Antioxidant and Immunomodulatory Activity of Hydroalcoholic Extract and its Fractions of Leaves of *Ficus benghalensis* Linn.

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**ABSTRACT**

**Background:** *Ficus benghalensis* is a folk medicine indigenous plant of India. Several studies on this plant focused and reported on the biological profile of the plant. **Objectives:** This study is aimed to evaluate the antioxidant and immunomodulatory activity of *F. benghalensis* leaf extract using various *in vitro* screening methods of both parameters. **Materials and Methods:** Hydroalcoholic (FB1) extract and its four fractions viz. n-hexane (FB2), n-butanol (FB3), chloroform (FB4), and water (FB5) of leaves of *F. benghalensis* investigated for their free radical scavenging activity using 1,1-diphenyl-2-picrylhydrazyl and 2,2'-azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) radicals. A dose-response curve was plotted and IC$_{50}$ values were determined to assess antioxidant activity. Nitroblue tetrazolium test, phagocytosis of killed *Candida albicans* and candidal assay were carried out to assess the immunomodulatory activity. Positive non-lymphoid cell number, mean particle number of killed *C. albicans*, percent value of killed *C. albicans* by neutrophils were calculated and presented. **Results:** Each extract showed antioxidant and prominent immunomodulatory activity with compared to standard. **Conclusions:** Hydroalcoholic (FB1) extract and its four fractions viz. n-hexane (FB2), n-butanol (FB3), chloroform (FB4), and water (FB5) showed promising antioxidant and immunomodulatory activity.

**Key words:** Antioxidant, *Candida albicans*, *Ficus benghalensis*, Free radicals, Immunomodulatory

**SUMMARY**

- Hydroalcoholic extract and its fractions of *F. benghalensis* Linn exhibited different DPPH and ABTS scavenging activity in concentration dependent manner.
- The extract, fractions and reference antioxidants showed DPPH scavenging effect in the order of Vit-C > Quercetin > FB2 > FB1 > FB5 > FB3 > FB4 and ABTS scavenging effect in the order of Vit-C > Quercetin > FB1 > FB5 > FB3 > FB2 > FB4.
- FB2 and FB3 showed promising immunomodulatory activity at all concentrations.

**INTRODUCTION**

Immune system dysfunction is responsible for various diseases such as arthritis, ulcerative colitis, asthma, allergy, parasitic diseases, cancer, and infectious diseases.[1] The degree to which the patient becomes abnormally susceptible to infections by the microbial environment depends on the extent of immunosuppression. The suppression of the immune system is characterized by a reduction in the number and phagocytic function of the neutrophils and macrophages, as well as an impairment of the intracellular bactericidal capacity of these cells. This immunosuppression allows opportunistic pathogens to overwhelm the host to cause secondary infections.[1] This problem can be overcome by boosting the immune system by the use of immunomodulatory drugs.[3] Immunomodulators improves the host defense mechanism.

According to Ayurveda, rasayana, is the drug used for enhancement of body resistance against infection acts as an adaptogen, immunomodulator, and antimutagenic.[4,5] Chemotherapeutic agents available today have mainly immunosuppressive activity. Most of them are cytotoxic and exerts a variety of side effects. This has given rise to stimulation in the search for investigating natural resources showing immunomodulatory activity. Many plants used in traditional medicine were found to have immunomodulatory properties it could be because of effects of phytoconstituents such as phenolics, terpenoids, steroids, flavonoids, etc. The endogenous antioxidant system prevents the deleterious influence of the free radical on the immune cells and preserves their normal function. Impairment in the immune system leads to overutilization of endogenous antioxidant.[6]

Many *Ficus* species have long been used in folk medicine as astringents, carminatives, stomachics, vermicides, hypotensives, anthelmintics, and anti-dysentery drugs.[6] It is believed that some *Ficus* species can be used as a remedy for visceral obstructive disorders, diabetes, leprosy, respiratory disorders and certain skin diseases[7] and as an absorbent for inflammatory swellings and burns.[8] *Ficus benghalensis* belongs to the family *Moraceae*, which is commonly known as Banyan tree. In earlier study glucoside, 20-tetraatriaconthene-2-one, bistriazine, and the new creations are licensed under the identical terms.

