The L-α-Lysophosphatidylinositol/G Protein–Coupled Receptor 55 System Induces the Development of Nonalcoholic Steatosis and Steatohepatitis

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BACKGROUND AND AIMS: G protein–coupled receptor (GPR) 55 is a putative cannabinoid receptor, and l-α-lysophosphatidylinositol (LPI) is its only known endogenous ligand. Although GPR55 has been linked to energy homeostasis in different organs, its specific role in lipid metabolism in the liver and its contribution to the pathophysiology of nonalcoholic fatty liver disease (NAFLD) remains unknown.

APPROACH AND RESULTS: We measured (1) GPR55 expression in the liver of patients with NAFLD compared with individuals without obesity and without liver disease, as well as animal models with steatosis and nonalcoholic steatohepatitis (NASH), and (2) the effects of LPI and genetic disruption of GPR55 in mice, human hepatocytes, and human hepatic stellate cells. Notably, we found that circulating LPI and liver expression of GPR55 were up-regulated in patients with NASH. LPI induced adenosine monophosphate–activated protein kinase activation of acetyl–coenzyme A carboxylase (ACC) and increased lipid content in human hepatocytes and in the liver of treated mice by inducing de novo lipogenesis and decreasing β-oxidation. The inhibition of GPR55 and ACCα blocked the effects of LPI, and the in vivo knockdown of GPR55 was sufficient to improve liver damage in mice fed a high-fat diet and in mice fed a methionine-choline–deficient diet. Finally, LPI promoted the initiation of hepatic stellate cell activation by stimulating GPR55 and activation of ACC.

CONCLUSIONS: The LPI/GPR55 system plays a role in the development of NAFLD and NASH by activating ACC. (HEPATOLOGY 2021;73:606–624).

Nonalcoholic fatty liver (NAFL) disease (NAFLD) is a major global health threat because of its growing incidence and prevalence. It is becoming the leading cause of liver disease and despite the many efforts to find therapeutic targets for this disease, there is no established therapy yet.10 In this regard, G protein–coupled receptors

Abbreviations: ACC, acetyl-CoA carboxylase; ACTA2, actin α2; AMPK, adenosine monophosphate–activated protein kinase; AST, aspartate aminotransferase; CB1, cannabinoid type 1 receptor; CoA, coenzyme A; COL1α1, collagen α1; COL1α2, collagen α2; CPT1A, carnitine palmitoyl transferase 1A; ECS, endocannabinoid system; GPR, G protein–coupled receptor; HFD, high-fat diet; HSC, hepatic stellate cell; LPI, l-α-lysophosphatidylinositol; MBOAT7, membrane-bound O-acyltransferase domain–containing 7; MCD, methionine-choline–deficient; NAFLD, nonalcoholic fatty liver; NAS, NAFLD activity score; NASH, nonalcoholic steatohepatitis; NEFA, nonesterified fatty acid; pACC, phosphorylated ACC; pAMPKα, phosphorylated AMPKα; shGPR55, shRNA against GPR55; shRNA, short hairpin RNA; TGFβ1, transforming growth factor β1.

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(GPRs) are the most intensively studied drug targets, given their heavy involvement in human pathophysiology and their pharmacological flexibility. In fact, approximately 34% of all approved drugs act in some way through GPRs, reflecting the relevance of activating or inhibiting signal transduction cascades by modulating the activity of GPRs in drug discovery.

GPR55 is, in addition to the classical cannabinoid type 1 receptor (CB1) and classical cannabinoid type 2 receptor, considered a putative cannabinoid receptor. The endocannabinoid system (ECS) is involved in a wide range of biological processes; however, one of the aspects that has attracted more attention over the last years is undoubtedly its ability to modulate energy metabolism. GPR55 is widely distributed in rodent and human tissues and mediates the actions of its endogenous ligand 1-α-lysophosphatidylinositol (LPI) and to a much lesser extent mediates the actions of anandamide and 2-arachidonoyl-glycerol. LPI belongs to the...
class of lysophospholipids and is generated by phosphatidylinositol hydrolysis through the action of the calcium-dependent phospholipase A2 (14) and calcium-independent phospholipase A1. (15) GPR55 is implicated in numerous physiological actions, including reproduction, angiogenesis, apoptosis, inflammation, and energy homeostasis, among others. (8,16) In the gastrointestinal tract, the activation of GPR55 resulted in the inhibition of neurogenic contractions in the mouse intestine (17) and is also involved in colon motility. (18) Regarding the role of the LPI/GPR55 system in metabolism, the LPI/GPR55 system has been positively associated with human obesity because LPI induces the expression of lipogenic genes in explants of visceral adipose tissue from patients with obesity. (19) In the pancreas, the exposure of rodent islets to GPR55 agonists enhanced insulin secretion and improved glucose tolerance. (20,21) Similarly, the stimulation of human islets with GPR55 ligands also potentiates insulin levels. (22)

