SYNTHESIS OF A NOVEL CHALCONE DERIVATIVE FROM MYRISTICIN FOR SKIN CANCER PREVENTIVE ACTIVITY

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ABSTRACT

Chalcone with the presence of methoxy and two oxygenated substituents is predicted to have good anticancer activity. Therefore, a novel chalcone derivative 3 from myristicin was synthesized through three synthesis steps 1, 2, characterized by GC-Ms, IR, and \(^1\)H-NMR. Molecular target screening of skin cancer from the myristicin derivatives are Heat Shock Protein 90 (HSP90A), Prostaglandin Synthase 2 (PTGS2), and Dihydroorotate Dehydrogenase (DHODH). Molecular docking performed using AutoDock-Tools 1.5.6. The results showed that HSP90A, PTGS2, and DHODH were predicted as potential macromolecule targets with good interactions with myristicin derivatives for skin cancer therapy. Chalcone derivatives from myristicin are predicted as potent compounds against skin cancer molecular protein targets by docking molecular studies.

Keywords: Myristicin, Chalcone Derivative, Anticancer Activity, Molecular Docking

INTRODUCTION

Myristicin (6-allyl-4-methoxybenzo-1,3\[dioxole]) is a major molecule in nutmeg essential oil.\(^1,2\) It represents about 9-38 %, depending on the distillate material used.\(^3\) It can be isolated by distillation at 423 K, 25 mmHg.\(^4\) Myristicin has an allyl group that can be converted into an aldehyde group to be a benzaldehyde derivative. This conversion has been done with similar molecules like methyl eugenol and anethole through isomerization and oxidation.\(^5,6\) These benzaldehyde derivatives from myristicin can then be reacted with methyl phenyl ketone to produce novel chalcone derivatives. Chalcones have diverse biological activities, such as anticancer\(^7,8,9\), antioxidant\(^10\), antimalaria\(^11\), and anti-allergic activities.\(^12\) Chalcones derivative structure as an anticancer correlation has been investigated and concluded that the presence of 2ʻ oxygenated substituents and the presence of methoxy substituents are preferred structures.\(^13,14\) Therefore, we recently reported the novel chalcone derivative with the presence of methoxy and methylenedioxy substituents 3-(7-methoxybenzo[1,3]dioxol-5-yl)-1-phenylprop-2-en-1-one 3 through three-step synthesis(Scheme-1).

Skin cancer is an emerging public health issue with 40% of current cancer patients. Skin cancer contains squamous cell carcinoma (SCC), basal cell carcinoma (BCC), and malignant melanoma. SCC and BCC are two cancers of the non-melanoma skin arising from epidermal keratinocytes that correlate with prolonged exposure to the light, while melanoma of the skin affects melanocytes with intermittent exposure to the sun.\(^15\) The incidence of skin cancer is growing worldwide. Most skin cancers induced by ultraviolet radiation penetration and protection at the government and person-level advised.\(^16\) In-silico molecular docking modeled possible anticancer activities of myristicin. In this research, myristicin derivative performed molecular docking analysis toward active cancer-protein targets. A new study is also needed to discover new skin cancer compounds. One of the most successful methods of inhibiting skin cancer is to identify a particular target medication.
Molecular docking is a method that combines small molecules with macromolecular structure and ligand of the target protein identify and fit, shape a binding pose, and have an affinity. This approach has been commonly used to research medicines and integrate molecular docking with in vitro studies that have significance for finding the new drug. In this report, we looked at the usage of molecular docking and testing of small molecular inhibitors.

**EXPERIMENTAL**

**Material and Methods**

Myristicin (isolated results with 92% purity), potassium hydroxide, ethanol, anhydrous Na$_2$SO$_4$, chloroform, tween 80, potassium permanganate, dichloromethane, methyl phenyl ketone, sodium hydroxide, all chemical e-Merck with p.a grade. $^1$H-NMR recorded in 400 MHz Agilent NMR spectrometer, IR recorded in IR Prestige-21, Shimadzu spectrophotometer, GC recorded in GC 2010, Shimadzu and GC-Ms recorded in QP-2010 Plus, Shimadzu.

**General Procedure of Synthesis of Isomyristicin (1)**

Myristicin and 20 % KOH in ethanol (1:1) was reflux for 5 hours. The result extracted by chloroform-water up to neutral pH, and chloroform evaporated to have the reaction result. The reaction methods adopted from eugenol, anethole, and safrole isomerization reaction $^{18-20}$. Yield: 87%, m/z, two peaks with similar Ms profile 5.88% area and 91.40% area (GC), Irel, %: 192 (100), 177 (8), 161 (20), 147 (17), 131 (19), 119 (29), 103 (8), 91 (52), 77 (19), 65 (29), 53 (17), 39 (12), 27 (7).

