, Azithromycin, Clarithromycin, COVID19.

Introduction

At the end of December 2019, a new outbreak occurred in Wuhan, Hubei province, in China, with unknown causes and treatment-resistant pneumonia findings. It was observed that the agent that caused this outbreak belonged to the same subfamily as SARS-CoV, which is a member of the Coronaviridae family and originated in China in 2002 (1).

Coronaviruses are enveloped RNA viruses. There are many subtypes of coronaviruses. Six subtypes of coronaviruses (229E, OC43, NL63, HKU1, SARS-CoV, MERSCoV) are known to cause disease in humans (2). Coronaviruses consists of four structural proteins: nucleocapsid, envelope, membrane, and spines.

On December 31, 2019, the World Health Organization (WHO) China Country Office reported pneumonia cases of unknown aetiology in Wuhan, Hubei province, China. On January 7, 2020, the causative agent was identified as a new Coronavirus (2019-nCoV), which has not previously been detected in humans. Later, the name of 2019-nCoV disease was accepted as COVID-19, and the virus was named as SARS-CoV-2 due to its similarity to SARS-CoV (3). Although various antiviral agents have been tried in its treatment, there is no specific treatment effective against this virus yet mostly symptomatic agents are used.

Recently, these drugs have attracted great attention, along with the reveal of the anti-viral effects of macrolides. Both azithromycin and clarithromycin bind to the 50S ribosomal subunit in pathogenic microorganisms, inhibiting protein synthesis. Clarithromycin is more effective against gram positive bacteria and tissue penetration is better than other antibiotics in the macrolide group (4). Clarithromycin is recommended as the first line in pneumonia treatment guides, since clarithromycin is more effective in the macrolide group as an antibiotic treatment in pneumonia and especially during epidemic periods, and it is better to use it in addition to penicillin group antibiotics (5). In a limited number of studies, the use of clarithromycin with hydroxychloroquine has been shown to be effective in the treatment (6, 7). Combination therapy consisting of oseltamivir, a neuraminidase inhibitor, and clarithromycin or azithromycin, healed a seasonal influenza virus infection earlier. Macrolides such as: erythromycin, clarithromycin and azithromycin have not only anti-bacterial activity, but also have anti-inflammatory and immunomodulatory effects. The efficiency of clarithromycin and azithromycin in the treatment of rhinovirus
and influenza virus has been proven (8). The mechanism of action of these drugs are as followed:

• Immunomodulatory effect on inflammatory cells, fibroblasts and epithelial cells,
• Modulation of cytokine / chemokine production,
• inhibition of mucus hypersecretion,
• Suppression of transcription factors and inflammatory cytokine gene expression.

Macrolides in the treatment of respiratory infection; 14-ring clarithromycin and 15-ring azithromycin are widely used. In the study on the effect of clarithromycin on viral respiratory infections; In prophylaxis and treatment of monkeys infected with influenza virus, interleukin (IL) -6, IL-1β and IL-8 levels in lung tissues after treatment were shown to be lower than lungs of untreated monkeys (9).

Studies in people infected with Covid-19 have been shown to reduce the viral load with hydroxychloroquine and azithromycin therapy. Gautret et al in his study on patients with COVID-19; patients treated with hydroxychloroquine and azithromycin have been shown to have better results than those treated with hydroxychloroquine alone (10). In some studies, it has been observed that azithromycin alone was not effective against covid-19 in in-vitro environment. However, more effective results have been obtained, when azithromycin and hydroxychloroquine were used together. The authors think that this is due to the synergistic effect of the two drugs (11). The effect of azithromycin in the in vitro environment may differ from the in vivo environment, since in vitro environments cannot completely mimic the in vivo environment. The use of both azithromycin and hydroxychloroquine alone has not been as effective as combined use in clinical trials. However, it should be kept in mind that some patients, who use azithromycin with hydroxychloroquine may develop cardiac arrhythmias (12).

