Effects of introducing Xpert MTB/RIF test on multi-drug resistant tuberculosis diagnosis in KwaZulu-Natal South Africa

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Abstract

Background: An algorithm instituted following Xpert MTB/RIF (Xpert) introduction in South Africa advocates for treating all Xpert rifampicin resistant patients as MDR-TB cases while awaiting confirmation by phenotypic or genotypic drug susceptibility testing. This study evaluates how the Xpert has influenced the diagnosis and management of drug resistant TB in the highest burdened district of KwaZulu-Natal Province.

Methods: Data was retrospectively collected from all patients with rifampicin resistance on Xpert performed between March 2011 and April 2012. Xpert results were compared with those of phenotypic and genotypic drug susceptibility testing. Patients’ records were used to determine the time to treatment initiation.

Results: Out of 637 patients tested by Xpert, 50% had confirmatory results, of which a third were sent on the same day as Xpert test. The rate of rifampicin discordance and monoresistance was 8.8% and 13.4% respectively and there was no difference between phenotypic and genotypic confirmation. Among those who had been initiated on treatment, 28%, 40%, 21% and 8% of patients commenced within 2 weeks, 1 month, 2 months and 3 months of Xpert testing respectively, while the remaining 3% were observed without treatment.

Conclusion: This study emphasizes the importance of complying with the algorithm in confirming all Xpert rif resistant cases so as to ensure proper management of these patients. Despite the rapidity of the Xpert results, only about 70% of patients had been initiated treatment at one month. Therefore there is a definite need to improve the health systems in order to improve on these delays.

Keywords: Xpert, Rifampicin, Resistant, MTB, South Africa

Background

Whilst South Africa is the third country with the highest burden of tuberculosis (TB) in the world, following India and China, it is also overburdened with an HIV prevalence of 17.9% equating to 17% of the global burden of HIV [1-3]. More than 60% of TB infected patients in South Africa are co-infected with HIV, further complicating the diagnosis and management of these two diseases. The country is ranked fifth in the world with highest drug resistant TB cases and the multi-drug resistant (MDR) TB prevalence is 1.8% among new TB cases and 6.7% among TB retreatment cases [4]. The KwaZulu-Natal (KZN) province alone which is also disproportionately afflicted with HIV and TB co-infections, carries 36% and 37% of the country’s burden of MDR and XDR-TB cases respectively [5]. It was this overwhelming burden of disease that prompted the South African Department of Health to introduce the Xpert MTB/RIF (Cepheid GeneXpert, Sunnyvale, Ca, USA) in 2011 to improve the diagnosis of TB and detection of drug resistant TB.

The introduction of the Xpert MTB/RIF (Xpert) test replaces the two sputum smears in the initial diagnoses of TB. All cases detected as having rpoB mutations on this assay are initiated on MDR-TB treatment while a second sample is sent for culture and drug susceptibility testing (DST) to confirm these results [5]. It was this overwhelming burden of disease that prompted the South African Department of Health to introduce the Xpert MTB/RIF (Cepheid GeneXpert, Sunnyvale, Ca, USA) in 2011 to improve the diagnosis of TB and detection of drug resistant TB.

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associated with culture results, thus reducing patient loss to follow up related to long turnaround time results [6]. The mean delay in starting MDR-TB treatment has previously been 12 weeks [7] prior to the introduction of decentralised MDR-TB treatment sites which increased the capacity for earlier treatment initiation. Decentralisation in KZN resulted in an improved but still unacceptable time to treatment of 72 versus 93 days in centralised site [8].

Despite the fact that rifampicin resistance has been used as a surrogate marker for MDR-TB as more than 90% of these cases are associated with isoniazid (INH) resistance [9-11], there is a growing concern for the increasing rifampicin monoresistance in South Africa [12,13]. This implies that if these patients were treated as having MDR-TB, a significant number would be inadvertently denied INH. Although several studies have shown a high Xpert rifampicin resistance specificity (94-100%), the positive predictive value remains low in areas with a low prevalence of MDR-TB like South Africa [14,15], hence the need for confirmatory culture. Our guidelines for drug resistant TB require that the confirmatory sample be sent to the laboratory when the patient returns for the Xpert results. There are no studies that have assessed the frequency of return for second sampling and the extent of delay thereof.

