Ultra-small pH-responsive Nd-doped NaDyF₄ Nanoagents for Enhanced Cancer Theranostic by in situ Aggregation

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Abstract

To achieve accurate tumor location and highly efficient cancer therapy effect, the properties of cancer theranostic agents should be optimized and enhanced. In this work, ultra-small Nd doped NaDyF₄ were firstly reported as novel contrast agents for near-infrared second window downconversion luminescence (NIR II DCL) and magnetic resonance imaging. Based on the optimization strategy, gallic acid-Fe(III) complex modified NaDyF₄:10%Nd (NaDyF₄:10%Nd-GA-Fe) was selected as the optimal agent with high transversal relaxivity, strong NIR II DCL, high photothermal conversion efficiency, and low toxicity. In vitro experiment found that it can be aggregated rapidly in low pH condition, leading to the particle size increasing. Due to the theranostic properties coupled in NaDyF₄:10%Nd-GA-Fe are size dependent, properties enhancement was observed within the pH responsive aggregation progress. Further study in small animal model bearing tumor demonstrated the enhanced cancer theranostic by in situ aggregation. The optimized nanoagents have potential applications in medical and also provide a novel strategy for future study of cancer theranostic enhancement.

Key words: neodymium, NaDyF₄, NIR II, magnetic resonance imaging, photothermal therapy.

Introduction

Meeting the requirement of real-time diagnosis, luminescence imaging recently became the research hotspot. With fascinating and unique properties, rare earth elements have a great reputation in this domain, which lead to the wide and deep exploration in lanthanide doped nanomaterials to develop high efficient agents for luminescence imaging. Since biological tissues has an "optical transparency window" in near-infrared second window (NIR II) region, NIR II luminescence imaging take the advantage of deep tissue penetration, reduced photo-damage effects, low photo-scattering, and high signal-to-noise ratio in turn. Recently, a brilliant strategy was come up, applying neodymium (Nd) as novel sensitizer which can be excited by 808 nm laser and emitted in NIR II region. Thanks to the long excitation and emission wavelength, Nd doped nanomaterials as NIR II downconversion luminescent (DCL) agents have the potential application in accurate real-time cancer diagnosis. However, many works pointed out that luminescence imaging still have limitation in low tissue resolution. Thence, developing multifunctional luminescence agents coupled with high resolution imaging should be considered as priority. Among the imaging methods, T₂-weighted magnetic resonance imaging (MRI) is widely used in clinical cancer diagnosis and can provide high
resolution 3D images of pathological phenomena such as tumors and inflammation\textsuperscript{11,12}. Previous works suggested dysprosium (Dy) doped agents can be applied for effective T\textsubscript{2}-weighted MRI \textit{in vitro} and \textit{in vivo}\textsuperscript{13-15}. For cancer ablation following comprehensive diagnosis, photothermal therapy (PTT) is also a novel targeted and non-invasive therapeutic intervention for cancer treatment, which can be easily achieved by the combination of photothermal conversion agents and suitable irradiation in the cancerous area, thereby killing ordinary tumor cells at a relative high temperature (> 42°C)\textsuperscript{16,17}. Among all the discovered photothermal conversion agents, mainly including noble metals\textsuperscript{18-20}, semiconductors\textsuperscript{21-24}, molecular dyes\textsuperscript{25-27}, conjugated polymers\textsuperscript{28-31}, and carbon-based materials\textsuperscript{32-34}, the molecular dyes constitute the largest and most diverse family, which provide they with special performance in nanomaterial-based PTT\textsuperscript{35}. Therefore, it is necessary to explore multifunctional nanoagents with outstanding NIR II DCL imaging and T\textsubscript{2}-weighted MRI properties, as well as photothermal therapeutic effect for cancer theranostic.

Apart from multifunction, the ultra-small hydrodiameter (sub 10 nm) was also required to avoid the accumulation in reticuloendothelial system, partly contributing to the long circulation time, high tumor accumulation amount, thus resulting in cancer theranostic improvement\textsuperscript{36,37}. However, the theranostic effect was still limited due to the small size. Therefore, a strategy for theranostic properties improvement of the accumulated nanoagents in tumor region was needed to be put forward. Considering the fact that rare earth based luminescence, T\textsubscript{2}-weighted MRI, and photothermal conversion properties were size dependent, developing a multifunctional nanoagent which can \textit{in situ} aggregate in tumor region would seem to be a reasonable strategy for the cancer theranostic enhancement\textsuperscript{38-40}. Since the tumor region was more acidic than normal tissues, the pH-induced \textit{in situ} aggregation of the nanoagent in tumor region by aid of pH sensitive ligands was feasible for highly cancer theranostic enhancement.

