The HBV Specially-Related Long Noncoding RNA HBV-SRL Involved in the Pathogenesis of Hepatocellular Carcinoma

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Hepatitis B virus (HBV) is one of the major risk factors for HCC (hepatocellular carcinoma) occurrence with a diverse role in the pathogenesis of HCC. More works need to be performed to elucidate a more thorough understanding of the molecular mechanisms involving in HBV-induced HCC, although some mechanisms such as genome integration have been reported. In the present study, aberrantly expressed lncRNAs were identified between HCC tumor tissues with or without HBV infection. Among these molecules, HBV-specially-related long noncoding RNA (HBV-SRL) was further found to correlate with poor prognosis and a shorter overall survival time in HCC patients with HBV infection. Additionally, HBV-SRL was found function as oncogene by upregulating the NF-κB expression. These data suggest that HBV infection altered gene expression pattern in liver cells which contributed to HBV-related HCC development, and HBV-SRL may serve as a new molecular marker or potential therapeutic target of HBV-related HCC.

1. Introduction

Hepatitis B virus (HBV) infection is one of the major risk factors for HCC (hepatocellular carcinoma) occurrence [1]. More than half of all patients with HCC are etiologically linked to an infection [2]. Improvement in the prognosis of this disease via anti-HBV therapy has highlighted the crucial role of HBV in HCC pathogenesis and progression [3]. The integration of HBV DNA into the genome of hepatocytes was reported to be the main reason of HBV-related HCC (HBV-HCC) development due to its role in promoting genomic instability and mutagenesis [4]. Viral proteins such as HBX contributed to HBV-HCC via signaling through Akt or NF-kB pathways, modulating DNA methylation and histone modification patterns, among other roles [5]. Persistent inflammation and antiviral response induced by HBV infection increased host susceptibility to malignant transformation [6]. Taken together, these findings suggest a diverse role of HBV in the pathogenesis of HCC. However, more work needs to be performed to elucidate a more thorough understanding of the molecular mechanisms of HBV-induced HCC.

Long noncoding RNAs (lncRNAs), a novel type of noncoding RNA with pivotal roles in epigenetic regulation, were found to function as regulators of viral replication or antiviral response [7, 8]. Both viral lncRNAs and cellular lncRNAs induced by infection were found in infected cells [9]. Viral lncRNAs promoted viral replication via autonomous replication [10] and decreased the antiviral response by transcriptionally regulating the viral and host genomes.
Cellular lncRNAs induced by infection were mainly found to participate in the antiviral response. Cellular lncRNAs such as NEAT1 (nuclear enriched abundant transcript 1) and BISPR (bone marrow stromal antigen 2 IFN-stimulated positive regulator) were found to activate the antiviral response by modulating antiviral factors [13], while lncRNAs such as IncRNA IL7R and THRIL (TNF-alpha and heterogeneous nuclear ribonucleoprotein L-related immunoregulatory LncRNA) counteracted the antiviral response by negatively controlling the IFN pathway [14–16]. Aside from their role in viral infection or suppression, lncRNAs have also been shown to be involved in diverse biological cellular processes including tumorigenesis [17]. Aberrantly expressed lncRNAs, such as H19 and MVH, had been identified in multiple tumors including HCC [18]. H19 in liver cancer cells induces drug resistance through regulation of multidrug resistance 1 (MDR1) promoter methylation and suppressed metastasis via a miR-220 dependent pathway [19]. MVH (micrornanucleotide invasion in HCC) promoted tumor growth and intrahepatic metastasis by activating angiogenesis [20]. In addition, our previous study found that ICR (ICAM-1-related IncRNA), which is expressed in liver cancer stem cells (CSC), regulated CSC properties of ICAM-1+ HCC cells and promoted tumor cell migration [21]. These findings highlight the crucial roles of IncRNA in the pathogenesis and progression of HCC. Thus, it is possible that lncRNAs impact HBV-induced HCC. While lncRNAs expressed in HBV-HCC such as highly upregulated in liver cancer (HULC) and HBX-long interspersed nuclear elements 1 (HBX-LINE1) have been identified, additional studies are required to fully characterize the effect of lncRNAs in HBV-induced HCC.

