The Untapped Potential of Ginsenosides and American Ginseng Berry in Promoting Mental Health via the Gut–Brain Axis

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Abstract: Despite the popularity of the ginseng (Panax) root in health research and on the market, the ginseng berry’s potential remains relatively unexplored. Implementing ginseng berry cultivations and designing berry-derived products could improve the accessibility to mental health-promoting nutraceuticals. Indeed, the berry could have a higher concentration of neuroprotective and antidepressant compounds than the root, which has already been the subject of research demonstrating its efficacy in the context of neuroprotection and mental health. In this review, data on the berry’s application in supporting mental health via the gut–brain axis is compiled and discussed.

Keywords: ginseng berry; ginsenosides; nutraceutical; gut microbiota; mental health; gut–brain axis

1. Introduction

Mental illness is debilitating and compromises the individual’s quality of life, as well as it has surprisingly far-reaching economic effects, costing the Canadian economy an estimated 51 billion dollars annually [1]. Globally, mental illness has been estimated to be responsible for 32.4% of years lived with disability, surpassing all other forms of diseases [2]. Mood and anxiety disorders are the most common mental illnesses globally and in Canada [3], where their estimated prevalence is 4.7% annually [4]. Unfortunately, many standard pharmaceutical treatments, such as antidepressants, have significant side effects that could affect adherence to the treatment, as well as mixed results with regards to efficacy [5]. Furthermore, it has recently been shown that antidepressant use could negatively impact the intestinal microbiota diversity and be detrimental to certain types of beneficial bacteria [6,7]. With the concept and applications of the microbiota–gut–brain axis gaining traction among the scientific community, complementary treatments targeting this axis to promote mental health are needed.

Ginseng, one of the most important herbs of traditional Chinese medicine, has an impressive track record of positive effects in both in vitro and in vivo models of mental health [8], while also displaying efficacy in clinical research [9–11]. However, in traditional Chinese medicine and even in modern research, the root has been the primary focus of health allegations and, by extension, the focus of ginseng culture. The berries, which are largely regarded as by-products of the ginseng root culture, have great potential for applications in health due to their pharmacological properties and distinct composition with respect to the root [12]. Despite the rationale strongly supporting the pharmacological properties of the berries [12], they remain underutilized and are frequently discarded in agriculture, while the root is marketed. As ginseng is a slowly growing crop, root cultures take multiple years to harvest. Furthermore, significant crop mortality following the replanting of new ginseng is an issue plaguing agriculture; thus, bioremedial efforts have been undertaken to mitigate this effect [13]. The berry culture, on the other hand,
presents numerous advantages. For instance, berries can be harvested from the same ginseng plant annually starting on the second year of growth, without any detriment to the crop. The root culture, in contrast, takes 4 to 10 years to achieve a minimal marketable maturity. Finally, the berry culture can be implemented without impacting the current root harvesting practices.

Here, we evaluate the potential of the ginseng berry as a promising source of bioactive compounds with mental health-promoting effects. This review also discusses the pharmacological mechanisms through the gut–brain axis in which ginseng could promote mental health, as shown in Figure 1.

Figure 1. An overview of the reported studies highlighting the interplay of ginseng berry compounds with the microbiome–gut–brain axis. Green upward arrows represent a significant increase, whereas red downward arrows represent a significant decrease. Green arrow: increase; red arrow: decrease.

2. The Berry Is a Highly Concentrated Source of Ginseng’s Therapeutic Compounds

The main bioactive compounds in the berry are of the same classes as those found in the root. Ginsenosides, usually denoted by a capital R followed by a lowercase letter and a number, if required (e.g., Rg1), are saponins present in an impressive diversity within the same type of ginseng and even in the same part of the plant. Their structural diversity is naturally accompanied by a diverse range of pharmacological functions and efficacy. Ginseng root polysaccharide extracts have also been researched in various contexts. As shown in Table 1, the berry could contain higher levels of neuroprotective and antidepressant bioactive compounds than the root.
Table 1. A review of American ginseng berry bioactive compounds—(*B*) content is significantly higher in the berry than root, (*B<*) content is significantly less than the root, (B~) content is not statistically different from the root, (nil) was not detected in the berry, (ND) not determined in the study. Significance was determined using a two-tailed t-test with p < 0.05. When multiple harvest times were available in a study, the harvest date closest to August 30th was chosen.

