Magnetic bead-based salivary peptidome profiling for periodontal-orthodontic treatment

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Abstract

Background: Periodontal-orthodontic treatment is usually needed to correct malocclusions, and may also improve periodontal health [1]. Patients who have chronic periodontal problems want to improve not only their malocclusions to obtain a perfect facial and tooth profile with orthodontic treatment, but also their periodontal condition. However, some doctors believe that orthodontic treatment aggravates periodontal damage [2]; the matter remains disputed. The difference in osteogenesis between periodontitis and non-periodontitis patients [3] may lead to differences in the effect of orthodontic treatment [4]. The aim of this study was to determine the differences between periodontal and non-periodontal orthodontic patients. Bacterial plaque is an etiologic factor in periodontitis [5]. The pathogenesis of this disease in adults results in a loss of connective tissue, bone support, and, ultimately, teeth [6]. The diagnosis of periodontitis and identification of at-risk patients are challenging. Periodontal condition is usually determined visually by clinical periodontal and X-ray examinations, which are subject to considerable measurement error due to clinical experience, and are often poorly tolerated by patients.

Saliva is a complex hypotonic hydrated solution [7] that contains 3397 different proteins with a variety of biological functions [8]. Whole saliva comprises the secretions of the major and minor saliva glands, non-salivary gingival crevicular fluid, bronchial secretions, bacteria and bacterial products, deciduous epithelial...

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cells, and food debris. Moreover, saliva contains molecules which could be diffused or filtrated from blood [9]. For example, human C-reactive protein (CRP), which is used to assess the coronary events, could be present in saliva [10]. Periodontitis can involve the modified production or release of various factors, such as hormones and cytokines. We sought to discover a candidate biomarker that can facilitate the detection of periodontal changes before pathological symptoms occur.

Since saliva contains multiple potentially informative components and its collection is non-invasive, low-cost, and simple, research is increasingly focusing on the analysis of oral and systemic conditions using saliva as a detection strategy [11,12]. Moreover, there are databases (http://bioinformatics.ua.pt/OralCard/ and http://www.hspp.ucla.edu/) that compile proteomics data from oral cavity, contributing to the salivary analysis. And protein profiling methods such as two-dimensional polyacrylamide gel electrophoresis for separation and MS for identification have been used to investigate various conditions and disorders, including breast cancer [13], Sjögren’s syndrome [14], rheumatoid arthritis [15], and oral pathologies such as oral cancer [16,17], dental caries [18], cleft palate [19], and periodontitis [20].

We used matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS), a sensitive MS-based proteomic technique, to detect peptides over a large mass range. The mass spectra generated by this technique are easy to interpret [21]. Here, MALDI-TOF MS was used in combination with WCX to select peptides in the range of 1000–10000 Da prior to further identification. The effectiveness of this combination of techniques has been confirmed in many serum-based peptide profile identification studies [22,23]. MALDI-TOF MS generated accurate salivary protein profiles of patients with fixed orthodontic appliances with and without periodontitis.

In this study, the peptide mass profiles of saliva samples were investigated using magnetic bead-based peptidome analysis. In this way, we aimed to identify a panel of differentially-expressed specific candidate biomarkers.

**Results**

To investigate the differences between periodontal-orthodontic and non-periodontal orthodontic patients, the entire mass spectra of the extracted peptide samples from 24 subjects (eight per group) were obtained by MALDI-TOF MS (Figure 1). Saliva peptidome fingerprint peaks were characterized in each patient by showing the maximum intensity within a particular m/z range. The molecular weight of the majority of the peptides was 1000–7000 Da. The mass spectra peaks were then quantified and compared.

An average of 109 protein mass peaks was found. The peak intensities of nine (1062.1, 1454.2, 2213.2, 2621.9, 3016.1, 3154.4, 3163.4, 5378.5, and 5435.2 Da) differed

**Table 1 Significant (p <0.05) m/z values discriminating samples from the two groups**

| Mean m/z value | p-value | Tendency* |
|----------------|---------|-----------|
| 3163.4         | 0.008   | ↓         |
| 1454.2         | 0.009   | ↑         |
| 3154.4         | 0.012   | ↓         |
| 2621.9         | 0.013   | ↓         |
| 5378.5         | 0.023   | ↓         |
| 1062.1         | 0.035   | ↑         |
| 3016.1         | 0.042   | ↓         |
| 5435.2         | 0.042   | ↓         |
| 2213.2         | 0.045   | ↓         |

*Tendency, m/z value intensity trend between the two groups: ↑, higher intensity in group B than A; ↓, lower intensity in group B than A. m/z, mass-to-charge ratio. A, the orthodontic patients without periodontitis; B, the orthodontic patients with periodontitis.
significantly among the three different groups (Table 1). Orthodontic patients in the group without periodontal disease showed higher mass peaks for peptides of 2213.2, 2621.9, 3016.1, 3154.4, 3163.4, 5378.5, and 5435.2 Da, whereas the mass peaks for the peptides of 1062.1 and 1454.2 Da were higher in the periodontal-orthodontic patients (Figures 2 and 3).

