INVESTIGATION

The Evolution and Consequences of Sex-Specific Reproductive Variance

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ABSTRACT Natural selection favors alleles that increase the number of offspring produced by their carriers. But in a world that is inherently uncertain within generations, selection also favors alleles that reduce the variance in the number of offspring produced. If previous studies have established this principle, they have largely ignored fundamental aspects of sexual reproduction and therefore how selection on sex-specific reproductive variance operates. To study the evolution and consequences of sex-specific reproductive variance, we present a population-genetic model of phenotypic evolution in a dioecious population that incorporates previously neglected components of reproductive variance. First, we derive the probability of fixation for mutations that affect male and/or female reproductive phenotypes under sex-specific selection. We find that even in the simplest scenarios, the direction of selection is altered when reproductive variance is taken into account. In particular, previously unaccounted for covariances between the reproductive outputs of different individuals are expected to play a significant role in determining the direction of selection. Then, the probability of fixation is used to develop a stochastic model of joint male and female phenotypic evolution. We find that sex-specific reproductive variance can be responsible for changes in the course of long-term evolution. Finally, the model is applied to an example of parental-care evolution. Overall, our model allows for the evolutionary analysis of social traits in finite and dioecious populations, where interactions can occur within and between sexes under a realistic scenario of reproduction.

IN the absence of mutation, the change in allele frequency is the result of natural selection and genetic drift. Natural selection favors alleles that maximize their representation within the gene pool, and a large body of work has investigated how alleles achieve this by increasing the expected number of offspring produced by an individual, thereby slowing down or even preventing adaptation altogether.

While many studies have investigated how natural selection affects the expected number of offspring produced by an individual, less attention has been given to the degree to which selection acts on the variance in offspring number, or reproductive variance. Gillespie (1974, 1975, 1977) investigated how natural selection dampens randomness in within-generation fertility in a haploid population. He demonstrated that between two alleles that on average produce the same number of offspring, natural selection favors the allele that produces offspring with lesser variance when the breeding adults of the next generation are sampled from a finite pool of offspring.

Reproductive variance also correlates with the intensity of genetic drift. By decreasing effective population size, reproductive variance mitigates the effect of selection (Wright 1931). Gillespie (1974)’s haploid model also revealed that the level of genetic drift affecting the segregation of two alleles increases with the reproductive variance they code for. As a consequence, fixation of the allele coding for lower fertility variance reduces the intensity of genetic drift and facilitates adaptive evolution in future generations.

The variance in fertility considered in Gillespie’s (1974, 1975, 1977) seminal articles had arbitrary causes and could have resulted from randomness at any stage of an individual’s life history, such as its development, its fertility, or the survival of its offspring. Extensions of Gillespie’s models have since investigated the effect of selection against reproductive variance...
in the context of more specific life histories. Shpak (2007) investigated the evolution of this variance in age-structured populations and showed that selection favors alleles that code for lower stochasticity in age-specific survival and fertility. Selection against reproductive variance has also been demonstrated to affect the evolution of traits as diverse as sex allocation in hermaphrodites (Proulx 2000), dispersal in structured populations (Shpak 2005; Shpak and Proulx 2007; Lehmann and Balloux 2007), and helping behaviors in social animals (Lehmann and Balloux 2007; Beckerman et al. 2011).

These models have highlighted that selection against reproductive variance may be a subtle yet significant force in the evolution of many different traits in natural populations (Rice 2008, for a general discussion). However, it remains unclear how selection on reproductive variance, and its feedback with genetic drift, affect the reproductive biology and life history of sexual organisms. Most models so far have omitted sex altogether. The only study that has taken into account selection on reproductive variance in a dioecious (two-sexes) population approximated reproduction as the random union of gametes and assumed that gamete production of different individuals was uncorrelated (Taylor 2009). These assumptions miss fundamental aspects of the reproductive biology of the vast majority of organisms, where it is individuals, rather than gametes, that unite to mate. But considering a realistic mating system would have significant consequences for the variances and covariances in offspring number among individuals of one sex and how these (co)variances differ across the sexes (Bateman 1948; Wade 1979, for examples). If the effect of the mating system on genetic drift has successfully been captured by calculations of the effective population sizes (Nunney 1993; Nomura 2002), a more general approach is needed to make evolutionary predictions that incorporate selection acting on the traits that generate this variance.

In this article, we present a population-genetic model of male and female phenotypic evolution that makes it possible to predict the evolution of sex-specific reproductive traits under the influence of selection and genetic drift. Using an individual-based approach, the model incorporates a description of the mating system based on the first and second moments (means and (co)variances) of the distribution of individual offspring number. The article is divided into two broad sections. In the first part of the article, we present our model. We start by deriving the probability of fixation of a mutant allele that affects male and/or female reproduction in a finite dioecious population. Our derivation accounts for sex-specific levels of reproductive variance, as well as for covariances between members of the same sex. We then extend the analysis of short-term evolutionary change by deriving predictions of long-term phenotypic evolution, which in turn makes it possible to calculate the equilibrium for sex-specific phenotypes based on the probability of fixation and properties of the mutational input. In the second part of the article, we provide an illustration of how our model can be applied and study the effects of sex-specific reproductive variance on the evolution of various parental-care strategies. These simple examples allow us to demonstrate that sex-specific reproductive variance can lead to differences between the probability of fixation for mutants affecting female traits and those affecting male traits and between the phenotypic equilibria of these traits.

Model and Results

Probability of fixation in dioecious populations

We derive the probability of fixation of a mutant allele (A) introduced as a single copy into a population fixed for a resident allele (a). The population is dioecious and of any but constant, size, with \( N_m \) adult males and \( N_f \) females. Generations are nonoverlapping. The mutant A allele alters the expression of a continuously varying phenotypic trait, which may affect one or more aspects of reproductive biology, such as mating success, fecundity, or offspring survival. The trait can have different values in males and females and we denote by \( z_m \) the phenotypic value of a male homozygous for the resident allele (genotype aa). The phenotype of a heterozygous male (Aa) is denoted by \( z_m + h\delta_m \), where \( h \) is the dominance coefficient of A. A male homozygous for the mutant allele (AA) has phenotype \( z_m + \delta_m \). Similarly, the phenotypes of the three genotypes in a female are \( z_f \) (aa), \( z_f + h\delta_f \) (Aa), and \( z_f + \delta_f \) (AA).

Weak selection approximation: The fixation probability of the mutant is derived using an individual-based approach that builds on previous works (Rousset 2003; Roze and Rousset 2004; Lessard and Ladret 2007; Lehmann and Rousset 2009; and supporting information, File S1). Under weak selection (small mutant deviations \( \delta_m \) and \( \delta_f \)), and if the mutation rate is the same in males and females, the fixation probability of a single mutant copy arising at random in a monomorphic population with phenotypes \( z_m \) in males and \( z_f \) in females is

\[
\pi(z_m, z_f, \delta_m, \delta_f, h) = \frac{1}{2N} + \delta_m K_m(z_m, z_f, h) G_m(z_m, z_f) + \delta_f K_f(z_m, z_f, h) G_f(z_m, z_f) + O(\delta^2),
\]

where \( N = N_m + N_f \) is the total number of adults, and \( \delta \) is such that \( \delta_m \sim O(\delta) \), \( \delta_f \sim O(\delta) \) (File S1, Equations SI.1–SI.39). The functions \( G_m(z_m, z_f) \) and \( G_f(z_m, z_f) \) are fitness gradients: they measure the effect of the mutant on male and female fitness and are further explained below (Fitness gradients section). The functions \( K_m(z_m, z_f, h) \) and \( K_f(z_m, z_f, h) \) are measures of the variance of mutant frequencies in males and females over the segregation process, from the appearance until the eventual fixation of the mutant. They are inversely proportional to the intensity of genetic drift and capture the efficacy with which selection acts on the mutant (see Efficacy of selection section). Whether a mutant is under positive selection \([\pi > 1/(2N)]\), evolves neutrally
They are given by

\[ G_m(z_m, z_l) = \left( \frac{\partial w_m}{\partial z_{m-i}} (z_m, z_{m-i}, z_l) \right) \frac{N_m}{N_l} \frac{\partial w_m}{\partial z_{m-i}} (z_m, z_{m-i}, z_l) \]

and

\[ G_l(z_m, z_l) = \left( \frac{\partial w_l}{\partial z_{l-i}} (z_l, z_{l-i}, z_m) \right) \frac{N_l}{N_m} \frac{\partial w_l}{\partial z_{l-i}} (z_l, z_{l-i}, z_m) \]

where \( w_m^u \) is the expected number of adult offspring of sex \( u \) of a focal individual of sex \( v \) \( \in \{m, f\} \). This fitness function depends on the phenotype \( z_v \) of the focal individual of sex \( v \), on the average phenotype \( \bar{z}_{-v} = \frac{\sum z_{-v}}{N_v} \) among sex \( v \), but excluding the focal, and on the average phenotype \( z_l \) in the population of the opposite sex \( \bar{z}_m = \frac{\sum z_{m}}{N_m} \) for males and \( \bar{z}_f = \frac{\sum z_{f}}{N_f} \) for females. The model can thus easily accommodate for sex-specific interactions based on games, like the classic battle of the sexes. The derivatives of focal fitness \( w_v^u \) are evaluated at the resident phenotypes \( z_{m-i} = \bar{z}_{-m} = \bar{z}_m, z_{l-i} = \bar{z}_{-l} = \bar{z}_l \), so that \( G_m \) and \( G_l \) measure the effects of phenotypic changes on male and female fitness with respect to the resident population.

The fitness gradients in Equation 2a indicate the direction of phenotypic evolution in each sex that is favored by selection. If \( G_m(z_m, z_l) \) and \( G_l(z_m, z_l) \) are positive, then selection favors an increase in the trait in males and females; if they are negative, then selection favors a decrease. Although the gradients are derivatives of the expected average number of adult offspring produced \( (w_m^u \text{ and } w_l^u) \), the direction of selection depends on how the phenotype affects the average number of juvenile offspring produced as well as reproductive variance. To demonstrate why this is the case, we derive the fitness of a focal individual \( i \) of sex \( v \) in terms of the distribution of juveniles in the population. Fitness is the expected number of \( i \)'s offspring that become part of the adult breeding population of the next generation. We separate fitness gained through male and female offspring. We write \( J_v^u \) for the number of juvenile offspring of sex \( u \) born to \( i \), itself of sex \( v \). In each generation, the set of reproductive individuals is established by independently sampling \( N_m \) males from a pool of surviving male offspring and \( N_f \) females from a pool of surviving female offspring. The conditional fitness of individual \( i \) of sex \( v \) gained through offspring of sex \( u \), i.e., the expected number of its adult offspring of sex \( u \), can then be calculated in terms of \( i \)'s reproductive success, relative to that of the total population

\[ w_v^u | J_v^u = N_u \frac{J_v^u}{w_v^u} + \sum_{k \neq i} J_v^u_k, \]

where \( J_v^u = (J_v^u_1, J_v^u_2, \ldots, J_v^u_k) \) is the realized offspring production of all parents in the population. Note that the total number of juveniles of either sex must be the same when counted as the offspring of males or females (i.e., \( \sum_v J_v^u = \sum_u J_u^v \)). We assume nonextinction of the population \( \sum_k J_v^u_k \geq N_v \). To describe the fitness of individual \( i \), we need to calculate the expectation of Equation 3 over the distribution of \( J_v^u \). Following the approach of previous work (Gillespie 1975; Proulx 2000; Shpak and Proulx 2007; Lehmann and Balloux 2007; Rice 2008), \( E[w_v^u | J_v^u] \) is approximated using the delta method (Oehlert 1992), so that the expected fitness \( w_v^u \) becomes

\[ w_v^u = N_u \left( \mu_v^u - \mu_v^u - D_v^u + \mu_v^u \frac{\mu_v^u}{\mu_v^2} \sum_{k \neq i} J_v^u_k \right) + R, \]

where \( \mu_v^u = \sum u J_v^u \) is the expected number of juveniles of sex \( u \) produced by individual \( i \), \( \mu_v^u = \sum u J_u^v \) is the expected total number of juveniles of sex \( u \) produced in the population, \( v^u \) is the variance of the number of offspring of individual \( i \) \((v^u_i = V(J_v^u_i)) \), and \( R^{u}_{v,k} \) is the covariance between the number of offspring of sex \( u \) of individuals \( i \) and \( k \) of sex \( v \) \( (R^{u}_{v,k} = C(J_v^u_i, J_v^u_k)) \). The fitness of an individual therefore takes into account all first and second moments of the probability distribution that describes individual reproduction (see Figure 1 for a depiction of those moments for a focal male).

The remainder \( R \) in Equation 4 is composed of central cross moments of \( J_v^u \) of order 3 and higher. These terms may be significant in certain scenarios (Rice 2008), but we omit them in this analysis by assuming that the distribution of \( J_v^u \) is well behaved as the population size \( N \) increases. Previous models used the central limit theorem to justify this assumption (Shpak and Proulx 2007; Lehmann and Balloux 2007, Equation A6). This is not strictly valid here because the number of offspring produced by different individuals is not necessarily independent. However, the remainder terms can be ignored if we assume that offspring numbers are close to dependence and that the “total” covariance between a given set of individuals decreases as the number of individuals in that set increases (see Appendix, Sex-Specific Reproductive Variance 237).
Figure 1 The moments of male reproduction. At generation \( t \), a focal male \( i \) sires an expected \( \mu_{mi} \) number of male offspring with variance \( \nu_{mi} \) and covariance \( \rho_{mik} \) with the number of male offspring sired by another male \( k \). The males of the next generation \( t + 1 \) are established by sampling the juveniles and the expected number of adult males of the focal male \( i \) is \( w^m_{mi} \). Similarly, the focal male sires an expected \( \mu_{vi} \) number of female offspring with variance \( \nu_{vi} \) and covariance \( \rho_{vik} \) with the number of females sired by male \( k \). Then, the expected number of adult females of the focal male \( i \) is \( w^f_{vi} \). Finally, the number of male and female offspring of male \( i \) covary by \( \rho^m_{mi} \).

Equation A1). In this case, the remainder terms in Equation 4 are of order \((1/N^2)\). Therefore, while the expression for the fixation probability (Equation 1) holds for any population size, the approximation for fitness (Equation 4) takes into account only the first-order effects of finite population size on offspring means and variances. If condition (A1) also holds for the first and second moments of the distribution of reproduction, then the effects of (co)variances on individual fitness vanish as \( N \to \infty \), in agreement with previous studies (Gillespie 1974).

Equation 4 shows that individual fitness depends on four terms. The first is the relative expected number of offspring produced (\( \mu_{vi}/\mu_{m1} \)), which has a positive effect on fitness. The remaining three terms capture the effects of reproductive variance. Fitness decreases with the variance in offspring number (\( \nu_{vi} \)), increases with the variance in offspring number produced by the remaining individuals in the population \((\sum_{k \neq i} \mu_{vk}^l \text{ and } \sum_{k \neq i} \rho_{vkl}^l)\), and decreases with the covariance between the number of offspring produced and that of the remaining individuals in the population \((\sum_{k \neq i} \rho_{mik}^l)\).

The fitness effects of the variance terms stem from the nonlinear relationship between fitness and the offspring production of the focal (\( J^l_{mi} \) see Figure 2 and Equation 3) and the rest of the population \((\sum_{k \neq i} J^l_{vk})\) see Figure 2B and Equation 3). For a given number of offspring produced by the rest of the population, the fitness returns of a focal individual diminishes with its production of more offspring as a consequence of the increased competition between related juveniles for access to breeding. This results in a net negative effect of variance in the focal individual’s reproductive output on its fitness \( (\nu_{vi} \text{ in Equation 4 and Figure 2A})\). Conversely, for a given offspring production by the focal, the advantage of competing within a population of individuals that are on average less fecund is expected to be greater than the disadvantage of competing in a more productive population. This leads to a net positive effect of population variance on the focal individual’s fitness \((\sum_{k \neq i} \nu_{vk}^l \text{ and } \sum_{k \neq i} \rho_{vkl}^l)\) in Equation 4 and Figure 2B). Finally, using arguments similar to those presented in Figure 2, one can see that the benefit of overperforming in a less competitive population is on average greater than the cost of underperforming in a more competitive population. As a consequence, the covariance between the numbers of offspring produced by the focal individual and the rest of the population has a negative impact on focal fitness \((\sum_{k \neq i} \rho_{mik}^l)\) in Equation 4).

Selection on a phenotype (Equation 2) then reflects the balance between the impact of the trait on the different terms of Equation 4. Since the difference between two phenotypes are small [of order \( O(\delta^2) \)], we can describe the dependence between the moments and phenotypes without explicitly characterizing the interactions between every individual in the population. Rather, we average the sums in Equation 2 over mean population phenotypes (File S1, Equation S1.4). Then, if the trait of interest affects all first and second moments of individual reproduction, the fitness function of a focal individual of sex \( v \) can be written as

\[
W^l_{vi} = \frac{\mu^l_{vi}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})}{\mu^l_{vi}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})} - \frac{1}{N_{vi}} \frac{\nu^l_{vi}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})}{\mu^l_{vi}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})} - \frac{\mu^l_{ki}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})}{\mu^l_{ki}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})} - \frac{\nu^l_{ki}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})}{\mu^l_{ki}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi})} + O(1/N^2, \delta^2),
\]

(5)

where \( \bar{z}_{0 vi} \) denotes the average phenotype \( \bar{z}_{0 vi} \) of the sex opposite to that of the focal, \( v_i \) (e.g., \( \bar{z}_{0 vi} = \bar{z}_{mi} \)). The function \( \mu^l_{vi}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi}) \) is the expected number of juveniles of sex \( u \) produced by a focal of sex \( v \) and \( \mu^l_{ki}(\bar{z}_{vi}, \bar{z}_{-vi}, \bar{z}_{0 vi}) \) is the average expected number of juveniles of sex \( u \) produced by sex \( v \) individuals in the population; similar interpretations are given to the variance and covariance functions \( (\nu^l_{vi} \text{ and } \rho^l_{vi}) \). Therefore, calculating the individual fitness functions \( W^l_{vi} \) that go into the fitness gradients (Equation 2) requires characterizing only the individual mean, variance, and covariance functions, \( (\mu^l_{vi}, \nu^l_{vi}, \text{ and } \rho^l_{vi}) \), and these depend only on the phenotype of the focal individual \( (\bar{z}_{vi}) \), the average phenotype in the opposite sex \( (\bar{z}_{0 vi}) \), and the average among other individuals of the same sex \( (\bar{z}_{-vi}, \text{ as the average } z_{\bar{v}i} \text{ is written as } \bar{z}_{vi} = (N_{vi} - 1)\bar{z}_{-vi}/N_{vi} + z_{0 vi}/N_{vi}) \). Examples of such calculations are given in the Example section.
number of offspring produced because sibs also compete against each other. Then, the benefits reaped when it produces more offspring than average (gray arrow) are outweighed by the cost when it produces less (black arrow). (B) Fitness of a focal individual graphed against the random number of offspring produced by the rest of the population and by holding the number of offspring of the focal constant. The rest of the population produces on average μ, offspring with variance σ². It then produces more or less than μ, offspring. But fitness is a relative measure of reproductive success (Equation 3). The advantage of producing more offspring depreciates with the

The first line in Equation 5 reflects the fact that an individual who produces on average a greater number of offspring than the average individual in the population has higher fitness. The second and third lines reflect the fact that an individual with a lower variance in progeny number than the average individual has higher fitness, as originally described by Gillespie (1974). Finally, the last two lines of Equation 5 reflect the fact that an individual whose offspring production covaries with that of another individual to a lesser degree than the average individual also has higher fitness. In addition, we see that the effect of covariance on fitness of order (N, − 1)/N, is potentially greater than that of the variance (of order 1/N).

