The Association of HLA-B*35 and GSTT1 Genotypes and Hepatotoxicity in Thai People Living with HIV

Noppadol Chanhom 1,*, Jiraphun Jittikoon 1, Sukanya Wattanapokayakit 2, Surakameth Mahasirimongkol 2, Angkana Charoenyingwattana 3, Wanvisa Udomsinprasert 1, Usa Chaikledkaew 4, Supharat Suwichapanich 1, Taisei Mushiroda 5, Sasisopin Kiertiburanakul 6, Archawin Rojanawiwat 7, Wittaya Wangsomboonsiri 8, Weerawat Manosuthi 9, Pacharee Kantipong 10, Anucha Apisarnthanarak 11, Wilawan Sangsirinakakul 12, Pawinee Wongprasit 13, Romanee Chaivarith 14, Woraphot Tantisiriwat 15, Somnuek Sungkanuparp 16 and Wasun Chanthira 3

Abstract: Glutathione s-transferase (GST) is a family of drug-metabolizing enzymes responsible for metabolizing and detoxifying drugs and xenobiotic substances. Therefore, deletion polymorphisms of GSTs can be implicated in developing several pathological conditions, including antiretroviral drug-induced liver injury (ARVDILI). Notably, GST polymorphisms have been shown to be associated with ARVDILI risk. However, data on GST polymorphisms in the Thai population are limited. Therefore, this study investigated possible associations between GST genetic polymorphisms and ARVDILI development. A total of 362 people living with HIV (PLHIV) and 85 healthy controls from multiple centers were enrolled. GSTM1 and GSTT1 genetic polymorphisms were determined using polymerase chain reactions. In addition, HLA genotypes were determined using a sequence-based HLA typing method. After comparing GST genotypic frequencies, there was no significant difference between PLHIV and healthy volunteers. However, while observing the PLHIV group,
type was significantly associated with a 2.04-fold increased risk of ARVDILI (95% CI: 1.01, 4.14; p = 0.045). Interestingly, a combination of GSTT1 wild type and HLA-B*35:05 was associated with a 2.28-fold higher risk of ARVDILI (95% CI: 1.15, 4.50; p = 0.02). Collectively, GSTT1 wild type and a combination of GSTT1 wild type plus HLA-B*35:05 were associated with susceptibility to ARVDILI in the Thai population.

**Keywords:** drug-induced liver injury; glutathione s-transferase; human immunodeficiency virus; hepatotoxicity; adverse drug reaction; genetic polymorphisms

### 1. Introduction

Since the discovery of the human immunodeficiency virus (HIV) in 1983 [1], the virus has spread across the globe, becoming one of the most life-threatening diseases. Increased incidences of HIV-related hospitalizations and deaths commonly occur due to prolonged immune deficiency associated with a decrease in CD4\(^+\) lymphocytes, leading to other disease complications. Although receiving antiretroviral (ARV) treatment can help mitigate the disease severity, patients can develop adverse drug reactions, such as hepatotoxicity, commonly known as ARVDILI [2]. Unfortunately, Reister et al. reported that 8.9–10.8% of patients have nevirapine (NVP) hepatotoxicity [3]. Interestingly, genetic polymorphism has been reported as one of the possible factors associated with the development and progression of HIV infection and ARVDILI [4,5], thereby highlighting the possibility of genetic variations as alternative markers for ARVDILI in PLHIV.

GSTs are phase II metabolizing enzymes for drug detoxification through conjugation specific to glutathione (GSH). They are gaining increasing interest as a cellular defender against drugs, carcinogens, and oxidative stress caused by excessive reactive oxygen species (ROS) production. GSTM1 and GSTT1 are particularly fascinating GST genes because these two genes were defined as polymorphic in humans [6]. Given that GSTM1 is located on chromosome 1p13.3, it has been shown that deletion of this gene can impair an individual’s ability to detoxify some carcinogens, ROS, or certain drugs [7,8]. The GSTT1 gene is located on chromosome 22q11.23. This enzyme participates in detoxifying drugs and their conjugation with electrophilic and hydrophobic compounds [9,10]. In PLHIV, it has been recognized that GSTs are responsible for mitigating cellular damage resulting from oxidative stress via conjugating glutathione to ROS [11]. Accordingly, investigating the influence of GSTs’ genetic polymorphisms on ARV-administered patients may provide insight into the safe use of this medication in HIV-infected individuals.

In addition to GST polymorphisms, human leukocyte antigen (HLA) has been reportedly associated with NVP rash and/or hepatitis across different populations [12–16]. For example, in the Thai population, Chantarangsu et al. demonstrated a strong association between HLA-B*35:05 and NVP-induced skin rash [13]. Moreover, a recent study illustrated severe cutaneous adverse reactions were frequently comorbid with hepatitis, especially acute generalized exanthematous pustulosis and drug reaction with eosinophilia and systemic symptoms [17]. It has been hypothesized that there is a significant association between specific HLA genotypes and hepatotoxicity in Thais.

