Total Synthesis of Decahydroquinoline Poison Frog Alkaloids \textit{ent-cis-195A} and \textit{cis-211A}

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1. Introduction

The skin extracts of Neotropical poison frogs contain a variety of lipophilic alkaloids, and over 800 alkaloids have been isolated or detected to date \([1]\). Many of these alkaloids show remarkable biological activities on the nervous system such as nicotinic acetylcholine receptors \([2,3]\). Methods for chemical synthesis of poison frog alkaloids are needed to investigate the biological activities of poison frog alkaloids, as only minute amounts of natural alkaloids can be obtained from skin extracts \([1]\). Decahydroquinolines are a relatively large subgroup of poison frog alkaloids, and over 50 types have been detected. Among them, the alkaloid \textit{cis-195A} is the parent member of this class that was originally isolated from a Panamanian population of \textit{Oophaga (Dendrobates) pumilio} in 1969 \([4]\). The structure and...
absolute configuration of cis-195A were determined by X-ray crystallography, and several total syntheses have also been reported for this compound [5–22]. The alkaloid cis-211A was isolated from skin extracts of the same species of poison frog in 1987 [23]. However, no total synthesis of this alkaloid has been reported, and its absolute configuration remains unknown to date (Figure 1). As part of a program directed at studying the synthesis of poison frog alkaloids [24–38], herein, we report the total synthesis of ent-cis-195A and cis-211A. Both syntheses proceed via the common and key intermediate 11. The synthesis of cis-211A also enabled the determination of its absolute configuration.

**Figure 1.** Structures of cis-195A and cis-211A.

2. Results and Discussion

Hydrogenation of known allyl derivative 1 [39] provided the ester 2, which was converted to enaminoester 4 via thiophenyl derivative 3. The Michael-type conjugate addition reaction to 4 [25] gave adduct 5 as a single isomer in excellent yield. The ester moiety of 5 was elongated by the Arndt–Eistert reaction sequence to afford the homologated ester 6, which was transformed into the methyl ketone 8 via the corresponding Weinreb amide 7. Lemieux–Johnson oxidation of 8 provided the aldehyde 9, which was subjected to cyclization by treatment with DBU in refluxing benzene to yield the cis-fused enone 10c [25] without generating the trans-fused enone 10t. Selective formation of 10c was explained by the preferential formation of conformer A in the starting material 9 owing to the A1,3 strain. Thus, epimerization at the 3-position of 9 occurred first, and then cyclization proceeded to provide the enone 10c as shown in Figure 2. With enone 10c in hand, the stage was set for the divergent synthesis of ent-cis-195A and cis-211A. The conjugate addition reaction to 10c with Me₂CuLi followed by treatment of the resulting enolate with Comins’ triflating agent [40] afforded the common and key intermediate 11 (Scheme 1).

**Figure 2.** Reaction mechanism of the cyclization with epimerization of 9.
Scheme 1. Synthesis of the common and key intermediate enol triflate 11.

Finally, global hydrogenation of 11 and deprotection of the methyl carbamate moiety in 12 using TMSI in CHCl₃ at 50 °C provided ent-cis-195A, as shown in Scheme 2. The ¹H and ¹³C-NMR spectra of synthetic ent-cis-195A were in good agreement with those reported in the literature [22].

Scheme 2. Total synthesis of ent-cis-195A.

The enol triflate 11 was converted to olefin 13 by palladium-catalyzed reduction. Epoxidation of 13 by mCPBA proceeded smoothly to give the epoxide 14, unfortunately, as a 1:1 mixture of epoxide 14. Hydroboration of 13 using BH₃-SMe₂ in toluene provided the alcohol 15 and the mixture of alcohols 16 and 17. The structure of 15 was determined by NOESY. The NOESY experiments of 15 revealed a syn relationship between the methyl group at C-5 and H-6 based on the NOESY correlations from H₃-5 to H-6. However, the separation of 16 and 17 was difficult at this stage. For completion of the synthesis of cis-211A, inversion of the hydroxyl group in 15 was necessary. For this purpose, we subjected 15 to the Mitsunobu reaction; however, all attempts failed and resulted in the recovery of 15. Next, we examined hydroxyl inversion of 15 via ketone 19. Any oxidations of 15 using Swern, a SO₃-pyridine complex, PCC, PDC, DMP, or TPAP were not successful, and the starting material was recovered. Only oxidation using AZADOL® [41] proceeded smoothly...
to yield the desired ketone 19 in good yield. In addition, the AZADOL® oxidation of the mixture of 16 and 17 afforded the ketones 19 and 20 in 34% and 66% yield, respectively, which could be easily separated by column chromatography. Thus, we succeeded to obtain the ketone 19 from 13 in 64% (49% + 15%) overall yield, as shown in Scheme 3. The conformation of ketone 19 is depicted in 19-A. The reduction of 19 from the concave face was needed to obtain the desired alcohol 16. We expected that the reduction of 19 would proceed from the concave face, as shown in 19-A, because of the steric hindrance of the α-axial methyl group. However, the use of a small reducing agent like NaBH₄ or LiAlH₄ reduced 19 from the convex face to afford 15 as the sole product. To secure the reduction from the concave face, we tried the large reducing agent L-Selectride®; however, the reduction did not proceed, and only ketone 19 was recovered. Fortunately, Super-Hydride®, a moderately sized reducing agent, was the best match for this substrate, and the reduction proceeded from the concave face to provide the desired alcohol 16 as the major product (16:15 = 9:1). The final deprotection of the urethane moiety in 16 was also troublesome. First, we applied the same reaction conditions used for ent-cis-195A (TMSI in refluxing CHCl₃) to cleave the methyl carbamate; however, the reaction did not proceed. Then, other reaction conditions, such as the use of n-PrSLi/HMPA or KOH/i-PrOH in a sealed tube at 130 °C, resulted in the recovery of the starting material. Finally, we used TMSI in refluxing MeCN, and the reaction proceeded cleanly to yield cis-211A, as shown in Scheme 3.

Scheme 3. Total synthesis of cis-211A.
The $^1$H- and $^{13}$C-NMR spectra of synthetic cis-211A were in good agreement with the reported values [23]. The absolute stereochemistry of natural cis-211A was determined unambiguously by the present synthesis to be 2R, 4aR, 5R, 6S, and 8aS by comparison of the optical rotation of synthetic cis-211A ([α]$_D^{25}$ −11.5 (c 0.2, CHCl$_3$)) with the reported value ([α]$_D^{19}$ −1.7 (c 1.0, CHCl$_3$)) [23]. Interestingly, cis-195A and cis-211A were both isolated from the same poison frog, Oophaga (Dendrobatidae) pumilio (Dendrobatidae) from Panama; however, the absolute stereochemistry of the parent decahydroquinoline nuclei of cis-195A is opposite to that of cis-211A. The NMR spectra ($^1$H-NMR, $^{13}$C-NMR) of all synthesized compounds are listed in Supplementary Materials.

To further investigate the effect of the stereochemistry of the hydroxyl group at the 6-position on the inhibitory activity against nicotinic acetylcholine (ACh) receptors, we also synthesized 6-epi-211A by deprotection of the methoxycarbonyl group in 15, as shown in Scheme 4.

