The impact of COVID-19 on the male genital tract: A qualitative literature review of sexual transmission and fertility implications

Pierangelo Verrienti, Gianmartin Cito, Fabrizio Di Maida, Riccardo Tellini, Andrea Cocci, Andrea Minervini, Alessandro Natali
Department of Urology, Careggi Hospital, University of Florence, Florence, Italy

Introduction

The male genital tract is a target organ for several viral infections, with potential detrimental consequences from the standpoint of individuals, their offspring, and demographics [1]. A new type of coronavirus (severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)) has rapidly spread among humans, causing a worldwide pandemic. This virus causes a disease known as coronavirus disease 2019 (COVID-19), which is characterized by acute bilateral interstitial pneumonia and severe acute respiratory syndrome, which may lead to death in a short time [2].

In order to contain the spread of this infection and to properly inform the public about appropriate safety measures, it is pivotal to identify all the possible pathways of viral transmission. Besides the typical well-known respiratory route of transmission through droplets, the oral-fecal route due to gastrointestinal viral involvement should also be considered. Moreover, sexual transmission and the immune privilege of the male reproductive tract may play a significant role in the spread of an infectious disease.

Some prior studies explored the potential sexual and vertical transmission of previous epidemic coronaviruses, focusing on possible viral effects on fetuses and newborns in infected women [3,4]. Therefore, it is crucial to provide proper counseling for couples with regard to their sexual behaviors, particularly in asymptomatic or recovered individuals.

Based on our knowledge of other single-stranded RNA viruses, such as Zika and Ebola, which have also been responsible for recent widespread epidemic viral infections [5,6], it could be hypothesized—despite the clear differences between SARS-CoV-2 and the above-mentioned viruses—that the male genitalia could represent...
a possible reservoir for SARS-CoV-2, leading to its sexual transmission by asymptomatic or cured men [7]. Indeed, Zika and Ebola viruses are able to cause viremia, overcome the blood-testis barrier, and ultimately become present in semen [5-7]. The presence of Zika virus in human semen has been widely demonstrated for over 188 days after primary infection, even though it is cleared in the serum after the initial viral symptoms subside [8]. As a result, an immune response might be activated, leading to inflammatory processes such as orchitis, resembling what happens in human immunodeficiency virus (HIV), hepatitis B virus (HBV), and mumps infection [9]. However, extensive information about its potential impact on male fertility is currently lacking. Ebola virus can also persist in bodily fluids during recovery, enabling viral transmission through semen [6]. Preliminary results have shown that male survivors of Ebola virus disease can have virus-positive semen for up to 9 months after the acute infection, as determined by reverse-transcription polymerase chain reaction (RT-PCR) [6]. However, it is unknown with absolute certainty how long the virus can persist in the seminal fluid. Likewise, there are no conclusive data about the medium- or long-term consequences of Ebola virus on male fertility.

In this light, several case series have assessed the possible role of SARS-CoV-2 in male fertility impairment, with inconsistent results; the findings are also difficult to interpret because transmission through droplets can clearly occur during intimate sexual contact [10,11]. As such, in the current paper, we attempted to thoroughly explore the impact of SARS-CoV-2 on male fertility and sexual health by conducting a non-systematic review of the literature.

Search strategy

Medline, Embase, and Scopus Library were searched to identify studies published between January 2020 and March 2021 that investigated the impact of SARS-CoV-2 on male fertility. We conducted a non-systematic critical assessment of the current literature focusing on the impact of SARS-CoV-2 on reproductive hormones and fertility parameters, as well as its presence in seminal fluid and the testis. The following string terms were used: (“COVID-19” OR “SARS-CoV-2”) AND (“reproductive hormones” OR “fertility” OR “semen” OR “testes”). We report a detailed description of the findings extracted from the included studies.

