Extreme Variations of pCO₂ and pH in a Macrophyte Meadow of the Baltic Sea in Summer: Evidence of the Effect of Photosynthesis and Local Upwelling

Vincent Saderne¹², Peer Fietzek³, Peter Maria Jozef Herman²

¹ Benthic Ecology group, GEOMAR: Helmholtz Center for Ocean Research in Kiel, Kiel, Schleswig-Holstein, Germany, ² Spatial Ecology, NIOZ: Royal Netherlands Institute for Sea Research, Yerseke, Zeeland, The Netherlands, ³ CONTROS Systems & Solutions GmbH, Kiel, Schleswig-Holstein, Germany

Abstract

The impact of ocean acidification on benthic habitats is a major preoccupation of the scientific community. However, the natural variability of pCO₂ and pH in those habitats remains understudied, especially in temperate areas. In this study we investigated temporal variations of the carbonate system in nearshore macrophyte meadows of the western Baltic Sea. These are key benthic ecosystems, providing spawning and nursery areas as well as food to numerous commercially important species. In situ pCO₂, pH (total scale), salinity and PAR irradiance were measured with a continuous recording sensor package dropped in a shallow macrophyte meadow (Eckernförde bay, western Baltic Sea) during three different weeks in July (pCO₂ and PAR only), August and September 2011. The mean (± SD) pCO₂ in July was 383±117 µatm. The mean (± SD) pCO₂ and pHeq in August were 239±20 µatm and 8.22±0.1, respectively. The mean (± SD) pCO₂ and pHeq in September were 1082±711 µatm and 7.83±0.40, respectively. Daily variations of pCO₂ due to photosynthesis and respiration (difference between daily maximum and minimum) were of the same order of magnitude: 281±88 µatm, 219±89 µatm and 1488±574 µatm in July, August and September respectively. The observed variations of pCO₂ were explained through a statistical model considering wind direction and speed together with PAR irradiance. At a time scale of days to weeks, local upwelling of elevated pCO₂ water masses with offshore winds drives the variation. Within days, primary production is responsible. The results demonstrate the high variability of the carbonate system in nearshore macrophyte meadows depending on meteorology and biological activities. We highlight the need to incorporate these variations in future pCO₂ scenarios and experimental designs for nearshore habitats.

Introduction

Human activities since the 19th century led to an increase of atmospheric pCO₂ from 280 to 392 µatm [1] and the trend is rising. Some scenarios expect an elevation of atmospheric pCO₂ up to 1000 µatm during the 21th century, peaking around 1400 µatm in the year 2300 [2,3]. As oceans equilibrate with the atmosphere, dissolution of CO₂ in water induces a decrease in pH. Global change has already led to an average seawater pH of 8.15 in the year 2005 [4]. Acidification is enhancing the corrosiveness of seawater to calcite and aragonite, the two isoforms of calcium carbonates composing the shells and skeletons of marine organisms. Corrosiveness is expressed by the saturation states Ωarag and Ωcalc. A saturation state below 1 indicates a tendency towards dissolution of the crystal. Aragonitic structures (e.g. scleratinian corals, nacre of bivalve shells) are more soluble than calcitic (e.g. outer shell of oyster) [4–5].

Once dissolved, CO₂ becomes part of the carbonate system, almost entirely composed of bicarbonate (HCO₃⁻) and carbonate ions (CO₃²⁻). Under actual atmospheric concentrations of CO₂, CO₂(aq) in the surface ocean represents less than 1% of the dissolved inorganic carbon (DIC) while HCO₃⁻ represents ~90% and CO₃²⁻ ~10%. The carbonate system in open oceanic environments is well known and most of the data forming the basis for the predictive models (see GLODAP database, [5,6]) are derived from the open ocean. Those oceanic actual and future pCO₂/pH values are the ones referred to when designing ocean acidification studies [7]. However, the biogeochemistry of nearshore ecosystems features more variations and differs widely than offshore conditions [8,9]. Shallow nearshore and estuarine areas are the habitat of numerous benthic calcifiers. As highlighted by Andersson and Mackenzie (2012) [10], investigations on the effects of ocean acidification on calcifiers are neglecting this natural variability.

In nearshore habitats, the few available investigations were conducted on: (1) estuaries, salt marshes, mangroves and mudflats where transfers of carbon from land are occurring (e.g. [11–13]) and (2) reefs formed by corals or calcifying algae because of the direct effect of calcification on the carbonate system (e.g. [14,15]). The carbonate chemistry of nearshore habitats dominated by macrophytes (kelp forests and seagrass and/or seaweed meadows)
Carbonate Chemistry in Macrophyte Meadows

is almost unknown, even though they represent ~5% of the primary production in the ocean and ⅓ of the vegetal biomass [16,17]. In these habitats, the carbonate system is driven by the photosynthetic physiology of the macrophytes, taking up carbon during the day and releasing it by respiration during the night [18,19]. Since CO₂ is a major component of DIC and its diffusion rate in seawater is 10,000 times lower than in air [20], most macrophytes are relying on HCO₃⁻ for photosynthesis [21–24]. Macrophyte meadows are highly productive and net autotrophic (carbon sink) [25,26] exporting their excess production to the neighboring ecosystems under the form of litter or dissolved organic carbon [27]. Microbial degradation of exported organic carbon leads to elevated pCO₂ and hypoxia of deeper offshore waters [28]. This phenomenon is particularly important in eutrophied ecosystems like the Baltic Sea [29]. In the inner bays of the western Baltic, local upwelling of hypercapnic water masses is regularly observed, increasing the surface pCO₂ up to 2500 μatm. This typically happens at the end of summer when offshore winds are upwelling deep waters after long periods of stratification [30–32].

