Early defoliation effects on water status, fruit yield and must quality of ‘Nerello mascalese’ grapes

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ABSTRACT: The effect of basal zone defoliation on vines influences yield and ripening depending on the phenological stage and its intensity. Early basal defoliation (EBD) was carried out at flowering on the autochthonous Vitis vinifera (L.) cultivar ‘Nerello mascalese’ grown on the eastern slopes of Mount Etna, Sicily. The effects were evaluated over a two-year period. In the first year, the canopy retained 67 % of its original leaf area after EBD and in the second year, 58 %. Compared with control vines, mid-day leaf water potentials in the EBD vines enjoyed higher water status throughout the growing seasons in both years. EBD had no significant negative effects on yield but did have significant positive effects on the levels of total polyphenols, total flavonols and total anthocyanins. Yield and its components are conditioned by a number of factors mainly linked to climate and to the vines’ general condition during berry growth and ripening. Even after two consecutive years of treatment, EBD does not have any negative effects on yield. However, EBD does have strong positive effects on berry quality attributes as a consequence of the following: greater exposure of the cluster to sunlight, higher vine water status and avoidance of high temperature stresses in the last phase of ripening during the analysis of anthocyanin.

Keywords: Vitis vinifera (L.), photosynthesis, canopy, sustainability, berry quality

Introduction

Nowadays, partial defoliation in the basal zone of the grapevine shoot is a common management practice used to control the microclimate in the cluster area. This practice regulates the shade/exposure balance and the ratio of the older less photosynthetically active leaves to the younger more photosynthetically active leaves [Intrieri et al., 2008]. The effects of basal zone defoliation depend on the phenological stage at which it is implemented and on its intensity [Candolfi-Vasconcelos and Koblet, 1990; Poni et al., 2006]. Early basal defoliation (EBD), implemented from the pre-flowering to the early fruit set only, can influence both yield [Poni et al., 2006] and ripening (Ferlito et al., 2014). Yield reduction is likely due to a lower fruit set and smaller berries [Nicolosi et al., 2012; Poni et al., 2008] as a consequence of upsetting the source-sink balance. Before veraison, basal defoliation can lower shoot photosynthesis [Kotseridis et al., 2012]. Hunter and Visser [1988] suggested that excess vegetative growth is detrimental to interior-canopy microclimate as well as the photosynthetic rate of the entire vine. However, this effect may not be strong because grapes have great resilience, showing compensatory increases in lateral growth and in the photosynthetic efficiency of younger leaves [Reynolds and Wardle, 1989]. Meanwhile, fruit quality parameters are positively affected by EBD and are key to the oenological wine profile as measured by the fruit’s biochemical characteristics [Feng et al., 2017; Pisciotto et al., 2013]. Increases in bunch exposure are generally associated with increases in the ratio of total soluble solids/titratable acidity (Pastore et al., 2017). The other fruit quality attributes are often more strongly affected by cultivation practices, the environment and the genotype [Ferlito et al., 2014; Downey et al., 2006]. Generally, anthocyanin accumulations are positively related to the increased light exposure associated with defoliation [Main and Morris, 2004; Chorti et al., 2010; Pastore et al., 2017] and negatively related to the associated higher berry temperatures. Effects of increased light exposure on berry flavonoids are unclear. Lemut et al. [2013] found that increased light has a positive effect on flavonoid synthesis while others have reported decreased levels of these compounds [Spayd et al., 2002; Tarara et al., 2008; Pisciotto et al., 2013; Scafidi et al., 2018].

The aim of this research was to investigate the effects of EBD as a tool for yield control and fruit quality improvement in the black grape cultivar ‘Nerello mascalese’ grown on the eastern slopes of Mount Etna.

Materials and Methods

Site description, plant material and experimental design

The trial was conducted over two consecutive seasons [2015 and 2016] in a commercial Vitis vinifera [L.] vineyard [lat. 37°41’13” N; long. 15°07’00” E; elevation 602 m a.s.l.] (Figure 1). Vines of the black cultivar ‘Nerello mascalese’ grafted onto 140 Ru. rootstock were planted in 2010 with north-south rows in Zafferana Etnea, Catania, on the eastern slopes of Mount Etna, Italy. Vines were planted at a spacing of 0.9 m [in row] x 2.50 m [between rows] and trained to a unilateral cordon system at a height of 0.5 m. All vines were spur pruned to three or four spurs per vine and two buds
per spur. All vines were later standardised by removing by hand all shoots derived from the bourillon (first small bud), the crown and any adventitious buds so as to retain six-eight shoots per vine. The shoots were positioned vertically, aligned in the row direction and were not hedged during the growing season. The vineyard was not irrigated and soil tillage was mechanical. The trial was a completely randomised design with three independent plots of five rows, each containing 20 vines. All measurements were taken from three ‘index’ vines per block.

