Case Report

Successful Handling of Disseminated BCG Disease in a Child with Severe Combined Immunodeficiency

Silvia Bacalhau,1 Cristina Freitas,1 Rosalina Valente,1 Deolinda Barata,1 Conceição Neves,2 Katrin Schäfer,3 Annelie Lubatschofski,3 Ansgar Schulz,3 and João Farela Neves2

1 Pediatric Intensive Care Unit, Hospital Dona Estefânia, 1169-045 Lisboa, Portugal
2 Infectology Unit, Hospital Dona Estefânia, Rua Jacinta Marto, 1169-045 Lisboa, Portugal
3 Department of Immunology, Rheumatology and Stem Cell Transplantation, University Children’s Hospital, 89073-89081 Ulm, Germany

Correspondence should be addressed to João Farela Neves, joao.farelaneves@chlc.min-saude.pt

Received 5 May 2011; Revised 7 August 2011; Accepted 19 August 2011

Academic Editor: T. Ottenhoff

Copyright © 2011 Silvia Bacalhau et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

In high-burden countries, Mycobacterium bovis Bacillus Calmette-Guérin (BCG) vaccine is administered in newborn to prevent severe Mycobacterium tuberculosis infection. Because life-threatening disseminated BCG disease may occur in children with primary immunodeficiency, vaccination strategy against tuberculosis should be redefined in non-high-burden countries. We report the case of a patient with X-linked severe combined immunodeficiency (SCID) who developed disseminated BCG disease, highlighting the specific strategies adopted.

1. Introduction

Bacillus Calmette-Guérin (BCG) vaccine is administered worldwide to prevent severe Mycobacterium tuberculosis infection. BCG vaccine has an excellent safety profile [1], but a wide range of complications can occur (from simple local lymphadenitis to almost universally lethal disseminated disease) [2]. It is now accepted that most patients with moderate to severe complications have underlying immunodeficiency [3]. Several genetic defects that correlate with BCG disease have now been identified, such as Mendelian susceptibility to Mycobacterial disease (IFN-γ-IL-12 pathway), hyper-IgM syndrome, chronic granulomatous disease, and SCID [4].

BCG is part of the Portuguese Immunization Program since 1965, being administered at birth to children with weight above 2000 grams.

We report a patient with SCID with disseminated BCG disease and highlight specific strategies adopted in patient’s treatment.

2. Case Presentation

A 5-months-old boy was admitted to the intensive care unit of a tertiary pediatric hospital in Lisbon for respiratory insufficiency following three days of fever, cough, and worsening respiratory distress. He presented with oral thrush, bilateral pulmonary rales, and hepatosplenomegaly. He had been vaccinated at birth with BCG (Danish strain 1331) and had had recurrent infections (conjunctivitis, otitis media, and persistent oral thrush).

There was no consanguinity. Two maternal uncles died of pneumonia in infancy.

The chest X-ray revealed absence of a thymus and showed bilateral areas of opacities (Figure 1). The immunologic panel confirmed the hypothesis of SCID (Table 1), later genetically characterized (missense mutation in exon 5, codon 226 CGC → CAC, leading to an arginine 226 substitution by histidine; common cytokine receptor gamma chain of IL-2—IL2RG gene). This mutation has previously been described as causing human X-linked SCID [5].
Although being asymptomatic, chimerism studies were performed to assess the presence of exogenous maternal lymphocytes, which can be important for an early diagnosis and for the choice of the conditioning regimen if bone marrow transplantation is considered. The results confirmed that 75% of total lymphocyte count was maternal.

The bronchoalveolar lavage was positive for *Candida albicans*, *Pneumocystis jiroveci*, and acid-fast bacilli. An ultrasound evaluation showed an enlarged spleen with nodules. The BCG primer-specific polymerase chain reaction quickly identified the infection by BCG (amplification of *pncA* gene and a multiplex PCR that targets the RD1 region for BCG [6–8]), and presumptive diagnosis of disseminated BCG was made.

He was treated with trimethoprim-sulfamethoxazole and a four-tuberculostatic drug regimen, comprised of isoniazid, rifampin, ethambutol, and levofloxacin, according to laboratory data regarding BCG strain susceptibility. The infant started antibacterial and antifungal prophylaxis and immunoglobulin replacement therapy.

