Immunotherapy for targeting cancer stem cells in hepatocellular carcinoma

Xiaomeng Dai#, Yixuan Guo#, Yan Hu, Xuanwen Bao, Xudong Zhu, Qihan Fu, Hangyu Zhang, Zhou Tong, Lulu Liu, Yi Zheng, Peng Zhao, Weijia Fang

Department of Medical Oncology, The First Affiliated Hospital, College of Medicine, Zhejiang University, Hangzhou, China.

#These authors contributed equally to this work.

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See http://ivyspring.com/terms for full terms and conditions.

Received: 2020.10.18; Accepted: 2020.12.21; Published: 2021.01.19

Abstract

The rapid development and remarkable success of checkpoint inhibitors have provided significant breakthroughs in cancer treatment, including hepatocellular carcinoma (HCC). However, only 15-20% of HCC patients can benefit from checkpoint inhibitors. Cancer stem cells (CSCs) are responsible for recurrence, metastasis, and local and systemic therapy resistance in HCC. Accumulating evidence has suggested that HCC CSCs can create an immunosuppressive microenvironment through certain intrinsic and extrinsic mechanisms, resulting in immune evasion. Intrinsic evasion mechanisms mainly include activation of immune-related CSC signaling pathways, low-level expression of antigen presenting molecules, and high-level expression of immunosuppressive molecules. External evasion mechanisms are mainly related to HBV/HCV infection, alcoholic/nonalcoholic steatohepatitis, hypoxia stimulation, abnormal angiogenesis, and crosstalk between CSCs and immune cells. A better understanding of the complex mechanisms of CSCs involved in immune evasion will contribute to therapies for HCC. Here we will outline the detailed mechanisms of immune evasion for CSCs, and provide an overview of the current immunotherapies targeting CSCs in HCC.

Key words: hepatocellular carcinoma; cancer stem cells; immune evasion; targeting; immunotherapy

Introduction

Hepatocellular carcinoma (HCC) is one of the leading causes of cancer-associated deaths worldwide, accounting for approximately 75-85% of primary liver cancers [1, 2]. Hepatitis B virus (HBV), hepatitis C virus (HCV), alcoholic, and nonalcoholic steatohepatitis (NASH) are major risk factors in the development of HCC [3]. The tumor burden is highest in East Asia (more than 50% in China) and Africa because of HBV infection, while HCC incidence and mortality are increasing rapidly in the United States and Europe due to alcohol consumption and NASH [4]. Most patients with HCC are diagnosed at advanced stages with liver disease and cirrhosis, missing the opportunity for surgery. Despite several advances in the treatment of HCC, particularly in targeted therapy and immunotherapy, the 5-year survival rate remains poor [5]. Drug resistance, tumor metastasis and recurrence are the major causes of poor prognosis in HCC patients.

Cancer stem cells (CSCs) have been shown to be responsible for recurrence, metastasis, and local and systemic therapy resistance in HCC [6]. Moreover, an overwhelming number of studies have suggested that CSCs can form an immunosuppressive microenvironment through both intrinsic and extrinsic mechanisms to induce ineffective antitumor immune responses [7]. Application of immunotherapy, especially programmed cell death protein 1 (PD-1) and programmed cell death ligand 1 (PD-L1) monoclonal antibodies, to a variety of solid tumors (including HCC) represents a major breakthrough in cancer treatment [8, 9]. However, most patients who
have received immunotherapy still experience progression and metastasis [10]. Considering that CSCs are a reservoir for the progression and metastasis of HCC, immunotherapy that targets CSCs may be an exciting research field.

In this review, we summarize the role of CSCs in the tumor immunosuppressive environment (Figure 1) and provide an overview of the current immunotherapies targeting CSCs in HCC (Figure 2).

CSCs and immune evasion in HCC

CSCs are a small population of cells that can self-renew and differentiate to initiate and maintain tumor growth [11]. T Lapidot and colleagues first observed the existence of CSCs by demonstrating that CD34+/CD38- myeloid leukemia (AML) cells have the ability to initiate tumors in NOD/SCID mice [12]. HCC stem cells were first identified as side population (SP) cells by Haraguchi and colleagues in 2006 [13, 14]. They found that SP cells in HCC were more resistant to chemotherapy drugs (including 5-fluorouracil, doxorubicin and gemcitabine) than non-SP cells. Chiba et al. confirmed that as few as 1000 HCC SP cells have tumorigenic ability in NOD/SCID mice, whereas up to 1×10⁶ non-SP cells were unable to initiate tumors [15]. Since then, according to xenotransplantation experiments, several cellular biomarkers of CSCs in HCC have been identified, including epithelial cell adhesion molecule (EpCAM), CD133, CD44, CD90, CD13, CD24, OV6, CD47, calcium channel α2δ1 isoform5, and intercellular adhesion molecule 1 (ICAM-1) [6, 16]. Moreover, related studies showed that high expression of these CSC markers was associated with poor prognosis in HCC patients [17-22].

Figure 1. The external and intrinsic mechanisms to mediate immunotherapy resistance for CSCs in HCC. External evasion mechanisms are mainly related to HBV/HCV infection, alcohol/nonalcoholic steatohepatitis, hypoxia stimulation, abnormal angiogenesis, and crosstalk between CSCs and immune cells. Intrinsic evasion mechanisms mainly include activation of the Wnt/β-Catenin signaling pathway and TGF-β signaling pathway, low-level expression of TAP and/or MHC molecules, and high-level expression of CD47 and PD-L1.

http://www.thno.org
The existence of HCC CSCs indicates tumor heterogeneity and hierarchy, which is a hallmark feature of resistance to immunotherapy [23, 24]. Zheng and colleagues observed that CSCs are also heterogeneous, as determined by single-cell transcriptome and functional analysis of HCC cells. They found that different CSC subpopulations have distinct molecular signatures that were independently correlated with poor prognosis in HCC patients [25]. After decades of research, CSCs were found to mediate immunotherapy resistance through various intrinsic and external mechanisms [26]. Intrinsic mechanisms of immune evasion include related stem cell pathway activation, the low-level expression of cellular antigen processing and presentation molecules, and the high-level expression of CD47 and PD-L1. External mechanisms of immune evasion include HBV/HCV infection, alcoholic/nonalcoholic steatohepatitis, hypoxia stimulation, abnormal angiogenesis, and infiltration of suppressive immune cells (Figure 1) [27-29].

