A vertex similarity-based framework to discover and rank orphan disease-related genes

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Abstract

Background: A rare or orphan disease (OD) is any disease that affects a small percentage of the population. While opportunities now exist to accelerate progress toward understanding the basis for many more ODs, the prioritization of candidate genes is still a critical step for disease-gene identification. Several network-based frameworks have been developed to address this problem with varied results.

Result: We have developed a novel vertex similarity (VS) based parameter-free prioritizing framework to identify and rank orphan disease candidate genes. We validate our approach by using 1598 known orphan disease-causing genes (ODGs) representing 172 orphan diseases (ODs). We compare our approach with a state-of-art parameter-based approach (PageRank with Priors or PRP) and with another parameter-free method (Interconnectedness or ICN). Our results show that VS-based approach outperforms ICN and is comparable to PRP. We further apply VS-based ranking to identify and rank potential novel candidate genes for several ODs.

Conclusion: We demonstrate that VS-based parameter-free ranking approach can be successfully used for disease candidate gene prioritization and can complement other network-based methods for candidate disease gene ranking. Importantly, our VS-ranked top candidate genes for the ODs match the known literature, suggesting several novel causal relationships for further investigation.

Background

In the USA, a rare or orphan disease (OD) is defined as a disease that affects fewer than 200,000 inhabitants [1]. According to an estimate, there are as many as 8000 ODs, many of which are known to be of genetic origin, affect children at a very early age and are life-threatening and/or chronically debilitating [2,3]. Although, the advent of next-generation sequencing technologies accelerates the disease gene discovery pipeline, the prioritization of candidate genes is still a critical step for disease-gene identification [4]. We [5], and several other earlier studies [6-9], have shown that genes associated with phenotypically close disorders tend to share molecular signatures which include similar expression profiles, participation in the same biological processes or pathways, protein interactions or complexes, literature co-citation. We have recently completed a global analysis of all ODs that have at least one known mutant gene associated (data from Orphanet [10] and the OMIM databases [11]) and show that the relationship between ODs cannot be fully captured by the gene-based network alone. Integrating diverse biomedical and genomic data types can facilitate hypotheses synthesis about disease causing mutant genes. Additionally, it can help in addressing an important question, namely, are there any candidate genes related to known causal genes for a disease? A useful way to approach this question is to rank the genes in a test set based on their similarity to a reference or ‘seed’ set. Such a “guilt by association” ranking approach has become an important way to prioritize candidate disease genes, such as the candidates found in genome-wide association or linkage studies [12]. The genes within a locus shown to be linked to a particular disease, for example, can be prioritized based on their similarities to a reference set of known genes for that disease. We and
Network-based analyses have been equally successful in the identification and prioritization of disease candidate genes [6,7,24-31] especially where the genes are relatively less annotated. Network-based candidate gene ranking approaches can be broadly grouped into two categories: parameter-based and parameter-free methods. The parameter-based methods, such as PageRank with Priors (PRP [28]), Random Walk (RW [27]) and PRInoritization and Complex Elucidation (PRINCE [7]), usually require additional auxiliary parameters that need to be trained by using available data sets. The PRP for instance needs a parameter $\beta$ to control the probability of jumping back to the initial node [28], and the PRINCE algorithm uses a parameter to describe the relative importance of prior information [7]. Since selecting optimal parameters could be a challenge, parameter-free approaches are preferred and considered as more user-friendly [29]. Additionally, most parameter-based approaches take into account the global information in the entire network which often requires extensive computation. For example, in PRP, scores of all the nodes need to be updated iteratively until they converge. This process typically becomes extremely slow and inefficient especially when the network size is large. The parameter-free methods (e.g. Interconnectedness or ICN [29]), on the other hand, measure closeness of each candidate gene to known disease genes by taking into account direct link and the shared neighbors between two genes and tend to be less intensive computationally. The performance of parameter-free methods however is usually not comparable to parameter-based ranking approaches. Here, we report a novel network-based parameter-free framework for discovering and prioritizing candidate orphan disease genes. We specifically focus on two aspects: a) enhance prioritizing performance compared to current parameter-free methods and b) achieve a comparable performance to the parameter-based ones. We test, in a leave-one-out cross-validation setting, the utility of our approach in prioritizing genes for 172 ODs with at least five known causal genes (from Orphanet database [10]). We compare the performance of our method to two approaches, one each from parameter-based and parameter-free methods. To demonstrate the utility of our approach, we rank the immediate neighbors of known OD genes as potential novel candidate genes. The immediate neighboring gene sets were compiled using (a) protein interactions; (b) functional linkage network [32,33]; and (c) literature co-citations.

