Increased Sodium Concentration in Substantia Nigra in Early Parkinson’s Disease: A Preliminary Study With Ultra-High Field (7T) MRI

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Pathophysiology of idiopathic Parkinson’s disease (iPD) is complex and still misunderstood. At a time when treatments with disease-modifying potential are being developed, identification of early markers of neurodegeneration is essential. Intracerebral sodium accumulation could be one of them. Indeed, it may be in relation to the mitochondrial dysfunction that early exists in iPD. For the first time, we used brain sodium (23Na) MRI to explore sodium concentration changes that have already been reported to be related to neurodegeneration in other diseases. We prospectively included 10 iPD patients (mean age 52.2 ± 5.9 years-old) with motor symptoms that started <36 months before inclusion and 12 healthy subjects (mean age 53 ± 6.4 years-old). Patients were scanned in OFF medication state by using proton (1H) and 23Na MRI at 7T. We then extracted quantitative Total Sodium Concentration (TSC) from five regions of interest [substantia nigra (SN), putamen, caudate nucleus, pallidum, thalamus] and in one region supposed to be relatively spared in the first stages of the disease [cortical gray matter (neocortex)]. Potential atrophy in these structures was also investigated with 1H MRI. Relative to healthy subjects, iPD patients showed higher TSC in the SN (43.73 ± 4.64 vs. 37.72 ± 5.62, p = 0.006 after Bonferroni correction). A trend of increase in sodium concentrations was found within the pallidum (45.80 ± 4.19 vs. 41.07 ± 4.94, p = 0.017), putamen (48.65 ± 4.58 vs. 43.66 ± 5.04, p = 0.041) and the cortical gray matter (56.34 ± 3.92 vs. 50.81 ± 5.50, p = 0.021). No significant brain atrophy was found in patients compared to controls. Thus, alteration of sodium homeostasis in the SN in the absence of atrophy could be considered as a potential early marker of cellular dysfunction in iPD.

Keywords: Parkinson’s disease, neurodegeneration, sodium, biomarker, ultra high field magnetic resonance imaging

INTRODUCTION

Idiopathic Parkinson’s disease (iPD) is a neurodegenerative disease affecting the central nervous system and is a major cause of disability and dependence. By 2030, the number of parkinsonian patients could increase by 56%, with 1 in 120 people over the age of 45 suffering from the disease (1). To date, there is no treatment to slow or stop the progression of the disease. At a time
when treatments with disease-modifying potential are being developed (for example anti-synuclein antibodies or iron chelators) (2), identification of early markers of neurodegeneration is essential and these are urgently lacking.

Although α-synuclein misfolding is part of a significant upstream pathway leading to dopaminergic degeneration (3, 4), mitochondrial dysfunction is another main upstream pathway to parkinsonism (5). Among the deleterious consequences of mitochondrial dysfunction, increased intracellular sodium (Na+) concentration has been reported (6) and reflects an alteration of the cellular homeostasis. Interestingly, Na+ concentration changes in the brain have already been related to neurodegeneration, for example in Huntington’s Disease (7) or Amyotrophic Lateral Sclerosis (8) with brain sodium (23Na) MRI which allows the exploration of sodium distribution through a non-invasive procedure. Thus, we hypothesize that sodium accumulation could be a potential early biomarker of iPD, especially in the substantia nigra (SN) where impairment of regulation of mitochondrial DNA copy number occurs and is thought to be related to neurodegeneration in iPD (9). This pathophysiological approach could lead to the identification of new therapeutic targets such as the Na+ K+ Cl− cotransporter isoform 1 (NKCC1) importer antagonist bumetanide, which has been reported to attenuate motor effects of dopamine deprivation (10).

In this exploratory study, we propose to study brain sodium concentrations using ultra-high field (7T) MRI in iPD patients relative to healthy subjects.

MATERIALS AND METHODS

Population Studied

We prospectively recruited 10 patients with iPD between February and November 2020. All patients met the iPD MDS diagnostic criteria (11) with motor symptoms that started <36 months before inclusion.

