Peri-Implant Crevicular Fluid Analysis, Enzymes and Biomarkers: a Systematic Review

Erhan Dursun¹, Tolga Fikret Tözüm²

¹Department of Periodontology, Faculty of Dentistry, Hacettepe University, Ankara, Turkey.
²Department of Periodontics, College of Dentistry, University of Illinois at Chicago, Chicago, Illinois, USA.

Corresponding Author:
Tolga Fikret Tözüm
Department of Periodontics, College of Dentistry
University of Illinois at Chicago
801 S Paulina Street, Room 469G, Chicago, IL 60612
USA
Phone: +1-312-996-0265
E-mail: ttozum@uic.edu

ABSTRACT

Objectives: To review the current understanding of the biomarkers and enzymes associated with different forms peri-implant diseases and how their level changes influence the pathogenesis of the inflammatory diseases around dental implants.

Material and Methods: An electronic search in two different databases was performed including MEDLINE (PubMed) and EMBASE between 1996 to 2016. Human studies analyse peri-implant crevicular fluid (PICF) biomarker and enzyme levels of implants having peri-implant mucositis and peri-implantitis published in English language, were evaluated. A systematic review was performed to assess which biomarkers and enzymes in PICF were used to identify the inflammatory conditions around dental implants.

Results: Fifty-one articles were identified of which 41 were further evaluated and included in the analysis. Due to significant heterogeneity between included studies, a meta-analysis could not be performed. Instead, a systematic descriptive review was performed.

Conclusions: Biomarkers and enzymes in peri-implant crevicular fluid have shown promising results in differentiating from peri-implant disease condition to health. However, due to inconsistent results and acquiring much evidence from cross-sectional studies, additional evidence supported by randomized-controlled trials is needed to validate the links reported.

Keywords: biomarkers; dental implants; diagnosis; enzymes; inflammation.
INTRODUCTION

Dental implant treatment is a successful, widespread and predictable treatment for tooth loss over the past 20 years however, an increasing number of implant failures caused by peri-implant diseases still take part in every day clinical dental practice [1]. Two forms of peri-implant inflammation have been identified in the literature: peri-implant mucositis and peri-implantitis. The American Academy of Periodontology (AAP) [1] stated that from a clinical standpoint, signs that determine the presence of peri-implant mucositis include bleeding on probing (BOP) and/or suppuration, which are usually associated with probing depths (PDs) ≥ 4 mm and no evidence of radiographic loss of bone beyond bone remodelling. Peri-implantitis is a progressive, irreversible disease of the bone and soft tissues around osseointegrated dental implants under masticatory function that is accompanied by bone resorption, reduced osseointegration, deep pocket formation and suppuration [2]. Despite divergences in the definition of peri-implantitis and the differential diagnosis of peri-implant diseases, studies have estimated that peri-implantitis affects approximately 10% of implants and 20% of patients [3]. According to a recent systematic review peri-implant mucositis and peri-implantitis have a prevalence ranging from 19 to 65% and from 1 to 47%, respectively. On the other hand, another systematic review reported mean prevalence for peri-implant mucositis and peri-implantitis as 43% and 22%, respectively [4].

The combination of clinical and radiographic parameters, such as PD, BOP, suppuration, mobility and marginal bone loss, are the commonly used parameters for the diagnosis of peri-implantitis [5]. However, these diagnostic processes might not be sensitive or specific enough to distinguish disease onset, development, and activity. Clinical measurements around implants as like natural teeth might be challenged by the force and direction of probing, implant geometry, prosthesis design and peri-implant soft tissue biotype. In addition, both peri-implant mucositis and peri-implantitis lesions can present with BOP and/or suppuration, with PDs greater than 4 mm. Therefore, clinicians and researchers may often observe the early, and sometimes the late diagnosis of peri-implantitis.

Early detection of peri-implant destruction, as well as monitoring progression of bone loss is extremely important. Currently, blunt surrogate markers are being used such as radiographs and peri-implant probing. These tests have obvious limitations as only history of disease may be detected. As main markers of peri-implantitis are bone destruction and inflammation, biomarkers and enzymes in implant sulcus fluid (PISF) focusing on these disease entities are of interest. Active components and mechanisms involved in the destructive process may thus be important perspectives within this field. Such knowledge may potentially lead to new diagnostic strategies and candidate disease markers for peri-implant conditions.

