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Detecting the Effect of Umckaloabo/EPs®7630 Liquid Extract after its Therapeutic Purposed Usage in Calves Showing Symptoms of Respiratory Tract Infection

Ahmet Ak & Mehmet Kale

ABSTRACT

Background: Respiratory tract diseases are commonly seen in beef cattle. Young calves are affected with many respiratory pathogens. Viral pathogens are particularly seen. There are many causative factors, e.g. environmental conditions, immune system of calves. Therefore, alternative treatments are needed for viral respiratory infections. The purpose of the current study was to investigate effectiveness of Umckaloabo/EPs®7630 liquid extract in some bovine viral pathogens of young beef calves.

Materials, Methods & Results: Antibody presence in terms of bovine herpesvirus type 1 (BHV-1), bovine viral diarrhea virus (BVDV), bovine parainfluenza virus type 3 (BPIV-3), bovine respiratory syncytial virus (BRSV) and bovine adenovirus type 3 (BAV-3) was searched in blood serum samples of 40 Holstein calves aged 6 months and over showing respiratory tract infection symptoms. All animals were found seronegative in terms of other factors except BRSV. Out of 20 BRSV seropositive calves, 10 of them were classified as control group and the other ten as testing group. BRSV antibody titers were also detected in blood samples of both groups on day 0. Umckaloabo/EPs®7630 liquid extract was given through oral route to animals in testing group according to their weights for 14 days morning, noon and night. No application was performed on animals in control group. BRSV antibody titers were detected in blood samples of animals in both groups taken on days 1st, 3rd, 5th, 7th, 10th and 14th. At the end of day 14th, BRSV antibody titer increased in 9 out of 10 animals (90%) in testing group that were given Umckaloabo/EPs®7630 liquid extract while one of them (10%) showed no variability. BRSV antibody titer increased in 6 out of 10 animals (60%) in control group while it decreased in one of them (10%) and 3 of them (30%) showed no variability. In testing group, BRSV antibody titer started to increase on day 3rd in 6 out of 10 animals (60%), on day 5th in one (10%), on day 10th in one (10%) and on day 14th in another (10%). In control group, BRSV antibody titer started to increase on day 3rd in 3 out of 10 animals (30%) and on day 5th in 3 of them (30%). When haematologic values of blood samples taken from animals in testing and control groups on day 14th were studied comparatively, no statistical importance ($P < 0.01$ or $P < 0.05$) and difference was detected. As a result, in stock calves showing respiratory tract infection symptoms, applying Umckaloabo/EPs® 7630 liquid extract helped BRSV antibody titer to increase and symptoms decreased. The increase in antibody titers started from day 3rd especially after applying Umckaloabo/EPs® 7630 liquid extract.

Discussion: Respiratory system infections are contagious and fatal in calves. Respiratory tract bacterial and viral vaccinations are applied in order to prevent losses of calves due to these problems. However, these vaccinations are not sufficient most of the time and antibiotics+supportive treatment applications follow afterwards. Therefore, we advise to use a herbal product, Umckaloabo/ EPs®7630 liquid extract, that has both an antiviral efficiency and a feature of supporting immune system in order to prevent mortalities in stock calves, to stop the infection from spreading on all the herd and to decrease the symptoms of respiratory system. We believe that the obtained data should be supported with more examples and more wide-ranging studies.

Keywords: respiratory infection, BRSV, Umckaloabo/ EPs®7630, Pelargonium sidoides, calf.
INTRODUCTION

Respiratory tract problems in cattle settled in plants can be caused by many pathogens [19]. Especially viral pathogens are the most commonly seen factors [20]. BRSV has an important place among pneumonia cases seen in cattle. It is frequently seen especially in calves and calves younger than 1 year. In calves younger than six months, it causes acute interstitial pneumonia proceeding with alveolitis and bronchiolitis [26].