Pathophysiology and the molecular mechanism of COVID-19 cell entry

The entrance of SARS-CoV-2 into target host cells has been shown to be mediated by the interaction between the surface spike viral protein (S) and the angiotensin-converting enzyme 2 receptor (ACE2), employing the cellular transmembrane protease serine 2 (TMPRSS2). Therefore, the co-expression of these is needed within the same cell to allow viral entry [12,13]. As a protease, TMPRSS2 is essential for SARS-CoV-2 to penetrate cells, but many cells in the male genital tract do not express these proteins simultaneously with ACE2 [13-15]. However, other authors demonstrated that there was almost no overlapping gene co-expression (<1%) in the human testicle [12]. The S1 site contains a receptor-binding domain, which then links to ACE2 and facilitates viral entry into the cell. ACE2 is expressed in various organs, including type II alveolar cells of lungs, heart, kidneys, and intestines [16]. In addition, ACE2 seems to be constitutively expressed in the testes, predominantly in spermatogonia, Sertoli cells and Leydig cells [17,18]. Most recently, Vishv karma and Rajender [19] found the presence of ACE2 transcripts in recent transcriptome sequencing of human spermatozoa, further validating its expression in germ cells. The physiological functions of ACE2 in Leydig cells include the regulation of testosterone production and local balance in interstitial fluid volume by modulating the conversion of angiotensin II to angiotensin I. In the COVID-19 infection process, ACE2 receptors are saturated by binding with the virus, giving rise to the increased availability of angiotensin II, which cannot be converted.

As confirmed by recent studies, the levels of ACE2 transcripts are extremely high in the normal adult testes [13]. Spermatogonia that express ACE2 have higher levels of genes associated with viral reproduction and transmission, but lower levels of genes related to spermatogenesis, than ACE2-negative spermatogonia [20]. This has generated the idea that COVID-19 could have possible implications for the male genital tract, which could be considered a potential target for SARS-CoV-2, as well as a possible reservoir of infection. In this regard, it might be hypothesized that the virus could have a negative influence on male gonads, resulting in spermatogenetic injury and endocrine dysregulation.

In 2006, Xu et al. [11] demonstrated a wide range of histological injuries to germ cells and spermatogenesis, through a complex inflammatory infiltrate, in males who died from SARS-CoV complications, implying a correlation between the disease and subsequent reproductive impairment. SARS-CoV, as well as HIV, HBV, and mumps, can trigger orchitis as a possible complication due to body temperature increase and the virus-induced autoimmune response [9]. Indeed, previous studies, have reported that apoptosis of meiotic germ cells occurs at high temperatures [21]. Therefore, high fevers related to COVID-19 could cause indirect damage to testicular function, leading to temporary sub-fertility.
COVID-19 and male sexual health: the state of the art

1. Impact on reproductive hormones

Several studies have investigated the impact of SARS-CoV-2 on male reproductive hormones (Table 1 [22-24]). Luteinizing hormone and prolactin concentrations appear to be significantly increased in COVID-19-infected men of reproductive age, compared to healthy age-matched controls, while follicle-stimulating hormone and estradiol levels seem to be comparable between these groups [22]. Thus, Leydig cells would seem to be more sensitive to viral attack, with possible implications regarding hypogonadism, while Sertoli cells may be more resistant. Schroeder et al. [23] reported that most patients with COVID-19 had low testosterone and dihydrotestosterone levels. Moreover, total testosterone levels have been shown to be inversely proportional to C-reactive protein (CRP) levels in COVID-19-recovered patients [24]. On the basis of these findings, we could speculate that hypogonadism may be related to the severity of COVID-19 infection, most likely due to a body temperature increase and virus-induced autoimmune response, with consequent apoptosis of meiotic germ cells [9,21]. Moreover, since the co-expression of ACE2 and TMPRSS2 occurs only in a small percentage of prostate epithelial cells, prostate infection is unlikely to explain the dysregulation of steroidogenesis mediated by SARS-CoV-2 infection [13,15,17]. Furthermore, the sexual hormonal alterations observed in SARS-CoV-2 patients may reflect the global stress response [9,21]. However, to date, very few scientific studies have investigated the presence of SARS-CoV-2 in the seminal fluid, or even within the testicles, as reported below.

