Research article

Fragments of the key flowering gene GIGANTEA are associated with helitron-type sequences in the Pooideae grass *Lolium perenne*

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Abstract

**Background:** Helitrons are a class of transposable elements which have been identified in a number of species of plants, animals and fungi. They are unique in their proposed rolling-circle mode of replication, have a highly variable copy-number and have been implicated in the restructuring of coding sequences both by their insertion into existing genes and by their incorporation of transcriptionally competent gene fragments. Helitron discovery depends on identifying associated DNA signature sequences and comprehensive evaluation of helitron contribution to a particular genome requires detailed computational analysis of whole genome sequence. Therefore, the role which helitrons have played in modelling non-model plant genomes is largely unknown.

**Results:** Cloning of the flowering gene GIGANTEA (GI) from a BAC library of the Pooideae grass *Lolium perenne* (perennial ryegrass) identified the target gene and several GI pseudogene fragments spanning the first five exons. Analysis of genomic sequence 5’ and 3’ of one these GI fragments revealed motifs consistent with helitron-type transposon insertion, specifically a putative 5’-A↓T-3’ insertion site containing 5’-TC and CTAG-3’ borders with a sub-terminal 16 bp hairpin. Screening of a BAC library of the closely related grass species *Festuca pratensis* (meadow fescue) indicated similar helitron-associated GI fragments present in this genome, as well as non-helitron associated GI fragments derived from the same region of GI. In order to investigate the possible extent of ancestral helitron-activity in *L. perenne*, a methylation-filtered GeneThresher® genomic library developed from this species was screened for potential helitron 3’ hairpin sequences associated with a 3’-CTRR motif. This identified 7 potential helitron hairpin-types present between at least 9 and 51 times within the *L. perenne* methylation-filtered library.

**Conclusion:** This represents evidence for a possible ancestral role for helitrons in modelling the genomes of *Lolium* and related species.

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Background

Helitrons are a class of transposons which are unique in their proposed rolling-circle mode of replication mediated either autonomously by an internally coded putative DNA replication-initiator-helicase protein, or non-autonomously. They have been identified in a number of species of plants, animals and fungi and can have a highly variable copy-number, from an infrequent representation in many mammals to contributing up to 5% of the genome size in some Drosophila species (see reviews by [1,2]). They show considerable size variation (0.5 – > 15 kb for Arabidopsis helitrons, [3]) and, unusually, helitron transposition does not give rise to duplication of target sites. Helitrons insert within 5'-A$$\rightarrow$$T-3' target sites within the genome and can be recognised by conserved 5'-TC.. and ..CTRR-3' termini with, typically, 16–20 bp hairpin motifs 8–12 bp from the 3' termini.

A feature of helitron transposons is their ability to incorporate multiple genomic gene fragments which can still show transcriptional activity – thus creating the potential for novel truncated, alternatively spliced and chimeric mRNAs and proteins [4]. The mechanism by which helitrons incorporate gene fragments is not clear, though it is presumably associated with mutation or misidentification of recognition sites during the replication process, and models which describe the acquisition of gene fragments both at the 5' and at the 3' end have been proposed [1-4]. In rice, Arabidopsis and maize, the extensive genome resources have facilitated in silico identification of helitrons in these and related genera [3,5-7]. Helitrons identified in maize [4,8-12] and Ipomoea tricolor [13] have generated particular interest due to their proposed actions in creating haplotypic diversity and influencing gene function.

Lolium perenne (perennial ryegrass) and Festuca pratensis (meadow fescue) are members of the 'Lolium/Festuca complex' of interfertile grasses which form the basis of many grassland agricultural and amenity systems in temperate areas of the world. They belong to the Pooidae sub-family of the Poaceae, along with the Triticeae cereal crops and Brachypodium distachyon, the rapidly developing model for monocot species. The haploid genome sizes of L. perenne and F. pratensis are estimated to be c. 2 Gb [14,15], less than half the size of barley and the constituent genomes of hexaploid wheat [16,17] but c. 6–7 times the size of B. distachyon and rice [16]. Consequently, the intermediate genome sizes of L. perenne and F. pratensis between B. distachyon and the Triticeae cereals and the close evolutionary interrelationships of these Pooidae species, makes the Lolium/Festuca grasses of great interest in terms of understanding the processes which influence the evolution of genome organisation and size in close relatives.

