Targeting ligand-dependent wnt pathway dysregulation in gastrointestinal cancers through porcupine inhibition

Dustin J. Flanagan a,b, Simon A. Woodcock c, Caroline Phillips c, Catherine Eagle c, Owen J. Sansom a,d,*

a Cancer Research UK Beatson Institute, Glasgow, UK
b Biomedicine Discovery Institute, Monash University, Melbourne, Australia
c Redx Oncology Ltd, Redx Pharma PLC, UK
d Institute of Cancer Sciences, University of Glasgow, Glasgow, UK

Abstract

Gastrointestinal cancers are responsible for more cancer deaths than any other system of the body. This review summarises how Wnt pathway dysregulation contributes to the development of the most common gastrointestinal cancers, with a particular focus on the nature and frequency of upstream pathway aberrations. Tumors with upstream aberrations maintain a dependency on the presence of functional Wnt ligand, and are predicted to be tractable to inhibitors of Porcupine, an enzyme that plays a key role in Wnt secretion. We summarise available pre-clinical efficacy data from Porcupine inhibitors in vitro and in vivo, as well as potential toxicities and the data from early phase clinical trials. We appraise the rationale for biomarker-defined targeted approaches, as well as outlining future opportunities for combination with other therapeutics.

© 2022 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

Keywords:
Porcupine inhibitor
Wnt
gastrointestinal cancer
RNF43
RSPO
clinical trials

Contents
1. Introduction .................................................................................................................. 2
2. Dysregulation of Wnt Pathways in Gastrointestinal Cancers .................................... 3
3. Wnt pathway dysregulation in colorectal cancer (CRC) ........................................ 3
4. Wnt pathway dysregulation in pancreatic cancer ..................................................... 4
5. Wnt pathway dysregulation in gastric cancer ............................................................ 4
6. Wnt pathway dysregulation in biliary cancer (Cholangiocarcinoma) ......................... 5
7. Wnt pathway dysregulation in hepatocellular carcinoma (HCC) ............................ 5
8. Pre-clinical evidence supporting use of porcupine inhibitors in gastrointestinal cancers .......................................................... 5
9. Effects of porcupine inhibitors on tumor growth and proliferation ......................... 5
10. Effect of porcupine inhibitors on cancer stem cells. .............................................. 7
11. Effect of porcupine inhibitors on invasion and metastasis ................................... 7
12. Effects of porcupine inhibitors on fibrosis. .............................................................. 7
13. Reversal of immune evasion by porcupine inhibition ............................................. 8
14. Potential toxicities of porcupine inhibitors identified in vivo ................................ 8
15. Porcupine inhibitors in the clinic ............................................................................ 8
16. Conclusions and future perspectives ..................................................................... 9

Abbreviations: APC, adenomatous polyposis coli; CAF, cancer associated fibroblast; CMS, consensus molecular subtype; CRC, colorectal cancer; CSC, cancer stem cell; EMT, epithelial-mesenchymal transition; Dkk, Dickkopf; HCC, hepatocellular carcinoma; LGR, leucine-repeat G-coupled receptor; LRP, low-density lipoprotein receptor-related protein; MSI, microsatellite instability; PCP, planar cell polarity; ROR, receptor tyrosine kinase-like orphan receptors; RSPO, R-spondin; SFRP, Secreted frizzled-related protein; TP53, transformation-related protein 53.

* Corresponding author at: Cancer Research UK Beatson Institute, Glasgow G61 1BD, UK.
E-mail address: o.sansom@beatson.gla.ac.uk (O.J. Sansom).
1. Introduction

Secreted Wnt proteins were originally discovered as key players in development and oncogenesis (Nusse & Varmus, 1982; Nüsslein-Volhard & Wieschaus, 1980; Sharma & Chopra, 1976) and subsequently found to be important in tissue homeostasis, with pivotal roles in stem cell-mediated regeneration of the intestinal epithelium and maintenance of bone integrity (Clevers & Nusse, 2012). Dysregulation of Wnt signaling has been implicated in various cancers, particularly those of the gastrointestinal tract (Jackstadt, Hodder, & Sansom, 2020; Sphyris, Hodder, & Sansom, 2021).

Extracellular Wnts can be divided into canonical or non-canonical according to the cellular pathways they activate (Fig. 1). Canonical Wnts bind Frizzled receptors which, in complex with low-density lipoprotein receptor-related protein (LRP) 5 or 6 and Dishevelled, release β-catenin from a destruction complex of Axin, adenomatous polyposis coli (APC), casein kinase 1α (CK1α) and glycogen synthase kinase 3β (GSK3β). This permits accumulation of β-catenin and its entry into the nucleus to initiate activation of target genes in combination with transcription factor/lymphoid enhancer binding factors (TCF/LEF) (Clevers & Nusse, 2012). The pathway is subject to several checks and balances, particularly in the extracellular space (Niehrs, 2012). The transmembrane E3 ligase RNF43 and its close relative, ZNRF3, negatively regulate canonical Wnt signaling through ubiquitylation of Frizzled receptors, which targets them to lysosomes for destruction (Koo et al., 2012). Soluble Dickkopf (Dkk) proteins compete with Wnt binding to LRPs/6 (Kagey & He, 2017) to block canonical signaling. Wnt activation can be fine-tuned by extracellular R-spondins (RSPOs), which bind to leucine-repeats G-coupled receptors (LGR/5/6) on the cell surface, bringing RSPO into contact with RNF43/ZNRF3. This interaction leads to membrane clearance of RNF43/ZNRF3 with a concomitant increase in membrane Frizzled receptors, thus enhancing Wnt signalling.
2. Dysregulation of Wnt Pathways in Gastrointestinal Cancers

Elevated β-catenin, indicative of canonical Wnt pathway activation, has been observed in many gastrointestinal tumors. Such activation may be driven by mutations in upstream pathway components such as RNF43 and RSPO fusions and tumors will remain dependent on Wnt ligand, or mutations in downstream Wnt pathway components such as APC or CTNNB1, the gene encoding β-catenin, where the pathway may be activated independent of Wnt ligand. Deregulation of extracellular Wnt regulators such as Dkk and SFRPs via epigenetic mechanisms may also play a role (for reviews see e.g. (Kleeman & Leedham, 2020; White, Chien, & Dawson, 2012)). Activation of the non-canonical pathway in cancers has also been associated with poor prognosis (Chen, Chen, Tang, & Xiao, 2021). Although proliferation and stem cell regeneration have been identified as potential cancer-promoting mechanisms, Wnt pathways also promote tumor evasion of host immunity (Luke, Bao, Sweis, Spranger, & Gajewski, 2019; Oderup, Lajevic, & Butcher, 2012; Valencia et al., 2011; Wang, Kalland, Ke, & Qu, 2018), as well as increasing migration and invasiveness via epithelial-mesenchymal transition (EMT) (Hlbek et al., 2007). These findings have led to a search for therapeutics targeting Wnt pathways. One of several emerging drug candidates is the membrane-bound protein-serine O-palmitoleoyltransferase Porcupine, encoded by the PORCN gene. Porcupine is located in the endoplasmic reticulum of stromal or epithelial cells and palmitoylates Wnt ligands, an enzymatic process essential for their secretion and binding to Frizzled receptors (Liu et al., 2013). Since Porcupine targets both canonical and non-canonical Wnts (Madan et al., 2016), inhibitors have the potential to hit multiple pro-tumorigenic mechanisms. However, they will only block signaling that remains dependent on secreted Wnts. In the next section, we summarise evidence for the involvement of Wnt pathway aberrations in five leading gastrointestinal cancers, with a focus on ligand-dependent mechanisms.

3. Wnt pathway dysregulation in colorectal cancer (CRC)

CRC is a Wnt-pathway driven disease, with 63-93% of patients having mutations in at least one component of the canonical Wnt pathway (Fig. 2) (Cancer Genome Atlas Network, 2012; Kleeman et al., 2020; Yaeger et al., 2018). The majority of mutations occur in downstream elements of the pathway. Most common are loss of function mutations in APC, which account for 60-80% of Wnt pathway aberrations in colorectal cancers (Kinzler & Vogelstein, 1996; Kleeman et al., 2020; Yaeger et al., 2018), and to a lesser extent, aberrations in other downstream components AXIN1, AXIN2, and CTNNB1. In APC-mutated cancer models, restoration of functional APC protein is effective in reversing tumorigenesis (Dow et al., 2015; Faux et al., 2004; Zilberberg, Lahav, & Rosin-Arbesfeld, 2010), but with current technologies, APC restoration is not a practical therapeutic option. In theory, cancers driven by such downstream activation mechanisms are unlikely to respond to Porcupine inhibition, as they do not depend on Wnt ligand, and are therefore termed ligand-independent. However, there are reports of APC-deficient and CTNNB1-mutant colorectal cell lines that remain dependent on Wnt ligands, and whose proliferation was reduced by Porcupine inhibition (J. Li, Zhang, Wang, & Zhang, 2019; Voloshanenko et al., 2013). Whether the dependency of APC- or CTNNB1-mutant CRC cells on Wnt extents beyond in vitro models requires further investigation.