**Intrinsic factors of immune evasion**

**CSC signaling pathways and immune evasion**

In HCC CSCs, signaling pathways involved in self-renewal and differentiation characteristics mainly include the Wnt/β-Catenin signaling pathway, Notch signaling pathway, Hedgehog signaling pathway, TGF-β signaling pathway, and AKT signaling pathway [26, 30, 31]. The Wnt/β-Catenin signaling pathway and TGF-β signaling pathway are closely related to immune evasion in HCC [32]. Intriguing studies have demonstrated that the aberrant activation of the tumor-intrinsic Wnt/β-Catenin signaling pathway correlates with a low proportion of T cell infiltration in the tumor microenvironment (TME) of HCC and melanoma tumor samples [33, 34]. Tang and colleagues suggested that there was a functional link between the TGF-β signaling pathway...
and IL-6 in HCC [35]. Moreover, IL-6 (Th2 cytokine) and TGF-β play an important role in the generation of an inhibitory immune microenvironment, antagonizing cytotoxic T lymphocytes (CTLs) and inducing antitumor immunity [36, 37]. Other studies have found that Notch pathway activation was associated with low CTL activity by recruiting tumor-associated macrophages (TAMs) or myeloid-derived suppressor cells (MDSCs) in other tumors (including pancreatic cancer, ovarian cancer, prostate cancer) [38-41].

**Immunological properties of CSCs in HCC**

A major mechanism by which CSCs avoid being attacked by the immune system involves minimization of antigenicity by downregulating key components of the cellular antigen processing and presentation machinery, mainly including transporters associated with antigen processing (TAP) and/or major histocompatibility complex (MHC) molecules [7, 32]. CSCs or other tumor cells of HCC lack targetability due to rare presentation by human leukocyte antigen (HLA) complexes [42]. In addition, Di Tomaso et al. found that glioblastoma CSCs were weakly positive and negative for MHC-I and MHC-II, leading to a lack of a T cell-mediated immune response [43]. Interestingly, downregulation or loss of HLA-I/II expression in spheres was also observed in tumor spheres (including colon, pancreas, and breast carcinoma), which were composed of CSCs [44]. Thus, downregulation or defects in antigen processing and presentation molecules provide a means for CSCs to evade CTL-mediated immune responses.

Additionally, CSCs have been found to express high levels of CD47 (the “don’t eat me” signal), which inhibits macrophage phagocytosis by binding to its cognate ligand, signal-regulatory-protein-α (SIRPα) [45, 46]. Lee and colleagues suggested that CD47 is preferentially expressed in liver CSCs, contributing to tumor initiation, self-renewal, and metastasis, and is significantly associated with poor clinical outcome [47]. Therefore, CD47 has been identified as a marker of CSCs in HCC [16]. Moreover, the high expression of CD47 in sorafenib-resistant HCC cells and samples is dependent on NF-κB expression [48]. TAM-derived IL-6 induced CD47 upregulation in HCC through activation of the STAT3 pathway and correlated with poor survival in HCC patients [49]. In summary, CSCs with high expression of CD47 in HCC can effectively avoid phagocytosis by macrophages and thus provide CSCs with a means of immune evasion.

Moreover, accumulating evidence has indicated that CSCs express high levels of PD-L1, which induce T cell apoptosis by binding to its cognate receptor PD-1. Hsu et al. demonstrated that epithelial-mesenchymal transition (EMT) enriched more PD-L1 in CSCs of breast and colon cancer cells by the EMT/β-catenin/STT3/PD-L1 signaling axis than non-CSCs [50]. In the case of squamous cell carcinoma of the head and neck (SCCHN), PD-L1 was also highly expressed on CD44+ cells (CSCs) compared to CD44− cells (non-CSCs), which was found to be dependent on the constitutive phosphorylation of STAT3 in CSCs [51]. Although CSCs have been found to overexpress PD-L1 in a variety of tumors [52], no relevant studies have focused on PD-L1 and CSCs in HCC. Recently, two types of anti-PD-1 monoclonal antibodies, nivolumab and pembrolizumab, have been FDA-approved as second-line therapies for advanced HCC, and a small percentage of patients have achieved complete remission (CR), resulting in long-term survival [53, 54]. Therefore, we speculate that anti-PD-1 therapy may be effective in clearing PD-L1-overexpressing CSCs in CR patients with HCC, which needs to be validated in future studies.

**External mechanisms of immune evasion**

**HBV/HCV infection**

HBV and HCV infections are major risk factors for HCC development and are also associated with the acquisition of a stem-like phenotype in HCC [55-58]. Hepatitis B virus X protein (HBx) is a 16.5 KDa protein, which has been shown to promote the expression of hepatoma stem cell markers (including EpCAM, CD133, CD90, etc.), contributing to tumor initiation and migration [59, 60]. In addition, chronic HCV infection can potentiate CSC generation by inducing CaM kinase-like-1 (DCAMKL-1), EMT, and hepatic stem cell-related factors [55, 56]. Moreover, chronic HBV/HCV infection promoted a viral-related inflammatory environment, which increased the expression of stemness-related properties (OCT4/Nanog, IGF-IR) by inflammatory cytokines in HCC [61]. Chang and colleagues also demonstrated that the activation of IL-6/IGFIR through induction of OCT4/NANOG expression was related to poor prognosis in HBV-related HCC [62]. Furthermore, a virus-associated inflammatory microenvironment can antagonize the antiviral immune response, as well as the antitumor response through recruitment of macrophages and the secretion of IL-6 [61, 62].

**Alcoholic and nonalcoholic steatohepatitis**

As we known, alcoholic, and nonalcoholic steatohepatitis (NASH) have emerged as an important risk factor in the development of HCC [63]. Chronic alcohol intake favors the formation of chronic inflammation, which induces reactive oxygen species (ROS) and DNA damage, thereby facilitating the activation of mutations in tumor stem cell-associated...
derived homolog (MYCN) high expression. Qin and colleagues demonstrated that neuroblastoma there is a close link between NASH and CSCs in HCC. Related studies also have verified that activation [67]. Moreover, NASH can lead to the reshaping of local TME, which weakens the antitumor functions of CD4+ T cells, cytotoxic CD8+ T cells, and natural killer (NK) cells and Th17 cells [70-74]. Additionally, alcohol or NASH-related HCC usually develops with advanced liver fibrosis and cirrhosis, which can induce the formation of hypoxia, contributing to CSC-mediated immune escape in HCC.