The aim of this study, conducted in the western Baltic Sea over three weeks of summer 2011, was to measure (1) the day/night variations of pCO₂ due to photosynthesis and respiration of a macrophyte bed, (2) the evolution of the baseline pCO₂ over summer and (3) the effect of local upwellings on the carbonate system and its diel variations. The variations of DIC, measured in the macrophyte meadow, were modeled with a statistical model considering wind direction and speed together with photosynthetically active radiation (PAR).

Materials and Methods

2.1 Study site

Physico-chemical parameters of seawater were recorded in a macrophyte meadow (3 m depth) in Eckernförde Bay (western Baltic Sea, Germany, 54°27’ N, 9°54’ E, see Fig. 1), during 3 weeks of summer 2011: July: 29.06–08.07, August: 29.07–05.08 and September: 09.09–16.09. In July, only pCO₂ and PAR irradiance were recorded by two independent sensors. For the August and September deployments, a multimeter recording pH, salinity and temperature was added to the CO₂ and PAR sensor. Wind speed (m s⁻¹) and direction (rad) with a resolution of 10 min were provided by the meteorological station of Aschau in Eckernförde Bay (54°27’40” N, 9°53’50” E) belonging to the division of marine technology and research of the German Navy. The nearshore habitat of Eckernförde is a mixed bottom type dominated by macrophyte vegetation. The macrophyte vegetation covers approximately 75% of the surface. Dominant species are the brown algae Fucus serratus (40–60% of the macrophytes), growing on stones and rocks and the seagrass Zostera marina (<10% of the macrophytes) growing on sandy bottoms [33]. The inner basin reaches a maximum depth of 28 m in gentle slope. The water column is stratified in summer, with a pycnocline around basin reaches a maximum depth of 28 m in gentle slope. The of the macrophytes) growing on sandy bottoms [33]. The inner

The pCO₂ was measured with a HydroCTM underwater sensor (CONTROS Systems & Solutions, Germany). The instrument measures the CO₂ mole fraction in a headspace behind a membrane equilibrator with a two-wavelength non-dispersive infra-red detector (NDIR). The equilibrator is composed of a flat silicone composite membrane, the NDIR unit and additional sensors for pressure, temperature and relative humidity used to correct the NDIR signal and calculate the partial pressure. A small internal pump insures the continuous circulation of air in the equilibrator. To correct the drifting of the instrument with time, regular zeroings are automatically performed by scrubbing the CO₂ from the internal gas stream. The zeroings are accounted during post processing of the data. A first description of the instrument can be found in [36] and deployments on various platforms are presented in [37]. The instrument was calibrated by the manufacturer prior to every deployment in the range of 100–1000 μatm at 17°C, 18°C and 18°C for the first, second and third deployment respectively. An overall accuracy of better than ±1% of the upper range value is assumed for post processed data. In order to achieve a constant and sufficiently short response time of the instrument, the HydroCTM is equipped with a water pump (Sea-Bird Electronics, USA) that provides a continuous water flow of approx. 35 ml sec⁻¹ to the membrane, leading to a response time T₆₃% of 3 min and a T₉₉% of approximately 15 min. These response times are sufficiently fast to resolve the observed signal changes without the need for signal deconvolution. To correct the pCO₂ series for the instrument drift, the zeroing signals were regarded as nodes and linearly interpolated in time to result in a smooth signal drift correction throughout the deployment time.

Salinity (psu), voltage (analogue signal from the pH measurement) and temperature (°C) were recorded automatically every 45 min with a pH-meter/conductimeter Mettler-Toledo SG 7/8 (Mettler Toledo, Switzerland). For the measurement of pH on total scale, seawater TRIS pH buffers for 15 psu were made according to the SOP 6a of Dickson et al. (2007) [38]. A combined electrode with a solid polymer electrolyte reference, equipped with an NTC (negative temperature coefficient) thermostor was used (Inlab expert pro, Mettler Toledo, Switzerland). The calibration of a new electrode was made 24 h before each deployment. To do so, the TRIS buffer was immersed in a thermostatic bath and the voltage of the pH electrode was measured with an accuracy of ±0.1 mV. The temperature of the buffer was varied by 1.5°C around seawater temperature. The temperature corresponding to every mV change was recorded with accuracy better than 0.01°C with a Fluke 5638 reference thermometer dotted of a 5608 platinum resistance sensor (Fluke, USA). This process was repeated by increasing and decreasing the temperature to get an average voltage (mV) versus 1°C reference curve for the electrode in the buffer. The NTC sensor of the pH sensor, of resolution 0.1°C, was calibrated against a reference thermometer and the resulting regression was achieved with a R² >0.99. The resulting equation was used to correct the sampled temperature. The sampled voltage and corrected temperature were converted to pH by making use of the initial TRIS buffer calibration of the electrode and by using the equations given in the SOP 6a. In the lab, work conducted on Certified Reference Material (CRM) (Andrew Dickson, Scripps Institution of Oceanography) together with 35 psu buffers demonstrated an accuracy of 0.003 to 0.005 pH units and a precision better than 0.001 pH unit. However, this accuracy was not reached in the field and therefore the pH series were not used to derive the carbonate system but instead the alkalinity from discrete samples (see 2.3 for method and 4.4 for discussion on the method).

