Comparison of matrix metalloproteinase 9 and 14 levels in vitreous samples in diabetic and non-diabetic patients: a case control study

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Abstract

Background: MMP-9 plays a prominent role in inflammation and MMP-14 take part in angiogenesis. The objective of this study is to compare MMP-9 and MMP-14 levels between diabetic and non-diabetic patients.

Methods: The patients who scheduled for pars plana vitrectomy were included in our study. Patients are divided into 2 groups: the diabetic group and non-diabetic group. Age, gender, intraocular pressure (IOP), visual acuity (VA) were reported. Color fundus photography, fundus fluorescein angiography, optic coherence tomography (OCT) were performed before and after the operation. MMP-9 and MMP-14 levels in vitreous samples were analyzed with a reader device by ELISA method. Mann–Whitney U test and logistic regressions were used in statistical analysis, p < 0.05 accepted as statistically significant.

Results: 70 eyes of 70 patients who received pars plana vitrectomy were enrolled in the study and divided into 2 groups: 34 patients in the diabetic group, 36 patients in the non-diabetic group. The average age of diabetic patients was 60.14 ± 10.20, and non-diabetic patients was 64.22 ± 11.16, respectively. The average MMP-9 (0.67 ± 0.66 ng/ml) and MMP-14 (0.16 ± 0.45 ng/ml) values in the diabetic group were significantly higher than the average MMP-9 (0.21 ± 0.05 ng/ml) and MMP-14 (0.07 ± 0.02 ng/ml) values in the non-diabetic group (P < 0.01). Also, it was observed that MMP-9 and MMP-14 levels increases as the diabetic disease duration increases. The risk of diabetes incidence increased with high levels of MMP-9 and MMP-14.

Conclusion: Due to the higher levels of MMP-9 and MMP-14 in the pathogenesis of diabetic retinopathy, these proteins may probably be among the therapeutic targets in the prevention and treatment of retinopathy.

Keywords: Matrix metalloproteinase, Diabetic retinopathy, Pars plana vitrectomy

Introduction

Diabetic retinopathy which is progressive and gradually damages the retinal vessels is one of the most common ocular findings in diabetic patients. This condition is asymptomatic in the initial stages of diabetes, but if diabetes is not treated, it may even result in blindness [1]. Diabetic retinopathy is a pathological and complex disease which causes retinal ischemia, increased retinal permeability, retinal neovascularization, macular edema, and microvascular changes [2, 3].

In the treatment of diabetic retinopathy, interventions aiming treatment of diabetes are sometimes insufficient. Thus, alternative treatments are required. The role of molecular mechanisms in the development of diabetic retinopathy should be explained and therapeutic goals might be set [4].

Matrix metalloproteinase (MMP) family is a zinc-dependent protease family with more than 25
members. MMPs family regulates the structure of the extracellular matrix in physiological and pathological processes [5]. It has a central role in organ development and tissue remodeling. Activation of MMPs accelerates apoptosis by a process involving mitochondria through various factors [6]. The MMPs family has a dual role in the development of diabetic retinopathy; it accelerates the apoptosis of retinal capillary cells, pericytes the microvessel regulation and disrupts endothelial cells in the early stages of diabetic retinopathy and it supports the formation of new vessels in the neovascular stage [6, 7].

MMP-9 is the largest and most complex molecule of the MMPs family [8]. The MMPs family regulates the physiological and pathological processes of the subcellular and intracellular region according to its location. MMP-9 is also involved in many inflammatory events with release activation in latent form [9]. MMP-9 is produced through very different stages such as gene transcription, synthesis, secretion, activation, inhibition and glycosylation [10]. In a recent study, MMP-9 gene modified rats were made diabetic and their retinas were examined. Rapid loss of retinal capillary cells was prevented and retinopathy in the early stages of diabetes was revealed [11].

Also, MMP-14, a transcollagenous protein, or type 1 membrane of MMPs family, has an important and critical role in the communication of cells with each other and in the remodeling and invasion of the cells into the extracellular matrix. MMP-14 has direct activity against some of the extracellular matrix proteins, such as gelatin, fibronectin, vitronectin, fibrillar collagen, and aggregates [12]. Studies have shown that MMP-14 plays a very critical and important role in angiogenesis [13]. MMP-14 is found in higher concentrations in many tumor tissues and is involved in tumor progression, angiogenesis, and metastasis [14, 15]. For example, in Kaimal et al’s study of mice with ovarian cancer, MMP-14 was increased in the serum of women with malignant ovarian tumors.