Climate and soil

Daily temperature and rainfall data and long-term climatic data of the site were provided by the Sicilian Water Observatory (www.osservatorioacque.it). The climate of the trial site is characterised by wet winters and semi-arid summers (Figures 2A, B and C). In the previous 30 years, according to the June-Aug rainfall, 14 seasons were characterized as dry (0-50 mm of rain), 9 seasons as normal (50-100 mm of rain) and 7 as wet (> 100 mm of rain). The solar radiation values were provided by the Sicilian Agrometeorological System (www.sias.regione.sicilia.it).

Soil was characterized at three different depths: 10-30 cm, 30-50 cm and 50-70 cm. According to the USDA scheme (Klingebiel and Montgomery, 1961) the vineyard soil was classified as loamy-sand at each depth. Briefly, the soil pH is neutral, the active carbonate is absent and the organic matter content is > 2% (data not shown).

Treatment and leaf and shoot measurements

[i] Early basal defoliation (EBD): All leaves were removed by hand from the shoot base (cordon) to the leaf just above the most distal cluster; [ii] Control [C]: No leaves were removed. The EBD treatment was imposed two weeks after emergence of the inflorescence (principal growth stage n. 6: flowering; BBCH 61) (Lorenz et al., 1994). At this stage, we recorded the total numbers and areas of the leaves removed from the main and lateral shoots (Table 1). Leaf areas were measured using a Leaf Area Meter. We calculated the total remaining leaf area (TLA) per vine, the removed leaf area per vine, per main shoot and per lateral shoot and their percentages as described by Nicolosi et al. [2012]. The lengths of the main and lateral shoots were also measured.
Physiological measurements

Mid-day leaf water potential ($\Psi_L$), leaf gas exchange and leaf SPAD index (amount of chlorophyll) were measured every 15 days from day of year (DOY) 181 (early summer) to DOY 256. The $\Psi_L$ measurements were taken on nine, fully-expanded leaves using a Scholander pressure chamber according to Matthews et al. (1987). At least 2 h before detachment from the vine, leaves were enclosed in small, black, hermetic plastic bags covered with aluminum foil. Nine replicates per treatment were measured selecting three leaves from each plot. Net photosynthesis ($Pn$), stomatal conductance (gs) and transpiration ($E$) rates of well-exposed, mature primary leaves were measured with a portable photosynthesis system. Nine leaves per plot inserted on the apical portion of each shoot were tagged and sampled. At each measurement, these leaves were monitored during the morning (10h00-12h00) on sunny days (~1500 μmol m$^{-2}$ s$^{-1}$) and had similar VPDmax (3.25 ± 0.25 kPa). The instrument was run in constant flow mode (500 μmol s$^{-1}$) and sample CO$_2$ concentration was held constant at 370 ppm using a mini CO$_2$ cartridge. A portable chlorophyll meter was used to measure the SPAD index. Data was collected on each measurement date from nine fully exposed main and lateral leaves.

Crop yield and biochemical analysis

The crops were harvested on 5 Oct in 2015 and 10 Oct in 2016. For yield assessment, the clusters on the index vines were counted and weighed. Total yields per vine and per shoot were determined. In the laboratory samples of nine clusters per plot were used to determine cluster length and weight, numbers and weights of berries, rachis and skin weights.

A 100-berry sample of the index plants per experimental unit was divided into three sub-samples. The first sub-sample was used to determine the concentration of total soluble solids (TSS) [as °Brix using a digital refractometer with temperature correction], the pH and the titratable acidity (TA) expressed as g L$^{-1}$ of tartaric acid equivalents, using an automatic titrator with 5.0 mL juice samples titrated against 0.1 M NaOH to pH 8.2.

