Desikan-Killiany-Tourville Atlas Compatible Version of M-CRIB Neonatal Parcellated Whole Brain Atlas: The M-CRIB 2.0

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Our recently published M-CRIB atlas comprises 100 neonatal brain regions including 68 compatible with the widely-used Desikan-Killiany adult cortical atlas. A successor to the Desikan-Killiany atlas is the Desikan-Killiany-Tourville atlas, in which some regions with unclear boundaries were removed, and many existing boundaries were revised to conform to clearer landmarks in sulcal fundi. Our first aim here was to modify cortical M-CRIB regions to comply with the Desikan-Killiany-Tourville protocol, in order to offer: (a) compatibility with this adult cortical atlas, (b) greater labeling accuracy due to clearer landmarks, and (c) optimisation of cortical regions for integration with surface-based infant parcellation pipelines. Secondly, we aimed to update subcortical regions in order to offer greater compatibility with subcortical segmentations produced in FreeSurfer. Data utilized were the T2-weighted MRI scans in our M-CRIB atlas, for 10 healthy neonates (post-menstrual age at MRI 40–43 weeks, four female), and corresponding parcellated images. Edits were performed on the parcellated images in volume space using ITK-SNAP. Cortical updates included deletion of frontal and temporal poles and ‘Banks STS,’ and modification of boundaries of many other regions. Changes to subcortical regions included the addition of ‘ventral diencephalon,’ and deletion of ‘subcortical matter’ labels. A detailed updated parcellation protocol was produced. The resulting whole-brain M-CRIB 2.0 atlas comprises 94 regions altogether. This atlas provides comparability with adult Desikan-Killiany-Tourville-labeled cortical data and FreeSurfer-labeled subcortical data, and is more readily adaptable for incorporation into surface-based neonatal parcellation pipelines. As such, it offers the ability to help facilitate a broad range of investigations into brain structure and function both at the neonatal time point and developmentally across the lifespan.

Keywords: newborn, parcellation, segmentation, infant, ground truth, manual tracing
INTRODUCTION

We recently published the M-CRIB (Alexander et al., 2017) neonatal parcellated brain atlas, comprising 100 regions in total, including 68 compatible with the Desikan-Killiany (DK; Desikan et al., 2006) adult cortical atlas, as well as basal ganglia, thalamus, and cerebellar regions. The DK atlas is one of the most commonly used parcellation schemes, thus an advantage of the M-CRIB atlas is that it provides compatibility of parcellated cortical regions between neonatal and later time points. This can help facilitate investigations into regional brain structure and function across the lifespan, potentially longitudinally. As we have discussed previously (see Alexander et al., 2017), in neonates, the major sulci and gyri seen in adults are present at term (Chi et al., 1977; Cowan, 2002; Griffiths, 2010; Hill et al., 2010). The M-CRIB atlas illustrates the ability of the DK cortical regions to be delineated at the neonatal time point, as well as the subcortical and cerebellar regions; while also capturing gross and subtle morphological differences between neonatal and adult brains. The M-CRIB atlas is also valuable in that it comprises 10 individual high-quality detailed manual parcellations based on high resolution T2-weighted images, providing a combination of detailed whole-brain ‘ground truth’ and individual variability in morphology not available previously. We have recently demonstrated the applicability of the M-CRIB atlas, reporting differences in neonatal regional brain volumes based on premature birth (Alexander et al., 2018).

A successor to the DK atlas is the Desikan-Killiany-Tourville (DKT; Klein and Tourville, 2012) adult cortical parcellated atlas, in which some regions with unclear or arbitrary boundaries were removed, and many existing boundaries were revised to conform to sulcal fundi. This provides greater anatomical consistency across individuals due to clearer and more reproducible landmarks. The use of sulcal-based landmarks also optimizes utility for application using surface-based labeling such as is performed in FreeSurfer (Fischl et al., 2002). Surface-based methods incorporate surface-based registration which aligns sulci and gyri more precisely than volume-based methods (Fischl et al., 1999; Makropoulos et al., 2018), thus facilitating more precise alignment of sulcally-bounded labels.

Multiple surface-based tools have been developed for infant data (e.g., Hill et al., 2010; Li et al., 2014, 2015; Kim et al., 2016; Makropoulos et al., 2018). FreeSurfer tools for infant parcellation are currently in development (e.g., Zollei et al., 2017). A key resource facilitating accurate surface-based parcellation at the neonatal time point is high-quality ground truth neonatal parcellated training data. Such data are currently in strong demand.

Here we firstly aimed to modify the cortical regions and protocol of the existing volumetric M-CRIB atlas to comply with the DKT cortical parcellation protocol, in order to (a) offer compatibility with data at older time points parcellated with the adult DKT atlas, (b) achieve greater anatomical consistency in labeling across brains due to some boundaries being revised to clearer landmarks in sulcal fundi, and (c) offer greater ease of adaptability for integration into neonatal surface-based parcellation pipelines due to the use of these sulcally-defined boundaries. Secondly, we aimed to update subcortical regions to offer greater compatibility with those segmented by FreeSurfer’s subcortical pipeline, including addition of the ‘ventral diencephalon,’ and removal of ‘subcortical matter’ labels. These cortical and subcortical updates together comprise the ‘M-CRIB 2.0’ neonatal atlas.

