A prospective study validating a clinical scoring system and demonstrating phenotypical-genotypical correlations in Silver-Russell syndrome

Salah Azzi,1,2,3,4 Jennifer Salem,5 Nathalie Thibaud,1,2,3 Sandra Chantot-Bastaraud,6 Eli Lieber,7 Irène Netchine,1,2,3 Madeleine D Harbison8

ABSTRACT

Background Multiple clinical scoring systems have been proposed for Silver-Russell syndrome (SRS). Here we aimed to test a clinical scoring system for SRS and to analyse the correlation between (epi)genotype and phenotype.

Subjects and methods Sixty-nine patients were examined by two physicians. Clinical scores were generated for all patients, with a new, six-item scoring system: (1) small for gestational age, birth length and/or weight ≤−2SDS, (2) postnatal growth retardation (height ≤−2SDS), (3) relative macrocephaly at birth, (4) body asymmetry, (5) feeding difficulties and/or body mass index (BMI) ≤−2SDS in toddlers; (6) protruding forehead at the age of 1–3 years. Subjects were considered to have likely SRS if they met at least four of these six criteria. Molecular investigations were performed blind to the clinical data.

Results The 69 patients were classified into two groups (Likely-SRS (n=60), Unlikely-SRS (n=9)). Forty-six Likely-SRS patients (76.7%) displayed either 11p15 ICR1 hypomethylation (n=35; 58.3%) or maternal UPD of chromosome 7 (mUPD7) (n=11; 18.3%). Eight Unlikely-SRS patients had neither ICR1 hypomethylation nor mUPD7, whereas one patient had mUPD7. The clinical score and molecular results yielded four groups that differed significantly overall and for individual scoring system factors. Further molecular screening led identifying chromosomal abnormalities in Likely-SRS—double-negative and Unlikely-SRS groups. Four Likely-SRS—double-negative patients carried a DLK1/GTL2 IG-DMR hypomethylation, a mUPD16; a mUPD20 and a de novo 1q21 microdeletion.

Conclusions This new scoring system is very sensitive (98%) for the detection of patients with SRS with demonstrated molecular abnormalities. Given its clinical and molecular heterogeneity, SRS could be considered as a spectrum.

INTRODUCTION

Silver-Russell syndrome (SRS, OMIM #180860; called also Russell-Silver syndrome, RSS, in the USA) is a clinically and genetically heterogeneous syndrome involving prenatal and postnatal growth retardation first described by Silver et al4 and Russell.5 Many other studies have since reported additional features, providing a complex clinical description of SRS.3,5 The clinical presentation of SRS is now known to cover a spectrum of signs that are easy to recognise in typical cases but may be difficult to diagnose clinically in less severely affected individuals. Furthermore, the facial characteristics of SRS tend to become attenuated as the patient grows up, making it difficult to diagnose SRS in older children and adults. Finally, some of the more common typical features of SRS overlap with those of other syndromic intrauterine growth retardation disorders (such as 3M or Mulholland syndromes, eg7).

There is growing evidence that a definition of SRS based on a compilation of features rather than a checklist of characteristics is required. Unfortunately, there is still no clear consensus about how SRS should be defined. This renders clinical diagnosis difficult, leading to underdiagnosis in some situations and overdiagnosis in others. Five attempts have been made to create a clinical definition of SRS, or an ‘SRS scoring system’.8,9,10–12 The various systems developed display some similarities, but also differences. In the paper describing the most recent of these systems, the Birmingham model,11 the four scoring systems that had previously been developed were evaluated, but on retrospective clinical data collected by different physicians, which could not, therefore, necessarily be considered reliable.

Despite the existence of these numerous clinical scoring systems, the identification of appropriate patients for molecular testing remains a challenge, because many of the features of SRS are non-specific or mild, and some may disappear over time.

Two primary molecular causes of SRS have been identified. In about 10% of cases, SRS is due to maternal UPD of chromosome 7 (mUPD7). The major abnormality, present in 50–60% of cases, was only recently identified: hypomethylation of the paternal allele of the 11p15 imprinting centre region 1 (ICR1) regulating the IGF2/H19 locus.9,10,12,13,14 Rare genetic or cytogenetic abnormalities have also been identified, but these abnormalities account for less than 2% of cases.14 The molecular cause of SRS thus remains unknown in about 30–40% of cases.