A biomarker is an indicator of a biological state and can help to distinguish between normal and pathologic processes [6]. Presently, radiographs and clinical parameters such as, PD, clinical attachment level and BOP generally used for peri-implant condition diagnosis. Research to look at associations between certain biomarkers with health and/or disease can give more tools to clinicians for better understanding the pathogenesis of such peri-implant diseases [6]. One of the main advantages of evaluating biomarkers is the repeatability and non-invasive nature of obtaining samples for analysis. Biomarkers can be measured in secretions such as saliva and gingival crevicular fluid, or in the case of implants, peri-implant crevicular fluid (PICF). An early pilot study demonstrated that this method of sampling provides reliable outcomes [7]. Since then, studies have been conducted to look at a vast array of biomarkers and enzymes around dental implants as an early sign of peri-implantitis.

Today; there is large variation for the threshold of diagnosis for peri-implantitis, which may explain the wide range of percentages reported for its prevalence. Researchers and clinicians are always looking for adjunctive measures to aid in proper disease diagnosis, and the measurement of levels of enzymes and biomarkers is possible tool, and has gathered a lot of interest. Therefore, the purpose of this article was to review the current understanding of the biomarkers and enzymes associated with peri-implant diseases and how their level changes took part in the pathogenesis of the disease.

MATERIAL AND METHODS

Protocol and registration

The methods of the analysis and inclusion criteria were specified in advance and documented in a protocol. The review was registered in PROSPERO, an international prospective register of systematic reviews. The protocol can be accessed at:

http://www.ejomr.org/JOMR/archives/2016/3/e9/v7n3e9ht.htm
The reporting of this systematic analysis adhered to the Preferred Reporting Items for Systematic Review and Meta-Analyses (PRISMA) Statement [8].

Focus questions

Which biomarkers and enzymes in PICF are used for distinguish between healthy implants and implants having peri-implant diseases? Do patients with peri-implant diseases (peri-implant mucositis or peri-implantitis) present higher levels of biomarkers and enzymes in PICF?

Types of publications

The review included studies on humans published in the English language. Letters, editorials, case reports, literature reviews, animal research, PhD theses, and abstracts were excluded.

Information sources

The search strategy incorporated the examination of electronic databases, supplemented by hand searches. A search was conducted on the National Library of Medicine database (MEDLINE) through its online site (PubMed) and EMBASE databases. Additionally, a hand search was conducted in the following journals: “Implant Dentistry”, “Clinical Oral Implants Research”; “Clinical Implant Dentistry and Related Research”, “European Journal of Oral Implantology”, “International Journal of Oral & Maxillofacial Implants”, “Journal of Oral Implantology”, “International Journal of Oral and Maxillofacial Surgery”, “International Journal of Oral and Maxillofacial Surgery”, “Journal of Periodontology”, “Journal of Clinical Periodontology”, “International Journal of Periodontics and Restorative Dentistry”, “Journal of Prosthetic Dentistry”, “International Journal of Endodontics, Journal of Endodontics”, “Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology”, “Endodontology” and “Turkish Journal of Medical Sciences”.

Search

The PubMed and EMBASE resource databases were explored through advanced searches. An organized and logical approach was used to categorize the studies dealing with the association between PICF biomarkers and peri-implant diseases. The keywords and search inquiries used during the primary stage were as follows: “peri implant crevicular fluid” OR “peri-implant crevicular fluid” AND “peri implant sulcus fluid” AND “peri-implant sulcus fluid” AND “peri-implantitis” OR “peri implantitis” AND “peri-implant inflammation” OR “peri implant inflammation” AND “peri-implant infection” OR “peri-implant infection” AND “peri-implant mucositis” OR “peri implant mucositis” AND “implant biomarkers” AND “implant enzymes”. The choice of keywords was intended to be broad to collect as much relevant data as possible without relying on electronic means alone to refine the search results. After advance search; the studies dealing with peri-implantitis or peri-implant mucositis and PICF biomarker or enzyme level analysis were included in the present review.

Selection of studies

Based on the inclusion criteria, the authors independently screened titles and abstracts derived from the literature search (Figure 1). Both reviewers compared decisions and their eligibility for this review was confirmed after discussion. Full articles were obtained for all the studies considered eligible for inclusion in this paper and further evaluated by both reviewers. If needed, a third party was consulted when consensus could not be reached.