Salinity was measured within 0.01 psu by a Mettler Toledo Inlab 738 conductivity probe after calibration at 25°C with KCl 0.1 mol L⁻¹ (Fischer Scientific, USA). The PAR irradiance (400 to
700 nm) was recorded every 5 min using a LI-192 quantum sensor connected to a LI-1400 data logger (Li-Cor Biosciences, USA). All series were extended to one minute interval series by linear interpolation.

2.3 Calculation of the carbonate system

Samples for alkalinity were taken at the beginning, middle and end of the measurement periods of August (the 29.07, 02.08 and 05.08, no replication) and September (the 09.09, 13.09 and 16.09, one replicate). Alkalinity was measured with an accuracy of \( \pm 5 \mu\text{mol kg}^{-1} \) using an open cell potentiometric titrator as described in the SOP 3b of [38] (Titrand 888, Methrom, Switzerland). A regression was calculated between the total alkalinity and salinity of the samples from August and September pooled together. The regression was highly significant (F-statistic: \( p < 0.001 \), \( R^2 = 0.83 \), \( n = 9 \)): 

\[
\text{Alkalinity} = 905.17 \times \text{Salinity} + 59.83,
\]

with both slope and intercept significant at \( p < 0.001 \) and \( p < 0.01 \) (t-test, \( n = 9 \)). Therefore, the alkalinity for the entire period was estimated from the salinity series. The DIC, \( V_{\text{arag}} \) and \( V_{\text{calc}} \) were derived from calculated alkalinity and the measured \( pCO_2 \) according to the equations described in [39] with the R package Seacarb [40] using first and second carbonate system dissociation constants for estuarine systems from [41] and the dissociations constants of HF and HSO_4\(^{-} \) of [42,43].

2.4 Statistical modeling

The calculated DIC series of August and September was explained through a statistical model. It considered that (1) the weekly trend of the DIC series is caused by the upwelling of DIC-rich bottom water, (2) the diel variation of the DIC series is caused by primary production and respiration of the meadow. The weekly trends of DIC (Cw) were separated from the diel oscillations (Cd) by a central running average with a time frame of 24 h. Upwelling was assumed to be driven by wind. Cw was fit by a function of wind speed weighted by wind direction (WWt).

\[
WW_t = \text{wind speed}[\text{m s}^{-1}] \times \cos(\text{wind direction}[\text{rad}]+W_{\text{up}}),
\]

With \( W_{\text{up}} \) being a parameter between 0 and \( 2\pi \), corresponding to the wind direction for which wind-induced upwelling is maximal. As examples, if \( W_{\text{up}} = 0 \) or \( 2\pi \), maximal weights are given to northern winds, if \( W_{\text{up}} = \pi \) maximal weights are given to Southern winds. A “left-sided” running integration was performed on the weighted wind time series over a period of \( k_w \):

\[
C_{\text{wt}} = \mu_w + \gamma_w \int_{t-k_w}^{t} WW(t) dt,
\]

Where \( C_{\text{w}} \) is the 24 h running-average series of DIC at time \( t \), \( WW_t \) is the weighted wind speed at time \( t \), \( \mu_w \) and \( \gamma_w \) are regression parameters and \( k_w \) is determined as the integration period yielding the best fit. Parameters in the regression were chosen to minimize residual variance. In the regression between time series, formal hypothesis tests, F-statistics and regression coefficients were not considered as they would be biased due to auto-correlation.

The within-day variations of DIC, Cd were modeled by exponentially weighted running integration of the PAR series over a period of 12 hours with an exponential decay rate \( \lambda \) (s\(^{-1}\)):

\[
Cd_t = \mu_d + \gamma_d \int_{t-12h}^{t} PAR(\tau) \cdot e^{-(\lambda(\tau-t))} d\tau
\]
In a final step, the irradiance and wind sub-models were summed to obtain the final model. The standard deviation of the residuals between model and DIC observation were considered for the parameterization of $W_{up}$, $k_0$, and $\lambda$.

**Results**

3.1 Measures

The three weeks revealed important day/night oscillations of pH and pCO$_2$. The pCO$_2$ increased during the night and decreased during the day reaching minima and maxima at 18:00 and 6:00 respectively. The pH inversely mirrored these pCO$_2$ variations.

In July, only pCO$_2$ and light were recorded. The mean pCO$_2$ of the week was 390 $\mu$atm (mean ± SD), corresponding to the atmospheric CO$_2$. However, diel oscillations were recorded, with a phase shift of ~6 h between light and pCO$_2$ cycles. This shift, common to all three measurement periods is illustrated for July in Fig. 2.

