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Seed germination of a rare neotropical canopy tree dormancy and the effects of abiotic factors
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ABSTRACT – The purpose of this study was to examine if germination is a critical phase on Enterolobium glaziovii regeneration. Hence, the germinative response of E. glaziovii seeds was investigated in relation to some of the main environmental factors (temperature, light and water stress) to which its seeds are subjected in the forest, as well as its dormancy and the longevity of its burial seeds. According to our results, its seeds may be regarded as photoblastic neutral. They do not need alternating temperatures to germinate and can germinate under a broad range of water stress. However, only about 10% of E. glaziovii seeds remain viable after one year. In other words, the annual fruiting, instead seed longevity, seems to maintain the long-term seed availability of this species. Consequently, the seed longevity could be a critical phase of E. glaziovii germination.

Keywords: Ecophysiology, Enterolobium glaziovii and Light.

1. INTRODUCTION

Seed germination is one of the most crucial and decisive phases for a plant species in establishing itself and growing under a set of environmental conditions, such as light, temperature variation, and water availability (FENNER and THOMPSON, 2005). Understanding seed ecology, a broad field encompassing processes associated with seed production, dispersal, predation and germination, are important prerequisites to understanding recruitment and regeneration in plant populations (PEARSON et al., 2002; CERVERA et al., 2006; SOUSA et al., 2008). Seed ecology provides an essential knowledge of population dynamics regarding the establishment of new individuals, and therefore the funding of new populations (REES, 1996; GODÍNEZ-ALVAREZ et al., 1999; VALVERDE et al. 2004). The analysis of the species’ features that determine their distribution, abundance and habitat specificity has long been the core of ecological research (KREBS, 1994). These ecological aspects determine a species’
level and type of rarity (RAWINOWITZ, 1981). This subject is considered of great relevance in the context of conservation biology, because a large number of endangered species are naturally rare and therefore more prone to decline and extinction than relatively more common ones (GASTON, 1994). A species is considered rare when its populations are biologically viable but naturally sparse, frequently limited in its distribution range and/or occupying specific habitats (RAMÍREZ-PADILLA and VALVERDE, 2005).

The reproductive biology and the requirements for seed germination may play an important role as causal factors for limited abundance or restricted distribution ranges (RAMÍREZ-PADILLA and VALVERDE, 2005). Seed germination responses have a direct impact on a species’ distribution and abundance, since it is a key element affecting population dynamics (GODOINEZ-ALVAREZ et al., 1999; VALVERDE et al., 2004, RAMÍREZ-PADILLA & VALVERDE, 2005).

Enterolobium glaziovii Bentham (Mimosaceae) is a rare canopy tree (Ramos et al. 2005a,b), endemic of the Brazilian Coastal Atlantic forests. This species occurs at low density, however, neither the growth nor survival of E. glaziovii offspring around conspecific adult trees are critical phases of its regeneration (Ramos et al, 2005a,b). Germination could be another possible phase that restrains regeneration of E. glaziovii. Knowledge of a critical factor that could inhibit or favor E. glaziovii germination may be important to conservation programs. Hence, the germinative response of E. glaziovii seeds was investigated in relation to some of the main environmental factors, including temperature, light and water stress, to which its seeds are subjected in the forest, as well as the dormancy and longevity of its burial seeds.

The purpose of this study was to study the germination responses to different factors of Enterolobium glaziovii (Benth.) Mesquita under controlled conditions to evaluate if their germination responses to these experimental factors are related to its rarity. The specific questions are: (1) Is seed coat responsible for seed dormancy?; (2) How do patterns of seed longevity in the soil vary?; (3) Are seeds capable of germinating in darkness?; (4) Are seeds capable of germinating under different temperatures? and (5) Are their seeds capable of germinating under different drought intensity?

2. MATERIALS AND METHODS

Mature fruit of E. glaziovii were collected on the floor (just after seed rain) under the only three individuals known at the Biological Reserve of Poco das Antas, (22,29' - 22,36'S, 42,13' - 42,21'W) Rio de Janeiro, SE, Brazil (RAMOS et al. 2005 a, b). The experiments were carried out in the Seed Laboratory of the Rio de Janeiro Botanical Garden after the seeds had been taken from the fruits. To prevent fungal infection, seeds were previously treated with sodium hypochlorite (10% v/v; Vetec Inc., Brazil) for 10 minutes, followed by rinsing three times in distilled water being stored at 15 °C in darkness, for periods ranging from 3 to 10 days, without any drying treatment. Seeds were selected eliminating those infested by insects or deteriorated.

