Dab2 attenuates brain injury in APP/PS1 mice via targeting transforming growth factor-beta/SMAD signaling

Lei Song¹, Yue Gu¹, Jing Jie², Xiaoxue Bai², Ying Yang², Chaoying Liu², Qun Liu¹

¹ Department of Neurology, Norman Bethune First Hospital of Jilin University, Changchun, Jilin Province, China
² Department of Respiratory Medicine, Norman Bethune First Hospital of Jilin University, Changchun, Jilin Province, China

Abstract

Transforming growth factor-beta (TGF-β) type II receptor (TβRII) levels are extremely low in the brain tissue of patients with Alzheimer’s disease. This receptor inhibits TGF-β1/SMAD signaling and thereby aggravates amyloid-beta deposition and neuronal injury. Dab2, a specific adapter protein, protects TβRII from degradation and ensures the effective conduction of TGF-β1/SMAD signaling. In this study, we used an adenoviral vector to overexpress the Dab2 gene in the mouse hippocampus and investigated the regulatory effect of Dab2 protein on TGF-β1/SMAD signaling in a mouse model of Alzheimer’s disease, and the potential neuroprotective effect. The results showed that the TβRII level was lower in APP/PS1 mouse hippocampus than in normal mouse hippocampus. After Dab2 expression, hippocampal TβRII and p-SMAD2/3 levels were significantly increased, while amyloid-beta deposition, microglia activation, tumor necrosis factor-α and interleulin-6 levels and neuronal loss were significantly attenuated in APP/PS1 mouse brain tissue. These results suggest that Dab2 can exhibit neuroprotective effects in Alzheimer’s disease by regulating TGF-β1/SMAD signaling.

Key Words: nerve regeneration; transforming growth factor-β1; Dab2; Alzheimer’s disease; amyloid-beta; neuron; SMAD2; SMAD3; microglia; neural regeneration

Introduction

Alzheimer’s disease is not only the most common neurodegenerative disease, but also the most common cause of dementia, and its prevalence increases with age, affecting approximately 4.4% of the population aged ≥ 65 years[1-4]. Owing to a lack of understanding of Alzheimer’s disease pathogenesis and treatment, there is no effective method to reverse or prevent disease progression[5]. Therefore, recognizing amyloid-beta protein generation and the potential molecular regulatory mechanism underlying the produced neurotoxicity is an important step toward developing novel drugs for Alzheimer’s disease[6].

The transforming growth factor-β1 superfamily mainly consists of transforming growth factor-β1, -β2 and -β3 subtypes[7-8]. Transforming growth factor-β exhibits a variety of biological effects, and participates widely in diverse pathophysiological processes, including cell apoptosis and proliferation, stem cell differentiation and embryonic development, extracellular matrix formation, wound repair and inflammatory reactions[9-12]. Transforming growth factor-β1 is the most important subtype in the central nervous system, being mainly expressed in neurons and microglia, and its level can be increased by stress after nerve injury. Accumulating evidence has demonstrated that transforming growth factor-β1, as a pleiotropic cytokine, exhibits important neuroprotective effects. In invertebrates, transforming growth factor-β1 can regulate synaptic growth and differentiation, neurotransmitter release and synaptic protein distribution and phosphorylation[13].

The Tgfb1+/− transgenic mouse exhibits a transforming growth factor-β1 mRNA level that is only 50% of the normal level and, compared with normal mice, nerve cells in Tgfb1−/− transgenic mice show an increased susceptibility to age and excitotoxins[14]. Tgfb1−/− transgenic mice show a wide range of degenerative diseases of the nervous system, accompanied by decreased levels of laminin and synaptophysin and microglial proliferation[14]. Transforming growth factor-β1 exhibits a synergic effect together with many neurotrophic factors, including nerve growth factor, brain-derived nerve growth, and neurotrophin-3, 4, maintains the growth of nerve cells, and protects them from injury[15].