MATERIALS AND METHODS

Data

Data utilized were the individual segmentation images and T2- and T1-weighted images comprising the M-CRIB atlas. This sample consisted of 10 healthy term-born (≥37 weeks’ gestation) neonates (four females, six males; gestational age at scanning 40.29–43.00 weeks, \(M = 41.71, SD = 1.31\)), selected from a larger cohort of controls with MRI scans recruited as part of preterm studies (Spittle et al., 2014; Walsh et al., 2014). T2-weighted images were acquired using a transverse T2 restore turbo spin echo sequence with: 1 mm axial slices, flip angle = 120°, TR = 8910 ms, TE = 152 ms, FOV = 192 mm × 192 mm, in-plane resolution 1 mm² (zero-filled interpolated to 0.5 mm × 0.5 mm × 1 mm), matrix = 384 × 384. Three-dimensional T1-weighted images were acquired using a magnetisation prepared rapid gradient-echo sequence with: 1 mm axial slices, flip angle = 9°, TR = 2100 ms, TE = 3.39 ms, FOV = 192 mm × 192 mm, in-plane resolution 1 mm² (zero-filled interpolated to 0.5 mm × 0.5 mm × 1 mm), matrix = 384 × 384. T2-weighted images were bias-corrected using N4ITK (Tustison et al., 2010), skull-stripped using BET (Smith, 2002; Smith et al., 2004), aligned to the anterior commissure-posterior commissure axis with 3D Slicer v4.1.1 (Fedorov et al., 2012), and resampled to 0.63 mm × 0.63 mm × 0.63 mm isotropic voxels (preserving voxel volume) using FLIRT (Jenkinson and Smith, 2001; Jenkinson et al., 2002; Greve and Fischl, 2009). Further information about the sample, data and preprocessing is listed in Alexander et al. (2017).

T1-weighted images are included in the M-CRIB and M-CRIB 2.0 datasets, however, they were not used for manual tracing, because of low contrast between tissue types due to partial myelination at the neonatal time point. Rather, they are included as they may provide additional intensity information leverageable in multimodal automated parcellation pipelines. The T2-weighted images, which confer higher tissue contrast, were used both for parcellation of the original M-CRIB, and for the edits performed here.

Manual Editing Procedure

The individual segmentation images comprising the M-CRIB atlas were edited in volume space using Insight Toolkit (ITK)-SNAP v3.6.0¹ (Yushkevich et al., 2006), by one operator (B.A.). ITK-SNAP displays axial, sagittal, and coronal views and a

¹http://www.slicer.org/
²www.itksnap.org
composite 3D mesh representation of utilized labels. The edits were performed and checked on a combination of the axial, sagittal, and coronal views, with reference to the 3D surface view. Edits were performed region-by-region rather than brain-by-brain to maximize consistency of labeling for each region across brains. An exception to this was in some areas where edits to multiple adjacent regions were required, as the alterations to one region sometimes necessitated specification of adjacent areas’ boundaries. For some regions such as the newly-specified ventral diencephalon, edits were performed for the whole sample, and then checked and edited where necessary to ensure consistency.

Parcellation Protocols
In the following cortical protocol, revised boundary descriptors are listed that aimed to replicate the DKT (Klein and Tourville, 2012) protocol as closely as possible within this volumetric neonatal sample. Where possible, verbatim DKT boundary descriptors have been utilized, and are indicated in bold font. Descriptors retained from the DK protocol are indicated in italics. Descriptors either retained from the M-CRIB protocol or newly specified here are indicated in regular font. Some anatomical axis descriptors (e.g., ‘anterior’) have been adjusted to retain anatomical accuracy in volume space.

When revising the boundary descriptors and editing the data, reference was made to Klein and Tourville (2012), the anatomical atlas by Petrides (2011) which describes many sulci used as DKT boundaries, other anatomical atlases (Duvernoy et al., 1999; Rubin and Safdieh, 2007; Duvernoy, 2013), the BrainInfo database (National Primate Research Center, 1991-present3), and individual papers describing anatomy (Nagata et al., 1988; Watson et al., 1993; Türe et al., 1999; Dumoulin et al., 2000).

Updates between the M-CRIB and M-CRIB 2.0 atlases pertain to the DKT cortical regions, ventral diencephalon (added), brainstem (edited in the course of defining ventral diencephalon), left and right ‘subcortical matter’ (removed), and left and right cerebral white matter (edited in removal of subcortical matter labels). Cerebellum, hippocampus, amygdala, and ventricles, were retained as per the original M-CRIB atlas, and parcellation protocols for these regions are listed in Loh et al. (2016) and Alexander et al. (2017). Basal ganglia and thalamus were not manually edited and protocols for these regions are retained from the M-CRIB atlas, however, post-processing performed on these segmentations was removed, as described below.

M-CRIB 2.0 Protocol
Cortical Regions
Frontal pole, temporal pole, and “banks of the superior temporal sulcus” regions were removed as per the DKT protocol, and replaced with surrounding gyral labels.

Temporal – medial aspect
**Entorhinal cortex** Boundaries: Anterior: Temporal incisure (rostral limit of collateral sulcus). Posterior: Posterior limit of the amygdala. Superior: Medio-dorsal margin of the temporal lobe anteriorly, amygdala and hippocampus posteriorly.

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Medial: Medial aspect of the temporal lobe. Lateral: Rhinal sulcus (collateral sulcus), or the collateral sulcus if the rhinal sulcus is not present.

**Parahippocampal gyrus** Boundaries: Anterior: Posterior limit of the amygdala. Posterior: Posterior limit of the hippocampus. Medial: Medial aspect of the temporal lobe. Lateral: Collateral sulcus.

Temporal pole (removed): The area included in the DKT temporal pole has been redistributed to the superior, middle and inferior temporal gyrus regions.

**Fusiform Gyrus** Boundaries: Anterior: Anterior limit of occipitotemporal sulcus (anterior limit of collateral sulcus). Posterior: First transverse sulcus posterior to the temporo-occipital notch. This is consistent with the posterior extent of the existing parcellation, which was based on the M-CRIB boundary listed as “posterior transverse collateral sulcus (Duvernoy et al., 1999).” Medial: Collateral sulcus. Lateral: Occipitotemporal sulcus.

Temporal – lateral aspect
**Superior temporal gyrus** Boundaries: Anterior: Anterior limit of the superior temporal sulcus or a projection from the superior temporal sulcus to the anterior limit of the temporal lobe. Posterior: Junction of posterior horizontal ramus of the lateral sulcus (or its posterior projection) and caudal superior temporal sulcus (1st segment of the caudal superior temporal sulcus). Note: The DKT protocol lists 1st, 2nd, or 3rd segment, however, the current parcellations of this region posteriorly conform specifically to the landmark that Petrides (2011) describes as the 1st segment, i.e., bounding the posterior extent of supramarginal gyrus (Petrides, 2011). Medial: Superior temporal sulcus. Lateral: Occipitotemporal sulcus.