Although a correlation between GST gene deletion and the risk of developing ARVDILI has been explored in the Indian population [5], to the best of our knowledge, no published studies have yet examined associations between GST and HLA polymorphisms with the risk of developing ARVDILI in the Thai population. Accordingly, the objective of this study was to determine whether genetic polymorphisms of GSTs or HLA were associated with ARVDILI risk in Thai PLHIV.
2. Materials and Methods

2.1. Study Subjects

This observational multicentered retrospective case-control study consisted of two study subject groups. The first group included newly diagnosed Thai HIV-positive patients being treated with NVP-based HIV treatment at Ramathibodi Hospital (Bangkok, Thailand), Bamrasnaradura Infectious Diseases Institute (Nonthaburi, Thailand), HRH Princess Maha Chakri Sirindhorn Medical Center (Nakhon-Nayok, Thailand), Thammasat University Hospital (Pathum-Thani, Thailand), Chiangrai Prachanukroh Hospital (Chiangrai, Thailand), Maharat Nakhon Ratchasima Hospital (Nakhon-Ratchasima, Thailand), Sawanpracharak Hospital (Nakhonsawan, Thailand), Buriram Hospital (Buri-Ram, Thailand), or Maharaj Nakorn Chiang Mai Hospital (Chiang-Mai, Thailand). The other was a healthy control group recruited from Ramathibodi Hospital (Bangkok, Thailand) from 2010 to 2012. This study protocol was performed in compliance with the International Guidelines for Human Research Protection, including the Declaration of Helsinki, and the Belmont Report. It was approved by the Institutional Review Board of the Faculty of Dentistry/Faculty of Pharmacy, Mahidol University (IRB No. 2020/PY067). Written informed consent was obtained from all patients before their admission to the study.

Four hundred and forty-seven study participants, including 362 PLHIV and 85 healthy controls, were enrolled (Figure 1). PLHIV were categorized into 50 ARVDILI cases and 312 non-ARVDILI cases. The inclusion criteria for non-ARVDILI PLHIV were as follows: (i) the patient agreed to sign an informed consent form after thoroughly considering the study protocol, (ii) male or female patients aged between 18–70 years old, (iii) a certified laboratory procedure confirmed the patient with HIV-1 infection, and (iv) the patient has never been received any antiretroviral treatment medication. The exclusion criteria for this group were as follows: (i) the patient requested to be excluded from the study, (ii) HIV-infected patient must breastfeed, (iii) the patient has ever received other HIV treatment medication, (iv) the patient’s liver enzymes were more than five times the upper limit of normal. Under Thailand National Guidelines on HIV/AIDS Diagnosis, Treatment, and Prevention 2010 [18], the inclusion criteria for hepatotoxicity were as follows: (i) the HIV patients who presented with hepatotoxicity symptoms, such as nausea, vomiting, fatigue, and myopathy, and had aminotransferase (AST) or alanine aminotransferase (ALT) levels higher than five times the upper limit of normal (ULN); (ii) HIV patients who had AST or ALT level of 5–10 times the ULN without presenting with symptoms. The exclusion criteria for the hepatotoxicity group were as follows: (i) the patient had concomitant administration of other potentially hepatotoxic drugs, as defined by the LiverTox database [19]; (ii) the patient had an underlying disease such as viral hepatitis, liver cirrhosis, hepatoma, or tuberculosis infection. The inclusion criteria for healthy control were as follows: (i) the volunteer agreed to sign an informed consent form after thoroughly considering the study protocol; (ii) male or female individuals aged 18–70 years old; (iii) the individual had never been infected with HIV-1. The exclusion criteria for this group were as follows: (i) the patient had requested to be excluded from the study. All PLHIV were treated under the Thailand National Guidelines on HIV/AIDS Diagnosis, Treatment, and Prevention 2010 [18]. In addition, clinical data and blood samples of the patients were collected by onsite associates and recorded in the project’s specific pre-defined clinical record forms.

![Figure 1. Timeline for patients’ recruiting process. ARVDILI, antiretroviral drug-induced liver injury; HIV, human immunodeficiency virus.](image-url)
2.2. DNA Sample Retrieval and DNA Quantification

As previously described, the leftover DNA samples were retrieved from Ramathibodi hospital [13]. Extracted DNA was analyzed using agarose gel electrophoresis and quantitatively quantified by ultraviolet spectrometer Nanodrop® 2000c (Thermo Scientific, Waltham, MA, USA). Furthermore, each hospital’s automated machine routinely measured all clinical parameters including CD4+, viral load, liver function test, pregnancy test, and HIV drug resistance.