Types of publications

The present review included only human studies published in the English language. Letters, editorials, case reports, literature reviews, animal research, PhD theses, and abstracts were excluded.

Types of studies

The present review included all human prospective, follow-up studies, clinical trials, cohort studies, case-control studies, case series studies, published between January 1, 1996 and March 1, 2016, were searched that reported on biomarkers and enzyme levels obtained by PICF and/or PISF analysis.

Types of participants/population

Individuals included in the studies should have had at least one osseointegrated screw-type dental implant that presented with clinical or radiologic signs of peri-implant mucositis or peri-implantitis and subjected to PICF biomarker analysis. However, studies presented with different definitions for the peri-implant diseases included.
Inclusion and exclusion criteria

For this systematic review, original cross-sectional and longitudinal prospective clinical studies with collection of different biomarkers and enzymes in PICF from individuals with peri-implantitis (P) or peri-implant mucositis (M) were selected. Letters, editorials, case reports, literature reviews, animal research, PhD theses, and abstracts were excluded. Only reports in English were included. Other exclusion criteria were as follows: 1) studies with quantification of biomarkers and enzymes in tissue biopsies, serum, saliva and other biologic sources; 2) assessment of only fluid volume but not biomarker and enzyme levels; 3) fluid collection and analyses for determining the effect of different implant designs not inflammation; 4) fluid collection during early osseointegration; 5) focus on gingival distances; 6) unreported implant inflammation criteria; 7) not clear information about patient/implant groups whether healthy or peri-implantitis; 8) studies assessed different biomarker genotypes.

Assessment of methodological quality

The quality of all selected trials was assessed using the recommended approach for assessing risk of bias in studies included in Cochrane reviews [9]. Evaluated parameters are: (1) random sequence generation, (2) allocation concealment, (3) blinding of participants and personnel, (4) blinding of outcome assessment, (5) incomplete outcome data, (6) selective reporting, and (7) other bias. The potential risk of bias was categorized as “low”, “unclear” or “high” depending on the quality and detailed explanation of provided information about all abovementioned parameters. All assessments were completed by a single examiner (ED).
Data analyses

Significant heterogeneity between publications in terms of diseases definitions, assessed parameters, study designs, as well as measured outcomes, among others, prevented the quantitative synthesis of the included studies and consequently a meta-analysis could not be completed. Instead, a qualitative descriptive analysis of the reported outcomes was performed and systematically reviewed in forms of tables.

RESULTS

Study selection and search results

Two reviewers based on the inclusion criteria independently screened titles derived from this comprehensive search. The reviewers compared decisions and resolved differences through discussion, consulting a third party when consensus could not be reached. The third party was an experienced senior reviewer. Full reports were obtained for all the studies judged eligible for inclusion in this paper. At the title and abstract stage, one reviewer accepted the citations that appeared to meet the inclusion criteria and sent them on for full-text review, with a second reviewer assessing only those citations the first reviewer believed ineligible.

The electronic search strategy provided 1318 titles. After title screening and abstract reading, 1267 studies were excluded because they did not fit the inclusion criteria. The participants of all studies selected were in good general health and had not received any medication (e.g. antibiotics, and/or anti-inflammatory and/or immunosuppressive agents) that could affect the peri-implantitis biologic process at the time of PICF sampling.

Study characteristics

The characteristics of the 41 included articles are summarized in Table 1. The most prevalent study design was cross-sectional (n = 32) followed by interventional (n = 5). A great variability in PICF collection and biomarker or enzyme assessment was observed, and many different biomarkers and enzymes have been reported across the studies. The participants of all included studies were in general health and had not received any medication that may affect the peri-implant disease process. A wide range of biomarkers and enzymes used to explore an association between studied marker and peri-implant diseases. After full text reading, the 10 studies were excluded due to following reasons: 1) focus on the comparison of PICF interleukin-1 beta (IL-1β) and plasma tumor necrosis factor-alpha (TNF-α) levels between systemically healthy and diabetic subjects [10], 2) directly compare the biomarkers and enzymes of PICF around implants with gingival crevicular fluid (GCF) around natural teeth [11-15], 3) cytokine evaluation with polymerase chain reaction instead of biomarker levels [16-18], 4) sample composed of only failing implants instead of peri-implant diseases [19].

