New insights into the regulatory role of microRNA in tumor angiogenesis and clinical implications

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Abstract

Angiogenesis is essential for tumor growth and metastasis. Understanding the regulation of tumor angiogenesis has become increasingly important. MicroRNAs (miRNAs) are small noncoding RNAs that function in diverse biological processes via post-transcriptional regulation. Extensive studies have revealed two important regulatory roles of miRNAs in tumor angiogenesis: miRNAs in tumor cells affect the activity of endothelial cells via non-cell-autonomous mechanisms, and miRNAs in endothelial cells regulate the cell-autonomous behavior. Recent advances have further highlighted the role of tumor-derived extracellular vesicles in the regulation of tumor angiogenesis via transferring miRNAs to endothelial cells. In this review, we summarize the regulatory role of miRNA in tumor angiogenesis, with a highlight on clinical implications of miRNAs as biomarkers for anti-angiogenic therapy response, and as therapeutic interventions against tumor angiogenesis in vivo.

Keywords: Tumor angiogenesis, microRNA, Extracellular vesicle, Biomarker, miRNA delivery

Background

Inducing angiogenesis is one of the hallmarks of cancer [1]. Tumor-associated neovasculature is important for delivering nutrients and oxygen to growing tumors, and also playing key roles in multi-aspect of tumor biology, including tumor dissemination/metastasis [2], metabolic deregulation [3] and cancer stem cell maintenance [4, 5]. For its fundamental role in tumor growth and progression, angiogenesis has become an appealing target in cancer treatment.

Angiogenesis is a complex process by which new blood vessels are formed from pre-existing ones by sprouting, remodeling and expansion of primary vascular networks [6]. In normal conditions, following this morphogenesis, the vasculature becomes largely quiescent. However, within tumors, an “angiogenic switch” is always activated, causing continuous generation of new vessels [7]. The “angiogenic switch” is governed by pro-angiogenic and anti-angiogenic signals elicited by tumor cells or tumor microenvironment [8, 9]. When pro-angiogenic signals are activated or anti-angiogenic signals are inhibited, the “angiogenic switch” is turned on. Understanding the regulation of tumor angiogenesis is critical to developing therapeutic strategies against cancer.

MicroRNAs (miRNAs) are small non-coding RNAs [10], that negatively regulate gene expression at post-transcriptional level. Through sequence-specific interaction with the 3′-untranslated region (UTR) of target mRNA, miRNAs suppress gene expression via transcript degradation or inhibition of protein translation. To date, miRNAs have been reported to play key roles in diverse biological processes of cancer, including tumor growth, metastasis, angiogenesis and drug resistance [11–14]. Extensive studies have shown that miRNAs could modulate the function of endothelial cells (ECs) via non-cell-autonomous (genetic and epigenetic alterations in a cell affect the phenotypes of other cells through changing microenvironment or intercellular interaction) and cell-autonomous (genetic and epigenetic alterations in a cell affect the cell’s own phenotypes) manner during tumor angiogenesis. MiRNAs in tumor cells regulate the expression of pro- or anti-angiogenic factors, thereby modulating the proliferation and migration of ECs in a...
paracrine manner. Endothelial miRNAs regulate the response of ECs to multiple angiogenic stimuli mainly by targeting growth factor receptors and signaling molecules autonomously. Recently, extracellular vesicles (EVs) are emerging as mediators of intercellular communication [15]. Cancer cell-derived miRNA-containing EVs could be transferred to ECs where they induce pro-angiogenic effects. Importantly, several miRNAs have been shown to serve as predictive biomarkers for anti-angiogenic therapy response, especially as non-invasive biomarkers in the circulation. Furthermore, with the development of RNA delivery technology, miRNA-based interventions may act as novel therapeutic means to target tumor angiogenesis.

In our review, we will summarize the non-cell-autonomous and cell-autonomous roles of miRNAs, as well as EV-transferred miRNAs in the regulation of tumor angiogenesis. Moreover, we highlight the clinical implications of miRNAs as predictive biomarkers for anti-angiogenic therapy response in both tissue and circulation, and several miRNA delivery approaches that have been applied to suppress angiogenesis in vivo.

