Soluble Vascular Cell Adhesion Molecule-1 (sVCAM-1) Is Elevated in Bronchoalveolar Lavage Fluid of Patients with Acute Respiratory Distress Syndrome

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Abstract

Introduction
Pulmonary vascular endothelial activation has been implicated in acute respiratory distress syndrome (ARDS), yet little is known about the presence and role of endothelial activation markers in the alveolar space in ARDS. We hypothesized that endothelial activation biomarkers would be differentially expressed in bronchoalveolar lavage fluid from patients with ARDS compared with healthy volunteers, and that biomarker concentrations would be associated with ARDS severity.

Methods
We performed a cross-sectional analysis of data from 26 intubated patients with ARDS undergoing evaluation for clinically suspected ventilator-associated pneumonia and five healthy volunteers. Patients underwent bronchoalveolar lavage a median of five days after intubation. Healthy volunteers also underwent bronchoalveolar lavage. Endothelial activation biomarkers (soluble vascular cell adhesion molecule-1 [sVCAM-1], soluble endothelial selectin [sESEL], angiopoietin-1 [Ang-1] and angiopoietin-2 [Ang-2]) were measured in bronchoalveolar lavage fluid. Clinically suspected ventilator-associated pneumonia was confirmed with microbiologic culture data.

Results
Patients with ARDS had significantly higher median sVCAM-1 concentrations in the bronchoalveolar lavage fluid compared with healthy volunteers (985 vs 119 pg/mL, p = 0.03).
Additionally, there was a trend toward greater bronchoalveolar lavage fluid sVCAM-1 concentrations among patients with moderate/severe compared to mild ARDS (1395 vs 209 pg/mL, p = 0.06). We did not detect significant differences in bronchoalveolar lavage fluid levels of sESEL, Ang-1 or Ang-2 between patients with ARDS and healthy volunteers. Median bronchoalveolar lavage fluid biomarker levels did not differ between patients with and without microbiologically-confirmed ventilator-associated pneumonia.

**Conclusions**

sVCAM-1 concentrations were significantly higher in the bronchoalveolar lavage fluid of patients with ARDS compared to healthy controls, and tended to be higher in moderate/severe ARDS compared to mild ARDS. Our findings add to the growing evidence supporting the concept that endothelial activation plays an important mechanistic role in the pathogenesis of ARDS. Further studies are necessary to characterize the role and/or clinical significance of sVCAM-1 and other endothelial activation markers present in the alveolar space in ARDS.

**Introduction**

Acute respiratory distress syndrome (ARDS) is a common cause of severe hypoxemia associated with mortality approaching 40% in U.S. hospitals [1]. ARDS is characterized by a robust inflammatory response encompassing both alveolar epithelial dysfunction and pulmonary vascular endothelial activation [2]. Pulmonary endothelial-epithelial barrier compromise contributes to increased vascular permeability and alveolar accumulation of inflammatory cells [3, 4]. Animal and in vitro models suggest that markers of endothelial activation may also accumulate in the alveolar space in ARDS [3, 5]. Whether a similar accumulation occurs in the lungs of mechanically ventilated patients with ARDS is unclear, and whether the presence of endothelial activation biomarkers in the alveolar space confers clinical significance is largely unknown.

Biomarkers of endothelial activation, such as endothelial selectin (ESEL) and vascular cell adhesion molecule-1 (VCAM-1) are expressed on the surfaces of activated human pulmonary microvascular endothelial cells [6], supporting that the cleaved, soluble forms of these molecules are relevant markers of endothelial activation in pulmonary pathology. Circulating serum/plasma soluble VCAM-1 (sVCAM-1) and angiopoietin-2 (Ang-2), another marker of endothelial activation, have been shown to be associated with ARDS [5, 7, 8]. However, studies measuring endothelial activation biomarkers in the bronchoalveolar lavage fluid (BALF) of patients with ARDS are limited and have yielded conflicting results. In one study, BALF soluble endothelial selectin (sESEL) was elevated among ARDS patients with chronic alcohol use [9]. Another study found higher Ang-2 and lower angiopoietin-1 (Ang-1) in the BALF of individuals with hyperoxia-induced acute lung injury [5]. A third demonstrated no difference in BALF soluble vascular cell adhesion molecule-1 (sVCAM-1) between patients with and without ARDS [8].

