INTRODUCTION

Date palm (*Phoenix dactylifera* L.), a monocotyledonous dioecious plant, is one of the most cultivated palms around the world (Abass, 2013). Date palm trees are cultivated in different regions worldwide, especially in the Middle East, North Africa, North and South America, Southern Europe, Pakistan and India (Al-Shahib and Marshall, 2003; Al-Khayri et al., 2015). Date palm grows very well under adverse conditions especially in arid and semiarid regions and produces highly valued date fruits. Date palm fruit is rich in carbohydrates and nutrients. It contains sugars in mostly inverted form, dietary fiber, high levels of essential amino acids and minerals (El-Far et al., 2016).

Propagation through offshoots is slow technique, limited number of offshoots, survival of low life and the risk of disease transmission rate (Al-Khalifah and Askari, 2011). Propagation of date palm via *in vitro* culture using shoot tip explants provides effective substitution to traditional methods (Quiroz-Figueroa et al., 2006). Somatic embryogenesis is the common method used for *in vitro* propagation of date palm (Abahmane, 2013) depending on the development of callus and embryos to salt stress under *in vitro* conditions.
Salinity stress is one of the most significant environmental stresses limiting agricultural production worldwide; and affects the anatomical, physiological and enzymatic features of plants (Arslan et al., 2016 and Nawaz et al., 2016). Salinity affects several physiological and biochemical processes; there are two types of effects, first the adverse osmotic effect, which is the presence of high concentrations of salts in the soil solution making it harder for roots to extract water and reducing the ability of the plant to take it up, leading to slower growth (Munns and Tester, 2008). Osmotic stress delays the growth of plant and affects cell division and elongation. The division of cells is crucial processes which determine the meristem activity and the overall plant growth rate (Bartels and Sunkar, 2005). Secondly, the toxicity effects, which are the presence of high concentrations of salt in the plant which can be toxic and lead to cellular damage (Munns, 2005). The salinity stress causes a negative impact on plant physiology including reduced photosynthetic capacity, impaired signaling and alterations to cellular metabolism (Munns and Tester, 2008). To understand the effect of salinity on the behavior of date palm in vitro culture, several studies have been conducted (Al-Khayri and Al-Bahrany, 2004; Al-Bahrany and Al-Khayri, 2012; Al-Khayri and Ibraheem, 2014). Responses of date palm in vitro to salt stress by using different concentrations of NaCl and the positive effect of NaCl at low concentration (25 µM) on the proliferation of shoot tip derived callus was examined. Accumulation of proline in callus tissue date palm cultured under salt stress has been noted with stress tolerance (Al-Khayri, 2002). Under stress the proline acts as an alternative resource for carbon and nitrogen, helps reducing oxidative damages and stabilizing DNA and membrane protein (Szabados and Savoure, 2010).

The aim of this study was to determine the effect of different concentrations of NaCl on growth, development and differentiation of in vitro embryogenic callus of Date Palm Hayani cv. and find out the effect of salt stress on the biochemical content of embryogenic callus and somatic embryos of date palm as well as, their content from total carbohydrate, total proline, total protein and minerals.

**MATERIALS AND METHODS**

This study was conducted in The Central Lab of Date Palm for Researches and Development - Agricultural Research Center, Egypt during the period from 2017 to 2018.

**Offshoot preparation**

The date palm offshoots Hayani cv. about 3-4 years old and weighing 5-7 kg were separated from healthy mother trees from date palm orchard in (Sakara), Giza governorate, Egypt. The shoot tips were washed with distilled water to eliminate any organic ruins then soaked in Benlate 1 g/l fungicide solution for 10 min and rinsed three times with sterile distilled water. The offshoots were thoroughly cleaned and the outer leaves were removed to expose the shoot tip under aseptic condition, the shoot tips separated and immersed in antioxidant solution (150 mg citric acid and 100 mg ascorbic acid). Shoot tips were surface sterilized with 70% ethanol for 1 min followed by sodium hypochlorite treatment (60 % v/v Clorox solution) containing 2 drops of tween 20 for 25 min after that washed three times with sterile distilled water, soaked in 0.2% HgCl₂ solution for 5 min. The shoot tips were then rinsed thoroughly with sterile distilled water, as shown in Fig. 1 (A). All leaf primordial were removed except 2 pairs surrounding the apical meristems and shoot tip was cut into 6 equal segments, then instantly moved to a prepared culture medium.