**Non-cell-autonomous regulation of tumor angiogenesis by miRNAs in tumor cells**

Angiogenesis is under the control of pro-angiogenic and anti-angiogenic factors [7]. Tumor cells could produce and secrete such factors into surrounding environment to promote vessel growth. Endogenous activators and inhibitors are frequently targeted in tumor cells by miRNAs, which thereby regulate angiogenesis in a non-cell-autonomous manner (Table 1).

**Tumoral miRNAs regulating VEGF-dependent angiogenesis**

The most predominant growth factor of angiogenesis is VEGF, consisting of VEGF-A/B/C/D and placental growth factor (PIGF) [16, 17]. VEGF-A (also known as VEGF) is a potent trigger for angiogenesis by signaling through VEGF receptor-2 (VEGFR-2) [18]. While VEGF-B specifically regulates the embryonic angiogenesis of myocardial tissue [19] and VEGF-C/D mainly participates in the genesis and maintenance of lymphatic vessels. Therefore, among the VEGF factors, VEGF-A is most widely studied in the regulation of tumor angiogenesis [20].

MiR-20 [21], miR-29b [22], miR-93 [23–25], miR-126 [26–30], miR-190 [31], miR-195 [32], miR-200 [33, 34], miR-203 [35], miR-497 [36, 37], miR-503 [38] and miR-638 [39] have been experimentally shown to directly target the 3’UTR region of VEGF-A mRNA in different types of cancer and subsequently impairing the pro-angiogenesis signaling of VEGF/VEGFR-2 in endothelial cells. These miRNAs are generally downregulated in cancer. Recently, the role of VEGF-C in initiating and potentiating neo-angiogenesis has been uncovered [40]. MiR-27b and miR-128 functioned as inhibitor of tumor progression and angiogenesis through targeting VEGF-C [41, 42].

Apart from directly targeting VEGF, a handful of miRNAs regulate VEGF-dependent tumor angiogenesis by targeting VEGF inducers, such as HIF pathway (miR-22 [43], miR-107 [44], miR-519c [45], miR-145 [46]), PI3K/AKT pathway (miR-26a [47], miR-145 [48]), mTOR pathway (miR-18a [49], miR-128 [50], miR-145 [51], miR-218 [52]), IGF1R pathway (miR-126, miR-181b, miR-148a, miR-152 [53–55]) and VEGF upstream transcription factors (STAT3: miR-874 [56], Bmi-1: miR-16 [57], E2F3: miR-34a [58], NF90: miR-590-5p [59]).

**Table 1 MiRNAs involved in non-cell-autonomous regulation of tumor angiogenesis in tumor cells**

| Targets | miRNAs | Expression in tumor cells | Ref |
|---------|--------|---------------------------|-----|
| VEGF    | miR-20, miR-29b, miR-93, miR-126, miR-190, miR-195, miR-200, miR-203, miR-497, miR-503, miR-638, miR-27b, miR-128 | down | [21–42] |
| VEGF inducers | HiF-1, miR-22, miR-107, miR-519c | down | [43–45] |
|          | HiF-2, miR-145 | down | [46] |
|          | PI3K/AKT pathway, miR-26a, miR-145 | down | [47, 48] |
|          | mTOR pathway, miR-18a, miR-128, miR-145, miR-218 | down | [49–52] |
|          | IGF1R pathway, miR-126, miR-181b, miR-148a, miR-152 | down | [53–55] |
| VEGF upstream TFs | STAT3: miR-874 | down | [56] |
|          | Bmi-1: miR-16 | down | [57] |
|          | E2F3: miR-34a | down | [58] |
|          | NF90: miR-590-5p | down | [59] |
| VEGF suppressor | PTEN: miR-21 | up | [60] |
|          | E-cadherin: miR-9 | up | [61] |
| VEGF independent factors | FGFR: miR-503 | down | [38] |
|          | HDGF: miR-214, miR-497 | down | [62, 63] |
|          | angiogenin: miR-409-3p | down | [64] |
|          | angiopoietin: miR-204 | down | [65] |
|          | MMPs: miR-9, miR-26b-5p, miR-98, miR-181a-5p | Down | [66–69] |
|          | TSP-1: miR-17-92 cluster, miR-182, miR-194, miR-467 | up | [75–78] |

**IGF1R** Insulin-like growth factor 1 receptor, Bmi-1: B-cell-specific Moloney murine leukemia virus integration site 1, E2F3: E2F transcription factor 3, NF90: Nuclear factor 90, TSP-1: Thrombospondin-1, HDGF: Hepatoma-derived growth factor
signaling and finally enhanced HIF-1α and VEGF expression in prostate cancer cells [60]. E-cadherin downregulation by miR-9 activated β-catenin signaling, in turn upregulating VEGF-A expression and promoting tumor angiogenesis [61].

