The role of ecological groups in the formation of cyanobacterial communities in the ecosystems of the North Azov region (Ukraine)

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The role of Cyanoprokaryota ecological groups in the ecosystems of the North Azov region was revealed in this work. On the territory of Pryazovsky National Nature Park, 9 experimental polygons were studied, which covered steppe areas or slopes, salt marshes, coastal sandy soils and water bodies (rivers, lakes, estuaries, sea bays, lagoons). As a result of research on the territory of Pryazovsky National Nature Park, 124 species of cyanoprokaryotes were identified, which include 127 intraspecific taxa. It was proved that the procedure of canonical correspondence analysis is the most suitable for the analysis of the species matrix. The axes identified as a result of the ordination procedure, which indicate the coordinated dynamics of the species, correlated with both synecological characteristics, such as diversity indicators, and with autoecological characteristics, such as ecotypes of cyanoprokaryotes in relation to habitat types or types of adaptation to salinity conditions. The first four canonical axes together explain 47.5% of species matrix variability. Canonical axis 1 explains 18.0% of the variability of the species matrix and is mostly marked by subaereal cyanobacteria and eurybionts. This axis indicates the presence of a gradient of salinity conditions where the most saline conditions correspond to the positive values of the axis, and the negative values correspond to less saline. Canonical axis 2 describes 12.1% of species matrix variability. This axis differentiates aquatic ecosystems from others. Canonical axis 3 explains 10.0% of the communities’ variability. This axis distinguishes freshwater ecosystems from saline ecosystems. Markers of freshwater communities are stenotopic halotolerants, which are narrow-range, common mainly in the temperate zone of Europe. The canonical axis 4 explains 7.3% of variability of the matrix of species and is able to differentiate sand ecosystems. The ecotopic structure and geographic range width of community species have the greatest independent value among the considered sources of variation. The independent role of adaptation to the salinity conditions of the ecotope and the role of the type of ecosystems is somewhat smaller. The interaction between the sources of variation is important in the variation of the structure of communities. The interaction between the ecotopic structure and the geographic range width of species and the triple interaction between the ecotopic structure of a community, the width of the geographic range of species and the ecosystem type plays the greatest role in the variation of community structure. Ecotropic groups, which indicate the preference of a particular habitat, correlate with the species composition of the communities. It is shown that the ratio of ecotopic groups in a community is a characteristic that reveals the features of the community as a whole.

Keywords: diversity; ecological groups; environmental gradients; geographical range; variation partitioning.

Introduction

Cyanoprokaryota (or Cyanophyceae) is a group of ubiquitous prokaryotic organisms that have a high ecological plasticity, which enables them to inhabit different, often even extreme, habitats (Pelekata et al., 2016). Cyanophyta are very widespread: in seas, rivers and hypersaline waters, in soils, on snow and ice, in hot springs, in aerophytes conditions, etc. (Hoffman, 1999; Liu et al., 2016). Cyanophyta are extremely tolerant of extreme conditions on the planet (Rampelotto, 2013). In model experiments, the blue algae stayed alive in the temperature range from −195 to +130 °C (Inoue et al., 2001; Červený et al., 2015), pressure from 0.05 to 300 atm (Qin et al., 2014; Kitahara et al., 2019), and can withstand exposure to 160 kR/hr of radioactive cobalt (Microcoleus vaginatus – up to 1280 kR/hr) (Kraus, 1969; Yatagai & Ishioka, 2014; Badri et al., 2015; Varshey et al., 2015). Blue algae play a planetary role in nitrogen balance (Jiang et al., 2017; Zhao et al., 2019). This is due to the ability of Cyanophyta’s representatives to absorb nitrogen directly from the atmosphere (Bothe et al., 2010). The phenomenon of restoration of soil fertility, preservation of fertility of virgin soils (Punkratov, 2006; Singh et al., 2016; Chittora et al., 2020), productivity of biologically complex of the World Ocean (Flornbaum et al., 2013; Hamilton et al., 2016; Fuchsman et al., 2019) are connected with nitrogen fixation activity of blue-green algae. Among Cyanophyta there are extremely harmful species which are the causative agents of “blooming” of water (Bhalla et al., 2009; Brookes & Carey, 2011; Carey et al., 2012; Cood et al., 2014; Rastogi et al., 2015). During blooming, the oxygen regime deteriorates sharply, the decomposing products of dead cells enter the water, living individuals emit toxic substances (Davidson et al., 2014; Rastogi et al., 2015; Walter et al., 2018; Kinnambo et al., 2019). Factors contributing to the development of "blooming" are high water temperature (+23…+32 °C), high concentration of nutrients (nitrogen, phosphorus, potassium), and lack of mixing of water masses (Cood et al., 2005; Posch et al., 2012; Backet et al., 2015; Zhang et al., 2016; Lürling et al., 2018; Kinnambo et al., 2019).

Cyanobacteria are known from a wide range of water bodies – from oligotrophic to eutrophic. Much attention has been focused on determining the role of cyanobacteria as indicators of the eutrophication process. Particular attention is paid to eutrophic and hypertrophic lakes, where cyanobacterial blooms are caused by potentially toxic, often invasive species (Nixdorf & Denke, 1997; Wiedner et al., 2002; Kokozski et al., 2010). A lot of interest in this problem is related to the negative effects of cyanobacterial bloom (Cood et al., 1999; Lopes & Vasconcelos, 2011). Cyanobacteria are a permanent element of the phytoplankton communities in lakes of low trophicity, where they usually have little importance in the community structure. However, some data show the dominance of the
cyanobacteria in oligotrophic lakes (Callieri & Stockner, 2000) or oligomesotrophic lakes (Napórowska-Krzebietke & Hutorovicz, 2013; Dadheech et al., 2014). In eutrophic lakes and water reservoirs, the domination of cyanobacteria in phytoplankton usually occurs in summer or autumn (Sommer et al., 1986). Strong eutrophic or hypertrophic waters show cyanobacterial domination regardless of season (Zohary & Breen, 1989; Wagner & Adrian, 2009; Beerendonck et al., 2015). Cyanobacteria species show different optimums with respect to environmental parameters such as temperature, light or nutrient content. Cyanobacteria are very well adapted to conditions of nutrient deficiency and limited availability of underwater light (Reynolds, 1984). The effect of nutrient concentrations on the development of blue-green algae is well documented, especially in terms of nitrogen (N), phosphorus (P) and the N to P ratio (Smith, 1983; Berman, 2001; Downing et al., 2001).