**Middle temporal gyrus** Boundaries: Anterior: Anterior limit of the superior temporal sulcus. Posterior: Anterior occipital sulcus. Note: this has also been described as the ascending limb of inferior temporal sulcus (Watson et al., 1993; Dumoulin et al., 2000; Petrides, 2011). This is described by Duvernoy et al. (1999) as only sometimes being present: “The inferior temporal sulcus is usually not continuous and does not provide easy identification. In the vicinity of the occipital lobe, its posterior end may occasionally run upward and be called the anterior occipital sulcus.” In cases where this sulcus segment did not occur, the boundary was a point on a theoretical line extending vertically from the occipito-temporal incisure on the cortical surface. Medial: Superior temporal sulcus anteriorly, posteriorly formed by caudal superior temporal sulcus third segment. Inferior: Inferior temporal sulcus.

**Inferior temporal gyrus** Boundaries: Anterior: Anterior limit of the inferior temporal sulcus. Posterior: Anterior occipital sulcus (see descriptor for posterior boundary of middle temporal gyrus). In cases where this sulcus segment did not occur, the boundary was a point on a theoretical line extending vertically from the occipito-temporal incisure on the cortical surface. Superior: Inferior temporal sulcus Inferior: Occipitotemporal sulcus (Duvernoy et al., 1999).
Transverse temporal cortex Description: Also termed Heschl's gyrus, this area lies along the superior temporal plane, extending from the retroinsular region to the lateral edge of the superior temporal gyrus. It can be a single gyrus, or divided into two gyri by an intermediate transverse temporal sulcus (Duvernoy et al., 1999; Rademacher, 2003).

Boundaries: Anterior: Anterior limit of first transverse temporal sulcus (also referred to as the anterior transverse temporal sulcus (Tamraz and Comair, 2006).) Posterior: Posterior limit of Heschl's sulcus [also referred to as the posterior transverse temporal sulcus (Rademacher, 2003; Tamraz and Comair, 2006), or transverse temporal sulcus (Duvernoy et al., 1999; Ono et al., 1990)]. Medial: Retro-insular area of the lateral fossa. Lateral: Lateral surface of the superior temporal gyrus.

Frontal

Superior frontal gyrus Boundaries: Anterior: Frontomarginal sulcus. Posterior: Precentral sulcus (lateral surface); paracentral sulcus (medial surface). Medial: Medial aspect of the frontal lobe. Inferior: Superior frontal sulcus.

Middle frontal gyrus – rostral division Description: Approximates the rostral-most three quarters of the middle frontal gyrus. Boundaries: Anterior: Anterior limit of the superior frontal sulcus. Posterior: A theoretical line separating the caudal-most quarter of the middle frontal gyrus. Medial: Superior frontal sulcus. Lateral: Inferior frontal sulcus; anterior to inferior frontal sulcus, the ventro-lateral boundary is formed by frontomarginal sulcus and lateral H-shaped orbital sulcus.

Middle frontal gyrus – caudal division Description: Approximates the caudal-most quarter of the middle frontal gyrus. Boundaries: Anterior: A theoretical line separating the caudal-most quarter of the middle frontal gyrus. Posterior: Precentral sulcus. Medial: Superior frontal sulcus. Lateral: Inferior frontal sulcus.

Inferior frontal gyrus Description: The inferior frontal gyrus comprises the three pars regions.

Inferior frontal gyrus – pars opercularis Boundaries: Anterior: Anterior ascending ramus of the lateral sulcus, which is also referred to as the ascending ramus (Türe et al., 1999). Posterior: Precentral sulcus. Superomedial: Inferior frontal sulcus. Inferomedial: Circular insular sulcus.

Inferior frontal gyrus – pars triangularis Boundaries: Anterior: Pretrigeminal sulcus. Posterior: Anterior ascending ramus of the lateral sulcus. Superomedial: Inferior frontal sulcus. Inferomedial: Anterior horizontal ramus of the lateral sulcus; if the anterior horizontal ramus of the lateral sulcus does not extend anteriorly to pretrigeminal sulcus, an anterior projection from anterior horizontal ramus of the lateral sulcus to pretrigeminal sulcus.

Inferior frontal gyrus – pars orbitalis Boundaries: Anterior: Pretrigeminal sulcus – if pretrigeminal sulcus does not extend ventrally to the lateral H-shaped orbital sulcus, a ventral projection from pretrigeminal sulcus to lateral H-shaped orbital sulcus completes the anterior boundary.

Posterior: Posterior limit of orbitofrontal cortex. Superomedial: Anterior horizontal ramus of the lateral sulcus – if the anterior horizontal ramus of the lateral sulcus does not extend anteriorly to the pretrigeminal sulcus, an anterior projection from anterior horizontal ramus of the lateral sulcus to pretrigeminal sulcus completes the lateral boundary. Inferomedial: Lateral H-shaped orbital sulcus.

Orbitofrontal cortex – lateral division Boundaries: Anterior: Frontomarginal sulcus. Posterior: Posterior limit of orbitofrontal cortex. Medial: Olfactory sulcus. Lateral: Lateral H-shaped orbital sulcus.

Orbitofrontal cortex – medial division Boundaries: Anterior: Frontomarginal sulcus. Posterior: Posterior limit of orbitofrontal cortex. Superomedial: superior rostral sulcus; if superior rostral sulcus merges with cingulate sulcus, the medial/dorsal boundary is formed by cingulate sulcus. Inferolateral: Olfactory sulcus.

Precentral gyrus Boundaries: Anterior: Precentral sulcus. Posterior: Central sulcus. Superomedial: Medial bank of the central sulcus. Inferomedial: Circular insular sulcus.