![Scheme 4. Synthesis of 6-epi-211A.](image_url)

Nicotinic ACh receptors are ligand-gated cation channels [42,43]. Homomeric α7- and heteromeric α4β2-pentamers are the major subtypes of nicotinic receptors found in the central nervous system [44]. It has been reported that (−)-cis-195A, a natural cis-decahydroquinoline alkaloid (formerly referred to as Pumiliotoxin C), blocks ganglionic nicotinic ACh receptors in pheochromocytoma PC12 cells [45] and that the synthetic analog (+)-cis-195A is more potent than (−)-cis-195A at inhibiting nicotinic receptor activity [3]. Here, we examined the effects of ent-cis-195A, cis-211A, and 6-epi-211A on α7- and α4β2-nicotinic ACh receptors ectopically expressed in Xenopus oocytes. If the criterion for partial inhibition by an alkaloid is defined as a ≥ 20% decrease in the peak amplitude of ACh-elicited currents, then ent-cis-195A (1–10 μM) showed no apparent inhibitory effects on α7- and α4β2-receptor-mediated currents (Figure 3A,B). cis-211A and 6-epi-211A at 10 μM partially inhibited α7-receptor-mediated currents by 38% and 31%, respectively, while both alkaloids at 10 μM showed negligible effects on α4β2-receptor-mediated currents (Figure 3C–F). Analysis of their structure–activity relationship suggested that the 6-hydroxy moiety of cis-211A and 6-epi-211A might contribute to the partial blockade of α7-nicotinic ACh receptors. The ligand-binding assays showed that none of these alkaloids affected $[^3]$Hnicotine and $[^3]$Hmethyllycaconitine binding to rat whole brain membranes (data not shown). Therefore, cis-211A and 6-epi-211A were believed to act as noncompetitive blockers of α7-nicotinic receptors, although they were less potent and not as highly selective.

Neuronal nicotinic ACh receptors play a role in cerebral and retinal physiology. The blood–brain barrier (BBB) and inner blood–retinal barrier (BRB) directly segregate the brain and retina, respectively, from the circulating blood. It has been reported that putative nicotinic- and verapamil-sensitive cationic drug transport systems at the BBB and inner BRB, respectively, are involved in the facilitative distribution of their substrates to the central nervous system [46,47]. To evaluate the recognition of ent-cis-195A and cis-211A as substrates for these cationic drug transport systems, we performed an inhibition study using conditionally immortalized rat BBB and inner BRB model cells, known as TR-BBB13 and TR-iBRB2 cells [48,49]. As shown in Table 1, ent-cis-195A exhibited an inhibitory effect on $[^3]$Hnicotine transport into TR-BBB13 cells and $[^3]$Hverapamil transport into TR-iBRB2 cells by more than 40%. In addition, the presence of cis-211A significantly attenuated $[^3]$Hnicotine and $[^3]$Hverapamil uptake by TR-BBB13 and TR-iBRB2 cells, respectively, by at least 29%. These results suggest that ent-cis-195A and cis-211A are recognized by the cationic drug transport systems at the BBB and inner BRB. It is possible that these
derivatives reach the brain and retina via the cationic drug transport systems and show neuronal effects in the CNS.

![Graph showing effects of ent-cis-195A, cis-211A, and 6-epi-211A on α7- and α4β2-nicotinic receptors](image.png)

**Figure 3.** The effects of ent-cis-195A, cis-211A, and 6-epi-211A on α7- and α4β2-nicotinic acetylcholine receptors expressed in Xenopus oocytes. (A–F). Concentration–inhibition curves for ent-cis-195A (A,B), cis-211A (C,D), and 6-epi-211A (E,F) on α7- (A,C,E) and α4β2-nicotinic receptors (B,D,F). Current response to acetylcholine (ACh) in the presence of alkaloid was normalized to the current elicited by ACh alone in the same oocyte and averaged. Values represent the mean ± S.E.M. (A–D): n = 3–5; E: n = 3–4; F: n = 4–5.

**Table 1.** Inhibitory effects of ent-cis-195A and cis-211A on [3H]nicotine uptake by TR-BBB13 cells and [3H]verapamil uptake by TR-iBRB2 cells.

| Conditions | [3H]Nicotine Uptake | [3H]Verapamil Uptake |
|------------|---------------------|----------------------|
| Control    | 100 ± 5             | 100 ± 4              |
| ent-cis-195A | 31.6 ± 1.7*         | 66.9 ± 2.5*          |
| cis-211A   | 60.0 ± 3.5*         | 70.6 ± 4.1*          |

[3H]Nicotine uptake (0.1 μCi/well, 6.0 nM) by TR-BBB13 cells was measured at 37 °C for 10 sec in the absence (control) or presence of test compounds at 200 μM with 1.0% dimethyl sulfoxide (DMSO). Similarly, [3H]verapamil uptake by TR-iBRB2 cells was performed at 37 °C for 3 min. Each value represents the mean ± standard error of the mean S.E.M. (n = 3–6). * p < 0.01, significantly different from the control in Dunnett’s test.

In summary, we achieved the total syntheses of ent-cis-195A and cis-211A in a divergent process from the key and common intermediate 11. The absolute stereochemistry of natural cis-211A was determined to be 2R, 4aR, 5R, 6S, and 8aS by comparison with the data obtained from our total synthesis. The inhibitory effects of ent-cis-195A, cis-211A, and 6-epi-211A on nicotinic ACh receptors were also investigated. The results showed that cis-211A and 6-epi-211A had better inhibitory effects on the α7-receptor than that of ent-cis-195A, and none of the compounds showed inhibitory effects on the α4β2-receptor at the same concentration. These results suggested that cis-211A and 6-epi-211A could be applied as important tools for studying the brain and nervous system. More interestingly, the absolute configuration of the decahydropyridine nuclei of cis-211A was a mirror image of that of cis-195A, even though both alkaloids were isolated from Ophaga (Dendrobates) pumilio from Panama.

3. Materials and Methods
3.1. Chemistry
3.1.1. General Information

Chemicals were purchased from Sigma–Aldrich, Merck (Darmstadt, Germany), FUJIFILM Wako Chemicals (Osaka, JAPAN), Nacalai Tesque, Tokyo Chemical Industry (Tokyo, Japan), and Kanto Chemical (Tokyo, Japan) and used without further purification. Col-
uum chromatography was done on Cica silica gel 60N (spherical, neutral; particle size, 63–210 nm, Kanto Chemical), while thin-layer chromatography was performed using Merck silica gel 60F<sub>254</sub> plates. Melting points were taken on a Yanaco micromelting point apparatus and are uncorrected. The nuclear magnetic resonance (NMR) spectra were acquired in the specified solvent in JEOL JNM-A400 (400 and 100 MHz for <sup>1</sup>H and <sup>13</sup>C, respectively) or JEOL JNM-ECX500 (500 and 125 MHz for <sup>1</sup>H and <sup>13</sup>C, respectively). The chemical shifts (δ) are reported in ppm downfield from TMS, and coupling constants (J) are expressed in Hertz. IR spectra were measured with a JASCO FT/IR-460 Plus spectrophotometer (JASCO Corp., Tokyo, Japan). The low-resolution and high-resolution mass spectra were obtained with a Shimadzu GCMS-QP 500 mass spectrometer (Shimadzu Corp., Kyoto, Japan), JEOL D-200, or JEOL AX505 mass spectrometer (JEOL Ltd., Tokyo, Japan) in the electron impact mode at the ionization potential of 70 eV.

