Process optimization and evaluation of novel baicalin solid nanocrystals

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Abstract: The objective of this study was to prepare baicalin solid nanocrystals (BCN-SNS) to enhance oral bioavailability of baicalin. A Box–Behnken design approach was used for process optimization. The physicochemical properties and pharmacokinetics of the optimal BCN-SNS were investigated. Multiple linear regression analysis for process optimization revealed that the fine BCN-SNS was obtained wherein the optimal values of homogenization pressure (bar), homogenization cycles (cycles), amount of TPGS to drug (w/w), and amount of MCCS to drug (w/w) were 850 bar, 25 cycles, 10%, and 10%, respectively. Transmission electron microscopy and scanning electron microscopy results indicated that no significant aggregation or crystal growth could be observed in the redispersed freeze-dried BCN-SNS. Differential scanning calorimetry and X-ray diffraction results showed that BCN remained in a crystalline state. Dissolution velocity of the freeze-dried BCN-SNS powder was distinctly superior compared to those of the crude powder and physical mixture. The bioavailability of BCN in rats was increased remarkably after oral administration of BCN-SNS (P < 0.05), compared with those of BCN or the physical mixture. The SNS might be a good choice for oral administration of poorly soluble BCN, due to an improvement of the bioavailability and dissolution velocity of BCN-SNS.

Keywords: baicalin, solid nanocrystals, optimization, in vivo/vitro evaluation

Introduction

Baicalin (BCN; 7-d-gluconic acid, 5,6-dihydroxy flavone) (Figure 1), one of the major bioactive flavone glucuronides present in the radix of Scutellaria baicalensis, is generally used in traditional Chinese medicine as a remedy for the treatment of inflammation, fever, and allergic diseases.1 Due to the glycosyl group on the ring, BCN is poorly absorbed after oral administration, which causes low bioavailability and limits its therapeutic efficacy and clinical application.2,3 Because of its poor water solubility, the oral administration of BCN often results in erratic pharmacological activity. The poor absorption issue of poorly water-soluble drugs is known as a severe challenge for pharmaceutical development for improving drug therapeutic effectiveness. Nanonization technology is a promising formulation strategy for poorly water-soluble drugs, which could have some appealing advantages, such as increased saturation, solubility, or drug-dissolution velocity, compared with coarse or micronized drug powder.4–6 So far, various nanonization formulations of BCN have been used to improve its bioavailability, such as nanoparticles, liposomes, and nanoemulsions.7–9 However, these approaches still had some disadvantages, such as poor entrapment efficiency and low drug-loading, and were not able to give satisfactory results for BCN as expected.
Recently, nanosuspension (NS) technology has been successfully used to tackle the formulation problem of poorly soluble drugs.\textsuperscript{10,11} Remarkable NS properties, such as increased drug solubility and drug-loading capability (Figure 2), simple production process, easy scale-up, and universal adaptivity enable its applications in the formulation of large amounts of poorly soluble compounds. Furthermore, to improve patient convenience and physical stability issues of NS,\textsuperscript{12} it is usually preferable to transform liquid NS (LNS) into solid nanocrystals (SNS) via freeze-drying, spray-drying, or vacuum-spraying, and SNS can be redispersed into fine NS instantaneously with aqueous media in vitro or in the gastrointestinal tract.\textsuperscript{13}

Commonly, NS techniques are classified as bottom-up methods\textsuperscript{14,15} or top-down methods,\textsuperscript{16,17} according to the differences in production principle. High-pressure homogenization technology is a promising method for preparing NS. However, there are many process factors that can affect the physicochemical characterizations of the obtained NS, such as particle size and particle-size distribution. Some of these factors are stabilizer or polymer amount, homogenization pressure, and homogenization circles.\textsuperscript{18} How to determine which actual values of these variables result in a response value that is near the optimum? Box–Behnken design methodology is a well-known approach for formulation-process optimization, which can get the most information from a few well-designed experiments.\textsuperscript{19} Box–Behnken design has been comprehensively adopted to optimize the process of formulations.\textsuperscript{20–23} Those studies demonstrated the apparent advantage of Box–Behnken design utilization in the formulation-optimization process.