Hypoxia and Angiogenesis

Hypoxia is common in HCC, especially in patients with liver cirrhosis [75]. Hypoxia can induce EMT and increase the expression of stemness-related genes, which further increases the proportion of CSCs in HCC [76-79]. HIF-1α is a major transcription factor involved in the hypoxic response of hepatoma cells. Ye et al. demonstrated that HIF-1α-induced EMT led to the creation of an immunosuppressive TME to promote the metastasis of hepatocellular carcinoma cells. They found that hypoxia-induced EMT of hepatoma cells recruited and educated suppressive indoleamine 2,3-dioxygenase (IDO)-overexpressing TAMs to inhibit T-cell responses and promote immune tolerance in a CCL20-dependent manner [76]. Zhang and colleagues found that under a hypoxic microenvironment, the HIF-1α/IL-1b signaling loop between hepatoma cells and TAMs can promote EMT of cancer cells and metastasis [80]. Therefore, hypoxia can induce the phenotype of CSCs and further promote a suppressive TME, allowing tumor cells to escape antitumor immunity [81, 82].

An overwhelming number of studies have described the close crosstalk between CSCs and angiogenesis in the TME of various tumors, including HCC [29, 82, 83, 84]. VEGF is an important pro-angiogenic factor that has been shown to play a key role in the generation of a pro-angiogenic TME. Tang and colleagues documented that CD133+ CSCs of HCC can promote tumor angiogenesis through neurotensin/interleukin-8/CXCL1 signaling. Moreover, HCC CSCs preferentially secrete exosomes to promote VEGF secretion from endothelial cells, which in turn promotes tumor angiogenesis [85]. Liu et al. found that VEGF increases the proportion of CD133+ CSCs by activating VEGFR2 and enhances their self-renewal capacity by inducing Nanog expression in HCC [86]. Meanwhile, VEGF plays an important role in attenuating antitumor effects by negatively affecting antigen-presenting cells (APCs, such as DCs) and effector T cells while positively affecting suppressor immune cells (e.g., TAMs, Tregs, and MDSCs) [87, 88]. In summary, the crosstalk between CSCs and angiogenesis may contribute to the suppressive immune microenvironment and immune evasion observed in HCC.

Intratumoral hypoxia is a key driver of tumor angiogenesis [89]. Related studies have suggested that HIF-1α can bind to the promoter region of the VEGF gene and promote VEGF expression [90]. In summary, the close link between hypoxia, CSCs, and angiogenesis may play an important role in antitumor immunity evasion for HCC patients.

CSC-suppressive immune cell interactions

Over the decades, a large number of studies have been accumulated that extensively describe the interaction of CSCs with the immune system [91, 92]. TAMs, as one of the most infiltrating inflammatory cells in the TME, are classified as M1 (tumor-suppressing phenotype) and M2 (tumor-promoting phenotype) macrophages (MΦs). In the TME, TAMs are mostly M2 MΦs that play an important role in attenuating the antitumor immune response [93, 94]. Several studies have revealed that CSCs and TAMs can interact closely with each other to suppress antitumor immune effects in various tumors [95, 96]. Prostate CSCs can secrete some immunosuppressive molecules, such as TGF-β and IL-4, to promote M2 MΦ polarization [97]. CSCs in glioblastoma multiforme can secrete peristin to recruit TAMs [98]. Emerging evidence has also revealed that TAMs play a predominant role in the induction and maintenance of CSCs in various tumors by some secretory proteins.
[96]. In HCC, TAM-derived IL-6 can promote the expansion of CD44+ CSCs via the STAT3 signaling pathway [99]. At the same time, TAMs can also secrete TGF-β to promote CSC-like properties by inducing EMT in HCC [100]. As previously described, CD47 has been identified as a marker of CSCs in HCC, which can escape phagocytosis by M1 MΦs in the TME [16], and hypoxia-induced CSCs can secrete CCL20 to recruit IDO+ TAMs to inhibit T-cell responses and promote immune tolerance [76]. Altogether, these findings indicate a predominant role of TAMs in driving the immune evasion of CSCs in HCC. Moreover, NK cells, as key components of the innate immune system, are anti-tumor effector cells, which also can be impaired by soluble cytokines present in the TME (including CSC-derived cytokines), such as PGE2, IL-10, TGF-β1, granulin-epithelin precursor (GEP), and IDO [101, 102]. In HCC, GEP is overexpressed in tumor tissue but not in the adjacent normal tissue, which regulated the expression of CSC-related signaling molecules β-catenin, Nanog, Oct4, and Sox2 [103]. And, GEP renders hepatocellular carcinoma cells resistant to NK cytotoxicity by down-regulating surface expression of MHC class I chain-related molecule A (MICA), ligand for NK activated receptor NK group 2 member D (NKG2D), and up-regulating human leukocyte antigen-E (HLA-E), ligand for NK inhibitory receptor CD94/NKG2A [102].

MDSCs are another type of suppressive immune cell that seems to enable CSCs to escape antitumor immunity [104]. Increasing evidence suggests that MDSCs can secrete inflammatory molecules such as prostaglandin E2 (PGE2), IL-6, and nitric oxide (NO) to foster stemness of tumor cells in cervical cancer or breast cancer [105-107]. Conversely, glioblastoma CSCs also promote the survival and immunosuppressive activities of MDSCs by secreting macrophage migration inhibitory factor (MIF) [108]. In HCC, MDSCs can inhibit NK cells in patients via the NKP30 receptor [109]; hypoxia can induce the recruitment of MDSCs in the TME through chemokine (C-C motif) ligand 26 [110]. Moreover, Xu et al. found that drug-resistant HCC cell-derived IL-6 can enhance the expansion and immunosuppressive function of MDSCs [111]. Additionally, HCC CSCs can enhance the production of VEGF, thereby promoting MDSC recruitment in the TME [87]. Therefore, the interaction between CSCs and MDSCs can further contribute to immune evasion in HCC. Furthermore, HCC CSCs can attenuate antitumor effects by interacting with Treg cells and cancer-associated fibroblasts (CAFs) [112].

**Immunotherapeutic approaches targeting CSCs**

**Antibody immunotherapy based on markers of CSCs**

During the decades, several monoclonal antibodies (mAbs) have been successfully used in clinical patients for the treatment of human cancer, such as antagonists of VEGF, bevacizumab for colorectal cancer, ramucirumab for HCC and so on [113, 114]. The mechanisms of antibody-based approaches for targeting CSCs are mainly divided into two parts: the direct inhibitory effect of mAbs and antibody-dependent cellular cytotoxicity (ADCC) [115]. Additionally, several bispecific mAbs (BiTE antibodies) consisting of CSC and T cell targets displayed good antitumor effects in some preclinical studies and clinical trials [116].