3.1.2. Synthesis of (6R)-2-Phenylsulfonyl-6-propyl-piperidine-1,2-dicarboxylic Acid Dimethyl Ester (3)

To a stirred solution of 1 [39] (2.51 g, 10.39 mmol) in EtOAc (30 mL) was added 10% Pd/C (30 mg), and the resulting mixture was hydrogenated at 1 atm for 16 h. The catalyst was removed through a celite pad and washed with EtOAc (5 mL × 3). The filtrate and washings were combined and evaporated to give 2, which was essentially pure and used directly in the next step. To a stirred solution of 2 in THF (30 mL) was added a solution of sodium bis(trimethylsilyl)amide (1.9 M in THF, 8.20 mL, 15.59 mmol) at −78 °C, and the reaction mixture was stirred at −78 °C for 30 min. To the reaction mixture was added a solution of diphenyl disulfide (3.40 g, 15.59 mmol) in THF (15 mL), and the resulting mixture was stirred at 0 °C for 30 min. The solvent was evaporated, and the residue was chromatographed on SiO<sub>2</sub> (50 g, acetone/n-hexane = 1/30) to give 3 (3.39 g, 9.66 mmol, 93% in 2 steps) as a yellow oil as a mixture of diastereomers.

<sup>1</sup>H-NMR (400 MHz CDCl<sub>3</sub>) δ: 0.90 and 0.94 (3H, each t, J = 7.2 Hz), 1.28–1.79 (8H, m), 3.49 and 3.62 (3H, each s), 3.70 (3H, s), 4.42 (1H, br), 6.06 (1H, t, J = 3.6 Hz); <sup>13</sup>C-NMR (125 MHz CDCl<sub>3</sub>) δ: 13.75, 19.10, 19.53, 25.70, 31.62, 50.99, 51.87, 52.85, 129.87, 154.63, 165.56; IR (neat): 1231, 1275, 1330, 1442, 1536, 3063, 2927, 1714, 1733 cm<sup>−1</sup>; MS (EI): m/z 241 (M<sup>+</sup>); HRMS (EI) Calcd for C<sub>12</sub>H<sub>15</sub>NO<sub>4</sub> 241.1314 (M<sup>+</sup>); Found 241.1315; [α]D<sup>19</sup> +68.0 (c 1.00, CHCl<sub>3</sub>).

3.1.3. Synthesis of (6R)-6-Propyl-5,6-dihydro-4H-pyridine-1,2-dicarboxylic Acid Dimethyl Ester (4)

To a stirred solution of 3 (1.27 g, 3.61 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (12 mL) was added 2,6-lutidine (0.84 mL, 9.03 mmol), and then mCPBA (70%, 1.50 g, 8.67 mmol) was added to the reaction mixture in four portions in 15 min intervals at 0 °C. The resulting mixture was stirred at room temperature for 8 h. The reaction was quenched with 10% Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub> in sat. NaHCO<sub>3</sub> (aq.) (25 mL), and the aqueous mixture was diluted with EtOAc. The layers were separated, and the aqueous layer was extracted with EtOAc (5 mL × 3). The organic layer and extracts were combined and washed with brine, 10% HCl (aq.), and brine, successively. The organic layer was dried and evaporated to give a pale yellow oil, which was chromatographed on SiO<sub>2</sub> (20 g, acetone/n-hexane = 1/30) to give 4 (871 mg, 3.61 mmol, 100%) as pale yellow oil.

<sup>1</sup>H-NMR (400 MHz CDCl<sub>3</sub>) δ: 0.93 (3H, t, J = 7.3 Hz), 1.17–1.28 (1H, m), 1.37–1.58 (3H, m), 1.69–1.76 (1H, m), 1.79–1.89 (1H, m), 2.15–2.22 (2H, m), 3.70 (3H, s), 3.76 (3H, s), 4.42 (1H, br), 6.06 (1H, t, J = 3.6 Hz); <sup>13</sup>C-NMR (125 MHz CDCl<sub>3</sub>) δ: 13.75, 19.10, 19.53, 25.70, 31.62, 50.99, 51.87, 52.85, 129.87, 154.63, 165.56; IR (neat): 1231, 1275, 1330, 1442, 1714, 1733 cm<sup>−1</sup>; MS (EI): m/z 241 (M<sup>+</sup>); HRMS (EI) Calcd for C<sub>12</sub>H<sub>15</sub>NO<sub>4</sub> 241.1314 (M<sup>+</sup>); Found 241.1315; [α]D<sup>19</sup> +68.0 (c 1.00, CHCl<sub>3</sub>).

3.1.4. Synthesis of (2R, 3S, 6R)-6-Propyl-3-vinyl-piperidine-1,2-dicarboxylic Acid Dimethyl ester (5)

To a stirred solution of Cul (1.31 g, 6.90 mmol) in Et<sub>2</sub>O (15 mL) was added a solution of vinyl lithium, prepared from tetravinyltin (0.61 mL, 3.45 mmol) and MeLi (1.13 M in Et<sub>2</sub>O, 12.20 mL, 13.80 mmol) in Et<sub>2</sub>O (15 mL) at 0 °C for 30 min, at −78 °C, and the reaction mixture was warmed to -35 °C for 30 min. The reaction mixture was recooled to −78 °C,
and a solution of 4 (555 mg, 2.30 mmol) in Et2O (7 mL) was added to the reaction mixture. The resulting mixture was gradually warmed to 0 °C and stirred at the same temperature for 1 h. The reaction was quenched with sat. NH4Cl (aq.) (30 mL). The aqueous mixture was diluted with CH2Cl2 (30 mL), and the resulting mixture was filtered. The filtrate was separated, and the aqueous layer was extracted with CH2Cl2 (10 mL × 3). The organic layer and extracts were combined, dried, and evaporated to give a colorless oil, which was chromatographed on SiO2 (20 g, acetone/r-hexane = 1/30) to give 5 (613 mg, 2.28 mmol, 99%) as a colorless oil.

1H-NMR (400 MHz CDCl3) δ: 0.90 (3H, t, J = 7.0 Hz), 1.25–1.56 (6H, m), 1.78–1.92 (2H, m), 3.08 (1H, br), 3.71 (3H, s), 3.74 (3H, s), 4.17–4.18 (1H, m), 4.88 (1H, br), 5.09–5.15 (2H, m), 5.81 (1H, ddd, J = 17.1, 10.7, 6.4 Hz); 13C-NMR (125 MHz CDCl3) δ: 13.86, 19.88, 21.03, 22.45, 34.59, 36.52, 50.96, 51.99, 52.75, 55.07, 115.16, 139.00, 157.08, 172.86; IR (neat): 1200, 1340, 1448, 1506, 1558, 1683, 1699, 1734 cm⁻¹; MS (EI): m/z 269 (M⁺); HRMS (EI) Calcd for C14H23NO4 269.1627 (M⁺); Found 269.1631; [α]D25 +53.6 (c 1.00, CHCl3).

