INTRODUCTION

Alzheimer’s disease (AD) was discovered in 1906 by Alois Alzheimer, a German brain scientist. The sickness was initially ascertained during a 51-year-old lady named Auguste D. Her family brought her to Dr. Alzheimer in 1901 once noticing changes in her temperament and behavior. Dr. Alzheimer represented Auguste as having associate degree aggressive variety of dementedness, manifesting in memory, language, and activity deficits [1]. There are two core kinds of diseases. Familial AD affects folks younger than 65, accounting for nearly 500,000 AD cases within the US alone. The rest of the AD cases occur in adults aged 65 and older and are classed as sporadic [2]. AD caused by genetic and environmental factors. Symptoms of AD include difficulty with speech, agitation, confusion, difficulty in retaining new memory, and loss of ability to communicate [3,4]. The main changes that take place in the brain are neurofibrillary tangle and beta-amyloid deposition. Beta-amyloid is an associate degree insoluble, cyanogenic peptide macromolecule of concerning 39–43 amino acids. Beta-amyloid accumulates into plaques into oligomers. These insoluble proteins clusters are found within the neural area of the brains of AD patients and block nerve cells [5]. The accumulation of neurofibrillary tangles (NFT) among neurons is because of hyperphosphorylated tau proteins. An abnormal number of additional phosphates are added to the tau protein, causing the tau proteins to release from the microtubules, inflicting the tau proteins to unharvested from the microtubules. These hyperphosphorylated proteins bind with each other and type tangles among the cell, referred to as NFT [6].

Currently, there is no cure for AD, the drugs currently used are tarecine hydrochloride (Cognex) and donepezil hydrochloride (Aricept), rivastigmine (exelon), and galantamine (reminyl) [7]. Many plants useful for the treatment of anti-Alzheimer’s in Ayurvedic system of medicine have been tested for their anti-Alzheimer effect in experimental animals. In plant phytochemicals can shield hominid useful for the treatment of anti-Alzheimer’s in Ayurvedic system of medicine. Plants produce these substances to save themself; however, fresh examination determines that various phytochemicals can guard creatures besides syndromes. There are several phytochemicals in pods and sages and each mechanism contrarily [8]. In the modern years, pioneers are more fascinated to shrub beginning medications as they are greatly biocompatible with minor side effects than the human-made medications. The ordinary yield and class are not reasonable due to slow development degree, overexploitation, and environment demolition which are possibly the key drawbacks to see the ever-growing marketplace request. Additional, farming of these rough therapeutically significant plants lacks satisfactory illness-free elite implanting resources due to great vulnerability of the crop for rhizome rot, leaf spot, and microbial wilt [9]. Costus pictus D. Don referred to as internal secretion plant or Painted Spiral Ginger. It is a category of the herb in Costaceae family native to the United Mexican States. Bhattacharya et al. (1972) confirmed that C. speciosus alkaloids possess anticholin activity in vitro and in vivo. Thus, C. pictus D. Don additionally possesses the anticholinesterase activity and they decrease the breakdown of acetylcholine so may help in treating Alzheimer diseases [10].

METHODS

Plant description

Leaf

The massive, smooth, simple kind, alternate organized, rectangular-shaped, and green color leaves have light-weight purple undersides and unit spirally organized around stems, forming partaking, and bowed clumps arising from underground rootstocks.

Flower

The plant flowers are of beautiful orange color, are created among the warmth months. Flower petals are quite sweet and nutritious. The plant to boot bears fruit, but its type is unknown.

Keywords: Alzheimer diseases, Costus pictus D. Don, Acetylcholinesterase, Scopolamine, Neurofibrillary tangles, Extract of Costus pictus D. Don.

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Plant

It reaches to relating to two feet tall, with the tallest stems falling over and lying on the lowest. The plant grows in partially shade/partially sun and it wants generally wet, clay, and sand. Costus does not have a disadvantage with pests and illness [11].

