Integrative analysis of AGR2 based on data mining reveals new insights in breast cancer

Xin Zhou
Department of breast surgery, Zibo Maternal and Child Health Hospital, Zibo, China

Fangyuan Zhang
Department of breast surgery, Zibo Maternal and Child Health Hospital, Zibo, China

Yan Liu
Department of breast surgery, Zibo Maternal and Child Health Hospital, Zibo, China

Dongxin Wei (✉ deschou1986@163.com)
Zibo Maternal and Child Health Hospital  https://orcid.org/0000-0001-7672-8952

Research

Keywords: AGR2, breast cancer, DNA methylation, copy number variation, ribosome biogenesis, prognosis

DOI: https://doi.org/10.21203/rs.3.rs-52934/v1

License: © This work is licensed under a Creative Commons Attribution 4.0 International License.
Read Full License
Abstract

**Background:** Belonging to the protein disulfide isomerase (PDI) family, anterior gradient 2 (AGR2) is overexpressed in mucus-secreting cells and endocrine organs such as breast, lung, and ovarian, but its exact function and regulation remain unclear. We aimed to perform integrative analysis of AGR2 in breast cancer.

**Methods:** In our study, a serious of bioinformatic online databases were used to analyze the expression, regulation, prognostic value, and function of AGR2 in breast cancer.

**Results:** The results suggested that AGR2 mRNA was overexpressed in non-basal-like or non-triple-negative breast cancer, but underexpressed in basal-like/ triple-negative breast cancer. AGR2 mRNA expression was correlated with its protein expression. Moreover, the expression of AGR2 was lower in more malignant breast cancer. Furthermore, we also found that DNA methylation, rather than copy number variation, led to AGR2 mRNA overexpression. Prognosis analysis showed no significant correlation between AGR2 level and survival, but in subtype investigation, patients with higher AGR2 level had a significantly better outcome in patients with basal-like / triple-negative breast cancer. Enrichment analysis for co-expression genes of AGR2 revealed that gene sets enriched for chromosome segregation, DNA conformation change, chromosomal region, and GABA receptor activity may play important roles in breast cancer, and that the most significant pathway was “ribosome biogenesis in eukaryotes”, in which negative co-expression genes of AGR2 were enriched.

**Conclusions:** Overexpression of AGR2 was found in less malignant breast cancer, for the first time, our results propose that DNA methylation rather than copy number variation leads to AGR2 overexpression, which can predict favorable prognosis in basal-like/ triple-negative breast cancer, and AGR2 could be a possible regulator of ribosome biogenesis in patients with breast cancer.

1. **Background**

As the most common cancer among women in the world, breast cancer (BRCA) has caused a serious burden to China and the rest of the world. The key measures to treat patients with BRCA include surgery, chemotherapy, hormone, targeted therapy, and radiotherapy. Due to its heterogeneous, medication options are varying according to different subtypes, which include Luminal A, Luminal B, Her-2 and Basal-like (or triple-negative breast cancer, TNBC) subtypes. About 30% of BRCA patients are stricken with metastasis and relapse, although treated at an early stage\(^1\). Therefore, it's crucial to explore biomarkers for prognosis judgment and treatment of BRCA.

AGR2, as a member of the protein disulfide isomerase (PDI) family which is responsible for the catalysis of disulphide bond formation and the maintenance of cellular homeostasis, may play a role in facilitating the spreading of metastatic tumor cells, and promoting cell survival and proliferation\(^2\). Studies demonstrated that AGR2 was a clinically relevant factor to modulate the behavior and response of hormone-dependent cancers (such as prostate, breast cancer) and hormone-independent cancers (such
as pancreatic, colorectal cancer)\textsuperscript{3}. It has been widely reported that AGR2 is associated with the development of estrogen receptor (ER) positive breast cancer\textsuperscript{4}. However, the actual functions and regulation of AGR2 are poorly comprehended, and the prognostic value of AGR2 remains inconsistent. With the development of bioinformation technology, diverse public databases and analytical strategies, data mining techniques has been widely accepted in medical researches. To develop a better understanding about the role of AGR2 in BRCA, we performed integrative analysis based on data mining by using public web tools to explore the expression, regulation, function, and prognostic value of AGR2 in BRCA.

2. Materials And Methods

2.1 AGR2 Expression Analysis

The expression of AGR2 mRNA was examined with two web-based databases, including GEPIA2 and Breast Cancer Gene Expression Miner v4.4 (bc-GenExMiner v4.4). First, we evaluated the AGR2 mRNA expression in pan-cancer and breast cancer by using GEPIA2, which is a web tool for Wide-ranged expression profiling and interactive analysis with data from The Cancer Genome Atlas (TCGA) database and the Genotype-Tissue Expression (GTEx) database (http://gepia.cancer-pku.cn/)\textsuperscript{5}. We also explored the AGR2 expression according to different clinicopathological characteristics of breast cancer, including hormonal receptors status, Her-2, age, nodal status, histological subtypes, PAM50 subtypes, Nottingham prognostic index (NPI), and Scarff-Bloom-Richardson(SBR) grade\textsuperscript{6,\textsuperscript{7}}, by using bc-GenExMiner v4.4, which is a user-friendly online mining platform with published annotated data of breast cancer (http://bcgenex.centregauducheau.fr)\textsuperscript{7}. The expression of AGR2 protein was studied with The Human Protein Atlas (HPA) database and UALCAN database. IHC results of AGR2 in breast cancer and normal breast tissue were obtained from HPA, which is an online program containing human transcriptome and proteome data generated from RNAsequencing analysis and immunohistochemistry analysis\textsuperscript{8}. We explored AGR2 protein expression in normal breast tissues and malignant counterparts by using UALCAN, which is a web portal for performing analyses of gene expression data from TCGA (http://ualcan.path.uab.edu)\textsuperscript{9}. Correlation between mRNA and protein expression of AGR2 was determined by using The cBioPortal database, which is a portal for integrative analysis of cancer genomics and clinical profiles (http://cbioportal.org)\textsuperscript{10}. P $<$ 0.05 was considered statistically significant.