**General Procedure of Synthesis of 7-methoxybenzo[1,3]dioxole-5-carbaldehyde (2).**

The mixture of I (3.90 g, 0.02 mol), sulfuric acid 50% (15 ml), tween 80 (0.1 g), dichloromethane added to 50 ml, and temperature adjusted under 10°C. Then 10% potassium permanganate solution was added dropwise into the mixture and keep the temperature. Then the reaction mixture heated to 40°C (till the purple color disappear). Then add sodium bisulfite (3 g). The result extracted by dichloromethane-water up to neutral pH, and dichloromethane evaporated to have the reaction result. Yield: 49 % m.p. 53°C. 97.47% area (GC).
General Procedure of Synthesis of 3-(7-methoxybenzo[1,3]dioxol-5-yl)-1-phenylprop-2-en-1-one (3). 2 (0.9 g, 0.005 mol) dissolve in 10 ml of ethanol, add methyl phenyl ketone (0.6 g, 0.005 mol), then add in dropwise 10 ml of 30% sodium hydroxide in ethanol and keep the temperature under 30°C. The mixture stirred for three hours. Solid results filtered and washed with water to neutral pH. Yield: 55% m.p. 88°C, 95.56% area (TLC).

GC-Ms Analysis Method
Helium as a carrier gas. 50°C of column oven temp, 300°C of injection temperature, split injection mode, pressure flow control, 13.0 kPa of pressure, 79.3 ml/min of total flow, 0.55 ml/min column flow, 26.8 cm/sec linear velocity, 3.0 ml/min of purge flow, 138.9 split ratios, Oven Temperature Program with rate 5°C from 50 – 240°C withholding time 5-7 min. The mass spectrometer operated in EI mode with 250°C of ion source temp, 300°C of interface temp, 3 min of solvent cut time, absolute detector gain mode, and 0.80 kV detector gain, 0 of the thresholds.

Molecular Docking
Chemical structures of 4 ligands, made of ChemDraw database manually. Three dimensional (3D) structures are HSP90 (PDB ID: 2VCJ resolution: 2.50 Å), DHODH (PDB ID: 5IKQ resolution: 2.41 Å), and PTGS2 (PDB ID: 2BXV with resolution of 2.15 Å). Macromolecules collected from the Protein Data Bank (PDB). A macromolecule was prepared using UCSF Chimera software. Docking investigations carried out using AutoDock Tools 1.5.6. Furthermore, the binding site study was performed by Discovery Studio.

RESULTS AND DISCUSSION
The isomyristicin 1 was synthesized. GC-Ms spectra of 1 show the presence of two peaks with the same m/z = 192 at 25.703 min 5.88% area and 27.057 min 91.40%. It indicated that reaction product 1 has isomers. The possible isomers for 1 are geometry isomer’s E and Z, the reason for the two diastereoisomer’s products explained by Hassam et al. as- “The rate of rotation in the conformation will determine the deprotonation rate (K_E and K_Z) so that the E and Z ratio of the reaction product will be related to the K_E / K_Z ratio. The configuration in the Z allylic anion transition state has higher energy than the E allylic anion. This is due to an unfavorable 1-3 interaction that will disturb the resonance stability of the phenyl” (Scheme-2).

Then from IR spectra shows the absorption that matches to isomyristicin functional group, Table-1. The 1H-NMR spectra of 1 show the perfect match for hydrogen chemical shift. The spectrum is very clear and convincing that the target molecule has obtained Table-2. The appearance of –CH3 protons signal at 1.84 ppm (dd) J: 1.2 Hz; J: 6.4Hz showing that the reaction was successful. The coupling value shows the E geometry isomer of 1 with J = 15.6 Hz.

For compound 2 synthesized, GC spectra show the presence of one major peak at 21.246 min 97.47% area. From IR spectra show the absorption of the aldehyde functional group that indicated the reaction was successful, Table-1. Hydrogen chemical shift of 2 also shows aldehyde proton type Table-2 at 9.82 ppm.

| Comp. | Type of Bond | C-H | –C-H | C=C | C=C Ar | C-O | C=O |
|-------|--------------|-----|------|-----|--------|-----|-----|
| 1     | C-H          | 2908 s, 1357 s | 3070 m, 3010 m, 925 s | 1627 s | 1504 s | 1087 s, 1126 s | - |
| 2     | C-H          | 2939 s, 1342 s | 3062 m, 3015 m, 925 s | 1581 s | 1512 s | 1087 s, 1134 s | 1651 s |
| 3     | C-H          | 2939 s, 1265 s | 3086 s, 3024 m, 964 s | 1589 s | 1504 s | 1018 s, 1134 s | 1674 s |

m = moderate-intensity, s = strong
For compound 3 synthesized, TLC scanner show \( r_f = 0.22 \) with 95.56% of purity. IR spectra show match absorption with molecule 3 functional group, Table-1. \(^1\)H-NMR spectra show that the reaction has done with the loss of proton aldehyde shift. The spectra also show that the geometry of this chalcone was \( E \). It showed that the \( J \) value of proton \( C_\alpha \) and \( C_\beta \) is higher than 15. Synthesis of chalcone derivatives using piperonal has a better % yield of 68% with 10% NaOH in ethanol and the same reaction time, as a comparison.\(^2\)