EpCAM is a common marker of CSCs in HCC. Sun et al. indicated that EpCAM+ circulating tumor cells were associated with poor prognosis in HCC patients after curative resection [117]. In HCC cells, EpCAM expression was found to be dependent the activation of the Wnt/β-catenin signaling pathway, and EpCAM could directly bind to the downstream transcription factor Tcf4, which contributed to the formation of the Tcf4/β-catenin complex [118]. In an HCC preclinical study, an EpCAM/CD3 bispecific antibody (anti-EpCAM bispecific T cell engager (BiTE) 1H8/CD3) induced strong peripheral blood mononuclear cell-dependent cellular cytotoxicity, inducing strong elimination of HCC cells in vitro and vivo [119]. Currently, several II/III stage clinical trials (NCT01320020, NCT00822809, NCT00836654, etc) have shown that BiTE catumaxomab (Anti-EpCAM x Anti-CD3) can effectively improve the quality of life and survival time of malignant ascites (MA) from ovarian and nonovarian (including gastric, pancreatic, and breast, etc.) cancer patients [120-122]. Moreover, according to the analysis of peritoneal fluid samples from 258 MA patients in a phase II/III study (NCT00836654), catumaxomab therapy can significantly promote the activation of peritoneal T cells and eliminate EpCAM+ tumor cells in a manner associated with the release of proinflammatory Th1 cytokines [123]. However, in a randomized phase II trial (NCT01504256), compared with chemotherapy alone, catumaxomab followed by chemotherapy did not decrease peritoneal metastasis in gastric cancer patients [124]. Therefore, although catumaxomab has achieved promising effects in MA, clinical trials need further exploration in solid tumors, including HCC.

Given that CD47 acts as a marker for HCC CSCs and is crucial for evading phagocytosis by macrophages; thus, targeting CD47 is a promising approach to affect CSCs [125]. Preclinical studies...
demonstrated that anti-CD47 antibody effectively inhibited the growth of HCC, while combination chemotherapy had a synergistic antitumor effect [126, 127]. Currently, several anti-CD47 antibodies are currently being studied in clinical trials for a variety of human cancers [128]. Related phase I trials suggest that CD47 blockade is well tolerated in patients with hematological malignancies and solid tumors [129, 130]. A phase 1b study (NCT02953509) involving patients with relapsed or refractory non-Hodgkin's lymphoma revealed that a total of 50% of the patients had an objective response, with 36% having a complete response, after receiving combination therapy of the Hu5F9-G4 antibody (CD47 blockade) and rituximab [130]. However, the effects of antibody-based therapies targeting CD47 need to be further explored in the future in large phase II/III randomized controlled clinical trials.

Based on other HCC CSC markers, such as CD133, CD44, and CD24, related mAbs have demonstrated their effectiveness in eliminating HCC CSCs in preclinical models. Jianhua Huang and colleagues demonstrated that cytokine-induced killer (CIK) cells bound with anti-CD3/anti-CD133 bispecific antibodies can effectively target and kill CD133+ HCC CSCs in vitro and in vivo [131]. Wang et al. showed that CD44 antibody-targeted liposomal nanoparticles can target and eliminate HCC CSCs in preclinical models [132]. In addition, a phase I trial (NCT01358903) involving patients with advanced, CD44-expressing solid tumors revealed that RG7356, an anti-CD44 humanized antibody, is well tolerated but has limited clinical efficacy (21% patients, stable disease) [133]. Ma et al. suggested that anti-CD24 antibody conjugating doxorubicin can improve antitumor efficacy and has less systemic toxicity in an HCC preclinical model [134]. At the same time, a high-affinity humanized anti-CD24 antibody (hG7-BM3-VcMMAE conjugate) was designed to target hepatocellular carcinoma in vivo [135]. However, these HCC CSC marker-specific, antibody-based therapies require further clinical trials for validation.

**Immune checkpoint inhibitors and antiangiogenic therapy**

The rapid development and remarkable success of checkpoint inhibitors in the activation of CTLs led to cancer immunotherapy being named the “Breakthrough of the Year” by Science in 2013 [136]. Considering that CSCs can induce T-cell apoptosis by high expression of PD-L1, which binds to PD-1, immune checkpoint inhibitors may play an important role in CSC targeted therapy. Two PD-1 inhibitors, nivolumab and pembrolizumab, have been approved by the FDA for HCC after treatment failure on sorafenib based on two phase II trials, the Checkmate-040 study and the Keynote-224 trial, respectively [137, 138]. Reportedly, these two trials demonstrated RECIST1.1 objective response in 15-20% of HCC patients, including a small number of these patients with durable responses.

As one of the most vascular solid tumors, the role of angiogenesis has been extensively studied in HCC, and CSCs play an important role in promoting angiogenesis in HCC. Multiple kinase inhibitors with anti-angiogenic activity, such as sorafenib and lenvatinib, have been approved by the FDA for the treatment of advanced HCC [139, 140]. Additional anti-angiogenic multi-kinase inhibitors, such as regorafenib and cabozantinib, have been approved for the treatment of advanced HCC after treatment failure with sorafenib [141, 142]. Moreover, according to the results of the phase III trials REACH and REACH-II, ramucirumab (an anti-VEGF antibody) has been approved for patients with unresectable HCC with AFP ≥ 400 ng/dL who experience sorafenib failure [143, 144]. In sum, these effective antiangiogenic therapies in HCC may exert antitumor effects by indirectly targeting CSCs. In addition, the close crosstalk between CSCs and angiogenesis in the TME of HCC supports an inhibitory immune microenvironment, leading to antitumor immune evasion. Therefore, the combination of immunotherapy with VEGF antagonists in HCC is another new promising direction [145]. Recently, a global, open-label, phase III trial (IMbrave150) showed that combining atezolizumab (PD-L1 inhibitor) with bevacizumab resulted in better overall and progression-free survival than sorafenib in patients with unresectable HCC (NCT03434379) [146]. Based on these exciting results, the FDA approved bevacizumab in combination with atezolizumab as an updated first-line systemic therapy for patients with unresectable HCC [147]. Additionally, the REGONIVO trial (phase Ib trial, NCT03406871) demonstrated that the combination of regorafenib plus nivolumab led to an objective response in 20 patients (40%) with gastric and colorectal cancer [148]. Taken together, these findings indicated that checkpoint inhibitors in combination with anti-angiogenic inhibitors may lead to the depletion of CSCs, which contributed to the success of these trials.