**Tumoral miRNAs regulating VEGF-independent angiogenesis**

Several other growth factors also play positive roles in tumor angiogenesis. MiRNAs have been reported to target these pro-angiogenic factors in tumor cells, such as FGF (miR-503 [38]), HDGF (miR-214 [62], miR-497 [63]), angiogenin (miR-409-3p [64]), angiopoietin (miR-204 [65]), and MMPs (miR-9 [66], miR-26b-5p [67], miR-98 [68], miR-181a-5p [69]). Moreover, some miRNAs were shown to simultaneously target more than one angiogenic factors. MiR-190 significantly suppressed tumor angiogenesis via targeting VEGF, HGF and IGF1, thus altering the local microenvironment and subsequently regulating neighboring ECs [31].

Angiogenesis is regulated by the balance between pro- and anti-angiogenic factors. Angiostatin, endostatin and thrombospondin-1 (TSP-1) are endogenous anti-angiogenic factors [70–72], among which TSP-1 is most well-known. TSP-1 binds to its receptor CD36 on the surface of ECs, as well as by binding to β1-integrins, leading to cell apoptosis [73]. In addition, TSP-1 inhibits the activity of MMP-9, thereby hindering the release of VEGF-A sequestered in extracellular matrix [74]. Several miRNAs, including miR-17-92 cluster [75], miR-182 [76], miR-194 [77] and miR-467 [78], have been reported to target TSP-1 in tumor cells, thus decreasing TSP-1 secretion and promoting tumor angiogenesis.

**Cell-autonomous regulation of tumor angiogenesis by miRNAs in endothelial cells**

The importance of miRNAs in ECs was demonstrated by the silencing of Dicer, an enzyme responsible for miRNA maturation. Knockdown of Dicer in ECs inhibited cell proliferation, migration and cord formation [79], indicating that miRNAs are important in the function of ECs. Many studies have shown that miRNAs in ECs regulate the cellular response to angiogenic factors by targeting surface receptors and signaling molecules (Table 2).

**MiRNAs targeting angiogenic factor receptors in endothelial cells**

Tyrosine kinase receptors VEGFR and platelet-derived growth factor receptor (PDGFR) are primarily located on the surface of ECs [80, 81]. The binding of VEGF-A to VEGFR-2 and PDGFR to PDGFR initiates a cascade of signals that result in endothelial cell proliferation and migration [80]. Several miRNAs have been reported to directly target these receptors in ECs, such as miR-128 [42] and miR-497 [82], blocking their downstream signaling to inhibit tumor angiogenesis. On the contrary, miRNAs targeting the suppressors of these receptors promoted tumor angiogenesis. MiR-296 contributed to angiogenesis by directly targeting the hepatocyte growth factor-regulated tyrosine kinase substrate (HGS) mRNA, thereby reducing HGS-mediated degradation of VEGFR-2 and PDGFRβ in ECs [83]. MiR-146a enhanced tumor angiogenesis by targeting BRCA1 which conferred a transcriptional repression on PDGFRA [84].

**MiRNAs targeting signaling molecules in endothelial cells**

MiRNAs have been implicated as key modulators of angiogenic signal transduction pathways. MiR-125b suppressed tube formation of ECs by inhibiting VE-cadherin translation [85]. MiR-34a was shown to inhibit the proliferation, migration and tube formation of ECs by downregulating a number of key proteins including E2F3, SIRT1, survivin and CDK4 [58]. Conversely, miRNAs promote angiogenesis by targeting negative regulators in angiogenic signaling pathways. MiR-132 was highly expressed in the endothelium of human tumors and hemangiomas but was undetectable in normal endothelium. MiR-132 induced Ras activation via suppressing p120RasGAP expression, thus promoting neovascularization [86]. MiR-130a is a positive regulator of ECs largely through targeting angiogenic homeobox gene GAX and HOXA5 [87]. Notably, miR-126 has been well known as an endothelial-specific miRNA and has pro-angiogenic action during embryo development [88]. However, a recent study illustrated an anti-angiogenic role of miR-126 during tumor angiogenesis by targeting a pro-angiogenic gene adrenomedullin (ADM) [89]. This study suggests that the role of miRNAs may vary depending on microenvironment.