Transforming growth factor-β1 is closely related to Alzheimer’s disease and amyloid-beta. The level of transforming growth factor-β1 in the brain and cerebrospinal fluid is increased, but its level in the serum is lower in Alzheimer’s disease patients than in unaffected people[16-17]. Wyss-Coray et al.[18-19] reported that, in APP transgenic mice, transforming growth factor-β1 overexpression in microglia can
Attenuate amyloid-beta deposition in the hippocampus; moreover, transforming growth factor-β1 can also promote amyloid-beta phagocytosis and degradation in vitro. In addition to its effects on amyloid-beta metabolism, transforming growth factor-β1 can protect neurons from amyloid-beta-induced neurotoxicity\[20\]. Because transforming growth factor-β1/SMADs are inhibited in multiple sites, the neuroprotective effect of transforming growth factor-β1 on Alzheimer’s disease cannot function normally\[21\]. In Alzheimer’s disease patients, the TβRII level is decreased and is related to the severity of Alzheimer’s disease. Age-related neuronal degeneration and hippocampal amyloid-beta deposition appear in TβRII knockout mice\[22\]. Lee et al.\[23\] found that SMAD2 could not be transported into nucleus after phosphorylation, resulting in abnormal intracellular aggregation of phosphorylated SMAD2 (phospho-SMAD2, p-SMAD2) molecules; this abnormal aggregation of p-SMAD2 is related to neurofibrillary tangles and granulovacuolar degeneration. Therefore, ensuring normal conduction of transforming growth factor-β1/SMAD signaling has become a problem to be solved in the treatment of Alzheimer’s disease using transforming growth factor-β1/SMAD\[24-25\].

TβRII expression is regulated by many factors, including microRNAs, ubiquitination, and membrane transport. In a previous study by Wang et al.\[26\], in APP/PS1 transgenic mice, hippocampal TβRII expression was found to be regulated by microRNA-106b; specifically, microRNA-106b aggravated amyloid-beta-induced neurotoxicity by down-regulating the TβRII level. Dab2, a specific adapter protein, can protect TβRII from degradation in vivo\[27\]. A previous study has reported that Dab2 expression is downregulated in Alzheimer’s disease mice\[28\]. Therefore, it is presumed that loss of Dab2 can lead to downregulation of TβRII expression, block the normal conduction of transforming growth factor-β1/SMAD signaling in Alzheimer’s disease, and thereby accelerate the progression of Alzheimer’s disease. To test this hypothesis, we used an adenoval viral vector to overexpress Dab2 in the APP/PS1 transgenic mouse model of Alzheimer’s disease, and investigated the regulatory effect of Dab2 protein on transforming growth factor-β1/SMAD signaling and brain injury in Alzheimer’s disease.

Results

Quantitative analysis of experimental animals

Ten BL6/C57 mice were used as negative controls receiving no treatments. Twenty-four APP/PS1 transgenic mice were randomly and evenly divided into Alzheimer’s disease, vector and Dab2 groups. The rats in these groups were administered physiological saline, adenovirus-associated virus (AAV)9-RFP plasmids and Dab2 plasmids, respectively, via AAV injection. (A) Mice in each group received hippocampal injection of AAV9 plasmid at 3 months of age, underwent behavioral tests at 9 months and were sacrificed at 10 months for molecular biological and pathological tests. (B) By fluorescence microscopy, AAV9 plasmids can be seen to be expressed in the mouse hippocampus, showing red fluorescence. Scale bar: 100 μm. (C) The hippocampal Dab2 level in the Alzheimer’s disease group was significantly lower than that in the control group, and the hippocampal Dab2 level in the Dab2 group was significantly higher than that in the vector group (\(P < 0.01\)). One-way analysis of variance and the SNK-q test were used to assess the significance of differences between groups. Data are represented as mean ± SEM from at least three independent experiments. The Dab2 level is expressed as the ratio of the target protein absorbance value to that for GAPDH.
bilateral hippocampi. In each group, mice at the age of 9 months underwent behavioral tests. All mice were sacrificed at 10 months of age and molecular biological and immunological tests were performed as shown in Figure 1A. No mice died throughout the entire experiment.

