Pathophysiology of chronic limb ischemia

Introduction to the topic

Chronic limb ischemia (CLI) generally occurs in old age and is a common reason for treatment. Due to the slow progression of symptoms, the transition to critical ischemia can appear to “come as a surprise.” It is important to have knowledge and understanding of the pathophysiological processes in limbs damaged by chronic ischemia in order to provide these patients with adequate treatment and prevent or at least delay further damage.

Introduction

Chronic tissue ischemia in the lower extremities is a common problem encountered in vascular surgical departments. Peripheral arterial occlusive disease (PAOD) affects more than 202 million people worldwide [9]. Approximately 3% of all hospitalizations in Germany are due to PAOD, with a sharp rise being seen in critical ischemia [16]. More than 20% of over 65-year-olds already have PAOD [7], whereby the rate of symptomatic forms in men exceeds 8% [15]. A PAOD is the result of a gradual process that progressively narrows the arterial lumen through which blood flows. This vasoconstriction is generally a sign of atherosclerotic changes in the vessel walls. Typical risk factors include advanced age, nicotine abuse, diabetes mellitus, hypercholesterolemia, and arterial hypertension [18]. The sequelae of chronic ischemia range from reduced pain-free walking distance to loss of the extremity. Therefore, restoring perfusion is the crucial goal and its success depends on numerous factors. The biological adaptation of chronically undersupplied tissue manifests as the formation of a collateral network. This process is more pronounced the longer and, above all, the slower vasoconstriction develops. Angiogenesis, i.e., the growth of small vessels from pre-existing vessels, and arteriogenesis, dilation of the lumen of pre-existing vessels, can lead to the development of a dense collateral network [27]; however, if PAOD progression persists, these biological adaptation mechanisms become overburdened, i.e., collateral supply is no longer sufficient to compensate for tissue hypoperfusion. This results in increasing cell loss, ultimately leading to a local inflammatory reaction. In the course of inflammation, damaged tissue is broken down and replaced by fibrosis. This additional barrier to oxygen diffusion in the tissue further exacerbates ischemia, resulting in a downward spiralling vicious circle. Depending on the degree of stenosis and the biological reaction, different degrees of PAOD severity develop and can affect patients to varying extents. The classification of PAOD according to Fontaine stages ranges from asymptomatic PAOD in stage I and varying degrees of reduced walking distance in stage II, to pain at rest as well as necrosis and gangrene in stages III and IV. As a result, approximately 25,000 patients per year in Germany require a major amputation as a last resort, of which around 70% are diabetics [12]. Other signs can include trophic changes in the affected limb due to impaired macrocirculation and microcirculation. In contrast to venous ulcers, which are primarily located above the medial malleolus, arterial ulcers are predominantly seen at acral sites. Further signs include cold sensation at these sites, as well as metabolic disorders involving skin appendage changes, such as loss of leg hair.

Approximately 25,000 patients per year in Germany require a major amputation

Irrespective of the stage of PAOD, effective treatment of impaired circulation, in conjunction with individual risk factor adjustment, not least due to the risk of ulcer infection, is crucial to the success of treatment for PAOD, which would progressively deteriorate if left untreated.

Molecular mechanisms of atherosclerosis and atheroma development

A number of factors are expressed in healthy endothelium, such as prostacyclin, nitric oxide (NO), thrombomodulin, and plasminogen activators, all of which have a strongly platelet-repellent, anticoagulant, and fibrinolytic effect. In addition, the endothelium has a high negative charge, explaining why the also negatively charged platelets are repelled from the vessel wall. This makes the luminal side of each vessel a strong barrier to thrombotic factors.
Cell-to-cell contacts are particularly important in the regulation of endothelial permeability. Important complexes for the regulation of these compounds include occludin, adhesion molecules, cadherins, gap junctions, and integrin receptors such as fibronectin [17].

