Horizontal Gene Transfer and Endophytes: An Implication for the Acquisition of Novel Traits

Pragya Tiwari and Hanhong Bae *

Department of Biotechnology, Yeungnam University, Gyeongsan, Gyeongbuk 38541, Korea; pragyamital02@gmail.com
* Correspondence: hanhongbae@ynu.ac.kr; Tel.: +8183-810-3031

Received: 14 January 2020; Accepted: 19 February 2020; Published: 1 March 2020

Abstract: Horizontal gene transfer (HGT), an important evolutionary mechanism observed in prokaryotes, is the transmission of genetic material across phylogenetically distant species. In recent years, the availability of complete genomes has facilitated the comprehensive analysis of HGT and highlighted its emerging role in the adaptation and evolution of eukaryotes. Endophytes represent an ecologically favored association, which highlights its beneficial attributes to the environment, in agriculture and in healthcare. The HGT phenomenon in endophytes, which features an important biological mechanism for their evolutionary adaptation within the host plant and simultaneously confers “novel traits” to the associated microbes, is not yet completely understood. With a focus on the emerging implications of HGT events in the evolution of biological species, the present review discusses the occurrence of HGT in endophytes and its socio-economic importance in the current perspective. To our knowledge, this review is the first report that provides a comprehensive insight into the impact of HGT in the adaptation and evolution of endophytes.

Keywords: endophytes; environmental adaptation; evolution; horizontal gene transfer; novel traits; prokaryotic genomes; whole genome sequencing

1. Introduction

Horizontal gene transfer (HGT) refers to the transmission of genetic material across the genomes of biological organisms by processes other than fertilization. HGT is a universal phenomenon observed in bacterial, fungal, and eukaryotic genomes [1,2]. However, it occurs infrequently and only serves as an alternative process for the exchange of genetic material between distantly related species. HGT is relatively more common in prokaryotes than eukaryotes [3–5]; studies have shown that approximately 81% of genes have transferred through HGT, in 181 sequenced prokaryotic genomes [6].

In the past decade, the availability of eukaryotic genomes through high-throughput sequencing has promoted the research on the occurrence and mechanism of HGT in eukaryotes [7]. Additionally, information about whole prokaryotic genomes has made it convenient to study HGT between distantly related species [8], specifically in terms of organismal evolution and ecological adaptation for survival [9,10]. HGT has been observed previously between Alternaria and Fusarium fungi [11], nematodes and insects [12,13], humans and bacterial pathogens [14], plants and silkworms (Bombyx mori) [15], and plants and fungi [16]. Extensive investigations on the significance of the HGT phenomenon in prokaryotic evolution (e.g., archaea and bacteria) revealed a possible mechanism for acquiring novel traits during the evolutionary course [17]; however, in eukaryotes, this phenomenon was presumed to be uncommon [18,19]. Moreover, the transmission and integration of the transferred genes might provide several beneficial attributes, including prokaryotic adaptation during environmental changes [20,21], acquisition of new traits/functions [22], and evolutionary adaptation in eukaryotes [23,24].
Endophytes represent the ecologically favored association between plants and microbes, providing multiple advantages for the plant and the environment. Endophytic associations are gaining momentum socio-economically due to their potential applications in agriculture, industries, and healthcare. The HGT phenomenon in endophytes, which highlights an important biological mechanism for their evolutionary adaptation within the host plant and simultaneously confers “novel traits” to the associated microbes, remains less studied. Studies in endophytic bacteria revealed its role in toluene biodegradation and disease reduction in wheat (*Triticum durum*) and corn (*Zea mays*) [25]. Moreover, scientists hypothesized that genetic recombination between plants and endophytes might have led to the inclusion of metabolic pathway genes in the host plant [26]. Table 1 provides a summarized account of the HGT events reported in plant–endophyte associations. Although HGT has been observed across different species, the implications of HGT occurring from a plant to its associated endophytic microbe presents an important and alternative process for addressing ecological and social concerns through environment-friendly approaches. Since there is very little information available about HGT in endophytes, the authors aimed to highlight the significance of HGT in endophytes, through a current perspective. The evolutionary importance of HGT across kingdoms, specifically its mechanisms and potential roles in addressing socio-economic and ecological concerns, has been extensively discussed. To our knowledge, this systematic review is the first report that emphasizes the emerging trends in HGT research on endophytes and its significance in the present time.

Table 1. A summarized account of the HGT events reported in plant–endophyte associations.

| No. | Endophyte                          | HGT Phenomenon                  | Beneficial Role(s)                           | Ref.  |
|-----|-----------------------------------|---------------------------------|---------------------------------------------|-------|
| 1.  | *Pseudomonas putida* W619-TCE     | Bacteria to bacteria            | Trichloroethylene degradation              | [24]  |
| 2.  | *Burkholderia cepacia* strain FX2 | Bacteria to bacteria            | Biodegradation of toluene, disease reduction on wheat and corn plants | [25]  |
| 3.  | *Epichloe* (syn. *Neotyphodium*)  | Endophytic fungi to grasses     | Adaptation for angiosperms                 | [27]  |
| 4.  | *Agrobacterium tumefaciens*       | Plants to endophyte             | Phytoremediation of arsenic contaminated soil | [28]  |
| 5.  | Endophytic fungi                  | Plants to endophyte             | Production of secondary metabolites         | [29]  |
| 6.  | Endophytic fungi                  | Plant to endophyte              | Production of secondary metabolites         | [30]  |
| 7.  | *Burkholderia cepacia* L.S.2.4    | Bacteria to bacteria            | Phytoremediation of toluene and organic pollutants | [31]  |
| 8.  | *Rhizophagus irregularis*         | Bacteria and plants to fungi    | Evolution and symbiotic adaptation of the arbuscular mycorrhizal fungi | [32]  |

2. HGT in Nature: An Overview

The phenomenon of HGT differs from the vertical transmission of genetic material from parent to offspring [33,34]. Distinct patterns in HGT were observed through endosymbiosis and introgressive hybridization [35]. HGT was first discovered in 1928 by Griffith, who demonstrated that virulence factor was transferred between pneumococcal strains in mice. Since this discovery, the accessibility to “big data” provided by the sequencing of complete genomes has led to the identification of transferred genes in multiple taxonomic groups [36]. However, the importance of HGT in eukaryotic genome evolution [37–39], is an emerging research area in the present time and is extensively explored.

