Background: Osteosarcoma is a common malignancy of bone with inferior survival outcome. Autophagy can exert multifactorial influence on tumorigenesis and tumor progression. However, the specific function of genes related to autophagy in the prognosis of osteosarcoma patients remains unclear. Herein, we aimed to explore the association of genes related to autophagy with the survival outcome of osteosarcoma patients.

Methods: The autophagy-associated genes that were related to the prognosis of osteosarcoma were optimized by LASSO Cox regression analysis. The survival of osteosarcoma patients was forecasted by multivariate Cox regression analysis. The immune infiltration status of 22 immune cell types in osteosarcoma patients with high and low risk scores was compared by using the CIBERSORT tool.

Results: The risk score model constructed according to 14 autophagy-related genes (ATG4A, BAK1, BNIP3, CALCOCO2, CCL2, DAPK1, EGFR, FAS, GRID2, ITGA3, MYC, RAB33B, USP10, and WIPI1) could effectively predict the prognosis of patients with osteosarcoma. A nomogram model was established based on risk score and metastasis.

Conclusion: Autophagy-related genes were identified as pivotal prognostic signatures, which could guide the clinical decision making in the treatment of osteosarcoma.

Keywords: prognosis, risk score, osteosarcoma, autophagy-related genes, nomogram

BACKGROUND

Osteosarcoma, also known as osteogenic sarcoma, is considered to be the most common malignancy in bone among children and adolescents [1, 2]. Although the metaphyseal region is the most common location of osteosarcoma, this disease usually progresses rapidly and is prone to metastasis [3, 4]. The prognosis of osteosarcoma is poor and it seriously threatens the life and health of adolescents [5]. Therefore, it is urgent to explore novel targets or signatures for improving the clinical practice of osteosarcoma patients in the future.

Autophagy is a biological process mediated by certain genes, in which the aberrant organelles as well as macromolecules are digested by lysosomes, and it is involved in multiple processes such as cell metabolism, renewal of organelles and intracellular homeostasis maintenance [4, 6]. In recent years, several studies have indicated the relationship of autophagy with the initiation and progression of various diseases, such as cancer and diseases associated with neurodegeneration and immunization [7]. In bladder cancer, autophagy could inhibit the tumorigenesis via limiting tissue damage and...
oncogenic signaling [8]. Furthermore, autophagy could eliminate the accumulation of damaged proteins and organelles, indicating its role in the prevention of tumorigenesis. However, other research has reported that autophagy was essential for enhancing the survival ability of tumor cells and suppressing the necrosis in some cancers, including melanoma and breast cancer [9, 10]. Recycling by autophagy is necessary for the maintenance of energy balance and mitochondrial metabolism for tumor growth and proliferation. In addition, suppression of autophagy is considered as a potential modality for tumor treatment [11]. Autophagy also plays a significant role in osteosarcoma. It has been proved that autophagy is induced in osteosarcoma, and several intermediates are implicated in this process [12]. Autophagy could be promoted by the increased expression of high mobility group box 1, contributing to the drug resistance during the treatment of osteosarcoma [13]. Sun et al. reported that the silence of autophagy-related gene 5 reduced the malignancy of osteosarcoma with anti-oncogenic effects [14]. Moreover, Liu et al. found that the expression of autophagy-related 4B was obviously elevated, which accelerated osteosarcoma development and suppressed the apoptosis of osteosarcoma cells [15]. Consequently, analysis and identification of autophagy-related genes are helpful to improve our knowledge on the association of autophagy with osteosarcoma.

Herein, we analyzed 210 genes associated with autophagy and identified 14 optimized autophagy-associated genes related to the survival outcome of osteosarcoma patients. A death risk model based on those 14 autophagy-related genes could effectively predict the prognosis of osteosarcoma patients. Finally, we established the nomogram model by including independent factors of prognosis (risk score and metastasis) and demonstrated its better performance in predicting the long-term prognosis of osteosarcoma patients.

**METHODS**

**Data Sources**

We downloaded the mRNA expression profiles of 88 osteosarcoma patients with their corresponding clinical information from the Therapeutically Applicable Research to Generate Effective Treatments (TARGET, https://ocg.cancer.gov/programs/target) database. Among them, 85 patients had complete survival information, whose clinicopathological features are depicted in Table 1. Moreover, by using osteosarcoma and survival as keywords in the Gene Expression Omnibus (GEO, https://www.ncbi.nlm.nih.gov/geo/) database, we obtained the mRNA expression information and clinical information of GSE21257 [16] and GSE16091 [17]. Through the SVA package of R software, the batch effect was removed between different datasets. The two datasets were combined to verify the prognostic model. GSE21257 included 53 osteosarcoma samples, and the mRNA expression profile data was detected using Illumina human-6 v2.0 expression beadchip. GSE16091 consisted of 34 osteosarcoma samples, and mRNA expression profile was quantified by Affymetrix Human Genome U133A Array. The clinicopathological characteristics of osteosarcoma patients in GEO datasets were shown in Supplementary Table S1. The mRNA expression data that was previously normalized was used in our study. In addition, we selected the 210 genes associated with autophagy by referring to the previous study [18] and Human Autophagy Database (HADb, www.autophagy.lu/project.html), whose details are displayed in the attached Supplementary Table S2.

**Cluster Analysis**

According to the mRNA expression levels of the 210 genes related to autophagy, the samples were clustered by using factoextra package in R software (https://CRAN.R-project.org/package=factoextra), followed by principal component analysis (PCA).