10-20% of CRC patients have pathway aberrations in which a requirement for Wnt ligand is maintained. The majority of these are in RNF43 or ZNRF3, with RNF43 mutations found in 9-18% of CRC patients, most of which associate with microsatellite instability and are mutually exclusive from genetic aberrations in APC but strongly associated with BRAF mutations (Giannakis et al., 2014; Yaeger et al., 2018). Many RNF43 mutations observed in CRC are truncations and hence loss of function, and therefore predicted to activate both canonical and non-canonical Wnt signaling. Certain truncation mutants have been reported to switch the protein from a tumor suppressor into an activator of downstream β-catenin signaling (Spit et al., 2020). Growth of tumorigenic intestinal organoids that lack functional RNF43 or ZNRF3 remain dependent on local Wnt ligand production (Koo, Van Es, Van Den Born, & Clevers, 2015), and their growth can be blocked by Porcupine inhibitors (Van De Wetering et al., 2015). However, there is debate concerning the most common RNF43 mutation, G659Vfs*41, a frameshift mutation which is almost exclusively found in microsatellite instability (MSI) patients. Initial reports suggested it was associated with cancer cell growth and/or survival, but it has also been reported to generate fully functional protein, although potentially one that is less stable than wild-type and associated with lower RNF43 expression (Giannakis et al., 2014; Liet al., 2020; Tu et al., 2019; Yu et al., 2020). Oncogenic effects of this mutation have been reported to be independent of Wnt signaling (L. Fang et al., 2021). It is therefore unclear whether tumors with this particular frameshift retain Wnt ligand dependency in patients or in fact have enhanced Wnt pathway signalling as a result of the mutation.

RSPO2 and RSPO3 fusions are mutually exclusive and are present in 2-10% of CRC, co-occurring with KRAS or BRAF mutations, but only very rarely with APC and mutually exclusive of CTNNB1 and RNF43 mutations (Giannakis et al., 2014; Seshagiri et al., 2012; Shimamura et al., 2014). The gene encoding RSPO3 fuses with the signal sequence and early section of the PTPRK gene leading to increased expression of RSPO3 and activation of the canonical Wnt pathway. Genetic fusions of RSPO2 have been observed with EHSF and PIZE01, and these were associated with increased RSPO2 expression (Hashimoto et al., 2019). Increased stromal RSPO3 expression has also been observed in the absence of gene fusions and this stromal overexpression of RSPO3 is strongly associated with the aggressive mesenchymal consensus molecular subtype 4 (CMS4) CRC phenotype (Kleeman et al., 2020). Targeting RSPO3 overexpression to the murine gut epithelium upregulates Wnt signaling and drives the formation of ectopic niche compartments and intestinal stem cells, leading to adenoma lesions (Hikens et al., 2017). Inhibition of RSPO3 with function-blocking antibodies in PTPRK-RSPO3-fusion tumor xenografts inhibited growth and promoted epithelial differentiation (Storm et al., 2016).
Excessive Wnt signaling can also arise by non-mutational mechanisms. For example, epigenetic silencing of negative Wnt regulators has been observed during CRC cancer progression and was associated with increased nuclear β-catenin (Caldwell et al., 2004; Silva et al., 2014; Suzuki et al., 2004; Zhang et al., 2008). Epigenetic suppression of Wnt feedback loops, including reduced Axin2 expression, has been observed in CRC and low Axin2 expression has been postulated to be a discriminatory biomarker for ligand-dependent disease (Kleeman et al., 2020). However, the expression of some negative Wnt regulators (Notum, Wif1, Dkk3) is very low/absent in normal murine gut tissue (Flanagan et al., 2021) which may suggest epigenetic silencing takes place prior to tumour formation, possibly confounding the premise of further epigenetic silencing in cancer tissue.

Non-canonical pathways are also dysregulated in colorectal cancer, with high expression of Wnt11a or ROR1 associated with poor prognosis (Nishioka et al., 2013; Zhou et al., 2017). Non-canonical Wnt signaling has been linked to proliferation, invasion, survival and metastasis of cancer stems cells (Katoh, 2017; Nishioka et al., 2013), although it should be noted that the role of non-canonical Wnts is context dependent. In certain situations, non-canonical Wnt5a has been shown to be tumor suppressive and promote differentiation, most likely via cross-inhibition of canonical Wnt signaling (Zhou, Kipps, & Zhang, 2017).

### 4. Wnt pathway dysregulation in pancreatic cancer

RNF43 mutations are the most common Wnt pathway mutations in pancreatic cancer, observed in 5–10% of cases (Fig. 2) (Bailey et al., 2016; Tu et al., 2019; Witkiewicz et al., 2015). Genetic lesions in APC or AXIN1/2 are less common, observed in 1–5% of cases. As expected, RNF43-mutant pancreatic cancer xenografts remained dependent on Wnt ligand. However, a significant proportion of organoids derived from PDAC patients with wild-type RNF43 also remained dependent on Wnt ligands, suggesting opportunities for Porcupine inhibitors and other ligand-dependent therapeutics exist beyond RNF43 mutations in pancreatic cancers (Seino et al., 2018).

At the expression level, upregulation of β-catenin, Wnts and Frizzled receptors was seen in a gene-array analysis of pancreatic ductal adenocarcinomas (Pasca Di Magliano et al., 2007) and elevated nuclear β-catenin is associated with high grade neoplasms and poor prognosis (Al-Aynati, Radulovich, Riddell, & Tsao, 2004; Sano et al., 2016). High epithelial expression of Wnts in pancreatic tumors was also associated with poor prognosis (Seino et al., 2018). Ectopic expression of Wnt1 or β-catenin accelerated tumor growth and decreased survival in a murine pancreatic cancer model (Sano et al., 2016).

Wnt5a is elevated in pancreatic cancer tissue versus surrounding non-cancerous tissues and promotes invasion and proliferation whilst inhibiting apoptosis. Although typically considered a non-canonical Wnt, these effects appear to be via β-catenin signaling (Bo, Gao, Chen, Zhang, & Zhu, 2016; Griesmann et al., 2013; Ripka et al., 2007). The non-canonical pathway receptor ROR1 is overexpressed in circulating tumor cells from pancreatic cancer patients and knockdown of ROR1 reduces invasiveness (Xu, Shen, Xu, Wang, & Ni, 2018). Consistent with non-canonical signaling playing a role in pancreatic cancer, ROR2 expression in tumor epithelium and stroma is associated with poor prognosis in this indication (J. Huang et al., 2015).

### 5. Wnt pathway dysregulation in gastric cancer

Gastric cancers have one of the highest incidence of Wnt pathway mutations among cancers of the digestive tract (Flanagan, Vincan, & Phesse, 2017). CTNNB1 mutations occur in up to 26% of gastric cancers and are associated with nuclear localization of β-catenin (Clements et al., 2002). Several CTNNB1 single-nucleotide polymorphisms are associated with gastric cancer susceptibility and prognosis (Wang et al., 2018).
2012). APC and AXIN1/2 mutations also occur in 5–10% of patients, although prevalence of APC mutations has been reported to be as high as 25% in some studies (Z. Fang et al., 2012; Horii et al., 1992; Pan, Liu, Zhang, You, & Lu, 2008). A significant proportion of gastric cancers contain ligand-dependent aberrations. RNF43 mutations occur in 10% of patients, rising to 54% of MSI tumors, a high proportion of which are the aforementioned G659Vfs*41 frameshift truncations (Wang et al., 2014). RSP2 fusions have also been reported in gastric tumor xenografts (Li et al., 2018), although prevalence data is not available. Interestingly, patient-derived gastric tumor organoids mimic the variety of Wnt-pathway mutations observed in the clinic and reveal novel mutational and methylation patterns (CPg island methylation phenotype (CIMP+)); DNA methylation) to achieve Wnt and/or RSP0 independence. Importantly, organoids independent of RSP0 (typically RNF43 loss of function mutants) remain sensitive to Wnt inhibition (Nanki et al., 2018).

Beyond mutations, gene expression analysis found 46% of gastric cancers had upregulation of canonical and non-canonical Wnts, or their downstream pathway components (Ooi, Ivanova, Wu, Lee, & Tan, 2009). Conversely, multiple SRFPs and DKKs are downregulated by promotor hypermethylation (Chiurillo, 2015). Of note, Fizzled7 and Wnt5a are highly expressed in gastric cancer and both are associated with poor outcomes (Li et al., 2018; Maeda et al., 2020). Fizzled7 was identified as the predominant receptor responsible for transmitting Wnt signaling in human gastric cancer cells and genetic deletion of Fizzled7 prevented growth of gastric adenomas with or without APC mutations (Flanagan et al., 2019). Inhibiting Wnt5a blocked fibroblast-induced gastric cancer cell proliferation and migration in vitro, and metastasis of gastric cancers in vivo (Hanaki et al., 2012; Kurayoshi et al., 2006; Maeda et al., 2020). Together, these observations suggest that, even in the presence of downstream Wnt mutations, upstream inhibitors including those targeting Porcupine could have utility in gastric cancer.