In the present study, we analyzed the expression profile of IncRNA and mRNA in HBV-related and unrelated HCC tissues using cDNA microarray and found differently expressed lncRNAs and mRNAs between these tissues. One of the differentially expressed lncRNAs, termed as HBV-related long noncoding RNA (HBV-SRL), was found to be upregulated in HBV-induced HCC. HBV-SRL correlated with the poor prognosis and reduced survival of HCC patients with HBV infection. Functionally, HBV-SRL promoted tumor cell proliferation by upregulating NF-kB2 via activating NF-kB2 transcription. Our findings suggest that HBV-SRL may represent a new molecular marker and potential therapeutic target of HBV-induced HCC.

2. Materials and Methods

2.1. Patients and Follow-Up. HCC patients with or without HBV (n = 232) who underwent radical cancer resection at Eastern Hepatobiliary Surgery Hospital (EHBH) were recruited for microarray analysis and validation, respectively, in the present study. The clinical characteristics of the patients are listed in Supplementary Table 1. The study was approved by the Institutional Review Board of the EHBH. Other details of patient information are listed in Supplementary Materials.

2.2. Statistical Analysis. Student’s t-tests were used to compare two groups unless otherwise indicated (χ2 test). Categorical data were analyzed using the Fisher exact test, and quantitative variables were analyzed using the t tests or Pearson’s correlation test. Survival was calculated with the log-rank test. The Cox regression model was used to perform multivariate analysis. P < 0.05 was considered statistically significant.

Detail of other materials and methods is listed in Supplementary Materials.

3. Results

3.1. HBV-SRL Was Higher in Hepatocellular Carcinoma with HBV. To identify lncRNAs associated with HBV-HCC, we collected HCC tumor tissues with (Group 1, n = 5, HBV+) or without HBV (Group 2, n = 5, HBV−) infection and analyzed lncRNA expression profiles in these tissues using cDNA microarrays.

Differentially expressed lncRNAs profiles were identified between tumor tissues (T) and the corresponding parenchyma tumor tissues (PT) in both groups (Supplementary Figure S1), indicating the involvement of lncRNAs in HCC development. To find out HBV specific-related lncRNAs, differentially expressed lncRNAs between HBV+ and HBV− tumors were also investigated. 183 upregulated lncRNAs and 241 downregulated lncRNAs were identified to be differentially expressed between HBV+ and HBV− tumors (Figures 1(a) and 1(b)). Among these lncRNAs, 72 lncRNAs were found upregulated in HBV+ tumors compared with the corresponding HBV− PTs and 88 lncRNAs were found downregulated (Figure 1(b)). Based on these findings, four upregulated lncRNAs with a 6-fold greater expression in HBV+ TVs and HBV+ PT and 2.5-fold greater in HBV− TVs and HBV− T were selected as candidate (Supplementary Table 2). We validated our microarray results using real-time PCR, highlighting the elevated expression of our candidate lncRNAs in T versus PT in both HBV+ (Figure 1(c)) and HBV+ patients (Figure 1(c)). AK128595 was selected for further analysis due to its greatest increase. Then, the AK128595 expression was verified using a validation cohort of 30 HBV+ tumors, and the result showed higher level of AK128595 in HBV+ T compared with HBV+ PT (Figure 1(d)). In the present study, AK128595 was identified as HBV-related long noncoding RNA (HBV-SRL).

3.2. HBV-SRL Produced No Protein HBV-SRL. An evaluation of the HBV-SRL sequence identified an open reading frame (ORF) with 504 nucleotides in length, suggestive of a potential protein product (Figure 2(a)). Bioinformatic analysis (CPC, CNCI, and PFAM) also found the coding potential of HBV-SRL (Figure 2(b)). To evaluate these findings, a pAdeno-HBV-SRL-His plasmid expressing HBV-SRL with polyhistidine (6XHis) insertion into the ORF just before the stop codon was transfected into CSQT-2 cells (Supplementary Figure S2). After validating enhanced HBV-SRL expression (Figure 2(c)), His antibody was used to detect whether His-tagged protein was produced via western...
blot. No specific signal was observed in cells transfected with pAdeno-His (Control) or pAdeno-HBV-SRL-His (HBV-SRL-His), while an enhanced signal was observed in cells transfected with pSMYD4-His (SMYD4-His) (Figure 2(c)), indicating that no His-tagged protein was produced by pAdeno-HBV-SRL-His. Moreover, antibodies against the potential HBV-SRL protein (anti-HBV-SRP) were produced using the synthesized peptide. Following HBV-SRL
upregulation by pAdeno-HBV-SRL-His transfection and subsequent downregulation by siRNA transfection in CSQT-2 cells (Figure 2(d)), anti-HBV-SRLP was used to detect protein changes. No significant differences in western blot analysis were observed despite changes in HBV-SRL mRNA (Figure 2(d)). The similar results were also observed

