Review

Gut Microbes and Neuropathology: Is There a Causal Nexus?

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Abstract: The gut microbiota is a virtual organ which produces a myriad of molecules that the brain and other organs require. Humans and microbes are in a symbiotic relationship, we feed the microbes, and in turn, they provide us with essential molecules. Bacteroidetes and Firmicutes phyla account for around 80% of the total human gut microbiota, and approximately 1000 species of bacteria have been identified in the human gut. In adults, the main factors influencing microbiota structure are diet, exercise, stress, disease and medications. In this narrative review, we explore the involvement of the gut microbiota in Parkinson’s disease, Alzheimer’s disease, multiple sclerosis and autism, as these are such high-prevalence disorders. We focus on preclinical studies that increase the understanding of disease pathophysiology. We examine the potential for targeting the gut microbiota in the development of novel therapies and the limitations of the currently published clinical studies. We conclude that while the field shows enormous promise, further large-scale studies are required if a causal link between these disorders and gut microbes is to be definitively established.

Keywords: brain-gut-microbiota axis; probiotics; psychobiotics; prebiotics; faecal microbiota transplantation; neurological disorders

1. Introduction

The adult human brain weighs around 1.5 kg, approximately the same as the gut microbiota. The number of bacteria in the microbiota is similar to the number of cells in the human body [1]. However, there is greater DNA content with a larger number of genes in the microbiota than in the human cell. Humans and microbes have co-evolved in a symbiotic relationship, and while we feed the microbes, in turn, they produce molecules that our brains and other organs require for survival. If all the microbial DNA required by us was directly incorporated in our cells, there would be inadequate space for such material. Thus, the symbiotic relationship is essential for our physiological functioning. Any significant derangement of the gut microbiota architecture can impact brain function and result in neuropathology [2].

2. Microbiota Structure

Advances in DNA sequencing and bioinformatics have enabled an increased understanding of gut microbiota architecture and function [3]. Bacteria are by far the dominant microbe within the microbiota architecture, but other microbes include viruses, fungi and archaea [4]. Currently, there are 25 recognised bacterial phyla, which include Firmicutes, Bacteroidetes, Actinobacteria, Cyanobacteria, Fusobacteria, Proteobacteria and Verrucomic. It is estimated that Bacteroidetes and Firmicutes phyla account for around 80% of the total gut microbiota. The genera within the Firmicutes phylum include Clostridium, Lactobacillus, Bacillus, Clostridium, Enterococcus and Ruminococcus. The Bacteroidetes phylum predominantly consists of the Bacteroides and Prevotella genera. The Actinobacterium phylum is dominated by the Bifidobacterium genus [5].

Approximately 1000 species of bacteria have been identified in the human gut. There remains a lack of consensus as to the structure of a normal gut microbiota and hence...
as to a definition of dysbiosis [6]. This lack of consensus is due to the significant inter-individual differences seen in the microbiota structure. However, it is generally accepted that greater microbial diversity in adulthood together with high levels of resistance and resilience are associated with a healthy state. Furthermore, some genera are classified as beneficial symbionts, while others have been classified as potential pathogens. An altered ratio of symbionts to pathogens may increase disease vulnerability. Bifidobacteria and lactobacilli are generally viewed as beneficial bacteria and are frequently sold as probiotic supplements (live biotherapeutics). Strains within the *Clostridium* genus or lipopolysaccharide (LPS)-forming taxa such as Enterobacteriaceae have been associated with disease states [7]. Though somewhat controversial, the Firmicutes/Bacteroidetes ratio, exploring the relationship between the two dominant phyla, has been connected with various pathologies [8]. It has been argued that exploring the functional state of the gut microbiota is of greater value in studying disease than describing the overall architecture.

In adulthood, the core gut microbiota is relatively stable. The main factors influencing structure are diet, exercise, stress, disease and medications [9]. It is generally accepted that a Mediterranean diet rich in fruit, vegetables, nuts, fermented foods and fish yields an optimal microbiota [10]. Up to 80% of medications prescribed by clinicians impact the gut microbiota, and many in a negative manner [11]. Broad-spectrum antibiotics have a dramatic impact, but after such treatment, the microbiota does gradually revert to a pre-antibiotic state. This may take up to twelve weeks.

The gut microbiota decreases in diversity with ageing [12]. Healthy elderly people tend to maintain microbial diversity, unlike those who age in an unhealthy manner. These significant shifts in microbiota composition may be at the heart of many elderly disease processes, including those involving the brain.

3. Microbe to Brain Communication

All the important neurotransmitters can be produced by microbes [13]. These neurotransmitters include GABA, norepinephrine, dopamine, serotonin and histamine. GABA, for instance, is produced to varying degrees by all lactobacilli [14]. However, even when produced by gut bacteria, it is highly unlikely that these neurotransmitters travel to and influence the brain. However, they may affect the enteric nervous system and other brain communication networks such as the vagus nerve. In the case of serotonin, it has been shown that bifidobacteria increase plasma tryptophan levels [15]. The latter is the building block of serotonin, and the human brain has limited storage capacity. It seems likely that diet and bifidobacterial synthesis play an important role in maintaining normal central serotonergic function. If diet was the sole source of tryptophan, it is likely that several days of starvation would have a profound effect on mood. In such circumstances, microbial production provides an important alternative to dietary tryptophan.

The routes of communication between gut microbes and the brain are not fully elucidated but do include neural, endocrine, immune and metabolic pathways (Figure 1) [16]. The vagus nerve, a long meandering nerve, is a major route of bidirectional information signalling between the gut and the brain [17]. Several important chemicals are produced in the gut and travel to the brain via the bloodstream. Many of these are manufactured by or have their production regulated by gut microbes. These include short-chain fatty acids, tryptophan, leptin and ghrelin. The short-chain fatty acids include butyrate, propionate and acetate. Butyrate is a known inhibitor of histone deacetylases (HDACs), and by doing so, acts as an epigenetic regulator [18]. It also has the ability to influence the G-protein coupled receptors, namely free fatty acid receptors [19], though these are sparsely distributed in the mammalian brain. Cytokines, key immune regulatory molecules produced at the level of the gut, can travel via the bloodstream and influence brain function, especially in brain regions where there is a deficient blood-brain barrier, such as the hypothalamus.
Figure 1. Connections within the brain-gut-microbiota axis, including the vagus nerve, and SCFAs such as butyrate, cytokines and tryptophan. Modified from Ref. [20] 2015 Springer Nature: More than a gut feeling: the microbiota regulates neurodevelopment and behavior. Neuropsychopharmacology 2015; 40: 241–242. Abbreviations: HPA, hypothalamic–pituitary–adrenal; CRH, corticotrophin-releasing hormone; ACTH, adrenocorticotropic hormone; GABA, gamma aminobutyric acid; SCFAs, short-chain fatty acids.

Over the past decade, several groups have examined the gut microbiota in a variety of neurological disorders, viewing such conditions as disorders of the brain-gut-microbiota axis, and there are several published reviews [21–24]. However, a limitation in most of these clinical studies is a failure to control for relevant variables such as diet, levels of exercise and medication usage. The impact of diet on the gut microbiota can be a significant confounder if not adequately controlled. This was clearly illustrated in the recent autism study by Yap et al. [25]. Here, we will examine the findings reported in some of the major neurological conditions. Most of the published reviews come from a preclinical perspective. In contrast, the current review begins from the angle of the clinician and translates back to the preclinical findings. We focus on Parkinson’s disease, Alzheimer’s disease, multiple sclerosis and autism because of their high prevalence in western society. Such a focus is not meant to diminish the studies in other therapeutic areas such as motor neurone disease and Huntington’s disease.

