The Sulwon Award for Scientific Achievement is the Korean Diabetes Association's highest scientific award and honors an individual who has excellently contributed to the progress in the field of diabetes and metabolism. Sulwon Award is named after an emeritus professor Eung Jin Kim, who founded Korean Diabetes Association.

Prof. Sung-Woo Park received the third Sulwon Award at 2012 International Conference on Diabetes and Metabolism, November 8-10, 2012 at Seoul, Korea.

INTRODUCTION

Cholesterol is an essential structural component for cell membrane integrity, permeability and fluidity. In mammals, it also serves the precursor for the synthesis of bile acids, vitamin D and steroid hormones. Furthermore, cholesterol plays as a critical signaling molecule. Therefore, dysfunctional regulation of cholesterol and its related molecules affect human health. Elevated blood cholesterol level is a major risk factor for atherosclerotic coronary heart disease, leading cause of death in developed countries [1,2].

Cholesterol homeostasis in the body is tightly balanced by de novo biosynthesis, intestinal absorption, and biliary and fecal excretion. Compared to well-defined process of cholesterol biosynthesis, the mechanisms for intestinal cholesterol absorption and excretion were poorly defined [3,4]. In 2004, Altman et al. [5] find that Niemann-Pick C1-Like 1 (NPC1L1) plays an important role in intestinal cholesterol absorption. Ezetimibe, first pharmacological inhibitor of cholesterol absorption has been shown to target NPC1L1 [5,6]. Recently, NPC1L1 has been implicated in hepatitis C virus (HCV) entry [7]. From clinical trials and animal studies, there are accumulated data showing that NPC1L1 and NPC1L1 associated cholesterol metabolism influence metabolic syndrome such as nonalcoholic fatty liver disease (NAFLD), diabetes, obesity, and atherosclerotic coronary heart disease. Here, I discuss NPC1L1, NPC1L1-dependent intestinal and hepatic cholesterol uptake and its associated metabolic disease.

DISCOVERY AND CHARACTERIZATION

NPC1L1 was first identified as a homolog of Niemann-Pick C1 (NPC1), a gene which defect causes inherited lipid storage disorder Niemann-Pick disease type C1 [8]. Like its ho-
mologue, NPC1L1 is a polytopic transmembrane protein consisting of 13 transmembrane domains, N-terminal domain (NTD) and N-linked glycosylation sites [9]. Five of 13 membrane domains consist of sterol sensing domain (SSD). Conserved SSD is also found in several other transmembrane proteins, all of which are involved in cholesterol metabolism. These proteins include NPC1, 3-hydroxy-3-methylglutaryl CoA reductase (HMG-CoA reductase), the rate-limiting enzyme in cholesterol biosynthesis, sterol regulatory element binding protein (SREBP)-cleavage activating protein, a protein that regulates transport and proteolytical activation of SREBPs which controls sterol and other lipid biosynthesis, and patched, 12-pass transmembrane protein receptor for cholesterol linked signaling peptide hedgehog [10,11]. Sterol binding pocket is localized in crystal structure of NTD of NPC1L1. NTD of NPC1L1 directly binds to cholesterol [12], which leads to confirmation change and cholesterol entry [13]. Extensive N-glycosylation sites consist of three extracellular/luminal loops of NPC1L1. As posttranslational modification, N-glycosylation affects maturation and function of NPC1L1 by folding, secretion and endoplasmic reticulum (ER) retention [14].

It has been demonstrated in several studies that NPC1L1-dependent cholesterol transport may be regulated by clathrin-mediated endocytosis [15-17]. At steady state, NPC1L1 proteins are mainly found in endocytic recycling compartment (ERC). When cholesterol is depleted, NPC1L1 proteins move from ERC to plasma membrane (PM) [15]. On cholesterol repletion, cholesterol is sensed by PM transported NPC1L1 [15] and incorporated into PM by the formation of NPC1L1-flotillin-cholesterol membrane microdomains [16]. Subsequently, this formation is internalized by clathrin/AP2 mediated endocytosis. The vesicles are then moved to ERC [16]. Excessive cholesterol could be transported into cells in this NPC1L1 dependent manner.