Paracentral lobule Description: Medial structure consisting of the superomedial ends of the pre-central and post-central gyri surrounding the superior end of the central sulcus (Duvernoy et al., 1999). Boundaries: Anterior: Paracentral sulcus. Posterior: Marginal ramus of cingulate sulcus. Inferomedial: Cingulate sulcus. Superolateral: Medial bank of the central sulcus.

Parietal

Post-central gyrus Boundaries: Anterior: Central sulcus. Posterior: Post-central sulcus. Superomedial: Medial bank of the central sulcus. Inferomedial: Circular insular sulcus – if the lateral limit of post-central sulcus extends anterior to circular insular sulcus, the posterior portion of the lateral/ventral boundary is formed by the lateral sulcus.

Supramarginal gyrus Description: Formed by sulci demarcating the cortical convolution surrounding the posterior ascending ramus of the lateral sulcus. Boundaries: Anterior: Post-central sulcus. Posterior: Primary intermediate sulcus superomedially, and caudal superior temporal sulcus (first segment) inferolaterally. Superomedial: Intraparietal sulcus. Inferior: Lateral sulcus anterior to posterior horizontal ramus of the lateral sulcus, posterior horizontal ramus of the lateral sulcus posteriorly.

Superior parietal cortex Boundaries: Anterior: Post-central sulcus. Posterior: Transverse sulcus lying immediately posterior to the parieto-occipital sulcus – this is described as the transverse occipital sulcus, medial segment, by Petrides (2011). Medial: Dorsomedial hemispheric margin. Lateral: Intraparietal sulcus.

Inferior parietal cortex Description: Includes the inferior parietal gyrus and the angular gyrus and lies inferior to the superior parietal gyrus. Boundaries: Anterior: Caudal superior temporal sulcus, first segment. Posterior: A theoretical line reaching from the parieto-occipital fissure to the temporoparietal incisure. Medial: Intraparietal sulcus. Lateral: lateral occipital sulcus anteriorly, transverse occipital sulcus lateral segment posteriorly.
The depth of the calcarine sulcus. *Cingulate sulcus*.

**Occipital**

**Lingual gyrus** *Boundaries*: Anterior: Posterior limit of the hippocampus. *Posterior*: Posterior limit of calcarine sulcus. *Medial*: Medial portion of the temporal and occipital cortices. *Lateral*: Superior parietal gyrus.

**Pericalcarine cortex** *Boundaries*: Anterior: Junction of calcarine sulcus and parieto-occipital sulcus. *Posterior*: Posterior limit of calcarine sulcus. *Superior*: Dorso-medial margin of calcarine sulcus. *Inferior*: Ventromedial margin of calcarine sulcus. *Medial*: Medial portion of the temporal and occipital cortices. *Lateral*: The depth of the calcarine sulcus.

**Cuneus cortex** *Boundaries*: Anterior: Parietooccipital sulcus. *Posterior*: Posterior limit of calcarine sulcus. *Ventral*: Dorso-medial margin of calcarine sulcus. *Dorsal*: Dorso-medial hemispheric margin.

**Lateral occipital cortex** *Boundaries*: Anterior: Temporo-occipital notch laterally, anterior occipital sulcus more medially, transverse occipital sulcus, medial segment, medial to intraparietal sulcus. *Posterior*: The last visible portion of occipital cortex. *Medial*: Cuneus/pericalcarine cortex. *Lateral*: The lateral surface of the hemisphere at this area’s anterolateral boundaries.

**Cingulate**

**Rostral anterior division** *Boundaries*: Anterior: Cingulate sulcus. *Posterior*: Corpus callosum genu. *Superior*: Subparietal sulcus. *Rostral/dorsal*: Mammillary bodies. *Ventral*: Dorsal to the corpus callosum, the ventral boundary is formed by the callosal sulcus. In the subgenual area, it is formed by the cingulate sulcus. In the case of “double parallel cingulate” sulcus that continues anteroventrally to join the “superior rostral sulcus” (listed in Klein and Tourville, 2012), the ventral boundary is the superior rostral sulcus, also termed ‘supraorbital sulcus’ (Duvernoy et al., 1999, p. 33).

**Caudal anterior division** *Boundaries*: Anterior: Corpus callosum genu. *Posterior*: Mammillary bodies. *Rostral/dorsal*: Cingulate sulcus; in the event of a “double parallel cingulate,” (e.g., Ono et al., 1990), the rostral/dorsal boundary of the cingulate is formed by the more rostral-dorsal branch of the cingulate sulcus. *Ventral*: Callosal sulcus.

**Posterior division** *Boundaries*: Anterior: Mammillary bodies. *Posterior*: Junction of the subparietal sulcus and cingulate sulcus (approximately).

**Superior**: Cingulate sulcus. *Ventral*: Callosal sulcus.

**Isthmus division** *Boundaries*: Anterior: Junction of the subparietal sulcus and cingulate sulcus (approximately). *Posterior*: The anterior calcarine sulcus (Duvernoy et al., 1999) if present, or the parieto-occipital fissure. *Lateral*: The depth of the calcarine sulcus.

**Insula**

**Insula Description**: Inverted-triangle-shaped area of mesocortex in the base of the lateral fossa covered by frontal, temporal, central and parietal opercula; and delineated from these by the circular insular sulcus (also termed perinsular or limiting sulcus) (Duvernoy et al., 1999; Türe et al., 1999). *Boundaries*: Anterior: Anterior peri-insular sulcus. *Superior*: Superior peri-insular sulcus. *Infero-Posterior*: Inferior peri-insular sulcus.