Growth

The spiral flag grows in either full sun or partial shade. It desires fertile soil and is typically planted on the brink of the water. Propagation is by the division of the clumps, cuttings, or by separating the offsets. That sort below the flower heads. Mites and nematodes are typically a retardant, significantly on light-weight, sandy soil. The plant has no diseases that are of major concern [12].

Chemical constituent

Pictus leaves disclosed that it is wealthy in macromolecule, iron, and inhibitor elements such as antioxidant, α-tocopherol, β carotene, terpenoids, steroids, and flavonoids. It was disclosed in another study that ethanolic extract was found to contain the very best range such as carbohydrates, triterpenoids, proteins, alkaloids, tannins, saponins, and flavonoids. Flavonoid and phenolic resin compound enamplifies a neuroprotective impact [13,14].

Medicinal properties

It possesses antimicrobial activity [15], hypoglycemic activity [16], ant-fertility activity [17], hyperlipidemic activity [18], ameliorative effect [19], diuretic effect [20], and putative activity [21].

Toxicity

The plant is usually eaten by animals that are additionally associate indicator to prove it’s less cyanogetic. In the literature survey, it had been found that the ethanolic extract of C. pictus D. Don was safe [22,23]. LD₅₀ of the ethanolic extract is according to be 2000 mg/kg. Once literature surveys of varied analysis papers, the ultimate doses for the study 100, 150, and 250 mg/kg were selected [24].

Plant material and extraction

The plant (C. pictus D. Don) specimens for the proposed study were collected from the medicinal garden of Oriental College Of Pharmacy. A sample specimen voucher was submitted to Dr. Rajendra. D. Shinde, Director, Blatter Herbarium of St. Xavier’s College, Mumbai. The leaves were washed with tap water and shade dried at normal room temperature with the aid of circulating airflow using fan. The continuous hot extraction methodology was used for getting plant extracts. The dried powder of C. pictus D. Don (150 g) was extract with 700 ml of ethanol using Soxhlet apparatus. The collected extract is evaporated to remove the ethanol rotary vacuum evaportor. The dried herbal extract is mixed with carboxy methylkellulose (CMC) and administered to the animals.

Animals

Male albino Wistar rats weighing 250 g–386 g were used in this study. Animals were housed in plastic cages in seven groups. They had free access to food and water, and they were kept in a regulated environment (23°C, 40–60 humidity). Experiments were carried out between 9:00 a.m. and 7:00 p.m., in an experimental room with in the animal facility. All animal procedures were conducted in strict under the rules of the ethical committee.

Equipment and chemicals

Electronic balance, elevated plus Maze, Y-maze, novel object apparatus, syringes, and needles, ethanol, CMC, and scopolamine. Donepezil tablet, 5,5-dithiobis-(2-nitrobenzoic acid) (DTNB), acetylcholine, thio-barbituric acid (TBA), Nicotinamide adenine dinucleotide phosphate (NADP), and NADPH. Sodium bicarbonate glutathione reductase, and sodium azide.

Preliminary phytochemical analysis [25]

Test for carbohydrate

Molisch test

The test solution is combined with a small amount of Molisch’s reagent (α-naphthol dissolved in ethanol) in a test tube. After mixing, a small amount of concentrated sulfuric acid is slowly added down the sides of the sloping test-tube, without mixing, to form a layer. A positive reaction is indicated by the appearance of a purple-red ring at the interface between the acid and test layers.

Fehlings test

In a test tube, add 2 ml of the test solution and add equal volumes of Fehling A and Fehling B and place it in a boiling water bath for few minutes. When the content of the test tube comes to boiling, mix them together and observe any change in color or precipitate formation.

Protein test

Ninhydrin test: Ninhydrin is the most commonly used method to detect fingerprints, as the terminal amines or lysine residues in peptides and proteins sloughed off in fingerprints react with ninhydrin. To 3 ml of the test solution add 3 drops of 5% ninhydrine solution—boil over a water bath for 10 min purple bluish color appears.