3.1.5. Synthesis of (2S, 3S, 6R)-2-Methoxycarbonylmethyl-6-propyl-3-vinyl-piperidine-1-carboxylic Acid Methyl Ester (6)

To a stirred solution of 5 (428 mg, 1.59 mmol) in MeOH (6 mL) and H2O (2 mL) was added LiOH·H2O (266 mg, 6.36 mmol), and the resulting mixture was refluxed for 2 h. After cooling, MeOH was evaporated, and the residue was acidified with 10% HCl (aq.) (5 mL). The aqueous mixture was extracted with EtOAc (3 mL × 5). The organic extracts were combined, dried, and evaporated to give a yellow oil, which was used directly in the next step. To a stirred solution of the above oil in THF (10 mL) were added ClCO2Et (0.18 mL, 1.91 mmol) and Et3N (0.27 mL, 1.91 mmol) at 0 °C, and the resulting mixture was stirred at 0 °C for 1 h. The reaction mixture was diluted with Et2O (3 mL), and Et3N·HCl was filtered off. The filtrate was evaporated to give a yellow oil, which was used directly in the next step. To a stirred solution of the above oil in Et2O (10 mL) was added a solution of CH2N2 in Et2O at 0 °C, and the reaction mixture was stirred at room temperature for 16 h. The solvent was evaporated to give a yellow oil, which was dissolved in MeOH (10 mL). To the MeOH solution were added AgCO2Ph (37 mg, 0.16 mmol) and Et3N (0.45 mL, 3.18 mmol), and the resulting mixture was stirred at room temperature for 24 h. The reaction mixture was diluted with Et2O, and the insoluble material was filtered off. The filtrate was evaporated to give a black oil, which was chromatographed on SiO2 (20 g, acetone/r-hexane = 1/30) to give 6 (409 mg, 1.45 mmol, 91% in 4 steps) as a colorless oil.

1H-NMR (400 MHz CDCl3) δ: 0.92 (3H, t, J = 7.3 Hz), 1.20-1.42 (4H, m), 1.43–1.52 (2H, m), 1.78–1.92 (2H, m), 2.32 (1H, br), 2.54 (1H, dd, J = 14.9, 4.8 Hz), 2.65 (1H, dd, J = 14.9, 10.1 Hz), 3.63 (3H, s), 3.68 (3H, s), 4.12 (1H, br), 4.61 (1H, br), 5.06 (1H, dt, J = 10.6, 1.4 Hz), 5.09 (1H, dt, J = 17.2, 1.4 Hz), 5.84 (1H, ddd, J = 17.2, 10.6, 6.6 Hz); 13C-NMR (125 MHz CDCl3) δ: 13.96, 20.05, 20.32, 22.18, 37.46, 39.73, 39.90, 50.73, 50.92, 51.67, 52.62, 115.07, 140.04, 156.76, 171.64; IR (neat): 1101, 1363, 1443, 1696, 1740 cm⁻¹; MS (EI): m/z 283 (M⁺); HRMS (EI) Calcd for C15H25NNO4 283.1784 (M⁺); Found 269.1780; [α]D19 +31.4 (c 1.00, CHCl3).
on SiO$_2$ (10 g, acetone/$n$-hexane = 1/7) to give 7 (610 mg, 1.95 mmol, 97% in 2 steps) as a colorless oil.

$^1$H-NMR (500 MHz CDCl$_3$) $\delta$: 0.91 (3H, t, $J = 7.3$ Hz), 1.16-1.43 (4H, m), 1.48 (2H, q, $J = 6.0$ Hz), 1.76-1.92 (2H, m), 2.37 (1H, br), 2.53-2.56 (1H, m), 2.80 (1H, m), 3.12 (3H, br), 3.65 (3H, s), 3.67 (3H, s), 4.12 (1H, br), 4.63 (1H, br), 5.04 (1H, dd, $J = 10.7$, 14.6 Hz), 5.07 (1H, dd, $J = 17.2$, 1.4 Hz), 5.84 (1H, dd, $J = 17.2$, 10.7, 1.4 Hz); $^{13}$C-NMR (125 MHz CDCl$_3$) $\delta$: 13.96, 19.85, 20.26, 22.15, 29.20, 32.12, 37.36, 39.15, 50.20, 50.55, 52.51, 61.22, 114.82, 140.30, 156.75, 172.01; IR (neat): 1102, 1277, 1361, 1407, 1443, 1640, 1694 cm$^{-1}$; MS (EI): $m/z$ 312 (M$^+$); HRMS (EI) Calcd for C$_{16}$H$_{26}$N$_2$O$_4$ 312.2049 (M$^+$); Found 312.2046; $[\alpha]_D^{23}$ $-$36.0 (c 1.00, CHCl$_3$).

3.1.7. Synthesis of (2S, 3S, 6R)-2-(2-Oxo-propyl)-6-propyl-piperidine-1-carboxylic Acid Methyl Ester (8)

To a stirred solution of 7 (188 mg, 0.60 mmol) in THF (3 mL) was added a solution MeMgBr (0.91 M in THF, 0.97 mL, 0.72 mmol) at $0^\circ$C, and the resulting mixture was stirred at $0^\circ$C for 1 h. The reaction was quenched with sat. NH$_4$Cl (aq.) (5 mL). The layers were separated, and the aqueous layer was extracted with CH$_2$Cl$_2$ (5 mL × 3). The organic extract and extracts were combined, dried, and evaporated to give a colorless oil, which was chromatographed on SiO$_2$ (7 g, acetone/$n$-hexane = 1/7) to give 8 (160 mg, 0.60 mmol, 99%) as a colorless oil.

$^1$H-NMR (500 MHz CDCl$_3$) $\delta$: 0.93 (3H, t, $J = 7.3$ Hz), 1.19-1.62 (6H, m), 1.79-1.91 (2H, m), 2.18 (3H, s), 2.24 (1H, br), 2.59-2.63 (1H, dd, $J = 12.0$, 2.8 Hz), 2.70-2.79 (1H, dd, $J = 12.0$, 8.4 Hz), 3.69 (3H, s), 4.12 (1H, br), 4.62-4.64 (1H, m), 5.07 (1H, dd, $J = 10.6$, 1.5 Hz), 5.09 (1H, dd, $J = 17.2$, 1.5 Hz), 5.86 (1H, dd, $J = 17.2$, 10.6, 1.5 Hz); $^{13}$C-NMR (125 MHz CDCl$_3$) $\delta$: 13.97, 19.82, 20.29, 22.00, 29.86, 37.30, 39.39, 49.44, 50.00, 50.26, 52.56, 115.08, 140.07, 156.74, 206.60; IR (neat): 1102, 1277, 1361, 1407, 1443, 1640, 1694 cm$^{-1}$; MS (EI): $m/z$ 267 (M$^+$); HRMS (EI) Calcd for C$_{15}$H$_{25}$NO$_3$ 267.1834 (M$^+$); Found 267.1835; $[\alpha]_D^{19}$ $-$70.0 (c 1.00, CHCl$_3$).

3.1.8. Synthesis of (2S, 3S, 6R)-3-Formyl-2-(2-oxo-propyl)-6-propyl-piperidine-1-carboxylic Acid Methyl Ester (9)

To a stirred solution of 8 (368 mg, 1.38 mmol) in 1,4-dioxane (6 mL) and H$_2$O (2 mL) was added 2,6-lutidine (0.32 mL, 2.75 mmol), OsO$_4$ (2% aqueous solution, 1.7 mL, 0.14 mmol) and NaI$_2$O (1.18 g, 5.51 mmol) at $0^\circ$C, and the resulting mixture was stirred at room temperature for 3 h. The reaction was quenched with 10% Na$_2$S$_2$O$_3$ in sat. NaHCO$_3$ (aq.) (10 mL), and the aqueous mixture was diluted with CH$_2$Cl$_2$. The layers were separated, and the aqueous layer was extracted with CH$_2$Cl$_2$ (5 mL × 3). The organic layer and extracts were combined; washed with brine, 10% HCl (aq.), and brine, successively; dried; and evaporated to give a yellow oil, which was chromatographed on SiO$_2$ (10 g, acetone/$n$-hexane = 1/10) to give 9 (349 mg, 1.29 mmol, 94%) as a colorless oil.