The discovery of HGT across diverse biological species suggests its strong evolutionary role and highlights its significance in species adaptation and survival. For example, fungi and ciliates acquired the genes for carbohydrate metabolism from a ruminant animal [40,41]. Studies have suggested the role of HGT in facilitating microbial adaptation to adverse conditions in the environment [42] and inside the host plant [43]. In the past few years, the information on HGT events in whole eukaryotic genomes was limited and genomic information at the taxonomic level was unavailable. However, studies utilizing whole genome sequences and high throughput technologies revealed that HGT events in eukaryotic genomes, particularly in the plant kingdom, have a crucial impact on plant evolution [44]. In particular, the comprehensive phylogenetic analyses of genomes in 6 plant species, together with
other prokaryotic and eukaryotic genomes, identified 1,689 genes that were similar to fungi, indicating
the exchange of genetic material between plants and fungi [44], consequently resulting in novel
traits arising in plant genomes. Moreover, studies reviewed the possibilities of HGT occurring from
plants to endophytes, to determine potential applications for the production of bioactive secondary
metabolites via the endophytic fungi [29]. Endophytic fungi can produce secondary metabolites,

hence, it was hypothesized that HGT between the host plant and associated fungi was responsible for
the production of bioactive secondary metabolites [30]. The study also discussed the possibilities of
different mechanisms on how an endophytic fungus acquired genetic traits from the host plant and
whether HGT was beneficial for the adaptation and survival of the organisms in association [29,30].

3. The Mechanisms of HGT

HGT differs from other mechanism of genetic transmission since it does not involve
parent–offspring inheritance, and therefore, might occur between distantly related species. Furthermore,
the transfer of genetic material is rapid, rendering it an important mechanism for microbial adaptation
to new environmental niches [45,46]. There are several mechanisms for HGT across different taxonomic
groups (with different frequencies) as determined by the genetic distance between two biological
organisms [47]. Several studies have shown HGT across biological species, specifically in archaea [48],
bacteria [2,45], and eukaryotes [49]. The HGT phenomenon has been widely studied in bacteria and
archaeal genomes; however, this is not the case in eukaryotes.

The HGT mechanisms in prokaryotes, including transformation, conjugation, and transduction,
have been well documented; however, HGT in eukaryotes might be more complex, particularly in
plants, and might involve vector-mediated or direct pathways [50]. The direct pathway might occur
through direct DNA exchange [18], while vector-mediated pathway requires the use of vectors such
as bacteria, fungi, virus, etc. [51,52]. Moreover, HGT between nuclear and plastid genomes have
been reported [53], i.e., between plant mitochondrial genomes [54] via bacteria-mediated [46,55] and
parasitic insect-mediated HGT [56]. Studies have also suggested the possibility of virus-mediated
HGT from plants to other genomes via pathogen, transgenic bacteria (e.g., *Agrobacterium tumefaciens*),
virus [57], fungi [58], and nematodes [59].

Studies have shown that adaptation of bacteria to host plant involves diversification and
evolutionary processes, while switching from parasitism to mutualism [60]. The bacterial transition
to intracellular lifestyle induce various ecological changes. The sequencing of bacterial genomes has
yielded significant insights about bacterial population dynamics and evolution, by highlighting gene
recombination, deletion and gene amplification events [60]. Moreover, it has been seen that bacteria
classified in different phylogenetic clades differ in host adaptation. The co-integration of endosymbiont
into host and adaptation to intracellular lifestyle render changes in bacterial genes, conferring the
function of mutualism to bacteria. The endophytic association further makes the bacterial genome
to co-evolve with the host genome, characterized by reduction in bacterial genome evolution [60].
Moreover, studies have shown that the HGT event occurring in symbiotic and pathogenic bacteria is
responsible for functional divergence in different phylogenetic clades [61,62]. The host adaptation by
endophytes confers distinct advantages to associated microbes and helps in its adaptation and survival.
The inclusion of new functions via HGT (gene duplication and functional divergence) facilitates greater
interaction with the respective host organism [63].

4. Significance of HGT in The Evolution of Biological Organisms

The HGT phenomenon, widely regarded as a mechanism favored by evolution, aids the acquisition
of novel traits by the associated species. Previous studies have reported the transfer of genetic
material between microorganisms [63,64] and many examples of gene transfer across species are also
well-documented [33]. Studies discussing the significance of HGT in the evolution of biological species
and its contradiction to phylogenetic relationships between organisms are deemed controversial.
Several studies have reported the transfer of genes via HGT between similar and different biological
species, between different domains, and across kingdoms [39]. The role of HGT in evolution has been well-reported in bacteria and archaea, in contrast to eukaryotic genomes.

4.1. HGT in The Evolution of Prokaryotes

The role of HGT in prokaryotic evolution depends on the amount of genes transferred, their integration in microbial genomes, and the phylogenetic relationship between different species [39]. Successful HGT is mediated by the transduction, conjugation, and transfection processes and depends on the stable integration of the transferred genetic material. The differences between the transferred genes might be observed due to the barriers limiting gene transfer and various selective forces acting on HGT. As an explanation, Jain et al. [65] hypothesized that the genes involved in maintaining cell functions are more likely to participate in HGT than those involved in DNA replication, translation, and transcription, which are responsible for transmitting genetic information [65]. However, other scientists suggested that functional categorization of genes as a prediction method for HGT is not absolute. Moreover, the genetic distance between biological species during the course of evolution is another factor influencing HGT; gene transfer is frequent in closely related species and less common in distantly related species [6,66]. Wagner and colleagues [67] studied 438 complete prokaryotic genomes and found 30 cases of gene transfer among distantly related clades, showing a lesser frequency of HGT occurrence in species with a larger evolutionary distance between them [68].

The occurrence of HGT in prokaryotic genomes challenged the construction of phylogenetic relationships between biological species. The existence of HGT events in microbial genomes make it difficult to establish a correct phylogenetic relationship between microbes; however, it was suggested that the main genes are not transferred and a phylogenetic tree might be constructed using this pattern [68,69]. Furthermore, the genes can be cloned in E. coli, indicating that HGT events were not limited by any barriers [70]. The HGT phenomenon, showing the transfer of genetic material between different organisms in one generation, displays a striking contrast to the neo-Darwinian concept of evolution. However, after the gene transfer by HGT, the natural selection process determines the gene selection for spread into other populations [71]. In the present context, the phenomenon of HGT, together with other mechanisms namely hybridization, gene duplication, and gene acquiring mechanisms might be regarded as an evolutionary process [2,72,73].

4.2. HGT in The Evolution of Eukaryotes

HGT has been well-documented and is considered to be a key evolutionary factor in prokaryotic genomes; however, its significance in eukaryotes remains undetermined [38,49]. The frequency of occurrence and role of HGT in eukaryotes are being increasingly discussed, in light of the evolution process. The availability of whole genome sequences revealed the occurrence of HGT in eukaryotes, with the transfer of bacterial genes from ruminant host to fungi and ciliates [40,41]. With the sequencing of whole genomes and increasing availability of “big data” on eukaryotes, the significance of HGT in the adaptation of eukaryotes is being widely recognized [33,38,74]. Phylogenomic studies on several fungal genomes have identified multiple HGT events, which is the largest among eukaryotic genomes [75]. The sequencing and analysis of 60 fungal genomes revealed the existence of approximately 700 genes of prokaryotic origin, transferred through HGT [75]. Several examples of HGT in fungal genomes have highlighted the benefits to the fungal host, including adaptation to extreme environments [76], clustering of secondary metabolic genes [77,78], and pathogenicity to the host plant [79], suggesting that HGT conferred beneficial traits to the fungi and assisted in the adaptation to new environments. These studies indicate that HGT events play an important role in eukaryotes and that investigating a large number of eukaryotic genomes would provide better insights about the significance of HGT in the adaptive divergence of eukaryotes [80]. In addition, 57 gene families were assumed to be transferred from prokaryotes or fungi to the genome of the moss, Physcomitrella patens via HGT [81]. These acquired genes afforded multiple beneficial functions, specifically the biosynthesis of hormones and plant defense, leading to plant adaptation from aquatic to terrestrial habitat [81].
5. Recent Approaches to Detect HGT in Genomes