**Initiation stage**

For callus culture initiation, the pieces of shoot tips were cultured into callus induction medium consisting of MS salts (Murashige and Skooge, 1962) supplemented with Na₂HPO₄ (170.0 mg/l), myo-inositol (100.0 mg/l), glutamine (200.0 mg/l), nicotinic acid (0.5 mg/l), pyridoxine-HCl (0.5 mg/l), thiamine (1.0 mg/l), sucrose (40.0 mg/l), agar (6.0 g/l) and 0.5 g/l of activated charcoal. Plant growth regulators were added as per requirement for each in vitro stage.

For initiation stage, MS growth medium supplemented with 10.0 mg/l of 2,4-dichlorophenoxy acetic acid (2,4-D), 3.0 mg/l of isopentenyl adenine (2ip) and the medium was adjusted to a pH of 5.7 and autoclaved. Explants were cultured on this medium and incubated in dark culture room at 25 ± 2°C for 4 months (four subcultures) to improve the initiation of callus during this period explants were re-cultured on the same medium each 4 weeks. Explants swell during the second subculture. After 4 months from culture the callus was initiated and the resultant callus
was transferred to callus proliferation medium, as shown in Fig. 1 (B, C).

**Callus formation stage**

The concentration of 2,4-D was reduced gradually to enhance the proliferation callus as shown in Table 1. The explants were cultured on MS medium having 2,4-D at 5.0 mg/l mixed with 2iP at 3.0 mg/l for three subcultures. After that the callus was transferred to embryogenic callus medium which consist of MS basal medium supplemented with 2,4-D at 3.0 mg/l with 2iP at 1.0 mg/l to produce the embryogenic callus, as shown in Fig. 1 (D).

All cultures were incubated in the growth room at 25±2°C for 3 months (three subcultures) at total darkness. Explants or calli were recultured on the same medium each for 4 weeks. Thereafter, embryogenic callus was maintained on callus maintenance medium which consists of MS basal medium supplemented with BA at 0.05 mg/l + NAA at 0.1 mg/l. These cultures served as a callus source for the salt induced stress study.

**Table 1. Plant growth regulators were requirement for date palm in vitro culture stages**

| Plant growth regulators | Callus induction | Callus initiation | Callus proliferation | Embryogenic callus | Callus maintained |
|-------------------------|------------------|-------------------|---------------------|-------------------|------------------|
| 2,4-D                   |                  | 10 mg/l           | 5 mg/l              | 3 mg/l            | -                |
| 2iP                     |                  | 3 mg/l            | 3 mg/l              | 1 mg/l            | -                |
| BA                      | -                | -                 | -                   | -                 | 0.05 mg/l        |
| NAA                     | -                | -                 | -                   | -                 | 0.1 mg/l         |

Fig. 1. Different stage of *in vitro* date palm Hayani cv., A: Offshoot preparation, B: swelling stage, C: callus formation stage, D: Embryogenic callus formation
Salinity stress

The embryogenic callus was used to begin in vitro salt tolerance experiments. For that, we used MS basal nutrient medium supplemented with BA at 0.05 mg/l + NAA at 0.1 mg/l with the addition of different concentrations of NaCl (0, 500, 1000, 1500 and 2000 ppm) with five replicates. Cultures were incubated in complete darkness at 25 ± 2°C for 8 weeks (two subcultures). Each treatment consisted of 8 cultures containing about 0.5 g embryogenic callus/jar.

a. Physiological traits

After the incubation period, all exposed callus was analyzed to determine their responses to salt stresses, the fresh weight of embryogenic callus was measured after 8 weeks from culture and also the number of embryos induction on callus was counted.

b. Biochemical analyses

Chemical analyses were done for embryogenic callus and somatic embryos including the following:

1. Determination of total carbohydrates

The total carbohydrates were determined in the embryogenic callus and somatic embryos of date palm according to the method of Watanabe et al. (2000) using a spectrophotometer which measure absorption spectrum at wave length of 620 nm and standard curve of glucose was made for calculation.

2. Determination of proline

Proline content was determined by a spectrophotometric assay as described by Bates et al. (1973).

3. Determination of total soluble protein

The method of Bradford (1976) was followed to measure the total soluble protein, Albumin was used to make a standard curve and absorbance measured at 595 nm.

4. Mineral analysis

The Na⁺ and K⁺ concentration in the treated calli were measured based on the method described by Skoog et al. (2007) using Jenway Model PEP7 flame photometer.

