Th2 cytokine profile in appendicular lavage fluid is compatible with allergy as an etiology for acute appendicitis

Nuno Carvalho\textsuperscript{1,2}, André Barros\textsuperscript{4}, Hélder O. Coelho\textsuperscript{3}, Catarina F. Moita\textsuperscript{4}, Ana Neves-Costa\textsuperscript{4}, Filipe C. Borges\textsuperscript{1}, Luís F. Moita\textsuperscript{4,5} Paulo M. Costa\textsuperscript{1,2}

\textsuperscript{1}Serviço Cirurgia Geral, Hospital Garcia de Orta, Almada, Portugal
\textsuperscript{2}Faculdade de Medicina, Universidade de Lisboa, Portugal
\textsuperscript{3}Serviço Anatomia Patológica, Hospital Garcia de Orta, Almada, Portugal
\textsuperscript{4}Innate Immunity and Inflammation Laboratory, Instituto Gulbenkian de Ciência, Oeiras, Portugal
\textsuperscript{5}Instituto de Histologia e Biologia do Desenvolvimento, Faculdade de Medicina, Universidade de Lisboa, Portugal

Correspondence should be addressed to L.F.M. (lmoita@igc.gulbenkian.pt)

Abstract

Acute appendicitis is the most frequent surgical abdominal emergency, but its etiology remains poorly understood. Histological examination of the appendix, following its removal due to acute appendicitis, consistently shows features in common with bronchial asthma, pointing to an allergic reaction as a candidate etiologic factor. Here we have developed the concept of appendicular lavage and used it to study the levels of key Th2 cytokines, IL-4, IL-5 and IL-9 in patients with a clinical diagnosis of acute appendicitis. The study group consisted of 20 phlegmonous appendicitis, 13 gangrenous appendicitis, and a control group of 8 patients with clinical diagnosis of appendicitis but with normal histology. Cytokine levels are higher in acute appendicitis. The difference was clearer when comparing phlegmonous appendicitis with non-pathological appendix (p=0.01), respectively 48.3 vs 21.3; 29.2 vs 8.0; 34.1 vs 16.6 pg/mL for IL-4, IL-5 and IL-9. This Th2 cytokine profile is compatible with the hypothesis of allergy as an etiologic factor for acute appendicitis and may have important implications for the diagnosis, prevention and treatment of this condition.

Key words: acute appendicitis, allergy, appendicular lavage, IL-4, IL-5, IL-9, Th2 cytokine
Introduction

Acute appendicitis (AA) is a complex disease whose etiology cannot be explained by any single factor. Luminal obstruction is believed to be the trigger event that culminates in inflammation of the appendix. Fecaliths are found in one-third of specimens. In the other cases, obstruction is thought to be caused by hypertrophy of mural lymphoid follicles in response to diverse causes [1]. The peak incidence of appendicitis coincides with the age when the immune response is most vigorous, and the lymphoid follicles are at their maximum development [2].

Aravandian has reported histological features in AA that are similar to bronchial asthma, a paradigm for an allergic reaction. Based on these findings, this author has proposed that AA is triggered by a hypersensitivity type I reaction and, therefore, could be caused by an allergic reaction [3]. Cytokines from Th2 lymphocytes are responsible for the histological features of asthma. [4]. Th2 effector cells secrete mainly interleukin-4 (IL-4), IL-5, IL-9 and IL-13, which are known to be involved in allergic responses [5].

Broncho-Alveolar Lavage (BAL) is a useful tool for investigating inflammatory cell and mediator profiles, like cytokines, in various bronchopulmonary diseases [6]. High levels of IL-4 and IL-5 are found in BAL in asthma [7]. Similar to BAL, we have used the concept of appendicular lavage (AL), where saline is instilled and collected in appendicular lumen of appendectomy surgical specimens. The aim of this study was to test the hypothesis that AA might be the consequence of an allergic reaction by evaluating the levels of Th2 cytokines in AL fluid of patients submitted to appendectomy due to a clinical diagnosis of AA.