Steroid test

Liebermann-Burchard test: The extracts were evaporated to dryness and the residues were extracted with petroleum ether and acetone. The insoluble residues left after extraction were dissolved in chloroform, and few drops of acetic anhydride were added along with a few drops of concentrated sulfuric acid from the side of the tube. The appearance of blue to blood red color indicates the presence of sterols in the extracts.

Glycoside test

Killer-kiliani test: 1 ml of glacial acetic acid containing traces of ReCl₃, and 1 ml of concentrated H₂SO₄ were added to the extracts carefully. A reddish-brown color is formed at the junction of two-layer and the upper layer turns bluish-green in the presence of glycosides.

Saponin test

Foam test: The extract (2 g) was shaken vigorously with 20 ml of water and observed for persistent foam, which indicates the presence of saponins.

Test for flavonoids and phenol

Shinoda test: To the dry extract (2 g), 5 ml of ethanol (95% v/v), 5 drops of hydrochloric acid, and 0.5 g of magnesium turnings were added. The appearance of pink-red purple color indicates the presence of flavonoids.
Test for phenol
0.50 g of phenol and 0.5 g of extract dissolved in 5 ml of distilled water. 
Add a few drops of ferric chloride solution dark green color indicates the presence of phenol.

Test for tannins
Ferric chloride test: To 3 ml of extract, 3 ml of 5% w/v ferric chloride solution was added. The blue-black color indicates the presence of tannins.

Lead acetate solution
2 ml ethanolic extract of C. pictus D. Don leaves was treated with few drops of lead acetate solution. White precipitate indicates the presence of tannins.

Test for triterpenoids
Salkowski test: To the 2 ml ethanol extract of C. pictus D. Don, 2 ml chloroform and 2 ml concentrated H₂SO₄ were added and shaken. The presence of red color in the chloroform layer and greenish-yellow fluorescence in the acid layer indicates the presence of steroids.

Test for alkaloids
Mayer's test: To 3 ml of filtrate, 1 ml of Mayer's reagent (potassium mercuric iodide) was added. The appearance of a white precipitate indicates the presence of alkaloids.

Wagner's test: To 3 ml of filtrate, 1 ml of Wagner's reagent (iodine in potassium iodide) was added. The appearance of reddish-brown precipitate indicates the presence of alkaloids.

Hager's test: To 3 ml of filtrate, 1 ml of Hager's reagent (saturated picric acid solution) was added. The appearance of yellow precipitate indicates the presence of alkaloids.

Experimental design
Grouping of Animals: Animals were divided into seven groups, each of six animals.

- Group I: Control group oral administered by saline
- Group II: Intraperitoneal injection by scopolamine hydrochloride (0.5 mg/kg) (Negative control)
- Group III: Animal oral administered by donepezil in CMC (250 mg/kg) and Alzheimer’s induced with scopolamine
- Group IV: Animals oral administered by extract which is dissolved in CMC (100 mg/kg) and Alzheimer’s induced with scopolamine
- Group V: Animals oral administered by extract which is dissolved in CMC (150 mg/kg) and Alzheimer’s induced with scopolamine
- Group VI: Animals oral administered by extract which is dissolved in CMC (250 mg/kg) and Alzheimer’s induced with scopolamine
- Group VII: Animals oral administered by extract which is dissolved in CMC (250 mg/kg) and Alzheimer’s induced with scopolamine

\[ \text{Retention part. On a primary day (habituation phase), rats were on an individual basis subject to one familiarization session of 10 min, throughout that they will be introduced within the empty space, so as to become familiar with the environment.} \]

On an ordinal day (acquisition phase), animals were subjected to one 10-min session, throughout that floor-fixed two objects (A and B) placed in an exceeding rhombohedral position within the central line of the world. 10 cm from each other and 8 cm from the closest wall (each object occupies roughly 5 cm area by its size).