2.2 AGR2 Gene Alteration Analysis

DNA copy number of AGR2 in breast cancer was analyzed within the Oncomine 4.5, which is an outstanding database for integrated data-mining with cancer microarray data (www.oncomine.org)\textsuperscript{11}. The cBioPortal database was used to analyze AGR2 gene alteration with data from the Breast Invasive Carcinoma (TCGA, Firehose Legacy) samples. An overview of genetic alterations per sample in AGR2 is displayed in the tab OncoPrint. In the tab Survival, the Kaplan-Meier analysis was used to evaluate the effect of AGR2 alterations on the patient’s overall survival (OS) and disease-free survival (DFS). The tab Comparison shows correlation between AGR2 mRNA expression and CNV, DNA methylation or protein
expression. We also analyzed correlation between AGR2 CNV and DNA methylation status. P < 0.05 was considered statistically significant.

2.3 AGR2 DNA Methylation Analysis

This part was performed by using three databases, including UALCAN, Wanderer and Methsurv. We first explored AGR2 DNA methylation status in normal breast tissues and malignant counterparts by using UALCAN, then obtained data of five probes from Methsurv (https://biit.cs.ut.ee/methsurv), which is an online database for multivariable prognostic analysis using DNA methylation data\textsuperscript{12}. We investigated AGR2 DNA methylation status in BRCA and normal tissues and relevance between AGR2 DNA methylation status and mRNA expression at all the five probes respectively by using Wanderer database, which is an interactive online tool to investigate data of DNA methylation and gene expression in human cancer from TCGA (http://maplab.cat/wanderer)\textsuperscript{13}. Within Methsurv database, We explored prognosis values of AGR2 DNA methylation in BRCA. P ≤ 0.05 was considered statistically significant.

2.4 Survival Analysis of AGR2 using GEPIA2, TCGAportal and bc-GenExMiner v4.4

Within GEPIA2 database, we performed survival analysis of AGR2 in breast cancer including OS and DFS. Then the prognostic value of AGR2 was evaluated in patients with different subtypes of breast cancer by using TCGAportal, which is a newly released web tool that allows for analysis of survival curve with data from TCGA (http://www.tcgaportal.org)\textsuperscript{14}. Afterword, we verified the results in bc-GenExMiner v4.4. P ≤ 0.05 was considered statistically significant.

2.5 Co-expression Genes of AGR2 and Functional Enrichment Analysis

In this part, we performed analysis by using LinkedOmics database, which is an online database that integrates data from the Clinical Proteomic Tumor Analysis Consortium (CPTAC) and contains multi-omics data and clinical data from TCGA (http://www.linkedomics.org/login.php)\textsuperscript{15}. The module of LinkFinder was used to analyze associated genes of AGR2 in the TCGA BRCA cohort (n = 1093). The module of Link-Interpreter module was used to conduct functional analysis of associated genes of AGR2 in the Web-based Gene SeT AnaLysis Toolkit (WebGestalt)\textsuperscript{16}. GSEA was employed to perform Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses. GO functional analysis contained three categories, including biological process (BP), cellular component (CC) and molecular function (MF). P ≤ 0.05 and FDR < 0.05 were considered statistically significant.

2.6 Protein-Protein Interaction (PPI) Network

PPI network for genes enriched in the pathway of ribosome biogenesis in eukaryotes was established by using GeneMANIA database, which is a web interface for fast gene network construction and function prediction (http://www.genemania.org)\textsuperscript{17}. The result contained the source of the edge of the network and
a number of bioinformatics methods, including physical interaction, gene co-location, gene co-expression, gene enrichment analysis and website prediction.