| Compounds | Pharmacological Effects | Content (Berry vs. Root, mg/g Dry Weight) |
|-----------|-------------------------|------------------------------------------|
| Ginsenosides | | |
| Rb1 | Neuroprotective [14], anti-diabetic [15], mitochondrial antioxidant [16] | *B< [12,17] 0.86 ± 0.09 vs. 25.36 ± 1.67 [17] |
| | Anti-diabetic, anti-viral, cardioprotective, neuroprotective [20] | ND vs. 47.96 ± 1.04 [19] |
| Rb2 | Anti-diabetic, anticonvulsant, antitumor, cardioprotective, antidepressant [22] | *B> [12,17,21] 1.54 ± 0.95 vs. 0.3 ± 0.02 [17] |
| Rb3 | Antiallergic [23], antioxidant [24], anti-inflammatory [25], SIRT1 activation [26] | *B> [12,21], *B< [17] 1.51 ± 0.11 vs. 7.03 ± 2.15 [17] |
| Rc | Cardioprotective [29], Neuroprotective [30,31], antidepressant [32] | Rb3, Re, Rb2, Rd, and Rc, in descending order of abundance (with occasional variation between Re and Rb2), are the ginsenosides that are the most abundant in the American ginseng berry, and this is consistent throughout the studies assessing its composition via high-performance liquid chromatography [12,42,50,51]. Of note, the berry ginsenoside content can be different depending on the variety of ginseng selected. For instance, ginsenoside Re is the most abundant in Korean ginseng berries and is approximately 8 times more concentrated than Rb2 [52], whereas in the American ginseng, Re is at most 1.2 times as concentrated as Rb2 [12]. The harvest time has also been shown to cause significant variance in ginsenoside content; American ginseng berries were shown to lose over half of their Rb1, Re, and Rg1 content during the season, strongly suggesting that the ginsenoside content may be at its peak before the berries are ripe [18]. Post-harvest treatment should also be considered, as steaming has been shown to cause a sharp decrease in the total content, consistently causing a loss of about 50% after 2 h of steaming at 120 °C [42,50]. Conversely, ginsenosides Rh1, Rg2, (20R)-Rg2, Rg3, and Rh2 sharply increased in content after a 2 h steaming treatment [42,50].
Given the berry’s high concentration of Rb3, Re, Rb2, and Rd, the ginsenosides with demonstrated antidepressant and neuroprotective effects [20,22,27,30,31], it could be expected that the berry has even a superior potential for mental health applications than the root. Still, root extracts and specific ginsenosides have been the subject of most research and have consistently demonstrated efficacy in vitro and in vivo models in the context of central nervous system diseases and depression [53,54]. Another aspect to consider when evaluating the berry’s antidepressant and neuroprotective potential is that the microbial community of the intestine metabolizes the ginsenosides into alternate forms with varying effects and degrees of bioactivity. For instance, Rb3, the berry’s main ginsenoside, and its deglycosylated metabolites Rg3, Rh2, compound K, and 20(S)-protopanaxadiol have had their antidepressant potential assessed, and it was shown that Rg3 and compound K have more powerful antidepressant effects which are brought upon by the modulation of corticosterone, adrenocorticotropic hormone, and noradrenaline levels [55]. Thus, the fact that the berry has an inherently higher concentration of Rg3 than the root and a higher concentration of Rb3, which can, in turn, be deglycosylated into Rg3 [12,55], is a fine example of the berry’s untapped potential as a mental health-promoting nutraceutical.

3. Pharmacological Effects in The Context of Mental Health

Most of the mental health-promoting effects attributed to the berry come from extrapolation of data from single ginsenoside or total ginsenoside extract experiments. Data from experiments directly involving the berry or its distinct ginsenoside composition are scarce in the context of mental health. The berry saponin extract was shown to regulate 5-HT and rescue depressive-like behaviour in a mouse model of myocardial infarction, though this was fruit from the Panax notoginseng [56]. In fact, ginseng berry experiments with application to mental health seem to be limited to the examination of serotonin regulation in comorbid myocardial infarction models [57,58] and one additional study involving scopolamine-induced memory impairment, where the berry extract was shown to have antioxidant effects and to preserve acetylcholine and brain-derived neurotrophic factor (BDNF) mRNA levels [59]. This section illustrates the current mental health-related findings for the predominant ginsenosides in the American ginseng berry.

3.1. Ginsenoside Rb3

Rb3, the berry’s most abundant ginsenoside, exerts pharmacological effects that benefit mental health through multiple mechanisms. Such neuroprotective mechanisms occur through varied antioxidant effects, such as suppressing inducible nitric oxide synthase in hypoxic hippocampal neurons [60], preserving superoxide dismutase (SOD) and catalase (CAT) levels [61], and inducing Nrf2 transcription activity [62], which is downregulated in neurological conditions, such as depression [63]. Likewise, ginsenoside Rb3 was shown to interact with multiple neurotransmitters and receptors, leading to neuroprotective effects through inhibiting the NMDA receptor [64,65], activating the GABA(A) receptor [66], or acting beneficially on the noradrenergic pathway to relieve depression in rodent models [55,67].

3.2. Ginsenoside Re

Ginsenoside Re also demonstrates neuroprotective effects. It could be effective at reducing neuroinflammation by inhibiting the CAMK/MAPK/NF-κB signaling, as demonstrated by Madhi et al. [30], as well as by attenuating NLRP3 activation, as reported by Wang et al. [31]. In the same study, ginsenoside Re was also able to counter the loss of the antioxidant enzymes SOD, CAT, and glutathione (GSH) and the loss of Nrf2 expression following chronic restraint stress [31]. The compound also induced the expression of genes involved in acetylcholine neurotransmission, elevated acetylcholine levels, and enhanced the differentiation of Neuro-2a cells, which could translate to benefit in Alzheimer’s disease [68]. The neuronal effects also extend to reversing the depression- and anxiety-associated behavioural changes in rat models of repeated immobilization [32] and the
learning and memory decline caused by chronic restraint in mice [31], while exerting BDNF-protecting effects in both studies.