**LASSO Cox Regression Analysis**

Univariate Cox regression analysis was carried out basing on the mRNA expression levels of 210 genes associated with autophagy, and by using the threshold of \( p < 0.05 \), the autophagy-associated genes that were related to the survival outcome of osteosarcoma patients were selected. Subsequently, LASSO Cox regression analysis was carried out with glmnet package in R software [19]. In the LASSO regression model, the lambda value which corresponded to the minimum value of partial likelihood deviance was considered as the best one, and the best tuning parameter lambda was used to screen the genes associated with autophagy that showed a significant relationship with the survival outcome of osteosarcoma.

\[
\text{Risk score} = \sum_{i=1}^{n} \text{Coef}_i * x_i
\]

Among them, the risk coefficients of all factors were computed using LASSO Cox model and expressed as \( \text{Coef}_i \), \( x_i \) represented

| Characteristics | Patients (N = 85) |
|-----------------|-----------------|
|                | No. | %    |
| Sex            |     |      |
| Female          | 37  | 43.53|
| Male            | 47  | 55.29|
| Unknown         | 1   | 1.18 |
| Age             |     |      |
| ≤ 14 (Median)   | 44  | 51.76|
| > 14 (Median)   | 40  | 47.06|
| Unknown         | 1   | 1.18 |
| Race            |     |      |
| Asian           | 6   | 7.06 |
| Black or African American | 7 | 8.24 |
| Unknown         | 21  | 24.71|
| Disease at diagnosis |     |      |
| Metastatic disease | 21 | 24.71|
| Non-metastatic disease | 63 | 74.12|
| Unknown         | 1   | 1.18 |
| Primary tumor site |    |      |
| Arm/hand        | 6   | 7.06 |
| Leg/foot        | 76  | 89.41|
| Pelvis          | 2   | 2.35 |
| Unknown         | 1   | 1.18 |
| Vital status    |     |      |
| Alive           | 58  | 68.24|
| Dead            | 27  | 31.76|
the mRNA expression levels of factors. We confirmed the optimal cutoff value of the risk score via survival and survminer packages in R software and bilateral log rank test, and then stratified these osteosarcoma patients into high and low risk groups by the above cutoff value.

**Survival Analysis**
The overall survival (OS) rate of different groups was assessed by using survival and survminer packages in R software. R language survival ROC package [20] was applied to draw the time-dependent ROC curve. The multivariate Cox regression model was constructed to verify whether risk score was an independent signature for osteosarcoma prognosis after adjusting for multiple factors. The OS of different groups were evaluated by using the Kaplan-Meier method [21] followed by OS comparison via log-rank test. The divergences in infiltrating immune cells between different groups were analyzed by Wilcoxon signed-rank test [22] using $p$ value less than 0.05 as the threshold. All analyses were carried out by R software (version 3.5.2).

**Immune Cell Infiltration Proportion Analysis**
The relative ratio of 22 immune cell types was computed using CIBERSORT [23], which characterized the composition of immune infiltration cells by deconvolution algorithm using the preset 547 barcode genes on the base of gene expression matrix. We set the sum of immune cell ratios as 1 for all samples.

**Gene Set Enrichment Analysis**
GSEA (version 4.0.3) [24] was used for gene set enrichment analysis with c2.cp.kegg.v7.0. symbols derived from Molecular Signatures Database (MSigDB) as the gene set. The significantly enriched KEGG pathway was screened with $p < 0.05$ as the threshold.
FIGURE 2 | The risk score model predicted the survival of patients with osteosarcoma. (A) Forest map of autophagy-related genes significantly related to overall survival in univariate analysis. HR was Hazard ratio, and 95% CI was 95% confidence interval. (B) Diagram of the optimal number of genes in the LASSO regression model. The horizontal axis represented log (lambda), and the vertical axis represented the partial likelihood deviance. The Lambda value corresponding to the minimum value was the best. (C) Coefficient spectrum of LASSO Cox regression model. (D) The risk scores distribution of samples in the TARGET dataset. A point indicated a sample, a red point represented a sample with a higher risk score, a green point indicated a sample with a lower risk score, and the intersecting point represented the optimal risk score. (E) The Cluster heat map of 14 autophagy-related gene expressions in the TARGET dataset. Behavioral genes were listed as samples. Red
expression of genes were conducive to patient prognosis. Other genes related to prognosis were protective genes, and high expression of genes led to poor prognosis.

1.1 selection criteria (\( \frac{\text{SSE}}{\text{SSE}} \)) were used to select the number of clusters \( k \) in the Voronoi diagram. The clustering diagram showed that samples within cluster 1 and cluster 2 could be well distinguished (Figure 1C). The expression calorimetry diagram (Figure 1D) showed the consistency of clustering, and these two types could be obviously distinguished. The principal component analysis (PCA) was carried out, and the result displayed that samples within cluster 1 and cluster 2 could be well distinguished (Figure 1D). Kaplan-Meier survival analysis revealed obviously worse survival outcome of osteosarcoma patients in cluster 1 than those in cluster 2 (Figure 1E). Autophagy-related genes could effectively distinguish the osteosarcoma patients with different prognosis.

**RESULTS**

**Autophagy-Related Genes Distinguished Osteosarcoma Patients With Different Prognoses**

For the 85 osteosarcoma samples with complete survival information in the TARGET database, we used the factoextra function package in R language to perform cluster analysis based on the expression levels of 210 genes which were related to autophagy. With reference to the sum of the squared errors (SSE), we selected the number of clusters \( k = 2 \) (Figure 1A) to cluster the samples into two types. The clustering diagram (Figure 1B) and the expression calorimetry diagram (Figure 1C) showed the consistency of clustering, and these two types could be obviously distinguished. The principal component analysis (PCA) was carried out, and the result displayed that samples within cluster 1 and cluster 2 could be well distinguished (Figure 1D). Kaplan-Meier survival analysis revealed obviously worse survival outcome of osteosarcoma patients in cluster 1 than those in cluster 2 (Figure 1E). Autophagy-related genes could effectively distinguish the osteosarcoma patients with different prognosis.