**CAR-T/TCR-T targeting CSCs**

The advent of chimeric antigen receptor (CAR) T cell immunotherapy opens a new avenue in adoptive cell therapy, indicating the next breakthrough in immunotherapy [149]. According to these unprecedented clinical outcomes of CD19-directed
CAR T-cells in patients with certain refractory B cell malignancies, the FDA approved two anti-CD19 CAR-T cell therapies (lisocabtagene and axicabtagene cloleucel) for the treatment of certain hematological malignancies in 2017. Then, the American Society of Clinical Oncology named CAR-T cell therapy “advance of the year” in 2018 [150]. Indeed, using CAR-T cells to target CSCs is an interesting and promising immunological approach for treating HCC [151]. According to the web of clinical trials (https://clinicaltrials.gov/), the most registered type of HCC-related CAR-T clinical trial is GPC-3-targeted CAR-T, mainly because GPC-3 is the specific cell surface marker of HCC [152]. In a phase I study (NCT02395250), the results showed that CAR-GPC3 T-cell therapy is well tolerated in GPC3-positive patients with refractory or relapsed HCC, in which two patients had partial responses [153]. Moreover, based on the CSC-associated surface markers of HCC, several CAR-T-related clinical trials are ongoing. Wang et al. conducted a phase I clinical study (NCT02541370) using autologous CAR-CD133 T-cells to treat 23 patients with advanced and CD133-positive tumors, including 14 advanced HCC patients. The results showed that CAR-CD133 T-cell therapy was feasible and had controllable toxicities; 3 patients achieved partial remission (including 1 HCC patient), and 14 patients (including 9 HCC patients) achieved stable disease; the 3-month disease control rate was 65.2%, and the median progression-free survival was 5 months [154]. Additionally, the efficacy of EpCAM-targeted CAR-T cells has been demonstrated preclinically for several solid tumors, such as colon, prostate, and peritoneal cancers [155-158]. Currently, one CAR-EpCAM T-cell clinical trial (NCT03013712) has been registered and is recruiting EpCAM-positive cancer (including HCC).

Another promising adoptive cell therapy, T-cell receptor (TCR)-engineered T-cell immunotherapy, has attracted widespread attention and been extensively studied. Compared with CAR-T cells, TCR-T cells can recognize intracellular tumor-associated antigens depending on the MHC complex. In HCC, targeting alpha-fetoprotein (AFP) or HBV/HCV-associated antigens with TCR-T therapies has shown powerful antitumor effects in preclinical models [159-162]. Moreover, a series of clinical trials targeting AFP (NCT03971747, NCT04368182) or virus-associated antigens (NCT02686372, NCT03899415) with TCR-T therapies for HCC are currently underway. Considering that HBV and HCV infections contribute to the acquisition of a stem-like phenotype in HCC, TCR-T cells targeting special viral antigens may effectively clear CSCs. In any case, viral antigen-specific TCR-T cell injection may be a promising strategy for HCC.

NK cell-based cancer immunotherapies

As mentioned previously, CSCs of HCC have low expression of MHC molecules, which contribute to immune escape. Interestingly, the inhibitory receptors of NK cells can recognize MHC-I molecules, hence, NK cells do not usually attack normal cells [163]. Therefore, the low-expression of MHC-I on CSCs will make them to be susceptible to be killed by NK cells [164], indicating that NK cell-based cancer immunotherapies may be a promising treatment strategy to target CSCs [32]. NK cell-based immunotherapies have achieved encouraging results in hematologic cancers, including IL2-activated haploidentical NK cells infusions [165], and anti-CD19 CAR-NK cell therapy [166, 167]. Although some progress is also being made to apply NK cell-based therapies against solid tumors, response rates in patients remain to be unsatisfied [163]. In HCC, several NK-cell based I/II phase clinical trials are in progress, such as, autologous/allogeneic NK cells infusion or in combination with other therapies (NCT03319459, NCT04162158, NCT03592706), anti-MUC1 CAR-NK cells (NCT02839954), and so on [168]. Moreover, related studies have showed that chemotherapy or radiation therapy can increase the amounts of CSCs in various tumor and induce up-regulating NKG2D ligands MICA and MICB on CSCs, indicating that NK cell-based immunotherapies in combination with radiation therapy or chemotherapy could better eradicate CSCs in HCC [169].

Vaccines targeting CSCs

CSC-directed immunotherapies to promote tumor cell recognition and elimination by the immune system are mainly focused on the use of DC vaccines [170]. Related studies have suggested that DC vaccination using CSC-associated antigens can elicit antigen-specific T-cell responses against CSCs in vitro and in vivo [171-173]. In HCC, Choi and colleagues suggested that DCs stimulated by EpCAM peptides enhance T cell activation and generate CTLs, thus effectively killing HCC CSCs [174]. In addition, SP cell lysate-pulsed DCs have been demonstrated to induce a special T cell response against HCC CSCs and suppress tumor growth in vivo [172]. To date, more than 200 completed clinical trials have involved the use of DC vaccines for cancer treatment. Sipuleucel-T (Provenge) is the only FDA-approved DC vaccine loaded with a fusion antigen protein composed of GM-CSF and prostatic acid phosphatase; it has been used to treat prostate cancer patients and has extended the median overall survival by
approximately 4 months [175]. Based on the remarkable success of checkpoint inhibitors in the treatment of various tumors in the clinic, DC vaccination in combination with checkpoint inhibitors may be an ideal immunotherapy to foster powerful initial specific effector T cell activation [176]. Moreover, as shown in the web of clinical trials (https://clinicaltrials.gov/), a series of clinical trials are ongoing based on DC vaccines (loaded with HCC neoantigens or virus-associated antigens) or combined PD-1 monoclonal antibodies. However, DC vaccination using CSC-associated antigens against HCC needs to be further investigated in future preclinical and clinical trials.

Conclusions

In summary, accumulating evidence has suggested that CSCs can create an immunosuppressive microenvironment through certain intrinsic and extrinsic mechanisms, resulting in immune evasion in HCC. The intrinsic mechanisms mainly include the following: 1. the activation of immune-related CSC pathways; 2. low-level expression of TAP and/or MHC molecules; and 3. high-level expression of CD47 and PD-L1. The external mechanisms mainly include the following: 1. HBV/HCV infection; 2. alcoholic/nonalcoholic steatohepatitis; 3. hypoxia stimulation; 4. abnormal angiogenesis; and 5. infiltration of suppressive immune cells (Figure 1). Currently, immunotherapeutic approaches targeting HCC CSCs mainly include antibody immunotherapy based on CSC markers, immune checkpoint inhibitors, antiangiogenic therapy, CAR-T/TCR-T cell therapy, NK cell-based cancer immunotherapies, and DC vaccines (Figure 2).