The aim of this study was to ascertain how the introduction of Xpert has influenced the health care practices in the management of MDR-TB in KZN by determining the rate of patient return for the confirmatory culture sample, whether it could be linked to the initial GXP result and the period it takes before starting MDR-TB treatment. In addition to this, we also aim to determine the rate of rifampicin monoresistance and concordance when compared with culture.

**Methods**

**Study design**

The study was an observational retrospective study using data of all patients who showed rifampicin resistance on the Xpert test performed on the GeneXpert at the Prince Mshiyeni Memorial Hospital (PMMH) laboratory over one year period between March 2011 and April 2012.

**Study setting**

PMMH is a public regional hospital located in the biggest township in KZN in Durban. It has 1200 beds which serve the surrounding community including 18 primary health clinics. It was selected for the placement of the GeneXpert 48 infinity due to the high burden of TB and HIV in the area. Surrounding hospitals in the sub-district also send their sputum specimens for processing at this Hospital. Patients diagnosed with drug resistant TB are further referred to a 320 beds hospital that manages drug resistant TB patients.

**Laboratory procedures**

The National Health Laboratory Service (NHLS) operates on a national laboratory information system (LIS) which allows tracking of patients results as all the public laboratories utilise this LIS. The Xpert MTB/RIF test was performed according to the manufacturer's instructions. The GeneXpert infinity is interfaced with the LIS; therefore all tests performed by the instrument are available on the system. All the data from the LIS is processed centrally by the central data warehouse (CDW). The Xpert results are printed directly at the patient facilities using SMS printers, while printed copies are distributed from the laboratory to the healthcare facilities on the next day.

The *Mycobacterium tuberculosis* (MTB) culture and drug susceptibility testing (DST) was performed at the Central Academic Laboratory using the standard laboratory procedures for culture and indirect line probe assay (LPA) [GenoType MTBDRplus assay, Hain Lifescience, Nehren, Germany] and/or the 1% indirect agar proportion method (APM) using Middlebrook 7H10 agar. This laboratory is a quality assured reference laboratory for the KZN province. Culture for TB was performed using BACTEC mycobacteria growth indication tubes (MGIT) 960 system [BACTEC MGIT Becton Dickinson, USA]. Positive cultures were tested for susceptibility to first line TB drugs using the LPA first in order to confirm resistance to rifampicin and isoniazid. Where these results demonstrated resistance to any of the drugs tested, further testing for both first and second line drugs was performed using APM. Samples coming from patients in the MDR-TB units were tested using APM with no preceding LPA.

**Data collection and analysis**

The Xpert rifampicin resistant data was obtained from the CDW and from the LIS. Demographic data was used to link Xpert results with the culture results. Patients that could not be reliably linked were regarded as untraceable and excluded from further analysis. All samples registered in the laboratory within 48 hours of each other were regarded as same day samples. Where both the LPA and APM were performed, the result of the APM was used for comparison with the Xpert result, where only LPA was done; it was used for comparison. The information containing the date of commencing MDR TB treatment was collected from the TB register at admitting Hospital. Patients were traced up to 3 months from the date of Xpert sample and cases which were not identified on the TB register after 3 months were considered as missing.

For those patients with both the Xpert and culture results, the period between the two samples was determined. Descriptive statistics were used to report proportions of patients who measured rifampicin monoresistant, MDR-TB and rifampicin susceptible on both LPA and DST tests. A Chi square test for agreement was used to compare the
agreement of results between the LPA and APM diagnostic methods. The amount of time taken to commence treatment was calculated by subtracting the date of Xpert registration in the laboratory from the date of starting TB treatment. Time to treatment was categorized and described. Analysis was performed using SAS version 9.3 (SAS Institute Inc., Cary) and graphs were drawn using Microsoft Excel.