2.3. Genetic Genotyping and Data Retrievals

GSTM1 and GSTT1 genotypes were determined using PCR. β-globin was used as an internal standard. The primers used for GSTM1 and GSTT1 were as follows: GSTM1 forward 5′-GAACTCCCTGAAAAGCTAAAGC-3′; GSTM1 reverse 5′-GTTGGGCTCAAATATACGGTG–3′; GSTT1 forward 5′-TTCTCTACTGGTCCTCACTC-3′; GSTT1 reverse 5′-TCACCGGATCATGCCCAGCA-3′ [20]. The PCR was performed with 20 ng of genomic DNA in a total volume of 20 µL using a T100 Thermal Cycler (BIORAD, Hercules, CA USA) and KAPA2G Fast Multiplex PCR Kit (KAPA Biosystems, Wilmington, MA, USA), with an initial denaturation step of 95 °C for 5 min, followed by 35 cycles of denaturation at 95 °C for 15 s, annealing at 63 °C for 20 s, and extension at 72 °C for 20 s. The PCR products were analyzed on 2% agarose gel pre-stained with ethidium bromide. GSTM1 polymorphism was identified by the presence or absence of bands at 219 base pairs, whereas GSTT1 polymorphism was interpreted by the presence or absence of bands at 459 base pairs (Figure S1). Moreover, both GSTM1 and GSTT1 genotypic data from healthy controls were derived from whole-genome sequencing data using ClinCNV software [21]. HLA genotypic data from a previous study were retrieved for analytical purposes [13].

2.4. Data Analysis

Statistical analyses were executed by Statistical Package for Social Sciences version 22.0 (SPSS, Inc., Chicago, IL, USA). Quantitative parameters, such as demographics and clinical data, normally distributed, were compared between groups using χ² tests and Student’s t-test, where appropriate. Comparisons in non-normally distributed continuous data among each group were evaluated by the Mann–Whitney U test or Kruskal–Wallis H test. Statistical differences in genotypic distributions of cases and controls were undertaken using Pearson’s Chi-square or Fisher’s exact test. The Mantel–Haenszel statistic method was used to combine the effects of genetic polymorphisms.

3. Results

3.1. Demographic and Clinical Characteristics of PLHIV

As detailed in Table 1, the demographic data before ARV treatment, 14 days after ARV treatment, and 42 days after ARV treatment were compared between ARVDILI or HIV+ with hepatotoxicity and non-ARVDILI or HIV+ without hepatotoxicity groups. There were no differences in demographic data, including age, sex, BMI, smoking, alcohol consumption, and received drug regimen between PLHIV with and without ARVDILI. In addition, we observed the viral loads and CD4+ T-cells count both before ARV treatment and 42 days after ARV treatment. The results indicate no difference between the ARVDILI and non-ARVDILI groups. Clinical parameters, including AST and ALT, were significantly lower in PLHIV without ARVDILI than in patients with ARVDILI (p < 0.001).
Table 1. Clinical characteristics of HIV patients with and without ARVDILI.

| Variables | Before ART Treatment | HIV+ without Hepatotoxicity | HIV+ with Hepatotoxicity | p-Value |
|-----------|----------------------|-----------------------------|--------------------------|---------|
| N (%)     | 312 (100.0)          | 50 (100.0)                  |                          |         |
| Age (years) | 36.32 ± 10.29       | 39.26 ± 11.29              | 0.080                    |         |
| Sex (M/F) | 180/132              | 25/25                       | 0.357                    |         |
| BMI (kg/m²) | 22.58 ± 21.52       | 20.52 ± 3.58               | 0.098                    |         |
| Smoking (%) | 70 (23.0)           | 9 (18.4)                    | 0.581                    |         |
| Alcohol (%) | 98 (34.4)           | 16 (35.6)                   | 0.868                    |         |
| CD4+ (cells/mm³) | 144.23 ± 105.11 | 167.49 ± 114.14 | 0.224 |         |
| Viral load (copies/mm³) | 129,027.37 ± 105,260.46 | 105,268.46 ± 170,374.59 | 0.332 |         |
| AST (IU/L) | 31.74 ± 15.47       | 32.76 ± 13.42              | 0.299                    |         |
| ALT (IU/L) | 33.48 ± 18.86       | 36.66 ± 18.33              | 0.235                    |         |

Abbreviation: ALT, alanine aminotransferase; ART, antiretroviral therapy; AST, aspartate aminotransferase; BMI, body mass index; HIV, human immunodeficiency virus.

