Research Article

Fluid Retention Caused by Rosiglitazone Is Related to Increases in AQP2 and αENaC Membrane Expression

Jinghua Xu, Mingyue Pan, Xiaoli Wang, Lishi Xu, Lanfang Li, and Cheng Xu

Department of Physiology, School of Life Science and Biopharmaceutics, Shenyang Pharmaceutical University, 103 Wenhua Road, Shenyang 110016, China

Correspondence should be addressed to Cheng Xu; 2443759092@qq.com

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1. Introduction

Rosiglitazone (ROS), a classic clinical oral antidiabetic drug, is a PPARγ agonist; studies have shown that the decreased phosphorylation of PPARγ due to ROS is the main reason for the increase in insulin sensitivity caused by this drug [1]. However, long-term clinical observations have revealed that ROS has the side effect of fluid retention, which leads to heart failure [2, 3]. SR1664, a novel compound developed by the Scripps Institute in the United States and other institutions, is a PPARγ ligand that activates PPARγ to suppress PPARγ Ser273 phosphorylation, leading to enhanced insulin sensitivity, thereby playing a role in treating type 2 diabetes [4]. However, in vivo experiments showed that SR1664 can increase insulin sensitivity and does not cause fluid retention [5].

Substantial research showed that fluid retention is closely related to AQP2 and αENaC proteins [6, 7], and in vitro experiments indicated that the expression of AQP2 and αENaC is upregulated after PPARγ agonist treatment [8, 9]. Nevertheless, the mechanism of ROS on membrane, cytoplasmic, and nuclear AQP2 and αENaC has yet to be clarified.

In this study, we investigated the effects of ROS, SR1664, and TNFα (increased phosphorylation of PPARγ) on p-PPARγ, AQP2, and αENaC in HEK293 and mIMCD-3 cells, being then coincubated with PPARγ antagonist GW9662; results showed that the effects disappeared. So we concluded that in vitro the decrease of PPARγ phosphorylation has little relationship with fluid retention, and the fluid retention induced by ROS is mainly related to the increase of membranes AQP2 and αENaC.

2. Materials and Methods

2.1. Chemicals and Reagents. Reagents were purchased from the following sources: rosiglitazone and GW9662 (Sigma, St Louis, MO); RPMI-1640 medium (GIBCO, Invitrogen) and fetal bovine serum (FBS; Shenyang Huibai Biotechnology Co., Ltd.); 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium (MTT; Biosharp); and ECL Western Blotting Detection Reagent (Bio-Rad, Hercules, CA, USA). Antibodies were selected using polyclonal antibody, PPARγ (Santa Cruz, CA, USA), p-PPARγ (Proteintech, CA, USA), αENaC (Santa Cruz, USA), AQP2 (Santa Cruz, USA), and β-tubulin (Santa...
Cruz, USA); FITC Alexa 488-conjugated goat anti-rabbit secondary antibodies were from Santa Cruz Biotechnology (USA); and horseradish peroxidase-conjugated anti-rabbit secondary antibodies were from Proteintech (USA).

2.2. Cell Culture and Administration. Mouse kidney inner medullary collecting duct (mIMCD) cells (Shanghai Bogu Biological Technology Co., Ltd.) and human embryo kidney (HEK293) cells (ATCC, Manassas, VA, USA) were routinely cultured in RPMI-1640 medium supplemented with 10% FBS and antibiotics (Serva & AMRESCO), in a humidified chamber containing 5% CO₂ at 37°C. Combined administration regimen is as follows: when the cells reached 80%–90% confluence, GW9662 (5 μM) was incubated for 6 h before the addition of ROS (1, 10 μM) or SR1664 (1, 10 μM). After 24 h, cells were collected for extraction of total, cytoplasmic, membrane, nucleus, or membrane protein, respectively.

