Prevention of tumorigenesis in mice by exercise is dependent on strain background and timing relative to carcinogen exposure

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Among cancer diagnoses, colorectal cancer (CRC) is prevalent, with a lifetime risk of developing CRC being approximately 5%. Population variation surrounding the mean risk of developing CRCs has been associated with both inter-individual differences in genomic architecture and environmental exposures. Decreased risk of CRC has been associated with physical activity, but protective responses are variable. Here, we utilized a series of experiments to examine the effects of genetic background (strain), voluntary exercise (wheel running), and their interaction on azoxymethane (AOM)-induced intestinal tumor number and size in mice. Additionally, we investigated how the timing of exercise relative to AOM exposure, and amount of exercise, affected tumor number and size. Our results indicated that voluntary exercise significantly reduced tumor number in a strain dependent manner. Additionally, among strains where exercise reduced tumor number (A/J, CC0001/Unc) the timing of voluntary exercise relative to AOM exposure was crucial. Voluntary exercise prior to or during AOM treatment resulted in a significant reduction in tumor number, but exercise following AOM exposure had no effect. The results indicate that voluntary exercise should be used as a preventative measure to reduce risk for environmentally induced CRC with the realization that the extent of protection may depend on genetic background.

As with any complex disease, cancer risk is affected by both genetic and environmental factors. Of the 90% of colorectal cancers (CRC) not caused by inherited mutations in specific cancer-associated genes1,2, environmental variables are thought to contribute at least half of the risk for developing CRC3,4. Environmental and lifestyle contributions to cancer risk have been broadly categorized and include disparities in cigarette smoking, radiation exposure, stress, exposure to environmental pollutants, diet, obesity, and physical activity (refs 2 and 5 and references therein). Surprisingly, there is no evidence for interaction between CRC susceptibility alleles identified in genome-wide association studies and environmental factors like body mass index, alcohol, smoking, and diet6. However, the interaction between genetic risk factors and physical activity has not been investigated.

Consistent evidence exists for an inverse correlation between the risk for developing cancer and the level of physical activity for many cancer types7-10. A meta-analysis of 52 studies suggested that physical activity reduces the risk of developing CRC by 25%11. Most investigations into the relationship between physical activity and carcinogenesis involve very heterogeneous physical activity exposures that are broadly categorized as occupational or recreational (see ref. 5 and references therein), and, often, essential components (frequency, intensity, duration) of physical activity are not consistently measured. Moreover, when activity parameters are reported utilizing self-assessment recall surveys (often the only feasible method, see ref. 12) considerable measurement error may be introduced13,14. Alternatively, rodent models15 have been effectively utilized in controlled settings to investigate
the relationship between forced or voluntary exercise and growth of transplantable, chemically induced, and/or spontaneous tumors (for a review see refs 16 and 17).

The effect of exercise on tumor growth inhibition in transplantable tumor systems 18, or tumor occurrence reduction in chemically induced or spontaneous tumor models 19, was especially striking in early studies using mice and rats. However, these studies frequently used exhaustive forced exercise protocols (i.e., 18 h of daily exercise in rotating drums, ref. 20) or high-dose inoculations of tumor cells, which limited extrapolation of the findings to humans. Recent mouse and rat studies utilizing more moderate forced and voluntary exercise and inoculation protocols have generally supported earlier findings, with the effect of exercise on cancer incidence and progression being more modest and variable 21–24.

In this report, we investigated the effects of, and interactions between, genetics and voluntary physical activity on azoxymethane (AOM)-induced colon tumor number and size in mice. Additionally, we examined whether the timing of access to exercise relative to AOM exposure influenced carcinogenesis. We also examined exercise parameters (daily running distance, time spent running, average running speed, and maximum running speed) and tumor number and size, at the level of the individual, to elucidate dose response relationships. To identify potential molecular mechanisms we utilized transcriptional analysis of tumor and unaffected adjacent tissue.

Results

AOM 1—Strain by exercise interactions. We investigated the impact of genetic background, exercise, and their interaction on tumor number and size (Fig. 1). As a result of azoxymethane (AOM) administration there was considerable strain variability in mortality rate prior to tumor harvest at 35 weeks. Mortality numbers (as totals and percent of strain) were as follows: CC001/Unc (n = 3, 6.7%), A/J (n = 4, 8.9%), C57BL6/J (n = 19, 42.2%), C58/J (n = 13, 28.9%), I/LNJ (n = 3, 6.7%), KK/HIJ (n = 11, 24.4%). The preceding individuals did not yield tumor data and were censured from analysis. Alternatively, some individuals were sacrificed prior to 35 weeks endpoint due to declining health but still yielded data for tumor number and size. Age at early sacrifice ranged from 19–34 weeks and included individuals from the following strains: CC001/Unc (n = 5), C57BL6/J (n = 2), I/LNJ (n = 1), KK/HIJ (n = 2). A separate set of identical analyses excluding individuals sacrificed prior to 35 weeks yielded results very similar to those presented below. Cancer incidence among all individuals was 99.5% with only one I/LNJ mouse (access to wheels during AOM injections) not developing CRC. For descriptive statistics see Supplemental Tables 2 and 3.
When toxicity is observed with AOM, it is seen acutely due to disruption of the intestinal mucosa and consequent endotoxin effects on the liver. Animals that died acutely were excluded since their tumor response is unknown. Any animal euthanized early for a cancer endpoint (rectal prolapse for example) was included (as noted above). A near 100% tumor incidence in the control group is standard when using the AOM model in susceptible strains. The number of tumors is used as a surrogate for effect on incidence since each initiated tumor arises independent of all other tumors.