The identification and annotation of complete genomes in biological organisms have shown the frequent occurrence of HGT events, resulting in a chimeric organism, with multiple DNA from different genomes. Considering the emerging importance of HGT events in the evolutionary process [82], understanding the HGT phenomenon is crucial to learn about novel functions, such as the emergence of antibiotic resistance [83] and prediction of gene functions [84]. Several approaches are available for the identification of HGT events in whole genomes, depending on the type of HGT process. These methods are enumerated below:

Studying gene distribution patterns: Gene transfer between different species results in the acquisition of new genes. Therefore, studying gene distribution patterns with uneven occurrence might lead to the identification of HGT events. However, uneven distribution patterns might also be caused by processes such as sequence divergence or gene loss [82]. Additionally, analysis of gene distribution patterns can be used to detect homologous gene recombination.

Comparison of phylogenetic trees: One possible method to determine the occurrence of HGT event is the analysis of phylogenetic trees of different genes, based on the assumption that HGT events might lead different genes to have different evolutionary trees. However, this method is not accurate since various factors, such as ortholog/paralog misidentification, occurrence of convergence, and incorrect alignment of gene sequences [85–87], might lead to incorrect conclusions. Although the phylogenetic methods do not always correctly predict the HGT events among closely related species, it is still a method of choice in analyzing the genomes of biological organisms. Moreover, this method is based on extensive information about the number of genes.

Studying unusual genome composition: A consistent uniformity is present in genome composition and phenotype and the presence of foreign genes due to HGT events can be detected by identifying genomic regions with unusual composition (e.g., codon usage) [88]. This method requires the complete genome sequence of one species for the estimation of HGT events. However, since non-uniform genome composition might also result due to mutation, natural selection, and HGT [89], as well as the presence of biological vectors (e.g., bacteriophages), this method is not reliable for predicting HGT events between species with similar genome composition and HGT events that occurred long ago [82].

Similarity search between genomes: A common method for the identification of HGT events in genomes is to search for maximum similarity between genes. HGT is very likely between genes of distantly related species. This is a common and fast method, however, its disadvantages include less accuracy and uncertainty regarding the number of best matches to search for, identification of orthologs, and uncertainty with multi-domain proteins [82,90]. For an effective and methodical study to identify HGT events in biological species, one or more of the above mentioned methods were used in combination. However, the methods depend on the different type of gene transfer and their occurrence during the course of evolution.

6. HGT and Endophytes

The mutualistic association between endophytes and host plants has beneficial consequences for both plant development and microbial adaptation. In the evolution of biological species, HGT events were considered to be a strong, evolutionary mechanism for conferring new traits and assisting adaptation to adverse environments. However, reports on gene transfer between plants and endophytes were limited, suggesting that a more comprehensive research is required for understanding HGT signatures and their possible implications on the associated organisms. Significant studies on HGT in endophytes include the following—genetic manipulation of P. putida W619-TCE, an endophyte of poplar plant for degradation of TCE soil contaminant [91]; arsenic hyperaccumulation in endophytic bacteria [28]; toluene phytoremediation by endophytic bacteria [92]; endophyte-mediated toluene degradation and growth promotion in T. durum, Z. mays, and B. cepacia [25]. These studies highlighted the potential of HGT events in conferring novel traits to either of the associated species and their significance in ecological and biotechnological studies. In addition, studies on HGT in plant-associated
bacteria, specifically *Rhizobium* and *Xanthomonas*, revealed its significance as a mechanism for plant adaptation and survival [93]. Studies have also suggested the transfer of four cluster genes, through HGT in sugarcane endophyte *Burkholderia seminalis* TC3.4.2R3, where it is suggested to be a possible adaptation mechanism. Further studies on the four-gene cluster would provide valuable information about the biosynthetic mechanism of antifungal compounds [94].

6.1. HGT as A Function of Ecological Adaptation

Plants are dynamic communities colonized by complex microbial communities that influence plant growth and development. The association of microbial communities with host plant provides multiple advantages to both the microbe and the host plant. These microorganisms spend a part of or an entire life cycle inside the plant and might be present externally (rhizosphere) or internally (endosphere). Studies have suggested that the association of a microbe influences plant growth and development and confers resistance to environmental stresses [95, 96], while in exchange, the plant assists in microbial adaptation and survival [97]. Some key studies have suggested the horizontal transmission of bacterial endophytes via soil [98], colonization of the root endosphere via the rhizosphere [99], through aerial tissues [100], and as host plant colonization.

The HGT phenomenon was a key event that conferred novel traits to both prokaryotes and eukaryotes, during evolution [36, 38, 101]. An example of HGT between plants is the presence of the mitochondrial intron sequence in a number of angiosperms [102]. A similar study showed the acquisition of mitochondrial genes from algae and angiosperms to *Amborella trichopoda*, through HGT events [103]. However, the HGT of nuclear genes from prokaryotes to plants is relatively rare [18, 36, 97]. Whole-genome sequencing of plants, namely *Arabidopsis thaliana*, *Oryza sativa*, *Populus trichocarpa*, and *Sorghum bicolor* revealed fewer HGT events, suggesting their less frequent occurrence. One remarkable example is the first reported HGT of the β-1,6-glucanase gene from endophytic fungi to grasses [27]. The study performed de novo sequencing and assembly of the ryegrass genome and discovered the presence of the β-1,6-glucanase gene. Moreover, the transfer of β-1,6-glucanase gene from endophytic fungi to perennial ryegrass, suggested an evolutionary adaptation in angiosperms [27]. In the current perspective, similar studies are essential to elucidate the mechanism and importance of HGT in the evolution of plant-microbe associations.

6.2. Perspectives in Environment and Agriculture

Endophytic fungi have a long evolutionary history of association with land plants, affecting almost all plant species [104]. Fungal association defines a diverse relationship with the host plant, ranging from parasitism to commensalism and symbiosis. These organisms show very high rates of plant infection and causes multiple diseases. Diverse fungal species might colonize the same plant, which suggests a highly specific interaction with the respective plant host. Moreover, comparative genomics studies suggested that these fungi display similar genomic content to their associated organism, as well as symbiosis-specific genes [105, 106]. The endophytic fungi can influence the physiology of the host plant by protecting against pathogenic fungi [104, 107, 108], promoting plant growth [97, 109, 110], and aiding nutrient and water uptake [111]. Moreover, studies have demonstrated the positive impacts of plant-endophyte associations in maintaining ecological balance and their potential biotechnological applications. Endophytic microbes have been engineered for promoting phytoremediation [91]; and the natural process of HGT has been studied as a possible mechanism to introduce novel traits to the host plant [112] and the associated microbe [28, 92].

