Comparative Leaf Anatomy and Micromorphology of *Thunbergia erecta* (Benth.) T. Anderson and *Thunbergia laurifolia* Lindl. in Peninsular Malaysia

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Highlights

- Five common characteristics of *Thunbergia erecta* and *Thunbergia laurifolia* were the presence of raphide, sinuous anticlinal walls, diacytic stomata, majority opened and minority closed venation in lamina and the presence of peltate glandular (unicellular terminal) trichome.

- Several variable characteristics to distinguish *Thunbergia erecta* and *Thunbergia laurifolia* were petiole and marginal outlines, types of vascular bundles, the presence of druse, marginal venation, stomata occurrence, types of wax, cuticular sculpturing and types of trichomes.

- Leaf anatomy and micromorphology are proven to be additional tools to provide useful information for the identification of *Thunbergia* species.
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Abstract: Comparative leaf anatomy and micromorphology study was carried out on two selected species from the genus *Thunbergia* Retz. of Acanthaceae subfamily Thunbergioideae. These two investigated species were *T. erecta* and *T. laurifolia* from Peninsular Malaysia. The leaf anatomical study involve several methods such as cross-section using sliding microtome on the petioles, midribs, lamina and marginal, leaf epidermal peeling, leaf clearing and observation under a light microscope. The leaf micromorphology method involve the observation under a scanning electron microscope (SEM). This study aimed to investigate the taxonomic value of leaf anatomy and micromorphology characteristics of genus *Thunbergia*. The results have shown that there were five common characteristics present in both species studied and several variable characters that might be useful for species differentiation of *T. erecta* and *T. laurifolia*. The five common characteristics recorded were the presence of raphide, sinuous anticlinal walls, diacytic stomata, majority opened and minority closed venation in lamina and the presence of peltate glandular (unicellular terminal) trichome. The variable characteristics included were petiole, and marginal outlines, types of vascular bundles, the presence of druse, marginal venation, stomata occurrence, types of wax, cuticular sculpturing and types of trichomes. In conclusion, findings in this study showed that leaf anatomical and
micromorphological characteristics possessed taxonomic value that can be used in the species identification for the genus Thunbergia specifically for T. erecta and T. laurifolia.

**Keywords:** Leaf Anatomy, Leaf Micromorphology, Taxonomic Significance, Thunbergia

**Abstrak:** Kajian perbandingan ciri anatomi dan mikromorfologi daun telah dijalankan ke atas dua spesies terpilih dalam genus Thunbergia Retz. (Acanthaceae) subfamili Thunbergioideae. Dua spesies yang telah dikaji ialah T. erecta dan T. laurifolia dari Semenanjung Malaysia. Kajian anatomi daun ini melibatkan beberapa kaedah seperti kaedah hirisan dengan mikrotom gelongsor pada bahagian petiol, tulang daun, lamina dan tepi daun, kaedah siasatan epidermis, kaedah penjerminan dan cerapan di bawah mikroskop cahaya. Kajian mikromorfologi melibatkan cerapan di bawah mikroskop imbasan elektron. Kajian ini bertujuan untuk melihat nilai taksonomi ciri anatomi dan mikromorfologi daun dalam genus Thunbergia. Hasil kajian menunjukkan terdapat lima ciri sepunya ke atas spesies yang dikaji dan beberapa ciri variasi yang boleh digunakan untuk pembezaan di antara T. erecta dan T. laurifolia. Lima ciri sepunya yang telah direkodkan ialah kehadiran rafid, dinding antiklinal sinuous, stomata diasitik, peruratan lamina daun majoriti terbuka dan minoriti tertutup serta kehadiran trikom kelenjar peltat (terminal unisel). Ciri variasi termasuklah bentuk petiol dan tepi daun, jenis tisu vaskular, kehadiran drus, kehadiran stomata, peruratan tepi daun, jenis lilin, ukiran kutikel dan jenis trikom. Kesimpulannya, hasil kajian menunjukkan ciri anatomi dan mikromorfologi daun mempunyai nilai taksonomi yang boleh digunakan dalam pengecaman spesies bagi genus Thunbergia khususnya pada T. erecta dan T. laurifolia.

**Kata kunci:** Anatomi Daun, Mikromorfologi Daun, Nilai Taksonomi, Thunbergia

**INTRODUCTION**

The Acanthaceae family is one of the plant families that belong to the order Lamiales. According to Wasshausen and Wood (2004), the Acanthaceae is a large and diverse pantropical family consist of approximately 240 genera and 3,250 species. Recently, the systematic position of Acanthaceae is divided into four subfamilies which are; Acanthoideae, Nelsonioideae, Thunbergioideae and Avicennioideae (Stevens 2016). The Acanthaceae is mainly herbaceous and shrubs but sometimes found as climbers or lianas especially in the genus Thunbergia, while rarely found as woody plants or trees (Metcalfe & Chalk 1965; Carlquist 1988; Scotland et al. 1995; Vollesen 2008). One of the most important criteria in recognising the Acanthaceae is the occurrence of cystoliths. However, Heywood et al. (2007) reported on the absence of cystoliths in the vegetative parts of subfamilies Nelsonioideae, Thunbergioideae and the tribe of Acanthae.