3.2. Genotypic Distributions of GSTs and HLA-B among Healthy Volunteers, ARVDILI and Non-ARVDILI Groups

When frequencies of GSTM1 and GSTT1 polymorphisms between healthy control and PLHIV were compared, no significant differences in GSTM1 and GSTT1 polymorphisms between groups were observed (Table 2). Although there were significant differences in levels of liver function tests between ARVDILI and non-ARVDILI groups after 14 and 42 days of HIV treatment, as demonstrated in Table 3, we found that GSTT1 wild type was significantly more prevalent in the ARVDILI group, with an odds ratio (OR) of 2.04 (95% CI, 1.01–4.14; p = 0.045) (Table 3). In addition to GST distributions, HLA-B*35:01, HLA-B*35:03, HLA-B*35:05, HLA-B*35:60, and HLA-B*58:01 were not significantly associated with ARVDILI, even though HLA-B*35:01 was predominant in the ARVDILI group.

Table 2. Genotypic distributions of GSTs in healthy volunteers and PLHIV.

| GST        | HIV+ with Hepatotoxicity | HIV+ without Hepatotoxicity | OR (95% CI) | p-Value |
|------------|--------------------------|----------------------------|-------------|---------|
| GSTM1 present | 151 (41.7%)           | 40 (47.1%)                  | 1           | ref.    |
| GSTM1 null   | 211 (58.3%)            | 45 (52.9%)                  | 1.257       | 0.346   |
| GSTT1 present | 237 (65.5%)           | 57 (67.1%)                  | 1           | ref.    |
| GSTT1 null   | 125 (34.5%)            | 28 (32.9%)                  | 1.097       | 0.720   |
| GSTM1 or GSTT1 present | 292 (80.7%) | 71 (83.5%)                  | 1           | ref.    |
| GSTM1 and GSTT1 null | 70 (19.3%)           | 14 (16.5%)                  | 1.268       | 0.467   |

Abbreviation: ARVDILI, antiretroviral drug-induced liver injury; GSTM1, glutathione S-transferase Mu 1; GSTT1, glutathione S-transferase theta 1; HIV, human immunodeficiency virus.

Table 3. Genotypic distributions of GSTs and HLA in PLHIV with and without ARVDILI.

| GST        | HIV+ with Hepatotoxicity | HIV+ without Hepatotoxicity | Odd Ratio (95% CI) | p-Value |
|------------|--------------------------|----------------------------|-------------------|---------|
| N (%)      | 50 (100%)                | 312 (100%)                 | Ref.              |         |
| GSTM1 present | 21 (42.0%)            | 130 (41.7%)                | 1                 | Ref.    |
| GSTM1 null   | 29 (58.0%)              | 182 (58.3%)                | 0.99 (0.54–1.81)  | 0.965   |
| GSTT1 present | 39 (78.0%)            | 198 (63.5%)                | 1                 | Ref.    |
| GSTT1 null   | 11 (22.0%)              | 114 (36.5%)                | 0.49 (0.24–0.99)  | 0.045   |
| GSTM1 or GSTT1 present | 44 (88.0%)        | 248 (79.5%)                | 1                 | Ref.    |
| GSTM1 and GSTT1 null | 6 (12.0%)            | 64 (20.5%)                 | 0.53 (0.22–1.29)  | 0.157   |

Abbreviation: ALT, alanine aminotransferase; ART, antiretroviral therapy; AST, aspartate aminotransferase; BMI, body mass index; HIV, human immunodeficiency virus.
Table 3. Cont.

| GST   | HIV+ with Hepatotoxicity | HIV+ without Hepatotoxicity | Odd Ratio (95% CI) | p-Value |
|-------|--------------------------|-----------------------------|-------------------|---------|
|       | HLA-B                    |                             |                   |         |
| *35:01| 0 (0.0)                  | 6 (1.9)                     | 0.47 (0.03–8.39)  | 1.000   |
| *35:03| 0 (0.0)                  | 4 (1.3)                     | 0.68 (0.04–12.80) | 1.000   |
| *35:05| 2 (4.0)                  | 1 (0.3)                     | 12.92             | 0.052   |
| *35:60| 0 (0.0)                  | 1 (0.3)                     | 2.06 (0.08–51.17) | 1.000   |
| *58:01| 5 (10.0)                 | 45 (90.0)                   | 1.415 (0.59–3.40) | 0.427   |

Abbreviation: ARVDILI, antiretroviral drug-induced liver injury; GSTM1, glutathione S-transferase Mu 1; GSTT1, glutathione S-transferase theta 1; HIV, human immunodeficiency virus; Ref, reference.

