Tracing the horizontal transport of microplastics on rough surfaces

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Abstract
Occurrence and distribution of microplastics in different ecosystems have recently become subjects of numerous studies. However, to date the research has focused mainly on marine and freshwater ecosystems and widely neglected terrestrial environments. Only recently, first studies investigated the microplastics contamination of soils. Therefore, we know little about the transport mechanisms of microplastics in soils and sediments and virtually nothing about their surface transport. In this study we investigate surface transport mechanisms by tracking fluorescent, irregularly shaped polymethyl methacrylate (PMMA) particles in real time in a laboratory setup. In 108 experimental runs, we vary the irrigation rates, inclinations and surface roughnesses. Additionally, we simulate the small-scale flow patterns to resolve the role of the roughness-induced microrelief. Our results suggest that microplastics are transported along preferential pathways resulting from the micro- and macrorelief, which can be correlated to the flow pattern observed in the computer simulation. Our model study facilitates a deeper insight into microplastic transport on different soil surfaces and serves as a pilot for investigating mechanisms of horizontal microplastic transport. However, microplastics are a diverse group of contaminants with varying shapes, densities and sizes. Therefore, for a full understanding of transport of microplastics in terrestrial environments, it is important to address these properties as well as more variable surfaces for horizontal migration and to include vertical transport mechanisms in future research.

Keywords: Microplastics, Surface transport, Surface roughness, PMMA, sCMOS camera, Particle tracking

Introduction
The large-scale production of synthetic polymers started in the 1950s and over the last seven decades the amount of produced plastic has exceeded eight billion tons [1]. In 2019, plastic production reached 57.9 million tons in Europe and 368 million tons globally [2]. Plastic products are durable and resist to degradation [1], however, the accompanying environmental challenges caused by the increasing amount of plastic waste entering the environment have been neglected for a long time. Due to its poor biodegradability, plastic waste could remain in the environment for centuries leading to an accumulation and a potential threat to our ecosystems [1, 3]. While for a long time only large items of plastic waste have been studied, microplastics came more into focus during the last decade [3]. Microplastics are in general defined as particles smaller than 5 mm [4] and occur as primary and secondary microplastics. Primary microplastics are intentionally produced micro particles like pellets and beads. Secondary microplastics originate from abrasion of plastic products or fragmentation of macroplastics by a combination of e.g. elevated temperature, light, exposure to water, friction from wind and water or exposure to organisms, resulting in thermo-oxidative degradation, photo-oxidation, hydrolysis, mechanical breakdown and biodegradation [1, 5, 6]. Due to their manifold origins and degradation pathways, microplastics are a diverse group of contaminants with a broad range of shapes, densities, sizes, polymers and additives [7]. It is best to think about...
their properties as a continuum rather than in discrete categories [8]. Today microplastics are found in every environmental compartment and region of the globe from megacities to even the remotest arctic areas [9–11].

While the contamination of oceans with microplastics and their transport in the marine environment have become a widely considered research field, our knowledge on the distribution and transport of microplastics in terrestrial environments is still limited [10, 12–14]. This is partly due to the analytical challenges because the extraction of microplastics from mineral and organic-rich soils and sediments is much more complicated than from aquatic samples [15–17]. Additionally, we still lack a standard analytical protocol which is a prerequisite for a proper comparison of data from different studies [18, 19]. Furthermore, the distribution of microplastics in the terrestrial environments depends on diverse sources and transport pathways. Sources can be manifold and include e.g. the introduction of microplastics via sewage sludge, compost application or plastic mulching in agriculture, littering or waste mismanagement and the fragmentation and abrasion of plastic particles from plastic products with outdoor use [17, 20, 21]. Once in the environment, microplastics may be transported by e.g. wind and water, and may finally be deposited and potentially accumulate in different environmental compartments [20, 22, 23]. Indeed, terrestrial soils are suggested as a large sink for microplastics [10, 24, 25].