MMPs pharmacological inhibitors make the inhibition by blocking the zinc-dependent site. Doxycycline is a broad spectrum MMP inhibitor that inhibits many MMPs, including MMP-9 [16]. Another inhibitor, bisphosphonates, make MMP inhibition by chelating with the zinc cation site [17]. Because MMP-9 is closely associated with inflammation, non-steroidal anti-inflammatory drugs, such as indomethacin, reduce prostaglandin E2 synthesis and MMP-9 levels and Trastuzumab reduce MMP-9 as confirmed in TOGA trial study [18, 19]. Pharmacological inhibition of MMPs in the development of diabetic retinopathy protects from retinal and choroidal neovascularization [20] and prevents MMP-9-related vascular permeability and inflammation [21]. Also, the use of synthetic MMP inhibitors prevents the induction of proliferative vitreoretinopathy [22]. Inhibition of MMP-9 with COX inhibitors prevents retinal pathologies [23].

Studies about MMP inhibitors and treatment in literature mostly relate to animal models or other in vitro studies, and not to humans. Inhibition of the MMP-9 and MMP-14 proteins may be an alternative for suppressing diabetic retinopathy. Thus, we aimed to compare MMP-9 and MMP-14 levels in diabetic and non-diabetic patients which could be the biomarkers of diabetic retinopathy.

**Methodology**

70 eyes of 70 patients who applied to Atatürk University Faculty of Medicine Ophthalmology Outpatient Clinic with the complaint of visual impairment and were scheduled for surgery with vitreoretinal pathology in the ophthalmological examination were included in our study. A systematic sample size calculation was not done. With the purposeful sampling, all patients who were to be operated on due to the problems mentioned in the study and who gave consent for the study in a 1-year period were included in the sample. All patients in the diabetic group were in the proliferative diabetic retinopathy stage and unresponsive to medical treatment. The patients were recruited based on diabetic status: the diabetic group and non-diabetic group. The study was conducted with the approval of the Ethics Committee of Atatürk University Faculty of Medicine, Clinical Research Ethics Committee, numbered B.30.2.ATA.0.01.00/335. Before the study, the patients were given detailed information about the purpose of the study and the procedures to be performed. The Helsinki declaration was followed and informed consent forms were obtained from the patients.

The pathologies requiring vitreoretinal surgery were grouped within themselves. Detailed medical history was taken from the patients. In the study group for whom vitreoretinal surgery was planned, patients who had received anti-VEGF treatment due to an ocular disease or had any previous ocular surgery and those who used medication for ocular disease were excluded from the study. Patients with pathology in both eyes requiring vitreoretinal surgery were also excluded from the study. The patients in the control group underwent vitreoretinal surgery because of the macular hole (n = 20) and epiretinal membrane (n = 16).

Best corrected visual acuity (BCVA), intraocular pressure (IOP) measurement with applanation tonometry (Haag Streit, Bern, Switzerland), anterior segment examination and fundus examination were performed before and after the operation. Also, HbA1c levels of the patients were measured.
Vitreous material collection was taken under appropriate operating conditions, under general or local anesthesia, during pars plana vitrectomy surgery. After conjunctival incisions made in the lower nasal and upper temporal regions with scissors, 3 cc subtenon anesthetic substance (2 cc Lidocaine and 1 cc Bupivacaine) was applied to all patients undergoing local anesthesia. The conjunctival sac was washed with povidone iodine 5%. Transsclerotomy was performed from the lower temporal, upper temporal and upper nasal 3–4 mm behind the surgical limbus and entered as a pars plana. At this stage, 0.3–0.6 ml of vitreous was taken by vitreous tap method using a vitrectomy cutter just before the infusion cannula was inserted [24]. Then, the vitreoretinal surgery was completed by entering the infusion cannula, light probe and vitrectomy probe. Silicone oil, gas material (C3F8), medical air or balanced salt solution were used as the tamponade. Vitreous samples taken were placed in Eppendorf tubes under sterile conditions. Tubes were frozen at −80°C without waiting to preserve the materials.

MMP-9 and MMP-14 levels in vitreous samples are taken into its special apparatus Sunlong ([Cat No: SL1157Hu for MMP-9, Cat No: SL2924Hu for MMP-14, Sunglong Biotech Co. Ltd, Zhejiang, China]) according to the standard protocol suggested by the manufacturer, Dynex brand automatic ELISA (Enzyme linked Immunosorbent Assay) was analyzed with a reader device (Dynex Technologies Headquarters, Chantilly, USA) by ELISA method. The pre-measurement dilution step recommended in the kit measurement protocol was not performed because the levels of these molecules in vitreous samples were expected to be low. The intra-assay coefficient of variation (CV) value of the kit was below 10% and the inter-assay CV below 12%.