2.1. Fruit and seed characteristics

Non-moldy and insect-free fruit were used to determine the mass of fruit and seeds, the seed / fruit ratio, as well as the fruit and seed size with the aid of a calliper. Seed water content was determined using five replicates of five seeds cut in quarters, using a dry oven at 105°C for 24 hours according to Ramos et al. (2000).

2.2. Germination tests

Germination tests were performed in transparent plastic boxes (gerbox) filled with heat-sterilized vermiculite, under temperature and light control chambers, with temperature kept constant within ± 1 °C and, unless light was an intended variable, an 8h light/16 h darkness regime (4 x 20W white fluorescent tubes; total fluence rate 90mmol m-2 s-1). For the water stress experiments, two filter papers wet in distilled water or osmotic solution were used instead of vermiculite. For all experiments, percentage values are means of four replicates of 25 seeds.

Number of germinated seeds was recorded three times a week, until all the seeds germinated or died. The criterion for germination was visible radicle protrusion. Seeds showing external signs of rotting were considered dead.

2.2.1 Seed dormancy

Scarified and non-scarified seeds were germinated at 25 °C. Seeds were scarified in the opposite side of the radicle, with wood sandpaper.
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2.2.2 Effect of temperature

Scarified seeds were germinated under constant (20, 25, 30 and 35 °C) and alternating temperatures (20-30 °C and 20-35 °C) subjected to a 8/16 h thermoperiod, according to Andrade et al. (2006).

2.2.3 Effect of light treatment regime

Scarified seeds were germinated at 25 °C, in black and transparent plastic boxes. Seed germination recordings were carried out in a dark room using green safety light.

2.2.4 Effect of the water stress

Scarified seeds were germinated at 25 °C, in gerbox moistened with different osmotic potentials. Aqueous (distilled water) solutions of mannitol mixed after Nobel (2005) were used to exert osmotic potentials (γ) of -0.2, -0.4, -0.8, -1.2, -1.6; -1.8; and -2.0 MPa. In order to avoid evaporation losses, the plastic boxes were sealed with adhesive tape.

2.2.5 Seed longevity

Non scarified seeds were placed with vermiculite into small nylon mesh bags (mesh size 1 mm) and they were buried at 3 cm below soil surface under the shade of an E. glaziovii adult tree. During 24 months, the bags were exhumed every 3 months, and the seeds were germinated at 25 °C, at the laboratory. At the end of the experiment, the seeds that did not germinate were cut open for examination using the tetrazolium test whether or not the non-germinated seeds had lost their viability or failed to germinate.

2.3. Analysis

The rate of seed germination was estimated using the Index of Germination Speed (IGV) (LABOURIAU, 1970):

\[
IGV = \frac{1}{t} = \sum_{i=1}^{n_i} \frac{t_i}{t_i - 1},
\]

where \(t_i\) is the number of days between the beginning of the experiment and the \(i\)th observation, and \(n_i\) is the number of seeds germinated within the time interval \(t_{i+1} - t_i\). Vertical lines and ± values represent one standard error.

Data were subjected to one-way ANOVA, two-way ANOVA or to t test after the arcsin transformation. Tukey's multiple comparison procedure was used to compare significant differences between treatment means (ZAR, 1996). Whenever the result of a replicate was zero or 100% the relative values of 1/4\(n\) or 1 - 1/4\(n\) were used, respectively, to calculate proportions before transformation (\(n\) is the number of seeds per replicate; ZAR, 1996). Throughout, results in which P < 0.05 are reported as significant.

3. RESULTS

3.1. Fruit and seed characteristics

The E. glaziovii fruit are indehiscent, black, contorted woody, large (4.3 - 8.9 cm x 3.0 – 4.4 cm) and heavy (14.6 g ± 6.4). They contain 9.4 (± 3.1) seeds per fruit. The seeds were 0.5 - 1.6 cm x 0.5 – 0.6 cm, 0.1 g (± 0.02) and showed a 14.9% (± 0.8) water content. No record of seed predation inside the fallen fruit was obtained.

3.2. Germination test

3.2.1 Seed dormancy

The hard seed coat influenced E. glaziovii seed germination. Scarified seeds present a higher germination percentage (T6 = 10.2; \(P = 0.001\)) and germination rate (T3.6 = 4.9; \(P = 0.01\)) than non-scarified ones (Table 1).