GPR55 mRNA has also been described in the liver of humans (19) and mice (20), however, the hepatic function of the LPI/GPR55 system is still unknown. Herein, we sought to investigate the potential role of this system in the development of NAFLD. We found that circulating LPI and liver GPR55 expression levels were increased in patients and animal models with NAFLD. LPI augmented lipid content by inducing de novo lipogenesis and decreasing β-oxidation, whereas down-regulation of GPR55 and acetyl–coenzyme A (CoA) carboxylase (ACC) α blocked the effects of LPI. The knockdown of GPR55 was sufficient to improve liver damage in mice fed a high-fat diet (HFD) and mice fed a methionine–choline–deficient (MCD) diet. Finally, LPI promoted the initiation of hepatic stellate cell (HSC) activation by stimulating GPR55 and ACC. These results highlight the importance of LPI and GPR55 in the development of NAFL and nonalcoholic steatohepatitis (NASH).

Participants and Methods

HUMAN LIVER SAMPLES

These samples were obtained from patients at the University Hospital of Salamanca, Santa Cristina University Hospital in Madrid (Spain) and Hospital Clinico Universidad Catlica (Chile). Baseline characteristics of each cohort and exclusion criteria are described in Supporting Participants and Methods. Informed consent in writing was obtained from each patient and the study protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki as reflected in a priori approval by the appropriate institutional review committee.

ANIMALS AND DIETS

Male C57BL/6J were used and fed with different diets as described in Supporting Participants and Methods. Animal protocols were approved by the Committee at the University of Santiago de Compostela and received humane care according to the criteria outlined in the “Guide for the Care and Use of Laboratory Animals.” For further details please see Supporting Participants and Methods.

