Antitoxic, Antifungal and Phytochemical Analysis of Medicinal Compounds of Guiera senegalensis Leaves

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ABSTRACT

Millions of people in the world depend on traditional medicinal plants for treatment of many diseases. They have curative properties due to presence of various complex chemical substances of different composition, which are found as secondary plant metabolites in one or more parts of these plants. These plant metabolites according to their composition are grouped as flavonoids, tannins, alkaloids, saponins etc. Guiera senegalensis (Gs), which grows in abundance in semi-desert area of Western Africa such as Nigeria, has been used for treating specific diseases and wounds. The study is carried out to investigate the phytochemical analysis, toxicity, and the antifungal activity of Gs leaves extract.

Keywords: Guiera senegalensis, Antifungal activity, Brine shrimp, toxicity, phytochemical analysis;

INTRODUCTION

Guiera senegalensis is called Sabara* (Hausa), [1]. It is a shrubby and can grow to a height of 3 to 5 m depending on the habitat [2]. The leaves which are 3 to 5 cm long and 1.5 to 3.0 cm broad are arranged opposite or sub opposite on the stem [3]. It is widely distributed in the savannah region of west and central Africa, Nigeria, Senegal, Gambia, Mali, Niger, Burkina Faso and Ghana, [4-6]. It is active against cough, respiratory congestion and fever [7,8], and is prescribed as an antitussive, hypertension and hypotension as well as venereal diseases [7], to ease breathing and to treat lung and bronchial disorders. It is also used against malaria fever [9].

Botanical Description

Kingdom: Plantae, Division: Magnoliophyta, Class: Magnoliopsida, Order: Myrtales, Family: Combretaceae, Genus: Guiera Species: senegalensis

D. Morphology

Guiera senegalensis is very well known in its native area, generally occurs as a shrub that can grow to a height of 3 to 5 m according to habitat. Its stem presents numerous knots that send out branches. The ash-grey stem and branches have fibrous or pubescent bark and bear opposing, short petiolated oval leaves, sometimes mucronate, sometimes even coriaceous at their base, about 2 to 4 cm long by 1 to 2 cm wide. These grey-green leaves, darker on their upper surface, display black spots on their lower surface and are slightly downy on both sides. These features lend the plant an overall silver green color that is conspicuous in brushland [2]. Flowering occurs almost throughout the year, when it is leafy. Often blooms twice a year, during the dry season and the rainy season. Each flower has a calcinal tube ovoid, welded to the ovary. This tube is topped by a bellflower blade with 5 teeth screened black and persistent points to fruiting. The liguliform corolla is composed of 5 petals also riddled with black spots. The stamens are 10 on two rows of 5, all inserted on the calyx. The ovary has a single box containing 4 to 6 eggs [11].

Habitat and Repartition

It is a plant that grows primarily in Sudanian Sahel area, on soils sandy, leached or exhausted, fallow and dry stations. Plant pioneer disseminated by cattle in the fallow land, it is also indicative of overgrazing. It is found from Nigeria, Senegal to Cameroon to Sudan. Widespread and common, locally gregarious and very abundant [12].

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METHODOLOGY

Plant materials
The leaves of *Gs* used in this study were previously collected from *Gs* bushes around Bakura village area of Zamfara State, Nigeria. The plant leaves were dried in the shade. After drying, the leaves were ground well into fine powder using a mechanical blender, and the powder was transferred into airtight containers with proper labeling for future use.

Water extraction
Five grams of the plant powder was placed in a beaker containing 100 ml distilled water; soaked and shaken well. The solution was filtered with the help of filter paper and the filtrate was kept and used for further phytochemical analysis. The extracts were then kept in sterile bottles, under refrigerated conditions, until further use.

Phytochemistry
Earlier works showed that leaves and roots of *G. senegalensis* contained Harman, tetrahydroharman, harmalain and Guieranone A [1]. The plant has been found to contain carboxydrates, steroids, flavonoids, saponins, alkaloids, tannins and mucilage [24, 26]. Ficarra et al. in 1997 found four flavonoids in the leaves of *G. senegalensis*, namely catechin, myricitrin, rutin and quercetin [13].