Pelargonium sidoides (PS) is a kind of plant used traditionally by native people in coastal areas of South Africa. The extract from the root of PS is a medicine whose effectiveness has been proven by various fitomedical studies (in vitro, in vivo and clinical). PS contains phenolic and cinnamic acids, tannins, flavonoids and coumarins. Towards the end of the 20th century, an effective and reliable herbal medicine called Umckaloabo was developed to cure acute bronchitis especially in children and to cure infections of upper respiratory tract obstructions [11]. Umckaloabo (EPs®7630) is an ethanolic root extract of PS roots [4] and its antibacterial, antiviral and immunomodulator features have been revealed by many studies and its therapeutic effects were determined.

This medicine is generally used for upper respiratory tract infections in human medicine [2,6]. The medicine was proved to be effective for microorganisms and immune systems [10,13], by every experimental, observational and clinical study performed with PS and its extracts. However, there is no study on its usage in veterinary medicine. In this regard, deeper knowledge and research are needed on the effectiveness of PS and its extracts for diseases both in veterinary and human medicine.

MATERIALS AND METHODS

Animals and properties of the management

In this study, we worked between 2015 and 2016 years in a stock cattle farming plant located in central Burdur Kışla district in Turkey. In this management, we worked on 40 Holstein calves aged 6-8 months old (weighing between 120-200 kg) showing respiratory system symptoms (cough, 40-41°C fever, inappetency, serious nasal flow, drying in nasolabial area, increase in respiratory rate, foamy nasal flow and dyspnea) depending on acute and peracute proceeding pneumonia. Symptoms of animals were determined by plant veterinary surgeon by getting help. The plant the study was applied was semi-outdoor with linoleum ceiling, brick side walls and soil ground. The animals were always in contact in three different sections (120 kg, 150 kg, 180-200 kg) seperated by iron paddocks. There was a feedbox and a waterbowl in each section. Within the plant, adult stock cattle were located in another area 200 meters away. During the study, no practice of vaccination or antibiotics was applied on testing and control group animals used in the study. The fact that the animals had no vaccination for bovine herpesvirus type 1 (BHV-1), bovine viral diarrhea virus (BVDV), bovine parainfluenza virus type 3 (BPIV-3), bovine respiratory syncytial virus (BRSV) and bovine adenovirus type 3 (BAV-3) before the study was confirmed by getting information from plant veterinary surgeon.

Blood sampling of animals to be used in the study

In the study, blood samples taken from v. jugularis of calves showing respiratory system symptoms (those under treatment and control groups) were collected in 10 mL sterile vacuum tubes containing no chemical substances. Blood samplings of BRSV seropositive animals (testing and control groups) were taken on days 1st, 3rd, 5th, 7th, 10th and 14th. Samples were taken to lab with cold chain and centrifuged at 720 g in due form. The obtained serum samples were taken into eppendorf tubes and tested within the same day. Besides, blood samples of the same animals were collected in 8 mL EDTA sterile vacuum tubes on day 14th in order to detect whether there were changes in haematological parameters.

Applying Umckaloabo/ EPs®7630 liquid extract on animals to be used in the study

In the study, Umckaloabo/ EPs®7630 liquid extract Umca solution was used. The compound of this commercial liquid extract includes 80 g of PS root liquid extract as the active substance per 100 g of solution and ethanol and glycerol as dissolver and protector. Commercial introduction of Umca solution includes 20 and 50 mL self-dropping glass bottles.

In our study, the usage of Umca solution in calves was determined by considering the average adult human weight. Around the world, the average adult human weight was found as 62.0 kg (condensed between 58.8 and 74.6) [27] and this figure was ac-
cepted as 60 kg. Accordingly, we applied 60 drops of Umca solution for calves weighing 120 kg, 75 drops for those weighing 150 kg and 90 drops for those weighing between 180-200 kg. Human beings are advised to consume Umca solution by dropping it into some amount of water. In the study, standard amount (100 mL) of sterilized distilled water was used.

