Editorial: Control of plant organ size and shape

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Organ size and shape are the most intuitive morphological characteristics of plants. Dissecting the mechanisms controlling plant organ size and shape could shed light on the nature of plant diversity. In addition, organ size and shape directly associate with crop yield and quality. Identification of key regulators of organ size and shape would provide new genetic targets for crop improvement. This Research Topic aims to illustrate the latest advances of plant organ size and shape regulation.

In this Research Topic, two review articles summarized morphology and organ size control in soybean and the role of salicylic acid (SA) in plant growth, respectively. During the domestication process of soybean, several significant morphology and organ size changes have occurred, such as twisted stems to erect stems and small seeds to large seeds. Zhou et al. summarized genes participated in stem growth habit, leaf size and shape, and seed size and weight in soybean. They also discussed the application of new technologies on the basic research and breeding of soybean and the challenges and hotspots of soybean research in the future. Salicylic acid (SA) is an important phytohormone that not only plays critical roles in plant immunity, but also is involved in plant growth regulation. Li et al. summarized current knowledge about the roles of SA on plant growth. The authors highlighted that SA mediates growth regulation by affecting both cell division and expansion. Particularly, they discussed the interactions of SA with other hormones in plant growth determination. Fiber length is one of the most important economic traits of cotton. In this Research Topic, Pandey et al. provided an opinion article that presents novel perspectives about evolution of fiber length. They expounded...
that concurrent transcriptional dynamics of cytoskeleton-associated structural genes in modern cotton fibers are very useful in understanding the evolutionary recruitment of cell wall-modifying gene clusters for shaping the floral organs and determining fiber length.

Identification of novel regulators of organ size and shape is of great importance in understanding the molecular mechanisms underlying plant growth and development control. In this Research Topic, several novel factors involved in organ size and shape control were characterized. Gao et al. described the role of BrCPS1 (ENT-COPALYL DIPHOSPHATE SYNTHASE 1) in leafy head formation, which is an important agronomic trait in Chinese cabbage (Brassica rapa L. ssp. pekinensis). BrCPS1 affects gibberellin biosynthesis, and mutants of BrCPS1 did not form leafy heads at the heading stage, revealing the critical role of BrCPS1-mediated gibberellin biosynthesis in leafy head formation.

FW2.2 is a classical regulator of fruit size in tomato (Frary et al., 2000). Wang et al. showed that the homolog of FW2.2, MdCNR8 (Cell Number Regulator 8), is sumoylated by the SUMO E3 ligase MdSIZ1 to control organ size in apple, suggesting that FW2.2/CNR8 have a conserved function in fruit size control in tomato and apple. TTG1 participated in trichome formation and anthocyanin accumulation in Arabidopsis (Zhang et al., 2003; Chen et al., 2015). Huang et al. showed that RrTTG1 may promote fruit prickle development through an MBW complex in Rosa roxburghii, which revealed the conserved roles of TTG1 in different species and laid a ground for genetic improvement of prickle-free R. roxburghii. Zhao et al. demonstrated that ectopic expression of poplar PstCYCD1;1 in Nicotiana tabacum regulated flower organ development, which provides new understanding for CYCD function. Besides for these new regulators of organ size and shape, Xie et al. identified a specific mutant (referred as mr1) with a reduced pala in rice, and mapped the causal gene of mr1, which will contribute to understanding of grain formation. Liu et al. identified QTLs associated with agronomic traits in tobacco via a biparental population and an eight-way MAGIC population, which could be applied for breeding new tobacco varieties using molecular marker-assisted selection. Based on a high-quality genetic map constructed by whole genome resequencing, An et al. identified 25 QTLs affecting leaf area of tea plants and developed the related molecular markers, providing useful tools for molecular breeding of tea plant.

Flowering time could affect the final morphology and biomass of plants. It is known that vernalization induces epigenetic silencing of the floral repressor gene FLC (FLOWERING LOCUS C) (Michaels and Amasino, 1999; Whittaker and Dean, 2017; Zhu et al., 2021). In this Research Topic, Maruoka et al. found that the histone demethylases JMJ30 and JM32 could brake vernalization through the activation of FLC, which provides novel insight about the role of repressive histone marks in environmental responses in plants. FT (Flowering locus T) is a key flowering regulator (Turck et al., 2008). Wu et al. introduced an FT homolog from Jatropha curcas into tobacco, and found that FT played crucial role in stem growth.