The principal cause of vasoconstriction is atherosclerosis, a chronic process that goes unnoticed for years. The process starts with stress factors such as hyperlipidemia (LDL), free radicals, arterial hypertension, infections, or subclinical inflammation, which impair endothelial homeostasis. This leads to impairment of the stressed endothelium and increased permeability to lipids, which build up in the intima causing further endothelial stress. This mutually reinforcing cycle enables lipids to penetrate deeper into the vascular wall layers and accumulate in the extracellular matrix. Oxidative stress and enzymatic modifications produce oxidized LDL, which triggers strong inflammatory activity [29]. Immune system cells such as monocytes are attracted by these lipid accumulations and migrate into the vascular wall; however, not only the innate but also the acquired immune system contributes to atheroma development by activating T-cells and antibody production. Cytokines such as interleukin 18 (IL-18) mediate this interaction between different immune cells [1].

Migrated monocytes differentiate into macrophages in the subendothelial tissue, where they can be polarized into different phenotypes depending on the composition of the environment into which they migrate. They express an abundance of receptors in order to absorb the LDL present, whereby the macrophages are converted into foam cells. The release of growth factors and cytokines induces smooth muscle cells to migrate from the media into the intima, where they then form extracellular matrix components, thereby contributing to the formation of the fibrous cap [14]. Many of the foam cells perish in the process themselves and are broken down by other macrophages. Due to the number of these degradation processes, the macrophage endoplasmic reticulum is overburdened leading to further macrophage death. As a result of this now uncontrolled decay, all intracellular components of macrophages, such as lipids, inflammatory cytokines, and metalloproteinases (MMP), are released.

This reaction, which has now become independent, leads to the destruction of matrix components and, thus, to plaque destabilization and ultimately to plaque rupture. This process is additionally driven by smooth muscle cell loss and ischemia-induced angiogenesis [31].

Another process that is already underway in the initial phase of atheroma development takes place simultaneously. As an initial reaction to endothelial stress, intimal thickening occurs with the migration of phenotypically altered vascular smooth muscle cells (VSMC) [32]. These cells produce a milieu rich in proteoglycans and extracellular matrix in their immediate environment, where lipoproteins can accumulate and undergo oxidation to a greater extent. These VSMCs possess an abundance of receptors, such as the low-density lipoprotein receptor-related protein 1 (LRP1) or the lectin-type oxidized LDL receptor 1 (LOX-1), which make increased endocytosis of oxidized lipoproteins possible. At the same time these smooth muscle cells lose transport systems in order to be able to release the enriched lipoprotein once again. These transporters include, e.g., the ATP-binding cassette transporter A1 (ABCA1) and apolipoprotein A1 (ApoA1). As a result of this imbalance, the lipoprotein content in VSMC increases. Thus, the smooth muscle cells are transformed into foam cells and undergo apoptosis. This foam cell decay causes an inflammatory reaction that attracts circulating monocytes via the MCP-1 receptor and further perpetuates inflammation through proinflammatory cytokines such as IL-1α and IL-1β. Thus, a vicious cycle is set in motion that ultimately results in atheromatous changes in the vessel wall [13].

The onset of atherosclerosis is an active process

From this it can be deduced that the risk of fibrous cap rupture has less to do with the degree of vascular stenosis than it does with plaque formation [2]. A thin cap measuring less than 65 μm, a necrotic core occupying more than 30% of the total plaque, hemorrhage, infiltration of inflammatory cells such as lymphocytes and macrophages, as well as the reduction in smooth muscle cells are unfavorable factors that result in rupture and subsequent thrombosis and occlusion of the vessel by destabilizing the plaque and fibrous cap [30]. The structure of the necrotic core is particularly important in this process.

The loss of stabilizing matrix components as well as the build-up of lipids and inflammatory cells causes the core to become mechanically unstable. The apoptotic death of smooth muscle cells as well as macrophages increases this instability and, due to fibrin production, promotes thrombus formation via the release of tissue factor (TF) in the case of rupture [28].