The most significant differences were exhibited by the 3163.4- and 1454.2-Da proteins ($p < 0.01$); the fit of the other combinations was not as good. Thus, we chose these two peptides to establish a fitted curve between the two groups (Figure 4). Samples from the two groups were well-separated, indicating that the fitting results were satisfactory.

Moreover, using LTQ-Orbitrap-MS detection, eight (1062.1, 1454.2, 2213.2, 2621.9, 3016.1, 3154.4, 3163.4, 5378.5, and 5435.2 Da) of the nine differentially-expressed peptides were successfully identified (Table 2).

Figure 2 Column view of the mass spectra of the three groups. The peak intensities of the three different groups showed differential expression of nine salivary peptides. Noticeably, significant differences were investigated between the two orthodontic groups. (*$p < 0.05$; **$p < 0.01$).
Discussion

Adult patients with periodontal disease exhibit tooth malocclusions such as flaring of the anterior segment. This unaesthetic appearance and/or dysfunction are the primary reasons for undergoing periodontal-orthodontic treatment using a multidisciplinary approach [24]. Combined orthodontic and periodontal treatment does not compromise the therapeutic effectiveness of the latter [25]. Indeed, this combination may be beneficial since it leads to increased bone resorption stability, a lack of incisor flaring, and improvements in bone defects [26]. However, some authors argue that orthodontic treatment is unsuitable in chronic periodontal patients because bacterial plaque is an etiologic factor in the development of periodontitis and the presence of orthodontic appliances facilitates plaque growth and maturation. Thus, the periodontal condition could worsen after orthodontic treatment [27]. Therefore, we examined periodontal-orthodontic patients with a view to elucidating the differences between them and non-periodontal orthodontic patients.

Such differences could be caused by inflammatory factors related to periodontitis or other protein factors [28], which may represent candidate biomarkers. A candidate biomarker is defined as an informative signal associated with a specific condition. The specificity and sensitivity of a biomarker describe its usefulness in diagnosing a specific condition or predicting its progress [29]. An effective biomarker should be measurable in an accessible body fluid, such as serum, urine, or saliva [30]. Saliva contains abundant proteins, peptides, and other small molecules [8]. Thus, the salivary peptide spectrum may be applied widely in the diagnosis and monitoring of oral diseases. Progress has been made in screening for not only oral diseases such as oral cancer [17], but also systemic conditions such as gastric cancer [31] and breast cancer [13]. Studies of periodontitis using proteomics [32,33] have been performed, and the demand for periodontal-orthodontic combination treatment is increasing. However, little research on the salivary peptide spectrum in the periodontal-orthodontic area has been conducted.

In the present study, we used MALDI-TOF MS-based proteomic methods and WCX magnetic beads to examine all 24 saliva samples. Nine peaks that differed significantly were found (Figure 2), of which two (3163.4 and 1454.2 Da) exhibited the most significant differences ($p < 0.01$). Orthodontic treatment is a prolonged procedure. And in our previous study, alterations were found in salivary proteins due to different orthodontic treatment durations [34]. In this study, the peak intensities of the three different groups showed differential expression of salivary peptides, indicating the orthodontic treatment could contribute to the change of salivary peptidome. Noticeably, significant differences were investigated between the two orthodontic groups. This suggested that differential expression of salivary peptidome profile did exist between orthodontic patients with and without periodontitis. Thus, this method provides a new tool for analyzing the effect of periodontitis on orthodontic treatment.

