Non-destructive mid-IR spectroscopy with quantum cascade laser can detect ethylene gas dynamics of apple cultivar ‘Fuji’ in real time

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Many plants, including fruits and vegetables, release biogenic gases containing various volatile organic compounds such as ethylene ($C_2H_4$), which is a gaseous phytohormone. Non-destructive and in-situ gas sampling technology to detect trace $C_2H_4$ released from plants in real time would be attractive for visualising the ageing, ripening, and defence reactions of plants. In this study, we developed a $C_2H_4$ detection system with a detection limit of 0.8 ppb (3σ) using laser absorption spectroscopy. The $C_2H_4$ detection system consists of a mid-infrared quantum cascade laser oscillated at 10.5 µm, a multi-pass gas cell, a mid-IR photodetector, and a gas sampling system. Using non-destructive and in-situ gas sampling, while maintaining the internal pressure of the multi-pass gas cell at low pressure, the change in trace $C_2H_4$ concentration released from apples (Malus domestica Borkh.) can be observed in real time. We succeeded in observing $C_2H_4$ concentration changes with a time resolution of 1 s, while changing the atmospheric gas and surface temperature of apples from the ‘Fuji’ cultivar. This technique allows the visualisation of detailed $C_2H_4$ dynamics in plant environmental response, which may be promising for further progress in plant physiology, agriculture, and food science.

Plants generate various phytohormones and substances in their tissues and organs at all growth stages, and their expression can change in response to infectious diseases. Ethylene ($C_2H_4$) is the lightest gaseous phytohormone. It is involved in many functions, such as leaf and flower senescence, promotion of seed germination and flowering, and softening and ripening of fruit1–4. Moreover, part of the $C_2H_4$ produced in the plant is released as a volatile organic compound (VOC). For example, ripe apples and avocados release approximately < 200 µL·kg$^{-1}$·h$^{-1}$ of $C_2H_4$5–8. Rice releases a low concentration of approximately 10 µL·kg$^{-1}$·h$^{-1}$ of $C_2H_4$ upon disease infection9. It has also been reported that soya beans and wheat release 10–60 ppb of $C_2H_4$ during periods of canopy expansion and rapid growth10. The amount of $C_2H_4$ released depends on the weight and growing conditions of the plants, and the concentration of $C_2H_4$ is strongly influenced by the gas sampling method. In many cases, non-destructive and real-time detection of ppb-level $C_2H_4$ allows the visualisation of $C_2H_4$ dynamics to plant environmental responses.

Some methods to detect trace $C_2H_4$ exist, such as gas chromatography-mass spectrometry (GC/MS), chemiresistive sensors, and laser sensing. GC/MS is a widely used method for detecting and analysing numerous VOCs, including $C_2H_4$. Nevertheless, GC/MS is not suitable for real-time detection of gas concentration changes, owing to the necessity of long measurement times and complex pretreatments. However, chemiresistive sensors made from single-walled carbon nanotubes are now available, which are tiny sensors that allow real-time detection of sub-ppm level $C_2H_4$. In addition, artificial metalloenzyme-based chemical biosensors have been used to detect ethylene gas spatially by fluorescence in fruits and leaves15. However, it is difficult to detect ppb-level $C_2H_4$. Moreover, these sensors respond to several gases (e.g., ethanol, acetaldehyde, and hexane) other than $C_2H_4$.

Optical sensing technologies based on laser spectroscopy allow rapid detection of trace VOCs. These include photoacoustic absorption spectroscopy (PAS)16,17, cavity ring-down spectroscopy (CRDS)18,19, and laser absorption spectroscopy (LAS)20,21 using tunable diodes or quantum cascade lasers (QCLs). A detection limit of 0.3 ppb.
(for 2σ) of C₂H₄ has been demonstrated with PAS using a CO₂ laser as the pump source. When a quantum cascade laser (QCL) was used as a pump source, a detection limit of 50 ppb of C₂H₄ was obtained, and C₂H₄ was successfully detected in the gases released from the apple. In this case, the gas released from the apple was collected in a slightly complicated manner using a sample bag. The apples were placed in the bag, and a pump fully evacuated the air inside the bag. Then, N₂ with 99.999% purity was introduced into the bag until atmospheric pressure was reached. After waiting for at least 15 min, the gas in the bag was used for the measurement. In CRDS, sub-ppb C₂H₄ detection has been realised using a near-IR distributed feedback (DFB) laser oscillating at 1.6 µm. However, these optical sensing methods are not yet robust and user-friendly because they require high-power lasers and optical cavities with precision adjustment.