NPC1L1 is widely expressed in many human tissues but highly expressed in the liver and small intestine [5,18,19]. According to species, distribution and pattern of NPC1L1 expression are different. Mouse and rat NPC1L1 are more abundant in small intestine than liver [5,19]. The reasons for different patterns of NPC1L1 expression among species remain elusive.

**TRANScriptionAL REGULATION OF NPC1L1**

Cholesterol transporter, NPC1L1 is reduced by cholesterol feeding and increased by NPC1L1 inhibitor, ezetimibe in animal models [20,21]. Several transcription factors involved in cholesterol metabolism are suggested as regulatory factor for NPC1L1 expression. SREBP2, a transcription factor for cholesterol biosynthesis shows positive relationship with mRNA expression of NPC1L1 in human hepatoma HepG2 cells and intestinal Caco2 cells [22-24]. SREBP2 together with hepatocyte nuclear factor 4α synergistically activates human NPC1L1 promoter [24]. In vivo and in vitro studies demonstrate the regulatory effects of nuclear receptors including liver X receptor (LXR), retinoid X receptor, and peroxisome proliferator-activated receptors (PPARs) on NPC1L1 transcription. PPARα agonist, fenofibrate administered mice remarkably decrease intestinal cholesterol absorption accompanied with the reduction in NPC1L1 mRNA expression [25]. PPARγ agonist also decreases mRNA level of NPC1L1 in small intestine and increases fecal sterol excretion. A single dose of LXR agonist mice and treatment of LXR activators, GW39665 and T0901317 in the human enterocyte cell line reduce mRNA expression of NPC1L1 [26]. However, the effects of nuclear receptors on NPC1L1 transcription are discrepant according to tissue and species.

**INTESTINAL NPC1L1**

NPC1L1 is abundantly expressed in the jejunum and proximal ileum and specifically located to the brush border membrane of intestinal enterocytes where cholesterol absorption takes place [5,27]. NPC1L1 deficient mice remarkably decrease intestinal cholesterol absorption by 70% [5] and attenuate diet induced hypercholesterolemia [28]. However, the genetic ablation of NPC1L1 doesn’t change triglyceride uptake by intestine [5].

As mentioned above, NPC1L1 is tightly regulated by intracellular itinerary. Cholesterol is transported into cells with NPC1L1 through clathrin/AP2-mediated endocytosis [15,16]. Delivered cholesterol to ER is packaged into cholesterol ester by acyl coenzyme A: cholesterol acyltransferase 2 (ACAT2), which in turn incorporated into chylomicrons for lymphatic secretion. Several studies demonstrate that NPC1L1 has substrate specificity on sterol absorption. In NPC1L1 deficient mice [5,19,20], ezetimibe treated mice [29] and patients with cholesterolemia or sitosterolemia [30,31], sterols are significantly decreased. In cell culture system, phytosterols show the decrease of NPC1L1 internalization and uptake compared to cholesterol [16,17]. This illustrates that phytosterol uptake is NPC1L1 dependent but NPC1L1 has high affinity with cho-
esterol compared to phytosterols.

**HEPATIC NPC1L1**

Hepatic NPC1L1 expression differs among mammalian species. Human and rat NPC1L1 are abundantly expressed in the liver, but hepatic NPC1L1 expression is undetectable in mouse liver [5,19,32]. Still the reasons for different tissue patterns of NPC1L1 expression and abundant hepatic NPC1L1 expression in humans are not fully determined.

