GGE Biplot Stability Analysis of Seed Yield in Teff (Eragrostis tef (zucc.)) Varieties in South West Ethiopia

Afework Legesse*, Tegegn Belete

Ethiopian Institute of Agricultural Research, Jimma Agriculture Research Center, Jimma, Ethiopia

Email address:
afework.legesse@yahoo.com (A. Legesse)
*Corresponding author

To cite this article:
Afework Legesse, Tegegn Belete. GGE Biplot Stability Analysis of Seed Yield in Teff (Eragrostis tef (zucc.) Varieties in South West Ethiopia. Advances in Biochemistry. Vol. 8, No. 4, 2020, pp. 62-67. doi: 10.11648/j.ab.20200804.12

Received: November 12, 2020; Accepted: November 30, 2020; Published: December 28, 2020

Abstract: Tef (Eragrostis tef (zucc.) is one of the major cereal crop grown in Ethiopia where it is staple food for about 50 million people. Among many factors contributed to low production and productivity of tef lack of widely adopted improved tef variety is one of the factors. The experiment was conducted in south western Ethiopia across six test locations during the 2019 cropping seasons to study the nature of GEI on grain yield of tef and to classify environments based on the performance of genotypes. A total of seven tef varieties were laid out randomized complete block design with three replicates at each site. The Analysis of variance revealed highly significant difference for environment, genotype and genotype by environment interaction (p<0.001). Large proportion of the variation was explained by the environmental effect (69.22%) followed by the GEI effect (20.19%) and genotypes (7.5%) of the overall variation. GGE biplot analysis showed that PC1 and PC2 accounted for 42.37% and 30.42% of GGE sum of squares, respectively, explained 72.79% of the total variance. The six locations were divided into three mega environments G28, G22 and G25 being the best varieties in each of the mega environments. However, G28 had the highest stability out of these three varieties. E6, E5 and E3 were ideal environments or the most suitable taste location for Teff breeding in the region, while E4 followed by E1 and E2 is a less desirable as testing environment.

Keywords: Biplots Analysis, Genotype by Environment Interaction, GGE, Grain Yield, Stability, Tef

1. Introduction

Tef (Eragrostis tef (zucc.) Trotter, 2n=40) is the poaceae family native to Ethiopia, which is centre of diversity for this important cereal crop [1]. It is one of the major cereal crop grown in Ethiopia where it is staple food for about 50 million people [2]. It is cultivated on approximately about 3 million hectare producing 5.02 million tons [3].

Tef is an important rain fed crop adopted wide range of climatic and edaphic conditions. It needs an optimum total annual rainfall of 750 to 850 mm, growing season rainfall of 450-550 mm and a temperature range of 10 to 27°C [4]. The suitable Tef growing agro-ecological zones of Ethiopia have different ranges of altitude (from sea level up to 2800 m.a.s.l.). The ideal altitude ranges between 1700 up to 2200 meter above sea level [4].

Major factors contributing to low productivity of Tef is susceptibility to lodging, low yield potential of landraces under widespread cultivation, poor agronomic management practices, biotic and abiotic stresses [5]. Nevertheless, it is possible to increase the yield up to 4.5 ton per hectare by using improved varieties and proper management practices [6]. Determining the magnitude and nature of the production environment is also the most important strategy to maximize grain yield and ensure stable performance of tef varieties across varying environments [7]. Genotype by environment interaction testing over diverse environment is very important to ensure that whether there is a need to develop a widely adapted cultivar for all environments of interest, or specifically adapted cultivars for specific target environments [8, 9]. In crop improvement programs multi-environment performance tests across a wide range of environments are conducted to reduce the effect of GEI and to ensure that the selected genotypes have a high and stable performance across several environments as it is easier and cost effective both in terms of variety evaluation and seed multiplication [10]. However, the need to develop a stable variety across the environment is dependent up on the kind of
interaction prevailing [9]. Previous studies of G x E interaction on Tef have illustrated significant interaction of genotypes with environment for yield and yield related traits and use different parametric methods for partitioning GEI [11-13], but only a few studies use the GGE- bi-plot model for stability analysis [14]. Therefore, the present study was conducted to examine the pattern of genotype by environment interaction (GEI) of tef yield by using GGE biplot analysis, to identify the most stable tef genotype for wide and/or specific adaptations.