The selected studies were based on a PubMed search of the following terms: gut microbiota, brain-gut-microbiota axis and each of the individual diseases. The analysis is based on papers published until June 2022. Preclinical studies were chosen on the basis that they provided potential leads to clinical pathophysiology.
4. Parkinson’s Disease

Parkinson’s disease (PD), the ‘shaking palsy’, is named after James Parkinson, the East London surgeon who provided the first detailed description of the disorder in 1817 in his paper “An Essay on the Shaking Palsy”. He was probably not the first to describe the disease, which may originally have been recognised by the Roman physician Galen. The term Parkinson’s disease was coined by the neurologist Jean-Martin Charcot. The condition is characterised by slow movements (bradykinesia), resting tremor, rigidity and postural instability, and can profoundly impact quality of life. Its underlying pathology is degeneration of dopamine neurones in the zona compact region of the substantia nigra. Current therapies largely focus on enhancing dopamine transmission in the nigrostriatal pathways. However, while initially helpful in managing symptoms, such treatments usually lose efficacy over time. Alternative therapies are clearly required, and the gut microbiota is viewed by many as a suitable therapeutic target.

It is argued that PD occurs because of toxins produced within the gut by microbes, or alternatively as a failure of the microbiota to manufacture neuronal dopamine-specific nutrients. The protein alpha-synuclein has been implicated in the pathogenesis of PD. It is speculated that aggregation of alpha-synuclein is central to the pathophysiology of the disorder and that the dopaminergic neurones in the substantia nigra are majorly sensitive and as a result degenerate with such accumulation [26]. Evidence is accumulating to support the view that the alpha-synuclein originates in the gut and spreads to the brain like a prion, probably spreading along the vagus nerve. The molecule can form aggregates, regarded as disease biomarkers, and may be the vehicle for the spread of disease from the peripheral to the brain. Furthermore, alpha-synuclein is viewed as being involved in the autonomic dysfunction seen in many patients with PD. It may also play a role in the gastrointestinal symptoms frequently seen in the condition.

Vagotomy was a common surgical procedure for the treatment of peptic ulcers prior to the introduction of H2 antagonists and proton pump inhibitors. As a treatment, it was used for decades, and for many patients, it did improve symptoms, although not entirely without side effects. Most of the patients who underwent such surgery and who are still alive are now elderly. A Swedish register study [27] investigated the risk of PD in patients who underwent vagotomy and hypothesised that truncal vagotomy is associated with a protective effect. They found that the risk of PD was indeed decreased in patients who underwent full truncal vagotomy, while there was no risk reduction in those who underwent a super-selective vagotomy. Risk of PD is also decreased after truncal vagotomy when compared to the general population. These epidemiological findings are important and raise several issues requiring further exploration. Overall, the data support the hypothesis that PD commences in the gut and not initially in the brain. This provides further evidence for the involvement of the vagus nerve in the development of the disorder.

To characterise the expression of alpha-synuclein in the innervation of the gut, both the postganglionic neurones and the preganglionic projections, by which the disease is postulated to retrogradely invade the brain, were examined [28]. It was found that some vagal preganglionic efferents expressing alpha-synuclein formed varicose terminal rings surrounding myenteric plexus neurones that were also positive for the protein. This fundamental finding provides a plausible alpha-synuclein-expressing pathway, enabling the retrograde transport of PD pathogens from the enteric nervous system to the brain.

The gut microbiome in PD has been investigated in several research settings. For a comprehensive review, see [29]. Sheperjans and colleagues compared the faecal microbiomes of 72 PD patients and 72 control subjects using 16S ribosomal RNA sequencing [30]. They explored the associations between clinical parameters and microbiota bioinformatics. They found that the abundance of Prevotellaceae in faeces of PD patients was reduced by over 70% as compared with controls. The relative abundance of Prevotellaceae of 6.5% or less had 86% sensitivity and 39% specificity for PD diagnosis. A discriminant analysis based on the abundance of four bacterial families and constipation severity identified PD patients with 67% sensitivity and 90% specificity. Of note is the fact that the relative abundance of
Enterobacteriaceae was positively correlated with the severity of postural instability and gait difficulties. These findings indicate that the intestinal microbiome is altered in PD and is related to the motor phenotype. However, the possibility that these microbiota changes are entirely epiphenomenal cannot be entirely ruled out. The findings nonetheless raise the exciting prospect that alterations in the gut microbiota are a potential diagnostic marker.

Mazmanian’s group have added considerably to the literature. They used the alpha-synuclein overexpressing mouse to investigate the genesis of PD [31]. These mice are genetically engineered to overexpress alpha-synuclein, and they go on to develop core features of PD. This is currently the most widely used animal model of the disorder. Surprisingly, when these mice are raised germ-free, their propensity to develop motor anomalies is significantly reduced. If such animals are given a combination of short-chain fatty acids, they show microglial activation in the brain and aggregation of alpha-synuclein with the onset of motor features. The changes are prevented by treatment with the antibiotic minocycline, which acts on both Gram-positive and Gram-negative bacteria. When germ-free, alpha-synuclein overexpressing mice are given a humanised microbiota from a patient with PD, the resultant pathology is greater than that seen following transplant with the microbiota from a healthy subject. The findings suggest multiple potential lines of treatment, which include short-chain fatty acid antagonists, antibiotics and microbiota transplantation.

In a recently published study, Nishiwaki and colleagues investigated the manner in which short-chain fatty acid-producing and mucin-degrading intestinal bacteria predict the progression of early PD [32]. They report that decreases in short-chain fatty acid-producing genera, Fusicatenibacter, Faecalibacterium and Blautia, as well as an increase of the mucin-degrading genus Akkermansia, predicted accelerated disease progression. This requires replication, but if replicated may be an important finding, helping the development of new innovative therapies.

Xue et al. examined the impact of faecal microbiota transplant therapy in patients with PD [33]. Fifteen PD patients were included: ten received FMT via colonoscopy and five received FMT via a nasal-jejunal tube. Overall, an improvement in both motor and psychological symptoms was reported. The colonic FMT group showed significant improvement and longer maintenance of efficacy compared with nasointestinal FMT. None of the patients were satisfied with nasointestinal FMT for more than three months. Overall, adverse events were mild. This was a preliminary open-label study which certainly provides support for larger-scale, double-blind interventions.

Tamtaji et al. [34] conducted a randomised, double-blind, placebo-controlled trial, in 60 people with PD. Subjects were randomly divided into two groups. Group 1 received a probiotic at $8 \times 10^9$ CFU/day, and Group 2 received a placebo (n = 30 in each group) for 12 weeks. Consumption of the probiotic decreased neurological symptoms relative to those seen in the placebo group. The probiotic supplementation also reduced high-sensitivity C-reactive protein, insulin and malondialdehyde, and increased glutathione levels. The authors conclude that the therapy has potential benefits for patients with PD.

In summary, the gut microbiota shows promise as a potential target for intervention in patients with PD. See Table 1.