Statistical analysis
Statistical analysis was performed using the SPSS 22.0 (SPSS, Chicago IL, United States) program. Because the study sample was small, the non-parametric Mann–Whitney U test was applied to define whether there is a significant difference of MMP-9 and MMP-14 between the diabetic and non-diabetic groups. P value ≤0.05 was considered to indicate statistical significance.

Results
A total of 70 patients who received pars plana vitrectomy were enrolled in the study and divided into 2 groups: 34 patients in the diabetic group, 36 patients in the non-diabetic group. Table 1 presents the gender and age distributions of the patients. 19 (55.9%) of the diabetic patients were male and 15 (44.1%) were female. The patients in the non-diabetic group consisted of 15 (41.7%) men and 21 (58.3%) women. The mean age of diabetic patients was 60.14 ± 10.20, while the mean age of non-diabetic patients was 64.22 ± 11.16. (Table 1).

The average HbA1c levels were 7.5059 ± 0.727,778 for diabetic group and 5.138,889 ± 0.322,736 for non-diabetic group. The patients in diabetic group were undergone vitreoretinal surgery for tractional retinal detachment (n = 16) and vitreous hemorrhage (n = 18). Vitrectomy indications of non-diabetic group were macular hole (n = 20) and epiretinal membrane (n = 16) (Table 1).

A significant difference was found between diabetic and non-diabetic groups in terms of MMP-9 level (U = 363.500; p = 0.003) and MMP-14 level (U = 404.500; p = 0.015). MMP-9 and MMP-14 values of the patients in the diabetic group were significantly higher than the non-diabetic group (p < 0.01). The average (SD) MMP-9 value x̄ = 0.6677 (±1.28602)

| Table 1 Demographic, clinic and laboratory informations of patients |
|---------------------------------------------------------------|
| **Gender** | **Diabetic (n = 34)** | **Non-diabetic (n = 36)** |
| Women | 15 (44.1%) | 21 (58.3%) |
| Men | 19 (55.9%) | 15 (41.7%) |
| Age | 60.14 ± 10.20 (38–77) | 64.22 ± 11.16 (33–81) |
| HbA1c level | 7.5059 ± 0.727,778 | 5.138,889 ± 0.322,736 |
| Vitrectomy indications | | |
| Tractional retinal detachment | 16 (47.1%) | – |
| Vitreous hemorrhage | 18 (52.9%) | – |
| Macular hole | – | 20 (55.5%) |
| Epiretinal membrane | – | 16 (44.4%) |
in diabetic group was significantly higher than the average (SD) MMP-9 value $\bar{x} = 0.2149 \pm 0.05381$ in non-diabetic group. Also, the average (SD) MMP-14 value $\bar{x} = 0.1641 \pm 0.45261$ in diabetic group was significantly higher than the average (SD) MMP-14 value $\bar{x} = 0.0718 \pm 0.01718$ in non-diabetic group.

There was no significant difference between Tractional Retinal Detachment and Vitreous Hemorrhage as diabetic subgroups in terms of MMP-9 and MMP-14 levels $p > 0.05$ (Table 2). Distribution of MMP-9 and MMP-14 values in diabetic and nondiabetic groups are shown in Figs. 1 and 2, respectively.

### Table 2 Comparison of MMP-9 and MMP-14 levels of diabetic and non-diabetic groups

|                     | Diabetic (Mean ± Std. Dev) (ng/ml) | Non-diabetic (Mean ± Std. Dev) (ng/ml) | Mann-Whitney U | $p$ value | Effect size (95% CI) |
|---------------------|------------------------------------|----------------------------------------|----------------|-----------|---------------------|
| MMP-9               | (0.668 ± 1.28)                     | (0.2149 ± 0.053)                       | 363.500        | 2.922     | 0.003* (0.025–0.881) |
| MMP-14              | (0.1641 ± 0.453)                   | (0.0718 ± 0.017)                       | 404.500        | 2.439     | 0.015* (0.058–0.243) |

|                     | Tractional retinal detachment | Vitreous hemorrhage | Mann-whitney U | $p$ value | Effect size (95% CI) |
|---------------------|-----------------------------|---------------------|----------------|-----------|---------------------|
| MMP-9               | (0.4205 ± 0.315)            | (0.8629 ± 1.692)    | 117.000        | 0.885     | 0.391 (−1.347–0.462) |
| MMP-14              | (0.0894 ± 0.035)            | (0.2231 ± 0.605)    | 141.000        | 0.052     | 0.973 (−0.453–0.186) |

![Fig. 1](image_url) Comparison of MMP-9 levels in diabetic/nondiabetic groups
According to Table 3, a significant positive correlation was found between HbA1c levels and MMP-9 and MMP-14 levels of patients. While HbA1c levels increase, the MMP-9 ($r = 0.661$, $p = 0.000$) and MMP-14 ($r = 0.468$, $p = 0.005$) levels also increase.