The main objectives of this study were: (1) to prepare acceptable BCN-NS, the Box–Behnken design approach was used for optimization of process variables, such as amounts of microcrystalline cellulose and carboxymethyl cellulose sodium (MCCS) or d-\textalpha-tocopherol polyethylene glycol 1000 succinate (TPGS), homogenization pressure, and homogenization circles; (2) to get BCN-SNS with good redispersibility, acceptable BCN-NS was freeze-dried; furthermore, the redispersibility of BCN-SNS after freeze-drying was investigated; (3) to systematically evaluate the physicochemical properties of BCN-SNS by particle-size analysis, transmission electron microscopy (TEM), differential scanning calorimetry (DSC), and powder X-ray diffraction (XRD). Furthermore, the dissolution and pharmacokinetic characteristics of BCN-SNS were also evaluated after oral administration.

**Materials and methods**

**Materials**

BCN was purchased from Zelang (>98%, Nanjing, People’s Republic of China). TPGS was purchased from Xi’an Healthful Biotechnology (Xi’an, People’s Republic of China). MCCS (Ceolus RC-A591 NF; Asahi Kasei, Tokyo, Japan) was commercially obtained.

**Methods**

**Nanosuspension production**

BCN-NS was prepared via high-pressure homogenization. Before producing BCN-NS, BCN coarse powder 1% (w/v), MCCS, and TPGS were dispersed in 100 mL water. The obtained mixture was disintegrated into coarse suspension by a high-shear homogenizer (FA25; Fluko, Shanghai, People’s Republic of China) at 16,000 rpm for 5 minutes. Then, the gained coarse BCN suspension was homogenized at predetermined pressure using a piston-gap high-pressure homogenizer (AH-1000D; ATS Engineering, Seeker, Canada). The premilling step was firstly carried out at 200 bar for five cycles, and then the BCN-NSs were homogenized at 600–1000 bar for 10–30 cycles.
Box–Behnken design

Independent and response variables

Preliminary screening trials were carried out for evaluating the formulation process parameters of BCN-NS. Such factors as homogenization pressure, homogenization cycles, amount of TPGS to drug, and amount of MCCS to drug were identified as critical to give a product in the nanorange and with the required stability. Results from the preliminary screening trials suggested that homogenization pressure, homogenization cycles, amount of TPGS to drug, and amount of MCCS to drug were the main factors that significantly affected particle size and zeta potential of the BCN-NS.

To reduce the number of trials and attain the most information on properties of the fine NS, the screening was planned applying a Box–Behnken design. According to the principles of Box–Behnken design, homogenization pressure (A, bar), homogenization cycles (B, cycles), amount of TPGS to drug (C, %, w/w), and amount of MCCS to drug (D, %, w/w), defined as independent values, were evaluated as four response values (Table 1), and the mean diameter (Y1, µm), span (Y2), and stability index (Y3) were defined as response values in the mathematical modeling, respectively. Each of the 29 formulations of a trial was produced three times in order to estimate the precision of the amount of TPGS to drug, amount of MCCS to drug, homogenization pressure, and homogenization cycles (see Table 2).

ANOVA analysis of model

A statistical model with interactive and polynomial terms was used to evaluate the response values using the formula:

\[
Y_{ij} = b_0 + \sum_{i} b_i x_i + \sum_{ij} b_{ij} x_i x_j + \sum_{ij} b_{ij} x_i^2 x_j + \sum_{ij} b_{ij} x_i^2 + \sum_{ij} b_{ij} x_i^2 + \varepsilon
\]

Table 1 Process variables and their levels used in the Box–Behnken design

| Levels    | Low | Medium | High |
|-----------|-----|--------|------|
| Coded values | -1  | 0      | 1    |
| **Independent variables** |     |        |      |
| A = homogenization pressure (bar) | 600 | 800    | 1,000 |
| B = cycle numbers | 10  | 20     | 30   |
| C = amount of surfactant to drug (w/w, %) | 5   | 10     | 15   |
| D = amount of polymer to drug (w/w, %) | 5   | 10     | 15   |
| **Dependent variables** |     |        |      |
| Y1 = particle size (nm) |     |        |      |
| Y2 = span |     |        |      |
| Y3 = stability index |     |        |      |