Full factorial analysis revealed a statistically significant exercise condition by strain interaction on tumor number \( (F_{10, 191} = 6.051; P < 0.0001) \). Analyses of average tumor size (mm) indicated a statistically significant effect of strain \( (F_{5, 189} = 17.102; P < 0.0001) \), with no effect of exercise condition \( (F_{2, 189} = 2.584; P = 0.078) \). Asterisks denote significant effects of exercise condition within a strain. Bars represent estimated marginal means ± standard errors.

**Figure 2. AOM 1 – Effects of strain and exercise condition on tumor number and size (mm).** General Linear Model revealed a statistically significant interaction between exercise condition [none, 5 weeks during azoxymethane (AOM) injections, 5 weeks following AOM injections] and strain on tumor number \( (F_{10, 191} = 6.051; P < 0.0001) \). Analyses of average tumor size (mm) indicated a statistically significant effect of strain \( (F_{5, 189} = 17.102; P < 0.0001) \), with no effect of exercise condition \( (F_{2, 189} = 2.584; P = 0.078) \). Asterisks denote significant effects of exercise condition within a strain. Bars represent estimated marginal means ± standard errors.

**AOM 2 – Timing of exercise exposure.** We next examined how the timing of exercise (relative to AOM treatment) influenced tumor number and size in both male and female A/J mice (Fig. 1b). Mortality prior to 40 weeks of age was observed in all groups except the group provided 5 weeks of wheel access coinciding with AOM treatment. Mortality numbers by group and sex were as follows: no access to a running wheels \( (n = 7, \text{ males}) \), 5 weeks of access to a running wheel prior to AOM treatment \( (n = 3, \text{ males}; n = 4, \text{ females}) \), 10 weeks of wheel access spanning the 5 weeks prior and 5 weeks during AOM treatment \( (n = 3, \text{ males}) \). The preceding individuals did not yield tumor data and were censured from analysis. Cancer incidence among all individuals was 100%. For descriptive statistics see Supplemental Table 4.

Analysis of tumor number revealed a statistically significant effect of only exercise condition \( (F_{2, 35} = 24.292; P < 0.0001) \) (Fig. 2). Post-hoc test (Tukey HSD) revealed that exercise during the 5 weeks of AOM treatment significantly reduced tumor number and average size relative to other treatments (Fig. 2).

For C57BL6/J, C58/J, I/LNJ, and KK/HIJ, exercise condition had no statistically significant effect on tumor number \( (F_{2, 35} = 7.992; P = 0.001) \) and average tumor size \( (F_{2, 35} = 6.277; P = 0.005) \). Post-hoc tests (Tukey HSD) revealed that exercise during the 5 weeks of AOM treatment significantly reduced tumor number as compared to other treatments (Fig. 2).

For A/J, exercise condition had a statistically significant effect on only tumor number \( (F_{2, 38} = 24.292; P < 0.0001) \). Post-hoc test (Tukey HSD) revealed that exercise during the 5 weeks of AOM treatment significantly reduced tumor number as compared to other treatments (Fig. 2).

For C57BL6/J, C58/J, I/LNJ, and KK/HIJ, exercise condition had no statistically significant effect on tumor number or average size. At the level of the individual, no wheel-running trait (distance run, time spent running, average running speed, or maximum running speed) explained a significant proportion of the variation in tumor number in either wheel access group for any of the strains.
For males, at the level of individual, no wheel running trait explained a significant proportion of the variation in tumor number. For females, mean wheel running distance (Fig. 4a), speed, and maximum speed explained a significant proportion of the variance in tumor number across all three groups (\(R^2 > 0.362\), 
\(R^2 > 0.131\), 
\(P < 0.042\)). Additionally, mean time spent running explained a significant proportion of the variance in tumor number within groups that had access to wheels prior to (\(R^2 = 0.751\), 
\(R^2 = 0.564\), 
\(P = 0.032\)) and during (\(R^2 = 0.666\), 
\(R^2 = 0.444\), 
\(P = 0.018\)) AOM treatment.