Then, we carried out LASSO Cox regression analysis based on these 46 autophagy-related genes. The optimal number of genes was determined as 14 with reference to the minimum lambda value (Figures 2B,C). We established the risk score model after weighting the autophagy-associated genes expression and the coefficients to predict the survival outcome of osteosarcoma patients. Risk Score = 0.3316 * Expression Value of BNIP3-0.1571 * Expression Value of ATG4A-0.0114 * Expression Value of BAK1-0.192 * Expression Value of CALCOCO2-0.0457 * Expression Value of CCL2-0.0938 * Expression Value of DAPK1-0.0738 * Expression Value of EGFR-0.0734 * Expression Value of FAS-0.1609 * Expression Value of GRID2-0.0313 * Expression Value of ITGA3 + 0.2591 * Expression Value of MYC-0.0593 * Expression Value of RAB33B-0.1215 * Expression Value of USP10-0.0467 * Expression Value of WIP1. We calculated the risk score of osteosarcoma patients from TARGET and GEO cohorts (GSE21257 and GSE16091 combined), and divided the samples into high and low risk groups with reference to respective optimal cut-off value. The distribution of risk score for the samples was shown in Figures 2D,H. Meanwhile, manifest difference in autophagy-associated gene expression between groups with distinct risk scores was observed (Figures 2E,I). Survival analysis revealed the survival outcome of osteosarcoma samples with high risk scores was worse than those with low risk score (Figures 2F,J). In addition, the time-dependent ROC analysis displayed that the AUC of osteosarcoma patients in TARGET dataset for 1-year, 3-years, and 5-years OS were 0.86, 0.843, and 0.85, respectively (Figure 2G), and 0.873, 0.683, and 0.622 for the GEO cohort (Figure 2K). The result indicated that the risk models in both datasets effectively predicted the prognosis of patients with osteosarcoma. Overall, those results suggested that the risk assessment models constructed based on 14 autophagy-related genes including ATG4A, BAK1, BNIP3, CALCOCO2, CCL2, DAPK1, EGFR, FAS, GRID2, ITGA3, MYC, RAB33B, USP10, and WIP1 were able to forecast the survival outcome of osteosarcoma patients.

**Immune Infiltration Analysis**

The distinction in infiltration of the 22 immune cell types between osteosarcoma patients with high and low risk scores was analyzed by using CIBERSORT and LM22 feature matrix. The immune infiltration landscape of 85 osteosarcoma patients was shown in Figure 3A. The proportions of immune cell infiltration in different patients were different, which probably reflected the inherent features of individuals. A weak correlation of the infiltration proportion among different immune cell types was found (Figure 3B), which indicated that there was a large
heterogeneity in the infiltration of different immune cells in tumor patients. Moreover, we found that activated dendritic cells, M2 macrophages and CD8 T cells had remarkable differences in the degree of infiltration between groups with different risk scores. As shown in Figure 3C, the infiltration proportions of activated dendritic cells, M2 macrophages, and CD8 T cells were significantly higher in the low risk group than those in the high risk group, which might be associated with the prognostic difference between these two groups.

Immune checkpoints have been the research hotspots in recent years, which show great clinical significance and provide promising treatment target in cancer. It was found that there was a significant relationship of the risk score with important immune checkpoints expressions (CTLA4, PDL1, TIM3, LAG3, TIGIT) in osteosarcoma patients (Figure 3D and Supplementary Table S3). In addition, five immune checkpoints expressions in high and low risk groups and different clusters of osteosarcoma patients were investigated.
The result revealed that CTLA4, TIM3, LAG3, and TIGIT expression levels in the group with low risk scores were obviously elevated compared with the group with high risk scores (p < 0.05) (Figure 3E), and the expressions of CTLA4, PD1, TIM3, LAG3, and TIGIT in Cluster 2 samples were strikingly increased compared with those in Cluster 1 samples (Figure 3F), indicating that the patients of the group with low risk scores and Cluster 2 were probably more sensitive to the treatment of immune checkpoint inhibitors.

GSEA Enrichment Analysis
The gene set enrichment analysis (GSEA) was carried out for osteosarcoma patients with different risk scores, and the significantly enriched KEGG pathway was screened with the threshold of p value less than 0.05. A total of 23 pathways were significantly enriched as shown in Table 2. The top six pathways were displayed in Figures 4A–F. It was found that the immune-related pathways were more likely to be enriched in osteosarcoma patients with low risk scores.