| RNA_ID   | Length | CPC  | CNCI | PFAM | CPAT |
|----------|--------|------|------|------|------|
| AK128595.1 | 3764   | 1.06 | -0.06 | Significance | 0.488 |

Figure 2: Investigation of HBV-SRL coding capacity. (a) Diagram of HBV-SRL transcript. Potential open reading frame (ORF) ranged from 830 to 1333. (b) Bioinformatic analysis of coding capacity of HBV-SRL using CPC, CNCI, PFAM, and CPAT. CPC, coding potential calculator; CNCI, coding-noncoding index; CPAT, coding potential assessment tool. Transcript with CPC score < 0, CNCI score < 0, PFAM = nonsignificant, and CPAT < 0.364 was a potential IncRNA. (c) Real-time PCR analysis of HBV-SRL (A) and western blot analysis of His expression (B) in CSQT-2 cells transfected with plasmid pAdeno-His (Control), pAdeno-HBV-SRL-His (HBV-SRL-His), or pSMYD4-His (SMYD4-His). (d) Real-time PCR analysis of HBV-SRL (A) and western blot analysis by anti-HBV-SRLP antibody (B) using CSQT-2 cells transfected with plasmid pAdeno-His (Control), pAdeno-HBV-SRL-His (HBV-SRL-His), or siRNAs targeting HBV-SRL (si-HBV-SRL). The error bars represent the standard deviation (SD) of data obtained in at least three independent experiments, **P < 0.01.
in Hep3B cells (Supplementary Figure S3). These data indicated that no protein can be produced by HBV-SRL, and HBV-SRL was a noncoding RNA.

3.3. HBV-SRL Functions as an Oncogene by Upregulating the NF-κB2 Expression. Having documented that HBV-SRL does not produce a protein product, we investigated how the HBV-SRL expression affects tumor cells. To this end, we aligned the HBV-SRL sequence with the gene promoters (upstream 1.5 kb) using Blat to find out promoter regions in which HBV-SRL can bind, since lncRNAs was previously reported to function as the transcription factor [22]. RELB and NF-κB2 were retrieved, while the RELB promoter had 324 bp nucleotides similar to HBV-SRL and the NF-κB2 promoter had 238 bp nucleotides complemented with HBV-SRL (Supplementary Table 3). The effects of HBV-SRL on NF-κB2 were further investigated to functionally validate whether HBV-SRL binds to the NF-κB2 promoter. The dual-luciferase reporter assay was performed to investigate whether HBV-SRL binds to the NF-κB2 promoter. After HBV-SRL was upregulated by pAdeno-HBV-SRL-His transfection into CSQT-2 cells, pGL3-NF-κB2 promoter-wild type (pGL3-WT) or pGL3-NF-κB2 promoter-mutant (pGL3-MUT) with the complementary sequence deleted was transfected. The dual-luciferase reporter assay showed an higher luciferase activity in cells transfected with pGL3-WT compared with those transfected with the pGL3-enhancer (pGL3) (Figure 3(a)), and no significant change in luciferase activity was observed in CSQT-2 cells transfected with pGL3-MUT, suggesting that HBV-SRL binds to the NF-κB2 promoter via sequence complement. To verify whether HBV-SRL regulates the NF-κB2 expression, NF-κB2 mRNA and protein were measured after upregulation or down-regulation of HBV-SRL. Higher levels of NF-κB2, at the both mRNA and protein levels, were observed when HBV-SRL was upregulated in both CSQT-2 and Hep3B cells (Figure 3(b) and Supplementary Figure S4(a)). Conversely, reduced NF-κB2 mRNA and protein were observed when HBV-SRL was downregulated (Figure 3(c) and Supplementary Figure S4(b)), suggesting that HBV-SRL regulated the expression of NF-κB2. As NF-κB2 was reported to regulate tumor cell growth [23, 24], cell cycle and proliferation capacity of tumor cells were assessed following HBV-SRL downregulation. When HBV-SRL was downregulated by siRNAs (si-HBV-SRL) in CSQT-2 cells, tumor cells displayed higher percentage of cells in G1 phase and lower percentage of cells in G2 phase (Figure 3(d) and Supplementary Figure S5(a)). This alteration of cell cycle induced by HBV-SRL downregulation was also observed in Hep3B cells (Supplementary Figures S5(b) and S5(c)), indicating that HBV-SRL affected cell cycle. Moreover, CSQT-2 cell growth was slowed down by HBV-SRL downregulation (Figure 3(d)). In addition, the mice model was established by injecting Hep3B cells into mice subcutaneously to investigate whether HBV-SRL downregulation inhibited tumor growth in vivo. When tumor nodes appeared, si-HBV-SRL or NC was administrated in situ twice a week for four weeks. During this period, the tumor size was measured every three days. Five weeks later, all the mice were sacrificed and tumor tissues were weighted and sent for further analysis. Tumors treated with si-HBV-SRL were much smaller in volume than those treated with NC (N = 6 for each group) (Figure 3(e)). Tumor growth curve showed that growth of tumors administrated by si-HBV-SRL was much slower than that of control tumors (Figure 3(e)), indicating that si-HBV-SRL administration slowed down tumor growth in mice. The results show that HBV-SRL can regulate tumor cell cycle and growth via NF-κB2 regulation.