Results

HEPATIC GPR55 IS OVEREXPRESSED IN HUMAN AND MURINE NAFL AND NASH

Analysis of liver biopsies from patients with NAFLD and obesity (with a body mass index [BMI] > 35 kg/m2) revealed elevated mRNA expression of GPR55 compared with individuals without obesity and NAFLD (Fig. 1A). (Characterization of cohort 1 is shown in Supporting Table S1.) GPR55 was similarly increased in patients with obesity and NAFL or NASH compared with individuals without obesity and without liver damage (Fig. 1A). Further, hepatic GPR55 mRNA was significantly elevated in another cohort of individuals with a BMI < 30 kg/m2 and NAFLD compared with individuals without obesity and without liver damage (Fig. 1A). Further, hepatic GPR55 mRNA was significantly elevated in another cohort of individuals with a BMI < 30 kg/m2 and NAFLD compared with individuals without liver disease (Fig. 1B) and showed a positive correlation with NAFLD activity score (NAS) (Supporting Fig. S1). (Characterization of cohort 2 is shown in Supporting Table S2.) Finally, serum analysis of LPI was conducted in a third cohort of patients with obesity and NAFLD or NASH. (Characterization of cohort 3 shown in Supporting Table S3.) Our results revealed higher levels of LPI 16:0, 18:1, and 18:1 isomers in patients with NASH compared with those with only fatty liver (Fig. 1C). Furthermore, we found that these species of LPI were increased as inflammation and...
**FIG. 1.** Hepatic GPR55 mRNA is up-regulated in patients with NAFLD. GPR55 mRNA levels in the liver of (A) participants without obesity and without NAFLD (n = 11) or participants with obesity and NAFLD (n = 35) (cohort 1), either grouped or considering the NAFL (n = 8) and NASH (n = 27) stages in two different groups, and (B) patients without obesity and without (n = 18) or with (n = 21) NAFLD (cohort 2), either grouped or considering the NAFL (n = 13) and NASH (n = 8) stages in two different groups. (C) Serum levels of the different species of LPI in patients with NASH (n = 78) compared with patients with fatty liver without NASH (n = 31) (cohort 3). HPRT was used to normalize mRNA levels. Data are presented as mean ± SEM. Statistical differences are denoted by *P < 0.05 and **P < 0.01. Abbreviations: GPR 55, G protein–coupled receptor 55; HPRT, hypoxanthine-guanine phosphoribosyltransferase; LPI, l-α-lysophosphatidylinositol; NAFL, nonalcoholic fatty liver; NAFLD, NAFL disease; NASH, nonalcoholic steatohepatitis.
FIG. 2. Hepatic GPR55 is increased in different models of NAFLD in vivo and in vitro. GPR55 mRNA levels, GPR55 protein levels, H&E staining, and Sirius Red staining in the liver of (A) mice fed an HFD with 45%-kcal fat content or an STD for 6, 12, and 52 weeks (n = 5-14); (B) mice fed a CD-HFD or STD for 6, 12, and 52 weeks (n = 5-10); (C) mice fed a vHFD with 60%-kcal fat content or an STD for 10 weeks (n = 9-10); and (D) mice fed an MCD diet or an STD for 4 weeks (n = 7-10 per group). (E) GPR55 mRNA levels in hepatocytes isolated from the liver of mice fed an MCD diet or STD for 4 weeks (n = 5 per group). (F) GPR55 mRNA levels in THLE2 cells cultured in oleic acid or BSA (n = 6). GAPDH and HPRT were used to normalize protein and mRNA levels. Dividing lines indicate splicing in the same gel. Data are presented as mean ± SEM. Statistical differences are denoted by *P < 0.05, **P < 0.01, and ***P < 0.001. Abbreviations: BSA, bovine serum albumin; CD-HFD, choline-deficient HFD; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; GPR55, G protein–coupled receptor 55; H&E, hematoxylin and eosin; HFD, high-fat diet; HPRT, hypoxanthine-guanine phosphoribosyltransferase; MCD, methionine-choline–deficient; NAFLD, nonalcoholic fatty liver disease; STD, standard diet; vHFD, very HFD.
ballooning scores augmented (Supporting Fig. S2A). When patients from cohort 3 were separated as with diabetes versus without diabetes, we found that serum levels of these species of LPI did not show any difference (Supporting Fig. S2B), suggesting that changes in LPI species are independent of dysfunctions in glucose metabolism.

We next studied the expression of GPR55 in livers from mice fed an HFD with 45%-kcal fat content, a choline-deficient HFD (CD-HFD), a very HFD (vHFD) with 60%-kcal fat content, and an MCD diet at different times. In all of these animal models, we detected the expected histological alterations and increases of both mRNA and protein levels of GPR55 (Fig. 2A-D). GPR55 mRNA and protein levels were also augmented in isolated hepatocytes from mice fed an MCD compared with mice fed a chow diet (Fig. 2E). Finally, we used the human hepatic cell line THLE2 and treated these hepatocytes with oleic acid. This treatment significantly increased GPR55 mRNA expression after 1 hour in both THLE2 (Fig. 3A) and HepG2 (Supporting Fig. S3A) cells. After 24 hours of its administration, LPI incrementally altered the lipid content in both cell lines (Fig. 3C and Supporting Fig. S3B). At the molecular level in the two cell lines, we found diminished levels of phosphorylated ACC (pACC) and phosphorylated adenosine monophosphate–activated protein kinase (AMPK) α (pAMPKα), ACC’s main regulator, and diminished protein levels of carnitine palmitoyl transferase 1a (CPT1A) (Fig. 3B and Supporting Fig. S3B), suggesting an activation of ACC and thus a stimulation of de novo fatty acid synthesis and inhibition of fatty acid oxidation, respectively. The used dose of LPI did not alter protein levels of apoptotic markers (Supporting Fig. S3C). To further characterize whether the effects of LPI were mediated by GPR55, we silenced the receptor and found that LPI was not able to increase the lipid content (Fig. 3C and Supporting Fig. S3D). Consistent with this, although LPI decreased protein levels of pAMPKα and pACC, it failed to do so after GPR55 silencing (Fig. 3C and Supporting Fig. S3D). Given that GPR55 was mediating LPI-induced lipid deposition, we next studied whether this receptor could be relevant for the deposition of lipids induced by a different stimulus. Oleic acid–induced lipid content was strongly attenuated after the inhibition of GPR55 (Fig. 3D). Finally, because LPI activates ACC, we next investigated whether the inhibition of ACC would affect LPI’s effects. LPI failed to increase lipid content in THLE2 and HepG2 cells after silencing of ACCα (Fig. 3E and Supporting Fig. S3E).