**Regulation of tumor angiogenesis by miRNAs in tumor-derived extracellular vesicles**

The discovery of extracellular vesicles (EVs) represents an exciting area of research that offers novel mechanisms for intercellular communication. EVs, generally including exosomes (typically 30–100 nm) and microparticles or microvesicles (typically 100 nm-1 μm), are small secretory vesicles that contain proteins, DNA, and coding and non-coding RNAs [90]. EVs can be released from various cell types under physiological and pathological conditions and transfer their cargo to proximal or distal cells [91]. In many published articles, the collective term “microvesicles” was used for all the EVs between 30 and 1000 nm in diameter.

Recent evidence has demonstrated that cancer cell-derived EVs can be transferred to ECs where they induce pro-angiogenic effects. Glioblastoma cells released exosomes containing mRNA, miRNAs and angiogenic proteins. These exosomes were taken up by brain microvascular endothelial cells and modulated angiogenesis
A set of pro-angiogenic mRNAs and miRNAs were enriched in the microvesicles of CD105-positive renal cancer stem cells, therefore conferring an activated angiogenic phenotype to ECs [93].

MiRNAs are the most abundant non-coding RNAs enriched in EVs, and studies have also showed that EVs contain components of the RNA-induced silencing complex (RISC), suggesting potent regulatory role of EV-miRNA in recipient cells [94]. Several studies have reported the regulation of signaling pathways in ECs by miRNAs delivered via tumor-derived EVs (Table 3). MiR-494 containing EVs secreted by tumor cells were taken up by ECs and promoted the migration of EC via targeting of PTEN and subsequent activation of Akt/eNOS pathway [95]. EV-encapsulated miR-9 transferred from tumor cells to ECs reduced SOC5 level, leading to JAK-STAT pathway activation to promote endothelial cell migration and sprout [96].

Hypoxia is a potent trigger of angiogenesis. Some studies have focused on clarifying how hypoxia affects tumor angiogenesis through tumor cell-derived EVs. Hsu et al. found that more exosomes were produced under hypoxic conditions than normoxic conditions by lung cancer cells. Tumor-derived exosomal miR-23a directly suppressed the expression of prolyl-hydroxylase 1 and 2 (PHD1 and 2), leading to the accumulation of HIF-1α in ECs and increased angiogenesis [98]. Tadokoro et al. reported that exosomes derived from leukemia cells under hypoxia are potent inducers of angiogenesis through transfer of miR-210 to ECs, downregulating the expression of receptor tyrosine kinase ligand Ephrin-A3 in HUVECs [99]. Neutral Sphingomyelinase 2 (nSMase2) was shown to regulate exosomal miR-210 secretion by metastatic cancer cells [100].

Collectively, these observations suggest that tumor-secreted EV miRNAs participate in intercellular communication and function as a novel pro-angiogenic mechanism.

**Table 2** MiRNAs involved in cell-autonomous regulation of tumor angiogenesis in ECs

| miRNAs | Targets | Function | Ref |
|--------|---------|----------|-----|
| pro-angiogenesis | miR-132 | p120RasGAP | promote ECs proliferation, tube formation | [86] |
| | miR-130a | GAX, HOXAS | promote ECs proliferation, migration, tube formation | [87] |
| | miR-146a | BRCA1 | promote ECs proliferation, migration, tube formation | [84] |
| anti-angiogenesis | miR-128 | VEGFR-2 | inhibit ECs tube formation | [42] |
| | miR-497 | VEGFR-2 | induce ECs apoptosis | [82] |
| | miR-296 | HGS | inhibit ECs tube formation | [83] |
| | miR-125b | VE-cadherin | inhibit ECs tube formation | [85] |
| | miR-34a | E2F3, SIRT1, survivin, CDK4 | inhibit ECs proliferation, migration and tube formation | [58] |
| | miR-126 | ADM | inhibit ECs migration and tube formation | [88] |