While first experimental evidence on factors affecting vertical transport processes of microplastics in soils and sediments like e.g. particle size or presence of preferential flow paths are published [26–30], studies on horizontal transport are lacking to date [20, 31]. To fill this gap, tracking microplastics during their transport could generate the necessary data and help to understand the relevant transport parameters and mechanisms. Particle tracking, also referred to as particle tracing [32], has a long history in sedimentology [33]. In this discipline it is understood as the analysis of the spatial and temporal distribution of single natural or artificial components of the sediment that are marked and observed during transport [32]. In environmental sciences, sand particles are tracked for example along fluvial trajectories [34], along coasts [35] or on tidal inlets [36]. Furthermore, particles can also be tracked under laboratory condition on a far smaller scale including vertical and horizontal transport pathways with a high temporal and spatial resolution [37–40].

Nowadays specific marking or tagging the particles of interest with an identifiable signature largely facilitates the tracking [32]. Since the use of radioactivity has shown its limitations and is seen critically due to environmental and health concerns, fluorescent dyes are commonly used [32, 41]. Therefore, rhodamine or anthracene serve frequently as fluorescent coatings in combination with additive binding agents for particle tracking [32, 42]. Advances in real-time imaging and image processing allow to trace temporal and spacial distribution of particles, at least under laboratory conditions. Hardy and colleagues [40], for example, developed a real-time approach to trace the movement of sand particles with fluorescent coating during rainfall events by using fluorescent videography techniques. They could visualise and measure transport parameters like direction and travel distance of sand particles. However, most of the studies dealing with particle tracing focus on the spatial distribution as a result of transport processes. Real-time tracking of (natural) particles as demonstrated by Hardy and colleagues in a laboratory and field setup for terrestrial environments remains an exception [40, 43]. Nizzetto and colleagues [31] suggested in their theoretical study that microplastics could be transported similarly to natural particles. However, data to confirm this statement and to parametrize models is lacking [44]. Nevertheless, such data are of utmost importance for a mechanistic understanding of transport processes of microplastics in the environment.

In this study, we constructed a laboratory setup for real-time particle tracing and observed fluorescent microplastics in irrigation experiments. We analysed the influence of different irrigation rates, inclinations and surface roughnesses on the movement of irregularly shaped polymethyl methacrylate microplastics.

Material and methods
Microplastic particles
We used UV-fluorescence-labelled polymethyl methacrylate (PMMA) particles. PMMA, also known as acrylic glass, has a density of 1.18 to 1.19 g cm$^{-3}$ and is thus heavier than water. The source material was purchased already fluorescence-labelled from LLV-shop (https://www.llv-
Fig. 2 Laboratory setup with the irrigation and illumination systems. The PMMA particles are positioned in row vertically below the camera.
Fig. 3 Examples of (a) trajectories of particles and (b) corresponding path lengths. These are results from one of the runs on the coarse surface with inclination 2.5° and an irrigation rate of 10.44 L h⁻¹ and from a run with the same parameters but on the fine surface. Particle ID corresponds to an automatic numbering of PMMA articles sorted by the y coordinate.

6.5 μm² and a Ricoh FL-BC2518-9M 25 mm objective. An sCMOS sensor has the advantage to combine a large field-of-view, low electrical noise and a high frame rate compared to a CCD (charge-coupled device) sensor [46]. The exposure time of the camera was set to 100 ms, resulting in a frame rate of 10 frames per second and 500 pictures during the 50 seconds of recording for each experimental run. The camera was positioned 30 cm vertically above the PMMA particles in the central part of the surface and recorded an area of 150 × 150 mm (Supplementary Figure S2). The camera was equipped with an EFFO-FLR-BN630-M40.5 optical filter (Midwest Optical Systems Inc., Palatine, Illinois, United States) to limit the recorded spectrum to a narrow spectral range around 630 nm, which corresponds to the fluorescence of the PMMA particles. Two EFFI-FLEX-15-465-SD-P2 LED bars (EFFILUX GmbH, Les Ulis, France) with an emission wave length of 465 nm (blue light) were fixed laterally above the rough surface to excite the fluorescence of the PMMA particles. Natural illumination was minimized by darkening the laboratory to reduce noise in the pictures.