The concept of functional groups was proposed to describe the structure and dynamic of phytoplankton (Reynolds et al., 2002; Pudzhal et al., 2009). For phytoplankton, species from similar habitats and with similar sensitivity to environmental factors were classified in one functional group. The freshwater phytoplankton species were distributed among 31 functional groups (Costa et al., 2009). Phytoplankton functional groups have been used to study the impact of changes in the aquatic environment on the physiological, morphological and ecological characteristics of phytoplankton in different rivers, lakes and waters around the world (Becker et al., 2009, 2010; Krak et al., 2017; Rodrigues et al., 2018).

The application of the methods of classifying the functional groups of phytoplankton in aquatic ecosystems provides the key data reflecting dynamics of phytoplankton communities (Santana et al., 2017; Cuperino et al., 2019; Yao et al., 2020).

The north-western coast of the Sea of Azov is a set of natural systems that are characterized by a high level of biological diversity, economic value and have a high recreational potential (Maisheva et al., 2019; Yorkina et al., 2019). Harmonization of the goals of nature protection, optimization of economic efficiency and maintenance of recreational value is an extremely complex scientific and scientific-practical task. The development of a strategy for the rational use of nature should be based on the study of natural complexes that are within the protected areas as reference areas. In addition to the role of markers of the level of biological diversity, the objects of the nature reserve fund act as factors of functional stability of natural complexes in general. Therefore, the establishment of patterns of formation of Cyanoprokaryota communities in Pryazovsky National Nature Park is an important scientific problem. In this paper, we consider the hypothesis that ecological groups of cyanoprokaryotes, established by different criteria, are a source of information that can explain the structure of their communities depending on environmental conditions.

**Methods**

*Characteristics of experimental polygons.* The research was conducted within Pryazovsky National Natural Park. The park area is located in the coastal strip of the Sea of Azov and includes the aqual (coastal part of the Azov Sea, estuaries, bays, channel and floodplain of small rivers, their mouths, small artificial reservoirs) and terrestrial (coastal spits, the coast, the Azov Sea, estuaries, bays, channel and floodplain of small rivers, their mouths, small artificial reservoirs) landscape complexes (Vorovka, 2011). There were 9 experimental polygons in the territory of Pryazovsky National Natural Park which embraced steep areas or slopes, saline areas, coastal sandy soils, and water bodies (rivers, lakes, lirams, sea bays, lagoons).

**Polygon No 1:** Tashchenakovsky Pod (46°39'29" N 35°16'14" E) is an ornithological reserve of local significance. Area is 280 hectares. The research polygon is located at the mouth of the River Maly Utjug and occupies its right bank. The polygon includes a steppe slope, a salt marsh, and a brackish water body. The length of the sampling profile is 350 m.

**Polygon No 2:** the River Maly Utjug (46°23'30" N 35°12'42" E). It is a small steppe river that flows into the Utjug Estuary. The polygon is located near the village Shelyugi. The width of the river at the sampling sites is 30–35 m, depth up to 2 m. The banks of the river are overgrown with reeds. The profile covers only the water body.

**Polygon No 3:** Upper part of the Utjug Estuary (46°30'41" N 35°11'09" E). It is a complex natural monument of national importance, with an area of 280 hectares. The research polygon is located at the mouth of the River Maly Utjug and occupies its right bank. The polygon includes a steppe slope, a salt marsh, and a brackish water body. The length of the sampling profile is 350 m.

**Polygon No 4:** the Sivashik Estuary (46°24'28" N 35°06'09" E) is a landscape reserve of national importance with an area of 2800 hectares. It is a brackish water body which has a connection with the Sea of Azov and the coastline (steppe slopes which pass into saline meadows and salt marshes). In summer, the upper part of the estuary dries out and the lower part is covered with a self depositing salt layer. The salinity of the estuary ranges 17–28‰ and in the drying lagoons of the estuary to 120‰. The polygon is located on the right bank of the estuary. The sampling profile is 360 m and includes steppe slopes, salt marshes, water estuary.

**Polygon No 5:** Fedotova Spit (46°17'14" N 35°17'38" E) is a landscape reserve of national importance occupying 1910 hectares. The spit is represented by sand-seashell sediments of accumulative origin and stretches almost 30 km to Brinichy Island, the width of the spit ranges 360-800 m. The scientific polygon is located near the village of Stepok. The length of sampling profile is 540 m, from the Utjug Estuary to the sandy alluvial bars of the Azov Sea. The profile includes the water of the Utjug Estuary with salinity of 8–11‰, steppe vegetation and sand soils.

**Polygon No 6:** Stepanovska spit (46°27'06" N 35°28'33" E) is a landscape reserve of local significance, with an area of 200 hectares. The spit separates the Molochny Estuary from the Sea of Azov. The scientific polygon covers the waters of the Molochny Estuary (25–40‰), the salt marshes of the Molochny Estuary shore, and the sandy soils of the alluvial bar along the Sea of Azov. The length of the sampling profile is 340 m.

**Polygon No 7:** the Tubalsky Estuary (46°35'28" N 35°42'56" E). It is a flat depression at the mouth of the drying river that flows into the Sea of Azov. The scientific polygon is located near the right slope in an area of temporary salty ponds. Temporary water bodies are separated from the Sea of Azov by a sandy shore 200-250 m wide. The sampling profile is 350 m and includes steppe slopes, salt marshes, alluvial sandbars of the Azov Sea and saline water bodies with the mineralization of 34‰.

**Polygon No 8:** the mouth of the Korsak River (46°38'38" N 35°51'28" E). Is a flat part of the territory of the Korsak River floodplain. The floodplain is periodically flooded with the waters of the Azov Sea. In the surrounding areas a temporary shallow saline water body forms. The level of mineralization in these water bodies ranges from 14–40‰. The surrounding area is represented by saline vegetation. The sampling profile is 352 m from the right slope to the Azov Sea and includes a steppe slope, salt marshes, saline temporary lakes and a sandbar along the sea.

**Polygon No 9:** the floodplain of the Berda River (46°47'49" N 36°52'24" E) is a landscape reserve of national significance of 1417 hectares. It is a flat area with thickets of reeds and meadow-halophilic vegetation. The scientific polygon includes steppe slopes, saline meadows, salt marshes, and mineralized temporary ponds with salinity of 12–15‰. Sampling profile is 660 m.
Methods of collection and processing of soil algological samples. 124 algological individual and combined water and soil samples served as the study material. The individual samples were selected to determine macroscopic differences on the surface of the soil or in the aquatic environment. The sample was carried out in nine defined scientific polygons in the spring-winter period during the 2013–2019 exploratory route studies. The coordinates of each polygon were defined using the GPS (Garmin GPS 12 MAP) browser. The polygons were selected in several sequences for each biotope (steep slope, salt, sandy littoral zone, water body). The material was collected according to the generally accepted basic algorithm by a sterile method (Hollerbach & Sina, 1969). Soil was collected in paper envelopes that were previously heated in a drying cabinet at 130–150 degrees for an hour. The collection of the soil was performed using a spatula that was sterilized directly in the sites of the selection: the instrument was wiped in spirit and then plunged into the research soil repeatedly. The samples were processed fresh for 1–2 days from the moment of selection, and dried in a dark place to dry air condition and kept for further processing for several months.

