Anosmia-hyposmia and dysgeusia in COVID-19 may be due to SARS-CoV-2 protein mimicry of olfactory receptors

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Abstract
Background: Anosmia-hyposmia and dysgeusia are common symptoms of mild-to-moderate COVID-19 cases. They are usually, but not always, reversible and the cause is unknown.

Methodology: Proteonomical similarity searching (BLAST) was used to test whether SARS-CoV-2 has an unusual degree of similarity to human olfactory receptors as compared with other viruses.

Results: SARS-CoV-2 displays two orders of magnitude more similarities to human olfactory receptors than does any other upper respiratory virus or rhinovirus and five times as many similarities as cold-causing coronaviruses.

Conclusions: It is hypothesized that IgA antibodies produced against SARS-CoV-2 proteins and secreted in saliva and mucus bind to and block human olfactory receptors. These antibodies cause the reversible form of anosmia-hyposmia associated with COVID-19 infection. The mimicry may also, rarely, lead to autoimmune disease against neurons expressing olfactory receptors, in which case oral or nasal steroids may be therapeutic.

Key words: SARS-CoV-2, COVID-19, olfactory, smell, taste, molecular mimicry, IgA, autoimmunity, autoimmune disease, immunity, cross-reactivity

Introduction
Loss of smell and taste (anosmia or hyposmia) and/or taste (dysgeusia) are now recognized to be significant, often presenting, clinical features of mild-to-moderate SARS-CoV-2 infection affecting, on average, more than 50% of patients (range 5 to 98% in different studies) with a preponderance in women (ca. 70%) and individuals under the age of 50 (e.g., 1,2). Various hypotheses have been proffered to explain the loss of olfactory receptor function including direct infection of olfactory neurons; infection of sustentacular (neuronal support) cells resulting in loss of olfactory receptor-containing cilia; and local inflammation of the olfactory epithelium (reviewed in 3) but no definitive explanation yet exists. Evidence is presented here that SARS-CoV-2 proteins exhibit multiple sequences that mimic highly conserved extracellular loop and binding regions of human olfactory proteins. It is hypothesized that these SARS-CoV-2 antigens induce immunoglobulin type A (IgA), that is released into saliva, mucus and tears to control the infection but which also bind to and block olfactory receptors.

Materials and methods
BLASTP (version 2.2.31+) on the www.expasy.org website was used to search for possible similarities between the entire sequences of SARS-CoV-2 proteins and human olfactory receptors with the following settings: BLOSUM80 was used to identify short sequences such as those presented by human leukocyte antigens and recognized by T cell receptors; gaps disallowed; low complexity regions screened out; E set to 1000 with 3000 best matches displayed. All SARS-CoV-2 proteins were tested along with those of coronavirus HKU1, SARS-CoV-1, Middle
Anosmia/dysgeusia due to olfactory receptor mimicry of SARS-CoV-2

Table 1. Summary of BLAST results comparing virus proteomes with human olfactory receptors. Unique sequences are similarities that are distinct in the region of the olfactory receptor that any given virus protein matches. In most cases, a given virus protein matched more than one olfactory receptor, which is indicated in the number of “total matches”.

| Viral genome                            | Unique sequences | Total matches |
|-----------------------------------------|------------------|---------------|
| Coxsackievirus A9 l6VPA8                | 0                | 0             |
| Coxsackievirus B3 P03313                | 0                | 0             |
| Echovirus E6 K4M118                     | 0                | 0             |
| Enterovirus E A0A193AUM2                | 0                | 0             |
| Hepatitis A Virus P06441                | 1                | 5             |
| HIV-1 group M subtype B ^               | 2                | 7             |
| Influenza H1N1 Wilson ^                 | 3                | 4             |
| Poliovirus Type 1 P03300                | 0                | 0             |
| Rhinovirus 1A P23008                    | 1                | 2             |
| Rhinovirus 2 P04936                     | 0                | 0             |
| Rhinovirus C3 A0A5Q0QNI0                 | 1               | 2             |
| Rhinovirus 14 P03303                    | 0                | 0             |
| Rhinovirus 16 Q82122                    | 1                | 2             |
| **AVERAGE**                             | **0.7**         | **1.7**       |
| Coronavirus HKU1 (Isolate N1) ^         | 14               | 48            |
| MERS (isolate United Kingdom/           | 15              | 54            |
| H1239900006/2012) +                     |                  |               |
| Human SARS coronavirus (SARS-CoV) ^     | 21              | 233           |
| Human SARS-CoV-2                        | 23              | 203           |
| Replicase 1a P0DTC1                     | 1                | 2             |
| Spike Protein P0DTC2                    | 1                | 1             |
| Protein 3a P0DTC3                       | 0                | 0             |
| Small Envelope Protein P0DTC4           | 4                | 14            |
| Membrane Protein P0DTC5                 | 4                | 10            |
| Non-Structural Protein P0DTC6           | 2                | 3             |
| Protein 7a P0DTC7                       | 9                | 168           |
| Protein 8 P0DTC8                        | 0                | 0             |
| Nucleoprotein P0DTC9                    | 0                | 0             |
| Replicase 1ab P0DTD1 ^                  | 1                | 2             |
| Protein 9b P0DTD2                       | 1                | 3             |
| Protein N514 P0DTD3                     | 0                | 0             |
| Protein 7b P0DTD8                       | 0                | 0             |