Embryos culture

After the experiment of salt stress, the embryo clusters were derived from callus cultures and transferred to fresh medium without NaCl consists of ¾ MS basal medium supplemented with 30 g/l sucrose, 0.1 mg/l NAA and 0.05 mg/l BA (Ibrahim et al., 2009) for continued embryo proliferation. The somatic embryo cultures were incubated in growth room at 26 ± 2°C with light conditions 16-h photoperiods, and recultured to the same fresh medium for two subcultures (8 weeks) to increase mass production of somatic embryos and germination.

Shoot elongation and rooting stage

Regeneration shoots were transferred to the ¾ MS medium supplemented with 0.5 mg/l gibberellic acid (GA₃) + 1.0 mg/l NAA to obtain shoot elongation and root formation (AL-Mayahi, 2015). The cultured jars were incubated at 25 ± 2°C with light intensity 3000 lux for 16h photoperiod in the growth room for 8 weeks and each 4 weeks the shoots were transferred to the same fresh medium. To enhance root formation, for two month (two subculture) the healthy regenerated plantlets were cultured on ½ MS medium supplemented with 0.1 mg/l IBA, 1.5 g/l activated charcoal and 30 g/l sucrose (Eke et al., 2005) and incubated under the same conditions as described above.

Statistical analysis

This experiment was designed as a randomized complete block design as described by (Gomez and Gomez, 1984). The obtained data were statistically analyzed using MSTAT Computer Program (MSTAT Development Team, 1989). To verify differences among means of various treatments, means were compared using Duncan's Multiple Range Test as described by (Duncan, 1955).

RESULTS AND DISCUSSION

Effect of salinity on callus fresh weight (g)

The embryogenic callus continued to grow normally without any sign of cell dehydration or salt induced damage showed tolerance to the lower concentration of NaCl added to the culture medium. This may point to the suitability of the osmotic potential of the medium for date palm callus growth and the best proliferation (Al-Mansoori et al., 2007).

Results presented in Table 2 and Fig. 2 show the effect of NaCl concentration on fresh weight of embryogenic callus. Callus fresh weight was determined after 8 weeks in salt treatments. The results showed a gradual decrease in fresh weights with an increase in salt concentration over 1000 ppm of NaCl as compared to the control treatment. Salt concentrations at 500 and 1000 ppm gave the highest fresh weight reached 4.84 and 4.79 g,
respectively compared with control treatment 3.22 g and significant difference with all treatments. Also, there was a significant decrease in fresh weight at concentrations of 1500 and 2000 ppm and the lowest fresh weight was recorded with concentration of 2000 ppm reached 1.82 g which differed significantly with control and all other treatments.

Perhaps low NaCl concentrations (500 and 1000 ppm) are optimal for the growth of the embryogenic callus and somatic embryos. It may also be the reason behind the significant enhancement of its fresh weight due to the maximum possible absorption of water and essential mineral elements by cells as a means of resistance to salt stress conditions (Rains et al., 1986). It was found that using NaCl at concentrations of 25-50 µM caused a significant increase in callus fresh weight of other date palm cultivars Barhee and Helali (Al-Khayri, 2002).

But the reduction in callus fresh weight at high salt stress 2000 ppm may be attributed to the negative effect of salt on cells grown at high salt concentrations. This finding was similar with previous reports under in vitro salinity stress (Al-Khayri, 2002; Queiros et al., 2007; Rafiq et al., 2008; Lokhande et al., 2011 and Golkar et al., 2017).

There was a significant decrease in fresh weight of callus with high concentration of NaCl at 320 µM. The effect on growth is probably due to osmotic effect and ion cytotoxicity, due to the accumulation of Na⁺ and Cl⁻ leading to growth inhibition. It has been suggested that the reduction of growth in response to salinity is the result of great portion of respiratory energy being transferred to processes resulting in salt tolerance rather than growth (Mass, 1986). Data indicated the passive effect of NaCl treatments on callus growth and frequency of somatic embryo germination percentage after 12 weeks. The Highest value of callus growth (1.93 and 2.00 g) was observed at 50 µM NaCl compared to control treatment (1.98 and 2.05 g), following by 100 µM (1.47 and 1.90 g), 200 µM (1.25 and 1.49 g), 300 µM (0.24 and 0.45 g) for cv. Barhi and cv. Khalas, respectively although callus of cv. Khalas recorded the minimum value of callus growth (0.13 g) under 400 µM NaCl (Al-Dhebiani et al., 2018). Adding NaCl to callus culture of N. sativa results in reducing the callus growth from control (0.17 mm) to (0.068 mm) significantly in the treatment with 250 µM NaCl under in vitro salt stress. In addition, salinity stress represents a negative osmotic stress, leading to a decrease in the water available in the callus cells, which consumes energy leading to the reduced growth of callus (Golkar et al., 2019). Salinity stress could reduce the plant productivity by unbalancing cellular ions (Tester and Davenport, 2003). It had been previously observed by many workers that the presence of salt in the medium generally reduced or even completely inhibited the plant regeneration (Lutts et al., 1999).