In this study, we aimed to investigate whether clarithromycin and azithromycin are effective molecules against COVID-19. In this study, the activities of analogues of FDA approved of azithromycin and clarithromycin against SARS-CoV2 virus were compared. Molecular structures of 381 analogues of clarithromycin and 122 analogues of azithromycin were downloaded from the PubChem website (https://pubchem.ncbi.nlm.nih.gov/). A total of 503 analogues of these two molecules were studied to target the RNA-dependent RNA polymerase protein of the SARS-CoV-2 virus. The proteins used for this purpose were 5W44 (13), 5FDD (14), 6E6V (15), 6FS8 (16), 6QX8 (17), 6NUR (18), and 6NUS (18). As the result of the calculations, the results of 503 analogues were compared with the results of azithromycin and clarithromycin using the numerical values of the parameters obtained. We tried to find more effective and high-activity molecules against SARS-CoV-2 virus’s RNA-dependent RNA polymerase protein.

Method

In this study, the activities of azithromycin and clarithromycin in total 503 analogues, against SARS-CoV-2 virus RNA-dependent RNA polymerase protein were compared. In the calculations for this comparison, molecular docking calculations were made in the interaction of the SARS-CoV-2 virus RNA-dependent RNA polymerase protein with a total of 503 analogues of azithromycin and clarithromycin. Maestro Molecular Modelling platform (version 12.2) by Schrödinger, LLC (19) was used for these calculations. The calculations made on the Maestro Molecular modelling platform consisted of several stages. Each stage is called a module, each of which works for a different task. The first module used in the Maestro Molecular modelling platform is the protein preparation module (20, 21), which is used to prepare the studied proteins for calculations.

In the protein preparation module, interactions of molecules with many others have been studied. Studied proteins were downloaded from the protein data bank site. IDs of these proteins are Crystal structure of the influenza virus PA endonuclease in complex (ID:5W44), Endonuclease inhibitor 1 bound to influenza strain H1N1 polymerase (ID:5FDD), The N-terminal domain of PA endonuclease from the influenza H1N1 virus (ID:6E6V), Influenza B/Memphis/13/03 endonuclease (ID:6FS8), Influenza A virus (ID:6QX8), SARS-Coronavirus NSP12 bound to NSP7 and NSP8 co-factors (ID:6NUR), and SARS-Coronavirus NSP12 bound to NSP8 co-factor (ID:6NUS). By using this module, at first, the water molecules in the structures of the proteins were deleted. Afterwards, optimizations of these proteins were made, binding methods and charges of proteins were calculated. After this optimization process, active regions of these proteins were determined. The proteins in the active regions of the studied proteins were given a mobility for interaction, because the mobility-free proteins can interact more easily with 503 analogues.

After that, a preparation of e for 503 analogues calculations with another module started, in the LigPrep module (22, 23). Physiological pH values (pH = 5 ± 4) of 503 analogues were calculated within this module. At this pH value, 3D structures of high-energy isomers of 503 analogues in accurate protonation were obtained and minimized at OPLS3e method.

The Glide ligand docking module was used to calculate the interactions of RNA-dependent RNA polymerase proteins of the SARS-CoV-2 virus with the 503 analogues. Many parameters were obtained from the calculations made using this module. Inhibition activities of molecules were predicted from the numerical values of these parameters. After interactions of 503 analogues with proteins, the parameters obtained were compared. ADME/T analysis (absorption, distribution, metabolism, excretion and toxicity) of molecules with a higher inhibition activity than the reference molecules azithromycin and clarithromycin were performed after this comparison. ADME/T analysis calculations were made with the Qik-prop module (24) of the Schrödinger software. The most important reason for these calculations with the Qik-prop module was that it was a guide for in vitro and in vivo experiments, because with these calculations, the effects and reactions of molecules in human metabolism were predicted.