**Subcortical Regions**

**Basal ganglia and thalamus**

The manual tracing protocol for the M-CRIB basal ganglia nuclei (caudate, putamen, pallidum, and nucleus accumbens) and thalamus is described in Loh et al. (2016). These regions are delineated in the M-CRIB 2.0 atlas.
### TABLE 1
Mean volume of each M-CRIB 2.0 region, and volume relative to equivalent M-CRIB region.

| Region                        | Structure                                | Mean volume in mm³ (standard deviation), % volume of original M-CRIB area |
|-------------------------------|------------------------------------------|------------------------------------------------------------------------|
| CSF (extraventricular)        |                                          | S 89668 (18597) 100                                                   |
| Brainstem                     |                                          | S 5394 (632) 85                                                       |
| Corpus callosum               |                                          | S 764 (179) 100                                                      |
| L/R ventral diencephalon      | L 1195 (107) N/A R 1216 (121) N/A       |                                                                       |
| L/R cerebral white matter     | L 79958 (7244) 106 R 80610 (6990) 106   |                                                                       |
| L/R hippocampus               | L 1473 (157) 100 R 1430 (170) 100       |                                                                       |
| L/R amygdala                  | L 840 (86) 100 R 802 (91) 100            |                                                                       |
| L/R lateral ventricle         | L 2965 (841) 100 R 2212 (771) 100       |                                                                       |
| 3rd ventricle                 | S 481 (47) 100                           |                                                                       |
| 4th ventricle                 | S 410 (70) 100                           |                                                                       |
| Cerebellum                    |                                          |                                                                       |
| Cerebellar vermis anterior    | L 214 (34) 100                           |                                                                       |
| Cerebellar vermis superior posterior | S 201 (39) 100 |                                                                       |
| Cerebellar vermis inferior posterior | S 214 (38) 100 |                                                                       |
| L/R cerebellar hemisphere     | L 11374 (1473) 100 R 11390 (1466) 100   |                                                                       |
| Basal ganglia and thalamus    |                                          |                                                                       |
| L/R caudate                   | L 1370 (174) 104 R 1333 (136) 103       |                                                                       |
| L/R putamen                   | L 1757 (142) 103 R 1772 (126) 102       |                                                                       |
| L/R accumbens area            | L 189 (38) 115 R 188 (31) 108           |                                                                       |
| L/R pallidum                  | L 775 (82) 102 R 770 (79) 102           |                                                                       |
| L/R thalamus                  | L 3904 (409) 100 R 3896 (419) 100       |                                                                       |
| Cortex                        |                                          |                                                                       |
| L/R unknown                   | L 202 (42) 103 R 224 (50) 104           |                                                                       |
| L/R caudal anterior cingulate | L 1080 (474) 120 R 1306 (259) 125       |                                                                       |
| L/R caudal middle frontal     | L 1487 (272) 100 R 1545 (226) 100       |                                                                       |
| L/R cuneus                    | L 1930 (348) 91 R 2072 (309) 94         |                                                                       |
| L/R entorhinal                | L 556 (98) 112 R 560 (93) 114           |                                                                       |
| L/R fusiform                  | L 2306 (425) 101 R 2650 (440) 100       |                                                                       |
| L/R inferior parietal         | L 4544 (1206) 97 R 4825 (1228) 99       |                                                                       |
| L/R inferior temporal         | L 3048 (934) 103 R 2831 (642) 106       |                                                                       |
| L/R isthmus cingulate         | L 1073 (232) 100 R 1154 (236) 100       |                                                                       |
| L/R lateral occipital         | L 4649 (653) 103 R 4520 (953) 103       |                                                                       |
| L/R lateral orbitofrontal     | L 2479 (425) 67 R 2766 (616) 73         |                                                                       |
| L/R lingual                   | L 2528 (566) 100 R 2874 (667) 100       |                                                                       |
| L/R mental orbitofrontal      | L 1864 (283) 97 R 1822 (320) 96         |                                                                       |
| L/R middle temporal           | L 3214 (514) 118 R 4045 (965) 111       |                                                                       |
| L/R parahippocampal           | L 576 (114) 100 R 579 (84) 100          |                                                                       |
| L/R paracentral               | L 1438 (319) 100 R 1444 (396) 100       |                                                                       |
| L/R pars opercularis          | L 1315 (130) 98 R 1419 (142) 102       |                                                                       |
| L/R pars orbitals             | L 1217 (395) 150 R 1015 (290) 141       |                                                                       |
| L/R pars triangularis         | L 1570 (309) 150 R 1522 (368) 152       |                                                                       |
| L/R pericalcarine             | L 2307 (647) 100 R 2606 (685) 99       |                                                                       |
| L/R post-central              | L 5038 (686) 100 R 4998 (655) 100       |                                                                       |
| L/R posterior cingulate       | L 1132 (335) 102 R 1301 (501) 101       |                                                                       |
| L/R precentral                | L 4294 (478) 101 R 4125 (304) 100       |                                                                       |
| L/R precuneus                 | L 3173 (658) 100 R 3191 (545) 100       |                                                                       |
| L/R rostral anterior cingulate| L 904 (267) 111 R 1121 (317) 138       |                                                                       |
| L/R rostral middle frontal    | L 4142 (693) 103 R 4066 (961) 103       |                                                                       |
| L/R superior frontal          | L 8136 (643) 102 R 8083 (1028) 101     |                                                                       |
| L/R superior parietal         | L 4700 (1065) 100 R 4590 (1208) 100    |                                                                       |
| L/R superior temporal         | L 4401 (805) 118 R 4306 (637) 120       |                                                                       |
| L/R supramarginal             | L 3615 (898) 100 R 3243 (633) 101       |                                                                       |
| L/R transverse temporal       | L 678 (115) 100 R 634 (131) 100         |                                                                       |
| L/R insula                    | L 1969 (218) 100 R 1918 (190) 100       |                                                                       |

*L/R* indicates one label in each hemisphere. ‘S’ indicates a single label that is not separated based on hemisphere. Percentages are % of the volume of the corresponding area in the original M-CRIB atlas. *The L/R ‘unknown’ labels are used for unspecified voxels (as utilized in the adult DK atlas). In the M-CRIB and M-CRIB 2.0 atlases, these labels are utilized in vicinity of the amygdala anteriorly.*Parcellation protocols are listed in Alexander et al. (2017). *Parcellation protocols for basal ganglia and thalamus are listed in Loh et al. (2016).*
were not manually edited here. However, in the original M-CRIB dataset, basal ganglia and thalamus segmentations underwent morphological smoothing. For the M-CRIB 2.0, the smoothed segmentations of these structures were replaced with non-smoothed segmentations in order to recover fine-scale, irregular, intensity-based anatomical detail such as is provided for the rest of the M-CRIB and M-CRIB 2.0 regions.