Material and methods of collection and study of water-algological samples. The water samples were collected in temporary waters, salt lakes, estuaries and rivers. Mostly sampling was carried out in shallow water up to 1 m depth, as the studied water bodies did not have significant depths. Therefore, sampling was carried out by hydrobiological methods of sampling phytoplankton and phytobenthos. Also, in cases of intensive development of algae on the surface of the water body and in the benthos, individual samples were collected. Phytoplankton was selected by filtration of water through a planktonic mesh, 50–100 liters of water from the surface layer of 15 cm was poured through the planktonic mesh, after which the water sample was collected in labeled sterile glass containers. When collecting plankton of surface water layers, the planktonic mesh was lowered into the water so that the upper hole of the net was 5–10 cm above its surface.

Ecological and areological groups of Cyanoprokaryota. In relation to preferential environment, cyanoprokaryotes were divided into aerobions, aquatic organisms, amphibhions, and teratobions. Teratobions or terrestrial forms are in turn divided into edaphobions, aerophytes and subaerophytes (Vinigradova, 2012). Subaerophytes include aquatic and out of water forms of cyanophyceae, which are characterized by high ecological valence. Cyanophyceae form transitional ecological groups according to the preferential type of habitat, namely aerophytes, aqual, aqua-subaerophytes, aqua-terrestrial, eurybions, subaerophytes, terrestrial forms, terres-trial-subaerophytes forms. Amphibiont forms include aqua-subaerophytes, subaerophytes, and terrestrial-subaerophytes forms. Such species can exist in both aquatic and terrestrial environments. Aqual forms occur exclusive-ly in the aquatic environment. Terrestrial forms inhabit the soil and aerophytes which are adapted to exist in conditions of moisture deficiency (Vinigradova, 2012).

The following ecological groups among Cyanobacteria are selected in relation to the level of environmental salinity (Kornarek & Anagnostidis, 1999). Halotolerant is a species that exists in habitats with normal (0–30‰) or increased salt concentration. Halobiont is a species that exists in habitats with a high (31–60‰) or very high (70–330‰) salt concentration. Halophiles prefer habitats with a high concentration of salt. The marine species live on the coast of the sea and ocean. In relation to salt, marine species take a transitional position between halotolerants and halophiles (Vinigradova, 2012). According to the spatial coverage of species geographic ranges, cyanoprocariotes can be divided into the following groups: European moderate zone residents, European species, Eurasian species, Circumcontinental or Circumeconico and Cosmopolitan (Vinigradova, 2012).

The dependence between species and genera numbers in communities can be described by a linear model (Palczewski et al., 2008): log(Species) = b0 + b1 log(Gen) + e, where “Species” is the number of species, “Genera” is the number of genera, b0 and b1 are regression coefficients. The residuals of the regres-sion model e were considered to be the index of the taxonomic ratio species/genus.

Methods of ordination of communities. Multidimensional ordination techniques were used for analyzing spatial-temporal variation in the species composition of the invertebrate complex. The data matrix was subjected to Correspondence Analysis (CA), Detrended Correspondence Analysis (CCA) and Constrained Redundancy Analysis (RDA) in order to extract the most important patterns of community variation (Legendre et al., 2005). The environmental factors were fitted into the CA-ordination axes by means of the enfit function from the vegan library (Oksanen et al., 2018). The Detrended Correspondences Analysis (DCA) was used to distinguish whether the species response to environment factors is pre-dominantly monotonous or mostly unimodal. To do this, length of the first most important gradient of the community variation was assessed (Ter Braak & Prentice, 1988). If the gradient length exceeds more than 2 standard deviations, then Constrained Correspondence Analysis (CCA) should be chosen as the routine procedure. Otherwise, Constrained Re-dundancy Analysis (RDA) is more appropriate. For statistical calculations, we used appropriate procedures for Statistica (Version 12.0, StatSoft Inc., www.statsoft.com) or R (version 3.5.2; R Core Team, 2018).

Results

As a result of our research within the territory of Pryazazovsky National Natural Park, 124 species of Cyanoprokaryota were found which include 127 intraspecies taxa. The largest axis extracted after Detrended Correspondences Analysis equalled 6.0, indicating that the Constrained Correspondence Analysis procedure is the most suitable for the ordination of the species matrix. The first four canonical axes are able to explain 47.5% of the species matrix variability (Table 1). The canonical axis 1 explains 18.0% of variability of the species matrix and is most marked by the aquatic-subaerophytes (positive values) and eurybions (negative values, Fig. 2a). The positive axis values correspond to communities with the higher diversity, especially the species/genus ratio. This indicates the presence of a gradient of the salinity regimes where the most saline conditions correspond to the positive axis values, and negative ones are less saline. So it is only natural that there are water and salt ecosystems in the area of higher canonical axis 1 values, and there are steppe and sand ecosystems in the area of negative ones. The conditions for higher salinization are more favourable for species with a wide geographical range. The conditions with less or no salinization are more favourable for Cyanoprokariotes species with a European range. Canonical axis 2 explains 12.1% of the species matrix variability. This axis differentiates the aquatic ecosystems (positive axis values) from the others (negative axis values). The consequence of this is that aquatic species positively correlate with the canonical axis 2. Other species, except for aqual-subaerophytes, whose correlation coefficient statistically does not significantly differ from the null alternative, negatively correlate with the canonical axis 2. Water ecosystems are characterised by lower rates of diversity, but by a high species/species ratio. Ecological groups of cyanoprokariotes in terms of response to salinization do not form a clear pattern so one can assume that correlative relationships are focused on community relationships and not directly on external influences.