3. Results

3.1 AGR2 expression in BRCA

3.1.1 mRNA expression of AGR2 in BRCA

Initially, we evaluated the mRNA expression of AGR2 in pan-cancer by using the GEPIA2 dataset. It indicated that AGR2 mRNA expression was higher in several cancers than matched normal tissues, such as breast cancer (BRCA), cholangiocarcinoma (Chol), lung adenocarcinoma (LUAD) and so on (Fig. 1A). AGR2 mRNA was significantly overexpressed in BRCA compared to normal tissues (p < 0.01) (Fig. 1B). In subtype analysis, the lowest level of AGR2 mRNA was found in base-like subtype (p < 0.01) (Fig. 1C), in which AGR2 lever was even lower than normal tissue (p < 0.01) (Fig. 1D). Within Breast Cancer Gene-Expression Miner v4.4 (bc GenExMiner v4.4) database, we examined expression profiles of AGR2 using 4712 patients with BRCA based on different clinical-pathological indicators. AGR2 mRNA expression was significantly upregulated in ER+, PR+, and HER-2 + subtypes than corresponding negative groups, respectively (p < 0.0001) (Fig. 2A-C) and downregulated in TNBC subtype than non-TNBC subtypes (p < 0.0001) (Fig. 2D). We also found significant differences between the ≤ 51- and > 51-year group, between lymphnode positive and negative group (p < 0.0001) (Fig. 2E-F). Mucinous group showed higher AGR2 expression level than invasive groups (p < 0.0001) (Fig. 2G). According to SBR grade and NPI, AGR2 level was the lowest in SBR3 group(Fig. 2H) and was lower in NPI3 group than NPI1 group (Fig. 2I).

3.1.2 Protein Expression of AGR2 in BRCA

The UALCAN database was used to assess the different levels of AGR2 in BRCA and normal breast tissues, and showed that AGR2 protein level was higher in BRCA (P = 8.59E-11, P) (Fig. 3C). To validate the IHC results, the HPA database was used to obtain AGR2 expression in BRCA and normal tissue, and showed that AGR2 level (antibody HPA007912) was upregulated in BRCA compared to normal tissue (Fig. 3A-B). Furthermore, correlation was found between AGR2 protein expression (mass spectrometry by CPTAC) and mRNA level (microarray) in cBioPortal database (Spearman: 0.81, P = 3.43E-18; Pearson: 0.79, P = 4.64E-17) (Fig. 3D).

3.2 Gene alteration of AGR2 in BRCA

The Oncomine database revealed that DNA copy number of AGR2 was significantly higher in invasive BRCA than corresponding normal tissues (P = 0.027) (Fig. 4C). Afterword, we used the cBioPortal database to determine the type and frequency of AGR2 alteration in BRCA patients based on data from Breast Invasive Carcinoma (TCGA, Firehose Legacy), then performed correlation analysis between AGR2 copy number variation (CNV) and mRNA expression, and prognostic analysis of AGR2 CNV. AGR2 was
altered in 76 of 1093 (7%) BRCA patients, and these alterations were mRNA amplification in 13 cases (1.19%), mRNA low in 61 cases (5.58%), and multiple alterations in 2 cases (0.18%) (Fig. 4A-B). Hence, the most common type of AGR2 CNV is amplification in BRCA. Correlation analysis showed that AGR2 mRNA expression was not associated with CNV (Spearman: 0.08, p = 9.166e-3; Pearson 0.07, p = 0.0313) (Fig. 4D). Survival analysis suggested that AGR2 CNV was not associated with overall survive (OS) (p = 0.776) or disease free survive (DFS) (p = 0.0974) in BRCA (Fig. 4E-F).

3.3 DNA methylation of AGR2 in BRCA

In this part, we first compared different status of AGR2 DNA methylation between BRCA and normal breast tissues by using UALCAN. It showed that AGR2 DNA methylation status was significantly lower in BRCA than normal breast tissues (P < 1E-12) (Fig. 5A). Then in cBioPortal database, we found that AGR2 mRNA expression (mircoarray) was negatively associated with its DNA methylation status (HM450) (Spearman: -0.76, P = 4.21E-42; Pearson: -0.65, P = 1.15E-27) (Fig. 5B). We also analyzed correlation between AGR2 DNA methylation status and CNV, but the result was negative (Fig. 5C). Furthermore, details of five probes were available by using MethSurv database (Supplementary Table 1). The methylation status of AGR2 at the five probes were all lower in BRCA than corresponding normal tissue according to Wanderer database (Fig. 5D). Survival analysis showed significant result only at probe cg25343204 (p = 0.036) by using Methsurv database, lower AGR2 DNA methylation status indicating better prognosis (p = 0.036) (Fig. 6F). Interestingly, among all the five probes, AGR2 DNA methylation status had the strongest negative correlation to its mRNA expression at probe cg25343204 (Pearson r=-0.646) (Fig. 6A).

3.4 Prognosis analysis of AGR2 in BRCA

We investigated prognostic value for AGR2 in BRCA by using GEPIA2, TCGAPotal and bc GenExMiner v4.4. According to GEPIA2, no correlation was found between AGR2 mRNA and OS (p > 0.05) or DFS (p > 0.05) (Fig. 7A-B). According to TCGAPotal, subtype analysis was performed and AGR2 showed no significant correlation to survival in Luminal A, Luminal B or Her-2 subtype respectively, but in basal-like subgroup, increased AGR2 was associated with better prognosis (p = 0.00061) (Fig. 7C-F). Furthermore, we verified it in bc GenExMiner v4.4 database, it also showed significant result only in basal-like subgroup (p = 0.0130), higher AGR2 expression indicating better prognosis (Fig. 7G-J).