However, there are still some hindrances to achieving efficacious immunotherapy targeting CSCs in HCC [11, 177]. First, the abovementioned immunotherapies targeting CSCs in HCC are based on CSC-specific molecular markers. However, almost all identified stem cell markers are not unequivocally exclusive CSC markers for HCC; in other words, they are also shared with normal stem cells. Second, the existence of intertumor, intratumor, and CSC heterogeneity is a daunting challenge in the development of immunotherapy targeting HCC CSCs [23, 178]. Additionally, several studies have demonstrated that the HCC CSCs are plastic and can be converted from tumor cells without a stem phenotype, which can be induced by virus infection, crosstalk between CSCs and tumor cells, hypoxia stimulation, and conventional therapies [76, 77, 106, 179, 180]. This plasticity and instability of the CSC phenotype in HCC is a major obstacle for effective immunotherapy targeting CSCs. Considering that CSCs are a rare subpopulation in tumor tissue, targeted CSC therapy alone is presumed to be inadequate for the effective elimination of tumors. Thus, the combination of CSC-targeted immunotherapy with currently used cancer therapies, such as chemotherapy, radiation therapy, antiangiogenic therapy, and checkpoint inhibitors, may effectively eradicate HCC tumors. Moreover, the success of the IMbrave150 trial in advanced HCC patients has illustrated the importance and necessity of combined therapy.

Finally, we think that the most attractive research prospects focused on CSC-targeted immunotherapy in HCC mainly include the following: a) the identification of unequivocal CSC-specific molecular markers through multiomics analyses, such as the combination of proteomics and single-cell analysis; b) dissection of the complex mechanisms of the crosstalk between CSCs and immune cells; and c) validation of the effects of combinatorial treatments in future preclinical and clinical trials, such as DC vaccination (loaded with a mixed CSC and non-CSC special antigens) in combination with checkpoint inhibitors, CAR-T/TCR-T therapy in combination with antiangiogenic therapy, anti-CD47 antibody combined with CAR-T, CAR-T combined NK cell–based cancer immunotherapies, and so on. In conclusion, based on the heterogeneity, plasticity and scarcity of HCC CSCs, it is suggested that combinatorial treatments will be more efficacious than anti-CSC treatment alone. Overall, future immunotherapy should serve as a model for combined therapy.

Acknowledgements

This study was supported by grants from the National Natural Science Foundation of China (No. 82074208, 81472346 and 81802355), and Zhejiang Natural Science Foundation (LY20H160033).

Consent for publication

All authors agreed to submit for consideration for publication in this journal.

Competing Interests

The authors have declared that no competing interest exists.

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018; 68: 394-424.
2. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin. 2020; 70: 7-30.
3. Singal AG, Lampertico P, Nahon P. Epidemiology and surveillance for hepatocellular carcinoma: New trends. J Hepatol. 2020; 72: 250-61.
91. Kise K, Kinugasa-Katayama Y, Takakura N. Tumor microenvironment for
90. Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet AL, et al.
88. Tang KH, Ma S, Lee TK, Chan YP, Kwan PS, Tong CM, et al. CD133(+) liver
85. Xiong XX, Qiu XY, Hu DX, Chen XQ. Advances in Hypoxia-Mediated
82. Nishida N, Kudo M. Immunological Microenvironment of Hepatocellular
75. Lin D, Wu J. Hypoxia inducible factor in hepatocellular carcinoma: A
72. Shalapour S, Lin XJ, Bastian IN, Brain J, Burt AD, Aksenov AA, et al.
73. Bhattacharjee J, Kirby M, Softic S, Miles L, Salazar-Mendoza RM, Shivakumar P.
70. Ma C, Kesarwala AH, Eggert T, Medina-Echeverz J, Kleiner DE, Jin P, et al.
69. Qin XY, Su T, Yu W, Kojima S. Lipid desaturation-associated endoplasmic
68. Machida K, Tsukamoto H, Mkrtchyan H, Duan L, Dynnyk A, Liu HM, et al.
65. Anstee QM, Reeves HL, Kotsiliti E, Govaere O, Heikenwalder M. From NASH
to HCC: current concepts and future challenges. Nat Rev Gastroenterol
2019; 16: 411-28.
63. Peng JM, Bera R, Chiou CY, Yu MC, Chen TC, Chen CW, et al. Actin
P21-induced factor-alpha 1 stimulating tumor growth factor-beta1-induced epithelial-
mesenchymal transition in Hepatocellular Carcinoma and Its Clinical Implication.
Onco Targets Ther. 2017; 10: 347-54.
61. Elaimy AM, Mercurio AM. Convergence of VEGF and YAP/TAZ signaling:
Implications for angiogenesis and cancer biology. Sci Signal. 2018; 11: eaat165.
60. Beck B, Driessens G, Goosens S, Youssef RK, Kuchnio A, Caauwe A, et al. A
vascular niche and a VEGF-NRP1 loop regulate the initiation and stemness
of skin tumours. Nature. 2011; 478: 399-403.
59. Tang KH, Ma S, Lee TK, Chan YP, Kwan PS, Tong CM, et al. CD133(+) liver
initiating cells promote tumor angiogenesis, growth, and self-renewal through
VEGFR2/neurotrophin-1/ERK signaling. Hepatology. 2012; 55: 870-7.
58. Liu K, Hao M, Ouyang Y, Zheng J, Chen D. CD133(+) cancer stem cells
promoted by VEGF-accelerate the recurrence of hepatocellular carcinoma. Scie
Repir. 2017; 7: 41497.
57. Rahma OE, Hodi FS. The intersection between Tumor Angiogenesis and
Immune Suppression. Curr Cancer Res. 2019; 25: 5449-57.
56. Veron T, Colussi O, Marcheteau E, Pernet S, Nizard M, Pointet AL, et al.
VEGF-A modulates expression of inhibitory checkpoints on CD8+ T cells in
tumors. J Exp Med. 2015; 212: 129-38.
55. Pettirillo M, Patella F, Pespana F, Suter MB, Ierardi AM, Angileri SA, et al.
Hypoxia and tumor angiogenesis in the era of hepatocellular carcinoma
translational loco-regional treatments. Future Oncol. 2018; 14: 2957-67.
54. Ping YF, Bie H, Zhang M, Wang Y, Duan L, Zhang R, et al. Circulating stem
cell-like epithelial cell adhesion molecule-positive tumor cells indicate poor
prognosis of hepatocellular carcinoma after curative resection. Hepatology.
2013; 57: 1458-68.
53. Yamashita T, Budhu A, Forgacs M, Wang XW. Activation of hepatic stem cell
marker EpCAM by Wnt-beta-catenin signaling in hepatocellular carcinoma. 
Cancer Res. 2007; 67: 10831-9.
52. Plaks V, Kong N, Werb Z. The cancer stem cell niche: how essential is the
niche in regulating stemness of tumor cells? Cell stem cell. 2015; 16: 225-38.
51. DeNardo DG, Ruflitt B. Macrophages as regulators of tumour immunity and
immunotherapy. Nat Rev Cancer. 2019; 19: 369-82.
50. Pathria P, Louis TL, Varner JA. Targeting Tumor-Associated Macrophages in
Cancer. Trends Immunol. 2019; 40: 310-27.
49. Muppala S. Significance of the Tumor Microenvironment in Liver Cancer
Progression. Crit Rev Oncol Hematol. 2016; 95: 1-9.
48. Muller L, Tunger A, Pesca I, Wehner R, Temme A, Westphal D, et al. 
Bidirectional Crossstalk Between Cancer Stem Cells and Immune Cell Subsets.
Front Immunol. 2020; 11: 140.
47. Vescovi A, Handle F, Santer FR, McNeill RV, Seed RI, Collins AT, et al. The
immunosuppressive cytokine interleukin-4 increases the clonogenic potential
of prostate stem-like cells by activation of ST6ST6 signaling. Oncogene.
2017; 36: e342.
46. Zhou W, Ke SQ, Huang Z, Flavahan W, Fang X, Paul J, et al. Periostin secreted
by glialblasta stem cells recruits M2 tumor-associated macrophages and
promotes malignant growth. Nat Cell Biol. 2015; 17: 170-82.
45. Wan S, Zhao E, Kryczek I, Vatan L, Sadowska Y, Ludema G, et al. Tumor-
associated macrophages produce interleukin 6 and signal via ST1A3 to
promote expansion of human hepatocellular carcinoma stem cells. 
Gastroenterology. 2014; 137: 1903-44.
44. Fan QM, Jing YY, Yu GF, Kou XR, Ye F, Gao L, et al. Tumor-associated macrophages
promote cancer stem cell-like properties via transforming growth factor-
beta/beta-1 signaling, which mediates tumor transition in hepatocellular
 carcinoma. Cancer Lett. 2014; 352: 160-8.
43. Dianat-Moghadam H, Rokni M, Marofi F, Panahi Y, Yousefi M. Natural killer
cell-based immunotherapy: From transplantation toward targeting cancer
immunotherapy. J Cell Physiol. 2018; 233: 259-73.
42. Cheung PF, Yip CW, Wong NC, Fong DY, Ng LW, Wan AM, et al. Granulin-epithelin
precursor renders hepatocellular carcinoma cells resistant to
natural killer cytotoxicity. Cancer Immunol Res. 2014; 2: 1209-19.
41. Cheung PF, Cheng CK, Wong NC, Ho JC, Yip CW, Lui VC, et al. Granulin-epithelin
precursor is an oncofetal protein defining hepatic cancer stem cells. 
PLoS One. 2011; 6: e28246.
40. Tannievor G, Ayata C. Mutualistic Effects of the Myeloid-Depressed Suppressors
and Cancer Stem Cells in the Tumor Microenvironment. Crit Rev Oncog. 2019;
24: 61-7.
39. Nogueira R, Soto MA, Lopes MF, et al. Tumor cell-secreted tumor necrosis factor
alpha induces the expression of CD44(+) cells and stimulates tumor
angiogenesis. Adv Drug Deliv Rev. 2016; 99: 197-205.
121. Wimberger P, Gilet H, Gonschior AK, Heiss MM, Moehler M, Oskay-Oezcelik, et al. The promise of adoptive cellular immunotherapies in hepatocellular carcinoma. Oncoimmunology. 2020; 9: 1673129.