Efficacy of selection: In addition to the fitness gradients, the fixation probability (Equation 1) also depends on Kₘ(zₘ, zₙ, h) and Kₙ(zₙ, zₙ, h), which weigh on the fitness gradients and measure the sex-specific efficacy of selection in males and females, respectively. The weight Kₘ(Kₙ) is the expected covariance between the phenotype in males (females), cumulated over the neutral segregation of the mutant. Mathematically, this is

$$K_u = \frac{1}{2N} \sum_{t=0}^{\infty} \mathbb{E}
C[z_{ui}, p_{ui}, t],$$

where C[zₜ, pₜ] is the covariance between the phenotype zₜ of an individual of sex u and the frequency pₜ of the mutant at generation t, and \(\mathbb{E}[\cdot]\) denotes expectation under neutral evolution, i.e., where only genetic drift affects fluctuations of genotypic frequency pₜ from one generation t to the next (File S1, Equations SI.20, SI.21, and SI.39). The factor 1/(2N) in Equation 6 is the initial frequency of the mutant, while the factor 1/4 is the product of the frequency of transmission of a gene by a parent to an offspring (i.e., 1/2; File S1, Equation SI.2) and the reproductive value of that class of offspring, which is here 1/2 for both males and females. If mutant effects are additive (h = 1/2), then C[zₜ, pₜ] = V[pₜ], and Kₜ reduces to the cumulative genetic variance in sex u, highlighting that the larger genetic variance is, the more efficient selection can be. In the simple case of asexual haploids, this variance reduces to the familiar \(\text{p}_t(1 - \text{p}_t)\), where \(\text{p}_t\) is the average frequency of the mutant at generation t. More generally, Kₘ(Kₙ) depends on dominance and captures the association between phenotypic and genetic variance in males (females) on which selection is then able to act.

Since the calculations for Kₘ and Kₙ are made over the segregation of a neutral mutant (Equation 6), they can be expressed in terms of coalescence times of neutral genes (File S1, Equations SI.37–SI.38), which themselves can be expressed in terms of how genes coalesce within individuals of different sexes. Doing so links Kₘ and Kₙ back to the mating system and hence to the evolving reproductive traits zₘ and zₙ that are under study. The general expressions for Kₘ and Kₙ in terms of the coalescence process depend on the probabilities that individuals share the same parent in the absence of selection, referred to here as “probabilities of sibship.” We find that the coalescence process can be described by 14 probabilities of sibship. Six of these describe the probability that a pair of individuals share the same parent; they are written as Θₘ', where \(u \in \{m, f, c\}\) indicates whether a pair of individuals consists of two males, two females or a male and a female (Figure 3). The remaining eight probabilities of sibship describe the probability of three individuals (three males, two males and a female, two females and a male, or three females) sharing the same parent (male or female). Providing a general characterization of the neutral coalescence process is complicated, but the system can be simplified by taking into account only the first-order effects of finite population size [O(1/N)]. In this case, we find that the eight three-way probabilities of sibship may be expressed in terms of the pairwise probabilities Θₘ'’s (File S1, Equations SI.40–SI.42), which are then sufficient to describe the entire coalescence process.

The pairwise probabilities of sibship Θₘ' capture different aspects of reproductive variance. The probabilities that two males have the same father (Θₘ'), that two females have the same father (Θₘ'), and that a male and female have the same
The paternal probabilities of sibship. With probability $\Theta_m^s$, two males sampled at generation $t+1$ have the same father from generation $t$. So, with probability $1 - \Theta_m^s$, they come from different fathers. Similarly, a male and a female sampled at generation $t+1$ have the same father with probability $\Theta_f^s$, and two females do so with probability $\Theta_f^s$.

Figure 3: The paternal probabilities of sibship. With probability $\Theta_m^s$, two males sampled at generation $t+1$ have the same father from generation $t$. So, with probability $1 - \Theta_m^s$, they come from different fathers. Similarly, a male and a female sampled at generation $t+1$ have the same father with probability $\Theta_f^s$, and two females do so with probability $\Theta_f^s$.

The probability of fixation in the face of increased drift (Equation 1).

Similar patterns are observed when reproductive variance varies with the sex of the parent and the sex of the offspring. Analytical results for additive mutants (Equation A2, Figure 4, A–C) and numerical results for nonadditive mutants show that $K_m$ and $K_f$ both decrease hyperbolically with all six probabilities of sibship. Numerical results also show that $K_m$ and $K_f$ increase linearly with dominance (Figure 4D). This stems from the fact that dominance increases the phenotype-genotype covariance at lower allele frequencies ($p_{ui} < 1/2$) and that the frequency of neutral mutants remains on average low.

Because $K_m$ and $K_f$ weight male ($G_m$) and female ($G_f$) fitness gradients independently in the probability of fixation (Equation 1), reproductive variance may affect male and female evolution differently. For example, selection on females has a greater impact on the probability of fixation than selection on males when $K_m < K_f$. In this case, a female-limited mutation ($\delta_m = 0$ or $G_m = 0$) would have a greater chance of reaching fixation than a male-limited mutation ($\delta_f = 0$ or $G_f = 0$), even if both improve fitness by the same amount. In the longer term, we would then observe a faster rate of adaptation in females than males. The reverse patterns are predicted when $K_m > K_f$.

Differences between $K_m$ and $K_f$ and hence differences between the efficacy of selection in males and females, occur whenever genetic variance is lower in one sex than in the other. For additive mutants (Equation A2 and Figure 4, A and B), $K_m < K_f$ requires that

$$\Theta_m^s + \Theta_f^s > \Theta_f^s + \Theta_f^s, \quad (8)$$

i.e., the probability that two males have at least one parent in common exceeds the probability that two females have at least one parent in common. This inequality (Equation 8) reflects that if at each generation male offspring are more related than female offspring, then genetic variance in males is lower than in females, and as a consequence, selection is less efficient in males.

Calculating the probability of fixation: Based on the above derivations, the probability of fixation can be explicitly calculated, taking into account the fitness change caused by the mutant, through its effect on first and second moments.

**Table 1 The probabilities of sibship**

| $\nu$ | $m$ | $f$ | $c$ |
|-------|-----|-----|-----|
| $\Theta_m^s$ | 1/N$_m$$(1 + \nu_m^s/(\mu_m^s)^2)$ | 1/N$_m$$(1 + \nu_m^s/(\mu_m^s)^2)$ | 1/N$_m$$(1 + \mu_m^t/\mu_m^s\mu_m^c)$ |
| $\Theta_f^s$ | 1/N$_f$$(1 + \nu_f^s/(\mu_f^s)^2)$ | 1/N$_f$$(1 + \nu_f^s/(\mu_f^s)^2)$ | 1/N$_f$$(1 + \mu_f^t/\mu_f^s\mu_f^c)$ |

The first row gives the paternal probabilities of sibship and the second row gives the maternal probabilities of sibship. The moments $\mu$ and $\nu$ terms are defined in the main text, except for $\rho_{ui}^m = C(p_{ui}^m, f_{ui}^m)$, which is the covariance between the number of male and offspring juveniles fathered by a resident male (Figure 3), and $\rho_{ui}^f = C(p_{ui}^f, f_{ui}^f)$, which is the covariance between the number of male and female offspring a resident female gives birth to. Therefore, the probabilities of sibship increase with reproductive variance.
of the distribution of offspring production and the impact of segregation in the two sexes on the efficacy of selection. To calculate the fixation probability, the probabilities of sibship (Table 1) are substituted into $K_m$ and $K_f$ (Equation A2 if the mutant is additive and File S1, SI.38 otherwise), and the expressions for focal fitness (Equation 4) are substituted into the fitness gradients $G_m$ and $G_f$ (Equation 2). Finally, $K_m$, $K_f$, $G_m$, and $G_f$ are substituted into the fixation probability (Equation 1).

**Long-term phenotypic evolution in dioecious populations**

The fixation probability of a mutant is useful for predicting short-term evolution and to understanding how the interplay between selection and genetic drift affects the fate of a new mutation. However, it is often desirable to predict the long-term evolution of phenotypes as a result of selection and drift acting on an influx of new mutations. In this section, we use the probability of fixation (Equation 1) to determine the phenotypes most likely to be observed in males and females at a selection–mutation–drift balance. To that aim, we assume that the autosomal locus mutates at a constant rate $\xi$. This rate is sufficiently weak compared to the rate of fixation to ensure that there are only ever two alleles segregating, thus complying with the weak-mutation strong-selection limit of population genetics (e.g., Gillespie 1994; Sella and Hirsh 2005) and/or the trait substitution sequence limit of evolutionary game and inclusive fitness theory (e.g., Metz et al. 1995; Champagnat and Lambert 2007; Lehmann 2012). The effects $(\delta_m, \delta_f, h)$ of a mutation are drawn from a distribution $u(\delta_m, \delta_f, h)$, which is such that the dominance $(h)$ of a mutant is independent from its homozygotic effects $(\delta_m, \delta_f)$, and mutants have on average no phenotypic effect $E[\delta_m] = E[\delta_f] = 0$. The rate at which a population monomorphic for $(z_m, z_f)$ is substituted by a population with traits $(z_m + \delta_m, z_f + \delta_f)$ can then be written as

$$k(z_m, z_f, \delta_m, \delta_f, h) = 2Nx u(\delta_m, \delta_f, h) \left( \frac{1}{2N} + \delta_f \frac{\partial \pi}{\partial \delta_f} + \delta_m \frac{\partial \pi}{\partial \delta_m} \right) + O(\delta^2),$$  \tag{9}$$

where $2N$ is the number of gene copies in the population, $u(\delta_m, \delta_f, h)$ is the probability that a single copy produces a mutation of type $(\delta_m, \delta_f, h)$, and the term within brackets is the probability that this mutation fixes in the population, which is given by Equation 1.

The substitution rate $k$ determines a jump process (Gardiner 2009), which describes the stochastic evolution of male and female traits as jumps between monomorphic states in phenotypic space. Ignoring terms of order $O(\delta^2)$ in Equation 9, the jump process can be described in continuous time by a diffusion process that eventually reaches a stationary distribution $\psi(z_m, z_f)$ (Appendix, Diffusion equation for phenotypic evolution in dioecious populations). This long-run stationary state reflects a balance between the forces of mutation, selection, and genetic drift, and the maxima of $\psi(z_m, z_f)$ correspond to phenotypes around which the populations spend the greatest amount of time. These maxima are the most likely outcomes of phenotypic evolution, and in single phenotype models, they are the “convergence stable” states of the system (Lehmann 2012). A phenotype is convergent stable if populations sitting close to this phenotype are attracted toward it. Convergence stability is an important concept of attainability of equilibrium points, common to evolutionary game and inclusive fitness theory (Rousset and Billiard 2000; Leimar 2009). Under the trait-substitution sequence limit, a phenotype that is convergent stable is also evolutionary stable (Wakano and Lehmann 2012).
Phenotypes that are multidimensional convergence stable can be found by considering the attractor points of the system of differential equations

\[
\frac{dz_m}{dt} = 2N\xi(\beta_{zm}K_m(z_m, z_l, \bar{H})G_m(z_m, z_l) + b_{zm}K_f(z_m, z_l, \bar{H})G_f(z_m, z_l))
\]

\[
\frac{dz_l}{dt} = 2N\xi(\beta_{zl}K_m(z_m, z_l, \bar{H})G_m(z_m, z_l) + b_{zl}K_f(z_m, z_l, \bar{H})G_f(z_m, z_l))
\]

(10)

which describes the deterministic trajectory associated to the underlying diffusion process. Here, \(\bar{H} = E[h]\) is the average dominance of the mutation distribution and \(b_{zm} = C[\delta_u, \delta_z]\) is the covariance between mutation effect in sex u and v. If none of the attractor points \((z_{m, l}^*, z_{l}^*)\) of system Equation 10 lie on the boundary of the phenotypic space, then large deviation theory shows that as the population size grows, the stationary distribution \(\psi(z_m, z_l)\) becomes peaked around these attractor points (use Theorem 4.3 of Freidlin and Wentzell 2012, p. 170, and observe that if none of the attractor points \((z_{m, l}^*, z_{l}^*)\) lies on the boundary of the phenotypic space, then a smooth domain can be drawn around all attractor points, thereby satisfying condition A of Freidlin and Wentzell 2012, p. 150). Therefore, when all attractor points \((z_{m, l}^*, z_{l}^*)\) of Equation 10 lie in the interior of the phenotypic space, they correspond to the convergence stable states and are the most likely phenotypic outcomes of evolution. Furthermore, in the infinite size limit \((N \rightarrow \infty)\), the stationary distribution becomes fully concentrated around a single of these convergence stable states (Theorem 4.2 of Freidlin and Wentzell 2012, p. 167), which corresponds to the highest peak of the adaptive landscape and the stochastic stable state of the system (e.g., Foster and Young 1990; Van Cleve and Lehmann 2013).

For an interior point \((z_{m, l}^*, z_{l}^*)\) of system Equation 10 to be convergence stable, two conditions must be satisfied. First, \(dz_m/dz = 0\) and \(dz_l/dz = 0\) must hold, but since \(K_m(z_m, z_l, \bar{H}) > 0\) and \(K_f(z_m, z_l, \bar{H}) > 0\), this condition is equivalent to

\[
G_m(z_{m, l}^*, z_{l}^*) = G_f(z_{m, l}^*, z_{l}^*) = 0,
\]

i.e., that the male and female fitness gradients vanish, which is equivalent to the condition for establishing singular points in deterministic evolution (Leimar 2009). Second, the real part of all the eigenvalues of the Jacobean matrix of Equation 10 must be negative. Combined with Equation 11 and the properties of \(b_{uw}\) (Leimar 2009), this latter condition is equivalent to the real part of the eigenvalues of

\[
\begin{bmatrix}
K_m \partial G_m / \partial z_m & K_m \partial G_m / \partial z_l \\
K_f \partial G_f / \partial z_m & K_f \partial G_f / \partial z_l
\end{bmatrix}
\]

(12)

being negative at \((z_{m, l}^*, z_{l}^*)\). Conditions (11) and (12) extend the one-dimensional condition of convergence stability for finite populations (Rousset and Billiard 2000; Lehmann 2012), and when \(K_m = K_f\), it is equivalent to the condition for multidimensional convergence stability for populations of infinite size (Leimar 2009), which depends only on the fitness gradients \(G_m\) and \(G_f\). When attractor points \((z_{m, l}^*, z_{l}^*)\) of Equation 10 lie on the boundary of the phenotypic space or outside of it, the shape of the equilibrium distribution cannot in general be assessed (to the best of our knowledge), and in this case, characterizing convergence stable points is not straightforward.

Example

In this section, we illustrate a possible application of our model by analyzing the evolution of maternal and paternal care behaviors. The emphasis is on highlighting the effects of selection on reproductive variance in driving the evolution of reproductive traits. We consider a dioecious species with a simple life cycle. An equal number \(N\) of adult males and females randomly pair up to mate. All females mate once with a single male, but males mate with harems of exactly \(H\) females. If \(H = 1\), the population is monogamous and all males mate. If \(H > 1\), then the population is polygynous and some males mate \(H\) times while others do not reproduce at all. Each female gives birth to exactly \(f\) offspring with an equal sex ratio. The offspring survive with probabilities that depend on the phenotypes \(z_m\) and \(z_l\) of their parents. Female offspring each survive independently from each other with probability \(s_f(z_m, z_l)\). In contrast, the survival of males is strongly correlated within broods and the males of a brood either all survive [with probability \(s^m(z_m, z_l)\)] or all die [with probability \(1 - s^m(z_m, z_l)\)]. This is close to the assumptions of the hermaphroditic model of Proulx (2000). The difference in male and female survival could reflect sex differences in the susceptibility to random variation in the breeding environment provided by the male’s territory, for example. As a consequence of their correlated survival, surviving males are more related than surviving females. The next generation of adults is randomly sampled from the population of surviving offspring as described in the previous section.

The phenotypes that evolve are the level \(z_m\) of paternal care provided by a male and the level \(z_l\) of maternal care provided by a female. The survival probabilities of daughters, \(s_f(z_m, z_l)\), and sons, \(s^m(z_m, z_l)\), are both increasing functions of \(z_m\) and \(z_l\). We analyze the fate of four different types of mutations altering the parental-care phenotypes. These types are characterized by the sex of the parent providing the care and the sex of the offspring receiving it. We distinguish between mutations that affect care for sons only and those that affect care for daughters only. For both of those, we consider sex-limited mutations that affect care only in males or only in females. In total, the evolution of four different traits is studied, paternal care for daughters, maternal care for daughters, paternal care for sons, and maternal care for sons.

The covariance between the survival of male offspring depends on the sex of the parent for which care evolves. If female care is evolving, then for each female, her entire male brood either survives or dies, independently from other females, even from those that have mated with the same male. When care is provided by males, then for each male,
his entire male brood either survives or dies, independently from other males, but his brood includes all the male offspring he has had with different females. We assume that the effect of mutations is additive and identical for all traits. Thus, compared to the resident homozygote, the level of care is increased by an amount \( \delta/2 \) in heterozygotes and by \( \delta \) in mutant homozygotes.

**Probability of fixation of new mutants**

To calculate the fitness of a focal individual (Equation 5) and evaluate the fixation probability of the different mutants, we need to express the means and covariances of offspring production in terms of care phenotypes. We first consider the offspring production of a focal female with phenotype \( z_f \). Because the evolution of male and female traits are treated separately, mutations for altered maternal care always occur in the presence of constant resident paternal care \( z_m \), and we can therefore omit paternal care from the survival functions. The expected number of daughters (\( \mu^f \)) and sons (\( \mu^m \)) the focal female produces are given by

\[
\mu^f_f(z_f) = \frac{f^f(z_f)}{2},
\]

(13a)

\[
\mu^m_f(z_f) = \frac{f^m(z_f)}{2},
\]

(13b)

where the factor 1/2 stems from the equal sex ratio.