Because these results indicated that LPI and GPR55 were modulating fatty acid synthesis and oxidation, we next directly measured these parameters using [3H]acetate and [14C]palmitate, respectively, and detected that LPI increased de novo triglyceride, diglyceride, free fatty acid, and phospholipids lipogenesis (Fig. 4A and Supporting Fig. S4A), whereas LPI decreased β-oxidation (Fig. 4B and Supporting Fig. S4B). These LPI-induced effects were blocked when GPR55 was silenced (Fig. 4). An important event that occurs during development of NAFLD/NASH is the reduction in mitochondrial respiratory-chain activity.(23) To investigate whether this decrease in β-oxidation alters mitochondrial respiration, we monitored the oxygen consumption rate (OCR) in cells treated with LPI. We found a significant reduction in the OCR 1 hour after the administration of LPI. This effect was blunted when etomoxir, a CPT1A inhibitor, was provided 30 minutes before the LPI (Supporting Fig. S4C).

LPI INDUCES LIVER STEATOSIS IN MICE THROUGH GPR55 AND ACC

C57BL/6J mice were treated with LPI (0.5 mg/kg) intraperitoneally for 7 days. As reported before,(19) this dose of LPI did not affect body weight (Supporting Fig. S5A) or food intake (Supporting Fig. S5B). Herein, we detected changes in neither liver mass nor aspartate aminotransferase (AST) or cholesterol, but serum triglycerides and nonesterified fatty acids (NEFAs) were increased after LPI treatment.
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The hepatic lipid content, GPR55 mRNA expression, and fatty acid synthase (FAS) protein levels in LPI-treated mice were significantly higher than in vehicle-treated mice (Supporting Fig. S5D,E), whereas the levels of pAMPKα, the ratio of pACC to ACC, and the levels of CPT1A were reduced (Supporting Fig. S5F).

To test whether GPR55 was mediating the in vivo effects of LPI, we performed the same pharmacological treatment but added a group of mice, which previously had a lentivirus encoding short hairpin RNA (shRNA) against GPR55 (shGPR55) injected into the tail vein. Although LPI increased GPR55 expression, shGPR55-treated mice showed a marked decrease in GPR55 expression in the liver (Fig. 5A). The LPI-induced lipid and triglyceride content in the liver was blunted in shGPR55-treated mice (Fig. 5B). Moreover, the reduction of pAMPKα and pACC caused by LPI did not occur in shGPR55-treated mice (Fig. 5C). Consistent with the activation of ACC by LPI, the levels of malonyl-CoA in the liver were increased by LPI and reduced after the knockdown of GPR55 (Fig. 5D).