The two objects, manufactured from an equivalent material with the similar color and smell, were completely different in form however identical in size. Rats were allowed to explore the objects within the open field. The exploration time on every object is shown (as seconds) to point the exploring activity of rats.

On the 3rd day (retention phase), rats are going to be allowed to explore the open field within the presence of two objects: The familiar object A and a completely unique object C in numerous shapes however in similar color and size (A and C). A recognition index \( RI \) (for retention session), calculated for each mouse, was expressed as \( RI = \frac{\text{Time exploring novel object} - \text{Time exploring familiar object}}{\text{Time exploring familiar object} + \text{Time exploring novel object}} \times 100\% \)

Elevated plus maze
Elevated plus maze function the sensibility behavior model to gauge learning and memory in rodents. The procedure, technique, and finish purpose for testing, learning and memory are followed as per the parameters represented by the investigators operating within the space.
of neuropsychopharmacology. The equipment consists of two open arms and two boxed arms the arms area unit extended from a central platform and therefore the maze is elevated to a height of 25 cm from the ground. On the 14th day, every mouse was placed at the top of an open arm, facing far from the central platform. Transfer latency (TL) is that the time taken by mouse with all its four legs to maneuver into of the boxed arms that are recorded on the primary day. If the animal did not enter into one of the enclosed arms within 90 s, its gently pushed into one of the two enclosed arms and the TL will be assigned as 90 s.

The mouse will be allowed to explore the maze for another 10 s and then returned to its home cage. Retention of this learned task will be examined 24 h after the 1st day trial (15th day). The inflexion ratio (IR) was calculated by the formula IR = Lo – Lt/Lo, where Lo is initial TL on 1st day and Lt is TL after 24 h. Decrease IR indicates the induction of amnesia, and increased IR indicates an improvement in cognition and memory impairment [27,28].

Y -maze spontaneous alternation could be an activity check for in the prosencephalon, and anterior cortex, are concerned during this task.

Rodents generally opt to investigate a brand new arm of the maze instead of returning to one that was antecedent visited. Several components of the brain, including the hippocampus, septum, basal prosencephalon, and anterior cortex, are concerned during this task. Y-maze created of black painted wood. Every arm is 40 cm long, 12 cm height, 3 cm wide at an all-time low and 10 cm wide at the highest and converged in an equal triangular central space. Each mouse was placed at the tip of one arm associated allowed to maneuver freely through the maze throughout an eight min session. The flexibility to alternate needs that the mice understand that arm they need already visited. The series of arm entries, as well as attainable returns into an equivalent arm, were recorded visually. Immediate memory performance is going to be assessed by recording spontaneous alternation behavior. Entry was going to be thought of to be completed once the hind paws of the mouse had fully entered the arm. Alternation was outlined as consecutive entries into the three totally different arms (A, B, and C) on overlapping triplet sets. The share of trials within which all three arms were drawn, i.e., ABC, CAB, or BCA however not BAB, was recorded as associate in nursing alternations’ to estimate remembering. On the 19th day, 90 min when the treatment of last dose arm entries was recorded and share alteration are going to be calculated [29] % Alteration = (No of alteration/total arm entry-2)².

Biochemical test

Preparation of brain sample

After assessing the learning and memory paradigms in scopolamine-induced amnesia, rats from each group were euthanized using a carbon dioxide chamber; brains were removed quickly and placed in ice-cold saline. Frontal cortex, hippocampus, and septum (and any other regions of interest) were quickly dissected out on a Petri dish chilled on crushed ice. The tissues were weighed and homogenized in 0.1M phosphate buffer (pH 8). The samples of mice brain homogenates were collected in a different test. The supernatant was used for enzymatic assays.