The variance terms that contribute to the fitness of a focal female are found by writing \( n_0 \) and \( n_z \) as the number of daughters and sons, respectively, at birth before survival selection. With the sex ratio being equal, \( n_0 \sim \text{Bin}(f, 1/2) \) and \( n_z = f - n_0 \). Given \( n_0 \), the variance in the number of female offspring born to a female is \( n_0s^f(z_f)(1 - s^f(z_f)) \), and the mean is \( n_0s^f(z_f) \). Therefore, applying the law of total variance, the variance in the number of female offspring born to a focal female is

\[
\nu^f_f(z_f) = E_{n_0} [n_0s^f(z_f)(1 - s^f(z_f))] + V_{n_0} [n_0s^f(z_f)]
\]

\[
= \frac{s^f(z_f)}{2} \left( 1 - \frac{s^f(z_f)}{2} \right).
\]

(14)

In contrast, since sons do not survive independently from each other, we have, given \( n_z \), that the variance in the number of male offspring is \( n_zs^m(z_f)(1 - s^m(z_f)) \) and that the mean is \( n_zs^m(z_f) \). Thus, the variance in the number of sons produced by a focal female differs from the variance in the number of daughters and is

\[
\nu^m_f(z_f) = E_{n_z} [n_zs^m(z_f)(1 - s^m(z_f))] + V_{n_z} [n_zs^m(z_f)]
\]

\[
= \frac{s^m(z_f)}{4} \left( 1 + f s^m(z_f)(1 - s^m(z_f)) \right).
\]

(15)

Finally, since females give birth to and care for their offspring independently of one another, the covariance between the number of offspring of two females is zero (\( \rho^f_f = \rho^m_m = 0 \)).

The means and variances of the offspring numbers produced by a focal male \( i \) are obtained similarly (see Appendix, Calculations for the evolution of parental care), but in contrast to singly mating females, polygyny leads to a negative covariance between the numbers of offspring sired by two males. The additional variance terms (\( \rho^m_m \) for male offspring and \( \rho^f_f \) for female offspring) must be taken into account when determining the fitness of a focal male and are calculated here. Since maternal care is now constant, we can omit the female care phenotype \( z_f \) from the survival functions. By definition, the covariance between the number of male offspring fathered by the focal male \( i \) and an “average” male other than \( i \) is

\[
\rho^m_m(z_m, \bar{z}_m) = \langle E[N_{mat_{i}}N_{mat_{-i}}] - 1 \rangle \frac{f^2}{4} s^m(z_m)s^m(\bar{z}_m),
\]

(16)

where \( E[N_{mat_{i}}N_{mat_{-i}}] \) is the expected product between the number of matings of the focal male \( i \), \( N_{mat_{i}} \), and that of another male, \( N_{mat_{-i}} \). Then, since \( N/H \) males are chosen at random without replacement to mate exactly \( H \) times, \( E \)

\[
E[N_{mat_{i}}N_{mat_{-i}}] = (N - H)/(N - 1),
\]

and

\[
\rho^m_m(z_m, \bar{z}_m) = \frac{(N - H)}{(N - 1)} \frac{f^2}{4} s^m(z_m)s^m(\bar{z}_m).
\]

(17)

Similarly, the covariance number of females fathered by the focal male \( i \) and an average male is

\[
\rho^f_f(z_f, \bar{z}_f) = \frac{(N - H)}{(N - 1)} \frac{f^2}{4} s^f(z_f)s^f(\bar{z}_f).
\]

(18)

As expected, these covariances vanish in a monogamous population (\( H = 1 \)), but become increasingly negative as fewer males mate. For large \( H \), they contribute significantly to the fitness of a focal male.

**Fitness gradients:** Substituting Equations 13a and 14 into Equation 5 and deriving according to Equation 2b give the fitness gradient for alleles that code for maternal care of daughters. Similarly, substituting Equations 13b and 15 into Equation 5 and differentiating according to Equation 2b give the fitness gradient for alleles that code for paternal care of sons. The fitness gradients for alleles that code for paternal care are obtained in the same way using Equation 2a.

To identify the different effects of sex-specific reproductive variance, we first consider a population that is strictly monogamous (\( H = 1 \)). If maternal and paternal care have the same effect on offspring survival [i.e., \( \partial s^f(z_m, z_f)/\partial z_f = \partial s^m(z_m, z_f)/\partial z_m \)], the fitness gradients for mutants that increase maternal (\( G_f \)) or paternal (\( G_m \)) care of daughters are identical and equal to

\[
G_u(z_m, z_f) = \frac{\partial s^f(z_m, z_f)}{\partial z_u} \frac{1}{s^f(z_m, z_f)} \left[ \left( 1 - \frac{1}{N} \right) + \frac{1}{Nf} \right].
\]

(19)

where \( u \in \{ m, f \} \) and \( z_m \) and \( z_f \) are the resident levels of paternal and maternal care. Likewise, there is no difference
between the fitness gradients for mutations that increase maternal \((G_f)\) or paternal \((G_m)\) care of sons, which are both

\[
G_u(z_m, z_f) = \frac{\partial s^m(z_m, z_f)}{\partial u} \frac{1}{s^m(z_m, z_f)} \left[ \left( 1 - \frac{1}{N} \right) + \frac{1}{N} \right]
\]

\[
= \frac{\partial s^m(z_m, z_f)}{\partial u} \frac{1}{s^m(z_m, z_f)}. \tag{20}
\]

\(u \in \{m, f\}\). The \((1 - 1/N)\) term of Equations 19 and 20 describe the balance between the advantage of increasing the expected number of offspring of the focal individual and simultaneously increasing the total expected number of offspring in the population and the level of competition between kin. This is equivalent to the first term of Equation 5. It is equal for gradients describing care for sons and daughters, in line with the fact that the effect of care on the expected numbers of male and female offspring is identical. The second term in the brackets of Equations 19 and 20 captures the increase in fitness due to the reduction in the variance of offspring number (reflecting the remaining terms of Equation 5).

The variance term is greater for mutants that alter the care of sons (Equation 20) because the variance in the number of surviving sons is inherently greater than that of surviving daughters. As a consequence, reducing that variance generates proportionately greater fitness benefits. The benefit of reducing the variance in male offspring number is so large that it completely offsets the reduction in fitness due to kin competition (Equation 20). The benefits of decreasing the variance in the number of surviving daughters vanish as brood size \(f\) becomes large (Equation 19), due to the fact that daughters survive independently of each other. As a result of these different effects, selection favors the fixation of mutations that increase the care of sons (Equation 20) with greater strength than those that increase the care of daughters (Equation 19), especially when fertility is high.

**Efficacy of selection:** Differences in the patterns of male and female survival also affect the efficacy of selection on paternal and maternal care, \(K_m\) and \(K_f\). Both coefficients are calculated using the probabilities of sibship (Equation A2), which are themselves expressed in terms of the moments of offspring production in Table 1. These moments are the same as those appearing in the calculation of focal fitness and in addition include the covariance between the number of male and female offspring produced by a resident individual (Appendix, Calculations for the evolution of parental care, for calculations). We find that \(K_m\) and \(K_f\) both increase with male and female survival but that their difference

\[
K_m(z_m, z_f) = K_f(z_m, z_f) \left( 1 - \frac{1 - s^m(z_m, z_f)}{2N s^m(z_m, z_f)} \right) \tag{21}
\]

\([w \sim O(N)]\) depends only male survival rate \(s^m(z_m, z_f)\). In the extreme case where all males survive \((s^m(z_m, z_f) = 1)\), selection is as efficient for male and female traits \((K_m = K_f)\).

But as male survival rate decreases, the efficacy of selection falls more rapidly in males than females. This is caused by the different modes of survival for male and female offspring. Because male offspring tend to be more related than female offspring, genetic variation in males is lower. As a consequence, the efficacy of selection in females is greater than that in males \((K_f > K_m)\), and alleles that code for maternal care are under more efficient selection than those for paternal care.

**Probability of fixation:** Combining the weights \(K_m\) and \(K_f\) (Equation 21) with the fitness gradients \(G_m\) and \(G_f\) (Equations 19 and 20) for the probability of fixation (Equation 1), we find that differences between male and female survival affect the evolution of paternal and maternal care in two ways. First, the correlation in male survival creates a stronger selection pressure on increased care for sons than for daughters. Second, the effect of more stochastic male survival increases drift in males and makes selection on paternal care less efficient than selection on maternal care. As a consequence of these effects, the most probable form of parental care to evolve in our model is maternal care for sons and the least probable is paternal care for daughters (Figure 5 for \(H = 1\)). Obviously, these conclusions are conditional on the assumptions underlying our analyses, most importantly that mutations affect only male and female care, and do so equally, and that increases in paternal and maternal care result in identical changes in the expected survival of sons and daughters.

**Mating system:** Polygyny generates a negative correlation between the reproductive output of different males, and this affects the evolution of parental care in two ways. First, reproductive skew in males decreases the strength of selection on male care, due to intensified sib competition among the surviving offspring of a male. This effect can be seen when inspecting the fitness gradients on paternal care for a female offspring

\[
G_m(z_m, z_f) = \frac{\partial s^f(z_m, z_f)}{\partial z_m} \frac{1}{s^f(z_m, z_f)} \left[ \left( 1 - \frac{1}{Ns} \right) + \frac{1}{Nf} - \frac{H}{N^2} \right] \tag{22}
\]

and that on a male offspring

\[
G_m(z_m, z_f) = \frac{\partial s^m(z_m, z_f)}{\partial z_m} \frac{1}{s^m(z_m, z_f)} \left( 1 - \frac{H}{N^2} \right). \tag{23}
\]

where both equations are here shown for a population that is strongly polygynous \([H \sim O(N)]\). Equations 22 and 23 correspond to the gradients in a monogamous population (Equations 19 and 20), with the exception of the last negative term. This term expresses the cost of intensified sib competition and increases with the level of polyandry \(H\). The gradients for the care of female offspring are unaffected by polygyny (see Equations 19 and Equation 20).

Second, polygyny affects the efficacies of selection \(K_m\) and \(K_f\) (Appendix, Calculations for the evolution of parental care).
care, for calculations). Polygyny, and the associated increase in male reproductive variance, generate additional genetic drift and reduce both $K_m$ and $K_f$ relative to monogamy. However, when male brood are brothers through their father, the genetic variance in male offspring decreases with harem size. As a consequence, the depreciation in $K_m$ with $H$ is steeper than in $K_f$:

$$K_m(z_m, z_f) = K_f(z_m, z_f) \left(1 - \frac{(1 + H)(1 - s^m(z_m, z_f))}{4N s^m(z_m, z_f)}\right),$$  

(24)

indicating that the evolution of paternal care is more sensitive to polygyny than the evolution of maternal care. The joint effects of reduced selection and lower efficacy of selection on males compromise the evolution of parental care in polygynous populations. Figure 5 shows analytical predictions of the probability of fixation for varying levels of polygyny. These show that mutants for parental care become less likely to fix as the level of polygyny increases, and this effect is exacerbated for paternal care, demonstrating the double effect of reproductive variance on both the intensity and the efficacy of selection on reproductive traits.

Long-term evolutionary equilibrium

The probability of fixation captures evolutionary dynamics over a short timescale. But as shown above (Equations 9–12), it can be used to predict long-term phenotypic outcome under a recurrent inflow of mutations. We explore here how reproductive variance affects the long-term evolution of paternal care.

In the above parental care example, the evolutionary dynamics governed by the selection gradients (Equations 19, 20, 22, and 23) eventually reach the trivial equilibrium phenotypes of maximum care for sons and daughters $[s^m(z_m, z_f) = s^m(z_m, z_f) = 1]$. We therefore introduce the realistic assumption that a parent’s resources are limited and that as a consequence, there is a trade-off between the efforts allocated to sons and daughters. The care provided to male offspring by a parent of sex $u$ is written $z_u$ and $1 - z_u$ is the care allocated to daughters (with $0 \leq z_u \leq 1$). As before, the survival $s^w(z_m, z_f)$ of an offspring of sex $v$ is a function of the paternal and maternal care received. Here, a simple additive function is assumed, where the survival of a male offspring is $s^m(z_m, z_f) = (z_m + z_f)/2$, while that of a female offspring is $s^f(z_m, z_f) = (1 - z_m + 1 - z_f)/2$. Then, because we consider the evolution of care in one sex while maintaining the care of the other sex constant, Equation 10 shows that the long-term evolution of sex-limited traits can be inferred from selection on that sex alone. In other words, we can predict the phenotypic equilibrium of a male-limited trait ($b_{mf} = 0$ and $b_{hf} = 0$) from the zeroes of the fitness gradient $G_m(z_m, z_f) = 0$ of males and that of a female-limited trait ($b_{mf} = 0$ and $b_{mn} = 0$) from the zeroes of $G_f(z_m, z_f) = 0$.

Then, convergent stable states are found in three steps. First, using calculations similar to those used in Equations 13–18 and Appendix, Calculations for the evolution of parental care, we calculate the moments of reproduction for a focal male and a focal female in the presence of trade-offs to find the fitness components of a focal individual ($w^m_v, w^f_v$, Equation 4). Second, we add the fitness components of a parent of sex $u$ derived from male and female offspring to obtain the gradient $G_u(z_m, z_f)$ as in Equation 2. Finally, solving for $G_u(z_m, z_f) = 0$, we find the convergence-stable level of investment in sons, $z^*_m$ is identical for male and female parents when the population is monogamous, and such that male survival is

$$s^m(z^*_m) = \frac{1}{2} + \frac{f - 1}{2 + 2f(2N - 1)}. \tag{25}$$

This equation show that the equilibrium investment approaches $1/2$ as population size goes to infinity ($N \to \infty$). This prediction is in line with the fact that kin competition vanishes in infinite populations and with it the selection pressures emanating from reproductive variance. Parents in very large populations are then expected to ensure an even survival of male and female offspring. As the population size decreases, however, reproductive variance starts to affect fitness and it becomes beneficial to invest more in the care of sons ($s^m(z^*_f) > 1/2$) to dampen the stochasticity in reproductive output that results from their mortality. The clutch size $f$ has an additional, weaker, effect on equilibrium care. At the extreme of single-offspring clutches,
f = 1, the differences between the patterns of male and female survival are irrelevant, and equilibrium care ensures equal survival in males and females when sex ratio is equal \((s^m(z^s_f)) = 1/2\). As clutch size increases, the effects of reproductive variance come into play and, for a given population size \(N\), larger clutch sizes result in male bias in care \((s^m(z^s_f)) > 1/2\). However, this effect rapidly plateaus with increasing \(f\) and is weaker than that of altering population size.

Theory about sex ratio predicts that to minimize the variance in offspring number, hermaphroditic females should make more offspring of the sex that is less variable in survival (Proulx 2000, 2004). In our model population, females should then produce more daughters. Using the same approach as above, we can calculate female fitness when sex ratio \(r\) at birth (ratio of males to total offspring) is maternally controlled; i.e., the number of females at birth of a focal female with phenotype \(z^s_f\) now is \(n_f \sim \text{Bin}(f; 1 - r (z^s_f))\). As before, limited care is provided by females, and covariance in survival is greater between related males. Calculating the moments of reproduction as in Equations 13–15 with appropriate sex ratio \(z^s_f\) and survival independent of the trait, we find as expected,

\[
r(z^s_f) = \frac{1}{2} \frac{1}{2} \frac{(f - 1)(1 - s^m)}{1 + s^m + f s^m (2N - 1) - f^2}
\]

that the evolutionary convergent sex ratio is biased toward females \([r(z^s_f) < 1/2]\).

**Discussion**

**Capturing sex-specific reproductive variance**

It has long been known that reproductive variance impedes adaptation by increasing genetic drift (Wright 1931; Nunney 1993; Nomura 2002; Charlesworth 2009). In parallel, a body of work has shown that reproductive variance is itself under selection, favoring less variable offspring production (Gillespie 1974; Courteau and Lessard 1999; Proulx 2000; Shpak 2005; Shpak and Proulx 2007; Lehmann and Balloux 2007; Rice 2008; Taylor 2009; Proulx and Adler 2010). Together, these studies have provided a solid theoretical basis for understanding the effects of selection on offspring distribution in a natural world that is inherently uncertain within generations. Despite these advances, models for the evolution of reproductive variance and its effects on adaptation have so far ignored biologically realistic cases of sexual reproduction, where the role of the variance can be expected to be most important. Closest to this, Taylor (2009) studied the effect of sex-specific reproductive variance on adaptation, but by modeling mating as the random union of gametes, key features of reproductive biology were neglected, since in most cases it is individuals, not gametes, that unite to mate. Finite numbers of matings and the structure of the mating system have important evolutionary effects. Not only do they generate correlations between the number of offspring of different individuals of the same sex, but they also often underlie disparities between male and female reproductive variance.

In this article, we used an individual-based approach to provide an analytical model for the evolution of male and female reproductive traits within a biologically realistic context of sexual reproduction. First, we calculated the probability of fixation of a mutant that perturbs male and female reproductive phenotypes (Equation 1), taking into account all first and second moments of the probability distribution that describes individual reproduction (Figure 1 and File S1). As a consequence, the fitness gradients, \(G_m\) and \(G_f\) (Equations 2 and 4), which express the direction and intensity of selection on a mutant, reveal the many components of reproductive variance that contribute to fitness and are hence under selection. These include the variance in the reproductive output of a focal individual (Equations 4 and 5), which decreases fitness (Figure 2A), and the variance in the total reproductive output of the rest of the population (Equations 4 and 5), which increases fitness (Figure 2B). The impact of these variances on fitness has been accounted for in previous studies (Appendix, *Link with previous work*, to see how previous works connect to the model presented here). However, our model also takes into account the covariance between the numbers of juveniles produced by different individuals of the same sex (Equations 4 and 5), which had been ignored so far and potentially have greater consequences for fitness than the variance alone (Equation 5). This covariance would emerge as a direct consequence to the biological constraints that the number of matings and female fecundity are finite and therefore cannot be ignored.

**Efficacy of selection in males and females**

The probability of fixation of a mutant (Equation 1) also depends on the efficacy with which selection can act on mutants. This is represented here by the scaling factors \(K_m\) and \(K_f\). They measure the degree to which neutral genetic variation results in phenotypic variation is then exposed to selection in males and females (Equation 6). As \(K_m\) and \(K_f\) increase, the probability of fixation of a mutant increasingly reflects the effects it has on male and female fitnesses, respectively.