2. Presence in seminal fluid and testis

A research team focused on this topic analyzed semen samples from a group of 34 men about 1 month after the diagnosis of COVID-19, and reported that viral RNA did not seem to be present [12]. Consistently with this finding, other authors found no presence of viral RNA by RT-PCR in semen or urine samples of patients with laboratory-confirmed SARS-CoV-2 infection, either symptomatic or asymptomatic [25-30]. A possible explanation for this may be the low likelihood of the virus crossing the blood-testis barrier. Indeed, SARS-CoV-2 appears to be present in the blood in only 1% of cases [31], which would suggest a low probability of developing viremia.

Conversely, Li et al. [32] enrolled a cohort of 38 SARS-CoV-2-positive patients for semen testing. Among them, 23 had achieved clinical recovery, while 15 were in the acute stage of infection. Six patients (15.8%) had results positive for SARS-CoV-2, including 4 of the 15 patients (26.7%) in the acute stage of infection and 2 of the 23 patients (8.7%) who were recovering. However, the small sample size makes these findings difficult to generalize universally. Furthermore, the single-center study design and the lack of accurate data about the baseline study cohort might have introduced non-negligible statistical bias, thus meaningfully undermining the reliability of the reported findings.

Another further study explored the presence of SARS-CoV-2 RNA in semen and testicular tissue of 12 men suffering from COVID-19 with symptoms of mild severity. In 10 patients with negative pharyngeal swabs, the presence of viral RNA, tested by RT-PCR, was not found in the seminal fluid. Moreover, a patient with a positive swab test was negative for the presence of viral RNA in the semen. Only one...
patient in the acute phase of disease, who died from COVID-19–related complications, underwent a testicular biopsy, which did not demonstrate the presence of the virus in the tissue [33]. In line with these findings, a recent paper by Flaifel et al. [34] analyzed specimens of the testes and epididymis from a series of 10 SARS-CoV-2-positive autopsies. Although the autopsies all showed the presence of SARS-CoV-2 in the respiratory tract, the testicular samples tested by RT-PCR were all negative. However, seven of 10 cases showed significant seminiferous tubular injury, including nuclear fragmentation of spermatocytes, elongation of spermatids, and vacuolization of the Sertoli cells, while 1 case showed an increased mononuclear inflammatory infiltrate (CD8+ dominant) in the interstitial space, compatible with orchitis [34]. In their analysis, the acute morphological changes reported could have been related to oxidative stress and microthrombosis in the testicular vasculature. In addition, the absence of the virus in the testes suggests that direct injury by SARS-CoV-2 infection is unlikely [34].

The characteristics of the studies exploring the presence of SARS-CoV-2 in semen, urine, and testicular tissue are reported in Table 2 [12,22,26-30]. The challenge is to understand whether SARS-CoV-2 can directly infect the testicles, as the main target of the male genital tract, even during the acute phase of the disease. To better understand this issue, it could be useful to have more data from autopsies of COVID-19 patients, since we do not know if SARS-CoV-2 is present only in the seminal fluid, binds to spermatozoa, or can even integrate into the cell genome. More recently, Yang et al. [35] examined postmortem testes from 12 COVID-19 patients using RT-PCR. The authors found positive viral detection only in 1 patient with a high viral load. Conversely, concerning morphological changes, the testes from COVID-19 patients exhibited significant seminiferous tubular injuries, reduced Leydig cells, and mild lymphocytic inflammation. Alternatively, hyperthermia, hypoxia, and steroid use could have played a crucial role in testicular damage.

A recent paper by Achua et al. [36] explored the presence of SARS-CoV-2 in the testes of 6 COVID-19-positive autopsies and 3 negative men by hematoxylin and eosin (H&E) histomorphology and transmission electron microscopy (TEM). They reported that 3 (50%) SARS-CoV-2 biopsies had normal spermatogenesis, while the other 3 had impaired spermatogenesis on H&E histomorphology. Moreover, TEM showed SARS-CoV-2 in testis tissue of 1 positive autopsy case and a biopsy obtained from a live patient who was previously diagnosed with SARS-CoV-2 and subsequently seroconverted. In addition, immunofluorescence-stained slides from the positive men demonstrated an association between increased quantitative ACE-2 levels and impairment of spermatogenesis [36]. These findings, especially the inverse association between ACE-2 receptor levels and spermatogenesis, may suggest that the testes could be a target of SARS-CoV-2, ultimately providing a possible mechanism of post-COVID-19 infertility. However, that study has several limitations, the main ones being the lack of a detailed analysis of seminal parameters and virus detection by RT-PCR, making these results unreliable.