3.3. Effects of Multiple Genes on ARVDILI

In this study, we aimed to observe the effect of multiple genes. Therefore, Mantel–Haenszel statistical analysis was applied in order to investigate the additional effect of interesting genes. After pooling the effects of GSTT1 and HLA-B*35:05 using fixed-effect Mantel–Haenszel statistics, results revealed that a combination of GSTT1 wild type and HLA-B*35:05 was significantly associated with a 2.28-fold increased risk of ARVDILI in the Thai population (pooled OR = 2.28; 95% CI: 1.01–4.14; p = 0.02), as illustrated in Figure 2.

Figure 2. Effects of GSTT1 and HLA-B*35:05 gene polymorphisms on ARVDILI combination using a fixed-effect model of Mantel–Haenszel statistics. The cyan squares and lines represent the point estimate of the odd ratio and 95% confidence intervals, while the pink diamonds represents the overall effect of all studies using the fixed and random effect models, respectively.

4. Discussion

HIV is widely recognized as one of the most affecting and life-threatening infectious diseases, impacting millions of individuals globally and hundreds of thousands in Thailand [22]. PLHIVs die and are hospitalized for various causes, including a weakened immune system, which eventually results in an increased risk of contracting opportunistic infections, malignancy, and even suicide [23]. Therefore, ARV therapy is an efficient strategy to mitigate HIV-related mortality to reduce the disease severity and increase survival chances. However, another cause of mortality in PLHIV is adverse drug reactions caused by ARV therapy or HIV treatment, such as ARVDILI [24,25]. In that context, the novel strategy for predicting ARVDILI risk is critical for long-term ARV prescription in various populations. Unfortunately, research on genetic variations associated with ARVDILI is sparse. They may, however, potentially serve as genetic biomarkers for identifying not only severe patients, but also ARVDILI development in PLHIV. GSTs and HLA, which are phase II xenobiotic-metabolizing enzymes and immune regulators, respectively, are exciting candidates for investigating associations between their polymorphisms and ARVDILI.

The present study is the first to uncover that the frequencies of GSTM1 and GSTT1 null genotypes in the Thai healthy population were 52.9% and 32.9%, respectively. Consistent with our results, Kasthurinaidu et al. revealed that GSTM1 deletion frequency was not equally distributed by race [26]. Besides this, another study by Lam et al. demonstrated that
GSTT1 null frequency was not equally distributed by race [27]. In line with our results, we found no significant differences in frequencies of GSTs deletions between healthy controls and PLHIV. This result led us to speculate that Thai people may be more prone to some adverse reactions due to medications, cancers, and oxidative stress [7,8,28] as a result of the high prevalence of GSTM1 null alleles in the Thai population.

Another striking aspect of GSTs’ relevance in HIV infection is ARVDILI. According to a previous study by Singh et al., it has been suggested that homozygous deletions of GSTM1 and GSTT1 may predict the acquisition of hepatotoxicity in PLHIV receiving ARV [5]. To address this hypothesis, our study investigated this matter and found no correlation between GST gene deletions and the risk of developing ARVDILI in Thai PLHIV. The possible reasons for the discrepancy between our findings and Singh et al. might be due to various factors. Firstly, our study included only NVP-based ART patients, while Singh’s study included both nevirapine-based and efavirenz-based ART patients. In addition to the standard regimen, genotypic frequencies of GST deletions in the Indian population were lower than those in the Thai population [26], which may contribute to our inconsequential findings.

On the contrary, we discovered that GSTT1 presence was significantly associated with an increased risk of ARVDILI in the Thai population. As previously mentioned, cells can employ GSTs to conjugate GSH in response to xenobiotics, oxidative stress, or carcinogens. According to Ivanov et al., it has been postulated that prolonged HIV infection may result in massive production of oxidative stress [11]. This mechanism may account for the depletion of GSH in GSTT1 carriers, which are capable of effectively using GSH in response to oxidative stress and exacerbating hepatocellular damage caused by NVP and its metabolites [29]. In addition to the GSTT1-ARVDILI relationship, combining the effects of GSTT1 wild type and HLA-B*35:05 can yield a greater significant level. This result confirmed the findings of Popovic et al., who demonstrated a correlation between immune response and hepatotoxicity due to NVP [30]. Furthermore, according to Srivastava et al., enhanced hyphenation of tissue proteins by reactive NVP metabolites under a GSH-depleted environment might also lead to the initiation of an immune response [31]. Therefore, treating ARVDILI patients with N-acetylcysteine, a precursor to GSH, may help to decrease the severity of this adverse event [32]. Unquestionably, further studies with a larger sample size are necessary to investigate this matter or combine the impacts of genetic polymorphisms in order to clarify our results.