6. Wnt pathway dysregulation in biliary cancer (Cholangiocarcinoma)

Only a small proportion of bile duct cancers carry genetic mutations in Wnt pathway components. RNF43 mutations were observed in 1–2% of patients (Weinberg et al., 2019), although higher rates (4–9%) have been seen in Asian cohorts, particular those associated with liver fluke infection (Chan-On et al., 2013; Ong et al., 2012). However, high expression of stromal Wnt7b and Wnt10a, and upregulation of several Fizzled receptors has been observed (Boulter et al., 2015), with greater than 75% of patient samples in another study showing positive expression of Wnt3a, Wnt5a and Wnt7b (Loilome et al., 2014). In addition Boulter and colleagues demonstrated efficacy in a biliary tract preclinical model with C-59, a Porcupine inhibitor, demonstrating that Wnt ligand is required for proliferation of these tumors (Boulter et al., 2015). Hypermethylation induced silencing of negative pathway regulators such as SFRP2 and DKK2 has also been observed (Goepert et al., 2014). These data are suggestive of highly activated extracellular Wnt signaling in biliary cancers, which may therefore be suitable targets for inhibitors of Porcupine.

7. Wnt pathway dysregulation in hepatocellular carcinoma (HCC)

CTNNB1 and AXIN1/2 mutations occur in a significant proportion of HCC patients (Miyoshi et al., 1998; Schulze et al., 2015; Taniguchi et al., 2002), and mice with liver-targeted disruption of APC or overexpression of an oncogenic form of β-catenin develop hepatic tumors indicating a role for downstream canonical Wnt pathway activation (Colnot et al., 2004; de La Coste et al., 1998). Another study showed that partial ablation of APC also induced liver tumorigenesis in mice (Buchert, Athineos, Abd, Burke, & Faux, 2016). In contrast, ZNRF3 mutations were observed in only 3% of HCC (Schulze et al., 2015) and RSP0 fusions are rare, although elevated RSP02 was seen in a subset of HCC patients. Co-amplification of RSP02 and MYC has been observed in HCC, although the functional consequence of this is unclear (Sanchez-Vega et al., 2018). Elevated Wnt3/4/5a and suppression of SFRPs have also been observed, an expression pattern that did not correlate with CTNNB1 mutations but was associated with loss of differentiation and cirrhosis, and activation of PCP pathways (Bengochea et al., 2008). Functional overexpression of RSP02 with combined loss of transformation-related protein 53 (TP53) led to development of liver cancer in mice models (Conboy et al., 2019), implicating a role for extracellular Wnt ligands in liver tumorigenesis.

8. Pre-clinical evidence supporting use of porcupine inhibitors in gastrointestinal cancers

The pivotal role of Wnt signaling in gastrointestinal cancers has led to a search for therapeutics that target this pathway. A major challenge has been finding tractable targets, given the non-enzymatic nature of key intracellular components such as APC, AXIN and β-catenin. The majority of pathway inhibitors explored to date are antibody or biologicals-based approaches such as Fizzled decoy receptors and Fizzled blocking antibodies. Clinical studies of these therapeutics have been limited, hampered by toxicity issues (Table 1). A handful of small molecules have also been developed, such as CWP32291, which degrades β-catenin via an unknown mechanism (Pak et al., 2019), and PRI-724, which has been reported to block β-catenin/CBP interactions (Table 1). However both compounds require intravenous dosing, and reported effects in the clinic to date are minimal, with no direct evidence of target engagement (El-Khoueiry et al., 2013; K. Kimura et al., 2017; Ko et al., 2016; Lee et al., 2020). Tankyrase inhibitors, which can antagonize Wnt signaling via Axin stabilisation, have failed to advance into the clinic due to lack of a therapeutic window versus gastrointestinal toxicity, as well as non-Wnt related effects (S.-M. A. Huang et al., 2009; Y. Zhong et al., 2016). Antibodies directed against Dkk-1, such as DKN-01 and BHQ880 are also being tested. Although Dkk-1 functions as a naturally occurring negative regulator of Wnt signaling, Dkk-1 expression is often associated with poor prognosis in gastrointestinal cancers, and Dkk-1 blocking antibodies can inhibit tumor growth in pre-clinical animal models (Kagey & He, 2017). Such effects may arise from the complexity of Wnt signaling; for example, Dkk-1 has been linked to promotion of metastasis via activation of the non-canonical PCP-pathway, and also to immune suppression. It is speculated that the role of Dkk1 may be altered in different contexts, including when the canonical Wnt pathway is constitutively activated (Kagey & He, 2017).

More recently, several small molecule drug candidates with attractive pharmacokinetic properties have been developed that target Porcupine (Table 2; Bhamra et al., 2017; B. Chen et al., 2009; J. Jiang et al., 2018; Liu et al., 2013; Madan, Ke, et al., 2016; Profitt et al., 2013). Pre-clinical evidence suggests that Porcupine inhibitors exert anti-tumor effects via multiple mechanisms, and data from gastrointestinal models is summarised below.

9. Effects of porcupine inhibitors on tumor growth and proliferation

Porcupine inhibitors have proven effective in inhibiting proliferation of cell lines and organoids derived from gastric, colorectal, pancreatic and biliary cancers. They also show potent inhibition of tumor xenograft growth in mice (Boulter et al., 2015; Li, Cao, et al., 2018; Madan, Ke, et al., 2016; Mo et al., 2013). Effects are limited to models derived from Wnt-dependent tumors, in particular those with RSP02/3 fusions or RNF43 mutations, consistent with the role of Porcupine in Wnt secretion (X. Jiang et al., 2013; Koo et al., 2015; Woodcock et al., 2019). Inhibition of tumor proliferation was
accompanied by downregulation of Wnt targets such as Axin2 and c-Myc, and loss of cell cycle and stem cell markers (Madan et al., 2016). In addition, Porcupine inhibitors reduce glucose uptake by cancer cells in vitro and in vivo (Phillips et al., 2021 and Phillips et al, manuscript submitted for publication) and several studies demonstrate elevated caspase 3/7 expression and induction of apoptosis (Bagheri, Tabatabae Far, Mirzaei, & Ghasemi, 2020; Boulter et al., 2015; Mo et al., 2013), all of which may contribute to inhibition of tumor growth. The multi-modal effects of Porcupine inhibitors may explain why established RSPO2/3-fusion tumours are rapidly cleared following treatment with the porcupine inhibitor LGK974, even though treatment had no effect on the normal growth of intestinal crypts (Han et al., 2017). Effects of Porcupine inhibitors are long-lasting, with no tumor regrowth observed within 6 weeks of treatment withdrawal (Madan, Ke, et al., 2016) and significant reduction in growth was seen when Porcupine-inhibitor treated tumors were reimplanted into naïve, untreated mice (Phillips et al., 2021 and Phillips et al, manuscript submitted for publication).

Table 1
Wnt Pathway Inhibitors in Cancer Clinical Trials

| Drug Identifier | Mode of Action | Trial Identifier | Regimen | Cancer Indication | Trial Phase | Study Status or Clinical Results |
|-----------------|---------------|-----------------|---------|-------------------|------------|----------------------------------|
| OMP-54F28 (Ipafricept) | Fzd8-Fc Decoy receptor | NCT01608867 | Monotherapy | Solid tumors | 1 | Dysgeusia and fragility fractures observed. Best response was SD (Jimeno et al., 2017) |
| OMP18R5 (Vantictumab) | Anti-Fzd7 antibody, also neutralizes binding to Wnts to Fzd 1, 2, 5,8 | NCT01345201 | In combination with Paclitaxel and Carboplatin | Ovarian cancer; Hepatocellular carcinoma | 1 | Study completed, no results reported |
| OTSA101 | Yttrium90 radiolabelled Anti-Fzd10 antibody | NCT01957007 | Monotherapy | Solid tumors | 1 | Study completed, no results reported |
| OMP131R10 | Anti-R-spondin3 antibody | NCT01197004 | With or without FOLFIRI | Synovial sarcoma | 1 | Best response was SD in 3/7 patients treated with chemo combination (Bendell et al., 2016) |
| Foxy-5 | Wnt5a mimetic | NCT02655952 | Monotherapy | Breast cancer; colorectal cancer; prostate cancer | 1 | Completed. No efficacy data reported |
| PRI-724 | Inhibitor of TCF-CBP interaction | NCT01032405 | Monotherapy | Advanced solid cancers | 1 | No responses reported (El-Khoueiry et al., 2013) |
| CWP232291 | Induces degradation of beta catenin. Molecular target not defined | NCT01398462 | Monotherapy | Advanced/metastatic pancreatic adenocarcinoma | 1 | Rest response was SD in 8/20 patients (Ko et al., 2016) |
| SM04755 | Small molecule Wnt inhibitor | NCT02191761 | Monotherapy | Advanced CRC, gastric, HCC and pancreatic cancer | 1 | Completed. No results reported |
| Cirmtuzumab | Anti-ROR1 antibody | NCT05135905 | Monotherapy | Metastatic castration resistant prostate cancer | 2 | Not yet recruiting |