In the study, for calves showing respiratory system symptoms and found BRSV seropositive in their blood serum samples, morning, noon and night between days 1st-14th, Umckaloabo/EPs®7630 commercial liquid extract was homogenized in plastic containers and applied through oral route by injectors as 60 drops in 100 mL water for calves weighing 120 kg (Testing 1, Testing 2, Testing 3), 75 drops in 100 mL water for those weighing 150 kg (Testing 4, Testing 5, Testing 6) and 90 drops in 100 mL water for those weighing between 180-200 kg (Testing 7, Testing 8, Testing 9). No treatment application was performed for animals in control group (Control 1-10).

ELISA (blood)-Serological diagnostic method of respiratory tract infections

Respiratory ELISA kit commercial testing product was used in order to detect antibody presence in the plant against BHV-1, BVDV, BRSV, BPIV-3 and BAV-3 in blood serum samples taken on day 1st from 40 stock calves showing respiratory system symptoms. The application was performed according to kit procedure.

ELISA (blood)-Calculating antibody titers with BRSV IgG

Twenty BRSV seropositive calves were classified as 10 for control group and 10 for testing group using Respiratory ELISA kit commercial testing product. In blood samples of both groups taken on days 1st, 3rd, 5th, 7th, 10th and 14th, BRSV titers were detected using BRSV IgG Antibody Test Kit. Blood samples were diluted with sterile saline solution at a rate of ½ according to log<sub>2</sub> base. Except for negative and positive controls, dilutions for each sample were performed within 8 compartments of the plate at rates of 1/2, 1/4, 1/8, 1/16, 1/32, 1/64, 1/128, 1/256. At the end of day 14th, in 9 out of 10 animals in testing group (90%) that were given Umckaloabo/EPs®7630 liquid extract, BRSV antibody titer increased and no change was observed in one of them (10%). In 6 out of 10 animals (60%) in control group, BRSV antibody titer increased, no change was observed in three of them (30%) and it decreased in one of them (10%). In testing group, BRSV antibody titer started to increase in 6 out of 10 animals (60%) on day 3rd, in 1 of them (10%) on day 5th, in 1 of them (10%) for haematological studies.

Statistical analysis

In this study, 2 sample t-test analysis was performed to compare haematological parameters between testing and control groups. The analysis was carried out using Minitab programme. Values smaller than $P < 0.01$ or $P < 0.05$ were statistically considered important.

RESULTS

Respiratory system symptoms of animals

In the study, respiratory system symptoms in 10 Holstein calves within the control group continued and subsided slightly. Respiratory system symptoms in 10 Holstein calves within the testing group subsided and got better. Respiratory system symptoms of animals were determined by the help of plant veterinary surgeon.

ELISA (blood) - Serological diagnostic results of respiratory tract infections

Antibody presence in the plant against BHV-1, BVDV, BRSV, BPIV-3 and BAV-3 in blood serum samples taken on day 0. from 40 stock calves showing respiratory system symptoms was examined. All samples were found as BRSV seropositive while other factors were found seronegative in terms of BHV-1, BVDV, BPIV-3 and BAV-3.

ELISA (blood)-Calculating antibody titers with BRSV IgG in testing and control groups

Twenty BRSV seropositive calves were classified as 10 for testing and 10 for control groups. BRSV titers were detected in blood samples taken from both groups on days 1st, 3rd, 5th, 7th, 10th and 14th (Tables 1 and 2). Blood samples were diluted at a rate of ½ according to log<sub>2</sub> base. Except for negative and positive controls, dilutions for each sample were performed within 8 compartments of the plate at rates of 1/2, 1/4, 1/8, 1/16, 1/32, 1/64, 1/128, 1/256. At the end of day 14th, in 9 out of 10 animals in testing group (90%) that were given Umckaloabo/EPs®7630 liquid extract, BRSV antibody titer increased and no change was observed in one of them (10%). In 6 out of 10 animals (60%) in control group, BRSV antibody titer increased, no change was observed in three of them (30%) and it decreased in one of them (10%). In testing group, BRSV antibody titer started to increase in 6 out of 10 animals (60%) on day 3rd, in 1 of them (10%) on day 5th, in 1 of them (10%)
Table 1. BRSV antibody titer results in the testing group.

