Media Optimization in Immobilized Culture to Enhance the Content of Curcumin in *Curcuma longa* (Zingiberaceae) and Protein Profile of Treated Samples in Static Culture

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**Abstract**

Plant cell culture system has served as an alternative to enhance the production of high value phyto-pharmaceuticals. In the described study, the immobilized culture and static culture of *Curcuma longa* (Zingiberaceae) were used to maximize the production of the bioactive compound Curcumin. It was observed that Curcumin content in *in vitro* studies (static as well as immobilize cultures) were enhanced in one week old cultures fed with 5 mg/100 ml of media (Static 8.71%; immobilize 2.03%). Statistically significant (seven fold) enhancement in Curcumin content was obtained in one week old static culture, which was maintained on Zenk production media incorporated with cinnamic acid (Control 1.57% and induced 8.717%). Quantitative estimation was done using HPTLC analysis with standard Curcumin. To evaluate the effect of treatment on total protein in Curcumin biosynthesis, we have examined Curcumin content as well as the protein profile of treated samples of *Curcuma longa* static culture. All the treated samples were analysed using SDS-PAGE for their proteomic profiles. A 23,420 Da protein was prominently expressed in all samples which may be a glycine rich protein (works on defence mechanism). Treated samples exhibited decreased expression of the protein as compared to control. This may be attributed to the formation of Reactive Oxygen species (ROS) in culture condition due to high concentration of sucrose (5%) in the *Curcuma longa* culture media that is known to induce oxidative stress and subsequent increase in Curcumin production. Further investigation is required to understand the actual protein involvement in Curcumin synthesis in different cultures treated with different compounds. The study signifies the use of plant explants to develop immobilized and static cultures rather than callus in Zenk media, which reduce the time as well as expenditure. The proteomic profile of *C. longa* has been discussed earlier, but its effect on proteomic profile of *in vitro* treated samples is a new study.

**Keywords:** Immobilize culture; *Curcuma longa*; Curcumin; Protein profile

**Introduction**

Plants in Zingiberaceae family are widely distributed in many countries of Southeast Asia. In Thailand at least two-hundred species of Zingiberaceous plants are found and these include members of various genera, such as *Alpinia*, *Annonum*, *Curcuma*, *Elettang*, *Kaempferia*, and *Zingiber* [1,2]. Zingiberaceous plants have been widely used in traditional medicine, as well as food flavouring and spice agents. Many studies have focused on the bioactive small organic compounds from these plants and have supported the traditional medicinal use of the plant extracts, such as Curcumin [3], sesquiterpene [4-6], and various essential oils [7-9], flavonoids and phenolic compounds [10,11]. Their actual physiological functions are likely to be in the defence against phytophagous predators (mostly insects) and phytopathogenic microorganisms [12,13]. Kumar, 2010 have reported the antioxidant protein turmerin from *Curcuma longa* by using SDS PAGE technique. This protein was examined for its ability to prevent oxidative organ damage against Naja naja venom phospholipase A, in male swiss wistar mice. Discoveries of cell cultures capable of producing specific medicinal compounds at a rate similar or superior to that of intact plants have accelerated in the last few years and has been well reviewed. In order to obtain high yields suitable for commercial exploitation, efforts have been focused on isolating the biosynthetic activities of cultured cells, achieved by optimizing the cultural conditions and employing precursor feeding [14]. The current yield and productivity cannot fulfill the commercial goal of plant cell-based bioprocess for the production of most secondary metabolites, hence we have developed the novel protocol to enhance the Curcumin content in tissue culture system as also to reduce the cost of its production. Cell cultures have a higher rate of metabolism than intact differentiated plants because the initiation of cell growth in culture leads to fast proliferation of cell mass and to a condensed biosynthetic cycle. This is the most important advantage of plant cell cultures as model systems for the study of biosynthetic pathways, as secondary metabolite formation can take place within a short cultivation time (about 2-4 weeks). With emerging trends of exploiting plant cell culture for the production of high value phyto–pharmaceuticals; immobilization of plant cell has a very important role. Immobilization can overcome many limiting factors of suspension culture with distinct advantage of easier separation from product and also being amenable for biotransformation of low value compounds to higher value product [14].