When the correlation between diabetes duration and MMP-9 and MMP-14 levels was examined, it was observed that the level of these proteins increased significantly as the disease duration increased ($p < 0.05$). (Table 4).

Binary logistic regression analysis was performed to determine the risk factors of the presence of diabetes and the effect of MMP-9 and MMP-14 levels on being diabetic/nondiabetic group. Age and gender variables also were included in the analysis. The risk of diabetes incidence was found to increase in cases of high MMP-9 ($\beta = -5.853$, $p = 0.006$) and high MMP-14 ($\beta = -31.613$, $p = 0.18$).

**Table 3** Correlation between HbA1c and MMP-9 and MMP-14 levels in diabetic group

|       | HbA1c | MMP-9 | MMP-14 |
|-------|-------|-------|--------|
| HbA1c | 1     | 0.661 | 0.468  |
| MMP-9 | 0.661 | 1     | 0.960  |
| MMP-14| 0.468 | 1     | 1      |

**Table 4** Correlation between the duration of the diabetes and MMP-9 and MMP-14 Levels

|       | MMP-9 | MMP-14 |
|-------|-------|--------|
| Duration | 0.665** | 0.518  |
Discussion
We compared MMP-9 and MMP-14 levels in vitreous between diabetic and non-diabetic groups to analyze novel markers of diabetic retinopathy as diabetes is a risk factor of diabetic retinopathy. Diabetic retinopathy is a systemic and epidemic disease affecting millions of people worldwide [25].

Current treatment includes insulin therapy in Type 1 diabetes and single or multiple combinations of sulfonylurea, diazolidinediones, biguanides, alpha glucosidase inhibitors, meglitinides and insulin in Type 2 diabetes [26]. However, many of these drugs may not treat the microvascular mechanisms that cause diabetic retinopathy. Hyperglycemic vascular stress persists even if blood glucose is brought within normal limits. The main reasons are the advanced glycation end products [27], mitochondrial damage and oxidative stress [28], the epigenetic changes that have occurred [28, 29].

Detected hyperglycemia is seemed to be the main determinant in diabetic retinopathy. However, it is not clear how hyperglycemia produces retinal pathology. In the pathogenesis of diabetic retinopathy, an accelerated apoptosis process is observed in retinal cells, capillary cells, Müller cells, and ganglion cells [30]. Apoptosis in capillary cells leads to characteristic microvascular pathologies in diabetic retinopathy. Accelerated apoptosis also expresses how the destruction of pericyte cells occurs [31]. Blood pressure control and blood sugar regulation are sometimes not possible and adjunctive treatments are required. Thus, molecular mechanisms in the development of diabetic retinopathy should be investigated [4].

MMPs family has a central role in organ development and tissue remodeling. Activation of MMPs accelerates apoptosis by a process involving mitochondria through various factors [6]. MMPs activation has been shown to accelerate the apoptosis process in some studies. MMP-9 activity affects retinal cells and their function in the eyes. MMP activity or physiological disturbances can lead to disruption of the ECM, resulting in ocular damage and malfunction of various cell types in the eyes. The MMPs family has a dual role in the development of diabetic retinopathy; It accelerates the apoptosis of retinal capillary cells, pericytes the microvessel regulation and disrupts endothelial cells in the early stages of diabetic retinopathy and it supports the formation of new vessels in the neovascular stage [6, 7].

In recent studies, MMP-9 gene modified rats which were made diabetic and their retinas were examined, rapid loss of retinal capillary cells was prevented and retinopathy in the early stages of diabetes was revealed [11]. Studies have shown that MMP-14 plays a very critical and important role in angiogenesis [13]. MMP-14 is found in higher concentrations in many tumor tissues and is involved in tumor progression, angiogenesis, and metastasis [14, 15].