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**Cite this article as:** Bhanwase AS, Alagawadi KR. Antioxidant and Immunomodulatory activity of Hydroalcoholic extract and its fractions of leaves of *Ficus benghalensis* Linn. Phcog Res 2016;8:50-5.
6-heptatriacontene-10-one, pentatriacontan-5-one, beta-sitosterolalpha-D-glucose, and meso-inositol have been isolated from the bark of *F. benghalensis*. Leaves contain crude protein 9.63%, crude fibres-26.84%, CaO-2.53%, and phosphorus-0.4%. It yields latex containing caooychoue (2.4%), resin, albumin, cerin, sugar, and malic acid. It is used in Ayurveda for the treatment of diarrhea, dysentery and piles,[9,10] teeth disorders,[11] rheumatism, skin disorders such as sores,[12] to boost immune system,[13] and as a hypoglycemic.[14-15] The extracts of *F. benghalensis* were also reported to inhibit insulinase activity from liver and kidney,[16] fruit extracts exhibited anti-tumor activity in the potato disc bioassay.[17] Two flavonoid compounds, viz. 5, 7-dimethyl ether of leucopelargonidin 3-0-alpha-Lrhamnoside and 5, 3'-dimethyl ether of luecocyandin 3-0-alpha-D-galactosyl cellobioside were obtained from the bark of *F. benghalensis* and were evaluated for anti-oxidant activity in hyperlipidemic rats.[18] It was also found to inhibit the lipid peroxidation.[19] Various extracts of *F. benghalensis* was screened for its anti-allergic and anti-stress potential in asthma by milk induced leukocytosis and milk induced eosinophilia.[20-21] In Ayurveda, *F. benghalensis* has been reported for their skin disorder, rhumatitis, diabetes, anti-septic, aphrodisiac, hemostatic, nerve disorder, and menorrhagia. This plant is used throughout India for the purpose of complication related to blood, bone, endocrine system, toothache, gastric disorder, reproductive disorder, urinary disorder, and fever.

*F. benghalensis* is a holy plant largely found in the region of the forest of Deccan, and Southern India. It is also known for the tribal area of Maharashtra for their holiness and medical uses. This is evergreen plant having large source of leaves; Nalpamaram is an important group of Ayurvedic formulation that constitutes bark of *F. benghalensis* used in the treatment of skin disease and other ailments. In view of the medical importance and literature, the present study is aimed to evaluate *F. benghalensis* leaves for antioxidant and immunomodulatory activity.

**MATERIALS AND METHODS**

**Collection of plant material**

Fresh leaves of *F. benghalensis* Linn. were collected from the Western Ghat of Belgaum region in Karnataka. The plant was identified and authenticated by Regional Medical Research Center (RMRC), Belgaum. The herbarium was prepared and deposited at RMRC, Belgaum, Karnataka with voucher specimen No. RMRC-918.

**Preparation of extracts**

Hydroalcoholic extract of leaves of *F. benghalensis* Linn. was prepared from 500 g of leaf powder by maceration at room temperature using 2000 mL 50% v/v ethanol for 7 days with occasionally shaking. The extract was evaporated and concentrated by using rotary vacuum evaporator (IKARV 10 digital) to get semisolid mass further drying was done by vacuum oven. Semisolid extract was further successively extracted with n-hexane, n-butanol, chloroform and water. All extracts were stored in desiccator until further use.

**Chemicals and reagents**

All the chemicals and solvents are of analytical grade and obtained from Qualigens and Sigma-Aldrich. 1-1-diphenyl-2-picrylhydrazyl (DPPH) radical, 2, 2’azino-bis (3-ethylbenzthiazoline-6-sulphonic acid) (ABTS) radical, potassium persulfate, nitroblue tetrazolium (NBT), *Candida albicans* suspension, hanks balanced salt solution, sodium deoxy cholate, methylene blue solution, supplemented minimum essential media, and wright stain.

**Antioxidant activity**

Antioxidant property of the extracts and fractions of leaves of *F. benghalensis* Linn. was investigated by using DPPH and ABTS radical scavenging activity.

1. **1, 1-diphenyl-2-picrylhydrazyl radical scavenging activity**

A solution of DPPH was prepared by dissolving 25 mg of DPPH in 10 mL of methanol and the solution kept in the dark at 4°C. Stock solution of extract and fractions of concentration 1,000 µg/mL were prepared. A volume of 100 µL of stock solution (1,000 µg/mL) was added to the well and it was further down diluted to the lowest concentration (7.8 µg/mL) by performing a serial 2-fold dilution in a 96-well microtiter plate. A row of negative DPPH control was developed in the same 96-well microtiter plates by adding 100 µL of methanol into each well. Quercetin and Vitamin C were used as positive controls. Five microliters of methanolic DPPH were added into each well and reaction was allowed to proceed for 30 min in the dark. The absorbance was measured at 492 nm by a microplate reader.[24] The DPPH radical scavenging effect (%) was calculated using the formula:

\[
\text{DPPH radical Scavenging effect } (\%) = \frac{\text{OD (DPPH)} - \text{OD (DPPH + Sample)}}{\text{OD (DPPH)}} \times 100
\]