2.3. Cell Viability Experiment. For the MTT assay, cells were seeded at 6 × 10⁶ cells per well onto 96-well culture plates and allowed to grow for 24 h after treatment with various concentrations of ROS, SR1664, GW9662, TNFα, or ROS and SR1664 combined with GW9662. After removing the medium, MTT solution (5 mg/ml in PBS) was added and incubated for 4 h and the resulting formazan was solubilized with DMSO (150 μl). The absorption was measured at 490 nm in a multifunctional enzyme marking instrument.

2.4. Preparation of Protein Samples

2.4.1. Preparation of Total Protein. RIPA lysis buffer was added to the cell precipitations and resuspended. After 30 min of lysis at 4°C, the lysates were centrifuged at 12,000 g for 20 min, and the obtained supernatant was used as the total protein.

2.4.2. Preparation of Cytoplasm and Nuclear Proteins. The reagents were dissolved at room temperature and put on the ice immediately. Then, 200 μl cytoplasmic protein extraction reagent A (to which 1 mM PMSF had been added a few minutes previously) was added to 20-μl cell precipitations. Five seconds’ vortex was performed to ensure adequate resuspension, followed by incubation on ice for 10–15 min. Then, 10 μM cytoplasmic protein extraction reagent B was added. Vortexing was again performed for 5 s, followed by incubation on ice for 1 min. Centrifugation was then performed at 12,000 g and 4°C for 5 min, and the obtained supernatant was used as cytoplasmic protein. Then, 50 μl of PMSF-added nucleoprotein extraction reagent was added to the nuclear pellet, followed by vortexing for 15–30 s, to ensure complete suspension and dispersal. Then, after incubation on ice, vortexing for 30 s was performed every 1-2 minutes for 30 min. This was followed by vortexing at 12,000 g at 4°C for 10 min, from which the obtained supernatant was used to represent nuclear protein. The nuclear and cytoplasmic extracts were then analyzed for protein content using BCA assay.

2.4.3. Preparation of Membrane Proteins. The cellular membrane fraction was prepared in accordance with the manufacturer's instructions. 1 ml membrane protein extraction reagent A with PMSF was added to 2–5 billion cells, for gentle and complete suspension, followed by incubation on ice for 10–15 min. Next, centrifuging was applied at 700 g for 10 min at 4°C. The supernatant obtained from this procedure was then centrifuged at 14,000 g for 30 min at 4°C to precipitate membrane fragments, with the obtained supernatant being used to represent cytoplasmic protein. The precipitate was also centrifuged at 14,000 g for 10 s at 4°C and exhausted the supernatant completely. Then, after the addition of 200 μl of membrane protein extraction reagent B, vortexing was performed for 5 s for resuspension, followed by incubation on ice for 5–10 min. The previous steps were then repeated 1-2 times to extract the membrane protein completely. Subsequently, centrifugation was performed at 14,000 g for 5 min at 4°C, with the obtained supernatant being used as the membrane protein. The membrane extracts were then analyzed for protein content using BCA assay.

2.5. Western Blot Analysis. Cells were first washed with cold PBS three times and lysed in RIPA buffer. The BCA protein assay was used to determine the protein concentrations of the samples. Equal amounts (25 μg) of cellular proteins were loaded into each well and separated by 10% SDS-PAGE after denaturation with 5x loading buffer and then transferred onto PVDF membranes, incubated in 5% nonfat dry milk for 2 h on shaker at room temperature and then incubated with PPARγ (1:500), p-PPARγ (1:500), AQP2 (1:800), and αENaC (1:800) antibodies, respectively; β-tubulin (1:1000) was used as internal control. Finally, blots were also incubated with secondary antibody (1:5000) and visualized using enhanced ECL luminous fluid.