**AOM 3–Timing of exercise exposure.** We replicated AOM 2 utilizing only females but changing the time of day for AOM administration (Fig. 1b). Mortality prior to 40 weeks of age was observed in all groups, albeit less than that observed in AOM 2, except for the group provided 5 weeks of wheel access prior to AOM treatment. Mortality numbers were as follows: no access to a running wheels (n = 2), 5 weeks of access to a running during AOM treatment (n = 1), 10 weeks of wheel access spanning the 5 weeks prior and 5 weeks during AOM treatment (n = 1). The preceding individuals did not yield tumor data and were censured from analysis. Cancer incidence among all individuals was 100%. For descriptive statistics see Supplemental Table 5.

Analysis of tumor number revealed a statistically significant effect of exercise condition (\(F_{3, 32} = 6.355\); 
\(P = 0.002\)) (Fig. 3b). Post-hoc tests (Tukey HSD) between exercise conditions indicated that access to a running wheel at any time statistically significantly reduced tumor number relative to no exercise (\(P < 0.03\), for all pairwise comparisons). Bars represent estimated marginal means ± standard errors.

For males, at the level of individual, no wheel running trait explained a significant proportion of the variation in tumor number. However, within the group that had access to wheels prior to the AOM treatment, mean wheel running distance explained a significant proportion of the variance in tumor number (\(R = -0.646\), 
\(R^2 = 0.418\), 
\(P = 0.044\)) (Fig. 4b).
AOM 4—Lasting effects of exercise. In the final experiment we examined if the protective effect of physical activity persists after individuals have stopped exercising (Fig. 1c). Mortality prior to 40 weeks of age was observed in both groups (no access to running wheels, \( n = 2 \); access to running wheels, \( n = 1 \)), and these individuals did not yield tumor data. Cancer incidence among all individuals was 100%. For descriptive statistics see Supplemental Table 6. Analysis of tumor number revealed that 5 weeks of wheel access (5 weeks prior to AOM treatment) had a statistically significant reduction on tumor number (\( F_{1, 35} = 18.524; P < 0.001 \)), but not size (Fig. 5). At the level of individual, the mean amount of wheel running (\( R = -0.572, R^2 = 0.327, P = 0.010 \)) and mean maximal running speed (\( R = -0.589, R^2 = 0.347, P = 0.008 \)) during the final week of wheel access each individually explained a significant proportion of the variance in tumor number (Supplemental Figure 1).

Transcriptional analysis. Hierarchical clustering and principal component analysis strongly indicate large expression differences by tissue followed by strain (Fig. 6). The first principal component (PC), which accounted for approximately 38% of the expression variance, was significantly associated with tissue differences between tumor and normal (ANOVA, \( P = 4.2E-32 \)). PC2 accounted for approximately 12% of the expression variance and was associated with strain differences between A/J and CC001/Unc (ANOVA, \( P = 8.6E-46 \)). In contrast to tissue and strain effects, wheel effects on gene expression were considerably smaller and marginally explained by PC3 (ANOVA, \( P = 0.05 \)).

Differential expression analysis using all 77 samples at FDR 5% detected 20,119 differentially expressed transcripts for tumor vs. normal tissue, 4,833 transcripts for CC001/Unc vs. A/J, none for wheel vs. no wheel, 159 transcripts for tissue by wheel interaction, and none for strain by wheel interaction (Supplemental Table 7). These results are consistent with to hierarchical clustering and PCA plots, where tissue and strain dominated PC1 and
Wheel running distances (in revolutions per day) for both A/J and KK/HIJ mice were very similar (mean ± standard errors). 

We performed functional enrichment analysis for the differentially expressed genes in Supplemental Table 7. Differentially expressed genes for tumor vs. normal at FDR q values (adjusted p-values) < 0.05 were enriched for GO categories of cell cycle, DNA repair, mRNA processing, and other pathways related to cell survival. As expected, genes with strain effect at FDR q < 0.05 were enriched for diverse pathways, but did not form interpretable pathways. Enriched pathways for each factor of interests are listed in a Supplementary File (GO_pathways).

To investigate whether the bulk of CC001/Unc vs. A/J strain effects are attributable to cis eQTL effects, we studied the impact of genetic sequence similarity on expression levels (Supplemental Table 8). Regardless of tissues, transcripts located in regions inherited from non-A/J CC founders were more likely to show differential expression between A/J and CC001/Unc genome-wide (P = 3.7E-35 for normal tissue; P = 3.3E-23 for tumor tissue).

We repeated similar analysis for each chromosome individually. In Chromosomes 2, 8, 17, and 18, we observed an enrichment of transcripts showing strain effect among regions inherited from the other seven CC founders. This is expected because those chromosomes have a higher fraction of A/J inherited regions (Supplemental Figure 3) and hence higher power to detect the strain effect than the other chromosomes.