HSG hepatocyte growth factor-regulated tyrosine kinase substrate, BRCA1 breast cancer 1, p120RasGAP Ras p21 protein activator 1, SIRT1 sirtuin 1, ADM adrenomedullin

**Table 3** MiRNAs involved in angiogenesis regulation by tumor-derived extracellular vesicles

| miRNAs in EVs | Targets | Function | Ref |
|---------------|---------|----------|-----|
| miR-494 | PTEN | promote ECs migration, tube formation | [95] |
| miR-9 | SOC5 | promote ECs migration, tube formation | [96] |
| miR-1246 | PML | promote ECs proliferation, migration, tube formation | [97] |
| miR-23a | PHD1, PHD2, ZO-1 | increase the number of tumor vessels, disrupt endothelial barriers | [98] |
| miR-210 | Ephrin-A3 | Promote ECs tube formation | [99] |
| miR-105 | ZO-1 | destroy tight junctions and integrity of vascular endothelial barriers | [102] |

SOC5 suppressor of cytokine signalling 5, PML promyelocytic leukemia, PHD prolyl hydroxylase, ZO-1 tight junction protein 1

**MiRNAs as biomarkers for anti-angiogenic therapy response**

As angiogenesis is essential for tumor growth and progression, anti-angiogenic therapies have been developed to improve the outcomes of cancer patients. Several agents,
including bevacizumab, aflibercept and most recently ramucirumab that target the VEGF/VEGFR signaling pathway, have been approved across some cancer types. In addition, multi-targeted tyrosine kinase inhibitors (TKIs), such as sorafenib and sunitinib, also block angiogenesis signaling. However, the overall response rate of these anti-angiogenic therapies is unsatisfactory in the clinic. Therefore, selecting out the patients, who would benefit from anti-angiogenic treatment, will help to promote therapy efficacy and avoid unnecessary toxic adverse effect. Due to the high tissue specificity and stability of miRNAs and their altered expression in tumor angiogenesis, miRNAs have been suggested as potent predictive biomarkers for therapeutic response (Table 4).

### MiRNA biomarkers of anti-angiogenic therapy response in tumor tissue

Bevacizumab, a humanized monoclonal antibody against VEGF-A, has been approved as combination therapy or monotherapy for first- and second-line treatment of several advanced cancers, including metastatic colorectal cancer (mCRC), metastatic renal cell carcinoma (mRCC), metastatic non-small cell lung cancer (NSCLC), progressive glioblastoma, metastatic cervical cancer and ovarian cancer etc. [103]. In TCGA dataset, high miR-378 expression was associated with poor progression-free survival (PFS) in recurrent ovarian cancer patients treated with bevacizumab (low vs. high, 9.2 vs. 4.2 months; *p* = 0.04). Multivariate analysis revealed that miR-378 expression was an independent predictor for PFS after anti-angiogenic treatment [104]. For patients with mCRC treated with first line CAPEOX (capecitabine and oxaliplatin) alone, or CAPEOX and bevacizumab (CAPEOXBVE), predictive miRNAs for therapy effectiveness were identified using PCR arrays. Higher miR-664-3p level and lower miR-455-5p level were predictive of improved outcome in the CAPEOXBVE cohorts and showed a significant interaction with bevacizumab effectiveness [105].

Several multi-target TKIs have potent anti-angiogenic effects due to the inhibition of VEGFR and PDGFR, and have been approved, including sorafenib for mRCC and unresectable hepatocellular carcinoma (HCC), and sunitinib for mRCC. High level of miR-425-3p was associated with longer PFS in HCC patients treated with sorafenib. Multivariate analysis confirmed the predictive significance of miR-425-3p, suggesting that assessment of miR-425-3p levels in liver biopsies could help in stratifying patients with advanced HCC for sorafenib treatment [106]. Khella et al. compared miRNA profile between patients with a short (≤12 months) versus prolonged (>12 months) PFS under sunitinib as first-line therapy for metastatic RCC. They developed miRNA statistical models that could accurately distinguish the two groups. MiR-221 overexpression was validated to be associated with a poor PFS, while its target VEGFR2 was associated with longer survival [107]. In another study, miRNA profiling in tumor tissues of sunitinib-treated mRCC patients was performed by TaqMan Low Density Arrays. Decreased tissue levels of miR-155 and miR-484 were significantly associated with increased time to progression (TTP), suggesting miR-155 and miR-484 are potentially connected with sunitinib resistance and failure of the therapy [108].