In August, seawater was undersaturated for CO$_2$ compared to the atmosphere with a week mean of 239 $\mu$atm only. However, the diel variations remained comparable to those of July with a mean of 219±24 $\mu$atm (± SD) (Fig. 3, top left panel). The mean ± SD daily maximal and minimal pCO$_2$ values were 374±67 and 155±32, respectively. The maximal amplitude was 416 $\mu$atm. The pH variations mirrored fluctuations in pCO$_2$ with a weekly mean of 8.21, mean diel amplitude of 0.34±0.15 (± SD) and mean daily minimum and maximum of 8.03±0.07 and 8.37±0.08 (± SD). The average salinity and temperature during the week were 14.5±0.3 psu and 19.1±0.7°C, respectively (Fig. 3, lower left panel).

The recordings of September exhibit a strong difference between the beginning (09.09 to 13.09) and the end of the week (13.09 to 16.09). The mean ± SD salinity and temperature during the first three days were 17.8±0.6 psu and 15.9±0.6°C while during the three last days they increased and decreased to 20.7±0.5 psu and 12.4±0.4°C respectively (Fig. 3, lower right panel). This sudden decrease in seawater density is revealing an upwelling occurring in the middle of the week. Consequently, we observed a large discrepancy of the pCO$_2$/pH between those two periods with means of 426 $\mu$atm/8.14 and 1593 $\mu$atm/7.46 respectively (Fig. 2, top right panel). In that second part of the week, oscillations of pCO$_2$ became extreme. The maximal daily amplitudes recorded were 2184 $\mu$atm pCO$_2$ and 1.15 pH units.

The night peaks (mean ± SD) of pCO$_2$ were 2397±425 $\mu$atm, with drops of pH to (mean ± SD) 7.26±0.07. During daytime, minimum pCO$_2$ levels were still much higher than atmospheric concentrations with minima of (means ± SD) 681±211 $\mu$atm and corresponding maxima of pH of 7.77±0.18.

3.2 Carbonate system

In August, means ± SD of the week and day/night variations of DIC were 1609±37 $\mu$mol kg$^{-1}$ and 131±45 $\mu$mol kg$^{-1}$ respectively (Fig. 4 central left panel). The seawater was always supersaturated with respect to both aragonite and calcite with diel means ± SD of 2.2±0.2 and 3.7±0.3 and means ± SD day/night variations of 1.4±0.6 and 2.4±0.9 respectively (see Fig. 4, top left panel).

In September the daily mean ± SD DIC concentrations preceding and during the upwelling event were 1829±43 and 2158±27 $\mu$mol kg$^{-1}$, respectively (Fig. 4 central right panel). The diel variations of DIC were higher before than during the upwelling, with mean ± SD of 259±10 $\mu$mol kg$^{-1}$ against 205±91 $\mu$mol kg$^{-1}$, respectively.

Before the upwelling, the seawater was almost always supersaturated for aragonite and calcite (see Fig. 4 top right panel, 09.09 to 13.09). However at the hours of maximal pCO$_2$ (6:00), aragonite saturation dropped below 1: mean $\Omega_{arag}$ 0.8±0.3 (± SD). Oppositely, seawater reached maxima for $\Omega_{calc}$ and $\Omega_{arag}$ around 18:00. The resulting diel oscillations were of 4.3±0.3 and 2.6±0.2 for calcite and aragonite, respectively.

During the upwelling event the seawater was constantly undersaturated with mean ± SD $\Omega_{calc}$ and $\Omega_{arag}$ of 1±0.5 and 0.6±0.3, exception made of the hours around 18 h, were modest maxima (mean ± SD) of 1.9±0.6 and 1.1±0.4 were reached (see Fig. 4 top right panel, 13.09 to 16.09).

3.3 Model

The DIC calculated from pCO$_2$ and alkalinity and the DIC predicted by the statistical model for August and September are presented in Fig. 4 central panels. The differences between both are presented in Fig. 4, lower panels (residuals). In August, the model predicted a diel DIC mean (± SD), respectively (Fig. 4 top left panel, 09.09 to 13.09). However at the hours of maximal pCO$_2$ (6:00), aragonite saturation dropped below 1: mean $\Omega_{arag}$ 0.8±0.3 (± SD). Oppositely, seawater reached maxima for $\Omega_{calc}$ and $\Omega_{arag}$ around 18:00. The resulting diel oscillations were of 4.3±0.3 and 2.6±0.2 for calcite and aragonite, respectively.

During the upwelling event the seawater was constantly undersaturated with mean ± SD $\Omega_{calc}$ and $\Omega_{arag}$ of 1±0.5 and 0.6±0.3, exception made of the hours around 18 h, were modest maxima (mean ± SD) of 1.9±0.6 and 1.1±0.4 were reached (see Fig. 4 top right panel, 13.09 to 16.09).

The DIC calculated from pCO$_2$ and alkalinity and the DIC predicted by the statistical model for August and September are presented in Fig. 4 central panels. The differences between both are presented in Fig. 4, lower panels (residuals). In August, the model predicted a diel DIC mean (± SD) of 1610±16 $\mu$mol kg$^{-1}$ and diel amplitudes of DIC of 86±14 $\mu$mol kg$^{-1}$ (mean ± SD). The differences between modeled and calculated DIC is 1 $\mu$mol kg$^{-1}$ for the diel average and 44 $\mu$mol kg$^{-1}$ for the diel amplitude.