Liver is the main site for cholesterol homeostasis in the body. In the liver, cholesterol de novo biosynthesis by HMG-CoA reductase, taking up lipoprotein cholesterol via low density lipoprotein receptors (LDLr) for regulating blood cholesterol levels, lipoprotein release, storage by esterification, degradation, and conversion into bile acid for cholesterol disposal and regulation of intestinal lipid absorption occur [3,4]. Compared to intestinal NPC1L1, still the functions of hepatic NPC1L1 in human remain largely unknown. To investigate the function of hepatic NPC1L1, mice expressing human NPC1L1 in the liver were generated. In these transgenic mice, human NPC1L1 localizes to the canalicular membrane of the hepatocytes as found in humans [15,32]. There are significant reduction of cholesterol in bile acid and elevation of liver cholesterol without changes of hepatic expression in canalicular cholesterol efflux transporter, ATP-binding cassette transporter G5 and G8 (ABCG5/8) [32]. In addition, ezetimibe treatment reduces biliary cholesterol in patients with gallstones and in mice [33,34]. These observations suggest that hepatic NPC1L1 may counterbalance ABCG5/8-mediated biliary secretion of cholesterol, thereby prevent extensive loss of cholesterol by transporting biliary cholesterol to hepatocytes, as illustrated in Fig. 1.

**NPC1L1 INHIBITOR: EZETIMIBE**

Ezetimibe was discovered as an active and potent metabolite of Schering-Plough’s SCH48461 substance and introduced as the first of a new class of selective cholesterol absorption inhibitor. Ezetimibe is absorbed and metabolized by intestine and liver to its glucuronide [35]. Both ezetimibe and its metabolite inhibit intestinal cholesterol absorption by preventing cholesterol transport from intestinal lumen to small intestinal enterocytes [36], resulting in lowering plasma cholesterol by 15% to 20% [6]. In the liver, ezetimibe reduces cholesterol delivery, which consequently increases cholesterol clearance from the circulation to hepatocytes via LDLr upregulation [21,32]. Thus, clinically proven ezetimibe has been used to treat hypercholesterolemia.

For a long time, the molecular target of ezetimibe has not been identified. In animal studies, NPC1L1 has a critical role in absorption of cholesterol and phytosterol, all of which is inhibited by ezetimibe [5,6]. Ezetimibe has no inhibitory effect on cholesterol transport in NPC1L1 knockout mice [5]. These findings suggest that NPC1L1 and ezetimibe act in the same pathway. In radioligand binding assay, ezetimibe binds to brush border membranes from various species and human NPC1L1 expressing cells but not those of NPC1L1 knockout mice [6]. Ezetimibe prevents the specific binding of NPC1L1 to clathrin/AP2 coated vesicles, internalization and endocytosis of cholesterol and thereby inhibits NPC1L1 mediated cholesterol uptake [16,17,37]. Taken together, NPC1L1 is a direct molecular target of ezetimibe.

**NPC1L1 AND DISEASE**

There is a strong positive correlation between high level of blood cholesterol and the incidence of atherosclerosis. Ezetimibe treatment and NPC1L1 deletion significantly reduce cholesterol absorption, decrease plasma cholesterol, and thereby inhibit the development of atherosclerosis in apolipoprotein E knockout mice, a model of atherosclerosis [28,38]. In subjects with hypercholesterolemia, ezetimibe monotherapy or ezetimibe–statin coadministration significantly decreases blood LDL-cholesterol (LDL-C) [39,40]. However, a 2-year clinical trial, Ezetimibe and Simvastatin in Hypercholesterolemia Enhances Atherosclerosis Regression (ENHANCE) shows no distinct change of carotid intima-media thickness between coadministration of ezetimibe and simvastatin and simvastatin alone in 720 heterozygous familial hypercholesterolemic patients [41]. This ENHANCE study has limitations such as subject number, subject selection, primary endpoints, treatment duration, and methodology. A long-term large clinical trial, Improved Reduction of Outcomes: Vytorin Efficacy International Trial (IMPROVE-IT) considers the problems of ENHANCE study and has been ongoing to demonstrate the cholesterol lowering effects of ezetimibe and its health outcomes. Although IMPROVE-IT is not completed, recent reported study of heart and renal protection demonstrates that the combination of simvastatin and ezetimibe is safe and effective to decrease cholesterol in 9,270 chronic kidney disease (CKD)
patients. Combination medication also helps to reduce atherosclerotic events [42]. However, simvastatin monotherapy shows the same extent of lowering cholesterol and major cardiovascular event, compared to simvastatin plus ezetimibe in CKD patients. Ongoing IMPROVE-IT trial will answer the contribution of ezetimibe to the reduction of cholesterol and the improvement of health outcomes.

