The Potential of Light Microscopic Features of the Oral Mucosa in Predicting Post-mortem Interval

Jaganath Patro,1 ‘Swagatika Panda,1 Neeta Mohanty,1 Uma S. Mishra2

ABSTRACT: Objectives: The post-mortem interval (PMI) refers to the amount of time elapsed between death and discovery of the body. This study aimed to evaluate light microscopic cellular changes in the oral mucosa and identify the potential of this method for predicting PMI. Methods: This prospective study was conducted between July 2016 and January 2018 at the Institute of Dental Sciences, Siksha ‘O’ Anusandhan University, Bhubaneswar, India. A total of 150 post-mortem (including 75 gingival and 75 buccal mucosa samples) and 40 ante-mortem (including 20 gingival and 20 buccal mucosa samples) tissue samples were compared using haematoxylin and eosin, periodic acid-Schiff (PAS) and van Gieson stains. Microscopic changes in the epithelium and connective tissue were categorised according to PMI stage as early (<12.5 hours since death), intermediate (12.5–20.5 hours since death) or late (>20.5 hours since death). Results: Most epithelial cellular changes occurred early, except for arc-shaped nuclei and epithelial shedding which were intermediate and late changes, respectively. However, microscopic changes in the connective tissue were only observable at ≥12.5 hours. There was a progressive decrease in intensity in van Gieson stains and an increase in intensity in PAS stains as PMI increased. Several microscopic features were found to be significant predictors of PMI including epithelial homogenisation, cytoplasmic vacuolisation, nuclear degeneration, arc-shaped nuclei, chromatin clumping, red blood cell clumping and lysis, melanin incontinence, myofibril degeneration, salivary gland acinar degeneration and epithelial connective tissue separation (P < 0.05 each). Conclusion: These findings indicate that microscopic evaluation of the oral mucosa may be helpful for PMI prediction.

Keywords: Post-mortem Changes; Light Microscopy; Oral Mucosa; Epithelial Cells; Lamina Propria; Salivary Glands; Histocytchemistry; Periodic Acid–Schiff Reaction; India.

The potential of light microscopy in the estimation of post-mortem interval, but also highlight the importance of special stains other than haematoxylin and eosin as a complementary tool in forensic investigation.

To the best of the authors’ knowledge, this study is the first of its kind to depict the sequential, temporal, cellular and architectural histological features of the oral mucosa after death in a large sample using two distinct kinds of mucosal samples and three staining techniques.
Thanatology refers to the philosophical and scientific study of various aspects of death including the forensic examination of bodily changes which take place at death and during the post-mortem period. While dozens of methods are currently used to determine the amount of time elapsed between death and discovery of the body—a period known as the post-mortem interval (PMI)—there is nevertheless still demand for more reliable and precise techniques. Currently, many approaches to PMI estimation focus on observable and measurable changes to the body after death such as autolysis, algor mortis, rigor mortis, livor mortis and the digestive status of the stomach contents.

Following death, many physical and chemical changes begin to take place and continue until the body eventually decomposes. At the onset of death, respiration ceases and cell glycolysis begins, leading to the production of lactic acid followed by a drop in the pH of the cellular contents. Eventually, cellular metabolism comes to an end and the subsequent predominance of lytic enzymes results in autolysis. The cell membranes disintegrate and a reduction in adenosine triphosphate levels causes loss of the cellular architecture and fatty acids. Cell swelling occurs due to a decrease in energy level; subsequently, the cells progressively detach from one another in a process known as necrosis. The reduction in cellular energy causes disruption of the mitochondrial electron transport system and leads to fermentation, resulting in the formation of anaerobic lactic acid from pyruvic acid.

Utilising light microscopy, cells are perceived as dead after they have undergone a series of specific morphological changes which occur as a result of two distinct cellular processes: (1) apoptosis, a programmed and orderly form of cell death and (2) necrosis, a disorderly and unpredictable form of cell death. Necrotic cells show an increased eosinophilia. Nucleus of the epithelial cells may undergo changes like karyolysis, pyknosis and karyorrhexis.