**Results and discussion**

**Vertex similarity (VS) based candidate gene ranking**

Hypothesizing that genes that are connected to one or more known disease genes (“seed genes”) are also probably implicated in the same disease, our goal is to find such novel candidate genes with “strong” associations to the seed genes. Our proposed VS-based candidate gene ranking approach is based on guilt-by-association principle. Two nodes or vertices are considered similar if their immediate neighbors in the network are themselves similar (common biological process, pathway, etc.). This principle is used to build a self-consistent matrix formulation of functional similarity that can be evaluated iteratively using only knowledge of the adjacency matrix of the network (based on functional annotations of genes). To this effect, we consider similarity between two vertices (genes) as a measure of their association strength in a network. Thus, two vertices with a high similarity are likely to be strongly related. In order to find the similarities between the seed and the candidate or test set genes, we introduce a vertex similarity measurement in our algorithm. Vertex similarity which defines the similarity of two vertices based on the structure of network has been used for information retrieval in World Wide Web [34] and in social network analysis [35]. Similarity measurements, such as cosine similarity, have been successfully applied for computing similarity between documents which are described as vectors of keywords [36]. However, to the best of our knowledge, there have been no reports of using it as a measure to compute similarity between two genes in a functional network and use it for ranking candidate disease genes.

In our approach, when two genes are connected, the gene vectors are constructed based on protein interactions with the other neighboring genes. The shortest path is considered in cases where two genes are not directly connected. As illustrated in Figure 1, the similarity score $\text{Sim}(A, B)$ between two genes A and B is defined as:

$$\text{Sim}(A, B) = \frac{\sum_{i=1}^{n} \omega_{A,i} \times \omega_{B,i}}{\sqrt{\sum_{i=1}^{n} (\omega_{A,i})^2} \times \sqrt{\sum_{i=1}^{n} (\omega_{B,i})^2}}$$  \hspace{1cm} (1)$$

where $\omega_{A,i}$ represents the edge weight of node A to node i, and we define $\omega_{A,A} = \omega_{B,B} = 1$ (protein interactions); $n$ is the number of the nodes which includes A, B and all the nodes that are directly associated with A and B. Equation 1 applies only when nodes A and B are connected and hence the value of $n$ can be derived by

$$n = \Gamma_A + \Gamma_B - \sigma_{shared}$$  \hspace{1cm} (2)$$

where $\Gamma_A$ and $\Gamma_B$ represent the degree (number of connections or edges the node has to other nodes) of nodes A and B respectively, and $\sigma_{shared} = | \Gamma_A \cap \Gamma_B |$.

Others have developed several computational approaches which perform this task automatically [4,13-23].
and represents the number of shared neighbor nodes by both A and B. When there is no direct connection between node A and node B, we try to find the shortest path between them. In this case, the similarity score will be derived by

$$Sim^*(A, B) = \begin{cases} \prod_{k=1}^{K} Sim(C_k, C_{k+1}) & \text{if } K \leq r \\ 0 & \text{otherwise} \end{cases}$$ (3)

where $C_k$ is the node on the shortest path of A and B, and $r$ is the discovery range that controls the maximum degree of separation (maximum $r$ hops). In other words if the shortest path length between nodes A and B is more than $r$ hops or if there is no shortest path between them, $Sim(A, B)$ equals to 0.

The candidate genes in the test set are prioritized based on the similarity scores calculated from equation 1 and equation 3. For example, for a given disease $d$, each candidate gene is scored by summing up the similarity scores between the candidate gene and each of the seed genes from the training or seed set $S_d$. The score of a candidate gene $i$ is calculated as:

$$score_i = \sum_{j \in S_d} Sim(i, j)$$ (4)

where $Sim(i, j)$ is the connection score between gene $i$ and $j$. All candidate genes are then ranked based on these scores.