On the basis of our considerable clinical experience with patients with SRS, we conducted a prospective study in which 69 patients with suspected SRS were assessed clinically and then underwent state-of-the-art molecular investigations. The results obtained were then used to validate a modified scoring system adapted from the original scoring...
system developed by Netchine et al in which small for gestational age (SGA) was no longer an obligate factor. We have shown that this new scoring system was highly sensitive for identification of the subjects most likely to test positive for one of the known molecular causes of SRS, and for distinguishing these subjects from those not likely to test positive. This new scoring system is easy to use and flexible enough to be run even if data are missing for one or more factors. The combination of variables in this scoring system may be considered an improvement over those previously published.

METHODS

Study population

The population consisted of 69 patients (37 boys and 32 girls; age range 1.05–20.06 years with a mean age of 6.61 years) clinically diagnosed as SGA or possibly SRS by a local physician. Sixty-three were Caucasians, two were of Asian origin, one was an Afro-American and three were of mixed origin. The patients received information about the study from the MAGIC Foundation, a patient support group for patients with growth disorders with separate divisions for SGA patients and patients with SRS. All the patients attending either the SGA or the SRS division at the MAGIC convention in July 2008 were invited to participate in the study. Each patient was examined by the same two paediatric endocrinologists (IN and MDH) with substantial experience in the field of SRS. Physical characteristics from an extensive list were recorded as present or absent. Photographs of each patient were taken at this time. For patients over the age of 3 years, we also requested additional photos of the child at an age of 1–3 years (face and profile). The parents were also asked to complete a multipage survey and growth records were obtained. Fifteen patients of the 69 have been reported in Azzi et al whereas the remaining patients are reported for the first time.

Molecular investigations

Briefly, after DNA extraction and sodium bisulfite treatment, we used TaqMan Allele-Specific Methylated Multiplex Real-Time Quantitative PCR (ASMM RTQ-PCR) to analyse the methylation status at 11p15 ICR1 CBS2, 11p15 DMR (H19 promoter), 11p15 ICR2, ZAC1 differentially methylated region (DMR), the GNAS locus SNRPN and DLK1/GTL2 IG-DMR locus, as previously described. We ruled out a maternal duplication involving the 11p15 centromeric domain by studying the ICR2 methylation status and mutations at the DLK1/GTL2 locus SNRPN and ZAC1 locus, as previously described. For more details, see online supplementary data.

SNP microarray: Chromosomal abnormalities were screened using CytoSNP12 microarray (Illumina, San Diego, California). For more details, see online supplementary data.

Clinical score

Our new scoring system, the Netchine-Harbison clinical scoring system (NH-CSS), is based on the original system developed by Netchine et al and no longer includes a requirement for SGA (see online supplementary table S1 and scoring system sheet). Six factors are included in the NH-CSS: (1) prenatal growth retardation (birth weight and/or length ≤−2 standard deviation score (SDS) for gestational age); (2) postnatal growth retardation (height at 24 ±1 months ≤−2 SDS according to the Centers for Disease Control and Prevention (CDC) growth charts (http://www.cdc.gov/growthcharts/); (3) relative macrocephaly at birth (head circumference at birth ≥1.5 SDS above birth weight and/or length SDS according to Usher and McLean statistical data); (4) protruding forehead (defined as a forehead that projects beyond the face on a side view as depicted on figure 1) as a toddler; (5) body asymmetry (defined as a leg length discrepancy (LLD) of ≥0.5 cm or arm asymmetry or LLD <0.5 cm with at least two other asymmetrical body parts (with one being a non-face part)); (6) feeding difficulties (use of a feeding tube or cyproheptadine for appetite stimulation) and/or low BMI (BMI ≤−2 SDS at 24 months) according to CDC growth charts (http://www.cdc.gov/growthcharts/). Midparental target height was calculated as follows: (father’s height+mother’s height)/2+6.5 cm for boys and −6.5 cm for girls.

A patient was classified as having ‘Likely-SRS’ if at least four of these six factors were present. A patient was assigned to the ‘Unlikely-SRS’ group if three or fewer of these factors were present. We demonstrated the reliability of the NH-CSS by also applying the Birmingham11 and Netchine et al scoring systems to our studied population, and comparing the results obtained with the three clinical scoring systems. Sensitivity and specificity were assessed for the identification of a recognised molecular cause of SRS (ie, 11p15 ICR1 hypomethylation or mUPD7). The details of each of the three clinical scoring systems are summarised in online supplementary table S1.

Statistical analysis

See online supplementary data.