3.3. Ginsenoside Rb2

Research evaluating the efficacy of ginsenoside Rb2 is scarce in the context of mental health, though it has been shown to protect against glutamate-mediated neurotoxicity in HT22 hippocampal cells [69]. Miao et al. have recently written a review compiling the pharmacological effects of Rb2, which include inhibition of oxidative stress, inflammation, and apoptosis through multiple pathways [20]. Although these effects were not tested in neurological models, some described pathways (SIRT1, AMPK, MAPK, and NF-κB) are relevant for many neurological conditions.

3.4. Ginsenoside Rd

Chen et al. have written a comprehensive review thoroughly describing the neuroprotective mechanisms of ginsenoside Rd, which was published a few months prior to this paper [27]. Some key reported data include anti-inflammatory effects via the regulation of iNOS, COX-2, MAPK, and NF-κB, antioxidant effects through increasing the SOD, GSH, and CAT, and antiapoptotic effects in several models of neuron stress [27]. Ginsenoside Rd was more recently shown to exert a significant antidepressant effect in the chronic unpredictable mild stress and behavioural despair mouse models via the hypoxia-inducible factor-1α and to increase the expression of SYN1 and PSD 95, two synaptic plasticity-related proteins [28]. Also of note, ginsenoside Rd alleviated both Escherichia coli K1-induced colitis and depression/anxiety in mice as measured by light/dark transition, forced swimming, and tail suspension tests, while significantly countering induced IL-6 expression in plasma and NF-κB activation (both colonic and hippocampal) [70]. In the same study, ginsenoside Rd also protected the hippocampal BDNF levels and even reversed some changes in intestinal microbiota, brought upon by the administration of Escherichia coli K1 [70].

Ginsenosides could also exert neuroprotective effects through the modulation of microRNA, and Rd modulating miR-144-5p in a glioblastoma model is one such example [71]. In this study, Rd upregulated miR-144-5p, which decreased both TLR2 and the proliferation of the glioblastoma cells [71]. Although it remains to be confirmed that ginsenoside Rd could systemically upregulate miR-144-5p in vivo at a significant level, by extrapolating this microRNA’s targets to other models, it could be hypothesized that ginsenoside Rd has the potential to act therapeutically where TLR2 antagonism has shown benefit. For instance, anti-TLR2 has proven beneficial in decreasing α-synuclein accumulation in neuronal and astroglia cells in Parkinson’s and dementia with Lewy bodies mouse models, accompanied by decreased neuroinflammation and behavioural deficits [72]. Notably, miR-144-5p has been downregulated in depression and anxiety relative to healthy controls and inversely correlated with depression scores [73]. Similarly, a psychological treatment that decreased depression scores decreased specific inflammation-associated proteins and increased miR-144-5p in another cohort of depression, anxiety, and stress-related disorder patients [74]. Recently, Hyun compiled research demonstrating the microRNA modulating effects of various ginsenosides [75], but it may be too early to further extend these findings to the context of mental health. As microRNAs continue to gain traction as therapeutic targets, more research evaluating ginseng’s ability to modulate microRNAs would be of benefit to the scientific community.

In summary, the American ginseng berry’s main ginsenosides are promising mental health-promoting compounds through multiple neuroprotective and anti-depressive mechanisms. As illustrated by a previously mentioned study involving Es. coli K1 administration [70], the ginseng berry’s bioactive components can additionally exert mental health benefits by modulating microbiota and other intestinal health parameters.
4. The Ginseng Berry and The Gut–Brain Axis

Beyond direct pharmacological action on the nervous system, another mechanism through which ginseng could promote mental health is through the gut–brain axis. This axis, relating the concepts of the intestinal microbiome, intestinal barrier function, endocrine and neurological factors, and mental health, is of great importance as new implications for a wide range of disease states have been emerging. For instance, links have been established between the gut–brain axis and neurological conditions, such as Alzheimer’s disease, Parkinson’s disease, amyotrophic lateral sclerosis, stroke, and major depressive disorder [76,77], highlighting the need for more research evaluating strategies to target the gut–brain axis in these contexts effectively. Given the close link between host nutrition and the intestinal microbiome, nutritional and nutraceutical strategies are promising avenues to explore in helping treat these conditions. Potential therapeutic targets along the axis include the positive modulation of the intestinal microbiota composition or the reversal of dysbiosis, the reduced permeability of the intestinal epithelium to inflammatory food-derived antigens and inflammatory microbial products, and even the mitigation of the negative impact that psychotropic drugs could exert on the gut microbiome [78].

4.1. Intestinal Permeability

It is well known that with increased intestinal permeability, inflammatory microbial products, such as lipopolysaccharides (LPS), are present in higher quantities in the systemic circulation [79]. This endotoxemia results in metabolic dysfunction and neuroinflammation, potentially leading to overt depressive and anxious behaviour [80,81]. Indeed, serum LPS has been shown to dose-dependently depress mood in humans [81]. Further, the translocation of bacterial LPS into the systemic circulation is a major driver in the “leaky gut” model of depression [82]. There are currently insufficient data to determine the effect of the ginseng berry on the intestinal barrier function, though there is room for extrapolation. The effects of different ginseng extracts on relevant intestinal barrier function parameters, such as the tight junction proteins Claudin-1, Occludin, and Zonula Occludens-1 (ZO-1), colonic inflammatory markers, and serum markers of permeability, such as LPS and D-lactate, are reported in Table 2.