3.4. HBV-SRL/NF-κB2 Is Differentially Expressed in Tumors Compared to Paired Liver in Different HBV DNA Levels. Having demonstrated that HBV-SRL upregulated the NF-κB2 expression and was involved in cell migration in vitro, we evaluated the relationship between the HBV-SRL and NF-κB2 expression and recurrence in HCC patients with different level of HBV DNA. For this purpose, 222 HCC patients were analyzed as a validation cohort. Real-time PCR was used to measure the expression of HBV-SRL and NF-κB2 in tumors (T) and paired liver tissues (PT). Both the expression of HBV-SRL and NF-κB2 were higher in tumor tissues than in paired liver tissues (Figure 4(a) and Supplementary Figure S6(a)). According to the results of auxiliary examination (HBV DNA quantitative test), 222 HCC patients were divided into two groups: high HBV DNA level (HBV DNA ≥ 10^4 copy/ml) (n = 95) and low HBV DNA level (HBV DNA < 10^4 copy/ml) (n = 127). We then compared the relative expression level of HBV-SRL (T Vs PT) between the two groups. Higher level of HBV-SRL was observed in the high HBV DNA level group (HBV-high) (Figure 4(b)). We then analyzed the relationship between the HBV-SRL expression and clinicopathologic features in HCC patients and identified association between the HBV-SRL expression and absent tumor encapsulation (P = 0.044) (Table 1). Univariate analysis found HBV-SRL as well as NF-κB2 and HBV DNA was associated with DFS and OS (Table 2). Further, multivariate Cox hazards analysis found the association of HBV-SRL with prognosis in HBV-related HCC patients (Figure 4(c)). The median DFS and OS were 20.13 months and 22.90 months, respectively, in patients with low HBV-SRL expression. These findings were significantly longer than those observed for patients with a high HBV-SRL expression (12.63 and 20.20 months, respectively; both P < 0.001). Kaplan–Meier analysis demonstrated that patients with low HBV-SRL expression had better prognosis than those with a high HBV-SRL expression (Figure 4(d)). The NF-κB2 expression was also associated with DFS and OS (Supplementary Figures 6(b) and 6(c)). The results suggested that both the high expressions of HBV-SRL and NF-κB2 correlate with the poor prognosis of HCC patients with HBV infection.