One of the main problems encountered is the lack of basic knowledge of the biosynthetic routes and mechanisms responsible for the production of plant metabolites, so the developed protocol is also helpful in understanding the biosynthetic pathway of the end product (natural product). The proteomic profile of *C. longa* has been discussed earlier [1], but proteomic profile of *in vitro* treated samples is a new kind of study. Keeping all the facts, the *Curcuma longa* tissue samples that have been induced by different compounds in *in vitro* system aseptically

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(static and immobilized) were analysed for Curcumin contents also for the protein profile which made a clear perception on tentative protein that has a role in biosynthesis of desire compound.

**Material and Method**

**Static culture of *Curcuma longa*** for enhancement of curcumin

Rhizome of *C. longa* has been collected from the local market Mumbai (Latitude: 18°96′50″N Longitude: 72°82′53″E) and identified by Botanist of Haffkine Institute. Here we are focusing on the enhancement of this poly phenolic compound in rhizome tissue rather than callus *in vitro* system. The experiment is based on the dual strategies that were used by Zenk et al., for alkaloid production in *Catharanthus roseus* tissue culture. The experiment involves the transfer of biomass into production media that favours the production and accumulation of secondary metabolites. Here we have used Zenk media as basal media supplemented with different concentration of cinnamic acid and Tyrosine separately and 5% sucrose. pH was 5.6 before autoclaving. Sterile media, Autoclaved on 121°C were used to aseptic inoculation in laminar hood. Here it is mentioned that both compounds are intermediate compounds in biosynthetic pathway of Curcumin [15].

The inoculated cultures were maintained at 26 ± 1°C, 70% humidity with 16 hours of light (1000 lux) and 8 hours of dark period in growth room. The different tissue samples were harvested at the age of 1 week separately, weighed and subjected for cold extraction in ethanol (w/v) at room temperature. After 48 hours the extracts were dried, weighed and subjected for High Performance Thin Layer Chromatography (HPTLC) analysis. Statistical analysis was done by using Anova test (Graph pad prism) in triplicate (Figure 1).

**Immobilize suspension culture for enhancement of curcumin**

The fresh rhizome was washed with teepol and rinsed in running tap water. Rhizome explant (2 gm) was sterilized with 0.1% mercuric chloride for 2 min followed by three times wash with sterile water, aseptically and homogenized in mortar and pestle. Homogenate materials were then passed through net (425 μm), to remove the large cell aggregate and fibrous material. Homogenate tissue was suspended in 2% sodium alginate (10 ml) and the suspension was added drop wise to 50 mM Calcium chloride through sterilized needle and syringe. The alginate beads of diameter 2-3 mm were formed and were left in CaCl₂ solution for 30 min for the stabilization of beads and then washed with sterilized water separately.

**Figure 1:** A: *Curcuma longa* rhizome; B: *C. longa* static culture; C: *C. longa* immobilize culture; D, E: HPTLC Chromatogram of immobilize culture at 425 nm induced with Standard of Curcumin; F,G: HPTLC Chromatogram of Curcumin in static culture of *Curcuma longa* with standard Curcumin standard at 525 nm.
Curcumin (Rf value 0.76). Inert gas was used as spray gas in CAMAG High Performance Thin Layer Chromatography (HPTLC).

High Performance Thin Layer Chromatography (HPTLC)

High Performance Thin Layer Chromatography of all samples was carried out in Khalsa College, Matunga (E) with standard compound of Curcumin (purchased from Sigma Alderich) by using High Performance Thin Layer Chromatography (HPTLC).