**Comparison with other network-based prioritization algorithms**

To compare the performance of our VS-based approach in candidate disease gene ranking, we select two methods, one each from parameter-based and parameter-free methods: PageRank with priors (PRP) [28] and Interconnectedness (ICN) [29]. Parts of implementation of PRP are done using JUNG (Java Universal Network/Graph; jung.sourceforge.net) framework [37] as described earlier [28]. To evaluate the performance of VS-based approach and compare it with two other methods, we used a leave-one-out cross-validation procedure. In each cross-validation trial, we removed a single OD causal gene ("target gene") from the data, and each of the 3 algorithms was evaluated by its success in assigning the rank to the "target gene" (see Methods for additional details).

We selected 172 ODs (ODs with 5 or more known causal genes) and 1598 OD causal genes for the cross-validation runs. Of the 1598 genes, we used 1312 which were in the protein interactome. The results from the leave-one-out cross-validation using the three approaches are presented in Figure 2. As can be seen...
from the Figure 2, when k (rank cut-off) = 1, both VS-based (parameter-free) and PRP (with back probability set to 0.3; parameter-based) methods, achieved the best performance with a success rate of ~43%. In other words, the target gene was top ranked in 568 out of 1312 cases (43.3%) using VS-based method. On the other hand with PRP (0.3 back probability), the target gene was top ranked 559 times out of a total 1312 cases (42.6%). ICN, another parameter-free method showed a lower success rate at 35.3% with the target gene ranked at top 463/1312 times. Expectedly, PRP (with back probability set to 0.05) showed a lower success rate than VS at 37.2% (488/1312) but was better than ICN.

When we increased the rank cut-off (k), VS-based approach performed equally well as PRP0.3. Additionally, compared to ICN, another parameter-free method, our VS-based approach performed better. We also note that VS outperformed PRP too when the back probability was set to 0.05.

The improved performance of VS over ICN we believe is because of the “extended guilt by association” [38] principle on which VS is based on. For example, if we consider a simple unweighted network (Figure 3; all edges equal to 1) where nodes A and B do not have a common neighbor and the shortest path connecting them is A-C-D-B. In this case, the ICN [29] score would be 0 because there is not even a single shared node between A and B. However, using VS, we can calculate the similarity between A and B (Sim(A, B) = Sim(A, C) * Sim(C, D) * Sim(D, B) = 0.276). Although, we have not performed an extensive analysis on disease gene connectivity, for the examples we have analyzed, we have found that several causal genes of a specific disease are connected indirectly (e.g., 3-step away).

However, since biological networks tend to have low diameters [39], we believe that low values of the steps/hops are preferable. Interestingly, a previous study provided examples of two real data applications where the number of hops or steps between disease causal genes (m) were set to two and reported that m = 2 was preferable over m = 1 [40]. Since the edge information between two genes may be noisy or incomplete, we believe that our VS-based approach for novel candidate disease gene ranking is desirable as it takes into account alternative measures of pairwise interconnectedness and is not just limited to direct interactions or having a shared neighbor node.

Identifying and ranking novel OD candidate genes with VS-based approach

Having validated our method, we proceeded to execute our algorithm on several ODs with the goal of identifying and ranking potential novel candidate genes for ODs. We ranked candidate genes over the entire protein-protein interaction (PPI) network, and analyzed our top-five predictions for ten select ODs which have known protein interactions for all of their causal genes. The test set genes were compiled using several different sources comprising
protein interactions, functional relatedness and literature co-citations. Briefly, for each causal gene of an OD, we extracted the immediate neighboring genes from the above mentioned resources (see Figure 4 and Methods for additional details). Table 1 shows the top-five predictions for ten ODs. We checked whether our VS top-ranked genes were already found to be associated with their query OD by searching online databases and scientific publications and found that most of the top candidate genes were already related to the respective OD. For example, the top five predictions for cone rod dystrophy are CRB1, RDH5, USH1C, EFEMP1, CABP4, which are genes associated with visual perception (Gene Ontology) and two of them (CRB1 and CABP4) are known to be involved in eye photoreceptor cell development and differentiation [41-43]. For this particular example, we also performed candidate gene ranking using PRP (with back probability set to 0.3) and ICN. We also used a functional annotation based candidate gene ranking method (ToppGene [16]) for ranking. When we compared the twenty top ranked genes from each of these three methods with VS-based ranking, there were five genes (CRB1, EFEMP1, NPHP4, CNGB1 and GUCA1B) common to all (Figure 5).