In this work, we firstly reported a series of multifunctional Na\textsubscript{3}Dy\textsubscript{4} nanoagents doped with different amount Nd for NIR II DCL imaging and T\textsubscript{2}-weighted MRI, further discussing and optimizing their imaging properties. The optimized agent was then modified with photothermal conversion and pH-responsive gallic acid-Fe(III) complex (GA-Fe) and tested their pH induced theranostic properties enhancement. Following efforts have been directed towards treatment of tumor bearing-mice with the optimized GA-Fe modified nanoagents to illustrate their \textit{in situ} aggregation-mediated enhanced theranostic effect and biosafety \textit{in vitro} and \textit{in vivo} (Scheme 1).

\textbf{Scheme 1.} Optimization of ultra-small Na\textsubscript{3}Dy\textsubscript{4}\textsubscript{x}%Nd nanoagents for enhanced cancer theranostic by pH-responsive \textit{in situ} aggregation.
Results and Discussion

Synthesis and characterization of NaDyF$_4$:x%Nd.

Hydrophobic oleic acid (OA) coated NaDyF$_4$:x%Nd (x = 2.5, 5, 10, 20) nanoagents were synthesized via a typical solvothermal method$^{15}$. Transmission electron microscopy (TEM) images showed that the NaDyF$_4$:x%Nd nanoagents are well dispersed in cyclohexane with good crystallinity and uniform shape (Figure 1A-D). All samples exhibit a narrow size distribution (6.6 ± 1.3 nm). The unobvious size change through Nd dopant may be interpreted to the low dopant amount and ultra-small size. The energy-dispersive X-ray analysis (EDXA) spectra and ICP-MS of the NaDyF$_4$ nanoagents confirmed the exclusive composition of Na, Dy, and F (Figure S1 and Table S1), which indicated the element formation of Nd doped NaDyF$_4$ nanoagents. High-resolution TEM (HR-TEM) images and powder X-ray diffraction (XRD) patterns of NaDyF$_4$:x%Nd nanoagents showed good crystallinity and confirmed their hexagonal phase (JCPDS: 027-0687 for NaDyF$_4$ and JCPDS: 35-1367 for NaNdF$_4$) (Figure S2). Selected area electron diffraction (SAED) pattern could also support this observation (Figure S3).

Figure 1. Characterizations and optimization of NaDyF$_4$:x%Nd (x = 2.5, 5, 10, 20) nanoagents. TEM images A-D) of hydrophobic NaDyF$_4$:x%Nd (x = 2.5, 5, 10, 20) nanoagents (Insert: size distribution of NaDyF$_4$:x%Nd). Hydrodiameter E), transversal relaxivity (r$_2$) F), and NIR II DCL G) properties of hydrophilic NaDyF$_4$:x%Nd (x = 2.5, 5, 10, 20) nanoagents. The properties variation H) as a function of Nd concentration in obtained nanoagents (HD: the abbreviation of hydrodiameter; NIR II DCL: the NIR II DCL intensity under 808 laser irradiation with power intensity of 4.5 W cm$^{-2}$). These results confirmed the successful controlled synthesis of ultra-small hexagonal phase NaDyF$_4$:x%Nd (x = 2.5, 5, 10, 20) nanoagents and demonstrated that the 10% Nd doped NaDyF$_4$ was preferred for further modification and applications.
Optimization of pH-responsive multifunctional nanoagents

Multifunctional imaging properties optimization

To obtain the hydrophilic NaDyF4:x%Nd for further application, nanoagents were treated using nitrosoniumtetrafluoroborate (NOBF₄) to remove OA ligands on the surface, and gallic acid (GA) was then used to coat the four types of ligand-free nanoagents. Fourier transform infrared (FTIR) spectra proved the successful ligand exchange (Figure S4). Zeta potential investigation and dynamic light scattering (DLS) measurement suggested that all the four type nanoagents can well dispersed in water with a similar average hydrodiameter (HD) (Figure 1E).

The water-dispersible NaDyF₄:x%Nd nanoagents were then investigated for positive longitudinal relaxivity (r₁) and negative transverse relaxivity (r₂) measurements, which were measured as a function of nanoagents molar concentrations at room temperature using a 3 T magnetic resonance imaging (MRI) system. All NaDyF₄:x%Nd nanoagents showed good r₂ enhancement and relative low r₁ as a result of the presence of the Dy³⁺ ions (Figure 1F and Figure S5). As the Nd³⁺ dopant amount increasing, r₂/r₁ of NaDyF₄:x%Nd decreased monotonously and surprisingly showed a linear relationship (Figure S6). Then, the near-infrared second window downconversion luminescence (NIR II DCL) was also taken into comparison. The NIR II DCL emission can be ascribed to the f-f electronic transition of Nd³⁺.