Image analysis
We analysed the images using Python 3.8 (Python Software Foundation, https://www.python.org/) and R 4.0.3 [47]. Some of the images contained artefacts due to residual ambient light which resulted in blurred particles’ edges. Additionally, some particles moved abruptly and their trajectories appeared as lines in the images (Supplementary Figure S4). To remove the artefacts, we first preprocessed the images by calculating the centre of mass of every particle and then simplified them to equally sized circles centered around their centres of mass. This step increased the accuracy of particle tracking. The latter was done with TrackPy version 0.4.2 [48], a Python package that calculates particle trajectories or paths. Sub-
subsequently, we derived the path lengths by calculating the Euclidean distances between particle positions in consecutive images. The calculated centre of mass could vary slightly from image to image by one or two pixels without any particle movement. Therefore, we ignored any trajectory of less than 3 pixels.

From every first image of the experiment, we estimated the sizes of the particles by segmenting the images at the grey value 200 (i.e. any pixel with a grey value equal to or larger than 200 was classified as particle). Based on these sizes in pixels, we calculated particle characteristics ‘equivalent diameter’ (i.e. the diameter that a completely spherical particle with the same area visible in the image would have) and ‘eccentricity’ which shows whether a particle resembles a sphere or an ellipse. Eccentricity is defined as the ratio between the focal points of an ellipse to its major axis. To calculate eccentricity, the particle’s shape is approximated as an ellipse with the same second-order moments (covariance) as its visible area. Both characteristics were calculated with the Python module ‘measure’ from the image processing library scikit-image (https://scikit-image.org/).

**Table 1** Estimated parameters of the linear model with confidence intervals shown in parentheses

| Model parameter                  | Estimate (CI)          |
|----------------------------------|------------------------|
| Intercept (mm)                   | 1.76 (1.2, 2.3)        |
| Equivalent diameter (mm μm⁻¹)    | -2.96E-03 (-4.49E-03, -1.54E-03) |
| Inclination (mm degree⁻¹)        | -0.46 (-0.69, -0.27)   |
| Irrigation (mm L⁻¹ h⁻¹)          | 0.89 (0.57, 1.3)       |
| Roughness fine (mm)              | 3.33 (1.97, 5.11)      |

The parameters correspond to β in Eq. 1. They represent slopes for numerical variables and an additive effect for the factor roughness.

**Statistical analysis**

We estimate the influence, i.e. effect sizes and confidence intervals, of the experimental parameters ‘roughness’, ‘inclination’, ‘irrigation’ rate and ‘equivalent diameter’ on the length of paths that the particles moved in the experimental runs. We follow the advice of the American Statistical Association and refrain from hypothesis testing [49, 50]. Instead, we estimate the effect sizes and their confidence intervals using bootstrap re-sampling [51] in the following linear model:

\[
y = \text{Int} + \beta_{\text{ED}} \cdot \text{ED} + \beta_{\text{Inc}} \cdot \text{Inc} + \beta_{\text{Irr}} \cdot \text{Irr} + \beta_{\text{Rough}} \cdot \text{Rough} + \epsilon
\]

where \(y\) is the path length in mm, Int the intercept in mm, ED the equivalent diameter in μm, Inc the inclination in degrees, Irr the irrigation rate in L h⁻¹, Rough the roughness (a factor, coarse being the reference level), \(\beta_i\) are the model parameters or effects (a slope for numerical variables and an additive effect for factors) and \(\epsilon\) are the residuals of the model. The intercept is the expected value of the path length if all other factors are zero. This is not

**Fig. 5** Velocity profile on a 1 cm × 1 cm section of the fine-grained microrelief illustrated with colored passive tracer particles (left). The 0.3 mm average water film thickness leads to a flow rate of 7.27 L h⁻¹. The flow direction is top to bottom. The flow pattern is stationary and forms streams of increased velocity around teardrop shaped regions of decreased velocity behind bumps in the microrelief. At the bottom left we show a side view of the microrelief and water layer above. At the right we show a heatmap of the microrelief elevation.
meaningful for our setting because this would extrapolate the model beyond the measured ranges of the experimental parameters. To have a meaningful intercept, we subtracted the mean from the irrigation, inclination and the equivalent diameter, i.e. we centered all experimental parameters around their respective mean values. Therefore, the intercept now equals the travelled path length in mm for a mean-sized particle (1817 µm) irrigated by the mean irrigation rate (7.5 L h⁻¹), on a mean-inclined (8.7°) coarse surface. This data normalization does not affect the estimation of the effects βᵢ, but of the intercept only. We note that our model aims at a phenomenological description of particle transport within a certain range of parameters around their mean values rather than a fully detailed physical prediction.