Recent studies highlighted the emerging importance of gene transfer events as a possible alternative mechanism, to acquire “novel traits” for potential socio-economic applications. Studies have been performed to understand the importance of HGT events between plants and endophytes, particularly the acquisition of “novel traits” by the associated organisms. Some endophytes were found to improve phytoremediation, with HGT as a possible mechanism of gene transfer. A recent study reported that the endophytic bacteria from *P. vittata* were effective arsenic accumulators inside the host plant.
The study discussed arsenic resistance and its associated genes in the endophytic bacteria, which are responsible for arsenic accumulation [28]. In total, 116 arsenite-resistant bacteria were isolated from *P. vittata* roots and characterized for arsenic tolerance. Results showed that the exchange of arsenite transporter genes *arsB/ACR3(2)*, possibly occurred through HGT between endophytic bacteria [28]. A similar study reported that the gene transfer between endophytic bacteria residing inside *T. durum* and *Z. mays*, induced toluene degradation and promoted plant growth [25]. The endophyte, *B. cepacia* FX2 containing the gene for catechol 2, 3-dioxygenase (C23O) on a plasmid, effectively degraded toluene in contaminated soil [25]. The characterization of this endophytic bacteria and its functional role in the environment was consequently established [25]. Moreover, several studies reported that the endophytic bacteria acquire new metabolic traits and adaptation to the environment, through HGT [42,43].

7. Significance and Directions for Future Research

HGT is considered to be a significant process in the evolution of prokaryotes, although it remains less studied as a controversial subject in eukaryotes [38]. Several studies have acknowledged HGT as a driving force in prokaryotic adaptation and survival, however, very few reports of HGT events in multicellular eukaryotes are available [18]. In eukaryotes, genomic content constitutes mobile genetic elements, known as transposable elements (TE), which can move from one region to another and duplicate themselves [113]. TEs are responsible for epigenetic changes and any genomic variation, thereby, influence the evolution of host species. Moreover, studies suggested that the movement of TEs across eukaryotic genomes might be caused by horizontal transposon transfer (HTT), a phenomenon linked to HGT, which plays a significant role in creating genomic variations [113].

The occurrence of HGT as a major evolutionary event has been described in various studies [36,94]. However, reports about HGT events between endophytes and plants are limited. Considering the significance of HGT in the adaptation and evolution of biological species, an extensive and in depth analysis on HGT events between endophytes and host plants is required. HGT research should focus on scientific methods utilizing available genomic data for analysis, to effectively detect the occurrence of HGT events. Current studies focusing on HGT events in eukaryotes depend on a comparative phylogenetic tree analysis and BLAST search methods. However, several genomic processes, such as gene duplication/deletion, introgression events, etc., have made the functional characterization of HGT difficult in biological species. Additionally, it is difficult to explain the occurrence and mechanism of HGT among distantly related species. No direct evidence is available on how an HGT event occurs across biological organisms; however, vector-mediated HGT via plasmids or transposable elements has been suggested for fungi [114]. Furthermore, studies regarding HGT in eukaryotes is limited, hindering the analysis of genomic data using statistical methods [115]; however, researchers have devised an advanced statistical method called coalescent statistics to study HGT in evolution [116]. The availability of high-quality genomic information and development of accurate phylogenetic methods can revolutionize and provide a better insight about the roles of HGT events in eukaryotes. Several challenges exist in defining the exact role of HGT in the evolution of biological species, such as determining phylogenetic relationships between organisms in the presence of HGT and identifying the preferred genes during HGT events, remain to be addressed.

In recent times, only a few studies have emphasized the advantages of utilizing HGT events in endophytes, and their potential applications in environmental and agricultural research. These studies include arsenic hyperaccumulation in endophytic bacteria [28], endophyte-mediated toluene degradation and growth promotion in *T. durum*, *Z. mays*, and *B. cepacia* [25], and toluene phytoremediation through endophytic bacteria [88]. These studies demonstrate that investigation of HGT events in endophytes highlights their potential in conferring novel traits to either of the associated species, consequently suggesting important ecological and biotechnological implications.
8. Conclusions

With the progress in whole genome sequencing, the availability of “big data” has enabled the study of entire genomes of biological organisms. The study of HGT in eukaryotic genomes has become feasible, leading to the discovery of genes linked with HGT in different taxonomic groups of eukaryotes [36]. In recent times, HGT events observed in plant lineages [28,108] were found to be more common than expected. Moreover, the significance of HGT events in the evolution and adaptation of prokaryotes and eukaryotes was established; it has emerged as a key mechanism driving several biological processes, including insect resistance, C4 photosynthesis in grasses, parasite dynamics, etc. [108]. In plant-associated microorganisms, the occurrence of HGT events was highlighted as an important evolutionary process for the acquisition of “novel traits” by either the host plant or the associated endophyte, which confers better adaptation and new functions to the organism and might have potential environmental and biotechnological applications. Studies regarding the HGT events in endophytes are very limited, hence, this review provides an insight on their functional role, significance, and potential socio-economic applications, in a current perspective.