2.6. Immunocytochemistry. Cells in the logarithmic growth phase were collected and, following adjustment of cell suspension density to 1 × 10⁵ cells/ml after digestion, the cells were inoculated on cover slips with concentrations of drugs diluted with culture medium, cultured at 37°C for 24 h, and blocked for 30 min without light at room temperature [4% paraformaldehyde (PFA), 0.2% Triton X-100, and 5% BSA]. Overnight incubation with primary antibodies (diluted with 1% BSA) was performed at 4°C followed by 30 min of incubation with secondary antibodies (diluted with 1% BSA). Washing with PBS was then performed three times for 10 min after each step, along with exposure to 100 ng/ml Hoechst 33258 dye for 10 min, then washing again with PBS for 5 min three times. Finally, confocal microscopy was performed using a 60x oil objective on a Nikon C2-si laser-scanning confocal microscope, and images were manipulated using Photoshop software.

2.7. Data Analysis. All experiments were repeated at least three times independently, and data are expressed as mean ± s.e.m. Statistical analysis was performed using SPSS 17.0 by one-way analysis of variance (ANOVA). The LSD test was used to compare differences in the means between groups; if
the variance is different, Dunnett’s $t$-test was used. A value of $P < 0.05$ was considered to represent a significant difference, while $P < 0.01$ was considered to present a very significant difference.

3. Results

3.1. Cell Viability Analysis. First, to consider the cytotoxic effects of ROS, SR1664, GW9662, and TNF-α, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) experiments were used to determine the doses of these drugs and combination therapies. The administration of ROS, SR1664, and GW9662 at 0.1, 1, and 10 μM (Figures 1(a) and 1(b)) and TNF-α at 1, 5, and 10 ng/ml (Figure 1(c)) to HEK293 cells and mIMCD-3 cells showed no significant effects on cell viability. The administration of these agents in combination (Figure 1(d)) also had no such effects.

3.2. Immunofluorescence Assay. Activation of nuclear receptor promotes the translocation of transcription factors from the cytoplasm to the nucleus to improve the transcriptional activity of transcription factor response element binding (CREB) protein. Therefore, a cell immunofluorescence experiment was used to detect the changes in the distribution of
3.3. Effects of ROS on PPARγ, p-PPARγ, AQP2, and αENaC in HEK293 Cells. To investigate the effects of ROS on PPARγ, p-PPARγ, AQP2, and αENaC at different locations, total, nuclear, membrane, and cytoplasmic proteins were extracted. The data show that ROS at 10 μM can significantly increase nuclear PPARγ (N-PPARγ) (Figure 3(a)) but downregulated total p-PPARγ (T-p-PPARγ) and nuclear p-PPARγ (N-p-PPARγ) and had no influence on cytoplasmic p-PPARγ (C-p-PPARγ) (Figure 3(b)). As for AQP2, membrane (M-AQP2) and cytoplasmic (C-AQP2) proteins levels were increased (Figure 3(c)), while, for αENaC, the findings show that just membrane (M-αENaC) translocation increased (Figure 3(d)). After coincubation with GW9662, the effects of ROS on PPARγ, p-PPARγ, AQP2, and αENaC disappeared (Figures 3(e)–3(h)).

3.4. Effects of ROS on PPARγ, p-PPARγ, AQP2, and αENaC in mIMCD-3 Cells. To validate the above findings in HEK293 cells, the same experiments were conducted in mIMCD-3 cells. The data indicate that ROS (10 μM) also critically increased nuclear PPARγ (Figure 4(a)), downregulated total and nuclear p-PPARγ in mIMCD-3 cells (Figure 4(b)), and increased the expression of AQP2 in cytoplasm and membrane (Figure 4(c)), as well as membrane expression of αENaC (Figure 4(d)). Upon coincubation with GW9662, these effects disappeared (Figure 4(e)–4(h)). The results are consistent with those in HEK293 cells.