| Time (Days) | Testing-1 | Testing-2 | Testing-3 | Testing-4 | Testing-5 | Testing-6 | Testing-7 | Testing-8 | Testing-9 | Testing-10 |
|-------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| 1st         | 1/8       | 1/256     | 1/32      | 1/128     | 1/64      | 1/128     | 1/64      | 1/128     | 1/64      | 1/128     |
| 3rd         | 1/16      | 1/256     | 1/16      | 1/256     | 1/128     | 1/256     | 1/64      | 1/128     | 1/64      | 1/256     |
| 5th         | 1/16      | 1/256     | 1/32      | 1/256     | 1/256     | 1/128     | 1/64      | 1/128     | 1/64      | 1/256     |
| 7th         | 1/64      | 1/256     | 1/32      | 1/256     | 1/256     | 1/128     | 1/64      | 1/128     | 1/64      | 1/256     |
| 10th        | 1/128     | 1/256     | 1/64      | 1/256     | 1/256     | 1/128     | 1/64      | 1/128     | 1/64      | 1/256     |
| 14th        | 1/256     | 1/256     | 1/128     | 1/256     | 1/256     | 1/128     | 1/256     | 1/128     | 1/256     | 1/256     |

Table 2. BRSV antibody titer results in the control group.

| Time (Days) | Control-1 | Control-2 | Control-3 | Control-4 | Control-5 | Control-6 | Control-7 | Control-8 | Control-9 | Control-10 |
|-------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| 1st         | 1/64      | 1/32      | 1/32      | 1/256     | 1/16      | 1/4       | 1/64      | 1/128     | 1/4       | 1/8       |
| 3rd         | 1/64      | 1/64      | 1/32      | 1/256     | 1/32      | 1/4       | 1/64      | 1/128     | 1/8       | 1/8       |
| 5th         | 1/256     | 1/256     | 1/128     | 1/256     | 1/32      | 1/4       | 1/64      | 1/128     | 1/8       | 1/8       |
| 7th         | 1/128     | 1/128     | 1/128     | 1/256     | 1/32      | 1/4       | 1/64      | 1/128     | 1/8       | 1/8       |
| 10th        | 1/128     | 1/128     | 1/128     | 1/256     | 1/32      | 1/4       | 1/64      | 1/128     | 1/8       | 1/8       |
| 14th        | 1/64      | 1/64      | 1/64      | 1/128     | 1/64      | 1/32      | 1/64      | 1/128     | 1/8       | 1/16      |

Haematological parameter results:

When haematological values (WBC, LYM, MID, GRA, LY%, MI%, GR%, RBC, HGB, HCT, MCV, MCH, MCHC, PLT) in blood samples taken on day 14th from animals in testing and control groups were comparatively studied, no statistical importance (P < 0.01 or P < 0.05) or difference was detected between both groups (Tables 3 and 4) (Table 5). In brief, the possibility of averages between both groups to be different from each other was found higher than 1% and 5%.
Table 3. Haematological values of the testing groups (Day 14th).