LAS is highly robust and user-friendly compared to PAS and CRDS, owing to its simple measurement setup. Therefore, LAS is used for various forms of gas absorption spectroscopy, although its detection sensitivity is inferior to that of PAS and CRDS. Recently, using a 1.62-µm DFB laser and a 3.27-µm interband cascade laser, respective detection limits of 6.6 and 53 ppb C₂H₄ have been demonstrated. However, the detection limits of LAS-based optical sensing systems have not yet reached the ppb level. When detecting C₂H₄ released from plants using optical sensing technologies, it is necessary to place the plant into a sample bag or glass chamber to collect gas. Moreover, a carrier gas such as N₂ must be used. Non-destructive optical sensing technology that uses in-situ gas sampling methods in an open environment would represent an advance in biogenic gas analysis technology for plants. For example, they would allow us to observe rapid changes in plant environmental response and identify the source of VOCs. The resulting technology would be useful for early disease diagnosis and stress susceptibility monitoring in the plant physiology and breeding fields, and would facilitate efficient quality control of fruits and vegetables in the postharvest industry.

In this study, we developed a C₂H₄ detection system based on LAS using a QCL and demonstrated the real-time detection of C₂H₄ released from apples by non-destructive and in-situ gas sampling in an open environment. When apples were stressed by changes in atmospheric gas concentration or surface temperature, we also succeeded in observing rapid changes in the concentration of C₂H₄ released from apples.

**Results and discussion**

**Trace gas detection system.** Figure 1 shows a schematic diagram of the C₂H₄ detection system, which is mainly composed of the mid-IR QCL (QD10500CM1, Thorlabs Inc.), the astigmatic Herriott multi-pass gas cell (AMAC-76, Aerodyne Research), the mid-IR detector (PVI-4TE-10.6, VIGO system), and a gas dilution system. According to the HITRAN database, C₂H₄ has the strongest absorption lines at approximately 10.5 µm. Therefore, we selected a single-frequency distributed feedback mid-IR QCL that oscillated at a central wavelength of 10.5 µm (~949.5 cm⁻¹). The mid-IR QCL was operated in continuous wave mode, and the typical output power was 10 mW. The mid-IR QCL was controlled at 24 °C by integrated thermoelectric cooling, and the lasing wavelength was swept around the absorption peak at 10.5 µm by current modulation. The QCL output was passed through a lens pair and coupled to a multi-pass gas cell. The multi-pass gas cell provided a long optical pass length of 76 m to detect trace C₂H₄. The volume of the gas cell was 0.5 L. The output beam from the multi-pass gas cell was focused on the mid-IR detector. The inlet and outlet of the multi-pass gas cell were connected to the apples and scroll pump, respectively. By operating the dry scroll pump and simply making contact between the PTFE tube and the apple, the gas released from the apple was sampled in situ in an open environment without destroying the sample. By controlling the needle valves and mass flows installed at the inlet and outlet of the multi-pass gas cell, it was possible to perform LAS during constant flow of the sampled gas and while maintaining the internal pressure of the gas cell at an arbitrary pressure. By placing the apples in the glass jar connected to the gas dilution system, the atmospheric gas of the apple was also controllable. The gas dilution system was composed of mass flows, reference gases (N₂, CO₂, C₂H₄), and a zero-air generator. The CO₂ and C₂H₄ concent-
Concentrations were adjusted by controlling the flow rate of each reference gas and the zero-air generator. The spectral data were acquired on a PC with a time resolution of 1 s.

Transmission spectra and detection limit. Figure 2 shows the results of the measured transmission spectra of C\textsubscript{2}H\textsubscript{4} and CO\textsubscript{2} using the C\textsubscript{2}H\textsubscript{4} detection system. In this measurement, the multi-pass gas cell was directly connected to the gas dilution system without the apple to obtain the spectral data of the reference gases C\textsubscript{2}H\textsubscript{4} and CO\textsubscript{2}. Here, the absorption intensity was weaker than that of C\textsubscript{2}H\textsubscript{4}, but the absorption spectrum of CO\textsubscript{2} also appears near 949.48 cm\textsuperscript{-1}. Therefore, this system was able to measure both the C\textsubscript{2}H\textsubscript{4} and CO\textsubscript{2} spectra. C\textsubscript{2}H\textsubscript{4} and CO\textsubscript{2} were supplied to the multi-pass gas cell from the gas dilution system at concentrations of 10 ppm and 99.995%, respectively, and the transmission spectra were measured while reducing the pressure inside the gas cell from 20 to 4 kPa. By reducing the internal pressure of the gas cell, the pressure broadening of the gas spectra was suppressed, and spectral narrowing of C\textsubscript{2}H\textsubscript{4} and CO\textsubscript{2} was observed. These results show that spectral measurement with pressure reduction helps prevent interference between C\textsubscript{2}H\textsubscript{4} and CO\textsubscript{2} absorption.