Source: ClinicalTrials.gov. Data collected on 10th January 2022. Withdrawn trial or trials terminated early are not included. Dkk blocking antibodies such as DkN01 and BHQ880 were not included due to observed effects on other (non-Wnt) pathways.
Most studies of Porcupine inhibitors have focused on tumor models with ligand-dependent Wnt pathway activation, under the assumption that downstream pathway activation such as that seen with mutations in APC and β-catenin would be unaffected by extracellular Wnt levels. Indeed, it has been reported that the Porcupine inhibitor LGK974 accelerates tumorigenesis in APC-deficient mice models by selectively blocking wild-type stem-cell proliferation, and thus favouring growth of APC-mutant cells (Huels et al., 2018). However, it has also been reported that Porcupine inhibitors could inhibit growth of a colon cancer cell line with mutations in CTNNB1 (Voloshanenko et al., 2013). Other colon cancer cells lines, including some with APC mutations also appeared dependent on Wnt/Fzd signaling for growth (Vincan et al., 2007). Small molecule Porcupine inhibition in these cell lines resulted in decreased pathway activation, anti-proliferative effects and reduced cell viability. Taken together these data suggest patients with cancers bearing a subset of downstream mutations in the Wnt pathway, traditionally expected to be insensitive to Porcupine inhibitors, may well in fact receive therapeutic benefit from such treatment. However, further research is needed to determine which APC/CTNNB1 mutation subtypes retain Wnt dependence, and the mechanism(s) involved.

### 10. Effect of porcupine inhibitors on cancer stem cells

Tumor recurrence following chemotherapy occurs through clonal replacement and reactivation of dormant cancer stem cells (CSCs) (Jahanban-Esfahan et al., 2019). Canonical Wnt signaling prevents differentiation of CSC and promotes their retention to fuel tumor expansion, under the assumption that downstream pathway activation by direct effects on cancer cells and/or cell-stromal/immune interactions (Regan et al., 2017; Vermeulen et al., 2010). These findings go some way to reconcile the paradoxical observation that despite all cells within a tumor harbour Wnt-activating mutation, only a subfraction of cancer cells display CSC properties. Non-canonical Wnt signaling also promotes survival and resistance to therapy of CSCs through PI3K-AKT signaling activation (reviewed in Katoh, 2017). The Porcupine inhibitor LGK974 inhibited the ability of breast cancer stem cells, but also via CSC-stromal/immune interactions (Hayashi et al., 2017). Effects of Porcupine inhibitors on migration (Mo et al., 2013), and other Porcupine inhibitors reduced invasion of renal cancer cells in vivo (Mo et al., 2013), and other Porcupine inhibitors reduced invasion of renal cancer cells in vivo (Hayashi et al., 2017). Effects of Porcupine inhibitors on migration in vivo may be due to direct effects on cancer cells and/or cell-nonautonomous effects related to interactions with neighboring stromal tissues.

### 11. Effect of porcupine inhibitors on invasion and metastasis

Epithelial-mesenchymal transition (EMT) has been linked to invasion and metastasis in intestinal cancers. Reactivation of key developmental pathways, including the Wnt pathway, leads to loss of epithelial architecture and disruption of the basement membrane, which enables cancer cells to invade healthy tissues and migrate into the lymphatic system or bloodstream, leading to metastasis.Canonical and non-canonical PCP Wnt pathways have been implicated in CRC cell migration (Al-Aynati et al., 2004; Katoh, 2017; Y. Zhang et al., 2016; Z. Zhang, Wang, Zhang, & Yang, 2018). The Porcupine inhibitor IWP-2 was shown to reduce migration and invasion of a gastric cancer cell line (Mo et al., 2013), and other Porcupine inhibitors reduced invasion of renal cancer cells in vivo (Li et al., 2020) and prolonged metastasis-free survival in an in vivo model of Ewing sarcoma (Hayashi et al., 2017). Effects of Porcupine inhibitors on migration in vivo may be due to direct effects on cancer cells and/or cell-nonautonomous effects related to interactions with neighboring stromal tissues.

### 12. Effects of porcupine inhibitors on fibrosis

Another corollary of EMT is the induction of fibrosis. Wnt signaling has long been implicated in fibrosis (Akhmetshina et al., 2012) and stromal signaling plays a key role in cancers, with growing evidence that cancer-associated fibroblasts (CAFs) play a role in immunosuppression as well as remodelling (Hilmi, Bousquet, Neuville, & Neuzillet, 2020). Wnt5a functions as a supportive niche factor in gastric cancer, where it is upregulated by the tumor microenvironment (Hayakawa et al., 2015; Maeda et al., 2020). Wnt5a is also associated with the strong desmoplastic reaction observed in pancreatic cancer (Pilarsky et al., 2008). In colorectal cancers, upregulation of endogenous RSPO2/3 was appeared dependent on Wnt/Fzd signaling for growth (Vincan et al., 2007). Small molecule Porcupine inhibition in these cell lines resulted in decreased pathway activation, anti-proliferative effects and reduced cell viability. Taken together these data suggest patients with cancers bearing a subset of downstream mutations in the Wnt pathway, traditionally expected to be insensitive to Porcupine inhibitors, may well in fact receive therapeutic benefit from such treatment. However, further research is needed to determine which APC/CTNNB1 mutation subtypes retain Wnt dependence, and the mechanism(s) involved.

### Table 2: Porcupine Inhibitors in Cancer Clinical Trials

| Drug Identifier | Trial identifier | Regimen | Cancer Indication | Clinical Phase |
|----------------|-----------------|---------|------------------|---------------|
| WNT974 (previously LGK974) | NCT02278133 | In combination with LGX818 and Cetuximab Monotherapy | Metastatic colorectal cancer with BRAFV600 mutations and RSPO fusions or RNF43 mutations | Phase I/II |
|                | NCT01351103 | OR Combination with PDR001 (Spartializumab) | Advanced solid tumors with Wnt pathway activation (monotherapy) | Phase I |
| ETC-159        | NCT02521844 | Monotherapy OR Combination with Pembrolizumab or Denosumab | Various advanced malignancies naïve or refractory to PD-L1 inhibitors (combination with PDR001) | Phase I |
| CGX1321        | NCT02675946 | Monotherapy OR Combination with Pembrolizumab | Advanced solid tumors (monotherapy) | Phase I |
| RXC004         | NCT03507998 | Monotherapy OR Combination with Pembrolizumab | Advanced colorectal cancer (combination) | Phase I |
|                | NCT03447470 | Monotherapy and in combination with Nivolumab | Advanced G1 tumors | Phase I |
|                | NCT04907539 | Monotherapy and in combination with Nivolumab | Advanced solid tumors | Phase II |
|                | NCT04907851 | Monotherapy and in combination with Nivolumab | Advanced solid tumors (combination) | Phase II |

Source: Clinicaltrials.gov. Data collected on 10th Jan 2022. Withdrawn trials not included
strongly associated with a defined CAF signature and with the CMS4 (mesenchymal) consensus molecular subtype (Kleeman et al., 2020). Although data on fibrosis is lacking from cancer models, Porcupine inhibitors have demonstrated profound anti-fibrotic effects in models of skin, lung and kidney fibrosis (Bunyard et al., 2019; C.-W. Chen et al., 2017; Madan, Patel, et al., 2016).

13. Reversal of immune evasion by porcupine inhibition

Immune evasion plays a key role in tumor development. The presence of Wnt ligands in the microenvironment in gastrointestinal tumors positions them perfectly to impact the local tumor immune response. There is strong association between upregulation of Wnt signaling and downregulation of immune markers in cancers, including those of the gastrointestinal tract (Luke et al., 2019; Spranger, Bao, & Gajewski, 2015). Tumors may be classed as “hot” (i.e. inflamed), or “cold”, the latter state characterized by a tumor microenvironment lacking infiltrating lymphocytes and with high levels of suppressive regulatory T-cells (Ochoa de Olza, Navarro Rodrigo, Zimmermann, & Coukos, 2020). Cold tumors do not respond to immunotherapy agents. Of note, several canonical and non-canonical Wnt ligands and Frizzled receptors were upregulated in melanoma patients that failed to respond to anti-PD-1 therapy. The Porcupine inhibitor C59 reversed the ability of melanoma cancer cell-conditioned media to induce generation of dendritic cell-mediated regulatory-T cells in vitro (DeVito et al., 2021). In the immune cold B16F10 (C57BL/6 mice) melanoma tumor model, anti-PD-1 therapy was not effective, but treatment with the Porcupine inhibitor RXC004 was able to reduce tumor size both as a monotherapy and in combination with an anti-PD-1 inhibitor. Flow cytometry showed that the RXC004 treatment reduced the myeloid derived suppressor cells in the tumor microenvironment (Phillips et al., 2019, Phillips et al, manuscript submitted for publication). A combination of the Porcupine inhibitor C59 with anti-CTLA4 antibodies was also shown to have synergistic anti-tumor effects in the same model (Holzthausen et al., 2015). These data suggest that Porcupine inhibitors may be effective in combination with immunotherapy in patients that currently do not respond to such therapies.