The second sub-sample was used to determine total polyphenols, expressed as mg of gallic acid equivalent (GAE) 100 g$^{-1}$ of berries and measured using the Folin-Ciocalteu reagent assay (Singleton et al., 1999). The third sub-sample was used to measure total anthocyanins and flavonoids expressed as mg kg$^{-1}$ fresh weight as described by Ferlito et al. (2014).

Statistical analyses

Analysis of variance (ANOVA) and mean separations by Tukey’s post hoc test were carried out using STATISTICA 6.0 statistical software. Significant treatment and genotype effects were shown by a factorial analysis of variance. Statistically significant differences were represented as $p$ values that define the probability that an observation indicates a true relationship between factors. Results are marked *, ** or ***, referring to $p \leq 0.05$, $p \leq 0.01$ or $p \leq 0.001$, respectively. In order to determine the relationships between the indicators, we carried out a Spearman’s rank-order correlation analysis applying the non-parametric version of the Pearson correlation using the same statistical software. A principal component analysis (PCA) was carried out on a dataset including yield components, morphological data and cluster biochemical characteristics of EBD and C vines to represent a multidimensional dataset, with the lowest number of variables explaining the main features of the dataset. PCA was carried out using PAST (Paleontological Statistics software package, 2001).

Results

Climate variables

The annual average temperature was 17 °C [Figures 2A, B and C]. During the two years of the study the lowest minimum temperatures were recorded in Jan and Feb. July was the hottest month [mean daily maximum temperature 25.9 °C in 2015 and 25.4 °C in 2016]. Mean daily maximum temperature values were always above 23 °C from June to Sept and above 14.5 °C from Mar to May. Annual rainfall was about 2,000 mm in 2015 and 1,500 mm in 2016. No rainfall was recorded in July in either year, Aug rainfall was 66 mm in 2015 and 49 mm in 2016 and Sept rainfall was 240 mm in 2015 and 230 mm in 2016. The solar radiation levels are somewhat high in both years. Values ranged between 25 and 30 MJ m$^{-2}$ from Apr to the end of Aug (Figure 3).

Early Basal Defoliation

After defoliation, the fraction of canopy leaf area remaining was 67 % in 2015 and 58 % in 2016. The lower fraction of remaining leaf in 2016, derived from the low value of TLA per vine. The low TLA per vine was due to smaller and fewer leaves, and not shorter shoots (shoot lengths were similar in the two years). The leaf area/vine removed was 10,400 cm$^2$ in 2015 and 8,500 cm$^2$ in 2016 (a difference of approximately 1,900 cm$^2$) while the reduction in total leaf area/vine was 11,500 cm$^2$. The second subsample was used to determine total polyphenols, expressed as mg of gallic acid equivalent (GAE) 100 g$^{-1}$ of berries and measured using the Folin-Ciocalteu reagent assay (Singleton et al., 1999). The third sub-sample was used to measure total anthocyanins and flavonoids expressed as mg kg$^{-1}$ fresh weight as described by Ferlito et al. (2014).
cm² in 2016 year [64 % less]. These percentages are due to the removal of 14 % [2015] and 15 % [2016] of leaves per shoot. The EBD had an effect on lateral leaf growth causing both TLA and removed leaf area to decrease in area terms but increased in percentage terms [Table 2].

Physiological measurements

Compared with the C vines, the EBD vines showed higher water status [less negative midday leaf water potential] at all times for both years [Figures 4A and B]. At the beginning and the end of the water potential monitoring period in the first year, there were no differences between the mid-day leaf water potentials of the EBD and C vines, but in the second year, the EBD vines had higher water status at the first monitoring in June. In the period from DOY 211 to DOY 241 [from veraison to ripening] mid-day leaf water potential differences between EBD and C vines were registered. In

the EBD vines, midday leaf water potential remained above -0.8 MPa in 2015, falling to -0.8 MPa only at DOY 226. In 2015, the midday leaf water potential of the C vines reached -0.8 MPa somewhat earlier, at DOY 211 and stayed below this value until DOY 241. In 2016, mid-day leaf water potential fell to ~0.8 MPa for C vines quite early, at DOY 196, and remained low until berry ripening [DOY 256].

Compared with the C vines, the EBD vines showed higher rates of net photosynthesis, stomatal conductance and transpiration in both years [Figure 5A-F]. In 2015, differences were recorded in these values from

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Figure 3 – Monthly minimum, mean and maximum global radiation in the experimental vineyard (lat. 37°41’13” N; long. 15°07’00” E; elevation 602 m a.s.l.).