The scalars \(K_m\) and \(K_f\) express effects similar to those captured by the traditional heritability of a trait (Falconer and Mackay 1996). However, while heritability is a snapshot of a population in time, \(K_m\) and \(K_f\) take into account the segregation of alleles and changes in frequency until loss or fixation of a mutant. This is illustrated by the interpretation of \(K_m\) and \(K_f\) in terms of coalescence times (File S1, Equation S1.37), which can themselves be expressed in terms of probabilities of sibship (File S1, Equation S1.38, and Figures 3 and 4), or how genes coalesce in different individuals of both sexes. The probabilities of sibship depend on the moments of offspring production (Table 1) and thereby establish a link between the mating system of
a population and the potential for selection to act on different traits in that population. High probabilities of sibship reflect a situation in which reproduction is monopolized by a subset of individuals. This reproductive skew entails a greater likelihood that a mutant is either transmitted or lost by chance and hence reduces levels of genetic variation. The factors $K_m$ and $K_f$ also increase with the dominance coefficient $h$ of the mutant. Dominance increases the covariation between genotype and phenotype at low allele frequency, which is the frequency dominating the segregation process of a new mutation, and therefore increases the visibility of mutants to selection.

An important feature of $K_m$ and $K_f$ are their sex specificity, respectively scaling on male and female fitness gradients. This reveals that genetic drift can influence male and female evolution with varying strength. Traditionally, population-genetic treatments of evolution in dioecious populations express the effect of genetic drift on the segregation of two alleles simply as the inverse of the overall effective population size or as some mutant frequency-dependent function (Ethier and Nagylaki 1988; Taylor 2009), but in both cases, the effect of genetic drift on male and female selection is the same. This simplification stems from the requirements to obtain a diffusion limit for the segregation process, which ignore some differences between male and female reproduction.

The method we used here to calculate the probability incorporates all second moments of male and female individual reproduction and shows that it is possible for genetic drift to affect selection on males and females differently. When $K_f$ is larger than $K_m$, selection on females contributes more to the probability of fixation than does selection on males, and vice versa. A variety of factors can lead to differences between male and female efficacy of selection. As shown in the Example section, discrepancies between $K_m$ and $K_f$ can occur as the result of differences between male and female patterns of mortality that generate a greater level of genetic variance in females than in males. This is not only a theoretical possibility. In the house finch, for example, mite ectoparasitism affects related males more strongly than related females, leading to male-biased mortality (Badyaev et al. 2006). As a consequence, we expect $K_m$ to be smaller than $K_f$ in this species.

**Long-term sex-specific evolution**

To predict the joint evolution of male and female phenotypes, we embedded our model into a trait substitution sequence process. We obtained a stochastic model of long-term phenotypic evolution for dioecious populations that allows one to conveniently evaluate convergence-stable states, which correspond to the most likely phenotypic outcomes of evolution at mutation–selection–drift balance (Equations 11 and 12 and Appendix, Diffusion equation for phenotypic evolution in dioecious populations). When the reproductive variances of males and females are such that there is no difference in the efficacy of selection between the sexes ($K_m = K_f$; Equations 11 and 12), then the conditions for phenotypes to be convergence stable depend only on the fitness gradients, in agreement with previous deterministic models (Leimar 2009). When the efficacy of selection differs between the sexes ($K_m \neq K_f$), however, they may affect the evolutionary trajectory and change the stability of internal equilibria (Equation 12). Therefore, the most likely phenotypes to be observed in natural populations can be significantly affected by sex-specific reproductive variance.

**Effects of sex-specific variance on parental care**

To illustrate the many effects of reproductive variance on the evolution of dioecious species, we calculated the probability of fixation of mutants coding for maternal and paternal care for sons and daughters in a situation where the survival of sons is highly correlated within broods. While very specific, this example allows us to illustrate some of the key effects that our model can capture. First, our results demonstrate how phenotypic evolution can be driven by selection against reproductive variance. Thus, care for sons evolves more readily than care for daughters, because the former alleviates the high degree of reproductive variance that arises as a consequence of correlated male survival (Equations 19 and 20). Second, we showed that the pattern of male survival reduced the efficacy of selection on male traits by decreasing the amount of genetic variation in males (Equation 21). This means that mutants coding for maternal care have a greater probability of fixation than those coding for paternal care, even if the effect of maternal and paternal care on offspring survival is identical. Finally, male polygyny generates a negative correlation between the reproductive outputs of different males, which in turn generates an additional selection pressure on the evolution of paternal care (Equations 23 and 22). These forces mitigate the strength of selection for paternal care because as fewer males monopolize reproduction, kin competition between the offspring of a male increases. The selection pressures generated by (co)variances might be minimal when populations are very large, polygamy extensive and fecundity effectively unlimited. However, in most biologically realistic scenarios, the complicated interactions between the different components of reproductive variance can be expected to affect the evolutionary process through selection and genetic drift.

The phenotypic equilibrium predicted for both parental sexes is, like the probability of fixation, affected by selection against reproductive variance. Thus, fathers and mothers will invest more in the care of sons to mitigate the detrimental effects of their stochastic survival on parental fitness (Equation 25). Interestingly, this prediction contrasts with other results on the evolution of sex-ratio allocation whereby females are expected to produce more daughters when the survival of males within a brood is highly correlated (Equation 26) (Proulx 2000, 2004). Therefore, selection against reproductive variance leads to the counterintuitive equilibrium whereby females produce fewer sons for which they care more. It would be interesting to further explore the effect of sex-specific variance on the evolution of
sex allocation. In particular, we expect that the sex ratio at birth would differ according to whether it is controlled by the male or female parent and that the difference between maternally and paternally controlled sex ratio depends on the mating system. For instance, with the life cycle given in the Example section of this article, if sex ratio is male controlled and the population is polygynous, then selection on males to minimize their reproductive variance would favor a bias toward females that is even more pronounced than when the sex ratio is female controlled (Equation 26).

Our analysis of the long-term evolution of male and female care behavior showed that both sexes evolve toward the same equilibrium level of care (Equation 25). This contrasts with the predicted short-term dynamics, where greater stochasticity in male survival caused a reduction in the probability of fixation of mutants for paternal care, compared to that seen for maternal care mutants. This discrepancy intuitively implies a lower rate of adaptation in the male than female trait and a longer time to reach the evolutionary equilibrium. In general, the stochastic model of phenotypic evolution (Appendix, Diffusion equation for phenotypic evolution in dioecious populations) suggests that the rate of adaptation in males and females scales with the efficacy of selection $K_m$ and $K_f$, respectively.

**Outlook**

The framework provided in this article is ideal for studying complex social interactions between individuals of sexual populations. Examples of such traits are those involved in evolutionary games between the male and female of a mating pair or strategies in games between individuals of the same sex, for example, in male–male competition for mating and fertilization success. In the latter case, the covariance between the numbers of offspring produced by different males is expected to have important effects. Use of our model to study the social and sex-specific frequency-dependent aspects of reproductive evolution is straightforward because all parameters in Equations 1 and 10 can be derived using only the phenotype of a focal individual and the average male and female phenotypes in the population. Another class of traits for which our model is particularly well suited are sexually antagonistic ones (Parker 1979; Lande 1980; Bonduriansky and Chenoweth 2009; Pennell and Morrow 2013). By taking into account the positive correlation of mutational effects in males and females ($b_{maf} > 0$ in Equation 10), different selection pressures in males and females ($G_m \neq G_f$), and different levels of reproductive variance in the sexes, the model is well adapted to investigating the evolution of these traits under the simultaneous influences of selection and drift.

With selection on variance being inversely proportional to the population size, selection on the variance will be mostly relevant in small panmictic populations, where genetic drift may therefore mitigate its effects. But if populations are structured into local breeding groups, then selection against reproductive variance is inversely proportional to local patch size (Shpak and Proulx 2007; Lehmann and Balloux 2007), while genetic drift inversely scales with the total population size, which can be very large. All the effects of selection on reproductive variance described in this article may then be particularly relevant in populations that are divided in small patches but are globally large. In fact, if density-dependent regulation takes place before dispersal (soft selection, Roze and Rousset 2003), then selection against reproductive variance is as described by our panmictic model, with fitness given by Equation 5 but with the number of individuals being those in a local patch (Shpak 2005; Lehmann and Balloux 2007).

Explicitly taking spatial structure into account in regimes of soft and hard selection may also reveal interesting examples of sex-specific evolution. In structured and dioecious populations, we expect that sex-specific local competition (e.g., Perrin and Mazalov 2000), but also reproductive variance, will drive the evolution of sex-specific dispersal. In turn, this will generate differences in genetic variation across the sexes (i.e., between $K_m$ and $K_f$), thereby influencing the evolution of sex-specific strategies. It would therefore be particularly interesting to extend the model to explicitly take into account spatial structure to investigate the evolution of sex-specific dispersal strategies and how it interacts with the evolution of other sex-specific traits.

Future development of the model should also accommodate for a greater variety of genetic architecture of traits. Because $K_m$ and $K_f$ depend on the covariance between genotype and phenotype, differences in the genetic determination of traits between the sexes would also translate into differences between the efficacy of selection in males and females. This is not unlikely as differences between male and female heritability have been reported for phenotypic traits in animals (e.g., Eisen and Legates 1966; Jensen et al. 2003), including humans (Weiss et al. 2006) as well as plants (e.g., Ashman 1999). Such differences would naturally arise for sex-linked genes. For instance, in species with an XY sex-determining system, where males are hemizygous for the X chromosome, dominance interactions can occur only between the two X chromosomes of females (Wayne et al. 2007). It would therefore be interesting to extend our model for sex-linked genes and test whether the interaction between selection on reproductive variance and the efficacy of selection in males and females lead to different evolutionary dynamics than on autosomes.

To conclude, using a population-genetic approach that takes into account all the relevant moments of reproduction in the two sexes, we have shown that the effect of sex-specific reproductive variance and covariances and selection on it influences the evolution of dioecious species. In particular, we have found that even if the fitness gradients on male and female traits have the same steepness but opposite directions, differences in male and female reproductive variance can lead to selection in one sex dominating selection in the other, and alter the trajectory of long term phenotypic evolution.
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Literature Cited

Ashman, T.-L., 1999  Quantitative genetics of floral traits in a gy- nodioecious wild strawberry Fragaria virginiana: implications for the independent evolution of female and hermaphrodite floral phenotypes. Nat. Genet. 83: 733–741.

Badyaev, A. V., T. L. Hamstra, K. P. Oh, and D. A. Acevedo Seaman, 2006  Sex-biased maternal effects reduce ectoparasite-induced mortality in a passerine bird. Proc. Natl. Acad. Sci. USA 103 (39): 14406–14411.

Bateman, A. J., 1948  Intra-sexual selection in Drosophila. Heredity 2(3): 349–368.

Beckerman, A. P., S. P. Sharp, and B. J. Hatchwell, 2011  Predation and kin-structured populations: an empirical perspec- tive on the evolution of cooperation. Behav. Ecol. 22(6): 1294–1303.

Bonduriansky, R., and S. F. Chenoweth, 2009  Intra-lus sexual conflict. Trends Ecol. Evol. 24(5): 280–288.

Champagnat, N., and A. Lambert, 2007  Evolution of discrete pop- ulations and the canonical diffusion of adaptive dynamics. Ann. Appl. Probab. 17(1): 102–155.

Charlesworth, B., 2009  Effective population size and patterns of molecular evolution and variation. Nat. Rev. Genet. 10(3): 195–205.

Courteau, J., and S. Lessard, 1999  Stochastic effects in LMC mod- els. Theor. Popul. Biol. 55(2): 127–136.

Eisen, E. J., and J. E. Legates, 1966  Genotype-sex interaction and the genetic correlation between the sexes for body weight in Mus musculus. Genetics 54: 611–623.

Ether, S. N., and T. Nagylaki, 1988  Diffusion approximations of Markov chains with two time scales and applications to population genetics. II. Adv. Appl. Probab. 20(3): 525–545.

Falconer, D. S., and T. C. F. Mackay, 1996  Introduction to Quan- titative Genetics, 4th ed. Longman, London.

Frank, S. A., 2011  Natural selection. I. Variable environments and uncertain returns on investment. J. Evol. Biol. 24(11): 2299–2309.

Foster, D., and P. Young, 1990  Stochastic evolutionary game dy- namics. Theor. Popul. Biol. 38(2): 219–232.

Freidlin, M. I., and A. D. Wentzell, 2012  Random Perturbations of Dynamical Systems, Springer-Verlag, Berlin.

Gardiner, C., 2009  Stochastic Methods: A Handbook for the Natural and Social Sciences, Springer Series in Synergetics, 4th ed. Springer-Verlag, Berlin.

Gillespie, J. H., 1974  Natural selection for within-generation var- iance in offspring number. Genetics 76: 601–606.

Gillespie, J. H., 1975  Natural selection for within-generation var- iance in offspring number II. Discrete Haploid models. Genetics 81: 403–413.

Gillespie, J. H., 1977  Natural selection for variances in offspring numbers: a new evolutionary principle. Am. Nat. 111(981): 1010–1014.

Gillespie, J. H., 1994  The Causes of Molecular Evolution, Oxford Series in Ecology and Evolution. Oxford University Press, New York.

Jensen, H., B.-E. Sæther, T. H. Ringsby, S. C. Tufto, J. Griffith et al., 2003  Sexual variation in heritability and genetic correlations of morphological traits in house sparrow (Passer domesticus). J. Evol. Biol. 16: 1296–1307.

Lande, R., 1980  Sexual dimorphism, sexual selection, and adap- tation in polygenic characters. Evolution 34(2): 292–305.

Lehnmann, L., 2012  The stationary distribution of a continuously varying strategy in a class-structured population under mutation–selection–drift balance. J. Evol. Biol. 25(4): 770–787.

Lehmann, L., and F. Balloux, 2007  Natural selection on fecundity variance in subdivided populations: kin selection meets bet hedging. Genetics 176: 361–377.

Lehmann, L., and F. Roussel, 2009  Perturbation expansions of mulitlocus fixation probabilities for frequency-dependent selection with applications to the Hill–Robertsen effect and to the joint evolution of helping and punishment. Theor. Popul. Biol. 76(1): 35–51.

Leimar, O., 2009  Multidimensional convergence stability. Evol. Ecol. Res. 11(2): 191–208.

Lessard, S., and V. Ladret, 2007  The probability of fixation of a single mutant in an exchangeable selection model. J. Math. Biol. 54(5): 721–744.

Metz, J. A. J., S. A. H. Geritz, G. Meszena, F. J. A. Jacobs, and Heerwaarden, 1995  Adaptive dynamics: a geometrical study of the consequences of nearly faithful reproduction. Technical Report, International Institute for Applied Systems Analysis A-2361. Laxenburg, Austria.

Nomura, T., 2002  Effective size of populations with unequal sex ratio and variation in mating success. J. Anim. Breed. Genet. 119(5): 297–310.

Nunney, L., 1993  The influence of mating system and overlapping generations on effective population size. Evolution 47(5): 1329–1341.

Oehlert, G. W., 1992  A note on the delta method. Am. Stat. 46 (1): 27–29.

Parker, G. A., 1979  Sexual Selection and Reproductive Competition in Insects. Academic Press, San Diego.

Pennell, T. M., and E. H. Morrow, 2013  Two sexes, one genome: the evolutionary dynamics of intrasexual conflict. Ecol. Evol. 3(6): 1819–1834.

Perrin, N., and V. Mazalov, 2000  Local competition, inbreeding, and the evolution of sex biased dispersal. Am. Nat. 155(1): 116–127.

Proulx, S., 2000  The ESS under spatial variation with applications to sex allocation. Theor. Popul. Biol. 58(1): 33–47.

Proulx, S. R., 2004  Sources of stochasticity in models of sex allo- cation in spatially structured populations. J. Evol. Biol. 17(4): 924–930.

Proulx, S. R., and F. R. Adler, 2010  The standard of neutrality: Still flapping in the breeze? J. Evol. Biol. 23(7): 1339–1350.

Rice, S., 2008  A stochastic version of the Price Equation reveals the interplay of deterministic and stochastic processes in evolution. BMC Evol. Biol. 8(1): 262.

Rousett, F., 2003  A minimal derivation of convergence stability measures. J. Theor. Biol. 221(4): 665–668.

Rousett, F., and S. Billiard, 2000  A theoretical basis for measures of kin selection in subdivided populations: finite populations and localized dispersal. J. Evol. Biol. 13(5): 814–825.

Roze, D., and F. Rousset, 2003  Selection and drift in subdivided populations: a straightforward method for deriving diffusion approximations and applications involving dominance, selving and local extinctions. Genetics 164(4): 2153–66.

Roze, D., and F. Rousset, 2004  The robustness of Hamilton’s rule with inbreeding and dominance: kin selection and fixation prob- abilities under partial sib mating. Am. Nat. 164(2): 214–231.
Appendix

Assumption on distribution of juveniles

Given an index set of individuals \( i \in I \) in the population and a corresponding set of powers defined by a mapping \( \xi: I \rightarrow \mathbb{Z}^+ \), it is assumed that the following

\[
E \left[ \prod_{i \in I} (J_{vi} - \mu_{\xi(i)})^{\xi(i)} \right] \sim O \left( \sum_{i=1}^{\xi(I)+1-|I|} \right),
\]

(A1)

holds, where \(|I|\) is the number of individuals in set \( I \). The remainder terms that appear in \( R \), given by the higher-order terms of the Taylor expansion of \( F \), are thus of order \( 1/N^2 \).

Weights for additive mutants

Using Equation S1.38 from File S1, the weights on male and female fitness gradients in the probability of fixation of an additive \((h = 1/2)\) mutant are given by

\[
K_m = \left( \frac{1}{2N_m + 2N_f} \right) \left( 1 - \frac{1}{N_m} \right) \left( \frac{4 + \Omega_m^c - \Omega_m^v}{D} \right),
\]

\[
K_f = \left( \frac{1}{2N_m + 2N_f} \right) \left( 1 - \frac{1}{N_f} \right) \left( \frac{4 + \Omega_f^c - \Omega_f^v}{D} \right),
\]

(A2)

where \( D = \Omega_m^c + \Omega_f^c + 2\Omega_m^v + \Omega_f^v + 2\Omega_c^v (\Omega_f^c - \Omega_m^v)/2 + \Omega_c^v (\Omega_m^c - \Omega_f^c)/2 + \Omega_c^c \Omega_f^v/2 - \Omega_f^c \Omega_m^v/2 \).

Calculations for the evolution of parental care

Here the remaining components of the probability of fixation of alleles coding for parental care are calculated. Because male survival is different between populations in which maternal and parental care are provided (see main text), it is simpler to consider separately the cases when care is maternal and when care is paternal.