Splenectomy was performed aiming for a reduction in BCG load which could worsen the prognosis following immune reconstitution. Spleen analysis revealed severe BCG infiltration (Figure 2), thus establishing the definitive diagnosis of disseminated BCG disease. Four and a half months after the start of tuberculostatic therapy, T-cell depleted bone marrow transplantation (BMT) from an unrelated 10/10 HLA matched donor following reduced intensity conditioning with treosulfan, fludarabine and campath was performed (graft contained $12.5 \times 10^6$/kilograms CD34+ stem cells). To enhance immune reconstitution, donor lymphocytes were administered at d + 31, d + 61, and d + 88 containing 5, 15, and $50 \times 10^4$ CD3+ T cells per kilograms, respectively. The transplant course was uneventful with the exception of repeated periods of fever that, in the absence of other reported infections, were probably related to active BCG infection, treated continuously by isoniazid, rifampin, and ethambutol. Haematological reconstitution occurred within normal time frame, and there were no signs of graft-versus-host disease. About three months after transplantation, relevant numbers of T cells become detectable. During this phase of immunological reconstitution, signs of BCG infection augmented with prominent nodules at the BCG inoculation site (Figure 3), enlarged thoracic and abdominal lymph nodes, fever, leukocytosis, and massive elevation of C-reactive protein. Linezolid was added to other antituberculous drugs, and some short courses of steroids were administered. The child remained in good clinical condition and is alive and well to date.

3. Discussion

We report a case of X-linked SCID with disseminated BCG disease. SCID is the most severe form of primary immunodeficiency disease, characterized by defective T-lymphocyte differentiation that leads to early death in the absence of hematopoietic stem cell transplantation. Patients usually present severe and life-threatening opportunistic infections early in infancy such as disseminated BCG infection [9].

Prevalence data of primary immunodeficiency disease (PID) are not available in Portugal. In the USA, the prevalence rate of PID is estimated to be 1:2000 children [10] and there are more than 200 cases of disseminated BCG infection in this group of patients [11]. Its estimated incidence is 0.1 to 4.3 per one million vaccinated children and is lethal in 50 to 71% of cases [2, 3, 12]. The availability of the new polymerase chain reaction primers that allow the distinction between *Mycobacterium tuberculosis* and *bovis* is a useful tool in the management of these patients, allowing for quicker diagnosis thus preventing the use of nonappropriate drugs.

Patients with SCID previously vaccinated with BCG are difficult to manage and should be kept under prophylactic three drug regimens (without pyrazinamide, for which BCG is primarily resistant [13]) after exclusion of disseminated disease and until complete immunologic reconstitution occurs after BMT.
Table 1: Laboratory results. *Maternal T cells in chimerism analysis. #Total chimerism 60% donor, 40% patient. Myeloid chimerism 100% patient, T and NK cell chimerism 100% donor.

| Laboratory test                      | Patient prior to transplantation | Patient 3 months after transplantation | Patient 6 months after transplantation | Reference value |
|--------------------------------------|----------------------------------|----------------------------------------|----------------------------------------|-----------------|
| Serum immunoglobulins                |                                  |                                        |                                        |                 |
| IgA (mg/mL)                          | <0.1                             | 0.12                                   | 0.15                                   | 0.2–0.6         |
| IgG (mg/mL)                          | 0.63                             | Substituted                            | Substituted                            | 2.3–4.4         |
| IgM (mg/mL)                          | 0.351                            | 0.79                                   | 0.32                                   | 0.3–0.9         |
| Vaccine titers                        |                                  |                                        |                                        |                 |
| Diphtheria (IU/mL)                   | 0.25                             | Not done                               | Not done                               | >0.5            |
| Tetanus (IU/mL)                      | 0.13                             | Not done                               | Not done                               | >0.5            |
| Isohemagglutinins                    | Undetected                       | Not done                               | Not done                               | >1/16           |
| Lymphocyte subsets                   |                                  |                                        |                                        |                 |
| CD3+ (cells/μL)                      | 345*                             | 124                                    | 1098                                   | 2300–6500       |
| CD3+CD4+ (cells/μL)                  | 8                                | 69                                     | 960                                    | 1500–5000       |
| CD3+CD4+CD45RA (%)                   | <1                               | 62%                                    | 84%                                    | 64–92           |
| CD3+CD8+ (cells/μL)                  | 339*                             | 13                                     | 91                                     | 500–1600        |
| CD3+CD8+CD45RA (%)                   | 0                                |                                        | 53–88                                  |                 |
| CD19+ (cells/μL)                     | 210                              | 1062                                   | 3477                                   | 600–1300        |
| CD56+ (cells/μL)                     | 18                               | 165                                    | 127                                    | 100–1000        |
| Lymphocyte proliferation             |                                  |                                        |                                        |                 |
| Phytohemagglutinin (SI)              | 3                                | 86                                     |                                        |                 |
| Anti-CD3 (SI)                        | 1                                | 24                                     |                                        |                 |
| PPD (SI)                             | 1                                | 9                                      |                                        |                 |
| Vβ TCR repertoire                    | restricted                       | Not done                               | Not done                               |                 |
| Chimerism                            | 75% maternal                     | 100% donor                             | 60% donor*                             |                 |
| Viral loads and PCR                  |                                  |                                        |                                        |                 |
| CMV, EBV, HSV, HIV1/2, HBV, HCV, HHV6/7, VZV, Enterovirus, Adenovirus | All negative | All negative | All negative |                  |
| PCR Norovirus                        | Not done                         | +                                      | +                                      |                 |