GIGANTEA (GI) was originally identified as a key gene in the perception of circadian rhythms and the photoperiodic control of flowering by mutation analysis in Arabidopsis [18,19] but it is only recently that detailed knowledge of the mode of action and interaction of this gene has become available [20-23]. Comparative genome analysis between dicots and monocots has indicated that orthologues of many of the key genes involved in flowering in Arabidopsis also exist in rice and other monocots [24-26] and experimental evidence indicates that similar control mechanisms may be involved in some cases [27-31] including for GI [32,33]. Consequently, the identification of the orthologues of GI in L. perenne and other monocot crop species has been a desirable goal, partly to ascertain if it is implicated in flowering control in current breeding populations through QTL/genetic mapping studies but also to identify allelic variants which may be useful in future population development.

In this study we describe how, in the process of cloning the L. perenne orthologue of GI from a BAC library, we identified GI pseudogene fragments associated with helitron-type sequences. Similar sequences were found to be also present in the F. pratensis genome. Additionally, we describe the use of a methylation-filtered L. perenne genomic library in an initial survey to ascertain the potential frequency of helitrons within the L. perenne genome.

Results

Identification of GI and GI pseudogene sequences from L. perenne and F. pratensis BAC libraries

A primer pair, GIG49660.6F/7R (see Table 1 for primer sequences) was designed based on conserved regions spanning the first and fourth exons in existing GI sequences from other monocot species. This primer pair was tested on a range of genotypes from a BAC library, we identified GI pseudogene fragments associated with helitron-type sequences. Similar sequences were found to be also present in the F. pratensis genome. Additionally, we describe the use of a methylation-filtered L. perenne genomic library in an initial survey to ascertain the potential frequency of helitrons within the L. perenne genome.

Table 1: PCR primer sequences, 5'...3'1

| Primer Pair | Forward Sequence | Reverse Sequence |
|-------------|-----------------|-----------------|
| GIG49660.6F | GTCCCGTCTATGTAGCTGTA | GIG49660.7R | CCAGTTTCATCAGCTGTCG |
| GiGt.1F | ATTCCTGCATCTGAAACCAC | GiGt.1R | CAGCCACCATACGGATC |
| GiGt.2F | GCAATCAATGGGAAAGTGAT | GiGt.2R | TGCAACTTGAAGATGGCC |

1Thermal cycling profile for all primer pairs was as follows: 1 minute at 94°C, followed by 10 cycles of 1 min at 94°C, 1 min at 60°C (with the temperature reduced by 1°C per cycle), 1 min at 72°C, followed by 30 cycles of 1 min at 94°C, 1 min at 50°C, 1 min at 72°C.

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band consisted of an apparent GI pseudogene fragment. PCR screening of an *L. perenne* BAC library (5 genome equivalents) with a second primer pair, GIGgt2F/2R, designed directly upon derived *L. perenne* genomic sequence, estimated between 4 and 5 GIGgt2F/2R priming sites per genome (see Additional File 1 for derivation of this estimate). Four GIGgt2F/2R-positive BAC clones were isolated from the library; one contained GIGANTEA (*LpGI*) and 3 contained apparently non-allelic GI pseudogene fragments (*Lp-psGI1–3*). Primer pair GIGgt2F/2R was also screened on the 2.5 genome equivalent *F. pratensis* BAC library and, again, an estimate of 4–5 priming sites per genome was obtained (see Additional File 1). However, the PCR products amplified from the *F. pratensis* BAC library were of two distinct types, one type in the expected range and the other type smaller than expected. This latter type was subsequently confirmed by sequencing to be a truncated version of the GI pseudogene.

Both BAC libraries were also screened with the GI specific primer pair GIGgt1F/1R and the assay results estimated 1–2 copies per genome for the *L. perenne* library and 1 copy per genome for the *F. pratensis* library (see Additional File 1). All the BAC library DNA screening pools identified by primer pair GIGgt1F/1R in both libraries were also identified by primer pair GIGgt2F/2R, indicating that both *Lp/Fp* and *Lp-psGI* sequences were amplified by the latter primer pair.