**Maternal care:** To calculate the weight \( K_t \), we need the probabilities of sibship (Table 1) in the resident population \((z_f = z_t)\). To calculate the maternal probabilities of sibship \( \bar{\Theta}^v \), in addition to Equation 13–15 of the main text, the covariance between number of male and female offspring that a female produces is also required, and it is given by

\[
\mu^{mf}_t(z_m, z_t) = E_{n_f} \left[ n_f s^m(z_m, z_t) (f - n_f) s^f(z_m, z_t) \right] - E_{n_m} \left[ n_m s^f(z_m, z_t) \right] E_{n_f} \left[ n_f s^m(z_m, z_t) \right]
\]

\[
= J_f (z_m, z_t) \vartheta^{(z_m, z_t)}, \quad \vartheta^{(z_m, z_t)}.
\]

The paternal probabilities of sibship \( \bar{\Theta}^c \) also influence \( K_t \) (Equation A2) and their components are derived below. The expected numbers of male and female offspring of a male are given by Equation 13 evaluated at male phenotype \( z_m \). The

Sella, G., and A. E. Hirsh, 2005 The application of statistical physics to evolutionary biology. Proc. Natl. Acad. Sci. USA 102: 9541–9546.

Shpak, M., 2005 Evolution of variance in offspring number: the effects of population size and migration. Theory Biosci. 124(1): 65–85.

Shpak, M., and S. Proulx, 2007 The role of life cycle and migration in selection for variance in offspring number. Bull. Math. Biol. 69(3): 837–860.

Taylor, J. E., 2009 The genealogical consequences of fecundity. Proc. Natl. Acad. Sci. USA 106(21): 8609–8611.

Wayne, M. L., M. Telonis-Scott, L. M. Bono, L. Harshman, A. Kopp et al., 2007 Simpler mode of inheritance of transcriptional variation in male Drosophila melanogaster. Proc. Natl. Acad. Sci. USA 104(47): 18577–18582.

Weiss, L. A., L. Pan, M. Abney, and C. Ober, 2006 The sex-specific genetic architecture of quantitative traits in humans. Nat. Genet. 38: 218–222.

Wright, S., 1931 Evolution in Mendelian populations. Genetics 16: 97–159.
variance in the number of females fathered by a male is found by conditioning on the random number of matings \(N_{\text{mat}}\) of a male

\[
\nu_{m}^{f}(z_{m}, z_{f}) = E_{\text{mat}} \left[ N_{\text{mat}} f \frac{s^{f}(z_{m}, z_{f})}{2} \left( 1 - \frac{s^{f}(z_{m}, z_{f})}{2} \right) \right] + V_{\text{mat}} \left[ N_{\text{mat}} f \frac{s^{f}(z_{m}, z_{f})}{2} \right].
\] (A4)

Because each male is equally likely to mate, and if a male does, it mates exactly \(H\) times, we have \(E[N_{\text{mat}}] = 1\) and \(V[N_{\text{mat}}] = H - 1\), so that

\[
\nu_{m}^{f}(z_{m}, z_{f}) = f \frac{s^{f}(z_{m}, z_{f})}{2} \left( 1 + f \frac{s^{f}(z_{m}, z_{f})}{2} (1 + f(H - 1)) \right).
\] (A5)

The variance in the number of males fathered by a male, given that each male brood entirely survives or dies, reads

\[
\nu_{m}^{m}(z_{m}, z_{f}) = E_{\text{mat}} \left[ N_{\text{mat}} f \frac{s^{m}(z_{m}, z_{f})}{4} \left( 1 + f \left( 1 - s^{m}(z_{m}, z_{f}) \right) \right) \right] + V_{\text{mat}} \left[ N_{\text{mat}} f \frac{s^{m}(z_{m}, z_{f})}{2} \right]
\]

\[
= f \frac{s^{m}(z_{m}, z_{f})}{4} \left( 1 + f \left( 1 + (H - 2) s^{m}(z_{m}, z_{f}) \right) \right).
\] (A6)

To calculate the covariance between number of males and of females produced by a male, we define \(X\) as the random product of males and females coming from the same mating. Then, since offspring survival is independent across matings, we have

\[
\rho_{m}^{m,f}(z_{m}, z_{f}) = E_{\text{mat},X} \left[ N_{\text{mat}} f \frac{s^{m}(z_{m}, z_{f})}{2} \right] E_{\text{mat}} \left[ N_{\text{mat}} f \frac{s^{m}(z_{m}, z_{f})}{2} \right],
\]

and since, \(E[X] = s^{m}(z_{m}, z_{f}) s^{f}(z_{m}, z_{f}) E_{\text{mat}}[n_{o}(f - n_{o})]\),

\[
\rho_{m}^{m,f}(z_{m}, z_{f}) = -f s^{m}(z_{m}, z_{f}) s^{f}(z_{m}, z_{f}) (1 - f(H - 1)).
\] (A8)

Finally, substituting Equations 13–15 and Equations A3–A8 into the probabilities of sibship (Table 1), and in turn, substituting the latter into Equation A2, we find the efficacy of selection on maternal care.

**Paternal care:** Most of the moments required to calculate focal male fitness and the probabilities of sibship to find \(K_{m}\) are the same as in the preceding section (evaluated at focal male phenotype \(z_{m}\) for focal fitness, and at resident male \(z_{m}\) instead of resident female \(z_{f}\)). However, because male survival is different in a population in which parental care is provided by males, the variance in the number of male offspring fathered by the focal male, and the covariance between the number of male and female offspring fathered by a resident male (required for the probabilities of sibship) are different. Using similar arguments as above, we find that they are respectively given by

\[
\nu_{m}^{m}(z_{m}, z_{f}) = \frac{1}{2} f s^{m}(z_{m}, z_{f}) \left( 1 + f H - f s^{m}(z_{m}, z_{f}) \right)
\]

\[
\rho_{m}^{m,f}(z_{m}, z_{f}) = s^{m}(z_{m}, z_{f}) s^{f}(z_{m}, z_{f}) f(H - 1 - f).
\] (A9)

Calculating the probabilities of sibship (Table 1) using the above and substituting into Equation A2, we find the efficacy of selection on paternal care.

**Diffusion equation for phenotypic evolution in dioecious populations**

The substitution rate \(k\) (Equation 9) is the jump rate of a so-called jump process (Gardiner 2009), which here describes a population “jumping” from a monomorphic phenotypic state to another. The moments of the infinitesimal jump of the evolving phenotypes in each sex \((\Delta z_{m}, \Delta z_{f})\) characterize the distribution of phenotypic changes over an infinitesimally small evolutionary time period, and they are found by integrating the phenotypic effects of a substitution over the p.d.f. of all substitution rates, \(E[(\Delta z_{m})^{i}(\Delta z_{f})^{j}] = \int \int \delta^{i}_{m} \delta^{j}_{f} k(z_{m}, z_{f}, \delta_{m}, \delta_{f}, h) d\delta_{m} d\delta_{f} dh\). To the first order of \(\delta\), we obtain for the first moment of \(\Delta z_{u}\) for \(u \in \{m, f\}\),
for \( v \in \{m, f\}, v \neq u \), and where \( b = C[\delta_u, \delta_v] \) are the second moments of mutant sex-specific effects. Because dominance is independent of \( \delta_m \) and \( \delta_f \) and \( K \) is linear in \( h \) (File S1, Equation S1.38), \( K_u(z_m, z_f, \overline{h}) \) is simply evaluated at expected dominance \( E[h] = \overline{h} \). Similarly, it is possible to show that the Equation for \( E[\Delta z_u] \) still holds if mutants have sex-specific dominance \( h_m \) and \( h_f \), as long as they are independent of \( \delta_m \) and \( \delta_f \) and that they are on average equal \( \overline{h_m} = \overline{h_f} = \overline{h} \). The second moments of infinitesimal phenotypic change of the first order of \( \delta \) are given by \( E[(\Delta z_u)^2] = \xi b_{uu} \) and \( E[(\Delta z_m)(\Delta z_f)] = \xi b_{mf} \).

Assuming that the phenotypic changes are continuous in probability, the first two moments of infinitesimal change \( \Delta z_u \) can be used to approximate the rate of phenotypic change by a Fokker–Planck equation (Gardiner 2009). This equation characterizes the change of the distribution of male and female phenotypes \( \psi(z_m, z_f; \tau) \) in evolutionary time \( \tau \). The distribution of phenotypes here is meant over many evolutionary trajectories or experiments, rather than over the population. The population remains monomorphic: fixation of loss of mutants is instantaneous. If the male and female phenotypes \((z_m, z_f)\) at evolutionary time \( \tau \) have p.d.f. \( \psi(z_m, z_f; \tau) \), then it satisfies

\[
\frac{\partial \psi(z_m, z_f; \tau)}{\partial \tau} = - \frac{\partial}{\partial z_m} \left[ a_m(z_m, z_f) \psi(z_m, z_f; \tau) \right] - \frac{\partial}{\partial z_f} \left[ a_f(z_m, z_f) \psi(z_m, z_f; \tau) \right] + \xi \left( b_{mn} \frac{\partial^2 \psi(z_m, z_f; \tau)}{\partial z_m^2} + b_{mf} \frac{\partial^2 \psi(z_m, z_f; \tau)}{\partial z_f^2} + b_{mf} \frac{\partial^2 \psi(z_m, z_f; \tau)}{\partial z_m \partial z_f} \right),
\]

where the functions \( a_m(z_m, z_f) \) and \( a_f(z_m, z_f) \) are the expected infinitesimal changes of male and female phenotypes given in Equation A10 and the stationary distribution is given by \( \psi(z_m, z_f) = \lim_{\tau \rightarrow \infty} \psi(z_m, z_f; \tau) \).

**Link with previous work**

Substituting Equation 4 into Equation 2, we find fitness gradients that are consistent with previous work on the evolution of reproductive variance. For instance, Lehmann and Balloux (2007) models the evolution of a helping trait \( z_f \) that disrupts the mean \( \mu_f(z_f) \sim O(N) \) and variance \( \nu^2_f(z_f) \sim O(N) \) in fertility. Mating is random, each female gives birth independently of one another, and sex ratio is equal at birth and in the population. Substituting Equation 4 into Equation 2, we have that the fitness gradient on \( z_f \) is proportional to

\[
G(z_f) \propto \frac{1}{2} \left[ 1 - \frac{C^2}{N_f} \right] \frac{d \ln \mu_f(z_f)}{d z_f} - \frac{1}{2} \frac{C^2}{N_f} \frac{d \ln \nu^2_f(z_f)}{d z_f} + O(1/N^2), \tag{A12}
\]

where \( C^2 = \nu^2_f/\mu^2_f \) is the squared coefficient of variation in fertility in the resident female, in agreement with Equation A37 of Lehmann and Balloux (2007).

The original fitness gradient by Gillespie (1975, Equation 11a) or equivalently, that derived for a dioecious population by Taylor (2009, Equation 14 for an additive mutant) may be found directly from Equation A12. These analyses use the diffusion approximation, which requires that the difference between the mean fertilities of the resident and mutant phenotypes tend to zero as the population size tends to infinity, i.e., that \( d \ln \mu_f(z_f)/dz_f \sim O(1/N) \). Applying this assumption to Equation A12, we have

\[
G(z_f) \propto \frac{1}{2} \frac{d \ln \mu_f(z_f)}{d z_f} - \frac{1}{2} \frac{C^2}{N_f} \frac{d \ln \nu^2_f(z_f)}{d z_f} + O(1/N^2), \tag{A13}
\]

where the deleterious effects of sib competition on expected fecundity fall victim to the order condition required by the diffusion approach of Gillespie (1975) and Taylor (2009).
The Evolution and Consequences of Sex-Specific Reproductive Variance

Charles Mullon, Max Reuter, and Laurent Lehmann
File S1

Derivation of the fixation probability of a mutant

Expected change of mutant frequency. In order to derive the probability of fixation of a mutant, we first evaluate the expected change of mutant frequency over one generation. The frequency of the mutant in a male indexed $i \in \{1, \ldots, N_m\}$ is written as $p_{mi} \in \{0, 1/2, 1\}$, and the frequency in a female $j \in \{1, \ldots, N_f\}$ is written $p_{fj} \in \{0, 1/2, 1\}$. The indicator variables $\mathbb{1}_{\delta^m_i}$ and $\mathbb{1}_{\delta^f_j}$ respectively take the value one if the paternally and maternally inherited alleles of individual $i$ are mutant, and zero otherwise. Then, the mutant frequencies in male $i$ and in female $j$ are

$$p_{mi} = \frac{\mathbb{1}_{\delta^m_i} + \mathbb{1}_{\delta^f_i}}{2} \quad \text{and} \quad p_{fj} = \frac{\mathbb{1}_{\delta^m_j} + \mathbb{1}_{\delta^f_j}}{2}. \quad (\text{SI.1})$$

We write $\bar{p}_{m,t} = \sum_{i=1}^{N_m} p_{mi,t}/N_m$ and $\bar{p}_{f,t} = \sum_{j=1}^{N_f} p_{fj,t}/N_f$ for the average mutant frequencies in males and females in the population and denote by $q_i$, the vector collecting the realization of mutant frequencies (the realized values of $\mathbb{1}_{\delta^m_i}$ and $\mathbb{1}_{\delta^f_i}$) in the population at time $t$.

If the mutant changes male and female phenotypes by $\delta_m$ and $\delta_f$ and a parent transmits its maternally or paternally inherited gene with equal probability, the expected average male and female mutant frequencies in the next generation is

$$E[\bar{p}_{m,t+1}|q_i] = \frac{1}{2N_m} \left( \sum_{i=1}^{N_m} p_{mi,t} w^m_{m_i}(\delta_m, \delta_f) + \frac{N_f}{N_f} \right)$$

$$E[\bar{p}_{f,t+1}|q_i] = \frac{1}{2N_f} \left( \sum_{i=1}^{N_f} p_{fj,t} w^f_{mj}(\delta_m, \delta_f) + \frac{N_m}{N_m} \right), \quad (\text{SI.2})$$

where $w^u_{vi}(\delta_m, \delta_f)$ is the expected number of adult offspring of sex $u$ of individual $i$ (itself is of sex $v$) (Price 1970). Eq. (SI.2) extends Rice (2008)'s "selection differential" to a two-sexes populations (his $\text{cov}(\phi, \hat{\Omega})$ term assuming a constant population size).

If selection is weak, it is sufficient to approximate allele frequency change to the first order of phenotypic effect in males and females $\delta_m$ and $\delta_f$. The fitness terms $w^u_{vi}$ are approximated as $w^u_{vi}(\delta_m, \delta_f) = w^u_{vi}(0) + \delta_m(\partial w^u_{vi}(0)/\partial \delta_m) + \delta_f(\partial w^u_{vi}(0)/\partial \delta_f) + O(\delta^2)$, with $(0) = (0, 0)$. There are two things to note about the fitness terms and their derivatives. First, in the absence of phenotypic differences, each individual is expected to contribute equally to the next generation, and so $w^u_{vi}(0) = N_u/N_u$. Second, the partial derivatives of an individual's fitness with respect to phenotypic effect in the other sex is zero $\partial w^u_{vi}(0)/\partial \delta_u = 0$ with $u \neq v$. For instance, when all males are the same ($\delta_m = 0$), changes in female phenotype have no effect on the expected number of adult offspring of a focal male. So substituting for $w^u_{vi}(\delta_m, \delta_f)$ in eq. (SI.2) gives

$$E[\bar{p}_{m,t+1}|q_i] = \frac{1}{2}(\bar{p}_{m,t} + \bar{p}_{f,t}) + \frac{1}{2N_m} \left( \delta_m \sum_{i=1}^{N_m} p_{mi,t} \frac{\partial w^m_{mi}(0)}{\partial \delta_m} + \delta_f \sum_{j=1}^{N_f} p_{fj,t} \frac{\partial w^f_{mj}(0)}{\partial \delta_f} \right) + O(\delta^2) \quad (\text{SI.3})$$

$$E[\bar{p}_{f,t+1}|q_i] = \frac{1}{2}(\bar{p}_{m,t} + \bar{p}_{f,t}) + \frac{1}{2N_f} \left( \delta_m \sum_{i=1}^{N_m} p_{mi,t} \frac{\partial w^m_{mi}(0)}{\partial \delta_m} + \delta_f \sum_{j=1}^{N_f} p_{fj,t} \frac{\partial w^f_{mj}(0)}{\partial \delta_f} \right) + O(\delta^2).$$

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Another consequence of weak selection is that the fitness phenotype of an individual in eq. (SI.3) can be approximated in terms of only three phenotypic values: the phenotype of an individual, the average male phenotype and the average female phenotype. To see this, consider the expected number of female adults produced by male $i$, $w_{mi}^f$. This depends on his phenotype $z_{mi}$, as well as the collection of the phenotypes of all the other males in the population, $z_{-mi} = \{z_{mk} : k : 1 \rightarrow N_m, k \neq i\}$, as well as those of all the females in the population, $z_i = \{z_{j} : j : 1 \rightarrow N_f\}$. Expanded about male population average, excluding male $i$, $\bar{z}_{-mi} = 1/(N_m - 1) \sum_{k \neq i} \sum_{1}^{N_m} z_{mk}$, and female population average $\bar{z}_i = \sum_{j} z_{j}/N_f$, $w_{mi}^f$ reads

$$w_{mi}^f(z_{mi}, z_{-mi}, z_i) \approx w_{mi}^f(z_{mi}, \bar{z}_{-mi}, \bar{z}_i) + \sum_{k=1;k\neq i}^{N_m} \frac{\partial w_{mi}^f}{\partial z_{mk}} (z_{mk} - \bar{z}_{-mi}) + \sum_{j=1}^{N_f} \frac{\partial w_{mi}^f}{\partial z_{j}} (z_{j} - \bar{z}_i),$$

(SI.4)

and the remainder is $O(\delta^2)$ because the difference between any two phenotypes of the same sex is of order $O(\delta)$. The effect of changing the phenotype of any female has the same effect on the fitness of male $i$, so that all $\partial w_{mi}^f/\partial z_{j}$ are equal, and

$$\sum_{j=1}^{N_f} (\partial w_{mi}^f/\partial z_{j})(z_{j} - \bar{z}_i) = (\partial w_{mi}^f/\partial z_{j}) \sum_{j=1}^{N_f} (z_{j} - \bar{z}_i),$$

but by definition, $\sum_{j=1}^{N_f} (z_{j} - \bar{z}_i) = 0$. A similar argument shows that $\sum_{k=1;k\neq i}^{N_m} (\partial w_{mi}^f/\partial z_{mk})(z_{mk} - \bar{z}_{-mi}) = 0$. Hence, the female component of fitness of male $i$, $w_{mi}^f(z_{mi}, z_{-mi}, z_i)$, can be approximated by $w_{mi}^f(z_{mi}, \bar{z}_{-mi}, \bar{z}_i)$; that is, as a function of its phenotype, $z_{mi}$, the average male phenotype excluding the focal, $\bar{z}_{-mi}$, and the average phenotype of females in the population. However, for computational purposes it may be more convenient to express $w_{mi}^f$ in terms of $z_{mi}$ and the average male phenotype $\bar{z}_m$. This can be done since $\bar{z}_{-mi} = (N_m \bar{z}_m - z_{mi})/(N_m - 1)$, so from now on we write the fitness of individual $i$ as $w_{mi}^f(z_{mi}, \bar{z}_m, \bar{z}_t)$, keeping in mind that with this notation $\partial w_{mi}^f(z_{mi}, \bar{z}_m, \bar{z}_t)/\partial z_{mi} = \partial w_{mi}^f(x, \bar{z}_m, \bar{z}_t)/\partial x + (\partial w_{mi}^f(z_{mi}, \bar{z}_m, \bar{z}_t)/\partial \bar{z}_m)/N_m$.