**Effect of salinity on somatic embryos induction**

There was a significant difference between different concentrations of NaCl and the number of somatic embryos. The number of somatic embryos of Hayani cv. was enhanced in response to low level of salinity as NaCl at 500 and 1000 ppm which resulted in the increase in the number of somatic embryos 12.45 and 11.60 embryo/jar, respectively compared with the control treatment which recorded low number of embryos 5.13 embryo/jar. However, increasing the concentration of NaCl over 1000 ppm decreased the number of embryos gradually; it was reduced from 6.29 to 4.03 embryos/jar with the treatments 1500 and 2000 ppm NaCl, respectively as shown in Table 2 and Fig. 2.

**Table 2.** Effect of NaCl concentrations on fresh weight of embryogenic callus and number of somatic embryos of the date palm Hayani cv. after 8 weeks from culture

| NaCl concentration (ppm) | Fresh weight (g) of embryogenic callus | Number of somatic embryos |
|--------------------------|----------------------------------------|---------------------------|
| 0                        | 3.22 b                                 | 5.13 b                    |
| 500                      | 4.84 a                                 | 12.45 a                   |
| 1000                     | 4.79 a                                 | 11.60 a                   |
| 1500                     | 2.47 c                                 | 6.29 c                    |
| 2000                     | 1.82 d                                 | 4.03 d                    |
These results were in agreement with several studies, low levels of salinity can enhance growth and development of in vitro date palm callus culture (Al-Khayri, 2002). Moreover, the number of somatic embryos of cv. Zaghloul was enhanced in response to low concentration (25 µM) of NaCl. However, higher concentrations of NaCl reduced callus growth and the number of somatic embryos (Ibraheem et al., 2012). In addition, using of seawater in all in vitro culture stages, proved to be highly effective for enhancing somatic embryogenesis of cv. Malkaby (Taha, 2014). According to Ibraheem et al. (2012) the number of somatic embryos of cv. Zaghloul was enhanced with adding 25 µM NaCl to the regeneration medium; however, at 75 µM NaCl the number of resultant somatic embryos was reduced and no embryo formed at 175 µM NaCl.

The results showed that increasing NaCl concentration significantly decreased the fresh weight of both the embryogenic callus and somatic embryos (Abbas et al., 2012). The addition of NaCl to the media caused a significant effect on the reduction of somatic embryo germination percentage (Al-Zubaidi et al., 2013 and Ibraheem, 2013). Adverse effect was reported by Jasim et al. (2010) where 0.5-2.0% NaCl inhibited callus growth and somatic embryogenesis of cv. Ashkar. The presence of salt in culture medium resulting in the loss of regeneration in organogenic callus and shoots were obtained from culture media with 42.7 mm and 85.57 mm NaCl (Sharry and Teixeira da Silva, 2006). Date palm regeneration through somatic embryogenesis is affected by the salt concentration of the culture medium. A low concentration of salt (0.4% NaCl) observed to increase the length of in vitro shoots of cvs. Bartamuda, Sewy and Samani. However, shoot growth reduction was noticed at 0.8% and 1.2% NaCl (El-Sharabasy et al., 2008).