Figure S5). As the Nd³⁺ dopant amount increasing, r₂/r₁ of NaDyF₄:x%Nd decreased monotonously and surprisingly showed a linear relationship (Figure S6). Then, the near-infrared second window downconversion luminescence (NIR II DCL) was also taken into comparison. The NIR II DCL emission can be ascribed to the f-f electronic transition of Nd³⁺. Upon 808 nm excitation, DCL emission peaks of Nd in the NIR II region occurred at 1050 nm and 1330 nm, which correspond to the transition from 4F₃/2 → 4I₁₁/2 and 4F₃/2 → 4I₁₃/2. (Figure 1G and Figure S7). The interplay between both Nd concentration-induced absorption increment and concentration-induced luminescence quenching result in the max NIR emission peak at 10% Nd dopant amount. Nevertheless, with continuous increasing of Nd doping amount (>13%), the cross-relaxation would lead to the luminescence quenching accordingly. NaDyF₄:10%Nd has a similar quantum field of ~19.4% with previous work (~22% for NaGdF₄:3%Nd) and the different optimized Nd concentration may be contributed to the various hosts, which influenced the surrounding of Nd and their optical properties by crystal field. Since the luminescence intensity at 1050 nm was much stronger than that at 1330 nm, we then chose the 1050 nm luminescence for further study and imaging application. Considering all the factors above, NaDyF₄:10%Nd among the four nanoagents has a preferable MR and NIR II DCL imaging effect, which can be optimized for the further study (Figure 1H).

pH-responsive aggregation and properties enhancement

Gallic acid-Fe(III) complex (GA-Fe) has strong absorption at the range of 500-700 nm, and shows pH-responsive properties which can be efficiently aggregated in acidic condition (tumor region). The aggregation property of nanoagents may contribute to the protonation of modified GA-Fe(III) and the formation of hydrogen bond between oxygen atoms in hydroxyl groups and carboxyl groups according to previous report. Hence, GA-Fe modified optimized NaDyF₄:10%Nd nanoagents (NaDyF₄:10%Nd-GA-Fe) can be expected to be photothermal agents and enhance cancer theranostic effect.

First, the NaDyF₄:10%Nd-GA-Fe was synthesized and characterized (Figure 8A,9). Due to the separation of GA-Fe absorption wavelength (500-700 nm) and NaDyF₄:10%Nd emission wavelength (1050 ± 25 nm), the GA-Fe modification has no obvious re-absorption effect on NIR II DCL of NaDyF₄:10%Nd, which can ensure the efficiency of DCL imaging (Figure 9). The NaDyF₄:10%Nd-GA-Fe can be well dispersed with a small hydrodiameter of 8.7 ± 0.9 nm and zeta potential of -28.47 mV. It also shows a similar absorption as reported free GA-Fe complex (Figure S10). Then, we studied the aggregation and enhancement effect under different pH condition (pH = 4.5-7.5). TEM and DLS analysis showed an increasing of the average size of NaDyF₄:10%Nd-GA-Fe from 6.5 ± 0.7 nm (HD: 8.7 ± 0.9 nm) to 50.4 ± 4.4 nm (HD: 59.3 ± 6.6 nm) in acidic condition (pH = 4.5) and no obvious aggregation was observed in neutral condition (pH = 7.5) within 30 min (Figure 2A, B, D, E). Besides, relative high concentration of NaDyF₄:10%Nd-GA-Fe showed more rapid aggregation than low concentration, which can be explained as the easy attachment of nanoparticles in high concentration solution (Figure 2F). These results suggest that NaDyF₄:10%Nd-GA-Fe can efficiently aggregate under acidic condition. Notably, the aggregation-decomposition progress can be repeated by changing the pH condition (Figure 2C). After five circle of aggregation-decomposition progress, NaDyF₄:10%Nd-GA-Fe can still show good solubility and small HD (<10 nm) in solution (Figure 2G).

Besides, the ultraviolet-visible-near infrared (UV-vis-NIR) spectra also provide the same consequence by an obvious increase of absorption in low pH condition (Figure 3A). Because the aggregation of nanoagents increase the average size and further increase the molar extinction coefficient (ε value), the photothermal conversion effect of NaDyF₄:10%Nd-GA-Fe under different pH condition was calculated using the following equation.
According to equation 1 and equation S1, the ε value of NaDyF₄:10%Nd-GA-Fe was increasing from $1.89 \times 10^6$ to $1.37 \times 10^8$ M⁻¹ cm⁻¹ with the pH decreasing from 7.5 to 4.5, monotonously. The photothermal conversion efficiency (η value) of NaDyF₄:10%Nd-GA-Fe (200 μg mL⁻¹) under the pH = 4.5 condition was calculated using the following equation.