2. Materials and Methods

2.1. Plant Materials and Test Locations

A multi environment trial was conducted using seven tef varieties (Table 3) for each location. The trial was conducted during the 2019 main cropping seasons at, Gechi, Omonada, Gooma, Somodo, Melko and Kersa. Average weather data and geographical coordinates of the test sites are presented in Table 1. Seven nationally released tef varieties were included in the study (Table 2). They were obtained from Debre Zeit Agricultural Research Center (DZARC).

Table 1. Description of the test environments.

| Locations | Altitude (m.a.s.l) | Coordinates | Soil type | Temp (°C) | Rainfall (mm) |
|-----------|-------------------|-------------|-----------|-----------|---------------|
| Gechi     | 2087              | 8°27’N 36°21’E | Nitosols  | 20.7      | 1800          |
| Gooma     | 1,560             | 7°51’N 36°35’E | Nitosols  | 19.7      | 1764          |
| Kersa     | >1780             | NA          | Nitosols  | 20.3      | 2000          |
| Mana      | 1770              | 7°45’N 36°45’E | Nitosols  | 18.9      | 1624          |
| Melko     | 1753              | 7°47’N 36°47’E | Nitosols  | 22.0      | 1639          |
| Omonada   | 1975              | 7° 41’N 37°12’E | Nitosols  | 20.0      | 1600          |

2.2. Experimental Design and Management

The trial was conducted using randomized complete block design (RCBD) with three replications at all locations under rain-fed conditions. Sowing was done manually. Fertilizer rate, seed rate, and crop cultivation were applied based on agronomic recommendations for each site. Spacing between plots was 1 m, whereas between replications was 1.5 m and the total plot size was 2mx2m. Seed rates was based on the recommendation which was 15kg/ha. Planting was done on the onset of rain in the respective locations. As per the recommendations, plots were fertilized with 40 kg of N and 60 kg of P₂O₅ per hectare for light soils and 60kg N and 60kg P₂O₅ per hectare for black soils (Vertisols). All DAP was applied at planting, while urea was applied in split half at planting and the remaining half at tillering stage. All other relevant field trial management practices were carried out throughout the experimentation period across all locations as per the recommendations for the respective locations.

2.3. Data Collection

Data were recorded on plot and single plant basis. Individual plant based data were taken from five plants in each plot taken randomly from the centre of each plot.

2.3.1. Data Collected on Plot Basis

Days to heading (DH): The number of days from 50% of the plots showing emergence of seedlings up to the emergence of the tips of the panicles from the flag leaf sheath in 50% of the plot stands.

Days to maturity (DM): The number of days from 50% of the plots showing seedling emergence up to 90% of the plants in the plot reaching phenological maturity stage (as evidenced by eye-ball judgment of the plant stands when the color is changed from green to yellow color of straw)

Grain filling period (GFP): The number of days from 50% heading to 90% maturity of the stands in each plot

Lodging index (X): The value recorded following the method of Caldicott and Nuttall (1979) who defined lodging index as the sum of product of each scale or degree of lodging (0-5) and their respective severity percentage divided by five, where 0 value is fully upright (90°), 1 = 0-15° lodging, 2=15-30° lodging 3 = 30-45° lodging, 4 = 45-60° lodging and 5 = 60-90° lodging and the plants become completely flat.

Total biomass yield (g/plot): The weight of all the central row plants including tillers harvested at the level of the ground

Grain yield (g/plot): The weight of grain for all the central row plants including tillers harvested at the level of the ground

Straw yield (g/plot): The weight of straw plus chaff of all the central row plants including tillers harvested at the level of the ground

Thousand seed weight (gram): It is the weight of thousand seeds at 12.5% moisture content

Harvest index: The value computed as the ratio of grain
yield to the total (grain plus straw) biomass multiplied by 100.

2.3.2. Data Collected on Plant Basis

Plant Height (cm): Measured as the distance from the base of the stem of the main tiller to the tip of the panicle at maturity

Panicle Length (cm): The length from the node where the first panicle branch starts up to the tip of the main panicle at maturity

Culm Length (cm): The length of the main shoot node from the ground level up to the point of emergence of the panicle branches

Fertile Tillers: The number of panicle-bearing fertile tillers produced per plant

Statistical analysis

Combined analysis over years and locations was done separately for Gechi, Omonada, Gooma, Somodo, Melko and Kersa SAS software (SAS 9.0) after testing for homogeneity of variance.

GGE biplot analysis was conducted on the mean best linear unbiased estimate (BLUE) values of eight Tef genotypes in the respective locations using GenStat 18 [15].

