Mitochondria, traditionally known for their role in power (ATP) generation, are important signalling organelles. To support these vital functions, mitochondria need to undergo continuous remodelling - a process known as "mitochondrial dynamics." Failure to do so compromises their function, leading to a spectrum of age-related human diseases, including diabetes, cardiovascular (ischaemia), neuronal (stroke, Parkinson’s and Alzheimer’s) and renal diseases, and cancer. It is generally acknowledged that a rise in oxidative stress, characterised by an increased production of reactive oxygen species (ROS), is responsible for abnormal mitochondrial dynamics. However, how ROS signal abnormal mitochondrial dynamics was unclear.

Mitochondrial dynamics involves continuous fission and fusion of the mitochondrial tubular network. Fission is initiated by the ER-assisted constriction of the mitochondrial tubules followed by their cleavage by Drp1 (dynamin-related protein1) and dynamin 2. Drp1 is normally localised to the cytoplasm where it needs to undergo modifications before it can be recruited to mitochondria. One such modification requires a Ca\(^{2+}\) signal from an undetermined source. By activating calcineurin, Ca\(^{2+}\) removes the inhibitory phosphate group from Drp1, thereby promoting its recruitment to mitochondria. Fission results in the generation of functional and dysfunctional fragments. The latter are removed by mitophagy. Functional mitochondrial fragments merge with the healthy mitochondrial network, with the help of mitofusin-1/2 (MFN-1/2) and OPA-1 (Optic atrophy type 1), which catalyse the fusion of the outer and inner membranes of mitochondria respectively.

Previous studies have shown that free fatty acids (FFAs), whose levels increase in obesity and contribute to type 2 diabetes, cause extensive fragmentation of the mitochondrial network in pancreatic β-cells by stimulating cellular ROS production. In our recent publication, we tested the possibility that the rise in ROS would activate Ca\(^{2+}\) channels to provide the Ca\(^{2+}\) required for Drp1 recruitment to mitochondria. We focussed our attention on TRPM2 (transient receptor potential melastatin2) channels because these are activated by ROS and conduct cations including Ca\(^{2+}\). Consistent with our prediction, inhibition of TRPM2 channels by chemical, siRNA and gene knockout approaches prevented FFA-induced mitochondrial fragmentation as well as β-cell death. ROS required for mitochondrial fission are provided by the FFA-mediated activation of cytoplasmic NADPH-oxidase-2. At first, these findings seemed to support our initial hypothesis that TRPM2 activation provides the Ca\(^{2+}\) required for Drp1 recruitment and the subsequent mitochondrial fragmentation. However, we have decided to test the role of Zn\(^{2+}\) because our previous study demonstrated that Zn\(^{2+}\) chelation prevents TRPM2-mediated β-cell death, and other studies suggested that mitochondrial fragmentation precedes β-cell death. Surprisingly, chelating Zn\(^{2+}\) alone was sufficient to prevent mitochondrial fission. On probing further, we found that free Zn\(^{2+}\)
found largely in the lysosomes was transferred to mitochondria. In a related study, we have demonstrated that TRPM2-mediated Ca\(^{2+}\) entry causes lysosomal membrane permeabilisation and Zn\(^{2+}\) release.\(^7\) Although we do not know how this Zn\(^{2+}\) is transferred to mitochondria, rise in the mitochondrial Zn\(^{2+}\) led to a marked loss of mitochondrial membrane potential, promoting Drp1 recruitment to mitochondria and \(\beta\)-cell apoptosis.

This study has thus led to the discovery of a novel signalling pathway where Ca\(^{2+}\) and Zn\(^{2+}\) collaborate to transduce the signal from FFA to the mitochondria of pancreatic \(\beta\)-cells to cause cell death (Fig. 1). Importantly, the two ions facilitate communication between different cellular compartments. Thus extracellular Ca\(^{2+}\) enters the cytoplasm via ROS-activated plasma membrane TRPM2 channels. The resultant rise in cytosolic Ca\(^{2+}\) triggers escape of lysosomal free Zn\(^{2+}\) to mitochondria. Zn\(^{2+}\), being an inhibitor of the electron transport chain, leads to the loss of mitochondrial membrane potential required for the recruitment cytoplasmic Drp1 to mitochondria. Drp1 then induces excessive mitochondrial fission, leading to increased apoptotic \(\beta\)-cell death.

These new findings raise a number of unanswered questions: (1) How does the rise in cytosolic Ca\(^{2+}\) induce lysosomal Zn\(^{2+}\) release? (2) How does Zn\(^{2+}\) enter mitochondria? Is it via the mitochondrial uniporter (MCU) or other transport molecules? (3) How does Zn\(^{2+}\) promote Drp1 recruitment? Which of the multiple posttranslational mechanisms required for mitochondrial Drp1 recruitment does Zn\(^{2+}\) affect? (4) Does Drp-1 recruitment to mitochondria involve Ca\(^{2+}\) signalling? If so, where does this Ca\(^{2+}\) come from? Interestingly, a recent study suggested Ca\(^{2+}\)-calcineurin signalling occurs at the ER-mitochondria junction.\(^8\) Answers to these key questions are important if we are to understand how the oxidative stress signals are translated into mitochondrial fragmentation, which is a common feature of most age-related human illnesses.

In conclusion, our findings highlight a previously unappreciated role for ionic signalling, in particular a role for Zn\(^{2+}\) signalling, in mitochondrial dynamics. Furthermore, they suggest that ionic signalling is not limited to communication between just two organelles (for example, between the plasma membrane and the ER, and mitochondria and the ER), but could involve multiple organelles. Our findings emphasise the importance of an integrated approach to investigate how abnormal inter-organelle signalling can affect organelle homeostasis and contribute to human diseases. Such approaches may reveal hitherto unknown therapeutic targets that may be common to a wide range of human diseases.

**Disclosure of potential conflicts of interest**

No potential conflicts of interest were disclosed.

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