### Circulating miRNA biomarkers of anti-angiogenic therapy response

Although tissue miRNA expression could serve as molecular classifier for clinical outcome, there are several drawbacks including small quantities of diagnostic tumor tissue, genetic tumor heterogeneity in space and time, and unavailable acquisition of repeated biopsies. Therefore, circulating biomarkers offer a non-invasive opportunity for early diagnosis, prognosis evaluation and drug response prediction in cancer. Circulating miRNAs are stable and reproducible in blood due to their incorporation in exosomes/microvesicles or protein complexes, protecting them from degradation by endogenous RNase [109, 110]. Technical advances in detection and profiling

| Table 4 Tissue and blood miRNA biomarkers for anti-angiogenesis therapeutic response |
|-----------------------------------------------|---------------|----------------|-------------------|-----------------|-------------------|
| Clinical Outcomes | miRNAs | Sample type | Detection time | Therapy | Cancer | Patient cohort<sup>a</sup> | Ref |
| Better outcomes | miR-378 | tissue | post-therapy | bevacizumab | ovarian cancer | 113/34 | [104] |
| | miR-455-5p | tissue | post-therapy | bevacizumab | mCRC | 212/121 | [105] |
| | miR-221 | tissue | post-therapy | sunitinib | mRCC | 30/27 | [107] |
| | miR-155, miR-484 | tissue | post-therapy | sunitinib | mRCC | 16/63 | [108] |
| | miR-126 | blood | pre- and post-therapy | bevacizumab | mCRC | 75/68 | [111] |
| | miR-665 | blood | pre-therapy | bevacizumab/erlotinib | NSCLC | 51/50 | [112] |
| Worse outcomes | miR-664-3p, | tissue | post-therapy | bevacizumab | mCRC | 212/121 | [105] |
| | miR-425-3p | tissue | pre- and post-therapy | sorafenib | HCC | 26/58 | [106] |
| | miR-223 | blood | pre-therapy | bevacizumab/erlotinib | NSCLC | 51/50 | [112] |

<sup>a</sup>mCRC metastatic colorectal cancer, HCC hepatocellular carcinoma, mRCC metastatic renal cell carcinoma, NSCLC non-small-cell lung cancer

<sup>a</sup>The pattern A/B represents patient number in training cohort and validation cohort respectively
makes them promising candidates. Recently, the occurrence of miRNAs in the serum or plasma of human has been repeatedly observed and become the focus of biomarker discovery.

Hansen et al. investigated the predictive value of circulating miR-126 in mCRC patients receiving first-line chemotherapy combined with bevacizumab. Blood samples of patients were collected before treatment (baseline), 3 weeks after treatment and at progression. Evaluation of miR-126 expression in plasma showed that non-responders had an increase in miR-126 level relative to baseline, while responders with decreased miR-126 alteration. The results indicated that changes in circulating miR-126 during treatment represented a possible biomarker for the response to anti-angiogenic containing therapy [111]. M. Joerger’s study assessed the predictive value of circulating miRNA in patients with non-squamous NSCLC, receiving treatment with first-line bevacizumab and erlotinib followed by platinum-based chemotherapy at progression. MiRNA profiling in pre-treatment blood showed that 12 miRNAs were significantly associated with tumor shrinkage following bevacizumab/erlotinib treatment, with miR-665 being the strongest predictive marker. Patients with high miR-665 expression had a higher chance for tumor shrinkage. Furthermore, miR-223 was found to be associated with TTP following bevacizumab/erlotinib treatment and TTP following second-line chemotherapy [112]. Collectively, these studies indicate that tissue and circulating miRNAs could serve as predictive biomarkers for response to anti-angiogenic therapy.

MiRNA-based therapeutics to target tumor angiogenesis

Given the important regulatory role of miRNAs in tumor angiogenesis, targeting or delivering miRNAs to tumors holds great promise as a novel therapeutic approach. In this case, it is essential to deliver antagoniRs or miRNA mimics into tumor cells or endothelial cells. As naked miRNAs are hydrophilic, unable to pass through cell membranes, and could be easily degraded by serum RNase [113], in vivo application of miRNA requires formulation into delivery systems. Here we summarize some miRNA delivery approaches that have been applied to suppress angiogenesis in vivo (Table 5).