Using the chain rule, the derivatives of fitness with respect to $\delta_v$ is $\partial w_{vi}^u/\partial \delta_v = (\partial w_{vi}^u/\partial z_{vi})(dz_{vi}/d\delta_v) + (\partial w_{vi}^u/\partial \bar{z}_v)(d\bar{z}_v/d\delta_v)$.

By observing that the average male phenotype is insensitive to changes in female mutant effects (d$\bar{z}_m$/d$\delta_t$ = 0), and that the average female phenotype is insensitive to changes in male mutant effects (d$z_{vi}$/d$\delta_m$ = 0), the derivatives of fitness collapse to $\partial w_{vi}^u/\partial \delta_v = (\partial w_{vi}^u/\partial z_{vi})(dz_{vi}/d\delta_v) + (\partial w_{vi}^u/\partial \bar{z}_v)(d\bar{z}_v/d\delta_v)$. This may be further simplified by noting that since the number of adults of either sex held constant at each generation, any fitness gain made by a focal individual due to a change of phenotype must be compensated by a decrease in fitness by the rest of the population (Rousset 2004, p. 96), i.e., $\partial w_{mi}^m/\partial z_{mi} + \partial w_{mi}^m/\partial \bar{z}_m = 0$ and $\partial w_{ti}^m/\partial z_{ti} + \partial w_{ti}^m/\partial \bar{z}_t = 0$. Thus, we eventually obtain for the derivatives of fitness

$$\frac{\partial w_{vi}^u}{\partial \delta_v} = \frac{\partial w_{vi}^u}{\partial z_{vi}} \left( \frac{dz_{vi}}{d\delta_v} - \frac{d\bar{z}_v}{d\delta_v} \right).$$

(SI.5)

Eq. (SI.5) is used to substitute for the derivatives of fitness in eq. (SI.3). To see how, consider the substitution for $\partial w_{mi}^m(\mathbf{0})/\partial \delta_m$ in

$$\frac{1}{N_m} \sum_{i=1}^{N_m} \frac{\partial w_{mi}^m(\mathbf{0})}{\partial \delta_m} = \frac{1}{N_m} \sum_{i=1}^{N_m} \frac{\partial w_{mi}^m(\mathbf{0})}{\partial z_{mi}} \left( \frac{d\bar{z}_m(\mathbf{0})}{d\delta_m} - \frac{d\bar{z}_m(\mathbf{0})}{d\delta_m} \right).$$

(SI.6)

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At \((\delta_m, \delta_i) = 0\), i.e. where all males are the same, the rate of change of fitness of a male \(i\) with respect to its phenotype is the same for all males \(\partial w^m_{mi}(0)/\partial z_{mi} = \partial w^m_{mk}(0)/\partial z_{mk}\). Thus, the index \(i\) denotes a representative male (or a focal male), rather than a specific one. Then, \(\partial w^m_{mi}(0)/\partial z_{mi}\) may be taken out of the sum in eq. (SI.6) and the index dropped for the function \(w^m_{mi}\) is dropped, giving

\[
\frac{1}{N_m} \sum_{i=1}^{N_m} p_{mi,t} \frac{\partial w^m_{mi}(0)}{\partial \delta_m} = \left( \frac{p_{mi} \cdot \overline{z}_{mi}}{d\delta_m} - \overline{p}_m \frac{d\overline{z}_m}{d\delta_m} \right) \frac{\partial w^m(0)}{\partial \overline{z}_m},
\]

where the overbar with index \(mi\) denotes averaging over all males \(\overline{x}_{mi} = \sum_{i=1}^{N_m} x_i/N_m\). Using a similar argument for all derivatives of fitness in eq. (SI.3), we obtain

\[
E[p_{m,t+1}|q_t] = \frac{1}{2} (\overline{p}_{m,t} + \overline{p}_t, t) + \frac{1}{2} \frac{N_i}{N_m} D_{m,t} \frac{\partial w^m(0)}{\partial \overline{z}_m} + \delta f \frac{N_i}{N_m} D_{m,t} \frac{\partial w^m(0)}{\partial \overline{z}_m} + 0(\delta^2)
\]

\[
E[p_{f,t+1}|q_t] = \frac{1}{2} (\overline{p}_{f,t} + \overline{p}_t, t) + \frac{1}{2} \frac{N_i}{N_f} D_{f,t} \frac{\partial w^m(0)}{\partial \overline{z}_m} + \delta f \frac{N_i}{N_m} D_{f,t} \frac{\partial w^m(0)}{\partial \overline{z}_m} + 0(\delta^2)
\]

where

\[
D_{m,t} = 2 \left( \frac{p_{mi} \cdot \overline{z}_{mi}}{d\delta_m} - \overline{p}_m \frac{d\overline{z}_m}{d\delta_m} \right), \quad \text{and} \quad D_{f,t} = 2 \left( \frac{p_{fi} d\overline{z}_f}{d\delta_t} - \overline{p}_f d\overline{z}_f \right)
\]

and the overbar with index \(f_j\) denotes averaging over all females \(\overline{x}_{fj} = \sum_{j=1}^{N_f} x_j/N_f\). We have added the subscript \(t\) in eq. (SI.9) to make the time dependence of \(D_{m,t}\) and \(D_{f,t}\) explicit, since they depend on the population genotypic realization at generation \(t\), \(q_t\).

The expectation of mutant frequencies in males and females from generation \(t\) to generation \(t + 1\) are found by marginalizing eq. (SI.8) over \(q_t\),

\[
p_{m,t+1} = E[p_{m,t+1}|q_t] = \sum_{q_t} E[p_{m,t+1}|q_t] Pr(q_t)
\]

\[
p_{f,t+1} = E[p_{f,t+1}|q_t] = \sum_{q_t} E[p_{f,t+1}|q_t] Pr(q_t)
\]

where \(Pr(q_t)\) is the distribution of allele frequencies at time \(t\). By inspection of eq. (SI.8), we see that only \(\overline{p}_{m,t}, \overline{p}_t, D_{m,t}\) and \(D_{f,t}\) depend on \(q_t\) and thus have to be marginalized over \(q_t\). Doing so will define the moments of the distribution \(Pr(q_t)\) required to calculate the expected allele frequency change over one generation. Since \(\overline{p}_{m,t}, \overline{p}_t, D_{m,t}\) and \(D_{f,t}\) are all evaluated in the absence of phenotypic differences \((\delta m, \delta i) = 0\), they are marginalized for a neutral process, and the expectation operator is written \(E^\circ[]\). We have \(E^\circ[\overline{p}_{m,t}] = p_m\) and \(E^\circ[\overline{p}_t, t] = p_t\) and evaluate \(E^\circ[D_{m,t}]\) and \(E^\circ[D_{f,t}]\) below.

We will calculate \(E^\circ[\overline{p}_{mi}(d\overline{z}_m/d\delta_m)]\) and \(E^\circ[\overline{p}_{ij}(d\overline{z}_f/d\delta_t)]\) together, and then \(E^\circ[\overline{p}_m(d\overline{z}_m/d\delta_m)]\) and \(E^\circ[\overline{p}_f(d\overline{z}_f/d\delta_t)]\), but first we note that the distribution of individual phenotype in terms of individual allele frequencies are given by \(\overline{z}_{mi} = z_m + \delta_m(2hp_{mi} + (1 - 2h)\overline{p}_m q), \) and \(\overline{z}_{fj} = z_f + \delta f(2hp_{fj} + (1 - 2h)\overline{p}_f q_j)\). So that average male and female phenotypic values are written as \(\overline{z}_m = \sum_{i} \overline{z}_{mi}/N_m = z_m + \delta m(2hp_{m,t} + (1 - 2h)\overline{p}_m q)\) and \(\overline{z}_f = \sum_{j} \overline{z}_{fj}/N_f = z_f + \delta f(2hp_{f,t} + (1 - 2h)\overline{p}_f q_j)\). We then obtain the derivatives with respect to \(\delta m\).
these averages and the phenotype of male \( i \), which are needed for the population statistics, as

\[
\frac{dz_{mi}}{dm} = 2hp_{mi} + (1 - 2h)1_{c_i}1_{\bar{q}_i} \\
\frac{dz_{mj}}{dm} = 2hp_{mj} + (1 - 2h)1_{c_j}1_{\bar{q}_j} \\
\frac{dz_{kt}}{dt} = 2hp_{kt} + (1 - 2h)1_{c_j}1_{\bar{q}_j}.
\]  

(SI.11)

Using eq. (SI.1) together with eq. (SI.11), we have

\[
E^0 \left[ \frac{dz_{mi}}{p_{mi}} \frac{dt}{dm} \right]_t = E^0 \left[ \frac{1_{c_i}1_{\bar{q}_i}}{2} \left( h(1_{c_i}1_{\bar{q}_i} + 1_{c_j}1_{\bar{q}_j}) + (1 - 2h)1_{c_i}1_{\bar{q}_i} \right) \right]_t
\]

(SI.12)

\[
= \frac{1}{N_m} E^0 \left[ \sum_{i=1}^{N_m} h/2(1_{c_i}1_{\bar{q}_i} + 21_{c_i}1_{\bar{q}_i} + 1_{c_j}1_{\bar{q}_i}) + (1 - 2h)1_{c_i}1_{\bar{q}_i} \right]
\]

(SI.13)

\[
= E^0 [h/2(1_{c_i}1_{\bar{q}_i} + 21_{c_i}1_{\bar{q}_i} + 1_{c_j}1_{\bar{q}_i}) + (1 - 2h)1_{c_i}1_{\bar{q}_i}]_t,
\]

where we have used that at neutrality, all males are expected to have the same genotypic composition. More succinctly, we write

\[
E^0 \left[ \frac{dz_{mi}}{p_{mi}} \frac{dt}{dm} \right]_t = h(p_{mi,t} + \eta_t) + (1 - 2h)\eta_t,
\]

(SI.14)

\[
E^0 \left[ \frac{dz_{kj}}{p_{kj}} \frac{dt}{dt} \right]_t = h(p_{kj,t} + \eta_t) + (1 - 2h)\eta_t,
\]

where \( \eta^H = E^0[1_{c_i}1_{\bar{q}_i}] \) is the probability that both the paternal and maternal alleles of an individual are mutants.

In the absence of phenotypic differences, this probability is equal for all individuals \( E^0[1_{c_i}1_{\bar{q}_i}] = E^0[1_{c_k}1_{\bar{q}_k}] \) for all \( i \) and \( k \) and irrespective of the sexes of the individuals. To see this, consider the recurrence for \( \eta \) over one generation: \( \eta_{t+1} = E^0[1_{c_i}1_{\bar{q}_i}]_{t+1} \). If individual \( i \) of generation \( t+1 \) has father indexed \( a \) and mother indexed \( c \) at generation \( t \),

\[
\eta_{t+1} = \frac{1}{4} E^0[(1_{c_a}1_{\bar{q}_a})(1_{c_c}1_{\bar{q}_c})]_t,
\]

(SI.15)

since the paternally inherited mutant of \( i \) is equally likely to be the paternally or the maternally inherited mutant of its father \( a \), and the maternally inherited mutant of \( i \) is equally likely to be the paternally or the maternally inherited mutant of its mother \( c \). This argument holds whatever the sex of \( i \), so \( \eta = E^0[1_{c_i}1_{\bar{q}_i}] \) does not depend on the sex of individual \( i \). A similar argument shows that \( \eta \) is also equal to the probability that a paternally inherited allele and a maternally inherited allele of two different, randomly sampled individuals are mutants, i.e. \( \eta = E^0[1_{c_i}1_{\bar{q}_j}] = E^0[1_{c_j}1_{\bar{q}_i}] \) with \( i \neq j \).
We now calculate $E^o[p_m(\delta \pi_m/\delta \delta_m)]$ and $E^o[p_\delta(\delta \pi_\delta/\delta \delta_\delta)]$. Using eq. (Sl.11) and rearranging to collect the terms that involve the same male $i$, and those that involve two different males $i$ and $k$, we have $E^o[p_m(\delta \pi_m/\delta \delta_m)]_t = E^o[2h/N_m^2(\sum_i \rho_m^i + \sum_{i,k,i\neq k} \rho_m^i \rho_m^k) + (1 - 2h)/(N_m^2)\sum_i \rho_m^i \rho_m^k]$. Letting expectation run through gives $2h/N_m(E^o[p_m^2]_t + (N_m - 1)E^o[p_m^2]_t) + (1 - 2h)/N_m(E^o[p_{m,k}^2]_t + (N_m - 1)E^o[p_m^2 \rho_m^k]_t)$ where $i \neq k$. Finally, factoring by $1/N_m$ yields

$$
E^o \left[ \frac{d \pi_m}{d \delta_m} \right]_t = \frac{1}{N_m} \left( 2h \left( E^o[p_m^2]_t - E^o[p_m^2]_t \right) + (1 - 2h) \left( E^o[p_m^2 \rho_m^k]_t - E^o[p_m^2\rho_m^k]_t \right) \right) + 2hE^o[p_m^2]_t + (1 - 2h)E^o[p_m^2 \rho_m^k]_t.
$$

Expanding the above in terms of indicator variables for paternally and maternally inherited alleles, we have $E^o[p_m^2]_t = E^o[2\eta + 2\eta_m, \eta_m]/2$, and we write $E^o[p_m^2]_t = (2\eta + \eta_m^2)/4$, where $\eta_m^2 = E^o[\rho_m^2 \rho_m^k]$ is the probability that two randomly sampled males $i \neq k$ both inherited the mutant allele from their fathers, and $\eta_m^2 = E^o[\rho_m^2 \rho_m^k]$ is the probability that they inherited the mutant allele from their mothers. Then, $E^o[p_m^2 \rho_m^k]_t = \eta_s$, and finally $E^o[p_{m,k}^2 \rho_m^k]_t = (\rho_m^2 + \rho_m^k)/2$, where $\rho_m^2 = E^o[\rho_m^2 \rho_m^k]$ is the probability that randomly sampled male $i$ has inherited the mutant from its father and that another randomly sampled male $k$ is homozygous for the mutant, and $\rho_m^k = E^o[\rho_m^2 \rho_m^k]$ is the probability that randomly sampled male $i$ has inherited the mutant from its mother and that another randomly sampled male $k$ is homozygous for the mutant. After using the similar argument for $E^o[p_{m,t}]_t$, we find that at generation $t$

$$
E^o \left[ \frac{d \pi_m}{d \delta_m} \right]_t = \frac{1}{N_m} \left\{ h \left( \frac{\rho_m^2 + \rho_m^k}{2} \right) + (1 - 2h) \left( \eta_s - \frac{\rho_m^2 + \rho_m^k}{2} \right) \right\}
$$

$$
E^o \left[ \frac{d \pi_t}{d \delta_t} \right]_t = \frac{1}{N_t} \left\{ h \left( \frac{\rho_t^2 + \rho_t^k}{2} \right) + (1 - 2h) \left( \eta_s - \frac{\rho_t^2 + \rho_t^k}{2} \right) \right\},
$$

where for two randomly sampled females $j \neq l, \eta_s = E^o[\rho_j^2 \rho_l^k], \eta_s = E^o[\rho_j^2 \rho_l^k], \eta_s = E^o[\rho_j^2 \rho_l^k]$ and $\rho_t^k = E^o[\rho_j^2 \rho_l^k]$.
for \( u \in \{m, f\} \). The latter can be interpreted as the neutral expectation of the covariance between genotype and phenotype at generation \( t \) in an individual of sex \( u \). Indeed, from eqs. (SI.6) and (SI.10), we have that \( K_u \) is also equal to

\[
K_u, t = \frac{1}{2} E^o \left[ \frac{1}{N_u} \sum_i p_{ui, t} \left( \frac{dz_{ui} (0)}{d\delta_u} - \frac{dz_{ui} (0)}{d\delta_u} \right) \right],
\]

and since \( z_{ui} = z_a + \delta_u (2h p_{ui} + (1 - 2h) \mathbb{1} \sigma^p i_{ui} ) \), this may be written as

\[
K_u, t = \frac{1}{2} \frac{1}{\delta_u} E^o \left[ \frac{1}{N_u} \sum_i p_{ui, t} (z_{ui} - z_a) \right]
= \frac{1}{2} \frac{1}{\delta_u} E^o \left[ \frac{1}{N_u} \sum_i (p_{ui, t} - \overline{p}_{ui, t}) (z_{ui} - z_a) \right].
\]

Therefore, \( K_u, t \) is proportional to the expected covariance \( E^o [\mathbb{1} p_{ui, t}, z_{ui}] \) at generation \( t \) between individual genotype and phenotype in sex \( u \), when mutant frequencies \( p_{ui, t} \) evolve neutrally.

**Closing the recursion.** Eq. (SI.18) gives the change of \( p_m \) and \( p_f \) over one generation, which depends on higher moments of the distribution of the mutant in the population \( (\eta_t, \kappa^p_{u, t}, \kappa^o_{u, t}, \rho^p_{u, t}, \rho^o_{u, t}) \). These latter also change from one generation to the next, and in order to evaluate the change of \( p_{m, t} \) and \( p_{f, t} \) over more than one generation, we need to characterize these recursions. Since they are evaluated at \( (\delta_m, \delta_f) = 0 \) in eq. (SI.18), it is sufficient to evaluate the recursions for \( \eta_t, \kappa^p_{u, t}, \kappa^o_{u, t}, \rho^p_{u, t}, \rho^o_{u, t} \) at neutrality, where they are only affected by genetic drift. We give these recursions below using standard population genetic methods (Karlin 1968, for example).

The probability that a gene sampled in an individual is mutant does not depend on the sex of the individual as it comes with equal probability from its father or its mother

\[
p_{m, t+1} = p_{f, t+1} = \frac{1}{2} (E^o [\mathbb{1} \sigma^p + \mathbb{1} \sigma^o] t) = \frac{1}{2} (p_{m, t} + p_{f, t}).
\]

The probability that the paternally and the maternally inherited allele of individual \( i \) at time \( t + 1 \) are both mutant, \( \eta_{t+1} \), is given in terms of neutral moments of gene frequency at generation \( t \) in eq. (SI.15) which, if expanded, gives

\[
\eta_{t+1} = \frac{1}{4} (2 \eta_t + \kappa^p_{u, t} + \kappa^o_{u, t}).
\]

where for a male \( i \) and a female \( j \), \( \kappa^p_u = E^o [\mathbb{1} \sigma^p_i \mathbb{1} \sigma^p_j] \), and \( \kappa^o_u = E^o [\mathbb{1} \sigma^o_i \mathbb{1} \sigma^o_j] \).