Further evidence that Porcupine inhibitors can reverse Wnt pathway-induced immune evasion comes from the immune hot CT26/BALBc colorectal syngeneic model, where RXC004 in combination with anti-PD-1 antibody resulted in a significant increase in the cytotoxic to regulatory T-cell ratio within the tumor. This was not seen in either monotherapy treatment arm (Phillips et al., 2019, Phillips et al, manuscript submitted for publication). The ability to reverse Wnt ligand induced immune evasion in the wider tumor microenvironment may provide an explanation for the observed anti-tumor effects of Porcupine inhibitors even in cases where Wnt pathway is activated by downstream mutations within the tumor cell.

14. Potential toxicities of porcupine inhibitors identified in vivo

Wnt ligands play a vital homeostatic role in stem cell renewal, which means targeting the pathway has the potential for adverse effects, particularly within the intestinal epithelium. Ectopic expression of the negative Wnt regulator Dkk1 in mice caused loss of intestinal architecture (Kuhnert et al., 2004; Pinto, Gregorieff, Begthel, & Clevers, 2003), and high doses of Porcupine inhibitors have adverse effects on stem cell renewal with associated disruption of villi architecture (Kabiri et al., 2018). However, anti-tumor effects of Porcupine inhibitors have been observed at doses that have no effect on the normal intestine (Koo et al., 2015; J. Liu et al., 2013; Phillips et al, manuscript submitted for publication), suggesting there may be a safe therapeutic window for these compounds. The effects of Dkk1 overexpression on intestinal architecture were reversible, consistent with other studies that indicate the intestine has the ability to regenerate after insult (Kuhnert et al., 2004). A possible explanation for this may be redundancy in the stem cell populations, with HOXP-expressing stem cells, typically located higher in the intestinal crypt, able to compensate for loss of LGR5 expressing stem cells at the base of crypts, and vice-versa (Takeda et al., 2011), although such plasticity may be a relatively short-lived response to injury. Stem cells located at the intestinal crypt base are exposed to the highest levels of Wnt ligand provided by the niche (Paneth cells and stromal cells), and therefore may be more protected from drug-induced reduction of Wnt secretion (Kosinski et al., 2007). Consistent with this, mice with genetic ablation of PORCN in intestinal epithelial cells retained normal intestinal homeostasis due to compensatory Wnt secretion from neighboring stromal cells (Kabiri et al., 2014). Furthermore, pericryptal stromal cells have high expression of drug transporters, protecting the stem cell niche from xenobiotics, including Porcupine inhibitors (Chee et al., 2018).

Other in vivo toxicities observed with Porcupine inhibitors include loss of bone volume and density, most likely due to effects on osteoclasts. However, these effects could be mitigated by prophylactic treatment with bisphosphonates (Madan et al., 2018).

15. Porcupine inhibitors in the clinic

To date, five Porcupine inhibitors have entered phase I/II clinical trials (Table 2) in patients with advanced solid tumors. In NCT02521844, a maximum tolerated dose of 30 mg for ETC-159 (previously known as ETC-1922159) was established, limited by high bone marker turnover and compression fractures. Bone effects at lower concentrations were mitigated by prophylactic treatment with denosumab. Dysgeusia (loss or change in taste) was common, even at lower doses. Target engagement was demonstrated by inhibition of Axin2 expression in hair follicles and increased infiltration of T-cells into the tumor microenvironment. There were no objective responses, although 17% (2/12) patients had stable disease (Tan et al., 2018).

The WNT974 (previously LGK974) monotherapy dose-finding study did not find a maximum tolerated dose. Dysgeusia was the most common side effect, although bone effects were also observed in a subset of patients. Target engagement was demonstrated by reduction of Axin2 expression in skin. There were no objective responses; 16% of 98 patients had stable disease (Rodon et al., 2021). In this study, 28 patients were genetically selected for Wnt pathway activation as part of a dose expansion arm, the majority of which had RNF43 mutations. Of these 28 patients, 10 patients had stable disease. This result has led to others concluding that monotherapy Porcupine inhibitor treatment failed even with this enrichment for upstream Wnt pathway aberrations. However, since this dose expansion arm was run in 2015, knowledge of exactly which RNF43 mutations result in loss of function has increased (Spit et al., 2020; Tu et al., 2019; Yu et al., 2020). With our current improved understanding, and excluding patients with co-occurring downstream Wnt pathway mutations, only 8 of these patients would now be considered to have RNF43 loss of function mutations. Of these 8 patients 7 had a best tumor response between -27 and +20%, suggesting a stringent patient selection strategy is likely needed in the clinic for Porcupine inhibitors (Rodon et al., 2021). When WNT974 was paired with the anti-PD-1 antibody spartalizumab, dose-limiting toxicities were reported in 2 patients, including one spinal compression fracture. One patient (4%) with triple-negative breast cancer had a partial response, 11 patients (41%) had stable disease, 13 patients (48%) had progressive disease. 75% of patients experienced a treatment-related adverse events, with dysgeusia and osteopenia among the most common (Janku et al., 2020).

Preliminary data from NCT03447470, a dose finding study for RXC004 in solid tumors, has recently been reported. The most common treatment related adverse events were fatigue, nausea, dysgeusia, vomiting and anorexia. No grade 4/5 adverse events or bone fragility events were reported, with patients receiving prophylactic denosumab. Five patients, all with Wnt pathway activated tumors, had stable disease, including one patient with biliary tract cancer, and two CRC
patients with either an RNF43 mutation or a RSPO fusion (Cook et al., 2021).

These early clinical studies have identified bone loss and dysgeusia as class effects of Porcupine inhibitors. The latter likely arises because Wnt signaling has been shown to be vital for proliferation of taste-progenitors and formation of taste buds (Gaillard et al., 2017; Prochazkova et al., 2017). Prophylactic treatment with bisphosphonates or denosumab has been shown to mitigate increases in bone turnover markers (Cook et al., 2021; Tan et al., 2018). Encouragingly, gastrointestinal side effects appear to be limited and manageable. Furthermore, the limited efficacy observed to date may be attributable to the studies being performed in heavily-pretreated patients with poor prognosis and lack of appropriate patient selection.

16. Conclusions and future perspectives

Porcupine inhibitors have the potential for multiple potentially beneficial effects on tumor biology (Fig. 3). There is substantial preclinical data supporting the use of Porcupine inhibitors in genetically-defined populations of patients with gastrointestinal cancers. Functionally relevant mutations are most common in CRC and pancreatic cancers. Patients with RNF43 or ZNRF3 truncations may be amenable to treatment and mutations in RNF43 are particularly common in MSI high CRC and pancreatic cancers, although it should be noted that the majority of RNF43 mutations found in MSI disease are the hotspot G659Vfs*41 frameshift, which has been reported to encode functional protein and may be simply a passenger mutation in this context (Tu et al., 2019). Tumors with RSP02/3 fusion and RSP03 stromal overexpression are also likely to be susceptible to Porcupine inhibition, an intriguing prospect for CMS4 CRC subtype patients, where RSP03 overexpression is common. There is also data to suggest Porcupine inhibitors may be effective in patients where Wnt signaling is activated by non-genetic/epigenetic mechanisms, and in certain patients with downstream Wnt pathway aberrations. However further work is needed to understand the processes involved, and to develop biomarkers to robustly identify such patients.

The effects of Porcupine inhibitors in reversing immune evasion provide a strong rationale for their use in combination with checkpoint inhibitors, particularly in patients previously unresponsive to anti-PD-1/PD-L1 therapy. MSI high CRC patients are currently indicated for anti-PD-1 therapy and exploration of anti-PD-1/Porcupine inhibitor combinations in these patients may also be warranted. Similarly, Porcupine and BRAF inhibitors may be an attractive combination in CRC patients with both BRAF and RNF43 mutations, particularly in light of the recent data from the BEACON study which demonstrated benefits of combining the BRAF inhibitor encorafenib with cetuximab in this setting (Tabernero et al., 2021). Additional combination opportunities are being explored pre-clinically, with synergy observed between ETC-159 and PI3K inhibitors or ETC-159 and PARP inhibition in Wnt-driven cancer models (Kaur et al., 2021; Z. Zhong et al., 2019).