**Dab2 plasmid injection increased hippocampal Dab2 protein expression**

To test whether the Dab2 plasmid is expressed in mouse hippocampus, we observed hippocampal slices under a fluorescence microscope. Expression of the Dab2 plasmid carrying RFP was observed in hippocampal slices (Figure 1B). Western blot analysis showed that the hippocampal Dab2 level was significantly lower in the Alzheimer’s disease group than in the control group. After hippocampal injection of AAV9-RFP plasmid (vector group), the hippocampal Dab2 level was similar to that in the Alzheimer’s disease group. The hippocampal Dab2 level was significantly higher in the Dab2 group, into which the AAV9-CMV-Dab2 plasmid was injected, than in the vector group ($P < 0.01$; Figure 1C). These results indicate that AAV can be used as a safe expression vector to overexpress Dab2 gene in the central nervous system.

**Dab2 regulated mouse hippocampal transforming growth factor-β1/SMAD signal transduction**

To investigate the regulatory effect of Dab2 on hippocampal transforming growth factor-β1/SMAD signal transduction, we detected hippocampal TβRII and phosphorylated SMAD2/3 levels by western blotting. The results showed that the hippocampal TβRII level was significantly lower, while the pSMAD2/3 levels were significantly increased in the Alzheimer’s disease group compared with the control group. These findings are consistent with those reported by Tesser et al.\textsuperscript{[22]} (Figure 2A, B). TβRII and pSMAD2/3 levels in the Dab2 group were significantly higher than in the vector group (Figure 2).

**Dab2 overexpression improved the abnormal behaviors of Alzheimer’s disease mice**

Morris water maze test results showed that the escape latency and swimming distance of mice in the Dab2 group were significantly shorter than those in the vector group ($P = 0.039, 0.039$; Figure 3A, B). These results demonstrate that learning ability in the Dab2 group was significantly better than that in the vector group. After removal of the platform, the percentage of time that mice spent in quadrant A was significantly greater in the Dab2 group than in the vector group.
Dab2 overexpression decreased the amyloid-beta level in brain tissue

ELISA results showed that the levels of soluble and insoluble amyloid-beta 40 and 42 in mouse brain tissue were not significantly different between the Alzheimer’s disease group and the vector group (P > 0.05). Soluble amyloid-beta 40 and amyloid-beta 42 levels were significantly lower in the Dab2 group than in the vector group (P < 0.05). One-way analysis of variance and the SNK-q test were used to assess the significance of differences between groups. Data are represented as mean ± SEM from at least three independent experiments.

Dab2 reduced microglial activation and inflammatory factor expression in the hippocampi of Alzheimer’s disease mice

Through immunofluorescence staining, Iba1-positive microglia with irregular morphology and many neurites were observed in the hippocampi of mice in the Dab2, vector and Alzheimer’s disease groups (Figure 5A). Under 100-fold magnification, the number of Iba1-positive microglia was significantly lower in the Dab2 group than in the vector group (P = 0.038) and Alzheimer’s disease group (P = 0.019). There was no significant difference in numbers of Iba1-positive cells between the vector group and the Alzheimer’s disease group (P = 0.904; Figure 5B).

To investigate the effect of Dab2 on inflammatory reactions in the brain tissue of Alzheimer’s disease mice, we detected hippocampal tumor necrosis factor-α and interleukin-6 levels by ELISA. The results showed that hippocampal tumor necrosis factor-α and interleukin-6 levels were significantly lower in the Dab2 group than in the vector group (P = 0.029, 0.040), and there were no significant differences in tumor necrosis factor-α and interleukin-6 levels between the vector group and the Alzheimer’s disease group (P = 0.886, 0.883; Figure 5D). These results suggest that Dab2...
overexpression can reduce hippocampal microglia activation and inflammatory factor release in Alzheimer’s disease mice, thereby providing anti-inflammatory effects.