Post-mortem cellular changes have been proven to reliably estimate PMI in several organs including the liver, kidney, pancreas and heart. As in other tissues, autolytic enzymes are released by the oral mucosal cells after death, resulting in abnormal morphology. Therefore, researchers have explored the possibility of identifying time since death via microscopic examination of the oral mucosa. Degenerative post-mortem changes to the epithelium and connective tissue in the oral mucosa may demonstrate specific patterns that could prove useful to determine PMI; however, few studies have sought to evaluate post-mortem cellular changes in the oral tissues including the gingiva, salivary glands and dental pulp. Moreover, inadequacies in terms of sample size and standardised methodology have often precluded confirmation of this approach. Therefore, the current study aimed to investigate post-mortem light microscopic cellular changes in the oral tissues with the objective of estimating their potential for PMI prediction.

**Methods**

This prospective study took place over an 18-month period between July 2016 and January 2018 at the Institute of Dental Sciences, Siksha ‘O’ Anusandhan University, Bhubaneswar, Odisha, India, in collaboration with two government medical colleges in Odisha. Oral mucosa samples were collected from the bodies of recently deceased individuals aged 18–45 years old. In each case, information was obtained concerning cause of death, age, gender, time and date of death and time spent outside or in a natural environment since death. The bodies of individuals with a history of chronic periodontitis, neoplastic growth or communicable disease were excluded, as were those of pregnant females, victims with facial burns and those who had been preserved after death.

The bodies were placed in a dry environment with adequate light and kept at room temperature. Oral tissues were collected under the direct supervision of a forensic medicine and toxicology specialist. The gingival and buccal mucosa were accessed using basic diagnostic instruments including a mouth mirror, a no. 5 interproximal explorer, a pair of tweezers and a pair of toothed forceps. Samples were collected using a round punch knife of 6 mm in diameter, a no. 15 Bard-Parker stainless steel blade (Aspen Surgical Corp., Caledonia, Michigan, USA) and a Langenbeck retractor to retract the buccal mucosa. A total of 150 mucosa samples of approximately 6 × 8 × 5 mm in size were collected, comprising both gingival (n = 75) and buccal (n = 75) samples. In addition, 40 antemortem samples of keratinised (n = 20) and non-keratinised (n = 20) oral mucosa were collected from living patients of the same age group admitted for surgical procedures.
such as gingivectomy, surgical impaction and biopsies of reactive lesions.

The excised tissues were fixed in 10% buffered formalin. Subsequently, formalin-fixed paraffin-embedded (FFPE) blocks were prepared for both post-mortem and antemortem samples. The FFPE blocks were sectioned into slides of 4 µm thickness and stained with routine haematoxylin and eosin (H&E) stain. In addition, 21 slides were stained with van Gieson stain (Hi-Media Laboratories Pvt. Ltd., Einhausen, Germany) and 15 slides were stained with periodic acid-Schiff (PAS) stain (Hi-Media Laboratories Pvt. Ltd.) to determine changes in the intensity of collagen/reticulin fibres and basement membranes/mucous acini, respectively, with time since death.

Stained tissue sections were viewed under a light microscope to evaluate epithelial and connective tissue changes. These changes were categorised according to PMI stage as either early (<12.5 hours since death), intermediate (12.5–20.5 hours since death) or late (>20.5 hours since death).

### Table 1: Light microscopic cellular changes according to time since death in oral mucosa samples (N = 150)

| Type of tissue | PMI stage* |
|---------------|------------|
|               | Early      | Intermediate | Late       |
| Epithelium    | • Cytoplasmic vacuolation (BM and GM) | • Arc-shaped nuclei (BM) | • Epithelial shredding (BM) |
|               | • Homogenisation (BM and GM) | | |
|               | • Nuclear degeneration (BM and GM) | | |
|               | • Chromatin clumping (BM and GM) | | |
|               | • Arc-shaped nuclei (GM) | | |
| Connective tissue | • Collagen lysis (BM) | • Melanin incontinency (BM and GM) | • RBC clumping (BM and GM) |
|               | • Loss of epithelial connective tissue interface (BM) | | • Collagen lysis (GM) |
|               | • Myofibril degeneration (BM) | | |
|               | • Mucous acini degeneration (BM) | | |
| Lamina propria | • Clumping and lysis of RBCs | • Clumping and lysis of RBCs | • Clumping and lysis of RBCs |
|               | • Collagen lysis | • Myofibril degeneration | • Melanin incontinency |
| Salivary gland | • Epithelial connective tissue separation | | |
|               | • Mucous acini degeneration | | |

PMI = post-mortem interval; BM = buccal mucosa; GM = gingival mucosa; RBC = red blood cell. *PMI stage was categorised as early (<12.5 hours since death), intermediate (12.5–20.5 hours since death) or late (>20.5 hours since death).