### Table 1. Microbiota changes in Parkinson’s disease.

| Reference                                      | ↓ or ↑ |
|------------------------------------------------|--------|
| Christensenella, Catabacter, Lactobacillus, Oscillospira, Bifidobacterium, Christensenella minuta, Catabacter hongkongensis, Lactobacillus mucosae, Ruminococcus bromii and Papillibacter cinnamivorans | ↑ [35,36]
| Dorea, Bacteroides, Prevotella, Faecalibacterium, Bacteroides massiliensis, Stoquefichus massiliensis, Bacteroides coprocola, Blautia glucerasea, Dorea longicatena, Bacteroides dorei, Bacteroides plebeus, Prevotella copri, Coprococcus eutactus and Ruminococcus callidus | ↓ [36]
| Clostridium cocoides, Bacteroides fragilis     | ↓ [37]
| Roseburia, Prevotella and Bifidobacterium      | ↓ [38] |
Table 1. Cont.

| Reference | Germ family or genus          |
|-----------|------------------------------|
| [39]      | Eubacterium ↓                |
| [40]      | Lactobacillaceae, Barnesiellaceae and Enterococcaceae ↑ |
| [41]      | Escherichia-Shigella, Streptococcus, Proteus and Enterococcus ↑ |
|           | Blautia, Faecalibacterium and Ruminococcus ↓ |
| [42]      | Bilophila and Paraprevotella ↑ |
| [36,43]   | Bifidobacteriaceae ↑ Lachnospiraceae ↓ |

5. Alzheimer’s Disease (AD)

AD is the most common cause of dementia and occurs in a senile and pre-senile form. The latter begins before the age of sixty-five, and the former occurs after this age. It was originally described in 1906 by the pathologist Alois Alzheimer and we now know that the neuropathology includes amyloid plaques and neurofibrillary tangles [44]. The World Health Organization (WHO) estimates that over 50 million people worldwide suffer from dementia, with AD accounting for 60–70% of all cases. In the USA alone, it costs the healthcare system over USD250 billion annually.

Germ-free mice who lack a gut microbiota have a marked absence of amyloid plaque and neuroinflammation. Transgenic mouse studies of AD have shown an altered microbiome [45,46]. AD histological and cognitive features in the APP/PS1 mouse model of AD correlated with gut microbiome alterations. Specifically, Helicobacteraceae and Desulfovibrionaceae at the family level and Odoribacter and Helicobacter at the genus level are increased relative to controls [47].

When APP/PS1 transgenic mice are treated with an antibiotic cocktail, a reduction in microglial and astrocyte accumulation surrounding amyloid plaques is observed. Furthermore, modulation of intestinal microbiota by the probiotic VSL#3 resets brain gene expression and ameliorates the age-related deficit [48]. With such animal findings, there is now an increasing focus on dietary interventions targeting the microbiome for the management of AD. In an animal model of AD, administration of *Bifidobacterium bifidum* and *Lactobacillus plantarum* for eight weeks with physical training had a positive effect. A decrease in amyloid-β protein and an improvement in spatial learning via an acetylcholine-mediated mechanism were found.

There are no prospective studies of the gut microbiota in patients with AD published so far. Cattaneo et al. [49] conducted a cross-sectional study which identified Escherichia/shigella bacterial taxa as increased in faecal samples from AD patients relative to control subjects. The data suggest that these taxa are associated with inflammation. Of note is the fact that microbiota changes correlate with pro-inflammatory blood cytokines levels. The authors suggest a causal link between dysregulation of the microbiota and systemic inflammation and speculate that such microbiota changes initiate or exacerbate the neurodegeneration in AD. See Table 2.

In a recent study, Wang et al. compared faecal microorganisms of patients with AD and healthy cohabiting caregivers [50]. They found that the abundance of several bacteria taxa in AD was altered at the genus level. The most notable changes were Anaerostipes, Mitsuokella, Prevotella, Bosea, Fusobacterium, Anaerotruncus, Clostridium and Coprobacillus. Surprisingly, Akkermansia, an available probiotic, was increased in the AD patients compared with controls. Additionally, the levels of Bifidobacteria were found elevated. The authors conclude that “patients with mild AD have unique gut microbial characteristics”. Such changes may present a novel target for therapeutic intervention.

Asaoka et al. [51] conducted a randomised clinical trial on 130 patients aged from 65 to 88 years old with mild cognitive impairment. They received either a daily probiotic (*B. breve* MCC1274, 2 × 1010 CFU) or a placebo for 24 weeks. The probiotic intervention produced marked changes relative to the placebo. The data suggest that the intervention suppressed
brain atrophy progression and ameliorated cognitive deterioration. Interestingly, the composition of the microbiota was not changed by the intervention.

Table 2. Microbiota changes in Alzheimer’s disease.

| Reference |
|-----------|
| Lachnocostridium, Bacteroides [52] |
| Enterococcaceae †, [52] |
| Akkermansia, Blautia, Dorea, Eggerthella, Streptococcus, Bifidobacterium, Lactobacillus † [53] |
| Alistipes, Bacteroides, Butyricimonas, Haemophilus, Parabacteroide † [53] |
| Enterobacteriaceae, Veillonellaceae † [54] |
| Clostridiaceae, Lachnospiraceae, Ruminococcaceae Genus: Blautia, Ruminococcus †, [54] |
| Bacteroidaceae, Rikenellaceae, Gemellaceae Genus: Blautia, Bacteroides, Alistipes, Bilophila, Gemella, Paraclostridium †, [55] |
| Ruminococcaceae, Bifidobacteriaceae, Clostridiaceae, Peptostreptococcaceae, Mogibacteriaceae, Turicibacteraceae Genus: Bifidobacterium, Dialister, Clostridium, Turicibacter, Adlercreutzii †, [55] |
| Escherichia, Shigella † [49] |

6. Multiple Sclerosis (MS)

MS is an autoimmune disorder associated with the demyelination of neurones in the brain and spinal cord [56]. The most common presentation shows a relapsing/remitting pattern, but some patients present with a more severe form of primary progressive disorder. The symptoms are multitudinous but usually involve movement, sensation or balance. Despite therapeutic advances, the condition can have a profound negative impact on quality of life.

Germ-free mice show increases in myelination patterns in the prefrontal cortex, suggesting that the gut microbiota plays a role in controlling myelination [57]. Furthermore, germ-free mice are especially resistant to developing experimental autoimmune encephalomyelitis (EAE), which is a well-established animal model of MS. Other studies indicate that the presence of certain Gram-positive segmented filamentous bacteria, known to activate Th17 cells, significantly alter EAE severity [58].

Navarro-López [59] analysed the gut microbiota of 15 patients with relapsing/remitting MS and compared samples with those from diet-matched healthy controls. They found that the gut microbiota of the MS group differed significantly from healthy controls in the levels of the Lachnospiraceae, Ezakiella, Ruminococcaceae, Hungatella, Roseburia, Clostridium, Shuttleworthia, Poephyromonas and Bilophila genera. The data provide support for the view that MS is associated with a dysbiosis and that Ezakiella and Bilophila genera in particular may be a risk factor in patients with relapsing/remitting MS. See Table 3.

FMT has been assessed in a preliminary study of patients with MS [60]. Nine patients with MS were recruited and underwent FMTs monthly for up to six months. The primary outcome measure was alteration in blood cytokines. The secondary outcomes were gut microbiota composition together with intestinal permeability. Microbiota enrichment was observed, and in two patients who had increased intestinal permeability at baseline, the FMT decreased the permeability. The study was preliminary and obviously limited by the small sample size. The overall conclusion is that FMT is safe in patients with MS.

