Cerebrospinal fluid features in SARS-CoV-2 RT-PCR positive patients

Mathilde Bellon1,2,7*, Cecilia Schweblin1,2,7*, Nathalie Lambeng3*, Pascal Cherpillo1,
Jessica Vazquez3, Patrice H Lalive4,5,6,7, Manuel Schibler4,5,6,7, Christine Deffert3,4

* Bellon Mathilde, Schweblin Cecilia, and Lambeng Nathalie contributed equally to this work.

# Schibler Manuel and Deffert Christine contributed equally to this work.

Affiliations:
1. Laboratory of Virology, Laboratory Medicine Division, Diagnostic Department, Geneva
   University Hospitals, Geneva, Switzerland
2. Division of Infectious Disease, Department of Medical Specialties, Geneva University
   Hospitals, Geneva, Switzerland
3. Laboratory of Biological Fluids, Laboratory Medicine Division, Diagnostic Department,
   Geneva University Hospitals, 1211 Geneva, Switzerland.
4. Laboratory Medicine Division, Department of Medical Specialties, Faculty of Medicine,
   University of Geneva, 1206 Geneva, Switzerland.
5. Department of Clinical Neurosciences, Division of Neurology, Geneva University Hospital,
   Geneva, Switzerland.
6. Department of Pathology and Immunology, Faculty of Medicine, University of Geneva,
   Geneva, Switzerland.
7. Faculty of Medicine of Geneva, University of Geneva, Geneva, Switzerland
#Corresponding author:

Mathilde Bellon

Laboratory of Virology, Division of Infectious Diseases

Geneva University Hospitals

Rue Gabrielle-Perret-Gentil 4

1205 Geneva 14

Switzerland

Tel: ++41 22 372 40 96 ; fax: ++41 22 372 40 97

Email: mathilde.bellon@unige.ch
Abstract

This study analyzed the cerebrospinal fluid features of 31 COVID-19 patients with neurological complications. We observed neither SARS-CoV-2 RNA in the cerebrospinal fluid, nor intrathecal IgG synthesis, but did observe signs of blood-brain barrier disruption. These results might serve as a basis for a better understanding of SARS-CoV-2 related neuropathogenesis.

Keywords: SARS-CoV-2, neurological manifestations, COVID-19, intrathecal synthesis, blood-brain barrier
Introduction
Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2) is responsible for the coronavirus disease 2019 (COVID-19) pandemic. Typical clinical manifestations include cough, dyspnea, fever and fatigue, with a minority of patients evolving towards bilateral pneumonia, sometimes leading to acute respiratory distress syndrome (1). Among less typical features, neurological complications have been described in SARS-CoV-2 infected patients, including meningo-encephalitis, encephalopathy, stroke and Guillain-Barré syndrome (2-5). Currently, the central nervous system tropism and neuropathogenesis of SARS-CoV-2 have not been fully elucidated. ACE-2 identified as the receptor required for virus entry (6) is expressed not only in the lungs, but also in the vascular endothelium and the central nervous system (4). Several articles have described the neurological manifestations of SARS-CoV-2 in large series of patients but cerebrospinal fluid (CSF) parameters have only been analyzed in a limited number of patients (4, 5, 7). The aim of this retrospective observational study was to report CSF features of 31 patients with documented SARS-CoV-2 infection associated with neurological disorders.

Material and methods

Patients
We retrospectively included 31 patients who were hospitalized or consulted in the emergency department between February 26, 2020 - the beginning of the epidemic in Switzerland - and May 7, 2020. Inclusion criteria were a positive rRT-PCR for SARS-CoV-2 in a respiratory specimen and a lumbar puncture on the same day or thereafter, due to acute neurological manifestations. We retrospectively collected epidemiological and biological data, the patient type (outpatient or hospitalized in an intensive or a non-intensive care unit) and the medical
history of COVID-19. Neurological manifestations confirmed by neurologists were reviewed and classified into 5 categories: encephalopathy (confusion, impaired consciousness, delayed awakening), isolated headache, meningeal syndrome, seizure, and peripheral nervous system manifestations (polyneuropathy).