Table 2. Direct effects of ginseng on the intestinal barrier function.

| Compounds                        | Models                                | Mechanism(s)                      | Significant Effects ($p < 0.05$)                                      |
|----------------------------------|---------------------------------------|-----------------------------------|----------------------------------------------------------------------|
| American Ginseng Root Polysaccharides | Antibiotic-associated Diarrhea in Rats (Lincomycin Hydrochloride) | MAPK Signaling                    | Reduces colonic IL-1β, IL-6, IL-17A and TNF-α and increases IL-4 and IL-10. Increases Claudin-1 and Occludin expression [83] |
| Korean Ginseng Root Polysaccharides          | DSS-induced Colitis in Rats | TLR4/MyD88/NF-κB-signaling pathway inhibition | Alleviates colitis symptoms, downregulates IL-1β, IL-2, IL-6, IL-17A, upregulates ZO-1 and Occludin [84] |
| Fermented Korean Ginseng Root Ginsenosides | Intraperitoneal LPS Injection in Mice | TLR4/MAPK                          | LPS-induced increases in ALT and AST, increases LPS-induced expression of Claudin-1 [85] |
| American Ginseng Ginsenosides                              | Cisplatin-induced intestinal injury in Mice | Decreased NF-κB activity          | Attenuates cisplatin-induced increases in TNF-α and IL-1β. Attenuates cisplatin-induced decreases in ZO-1 and Occludin [86] |
Table 2. Cont.

| Compounds                               | Models                                      | Mechanism(s)                  | Significant Effects ($p < 0.05$)                                                                 |
|-----------------------------------------|---------------------------------------------|-------------------------------|-------------------------------------------------------------------------------------------------|
| Korean Red Ginseng Root                 | MPTP-induced Intestinal Permeability in Mice| -                             | Prevents MPTP-induced decrease in Occludin and ZO-1, and MPTP-induced colonic increase in TNF-α and IL-1β [87] Increases jejunal villus height and expression of Occludin and Claudin in both LPS-treated and control groups. Alleviates LPS-induced increases in ALT, AST, TNF-α, and IL-1β [88] Decreases serum LPS levels and decreases plasma FITC-dextran. Pretreatment prevented plasma IL-6 decrease and TNF-α increase. Treatment dose-dependently increases ZO-1 and Occludin post-radiation injury [89] Dose-dependently reduces serum D-lactate and intestinal clearance of FITC-dextran [90] Both treatments decrease IL-4 and TNF-α mRNA expression.  |
| Ginseng Polysaccharides (Unspecified Variety) | Intraperitoneal LPS Injection in Piglets | Decreased LPS-induced NF-κB activity | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Korean Ginseng Root Oligopeptides       | Irradiation induced intestinal injury in mice| -                             | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Ginsenoside Rb1                         | Peritoneal air exposure intestinal damage in Rats | -                             | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Fermented and Unfermented Korean Red Ginseng Root | Ovalbumin-induced allergy in sensitized mice | Th1/Th2 balance, IgE suppression | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Fermented Wild Ginseng Root             | DSS-induced colitis Mouse Model             | Decreased DSS-induced NF-κB activity | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Korean Ginseng                          | Healthy Mouse Model                         | -                             | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Ginsenoside Rk3                         | High-fat diet Mouse Model                   | TLR4/NF-κB signaling pathway inhibition | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Ginsenoside Rh2                         | T-cell acute lymphoblastic leukemia mouse model | Decreased TLR4/MyD88 expression | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Ginsenosides Rb3 and Rd                 | ApcMin/+ mice (colon cancer model)         | -                             | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
| Ginsenoside Rk3                         | Hepatocellular carcinoma mouse model       | TLR4 pathway inhibition       | Both treatments prevented an allergy-induced increase in serum beta-lactoglobulin after gastric administration [91] Alleviates colitis, prevents DSS-induced loss of ZO-1, downregulates DSS-induced IL-1β, IL-6, TNF-α, and IFN-γ mRNA expression. Decreases colonic levels of TNF-α [92] Increased Muc2 expression [93] Reduced colonic inflammatory cytokines and oxidative stress. Increases ZO-1, Occludin, and Claudin expression [94] Decreased IL-1β, IL-6, and TNF-α. Increased IL-10 and TGF-β. Increased mRNA expression of ZO-1, Claudin, and Occludin [95] Increased Goblet and Paneth cell count [96] Visual restoration of the intestinal barrier, increased expression of ZO-1, Occludin, and Claudin [97] |
Table 2. Cont.