**LpGI sequence analysis**

The region of one of the BACs containing the *LpGI* gene (identified by the GIGgt1F/1R screen) was sequenced directly and the genomic region containing *LpGI* identified. The gene structure was predicted with FGENESH+, using an existing *L. perenne* GI protein sequence (ABF83898) as template and spanned 6024 bp from initiator to terminator codons. Fourteen exons coded for a protein of 1148aa which showed 99% homology with the *L. perenne* GI protein sequence (ABF83898) and 92%, 91% 88% and 66% with homologous GI sequences from barley (AAW66946), wheat (AAO11738), rice (BAF04134) and *Arabidopsis* (ABP96502), respectively (Additional File 2). *LpGI* was mapped to chromosome 3 of *L. perenne* mapping family to a position compatible with the known syntenic relationship between *L. perenne* chromosome 3 and rice chromosome 1 (King et al., 2007; J. King, unpublished data).

**Helitron-like sequences in Lp-psGI.1–3**

Between c. 8 and 11 kb of the 3 BACs containing the different *Lp-psGI* fragments (*Lp-psGI1–3*) were sequenced directly from the BAC. Alignment of these sequences identified regions of partial homology between *Lp-psGI1* and *Lp-psGI2* of c. 6 kb and between *Lp-psGI1* and *Lp-psGI3* of c. 5.6 kb. Insertions of c. 0.8 kb and 0.2 kb interrupted the homologous regions in *Lp-psGI1* and *Lp-psGI2*, respectively. The 3’ end of the homologous regions were terminated in all the *Lp-psGI* sequences by conserved regions containing a 14 bp motif (16 bp in *Lp-psGI1* and *Lp-psGI3*) capable of forming a hairpin structure – characteristic of the 3’ termini of helitron-like transposons (Fig. 1).

BLAST comparisons of the *Lp-psGI* sequences against the *L. perenne* GeneThresher* (LpGT) library identified 10 individual *LpGT* sequences with homology to *Lp-psGI1* both at the 5’ and 3’ ends, with the homology interrupted by a 7501 fragment inserted into a potential helitron 5’-A↓T-3’ target motif (Fig. 2). The borders of the 7501 bp insert consisted of a 5’-TC and 3’ 16 bp conserved hairpin and CTAG motifs, consistent with known helitron structures (Fig. 1 and 2). No evidence of a potential DNA replication-initiator-helicase protein coding sequence was identified within the 7501 bp fragment, indicating that it was likely to represent a non-autonomous helitron. No LpGT sequences could be identified which spanned potential intact helitrons in *Lp-psGI2* or *Lp-psGI3*, indicating that the 5’ regions of these putative helitrons may have been displaced. However, 2 different *LpGT* sequences were identified with homology beginning immediately beyond the conserved CTAG 3’ helitron terminus of *Lp-psGI2*. In both these *LpGT* fragments the homologous regions began at a potential 5’-A↓T-3’ helitron insertion site (Fig. 3). Three further *LpGT* sequences were identified with partial homology to the same internal region of *Lp-psGI2*. In each of these fragments, the homology ended at potential 5’-A↓T-3’ helitron insertion sites (Fig. 3). This may represent the border of a smaller ancestral helitron, which subsequently expanded in the 5’ direction.

**Gene fragments within the Lp-psGI helitron sequences**

Within all the *Lp-psGI* sequences, the *Lp-psGI*-like fragment consisted of a continuous region of c. 0.9 kb from 35 bases 5’ of the ATG initiation codon to 91 bases into the fifth exon (Fig. 4). Clustal alignments of the 3*Lp-psGI* sequences with *LpGI* over the c.0.9 kb conserved region indicated different degrees of sequence conservation in exonic and intron-derived regions. Excluding base insertions and deletions, *LpGI* showed 83–86% sequence conservation with the *Lp-psGI* sequences within the exonic regions but this dropped to 72–73% within the intronic regions. Within the 3 *Lp-psGI* sequences the ranges of sequence conservation within ‘exonic’ and ‘intrinsic’ regions were 94–98% and 95–97%, respectively (Table 2).