$$\eta = \frac{h\Delta T_{\max} - Q_0}{I(1-10^{-A})}$$  \hspace{1cm} (2)

According to equation 2 and equation S2, the η value of the NaDyF₄:10%Nd-GA-Fe is 60.12%, which is much higher than that of recently reported photothermal coupling agents (47.83% for rGO-Bi₂S₃, 36.10% for SnS) (Figure S11)¹³,⁴⁴. Moreover, the enhancement of diagnosis signals was also studied. With the condition turned from neutral to acidic, the T₂-weighted MRI and NIR II DCL signal intensity can be efficiently increased, which proved that NaDyF₄:10%Nd-GA-Fe can be used for enhanced cancer theranostic (Figure 3E-H). The increasing of MRI and NIR II DCL property may contribute to the decrease of surface area, which result in the weakening of spin-canting effect and surface quenching effect.

Figure 2. pH-induced aggregation-decomposition progress of NaDyF₄:10%Nd-GA-Fe. A-C) TEM images of NaDyF₄:10%Nd-GA-Fe in aggregation-decomposition progress. D,E) Hydrodiameter vs time plots of NaDyF₄:10%Nd-GA-Fe under different pH condition. F) Hydrodiameter vs concentration and pH plots of NaDyF₄:10%Nd-GA-Fe. G) Hydordiameter variations of NaDyF₄:10%Nd-GA-Fe under the different pH condition for five cycles. These results suggested NaDyF₄:10%Nd-GA-Fe can response to different pH condition and hence aggregate in acidic condition.
Figure 3. pH-induced properties enhancement of NaDyF₄:10%Nd-GA-Fe. A) UV-vis-NIR spectra of NaDyF₄:10%Nd-GA-Fe under different pH condition within 30 min (pH = 4.5, 5.5, 6.5, and 7.5). Insert: relative intensity at 808 nm. Photothermal temperature change B) and photothermal images C) of NaDyF₄:10%Nd-GA-Fe under laser irradiation with different power intensity. D) Temperature variations of NaDyF₄:10%Nd-GA-Fe (200 ppm) under the continuous irradiations of 808 nm laser for four cycles. E,F) Relative MR intensity vs time plots of NaDyF₄:10%Nd-GA-Fe under different pH condition (Insert: MRI images in tube). G,H) Relative DCL intensity vs time plots of NaDyF₄:10%Nd-GA-Fe under different pH condition (Insert: NIR II DCL images in tube). These results suggested NaDyF₄:10%Nd-GA-Fe has enhanced properties in acidic condition and the potential to achieve enhanced cancer theranostic.

The stability of NaDyF₄:10%Nd-GA-Fe was also investigated. UV-vis-NIR spectra and DLS of NaDyF₄:10%Nd-GA-Fe after several days standing and after several hours irradiation were measured, and suggested great stability under neural condition and different biological solution (Figure S12, Table S2, S3). Inductively coupled plasma mass spectrometry (ICP-MS) results suggested that no noticeable Fe(III) and Dy (III) leaching from NaDyF₄:10%Nd-GA-Fe after 60 days standing or 1 hour irradiation in both neural and acidic condition. It was also found that NaDyF₄:10%Nd-GA-Fe was a robust photothermal heater after five cycles of NIR laser-induced heating (Figure 3D). In all, the pH-responsive enhanced multifunctional properties and stability of NaDyF₄:10%Nd-GA-Fe nanocomposites make them superior as a promising pH-responsive multifunctional agent for enhanced cancer theranostic.

Power density optimization
Since the body temperature of human beings was ~37°C and the temperature over 42°C may increase the potential risk of overheat to normal tissues, temperature change less than 5°C will provide a safe condition for cancer diagnosis. To avoid the potential damage to normal tissues during the cancer diagnosis and obtain a high photothermal temperature change during the cancer therapy, the power density of irradiation laser was also optimized (Figure 3B, C). When the power density was 0.25 W cm⁻² or lower, there are no obvious temperature increase (<3°C), and thus NIR II DCL imaging can be performed under this power density stage (Figure 3B i). More importantly, the temperature change was lower than 5°C under 0.25-0.44 W cm⁻² and can show a significant difference between acidic condition and neural condition. Hence, the power intensity range
guarantees the safety of photothermal imaging (Figure 3I ii). While the power intensity increased to the range of 0.44-0.74 W cm⁻², the temperature can increase by over 5°C in acidic condition and only by less than 5°C in neutral condition, which suggested that the suitable laser power intensity range can both ensure the therapeutic effect and reduce the heat threaten to normal tissue (Figure 3B iii). Under the power intensity >0.74 W cm⁻², the temperature change can achieved over 5°C in both the acidic and neural condition, which may cause potential damage risk and hence not suitable for photothermal therapy (PTT) (Figure 3B iv). The low temperature change would lead to cell death, but the surviving cells would have the capacity to resistant further heat damage by activating the heat-shock proteins. Meanwhile, the high temperature can result in irreversible injury, by accelerating biological reaction and biomelecule damage. Therefore, the nanoagents would show high cytotoxicity under acidic condition rather than neutral condition. This result suggested that the using of 808 nm laser with different power density can both ensure the cancer diagnosis safety and the theranostic effect.