Twelve age- and gender-matched healthy subjects were also recruited prospectively during the same period.

Written informed consent was obtained from all participants. The study was approved by the local Ethics Committee (Comité de Protection des Personnes Sud Méditerranée 1), in accordance with the Declaration of Helsinki.

Clinical Data Collection

All patients had been examined in a standardized manner within a half-day by a movement disorders specialist working in the Department of Neurology and Movement Disorders, Marseille University Hospital. We collected scores using different scales to evaluate the severity of symptoms [i.e., UPDRS III, Schwab & England, Hoehn & Yahr, SCOPA-AUT and REM-sleep behavior disorders (RBD) Screening Questionnaire (12)], cognition (MoCA, Lexical and semantic fluency, Benton Judgment of Line Orientation Test-15 items), mood disorder [Hospital Anxiety and Depression (HAD) scale] and apathy (Starkstein motivation scale). Levodopa equivalent dose was calculated (13). The clinical data are summarized in Table 1. After receiving the patient’s agreement, dopaminergic treatment was suspended 72 h before scanning to evaluate the severity of the disease and levodopa responsiveness (good if >50%/bad). Indeed, the effect of exogenous dopaminergic intake on a possible modification of sodium concentrations and metabolism is unknown.

MRI Acquisition and Post-processing

MRI acquisition was performed with a 7-T Magnetom system (Siemens, Erlangen, Germany). For 23Na MRI exploration, we used a dual-tuned 23Na/1H QED birdcage coil and a multi-echo density adapted 3D projection reconstruction pulse sequence (TR = 120 ms, 5,000 spokes, 384 radial samples per spoke, 3 mm nominal isotropic resolution; to ensure a sufficient number and distribution of TEs, while taking into account the 5 ms readout of the sequence, we applied the sequence three times within the same exam in order to obtain 24 TEs ranging from 0.2 ms to 70.78 ms: 1st acquisition: 0.20 - 9.70 - 19.20 - 28.70 - 38.20 - 47.70 - 57.20 - 66.70 ms, 2nd acquisition: 1.56 - 11.06 - 20.56 - 30.06 - 39.56 - 49.06 - 58.56 - 68.06 ms, 3rd acquisition: 4.28 - 13.78 - 23.28 - 32.78 - 42.28 - 51.78 - 61.28 - 70.78 ms, total acquisition time = 3 × 10 min) adapted from (14). Six tubes with known sodium concentrations (from 25 to 100 mmol/L within 2% of agar gel) were placed within the field of view to

| TABLE 1 | Patients’ characteristics. |
|-----------------|--------------------------|
|                | Mean | SD |
| Age (years)    | 55.2 | 5.9 |
| Disease duration (months) | 20.3 | 11.2 |
| Schwab & England | 97 | 4.8 |
| Hoehn & Yahr    | 1    | 0   |
| UPDRS III       | 12.9 | 3.8 |
| MoCA            | 27.9 | 1.3 |
| Total Lexical fluency (number in 2 min) | 20.2 | 7.4 |
| Total Semantic fluency (number in 2 min) | 28.8 | 3.9 |
| Benton Test (15-items) | 30 | 2.7 |
| Starkstein motivation scale | 19.9 | 5.7 |
| HAD             | 11.8 | 5.9 |
| SCOPA-AUT       | 9.7  | 7.8 |
| RBD screen questionnaire | 3.7 | 2.2 |
| Levodopa (md/day) | 80 | 188.9 |
| Levodopa Equivalent Dose (mg/day) | 175.5 | 259.2 |
| Gender          | 8 M (80%) and 2 F (20%) |
| Orthostatic hypotension | 2 (20%) |
| Dopamine agonist | 4 (40%) |
| Levodopa        | 3 (30%) |
| IMAO-B          | 6 (60%) |
| ICOMT           | 0 (0%) |
| Anticholinergic | 0 (0%) |
| Levodopa Responsivness | Good for 10 (100%) |