Among other examples, HESI, the top ranked gene for Fanconi anemia is a novel interacting protein of the Fanconi anemia core complex and cells depleted of HESI exhibit a Fanconi anemia-like phenotype [44]. The two top-ranked genes for gonadal dysgenesis, ZFX and ZFY, are known to function in sex differentiation and Zfx mutant mice are reported to have fewer germ cells than wild-type mice [45]. Likewise, maturity-onset diabetes of the young type (MODY syndrome) is linked to kinetic alterations and regulation of glucokinase activity [46,47] and in our ranking glucokinase receptor is the top ranked gene for MODY syndrome. Interestingly, a recent study in the Japanese families proposes GCKR as a susceptibility gene for familial diabetes [48]. While our ranking provides further support for the involvement of the top-ranked ranked genes in the investigated ODs, it also suggests that the top scoring candidates that are not previously associated with these ODs could be potential candidates for further research.

Conclusion

The vertex similarity method (VS) is parameter-free approach for prioritizing candidate disease genes, where it calculates the similarity between nodes other than updating and training the parameters and data sets in every step. Through cross-validation experiments we show that VS outperforms ICN, another parameter-free method and that it is comparable to parameter-based methods such as PRP. We demonstrate the utility of VS-based parameter-free ranking approach in ranking OD candidate genes and importantly, these top ranked candidate genes for the ODs match the known literature, suggesting several novel causal relationships for further investigation.

Our approach however has some limitations. First, as with any training set dependent candidate gene ranking approaches, we assume that the OD causal genes we have yet to discover will be consistent with what is already known about an OD and/or its genetic basis which may...
Figure 4 Workflow for identifying and ranking novel OD candidate genes using VS.

Table 1 Examples of orphan diseases and VS-ranked top 5 candidate genes

| Orphan disease                                             | No. of known causal genes | VS ranked top 5 candidate genes                  |
|------------------------------------------------------------|---------------------------|-------------------------------------------------|
| Cone rod dystrophy                                        | 20                        | CRB1, RDH5, USH1C, EFEMP1, CABM                 |
| Severe combined immunodeficiency                           | 17                        | CD3G, JAK1, ZAP70, IL2RB, IL4                   |
| Fanconi anemia                                             | 15                        | HES1, SAMD3, CYP19A1, XRCC3, USP1               |
| Zellweger syndrome                                        | 14                        | PEX7, PHEX, ABCD2, ABCD1, ABCD3                |
| Autosomal dominant Charcot-Marie-Tooth disease, type 2     | 12                        | STAT4, FAIM, MARCH5, STAT6, CRYGC              |
| Gonadal dysgenesis                                         | 12                        | ZFY, ZFX, PTC12, SOX9, AMH                      |
| Hereditary nonpolyposis colon cancer                       | 11                        | MRC1, MSH3, CARKD, TRIT1, EXO1                 |
| Papillary or follicular thyroid carcinoma                   | 11                        | CORO2A, ZBTB33, KIF11, AAS5, SEH1L              |
| Romano-Ward syndrome                                       | 11                        | KCNE3, MINK1, KCNJ3, ALG10B, KCNJ9             |
| MODY syndrome                                              | 10                        | GCKR, IDDM7, MAFA, ST6GAL1, INSRL               |
not always be the case. Additionally, this also means that our approach currently cannot be used to rank novel candidate OD genes if an OD lacks known causal genes. Similarly, even if an OD has known causal genes but if there is no protein interactome data available then we cannot use VS for such cases. An alternative approach would be to consider other types of networks (coexpression or functional networks). Second, it is important to note that the prioritization by our approach can only be as accurate as the current protein interactome data are. Third, if a seed gene has only one known interaction then that interactant will be ranked higher.