UniProt accession numbers are provided for each virus proteome utilized in the searches: ^ P03470, P05777, P03427, P03430, P05777, P03454, Q82506, P15659, P15682. ^ P06X2, QSMQD0, QSMQD1, QSMQC7, QSMQC6, QSMQ8, QSMQ5, QSMQ9; ~ P59635, P59632, P59594, P59637, P59595, P59636, P59634, P59596, P59633. Q809H3, Q7TF0A1, Q7TLC7, Q7TFA0; ~ K9N7C7, K9N5Q8, K9N7A1, K9N643, K9N4V0, K9N3R3, K9N4V7, K9N7D2, K9N796; ^ P04585, K7X064, P69726, A0EV89, P05919, O43719, P04618, P69723 & P0DTC1 overlaps P0DTC1 so only additional matches are listed in this row.

Results

SARS-CoV-2 proteins exhibited almost two orders of magnitude greater numbers of similarities to human olfactory proteins than did any other virus tested except for the coronavirus HKU1 (a cold virus), SARS-CoV-1 and MERS. Controls were mainly other respiratory viruses including influenza H1N1 (which was typical of other influenza viruses, not shown), coxsackieviruses (only two shown), and rhinoviruses four of which are presented. Most SARS-CoV-2 similarities were associated with the small envelope protein (P0DTC4), membrane protein (P0DTC5), and protein 7a (P0DTC7), which mimicked extracellular and transmembrane binding regions of more than two hundred different human olfactory receptors. The same pattern was duplicated in SARS-CoV-1 and MERS, which were also characterized by anosmia/hyposmia. This highly conserved mimicry would assure broad loss of olfaction. It is unlikely that cytoplasmic regions of similarity (Figure 1) are accessible to antibodies though they might become so during autoimmune disease.

The vast majority of the sequence similarities between SARS-CoV-2 proteins and olfactory receptors (and all of those shown in Figure 1) satisfy the criterion that at least six of the amino acids in a sequence of ten are identical (where a pair of acceptable substitutions counts as one identity), making them excellent candidates for antigenic cross-reactivity as determined previously by experiment.

Discussion

Infection with SARS-CoV-2 elicits early, high titers of immunoglobulin type A (IgA) antibodies that rapidly decrease after the virus is eliminated with a titer half-life of about ten days. These IgA antibodies are the primary means of virus neutralization in mucosa. I propose the novel hypothesis that IgA cross-react with olfactory receptors blocking receptor function without killing the associated neurons. The better the immune response, the more complete the anosmia but, correspondingly, the better-controlled the infection. This hypothesis explains why gustatory and olfactory impairment are associated with mild to moderate COVID-19 symptoms and why most patients recover smell and taste within four weeks as the virus is cleared from the body and IgA antibody levels decrease. The hypothesis is testable by examining the saliva of anosmic, hyposmic or dygeusitic COVID-19 patients for IgA antibodies that bind to...
olfactory receptors. This hypothesis might also explain transient hyposmia and anosmia associated with colds associated with other coronaviruses and rhinoviruses.

A corollary to the hypothesis could explain why some COVID-19 patients (as many as 25%) develop chronic anosmia or hyposmia that does not resolve even after three or more months \(^\text{11}\). Such patients may develop an autoimmune disease against their olfactory neurons triggered by the molecular mimicry illustrated in Figure 1 between SARS-CoV-2 proteins and olfactory receptors. While molecular mimicry is often associated with induction of autoimmune diseases, however, it is generally considered to be insufficient to trigger disease \(^\text{11}\). Current theory suggests that molecular mimicry requires an associated bystander or complementary infection that prevents T-cell tolerization needed to protect “self” cells \(^\text{11}\). Patients experiencing chronic smell and taste loss may therefore differ from those with transient symptoms in having evidence of a viral or bacterial co-infection along with SARS-CoV-2. Such patients should be tested for complement-activating autoantibodies with specificity to olfactory receptors and loss of T-cell tolerance to olfactory neurons.

Nasal or oral steroids might be useful in treating the autoimmune form of the disease after a patient has repeatedly tested negative for SARS-CoV-2 but are contraindicated during active infection and in cases of transient smell and taste loss because of interference with active immunity against SARS-CoV-2 \(^\text{11}\). Indeed, for most patients, a strong IgA response resulting in anosmia-hyposmia appears to be a predictor of mild COVID-19 symptoms \(^\text{1, 2, 6, 7}\).

**Conclusions**

In sum, it is demonstrated that SARS-CoV-2 shares significant similarities with a huge number of human olfactory receptors and it is proposed that IgA antibody produced in response to the viral infection then blocks these receptors. This explains...
both why the anosmia/hyposmia and dysgeusia are transient and associated with mild cases of COVID-19. The hypothesis also proposes that chronic (>3 month) loss of smell and taste may be due to an autoimmune disease against olfactory neurons triggered by the SARS-CoV-2-olfactory receptor mimicry. The hypothesis may also apply to other coronaviruses that share many olfactory receptor similarities that are not present in most rhinoviruses, influenza viruses and other upper respiratory viruses. Means of distinguishing between the transient and autoimmune forms of anosmia/hyposmia are suggested as well as differences in treatment.

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