*Thunbergia* Retz. is a large genus of the Acanthaceae subfamily Thunbergioideae (Takhtajan 1997). The genus *Thunbergia* is comprised of more than 100 species (Chia-chi et al. 2011) distributed in the tropical and subtropical regions of Africa, Madagascar, Asia and Australia (Borg et al. 2008). Kar et al. (2013) explained that *Thunbergia* was named in 1780 by Retzius, in the honours of Carl Peter Thunberg (1743–1828), a Swedish botanist, doctor and naturalist.
Most of the *Thunbergia* species are known as clock vine which referred to its clockwise twinning habit (Retief & Reyneke 1984). The Asia *Thunbergia* is taxonomically characterised by a few morphological characters such as perennial herbaceous or woody climbers, shrubs, rarely erect or trailing herbs without the occurrence of cystoliths (Suwanphakdee & Vajridaya 2018). To be noticed, many of the *Thunbergia* species preferred full sun exposure and well-drained soil but somehow can bloom in the partial shady areas (Sultana et al. 2015).

Moreover, the leaves are usually simple, petiolate, opposite arranged, ovate or lanceolate to hastate or sagitate shaped and the margins are entire to lobed to dentate. Borg et al. (2008) also mentioned that this genus is differed from other members of the family by their reduced calyx and enlarged bracteoles. Apart, seeds of the Acanthaceae are usually borne on hook-like retinacula that attached to the septa of the capsules but sometimes lacking in *Thunbergia* (Chia-chi et al. 2011). The present research thereby deals with two species of *Thunbergia* namely; *Thunbergia erecta* (Benth.) T. Anderson and *Thunbergia laurifolia* Lindl., which can be differentiated obviously by the characteristics of their floral part. *T. erecta* showed a solitary type of inflorescence with purplish-blue corolla, whereas *T. laurifolia* as in raceme inflorescence with blue-violet corolla. However, taxonomists might face difficulty to identify the species if the absence of the floral part since both *Thunbergia* species studied shared similarities in the vegetative parts. Even Nurul-Aini et al. (2018) mentioned that the identification and classification of the Acanthaceae species is quite challenging due to morphological similarities shared with other species in the same genus. Leaf anatomy and micromorphology hence are very useful tools to provide the additional data for the identification purposes as well to support the taxonomic study of the species, especially when the absence of floral and fruiting materials (Khatijah & Ruzi 2006).

According to Sultana et al. (2015), most of the *Thunbergia* species possessed the ornamental values because of their attractive flowers and twinning characters. Previous studies also reported on the utilisation of certain *Thunbergia* species in India for medicinal purposes. For instance, *Thunbergia grandiflora* has been useful in treating minor eyesore and against snake bite as reported by Teron (2005), whereby Nath and Dutta (2010) explained on the uses of *T. grandiflora* stem juice against conjunctivitis among the indigenous tribe. Also, Sarma (2006) reported on the usage of *Thunbergia coccinea* roots against dysentery, stomach ache and fever. Additionally, Aritajat et al. (2004) reported on the usage of *T. laurifolia* in treating diabetes among Thai traditional healers. Besides, the herbal teas and capsules of *T. laurifolia* are produced and marketed by the local companies in Thailand (Chan et al. 2011). The growing interest in the potential medicinal sources from the genus *Thunbergia* in India and Thailand is considered as a good starting point for pharmacological research. However, the lack of attempt is reported on the potential medicinal sources from the genus *Thunbergia* in Peninsular Malaysia. Therefore, this research is expected to be a good platform to encourage more discoveries on the potential medicinal plants in Peninsular Malaysia. In fact, the present
comparative study is significant to avoid misidentification of *Thunbergia* species that might lead to incorrect harvesting of raw materials and drug authentication, especially to produce medicines. Thereby, this present study is conducted to investigate the comparative leaf anatomy and micromorphology of two selected *Thunbergia* species; *T. erecta* and *T. laurifolia* in Peninsular Malaysia, and later determine whether they possessed taxonomic values that might be useful in the identification and classification of *Thunbergia*, especially at the species level. Also, this research aims to contribute to the knowledge and documentation of these selected *Thunbergia* species from Peninsular Malaysia.