UPDRS, Unified Parkinson’s Disease Rating Scale; MoCA, Montreal Cognitive Assessment; HAD, Hospital Anxiety and Depression scale; SCOPA-AUT, Scales for Outcomes in Parkinson’s disease - AUTonomy; RBD, REM Sleep Behavior Disorder Screening Questionnaire; SD, standard deviation.
serve as a reference for quantification (14). A 32-channel phased-array $^1$H head coil (1Tx/32 Rx; Nova) was used to acquire a sub-millimeter $^1$H three-dimensional Magnetization-Prepared 2 Rapid Acquisition Gradient-Echoes (MP2RAGE) sequence (TR = 5,000 ms/TE = 3 ms/T1 = 900 ms/T2 = 2,750 ms, 256 slices, 0.6 mm isotropic resolution, acquisition time = 10 min 12 s).

Sodium images were reconstructed offline, fitted using a bi-exponential model, and finally normalized relative to signal from reference tubes to obtain quantitative Total Sodium Concentration (TSC) maps of the whole brain. We followed the methodology previously described in (14). This multi-echo approach allows to account for the complex relaxation processes of $^{23}$Na through a bi-exponential fitting enabling to determine the ordinate at TE = 0 ms of the curve corresponding to the TSC. In the present preliminary study with limited sample size and in order to minimize the statistical type II error, we decided to focus the analysis solely on TSC, the metric mainly expressed in the literature. Increase in sample size will allow to evaluate relative variations of short and long T2* pool fractions of $^{23}$Na. The first $^{23}$Na echo (TE = 0.20 ms) and the $^1$H images were coregistered using a rigid transformation (15). $^1$H images were segmented into Gray Matter (GM), White Matter (WM) and cerebrospinal fluid (CSF) (0.9 tissue probability threshold) using the Statistical Parametric Mapping 12 “New Segment” tool (16), into deep gray matter (DGM) using FSL-FIRST tool (accumbens, amygdala, caudate, hippocampus, pallidum, putamen and thalamus) (17) and into substantia nigra by registering the $^1$H images to the Montreal Neurologic Institute (MNI) 152 space and bringing back the SN-AAL3 mask to the subject space (15). without resectioning (SPM8; https://www.fil.ion.ucl.ac.uk/spm/software/spm8/). The $^1$H images were then segmented and normalized into the MNI template, and the resulting transformation was applied to the quantitative $^{23}$Na maps. Quantitative TSC values were extracted from five regions of interest known to be early impaired in iPD (SN, putamen, caudate nucleus, pallidum, thalamus) and in one region supposed to be relatively spared in the first stages of the disease, the cortical gray matter (neocortex). Cerebral volumes were normalized for head size using the intracranial volume (18).

**Statistical Analysis**

Categorical variables are presented as numbers and percentages, and the quantitative results as a mean with standard deviation. Comparisons between groups were made with Chi2 or Fisher tests for categorical data and Wilcoxon for continuous data, as appropriate. Correlations between clinical features and MRI parameters were looked for with Spearman correlation analysis. A two-sided p-value < 0.05 was considered statistically significant in univariate analysis and p < 0.008 after Bonferroni correction. Statistical analyses were performed using JMP software JMP 9.0.1 (SAS Institute).

**RESULTS**

**Patients’ and Healthy Subjects’ Characteristics**

Eight males and 2 females with iPD were included in this study with a mean age of 55.2 ± 5.9 years-old and a mean disease duration of 20.3 ± 11.2 months. All of them were completely independent in their daily life as shown by scores of Schwab & England (97 ± 4.8) and Hoehn & Yahr (1 ± 0). No cognitive impairment was identified (MoCA score at 27.9 ± 1.3). Mean UPDRS III OFF anti-parkinsonian drugs was 12.9 ± 3.8. Clinical data are summarized in Table 1.

iPD patients were comparable with healthy subjects concerning age (mean age of 53.0 ± 6.4, p = 0.35) and gender (6 F and 6 M, p = 0.20).