### Table 2: Predictive potential of selected light microscopic cellular features to estimate post-mortem interval in oral mucosa samples* from recently deceased individuals (N = 75)

| Type of tissue | Feature | Frequency according to PMI stage†, n (%) | P value |
|---------------|---------|----------------------------------------|---------|
| Epithelium    | GM      | Homogenisation                         | 0.011   |
|               | BM      | Nuclear degeneration                    | 0.021   |
|               | GM      | Cytoplasmic vacuolation                 | 0.012   |
|               | BM      | Chromatin clumping                      | 0.029   |
|               | BM      | Arc-shaped nuclei                       | 0.038   |
| Lamina propria| BM      | Clumping and lysis of RBCs              | 0.049   |
|               | GM      | Clumping and lysis of RBCs              | 0.010   |
|               | GM      | Collagen lysis                          | <0.001  |
|               | BM      | Myofibril degeneration                  | 0.003   |
|               | BM      | Melanin incontinency                    | 0.017   |
| Salivary gland| BM      | Epithelial connective tissue separation | 0.028   |
|               | BM      | Mucous acini degeneration               | 0.013   |

PMI = post-mortem interval; BM = buccal mucosa; GM = gingival mucosa; RBC = red blood cells. *Including one sample each of BM and GM. †PMI stage was categorised as early (<12.5 hours since death), intermediate (12.5–20.5 hours since death) or late (>20.5 hours since death).
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*Jagganath Patro, Swagatika Panda, Neeta Mohanty and Uma S. Mishra*

**Figure 1:** Haematoxylin and eosin stains at 10x magnification of post-mortem samples of oral mucosa epithelial cells showing (A) arc-shaped nuclei (arrow), (B) epithelial homogenisation (arrow), (C) nuclear degeneration (arrow) and (D) cytoplasmic vacuolation (arrow). These cellular and architectural microscopic changes were found to be significant predictors of post-mortem interval.

**Figure 2:** Haematoxylin and eosin stain at 10x magnification of post-mortem samples of oral mucosa connective tissue cells showing (A) epithelial connective tissue separation (arrow), (B) clumping (arrow) and lysis (arrowhead) of the red blood cells, (C) myofibril degeneration (arrow), (D) collagen lysis (arrow), (E) melanin incontinency (arrow) and (F) degeneration of the mucous acini (arrow). These cellular and architectural microscopic changes were found to be significant predictors of post-mortem interval.
Results

A total of 150 oral mucosa samples comprising 75 gingival and 75 buccal mucosa samples were obtained from 75 deceased individuals. Of these, 51 (68%) were male and 24 (32%) were female. The most common cause of death was road traffic accidents (n = 37; 49%), followed by poisoning (n = 19; 25%), hypoxia (n = 15; 20%) and cardiac arrhythmia (n = 1; 1%). The reason for death in the remaining three patients was unknown. In terms of PMI stage, 25 (33%) individuals had died <12.5 hours previously, 28 (37%) had died between 12.5–20.5 hours previously and 22 (29%) had died >20.5 hours previously.

In the epithelial cells, early microscopic changes seen within 12.5 hours of death included epithelial homogenisation, cytoplasmic vacuolation, nuclear degeneration and chromatin clumping with a progressive increase in the frequency of these features in the intermediate and late stages. Arc-shaped nuclei and epithelial shredding were intermediate and late changes, respectively. Interestingly, microscopic features of the connective tissue such as melanin incontinency and mucous acini degeneration were observed in the intermediate stage, whereas RBC clumping and lysis, and collagen lysis were late changes. Epithelial connective tissue separation and myofibril degeneration were observed in <40% of the cell population at all three PMI stages. In addition, inflammatory cells in the connective tissue stroma were observed in very few post-mortem tissues. Time-specific microscopic changes according to PMI stage are summarised in Table 1. The appearance of these features in the epithelium and connective tissue are shown in Figures 1 and 2, respectively. None of these architectural or cytological abnormalities were identified in the antemortem specimens.

