Therapeutic Applications of Coconut Water Mitigates Diabetes Mellitus in Oryctolagus cuniculus (Rabbit)

Shahab Uddin1, Imdad Ali1, Mazhar Hassan1, Laila Hamayun1, Uroba Ali1, Shah Saud1, Ibrahim Khan1, Waqas Ahmad1, Abdul Bari Shah2, Amir Zeb*3 and Ayaz Ali Khan*1

1Department of Biotechnology, University of Malakand, Pakistan
2Department of Chemistry, University of Malakand, Pakistan
3Department of Biotechnology, Quaid-i-Azam University, Pakistan
*These authors contributed equally to this work
*Corresponding author: Ayaz Ali Khan, Department of Biotechnology, Charkdara, Dir (L), Khyber Pakhtunkhwa, Pakistan

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ABSTRACT

Diabetes mellitus (DM) is a pandemic disease and causes significant contribution to human mortality across the globe. Since, natural products have been proved to be safe and effective remedies in human healthcare practice; therefore, the present study has investigated the anti-diabetic potential of coconut water against the diabetic rabbits. Rabbits were intoxicated with streptozotocin (80 mg/kg body weight) and diabetes mellitus was induced. To counteract initial hypoglycemia, 10% of glucose was administrated in drinking water for 24 hours. One group was kept as negative control, another was diabetic and un-medicated control, two groups were fed with coconut water dose 2.2g/kg and 4.1g/kg, and last group was medicated with Metformin. Several analyses such as body weight, physical activities, urination, and different biochemical properties and characterization of blood samples were performed. The intoxicated rabbits were characterized by weight loss, frequent urination, sluggish physical activities, low level of high-density lipoprotein, and elevated levels of blood glucose, cholesterol, and triglyceride. Upon administration of coconut water at a dose rate of 2.2 g/kg and 4.1g/kg for two weeks, all the parameters became normal and/or near to normal when compared to positive control. Therefore, we recommend that coconut water can be developed as a nutraceutical or functional food for the management of diabetes and its associated complications.

Abbreviations: SGOT: Serum the Level of Glutamic-Oxaloacetic Transaminase; SGPT: Serum Glutamic Pyruvic Transaminase; DM: Diabetes Mellitus; WHO: World Health Organization; OAAs: Oral Anti-Diabetic Agents; HDL: High Density Lipoprotein

Introduction

Diabetes mellitus (DM) is an epidemic disease and causes severe problem to human health [1]. By definition, diabetes mellitus is a group of metabolic imbalances, results in hyperglycemia, which is either caused by abnormal production of insulin or its harmful physiology [2]. Diabetic patients are at more risk to cardiac, peripheral arterial and cerebrovascular diseases [2,3]. The chronic hyperglycemia of diabetes is associated with long-term damage, dysfunction, and failure of various organs including eyes, kidneys, nerves, heart, and blood vessels [4]. DM also disturbs function of liver; therefore, in blood serum the level of glutamic-oxaloacetic transaminase (SGOT) and Serum Glutamic Pyruvic Transaminase (SGPT) increases [5]. There is usually persistent thirst, polyuria, blurred vision, and weight loss followed by ketoadiosis or non-ketotic hyperosmolar condition that leads to stupor, coma and, if untreated causes death [6]. In 1993, the World Health Organization (WHO) Diabetes reporting group published global estimates for the prevalence of diabetes from 75 communities in 32 countries [7].

Worldwide, the prevalence rate of obesity, diabetes, and other metabolic syndromes is high [8,9]. There have been several previous estimates of the number of persons with diabetes. Prevalence of diabetes among adults (age 20–79 years) will increase to 7.7% (439 million) of the world population by 2030. This estimation is
alarmingly high for developing countries where diabetes patients will increase to 69% by 2030 [10]. According to World Health Organization (WHO), diabetes will be the 7th leading cause of death in 2030 [11]. Currently, Pakistan occupies 7th position of high-risk diabetes-suffered countries [12]. The current prevalence of type 2 diabetes (T2D) in Pakistan is 11.77% [13]. Furthermore, according to National Survey (2016-17), 26% of Pakistan population is suffering from diabetes. Therefore, prevention of diabetes mellitus needs serious attention in health policy. By far the most popular approach to treat T2D is glycemic control in an attempt to reduce complications and death. In the past, several drugs for T2D including oral anti-diabetic agents (OADs), insulin, and incretin-based drugs have been developed.