**MATERIALS AND METHODS**

This study was carried out on two selected taxa from the genus *Thunbergia* of Acanthaceae subfamily Thunbergioideae which were; *T. erecta* and *T. laurifolia*. Fresh specimens were obtained from several forest reserves in Peninsular Malaysia such as in Hutan Lipur Lata Tembakah, Besut, Terengganu and Hutan Lipur Lata Kinjang, Perak. Three replicates of each plant species were collected throughout this research. The voucher specimens were deposited at Universiti Kebangsaan Malaysia Herbarium (UKMB) for future reference. Fresh specimens collected were fixed in 3:1 AA solutions (70% Alcohol: 30% Acetic Acid). For the leaf anatomical methods, part of petioles, midribs, leaf lamina and marginal were sectioned in a range of thickness (30 µm–40 µm) using sliding microtomes. The epidermal peels were conducted manually by scrapping off the underside of leaf surfaces by using a razor blade. The leaf clearing involved the immersion of leaf lamina and margin into the Basic Fuchsin solution. All the anatomical procedures including the staining and dehydration process followed the suitable modifications methods by Johansen (1940) and Saas (1958). Anatomical images were captured using a video (3CCD) camera attached to a Leitz Diaplan microscope using Cell^B software. For the leaf micromorphology method, the specimens were obtained from dried samples of herbarium. Samples lamina were cut about 1 cm². The specimens were then mounted on a mounting holder and coated with gold by using a sputter-coated machine. Micromorphological images were captured by using a scanning electron microscope Philips XL Series XL 30 with magnifications of 150×, 300×, 700× and 1,000×.

**RESULTS AND DISCUSSIONS**

The findings of this study showed the presence of five common characters of leaf anatomy and micromorphology among two selected *Thunbergia* species. The common characteristics are the presence of raphide in the parenchyma cortex of petiole and midrib (Fig. 1A), sinuous anticlinal walls on adaxial and abaxial surfaces (Fig. 1B), presence of diacytic stomata (Figs. 1C and 1D),
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lamina venation with majority opened and minority closed with swollen tracheids (Fig. 1E) and presence of peltate glandular trichomes (unicellular terminal) (Figs. 1F and 1G).

Figure 1: Common characteristics of leaf anatomy and micromorphology in Thunbergia (A) Presence of raphide, (B) Sinuous anticlinal walls on adaxial surface, (C) Sinuous anticlinal walls on abaxial surface, (D) Presence of diacytic stomata, (E) Lamina venation with majority opened and minority closed with swollen tracheid, (F and G) Presence of peltate glandular trichomes (unicellular terminal). Scales: (E) 500 µm; (B) 100 µm; (A, D, F and G) 50 µm; (C) 10 µm.

The variation characteristics examined are petiole outlines; horns-like shape at the right and left upper corners of adaxial surface with U-shaped of abaxial surface in T. erecta (Fig. 2A). While in T. laurifolia, lobes at the right and left upper corners of adaxial surface and flat in the middle of adaxial surface with ¾ round shaped of abaxial surface (Fig. 2B). Besides, the main vascular
bundle (opened system with continuous ring of vascular bundle) and additional vascular bundles (opened system with two continuous rings of vascular bundles) are recorded in petiole of *T. erecta* (Fig. 2C). Whereas main vascular bundle (closed system with continuous ring of vascular bundle) and additional vascular bundles (opened system with two continuous rings of vascular bundle) are observed in petiole of *T. laurifolia* (Fig. 2D). For marginal outlines; round-shaped with 90° curved in *T. erecta* (Fig. 2E) whilst round shaped and straight in *T. laurifolia* (Fig. 2F). The presence of druses also observed in lamina part of *T. erecta* (Fig. 2G). For marginal venation; complete venation is recorded in *T. erecta* (Fig. 2H) whereas incomplete venation in *T. laurifolia* (Fig. 2I). Four types of waxes present in this study which are; verrucate, flakes and granules observed in *T. erecta* (Fig. 2J) while crustose and granules in *T. laurifolia* (Fig. 2K). Anticlinal and periclinal walls can be slightly differentiated in *T. erecta* (Fig. 2L) but sunked anticlinal walls and raised periclinal walls are recorded for *T. laurifolia* (Fig. 2M). Stomata are absent on adaxial surface of *T. erecta* (hypostomatic) (Fig. 2N) yet present on adaxial surface of *T. laurifolia* (amphistomatic) (Fig. 2O).