Author Contributions: P.T. and H.B. discussed the concept of the manuscript. P.T. contributed to literature collection and writing of the manuscript. H.B. critically reviewed and revised the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by a grant from the Next-Generation Biogreen 21 Program (Project No. PJ013655022020), Rural Development Administration, Republic of Korea.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Bansal, A.K.; Meyer, T.E. Evolutionary analysis by whole-genome comparisons. J. Bacteriol. 2002, 184, 2260–2272. [CrossRef]
2. Vos, M.; Hesselman, M.C.; Te Beek, T.A.; van Passel, M.W.J.; Eyre-Walker, A. Rates of Lateral gene transfer in prokaryotes: High but why? Trends Microbiol. 2015, 23, 598–605. [CrossRef]
3. Rolland, T.; Neuveglise, C.; Sacerdot, C.; Dujon, B. Insertion of horizontally transferred genes within conserved syntenic regions of yeast genomes. PLoS ONE 2009, 4, e6515. [CrossRef]
4. Mc Ginty, S.E.; Rankin, D.J.; Brown, S.P. Horizontal gene transfer and the evolution of bacterial cooperation. Evolution 2011, 65, 21–32. [CrossRef]
5. Hotopp, J.C.D. Horizontal gene transfer between bacteria and animals. Trends Genet. 2011, 27, 157–163. [CrossRef]
6. Dagan, T.; Artzy-Randrup, Y.; Martin, W. Modular networks and cumulative impact of lateral transfer in prokaryote genome evolution. Proc. Natl. Acad. Sci. USA 2008, 105, 10039–10044. [CrossRef]
7. Deschavanne, P.; Mallet, L.V.; Becq, J. Whole genome evaluation of horizontal transfers in the pathogenic fungus Aspergillus fumigatus. BMC Genom. 2010, 11, 71.
8. Gogarten, J.P.; Doolittle, W.F.; Lawrence, J.G. Prokaryotic evolution in light of gene transfer. Mol. Biol. Evol. 2002, 19, 2226–2238. [CrossRef]
9. Kurdan, C.G.; Canback, B.; Berg, O.G. Horizontal gene transfer: A critical view. Proc. Natl. Acad. Sci. USA 2003, 100, 9658–9662. [CrossRef]
10. Tamames, J.; Moya, A. Estimating the extent of horizontal gene transfer in metagenomic sequences. BMC Genom. 2008, 9, 136. [CrossRef]
11. de Wit, P.J.G.M.; Mehrabi, R.; Bahkali, A.H.; Abd-Elsalam, K.A.; Moslem, M.; Ben M’Barek, S.; Gohari, A.M.; Jashni, M.K.; Stergiopoulos, I.; Kema, G.H.J. Horizontal gene and chromosome transfer in plant pathogenic fungi affecting host range. FEMS Microbiol. Rev. 2011, 35, 542–554.
12. Fournier, P.E.; Moliner, C.; Raoult, D. Evidence of horizontal gene transfer between amoeba and bacteria. Clin. Microbiol. Infect. 2009, 15, 178–180.
13. Dao, Y.P.; Qi, Y.M.; Ma, Y.F.; Xia, A.; Sharakhov, I.; Chen, X.G.; Biedler, J.; Ling, E.; Tu, Z.J. Next-generation sequencing reveals recent horizontal transfer of a DNA transposon between divergent mosquitoes. PLoS ONE 2011, 6, e16743. [CrossRef]
14. Seifert, H.S.; Anderson, M.T. Opportunity and means: Horizontal gene transfer from the human host to a bacterial pathogen. *mBio* 2011, 2, 5–11.
15. Li, B.; Zhu, B.; Lou, M.M.; Xie, G.L.; Zhang, G.Q.; Zhou, X.P.; Jin, G.L. Horizontal gene transfer in silkworm, *Bombyx mori*. BMC Genom. 2011, 12, 248.
16. Armijos Jaramillo, V.D.; Vargas, W.A.; Sukno, S.A.; Thon, M.R. Horizontal transfer of a Subtilisin gene from plants into an ancestor of the plant pathogenic fungal genus *Colletotrichum*. *PLoS ONE* 2013, 8, e59078. [CrossRef]
17. Babic’, A.; Lindner, A.B.; Vulic´, M.; Stewart, E.J.; Radman, M. Direct visualization of horizontal gene transfer. *Science* 2008, 319, 1533–1536. [CrossRef]
18. Bock, R. The give-and-take of DNA: Horizontal gene transfer in plants. *Trends Plant Sci.* 2010, 15, 11–22. [CrossRef]
19. Huang, J. Horizontal gene transfer in eukaryotes: The weak-link model. *Bioessays* 2013, 35, 868–875. [CrossRef]
20. Popa, O.; Dagan, T. Trends and barriers to lateral gene transfer in prokaryotes. *Curr. Opin. Microbiol.* 2011, 14, 615–623. [CrossRef]
21. Juhas, M. Horizontal gene transfer in human pathogens. *Crit. Rev. Microbiol.* 2015, 41, 101–108. [CrossRef] [PubMed]
22. Lacroix, B.; Citovsky, V. Transfer of DNA from bacteria to eukaryotes. *mBio* 2016, 7, e00863-16. [CrossRef] [PubMed]
23. Husnik, F.; McCutcheon, J.P. Functional horizontal gene transfer from bacteria to eukaryotes. *Nat. Rev. Microbiol.* 2017, 16, 67–79. [CrossRef] [PubMed]
24. McAnerney, J.O. Horizontal gene transfer is less frequent in eukaryotes than prokaryotes but can be important. *Bioessays* 2017, 39, 1700002. [CrossRef] [PubMed]
25. Wang, Y.; Li, H.; Zhao, W.; He, X.; Chen, J.; Geng, X.; Xiao, M. Induction of toluene degradation and growth promotion in corn and wheat by horizontal gene transfer within endophytic bacteria. *Soil Biol. Biochem.* 2010, 42, 1051–1057. [CrossRef]
26. Tan, R.X.; Zou, W.X. Endophytes: A rich source of functional metabolites. *Nat. Prod. Rep.* 2001, 18, 448–459. [CrossRef]
27. Shinozuka, H.; Hettiarachchige, I.K.; Shinozuka, M.; Cogan, N.O.I.; Spangenberg, G.C.; Cocks, B.G.; Forster, J.W.; Sawbridge, T.I. Horizontal transfer of a β-1,6-glucanase gene from an ancestral species of fungal endophyte to a cool-season grass host. *Sci. Rep.* 2017, 7, 9024. [CrossRef] [PubMed]
28. Gu, Y.; Wang, Y.; Sun, Y.; Zhao, K.; Xiang, Q.; Yu, X.; Zhang, X.; Chen, Q. Genetic diversity and characterization of arsenic-resistant endophytic bacteria isolated from *Pteris vittata*, an arsenic hyperaccumulator. *BMC Microbiol.* 2018, 18, 42. [CrossRef]
29. Sachin, N.; Manjunatha, B.L.; Kumara, P.M.; Ravikanth, G.; Shweta, S.; Suryanarayanan, T.S.; Ganeshaiah, K.N.; Shaanker, R.U. Do endophytic fungi possess pathway genes for plant secondary metabolites? *Curr. Sci.* 2013, 104, 178–182.