Table 2 The Box–Behnken design and the corresponding responded measurements

| Run | A   | B   | C   | D   | Y1  | Y2  | Y3  |
|-----|-----|-----|-----|-----|-----|-----|-----|
| 1   | -1  | -1  | 0   | 0   | 1.369 | 1.738 | 0.612 |
| 2   | -1  | 1   | 0   | 0   | 1.387 | 1.273 | 0.637 |
| 3   | -1  | 0   | 0   | -1  | 1.417 | 1.349 | 0.511 |
| 4   | -1  | 0   | 0   | 1   | 1.383 | 1.53   | 0.577 |
| 5   | -1  | 0   | -1  | 0   | 1.454 | 1.225 | 0.596 |
| 6   | -1  | 0   | 1   | 0   | 1.105 | 1.336 | 0.592 |
| 7   | 0   | 0   | -1  | -1  | 0.714 | 1.621 | 0.637 |
| 8   | 0   | 0   | 1   | -1  | 0.574 | 1.644 | 0.677 |
| 9   | 0   | 0   | -1  | 1   | 0.603 | 1.591 | 0.728 |
| 10  | 0   | 0   | 1   | 1   | 0.708 | 1.672 | 0.795 |
| 11  | 0   | -1  | -1  | 0   | 1.226 | 2.818 | 0.754 |
| 12  | 0   | -1  | 1   | 0   | 0.655 | 1.515 | 0.734 |
| 13  | 0   | 1   | -1  | 0   | 0.736 | 2.839 | 0.782 |
| 14  | 0   | 1   | 1   | 0   | 0.566 | 1.569 | 0.792 |
| 15  | 0   | -1  | 0   | -1  | 0.879 | 2.877 | 0.621 |
| 16  | 0   | 1   | 0   | -1  | 0.67  | 1.681 | 0.653 |
| 17  | 0   | -1  | 0   | 1   | 0.915 | 2.758 | 0.754 |
| 18  | 0   | 0   | 0   | 0   | 0.625 | 1.536 | 0.715 |
| 19  | 0   | 0   | 0   | 0   | 0.541 | 1.468 | 0.733 |
| 20  | 0   | 0   | 0   | 0   | 0.503 | 1.58  | 0.742 |
| 21  | 0   | 0   | 0   | 0   | 0.506 | 1.617 | 0.751 |
| 22  | 0   | 0   | 0   | 0   | 0.516 | 1.599 | 0.738 |
| 23  | 0   | 0   | 0   | 0   | 0.588 | 1.655 | 0.741 |
| 24  | 1   | -1  | 0   | 0   | 0.535 | 3.251 | 0.642 |
| 25  | 1   | 1   | 0   | 0   | 0.542 | 2.272 | 0.757 |
| 26  | 1   | 0   | 0   | -1  | 0.513 | 2.617 | 0.615 |
| 27  | 1   | 0   | 0   | 0   | 0.602 | 2.642 | 0.728 |
| 28  | 1   | 0   | -1  | 0   | 0.588 | 2.783 | 0.686 |
| 29  | 1   | 0   | 1   | 0   | 0.539 | 2.631 | 0.777 |

where \( Y \) represented the response variables, \( x \) represented the variables of the system, \( i \) and \( j \) were design variables, \( b \) represented model coefficients that were estimated using a least square fit of the model to the experimental results obtained during the design runs, and \( \varepsilon \) was the error.

To ensure a desirable model, test for significance of the regression model, test for significance on individual model coefficients, and test for lack of fit were performed. In order to summarize the tests performed, analysis of variance (ANOVA) was commonly used. The fitted model is adequate if the model is significant and the lack of fit is not significant.

Optimization

The desirability-function approach can be used to optimize the four response variables simultaneously.\(^{23}\) The desirability
function was achieved with Design-Expert (Stat-Ease, Minneapolis, MN, USA). Each response variable can be assigned an importance relative to the other responses. Importance \( r_i \) varies from the least important (\( \approx \)), a value of 1, to the most important (\( \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv \equiv 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Powder X-ray diffraction
The characteristics of the stabilizers, coarse BCN powder, BCN-SNS powder, and physical mixture were analyzed by powder XRD (D8 Advance; Bruker Optik, Ettlingen, Germany) with Cu source of radiation. Measurements were carried out at a voltage of 40 kV and 25 mA. The scanned angle was performed from 3°<20<60°, and the scanning rate was 2°/minute. Measurements were carried out in triplicate.