**Cancer theranostic enhancement in vitro and in vivo**

T₂-weighted MR image can reveal pathological phenomena and high-resolution 3D images, which would be benefit to the location of tumor.⁴¹ Luminescence imaging achieves dynamic real-time imaging to monitor the ablation of tumor and photothermal images are used to record the real-time temperature changes.⁴⁵,⁴⁶ Therefore, the combination of imaging modalities can provide complementary information in cancer theranostic. To confirm the potential application of NaDyF₄:10%Nd-GA-Fe as a multifunctional imaging probe and show enhanced imaging effect in tumor region than normal tissue, a series of in vitro and in vivo experiments were carried out. A typical non-pH-responsive citric acid-Fe (III) complex modified NaDyF₄:10%Nd (NaDyF₄:10%Nd-CA-Fe, HD: 9.1 nm, zeta potential: -27.60 mV) with similar imaging properties was designed and optimized for comparison (Figure S13).

To demonstrate the feasibility of NaDyF₄:10%Nd-GA-Fe as an enhanced multimodal imaging probe, we firstly compared the T₂-weighted MRI in vitro. In comparison with in neutral condition, NaDyF₄:10%Nd-GA-Fe in acidic condition showed a sharper contrast and NaDyF₄:10%Nd-CA-Fe showed no obvious difference between those two conditions (Figure 4A). Further study was taken on HCT116 tumor-bearing mice model. Post-injection T₂-weighted MRI images at 0 and 120 min showed clearly contrast in the tumor region, which illustrated the possibility as a T₂-weighted MRI contrast agent for bioapplication (Figure 4B). Besides, T₂-weighted MRI signals in tumor and liver region were measured 1 hour post-injection and the tumor/liver ratio from the MRI images were also calculated. The result suggested that mice receiving NaDyF₄:10%Nd-GA-Fe had much more obvious tumor/liver ratio change than NaDyF₄:10%Nd-CA-Fe (Figure S14). To further demonstrate the pH-induced T₂-weighted MRI enhancement in vivo, nanoagents was intratumorally (2 mg per kg body weight of mouse) injected to tumor-bearing mice, respectively. By intratumorous injection, mice receiving NaDyF₄:10%Nd-GA-Fe showed much more significant T₂-weighted MRI signal enhancement in tumor region than that receiving NaDyF₄:10%Nd-CA-Fe (Figure 4C). The same result was also found in NIR II DCL imaging (Figure 4D,E and Figure S15). The above results suggested that NaDyF₄:10%Nd-GA-Fe can selectively enhanced the MRI and NIR II DCL signal in tumor region due to the acidic microenvironment, which illustrated the pH-responsive enhanced cancer diagnosis by in situ aggregation progress in vivo (Figure S16).

Since NaDyF₄:10%Nd-GA-Fe possessed good solubility, stability, and photothermal conversion efficiency in biological aqueous media, it could be also considered as a photothermal therapeutic agent. NaDyF₄:10%Nd-GA-Fe under the laser irradiation with an optimized power intensity of 0.64 W cm⁻² showed enhanced photothermal therapeutic effect on cancerous cells (cell viability: <10%) compared to normal cells (cell viability: >75%) (Figure 4F). Considering the fact that NaDyF₄:10%Nd-GA-Fe has enhanced photothermal therapeutic effect in tumor cells and the thermal signal of tumors increased over time and showed an obvious contrast-distinction with surrounding tissue after NaDyF₄:10%Nd-GA-Fe injection (Figure 4G), PTT in vivo was further studied with several groups of HCT116 tumor-bearing mice. In the test group, mice were intravenously injected with a PBS solution of NaDyF₄:10%Nd-GA-Fe and irradiated by a 808 nm continuous laser (Material + Laser, n = 5). For comparison, mice without injection of NaDyF₄:10%Nd-GA-Fe (Laser, n = 5), without laser irradiation (Materials, n = 5), and without injection or laser irradiation were chosen as the control groups. Upon irradiation at 808 nm (0.64 W cm⁻²) for 6 min, the tumors of the test groups shrank within 9 days. After 11 days therapy period, the tumors were eliminated, with residual black scars and no obvious swollen parts, and the relative tumor volume (V/V₀) shrank to 0. In contrast, the V/V₀ of tumors in the control and blank groups showed rapid growth over time (Figure 4H). Besides, the bodyweight of mice
were also determined. Within the first 3 days, the mice in test group showed a decrease in bodyweight, which demonstrated the efficient ablation of tumor by PTT. However, their bodyweight increase rapidly in the following 11 days, suggesting that the mice receiving PTT were in the recovery process from therapy and make a supplementary to the low toxicity of nanoagents (Figure S17).