Methods

Data resources

The ODs and causal gene information was downloaded from Orphanet [10]. We merged some of the OD subtypes of a single disease based on their given disorder names as described previously [5,8]. From this, we selected 172 ODs that have at least five causal genes. The total number of genes across 172 selected diseases was 1598. The human protein interactome used in this study was compiled from several resources [49-54] with both redundant interactions and self-loops removed. The assembled PPI network consists of 11,765 proteins and 69,167 interactions. During each set of a validation trial, one seed gene ("target gene") from one of the selected 172 ODs was picked out and mixed with 99 random genes from PPI network to form a test set of 100 candidate genes. The remaining seed genes of an OD were used as the training set. The test set genes were then prioritized using the three approaches: PRP (with back probabilities 0.3 and 0.05), ICN, and VS-based approach. During each run, the rank of the "target gene" was noted. We evaluated the performance of each algorithm in terms of the success rate versus rank cut-off (k). If the "target gene" is ranked among the top k in a particular validation run, it is considered as a 'success'. The validation runs are repeated until all the seed genes have been used as the target gene and their ranks are obtained. The "success rate" is defined as the ratio of successful validation runs and the total validation runs for all the existing OD genes from 172 ODs. The same strategy was followed for all the three algorithms. In case of PRP which is a parameter-based method, we selected a back probability of 0.3 since we have shown previously that the performance of PRP in ranking candidate disease genes was best at p = 0.3 [28].

Figure 5 Venn diagram comparing the top 20 ranked candidate genes for cone rod dystrophy using PRP (0.3), ICN, VS, and ToppGene.

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http://www.biomedcentral.com/1752-0509/6/S3/S8
Test set genes for identifying and ranking OD candidate genes

For identifying and ranking novel OD candidate genes, we used the immediate neighbors of known OD genes as the test set. The immediate neighboring genes of selected ODs’ causal genes were compiled based on (a) protein interactions; (b) functional linkage network [32,33]; and (c) literature co-citations. The protein interactome data as described earlier was compiled from several resources [49-54]. The functional linkage network-based candidate gene sets were derived from two resources: (i) HumanNet, a probabilistic functional gene network of Homo sapiens [33] and (ii) functional protein interaction network built upon expert-curated pathways [32]. The test set genes based on literature co-citations were compiled using the OMIM database. Briefly, for the selected ODs, we identified the corresponding OMIM records, which summarize results from publications about gene-disease relationships. For the OD mapped OMIM mapped records, we first extracted the cited literature (links to PubMed records for the references cited in an OMIM entry) in the OMIM records. Using this OD-related PubMed records, we extracted the related genes from the ‘gene2pubmed’ file from NCBI [55]. For a given OD with known causal genes, we pooled all neighboring genes (immediate neighbors or direct interactants) of causal genes from different sources and used it as a test set for ranking in the global protein interactome using VS-based approach.