3. Results and Discussions

The Analysis of variance revealed highly significant difference for environment, genotype and genotype by environment interaction (Table 1). Highly significant variation was observed for genotype by environment interaction of Tef grain yield, indicating that possibility of stability analysis. Large proportion of the variation was explained by the environmental effect (69.22%) followed by the GEI effect (20.19%). Genotypes accounted for (7.5%) of the overall variation. There still remains some proportion of variation left unexplained by the model pooled into the error term (3.08%). The high percentage of the environment sum square is an indication that the major factor that influence yield performance of Tef genotypes is the environment. The relatively large percentage of the Genotype x Environment interaction sum square, when compared to that of genotypes as a main effect, is a very important consequence. The G x E interaction is highly significant (p<0.01) accounting for 69.22% of the sum of squares implying the need for investigating the nature of differential response of the genotypes to environments.

Highly significant variations observed for most of the traits (Days to heading, days to maturity, plant height, Panicle length, culm length, lodging index, shoot biomass, grain yield, harvest index) tested among genotypes across all locations, indicating the existence of variability among the tested genotypes (Table 4).

### Table 3. ANOVA for grain yield (kg/ha) of eight Tef genotypes tested at six environments.

| Source of variation | Df | Mean square | Pr>|f | Proportion of TSS |
|---------------------|----|-------------|-----|----------|------------------|
| Genotype (G)        | 7  | 13.85**     | <.0001 | 7.50%    |
| Location            | 5  | 17.8211**   | <.0001 | 69.22%   |
| Rep within location | 2  | 0.04778     | 0.8934 |          |
| Genotype X Environment (GxE) | 35 | 7.452**     | <.0001 | 20.19%   |
| Pooled error        | 94 | 0.4232      |        | 3.08%    |

*CV= coefficient of variation df=degree of freedom, TSS= total sum square

### Table 4. Combined Analysis of variance and mean performance of different traits of tef varieties tested at different locations.

| Varieties | HD (Days to heading) | MD (Days to maturity) | PH (Plant height) | PL (Panicle length) | CL (Culm length) | LI (% | SHB (Shoot biomass) | GY (Grain yield) | HI (Harvest index) |
|-----------|----------------------|-----------------------|-------------------|--------------------|------------------|-------|---------------------|------------------|------------------|
| Dagim     | 56.4                 | 107.3                 | 106.1             | 41.4               | 64.6             | 54.9  | 35.8                | 8.3              | 23.2             |
| Negus     | 54.3                 | 106.6                 | 96.9              | 38.2               | 58.7             | 58.9  | 37.4                | 7.8              | 22.6             |
| Tesfa     | 55.6                 | 106.9                 | 97.8              | 36.5               | 61.3             | 56.2  | 35.8                | 6.5              | 18.2             |
| Felagot   | 54.8                 | 101.8                 | 85.3              | 31.7               | 53.6             | 62.2  | 35.2                | 7.9              | 23.4             |
| Abola     | 55.6                 | 107.4                 | 101.7             | 39.3               | 62.7             | 57.2  | 40.6                | 7.6              | 19.7             |
| Heber-1   | 55.3                 | 109.2                 | 106.2             | 42.6               | 63.5             | 54.6  | 40.1                | 7.7              | 20.1             |
| Gibe      | 55                  | 110.2                 | 95.4              | 39.3               | 56.1             | 61.7  | 32.3                | 5.8              | 19.1             |
| Kora      | 55.4                 | 108.6                 | 110.7             | 43.7               | 67               | 58.8  | 35.7                | 8.3              | 24.0             |
| Mean      | 55.3                 | 107.3                 | 100.2             | 39.1               | 60.9             | 58.1  | 36.6                | 7.5              | 21.2             |
| F test    | <.0001               | <.0001                | <.0001            | <.0001             | <.0001          | <.0001| 0.0002              | <.0001          | <.0001          |
| LSD value | 0.67                 | 0.89                  | 3.1               | 1.8                | 2.3             | 2.51  | 3.52                | 0.445            | 0.0218          |
| CV (%)    | 1.82                 | 1.25                  | 4.6               | 7.3                | 5.7             | 6.5   | 14.5                | 8.98             | 15.5             |
| R-square  | 0.94                 | 0.953                 | 0.94              | 0.83               | 0.92            | 0.89  | 0.89                | 0.97             | 0.86             |

HD=Days to heading, MD= days to maturity, PH=plant height, PL=Panicle length, CL=Culm length, LI=Lodging index, SHB=Shoot biomass, GY=Grain yield, HI=Harvest index