Candolle (1879) described on the several fundamental concepts of petiole anatomy and distinguished between two systems of vascular bundles which are; open system and close system of vascular bundles. Even Hare (1942) emphasised on two types of vascular tissue existed in petiole which are open system (U-shaped) and close system (O-shaped). Previous studies on the petiole anatomy of some Philippines *Shorea* species also recognised the taxonomic distinction in petiole vascular patterns (Rojo 1987). Not only that, few studies affirmed that petiole anatomy can be used for grouping genera and identification of species (Agbagwa & Ndukwa 2004; Noraini & Cutler 2009). In fact, the petiole anatomy can be used to form groups in the genus *Dipterocarpus* (Ruzi et al. 2009). Recently, Nurul-Aini et al. (2018) also agreed on the significance of vascular bundles types in taxonomic study as in three selected Acanthaceae species studied, namely *Acanthus ebracteatus* (Vahl), *Andrographis paniculata* (Burm.f.) Wall. Ex Nees and *Chroesthes longifolia* (Wight) B. Hansen. The importance of midrib vascular bundles characterisctics also proven to be useful to identify and differentiate the genus and species as in Rubiaceae (Nurul-Syahirah et al. 2016). The present study hence reported on the main vascular bundle (opened system with continuous ring of vascular bundle) and additional vascular bundle (opened system with two continuous rings of vascular bundles) are recorded in petiole of *T. erecta*, whereas the main vascular bundle (closed system with continuous ring of vascular bundle) and additional vascular bundles (opened system with two continuous rings of vascular bundles) in *T. laurifolia*. Therefore, the character of vascular bundles might be useful and provide additional data to differentiate between these two species.
Figure 2: Variation characteristics of leaf anatomy and micromorphology in *Thunbergia* (A and B) Petioles outlines (C) Opened vascular bundles, (D) Closed vascular bundles, (E) Marginal outline; round shaped with 90° curved, (F) Marginal outline; round shaped and straight, (G) Presence of druses, (H) Complete marginal venation, (I) Incomplete marginal venation, (J) Types of wax; verrucate, flakes and granules, (K) Types of wax; crustose and granules, (L) Cuticular ornamentation; anticlinal and periclinal walls can be slightly differentiated, (M) Cuticular ornamentation; Sinked anticlinal walls; raised periclinal walls, (N) Stomata absent on adaxial surface (hypostomatic) and (O) Stomata present on adaxial surface (amphistomatic).

*Scales:* (H and I) 1000 µm; (A and B) 500 µm; (C, D and F) 200 µm; (E) 100 µm; (G) 50 µm; (K and M) 20 µm; (N and O) 10 µm; (J and L) 2 µm.
Recently, the leaf anatomy study conducted on selected Acanthaceae taxa in Peninsular Malaysia showed several characteristics that can be used in the identification of Acanthaceae including petiole and midrib outlines (Nurul-Aini et al. 2018). This present study hence proved that *Thunbergia* studied also can be distinguished by the petiole and marginal outlines. Petiole outlines in *T. erecta* appeared as a horns-like shape at right and left upper corners of adaxial surface; U-shaped of abaxial surface, whereby lobes at right and left upper corners and flat in the middle of adaxial surface; ¾ round-shaped of abaxial surface in *T. laurifolia*. The marginal outlines in *T. erecta* occurred in round-shaped with 90° curved, whereas round-shaped and straight in *T. laurifolia*. Findings on the taxonomic value of the petiole and marginal outlines in *Thunbergia* species also supported by Nurul-Aini et al. (2013) in the classification species from the genus *Microcos* (Tiliaceae) by using petiole outlines criteria.

Beck (2010) explained that protoplast consists of two major categories of waste metabolites which are; calcium oxalate crystals and tanniferous substances. According to Franceschi and Nakata (2005), the presence of these small crystals in higher plants is common and usually related to the physical protection, storage of calcium, removal of oxalate from the metabolic system and regulation of light during photosynthesis in plants that grow under shade. Noraini et al. (2019) even stressed that the occurrence of ergastic substances such as calcium oxalate and calcium carbonate crystals are usually related to the physiological activities of the plants. There are two typical types of crystals found in plants known as druse and raphide. The compact and spherical aggregates of angular crystals are known as druse, whereas the aggregations of needle-like crystal known as raphide (Beck 2010). However, the occurrence of crystals, distributions, or types of prismatic either druse or raphide can be used as a taxonomic character, especially for species identification. Metcalfe and Chalk (1985) mentioned the presence of raphide, druse, styloid, prismatic and sand-like crystal idioblasts in species of Rubiaceae. The presence of druse only in the lamina part of *T. laurifolia*, and raphide in the parenchyma cortex of *Thunbergia* studied thereby suggests that this might be a significant characteristic for the genus and also to differentiate these two species.