Estimation acetylcholinesterase enzyme levels in the brain

0.4 ml aliquot of the material was added to a cuvette containing a pair of 6 cc phosphate buffer (0.1M, pH 6) and 100 μl of DTNB. The contents of the cuvette were mixed completely by effervescent air, and absorbance is measured at 412 nm in a very photometer. Once absorbance reaches a stable price, it absolutely was recorded because of the basal reading. 20 μl of substrate, i.e., acetylthiocholine were added and alter in absorbance was recorded. Amendment within the absorbance per minute was therefore determined [30].

\[
R = \frac{5.74 \times 10^{-4} A}{CD}
\]

R= rate in moles of substrate hydrolyzed (min/g tissue)
A=change in absorbance/min
CD= original concentration of the tissue (mg/ml).

\[
\text{Cat(U)}/100 \text{ml of sample} = \frac{(dy/dx) \times 0.003/38.3956 \times 10^{-6}}{0.003/38.3956 \times 10^{-6} \text{molar extinction coefficient of H}_2\text{O}_2} = \frac{dy/dx}{0.003/38.3956 \times 10^{-6}}
\]

dy/dx = change in absorbance/minute

Determination of catalase (cat) principle

Catalase activity was measured by the strategy of Aebi 0.1 metric capacity unit of supernatant was accessorial to a cuvette containing 1.9 metric capacity unit of 50 mm phosphate buffer (pH 7.0). The reaction was started by the addition of 1.0 ml of freshly ready 30 mm H2O2. The speed of decomposition of H2O2 was measured spectrophotometrically from changes in absorbance at 240 nm. The activity of enzyme was expressed as units/mg macromolecule. The reaction happens now when the addition of H2O2 Solutions were mixed well and also the initial absorbance (A1) was browse when 15 s (t1) and also the second absorbance (A2) when 30 s (t2). The absorbance was browse at wavelength 240 nm [31].

\[
\text{Cat(U)} = \frac{0.4 \times 10^{-6} \times 10^{-6} \times 10^{-6}}{0.003/38.3956 \times 10^{-6} \text{molar extinction coefficient of H}_2\text{O}_2}
\]

Determination of malondialdehyde (MDA)

Procedure MDA was measured in step with the strategy of Ohkawa et al. 1 ml of suspension medium was taken from the tissue material in an exceeding tube. 0.5 ml of trichloroacetic acid (TCA) was further to that, followed by 0.5 ml 8% of TBA chemical agent. The tubes were coated with tin foil and unbroken within the water bathtub for 30 min at 90°C when 30 min over the tubes were taken out and placed within the cold water for 30 min. These tubes were centrifuged for 15 min at 3000 rate. The absorbance of the supernatant was taken at 540 nm, at
temperature against applicable blank answer (1 cc H₂O, 0.5 cm³ of half-hour TCA, and 0.5 ml of 0.8% TBA). MDA worth was expressed as n moles MDA/mg of supermolecule [32].

\[
MDA = \left( A_{\text{540}} * 100 / 1.56 \right) * V_t / \text{weight of tissue} * 1000
\]

\( V_t \) = total volume.

**Glutathione peroxidase assay**

3-ml cuvette containing a pair of 2.0 of phosphate buffer (75 mmol/L, PH 7.0), 50 μl of (60 mmol/L) glutathione enzyme resolution, 50 μl of (0.12 mol/L) NaN₃, 0.1 ml of (0.15 mmol/L) Na₂ ethylenediaminetetraacetic acid, 100 μl of (3.0 mmol/L) NADPH, and 100 μl of tissue supernatant was additional. Water was additional to form a complete volume of 2.9 ml. The reaction was started by the addition of 100 μl of (7.5 mmol/L) H₂O₂ and also the conversion of NADPH to NADP was monitored by a continuous recording of the change of absorbance at 340 nm at 1-min interval for 5 min. Enzyme activity of GSHP was expressed in terms of mg of proteins [33].

