Research Article

In Silico Screening of Mutated K-Ras Inhibitors from Malaysian Typhonium flagelliforme for Non-Small Cell Lung Cancer

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K-ras is an oncogenic GTPase responsible for at least 15–25% of all non-small cell lung cancer cases worldwide. Lung cancer of both types is increasing with an alarming rate due to smoking habits in Malaysia among men and women. Natural products always offer alternate treatment therapies that are safe and effective.

Typhonium flagelliforme or Keladi Tikus is a local plant known to possess anticancer properties. The whole extract is considered more potent than individual constituents. Since K-ras is the key protein in lung cancer, our aim was to identify the constituents of the plant that could target the mutated K-ras. Using docking strategies, reported potentially active compounds of Typhonium flagelliforme were docked into the allosteric surface pockets and switch regions of the K-ras protein to identify possible inhibitors. The selected ligands were found to have a high binding affinity for the switch II and the interphase region of the ras-SOS binding surface.

1. Introduction

Cancer is a major health problem in Malaysia with a total of 2,048 cases registered with National Cancer Registry (NCR) in 2006 [1]. The disease is now the third leading cause of premature deaths in our country. Lung cancer is among the top five cancers affecting both male and female in Malaysia at 9.4 percent in peninsula Malaysia with 2100 Malaysians diagnosed every year [2, 3]. Smoking related diseases are increasing in Malaysia especially lung cancer [3, 4]. A recent study conducted by Liam et al. (2013) highlighted adenocarcinomas as the most frequent types of cancer among Malaysian men and women, smoker, and nonsmokers. With an incidence rate of 109.8 cases of cancer per 100,000, it is imperative to find treatments that are safe and effective [5].

Lung cancer can be divided into two major classes based on its biology, therapy, and prognosis, namely, non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC). NSCLC accounts for 75% to 80% of all lung cancer incidents while small cell lung cancer accounts for 15% to 25% of all lung cancer [6].

The ras pathway is an important signaling pathway that allows cell proliferation in response to stimulation of the epidermal growth factor receptor [7, 8]. These signals affect the production and regulation of other key proteins involved in cell proliferation. Studies have reported that K-ras mutation occurring in NSCLC varies between 16% and 40% [8]. K-ras, a GTPase, also known as V-Ki-ras-2 (Kirsten rat sarcoma viral oncogene) is a protein that in human is encoded by the K-Ras gene [9]. The normal protein is an essential part of the ras signaling pathway acting as a molecular switch. In the “off” state, it is bound to the guanine diphosphate nucleotide (GDP). It is turned on via the growth factor stimuli. The guanine nucleotide exchange factor, also known as the son of sevenless (SOS) protein, and the growth factor receptor-bound protein 2 (Grb-2) together enable the K-ras to become activated by exchanging the GDP molecule for the more active guanine triphosphate nucleotide (GTP). Once turned “on,” it recruits and activates downstream proteins...
necessary for the propagation of growth factor and other receptor signals. It possesses an intrinsic weak enzymatic activity which is enhanced by interaction with the GTPase activating protein (GAP) leading to cleavage of the terminal phosphate of the nucleotide guanine triphosphate (GTP) converting it into the guanine diphosphate (GDP). Upon conversion of GTP to GDP, K-ras is switched “off.” Despite being a weak GTPase, K-ras possesses two very important features involved in its switching “on” and “off.” Called switches because of their ability to change the conformation of the protein in the active and inactive state, the regions are involved in interacting with the nucleotides. The guanine nucleotide pocket of the K-ras is highly conserved and is lined with residues 11–16 [10, 11]. Because of the specific interactions of amino acid residues of this region with the GTP, mutations at the 12 and 13 amino acid positions in the enzyme lead to permanent cell proliferation because it cannot be hydrolysed and hence, the ras signaling function is unable to be turned “off” [12–14]. The mutated K-ras (mut-Kras) is an interesting drug target of several studies [15–18]. The major reason being that it provides fast resistance to the available drug therapy. Several EGFR, MEK inhibitors have been tried in single and combination. However, drug resistance develops quickly [17, 19–21].

Medicinal plants with anti-cancer effects are commonly used as alternative medicine because of their safety and toxicity profiles. Several herbal medicines have been studied for finding effective treatment of lung cancer. Only few toxicity profiles. Several herbal medicines have been studied used as alternative medicine because of their safety and

2. Materials and Methods

The three dimensional structures of the G12D mutated K-ras structure inhibitor bound to SOS pocket (PDB ID: 4DST), and two G12C mutated K-ras structure bound to allosteric sites (PDB ID: 4LUC and 4LYF). Autodock Vina 4.2 [27] was used to dock all ligands to the K-ras protein. Before that, we used Autodock tools downloaded from The Scripps Research Institute to prepare the ligand and protein file [29, 30]. All water molecules were removed and Kollman charges added as described in the Autodock Vina 4.2 manual [31, 32]. The grid box dimensions were obtained from the grid box widget by keeping the bound ligand sites as box centers. Control studies were performed with all ligand bound in the crystal structures before docking with test ligands from T. flagelliforme. Pheophorbide a and two related epimers were drawn in ChemSketch [33] based on reported structure by Lai et al. [22] while hexadecane and hexadecanoic acid structure were obtained from Pubchem. The 2D structures of the active constituents are tabulated in Table 1.