**Figure 4. Enhanced cancer theranostics effect of NaDyF4:10%Nd-GA-Fe in vitro and in vivo.** T1-weighted MRI images in pre- and post-incubated cells A) and tumor-bearing mice within 120 min post-injection B). C) T2-weighted MRI signal in tumor region after intratumorous injection of NaDyF4:10%Nd-GA-Fe. NIR II DCL images in pre- and post-incubated cells D) and tumor-bearing mice E). F) Photothermal therapy efficiency in vitro. G) Photothermal images in pre- and post-injection tumor-bearing mice. H) Tumor growth rates of test group (Material + Laser), control groups (Laser only, Materials only) and blank group (Blank). I) Tissue distribution of NaDyF4:10%Nd-GA-Fe within 1 and 4 hour post-injection intravenously. NaDyF4:10%Nd-CA-Fe was used for comparison (control in 4A and 4D). Statistical significance was determined from one-way t tests. Note: *p < 0.05, **p < 0.01, ***p < 0.001
The tissue distribution of both NaDyF₄:10%Nd-GA-Fe and NaDyF₄:10%Nd-CA-Fe was also quantitatively analyzed by ICP-MS and the Dy³⁺ is an indicator of the nanoagents which are not directly measured. Figure 4I shows that high dose of Dy³⁺ were uptake in both tumor region. These results demonstrated that the nanoagents can passive target and accumulate in the tumor region. More importantly, both of the nanoagents have a similar distribution in mice within 1 hour post-injection, indicating the cancer theranostic enhancement of NaDyF₄:10%Nd-GA-Fe in tumor-bearing mice model based on their pH responsive properties. Besides, the relatively long blood circulation half-live time was determined to be ~56 min benefit from their ultra-small HD. Thence, to sum up the points which we have just indicated, the NaDyF₄:10%Nd-GA-Fe can be applied to enhance cancer theranostic in vitro and in vivo.

Toxicity study

We then studied the in vivo toxicity of NaDyF₄:10%Nd-GA-Fe to determine the safety of bioapplication. A MTT assay shows no significant difference in proliferation of both HCT116 and CCC-HEL-1 cells in the absence or presence of NaDyF₄:10%Nd-GA-Fe (0-1.0 mg mL⁻¹) within 24 and 48 hours (Figure 5A,B). Cellular viabilities were estimated to over 85% even at high dose. The half maximal (50%) inhibitory concentration (IC50) was also investigated via the MTT assay, and was calculated to be 3.97 mg mL⁻¹ (24 hours) and 3.19 mg mL⁻¹ (48 hours) for HCT116 cells, 3.94 mg mL⁻¹ (24 hours) and 3.42 mg mL⁻¹ (48 hours) for CCC-HEL-1 cells (Figure S18). The oxidation stress to HCT116 and CCC-HEL-1 cells was also determined by fluorescence method using singlet oxygen sensor green (SOSG). The result showed no obvious reactive oxygen species accumulation in both NaDyF₄:10%Nd-GA-Fe incubated HCT116 and CCC-HEL-1 cells, which suggested low oxidation stress of NaDyF₄:10%Nd-GA-Fe to cells (Figure S19).

Moreover, healthy mice were injected intravenously with NaDyF₄:10%Nd-GA-Fe at the dose of 20 mg per kg body weight of mouse. Neither mouse death nor noticeable abnormal behavior was observed. Serum biochemistry assays and complete blood panel tests were carried out on NaDyF₄:10%Nd-GA-Fe-injected mice at 24 hours, 7 days, 30 days, and 60 days. The liver and kidney function indices, including alanine aminotransferase (ALT), aspartate aminotransferase (AST), total bilirubin (TBIL), albumin (ALB), total protein (TP), uric acid (UA) and creatinine (CREA) were all tested to be normal, which suggested no hepatic or kidney function influenced by NaDyF₄:10%Nd-GA-Fe (Figure 5C-I). Besides, hematoxylin-eosi (H&E) stained tissues sections were also taken to testify histological examination, which suggested no appreciable adverse effect of NaDyF₄:10%Nd-GA-Fe to examined major organs, including liver, spleen, kidney, lung, and heart at 24 hours, 7 days, 30 days, and 60 days post-injection (Figure 5J). Dy amount in urine and feces was also determined by ICP-MS to study their metabolic pathway. The results suggested that NaDyF₄:10%Nd-GA-Fe can be rapidly metabolized by renal and fecal pathway (>80% within 48 hours), which greatly ensured their biosafety (Figure S20). Our results indicated that NaDyF₄:10%Nd-GA-Fe was nontoxic to mice at our tested dose.

