Applications of Bioadhesives: A Mini Review

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Bioadhesives have demonstrated their superiority in clinical applications as tissue adhesives, hemostats, and tissue sealants. Because of the intrinsic stickiness, the applications have been expanded to various areas, such as functional wound dressing, factor delivery vehicles, and even medical device fixation. While many literature works discussed the mechanism of bioadhesives, few of them specifically summarized the applications of bioadhesives. To fill in the blanks, this review covers recent research articles and focuses precisely on the applications of bioadhesives which can be generally classified as follows: 1) wound closure, 2) sealing leakage, and 3) immobilization, including those already in the clinic and those showing great potential in the clinic. It is expected that this article will provide a whole picture on bioadhesives’ applications and lead to innovations in the application of bioadhesives in new fields.

Keywords: bioadhesive, sealant, wound closure, functional wound dressing, medical device fixation

INTRODUCTION

Bioadhesives have been changing the surgical process with increasing importance and rapid development over the past 30 years (Ge and Chen, 2020; Taboada et al., 2020). The growing interest in producing adhesives and sealants makes them constitute a market share of $38 billion currently (Spotnitz and Burks, 2012; Qu et al., 2018; Liang et al., 2019). Compared with traditional invasive wound closure methods, including sutures, wires, and staples, bioadhesives have less possibility to damage the tissues and can promote wound healing through different mechanisms. For example, the bioadhesives possess antibacterial, anti-inflammatory, and antioxidant properties (Giano et al., 2014; Zhao et al., 2020). Other properties like self-healing and injectability significantly increase bioadhesives’ ease of use (Sun et al., 2020). Preventing leakage is also an essential role of bioadhesives. Leakage happens easily after the surgical process, which is up to 30% in some challenging situations. The leakage will easily lead to pain, inflammation, infection, and a high mortality rate (Artzi, 2013; Sliker et al., 2013; Pausch et al., 2020). With an aim to prevent those postoperative leakages, different bioadhesives have been developed accordingly. FocalSeal® was developed to avoid air leakage during lung surgery. DuraSeal® was designed for the spine and dura sealing. Coseal® was used as an adjunct of suture to prevent the leakage of blood vessels.

Moreover, they can remain stable on the site of application because of the intrinsic adhesion property. So, another important function of bioadhesives is immobilization. They can immobilize themselves as functional wound dressings to promote wound healing without other fixation methods (Yang et al., 2021). They can also be employed as vehicles to deliver functional items like drugs or cells to realize local delivery (Patel et al., 2014; Hu et al., 2021). With the development of smart biomedical devices, like wearable devices, implantable detectors, or sensors, a question has been raised about how to fix those devices on/in the body through noninvasive methods without damaging
Bioadhesives have been explored in human bodies in different organs, including the brain, eyes, heart, liver, and skin. Their applications can be grouped into three categories. (A) Wound closure, which has been used in topical skin, and brittle/hard tissues. (B) Sealing leakage, including most explored blood leakage and other fluids or gas leakage. (C) Immobilization for wound dressing, drug/cell delivery, and fixation of devices.

FIGURE 1 | The applications of bioadhesives in human bodies and their categories. Bioadhesives have been explored in human bodies in different organs, including the brain, eyes, heart, liver, and skin. Their applications can be grouped into three categories. (A) Wound closure, which has been used in topical skin, and brittle/hard tissues. (B) Sealing leakage, including most explored blood leakage and other fluids or gas leakage. (C) Immobilization for wound dressing, drug/cell delivery, and fixation of devices.

WOUND CLOSURE

Wound closure is one of the most widely used applications of bioadhesives (Table 1). Sutures, wires, and staples have been the routine practice of wound closure for many years (Mehdizadeh and Yang, 2013). However, concerns about the scar tissues, secondary injury, foreign body reaction, wicking-induced infection, impaired wound healing process, and complex postoperative care are still waiting to be addressed (Harsha and Vasudha, 2018). As a good alternative, bioadhesives can adhere two wounds together through a noninvasive behavior. Typically, bioadhesives close the wounds by three methods: bringing the two sides of an injury together from the wound surface (Figure 2A), bringing the tissues beneath the surface together (Figure 2B), or closing wounds in both ways (Figure 2C). Firm adhesion is the property needed for all the three types. Moreover, the bioadhesives applied to wounds (Figures 2A,C) should be biodegradable and biocompatible and should not hinder the healing process (Li et al., 2020). The bioadhesives used on the surface are generally tape-like ones with strong cohesion strength (Bae et al., 2013; Yang et al., 2013). Cohesion, which is defined as the internal strength of an adhesive, together with adhesion creates a strong bond; few people conducted in-depth research on cohesion strength alone. However, it is reported that the photo-crosslinking strategy is commonly used to develop tape-like bioadhesives with high cohesion strength. Besides, the double network strategy has also been used to develop bioadhesive tapes with good wound closure efficacy by increasing the cohesion strength (Liu et al., 2018; Yuk et al., 2019; Pausch et al., 2020).