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Authors’ contributions

CZ, KB and AJ conceived the study design which was coordinated by AJ. CZ designed and implemented the VS-based candidate gene ranking approach and along with AJ and AK participated in the analysis and interpretation of results. CZ and AJ drafted the manuscript. All the authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Dear IWP, Webb DJ. Are rare diseases still orphans or happily adopted? The challenges of developing and using orphan medicinal products. Br J Clin Pharmacol 2006, 62(3):264-271.
2. Schieppati AH, Daina E, Aperia A: Why rare diseases are an important medical and social issue. Lancet 2008, 371(9629):2039-2041.
3. Stolk P, Willemen MJ, Leufkens HG: Rare essentials: drugs for rare diseases as essential medicines. Bull World Health Organ 2006, 84(9):745-751.
4. Firo RM, Di Cunto F: Computational approaches to disease-gene prediction: rationale, classification and successes. FEBS J 2012.
5. Zhang M, Zhu C, Jacomy A, Lu LJ, Jegga AG: The orphan disease networks. Am J Hum Genet 2011, 88(6):755-766.
6. Wu X, Jiang R, Zhang MQ, Li S: Network-based global inference of human disease genes. Mol Syst Biol 2008, 4:189.
7. vanunu O, Nagger O, Ruppin E, Shlomi T, Sharan R: Associating genes and protein complexes with disease via network propagation. PLoS Comput Biol 2010, 6(11):e1000641.
8. Goh KI, Cusick ME, Valle D, Childs B, Vidal M, Barabasi AL: The human disease network. Proc Natl Acad Sci USA 2007, 104(21):8685-8690.
9. Feldman I, Rzhetsky A, Vitkup D: Network properties of genes harboring inherited disease mutations. Proc Natl Acad Sci USA 2008, 105(11):4323-4328.
10. Ayme S: [Orphanet, an information site on rare diseases]. Orphanet J Rare Dis 2003, 672:46-47.
11. Hamosh A, Scott AF, Amberger J, Valle D, McKusick VA: Online Mendelian Inheritance in Man (OMIM). Hum Mutat 2000, 15(1):57-61.
12. Hardy J, Singleton A: Genomic variation studies and human disease. N Engl J Med 2009, 360(17):1759-1768.
13. Adele EA, Adams RR, Evans KL, Porteous DJ, Pickard BS: Speeding disease gene discovery by sequence based candidate prioritization. BMC Bioinformatics 2005, 655.
14. Adele EA, Adams RR, Evans KL, Porteous DJ, Pickard BS: SUSPECTS: enabling fast and effective prioritization of positional candidates. Bioinformatics 2006, 22(8):773-774.
15. Aerts S, Lambrechts D, Maity S, Van Loo P, Coessens B, De Smet F, Tranchevent LC, De Moor B, Marynen P, Hassan B, Carmeliet P, Moreau Y: Gene prioritization through genomic data fusion. Nat Biotechnol 2006, 24(5):537-544.
16. Chen J, Barden EE, Aranow BJ, Jegga AG: ToppGene Suite for gene list enrichment analysis and candidate gene prioritization. Nucleic Acids Res 2009, 37 Web server: W305-311.
17. Chen J, Xu H, Aranow BJ, Jegga AG: Improved human disease candidate gene prioritization using mouse phenotype. BMC Bioinformatics 2007, 8 392.
18. Freudenberg J, Prapping P: A similarity-based method for genome-wide prediction of disease-relevant human genes. Bioinformatics 2002, 18(Suppl 2):i110-i115.
19. Thornblad TA, Elliott KS, Jowett J, Visscher PM: Prioritization of positional candidate genes using multiple web-based software tools. Twin Res Hum Genet 2007, 10(6):861-870.
20. Tiffin N, Adele E, Turner F, Brunner HG, van Driel MA, Oti M, Lopez-Bigas N, Ouzounis C, Perez-iraneta C, Andrade-Navarro MA, Adeyemo A, Patti ME, ToppGene Suite for gene list enrichment analysis and candidate gene prioritization. Nucleic Acids Res 2006, 34(10):3067-3081.
21. Tiffin N, Kelso JF, Powell AR, Pan H, Bajic VB, Hide WA: Integration of text- and data-mining using ontologies successfully selects disease gene candidates. Nucleic Acids Res 2005, 33(Suppl 1):1544-1552.
22. Turner FS, Clutterbuck DR, Semple CA: POCUS: mining genomic sequence annotation to predict disease genes. Genome Biol 2003, 4(11):R75. 23. Zhu M, Zhao S: Candidate gene identification approach: progress and challenges. International journal of biological sciences 2007, 3(7):420-427.
24. Sam L, Lu Y, Li J, Friedman C, Lussier YA: Discovery of protein interaction networks shared by diseases. Pacific Symposium on Biocomputing 2007, 76-87.
25. Goehler H, Lalowski M, Stelzl U, Waelter S, Stroedicke M, Worm U, Bussow K, Coleman SH, Gutkunst CA, Landwehrmeier BG, Lehrach H, Wanker EE: A protein interaction network links G711, an enhancer of huntingtin aggregation, to Huntington’s disease. Molecular cell 2004, 15(6):853-865.
26. Lage K, Karliberg EO, Stirling ZM, Olson PI, Pedersen AG, Rigna O, Hinsby AM, Turner Z, Pociot F, Tommerup N, et al: A human phenome-interactome network of protein complexes implicated in genetic disorders. Nat Biotechnol 2007, 25(3):309-316.
27. Kohler S, Bauer S, Horn D, Robinson PN: Walking the interactome for prioritization of candidate disease genes. Am J Hum Genet 2008, 82(4):949-958.