In September, the model predicted a diel DIC mean (± SD) of 1852±83 $\mu$mol kg$^{-1}$ before the upwelling and 2143±47 $\mu$mol kg$^{-1}$ during the upwelling. The differences with the calculated DIC are 23 $\mu$mol kg$^{-1}$ and 15 $\mu$mol kg$^{-1}$, respectively. Before and during upwelling, the diel amplitudes of DIC predicted by the model are 141±61 $\mu$mol kg$^{-1}$ and 150±107 $\mu$mol kg$^{-1}$ (mean ± SD), respectively. The difference from the calculated DIC is 117 $\mu$mol kg$^{-1}$ and 55 $\mu$mol kg$^{-1}$.

The set of parameters producing the best fitting models are $W_{up} = 3\pi/2$, corresponding to westerly wind, a period of integration $k_0$ of 55 h and $\lambda$ of 0.0025 min$^{-1}$ and 0.001 min$^{-1}$ for August and September respectively. The standard deviation of the residuals between modeled and calculated DIC are 25 $\mu$mol kg$^{-1}$ and 49 $\mu$mol kg$^{-1}$, corresponding to percentages of unexplained variation of 1.6% and 2.5% for August and September respectively (Fig. 3 lower panels). Fig. 5 presents, for September only, the evolution of the model residuals as function of the wind direction ($W_{up}$) and the integration period ($k_0$). Easterly winds as maximal weights produce the worst fitting models while westerly winds the best fitting ones, southerly and northerly winds are intermediate. The parameter $\lambda$ had less influence on the outcome of the model (Fig. 6), increasing its accuracy by a maximum of 2 to
Despite its poor effect on the model, the presence of this parameter is justified by the consideration of the decrease of the day length between August and September (the period of integration of the irradiance is fixed to 12 h in both).

Discussion

In our study, we showed the importance of the diel variations of $\text{pCO}_2$ and pH due to photosynthesis and the importance of the variations of the carbonate chemistry baseline over summer. We demonstrated the interactive effects of upwelling and algal metabolism on the carbonate chemistry with a simple statistical model.

4.1 Inter-weekly dynamics

The $\text{pCO}_2$ of our measurement site exhibited very different weekly trends over the summer. In August, we observed an important and stable $\text{pCO}_2$ undersaturation. This reflects the conditions of the whole Baltic at this period, generated by the succession of blooms of phytoplankton and cyanobacteria [44,45]. In September this stable condition is interrupted by the strengthening of the westerly winds, leading to the upwelling of the water masses isolated until then below the pycnocline. The chemistry of the deep waters of Eckernförde bay are monitored since 1957 and $\text{pCO}_2$ of about 2500 $\text{mol kg}^{-1}$ are yearly observed in summer (Boknis Eck time series: Figure 3. In situ measurements for August and September. $\text{pCO}_2$ (cyan), pH (red) Salinity (black) and temperature (dark blue) recorded in August (left) and September (right). The recording frequency was one minute for $\text{pCO}_2$ and 45 minutes for the 3 other parameters. Symbols in the plots mark the recording events; linkage between the measurements every 45 min was achieved by linear interpolation. Secondary vertical gridlines unit: 6 h. doi:10.1371/journal.pone.0062689.g003

Carbonate Chemistry in Macrophyte Meadows

Figure 4. Calculated and modeled carbonate chemistry. Top panels: Saturation states for calcite (light brown) and aragonite (dark brown) for August (left) and September (right). Dashed line: Saturation threshold. Central panels: Observed (cyan) and modeled (red) DIC for August (left) and September (right). The residuals between model and observations are presented in bottom left and right for August and September respectively. Secondary vertical gridlines unit: 6 h. doi:10.1371/journal.pone.0062689.g004
and submitted to upwelling. We recorded diel oscillations of buffered and eutrophied ecosystem dominated by macrophytes at our study site cumulates those three characteristics: brackish, weakly stratified, low productivity, upwelling areas and estuaries [50]. Our study site is one of the few algal meadows of the Danish islands [18]. Among nearshore meadows in the Baltic region, the nearshore biota in shallow water is exposed to high concentrations during upwelling events. The deep-water pCO$_2$ is presumed to increase in the future by the consequence of higher atmospheric pCO$_2$ and reduced salinity and alkalinity of the Baltic Sea [32]. Melzner et al. (2012) [29] are expecting this deep water body to reach pCO$_2$ of 4000 to 5000 atm and the hypercapnic events of September to increase in magnitude. Besides, the frequency and duration of those upwelling events could also increase as a reinforcement of the westerly winds in the Baltic region is expected with global warming [47].

4.2 Daily oscillations

Diel oscillations, related to photosynthesis and respiration, are superimposed to the week scale dynamics. In normal summer conditions (without upwelling), the mean (± SD) amplitudes of the diel variations were 243±95 µatm CO$_2$ (July, August) and 0.34±0.15 pH units (August only). Such diel variations have already been observed in macrophyte stands worldwide: in seagrass beds of Australia [16], Mediterranean [48] and Zanzibar [19], in tidal rocky-shores of the northeastern Pacific [49] and in algal meadows of the Danish islands [18]. Among nearshore ecosystems, the highest diel variations are occurring in macrophyte dominated ecosystems, upwelling areas and estuaries [50]. Our study site cumulates those three characteristics: brackish, weakly buffered and eutrophied ecosystem dominated by macrophytes and submitted to upwelling. We recorded diel oscillations of 1604.9±795.7 µatm (mean ± SD) during the upwelling event of September. To our knowledge only near-shore mangroves exhibit wider diel variations with 3500 µatm recorded in the Bermuda [10].