Cytotoxic degradation products, and long degradation time (Bu et al., 2017; Harsha and Vasudha, 2018). So, special attention should be paid to avoid pushing cyanoacrylate-based bioadhesives into the wound, which can cause irritation and foreign body reaction. There are indications of holding wound edges together for at least 30 s before releasing. Because of the brittle property of the barriers formed by cyanoacrylate, it is suggested that cyanoacrylates are not suitable for wounds over joints, like the knees, groins, or hands, where adhesion easily fails because of the skin torsion (Harsha and Vasudha, 2018).

Skin Closure

Skin closure is one of the main goals for wound closure-targeting bioadhesives which is in high demand because of the increasing workload of general surgery (Lu et al., 2020). This application has expanded popularity also because people pay more attention to their physical appearance. Dermal surgeons prefer using bioadhesives to improve their work efficiency. Patients tend to use noninvasive methods because there is usually less pain and a better cosmetic outcome (Ge and Chen, 2020). Luo et al. developed a new bioadhesive from the skin secretion of Chinese giant salamander. Later, the ability to close the wound was tested on the back of the rats with four incisions (2 cm). At the 5th day of postoperation, the bioadhesive-treated group showed the best healing effect among all groups, with no scar formation, infection, and inflammation (Deng et al., 2019). Du et al. fabricated an adhesive patch with poly-(ethylene glycol) diacrylate/quaternized chitosan/tannic acid based on mussel-inspired chemistry. The efficacy of the wound closure was...
tested on a full-thickness incision model. It was proved that at day 7 postsurgery, the patch-closed skin incisions exhibited more complete epidermis and dermis structures, and higher collagen deposition levels than the untreated tissues (Du et al., 2019).

## Wound Closure of Hard or Brittle Tissues

Other kinds of wound closure, in which bioadhesives show super advantages, are closing wounds of hard and extremely brittle tissues. In hard tissues like bones, bioadhesives provide a quick and straightforward method to fix the broken pieces, especially for small bone fragments (Farrar, 2012). For example, comminuted bone fracture is a severe orthopedic condition. The difficulty in fixation of the small bone pieces often leads to bone reduction, further resulting in bone displacement, bone union deformation, and nonunion. Based on citrate, Xie et al. developed an injectable bioadhesive to fix small bone pieces in comminuted bone fractures (Xie et al., 2015). Hydroxyapatite was added to the system to improve the healing efficacy. It was demonstrated that the bioadhesive increased bone formation with markedly enhanced three-point bending strength compared with the negative control. In extremely brittle or sensitive tissues like nerves, traditional sutures can cause irreversible damage. Besides, skilled surgeons are required for suturing those tissues, which entails prolonged surgical time and surgical skills. In our previous work, the octa-PEG–based bioadhesives have been used to close the nerve transection. After adding lithium chloride, the adhesive-reconnected nerves showed a low level of fibrosis, inflammation, and myoatrophy, as well as robust axonal regeneration and functional recovery (Bu et al., 2020). Corneal is another brittle tissue in which closure can
be achieved by the bioadhesives. Shirzaei Sani et al. had engineered a gelatin-based adhesive biomaterial GelCORE to close the eye incision in an *ex vivo* model. It was found that the mean leak pressure of glue was more significant than that of commercial control groups (Shirzaei Sani et al., 2019).

**SEALING LEAKAGE**

Leakage is a common complication of surgeries and injuries. After lung resections, the incidence of air leakage was reported to be around 50% (Mueller and Marzluf, 2014). Cerebrospinal fluid leakage, caused by injuries or brain and sinus surgery, can lead to headaches, meningitis, and seizures. Gastric fluid leakage can cause severe tissue damage and infection, which happens easily after surgical procedures. So leakage prevention is vital in reducing operative risks, and decreasing the complications and the cost. Bioadhesives for leakage prevention are also called tissue sealants, which attracted the attention of researchers and have shown great potential in the clinic (Ryu et al., 2011; Nie et al., 2013; Behrens et al., 2014; Chan Choi et al., 2014; Kim et al., 2015; Chen et al., 2017; Yan et al., 2018; Kim et al., 2020). Some examples of the tissue sealants are summarized in *table 2*.