3. Results and Discussions

The results of highest binding affinity of the five active constituents for the GTP bound K-ras as well as the mutated protein structures are tabulated in Table 2. T. flagelliforme is a local Malaysian plant with anticancer activity when taken as a fresh juice prepared from freshly crushed plant. Researchers have reported its activity in lung cancer as a whole extract in dichloromethane [22, 23]. Lai et al. (2010) reported that the extract contained at least 11 chemical compounds of hydrophobic character [22]. The most predominant compounds were pheophorbide a, pheophorbide a’, pyropheophorbide a, methyl pyropheophorbide a, hexadecanoic acid, oleic acid, linoleic acid, linolenic acid, campesterol, stigmsterol, and β-sitosterol. The in vitro studies conducted by the group on the NCI-H23 lung cancer cell line had concluded that individual compound isolates had failed to show a significant anti-cancer behavior. However, the whole extracts were found to have an IC50 of 2.7 μg/mL suggesting that components had a synergistic effect on antiproliferation of cancerous cells. They also suggested that the constituents showed increased activity upon exposure to light. The authors had earlier reported an active whole extract of the plant comprising of hexadecanoic acid, 1-hexadecene, phytoh, and a phytol derivative with an IC50 of 7.5 μg/mL against NCI-H23 cell lines [23].

Mohan et al. (2010) conducted their study on the leukemic cells and showed the selectivity of the dichloromethane fractions for the cancerous cells. However, they did not indicate the effect of any particular constituent on the cancer cells [34].

In our study we focused on determining whether the proposed constituents by Lai et al. (2008, 2010) were able to target the K-ras protein either directly or allosterically. For direct inhibition we docked the selected ligands at or near the SOS binding pocket using the PDB structure 4DST [24]. The target binding site proposed by the authors is near the ras switches I and II and binding to this area was indicated
Table 1: Two dimensional structures of the constituents used in the study.

| Name            | Structure                      |
|-----------------|--------------------------------|
| Hexadecanoic acid | ![Structure](image1)            |
| Hexadecene      | ![Structure](image2)           |
| Pheophorbide a epimer | ![Structure](image3)          |
| Pheophorbide a’ epimer | ![Structure](image4)        |
Table 1: Continued.

| Name                   | Structure  |
|------------------------|------------|
| Pyropheophorbide epimer| ![Structure](image) |

Table 2: Binding affinity values obtained for the control and test ligands.

| Protein                        | Binding affinity Kcal/mol |
|--------------------------------|---------------------------|
|                                | Control | Pheophorbide A | Pheophorbide A’ | Pyropheophorbide A | 1-Hexadecene | Hexadecanoic acid |
| G12D mutated K-ras GDP SOS binding pocket (4DST) | −5.4 | −7.2 | −7.5 | −7.1 | −4.0 | −4.1 |
| G12C mutated K-ras GDP allosteric binding pocket (4LUC) | −8.0 | −7.0 | −6.8 | −7.3 | −4.8 | −4.6 |
| G12C mutated K-ras GDP allosteric binding pocket (4LYF) | −6.7 | −6.7 | −6.6 | −6.5 | −4.5 | −5.0 |

as interfering with ras-SOS binding surface [24]. This finding is important because the Ras-SOS complex is essential for activation of the K-ras since SOS initiates the GTP exchange process to the protein. According to the resolved structure of the ras-SOS complex (PDB ID: 1BKD) [35], the CDC25 binding region is tightly bound to the Switch II of ras and causes the disruption of the GDP bound structure [35]. Tyr 64 of the ras appears to be the anchoring residue for the Ras-SOS complex. Hence, the inhibitors should be designed to target the switch regions or the binding surface between the protein-protein complex. Where small molecules can modulate the switch regions of the ras due to space confinement, larger molecules can target the accessible surface areas between the protein-protein complex.

Our semiflexible docking experiment on the K-ras molecule with control ligand 4,6-dichloro-2-methyl-3-aminoethyl-indole (4DST) [24] had a binding affinity of −5.4 Kcal/mol. Among the test ligands the highest binding affinities were shown by the pheophorbide epimers. The observed docked poses, given in Figure 1, showed that the epimers were not near the binding site of the control ligand.