2. **2’,2’-azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) radical scavenging activity**

The ABTS radical was prepared by reacting equal volumes of 1.1 mg/mL aqueous ABTS and 0.68 mg/mL potassium per sulfate (K₃S₄O₆) and stored in dark for 6 h at room temperature. A stock solution of extracts and fractions at a concentration of 1,000 µg/mL was prepared. A volume of 100 µL of stock solution (1,000 µg/mL) was added to the well and it was further down diluted to the lowest concentration (7.8 µg/mL) by performing a serial 2-fold dilution in a 96-well microtiter plate. A row of negative ABTS control was developed in the same 96-well microtiter plates by adding 100 µL of methanol into each well. Quercetin and Vitamin C were used as positive controls. Twenty-five microliters of ABTS radical were added into each well and reaction was allowed to proceed for 30 min in the dark. The absorbance was measured at 734 nm by a microplate reader.[24] The ABTS radical scavenging effect (%) was calculated using the formula:

\[
\text{ABTS radical Scavenging effect } (\%) = \frac{\text{OD (ABTS)} - \text{OD (ABTS + Sample)}}{\text{OD (ABTS)}} \times 100
\]

**In vitro immunomodulatory activity**

NBT screening test, phagocytosis of killed *C. albicans* and candididial assay were carried out at KLE University's Dr. Prabhakar Kore Basic Science Research Center, Belgaum to assess *in vitro* immunomodulatory activity. Ethical clearance was obtained from the Research and Ethical Committee, KLE University's V.K. Institute of Dental Sciences, Belgaum prior to study.

**Nitroblue tetrazolium screening test**

One part of 0.3% NBT solution was prepared in 0.35% sucrose solution and was used fresh. Hundred microliters of whole blood and 100 µL of Hank’s Balanced Salt Solution (HBSS) were added to each Eppendorf tube and mixed well. The stock solution of hydroalcoholic extract and its fractions of leaves of *F. benghalensis* Linn. in different concentrations of 1,000 µg/mL, 500 µg/mL, 250 µg/mL, 125 µg/mL, 62.5 µg/mL were added individually to tubes containing suspension of blood cells and 50 µL of NBT solution. In another tube 50 µL of *Escherichia coli* endotoxin
was added to 50 μL NBT solution and suspension of blood cells, which served as positive control (standard). A normal control was maintained in another tube with only suspension of blood cells and NBT solution. All tubes were incubated at 37°C for 20 min and were gently shaken at room temperature for 20 min. A smear of fluid from each tube was taken on microscope slide, air dried, fixed in methanol for 2 min, stained with Giemsa stain for 15 min, rinsed with distilled water, dried, and observed under microscope. Hundred neutrophils were counted and percentage of NBT-positive cells containing blue deposits was determined.^[25]\(^2\)

**Phagocytosis of killed Candida albicans Preparation of Candida albicans suspension**

The *C. albicans* culture was incubated in Sabouraud broth overnight and then centrifuged to form a cell button at the bottom and supernatant was discarded. The cell button was washed with sterile HBSS and centrifuged again. This step was repeated 3–4 times. The final cell button was mixed with a mixture of sterile HBSS and human serum in the proportion of 4:1. The cell suspension of concentration 1 × 10⁶/mL was used for the experiment.

**Phagocytosis evaluation**

Fifty microliters of concentrations of the hydroalcoholic extract and its fractions of leaves of *F. benghalensis* Linn. (1,000 μg/mL, 500 μg/mL, 250 μg/mL, 125 μg/mL, 62.5 μg/mL) were taken in separate Eppendorf tubes and 100 μL of HBSS, 100 μL of *C. albicans* suspension and 100 μL of leucocytes suspension (2 × 10⁶/mL) were added to each tube. A positive control (standard) and a normal control were maintained. All the tubes were shaken gently, incubated at 37°C for 30 min and centrifuged at 200 g for 5 min. The supernatant was removed leaving a small droplet into which sediment was resuspended. Smears were made, air-dried and stained with Giemsa stain. Two hundred neutrophils were observed under a microscope, a number of ingested *C. albicans* associated with each cell was counted and mean particle number (MPN) associated with each neutrophil was calculated.^[26]\(^2\)