The study protocol was approved by the University of KwaZulu-Natal Biomedical Research Ethics Committee.

**Results**

Out of 34,444 patients tested for TB using Xpert at PMMH laboratory between March 2011 and April 2012, 5,870 were positive for TB. Of these, 637 showed rifampicin resistance but after excluding the untraceable results, only 268 (42%) had a culture sample for comparison with the Xpert and therefore eligible for further analysis (Figure 1). About a third (32.8%) of the confirmatory samples were submitted simultaneously with the Xpert samples (paired samples) with an additional 20% being sent within two weeks of Xpert (Table 1). Most of the confirmatory samples submitted within 2 weeks of Xpert (63%) were from the same hospital where Xpert sample was collected while those collected at a different hospital to that of Xpert accounted for the majority of those collected later (Figure 2).

Most of the rifampicin resistance confirmation was performed using APM. Table 2 shows the summary of the LPA and APM results for the Xpert rifampicin resistant samples. The Xpert MTB/RIF assay showed 8.8% discordance when compared to culture and LPA/APM and the overall rate of rifampicin monoresistance was 13.4%.

When comparing the LPA and APM results for the 144 samples where both were available, 87% (126) of LPA results agreed with APM (Table 3). An agreement test failed to show a significant difference in agreement between LPA and APM, however there does appear to be a trend ($p = 0.062$).

Out of the 268 patients with confirmatory samples, 98 (36%) were not found on the MDR-TB treatment register within 3 months of Xpert samples. Of the remaining 170 patients that started treatment, 28% were started within two weeks of Xpert test, with the majority (68%)

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**Figure 1 Flow diagram of results of patients with Rifampicin resistance on Xpert.**
already started treatment within a month of Xpert (Table 4).

Discussion
This study reveals lessons that have been learnt during the initial introduction of Xpert. Considerable gaps were identified regarding compliance with the current national guidelines in the diagnosis and management of MDR-TB using Xpert [16].

There was poor adherence to the Xpert algorithm especially with sending a confirmatory sample. Even though we could not show the reasons behind this finding, several explanations could be attributed to this non-compliance, including poor training of health care workers at the time of Xpert introduction. The rollout of Xpert preceded the training of clinicians; this may have lead to poor understanding of the role of the assay in the diagnosis of DR TB. Other reasons could be patient related, as shown in several studies conducted in South Africa where the initial defaulter rate was between 16 and 17% [17,18]. The majority of the patients in these studies could not be interviewed in order to find reasons behind their default because they were either lost to follow up (45%), or demised prior to starting treatment (24%). In recognition of this, the provincial department of health in collaboration with the laboratory services has recently established an alert system where health care workers at patient facilities are alerted of MDR TB results. This should lead to prompt tracing of patients for further management.

A significant number of samples were submitted simultaneously for both Xpert and culture. While this may have ensured timely confirmation of Xpert rifampicin resistant cases, both tests were performed concurrently without considering the Xpert results. This practice could lead to wastage of resources where Xpert results do not require a confirmatory test. The Western Cape which is one of the provinces in South Africa has a protocol where two samples are submitted concurrently, one for Xpert and should the rifampicin be resistant, the other one is used for confirmation using LPA or APM [19]. Should this practice be adopted by the rest of the country, it could improve the confirmation of Xpert resistant cases.

Our study shows a slightly lower concordance between Xpert and culture than previously reported despite the previous assay improvements made in 2010 [20]. Several studies conducted in South Africa have shown excellent Xpert rifampicin resistance specificity of between 99.4 and 99.5% [19,21]. However these studies used molecular assays (LPA and sequencing) which have recently been proven to show a better concordance to Xpert than the phenotypic assays [22]. Interestingly, as shown in Table 3, out of the seven patients who had discordant Xpert rifampicin resistance where both the LPA and APM were performed for confirmation, only two were sensitive to rifampicin on LPA while all were sensitive on APM. We were unable to show any significant difference between using the phenotypic or genotypic test for confirmation because of the small number of cases.