### Polygon View of GGE biplot Analysis /Which Won Where Pattern

The polygon view of GGE biplot (Figure 1) is important for studying the possible existence of different mega environments in a region [16, 17]. In the present investigation, the partitioning of GE interaction through GGE biplot analysis showed that PC1 and PC2 accounted for 42.37% and 30.42% of GGE sum of squares, respectively, explained 72.79% of the total variance (Figure 1). The polygon view of GGE biplot was formed by connecting the vertex genotypes with straight lines to form a polygon.
lines and the rest of the genotypes were placed within the polygon. The rays in Figure 1 were formed as perpendicular to the sides of the polygon or their extensions. There are four rays, which divided the biplot into four sections. The genotypes fell into four sections and all the tested environments fell into three sections (mega environments) as shown in Figure 1.

The first section contains two genotypes G24, and G26 and the vertex genotype for this section was G24, suggesting the high yielding genotype for the environments that fall in this sector. The second section contains two genotypes G27 (vertex genotypes) and G23 (vertex genotypes) were poorest yielding genotypes in any environments. The other vertex genotype G21 gave high yield in environment (E3 & E4) which fell in section 3. On the other hand, the genotype, which was located near the origin, was less responsive than the corner (vertex) genotypes. Hence, the G25, G28 and G26 were located apparently near the biplot origin showed Poorest, Moderately and average; respectively performance and these genotypes were less responsive to environments than the vertex genotypes. According to the findings of Yan and Tinker (2006), the vertex genotypes were the most responsive genotypes, as they have the longest distance from the origin in their direction. The vertex genotypes were G22, G24, G27, G23 and G21 far from the origin. These genotypes are the best or poorest in some or all environments because they are farthest from the origin of biplot [9], which were more responsive to environmental change and are considered as specially adapted genotypes.

**Figure 1.** The polygon view of GGE biplot to the identification of winning genotypes and their related Mega environments (where G=genotype, E=environment). Where G21= Dagim, G22=Negus, G23=Tesfa, G24= Felagot, G25=Abola, G26=Heber-1, G27=Gibe, G28=Kora, E1= Kersa, E2=Melko, E3=Somodo, E4=Gooma, E5= Omonada and E6= Gechi

**Genotypes Mean Yield and their Stability**

Visualization of mean performance and stability analysis of genotypes is an important issue in crop genotype evaluation. The estimation of yield and stability of genotypes were done by using the average environment (tester) coordinate (AEC) methods (Figure 2) [19, 20]. The line passing through the biplot origin is called the average environment (tester) coordinate (AEC), which is defined by the average PC1 and PC2 scores for all environments [9]. More close to concentric circle indicates higher mean yield. The line, which passes through the origin and is perpendicular to the AEC with double arrows, represents the stability of genotypes. Either direction away from the biplot origin, on the axis, indicates greater GE interaction and reduced stability. For selection, the ideal genotypes are those with both high mean yield and high stability. In the biplot, they are close to the origin and have the shorter vector from the AEC. Thus, genotypes G27, G23, G24 and G25 were the least stable and genotypes G28, G22, G26 and G25 were the most stable. On the other hand, the genotypes on the right side of the line with double arrows have yield performance greater than mean yield and the genotypes on the left side of this line had yields less than mean yield. In this study, the genotypes G22, G28 and G25 had the higher stability as well as higher mean yield and eventually had been constellated into the same group. However, G28 had the highest stability out of these three genotypes. These results are in agreement with those obtained by [21] in rice and [22] in wheat.

**Figure 2.** GGE biplot visualization of the genotypes ranking for both yield and stability performance over environments. Where G21= Dagim, G22=Negus, G23=Tesfa, G24= Felagot, G25=Abola, G26=Heber-1, G27=Gibe, G28=Kora, E1= Kersa, E2=Melko, E3=Somodo, E4=Gooma, E5= Omonada and E6= Gechi

**Evaluation of Genotypes Relative to Ideal Genotypes**

The ideal genotype as virtual genotype is one that has both high mean yields across test environments and is absolutely stable in performance [23, 9, 24]. This genotype has large PC1 scores (high mean yield) and small (absolute) PC2 scores (high stability). The center of the concentric circles (Figure 3) represents the position of an ideal genotype, which is defined by a projection onto the mean-environment axis that equals
the longest vector of the genotypes that had above-average mean yield and by a zero projection onto the perpendicular line (zero variability across environments). A genotype is more desirable if it is closer to the ideal genotype [25, 18, 26]. Although such an ideal genotype may not exist in reality, it can be used as a reference for genotype evaluation. Therefore, G28 was closer to the ‘ideal’ genotype followed by G22 and G25 being more desirable than other genotypes (Figure 3). On the other hand, the low yielding genotypes (G27 and G23) were considered to be undesirable because they are placed far from the ideal genotypes. It seems that identification of ideal genotype through GGE biplot methodology is a proper tool for identifying most stable high yielding genotypes.