| No | WBC 4-12x10⁹ cell/µL | LYM 2.5-7.5x10⁹ cell/µL | MID 0-0.84x10⁹ cell/µL | GRA 0.6-6.7x10⁹ cell/µL | LY% 45-75% | MI% 2-7% | GR% 15-65% | RBC 5-10x10¹²/L | HGB 8-15 g/dL | HCT 24-46% | MCV 40-60 µ | MCH 11-17 pg | MCHC 30-36 g/dL | PLT 100-800x10⁹/L |
|----|----------------------|--------------------------|----------------------|--------------------------|------------|---------|-----------|----------------|-------------|-----------|---------|----------|-----------|-------------|----------------|
| T-1 | 10.09 | 3.79 | 0.10 | 6.21 | 37.5 | 1.0 | 61.5 | 9.08 | 8.4 | 26.28 | 29 | 9.2 | 31.9 | 629 |
| T-2 | 11.46 | 3.48 | 1.01 | 6.97 | 30.4 | 8.8 | 60.8 | 8.13 | 8.0 | 24.90 | 31 | 9.8 | 32.1 | 590 |
| T-3 | 9.91 | 6.08 | 0.48 | 3.35 | 61.4 | 4.8 | 33.8 | 9.67 | 9.4 | 29.34 | 30 | 9.7 | 31.9 | 796 |
| T-4 | 8.52 | 3.35 | 0.08 | 5.09 | 39.3 | 1.0 | 59.7 | 8.12 | 8.8 | 28.66 | 35 | 10.9 | 30.8 | 691 |
| T-5 | 10.75 | 4.80 | 0.09 | 5.86 | 44.7 | 0.8 | 54.5 | 9.83 | 9.7 | 30.06 | 31 | 9.8 | 32.2 | 676 |
| T-6 | 7.80 | 4.57 | 0.57 | 2.65 | 58.7 | 7.4 | 34 | 9.53 | 9.8 | 29.55 | 31 | 10.3 | 32.2 | 693 |
| T-7 | 11.28 | 7.44 | 0.11 | 3.73 | 66.0 | 1.0 | 33.1 | 8.20 | 8.6 | 28.08 | 34 | 10.5 | 30.6 | 330 |
| T-8 | 10.11 | 3.63 | 0.08 | 6.40 | 35.9 | 0.8 | 63.3 | 7.83 | 7.8 | 24.56 | 31 | 9.9 | 31.7 | 475 |
| T-9 | 9.37 | 6.60 | 0.53 | 2.24 | 70.5 | 5.6 | 23.9 | 6.62 | 7.7 | 22.96 | 35 | 11.6 | 33.5 | 213 |
| T-10 | 9.73 | 4.05 | 0.22 | 5.46 | 41.6 | 2.3 | 56.1 | 8.69 | 8.1 | 24.41 | 28 | 9.3 | 32.2 | 689 |

T: Testing; WBC: Leucocyte; LYM: Lymphocyte; MID: Monocyte; GRA: Granulocyte; LY%: Lymphocyte%; MI%: Monocyte%; GR%: Granulocyte%; RBC: Erythrocyte; HGB: Hemoglobin; HCT: Hemotocrit; MCV: Average cell volume; MCH: Average cell hemoglobin; MCHC: Average cell hemoglobin concentration; PLT: Platelet.

Table 4. Haematological values of the control groups (Day 14th).

| No | WBC 4-12x10⁹ cell/µL | LYM 2.5-7.5x10⁹ cell/µL | MID 0-0.84x10⁹ cell/µL | GRA 0.6-6.7x10⁹ cell/µL | LY% 45-75% | MI% 2-7% | GR% 15-65% | RBC 5-10x10¹²/L | HGB 8-15 g/dL | HCT 24-46% | MCV 40-60 µ | MCH 11-17 pg | MCHC 30-36 g/dL | PLT 100-800x10⁹/L |
|----|----------------------|--------------------------|----------------------|--------------------------|------------|---------|-----------|----------------|-------------|-----------|---------|----------|-----------|-------------|----------------|
| C-1 | 14.72 | 6.73 | 0.13 | 7.86 | 45.7 | 0.9 | 53.4 | 10.18 | 9.7 | 29.45 | 29 | 9.5 | 32.8 | 748 |
| C-2 | 4.80 | 0.87 | 0.06 | 3.88 | 18.1 | 1.1 | 80.8 | 7.69 | 8.5 | 26.73 | 31 | 11.0 | 31.7 | 730 |
| C-3 | 14.22 | 4.99 | 1.34 | 7.89 | 35.1 | 9.5 | 55.4 | 8.72 | 8.0 | 25.16 | 29 | 9.2 | 31.8 | 820 |
| C-4 | 7.27 | 1.94 | 0.18 | 5.15 | 26.7 | 2.5 | 70.8 | 9.02 | 8.4 | 25.69 | 28 | 9.4 | 32.9 | 505 |
| C-5 | 9.27 | 2.18 | 0.82 | 6.26 | 23.6 | 8.9 | 67.6 | 8.98 | 8.7 | 26.45 | 29 | 9.7 | 32.8 | 588 |
| C-6 | 8.30 | 2.40 | 0.72 | 5.17 | 29.0 | 8.7 | 62.3 | 8.18 | 7.7 | 24.33 | 30 | 9.4 | 31.6 | 793 |
| C-7 | 11.71 | 6.90 | 0.42 | 4.39 | 58.9 | 3.6 | 37.5 | 9.77 | 9.2 | 27.83 | 28 | 9.4 | 32.9 | 803 |
| C-8 | 8.97 | 3.79 | 0.78 | 4.40 | 42.2 | 8.7 | 49.1 | 9.34 | 8.6 | 25.32 | 27 | 9.3 | 34.1 | 351 |
| C-9 | 9.60 | 4.47 | 0.13 | 5.00 | 46.6 | 1.3 | 52.1 | 8.64 | 8.2 | 23.97 | 28 | 9.5 | 34.1 | 728 |
| C-10 | 13.03 | 7.51 | 0.72 | 4.80 | 57.7 | 5.5 | 36.8 | 9.00 | 9.3 | 28.08 | 31 | 10.4 | 33.3 | 504 |

