Molecular characterization of *Klebsiella pneumoniae* isolates from stool specimens of outpatients in sentinel hospitals Beijing, China, 2010–2015

Bing Lu¹, Haijian Zhou²,³, Xin Zhang¹, Mei Qu¹, Ying Huang¹ and Quanyi Wang¹*

Abstract

**Background:** *Klebsiella pneumoniae* (*K. pneumoniae*) is an opportunistic pathogen associated with community-acquired infections and nosocomial infections. From 2010 to 2015, *K. pneumoniae* testing was included into the exiting diarrhea-syndrome surveillance with objective to estimate the prevalence of *K. pneumoniae* in diarrhea-syndrome patients, test antibiotics susceptibility and investigate molecular characteristics.

**Methods:** Stool specimens from diarrhea-syndrome outpatients were cultured and identified the pathogens by the Vitek2 Compact instrument. The isolated *K. pneumoniae* strains were tested for antibiotics susceptibility by minimal inhibitory concentration (MIC) method, and subtyped by pulsed field gel electrophoresis (PFGE) and multilocus sequence typing (MLST).

**Results:** 22 *K. pneumoniae* strains were identified from 4340 stool specimens of outpatients who visited sentinel hospitals in Beijing during 2010–2015. All strains were sensitive to gentamicin, nalidixic acid, ciprofloxacin, ceftriaxone, ceftaxime, cefepime, imipenem. The highest resistance rate of *K. pneumoniae* strains was 100% to amoxicillin–clavulanate, followed by 72.7% to ampicillin. These 22 *K. pneumoniae* strains were characterized into 21 different PFGE types and 20 MLST types with less similarity.

**Conclusions:** The detection rate of *K. pneumoniae* in stool specimens from outpatients with diarrhea syndromes was about 0.5% in Beijing. Less similarity of the isolated strains indicated the unlikely long-term circulating of *K. pneumoniae* in the community. ST23 was the most common genotype. Drug resistance of the community-acquired *K. pneumoniae* was not a serious problem in comparing with hospital-acquired infections. High vigilance in the community-acquired *K. pneumoniae* strains and investigation of pathogens’ microbiological characteristics are valuable for signals detection for drug resistance in population and strains variation.

**Keywords:** *Klebsiella pneumoniae*, Drug resistance, Multilocus sequence typing

**Background**

*Klebsiella* spp. is ubiquitous in nature. *Klebsiella pneumoniae* (*K. pneumoniae*) is an opportunistic pathogen associated with both community-acquired and nosocomial infections, causing pneumoniae, abscess, bacteremia, and urinary tract infections [1], and occasionally causes diarrhoea in humans [2, 3].

The surveillance on diarrhoea-syndrome outpatients included any age people in Beijing, involving 245 sentinel hospitals from all 16 districts. Stool specimens were collected from diarrhoea-syndrome outpatients in sentinel hospitals [4], and tested the common diarrhoea-induced pathogens including rotavirus, norovirus, diarrheagenic *Escherichia coli*, *Salmonella*, and *Shigella* spp. [5]. Because the...
increasingly nosocomial infections caused by *K. pneumoniae* might impose an increasing risk of infections in communities, from 2010 to 2015, *K. pneumoniae* testing was included into the exiting diarrhea-syndrome surveillance with objective to estimate the prevalence of *K. pneumoniae* in diarrhea-syndrome outpatients, test antibiotics susceptibility and investigate microbiological characteristics.

**Methods**

**Identification of bacterial strains**

Stool specimens from diarrhea-syndrome outpatients were collected through Beijing diarrhea-syndrome surveillance from 2010 to 2015. Diarrheagenic bacteria strains were cultured and isolated from stool specimens firstly. Any isolated positive bacteria strains were further identified the pathogens (e.g., *Salmonella*, *Shigella* spp., diarrheagenic *Escherichia coli*, *Vibrio parahemolyticus*, and *K. pneumoniae*) by the Vitek2 Compact instrument (bioMérieux, Marcy, France). The isolated *K. pneumoniae* strains were proceeding for antibiotics susceptibility test and molecular characterization.