Next, we assessed whether ACC was also playing a relevant role as a mediator of LPI-induced actions.
in the liver. For this, we treated one group of mice with LPI and treated another group with LPI and the ACC inhibitor GS-0976, which is in clinical trials for the treatment of NAFLD.\(^{(24)}\) We injected GS-0976 intraperitoneally at a dose of 10 mg/kg for 1 week as reported\(^{(25)}\) and found that it blocked the hepatic
FIG. 5. LPI induces hepatic steatosis in a GPR55-dependent and ACC-dependent manner in vivo. C57BL/6J mice fed a standard diet randomly received a TVI with Lentiv-sh scrambled or Lentiv-shGPR55 and 3 weeks later were treated with either vehicle or LPI (0.5 mg/kg) for 7 days (n = 6). (A) GPR55 mRNA levels in the liver. (B) H&E staining (upper panel) and Oil Red O staining (lower panel) of liver sections. Lipids were quantified and normalized to the total number of nuclei per field. Hepatic triglyceride content was also directly measured. (C) Hepatic protein levels of pAMPKα, pACC, and ACC. (D) Hepatic malonyl-CoA levels. C57BL/6J mice fed a standard diet were treated daily with the ACC inhibitor GS-0976 at a dose of 10 mg/kg (or vehicle) for 7 days and were co-administered either vehicle or LPI (0.5 mg/kg) for the same period (n = 8). (E) H&E staining (upper panel) and Oil Red O staining (lower panel) of liver sections. Lipids were quantified in the Oil Red O staining sections. Hepatic triglyceride content was also directly measured. (F) Hepatic malonyl-CoA levels. HPRT and GAPDH were used to normalize, respectively, mRNA and protein levels. Dividing lines indicate splicing in the same gel. Data are presented as mean ± SEM. Statistical differences are denoted by *P < 0.05, **P < 0.01, and ***P < 0.001. Abbreviations: ACC, acetyl-CoA carboxylase; CoA, coenzyme A; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; GPR55, G protein–coupled receptor 55; H&E, hematoxylin and eosin; HPRT, hypoxanthine-guanine phosphoribosyltransferase; i.p., intraperitoneally; Lentiv-shGPR55, lentiviral vector encoding short hairpin RNA against GPR55; Lentiv-sh scrambled, lentiviral vector encoding scrambled short hairpin RNA; LPI, l-α-lysophosphatidylinositol; pACC, phosphorylated ACC; pAMPKα, phosphorylated adenosine monophosphate–activated protein kinase α; TVI, tail-vein injection.