Some findings on the plant elsewhere showed the presence of alkaloids (Hyoscyamin I and solanine, II), tannins, terpenoids menthol, coumarins, saponins, flavonoids (quercetin, III), cardiotonics and cyanogen heterosides which were assayed in various organs of the plant leaves, stem bark, fruits and roots [12]. Hyoscyamin I, solanine II and quercetin III are represented in figure 2. While in the gall of the plant from Ouagadougou, Burkina Faso, alkaloids, polyphenols and saponins were detected [23]. The ashes are poor in alkali but rich in alkaline earth metals. We found especially Mg, Ca, Sr, Ti, Fe, Al and in lesser amounts sometimes traces: Cu, Ni, Co, Zn [11]. Previous studies indicated the presence in leaves of two alkaloids (harman, tetrahydroharman or ecleagnine), flavonoids, naphthopyrans, tannins, and a naphthyl butenone (guieranone A); in roots, were only obtained tannins and the same beta-carboline than in leaves [9, 11, 29, 30, 31, 32, 33]

Antifungal test
Antimicrobial activity testing was carried out by using agar spread method. One ml of the *Gs* extract (5%) was spread to each one of seven (PDA) that were cultured with *Phytophthora capsici*, *Aspergillus flavus*, *Trichoderma viride*, *Pseudomonas aeruginosa*, *Cladosporium cladosporioides*, *Fusarium solani*. The petri dishes were incubated for seven days at room temperature. At the end of incubation, zone of inhibition that developed were measured with a ruler. Distilled water was used as a control.

Test for flavonoids
For the confirmation of flavonoids in the *Gs* leaf powder, 0.5 g of plant extract was placed in a test tube and 10 ml of distilled water was added, 5 ml of dilute ammonia solution were added to a portion of the aqueous filtrate of the plant extract followed by addition of 1 ml concentrated H2SO4. Indication of a yellow color shows the presence of flavonoids in each extract [34].

Test for Alkaloids
The procedure used for determination of alkaloids in *Gs* sample is similar to that described by [9, 4]. Briefly, 0.2 g of *Gs* leaf powder was added to a glass test tube that has 3 ml of hexane. The powder and hexane were thoroughly mixed, shaken well, and filtered. To the hexane and *Gs* leaves extract mixture, 5 ml of 2% HCl were added, and the mixture heated until boiling. The mixture was then filtered, and 1-3 drops of picric acid were added to the hexane, HCl and *Gs* leaf extract filtrate. The presence of alkaloids in the *Gs* sample was confirmed by the yellow-colored precipitate that was formed.

Test for Terpenoids
An amount of 0.8 gram of the *Gs* leaf powder was placed in a test tube, then 10 ml of methanol was poured in it, shaken well and filtered. Five ml of the plant extract of plant sample was taken. Then 2 ml of chloroform were mixed with the extract and 3 ml of sulphuric acid were added. Formation of reddish brown color indicates the presence of terpenoids in the selected plants [34].

Test for Tannins
Crude extract of *Gs* powder was mixed with 2 ml of 2% solution of FeCl3. A blue-green or black coloration indicated the presence of tannins [35].

Test for proteins, ninhydrin test
The Crude extract of *Gs* leaves powder was boiled with 2 ml of 0.2% solution of Ninhydrin. Violet color appearance will suggest the presence of amino acids and proteins [35].

Test for Carbohydrates, Fehling’s test
Two ml of equal volumes of Fehling A and Fehling B reagents mixed together were added to the crude extract and gently boiled. A brick red precipitate appearing at the bottom of the test tube indicates the presence of reducing sugars [35].

Test for Steroids
*Gs* leaves powder was mixed with 2 ml of chloroform and concentrated H2SO4 was added sidewise. A red colour produced in the lower chloroform layer indicated the presence of steroids. Another test was performed by mixing crude extract with 2 ml of chloroform. Then 2 ml of each of concentrated H2SO4 and acetic acid were poured into the mixture. The development of a greenish coloration indicated the presence of steroids [35].

Test for Saponins
The crude extract was mixed with 5 ml of distilled water in a test tube and it was shaken vigorously. The formation of stable foam was taken as an indication for the presence of saponins [35].