Given the role of Wnt pathway in CSC maintenance, combination of Porcupine inhibitors with chemotherapy in selected patients may provide an interesting avenue of research, based on the rationale that Porcupine inhibitors may overcome chemoresistance. Initial clinical data is promising in that, despite the homeostatic role of Wnt in the digestive tract, gastrointestinal side-effects at doses that engage the target are limited, indicating the presence of a therapeutic window. Effects on bone turnover can be mitigated with Receptor activator of nuclear factor-κB ligand (RANKL) inhibitors or bisphosphonate treatment, although the additional impact on taste may need to be managed in order to minimize associated weight loss. Since future strategies may include combining Porcupine inhibitors with other therapeutics, the potential for combined toxicities will also need to be managed. However initial data from NCT01351103, where WNT974 was used in combination with anti-PD-1 therapy showed the combination to be well-tolerated (Janku et al., 2020).

![Porcupine Inhibitors Block Proliferation in Genetically Defined Tumors, Promote Differentiation and Stimulate Immune Responses](image.png)
Conclusive clinical efficacy data on Porcine inhibitors is currently lacking, with studies to date limited to small numbers of late stage, previously-treated patients with relevant somatic mutations. Key questions remain concerning optimal patient selection strategies for Porcine inhibitors. Different approaches are being explored, including prospectively targeting patients with RSP02/3 fusions and RNF43/ZNRF3 mutations with monotherapy, but also broader populations. WNT974 is under investigation in combination with anti-PD-1 in patients refractory to prior anti-PD-1 therapy, as well as other targeted populations. It is also interesting to note differences in dosing strategies between various Porcine inhibitors; In NCT01351103, WNT974 was given as a priming dose when used in combination with anti-PD-1. RXC04 is dosed one daily, and ETC-159 demonstrated interpatient variability in pharmacokinetics and was dosed every other day. The impact of distinct dosing regimens and patient selection strategies is yet to be determined. Large scale, randomized, controlled clinical trials with robust biomarker assessments will be needed to elucidate whether Porcine inhibitors will prove useful in gastrointestinal cancers, and to identify which patients are most likely to benefit from such treatments.

Declaration of Competing Interest

O.J.S acknowledges core grant A17196 and A31287 from Cancer Research UK. Medical writing assistance was provided by Marianne Ratcliffe, PhD, of EmWrite Ltd, and was funded by Redx Pharma PLC

References

Alhmethysta, A., Palumbo, K., Dees, C., Bergmann, C., Venalis, P., Zerr, P., ... Distler, J. H. W. (2012). Activation of canonical Wnt signalling is required for TGF-beta-mediated fibrosis. Nature Communications 3, 735.

Al-Aynati, M. M., Radulovich, N., Riddell, R. H., & Tsao, M. S. (2004). Epithelial-cadherin

Al-Aynati, M. M., Radulovich, N., Riddell, R. H., & Tsao, M. S. (2004). Epithelial-cadherin

Acknowledgements

O.J.S acknowledges core grant A17196 and A31287 from Cancer Research UK. Medical writing assistance was provided by Marianne Ratcliffe, PhD, of EmWrite Ltd, and was funded by Redx Pharma PLC

Cancer Genome Atlas Network. (2012). Comprehensive molecular characterization of human colon and rectal cancer. Nature 487, 330–337.

Cerami, E., S. L. Davis, S. B. L. Kaufmann, G., Koes, N., & et al. (2012). The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discovery 2(5), 401 LP – 404.

Chan-On, W., Nair, S. R., Kim, H., Lim, W. K., Kim, K. W., & et al. (2018). Exome sequencing identifies distinct mutational patterns in liver fibrosis and non-infection-related bile duct cancers. Nature Genetics 45(12), 1474–1478.

Chee, Y. C., Pakhrin, J., Bunte, R., Adoil, V. A., Madan, B., & Virshup, D. M. (2018). Intrinsic Wnt-mediated resistance of the intestinal stem cell niche. Developmental Cell 46(6), 681–695.e5.

Chen, B., Dodge, M. E., Tang, W., Liu, J., Ma, Z., Fan, C., ... Lum, L. (2009). Small molecule-mediated disruption of Wnt-dependent signaling in tissue regeneration and cancer. Science 322(5898), 1478–1482.

Chen, C.-W., Beyer, C., Liu, E., Maier, C., Li, C., Trinh-Minh, T., ... Distler, J. H. W. (2017). Pharmacological inhibition of porcine induces regression of experimental skin fibrosis by targeting Wnt signalling. Annals of the Rheumatic Diseases 76(4), 773–778.

Chen, N., Chen, Z., Yang, X., & Xiao, Q. (2021). The involvement of noncanonical Wnt signalling in cancers. Biomembrane and Pharmacology 133, Article 110946.

Chiuriolo, M. A. (2015). Role of the Wnt/b-catenin pathway in gastric cancer: An in-depth literature review. World Journal of Experimental Medicine 5(2), 84–102.

Clements, W. M., Wang, J., Sarnaik, A., Kim, O., J. Ma, D., Fennoglio-Preser, C., ... Lowy, A. M. (2002). β-catenin mutation is a frequent cause of Wnt pathway activation in gastric cancer. Cancer Research 62(12), 3501–3506.

Clevers, H., & Nusse, R. (2012). Wnt/b-catenin signaling and disease. Cell 149(6), 1192–1205.

Colnot, S., Decaens, T., Niwa-Kawakita, M., Godard, C., Hamard, G., Kahn, A., Giovannini, M., & Perret, C. (2004). Liver-targeted disruption of Apc in mice activates-catenin signaling and leads to hepatocellular carcinomas. Proceedings of the National Academy of Sciences USA 101(49), 17216–17221.

Conboy, C. B., Velez-Reyes, G. L., Tsirka, B. R., Hu, H., Kaufmann, G., Koes, N., ... Conboy, C. B., Velez-Reyes, G. L., Tsirka, B. R., Hu, H., Kaufmann, G., Koes, N., ... Tsirka, B. R., Hu, H., Kaufmann, G., Koes, N., ... and others (2004). Liver-targeted disruption of Apc in mice activates-catenin signaling and leads to hepatocellular carcinomas. Proceedings of the National Academy of Sciences USA 101(49), 17216–17221.

Cook, N., Blagdon, S., Lopez, J., Sarker, D., Greystoke, A., Harris, N., ... Cook, N., Blagdon, S., Lopez, J., Sarker, D., Greystoke, A., Harris, N., ... and others (2004). Liver-targeted disruption of Apc in mice activates-catenin signaling and leads to hepatocellular carcinomas. Proceedings of the National Academy of Sciences USA 101(49), 17216–17221.

De Laou, W., Peng, W. C., Gros, P., & Clevers, H. (2021). The R-spondin/Lgr5/Rnf43 module: regulator of Wnt signal strength. Genes and Development 28, 305–314.

DeVito, N. C., Sturdivant, M., Thievanthiran, B., Xiao, C., Plebanek, M. P., Salama, A. K., ... and others (2021). Pharmacological Wnt ligand inhibition overcomes key tumor-mediated resistance pathways to anti-PD-1 immunotherapy. Cell Reports 35, Article 109071.

Diamond, J. R., Becerra, C., Richards, D., Mitra, A., Osborne, C., O'Shaughnessy, J., ... and others (2020). Phase Ib clinical trial of the anti-frizzled antibody vantictumab (OMP-18R5) plus paclitaxel in patients with locally advanced or metastatic HER2-negative breast cancer. EBCTCG Best Cancer Research and Treatment 184(1), 51–62.

Dotan, E., Cardin, D. B., Shahda, S., Lenz, H., ... Dotan, E. and others (2020). Phase Ib dose escalation study of Wnt pathway inhibitor vantictumab in conjunction with nab-paclitaxel and gemcitabine in patients with previously untreated metastatic pancreatic cancer. Investigational New Drugs 38, 821–830.

De Laau, W., Peng, W. C., Gros, P., & Clevers, H. (2021). The R-spondin/Lgr5/Rnf43 module: regulator of Wnt signal strength. Genes and Development 28, 305–314.

Dow, L. E., O'Rourke, K. P., Simon, J., Tschaharganeh, D. F., Van Es, J. H., ... Dow, L. E., O’Rourke, K. P., Simon, J., Tschaharganeh, D. F., Van Es, J. H., ... and others (2015). Apc restoration promotes cellular differentiation and reestablishes crypt homeostasis in colorectal cancer. Cell 161(7), 1539–1552.

El-Khtayyat, B., Khan, N., Yang, Y., Cole, S., Kahn, M., Zaghloul, N., ... and others (2021). Evaluation of anti-tumor effects of aspirin and LGK974 drugs on cellular signaling pathways, cell cycle and apoptosis in colorectal cancer cell lines compared to oxaliplatin drug. Fundamental and Clinical Pharmacology 34(1), 51–64.

El-Khtayyat, B., Khan, N., Yang, Y., Cole, S., Kahn, M., Zaghloul, N., ... and others (2021). Evaluation of anti-tumor effects of aspirin and LGK974 drugs on cellular signaling pathways, cell cycle and apoptosis in colorectal cancer cell lines compared to oxaliplatin drug. Fundamental and Clinical Pharmacology 34(1), 51–64.