For instance, sulfonylurea-type drugs, insulin sensitizers such as TZDs (e.g., rosiglitazone and pioglitazone) and a biguanide, metformin, can directly lower insulin resistance and, subsequently, blood glucose. Glucose (re)absorption is viewed as an alternative way to lower blood glucose level. Inhibitors of SGLT 2 and glucosidase such as dapagliflozin, emagliflozin, and a carboxylic acid, inhibit the activity of sodium-glucose cotransporter-2 and glucosidases, respectively. Such inhibition leads to decrease glucose (re)absorption via the renal tubules and the intestine, respectively [14,15]. The available anti-diabetic drugs have limited rules and cause serious side effects. For instance, metformin [16], pioglitazone [17], and SGLT-2 inhibitors [18] cause lactic acidosis, edema and heart failure, and dehydration, urinary and genital infections, respectively. Lowering blood glucose and lipid profiles by using natural products of plant origin as a possible therapeutic measure has become a promising scientific investigation. It has been reported that essential oils may be effective against type 2 diabetes mellitus. The daily consumption of olive oil had positive effect on fasting blood glucose and lipid profiles of healthy controls [19].

Experimental studies have shown that Hibiscus rosasinensis Linn. ethanol flower extract possesses positive effect on blood glucose and lipid profile in streptozotocin induced diabetes in rats [20]. Aloe vera gel extract has hypoglycemic, hypotriglyceremic properties in STZ-diabetic rats [21]. The natural Egyptian Morus alba root bark extracts protective effects in diabetes by decreasing glucose level, moderate lipid peroxidation, and preservation of pancreatic β cell integrity [22]. CW is a rich source of essential metabolites including sugars, sugar alcohols, vitamin C, folate acid, free amino acids, phytohormones (auxin, cytokinin), enzymes (acid phosphatase, catalase, dehydrogenase, diastase, peroxidase, RNA-polymersases) and growth promoting factors [23]. Quantitative difference in the composition of the active components available in coconut water are 95.3 % water; 0.005% nitrogen; 0.56% Phosphoric acid; 0.25% Potassium; 0.69% Calcium oxide; 0.59% Magnesium oxide; 0.5% Iron; 0.8% reducing sugar and a total sugar of 2.08% [24]. Since, the coconut water has several medicinal properties such as hypcholesterolemic, antihypertensive, cardio protective, hepatoprotective, and hypolipidemic properties [25,26]. Therefore, the current study has evaluated the therapeutic potential of CW against the diabetic rabbits.

Materials and Methods

Selection of Animals

Oryctolagus cuniculus (Rabbit) was selected as experimental animals. Rabbits were purchased from local markets and divided into several groups on the basis of their body weights.

Induction of Diabetes Mellitus (DM)

Streptozotocin was used for the induction of diabetes mellitus in rabbits. The rabbits were made diabetic by injecting streptozotocin via peritoneum route [27]. Streptozotocin dose was administered body weight dependent. To counteract initial hypoglycemia, glucose was given orally, also 10% glucose was provided in drinking water for next 24 hours [28].

Drug Administration

a. Group A was medicated with coconut water at the dose rate of 2.2g/kg body weight.
b. Group B was medicated with coconut water at the dose rate of 4.1g/kg body weight.
c. Group C was kept as null control without any treatment.
d. Group D was kept as negative control (diabetic but unmedicated).
e. Group E was considered as positive control and medicated with Glucophage (Metformin HCl) at the dose rate of 14 mg/kg body weight.
f. Drug administration was continued for regular 15 days after confirmation of diabetes mellitus.

Collection of Blood Samples

Blood samples were collected from all the groups with the following schedule: Day zero, day 03, day 08, day 13, and day 18. Blood samples were collected, stored in slant position and serum was oozed out upon clotting. Serum was collected in falcon tubes and was analyzed for glucose, triglyceride, cholesterol, HDL and LDL levels.

Analysis of biochemical parameters. Blood samples were analyzed for different parameters using their respective standard available kits (Semi Auto Chemistry Analyzer).