Additionally, there is a paucity of published data exploring the role of endothelial activation among patients with concomitant ARDS and ventilator-associated pneumonia (VAP). Up to 20% of patients receiving mechanical ventilation develop VAP, with greater frequency among patients with ARDS [10]. VAP is associated with greater organ dysfunction and higher mortality in severe ARDS [11, 12]. However, these studies primarily investigated epithelial cell...
activation, and limited data are available regarding the role of pulmonary endothelial activation on outcomes from ARDS and VAP.

Endothelial activation biomarkers in the alveolar space may be more specific for pulmonary, rather than systemic, inflammation and may provide insights into the pathogenesis of ARDS, particularly when associated with VAP. Therefore, in this study, we compared BALF concentrations of sVCAM-1, sESEL, Ang-1 and Ang-2 between patients with ARDS undergoing evaluation for VAP and healthy volunteers, and investigated whether biomarker concentrations were associated with ARDS severity. We hypothesized that these endothelial activation biomarkers would be differentially expressed in BALF from patients with ARDS compared to healthy volunteers, and that concentrations would differ by ARDS severity.

Materials and Methods

Cohort Study Participants

We performed a secondary analysis of data collected from adult intubated patients with ARDS undergoing bronchoscopy for suspected VAP. Bronchoscopy for diagnosis of clinically suspected VAP is standard practice at our institution, and these patients represented a convenience sample of ARDS patients cared for in the intensive care units of our institution in 2008 and 2009. ARDS was defined as: 1) PaO2/FiO2 <300 mmHg while receiving ≥5 cmH2O positive end-expiratory pressure, 2) diffuse parenchymal infiltrates, 3) pulmonary arterial wedge pressure <18 mmHg or lack of clinical evidence of congestive heart failure, and 4) no other obvious diagnosis explaining these findings. ARDS severity was based on the Berlin consensus definition as mild (200 mmHg < PaO2/FiO2 ≤ 300 mmHg) or moderate/severe (PaO2/FiO2 ≤200 mmHg) [13]. Clinical criteria for suspected VAP, based upon American Thoracic Society/Infectious Disease Society of America guidelines [14], included mechanical ventilation ≥48 hours, new or progressive pulmonary infiltrates on imaging, and ≥1 of the following: fever, leukocytosis or leukopenia, increased purulent endotracheal secretions, and no antibiotic changes for 72 hours. Control samples were obtained from healthy, non-smoking volunteers.

Written informed consent was obtained from participants or legal next of kin. Study procedures were approved by the Institutional Review Board at the University of Washington.

BALF Collection

Participants with ARDS and healthy volunteers underwent protocolized bronchoscopy with BALF sample collection as previously described [15, 16]. Briefly, five separate 30 mL aliquots of 0.9% sterile saline were instilled in the most affected lobe radiographically, or the right middle lobe or left lingula, then aspirated and pooled. BALF was immediately centrifuged at 14,000 g for 20 minutes at 4°C and cell-free supernatant was aliquoted and stored at -80°C. Clinically suspected VAP was confirmed when bacterial cell culture grew ≥10^4 colony forming units per milliliter (cfu/mL) or protected specimen brush culture grew ≥10^3 cfu/mL [17].

Biomarker Quantification

Concentrations of Ang-1, Ang-2, sESEL and sVCAM-1 were measured in aliquots of BALF supernatants using enzyme-linked sandwich immunoassay (ELISA), utilizing validated human-specific monoclonal antibody pairs (R&D Systems Duoset kits, Minneapolis, MN, USA) as previously described [18, 19]. Concentrations were extrapolated from simultaneously run standard curves.
Clinical Data

Baseline clinical, physiologic and demographic data were obtained from the clinical electronic medical record for intubated patients with ARDS via standardized chart abstraction methods. Collected data included: duration of mechanical ventilation, PaO₂:FiO₂ ratios, microbiologic culture results, in-hospital mortality, age and gender.