3.5. Effects of SR1664 and TNFα on PPARγ, p-PPARγ, AQP2, and αENaC in mIMCD-3 Cells. SR1664 is a new type of PPARγ ligand that blocks the cyclin-dependent kinase 5-(Cdk5-) mediated phosphorylation of PPARγ. Research has shown that it improves insulin sensitivity by lowering glucose and has no side effects, as is the case for ROS. To confirm these effects, we detected PPARγ, p-PPARγ, AQP2, and αENaC proteins in mIMCD-3 cells. The data suggest that SR1664 (10 μM) can dramatically limit the p-PPARγ (Figure 5(a)); upon coincubation with GW9662, the effects on p-PPARγ disappeared (Figure 5(b)). Nevertheless, the expression of AQP2 in the cytoplasm and membrane and the membrane expression of αENaC exhibited no significant difference (Figures 5(c) and 5(d)).

Obesity-linked insulin resistance is associated with inflammation in adipocytes. Among the different types of proinflammatory cytokines, TNFα is the first one identified to connect obesity, inflammation, and insulin resistance. Notably, TNFα at both 5 and 10 ng/ml can upregulate p-PPARγ (Figure 5(e)), while it had no effect on αENaC in membrane and AQP2 in the cytoplasm and membrane (Figures 5(f) and 5(g)).

4. Discussion

In this study, we determined the reason for the relationship between fluid retention caused by ROS and the expression of AQP2 and αENaC in HEK293 and mIMCD-3 cells. Immunofluorescence experiment reveals that the green fluorescence in the nucleus increased after ROS application compared with that in the control, illustrating that ROS can activate the transcriptional activity of transcription factors
Figure 3: Continued.
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Figure 3: Effects of rosiglitazone and coincubation with GW9662 in HEK293 cells. (a–d) The effects of rosiglitazone on PPARγ, p-PPARγ, AQP2, and αENaC. (e–h) The changes of PPARγ, p-PPARγ, AQP2, and αENaC upon coincubation with GW9662 (10μM ROS + 5μM GW9662). The results show that rosiglitazone at 10 μM can significantly decrease total and nuclear p-PPARγ but critically upregulated nuclear PPARγ and AQP2 and αENaC membrane transposition. After coincubation with GW9662 (10μM ROS + 5μM GW9662), the effects of rosiglitazone on PPARγ, p-PPARγ, AQP2, and αENaC disappeared. The results are shown as mean ± s.e.m., n = 3. *+/# P < 0.05. **+/## P < 0.01. * means compared to control; # means compared to rosiglitazone.

and promote their transfer from the cytoplasm to the nucleus. Next, we studied the effects of ROS on p-PPARγ at the protein level in HEK293 and mIMCD-3 cells. As shown in Figures 3(b) and 4(b), in both HEK293 and mIMCD-3 cells, ROS (10 μM) can critically inhibit PPARγ phosphorylation, in terms of both the total level and that in the nucleus; upon coincubation with GW9662, an antagonist of PPARγ, all of these effects disappeared. Taking these findings together, ROS activated PPARγ, leading to the reduction of p-PPARγ.

Against this background, to determine whether the reduction of p-PPARγ could lead to fluid retention, we examined the expression of AQP2 and αENaC proteins, which are related to body fluid homeostasis [10, 11]. The trafficking mechanism of AQP2 was mainly induced by arginine vasopressin (AVP); when AVP increased, the cytoplasmic vesicles and lumen membrane fused, and AQP2 was transferred to the luminal membrane, increasing the permeability to water [12]. Based on these findings, an abnormal mechanism of AQP2 trafficking would affect the number of AQP2 molecules in the luminal membrane [13]. In diabetic model mice in vivo, after feeding on PPARγ agonist, PCR detection results showed that ADH had no significant changes, but AQP2 significantly increased [14]. Our results revealed that ROS (10 μM) can increase the expression of AQP2 in the cytoplasm and facilitate AQP2 vesicles fusing to the cell membrane; in addition, after coincubation with GW9662, these effects were all offset. These findings may be explained by ROS increasing...
Figure 4: Continued.
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the expression of AQP2 in the collecting duct membrane and increasing water reabsorption, leading to fluid retention.