RESULTS

Netchine-Harbison clinical scoring system results

The 69 study participants were examined by two paediatric endocrinologists with considerable experience in the diagnosis and care of patients with SRS. These physicians together agreed upon a series of phenotypical characteristics for each patient. The presence or absence of a protruding forehead was determined during the initial examination. For patients over the age of 3 years, the two physicians also evaluated this facial characteristic together, on photos of the child aged 1–3 years (face and profile). This method made it possible to collect homogeneous phenotypical data confirmed by two examiners. Following the review of the photos, each patient was then scored for the new NH-CSS (see online supplementary table S1). Sixty of the 69 patients (35 boys and 23 girls; mean age 6.85 years) included in the study satisfied at least four of the criteria (mean factor frequency=5.29) of the NH-CSS and were consequently classified as ‘Likely-SRS’. Nine patients (two boys and seven girls; mean age 4.96 years) were considered as satisfying up to three of these criteria (mean factor frequency=2.89) and were classified as ‘Unlikely-SRS’.

We then compared these two groups (Likely-SRS and Unlikely-SRS). We initially studied only those patients for whom we had data for all six criteria (n=61). We compared 52 patients from the Likely-SRS group with the 9 patients from the Unlikely-SRS group. As expected, these two groups displayed statistically significant clinical differences (t test; p=0.000). We also compared the two groups for each NH-CSS criteria separately. The two groups did not differ significantly in terms of birth weight or length or postnatal growth failure. By contrast, they differed significantly for the other four criteria of the NH-CSS (Pearson’s χ²; p=0.000) (table 1).

Molecular analysis

11p15 methylation and mUPD7 screening

We assessed 11p15 ICR1 region methylation status and mUPD7 in the 69 patients. We detected 11p15 ICR1 and mUPD7

Azzi S, el al. J Med Genet 2015;52:446–453. doi:10.1136/jmedgenet-2014-102979
abnormalities by carrying out ASMM RTQ-PCR to determine the methylation status of ICR1 CBS2 and H19 DMR (H19 promoter) for 11p15 ICR1 and of PEG/MEST in the 7p32 region for mUPD7. We identified molecular abnormalities in 46 of the 60 (76.7%) Likely-SRS patients (35 (58.3%) (22 boys and 13 girls; mean age 6.9 years) due to 11p15 ICR1 hypomethylation and 11 (18.3%) (7 boys and 5 girls; mean age 7.69 years) due to mUPD7). Fourteen of the 60 Likely-SRS patients (23.3%) (6 boys and 8 girls; mean age 5.79 years) were negative for both abnormalities and are described hereafter as Likely-SRS double-negatives (L-SRS-dblneg). No molecular abnormality of either the 11p15 ICR1 region or mUPD7 was identified in eight of the nine (88.9%) patients in the Unlikely-SRS group (UL-SRS-dblneg). However, one patient who was positive for three criteria of the NH-CSS was found to have mUPD7.

Overall, NH-CSS successfully picked up 97.9% of the patients with known molecular abnormalities associated with SRS (100% of the 35 11p15 ICR1 patients and 91.7% of the mUPD7 patients), missing only a single mUPD7 patient.

Multilocus imprinting disturbance in patients with SRS with ICR1 11p15 hypomethylation
Multilocus imprinting disturbance (MLID) is increasingly being reported in human imprinting diseases, including SRS. We therefore screened for MLID in this well clinically characterised cohort of patients with SRS. We investigated the methylation status of 11p15 ICR2, the DLK1/GTL2 IG-DMR locus on

| Table 1 | Classification of the SRS population according to the NH-CSS and statistical comparison of the two clinical groups |
|---------|------------------------------------------------------------------------------------------------------------|
| Mean of factors recorded ‘yes’ for each group (subjects with missing data excluded)*† | Likely-SRS (≥4 factors present) | Unlikely-SRS (≤3 factors present) | p Value |
| Mean or Freq. | Mean or Freq. | | |
| Mean of factors recorded ‘yes’ for each group (subjects with missing data excluded)*† | 5.29 (n=52) | 2.89 (n=9) | 0.000 |
| SGA (birth weight and/or birth length‡§) | 55 of 60 (91.67%) | 7 of 9 (77.78%) | NS |
| Postnatal growth failure‡§ | 55 of 58 (94.83%) | 7 of 9 (77.78%) | 0.07 (NS) |
| Relative macrocephaly at birth‡§ | 46 of 56 (82.14%) | 2 of 9 (22.22%) | 0.000 |
| Protruding forehead‡§ | 55 of 58 (94.83%) | 5 of 9 (55.56%) | 0.000 |
| Body asymmetry‡§ | 44 of 60 (73.33%) | 1 of 9 (11.11%) | 0.000 |
| Feeding difficulties and/or BMI ≤−2SDS | 58 of 59 (98.31%) | 3 of 9 (33.33%) | 0.000 |