CSF analysis

Cytological examination of CSF was performed for 33 CSF samples from 31 patients. The blood- brain- barrier (BBB) integrity was evaluated using the albumin quotient (CSF albumin/serum albumin) with age- corrected cut- offs and the Reiber formula.

Intrathecal synthesis SARS-CoV-2 IgG

The IgG antibody index (AI) calculation were determined according to Reiber’s method. Eighteen patient’s paired CSF and serum, sampled less than 2 days apart, were analyzed on an Agility ELISA system (Dynex, Denkendorf, Germany) with the Euroimmun Anti-SARS-CoV-2-ELISA (IgG) kit (Euroimmun, Luebeck, Germany). An AI > 2.0 reflects a specific anti-SARS-CoV-2 IgG intrathecal production.

Results

Patients

The study included COVID-19 patients (n=31), confirmed by a positive SARS-CoV-2 rRT-PCR in a respiratory specimen, who displayed new neurological manifestations at the time of or after diagnosis, and who underwent lumbar puncture. The majority of patients (29/31) were hospitalized, 58% in the intensive care unit (ICU) and 35% in the non-intensive care unit (Table 1). The median age of patients was 66 years (range 39 to 85 years) and the oldest
patients were those in the ICU (69 years; range 42 to 83 years). The majority were male (71%), particularly in the ICU (83%).

**Neurological manifestations**

Neurological manifestations were classified into 5 groups: 19 patients (61%) had encephalopathy, 3 (10%) isolated headache, 4 (13%) meningeal syndrome, 1 (3%) seizures, 3 (10%) Guillain-Barré syndrome and 1 (3%) a critical illness polyneuropathy (Table 1).

Encephalopathy, the most observed neurological manifestation, was more frequent in patients hospitalized in the ICU (83%). The median time from the onset of symptoms associated with COVID-19 to the time of lumbar puncture was 23 days; it was longer in patients hospitalized in the ICU (26 days), compared to patients not requiring ICU admission (8 days).

**CSF analysis**

All patients test results were negative for SARS-CoV-2 RNA in the CSF by rRT-PCR (Table 1). Among 33 CSF specimens, elevated white blood cell (WBC) counts were noted for 3 patients. The first had concomitant pneumococcal meningitis, and the second had a Waldenstrom macroglobulinemia and suspicion of Bing-Neel syndrome. Interestingly, the third patient was a 45 year old female presenting with a meningeal syndrome. CSF WBC count was 99 M/L, with 99% lymphocytes. An extensive microbiological workup was negative, including SARS-CoV-2 rRT-PCR in CSF, as well as the AI.

The predominant cells were lymphocytes. Macrophages were observed in 60% of CSF specimens. CSF protein and albumin levels were increased in 39% and 23%, respectively. CSF/plasma albumin ratio (Qalb), was increased in 58% of 19 tested patients, indicating a disturbance of the BBB integrity (S5). CSF protein, albumin levels and Qalb were well correlated, but as all patients showed low levels of serum albumin (supplementary Table 1)
following intensive care hospitalization, only Qalb could be used to estimate the BBB integrity. This serum albumin decrease was probably induced by feeding difficulties during prolonged disease.

All analyzed patients tested positive for anti-SARS-CoV-2 IgG antibodies in sera and 78% of them also showed these specific antibodies in CSF (supplementary Table 1). Sixteen of 19 tested patients (84%) showed IgG oligoclonal bands (OBs) with an identical electrophoretic pattern in serum and CSF (OB IgG type IV pattern). None of the tested patients presented CSF-specific IgG OBs (type II or III) nor an elevated IgG index indicating intrathecal IgG synthesis (data not shown). In addition, no intrathecal IgG-anti-SARS-CoV-2 synthesis was detected.