Analysis of the configuration of ecological factors and synecological indicators in the space of the first two canonical axes allows us to obtain a comprehensive picture of the ecological structure of the cyanoprokaryotic communities. Communities of aquatic ecosystems occupy the upper right quadrant of the plane that is defined by these two axes and are marked with aqual forms of Cyanophyceae. The counterclockwise movement from this quadrant sequentially moves us from aqua-subaerophytes forms to subaerophytic ones. The closest biotope marker to this area is salt marshes, but the corresponding zone is very wide, which indicates a significant floristic heterogeneity of salt marsh communities. The right side of the considered area corresponds to more saline stations, the left – less saline. Solonchak communities occupy the area from highly saline to moderately saline. Communities with a predominance of subaerophytes are characterized by higher species diversity and form a special kind of salt marsh flora. It is formed under conditions of moderate salinity and mainly by representatives of Eurasian species. It should be noted that saline conditions are most often inhabited by species with a wide geographical distribution, especially cosmopolitans.

The lower right quadrant is the zone of the greatest diversity of communities both in terms of the number of species and the number of genera.
These two indicators of diversity are characterized by consistent dynamics. The canonical Axis 3 explains 10.0% of the community variability. This axis distinguishes freshwater ecosystems (right side of the Fig. 2b) from the saline ecosystems (left side). Accordingly, steppe communities occupy the central part of the space, which is set by the canonical axes 3 and 4. It is natural that the markers of freshwater communities are the stenotopic halotolerants. These species also have mainly narrow ranges within the temperate zone of Europe. The canonical axis 4 explains 7.3% of the variability of a species matrix. The negative value of this axis denotes sand ecosystems. The results of the ordination indicate that communities in sandy soils tend to be represented by wide-ranging species.

**Table 1**

Correlation of external predictors and canonical axes

| Predictors | Canonical Axes, % explained inertia |
|------------|-------------------------------------|
|            | CCA1, 18.0% | CCA2, 12.1% | CCA3, 10.0% | CCA4, 7.3% |
| Ecotypic groups | | | | |
| Aq | 0.40 | 0.67 | 0.62 | 0.01 | 0.72 | 0.001 |
| AqSA | 0.89 | 0.01 | -0.33 | -0.30 | 0.41 | 0.001 |
| SA | 0.52 | -0.71 | -0.30 | 0.36 | 0.56 | 0.001 |
| TrSA | -0.72 | -0.46 | -0.46 | 0.23 | 0.59 | 0.001 |
| AqTr | -0.71 | -0.46 | -0.51 | -0.13 | 0.27 | 0.009 |
| Tr | -0.52 | -0.68 | -0.50 | -0.13 | 0.40 | 0.001 |
| Ae | -0.41 | -0.69 | -0.51 | 0.31 | 0.29 | 0.004 |
| Eu | -0.95 | -0.17 | 0.20 | -0.18 | 0.57 | 0.001 |

Diversity indexes

|           |               |
|-----------|---------------|
| Species | 0.48 | -0.80 | -0.25 | 0.27 | 0.22 | 0.032 |
| Genus | 0.10 | -0.99 | 0.00 | -0.06 | 0.26 | 0.012 |
| Resid | 0.71 | 0.48 | -0.25 | 0.45 | 0.59 | 0.001 |

Ecological groups in the salinity gradient

|           | H1 | H2 | H3 | H4 |
|-----------|----|----|----|----|
| Geographic range width | -0.75 | 0.34 | 0.53 | 0.21 | 0.78 | 0.001 |
| | 0.87 | -0.33 | -0.28 | -0.22 | 0.78 | 0.001 |
| | 0.97 | 0.22 | 0.01 | -0.06 | 0.82 | 0.001 |
| | 0.94 | -0.14 | -0.10 | 0.29 | 0.75 | 0.001 |

Geographic range width

|           |                  |
|-----------|------------------|
| Geographical groups | G1 | G2 | G3 | G4 | G5 |
| Biotopes | -0.26 | 0.70 | 0.59 | 0.30 | 0.11 | 0.262 |
| Sand | -0.93 | -0.25 | -25 | -0.06 | 0.62 | 0.001 |
| Solonchak | 0.72 | -0.51 | 0.02 | 0.47 | 0.47 | 0.001 |
| Steppe | -0.84 | 0.15 | 0.64 | -0.27 | 0.22 | 0.032 |
| Water | 0.79 | 0.34 | -0.29 | -0.42 | 0.35 | 0.003 |

Biocenotic traits (centroids coordinates)

|           | H1 | H2 | H3 | H4 |
|-----------|----|----|----|----|
| Location of cyanoprokaryotic communities in the space of canonical variables 1 and 2 (a) and 3 and 4 (b) and assessment of the impact of ecological variables on communities: Cyanoprokaryota species are presented by points; arrows show ecological variables: Aq – aqual; AqSA – aqual-subaerophytic; SA – subaerophytic; TrSA – terrestrial-subaerophytic; AqTr – aqual-terrestrial; Tr – terrestrial; Ae – aerophytic; Eu – eurytopic; Species – number of species; Genus – number of genus; Resid – depend ratio of species/genus; H1 – halotolerant (stenotopic); H2 – halotolerant (eurytopic); H3 – halobionts; H4 – halophiles; G1 – range within the Europe temperate zone; G2 – European; G3 – Eurasian; G4 – Circumcontinental or Circumoceanic; G5 – Cosmopolitans.

Inertia unexplained by the ecological predictors is characterized by $\chi^2$, which is equal to 1.353, which corresponds to 47.2% of the total inertia of the species matrix. Thus, the ecological predictors, which were chosen for analysis, well explain the variation of the Cyanoprokaryota community structure. The fractionation of the $\chi^2$ statistic showed that among the examined sources of variation, both the ecotopic structure of the community (explains $\chi^2 = 0.243$, or 8.5% of the total inertia) and the width of the range (explains $\chi^2 = 0.175$, or 6.1% of the total inertia) have the greatest independent importance to explain community structure. The independent importance of the adaptation to the salinity conditions and ecosystem type is slightly lower ($\chi^2 = 0.077$, or 2.7% of total inertia and $\chi^2 = 0.041$, or 1.5% of total inertia, respectively). The interaction between the sources of variation is important in the variation of the structure of community. The largest role in variation of the community structure is played by the interaction between the ecotopic structure and width of species ranges ($\chi^2 = 0.395$, or 13.8% of total inertia) and the triple interaction between the ecotopic structure of the community, the width of species ranges and ecosystem type ($\chi^2 = 0.266$, or 9.3% of total inertia).