Statistical Analyses

Patient baseline characteristics were compared using Wilcoxon rank-sum tests for continuous and chi-square (\(\chi^2\)) or Fisher’s exact test for categorical variables. To account for non-parametric data, we used Wilcoxon rank-sum tests to compare median biomarker levels between patients with ARDS and healthy volunteers as well as by ARDS severity and microbiologically-confirmed VAP diagnosis. We repeated similar analyses restricted to patients without microbiologically-confirmed VAP as a sensitivity analysis to further evaluate the association of BALF endothelial activation biomarkers with ARDS independent of the effects of VAP. We also compared continuous values of BALF biomarkers with PiO₂:FiO₂ ratios, calculating the Pearson correlation coefficients (\(r\)). Finally, we compared biomarker levels with duration of mechanical ventilation and by in-hospital death, using Pearson correlation analyses and Wilcoxon rank-sum tests, respectively. A p-value of <0.05 was considered statistically significant. Analyses were performed in STATA version 13.1 (StataCorp LP, College Station, TX).

Results

Baseline characteristics were compared between 26 patients with mild and moderate/severe ARDS (Table 1). Most patients were men (n = 16, 55%) with a median age of 65 years (interquartile range [IQR] 49–75). As expected, patients with mild ARDS had higher median PaO₂:FiO₂ ratios compared to patients with moderate/severe ARDS (220 mmHg vs. 131 mmHg, \(p<0.001\)). BALF was collected on day 7 (IQR 2–13) of mechanical ventilation among patients with mild ARDS and day 5 (IQR 3–8) in those with moderate/severe ARDS. There was no significant difference between the proportion of microbiologically-confirmed VAP among patients with mild ARDS and day 5 (IQR 3–8) in those with moderate/severe ARDS. There was no significant difference between the proportion of microbiologically-confirmed VAP among patients with mild compared to moderate/severe ARDS (14% vs. 32%, \(p = 0.6\)). The most

Table 1. Baseline characteristics of ARDS patients stratified by ARDS severity.

|                      | All ARDS (n = 26) | Moderate/Severe ARDS (PaO₂:FiO₂ <200, n = 19) | Mild ARDS (PaO₂:FiO₂ 201–300, n = 7) |
|----------------------|-------------------|-----------------------------------------------|-------------------------------------|
| Age, years (median (IQR)) | 65 (49–75)        | 67 (51–79)                                    | 56 (47–62)                          |
| Male, n (%)           | 15 (58)           | 11 (58)                                       | 4 (57)                              |
| Race/Ethnicity, n (%) |                   |                                               |                                     |
| White                 | 22 (85)           | 18 (95)                                       | 4 (57)                              |
| Black                 | 3 (12)            | 0 (0)                                         | 3 (43)                              |
| Hispanic              | 1 (4)             | 1 (5)                                         | 0 (0)                               |
| Ventilator-associated pneumonia, n (%) | 7 (27) | 6 (32)                                       | 1 (14)                              |
| PaO₂:FiO₂ ratio (median (IQR)) | 155 (105–203) | 131 (97–156)                                 | 220 (219–238)                       |
| Total duration of mechanical ventilation, days (median (IQR)) | 12 (9–20) | 12 (9–17)                                    | 18 (10–27)                         |
| In-hospital death, n (%) | 9 (35)           | 8 (42)                                        | 1 (14)                              |

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common organism isolated was *Haemophilus influenzae* followed by methicillin-sensitive *Staphylococcus aureus*. In-hospital mortality was 14% among patients with mild ARDS and 42% among those with moderate/severe ARDS ($p = 0.4$).