Additional gene fragments were identified 5’ of the GI conserved region. A ribosomal protein S7 fragment was present approximately 1 kb upstream of GI in all of the *Lp-psGI* sequences while a succinate dehydrogenase (SDH) fragment was found close to the 5’ end of the helitron in *Lp-psGI1* alone. Both of these fragments contained exon
and intron sequences. A 0.8 kb insert specific to \textit{Lp}-psGl.2 was found to contain a fragment of a non-LTR retroelement, including a partial reverse transcriptase reading frame, which most likely results from a retrotransposition event unrelated to helitron activity (e.g. TBLASTX match with AF474071.1, barley clone) (Fig. 1).

**Comparison of psGl sequences from \textit{L. perenne} and \textit{F. pratensis}**

Three different psGl-type sequences (\textit{Fp}-psGl.1–3) were cloned from the \textit{F. pratensis} BAC library on the basis of identification with primer pair GIGtg2F/2R. Comparison of these with the \textit{Lp}-psGl sequences showed that one, \textit{Fp}-
Diagrammatic representation of region of *GIGANTEA* (*GI*) that has been ancestrally incorporated into a helitron. Black horizontal bar = *L. perenne* genomic sequence spanning the complete *GI* coding sequences; predicted exons are indicated by the thick bar. Grey horizontal bar indicates putative complete helitron sequence from *Lp-psGl.1*; relative position of the *GI* fragment incorporated into the helitron is indicated by the thick grey bar. Sequence detail shows 3' border of conserved *GI* region with putative helitron A-T insertion site at the border.

Figure 3
Diagrammatic representation and sequence details of alignments between *LpGT* sequences and *Lp-psGl.2*, indicating possible ancestral 5' (left) and 3' (right) helitron borders. Diagram: sequence within helitron borders conserved (thick black bar) and not conserved (thin black bar) between *Lp-psGl.1* and *Lp-psGl.2*; non-helitron genomic sequence (thin grey bar). *LpGT* sequences homologous (thick grey bar) and non-homologous (thick white bar) with *Lp-psGl.2*. Sequence details: alignments between *Lp-psGl.2* and *LpGT* sequences showing potential A-T helitron insertion sites; these indicate possible ancestral 3' and 5' borders for different helitron insertion events and also mark the borders of *Lp-psGl.1* and *Lp-psGl.2* homology. *LpGT* sequences: a) FLPB0022289H22-b0FSP_20020409, b) FLPB002413G09-b0FSP_20011203, c) FLPB002264M19-g0RSP_20011109, d) FLPB002078109-b0FSP_20010827, e) FLPB002029F17-g1RSP_20010827 (see Additional File 5).
psGI.1, represented a helitron remnant sequence which was highly similar to the Lp-psGI sequences, indicating a likely similar origin (the 6686 bp putative helitron region of Fp-psGI.1 showed 90% homology with Lp-psGI.1). Fp-psGI.1 contained a similar 3' terminus to the Lp-psGI sequences and the same SDH fragment near its 5' terminus (Fig. 1). However, Fp-GI.2 and .3 were noticeably different. Fp-psGI.2; they contained a GI fragment slightly longer than that found in the Lp-psGI sequences, extending more or less continuously from 231 bp 5' of the ATG initiation codon to 16 bp before the end of the 5th exon, with subsequent partial homology up to the beginning of the 6th exon (Additional File 3). The GI fragment in Fp-psGI.3 was similar to that in Fp-GI.2, except that it contained a 447 bp deletion covering the 3rd and 4th exons of the GI fragment. This truncated GI fragment corresponded to the smaller PCR product obtained in some of the DNA pools from the F. pratensis BAC library screened with GIGgt2f/2r. In total, Fp-psGI.2 and .3 shared sequence homology, interrupted by two major deletions in Fp-GI.3, over c. 5.1 kb region of Fp-GI.2 but showed no apparent homology with either Fp-psGI.1 or the Lp-psGI sequences outside of the GI region.

The conservations of exon- and intron-derived sequences within the GI fragment in Fp-GI.2 in comparison to GI were 92% and 78%, respectively, indicating slightly greater conservation of exon and intron sequence than was observed for the Lp-psGI sequences (83–86% and 72–73%); Table 1. The equivalent figures for Fp-psGI.2 in relation to the Lp-psGI sequences were 79–81% for exons and 71–73% for introns.