**Discussion**

The main challenge for ecology is to identify the determinants of the structure of environmental communities (Gilbert & Bennett, 2010). Our study covers a wide range of ecosystem types – from steppe to aquatic ecosystems. The Detrended Correspondence Analysis indicates that the species of which the Cyanoprokaryota community consists are represented within significant environmental gradients in such a way that the form of the function of their response to the impact of environmental factors is mostly unimodal. For this type of data, Canonical Correspondence Analysis is most appropriate. A list of community ecological characteristics calculated on the basis of the autecological parameters of the
Cyanoprokaryota species was used for the explanatory variables. This approach allows us to test the hypothesis that the autoecological parameters of the Cyanoprokaryota species are important for the formation of the community structure. In our study, it was shown that the axes extracted as a result of the ordination procedure, which indicate the coordinated dynamics of species, correlate with both synecological characteristics, such as diversity indicators (species and genus diversity and detrended species/genus ratio), and with autoecological characteristics, such as cyanoprokaryotic ecotypes in terms of habitat condition or salinity adaptation. Other studies have shown that within the region, diversity in the number of species and number of genera is characterized by a high level of correlation (Andersen, 1995). Genus diversity can be used as a surrogate for species diversity (Baldi, 2003) and species composition (Alves et al., 2016). There is an opinion that the use of the lower taxonomic resolution in taxonomic determination of plankton in combination with ecological classification can be used as a strategy to minimize difficulties in plankton identification (Machado et al., 2015). Our results suggest that only for part of the cyanoprokaryotic community can the genus level be a good approximation of species diversity.

That the number of species is an increasing function of the area is one of the most significant generalizations of the ecology (Arrhenius, 1921; Gleason, 1922; Fisher et al., 1943). No clear trends in the number of species and genera with geographical latitude have been found for the flora of North America, but the residuals of this model show a clear dependence on latitude (Palmer et al., 2008). Also an important result is that the direction corresponding to the variation of diversity, which is indicated by the number of species and genera, is orthogonal to the direction denoted by the detrended species/genus ratio. Orthogonality indicates the independence of variables which also indicates that the detrended species/genus ratio has specific information. The intensity of interspecies competition is regarded as a possible ecological interpretation of the species/genus ratio. A higher value of the ratio indicates a saturation of the community of taxonomically similar species, between which the intensity of interspecific competition is higher than between more taxonomically distant species (Enquist et al., 2002; Webb et al., 2002). According to such interpretation, the ecosystems with a high level of salinity (water or solonchak) are able to carry the largest number of species that can actively compete. It should be noted that cyanoprokaryote species in these ecosystem types are represented mainly by aqua-subaerophile forms with a wide (cosmopolitan) range. In turn, there is a competitive exclusion of species that is labeled with a lower detrended species/genus ratio in the steppe or sandy ecosystems that are subjected to less salinity. These Cyanoprokaryota communities are represented by terrestrial or eurytopic species, the range of which predominantly covers Europe.

The key goal of experimental biology is to quantify the relationship between environment predictors and ecosystem response (Sokolov & Zhukov, 2017). A statistical analysis that allows one to achieve this goal is called variation partitioning (Borcard et al., 1992; Legendre et al., 2005; Fischer, 2019). Our results confirm the importance of cyanoprokaryotic adaptations for living in certain environments as a community structuring factor. It is also worth noting the structuring role of species response types in the gradient of salinization conditions, as well as the width of species ranges. It is important not only to recognize the impact itself, but also to quantify it in comparison with other sources of variation. This task can be solved by variance partitioning (Borcard & Legendre, 2002; Sattler et al., 2010; Shi et al., 2015; Tonkin et al., 2016; Astor et al., 2017). The variation partitioning procedure is used to weight balance between importance of the neutral and niche-dependent factors for community structuring (Smith & Lundholm, 2010; Yorke et al., 2018). The variation partitioning of a community is aimed at studying the beta diversity. The beta diversity characterizes variability in composition of species between sites in a geographic region (Zhukov et al., 2018). Beta diversity is a key concept for understanding the functioning of ecosystems, their protection and management (Legendre, 2007). The use of the redundancy analysis of and variation partitioning in communities of aquatic macrophytes allows one to estimate the relative importance of the local environment variables, climatic factors and spatial position (Alahuhina et al., 2018).

Analysis of the species list of a respective territory allows one to establish the ecological profile of its flora (Vinigradova, 2012). The basis of solonchak flora was shown to be formed by the species with a high adaptive potential such as amphibient and terrestrial forms (Vinigradova, 2006). A significant information importance of the structure of the ecotopic groups was revealed to explain variations of species composition at the community level. The considerable adaptation potential of species contributes to the variability in species composition of cyanoprokaryotic communities (specia particular ecosystem type). The diversity of conditions within ecosystems forms a wide variety of environmental regimes, according to which the species composition of the community changes. But the degree of freedom of such variability is restricted by the adaptation models, which can be specified by the spectra of ecotopic groups. This explains that the role of ecotypic diversity in the determination of species composition is more important than that of ecosystem types. Such variability of the Cyanoprokaryota communities allows us to consider this taxonomic group as a source of independent information, which does not repeat the bioindicative information from other sources.

The type of ecosystems is determined based on the material “macroscopic” parameters, such as relief, soil conditions, and vegetation structure represented by higher plants. The dynamics of the Cyanoprokaryota communities take place at another spatial and time level of ecosystems. Thus, the functional-hierarchical levels of organization of superorganismic systems form relatively independent space-time ecosystems, which differ in their spatial and temporal aspects of dynamics (Pakhrzhevskii, 1996). The measure of temporal dynamics is ecological time (the duration of existence in units of astronomical time of one stage of succession dynamics) and time of complete biological cycle (time during which the flow of nutrients passing through the ecosystem becomes equal to their mass in ecosystem biomass) (Gongalsky, 2014). In terrestrial soil ecosystems there are bacterial-alga-protozoan (ecological time – from 1 week, size – up to several centimeters), fungal-microarthropodic (ecological time – from 1 week to 1 year, size is commensurate with the rhizosphere of one plant) and lumbicide-plant (ecological time – from 1 month to decades, size within the biogeocenosis) (Pakhrzhevskii, 1996). Similar spatio-temporal structuring can be performed for other living environments – aquatic, terrestrial and amphibiotic. Thus, according to the dimensional features, the spatio-temporal dynamics of cyanoprokaryotic communities causes their significant diversity within a particular type of ecosystem. Such diversity is the basis for obtaining specific information about the course of processes at the level of a particular ecosystem.

As sources of variation in the species composition of Cyanoprokaryota communities, we consider indicators that differ significantly in their nature: Ecotope groups and groups that characterize the preferences of species for the salinity conditions of ecotopes characterize the autoecological strategies of species for adaptation to the conditions of existence. It is important to note that ecotopic groups that indicate preference for a particular habitat correlate with the species composition of communities. Thus, the correlation of ecotopic groups in a community is a characteristic that reveals the features of the community as a whole. This makes it possible to consider ecotypic spectra as a synecological characteristic of a Cyanoprokaryota community.