**Discussion**

We have demonstrated that exercise significantly reduced tumorigenesis and the preventative effect was dependent on genetic background and the timing of physical activity relative to the cancer-inducing agent (azoxymethane, AOM). Furthermore, we have shown that the protective effect of physical activity persists after individuals have stopped exercising. Importantly, we saw no evidence that physical activity was an effective agent (azoxymethane, AOM). Furthermore, we have shown that the protective effect of physical activity persists after the completion of the exposure to AOM. We observed heterogeneity across experiments in the relationship between amount of exercise and the extent of tumor formation (i.e., the dose-response relationship). We discuss these findings in the context of human cohort studies and mouse model investigations into the association between physical activity and CRC risk and prognosis.

Literature examining CRCs and physical activity in humans has yielded variable conclusions as to the efficacy of exercise as a preventative measure for tumorigenesis. For example, Sanchez et al. concluded that one-hour of exercise per week lowered prevalence of polyps and adenomas in a multiethnic population. Conversely, Spence et al. systematically reviewed 20 cohort studies (11 that were “high-quality”) and concluded that evidence was not “sufficient to claim a convincing [sic] relationship exists between PA and CRC risk.” However, the authors acknowledge heterogeneity in the results of the reviewed studies—with 64% of the highest quality studies reporting at least one significant association between physical activity and CRC risk. Additionally, Spence et al. observed a non-significant trend across studies (n = 11) where cancer risk decreased as physical activity increased. It is worth noting that these dose-response relationship examinations are not true measures at the level of the individual (i.e., linear regression or correlational analyses), but rather a categorical comparison of multiple levels of ranges of physical activity (e.g., no activity, “low” activity, “high” activity). These types of broad physical activity exposure definitions (often out of necessity) may contribute to disparities between human cohort findings.

We demonstrated exercise (an altered environment) reduces AOM-induced tumor formation in a strain dependent manner. Among the current strains, the one that was most susceptible in the absence of exercise, A/J, was also the most responsive to exercise as indicated by a nearly 50% reduction in tumor number. The response of KK/HIJ mice compared to A/J epitomizes the importance of gene-by-environment interactions associated with physical activity and cancer risk. When granted wheel access (and the opportunity to exercise) during AOM treatment, tumor number increased substantially in KK/HIJ mice albeit just short of significant (effects of exercise condition, P = 0.055). Exercise quantity (or type) could not explain the opposing effects in these two strains as wheel running distances (in revolutions per day) for both A/J and KK/HIJ mice were very similar (mean ± SE
Figure 6. (a) Array-level, unsupervised descriptive summaries of gene expression using higherarchical clustering. (b) Eigenvalues and projection plots for the first three principal components (PC). Samples were from CC001/Unc and A/J mice provided no access to a running wheel for the duration of the experiment (no wheel) or 5 weeks of access to a running wheel coinciding with AOM treatment (wheel).
were 7274 ± 249 and 7314 ± 362, respectively). This discrepancy between strains in response to exercise may be indicative of the heterogeneity frequently observed between human cohort studies. And, the strain and the strain-by-exercise variation may potentially be mediated through immunological variation between strains and variation in immunoreactivity to wheel running.

It is noteworthy that in AOM 1 exercise reduced tumor number in a strain dependent manner only when the exercise occurred concurrent with AOM treatment. No strain showed a significant reduction in tumor number when exercise was administered following the AOM treatment. This conflicts with previous studies where pre and post inoculation exercise resulted in a marked protective effect. In all our subsequent experiments AOM treatments were administered during or after wheel running, and we found remarkably consistent and highly repeatable results showing that exercise significantly reduced tumor number regardless of the timing. Our findings in A/J mice concurred with Colbert et al., who found negative energy balance induced by voluntary wheel running for 10 weeks significantly inhibited intestinal polypl formation in ApcMin mice. Unique to the current experiments, we demonstrated that 5 weeks of exercise in the 5 weeks prior to AOM treatment resulted in a significant reduction in tumor number. This is an important first step to demonstrating the lasting effects of physical activity in CRC prevention, and may indicate that the positive effect of exercise on reducing cancer risk occurs during the carcinogenesis process, rather than on progression of initiated tumors.

Similar to previous findings in breast cancer murine models, we found a significant negative relationship between mean running distance (during the final weeks of wheel access) and tumor number among individuals during AOM 3 and AOM 4. However, in AOM 2, which was methodologically identical to AOM 3 except for the timing of AOM exposure, we observed a significant positive relationship between tumor number and mean running distance. In an attempt to better understand the inconsistency found in AOM 2, we examined all weeks of wheel running (not just the cumulative amount of running revolutions summed across all days). The significant positive relationship between running distance and tumor number varied by group and week (see Fig. 1B for treatment schedule)—with the group with wheel access before AOM treatment revealing the most consistent positive relationships (weeks 2–5, and total wheel running across all weeks). There were no significant relationships observed within the group granted wheel access during AOM treatment. And, we only observed significant positive relationships during weeks 9 and 10 among the group with access to running wheels before and during AOM treatment. These inconsistencies may potentially be attributable to the overestimation of the duration of continuous running or high levels of inter-individual variability in “coasting” (not measured here but see ref. 35). Although there are differences between experiments at the level of the individual (i.e., correlation between exercise amount and tumor number), which require further investigation, across every experiment we clearly demonstrate that exercise (prior to or during AOM exposure) reduces tumor number at the ‘group’ level.