**Bleeding**

In this review, bleeding is considered as the leakage of the blood, resulting from trauma, surgical process, diseases, and even some medicines. It is one of the most frequent complications in patients. There are many sealants available in the market for hemostasis, such as Tisseel® (Fibrin sealant), Coseal® (PEG sealant), and Bioglue® (Albumin and Glutaraldehyde). However, they have separate limits. In their indications, Tisseel® is not suggested for massive bleeding; Coseal® and Bioglue® are suggested to be used as adjunctions to sutures or staples. So, sealants with high efficacy are still highly desired for uncontrollable or massive bleeding. Different strategies have been used to develop bioadhesives for hemostasis. Cui et al. developed a hyperbranched polymer sealant with a hydrophobic backbone and hydrophilic adhesive catechol side. By introducing long alkylamine chain into the structure, their sealant showed efficient hemostasis in the rat’s femoral artery bleeding and liver bleeding model (Cui et al., 2019). In our previous work, the concept of fabricating sealants with strong cohesion strength has been used (Bu et al., 2016; Bu et al., 2019). Tough sealants based on ammonolysis-based Tetra-PEG hydrogels were fabricated, which showed promising efficacy in pigskin massive bleeding and rabbit femoral artery section models (Bu et al., 2016). Hemostasis is another critical situation for patients with coagulation disorders, such as hemophilia, Von Willebrand disease, and aged patients taking anticoagulation drugs. Shin et al. presented a hemostatic hypodermic needle that will be able to prevent bleeding following tissue puncture. The surface of the needle was coated with catechol-functionalized chitosan that would be transformed from the solid to the gel phase *in situ* to seal punctured tissues (Shin et al., 2016). Later, Kim et al. used the catechol-conjugated chitosan to fabricate a hemostatic sponge (Kim et al., 2021). They used preclinical models to evaluate the hemostatic efficacy, including the heparinized rabbit model of femoral artery bleeding, the pig model of traumatic blunt liver injury with hemodilutional and hypothermic coagulopathy, and the anticoagulant-treated rabbit model of liver resection bleeding. A further clinical study performed on 15 patients showed that this sponge demonstrated an excellent hemostatic effect compared with the commercialized controls.
TABLE 2 | Bioadhesives for sealing leakage.

| Materials used                                                                 | Type of the model                                                                 | Animal species              | References                                                                 |
|--------------------------------------------------------------------------------|----------------------------------------------------------------------------------|-----------------------------|---------------------------------------------------------------------------|
| Hemostasis                                                                     | The rat femoral artery was punctured with a 1-ml syringe needle                   | Male SD rats                | Cui et al. (2019)                                                         |
| Multi-vinyl monomers, dopamine                                                  | One-quarter of the liver lobe was sheared off with a 20-G needle                 | Male SD rats; C57BL/6 mice; Kunming mice | Cui et al. (2019), Liu et al. 2012, Zhu et al. 2017, and Qu et al. 2018 |
| Multi-vinyl monomers, dopamine                                                  | Liver bleeding was triggered by puncture with an 18-G needle                     | Normal ICR mice; SD rats; Female balb-c mouse | Ryu et al. 2011, Nie et al. 2013, Behrens et al. 2014, Chan Choi et al. 2014, Kim et al. 2015, Chen et al. 2017, Yan et al. 2018, and Kim et al. 2020 |
| Poly (ethylene glycol), tyramine, chitosan; bovine serum albumin (BSA), Citrate acid, dopamine; chitosan, Pluronic®F127 (PF127-CHO) |                                                                                   |                                                                           |                                                                           |
| Tannic acid, poly (ethylene glycol); chitosan/pluronic composite hydrogel; chitosan/poly-lysine hydrogel; poly (γ−glutamic acid), dopamine hydrochloride (DA); N-[3-aminopropyl] methacrylamide (APM); DOPE-modified gelatin; hydrazide-modified poly (L-glutamic acid) (PLGA−ADH), dual-functionalized alginate; epigallocatechin gallate (EGCG); tyramine, hyaluronic acids, tyrosinase |                                                                                   |                                                                           |                                                                           |
| DNA from salmon testes, tannic acid                                            |                                                                                   |                                                                           |                                                                           |
| Glycol chitosan (GC), 3-[4-(hydroxyphenyl) propionic acid                       |Liver bleeding was triggered by puncture with a 23-G needle                       | ICR mice                    | Shin et al. (2015)                                                        |
| Tetra-armed poly (ethylene glycol) amine, tetra-armed poly (ethylene glycol) succinimidyl succinate | Liver bleeding was triggered by puncture with a 28-G needle | Male BALB/c mice            | Lu et al. (2018)                                                          |
| Tetra-armed poly (ethylene glycol) amine, tetra-armed poly (ethylene glycol) succinimidyl succinate | An incision with a length to be 20 mm and a depth of 5 mm was made on the left lobe of the liver | New Zealand white rabbits | Bu et al. (2019)                                                          |
| TachoSil (fibrogen-in-impregnated sealant), TissuFleece and Tissucol Duo (fibrin glue) | A wound with a diameter of 25 mm and a depth of 10 mm was made on the spleen | Bama miniature pigs         | Bu et al. (2019)                                                          |
| Chitosan                                                                       | A standardized left hemihepatectomy was performed by resecting the left and medial segment of the liver | Landrace pigs                | Fonouni et al. (2017)                                                     |
| Chitosan                                                                       | The femoral vein was transected                                                  | Male Long-Evans rats        | Dowling et al. (2011)                                                     |
| N-[3-aminopropyl] methacrylamide hydrochloride (APM)                            | The femoral artery was transected                                               | Yorkshire crossbred swine, SD rats | Dowling et al. (2011)                                                     |
| Gelatin (Type A); methacrylic anhydride, polyethylene glycol diacrylate (PEGDA-Mn 700) | Tail amputation at 50% tail length was completed using surgical scissors         | Male Wistar rats            | Krishnadoss et al. (2019)                                                 |
| N-[3-aminopropyl] methacrylamide hydrochloride (APM)                            | Rat tails were marked 4 cm from the tip and transected with a scalpel           | Adult Dorsett hybrid sheep  | Behrens et al. (2014)                                                     |
| Carboxymethyl chitosan (CMC), gelatin, oxidized alginate (OSA)                 | An incision of 5 cm in length and 1 cm in depth was made with a surgical scalpel on the right lobe of the sheep’s liver | Normal SD rats              | Cao et al. (2019)                                                         |
| Methacrylated gelatin (GelMA), N-[2-aminoethyl]-4-(4-(hydroxyethyl)-2-methoxy-5-nitrosophenoxy) butanamide (NB), glycosaminoglycan hyaluronic acid, lithium phenyl-2,4,6-trimethylbenzoxophosphinate | A wound about 1 cm in length and 2 mm in depth was made in one lobe of the liver | Male Bama Miniature pigs       | Hong et al. (2019)                                                        |
| Methacrylated gelatin (GelMA), N-[2-aminoethyl]-4-(4-(hydroxyethyl)-2-methoxy-5-nitrosophenoxy) butanamide (NB), glycosaminoglycan hyaluronic acid, lithium phenyl-2,4,6-trimethylbenzoxophosphinate | A 6-mm inner diameter needle was used to pierce the ventriculus sinister of pig hearts; an incision (4–6 mm) was created by needle puncture in the femoral artery | Male New Zealand white rabbits | Hong et al. (2019)                                                        |
| 4-Arm poly (ethylene glycol), 4-Arm poly (ethylene glycol) succinimidyl, 4-Arm poly (ethylene glycol) amine, vancomycin | An incision of 1 cm in length and 0.5 cm in depth was made in the liver | New Zealand white rabbits | Bu et al. (2016)                                                          |
| 4-Arm poly (ethylene glycol), 4-Arm poly (ethylene glycol) succinimidyl, 4-Arm poly (ethylene glycol) amine, vancomycin | Femoral artery transection                                                      | New Zealand white rabbits | Bu et al. (2016)                                                          |
| Chitosan hydrochloride (ChitHCl), dextran dialdehyde (DDA)                     | Liver lobe edge resection of approximately 1.5 cm length at two sites; liver lobe circular excision of approximately 1 cm diameter at one site | New Zealand white rabbits | Balakrishnan et al. (2017)                                               |