We estimated the confidence intervals by bootstrapping the data (stratified by roughness) 10000 times and re-calculated the model on these bootstrap samples [52]. Different methods to estimate confidence intervals from bootstrapping exist and we compared three of them, namely the percentile, student-t and bca (bias-corrected and accelerated) methods, all implemented in the R package rsample [53]. All calculations were done in R [47] using the package collection tidymodels [54], V.0.1.0.

Simulation of the water flow
To better understand the water flow on the sediment surfaces, we qualitatively examined the flow patterns in a simulation on a microscopic level of detail. We assumed a free surface flow totally wetting an inclined plate with roughness at the scale of the grains of sand used in preparation of the fine surface. The liquid layer was very thin, about 0.1 – 0.3 mm thickness.

To model the flow, we used the Volume-of-Fluid [55-57] lattice Boltzmann method [58, 59] implemented in the software FluidX3D [55, 60, 61]. We simulated an area of 10 mm × 10 mm at 2.5° inclination. The boundaries in the lateral directions were periodic. We used the parameters of water at 20 °C (kinematic shear viscosity \( \nu = 1.004 \cdot 10^{-6} \text{ m}^2 \text{ s}^{-1} \), density \( \rho = 998.21 \text{ kg m}^{-3} \), surface tension \( \sigma = 72.75 \cdot 10^{-3} \text{ kg s}^{-2} \) [62]). We generated the microrelief of the surface caused by its roughness with a 2D periodic Perlin noise [63]. The grain size for the Perlin noise was set to approximately 0.4 mm diameter and the thickness of the water film to 0.3 mm on average. This resulted in a simulated flow rate (scaled up to the entire 400 mm width of the plate in the experiment) of 7.27 L h⁻¹ which is in line with experimental parameter of 7.2 L h⁻¹.

Additionally, to visualize the flow patterns, we irrigated the surfaces without any PMMA particles with a methanol solution stained with Nile Red, a fluorescent dye [64, 65]. Using Nile Red allowed us to capture the flow patterns with the same LED lights and camera setup and filter as in the experimental runs with particles. We are aware that methanol has different wetting properties than water. Therefore, we use the videos for visualisation purposes only. We recorded the initial wetting process as well as the flow patterns once stationary flow set in. The coarse surface was irrigated at an inclination of 7.5° with an irrigation rate of 7.2 L h⁻¹ and the fine surface at an inclination of 2.5° with an irrigation rate of 7.2 L h⁻¹. The videos are provided in the Additional files 2 and 3.

Results and discussion
Tracking microplastics under laboratory conditions
Our experiments allowed for a high-resolution tracking of microplastic particles under different inclinations, surface roughness and irrigation scenarios. Next to their respective trajectories, the image analysis provided information about the characteristics of the particles in the experimental runs. The equivalent diameter of PMMA particles ranged between 453 and 3387 µm (Supplementary Figure S3). They differed from spheres with eccentricities varying between 0.1 and 0.9. An eccentricity of 0 corresponds to a perfect circle, a value of 1 indicates an “extreme ellipse”, a parabola. Thus, the larger the value the less circular the PMMA particle. The distribution of both characteristics were comparable between different roughnesses and irrigation rates and shows that the particles in the experiments were comparable (Supplementary Figure S3). Therefore, no differences between the treatments as a consequence of a systematically varying particle morphology are to be expected.

576 out of 1081 particles remained motionless and the other 505 recorded particle trajectories were mostly short (Fig. 3). Most of the recording time, particles did not move at all. However, from time to time they travelled a larger distance, often very quickly. Thus, the overall path lengths and the particle’s maximum path length per second correlated (Spearman correlation coefficient of 0.9).

