Effect of genetic variants in cell adhesion pathways on the biochemical recurrence in prostate cancer patients with radical prostatectomy

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Abstract
The aberrant expression of cell adhesion molecules is a hallmark of epithelial-to-mesenchymal transition, resulting in the transformation of cancer cells to a more aggressive phenotype. This study investigated the association between genetic variants in cell adhesion pathways and the prognosis of patients with prostate cancer following radical prostatectomy (RP). A total of 18 haplotype-tagging single-nucleotide polymorphisms (SNPs) in eight cancer-related adhesion molecules were genotyped in 458 patients.
1 | INTRODUCTION

Prostate cancer is the second most common cancer and the fourth leading cause of cancer death in men, worldwide. Most early stage prostate cancers tend to develop slowly and show an indolent clinical course. However, some prostate cancers display aggressive behavior and metastasize to other organs. While localized prostate cancer can be well controlled by active surveillance, radical prostatectomy (RP), or radiotherapy, metastatic tumors remain a lethal disease. Therefore, identification of key molecules and accurate prediction of patient prognosis are particularly important for prognostic and therapeutic purposes.

Recent evidence indicates that epithelial-to-mesenchymal transition (EMT) is a critical step for cancer progression. During EMT, polarized epithelial cells alter cell adhesion molecules and generate a new microenvironment, acquiring the aggressive behavior of metastatic competence, such as stem cell-like features and treatment resistance. It is believed that the dysfunction of cell adhesion molecules, such as cadherins and integrins, is involved in cancer progression, based on the correlation of their expressions and tumor stage, metastasis, as well survival. Given the important role of EMT in carcinogenesis, we hypothesized that genetic variants in cell adhesion molecules might influence the prognosis of prostate cancer. Therefore, we conducted a two-stage study to investigate the impact of single-nucleotide polymorphisms (SNPs) in eight cancer-related cell adhesion molecules on the biochemical recurrence (BCR) in patients with localized prostate cancer after RP.

2 | MATERIALS AND METHODS

2.1 | Patient recruitment and data collection

A total of 643 patients, with histopathologically confirmed prostate cancer, that underwent RP were recruited from three Taiwan medical centers: Kaohsiung Medical University Hospital, Kaohsiung Veterans General Hospital, and National Taiwan University Hospital, as described previously. A two-stage approach was applied to evaluate the effect of genetic variants in cell adhesion pathway genes on patient prognosis. Therefore, the study population was randomly divided into discovery and replication sets with a 7:3 ratio. The demographic data were collected through in-person interviews using a structured questionnaire, which intends to identify individuals and/or families probably at-risk for prostate cancer, and the clinicopathologic information was retrieved from patients’ medical records. All participants in the study were unrelated. BCR was defined according to two consecutive prostate-specific antigen (PSA) measurements of 0.2 ng/mL or more after RP. BCR-free survival was defined as the duration from RP to the date of BCR. This study was approved by the institutional review board of Kaohsiung Medical University Hospital, and all participants signed the informed consent form according to institutional guidelines.

2.2 | SNP selection and genotyping

Several adhesion molecules commonly participate in the cancer metastasis process, including CD276 molecule...
CD276), CD6 molecule (CD6), CD8a molecule (CD8A), cadherin 2 (CDH2), claudin 11 (CLDN11), integrin subunit beta 1 (ITGB1), integrin subunit beta 7 (ITGB7), and poliovirus receptor (PVR). We initially selected 25 common tag SNPs in these cell adhesion genes using SNPinfo, based on the following criteria: a minor allele frequency of > 0.05 in the HapMap CHB (Han Chinese in Beijing) population, a pairwise linkage disequilibrium squared correlation coefficient ($r^2$) of > 0.8, whether they were potentially functional, and a maximum of five tag SNPs per gene. Genomic DNA was extracted from peripheral blood samples using the QIAamp DNA Blood Mini Kit (Qiagen, Valencia, CA, USA) according to manufacturer's instructions. Genotyping was carried out using Agena Bioscience iPLEX matrix-assisted laser desorption/ionization time-of-flight mass-spectrometry technology at the National Centre for Genome Medicine, Taiwan, as described previously. The average genotype call rate for these SNPs was 92.6% and the average concordance rate was 99.9% among 35 blind duplicated quality control samples. Any SNP that failed at assay design ($N = 1$), below a genotyping call rate of 80% ($N = 4$), or did not conform to Hardy-Weinberg equilibrium ($P < 0.005$, $N = 2$), was removed. Thus, a total of 18 SNPs were included for further analyses.