KNOCKDOWN OF HEPATIC GPR55 AMELIORATES MCD DIET–INDUCED AND CCl4–INDUCED STEATOHEPATITIS

Because GPR55 mRNA expression in the liver was up-regulated in humans and rodents with NASH, we evaluated the potential of the inhibition of GPR55 to ameliorate NASH. To test this, we used mice fed an MCD diet, which induces macrovesicular steatosis and is widely used in NASH research.(26) Mice underwent tail-vein injection of a lentivirus encoding shGPR55 and were fed an MCD diet 4 weeks later for 4 weeks until being sacrificed. The efficiency of the infection was assessed by the significant reduction in GPR55 mRNA and protein levels when compared with scrambled shRNA–treated mice (Supporting Fig. S6A). Although liver mass and serum levels of triglycerides, NEFAs, and cholesterol were not affected by the inhibition of hepatic GPR55 (Supporting Fig. S6B), circulating AST was significantly lower compared with that of the control group (Fig. 7A). In line with this, the inhibition of GPR55 reduced hepatic lipid and triglyceride content, hydroxyproline, F4/80, and collagen (Fig. 7B,C) and inactivated ACC, as demonstrated by increased pAMPKα and pACC levels and decreased malonyl-CoA and collagen mRNA (Fig. 7C). According to the improvement in liver condition, levels of different markers of ER stress, inflammation, fibrosis, and apoptosis were significantly reduced after the suppression of hepatic GPR55 (Supporting Fig. S6C-F).
Next, we used a second model of fibrosis, similar to CCl₄-induced liver fibrosis, in which we also injected a lentivirus encoding shGPR55. We found that after 6 weeks, CCl₄ induced the expression of GPR55 (Fig. 7D). This effect was concomitant with higher levels of serum AST, a higher level of collagen in the liver,
FIG. 6. The down-regulation of hepatic GPR55 ameliorates NAFL induced by a vHFD (60%-kcal fat content) in mice. C57BL/6J mice fed a vHFD diet for 6 weeks randomly received a TVI with Lentiv-sh scrambled or Lentiv-shGPR55, maintaining the vHFD for 4 weeks more. (A) GPR55 mRNA levels in the liver. (B) Body-weight change and food intake. (C) Serum levels of AST, triglycerides, NEFAs, and cholesterol. (D) H&E staining (upper panel), Oil Red O staining (middle panel), and F4/80-immunohistochemistry staining (lower panel) of liver sections. Lipids were quantified in the Oil Red O staining sections, and the F4/80-positive area was quantified in the F4/80-immunohistochemistry staining sections. Hepatic triglyceride content was also directly measured. (E) Hepatic levels of proteins involved in lipid metabolism and hepatic levels of malonyl-CoA. (F) Protein levels of ER stress markers, gene expression of inflammatory markers, and cleaved isoforms of apoptotic markers caspase 3 and caspase 7 in the liver. HPRT and GAPDH were used to normalize mRNA and protein levels, respectively. Dividing lines indicate splicing in the same gel. Data are presented as mean ± SEM (n = 7). Statistical differences are denoted by *P < 0.05, **P < 0.01, and ***P < 0.001. Abbreviations: AST, aspartate aminotransferase; CoA, coenzyme A; CPT1a, carnitine palmitoyl transferase 1a; ER, endoplasmic reticulum; FAS, fatty acid synthase; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; GPR55, G protein–coupled receptor 55; H&E, hematoxylin and eosin; HPRT, hypoxanthine-guanine phosphoribosyltransferase; i.p., intraperitoneally; Lentiv-shGPR55, lentiviral vector encoding short hairpin RNA against GPR55; Lentiv-sh scrambled, lentiviral vector encoding scrambled short hairpin RNA; NAFL, nonalcoholic fatty liver; NEFA, nonesterified fatty acid; pEIF2α, phosphorylated eukaryotic initiation factor 2α; pIRE, phosphorylated iron-responsive element; pPERK, phosphorylated protein kinase R-like ER kinase; shCTL, short hairpin RNA control; shGPR55, short hairpin RNA against GPR55; TVI, tail-vein injection; vHFD, very high-fat diet; XBP1S, X-box binding protein 1S.
FIG. 7. Hepatic silencing of GPR55 ameliorates NASH induced by an MCD diet and fibrosis induced by CCl₄ in mice. C57BL/6J mice randomly received a TVI with Lentiv-shGPR55 or Lentiv-sh scrambled and 4 weeks later were fed an MCD diet for 4 weeks (n = 7). (A) Serum levels of AST. (B) H&E staining (first panel), Oil Red O staining (second panel), Sirius Red staining (third panel), and F4/80-immunohistochemistry staining (fourth panel) of liver sections. Staining areas were quantified. (C) Triglyceride content; levels of malonyl-CoA; and mRNA levels of fibrosis markers in the liver. C57BL/6J mice were treated with CCl₄ (0.6 mL/kg administered i.p.) or vehicle once a week for 6 weeks, receiving a TVI with Lentiv-shGPR55 or Lentiv-sh scrambled during the first week. (D) GPR55 mRNA levels in the liver and serum levels of AST (n = 8). (E) H&E staining (upper panel) and Sirius Red staining (lower panel) of liver sections. Collagen deposition was quantified in the Sirius Red staining sections (n = 8). (F) Protein levels of pAMPKα, pACC, and ACC and mRNA levels of fibrosis markers in the liver (n = 4-8). HPRT and GAPDH were used to normalize mRNA and protein levels, respectively. Dividing lines indicate splicing in the same gel. Data are presented as mean ± SEM. Statistical differences are denoted by *P < 0.05, **P < 0.01, and ***P < 0.001. Abbreviations: ACC, acetyl-CoA carboxylase; AST, aspartate aminotransferase; CoA, coenzyme A; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; GPR55, G protein–coupled receptor 55; H&E, hematoxylin and eosin; HPRT, hypoxanthine-guanine phosphoribosyltransferase; i.p., intraperitoneally; Lentiv-sh scrambled, lentiviral vector encoding scrambled short hairpin RNA against GPR55; Lentiv-sh GPR55, lentiviral vector encoding short hairpin RNA against GPR55; MCD, methionine-choline–deficient; NASH, nonalcoholic steatohepatitis; pACC, phosphorylated ACC; pAMPKα, phosphorylated adenosine monophosphate–activated protein kinase α; shCTL, short hairpin RNA control; shGPR55, short hairpin RNA against GPR55; sh scrambled, scrambled short hairpin RNA; TVI, tail-vein injection.

Discussion

The findings presented herein show that GPR55 is overexpressed in the liver of patients and animal models with NAFLD and that LPI-induced GPR55 activation promotes lipid storage, thereby participating in the progression of liver steatosis and steatohepatitis. We also demonstrated that the knockdown of hepatic GPR55 ameliorates HFD-induced NAFL and both MCD diet–induced and CCl₄-induced NASH. Pathologic overactivation of the ECS is associated with the development of dyslipidemia, obesity, and type 2 diabetes. These actions of the ECS have been mainly attributed to the activation of central and peripheral CB1 receptors. (30) HFD increased the hepatic levels of the endocannabinoid anandamide CB1 receptors and basal rates of fatty acid synthesis, and the latter is reduced by CB1 blockade. (29,31) More specifically, the activation of hepatic CB1 stimulates de novo lipogenesis through induction of the lipogenic transcription factor sterol regulatory element binding protein 1c and its target enzymes ACC and FAS. (25) Our present results indicate that hepatic activation of GPR55 by its endogenous ligand LPI is also able to promote lipid deposition in the liver by its actions on both hepatocytes and HSCs.