Figure 3 shows the detection signal of the mid-IR detector for different concentrations of C\textsubscript{2}H\textsubscript{4}. The pressure inside the gas cell was constant at 12 kPa. The inset shows the histogram plot of the detection signal using the N\textsubscript{2} purged gas cell. The standard deviation (σ) was 0.32 mV.
The histogram plot, which corresponded to 0.8 ppb when compared with the linear fitting line in Fig. 3. However, in the lower concentration region (< 100 ppb), some measurement points deviated from the linear fitting line. When supplying low concentration C2H4 using the gas dilution system, the mass flow meter and pressure gauge have to read out a lower flow rate and pressure. Therefore, in the low concentration region, as compared with the high concentration region, the effective measurement accuracies of the mass flow meter and pressure gauge were lowered, which caused the deviation in the low concentration region. The effective quantification limit was estimated to be approximately 2.8 ppb, which was equivalent to ten-standard deviations (10σ). The detection ability of ppb-level C2H4 was sufficient to apply this system to the analysis of C2H4 released from plants.

Non-destructive and in-situ sampling of C2H4 released from the apples. We have demonstrated trace C2H4 detection released from apples by non-destructive and in-situ gas sampling in an open environment. Apples are typical climacteric fruits, characterised by an increase in C2H4 generation and respiration during ripening. In this test, the cultivar 'Fuji' was used to clearly detect the source of C2H4 generation in the apple, because it is known to produce remarkably low levels of ethylene gas. Figure 4 shows the locations of C2H4 concentrations. The gases were sampled in situ at the top and bottom of the apple cores and side surfaces of the apples. The sampling flow rate and time were 30 cc/min and 5 min, respectively, and the internal pressure of the gas cell was 12 kPa. Ten apples were used for the measurements. Error bars represent standard deviation. Two apples had cracks on the top of the core, and the other eight apples had no visible cracks. The cracks were from existing natural injuries; they were not intentionally made for the experiment. When using the cracked apples as samples, a C2H4 concentration of 340 ± 160 ppb was detected only at the top of the core. Five of the eight apples without cracks released C2H4 at a concentration of 580 ± 360 ppb from the bottom. In the three remaining apples, C2H4 was not detected in any position. No C2H4 was observed on the side surfaces of any apples. The reason for the absence of C2H4 detection may be that C2H4 released from the side surface was at a much lower concentration than the detection limit. C2H4 release at ~ 100 ppb concentration was observed from the side surface of a few apples when using the PTFE tube with a sucking disk at the tip for gas sampling. However, in this case, the sucking disk quickly sticks to the apple surface, and the apples become stressed by forced gas suction. This sampling method was not stress-free. Additionally, the method is not effective when using plants with bumpy surfaces as samples. Based on the above results, apples without cracks were used as samples in subsequent experiments, and the gas was sampled from the bottom of the core without the sucking disk.

Figure 5 shows the transmittance spectra of C2H4 and CO2 with changing concentrations of CO2 in the atmospheric gas of the apples. The apples were placed in a glass jar, and the concentration of CO2 supplied to the glass jar was controlled between 2.0 and 10.6% using a gas dilution system. The mid-IR absorptions of both C2H4 and CO2 were observed simultaneously in the wavelength region of 949.32–949.52 cm⁻¹. As the CO2 concentration increases from 2.0 to 10.6%, CO2 transmission near 949.48 cm⁻¹ decreases from 75 to 5%, which indicates that CO2 absorption has increased. At the same time, an increase in C2H4 transmission around 949.35 cm⁻¹ was observed. This result shows that C2H4 release from apples decreases as the atmospheric CO2 concentration of
the apple increases. It was confirmed that the C$_2$H$_4$ concentration decreased from 2200 to 1200 ppb as the CO$_2$ concentration increased from 2.0 to 10.6%. Using our C$_2$H$_4$ detection system, we succeeded in visualising the suppression of C$_2$H$_4$ production by CO$_2$ treatment, which is a commonly used controlled atmosphere storage method$^{32,33}$. The same phenomenon was observed in another cultivar termed “Kiou” (data not shown).