### 2.3 | Bioinformatics analysis

The regulatory annotation of the risk SNPs and their proxies ($r^2 \geq 0.8$ in the Asian population from 1000 Genomes Project) was conducted by HaploReg v4.1. Expression quantitative trait loci (eQTL) analysis was performed by using Genotype-Tissue Expression (GTEx). The prognostic significance of prostate cancer was analyzed using the publicly available GSE70769 microarray dataset.

### 2.4 | Statistical analysis

All statistical analyses were undertaken with Statistical Package for the Social Sciences software version 19.0.0 (IBM). Kaplan-Meier analysis with log-rank test was used to compare survival time between subgroups. Additive, dominant, and recessive genetic models were applied to analyze the prognostic effects of cell adhesion gene SNPs. The hazard ratios (HRs) and the corresponding 95% confidence intervals (CIs) were calculated by multivariate Cox regression analyses, which were adjusted for age, PSA at diagnosis, pathologic Gleason score, stage, and surgical margin. The combined HRs and 95% CIs were calculated by random effect models when heterogeneity of between-set existed; otherwise a fixed effect model was used. All tests were two-sided, and $P$-values < 0.05 were regarded as statistically significant.

### 3 | RESULTS

The clinicopathologic characteristics of patients in the discovery and replication sets are shown in Table 1. Most characteristics are comparable between the two sets. One hundred eighty-four (40.2%) and 90 (48.6%) patients experienced BCR during the median follow-up times of 54 and 74 months in the discovery and replication sets, respectively.

Of the 18 SNPs in eight cell adhesion molecules analyzed in the discovery set, four SNPs in CDH2 showed marginal association with BCR (Table S1). After adjusting for age, PSA at diagnosis, pathologic Gleason score, stage, and surgical margin, CDH2 rs643555 remained significant ($P = 0.039$, Table 2 and Figure 1A). Consistent with the results of discovery set, CDH2 rs643555 was found to be associated with BCR in the same direction in an independent replication set ($P = 0.046$, Table 2 and Figure 1B). In combined analysis, patients with rs643555 TT genotype had a significantly

### TABLE 1  Clinicopathologic characteristics of the study populations

| Characteristics                  | Discovery | Replication |
|----------------------------------|-----------|-------------|
| No. of patients                  | 458       | 185         |
| Median age, years (IQR)          | 66 (61-70)| 66 (61-70)  |
| Median PSA at diagnosis, ng/mL (IQR) | 11.1 (7.1-17.5) | 11.0 (6.9-18.7) |
| Gleason score, N (%)             |           |             |
| 2                                | 3 (0.7)   | 2 (1.1)     |
| 4                                | 8 (1.8)   | 5 (2.8)     |
| 5                                | 30 (6.6)  | 13 (7.2)    |
| 6                                | 119 (26.3)| 52 (28.7)   |
| 7                                | 232 (51.2)| 83 (45.9)   |
| 8                                | 25 (5.5)  | 8 (4.4)     |
| 9                                | 32 (7.1)  | 17 (9.4)    |
| 10                               | 4 (0.9)   | 1 (0.6)     |
| Pathologic stage, N (%)          |           |             |
| 1                                | 59 (13.1) | 25 (13.9)   |
| 2                                | 247 (54.9)| 102 (56.7)  |
| 3                                | 134 (29.8)| 47 (26.1)   |
| 4                                | 10 (2.2)  | 6 (3.3)     |
| Surgical margin, N (%)           |           |             |
| Negative                         | 241 (72.6)| 104 (75.4)  |
| Positive                         | 91 (27.4) | 34 (24.6)   |
| Biochemical recurrence, N (%)    | 184 (40.2)| 90 (48.6)   |
| Median follow-up, months         | 54        | 74          |