**Chemical analysis**

Plant acclimation under abiotic stresses such as salinity has been examined through biochemical analysis. There is much studies of embryos subjected to salt treatment (Munns, 2002 and Santos et al., 2011). The tolerant cells make osmotic adjustments in response to abiotic stress; such as, highly soluble compounds like sugars, amino acids and proline accumulate in higher plants under salinity stress (Ashraf, 1994). They serve as organic compatible solutes, which cause osmotic adjustments and play a role in stabilization of proteins, protection of membrane and help in regulating ionic sequestration (Patade et al., 2011).
**Determination of total carbohydrates**

Data in Fig. 3 revealed that, there was a significant difference between different concentrations of NaCl and total carbohydrate content in the embryogenic callus. Added NaCl to culture medium at concentration 500 ppm increased the carbohydrate content (21.80 mg/g fw) in the embryogenic callus compared with the control treatment (20.16 mg/g fw). Increasing NaCl concentration to 1000 ppm increased total carbohydrate content to (22.5 mg/g fw); however, increasing the concentration of salinity stress over 1000 ppm to 1500 ppm was resulted reduced the total carbohydrate content to (20.00 mg/g fw).

While, the lowest carbohydrate content (17.30 mg/g fw) was recorded at the highest concentration of NaCl at 2000 ppm.

There was significant difference between different concentrations of NaCl and total carbohydrate content in the somatic embryos. The highest total carbohydrate content (20.76 and 21.60 mg/g fw) was recorded with the concentration of NaCl at 500 and 1000 ppm compared with the control treatment which was (19.20 mg/g fw). The content of carbohydrate was reduced due to increasing the salinity concentration to 1500 and 2000 ppm and recorded (18.62 and 16.80 mg/g fw), respectively.

Fig. 3. Effect of different concentrations of NaCl on carbohydrate content (mg/g fw) in embryogenesis callus (A) and somatic embryos (B) after 8 weeks in culture

These results were in agreement with (Mehr, 2013) who reported the increase in the amount of production of some compounds in plants as exposure to salt stress and this is one of the mechanisms by which plants resist salt stress examples of such compounds are carbohydrates. Jasim et al. (2010) also found that an increase of NaCl concentrations in the medium led to a decrease of total soluble carbohydrates and proteins of embryogenic callus and somatic embryos of date palm, but it increased free proline.

The decrease in carbohydrates under salt stress condition might be related to the salt stress that make cells spend more energy for osmoregulation to endure the turbulence caused by salt accumulation inside cells, also high salt condition increase respiration rate through the effect of sodium ions on respiration cycle that led to the decrease in carbohydrates (Wang et al., 1999 and Huang and Liu, 2002). Salinity induced soluble sugar accumulation has also been observed in *P. euphratica* (Watanabe et al. 2000; Zhang et al. 2004). The best characterized biochemical response of plant cells to osmotic stress is accumulation of some compatible organic solutes like soluble sugars (Flowers and Colmer, 2008). The results showed clearly that salinity significantly inhibited fresh weight of the callus of *Citrus sinensis* and decreased the concentration of carbohydrates (Abbas et al., 2013). The carbohydrate concentration of *Salsola arbuscula* callus increased with used 300 µM of NaCl, but decreased significantly in the 400 µM of NaCl (Amini et al., 2017).

Salinity also induces oxidative stress, which influences enzyme activity causing a reduction in carbohydrate metabolism (Huang and Liu, 2002). It was observed that low salinity caused by NaCl at 10 µM significantly increased total soluble carbohydrate in plantlet leaves as reported by Al-Kabi (2004) for date palm. As the concentration of NaCl increased, the concentration of soluble carbohydrates was significantly decreased. The salt tolerant callus of sugarcane accumulated more soluble sugars under NaCl stress (Gandonou et al.,...
However, the soluble sugars did not accumulate in calli from salt-resistant wheat cv. Belikh (Lutts et al., 2004). Increasing seawater levels increased total soluble sugars content compared with the control. Total soluble sugars in leaves of two young Iranian commercial olive cultivars increased with an increase in salinity up to 80 µM but decreased with additional increase in salinity (Mousavi et al., 2008). In addition, soluble carbohydrate concentration of two sugar beet cultivars significantly increased with increasing salt stress. Palma et al. (2009) suggested that the response to salinity (100 µM NaCl) was improved by increasing plant dry weight and decreasing the contents of organic solutes like total soluble sugar contents in Phaseolus vulgaris L. Dhanapackiam and Ilyas (2010) reported that the increase of total carbohydrates in the shoot is considered to be playing an important role in the osmotic adjustment. To evaluate osmotic adjustment of the callus, the contents of soluble sugar were determined. The soluble sugar contents were increased by 1.6-fold under 50 µM NaCl and 1.8-fold under 150 µM NaCl treatment. The soluble sugar content was induced in the alkaligrass calli and leaves under stresses of NaCl (Yu et al., 2011). The decrease in the photosynthesis rate occurred at high salinity, also inhibited cell expansion and cell division stomatal conductance and closure in Bruguiera parviflora followed by the decrease in chlorophyll and carbohydrates content (Sofy and Fouda, 2013).