| KEGG pathway | Normalized enrichment score | NOM p-val |
|---------------|-----------------------------|-----------|
| KEGG_SYSTEMIC_LUPUSERYTHEMATOSUS | 1.7960994 | 0.012793177 |
| KEGG_T_CELL_RECEPTOR_SIGNALING_PATHWAY | 1.7504385 | 0.016666668 |
| KEGG_CYTOKINE_CYTOKINE_RECEPTOR_INTERACTION | 1.7406833 | 0 |
| KEGG_B_CELL_RECEPTOR_SIGNALING_PATHWAY | 1.7296787 | 0.028906451 |
| KEGG_COMPLEMENT_AND_COAGULATION_CASCADES | 1.7081273 | 0 |
| KEGG_CHEMOKINE_SIGNALING_PATHWAY | 1.7065754 | 0.02306752 |
| KEGG_IMMUNE_NETWORK_FOR_GA_PRODUCTION | 1.700867 | 0.006429383 |
| KEGG_HEMATOPOIETIC_CELL_LINEAGE | 1.6924057 | 0.015151516 |
| KEGG_CELL_ADEHESION_MOLECULES_CAMS | 1.6835115 | 0.014861995 |
| KEGG PRIMARY_IMMUNODEFICIENCY | 1.6811936 | 0.010845987 |
| KEGG_ALLOGRAFT_REJECTION | 1.6579281 | 0.031982943 |
| KEGG_ASTHMA | 1.6458044 | 0.02258727 |
| KEGG_NATURAL_KILLER_CELL_MEDIATED_CYTOTOXICITY | 1.6449951 | 0.03068803 |
| KEGG_LEUKOCYTE_TRANSIENT_ENDPOINTIAL_MIGRATION | 1.6381815 | 0.019480519 |
| KEGG_AUTOIMMUNE_THYROID_DISEASE | 1.6160316 | 0.020220589 |
| KEGG_TYPE_1_DIABETES_MELLITUS | 1.6096383 | 0.04621849 |
| KEGG_RENIN_ANGIOTENSIN_SYSTEM | 1.6096383 | 0.04621849 |
| KEGG_LEUKOCYTE_ADHESION_MOLECULES_CAMS | 1.6096383 | 0.04621849 |
| KEGG PRIMARY_IMMUNODEFICIENCY | 1.6096383 | 0.04621849 |
| KEGG_ASTHMA | 1.6096383 | 0.04621849 |
| KEGG_GLYCOSAMINOGLYCAN_DEGRADATION | 1.6096383 | 0.04621849 |
| KEGG_ENDOCYTOSIS | 1.5964051 | 0.02631579 |
| KEGG_CYTOKINE_CYTOKINE_RECEPTOR_INTERACTION | 1.5839647 | 0.02631579 |
| KEGG_COMPLEMENT_AND_COAGULATION_CASCADES | 1.5839647 | 0.02631579 |
| KEGG Regulation_of_Actin_Cytoskeleton | 1.5839647 | 0.02631579 |
| KEGG_NATURAL_KILLER_CELL_MEDIATED_CYTOTOXICITY | 1.5839647 | 0.02631579 |
| KEGG_ASTHMA | 1.5839647 | 0.02631579 |
| KEGG PRIMARY_IMMUNODEFICIENCY | 1.5839647 | 0.02631579 |
| KEGG_ASTHMA | 1.5839647 | 0.02631579 |
| KEGG_CYTOKINE_CYTOKINE_RECEPTOR_INTERACTION | 1.5839647 | 0.02631579 |
| KEGG_COMPLEMENT_AND_COAGULATION_CASCADES | 1.5839647 | 0.02631579 |
| KEGG_REGULATION_OF_ACTIN_CYTOSKELETON | 1.5839647 | 0.02631579 |

Nomogram Model Could Better Forecast the Survival of Osteosarcoma Patients
The nomogram model was established based on risk score and metastatic status (Figure 6A). Then, the nomogram model was verified by proportional hazards (PH) assumption, and conformed to the PH assumption test (Supplementary Figure S1). Three lines were drawn upward to measure the points of each factor in the nomogram. Subsequently, we plotted a line downward from the total points, which represented the sum of all points here, to obtain the actual result (Figures 6B–D). When predicting the
FIGURE 4 | The top six significantly enriched pathways in GSEA enrichment analysis. (A) B cell receptor signaling pathway; (B) Chemokine signaling pathway; (C) Systemic lupus erythematosus; (D) T cell receptor signaling pathway; (E) Complement and coagulation cascades; (F) Cytokine cytokine receptor interaction.
survival outcome of osteosarcoma patients at 1, 3, and 5 years, the AUC value of the nomogram model based on two independent factors for prognosis was higher than that based on one (Figures 6E–G), suggesting the better performance of the nomogram model in the prognostic prediction of osteosarcoma patients.

**DISCUSSION**

Osteosarcoma is a frequent malignancy in bone among children and adolescents, with an estimated incidence rate of 3 per million per year all over the world. Osteosarcoma is often accompanied with early metastasis, thus being considered as an invasive tumor [26, 27]. Although radiotherapy and neo-adjuvant chemotherapy have been developing over the past years, the OS at 5 years in metastatic cases remains only about 20%. The cure rate for osteosarcoma patients that have focal tumor increases up to 70% [28, 29]. Therefore, it is urgent to explore and identify novel prognostic biomarkers for proper clinical decision making, which may provide a searchable idea to ameliorate treatment status and survival outcome of osteosarcoma patients.

Autophagy is self-degradation targeting defective proteins and organelles, and maintains the function of mitochondria under the condition of stress. Extensive research has reveal that autophagy is associated with tumor initiation and progression. Indeed, autophagy could contribute to suppression of tumorigenesis of liver tumor through cell-intrinsic p62 accumulation [30]. Another study found that autophagy suppressed the pancreatic tumor formation by p53 loss [31]. Nevertheless, autophagy was actually required for growth, survival, and tumorigenesis of pancreatic cancers [32]. Autophagy-related genes were upregulated in breast cancer cells transformed with RAS and promoted the invasion of cancer cells [33]. Hence, autophagy provides a potential alternative for osteosarcoma treatment.