According to Cutler et al. (1978), the characters on leaf epidermis can be used as a diagnostic feature in the identification of plant species. Fahn (1967) explained that many of the epidermal cells appeared in tabular form by which mostly sinuous anticlinal walls occurred in the leaf blade of dicotyledons, whereby elongated in monocotyledons. Moreover, the value of the sinuous wall is reported to increase the tensile strength of the epidermis. For instance, the geometric features of the sinuous shape of *Dryopteris thelypteris* L. are reported to provide a more secure attachment between the epidermal and mesophyll cells (Korn 1976). The present study of *Thunbergia* species has recorded on the sinuous anticlinal walls of abaxial and adaxial surfaces, thus considered as a common character for the genus. Apart, the results of leaf surface studies also reported on the occurrence of hypostomatic leaves in *T. erecta* whereas amphistomatic leaves in *T. laurifolia*. Fahn (1967) explained that hypostomatic
leaves referred to the presence of stomata only on the upper surface of leaves, whereby stomata present on both sides of leaves are known as amphistomatic. The basis of leaf epidermal structure is important for species identification as reported by Nurul-Aini et al. (2014) on the presence of hypostomatic and amphistomatic leaves among eight chosen taxa of Acanthaceae. Even Robbrecht (1988) mentioned that hypostomatic leaves are common in Rubiaceae. Therefore, results on the occurrence of stomata are considered as important criteria to distinguish Thunbergia species. Not only that, Metcalfe and Chalk (1950) categorised the dicotyledons stomata into four main types based upon the basis arrangement of the subsidiary cells which are; anomocytic, anisocytic, paracytic and diacytic. Recently, Noraini et al. (2019) divided the stomatal types in plants into 10 categories which are; polocytic, amphidiacytic, diallelocytic, tetracytic, staurocytic and cyclocytic. The present leaf epidermal study on Thunbergia hence might confirm that the diacytic stomata found in both species studied are common in Thunbergia species as reported by Ahmad (1974).

Hickey (1973) claimed that the criteria of leaf venation is related to plant evolution and possessed as an important taxonomic value in plant systematic. The main functions of leaves including light-harvesting, gaseous exchange, water transport, and photosyntheate distribution depend on the architectural elements that are related to the arrangement of veins or called a venation pattern. Ummu-Hani et al. (2014) classified two types of ultimate marginal venation in the genus Ficus (Moraceae) which are; incomplete ultimate marginal venation (freely ending veinlets directly towards adjacent of the margin) and complete ultimate marginal venation (higher vein orders fused into a vein running just inside the margin). Furthermore, Sehgal and Paliwal (2008) also mentioned that anatomical studies on the ornamentation of leaf veins have significant values for the identification of Euphorbia species. Therefore, the complete marginal venation in T. erecta and incomplete venation in T. laurifolia can be used as a distinctive character to distinguish these two species. The present study also reported on the lamina venation with majority opened and minority closed with swollen tracheids in both Thunbergia studied, hence might be considered as a common character for the genus.

Leaf surface investigations have been more focussed compared to other plant surfaces because it comprised of variable features that occurred within taxa and useful in taxonomic applications. The results of the leaf surface studies are reported to be useful in the examination of genera and species of various families such as Podocarpaceae (Mill & Schilling 2009) and Rubiaceae (Moraes et al. 2009). Even Barthlott (1998) affirmed that the epicuticular wax structures possessed a considerable ultrastructural and chemical diversity, and significant in taxonomic study. The waxes have ecological importance especially for the interaction between the environment and plants. The appearance of verrucate, flakes and granules cuticular wax of T. erecta, and crustose and granules cuticular wax in T. laurifolia proved that the cuticular wax can be one of the important characters to distinguish these two species. The significance of the cuticular wax character in Thunbergia studied is supported by Nurul-Aini et al.
(2014) on the recognition of six types of epicuticular wax found in eight chosen taxa of Acanthaceae. Furthermore, Nurhanim et al. (2014) also mentioned that cuticular sculpturing is a criterion that can be used to distinguish plant species in Schoutenia Korth. (Malvaceae). There are three types of cuticular sculpturing that have been reported in five selected species of Schoutenia. The present study however reported that cuticular sculpturing can be slightly differentiated in T. erecta, whereas appeared as sinked anticlinal walls and raised periclinal walls in T. laurifolia. Somehow, the micromorphological aspects of the leaf surface are influenced by its habitat (Dunn et al. 1965). Moraes et al. (2011) also mentioned that cuticular ornamentation in some of Psychotria species does not appear to occur because of the sunlight exposure factor as the individuals grow in the understory of the forest.