**Antibiotics susceptibility testing**

Antibiotics susceptibility testing for the *K. pneumoniae* strains was measured by minimal inhibitory concentration (MIC) method. The MICs were interpreted by the standards of Clinical and Laboratory Standards Institute (CLSI) document M100-S26:2016 [6]. The following 14 antibiotics sourced from Shanghai Xingbai Co. (AST panel for aerobic gram negative bacilli) were used for antimicrobial susceptibility test: ciprofloxacin, ampicillin, amoxicillin–clavulanate, chloramphenicol, sulfisoxazole, trimethoprim–sulfamethoxazole, nalidixic acid, ceftriaxone, gentamicin, tetracycline, cefotaxime, cefoxitin, cefepime, imipenem. *Escherichia coli* strain ATCC 25922 was in use as quality-control strains for the antibiotics susceptibility testing. MIC level at or above 2 μg/mL for cefotaxime and ceftriaxone indicates the strains possibly produced extended-spectrum beta-lactamases (ESBL), which needs further confirmation. MIC for ceftazidime combination with clavulanate decrease at least three twofold concentration in contrast with the MIC value for ceftazidime alone (e.g., ceftazidime MIC = 8 μg/mL; ceftazidime–clavulanate MIC = 1 μg/mL) confirms the existing ESBL production strains [6].

**Pulse field gel electrophoresis**

*K. pneumoniae* strains were subtyped by pulse field gel electrophoresis (PFGE) method using restriction endonucleases *XbaI* [7]. The restriction endonuclease *XbaI* (Fermentas) was used to digest the prepared genomic DNA and resultant DNA fragments were separated in a PFGE CHEF-DR III system (Bio-Rad Laboratories) in 0.5× Tris–borate–EDTA buffer at 120 V for 18.5 h, with pulse times ranging from 6 to 36 s. PFGE dendrogram was generated by BioNumerics version 7.1 (Applied Math, Belgium) with the unweighted pair-group method (UPGMA) and Dice coefficient. Isolates that exhibited a PFGE profile with more than 80% similarity were considered as closely related strains [8, 9].

Multilocus sequence typing

Multilocus sequence typing (MLST) was performed to subtype *K. pneumoniae* strains using seven housekeeping genes (gapA, infB, mdh, pgI, phoE, rpoB, and tonB) [10]. Seven gene fragments of the *K. pneumoniae* strains were amplified by PCR and sequenced. The allele sequences and sequence types (STs) were determined by the Institute Pasteur Klebsiella MLST database (http://bigsdb.web.pasteur.fr/klebsiella/klebsiella.html). BioNumerics version 7.1 software was used to create the minimum spanning tree. In the minimum spanning tree, the founder ST was defined as the greatest number of single-locus variants. Types were represented by circles and the size of a circle indicated the number of strains with this particular type.

**Results**

**Identification of bacterial strains**

In the 4340 stool specimens collected in the surveillance period from 2010 to 2015, 22 *K. pneumoniae* strains were identified: one strain in 2010, three strains in 2011 and 2012 respectively, five strains in 2013, nine strains in 2014, and one strain in 2015; 16 out of 22 positive *K. pneumoniae* were isolated from April to October, while six positives identified in low-incidence seasons 2010–2015.

**Antibiotics susceptibility results**

Antibiotics susceptibility is shown in Table 1. The result showed that 22 *K. pneumoniae* strains were all sensitive

| Antibiotics agent | R (%) | I (%) | S (%) |
|-------------------|-------|-------|-------|
| Ampicillin        | 16 (72.7%) | 6 (27.3%) | 0     |
| Amoxicillin–clavulanate | 22 (100%) | 0     | 0     |
| Ceftriaxone       | 0     | 0     | 22 (100%) |
| Cefotaxime        | 0     | 0     | 22 (100%) |
| Cefepime          | 0     | 0     | 22 (100%) |
| Ceftoxitin        | 1 (4.5%) | 0     | 21 (95.5%) |
| Imipenem          | 0     | 0     | 22 (100%) |
| Tetracycline      | 2 (9.1%) | 7 (31.8%) | 13 (59.1%) |
| Nalidixic acid    | 0     | 0     | 22 (100%) |
| Ciprofloxacin     | 0     | 0     | 22 (100%) |
| Trimethoprim–sulfamethoxazole | 1 (4.5%) | 0 | 21 (95.5%) |
| Sulfisoxazole     | 8 (39.3%) | 0     | 14 (63.6%) |
| Chloramphenicol   | 4 (18.2%) | 7 (31.8%) | 11 (50%) |
| Gentamicin        | 0     | 0     | 22 (100%) |

*R* resistant, *I* intermediate, *S* susceptible
to gentamicin, nalidixic acid, ciprofloxacin, ceftriaxone, ceftaxime, cefepime, imipenem, followed by 95.5% to trimethoprim–sulfamethoxazole, cefoxitin, and 63.6% to sulfisoxazole. The highest resistance rate of *K. pneumoniae* strains was 100% to amoxicillin–clavulanate, and followed by 72.7% to ampicillin. Moreover, all 22 *K. pneumoniae* strains were sensitive to ceftriaxone, ceftaxime, the MIC was less than 1 μg/mL, and no ESBL production strain was identified.