(Continued on following page)
TABLE 2 | (Continued) Bioadhesives for sealing leakage.

| Materials used                                      | Type of the model                                      | Animal species | References                      |
|-----------------------------------------------------|--------------------------------------------------------|----------------|---------------------------------|
| Dextran sodium periodate                            | An incision of ~1 cm in length and ~0.2 cm in depth was fabricated with a surgical scalpel on the ear-vein of the rabbit; the uncontrolled hemorrhage model was created by cutting a wound on the rabbit’s femoral artery by using ophthalmic scissors | Male New Zealand white rabbits | Liu et al. (2019)               |
| 4-Arm-poly (ethylene glycol) succinimidyl, Lysozyme  | The iatrogenic injury of the blood vessel was created by a 0.5 x 20-mm medical needle | Rabbits        | Tan et al. (2019)               |
| Glycerol, sebacic acid                              | The laceration was made on a porcine lung lobe with a razor blade (3 cm in length); the air was applied through the tubing connected to the upper part of the trachea (25-mmHg pressure) to visualize air leakage with or without bioadhesive | Yorkshire pigs | Lang et al. (2014)              |
| Other leakage prevention Bovine serum albumin (BSA), citrate acid, dopamine; gelatin, dopamine, genipin | Carotid artery defects model | Female SD rats | Zhu et al., 2017 and Yanagihara et al., 2021 |
| Gelatin type A, methacrylic anhydride (MA), tannic acid (TA) | An incision (~1 cm) was made on the mouse’s stomach | C57BL/6J mice | Liu et al. (2018)               |
| Polydextran aldehyde (PDA), branched polyethyleneimine (PEI) | Celiac ligation and puncture model | C57BL/6 mice | Giano et al. (2014)             |
| Poly (lactic-co-glycolic acid) (PLGA), poly (ethylene glycol) | Celiac intestinal anastomosis survival model | C57BL/6J mice | Behrens et al. (2015)           |
| Methacryloyl-substituted tropoelastin (MeTro)        | Standard incision (15 mm x 15 mm x 1 mm) was generated on the lung with a scalpel | Yorkshire pigs | Annabi et al. (2017)            |
| Gelatin                                             | Pleural defects in ex vivo and in vivo porcine models | Pigs           | Elvin et al. (2010)             |
| Methacrylated gelatin (GeIMA)                       | A standardized lung lobe incision (3 mm in length; 5 mm in depth toward the hilum) was generated | Male Wistar rats | Assmann et al. (2017)           |
| Methacrylated gelatin (GeIMA)                       | Standardized visceral pleural defect (15 mm in length; 15 mm in width; 1 mm in depth) was generated | Pigs           | Assmann et al. (2017)           |
| Gelatin, dopamine-conjugate gelatin (GeDA)          | A small (3 mm) incision was created in the murine small bowel; a surgical incision (2–4 mm) was made in one of the uterine horns | C57BL6 mice   | Hong et al. (2016)              |
| Polyvinyl alcohol (PVA), poly (acrylic acid) (PAA), N-hydroxysuccinimide (NHS) ester, sodium bicarbonate (SBC), glutathione | A laceration was made on a porcine lung lobe with a razor blade (3 cm in length); the air was applied through the tubing connected to the upper part of the trachea (25-mmHg pressure) to visualize air leakage with or without bioadhesive | Pig            | Chen et al. (2020)              |