| Compounds                        | Models                                | Mechanism(s)                                 | Significant Effects (p < 0.05)                                                                 |
|----------------------------------|---------------------------------------|----------------------------------------------|------------------------------------------------------------------------------------------------|
| Ginsenoside Rk3                  | Lincomycin-treated mice               | -                                            | Increased expression of ZO-1, Occludin, and Claudin-1, and reversed structural changes to the epithelium. Prevented increased IL-1β, IL-6, IL-17, IFN-γ and TNF-α and prevented decreased IL-10 [98] |
| Ginsenoside Rg5                  | db/db diabetes mouse model            | TLR4/NF-κB signaling pathway inhibition      | Increased Occludin and ZO-1 protein expression, decreased serum LPS [99]                                                                          |
| Panax Notoginseng saponins       | Lepob mice on a high-fat diet         | TLR4 pathway inhibition                      | Increased expression of ZO-1 and Claudin-1 [100]                                                                                                  |
| Ginsenoside Rh4                  | Antibiotic intestinal inflammation mouse model | Decreased TLR4/NF-κB/MyD88 expression       | Increased expression of ZO-1 and Claudin-1. Decreased IL-1β, IL-6, IL-17, IFN-γ and TNF-α. Prevented increase in IL-10. Reduced serum LPS [101] Both ginsenosides and polysaccharides independently increased mucin area, goblet cell count, and increased expression of ZO-1 and Occludin, but the combination had higher effect [102] |
| American ginseng polysaccharides and ginsenosides | Cyclophosphamide-Induced Intestinal Damage in Mice | -                                            | Decreased levels of IL-6, IL-33, TNF-α and increased IL-4 and IL-10 [103]                                                                 |
| Ginsenoside Rg1                  | DSS-induced colitis mouse model       | -                                            | Increased expression of ZO-1 and Occludin mRNA expression. Decreased serum LPS [104]                                                            |
| Korean Ginseng Ginsenosides      | Mice on a high-fat diet               | -                                            | Overall, ginsenosides have remarkable potential as therapeutic products for preserving the intestinal barrier function in various stress situations through anti-inflammatory and transcriptional effects, favourably modulating tight junction protein expression. For instance, Seong et al.’s study involving a fermented ginseng root in a dextran sodium sulfate-induced murine colitis model has shown that the extract prevents the loss of the tight junction protein Zonula Occludens-1 while inhibiting the NF-κB inflammatory pathway [92]. The ginsenosides’ effects appear to go beyond mitigating the loss of tight junction proteins amidst inflammatory insult; ginsenosides have also been shown to upregulate the expression of tight junction protein expression and mRNA expression [97,99–101,104]. Ginseng has also been shown to increase Muc2 expression [93], though it is unclear if this is a direct upregulation or due to the increase in goblet cell count that ginsenosides have also been shown to induce [96]. The effects discussed so far are complemented by improvements in histological parameters, such as the mucin barrier area or the thickness and reversal of epithelial damage from an impressive range of causes. Functional experiments of intestinal permeability have also demonstrated efficacy with decreases in serum LPS, replicated in a few different studies [99,101,104]. The ginseng root has also been shown to decrease serum beta-lactoglobulin following gastric administration in a mouse model of allergy [91] and decrease serum D-lactate and FITC-translocation across the intestinal epithelium in a rat model of intestinal injury by peritoneal air exposure [90]. However, |
research examining the impact of whole ginseng berries specifically on the intestinal barrier function is lacking. Furthermore, it is unknown whether berry polysaccharides or other non-saponin compounds could also exert a protective effect on the intestinal barrier, as root polysaccharides and oligopeptides have been shown to do.

4.2. Prebiotic Effects and Modulation of The Intestinal Microbiota

Another way the ginseng can act on the gut–brain axis is through modulation of the intestinal microbiome. This type of benefit to the axis contrasts with the previously discussed pharmacological effects, as they are indirect. Microbiome modulation is a multifaceted phenomenon whose results on the gut–brain axis depend on the initial host microbiome, diet, immunological factors, etc. Table 3 provides several examples of ginseng components reversing induced dysbiosis. However, it should be noted that ginseng also induces positive microbiome changes in healthy models, as well as improved intestinal metabolism and immunity, as shown by Sun et al. [105].

Table 3. Direct effects of ginseng on the intestinal microbiome.

| Compounds | Models | Significant Effects ($p < 0.05$) |
|-----------|--------|---------------------------------|
| American Ginseng Root Polysaccharides | Antibiotic-associated Diarrhea in Rats (Lincomycin Hydrochloride) | Increased production of acetate and propionate, improved the relative richness of *Lactobacillus* and Bacteroides, and reduced the relative richness of *Blautia* and *Coprococcus* [83] |
| Korean Ginseng Ginsenoside Rk3 | Healthy Mouse Model | Increased total bacterial count and *Lactobacillus* count [95] |
| 25-hydroxyl-protopanaxatriol | High-fat diet Mouse Model with streptozotocin | Increased abundance of *Bacteroides* and *Bifidobacteria*, decreased abundance of *Firmicutes* [94] |
| Fermented Wild Ginseng root | Antibiotic-associated diarrhea mouse model | Partly reversed an increase in Firmicutes/Bacteroides ratio, increased relative abundance of *Lachnospiraceae* [106] |
| Ginsenoside Rh2 | T-cell acute lymphoblastic leukemia mouse model | Increased relative abundance of *Akkermansia*, *Lactobacillus*, and *Lachnospiraceae* [95] |
| Korean red ginseng root insoluble fiber | In vitro colon-simulated fermentation using swine fecal bacteria | Increased production of short-chain fatty acids, decreased alpha-diversity, and increased relative abundance of *Bifidobacterium* and *Prevotella* compared to control fermentation with cellulose [108] |
| Fermented Korean Ginseng Root | Alcoholic injury mice (ethanol diet) | Prevented relative abundance loss of *Akkermansia* and *Allobaculum*. Decreased relative abundance of *Parabacteroides* [109] |
| Ginseng Root Polysaccharides (Unspecified variety) | Healthy Piglets with supplemented diet | Increased colonic acetic acid, isobutyric acid, and butyrate. Decreased abundance of *Malainabacteria* [110] |
| Water Soluble Neutral Ginseng Polysaccharides | Antibiotic-associated Diarrhea in Mice (Lincomycin Hydrochloride) | Increased abundance of *Lactobacillus*, decreased abundance of *Bacteroides*, *Streptococcus*, *Ochrobactrum*, and *Pseudomonas* [111] |
| Unspecified Ginseng Extracts (Article in Chinese) | Healthy Rats | Increased abundance of *Bifidobacterium*, *Lactobacillus*, *Allobaculum*, and *Clostridium*. Decreased abundance of *Butyricimonas*, *Parabacteroides*, *Alistipes*, and *Helicobacter* [112] |
| Korean Red Ginseng Root Polysaccharides and Ginsenoside Rb1 | Streptozotocin-Induced Diabetes Mouse Model | Polysaccharide treatment reversed the dysbiosis caused by the treatment, as evidenced by reversal of loss of relative abundance of Firmicutes and reversal of increase of the relative abundance of *Bacteroides* [113] |
### Table 3. Cont.