In the buccal mucosa samples, a logistic regression analysis indicated that epithelial features such as nuclear degeneration, arc-shaped nuclei and chromatin clumping along with connective tissue features such as RBC clumping and lysis, melanin incontinency, degeneration of the myofibrils and salivary gland acini and epithelial connective tissue separation were significant predictors of PMI (P <0.050 each). In turn, epithelial homogenisation, changes to the nuclear outline, nuclear vacuolation in the epithelial cells and collagen lysis and RBC clumping and lysis in connective tissue cells were significant predictors of PMI in the gingival samples (P <0.050 each) [Table 2].

There was a progressive decrease in staining intensity of collagen fibres in van Gieson stains [Figure 3]. The intensity of van Gieson stains in collagen fibres was found to be a significant predictor of time since death in both buccal mucosa and gingival samples (P = 0.029). While there was no progressive change in PAS staining intensity in the basement membranes of either buccal mucosa or gingival samples as PMI increased, PAS staining intensity in salivary gland acini increased significantly with PMI (P = 0.021) [Figure 4].

Discussion

The current study was conducted on post-mortem samples of otherwise healthy buccal and gingival mucosae collected at three different intervals of time since death. The first objective was to identify
differences between antemortem and post-mortem samples of oral mucosa among adults of similar age groups, while the secondary objective was to evaluate early, intermediate and late microscopic changes in oral tissues according to PMI stage. Finally, the study sought to determine the potential of these microscopic features in predicting time since death using both routine and special staining techniques.

With regards to the first objective, various degenerative microscopic features in the epithelium and connective tissue of the oral mucosa were observed in post-mortem samples, but not in antemortem samples. In particular, cytoplasmic vacuolation, epithelial homogenisation, nuclear degeneration and chromatin clumping were early changes in both the buccal and gingival mucosa. However, while arc-shaped nuclei were seen at <12.5 hours since death in the gingival mucosa, this feature was only seen at ≥12.5 hours in the buccal mucosa. Similarly, melanin incontinency was the only microscopic change observed in both the buccal and gingival mucosa at ≥12.5 hours, whereas collagen lysis, degeneration of the myofibrils and salivary gland acini and epithelial connective tissue separation were seen at this stage only in the buccal mucosa. These findings support those of previous research conducted by Pradeep et al. and Yadav et al.7

In addition, the present study also focused on additional features such as arc-shaped nuclei as an early change in the gingiva and as an intermediate change in the buccal mucosa. The identification of epithelial shredding as a late microscopic change in the buccal mucosa is in agreement with results reported by Yadav et al. in the gingival and labial mucosa.8,15 Microscopic changes in the connective tissue such as RBC clumping and lysis in both types of mucosae and collagen lysis in the gingiva occurred at >20.5 hours. The latter finding supports previous research by Yadav et al. indicating that collagen disruption becomes increasingly apparent over time.6,15 However, Yadav et al. reported no abnormalities in the RBCs, a finding which contradicts that of the present study.7

The logistic regression analysis indicated that certain microscopic features in the epithelium—including homogenisation and cytoplasmic vacuolation in the gingiva and nuclear degeneration, arc-shaped nuclei and chromatin clumping in the buccal mucosa—were significant predictors of PMI. Similarly, connective tissue features of RBC clumping and lysis, melanin incontinency, degeneration of the myofibrils and salivary gland acini and epithelial connective tissue separation in the buccal mucosa and RBC clumping and lysis and collagen lysis in the gingiva were significant PMI predictors. While the role of melanin incontinency in estimating PMI has been evidenced in the skin, no previous study had yet explored its potential in the oral mucosa.16