Statistical Analysis

The results obtained were analyzed statistically. Mean and standard deviation were determined. All the values of different groups were compared at different days using ANOVA. Online software, Prism, and Demo v05 were used for statistical analysis (www.graphpad.com).
Results

Induction of Diabetes Mellitus

All the rabbits of group A, B, D and E were intoxicated with streptozotocin (80mg/kg). Diabetes mellitus was confirmed on day 3rd of the experiment. Our results suggested that diabetic-induced rabbits lost their body weight (Table 1). Two groups (A and B) were treated with coconut water at dose rate of 2.2g/kg and 4.1g/kg, while, one group (E) was medicated with 14mg/kg metformin HCl. Our results observed significant increase (P<0.0001) in the body weight of diabetic treated groups (Table 1). Upon treatment of diabetic rabbits with CW, it was observed that the treated rabbits were gaining body weight (Table 1). Conversely, significant decrease (P<0.0001) was observed in the body weight of positive control (Table 1). Our results observed that that coconut water and metformin increase the body weight in diabetic groups (Table 1).

Table 1: Time-scale analysis of body weight of rabbits (n=3).

| Groups | Medication       | Weight of rabbits in gram (n=3) (M±SD) | Days  |
|--------|------------------|--------------------------------------|-------|
| A      | CW* 2.2g/kg      | 563±3.79 553±3.21 543±3.51 544±2.08 547±1.00 554±2.65 561±3.06 574±2.52 |       |
| B      | CW 4.1g/kg       | 532±1.53 519±1.00 507±2.08 510±3.21 513±2.65 520±3.00 533±3.21 542±2.08 |       |
| C      | NC               | 540±2.52 542±1.00 540±2.08 540±3.00 540±2.52 543±2.08 540±3.61 541±2.65 |       |
| D      | PCΨ 14mg/kg      | 569±3.08 560±3.21 557±1.00 549±2.00 539±2.08 534±2.65 530±1.53 523±2.52 |       |
| E      | Meformin 14 mg/kg| 550±1.53 548±2.65 537±3.21 537±2.00 541±2.52 550±1.53 551±2.65 556±1.00 |       |

Note: #M: Mean
$SD$: Standard deviation
*CW: Coconut water
NC: Negative control
ΨPC: Positive control

Analysis of Blood Glucose Level

Blood glucose level of all rabbits was analyzed prior to exposure to STZ. The glucose levels were 160±2.52, 161±2.08, 161±2.61, and 161±1.53 mg/dl of group A, B, C, D, and E, respectively (Figures 1 & 2). After intoxication with STZ (80mg/kg), diabetes was further confirmed by blood glucose analysis. Blood glucose levels were found 287±3.00, 290±3.21, 269±3.61, and 260±2.00 mg/dl for group A, B, D, and E, respectively (Figures 1 & 2). Our results observed elevated glucose level in all the diabetic rabbits as compared to negative control (160±1.53mg/dl). Our findings suggested that administration of CW to diabetic groups at dose rate of 2.2g/kg and 4.1g/kg for eight days decreased the blood glucose level to 254±3.61 and 247±2.00 mg/dl, respectively. Moreover, the glucose level of metformin-treated group dropped down to 229±1.53 mg/dl. Furthermore, on day 13th and 18th, further decrease was observed in the glucose level of CW-treated groups A and B to 188±2.00 and 187±2.00 mg/dl and 163±1.53 and 162±1.53 mg/dl, respectively (Figures 1 & 2). Metformin also decreased the glucose level significantly. Hence, our findings demonstrated a significant difference (P<0.0001) in the blood glucose level of CW-treated and -untreated diabetic rabbits.

![Figure 1: Blood glucose level of rabbits medicated with CW at dose rate of 2.2 g/kg.](image-url)
Figure 2: Blood glucose level of rabbits medicated with CW at dose rate of 4.1 g/kg.

Analysis of Blood Cholesterol Level

Before induction of diabetes, blood cholesterol level was determined and observed that group A, B, C, D, and E has blood cholesterol level of 55.0±2.65, 55.7±2.52, 57.7±2.08, 57.3±3.79, and 54.7±2.52 mg/dl, respectively (Figures 3A & 4A). Our results observed that upon induction of diabetes, cholesterol level was increased to 81.7±3.79, 80.3±3.21, 81.2±2.65, and 80.0±1.00 mg/dl as compared to negative control 62.0±2.65 mg/dl (Figures 3A & 4A) which show significant difference in all diabetic groups. Thereafter, the diabetic group A and B, were treated with CW at dose rate of 2.2 g/kg and 4.1 g/kg, and group E were treated with metformin 14 mg/kg. Our analysis of blood samples on day 8th observed that cholesterol level was reduced to 74.7±2.08, 73.0±2.00 and 70.7±2.08 mg/dl in medicated group. Moreover, further decrease was observed at day 13th to 68.3±1.53 and 66.3±1.53 in the cholesterol level of CW-treated groups A and B and in metformin-administrated group 65.0±2.00 mg/dl. Further reduction was noted at day 18th in the cholesterol level to 58.0±1.00, 57.0±1.00, and 56.3±3.06 mg/dl of CW-treated groups A and B and metformin-administrated group. Therefore, we suggested that CW at the dose rate of 2.2 and 4.1 g/kg leads to decrease in cholesterol level (Figures 3A & 4A).