### Ventral diencephalon

The protocol for this region is based on that of de Macedo Rodrigues et al. (2015). **Boundaries:** Anterior: Anterior commissure (however, unlike the protocol of de Macedo Rodrigues et al., 2015, where the infero-rostral boundary is designated as the infundibular recess, we have referred solely to the anterior commissure as an anterior boundary, as much of the optic recess was also visible posterior to the anterior commissure). Posterior: Medially, the posterior commissure. Laterally, the posterior extent of the lateral geniculate nucleus. However, the lateral geniculate nucleus itself was retained as part of the thalamus label. Superior: The inferior surface of the thalamus, posteriorly (as per de Macedo Rodrigues et al., 2015). Inferior: A line extending from the pontomesencephalic sulcus anteriorly, to the posterior commissure posteriorly. Lateral: The optic pathways (de Macedo Rodrigues et al., 2015).

### Brainstem

The M-CRIB brainstem label was originally derived via the initial automated MANTIS (Beare et al., 2016) tissue segmentation, and refined during the process of manually delineating surrounding structures. Here partial sections of the cerebral peduncles, red nucleus, and substantia nigra have been reassigned from the brainstem label to form part of the ventral diencephalon label.

### RESULTS

The M-CRIB 2.0 atlas comprises 94 regions: 62 cortical regions, and subcortical and cerebellar regions from the M-CRIB atlas. Figures 1, 2 illustrate some of the updates made, displayed on surface meshes and axial slices, respectively. Atlas colors and corresponding label names are shown in Supplementary Figure S1.

In Figure 2, altered regions surrounding basal ganglia and thalamus primarily reflect the removal of the ‘subcortical matter’ label, which was replaced with ‘ventral diencephalon’ and cerebral white matter labels.

Table 1 lists the mean volume of each M-CRIB 2.0 region, and the volume relative to the equivalent structure, where applicable, from the original M-CRIB atlas.

### DISCUSSION

The M-CRIB and M-CRIB 2.0 atlases provide neonatal parcellated regions compatible with those in adults, while also representing the gross and subtle morphological differences between neonatal and adult brains. The adult DK and DKT cortical atlases are comprised of major gyri and large-scale regions that are extant at term in infants. The DKT protocol involved removal of some abstractly-bounded regions, and specification of additional sulci as regional boundaries. These sulci were generally readily identifiable in the current data. In a few instances, however, boundaries consisting of minor sulci were not identifiable in the neonatal data. In these cases boundaries were adjusted to the most closely-equivalent boundary in the neonatal data to correspond with that specified in adults. For example, the posterior boundary of the supramarginal gyrus was defined as the first segment of the caudal superior temporal sulcus in the neonatal data, because the second and third segments (specified as alternatives in the adult DKT protocol) were not consistently identifiable. Such differences reflect morphology specific to neonates, highlighting the value of delineating atlases in neonatal data, rather than projection of atlases defined in adults into neonatal space. Similarly, a small number of non-cortical regions in the M-CRIB and M-CRIB 2.0 atlases are necessarily inconsistent between neonates and adults. For example, cerebellar white and gray matter can be parcellated separately in adults. However, these finely-interbranched structures are not delineable in neonatal data at the current resolution due to partial voluming, so cerebellar hemispheres are provided as single structures. Conversely, the CSF label in neonates includes the cavum septum pellucidum – the CSF-filled space between the two septa pellucida, however, this structure is infrequently seen in adults (Tubbs et al., 2011). Such differences necessitate clear protocol descriptions in neonatal data as we have provided, to facilitate clear understanding of the parcellated anatomy. The M-CRIB 2.0 atlas incorporates updates that increase compatibility with adult subcortical segmentations derived via FreeSurfer, namely the addition of ventral diencephalon, and removal of ‘subcortical matter’ labels.

The parcellated M-CRIB 2.0 images will be more readily adaptable for potential incorporation into surface-based neonatal parcellation pipelines. Indeed, forthcoming work from our lab consists of the production of surface-based templates of the DK-compatible M-CRIB cortical regions, and the DKT-compatible M-CRIB 2.0 cortical regions. Accompanying the templates will be a protocol for labeling neonatal data using these atlases in combination with existing infant surface-based tools.

The current volumetric parcellations may also be used in combination with labeling tools such as STAPLE (Zou et al., 2004; Akhondi-Asl and Warfield, 2013; Akhondi-Asl et al., 2014) that apply labels to new data in a probabilistic, voxelwise manner (see Alexander et al., 2018, for an example).

The individual volumetric parcellated images and T1- and T2-weighted images comprising the M-CRIB 2.0 atlas will be publicly available.

### CONCLUSION

We updated the M-CRIB neonatal parcellated brain atlas to be compatible with the DKT adult cortical parcellated atlas, and to incorporate updates to subcortical regions facilitating greater compatibility with FreeSurfer’s subcortical segmentation.
We achieved this via manual volumetric edits to the individual parcellated images, and via the production of a detailed, revised whole-brain parcellation protocol. The resulting M-CRIB 2.0 atlas offers greater compatibility with adult parcellated data, greater accuracy due to more reproducible landmarks, and greater optimisation for integration with surface-based infant cortical parcellation pipelines. This high-quality dataset can therefore help facilitate a broad range of investigations into brain structure and function both at the neonatal time point and developmentally across the lifespan.