4. Discussion

In the present study, we performed a transcriptional analysis of lncRNA expression in tumor tissues with and without
HBV infection. Among differentially expressed lncRNAs, HBV-SRL was investigated due to in silico predictions of interactions with the NF-κB2 promoter and subsequent in vitro studies validating the effects of HBV-SRL on the NF-κB2 expression. Additionally, the HBV-SRL level was found to positively correlate with HBV DNA status and associate with poor prognosis of HCC patients with HBV infection. The results suggest that HBV infection induced the aberrant expression of lncRNAs that contributed to HCC development.
Figure 4: Correlation of the HBV-SRL expression with prognosis of HCC patients with HBV infection. (a) Real-time PCR analysis of HBV-SRL expression in tumor tissues (T) and corresponding parenchyma tumor tissues (PT) from 222 HCC patients with HBV. The error bars represent the standard deviation (SD) of data obtained in at least three independent experiments, ** P < 0.01. (b) Expression of HBV-SRL in tumor tissues with high (≥10^4 copy/ml, HBV-High) or low (<10^4 copy/ml, HBV-Low) HBV DNA level. (c) Association of HBV-SRL with disease-free survival (DFS) or overall survival (OS) of HCC patients with HBV infection analyzed by multivariate Cox hazards model. (d) Correlations of HBV-SRL with DFS and OS of 222 HCC patients analyzed by Kaplan–Meier’s analyses.
Nowadays, several studies have reported that HBV infection altered the expression of noncoding RNAs such as miRNAs and lncRNAs in liver cells. Our previous study and others found aberrant miRNA expression in the liver of HBx transgenic mice contributed to the HCC development [25, 26]. Similarly, microarray analysis of HBV-related HCC...
and normal liver tissues showed an altered lncRNA expression profile in tumor tissues, further indicating the crucial role of lncRNA in HBV-related HCC [27, 28]. LncRNAs such as highly upregulated LncRNAs in liver cancer (HULC) and HBX-long interspersed nuclear elements1 (HBX-LINE1) have been also reported to contribute to HBV-induced HCC development via different ways [29]. By comparing with gene expression profiles from HBV+ HCC tumor tissues and HBV-HCC tumor tissues, we identified a unique gene expression profile in HBV+ HCC tumor tissues, which included lncRNAs. Moreover, our analysis highlighted the association of genes involved in tumor development and recurrence with aberrantly expressed lncRNAs induced by HBV infection (data not shown). These findings suggested a crucial and complex role by which HBV infection lead to the development of HCC. Besides, different gene expression profiles were also found between HBV-HCC tumors and the corresponding parenchyma tumor tissues, indicating that there many genes contributed to tumor development without HBV affection.

HBV infection is one of the most important factors associated with the poor prognosis of HBV-related HCC. In China, more than 90% of patients are found to have advanced HCC with HBV when the liver cancer is first diagnosed [30]. Although TACE, radiotherapy, and symptomatic treatment were used, hepatectomy still remains the most effective treatment [31]. However, not all patients have a bad prognosis due to the high HBV DNA level. Our results showed that HBV-SRL may be a novel predictor associated with malignant features of HBV-related HCC and is correlated with patients’ prognosis. Moreover, HBV-SRL was found to associate with absent tumor encapsulation which suggested that HBV-SRL may be correlated with malignant tumors progression. HCC patients with high levels of HBV-SRL in parenchyma tumor should be monitored at increased intervals to prevent tumor metastasis or recurrence.

Long noncoding RNAs function as key regulators in multiple cellular processes including tumorigenesis via numerous molecular mechanisms [32]. In the present study, we found that lncRNA HBV-SRL functions as an oncogene regulating NF-κB2 expression via binding to the NF-κB2 promoter. HBV-SRL acts as a transcription factor with a detailed understanding of the molecular mechanism of its actions remains to be elucidated. Moreover, we found the slowdown of tumor cell growth by HBV-SRL down-regulation via affecting cell cycle. However, HBV-SRL may