Figure 3: Analysis of blood of rabbits medicated with CW at dose rate of 2.2 g/kg.
**Analysis of Triglyceride Level in Blood**

Measurement of blood triglyceride (TG) level of healthy rabbits observed 62.0±2.00, 61.3±3.21, 60.3±2.52, 63.0±3.61, and 62.7±2.08 mg/dl in group A, B, C, D, and E, respectively (Figures 3B & 4B). Analysis of STZ-induced diabetic rabbits confirmed that blood TG level was increased as compared to control. At day 3rd, the TG level of blood samples of diabetic rabbits increased to 90.3±3.21, 91.0±2.65, 76.3±3.21, and 95.0±3.61 mg/dl for groups A, B, D, and E, as compared to negative control 62.3±3.06 mg/dl for group C, respectively (Figures 3B & 4B). Treating diabetic rabbits, A and B with CW at the dose rate of 2.2 g/kg and 4.1 g/kg, and group E were treated with metformin at 14 mg/kg, significantly reduced TG level to 80.3±2.52, 79.3±1.53, and 78.3±3.06 mg/dl as compared to positive control 87.0±3.00 mg/dl at day 8th. With continuous treatment, TG levels were reduced to 74.3±2.08, 73.0±3.61 and 72.0±2.00 mg/dl as compared to positive control 96.7±2.52 mg/dl after blood analysis at day 13th. At day 18th, further reduction was noted in TG level (64.7±2.08, 64.0±1.00 and 63.7±1.53 mg/dl) as compared to the day 13th as shown in Figures 3B & 4B. Therefore, it is concluded that CW at the dose rate of 2.2 and 4.1 g/kg decreases TG level.

**Evaluation of High-Density Lipoprotein (HDL) in blood**

Prior to diabetes induction, the blood high density lipoprotein (HDL) level of all the groups were measured as 31.3±1.53, 29.3±2.08, 30.3±1.53, 31.0±2.65, and 31.3±2.52 mg/dl for groups A, B, C, D, and E, respectively (Figures 3C & 4C). At day 3rd of diabetes induction, the HDL level was reduced to 20.3±2.08, 20.0±2.00, 22.7±2.08, and 20.7±2.08 mg/dl in diabetic group A, B, D, and E. The blood HDL level of negative control C (31.0±2.00 mg/dl) remained unchanged. Our results demonstrated that CW-treated rabbits (dose rate of 2.2 g/kg and 4.1 g/kg) showed elevated level of blood HDL of 18.7±2.52 and 17.7±2.52 mg/dl for diabetic groups at the dose rate of 2.2 g/kg and 4.1 g/kg, respectively (Figures 3C & 4C). Consistently, the metformin-treated groups showed 23.0±1.00 and 18.0±2.00 mg/dl HDL concentration at day 13 and 18, respectively (Figures 3C & 4C). So, we recommend that CW increases the blood HDL level of diabetic rabbits (Figures 3C & 4C).