Result and discussion

In this study, the activities of azithromycin and clarithromycin in total 503 analogues were compared against the RNA-dependent RNA polymerase protein of the SARS-CoV-2 virus. It should be
known very well that as the interaction of these 503 analogues with the RNA-dependent RNA polymerase protein of the SARS-CoV-2 virus, the inhibition activity of the molecule with a higher interaction will be higher (25-27). The RNA-dependent RNA polymerase protein of the SARS-CoV-2 virus of the molecule with a high inhibition activity will adhere more. This will prevent the SARS-CoV-2 virus from replicating the RNA-dependent RNA polymerase protein itself, and this virus will be isolated. In this direction, many parameters have been obtained in the molecular docking calculations.

Azithromycin (PubChem ID: 447043) whose UI-PAC name is (2R,3S,4R,5R,8R,10R,11R,12S,13S,14R)-11-
[(2S, 3R, 4S, 6R)-4-(dimethylamino)-3-hydroxy-6-methyl-oxan-2-yl]oxy-2-ethyl-3,4,10-trihydroxy-13-[(2R, 4R, 5S, 6S)-5-hydroxy-4-methoxy-4,6-dimethyloxan-2-yl]oxy-3,5,6,8,10,12,14-heptamethyl-1-oxa-6-azacyclopentadecan-
15-one and clarithromycin (PubChem ID: 84029) whose UIPAC name is (3R, 4S, 5S, 6R, 7R, 9R, 11R, 12R, 13S, 14R)-6-[(2S, 3R, 4S, 6R)-4-(dimethylamino)-3-hydroxy-6-metyl-
voxan-2-yl]oxy-14-ethyl-12,13-dihydro-
xy-4-[(2R, 4R, 5S, 6S) -5-hydroxy-4-me-
thoxy-4,6-dimethyloxan-2-yl]oxy-7-me-
thoxy-3, 5, 7, 9, 11, 13-hexamethyl-oxa-
cyclo tetradecane-2,10-dione are presented
in Figure 1.

In this study, the inhibitory activities
of azithromycin and clarithromycin’s 503
analogues against SARS-CoV2 proteins
were compared. In all coronavirus varieties, RNA-dependent RNA polymerase (RdRp) is the most important enzyme in the cell that catalyses the replication of RNA from RNA templates. When the previous studies were examined, severe acute respira-

![Fig. 3. The interaction schema at analogue 9851445 against 6NUR.](image-url)

![Fig. 4. The interaction schema at analogue 9851445 against 6NUS.](image-url)
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...syndrome coronavirus (SARS-CoV), SARS-CoV-2, Middle East respiratory syndrome coronavirus (MERS-CoV), Human coronavirus OC43 (HCoV-OC43), Human coronavirus HKU1 (HCoV-HKU1) RNA-dependent RNA polymerase protein sequences of Human coronavirus NL63 (HCoV-NL63), and Human coronavirus 229E (HCoV-229E) have similar sequences, when compared to RNA-dependent RNA polymerase protein sequences and have structurally similar codes as shown in Figure 2 (28–30).

Molecular docking calculations were made as the result of interaction of these 503 analogues against SARS-CoV2 RNA-dependent RNA polymerase protein. The inhibition activities of these 503 analogues against SARS-CoV2 RNA-dependent RNA polymerase protein were compared. Calculations for this comparison were made using the numerical values of the obtained parameters. The first parameter used for this comparison is the Docking Score (31–34), which is the most important parameter used to compare the inhibition activities of molecules. The molecule, which is the most negative molecule of this parameter, has the highest inhibition activity value. Analogues studied may have more than one stable structure. Accordingly, more than one structure of an analogue can interact with different points in the active

Fig. 5. The interaction schema at analogue 118859401 against 6NUR.

Fig. 6. The interaction schema at analogue 121373522 against 6NUS.
region of proteins. Interactions can occur both with the original structure and with the possible structure of the analogues. The structures of the interactions obtained as the result of the calculations are given in Table 1. If one or more isomers of an analogue interact more than once, the interaction with the most negative value is taken into account in the comparison operations.