Solubility measurement of BCN-SNS
Equilibrium solubility of coarse BCN, BCN-SNS, or physical mixture was performed by shake-flask method.27,28 Excess amounts of coarse BCN, BCN-SNS, and physical mixture were added in water and equilibrated in a shaker water bath (HZ-92 1K; Taicang Instrument, Taicang, People’s Republic of China) at 37°C for 48 hours. Vials were sealed to avoid loss due to evaporation and shielded from light to prevent any degradation of BCN. As soon as equilibrium was reached, suspensions were firstly subjected to ultracentrifuge and then filtered through 0.22 µm syringe filters. An aliquot of filtrate was determined by high-performance liquid chromatography (HPLC) to evaluate the dissolved amount of BCN.

In vitro dissolution studies of BCN-SNS
Dissolution characterization of BCN-SNS, coarse BCN powder, and physical mixture containing the same amount of BCN (100 mg) was evaluated. According to the China Pharmacopoeia Appendix XC paddle method, a dissolution apparatus (RC-8; Tianjin Guoming Medicine and Equipment, Tianjin, People’s Republic of China) was used. Phosphate-buffered saline (900 mL, pH 7.4) at 37°C was used as a dissolution medium. The rotation speed of the paddles was set at 100 rpm. At predetermined time intervals (5, 10, 20, 30, 45, and 60 minutes), 2 mL samples were withdrawn and filtered through 0.22 µm filter membrane immediately. Simultaneously, equal blank medium was compensated immediately after withdrawal. The amount of dissolved BCN in the sample solution was assayed by HPLC.

Pharmacokinetic studies in vivo of BCN-SNS
Blood-sample preparation
Wistar rats (body weight 280 ± 20 g) were supplied by the Experimental Animal Center of Jiangxi University of Traditional Chinese Medicine (Nanchang, People’s Republic of China). The experimental protocol was approved by the University Ethics Committee for the use of experimental animals and conformed to the Guide for the Care and Use of Laboratory Animals. All rats were divided randomly into two groups of six animals each. Rats were fasted for 12 hours prior to the study. BCN was administered to them by gavages (30 mg/kg of body weight) in the form of either BCN suspension (dispersed in 10% [w/w] TPGS and MCCS aqueous solution) or BCN-NS. Blood samples of approximately 0.25 mL were collected by retroorbital puncture at predetermined intervals 0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 6, 8, 10, and 24 hours. Plasma was obtained from whole blood in heparinized tubes by centrifugation at 3,000 rpm for 15 minutes and frozen at −20°C until analysis.

To a 100 µL aliquot of plasma sample, 300 µL methanol and 300 µL acetonitrile were added and vortexed for 3 minutes in a 2.0 mL polypropylene tube. The mixture was centrifuged at 12,000 rpm for 10 minutes, and then the supernatant was transferred to a clean tube and evaporated to dryness under a nitrogen gas stream in a 37°C water bath. The dried residue was then redissolved in 100 µL of mobile phase and centrifuged at 12,000 rpm for 10 minutes, and the aliquot of the supernatant was injected into the HPLC system for analysis. The mobile phase was modulated to acetonitrile and 0.2% (v/v) aqueous phosphoric acid (47:53) at a flow rate of 1 mL/minute and a detection wavelength of 280 nm.

Statistical analysis
The plasma concentration-versus-time profile was analyzed by statistical software (DAS 2.0; Boying, Beijing, People’s Republic of China). A noncompartmental model was chosen to calculate the main pharmacokinetic parameters. The results were expressed as means ± standard deviation. Differences in pharmacokinetic parameters among the three groups were tested by one-way ANOVA. Values of P < 0.05 were considered statistically significant.

Results and discussion
Box–Behnken design
Table 2 shows the experimental conditions of the Box–Behnken design along with the corresponding values observed for the three response values. Experimental data were fitted to the cubic model by ANOVA. The ANOVA results for the three responses are shown in Tables 3–5. The results indicated that all the three models were significant at 95% confidence and the lack of fit was not significant. This
indicated that the fitted three models were adequate. The fitted models in terms of coded factors are demonstrated in Equations (3)–(5). Coefficient of determination ($R^2$) values for particle size, span, and SI are 0.9880, 0.9972, and 0.9936, respectively. The closer $R^2$ is to 1, the better the empirical models fit the actual data. Figure 3 shows surface-response graphs obtained using the fitted model.

\[ Y_1 = 0.59 - 0.42A - 0.16B - 0.084C - 0.0072AB + 0.075AC + 0.30A^2 + 0.13B^2 + 0.055C^2 + 0.16A^2B + 0.063AC^2 \]  
\[ Y_2 = 1.66 + 0.61A - 0.62B + 0.011C - 0.13AB - 0.066AC + 0.24A^2 + 0.42B^2 + 0.060C^2 + 0.26A^2B + 0.10AC^2 \]