**Candidacidal assay**

Fifty microliters each of different concentration of hydroalcoholic extract and its fractions of leaves of *F. benghalensis* were taken in separate Eppendorf tubes and 100 μL of HBSS, 50 μL of *C. albicans* suspension (1 × 10⁶/mL) and 100 μL of leucocytes suspension (7 × 10⁶/mL) were added to each tube. In another tube, 50 μL of pooled serum was added to 100 μL of HBSS, 50 μL of *C. albicans* suspension and 100 μL of leucocytes suspension which served as positive control (standard). A normal control was maintained in another tube containing leucocytes suspension, HBSS and *C. albicans* suspension. Each tube was incubated at 37°C for 60 min with shaking every 15 min. After 30 min, 100 μL solutions of 0.5% sodium deoxycholate were added to each tube and mixed causing lyses of leucocytes without damage to *Candida* cells. One milliliter of 0.01% methylene blue was added to each tube, mixed and centrifuged at 1,500 g for 5 min. The supernatant containing methylene blue was carefully removed with Pasteur pipette leaving behind droplet to resuspend the organism. The suspension was put in ice bath until ready for counting. The suspension was observed on Neubauer counting chamber under a microscope. The proportion of dead cells (stained blue) was determined.^[26]\(^2\)

**Statistical analysis**

The values were expressed as mean ± standard error of the mean (n = 3). The results were analyzed using one-way analysis of variance followed by Dunnet’s t-test to determine the statistical significance.^[27]\(^2\)

**RESULTS**

Extraction of leaves of *F. benghalensis* with cold maceration gives hydroalcoholic extract 25% w/w, and successive fractions of n-hexane, n-butanol, chloroform and water 7% w/w, 2.8% w/w, 1.5% w/w and 14% w/w, respectively.

1-1-diphneyl-2-picrylhydrazyl radical scavenging activity

The free radical scavenging activity of hydroalcoholic extract and its different successive fractions of a leaf of *F. benghalensis* was determined according to the DPPH radical scavenging method and is shown in Figure 1. According to this method, a compound with high antioxidant activity effectively bind with the radical hence prevent its propagation and the resultant chain reaction. Different concentrations of five different samples, that is, hydroalcoholic extract (FB1), n-hexane fraction (FB2), n-butanol fraction (FB3), chloroform fraction (FB4) and water fraction (FB5) were analyzed. The results were expressed as IC₅₀ values and calculated from regression lines as presented in Table 1.

2, 2’-azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) radical scavenging activity

The free radical scavenging activity of hydroalcoholic extract and its different successive fractions of a leaf of *F. benghalensis* was determined according to the ABTS radical scavenging method and is shown in Figure 2. According to this method, a compound with high antioxidant activity effectively bind with the radical hence prevent its propagation and the resultant chain reaction. Different concentrations of five different extracts, that is, hydroalcoholic (FB1), n-hexane (FB2), n-butanol (FB3), chloroform (FB4) and water (FB5) extracts were analyzed. The results

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**Table 1: IC₅₀ values of hydroalcoholic extract and its fractions of FB leaves and reference compounds for scavenging of DPPH free radicals**

| Sample               | IC₅₀ (μg/ml) |
|----------------------|-------------|
| Vitamin-C            | 11.5±0.052  |
| Quercetin            | 15.4±0.120  |
| Hydroalcoholic (FB 1)| 32.3±1.320  |
| n-Hexane (FB 2)      | 28.2±0.993  |
| n-Butanol (FB 3)     | >1000       |
| Chloroform (FB 4)    | >1000       |
| Water (FB 5)         | 125.0±0.025 |

DPPH: 1-1-diphenyl-2-picrylhydrazyl; FB: Ficus benghalensis
were expressed as IC\textsubscript{50} values and calculated from regression lines as presented in Table 2.

**Nitroblue tetrazolium test**

The hydroalcoholic (FB1) extract of leaves of *F. benghalensis* has stimulated the neutrophils to phagocytic activity to the extent of 45% at concentration of 1,000 µg/mL and 41.66% at concentration of 500 µg/mL very significantly. n-Hexane (FB2) fraction showed 80.33% at concentration of 1,000 µg/mL, 78.00% at concentration of 500 µg/mL, 76.66% at concentration of 250 µg/mL, 44.00% at concentration of 125 µg/mL. n-Butanol (FB3) fraction showed very highly significant result at all concentrations. Chloroform (FB4) fraction has stimulated the neutrophils to phagocytic activity to the extent of 55% at concentration of 1,000 µg/mL, 54.00% at concentration of 500 µg/mL and 36% at concentration of 250 µg/mL, while water (FB5) fraction does not show significant activity. The results are presented in Table 3.