The probability that two paternally inherited alleles randomly sampled in two different males are both mutants at generation \( t + 1 \), \( \kappa^p_m, t+1 \), depends on whether the two males have the same father, which occurs with a probability denoted \( \Theta^p_m \) or not (which occurs with probability \( 1 - \Theta^p_m \)). These probabilities are referred to as probabilities of sibships. If the two males have the same father, which we index \( a \), then their paternal alleles can be either both copies of the paternal gene of \( a \) (with probability \( 1/4 \)), both copies of the maternal gene of \( a \) (with probability
or one is a paternal copy and one is a maternal copy (with probability 1/2). So, if two males have the same father, their two paternally sampled genes are mutants with probability \((1/4)\mathbb{E}[(\|s_a\| + \|q_a\|)^2]_t\). If they have different fathers, indexed \(a\) and \(b\), then the paternal copy of the first male may be the paternal or maternal copy of \(a\) (each with probability 1/2), and the paternal copy of the second male may be the paternal or maternal copy of \(b\) (also each with probability 1/2). In this case, the paternal alleles of the two individuals are both mutants with probability \((1/4)\mathbb{E}[(\|s_a\| + \|q_a\|)(\|s_b\| + \|q_b\|)]_t\). Combining these two cases, the probability that two randomly sampled paternal alleles of different males at generation \(t + 1\) are mutants is \(\kappa^\sigma_{m,t+1} = \Theta^\sigma_{m} (1/4)\mathbb{E}[(\|s_a\| + \|q_a\|)^2]_t + (1 - \Theta^\sigma_{m})(1/4)\mathbb{E}[(\|s_a\| + \|q_a\|)(\|s_b\| + \|q_b\|)]_t\) which, after letting expectation \(\mathbb{E}[]\) run through and using previous definitions, gives \(\kappa^\sigma_{m,t+1} = \Theta^\sigma_{m} (2\eta_t + p_{m,t} + p_{t,t})/4 + (1 - \Theta^\sigma_{m})(2\eta_t + \kappa^\sigma_{m,t} + \kappa^\varnothing_{m,t})/4\). In fact, we find more generally that the probabilities that the paternal alleles of two males \((x = m)\), or of two females \((x = f)\), or of a male and female \((x = c)\) are mutants at generation \(t + 1\) are given by

\[
\kappa^{m}_{x,t+1} = \Theta^{\sigma}_{x} \left(2\eta_t + p_{m,t} + p_{t,t} \right) / 4 + \frac{1 - \Theta^{\sigma}_{x}}{4} (2\eta_t + \kappa^{\sigma}_{m,t} + \kappa^{\varnothing}_{m,t})
\]  

(24)

where \(\Theta^{\sigma}_{x}\) is the probability that two females have the same father and, \(\Theta^{\sigma}_{c}\) is the probability that a male and a female have the same father.

Using a similar argument, we find that the probabilities that the maternal alleles of two males \((x = m)\), or of two females \((x = f)\), or of a male and female \((x = c)\) are mutants at generation \(t + 1\) are given by

\[
\kappa^{0}_{x,t+1} = \Theta^{\varnothing}_{x} \left(2\eta_t + p_{m,t} + p_{t,t} \right) / 4 + \frac{1 - \Theta^{\varnothing}_{x}}{4} (2\eta_t + \kappa^{\sigma}_{c,t} + \kappa^{\varnothing}_{c,t})
\]  

(25)

where \(\Theta^{\varnothing}_{x}\) is the probability that two individuals, whose sexes are given by \(x\), have the same mother.

The probability \(\rho^{s}_{m,t+1} = \mathbb{E}[(\|s_a\| - \|s_k\|)(\|s_a\| - \|s_k\|)]_t+1\) that two (different) paternally inherited alleles and one maternally inherited allele at generation \(t + 1\) are mutants depends on whether the males from which the paternal alleles are sampled (males \(i\) and \(k\) here) have the same father (indexed \(a\)) or different fathers \((a\) and \(b\)). Using a similar argument as in the preceding section, and indexing by \(c\) the mother of the male who holds the maternal allele, we have \(\rho^{s}_{m,t+1} = \Theta^{\sigma}_{m} (1/8)\mathbb{E}[(\|s_a\| + \|q_a\|)^2(\|s_c\| + \|q_c\|)]_t + (1 - \Theta^{\sigma}_{m})(1/8)\mathbb{E}[(\|s_a\| + \|q_a\|)(\|s_b\| + \|q_b\|)]_t\). Then, expanding and letting expectation run through, we have: \(\rho^{s}_{m,t+1} = \Theta^{\sigma}_{m} \left(2\eta_t + \kappa^{\sigma}_{c,t} + \kappa^{\varnothing}_{c,t} + 2\rho^{\sigma}_{c,t} + 2\rho^{\varnothing}_{c,t} \right) / 8 + (1 - \Theta^{\sigma}_{m}) \left(\frac{\kappa^{\sigma}_{2m,t} + \kappa^{\varnothing}_{2m,t} + 2\rho^{\sigma}_{2m,t} + 2\rho^{\varnothing}_{2m,t} + \rho^{\sigma}_{m,t} + \rho^{\varnothing}_{m,t}}{8} \right)\), where \(\kappa^{\sigma}_{2m,t} = \mathbb{E}[(\|s_a\| + \|q_a\|)(\|s_a\| + \|q_a\|)]_t\) and \(\kappa^{\varnothing}_{2m,t} = \mathbb{E}[(\|q_a\| + \|q_b\|)(\|q_a\| + \|q_b\|)]_t\) are the probabilities that the paternal and maternal alleles, respectively, of two randomly sampled (without replacement) males \(a\) and \(b\) and a female \(c\) at generation \(t\) are all mutants. We find in general that for \(x \in \{m, f, c\}\)

\[
\rho^{\sigma}_{x,t+1} = \Theta^{\sigma}_{x} \left(2\eta_t + \kappa^{\sigma}_{c,t} + \kappa^{\varnothing}_{c,t} + 2\rho^{\sigma}_{c,t} + 2\rho^{\varnothing}_{c,t} \right) / 8
\]  

(26)

\[
+ \frac{1 - \Theta^{\sigma}_{x}}{8} \left(\kappa^{\sigma}_{2m,t} + \kappa^{\varnothing}_{2m,t} + 2\rho^{\sigma}_{2m,t} + 2\rho^{\varnothing}_{2m,t} + \rho^{\sigma}_{m,t} + \rho^{\varnothing}_{m,t} \right)
\]
Similarly, the probability that two (different) maternally inherited alleles and one paternally inherited allele from two individuals are mutants at generation \( t + 1 \), 
\[
\rho^Q_{x,t+1} = E^t[\mathbb{1}_{Q_a} \mathbb{1}_{Q_b} \mathbb{1}_{Q_c}] 
\]
develops on whether individuals \( i \) and \( j \) from which maternal genes are sampled have the same mother (indexed \( c \)) or different mothers (\( c \) and \( d \)), where \( a \) is the father of the individual whose paternal gene is sampled. Then for \( x \in \{m, f, c\} \)
\[
\begin{aligned}
\rho^Q_{x,t+1} &= \Theta_x^Q \left( \frac{2\eta_t + \kappa^Q_m + \kappa^Q_d + 2\rho^Q_m + 2\rho^Q_d}{8} \right) \\
&\quad + \frac{1 - \Theta_x^Q}{8} \left( \frac{\eta^Q_{2m,t} + \eta^Q_{2f,t} + 2\rho^Q_{2m,t} + 2\rho^Q_{2f,t}}{2} + \rho^Q_{m,t} + \rho^Q_{d,t} \right) 
\end{aligned}
\]  
(\text{SI.27})

where \( \eta^Q_{2f,t} = E^t[\mathbb{1}_{Q_a} \mathbb{1}_{Q_b} \mathbb{1}_{Q_d}] \) and \( \eta^Q_{2m,t} = E^t[\mathbb{1}_{Q_a} \mathbb{1}_{Q_c} \mathbb{1}_{Q_d}] \) are the probabilities that the paternal and maternal alleles, respectively, of a male \( a \) and two different females \( c \) and \( d \) at generation \( t \) are all mutants.

The probability that three alleles sampled from different individuals are mutants depends on the probabilities of sibship of three individuals. In order to consider the iteration of the probability \( \xi^Q_t \), i.e. that three randomly chosen paternally inherited genes are mutants, we need to separate the cases where all three individuals are males (subscript \( x = 3m \)), all three are females (\( x = 3f \)), two are males and one is female (\( x = 2m \)), or two are females and one is male (\( x = 2f \)). The probabilities that three paternal alleles are mutants then depend on whether all three individuals have the same father, which occurs with a probability we write as \( \Xi^Q_2 \), whether only two have a same father (with probability \( \Xi^Q_1 \)), or if none of the three have the same father (with probability \( 1 - \Xi^Q_2 - \Xi^Q_1 \)). If they all have the same father (indexed \( a \)), then they are all mutants if they have inherited the mutant gene from the maternal or paternal locus from \( a \). And similar arguments apply for the case when only two have the same father (indexed \( a \), and the other father is indexed \( b \) or if they have three different fathers (indexed \( a, b \) and \( c \)) to give \( \eta^Q_{3f,t} = \Xi^Q_1 E^t[\mathbb{1}_{Q_a} + \mathbb{1}_{Q_b} + \mathbb{1}_{Q_c}] / 8 + \Xi^Q_2 E^t[\mathbb{1}_{Q_a} + \mathbb{1}_{Q_c}] / 8 + (1 - \Xi^Q_1 - \Xi^Q_2) E^t[\mathbb{1}_{Q_b} + \mathbb{1}_{Q_c}] / 8 \), which, expanding and letting expectation run through, results in
\[
\begin{aligned}
\xi^Q_{3f,t+1} &= \Xi^Q_1 \frac{\eta^Q_{3m,t} + \rho^Q_{3m,t}}{4} + \frac{\Xi^Q_2}{4} \left( \frac{\eta^Q_{3m,t} + \rho^Q_{3m,t} + 3\rho^Q_{3m,t}}{4} \right) + \frac{1 - \Xi^Q_1 - \Xi^Q_2}{4} \left( \frac{\eta^Q_{3f,t} + \rho^Q_{3f,t} + 3\rho^Q_{3f,t}}{4} \right) 
\end{aligned}
\]  
(\text{SI.28})

Similarly, the probability that three randomly chosen maternally inherited genes \( \xi^Q_t \) are mutants can be expressed in terms of the probabilities that the individuals have the same mother,
\[
\begin{aligned}
\xi^Q_{x,t+1} &= \Xi^Q_1 \frac{\eta^Q_{3m,t} + \rho^Q_{3m,t}}{4} + \frac{\Xi^Q_2}{4} \left( \frac{\eta^Q_{3m,t} + \rho^Q_{3m,t} + 3\rho^Q_{3m,t}}{4} \right) + \frac{1 - \Xi^Q_1 - \Xi^Q_2}{4} \left( \frac{\eta^Q_{3f,t} + \rho^Q_{3f,t} + 3\rho^Q_{3f,t}}{4} \right) 
\end{aligned}
\]  
(\text{SI.29})

where \( \Xi^Q_2 \) is the probability that the three holders (whose sexes are given by \( x \in \{3m, 3f, 2m, 2f \} \)) have the
same mother, and $\Xi^3_2$ is the probability that out of the three individuals, two have the same mother. The moments $\varsigma_{m,t+1}$ and $\varsigma_{f,t+1}$ ($x \in \{m, f\}$) also satisfy the recurrences given by eqs. (SI.28),(SI.29), and complete the necessary moments to iterate eq. (SI.18).

**Probability of fixation of an autosomal mutant.** We proceed to calculate the probability of fixation of the mutant by iterating its expected change over many generations. Eqs. (SI.22) - (SI.29) define the changes in the moments of the population genotypic distribution of a neutral mutant. Since eqs. (SI.22) - (SI.29) are all linear in the relevant moments, we may express the set of recurrences as a matrix operation: $\mathbf{p}_{t+1} = \mathbf{A}^t \mathbf{p}_t$, where $\mathbf{p}_t$ is a $23 \times 1$ vector which collects the necessary moments of $\Pr(\mathbf{q}_t)$ ($\mathbf{p}_m, \mathbf{p}_f, \eta, \kappa_0^m, \kappa_0^f, \rho_0^m, \rho_0^f, \varsigma_{m,t+1}, \varsigma_{f,t+1}$) for $x \in \{m, c, f\}$, $y \in \{3m, 3f, 2m, 2f\}$, and $\mathbf{A}^t$ is a $23 \times 23$ matrix defined by eqs. (SI.22) - (SI.29).

Eq. (SI.18) adds the effects of selection to the expected mutant frequency change. Since it is also linear in $\mathbf{p}_{m,t}, \mathbf{p}_{f,t}, \eta, \kappa_0^m, \kappa_0^f, \rho_0^m, \rho_0^f$, it may also be represented as a matrix operation, giving

$$\mathbf{p}_{t+1} = \mathbf{A}^t \mathbf{p}_t \quad \text{with} \quad \mathbf{A} = \mathbf{A}^0 + \delta^m \mathbf{A}_m + \delta^f \mathbf{A}_f + O(\delta^3),$$  

(SI.30)

where the $23 \times 23$ matrices $\mathbf{A}_m$ and $\mathbf{A}_f$ describes the first order perturbation of average frequency change due to mutant effect in males and females respectively. Eq. (SI.30) fully characterizes the expected frequency change of a mutant in a sexually dimorphic population at any generation i.e., the model is dynamically sufficient.

Explicit expression for these large matrices are omitted from this paper, but they can be found straightforwardly from eqs. (SI.22) - (SI.29) for $\mathbf{A}^0$ and from eq. (SI.18) for $\mathbf{A}_m$ and $\mathbf{A}_f$. Their entries will of course depend on the order chosen for the entries of $\mathbf{p}_t$. We will assume here that the first 15 entries of $\mathbf{p}_t$ are

$$\mathbf{p}_t = (\mathbf{p}_m, \mathbf{p}_f, \eta, \kappa_0^m, \kappa_0^f, \rho_0^m, \rho_0^f, \varsigma_{m,t+1}, \varsigma_{f,t+1}, \ldots)^T.$$

We derive the expression for the fixation probability $\pi$ of the mutant by estimating the asymptotic sum of expected allele-frequency change of the allele in males and females (Leturque and Rouset 2002; Rouset 2004; Lessard and Ladret 2007; Lehmann and Rouset 2009). The fixation probability of the mutant $\pi_m$ in males, and $\pi_f$ in females is the asymptotic average frequency of the mutant in each sex

$$\pi_m = \lim_{t \to \infty} \mathbf{p}_{m,t}, \quad \pi_f = \lim_{t \to \infty} \mathbf{p}_{f,t}.$$  

(SI.31)

Because the mutant allele eventually is either eliminated or fixated in the population, the fixation probability in males and females is the same $\pi_m = \pi_f = \pi$. The fixation probabilities in males and females could be obtained from the asymptotic vector $\lim_{t \to \infty} \mathbf{A}^t \mathbf{p}_0$, but this is difficult as it requires the calculation of $\mathbf{A}^t$’s eigenvectors. We rely on an alternative scheme to obtain $\pi$. To that aim, it is convenient to express the fixation probability of the mutant as the average

$$\pi = \alpha \pi_m + (1 - \alpha) \pi_f,$$

(SI.32)

where the weight $\alpha$ is chosen such that the expected frequency change of a neutral mutant in any generation $t$ is
zero: \( \alpha E[\Delta p_{m,t}] + (1 - \alpha)E[\Delta p_{t,t}] = 0 \). In this case, \( \alpha = 1/2 \) for a diploid, autosome genetic system. Together, eqs. (SI.31) & (SI.32) imply that \( \pi \) is the average sum of gene frequency change in males and females, from the appearance to the eventual fixation or loss of the mutant

\[
\pi = \alpha p_{m,0} + (1 - \alpha)p_{t,0} + \sum_{t=0}^{\infty} \left( \alpha E[\Delta p_{m,t}] + (1 - \alpha)E[\Delta p_{t,t}] \right).
\]

The probability of fixation of a mutant with initial frequencies \( p_{m,0} \) in males and \( p_{t,0} \) females is approximated to the first order of \( \delta: \pi = \alpha p_{m,0} + (1 - \alpha)p_{t,0} + \alpha \delta_m (\partial \pi(0) / \partial \delta_m) + (1 - \alpha)\delta_t (\partial \pi(0) / \partial \delta_t) + O(\delta^2) \). We begin by considering the first order effects of male phenotype on \( \pi \). Using eq. (SI.33), it is \( \partial \pi(0) / \partial \delta_m = \langle \partial / \partial \delta_m \rangle \sum_{t=0}^{\infty} \left( \alpha E[\Delta p_{m,t}] + (1 - \alpha)E[\Delta p_{t,t}] \right) \delta_m = \alpha \sum_{t=0}^{\infty} (\partial / \partial \delta_m) (p_{t+1} - p_t) \delta_m = 0 \), where \( p = p_m, p_t, \ldots \) and \( \alpha = (\alpha, 1 - \alpha, 0, \ldots, 0) \) is such that when dot multiplied with \( p \), it collects and sums \( p_{m,t} \) and \( p_{t,t} \) weighted by the reproductive values. Then, using eqs. (SI.30), we have \( \partial (p_{t+1} - p_t) / \partial \delta_m = \hat{A}_m p_t \). So the male perturbation of the probability of fixation may be written as

\[
\frac{\partial \pi(0)}{\partial \delta_m} = \alpha \cdot \sum_{t=0}^{\infty} \hat{A}_m p_t \bigg|_{\delta_m = \delta_t = 0}.
\]

The sum \( \sum_{t=0}^{\infty} p_t |_{\delta_m = \delta_t = 0} \), which we write as \( \sum_{t=0}^{\infty} p_t^\circ \) where \( p_t^\circ = A^\circ p_t^\circ \), does not converge as \( A^\circ \) is not regular. This means \( \hat{A}_m \) cannot be factored out of the sum in eq. (SI.34). To circumvent this problem, we construct an iteration around a centred variable using the zero row-sum property of matrix \( \hat{A}_m \) (Lehmann and Roussel 2009).