Compared to five healthy controls, ARDS patients had significantly higher median BALF sVCAM-1 levels (985 pg/mL, IQR 134–2000, vs 119 pg/mL, IQR 86–124, $p = 0.03$; Fig 1, Panel A). Further, there was a trend toward greater BALF sVCAM-1 levels among patients with moderate/severe ARDS compared to those with mild ARDS (1395 pg/mL, IQR 418–7232 vs 209 pg/mL, IQR 42–1276, $p = 0.06$). Median BALF Ang-1, Ang-2 and sESEL concentrations did not differ significantly between ARDS patients and controls or by ARDS severity (Fig 1, Panel A and Table 2). When comparing continuous levels of BALF biomarkers with the PaO2:FIO2 ratio among ARDS patients, sVCAM-1 tended to be weakly inversely correlated with continuous PaO2/FiO2 ($r = -0.30$, $p = 0.1$); we detected no other associations between BALF biomarker levels and PaO2/FiO2 ratios (Fig 2). Additionally, there were no significant correlations between days elapsed prior to BALF collection and endothelial activation marker levels (sVCAM, $r = 0.05$, $p = 0.8$; sESEL, $r = -0.06$, $p = 0.8$; Ang-1, $r = -0.13$, $p = 0.6$; Ang-2, $r = 0.20$, $p = 0.4$).

Median BALF biomarker levels also did not differ significantly between patients with and without VAP diagnoses (Fig 1, Panel B). The associations described above were similar when we restricted analyses to patients who did not have microbiologically-confirmed VAP (Table 3).

We did not detect significant associations between BALF biomarker levels and the clinical outcomes of either duration of mechanical ventilation (sVCAM, $r = -0.21$, $p = 0.3$; sESEL, $r = -0.16$, $p = 0.5$; Ang-1, $r = -0.15$, $p = 0.5$; Ang-2, $r = 0.05$, $p = 0.8$) or in-hospital deaths in our cohort (Table 4).

**Discussion**

In this study, we found that BALF sVCAM-1 levels measured several days into the course of ARDS were significantly elevated compared to levels in healthy controls. We also analyzed endothelial activation biomarkers by ARDS severity and found that BALF sVCAM-1 concentrations tended to be higher among patients with moderate/severe ARDS compared to mild ARDS.

Our findings build upon the results of a prior study that found no difference in BALF sVCAM-1 levels between mechanically ventilated patients with and without ARDS [8]. Importantly, in that cohort, BALF samples were collected within two hours of intensive care unit admission, markedly earlier than in our study and likely relatively earlier in the pathogenesis of ARDS. Early sampling of the alveolar space may not fully reflect the complex contribution of endothelial activation to the pulmonary endothelial-epithelial (i.e., vascular-alveolar) barrier dysfunction in ARDS. Upon activation, endothelial cells upregulate cell surface adhesion molecule expression, and soluble isoforms, such as sVCAM-1, are subsequently shed. These isoforms accumulate in the plasma and alveolar fluid, modulating the inflammatory response [20] which is likely to impact pulmonary endothelial-epithelial barrier dysfunction and ARDS severity. Further studies exploring early versus late sampling strategies are necessary to elucidate the kinetics, ideal sampling frame and test characteristics of BALF sVCAM-1 as a biomarker of ARDS.

VCAM-1 is an endothelial adhesion molecule that regulates leukocyte recruitment during inflammation. VCAM-1 is cleaved from the endothelial cell surface to form soluble sVCAM-1. sVCAM-1 has biologic activity and can block adhesion of inflammatory cells to the endothelium and function to dampen the inflammatory response [20]. Regulation of VCAM-1 expression and sVCAM-1 shedding in ARDS, especially in the setting of VAP, is not extensively...
studied. Data available from patients with sepsis, the most common cause of ARDS, suggest that higher sVCAM-1 levels correlate with severity of sepsis [21]. Other studies, however, report an association between mid-range levels of soluble adhesion molecules and worse outcomes [20]. The blunted elevations in soluble isoforms were attributed to inadequate or aberrant shedding, and may lead to unresolved inflammation and persistent endothelial activation [20]. Interestingly, a post-mortem study demonstrated over-expression of endothelial cell surface VCAM-1 in the pulmonary vessels of patients who died from ARDS [22], but BALF sVCAM-1 levels were not measured in that study. It is unclear whether our findings of elevated BALF sVCAM-1 amongst ARDS patients reflect expected levels of shedding or, conversely, a dysregulated pathophysiologic mechanism, especially in the presence of concomitant VAP.