Table 2 – Intensity and incidence of early basal defoliation on vines.

| Parameter                                      | 2015               | 2016               |
|------------------------------------------------|--------------------|--------------------|
| Total leaf area/vine [cm²]                     | 32,000 ± 9,700     | 20,500 ± 3,970     |
| Removed leaf area/vine [cm²]                   | 10,400 ± 1,940     | 8,500 ± 1,160      |
| Removed leaf area/vine [%]                     | 35 ± 17            | 43 ± 9             |
| Main shoot length [cm]                         | 116 ± 2            | 117 ± 4            |
| Total leaf area main shoot [cm²]               | 3,470 ± 852        | 2,600 ± 431        |
| Removed leaf area/main shoot [cm²]             | 874 ± 327          | 1,020 ± 216        |
| Removed leaf area/main shoot [%]               | 27 ± 16            | 40 ± 10            |
| Lateral shoot length [cm]                      | 28 ± 7             | 26 ± 5             |
| Total leaf area lateral shoot [cm²]            | 1,140 ± 123        | 730 ± 170          |
| Removed leaf area/lateral shoot [cm²]          | 844 ± 200          | 450 ± 109          |
| Removed leaf area/lateral shoot [%]            | 74 A ± 16          | 64 B ± 20          |

For each year and parameter means indicated by different letters are significantly different (lowercase letter: p ≤ 0.05, uppercase letters: p ≤ 0.001) based on Tukey’s HSD test within year (± standard deviation).
DOY 196 onwards. Compared with the C vines, in the EBD vines, \( P_n \) was higher till the last measurement in Sept (with the exception of the measurement on DOY 226). Meanwhile, values of \( g_s \) were different till DOY 211, and those of \( E \) were different till DOY 226. In 2016, all gas exchange values for the EBD vines were higher from DOY 196 to DOY 256. As the season progressed, a worsening of all gas exchange values was seen in both years but these were generally lower in 2016. This was probably due to the higher average temperatures in the second year. Compared to the C vines, the SPAD index values in the EBD vines were generally more favorable in the leaves of both the main shoots and the laterals (Figure 6A-D). In both years SPAD differences between vines were recorded for the main leaves only (in 2015 at each date).

**Yield and biochemical analyses**

In 2015 and 2016, compared with the C vines, vine productive performance in the EBD vines was not different. Thus, there was no influence of EBD or year, and no interaction between factors. Similarly, there were no differences in yield/main shoot. However, cluster weights in 2015 were considerably higher in the EBD vines than the C ones, but no differences were recorded in 2016. The EBD also had no effect on yield/main shoot. Conversely, an increase in cluster weight was detected in 2015 and a year vs treatment effect was also found. The EBD treatment influenced cluster length in 2015 but not in 2016. There were no differences in berry weight with EBD but the difference between one year and the next was noted as was also observed in the case of the strong effect of EBD treatment on the rachis weight, in addition to increased skin weight (Table 3). The TSS did not increase in the EBD vines in either year, while the TA showed a difference. In the first year TA was generally higher in the EBD vines while in 2016 it decreased with the meteorological trend characterized by a strong increase in rainfall (Table 3).

The EBD treatment improved the quality attributes including the contents of total polyphenols, total flavonoids and total anthocyanins. In 2015 only the total polyphenols increased, while in 2016 all the quality attributes showed improvement. In 2016, both EBD and C vines showed a general decrease in their quality attributes (Figure 7A and B).
According to Spearman’s analysis we found significant correlation between the variables related to the size of the bunch, as expected. Rachis and skin weight also showed significant correlation ($p \leq 0.05$) with the majority of the qualitative traits studied (Table 4). In particular, the rachis weight was strongly negative correlated with anthocyanins ($r_s = -0.70$) and moderately with flavonoids, polyphenols and titratable acidity ($r_s = -0.55$, $-0.45$ and $-0.53$, respectively; $p \leq 0.05$). Skin weight was strongly positively correlated with anthocyanins and flavonoids ($r_s = 0.61$ and $0.63$), and moderately with polyphenols and titratable acidity ($r_s = 0.40$ and 0.46, respectively).