However, it should be noted that this study had some limitations. Firstly, this was a retrospective case-control study that might preclude the determination of cause-and-effect relationships. Therefore, it is recommended that multi-center prospective cohorts are needed to verify any associations. Furthermore, this study could not identify the specific drug-induced liver injury because antiretroviral treatment guidelines indicate all three antiretroviral drugs, two nucleoside reverse transcriptase inhibitors and one non-nucleoside reverse transcriptase inhibitor, can be concurrently delivered to HIV patients, and rechallenge histories are often lacking [18]. On the other hand, the strength of our study is that our findings might be used to support the view that genetic polymorphisms could be used as alternative markers along with a conventional tool for monitoring ARVDILI progression in NVP-administered patients.

5. Conclusions

Our study revealed that GSTT1 and HLA-B*35:05 genotypes are associated with ARVDILI in Thai PLHIV, which may help predict ARVDILI risk in the patients and have predictive value as a genetic biomarker. However, further studies are necessary and encouraged to draw a more decisive conclusion and justify using GSTT1 and HLA-B*35:05 genotypes in combination as an additional genetic biomarker.
Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/jpm12060940/s1, Figure S1. Agarose gel electrophoresis (2% agarose) of PCR amplified products using species-specific PCR primer sets. Lanes 1–24 are examining the presence of GSTM1 and GSTT1 using β-globin as internal control. Lane M, 100 base-pair DNA size marker.

Author Contributions: Conceptualization, J.J., W.C., W.U., U.C., S.M., S.W., T.M. and A.C.; data curation, N.C. and J.J.; formal analysis, N.C., S.M., S.S. (Supharat Suvichapanich), W.U. and J.J.; funding acquisition, J.J., W.C. and S.S. (Supharat Suvichapanich); investigation, N.C., S.W., A.C., W.U. and J.J.; methodology, J.J., W.U., U.C., S.M. and S.S. (Supharat Suvichapanich); project administration, J.J. and W.C.; resources, S.K., A.R., W.W., W.M., P.K., A.A., W.S., P.W., R.C., W.T., S.W., A.C. and S.S. (Somnuek Sungkanuparph); supervision, J.J. and W.C.; validation, J.J., W.U. and S.S. (Supharat Suvichapanich); visualization, N.C.; writing—original draft, N.C.; writing—review and editing, J.J., W.U., S.S. (Somnuek Sungkanuparph) and N.C. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the International Research Network—The Thailand Research Fund (IRN60W003), e-ASIA Joint Research Program (the e-ASIA JRP), Health Systems Research Institute (HSRI), Thailand Center of Excellence for Life Sciences (TCERSL), and the Medical Scholar Program of Mahidol University.

Institutional Review Board Statement: The study was approved by the Institutional Review Board of the Faculty of Dentistry/Faculty of Pharmacy, Mahidol University (IRB No. 2020/PY067).

Informed Consent Statement: Written informed consent has been obtained from the participant(s) to publish this paper.

Data Availability Statement: The data analyzed and generated during the current study are available from the corresponding author on reasonable request.