**Evaluation of Low-Density Lipoprotein (LDL) in Blood**

The healthy samples showed normal concentration of low-density lipoprotein (LDL) and obtained 20.0±3.00, 20.7±2.08, 21.0±2.65, 20.0±2.00, 21.7±2.52 mg/dl for groups A, B, C, D, and E. Further increase was observed in LDL level in the groups treated either with CW or metformin. At day 13th and 18th, the LDL level was increased to 23.0±1.00 and 24.0±1.00, and 28.0±2.00 and 28.7±1.53 for diabetic groups treated with CW at the dose rate of 2.2 g/kg and 4.1 g/kg, respectively (Figures 3C & 4C). Consistently, the metformin-treated groups showed 23.0±1.00 and 28.0±2.00 mg/dl LDL concentration at day 13 and 18, respectively (Figures 3C & 4C). So, we recommend that CW increases the blood LDL level of diabetic rabbits (Figures 3C & 4C).
E, respectively. After induction of diabetes at day 3rd, the LDL level was increased to 28.0±1.00, 30.0±1.00, 28.3±2.08, and 31.0±2.65 mg/dl for groups A, B, D, and E, respectively (Figures 3D & 4D). Upon treatment with CW at the dose rate of 2.2g/kg and 4.1g/kg, group A and B showed slightly low LDL level of 29.3±1.53, 31.0±2.65, respectively. In parallel, metformin-treated diabetic group (E) at the dose rate of 14 mg/kg obtained 32.7±2.31 mg/dl. At day 8th, LDL level were decreased to 29.3±1.53, 31.0±2.65, 32.7±2.31 mg/dl, as compared to positive control 33.0±1.00 mg/dl. At day 13th, LDL levels were reduced in CW as well as Metformin-treated groups to 24.3±2.52, 22.7±2.52, and 22.7±2.08 mg/dl, respectively. At day 18th, further decrease was recorded in LDL values in both the CW as well as Metformin-treated groups to 21.3±2.08, 20.0±2.00, and 19.7±3.79 mg/dl, respectively. Conversely, the LDL level remained higher for positive control (40.0±3.00 mg/dl) and lower for negative control (21.3±2.08 mg/dl) as shown in Figure 3D & 4D. So, it is concluded that CW at the dose rate of 2.2g/kg and 4.1g/kg decreases LDL level in diabetic rabbits (Figures 3D & 4D).

Discussion

Diabetes mellitus (DM) can mainly be attributed to the sedentary lifestyle and calorie-rich diet. DM is linked with abnormal lipid metabolism and is considered as a major factor for the development of atherosclerosis and cardiovascular complication [29]. STZ causes hyperglycemia along with hyperglycemic condition [21,30,31]. Previously, it was investigated that STZ causes diabetes by rapid depletion of beta-cells, which leads to reduction of insulin secretion [32]. In rats, intravenous injection of STZ (70 mg/kg of body weight) induces diabetes mellitus [33]. Moreover, Mozaffari et al., induced DM in rats by injecting STZ at dose rate of 90 mg/kg of the body weight [31]. In present study, STZ was used as DM inducer at the dose rate of 80 mg/kg body weight. Our results confirmed that STZ-administrated rabbits lost their body weights, which is the ultimate symptom of diabetic rabbits [34-36]. After treatment with CW at dose rate of 2.2 g/kg and 4.1 g/kg, an increase occurred in the body weight of diabetic rabbits. Our results are in strong agreement of Preetha et al. and Saat et al. where they observed an increase the body weights of virgin coconut oil-treated diabetic rabbits [37,38].

Additionally, administration of STZ 80 mg/kg increased the glucose level of STZ-administrated rabbits. Several reports have investigated that STZ administration increases blood glucose level upon reduction of insulin secretion [34,39,40]. Our study follows Aly et al. when they induced hyperglycemia by injecting STZ at a dose rate of 65 mg/kg body weight [41]. In another study, STZ increased blood glucose level when injected at a dose rate of 60 mg/kg body weight [42]. Our results demonstrated that CW at the dose rate of 2.2 g/kg and 4.1 g/kg decreased blood glucose level of diabetic rabbits. Preetha et al. investigated that Albizia myriophylla and virgin coconut oil have significant hypoglycemic potential [37]. Additionally, a couple of studies have reported that mature coconut water and coconut kernel protein reduced the blood glucose level of diabetic rats [38,43]. Therefore, our study recommends that coconut water decreased the blood glucose level of diabetic rabbits. Our results observed elevated level of cholesterol in STZ-induced diabetic rabbits. Our results are in strong agreement that STZ accumulates high concentration of cholesterol diabetic rats [41,44].