Conclusions

In this work, we reported four types of ultra-small Nd doped NaDyF₄ nanoagents via typical solvothermal method. Considering their theranostic properties, NaDyF₄:10%Nd was optimized for further GA-Fe modification. The obtained NaDyF₄:10%Nd-Fe-GA exhibited high r2 (27.9 mM⁻¹ s⁻¹), strong NIR II DCL (1050 nm and 1330 nm), outstanding photothermal conversion efficiency (60.12%), and unobservable toxicity. More importantly, the effective pH-responsibility properties make the nanoagents aggregate in acidic condition. Moreover, the in vitro and in vivo studies showed enhanced contrast in tumor region, which demonstrated that ultra-small NaDyF₄:10%Nd-Fe-GA has the potential to be applied in efficient cancer theranostic by pH-responsive in situ aggregation. Our results provided a novel optimization strategy of developing multifunctional nanoagents, as well as encouraged the further research in the subfield of cancer theranostic enhancement.

Experimental

Materials

Rare-earth oxides Nd₂O₃ (99.99%) and Dy₂O₃ (99.9%) were purchased from Beijing Lansu Co. China. Sodium hydroxide (NaOH), hydrochloride (HCl), ethanol, cyclohexane, and dichloromethane were purchased from Beijing Chemical Reagent Company. Oleic acid (OA) was purchased from Aldrich. 1-Octadecene (ODE), ammonium fluoride (NH₄F), and nitrosoniumtetrafluoroborate (NOBF₄) were purchased from Alfa Aesar Chemical (Tianjing) Co. Ltd. Iron (III) chloride (FeCl₃·6H₂O) was purchased from Thermo Fisher Scientific (China) Co Ltd. Gallic acid (GA) was purchased from Energy Chemical (Shanghai) Co. Ltd. Rare earth chlorides (LnCl₃, Ln: Nd, Dy) were prepared by dissolving the...
corresponding metal oxide in HCl solution at elevated temperature and then evaporating the water completely under reduced pressure. All other chemical reagents were of analytical grade and were used directly without further purification. Deionized (DI) water was used throughout.

Figure 5. Toxicity of NaDyF₄:10%Nd-GA-Fe in vitro and in vivo. The viability of HCT116 A) and CCC-HEL-1 B) cells incubated with different concentrations NaDyF₄:10%Nd-GA-Fe within 24 and 48 hours. Serum biochemistry results obtained from mice injected with NaDyF₄:10%Nd-GA-Fe (20 mg kg⁻¹) different time post-injection (24 hours, 7 days, 30 days, and 60 days) and mice receiving no injection (control). C) ALT (alanine aminotransferase); D) AST (aspartate aminotransferase); E) TBIL, total bilirubin; F) ALB, albumin; G) TP, total protein; H) CREA, creatinine; I) UA, uric acid. J) H&E-stained tissue sections from mice injected with NaDyF₄:10%Nd-GA-Fe (20 mg kg⁻¹) different time post-injection (24 hours, 7 days, 30 days, and 60 days) and mice receiving no injection (control). Tissues were harvested from heart, liver, spleen, lung, and kidney.
Synthesis of the OA-coated NaDyF₄:x%Nd (x = 2.5, 5, 10, 20)

In a typical experiment, a mixture of 1 mM LnCl₃ (Ln: Nd, Dy), 6 mL OA, and 15 mL ODE were added into a 100 mL three-necked flask. Under the vacuum, the mixture was heated to 160°C to form a clear solution, and then cooled to room temperature. After the solution cooling down, NaOH (0.025 mmol, 0.1 g) and NH₄F (0.04 mmol, 0.1481 g) were added into the flask directly and stirred for 30 min. The solution was slowly heated with gently stirred, degassed at 100°C, and then heated to 300°C and maintained for 1 h under the argon atmosphere. After the solution was cooled naturally, the nanoparticles were washed with ethanol/cyclohexane (1:1 v/v) several times and dispersed in cyclohexane15.