Evaluation of Environments Relative to Ideal Environments

Discriminating ability and representativeness are the important properties of a test environment. An ideal environment should be highly differentiating for the tested genotypes and at the same time representative of the target environment [9, 19]. Similar to ideal genotype, an ideal environment is defined and showed by the small circle with an arrow pointing to it. As shown in Figure 4 the environments E6, E5 and E3 were ideal environments. The environments E2, E1 and E4 rank first, second and third, respectively based on the ideal nature of environments (Figure 4). The environment E6, E5 and E3 has large PC1 score and small PC2 score. Hence, this environment is more stable and suitable for all genotypes following E2. On the other hand, E4 is a less desirable as testing environment because it has large PC2 score. The discriminating ability of a location is concerned with the composition of genotypes, but the presence of GE interaction complicates the identification of an ideal test location [27]. The test environments should have large PC1 scores in order to discriminate genotypes in terms of the genotypic main effect and absolute small PC2 scores in order to be more representative of the overall locations [23].

4. Conclusion

The Analysis of variance revealed highly significant difference for environment, genotype and genotype by environment interaction. In the present investigation, the partitioning of GE interaction through GGE biplot analysis showed that PC1 and PC2 accounted for 42.37% and 30.42% of GGE sum of squares, respectively, explained 72.79% of the total variance. The polygon views of the GGE biplot pointed out that there existed three possible mega environments. The first mega environment consisted of two environments (E6 and E5), the second mega environment consisted of two environments (E1 and E2) and the third mega environment consisted of two environments (E3 and E4). The vertex genotypes were G22, G24, G27 and G25 having the largest distance from the origin. In this study, the genotypes G22, G28 and G25 had the higher stability as well as higher mean yield and eventually had been constellated into the same group. However, G28 had the highest stability out of these three genotypes. E6, E5 and E3 were ideal environments or the most suitable taste location for Tef breeding in the region. However, one season research data is not enough to identify the ideal environments so research across many years is
required to firmly conclude the identified mega environments and device a breeding strategy for those mega environments separately.

Acknowledgements

The authors would like to acknowledge Ethiopian Institute of Agricultural Research (EIAR) for financial support. The tef breeding team at Debre zeit and cereal case team of Jimma and device a breeding strategy for those mega environments required to firmly conclude the identified mega environments.

References

[1] Vavilov, I. (1951). The origin, variation, immunity and breeding of cultivated plants. Translated from the Russian by K. Starrchon Ronald Press, New York.

[2] Kebebew A., Gina C., Dejene G., Rizqah K., Solomon C., Sonia Pl., Regula Blösch, Abiel R., Suhail R. and Zerihun T. (2015). Genetic diversity in tef [Eragrostis tef (Zucc.) Trotter]. Frontiers in Plant Science 6 (177): 1-13.

[3] CSA (Central Statistical Agency) (2017). Agricultural Sample Survey 2016/17 (2009 E. C.), Vol. I. Report on Area and Production of Major Crops, (Private Peasant Holdings, Meher Season), Statistical Bulletin 584, Addis Ababa, Ethiopia.

[4] Seyfu, K. (1997). Tef. (Eragrostis tef (Zucc.) Trotter) Promoting Genetics and Improvement. 145-156. October 16-19, 2000, Rome, Italy.

[5] Kebebew, A., Yu, J. K., Zeid, M., Getachew B., Hailu T. and Sorrells, M. E. (2011). Breeding tef [Eragrostis tef (Zucc.) Trotter]; Conventional and molecular approaches. Plant Breeding, 130: 1-9.

[6] Likyelesh, G. and Loerz, H. (2013). Male sterility and gametoclonal variations from gynogenically derived polyploids of tef (Eragrostis tef), Zucc. Trotter. African Journal of Plant Science, 7 (2): 53-60.