The Supplementary Figure S5 shows this relationship. The maximum overall recorded path length equalled 136 mm (fine surface, inclination 10°, irrigation rate 10.44 L h⁻¹) and the maximum recorded path length in one second 135 mm (fine surface, inclination 2.5°, irrigation rate 7.2 L h⁻¹).

Influence of experimental parameters on path lengths
The observed path lengths increased with increasing irrigation rate and were larger on the fine than on the coarse surface (Fig. 4). However, the dot plots show that the most frequent paths were small or zero and that only few particles traveled far.

The estimated parameters of the linear model support this observation (Table 1). Confidence intervals estimated with different bootstrap methods were comparable
(Supplementary Figure S6) and we report the student-t intervals only. All confidence intervals excluded zero. With increasing inclination and increasing equivalent diameter of the microplastic particles, the path length decreased. Indeed, for a constant flow rate, the water film thickness is approximately proportional to the $\sin(\alpha)^{-1/3}$, with $\alpha$ being the inclination [66][eq.(4.2.13)] and the larger the film thickness the larger the flow rate (see also “Water flow” section). The effects of inclination and equivalent diameter seem smaller compared to the irrigation rate or the roughness. However, the experimental parameters are measured on different scales and the absolute values of the effects depend on it. Thus, it is better to compare the effects relative to the measurement units of the parameters. If, for example, the inclination is increased from 5 to 10 degrees, the path length would decrease by 2.3 mm. For an increase of irrigation rate from 4 L h$^{-1}$ to 8 L h$^{-1}$, the path length would increase by 3.6 mm. By increasing the equivalent diameter of the microplastic particles from 1000 to 2000 μm, the path length decreases by 3 mm. And finally, on the fine surface, the particles travelled 3.33 mm farther which is in accordance with several observations from laboratory and environmental studies (e.g. in fluvial context and at valley slopes [67,68] or on beaches [69,70]).

The overall goodness of fit of the model was low ($R^2_{adj} = 0.07$) indicating that the assumption of the linear relationship between the experimental parameters and the path length is overly simplistic. However, because our intention is not to predict path lengths but to analyse the influence of experimental parameters (size effect and confidence intervals), the model is still useful.

**Water flow**

To better understand why most particles are barely advected whilst a few particles are moved across a long distance (Fig. 4), we qualitatively examined the flow pattern in a simulation. For the typical flow rates used in the experiment, the water film thickness is on the same scale as the grain size of the roughness-induced microrelief. The microrelief therefore greatly influences the flow pattern on a microscopic level. For the setup simulated here, where we target 7.2 L h$^{-1}$ to match the experiment, we find that the film thickness is 0.3 mm on average. For comparison, for the targeted irrigation rates of 4.8 and 10.4 L h$^{-1}$, we determine the film thickness to 0.252 mm and 0.348 mm respectively. The resulting simulation flow rates are 4.87 and 10.97 L h$^{-1}$. The flow rate is proportional to film thickness to the third power for a laminar flow over a plane and smooth surface [66][eq.(4.2.12)]. With increasing inclination, the film thickness decreases. If the film is thinner than the diameter of the microplastic particles, the water surface wetting the particles bulges up on them and pushes them down onto the microrelief, increasing friction and reducing mobility. In contrast, at large film thickness, microplastic particles may be entirely engulfed into the liquid and not touch the microrelief at all, reducing friction and increasing mobility. So although the flow velocity at the surface is faster (proportional to $\sin(\alpha)^{1/3}$, with $\alpha$ being the inclination) at larger inclination, the friction between the particles and the plate is increased and particles are less mobile overall. The roughness changes the flow conditions and influences the flow rate. Therefore, this relationship holds only approximately.