We detected no significant differences in sESEL, Ang-1 or Ang-2 BALF concentrations between patients with ARDS who were undergoing evaluation for clinically suspected VAP and healthy volunteers. In a study of patients with chronic alcohol abuse, although BALF sESEL levels were elevated, levels did not differ significantly between patients with and without ARDS [9]. Also, previous investigators demonstrated elevated Ang-2 and decreased Ang-1 levels in the BALF of patients with hyperoxia-induced acute lung injury [5]. However, in our study, we did not identify appreciable differences in the levels of Ang-2 and Ang-1 in BALF, suggesting that alveolar accumulation of angiopoietin may not be uniform across types of lung injury. We also did not identify differences in median BALF biomarker levels between patients

Table 2. Levels of endothelial activation biomarkers (pg/mL) in bronchoalveolar lavage fluid of ARDS patients and controls.

|                  | All ARDS n = 26 | Moderate/Severe ARDS (PaO2:FIO2 < 200) n = 19 | Mild ARDS (PaO2:FIO2 201–300) n = 7 | Controls n = 5 |
|------------------|-----------------|---------------------------------------------|-----------------------------------|--------------|
| sVCAM, median (IQR) | 985 (134–2000)* | 1395 (418–7232)** | 208 (42–1276) | 119 (86–124) |
| sESEL, median (IQR) | 12 (12–222) | 100 (12–244) | 12 (12–12) | 93 (72–103) |
| Ang-1, median (IQR) | 20 (20–53) | 20 (20–103) | 20 (20–20) | 20 (20–21) |
| Ang-2, median (IQR) | 14 (14–14) | 14 (14–22) | 14 (14–14) | 14 (14–49) |

*Control vs. ARDS, $\chi^2 p = 0.03$.

**Mild vs. Moderate/Severe ARDS $\chi^2 p = 0.06$
with and without microbiologically-confirmed VAP. The small sample size of our study may have impacted our ability to detect associations between the studied BALF biomarker levels and ARDS. Further, our study was not sufficiently powered to detect differences in clinical outcomes, such as ventilator days or in-hospital mortality associated with BALF biomarker concentrations.

Although we did not measure levels of cytokines in BALF, inflammatory cytokines have been implicated in ARDS pathogenesis, severity and outcomes, and mediate the expression of endothelial activation biomarkers [2, 23]. For instance, VCAM-1 expression on endothelial cell surfaces has been shown to be induced by IL-4 and IL-13 [24, 25] as well as other inflammatory exposures [26], and our finding of elevated sVCAM-1 in BALF is consistent with these published data. Conversely, the expression of E-selectin is downregulated over time by IL-4 and IL-13 [27], supporting our finding that E-selectin was not markedly elevated in the BALF of patients with ARDS collected a median of 5 or 7 days after intubation. Further, among other inflammatory cytokines, IL-4 has been described to rise in BALF over the course of ARDS progression, particularly in the setting of sepsis [28].