Table 3 Analysis of variance for response surface quadratic model of $Y_1$

| Source   | Sum of square | df | Mean square | $F$       | $P$-value |
|----------|---------------|----|-------------|-----------|-----------|
| Model    | 3.04          | 17 | 0.18        | 53.16     | <0.0001   | Significant |
| A        | 1.41          | 1  | 1.41        | 420.6     | <0.0001   |            |
| AC       | 0.022         | 1  | 0.022       | 6.68      | 0.0254    |            |
| BC       | 0.04          | 1  | 0.04        | 11.94     | 0.0054    |            |
| A^2      | 0.66          | 1  | 0.66        | 194.55    | <0.0001   |            |
| C^2      | 0.034         | 1  | 0.034       | 10        | 0.0090    |            |
| D^2      | 0.051         | 1  | 0.051       | 15.23     | 0.0025    |            |
| B^2C     | 0.037         | 1  | 0.037       | 10.99     | 0.069     |            |
| BC^2     | 0.073         | 1  | 0.073       | 21.78     | 0.0007    |            |
| BD^2     | 0.034         | 1  | 0.034       | 10.19     | 0.0086    |            |
| Residual | 0.037         | 11 | 0.004       |           |           |            |
| Lack of fit | 0.032       | 7  | 0.005       | 3.68      | 0.1129    | Not significant |

Pure error 0.005 4 0.001
Total 3.08 28

\[ Y_3 = 0.74 + 0.066A - 0.0021B + 0.023C + 0.049D + 0.023AB + 0.024AC - 0.018BD - 0.086A^2 + 0.0039B^2 + 0.016C^2 - 0.051D^2 + 0.037A^2B - 0.029AB^2 \]

(5)

The results corresponding to the three response values are discussed as follows.

The effect of process parameters on the particle size of BCN-NS

Figures 3A–C show the surface-response plot of the effect of process parameters on the particle size of BCN-NS. The surface-response plot of interaction homogenization pressure × cycles (Figure 3A) shows that the particle size was attained with homogenization pressure between 600 and 1000 bar. This increased as power increased from 600 to 800 bar. Homogenization cycles significantly affected particle size (Figure 3A). As can be seen from Figure 3A, the particle size decreased from 1.454 µm to 0.503 µm as the homogenization pressure and homogenization cycles increased. As can be seen from Figures 3B and C, the amount of TPGS affected particle size; particle size decreased with increasing TPGS amount. Figure 3B demonstrates that particle size decreased as the TPGS amount increased to 15%.

The effect of process parameters on the span of BCN-NS

Figures 3D–F show the surface-response plot of the effect of process parameters on the span of BCN-NS. Figure 3D

Table 5 Analysis of variance for response surface quadratic model of $Y_3$

| Source   | Sum of square | df | Mean square | $F$       | $P$-value |
|----------|---------------|----|-------------|-----------|-----------|
| Model    | 0.16          | 19 | 0.008       | 73.48     | <0.0001   | Significant |
| A        | 0.016         | 1  | 0.016       | 144.74    | <0.0001   |            |
| C        | 0.005         | 1  | 0.005       | 41.46     | <0.0001   |            |
| D        | 0.02          | 1  | 0.02        | 181.65    | <0.0001   |            |
| AB       | 0.002         | 1  | 0.002       | 18.03     | 0.0022    |            |
| AC       | 0.002         | 1  | 0.002       | 20.90     | 0.0015    |            |
| BD       | 0.001         | 1  | 0.001       | 11.22     | 0.0085    |            |
| A^2      | 0.048         | 1  | 0.048       | 431.29    | <0.0001   |            |
| C^2      | 0.002         | 1  | 0.002       | 14.94     | 0.0038    |            |
| D^2      | 0.017         | 1  | 0.017       | 151.94    | <0.0001   |            |
| A^2 B    | 0.004         | 1  | 0.004       | 22.72     | 0.0003    |            |
| AB^2     | 0.002         | 1  | 0.002       | 12.27     | 0.0067    |            |
| Residual | 0.0001        | 9  | 0.0001      |           |           |            |
| Lack of fit | 0.0008      | 5  | 0.0002      | 3.85      | 0.1081    | Not significant |