3.5 Co-expression Genes of AGR2 and enrichment analyses in BRCA

In the LinkedOmics database, we used the Function module to analyze mRNA sequencing data 1093 BRCA patients from TCGA. Volcano plots showed the significant positive (red dots) and negative (green dots) co-expression genes of AGR2 (false discovery rate [FDR] < 0.05) (Fig. 8A). Heat maps showed the top 50 positive and the top 50 negative associated genes respectively (Fig. 8B-C). Then gene set enrichment analysis (GSEA) was used to perform GO and KEGG pathway analyses in the LinkInterpreter
The Web-based Gene SeT AnaLysis Toolkit (WebGestalt) was applied. The significant results of GO term analysis (p < 0.05 and FDR < 0.05) were shown in Supplementary Fig. 1A-C. According to volcano plots shown in Fig. 8D-F, we speculated that gene sets enriched for chromosome segregation (BP), DNA conformation change (BP), chromosomal region (CC), and GABA receptor activity (MF) may play important roles in BRCA. Details of these gene sets are shown in Supplementary Table 2–5. The significant result of KEGG pathway analysis (p < 0.05 and FDR < 0.05) was shown in Supplementary Fig. 1D, and the volcano plot shown in Fig. 8G suggested that the pathway of ribosome biogenesis in eukaryotes (Supplementary Fig. 2) may play an important role in BRCA. Details of genes enriched in this pathway are shown in Supplementary Table 6. Finally, the protein-protein interaction network (PPI) containing genes enriched in ribosome biogenesis was constructed by GeneMANIA. It showed that these genes were mainly responsible for ribosome biogenesis, small nucleolar ribonucleoprotein and ribonucleoprotein complex biogenesis (Fig. 9).

4. Discussion

In the study, we firstly evaluated AGR2 expression in BRCA. Generally, AGR2 upregulates in non-basal-like BRCA compared to normal tissues, which is consistent with previous studies18,19. We also found that AGR2 level was the lowest in basal-like BRCA and was even lower than in normal tissues. Consistently, Jilong Guo et al. performed immunohistochemistry and revealed that AGR2 was significantly upregulated in non-TNBC tissues than TNBC tissues, and was also significantly downregulated in TNBC than normal tissues20. Basal-like breast cancer, also known as triple-negative breast cancer (TNBC), lacks estrogen receptor (ER) and progesterone receptor (PR) expression as well as human epidermal growth factor receptor 2 (HER2) amplification. Compared with other subtypes, the TNBC subtype has poorer prognosis, higher mortality and the highest probability of recurrence after treatment. Its poor outcome is due to the lack of available targeted therapies, and its intrinsically aggressive nature21. We also found that AGR2 expression was higher in mucinous breast cancer (MBC) than invasive ductal cancer (IDC) and invasive lobular cancer (ILC). As a rare histological cancer type, MBC exhibits lower malignancy and a comparatively better prognosis. A study showed that patients with MBC had a significantly better outcome than patients with IDC22. Plus, lower AGR2 level was found in SBR3 group and NPI3 group. It appears that AGR2 expressions are lower in more invasive tumors, though Wang Z suggested that increased AGR2 contributes to tumor growth and invasion and metastasis formation23.

To understand the reasons behind AGR2 overexpression in BRCA, we explored the relationship between AGR2 mRNA expression and CNV or DNA methylation. Because of changes in gene sequence location, CNV can affect gene expression, increase disease susceptibility and aggravate disease progression24. Changes in DNA methylation have an impact on regulating gene expression during cancer progression and metastasis25. Hence, we planned to explore what may be the potential reason behind changes of AGR2 mRNA expression in BRCA at the DNA level. Our study found that AGR2 copy number was significantly increased in BRCA, and the main type of AGR2 alteration was amplification. However, AGR2 CNV was not associated with mRNA expression or survive. But we found AGR2 mRNA expression was
negatively associated with DNA methylation status, lower methylation status indicating higher mRNA level. HYE YOUNG SUNG et al analyzed AGR2 mRNA expression and DNA methylation data in metastasized ovarian tumor in the mice and showed mRNA overexpression and hypomethylation at CpG sites in its promoter region. Furthermore, they revealed that treatment with 5-aza-2′-deoxycytidine, a kind of the DNA methyltransferase inhibitor, could reduce the level of CpG methylation in the AGR2 promoter and increase the level of AGR2 expression. We carried out further exploration and found AGR2 methylation status were lower in BRCA than normal tissues, and it showed strong negative correlation between AGR2 methylation and mRNA expression at probe cg25343204, where also showed significant association with survival, lower methylation status showing better outcome. Thus, for the first time, we propose that upregulation of AGR2 mRNA in BRCA may be caused by DNA methylation rather than CNV.

The prognostic value of AGR2 remains largely inconclusive. A study demonstrated that higher level of AGR2 was related to less aggressive tumors and better prognosis in epithelial ovarian carcinoma, which is in line with Ames and colleagues who found absence of AGR2 was associated with increased malignancy and tumor progression, and relapse in endometrioid and mucinous subtypes. AGR2 overexpression was found to contributed to the poor OS in lung cancer, whereas another study showed that AGR2 overexpression did not show any prognostic value in non-small-cell lung cancer. Fritzsch FR et.al found that for patients with AGR2-expressing breast carcinomas, significantly longer OS became apparent than those without AGR2-expression. However, other reports suggested that high AGR2 expression was related to poor outcome in BRCA. In our study, no significant prognostic values of AGR2 were found according to GEPIA2 database. Interestingly, in subtype analysis, for patients with basal-like BRCA, higher AGR2 level showed better outcome according to TCGAPortal and bc-GenExMiner v4.4 databases. Combining data from our expression analysis of AGR2, it seems like that AGR2 plays a protective role in BRCA due to its higher level in less aggressive tumors such as Luminal subtype and MBC BRCA.

