Effects of huoxiangzhengqi liquid on enteric mucosal immune responses in mice with Bacillus dysenteriae and Salmonella typhimurium induced diarrhea

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Abstract

AIM: To explore effects of huoxiangzhengqi liquid (HXZQ) on enteric mucosal immune responses in mice with Bacillus dysenteriae and Salmonella typhimurium induced diarrhea (BSD).

METHODS: BSD was induced in Balb/c mice by oral administration with Bacillus dysenteriae and Salmonella typhimurium. HXZQ was administrated from the day of diarrhea induction at dosages of 5.21 g/kg and 0.52 g/kg, respectively. The onset of diarrhea and lasting time were recorded. Peyer’s patches and peripheral lymphocytes were prepared for flow cytometry, and levels of TNF-α in peripheral blood and enteric tissue homogenates were determined with ELISA. Student’s t test was employed for statistics.

RESULTS: Mice in BSD group started showing continuous diarrhea on the day of induction until the fourth day when they were sacrificed. Diarrhea in the mice of HXZQ high and low dose groups lasted for 36 and 54 h, respectively. There were more CD4+ and CD8+ cells in peripheral blood, fewer CD4+ cells in Peyer’s patches in BSD mice compared to normal mice. Fewer CD4+ and CD8+ cells was shown in the mice in HXZQ high group compared to BSD mice. In Peyer’s patch, there were more CD8+ cells in mice in HXZQ high and low dose groups and more CD4+ in mice in HXZQ high group. Higher levels of TNF-α in peripheral blood and intestinal tissue homogenates in BSD group were observed. Mice in HXZQ high group showed decreased levels of TNF-α in peripheral blood and enteric tissue homogenates.

CONCLUSION: The immune regulation of CD4+ and CD8+ cells in Peyer’s patch and suppression of TNF-α levels in enteric homogenates may partially explain the effect of HXZQ on improvement of BSD.

Key words: Diarrhea; Peyer’s patches; TNF-α; Huoxiangzhengqi liquid

INTRODUCTION

Huoxiangzhengqi liquid (HXZQ), consisting of Agastache rugosa, Ammi majus, areca peel, hoelen, beefsteak plant leaf, aurantii nobilis pericarpium, atractylodes macrocephala, magnolia bark, pinelliae tuber, platycodon root and Glycyrrhiza uralensis, has been used in China for thousands of years for the treatment of various diarrhea[1]. It is one of earliest marketed herbal medicines and is produced by China Tongrentang Pharmaceuticals Corp(GMP certificated), Beijing. Pharmacological research has shown that HXZQ has the activities of improving gastrointestinal dysfunction, anti-inflammation and immune regulation[2,3].

Enteric mucosal responses play an important role in anti-inflammation of the digestive tract[4,5]. However, the effects of HXZQ on enteric mucosal immune responses are largely unknown. Dysenteriae and Salmonella typhimurium induced diarrhea (BSD) is one of widely used animal models for
human infectious diarrhea study[6,7]. In the present study, we investigated the effects of huoxiangzhengqi liquid on enteric mucosal immune responses in mice with BSD.

MATERIALS AND METHODS

Animals
Forty Balb/c mice, female and male in equal number, 8-10 wk old, were purchased from the Research Institute of Experimental Animals, Chinese Academy of Medical Science, Beijing. Mice were randomly divided into four groups with 10 in each group: control group (Control), BSD group, Huoxiangzhengqi liquid treated BSD groups at high dosage and low dosage (HXZQ high, HXZQ low). Mice were housed in a temperature-, humidity- and light-controlled environment with free access to rodent chow and water. The light-dark cycle was 12 h:12 h with the light phase from 06:00 am to 18:00 pm.

Induction of BSD
BSD were induced by oral administration with sodium ampicillin (Northern China Pharmaceutical Limited Company, Guangzhou, China) at a dosage of 20 g/kg body weight, and 8 h later, were orally administered with Bacillus dysenteriae and Salmonella typhimurium (National Institute for Control of Drugs and Biological Products, Beijing, China) at the proportion of 1:1 with 0.2 mL/mouse (concentration at 1010 CFU/mL). Mice in BSD, HXZQ high, and HXZQ low groups were induced with BSD, and mice in Control group were treated with equal volume physiological saline. The onset of diarrhea and its lasting time were recorded as previously reported[6,7].