Table 1 Time between date of Xpert and culture samples

| Period between samples | Number of patients | Percentage |
|------------------------|--------------------|------------|
| Culture prior to XGP   | 22                 | 8.2%       |
| Paired samples         | 88                 | 32.8%      |
| 0-2 weeks              | 53                 | 19.8%      |
| >2-4 weeks             | 55                 | 20.5%      |
| >4-8 weeks             | 26                 | 9.7%       |
| >8-12 weeks            | 9                  | 3.4%       |
| >12 weeks              | 15                 | 5.6%       |
| Total                  | 268                | 100%       |

Figure 2 Site and timing of collection of Xpert and Confirmatory samples.
Traditional phenotypic DST is considered the gold standard for TB drug resistance testing; however, studies are now showing cases of TB isolates that have rpoB mutation that are not detected by phenotypic assays [23-26]. Williamson et al. [26] reported cases with INH mono-resistance that demonstrated genotypic rifampicin resistance while phenotypically susceptible to rifampicin on proportion method using MGIT. These patients subsequently failed treatment. Almost half of our culture confirmed rifampicin susceptible (discordant) cases were resistant to INH. The isolates were not available for sequencing, but further studies that look at the presence of these low level mutations are warranted.

The discordance of rifampicin resistance can lead to diagnostic and management dilemmas because DR-TB management has serious consequences. Patients are not only given more toxic and less effective second line drugs, but they are also subjected to socioeconomic disruption as they have to get their treatment from the designated MDR-TB treatment facilities. Given the limitations of the available laboratory diagnostic assays, discordance between Xpert and phenotypic testing require confirmation by another molecular assay. Additionally, larger studies are essential to ascertain the clinical significance of such cases.

Our findings demonstrate a high rate of rifampicin monoresistance (13.4%) and there was no statistical difference between using LPA and APM, hence both these test were reliable in confirming MDR TB. A study conducted by Coovadia and colleagues in their setting between 2007 and 2009, showed the overall rifampicin monoresistance rate of 8.8% [13]. This increasing rifampicin resistance show that rifampicin can no longer predict MDR TB in a significant number of patients, further emphasizing the importance of performing confirmatory DST. Whereas these patients might benefit from adding isoniazid on their MDR TB treatment regimen while awaiting culture and DST results, the majority of patients would be exposed to this drug unnecessarily, particularly because most of these patients are already on multiple drugs due to co-infection with HIV.

Among patients found on the TB register, 68% started treatment within one month of Xpert test. Although this is a major improvement when comparing to the pre-Xpert period [7,27], the delay is still substantial considering the rapidity of the Xpert together with the decentralization of the MDR TB treatment. In a feasibility study conducted by Boehme et al. [28], Xpert reduced time to treatment in smear negative TB from 56 days to 5 days, the latter has also been advocated by the SA department of Health as the cut off period for starting MDR TB treatment. The TB/HIV care project in rural KZN was able to put 89% of patients on treatment within five days [29]. This was achieved by using Xpert for diagnosis, while the results were communicated using cell phones to link patients with community health workers, clinics and laboratory. This could be adopted by the department of health in order to improve time to treatment initiation. An efficient health care system is crucial to achieve a significant impact on drug resistant TB management.