Although the risk of the presence of SARS-CoV-2 in semen appears to be low, future studies need to focus on whether complete viral particles can be observed in semen and the possibility of sexual transmission. Furthermore, the embryology community needs to establish the implications of SARS-CoV-2 for assisted reproductive technology and whether the virus can be removed by sperm washing techniques, as with HIV or hepatitis C virus. Moreover, data about how long the virus remains detectable in the seminal fluid, its ability to actively replicate, and its potential for sexual transmission are still lacking. As such, there are still many open questions to be discussed.

3. Impact on fertility parameters

Due to the reported high expression of ACE2 in seminiferous tubule cells, spermatogonia, adult Leydig cells, and Sertoli cells of the human testis, and TMPRSS2 expression in prostate epithelial cells, SARS-CoV-2 may be involved in the dysregulation of steroidogenesis [17,37]. These findings imply that SARS-CoV-2 infection may have risks for the male reproductive system in terms of impaired spermatogenesis.

In detail, sperm cells express all types of ACEs (1–7) and recent publications reported angiotensin II type 1 receptor (AT1R) and angiotensin II type 2 receptor (AT2R) expression in sperm, suggesting that sperm cells could act as a direct target of SARS-CoV-2 infection [12,13,16-20,38,39]. In this scenario, SARS-CoV-2 infection would be expected to impact ACE2 activity, leading to an increase in the availability of angiotensin II in sperm cells, stimulating the acrosome reaction [13,18,19,37,38]. This may lead to premature acrosomal exocytosis and sperm senescence [40]. Furthermore, angiotensin II may affect sperm fertilization and motility by stimulating AT1R and AT2R [39].

Although Holtmann et al. [28] in their aforementioned study did not detect SARS-CoV-2 RNA in the semen samples of recovered or acutely infected patients, they found significant impairment of sperm quality (in terms of sperm concentration and motility) in patients with a moderate infection, as compared with men recovered from mild infection and a control group. Similarly, Ma et al. [22,25] reported altered semen quality in four patients (33.3%), who showed decreased sperm concentration and motility and a higher DNA fragmentation index.

A recent paper from Zhang et al. [41] failed to isolate SARS-CoV-2 in prostatic secretions of patients with active disease; however, they described a significant increase of inflammatory markers such as CRP and interleukin-6 in these biologic samples. These results, taken together, suggest a potential impact of SARS-CoV-2 on semen param-