The important result is that with the width of geographic range of Cyanoprokaryota species, as quantitative indicators, correlate certain features of the structure of species ensembles. The problem of geographical range of species is key in modern integrative biology (Bozinovic & Naya, 2015). There is a consensus in the idea that the boundaries of the distribution range of species are the consequence of complex relationships between the species specific physiological, phenological and ecological characteristics, as well as the ability to dispersion and environmental inter-actions between species (Bozinovic et al., 2011). A number of climate hypotheses were advanced to explain the variability in species ranges (Päther, 2003). The key idea is that variability of physiological properties along geographical gradients affects species tolerance (Sperber & Gaston, 2009). Thus, the correlation of causes that affect the position of a species in a community and its geographical distribution is the reason why the community structure depends on the biogeographical features of the species that constitute it. Dispersion, geographical isolation, range limitation or expansion in the past, drifting processes, the originator's effect and the role of selection all leave their traces in the appearance and features of the geographical distribution of the modern cyanobacteria flora (van
Grenberg et al., 2011). There is a standpoint that there are no limitations to the geographical distribution of bacteria (Foissner, 2006). The lack of spatial structure in bacterial communities is confirmed by molecular data for soil (Fierer & Jackson, 2006), marine (Mullins et al., 1995) and freshwater bacteria (Glodek et al., 2000; Van Der Gucht et al., 2007), including cyanobacteria (Junghalt et al., 2010). However, several studies have reported a clear spatial structure for other prokaryotes, including marine (Zoioglithra et al., 2008), soil (Cho & Tiedje, 2000) and freshwater bacteria (Pearson et al., 2009). For prokaryotes occurring under extreme conditions, such as hot volcanic vents or deep hydrothermal vents, the phytogeographic structure indicates effects of strong geographical isolation and limited distribution (Whitaker et al., 2003; Papke et al., 2003; Miller et al., 2007), although not all thermophilic cyanobacteria show a clear spatial structure (Ionescu et al., 2010). For more widely distributed bacteria, biogeographic patterns may be the result of historical (e.g., limiting distribution) and/or ecological processes (e.g., local selection) (Whitaker, 2006; Ramette & Tiedje, 2007). The relative importance of these processes in the structuring of microbial systems has so far been little studied (Martiny et al., 2006). Apart from cosmopolitan species, taxa with more restricted distribution also exist among Cyanoprocrayotes. Many of them have a Holarctic or pantropical distribution. In addition to the distribution of ecological niches, temperature is one of the main control factors limiting the spread of Cyanoprocrayotes species in certain latitudes. In addition, the presence of species with regional distribution (endemics) cannot be excluded, which points to the need to take into account other factors affecting the geographical distribution of Cyanoprocrayotes (Hoffmann, 1996). A relationship has been found between geographical distribution and ecological properties of Cyanoprocrayotes species. Thus, for Cyanoprocrayotes of the Black Sea coast of Ukraine the proportion of cosmopolitans is higher among halobionts, marine and saltwater species. In turn, the number of European species and species of temperate latitudes is greater among aerophytes (Vinogradova & Bryantseva, 2017). The results show that not only among the total list of regional flora species there is an unequal ratio of representatives with different range width depending on the position of the species in the gradient of salinity conditions. It is shown that the composition of species of a particular community is due to the ecological properties of species, which also affect the overall geographical distribution of the species. The presence of ecological determinants which influence the geographical distribution as well as the success of a species in a particular community is confirmed by the statistically significant influence of the interaction of species range width and other factors of community variation, namely ecotopic specialization, preference of salinity conditions and habitat type. Thus, biogeographic characteristics of species, which are traditionally considered as flora characteristics within a large area, are an indicator that plays an important role in forming the structure of a particular plant community. This role is conditioned by the fact that the geographical distribution of a species depends on the ontological characteristics of the species, which also determine the success of the species in communities within the range.

**Conclusion**

Cyanoprocrayotes are represented by various communities in aquatic, terrestrial and amphibiotic ecosystems of the investigated area. The variation in species composition in Cyanoprocrayotes communities is driven by the peculiarities of ecological regimes of the environment and is conditioned by the diversity of ecosystem types. Variation of the Cyanoprocrayota species structure correlates with changes in the structure of ecotopic groups, ecological groups in the gradient of salinity conditions of ecotopes and with the width of species ranges. The communities of aquatic and terrestrial ecosystems are the most distinct among themselves. Aquatic ecosystems are clearly distinguished by lower diversity, but high species/genus ratios. Aquatic communities are typically characterized by the dominance of aquatic and aquatic-subaerophyte species. Terrestrial and amphibious ecosystems are known to be characterized by higher prevalence of eurybionts. Solonchak ecosystems, as an amphibious habitat, are characterized by a significant diversity of conditions, so their Cyanoprocrayote communities are significantly diverse. Saline conditions are most often inhabited by species with wide geographical distribution, especially cosmopolitans. Specific conditions for Cyanoprocrayote life are formed in sandy soils. Communities in sandy soils tend to be represented by wide range species.