The majority of the studies used the following criteria to define the peri-implantitis: at least one peri-implant site with PD > 4 mm with clinical signs of inflammation (BOP, suppuration and bone loss). Health implants were generally accepted as having no bone loss and no signs of inflammation. Smokers were excluded from the majority of the studies. Two studies included smokers and match the number of smokers in healthy and peri-implantitis groups [20,21].

Most articles reported the subject numbers as well as the implants evaluated in the studies however, some of the studies reported only the number of patients and some studies reported only the number of implant sites. Of the included biomarkers and enzymes in PICF, IL-1β was the most studied parameter (19 studies) followed by TNF-α (11 studies), IL-6 (11 studies) and IL-8 (6 studies). Five studies reported oxidative stress parameters associated with peri-implant inflammation and 6 studies reported matrix metallo proteinase (MMP) levels. Twenty studies compared the results between P and H sites and most studies evaluated healthy and diseased implants from different patients. Only one study performed in-patient evaluation between H and P sites [22].

Results of individual studies

One interventional study evaluated the levels of IL-6, OPG, osteocalcin (OC), leptin, osteopontin (OPN), parathyroid hormone (PTH), TNF-α, adiponectin and insulin levels after surgical treatment of peri-implantitis [23]. They reported no change according to the levels of OC, OPN, PTH, TNF-α and insulin levels and significant reduction according to total protein, MMP-8, IL-6, OPG, leptin and adiponectin levels after surgical treatment [23]. Another interventional study also reported total amount of TNF-α was significantly reduced at 3 and 12 months after therapy (open flap debridement) compared to baseline associated with improvements in clinical parameters [24]. Another interventional study exposed healthy implants to de novo plaque accumulation and no significant changes observed in the total amount of TNF-α, IL-1β and TGF-β2 compared to baseline in PICF [25].
Table 1. Assessed biomarkers and enzymes in PICF for peri-implant diseases