Mirashrafi et al. [61] conducted a meta-analysis of probiotic trials where probiotics were used as adjunctive therapy in patients with relapsing/remitting MS. A total of 106 patients were randomly assigned to a probiotic treatment, and 107 were randomly assigned to a placebo. The probiotics used included Lactobacillus acidophilus, Lactobacillus casei, Bifidobacterium bifidum, and Lactobacillus fermentum, Bifidobacterium infantis, Bifidobacterium lactis and Lactobacillus reuteri. A significant improvement with probiotic supplementation
was found in the Expanded Disability Status Scale as well as on psychological measures. This meta-analysis is of interest but limited by the large variety of probiotics used and the different durations of therapy. A large-scale study with the most promising probiotic needs to be conducted.

Table 3. Microbiota changes in multiple sclerosis.

| References |
|------------|
| Dorea, Pedobacter, Flavobacterium, Prevotella, Parabacteroides, Collinsella ↓ | [62] |
| Methanobrevibacter, Akkermansia ↑ Butyriramonas, Prevotella ↓ | [63] |
| Prevotella ↓ | [64] |
| Streptococcus ↑ Prevotella, Faecalibacterium ↓ | [65] |
| Lawsonella ↑ Faecalibacterium Prausnitzii, Bacteroides fragiils, Eubacterium rectale, Blutyrirbrio, Clostridium, Coprococcus, Roseburia ↓ | [66] |
| Faecalibacterium, Ruminococcus ↑ Blautia, Anaerostipes, Bifidobacterium, Prevotella | [21,67] |

7. Autism

Autism is a disorder with an early age of onset, characterised by poorly developed social interactions, difficulty with receptive and expressive communication and the presence of repetitive behaviours [68]. Epidemiological evidence suggests that the prevalence of the disorder is on the rise, though this may represent changes in diagnostic patterns. Gastrointestinal disturbance is frequently co-morbid. Constipation is the most common symptom, followed by abdominal pain, diarrhoea and occasionally vomiting. Overall, the condition is more common in males than females, and in general autistic children grow into autistic adults, though early behavioural interventions can ameliorate symptoms.

In a recent paper, Lu et al. [69] argue that rebalancing the gut microbiota is the critical step in treating autism and the broader autism spectrum disorder (ASD). Germ-free studies provide support for the link between gut microbiota and autism. Such animals show behavioural changes similar to in autism [70]. When allowed to interact with another mouse or an inanimate object, germ-free mice are as likely to interact with the latter. Such behaviour is abnormal for socially interactive animals. If germ-free animals undergo conventional colonisation at an early stage, the emergence of behavioural changes can be prevented. In fact, the administration of single bacterial strains such as Bacteroides Fragilis or Lactobacillus reuteri can reverse many of the behavioural changes in these animals [71].

Numerous types of dietary interventions have been employed in treating autism, including the Mediterranean diet, gluten-free diet and ketogenic diet. For a review, see [72]. Here, we will focus our attention on directly impacting microbial interventions such as probiotics, which are sometimes referred to as psychobiotics when associated with the management of syndromes such as this with a behavioural component. A psychobiotic is defined as bacteria that, when ingested in adequate amounts, has a positive mental health benefit [73]. An intervention study found that Lactobacillus plantarum WCSF1 increased the number of Lactobacillus and Enterococcus bacteria in autism, and significantly decreased the level of Clostridium cluster XIVa, viewed as a harmful bacterium. More importantly, from the perspective of the patient, after the probiotic, the scores of destructive behaviour, anxiety, self-focused behaviour and poor communication were improved. A different strain of Lactobacillus plantarum (PS128) was shown to reduce the scores on the social responsiveness...
Lactobacillus acidophilus is also found to improve the concentration of ASD children. See Table 4.

Some groups have studied synbiotics, the combination of probiotic and prebiotic fibre. The latter promotes the growth of good bacteria. One month of treatment with the polybiotic combination Bifidobacterium infantis Bi-26, Lactobacillus rhamnosus HN001, Bifidobacterium lactis BL-04 and Lactobacillus paracasei LPC-37, together with fructooligosaccharide, brought about improvements in speech/language and overall social interaction [75]. This observation is supported by a recent systematic review which found that supplementing with multiple probiotic species or the addition of a prebiotic fibre produced a better result than the use of a single strain [76].

Given the potential of using the gut microbiota as a target [77], it is not surprising that FMT has been used as a potential therapy. FMT was found to improve behavioural problems and gastrointestinal symptoms in younger patients but not in those over twenty-one years [69]. In a further FMT study, Zhao et al. [53] conducted a randomised controlled study on 48 patients with ASD. The study was not double-blind. The FMT group had two FMT treatments, and the control group received a behavioural intervention. After the first FMT treatment, the Childhood Autism Rating Scale (CARS) scores in the FMT group decreased by 10.8%, compared with 0.8% in the control group. After the second FMT, further improvement was observed. However, FMT was associated with some transient side effects, such as fever.

In a further study, the vancomycin antibiotic was first administered, followed by a bowel preparation and then FMT [78]. Eighteen ASD children aged 7–16 years old were included and, at baseline, were compared with twenty healthy age-matched controls. FMT not only improved their behavioural symptoms but also increased bacterial diversity with a decrease in gastrointestinal symptoms. There were increases in the abundance of Bifidobacterium, Prevotella and Desulfovibrio. The behavioural improvements were sustained for up to two years. Although the sample size is relatively small, the findings show enormous promise for FMT.

Table 4. Microbiota changes in autism.

| Reference |
|------------------|------------------|
| Clostridium and Ruminococcus spp. ↑ | [79] |
| Bacteroidetes, Proteobacteria, Alkalifexus, Desulfovibrio, Acetanaerobacterium, Bacteroides, Parabacteroides, Desulfovibrio spp. ↑ | [80] |
| Actinobacteria, Turicibacter Clostridium, Firmicutes, Weissella, Helcococcus, Alkaliphilus, Anaerofilum, Pseudoramibacter, Ruminococcus, Streptococcus, Anaerovorax, Dialister, Lactococcus ↓ | |
| Fimbicutes↓, Proteobacteria↓, Verrucomicrobia ↓ | |
| Bacteroidetes/Fimbicutes↑, Dialister↓ | |
| Prevotella↑ Bacteroides, Meganonas, Escherichia/Shigella, Lachnospiraceae_incertae_sedis↑ | [81] |
| Clostridium XIVa, Eisenbergiella, Clostridium IV, Flavonifractor, Haemophilus, Akkermansia↓ | |
| Lachnospiraceae, Clostridiales, Dorea, Erysipelotrichaceae, Collinsella, Lachnolavidum↑ | [82] |
| Bacteroides, Faecalibacterium, Paraprevotella, Paraprevotella↓ | |
| Firmicuteses, Meganonas, Proteobacteria, Actinobacteria, Dialister, Escherichia-Shigella, Bifidobacterium↑ Bacteroidetes↓ | [83] |
| Clostridium, Dialister, Coprobacillus↑ Faecalibacterium↓ | [84] |

8. Discussion

The field of the microbiota-gut-brain axis and neurological disease is novel and in most respects, an area in the early stages of development. See Figure 2. For this reason and
because of the complexity of conducting well-controlled clinical studies, the literature is skewed in favour of preclinical studies. Consequently, there are now more published speculative reviews than papers focusing directly on carefully phenotyped patient populations. Here, we have pinpointed the clinical studies required to bring the field forward.

**Alzheimer’s Disease**
- amyloid plaques
- neurofibrillary tangles

**Parkinson’s disease**
- loss of DA neurons in substantia nigra

**Multiple sclerosis**
- neuronal demyelination

**Autism**
- poorly defined pathology

**Consequences of dysbiosis**
- ↑ inflammatory cytokines
- Altered short-chain fatty acids
- Pathogen spread via vagus nerve
- Altered gut barrier
- Altered BBB

**Figure 2.** Bidirectional communication between the gut and the brain and how a gut dysbiosis can result in several central pathologies causing neurological dysfunction. Abbreviations: DA, dopamine; BBB, blood–brain barrier.