**References**

Alahauté, J., Lindholm, M., Vivo, C. P., Chaguais, E., Clayton, I., de Viento, M., Feldmann, T., Ecke, F., Gacia, E., Grillas, P., Hoyers, M. V., Johnson, L. B., Kolada, A., Kosten, S., Lauridsen, T., Luikens, B. A., Mijdele, M., Momdal, R. P., Rhazi, L., Rhazi, M., Sass, L., Sonderegger, M., Xu, J., & Heinso, J. (2018). Global patterns in the metacommunity structuring of lake macrophytes: Regional variations and driving factors. Oecologia, 188(4), 1167–1182. Alves, C., Vieira, C., Almeida, R., & Hespanhol, H. (2016). Genera as surrogates of bryophyte species richness and composition. Ecological Indicators, 63, 82–88. Andersen, A. N. (1955). Measuring more of biodiversity: Gnera richness as a surrogate for species richness in Australian ant faunas. Biological Conservation, 73(1), 39–43. Armeanu, O. (1921). Species and area. The Journal of Ecology, 91(1), 95. Astor, T., von Proschwitz, T., Stranghorn, J., Berg, M. P., & Bengtsson, J. (2017). Importance of environmental and spatial components for species and trait composition in terrestrial snail communities. Journal of Biogeography, 44(6), 1362–1372. Becker, L. J., Manussan-Baptiste, D., LePeull, R., & Bolton, B. (2015). Cyanobacteria and algae blooms: Review of health and environmental data from the harmful algal bloom-related illness surveillance system (HABISS) 2007–2011. Toxins, 7(4), 1088–1064. Badi, H., Monsieurs, P., Conino, I., Wattiez, R., & Loys, N. (2015). Molecular investigation of the radiation resistance of edible cyanobacteria: Anabaena sp. PCC 8005. Microbiology Open, 4(2), 187–207. Bládi, A. (2003). Using higher taxa as surrogates of species richness: A study based on 3700 Coleoptera, Diptera, and Acari species in Central-Hungarian reserves. Basic and Applied Ecology, 4(6), 589–593. Becker, V., Caputo, L., Ordóñez, J., Maró, Á., Arnegild, J., Crosseti, L. O., & Huszar, V. L. M. (2010). Driving factors of the phytoplankton functional groups in a deep Mediterranean reservoir. Water Research, 44(11), 3345–3354. Becker, V., Huszar, V. L. M., & Crossetti, L. O. (2009). Responses of phytoplankton functional groups to the mixing regime in a deep sub-tropical reservoir: Hydrobiologia, 628(1), 137–151. Berkman, T. (2001). The role of DON and the effect of N-P ratios on occurrence of cyanobacterial blooms: Implications from the outgrowth of Aphanothece limnetica in Lake Kinneret. Limnology and Oceanography, 46(2), 443–447. Bevesendorf, L. J., Miller, T. R., & McManus, K. D. (2015). Long-term monitoring reveals carbon-nitrogen metabolism key to microcystin production in eutrophic lakes. Frontiers in Microbiology, 6, 456. Bláhová, L., Babica, P., & Martášek, B. (2009). Toxins produced in cyanobacterial water blooms – toxicity and risks. Interdisciplinary Toxicology, 2(2), 36–41. Borcard, D., & Legendre, P. (2002). All-scale spatial analysis of ecological data by means of principal coordinates of neighbour matrices. Ecological Modelling, 153, 51–68. Borcard, D., Legendre, P., & Drapeau, P. (1992). Partiaulling out the spatial component of ecological variation. Ecology, 73(3), 1045–1055. Bothe, H., Schmitz, O., Yates, M. G., & Newton, W. E. (2010). Nitrogen fixation and hydrogen metabolism in Cyanobacteria. Microbiology and Molecular Biology Reviews, 74(4), 529–551. Boziofific, F., & Yaye, D. E. (2015). Linking physiology, climate, and species distributional ranges. In: Integrative organismal biology. Wiley Blackwell. Pp. 277–290. Boziofific, F., Calosi, P., & Spicer, J. J. (2011). Physiological correlates of geographic range in animals. Annual Review of Ecology, Evolution, and Systematics, 42(1), 155–179. Brookes, J. D., & Carey, C. C. (2011). Ecology: Resilience to blooms. Science, 334(6065), 46–47. Callaini, C., & Stockner, J. (2000). Phycocyanobacteria success in oligohaline lakes: Fact or fiction? Journal of Limnology, 59(1), 72–76. Carey, C., Belinga, B. W., Hoffmanc, E. P., Hamilton, D. P., & Brookes, J. D. (2012). Eco-physiological adaptations that favour freshwater cyanobacteria in a changing climate. Water Research, 46(5), 1394–1407. Červený, J., Sinetova, M. A., Zavřel, T., & Los, D. A. (2015). Mechanisms of high temperature resistance of Synechocystis sp. PCC 6803: An impact of histidine kinase 34. Life, 5(1), 676–699. Chittor, D., Moina, M., Barup, T., & Swarn, P. (2020). Cyanobacteria as a source of biofertilizers for sustainable agriculture. Biochemistry and Biopysics Reports, 22, 100737. Cho, J. C., & Tiedje, J. M. (2000). Biogeography and degree of endemicity of fluorescent Pseudomonas strains in soil. Applied and Environmental Microbiology, 66(12), 5448–5456.
Jiang, Y. L., Wang, X. P., Sun, H., Han, S. J., Li, W. F., Cui, N., Lin, G. M., Zhang, Hollerbach, M. M., & Stina, E. A. (1969). Soil algae. Science, Leningrad.

Glockner, F. O., Zaichikov, E., Belkova, N., Denissova, L., Pernthaler, J., Pernthaler, Gleason, H. A. (1922). On the relation between species and area. Ecology, 3(2), 272–272.

Hamilton, T. L., Bryant, D. A., & Macalady, J. L. (2016). The role of biology in plankton blooms in freshwater ecosystems and their link with hydro-meteorological and environmental variations in Tanzania. Hydrology, 5(3), 63132.

Katoh, R., Oyama, K., Kawanishi, T., Kitaizumi, S., Yasasaka, K., Sagara, N., Fujimoto, M., & Tenouchi, K. (2019). Pressure accelerates the cica- clid clock of cyanobacteria. Scientific Reports, 9(1), 1–8.

Kolociniski, M., Stefanik, K., Munkucziovecz-Boczok, J., Lydorczyk, D., & Szinnien, J. (2010). The ecology of the invasive cyanobacteria Cylindrospermopsis raciborskii (Nostocales, Cyanophyta) in two hypertrophic lakes dominated by Planktothrix agardhii (Oscillatoriales, Cyanophyceae). European Journal of Phycology, 46(1), 357–354.

Komarké, J., & Aragostidis, K. (1999). Cyanoprokaryota. I. Chlorococcales. Spektrum, Akademischer Verlag, Heidelberg, Berlin.

Kraus, M. P. (1969). Resistance of blue-green algae to Co gamma radiation. Radiation Botany, 9(6), 481–489.

Kruk, C., Devercelli, M., Huszar, V. L., M., Hernández, E., Beamud, G., Diaz, M., Silva, L. H. S., & Segura, A. M. (2017). Classification of Reynolds plankton functional groups using individual traits and machine learning techniques. Freshwater Biology, 62(10), 1661–1692.

Legendre, P. (2007). Studying beta diversity: Ecological variation partitioning by multiple regression and canonical analysis. Chinese Journal of Plant Ecology, 31(5), 976–981.

Legendre, P., Borceaud, E., & Peres-Neto, P. R. (2005). Analyzing beta diversity: Partitioning the spatial variation of community composition data. Ecological Monographs, 75(4), 435–451.

Liu, X., Hou, W., Dong, H., Wang, S., Jiang, H., Wu, G., Yang, J., & Li, G. (2016). Distribution and diversity of cyanobacteria and eukaryotic algae in Qingshui–Tibetan Lakes. Geomicrobiology Journal, 33(10), 808–809.