*† Test, equal variances not assumed; t=15.279; df 27.619; Sig (2-tailed) 0.000.
†Only subjects with data for all 6 factors were included in this ‘overall’ top-level analysis, to ensure that group mean factor numbers were comparable.
†Pearson’s χ² test.
§The Ns for each scoring system factor can be less than the total of 60, because some subjects were missing data for one or more factors (but still had enough data to qualify for ‘4 or more factors recorded as yes’).
NH-CSS, Netchine-Harbison clinical scoring system; NS, not significant; SGA, small for gestational age; SRS, Silver-Russell syndrome.

448 Azzi S, et al. J Med Genet 2015;52:446–453. doi:10.1136/jmedgenet-2014-102979

Figure 1  Representative pictures of children with a high forehead not protruding (A) or with protruding foreheads of various degrees, mild (B), moderate (C) or marked (D).
chromosome 14q32, ZAC1 DMR on 6q24, SNRPN on 15q11.2 and the GNAS locus (XLa DMR, AB DMR and NESP55 DMR) on 20q13.32, by ASMM RTQ-PCR. Four of 35 patients (11.4%) were found to display MLID: one with hypomethylation at ZAC1 DMR (MI=36%; range of normal values 45–57%); one with a methylation profile similar to that found in hypoparathyroidism type 1b (PHP1b) patients (ie, hypomethylation at XLa DMR (MI=18%) and AB DMR (MI=21%) and hypermethylation at NESP55 DMR (MI=73%)) and two patients displayed hypomethylation of the DLK1/GTL2 IG-DMR locus (MIs=17% and 8%).

Further findings
For all patients negative for the conventional molecular aetiologies of SRS,21 22 we searched for 11p15 ICR2 duplication, CDKN1C mutation or copy number variation (CNV) or imprinting abnormalities in other imprinted regions that might account for the SRS phenotype in the L-SRS-dblneg group or the growth retardation observed in the UL-SRS-dblneg group. We carried out genome-wide screening of DNA with the CytoSNP12 microarray from all the patients of the L-SRS-dblneg and UL-SRS-dblneg groups. Methylation abnormalities in imprinted regions were assessed by ASMM RTQ-PCR. We identified four independent molecular defects in four patients from the L-SRS-dblneg group. Two patients displayed maternal UPD, one of chromosome 16 (mUPD16) and the other of chromosome 20 (mUPD20). A third patient carried a de novo 1q21 3 Mb microdeletion (figure 2). The fourth patient displayed a loss of methylation at the DLK1/GTL2 IG-DMR locus; and the CytoSNP12 microarray excluded mUPD14, microdeletion or microduplication involving the 14q32 domain.

In the UL-SRS-dblneg group, we identified three independent chromosomal rearrangements in three patients: one patient carried a de novo 20.2 Mb 1q24.3q31.2 microdeletion, the second carried a de novo 5.2 Mb 11p13-p12 microdeletion centromeric to the WT1 gene and the third carried a paternally transmitted 2.6 Mb 22q11.21 microduplication.

The clinical features of these four L-SRS-dblneg and three UL-SRS-dblneg patients plus the UL-SRS mUPD7 patient are summarised in online supplementary table S7 and supplementary data.

Comparison of NH-CSS criteria between the different molecular groups
We reclassified the Likely-SRS and Unlikely-SRS patients according to the molecular data which results in four groups (11p15 hypomethylation, mUPD7, L-SRS-dblneg and Unlikely-SRS) and then investigated the differences between them in terms of NH-CSS factors. We excluded the mUPD7 false-negative patient from this analysis for the sake of clarity. However, it should be noted that its exclusion had no effect on the results of the statistical analysis (data not shown). Overall scores differed significantly between the four groups (table 2; p=0.000). By including only subjects for whom data were available for all six factors, we ensured that comparisons between groups in terms of the overall mean number of NH-CSS positive criteria were carried out correctly. We then compared scoring system factors separately across the four groups. Statistically significant differences between the four groups were found for all the criteria except for one of these factors, postnatal growth failure ($\chi^2$ test; p=0.06).