C: Control; WBC: Leucocyte; LYM: Lymphocyte; MID: Monocyte; GRA: Granulocyte; LY%: Lymphocyte%; MI%: Monocyte%; GR%: Granulocyte%; RBC: Erythrocyte; HGB: Hemoglobin; HCT: Hemotocrit; MCV: Average cell volume; MCH: Average cell hemoglobin; MCHC: Average cell hemoglobin concentration; PLT: Platelet.
DISCUSSION

BRSV played the biggest role in respiratory system epidemics in calves in 20 herds located in Middle and Southeast France [24]. Similarly, BRSV and BPIV-3 factors were regarded responsible for respiratory infection epidemics in calves in Ireland [5]. Within naturally developed infections in BRSV endemic areas, the infection mostly develop in calves smaller than 6 months old and adult animals become immune. However, together with virus introduction into areas where BRSV non-endemic and BRSV seronegative adult animals are located, animals of all ages might become infected [8,25].

On our day, when vaccinations against RSV infections are not available, some treatment applications that have moderate or limited efficiency such as palivizumab (monoclonal antibody against RSV fusion protein) and ribavirin (nucleoside analogue) can be performed. That’s why developing a new antiviral medicine against RSV infections is needed. In this regard, some herbal origin natural products as an antiviral activity have been developed. Among these herbs, especially tannin substances inactivate RSV particles and prevent the virus from binding and entering into the cell including fusion stages. Therefore, respiratory system symptoms resulting from RSV, including inflammations in respiratory tracts, can be cured by some natural herbal products [12]. Applications of herbal origin medicine on many respiratory system infections to this day have been performed on humans and experimental animals. About this topic, no study is available on calves and cattle. That’s why our results from the study were mostly evaluated and compared with data obtained from people.

In our study, at the end of day 14th, in 9 out of 10 animals in testing group (90%) that were given Umckaloabo/EPs®7630 liquid extract, BRSV antibody titer increased and no change was observed in one of them (10%). In 6 out of 10 animals (60%) in control group, BRSV antibody titer increased, no change was observed in three of them (30%) and it decreased in one of them (10%). In testing group, BRSV antibody titer started to increase in 6 out of 10 animals (60%) on day 3rd, in 1 of them (10%) on day 5th, in 1 of them (10%) on day 10th and in one of them (10%) on day 14th. In control group, BRSV antibody titer increased in 3 out of 10 animals (30%) on day 3rd and in 3 of them (30%) on day 5th. In case of naturally developed