Other Leakages
Except for blood leakage, there are also some other leakage types. In lung surgery, prolonged air leakage is the most common complication after surgical dissection and resection. The criteria of an ideal sealant for lung leakage include the following: 1. The sealant can stand higher burst pressure than that generated during physiological breathing; 2. the sealant should be elastic with a proper elastic modulus to support the inflation and deflation of lung tissue. Annabi et al. used methacryloyl-substituted tropoelastin (MeTro) to engineer a highly flexible sealant (Assmann et al., 2017). After applying MeTro to a porcine model, it was found that the sealant completely sealed the severely leaking lung tissue in the absence of sutures or staples. Urinary fistulas have been considered a severe socioeconomic problem, which occurs most commonly as a result of prolonged obstructed labor, which causes pelvic floor ischemia and, at times, substantial tissue loss (Margules and Rovner, 2019). Kim et al. developed water-immiscible mussel protein–based bioadhesive, which successfully sealed ex vivo urinary fistulas and provided good durability and high compliance (Kim et al., 2015). Liu et al. developed gelatin methacrylate–based double-network hydrogel to manage the leakage of gastric contents without sutures successfully (Liu et al., 2018).

IMMOBILIZATION
The last category for bioadhesives includes those for immobilization (Table 3). Because of the intrinsic adhesion property, they can immobilize themselves as functional wound dressing or delivery vehicles. By adhering items together, they even can fix other medical devices.