Many findings are in favor of the hypothesis that AGR2 is an oncogene and can promote tumor cell survival, migration, and invasion; drug resistance; angiogenesis; and metastasis. However, Shao-bo Tian and colleagues performed a systematic review and meta-analysis about the prognostic value of AGR2 expression in solid tumors, and speculated that AGR2 might act as a tumor suppressor in some tumor types, which needed further verifications. Hence, we performed functional analysis of AGR2 for better understanding. GO and KEGG pathway enrichment analyses of AGR2 associated genes in BRCA were performed by using GSEA. Go term analysis indicated that these genes were mainly enriched in chromosome segregation (BP), DNA conformation change (BP), chromosomal region (CC), and GABA receptor activity (MF). KEGG pathway analysis demonstrated that “ribosome biogenesis in eukaryotes” was the most significant signaling pathway, and genes enriched in this pathway were negatively associated with AGR2. Ribosomes play important roles in translating information contained in mRNAs into functional proteins, the final step in the genetic programme. Ribosome biogenesis can increase the size of nucleolar organizing regions (NORs) which have long been used as a marker of tumor cell proliferation that negatively correlates with patient survival. Richard Y. et al. indicated that the
increasing of ribosomal content in epithelial cell fates may enhance their metastatic potential in breast cancer\textsuperscript{40}. Studies also demonstrated that drugs inhibiting ribosome biogenesis might provide a feasible therapeutic method for cancer treatment\textsuperscript{41}. Moreover, Varsha Prakash et al. demonstrated that upregulation of ribosome biogenesis could fuel the execution of Epithelial-to-Mesenchymal Transition (EMT), which is a migratory cellular program connected with development and tumor metastasis, and inhibition of rRNA synthesis in vivo turns primary tumors into a benign, Estrogen Receptor-alpha (ER\textalpha) positive, Rictor-negative (a fundamental constituent of the EMT-promoting mammalian target of rapamycin complex 2 ) phenotype and reduced metastasis\textsuperscript{42}. Lucia Sommerova et al. described that loss of intracellular AGR2 in cell lines caused a variation from epithelial to mesenchymal phenotype and establishment of intracellular AGR2 expression launched EMT\textsuperscript{43}. However, other studies suggested that extracellular AGR2 could interrupt adherens junctions, disrupt basal laminin and activate fibroblast-associated cancer invasion, promoting epithelial morphogenesis and tumorigenesis\textsuperscript{44}. To date, no researches about correlation between AGR2 and ribosome biogenesis have been reported. For the first time, we speculate that AGR2 may be a potential regulator of ribosome biogenesis in BRCA. In our opinion, the function of AGR2 should be explored according to different ER status and its different locations: intracellular or extracellular.

There were still limitations to the current study. Firstly, all the results were validated by using web tools, meaning a lack of in vitro and in vivo experiments. Secondly, all the researches should be performed in different subtypes respectively, due to results may be affected by ER, PR, or Her2 status.

5. Conclusion

By using bioinformatics analysis, we found that higher AGR2 expression was associated with less aggressive breast cancer. Our results was the first to demonstrated that DNA methylation rather than copy number variation led to AGR2 overexpression, which could predict favorable prognosis in TNBC, and AGR2 could be a possible regulator of ribosome biogenesis in patients with breast cancer.

Abbreviations

AGR2 anterior gradient 2

BP biological process

BRCA breast cancer

CC cellular component

CNV copy number variation

DFS disease free survive
ER estrogen receptor
FDR false discovery rate
GABA gamma-aminobutyric acid
GO Gene Ontology
Her2 human epidermal growth factor receptor 2
KEGG Kyoto Encyclopedia of Genes and Genomes
LeadingEdgeNum the number of leading edge genes
MBC mucinous breast cancer
MF molecular function
NPI Nottingham prognostic index
OS overall survive
PDI protein disulfide isomerase
PPI protein-protein interaction
PR progesterone receptor
SBR Scarff-Bloom-Richardson
TNBC triple-negative breast cancer

**Declarations**

**DATA AVAILABILITY STATEMENT**

The data that support the findings of this study are openly available in all the online databases we used, and the websites are included within this article.

**Ethics approval and consent to participate**

There was no ethics associated with this work because the study didn’t involve animal experiments or human specimens.
Consent for publication

Not applicable.

Competing interests

The authors have no conflicts of interest or disclosures relevant to this work.

Funding

Not applicable.

AUTHOR CONTRIBUTIONS

Xin Zhou designed and performed the study, analyzed and interpreted data, and drafted the manuscript. Fangyuan Zhang, Yan Liu participated in data analysis and figure preparation. Dongxin Wei revised the manuscript critically. All authors read and approved the final manuscript.

ACKNOWLEDGMENTS

We sincerely thank all the online databases we used for their user-friendly features, and the support from Zibo Maternal and Child Health Hospital.

References

1. Cao X, He Q. Anti-tumor activities of bioactive phytochemicals in sophora flavescens for breast cancer. Cancer Manag Res. 2020;12:1457–67. doi:10.2147/CMAR.S243127.