MMPs pharmacological inhibitors make the inhibition by blocking the zinc-dependent site. Doxycycline is a broad spectrum MMP inhibitor that inhibits many MMPs, including MMP-9 [16]. Another inhibitor, bisphosphonates, make MMP inhibition by chelating with the zinc cation site [17]. Because MMP-9 is closely associated with inflammation, non-steroidal anti-inflammatory drugs, such as indomethacin, reduce prostaglandin E2 synthesis and MMP-9 levels [32]. Pharmacological inhibition of MMPs in the development of diabetic retinopathy prevents from retinal and choroidal neovascularization [7] and prevents MMP-9-related vascular permeability and inflammation [33]. Also, the use of synthetic MMP inhibitors prevents the induction of proliferative vitreoretinopathy [22]. Inhibition of MMP-9 and MMP-2 with COX inhibitors prevents retinal pathologies [23], but there is no clinical study on the treatment of diabetic retinopathy patients with MMP inhibitors.

In the treatment of diabetic retinopathy, interventions aiming treatment of diabetes are sometimes insufficient. Thus, alternative treatments are required. Inhibition of the MMP-9 and MMP-14 proteins may be an alternative for suppressing diabetic retinopathy. Many different protein levels in the vitreous can be examined to the extent which technology allows. The MMPs family is a protein found in many regions of the body at crucial stages in wound remodeling, inflammation, angiogenesis, and tumor tissues. In the expert opinion presented by Kowluru et al. based on their literature review, MMP-14 is recommended as a biomarker in the treatment of diabetic retinopathy. MMP-9 is often found elevated levels in inflammation, autoimmune degenerative diseases and conditions requiring angiogenesis [34].

In some studies, it has been found that the level of MMPs in diabetic retinopathy and proliferative diabetic retinopathy is higher than the non-diabetic vitreous [35, 36]. The findings of our study are consistent with findings of studies of El-Asrar et al. [24, 35] and Jin et al. [36] in which MMP-9 levels were higher in diabetic patients than non-diabetic patients. In the study of Abu El-Esrar et al., MMP-14 is recommended as a bio-marker, MMP-9 is additionally recommended in our study.

In our study, similar to the study of Giebel et al. [6] in bioequivalent rats samples, MMP-14 level was found to be significantly higher in diabetic group compared to the non-diabetic group in human samples. Kowluru et al. [37] in 2014 showed that the increase in MMP-9 level in diabetes increased oxidative stress in retinal capillary cells via mitochondria and consequently accelerated apoptosis in retinal capillary cells. Retinal mitochondrial
hemostasis continues normally in rats blocked the MMP-9. It has been shown that the development of diabetic retinopathy in these rats was inhibited. Also, regulation of oxidative stress by pharmacological/genetic approaches maintains retinal mitochondrial homeostasis in diabetic rats by improving epigenetic modifications in the MMP-9 promoter region in a study [38].

Our results showed that MMP-9 and MMP-14 enzymes were significantly higher in diabetic patients compared to nondiabetic patients. Also, it was observed that MMP-9 and MMP-14 levels increases as the disease duration increases. However, no difference was found between different diabetic medical conditions. The risk of diabetes incidence increased with high levels of MMP-9 and MMP-14. Also, we found a positive correlation between HbA1c levels and MMP-9 and MMP-14 levels in diabetic group. This result supports that MMP-9 and MMP-14 could be a biomarkers of diabetic retinopathy. Diabetic retinopathy can be suppressed by inhibition of MMP-9 and MMP-14 levels which indicates a new form of treatment. MMP-9 has a very important role in diabetic retinopathy and its increased levels are one of the main factors causing retinal capillary apoptosis. Blocking the cascade at any stage prevents the effects of diabetic retinopathy in theoretical and experimental terms. Jayashree et al., found higher serum MMP-9 levels in Diabetes Mellitus with type 2 patients with retinopathy, as compared with those without retinopathy. While we found high levels of MMP-9 in the vitreous samples of diabetic patients in our study, these researchers found high serum MMP-9 levels in diabetic rats. They conclude that rise in MMP-9, and associated serum markers promote disease progress in DR [39]. In an experimental study by Bhatt and Addepalli, the combination of minocycline and aspirin inhibited MMP-2 and MMP-9, and improved retinal thickness and blood-retinal barrier in diabetic mice. Authors suggest the beneficial effects of treatment with this combination in diabetic neuropathy [23]. Thus, treatments targeting the inhibition of MMP-9 and MMP-14 can be used in the treatment of diabetic retinopathy. Experimental studies with the rats mentioned above, while the first study found the higher serum MMP-9 level, the second study determined the MMP-9 levels by histological examination of the retinal section. Although the common diabetes context, the fact that our study was carried out with human vitreous samples constitutes its distinctive feature. At this point, this study constitutes an important pillar in the efforts to create a biomarker in the treatment of diabetic retinopathy.