RESULTS AND DISCUSSION

Toxicity test
In the present study, brine shrimp (artemia) toxicity test was used to find out if the leaves extract is safe to consume. The artemia toxicity test is used routinely because it is convenient to use in addition to being inexpensive. As a routine test, it is employed as an initial step in the assessment and evaluation of the toxic characteristics of a substance [36], for screening drugs, and for determination of lethal doses, and for assessing health hazards that may arise from short term exposure to drugs [37]. On these grounds, we decided to use the brine shrimp (artemia) toxicity test to evaluate if the traditional medicinal plants *Gs* leave extract has potential toxicity. In the present study, the leave extracts from *Gs* showed no toxicity to brine shrimps at concentrations of 250,500, and 1000 μg in 40 ml of distilled water, and that was confirmed by the number of dead brine

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shrimp larvae, which was zero. From this result, we can conclude that the Gs leaves extract is apparently not toxic. Our result are in general agreement with that obtained by Ref. [37] who observed that water extract of Gs given at the rate of 2 g/kg to Wister male and female rats showed no toxicity as confirmed by the evaluation of pathological changes in different organs, hematological parameters, and urinalysis. The results of the present study are also in agreement with Ref. [38] who concluded that Gs aqueous leave extract is not toxic and may be safe, especially at the dose used for treatment, which is greatly lower than the doses used in their experiments. However, the findings of the present study are not in agreement with that found by Ref. [39] who reported signs of toxicity when experimental animals were drenched with the leaves powder of Gs. At the dosage of 1 or 2 g/k/day, this disagreement could be explained by the fact that the present study used the aqueous extract of the leaves while Ref. [39] used the actual leaves powder. Interestingly, Ref. [39] observed that the treated animals became drowsy suggesting an effect of Gs powder on the cardiovascular system function and blood pressure. This may explain the folk use of the plant for hypertension control. Further, Ref. [40] reported that spontaneous motor activity was decreased in mice by administration of the aqueous extract of Gs and that the sleeping time induced by pentobarbital was prolonged in treated mice.

**Antifungal activity**

In developing countries, medicinal plants are gaining popularity because of their perceived effects against microbes, and cost or invariability of health care. The World Health Organization (WHO) reported that approximately 80% of the world population use traditional medicinal plants the extracts or the active ingredients in medicinal plants [41]. On the other hand, more than 60% of the successful drugs developed against infections during the last twenty years utilized natural products [42]. Therefore, the interest in traditional medicinal plants will continue to grow as emerging infectious diseases continue to expand, and the numbers of drug resistant microbes continue to escalate. Further, plant extract are often consumed without paying attention to safety issues. In the present work, the antifungal activity of the extracts of Gs was studied against seven fungal strains. For the antifungal susceptibility test, the leaves extracts obtained from Gs showed no activity (No inhibition) against all of the tested fungal strain. Ref. [43] indicated that since solvents used for extraction are expected to have different range of solubility for the phytochemical component of the medicinal plant, ethanolic extract would be more effective in treatment than an aqueous extract. In Western Sudan, people use the aqueous extract of the leaves as a medicine. However, from drug discovery point of view, it would be interesting to quantify antimicrobial activity of Gs ingredients extracted using ethanolic solvent in which plant material is expected to dissolve better.

**Phytochemical analysis**

This study also showed the presence of different phytochemicals whose biological activity can be of valuable therapeutic index. The result of the presence of the phytochemicals in Gs extract, shown in Table 1, is indicated by the colour intensity in a scale of 4 (+, ++, ++++, ++++).

Since medicinal plants, such as Gs, are expected to be a valuable source for a variety of drugs, no wonder that 80% of individuals from developed countries use traditional medicines extracted from plants according to W.H.O [41]. Therefore, it is important to investigate traditional medicinal plants to assess their properties, safety, and efficacy of treatment, and the optimum dose [44]. Table 1. Shows that plant under investigation contains flavonoids, alkaloids, terpenoids, tannins, proteins, carbohydrates, steroids and saponins. Our result is in line with the findings obtained by Ref. [45] who indicated that Gs leave extract had been found to contain alkaloids, tannins, flavonoids, amino acids, in addition to its antimicrobial activity. The results of the present study also agree with that reported by Ref. [46] who also confirmed the presence in Gs leave extract of flavonoids, saponins, tannins, terpenes, and carbohydrates. One of ingredients that we found in Gs leave extract are terpenoids, which are found to have a variety of pharmacological activities such anti-bacterial, anti-inflammatory and anti-malarial activities [47, 48]. The presence of flavonoids in Gs leaves extract documented in this study supports the findings of Ref. [49] who stated that the antimicrobial properties in some traditional medicinal plants can be due to the presence of flavonoids. Furthermore, several biological effects such as antioxidants abilities, and anti-inflammatory effects are attributed to flavonoids and phenolic compounds in medicinal plants [50]. These observations and the findings of the present study suggest there is an urgent need to investigate why Gs extract is credited for curing several diseases and infections in Africa. It would be important to find out which phytochemical compound in Gs is responsible for what activity.