Based on our gene expression results we preliminarily conclude that the protective effects of wheel running on tumor multiplicity are not mediated through changes in the overall level of expression of genes in the colon. It is possible that the protective effects are due to transcriptional changes that are undetectable by microarray analysis (for example alternative splicing) or to posttranscriptional processes. However, it is more likely that the benefits of exercise in CRC are mediated through systemic changes. For example, Pedersen et al. demonstrated that voluntary wheel running suppressed lung, liver, and skin tumor growth in C57BL/6 and the effects were mediated through epinephrine and IL-6 dependent NK cell mobilization.

In conclusion, our results indicate that voluntary exercise should be used as a preventative measure for colorectal cancer, with the realization that the extent of the protection may depend on genetic background and the timing of the activity relative to the onset of CRC. Our results also indicate that some exercise is better than none, but not necessarily that more exercise is consistently better than less. Nonetheless, these studies should be important in guiding future public health recommendations as this is the first demonstration that genetic background can interact with an environmental exposure (exercise) to modulate CRC risk.

Materials and Methods

Animals and carcinogen treatment. Mice were obtained from The Jackson Laboratory (A/J, C57BL6/J, C58/J, KK/HII, I/LN); selected for known susceptibility to CRC) or bred in house [CC001/Unc, a strain of the Collaborative Cross (CC) that had previously demonstrated high levels of voluntary wheel running]. The CC is a multiparental recombinant inbred panel with A/J being one of the eight founder strains. Mice were housed by gender in groups of four in a viral free facility and maintained in the same room. At approximately eight-weeks of age individuals from each strain were randomly assigned to their respective treatment group. Mice assigned to exercise groups were individually housed during wheel exposure in a room dedicated to collecting activity measures, within the same facility. At the end of the 5 or 10 week wheel exposure mice were regrouped and housed in their original room. Mice were weighed weekly for 5 weeks with intraperitoneal (IP) injections of azoxymethane (AOM, Sigma-Aldrich) at a dose of 10 mg/kg body weight (Fig. 1). This dose has been previously shown to maximize inter-strain differences while minimizing intra-strain variability. AOM exposures were in the mornings for experiments AOM 1, AOM 3, and AOM 4, while exposure was in the afternoon for AOM 2. Following AOM treatment mice were weighed weekly and health status monitored continuously. At the conclusion of the experiments, mice were weighed, sacrificed, and colons removed from the cecum to the rectum and flushed with phosphate-buffered saline (PBS). Tumors were counted and maximum diameters measured utilizing a Leica MZ FL III microscope. This is the most widely used method for tumor size determination in the AOM model. Since the vast majority of tumors are polypoid adenomas, they are largely symmetrical so taking the longest axis provides an appropriate measure of relative tumor size. We did not histopathologically classify tumors, but previous pathology has shown that the AOM model predominantly induces adenomas that when given a promoter progress to carcinoma. Here, we focused on initiation and as such, used an adenoma endpoint. The AOM model is one of the most widely used models of sporadic colon cancer despite being largely adenomas.
Throughout all experiments mice were provided with water and food (Purina PMI pelleted rodent diet) ad libitum. All animal procedures were approved by and are in accordance with guidelines set forth by the Institutional Animal Care and Use Committee at The University of North Carolina at Chapel Hill.

Exercise measurements. For individuals with access to running wheels, daily activity was monitored with Running Wheel Activity Software (AWM V9.2, Lafayette Instruments) via Activity Wheel Counters (model 86061, Lafayette Instruments) interfaced with computers. Wheel-running activity was recorded in 1-min intervals for 23–24 h each day of wheel access. From these recordings, the following daily traits were calculated: total daily revolutions, time spent running (i.e., cumulative 1-min intervals in which at least one revolution was recorded), average speed (total revolutions/time spent running), and maximum speed (highest number of revolutions in any 1-min interval within a 24 h period). We utilized mean values from the final weeks of exposure to wheels to examine relationships between amount/intensity of activity and tumor number and size at the level of the individual. Linear regression analysis was used to examine the relationship between running amount (total daily revolutions and time spent running) and intensity (average running speed, maximum running speed) and tumor number at the level of the individual.

Experimental cohorts. AOM 1 - Strain by exercise interactions. Fifteen female mice from each of the six strains were used in one of three exercise conditions: no access to a running wheel for the duration of the experiment, 5 weeks of access to a running wheel coinciding with AOM treatment, or 5 weeks of wheel access following AOM treatment. Females were initially chosen because they typically run greater distances, run at higher average speeds, and for more minutes per day than males. At sacrifice, a single tumor and unaffected adjacent colon tissue were harvested from each A/J and CC001/Unc mouse in the non-running wheel group and the group given concurrent access to running wheels during AOM exposure for transcriptional analysis. A General Linear Model (GLM) [Univariate GLM ANOVA (SPSS, Chicago, IL)] was utilized to examine the effects of the three exercise conditions and six strains as fixed effects on tumor number and size (mm). We also simultaneously examined interactions between exercise condition and strain. Distributions for tumor number and size were checked for normality prior to and following each analysis to determine if transformation was needed to stabilize variances among groups and/or improve normality of residuals. Statistical significance was judged at P < 0.05, and all P-values presented are two-tailed.