Pure error 0.0002 4 0.00004
Total 0.16 28

\[ Y_3 = 0.74 + 0.066A - 0.0021B + 0.023C + 0.049D + 0.023AB + 0.024AC - 0.018BD - 0.086A^2 + 0.0039B^2 + 0.016C^2 - 0.051D^2 + 0.037A^2B - 0.029AB^2 \]
indicating that homogenization pressure and cycle significantly affected the span of BCN-NS. The optimum homogenization pressure was between 600 and 800 bar, while the optimum cycles were located between 20 and 25 cycles. Homogenization pressure had a significant effect on the span of BCN-NS, and the minimized span was achieved at pressure 600 bar for 20 cycles (Figure 3D). As can be seen from Figures 3E and F, the TPGS amount as well as the MCCS amount did not significantly affect the span of BCN-NS.

### The effect of process parameters on the stability index of BCN-NS

Figures 3G–I show the surface-response plot of the effect of process parameters on the SI of BCN-NS. Figure 3G shows that homogenization pressure more significantly affected the SI compared with homogenization cycles. The optimum homogenization pressure was located between 800 and 900 bar. As can be seen from Figure 3H, the SI was not significantly affected by the TPGS amount. Meanwhile, Figure 3I indicates that MCCS significantly affected the SI. The SI increased as the MCCS amount increased to 15%.

### Desirability optimization

The aim of optimization was to find good conditions that give the maximum SI and minimum particle size and span. The desirability-function approach was used to achieve this goal. Constraints for this optimization that were set in the software can be seen in Table 6. Figures 3J–L show the surface-response graphs of the desirability. The desirability was 0.77. The optimum values of homogenization pressure, cycles, TPGS amount, and MCCS amount were 850 bar, 25 cycles, 10%, and 10%, respectively.

### Verification

In order to evaluate the optimization capability of the models of the Box–Behnken design, the BCN-NSs were

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**Figure 3** (A–L) Fitted surface for response values as a function of the pressure (A), homogenization cycles (B), TPGS amount (C), and MCCS amount (D). (A–C) Particle size (D50); (D–F) span; (G–I) stability index; (J–L) desirability.

**Abbreviations:** MCCS, microcrystalline cellulose and carboxymethyl cellulose sodium mixture; TPGS, d-α-tocopherol polyethylene glycol 1000 succinate; SI, stability index.
prepared according to the optimal process-variable settings, wherein A, B, C, and D were equal to 850 bar, 20 cycles, 10%, and 10%, respectively. The particle size, span, and SI of BCN-NS obtained by predicted models are shown in Table 7. The results indicated that there was good agreement on preparation properties with theoretical predictions.

**Freeze-drying of BCN-NS**

It is well known that protectants such as polysaccharides were usually used to protect NS from the impairments of freezing and desiccation. However, apart from polysaccharides, the stabilizers were effective enough to prevent damage during freeze-drying. The results (Table 8) showed that there was no significant difference in RDI of BCN-SNS after freeze-drying, compared with that of BCN-SNS with different concentrations of sucrose. BCN-SNS stabilized with TPGS and MCCS possessed good redispersibility after freeze-drying. These results indicated that TPGS and MCCS were prominent stabilizers for BCN-NS, and played an important role in protecting BCN-NS from irreversible agglomeration during freeze-drying.

**Morphology of BCN-SNS**

Unsuitable freeze-drying procedure of NS can induce destabilization of the particles, leading to irreversible aggregation.

**Table 6** Constraints of factors and responses for optimization

| Name                  | Goal          | Lower limit | Upper limit | Importance |
|-----------------------|---------------|-------------|-------------|------------|
| Homogenization        | Is in a range | 800         | 1000        |            |
| pressure (bar)        |               |             |             |            |
| Homogenization cycles | Is in a range | 15          | 25          |            |
| TPGS amount (%)       | Is in a range | 10          | 15          |            |
| MCCS amount (%)       | Is in a range | 8           | 12          |            |
| Mean diameter (nm)    | Minimize      |             |             | 5          |
| Span                  | Minimize      |             |             | 3          |
| Stability index       | Maximize      |             |             | 4          |