Enzyme activity (M/min/ml) = \( \left( A_{340} / \text{min} * V_t \right) / (\varepsilon) * d * V_s \)

\( \varepsilon = 6.22 * 10^9 \text{ M}^{-1} \text{ cm}^{-1} \)

\( d = 1 \text{ cm} \)

\( V_t = \text{total volume} \)

\( V_s = \text{sample volume} \)

**Statistical analysis**

The data obtained from animal experiments were analyzed with InStat Software by GraphPad (version 3.10). It was expressed as mean±standard error of mean. For statistical analysis, the data were subjected to analysis of variance followed by Dunnett’s t-test. Results were considered to be statistically significant at p≤0.05. Significance levels were as follows:

* indicates ps0.5 as significant

** indicates ps0.01 as highly significant

*** indicated ps0.001 as very significant

* is used to denote the comparison of standard and test groups with the toxicant control group. Experimental data obtained from all the animals in different groups, recorded, and tabulated on a broadsheet using MS Excel program, to prepare the graphs.

**RESULTS AND DISCUSSION**

**Preliminary phytochemical investigation**

**Table 1: Preliminary phytochemical result**

| S. No. | Phytoconstituents | Test  | Ethanolic extract of the leaves Costus pictus |
|-------|------------------|------|---------------------------------------------|
| 1.    | Test for carbohydrate | Molisch test  | +                                           |
| 2.    | Protein test | Ninhydrin test | +                                           |
| 3.    | Steroid test | Liebermann-bur test | +                                           |
| 4.    | Glycoside test | Kjeller-kiliiani test | +                                           |
| 5.    | Saponin test | Foam test | +                                           |
| 6.    | Test for flavonoids | Shinoda test | +                                           |
| 7.    | Test for tannin | Ferric chloride test | –                                           |
| 8.    | Test for triterpenoids | Salkowski test | +                                           |
| 9.    | Test for alkaloids | Mayer’s test | +                                           |
|       |                  | Wagner’s test | +                                           |
|       |                  | Hagers test | +                                           |

The revealed results of the preliminary phytochemical screening of ethanolic extract of the whole plant of Costus pictus D. Don are shown in Table 1.

**Fig. 6: (a-b) Qualitative phytochemical of Costus pictus D. Don**

**NOR test**

The exploring time for novel objects expressed as RI. An increase in RI indicates anti-Alzheimer activity. From Table 2, we are able to see that each one of the animals altogether the teams spent longer in exploring the novel object except the negative control (Scopolamine) group. The control group was 66.65±7.881%. Negative group showed a vital decrease in recognition 32.98±4.335%. The upper dose showed vital (p<0.01) increase within the novel object exploration indicate as increase RI as compared with an intermediate and lower group. The intermediate dose (74.29±0.687%) and lower dose (65.17±1.291%) showed a vital increase compared with the negative group. Conjointly, the extract only group (75.93±1.695) showed an increase in RI indicates as improvement of memory. The standard and treatment group antagonized the impact of scopolamine by increasing novel object exploring time.

**Table 2: Effect of ECPDD on RI**

| Groups | RI |
|--------|----|
| Control group | 66.65±7.881 |
| Negative control (0.5 mg/Kg) | 32.98±4.335 |
| Standard (2.5 mg/kg) | 81.63±0.833 |
| Low dose (100 mg/kg) | 65.17±1.291 |
| Medium dose (150 mg/kg) | 74.29±0.687 |
| High dose (250 mg/kg) | 80.83±1.681 |
| Extract only (250 mg/kg) | 75.93±1.695 |

Values are expressed as the mean±standard error of mean of n=6 rats/treatment. Significance **ps0.01. RI: Recognition index

**Fig. 7: Effect of ethanolic ECPDD leaves on recognition index**

**Elevated plus maze**

**Table 3: Effect of ethanolic ECPDD on IR**

| Groups | TL Day:14 | TL Day:15 | IR |
|--------|-----------|-----------|----|
| Control group | 15.83 | 6.5 | 0.594±0.079 |
| Negative control (0.5 mg/kg) | 37.66 | 26.83 | 0.480±0.117 |
| Standard (2.5 mg/kg) | 25.33 | 11.33 | 0.696±0.089 |
| Low dose (100 mg/kg) | 26.66 | 15.3 | 0.512±0.056 |
| Medium dose (150 mg/kg) | 23 | 12.33 | 0.530±0.134 |
| High dose (250 mg/kg) | 27 | 9.833 | 0.598±0.053 |
| Extract only (250 mg/kg) | 30.833 | 9.33 | 0.654±0.071 |