The PCA analyses (Figure 8) were carried out on a dataset, that included yield components, morphological data and cluster biochemical characteristics collected over the two-year trial (2015-2016) on EBD and C vines. Four principal components were extracted with eigenvalues $\geq 1$. These accounted for $72\%$ of the total variance. The first two components accounted for $51\%$ of the total variance. The PCA displays the relationships between the samples based on similarity in pattern of their original attributes leading to the highlighting of a number of groupings. All the 2015 observations were included in the positive part of PC1 ($35\%$ of the variance) and all the 2016 observations had negative scores. In the positive part of PC1 there was no separation between the EBD and C samples of 2015, whereas for 2016 the EBD and C samples were well separated. To identify the variables driving this pattern we generated a biplot (Figure 9). The first principal component was positively and well correlated with skin weight ($r = 0.8706$), anthocyanins ($r = 0.7687$), flavonoids ($r = 0.6225$) and polyphenols ($r = 0.5778$) while negative and less strong correla-

Table 3 – Influence of early basal defoliation on yield characteristics and main berry quality attributes.

| Parameters                  | 2015 Early basal defoliation | 2015 Control | 2016 Early basal defoliation | 2016 Control | Treatment (T) × Year (Y) |
|----------------------------|------------------------------|--------------|------------------------------|--------------|--------------------------|
| Yield/vine (kg)            | $2.44 \pm 0.36$              | $2.46 \pm 0.42$ | $1.94 \pm 0.32$              | $2.14 \pm 0.36$ |                          |
| Yield/main shoot (kg)      | $0.36 \pm 0.13$              | $0.42 \pm 0.11$ | $0.32 \pm 0.07$              | $0.36 \pm 0.11$ |                          |
| Cluster weight (g)         | $298.22 \pm 64.95$           | $233.33 \pm 30.96$ | $318.14 \pm 74.48$           | $319.79 \pm 70.11$ | ***                      |
| Cluster length (cm)        | $16.72 \pm 1.60$             | $15.95 \pm 1.42$ | $18.22 \pm 2.03$             | $16.85 \pm 1.39$ | *                        |
| Berries number             | $106.67 \pm 32.33$           | $112.67 \pm 35.69$ | $130.78 \pm 20.34$           | $139.33 \pm 30.04$ | *                        |
| Berry weight (g)           | $2.60 \pm 0.42$              | $2.62 \pm 0.36$  | $2.81 \pm 0.08$              | $2.86 \pm 0.09$  | *                        |
| Rachis weight (g)          | $8.38 \pm 3.62$              | $9.64 \pm 3.87$  | $26.26 B \pm 0.44$           | $32.89 A \pm 2.24$ | ***                      |
| Skin weight (g)            | $0.19 \pm 0.02$              | $0.17 \pm 0.07$  | $0.54 A \pm 0.08$            | $0.42 B \pm 0.03$ | ***                      |
| Total soluble solids (ºBrix)| $21.94 \pm 1.07$             | $20.79 \pm 1.23$ | $21.66 \pm 1.23$             | $21.49 \pm 0.66$  | ***                      |
| pH                         | $3.25 \pm 0.10$              | $3.31 \pm 0.13$  | $3.27 \pm 0.07$              | $3.27 \pm 0.06$  | **                       |
| Titratable acidity (g 100 mL–1)| $1.15 \pm 0.15$          | $0.93 \pm 0.15$  | $0.71 \pm 0.07$              | $0.76 \pm 0.14$  | **                       |

For each year and parameter, means indicated by different letters are significantly different (lowercase letters: $p \leq 0.05$; uppercase letters: $p \leq 0.001$) based on Tukey’s HSD test within year (± standard deviation). Interactions are indicated by: *Significant $p < 0.05$; **Significant $p < 0.01$; ***Significant $p < 0.001$. 

Figure 6 – SPAD index measured in early basal defoliated (EBD) and control (C) vines in main and lateral leaves. 2015 (A, B) and 2016 (C, D). For each year, treatment and day of year means indicated by different lowercase letters are significantly different ($p \leq 0.001$) based on Tukey’s HSD test (error bars indicate standard deviations).
tions were found with rachis weight ($r = -0.9499$), berry number ($r = -0.5972$), cluster weight ($r = -0.5643$) and berry weight ($r = -0.5164$). These variables discriminated between years, with small clusters with increased concentrations of antioxidants, alternating with higher yields with lower levels of antioxidants. In 2016, the separation between EBD and C was instead related to the variables associated with cluster size, including rachis weight, berry number, cluster weight and berry weight. The PC2 (16 % of the variance) showed a certain nega-

![Figure 7](image)

**Figure 7**– Total polyphenols, flavonoids and anthocyanins measured in early basal defoliated (EBD) and control (C) vines in 2015 (A) and 2016 (B). For each year, treatment and parameter columns indicated by different lowercase letters are significantly different ($p \leq 0.05$) or numerals ($p \leq 0.001$) based on Tukey’s HSD test (error bars indicate standard deviations).