We observed that CW reduced the blood cholesterol level of diabetic rabbits. Previously, palm kernel oil and coconut oil were used to reduce the total cholesterol levels of diabetic rats [45,46]. Our study also observed that diabetic rabbits were symptomized by elevated levels of triglycerides. Increase in triglyceride concentration is an essential indication of diabetic subjects [41,44,47]. Our study confirmed that CW caused a significant reduction in the concentration of triglyceride level of diabetic rabbits. Other studies have also reported that coconut oil and coconut water reduces triglyceride level and have anti-diabetic activities [45,46,48]. STZ administration decreases HDL levels in animals [30,44]. Our study observed that CW increased HDL level in CW-treated rabbits. Same results were also documented by several other reports [48-50]. Finally, the present study investigated that STZ increased the LDL level STZ-intoxicated rabbits. STZ has been reported to cause a significant increase in LDL concentration which leads to DM [41,44,51]. Our study investigated that CW reduces LDL level in diabetic rabbits. Other studies have also observed that coconut water was given to diabetic rats and same reduction in LDL level were found [48,52]. In another study, the synergistic effects of coconut oil and olive oil has demonstrated to reduce the LDL level [53]. Overall, our study demonstrated that CW has anti-diabetic potential and could be used as a nutraceutical or functional food for the management of diabetes and its associated complications.

Conclusion

STZ induced a characteristic multiphasic immediate response in rabbits. There was frequent urination, decreased physical activities and sluggishness in comparison to negative control. In addition, marked individual variations in response exist in terms of onset and severity of glycemic changes. The body weights were reduced in diabetic rabbits. There was also increase in cholesterol, triglyceride and low-density lipoprotein values in all the intoxicated rabbits with low values of high-density lipoprotein. After treatment with coconut water as well as metformin for two weeks, all the parameters became normal or near to normal when compared to positive control. Form the results of our study prove that coconut water is effective against experimentally diabetic. The present work demonstrates that both concentrations are capable of improving the diabetic complications. Conversely, coconut water showed an adverse effect in relation to the diabetic complications.

References

1. Narayan KV (2005) The diabetes pandemic: looking for the silver lining. Clinical diabetes 23: 51-53
2. Alberti KGMM, Zimmet P (1998) Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus. Provisional report of a WHO consultation. Diabetic medicine 15(7): 539-553.
3. Fox CS, Goady S, Sorlie PD, D'Agostino RB, Pencina MJ, et al. (2007) Increasing cardiovascular disease burden due to diabetes mellitus the Framingham Heart Study. Circulation 115: 1544-1550.

4. Gavin III JR, Alberti K, Davidson MB, De Fronzo RA (1997) Report of the expert committee on the diagnosis and classification of diabetes mellitus. Diabetes care 20(7): 1183-1197.

5. Chatila R, West BA (1996) Hepatomegaly and abnormal liver tests due to glycogenosis in adults with diabetes. Medicine 75(6): 327-333.

6. Mc Cane D, Hanson RL, Charles MA, Jacobsen LT, Pettitt D, et al. (1994) Comparison of tests for glycated haemoglobin and fasting and two-hour plasma glucose concentrations as diagnostic methods for diabetes. Bmj 308(6940): 1233-1238.

7. King H, Rewers M (1993) Global estimates for prevalence of diabetes mellitus and impaired glucose tolerance in adults. Diabetes care 16(1): 157-177.

8. Boyle JP, Honeycutt AA, Narayan KV, Hoerger TJ, Geiss LS, et al. (2001) Projection of diabetes burden through 2050 impact of changing demography and disease prevalence in the US. Diabetes care 24(11): 1936-1940.

9. Benjamin SM, Valder R, Geiss LS, Rolka DB, Narayan KV (2003) Estimated Number of Adults With Prediabetes in the US in 2000 Opportunities for prevention. Diabetes care 26(3): 645-649.

10. Shaw JE, Sircue RA, Zimmet PZ (2010) Global estimates of the prevalence of diabetes for 2010 and 2030. Diabetes research and clinical practice 87(1): 1-14.

11. Mathers CD, Loncar D (2006) Projections of global mortality and burden of disease from 2002 to 2030. Plos med 3(1): e44-2.

12. Khan MA, Sultan SM, Nazli R, Akhtar T, Khan MA, et al. (2014) Depression among patients with type-II diabetes mellitus. J Coll Physicians Surg Pak 24(10): 770-771.

13. Meo SA, Zia I, Bakhari IA, Arain SA (2016) Type 2 diabetes mellitus in Pakistan: Current prevalence and future forecast. JPMA. The Journal of the Pakistan Medical Association 66(12): 1637-1642.

14. Jones D (2011) Diabetes field cautiously upbeat despite possible setback for leading SGLT2 inhibitor. Nature Publishing Group.