2. Salmans ML, Zhao F, Andersen B. The estrogen-regulated anterior gradient 2 (AGR2) protein in breast cancer: A potential drug target and biomarker. Breast Cancer Res. 2013;15(2). doi:10.1186/bcr3408.

3. Brychtova V, Vojtesek B, Hrstka R. Anterior gradient 2: A novel player in tumor cell biology. Cancer Lett. 2011;304(1):1–7. doi:10.1016/j.canlet.2010.12.023.

4. Li Z, Zhu Q, Chen H, et al. Binding of anterior gradient 2 and estrogen receptor-α: Dual critical roles in enhancing fulvestrant resistance and IGF-1-induced tumorigenesis of breast cancer. Cancer Lett. 2016;377(1):32–43. doi:10.1016/j.canlet.2016.04.003.

5. Tang Z, Kang B, Li C, Chen T, Zhang Z. GEPIA2: an enhanced web server for large-scale expression profiling and interactive analysis. Nucleic Acids Res. 2019;47(W1):W556–60. doi:10.1093/nar/gkz430.
6. Jézéquel P, Campone M, Gouraud W, et al. Bc-GenExMiner: An easy-to-use online platform for gene prognostic analyses in breast cancer. Breast Cancer Res Treat. 2012;131(3):765–75. doi:10.1007/s10549-011-1457-7.

7. Jézéquel P, Frénel JS, Campion L, et al. bc-GenExMiner 3.0: New mining module computes breast cancer gene expression correlation analyses. Database. 2013;2013:1–10. doi:10.1093/database/bas060.

8. Uhlén M, Fagerberg L, Hallström BM, et al. Tissue-based map of the human proteome. Science. 2015;347(6220). doi:10.1126/science.1260419.

9. Chandrashekar DS, Bashel B, Balasubramanya SAH, et al. UALCAN: A Portal for Facilitating Tumor Subgroup Gene Expression and Survival Analyses. Neoplasia (United States). 2017;19(8):649–58. doi:10.1016/j.neo.2017.05.002.

10. Manuscript A. 27 Cbio-. Sci Signal. 2014;6(269):1–34. doi:10.1126/scisignal.2004088.Integrative.

11. Rhodes DR, Kalyana-Sundaram S, Mahavisno V, et al. Oncomine 3.0: Genes, pathways, and networks in a collection of 18,000 cancer gene expression profiles. Neoplasia. 2007;9(2):166–80. doi:10.1593/neo.07112.

12. Modhukur V, Iljasenko T, Metsalu T, Lokk K, Laisk-Podar T, Vilo J. MethSurv. A web tool to perform multivariable survival analysis using DNA methylation data. Epigenomics. 2018;10(3):277–88. doi:10.2217/epi-2017-0118.

13. Díez-Villanueva A, Mallona I, Peinado MA. Wanderer, an interactive viewer to explore DNA methylation and gene expression data in human cancer. Epigenetics Chromatin. 2015;8(1):1–8. doi:10.1186/s13072-015-0014-8.

14. Xu S, Feng Y, Zhao S. Proteins with Evolutionarily Hypervariable Domains are Associated with Immune Response and Better Survival of Basal-like Breast Cancer Patients. Comput Struct Biotechnol J. 2019;17:430–40. doi:10.1016/j.csbj.2019.03.008.

15. Vasaikar SV, Straub P, Wang J, Zhang B. LinkedOmics. Analyzing multi-omics data within and across 32 cancer types. Nucleic Acids Res. 2018;46(D1):D956–63. doi:10.1093/nar/gkx1090.

16. Wang J, Vasaikar S, Shi Z, Greer M, Zhang B. WebGestalt. 2017: A more comprehensive, powerful, flexible and interactive gene set enrichment analysis toolkit. Nucleic Acids Res. 2017;45(W1):W130-W137. doi:10.1093/nar/gkx356.

17. Montojo J, Zuberi K, Rodriguez H, Bader GD, Morris Q. GeneMANIA: Fast gene network construction and function prediction for Cytoscape. F1000Research. 2014;3:1–7. doi:10.12688/f1000research.4572.1.

18. Fletcher GC, Patel S, Tyson K, et al. hAG-2 and hAG-3, human homologues of genes involved in differentiation, are associated with oestrogen receptor-positive breast tumours and interact with metastasis gene C4.4a and dystroglycan. Br J Cancer. 2003;88(4):579–85. doi:10.1038/sj.bjc.6600740.
19. Fritzsche FR, Dahl E, Pahl S, et al. Prognostic relevance of AGR2 expression in breast cancer. Clin Cancer Res. 2006;12(6):1728–34. doi:10.1158/1078-0432.CCR-05-2057.

20. Guo J, Gong G, Zhang B. Identification and prognostic value of anterior gradient protein 2 expression in breast cancer based on tissue microarray. Tumor Biol. 2017;39(7):1–9. doi:10.1177/1010428317713392.

21. Cichon MA, Moruzzi ME, Shqau TA, et al. MYC is a crucial mediator of TGFβ-induced invasion in basal breast cancer. Cancer Res. 2016;76(12):3520–30. doi:10.1158/0008-5472.CAN-15-3465.