Values are expressed as the mean±standard error of mean of n=6 rats/treatment. Significance **ps0.01. TL: Transfer latency, IR: Inflexion ratio
The impact of all the drug-treated groups was evaluated at the top of 14th day. TL was recorded. It absolutely was seen that TL for all the drug-treated groups was less on the 15th day as compared to the 14th day. Decrease IR indicates the induction of a state of mind, and exaggerated IR indicates improvement in psychological features and memory impairment. Negative control group (scopolamine) animals were considerably shrunken compared with all the groups that indicated that state of mind is evoked. Furthermore, high dose showed an increase in IR compared with intermediate and low dose separately and also the extract the only group showed significant increase compared with the negative group.

Y MAZE

Table 4: Effect of ethanolic ECPDD on % alteration

| Groups               | Percentage alteration |
|----------------------|-----------------------|
| Control group        | 51.38±1.338           |
| Negative control (0.5 mg/kg) | 31.42±2.016         |
| Standard (2.5 mg/kg) | 45.13±2.05            |
| Low dose (100 mg/kg) | 41.75±0.705           |
| Medium dose (150mg/kg) | 44.5±1.361         |
| High dose (250 mg/kg) | 47.5±0.530            |
| Extract only (250 mg/kg) | 48.7±3.17          |

Values are expressed as the mean±standard error of mean of n=6 rats/treatment. Significance **p≤0.01

Biochemical test

Estimation of acetylcholinesterase enzyme levels in the brain

Table 5: Effect of ethanolic ECPDD on acetylcholine esterase level

| Groups               | Enzyme level |
|----------------------|--------------|
| Control group        | 0.0711±0.001 |
| Negative control (0.5 mg/kg) | 0.095±0.001    |
| Standard (2.5 mg/kg) | 0.067±0.0015  |
| Low dose (100 mg/kg) | 0.071±0.0025  |
| Medium dose (150mg/kg) | 0.068±0.0001  |
| High dose (250 mg/kg) | 0.065±0.0005  |
| Extract only (250 mg/kg) | 0.069±0.0013  |

Values are expressed as the mean±standard error of mean of n=6 rats/treatment. Significance **p≤0.01
All the groups showed attenuate acetylcholinesterase accelerator activity as compared with hyoscine, as indicative in Table 5. The acetyl enzyme activity was considerably hyperbolic by hyoscine as compared to the control group. The rise in AChE activity by hyoscine was considerably reduced by drug-treated groups. High dose animals showed a vital decrease in acyl-enzyme activity compared with low and medium dose, whereas extract only group showed a vital decrease in acetylcholinesterase activity compared with medium and low. The decrease in a acetylcholinesterase enzyme activity showed improvement in memory and provides anti-Alzheimer’s activity.

**Determination of catalase**

**Table 6: Effect of ethanolic ECPDD on catalase activity**

| Groups                  | Catalase level  |
|-------------------------|-----------------|
| Control group           | 38.51±0.235     |
| Negative control (0.5 mg/kg) | 33.3±0.500     |
| Standard (2.5 mg/kg)    | 40.50±0.115     |
| Low dose (100 mg/kg)    | 37.61±0.035     |
| Medium dose (150 mg/kg) | 38.5±0.315      |
| High dose (250 mg/kg)   | 39.41±0.080     |
| Extract only (250 mg/kg)| 38.79±0.040     |

Values are expressed as the mean±standard error of mean of n=6 rats/treatment. Significance **p≤0.01

The enzyme level was reduced in the negative control group compared with the control group animals considerably (p<0.001). Low, medium, and high dose animal treated with (100, 150, and 250 mg/kg), the enzyme level was increased in ECPPD treated group compared with negative control group animals. The extract only group showed an increase in enzyme level. Donepezil and CPDD showed significantly increased enzyme level that indicates protection from oxidation and also the decreased enzyme level indicates the reaction of the neuronic cell.