Discussion

Neurological complications associated with SARS-CoV-2 are frequent and diverse, and involve both the central and the peripheral nervous systems (2). To date, no specific CSF pattern in SARS-CoV-2 infected patients has been described. This retrospective study, despite the limited number of patients included, is the largest to date to describe CSF features of SARS-CoV-2 infected patients with neurological complications in detail, to the best of our knowledge.

Our findings suggest that SARS-CoV-2 infection is an indirect cause of the neurological complications, independently of the neurological diagnosis. Indeed, CSF analysis did not reveal any evidence of direct viral infection of the central nervous system: 1) SARS-CoV-2 RNA was not detected in CSF using rRT-PCR; 2) no sign of CSF inflammation except in rare case of co-infection; 3) absence of intrathecal synthesis using IgG Index and electrophoretic IgG oligoclonal bands; and finally, 4) absence of specific intrathecal anti-SARS-CoV-2 IgG synthesis. Currently, there are few case reports of patients with encephalitis associated with
SARS-CoV-2 RNA positive in CSF (4, 8, 9). One could argue that SARS-CoV-2 rRT-PCR assays are not validated in CSF, and that their sensitivity may be affected by delays in storage time. However, CSF is generally a type of sample for which RNA viruses rRT-PCR assays are easy to validate. Furthermore, since the virology laboratory is located inside the hospital in which the patients included were hospitalized, there has not been any particular delay in sample acquisition, and the storage modalities were not altered, in respect with the pre-COVID-19 era. There is therefore no theoretical reason why the rRT-PCR assay should yield false negative results in this material. The concomitant absence of total IgG and specific SARS-CoV-2 intrathecal synthesis strengthen the hypothesis that the neurological manifestations observed during COVID-19 are probably induced by indirect mechanisms in the majority of cases (4, 10), as already demonstrated for other viruses (4).

Interestingly, in our study, most of the CSF specimens analyzed showed a type IV oligoclonal pattern. This pattern, already reported in some patients with neurologic disorders in severe SARS-CoV-2 infection (5, 7), is consistent with passive diffusion of oligoclonal IgG from a systemic inflammatory state such as a systemic viral infection. The vast majority (92%) of patients had both type IV oligoclonal pattern and anti-SARS-CoV-2 antibodies in CSF, leading to the hypothesis that IgG oligoclonal bands are at least in part anti-SARS-CoV-2 IgG.

Most of our patients (58%) with neurological manifestations during their COVID-19 illness showed BBB leakage. BBB disruption could be induced by a direct effect of SARS-CoV-2, but is more likely caused by indirect effects via generalized endotheliitis (11). The abnormal presence of macrophages in 60% of the CSF specimens suggests a microglial activation known to induce the expression of pro-inflammatory cytokines, chemokines and matrix metalloproteases leading to the BBB disruption (12, 13). This mechanism resembles that described in septic encephalopathy (14).
Limitations of this study include selection biases during inclusion of patients with neurological manifestations. Indeed, the short study period in the initial phase of the pandemic that allowed only patients with rapid onset of neurological disorders to be targeted. Furthermore, patients with neurological manifestations who did not undergo lumbar puncture were not included. Finally, CSF analyses must be performed rapidly and requires CSF and serum samples collected less than two days apart, which prevented the retrospective inclusion of a large cohort.

In summary, our results mainly showed the absence of SARS-CoV-2 RNA detection in the CSF and specific intrathecal IgG synthesis in the tested samples, and highlighted a reproducible CSF pattern in SARS-CoV-2 infection: normal WBC count, presence of macrophages, signs of BBB permeability and type IV oligoclonal pattern. Neurological manifestations associated with COVID-19, especially encephalopathy, are more likely due to a systemic process, such as exacerbated inflammation resulting in endotheliitis, cytokine storm and microglia activation, rather than direct invasion of the CNS by SARS-CoV-2. If confirmed by other groups, these findings could orientate therapeutic strategies towards anti-inflammatory, rather than antiviral drugs, with regards to COVID-19 related neurological manifestations.
NOTES

Acknowledgments

The authors would like to thank the patients from the University Hospitals of Geneva, Neftali Rodrigo for his review of CSF cytospin slides and Erik Boehm for his valuable help in manuscript editing.