Acknowledgments: The authors would like to thank the International Research Network team for their encouragement and technical support. In addition, the authors would like to acknowledge the Genomic Medicine Centre, Ramathibodi Hospital, for their samples and clinical data regarding HIV patients.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Barré-Sinoussi, F.; Chermann, J.C.; Rey, F.; Nugeyre, M.T.; Chamaret, S.; Gruest, J.; Dauguet, C.; Axler-Blin, C.; Vézinet-Brun, F.; Rouzioux, C.; et al. Isolation of a T-lymphotropic retrovirus from a patient at risk for acquired immune deficiency syndrome (AIDS). Science 1983, 220, 868–871. [CrossRef] [PubMed]
2. Stern, J.O.; Robinson, P.A.; Love, J.; Lanes, S.; Imperiale, M.S.; Mayers, D.L. A Comprehensive Hepatic Safety Analysis of Nevirapine in Different Populations of HIV Infected Patients. JAIDS J. Acquir. Immune Defic. Syndr. 2003, 34, S21–S33. [CrossRef] [PubMed]
3. Reiter, G.S. Hepatitis in an HIV-infected man. AIDS Clin. Care 1997, 9, 78–81. [PubMed]
4. Parsons, M.; Campa, A.; Lai, S.; Li, Y.; Martinez, J.D.; Murillo, J.; Greer, P.; Martinez, S.S.; Baum, M.K. Effect of GSTM1-Polymorphism on Disease Progression and Oxidative Stress in HIV Infection: Modulation by HIV/HCV Co-Infection and Alcohol Consumption. J. AIDS Clin. Res. 2013, 4, 1002337. [CrossRef] [PubMed]
5. Singh, H.O.; Lata, S.; Angadi, M.; Bapat, S.; Pawar, J.; Nema, V.; Ghate, M.V.; Sahay, S.; Gangakhedkar, R.R. Impact of GSTM1, GSTT1 and GSTP1 gene polymorphism and risk of ARV-associated hepatotoxicity in HIV-infected individuals and its modulation. Pharm. J. 2017, 17, 53–60. [CrossRef]
6. Ebeshi, B.U.; Bolaji, O.O.; Masimirembwa, C.M. Glutathione-S-transferase (M1 and T1) polymorphisms in Nigerian populations. J. Med. Genet. Genom. 2011, 3, 56–60.
7. Soto-Quintana, O.; Zuñiga-Gonzalez, G.M.; Ramírez-Patiño, R.; Ramos-Silva, A.; Figuera, L.E.; Carrillo-Moreno, D.I.; Gutiérrez-Hurtado, I.A.; Puebla-Pérez, A.M.; Sánchez-Llamas, B.; Gallegos-Arreola, M.P. Association of the GSTM1 null polymorphism with breast cancer in a Mexican population. Genet. Mol. Res. 2015, 14, 13066–13075. [CrossRef]
8. Chanhom, N.; Udomsinprasert, W.; Chaikeldkaew, U.; Mahasirimongkol, S.; Wattanapokayakit, S.; Jittikoon, J. GSTM1 and GSTT1 genetic polymorphisms and their association with antituberculosis drug-induced liver injury. Biomed. Rep. 2020, 12, 153–162. [CrossRef]
9. Shokeer, A.; Mannervik, B. Residue 234 is a master switch of the alternative-substrate activity profile of human and rodent theta class glutathione transferase T1-1. Biochim. Biophys. Acta 2010, 1800, 466–473. [CrossRef]
10. Tars, K.; Larsson, A.K.; Shokeer, A.; Olin, B.; Mannervik, B.; Kleywegt, G.J. Structural basis of the suppressed catalytic activity of wild-type human glutathione transferase T1-1 compared to its W234R mutant. *J. Mol. Biol.* 2006, 355, 96–105. [CrossRef]

11. Ivanov, A.V.; Valuiev-Elliston, V.T.; Ivanova, O.N.; Kochetkov, S.N.; Starodubova, E.S.; Bartosch, B.; Isagulians, M.G. Oxidative Stress during HIV Infection: Mechanisms and Consequences. *Oxid. Med. Cell. Longev.* 2016, 2016, 8910396. [CrossRef] [PubMed]

12. Martin, A.M.; Nolan, D.; James, I.; Cameron, P.; Keller, J.; Moore, C.; Phillips, E.; Christiansen, F.T.; Mallal, S. Predisposition to nevirapine hypersensitivity associated with HLA-DRB*10101 and abrogated by low CD4 T-cell counts. *AIDS* 2005, 19, 97–99. [CrossRef] [PubMed]

13. Chantarangsu, T.; Mushiroda, T.; Mahasirimongkol, S.; Kiertiburanakul, S.; Sungkanuparp, S.; Manosuthi, W.; Tantisirirawit, W.; Charoenyingwattana, A.; Sura, T.; Chatrattita, W.; et al. HLA-B*3505 allele is a strong predictor for nevirapine-induced skin adverse drug reactions in HIV-infected Thai patients. *Pharm. Genom.* 2009, 19, 139–146. [CrossRef]

14. Gatanaga, H.; Yazaki, H.; Tanuma, J.; Honda, M.; Genka, I.; Teruya, K.; Tachikawa, N.; Kikuchi, Y.; Oka, S. HLA-Cw8 primarily associated with hypersensitivity to nevirapine. *AIDS* 2007, 21, 264–265. [CrossRef]

15. Littera, R.; Carcassi, C.; Masala, A.; Piano, P.; Serra, P.; Ortu, F.; Corso, N.; Casula, B.; La Nasa, G.; Contu, L.; et al. HLA-dependent hypersensitivity to nevirapine in Sardinian HIV patients. *AIDS* 2006, 20, 1621. [CrossRef]

16. Yuan, J.; Guo, S.; Hall, D.; Cammott, A.M.; Jayadev, S.; Distel, M.; Storfer, S.; Huang, Z.; Moostikapun, P.; Ruxrungtham, K.; et al. Toxicogenomics of nevirapine-associated cutaneous and hepatic adverse events among populations of African, Asian, and European descent. *AIDS* 2011, 25, 1271–1280. [CrossRef]

17. Esmaeilzadeh, H.; Farjadian, S.; Alyasian, S.; Nemati, H.; Nabavizadeh, H.; Esmaeilzadeh, E. Epidemiology of Severe Cutaneous Adverse Drug Reaction and Its HLA Association among Pediatrics. *Iran. J. Pharm. Res.* 2019, 18, 506–522.