$^1$H-NMR (500 MHz CDCl$_3$) $\delta$: 0.90 (3H, t, $J = 7.3$ Hz), 1.16-1.26 (1H, m), 1.26-1.36 (1H, m), 1.38-1.57 (4H, m), 1.74-1.82 (1H, m), 1.93-1.99 (1H, m), 2.17 (3H, s), 2.37 (1H, br), 2.70-2.80 (2H, m), 3.67 (3H, s), 4.06 (1H, br), 5.15 (1H, br), 9.66 (1H, s); $^{13}$C-NMR (125 MHz CDCl$_3$) $\delta$: 13.94, 14.37, 20.22, 23.39, 30.15, 36.61, 45.10, 48.19, 48.90, 49.93, 52.74, 156.29, 202.79, 206.29; IR (neat): 1100, 1328, 1354, 1447, 1684, 1694, 1717, 2873, 2957 cm$^{-1}$; MS (EI): $m/z$ 269 (M$^+$); HRMS (EI) Calcd for C$_{14}$H$_{23}$NO$_4$ 269.1627 (M$^+$); Found 269.1629; $[\alpha]_D^{23}$ $-$114.8 (c 1.00, CHCl$_3$).

3.1.9. Synthesis of (2R, 4aR, 8aS)-7-Oxo-2-propyl-3,4,4a,7,8,8a-hexahydro-2H-quinoline-1-carboxylic Acid Methyl Ester (10e)

To a stirred solution of 9 (349 mg, 1.30 mmol) in benzene (30 mL) was added DBU (0.78 mL, 5.18 mmol) and MS 4 A (50 mg), and the resulting mixture was refluxed for 48 h. After cooling, benzene was evaporated, and the residue was acidified with 10% HCl (aq.) (5 mL). The aqueous mixture was extracted with EtOAc (3mL × 5). The organic extracts
were combined, dried, and evaporated to give a brown oil, which was chromatographed on SiO\(_2\) (25 g, EtOAc/\(n\)-hexane = 1/10) to give \(10c\) (235 mg, 0.94 mmol, 72%) as a yellow oil.

\(^1\)H-NMR (500 MHz CDCl\(_3\)) \(\delta\): 0.91 (3H, t, \(J = 7.2\) Hz), 1.24–1.41 (2H, m), 1.44–1.55 (2H, m), 1.59 (1H, td, \(J = 10.0, 2.4\) Hz), 1.67 (1H, tdd, \(J = 10.0, 4.8, 2.4\) Hz), 1.73–1.78 (1H, m), 1.78–1.83 (1H, m), 2.40–2.45 (1H, m), 2.61 (2H, br), 3.70 (3H, s), 4.25 (1H, br), 4.63 (1H, br), 6.13 (1H, d, \(J = 9.7\) Hz), 6.77 (1H, dd, \(J = 9.7, 5.7\) Hz); \(^1^3\)C-NMR (125 MHz CDCl\(_3\)) \(\delta\): 13.88, 19.95, 20.24, 27.09, 36.53, 37.00, 40.30, 48.43, 49.73, 52.60, 128.64, 152.18, 156.04, 156.39, 218.22; IR (neat): 771, 1089, 1115, 1246, 1275, 1314, 1444, 1685, 2934 cm\(^{-1}\).

3.1.10. Synthesis of (2\(R\), 4a\(R\), 5\(R\), 8a\(R\))-5-Methyl-2-propyl-7-trifluoromethane-sulfonyloxy-3,4,4a,5,8,8a-hexahydro-2H-quinoline-1-carboxylic Acid Methyl Ester (11)

To a stirred solution of Cul (147 mg, 0.77 mmol) in Et\(_2\)O (3 mL) was added a solution of MeLi (1.17 M in Et\(_2\)O, 3.12 mL, 1.54 mmol) at –78 \(^\circ\)C, and the reaction mixture was warmed to 0 \(^\circ\)C for 30 min. The reaction mixture was recooled to –78 \(^\circ\)C, and a solution of \(10c\) (97 mg, 0.39 mmol) in Et\(_2\)O (3 mL) was added to the reaction mixture. The reaction mixture was gradually warmed to 0 \(^\circ\)C, and then a solution of Comins’ reagent (303 mg, 0.77 mmol) in Et\(_2\)O (3 mL) was added to the reaction mixture. The resulting mixture was stirred at room temperature for 2 h, and the reaction was quenched with sat. NH\(_4\)Cl (aq.) (5 mL). The aqueous mixture was diluted with CH\(_2\)Cl\(_2\) (5 mL), and the resulting suspension was filtered. The filtrate was separated, and the aqueous layer was extracted with CH\(_2\)Cl\(_2\) (3 mL × 3). The filtrate and extracts were combined, dried, and evaporated to give a colorless oil, which was chromatographed on SiO\(_2\) (10 g, acetone/\(n\)-hexane = 1/50) to give 11 (144 mg, 0.36 mmol, 94%) as a colorless oil.

\(^1\)H-NMR (400 MHz CDCl\(_3\)) \(\delta\): 0.90 (3H, t, \(J = 7.2\) Hz), 1.12 (3H, d, \(J = 7.2\) Hz), 1.19–1.39 (4H, m), 1.39–1.50 (1H, m), 1.50–1.72 (4H, m), 2.26 (1H, t, \(J = 6.0\) Hz), 2.50 (2H, br), 3.69 (3H, s), 4.10 (1H, br), 4.50 (1H, br), 5.66 (1H, d, \(J = 6.0\) Hz); \(^1^3\)C-NMR (100 MHz CDCl\(_3\)) \(\delta\): 13.83, 19.15, 20.68, 21.49, 27.88, 29.67, 35.23, 38.04, 40.26, 46.91, 50.41, 52.63, 118.45 (q, \(J = 318.5\) Hz), 121.19, 144.70, 156.65; IR (neat): 1303, 1317, 1443, 1695, 2864, 2929, 2955 cm\(^{-1}\); MS (EI): \(m/z\) 399 (M\(^+\)); HRMS (EI) Calcd for C\(_{16}\)H\(_{24}\)F\(_3\)NO\(_5\)S 399.1327 (M\(^+\)); Found 399.1337; [\(\alpha\)]\(_D\)^23 +58.6 (c 1.35, CHCl\(_3\)).

3.1.11. Synthesis of (2\(R\), 4a\(R\), 5\(R\), 8a\(R\))-5-Methyl-2-propyl-octahydro-quinoline-1-carboxylic Acid Methyl Ester (12)

To a stirred solution of 11 (60 mg, 0.15 mmol) in MeOH (3 mL) was added 20% Pd(OH)\(_2\)/C (5 mg), and the resulting mixture was hydrogenated at 1 atm for 16 h. The catalyst was removed through a celite pad and washed with MeOH (3 mL × 3). The filtrate and washings were combined and evaporated to give a pale yellow oil, which was chromatographed on SiO\(_2\) (8 g, acetone/\(n\)-hexane = 1/30) to give 12 (33 mg, 0.13 mmol, 88%) as a colorless oil.