![Figure 8](image)

**Figure 8**– Scatter plot with convex hulls of the Principal Component Analysis (PCA) carried out on the dataset collected during the two-year trial (2015-2016) on early basal defoliated (EBD) and control (C) vines.

![Figure 9](image)

**Figure 9**– Biplot of the Principal Component Analysis (PCA) carried out on the dataset collected during the two-year trial period (2015-2016) on early basal defoliated (EBD) and control (C) vines.

**Table 4** – Spearman’s rank-order correlation, significant correlations ($r$) for $p \leq 0.05$ are shown in bold characters.

| Parameter                      | Yield/vine kg | Yield/main shoot kg | Cluster weight g | Cluster length cm | Berries number | Berry weight g | Rachis weight g | Skin weight | Total soluble solids °Brix | Titratable acidity g 100 mL$^{-1}$ | Total polyphenols mg kg$^{-1}$ | Total anthocyanins mg kg$^{-1}$ | Total flavonoids mg kg$^{-1}$ |
|--------------------------------|---------------|---------------------|------------------|-------------------|----------------|----------------|------------------|-------------|---------------------------|-----------------------------------|----------------------------------|----------------------------------|-----------------------------|
| Yield/vine kg                  | 1.00          |                     |                  |                   |                |                |                  |             |                           |                                   |                                  |                                  |                             |
| Yield/main shoot kg            | **0.82**      | 1.00                |                  |                   |                |                |                  |             |                           |                                   |                                  |                                  |                             |
| Cluster weight g               | -0.02         | -0.13               | 1.00             | 0.68              | 0.18           | 0.51           | -0.52            | -0.23        | -0.23                     | 0.04                              | -0.17                            | -0.03                           | -0.44                       |
| Cluster length cm              | -0.07         | -0.13               | 0.68             | 1.00              | 0.12           | 0.24           | 0.22             | -0.24        | -0.24                     | 0.02                              | -0.09                            | 0.17                            | -0.12                       |
| Berries number                 | -0.21         | -0.20               | 0.17             | 0.24              | 0.35           | 0.56           | -0.41            | -0.25        | -0.25                     | -0.23                             | -0.04                            | -0.04                           | -0.41                       |
| Berry weight g                 | -0.18         | -0.20               | 0.18             | 0.24              | 0.35           | 1.00           | 0.55             | -0.28        | -0.28                     | -0.02                             | -0.35                            | -0.36                           | -0.31                       |
| Rachis weight g                | -0.31         | -0.27               | 0.51             | 0.22              | 0.56           | 0.55           | 1.00             | -0.84        | -0.84                     | 0.02                              | -0.53                            | -0.45                           | -0.70                       |
| Skin weight g                  | **0.35**      | 0.26                | **-0.52**        | -0.24             | -0.41          | -0.28          | -0.84            | 1.00         | 0.46                      | 0.46                              | 0.61                             | 0.63                            |                             |
| Total soluble solids °Brix     | -0.23         | -0.23               | 0.04             | 0.02              | -0.25          | -0.02          | -0.03            | -0.12        | 1.00                      | 0.00                              | 0.04                             | 0.00                            | -0.03                       |
| Titratable acidity g 100 mL$^{-1}$ | 0.23         | 0.06                | -0.17            | -0.09             | -0.23          | -0.35          | -0.53            | 0.46         | 0.00                      | 1.00                              | **0.38**                         | 0.48                            | 0.23                        |
| Total polyphenols mg kg$^{-1}$ | 0.19          | 0.10                | -0.03            | 0.17              | -0.04          | -0.36          | -0.45            | 0.40         | 0.04                      | 0.38                              | **0.56**                         | 1.00                            | 0.56                        |
| Total anthocyanins mg kg$^{-1}$ | -0.02         | -0.12               | -0.44            | -0.12             | -0.41          | -0.31          | -0.70            | 0.61         | 0.00                      | **0.48**                          | 0.56                             | 1.00                            | **0.36**                     |
| Total flavonoids mg kg$^{-1}$  | **0.39**      | 0.29                | **-0.35**        | -0.12             | -0.24          | -0.23          | **-0.55**        | 0.63         | -0.03                     | 0.23                              | 0.27                             | **0.36**                        | 1.00                        |
Discussion