Materials and Methods

Study population

The study group, evaluated between April 2016 and June 2017, consisted of patients with the clinical diagnosis of AA, admitted to emergency department when one of the authors (NC) was on call to perform the appendicular lavage. The only exclusion criterion was the absence of the author (NC). The histological diagnosis of AA discriminates acute phlegmonous appendicitis (APA) and acute gangrenous appendicitis (AGA). The control group consisted of patients admitted with the clinical diagnosis of AA, submitted to appendectomy, but with normal histology (Non-Pathological Appendix – NPA). No type of allergy test was performed. Laparoscopic appendectomy was performed in 33 patients and open surgery in 8 patients, including 3 conversions from laparoscopy. There were 9 localized and 3 generalized peritonitis.

Ethics Considerations

The study is part of a research project approved by the Ethics Committee of Garcia de Orta Hospital (Reference 05/2015) and each enrolled subject gave written informed consent. The work has been carried out in accordance with The Code of Ethics of the World Medical Association (Declaration of Helsinki). The authors declare no conflict of interest.
**Appendicular lavage**

After removal of the appendicular specimen, a gauge was inserted in the proximal luminal aspect of the appendix and 3 mL of saline 0.9 % were instilled and collected for AL. Saline was re-instilled and collected 3 times. Appendicular lavage process was standardized and performed exclusively by one of the authors (NC). The appendicular fluid samples were collected to a Sarstedt Monovette tube and centrifuged. 1 mL of the supernatant was extracted and stored at -20 °C. ELISA protocol was used for IL-4, IL-5 and IL-9 determinations (Human IL-4, IL-5 and IL-9 MAX, Biolegend, San Diego, CA 92121 USA) according to manufacturer’s protocol. The cytokine levels, IL-4, IL-5 and IL-9 were expressed in pg/mL.

**Pathologic analysis**

After AL procedures, the appendices were preserved in 10% formalin for histopathological examination. A minimum of 24 hours was allowed for adequate tissue fixation. Appendicular sections were taken from the tip, base and intermediate length for fixation and paraffin processing. Two sections of 5-micron thickness were cut from each paraffin block and stained by hematoxylin & eosin [8]. The criteria for AA was polymorphous nuclear neutrophils infiltration at muscularis propria [9]. APA was defined by the presence of neutrophils infiltrate in muscular propria and AGA was defined by the presence of necrosis of the wall of the appendix in a background of transmural inflammation [10]. The presence of neutrophils in the mucosa was considered as a variant of normal with no clinical relevance, when no other inflammatory cells were detected in abnormal numbers. The specimens were classified as negative for appendicitis when no neutrophil infiltrate was show in muscular propria (NPA) [10]. All the histopathological analyses were performed by one of the authors masked to the cytokines results (CH).

**Statistical analysis**

Data are presented as descriptive statistics, mean and standard deviation. For continuous variables, considering the distribution of number of cases among the categories, a non-parametric approach was followed to assess statistical differences among the considered groups: Kruskal-Wallis tests were used, with *a posteriori* pairwise Wilcoxon tests; p-values were then corrected for multiple comparison using the Holm correction. For categorical variables, such as gender, a strategy based on Fisher’s exact test overall and pairwise, using the aforementioned correction, were used. Statistical Analysis was performed on R ([https://cran.r-project.org](https://cran.r-project.org)), using the stats package for hypothesis testing and ggplot2 for the plots.

**Results**
We analyzed 33 patients with a histological diagnosis of AA, 20 patients with APA, 13 patients with AGA and 8 patients, the control group, with normal histology. History of allergy was present in 7 patients (4 for antibiotics, 1 for metibasol®, and 2 with allergic rhinitis), no differences between groups were identified. None took medication. No differences in age, gender and BMI among groups were found (p=0.898, p= 0.054 and p=0.211, respectively) (Table1). A significant difference was found for C-Reactive – Protein levels and Length of stay (p=0.001, p= 0.002, respectively) (Table 1). Cytokines levels in AL according to the histologic groups are present in Table 2. In all the studied subjects, cytokines IL-4, IL-5, IL-9 were detected in AL fluid. For IL-4 there were significant differences among the histological groups (p=0.034). The difference between APA and NPA groups was significant (p=0.017). No significant differences for AGA group with the remaining groups were found (AGA vs APA p=0.421; AGA vs. NPA; p=0.421) (Figure 1). For IL-5, the differences were subtle (p=0.056). Differences among groups showed a tendency for different levels of IL-5 between APA and NPA (p=0.05) (Figure 2). As for IL-9, there was no clear evidence of differences for AGA group (AGA vs APA; p=0.587; AGA vs. NPA; p=0.587). IL-9 was the cytokine whose levels revealed less differences among groups (p=0.083). However, while not statistically significant, there was a tendency indicating that differences exist between APA and NPA groups for this cytokine (p= 0.062) (Figure 3). No differences were found between groups for IL-4 and IL-5 blood levels.