Nanoparticles for therapeutic miRNA delivery

Nanoparticle delivery systems, including viral and non-viral ones, have been addressed to protect miRNAs from degradation by serum nucleases when administered systemically [114]. The advantage of non-viral approach is safety and avoiding induction of toxic immune response compared with viral delivery. Non-viral delivery systems mainly use liposomes, lipoplexes and polyplexes as miRNA carriers. The encapsulation of miRNAs into polyacation liposomes has been well studied. Muramatsu et al. injected miR-125b directly into the subcutaneous tumor using non-viral vectors composed of the cationic polymer polyethyleneimine (PEI). Administration of miR-125b induced formation of non-functional blood vessels and inhibited in vivo tumor growth by targeting VE-cadherin, suggestive of therapeutic potential [85]. Another kind of liposome is neutral liposomal particle 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine (DOPC) [115]. Wu et al. reported that the delivery of miR-192, which targets EGR1 and HOXB9, to tumors using the DOPC nanoliposomes significantly inhibited tumor angiogenesis of multiple ovarian and renal cancer models [116]. This delivery platform has also been used for miR-200 and miR-520d in several cancer types in preclinical models to inhibit tumor angiogenesis in vivo [117, 118].

To turn miRNA into therapeutics, it is critical to deliver antagoniRs or miRNA mimics specifically into target cells. Endothelial cells in the tumor microenvironment have been actively targeted in vivo for miRNA delivery. The cyclic Arginine-Glycine-Aspartic acid (cRGD) peptide coupled nanoparticle is most widely used. The cRGD could bind to integrin αvβ3 and αvβ5, which are present on tumor ECs, as well as on certain tumor cells. Studies have reported that systemic administration of anti-miR-296, anti-miRNA-132 or miR-7 mimics using the cRGD

| miRNAs  | Delivery systems | Cancer                      | Targets          | Ref |
|---------|-----------------|-----------------------------|------------------|-----|
| miR-125b | PEI             | lung cancer, colorectal cancer | VE-cadherin     | [85]|
| miR-192  | DOPC            | ovarian cancer, renal tumor  | EGR1, HOXB9      | [116]|
| miR-200  | DOPC            | lung cancer, ovarian cancer, renal cancer | IL8, CXCL1 | [117]|
| miR-520d | DOPC            | ovarian cancer              | EphA2           | [118]|
| miR-132  | cRGD            | breast cancer               | p120RasGAP      | [86]|
| miR-499  | APRPG-PEG       | colorectal cancer           | VEGF            | [122]|
| miR-29a/c| CMVs            | gastric cancer              | VEGF            | [127]|
| miR-150  | CMVs            | sarcoma                     | VEGF            | [128]|

*PEI* polymer polyethylenimine, *DOPC* 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine, *cRGD* cyclicArginine-Glycine-Aspartic acid, *APRPG-PEG* Ala-Pro-Arg-Pro-Glyepptide-conjugated polyethylene glycol, *CMVs* Cell-derived membrane vesicles
modified nanoparticles strongly reduced angiogenesis and tumor burden in mice, offering promise for miRNA-based anti-tumor therapeutic [86, 119, 120]. In addition to cRGD modification, anti-angiogenic effect was achieved by using miRNA- incorporated nanoparticles bearing Ala-Pro-Arg-Pro-Gly (APRPG) peptide as well. APRPG has an affinity for VEGFR-1, which is overexpressed on ECs and certain cancer cells [121]. In vivo delivery of miR-499 via APRPG-PEG-modified lipoplexes accumulated in both angiogenic vessels and cancer cells, resulting in the downregulation of miR-499 targeted proteins and inhibition of tumor growth and angiogenesis [122].

**Cell-derived membrane vesicles for therapeutic miRNA delivery**

Cell-derived membrane vesicles (CMVs), including exosomes and microvesicles, are endogenous carriers transporting biological molecules to recipient cells. In contrast to established synthetic nanoparticles, CMVs are natural transporters and hence, less likely to exert toxicity or immune response, which probably due to their lowhoming to the liver [123, 124]. Moreover, CMVs are shown to contain ribonucleoprotein involved in cellular RNA transport as well as RISC components. Exosomes have already been utilized to transfer therapeutic RNAi to treat diseases [125, 126]. Therefore, CMVs hold great promise as a novel class of miRNA delivery system.