**Identification of additional conserved hairpin motif-like sequences in the LpGT library**

SEEDTOP searches of the LpGT library identified 98 out of 16384 patterns with > 10 LpGT sequence alignments. Examination of these identified 7 possible helitron hairpin types (Fig. 5, Additional File 4). The most common type, represented 51 times in the LpGT library (using the criterion of clearly non-homologous sequences, at least 40 bp of sequence present both 5' and 3' of the hairpin motif and no N scores), was the 5'-GTGCAGGGGAGG-3' ‘Type 1’ motif present in the Lp-psGI sequences. In addition to the 16 bp hairpin and the CTAG↓T terminal motifs, the 11 bases 5' of the hairpin and the 8 bases between the hairpin and the CTAG↓T were also strongly conserved. There was no apparent homology between any of the 51 sequences 3' of the CTAG↓T and only limited homology 5' of the hairpin which was probably due to the AT-rich nature of this sequence. Between the different hairpin types, the length of the hairpin sequence varied from between predominantly 16 bp (types 1, 4–7) to predominantly 20 or 21 bp (types 2 and 3 respectively) with 1 to 4, but usually 2 non-complementary bases separating the 7–9 mer complementary sequence stretches. The hairpin was separated from the CTAG↓T motif by 7 to 9 bases for all hairpin types identified.

**Discussion**

The discovery of the helitron families of transposons in plant species over the last few years has largely been a consequence of the availability of comprehensive genome sequence for the models rice and *Arabidopsis*, and latterly for maize. The significance of this has been demonstrated by recent analyses in maize, which have shown the potential of helitron transposition for generating haplotypic diversity and disrupting gene function [4,8,10-12]. There are still few reports of helitron-like transposons in the Pooidae grasses, a sub-family that includes the Triticeae cereals and the Poaceae forage and amenity grasses, probably as a consequence of a necessary focus on transcrptional sequencing within these medium and large genome species. Consequently, the extent to which helitrons are present in, or may have had a role in modelling these genomes is at the moment unknown (though information for *B. distachyon* another Pooidae grass, should soon become available). Therefore, the identification of a putatively complete, non-autonomous helitron sequence as well as a number of partial helitron-like sequences in the species *L. perenne* and *F. pratensis* is important in con-
ferring that helitron activity may have played a significant role in genome modelling within the Pooidae.

The complete non-autonomous helitron sequence, *Lp-psGL1*, is not dissimilar to helitron-type transposons from other plant species, in that it has the expected 5'-TC and 3' hairpin and CTRR terminal motifs as well as showing apparent transposition into an AT target sequence (Figs. 1 and 2). Additionally, again as with similar helitron sequences, there is evidence that gene fragments have been captured within the helitron, in the present case fragments from a succinate dehydrogenase gene, a ribosomal protein gene and a fragment derived from the gene *GI* (Fig. 1). The partial helitron sequences *Lp-psGL2*, *Lp-psGL3* and *Fp-psGI* show a highly similar internal structure to *Lp-psGL1* towards the 3' end and so were, presumably, derived by transposition of the same ancestral helitron before the divergence of the *Lolium* and *Festuca* genomes; for *Lp-psGL2* and *Lp-psGL3*, the fact that there is little homology between the 3 sequences beyond the 3' and 5' termini would indicate that they represent separate transposition events, as opposed to haplotypic variants. Whether *Lp-psGL2/3* and *Fp-psGI* represent partial sequences of complete helitrons or the complete sequences of helitron remnants has not yet been established.

There is no clear relationship between the helitron associated *GI* sequences (*Lp-psGL1/2/3* and *Fp-psGI*) and the two independent fragments (*Fp-psGL2* and *Fp-psGL3*). The latter are relatively more closely related to the intact *LpGI* gene, with the helitron fragments being significantly diverged both from *LpGI* and the available *Triticaceae* sequences. *GI* is a single copy gene in rice and only a single *GI* copy exists.
in the current Brachypodium genome draft, but two divergent and unlinked GI loci have recently been described in maize [34]. The ryegrass and fescue GI fragments may therefore be remnants of similar ancestral duplications in the temperate grass genomes, whose intact descendants have been lost. It is surprising, however, that two apparently different GI lineages should both have become extinct leaving similar sized fragments preserved simultaneously in at least one genome (fescue), particularly if helitron activity was responsible for one fragmentation but not the other.