30. Frisvad, J.C.; Andersen, B.; Thrane, U. The use of secondary metabolite profiling in chemotaxonomy of filamentous fungi. *Mycol. Res.* 2008, 112, 231–240. [CrossRef]
31. Barac, T.; Taghavi, S.; Borremans, B.; Provoost, A.; Oeyen, L.; Colpaert, J.V.; Vangronsveld, J.; van der Lelie, D. Engineered endophytic bacteria improve phytoremediation of water-soluble, volatile, organic pollutants. *Nat. Biotechnol.* 2004, 22, 583–588. [CrossRef] [PubMed]
32. Li, M.; Zhao, J.; Tang, N.; Sun, H.; Huang, J. Horizontal gene transfer from bacteria and plants to the arbuscular mycorrhizal fungus *Rhizophagus irregularis*. *Front. Plant Sci.* 2018, 9, 701. [CrossRef] [PubMed]
33. Syvanen, M. Cross-species gene transfer; implications for a new theory of evolution. *J. Theor. Biol.* 1985, 112, 333–343. [CrossRef]
34. Smith, M.W.; Feng, D.F.; Doolittle, R.F. Evolution by acquisition: The case for horizontal gene transfers. *Trends Biochem. Sci.* 1992, 17, 489–493. [CrossRef]
35. Martin, W.; Rujan, T.; Richly, E.; Hansen, A.; Cornelsen, S.; Lins, T.; Leister, D.; Stoebe, B.; Hasegawa, M.; Penny, D. Evolutionary analysis of *Arabidopsis*, cyanobacterial, and chloroplast genomes reveals plastid phylogeny and thousands of cyanobacterial genes in the nucleus. *Proc. Natl. Acad. Sci. USA* 2002, 99, 12246–12251. [CrossRef]
36. Soucy, S.M.; Huang, J.; Gogarten, J.P. Horizontal gene transfer: Building the web of life. *Nat. Rev. Genet.* 2015, 16, 472–482. [CrossRef]
37. Kurland, C.G. What tangled web: Barriers to rampant horizontal gene transfer. *Bioessays* 2005, 27, 741–747. [CrossRef]
38. Keeling, P.J.; Palmer, J.D. Horizontal gene transfer in eukaryotic evolution. *Nat. Rev. Genet.* 2008, 9, 605–618. [CrossRef]
39. Boto, L. Horizontal gene transfer in evolution: Facts and challenges. *Proc. R. Soc. B* 2010, 277, 819–827. [CrossRef]
40. Ricard, G.; McEwan, N.R.; Dutilh, B.E.; Jouany, J.; Macheboeuf, D.; Mitsumori, M.; McIntosh, F.M.; Michalowski, T.; Nagamine, T.; Nelson, N.; et al. Horizontal gene transfer from bacteria to rumen ciliates indicates adaptation to their anaerobic, carbohydrates-rich environment. *BMC Genom.* 2006, 7, 22. [CrossRef]
41. Garcia-Vallve, S.; Romeu, A.; Palau, J. Horizontal gene transfer of glycosyl hydrolases of the rumen fungi. *Mol. Biol. Evol.* 2000, 17, 352–361. [CrossRef] [PubMed]
42. Dong, Q.; Springeal, D.; Schoeters, J.; Nuyts, G.; Mergeay, M.; Diels, L. Horizontal transfer of bacterial heavy metal resistance genes and its applications in activated sludge systems. *Water Sci. Technol.* 1998, 37, 465–468. [CrossRef]
43. van der Lelie, D.; Barac, T.; Taghavi, S.; Vangronsveld, J. Response to Newman: New uses of endophytic bacteria to improve phytoremediation. *Trends Biotechnol.* 2005, 23, 8–9. [CrossRef]
44. Richards, T.A.; Soanes, D.M.; Foster, P.G.; Leonard, G.; Thornton, C.R.; Talbot, N.J. Phylogenomic analysis demonstrates a pattern of rare and ancient Horizontal gene transfer between plants and fungi. *Plant Cell* 2009, 21, 1897–1911. [CrossRef] [PubMed]
45. Ochman, H.; Lawrence, J.G.; Grolsman, E.A. Lateral gene transfer and the nature of bacterial innovation. *Nature* 2000, 405, 299–304. [CrossRef]
46. Pal, C. Adaptive evolution of bacterial metabolic networks by horizontal gene transfer. *Nat. Genet.* 2005, 37, 1372–1375. [CrossRef]
47. Thomas, C.M.; Nielsen, K.M. Mechanisms of and barriers to horizontal gene transfer between bacteria. *Nat. Rev. Microbiol.* 2005, 3, 711–721. [CrossRef]
48. Polz, M.F. Horizontal gene transfer and the evolution of bacterial and archaeal population structure. *Trends Genet.* 2013, 29, 170–175. [CrossRef]
49. Andersson, J.O. Lateral gene transfer in eukaryotes. *Cell. Mol. Life Sci.* 2005, 62, 1182–1197. [CrossRef]
50. Gao, C.; Ren, X.; Mason, A.S.; Liu, H.; Xiao, M.; Li, J.; Fu, D. Horizontal gene transfer in plants. *Funct. Integr. Genom.* 2014, 14, 23–29. [CrossRef]
51. Trobridge, G.D. Foamy virus vectors for gene transfer. *Expert Opin. Biol. Ther.* 2009, 9, 1427–1436. [CrossRef] [PubMed]
52. Danchin, E.G.J.; Haegeman, A.; Jones, J.T. Horizontal gene transfer in nematodes: A catalyst for plant parasitism? *Mol. Plant Microbe Interact.* 2011, 24, 879–887.
53. Thyssen, G.; Svab, Z.; Maliga, P. Cell-to-cell movement of plastids in plants. *Proc. Natl. Acad. Sci. USA* 2012, 109, 2439–2443. [CrossRef] [PubMed]
54. Archibald, J.M.; Richards, T.A. Gene transfer: Anything goes in plant mitochondria. *BMC Biol.* 2010, 8, 147. [CrossRef]
55. Vogel, T.M.; Pontirolli, A.; Rizzi, A.; Simonet, P.; Daffonchio, D.; Monier, J.M. Visual evidence of horizontal gene transfer between plants and bacteria in the phytosphere of transplastomic tobacco. *Appl. Environ. Microb.* 2009, 75, 3314–3322.
56. Moran, N.A.; Jarvik, T. Lateral transfer of genes from fungi underlies carotenoid production in aphids. *Science* 2010, 328, 624–627. [CrossRef]
57. Broothaerts, W.; Mitchell, H.J.; Weir, B.; Kaines, S.; Smith, L.M.A.; Yang, W.; Mayer, J.E.; Roa-Rodriguez, C.; Jefferson, R.A. Gene transfer to plants by diverse species of bacteria. *Nature* 2005, 433, 629–633. [CrossRef]
58. Berghorsson, U.; Adams, K.L.; Thomason, B.; Palmer, J.D. Widespread horizontal transfer of mitochondrial genes in flowering plants. *Nature* 2003, 424, 197–201. [CrossRef]
59. Sommer, R.J.; Mayer, W.E.; Schuster, L.N.; Bartelmes, G.; Dieterich, C. Horizontal gene transfer of microbial cellulases into nematode genomes is associated with functional assimilation and gene turnover. *BMC Evol. Biol.* 2011, 11, 13.
60. Toft, C.; Andersson, S.G.E. Evolutionary microbial genomics: Insights into bacterial host adaptation. *Nat. Rev. Genet.* 2010, 11, 465–475. [CrossRef]