**Table 7** Model-predicted and observed values of particle size, span and SI of BCN-LNS prepared according to the optimal experimental conditions (A = 850 bar, B = 25, C = 10%, D = 10%) (n = 5)

| Dependent variable   | Predicted | Observed | Bias/% |
|----------------------|-----------|----------|--------|
| Mean diameter (Y₁)   | 477.2 nm  | 483.4 nm | -1.29  |
| Span (Y₂)            | 1.307     | 1.243    | 4.89   |
| SI (Y₃)              | 81.48%    | 80.56%   | 1.12   |

**Table 8** The RDI of BCN-SNS with different concentrations of sucrose after freeze-drying

| Protectants | Sucrose (relative to the weight of baicalin, %) | RDI       |
|-------------|-----------------------------------------------|-----------|
|             | 0                                             | 1.07 ± 0.06|
|             | 50                                            | 1.05 ± 0.06|
|             | 100                                           | 1.11 ± 0.07|
|             | 200                                           | 1.04 ± 0.07|

**Abbreviations**: RDI, redispersibility index; BCN-SNS, baicalin solid nanocrystals.

**Note**: Values are mean ± standard deviation.

Therefore, morphology evaluation of the redispersed BCN-SNS is needed. In this study, the morphology of the freshly prepared BCN-NS and redispersed BCN-NS after lyophilization was evaluated by TEM (Figure 4). TEM images of freshly prepared BCN-NS (Figure 4A) and redispersed BCN-NS (Figure 4B) were similar. This experiment revealed that the particle size of the freeze-dried BCN-SNS was distributed homogeneously as single particles. The morphology of freeze-dried BCN-SNS was evaluated by SEM. As shown in Figure 5, the BCN-SNS stabilized by TPGS postlyophilization were flocculent in shape and formed slight aggregations (Figures 5A and B). However, no significant aggregation was observed in the redispersed BCN-NS. TPGS homogeneously adsorbed on the surface of the baicalin nanocrystals and effectively prevented possible polymer-chain entanglement during lyophilization. Therefore, a slight aggregation after lyophilization did not impair the redispersibility of BCN-SNS. This result also agreed well with particle-size values of redispersed NS from BCN-SNS determined by laser diffractometry (Figure 6). Therefore, the BCN-SNS after lyophilization possessed good redispersibility.

**Crystalline state analysis of BCN-SNS**

Crystalline state is a very important factor, which can influence the dissolution and stability behavior of a compound. High power density in the piston-gap homogenizer leads to high-energy input during the high-pressure homogenization process. In some cases, this may induce changes in the...
crystalline state, increasing the amorphous fraction or even creating completely amorphous particles. The extent of such changes depends on the chemical and on the physical hardness of the active ingredient, as well as on the applied power density. Therefore, before and following particle-size reduction, DSC and XRD analyses were conducted to assess if the initial crystalline state was preserved.

Figure 7 shows the XRD pattern of BCN-SNS. The powder XRD study of BCN-SNS showed no significant shift in the main peaks, compared with coarse BCN. The characteristic peak for BCN-SNS was observed at the same 2θ value as that of coarse drug. A slight decrease in intensity of peaks was observed with BCN-SNS.

The results of thermal analysis by DSC are shown in Figure 8. The diagram reveals a similarity in melting points. As shown in the DSC thermograms, the melting temperatures of BCN (coarse BCN and BCN-SNS) were similar. According to these results, neither the stabilizer nor the applied physical treatment affected the crystalline state of BCN.

From the DSC and XRD results (Figures 7 and 8), it was shown that BCN remained in a crystalline state that was physicochemically and thermodynamically more stable than BCN in an amorphous state.

**Solubility and dissolution-velocity studies of BCN-SNS**

NS could effectively enhance solubility for poorly water-soluble drugs. BCN-SNS possessed enhanced solubility (188.6 ± 0.33 µg/mL) compared with the solubility (53.51 µg/mL) of coarse BCN. As shown in Figure 9, dissolution velocities of the freeze-dried BCN-SNS were distinctly superior compared to those of the crude powder and physical mixture. Within 60 minutes, approximately 80.87% of BCN
Figure 7 X-ray diffraction pattern of coarse baicalin (A), MCCS (microcrystalline cellulose and carboxymethyl cellulose sodium mixture) (B), the physical mixture (C) and baicalin solid nanocrystals (D).

Figure 8 Differential scanning calorimetry thermograms of MCCS (A), TPGS (B), coarse baicalin (C), the physical mixture (D), and baicalin solid nanocrystals (E). Abbreviations: MCCS, microcrystalline cellulose and carboxymethyl cellulose sodium mixture; TPGS, d-α-tocopherol polyethylene glycol 1000 succinate.