| Factor                   | OS  | DFS |
|--------------------------|-----|-----|
| Age (yr)                 |     |     |
| >60 Vs ≤ 60              | 0.427 | 0.008 |
| Sex                      |     |     |
| Male vs. female          | 0.417 | 0.107 |
| HBsAg                    |     |     |
| Positive vs. negative    | 0.53  | 0.374 |
| HBV DNA                  |     |     |
| >10^4 Vs ≤ 10^4          | <0.001 | <0.001 |
| AFP (μg/L)               |     |     |
| >20 Vs ≤ 20              | 0.004  | 0.003 |
| ALT (U/L)                |     |     |
| >41 Vs ≤ 41              | 0.209  | 0.028 |
| AST (U/L)                |     |     |
| >37 Vs ≤ 37              | 0.002  | <0.001 |
| Tumor number             |     |     |
| Multiple vs. single      | 0.009  | <0.001 |
| Liver cirrhosis          |     |     |
| Yes vs. no               | 0.156  | 0.598 |
| Tumor size (cm)          |     |     |
| >5 Vs ≤ 5                | <0.001 | <0.001 |
| Micrometastases          |     |     |
| Yes Vs. no               | <0.001  | <0.001 |
| Tumor encapsulation      |     |     |
| Yes Vs. no               | <0.001  | <0.001 |
| Microvascular invasion   |     |     |
| Yes Vs no                | 0.013  | <0.001 |
| HBV-SRL                  |     |     |
| High Vs. low             | 0.002  | 0.043 |
| NF-κB2                   |     |     |
| High Vs. low             | 0.001  | 0.041 |

OS, overall survival; DFS, disease-free survival; PVTT, portal vein tumor thrombus; HBsAg, hepatitis B surface antigen; HBV DNA, hepatitis B virus deoxyribonucleic acid; TNM, tumor-node metastasis; AFP, α-fetoprotein; NA, not adopted; NS, not significant; Low/high, HBV-SRL, and NFκB levels were lower/higher than the median value.
have roles in other biological processes such as inflammation and immune response due to its regulation of NF-κB2 which have crucial roles in these processes [33].

Additionally, the coding capacity of HBV-SRL was investigated by both bioinformatic analysis and in vitro experiments. No protein produced by pAdeno-HBV-SRL-His was observed in pAdeno-HBV-SRL-His transfected tumor cells although three out of four bioinformatic analysis tools indicated that HBV-SRL could have protein coding ability (Figure 2). It has been reported that several RNA transcripts annotated as lncRNAs produce micropeptides with biological functions [34, 35]. Thus, evaluating the protein coding capacity using experiments is necessary.

In conclusion, we describe a novel HBV-related lncRNA HBV-SRL that contributes to the pathogenesis of HCC in patients with HBV infection. High tumor HBV-SRL expression level was associated with higher rates of recurrence and poor prognosis after hepatectomy in HBV-related HCC, suggesting that HBV-SRL may serve as a new diagnostic marker for tumor recurrence and a potential target for HBV-related oncogenes inhibition.

Data Availability

The data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors have declared that there are no conflicts of interest.

Authors’ Contributions

Cunzhen Zhang, Lei Lu, and Haibei Xin carried out the studies, participated in collecting data, and drafted the manuscript. Minfeng Zhang performed the statistical studies, participated in collecting data, and drafted the manuscript. Cunzhen Zhang, Lei Lu, and Haibei Xin contributed equally to this work.

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Supplementary Materials

The supplementary materials for this article include 6 figures and 4 tables, and the contents are shown by the figures: Supplementary Figure 1: altered expressed lncRNAs between tumor and corresponding parenchyma tumor tissues. Supplementary Figure 2: diagram of the constructs used for the HBV-SRL-His expression. Supplementary Figure 3: expression of HBV-SRL in Hep3B cells transfected with plasmids or siRNAs. Supplementary Figure 4: expression of NF-κB2 in Hep3B cells transfected with plasmids or siRNAs. Supplementary Figure 5: cell cycle analysis of tumor cells using flow cytometry. Supplementary Figure 6: expression of NF-κB2 in tumor tissues and its correlation with prognosis of HCC patients with HBV infection. The contents are shown by tables: Supplementary Table 1: clinicopathologic features of 222 HCC patients with HBV. Supplementary Table 2: upregulated lncRNAs in HBV + tumors selected as candidate molecules. Supplementary Table 3: the promoters associated with HBV-SRL from Blat analysis. Supplementary Table 4: primers and siRNA sequences. Also, a document describing patients’ information and some experimental methods is included. (Supplementary Materials)