Several connective tissue stains are currently used in forensic pathology practice to order to identify the nature of death.17,18 However, to the best of the authors’ knowledge, no other studies have sought to determine their role in predicting PMI. The current study found that, with an increase in time elapsed since death, there was a significant progressive decrease in the staining intensity of collagen fibres using van Gieson stains, whereas there was a significant progressive increase in the staining intensity of salivary gland acini using PAS stains. Therefore, either of these two staining techniques may be useful in the estimation of PMI. The van Gieson stain is a simple stain which tints collagen red and other tissues yellow and is often used for the differential demonstration of collagen. Varying shades of red are produced depending on the density of the collagen fibres. A progressive decrease in stain intensity can be attributed to collagen lysis, a feature observed in both the intermediate and late PMI stages; alternatively, this may be due to a decrease in the thickness of the submucosa or lamina propria.15,19

In a previous study, Mahalakshmi et al. evaluated histological changes in gingival tissue based on an evaluation of unfixed antemortem tissues alone.6 Yadav et al. also studied post-mortem changes in oral mucosa without comparison with antemortem mucosa.15 In comparison, other researchers have compared antemortem and post-mortem mucosal samples to determine post-mortem cellular changes.7,8,10,16 Three studies have been conducted to associate changes in salivary gland oral mucosa with time since death.11,12,20 Based on evaluations of the cellular changes in antemortem and post-mortem mucosa, Gururaj suggested that the decomposition process begins in the initial 24 hours, while Pradeep et al. concluded that the decomposition process at the cellular level begins within 10 hours of death.7,21 Yadav et al. investigated 10 antemortem mucosa samples to determine nuclear and cytoplasmic changes at various time intervals and proposed the usefulness of histological changes in the oral mucosa for estimating PMI.6 In a recent study, Muthukrishnan et al. described early and late post-mortem changes in five antemortem and five post-mortem gingival samples.10

Due to sample size limitations, none of the aforementioned studies were able to provide statistical confirmation of the usefulness of light microscopic cellular changes in the oral mucosa as a method of predicting time since death.7–11,15,20,21 Moreover, several studies either evaluated post-mortem cellular changes in the gingiva alone or in the labial mucosa.7,8,15 No studies have yet sought to assess the role of these...
changes in the buccal mucosa. Indeed, the present study is the first to establish the usefulness of this approach in both the gingiva and buccal mucosa, which are distinct types of oral mucosal tissues which are different from both an anatomical and structural perspective. For instance, the mean thickness of the epithelium in the buccal mucosa and gingiva are 294 ± 68 µm and 239 ± 57 µm, respectively.2 In addition, the thickness of the connective tissue also differs in these two zones as submucosa is absent in the gingiva, but present in the buccal mucosa. As such, characterisation of the role of the oral mucosa in identifying PMI requires the individual evaluation of different mucosal regions.

This study is subject to certain limitations, such as the absence of quantifying the cellular and nuclear changes to establish any association with PMI and observing the post-mortem cellular and nuclear changes in different environments. These factors would have led to further accuracy in determining PMI. However, the strengths of this study lie in the inclusion of a large sample size, the evaluation of two different kinds of oral mucosa tissue—including the buccal mucosa—and examination of the role of microscopic features in routine H&E stains as well as special stains such as van Gieson and PAS. Based on these findings, further research is recommended to determine the point in time after death at which microscopic changes cease. In addition, advanced histological techniques, such as immunohistochemistry and electron microscopy, should also be investigated as other potential methods of determining post-mortem changes in the oral tissues.

**Conclusion**

Several microscopic features of the gingiva and buccal mucosa were found to be significant predictors of PMI including epithelial homogenisation, cytoplasmic vacuolation, nuclear degeneration, arc-shaped nuclei, chromatin clumping, RBC clumping and lysis, melanin incontinency, myofibril degeneration, salivary gland acini degeneration and epithelial connective tissue separation. In addition, there was a progressive decrease and increase in staining intensity with the use of van Gieson and PAS stains, respectively. To the best of the authors’ knowledge, this study is the first of its kind to identify light microscopic features in two distinct kinds of oral mucosae using two special staining techniques in addition to conventional H&E staining.

**CONFLICT OF INTEREST**
The authors declare no conflicts of interest.

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