Functional Wound Dressings
Advanced fixation methods are still in need because traditional wound dressing methods lack the ability of adhesion to wounds, which increases the operative difficulty index for both the patients and doctors. Compared with these methods, bioadhesives can easily manage the wound area, contributing to the increasing popularity of bioadhesives to be used as a functional wound dressing (Liang et al., 2019; Zhao et al., 2017; Blacklow et al., 2019; Han et al., 2019).
| Materials used | Type of the model | Animal species | References |
|----------------|-------------------|----------------|------------|
| Functional wound dressings—skin defects | | | |
| Quaternized chitosan (QCS), benzaldehyde-terminated Pluronic®F127 (PF127-CHO) | About 1 cm diameter full-thickness round skin wounds were created by a needle biopsy | Female Kunming mice | Qu et al. (2018) |
| Quaternized chitosan-g-polyaniline (QCSP), benzaldehyde group functionalized poly (ethylene glycol)-co-poly (glycerol sebacate) (PEGS FA) | 7 mm diameter full-thickness round skin wounds were created by a needle biopsy | Female Kunming mice | Lian et al. (2019) |
| Hyaluronic acid-graft-dopamine (HA-DA), Reduced graphene oxide (rGO), polydopamine | 7 mm diameter full-thickness round skin wounds were created by a needle biopsy | Female Kunming mice | Han et al. (2017) |
| Poly (N-isopropyl acrylamide) (PNIPAm), alginate, chitosan | A full-thickness dorsal excisional skin wound was created on the mice with a sterile 8-mm-diameter biopsy punch following the removal of hair | Female Kunming mice | Han et al. (2017) |
| Skin secretion of Andrias davidianus (SSAD) | A disposable biopsy punch was used to create a full-thickness round skin wound (diameter = 10 mm) on the back | Streptozotocin-induced diabetic SD rat | Deng et al. (2019) |
| Polydopamine-clay-polyacrylamide (PDA-clay-PAM) hydrogel | Full-thickness skin wounds were created on the dorsal area of the rats | Male SD rats | Han et al. (2017) |
| Polydopamine-polyacrylamide (PDA–PAM) hydrogel | Four full-thickness circular wound (5 mm in diameter) were created on the back of each mouse by a disposable 5 mm skin biopsy punch | Male SD rats | Han et al. (2017) |
| Ag-Lignin NPs-PAA–pectin hydrogel | Four full-thickness circular wounds (8 mm in diameter) were created on the upper back of the rats | Male SD rats | Han et al. (2017) |
| Functional wound dressings—corneal defects | Gelatin, methacrylic anhydride (MA) | A 3-mm biopsy punch was used to make a partial trephination (cut) in the central cornea of the right eye to a depth of approximately 50% | Male New Zealand white rabbits | Shirzaei Sani et al. (2019) |
| Functional wound dressings—cartilage defects | Polydopamine-chondroitin sulfate-polyacrylamide (PDA-CS-PAM) hydrogel | A full-thickness defect (diameter: 3.5 mm; thickness: 5 mm) was created through the articular cartilage and subchondral bone of the patellar groove in the right leg of the rabbits using an electric drill | Japanese white rabbits | Han et al. (2018) |
| Functional wound dressings—calvarial defects | Acrylate b-cyclodextrin (Ac-b-CD), methacrylated gelatin (MeGel) | Two 5-mm-diameter craniotomy defects were created in the parietal bones of the skull on each side of the sagittal suture line | Male SD rats | Feng et al. (2016) |
| Functional wound dressings—myocardial infarction (MI) | Gelatin methacryloyl (GelMA), choline-based bio-ionic liquid (Bio-IL) | Immediately after the induction of MI, the scaffolds were delivered to the surface of the left ventricle, distal to the site of MI, and photo-crosslinked for 3000 s using UV light | Balb/C mice | Walker et al. (2019) |
| Waxy starch | The materials were patched onto the MI site of the heart. | SD rats | Lin et al. (2019) |
| Drug/Cell delivery | Methacrylated alginate (Alg-DA-MA), Gingival mesenchymal stem cells (GMSCs), HAp microparticles (MPs) | Ex vivo–expanded human GMSC aggregates/HAp MPs (4 × 106) were encapsulated in adhesive hydrogel and implanted subcutaneously (0.50 ml) into the dorsal surface of a 5-month-old Beige nude XID III (nu/nu) (Harlan, United States) mice; titanium implants (ACE Surgical Supply, Brockton, MA) were used to introduce a well-characterized strain of A. actinomycetemcomitans biofilm transmucosally into rats | Beige nude XID III (nu/nu) (Harlan, United States) mice; Male and female SD rats | Hasani-Sadrabadi et al. (2020) |
| Poly (ethylene glycol), catechol | Islet transplantation: approximately 100 ml cPEG was applied following islet deposition directly on this tissue surface | Streptozotocin-induced diabetic mice | Brubaker et al. (2010) |
| HA-catechol (HA-CA) hydrogel | Hepatocyte transplantation: hepatocytes encapsulated in HA-CA hydrogel were transplanted onto the lobe of the native liver or liver with partial hepatectomy of athymic mice using a pipette; HA-CA hydrogel was painted onto the infarction site immediately after induction of hydrogel crosslinking | Female athymic mice (Balb/cnu); Male Hsd RH-mu rats with myocardial infarction | Shin et al. (2015) |

(Continued on following page)
Compared with the traditional hydrogel delivery system, the bioadhesive delivery systems have also been developed to load drugs to achieve better efficacy (Zhang et al., 2019; Bu et al., 2020). Cells can delivered items on the site. Mucoadhesion is very useful in increasing the bioavailability of poorly absorbed drugs by prolonging the residence time in the gastrointestinal tract, leading to reduced dose and dosing frequency (Han et al., 2012; Gong et al., 2017). A lot of mucoadhesive-based delivery systems were developed with some well-summarized reviews (Reddy et al., 2011; Zhang et al., 2016; Zhang et al., 2020; Pathak and Malviya, 2020). Hu et al. encapsulated camptothecin into poly(lactic acid)-hyperbranched polyglycerol-based nano-bioadhesive particles (NPs). Because of the strong bonding of these NPs to squamous cell carcinoma tumor cells, the system significantly reduced the tumor burden and enhanced survival (Hu et al., 2021). Except for the nano/micro scale mucoadhesion, macro-bioadhesives have also been developed to load drugs to achieve better healing efficacy (Zhang et al., 2019; Bu et al., 2020). Cells can also be loaded into the bioadhesives. The use of an appropriate scaffold biomaterial as a cell delivery vehicle can provide a suitable microenvironment to prolong cell viability and present essential factors to direct cell differentiation toward the desired lineages (Khademhosseini and Langer, 2016). Currently, however, a major drawback of the reported cell-laden hydrogels is the weak adhesion to the host tissue at the defective site. Hasani-Sadrabadi et al. used alginate-based photo-crosslinkable bioadhesives to load mesenchymal stem cells. It was found that the cell-loaded adhesive system leads to complete bone regeneration around the ailing dental implants with peri-implant bone loss (Hasani-Sadrabadi et al., 2020).