**Table 5.** Comparing the average haematological values of the testing and the control groups statistically (Day 14th).

|     | WBC | LYM | MID | GRA | LY% | MI% | GR% | RBC | HGB | HCT | MCV | MCH | MCHC | PLT |
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| **No** | 2.5 | 0.61 | 0.15 | 0.61 | 45.1 | 27.7 | 17.1 | 61.1 | 8.5 | 2.4 | 6.9 | 32.9 | 11.3 | 324.1 |
| **Mean** | 7.5 | 3 | 2.3 | 1.5 | 3 | 3 | 3 | 3 | 1 | 1 | 1 | 1 | 1 | 1 |
| **SD** | 1 | 0.3 | 0.5 | 0.7 | 0.2 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 | 0.1 |

In testing group, BRSV antibody titer increased, no change was observed in one of them (10%). In 6 out of 10 animals (60%) in control group, BRSV antibody titer increased, no change was observed in three of them (30%) and it decreased in one of them (10%). In testing group, BRSV antibody titer started to increase in 6 out of 10 animals (60%) on day 3rd, in 1 of them (10%) on day 5th, in 1 of them (10%) on day 10th and in one of them (10%) on day 14th. In control group, BRSV antibody titer increased in 3 out of 10 animals (30%) on day 3rd and in 3 of them (30%) on day 5th. In case of naturally developed infections to this day have been performed on humans and experimental animals. About this topic, no study is available on calves and cattle. That’s why our results from the study were mostly evaluated and compared with data obtained from people.
BRSV infection in calves aged 6-7 months old, IgM and IgG antibody titers increased. In case of natural infections based on the respiratory system developing during acute period especially in calves, IgM and IgG respond improved [28]. PS extracts were confirmed by some testing to have displayed decent activations towards influenza viruses (H1N1 and H3N2), coxsackie A9 virus, human coronavirus, RSV, PI-3, HSV-1 and 2 [15,17,22]. These two research groups [15,22] emphasized that EPs®7630 extract was more active for membranous viruses rather than non-membranous ones. Michaelis et al. [15] searched the effectiveness of EPs®7630 extract in replication of respiratory viruses. In the study, EPs®7630 extract was practised on liquid extract (10 out of 108 patients), placebo group patients (10 out of 109) and changes in leucocyte count appeared in patients using EPs®7630 liquid extract (4 out of 108 patients), placebo group patients (5 out of 109). In this study, when the haematological values (leucocyte, lymphocyte, monocyte, granulocyte, lymphocyte%, monocyte%, granulocyte%, erythrocyte, hemoglobin, hematocrit, average cell volume, average cell hemoglobin, average cell hemoglobin concentration, platelet) in blood samples taken on day 14th (post treatment) from animals in testing and control groups were comparatively examined, no statistical importance (P < 0.01 or P < 0.05) and difference was found between both groups.

An acute cough is usually seen in viral infections (common cold) in respiratory systems. Sore throat, red nose, headache, body pain, fatigue and fever might also exist. Coughing reflections start due to damage on respiratory tract epithelium nerve sensors or oscillation of inflammatory mediators. When Agbabiaka et al. [1] examined 5 studies performed in various years, they performed PS extracts as standard treatment for patients with acute bronchitis. They also included placebo group in the study. They compared a group on which PS extract was applied with another group on which antibiotics and acetylcysteine was not applied and 5 groups on which PS extract was applied with another group on which placebo was applied. In the study, they found that PS was effective in the group with acute bronchitis and acute bronchitis symptoms decreased significantly in these patients on day 7th.

Tahan and Yaman [21] applied PS extract (EPs®7630) for 5 days on 61 children with asthma progressing due to upper respiratory tract viral infections. In the study, fever and muscle pains in the group with PS treatment were statistically (P > 0.05) not found more important than in the group with no treatment and coughing frequency, nasal congestion and asthma attack cases in the group with PS treatment were statistically (P > 0.05) found more important.
than in the group with no treatment. They stated that asthma attacks in the group with PS treatment were seen less frequently.