3.2.2 Effect of temperature

E. glaziovii seeds did not need alternating temperatures to germinate. There was no difference in germination percentage of scarified seeds at constant

| Table 1 – Means (± SD) of germination percentage (Germ) and rate (IGV) of non-scarified and scarified seeds; and of seeds germinated under different light regimes. Similar letters (upper-case for dormancy treatment, lower-case for light treatment) do not differ by t test. Both non-scarified and scarified seeds were incubated in light, while for the light/dark experiment all seeds were scarified. |
|---------------------------------|
| Germ (%) | Non-scarified | Scarified | Light | Darkness |
| 12 (± 5.6) A | 71 (± 8.9) B | 82 (± 4.0) a | 81 (± 5.0) a |
| IGV (d⁻¹) | 0.12 (± 0.03) A | 0.20 (± 0.01) B | 0.17 (± 0.02) a | 0.18 (± 0.01) a |

E. glaziovii seeds did not need alternating temperatures to germinate. There was no difference in germination percentage of scarified seeds at constant
and alternating temperatures ($F_{5,18} = 0.25; P = 0.93$) (Figure 1), nevertheless germination rate was highest at 30 °C and lowest at 20 °C ($F_{5,18} = 7.26; P = 0.001$).

3.2.3 Effect of the light regime

Scarified seeds germinated at light and darkness did not present significant difference neither in its germination percentage ($T_{6} = 0.28; P = 0.79$), or rate ($T_{6} = 0.65; P = 0.54$) (Table 1).

3.2.4 Effect of the water stress

Although the germination percentage decreased with lower osmotic potentials ($F_{7,24} = 7.7; P = 0.001$), scarified seeds germinated at all osmotic treatments (Figure 2), even as strongly negative as – 2.0 MPa.

3.2.5 Seed longevity

$E. glaziovii$ seeds remain viable after almost two years (Figure 3). Although more than 50% of them lost their viability in the first 6 months, and only about 10% of them remain viable after one year ($F_{7,24} = 72.8; P = 0.001$).

4. DISCUSSION

As well as other Enterolobium species (EIRA et al. 1993, LIMA et al. 1997, MALAVASI and MALAVASI, 2004), seeds of $E. glaziovii$ present hard coats. Species with hard seed coats are common in many species of Leguminosae (BASKIN and BASKIN, 2001, 2004) tree species that produced these types of seeds and generally have low moisture content, high longevity, and take longer to germinate, like $E. glaziovii$ (VAZQUEZ- YANES and OROZCO-SEGOVIA, 1984).

Following the dispersal of seeds in late autumn/early winter, a majority of $E. glaziovii$ seeds are dormant. When dormancy is broken, some seeds may germinate. In the laboratory, dormancy of $E. glaziovii$ seeds could be overcome by mechanical scarification. This suggests that, in nature, seed dormancy may be ameliorated with mechanical abrasion, passage through the digestive tract of animals (BASKIN and BASKIN 2000), by fire or by microbial attack (BASKIN and BASKIN, 2000; TAYLOR, 2005) and, in some case, by high temperatures and thermal fluctuation.
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E. glaziovii seeds are also tolerant to a wide range of water stress. Water deficit could inhibit seed germination of many tree species, because water is the initial factor for seed germination and is directly and indirectly involved with subsequent germination metabolic stages (CAVALCANTE & PEREZ, 1995). Certain species were able to germinate at low water potentials. The range of water stress tolerance is different, among leguminous species. While some of them present lower tolerance, such as Acacia tortilis that did not germinate in osmotic potential below -0.6 MPa (CHOINSKI AND TUOHY, 1991), other species present higher tolerance, such as bean seeds (-1.2 MPa, MACHADO NETO et al., 2006), Leucaena leucocephala (-1.4 MPa, CAVALCANTE AND PEREZ, 1995), Indigofera astragalina (-1.5 MPa, SY et al., 2001), Enterolobium contortisiliquum (-1.6 MPa, HEBLING AND PEREZ, 1997), Prosopis juliflora (-1.9 MPa, PEREZ AND MORAES, 1991) and E. glaziovii (-2.0 MPa, present study). Since its germination is spread along the year, this tolerance could be very important to the E. glaziovii regeneration, because its seeds will be able to germinate even at times with low soil water potential. The replacement of osmotic liquids by distilled water did not change the pattern found in water stress, because the non-germinated seeds had lost their viability in each osmotic potential (data not showed).

In summary, E. glaziovii seeds are disseminated with a hard coat that inhibits water imbibition. Abrasion due to scarification of the E. glaziovii seed coat seems to be the most important requirement to its germination. Thus, seeds probably may not germinate readily in nature until the seed coats are made permeable by frugivorous vertebrates or weathering. Its seeds may be regarded as photoblastic neutral, so they do not need alternating temperatures to germinate and can germinate under a broad range of water stress. Due to its hard coat, E. glaziovii germination could be temporally distributed within a year, but could not maintain viable seeds in a soil seed bank for long periods. In other words, the annual fruiting, instead of seed longevity, seems to maintain the long-term seed availability of E. glaziovii on the seed bank. Consequently, the seed longevity could be a critical phase of E. glaziovii germination.
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