\(^1\)H-NMR (500 MHz CDCl\(_3\)) \(\delta\): 0.90 (3H, t, \(J = 7.2\) Hz), 1.06 (3H, d, \(J = 6.9\) Hz), 1.17–1.68 (14H, m), 1.79–1.88 (2H, m), 3.70 (3H, s), 4.06 (1H, br), 4.22 (1H, br); \(^1^3\)C-NMR (125 MHz CDCl\(_3\)) \(\delta\): 14.17, 19.41, 20.41, 20.72, 21.36, 26.82, 28.19, 28.57, 34.61, 37.81, 42.19, 48.91, 50.49, 52.36, 156.65; IR (neat): 1303, 1317, 1443, 1695, 2864, 2929, 2955 cm\(^{-1}\); MS (EI) \(m/z\) 253 (M\(^+\)); HRMS (EI) Calcd for C\(_{15}\)H\(_{27}\)NO\(_5\) 253.2042 (M\(^+\)); Found 253.2043; [\(\alpha\)]\(_D\)^23 -20.2 (c 1.00, CHCl\(_3\)).

3.1.12. Synthesis of (2\(R\), 4a\(R\), 5\(R\), 8a\(R\))-5-Methyl-2-propyldecahydroquinoline (ent-cis-195A)

To a stirred solution of 12 (42 mg, 0.17 mmol) in CHCl\(_3\) (3 mL) was added NaI (197 mg, 1.32 mmol) and TMSCl (0.10 mL, 0.83 mmol), and the resulting mixture was heated to 50 \(^\circ\)C for 24 h. After cooling, the reaction was quenched with 10% Na\(_2\)S\(_2\)O\(_3\) in sat. NaHCO\(_3\) (aq.) (3 mL), and aqueous mixture was extracted with CH\(_2\)Cl\(_2\) (2 mL × 10).
The organic extracts were combined, dried, and evaporated to give a pale yellow oil, which was chromatographed on SiO2 (3 g, MeOH/CH2Cl2 = 1/20) to give ent-cis-195A (30 mg, 0.15 mmol, 88%) as a pale yellow oil. ent-cis-195A·HCl was obtained in quantitative yield by treatment with HCl (ca. 1 mol/L in Et2O) followed by evaporation.

ent-cis-195A: 1H-NMR (400 MHz CDCl3) δ: 0.83 (3H, d, J = 6.6 Hz), 0.90 (3H, t, J = 7.0 Hz), 0.95–1.02 (1H, m), 1.03–1.14 (2H, m), 1.25–1.49 (8H, m), 1.52–1.70 (4H, m), 1.79–1.90 (1H, m), 1.90–1.97 (1H, m), 2.53 (1H, dt, J = 11.4, 5.8, 2.8 Hz), 2.84 (1H, q, J = 2.8 Hz); 13C-NMR (100 MHz CDCl3) δ: 14.30, 19.14, 19.91, 21.21, 26.99, 27.24, 27.35, 33.28, 35.88, 39.59, 42.49, 55.96, 57.72; IR (neat): 857, 890, 962, 1024, 1044, 1081, 1124, 1167, 1256, 1317, 1348, 1380, 1451, 1734, 2803, 2873, 2935 cm−1; MS (EI) m/z 195 (M+); HRMS (EI) Calcd for C13H25N O2 195.1987 (M+); Found 195.1985; ent-cis-195A·HCl: [α]D20 +12.7 (c 0.35, MeOH).

The 1H- and 13C-NMR spectra and optical rotation of the synthetic sample were identical with those of the literature data.

1H-NMR (500 MHz CDCl3) δ: 0.83 (3H, d, J = 6.6 Hz), 0.90 (3H, t, J = 7.0 Hz), 0.94–1.03 (1H, m), 1.05–1.14 (2H, m), 1.22–1.49 (9H, m), 1.77–1.90 (1H, m), 1.90–1.99 (1H, m), 2.53 (1H, dt, J = 11.4, 5.8, 2.7 Hz), 2.84 (1H, q, J = 2.8 Hz); 13C-NMR (75 MHz CDCl3) δ: 14.5 (CH3), 19.3 (CH3), 20.1 (CH3), 21.4 (CH2), 27.2 (CH2), 27.54 (CH), 27.55 (CH2), 33.6 (CH2), 36.1 (CH2), 39.9 (CH2), 42.8 (CH), 56.1 (CH), 57.9 (CH); ent-cis-195A·HCl: [α]D20 +12.9 (c 0.36, MeOH).[22]

3.1.13. Synthesis of (2R, 4aR, 5R, 8aR)-5-Methyl-2-propyl-3,4,4a,5,8,8a-hexahydro-2H-quinoline-1-carboxylic Acid Methyl Ester (13)

To a stirred solution of 11 (144 mg, 0.36 mmol) in THF (3 mL) was added PPh3 (7 mg, 0.03 mmol) and Pd(OAc)2 (3 mg, 0.01 mmol) at room temperature, and the reaction mixture was refluxed for 18 h. The reaction was quenched with brine (5 mL). The layers were separated, and the aqueous layer was extracted with CH2Cl2 (3 mL × 3). The organic layer and extracts were combined, dried, and evaporated to give a colorless oil, which was chromatographed on SiO2 (8 g, acetone/n-hexane = 1/10) to give 13 (85 mg, 0.36 mmol, 100%) as a colorless oil.

1H-NMR (400 MHz CDCl3) δ: 0.89 (3H, t, J = 7.2 Hz), 1.05 (3H, d, J = 6.4 Hz), 1.23–1.66 (9H, m), 2.03 (1H, m), 2.15 (2H, m), 3.67 (3H, s), 4.07 (1H, br), 4.35 (1H, br), 5.44–5.53 (2H, m); 13C-NMR (100 MHz CDCl3) δ: 14.02, 20.77, 21.96, 22.42, 27.34, 28.21, 29.67, 37.03, 41.33, 47.11, 50.53, 52.33, 122.82, 130.53, 156.39; IR (neat): 1093, 1304, 1320, 1444, 1695, 2871, 2930, 2955 cm−1; MS (EI) m/z 251 (M+); HRMS (EI) Calcd for C15H25NO2 251.1885 (M+); Found 251.1877; [α]D20 +57.4 (c 0.90, CHCl3).

3.1.14. Synthesis of (2R, 4aR, 5R, 6R, 8aR)-6-Hydroxy-5-methyl-2-propyl-octahydro-quinoline-1-carboxylic Acid Methyl Ester (15)

To a stirred solution of 13 (18 mg, 0.08 mmol) in toluene (1.5 mL) was added BH3·SMe2 (0.02 mL, 0.23 mmol) at 0 °C. The reaction mixture was stirred at room temperature for 24 h and then cautiously quenched with 10% NaOH (0.25 mL) at 0 °C, followed by the slow addition of H2O2 (30%, 0.25 mL) at 0 °C. The reaction mixture was stirred at room temperature for 24 h. The layers were separated, and the aqueous layer was extracted with CH2Cl2 (3 mL × 3). The organic layer and extracts were combined, dried, and evaporated to give a colorless oil, which was chromatographed on SiO2 (5 g, acetone/n-hexane = 1/10) to give 15 (10 mg, 0.04 mmol, 50%) as a colorless oil and the inseparable mixture of alcohols 16 and 17 (9 mg, 0.03 mmol, 45%) as a colorless oil.