Figure 5 shows the emerging flow pattern after 0.1 s of simulation, illustrated with colored passive tracer point-particles. These are passively advected points without influence on the flow itself. They are illustrated as colored pixels to show the magnitude of local velocity.
Although the microrelief has no directionality in itself, the flow pattern shows distinctive channels with increased flow velocity on the scale of the microrelief, while in teardrop-shaped regions behind larger bumps of the microrelief, flow velocity is significantly reduced. This pattern is stationary. The asymmetry of the flow pattern indicates deviations from purely laminar flow. There are regions where particles will move down the plate with increased probability (fast flow channels) while in other regions the hydrodynamic force can never overcome friction.

At the low flow rates in the experiment, there are only little surface instabilities (non-stationary, bow-shaped, capillary waves) present. These mainly occur at higher flow rate and larger film thickness [71–77].

On top of the microrelief, the surfaces also possess a macrorelief. The latter emerged during construction of the plates when sand was glued to the chipboards. In regions where there is more glue, the sand layer may be thicker, and in other regions the sand layer may be thinner. This macrorelief also creates preferential flow channels (on a larger scale) where PMMA particles are transported particularly far. Figure 6 shows that on the coarse surface, particles starting on the right moved the farthest, while on the fine surface, preferential flow existed on both the left and right sides.

These macroscopic flow channels would be much more difficult to model in a simulation, since on the one hand both micro- and macrorelief would have to be simultaneously resolved on very different length scales and on the other hand the exact topography is of key importance because it directly affects experimental results. So to confirm those preferential macroscopic flow channels in the experimental setup, we used the recordings of methanol solution stained with Nile Red. Figure 7 confirms the preferential flow paths on the macro scale on the fine surface. The imaged part in the videos is slightly shifted compared to the imaged part in the experiments because the experimental setup had to be moved under the fume hood. The videos are provided in the Supporting Information Additional files 2 and 3.

In the video of the fine surface, we also observed some surface ripples, mainly along the preferential flow channels in the macrorelief. These may be caused by the locally increased flow rate, but could also be a result of the irrigation system perturbing the surface. Such surface ripples could temporarily increase the hydrodynamic force enough to unlock a microplastic particle from the microrelief and initiate the motion.

**Transport processes and environmental implications**

In our experiments, we observe that the transport of the PMMA particles is based on two different phenomena: transport-inhibiting interaction with the microrelief and transport-enhancing interaction with preferential flow channels of the macrorelief. In the first case flow patterns with areas of larger flow velocities and therefore larger hydrodynamic forces acting on the particles emerge due to the microrelief. The size of PMMA particles is at a similar scale as the microrelief sand grains and generally larger than the water film. This means that the particles on the one hand are able to interlock with the microrelief and are not transported, and on the other hand have constant friction if rolled down the plate by the flow, increasing the probability to lock into the microrelief again and get stuck after a short travel distance. Therefore, most particles did not move at all or travelled only a very short distance. Second, the macrorelief that overlays the microrelief provides macroscopic preferential flow channels for the particles. If a particle is located in or reaches a preferential flow channel, the travelled path lengths are large.

Under natural conditions, the fast flow pathways (both micro and macro) will potentially lead to increased erosion, carving channels into the soil surface [78, 79]. Gomez et al. [78], for example, observed a larger erosion on rougher surfaces and attributed this to “concentration of flow around roughness elements”. This is comparable to the flow patterns around rough surface elements that we obtained in the simulation on the microscale and the observed preferential flow paths on the macroscale. This kind of ‘flow concentration’ could finally lead to rill erosion [80]. Additionally, under natural conditions, interrill erosion due to the detachment of particles by raindrop impact and transport to rills (i.e. preferential pathways), an interplay between overland flow, water infiltration and surface sealing could be important processes affecting particle transport. At least in the beginning, interrill erosion could be size-selective and small particles are eroded preferentially [81, 82]. However, the exact distribution of eroded particles for a particular soil depends on different soil properties like particle size distribution or aggregate stability [83, 84]. The general dependence of soil erodibility is reflected in the well known Revised Universal Soil Loss Equation (RUSLE) [85]. It is still unclear whether erosion processes would affect microplastic particles in the same way as natural sediments because of the broad range of shapes, densities and sizes of microplastics [7, 8].