Viruses, as the primary factor, are usually considered to be responsible for appearing respiratory tract infections and trigger asthma attacks. Increasing respiratory tract inflammations were the sudden reason of asthma attacks and respiratory tract inflammations increasing during viral infections. As a result of decreasing the duration of viral infection efficiency, development of respiratory tract inflammations could be decreased and asthma attacks could be discarded [21]. Bao et al. [3] searched antitussive, secretolytic and anti-inflammatory effects of EPs®7630 in experimental animals. After the study, the coughing frequency of experimental animals with cough decreased significantly depending on the dosage and cough delay period became longer. Secretolytic activity similarly decreased in rats. Lesions in lungs, trachea and bronch tissues caused by acute bacterial bronchitis depending on EPs®7630 in rats regressed histopathologically. As a result, researchers were of the opinion that using EPs®7630 therapeutically in respiratory system infections was effective. For 7 days, Umckaloabo solution was applied as 3x30 drop/day for patients over 18 years of age and showing acute bronchitis symptoms. Especially bronchitis severity score (BSS) [cough, mucus, dyspnea, chest pain while coughing, fever over 38.5°C] was determined for patients. At the end of the study, on day 7, BSS score decreased and individual symptoms decreased more distinctly than placebo group. On the fourth day of treatment applications, 69% of patients and 33% of placebo group showed improvement [18].

Viruses are the most commonly seen factors in most of acute respiratory tract infections. Acute coughing usually accompanies respiratory tract viral infections. Moreover, other symptoms such as sore throat, nasal flow, headache, joint pains, fatigue and fever can be seen [3]. In our study, in 10 Holstein calves in the control group, respiratory system symptoms (cough, fever, inappetency, serous nasal flow, drying on nasolabial area, increase in respiration rate, foamy nasal flow and dyspnea) continued and decreased slightly. In 10 Holstein calves in the testing group, respiratory system symptoms decreased and got better.

PS not only has an antiviral efficiency but also can increase immune response through secretory Ig A, IL-15, IL-6, IFN-γ ve TNF-α and shorten disease period [13]. Effects of PS on mucociliary system is believed to play a role in decreasing coughing frequency. The relation between EPs®7630 and cilia pulse frequency was searched in human cilia nose epithelial cell cultures. At the end of the study, EPs®7630 was thought to have increased cilia pulse frequency significantly depending on the dosage [16]. In conclusion, PS is thought to be effective through many mechanisms such as cilia functions, immunoglobulins, cytokines and chemokines, phagocytic functions, intracellular killing mechanisms and antiviral efficiency, to shorten viral upper respiratory tract infection period and to suppress attack development by preventing inflammatory cases from progressing [7,22].

In this study, by using ELISA (IgG), antibody presence against BRSV was detected in Holstein calves aged 6 months and over showing symptoms of respiratory problems. Out of 20 BRSV seropositive calves, 10 of them were classified as control group and the other ten as testing group. BRSV antibody titers were also detected in blood samples of both groups on day 1st Umckaloabo/EPs®7630 liquid extract was given through oral route to animals in testing group according to their weights for 14 days morning, noon and night. No application was performed on animals in control group. BRSV antibody titers were detected in blood samples of animals in both groups taken on days 1st, 3rd, 5th, 7th, 10th and 14th.

In BRSV seropositive stock calves showing respiratory system symptoms, applying Umckaloabo/EPs®7630 liquid extract helped antibody titer to increase and symptoms (cough, fever, inappetency, serous nasal flow, drying on nasolabial area, increase in respiration rate, foamy nasal flow and dyspnea) decreased. The increase in antibody titers started from day 3th especially after applying Umckaloabo/EPs®7630 liquid extract. Besides, when haematological values in blood samples taken on day 14th from animals in testing and control groups were comparatively studied, no statistical difference was detected between both groups. During clinical observations, no side effect was observed either in testing group on which extract was applied.

**Manufacturers**

1Vacutest. Arzgerande, Italy.
2Labor-Teknik. Istanbul, Turkey.
3Abdi Ibrahim Ilac Pazarlama A.S. Istanbul, Turkey.
4BioX Diagnostics, Pentakit BIO K 028, Belgium.
5IDEXX Laboratories. Saint Denis, France.
6Abacus Junior Vet Hematology Analyzer. Wien, Austria.
7Minitab15 (version15). State College, PA, USA.
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