A greater understanding of the role of gut microbes in disease pathogenesis raises the possibility of nutritional interventions. Mohan et al., for instance, explore the potential role of dietary gluten in neurodegeneration [85]. Cortical excitability to transcranial magnetic stimulation, after a lengthy gluten-free diet, modulated the electrocortical changes in celiac disease [86]. However, the use of gluten-free diets in managing cognitive impairment remains very controversial.

There are, however, a multitude of other nutritional interventions that could be undertaken for the aforementioned diseases. Conducting such studies on a large scale will be expensive and necessitate multiple research centres. For meaningful findings, the research needs to be powered in a similar manner to that for pharmaceutical interventions, and the studies should be double-blind, random-allocation, with a parallel-group design. Adequate powering is essential, but such large-scale non-pharmaceutical industry funding will be challenging to acquire.

**9. Conclusions**

Enormous progress has been made, over the past decade or two, in examining the impact of the gut microbiota on brain function. As with any emerging field, there are far more data available from preclinical than clinical studies, and the latter are largely cross-sectional in nature. What is now required are prospective, longitudinal studies exploring the microbiota in well-phenotyped patients. Whether the microbiota changes are a cause or a consequence of disease has yet to be definitively shown. Furthermore, we require large-scale intervention studies with a parallel-group, double-blind, placebo-controlled design to examine the impact of microbiota-targeted interventions, including FMT.

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References

1. Dinan, T.G.; Butler, M.J.; Cryan, J.F. Psychobiotics: Evolution of Novel Antidepressants. Mod. Trends Psychiatry 2021, 32, 134–143. [CrossRef] [PubMed]
2. Dinan, T.G.; Cryan, J.F. Gut microbiota: A missing link in psychiatry. World Psychiatry 2020, 19, 111–112. [CrossRef] [PubMed]
3. Shi, H.; Grodner, B.; De Vlaminck, I. Recent advances in tools to map the microbiome. Curr. Opin. Biomed. Eng. 2021, 19, 100289. [CrossRef] [PubMed]
4. Schwartz, D.J.; Langdon, A.E.; Dantas, G. Understanding the impact of antibiotic perturbation on the human microbiome. Genome Med. 2020, 12, 82. [CrossRef]
5. Wang, W.J.; Zhou, Y.L.; He, J.; Feng, Z.Q.; Zhang, L.; Lai, X.B.; Zhou, J.X.; Wang, H. Characterising the composition of intestinal microflora by. World J. Gastroenterol. 2020, 26, 614–626. [CrossRef]
6. Shanahan, F.; Ghosh, T.S.; O’Toole, P.W. The Healthy Microbiome—What Is the Definition of a Healthy Gut Microbiome? Gastroenterology 2021, 160, 483–494. [CrossRef]
7. Chia, J.H.; Feng, Y.; Su, L.H.; Wu, T.L.; Chen, C.L.; Liang, Y.H.; Chiu, C.H. Clostridium innocuum is a significant vancomycin-resistant pathogen for extraintestinal clostridial infection. Clin. Microbiol. Infect. 2017, 23, 560–566. [CrossRef]
8. Wu, N.; Zhou, J.; Mo, H.; Mu, Q.; Su, H.; Li, M.; Yu, Y.; Liu, A.; Zhang, Q.; Xu, J.; et al. The Gut Microbial Signature of Gestational Diabetes Mellitus and the Association With Diet Intervention. Front. Cell. Infect. Microbiol. 2021, 11, 800865. [CrossRef]
9. Weiss, G.A.; Hennet, T. Mechanisms and consequences of intestinal dysbiosis. Cell. Mol. Life Sci. 2017, 74, 2959–2977. [CrossRef]
10. Rejeski, J.J.; Wilson, F.M.; Nagpal, R.; Yadav, H.; Weinberg, R.B. The Impact of a Mediterranean Diet on the Gut Microbiome in Healthy Human Subjects: A Pilot Study. Digestion 2022, 103, 133–140. [CrossRef]
11. Liu, J.; Lahousse, L.; Nivard, M.G.; Bot, M.; Chen, L.; van Klinken, J.B.; Theising, C.S.; Beekman, M.; van den Akker, E.B.; Slieter, R.C.; et al. Integration of epidemiologic, pharmacologic, genetic and gut microbiome data in a drug-metabolite atlas. Nat. Med. 2020, 26, 110–117. [CrossRef] [PubMed]
12. Claesson, M.J.; Jeffery, I.B.; Conde, S.; Power, S.E.; O’Connor, E.M.; Cusack, S.; Harris, H.M.; Coakley, M.; Lakshminarayan, B.; O’Sullivan, O.; et al. Gut microbiota composition correlates with diet and health in the elderly. Nature 2012, 488, 178–184. [CrossRef] [PubMed]
13. Chen, M.; Ruan, G.; Chen, L.; Ying, S.; Li, G.; Xu, F.; Xiao, Z.; Tian, Y.; Lv, L.; Ping, Y.; et al. Neurotransmitter and Intestinal Interactions: Focus on the Microbiota-Gut-Brain Axis in Irritable Bowel Syndrome. Front. Endocrinol. 2022, 13, 817100. [CrossRef]
14. Patterson, E.; Ryan, P.M.; Wiley, N.; Carafa, I.; Sherwin, E.; Moloney, G.; Franciosi, E.; Mandal, R.; Wishart, D.S.; Tuohy, K.; et al. Gamma-aminobutyric acid-producing lactobacilli positively affect metabolism and depressive-like behaviour in a mouse model of metabolic syndrome. Sci. Rep. 2019, 9, 16323. [CrossRef] [PubMed]
15. Desbonnet, L.; Barrett, L.; Clarke, G.; Bienenstock, J.; Dinan, T.G. The probiotic Bifidobacterium infantis: An assessment of potential antidepressant properties in the rat. J. Psychiatr. Res. 2008, 43, 164–174. [CrossRef] [PubMed]
16. Dinan, T.G.; Cryan, J.F. Brain-Gut-Microbiota Axis and Mental Health. Psychosom. Med. 2017, 79, 920–926. [CrossRef] [PubMed]
17. Bravo, J.A.; Forsythe, P.; Chew, M.V.; Escaravage, E.; Savigia, H.M.; Dinan, T.G.; Bienenstock, J.; Cryan, J.F. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. Proc. Natl. Acad. Sci. USA 2011, 108, 16050–16055. [CrossRef] [PubMed]
18. Maes, H.; Kitahara, M.; Takamatsu, Y.; Mani, H.; Inoue, T. Effects of exercise and pharmacological inhibition of histone deacetylases (HDACs) on epigenetic regulations and gene expressions crucial for neuronal plasticity in the motor cortex. Brain Res. 2021, 1751, 147191. [CrossRef]
19. Kimura, I.; Ichimura, A.; Ohue-Kitano, R.; Igarashi, M. Free Fatty Acid Receptors in Health and Disease. Physiol. Rev. 2020, 100, 171–210. [CrossRef]
20. Cryan, J.F.; Dinan, T.G. More than a gut feeling: The microbiota regulates neurodevelopment and behavior. Neuropsychopharmacology 2015, 40, 241–242.
21. Castillo-Alvarez, F.; Marzo-Sola, M.E. Role of the gut microbiota in the development of various neurological diseases. Neurology 2022, 37, 492–498. [CrossRef] [PubMed]
22. Cryan, J.F.; O’Riordan, K.J.; Sandhu, K.; Peterson, V.; Dinan, T.G. The gut microbiome in neurological disorders. Lancet Neurol. 2020, 19, 179–194. [CrossRef]
23. Korf, J.M.; Ganesh, B.P.; McCullough, L.D. Gut dysbiosis and age-related neurological diseases in females. Neurobiol. Dis. 2022, 168, 105695. [CrossRef] [PubMed]
24. Guert, C.; Kong, G.; Renoir, T.; Hannan, A.J. Exercise, diet and stress as modulators of gut microbiota: Implications for neurodegenerative diseases. Neurobiol. Dis. 2020, 134, 104621. [CrossRef]
25. Yap, C.X.; Henders, A.K.; Alvares, G.A.; Wood, D.L.A.; Krause, L.; Tyson, G.W.; Restuadi, R.; Wallace, L.; McLaren, T.; Hansell, N.K.; et al. Autism-related dietary preferences mediate autism-gut microbiome associations. *Cell* **2021**, *184*, 5916–5931.e17. [CrossRef]