In this study, 210 autophagy-related genes were collected which could distinguish the osteosarcoma patients with significantly different prognoses. LASSO Cox regression analysis was carried out and identified 14 optimized autophagy-related genes, including ATG4A, BAK1, BNIP3, CALCOCO2, CCL2, DAPK1, EGFR, FAS, GRID2, ITGA3, MYC, RAB33B, USP10, and WIPI1, for the prognosis of osteosarcoma. ATG4A, a redox-regulated cysteine protease, is a vital autophagy regulator. ATG4A promoted the transition from epithelium to mesenchyme partly by the Notch signaling pathway in osteosarcoma cells [34] and was related to reduced risk for lung cancer [35]. Another report showed that hypomethylation of ATG4A predicted a poor prognosis for ovarian cancer patients [36]. BAK1, a member of the B cell lymphoma family containing BH3 domain, could induce the mitochondria-mediated apoptosis by interacting with other proteins. Studies demonstrated that BAK1 played a role in drug resistance and tumor proliferation in many cancers including breast, lung and cervical cancers [37–39]. An established prognostic signature based on seven genes including BAK1 was able to predict the survival outcome of head and neck squamous cell carcinoma patients [40]. BNIP3, also a member of B cell lymphoma family, could regulate the cell survival, autophagy,
and cytoprotection. Moreover, research suggested that the elevated BNIP3 levels were correlated with progression to metastasis and poor prognosis in multiple cancers, such as breast and lung cancers, and uveal melanoma [41–43]. BNIP3 overexpression could induce the apoptosis of osteosarcoma cells, and BNIP3 inhibition plays a suppressive role in osteosarcoma cells apoptosis [44, 45]. CALCOCO2, also known as nuclear domain 10 protein 52, is implicated in autophagy factors recruitment and TANK-binding kinase 1 (TBK1) activation [46]. The model based on 16 autophagy related genes including CALCOCO2 could discriminate the multiple myeloma patients with distinct clinical outcomes, presenting potential prognostic value in multiple myeloma research [47]. CCL2 belongs to the CC chemokine family and is secreted by various cells including endothelial cells, fibroblasts, monocytes and tumor cells [48]. A previous study indicated that CCL2 could promote the invasion of pancreatic ductal adenocarcinoma [49] and the metastasis in cervical cancer [50]. In addition, high levels of CCL2 were related to the inferior survival outcome in gastric cancer [51]. Compared with the low-grade osteosarcoma, CCL2 expression was elevated in the osteosarcoma with high grade, which enhanced the proliferative and invasive abilities of osteosarcoma cells [52]. DAPK1 belongs to the Ser/Thr kinase family and is considered as a key regulator of autophagy and apoptosis [53]. It was found that DAPK1 expression could significantly inhibit the tumor growth and metastasis [54]. Down-regulation of DAPK1 expression may be a prognostic factor in many tumors, such as diffuse large B-cell lymphoma [55] and liver cancer [56]. EGFR is a receptor tyrosine kinase (RTK) for ErbB family, and exhibits over-

![Figure 6](image-url)
expression in various tumor cells [57]. Overexpression of EGFR is related to survival, invasion, metastasis, drug resistance, and poor prognosis of tumor [58], for example, EGFR is considered as an indicator of inferior prognosis in node-negative breast cancer [59]. EGFR was reported to be abnormally expressed in osteosarcoma, and the expression as well as amplification of EGFR were observed in the osteosarcoma with high grade (PMID: [60, 61]). FAS is an essential enzyme in the process of lipogenesis, and could effectively maintain the energy homeostasis. FAS expression level is up-regulated in several cancers and exhibits a strong effect on tumor cell proliferation and apoptosis. It was found that inhibition of FAS could obviously inhibit the growth and migration of bladder cancer cells [62], and Fas was proved to be a significant marker for the prognosis of breast cancer [63]. GRID2, belongs to the ionic glutamate receptor family, and regulates excitatory synaptic transmission [64]. Previous study has confirmed that GRID2 was related to the inferior survival outcome of prostate and gastric cancers [65, 66]. ITGA3 belongs to the integrin family of cell surface receptors and is involved in the survival, proliferation, and migration of cells. In gastric carcinomas, ITGA3 expression could facilitate the invasion [67] and was considered as a key signature for colon cancer [68]. Interestingly, ITGA3 was also a component of the prognostic signature for head and neck squamous cell carcinoma, with a similar role like BAK1, as described above [40]. ITGA3 polymorphisms might influence the osteosarcoma in terms of the incidence rate, metastatic status and prognosis, which was considered as a potential signature for osteosarcoma [69]. MYC, an oncogenic transcription factor, regulates cell proliferation, apoptosis, and carcinogenesis [70]. Research revealed that mutations of c-MYC could result in tumorigenesis [71]. The amplification of c-MYC was observed during the development of hepatocellular carcinoma, which was associated with impaired survival [72]. Moreover, increased expression of c-MYC was proved to enhance the invasive ability of osteosarcoma cells by targeting MEK-ERK pathway [73]. RAB33B belongs to the Rab family of small GTP binding proteins, and regulates the fusion of autophagosomes and membrane trafficking [74]. In addition, RAB33B was identified as a biomarker for lung cancer diagnosis [75]. However, its prognostic value or relationship with the survival outcome of cancer is rarely reported. USP10 belongs to the ubiquitin-specific protease family, modulates DNA damage response and autophagy [76, 77]. Moreover, USP10 inhibited cell growth and invasion in lung cancer [78], and was an independent factor for the prognosis of gastric carcinoma [79]. WIP1, a member of WD-repeat protein which interacts with phosphoinositides (WIPI) family, participates in the formation of autophagosome [80]. Furthermore, WIP1 was a relevant novel melanoma marker [81], and the increased expression of WIP1 indicated poor clinical outcome in uveal melanoma [82] In osteosarcoma, WIP1 expression was obviously elevated, which promoted the proliferation of osteosarcoma cells through regulating CDKN1A expression [83]. This research further confirmed the potential prognostic value of the identified genes in osteosarcoma.