Potential conflict of interest statement

Pr P. H. Lalive received honoraria for speaking from Biogen-Idec, CSL Bering, Merck Serono, Novartis, Sanofi-Aventis, Teva, Roche; consulting fees from Biogen-Idec, Geneuro, Genzyme, Merck Serono, Novartis, Sanofi-Aventis, Teva; research grants from Biogen-Idec, Merck Serono, Novartis.
References

1. Vetter P, Vu DL, L'Huillier AG, Schibler M, Kaiser L, Jacquieroz F. Clinical features of covid-19. BMJ. 2020;369:m1470.
2. Mao L, Jin H, Wang M, Hu Y, Chen S, He Q, et al. Neurologic Manifestations of Hospitalized Patients With Coronavirus Disease 2019 in Wuhan, China. JAMA Neurol. 2020.
3. Lascano AM, Epiney JB, Coen M, Serratrice J, Bernard-Valnet R, Lalive PH, et al. SARS-CoV-2 and Guillain-Barre syndrome: AIDP variant with favorable outcome. Eur J Neurol. 2020.
4. Koralnik IJ, Tyler KL. COVID-19: a global threat to the nervous system. Ann Neurol. 2020.
5. Helms J, Kremer S, Merdji H, Clere-Jehl R, Schenck M, Kummerlen C, et al. Neurologic Features in Severe SARS-CoV-2 Infection. N Engl J Med. 2020.
6. Yan R, Zhang Y, Li Y, Xia L, Guo Y, Zhou Q. Structural basis for the recognition of SARS-CoV-2 by full-length human ACE2. Science. 2020;367(6485):1444-8.
7. Andriuta D, Roger PA, Thibault W, Toublanc B, Sauzay C, Castelain S, et al. COVID-19 encephalopathy: detection of antibodies against SARS-CoV-2 in CSF. J Neurol. 2020.
8. Moriguchi T, Harii N, Goto J, Harada D, Sugawara H, Takamino J, et al. A first case of meningitis/encephalitis associated with SARS-Coronavirus-2. Int J Infect Dis. 2020;94:55-8.
9. Domingues RB, Mendes-Correa MC, de Moura Leite FBV, Sabino EC, Salarini DZ, Claro I, et al. First case of SARS-COV-2 sequencing in cerebrospinal fluid of a patient with suspected demyelinating disease. J Neurol. 2020.
10. Ahmad J, Rathore FA. Neurological manifestations and complications of COVID-19: A literature review. J Clin Neurosci. 2020.
11. Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, et al. Endothelial cell infection and endotheliitis in COVID-19. The Lancet. 2020;395(10234):1417-8.
12. Bindoli S, Felicetti M, Sfriso P, Doria A. The amount of cytokine-release defines different shades of Sars-Cov2 infection. Exp Biol Med (Maywood). 2020;1535370220928964.
13. Al-Obaidi MMJ, Bahadoran A, Wang SM, Manikam R, Raju CS, Sekaran SD. Disruption of the blood brain barrier is vital property of neurotropic viral infection of the central nervous system. Acta Virol. 2018;62(1):16-27.
14. Tauber SC, Eiffert H, Bruck W, Nau R. Septic encephalopathy and septic encephalitis. Expert Rev Anti Infect Ther. 2017;15(2):121-32.
Table 1. Clinical and biological characteristics of the Patients with COVID-19 and neurological manifestations.