122. Baumann K, Pfisterer J, Wimberger P, Burchardi N, Kurzeder C, du Bois A, et al. Lenvatinib versus placebo as second-line treatment in patients with hepatocellular carcinoma following first-line therapy with sorafenib. Lancet. 2017; 389: 2492-502.

123. Kudo M, Finn RS, Qin S, Han KH, Piscaglia F, et al. Lenvatinib versus sorafenib in patients with hepatocellular carcinoma: a phase III trial. Lancet. 2016; 387: 743-51.

124. Bruix J, Qin S, Han KH, Piscaglia F, Ikeda K, Piscaglia F, et al. Lenvatinib versus sorafenib in patients with hepatocellular carcinoma who progressed on sorafenib therapy (REACH): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet. 2017; 389: 2492-502.

125. Wang Y, Chen M, Wu Z, Tong C, Dai H, Guo Y, et al. CD133-humanized anti-CD24 antibody to target hepatocellular carcinoma by a novel doxorubicin via conjugating to anti-CD24 antibody results in enhanced antitumor potency for hepatocellular carcinoma both in vitro and in vivo. J Cancer Clin Oncol. 2013; 14: 156-68.

126. Feng M, Jiang W, Kim BYS, Zhang CC, Fu YX, Weissman IL. Phagocytosis inhibition of tumor cells by SIRPα Immune Checkpoint. J Immunol. 2020; 164: 1-7.

127. Lo J, Lau EY, So FT, Lu P, Chan VS, Cheung VC, et al. Anti-CD47 antibody approaches in the treatment of hepatocellular carcinoma. Liver Int. 2016; 36: 737-45.

128. Logtenberg MEW, Scheeren FA, Schumacher TN. The CD47-SIRPα axis: a step forward. Lancet Oncol. 2020; 21: 412.

129. Xu Q, Liu G, Yuan X, Xu M, Wang H, Ji J, et al. Antigen-specific T-cell response from dendritic cell vaccination using cancer stem-cell-associated antigens. Stem Cells. 2009; 27: 1734-43.

130. Li X, Zhang P, Lin G, Gao Y, Pan Y, Yin H, et al. Antigen-specific T-cell response from dendritic cell vaccination using side population cell-associated antigens targets hepatocellular carcinoma. Tumour Biol. 2016; 37: 11267-78.

131. Huang J, Li C, Wang Y, Lv H, Guo Y, Dai H, et al. Cytokine-induced killer (CIK) cells bound with anti-CD3/anti-CD33 bispecific antibodies target CD133(high) cancer stem cells in vitro and in vivo. Clin Immunol. 2013; 149: 1-13.

132. Loj L, Lau EY, So FT, Lu P, Chan VS, Cheung VC, et al. Anti-CD47 antibody suppresses tumour growth and augments the effect of chemotherapy treatment in hepatocellular carcinoma. Liver Int. 2016; 36: 737-45.

133. Logtenberg MEW, Scheeren FA, Schumacher TN. The CD47-SIRPα axis: a step forward. Lancet Oncol. 2020; 21: 412.

134. Zou Y, Wen P, Li M, Li Y, Li XA. Construction of chimeric antigen receptor-modified T cells targeting EpCAM and assessment of their anti-tumor effect on cancer cells. Mol Med Rep. 2019; 20: 2355-64.

135. Peng Z, Wu Y, Ma W, Zhang H, Yang YG. Adaptive T-cell therapy of prostate cancer targeting the cancer stem cell antigen EpCAM. BMC Immunol. 2015; 16: 1.