AOM 2 - Timing of exercise exposure. Twelve A/J males and 12 A/J females were randomly assigned to one of four exercise conditions: no access to a running wheel for the duration of the experiment, 5 weeks of access to a running wheel prior to AOM treatment, 5 weeks of wheel access during AOM treatment, or 10 weeks of wheel access spanning the 5 weeks prior and 5 weeks during AOM treatment. Males were included to test for any potential gender or gender by exercise effects. A GLM was utilized to examine the effects of the four exercise conditions and sex as fixed effects on tumor number and size (mm). We also simultaneously examined interactions between exercise condition and sex. Regressions were performed separately for each sex. Within a sex, regressions were performed across all groups and within each of the wheel access treatments.

AOM 3 - Timing of exercise exposure. Because AOM injections in AOM 1 were performed in the morning while those for AOM 2 were performed in the afternoon and resulted in increased acute mortality, we replicated AOM 2 but with morning AOM injections using 10 A/J females assigned to each of the same four treatment groups as described above. All other methods and analyses were the same as for AOM 2.

AOM 4 - Lasting effects of exercise. Twenty A/J females were assigned to one of two exercise conditions: exposed to running wheels for 5 weeks, five weeks prior to AOM treatment or were never exposed to a running wheel. A GLM was utilized to examine the effects of exercise condition on tumor number and size (mm).

Transcriptional analysis. Sample processing and quality control. Gene expression assays were conducted at the University of North Carolina Functional Genomics Core. RNA samples from a single tumor and unaffected adjacent colon tissue from each A/J and CC001/Unc mouse used in AOM 1 were hybridized to an Affymetrix Mouse Gene 2.1 ST 96-Array plate. Array hybridization, washing, staining, and scanning were carried out using the Affymetrix GeneTitan system according to the manufacturer’s protocol. Following our prior reports, we used the robust multiarray average method (RMA) implemented in the Affymetrix gene expression console (default settings, median polish and sketch-quantile normalization) to estimate normalized expression levels of transcripts. We excluded probes containing any SNPs in the A/J and CC001/Unc genomes, control probesets, probesets without mRNA annotation, and lowly expressed probesets [maximum of log, (expression level across samples) < 4]. To evaluate the overall performance of the arrays and identify outliers, we applied principal component analysis (PCA) and hierarchical clustering using the R function hclust with the average link function. The dataset for primary analyses consists of 30,953 probesets and 77 samples (20 A/J mice and 19 CC001/Unc mice) after careful quality control procedures on probe, probesets, and samples. The experimental design is shown in Supplemental Table 1. Samples were partitioned into 8 groups (2 × 2 × 2) for each combination of strain, wheel, and tissue. The sample sizes in each group ranges from 8 to 11.

Differential expression test. We used a linear mixed effect model to jointly analyze all 77 samples, while accounting for the correlation in adjacent tissues of each mouse. Let y be the expression of one probeset in n samples, y = (y₁, y₂, ..., yₙ)ᵀ. The linear mixed effect model can be written as:
$$y_i = \beta_0 + \beta_1 \text{tissue} + \beta_2 \text{strain} + \beta_3 \text{wheel} + \beta_4 \text{tissue} * \text{wheel} + \beta_5 \text{strain} * \text{wheel} + \beta_6 \text{RIN} + Zu + \varepsilon,$$

Fixed effects include tissue (tumor vs. normal), strain (CC001/Unc vs. A/J), wheel (wheel vs. no wheel), two-way interactions, and RNA integrity number (RIN). The random effect ($Zu$) includes a random intercept to account for dependencies between two tissues of each mouse. We also fitted linear fixed effect models within each tissue separately to study strain, wheel, and their interaction effects. The model is:

$$y_i = \beta_0 + \beta_1 \text{strain} + \beta_2 \text{wheel} + \beta_3 \text{strain} * \text{wheel} + \beta_4 \text{RIN} + \varepsilon$$

Within each strain, we used a linear mixed effect model to account for correlation between two tissues of each mouse. The model is:

$$y_i = \beta_0 + \beta_1 \text{tissue} + \beta_2 \text{wheel} + \beta_3 \text{tissue} * \text{wheel} + \beta_4 \text{RIN} + Zu + \varepsilon$$

False discovery rate (FDR) correction was applied to transcript-based $P$-values to correct for multiple statistical comparisons (R package qvalue).44

**Functional clustering analysis.** We used ConsensusPathDB to test differentially expressed genes for enrichment in Gene Ontology (GO) and canonical pathways from the Kyoto Encyclopedia of Genes and Genomes (KEGG), Reactome, WikiPathways, and MouseCyc. For each of the functional categories, a hypergeometric test was performed to examine whether the overlap between our list of genes and those present in each reference category was higher than expected by chance.