**Phagocytosis of killed *Candida albicans***

The hydroalcoholic (FB1) extract and its four fractions n-hexane (FB2), n-butanol (FB3), chloroform (FB4) and water (FB5) of leaves of *F. benghalensis* stimulated the phagocytosis of killed *C. albicans*. The MPN was found to be 5 at concentration of 1,000 µg/mL for hydroalcoholic (FB1) extract; 5, 5, 4, 3-4 for n-hexane (FB2) fraction; 5-6, 5, 4, 3-5, 3 for n-butanol (FB3) fraction at down diluted concentrations from 1,000 µg/mL to 62.5 µg/mL; 4, 3-4 for Chloroform (FB4) fraction at concentration of 1,000 µg/mL and 500 µg/mL, respectively. Results are compared with positive control (standard) that is pooled serum (6, 5-6, 5, 5, and 4 at down diluted concentrations from 1,000 µg/mL to 62.5 µg/mL). Water (FB5) fraction does not show significant stimulation of phagocytic activity. The results are represented in Table 4.

**Candidacial assay**

The hydroalcoholic (FB1) extract of leaves of *F. benghalensis* has shown significant candidacidal activity to the extent of 29% at concentration of 1,000 µg/mL; n-hexane (FB2) fraction has shown 36.33% and 32.66% at concentrations of 1,000 µg/mL and 500 µg/mL respectively. n-Butanol (FB3) fraction has shown 35.33% and 33% at concentration of 1,000 µg/mL and 500 µg/mL respectively when compared to standard, that is pooled serum (40% and 37.33%) and to normal control (21.66% and 19.66%) at the same concentrations. Chloroform (FB4) fraction and water (FB5) fraction does not showed significant stimulation of candidacidal activity. The results are represented in Table 5.

**DISCUSSION**

Antioxidant activity usually means the ability of a compound to delay, inhibit or prevent the oxidation of oxidizable materials by scavenging free radicals and reducing oxidative stress. Oxidative stress has been implicated in the pathology of many diseases and condition, including diabetes, cardiovascular disease, inflammatory condition, cancer, ageing, etc. The antioxidant may offer resistance against the oxidative stress by scavenging the free radicals, inhibiting lipid peroxidation and by many other mechanisms and thus prevents diseases.

The DPPH, which possesses an unpaired electron and exhibits a stable violet color in methanol solution (peak absorbance at 517 nm), is commonly used as a reagent for evaluation of the free radical scavenging activity of antioxidants. The DPPH assay is based on the reduction of 2′-azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) (ABTS) in the presence of a hydrogen-donating antioxidant due to the formation of the nonradical form (DPPH-H).

Table 2: IC\textsubscript{50} values of hydroalcoholic extract and its fractions of FB leaves and reference compounds for scavenging of ABTS free radicals

| Sample                  | IC\textsubscript{50} value (µg/ml) |
|-------------------------|-----------------------------------|
| Vitamin-C               | 6.40±0.015                        |
| Quercetin               | 7.05±0.115                        |
| Hydroalcoholic (FB 1)   | 52.0±0.722                        |
| n-Hexane (FB 2)         | 58.2±0.714                        |
| n-Butanol (FB 3)        | 491±0.555                         |
| Chloroform (FB 4)       | >1000                             |
| Water (FB 5)            | 20.3±0.133                        |

ABTS: 2′-azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) assay. Results are mean ± standard error of the mean (n = 3)

Table 3: Percentage of reduced neutrophils after treatment with extract and fractions of FB by NBT test

| Sample                        | 1000          | 500           | 250           | 125           | 62.5          |
|------------------------------|---------------|---------------|---------------|---------------|---------------|
| Blood without endotoxin      | 20.66±1.20    | 21.33±0.57    | 21±1.15       | 20±0.57       | 22±0.57       |
| Blood with endotoxin         | 92.33±1.45    | 91.66±1.20    | 88.66±0.88    | 88±1.35       | 87.66±0.33    |
| Hydroalcoholic (FB 1)        | 45.0±0.57**   | 41.66±0.88**  | 25.0±2.30**   | 21±0.57***    | 20.3±0.66**   |
| n-Hexane (FB 2)              | 80.33±0.66*** | 78±0.57***    | 76.66±0.33*** | 44±0.57***    | 26.66±2.84**  |
| n-Butanol (FB 3)             | 89.66±0.33*** | 83.5±2.85***  | 82.66±0.88*** | 61±1.00***    | 32.66±0.88**  |
| Chloroform (FB 4)            | 55±1.00***    | 54±0.57***    | 36±0.57***    | 26±1.52***    | 21.33±0.88    |
| Water (FB 5)                 | 20.3±0.33**   | 20.5±0.70**   | 18±0.66**     | 18±0.00**     | 16.6±0.63     |