Several prognostic models of osteosarcoma have been established in previous research. For example, Qu et al. constructed a 5-gene-signature for the prognosis prediction of osteosarcoma based on the super-enhancer-associated genes [84]. The model basing on the biomarkers including RBC, PNI, CRE, Ca²⁺ and LSR in blood presented good performance in predicting the overall survival of osteosarcoma patients [85]. Lin et al. established a predictive model with five differentially expressed genes related to metastasis between the metastatic and non-metastatic samples for the prognosis of osteosarcoma patients [86]. To our knowledge, we are the first to construct a prognostic model of osteosarcoma with autophagy related genes via integrated methods of bioinformatics and machine learning.

Except for risk score, metastasis was also an independent factor for osteosarcoma prognosis. We applied risk score and metastasis as independent prognostic factors to construct a nomogram model for OS prediction in osteosarcoma patients. The result indicated that the nomogram model with two independent factors showed better performance in OS prediction than that with one factor. Furthermore, three immune cell types presented obvious distinction in infiltration proportion between samples with high and low risk scores. The infiltration proportions of activated dendritic cells, M2 macrophages and CD8 T cells were significantly higher in the low risk group than those in the high risk group. Zhang et al. indicated that the osteosarcoma patients with superior survival outcome had higher levels of M2 macrophages, compared with those with inferior survival outcome [87], which was consistent with our finding that the low-risk osteosarcoma patients with improved prognosis had higher proportion of M2 macrophage. Gomez-Brouchet et al. found that CD8 T cells were related to the non-metastatic osteosarcoma [88], and higher infiltration rate of CD8 T cells indicated improved survival outcome [89], showing consistency with our result that the low-risk osteosarcoma patients with superior prognosis had higher infiltration proportion of CD8 T cells. In terms of activated dendritic cells, Wang et al. demonstrated that activated dendritic cell was an independent predictor of osteosarcoma and the model basing on several immune cell types including activated dendritic cell could reliably predict the prognosis of osteosarcoma patients [90], suggesting the potentially important role of activated dendritic cells in osteosarcoma. However, the specific functions of these significantly distinct infiltrating immune cells between the osteosarcoma patients with high and low risk scores and the underlying mechanism still need further research. In addition, analysis of five immune checkpoints in osteosarcoma patients showed that the expressions of CTLA4, TIM3, LAG3, and TIGIT in the samples with low risk scores were markedly elevated compared with those with high risk scores, suggesting that patients with low risk scores might be sensitive to the treatment targeting immune checkpoints.

However, there are some limitations in our study. First, it lacks experimental work, which would be helpful for further validation of the results. Second, the sample size is relatively small. Further research with more samples is needed to better evaluate the performance of the model and elucidate the underlying mechanism in the future.
CONCLUSION

In brief, we identified a 14-autophagy-gene-based prognostic signature in osteosarcoma. Based on these 14 genes associated with autophagy, a model was established and risk score was able to predict the prognosis of osteosarcoma patients independently. Importantly, a nomogram model based on risk score and metastasis was established and exhibited better performance to predict the OS at 1, 3, and 5 years for osteosarcoma patients.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

WQ and KT proposed the research ideas for this article and analyzed and interpreted the data. QY and ML performed the data analysis, methodology and software. DS and WX contributed to the project administration and data presentation. All authors read and approved the final manuscript.