Besides the effects of ezetimibe treatment or NPC1L1 deficiency on hypercholesterolemia, there are accumulated data showing beneficial outcomes of ezetimibe on health. Ezetimibe improves hepatic steatosis in wild type mice fed a high fat, high cholesterol diet [43], and a high fat diet [44,45]. And it also shows similar effects on leptin receptor deficient \(db/db\) mice [46] and Zucker obese fatty rats [47,48] resulting in the improvement of fatty liver and metabolic syndrome. In a rodent model of NAFLD induced by a methionine choline-deficient diet, ezetimibe administration ameliorates NAFLD, accompanied with the decrease of hepatic triglyceride [49]. Similar to ezetimibe treatment, NPC1L1 ablation prevents the development of fatty liver in animals [50,51]. In humans, ezetimibe shows favorable effects on fatty liver in obese subjects on a weight loss diet [52], patients with nonalcoholic steatohepatitis and dyslipidemia [53] and nonobese participants with NAFLD [54]. Given that strong correlation between NAFLD and insulin sensitivity, ezetimibe improves insulin sensitivity in animals with NAFLD [48-50]. High fat diet induced glucose intolerance and insulin resistance are also ameliorated in NPC1L1 knockout mice [50,51]. This demonstrates that ezetimi-

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**Fig. 1.** The role of Niemann-Pick C1-Like 1 (NPC1L1) in cholesterol transport in the intestine and liver. NPC1L1 is located at intestinal brush border and hepatocanicular membrane. In the lumen of small intestine, biliary and dietary cholesterol are mixed and solubilized to form micelles. NPC1L1 transports sterols from brush border membrane to intracellular compartments. Most free cholesterol (FC) is esterified by acyl coenzyme A: cholesterol acyltransferase 2 (ACAT2) to form cholesterol esters (CE), packed into chylomicrons and then secreted into mesenteric lymph. Liver NPC1L1 facilitates the transfer of secreted biliary cholesterol back into hepatocytes and prevents cholesterol loss. The cholesterol efflux transporter, ATP-binding cassette transporter G5 and G8 (ABCG5/ABCG8) also localizes at apical membrane of hepatocytes. Ezetimibe inhibits NPC1L1-mediated cholesterol transport in intestine and liver.
mibe or NPC1L1 deletion has beneficial impacts on NAFLD by insulin sensitivity.

One of proposed mechanisms is that ezetimibe treatment or NPC1L1 deletion prevents NAFLD by reducing reactive oxygen species, activating C-Jun-N-terminal kinase, and decreasing ER stress [47,48]. All of these alterations explain how NPC1L1 ablation and ezetimibe act cholesterol uptake in the liver, rather than improve steatosis. Jia et al. [51] demonstrate inhibitory effects of ezetimibe on lipogenesis in the liver. Genetic dysfunction of NPC1L1 or ezetimibe treatment reduces intestinal fat absorption and decreases cholesterol dependent LXR activation. Inactivation of LXR subsequently suppresses hepatic lipogenesis by modulating gene expression of SREBP-1c thereby prevents high fat induced hepatic steatosis. Focusing on favorable regulation of ezetimibe in glycemic control, our team investigated the mechanism how ezetimibe administration improves glycemic control. Chronic ezetimibe administration in male hyperphagic Otsuka Long-Evans Tokushima Fatty (OLETF) rats improves hyperglycemia compared to OLETF rats with vehicle. We also find that ezetimibe increases pancreatic size and beta cell mass. In addition, 20-week chronic ezetimibe administration increases serum glucagon-like peptide-1 (GLP-1) and decreases serum dipeptidyl peptidase-4 (DPP4) activity [55]. GLP-1 is released from β-cells in response to meal and affects metabolic syndrome by increasing insulin secretion and reducing glucagon production. DPP4 plays a critical role in glucose homeostasis by its action to GLP-1 degradation [56]. From these data, we suggest that NPC1L1 inhibition by ezetimibe may improve glucose homeostasis by mediating GLP-1.