El-Khtayyat, B., Khan, N., Yang, Y., Cole, S., Kahn, M., Zaghloul, N., ... and others (2021). Evaluation of anti-tumor effects of aspirin and LGK974 drugs on cellular signaling pathways, cell cycle and apoptosis in colorectal cancer cell lines compared to oxaliplatin drug. Fundamental and Clinical Pharmacology 34(1), 51–64.

El-Khtayyat, B., Khan, N., Yang, Y., Cole, S., Kahn, M., Zaghloul, N., ... and others (2021). Evaluation of anti-tumor effects of aspirin and LGK974 drugs on cellular signaling pathways, cell cycle and apoptosis in colorectal cancer cell lines compared to oxaliplatin drug. Fundamental and Clinical Pharmacology 34(1), 51–64.

El-Khtayyat, B., Khan, N., Yang, Y., Cole, S., Kahn, M., Zaghloul, N., ... and others (2021). Evaluation of anti-tumor effects of aspirin and LGK974 drugs on cellular signaling pathways, cell cycle and apoptosis in colorectal cancer cell lines compared to oxaliplatin drug. Fundamental and Clinical Pharmacology 34(1), 51–64.
Holtzhausen, A., Zhao, F., Evans, K. S., Tsutsui, M., Orabona, C., Tyler, D. S., & Hanks, B. A. (2017). Canonical and non-canonical WNT signaling in cancer stem cells and immunotolerance: opportunities for pharmacologic enhancement of immunotherapeutic response. Molecular Cancer Therapeutics 16(11), 2986–3007.

Hoshimura, T., Ogawa, R., Yoshiha, H., Taniguchi, K., Kojima, M., Saito, Y., & Sekine, S. (2019). EIF1E–RSPO2 and PIEZO1–RSPO2 fusions in colorectal traditional serrated adenoma. Histopathology 75(2), 266–273.

Hayaoka, Y., Aoyama, H., Stanicakova, J., Sakahara, K., Asahara, Y., Renz, B. W., & Wang, T. C. (2019). Wnt6 expression is related to the normal and neoplastic gastric epithelial tissue and is supported by a perivascular stem cell niche. Cancer Cell 28(6), 800–814.

Hayasli, M., Baker, G., Goldstein, S. D., Albert, C. M., Jackson, K. W., McCarty, C., & Loeb, D. M. (2017). Inhibition of Wnt/β-Catenin pathway parallels early survival in a mouse xenograft model of Ewing sarcoma. Oncotarget (84), 78265–78276.

Hilken, J., Timmer, N. C., Boer, M., Ilink, G. J., Schewe, M., Sacchetti, A., & Bakker, E. R. M. (2017). RSPO3 expands intestinal stem cell niche and imparts properties of stemness. Gut 66, 1264–1274.

Hilmi, M., Nicolle, R., Bousquet, C., & Neuzillet, C. (2020). Cancer-associated fibroblasts accomplices in the tumor immune evasion. Cancers 12, 2996.

Huibek, F., Spaderna, S., Schmalhofer, O., Jung, A., Kirchner, T., & Brabletz, T. (2007). Wnt/FZD signaling and colorectal cancer morphogenesis. Frontiers in Bioscience : A Journal and Virtual Library 12, 458–470.

Holzhausen, A., Zhao, F., Evans, K. S., Tsutsui, M., Orabona, C., Tyler, D. S., & Hanks, B. A. (2017). Melanoma-derived Wnt5a promotes local dendritic-cell expression of IDO and immune-suppressive crosstalk that provides a potential approach for enhanced immunotherapy. Cancer Immunology Research 3(9), 1082–1095.

Horii, A., Nakatsuji, S., Miyoshi, Y., Ichii, S., Nagase, H., Kato, Y., Yanagisawa, A., & Nakamura, Y. (1992). The APC gene, responsible for familial adenomatous polyposis, is mutated in human gastric cancer. British Journal of Cancer 66, 655–661.

Huang, J., Fan, X., Wang, L., Yu, Y., Zhou, H., Wang, C., Zhong, L., & Wang, T. (2015). High ORR expression in tumor stroma and cromor is correlated with poor prognosis in pancreatic ductal adenocarcinoma. Scientific Reports 5(1), 1–9.

Huang, S., Xie, Q., Xie, S., Chen, A., Jie, M., Zhang, R., & Zhang, X. (2020). PTEN regulates the Wnt signaling pathway and inhibits clear cell renal cell carcinoma. Bone Research 8, 471–482.

Huebs, D. J., Brussel, L., Hoddler, M., C. M., Cammareri, P., Campbell, A. O., Ridgway, R. A., & Sanatar, J. C. (2017). Increased Wnt7A expression contributes to the growth and differentiation of the murine intestinal stem cells. Scientific Reports 7(1), 1–10.

Jackstadt, R., Hoddler, M., & Sansom, O. J. (2020). Wnt and β-Catenin in Cancer: Genes and Therapy. The Annual Review of Cancer Biology 4, 177–196.

Jabbaraghian, R., Seki, S., Djavadi, M., Bettahar, A., A., & A., P. (2019). Tumor cell dormancy: threat or opportunity in the fight against cancer. Cancers 11, 1207.

Janku, F., de Vos, F., de Miguel, M., Forde, P., Ribas, A., Nagasaka, M., & De Jonge, M. (2020). Abstract CT034: Phase I study of WNT974 + spartalizumab in patients (pts) with advanced solid tumors. Cancer Research 80 (16 Supplement), CT034.

Jiang, J., Lan, C., Li, L., Yang, D., Xiao, X., Liao, Q., & Zeng, C. (2018). A novel porcupine inhibitor blocks WNT pathways and attenuates cardiac hypertrophy. Biochimica et Biophysica Acta - Molecular Basis of Disease 1864 (10), 3459–3467.

Jiang, X., Hao, H. X., Gouwens, J. D., Wolttenden, S., Bottiglio, C., Li, N., & Cong, F. (2019). Tankyrase inhibition stabilizes axin and antagonizes Wnt signaling. Nature 561(7726), 614–620.

Hubsch, N., J., Bruys, W., Hoddler, M., Cammareri, P., Campbell, A. O., Ridgway, R. A., & Sansom, J. C. (2017). Increased Wnt7A expression contributes to the growth and differentiation of the murine intestinal stem cells. Scientific Reports 7(1), 1–10.

Jackstadt, R., Hoddler, M., & Sansom, O. J. (2020). Wnt and β-Catenin in Cancer: Genes and Therapy. The Annual Review of Cancer Biology 4, 177–196.

Jabbaraghian, R., Seki, S., Djavadi, M., Bettahar, A., A., & A., P. (2019). Tumor cell dormancy: threat or opportunity in the fight against cancer. Cancers 11, 1207.

Janku, F., de Vos, F., de Miguel, M., Forde, P., Ribas, A., Nagasaka, M., & De Jonge, M. (2020). Abstract CT034: Phase I study of WNT974 + spartalizumab in patients (pts) with advanced solid tumors. Cancer Research 80 (16 Supplement), CT034.

Jiang, J., Lan, C., Li, L., Yang, D., Xiao, X., Liao, Q., & Zeng, C. (2018). A novel porcupine inhibitor blocks WNT pathways and attenuates cardiac hypertrophy. Biochimica et Biophysica Acta - Molecular Basis of Disease 1864 (10), 3459–3467.

Jiang, X., Hao, H. X., Gouwens, J. D., Wolttenden, S., Bottiglio, C., Li, N., & Cong, F. (2019). Tankyrase inhibition stabilizes axin and antagonizes Wnt signaling. Nature 561(7726), 614–620.

Jiang, X., Hao, H. X., Gouwens, J. D., Wolttenden, S., Bottiglio, C., Li, N., & Cong, F. (2019). Tankyrase inhibition stabilizes axin and antagonizes Wnt signaling. Nature 561(7726), 614–620.

Jiang, X., Hao, H. X., Gouwens, J. D., Wolttenden, S., Bottiglio, C., Li, N., & Cong, F. (2019). Tankyrase inhibition stabilizes axin and antagonizes Wnt signaling. Nature 561(7726), 614–620.

Jiang, X., Hao, H. X., Gouwens, J. D., Wolttenden, S., Bottiglio, C., Li, N., & Cong, F. (2019). Tankyrase inhibition stabilizes axin and antagonizes Wnt signaling. Nature 561(7726), 614–620.

Jiang, X., Hao, H. X., Gouwens, J. D., Wolttenden, S., Bottiglio, C., Li, N., & Cong, F. (2019). Tankyrase inhibition stabilizes axin and antagonizes Wnt signaling. Nature 561(7726), 614–620.
Madan, B., Patel, M. B., Zhang, J., Bunte, R. M., Rudemiller, N. P., Griffiths, R., ... Crowley, S. D. (2016). Experimental inhibition of porcine-converted Wnt O-acetylation attenuates atherosclerosis in hyperlipidemic Rsag8/fli1 Δ/Δ mice. Circulation Research 119(11), 1709–1719.