In line with similar studies on EBD, canopy management has been shown to influence grape yield and quality attributes [Teixeira et al., 2013]. In a previous study in a warmer area, we identified the beneficial effects of early leaf removal for yield control, mainly in less vigorous cultivars, and also for improved color in the exposed clusters. Moreover, common endemic diseases were better contained due to freer air circulation [Nicolosi et al., 2012; Ferlito et al., 2014]. In this study, the EBD was carried out in a vineyard characterised by a loamy-sand, sub-acid and rich in organic matter soil. This is located in an area of Mount Etna that is generally cooler than coastal areas and wetter during the latter stages of berry ripening from mid Aug to Sept. These conditions, together with the high level of solar radiation for a relatively long season including most of the vine’s flowering-ripening stages, allow for the attainment of very high qualitative traits in berries, in terms of pigmentation and sugar content. After EBD, the total leaf area per vine was very different in the two years of this study, mostly the result of reduced cultivar vigor and productivity [Bekar et al., 2017]. This effect lasts for at least two years [Bledsoe et al., 1988] where reduced vigor is a result of lowered carbohydrate reserves. The EBD applied in the early stage of the flowering clearly changed the leaf area / fruit yield ratio. Despite this, it was proposed as a useful tool for yield regulation and quality improvement [Ferlito et al., 2014]. In our trial, the effect on yield was negligible, the percentage of leaf removal (33 % and 42 %) did not affect yield. This contrasts with the findings of Hunter and Visser [1990] who found a lower yield with 33 % defoliation during early fruit development (before pea size) and 66 % defoliation before veraison. These conflicting results may be related to the trial location - a cooler area and despite the berries being well exposed to the sun, the duration of exposure was shorter being located on an eastern slope. These results agree with those reported by Bledsoe et al. [1988] for a cooler region. Here, ‘Nerello mascalese’ is considered a mid-high vigor vine and at our location the period leading up to full ripening (between the end of Sept and the first week of Oct) is usually mild and with frequent rain (Figure 1). Under these conditions berry weight can increase rapidly.

The role played by the lateral shoots in the vine response to EBD would seem to be important. The percentage of leaf area removed was very high (74 % in 2015 and 64 % in 2016) even though this involved only basal leaves. The early timing of the EBD meant that these basal leaves were the only fully developed leaves on the vine. During the following weeks, the laterals from the upper productive canopy grew very rapidly, greatly increasing vine leaf area. This behavior has previously been noted by Poni et al. (2006) who observed significant increases in lateral shoot number as a result of severe early leaf removal. Generally, this response can influence canopy performance as shown by Candolfi-Vasconcelos and Koblet [1990] who reported that early defoliation caused an increase in leaf photosynthetic efficiency in the remaining leaves. This compensated for the stress of the early reduction in source/sink ratio [Pisciotta et al., 2013]. In our trial, the effect explains the best photosynthetic efficiency expressed in terms of SPAD values registered in the main and lateral leaves of defoliated vines. Moreover, it is possible that this type of defoliation improves berry metabolic activity increasing their leaf photosynthesis as has been reported by Hunter and Visser [1990]. As a consequence, according to Candolfi-Vasconcelos and Koblet [1990] and Bledsoe et al. [1988], this response to early defoliation did not have a strong effect on yield and its components. In 2016 defoliation caused an increase in skin weight. We suggest that the remaining leaves were well able to supply carbohydrates during berry growth [Intrieri et al., 2008] and this is probably advantageous in terms of protection against fungal diseases.