The peptide sequence identifications made in this study have led to interesting speculations. The 3163.4-Da peptide was identified as F2 prothrombin precursor. Thrombin is a ‘trypsin-like’ serine protease protein encoded by the $F2$ gene. Beyond its key role in the dynamic process of thrombus formation, thrombin has the potential to exert actions such as inflammation and leukocyte recruitment [35]. Moreover, isoform 1 of fibrinogen alpha chain precursor (FGA), which was the predicted identity of the 3154.4- and 2621.9-Da peptides, is encoded by $FGA$. It is cleaved by thrombin to form fibrin, indicating that it interacts with thrombin. Some studies [36] suggested epithelial thrombomodulin (TM), which binds to thrombin and converts it from a
procoagulant protease to an anticoagulant enzyme, increased in gingival crevicular fluid of individuals with chronic periodontitis. Moreover, gingipains, the major periodontopathic bacterium Porphyromonas gingivalis-derived cysteine proteases, lead to the degradation of endothelial TM. In addition, reduced expression of TM was found in gingival microvascular endothelia in patients with periodontitis [37], and this may be involved in the pathogenesis of periodontitis. Thus, these differential expression patterns of altered proteins may have originated from periodontitis-associated inflammation or differences in bone metabolism between orthodontic patients with and without periodontitis. Ultimately, our aim is to determine the protein or gene from which a peptide is derived; however, this is complex. When the distribution of the peptides with low molecular weight intended to match to the mass spectrometry spectra relative peak area, it should be noted that these peptides had complicated origins. Peptides in saliva could be secreted peptides and proteolytic fragments of related proteins [38]. Moreover, these components are subject to secondary modifications from distinctive protein families. Thus, a peptide sequence usually does not exclusively define a single protein.

As these specific peptides were investigated using this relatively new combined method with no validation using other techniques, thus, a larger sample size and repeated trial or trial using other techniques are needed to confirm the significant differences in peptide mass peaks found in this study and its reproducibility. The establishment of a relatively complete protein-peptide spectrum database will facilitate the determination of both the source of the salivary protein profile variation and the mechanism thereof. Moreover, the analysis of saliva is inherently challenging because of the wide protein concentration range therein [29] and the presence of multiple post-translational modifications [39]. Further research into the composition of saliva will likely provide novel tools for investigations of physiological and pathophysiological states.

**Conclusions**

In conclusion, our data suggest that the salivary protein profiles of periodontal, non-periodontal orthodontic patients and periodontitis patients exhibit differential mass spectra peak intensities (i.e., the peptide profile is altered in cases of periodontitis). This represents a new method of analyzing the effect of periodontitis on orthodontics and orthodontic contribution to the alterations of salivary peptidome profile, ultimately leading to increased treatment efficacy. However, expansion of the orthodontic patient dataset and the identification of additional candidate biomarkers are necessary to establish an effective diagnosis and monitoring model for periodontal-orthodontic treatment.

**Methods**

**Ethics statement**

This study was approved by the Peking University Biomedical Ethics Committee. Adult subjects and parents of pediatric subjects signed an informed consent form before participating in the study.

**Subjects**

Patients seeking treatment at the Stomatology School of Peking University were recruited in November, 2011. All 24 study subjects were generally ensured systemically healthy via testing of serum basic biochemical items and inquiring medical history, and those who presented with caries, diseases of the oral mucosa, or oral cancers were excluded. Recorded clinical periodontal parameters of the subjects included bleeding on probing (BOP), probing

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**Table 2 Sequences of the differentially-expressed peptides**

| Mean m/z value | Peptide name | Peptide sequence |
|----------------|--------------|------------------|
| 3163.4         | F2 Prothrombin precursor | LRLPFEKKSLEDKTERELLESYIDGR |
| 1454.2         | SERPINA1(PRO2275) | PLFMGKWNPTQK |
| 3154.4         | FGA Isoform 1 of fibrinogen alpha chain precursor | LIGQIVSITASLR |
| 2621.9         | FGA Isoform 1 of fibrinogen alpha chain precursor | SYKM*ADEAGSEADHEGTKRKGHAKGRP |
| 5378.5         | VWC Domain containing protein precursor | APVNCSSCPGPTASPSRPVHLILQQLRRTNLMKTQTLPSPAGAHGPHSLA |
| 1062.1         | ITIH4 Isoform 2 of inter-alpha-trypsin inhibitor heavy chain H4 precursor | SEMWAGKLO |
| 3016.1         | 280-kDa protein | LGVSPPPGAVLHSLPLEFPLAM*FAEQ |
| 5435.2         | Unknown peptide-identification failure | |
| 2213.2         | ACTB Actin, cytoplasmic | DLYANTVLSGGTTMYPGIADR |

m/z, mass-to-charge ratio.

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pocket depth (PPD) and clinical attachment level (CAL). Chronic periodontitis was diagnosed in patients who had six sites BOP on different teeth with PPD≥6 mm and CAL≥5 mm. Control orthodontic patients had <10% of sites BOP and PPD<3 mm. And all the periodontitis patients manifested to be moderate degree of chronic periodontitis. Then the patients were divided into three groups according; that is, periodontal-orthodontic, non-periodontal orthodontic groups and patients with periodontitis but no orthodontic treatment. The characteristics of the subjects are presented in Table 3.