**Dab2 inhibited hippocampal neuronal loss**

To investigate the effects of Dab2 on hippocampal neurons in Alzheimer’s disease mice, we detected hippocampal neuron-specific enolase-positive cells by immunofluorescence staining. The results showed that, in the normal mouse hippocampal tissue, neurons were arranged in an orderly, dense fashion and the cytoplasm was stained green. Compared with normal mice, different degrees of neuronal loss appeared in the hippocampus tissue of mice in the Alzheimer’s disease, vector and Dab2 groups (Figure 6A). Under 400-fold magnification, significantly more neuron-specific enolase-positive cells were seen in the Dab2 group than in the vector group ($P = 0.035$), and there were no significant differences between the Alzheimer’s disease and vector groups ($P = 0.842$; Figure 6B). These results suggest that Dab2 overexpression can attenuate pathogenic factor-induced mouse hippocampal neuronal loss.

**Discussion**

AAV is a defective, non-pathogenic human parvovirus with a length of approximately 4.7 kb, and, after recombination, the AAV vector retains only 4% of the original viral genes [29-31]. Compared with other viral vectors, such as adenovirus and lentivirus, AAV vector is highly safe, has a wide range of host cells, and can be used to express exogenous genes for a long time period *in vivo* [32-33]. AAV9 is an important AAV vector subtype. Numerous studies have documented that, *via* direct injection, the genes carried by AAV9 vectors are highly expressed in hippocampal nerve cells and, therefore, the vector can be used for Alzheimer’s disease studies [34]. The confocal laser scanning microscopy results in this study showed that, 3 months after AAV9 plasmid injection, RFP expression was still very high in the mouse hippocampus, indicating that AAV9 vectors express exogenous genes in brain tissue for a long period. However, 7 months after AAV9 plasmid injection, Dab2 protein level was higher in the Dab2 group than in the Alzheimer’s disease and vector groups, as shown by western blot analysis. The Dab2 gene carried by AAV9 plasmid can be expressed for a long period of time, which is required by many experiments.

Transforming growth factor-β1 binds to the cell membrane surface receptor TβRII, leading to a conformational change in TβRII, and then the complex binds to a conserved domain enriched in serine and glycine in the TβRI cytoplasmic region, namely, the GS domain, to form a TβRII-TG-
Fβ-TβRII complex. Then, TβRII autophosphorylates and is subsequently activated; thus, TβRI can specifically identify SMAD2/3 protein, phosphorylates it (to form p-SMAD2/3) and then mediates an intracellular signaling cascade. A previous study reported that the TβRII level is extremely low in the hippocampus of Alzheimer’s disease patients, which hinders the effective conduction of transforming growth factor-β/SMAD signaling, and, after Tβ RII knockout, a large amount of amyloid-beta deposition is seen in the mouse hippocampus, which accelerates neuronal aging. The results of the present study showed that T RII expression in the brain tissue of APP/PS1 mice was significantly lower than that in wild-type mice, consistent with the results of a previous report. Our results also showed that, although TβRII expression is decreased, the p-SMAD2/3 level in brain tissue was significantly higher in APP/PS1 mice than in wild-type mice. Combined with previous findings, these findings suggest that, although the p-SMAD2/3 level is increased, the expression of genes involved in intranuclear regulation is blocked; therefore, transforming growth factor-β1/SMAD signal transduction in the brains of APP/PS1 mice is inhibited.