REFERENCES

1. Picci P. Osteosarcoma (Osteogenic Sarcoma). Orphanet J Rare Dis (2007) 2:6. doi:10.1186/1750-1172-2-6
2. Mirabello L, Troisi RJ, and Savage SA. Osteosarcoma Incidence and Survival Rates from 1973 to 2004. Cancer (2009) 115(7):1531–43. doi:10.1002/cncr.24212
3. Singh HB, Singh H, and Chakraborty M. Metastatic Osteosarcoma of the Maxilla. J Laryngol Otol (1978) 92(7):619–22. doi:10.1017/s0022215100085832
4. Wolfe TD, Pillai SPS, Hildreth BE, 3rd, Lanigan LG, Martin CK, Werbeck JL, et al. Effect of Zeolodronic Acid and Amputation on Bone Invasion and Lung Metastasis of Canine Osteosarcoma in Nude Mice. Clin Exp Metastasis (2011) 28(4):377–89. doi:10.1007/s10683-011-9377-9
5. Smeland S, Bielack SS, Whelan J, Bernstein M, Hogendoorn P, Krailo MD, et al. Survival and Prognosis with Osteosarcoma: Outcomes in More Than 2000 Patients in the EURAMOS-1 (European and American Osteosarcoma Study) Cohort. Eur J Cancer (2019) 109:36–50. doi:10.1016/j.ejca.2018.11.027
6. White E, Mehnert JM, and Chan CS. Autophagy, Metabolism, and Cancer. Clin Cancer Res (2015) 21(22):5037–46. doi:10.1158/1078-0432.ccr-15-0490
7. Dong W, and Wang R. [Effects of Resveratrol-Induced Cellular Autophagy in Functional Role of Autophagy-Mediated Proteome Remodeling in Cell
8. Wu S-Y, Lan S-H, Cheng D-E, Chen W-K, Shen C-H, Lee Y-R, et al. Gene Set Enrichment Analysis: a Knowledge-Based Approach for
9. Degenhardt K, Mathew R, Beaudoin B, Bray K, Anderson D, Chen G, et al. Survival and Prognosis with Osteosarcoma: Outcomes in More Than 2000 Patients in the EURAMOS-1 (European and American Osteosarcoma Study) Cohort. Eur J Cancer (2019) 109:36–50. doi:10.1016/j.ejca.2018.11.027
10. Mathew R, Khor S, Hackett SR, Rabinowitz JD, Perlman DH, and White E. Functional Role of Autophagy-Mediated Proteome Remodeling in Cell Survival Signaling and Innate Immunity. Mol Cell (2014) 55(6):916–30. doi:10.1016/j.molcel.2014.07.019
11. Onorati AV, Dyczynski M, Ojha R, and Amaravadi RK. Targeting Autophagy in Cancer. Cancer (2018) 124(16):3307–18. doi:10.1002/cncr.31335
12. O’Farrell JS, and Gordon N. Autophagy in Osteosarcoma. Adv Exp Med Biol (2014) 804:147–60. doi:10.1007/978-3-319-04843-7_8
13. Huang J, Liu K, Yu Y, Xie M, Kang R, Vernon PI, et al. Targeting HMGB1-Mediated Autophagy as a Novel Therapeutic Strategy for Osteosarcoma. Autophagy (2012) 8(2):275–7. doi:10.4161/auto.8.2.18890
14. Huang Y, Liu W, He B, Wang L, Zhang F, Shu H, et al. Exosomes Derived from Bone Marrow Mesenchymal Stem Cells Promote Osteosarcoma Development by Activating Oncogenic Autophagy. J Bone Oncol (2020) 21:100280. doi:10.1016/j.jbono.2020.100280
15. Liu Y, Gu S, Li H, Wang J, Wei C, and Liu Q. SNHG16 Promotes Osteosarcoma Progression and Enhances Cisplatin Resistance by Sponging miR-16 to Upregulate ATG4B Expression. Biochem Biophys Res Commun (2019) 518(1):127–33. doi:10.1016/j.bbrc.2019.08.019
16. Buddingh EP, Kuijjer ML, Duim RAL, Bürger H, Agelepoulos K, Myklebost O, et al. Tumor-infiltrating Macrophages Are Associated with Metastasis Suppression in High-Grade Osteosarcoma: a Rationale for Treatment with Macrophage Activating Agents. Clin Cancer Res (2011) 17(8):2110–9. doi:10.1158/1078-0432.ccr-10-2047
17. Paoloni M, Davis S, Lana S, Withrow S, Sangiorgi L, Picci P, et al. Canine Tumor Cross-Species Genomics Uncovers Targets Linked to Osteosarcoma Progression. BMC Genomics (2009) 10:625. doi:10.1186/1471-2164-10-625
18. Mao D, Zhang Z, Zhao X, and Dong X. Autophagy-related Genes Prognosis Signature as Potential Predictive Markers for Immunotherapy in Hepatocellular Carcinoma. PeerJ (2020) 8:e8383. doi:10.7717/peerj.8383
19. Friedman J, Hastie T, and Tibshirani R. Regularization Paths for Generalized Linear Models via Coordinate Descent. J Stat Softw (2010) 33(1):1–22. doi:10.18637/jss.v033.i01
20. Heagerty PJ, Lumley T, and Pepe MS. Time-dependent ROC Curves for Censored Survival Data and a Diagnostic Marker. Biometrics (2000) 56(2): 337–44. doi:10.1111/j.0006-341x.2000.00357.x
21. Jager KJ, van Dijk PC, Zoccali C, and Dekker FW. The Analysis of Survival Data: the Kaplan-Meier Method. Kidney Int (2008) 74(5):560–5. doi:10.1038/ ki.2008.217
22. Rey D, and Neuhäuser. M. Wilcoxon-signed-rank Test. Springer Berlin Heidelberg (2014).
23. Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, et al. Robust Enumeration of Cell Subsets from Tissue Expression Profiles. Nat Methods (2015) 12(5):453–7. doi:10.1038/nmeth.3337
24. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene Set Enrichment Analysis: a Knowledge-Based Approach for

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.por-journal.com/articles/10.3389/pore.2021.1609782/full#supplementary-material

Supplementary Figure 1 | Schoenfeld residual plot of proportional hazards (PH) assumption. The absissa represented the time, and the ordinate represented the Beta value of each variable. The solid line was the fitted smooth curve, and the dotted line was the standard deviation of 2 units above and below the fitted curve. If the curve deviated by 2 units of standard deviation, then the proportional risk assumption was not satisfied.
66. Qi J, Sun M, Wang Y, and Chen B. Identification and Validation of an Individualized Autophagy-Clinical Prognostic index in Gastric Cancer Patients. *Cancer Cell Int* (2020) 20:178. doi:10.1186/s12935-020-01267-y

67. Ura H, Denno R, Hirata K, Yamaguchi K, and Yasoshima T. Separate Functions of a2β1 and a2β1 Integrins in the Metastatic Process of Human Gastric Carcinoma in vivo. *Metastasis* (2008) 28(10):1001–6. doi:10.1007/s00269-008-9491-1

68. Bauer KM, Watts TN, Buechler S, and Hummon AB. Proteomic and Functional Investigation of the colon Cancer Relapse-Associated Genes NOX4 and ITGA3. *J Proteome Res* (2014) 13(11):4910–8. doi:10.1021/pr500557n

69. Yang W, He M, Zhao J, and Wang Z. Association of ITGA3 Gene Polymorphisms with Susceptibility and Clinicopathological Characteristics of Osteosarcoma. *Med Oncol* (2014) 31(2):826. doi:10.1007/s12032-013-0826-y

70. Elliott B, Millena AC, Matyunina I, Zhang M, Zou J, Wang G, et al. Essential Role of JunD in Cell Proliferation Is Mediated via MYC Signaling in Prostate Cancer Cells. *Cancer Lett* (2019) 448:155–67. doi:10.1016/j.canlet.2019.02.005

71. Gabay M, Li Y, and Felsher DW. MYC Activation Is a Hallmark of Cancer Initiation and Maintenance. *Cold Spring Harb Perspect Med* (2014) 4(6):a014241. doi:10.1101/cshperspect.a014241