| Compounds                                      | Models                                              | Significant Effects ($p < 0.05$)                                                                                                                                                                                                                                                                                                                                                                                                                                                                 |
|------------------------------------------------|-----------------------------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Ginseng Root Polysaccharides                   | DSS-induced Colitis Mouse Model                     | Reverses DSS-induced changes; increases abundance of *Bifidobacterium*, *Lactobacillus*, and the bacteria *Clostridium leptum* and *Clostridium cocoides*. Reduces abundance of *Enterobacteriaceae* and the bacterium *Bacteroides fragilis* [114]                                                                                                                                                                                                                           |
|                                                 |                                                     | Decreased abundance of *Dysgonomonas*, *Porphyromonas*, and *Parabacteroides*. Increased abundance of *Prevotella* and *Paraprevotella* (Rd only). Increased richness of family *Bacteroidaceae*; promoted growth of *Bacteroides vulgatus*, *Bacteroides xylanisolvens*, *Bacteroides gallinarum*, and *Bacteroides acidifaciens* [96]                                                                                     |
| Ginsenosides Rb3 and Rd                         | Apc<sup>Min/+</sup> mice (colon cancer model)       | Bacteroidaceae; increased Lactobacillus, Actinobacillus, and the bacteria *Bacteroides acidifaciens*. Reduced abundance of *Enterobacteriaceae* and the bacterium *Bacteroides fragilis* [96]                                                                                                                                                                                                                                                                                     |
| American Ginseng Root                           | AOM/DSS intestinal inflammation and                | Gradual reversal of loss of alpha-diversity and beta-diversity following DSS treatment. Reversed increase in *Bacteroidaceae*, *Porphyromonadaceae*, *Enterobacteriaceae*, and *Verrucomicrobiaceae*, and reversed the decrease in *Clostridiales*, *Catabacteriaceae*, *Lachnospiraceae*, and *Ruminococcaceae* [115]                                                                                               |
|                                                 | tumorigenesis mouse model                           |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                               |
| Ginsenoside Rk3                                 | DEN- and CCl4-induced Hepatocellular carcinoma      | Reversed increase in *Helicobacter* and reversed the decrease in *Akkermansia*, *Lactobacillus*, *Oscillibacter*, and *Bifidobacterium* [97]                                                                                                                                                                                                                                                                                                          |
| Korean Ginseng Root Polysaccharides             | DSS-induced colitis in Mice                        | Restored loss of alpha diversity (Shannon Index). Reversed relative increase in *Bacteroidetes*, *Verrucomicrobiia*, *Proteobacteria*, *Tenericutes*, *Cyanobacteria*, *Prevotella* and *Defervibacterae* and reversed the decrease of *Firmicutes* and *Akkermansia* [116]                                                                                             |
|                                                 |                                                     | Preserved Simpson, Shannon, ACE and Cha01 index at levels of control. Increased levels Bacillaceae, *Bacteroidaceae* and *Prevotellaceae*. Increased levels of *Aerob痞spites*, *Alloprevotella*, *Lachnoolstiridium* and *Blautia*. Decreased loss of acetic acid production, prevented decrease of propionic acid, butyric acid, isobutyric acid, and valeric acid production [98]                                                                 |
| Ginsenoside Rk3                                 | Lincomycin-treated mice                             | Reversed relative loss of Bacteroidetes and reversed relative increase of Firmicutes. Increased Lactobacillus and Bacteroides, decreased Anaerotruncus. Reversed loss of *Bifidobacterium*, *Streptococcus*, *Coprococcus*, and *Clostridium* [117]                                                                                                                 |
| Ginseng Root Water-Soluble Extract (Unspecified Variety) | Exercise-Fatigue Mouse Model                        | Increased relative abundance of *Escherichia-Shigella*, decreased relative abundance of *Dorea*, *Prevotella*, and *Megasphaera*. Increased abundance of *Lachnospiraceae*, *Streptococcaceae* . . . (Abridged) [118]                                                                                                                                                                                                 |
| Protopanaxadiol-type Ginsenosides Extracted from Korean Ginseng Root | Human Fecal Microbiota In Vitro Fermentation       |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                               |
| Korean Ginseng Root                             | Middle-Aged Korean Women with Obesity              | Decreased relative abundance of *Anaerostipes* [119]                                                                                                                                                                                                                                                                                                                                                                                                                                           |
| Korean Red Ginseng Root                         | Patients with non-alcoholic steatohepatitis         | Increased *Lactobacillus* in subgroup who experienced improvements in ALT [120]                                                                                                                                                                                                                                                                                                                                                                                                                 |
| Ginsenoside Rg5                                 | db/db diabetes mouse model                          | Reversed decrease in abundance of *Alloprevotella*, *Barnesiella*, *Coprobacter*, *Lactobacillus*, *Lactococcus*, and *Parasutterella*, reversed increase in abundance of *Oscillibacter*, *Clostridium*, *Helicobacter*, and *Dorea* (abridged) [99]                                                                                                                                 |
| Panax notoginseng saponins                      | Diet-induced obesity mice                           | Increased abundance of *Akkermansia muciniphila* and *Parabacteroides distosinosis* [121]                                                                                                                                                                                                                                                                                                                                                                                                         |
| Ginsenoside Rb1                                 | Diet-induced obesity mice                           | Decreased *Helicobacteraceae* and *Ruminococcaceae*, and enriched *Rikenelaceae*. Decreased abundance of *Dorea*, *Helicobacter* and *Oscillospira* [122]                                                                                                                                                                                                                                                                                     |
Table 3. Cont.