However, the importance of these variations is relative, as the fraction of DIC present as CO$_2$ in high pCO$_2$/low pH seawater is higher than in low pCO$_2$/high pH seawater (see Bjerrum plot in e.g. [39]). Thus, in September, the diel variations of CO$_2$ are stronger during the upwelling than before but we observe the opposite for DIC (mean ± SD DIC: 258.6±10.5 µmol kg$^{-1}$ against 205.1±90.7 µmol kg$^{-1}$), possibly revealing a reduction of photosynthesis during the upwelling. This could be explained by an osmotic stress engendered by a rapid increase of salinity [51,52] rather than by the elevated pCO$_2$. Indeed, any increase in DIC and pCO$_2$ is presumed to be beneficial to marine macrophytes [53], even if this have never been formally proven [54].

4.3 Model

We were able to explain the DIC variations of August and September to an accuracy of ±2.50 µmol kg$^{-1}$ with a simple statistical model (three parameters only), based on wind and PAR. The weighing procedure of the wind speed, although simple in its geometric approach, appeared to work very well as an explanation of the multi-day trends. Our model designates westerly winds as responsible for upwelling. This result is slightly different from previous observations and simulations in other regions of the Baltic Sea identifying south-westerly winds as being the most effective [50]. This small discrepancy is certainly due to the east-west orientation of Eckernförde bay compared to the general north-east/south-west orientation of the Baltic Sea. The use of exponentially weighted integration of irradiance as a proxy for primary production was less effective than the similar approach used for the upwelling. Indeed, while the daily DIC means are quite accurately predicted (<20 µmol kg$^{-1}$), the diel amplitude is systematically underestimated by 40 to 70 µmol kg$^{-1}$ DIC. This variation, unexplained by the model, could be due to heterotrophic respiration, air-sea exchange of CO$_2$ [17] or diel variation of alkalinity as discussed in 4.1. However, despite the simplistic nature of our model, it is a first step in the understanding and prediction, in a context of global change, of the carbonate system dynamics in the Baltic nearshore areas.

4.4 Measurement reliability

The accuracy of the post-processed pCO$_2$ data is expected to be ±2.5 µatm for values within the calibrated measuring range of 100–1000 µatm. An additional error can be expected when the pCO$_2$ exceeds the measuring range as in this case the calibration polynomial of the instrument is extrapolated. We expect this error to be ±150 µatm at maximum for the highest pCO$_2$ recorded in September at around 2600 µatm.
Despite our efforts, we did not achieve in the field the pH accuracy necessary to use it as input parameter for the derivation of the carbonate system, as we are able to in laboratory. We estimate the error in pH of the order of 0.01 due to uncorrectable drift during the measurement periods. In the conditions of temporal and spatial variation of the western Baltic, such inaccuracy in pH produces anomalies in derived alkalinity of the order of 10–100 μmol kg⁻¹ at pH/pCO₂ inferior to 7.8/300 μatm and up to 1000 at higher pH/lower pCO₂. For future field studies, Durafets sensors [55] or spectrophotometric sensors [56] represent promising alternatives to glass electrodes, both capable of reaching accuracies of 0.001 to 0.0001 pH units. We had to perform alkalinity titrations and rely on a salinity to alkalinity relationship to achieve the calculations. This method is relevant [57,58] and we estimate the error on the derived DIC to be <15 μmol kg⁻¹ DIC. However, it ignores any changes of alkalinity at constant salinity. This phenomenon is very important in coral and shellfish reefs due to calcification or dissolution [59,60]. However, in macrophyte ecosystems, we expect very marginal diel variations of alkalinity due to photosynthesis and respiration [61].

4.5 Effect on fauna

The daily oscillations of pCO₂ generated by photosynthesis could be of prime importance for calcifiers, creating at daytime periods of high saturation states favourable to CaCO₃ precipitation. Such coupling between photosynthesis and calcification has already been observed in a Hawaiian coral reef by Drupp et al. (2011) [59] and Shamberger et al. (2011) [60] where calcification is already been observed in a Hawaiian coral reef by Drupp et al. (2011) [59] and Shamberger et al. (2011) [60] where calcification is minimal due to planktonic photosynthesis. In that reef, the intensity of the photosynthesis is modulated by wind driven inputs of nutrients from the flume of a neighboring estuary.