The content of alkaloids, tannin and saponins found in the present study agree with that reported by Ref. [51] who reported high levels of alkaloids and low tannins and saponins content in Gs. Alkaloids found in medicinal plants used as anesthetic agents [52], a premise that was not investigated in the present study. In addition, tannins have been found by Ref. [53] to have a wide range of antimicrobial and anti-inflammatory effects. The anti-carcinogenic

![Fig 2:Chemical structure from Guiera senegalensis [34].](image-url)
potentials of tannins could be attributed to its anti-oxidative properties that protect living cells from oxidative damage [54]. It is documented in this study that steroids were present in Gs leaf extract. These steroids have antibacterial effects [55], and are important because of its relationship with steroid hormones such as sex hormones estrogen and testosterone [56].

The observed antibacterial effects of the medicinal plants extracts have been related to the presence of tannins, flavonoids and saponins [57]. This is in agreement with Ref. [58] who investigated the effect Eucalyptus Camaldulensis extract against pathogenic bacteria (Salmonella typhi and Escherichia coli) and attributed the effect of the plant extract to the active phytochemical compounds it has, namely tannins, flavonoids and saponins. Gs have been shown to contain compounds such as alkaloids, tannins, flavonoids that demonstrated their antimicrobial activity [59].

Traditional Uses

G. senegalensis is considered by traditional practitioners of Nigeria, Burkina Faso, Senegal, and Mali as a panacea, both its medicinal properties are significant and varied. The usual form of preparation for internal use is in decoction or mixed with food preparations. G. senegalensis leaves are widely administered for pulmonary and respiratory complaints, for coughs, as a febrifuge, colic and diarrhea, syphilis, beriberi, leprosy, impotence, rheumatism, diuresis and expulsion [60,61].

Pharmacological and Toxicological Activities

Guiera senegalensis is a popular medicinal plant for both human and veterinary use in West Africa. Its leaves have been reportedly used to treat rhinitis, bronchitis and fever, and the roots to treat diarrhea and dysentery. It is recognized as being active against cough, respiratory congestion and fever [14], and is prescribed as an antitussive [11,15,16,17,18], to ease breathing and to treat lung and bronchial disorders. It is also used against malaria [9,19,20]. The syrup D2 from G. senegalensis has been screened for its antitussive clinical essay [10]. Aqueous extracts from its roots and leaves have also been screened for toxicity [21]. The galls of G. senegalensis are used in Burkina Faso as an ethnoveterinary product to increase milk production in cows and to treat fowlpox infection in chickens [22]. Guiera senegalensis leaves extract or fractions may be used for the treatment of various disease caused by Escherichia coli, Pseudomonas aeruginosa and Klebsiella pneumonia [24]. View in toxicity, aqueous extracts are overall low toxicity; “extract of leaves being slightly more active than the extract from roots in the Guinea pig, rabbit, dog and less active in mice and rats [11]. G. Senegalensis at lower doses is not harmful to the liver and therefore can be exploited as it is served in the treatments of some illnesses [25]. Treatment with water and methanol extracts from G. senegalensis leaves resulted in endotheliotoxicity, hepatonephrophyph and pancreatic hyperplasia [27]. Guieranine A from Guiera senegalensis showed a strong antiplasmodial activity associated with a high cytotoxicity toward human monocytes [1]. Methanol fraction from leaves extract only had antiplasmodial activity [28].