Table 3. Levels of endothelial activation biomarkers (pg/mL) in bronchoalveolar lavage fluid restricted to patients without microbiologically-confirmed VAP and controls.

|                      | All ARDS (without VAP) | Moderate/Severe ARDS (PaO₂:FiO₂ ≤ 200) | Mild ARDS (PaO₂:FiO₂ 201–300) | Controls |
|----------------------|------------------------|----------------------------------------|-------------------------------|----------|
|                      | n = 19                 | n = 13                                 | n = 6                         | n = 5    |
| sVCAM, median (IQR)  | 748 (94–2203)          | 1515 (515–7238)                        | 147 (42–649)                  | 119 (86–124) |
| sESEL, median (IQR)  | 12 (12–229)            | 109 (12–244)                           | 12 (12–12)                    | 93 (72–103) |
| Ang-1, median (IQR)  | 20 (20–89)             | 25 (20–103)                            | 20 (20–20)                    | 20 (20–21) |
| Ang-2, median (IQR)  | 14 (14–22)             | 14 (14–72)                             | 14 (14–14)                    | 14 (14–49) |
Our study has several other limitations, and, accordingly, detected associations or the lack thereof should be interpreted with caution. Firstly, only patients with ARDS and clinical suspicion for VAP underwent bronchoalveolar lavage. This may have introduced bias if a process mimicking VAP affects BALF endothelial activation biomarkers. However, we found no significant differences in BALF biomarker concentrations between patients who did and did not have microbiologically-confirmed VAP. Secondly, BALF samples were collected at a single time point, owing to the pragmatic nature of this study, which analyzed biomarkers collected from clinically indicated bronchoscopies. Thirdly, the use of BALF rather than undiluted pulmonary edema fluid for our analyses may have accounted for some of the differences observed in our study compared with prior studies; we were unable to calculate the dilution factor of BALF. Finally, although we could not exclude masking of proteins by binding to matrix components, we have not encountered difficulties in prior measurements of select proteins in the BALF of patients with ARDS [29–31].

Nonetheless, our study’s noteworthy strength is that we analyzed biomarkers of endothelial activation from the BALF of patients with ARDS. Findings from samples obtained directly from the alveolar space may provide new insights specific to the effects of endothelial activation in the lungs in the face of the often systemic involvement of ARDS. Recent data suggest that bronchoalveolar sampling for inflammatory biomarkers may better predict VAP compared to serum cytokine analysis [32], and this may extend to endothelial activation markers in ARDS.

Conclusions

We found that BALF sVCAM-1 concentrations were significantly elevated in ARDS and tended to be greater in moderate/severe compared to mild ARDS. Our findings add to the growing evidence supporting the concept that endothelial activation plays an important mechanistic role in the pathogenesis of ARDS. Further studies are necessary to understand the role of endothelial activation in the pathogenesis of ARDS in VAP and other conditions.

Supporting Information

S1 Data. Raw data collected and interpreted for this study.
(XLSX)

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Author Contributions
Conceived and designed the experiments: EFA SEJ LMS WCL. Performed the experiments: LMS WCL. Analyzed the data: EFA SEJ. Contributed reagents/materials/analysis tools: LMS WCL. Wrote the paper: EFA SEJ KC LMS WCL. Contributed to interpretation of the analyzed data: EFA SEJ KC LMS WCL. Revised the manuscript critically for important intellectual content: KC LMS WCL.