Synthesis of the GA-coated NaDyF₄:x%Nd (x = 2.5, 5, 10, 20)

Ligand-free NaDyF₄:x%Nd (x = 2.5, 5, 10, 20) was obtained by a reported method47. In a typical experiment, 5 mL of dichloromethane solution of NOBF₄ (0.01 M) was dropped into 5 mL of OA-coated NaDyF₄:x%Nd (x = 2.5, 5, 10, 20) dispersion in hexane (~5 mg mL⁻¹) at room temperature. Then, the ligand-free NaDyF₄:x%Nd (x = 2.5, 5, 10, 20) were washed with ethanol several times and dispersed in DI water. Ligand-free NaDyF₄:x%Nd (x = 2.5, 5, 10, 20) were then dispersed in a saturated solution of GA as the host reagent, being ultrasonically treated for 10 min at room temperature. The resultant mixture was separated to obtain GA-coated NaDyF₄:x%Nd (x = 2.5, 5, 10, 20) (NaDyF₄:x%Nd-GA) via centrifugation and washed with DI water several times to remove the extra GA.

Synthesis of the NaDyF₄:10%Nd-GA-Fe

NaDyF₄:10%Nd-GA-Fe were fabricated by simply mixing 5 mL 1.0 mg mL⁻¹ FeCl₃ with 10 mL 1 mg mL⁻¹ NaDyF₄:10%Nd-GA suspension (pH = 7.4) at room temperature. After vigorous stirring for another 50 min, the resulting colloidal solution was centrifuged and washed with DI water for several times.

Photothermal properties measurement

Photothermal imaging system was designed as described in previously literature58. To investigate the photothermal effect, NaDyF₄:x%Nd (x = 2.5, 5, 10, 20) and NaDyF₄:10%Nd-GA-Fe suspensions (200 μg mL⁻¹) were poured in specimen bottles (total volume of 2.0 mL), irradiated by continuous-wave diode NIR laser with a center wavelength of 808 nm and output power of 0-1.5 W cm⁻² for 2 min or 6 min. The temperature was measured by a digital thermometer with a thermo couple probe every 0.133 s. The photothermal images of NaDyF₄:x%Nd (x = 2.5, 5, 10, 20) and NaDyF₄:10%Nd-GA-Fe in solution were obtained using FLIR E40 equipment running on FLIR tools systems.

In vivo NIR II DCL imaging

The mice were scanned before and after the administration of contrast agent. The mice were injected with the solution of NaDyF₄:10%Nd-GA-Fe intravenously (10 mg per kg body weight of mouse) and applied to NIR II DCL imaging, which was performed with a modified DCL in vivo imaging system designed as described in previously literature34. In vivo NIR II DCL signals were collected at 1060 ± 12 nm. NaDyF₄:10%Nd-GA-Fe was used for comparison.

Oxidation stress study

Singlet oxygen sensor green reagent (SOSG) solution (5 mL, 0.1 mM; molecular probes) was added to the medium containing HCT116 cells, NaDyF₄:10%Nd-GA-Fe incubated HCT116 cells, CCC-HEL-1 cells, and NaDyF₄:10%Nd-GA-Fe incubated CCC-HEL-1 cells. The fluorescence intensity of SOSG was determined at 531 nm with an excitation wavelength of 488 nm. Fluorescent measurements were obtained using a fluorescence spectrophotometer (Hitachi F-7000).

Supplementary Material

Supplementary experiment section and figures. http://www.thno.org/v07p4217s1.pdf

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Competing Interests

The authors have declared that no competing interest exists.

References

[1] Chen G, Roy I, Yang C, et al. Nanochemistry and Nanomedicine for Nanoparticle-based Diagnostics and Therapy. Chem Rev. 2016; 116:2826-2885.
[2] Auzel F. Upconversion and Anti-stokes Processes with f and d Ions in Solids. Chem Rev. 2004; 104:139-173.
[3] Li XM, Zhang F, Zhao DY. Lab on Upconversion Nanoparticles: Optical Properties and Applications Engineering via Designed Nanostructure. Chem Soc Rev. 2015; 44:1346-1378.
[4] Zhang Y, Hong G, Zhang Y, et al. Ag₅S Quantum Dot: A Bright and Biocompatible Fluorescent Nanoprobe in the Second Near-Infrared Window. ACS Nano. 2012; 6:3695-3702.
Pang B, Zhao Y, Luehmann H, et al. 64
Wang J, Liu J, Liu Y, et al. Gd-Hybridized Plasmonic Au-Nanocomposites
Kang S, Lee J, Ryu S, et al. Gold Nanoparticle/Graphene Oxide Hybrid Sheets
Hu DH, Zhang JN, Gao GH, et al. Indocyanine Green-Loaded
Cao Y, Dou J, Zhao N, et al. Highly Efficient NIR-II Photothermal Conversion
Ju E, Dong K, Liu Z, et al. Tumor Microenvironment Activated Photothermal
Liang XL, Li YY, Li XD, et al. PEGylated Polypyrrole Nanoparticles
Wang Y, Yang T, Ke H, et al. Dual-Modal-Imaging Guided Synergistic Chemo-Thermal Tumor Therapy. Adv Funct Mater. 2015; 25:2520-2529.