Another parameter is the Glide ligand efficiency, which shows the numerical value of the activity of molecules. The next parameter is the Glide hbond (35), which is the numerical value of hydrogen bonds formed by interactions between molecules and proteins. The next parameters are the Glide evdw and Glide ecoul (36), which is a numerical value of Van Der Walls bonds and Coulomb interactions between the molecules and proteins. The next parameter is the Glide emodel (37), which shows the energy of the docking model resulting from the interaction. The next parameter is the Glide energy (38), which is a modified Coulomb-van der
Waals interaction energy. The last parameter is the Glide einternal (39), which is Internal torsional energy.

Interactions between the analogues and protein are the most important factors affecting the inhibition activity of molecules in Figure 3, 4, 5 and 6. Other interactions are provided with supplementary data in Figure S1-S5. These interactions have many interactions such as: hydrogen bonds, polar and hydrophobic interactions, π-π and halogen bonds (40–43).

The results of the molecular docking calculations are evaluated in the Tables 1 and 2. When the inhibition activities of the 503 analogues of azithromycin and clarithromycin against the SARS-CoV2 virus were compared, it was observed that the analogues had a similar order of docking score and the Glide energy parameters. Since the ID numbers of the molecules given in Table 2 are long, their names are as follows; 1 (9851445), 2 (118859401), 3 (118859402), 4 (121370516), 5 (121370518), 6 (121370520), 7 (121373522). These molecules in Table 1 and 2 have been chosen because they have both numerical values of these two parameters and the number of structures of analogues interacting with proteins, which are better than those of other analogues.

In this study, the SARS-CoV2 virus is 6NUR and 6NUS, which is the RNA-dependent RNA polymerase protein. Among these proteins, the best inhibitor among the 503 analogues according to the docking score parameter is 9851445 with a great difference. This analogue is the analogue of azithromycin. On the other hand, 118859401 and 121373522 have the highest inhibitory activity among 381 analogues of clarithromycin, but these analogues did not show a good inhibitory activity like 9851445. Another parameter is the Glide ligand efficiency, the numerical values of this parameter are very similar to the docking score parameter. The numerical value of this parameter is the analogue with the most negative value and the highest biological activity. Accordingly, 9851445 and 118859401 analogues were the most negative analogues for the 6NUR, 9851445 and 121373522 for 6NUS ligand activity. The calculations showed that 9851445, 118859401, and 121373522 analogues indicated that the results against the SARS-CoV2 virus RNA-dependent RNA polymerase protein were better than all the analogues of clarithromycin and azithromycin. These analogues were found to have better docking score parameters than other molecules. In this study, it was found that analogues with the best inhibition activity against proteins were found to stop the virus by blocking the RNA-dependent RNA polymerase protein of the SARS-CoV-2 virus. In this study, 37 out of 503 analogues interacted with proteins, among which 7 the SARS-CoV2 virus RNA-dependent RNA polymerase protein were better than all the analogues of clarithromycin and azithromycin.