26. Just, M.K.; Gram, H.; Theologidis, V.; Jensen, P.H.; Nilsson, K.P.R.; Lindgren, M.; Knudsen, K.; Borghammer, P.; Van Den Berge, N. Alpha-Synuclein Strain Variability in Body-First and Brain-First Synucleinopathies. *Front. Aging Neurosci.* **2022**, *14*, 907293. [CrossRef]

27. Liu, B.; Fang, F.; Pedersen, N.L.; Tillander, A.; Ludvigsson, J.F.; Ekborn, A.; Svenningson, P.; Chen, H.; Würdefeldt, K. Vagotomy and Parkinson disease: A Swedish register-based matched-cohort study. *Neurology* **2017**, *88*, 1996–2002. [CrossRef]

28. Phillips, R.J.; Walter, G.C.; Wilder, S.L.; Baronovsky, E.A.; Powley, T.L. Alpha-synuclein-immunopositive myenteric neurons and vagal preganglionic terminals: Autonomic pathway implicated in Parkinson’s disease? *Neuroscience* **2008**, *153*, 733–750. [CrossRef] [PubMed]

29. Huang, Y.; Liao, J.; Liu, X.; Zhong, Y.; Cai, X.; Long, L. Review: The Role of Intestinal Dysbiosis in Parkinson’s Disease. *Front. Cell. Infect. Microbiol.* **2021**, *11*, 615075. [CrossRef] [PubMed]

30. Scheperjans, F.; Aho, V.; Pereira, P.A.; Koskinen, K.; Paulin, L.; Pekkonen, E.; Haapaniemi, E.; Kaakkola, S.; Eerola-Rautio, J.; Pohja, M.; et al. Gut microbiota are related to Parkinson’s disease and clinical phenotype. *Mov. Disord.* **2015**, *30*, 350–358. [CrossRef]

31. Sampson, T.R.; Debelius, J.W.; Thron, T.; Janssen, S.; Shastru, G.G.; Ilhan, Z.E.; Challis, C.; Schretter, C.E.; Rocha, S.; Gradinaru, V.; et al. Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson’s Disease. *Cell* **2016**, *167*, 1469–1480.e1412. [CrossRef] [PubMed]

32. Nishiwaki, H.; Ito, M.; Hamaguchi, T.; Maeda, T.; Kashihara, K.; Tsuboi, Y.; Ueyama, J.; Yoshiya, T.; Hanada, H.; Takeuchi, I.; et al. Short chain fatty acids-producing and mucin-degrading intestinal bacteria predict the progression of early Parkinson’s disease. *NPJ Parkinson’s Dis.* **2022**, *8*, 65. [CrossRef] [PubMed]

33. Xue, L.J.; Yang, X.Z.; Tong, Q.; Shen, P.; Ma, S.J.; Wu, S.N.; Zheng, J.L.; Wang, H.G. Fecal microbiota transplantation therapy for Parkinson’s disease: A preliminary study. *Medicine* **2020**, *89*, e22035. [CrossRef] [PubMed]

34. Tamtaji, O.R.; Taghizadeh, M.; Daneshvar Kakhani, R.; Kouchaki, E.; Bahrani, F.; Borzabadi, S.; Oryan, S.; Mafi, A.; Asemi, Z. Clinical and metabolic response to probiotic administration in people with Parkinson’s disease: A randomised, double-blind, placebo-controlled trial. *Clin. Nutr.* **2019**, *38*, 1031–1035. [CrossRef]

35. Petrov, V.A.; Saltykova, I.V.; Zhukova, I.A.; Alifirova, V.M.; Zhukova, N.G.; Dorofeeva, Y.B.; Tyakht, A.V.; Kovarsky, B.A.; Alekseev, D.G.; Kostryukova, E.S.; et al. Analysis of Gut Microbiota in Patients with Parkinson’s Disease. *Bull. Exp. Biol. Med.* **2017**, *162*, 734–737. [CrossRef]

36. Vidal-Martinez, G.; Chin, B.; Camarillo, C.; Herrera, G.V.; Yang, B.; Sarosiek, I.; Perez, R.G. A Pilot Microbiota Study in Parkinson’s Disease Patients versus Control Subjects, and Effects of FTY720 and FTY720-Mitoxo Therapies in Parkinsonian and Multiple System Atrophy Mouse Models. *J. Parkinson’s Dis.* **2020**, *10*, 185–192. [CrossRef]

37. Hasegawa, S.; Goto, S.; Tsuji, H.; Okuno, T.; Asahara, T.; Nomoto, K.; Shibata, A.; Fujisawa, Y.; Minato, T.; Okamoto, A.; et al. Intestinal Dysbiosis and Lowered Serum Lipopolysaccharide-Binding Protein in Parkinson’s Disease. *PLoS ONE* **2015**, *10*, e0142164. [CrossRef]

38. Aho, V.T.E.; Pereira, P.A.B.; Voutilainen, S.; Paulin, L.; Pekkonen, E.; Auvinen, P.; Scheperjans, F. Gut microbiota in Parkinson’s disease: Temporal stability and relations to disease progression. *Ebiomedicine* **2019**, *44*, 691–707. [CrossRef]

39. Manfreday, R.A.; Forsyth, C.B.; Voigt, R.M.; Hall, D.A.; Goetz, C.G.; Keshavarzian, A. Gut-Brain Communication in Parkinson’s Disease: Enteroendocrine Regulation by GLP-1. *Curr. Neurol. Neurosci. Rep.* **2022**, *22*, 335–342. [CrossRef]

40. Hopfner, F.; Künstner, A.; Mülher, S.H.; Künzel, S.; Zeuner, K.E.; Margraf, N.G.; Deuschl, G.; Baines, J.E.; Kuhlpenbäumer, G. Gut microbiota in Parkinson disease in a northern German cohort. *Brain Res.* **2017**, *1667*, 41–45. [CrossRef]

41. Li, W.; Wu, X.; Hu, X.; Wang, T.; Liang, S.; Duan, Y.; Jin, F.; Qin, B. Structural changes of gut microbiota in Parkinson’s disease and its correlation with clinical features. *Sci. China Life Sci.* **2017**, *60*, 1223–1233. [CrossRef] [PubMed]

42. Baldini, F.; Hertil, J.; Sandt, E.; Thines, C.C.; Neuberger-Castillo, L.; Pavelka, L.; Betsoiu, F.; Krüger, R.; Thiele, L.; Consortium, N.-P. Parkinson’s disease-associated alterations of the gut microbiome predict disease-relevant changes in metabolic functions. *BMC Biol.* **2020**, *18*, 62. [CrossRef]