**Determination of proline**

The data indicate that salt stress had a significant effect on proline content in embryogenic callus cultured in MS medium with different concentration of NaCl. An increase in NaCl concentration cause a significant increase in proline content to 2.06 mg/g fw with adding 2000 ppm of NaCl in culture medium compared with control treatment which recorded (1.03 mg/g fw). Whereas low concentration of salt stress at 500 ppm gave less proline content (1.22 mg/g fw) and differed significantly with control treatment and with all treatments as showed in Fig. 4.

There was a significant difference between different concentration of NaCl and the proline content in somatic embryos. The increase in proline content was found to be gradual with the increase of NaCl concentration in the culture medium. The somatic embryos grown on MS medium supplemented with 1500 and 2000 ppm NaCl showed the highest proline content (1.93 and 2.20 mg/g fw, respectively) in comparison to control treatment which recorded (1.34 mg/g fw). As shown in Fig. 4. The low proline content (1.34 mg/g fw) resulted with low level of salt stress at 500 ppm compared to control.

![Fig. 4. Effect of different concentrations of NaCl on Proline content (mg/g fw) in embryogenesis callus (A) and somatic embryos (B) after 8 weeks in culture](image)

These results agree with the results of Watanabe et al. (2000) in which salinity stress showed to increase the proline content in grape var. Khoshnaw. The highest concentration of free proline was obtained at the 100 µM salinity level, while the lowest was recorded in the control treatment. The calli of Sesuvium portulacastrum grown on MS medium supplemented with 200 and 400 µM salt showed significantly the highest proline content in comparison to control calli (Lokhande et al., 2010).

Physiological studies on callus exposed to salt stress have shown an increase in proline accumulation in response to increased salinity. Proline accumulation was correlated to callus growth inhibition (Al-Khayri, 2002) and hence is useful as a biochemical marker to understand how.
plant tolerates the abiotic stresses (El-Hadrani et al., 2011). The role of proline accumulation in tolerance to salt stress is thought to play role in osmotic phase of salt stress at early stages of salt treatment or at mild salt stress conditions (Arzani, 2008). Proline acts as osmotsis that protects the cytosol from dehydration as symptomatic salt stress damage (Ashraf and Foolad, 2007), proline may act as a regulatory molecule able to activate multiple responses that are component of the adaptation process (Ashraf and Harris, 2004) and act as a osmolyte, stabilizer of the proteins structure, thereby protecting cells from damage caused by stress (Szabados and Savoure, 2010). Under salinity stress, higher levels of glutamate as a preproduct for chlorophyll and proline were consumed to generate proline (Ahmad, 2014). Proline can adjust the osmotic pressure of cells under various stresses. Additionally, free proline can remove and detoxify the reactive oxygen species (ROS) generated as a result of stresses, thus protecting cell membranes against these radicals (Saed-Moucheshi et al., 2014). The response of proline and carbohydrates to salt stress may be due to these osmoprotectants that play an important role in decreasing stress-induced cellular acidification and osmotic adjustments, stabilizes sub-cellular structures for recovery (Tan et al., 2008). In Plantago ovata callus, the proline content averaged over all the genotypes was found to increase with increasing salt from 0.1 mg/l FW in control to 1.1 mg/l FW in 200 μM NaCl (Golkar et al., 2017).

**Determination of total soluble protein**

There was a significant difference between different levels of NaCl and protein content in both of embryogenic callus and somatic embryos. Results in Fig. 5 indicated that increase in NaCl concentration in culture medium led to decrease in protein content in both embryogenic callus and somatic embryos especially under high level of salt stress 2000 ppm where the protein content reached to (0.72 mg/g fw) compared to (2.10 mg/g fw) under control treatment for embryogenic callus. While, in somatic embryos resulted (0.65 mg/g fw) at 2000 ppm salt stress compared to (1.8 mg/g fw) under control treatment.