| Compounds                        | Models                                      | Significant Effects ($p < 0.05$) |
|----------------------------------|---------------------------------------------|---------------------------------|
| Panax Notoginseng saponins       | Lep$^{ab}$ mice on High-fat diet            | Increased fecal acetic acid, butyric acid, propionic acid, isobutyric acid, valeric acid and isovaleric acid [100]. |
| Ginsenoside Rh4                  | Antibiotic intestinal inflammation mouse model | Decreased Firmicutes/Bacteroidetes ratio. Increased fecal acetic acid, butyric acid, propionic acid, isobutyric acid, valeric acid and isovaleric acid [101]. The combination increased abundance of Clostridiales, Bifidobacterium, and Lachnospiraceae, and decreased abundance of Escherichia-Shigella and Peptococaceae (reversing detrimental changes in microbiota). Polysaccharides and ginsenosides had different and synergistic effects [102]. |
| American ginseng polysaccharides and ginsenosides | Cyclophosphamide-Induced Intestinal Damage in Mice | The combination reversed the changes in microbiota. Polysaccharides and ginsenosides had different and synergistic effects [123]. |
| Korean Ginseng polysaccharides and ginsenosides | Exhaustion by forced swimming and human hepatoma HepG2 cells xenograft | Increased relative abundance of Firmicutes and Proteobacteria and decreased relative abundance of Bacteroidetes. Increased abundance of Escherichia, Streptococcus and Ruminococcus. Decreased abundance of Dorea, Sutterella, Prevotella and Megasphaera [124]. |
| Ginsenosides                     | Human Fecal Microbiota In Vitro Fermentation | Increased relative abundance of Lachnospiraceae and decrease of Staphylococcus, Bacteroid and Ruminococcaceae [103]. |
| Ginsenoside Rg1                  | DSS-induced colitis mouse model              | Increased abundance of Parabacteroides, Muribaculaceae, Akkermansia, and Ruminococcus. Decreased abundance of Lachnospiraceae and Helicobacter [104]. |
| Korean Ginseng Ginsenosides      | Mice on High-fat diet                        | Increased abundance of Bifidobacterium and Lactobacillus [105]. |
| Korean Ginseng                   | Healthy Rats                                 | Increased abundance of Bifidobacterium and Lactobacillus [105]. |

Ginseng and ginseng extracts have shown remarkable microbiome modulatory effects in an incredible amount of disease states and experimental diets. The reversal of deleterious microbiome changes is a persistent observation. For instance, the ginseng extracts have either increased or decreased the Firmicutes/ Bacteroidetes ratio at the phylum level, whichever would reverse the changes brought upon by metabolic or intestinal dysfunction [94,97,101,106,113,116,117]. Based on the available literature, there appears to be a distinction between the effects of the root polysaccharides and the ginsenosides that can be made with regard to this ratio. Indeed, the ginsenosides appear to decrease the Firmicutes/ Bacteroidetes ratio [94,97,101,106,117], whereas the root polysaccharides appear to increase it [113,116]. However, this distinction may be an oversimplification. It should be noted that the disease models in the experiments using ginsenosides were characterized by an increase in the Firmicutes/Bacteroidetes ratio, which the ginsenosides effectively reversed. In contrast, the models in the experiments using polysaccharides were characterized by a decrease in this ratio, which the polysaccharides also effectively reversed. It is unclear if the ginsenosides could also have acted to therapeutically increase the ratio if they had been used in a context where the increase would have acted as a reversal of dysbiosis, though it remains plausible. In supporting this idea, in vitro human microbiota-simulated fermentation with ginsenosides increased this ratio [124]. Together, these findings portend evidence that ginseng’s effects on the gut microbiome could be contextually adaptable to exert benefits.