Because TβRII is a key factor in transforming growth factor-β1/SMAD signal transduction and the progression of Alzheimer’s disease, understanding the regulation of TβRII expression in an Alzheimer’s disease model can help clarify the pathological mechanism of Alzheimer’s disease and aid the development of drugs for treating Alzheimer’s disease in the clinic. The TβRII post-transcriptional level is affected by various factors, and TβRII membrane transport is an important one. Under the physiological or pathological state, TβRII receptors on the cell membrane surface are included in intracellular early endosome antigen 1 (EEA1(+)) by endocytosis, and some intracellular TβRII receptors are recycled to the membrane surface. The remainder are directly degraded in cells, thereby regulating the TβRII level and transforming growth factor-β1/SMAD signaling transduction. Dab2, a member of the Dab protein family, mainly regulates the endocytosis of cell surface proteins and is closely related to cellular adhesion and proliferation as well as actin formation. A previous study has shown that Dab2 expression is decreased in Alzheimer’s disease mice. Our results also confirm that Dab2 expression is downregulated in the brain tissue of APP/PS1 mice. It is presumed that this decrease in Dab2 expression likely causes a TβRII circulatory disorder and subsequent degradation, leading to a decrease in TβRII level on the cellular membrane surface and blocking of transforming growth factor-β1/SMAD signaling transduction. Our results showed that Dab2 overexpression in hippocampal tissue leads to increased TβRII and p-SMAD2/3 levels, indicating that Dab2 can indeed regulate transforming growth factor-β1/SMAD signal transduction in a mouse model of Alzheimer’s disease, exhibiting a neuroprotective effect.

To investigate the neuroprotective effect of Dab2, we further detected its regulatory effect on amyloid-beta levels and neurotoxicity. Amyloid-beta is an initial factor and a key link in the pathological mechanism of Alzheimer’s disease. Amyloid-beta overexpression and abnormal accumulation in brain tissue can destroy intracellular calcium homeostasis, lead to excessive oxygen free radical formation, activate inflammatory factors, cause inflammation of focal brain tissue, lead to synapse reduction and neuronal loss, and finally result in an abnormal integrative function of the central nervous system. There is strong evidence that transforming growth factor-β1 signaling shows an inhibitory effect on either amyloid-beta generation or neurotoxicity.
The results from this study showed that, when Dab2 protein was overexpressed in the hippocampal tissue of Alzheimer’s disease mice, both soluble and insoluble amyloid-beta levels are reduced, indicating that Dab2 can reduce amyloid-beta generation. Amyloid-beta can induce neuronal apoptosis by abnormally activating the cell cycle, and is closely related to late-state degradation of β-catenin and excessive Tau phosphorylation. After binding to TβRII, transforming growth factor-β1 activates components of the SMAD and non-SMAD pathways, and thereby inhibits the abnormal activation of the cell cycle. In this study, immunofluorescence analysis revealed a significant reduction in the number of cells expressing neuron-specific enolase in the hippocampi of Alzheimer’s disease mice, while the numbers of neurons overexpressing Dab2 were not significantly reduced. Based on this and previous findings, we consider that Dab2 can regulate transforming growth factor-β1/SMAD signaling transduction by increasing the numbers of cell membrane surface TβRII receptors, and thereby attenuates amyloid-beta generation and neutrotic effects on nerve cells. In this study, we also performed a Morris water maze test, which is a means for studying the mechanism underlying cerebral learning and memory abilities and is widely used in the study of Alzheimer’s disease. The results from this study showed that, compared with the vector group, escape latency and swimming distance were significantly shortened, and the time spent within the quadrant with the platform removed was significantly prolonged, in the Dab2 group, suggesting that Dab2 overexpression can improve the memory ability and spatial orientation ability of Alzheimer’s disease mice, verifying the neuroprotective effect of Dab2.

More attention has been paid to the role of inflammatory reactions in the occurrence of Alzheimer’s disease. Increasing evidence has demonstrated that a chronic inflammatory reaction consistently exists in the brain tissue of Alzheimer’s disease patients and is possibly a factor involved in inducing the formation and development of other pathological characteristics. Transforming growth factor-β1, as an inflammatory regulator, can inhibit inflammatory reactions of the central nervous system and microglial activation. In various neurodegenerative diseases, inhibition of transforming growth factor-β1/SMAD signaling is related to T cells and microglia-mediated local inflammation. Results from this study showed that the numbers of activated microglia and tumor necrosis factor-α and interleukin-6 levels in the brain tissue of Alzheimer’s disease mice were significantly greater than in common mice. After Dab2 overexpression in the brain tissue of Alzheimer’s disease mice, the numbers of cells expressing neuron-specific enolase in the hippocampi of Alzheimer’s disease mice were not significantly reduced.