Pairwise comparison analyses showed that the 11p15 ICR1 hypomethylated group was statistically different overall and for all six factors considered separately. The Unlikely-SRS group also differed clinically from the other groups (see online supplementary table S2). Relative macrocephaly was present in a significantly higher percentage of individuals from the SRS 11p15 and SRS mUPD7 groups than in the UL-SRS group (see online supplementary table S3). The percentage of subjects with a protruding forehead was also significantly higher in the 11p15 and SRS mUPD7 groups than in the UL-SRS group (see online supplementary table S4). The percentage of subjects with body asymmetry was also significantly higher in the 11p15 group than in all the other groups but this percentage was significantly higher in the L-SRS-dblneg than in the UL-SRS group (see online supplementary table S5). Finally, feeding difficulties and

Figure 2 Chromosomal abnormalities found in Likely-Silver-Russell syndrome (SRS) double-negative patients (left panel) and Unlikely-SRS double-negative patients (right panel).
low BMI were significantly more frequent in the SRS 11p15, SRS mUPD7 and L-SRS-dblneg groups than in the UL-SRS group (see online supplementary table S6).

The NH-CSS failed to identify only one patient for whom subsequent molecular analysis demonstrated SRS mUPD7. Therefore, we sought to identify additional characteristics useful for the clinical diagnosis of SRS for borderline cases with the NH-CSS score of 3–4. We compared the four molecular groups for a number of cognitive and physical variables previously reported or considered by the investigators (IN and MDH) to be common in children with SRS (see online supplementary table S8). The four groups differed significantly in terms of a number of quantifiable physical characteristics; however, incidence calculations showed that the significance of these characteristics was often limited to specific groups (see online supplementary data).

Comparison of the clinical scoring systems for their sensitivity and specificity

We compared our NH-CSS with the scoring system previously reported by Netchine et al10 and the Birmingham clinical scoring system.11 We applied these three clinical scoring systems to the 69 patients studied here. We considered only the two common molecular aetiologies of SRS, mUPD7 and 11p15 ICR1 hypomethylation, for SRS-positive molecular testing. Our system classified 60 patients as Likely-SRS (table 3), whereas the Netchine et al10 and Birmingham systems classified only 55 and 47 patients, respectively, as Likely-SRS. The number of Unlikely-SRS patients was therefore higher with these two systems, 14 and 16 patients, respectively than with our new system, 9 patients.

Interestingly, whereas our new scoring system and that of Netchine et al10 identified 100% of the patients with 11p15 ICR1 hypomethylation, the Birmingham scoring system missed two of the patients (93.7% of identification). In addition, our new scoring system identified all but one of the mUPD7 patients (91.7% identified), whereas the Netchine et al10 system missed four mUPD7 (66.7% identified) and the Birmingham system missed five mUPD7 (54.6% identified). Overall, the number of false-negative results was reduced in the new scoring system. The Birmingham system uses only four factors making it more prone to the negative effect of missing data points. This resulted in the exclusion of six additional patients due to missing data including three 11p15 ICR1 hypomethylation and one mUPD7 patient. Overall, the Birmingham system failed to identify 11 subjects with molecularly confirmed SRS from our study population.

The new NH-CSS was more sensitive (97.9%) than the Netchine et al10 (91.5%) and Birmingham (83.7%) clinical scoring systems (table 4). By contrast, our new NH-CSS is less specific (specificity=36%) than the other systems. The three tests had similar positive predictive values, but the NH-CSS had the highest negative predictive value (NPV 88.9%). Thus, we can have a high degree of confidence that a score of less than four on the NH-CSS screen has a high likelihood of being truly SRS-negative. The Birmingham scoring system had the lowest NPV of the three clinical scoring systems tested (56.3%), due to the number of patients with positive molecular findings who would not have been tested if the decision to test were based on this scoring system.

DISCUSSION

SRS is a clinically and genetically heterogeneous condition. SRS has been extensively studied, but there is still no consensus on the clinical definition of this disorder. Consequently, large numbers of patients not actually meeting typical SRS scoring system criteria, regardless of the scoring system used, are referred to diagnostic laboratories for genetic testing, simply because they may have some of the extensive long list of typical SRS features. This approach is well documented by the low percentage of positive molecular results for both known molecular defects of SRS in molecular diagnostic laboratories which leads to an increase in healthcare costs.23

Several genotype/phenotype correlation studies have shown that not all patients with SRS are born SGA, and this is particularly true for those with mUPD7.24 25 Based on these observations and on our own experience, we revised the Netchine et al10 scoring system and developed a new six-factor NH-CSS. Patients classified as Likely-SRS or Unlikely-SRS with the NH-CSS differed significantly for four of the factors of the scoring system. Furthermore, our new scoring system was found to be highly sensitive and had a strong NPV. Although these values of the NH-CSS might be different in a more heterogeneous population recruited in a growth clinic for short stature, we still could compare for our cohort the sensitivity and the NPV of the NH-CSS with those obtained using other scoring systems previously described. We therefore also applied the Birmingham clinical scoring system11 to our cohort and