Protein extraction

Fresh treated tissue of Curcuma longa rhizome samples were grinded to a powder with liquid nitrogen in a mortar and pestle separately. Briefly, the plant powder (5 g) was extracted by suspension in 20 mL of extraction buffer (0.5 M Tris, 30 mM HCl, 0.1 M KCl, 0.7 M sucrose and 1% (v/v) β-mercaptoethanol) for 30 min at 4°C, whereupon the supernatant was then collected by centrifugation at 4,000×g for 10 min. The precipitate was extracted twice in extraction buffer and the extracts were then extracted with a 1:5 (v/v) ratio of water-saturated phenol at 4°C for 60 min. After phase separation the phenol phase were collected by centrifugation at 4,000×g for 10 min, re suspended in 20 mL of extraction buffer (0.5 M Tris, 30 mM HCl, 0.1 M KCl, 0.7 M sucrose and 1% (v/v) β-mercaptoethanol) for 30 min at 4°C, whereupon the supernatant was separated in centrifugation at 4,000×g for 10 min. The precipitate was extracted twice in extraction buffer and the extracts were then extracted with a 1:5 (v/v) ratio of water-saturated phenol at 4°C for 60 min. After phase separation the phenol phase were then harvested and proteins were precipitated from the phenol phase by the addition of a four-volume of 0.1 M ammonium acetate in methanol and left overnight at -20°C. The resulting phenol-soluble protein pellet were collected by centrifugation at 4,000×g for 10 min, re suspended in cold water with sonication for 3 min and then precipitated again in nine volumes of cold acetone at -20°C for 2 h and centrifuged at 4,000×g for 10 min. The protein pellet was air-dried to remove the acetone. The extracts were separately extracted with the equal volume of ethyl acetate and made a layer of EA, was in rich volume of pigments ,the rest of the extracts were separately extracted with the equal volume of ethyl acetate and made a layer of EA, was in rich volume of pigments ,the rest of the extracts were then extracted with a 1:5 (v/v) ratio of water-saturated phenol at 4°C for 60 min. After phase separation the phenol phase were then harvested and proteins were precipitated from the phenol phase by the addition of a four-volume of 0.1 M ammonium acetate in methanol and left overnight at -20°C. The resulting phenol-soluble protein pellet were collected by centrifugation at 4,000×g for 10 min, re suspended in cold water with sonication for 3 min and then precipitated again in nine volumes of cold acetone at -20°C for 2 h and centrifuged at 4,000×g for 10 min. The protein pellet was air-dried to remove the acetone. The extracts were separately extracted with the equal volume of ethyl acetate and made a layer of EA, was in rich volume of pigments ,the rest of the lower layer was separated and the hence we obtained the protein rich portion of the extracts. The amount of protein in each sample was determined by the Bradford assay [16] using BioTeK multimode reader. The Bradford assay was determined by the Bradford assay [16] using BioTeK multimode reader. The amount of protein in each sample was determined by the Bradford assay [16] using BioTeK multimode reader. The amount of protein in each sample was determined by the Bradford assay [16] using BioTeK multimode reader.
Formulation of Reactive Oxygen species (ROS) in culture condition and which modified the protein. In this case, increased level of sucrose than MS media (5%) has been added to Curcuma longa culture media, which is known inducer of oxidative stress (abiotic stress) and eventually the Curcumin production is increased. The protein profile expression was down regulated in all treated samples. Maximum intensity is expressed by tissue grown in Tyrosine 40 mg/100 ml (Relative intensity of Glycine rich protein: 65.53% w.r.t control) which is comparable to control (Table 2). The significant part of the research is to use the plant explants as source to develop immobilize culture and static culture rather than callus in Zenk production media, which reduce the time as well as expenditure and is new protocol; (Table 1). Large number of studies has focussed on the production of secondary metabolites in C. longa. However the exact mechanisms of ROS generation and its implication on the synthesis of stress specific metabolites have remained unanswered. The present study is a preliminary indicator of the changes in stress related proteins (e.g. possibly Glycine Rich Protein) with respect to induction of abiotic stress. Further studies encompassing complete proteome profiles are warranted to understand the mechanisms behind such phenomenon. After observing the results, it can be assumed that in stress condition, the protein may take part in the biosynthesis of Curcumin and itself get reduced. The present study is useful for the pharmaceutical industries which can enhance the Curcumin production by giving abiotic stress. These will not only enlighten us regarding the molecular mechanisms of plant stress physiology, but shall also provide new insights into developing protocols to enhance plant secondary metabolites.

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