Abbreviations: IQR, interquartile range; PSA, prostate-specific antigen.
TABLE 2 SNPs associated with BCR in patients receiving RP

| Gene SNP | Discovery | | | Replication | | | Combined | |
|---|---|---|---|---|---|---|---|---|
| | N | BCR | HR (95% CI) | P | N | BCR | HR (95% CI) | P | N | BCR | HR (95% CI) | P |
| Genotype | | | | | | | | |
| $CDH2$ rs1944294 | | | | | | | | |
| AA | 234 | 97 | 1.00 | | 104 | 49 | 1.00 | | 1.00 | | | | |
| AT | 181 | 64 | 0.779 (0.524-1.158) | 0.217 | 67 | 31 | 0.808 (0.434-1.506) | 0.503 | 0.79 (0.56-1.10) | 0.16 | | | |
| TT | 39 | 19 | 1.477 (0.818-2.665) | 0.196 | 14 | 10 | 2.430 (1.087-5.431) | 0.031 | 1.76 (1.09-2.83) | 0.02 | | | |
| AT/TT vs AA | 0.885 (0.614-1.274) | 0.510 | 1.040 (0.597-1.814) | 0.889 | 0.93 (0.68-1.26) | 0.64 | | | | | | |
| TT vs AA/AT | 1.641 (0.929-2.899) | 0.088 | 2.640 (1.221-5.707) | 0.014 | 1.94 (1.23-3.07) | 0.005 | | | | | | |
| $CDH2$ rs3745045 | | | | | | | | |
| TT | 132 | 57 | 1.00 | | 60 | 31 | 1.00 | | 1.00 | | | | |
| TC | 242 | 87 | 0.952 (0.630-1.437) | 0.813 | 94 | 37 | 0.820 (0.424-1.589) | 0.557 | 0.91 (0.64-1.30) | 0.61 | | | |
| CC | 82 | 38 | 0.955 (0.564-1.617) | 0.864 | 31 | 22 | 2.102 (0.996-4.345) | 0.051 | 1.24 (0.81-1.91) | 0.33 | | | |
| TC/CC vs TT | 0.953 (0.645-1.406) | 0.807 | 1.103 (0.607-2.002) | 0.748 | 1.00 (0.72-1.38) | 0.98 | | | | | | |
| CC vs TT/TC | 0.985 (0.620-1.565) | 0.948 | 2.351 (1.219-4.534) | 0.011 | 1.31 (0.90-1.92) | 0.16 | | | | | | |
| $CDH2$ rs643555 | | | | | | | | |
| CC | 207 | 85 | 1.00 | | 85 | 41 | 1.00 | | 1.00 | | | | |
| CT | 185 | 64 | 0.793 (0.524-1.202) | 0.274 | 83 | 38 | 0.949 (0.522-1.727) | 0.865 | 0.84 (0.60-1.18) | 0.32 | | | |
| TT | 59 | 29 | 1.490 (0.901-2.462) | 0.120 | 17 | 11 | 2.138 (0.943-4.847) | 0.069 | 1.65 (1.07-2.53) | 0.02 | | | |
| CT/TT vs CC | 0.956 (0.662-1.381) | 0.812 | 1.126 (0.647-1.960) | 0.674 | 1.01 (0.74-1.37) | 0.97 | | | | | | |
| TT vs CC/CT | 1.645 (1.024-2.642) | 0.039 | 2.191 (1.015-4.731) | 0.046 | 1.78 (1.19-2.67) | 0.005 | | | | | | |
| $CDH2$ rs8084948 | | | | | | | | |
| TT | 235 | 90 | 1.00 | | 126 | 66 | 1.00 | | 1.00 | | | | |
| TA | 98 | 29 | 0.690 (0.400-1.189) | 0.182 | 51 | 20 | 0.489 (0.234-1.022) | 0.057 | 0.61 (0.39-0.95) | 0.03 | | | |
| AA | 16 | 5 | 0.538 (0.185-1.566) | 0.255 | 8 | 4 | 0.939 (0.325-2.718) | 0.908 | 0.71 (0.34-1.51) | 0.38 | | | |
| TA/AA vs TT | 0.660 (0.395-1.103) | 0.113 | 0.573 (0.302-1.088) | 0.089 | 0.62 (0.42-0.93) | 0.02 | | | | | | |
| AA vs TT/TA | 0.610 (0.212-1.751) | 0.358 | 1.123 (0.393-3.206) | 0.829 | 0.83 (0.39-1.75) | 0.62 | | | | | | |

Note. HRs were adjusted by age, PSA at diagnosis, pathologic Gleason score, stage, and surgical margin. P < 0.05 are in boldface.