Procházková, M., Hájková, T., Procházka, J., Spurná, J., Trojanová, S., Vlček, R., ... Smalová, J. (2016). Establishment of transgenic pigs expressing modified porcine Wnt5a. Scientific Reports 6, 32499.

Procházková, M., Hájková, T., Procházka, J., Spurná, J., Trojanová, S., Vlček, R., ... Smalová, J. (2016). Establishment of transgenic pigs expressing modified porcine Wnt5a. Scientific Reports 6, 32499.

Procházková, M., Hájková, T., Procházka, J., Spurná, J., Trojanová, S., Vlček, R., ... Smalová, J. (2016). Establishment of transgenic pigs expressing modified porcine Wnt5a. Scientific Reports 6, 32499.

Procházková, M., Hájková, T., Procházka, J., Spurná, J., Trojanová, S., Vlček, R., ... Smalová, J. (2016). Establishment of transgenic pigs expressing modified porcine Wnt5a. Scientific Reports 6, 32499.

Procházková, M., Hájková, T., Procházka, J., Spurná, J., Trojanová, S., Vlček, R., ... Smalová, J. (2016). Establishment of transgenic pigs expressing modified porcine Wnt5a. Scientific Reports 6, 32499.

Procházková, M., Hájková, T., Procházka, J., Spurná, J., Trojanová, S., Vlček, R., ... Smalová, J. (2016). Establishment of transgenic pigs expressing modified porcine Wnt5a. Scientific Reports 6, 32499.

Procházková, M., Hájková, T., Procházka, J., Spurná, J., Trojanová, S., Vlček, R., ... Smalová, J. (2016). Establishment of transgenic pigs expressing modified porcine Wnt5a. Scientific Reports 6, 32499.
Van De Wetering, M., Frances, H. E., Francis, J. M., Bounova, G., Iorio, F., Pronk, A., ... Clevers, H. (2015). Prospective derivation of a living organoid biobank of colorectal cancer patients. Cell 161(4), 933–945.

Vermeulen, L., De Sousa, E., Melo, F., Van Der Heijden, M., Cameron, K., De Jong, J. H., ... Medema, J. P. (2010). Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. Nature Cell Biology 12(5), 468–476.

Vinci, E., Darcy, P. K., Farrelly, C. A., Faux, M. C., Brabletz, T., & Ramsay, R. G. (2007). Frizzled-7 dictates three-dimensional organization of colorectal cancer cell carcinoids. Oncogene 26, 2340–2352.

Voloshanenko, O., Erdmann, G., Dubash, T. D., Augustin, I., Metzig, M., Moffa, G., ... Boutros, M. (2013). Whole-genome sequencing and comprehensive molecular profiling identify new driver mutations in gastric cancer. Nature Genetics 46(6), 573–582.

Wang, K., Yuan, S. T., Xu, J., Lee, S. P., Yan, H. N. S., Shi, S. T., ... Leung, S. Y. (2014). Whole-genome sequencing and comprehensive molecular profiling identify new driver mutations in gastric cancer. Nature Genetics 46(6), 573–582.

Wang, S., Tian, Y., Wu, D., Zhuo, Y., Zhou, Y., Zhou, J., & Zhang, Z. (2012). Wnt secretion is required to maintain high levels of Wnt activity in colon cancer cells. Nature Communications 4, 2610.

Wang, K., Yuan, S. T., Xu, J., Lee, S. P., Yan, H. N. S., Shi, S. T., ... Leung, S. Y. (2014). Whole-genome sequencing and comprehensive molecular profiling identify new driver mutations in gastric cancer. Nature Genetics 46(6), 573–582.

Wang, T., Kalland, K., -H, Ke, X., & Qu, Y. (2018). Targeting Wnt/β-Catenin signaling for cancer immunotherapy. Trends in Pharmacological Sciences 39(7), 648–658.

Weinberg, B. A., Xu, J., Lindberg, M. R., Shields, A. F., Hwang, J. J., Poorman, K., ... Morse, M. A. (2019). Molecular profiling of biliary cancers reveals distinct molecular alterations and potential therapeutic targets. Journal of Gastrointestinal Oncology 10(4), 652–662.

White, B. D., Chien, A. J., & Dawson, D. W. (2012). Dysregulation of Wnt/β-catenin signaling in gastrointestinal cancers. Gastroenterology 142(2), 219–232.

Witkiewicz, A. K., McMillan, E. A., Balaji, U., Baek, G. H., Lin, W. C., Mansour, J., ... Knudsen, E. S. (2015). Whole-exome sequencing of pancreatic cancer defines genetic diversity and therapeutic targets. Nature Communications 6, 1–11.

Woodcock, S., Bhamra, I., Jones, C., Cook, A. E., Eagle, C., & Phillips, C. (2019). Abstract 3874: Efficacy of the Wnt/β-Catenin pathway inhibitor RXC004 in genetically-defined models of cancer. Cancer Research 79(13 Supplement), 3874.

Xavier, C. P., Melikova, M., Chuman, Y., Uren, A., Baljinnyam, B., & Rubin, J. S. (2014). Secreted Frizzled-related protein potentiation versus inhibition of Wnt3α/β-catenin signaling. Cellular Signalling 26(1), 94–101.

Xu, G. L., Shen, J., Xu, Y. H., Wang, W. S., & Ni, C. (2018). ROR1 is highly expressed in circulating tumor cells and promotes invasion of pancreatic cancer (GU-LiLe). Molecular Medicine Reports 18, 5087–5094.

Yaeger, R., Chatila, W. K., Lipsy, M. D., Hechtman, J. F., Cercek, A., Sanchez-Vega, F., ... Schultz, N. (2018). Clinical sequencing defines the genomic landscape of metastatic colorectal cancer. Cancer Cell 33(1), 125–136.

Yu, J., Mohamed Yusoff, P. A., Woutersen, D. T. J., Goh, P., Harmston, N., Smits, R., ... Madan, B. (2020). The functional landscape of patient-derived RNF43 mutations predicts sensitivity to Wnt inhibition. Cancer Research 80(24), 5619–5632.

Zhang, W., Glockner, S. C., Guo, M., Ota Machida, E., Wang, D. H., Easwaran, H., ... Baylin, S. B. (2008). Epigenetic Inactivation of the Canonical Wnt Antagonist SFRP-Box Containing Gene 17 in Colorectal Cancer. Cancer Research 68(5), 2764–2772.

Zhang, Y., Lin, L., Jin, Y., Liu, Y., Cao, Y., & Zheng, C. (2016). Overexpression of WNT3B promotes COLO 205 cell migration and invasion through the JNK signaling pathway. Oncology Reports 26(1), 23–30.

Zhang, Z., Wang, Y., Zhang, J., Zhong, J., & Yang, R. (2018). COL1A1 promotes metastasis in colorectal cancer by regulating the WNT/PCP pathway. Molecular Medicine Reports 17, 5037–5042.

Zhao, Z., Lu, P., Zhang, H., Xu, H., Gao, N., Li, M., & Liu, C. (2014). Nestin positively regulates the Wnt/β-catenin pathway and the proliferation, survival and invasiveness of breast cancer stem cells. Breast Cancer Research 16(4), 1–12.

Zhong, Y., Kavalkolos, P., Nguyen, T., Lau, T., Boggs, J., Sambrone, A., Kan, D., Merchant, M., Harstad, E., Diaz, D., Costa, M., & Schutteren, M. (2016). Tankyrase Inhibition Causes Reversible Intestinal Toxicity in Mice with a Therapeutic Index < 1. Toxicologic Pathology 44(2), 267–278.

Zhong, Z., Sepramaniam, S., Chew, X. H., Wood, K., Lee, M. A., Madan, B., & Virshup, D. M. (2019). PORCN inhibition synergizes with PI3K/mTOR inhibition in Wnt-addicted cancers. Oncogene 38(46), 6692–6677.

Zhou, J., -K, Zheng, Y., -Z, Liu, X. -S., Gou, Q., Ma, R., Guo, C. -L., ... Peng, Y. (2017). ROR1 expression as a biomarker for predicting prognosis in patients with colorectal cancer. Oncotarget 8(20), 32864–32872.

Zhou, Kipps, T. J., & Zhang, S. (2017). Wnt5a signaling in normal and cancer stem cells. Stem Cells International 2017, 1–6. Article ID 5295286.

Zilberberg, A., Lahav, L., & Rosin-Arbesfeld, R. (2010). Restoration of APC gene function in colorectal cancer cells by aminoglycoside- and macrolide-induced read-through of premature termination codons. Gut 59(4), 496–507.

Zimmerli, D., Cecconi, V., Valenta, T., Hausmann, G., Cantó, C., Restivo, G., ... Van Den Broek, M. (2018). WNT ligands control initiation and progression of human papillomavirus-driven squamous cell carcinoma. Oncogene 37, 3753–3762.