The treatment procedure for all subjects involved the use of a fixed appliance, followed by alignment, leveling, and refinement procedures. All patients were asked to maintain good oral hygiene during treatment and were counseled regarding oral hygiene and tooth brushing both before treatment and at each visit.

Saliva collection and processing
All individuals were asked to rest for 15 min before saliva collection at 8:30 am, and not to eat or drink after dinner the previous evening or to brush their teeth on the collection day morning. The subjects sat upright in a quiet room and were required to put the tip of their tongue against the sublingual caruncle without straining. Thus, the saliva, which was received in a paper cup for the first 5 min, could run from the mouth, and we collected 6 mL of the spontaneous saliva flow in a 50-mL centrifuge tube. During the collection procedure, patients were asked not to speak. Immediately after collection, the unstimulated whole saliva samples were kept on ice and then centrifuged at 9000 × g for 7 min at 4°C to remove insoluble materials, cells, and debris. Ethylene diamine tetracetic acid (1 mM; Sigma, St. Louis, MO, USA) and 1 mM phenylmethyl sulfonylfluoride (Sigma) were added to the sample supernatants to inhibit protease activity. Protein concentrations were determined using the Lowry assay and ELx808 Protein Assay (BioTek, Hercules, CA, USA). The supernatants were kept at −80°C for further analysis.

Reagents and instruments
The WCX magnetic bead kit (SPE-C; Bioyong Tech, Beijing, China), alpha-cyano-4-hydroxycinnamic acid (HCCA), MALDI-TOF MS (Bruker Bio-sciences, Bremen, Germany), 100% ethanol (chromatographic grade), and 100% acetone (chromatographic grade) were freshly prepared.

WCX fractionation and MALDI-TOF MS
The low-molecular-weight (LMW) salivary peptidome contains enormous and important biological information and they should be a rich source of specific candidate biomarkers [13]. Unlike 2D-PAGE analysis, which only allows the separation of proteins with molecular weight in a range of 20–300 kDa, the magnetic bead-based profiling MS technologies could cover a range of 1–250 kDa [29]. Moreover, the high-throughput nature of MALDI-TOF MS makes fast screening for novel candidate biomarkers possible, and this method is cost effective and can be easily adopted [31]. Besides, this combined method only required 20ul processed sample for generating enough proteins peaks profiling by MALDI-TOF-MS.

The WCX magnetic bead kit suspension was mixed by shaking. After eluting and beating, the magnetic beads were separated from the protein and the eluted peptide samples were transferred to a clean 0.5-mL sample tube for further analysis by MS.

Five microliters of HCCA substrate solution (0.4 g/L, dissolved in acetone and ethanol) and 0.8-1.2 μL of eluate were mixed. Next, 0.8-1.2 μL of this mixture was applied to a metal target plate and dried at room temperature. Finally, the prepared samples were analyzed by MALDI-TOF MS. Peptides with a molecular weight in the range of 1000–10000 Da were collected, and 400 shots of laser energy were used. Peptide mass fingerprints were obtained by accumulating 50 single MS signal scans.

Identification of differentially expressed candidate biomarkers
The sequences of peptides expressed differentially between the two groups were determined by nano-liquid chromatography-electrospray ionization-tandem mass spectrometry (nano-LC/ESI-MS/MS) using a setup consisting of an Aquity UPLC system (Waters) and an LTQ Orbitrap XL mass spectrometer (Thermo Fisher) equipped with a nano-ESI source. The obtained chromatograms were analyzed with BioworksBrowser 3.3.1 SP1, and the resulting mass lists were used in a database search with Sequest™ [IPI Human (3.45)]. The parameters used to generate the peak list were: parent ion and fragment mass relative accuracy (50 μg/g and 1 Da, respectively).

Statistical analysis
A two-tailed t-test and Student’s t-test were used to compare peptide peak intensities between the two groups.
Data were analyzed using the BioExplorer statistical package (BioyongTech). A p-value < 0.05 was considered significant.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
LX and CF conceived of the idea for the peptidomic study and participated in its design. ZHN and ZSN carried out a major portion of the data analysis and drafted the manuscript. LRX, ZH and WXD carried out the sample collection and extraction. ZYH and DN carried out the data statistical analysis and a portion of the data analysis. CT provided the orthodontic patients. All authors read and approved the final manuscript.

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