We considered whether capture by a helitron may have accelerated the divergence of the Lp-psGL.1/2/3 and Fp-psGL.1 lineage from a Fp-psGL.2/3 fragment progenitor but this seems unlikely for at least two reasons. Firstly, comparing the divergence between the helitron GI sequences indicates that they have acquired a relatively large number of indels since their origin from a common ancestor, but that the number of point mutations is not remarkable (there are 7 indel differences between Lp-psGL.1 and Fp-psGL.1, for example, and only 2.7% sequence variation despite separation of the two host species by ~2.8 myr, compared with 6 indels and 13.5% sequence variation between the same region of LpGI and the gene from barley, whose last common ancestor was ~35 myr [35]). Secondly, divergence from LpGI is significantly higher in the intron sequences of the helitron GI fragments than in their exons, consistent with the expected selection for GI protein function. However, this contrasts dramatically with the large proportion of non-synonymous mutations, particularly generating frameshifts and stop codons, within the exons, indicating strong selection against this function. This suggests that the progenitor of the helitron GI sequences did indeed evolve gradually as an intact and functional GI gene, giving rise to a lineage distinct from LpGI and Fp-psGI.2/3 but that at some stage its coding function became severely deleterious. This may have occurred before capture by the helitron or relatively soon after, as most inactivating mutations are shared by the elements described here.

The closer relationship between Fp-psGL.2/3 and LpGI suggests that the independent GI fragments may derive from a more recent duplication which also suffered a subsequent extinction under selective pressure. Consistent with this, there is less divergence between Fp-psGL.2 and Fp-psGL.3 than between any two of the helitron GI fragments, while there is still a high level of non-synonymous differences from the LpGI and Triticeae GI sequences. An interesting question is whether the pre-existing helitron fragments could in some way have been responsible for the coincident fragment size of Fp-psGL.2 and Fp-psGL.3 or whether there is some inherent reason for GI to be disrupted in this way. In order to address this, we are currently investigating whether intact or recently fragmented GI genes related to either of the two extinct Lolium/Festuca lineages still exist in related species.

The observation that the common ancestral helitron from which Lp-psGI.1–.3 and Fp-psGI.1 were derived had captured GI and other gene fragments is of interest from two angles. Firstly, although these sequences are only fragments, replication and transposition following their capture has increased their copy number. Whether this had any direct consequence in terms of the perception and response to photoperiod is unknown, but the observation of apparently independent extinction of a subsequent GI duplication does suggest that the helitron capture and/or fragmentation may be beneficial to the host genome in helping to eliminate expression of unnecessary or deleterious duplicated genes, possibly in response to new selective pressures. A further question remains as to the positions of the Lp-psGI sequences within the L. perenne genome relative to each other and to GI itself, which maps to chromosome 3. To resolve this, attempts were made to identify allelic polymorphism across the 3’ and 5’ borders of the Lp-psGI.1–3 sequences in the mapping family, but amplified PCR products showed no sequence variation (data not shown) and, so, the Lp-GI sequences could not be assigned a genetic position.

The process(es) by which helitrons capture foreign sequences has yet to be clarified and either ‘read-through’ errors at the 3’ terminus or a mechanism based upon non-homologous repair of double-stranded DNA breaks have been suggested [1,2]. Comparison of the Lp-psGI and GI sequences identified here provides some suggestion that the original capture of the GI fragment may have occurred by helitron expansion at the 5’ end, a possibility referred to by [4]. Alignment of the Lp-psGI fragments with the equivalent GI gene sequence shows that the 3’ border terminates with a potential A↓T helitron insertion site (Fig 4). It is therefore possible that helitron insertion originally occurred within this site in GI and upon subsequent transposition there was ‘slippage’ of the 5’ helitron border resulting in incorporation of a fragment of GI. A similar mechanism is a possibility for the incorporation into Lp-psGL.2 of a sequence homologous to LpGT fragments a, b, and c, as illustrated in Fig 3.