Discussion

The most common theory for the etiology of AA is intraluminal obstruction, which is not supported by histological findings in surgical specimens. In most cases, no obstruction is found [11]. Only in 0.4 % of 1969 appendectomy specimens was there evidence of luminal obstruction by vegetable fibers [12].

Based on histologic findings, specifically eosinophilic infiltration, mastocyte degranulation and muscular edema, an allergic reaction, was proposed as a possible etiology of AA [3]. The concept is attractive: the appendix is a lymphoid organ, therefore, an immune response to a local antigen could be a factor in the pathogenesis of AA [13]. The gastrointestinal system is one of the main entrances into the body for allergens during all life stages [9]. Atopy may be a risk factor for appendicitis [9]. By analogy with asthma, the contraction of the muscular wall of the appendix in response to antigenic stimulation, may result in luminal obstruction culminating in AA. This response may occur in any segment of the intestine, but the appendix is more vulnerable because of its small lumen size and limited capacity to accumulate fluids [3]. The identification of the offending allergen(s) is not always straightforward [14].

Allergies are inflammatory diseases dependent on Th2 activation, mediated by IL-4, IL-5 and IL-9, in response to environmental allergens [7, 15, 16]. IL-4 acts as a growth factor for Th2 cells and promotes the production of IgE. IL-5 induces the differentiation, the activation, and the survival of eosinophils. IL-4 and IL-9 induce the growth of mast cells and basophils [17, 18].
BAL has been performed for several years for lung diseases. We extend and adapt this approach to monitor the levels of cytokines in the appendix: appendicular lavage. The fluid collected from appendicular lumen may reflect local inflammatory alterations. We used 3 lavages with NaCl 0,9 % at appendicular lumen to mobilize cytokines that are adherent to the mucosa. All steps of the process were standardized, from the surgical handling of the specimens to the processing of lavage and quantitative determinations. Differences in cytokines should be accepted as reflecting local inflammatory changes. Elevated levels of IL-4 and IL-5, are present in BAL of patients with allergic asthma [6]. Higher levels are seen in symptomatic patients [19]. The results of our study show a strong statistical difference in the levels of IL-4 between phlegmonous and non-pathologic appendicitis. IL-4 elevation reflects a putative allergic reaction in AA. In the case of IL-5, our data suggests differences between groups, mainly between phlegmonous and non-pathological appendix. For IL-9 there are also indications of differences between groups, mainly between phlegmonous and non-pathological appendix.

No significant differences for Th2 cytokines profile were founded between AGA and APA or NPA. In fact, some authors claim that simple inflammatory appendicitis and necrosis represent different diseases, or different patient response to disease, with distinct epidemiology, natural history, microbiology and different Th17 cytokine profile [20,21,22]. Blood inflammatory response in AA showed a positive association of Th1 mediated immunity and gangrenous appendicitis [21]. AGA etiology may be different from APA, without allergic component and so, Th2 cytokines are not elevated in AL. Another possible explanation for IL-4 and IL-5 similar values in AGA and NPA is that the cellular destruction in AGA is so marked that Th2 cells can no longer produce IL-4 and IL-5 and the values fall down to values found in NPA.

In asthma, the paradigm of allergic disease, cytokine profile in BAL shows elevation of IL-4, IL-5 and IL-9 compared to control groups [6]. Our results with AL are similar to those findings in BAL in the presence of allergy. In fact, our study showed elevations of Th2 cytokines in AL fluid of patients with appendicitis, compared with the control group (non-pathologic appendicitis).

Strengths: Using a novel methodology, appendicular lavage, our study provides objective and original data demonstrating a correlation between cytokines and appendicular histological features that show that an allergic component is presented in AA.

Limitation: Appendicular lavage should be reproduced by other researchers. The sample size is still limited. A larger dataset of patients, originating from multiple centers is strongly recommended because it is likely to validate and extend the conclusions of the current study.