To that aim, we define a vector \( q^\circ \) and a matrix \( Q^\circ \) such that (i) \( \sum_{t=0}^{\infty} \hat{A}_m p_t^\circ = \sum_{t=0}^{\infty} \hat{A}_m (p_t^\circ - q_t^\circ) \), (ii) \( p_t^\circ - q_t^\circ = (A^\circ - Q^\circ)(p_t^\circ - q_t^\circ) \), and (iii) \( \lim_{t \to \infty} (p_t^\circ - q_t^\circ) = 0 \). The choice of \( q_t^\circ \) with all vector elements being equal to \( \alpha t, t + 1 \) \( p_{m,t} \), which acts as a reference variable, and \( Q^\circ = (q_{ij}) \) with all elements of column 2 being equal to \( \alpha \), all elements of column 2 being equal to \( 1 - \alpha \), and zero otherwise satisfies all three conditions. In effect, this choice of the vector \( q_t^\circ \) centers the iteration around the mutant frequency averaged across the sexes according to their reproductive class (this average is the reference variable), while \( Q^\circ \) provides the iteration of the reference variable.

Using properties (i)-(iii) in the preceding paragraph, we can now factorize \( \sum_{t=0}^{\infty} \hat{A}_m p_t = \hat{A}_m \sum_{t=0}^{\infty} (p_t^\circ - q_t^\circ) = \hat{A}_m \sum_{t=0}^{\infty} (A^\circ - Q^\circ)(p_t^\circ - q_t^\circ) \). With all eigenvalues of \( (A^\circ - Q^\circ) \) being less than 1 in absolute value (Lehmann and Roussel 2009, p. 47), the sum \( d^\circ = \sum_{t=0}^{\infty} (A^\circ - Q^\circ)(p_t^\circ - q_t^\circ) \) can be evaluated as \( |I - A^\circ + Q^\circ|^{-1} \), where \( I \) is the identity matrix, so we have

\[
\frac{\partial \pi(0)}{\partial \delta_m} = \alpha \cdot \hat{A}_m d^\circ, \quad \text{where} \quad d^\circ = |I - A^\circ + Q^\circ|^{-1} (p_0 - q_0).
\]

All the arguments used to derive eq. (SI.35) can be used for \( \partial \pi(0) / \partial \delta_t \), and we find \( \partial \pi(0) / \partial \delta_t = \alpha \cdot \hat{A}_t d^\circ \). Hence,
the fixation probability to the first order in selection intensity is

$$\pi = \alpha p_{m,0} + (1 - \alpha) p_{f,0} + \delta_m \alpha \cdot \hat{A}_m \cdot d^0 + \delta_f \alpha \cdot \hat{A}_f \cdot d^0 + O(\delta^2).$$  \(\text{(SI.36)}\)

The entries of \(d^0\) can be interpreted in terms of mean coalescence times in the resident population. To see this, we first note that if the expected initial frequency of the mutant is the same in males and females, then \(p_{m,0} = p_{f,0} = p_0\), which is equivalent to assuming that mutation rate is the same in males and females. Then, if the mutant arose as a single copy, \(p_0 = 1/(2N)\), where \(N = N_m + N_f\), and we have \(p_0 - q_0 = (0, 0, -1/(2N), -1/(2N), \ldots, -1/(2N))^T\). In this case, element \(d_i^t\) for \(i \geq 3\) of \(d^0\) is

$$d_i^t = -T_{(i)}/(2N),$$  \(\text{(SI.37)}\)

where \(T_{(i)}\) is the mean coalescent time into a single individual of a set of gene lineages initially residing in state \(i\) (Lehmann and Rousset 2009, eqs. A-28 & A-29). State here refers to the configuration of the sampled gene lineages, which are given by the entries of \(p_i\), e.g., for \(i = 3\), if the third entry of \(p_i\) corresponds to \(\eta_3\), the probability that an individual’s paternal and maternal alleles are both mutant, so \(d_3^3 = -T_{(3)}/(2N)\), where \(T_{(3)}\) is the expected number of generations taken for the paternal and maternal genes of an individual to coalesce.

Substituting for \(\alpha = 1/2\) (for an autosomal gene) and for matrices \(\hat{A}_m\) and \(\hat{A}_f\) into eq. (SI.36), the probability of fixation of a single copy mutant \((p_{m,0} = p_{f,0} = 1/(2N))\) can be expressed as eq. (1) in the main text, where if \(\mathbf{p}_t = (p_m, p_f, \kappa_m^c, \kappa_m^c, \kappa_f^c, \kappa_f^c, \kappa_f^m, \kappa_f^m, \kappa_f^m, \kappa_f^m, \rho_m^c, \rho_f^c, \rho_m^c, \rho_f^c, \rho_m^f, \rho_f^f, \rho_m^f, \rho_f^f, \ldots)^T\), the sex-specific weights \(K_m\) and \(K_f\) are given by

$$K_m = \frac{1}{4} \left(1 - \frac{1}{N_m}\right) \left[ -h \left( \frac{d_{11}^m + d_{12}^m}{2} - (1 - 2h) \left( \frac{d_{11}^m + d_{12}^m - d_3^m}{2} \right) \right) \right]$$

$$K_f = \frac{1}{4} \left(1 - \frac{1}{N_f}\right) \left[ -h \left( \frac{d_{11}^f + d_{12}^f}{2} - (1 - 2h) \left( \frac{d_{11}^f + d_{12}^f - d_3^f}{2} \right) \right) \right],$$  \(\text{(SI.38)}\)

with \(d_i\) as the \(i\)th entry of the vector \(d^0\) defined in eq. (SI.35). This shows that \(K_m\) and \(K_f\) may be interpreted in terms of coalescent times for sampled genes (eq. SI.37). Alternatively, using eq. (SI.21), we see that \(K_m\) and \(K_f\) can be interpreted as the expected covariance between between genotype and phenotype in males and females respectively, cumulated over the neutral segregation of the mutant

$$K_u = \frac{1}{2} \sum_{t=0}^{\infty} K_{u,t} = \frac{1}{4} \sum_{t=0}^{\infty} \sum_{t=0}^{\infty} E^0 [C(p_{ui,t}, z_{ui})]$$  \(\text{(SI.39)}\)

where the sum runs from the appearance to the eventual fixation or loss of the mutant.

**Probabilities of sibships of three individuals.** Until now, all our results hold for any arbitrary population size, but this implies tracking many gene associations. Indeed, as eqs. (SI.22) - (SI.29) show, the iteration of eq. (SI.18) over multiple generations depends on the six probabilities of sibships over two individuals, \(\Theta^c_x\) and \(\Theta^f_x\) \((x \in \{m, c, f\})\), and the eight probabilities of sibships over three individuals \(\Xi v^c_m\) and \(\Xi v^f_m\) \((v \in \{2, 3\}, w \in \{m, f\})\). Therefore,
$K_m$ and $K_1$ (eq. SI.38) also depend on these fourteen probabilities. As we show below, we can significantly reduce the number of necessary probabilities of sibships by approximating the probabilities of sibship of three individuals $\Xi_\nu^{c_3}$ and $\Xi_\nu^{c_2}$ as functions of the probabilities of sibship of two individuals $\Theta^{c_2}$ and $\Theta^{c_1}$ when we only consider the first order effects of finite population size $O(1/N)$.

The probability that three randomly sampled adult males have the same father is $\Xi_3^{c_3} = E^\nu[\sum N_m (W_m^3)/(N^3)]$. In the absence of phenotypic differences, each male has the same distribution of reproductive output and $\Xi_3^{c_3} = 1/((N_m - 1)(N_m - 2))E^\nu[W_m^3 - 3W_m^2 + 2W_m]$. If we assume that the distribution for $W_m$ is sufficiently well-behaved, and that the number of adult descendants of a male stays bounded as populations size ($N$) tends to infinity (or that $E^\nu[W_m^x]$, $x \geq 0$, remains bounded as $N \to \infty$), we find that none of the terms in $\Xi_3^{c_3}$ are of order $1/N$ or more, i.e. $\Xi_3^{c_3} = 0 + O(1/N^2)$, so the probability that three randomly sampled adult males have the same father can be approximated to being zero when $N$ is large. Similarly, we find that all probabilities of sibship three genes in the same individual are approximately zero, $\Xi_3^{c_2} = \Xi_3^{c_1} = 0 + O(1/N^2)$ for $x \in \{3m, 3f, 2m, 2f\}$.

Rather than calculating $\Xi_3^{c_3}$, the probability that out of three males only two have the same father directly, it is easier to consider the probability that out of three males, none have the same father. These two probabilities are related by $1 - \Xi_3^{c_3} - \Xi_3^{c_2} = 1 - \Xi_3^{c_2}$ (since $\Xi_3^{c_3} = 0 + O(1/N^2)$). The probability that out of three males, none have the same father is given by the expected value of the ratio of the number of ways three individuals may be sampled from the male offspring of three different adult males to the number of ways of sampling three males out of the entire male population $1 - \Xi_3^{c_2} = [\sum_{i,j,k} N_m W_m^3 W_m W_k/(N_m^2)]_{i \neq j \neq k \neq i}$, which after taking the sum and denominator outside reduces to $E^\nu[W_m^3 W_m^3 W_m W_m]_{i \neq j \neq k \neq i}$. Again by assuming that the number of adult descendants of a male stays bounded as populations size tends to infinity, using the delta method (Oehlerl 1992), and observing that $E^\nu[W_m^3] = 1$, we obtain $1 - \Xi_3^{c_2} = 1 + 3\Theta^{c_2} [W_m W_m W_m]_{i \neq j} + O(1/N^2)$.

The covariance term $C^{c_2} [W_m W_m W_m]_{i \neq j}$ may be expressed in terms of $\Theta^{c_2}$. The probability that two males do not have the same father is, by definition, $1 - \Theta^{c_2}$, but it is also given by $E^\nu[\sum_{i,j} W_m^3 W_m^3 W_m^3/(N_m^2)] = E^\nu[W_m W_m W_m]_{i \neq j} = C^{c_2}[W_m W_m W_m]_{i \neq j} + 1$, so that $C^{c_2}[W_m W_m W_m]_{i \neq j} = -\Theta^{c_2}$. Hence substituting back into the probability that out of three males none have the same father, and solving for $\Xi_3^{c_2}$, we obtain that the probability that out of three males only two have the same father is

$$\Xi_3^{c_2} = 3\Theta^{c_2} + O(1/N^2). \quad (SI.40)$$

The remaining probabilities can be derived by using the same argument, and that $E^\nu[W_m^4] = N_f/N_m$, producing

$$\Xi_2^{c_2} = 3\Theta^{c_2} + O(1/N^2)$$

$$\Xi_2^{c_1} = \frac{2}{3}N_m + \frac{4}{3}\Theta^{c_2} + \frac{1}{3}\Theta^{c_1} + O(1/N^2) \quad (SI.41)$$

$$\Xi_2^{c_2} = \frac{2}{3} \left(\frac{2}{N_m} - \frac{1}{N_f}\right) + \frac{4}{3}\Theta^{c_2} + \frac{1}{3}\Theta^{c_1} + O(1/N^2).$$
By symmetry, we find that the probabilities of sibship of three maternal genes are given to the order $O(1/N)$ by

$$\Xi_{3m}^2 = 3\Theta_m^2 + O(1/N^2)$$

$$\Xi_{3f}^2 = 3\Theta_f^2 + O(1/N^2)$$

$$\Xi_{2m}^2 = \frac{2}{3}\left(\frac{2}{N_f} - \frac{1}{N_m}\right) + \frac{4}{3}\Theta_e^2 + \frac{1}{3}\Theta_f^2 + O(1/N^2)$$

$$\Xi_{2f}^2 = \frac{2}{3N_f} + \frac{4}{3}\Theta_e^2 + \frac{1}{3}\Theta_f^2 + O(1/N^2).$$

So assuming the population is large, the iteration of eq. (SI.18) over many generations depends only on the six probabilities of sibships over two individuals, $\Theta_{e,f}^2$ and $\Theta_{m,f}^2 (x \in \{m, c, f\})$.

**Solving for $K_m$ and $K_f$ in terms of the probabilities of sibships of two individuals.** Having expressed the eight probabilities of sibships of three individuals in terms of the probabilities of sibships of two individuals $\Theta_{e,f}^2$, the matrix $A^e$ now only depends on these latter six probabilities of sibships, and therefore, so do $K_m$ and $K_f$ (eq. SI.38). Despite this simplification, solving explicitly for $K_m$ and $K_f$ still requires inverting a 23x23 matrix, $(I - A^e + Q^e)^{-1}$, which is computationally expensive and unlikely to yield results easy to interpret. Numerical results for $K_m$ and $K_f$ with arbitrary dominance are shown in fig. 4.D of the main text. However, if $h = 1/2$, only the first nine entries of $p$, are required to generate the expected frequency change over many generations, and hence the probability of fixation. Thus, $A^e$ reduces to a 9x9 matrix. In this case, $(I - A^e + Q^e)^{-1}$ can be inverted analytically, and using (SI.38) with $h = 1/2$, $K_m$ and $K_f$ are as eq. (A.2) in the main text.

**Probabilities of sibship of two individuals.** The probability of fixation of a mutant depends on the probabilities of sibship of two individuals in the resident population. Here, the probabilities of sibship are expressed in terms of the first ($\mu$’s) and second ($\nu$ and $\rho$) moments of the distribution of offspring produced by a resident male and a resident female to give table 1 of the main text.

The probability that two randomly sampled adult males have the same father, $\Theta_m^2$, is given by the expected value of the ratio of the number of ways two individuals may be sampled from the number of adult males produced by each male, to the number of ways of sampling two males out of the entire male population, i.e.,

$$\Theta_m^2 = E^{\left[\sum_{i=1}^{N_m} \left(\frac{W_m^i}{N_m}\right)\right]}(\frac{N_m}{N_m}) - 1\right) [V^e[W_m^m] + E^e[W_m^m]E^e[W_m^m] - 1]$. The expected number of male adults produced by a male in the absence of phenotypic differences, $E^e[W_m^m] = 1$, so the probability that two randomly sampled adult males have the same father reduces to $\Theta_m^2 = V^e[W_m^m]/(N_m - 1)$. Conditioning on the number of male juveniles produced in the population, and using the law of total variance, this gives

$$\Theta_m^2 = \frac{1}{N_m - 1} \left( N_m^2 V^e \left[\frac{J_m}{J_m}\right] + E^e \left[\frac{V^e[W_m^m]}{J_m}\right] \right).$$

The second variance term in eq. (SI.43) depends on how culling or regulation is assumed to take place, which is
assumed here to occur by sampling juveniles without replacement. In this case, $W_{m_i}^m$ follows a hypergeometric distribution with $N_m$ draws and parameters given by the realization of $J_{m_i}^m$, with initial probability of success $J_{m_i}^m/J_m$ and a total population size of $J_m$. Then, $E^o[V^o[W_{m_i}^m/J_{m_i}^m, J_m]] = E^o[N_m J_{m_i}^m (J_m - J_{m_i}^m) (J_m - N_m) / (J_m^2 (J_m - 1))]$.

Both variance terms in eq. (SI.43) are approximated omitting terms of order $1/N^2$ using the delta method. With assumption eq. (A.1) in the main text, the second variance term can be approximated as

$$\frac{1}{N_m - 1} E^o \left[ \frac{N_m J_{m_i}^m (J_m - J_{m_i}^m) (J_m - N_m)}{J_m^2 (J_m - 1)} \right] \approx \frac{E^o [J_{m_i}^m]}{E^o[J_m]} = \frac{\mu_{m_i}^m}{\mu_T} = \frac{1}{N_m}. \quad (SI.44)$$

Then, using the delta method with the variance operator, the first variance term in eq. (SI.43) is

$$\frac{N_m^2}{N_m - 1} V^o \left[ \frac{J_{m_i}^m}{J_m} \right] = N_m V^o \left[ \frac{J_{m_i}^m}{J_m} \right]^2 + O(1/N^2) = N_m \frac{\mu_{m_i}^m}{\mu_T^2} + O(1/N^2). \quad (SI.45)$$

Finally, substituting eqs. (SI.44)(SI.45) into eq. (SI.43) gives $\Theta_{m_i}^{c'}$ in table 1 of the main text. Using the same argument, we find a similar form for the probabilities that two females have the father $\Theta_{f}^{c'}$, that two males have the same mother $\Theta_{m}^{c}$, and that two females have the same mother $\Theta_{f}^{c}$ (see table 1 in the main text).

The probability that a male and a female have the same father $\Theta_{c}^{c'}$ is given by $E^o[\sum_{i=1}^{N_m} W_{m_i}^m J_{m_i}^f / (N_m N_i)]$, where $W_{m_i}^f$ is the random variable for the number of female breeders produced by male $i$. By conditioning on the juvenile production of every individual and using the assumption that male and female offspring are culled independently, we have $\Theta_{c}^{c'} = N_m E^o[J_{m_i}^m J_{m_i}^f / (J_m J_i)]$. The delta method is used to approximate the latter. Then, expanding about the means of $J_{m_i}^m, J_{m_i}^f, J_m$, and $J_i$ and using condition eq. (A.1) in the main text, we have

$$\Theta_{c}^{c'} = \frac{1}{N_m} + N_m \frac{C[J_{m_i}^m, J_{m_i}^f]}{E[J_m] E[J_i]} = \frac{1}{N_m} \left( 1 + \frac{\rho_{m_i}^m f}{\mu_{m_i}^m \mu_{m_i}^f} \right), \quad (SI.46)$$

where $\rho_{m_i}^m f = C[J_{m_i}^m, J_{m_i}^f]$ is the covariance between the number of male and offspring juveniles fathered by a male. Using a similar argument, the probability that a male and a female have the same mother is found as in table 1 of the main text.

References

KARLIN, S., 1968, Equilibrium Behavior of Population Genetic Models with Non-Random Mating: Part II: Pedigrees, Homozygosity and Stochastic Models. Journal of Applied Probability 5(3): 487+.

LEHMANN, L. and F. ROUSSET, 2009, Perturbation expansions of multilocus fixation probabilities for frequency-dependent selection with applications to the Hill-Robertson effect and to the joint evolution of helping and punishment. Theoretical population biology 76(1): 35–51.

LESSARD, S. and V. LADRET, 2007, The probability of fixation of a single mutant in an exchangeable selection model. Journal of mathematical biology 54(5): 721–744.
LETURQUE, H. and F. ROUSSET, 2002, Dispersal, kin competition, and the ideal free distribution in a spatially heterogeneous population. Theoretical population biology 62(2): 169--180.

OEHLERT, G. W., 1992, A Note on the Delta Method. The American Statistician 46(1): 27--29.

PRICE, G. R., 1970, Selection and covariance. Nature 227(5257): 520--521.

RICE, S., 2008, A stochastic version of the Price equation reveals the interplay of deterministic and stochastic processes in evolution. BMC Evolutionary Biology 8(1): 262+.

ROUSSET, F., 2004, Genetic Structure and Selection in Subdivided Populations. Princeton University Press.