There remains the major question as to how ubiquitous helitrons are in the L. perenne and other Pooidaeae genomes – a question that will only be definitively answered by the accumulation of contiguous genomic sequence for these species. However, the LpGT library does represent a collection of hypomethylated, presumed gene-rich [36,37] though relatively short (mean = 502 bp) genomic sequences. This size-range limitation means that they are unlikely to contain complete helitrons, but could
contain recognisable helitron 3’-border motifs. Searches of the LpGT library for short sequence stretches containing potential hairpins and the CTRR 3’ helitron border motif identified 7 sets of sequences (Fig. 5 and Additional File 4). If these do represent true 3’ helitron borders, this indicates that helitron activity in *L. perenne* may have been relatively widespread in recent evolutionary history, as evidenced by the presence of these sequences in presumed hypomethylated regions of the genome (i.e., their representation in the LpGT library) and by the sequence conservation across the hairpin types identified. The SEEDTOP search identified 172 non-homologous sequences containing potential 3’ helitron termini. However, it should be born in mind that this is very limited survey of the containing potential 3’ helitron termini. However, it should be born in mind that this is very limited survey of the

interestesting to see the extent to which helitron activity may have been responsible for modifying and diversifying these grass and cereal genomes.

**Conclusion**

An apparently complete non-autonomous helitron and a related series of incomplete helitron sequences have been identified in the Pooidae grasses *Lolium perenne* and *Festuca pratensis*. The identified helitrons had captured a number of gene fragments, including a fragment of the key flowering gene GIGANTEA. Searches of a *L. perenne* GeneThresher® DNA sequence library identified a number of possible 3’ helitron borders in unrelated sequences. This represents evidence for a possible ancestral role for helitrons in modelling the genomes of *Lolium* and related species.

**Methods**

**Genomic libraries**

The *L. perenne* (c. 5 × genome coverage) and *F. pratensis* BAC libraries (c. 2.5× genome coverage) have been described previously [38,39]. Derivation of copy number estimates from PCR screening of the BAC libraries is described in Additional File 1. The *L. perenne* GeneThresher® (LpGT) DNA sequence library database was obtained on license from ViaLactia Biosciences, Auckland, New Zealand and was described previously [40,41].

**Identification of *L. perenne* GIGANTEA and BAC sequencing**

Primer pair GIG49660.6F (GTCCCGTCTATGATGCGTGA), GIG49660.7R (CCAGTCTCATCAGTCTCGG) was designed on the basis of conserved sequences in exons 2 and 4 of the rice GI gene (LOC_Os01g08700) and wheat and barley ESTs (GenBank: BJ245948 and BI481891, respectively) and the identity of the PCR product confirmed by sequencing. This primer pair was then used to PCR screen the *L. perenne* BAC library to identify clones containing GI and GI-like sequences (Pseudo-GIGANTEA; Lp-psGI) which were sequenced directly from the BACs. Subsequently, both the *L. perenne* BAC library and the *F. pratensis* BAC libraries were screened with further primer sets based directly upon the derived *L. perenne* BAC sequences: primer pair GIGgt.2F (GCATCAATTGGGAAGTGAGGAT), GIGgt.2R (TGCAACTTTGAAAGATTGGCC), anchored in the first and fifth exons of GI and which amplified c. 800 bp PCR products from both GI and psGI containing BACS and primer pair GIGgt.1F (ATTCCTGCACTCTGAAACCAC), GIGgt.1R (CAGCCAGCACATACGAGT), which amplified c. 600 bp fragment from the 10th exon of GI and identified just GI containing BACs. Thermal cycling profile for all primer pairs was as follows: 1 minute at 94°C, followed by 10 cycles of 1 min at 94°C, 1 min at 60°C (with the temperature reduced by 1°C per cycle), 1 min at 72°C, followed by 30 cycles of 1 min at 94°C, 1 min at 50°C, 1 min at 72°C.

**Genetic mapping**

The F2 *L. perenne* mapping population (n = 187) and framework map has been described previously [42]. GI was mapped as a segregating CAPS marker detected as a Tat1 (Fermentas, York, UK) restriction enzyme polymorphism in a PCR product amplified from the the 10th exon of the GI gene using primer pair GIGgt.1F/1R. The marker was placed on the existing genetic map using Joinmap v. 3.0 [43].