136. Ang WX, Li Z, Chi Z, Du SH, Chen C, Tay JC, et al. Intraperitoneal immunotherapy with T cells stably and transiently expressing anti-EpCAM CAR in xenograft models of peritoneal carcinomatosis. Oncotarget. 2017; 8: 13545-59.

137. Zheng BL, Li D, Gong YL, Huang Y, Qin DY, Jiang L, et al. Preclinical Evaluation of Chimeric Antigen Receptor-Modified T Cells Specific to Epithelial Cell Adhesion Molecule for Treating Colorectal Cancer. Hum Gene Ther. 2019; 30: 492-502.

138. Bertolotti A, Brunetto M, Maini MK, Bonino F, Qasim W, Stass H. T-cell receptor-therapy in HBV-related hepatocellular carcinoma. Oncoimmunology. 2015; 4: e100854.

139. Zhu W, Peng Y, Wang L, Hong Y, Jiang X, Li Q, et al. Identification of a-fibronectin-specific T-cell receptors for hepatocellular carcinoma immunotherapy. Hepatology. 2016; 63: 574-89.

140. Tan AT, Yang N, Lee Krishnamoorthy T, Oei V, Chua A, Zhuo X, et al. Use of Expression Profiles of HBV-DNA Integrated Into Genomes of Hepatocellular Carcinoma Cells to Select T Cells for Immunotherapy. Gastroenterology. 2019; 156: 1594-606.

141. Spear TT, Callender GG, Roszkowski JX, Molesky KM, Simms PE, Foley KC, et al. TCR gene-modified T cells can efficiently treat established hepatocellular carcinoma tumors. Cancer Immunol Immunother. 2019; 68: 255-68.

142. Wimberger P, Gilet H, Gonschior AK, Heiss MM, Moehler M, Oskay-Oezcelik, et al. The promise of adoptive cellular immunotherapies in hepatocellular carcinoma. Oncoimmunology. 2020; 9: 1673129.

143. Complete Remission with Reduction of High-Risk Clones following Haploidentical NK-Cell Therapy against MDS and AML. Clin Cancer Res. 2018; 24: 1834-44.

144. Imai C, Iwamoto S, Campana D. Genetic modification of primary natural killer cells overcomes inhibitory signals and induces specific killing of leukemic cells. Blood. 2005; 106: 376-83.

145. Liu E, Marin D, Banerjee PC, Macapinlac HA, Thompson P, Basar R, et al. Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphomas. N Engl J Med. 2010; 362: 545-53.

146. Mantovani S, Olivierio B, Varchetta S, Mele D, Mondelli MU. Natural Killer Cell Responses in Hepatocellular Carcinoma: Implications for Novel Immunotherapeutic Approaches. Cancers. 2020; 12.

147. Ames E, Canter RJ, Grossenbacher SK, Mac S, Chen M, Smith RC, et al. NK Cells Preferentially Target Tumor Cells with a Cancer Stem Cell Phenotype. J Immunol. 2015; 195: 4010-9.

148. Björklund AT, Carlsten M, Sohlberg E, Liu LL, Clancy T, Karimi M, et al. Complete Remission with Reduction of High-Risk Clones following Haploidentical NK-Cell Therapy against MDS and AML. Clin Cancer Res. 2018; 24: 1834-44.

149. Iwamoto S, Campana D. Genetic modification of primary natural killer cells overcomes inhibitory signals and induces specific killing of leukemic cells. Blood. 2005; 106: 376-83.

150. Liu E, Marin D, Banerjee PC, Macapinlac HA, Thompson P, Basar R, et al. Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphomas. N Engl J Med. 2010; 362: 545-53.

151. Mantovani S, Olivierio B, Varchetta S, Mele D, Mondelli MU. Natural Killer Cell Responses in Hepatocellular Carcinoma: Implications for Novel Immunotherapeutic Approaches. Cancers. 2020; 12.

152. Ames E, Canter RJ, Grossenbacher SK, Mac S, Chen M, Smith RC, et al. NK Cells Preferentially Target Tumor Cells with a Cancer Stem Cell Phenotype. J Immunol. 2015; 195: 4010-9.

153. Björklund AT, Carlsten M, Sohlberg E, Liu LL, Clancy T, Karimi M, et al. Complete Remission with Reduction of High-Risk Clones following Haploidentical NK-Cell Therapy against MDS and AML. Clin Cancer Res. 2018; 24: 1834-44.

154. Björklund AT, Carlsten M, Sohlberg E, Liu LL, Clancy T, Karimi M, et al. Complete Remission with Reduction of High-Risk Clones following Haploidentical NK-Cell Therapy against MDS and AML. Clin Cancer Res. 2018; 24: 1834-44.
173. Szaryńska M, Olejniczak A, Kobiela J, Laski D, Śledziński Z, Kmiec Z. Cancer stem cells as targets for DC-based immunotherapy of colorectal cancer. Sci Rep. 2016; 8: 12042.

174. Choi YJ, Park SJ, Park YS, Park HS, Yang KM, Heo K. EpCAM peptide-primed dendritic cell vaccination confers significant anti-tumor immunity in hepatocellular carcinoma cells. PloS One. 2018; 13: e0190638.

175. Cheever MA, Higano CS. PROVENGE (Sipuleucel-T) in prostate cancer: the first FDA-approved therapeutic cancer vaccine. Clin Cancer Res. 2011; 17: 3520-6.

176. Garg AD, Coulie PG, Van den Eynde BJ, Agostinis P. Integrating Next-Generation Dendritic Cell Vaccines into the Current Cancer Immunotherapy Landscape. Trends in immunology. 2017; 38: 577-93.

177. Sukowati CHC. Heterogeneity of Hepatic Cancer Stem Cells. Adv Exp Med Biol. 2019; 1139: 59-81.

178. Gao Q, Wang XY, Zhou J, Fan J. Heterogeneity of intermediate-stage HCC necessitates personalized management including surgery. Nat Rev Clin Oncol. 2015; 12: 10.

179. Karagonlar ZF, Akbari S, Karabicici M, Sahin E, Avci ST, Ersoy N, et al. A Novel Function for KLF4 in Modulating the De-differentiation of EpCAM(-)/CD133(-) nonStem Cells into EpCAM(+)/CD133(+) Liver Cancer Stem Cells in HCC Cell Line HuH7. Cells. 2020; 9: 1198.

180. Jayachandran A, Dhungel B, Steel JC. Epithelial-to-mesenchymal plasticity of cancer stem cells: therapeutic targets in hepatocellular carcinoma. J Hematol Oncol. 2016; 9: 74.