**Sodium Brain Accumulation and Brain Volume**

As shown in Table 2, Figures 1 and 2, relative to healthy subjects, patients with iPD showed higher TSC in the SN (43.73 ± 4.64 vs. 37.72 ± 5.62, p = 0.006 after Bonferroni correction). A trend of increase in sodium concentrations was found within the pallidum (45.80 ± 4.19 vs. 41.07 ± 4.94, p = 0.017), putamen (48.65 ± 4.58 vs. 43.66 ± 5.04, p = 0.08) and caudate nucleus (54.22 ± 4.36 vs. 52.35 ± 5.50, p = 0.49) (Table 1).

**Table 2: Mean Total sodium concentration (TSC) (wet tissue volume, mmol/l) and normalized volumes of brain regions of interest.**

| Brain areas | Group | TSC ± SD | p     | Normalized volume ± SD | p     |
|-------------|-------|----------|-------|------------------------|-------|
| Substantia Nigra | iPD    | 43.73 ± 6.4 | 0.006* | 0.07 ± 0.01            | 0.149 |
|              | Healthy subjects | 37.72 ± 6.2 |       | 0.08 ± 0.01            |       |
| Putamen     | iPD    | 48.65 ± 4.5 | 0.041 | 0.62 ± 0.14            | 0.187 |
|              | Healthy subjects | 43.66 ± 5.0 |       | 0.69 ± 0.11            |       |
| Caudate nucleus | iPD    | 54.22 ± 4.3 | 0.055 | 0.48 ± 0.07            | 0.553 |
|              | Healthy subjects | 52.35 ± 5.5 |       | 0.50 ± 0.07            |       |
| Pallidum    | iPD    | 45.80 ± 4.1 | 0.017 | 0.23 ± 0.03            | 0.461 |
|              | Healthy subjects | 41.07 ± 4.9 |       | 0.24 ± 0.04            |       |
| Thalamus    | iPD    | 50.60 ± 5.9 | 0.129 | 0.97 ± 0.14            | 0.468 |
|              | Healthy subjects | 46.29 ± 5.1 |       | 1.01 ± 0.15            |       |
| Cortical Gray Matter | iPD    | 56.34 ± 3.9 | 0.021 | 42.53 ± 5.67           | 0.714 |
|              | Healthy subjects | 50.81 ± 5.5 |       | 43.37 ± 4.98           |       |

*pSurvive after multiple comparisons. TSC, Total sodium concentration; SD, standard deviation; iPD, idiopathic Parkinson’s Disease.*
vs. 43.66 ± 5.04, \( p = 0.041 \)) and the cortical gray matter (56.34 ± 3.92 vs. 50.81 ± 5.50, \( p = 0.021 \)).

No significant difference in ROI normalized volumes was found between patients and controls.

There was no correlation between clinical features, levodopa equivalent doses and MRI parameters.

**DISCUSSION**

Despite the limited number of subjects included in this study, we have been able to demonstrate an accumulation of sodium in patients with iPD. This accumulation is significant in the SN at this early stage of the disease.

SN is recognized as the origin of the nigro-basal ganglia-thalamic-cortical motor pathway dysfunction in iPD. Indeed, the most consistent finding of the neuropathology of iPD is a loss of dopaminergic neurons in the SN pars compacta which modulates activity of neurons in the basal ganglia. In the end, inadequate facilitation of the corticospinal tracts produces akinesia and bradykinesia which are part of parkinsonian syndrome (19). Pathophysiology of iPD is still misunderstood but along with \( \alpha \)-synuclein misfolding, mitochondrial dysfunction is considered to have an important role in neurodegeneration (5). Changes in mitochondrial gene expression, well-documented in the SN (9), seem to contribute to cellular energy failure, which in turn leads to loss of function of \( \text{Na}^+ / K^+ \) ATPase and impaired ability of the cell to maintain resting potential and to export \( \text{Na}^+ \) (6).
**FIGURE 2** | Violin plots of Mean Total sodium concentration (TSC) (wet tissue volume, mmol/l) for iPD and HC. *Survive after multiple comparisons. iPD, idiopathic Parkinson’s Disease; HC, Healthy Controls; TSC, Total sodium concentration.
Then, the significant sodium accumulation in the SN is believed to be a sign of early dopaminergic neurons dysfunction in iPD. Also, the trend of increase in sodium concentrations within the pallidum corroborates the early SN-basal ganglia neural circuits impairment in iPD but more investigations are warranted to support these results.