**Impact of genomic background on differential expression.** We expect that transcripts located in genetic sequence dissimilarity regions between A/J and CC001/Unc are more likely to be differentially expressed between the two strains due to cis eQTL effects. To test this hypothesis, we classified the genome of CC001/Unc into two categories (i.e., regions inherited from the A/J founder and regions inherited from the other seven CC founders). We also classified all the tested transcripts into two groups (i.e., differentially expressed transcripts or non-differentially expressed transcripts) for each factor of interest. Then, Fisher’s exact test was performed to test whether differentially expressed transcripts were enriched in regions inherited from the other seven CC founders.

**References**

1. Kolonel, L. N., Altshuler, D. & Henderson, B. E. The multiethnic cohort study: exploring genes, lifestyle and cancer risk. *Nat. Rev. Cancer*. 4, 519–527 (2004).

2. Anand, P. et al. Cancer is a preventable disease that requires major lifestyle changes. *Pharm. Res.* 25, 2097–2116 (2008).

3. Lichtenstein, P. et al. Environmental and heritable factors in the causation of cancer–analyses of cohorts of twins from Sweden, Denmark, and Finland. *N. Engl. J. Med.* 334, 78–85 (2000).

4. Risch, N. The genetic epidemiology of cancer: interpreting family and twin studies and their implications for molecular genetic approaches. *Cancer Epidemiol. Biomarkers Prev.* 10, 733–741 (2001).

5. Friedenreich, C. M. & Orenstein, M. R. Physical activity and cancer prevention: etiologic evidence and biological mechanisms. *J. Nutr.* 132, 3456S–3464S (2002).

6. Kantaor, E. D. et al. Gene-environment interaction involving recently identified colorectal cancer susceptibility Loci. *Cancer Epidemiol. Biomarkers Prev.* 23, 1824–1833 (2014).

7. Bergström, A. et al. Physical activity and risk of renal cell cancer. *Int. J. Cancer* 92, 155–157 (2001).

8. Michaud, D. S. et al. Physical activity, obesity, height, and the risk of pancreatic cancer. *J. Natl. Cancer Inst.* 93, 921–929 (2001).

9. Rossing, M. A., Remler, R., Voigt, L. F., Wicklund, K. G. & Daling, J. R. Recreational physical activity and risk of papillary thyroid cancer (United States). *Cancer Causes Control* 12, 881–885 (2001).

10. Shors, A. R., Solomon, C., McTiernan, A. & White, E. Melanoma risk in relation to height, weight, and exercise (United States). *Cancer Causes Control* 12, 599–606 (2001).

11. Wolin, K. Y., Yan, Y., Colditz, G. A. & Lee, I. M. Physical activity and colon cancer prevention: a meta-analysis. *Br. J. Cancer* 100, 611–616 (2009).

12. Kohl, H. W., Fulton, J. E. & Caspersen, C. J. Assessment of physical activity among children and adolescents: A review and synthesis. *Prev. Med.* 31, 54–76 (2000).

13. Shepard, R. J. & Vuillemin, A. Limits to the measurement of habitual physical activity by questionnaires. *Br. J. Sports Med.* 37, 197–206 (2003).

14. Tomaz, S. A., Lambert, E. V., Karpul, D. & Kolbe-Alexander, T. L. Cardiovascular fitness is associated with bias between self-reported and objectively measured physical activity. *Eur. J. Sport Sci.* 16, 149–157 (2016).

15. Uronis, J. M. & Threadgill, D. W. Murine models of colorectal cancer. *Mamm. Genome* 20, 261–268 (2009).

16. Thompson, H. J. Pre-clinical investigations of physical activity and cancer: a brief review and analysis. *Carcinogenesis* 27, 1946–1949 (2006).

17. Washington, M. K. et al. Pathology of rodent models of intestinal cancer: progress report and recommendations. *Gastroenterology* 144, 705–717 (2013).

18. Hoffmann, S. A., Paschke, K. E., DeBias, D. A., Cantarow, A. & Williams, T. L. The influence of exercise on the growth of transplanted rat tumors. *Cancer Res.* 22, 597–599 (1962).

19. Good, R. A. & Fernandes, G. Enhancement of immunologic function and resistance to tumor growth in Balb/c mice by exercise. *Fed. Proc.* 110, 1040 (1981).

20. Rusch, H. P. & Kline, B. E. Exercise effects on growth of mouse tumour. *Cancer Res.* 4, 116–118 (1944).

21. Basterfield, L., Reul, J. M. & Mathers, J. C. Impact of physical activity on intestinal cancer development in mice. *J. Nutr.* 135, 3002S–3028S (2005).