A number of studies performed in South Africa have shown that when Xpert is done closest to the point of care, treatment can be initiated on the same day [30-32]. An average of 71% of results achieved the targeted turnaround time of 48 hours at PMMH laboratory over the past year. Lawn and colleagues showed that performing Xpert at a centralised laboratory increases the turnaround time of results by up to four days, which could impact on the initiation of treatment [14,33]. Whilst these studies mainly involved patients with drug sensitive TB, further decentralisation of MDR TB treatment sites together with

**Table 2 Summary of Culture and LPA/APM in patients with rifampicin resistant Xpert**

| Test                              | Rifampicin sensitive | Rifampicin monoresistant | MDR-TB    | Inconclusive |
|-----------------------------------|----------------------|--------------------------|-----------|--------------|
| Culture and LPA (N = 180)         | 14 (7.8%)            | 30 (16.7%)               | 130 (72.2%) | 6 (3.3%)     |
| Culture and APM (N = 226)         | 11 (4.9%)            | 31 (13.7%)               | 184 (81.4%) | 0 (0.0%)     |
| Total Confirmed (N = 262)         | 23 (8.8%)            | 35 (13.4%)               | 204 (77.8%) | N/A          |
|                                   | 5.4-12.2%            | 9.2-17.5%                | 72.8-82.9% | N/A          |

**Table 3 Comparison of LPA and APM results**

| Number of samples | LPA results | APM results |
|-------------------|-------------|-------------|
|                   | RIF susceptible | RIF monoresistant | MDR-TB | Inconclusive |
| RIF susceptible   | 2           | 0           | 0     | 0           |
| RIF monoresistant | 3           | 17          | 6     | 0           |
| MDR-TB            | 1           | 2           | 107   | 0           |
| Inconclusive      | 2           | 3           | 1     | 0           |

Boldface shows agreement between LPA and APM results.
transfer of Xpert to the near point of care could bring significant improvement to these treatment initiation delays. The number of patients that were untraceable is a cause for concern. These were patients who did not have enough details on the computer system to link the Xpert result to the culture result. This reflects poor data capturing and matching; these results would erroneously cause increase in the rate of discordant result. Given that patients with MDR TB are treated in designated facilities which are different to the patient’s local facility, it becomes very important to have correct and complete patient details so as to follow up on the results. Failing to do so may result in repeated testing which not only leads to wastage but may also cause delays in changing to appropriate treatment where necessary.

There were several limitations to this study. Even though the matching of Xpert and culture results was carefully carried out and results with insufficient details were excluded from analysis, this method is not perfect; hence there may still be minor mismatching. We studied the period of the initial roll out of the Xpert; there may have been subsequent improvements in the management of MDR TB patients as the test was getting more recognised throughout the country. Finally, while the area studied is a high burden TB region, our findings may not be representative of the whole province.

### Conclusion

A few lessons have been learnt following the introduction of Xpert. Introducing a new test requires proper preparation and training of those involved in order to ensure compliance with the stipulated guidelines. Our findings show a significant amount of discordance between Xpert and phenotypic and/ or genotypic drug susceptibility testing which underscores the importance of taking a second sample in cases of Xpert rifampicin resistance. This will also assist in detection of the increasing rifampicin monoresistance. Studies ascertaining the causes and clinical significance of discordance between phenotypic and genotypic assays are warranted in order to solve the diagnostic dilemmas that often accompany such results. Despite having a rapid diagnostic tool which can generate results in a few hours, system associated challenges continue to result in delays in treatment initiation.

### Abbreviations

APM: Agar proportion method; CDW: Central data warehouse; DST: Drug susceptibility testing; INH: Isoniazid; KZN: KwaZulu-Natal; LIS: Laboratory information system; LPA: Line probe assay; MDR-TB: Multidrug resistant tuberculosis; MGIT: Mycobacteria growth indicator tube; NHLS: National Health Laboratory Services; PMMH: Prince Mshiyeni Memorial Hospital; Xpert: Xpert MTB/RIF.

### Competing interests

The authors declare that they have no competing interests.

### Authors’ contributions

ND and RP came up with the concept and developed the study protocol. ND contributed to the study design, collection and analysis of the data and wrote the manuscript. LW analysed the data and contributed to the writing of the manuscript. LC assisted with the collection and analysis of data. KM is the supervisor who provided intellectual content in the study design, analysis of data and the writing of the manuscript. All authors read and approved the manuscript.

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