**Determination of MDA level**

The MDA level of whole brain was markedly elevated once scopolamine treatment as compared with control. Within the treated groups there was a major (p<0.001) reduction in protein levels in 100 mg/kg, 150 mg/kg, and 200 mg/kg. Donepezil treated group showed important decrease in MDA levels. Results are shown in Table 7.

**Table 7: Effect of ethanolic ECPDD on MDA level**

| Groups                  | MDA Level  |
|-------------------------|------------|
| Control group           | 0.019±0.002|
| Negative control (0.5 mg/kg) | 0.031±0.0005|
| Standard (2.5 mg/kg)    | 0.008±0.00025|
| Low dose (100 mg/kg)    | 0.013±0.0006|
| Medium dose (150 mg/kg) | 0.011±0.000|
| High dose (250 mg/kg)   | 0.004±0.0004|
| Extract only (250 mg/kg)| 0.011±0.00025|

Values are expressed as the mean±standard error of mean of n=6 rats/treatment. Significance **p≤0.01

**Determination of glutathione peroxidase**

**Table 8: Effect of ethanolic ECPDD on glutathione peroxides**

| Groups                  | GSH level  |
|-------------------------|------------|
| Control group           | 1.75±0.005|
| Negative control (0.5 mg/kg) | 1.475±0.0005|
| Standard (2.5 mg/kg)    | 2.249±0.031|
| Low dose (100 mg/kg)    | 1.564±0.002 |
| Medium dose (150 mg/kg) | 1.81±0.002 |
| High dose (250 mg/kg)   | 2.162±0.002|
| Extract only (250 mg/kg)| 2.090±0.000|

Values are expressed as the mean±standard error of mean of n=6 rats/treatment. Significance **p≤0.01
Glutathione plays a key role in maintaining correct perform and preventing apophtosis stress in human cells. It was act as a scavenger for chemical group radicals, singlet oxygen, and varied electrophiles. This narrow balance is maintained by glutathione reduates that catalyzes the reduction of GSSG to GSH. The decrease in the level of glutathione cause chemical reaction of the cell. The glutathione level within the brain markedly decreases in the negative group. Low, medium, and high doses treated with (100, 150, and 200 mg/kg) of CPDD showed considerably (≤0.01) increase in glutathione level. The standard group of animals showed markedly increase in glutathione level. The group within which scopolamine is not given additionally show an increase in glutathione level as compared to the control group.

CONCLUSION

From this study, it is clear that herbs plays a key role against poor memory. Various herbal plants and plants extracts have significant memory improving activity in animal models. C. pictus D. Don have cholinesterase inhibitor property and useful anti-Alzheimer drug in delaying the onset and reducing the severity of AD when compared with that of reference drugs. The memory-improving activity is probably due to the presence of flavonoids. These memory-enhancing drugs showed that potential acting on cognitive functions by maintaining the acetylcholine level in the brain activity is of particular therapeutic importance. The Ethanolic extract of C. pictus D. Don showed a significant increase in the onset of action and decrease in duration of action and recovery of time as compared to negative control thus justifying its anti-Alzheimer activity which may be due to the presence of alkaloid and flavonoid as a phytoconstituent present.

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AUTHORS’ CONTRIBUTION

We declare that this work was done by the authors named in this article and all liabilities pertaining to claims relating to the content of this article will be borne by the authors. Miss Khoshbu Pasha collected the data and analyzed the data. Prof. Imtiyaz Ansari proof-read the whole manuscript, and suggested the necessary changes, and helped in designing the manuscript.

CONFLICTS OF INTEREST

The authors declare that there are no conflicts of interest regarding the publication of this paper.

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