Abbreviations: BCR, biochemical recurrence; CI, confidence interval; HR, hazard ratio; RP, radical prostatectomy; SNP, single-nucleotide polymorphism.

FIGURE 1 Kaplan-Meier survival curves of biochemical recurrence-free survival according to $CDH2$ rs643555 genotypes in (A) discovery set, (B) replication set, and (C) combined analysis. Numbers in parentheses indicate the number of patients.
shorter BCR-free survival compared with those with CC/CT genotypes (adjusted HR 1.78, 95% CI 1.19-2.67, P = 0.005, Table 2 and Figure 1C).

According to the functional annotation using HaploReg v4.1, rs643555 is an eQTL for CDH2, and is predicted to alter multiple regulatory motifs (Table S2). In eQTL analysis from the GTEx dataset, the risk allele T of rs643555 showed increased CDH2 expression in transformed human fibroblasts (P = 0.026, Figure 2A). We further investigated the prognostic effects of CDH2 expression on BCR-free survival after RP for prostate cancer. Based on the publicly available GSE70769 dataset, high expression of CDH2 was associated with higher pathologic stages (P = 0.043, Figure 2B) and poorer BCR-free survival (P = 0.005, Figure 2C).

4 | DISCUSSION

In this exploratory study, we investigated associations between genetic variants in eight cancer-related cell adhesion molecules and prostate cancer prognosis using a two-stage study design. CDH2 rs643555 was significantly associated with prostate cancer prognosis in both stages and remained significant after controlling for known risk factors. In addition, rs643555T risk allele was shown to upregulate the expression of CDH2, which was then linked to unfavorable BCR-free survival outcomes, further supporting for the biological plausibility of our findings.

HaploReg identified that CDH2 rs643555, an intronic variant, may be functional via a direct eQTL regulating the expression of CDH2. Several other potential causal variants in the proxy of rs643555 were also predicted to locate in enhancer histone marks and have regulatory effects on CDH2 through eQTL. According to GTEx dataset, the risk allele T of rs643555 was correlated with increased expression of CDH2. CDH2, also known as N-cadherin, is a transmembrane glycoprotein that mediates calcium-dependent cell adhesion and is mainly expressed in multiple cell types, including nerve, vascular, myocardial, and mesenchymal cells. Evidence suggests that increased expression of CDH2 together with the loss of E-cadherin (cadherin switching) plays an essential role during the progression of several human cancers, including prostate cancer. More importantly, aberrant CDH2 expression has been reported, not only in metastatic but also in castration-resistant prostate cancer. Mechanistic studies demonstrated that CDH2 promotes prostate cancer cells EMT, stemness, and metastatic ability by activating the ErbB signaling pathway. Monoclonal antibodies against CDH2 inhibited androgen-independent growth, local invasion, and metastasis in castration-resistant prostate cancer models. Furthermore, a CDH2 antagonist, ADH-1, is currently being used in clinical trials for treatment of CDH2-expressing solid tumors. Together, this evidence further supports the hypothesis that CDH2 could be a promising therapeutic target for prostate cancer.

There are some inherent limitations in the present study. First, both study populations are Taiwanese; therefore, our findings may not be generalized to other ethnic groups. Second, we are unable to explore the biological mechanisms behind the SNPs of cell adhesion pathway genes and disease progression because the prostate cancer tissues from study participants were unavailable. Third, we used haplotype-tagging SNPs to capture most of the genomic diversity, but the linked causal SNPs need to be further identified. Finally, the sample size of both cohorts is relatively small and does not have optimal power for discovering and replicating the associations. We had
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CONFLICT OF INTEREST

None declared.

DATA AVAILABILITY

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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SUPPORTING INFORMATION

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