22. Fu J, Wu L, Jiang M, et al. Clinical nomogram for predicting survival outcomes in early mucinous breast cancer. PLoS One. 2016;11(10):1–16. doi:10.1371/journal.pone.0164921.

23. Wang Z, Hao Y, Lowe AW. The adenocarcinoma-associated antigen, AGR2, promotes tumor growth, cell migration, and cellular transformation. Cancer Res. 2008;68(2):492–7. doi:10.1158/0008-5472.CAN-07-2930.

24. Jakobsson M, Scholz SW, Scheet P, et al. Genotype, haplotype and copy-number variation in worldwide human populations. Nature. 2008;451(7181):998–1003. doi:10.1038/nature06742.

25. Portela A, Esteller M. Epigenetic modifications and human disease. Nat Biotechnol. 2010;28(10):1057–68. doi:10.1038/nbt.1685.

26. Sung HY, Choi EN, Lyu D, Park AK, Ju W, Ahn JH. Aberrant hypomethylation-mediated AGR2 overexpression induces an aggressive phenotype in ovarian cancer cells. Oncol Rep. 2014;32(2):815–20. doi:10.3892/or.2014.3243.

27. Alves MR, e Melo NC, Barros-Filho MC, et al. Downregulation of AGR2, p21, and cyclin D and alterations in p53 function were associated with tumor progression and chemotherapy resistance in epithelial ovarian carcinoma. Cancer Med. 2018;7(7):3188–99. doi:10.1002/cam4.1530.

28. Armes JE, Davies CM, Wallace S, Taheri T, Perrin LC, Autelitano DJ. AGR2 expression in ovarian tumours: A potential biomarker for endometrioid and mucinous differentiation. Pathology. 2013;45(1):49–54. doi:10.1097/PAT.0b013e32835bd561.

29. Alavi M, Mah V, Maresh EL, et al. High expression of AGR2 in lung cancer is predictive of poor survival. BMC Cancer. 2015;15(1):1. doi:10.1186/s12885-015-1658-2.

30. Fritzsche FR, Dahl E, Dankof A, et al. Expression of AGR2 in non small cell lung cancer. Histol Histopathol. 2007;22(7–9):703–8. doi:10.14670/HH-22.703.

31. Barraclough DL, Platt-Higgins A, Rudland SDS, et al. The metastasis-associated anterior gradient 2 protein is correlated with poor survival of breast cancer patients. Am J Pathol. 2009;175(5):1848–57. doi:10.2353/ajpath.2009.090246.

32. Hrstka R, Nenutil R, Fourtouna A, et al. The pro-metastatic protein anterior gradient-2 predicts poor prognosis in tamoxifen-treated breast cancers. Oncogene. 2010;29(34):4838–47. doi:10.1038/onc.2010.228.

33. Innes HE, Liu D, Barraclough R, et al. Significance of the metastasis-inducing protein AGR2 for outcome in hormonally treated breast cancer patients. Br J Cancer. 2006;94(7):1057–65. doi:10.1038/sj.bjc.6603065.
34. Chevet E, Fessart D, Delom F, et al. Emerging roles for the pro-oncogenic anterior gradient-2 in cancer development. Oncogene. 2013;32(20):2499–509. doi:10.1038/onc.2012.346.

35. Tian S, Hu J, Tao K, et al. Secreted AGR2 promotes invasion of colorectal cancer cells via Wnt11-mediated non-canonical Wnt signaling. Exp Cell Res. 2018;364(2):198–207. doi:10.1016/j.yexcr.2018.02.004.

36. Jia M, Guo Y, Zhu D, et al. Pro-metastatic activity of AGR2 disrupts angiogenesis target bevacizumab efficiency via direct interaction with VEGFA and activation of NF-κB pathway. Biochim Biophys Acta - Mol Basis Dis. 2018;1864(5):1622–33. doi:10.1016/j.bbadis.2018.01.021.

37. Tian SB, Tao KX, Hu J, et al. The prognostic value of AGR2 expression in solid tumours: A systematic review and meta-analysis. Sci Rep. 2017;7(1):1–10. doi:10.1038/s41598-017-15757-z.

38. Noller HF. Evolution of protein synthesis from an RNAworld. Cold Spring Harb Perspect Biol. 2012;4(4). doi:10.1101/cshperspect.a003681.

39. Donati G, Montanaro L, Derenzini M. Ribosome biogenesis and control of cell proliferation: p53 is not alone. Cancer Res. 2012;72(7):1602–7. doi:10.1158/0008-5472.CAN-11-3992.

40. Ebright RY, Lee S, Wittner BS, et al. Deregulation of ribosomal protein expression and translation promotes breast cancer metastasis. Science. 2020;367(6485):1468–73. doi:10.1126/science.aay0939.

41. Bywater MJ, Poortinga G, Sanij E, et al. Inhibition of RNA Polymerase I as a Therapeutic Strategy to Promote Cancer-Specific Activation of p53. Cancer Cell. 2012;22(1):51–65. doi:10.1016/j.ccr.2012.05.019.

42. Prakash V, Carson BB, Feenstra JM, et al. Ribosome biogenesis during cell cycle arrest fuels EMT in development and disease. Nat Commun. 2019;10(1). doi:10.1038/s41467-019-10100-8.