43. Lin, A.; Zheng, W.; He, Y.; Tang, W.; Wei, X.; He, R.; Huang, W.; Su, Y.; Huang, Y.; Zhou, H.; et al. Gut microbiota in patients with Parkinson’s disease in southern China. *Parkinsonism Relat. Disord.* **2018**, *53*, 82–88. [CrossRef] [PubMed]

44. Schneider, J.A. Neuropathology of Dementia Disorders. *Continuum* **2022**, *28*, 834–851. [CrossRef]

45. Harach, T.; Marungruang, N.; Duthilleul, N.; Cheatham, V.; Mc Coy, K.D.; Frisoni, G.; Neher, J.J.; Fäk, F.; Jucker, M.; Lasser, T.; et al. Reduction of Abeta amyloid pathology in APPPS1 transgenic mice in the absence of gut microbiota. *Sci. Rep.* **2017**, 7, 41802. [CrossRef]

46. Brandscheid, C.; Schuck, F.; Reinhardt, S.; Schäfer, K.H.; Pietrzik, C.U.; Grimm, M.; Hartmann, T.; Schwierz, A.; Endres, K. Altered Gut Microbiome Composition and Tryptic Activity of the 5xFAD Alzheimer’s Mouse Model. *J. Alzheimer’s Dis.* **2017**, *56*, 775–788. [CrossRef]

47. Shen, T.; Yue, Y.; He, T.; Huang, C.; Qu, B.; Lv, W.; Lai, H.Y. The Association Between the Gut Microbiota and Parkinson’s Disease, a Meta-Analysis. *Front. Aging Neurosci.* **2021**, *13*, 636545. [CrossRef]
48. Distrutti, E.; O’Reilly, J.A.; McDonald, C.; Cipriani, S.; Renga, B.; Lynch, M.A.; Fierucci, S. Modulation of intestinal microbiota by the probiotic VSL#3 resets brain gene expression and ameliorates the age-related deficit in LTP. PLoS ONE 2014, 9, e106503. [CrossRef]

49. Cattaneo, A.; Cattane, N.; Galluzzi, S.; Provasi, S.; Lopizzo, N.; Festari, C.; Ferrari, C.; Guerra, U.P.; Paghera, B.; Muscio, C.; et al. Association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. Neurobiol. Aging 2017, 69, 60–68. [CrossRef]

50. Wang, Y.; Li, L.; Zhao, X.; Sui, S.; Wang, Q.; Shi, G.; Xu, H.; Zhang, X.; He, Y.; Gu, J. Intestinal Microflora Changes in Patients with Mild Alzheimer’s Disease in a Chinese Cohort. J. Alzheimer’s Dis. 2022. pre-press. [CrossRef]

51. Asaoka, D.; Xiao, J.; Takeda, T.; Yanagisawa, N.; Yamazaki, T.; Matsubara, Y.; Sugiyama, H.; Endo, N.; Higa, M.; Kaskanuki, K.; et al. Effect of Probiotic Bifidobacterium breve in Improving Cognitive Function and Preventing Brain Atrophy in Older Patients with Suspected Mild Cognitive Impairment: Results of a 24-Week Randomised, Double-Blind, Placebo-Controlled Trial. J. Alzheimer’s Dis. 2022, 88, 75–95. [CrossRef] [PubMed]

52. Zhuang, Z.Q.; Shen, L.L.; Li, W.W.; Fu, X.; Zeng, F.; Gui, L.; Lü, Y.; Cai, M.; Zhu, C.; Tan, Y.L.; et al. Gut Microbiota is Altered in Patients with Alzheimer’s Disease. J. Alzheimer’s Dis. 2018, 63, 1337–1346. [CrossRef] [PubMed]

53. Li, B.; He, Y.; Ma, J.; Huang, P.; Du, J.; Cao, L.; Wang, Y.; Xiao, Q.; Tang, H.; Chen, S. Mild cognitive impairment has similar alterations as Alzheimer’s disease in gut microbiota. Alzheimer’s Dement. 2019, 15, 1357–1366. [CrossRef] [PubMed]

54. Liu, P.; Wu, L.; Peng, G.; Han, Y.; Tang, R.; Ge, J.; Zhang, L.; Jia, L.; Yue, S.; Zhou, K.; et al. Altered microbiomes distinguish Alzheimer’s disease from mild cognitive impairment and health in a Chinese cohort. Brain Behav. Immun. 2019, 80, 633–643. [CrossRef]

55. Vogt, N.M.; Kerby, R.L.; Dill-McFarland, K.A.; Harding, S.J.; Merluzzi, A.P.; Johnson, S.C.; Carlsson, C.M.; Asthana, S.; Zetterberg, H.; Blennow, K.; et al. Gut microbiome alterations in Alzheimer’s disease. Sci. Rep. 2017, 7, 13537. [CrossRef]

56. Talanki Manjunatha, R.; Habib, S.; Sangaraju, S.L.; Yepez, D.; Grandes, X.A. Multiple Sclerosis: Therapeutic Strategies on the Horizon. Curr. Top. 2022, 14, e24895. [CrossRef]

57. Hoban, A.E.; Stilling, R.M.; Ryan, F.J.; Shanahan, F.; Dinan, T.G.; Claesson, M.J.; Clarke, G.; Cryan, J.F. Regulation of prefrontal cortex myelination by the microbiota. Transl. Psychiatry 2016, 6, e774. [CrossRef]

58. Lee, Y.K.; Menezes, J.S.; Umesaki, Y.; Mazmanian, S.K. Proinflammatory T-cell responses to gut microbiota promote experimental autoimmune encephalomyelitis. Proc. Natl. Acad. Sci. USA 2011, 108 (Suppl. S1), 4615–4622. [CrossRef]

59. Navarro-López, V.; Méndez-Miralles, M.; Vela-Yebra, R.; Fries-Ramos, A.; Sánchez-Pellicer, P.; Ruzafa-Costas, B.; Núñez-Delegido, E.; Gómez-Gómez, H.; Chumillas-Lidón, S.; Picó-Monllor, J.A.; et al. Gut Microbiota as a Potential Predictive Biomarker in Relapsing-Remitting Multiple Sclerosis. Genes 2022, 13, 930. [CrossRef]

60. Al, K.F.; Craven, L.J.; Gibbons, S.; Parvathy, S.N.; Wing, A.C.; Graf, C.; Parham, K.A.; Kerfoot, S.M.; Wilcox, H.; Burton, J.P.; et al. Fecal microbiota transplantation is safe and tolerable in patients with multiple sclerosis: A pilot randomised controlled trial. Mult. Scler. J. Exp. Transl. Clin. 2022, 8. [CrossRef]

61. Mirashrafi, S.; Hejazi Taghanaki, S.Z.; Sarlak, F.; Moravejolahkami, A.R.; Hojjati Kermani, M.A.; Haratian, M. Effect of probiotics supplementation on disease progression, depression, general health, and anthropometric measurements in relapsing-remitting multiple sclerosis patients: A systematic review and meta-analysis of clinical trials. Int. J. Clin. Pract. 2021, 75, e14724. [CrossRef] [PubMed]

62. Boussamet, L.; Rajoka, M.S.R.; Berthelot, L. Microbiota, IgA and Multiple Sclerosis. Microorganisms 2022, 10, 617. [CrossRef] [PubMed]