[7] Tiruneh, K. (2000). Genotype × Environment Interaction in tef [Eragrostis tef (Zucc.) Trotter]. In: Hailu T., Getchew, B. and M. E. Sorrells. (eds.) Narrowing the Rift: Tef Research and Development. Proceeding of the international workshop on tef genetics and improvement. 145-156. October 16-19, 2000, Addis Ababa, Ethiopia.

[8] Yan W, M Kang, B Ma, S Woods, and P Cornelius (2007). GGE biplot vs AMMI analysis of genotype-by-environment data. Crop Sci. 47, 643–655.

[9] Yan W, and M S Kang (2003). GGE biplot analysis: A graphical tool for breeders, geneticists, and agronomists. CRC Press LLC.

[10] Matus-Cadiz M A, P Hucl, C E Perron, and R T Tyler (2003). Genotype x Environment interaction for grain color in hard white spring wheat. Crop Sci. 43, 219-226.

[11] Fufa, H., Hailu T., Kebebew A., Tesfaye T., Tiruneh K. and Girma T. (2000). Grain yield stability in late maturing genotypes of tef [Eragrostis tef (zucc.) Trotter]. Journal of Genetics and Breeding, 54: 13-18.

[12] L. D. Kassa, Marie F. Smith & H. Fufa (2006). Stability analysis of grain yield of tef (Eragrostis tef) using the mixed model approach, South African Journal of Plant and Soil, 23: 1, 38-42.

[13] Mathewos, A, and Getachew, B. (2012). Genotype x Environment interaction analysis of tef grown in southern Ethiopia using Additive Main Effects and Multiplicative Interaction Model. Journal of Biology, Agriculture and Healthcare, 2 (1): 66-72.

[14] Yazaychew Genet, Tsion Fikre, Worku Kebede, Solomon Chanyalew, Kidist Tolosa, Kebebew Assefa (2020). Performance of Selected Tef Genotype for High Potential Areas of Ethiopia. Ecology and Evolutionary Biology. Vol. 5, No. 3, pp. 35-42.

[15] GenStat. (2015). GenStat for Windows (18th Edition) Introduction. VSN International, Hemel Hempstead.

[16] Gauch, H. G. and Zobel, R. W. (1997). Identifying mega-environments and targeting genotypes. Crop Science 37 (2): 311-326.

[17] Yan, W., Hunt, A., Sheng Q. & Slianzvics Z. (2000). Cultivar evaluation and mega environment investigation based on the GGE biplot. Crop Sci. 40: 597-605.

[18] Yan & Tinker., (2006). Biplot analysis of multi-environment trial data: Principles and applications. Canadian J. Plant Sci., 6 (3): 623-645.

[19] Yan, W. (2001). GGE biplot a Windows application for graphical analysis of multi-environment trial data and other types of two-way data. Agro, J. 93: 1111–1118.

[20] Yan W, Hunt LA (2001). Interpretation of genotype × environment interaction for winterwheat yield in Ontario. Crop Sci. 41: 19-25.

[21] Akter, A., Hasan, M. J., Kulsum, U., Rahman, M. H., Khattun, M. and Islam, M. R., (2015). GGE biplot analysis for yield stability in multi-environment trials of promising hybrid rice (Oryza sativa L.). Bangladesh Rice Journal, 19 (1), pp. 1-8.

[22] Naheef, E, M Mohammad, 2013. Genotype x Environment interactions for grain yield in bread wheat (Triticum aestivum L.). Global Sci. Res. J. 1 (1): 045-052.

[23] Yan, W., and I. Rajcan (2002). Biplot evaluation of test sites and trait relations of soybean in Ontario. Crop Sci. 42: 11-20.

[24] Farshadfar, E., Geravandi, M. and Vaisi, Z. (2012). Chromosomal localization of QTLs controlling genotype × environment Interactions in barley, International Journal of Agriculture and Crop Sciences, 4 (6): 317-324.

[25] Kaya, Y., Akcua, M. and Taner, S. (2006). GGE-biplot analysis of multi-environment yield trials in bread wheat. Turkish Journal of Agriculture and Forestry, 30 (5), pp. 325-337.

[26] Mitrovic B, Stanisavljevi D, Treski S, Stojakovic M, Ivanovic M, Bekavac G, and Rajkovic M. (2012). Evaluation of experimental Maize hybrids tested in Multi-location trials using AMMI and GGE biplot analysis. Turkish Journal of Field Crops, 17 (1): 35-40.

[27] Yan, W., L. A. Hunt, Q. Sheng and Z. Szlavnics (2000). Cultivar evaluation and mega environment investigation based on the GGE biplot. Crop Science, 40: 597-605.