**Real-time detection of C$_2$H$_4$.** Figure 6 shows the results of real-time monitoring of how the C$_2$H$_4$ concentration released by the apple changes with the environmental response. As shown in Fig. 6a, the C$_2$H$_4$ concentration was measured in real time by changing the atmospheric CO$_2$ concentration of the apple. The experimental setup was the same as that in Fig. 5. The CO$_2$ concentration supplied to the glass jar containing the apple was controlled using a gas dilution system. The C$_2$H$_4$ concentration decreased from 1000 to 700 ppb as the CO$_2$ concentration increased to 12%. Then, it returned to 1000 ppb when the CO$_2$ concentration returned to atmospheric concentration. It was observed in real-time that the change in C$_2$H$_4$ concentration was inversely correlated with the change in CO$_2$ concentration. Figure 6b shows the change in the C$_2$H$_4$ concentration when the apple was heated. The apple was soaked in a Dewar vessel filled with water at 22 °C, and a thermocouple for temperature measurement was attached to the apple surface. When the temperature was rapidly increased to 35.5 °C by adding boiling water, the C$_2$H$_4$ concentration increased from 1000 to 1500 ppb. As the temperature decreased, the C$_2$H$_4$ concentration also decreased. We observed for the first time that the C$_2$H$_4$ concentration released from apples changed with a time constant of several minutes in response to environmental changes.

**C$_2$H$_4$ identification by TD-GC/MS and bioassay.** We confirmed the existence of C$_2$H$_4$ in the gas released from the apple by thermal desorption gas chromatography-mass spectroscopy (TD-GC/MS) and bioassays. Adsorbents (MonoTrap RGPS TD, GL Science) were inserted inside the PTFE tube while sampling the apple-released gas to prepare analytical samples for TD-GC/MS. The flow rate in the PTFE tube was 30 cc/min, and the time for the gas to flow through the tube was 90 min. The chromatogram of the gas released from the apples is shown in Fig. 7a. The sampled gas contained various VOCs (e.g., ethylene, acetaldehyde, and ethanol), and each peak identification of the VOCs is shown in Fig. 7a. The peak retention time of C$_2$H$_4$ was observed at 2.2 min. The mass spectrum at 2.2 min shows good agreement with that of C$_2$H$_4$.

Next, we demonstrated a banana ripening test as a bioassay using the sample gas, because bananas are also typical climacteric fruits. Figure 7b shows the results of ripening bananas with apple-releasing gas collected by in-situ sampling. We used unripened green bananas as the samples. Gas sampled in situ at a flow rate of 30 cc/min from the apple was allowed into the gas chamber at a volume of 2 L, and the green banana was placed in the glass chamber and exposed to the sampling gas for 24 h. After sample gas exposure, the green banana was removed from the glass chamber, and the ripening process was compared with another green banana which was not exposed to the sample gas. A comparative test was performed at room temperature. As a result, a clear visual difference between both bananas was observable on the third day, confirming that bananas exposed to gas released from the apple ripened faster than the other bananas. This result shows that C$_2$H$_4$ in the apple-releasing gas causes banana ripening by inducing climacteric rise$^{34-36}$. The TD-GC/MS and banana ripening tests showed
that the gases from the apple we sampled contained \(\text{C}_2\text{H}_4\). We confirmed that our \(\text{C}_2\text{H}_4\) detection system is effective for the non-destructive real-time monitoring of trace \(\text{C}_2\text{H}_4\) released from apples.

**Conclusions**

In this study, we succeeded in real-time monitoring of \(\text{C}_2\text{H}_4\) concentration changes in gas released from apples by non-destructive and in-situ gas sampling in an open environment. Monitoring was demonstrated using an original \(\text{C}_2\text{H}_4\) detection system based on LAS. We also demonstrated the rapid concentration changes of \(\text{C}_2\text{H}_4\) released from apples when apples were stressed by changes in atmospheric gas concentration or surface temperature. Furthermore, a special sampling chamber or bag is not required, which provides a great advantage in that gas can be sampled from an arbitrary place of fruit. Gas sampling from wounds, lesions caused by diseases and pests, and other localised areas of plants are possible. We expect that our method will allow real-time visualisation of \(\text{C}_2\text{H}_4\) dynamics following changes in environmental response. By changing the lasers that make up this gas detection system, various VOCs other than \(\text{C}_2\text{H}_4\) can be detected. More detailed information about the plant gas response can be obtained by combining it with conventional GCMS research. Our results show promise in advancing analytical techniques for plant environmental responses.