Treatment of BSD with HXZQ
The administrations of HXZQ started from the day of induction of BSD, once a day for three days. The dosage of HXZQ was determined according to our previous study[5]. The remaining cells were then washed in saline twice, and resuspended for flow cytometry.

Preparation of enteric tissue homogenates
Colons were washed and put into homogenate medium (dehydrated alcohol and saline at a ratio of 1:4) at 300 g/L. After mixed sufficiently, they were placed at 4°C for 2 h, and centrifuged at 3500 rpm for 15 min. The supernatant was collected, and stored at 4°C for ELISA.

Flow cytometry for CD4+ and CD8+
Flow cytometry was performed on freshly isolated plasma lymphocytes and Peyer’s patch cell suspension. Immunofluorescence staining of cell surface markers was performed using R-Phycoerythrin (PE)-conjugated OX-38 (BD Biosciences Pharmingen) for CD4+ staining, Peridinin Chlorophyll-a Protein (PerCP)-conjugated OX-8 (BD Biosciences Pharmingen) for CD8+ staining. Each sample of 0.1 mL peripheral lymphocytes or Peyer’s patch cell suspension was incubated for 30 min at 4°C in the dark, with a solution consisting of 6 μL PBS 0.01 mol/L, and an appropriate concentration of antibodies. After washing in saline, the cells were resuspended and fixated. Flow cytometric analysis was performed using an FACS Caliburflow cytomter utilizing CELLQuest software (Becton Dickinson, San Jose, CA, USA). Analysis was carried out on 1 × 10⁶ cells for each sample.

ELISA for TNF-α
Detection of TNF-α in peripheral blood and intestinal tissue homogenates was conducted with ELISA kit (Cat. No.88-7340 ebioscience, Inc) according to instructions of the manuals.

Statistical analysis
Data were expressed as mean ± SD. ANOVA was used to determine significance in the data set. Student-Newman-Keuls test was employed for variables between both groups when equal variances were assumed and Dunnetts’s t test when equal variances not assumed. P < 0.05 was taken as significant.

RESULTS

Incidence and lasting time of diarrhea
After induction of BSD, all mice showed diarrhea within 6 h on the day of induction. The diarrhea in the mice in BSD group lasted until the fourth day when the mice were sacrificed. Diarrhea lasting time of the mice in HXZQ high and HXZQ low groups were 36 and 54 h, respectively.

Changes of CD4+ and CD8+ cells in peripheral blood and Peyer’s patch
Figure 1 shows that there were more CD4+ and CD8+ cells in BSD mice in periphery (P < 0.05 vs control group), less CD4+ and more CD8+ cells in Peyer’s patch (P < 0.05 vs control group). Less CD4+ and CD8+ cells showed in the mice in HXZQ high group (P < 0.05 vs BSD group), and less CD8+ cells in HXZQ low group (P < 0.05 vs BSD
group) in periphery. In Peyer’s patch, there were more CD8⁺ cells in mice in HXZQ high and low groups (P < 0.05 vs BSD group), and more CD4⁺ in mice in HXZQ high group (P < 0.05 vs BSD group).

**TNF-α in peripheral blood and intestinal tissue homogenates**

Table 1 shows that there were higher levels of TNF-α in peripheral blood and intestinal tissue homogenates in BSD group (P < 0.05 vs control group). Mice in HXZQ high group showed decreased levels of TNF-α in peripheral blood and enteric tissue homogenates (P < 0.05 vs BSD group), and mice in HXZQ low group showed decreased levels of TNF-α in enteric tissue homogenates (P < 0.05 vs BSD group).