**DNA sequence alignments**

GI and psGI sequences derived from *L. perenne* and *F. pratensis* were aligned with other plant sequences in GenBank and with the local LpGT library database using BLASTN. Further alignments and manual adjustments were performed using ClustalW [44] and Macaw version 2.0.5 [45]. Exon and intron sequence similarities between GI and the psGI fragments inserted in the Lp/Fp-psGI sequences (Table 2) were calculated after ClustalW alignment and manual adjustment both directly on the complete sequence alignments and after exclusion of base insertions and deletions (i.e., reflecting base substitutions).

Potential helitron 3’ hairpin and CTRR motifs were identified by searching the LpGT library with SEEDTOP (part of the stand alone BLAST executables package [46]) for sequences of the form N^superscript 1N^subscript 2N^superscript 3N^subscript 4N^subscript 5N^superscript 6N^subscript 0[7x(0,5)]N^superscript 7N^superscript 8N^subscript 9N^subscript N^subscript(Nx(0,12):CT) [GA] [GAT], where N^superscript is a defined base and N^subscript is its complement, x(n1, n2) is a number (n) of undefined bases between n1 and n2 (inclu-
sive) and [GA] is either G or A. N1–N7 consisted of all possible nucleotide 7 mers, giving 16384 search patterns. Where > 10 different LpGT sequences were identified by an individual search pattern, the LpGT database was additionally searched with the reverse complement of the search pattern and the sequences were examined for possible helitron 3’ motifs using Macaw sequence alignments. Identical or near identical LpGT sequences with different identifiers were only included once in the analysis. Possible helitron motifs were identified on the basis of sequence conservation across potential hairpin and CTRR motifs with low sequence homology 5’ and 3’ of these motifs. For illustration, c. 110 bp of sequence flanking the putative helitron motifs were aligned using ClustalW with manual adjustment in GenDoc (Figure 5, Additional File 4).

*LpGI* and all cited *Lp-,* *Fp-psGI* and LpGT sequences cited are given in Additional File 5 along with their EMBL accession numbers

**List of Abbreviations**

LpGT: *L. perenne* GeneThresher® genomic library; LpGI: *L. perenne* GIGANTEA; FpGI: *F. pratensis* GIGANTEA; Lp-psGI: *L. perenne* genomic sequence containing GIGANTEA pseudogene fragment; Fp-psGI: *F. pratensis* genomic sequence containing GIGANTEA pseudogene fragment

**Authors’ contributions**

IA and TL designed the study and analysed the data, all authors contributed to the execution of the study, IA, TL and KF contributed to the drafting of the manuscript and all authors read and approved the final version.

**Additional material**

**Additional File 1**

PCR-screening of the *L. perenne* and *F. pratensis* BAC libraries and derived copy number estimates. details the methods used and assumptions made in deriving sequence copy number estimates from PCR screening of the BAC libraries. References included.

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**Additional File 2**

Alignments of predicted protein sequences for GIGANTEA. Figure illustrating the alignments of GIGANTEA protein sequences from *L. perenne, wheat, barley, rice* and *Arabidopsis.*

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**Additional File 3**

Alignments of partial *Lp* and *Fp-psGI* illustrating regions of sequence conservation with *LpGI* genomic and coding sequence. Figure illustrating the regions of sequence conservation between *LpGI* genomic sequence and CDS and the GI fragments contained within the *Lp* and *Fp-psGI* sequences.

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**Additional File 4**

Type 1 ~ 7 putative 3' helitron sequence motifs identified in the *L. perenne* GeneThresher® library. Figure illustrating all of the putative 3’ helitron sequence motifs identified in the *L. perenne* GeneThresher® library by the SEEDTOP search, including the sequences not illustrated in Figure 5 (main text).

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**Additional file 5**

*L. perenne* and *F. pratensis* GI, psGI and GeneThresher® sequences. FASTA formatted *L. perenne* GI and *L. perenne* and *F. pratensis* ps-GI and *L. perenne* GeneThresher® library sequences referred to in the paper. Each sequence is accompanied by an EMBL accession numbers in brackets. GeneThresher® library sequences are also described with their original library reference number.

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