The decrease in protein content under salt stress might be related to the inhibition of protein synthesis high salty condition (Kaouthar et al., 2001), also high salts in the culture media might inhibit absorption of necessary elements for protein synthesis such as nitrogen, besides the negative effect of salinity on the biosynthesis of mRNA which lead to the disturbance of protein synthesis (Whitting and Wilson, 2003) whereas, the increase in protein content at 20, 40, 60 and 80 μM may be due to stress induced synthesis of proteins (Cherian and Reddy, 2003). Proteins are a potential biochemical useful in salt resistant of plant cells and protect high plants against stresses by adjusting osmotic pressure by many compatible solutes (Shonjani, 2002).

**Fig. 5. Effect of different concentrations of NaCl on total soluble protein (mg/g fw) in embryogenesis callus (A) and somatic embryos (B) after 8 weeks in culture**

Rahnama and Ebrahimzadeh (2004) also observed a decrease in protein in shoots and callus of potato with increasing NaCl concentrations. The effects of NaCl treatments (100 - 200 μM) on protein contents in potato leaves were studied by Fidalgo et al. (2004). They observed a significant decrease in protein content under salt stress.

Shankhdhar et al. (2000) indicated that total protein contents of callus cultures decreased with an increase in salt concentration after 4 weeks of culture in six cultivars of rice callus. Agastian et al. (2000) reported that soluble proteins increased at low salinity but decreased at high salinity in mulberry. Similarly, Khedr et al. (2003) also
reported a decrease in growth and protein contents due to salt stress in the desert plant *Pancratium maritimum* L. Soluble protein contents of leaves were shown to have decreased in response to salinity in *Oryza sativa* L. (Alamgir and Ali, 1999), *Raphanus sativus* (Muthukumarasamy et al., 2000) and *Bruguiera parviflora* (Parida et al., 2002). Total protein content was significantly the highest in callus of *Sesuvium portulacastrum* stressed with 100 μM NaCl compared to control and further decreased with increased salt concentrations (Lokhande et al., 2010). Protein synthesis and lipid metabolism are affected under salt stress, positive antioxidant response might be responsible for a higher tolerance to stress in Carrizo citrange. The photosynthesis, respiration rate and total protein content of plants decreased under salt stress (Yaish, 2015).

In contrast to above, the increase in protein contents under salinity stress was observed with Bekheet et al. (2000) who selected two cultivars of *Asparagus officinalis* by culture shoot segments on callus induction medium supplemented with salt mixture. The cultivars showed better growth, high protein content, fresh and dry weight as salt concentration increased up to 6000 ppm.

**Minerals analysis**

In the current study, the effect of different concentrations of salinity on mineral contents in Hayani cv. was investigated. The results indicated a positive relationship between salinity levels and Na⁺ ion content, while on the other side there was an inverse relation between salinity level and K⁺ ion content.

**Sodium ion content**

In the embryogenic callus, the Na⁺ concentration increased significantly with increasing the concentration of NaCl in the culture medium, especially at the concentrations of 1500 and 2000 ppm NaCl which recorded (4.26 and 4.33 mg/g dw, respectively) compared with the control treatment (3.42 mg/g dw) as shown in Fig. 6.

Na⁺ concentration in the somatic embryos increased significantly with increasing salt concentration in culture medium, as shown in Fig. 6. The increase in Na⁺ concentration was observed when the embryos were cultured on a medium containing high level of salt stress at 2000 ppm NaCl which resulted (4.02 mg/g dw) compared with the control treatment (3.13 mg/g dw).

This increase in Na⁺ is due to its high concentration in the culture medium, which increases its absorption over other essential elements, such as K⁺. This study was confirmed with previous reports on different plant species as safflower (Golkar and Taghizadeh, 2018), sunflower (Alvarez et al., 2003) and *P. ovata* (Golkar et al., 2017) subjected to in vitro salinity stress. The increase in Na⁺ concentration in other date palm cultivars has been reported by Al-Khayri (2002) and also for tomato and grapes under salt stress (El-Hammady et al., 1999). Salinity has a negative effect on growth and development of plants through the osmotic stress due to the toxic effect of sodium ion, besides the ionic imbalance induced by accumulation of this ion (Sairam and Tyagi, 2004). The callus Na⁺ content increased with increasing salt level, whereas callus K⁺ content decreased under the same conditions. The increase in Na⁺ contents is possibly an osmotic adjustment mechanism, which enable the tissues to adapt to the low water potential of the external environment (Sabbah and Tal, 1990).