In general, studies conducted on western Baltic populations of animals, calcifying or not, tend to demonstrate their tolerance to acidic conditions [62]. Also, the Baltic population of the mussels Mytilus edulis experiences reduced growth and dissolution of the shell only when Ωarag ≤0.15 corresponding to a pCO₂ of ~4000 μatm [31]. Despite this weakening of their shells, the predation by sea stars Asterias rubens and crabs Carcinus maenas maintained at similar pCO₂ are reduced by 56% and 41% respectively [63]. Besides, the growth of the Baltic brackish barnacle Amphibalanus imbornos, competitor of Mytilus for space, remains unaffected at both larval and adult stage at pCO₂ >3000 μatm [64]. Macrophyte meadows are also transient habitats, sheltering early life stages of numerous animal species. These might exhibit more tolerance to ocean acidification as well. As example, the spawns of Baltic herrings Clupea harengus, deposited on macrophyte beds [65], are not affected in their embryonic development by high pCO₂/low pH [66]. The volatility of the carbonate system and the extreme acidic events we observed could have exerted a selective pressure on Baltic populations, explaining the resistance of the local fauna to acidification stress in laboratory. Nevertheless, all the studies quoted previously were conducted under stable elevated pCO₂. None coupled elevated pCO₂ baseline and the diel variations we observed.

Conclusion

Our study represents one of the first attempts of high resolution continuous measurement of the carbonate system in the highly variable environment that is the Baltic Sea’s nearshore. The three weeks showed quite different results related to the dynamics of the whole Baltic carbonate system, to the meteorological condition and to very local processes of photosynthesis and respiration. This study highlights the importance of the natural variations of pCO₂ and pH and emphasizes the consideration of these in ocean acidification studies on nearshore organisms.

Acknowledgments

We thank M. Fischer, C. Hiebenthal, C. Howe, B. Huang Xuan, C. Lieberum, K. Maczassek and C. Pansch, Y. Sawall and the crew of the F. B. Polarfuchs for their field assistance.

Author Contributions

Manuscript improvement: PF PMJH. Conceived and designed the experiments: VS. Performed the experiments: VS. Analyzed the data: VS PF PMJH. Contributed reagents/materials/analysis tools: VS PF PMJH. Wrote the paper: VS.