Table-2: \(^1\)H-NMR Value (ppm) of 1, 2 and 3

| Comp. | -OCH\(_3\) | O-CH\(_2\)-O | Allyl Protons | Aldehyde Proton | Aromatic Proton (1) | Aromatic Proton (2) |
|-------|-------------|--------------|---------------|------------------|---------------------|--------------------|
| 1     | 3.89 s (3H) | 5.93 s (2H)  | 1.84 dd (3H, -CH-C-H\(_2\), J= 1.2, 6.4 Hz); 6.07 dq (1H, =C-H-CH\(_2\), J= 15.6, 6.4 Hz); 6.28 dd (1H, =C-H-Ar, J= 2, 15.6 Hz) | - | 6.46 (1H); 6.56 (1H) | - |
| 2     | 3.92 s (3H) | 6.93 s (2H)  | - | 9.82 s (1H) | 7.38 (1H); 7.42 (1H) | - |
| 3     | 3.94 s (3H) | 6.89 s (2H)  | 7.38 d (1H, C\(_\alpha\)-H, J= 15.6 Hz); 7.76 d (1H, C\(_\beta\)-H, J= 15.6 Hz) | - | 7.16 (1H); 7.24 (1H) | 8.01 (2H); 7.57 (2H); 7.50 (2H) |

HSP90 is a chaperone that plays an important role in conformational ripening and protein activity inside the substrate membrane. ATP interaction with its HSP90 contributes to some autophosphorylation tyrosine residues. Earlier screened hits show an inhibitory HSP90 effect.\(^23\) UV-B-induced tumorigenesis is related to altered metabolism, and that DHODH fuels mitochondrial respiration for DNA repair and ATP synthesis coordination.\(^24\) HSP90 is essential in regulating several cell proteins. It was a good goal for many diseases like tumors and protein misfolding disorder.\(^25\)

Three compounds docked to binding sites of the protein target. The best-docked ligand in Hsp90A was 3. The binding energy score is -7.5 kcal/mmol. The best binding score in the DHODH compound is -10.3 kcal/mol with no hydrogen bond interaction.

The docking results between the PTGS2 protein and ligands from myristicin derivatives showed all ligands could interact with PTGS2. The lowest \( \Delta G_{\text{bind}} \) was 3, which has a binding energy score of -8.6 kcal/mol. This compound occupies the same binding site as the native ligand. There is no hydrogen bond with amino
acid residues at the binding site. Residu Tyr385 and Ser530 only have Van der Walls interaction for PGTS2 inhibitory activity.

| No | Compound | Macromolecule | ΔG Binding (kcal/mol) | Residue Interaction (H-bonds) | Non-hydrogen Interaction |
|----|----------|---------------|-----------------------|------------------------------|--------------------------|
| 1  | 1        | HSP90         | -6.3                  | Asn51                        | Ala55, Leu48             |
| 2  | 2        | HSP90         | -6.4                  | Asn51, Thr184                | Ala55, Leu48             |
| 3  | 3        | HSP90         | -7.5                  | Asn51                        | Ala55, Leu48             |
| 4  | 1        | DHODH         | -8.0                  | Ser305                       | Tyr256, Val143, Pro52, Val134 |
| 5  | 2        | DHODH         | -7.4                  | Asn145, Ser305               | Tyr356, Val143           |
| 6  | 3        | DHODH         | -10.3                 | -                            | Val143, Thr360, Ala55, Ala59 |
| 7  | 1        | PTGS2         | -6.5                  | Ser530                       | Val523, Lue527, Ala527, Gly526, Val349 |
| 8  | 2        | PTGS2         | -6.5                  | Ser530                       | Val523, Ala527, Lue532, Gly526 |
| 9  | 3        | PTGS2         | -8.6                  | -                            | Val523, Ala527, Lue539, Leu531, Val349 |

CONCLUSION

A novel chalcone derivative 3 from myristicin synthesized through three synthesis steps 1, 2, characterized by GC-Ms, IR, and 1H NMR. Compound 3 at HSP90A has ΔGbind -7.5 kcal/mol. The docking results between PTGS2 and DHODH proteins with ligands from myristicin-derived compounds showed that almost all ligands could interact with both targets. Ligan has the lowest ΔGbind value and has the best interaction of -10.3 kcal /mol and -8.6 kcal /mol in DHODH and PTGS2. Myristicin chalcone derivatives are predicted as potent compounds against skin cancer molecular protein targets by docking molecular studies.

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