Table 2  Statistical comparison between the four molecular groups, overall and individually, for the six factors of the NH-CSS

| 11p15 ICR1 hypomethylation | mUPD7* | L-SRS-dblneg | UL-SRS* | p Value |
|-----------------------------|--------|--------------|---------|---------|
| Mean number of factors positivet | 5.86 (n=29) | 4.73 (n=11) | 4.42 (n=12) | 2.88 (n=8) | 0.000 |
| SGA (birth weight and/or length)§ | 35 of 35 (100%) | 8 of 11 (72.7%) | 12 of 14 (85.7%) | 7 of 8 (87.5%) | 0.034 |
| Postnatal growth failure§ | 34 of 34 (100%) | 10 of 11 (90.9%) | 11 of 13 (84.6%) | 6 of 8 (75%) | 0.060 |
| Relative macropcephaly at birth§ | 31 of 32 (96.9%) | 9 of 11 (81.8%) | 6 of 13 (46.2%) | 2 of 8 (25%) | 0.000 |
| Protruding forehead§ | 32 of 33 (97.0%) | 11 of 11 (100%) | 12 of 14 (85.7%) | 4 of 8 (50%) | 0.001 |
| Body asymmetry§ | 33 of 35 (94.3%) | 3 of 11 (27.3%) | 8 of 14 (57.1%) | 1 of 8 (12.5%) | 0.000 |
| Feeding difficulties and/or BMI <-2SDS§ | 35 of 35 (100%) | 11 of 11 (100%) | 12 of 13 (92.3%) | 3 of 8 (37.5%) | 0.000 |

†One-way ANOVA.
§2 test.
*The mUPD7 Unlikely SRS’ subject was removed from this analysis. Even if this subject was included in the ‘Unlikely SRS’ group, the significance level of the overall system remained at 0.000 and the level of significance of the factors remained within their overall significance level categories (<0.001, 0.05 or 0.1).
‡Only subjects with data for all 6 factors were included in this ‘overall’ top-level analysis, to ensure that group mean factor numbers were comparable.

ANOVAs, analysis of variance; L-SRS-dblneg, Likely SRS double-negative; mUPD, maternal UPD of chromosome 7; NH-CSS, Netchine-Harbison clinical scoring system; SGA, small for gestational age; SRS, Silver-Russell syndrome; UL-SRS, Unlikely-SRS.
Table 3  Assessment of the performance of the three clinical scoring systems on our cohort of 69 patients

| Scoring System | Likely SRS (%) | Unlikely SRS (%) |
|---------------|---------------|-----------------|
| 11p15 ICR1 hypomethylation or mUPD7 | 83.7 | 6.3 |
| Netchine-Harbison clinical scoring system | 75.0 | 25.0 |
| Birmingham clinical scoring system | 67.2 | 32.8 |

Table 4  Sensitivity and specificity of the three clinical scoring systems calculated for our cohort of patients

| System            | NH CSS (%) | Netchine-Harbison (%) | Birmingham (%) |
|-------------------|------------|-----------------------|----------------|
| Sensitivity        |            | N/A                   | 70.0           |
| Specificity        |            | N/A                   | 81.0           |
| Sensitivity        |            | 97.9                  | 91.5           |
| Specificity        |            | 36.4                  | 45.5           |
| Positive predictive value | 76.7 | 78.2 | 76.6 |
| Negative predictive value | 88.9 | 71.4 | 56.3 |

The 'Likely-SRS' patients with molecular causes other than mUPD7 or 11p15 ICR1 hypomethylation are considered as false positives for the purposes of this assessment.
molecular diagnosis is detailed in figure 3. However, the absence of a positive molecular test result should not rule out the clinical diagnosis of SRS in the patient concerned. This new scoring system missed one patient with genuine SRS who fulfilled only three of the NH-CSS criteria. The heterogeneous clinical presentation of SRS makes it very difficult to diagnose those patients with a mild SRS phenotype, particularly in some mUPD7 cases. Consequently, we looked for additional quantitative variables and typical physical SRS characteristics but did not find any that improve the scoring system but only helped to distinguish mUPD7 and 11p15 hypomethylation groups.