**TABLE 3** (Continued) Bioadhesives for immobilization.

| Materials used | Type of the model | Animal species | References |
|----------------|-------------------|----------------|------------|
| Tetra-PEG/agar hydrogel (PA) | The drug containing hydrogel was formed in situ on the surface of the rats’ skin | SD rats | Zhang et al. (2019) |
| Tannic acid, poly (ethylene glycol) | Each mouse was fed on 0.02 cc of the ICG-encapsulated TAPE-ÖH for the adhesion to the esophagus without any anesthesia | BALBc nude mice | [Kim et al., 2015; Shin et al., 2018] |
| Wheat germ agglutinin (WGA)-conjugated liposomes (WGA-liposomes) | The OKF6/TERT-2 cell suspension (1 x 104 cells) was seeded onto poly-d-lysine coated glass bottom micro-well dishes (MatTek Corporation) and allowed to grow in cell culture media for 24 h. WGA-conjugated CFPE liposomes (WGA-CFPE-liposomes) were added to the micro-well dishes (45 μg/ml lipid) and incubated at 37° C for 2 h | OKF6/TERT-2 cell | Wijetunge et al. (2018) |
| GO (graphene oxide) hybrid supramolecular hydrogels (GO-HSH) | DCX-loaded GO–HSH hydrogel coating on titanium substrate and drug release to kill Hela PDVC57 cells were harvested, washed, and resuspended, then injected into the dorsal right flank. Tumors were injected with BNP-CPT and visualized particle distribution via confocal microscope 72 h after injection | Hela | Chen et al. (2018) |
| Poly (lactic acid)-hyperbranched polyglycerol (PLA-HPG), camptothecin (CPT) | The heart was exposed via a thorotomy, bioadhesive electrodes were used to record epicardial ECG. | Female Sprague–Dawley rats. | Deng et al. (2021) |
| Medical device fixation | The heart was exposed via a thorotomy, bioadhesive electrodes were used to record epicardial ECG. | Ex vivo porcine heart | |
| GO (graphene oxide)-PVA (poly (vinyl alcohol)) hydrogel, GO (graphene oxide)-PVA (poly (vinyl alcohol)) PAA (poly (acrylic acid)) -NHS (N-hydroxysuccinimide) ester hydrogel | A circuit with light emitting diodes (LEDs) was applied to the ex vivo porcine heart (by introducing cyclical, pressurized air inputs into the heart chambers to mimic heartbeats) to test if electrical communication was stable enough. | | |

2017; Han et al., 2017; Gan et al., 2019). They are favorite candidates for skin damage, one of the most common physical injuries in human history. Based on quaternized chitosan (QCS) and benzaldehyde-terminated Pluronic®F127, Qu et al. developed antibacterial bioadhesives with rapid self-healing, extensibility, and compressibility for joints and skin wound healing (Qu et al., 2018). They loaded curcumin into the bioadhesive and found that it significantly accelerated wound healing with a higher granulation tissue thickness in a full-thickness skin defect model. Inspired by embryonic wound closure, Blacklow et al. fabricated mechanically active dressings to accelerate wound healing (Blacklow et al., 2019). The bioadhesive dressing will contract at body temperature, which further applies force to draw the wound edges together in a purse-string–like manner. Adhesive dressings are beneficial in places where the fixation is difficult, like brittle tissues. Lin et al. developed a viscoelastic adhesive patch that accommodates the cyclic deformation of the myocardium. It was found that the patch outperformed most existing acellular epicardial patches in reversing left ventricular remodeling and restoring heart function after both acute and subacute myocardial infarctions in rats (Lin et al., 2019). In addition to the heart, defects from the corneal, cartilage, and calvarial were explored to be treated with bioadhesives with good outcomes (Feng et al., 2016; Han et al., 2018; Lin et al., 2019; Shirzaei Sani et al., 2019).

**Delivery Systems**

Compared with the traditional hydrogel delivery system, the advantage of bioadhesives in delivery is that they can fix
Fixation of Other Medical Devices

Nowadays, a growing interest is centered on implantable and wearable medical devices with excellent translational potential in the clinic, like tissue scaffolds, biosensors, and bionanobots. However, it is crucial to establish conformal and stable contact between those devices and the target tissue (Schiavone and Lacour, 2019; Yuk et al., 2019). Wires and sutures are required for this fixation, which raises concerns of infection, secondary tissue injury, and scaffold damage. As a noninvasive adhesion method, bioadhesives have the potential to replace these invasive fixation methods. Based on a thin layer of a graphene nanocomposite, Deng et al. developed an electrical bioadhesive that can provide rapid, robust, and on-demand detachable integration of bioelectronic devices on diverse wet dynamic tissues (Deng et al., 2021). Later, they successfully used the e-bioadhesive to record an in situ epicardial electrocardiogram and electrically stimulated a sciatic nerve on a rat model. This technique offers a promising solution for addressing the long-standing challenges in tissue–device integration.