ETHICS STATEMENT

This study was carried out in accordance with the recommendations of the National Statement on Ethical Conduct in Human Research (2007), and Royal Children's Hospital Human Research Ethics Committee, with written informed parental consent provided for all subjects. Written informed parental consent was given in accordance with the Declaration of Helsinki. The protocol was approved by the Royal Children's Hospital Human Research Ethics Committee.

AUTHOR CONTRIBUTIONS

BA, WL, LM, and AM performed manual parcellation of the original atlas. CA, RB, and JC provided technical and methodological support in its development. PA, LD, AS, JLYC, MS, and DT facilitated collection of the data. DT contributed conception of the atlas. CA, RB, and JC provided technical and methodological support in its development. BA, WL, LM, and AM performed manual parcellation of the individual atlases. CA, RB, and JC provided technical and methodological support in its development. PA, LD, AS, JLYC, MS, and DT facilitated collection of the data. DT contributed conception of the atlas. BA performed edits to the data and wrote the manuscript. CK provided conceptual consultation. All authors contributed to manuscript revision, read and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fnins.2019.00034/full#supplementary-material

REFERENCES

Akhondi-Asl, A., Hoyte, L., Lockhart, M. E., and Warfield, S. K. (2014). A logarithmic opinion pool based STAPLE algorithm for the fusion of segmentations with associated reliability weights. Med. Imaging IEEE Trans. 33, 1997–2009. doi: 10.1109/TMI.2014.2329603
Akhondi-Asl, A., and Warfield, S. K. (2013). Simultaneous truth and performance level estimation through fusion of probabilistic segmentations. Med. Imaging IEEE Trans. 32, 1840–1852. doi: 10.1109/TMI.2013.2266258
Alexander, B., Kelly, C. E., Adamson, C., Beare, R., Zannino, D., Chen, J., et al. (2018). Changes in neonatal regional brain volume associated with preterm birth and perinatal factors. NeuroImage 185, 654–663. doi: 10.1016/j.neuroimage.2018.07.021
Alexander, B., Murray, A. L., Loh, W. Y., Matthews, L. G., Adamson, C., Beare, R., et al. (2017). A new neonatal cortical and subcortical brain atlas: the Melbourne Children’s Regional Infant Brain (M-CRIB) atlas. NeuroImage 147, 841–851. doi: 10.1016/j.neuroimage.2016.09.068
Beare, R., Chen, J., Alexopoulos, D., Kelly, C. E., Smyser, C., Rogers, C., et al. (2016). Neonatal brain tissue classification with morphological adaptation and unified segmentation. Front. Neuroinform. 10:12. doi: 10.3389/fninf.2016.00012
Chi, J. G., Dooling, E. C., and Gilles, F. H. (1977). Gyral development of the human brain. Ann. Neurol. 1, 86–93. doi: 10.1002/ana.410010109
Cowman, F. (2002). Magnetic Resonance Imaging of the Normal Infant Brain: Term to 2 Years. MRI of the Neonatal Brain. London: WB Saunders, 58–59.
de Macedo Rodrigues, K., Ben-Avi, E., Silva, D., Choe, M.-S., Drottar, M., Wang, R., et al. (2015). A freesurfer-compliant consistent manual segmentation of infant brains spanning the 0–2 year age. Brain 16, 1565–1579. doi: 10.3389/fnhum.2015.00021
Desikan, R. S., Ségonne, F., Fischl, B., Quinn, B. T., Dickerson, B. C., Blacker, D., et al. (2006). An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. NeuroImage 31, 968–980. doi: 10.1016/j.neuroimage.2006.01.021
Dumoulin, S. O., Bittar, R. G., Kabani, N. J., Baker, C. L., Le, Goualher G, Bruce, P., G et al. (2008). A new anatomical landmark for reliable identification of human area V5/MT: a quantitative analysis of sulcal patterning. Cereb Cortex 10, 454–463. doi: 10.1093/cercor/10.5.454
Duvernoy, H. M. (2013). The Human Hippocampus: Functional Anatomy, Vascularization and Serial Sections With MRI, 4th Edn. Berlin: Springer Science & Business Media, doi: 10.1007/978-3-642-33603-4
Fedorov, A., Beichel, R., Kalpathy-Cramer, J., Finet, J., Fillion-Robin, J.-C., Pujol, S., et al. (2012). 3D Slicer as an image computing platform for the quantitative imaging network. Magn. Reson. Imaging 30, 1323–1341. doi: 10.1016/j.mri.2012.05.001
Fischl, B., Salat, D. H., Busa, E., Albert, M., Dienerich, M., Haxby, C., et al. (2002). Whole brain segmentation: automated labeling of neuroanatomical structures in the human brain. Neuron 33, 341–355. doi: 10.1016/S0896-6273(02)00569-X
Fischl, B., Sereno, M. I., Tootell, R. B., and Dale, A. M. (1999). High-resolution intersubject averaging and a coordinate system for the cortical surface. Hum. Brain Mapp. 8, 272–284. doi: 10.1002/(SICI)1097-0193(19998<4:272::AID-HBM10>3.0.CO;2-4
Greve, D. N., and Fischl, B. (2009). Accurate and robust brain image alignment using boundary-based registration. Neuroimage 48, 63–72. doi: 10.1016/j.neuroimage.2009.06.060

Griffiths, P. (2010). Atlas of Fetal and Postnatal Brain MR. San Diego, CA: Elsevier Health Sciences.