Werker (2000) defined trichomes as unicellular or multicellular appendages that originated from the epidermal cells and develop outwards on the surface of various plant organs. Also, Navarro and El-Oualidi (2000) stated that trichomes are hairs which commonly can be found on the epidermis of plants. To be noticed, the trichomes morphology served an important taxonomic character and useful in the plant systematic. Beiras and Sajo (2009) also used the types of trichomes for the identification and classification of woody plants in Brazilian savannah. In fact, trichomes morphology is not only served in the identification of species but also important in pharmacognosy, archaeobotany, paleobotany and agronomy (Rao & Ramayana 1977). Apart, Kim et al. (2012) mentioned that trichomes also act as a physical and chemical barrier against biotic and abiotic stresses. For convenience purposes, the following structures of trichomes in Thunbergia studied are categorised into six recognisable types in Table 1. The types of trichomes are illustrated by drawings to avoid complex descriptions. Findings of the study recognised the five types of trichomes found in T. erecta which are; Types 1, 2, 3, 5 and 6, compared to T. laurifolia which consists of only two types of trichomes; Types 1 and 4. Moreover, the common type of trichomes found in Thunbergia studied is peltate glandular (unicellular terminal) trichome (Type 1). The criteria of trichomes morphology is proven to be useful to distinguish both species of Thunbergia studied, hence supported by previous research on the significance of trichomes morphology on petals of some Acanthaceae species (Amirul-Aiman et al. 2014).
| Type  | Trichomes descriptions                  | Pictures | Illustrations |
|-------|-----------------------------------------|----------|---------------|
| Type 1| Peltate glandular (unicellular terminal) | ![Picture](image1.png) | ![Illustration](image2.png) |
| Type 2| Peltate glandular (multicellular terminal) | ![Picture](image3.png) | ![Illustration](image4.png) |
| Type 3| Simple unicellular (short, blunt end)   | ![Picture](image5.png) | ![Illustration](image6.png) |
| Type 4| Simple unicellular (short, pointed end) | ![Picture](image7.png) | ![Illustration](image8.png) |
| Type 5| Simple multicellular (short, blunt end) | ![Picture](image9.png) | ![Illustration](image10.png) |
| Type 6| Simple multicellular (short, pointed end, echinate) | ![Picture](image11.png) | ![Illustration](image12.png) |
CONCLUSION

In conclusion, this study indicates that leaf anatomy and micromorphology characters are important in the taxonomic study, especially to distinguish some species in *Thunbergia*. Findings from this study showed five typical characters occurred in *T. erecta* and *T. laurifolia* including the presence of raphide in the parenchyma cortex of petiole and midrib, sinuous anticlinal walls on abaxial and adaxial surfaces, presence of diacytic stomata, lamina venation with majority opened and minority closed with swollen tracheids and presence of peltate glandular ( unicellular terminal) trichomes. These common characteristics can be used as taxonomic criteria in characterise species from the genus *Thunbergia*. Furthermore, the absence of cystoliths in *Thunbergia* studied confirms that subfamily Thunbergioideae does not possess cystoliths, hence can be a significant taxonomic criterion to recognise species from the genus *Thunbergia*. Results from this study also revealed several interesting features, with that might be useful to distinguish *T. erecta* and *T. laurifolia*. The variable characteristics examined are petioles and marginal outlines, types of vascular bundles, the presence of druses in the lamina part, types of marginal venation, cuticular wax, cuticular sculpturing, stomata occurrence and types of trichome. This study confirms that leaf anatomy and micromorphology characteristics possessed taxonomic importance and act as supportive evidence in the identification and classification of plants either at species or genus level.

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REFERENCES

Agbagwa I O and Ndukwa B C. (2004). The value of morpho-anatomical features in the systematic of *Cucurbita* L. (Cucurbitaceae) species in Nigeria. *African Journal of Biotechnology* 3(10): 541–546. https://doi.org/10.5897/AJB2004.000-2106

Ahmad K J. (1974). Cuticular studies in some of *Mendoncia* and *Thunbergia* (Acanthaceae). *Botanical Journal of the Linnee Society* 69: 53–63. https://doi.org/10.1111/j.1095-8339.1974.tb01614.x

Amirul-Aiman A J, Noraini T and Nurul-Aini C A C. (2014). Trichomes morphology on petals of some Acanthaceae species. *A Journal on Taxonomic Botany, Plant Sociology and Ecology* 14(1): 79–83.
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Aritajat S, Wutteerapol S and Saenphet K. (2004). Anti-diabetic effect of *Thunbergia laurifolia* Linn. aqueous extract. *Southeast Asian Journal Tropical Medicine and Public Health* 35: 53–58.

Barthlott W, Neinhuis C and Cutler D. (1998). Classification and terminology of plant epicuticular waxes. *Botanical Journal of the Linnean Society* 126(3): 237–260. https://doi.org/10.1111/j.1095-8339.1998.tb02529.x

Beck C B. (2010). *An introduction to plant structure and development plant anatomy for the twenty-first century*, 2nd ed. New York: Cambridge University Press. https://doi.org/10.1017/CBO9780511844683

Bieras A C and Sajo M G. (2009). Leaf structure of the Cerrado (Brazilian Savanna) woody plants. *Trees* 23: 451–471. https://doi.org/10.1007/s00468-008-0295-7

Borg J A, McDade A L and Schörenberger J. (2008). Molecular phylogenetics and morphological evolution of Thunbergioideae (Acanthaceae). *Taxon* 57(3): 811–822. https://doi.org/10.1002/tax.573012

Candolle C D E. (1879). *Anatomie compare des feuilles chez quelques familles de Dicotyledones*. Mémoires de la Société de Physique et d'Histoire Naturelle de Genève 26: 427–480.