Materials and Methods

Design
A randomized, controlled molecular biology trial.

Time and setting
This study was performed in the Laboratory of Immunology, Centre for Translational Medicine, First Hospital of Jilin University Bethune Medical Center, China from May 2012 to February 2013.

Materials
Animals
Twenty-four 2-month-old male APP/PS1 double transgenic mice and ten negative control 2-month-old BL6/C57 mice (5 male and 5 female) were purchased from the Animal Research Institute, Nanjing University, China. All animals were housed separately in a 21 ± 2°C and 30–35% relative humidity environment with artificial 12-hour illumination. There was no difference in exposure factors between groups. The experimental protocol was approved by the Animal Ethics Committee of Jilin University, China. After raising for 1 month, animals were used for further experiments.

Plasmids
AAV9-Dab2 plasmids and negative control AAV9-RFP plasmids were synthesized by Jikai Gene Chemical Technology Co., Ltd. (Shanghai, China).

Methods
Hippocampal injection of plasmids
Plasmids were injected into the mouse hippocampus according to a previously described method. After anesthesia by intraperitoneal injection of ketamine (100 mg/kg) and xylazine (20 mg/kg), 3-month-old mice were fixed in a small animal stereotaxic instrument (Ruiwode Life Science Co., Ltd., Shenzhen, China) and the skin was disinfected with 75% alcohol. Physiological saline or AAV plasmid (1 × 10^7 viral particles (VP)) was injected into the hippocampus via a needle inserted at a depth of 1.8 mm (Hamilton Co, Reno, NV, USA), 2.1 mm posterior to and 1.8 mm left and right lateral to the bregma, 2 μL each side. After injection, the needle was slowly...
withdrawn, the drill hole was sealed and the skin was sutured; then, mice were housed separately in a constant temperature environment and given free access to water and food.

**Behavioral tests**

Six months after AAV plasmid injection (i.e., at the age of 9 months), behavioral tests were performed using a Morris water maze test according to a previously described method [57]. Briefly, a cylindrical tank with a diameter of 100 cm and a height of 40 cm contained 23 ± 1°C water to a depth of 25 cm. Four points were marked on the tank wall to divide the tank into four equal quadrants (A, B, C, D). A small platform was placed in the center of quadrant A and hidden 1 cm deep away from the water surface. Initially, mice were directly placed on the platform for 30 seconds to feel the spatial location of the platform. When testing, a random quadrant was designed as the starting area and mice were placed in a water tank, on the top of which, a camera was installed to record the swim path. Then, the platform was taken out and mice were re-placed in the tank for automatic re-recording of the data by a computer. The maximum recording time was 200 seconds. Mouse spatial learning ability and memory ability in each group were measured within 4 days. On day 4, escape latency and escape distance were recorded. The percentage of mice staying in quadrant A after removing the platform in the whole swim path was calculated.

**Detection of amyloid-beta, tumor necrosis factor-α and interleukin-6 levels in brain tissue by ELISA**

According to Donkin’s method [58], amyloid-beta, tumor necrosis factor-α and interleukin-6 levels in brain tissue were detected by ELISA. Briefly, 500 μL of brain tissue, eight volumes of ice-cod bicarbonate buffer (100 mmol/L Na₂CO₃, 50 mmol/L NaCl pH 11.5) and protease inhibitor were added into a homogenizer for homogenization for 20 seconds. Then, the mixture was centrifuged at 12,000 × g for 45 minutes. Amyloid-beta, tumor necrosis factor-α and interleukin-6 levels in the supernatant were determined according to the instructions provided in the amyloid-beta (Invitrogen, Camarillo, CA, USA), tumor necrosis factor-α and interleukin-6 kits (Abcam, Hong Kong, China). The remaining precipitate was re-suspended with 50 mmol/L Tris-HCl (pH 8.0) containing 5 mol/L guanidine hydrochloride (pH 8.0). Insoluble amyloid-beta level was measured by kit instruction (Invitrogen).