The alteration of the phenotype of smooth muscle cells and macrophages or their polarization also leads to the development of osteoclast-like cells. These cells, in combination with a change in Ca++ and phosphate levels and a decrease in inhibitory cytokines, induce osteogenesis. Calcium-binding protein, osteopontin, and fetuin A are among the messengers that could prevent calcification [21]. Finally, hydroxyapatite crystals are deposited in the tissue. It is important here whether the crystals are deposited in the intima or in the media. Intimal sclerosis is closely linked to atheromatous plaque formation and is assumed to protect the intima from pathological processes in the media, which, due to progressive luminal narrowing, further exacerbates stenosis [21]. Furthermore, calcium deposits in the fibrous cap increase the risk of plaque rupture. This contrasts with medial calcification, which essentially takes place in close relation to smooth muscle cells.

This causes calcium salt deposits, which are subject to constant transformation until bone structures have formed. These deposits contribute more to the reduced elasticity and compliance of the vascular wall itself than they do to the progression of stenosis.
Plaque growth causes the oxygen diffusion distance to increase significantly. Starting at a vessel wall thickness of approximately 100 μm, hypoxia-induced factor (HIF-1α) expression is enhanced due to the lack of oxygen, leading to an increase in growth factors such as vascular endothelial growth factor (VEGF). This promotes angiogenesis to sprout new vessels from pre-existing vessels from the adventitia into the intima. The further the process progresses, the more chaotic growth becomes, leading to immature and unstable vessels that continue to enlarge and destabilize the nucleus through hemorrhage [20].

Other factors that influence atherosclerotic plaques include stress and shearing forces acting on the vessels. Atherosclerotic changes occur primarily in vascular branches and sections where blood flow is not linear. Low shear stress in particular results in an increased tendency towards inflammation in endothelial cells and leukocytes [22]. Differences in the distribution of inflammatory cells and wall thickening result in characteristic predilection sites for rupture of the various plaques. Plaque rupture caused by physical exertion is more likely to occur at the middle of the atheroma, whereas rupture occurring at rest is more likely to be found in the shoulder region of the plaque. This is explained by the fact that a particularly high number of inflammatory cells settle there, whereas central hardening caused by calcification leads to reduced vessel and plaque elasticity and hence to tearing under stress in particular [3].

If the endothelium breaks down or a plaque layer tears, a chain reaction occurs that culminates in platelet-rich thrombus formation, which further narrows or shifts the vascular lumen. Interaction between a platelet surface glycoprotein (GPIba) and the von Willebrand factor (vWF) A1 domain occurs in a first step. There are other binding possibilities to leukocytes, macrophages, and P-selectin. In a second step, this hitherto reversible adhesion is transformed by the interaction between two platelet collagen receptors (GPIa/IIa and GPVI) into stable thrombus adhesion. This leads, among other things, to the activation of

corresponding receptor. This leads, among other things, to the activation ofstable thrombus adhesion. This leads, among other things, to the activation of
phospholipase C, the mobilization of calcium, as well as the activation of integrins, laminin, and fibronectin, resulting in further platelet activation [10].

**Angiogenesis and arteriogenesis: the basics of collateral formation**

The task of blood vessels is to transport oxygen and nutrients to the organs and individual cells, as well as to remove waste products. It is not surprising, therefore, that there is more than one way to ensure perfusion both in healthy and diseased tissue. Vasculogenesis is a process by which vessels with a wholly adequate three-layered wall structure are created without the need for pre-existing vessels. This process is the primary process in the development and growth of new life and plays only a minor role in the adult individual; however, the recruitment of endothelial progenitor cells (EPC) that produce numerous growth factors can result in a milieu of vasculogenesis and angiogenesis in previously avascular tissue [11]. Angiogenesis and arteriogenesis are paramount in wound healing and especially in the chronic hypoperfusion discussed here.

