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References

1. Tang YW, Li YL, Ye KL, Xu ZY, Ruo cal Academy Haydarpasa Training Hospi-

2. Khan AS, Ksizaeck TG, Peters CJ. Hantavirus pulmonary syndrome. Lancet. 1996;347:739–41. DOI: 10.1016/S0140-6736(96)90082-3

3. Klempa B, Tkachenko EA, Dzagurova TK, Yunicheva YV, Morozov VG, Okulova NM, et al. Hemorrhagic fever with renal syndrome caused by 2 lineages of Dobrava hantavirus, Russia. Emerg Infect Dis. 2008;14:617–25. DOI: 10.3201/eid1404.071310

4. Ertek M, Buzgan T; Re 

5. European Network for Diagnostics of Imported Viral Diseases (ENIVD) diagnosis criteria [cited 2010 Sep 22]. http://www. enivd.de/FS/FS_endcdisases.htm

6. Laakkonen J, Kallio-Kokko H, Oktem MA, Bladell K, Plyasmina A, Niemi maa J, et al. Serological survey for viral pathogens in Turkish rodents. J Wildl Dis. 2006;42:672–6.

7. Oktem MA. Hantavirus and tick-borne encephalitis infections [in Turkish]. Ankem Derg. 2009;23(Suppl 2):245–8.

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Maternal-Fetal Transmission of Cryptococcus gattii in Harbor Porpoise

To the Editor: We report maternal-fetal transmission of Cryptococcus gattii and death in a wild porpoise. Cryptococcus neoformans and C. gattii are 2 environmental, encapsulated yeasts that cause invasive, potentially life-threatening infections in humans and animals (1). C. neoformans causes disease in immunocompromised hosts, and C. gattii is also pathogenic in immunocompetent hosts (2). Since 1999, cryptococcosis caused by C. gattii has appeared on southern Vancouver Island (British Columbia, Canada) and nearby surrounding areas (2,3).

Spread beyond Vancouver Island has been documented along the Pacific Northwest Coast, but the mechanism remains undetermined (4).

A pregnant, dead, stranded, harbor porpoise (Phocoena phocoena) was reported on February 22, 2007, on western Whidbey Island, in Puget Sound, Washington State (48.2833°N, 122.7283°W). The carcass was iced and necropsy was performed on February 24. Sampled tissues from the adult and fetus were divided: half fixed in 10% formalin for histopathologic analysis, and half frozen for ancillary studies.

For histologic analysis, tissues were embedded in paraffin, sectioned to 3–5 μm, and stained with hematoxylin and eosin. Selected sections were stained with mucicarmin. The adult porpoise (length 177 cm, weight ≈57.7 kg) was in poor condition (reduced blubber layer). Both lungs were exposed, extensively scavenged, firm, and nodular; a sectioned surface exud ed clear to slightly opaque gelatinous to mucinuous discharge. Mediastinal lymph nodes were grossly enlarged, multinodular, and firm with large numbers of yeasts visible by microscopy (online Appendix Figure, panel A, www.cdc.gov/EID/content/17/2/302- appF.htm). The first stomach chamber contained two 3.5 cm × 2.5 cm raised, centrally umbilicated ulcers and several embedded anisakid nematodes. The uterus was gravid in the right horn with a mid-term fetus. No other gross lesions were identified. Microscopically, the lung lesions correlated with granulomatous to pyogranulomatous infiltrates, often with a myriad of yeasts.

The male fetus (length 30 cm, weight 2.4 kg), was examined separately at a different facility than the dam. It appeared grossly normal exter-nally and was at a gestation of 5–6 months. Mediastinal lymph nodes had mild granulomatous inflammation and contained numerous yeasts morphologically consistent with Cryptococcus spp. (online Appendix Figure, panel B). The lymph nodes were partially replaced with intracellular and extracellular multilobulated yeast aggregates (length 8–20 μm) with pale eosinophilic central regions and a thin refractile wall peripherally bound by a 5-μm nonstaining capsule. Around the periphery of these aggregates, there were small numbers of macrophages and lymphocytes and fewer neutrophils. Specific staining showed a prominent mucicarminophilic capsule consistent with Cryptococcus spp.

Yeasts were found in the amniotic fluid and interspersed within the choroidal villi and submucosal vasculature of the placenta. Mild multifocal nonsuppurative myocarditis was detected. However, no yeasts were seen in inflamed areas. There were no overt lesions in the remaining organs.