The regulation of renal sodium (Na\(^+\)) handling is a key determinant of the long-term control of extracellular fluid volume homeostasis. Na\(^+\) reabsorption is mediated via the amiloride-sensitive epithelial sodium channel (ENaC), which exhibits high selectivity for sodium [15] and is a central requirement for Na\(^+\) reabsorption across renal epithelia. ENaC expression and translocation to the plasma membrane are tightly regulated by a diverse array of hormonal [16, 17] and physical factors [18]. Our experiments show that ROS (10 μM) remarkably raised the membrane level of αENaC, and thus more and more Na\(^+\) flowed into the lumen, which accelerated water reabsorption and caused more serious fluid retention. After incubation with GW9662, this effect dissipated.

SR1664 is a novel PPARγ ligand (a nonagonist PPARγ ligand) that blocked the cyclin-dependent kinase 5 (CDK5)-mediated phosphorylation of PPARγ. In vivo experiments show that it can increase insulin sensitivity and does not cause fluid retention [5]. This study focused on the relationship between SR1664 and AQP2/αENaC proteins in vitro, indicating that SR1664 downregulated p-PPARγ in the membrane, demonstrating that SR1664 increased insulin sensitivity without affecting water and sodium channel protein expression.

Figure 4: Effects of rosiglitazone and coincubation with GW9662 in mIMCD-3 cells. (a–d) The effects of rosiglitazone on PPARγ, p-PPARγ, AQP2, and αENaC. (e–h) The changes of PPARγ, p-PPARγ, AQP2, and αENaC upon coincubation with GW9662 (10 μM ROS + 5 μM GW9662). The results show that rosiglitazone at 10 μM can significantly decrease total and nuclear p-PPARγ but critically upregulated nuclear PPARγ and AQP2 and αENaC membrane transpositions. After coincubation with GW9662 (10 μM ROS + 5 μM GW9662), the effects of rosiglitazone on PPARγ, p-PPARγ, AQP2, and αENaC disappeared. The results are shown as mean ± s.e.m., n = 3. * p < 0.05. **/## p < 0.01. * means compared to control; # means compared to rosiglitazone.
Figure 5: Continued.
Figure 5: Continued.
Activated PPARγ suppresses the expression of TNFα [19, 20], and TNFα increases the level of insulin antagonistic hormones by phosphorylating serine residue of the substrates of the insulin receptor, inhibiting tyrosine phosphorylation of this receptor, which in turn limits signal transmission [21]. Furthermore, both ROS and SR1664 are reported to eliminate p-PPARγ upregulated by TNFα [22]. Prompted by these findings, we also studied the correlation of phosphorylated PPARγ induced by TNFα with AQP2 and αENaC expression. As shown in Figure 5, TNFα at both 5 and 10 ng/ml can upregulate p-PPARγ, but the expression of AQP2 and αENaC in the cytoplasm and membrane was not changed by it.

In conclusion, in vitro the fluid retention induced by ROS is closely associated with two major aspects: the first includes the increase of cytoplasmic AQP2 and promotion of AQP2 vesicles to undergo membrane fusion, thereby increasing water reabsorption; the second involves ROS enhancing membrane αENaC expression, thus accelerating Na+ reabsorption, which further increases the absorption of water. In contrast, SR1664 and TNFα experiments reveal that whether activated PPARγ was up- or downregulated did not affect AQP2 and αENaC expression. From the former evidence, we deduced that in vitro there was little association between the fluid retention induced by ROS and the phosphorylation of PPARγ.

**Conflicts of Interest**

The authors declare that there are no competing or financial interests.

**Authors’ Contributions**

Jinghua Xu and Mingyue Pan performed the molecular biology experiments and analyzed data, and Mingyue Pan wrote the manuscript. Xiaoli Wang designed the methodology. Lishi Xu and Lanfang Li prepared the figures and images. Cheng Xu guided the whole experiments. Jinghua Xu and Mingyue Pan contributed equally to the work.

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