15. Egan JM, Burotta A, Hui H, Perfetti R (2003) GIP-1 receptor agonists are growth and differentiation factors for pancreatic islet beta cells. Diabetes/metabolism research and reviews 19(2): 115-123.

16. Kim MJ, Han JY, Shin JY, Kim SL, Lee JM, et al. (2015) Metformin-associated lactic acidosis: predisposing factors and outcome. Endocrinology and Metabolism 30(3): 78-83.

17. Mundaliar M, FACE Sunder Chang M, Anna R, Henry M, Robert R (2003) Thiazolidinediones, peripheral edema, and type 2 diabetes: incidence, pathophysiology, and clinical implications. Endocrine Practice 9(5): 406-416.

18. Vasilaikou D, Karagiannis T, Athanasiadou E, Mainou M, Liakos A, et al. (2013) Sodium–glucose co-transporter 2 inhibitors for type 2 diabetes: a systematic review and meta-analysis. Annals of internal medicine 159(4): 262-274.

19. Al Jamal AR, Ibrahim A (2011) Effects of olive oil on lipid profiles and blood glucose in type 2 diabetic patients. Int J Diabetes Metab 19: 19-22.

20. Sachdeva A, Khemani L (2003) Effect of Hibiscus rosa sinensis Linn. ethano I flower extract on blood glucose and lipid profile in streptozotocin induced diabetes in rats. Journal of Ethnopharmacology 89(1): 61-66.

21. Rajasekaran S, Ravi K, Sivagnanam K, Subramanian S (2006) Beneficial effects of Ake vera leaf gel extract on lipid profile status in rats with streptozotocin diabetes. Clinical and Experimental Pharmacology and Physiology 33(3): 232-237.

22. Singab ANB, El Beshbishy HA, Yonelkawa M, Nomura T, Fukai T (2005) Hypoglycemic effect of Egyptian Morus alba mot bark extract: effect on diabetes and lipid peroxidation of streptozotocin-induced diabetic rats. Journal of ethnopharmacology 100(3): 333-338.

23. Yong JW, Lee Y, Ng YE, Tan SN (2009) The chemical composition and biological properties of coconut (Cocos nucifera L.) water. Molecules 14(12): 5144-5164.

24. Nwanwga BK (2012) The reno-protective effects of coconut water on the kidneys of diabetic wistar rats. Journal of Health Science 21(1): 1-4.

25. Bhagya D, Prema L, Ramajahan T (2010) Beneficial effects of tender coconut water on blood pressure and lipid levels in experimental hypertension. Journal of cell and tissue research 10: 2139.

26. Pratapahan A, Ramajahan T (2011) Antioxidant and antiinflammatory activity of tender coconut water in experimental myocardial infarction. Journal of Food Biochemistry 35(5): 1501-1507.

27. Ruiz S, Bernal MB, Chacón SDCC, Estrada DV (2018) Glucose Response in Animals Induced With Experimental Diabetes Type 1 after Treatment with Human Insulin Exposed To High Temperatures. Transsylvanian Review 1.

28. Saleem Mir M, Maqbool Darzi M, Musadiq Khan H, Ahmad Kamil S, Hassan S, et al. (2014) Pre-diabetic clinical changes induced by low doses of afo-soro-streafruit cocktail in rabbits. Iranian Journal of Pathology 9(2): 197-112.

29. Biddinger SB, Kahn CR (2006) From mice to men: insights into the insulin resistance syndromes. Annu Rev Physiol 68: 123-158.

30. Chattopadhyay R, Bandopadhyay M (2005) Effect of Azadirachta indica leaf extract on serum lipid profile changes in normal and streptozotocin induced diabetic rats. African Journal of Biomedical Research 8: 101-104.

31. Emam MA (2012) Comparative evaluation of antidiabetic activity of Rosmarinus officinalis L. and Chamomile recutita in streptozotocin induced diabetic rats. Agric Biol J North Am 3(6): 247.

32. Weir G, Clore E, Zmачnichs C, Bonner Weir S (1981) Islet secretion in a new experimental model for non-insulin-dependent diabetes. Diabetes 30(7): 590-595.

33. Karunanayake EH, Hearse DJ, Mellows G (1975) The metabolic fate and elimination of streptozotocin. Portland Press Limited 3(3): 410-414.

34. Adeyemi DO, Komolafe OA, Adewole OS, Obuotor EM, Adenowo TK (2009) Anti hyperglycemic activities of Annona muricata (Linn) 6(1): 62-69.