There are few studies in the literature examining the relationships between diabetic retinopathy and MMP's family. Also, most of these studies used bioequivalent living vitreous instead of human vitreous. The sample size in this study is higher than in the few studies conducted with patients. While some of the previous studies emphasized MMP-9 or MMP-14, both were analyzed in this study. Also, both MMP-9 and MMP-14 levels were found significantly higher in the diabetic group compared to the non-diabetic group. In the analyses, the effects of some variables such as age and gender were controlled. It was also investigated whether there was a difference in MMP-9 and MMP-14 levels in terms of two disease categories in the diabetic group. These proteins may probably be among the therapeutic targets in the prevention and treatment of diabetic retinopathy.

Following researches involving experimental interventions, levels of MMP-9 and MMP-14 can be used in the diagnosis and assessment of the severity and in treatments that can reduce or slow eye damage caused by diabetic retinopathy by suppressing the level of these proteins that represents the clinical contribution of this study. High MMP-9 and MMP-14 levels are associated with rapid progression of diabetes which can be considered as prognostic factors. Drugs targeting this protein can be used to slow or stop the progression of diabetes that may be useful for clinicians (see Table 5).

| Step | Variables | β     | Standart error | Wald  | p value | Exp(β) | 95% CI for OR |
|------|-----------|-------|----------------|-------|---------|---------|----------------|
| 1    | Gender    | -0.506| 0.608          | 0.693 | 0.405   | 0.603   | 0.183 1.985    |
|      | Age       | 0.056 | 0.029          | 3.752 | 0.053   | 1.057   | 0.999 1.119    |
|      | MMP-9     | -5.839| 2.190          | 7.111 | 0.008   | 0.003   | 0.000 0.213    |
|      | MMP-14    | -32.949| 13.592        | 5.876 | 0.015   | 0.000   | 0.000 0.002    |
|      | Constant  | 1.156 | 1.905          | 0.369 | 0.544   | 3.178   |                 |
| 2    | Age       | 0.052 | 0.028          | 3.361 | 0.067   | 1.053   | 0.996 1.113    |
|      | MMP-9     | -5.853| 2.138          | 7.495 | 0.006   | 0.003   | 0.000 0.190    |
|      | MMP-14    | -31.613| 13.365        | 5.595 | 0.018   | 0.000   | 0.000 0.004    |
|      | Constant  | 1.054 | 1.892          | 0.310 | 0.578   | 2.868   |                 |
Limitations

Other systemic and local diseases that could affect these protein levels were excluded from the study. Thus, we believe the sample size is sufficient to obtain these results. Patients in the nondiabetic group in this study have different diseases. However, these levels could only be measured after vitreoretinal surgery which is operated in some pathological conditions. It is not possible to make these measurements with completely healthy individuals.

Conclusion

In conclusion, this study showed that the diabetic retinopathy have increased MMP-9 and MMP-14 protein levels. These proteins could be the biomarkers of diabetic retinopathy. Thus, MMP-9 and MMP-14 protein levels can be used for the diagnosis and treatment in diabetic retinopathy. In addition to diabetes treatments, MMP-9 and MMP-14 inhibitors can be used to slow down the progression of diabetic retinopathy and to prevent retinal damage. These results should be confirmed by larger studies.

Abbreviations

MMP: Matrix metalloproteinase; anti-VEGF treatment: Anti–vascular endothelial growth factor treatment; BCVA: Best corrected visual acuity; IOP: Intraocular pressure; HbA1c levels: Hemoglobin A1c levels.

Acknowledgements

We would like to thank Atatürk University Faculty of Medicine for enabling us to carry out the laboratory studies of our research.

*This study is adapted from the specialization thesis named “Comparison of Matrix Metalloproteinase 9 and 14 Levels in Vitreous Sample in Diabetic and Non-Diabetic Patients” published in 2019.

Author contributions

We confirm that all authors contributed equally to this research. All authors approved the final manuscript and the order of authors listed in the manuscript. All the authors read and approved the final manuscript.

Funding

The authors received no specific funding for this research.

Availability of data and materials

The data of this study are available on request from the corresponding author. All the authors read and approved the final manuscript and the order of authors listed in the manuscript. All the authors received no specific funding for this research.

Declarations

Ethics approval and consent to participate

The study was approved by the Research Ethics Committee of Atatürk University (07–18). The study was conducted in accordance with the regulations of Helsinki Declaration. Written informed consent form was taken from all participants in the study.