| Tab. 3. ADME properties of molecules. |
|--------------------------------------|
| 1       | 2         | 3         | 4         | 5         | 6         | 7       | Reference Range |
|--------------------------------------|
| Solute Molecular Weight              | 590       | 612       | 670       | 626       | 668       | 668     | 720             | 130–725                      |
| Solute Dipole Moment (D)              | 9.2       | 8.22      | 5.70      | 6.78      | 3.79      | 3.79    | 6.37            | 1.0–12.5                     |
| Solute Total SASA                    | 825       | 804       | 803       | 824       | 871       | 871     | 899             | 300–1000                     |
| Solute Hydrophobic SASA              | 678       | 692       | 679       | 693       | 717       | 717     | 774             | 0–750                        |
| Solute Hydrophilic SASA              | 146       | 76        | 97        | 92        | 129       | 129     | 125             | 7–330                        |
| Solute Carbom Ptas SASA              | 0         | 36        | 60        | 40        | 25        | 25      | 0               | 0–450                        |
| Solute Weakly Polar SASA             | 0         | 0         | 0         | 0         | 0         | 0       | 0               | 0–175                        |
| Solute Molecular Volume (Å3)         | 1755      | 1752      | 1892      | 1841      | 1916      | 1916    | 2017            | 500–2000                     |
| Solute as Donor-Hydrogen Bonds       | 5.0       | 2.5       | 2.5       | 2.5       | 1.5       | 1.5     | 5.0             | 0.0–6.0                      |
| Solute as Acceptor-Hydrogen Bonds    | 16.00     | 13.30     | 15.30     | 13.30     | 15.60     | 15.60   | 18.15           | 2.0–20.0                     |
| Solute Globalularity (Sphere =1)     | 0.85      | 0.87      | 0.88      | 0.88      | 0.86      | 0.86    | 0.86            | 0.75–0.95                    |
| QP Polarizability (Angstroms ″3)     | 58.31     | 57.25     | 62.42     | 60.83     | 63.70     | 63.70   | 67.51           | 13.0–70.0                    |
| QP log p for hexadecane/gas          | 16.24     | 15.32     | 16.66     | 16.11     | 16.73     | 16.73   | 18.09           | 4.0–18.0                     |
| QP log p for octanol/gas             | 36.09     | 30.23     | 33.08     | 31.48     | 32.12     | 32.12   | 40.11           | 8.0–35.0                     |
| QP log p for water/gas               | 23.80     | 16.75     | 18.89     | 16.93     | 18.00     | 18.00   | 25.51           | 4.0–45.0                     |
| QP log p for octanol/water           | 1.30      | 3.65      | 3.59      | 4.09      | 3.33      | 3.33    | 2.73            | 2.0–6.5                      |
| QP log S aqueous solubility          | –1.90     | –3.09     | –2.77     | –3.41     | –3.14     | –3.14   | –3.24           | –6.5–0.5                     |
| QP log S-conformation independent    | –1.81     | –4.50     | –4.85     | –4.78     | –4.75     | –4.75   | –4.54           | –6.5–0.5                     |
| QPlogHERG                            | –5.36     | –4.46     | –4.42     | –4.37     | –4.70     | –4.70   | –4.54           | (concern below –5)            |
| QPPCaco (nm/sec)                     | 24        | 274       | 298       | 333       | 149       | 149     | 160             | *                            |
| QPlogBB (nm/sec)                     | –0.52     | –0.36     | –0.59     | –0.49     | –0.87     | –0.87   | –0.83           | –3.0–1.2                     |
| QPPMDCK (nm/sec)                     | 11        | 243       | 148       | 167       | 70        | 70      | 75              | *                            |
| QPlogKp                              | –7.48     | –3.80     | –4.08     | –4.81     | –4.81     | –4.84   | –4.84           | Kp in cm/hr                  |
| IP (ev)                              | 8.67      | 9.41      | 9.33      | 9.21      | 9.12      | 9.12    | 8.90            | 7.9–10.5                     |
| EA (eV)                              | –0.75     | –1.08     | –0.78     | –1.11     | –0.38     | –0.38   | –0.87           | –0.9–1.7                     |
| #metab                              | 8         | 8         | 8         | 8         | 8         | 8       | 7               | 1–8                          |
| QPlogKhsa                            | 0.00      | 0.35      | 0.30      | 0.58      | 0.21      | 0.21    | 0.19            | –1.5–1.5                     |
| Human Oral Absorption                | 2         | 2         | 2         | 2         | 2         | 2       | 2               | –                            |
| Percent Human Oral Absorption        | 33        | 83        | 66        | 83        | 59        | 59      | 56              | **                           |
| PSA                                  | 138       | 100       | 131       | 99        | 157       | 157     | 128             | 7–200                        |
| RuleOfFive                           | 2         | 1         | 2         | 2         | 2         | 2       | 2               | Maximum is 4                  |
| RuleOfThree                          | 1         | 1         | 1         | 1         | 1         | 1       | 1               | Maximum is 3                  |
| Jm                                   | 0.00      | 0.08      | 0.11      | 0.02      | 0.01      | 0.01    | 0.01            | –                            |