Conclusion

Differences in IL-4, IL-5 and IL-9 in AL fluid were found between the 3 study groups and especially significant between phlegmonous and negative appendicitis. Therefore, in AL fluid, we found a Th2 cytokine profile compatible with allergy. Further studies are
required to assess the importance and robustness of our results regarding the allergic components in AA. The identification of an allergen will be a particularly difficult task, but our results open the possibility for novel management strategies in AA that might not always include a surgical procedure.

Acknowledgements
We especially thank patients, surgical residents, surgeons, anesthesiologists, pathologists, nurses, medical and nursing students. Without their generous collaboration and hard work, this study would not have been possible. This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Author Contributions
N.D.C. designed the experiment, acquired data and wrote the manuscript. A.B.B., C.F.M. and A.N.C. did the analyses of the data. H.O.C. and F.C.B. acquired data including the pathology analyses of surgical samples. L.F.M and P.M.C. advised the project and reviewed the manuscript.

Conflict of interest
The authors have no conflict of interest in relation to this work.

References
[1] R. B., “Epidemiologic Features of Appendicitis,” Append. - A Collect. Essays
from Around World, pp. 1–20, 2012.

[2] T. Kuga, S. Taniguchi, T. Inoue, N. Zempo, and K. Esato, “Immunocytochemical Analysis of Cellular Infiltrates in Human,” Surg. Today, pp. 1083–1088, 2000.

[3] K. Aravindan, “Eosinophils in Acute Appendicitis: possible significance,” Indian J. Pathol. Microbiol., vol. 40 (4): 49, 1997.

[4] J. Just, L. Fournier, I. Momas, C. Zambetti, F. Sahraoui, and A. Grimfeld, “Clinical significance of bronchoalveolar eosinophils in childhood asthma,” J. Allergy Clin. Immunol., vol. 110, no. 1, pp. 42–44, 2002.

[5] K. C. L. Torres, W. O. Dutra, and K. J. Gollob, “Endogenous IL-4 and IFN-γ are essential for expression of Th2, but not Th1 cytokine message during the early differentiation of human CD4+ T helper cells,” Hum. Immunol., vol. 65, no. 11, pp. 1328–1335, 2004.

[6] J. C. Virchow, C. Kroegel, C. Walker, and H. Matthys, “Inflammatory determinants of asthma severity: mediator and cellular changes in bronchoalveolar lavage fluid of patients with severe asthma.,” J. Allergy Clin. Immunol., vol. 98, no. 5 Pt 2, pp. S27-33; discussion S33-40, 1996.

[7] M. Yazdanbakhsh, “Th2 Responses without Atopy: Immunoregulation in Chronic Helminth Infections and Reduce Allergic Disease.,” Trends Immunol., vol. 22, no. 7, pp. 372–377, 2001.

[8] A. Kolur, A. Patil, V. Agarwal, S. Yendigiri, and B. Sajjanar, “The Significance of Mast Cells and Eosinophils Counts in Surgically Resected Appendix,” J. Interdiscip. Histopathol., vol. 2, no. 3, p. 1, 2014.

[9] A. Harlak et al., “Atopy is a risk factor for acute appendicitis? A prospective clinical study,” J. Gastrointest. Surg., vol. 12, no. 7, pp. 1251–1256, 2008.

[10] L. W. Lamps, “Beyond acute inflammation: a review of appendicitis and infections of the appendix,” Diagnostic Histopathol., vol. 14, no. 2, pp. 68–77, 2008.

[11] L. S. Bernstein, B. Surick, and I. M. Leitman, “Is acute appendicitis in the weather forecast?,” J. Surg. Res., vol. 185, no. 1, pp. e23–e25, 2013.

[12] O. Engin, M. Yildirim, S. Yakan, and G. A. Coskun, “Can fruit seeds and undigested plant residuals cause acute appendicitis,” Asian Pac. J. Trop. Biomed., vol. 1, no. 2, pp. 99–101, 2011.

[13] M. Tsuji, G. McMahon, D. Reen, and P. Puri, “New insights into the pathogenesis of appendicitis based on immunocytochemical analysis of early immune response.,” vol. 25, no. 4, pp. 449–452, 1990.