Maternal and fetal tissues were cultured for fungi, and diagnosis was based on Gram stain (budding yeast-like cells), India ink stain (positive for encapsulated cells), hydrolysis of urea (positive), and final confirmation by using API 20C Aux V3.0 (bioMérieux, Marcy l’Etoile, France). Canavanine-glycine-bromthymol blue agar was used to differentiate between C. gattii.
and *C. neoformans* (5). Molecular typing by restriction fragment length polymorphism was used to definitively speciate and subtype *C. gattii* (6). Fungal culture showed heavy growth of *Cryptococcus* spp. from the dam (lungs, mediastinal lymph nodes, and placenta) and fetus (mediastinal lymph nodes). Genotyping of primary isolates identified VGIIa *C. gattii* in both animals. Test results for enteric pathogens, intestinal nematodes, morbillivirus, and *Brucella* spp. were negative.

Fetal infection was most likely hematogenous, disseminated from a primary maternal pulmonary source to the uterus and subsequently to placental vasculature and internal fetal tissues. Infection by aspiration or ingestion of contaminated amniotic fluid was also possible. Although close evaluation of the lung did not show any discernible yeasts, the organism may have been present in an area other than that sectioned.

During 1998–2007, ≈450 harbor and Dall’s porpoises (*Phocoenoides dalli*) and Pacific white-sided dolphins (*Lagenorhynchus obliquidens*) along the Pacific Northwest Coast were recovered and subjected to necropsy. Disseminated cryptococcosis caused by *C. gattii* since 2000 was diagnosed in 15 harbor porpoises, 10 Dall’s porpoises, 2 adult Pacific white-sided dolphins, and 3 unrecorded species (10 females, 15 males, and 5 unknown sex; 24 adults, 4 juveniles, 1 fetus, and 1 undocumented age; S. Raverty, unpublished data).

Wild porpoises in the Pacific Northwest Region, being near shore inhabitants in waters surrounding Vancouver Island, may come into contact with air containing *C. gattii* at the air–water interface or ingest seawater containing yeasts while feeding (7). Their proximity to a habitat containing Coastal Douglas fir (*Pseudotsuga menziesii*) and Western hemlock (*Tsuga heterophylla*) may play a role in the epidemiology of *C. gattii* because these trees have been associated with cases of *C. gattii* (8). Cryptococcal infection during pregnancy has been reported in humans and horses (9,10).

This fetal case of cryptococcosis may have major human and animal health implications. Further studies should be undertaken to assess possible fetal involvement, identify infections in pregnant females, and provide information on risk reduction and improving diagnosis and treatment.

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References

1. Ellis DH. *Cryptococcus neoformans* var. *gattii* in Australia. J Clin Microbiol. 1987;25:430–1.
2. MacDougall L, Fyfe M. Emergence of *Cryptococcus gattii* in a novel environment provides clues to its incubation period. J Clin Microbiol. 2006;44:1851–2. DOI: 10.1128/JCM.44.5.1851-1852.2006
3. Stephen C, Lester S, Black W, Fyfe M, Raverty S. Multispecies outbreak of cryptococcosis on southern Vancouver Island, British Columbia. Can Vet J. 2002;43:792–4.
4. MacDougall L, Kidd SE, Galanis E, Mak S, Leslie MJ, Cieslak PR, et al. Spread of *Cryptococcus gattii* in British Columbia, Canada, and detection in the Pacific Northwest, USA. Emerg Infect Dis. 2007;13:42–50. DOI: 10.3201/eid1301.060827
5. Kwon-Chung KJ, Polacheck I, Bennett JE. Improved diagnostic medium for separation of *Cryptococcus neoformans* var. *neoformans* (serotypes A and D) and *Cryptococcus neoformans* var. *gattii* (serotypes B and C). J Clin Microbiol. 1982;15:535–7.
6. Kidd SE, Hagen F, Tscharke RL, Huynh M, Bartlett KH, Fyfe M, et al. A rare genotype of *Cryptococcus gattii* caused the cryptococcosis outbreak on Vancouver Island (British Columbia, Canada). Proc Natl Acad Sci U S A. 2004;101:17258–63. DOI: 10.1073/pnas.0402981101
7. Kidd SE, Chow Y, Mak S, Bach PJ, Chen H, Hingston AO, et al. Characterization of environmental sources of the human and animal pathogen *Cryptococcus gattii* in British Columbia, Canada, and the Pacific Northwest of the United States. Appl Environ Microbiol. 2007;73:1433–43. DOI: 10.1128/AEM.01330-06
8. Kidd SE, Bach PJ, Hingston AO, Mak S, Chow Y, MacDougall L, et al. *Cryptococcus gattii* dispersal mechanisms, British Columbia, Canada. Emerg Infect Dis. 2007;13:51–7. DOI: 10.3201/eid1301.060823
9. Blanchard PC, Filkins M. Cryptococcal pneumonia and abortion in an equine fetus. J Am Vet Med Assoc. 1992;201:1591–2.
10. Molnar-Nadasgy G, Haesky I, Reed J, Altshuler G. Placental cryptococcosis in a mother with systemic lupus erythematosus. Arch Pathol Lab Med. 1994;118:757–9.

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Letters

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