Some types of circulating lysophospholipids have emerged as signaling molecules with important effects on inflammation and fatty liver disease. (32-34) In line with this, our present results indicate that LPI levels are increased in patients with NASH. The mechanisms by which LPI triggers lipid accumulation seem to involve alterations in hepatic fatty acid metabolism because LPI is also able to increase de novo lipogenesis by decreasing the phosphorylation of ACC, which leads to more active ACC in both hepatocytes and HSCs. Furthermore, we also found that LPI inhibited β-oxidation, which is consistent with the decreased levels of CPT1A that catalyze the primary regulated step in overall mitochondrial fatty acid oxidation in the liver mitochondrial outer membrane. Our current data are in agreement with those of a previous study showing that postprandial lysophospholipids suppress hepatic fatty acid oxidation and promote hepatic inflammatory and fibrotic transcriptional changes. (35) In this sense, it has been recently reported that membrane-bound O-acyltransferase domain–containing 7 (MBOAT7), which encodes an acyltransferase enzyme that specifically esterifies arachidonyl-CoA to LPI lipids, is involved in NAFLD. In particular, Mboat7 knockdown caused liver-disease progression and was associated with a significant accumulation of 16:0 and 18:1 LPI species in the liver; importantly, these were the two species we found to be positively correlated with NAS. Therefore, these two studies involved LPIs as important mediators in the development of NAFLD.

The LPI receptor GPR55 is increased in the liver not only in mice fed an HFD but also in mice exposed to different diets, such as the CD-HFD and the MCD diet, or in mice exposed to treatments like CCl₄, representing models of steatosis and NASH respectively. Importantly, the results are in agreement with human data because GPR55 levels were equally up-regulated in the liver of patients with obesity and...
with only steatosis or with NASH. This up-regulation is independent of obesity because patients without obesity and with NAFLD also showed higher expression of GPR55. Overall, these results support the hypothesis that increased levels of endogenous LPI and GPR55 are early events in the progression of liver disease by causing dyslipidemia and metabolic dysregulation of the liver. It is important to highlight that the data on GPR55 in the liver are homogenous in both rodents and humans. This is not the case for
GPR55 in white adipose tissue, where its mRNA expression was decreased in rodents with obesity but increased in the adipose tissue of patients with obesity.\(^{(19)}\) Moreover, although the LPI/GPR55 system induced lipogenic gene expression in human adipose tissue, it failed to promote adipocyte differentiation in mouse-derived 3T3-L1 cells.\(^{(19)}\)

In agreement with the stimulation of lipid storage induced by LPI in two different human hepatic cell lines (THLE2 and HepG2) and rodents, the knockdown of hepatic GPR55 ameliorated oleic acid–induced, LPI-induced, and HFD-induced steatosis, as well as MCD diet–induced and CCl4–induced NASH. This is consistent with the results of a previous report indicating that direct injection of exogenous LPIs can rapidly increase the expression of inflammatory and fibrotic genes in HFD–fed mice with Mboat7 knockdown.\(^{(36)}\) Targeting specific tissues is a relevant issue because GPR55 is widely expressed in relevant tissues.\(^{(8,37)}\) Several studies have reported on the metabolic phenotype of mice lacking GPR55, showing inconsistent results. Although two studies have found increased adiposity and insulin resistance in GPR55-deficient mice,\(^{(38,39)}\) others have reported that their tendency toward increased obesity was not significant.\(^{(40)}\) Moreover, differences in body composition were also not reproducible.\(^{(38,39)}\) From among these studies assessing the lack of GPR55, one has shown that GPR55 deficiency was associated with impaired insulin signaling in the liver.\(^{(38)}\) However, it is likely that all GPR55-expressing organs differentially contribute to the phenotype of these global knockout mice, and the lack of GPR55 thus does not prove that hepatic GPR55 was contributing to this insulin-signaling impairment. Although we failed to detect any difference in glucose tolerance after the inhibition of hepatic GPR55 in mice fed an HFD (data not shown), we found a clear reduction in the lipid content, as well as a decreased expression of inflammatory and apoptotic markers. Our viroge-netic tool showed that with just a partial (60%-70%) reduction in GPR55 levels in the liver of mice, it is possible to achieve a clear-cut amelioration of liver damage in different models of NAFLD and NASH. These in vivo results were consistent with results obtained in human hepatic cell lines, indicating that the LPI/GPR55 system can directly regulate lipid metabolism in the liver.