The trend of sodium accumulation in cortical gray matter was not fully expected at this stage, in the absence of obvious cognitive impairment. However, early changes in personality and mild cognitive impairment have been described in iPD (20). Characteristically, these changes are often subtle at the beginning and difficult to detect without specific neuropsychological tests. Deficits mainly affect executive function including working memory and visuospatial capacity. Here, 3 patients had a MoCA score at 26/30 which is the limit for normal values (21). Lewy pathology, hallmark of iPD, in the cerebral cortex does not correlate with cognitive impairment. Nevertheless, recent studies have shown abnormal mitochondria content and function, and increased oxidative stress and oxidative responses in the frontal cortex in iPD (22) that could be related to the tendency of sodium accumulation associated with early cortical dysfunction.

A variety of consequences of Na⁺ accumulation may be relevant in PD pathophysiology. Mitochondrial dysfunction, ionic disturbances and neurodegeneration are interrelated biological processes already described in PD (5). Mitochondrial dysfunction may result in intraneuronal sodium overload through reversed activity of the sodium-calcium exchanger and axonal calcium import (6). In turn, this calcium overload may force the opening of mitochondrial permeability transition pore, leading to retrograde electron flux through the electron transport chain, resulting in increased reactive oxygen species production, release of cytochrome c, and activation of apoptosis (23, 24) particularly in SN dopaminergic neurons which project their axons on basal ganglia nuclei (25). We hypothesize that this accumulation of Na⁺ may affect the activity of other structures in the SN-basal ganglia-thalamic-cortical circuit. For example, sodium ions are essential for spike generation in midbrain dopaminergic neurons (26) and pallidum neurons (27). A computational modeling study recently demonstrated a reduced availability of sub-threshold activated sodium and potassium channels, resulting in a decrease in firing regularity in pallidum neurons (28). Also, dopamine transporter (DAT) is a sodium-coupled transmembrane protein that mediates the reuptake of dopamine from the synaptic cleft, and is localized to presynaptic nigrostriatal terminals (29). It operates by coupling transport of Na⁺ along its concentration gradient from the extracellular compartment to the intracellular compartment allowing the transport of the substrate in the same way (30). Intracellular Na⁺ overload might deteriorate its functioning and worsen neurological symptoms.

From a therapeutic point of view, interestingly, it has been shown that the neuronal Na⁺ K⁺ Cl⁻ cotransporter isoform 1 (NKCC1) importer antagonist bumetanide, which reduces intracellular Cl⁻ levels (but also reduces Na⁺ and K⁺ intracellular import) attenuates motor effects of dopamine deprivation by restoring GABAergic inhibition including the cortico-striatal pause-rebound response (31). Damier et al. (10), reported an improvement of iPD motor symptoms in the 4 patients treated with bumetanide which also improved gait and freezing in 2 of these patients calling for double-blind, placebo-controlled, randomized trials to confirm the therapeutic efficacy of bumetanide. NKCC1 importer antagonist has been postulated to have also a neuroprotective effect in iPD through its action on astrocytes, microglia and oligodendrocytes by limiting the intracellular accumulation of Na⁺, Cl⁻ and K⁺ (32).

We did not find any significant atrophy when we compared the volumes of brain structures in iPD patients and healthy subjects although sodium concentration was abnormally elevated in patients. This suggests that abnormal elevated sodium concentration more likely reflects cellular dysfunction rather than cell death. This finding is in favor of the hypothesis that elevated sodium concentration could be a potential marker of early processes of neurodegeneration before neuronal death.