[14] D. G. Ebo et al., “Flow-assisted allergy diagnosis: Current applications and future perspectives,” Allergy Eur. J. Allergy Clin. Immunol., vol. 61, no. 9, pp. 1028–1039, 2006.

[15] K. Takatsu and H. Nakajima, “IL-5 and eosinophilia,” Curr. Opin. Immunol., vol. 20, no. 3, pp. 288–294, 2008.

[16] C. K. Kim, S. W. Kim, C. S. Park, B. Kim, H. Kang, and Y. Y. Koh, “Bronchoalveolar lavage cytokine profiles in acute asthma and acute bronchiolitis,” J. Allergy Clin. Immunol., vol. 112, no. 1, pp. 64–71, 2003.

[17] S. Romagnani, “The role of lymphocytes in allergic disease,” J. Allergy Clin. Immunol., vol. 105, no. 3, pp. 399–408, 2000.
[18] J. Gilmour and P. Lavender, “Control of IL-4 expression in T helper 1 and 2 cells,” *Immunology*, vol. 124, no. 4, pp. 437–444, 2008.

[19] S. E. Wenzel, “Abnormalities of cell and mediator levels in bronchoalveolar lavage fluid of patients with mild asthma,” *J. Allergy Clin. Immunol.*, vol. 98, no. 5 Pt 2, pp. S17-21; discussion S33-40, 1996.

[20] M. Rubér, M. Andersson, B. F. Petersson, G. Olaison, R. E. Andersson, and C. Ekerfelt, “Systemic Th17-like cytokine pattern in gangrenous appendicitis but not in phlegmonous appendicitis,” *Surgery*, vol. 147, no. 3, pp. 366–372, 2010.

[21] M. Rubér, A. Berg, C. Ekerfelt, G. Olaison, and R. E. Andersson, “Different cytokine profiles in patients with a history of gangrenous or phlegmonous appendicitis,” *Clin. Exp. Immunol.*, vol. 143, no. 1, pp. 117–124, 2006.

[22] García-Marín A, Pérez-López M, Martínez-Guerrero E, Rodríguez-Cazalla and Compan-Rosique A. Microbiologic Analysis of Complicated and uncomplicated Acute Appendicitis. Surgical Infections, Volume 19, Number 1, 83-86, 2018

**Figure 1.** Box plots of IL-4 levels according to the different histological categories.

![](image1)

**Figure 2.** Box plots of IL-5 levels according to the different histological categories.
Figure 3. Box plots of IL-9 levels according to the different histological categories.
Table 1. Characterization of the population according to Histological Categories

| Variable          | APA   | AGA   | NPA   | p-value |
|-------------------|-------|-------|-------|---------|
| N (%)             | 20 (49) | 13 (32) | 8 (19) |         |
| Age (y)           | 36.2±16.9 | 20.0±15.1 | 34.2±12.9 | 0.898   |
| Gender (M/F)      | 14/6  | 4/9   | 6/2   | 0.054   |
| BMI               | 25.3±6.0 | 26.6±3.1 | 23.7±3.7 | 0.211   |
| Allergy           | 3     | 2     | 2     | NA      |
| WBC               | 14.3±4.4 | 14.4±3.8 | 12.3±2.9 | 0.161   |
| CRP               | 3.7±5.6 | 9.4±6.2 | 4.6±6.3 | 0.001   |
| LOS               | 2.8±1.8 | 5.4±3.9 | 3.3±3.1 | 0.002   |

Results presented as number (valid percentage) or mean ± standard deviation. APA= acute phlegmonous appendicitis; AGA= acute gangrenous appendicitis; NPA= Non Pathological Appendix; M=Male; F=Female; BMI= Body Mass Index; NA= Not applicable; WBC= White Blood Count; CRP= C-Reactive-Protein; LOS= Length of stay;

Table 2. Cytokine levels (pg/mL) according to the histological categories

|            | IL-4     | IL-5      | IL-9      |
|------------|----------|-----------|-----------|
|            | Mean±SD  | Mean±SD   | Mean±SD   |
| Phlegmonous| 158,1±228 | 76,3±106,5 | 101,9±141,7 |
| Gangrenous | 109,2±154,7 | 49,2±81,6 | 72,5±110,3 |
| Non-Pathological | 23,5±6,8 | 9,9±4,0 | 17,9±7,0 |