Figure 3: BCG vaccination site at the time of immune reconstitution (a) 5 months after transplantation and (b) 5.5 months after transplantation, showing skin ulceration with massive infiltration of BCG.
There are no clear guidelines on the most suitable treatment for disseminated BCG disease [14, 15]. In this case, aggressive therapy (comprising four drugs: isoniazid, rifampicin, ethambutol, and levofloxacin) was administered for four and a half months prior to BMT to reduce the BCG load. For the same reason, splenectomy was performed prior to BMT. Since BCGitis is known to exacerbate clinically during immune reconstitution leading to sepsis and fatal multiple organ failure when T cells rapidly rise after BMT [16], the following modifications of transplant procedure were performed in this case: (a) reduced intensity conditioning using treosulfan and fludarabine instead of the standard regimen (busulfan and cyclophosphamide) to reduce toxicity; (b) T cell depletion of the HLA-identical graft to circumvent BCGitis exacerbation due to rapidly expanding peripheral T cells in the graft; (c) donor lymphocytes in escalating doses to induce a slow but stable T-cell reconstitution; (d) immunosuppression with prednisolone to treat cytokine reactions at the time of overwhelming immune reconstitution. Of course, tuberculostatic drugs have to be administered during the BMT procedure until clinical signs of BCG disappear, which may take several months. In case of empirical treatment, it should be taken into account that BCG vaccine strains differ in their susceptibility pattern to antituberculous drugs and this should be sought.

In countries where the diagnosis of cellular PID (especially SCID) is well below expected, the possibility of a fatal outcome due to the administration of BCG vaccine in the neonatal period (before the suspicion of cellular PID) is a reality. The incidence of tuberculosis in Portugal has been drastically decreasing (from 60 : 100,000 in 1990 to 24 : 100,000 in 2009, of which 5 : 100,000 in children under five years old), and the extrapulmonary forms comprise about 30% of cases (from 60 : 100,000 in 1990 to 24 : 100,000 in 2009, of which 5 : 100,000 in children under five years old), and the extrapulmonary forms comprise about 30% of cases (notably, in 2009 only two cases of "severe" extrapulmonary tuberculosis were reported in children) [17]. This is due to improved public health and social measures, but probably also to the protective effect of BCG vaccination, which is difficult to assess.

Based on this trend and being aware that the diagnosis of cellular PID remains below expected, the authors underline the need to redefine the vaccination strategy against tuberculosis in Portugal, namely, postponing its administration to the second semester of life in regions where the incidence of the disease is still above 20 : 100,000.

Conflict of Interests

We did not receive any funding and there is no commercial relationship or potential conflict of interests related to the submission.

Consent

Parents of patient gave their informed consent for the case report to be published.

Acknowledgments

The authors would like to thank the following for the important technical support: Dr. António Martinho and the Histocompatibility Laboratory of centre Portugal, Professor Ana Espada de Sousa, Professor Adriana Albuquerque, and the Immunology Laboratory of the Molecular Medicine Institute, Hospital Santa Maria, Lisbon, the team of the Department of Immunology, Rheumatology and Stem Cell Transplantation, Ulm.