The test ligand that showed affinity for the same binding site as the control molecule was hexadecanoic acid with

![Image: docked poses of flagelliforme constituents to GTP bound K-ras.](image)
binding affinity $-4.1 \text{ Kcal/mol}$. It appeared to overlap the control molecule at the same position. The authors suggested that the binding pocket involves residues Lys5, Leu6, Val7, Ile55, Leu56, and Thr74 [24]. Our results showed mostly the same binding site except that Thr 74 was placed a bit further from the ligands but within 5 Å. Our results showed the binding pocket residues to be Lys5, Leu6, Val7, Glu57, Ser59, Arg40, Asp54, Ile55, Leu56, Gln70, Tyr71, Met72, Thr74, and Gly75. They further elaborated that the binding of the indole derivative expanded the pocket to accommodate the ligand. A recent report by Grant et al. (2011) also established this region as one of the allosteric binding pockets important in finding inhibitors for K-ras [25]. Hexadecanoic acid is a long chained hydrocarbon that folds into a U-shape when docking into the pocket. This folding makes the molecule snugly fit into the pocket. From this result we can assume that when the cancer cells were incubated with the T. flagelliforme extract [23], the hexadecanoic acid could possibly target the K-ras at this surface pocket.

The other ligands presented another interesting position. The pheophorbide epimers docked strongly into a depression on the surface of the K-ras that is also the surface for interaction with the SOS protein [35] The residues involved that formed the binding pocket for the epimers and 1-hexadecene were Arg73, Thr74, Gly75, Glu76, and Gly77. The strong binding affinity between the protein and the epimers was the result of the hydrogen bonding between the ligands and Arg73 and Gly75.

For another docking experiment we used two of the PDB structures, 4LUC and 4LYF, reported by Ostrem et al. (2013) since it presented two different ligands, a sulphonamide and a vinyl sulphonamide that caused changes in the switch II region to accommodate the ligand [28]. The researchers focused on finding inhibitors that could bind to mutated GDP bound K-ras and change its structure such that it would not be able to exchange the GDP molecule for the GTP to be activated.

The authors showed that their test ligands targeted the switch II region that falls in the loop region between the central β-sheet, a2 and a3 helices of the ras. The pocket had been earlier reported by Taveras et al. [36]. We used the same binding pocket lined with residues Val7, Val9, Gly10, Ala11, Thr58, Ala59, Gly60, Gln61, Glu62, Glu63, Arg68, Tyr71, and Met72. Our results revealed that the pheophorbide epimers had the strongest affinity. However, the docked poses presented an entirely different picture.

In case of the structure PDB ID: 4LYF, pheophorbide a could insert its side chain (–CH$_2$–CH$_2$–COOH) into the pocket (Figure 2). The same side chain of pheophorbide a’ did not penetrate much. Pheophorbide a occupied position lined by residues Phe90, Glu91, His94, His95, Gln129, Asp132, Leu133, and Ser136. Being bulky molecules they could not penetrate inside the pocket, however could make a hydrogen bond with Glu63, His95 and Tyr96 that contributed to the binding affinity of $-7.0$ for pheophorbide a, 6.8 for pheophorbide a’ and $-7.3 \text{ Kcal/mol}$ for pheophorbide a’. Hexadecanoic acid and hexadecene could easily slide into the pocket and make polar contact with Arg68. This was perhaps due to the long hydrophobic chain despite the low

![Figure 2:Docked poses of the flagelliforme constituents to the binding pocket of K-ras (PDB ID 4LUC). The ligands are hexadecanoic acid (red), pheophorbide a (yellow), phephorbide a’ (blue pyropheophorbide a, and magenta), and 1-hexadecene (green)).](image-url)
state (4LUC, 4LYF). The hexadecanoic acid and 1-hexadecene looked for hydrophobic pockets that it could slide into and their preferable hotspot was switch II.

4. Conclusion

Hence, we can conclude that *T. flagelliforme* constituents could target several allosteric sites on the K-ras. Since this protein is the most important signaling molecule in cancer cells inhibiting this protein would induce apoptosis of lung cancer cells. Combining our results with experimental evidence from Lai et al. (2008, 2010) where whole extracts are more potent than individual constituents, we can hypothesize that K-ras could have been the probable target of pheophorbides and other constituents. Pheophorbides bind to the SOS binding site on K-ras and could possibly prevent a strong interaction between the nucleotide exchange protein SOS and K-ras, while 1-hexadecane and hexadecanoic acid bind to switch II region of the K-ras. Both events combined would eventually inhibit the growth signals in the cancerous cells. Further studies are required to conclusively indicate K-ras at the target for the pheophorbides and other constituents.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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