As shown in Figure 2, the ginseng extracts are relatively consistent in their effects on beneficial and detrimental bacteria from various genera. Indeed, ginseng’s prebiotic effect on beneficial bacteria of the genera Akkermansia, Bifidobacterium, and Lactobacillus is consistent throughout studies. The increase of Akkermansia, particularly Akkermansia muciniphila, is a beneficial characteristic associated with intestinal barrier functions [125,126]. The positive effects of Bifidobacterium and Lactobacillus are well known; these bacteria are associated with increased barrier function [127,128], which extends to metabolic benefits [129]. Of
note, *Bifidobacterium* and *Lactobacillus* could have antidepressant effects \[130,131\] in their own right. Furthermore, consistent increases in short-chain fatty acid (SCFA) production following supplementation of extracted ginsenosides suggest that the saponins preferentially support the growth of SCFA-producing bacteria \[98,100,101\]. Increased production of luminal butyrate is of great benefit to the intestinal barrier function, as it has been shown to promote mucosal healing and production of protective mucus along the intestinal epithelium, and to the decrease in intestinal permeability by modulating epigenetic and transcriptional activity in the cells of the intestinal epithelium \[132–136\]. Another important piece of information to be extracted from this compilation of data is that despite ginseng’s prebiotic effects on beneficial bacteria, there is a homogenous observation that it exerts selective antibacterial effects on bacteria that are considered detrimental, such as those from the genera *Dorea* and *Helicobacter*. Of importance in this context is *Dorea’s* association with major depressive disorder \[137\].

**Figure 2.** Reported prebiotic and antibacterial effects of ginseng and ginsenosides on microbial genera.

### 4.3. Improved Health Functionality through Bioconversion

Most ginsenosides have inherently low bioavailability, therefore, ginseng bioconversion by the gut microbiome is critical for absorption by the host. Through microbial bioconversion processes, such as deglycosylation, the ginsenosides can achieve higher absorption rates and pharmacological activity \[138,139\]. Examples of metabolized ginsenosides include the most bioactive compound K and ginsenoside Rg3 \[140\]. Since this is a microbial process, the host’s microbiome significantly impacts the outcome of an administered dose of ginseng extract; thus, the bioconversion has varying effects between individuals \[138\]. Another aspect to consider in preparing extracts to improve health parameters is the apparent synergy between different plant components. For example, ginseng root polysaccharides have been shown to promote the microbial metabolism of co-administered ginsenoside Rb1 by a prebiotic effect in vivo while upregulating the intestinal uptake of Rb1 in vitro \[114\]. The observations of synergy have also been echoed in the context of efficacy for reversing cyclophosphamide-induced intestinal damage, where
American ginseng ginsenosides and root polysaccharides were shown to have slightly different effects on inflammation, but were synergistic when co-administered [102]. To date, it remains unknown if berry polysaccharides and ginsenosides share the same synergistic relationship as root polysaccharides and ginsenosides.

5. Safety

In 2021, a systematic review aiming to include all clinical trials involving all forms of ginseng was published [141]. Of the 121 retained studies that evaluated safety, 41.6% reported no adverse events, 31.6% reported no significant difference between groups in adverse events, and 26.6% reported no serious adverse events [141]. Mild adverse events included dizziness, headaches, diarrhea, insomnia, hypoglycemia, and nausea [141]. Due to the scarcity of human research involving the ginseng berry and its extracts, establishing clinical safety remains essential. A literature search yields two clinical studies involving the ginseng berry; the first is a 12-week study examining the efficacy and safety of a berry extract on a glycemic control [142]. The extract-treated group of 34 patients did not have any statistical difference in measured safety parameters except for a decrease in diastolic blood pressure compared to the placebo group of 38 [142]. There was also no statistically significant difference in the occurrence or type of adverse events in this study [142]. The second clinical trial, which lasted 8 weeks, similarly did not report any adverse events related to the use of the berries in the 59 volunteers, nor any changes in blood biochemistry and hormone and lipid panels relative to the placebo [143]. It should be noted that both studies used Korean ginseng berries. In summary, the safety can be extrapolated from clinical research involving different parts of the American ginseng (i.e., the root) or from clinical research using Korean ginseng berries. Both perspectives suggest a good safety profile, but confirmation of the American ginseng berry’s safety through human trials remains undone.

6. Conclusions

By examining the American ginseng berry’s saponin profile and extrapolating from ginseng extract research, it can be determined that it has promising potential as a mental health-promoting nutraceutical. Indeed, through its neuroprotective and antidepressant effects that are amplified through microbial bioconversion, its microbiota-modulating effects that reverse deleterious alterations in composition, as well as promote the growth of beneficial bacteria, and finally, its positive effects on intestinal epithelium inflammation and tight junction protein expression, ginseng could broadly impact the gut–brain axis. However, research evaluating the berry’s efficacy is in short supply and limited to preclinical studies. Some questions remain regarding the berry’s non-saponin compounds, such as polysaccharides and their bioactivity, as research is scarce in this area. Considering the berry’s numerous advantages at the agricultural level, as well as potential advantages in terms of ginsenoside composition, the implementation of the ginseng berry culture and the increase in research evaluating the berry’s clinical efficacy are strongly encouraged.

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