72. Wang Y, Wu M-C, Sham JST, Zhang W, Wu W-Q, and Guan X-Y. Prognostic Significance of Myc and AIB1 Amplification in Hepatocellular Carcinoma. *Cancer* (2002) 95(11):2346–52. doi:10.1002/cncr.10963

73. Han G, Wang Y, and Bi W. C-myc Overexpression Promotes Osteosarcoma Cell Invasion via Activation of MEK-ERK Pathway. *Oncol Res Featuring Preclinical Clin Cancer Ther* (2012) 20(4):149–56. doi:10.3727/096504112x13522272332337

74. Morgan NE, Cutriona MB, and Simpson JC. Multitaskin Rab Proteins in Autophagy and Membrane Trafficking: A Focus on Rab33b. *Int J Mol Sci* (2019) 20(16):3916. doi:10.3390/ijms20163916

75. Zhang C, Leng W, Sun C, Lu T, Chen Z, Men X, et al. Urine Proteome Profiling Predicts Lung Cancer from Control Cases and Other Tumors. *Elife* (2018) 7:30208–8. doi:10.1016/j.ijrobio.2018.03.009

76. Zhang M, Hu C, Tong D, Yamaguchi K, and Yasoshima T. Separate Functions of a2β1 and a2β1 Integrins in the Metastatic Process of Human Gastric Carcinoma. *Surg Today* (1998) 28(10):1001–6. doi:10.1007/s00269-008-9491-1

77. Liu J, Xia H, Kim M, Xu L, Li Y, Zhang L, et al. Beclin1 Controls the Levels of p53 by Regulating the Deubiquitination Activity of USP10 and USP13. *Cell* (2011) 147(1):223–34. doi:10.1016/j.cell.2011.08.037

78. Sun J, Li T, Zhao Y, Huang L, Sun H, Wu H, et al. USP10 Inhibits Lung Cancer Cell Growth and Invasion by Upregulating PTEN. *Mol Cell Biochem* (2018) 441(1-2):1–7. doi:10.1007/s11010-017-3170-2

79. Zeng Z, Wu H-X, Zhan N, Huang Y-B, Wang Z-S, Yang G-F, et al. Prognostic Significance of USP10 as a Tumor-Associated Marker in Gastric Carcinoma. *Tumor Biol* (2014) 35(4):3845–53. doi:10.1007/s13277-013-1509-1

80. Tsuysuki S, Takabayashi M, Kawaue M, Kudo K, Watanabe A, Nagata Y, et al. Detection of WIP1 mRNA as an Indicator of Autophagosome Formation. *Jpn J Cancer Res* (2014) 105(5):597–63. doi:10.1111/j.1349-7006.2014.07419

81. D’Arcangelo D, Giampietri C, Muscio M, Satozzi F, Facchiano F, and Facchiano A. WIP1, BAG1, and PEX5 Autophagy-Related Genes Are Relevant Melanoma Markers. *Oxid Med Cell Longev* (2018) 2018:1471682. doi:10.1155/2018/1471682

82. Sun Y, Wang X, Chen B, Xiong L, and Liu Y. Prognostic Value of Autophagy-Related Genes Correlated with Metastasis in Uveal Melanoma Patients (2021).

83. Ran T, Ke S, Song X, Ma T, Xu Y, and Wang M. WIP1 Promotes Osteosarcoma Cell Proliferation by Inhibiting CDKN1A. *Gene* (2021) 8:145537. doi:10.1016/j.gene.2021.145537

84. OuYang Z, Li G, Zhu H, Wang J, Qi T, Qu Q, et al. Construction of a Five-Super-Enhancer-Associated-Genes Prognostic Model for Osteosarcoma Patients. *Front Cell Dev Biol* (2020) 8:598660. doi:10.3389/fcell.2020.598660

85. Tian L, Chen S, Li C, Zhong D, and Xu C. A Novel Prognostic Model to Predict Prognosis of Patients with Osteosarcoma Based on Clinical Characteristics and Blood Biomarkers (2020).

86. Yang B, Su Z, Chen G, Zeng Z, Tan J, Wu G, et al. Identification of Prognostic Biomarkers Associated with Metastasis and Immune Infiltration in Osteosarcoma. *Oncol Lett* (2021) 21(3):180. doi:10.3892/ol.2021.12441

87. Zhang C, Zheng J-H, Lin Z-H, Lv H-Y, Ye Z-M, Chen Y-P, et al. Profiles of Immune Cell Infiltration and Immune-Related Genes in the Tumor Microenvironment of Osteosarcoma. *Aging* (2020) 12(4):3486–501. doi:10.18632/aging.102824

88. Gomez-Brouchet A, Illac C, Gilhodes J, Bouvier C, Aubert S, Guinebretiere JM, et al. CD163-positive Tumor-Associated Macrophages and CD8-Positive Cytotoxic Lymphocytes Are Powerful Diagnostic Markers for the Therapeutic Stratification of Osteosarcoma Patients: An Immunohistochemical Analysis of the Biopsies From the French OS2006 Phase 3 Trial. *Oncoimmunology* (2017) 6(9):e1331193. doi:10.1080/2162402X.2017.1331193

89. Fritzsching B, Fellenberg J, Moskovszky L, Sapi Z, Krenacs T, Machado I, et al. Functional Investigation of the colon Cancer Relapse-Associated Genes WIPI1, BAG1, and PEX3 Autophagy-Related Genes Correlated with Metastasis in Uveal Melanoma Patients.

90. Chen Y, Zhao B, and Wang X. Tumor Infiltrating Immune Cells (TIICs) as a Biomarker for Prognosis Benefits in Patients with Osteosarcoma. *BMC Cancer* (2020) 20(1):1022. doi:10.1186/s12885-020-07536-3