61. Ma, W.; Dong, F.F.; Stavrini, J.; Guttman, D.S. Type III effector diversification via both pathoadaptation and horizontal transfer in response to a coevolutionary arms race. *PLoS Genet.* 2006, 2, e209. [CrossRef]

62. Frank, A.C.; Alsmark, C.M.; Thollesson, M.; Andersson, S.G. Functional divergence and horizontal transfer of type IV secretion systems. *Mol. Biol. Evol.* 2005, 22, 1325–1336. [CrossRef] [PubMed]

63. Lederberg, J.; Tatum, E.L. Gene recombination in *Escherichia coli*. *Nature* 1946, 158, 558. [CrossRef] [PubMed]

64. Zinder, N.D.; Lederberg, J. Genetic exchange in *Salmonella*. *J. Bacteriol.* 1952, 64, 679–699. [CrossRef] [PubMed]

65. Jain, R.; Rivera, M.C.; Lake, J.A. Horizontal gene transfer among genomes: The complexity hypothesis. *Proc. Natl. Acad. Sci. USA* 1999, 96, 3801–3806. [CrossRef]

66. Choi, I.G.; Kim, S.H. Global extent of horizontal gene transfer. *Proc. Natl. Acad. Sci. USA* 2007, 104, 4489–4494. [CrossRef]

67. Wagner, A.; De la Chaux, N. Distant horizontal gene transfer is rare for multiple families of prokaryotic insertion sequences. *Mol. Genet. Genom.* 2008, 280, 397–408. [CrossRef]

68. Wolf, Y.I.; Rogozin, I.B.; Grishin, N.V.; Koonin, E.V. Genome trees and the tree of life. *Trends Genet.* 2002, 18, 472–479. [CrossRef]

69. Brown, J. Ancient horizontal gene transfer. *Nat. Rev. Genet.* 2003, 4, 121–132. [CrossRef]

70. Sorek, R.; Zhu, Y.; Creevey, C.J.; Francino, M.P.; Bork, P.; Rubin, E.M. Genome-wide experimental determination of barriers to horizontal gene transfer. *Science* 2007, 318, 1449–1452. [CrossRef]

71. Goldenfeld, N.; Woese, C. Biology’s next revolution. *Nature* 2007, 445, 369. [CrossRef] [PubMed]

72. O’Malley, M.A.; Boucher, Y. Paradigm change in evolutionary microbiology. *Stud. Hist. Philos. Biol. Biomed. Sci.* 2005, 36, 183–208. [CrossRef] [PubMed]

73. Koonin, E.V. Darwinian evolution in the light of genomics. *Nucleic Acids. Res.* 2009, 37, 1011–1034. [CrossRef] [PubMed]

74. Keeling, P. Functional and ecological impacts of horizontal gene transfer in eukaryotes. *Curr. Opin. Genet. Dev.* 2009, 19, 613–619. [CrossRef] [PubMed]

75. Marcet-Houben, M.; Gabaldon, T. Acquisition of prokaryotic genes by fungal genomes. *Trends Genet.* 2010, 26, 5–8. [CrossRef] [PubMed]

76. Novo, M.; Bigey, F.; Beyne, E.; Galeote, V.; Gavory, F.; Mallet, S.; Cambon, B.; Legras, J.L.; Wincker, P.; Casaregola, S.; et al. Eukaryote-to-eukaryote gene transfer events revealed by the genome sequence of the wine yeast *Saccharomyces cerevisiae*. *Mol. Genet. Genom.* 2008, 280, 409–420. [CrossRef] [PubMed]

77. Walton, J.D. Horizontal gene transfer and the evolution of secondary metabolite gene clusters in fungi: An hypothesis. *Fungal Genet. Biol.* 2000, 30, 167–171. [CrossRef] [PubMed]

78. Slot, J.C.; Andersson, S.G.E. Evolutionary microbial genomics: Insights into bacterial host adaptation. *Nat. Rev. Genet.* 2010, 11, 465–475. [CrossRef]

79. Gardiner, D.M.; McDonald, M.C.; Covarelli, L.; Solomon, P.S.; Rusu, A.G.; Marshall, M.; Kazan, K.; Slot, J.C.; Rokas, A. Multiple GAL pathway gene clusters evolved independently and by different mechanisms in *Saccharomyces cerevisiae*. *Proc. Natl. Acad. Sci. USA* 2010, 107, 10136–10141. [CrossRef]

80. Gardiner, D.M.; McDonald, M.C.; Covarelli, L.; Solomon, P.S.; Rusu, A.G.; Marshall, M.; Kazan, K.; Chakraborty, S.; McDonald, B.A.; Manners, J.M. Comparative pathogenomics reveals horizontally acquired novel virulence genes in fungi infecting cereal hosts. *PLoS Pathog.* 2012, 8, e1002952. [CrossRef] [PubMed]

81. Yue, J.; Hu, X.; Sun, H.; Yang, Y.; Huang, J. Widespread impact of horizontal gene transfer on plant colonization of land. *Nat. Commun.* 2012, 3, 1152. [CrossRef] [PubMed]

82. Eisen, J.A. Horizontal gene transfer among microbial genomes: New insights from complete genome analysis. *Curr. Opin. Genet. Dev.* 2000, 10, 606–611. [CrossRef]

83. Spellberg, B.; Guidos, R.; Gilbert, D.; Bradley, J.; Boucher, H.W.; Scheld, W.M.; Bartlett, J.G.; Edwards, J., Jr.; Infectious Diseases Society of America. The epidemic of antibiotic-resistant infections: A call to action for the medical community from the Infectious Diseases Society of America. *Clin. Infect. Dis.* 2008, 46, 155–164. [CrossRef] [PubMed]