Lopes, V. R., & Vasconcelos, V. M. (2011). Planktonic and benthic cyanobacteria of European brackish waters: A perspective on estuaries and brackish seas. European Journal of Phycology, 46(3), 292–304.

Lürling, M., de Mello, M. M., van Oosterhout, F., de Senerpont Domis, L., & Marmiroi, M. M. (2018). Response of natural cyanobacteria and algae assemblages to a nutrient pulse and elevated temperature. Frontiers in Microbiology, 9, 1951.

Machado, K. B., Borges, P. P., Cameiro, F. M., de Santana, J. F., Vieira, L. C. G. de, Moraes Huszar, V. L., & Nabot, J. C. (2015). Using lower taxonomic resolution and ecological approaches as a surrogate for plankton species. Hydrobiologia, 743(1), 255–267.

Maloše, L., Yanova, T., Ambadzhi-Tepeniko, L., Pasenko, O., Yakovchuk, O., Zinenko, A., & Muravkova, M. (2019). Innovative technologies for ensuring ecological safety of maritime recreation. In: Eckert, M., Nestorenko, O., & Szynek, M. (Eds.). Modern innovative and information technologies in the development of society. Wydawnictwo Wyższej Szkoły Technicznej w Katowicach, Katowice, Pp. 29–38.

Marty, J. B. H., Boharman, J. B. M., Brown, J. H., Colwell, R. K., Farmer, J. A., Green, J. L., Homer-Denve, M. C., Kane, M., Kramins, A. J., Kusce, C. R., Morin, P. J., Nacem, O., Orlovs, L., Resibush, A. L., Smith, V. H., & Staley, J. T. (2006). Microbial biofilm: Putting microorganisms on the map. Nature Reviews Microbiology, 4(2), 102–112.

Miller, S. R., Castenholz, R. W., & Pedersen, D. (2007). Phylogeny of the thermophilic cyanobacterium Mastigocladus laminosus. Applied and Environmental Microbiology, 66(11), 4055–4055.

Gongalsky, K. B. (2014). Wildfires and soil fauna. KMK Scientific Press Ltd, Moscow.

Hamilton, T. L., Bryant, D., A., & Macalady, J. L. (2016). The role of biology in planetary evolution: Cyanobacterial primary production in low-oxygen Proterozoic oceans. Environmental Microbiology, 18(2), 325–340.

Hoffmann, L. (1999). Marine cyanobacteria in tropical regions: Diversity and ecology. European journal of Phycology, 34(1), 1–12.

Hoffmann, L. (1999). Geographical distribution of freshwater blue-green algae. Hydrobiologia, 336, 33–40.

Hollerbach, M. M., & Stina, E. A. (1969). Soil algae. Science, Leningrad.

Houe, N., Tiaa, Y., Emi, T., Yamane, Y., Kishino, Y., Kohke, H., & Sachet, K. (2001). The relationship between the growth temperature and the high-temperature stress on photosystem II and plasma membranes in a mesophilic cyanobacterium, Synechocystis sp. PCC6803. Plant and Cell Physiology, 42(10), 1140–1148.

Ikonen, L., Lindhjem, M., Malkavi, H., & Oren, A. (2010). Biogeography of thermophilic cyanobacteria: Insights from the Zenka Mai hot springs (Jordan). FEMS Microbiology Ecology, 72(1), 103–112.

Jiang, Y., Li, W., Zhang, Z., Cui, N., Lin, G. M., Zhang, J. Y., Cheng, W., Cao, D. D., Zhang, Z. Y., Zhang, C. C., Chen, Y., & Zhou, C. Z. (2017). Coordinating carbon and nitrogen metabolic signaling through the cyanobacterial global repressor NtrB. Proceedings of the National Academy of Sciences of the United States of America, 115(2), 403–408.

Jungblut, A. D., Lovejoy, C., & Vincent, W. F. (2000). Global distribution of cyanobacterial ecotypes in the cold biosphere. ISME Journal, 4(2), 191–202.

Kimambo, O. N., Gumbo, J. R., & Chikoore, H. (2019). The occurrence of cyanobacteria blooms in freshwater ecosystems and their link with hydro-meteorological and environmental variations in Tanzania. Hydrology, 5(3), 60312.

Kotara, R., Oyama, K., Kawanishi, T., Kitaizumi, S., Yasasaka, K., Sagara, N., Fujimoto, M., & Tenouchi, K. (2019). Pressure accelerates the cica- clid clock of cyanobacteria. Scientific Reports, 9(1), 1–8.

Kolociniski, M., Stefanik, K., Munkucziovecz-Boczok, J., Lydorczyk, D., & Szinnien, J. (2010). The ecology of the invasive cyanobacteria Cylindrospermopsis raciborskii (Nostocales, Cyanophyta) in two hypertrophic lakes dominated by Planktothrix agardhii (Oscillatoriales, Cyanophyceae). European Journal of Phycology, 46(4), 357–354.

Kotara, R., Oyama, K., Kawanishi, T., Kitaizumi, S., Yasasaka, K., Sagara, N., Fujimoto, M., & Tenouchi, K. (2019). Pressure accelerates the cica- clid clock of cyanobacteria. Scientific Reports, 9(1), 1–8.

Kolociniski, M., Stefanik, K., Munkucziovecz-Boczok, J., Lydorczyk, D., & Szinnien, J. (2010). The ecology of the invasive cyanobacteria Cylindrospermopsis raciborskii (Nostocales, Cyanophyta) in two hypertrophic lakes dominated by Planktothrix agardhii (Oscillatoriales, Cyanophyceae). European Journal of Phycology, 46(4), 357–354.

Kotara, R., Oyama, K., Kawanishi, T., Kitaizumi, S., Yasasaka, K., Sagara, N., Fujimoto, M., & Tenouchi, K. (2019). Pressure accelerates the cica- clid clock of cyanobacteria. Scientific Reports, 9(1), 1–8.

Kolociniski, M., Stefanik, K., Munkucziovecz-Boczok, J., Lydorczyk, D., & Szinnien, J. (2010). The ecology of the invasive cyanobacteria Cylindrospermopsis raciborskii (Nostocales, Cyanophyta) in two hypertrophic lakes dominated by Planktothrix agardhii (Oscillatoriales, Cyanophyceae). European Journal of Phycology, 46(4), 357–354.

Kotara, R., Oyama, K., Kawanishi, T., Kitaizumi, S., Yasasaka, K., Sagara, N., Fujimoto, M., & Tenouchi, K. (2019). Pressure accelerates the cica- clid clock of cyanobacteria. Scientific Reports, 9(1), 1–8.