* < 25 is poor and > 500 is great, ** < 25 % is poor and > 80 % is high
CoV-2 virus RNA-dependent RNA polymerase protein inhibition activity was higher than the others. After examining the docking parameters of these analogues, ADME/T analysis (Absorption, distribution, metabolism, excretion, and toxicity) (44-47) should be done to examine the effects and responses of these analogues in human metabolism. As the result of this analysis, many parameters were obtained. The numerical values of these parameters showed how these analogues act in human metabolism and how they react in organs.

There are many important parameters among the parameters obtained as the result of ADME/T analysis. The most important of these are: The Solute Molecular Weight (48) parameter tells the numerical value of the molecular weight of the molecules; Solute Hydrophobic SASA (49) is the hydrophobic component of the SASA (saturated carbon and attached hydrogen); The Solute Molecular Volume (50) parameter tells the molecular volume of analogues; Solute as Donor-Hydrogen Bonds (51) shows the numerical value of the number of hydrogen bonds formed between analogues and proteins; QP log p for octanol/gas (52) is predicted octanol/gas partition coefficient; QPlogHERG (53) is predicted IC50 value for blockage of HERG K+ channels; QPPCaco (54) is predicted apparent Caco-2 cell permeability in nm/sec. Caco-2 cells are a model for the gut-blood barrier; QPPMDCK (55) is predicted apparent MDCK cell permeability in nm/sec. MDCK cells are considered to be a good mimic for the blood-brain Barrier. QPlogKhsa (56) is prediction of binding to human serum albumin; RuleOffive (57) is number of violations of Lipinski’s rule of five, which is also known as the 5 rules of Pfizer. The rules are: mol_MW <500, QP Caco/p <5, donorHB ≤5, acceptHB ≤10. Compounds that satisfy these rules are considered drugglike; RuleOffThree (58) is number of violations of Jorgensen’s rule of three. The three rules are: QP logG <−5.7, QP Caco/p >22 nm/s, # Primary Metabolites<7. Considering the numerical values of these parameters, it was observed that they did not provide the conditions for some parameters.

Conclusions

Inhibition activities of azithromycin and clarithromycin analogues against SARS-CoV-2 virus against RNA-dependent RNA polymerase proteins were compared. Against the 6SUR, the SARS-CoV-2 Coronavirus virus protein, the two most negative analogues of the docking score parameter were 9851445 whose IUPAC Name is (2R,3S,4R,5R,8R,10R,11R,12S,13S,14R)-11-[(2S,3R,6R)-3-hydroxy-4,6-dimethyloxan-2-yl]oxy-3,5,6,8,10,12,14-heptamethyl-1-oxa-6-azacyclopentadecan-15-one and the other was 11885941 whose IUPAC Name is (3R,4S,5S,6R,7R,9R,11R,13S,14R)-6-[(4S,6R)-4-[but-3-ynyl](methyl)amino]-6-methylxan-2-yl]oxy-14-ethyl-4,12,13-trihydroxy-7-methoxy-3,5,7,9,11,13-hexamethyl-oxacyclotetradecane-2,10-dione. Against the 6NUS, another SARS-CoV-2 Coronavirus virus protein, the two most negative analogues of the docking score parameter were 9851445 and 12137352 whose IUPAC Name is (2R,3S,4R,5R,8R,10R,11R,12S,13S,14R)-2-ethyl-3,4,10-

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