84. Doolittle, W.F.; Logsdon, J.M., Jr. Do Archaea have a mixed heritage? *Curr. Biol.* 1998, 8, 209–211. [CrossRef]

85. Gatesy, J.; Desalle, R.; Wheeler, W. Alignment-ambiguous nucleotide sites and the exclusion of systematic data. *Mol. Phylog. Evol.* 1993, 2, 152–157. [CrossRef]
86. Eisen, J.A. The RecA protein as a model molecule for molecular systematic studies of bacteria: Comparison of trees of RecAs and 16s rRNAs from the same species. *J. Mol. Evol.* **1995**, *41*, 1105–1123. [CrossRef]
87. Philippe, H.; Forterre, P. The rooting of the universal tree of life is not reliable. *J. Mol. Evol.* **1999**, *49*, 509–523. [CrossRef]
88. Mrazek, J.; Karlin, S. Detecting alien genes in bacterial genomes. *Ann. N.Y. Acad. Sci.* **1999**, *870*, 314–329. [CrossRef]
89. Lafay, B.; Lloyd, A.T.; McLean, M.J.; Devine, K.M.; Sharp, P.M.; Wolfe, K.H. Proteome composition and codon usage in spirochaetes: Species-specific and DNA strand-specific mutational biases. *Nucleic Acids Res.* **1999**, *27*, 1642–1649. [CrossRef]
90. Eisen, J.A. Phylogenomics: Improving functional predictions for uncharacterized genes by evolutionary analysis. *Genome Res.* **1998**, *8*, 163–167. [CrossRef]
91. Weyens, N.; van der Lelie, D.; Artois, T.; Smeets, K.; Taghavi, S.; Newman, L.; Carlee, R.; Vangronsveld, J. Bioaugmentation with engineered endophytic bacteria improves contaminant fate in phytoremediation. *Environ. Sci. Technol.* **2009**, *43*, 9413–9418. [CrossRef] [PubMed]
92. Taghavi, S.; Barac, T.; Greenberg, B.; Borremans, B.; Vangronsveld, J.; van der Lelie, D. Horizontal gene transfer to endogenous endophytic bacteria from poplar improves phytoremediation of toluene. *Appl. Environ. Microbiol.* **2005**, *71*, 8500–8505. [CrossRef] [PubMed]
93. van Elsas, J.D.; Turner, S.; Bailey, M.J. Horizontal gene transfer in the phytosphere. *New Phytol.* **2003**, *157*, 525–537. [CrossRef]
94. Tsui, S. Evidence of horizontal gene transfer of a four-genes cluster exclusively in the sugarcane endophytic strain *Burkholderia seminalsis* TC3.4.2R3. *J. Med. Microbiol. Diagn.* **2018**, *7*. [CrossRef]
95. Santoyo, G.; Moreno-Hagesild, G.; Orozco-Mosqueda, M.; Glick, B.R. Plant growth-promoting bacterial endophytes. *Microbiol. Res.* **2016**, *183*, 92–99. [CrossRef]
96. Domka, A.M.; Rozpadek, P.; Turnau, K. Are fungal endophytes merely mycorrhizal copycats? The role of fungal endophytes in the adaptation of plants to metal toxicity. *Front. Microbiol.* **2019**, *10*, 371. [CrossRef]
97. Suman, A.; Yadav, A.N.; Verma, P. Endophytic microbes in crops: Diversity and beneficial impact for sustainable agriculture. In *Microbial Inoculants in Sustainable Agricultural Productivity*, Singh, D.P., Abhilash, P.C., Prabha, R., Eds.; Springer: Berlin, 2016; pp. 117–143.
98. Zarraonaindia, I.; Owens, S.M.; Weisenhorn, P.; West, K.; Hampton-Marcell, J.; Lax, S.; Bokulich, N.A.; Mills, D.A.; Martin, G.; Taghavi, S.; et al. The Soil Microbiome Influences Grapevine-Associated Microbiota. *mBio* **2015**, *6*, e02527-14. [CrossRef]
99. Vergani, L.; Mapielli, F.; Zanardini, E.; Terzaghi, E.; Di Guardo, A.; Morosini, C.; Raspa, G.; Borin, S. Phyto-rhizoremediation of polychlorinated biphenyl contaminated soils: An outlook on plant-microbe beneficial interactions. *Sci. Total Environ.* **2017**, *575*, 1395–1406. [CrossRef]
100. Bodenhausen, N.; Horton, M.W.; Bergelson, J. Bacterial communities associated with the leaves and the roots of *Arabidopsis thaliana*. *PLoS ONE* **2013**, *8*, e66329. [CrossRef]
101. Schonknecht, G.; Weber, A.P.M.; Lercher, M.J. Horizontal gene acquisitions by eukaryotes as drivers of adaptive evolution: Insights and perspectives. *BioEssays* **2014**, *36*, 9–20. [CrossRef]
102. Sanchez-Puerta, M.V.; Abbona, C.C.; Zhuo, S.; Tepe, E.J.; Bohs, L.; Olmstead, R.G.; Palmer, J.D. Multiple recent horizontal transfers of the coxl intron in Solanaceae and extended co-conversion of flanking exons. *BMC Evol. Biol.* **2011**, *11*, 277. [CrossRef] [PubMed]
103. Rice, D.W.; Alverston, A.J.; Richardson, A.O.; Young, G.J.; Sanchez-Puerta, M.V.; Munzinger, J.; Barry, K.; Boore, J.L.; Zhang, Y.; dePamphilis, C.W.; et al. Horizontal transfer of entire genomes via mitochondrial fusion in the angiosperm *Amborella*. *Science* **2013**, *342*, 1468–1473. [CrossRef] [PubMed]
104. Hardoim, P.R.; van Overbeek, L.S.; Berg, G.; Pirrillä, A.M.; Compan, S.; Campisano, A.; Döring, M.; Sessitsch, A. The hidden world within plants: Ecological and evolutionary considerations for defining functioning of microbial endophytes. *Microbiol. Mol. Biol. Rev.* **2015**, *79*, 293–320. [CrossRef] [PubMed]
105. Kuo, A.; Kohler, A.; Martin, F.M.; Grigoriev, I.V. Expanding genomics of mycorrhizal symbiosis. *Front. Microbiol.* **2014**, *5*, 582. [CrossRef]
106. Kohler, A.; Kuo, A.; Nagy, L.G.; Morin, E.; Barry, K.W.; Buscot, F.; Canbäck, B.; Choi, C.; Cichocki, N.; Clum, A.; et al. Convergent losses of decay mechanisms and rapid turnover of symbiosis genes in mycorrhiza mutualists. *Nat. Genet.* **2015**, *47*, 410–415. [CrossRef]
107. Arnold, A.E.; Mejía, L.C.; Kyllo, D.; Rojas, E.I.; Maynard, Z.; Robbins, N.; Herre, E. Fungal endophytes limit pathogen damage in a tropical tree. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 15649–15654. [CrossRef]

108. Kyekyeku, J.O.; Kusari, S.; Adosraku, R.K.; Bullach, A.; Golz, C.; Strohmann, C.; Spiteller, M. Antibacterial secondary metabolites from an endophytic fungus, *Fusarium solani* JK10. *Fitoterapia* **2017**, *119*, 108–114. [CrossRef]

109. Rodriguez, R.J.; White, J.F., Jr.; Arnold, A.E.; Redman, R.S. Fungal endophytes: Diversity and functional roles. *New Phytol.* **2009**, *182*, 314–330. [CrossRef]

110. Acuña-Rodríguez, I.S.; Hansen, H.; Gallardo-Cerda, J.; Atala, C.; Molina-Montenegro, M.A. Antarctic extremophiles: Biotechnological alternative to crop productivity in saline soils. *Front. Bioeng. Biotechnol.* **2019**, *7*, 22. [CrossRef]

111. Naranjo-Ortiz, M.A.; Gabaldon, T. Fungal evolution: Major ecological adaptations and evolutionary transitions. *Biol. Rev.* **2019**, *94*, 1443–1476. [CrossRef]

112. Wickell, D.A.; Li, F. On the evolutionary significance of horizontal gene transfers in plants. *New Phytol.* **2019**, *225*, 113–117. [CrossRef] [PubMed]

113. Schaack, S.; Gilbert, C.; Feschotte, C. Promiscuous DNA: Horizontal transfer of transposable elements and why it matters for eukaryotic evolution. *Trends Ecol. Evol.* **2010**, *25*, 537–546. [CrossRef] [PubMed]

114. Rosewich, U.L.; Kistler, H.C. Role of horizontal gene transfer in the evolution of fungi. *Annu. Rev. Phytopathol.* **2000**, *38*, 325–363. [CrossRef] [PubMed]

115. Dupont, P.Y.; Cox, M.P. Genomic data quality impacts automated detection of lateral gene transfer in fungi. *G3 Genes Genome Genet.* **2017**, *7*, 1301–1314. [CrossRef] [PubMed]

116. Du, P.; Ogilvie, H.; Nakheh, L. Unifying gene duplication, loss, and coalescence on phylogenetic networks. *bioRxiv* **2019**, 589655. [CrossRef]