| Study | Year of publication | Study design | Number of patients (implants) | Assessed PISF biomarker/enzyme Type of assay | Main findings |
|-------|--------------------|-------------|-------------------------------|---------------------------------------------|--------------|
| Panagia et al. [7] | 1996 | CS | 13 (7H, 2M, 26 P) | IL-1β, TNF-α, proc. IL-1β | ELISA |
| Hiltunen et al. [20] | 2002 | CS | 37 (14H, 42 P) | IL-1β | ELISA |
| Wang et al. [22] | 2012 | CS | 68 (32 M, 36 P) | IL-1β, VEGF, MMP-8, TIMP-2, and OPG | ELISA |
| Yaghoubi et al. [22] | 2014 | CS | 8 (116 E, 8 H, 112 P) | IL-1β, TNF-α, IL-6 | ELISA |
| Wohlfart et al. [23] | 2014 | INT | 32 (P) | IL-6, OPG, OC, leptin, OPG, PTI, TNF-α, adenosine, and adenosine | ELISA and Luminex |
| de Mendonça et al. [24] | 2000 | INT | 10P | TNF-α | ELISA |
| Scharnag et al. [25] | 2008 | INT | 23 (21 H) | TNF-α, TGF-β2 (IL-1β) | ELISA |
| Durett et al. [26] | 2009 | INT | 35 (10 H, 10 M, 20 P) | IL-4, IL-10, IL-2, TNF-α, RANKL, OPG | ELISA |
| Luchman et al. [27] | 2007 | INT | 21 (42) | IL-1β and PGE2 | ELISA |
| Baezquez et al. [29] | 2010 | Longitudinal | 28 (72) | PGE2 and MMP-8 | ELISA |
| Ramseier et al. [30] | 2016 | CS (504 implant 493 adjacent teeth) | IL-1β, MMP-8, MMP-9, MMP-11, and MMP-1 bound to tissue inhibitors of MMP (TIMP-1) | ELISA |
| Nomura et al. [31] | 2000 | CS | 6 (10) | MMP-8 | ELISA |
| Ma et al. [32] | 2003 | CS | 13 (49) | Collagenase 2 and 3 | ELISA |
| Ma et al. [33] | 2000 | CS | 12 (46) | Collagenase 2 and 3 | ELISA |
| Casado et al. [35] | 2013 | CS | 30 (10 H, 10 M, 10 P) | IL-1β and IL-10 | ELISA |
| Petrovski et al. [36] | 2010 | INT | 40 (49 H, 30 M, 11 advanced M) | IL-1β, TNF-α, IL-8, and IL-14 | ELISA |
| Ata-Ali et al. [38] | 2013 | CS | 34 (23 H, 8 M) | IL-1β and IL-8 | ELISA |
| Luchman et al. [39] | 2007 | CS | 29 (16 H, 17 P) | IL-1β, PAI-2, and PGE2 | ELISA |
| Milos et al. [40] | 2012 | CS | 31 (13 H, 18 P) | IL-1β and IL-6 | ELISA |
| Ravni et al. [41] | 2011 | CS | 35 (24 H, 12 P) | Flow cytometry | ELISA |
| Yaghubi et al. [42] | 2013 | CS | 32 (41 implant 41 contralateral teeth) | IL-1β | ELISA |
| Fonseca et al. [43] | 2014 | CS | 22 (60 M, 50 P) | IL-1β, IL-2, IL-10, IL-4, TNF-α, IL-12, IL-12, IFN-γ, and OPG | ELISA |
| Gincici et al. [44] | 2012 | CS | 28 (20 H, 7 M) | IL-1β, IL-10, RANKL, and OPG | ELISA |
| Abouyousef et al. [44] | 2012 | CS | 37 (17 H, 10 M) | IL-1β and PGE2 | ELISA |
| Lani et al. [46] | 2010 | CS | 10 (25 H, 7 M) | IL-1β, IL-2, TNF-α | ELISA |
| Ata-Ali et al. [47] | 2015 | CS | 35 (54 H, 24 P) | IL-4, IL-10, IL-12, and IL-19 | ELISA |
| Dasdo et al. [48] | 2013 | CS | 35 (15 H, 20 P) | TNF-α, IL-17 | ELISA |
| Gincici et al. [50] | 2010 | RC | 1111 tooth site, 109 implant sites | MPO | Spectrophotometrically |
| Looman et al. [51] | 2006 | CS | 25 (64) | MPO | Spectrophotometrically |
| Tusaie et al. [52] | 2013 | CS | 23 (67 tooth site, 41 implant sites) | MPO and NO | Spectrophotometrically |
| Plagh et al. [53] | 2012 | CS | 8 (10 P) | IL-1β, TNF-α | ELISA |
| Yamal et al. [54] | 2011 | CS | 596 tooth, 6 implant | Elastase, alpha2-macroglobulin and alkaline phosphatase | Spectrophotometrically |
| Panagia et al. [55] | 2011 | CS | 100 tooth, 6 implant | Elastase, alpha2-macroglobulin and alkaline phosphatase | Spectrophotometrically |
| Arik et al. [56] | 2014 | CS | 12 (18 H, 61 H) | ICP, RANKL, and OPG | ELISA |
| Bakie et al. [56] | 2013 | CS | 70 (23 H, 25 H) | sRANKL, RANKL, and OPG | ELISA |
| Murata et al. [57] | 2002 | CS | 16 (10 P, 16 M, 20 H) | OPG, bone-sourced protein, and IL-1β | ELISA |
| Turner et al. [58] | 2008 | CS | 15 (10 P) | ICTP and OC | Radiommununoassay |
| Bakie et al. [59] | 2014 | CS | 41 (18 P, 45 M, 58 H) | RANKL, soluble RANKL, OPG, cauthepin-B, and cathepin-C | ELISA |
| Severson et al. [60] | 2011 | CS | 14 (20 P, 11 D) | IL-6, IL-10 and IL-17 and the chemokine IL-8 | ELISA |
| Momose et al. [61] | 2006 | CS | 16 (19) | Immunoassay | ELISA |
| Kornfill et al. [62] | 2000 | CS | 20 (39) | A1F | Spectrophotometrically |
| Zhang et al. [63] | 2005 | CS | 56 (24 H, 33 M, 8 P) | IL-6 | ELISA |