63. Jang, S.; Gandhi, R.; Cox, L.M.; Li, N.; von Glehn, F.; Yan, R.; Patel, B.; Mazzola, M.A.; Liu, S.; Glanz, B.L.; et al. Alterations of the human gut microbiome in multiple sclerosis. Nat. Commun. 2016, 7, 12015. [CrossRef] [PubMed]

64. Oezguen, N.; Yalcinkaya, N.; Kücükali, C.I.; Dahdouli, M.; Hollister, E.B.; Luna, R.A.; Türkoglu, R.; Kürtüncü, M.; Eraksoy, M.; et al. Association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. Neurobiol. Aging 2017, 69, 60–68. [CrossRef]

65. Grassi, D.; Marraudino, M.; García-Segura, L.M.; Panzica, G.C. The hypothalamic paraventricular nucleus as a central hub for the association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. Neurobiol. Aging 2017, 69, 60–68. [CrossRef]

66. Castillo-Alvarez, F.; Pérez-Matute, P.; Oteo, J.A.; Marzo-Sola, M.E. The influence of interferon β-1b on gut microbiota composition in patients with multiple sclerosis. Neurologia 2021, 36, 495–503. [CrossRef]

67. Tang, J.W.; Li, J.W.; Baulderstone, D.; Jeyaseelan, D. Presenting age and features of females diagnosed with autism spectrum disorder. J. Paediatr. Child Health 2021, 57, 1182–1189. [CrossRef]

68. Tang, J.W.; Li, J.W.; Baulderstone, D.; Jeyaseelan, D. Presenting age and features of females diagnosed with autism spectrum disorder. J. Paediatr. Child Health 2021, 57, 1182–1189. [CrossRef]

69. Lu, C.; Rong, J.; Fu, C.; Wang, W.; Xu, J.; Xu, X.D. Overall Rebalancing of Gut Microbiota Is Key to Autism Intervention. Front. Psychol. 2022, 13, 862719. [CrossRef]

70. Desbonnet, L.; Clarke, G.; Shanahan, F.; Dinan, T.G.; Cryan, J.F. Microbiota is essential for social development in the mouse. Mol. Psychiatry 2014, 19, 146–148. [CrossRef] [PubMed]

71. Desbonnet, L.; Clarke, G.; Traplin, A.; O’Sullivan, O.; Crispie, F.; Moloney, R.D.; Cotter, P.D.; Dinan, T.G.; Cryan, J.F. Gut microbiota depletion from early adolescence in mice: Implications for brain and behaviour. Brain Behav. Immun. 2015, 48, 165–173. [CrossRef] [PubMed]
72. Yu, Y.; Huang, J.; Chen, X.; Fu, J.; Wang, X.; Pu, L.; Gu, C.; Cai, C. Efficacy and Safety of Diet Therapies in Children With Autism Spectrum Disorder: A Systematic Literature Review and Meta-Analysis. *Front. Neurol.* 2022, *13*, 844117. [CrossRef] [PubMed]

73. Dinan, T.G.; Stanton, C.; Cryan, J.F. Psychobiotics: A novel class of psychotrophic. *Biol. Psychiatry* 2013, *74*, 720–726. [CrossRef] [PubMed]

74. Liu, Y.W.; Liong, M.T.; Chung, Y.E.; Huang, H.Y.; Peng, W.S.; Cheng, Y.F.; Lin, Y.S.; Wu, Y.Y.; Tsai, Y.C. Effects of. *Nutrients* 2019, *11*, 820. [CrossRef] [PubMed]

75. Fang, X.; Wang, Y.; Huang, Z. Oxytocin Neurons Are Essential in the Social Transmission of Maternal Behavior. *Front. Behav. Neurosci.* 2022, *16*, 847396. [CrossRef] [PubMed]

76. Davies, C.; Mishra, D.; Eshraghi, R.S.; Mittal, J.; Sinha, R.; Bulut, E.; Mittal, R.; Eshraghi, A.A. Altering the gut microbiome to potentially modulate behavioral manifestations in autism spectrum disorders: A systematic review. *Neurosci. Biobehav. Rev.* 2021, *128*, 549–557. [CrossRef]

77. Liu, J.; Gao, Z.; Liu, C.; Liu, T.; Gao, J.; Cai, Y.; Fan, X. Alteration of Gut Microbiota: New Strategy for Treating Autism Spectrum Disorder. *Front. Cell. Dev. Biol.* 2022, *10*, 792490. [CrossRef] [PubMed]

78. Kang, D.W.; Adams, J.B.; Vargason, T.; Santiago, M.; Hahn, J.; Krajmalnik-Brown, R. Distinct Fecal and Plasma Metabolites in Children with Autism Spectrum Disorders and Their Modulation after Microbiota Transfer Therapy. *mSphere* 2020, *5*, e00314-20. [CrossRef]

79. Finegold, S.M.; Molitoris, D.; Song, Y.; Liu, C.; Vaisanen, M.L.; Bolte, E.; McTeague, M.; Sandler, R.; Wexler, H.; Marlowe, E.M.; et al. Gastrointestinal microflora studies in late-onset autism. *Clin. Infect. Dis.* 2002, *35* (Suppl. S1), S6–S16. [CrossRef]

80. Finegold, S.M.; Dowd, S.E.; Gontcharova, V.; Liu, C.; Henley, K.E.; Wolcott, R.D.; Youn, E.; Summanen, P.H.; Granpeesheh, D.; Dixon, D.; et al. Pyrosequencing study of fecal microflora of autistic and control children. *Anaerobe* 2010, *16*, 444–453. [CrossRef]

81. Zou, R.; Xu, F.; Wang, Y.; Duan, M.; Guo, M.; Zhang, Q.; Zhao, H.; Zheng, H. Changes in the Gut Microbiota of Children with Autism Spectrum Disorder. *Autism. Res.* 2020, *13*, 1614–1625. [CrossRef] [PubMed]

82. Ding, X.; Xu, Y.; Zhang, X.; Zhang, L.; Duan, G.; Song, C.; Li, Z.; Yang, Y.; Wang, Y.; Wang, X.; et al. Gut microbiota changes in patients with autism spectrum disorders. *J. Psychiatr. Res.* 2020, *129*, 149–159. [CrossRef] [PubMed]

83. Williams, B.L.; Hornig, M.; Buie, T.; Bauman, M.L.; Cho Paik, M.; Wick, I.; Bennett, A.; Jabado, O.; Hirschberg, D.L.; Lipkin, W.I. Impaired carbohydrate digestion and transport and mucosal dysbiosis in the intestines of children with autism and gastrointestinal disturbances. *PLoS ONE* 2011, *6*, e24585. [CrossRef]

84. Wan, Y.; Zuo, T.; Xu, Z.; Zhang, F.; Zhan, H.; Chan, D.; Leung, T.F.; Yeeoh, Y.K.; Chan, F.K.L.; Chan, R.; et al. Underdevelopment of the gut microbiota and bacteria species as non-invasive markers of prediction in children with autism spectrum disorder. *Gut* 2022, *71*, 910–918. [CrossRef]

85. Mohan, M.; Okeoma, C.M.; Sestak, K. Dietary Gluten and Neurodegeneration: A Case for Preclinical Studies. *Int. J. Mol. Sci.* 2020, *21*, 5407. [CrossRef]

86. Pennisi, M.; Bramanti, A.; Cantone, M.; Pennisi, G.; Bella, R.; Lanza, G. Neurophysiology of the “Celiac Brain”: Disentangling Gut-Brain Connections. *Front. Neurosci.* 2017, *11*, 498. [CrossRef] [PubMed]