References

[1] A. G. Singal, P. Lampertico, and P. Nahon, “Epidemiology and surveillance for hepatocellular carcinoma: new trends,” Journal of Hepatology, vol. 72, no. 2, pp. 250–261, 2020.
[2] P. Mettikanont, C. Bunchornvatavkul, and K. R. Reddy, “Systematic review: epidemiology and response to direct-acting antiviral therapy in genotype 6 chronic hepatitis C virus infection,” Alimentary Pharmacology & Therapeutics, vol. 49, no. 5, pp. 492–505, 2019.
[3] G. K. Abou-Alfa, Q. Shi, J. J. Knox et al., “Assessment of treatment with sorafenib plus doxorubicin vs sorafenib alone in patients with advanced hepatocellular carcinoma: phase 3 calgb 80802 randomized clinical trial,” JAMA Oncology, vol. 5, 2019.
[4] K. M. Sze, D. W. Ho, Y. T. Chiu et al., “HBV-TERT promoter integration harnesses host ELF4 resulting in TERT gene transcription in hepatocellular carcinoma,” Hepatology, vol. 73, 2020.
[5] X. D. Zhang, Y. Wang, and L. H. Ye, “Hepatitis B virus X protein accelerates the development of hepatoma,” Cancer biology and medicine, vol. 11, no. 3, pp. 182–190, 2014.
[6] Q. Gao, H. Zhu, L. Dong et al., “Integrated proteogenomic characterization of HBV-related hepatocellular carcinoma,” Cell, vol. 179, pp. 561–577.e22, 2019.
[7] P. Fortes and K. V. Morris, “Long noncoding RNAs in viral infections,” Virus Research, vol. 212, pp. 1–11, 2016.
[8] L. Stojic, A. T. L. Lun, P. Mascalchi et al., “A high-content RNAi screen reveals multiple roles for long noncoding RNAs in cell division,” Nature Communications, vol. 11, no. 1, 2020.
[9] J. Ouyang, J. Hu, and J. L. Chen, “lncRNAs regulate the innate immune response to viral infection,” WIREs RNA, vol. 7, no. 1, pp. 129–143, 2016.
[10] S. Gago-Zachert, “Viroids, infectious long non-coding RNAs with autonomous replication,” Virus Research, vol. 212, pp. 12–24, 2016.
[11] V. M. Noriega, K. K. Haye, T. A. Kraus et al., “Human cytomegalovirus modulates monocyte-mediated innate immune responses during short-term experimental latency in vitro,” Journal of Virology, vol. 88, no. 16, pp. 9391–9405, 2014.
[12] S. L. Moon, J. G. Blackinton, J. R. Anderson et al., “XRN1 stalling in the 5’ UTR of Hepatitis C virus and Bovine Viral Diarrhea virus is associated with dysregulated host mRNA stability,” PLoS Pathogens, vol. 11, no. 3, Article ID e1004708, 2015.
I. Ulitsky and D. P. Bartel, “lincRNAs: genomics, evolution, and mechanisms,” *Molecular Cell*, vol. 54, 2014.

H. Cui, N. Xie, Z. Tan et al., “The human long noncoding RNA Inc-IL7R regulates the inflammatory response,” *European Journal of Immunology*, vol. 44, no. 7, pp. 2085–2095, 2014.

Z. Li, T. C. Chao, K. Y. Chang et al., “The long noncoding RNA THRIL regulates TNFα expression through its interaction with hnRNPL,” *Proceedings of the National Academy of Sciences*, vol. 111, pp. 1002–1007, 2014.

X. Yang, M. Bam, W. Becker, P. S. Nagarkatti, and M. Nagarkatti, “Long noncoding RNA AW112010 promotes the differentiation of inflammatory T cells by suppressing IL-10 expression through histone demethylation,” *The Journal of Immunology*, vol. 205, no. 4, pp. 987–993, 2020.

J. K. Kulski, “Long noncoding RNA HCP5, a hybrid HLA class I endogenous retroviral gene: structure, expression, and disease associations,” *Cells*, vol. 8, 2019.

M. A. Parasramka, S. Maji, A. Matsuda, I. K. Yan, and T. Patel, “Long non-coding RNAs as novel targets for therapy in hepatocellular carcinoma,” *Pharmacology & Therapeutics*, vol. 167, 2016.

L. Zhang, F. Yang, J. H. Yuan et al., “Epigenetic activation of the MiR-200 family contributes to H19-mediated metastasis suppression in hepatocellular carcinoma,” *Carcinogenesis*, vol. 34, no. 3, pp. 577–586, 2013.