Consent for publication

I, the corresponding author, give my consent for the publication of information, which can include me, to be published in the International Journal of Retina and Vitreous.

Competing interests

The authors have no conflicts of interests to disclose.

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Received: 25 March 2022 Accepted: 4 June 2022

Published online: 21 June 2022

References

1. Williams R, Airey M, Baxter H, Forrester J, Kennedy-Martin T, Girach A. Epidemiology of diabetic retinopathy and macular edema: a systematic review. Eye (Lond). 2004;18(10):963–83.

2. Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. Diabetes Care. 2004;27(5):1047–53.

3. Klein R, Klein BE, Moss SE, Cruickshanks KJ. The Wisconsin Epidemiologic Study of diabetic retinopathy. XIV. Ten-year incidence and progression of diabetic retinopathy. Arch Ophthalmol. 1994;112(9):1277–8.

4. Brownlee M. The pathobiology of diabetic complications: a unifying mechanism. Diabetes. 2005;54(6):1615–25.

5. Yang Y, Hill JW, Rosenberg GA. Multiple roles of metalloproteinases in neurological disorders. Prog Mol Biol Transl Sci. 2011;99:241–63.

6. Giebel SJ, Menicucci G, McGuire PG, Das A. Matrix metalloproteinases in early diabetic retinopathy and their role in alteration of the blood-retinal barrier. Lab Invest. 2005;85(5):597–607.

7. Kowluru RA, Zhong Q, Santos JM. Matrix metalloproteinases in diabetic retinopathy: potential role of MMP-9. Expert Opin Investig Drugs. 2012;21(6):797–805.

8. Malermud CJ. Matrix metalloproteinases (MMPs) in health and disease: an overview. Front Biosci. 2006;11:1696–701.

9. Ovechkin AV, Tyagi N, Rodriguez WE, Hayden MR, Moshal KS, Tyagi SC. Role of matrix metalloproteinase-9 in endothelial apoptosis in chronic heart failure in mice. J Appl Physiol. 2005;99(6):2398–405.

10. Van den Steen PE, Dubois B, Nellissen I, Rudd PM, Dewer RA, Opdenakker G. Biochemistry and molecular biology of gelatinase B or matrix metalloproteinase-9 (MMP-9). Curr Rev Biochem Mol Biol. 2002;37(6):375–536.

11. Klein R, Klein BE, Moss SE, Davis MD, DeMets DL. The Wisconsin epidemiologic study of diabetic retinopathy. II. Prevalence and risk of diabetic retinopathy when age at diagnosis is less than 30 years. Arch Ophthalmol. 1984;102(4):520–6.

12. Chun TH, Sabeh F, Ota I, Murphy H, McDonagh KT, Holmbeck K, et al. MT1-MMP-dependent neovessel formation within the confines of the three-dimensional extracellular matrix. J Cell Biol. 2004;167(4):757–67.

13. Collen A, Hanemaaijer R, Lupu F, Quax PH, van Lent N, Grimbinger J, et al. Membrane-type matrix metalloproteinase-mediated angiogenesis in a fibrin-collagen matrix. Blood. 2003;101(5):1810–7.

14. Sakamoto T, Seiki M. Integrated functions of membrane-type 1 matrix metalloproteinase in regulating cancer malignancy: Beyond a proteinase. Cancer Sci. 2017;108(6):1095–100.

15. Kaimal R, Aljumaily R, Tressel SL, Pradhan RV, Covic L, Kuliopulos A, et al. Membrane-type matrix metalloproteinase-mediated angiogenesis in human monocytes. Pharmacology. 2009;83(1):18–25.

16. Bang YJ, Van Cutsen E, Feyereislova A, Chung HC, Shen L, Sawaki A, et al. Trastuzumab in combination with chemotherapy versus chemotherapy
alone for treatment of HER2-positive advanced gastric or gastro-oesophageal junction cancer (ToGA): a phase 3, open-label, randomised controlled trial. Lancet. 2010;376(9742):687–97.

20. Samtani S, Amaral J, Campos MM, Fariss RN, Becerra SP. Doxycycline-mediated inhibition of choroidal neovascularization. Invest Ophthalmol Vis Sci. 2009;50(11):5098–106.

21. Navaratna D, McGuire PG, Menicucci G, Das A. Proteolytic degradation of VE-cadherin alters the blood-retinal barrier in diabetes. Diabetes. 2007;56(9):2380–7.