Despite the disturbance of the first stage of berry development by defoliation, the later performance of the vines was enhanced by elevated water status. From June until the harvest, the midday leaf water potential was above -0.8 MPa. According to the literature, a reasonable mid-day leaf water potential value for a well-irrigated vine would be approximately -0.8 MPa, while -1.2 MPa is considered moderately stressed and -1.5 severely stressed [Olivo et al., 2009; Intrigliolo et al., 2012; Triolo et al., 2018]. The C vines showed at least moderate stress in mid-summer and their water status was worse than the EBD vines at the same stage. The differences in canopy management throughout the season indicate differences in vine evapotraspiration. Therefore, the reduced number of leaves in the EBD vines improved their efficiency. The EBD improved the fruit qualitative traits. Moderate water stress usually improves berry quality, although the mechanisms responsible are not clear and the response is not consistent from year to year [Girona et al., 2006].

The most important period for TSS increase is ripening [Coombe, 1995]. The absence of differences in TSS, pH and TA probably depend on rainfall which in 2015 occurred mainly in Sept. Nevertheless, despite there being no differences in qualitative parameters, the results indicate good ripeness at harvest for the EBD clusters.

The positive effects of EBD on total polyphenols, flavonoids and anthocyanins are interesting as these increased levels of biologically active substances are important both for the quality of wine as well as human health. Moreover, these qualitative parameters are positively correlated with skin weight (Table 4), suggesting...
a role for solar radiation on the ripening processes and skin/flesh ratio increase. The placement in the positive part of PC1 indicates these compounds were at higher levels than the 2015 samples, while the 2016 samples had heavier clusters with more berries. Tardaguila et al. (2010) reported that EBD substantially increased phenol levels in the fruit and wine of two cultivars. In particular for ‘Carignan’, EBD resulted in heavier berries and enhanced grape and wine color. Also, much research has shown a correlation between berry light exposure and the accumulation of flavonols (Spayd et al., 2002). However, it is very difficult to establish a direct effect of defoliation on flavonol accumulation as there are numerous factors involved in their synthesis (Downey et al., 2006). Despite the reported negative effects of excessive light exposure on flavonol accumulation (Chorti et al., 2010), in our study, exposure of the fruit to sunlight seems to be the main factor responsible for the increase in flavonol, as had also been reported by Spayd et al. (2002). Flavonol accumulation probably also influences total anthocyanins, since despite flavonols being colourless, their action is explained as a co-pigment for anthocyanins (Downey et al., 2003a). This correlation suggests these compounds are synthesised together. This is somewhat surprising given that the effects of light and temperature are opposite-going in respect of the synthesis of these compounds. However, the above interactions are eliminated because flavonol accumulation occurs during flowering and from 15 days after veraison until ripening (Downey et al., 2003b), while anthocyanins are synthesised from veraison until full ripening (Lo Cicero et al., 2016). The latter period is certainly cooler and thus no temperature limitations were observed. Moreover, it is generally true that with low yields/vine, anthocyanins usually increase (Petrie and Clingeleffer, 2006).

Conclusion

In Nerello mascalese, the most important black variety in the Mount Etna district, the effectiveness of EBD in modifying important physiological and qualitative parameters was demonstrated. As for the physiological parameters, the vines subjected to EBD showed a better water status and higher levels of photosynthetic efficiency during berry growth and ripening and did not exhibit any reduction in yield. Leaves from laterals in EBD vines were able to supply carbohydrates during berry growth in both years after treatment. On the other hand, EBD had strong positive effects on berry quality attributes which were particularly significant for total polyphenols, anthocyanins and flavonoids. This was probably due to better cluster exposure to the sun coupled with improved vine water status and (for the anthocyanins) no excessive temperatures in the last phase of ripening. Although fungal disease was not a factor recorded in this study, at no stage did we see any evidence of disease in leaves or clusters of either EBD or C vines.

Acknowledgments

The authors would like to thank the farmers Costa and Indelicato for hosting the trials.

Authors’ Contributions

Conceptualization: Ferlito, F.; Nicolosi, E. Data acquisition: Ferlito, F.; Allegra, M.; Continella, A.; Stagno, F.; Nicolosi, E. Data analysis: Ferlito, F.; Allegra, M.; Torrisi, B.; Continella, A.; Stagno, F.; Nicolosi, E.; Pappalardo, H. Design of methodology: Ferlito, F.; Gentile, A.; La Malfa, S.; Nicolosi, E. Software development: Ferlito, F.; Allegra, M.; Nicolosi, E. Writing and editing: Ferlito, F.; Continella, A.; Gentile, A.; La Malfa, S.; Nicolosi, E.

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