Another good aspect of bioadhesives to be used in these situations is that different functions can be added into the bioadhesives to improve the outcome of the medical devices or reduce the potential complications. For example, the antibacterial property can be introduced to reduce the chance of medical devices’ infection (Hwang et al., 2018). In fact, there is still a vast area of bioadhesives in medical device fixation waiting to be explored. However, one should be careful because the bioadhesives may also adversely influence the medical devices. Macnab et al. showed that Tisseel® significantly attenuated NIR light of a near-infrared spectroscopy during in vitro transmittance and critically compromised photo transmission in vivo (Macnab et al., 2018). Another fixation method is also required when there is a need for tissue transplantation. Islet transplantation is used to treat type I diabetes by replacing the lost beta cell function. Brubaker et al. directly immobilized islets onto intra-abdominal tissue surfaces using a thin layer of a mussel-inspired bioadhesive (Brubaker et al., 2010). On the one hand, the fixation approach offers the potential advantages for convenient, rapid, and minimally invasive islet transplantation by direct apposition of the islet bolus onto tissue surfaces. On the other hand, the technique avoids the intravascular engraftment site, eliminating adverse effects of first-pass blood exposure in the liver while maintaining the capability of rapid islet revascularization and the benefits of direct insulin secretion into the portal circulation.

PERSPECTIVE

Bioadhesives are believed to revolutionize the surgical process (Mehdizadeh and Yang, 2013; Taboada et al., 2020). They have already been widely used as adhesives and sealants in the clinic to reduce complications and improve outcomes. However, those commercialized products are still far from satisfactory. Fibrin-based, PEG-based, and cyanoacrylate-based bioadhesives are the most commonly used ones. Fibrin-based and PEG-based bioadhesives have good biocompatibility but weak adhesion strength. So, most of them are only used as adjunctions for traditional wound closure or sealing methods. Cyanoacrylate-based bioadhesives have strong adhesion strength, but their potential safety concerns limit their wide applications, especially internal applications. Thus, more powerful and commercially transformable bioadhesives for wound closure and sealing leakage are still needed.

Compared with traditional wound dressings, bioadhesives get easily attached to the parts where they are applied because of their intrinsic adhesion property (Li and Mooney., 2016). So, there is a growing interest in using bioadhesives as a functional wound dressing. This application is beneficial for tissues where the fixation of traditional wound dressing fails to work, like a beating heart and brittle brain (Lin et al., 2019). However, the absence of removability makes it hard for further wound care or dressing change, resulting in more potential troubles when mechanical debridement is involved. So controllably removable property is also explored for bioadhesives (Chen et al., 2020; Bu et al., 2019; Villa-Camacho et al., 2015; Koniecynska et al., 2016).

Using bioadhesives for the local delivery of functional items like drugs or cells is also a promising way to realize specialized and prolonged effectiveness. Compared with conventional hydrogel vehicles, bioadhesives can adhere to tissues, making them more stable in special tissues like the beating heart and esophagus (Lin et al., 2019). By mixing Tannic and PEG, Lee et al. developed a new medical glue called TAPE, which had been applied to the esophagus and demonstrated the ability to detect gastroesophageal reflux diseases because it maintained wet-adhesive properties (Kim et al., 2015). Bioadhesives are also used to fix medical devices or tissues, of which the importance is increasing with an increasing number of implantable medical devices and tissue transplantation. The fixation using bioadhesives will not damage either medical devices or the tissues. Although very promising, there is a difficulty in avoiding the interference between the functions of medical devices and bioadhesives. Besides, for tissue transplantation, the adhesion strength of bioadhesives available might not be sufficient for large pieces of tissues.

Although massive efforts have been spent on developing bioadhesives, there are only a handful of products available in the market (Taboada et al., 2020). First, the researcher might care too much about the adhesion mechanism, while cohesion is ignored. Cohesion dramatically influences how the bioadhesives would be used, which is particularly important for clinical translation. In the market, ease of use has a positive influence on people’s choices. Second, one bioadhesive never fits all the applications. The requirement of bioadhesives for wound closure differs from those for sealing leakage. So, it is suggested to choose the unmet clinical target first and then the relative characterization methods to fabricate bioadhesives for translation.

AUTHOR CONTRIBUTIONS

WD did the literature search and paper writing. XB did the literature research and helped revise the paper. YB was responsible for the whole paper design and manuscript organization.

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