35. Corrigton RN, Collins HL, Chandler MP, Lemon PW, DiCARLO SE (1996) Diabetes reduces growth and body composition more in male than in female rats. Physiology & behavior 60(5): 401-426.

36. Yoshida T, Nishioka H, Nakamura Y, Kondo M (1985) Reduced noradrenaline turnover in streptozotocin-induced diabetic rats. Diabetologia 28(9): 692-696.

37. Saat A, Rosalı R, Syalroni N (2013) Potential hypoglycemic property of Ailhiza myriophylla and virgin coconut oil in streptozotocin induced diabetic rats. Int J Pharm Pharm Sci 5(4): 199-202.

38. Preetha P, Devi VG, Ramajohan T (2012) Hypoglycemic and antioxidant potential of coconut water in experimental diabetes. Food & function 3(7): 753-757.

39. Habibudin M, Daghriri HA, Humaira T, Al Qahtani MS, Hefzi AAH (2008) Antidiabetic effect of alcoholic extract of Caralluma sinaica L. on streptozotocin-induced diabetic rabbits. Journal of ethnopharmacology 117(2): 215-220.

40. MotyI K, McCabe L R (2009) Streptozotocin, type I diabetes severity and bone. Biological procedures online 11: 296-315.

41. Aly HE, Ibrahim MR, Metawa HM, Hosni EMA, Ibrahim FM (2010) In vivo and in vivo evaluation of the antidiabetic effect of different extracts of Nepeta cataria in streptozotocin induced diabetic rats. J Am Sci 6(10): 364-386.

42. El Sayyed H, Badawy G, Elnabi SH, El Elaimy I, Al Shehari E (2015) Ameliorative effect of Morus alba leaves extract against development retinopathy in pups of diabetic and aluminum intoxicated pregnant albino rats. Asian Pacific Journal of Tropical Biomedicine 5(4): 300-309.
43. Salil G, Nevin K, Rajamohan T (2011) Arginine rich coconut kernel protein modulates diabetes in alloxan treated rats. Chemico-biological interactions 189(2): 107-111.

44. Adewole S, Caston Martins E, Ojewole J (2006) Histochemical and biochemical effects of melatonin on pancreatic β-cells of streptozotocin-treated diabetic rats. Pharmacologyonline.

45. Kitchensky D, Tepper SA, Lloyd LM, Davidson LM, Klurfeld DM (1988) Serum and liver lipids of rats fed cocoa butter, corn oil, palm kernel oil, coconut oil and cholesterol. Nutrition research 8(3): 287-294.

46. Nevin K, Rajamohan T (2008) Influence of virgin coconut oil on blood coagulation factors, lipid levels and LDL oxidation in cholesterol fed Sprague-Dawley rats. e-SPEN, the European e-Journal of Clinical Nutrition and Metabolism 3(1): e1-e8.

47. Lenich CM, Chobanian AV, Brecher P, Zannis V (1991) Effect of dietary cholesterol and alloxan diabetes on tissue cholesterol and apolipoprotein E mRNA levels in the rabbit. Journal of lipid research 32(3): 431-438.

48. Preetha P, Devi VG, Rajamohan T (2014) Antihyperlipidemic effects of mature coconut water and its role in regulating lipid metabolism in alloxan-induced experimental diabetes. Comparative Clinical Pathology 23(5): 1331-1337.

49. Naskar S, Mazumder UK, Pramanik G, Gupta M, Kumar RS, et al. (2011) Evaluation of antihyperglycemic activity of Cocos nucifera Linn. on streptozotocin induced type 2 diabetic rats. Journal of ethnomedicine 138(8): 769-773.

50. Marina A, Man YC, Amin I (2009) Virgin coconut oil: emerging functional food. Trends in Food Science & Technology 20(10): 481-487.

51. Brunzell JD, Ayobi AF (2003) Dyslipidemia in the metabolic syndrome and type 2 diabetes mellitus. The American journal of medicine 115(1): 24-28.

52. Eyres L, Eyres MF, Chisholm A, Brown RC (2016) Coconut oil consumption and cardiovascular risk factors in humans. Nutrition reviews 74(4): 267-280.

53. Nagaraju A, Lokesh BR (2007) Interesterified coconut oil blends with groundnut oil or olive oil exhibit greater hypocholesterolemic effects compared with their respective physical blends in rats. Nutrition research 27(9): 580-586.