References
1. Rubenfeld GD, Caldwell E, Peabody E, Weaver J, Martin DP, Neff M, et al. Incidence and outcomes of acute lung injury. N Engl J Med 2005; 353:1685–1693. PMID: 16236739
2. Matthay MA, Ware LB, Zimmerman GA. The acute respiratory distress syndrome. J Clin Invest 2012; 122:2731–2740. doi: 10.1172/JCI60331 PMID: 22850883
3. Bhargava M, Wendt CH. Biomarkers in acute lung injury. J Lab Clin Med 2012; 159:205–217.
4. Orfanos SE, Mavrommati I, Korovesi I, Roussos C. Pulmonary endothelium in acute lung injury: from basic science to the critically ill. Intensive Care Med 2004; 30:1702–1714. PMID: 15258936
5. Bhandari V, Choo-Wing R, Lee CG, Zhu Z, Nedrelow JH, Chupp GL, et al. Hyperoxia causes angiopoietin 2-mediated acute lung injury and necrotic cell death. Nat Med 2006; 12:1286–1283. PMID: 17086189
6. Muller AM, Hermanns MI, Cronen C, Kirkpatrick CJ. Comparative study of adhesion molecule expression in cultured human macro- and microvascular endothelial cells. Exp Mol Pathol 2002; 73:171–180. PMID: 12565792
7. van der Heijden M, van Nieuw Amerongen GP, Koolwijk P, van Hinsbergh VW, Groeneveld AB. Angiopoietin-2, permeability oedema, occurrence and severity of ALI/ARDS in septic and non-septic critically ill patients. Thorax 2008; 63:903–908. doi: 10.1136/thx.2007.087387 PMID: 18559364
8. Agouridakis P, Kyriakou D, Alexandrakis MG, Prekates A, Perisinakis K, Karkavitsas N, et al. The predictive role of serum and bronchoalveolar lavage cytokines and adhesion molecules for acute respiratory distress syndrome development and outcome. Respir Res 2002; 3:25. PMID: 12537603
9. Burnham EL, Moss M, Harris F, Brown LA. Elevated plasma and lung endothelial selectin levels in patients with acute respiratory distress syndrome and a history of chronic alcohol abuse. Crit Care Med 2004; 32:675–679. PMID: 15090946
10. Markowicz P, Wolff M, Djedaini K, Cohen Y, Chastre J, Delclaux C, et al. Multicenter prospective study of ventilator-associated pneumonia during acute respiratory distress syndrome. Incidence, prognosis, and risk factors. ARDS Study Group. Am J Resp Crit Care Med 2000; 161:1942–1948 PMID: 10852771
11. Forel JM, Voillet F, Pulina D, Gacouin A, Perrin G, Barrau K, et al. Ventilator-associated pneumonia and ICU mortality in severe ARDS patients ventilated according to a lung-protective strategy. Crit Care 2012; 16:R65. doi: 10.1186/cc11312 PMID: 22524447
12. Papazian L, Bregeon F, Thirion X, Gregoire R, Saux P, Denis JP, et al. Effect of ventilator-associated pneumonia on mortality and morbidity. Am J Resp Crit Care Med 1996; 154:91–97. PMID: 8680705
13. Ranieri VM, Rubenfeld GD, Thompson BT, Ferguson ND, Caldwell E, Fan E, et al. Acute respiratory distress syndrome: the Berlin definition. JAMA 2012; 307:2526–2533. doi: 10.1001/jama.2012.5669 PMID: 22797452
14. American Thoracic Society, Infectious Diseases Society of America. Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. Am J Resp Crit Care Med 2005; 171:388–416. PMID: 15699079
15. Steinberg KP, Milberg JA, Martin TR, Maunier RJ, Cochrell BA, Hudson LD. Evolution of bronchoalveolar cell populations in the adult respiratory distress syndrome. Am J Resp Crit Care Med 1994; 150:113–122. PMID: 8025736
16. Chen J, Ryu S, Gharib SA, Goodlett DR, Schnapp LM. Exploration of the normal human bronchoalveolar lavage fluid proteome. Proteomics Clin Appl 2008; 2:585–595. doi: 10.1002/prca.200780006 PMID: 21136857
17. Chastre J, Fagon JY: Ventilator-associated pneumonia. Am J Resp Crit Care Med 2002; 165:867–903. PMID: 11934711
18. Graham SM, Rajwans N, Jaoko W, Estambale BB, McClelland RS, Overbaugh J, et al. Endothelial activation biomarkers increase after HIV-1 acquisition: Plasma vascular cell adhesion molecule-1 predicts...
disease progression. AIDS 2013; 27:1803–1813. doi: 10.1097/QAD.0b013e328360e9fb; PMID: 23807276

19. Graham SM, Rajwans N, Tapia K, Jaoko W, Estambale BB, McClelland RS, et al. A prospective study of endothelial activation biomarkers, including plasma angiopoietin-1 and angiopoietin-2, in Kenyan women initiating antiretroviral therapy. BMC Infect Dis 2013; 13:263. doi: 10.1186/1471-2334-13-263; PMID: 23807276