15: 1H-NMR (400 MHz CDCl3) δ: 0.91 (3H, t, J = 7.6 Hz), 1.05 (3H, d, J = 7.6 Hz), 1.20–1.26 (1H, m), 1.28–1.36 (2H, m), 1.46–1.68 (7H, m), 1.78 (1H, qdd, J = 14.3, 3.9, 2.8 Hz), 1.88 (1H, qd, J = 7.6, 1.7 Hz), 2.01 (1H, qd, J = 13.2, 3.9 Hz), 2.28 (1H, qd, J = 13.6, 3.2 Hz), 3.67 (3H, s), 4.02–4.10 (1H, m), 4.18–4.25 (1H, m); 13C-NMR (100 MHz CDCl3) δ: 14.05, 18.89, 20.56, 22.77, 28.35, 28.64, 29.68, 37.33, 41.32, 41.67, 49.86, 50.51, 52.32, 70.97, 156.69; IR (neat):
1319, 1447, 1456, 1670, 1697, 2872, 2934, 2957 cm⁻¹; MS (EI) m/z 269 (M⁺); HRMS (EI) Calcd for C₁₅H₂₇NO₂ 269.1991 (M⁺); Found 269.1990; [α]D²⁵ -17.1 (c 0.80, CHCl₃).

3.1.15. Synthesis of (2R, 4aR, 5R, 8aR)-5-Methyl-6-oxo-2-propyl-octahydro-quinoline-1-carboxylic Acid Methyl Ester (19)

To a stirred solution of 15 (46 mg, 0.17 mmol) in CH₂Cl₂ (1.5 mL) and sat. NaHCO₃ (aq.) (1.5 mL) was added NaOCl 5H₂O (55 mg, 0.34 mmol) and AZADOL® (0.3 mg, 0.0017 mmol) at 0 °C, and the resulting mixture was stirred at room temperature for 1 h. The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (1 mL × 3). The organic layer and extracts were combined, dried, and evaporated to give a yellow oil, which was chromatographed on SiO₂ (8 g, acetone/n-hexane = 1/5) to give 19 (43 mg, 0.16 mmol, 98%) as pale yellow oil.

1H-NMR (400 MHz CDCl₃): δ 0.88 (3H, t, J = 6.8 Hz), 1.24 (3H, d, J = 7.6 Hz), 1.26–1.69 (8H, m), 1.76–1.84 (1H, m), 1.92–2.00 (1H, m), 2.06 (1H, qd, J = 12.8, 4.8 Hz), 2.16–2.22 (1H, m), 2.26-2.33 (1H, m), 2.60 (1H, td, J = 14.8, 6.8 Hz), 3.69 (3H, s), 4.05–4.12 (1H, m), 4.58–4.61 (1H, m); 13C-NMR (100 MHz CDCl₃): δ 13.99, 17.20, 20.53, 21.91, 27.40, 27.53, 29.68, 36.94, 37.45, 43.75, 48.89, 50.12, 50.45, 52.58, 156.57, 213.93; IR (neat): 1093, 1133, 1189, 1244, 1275, 1312, 1348, 1409, 1444, 1699, 1717, 2855, 2929, 2954 cm⁻¹; MS (EI) m/z 267 (M⁺); HRMS (EI) Calcd for C₁₃H₂₅NO₂ 267.1834 (M⁺); Found 267.1834; [α]D²⁵ -14.4 (c 0.50, CHCl₃).

3.1.16. Synthesis of (2R, 4aR, 5R, 6S, 8aR)-6-Hydroxy-5-methyl-2-propyl-octahydro-quinoline-1-carboxylic Acid Methyl Ester (16)

To a stirred solution of 19 (18 mg, 0.07 mmol) in THF (2 mL) was added a solution of Super-Hydride® (1.0 M in THF, 0.20 mmol, 0.20 mmol) at 0 °C, and the resulting mixture was stirred at room temperature for 1 h. The reaction was quenched with sat. NH₄Cl (aq.) (2 mL). The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (1 mL × 3). The organic layer and extracts were combined, dried, and evaporated to give a colorless oil, which was chromatographed on SiO₂ (5 g, acetone/n-hexane = 1/5) to give 16 (16 mg, 0.06 mmol, 89%) as a colorless oil and 15 (2 mg, 0.01 mmol, 11%) as a colorless oil.

1H-NMR (400 MHz CDCl₃): δ 0.91 (3H, t, J = 7.2 Hz), 1.06 (3H, d, J = 7.2 Hz), 1.23–1.71 (13H, m), 1.83 (1H, qd, J = 13.6, 2.8 Hz), 1.99 (1H, t, J = 6.0 Hz), 3.68 (3H, s), 4.02–4.11 (1H, m), 4.19–4.29 (1H, m); 13C-NMR (100 MHz CDCl₃): δ 12.22, 14.03, 20.58, 21.03, 28.12, 28.64, 29.68, 37.58, 40.53, 43.06, 48.75, 50.33, 52.38, 68.66, 156.63; IR (neat): 1097, 1319, 1448, 1670, 2870, 2932, 2955 cm⁻¹; MS (EI) m/z 269 (M⁺); HRMS (EI) Calcd for C₁₅H₂₇NO₂ 269.1991 (M⁺); Found 269.1990; [α]D²⁵ -16.5 (c 0.50, CHCl₃).

3.1.17. Synthesis of (2R, 4aR, 5R, 6S, 8aR)-5-Methyl-2-propyldecahydroquinoline-6-ol (cis-211A)

To a stirred solution of 16 (15 mg, 0.06 mmol) in MeCN (2 mL) was added NaI (67 mg, 0.45 mmol) and TMSCl (0.04 mL, 0.28 mmol), and the resulting mixture was refluxed for 1 h. After cooling, the reaction was quenched with 10% Na₂S₂O₃ in sat. NaHCO₃ (aq.) (2 mL). The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (1 mL × 10). The organic layer and extracts were combined, dried, and evaporated to give a pale yellow oil, which was chromatographed on SiO₂ (3 g, MeOH/CH₂Cl₂ = 1/5) to give cis-211A (13 mg, 0.06 mmol, 100%) as pale yellow oil.

1H-NMR (400 MHz CDCl₃): δ 0.90 (3H, t, J = 6.4 Hz), 0.95 (3H, d, J = 6.8 Hz), 1.10–1.18 (2H, m), 1.27–1.45 (6H, m), 1.54–1.66 (3H, m), 1.80–1.87 (1H, m), 1.91–2.03 (2H, m), 2.06–2.15 (1H, m), 2.53–2.62 (1H, m), 2.89 (1H, br), 3.83 (1H, br); 13C-NMR (100 MHz CDCl₃): δ 14.22, 15.82, 19.09, 26.37, 26.45, 28.08, 29.68, 31.02, 34.55, 39.09, 55.94, 57.95, 72.11; IR (neat): 753, 812, 883, 946, 997, 1029, 1100, 1158, 1191, 1257, 1317, 1339, 1376, 1444, 2806, 2879, 2934, 3659 cm⁻¹; MS (EI) m/z 211 (M⁺); HRMS (EI) Calcd for C₁₃H₂₅NO 211.1936 (M⁺); Found 211.1943; [α]D²⁵ -11.5 (c 1.00, CHCl₃).