Angiogenesis creates new capillary vessels from existing capillaries, with wound healing or the undersupply of oxygen to tissue representing the driving forces here. To this end, a variety of factors such as fibroblast growth factor (FGF), VEGF, and HIF are released, with HIF stimulating the transcription of VEGF [8]. This causes the proliferation and lateral migration of endothelial cells. The basement membrane and adjacent tissue are eroded by proteases such as metalloproteinases, and the pericytes surrounding the endothelium become detached. The VEGF renders endothelial cells more permeable to plasma proteins, thereby enabling the formation of a rudimentary extracellular matrix in which endothelial cells can in turn be anchored. A pseudopodium extension develops, enabling endothelial cell growth into the surrounding connective tissue. This extension is guided by signal molecules such as ephrin and semaphorin in order to maintain a particular direction [19].

The capillary shoot is the starting point for the accumulation of further cells and, after continuous growth, ultimately forms a lumen. In order to stabilize the new vessel, the endothelial cells return to a resting state, and pericytes, via signal molecules such as platelet derived growth factor B (PDGF-B), transforming growth factor-β (TGF-β), and Notch (transmembrane proteins involved in cell-to-cell communication), adhere to and surround the new vessel. Due to the activation of protease inhibitors such as tissue inhibitors of metalloproteinases (TIMP) and plasminogen activator inhibitor-1 (PAI-1), a basement membrane and new cell contacts are formed [4]; however, angiogenesis is not sufficient as a means of capillary formation to replace a stenosed or occluded artery. This is due to the low flow resistance in arteries that capillaries are unable to compensate with their much higher resistance. A further reason is the long distance between stenosis and hypoperfused tissue, hence the greater relevance attached to arteriogenesis [26].

»**Hypoxia is a key stimulus of angiogenesis**

Arteriogenesis refers to the expansion of existing collateral vessels [24], which are predominantly arched, bypass arterioles within the boundary of various supply regions with low flow and play only a minor role in organ perfusion under healthy conditions. By a process of remodelling, these vessels change their diameter, thereby forming a type of biological bypass that is then extended. The driving forces here include arterial blood pressure, the pressure gradient above and below the stenosis, and the shear stress acting on the vessel walls [25]. The arterioles are expanded by the pressure gradient in a first step, resulting in overexpansion and leakage of plasma proteins. Endothelial cells then enter the cell cycle and smooth muscle cells proliferate and express metalloproteinases (MMPs), which break down the basement membranes and collagen fibers and macerate the tissue. Lymphocytes also enter the tissue and, by killing off surrounding cells, create space for the pending growth of arterioles. Ultimately, the cell mass and total diameter of the individual collaterals can be up to 50 and 20 times, respectively, greater than those of the original arterioles [23]. As a result of endothelium activation, monocytes are attracted by cytokines, such as monocytes of chemoattractant protein 1 (MCP-1), which produce growth factors and proteolytic enzymes that additionally enable smooth muscle cells to migrate and divide. Other factors such as granulocyte macrophage colony-stimulating factor (GM-CSF) prolong the lifespan of monocytes and represent important arteriogenic factors. Since the vessels increase not only in diameter but also in length, yet are located in a restricted environment, the arterioles are subject to substantial distortion, ultimately resulting in the characteristic appearance of collaterals on angiography. All arterioles participate in early arteriogenesis; however, depending on flow, main collaterals quickly emerge that are subjected to maximum stimulation and can then undergo the complete arteriogenic remodelling process. In the case of long-term occlusion, these vessels are able take over a significant proportion of the perfusion of main vessels [26].