Zhang et al. revealed that CMVs derived from HEK293T cells are capable of packing and transporting miR-29a/c into gastric cancer cells. The in vivo experiments illustrated that CMVs could stably transfer miR-29a/c into the implanted tumor cells in mice, playing anti-angiogenic role by directly targeting VEGF [127]. Their data proved that CMVs function as a potential carrier of miRNAs for targeted therapy in gastric cancer. Similarly, Liu et al. used a MV-delivered antisense RNA against miRNA to treat tumors. With the utilization of CMVs derived from HEK293T cells, they transferred anti-miR-150 into mice and found that the neutralization of miR-150 downregulated VEGF levels in vivo and attenuates angiogenesis and tumor growth [128]. These results propose a novel anti-cancer strategy using miRNA-containing CMVs to control tumor angiogenesis.

**Conclusions**

Significant advances have been made in exploring the regulatory role of miRNAs in tumor angiogenesis. The rapidly increasing discoveries shall pave the way in the use of miRNAs as predictive biomarkers for anti-angiogenic treatments and as miRNA-based strategy against tumor angiogenesis in the future, though there are some challenges.

Given the low response rate of anti-angiogenic agents, a step forward would be discovering predictive biomarkers to distinguish responders from non-responders. So far, several candidate miRNA biomarkers have been identified, but they emerged from small studies and should be confirmed in large prospective trials. Besides, there has been no unified control miRNAs for normalization in the analysis of circulating miRNA expression. A proposed solution to this problem may be the introduction of exogenous control [129].

Multi-targeted anti-angiogenic approach might exert increasing antitumor efficacy. Since a single miRNA has the potential to regulate angiogenesis by targeting multiple miRNAs, miRNA holds great promise as therapeutic approach for tumor angiogenesis treatment [130]. However, targeting miRNAs systemically may affect normal angiogenesis in which these miRNAs might play regulatory roles as well. In this regard, it is important to identify and target miRNAs that could distinguish angiogenic endothelial cells in tumor vasculature from those in normal tissues [131, 132], thus achieving more specific therapeutic effect and reducing side effect.

At the same time, developing technologies to deliver miRNAs to specific cells in vivo is highly essential for their therapeutic application. Despite having many strategies reported, the successful delivery approach in vivo is still limited. Clinical translation has been hampered by dose-dependent toxicity upon systemic administration [133, 134]. Intriguingly, Kalluri’s recent study showed that exosomes derived from normal fibroblast-like mesenchymal cells exhibited a superior ability to deliver RNAi to suppress pancreatic cancer growth, and showed minimum cytotoxic effects in vivo compared to synthetic nanoparticles. The engineered exosomes (known as iExosomes) target oncogenic KRAS with enhanced efficacy probably due to that CD47 on exosomes surface contributed to the evasion from immune clearance in the circulation, and KRAS-stimulated macropinocytosis increased pancreatic cancer cells uptake of iExosomes [126]. This study offers insight into the therapeutic potential of exosomes in the delivery of nucleic acid-based drugs. However, in terms of using exosomes for specific targeting of angiogenesis-related miRNAs in tumor cells or ECs, there are several aspects to consider. New targeting moieties should be tested for selective homing of exosomes to certain tumor cells and tumor-associated ECs. New targeting moieties, such as peptides and antibody fragments, could be cloned into exosomal surface proteins, or directly attached to the surface of exosomes through chemical conjugation, therefore bypassing time-consuming cloning procedures. Furthermore, the use of miRNA sponge would allow for persistent suppression of miRNA in target tissues following exosome delivery. Additionally, it is critical to find a robust cell source that produces high quantities of exosomes for use in miRNA delivery. For instance, Myc-immortalized human ES-derived mesenchymal
stem cells (MSCs) have been demonstrated as a scalable source for production of therapeutic exosomes for regenerative medicine [135].

In summary, better understanding of the regulatory network of miRNAs in tumor angiogenesis provides new insights in the biological process of tumor-associated neovascularization. Advances in profiling and delivery system offer clinical translation potential of miRNAs as predictive biomarkers for anti-angiogenic therapy response and therapeutics against tumor angiogenesis.

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