20. Zonneveld R, Martinelli R, Shapiro NI, Kuipjers TW, Plotz FB, Carman CV. Soluble adhesion molecules as markers for sepsis and the potential pathophysiological discrepancy in neonates, children and adults. Crit Care 2014; 18:204. doi: 10.1186/cc13733; PMID: 24602331

21. Shapiro NI, Schuetz P, Yano K, Sorasaki M, Parikh SM, Jones AE, et al. The association of endothelial cell signaling, severity of illness, and organ dysfunction in sepsis. Crit Care 2010; 14:R182. doi: 10.1186/cc9290; PMID: 20942957

22. Muller AM, Cronen C, Muller KM, Kirkpatrick CJ. Heterogeneous expression of cell adhesion molecules by endothelial cells in ARDS. J Pathol 2002; 198:270–275. PMID: 12237888

23. Binnie A, Tsang JL, dos Santos CC. Biomarkers in acute respiratory distress syndrome. Curr Opin Crit Care 2014; 20:47–55. doi: 10.1097/MCC.0000000000000048; PMID: 24296379

24. Schleimer RP, Sterbinsky SA, Kaiser J, Bickel CA, Klunk DA, Tomioka K, et al. IL-4 induces adherence of human eosinophils and basophils but not neutrophils to endothelium. Association with expression of VCAM-1. J Immunol 1992; 148:1086–1092. PMID: 1371130

25. Bocchner BS, Klunk DA, Sterbinsky SA, Coffman RL, Schleimer RP. IL-13 selectively induces vascular cell adhesion molecule-1 expression in human endothelial cells. J Immunol 1995; 154:799–803. PMID: 7529288

26. Lee KS, Lee KY, Kim SR, Park HS, Park SJ, Min KH, et al. Blockade of airway inflammation and hyperresponsiveness by an angiopoietin-1 variant, COMP-Ang1. Exp Mol Med 2007; 39:733–745. PMID: 1816084

27. Etter H, Althaus R, Eugster HP, Santamaria-Babi LF, Weber L, Moser R. IL-4 and IL-13 downregulate rolling adhesion of leukocytes to IL-1 or TNF-alpha-activated endothelial cells by limiting the interval of E-selectin expression. Cytokine 1998; 10:395–403. PMID: 9632524

28. Meduri GU, Kohler G, Headley S, Tolley E, Stentz F, Postlethwaite A. Inflammatory cytokines in the BAL of patients with ARDS. Persistent elevation over time predicts poor outcome. Chest 1995; 108:1303–1314. PMID: 7587434

29. Nguyen EV, Gharib SA, Palazzo SJ, Chow YH, Goodlett DR, Schnapp LM. Proteomic profiling of bronchoalveolar lavage fluid in critically ill patients with ventilator-associated pneumonia. PLoS One 2013; 8:e58782. doi: 10.1371/journal.pone.0058782; PMID: 2350556

30. Palazzo SJ, Simpson TA, Simmons JM, Schnapp LM. Soluble triggering receptor expressed on myeloid cells-1 (sTREM-1) as a diagnostic marker of ventilator-associated pneumonia. Respir Care 2012; 57:2052–2058. doi: 10.4187/respcare.01703; PMID: 22613763

31. Park WY, Goodman RB, Steinberg KP, Ruzinski JT, Radella F 2nd, Park DR, et al. Cytokine balance in the lungs of patients with acute respiratory distress syndrome. Am J Respir Crit Care Med 2001; 164:1896–1903. PMID: 11734443

32. Hellyer TP, Morris AC, McAuley DF, Walsh TS, Anderson NH, Singh S, et al. Diagnostic accuracy of pulmonary host inflammatory mediators in the exclusion of ventilator-acquired pneumonia. Thorax 2015; 70:41–47. doi: 10.1136/thoraxjnl-2014-205766; PMID: 25298428