22. Mehl, K. A. et al. Decreased intestinal polyp multiplicity is related to exercise mode and gender in ApcMin/+ mice. *J. Appl. Physiol.* 98, 2129–2125 (2005).

23. Colbert, L. H. et al. Negative energy balance induced by voluntary wheel running inhibits polyp development in APCMin mice. *Carcinogenesis* 27, 2103–2107 (2006).
24. Reddy, B. S., Sugie, S. & Lowenfels, A. Effect of voluntary azoxymethane-induced colon carcinogenesis in male F344 rats. Cancer Res. 48, 7079–7081 (1988).
25. Wahlsten, D. Insensitivity of the analysis of variance to heredity-environment interaction. Behav. Brain Sci. 13, 109–161 (1990).
26. Wahlsten, D. Sample size to detect a planned contrast and a one degree-of-freedom interaction effect. Psychol. Bull. 110, 587–595 (1991).
27. Kelly, S. A. et al. Exercise training effects on hypoxic and hypercapnic ventilator responses in mice selectively bred for increased voluntary wheel running. Exp. Physiol. 99, 403–413 (2014).
28. Sanchez, N. F. et al. Physical activity reduces risk for colon polyps in a multiethnic colorectal cancer screening population. BMC Res. Notes 5, 312 (2012).
29. Spence, R. R., Heesch, K. C. & Brown, W. J. A systematic review of the association between physical activity and colorectal cancer risk. Scand. J. Med. Sci. Sports 19, 764–781 (2009).
30. Clague, J. & Bernstein, L. Physical activity and cancer. Curr. Oncol. Rep. 14, 550–558 (2012).
31. Sellers, R. S., Cliffor, C. B., Treuting, P. M. & Brayton, C. Immunological variation between inbred laboratory mouse strains: points to consider in phenotyping immunomodified mice. Vet. Pathiol. 49, 32–43 (2012).
32. Pedersen, L. et al. Voluntary running suppresses tumor growth through epinephrine and IL-6-dependent NK cell mobilization and redistribution. Cell Metab. 23, 554–562 (2016).
33. Uhlenbruck, G. & Order, U. Can endurance sports stimulate immune mechanisms against cancer and metastasis? Int. J. Sports Med. 12, S53–S68 (1991).
34. Goh, J., Endicott, E. & Ladiges, W. C. Pre-tumor exercise decreases breast cancer in old mice in a distance-dependent manner. Am. J. Cancer Res. 4, 378–384 (2014).
35. Koteja, P., Garland, T. Jr., Sax, J. K., Swallow, J. G. & Carter, P. A. Behaviour of house mice artificially selected for high levels of voluntary wheel running. Animal Behav. 58, 1307–1318 (1999).
36. Pigué, A. C. et al. Regular exercise decreases liver tumors development in hepatocyte-specific PTEN-deficient mice independently of steatosis. J. Hepatol. 62, 1296–1303 (2015).
37. Mathes, W. F. et al. Architecture of energy balance traits in emerging lines of the Collaborative Cross. Am. J. Physiol. Endocrinol. Metab. 300, E1124–E1134 (2011).
38. Collaborative Cross Consortium. The genome architecture of the Collaborative Cross mouse genetic reference population. Genetics 190, 389–401 (2012).
39. Bissahoyo, A. et al. Azoxymethane is a genetic background-dependent colorectal tumor initiator and promoter in mice: effects of dose, route, and diet. Toxicol. Sci. 88, 340–345 (2005).
40. Kelly, S. A. et al. Parent-of-origin effects on voluntary exercise levels and body composition in mice. Physiol. Genomics 40, 111–120 (2010).
41. Sun, W. et al. Transcriptome atlas of mouse brain reveals differential expression across brain regions and genetic backgrounds. G3 2, 203–211 (2012).
42. Crowley, J. J. et al. Analyses of allele-specific gene expression in highly divergent mouse crosses identifies pervasive allelic imbalance. Nat. Genet. 47, 353–360 (2015).
43. Keane, T. M. et al. Mouse genomic variation and its effect on phenotypes and gene regulation. Nature 477, 289–294 (2011).
44. Storey, J. D. & Tibshirani, R. Statistical significance for genomewide studies. Proc. Natl. Acad. Sci. USA 100, 9440–9445 (2003).
45. Kamburov, A. et al. ConsensusPathDB: toward a more complete picture of cell biology. Nucleic Acids Res. 39, D712–D717 (2011).
46. Eviakov, A. V., Dolan, M. E., Genrich, M. P., Patek, E. & Bult, C. J. MouseCyc: a curated biochemical pathways database for the laboratory mouse. Genome Biol. 10, R84 (2009).

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Author Contributions
D.P. and D.W.T. designed research; L.Z., K.-C.J., K.H., and Y.K. performed research; S.A.K., Y.K., and F.P.M. analyzed data; S.A.K. wrote the paper; and all authors critically revised the manuscript.

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