**PFGE typing**

22 isolates showed 21 different PFGE types. The dendrogram of the PFGE image showed that four *K. pneumoniae* strains isolated during 2011–2014 had over 80% similarity, which categorized into one PFGE cluster (cluster A), while other strains had less similarity (Fig. 1).

**MLST typing**

MLST was performed for all 22 isolates to investigate the genetic relationships. MLST analysis revealed 20 different sequence types (STs), including three ST23 strains and one ST65. In addition to identifying ST23 and ST65 which are prevalent in Asia countries, ST2362, ST2363, ST2364, ST2365, ST2366, ST2367, ST2368, ST2369, and ST2370 were discerned for the first time. In the minimum spanning tree, there were two clone groups: one clone group contained ST23 and ST1660, and the other clone group contained ST17 and ST20 (Fig. 2).

**Discussion**

*K. pneumoniae* is one of the most common Gram-negative pathogen found in human's nasopharynx and in the intestinal tract [1]. It existed with 60–70% the carrier rate
in the hospital environment, and was notified as a common pathogen caused nosocomial infection [1]. However, the community-acquired \textit{K. pneumoniae} infection was rare reported. This study, through inclusion of \textit{K. pneumoniae} into the existing diarrhea-syndrome surveillance, was able to detect \textit{K. pneumoniae} infection in community and its risk for possible persistent transmission in population in Beijing.

The detection rate of \textit{K. pneumoniae} in stool specimens from outpatients with diarrhea syndromes was about 0.5% (22/4340), which further demonstrated the existence of the community-acquired \textit{K. pneumoniae} infection [11]. The prevalence in this study was much lower than the previous detection rate of \textit{K. pneumoniae} in stool samples ranges from 5 to 38% in hospital patients [11], which emphasized the importance of nosocomial infection control as well necessary vigilance for detecting community-acquired \textit{K. pneumoniae}.

Less similarity of the strains typed by PFGE or MLST indicated the unlikely long-term transmission of \textit{K. pneumoniae} in the community. ST23 was the most common genotype. Drug resistance of the community-acquired \textit{K. pneumoniae} was not a serious problem in comparing with hospital-acquired infections. High vigilance in the drug resistance of the community-acquired \textit{K. pneumoniae} strains can provide a significant signal of drug resistance in population.

**Fig. 2** Minimum spanning tree of MLST typing for 22 \textit{K. pneumoniae} strains

\textbf{Abbreviations}

CLSI: Clinical and Laboratory Standards Institute; ESBL: extended-spectrum beta-lactamases; \textit{K. pneumoniae}: \textit{Klebsiella pneumoniae}; MIC: minimum inhibitory concentration; MLST: multilocus sequence typing; PFGE: pulsed field gel electrophoresis; STs: sequence types; UPGMA: the unweighted pair-group method.
Authors' contributions
BL participated in molecular genetic studies and drafted the manuscript; XZ and YH participated in sample collection and antimicrobial susceptibility testing; HZ carried out the data analysis; MQ and QW participated in the design of the study. All authors read and approved the final manuscript.

Author details
1 Institute for Infectious Disease and Endemic Disease Control, Beijing Center for Disease Prevention and Control, Beijing Center for Prevention Medical Research, Beijing Key Laboratory of Diagnostic and Traceability Technologies for Food Poisoning, Beijing 100013, China. 2 State Key Laboratory for Infection Disease Prevention and Control, National Institute for Communicable Disease Prevention and Control, Chinese Center for Disease Control and Prevention, Beijing 102206, China. 3 Collaborative Innovation Center for Diagnosis and Treatment of Infectious Diseases, Hangzhou, China.

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Competing interests
The authors declare that they have no competing interests.

Availability of data and materials
The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Consent for publication
This manuscript did not include any details, images, or videos relating to individual participants.

Ethics approval and consent to participate
The study was approved by the Ethics Committee of the Beijing Center for Disease Control and Prevention. All patients in the study signed informed consent forms.

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