In the experiments and the simulation investigated here, erosion and infiltration are explicitly prevented. Furthermore it should be considered that the irregular PMMA particles of a mean particle length of 1215 μm that we used to conduct our experiments are only one exemplary polymer and microplastics shape. Because microplastics comprise a large range of different particle sizes, densities, shapes and degradation stages, our results can reflect a small part of the reality only [7, 8]. Particles of different shapes (e.g. spheres, fibres or films) and with different
surface properties (e.g. due to degradation or biofilm) might interact differently with rough surfaces and be (re)mobilized and transported in a different way.

Nevertheless, the results of our laboratory setup contribute to understanding the horizontal transport of microplastics in nature. Sandy sediment and soil surfaces with sparse vegetation cover resemble to a certain extent to our surfaces. Comparable environments can be found for example in floodplains and on riverbanks [86, 87], on agricultural fields, in clearing areas, in freshly developed building sites and in ruderal habitats [88]. Because rivers are important pathways of microplastics [13, 89], the adjacent riparian zones and floodplains act as an interface between aquatic and terrestrial ecosystems. Therefore, they play an important role in the distribution of microplastics within the “plastic cycle”, a term coined by Horton et al. [20] to emphasize the connectivity between environmental compartments. During runoff events caused by floods or precipitation excess, microplastics can be (re-)mobilised, transported and accumulated in these highly dynamic areas [44, 87, 90].

The transport parameters of microplastics under natural conditions such as the flow velocities necessary to move the particles, the influence of land cover (e.g. vegetation) and sedimentation rates [90] are still not deciphered. While Nizzetto and colleagues [31] suggested that microplastics moved similarly to natural particles and organic matter, there is still a significant lack of data to understand the transport patterns of microplastics “across the compartments of hydrological catchments” [44]. Analysing transport patterns under laboratory conditions may facilitate future studies on microplastic distribution and mobility. For future research additional important factors, e.g. topography, hydrology, land cover and the exposure of microplastics to physical, biological and chemical processes should be considered [44] in order to improve the understanding of microplastic transport and the resulting contamination of the environment.

Conclusions

We developed a laboratory setup to reliably trace fluorescent microplastic particles in real-time during irrigation experiments. By limiting the experimental variables to irrigation rate, inclination and surface roughness the driving factors of the particle movement could be deciphered. Especially the roughness and the irrigation rate turned out to be important. In our experiments, we could show that the transport of the microplastics was inhibited by the interaction of the microplastics with the microrelief and enhanced by preferential flow channels of the macrorelief of the rough surfaces. The computer simulated flow patterns showed variable flow velocities on the scale of the microrelief and thus spatially variable hydrodynamic forces acting on the particles. Our laboratory results are a first step to gain a better understanding of the horizontal transport of microplastics on natural sediment and soil surfaces. However, microplastics are a diverse group of contaminants with varying shapes, densities and sizes. This continuity of properties, alteration of the microplastic particles’ surface properties as a consequence of biological and chemical processes in the environment and additional factors influencing soil surface, e.g. topography or land cover, could not be taken into account here and need to be part of further research.

Abbreviations

PMMA: Polymethylmethacrylate; sCMOS: Scientific complementary metal–oxide–semiconductor; CCD: Charge-coupled device
Supplementary Information

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Additional file 1: A pdf file with Supplementary Figures referenced in the main text.

Additional file 2: A video of runoff stained with Nile Red on the coarse surface. The surface was inclined by 7.5 degrees and irrigated with 7.2 L h⁻¹.

Additional file 3: A video of runoff stained with Nile Red on the fine surface. The surface was inclined by 2.5 degrees and irrigated with 7.2 L h⁻¹.

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Authors’ contributions

CB, HL and MK designed the study, HL supervised the experimental work, ML simulated the water flow, MK did the irrigation experiments, MGJ prepared and characterized the PMMA particles, CB wrote the code and analysed the image data, all authors discussed the results, HL, CB and ML wrote the first draft of the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets and code used and/or analysed during the current study are published on Zenodo, https://doi.org/10.5281/zenodo.4911185.

Declarations

Competing interests

The authors declare that they have no competing interests.

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