| Variable                                             | All Patients\(^\d\) (N = 31) | ICU Patients (N = 18) | Non-ICU Patients (N = 11) |
|------------------------------------------------------|-------------------------------|-----------------------|---------------------------|
| **Epidemiological data**                             |                               |                       |                           |
| Age (median +/- IQR)                                 | 66.3 +/- 18.8                 | 69.5 +/- 12.3         | 61.2 +/- 25.4             |
| Male (%)                                             | 71                            | 83                    | 55                        |
| **Neurological manifestations**                      |                               |                       |                           |
| Encephalopathy n (%)                                 | 19 (61%)                      | 15 (83%)              | 4 (36%)                   |
| PNS involvement n (%)                                | 4 (13%)                       | 1 (5%)                | 3 (27%)                   |
| Headache n (%)                                       | 3 (10%)                       | 0 (0%)                | 1 (9%)                    |
| Meningeal syndrome n (%)                             | 4 (13%)                       | 2 (11%)               | 2 (18%)                   |
| Seizures n (%)                                       | 1 (3%)                        | 0 (0%)                | 1 (9%)                    |
| Neurological symptoms may be related to COVID-19 n (%)| 28 (90%)                      | 16 (89%)              | 10 (91%)                  |
| Delay between onset general symptoms and LP median (min-max) | 23 (0-50) | 26 (4-50) | 8 (0-40) |
| **CSF analysis**                                     |                               |                       |                           |
| WBC (M/L) (median +/- IQR)                           | 2.0 +/- 2.0                   | 2.0 +/- 1.8           | 2.0 +/- 4.0               |
| Lymphocytes (%) (median +/- IQR)                     | 92.0 +/- 9.2                  | 92.0 +/- 9.5          | 95.0 +/- 9.0              |
| Monocytes (%) (median +/- IQR)                       | 4.5 +/- 6.0                   | 4.0 +/- 5.5           | 5.0 +/- 5.0               |
| Neutrophils (%) (median +/- IQR)                     | 1.0 +/- 3.5                   | 1.0 +/- 3.0           | 1.0 +/- 5.0               |
| Detection of Plasma cells n (%)                      | 5 (17%)                       | 3 (18%)               | 2 (18%)                   |
| Detection of Macrophages n (%)                       | 18 (60%)                      | 9 (53%)               | 8 (72%)                   |
| Positive SARS-CoV-2 RT-PCR n                         | 0                             | 0                     | 0                         |
| Protein level (g/L) (median +/- IQR) (Normal value:0.15 to 0.45g/L) | 0.42 +/- 0.36 | 0.45 +/- 0.36 | 0.39 +/- 1.07 |
| Increased Albumin Quotient (>Qlim) n/n total (%)     | 11/19 (58%)                   | 9/16 (56%)            | 2/3 (67%)                 |
| Detection of CSF anti-SARS-CoV-2 IgG n/n total (%)   | 14/18 (77%)                   | 11/12 (92%)           | 3/6 (50%)                 |
| Presence of oligoclonal bands type IV (same pattern in serum and CSF) n/n total (%) | 16/19 (84%) | 13/16 (81%) | 3/3 (100%) |
| Positive intrathecal anti-SARS-CoV-2 IgG antibody index (> 2.0) n | 0 | 0 | 0 |

\(^\d\)Among the patients, two were not hospitalized. Both had headache and were not tested for CSF IgG antibodies nor oligoclonal bands.

CSF: cerebrospinal fluid; PNS: Peripheral nervous system; RT-PCR: reverse transcriptase-Polymerase Chain Reaction; SARS-CoV-2: Severe Acute Respiratory Syndrome Coronavirus-2; LP: Lumbar Puncture; IgG: Immunoglobulin G; Qlim: albumin quotient cut-off corrected by age; IQR: Interquartile range; M/L: Mega per liter (equivalent cells per microliter); Positive intrathecal anti-SARS-CoV-2 IgG antibody index: an AI value below 2 means that specific SARS-CoV-2 antibodies were detected in the CSF, but they were not locally produced.