S. X. Yuan, F. Yang, Y. Yang et al., “Long noncoding RNA associated with microvascular invasion in hepatocellular carcinoma promotes angiogenesis and serves as a predictor for hepatocellular carcinoma patients’ poor recurrence-free survival after hepatectomy,” *Hepatology*, vol. 56, no. 6, pp. 2231–2241, 2012.

W. Guo, S. Liu, Y. Cheng et al., “ICAM-1-related noncoding rna in cancer stem cells maintains ICAM-1 expression in hepatocellular carcinoma,” *Clinical cancer research: an official journal of the American Association for Cancer Research*, vol. 22, 2015.

I. Ulitsky and D. P. Bartel, “lincRNAs: genomics, evolution, and mechanisms,” *Cell*, vol. 154, no. 1, pp. 26–46, 2013.

B. Barre, O. Coqueret, and N. D. Perkins, “Regulation of activity and function of the p52 NF-κB subunit following DNA damage,” *Cell Cycle*, vol. 9, no. 24, pp. 4795–4804, 2010.

R. De, S. Sarkar, S. Mazumder et al., “Macrophase migration inhibitory factor regulates mitochondrial dynamics and cell growth of human cancer cell lines through CD74–NF-κB signaling,” *Journal of Biological Chemistry*, vol. 293, no. 51, pp. 19740–19760, 2018.

X. Zhang, S. Liu, T. Hu, S. Liu, Y. He, and S. Sun, “Up-regulated microRNA-143 transcribed by nuclear factor kappa B enhances hepatocarcinoma metastasis by repressing fibronectin expression,” *Hepatology*, vol. 50, no. 2, pp. 490–499, 2009.

N. Sarkar and R. Chakravarty, “Hepatitis B virus infection, MicroRNAs and liver disease,” *International Journal of Molecular Sciences*, vol. 16, no. 8, pp. 17746–17762, 2015.

Y. F. Pan, T. Qin, L. Feng, and Z. J. Yu, “Expression profile of altered long non-coding RNAs in patients with HBV-associated hepatocellular carcinoma,” *Journal of Huazhong University of Science and Technology—Medical sciences*, vol. 33, no. 1, pp. 96–101, 2013.

Y. Deng, Z. Wei, M. Huang et al., “Long non-coding RNA F11-AS1 inhibits HBV-related hepatocellular carcinoma progression by regulating NR1I3 via binding to microRNA-211-5p,” *Journal of Cellular and Molecular Medicine*, vol. 24, no. 2, pp. 1848–1865, 2020.

B. Moyo, S. A. Nicholson, and P. B. Arbuthnot, “The role of long non-coding RNAs in hepatitis B virus-related hepatocellular carcinoma,” *Virus Research*, vol. 212, pp. 103–113, 2016.

J. Shi, E. C. H. Lai, N. Li et al., “A new classification for hepatocellular carcinoma with portal vein tumor thrombus,” *Journal of Hepato-Biliary-Pancreatic Sciences*, vol. 18, no. 1, pp. 74–80, 2011.

K. Sakamoto and H. Nagano, “Surgical treatment for advanced hepatocellular carcinoma with portal vein tumor thrombus,” *Hepatology Research*, vol. 47, no. 10, pp. 957–962, 2017.

O. M. Rogoyski, J. I. Pueyo, J. P. Couso, and S. F. Newbury, “Functions of long non-coding RNAs in human disease and their conservation in Drosophila development,” *Biochemical Society Transactions*, vol. 45, no. 4, pp. 895–904, 2017.

T. Brue, M. H. Quentien, K. Khetchoumian et al., “Mutations in NFKB2 and potential genetic heterogeneity in patients with DAVID syndrome, having variable endocrine and immune deficiencies,” *BMC Medical Genetics*, vol. 15, p. 139, 2014.

D. M. Anderson, K. M. Anderson, C. L. Chang et al., “A micropeptide encoded by a putative long noncoding RNA regulates muscle performance,” *Cell*, vol. 160, no. 4, pp. 595–606, 2015.

B. R. Nelson, C. A. Makarewich, D. M. Anderson et al., “A peptide encoded by a transcript annotated as long noncoding RNA enhances SERCA activity in muscle,” *Science*, vol. 351, pp. 271–275, 2016.