![Fig. 6. Effect of different concentrations of NaCl on sodium ion content (mg/g dw) in embryogenesis callus (A) and somatic embryos (B) after 8 weeks in culture](image-url)
Sotiropoulos et al. (2006) reported that explants are under stress in two ways under in vitro salinity by the increase of culture media osmotic potential as a result of high Na⁺ content and by Na toxic effects. Also, Rus et al. (2000) studied the rising of Na⁺ concentration in N. sativa calli under salinity stress. High concentrations of sodium ions decreased osmotic balance and membrane structure, reduced growth and inhibited cell division. In addition, high concentration of Na⁺ ion in nutritional solutions destroy hydraulic conductivity and tissue permeability related to water, thus decreasing plant survival (Abadí´a et al., 1999). Such effect on growth is probably due to the effects of salinity, include osmotic effect and ion cytotoxicity; the accumulation of Na⁺ and Cl⁻ in culture cell resulted in the growth inhibition. Furthermore, it has been suggested that the reduction of growth in response to salinity is the result of great portion of respiratory energy being diverted to processes which resulted in salt tolerance (Mass, 1986).

Potassium ion content

K⁺ has an important role in enzyme activation in plant cells (Tester and Davenport, 2003). Fig. 7 shows the effect of NaCl on the potassium concentration in the embryogenic callus and somatic embryos of date palm Hayani cv. It is clear that different concentrations of NaCl significantly decreased the level of K⁺.

The highest concentration of K⁺ (4.35 mg/g dw) recorded in the control treatment; however, with added NaCl at different concentration (500, 1000, 1500 and 2000 ppm) the level of K⁺ decreased gradually (4.22, 4.18, 4.10 and 4.02 mg/g dw, respectively) in embryogenic callus. On the other hand, in the somatic embryos, as shown in Fig. 7 the increase of NaCl concentration at 1500 and 2000 ppm led to a significant decrease in K⁺ concentration ranged from (4.52 mg/g dw) under the control treatment to (4.31 and 4.28 mg/g dw, respectively) under the highest level of salinity.

This reduction in levels of K⁺ is due to excessive accumulation of Na⁺ and Cl⁻, which reduces the accumulation of K⁺. Furthermore, osmotic stress induced by salinity also reduces water uptake and hence accumulation of potassium (Abbas et al., 2012). There is a significant reduction of the concentration of K⁺ in the callus tissues of Citrus sinensis (L.) and increasing the concentration of NaCl in the culture medium. This decrease in K⁺ concentration is due to the presence of Na⁺ at high concentration, which competes with K⁺ uptake and may also block the K⁺ specific transporters of the callus cells under salinity (Zhu, 2003) resulting in toxic levels of Na⁺ as well as insufficient concentration of K⁺ for enzymatic reactions and osmotic adjustment.

The reduction in K⁺ concentration in callus cells under salt stress could be explained by the change in expression and function of transporter as well as the ion channels especially those related to K⁺ (Arzani and Ashraf, 2016). The increase nutritional uptake of both Na⁺ and K⁺ might have led to the retention of water in the callus (Chaudhary et al., 1997).

![Fig. 7. Effect of different concentrations of NaCl on potassium ion content (mg/g dw) in embryogenesis callus (A) and somatic embryos (B) after 8 weeks in culture](image-url)
Increasing the concentration of NaCl in the culture medium generally resulted in increase the Na\(^+\) and reduction in K\(^+\) concentrations. However, at 25 µM NaCl, the only level at which callus growth was significantly enhanced; an increase in K\(^+\) content was noted, in comparison to the NaCl free control (Al-Khayri, 2002).

Na\(^+\) accumulation in tissues under salinity stress is generally considered as a major factor behind the adverse effect of salinity on nutrient uptake and growth (Shibli et al., 2001). The increase in Na\(^+\) content of cells was accompanied by a decrease in K\(^+\) accumulation and differences in Na/K ratio under saline conditions (Cherian and Reddy, 2003).

The somatic embryos cultures after treatment with different level of salinity stress for two months were transferred to optimized regeneration medium without NaCl. The recovery percentage decreased with increase in NaCl concentration. It was observed that regeneration of embryo cultures without salt or with low concentrations of NaCl was better than treated with high concentrations of NaCl (1500 and 2000 ppm). The shoots produced from the experiment of salinity with using low concentration of NaCl were the strongest in growth and ability to continue to rooting stage, as shown in Fig. 8.

Fig. 8. Different stages of somatic embryos to produce the shoots. A: Embryos culture, B: Shoots proliferation from embryos, C: Shoot elongation and rooting stage
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