43. Sommerova L, Ondrouskova E, Vojtesek B, Hrstka R. Suppression of AGR2 in a TGF-β-induced Smad regulatory pathway mediates epithelial-mesenchymal transition. BMC Cancer. 2017;17(1):1–11. doi:10.1186/s12885-017-3537-5.

44. Tsuji T, Satoyoshi R, Aiba N, et al. Agr2 mediates paracrine effects on stromal fibroblasts that promote invasion by gastric signet-ring carcinoma cells. Cancer Res. 2015;75(2):356–66. doi:10.1158/0008-5472.CAN-14-1693.

**Figures**
Figure 1

Expression of AGR2 in breast cancer (BRCA) based on TCGA data analyzed by GEPIA2. (A) The expression of AGR2 in pan-cancer. (B) The expression of AGR2 in BRCA and normal tissues (*p<0.01). (C) AGR2 expression in different subtypes of BRCA (*p<0.01). (D) Comparisons of AGR2 expression between different subtypes and normal tissues (*p<0.01).
Figure 2

Expression of AGR2 in breast cancer (BRCA) based on TCGA data analyzed by GEPIA2. (A) The expression of AGR2 in pan-cancer. (B) The expression of AGR2 in BRCA and normal tissues (*p<0.01). (C) AGR2 expression in different subtypes of BRCA (*p<0.01). (D) Comparisons of AGR2 expression between different subtypes and normal tissues (*p<0.01).
Figure 3

The expression of AGR2 protein in breast cancer. (A) The IHC expression of AGR2 in normal breast tissue obtained from HPA database. (B) The IHC expression of AGR2 in breast cancer obtained from HPA database. (C) Comparison of AGR2 protein expression between breast cancer and normal tissue determined by UALCAN database (p=8.59E-11). (D) Correlation between AGR2 mRNA expression and protein expression by using cbioportal database (Spearman: 0.80, p=1.33e-17; Pearson: 0.69, p=8.00e-12).
Gene alteration of AGR2 in BRCA (A-B) Gene alteration of AGR2 occurred in 76 (7%) of total 1093 sequenced cases, including 13 cases of amplification (1.19%), 61 cases of mRNA low (5.58%) and 2 cases of multiple alterations (0.18%) (cbioportal database). (C) DNA copy number of AGR2 was significantly higher in BRCA than normal tissues (p=0.027) (Oncomine database). (D) No correlation was found between AGR2 copy number variation (CNV) and mRNA expression (Spearman: 0.08, p=9.166e-3; Pearson 0.07, p=0.0313) (cbioportal database). (E-F) AGR2 CNV was not associated with overall survive (OS) (p=0.776) or disease free survive (DFS) (p=0.0947) in BRCA (cbioportal database).
Figure 5

DNA methylation status of AGR2 in BRCA. (A) The methylation status of AGR2 was significantly lower in BRCA than in normal tissues (p<1E-12) by using UALCAN database. (B) AGR2 mRNA expression was negatively associated with its methylation status (Spearman: -0.78, p=6.75e-160; Pearson: -0.60, p=1.28e-76) determined by cbioportal database. (C) No correlation was found between AGR2 methylation status and CNV (Spearman: -0.04, p=0.273; Pearson: -0.04, p=0.242). (D) According to Wanderer database, AGR2 methylation level was significantly lower in BRCA than in normal tissues at five probes (*p<001).
Figure 6

AGR2 DNA methylation analysis by using Wanderer database and Methsurv database. (A-E) Correlations between AGR2 mRNA and methylation status at five probes obtained by Wanderer base, it showed the strongest association at probe cg 25343204 (Pearson $r = 0.646$). (F-K) Survival analyses showed that lower AGR2 methylation status indicated better prognosis only at probcg 25343204 ($p=0.036$) by using Methsurv database.
Figure 7

Prognostic Value of AGR2 in BRCA. (A) According to GEPIA2 database, AGR2 was not a prognostic marker in BRCA (OS, p>0.05; DFS, p>0.05). (F) Subtype analysis suggested that low expression of AGR2 indicated poor prognosis in base-like BRCA (p=0.00061), but no prognostic value of AGR2 was found in other subtypes (TCGApolicy database). (G-J) Subtype analysis suggested that low expression of AGR2
indicated poor prognosis in base-like BRCA (p=0.0130), but no prognostic value of AGR2 was found in other subtypes (bc-GenexMiner database).

**Figure 8**

GO and KEGG pathway analyses of co-expression genes correlated with AGR2 in BRCA. (A) Correlations between RBM8A and genes differentially expressed in BRCA by using Pearson test. (B-C) Heat maps showing genes positively and negatively correlated with AGR2 in BRCA (TOP 50). Volcano plots of the most significant results of GO analysis in BRCA: (D) Biological processes (BP); (E) Cellular components (CC); (F) Molecular functions (MF). (G) Volcano plot of the most significant result of KEGG pathway analysis in BRCA.
Figure 9

Protein-protein interaction network of the gene set that was enriched in the pathway of ribosome biogenesis in eukaryotes (GeneMANIA). Different colors of the network edge indicate the bioinformatics methods applied: co-expression, website prediction, pathway, physical interactions and co-localization. The different colors for the network nodes indicate the biological functions of the set of enrichment genes.
Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- sfig2.pdf
- sfig1.pdf
- Supplementary.docx