CONCLUSION

In conclusion, medicinal plants are used for discovering and screening of the phytochemical constituents which are very helpful for the manufacturing of new drugs for treatment of various diseases. The results obtained in the present study have shown that Gs leaves have high concentration of alkaloids and low concentration of tannins and saponins and have no toxic effect and show no antimicrobial activities in case of fungi. These observations and the findings of the present study suggest an urgent need for investigating why Gs extract is credited for curing several disease and infections in Africa. It would be of important to find out which phytochemical compound in Gs is responsible for what activity. Thus, we hope that the important phytochemical compounds identified by our study in the Gs, will be helpful in treating different diseases of this particular region of Africa, and may be in other regions. The plant extract may be safe, especially at the therapeutic dose which is far lower than the tested doses. The results could serve for further pharmacological and phytochemical research. Also more research needed to evaluate the potential effectiveness of the crude extracts as the antimicrobial agents.

REFERENCES

1. Fiot J, Sanon S, Asaz N, et al. Phytochemical and pharmacological study of roots and leaves of Guiera senegalensis J.F. Gmel (Combretaceae). Journal of ethnopharmacology 2006; 106(2):173-178.
2. Silva O, Serrano R, Gomes ET. Botanical characterization of Guiera senegalensis leaves. Microscopy and Microanalysis. Official journal of microscopy society of America 2008; 14(5):398-404.
3. Hutchinson J, Daijik JM. Flora of West Tropical Africa. London: Crown Agents, 1972.
4. Zeljan M, Marica M, Franz B. Flavonoida of G. senegalensis – Thin layer Chromatography and Numerical Methods. Croatica Chemica Acta 1998; 71(1):69-79.
5. Shertima YA, Tijiani MA, Kanumi Y, et al. Phytochemical and anti diarrhoeal properties of methanol extract of Guiera senegalensis J.F. Gmel International Journal of Pharmacy, 2012; 3(11).
6. Tijiani A, Sallai MS, Sunusi I. Synergistic activity of methanolic extract of Adeniium obesum (Apocynaceae) stem-bark and azytetracycline against some clinical bacterial isolates. Bayero Journal of Pure and Applied Sciences 2011; 4(1):79-82.
7. Adedapo AA, Sofidja MO, Afolayan AJ. Antiinflammatory and analgesic activities of the aqueous extracts of Margaretaria discoidea (Euphorbiaceae) stem bark in experimental animal models. Int J Trop Biol. 2009; 57(4):1193-1200.
8. Ali AJ, Akanya HO, Dauda BN, et al. Antiplasmodial, analgesic and anti-inflammatory effects of crude Guiera senegalensis J.F. Gmel (Combretaceae) leaf extracts in mice infected with Plasmodium berghii. Journal of Pharmacognosy and Phytotherapy, 2011, 3(10).
9. Ancolo C, Asaz N, Mahiou V, et al. Antimalarial activity of extracts and alkaloids isolated from six plants used in traditional medicine in Mali and Sao Tome. Phytotherapy Research. 2002; 16(7):64-69.
10. Dénou A. Contribution à l’étude des problèmes de conservation de la forme sirop des médicaments traditionnels. Thèse de Pharmacie, FMPOS, Bamako. 2008, 82p.
11. Koumaré M. Contribution à l’étude pharmacologique du Guiera senegalensis Lam, Combretaceae. Thèse de Doctorat en pharmacie, Toulouse – France, 1968.
12. Somboro A A, Patel K, Diallo D, Sidibe L, Chakhat JC, Figueredo MR, Calabro ML, De Pasquale R, Germano MP, Sanogo R, Guiera senegalensis: a plant with potential therapeutic activity. Sri Lanka J. Nat. Med. 2014; 18(1):1-9.
13. Ficarra R, Ficarra P, Tommasini S, Carulli M, Melardi S, Di Bella Somboro A A, Patel K, Diallo D, Sidibe L, Chakhat JC, Figueredo MR, Calabro ML, De Pasquale R, Germano MP, Sanogo R, Guiera senegalensis: a plant with potential therapeutic activity. Sri Lanka J. Nat. Med. 2014; 18(1):1-9.
14. Kerharo J, Adam JG. La Pharmacopée Sénégalaise traditionnelle. Plantes Médicinales et Toxiques. Editions Vigot Frères, Paris, 1974, p. 101.1
15. Negrevorge G. New medicinal based on a dry aqueous extract of Guiera senegalensis, especially useful as a hypotensive and antitussive agent. Fr. M. 1968, p. 2.
16. Faye O, Olschwang D, Gono-Barber H, Pousett JL. Action antitussive d’un extrait lyophilisé de Guiera senegalensis. Plantes médicinales Africaines II, 1960; 25(4):285-292.
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