References

1. NOAA – ESRL (2011). Available: http://www.esrl.noaa.gov/gmd/ccgg/trends.
2. Solomon S, Qin D, Manning M, Chen Z, Marquis M, et al. (2007) IPCC, 2007: Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change, Cambridge: Cambridge University Press. 996 p.
3. Ridgwell A, Zondervan I, Hargreaves J, Bijma J, Lenton T (2007) Assessing the potential long-term increase of oceanic fossil fuel CO₂ uptake due to CO₂ – calcification feedback. Biogeosciences 4: 481–492.
4. Orr JC, Fabry VJ, Aumont O, Bopp L, Doney SC, et al. (2005) Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. Nature 437: 618–620.
5. Feely RA, Sabine CL, Lee K, Berelson W, Kleypas JA, et al. (2004) Impact of anthropogenic CO₂ on the CaCO₃ system in the oceans. Science 305: 362–363.
6. Key RM, Kozyr A, Sabine CL, Lee K, Wanninkhof R, et al. (2004) A global ocean carbon climatology: Results from Global Data Analysis Project (GLODAP), Glob Biogeochem Cy 18: 1–23.
7. Riebesell U, Fabry VJ, Hansson L (2010) Guide to best practices for ocean acidification research and data reporting. Luxembourg: Publication office of the European Union. 260 p.
8. Borges AV, Dellille B, Frankignoulle M (2005) Budgeting sinks and sources of CO₂ in the coastal ocean: Diversity of ecosystems counts. Geophys Res Lett 32: 1–6.
9. Chen CTA, Borges AV (2009) Reconciling opposing views on carbon cycling in the coastal ocean: Continental shelves as sinks and near-shore ecosystems as sources of atmospheric CO₂. Deep Sea Res Pt II 56: 578–590.
10. Anderson AJ, Mackenzie FT (2012) Revisiting four scientific debates in ocean acidification research. Biogeochemistry 9: 893–905.
11. Brasse S, Reimer A, Seifert R, Michaelis W (1999) The influence of intertidal mudflats on the dissolved inorganic carbon and total alkalinity distribution in the German Bight, southeastern North Sea. J Sea Res 42: 93–103.
12. Gattuso JP, Frankignoulle M, Wolfat R (1998) Carbon and carbonate metabolism in coastal aquatic ecosystems. Ann Rev Ecol Syst 29: 405–434.
13. Gazeau F, Borges AV, Barron C, Duarte CM, Iversen N, et al. (2005) Net ecosystem metabolism in a micro-tidal estuary (Randers Fjord, Denmark): evaluation of methods. Mar Ecol Prog Ser 301: 23–41.
14. Rees SA, Opdyke BN, Wilson PA, Henstock TJJ (2006) Significance of Halimeda bioherms to the global carbonate budget based on a geological sediment budget for the Northern Great Barrier Reef, Australia. Coral Reefs 26: 177–188.
15. Kleypas JA, Anthony KRRN, Gattuso JP (2011) Coral reefs modify their seawater carbon chemistry – case study from a barrier reef (Moorea, French Polynesia). Glob Change Biol 17: 3667–3678.
16. Smith S (1981) Marine macrophytes as a global carbon sink. Science 211: 838–840.
17. Gattuso JP, Payri CE, Pichon M, Delsalle B, Frankignoulle M (1995) Primary production, calcification and air-sea CO₂ fluxes of a macroalgal-dominated coral-reef community (Moorea, French Polynesia). J Phycol 31: 729–738.
18. Middelboe AL, Hansen PJ (2007) High pH in shallow-water macroalgal habitats. Mar Ecol Prog Ser 338: 107–117.
19. Sernesi IS, Beer S, Björk M (2009) Seagrass photosynthesis controls rates of calcification and photosynthesis of calcareous macroalgae in a tropical seagrass meadow. Mar Ecol Prog Ser 362: 41–48.
20. Lobban CS, Harrison PJ (1994) Seaweed Ecology and Physiology. Cambridge: Cambridge University Press. 366 p.
21. Beer S, Rhenberg J (1997) The acquisition of inorganic carbon by the seagrass Zostera marina. Aquat Bot 56: 277–283.
22. Badger MR (2003) The roles of carbonic anhydrases in photosynthetic CO₂ concentrating mechanisms. Photosynth Res 77: 83–94.
23. Giordano M, Beardall J, Raven JA (2005) CO₂ concentrating mechanisms in algae: mechanisms, environmental modulation, and evolution. Annu Rev Plant Biol 56: 99–131.
24. Hepburn CD, Pritchard DW, Cormwall CE, McLeod RJ, Beardall J, et al. (2011) Diversity of carbon use strategies in a kelp forest community: implications for a high CO₂ ocean. Glob Change Biol 17: 2488–2497.
25. Barrón C, Marbá N, Duarte CM, Pedersen MF, Linnblad C, et al. (2003) High Organic Carbon Export Precedes Eutrophication Responses in Experimental Rocky Shore Communities. Ecosystems 6: 144–153.
26. Duarte CM, Marbá N, García E, Fourquarean JW, Beggins J, et al. (2010) Seagrass community metabolism: Assessing the carbon sink capacity of seagrass meadows. Glob Biogeochem Cy 24: 1–8.
27. Barró C, Duarte CM (2009) Dissolved organic matter release in a Pseudom不要 almaco meadow. Mar Ecol Prog Ser 374: 73–84.
28. Gray JS, Wu RS, Os YY (2002) Effects of hypoxia and organic enrichment on the coastal marine environment. Mar Ecol Prog Ser 238: 249–279.
29. Melzner F, Thomsen J, Koeve W, Oschlies A, Gutiérrez MA, et al. (2012) Future ocean acidification will be amplified by hypoxia in coastal habitats. Mar Biol. Available: http://www.springerlink.com/index/10.1007/s00227-012-1954-1. Accessed 5 June 2012.
30. Myrberg K, Andrejev O (2003) Main upwelling regions in the Baltic Sea – a statistical analysis based on three-dimensional modelling. Boreal Environ Res 8: 97–112.
31. Thomsen J, Gutiérrez MA, Saphores J, Heinemann A, Trübenbach K, et al. (2009) Littoral Raumboeren – Analysen der Ostseeküste – Schwerpunkte: The seasonal cycle of carbon dioxide in Baltic Sea surface waters. J Mar Syst 22: 53–67.
32. Schneider B (2011) The CO₂ system of the Baltic Sea: Biogeochemical control and impact of anthropogenic CO₂. In: Schermerski G, Hofstedel J, Neumann T, editors. Global Change and Baltic Coastal Zones. Dordrecht: Springer Netherlands. Vol. 1. 33–49.
33. Melzner F, Hofstedel J, Neumann T, editors. Global Change and Baltic Coastal Zones. Dordrecht: Springer Netherlands. Vol. 1. 23–32.
34. Invers O, Romero J, Pizem P (1997) Effects of pH on seagrass photosynthesis: a laboratory and field assessment. Aquat Bot 59: 185–194.
35. Thomas H, Schneider B (1999) The seasonal cycle of carbon dioxide in Baltic Sea surface waters. J Mar Syst 22: 53–67.
36. Thomas H, Schneider B (2011) The CO₂ system of the Baltic Sea: Biogeochemical control and impact of anthropogenic CO₂. In: Schermerski G, Hofstedel J, Neumann T, editors. Global Change and Baltic Coastal Zones. Dordrecht: Springer Netherlands. Vol. 1. 33–49.
37. Bange H, Hansen H, Malen F, Lauf K, Karstensen J, et al. (2011) Boknis Eck time series station (SW Baltic Sea): Measurements from 1957 to 2010. LOICZ inprint: 16–22.
38. Neumann T (2011) Climate Change Impacts on the Baltic Sea. In: Schermerski G, Hofstedel J, Neumann T, editors. Global Change and Baltic Coastal Zones. Dordrecht: Springer Netherlands. Vol. 1. 23–32.
39. Fietzek P, Koertzing A (2010) Optimization of a membrane-based NDIR sensor and sensor development. Oceans 11 MTS/IEEE Conference. 1–9.
40. Fietzek P, Koertzing A (2010) Optimization of a membrane-based NDIR sensor and sensor development. Oceans 11 MTS/IEEE Conference. 1–9.
41. Fietzek P, Kramer S, Esser D (2011) Deployment of the HydroCTM (CO2/CH4) package version 2.3. 3. Software. Available: http://scholar.google.com/scholar?hl=en&btnG=Search&q=intitle:seacarb:...+package+version+2.3+&sa=X&ei=K3jWVoHbK4Qu4wOH3OGQDw&ved=0CCwQ...