18. Phanuphak, P.; Leechawengwongs, M.; Siraprapasiri, T.; Chantratita, W.; Techasathit, W.; Teeraratkul, A.; Chokephaibulkit, K. Oxidative stress in viral hepatitis and AIDS. *Exp. Mol. Pathol.* 2019, 122–132. [CrossRef]

19. Hoofnagle, J.H.; Navarro, V.J. LiverTox: Clinical and Research Information on Drug-Induced Liver Injury; National Institute of Diabetes and Digestive and Kidney Diseases: Bethesda, MD, USA, 2012.

20. Kiran, B.; Karkucak, M.; Ozan, H.; Yakut, S.; Ozekan, K.; Sag, S.; Ture, M. GST (GSTM1, GSTT1, and GSTP1) polymorphisms in the genetic susceptibility of Turkish patients to cervical cancer. *J. Gynecol. Oncol.* 2010, 21, 169–173. [CrossRef]

21. Demidov, G.; Ossowski, S. ClinCNV: A Tool for Large-Scale CNV and CNA Detection. 2019; Unpublished work.

22. UNAIDS Data 2020. Available online: https://www.unaids.org/en/resources/documents/2020/unaids-data (accessed on 6 January 2021).

23. Nishijima, T.; Inaba, Y.; Kawasaki, Y.; Tsukada, K.; Teruya, K.; Kikuchi, Y.; Gatanaga, H.; Oka, S. Mortality and causes of death in people living with HIV in the era of combination antiretroviral therapy compared with the general population in Japan. *AIDS* 2020, 34, 913. [CrossRef]

24. Kovari, H.; Sabin, C.A.; Ledergerber, B.; Ryom, L.; Worm, S.W.; Smith, C.; Phillips, A.; Reiss, P.; Fontas, E.; Petoumenos, K.; et al. Antiretroviral drug-related liver mortality among HIV-positive persons in the absence of hepatitis B or C virus coinfection: The data collection on adverse events of anti-HIV drugs study. *Clin. Infect. Dis.* 2013, 56, 870–879. [CrossRef]

25. Kowalska, J.D.; Friis-Møller, N.; Kirk, O.; Bannister, W.; Mocroft, A.; Sabin, C.; Reiss, P.; Gill, J.; Lewden, C.; Phillips, A.; et al. The Coding Causes of Death in HIV (CoDe) Project: Initial results and evaluation of methodology. *Epidemiology* 2011, 22, 516–523. [CrossRef] [PubMed]

26. Kashturinaidu, S.P.; Ramasamy, T.; Ayyavoo, J.; Dave, D.K.; Adroja, D.A. GST M1-T1 null allele frequency patterns in geographically assorted human populations: A phylogenetic approach. *PLoS ONE* 2015, 10, e0118660. [CrossRef] [PubMed]

27. Lam, Y.W.F.; Cavallari, L.H. Chapter 1—Principles of Pharmacogenomics: Pharmacokinetic, Pharmacodynamic, and Clinical Implications. In *Pharmacogenomics*; Lam, Y.-W.F., Cavallari, L.H., Eds.; Academic Press: San Diego, CA, USA, 2013; pp. 1–44.

28. Ntais, C.; Polycarpou, A.; Ioannidis, J.P. Association of GSTM1, GSTT1, and GSTP1 gene polymorphisms with the risk of prostate cancer: A meta-analysis. *Cancer Epidemiol. Biomark. Prev.* 2005, 14, 176–181.

29. Stehbens, W.E. Oxidative stress in viral hepatitis and AIDS. *Exp. Mol. Pathol.* 2007, 74, 121–132. [CrossRef] [PubMed]

30. Popovic, M.; Caswell, J.L.; Mannargudi, B.; Shenton, J.M.; Uetrecht, J.P. Study of the sequence of events involved in nevirapine-induced skin rash in Brown Norway rats. *Chem. Res. Toxicol.* 2006, 19, 1205–1214. [CrossRef] [PubMed]

31. Srivastava, A.; Lian, L.-Y.; Maggs, J.L.; Chaponda, M.; Pirmohamed, M.; Williams, D.P.; Park, B.K. Quantifying the metabolic activation of nevirapine in patients by integrated applications of NMR and mass spectrometries. *Drug. Metab. Dispos.* 2010, 38, 122–132. [CrossRef]

32. Claes, P.; Wintzen, M.; Allard, S.; Simons, P.; De Coninck, A.; Lacor, P. Nevirapine-induced toxic epidermal necrolysis and toxic hepatitis treated successfully with a combination of intravenous immunoglobulins and N-acetylcysteine. *Eur. J. Intern. Med.* 2004, 15, 255–258. [CrossRef]