The 1H- and 13C-NMR spectra and optical rotation of the synthetic sample were identical with those of the literature data.
H-NMR (400 MHz CDCl₃) δ: 0.90 (3H, t), 0.95 (3H, d), 1.06–1.16 (2H, m), 1.29–1.43 (6H, m), 1.56–1.67 (3H, m), 1.81–1.87 (1H, m), 1.91–2.03 (2H, m), 2.06–2.15 (1H, m), 2.53–2.61 (1H, m), 2.88 (1H, br); 13C-NMR (100 MHz CDCl₃) δ: 14.3, 15.8, 19.1, 26.7, 26.8, 26.9, 28.9, 31.1, 34.9, 39.3, 55.8, 57.8, 72.3; [α]D²⁵ -11.7 (c 1.00, CHCl₃) [23].

3.1.18. Synthesis of (2R, 4aR, 5R, 6R, 8aR)-5-Methyl-2-propyldecahydroquinoline-6-ol (6-epi-211A)

To a stirred solution of 15 (18 mg, 0.07 mmol) in MeCN (2 mL) was added NaI (80 mg, 0.54 mmol) and TMSCl (0.04 mL, 0.33 mmol), and the resulting mixture was refluxed for 1 h. After cooling, the reaction was quenched with 10% Na₂S₂O₃ in sat. NaHCO₃ (aq.) (2 mL). The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (1 mL × 10). The organic layer and extracts were combined, separated, and evaporated to give a pale yellow oil, which was chromatographed on SiO₂ (3 g, MeOH/CH₂Cl₂ = 1/5) to give 6-epi-211A (10 mg, 0.05 mmol, 72%) as pale yellow oil.

H-NMR (400 MHz CDCl₃) δ: 0.95 (3H, t, J = 7.2 Hz), 1.03–1.05 (1H, m), 1.07 (3H, d, J = 6.0 Hz), 1.31–1.60 (5H, m), 1.68–2.08 (4H, m), 2.14–2.20 (1H, m), 2.36–2.40 (1H, m), 3.11 (1H, m), 3.29 (1H, br), 3.34 (1H, td, J = 9.2, 3.6 Hz); 13C-NMR (100 MHz CDCl₃) δ: 13.84, 15.11, 19.01, 23.91, 25.14, 27.48, 28.89, 35.47, 36.00, 39.18, 57.06, 59.69, 74.85; IR (neat): 1375, 1456, 1636, 1647, 2866, 2926, 2955, 3647 cm⁻¹; MS (EI) m/z 211 (M⁺); HRMS (EI) Calcd for C₁₃H₂₅NO: 211.1936 (M⁺); Found 211.1928; [α]D²⁰ −5.3 (c 0.4, CHCl₃)

3.2. Electrophysiological Recording of Nicotinic ACh Receptor-Mediated Current in Xenopus Oocytes

Xenopus oocytes expressing recombinant mouse α7- and α4β2-nicotinic ACh receptors were prepared by injection of the plasmid containing respective subunit cDNAs (provided by Dr. J. A. Stitzel, University of Michigan Medical Center), according to the protocols described previously [29]. The oocytes were cultured at 19 °C for 3 to 6 days in 50% Leibovitz’s L-15 Medium (11415064, Thermo Fisher Scientific, Waltham, MA, USA, pH 7.5) containing 1 µg/mL insulin and 100 µg/mL gentamicin (078-06061, Fujifilm Wako Pure Chemical Corp., Osaka, Japan). Two-electrode voltage-clamp recordings were then performed, as described previously [29]. In brief, an oocyte was placed in a 300 µL tube-like chamber where Ringer solution (82.5 mM NaCl, 2.5 mM KCl, 2.5 mM CaCl₂, 1 mM MgCl₂, and 5 mM HEPES, pH 7.4) containing 1 µg/mL atropine (A0257, Sigma–Aldrich, MA, USA) was perfused at a rate of 15 mL/min. Membrane potential was held at −60 mV, and currents were measured using a GeneClamp 500 amplifier, Digidata1322A, and pClamp9 software (Axon Instruments, Union City, CA, USA). The oocyte was pretreated with an alkaloid for 3 min and then treated with ACh (011-00592, Fujifilm Wako Pure Chemical Corp., Osaka, Japan). To analyze the effects of alkaloids, current response to ACh in the presence of alkaloid was normalized to control response (i.e., current elicited by ACh (100 µM for α7 and 1 µM for α4β2) alone) in each oocyte and then averaged.

3.3. Ligand-Binding Assays

The [³H]nicotine and [³H]methyllycaconitine binding assays using membrane suspensions from whole rat brain (excluding cortex and cerebellum) were performed as described previously [50].

3.4. In Vitro Effect of the Compound on the Transport of Cationic Compounds at the BBB and Inner BRB

As model cells of the rat BBB and inner BRB, TR-BBB and TR-iBRB2 cells were utilized [48,49]. These cells were cultured following the previous manuscript [51] and seeded onto collagen type I-coated 24-well plate (Corning, Kennebunk, ME, USA) at a density of 5 × 10⁴ cells/cm². The cells were cultured for 2 days and then washed with
extracellular fluid buffer (122 mM NaCl, 25 mM NaHCO₃, 3 mM KCl, 1.4 mM CaCl₂, 1.2 mM MgSO₄, 0.4 mM K₂HPO₄, 10 mM D-glucose, 10 mM 2-[4-(2-hydroxyethyl)-1-piperazinyl]ethanesulfonic acid-NaOH, pH 7.4) at 37 °C. The extracellular fluid buffer containing ³H]-nicotine (85 Ci/mmol; American Radiolabeled Chemicals, St. Louis, MO, USA) or ³H]-verapamil (80 Ci/mm; American Radiolabeled Chemicals) at a concentration of 0.5 µCi/mL in the absence or presence of test compounds at 200 µM with 1.0% DMSO was applied to the cells and incubated at 37 °C for designed time (³H]-nicotine, 10 sec; ³H]-verapamil, 3 min). After the uptake reaction, the cells were rinsed with the extracellular fluid buffer at 4 °C and solubilized in 1 N NaOH. The solubilized solution was neutralized with 1 N HCl. The ³H-radioactivities derived from the cell-solubilized solution and reaction buffer were measured using an AccuFLEX LSC-7400 liquid scintillation counter (Hitachi, Kashiwa, Japan). Protein concentration in the solubilized solution was quantified by a DC protein assay kit II (BIO-RAD, Hercules, CA, USA). The uptake activities were normalized by the concentration of the radiolabeled compound in the transport buffer and the cellular protein amount in each well. The data were expressed as the percentage of uptake activity in the control group and mean ± S.E.M. Statistic difference was evaluated using a one-way analysis of variance followed by Dunnett’s test.

Supplementary Materials: The NMR spectra (¹H-NMR, ¹³C-NMR) of all new compounds are available online.

Author Contributions: T.O., K.T. and J.I. performed the experiments of the synthesis of ent-cis-195A and cis-211A and analyzed synthetic data; H.M., T.I. and T.K. determined the stereochemistry of the intermediate 15; N.W. performed the electrophysiological experiments under supervision by H.T., T.W., T.S. (Toshiyasu Sasaoka), T.S. (Takahiro Shimizu) and H.S. and H.T. partly wrote the corresponding parts; L.P.D. performed the binding assays; Y.Y., S.-i.A., Y.K. and K.-i.H. performed the experiments on nicotine and verapamil transport and partly wrote the corresponding parts; S.R.H. and R.A.S. partly wrote the introduction parts; N.T. conceived the need to synthesize ent-cis-195A and cis-211A and designed the experiment; T.O. and N.T. wrote the paper. All authors have read and agreed to the published version of the manuscript.

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Sample Availability: Samples of cis-195A and cis-211A are available from the authors.

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