**Critical ischemia: the final stage of atherosclerosis**

In the case of progressive stenosis, the compensatory capacity of arteriogenesis can be compromised, and manifest tissue hypoxia occurs. Physical activity is restricted due to ischemic pain. This results in reduced stimulation of arteriogenesis and degeneration of the newly formed vascular structures as well as to progressive muscle cell loss, thereby further reducing exercise capacity and setting a downward spiral in motion [5]. Chronic inflammation severely impairs endothelial function. As part of this, free radicals (ROS) attack the vessels’ capacity to relax by intercepting the NO produced by the endothelium for vascular relaxation. Reduced muscle activity leads to lower shear stress in the collaterals, as a result of which the endothelium produces even less NO and superoxide.
If excessive demands are put on the vessels and the tissue to be perfused and, in the further course, to atrophy of the vessels and the tissue to be perfused [26].

**Conclusion**

- Chronic limb ischemia is a multifactorial phenomenon.
- The onset of atherosclerosis is an active process in which foam cell formation and cell apoptosis play a crucial role.
- Hypoxia is a key stimulus of angiogenesis, whereas shear stress induces arteriogenesis.
- By means of vessel expansion, arteriogenesis is able to take over a significant proportion of the perfusion of major vessels.
- If excessive demands are put on these compensatory mechanisms, tissue atrophy and scar remodelling processes occur.

**References**

1. Badimon L (2012) Interleukin-18: a potent pro-inflammatory cytokine in atherosclerosis. Cardiovasc Res 96:172–175 (discussion 6–80)
2. Badimon L, Vilahur G (2014) Thrombosis formation on atherosclerotic lesions and plaque rupture. J Intern Med 276:618–632
3. Burke AP, Farb A, Malcom GT, Liang Y, Smialek JE, Virmani R (1999) Plaque rupture and sudden death related to exertion in men with coronary artery disease. JAMA 281:921–926
4. Carmeliet P, Jain RK (2011) Molecular mechanisms of angiogenesis. Nature 473:298–307
5. Coats P, Jarajapu YPR, Hillier C, McGrath JC, Daly C (2003) The use of fluorescent nuclear dyes and laser scanning confocal microscopy to study the cellular aspects of arterial remodelling in critical limb ischaemia. Exp Physiol 88:547–554
6. Coats P, Wadsworth R (2005) Marriage of resistance and conduit arteries breeds critical limb ischemia. Am J Physiol Heart Circ Physiol 288:H1044–H1050
7. Darius H, Trampisch H, Pittrow D, Allenberg JR, Haberl RL, Mahn M, Tepohl HG, Burhaus I, Diehm C (2008) Vergleich zweier Koronaruquivalente: Risikoorhohung unter Diabetes mellitus und Peripherer Arterieller Verschlusskrankheit. Dtsch Med Wochenschr 133(45):2317–2322
8. Draganeva G, Kopsila P, Vla-Hertuala S (2013) Promoting blood vessel growth in ischemic diseases: challenges in translating preclinical potential into clinical success. Dis Model Mech 6:312–322
9. Fowkes FG, Rudan D, Rudan I, Aboyans V, Denenberg J, McDermott M, Norman P, Sampson U, Williams L, Mensah G, Criqui M (2013) Comparison of global estimates of prevalence and risk factors for peripheral artery disease in 2000 and 2010: a systematic review and analysis. Lancet 382:1329–1340
10. Freynhofer MK, Bruno V, Wojta J, Huber K (2012) The role of platelets in athero-thrombotic events. Curr Pharm Des 18:5197–5214
11. Fujita Y, Kawamoto A (2017) Stem cell-based peripheral vascular regeneration. Adv Drug Deliv Rev 17:30182–30185