Carlquist S. (1988). Wood anatomy of Acanthaceae: A survey. *Journal of Systematic and Evolutionary Botany* 12(1): 201–227. https://doi.org/10.1007/978-3-662-21714-6

Chan E W C, Eng S Y and Tan Y P. (2011). Phytochemistry and pharmacological properties of *Thunbergia laurifolia*: A review. *Pharmacognosy Journal* 3(24): 1–6. https://doi.org/10.5530/pj.2011.24.1

Chia-Chi H, Yunfei D and Wood J R I. (2011). *Flora of China: Acanthaceae*. United States of America: Missouri Botanical Garden Press.

Cutler D F. (1978). *Applied plant anatomy*. London: Longman Group Limited.

Dunn D B, Sharma G K and Campbell C C. (1965). Stomatal patterns of dicotyledons and monocotyledons. *American Midland Naturalist* 74: 185–195. https://doi.org/10.2307/2423132

Fahn A. (1967). *Plant anatomy*. Israel: Hakkibutz Hameuhad Publishing House Ltd.

Franceschi V R and Nakata P A. (2005). Calcium oxalate in plants: Formation and function. *Annual Review of Plant Biology* 56: 41–71. https://doi.org/10.1146/annurev.arplant.56.032604.144106

Hare C L. (1942). On the taxonomic value of the anatomical structure of the vegetative organs of the dicotyledons: The anatomy of the petiole and its taxonomic value. *Proceedings of the Linnean Society of London* 555(3): 223–229.

Heywood V H, Brummitt R K and Culham A. (2007). *Flowering plants families of the world*. Canada: Firefly Books.

Hickey L J. (1973). Classification of the architecture of dicotyledonous leaves. *American Journal of Botany* 60: 1–33. https://doi.org/10.1002/j.1537-2197.1973.tb10192.x

Johansen D A. (1940). *Plant microtechnique*. New York: McGraw-Hill.

Kar A, Goswami N K and Saharia D. (2013). Distribution and traditional uses of *Thunbergia Retzius* (Acanthaceae) in Assam, India. *Pleione* 7(2): 325–332.

Khatijah H and Ruzi M A R. (2006). *Anatomical atlas of Malaysian medicinal plants*. Bangi: UKM Press.

Kim H J, Seo E Y and Kim J H. (2012). Morphological classification of trichomes associated with possible biotic stress resistance in the genus *Capsicum*. *The Plant Pathology Journal* 28(1): 107–113. https://doi.org/10.5423/PPJ.NT.12.2011.0245

Korn R W. (1976). Concerning the sinuous shape of leaf epidermal cells. *New Phytologist* 77: 153–161. https://doi.org/10.1111/j.1469-8137.1976.tb01510.x
Metcalfe J D and Chalk L. (1950). *Anatomy of the dicotyledons*, Vol. 1 and 2. Oxford: Clarendon Press.

———. (1965). *Anatomy of the dicotyledons*. Oxford: Clarendon Press.

———. (1985). *Anatomy of dicotyledons: Wood structure and conclusion of the general introduction*. Oxford: Clarendon Press.

Mill R R and Schilling D M S. (2009). Cuticle micromorphology of *Saxegothaea* (Podocarpaceae). *Botanical Journal of the Linnean Society* 159: 58–67. https://doi.org/10.1111/j.1095-8339.2008.00901.x

Moraes T M, Barros C F and Silva-Neto S J. (2009). Leaf blade anatomy and ultrastructure of six *Simira* species (Rubiacae) from the Atlantic Rain Forest, Brazil. *Biocell* 33(3): 155–165. https://doi.org/10.32604/biocell.2009.33.155

Moraes T M, Rabelo G R and Alexandrino C R. (2011). Comparative leaf anatomy and micromorphology of *Psychotria* species (Rubiacae) from the Atlantic rainforest. *Acta Botanica Brasilia* 25(1): 178–190. https://doi.org/10.1590/S0105-308901100000021

Nath M and Dutta-Choudhury M. (2010). Ethno-medico botanical aspects of Hmar tribe of Cachar district, Assam (Part I). *Indian Journal Traditional Knowledge* 9(4): 760–764.

Navarro T and El-Oualidi J. (2000). Trichome morphology in *Teucrim* L. (Labiateae). A taxonomic review. *Anales Jardin Botanico De Madrid* 57(2): 277–297. https://doi.org/10.3989/ajbm.1999.v57.i2.203

Noraini T and Cutler D F. (2009). Leaf anatomical and micromorphological characters of some Malaysian *Parashorea* (Dipterocarpaceae). *Journal of Tropical Forest Science* 21(2): 156–167.

Noraini T, Ruzi A R and Amirul-Aiman A J. (2019). *Anatomi dan mikroskopik tumbuhan*. Bangi: Universiti Kebangsaan Malaysia.

Nurhanim M N, Noraini T and Chung R C K. (2014). Nilai taksonomi ciri anatomi daun genus *Schoutenia* Korth. (Malvaceae subfam. Brownlowioideae). *Sains Malaysiana* 43(3): 331–338.