References

[1] J. B. Milstien and J. J. Gibson, “Quality control of BCG vaccine by WHO: a review of factors that may influence vaccine effectiveness and safety,” Bulletin of the World Health Organization, vol. 68, no. 1, pp. 93–108, 1990.
[2] E. A. Talbot, M. D. Perkins, S. F. M. Suva, and R. Frothingham, “Disseminated bacille Calmette-Guérin disease after vaccination: case report and review,” Clinical Infectious Diseases, vol. 24, no. 6, pp. 1139–1146, 1997.
[3] J. L. Casanova, S. Blanche, J. F. Emile et al., “Idiopathic disseminated bacillus Calmette-Guérin infection: a French national retrospective study,” Pediatrics, vol. 98, no. 4, pp. 774–778, 1996.
[4] W. I. Lee, F. C. Liang, J. L. Huang et al., “Immunologic analysis of HIV-uninfected Taiwanese children with BCG-induced disease,” Journal of Clinical Immunology, vol. 29, no. 3, pp. 319–329, 2009.
[5] J. M. Puck, A. E. Pepper, P. S. Henthorn et al., “Mutation analysis of IL2RG in human x-linked severe combined immunodeficiency,” Blood, vol. 89, no. 6, pp. 1968–1977, 1997.
[6] L. E. Espinosa De Los Monteros, J. C. Galán, M. Gutiérrez et al., “Allele-specific PCR method based on pncA and oxyR sequences for distinguishing Mycobacterium bovis from Mycobacterium tuberculosis: intraspecific M. bovis pncA sequence polymorphism,” Journal of Clinical Microbiology, vol. 36, no. 1, pp. 239–242, 1998.
[7] E. A. Talbot, D. L. Williams, and R. Frothingham, “PCR identification of Mycobacterium bovis BCG,” Journal of Clinical Microbiology, vol. 35, no. 3, pp. 566–569, 1997.
[8] W. J. Su, C. Y. Huang, C. Y. Huang, and R. P. Perng, “Utility of PCR assays for rapid diagnosis of BCG infection in children,” International Journal of Tuberculosis and Lung Disease, vol. 5, no. 4, pp. 380–384, 2001.
[9] A. Fischer, “Severe combined immunodeficiencies (SCID),” Clinical and Experimental Immunology, vol. 122, no. 2, pp. 143–149, 2000.
[10] J. M. Boyle and R. H. Buckley, “Population prevalence of diagnosed primary immunodeficiency diseases in the United States,” Journal of Clinical Immunology, vol. 27, no. 5, pp. 497–502, 2007.
[11] E. A. Bernatowska, B. Wolska-Kusnierz, M. Pac et al., “Disseminated bacillus Calmette-Guérin infection and immunodeficiency [letter],” Emerging Infectious Diseases, vol. 13, no. 5, pp. 799–801, 2007.
[12] M. Sadeghi-Shanbestari, K. Ansarin, S. H. Maljaei et al., “Immunologic aspects of patients with disseminated bacille Calmette-Guérin disease in north-west of Iran,” Italian Journal of Pediatrics, vol. 35, no. 42, pp. 1–5, 2009.
[13] N. Ritz, M. Tébruegge, T. G. Connell, A. Sievers, R. Robins-Browne, and N. Curtis, “Susceptibility of Mycobacterium bovis BCG vaccine strains to antituberculous antibiotics,”
[14] A. Ikincioğullari, F. Doğu, E. Çiftci et al., “An intensive approach to the treatment of disseminated BCG infection in a SCID patient,” Bone Marrow Transplantation, vol. 30, no. 1, pp. 45–47, 2002.

[15] W. Pariyaprasert, P. Pacharn, N. Visitsunthorn et al., “Successful treatment of disseminated BCG infection in a SCID patient with granulocyte colony stimulating factor,” Asian Pacific Journal of Allergy and Immunology, vol. 26, no. 1, pp. 71–75, 2008.

[16] R. Skinner, A. L. Appleton, M. S. Sprott et al., “Disseminated BCG infection in severe combined immunodeficiency presenting with severe anaemia and associated with gross hypersplenism after bone marrow transplantation,” Bone Marrow Transplantation, vol. 17, no. 5, pp. 877–880, 1996.

[17] Relatório para o dia mundial da Tuberculose, “Programa nacional de luta contra a tuberculose (PNT), Março de 2010,” 2010, http://www.min-saude.pt/NR/rdonlyres/8B5396AB-E7B3-4B81-A368-F2B5F7E28F95/0/RelatorioTuberculose-DGS.pdf.