Our study has some limitations. The main one is the limited number of participants. This preliminary study has therefore allowed us to demonstrate the feasibility and relevance of assessing brain sodium accumulation in iPD. Unfortunately, several structures implicated in iPD pathophysiology are too small to be evaluated individually in sodium imaging such as locus coeruleus, subthalamic-nucleus or SN pars compacta and SN pars reticulata separately due to the limited spatial resolution of sodium MRI even at high field.

**CONCLUSION**

To our knowledge, this is the first study using sodium MRI at 7T providing evidence of abnormal sodium concentration in SN which is a region known to be particularly impaired early in iPD. The evidence of such an alteration of sodium homeostasis in the absence of atrophy could be considered as a potential marker of early processes of cellular dysfunction and neurodegeneration before neuronal death in iPD. Further investigations are needed to confirm these results and to explore mapping of sodium homeostasis that might be different in other parkinsonian syndromes.

**DATA AVAILABILITY STATEMENT**

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

**ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by Comité de Protection des Personnes Sud Méditerranée 1. The patients/participants provided their written informed consent to participate in this study.

**AUTHOR CONTRIBUTIONS**

SG: conception and design of the study, acquisition and analysis of data, drafting the manuscript and figures, and
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REFERENCES

1. Wanneveich M, Moisan F, Jacqmin-Gadda H, Elbaz A, Joly P. Projections of prevalence, lifetime risk, and life expectancy of Parkinson’s disease (2010-2030) in France: projections of PD in France. Mov Dis. (2018) 33:1449–55. doi: 10.1002/mds.27447
2. Silva M, Caro V, Guzmán C, Perry G, Areche C, Cornejo A, a-Synuclein and tau, two targets for dementia. Stud Nat Prod Chem. (2021) 67:1–25. doi: 10.1016/B978-0-12-819483-6-00001-1
3. Dauer W, Przedborski S. Parkinson’s disease: mechanisms and models. Neuron. (2003) 39:889–909. doi: 10.1016/S0896-6273(03)00568-3
4. Schapira AH, Jenner P. Etiology and pathogenesis of Parkinson’s disease. Mov Disord. (2011) 26:1049–55. doi: 10.1002/mds.23732
5. Abou-Sleiman PM, Muqit MMK, Wood NW. Expanding insights of mitochondrial dysfunction in Parkinson’s disease. Nat Rev Neurosci. (2006) 7:207–19. doi: 10.1038/nrn1868
6. Waxman SG. Axonal conduction and injury in multiple sclerosis: the role of sodium channels. Nat Rev Neurosci. (2006) 7:932–41. doi: 10.1038/nrn2023
7. Reetz K, Romanzetti S, Dogan I, Saß C, Werner CJ, Schiefer et al. Increased brain tissue sodium concentration in Huntington’s disease – a sodium imaging study at 4T. NeuroImage. (2012) 63:517–24. doi: 10.1016/j.neuroimage.2012.07.009
8. Grapperon A-M, Ridley B, Verschueren A, Maarouf A, Conforti et al. Distribution of brain sodium long and short relaxation times and concentrations: a multi-echo ultra-high field 23Na-MRI study. Sci Rep. (2018) 8:4357. doi: 10.1038/s41598-018-22711-0
15. Avants BB, Tsustion NJ, Stauffer M, Song G, Wu B, Gee JC. The insight Toolkit image registration framework. Front Neuroinform. (2014) 8:44. doi: 10.3389/fninf.2014.00044
16. Ashburner J, Friston KJ. Unified segmentation. Neuroimage. (2005) 26:839–51. doi: 10.1016/j.neuroimage.2005.02.018