Because GPR55 expression was augmented in both isolated hepatocytes and HSCs from mice challenged with MCD diet–induced NASH, an important point was to investigate whether the LPI/GPR55 pathway was relevant in both cell types. Our findings indicate that LPI increases lipid content in hepatocytes, an effect that was blunted by silencing GPR55. In addition, LPI increased the expression of profibrotic genes, such as ACTA2, COL1α1, and COL1α2, an effect that was again suppressed when GPR55 was silenced in HSCs. These in vitro findings agree with the amelioration of fibrosis when GPR55 was knocked down in mice fed an MCD diet or treated with CCl4. Therefore, these results indicate that the activation of GPR55 is involved in fibrosis.

ACC is a biotin-dependent enzyme that catalyzes the irreversible carboxylation of acetyl-CoA to
produce malonyl-CoA, therefore providing the substrate for the biosynthesis of fatty acids. Preclinical studies have demonstrated that the lack or inhibition of ACCα and ACCβ reduced steatosis.\(^{(25,41-44)}\) Clinical trials showed that GS-0976, an inhibitor of ACC, also reduced hepatic steatosis and fibrosis markers in patients with NASH.\(^{(24,45)}\) Taking into account that (1) pACC is regulated by LPI/GPR55, (2) genetic silencing of ACC blunted the effects of LPI in hepatocytes and HSCs, (3) mice treated GS-0976 did not show LPI-induced steatosis, and (4) GPR55 was consistently expressed in mice and patients with NAFLD, our results suggest that the LPI/GPR55 system modulates hepatic lipid metabolism by activating ACC. Our results obtained in human hepatocytes and animal models indicate that the mechanism mediating that activation involves AMPKα, the main enzymatic upstream regulator of ACC. Indeed, an inability to describe how LPI/GPR55 regulates the phosphorylation of AMPKα is a limitation of the present study, and the potential role of some upstream modulators of AMPK, such as liver kinase B1 and/or calmodulin-dependent protein kinase kinase β, just to name a few, should be further assessed.

Although the promotion of steatosis by LPI is an event somewhat expected for a molecule activating ACC and thereby inducing \textit{de novo} lipogenesis in hepatocytes, the consequences of the activation of ACC in HSCs are still not understood. Quiescent HSCs contain ample triglycerides\(^{(46)}\) and express ACC and other genes involved in \textit{de novo} lipogenesis,\(^{(47)}\) suggesting that \textit{de novo} lipogenesis was operative during quiescence and restrained during activation of HSCs.\(^{(47,48)}\) However, a recent report has shown that activated HSC have increased levels of ACCα.\(^{(49)}\) In addition, another study has demonstrated a direct role for ACC in driving hepatic fibrosis through the activation of HSCs into collagen-secreting myofibroblasts, whereas ACC inhibition had an antifibrotic activity.\(^{(50)}\) Our results indicated that the activation of the LPI/GPR55 system triggered ACC activity and the expression of profibrotic markers in HSCs, and these effects were blocked after silencing ACC in these cells. Thus, our results support a role for ACC in the activation of HSCs.

In summary, these results indicate that (1) GPR55 expression is increased in the liver of patients with liver damage and in animal models of liver damage; (2) the endogenous ligand of GPR55 called LPI is, notably, positively correlated with the NAS in humans and causes steatosis in a GPR55 manner; (3) the knockdown of hepatic GPR55 ameliorates LPI-induced and HFD-induced NAFLD, as well as MCD-induced and CCl\(_4\)-induced NASH; and (4) the LPI/GPR55 system activates ACC through AMPK to cause liver disease.

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