22. Özdemir U, Mach-Hoface B, Cheng L, Chaidhawangul S, Keefe K, McDermott CD, et al. The effect of primomastat (AG334D), a potent inhibitor of matrix metalloproteinases, on a subacute model of proliferative vitreoretinopathy. Curr Eye Res. 2000;20(6):447–53.

23. Bhatt LK, Addepalli V. Attenuation of diabetic retinopathy by enhanced inhibition of MMP-2 and MMP-9 using aspirin and minocycline in streptozotocin-diabetic rats. Am J Transl Res. 2010;2(2):181–9.

24. Abu El-Asrar AM, Mohammad G, Allegaert E, Ahmad A, Siddiquei MM, Van den Eynde K, Moussa A, et al. Relationship between vitreous levels of matrix metalloproteinases and vascular endothelial growth factor in proliferative diabetic retinopathy. PLoS ONE. 2013;8(3):e63018.

25. Crews JE, Chou CF, Sekar S, Saadallah IN, Ahmed MA, Siddiquei MM, et al. Differential roles of vascular endothelial growth factor in proliferative diabetic retinopathy. Invest Ophthalmol Vis Sci. 2014;55(9):5653–60.

26. Jonas D, Van Scoyoc E, Geral M, Wines R, Arick H, Triplett M, et al. Drug class reviews. Drug class review: newer diabetes medications, TZDs, and combinations: final original report. Portland (OR): Oregon Health and Science University. 2011.

27. Chilelli NC, Burlina S, Lapolla A. AGES, rather than hyperglycemia, are responsible for microvascular complications in diabetes: a ‘glycoxidation-centric’ point of view. Nutr Metab Cardiovasc Dis. 2013;23(10):913–9.

28. Kowluru RA. Mitochondria damage in the pathogenesis of diabetic retinopathy and in the metabolic memory associated with its continued progression. Curr Med Chem. 2013;20(26):3226–33.

29. Perrone L, Matrone C, Sinha LP. Epigenetic modifications and potential new treatment targets in diabetic retinopathy. J Ophthalmol. 2014;2014:789120.

30. Feit-Leichman RA, Kinoshi R, Takeda M, Fan Z, Mohr S, Kern TS, et al. Vascular damage in a mouse model of diabetic retinopathy: relation to neuronal and glial changes. Invest Ophthalmol Vis Sci. 2005;46(11):4281–7.

31. Armulik A, Genové G, Betsholtz C. Pericytes: developmental, physiological, and pathological perspectives, problems, and promises. Dev Cell. 2011;21(2):193–215.

32. Hua Y, Xue J, Sun F, Zhu L, Xie M. Aspirin inhibits MMP-2 and MMP-9 expressions and activities through upregulation of PPARalpha/gamma and TIMP gene expressions in ox-LDL-stimulated macrophages derived from human monocytes. Pharmacology. 2009;83(1):18–25.

33. Reddy AB, Ramana KV, Srivastava S, Bhattacharya A, Srivastava SK. Aldose reductase regulates high glucose-induced ectodomain shedding of tumor necrosis factor (TNF)-alpha via protein kinase C-delta and TNF-alpha converting enzyme in vascular smooth muscle cells. Endocrinology. 2009;150(1):63–74.

34. Shibuya M. Differential roles of vascular endothelial growth factor receptor-1 and receptor-2 in angiogenesis. J Biochem Mol Biol. 2006;39(5):469–78.

35. Abu El-Asrar AM, Mohammad G, Nawaz MI, Siddiquei MM, Van den Eynde K, Moussa A, et al. Matrix metalloproteinases and vascular endothelial growth factor in proliferative diabetic retinopathy. PLoS ONE. 2013;8(12):e85857.

36. Jin M, Kashiwagi K, Izuka Y, Tanaka Y, Imai M, Tsukahara S. Matrix metalloproteinases in human diabetic and nondiabetic vitreous. Retina. 2001;21(1):28–33.

37. Kowluru RA, Santos JM, Zhong Q, Sirt1, a negative regulator of matrix metalloproteinase-9 in diabetic retinopathy. Invest Ophthalmol Vis Sci. 2014;55(9):5653–60.

38. Kowluru RA, Shan Y. Role of oxidative stress in epigenetic change of MMP-9 promoter in the development of diabetic retinopathy. Graefes Arch Clin Exp Ophthalmol. 2017;255(5):955–62.

39. Jayashree K, Yasar M, Senthilkumar GP, Ramesh Babu K, Mehalingam V, Mohanraj PS. Cirulating matrix modulators (MMP-9 and TIMP-1) and their association with severity of diabetic retinopathy. Diabetes Metab Syndr. 2018;12(6):869–73.