NPC1L1 MEDIATED HCV ENTRY

As HCV leads to liver disease, it is very critical to understand the mechanisms of viral entry, virus-host cell interactions and escape for drug development. In 2011, Sainz et al. [7] find that cholesterol transporter, NPC1L1 is involved in HCV entry. Silencing antibody specifically against NPC1L1 suppresses HCV infection by regulating replication or secretion. They also discover that NPC1L1 inhibitor, ezetimibe blocks HCV uptake into cells in a cholesterol dependent manner and delays the establishment of HCV infection in mice pretreated for 2 weeks before infection, confirming the ability of this drug to inhibit HCV infection in vivo. Despite new interesting finding that NPC1L1 affects HCV entry, clinical practice studies are needed to evaluate efficacy of NPC1L1 as antiviral target.

Developement of Other cholesterol absorption inhibitors

Ezetimibe, NPC1L1 inhibitor, effectively reduces cholesterol absorption but it alone has not shown enough improvement in atherosclerotic or vascular events in patients with cardiovascular disease. Therefore, ezetimibe has been used with other cholesterol lowering agents such as statins. The inhibitory effects of dual combination on cholesterol is more effective, even small dose combination, than monotherapy [57]. U.S. Food and Drug Administration has approved a new fixed dose combination agent including atorvastatin and ezetimibe for the treatment of hyperlipidemia next to simvastatin/ezetimibe combination.

Table 1 summarizes the development of other NPC1L1 inhibitors. Spiroimidazolidinone was identified by similarity-based virtual screening and has low binding affinities of brush border membranes. Spiroimidazolidinone significantly inhibits mouse cholesterol absorption by 67% [58]. MD-0727 is introduced as an analog of ezetimibe and effectively decreases LDL-C in clinical trials [59]. It might be a next novel cholesterol absorption inhibitor. Other compounds such as AZD4121, canosimibe, KT6-971, and MK-6213 were developed and have been discontinued due to several issues including target product profile, bioavailability and efficacy [60].

conclusions

As a homolog of NPC1 protein, NPC1L1 was first introduced. NPC1L1 contains multiple conserved transmembrane domains, involved in cholesterol metabolism. NPC1L1 is expressed in not only intestinal brush border but also hepatocanalicular membrane. It mediates intestinal cholesterol absorption and
Intestinal and hepatic NPC1L1

hepatic biliary cholesterol secretion. NPC1L1 is a molecular target of ezetimibe, the first hypolipidemic agent. Ezetimibe inhibits NPC1L1 dependent cholesterol uptake by preventing its intracellular trafficking and clathrin mediated endocytosis. Besides NPC1L1 actions to cholesterol transport, NPC1L1 is recently found to be responsible for HCV virus infection. Still, the molecular mechanisms for NPC1L1 dependent cholesterol uptake in small intestine and liver in vivo and sequential cholesterol transport to ER remain elusive. It is also undetermined how NPC1L1 proteins are responsible for the incidence of metabolic disorders such as NAFLD, obesity diabetes, and virus entry. Future studies are required to investigate molecular mechanisms of NPC1L1.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

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