28. Chen J, Aronow BJ, Jegga AG: Disease candidate gene identification and prioritization using protein interaction networks. BMC Bioinformatics 2009, 10(73).

29. Hsu C, Huang Y, Hsu C, Yang U: Prediction of human disease-related gene clusters by clustering analysis of network data using JUNG. Journal of Computer Software 2005, 10(2):135.

30. Chen X, Yan GY, Liao XP: Prioritizing disease candidate genes by network-based boosting of genome-wide association data. Genome Res 2011, 21(7):1109-1121.

31. Kleinberg J: Authoritative sources in a hyperlinked environment. Journal of the ACM (JACM) 1999, 46(5):29.

32. Leicht EA, Holme P, Newman MEJ: Vertex similarity in networks. Physical Review E 2006, 73(2).

33. Lee I, Blom UM, Wang P, Shm J, Marcotte EM: Clustering on the unit hypersphere using von mises-sher distributions. Journal of Machine Learning Research 2005, 6:1345-1382.

34. Madadhain J, Fisher D, Smyth P, White S, Boey Y: Gene network interconnectedness and the disease genes by network-based boosting of genome-wide association data. SIAM Review 2003, 45(2):167-256.

35. Yip AM, Horvath S: Gene network interconnectedness and the generalized topological overlap measure. BMC Bioinformatics 2007, 8:22.

36. Littink KW, Koeneekoop RW, van den Born LI, Collin RW, Moruz L, Veltman JA: Homozygosity mapping in patients with cone-rod dystrophy: novel mutations and clinical characteristics. Invest Ophthalmol Vis Sci 2010, 51(11):5943-5951.

37. Pellikka M, Tanentzapf G, Pinto M, Smith C, McGlade CJ, Ready DF, Tepass U: The role of indirect connections in gene networks in developmental biology. OMICS 2008, 12(7):293-308.

38. Abraham C, Bock N, Kiertzmann S, Goedde A, Tokesz E, Drogea E, Krotsch B, Kom B, Birchmeier W, Lehrach H, Wanker EE: A human protein-protein interaction network: a resource for annotating the proteome. Cell 2005, 122(6):957-968.

39. Rual JF, Venkatesan K, Hao T, Hirozane-Kishikawa T, Dricot A, Li N, Berriz GF, Gibbons FD, Dreze E, Ayivi-Guedehoussou N, Kitgord N, Simon C, Boxem M, Milstein S, Rosenberg J, Goldberg DS, Zhang LV, Wong SL, Franklin G, Li S, Albona LS, Lim J, Faughton CL, Llanosavas E, Cevik S, Rex B, Lamesch P, Skorski RS, Vanderhaute J, Zoghbi HY, Smolyar A, Bosak S, Sequerra R, Doucette-Stam L, Sussick ME, Hill DE, Roth FP, Vidal M: Towards a proteome-scale map of the human protein-protein interaction network. Nature 2005, 437(7062):1173-1178.

40. Ramani AK, Bunesch RC, Mooney RJ, Marcotte EM: Consolidating the set of known human protein interactions in preparation for large-scale mapping of the human interactome. Genome Biol 2005, 6(5):R40.

41. Spiridov V, Aronow BJ, Jegga AG: Human Protein Reference Database and Human Proteininterpedia as discovery tools for systems biology. Methods Mol Biol 2009, 577:67-79.

42. Zhu et al: A vertex similarity-based framework to discover and rank orphan disease-related genes. BMC Systems Biology 2012, 6(Suppl 3):S8.