**DISCUSSION**

HXZQ is one of ancient compound liquids of traditional Chinese herbal preparations. In China, it has been used for the effective treatment of various common gastrointestinal infectious diseases (GID), such as acute gastroenteritis, diarrhea and gastrointestinal type Frigorism[2]. It has been reported that HXZQ can promote gastrointestinal motility, ameliorate myoelectricity of enteric smooth muscle and regulate gastrointestinal function[2]. Huoxiangzhengqi liquid has also been shown to have regulatory effects on specific immune responses[3]. Agastache rugosa is the essential component of huoxiangzhengqi liquid, and the extract of Agastache rugosa has been reported to have immune modulatory activities[3, 4]. BSD in mice is induced by administration of attacking pathogenic bacteria, and shares similar pathogenesis and symptoms with GID[6, 7]. Our study showed that HXZQ at 5.21 g/kg and 0.521 g/kg could significantly ameliorate diarrhea in BSD mice.

Although bacteria are usually regarded as harmful, it is now recognized that the breakdown or a failure of components of the indigenous microflora is responsible for allergic diseases or food enteropathies[6-10]. Intestinal microbial species and its product can partially modulate mucosal immune responses and influence the expansion and structure of lymphoid tissues, the activation of TH1 and TH2 T cells[11, 12]. Intestinal bacteria are required for the development of gut-associated lymphoid tissues (GALT), which mediate a variety of host immune functions, such as mucosal immunity and oral tolerance[13]. Peyer’s patch lymphocytes, as one important part of GALT, are considered to be the effectors in reacting with intestinal antigens, and mediate immune tolerance[14]. It has been known that some drugs can reestablish the microenvironment of the intestinal tract, thereby enhancing mucosal as well as systemic immune response[15-17]. Intestinal immunity has been evaluated with respect to CD4⁺ and CD8⁺ T cells both in Peyer’s patch and peripheral blood in BSD mice, and HXZQ could regulate the proportion of CD4⁺ and CD8⁺. These results further supported that dysbacteria might take some active role in immune response in BSD mice, and HXZQ might have potential regulative activities on enteric mucosal immune responses in diarrhea. The results also indicate that CD4⁺ and CD8⁺ T cells in Peyer’s patch may serve as one of the targets of the immunoregulatory activity of huoxiangzhengqi liquid.

**TNF-α** is a pleiotropic cytokine with strong proinflammatory, immunomodulatory properties, and plays a critical role in inflammation and inflammatory bowel disease[18, 19]. Anti-TNF therapy has been proven to be a milestone in the treatment of inflammatory bowel disease, and equally important in other inflammation-mediated conditions, including viral infections and mucosal inflammation[20, 21]. Different strategies have been explored aiming at inhibiting TNF. Infliximab, a chimeric antibody to TNF-α, holds much promise for the treatment of Crohn’s disease[22]. TNF-α levels in enteric tissue homogenates and peripheral blood in BSD mice were higher in our study, while HXZQ could decrease TNF-α. The results could partially explain the immunosuppressive activities of HXZQ in immune response of mice, and further support that dysbacteria could lead to increased inflammatory response in BSD mice.

**Table 1 Changes of TNF-α in peripheral blood and enteric tissue homogenates**

| Group     | Dosage (g/kg) | Peripheral blood (ng/mL) | Enteric tissue homogenates (ng/mL) |
|-----------|---------------|--------------------------|-----------------------------------|
| Control   | -             | 2.24 ± 0.26              | 1.83 ± 1                          |
| BSD       | 10            | 2.76 ± 0.26              | 4.13 ± 1.73                       |
| HXZQ high | 10            | 2.31 ± 0.27              | 2.00 ± 0.53                       |
| HXZQ low  | 0.521         | 2.64 ± 0.54              | 2.7 ± 1.53                        |

Data represent mean ± SE. *P < 0.05 vs control group, †P < 0.05 vs BSD group. TNF-α in peripheral blood and enteric tissue homogenates was analyzed with ELISA.
In conclusion, enteric immune response might be actively involved in diarrhea and regulation of CD4 and CD8 T cells in Peyer’s patch and suppression of TNF-α levels in enteric homogenates might be one of the mechanisms of HXZQ in the treatment of diarrhea.

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