**Detection of hippocampal Dab2, TBR2II and p-SMAD2/3 levels by western blot analysis**

Hippocampal Dab2, TBR2II and p-SMAD2/3 protein levels were detected by western blot analysis [59]. A small amount of hippocampal tissue was chopped into small pieces, homogenized, and lysed on ice with lysis buffer here (Beyotime, Shanghai, China) for 30 minutes. The lysate was centrifuged at 12,000 × g for 20 minutes and then the supernatant was taken to determine Dab2, TBR2II and p-SMAD2/3 levels. An aliquot (15 μg) of each sample was thoroughly mixed with 4 μL of 5 × loading buffer and denatured in boiling water for 5 minutes. Protein samples were subjected to 10% SDS-PAGE (Beyotime) at 80 V for 40 minutes and at 110 V for 90 minutes, and then transferred onto PVDF membranes at 200 mA for 60 minutes and enveloped with blocking solution for 2 hours. After washes with western cleaning solution (Beyotime), protein samples were incubated with goat anti-IgG polyclonal antibody (1: 200; Santa Cruz Biotechnology, Santa Cruz, CA, USA) at 4°C overnight. The next day, after washes, sample was incubated with horseradish peroxidase-labeled rabbit anti-goat polyclonal antibody at room temperature for 2 hours. Thereafter, the bands were developed by enhanced chemiluminescence. The absorbance of the scanned bands was determined using Image J (National Institutes of Health, Rockville, MD, USA). The results are expressed as the ratio of the target protein absorbance value to that for GAPDH.

**Determination of the numbers of microglia and neurons by immunofluorescence staining**

According to a method described previously [60], mouse hippocampal slices were stained for Iba1 and neuron-specific enolase. After washes with 0.01 mol/L PBS, brain tissue was fixed with methanol containing 0.3% hydrogen peroxide for 20 minutes, blocked with 10% bovine serum albumin (Gibco, Grand Island, NY, USA), washed with 0.01 mol/L PBS, treated with rabbit anti-mouse polyclonal antibody IgG (which was diluted with 0.03% Triton-X-100-containing PBS at 1:200; Santa Cruz Biotechnology) overnight at 4°C. The next day, after washes with 0.01 mol/L PBS, brain tissue was treated with 3% bovine serum albumin-diluted (1:200) goat anti-rabbit monoclonal antibody IgG-FITC (Santa Cruz Biotechnology) and incubated for 1 hour at room temperature. After three washes with 0.1 mol/L PBS for 5 minutes each time, brain tissue was mounted with anti-quenching fluorescence mounting medium. Finally, brain tissue was observed under the fluorescence microscope (Olympus, Japan) and photographed. Five fields of view were taken from each of six consecutive slices and Iba1- and neuron-specific enolase-positive cells were counted. The mean value was taken as the final result.

**Statistical analysis**

All data were statistically processed using SPSS 18.0 software (SPSS, Chicago, IL, USA) and measurement data are expressed as mean ± SEM. One-way analysis of variance and the SNK-q test were used for intergroup comparisons. A level of α = 0.05 was considered statistically significant.

**Author contributions**: Song L and Liu Q were responsible for experimental design, evaluation, data analysis and manuscript writing. Gu Y, Jie J, Bai XX, Yang Y and Liu CY participated in experiment conduction. All authors approved the final version of this paper.

**Conflicts of interest**: None declared.

**Peer review**: This study was the first to report that Dab2 can regulate transforming growth factor-β1/SMADs signal pathway in an Alzheimer’s disease model and exhibit a neuroprotective effect through this pathway.

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