The data are expressed as mean percentage reduced neutrophils±SEM. Significant difference from positive control (standard) by one-way ANOVA followed by Dunnett’s t-test. (n=3). *P<0.05, **P<0.01, ***P<0.001. Blood without endotoxin (normal control), blood with endotoxin (positive control). **Not significant, NBT: Nitroblue tetrazolium; SE: Standard error of mean; FB: *Ficus benghalensis*; ANOVA: Analysis of variance
in the reaction. Figure 1 shows the DPPH scavenging activities of five different extracts, that is hydroalcoholic (FB1), n-hexane (FB2), n-butanol (FB3), chloroform (FB4), water (FB5) and reference antioxidants at down diluted concentrations from 1000 μg/ml to 7.8 μg/ml. The extracts exhibited different DPPH scavenging activities in a concentration-dependent manner. The scavenging effects of extracts and reference antioxidants on DPPH observed in the following order: Vitamin C > quercetin > FB2 > FB1 > FB5 > FB4 > FB3. The IC_{50} of various fraction and reference antioxidants are listed in Table 1. FB1, FB2 exhibited effective radical scavenging activity while FB5 showed less activity. FB3 FB4 showed very weak activity. The ABTS radical, which has a peak absorbance at 734 nm, should be performed by mixing ABTS and K₅⁵O₃. When antioxidants were added, the ABTS radical, which has a blue-green color, is reduced to ABTS (no color). Figure 1 shows the ABTS scavenging activities of five different extracts, that is hydroalcoholic (FB1), n-hexane (FB2), n-butanol (FB3), chloroform (FB4), water (FB5) and reference antioxidants at different concentrations (7.8–1,000 μg/ml). The extracts exhibited different ABTS scavenging activities in a concentration-dependent manner. The scavenging effects of extracts and reference antioxidants on ABTS decreased in the following order: Vitamin C > Quercetin > FB1 > FB2 > FB5 > FB4 > FB3. The IC_{50} of extract, fractions, and reference antioxidants are listed in Table 2. FB1 FB2 exhibited very strong radical scavenging activity while FB3, FB4 and FB5 showed weak activity.

Immunity plays an important role in the protection of the human body from foreign toxic matter. Many plants from Ayurveda have been used for the purpose of increase immunity. Immunomodulatory agents of plant and animal origin increase the immune responsiveness of the body against pathogens by activating the nonspecific immune system. However, there is a need to systemic studies on medicinal plants to substantiate the therapeutic claims made regarding their clinical utility. In this study, the hydroalcoholic (FB1) extract and its four fractions n-hexane (FB2), n-butanol (FB3), chloroform (FB4) and water (FB5) of leaves of *F. benghalensis* significantly increased the phagocytic function of human neutrophils, when compared with control indicating the possible immunostimulating effect. The engulfment of microorganisms by leukocytes called phagocytosis, which is one of the main defense mechanisms of an organism. Hydroalcoholic extract and its fractions *F. benghalensis* have significantly increased the intracellular reduction of NBT dye to formazan (deep blue compound) by the neutrophils confirming the intracellular killing property and overall metabolic integrity of phagocytizing neutrophils.

Phagocytosis of killed *C. albicans* and candidacidal assay has supported the immunomodulatory activity of *F. benghalensis*, claimed as an enhancer of general immunity against various physical and mental disorders in the indigenous system of medicine.

**CONCLUSIONS**

The results clearly indicate that the leaves of *F. benghalensis* possess immune boosting properties and suggest usefulness in the disorder of immunological origin where the antioxidant system is adversely affected. Further study regarding the isolation of active chemical constituents responsible for immunomodulatory activity and its mode of action need to be determined.

**Acknowledgments**

The authors are thankful to Dr. Prabhakar Kore Basic Science Research Center, Belgaum for providing research facilities.

**Financial support and sponsorship**

Nil.

**Conflicts of interest**

There are no conflicts of interest.

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Pharmacognosy Research, Vol 8, Issue 1, Jan-Mar, 2016