12. Heller G, Günster C, Schellsmidt H (2004) Wie häufig sind Diabetes-bedingte Amputationen unterer Extremitäten in Deutschland? Eine Analyse auf Basis von Routinedaten. Dtsch Med Wochenschr 129(9):429–433
13. Johnson JL (2014) Emerging regulators of vascular smooth muscle cell function in the development and progression of atherosclerosis. Cardiovasc Res 103:425–460
14. Koga J, Alkawa M (2013) Crosstalk between macrophages and smooth muscle cells in atherosclerotic vascular diseases. Vascul Pharmacol 57:24–28
15. Kroger K, Stang A, Konratiave J, Moebus S, Beck E, Schmermund A, Möhlenkamp S, Draganos N, Siegrist J, Joekel KH, Erbel R (2006) Prevalence of peripheral arterial disease – results of the Heinz Nixdorf Recall Study. Eur J Epidemiol 21:279–285
16. Malysar N, Furstenberg T, Wellmann J, Meyborg M, Lüders F, Gebauer K, Bunzeimeier H, Roeder N, Reinecke H (2013) Recent trends in morbidity and in-hospital outcomes of in-patients with peripheral arterial disease: a nationwide population-based analysis. Eur Heart J 34:2706–2714
17. Mehta D, Malik AB (2006) Signaling mechanisms regulating endothelial permeability. Physiol Rev 86:279–367
18. Meijer WT, Grobbée DE, Hunink MG, Hofman A, Hoes AW (2000) Determinants of peripheral arterial disease in the elderly: The Rotterdam Study. Arch Intern Med 160(19):2934–2938
19. Michaels UR (2014) Mechanisms of endothelial cell migration. Cell Mol Life Sci 71:4131–4148
20. Moreno PR, Purushothaman M, Purushothaman KR (2012) Plaque neovascularization: defense mechanisms, betrayal, or a war in progress. Ann NY Acad Sci 1254:7–17
21. Sage A, Tintut Y, Demer L (2010) Regulatory mechanisms in vascular calcification. Nat Rev Cardiol 7:528–536
22. Seneviratne A, Hulsmans M, Holvoet P, Monaco C (2013) Biomechanical factors and macrophages in plaque stability. Cardiovasc Res 99:284–293
23. Schaper W, DeBrabander M, Lewi P (1971) DNA-synthesis and mitoses in coronary collateral vessels of the dog. Circ Res 28:671–679
24. Schaper W (2009) Collateral circulation: past and present. Basic Res Cardiol 104:5–21
25. Scholz D, Cai WJ, Schaper W (2001) Arteriogenesis, a new concept of vascular adaptation in occlusive disease. Angiogenesis 4(4):247–257
26. Tang GL, Chang DS, Sarkar R, Wang R, Messina LM (2005) The effect of gradual or acute arterial occlusion on skeletal muscle blood flow, arteriogenesis, and inflammation in rat hindlimb ischemia. J Vasc Surg 41:312–320
28. Tedgui A, Mallat Z (2001) Apoptosis as a determinant of atherothrombosis. Thromb Haemost 86:420–426

29. Virginio VW, Nunes VS, Moura FA, Menezes FH, Andreollo NA, Rogerio F, Scherrer DZ, Quintao EC, Nakandakare E, Petrucci O (2013) Arterial tissue and plasma concentration of enzymatic-driven oxysterols are associated with severe peripheral atherosclerotic disease and systemic inflammatory activity. Free Radic Res 49:199–203

30. Virmani R, Kolodgie FD, Burke AP, Farb A, Schwartz SM (2000) Lessons from sudden coronary death: a comprehensive morphological classification scheme for atherosclerotic lesions. Arterioscler Thromb Vasc Biol 20:1262–1275

31. Virmani R, Kolodgie FD, Burke AP, Finn AV, Gold HK, Tulenko TN, Wrenn SP, Narula J (2005) Atherosclerotic plaque progression and vulnerability to rupture: angiogenesis as a source of intraplaque hemorrhage. Arterioscler Thromb Vasc Biol 25:2054–2061

32. Zhang MJ, Zhou Y, Chen L, Wang X, Pi Y, Long CY, Sun MJ, Chen X, Gao CY, Li JC, Zhang LL (2016) Impaired SIRT1 promotes the migration of vascular smooth muscle cell-derived foam cells. Histochem Cell Biol 146:33–43