Figure 9 Dissolution profiles of baicalin coarse powder (A), physical mixture of baicalin (B), and freeze-dried baicalin solid nanocrystal powder (C).
Process optimization and evaluation

was dissolved from the freeze-dried BCN-SNS. In contrast to this, only 41.15% and 43.37% was dissolved from the coarse BCN and physical mixture, respectively. We concluded that particle-size reduction led to enhanced dissolution velocity. This could be explained by the Noyes–Whitney equation.33,34 As when drug particles are smaller, the corresponding surface area is larger, so the nanoparticles provided a large increase in the surface area. Furthermore, according to the Prandtl equation,35 the diffusion distance, \( h \), decreases for very small particles. The increase of surface area and the simultaneous decrease of diffusion distance could lead to enhancement in dissolution velocity of the drug.

Pharmacokinetic study in rats of BCN-SNS

To confirm the advantage of NS in improving the bioavailability of BCN, an in vivo pharmacokinetic study of BCN-SNS was performed in rats. The plasma concentration–time curves of the BCN coarse suspension (BCN-CS) and BCN-SNS are displayed in Figure 10, and the pharmacokinetic parameters are shown in Table 9. After oral administration of these three different BCN formulas, their individual mean plasma concentration–time curve profile showed a biphasic absorption phenomenon. The first absorption peak presented at about 30 minutes, and then a second absorption peak occurred at about 6 hours. This might be explained by enterohepatic circulation of BCN, wherein BCN was hydrolyzed to baicalein by intestinal bacteria and then restored to its original form from the absorbed baicalein in the body, resulting in the second absorption peak.36,37

The results showed that coarse BCN and NS were different from each other in the corresponding parameters. Compared with the BCN-CS, the BCN-SNS had higher maximum concentration (\( C_{\text{max}} \); 5.808 ± 0.785 mg/L), higher area under the curve, 0 to time \( t \) (\( \text{AUC}_{0–t} \); 73.384 ± 8.1847 mg/L · hour), and shorter mean residence time (10.758 ± 1.348 hours), which indicated BCN-NS was easier to be absorbed in vivo.

BCN-NS was absorbed easily, which induced an increase in \( \text{AUC}_{0–t} \) and \( C_{\text{max}} \) and decreased mean residence time. These phenomena were probably caused by the absorption pattern of BCN. Greater exposure of the drug

![Graph showing mean plasma concentration-time curve of (A) baicalin coarse suspension, (B) baicalin physical mixture, and (C) baicalin solid nanocrystals after oral administration (n = 6).](image-url)
function size has been recognized as a crucial parameter for bioadhesion and absorption from gastrointestinal tissue. The results showed that drug NS had the higher dissolution rate compared with the coarse drug. Because drug NS had a greater surface area compared with the coarse particles drug under equivalent conditions, they dissolved easily into intestinal fluid, and smaller particles showed a higher extent of uptake than larger ones via both follicle-associated epithelia and absorptive enterocytes.

Conclusion

Process optimization of NS prepared by high-pressure homogenization was a complex process, since it involved a large number of factors that could affect the characteristics of NS. The Box–Behnken design was applied successfully to optimize formulation and process parameters for NS. From this study, it was concluded that homogenization pressure, homogenization cycles, amount of TPGS to drug, and amount of MCCS to drug played a significant role in controlling the particle size, span, and SI of NS. Homogenization pressure and homogenization cycles were considered to be significant factors that affected particle-size D50 and span of NS. The graphical optimization method helped in finding the “sweet spot” or design space to get NS with the desired physicochemical properties. This study also helped in identifying certain formulation and processing parameters, such as high polymer concentration and high homogenization pressure, which may affect the manufacturing of NS on a larger scale. The SI of the NS was found to be more dependent on polymer concentration compared to surfactant concentration.

The fabricated BCN-NS by optimization presented a sphere-like shape under TEM. SEM results demonstrated that no significant aggregation or crystal growth was observed in the redispersed BCN-SNS. By reducing BCN particle size into the nanometer range, its dissolution rate and oral bioavailability exhibited greater enhancement compared with coarse BCN. Through this study, it has been demonstrated that SNS might be a good choice for oral administration of poorly soluble BCN.

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Disclosure

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