Nurul-Aini C A C, Noraini T, Amirul-Aiman AJ and Ruzi A R. (2013). Taxonomic value of leaf micromorphology in some selected Acanthus L. (Acanthaceae) species in Peninsular Malaysia. Proceedings of 22nd Scientific Conference of Microscopy Society Malaysia (MSM2013), Primula Beach Hotel, Terengganu, 26–28 November, 40–44.

Nurul-Aini C A C, Noraini T and Latiff A. (2014). Taxonomic significance of leaf micromorphology in some selected taxa of Acanthusae (Peninsular Malaysia). *The 2014 UKM FST POSTGRADUATE COLLOQUIUM: Proceedings of the Universiti Kebangsaan Malaysia, Faculty of Science and Technology 2014 Postgraduate Colloquium*, 727–733. https://doi.org/10.1063/1.4895291

Nurul-Aini C A C, Nur-Shuhada T and Rozilawati S. (2018). Comparative leaf anatomy of selected medicinal plant in Acanthaceae. *IIUM Medical Journal Malaysia* 17(2): 17–24.

Nurul-Syahirah M, Noraini T and Latiff A. (2016). Characterization of midrib vascular bundles of selected medicinal species in Rubiaceae. Proceedings of the Universiti Kebangsaan Malaysia, Faculty of Science and Technology 2016 Postgraduate Colloquium, Selangor, Malaysia, 13–14 April 2016.

Rao S R S and Ramayana N. (1977). Structure distribution and taxonomic importance of trichomes in the Indian species of *Malvastrum*. *Phytomorphology* 27: 40–44.

Retief E and Reyneke W F. (1984). The genus *Thunbergia* in Southern Africa. *Bothalia* 15: 107–116. https://doi.org/10.4102/abc.v15i1/2.1109
Robbrecht E. (1988). *Tropical woody Rubiaceae. Characteristic features and progressions. Contributions to a new subfamilial classification*. Opera Botanical Belgica I. Meise: National Botanical Garden of Belgium.

Rojo J P. (1987). Petiole anatomy and infrageneric in interspecific relationship Philippines Shorea (Dipterocarpaceae). In A J G H Kostermans (ed.). *Proceedings of the Third Round Table Conference on Dipterocarps*. Mulawarman University, Indonesia.

Ruzi A R, Hussin K and Noraini T. (2009). Systematic significance of the petiole vascular bundles types in Dipterocarpus Gaertn. F. (Dipterocarpaceae). *Malaysian Applied Biology* 38(2): 11–16.

Saas J E. (1958). *Botanical microtechnique*, 3rd ed. Ames: Iowa State University. https://doi.org/10.31274/isudp.25

Sarma J. (2006). *Medicinal and aromatic plants of Assam with special reference to Karbi Anglong*. India: Silviculture Division, Department of Forest, Diphu.

Scotland R W, Sweere J A and Reeve P A. (1995). Higher level systematics of Acanthaceae determined by chloroplast DNA sequences. *American Journal of Botany* 82(2): 266–275. https://doi.org/10.1002/j.1537-2197.1995.tb11494.x

Sehgal L and Paliwal G S. (2008). Studies on the leaf anatomy of Euphorbia: II Venation Patterns. *Botanical Journal of Linnean Society* 68(3): 173–208. https://doi.org/10.1111/j.1095-8339.1974.tb01758.x

Stevens P F. (2016). *Angiosperm Phylogeny Website: Lamiales (Acanthaceae)*. http://www.mobot.org/MOBOT/research/APweb/ (accessed on 3 October 2020).

Sultana K W, Chatterjee S and Roy A. (2015). An overview on ethnopharmacological and phytochemical properties of Thunbergia sp. *Medicinal and Aromatic Plants* 4(5): 1–6.

Suwanphakdee C and Vajrodaya S. (2018). *Thunbergia impatienoides* (Acanthaceae), a new species from Thailand. *Blumea* 63: 20–25. https://doi.org/10.3767/blumea.2018.63.01.03

Takhtajan A. (1997). *Diversity and classification of flowering plants*. New York: Columbia University Press.

Teron R. (2005). Bottle gourd: Part and parcel of Karbi culture. *Indian Journal Traditional Knowledge* 4(1): 86–90.

Ummu-Hani B, Noraini T and Abdul-Latiff M. (2014). Studies on leaf venation in selected taxa of the genus Ficus L. (Moraceae) in Peninsular Malaysia. *Tropical Life Sciences Research* 25(2): 111–125.

Vollesen K. (2008). *Flora of Tropical East Africa: Acanthaceae*. Kew: Royal Botanic Gardens. Washington: Department of Botany, National Museum of Natural History.

Werker E. (2000). Trichome diversity and development. *Advances in Botanical Research* 31: 1–35. https://doi.org/10.1016/S0065-2296(00)31005-9