Hill, J., Dierker, D., Neil, J., Inder, T., Knutsen, A., Harwell, J., et al. (2010). A surface-based analysis of hemispheric asymmetries and folding of cerebral cortex in term-born human infants. J. Neurosci. 30, 2268–2276. doi: 10.1523/JNEUROSCI.4682-09.2010

Jenkinson, M., Bannister, P., Brady, M., and Smith, S. (2002). Improved optimization for the robust and accurate linear registration and motion correction of brain images. Neuroimage 17, 825–841. doi: 10.1006/nimg.2002.1132

Jenkinson, M., and Smith, S. (2001). A global optimisation method for robust affine registration of brain images. Med. Image Anal. 5, 143–156. doi: 10.1016/S1361-8415(01)00036-6

Kim, H., Lepage, C., Maheshwary, R., Jeon, S., Evans, A. C., Hess, C. P., et al. (2016). NEOCIVET: towards accurate morphometry of neonatal gyriﬁcation and clinical applications in preterm newborns. NeuroImage 138, 28–42. doi: 10.1016/j.neuroimage.2016.05.034

Klein, A., and Tourville, J. (2012). 101 labeled brain images and a consistent human cortical labeling protocol. Front. Neurosci. 6:171. doi: 10.3389/fnins.2012.00171

Li, G., Wang, L., Shi, F., Lin, W., and Shen, D. (2014). Simultaneous and consistent labeling of longitudinal dynamic developing cortical surfaces in infants. Medical image analysis 18, 1274–1289. doi: 10.1016/j.media.2014.06.007

Loh, W. Y., Connelly, A., Cheong, J. L., Spittle, A. J., Chen, J., Adamson, C., et al. (2016). A new MRI-based pediatric subcortical segmentation technique (PSSST). Neuroinformatics 14, 69–81. doi: 10.1007/s12021-015-9279-0

Makropoulos, A., Robinson, E. C., Schuh, A., Wright, R., Fitzgibbon, S., Bozek, J., et al. (2018). The developing human connectome project: a minimal processing pipeline for neonatal cortical surface reconstruction. Neuroimage 173, 88–112. doi: 10.1016/j.neuroimage.2018.01.054

Nagata, S., Rhoton, A. L., and Barry, M. (1988). Microsurgical anatomy of the choroidal ﬁssure. Surg. Neurol. 30, 3–59. doi: 10.1016/0038-1615(88)90180-2

National Primate Research Center (1991-present). BrainInfo. Available at: http://www.braininfo.org

Ono, M., Kubik, S., and Abernethy, C. D. (1990). Atlas of the Cerebral sulci. Stuttgart: Thieme.

Petrides, M. (2011). The Human Cerebral Cortex: An MRI Atlas of the Sulci and Gyri in MNI Stereotaxic Space. London: Academic Press.

Rademacher, J. B. (2003). “Topographical Variability of Cytoarchitectonic Areas,” in Cortical Areas: Unity and Diversity, eds A. Schau and R. Miller (London: Taylor & Francis), 53–78.

Rubin, M., and Saffleh, J. E. (2007). Netter’s Concise Neuroanatomy. Philadelphia: Saunders.

Smith, S. M. (2002). Fast robust automated brain extraction. Hum. Brain Mapp. 17, 143–155. doi: 10.1002/hbm.10062

Smith, S. M., Jenkinson, M., Woolrich, M. W., Beckmann, C. F., Behrens, T. E., Johansen-Berg, H., et al. (2004). Advances in functional and structural MR image analysis and implementation as FSL. Neuroimage 23, S208–S219. doi: 10.1016/j.neuroimage.2004.07.051

Spittle, A. J., Thompson, D. K., Brown, N. C., Treyvaud, K., Cheong, J. L., Lee, K. J., et al. (2014). Neurobehaviour between birth and 40 weeks’ gestation in infants born < 30 weeks’ gestation and parental psychological wellbeing: predictors of brain development and child outcomes. BMC Pediatr. 14:1. doi: 10.1186/1471-2431-14-111

Tamraz, J. C., and Comair, Y. G. (2006). Atlas of Regional Anatomy of the Brain Using MRI: With Functional Correlations. Berlin: Springer.

Tubbs, R. S., Krishnamurthy, S., Verma, K., Shoja, M. M., Loukas, M., Mortazavi, M. M., et al. (2011). Cavum velum interpositum, cavum septum pellucidum, and cavum vergae: a review. Child’s Nerv. Syst. 27, 1927–1930. doi: 10.1007/s00381-011-1457-2

Türe, U., Yasargil, D. C., Al-Mefty, O., and Yasargil, M. G. (1999). Topographic anatomy of the insular region. J. Neurosurg. 90, 720–733. doi: 10.3171/jns.1999.90.4.0720

Tustison, N. J., Avants, B. B., Cook, P. A., Zheng, Y., Egan, A., Yushkevich, P. A., et al. (2010). N4ITK: improved N3 bias correction. Med. Imaging IEEE Trans. 29, 1310–1320. doi: 10.1109/TMI.2010.2046908

Walsh, J. M., Doyle, L. W., Anderson, P. J., Lee, K. J., and Cheong, J. L. (2014). Moderate and late preterm birth: effect on brain size and maturation at term-equivalent age. Radiology 273, 232–240. doi: 10.1148/radiol.14132410

Watson, J. D. G., Myers, R., Frackowiak, R. S. J., Hajnal, J. V., Woods, R. P., Mazziotta, J. C., et al. (1993). Area V5 of the human brain: evidence from a combined study using positron emission tomography and magnetic resonance imaging. Cereb. Cortex 3, 79–94. doi: 10.1093/cercor/3.2.79

Yushkevich, P. A., Piven, J., Hazlett, H. C., Smith, R. G., Ho, S., Gee, J. C., et al. (2006). User-guided 3D active contour segmentation of anatomical structures: signiﬁcantly improved efﬁciency and reliability. Neuroimage 31, 1116–1128. doi: 10.1016/j.neuroimage.2006.01.015

Zollei, L., Ou, Y., Iglesias, J., Grant, P. E., and Fischl, B. (2017). “FreeSurfer image processing pipeline for infant clinical MRI images;,” in Paper presented at the Human Brain Mapping. (Vancouver, BC).

Zou, K. H., Warﬁeld, S. K., Bharatha, A., Temppany, C. M., Kaus, M. R., Haker, S. J., et al. (2004). Statistical validation of image segmentation quality based on a spatial overlap index 1: scientiﬁc reports. Acad. Radiol. 11, 178–189. doi: 10.1016/j.sr.2005.01.015

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or ﬁnancial relationships that could be construed as a potential conﬂict of interest.

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