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| Study             | Study design               | No. of cases | Patient group | Age (yr) | Clinical stage at time of sample collection | Urogenital symptoms or urological disease | No. of cases with semen/urine/tissue samples | Positive semen samples | Positive urine samples | Positive tissue samples | Time between disease onset/diagnosis and samples collection | Correlation with NP swab at the time of collection |
|-------------------|----------------------------|--------------|---------------|----------|---------------------------------------------|------------------------------------------|-----------------------------------------------|-----------------------|----------------------|-----------------------|----------------------------------------------------------|--------------------------------------------------|
| Li et al. [32]    | Monocentric cohort         | 38           | Recovery, 23 (60.5) | NA       | Recovery                                    | 1 (4.4)                                 | 23                                            | 2 (8.7)               | NA                   | NA                    | 27 day (12–64)                                             | Positive NP swab, 9 (52.9)                               |
| Ning et al. [30]  | Monocentric cohort         | 112          | Mild disease, 40 (35.7) | 55.5 (23–83) | Recovery                                  | 3/112 (2.7)                             | 17                                            | 0                     | NA                   | NA                    | None                                                       |                                                   |
| Holtmann et al. [28] | Monocentric              | 34           | Mild/moderate disease, 18 case | 42.2 ± 9.91 | Recovery                                    | 1/18                                    | 18                                            | 0                     | NA                   | NA                    | 43.5 ± 6.2 day                                              | None                                               |
| Pan et al. [12]   | Multicentric case control | 14           | Control, 14 case                          | NA       | Recovery                                    | 14                                       | 0                                             | NA                   | NA                   | NA                    | 47 ± 5.3 day                                               | None                                               |
| Paoli et al. [26] | Case report                | 1            | Mild disease                          | 37 (31–49) | Recovery                                    | 0                                       | 1                                             | NA                   | NA                   | NA                    | 8 day                                                      | None                                               |
| Song et al. [33]  | Monocentric cohort         | 34           | Mild/moderate disease, 12; severe disease, 1 | 22–38 | Recovery                                    | 0                                       | 12                                            | 0                     | NA                   | NA                    | None                                                       | Positive NP swab, 1                                 |
| Rawlings et al. [27] | Monocentric cohort       | 6            | NA                                   | 28–45 | Recovery                                    | 0                                       | 6                                             | 0                     | NA                   | NA                    | 12 day (6–17)                                              | Positive NP swab, 3 (50)                              |
| Ma et al. [22]    | Multicentric case control | 12           | Mild disease, 1/12                     | 31.5 (25–46) | Recovery                                    | 0                                       | 12                                            | 0                    | NA                   | NA                    | 75.5 day (56–109)                                          | None                                               |
| Kayaaslan et al. [29] | Monocentric cohort      | 56           | Mild/moderate disease, 11/12            | 33.5 (18–54) | Acute stage                                | 0                                       | 16                                            | 0                     | NA                   | NA                    | 1 day (0–7)                                                | Positive NP swab, 5                                 |
| Yang et al. [35]  | Monocentric cohort         | 12           | Severe disease                        | 65 (42–87) | Died                                       | 0                                       | 10                                            | NA                   | NA                   | 1                     | 1 hr                                                   | NA                                                 |
| Flaifel et al. [34] | Monocentric cohort       | 10           | Severe                                | 49.5 (22–83) | Died                                       | 0                                       | 10                                            | 0                    | NA                   | 0                     | 15 day (7–27)                                              | Positive 10%                                      |

Values are presented as number (%), median (interquartile range), or mean±standard deviation. All patients were diagnosed with positive RT-PCR results from NP swabs and/or serum IgM/IgG.

SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; NP, nasopharyngeal; NA, not applicable; RT-PCR, reverse-transcription polymerase chain reaction; Ig, immunoglobulin.

*Urogenital symptoms reported only in patients with severe disease; †Alteration of semen quality reported in 4 patients (33.3%); ‡Since confirmed death; §Morphological alteration of testes reported in 7 patients, 1 case of orchitis detected; ††RT-PCR on lung autopsies.
eters and ultimately on male reproductive capability. Therefore, given the widespread SARS-CoV-2 pandemic, direct or indirect fertility decline post-COVID-19 seems to be a possible and significant issue, particularly in the hardest-hit countries [42,43]. Given the relevance of these findings, potentially impacting the world’s demographics in the near future, we need further and stronger evidence on the real impact of SARS-CoV-2 on male reproductive health.

**Conclusion**

To date, there is very limited evidence reporting the presence of SARS-CoV-2 in semen and testicular samples. The studies available do not allow us to draw definitive conclusions or exclude the possibility of viral sexual transmission. Despite the small sample size and several selection biases, the male genital tract represents a potentially susceptible organ to viral infection. However, the real impact of SARS-CoV-2 on male reproductive function still remains to be fully determined.

**Conflict of interest**

No potential conflict of interest relevant to this article was reported.

**ORCID**

Pierangelo Verrienti https://orcid.org/0000-0002-7596-5961
Gianmartin Cito https://orcid.org/0000-0001-7526-4025
Fabrizio Di Maida https://orcid.org/0000-0003-1885-4808
Riccardo Tellini https://orcid.org/0000-0003-2189-5352
Andrea Cocci https://orcid.org/0000-0003-0138-6294

**Author contributions**

Conceptualization: PV, GC, AN. Data curation: PV, GC, FDM, RT. Formal analysis: PV, GC, RT. Methodology: PV, GC, FDM, RT. Project administration: AC, AM, AN. Visualization: AC, AM, AN. Writing—original draft: PV, GC, FDM. Writing—review & editing: PV, RT, AC, AM, AN.

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