A precise and simple clinical definition of SRS is important in order to establish a prevalence of this rare condition, propose common clinical guidelines and possible common clinical trials for this group of patients and allow the research to progress for the patients with no molecular cause identified. Sixty years after the initial description of SRS, 1,2 two molecular defects are considered to be common. In the future potentially new molecular aetiologies may be added to this list because they can be identified in patients diagnosed with SRS on our simple clinical scoring system who share enough similar clinical presentations so that they benefit from the same clinical management. This implies that SRS should remain a clinical diagnosis, based on the NH-CSS criteria. Because phenotypical differences exist between the groups of patients with SRS with different molecular aetiologies, and yet the overall clinical SRS group has been found to be statistically different from those unlikely to have SRS, we suggest that SRS should be considered as a spectrum based on positive NH-CSS screening. Similar clinical guidelines for treatment are appropriate for idiopathic SRS, once the differential diagnoses that question the efficacy of growth hormone (GH) therapy (ie, 3M syndrome, 37 IGF1R mutation 38 and some that question the safety or preclude GH therapy (ie, Bloom, 39 Mulibrey-nanism 40 and Fanconi 40 syndromes) have been ruled out.

Author affiliations
1INSERM, UMR_S 938, CDR Saint-Antoine, Paris, France
2Sorbonne Universités, UPMC Univ Paris 06, UMR_S 938, CDR Saint-Antoine, Paris, France
3Department of Pediatric Endocrinology, APHP, Armand Trousseau Hospital, Paris, France
4Epigenetics Programme, The Babraham Institute, Cambridge, UK
5MAGIC Foundation, RSS/SGA Research & Education Fund, Oak Park, Illinois, USA
6AP-HP, Hôpital Trousseau, Service de Génétique et d’Embryologie Médicales, Paris, France
7Department of Psychiatry and Biobehavioral Sciences, Semel Institute, University of California, Los Angeles, California, USA
8Department of Pediatrics, Ichan School of Medicine at Mount Sinai, New York, New York, USA
9Acknowledgements The authors thank the patients, their families and the physicians for referring patients; the MAGIC Foundation and Dayna Carney for their help with the patients recruited in the study; Frédéric Brioude, Sylvie Rossignol and Yves Le Bouc for helpful discussions; Boris Keren for the SNP arrays analysis and Cristina Das Neves, Marilyne Le Jule and Virginie Steunou for technical assistance.

Contributors SA participated in data analysis, performed the multilocus methylation study and wrote the paper. JS set up the clinical database and managed it, contributed to the statistical analysis and reviewed the paper. EL performed the statistical analysis. NT performed the molecular diagnosis: 11p15 region methylation and mUPD7. SC-B performed the SNP microarray study and reviewed the paper. IN and MDH designed the study, performed the clinical and phenotypical investigations on the entire cohort, analysed the data and reviewed the paper. IN validated the molecular studies.

Funding This work was supported by INSERM, UPMC-Paris6 funding, ANR EPIFEGR0 2010, a Pfizer grant, an Agence de Biomedecine 2010 grant, the MAGIC Foundation, the People Programme (Marie Curie Actions) of the European Union’s Seventh Framework Programme FP7/2007-2013/ under REA grant agreement n° 290123. SA was supported by Novonordisk and the INSERM-ANR EPIFEGR0 2010. IN is member of the COST Action BM1208.

Competing interests None declared.

Patient consent Obtained.

Ethics approval IRB 100000204 at the Mount Sinai School of Medicine, NYC, USA.

Provenance and peer review Not commissioned; externally peer reviewed.

Open Access This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

REFERENCES
1 Silver HK, Kiyasu W, George J, Deamer WC. Syndrome of congenital hemihypertrophy, shortness of stature, and elevated urinary gonadotropins. Pediatrics 1953;12:368–76.
2 Russell A. A syndrome of intra-uterine dwarfism recognizable at birth with cranio-facial dysostosis, disproportionately short arms, and other anomalies (5 examples). Proc R Soc Med 1954;47:1040–4.
3 Escobar V, Gleiser S, Weaver DD. Phenotypic and genetic analysis of the silver-Russell syndrome. Clin Genet 1978;12:278–88.
4 Price SM, Stanhope R, Garrett C, Preece MA, Trembath RC. The spectrum of Silver-Russell syndrome: a clinical and molecular genetic study and new diagnostic criteria. J Med Genet 1999;36:837–42.