**Methods**

**Plant materials.** We purchased ‘Fuji’ apples (LOPIA Co. Ltd, Saitama, Japan) and used the same cultivar for all experiments and analyses. For the bioassay of banana ripening, we used green bananas (‘Cavendish’) without exposure to \(\text{C}_2\text{H}_4\). Green bananas were purchased from Konmatsu Co., Ltd. (Iwate, Japan). During the experiment, both the apples and bananas were stored at 20–22 °C.

**Reference gas preparation and internal pressure control of multi-pass gas cell.** We prepared three reference gases and a zero-air generator (Zero Air Supply Model-111, Thermo Fisher Scientific, Waltham, MA, USA) for the gas dilution system. The three reference gases were \(\text{C}_2\text{H}_4\) (10 ppm), G1-CO\(_2\) (99.995%), and G1-N\(_2\) (99.99995%), and all gases were charged in 10 L high-pressure gas cylinders. According to the manufacturer specifications, the zero-air generator provides dry air without NO\(_x\), SO\(_2\), O\(_3\), CO, and hydrocarbons. All gas supply lines had mass flow meters (Standard Mass Flow Controller MODEL 3660 SERIES, KOFLOC, Kyoto, Japan), and the flow rate could be controlled. By changing the mixing ratio with N\(_2\) or zero air, \(\text{C}_2\text{H}_4\) and CO\(_2\)
can be diluted to any concentration. The reference and sampling gases were introduced into the multi-pass gas cell using a dry scroll pump connected to the gas cell. Then, using a needle valve attached to the inlet and outlet of the gas cell, the internal pressure of the gas cell can be adjusted between 2 and 100 kPa while maintaining a constant flow rate in the gas cell.

Thermal desorption-gas chromatography/mass spectrometry (TD-GC/MS). We used thermal desorption gas chromatography-mass spectroscopy (TD-GC/MS) to analyse numerous VOCs present in the apple-releasing gas. The TD-GC/MS consisted of a GC/MC (GCMS-TQ8040, Shimadzu, Kyoto, Japan) and a high-performance multimode injector (OPTIC-4, Shimadzu). For the thermal desorption analysis, we used MonoTrap RGPS TD (GL Science) as the absorbent. The adsorbents were inserted into a glass tube (MonoTrap TD Liner for OPTIC/LINEX) and introduced into the injector port for thermal desorption. The initial temperature of the injector port was set at 40 °C, and then the temperature was increased to 230 °C at a rate of 10 °C/s to desorb the VOCs in splitless mode. The desorbed VOCs were concentrated at the headspace of the GC column using cryofocus trapping cooled to −150 °C. After cryofocus trapping, the GC oven temperature was maintained at 35 °C for 3 min, increased to 200 °C (rising rate = 10 °C/min), and then held at 200 °C for 2 min. An Rt-Q-BOND column (ID = 0.32 mm, length = 30 m, df = 10 μm, RESTEK) was used to separate the eluting compounds, and the carrier gas was high purity He (99.9999%) with a constant flow of 5 ml/min. The MS detector was operated in scan mode from 20 to 300 amu at 200 °C for electron ionisation. C2H4 identification was achieved by comparing the mass spectra and retention times with the reference gases of C2H4. We used GCMSolution software (Ver. 4.45) and Mass Spectral library (NIST 14) to analyse the retention time and mass spectra, respectively. In this measurement, an apple confirmed to absorption spectral peak of C2H4 by our C2H4 detection system was used as a sample. The thermal desorption analysis using the absorbent was carried out three times and the C2H4 was detected in all measurements.

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Figure 7. C2H4 identified in sample gas released from an apple. (a) Chromatograms of TD-GC/MS analysis for the sample gas. The insets show the C2H4 chromatogram peak at 2.2 min retention time and the mass spectrum of the C2H4. (b) Bioassay of banana ripening. Group (A): bananas without exposure to the gas released from apples. Group (B): bananas with exposure to the gas released from an apple over 24 h.
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Author contributions
M.Y. conceived and conducted the experiments and wrote the manuscript with contributions from all authors. Y.K. and T.A. performed the experiments and analysed the data. T.M. proposed the concept of GC/MS and bioassay experiments. S.W. supervised the Project. All the authors contributed to the discussion and reviewed the manuscript.

Competing interests
The authors declare no competing interests.

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