17. Patenaude B, Smith SM, Kennedy DN, Jenkinson M. A bayesian model of shape and appearance for subcortical brain segmentation. Neuroimage. (2011) 56:907–22. doi: 10.1016/j.neuroimage.2011.02.046
18. Barnes J, Ridgway GR, Bartlett I, Henley SMD, Lehmann M, Hobbs N, et al. Head size, age and gender adjustment in MRI studies: a necessary nuisance? Neuroimage. (2010) 53:1244–55. doi: 10.1016/j.neuroimage.2010.06.025
19. Hutchison WD, Lozano AM, Davis KD, Saint-Cyr JA, Lang AE, Dostrovsky JO. Differential neuronal activity in segments of globus pallidus in Parkinson’s disease patients. Neuroreport. (1994) 5:1533–7. doi: 10.1097/00001756-1994100700-00031
20. Broders M, de Bie RMA, Velseboer DC, Speelman JD, Muslimovic D, Schmand B. Evolution of mild cognitive impairment in Parkinson disease. Neurology. (2013) 81:346–52. doi: 10.1212/WNL.0b013e31829c586
21. Gill DJ, Freshman A, Blender JA, Ravina B. The montreal cognitive assessment as a screening tool for cognitive impairment in Parkinson’s disease. Mov Disord. (2008) 23:1043–6. doi: 10.1002/mds.22017
22. Ferrer I. Early involvement of the cerebral cortex in Parkinson’s disease: convergence of multiple metabolic defects. Prog Neurobiol. (2009) 88:89–103. doi: 10.1016/j.pneurobiol.2009.02.004
23. Mattis MP, Chan SL. Calcium channels. Nat Cell Biol. (2003) 5:1041–3. doi: 10.1038/ncb1203-1041
24. Ludtmann MHR, Abramov AY. Mitochondrial calcium imbalance in Parkinson’s disease. Neurosci Lett. (2018) 663:86–90. doi: 10.1016/j.neulet.2017.08.044
25. Surmeier DJ, Schumacker PT, Guzman JD, Ilijic E, Yang B, Zampese E, Calcium and Parkinson’s disease. Biochem Biophys Res Comm. (2017) 483:1013–9. doi: 10.1016/j.bbrc.2016.08.168
26. Grace AA, Bunney BS. Intracellular and extracellular electrophysiology of nigral dopaminergic neurons–2. Action potential generating mechanisms and morphological correlates. Neuroscience. (1983) 103:317–31. doi: 10.1016/0306-4522(83)90136-7
27. Iyer R, Ungless MA, Faisal AA. Calcium-activated SK channels control firing regularity by modulating sodium channel availability in midbrain dopamine neurons. Sci Rep. (2017) 7:5248. doi: 10.1038/s41598-017-05578-5
28. Deister CA, Chan CS, Surmeier DJ, Wilson CJ. Calcium-activated SK channels influence voltage-gated ion channels to determine the precision of firing in globus pallidus neurons. J Neurosci. (2009) 29:8452–61. doi: 10.1523/JNEUROSCI.0576-09.2009
29. Borre L, Andreassen OE, Shi L, Weinstein H, Gether U. The second sodium site in the dopamine transporter controls cation permeation and is regulated by chloride. J Biol Chem. (2014) 289:25764–73. doi: 10.1074/jbc.M114.574269
30. Torres GF, Gainetdinov RR, Caron MG. Plasma membrane monoamine transporters: structure, regulation and function. Nat Rev Neurosci. (2003) 4:13–25. doi: 10.1038/nrn1008
31. Lozovaya N, Eftekhari S, Cloarec R, Gouty-Colomer LA, Dufour A, Riffault B, et al. GABAergic inhibition in dual-transmission cholinergic and GABAergic striatal interneurons is abolished in Parkinson disease. *Nat Commun.* (2018) 9:1422. doi: 10.1038/s41467-018-03802-y

32. Boscia F, Begum G, Pignataro G, Sirabella R, Cuomo O, Casamassa A, et al. Glial Na(+)-dependent ion transporters in pathophysiological conditions. *Glia.* (2016) 64:1677–97. doi: 10.1002/glia.23030

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