Figure 3 Flow chart detailing the molecular testing orientation when a Silver-Russell syndrome (SRS) is clinically suspected.

| Patients with SRS suspicion |
|----------------------------|
| Netchine-Harbison clinical scoring system (NH-CSS) |
| Likely SRS Patients |
| Molecular testing |
| 11p15 ICR1 hypomethylation |
| CDKWN1 mutation |
| DLK1/GTL2-DMR hypomethylation |
| SNAP microarray for Chromosomal rearrangement/disomy assessment |
| Unlikely SRS Patients |
| Molecular testing |
| mUPD7 |
| Double negatives |

Image 228x546 to 553x745
Genotype-phenotype correlations

Anderson J, Viskochil D, O’Gorman M, Gonzales C. Gastrointestinal complications of Russell-Silver syndrome: a pilot study. Am J Med Genet 2002;113:15–19.

Hamalainen RH, Moqvist D, Gabbett MB, Otten JJ, Brummer KG, van der Burgt I. 3-M syndrome: description of six new patients with review of the literature. Clin Dysmorphol 2006;7:473–50.

van der Wal G, Otten JJ, Brummer KG, van der Burgt I. 3-M syndrome: description of six new patients with review of the literature. Clin Dysmorphol 2001;10:241–52.

Lai KY, Skuse D, Stanhope R. Prenatal alcohol and cognitive abilities associated with the Silver-Russel syndrome. Arch Dis Child 1994;71:490–6.

Netchine I, Rossignol S, Baciller J, Brachet C, Heinrichs C, Carel JC, Loeuille GA, Fingholt T, Girard Y, Menten B. 11p15 imprinting center region 1 loss of methylation is a common and specific cause of typical Russell-Silver syndrome: clinical scoring system and epigenetic-phenotypic correlations. J Clin Endocrinol Metab 2007;92:3148–54.

Bartholdi D, Krajewska-Walasek M, Ounap K, Gaspar H, Chrzanowska KH, Ilyana H, Dias RP, Nightingale P, Hardy C, Kirby G, Tee L, Price S, Macdonald F, Barrett TG, Schonherr N, Meyer E, Eggermann K, Ranke MB, Wollmann HA, Eggermann T. (Epi) Reciprocal 11p15 ICR1 hypomethylation. Hum Mol Genet 2005;14:2121–20.

Maupetit-Mehouas S, Azzi S, Steunou V, Rousseau A, Rossignol S, Thibaud N, Danton F, Le Jule M, Kere J. Submicroscopic genomic alterations in Silver-Russell syndrome and consequences for genetic counseling in children with Silver-Russell syndrome: improving the recognition of an underdiagnosed chromosome 14 syndrome: from new molecular insights to the comprehension of imprinting regulation. Curr Opin Endocrinol Diabetes Obes 2014;21:30–8.

Azzi S, Blaise A, Steunou V, Harbison MD, Salem J, Brioude F, Rossignol S, Habib WA, Thibaud N, Neves CD, Jule ML, Brachet C, Heinrichs C, Bouc Y, Netchine I. Complex issue-specific epigenotypes in Russell-Silver Syndrome associated with 11p15 ICR1 hypomethylation. Hum Mutat 2014;35:1211–20.

Azzi S, Azzi S, Steunou V, Sax T, Thibaud N, Danton F, Le Jule M, Heinrichs C, Brachet C, Gicquel C, Le Bouc Y, Netchine I. Multicentric methylation analysis in a large cohort of 11p15-related foetal growth disorders (Russell Silver and Beckwith-Wiedemann syndrome) reveals simultaneous loss of methylation at paternal and maternal imprinted loci. Hum Mol Genet 2008;17:4724–33.

Maupetit-Mehouas S, Azzi S, Steunou V, Sakakini N, Silve C, Reynes C, Perez de Moria E, Kere J. Epigenotype-phenotype correlations in Silver-Russell syndrome. J Med Genet 2010;47:760–8.

Fuke T, Mizuno S, Nagai T, Hasegawa R, Honikawa R, Miyoshi Y, Muroya K, Kondoh T, Numakura C, Sato T, Nakabayashi K, Tayama C, Hata K, Sano S, Matusbaka K, Kagami M, Yamazawa K, Ogata T. Molecular and clinical studies in 136 Japanese patients with Silver-Russel syndrome. PLoS ONE 2013;8:e66105.

Kotzot D, Utermann G. Uniparental disomy (UPD) other than 15: phenomenotypes and bibliography updated. Am J Med Genet A 2005;136:287–305.

Brunetti-Pierri N, Berg JS, Scaglia F, Belmont J, Baccino F, Silavat S, Matsumoto M, Thomson P, Gallicchio F,等内容...