In addition to the above articles, the other six articles provided some novel information about organ size and shape from other aspects. By performing systematic analyses of the cytological characteristics and underlying mechanism of morphological variation in culms of Phyllostachys nidularia f. farcita (Shidu bamboo), Wang et al. found that the decrease in the total number of internodes and the decrease in dry matter content may contribute to the sharp decline in culm biomass of Shidu bamboo. Langer et al. investigated twist-to-bend ratios and safety factors of petioles with various geometries, sizes and shapes, and provided an equation to calculate the safety factor of naturally horizontally oriented petioles for the first time. KNOX genes encode homeobox transcription factors that play critical roles in determining cell fate in shoot apical meristem (Meng et al., 2020), Zhang et al. performed genome-wide identification of KNOX gene family in Orchidaceae, and provided a comprehensive analysis to uncover the underlying function of KNOX genes in Orchidaceae. Tu et al. performed a comprehensive analysis of the role of long non-coding RNAs, microRNAs, and transcription factors in regulating leaf and flower development in Liriodendron chinense, which laid a foundation for further investigation into the regulatory mechanisms of leaf and flower development in Liriodendron chinense. By CRISPR/Cas9-mediated editing in the cis-regulatory element of KLUH promoter, the fruit weight of tomato was significantly increased (Li et al). By employing mutants with thicker lateral roots, Kawai et al. found that auxin distribution in lateral root primordium affects the size and lateral root diameter in rice. Altogether, this Research Topic provides the latest progresses on the regulation of plant organ size and shape by uncovering new molecular mechanisms and key regulators.

Author contributions

All authors listed have made a substantial and direct contribution to the work and approved it for publication.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
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References

Chen, M., Zhang, B., Li, C., Kulaveerasingam, H., Chew, F. T., and Yu, H. (2015). TRANSPARENT TESTA GLABRA1 regulates the accumulation of seed storage reserves in Arabidopsis. Plant Physiol. 169 (1), 391–402. doi:10.1104/pp.15.00943

Frary, A., Nesbitt, T. C., Grandillo, S., Knaap, E., Cong, B., Liu, J., et al. (2000). fw2.2: a quantitative trait locus key to the evolution of tomato fruit size. Science 289 (5476), 85–88. doi: 10.1126/science.289.5476.85

Meng, L., Liu, X., He, C., Xu, R., Li, Y., and Hu, Y. (2020). Functional divergence and adaptive selection of KNOX gene family in plants. Open Life Sci. 15 (1), 346–363. doi:10.1515/biol-2020-0036

Michaels, S. D., and Amasino, R. M. (1999). FLOWERING LOCUS c encodes a novel MADS domain protein that acts as a repressor of flowering. Plant Cell 11 (5), 949–956. doi:10.1105/tpc.11.5.949

Turck, F., Fornara, F., and Coupland, G. (2008). Regulation and identity of floreigen: FLOWERING LOCUS T moves center stage. Annu. Rev. Plant Biol. 59, 573–594. doi:10.1146/annurev.arplant.59.032607.092755

Whittaker, C., and Dean, C. (2017). The FLC locus: a platform for discoveries in epigenetics and adaptation. Annu. Rev. Cell Dev. Biol. 33, 555–575. doi:10.1146/annurev-cellbio-100616-060546

Zhang, F., Gonzalez, A., Zhao, M., Payne, C. T., and Lloyd, A. (2003). A network of redundant bHLH proteins functions in all TTG1-dependent pathways of Arabidopsis. Development 130 (20), 4859–4869. doi:10.1242/dev.00681

Zhu, P., Lister, C., and Dean, C. (2021). Cold-induced Arabidopsis FRIGIDA nuclear condensates for FLC repression. Nature 599 (7886), 657–661. doi:10.1038/s41586-021-04062-5