CS = cross-sectional; INT = interventional; CC = case-control; RCT = randomized clinical trial; PICF = peri-implant crevicular fluid; PISF = peri-implant sulcus fluid; IL = interleukin; P = peri-implantitis; M = mucositis; H = healthy; TNF = tumour necrosis factor; VEGF = vascular endothelial growth factor; MMP = matrix metallo proteinase; TIMP = tissue inhibitor of MMP; OPG = osteoprotegerin; PTH = parathyroid hormone; OC = osteocalcin; OPG = osteoprotegurin; TGF = transforming growth factor; RANKL = receptor activator of factor kapp a H ligand; PGE = prostaglandin; MIP = macrophage inflammatory protein-1alpha; PA-I = plasminogen activator inhibitor type 2; PDGF = platelet derived growth factor; IFN = interferon; MPO = myeloperoxidase; NO = nitric oxide; ICTP = C-terminal prolyl-hydroxylase crosslinking of Type I collagen; AST = Aspartate aminotransferase; PD = probing depth; GI = gingival index; PI = plaque index; MBI = modified bleeding index; MPI = modified plaque index.
Another study included three groups (H, M, P) of implants and reported levels of TNF-α was significantly higher in P and M, TNF-α levels of diseased implants decreased from baseline to three months after therapies, no differences among groups for IL-4, IL-10, IL-12 and the osteoprotegerin (OPG) and receptor activator of NFκB ligand (RANKL) ratio was higher for healthy implants than for untreated peri-implantitis [26].

Another interventional study reported no difference between peri-implant health and disease condition according to IL-1β and PGE2 levels [27].

Assessment of methodological quality

The results of risk of bias assessment for included studies were summarized in Table 2.

Table 2. Risk of bias within the included studies

| Study                          | Random sequence generation | Allocation concealment | Blinding | Incomplete outcome data | Selective reporting | Other sources of bias |
|-------------------------------|---------------------------|------------------------|----------|-------------------------|---------------------|-----------------------|
| Panagos et al. [7]            | ?                         | ?                      | -        | +                      | +                   | +                     |
| Hultin et al. [20]            | ?                         | -                      | -        | +                      | +                   | +                     |
| Wang et al. [21]              | +                         | +                      | +        | +                      | +                   | +                     |
| Yaghobee et al. [22]          | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Wohlfahrt et al. [23]         | +                         | +                      | ?        | +                      | +                   | +                     |
| de Mendonça et al. [24]       | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Schienero et al. [25]         | +                         | ?                      | +        | +                      | +                   | +                     |
| Duarte et al. [26]            | +                         | +                      | +        | +                      | +                   | +                     |
| Lachmann et al. [27]          | ?                         | +                      | +        | ?                      | +                   | +                     |
| Basegmez et al. [29]          | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Rameisier et al. [30]         | +                         | +                      | ?        | +                      | +                   | +                     |
| Nomura et al. [31]            | ?                         | ?                      | ?        | +                      | +                   | ?                     |
| Ma et al. [32]                | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Ma et al. [33]                | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Casado et al. [35]            | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Petković et al. [36]          | ?                         | ?                      | -        | ?                      | +                   | ?                     |
| Ata-Ali et al. [38]           | ?                         | ?                      | ?        | +                      | ?                   | +                     |
| Lachmann et al. [39]          | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Melo et al. [40]              | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Renvert et al. [41]           | ?                         | +                      | +        | +                      | +                   | +                     |
| Yaghobee et al. [42]          | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Fonseca et al. [43]           | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Günücü et al. [44]           | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Aboyoussef et al. [45]        | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Luo et al. [46]               | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Ata-Ali et al. [47]           | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Darabi et al. [48]            | ?                         | ?                      | -        | ?                      | ?                   | ?                     |
| Günücü et al. [50]            | +                         | ?                      | ?        | +                      | +                   | +                     |
| Liskmann et al. [51]          | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Tözüm et al. [52]             | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Plagnat et al. [53]           | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Yamanalik et al. [54]         | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Arikan et al. [55]            | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Rakic et al. [56]             | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Muratu et al. [57]            | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Tümer et al. [58]             | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Rakic et al. [59]             | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Severino et al. [60]          | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Monov et al. [61]             | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Fiorellini et al. [62]        | ?                         | ?                      | ?        | +                      | +                   | +                     |
| Zhang et al. [63]             | ?                         | ?                      | ?        | +                      | +                   | +                     |

+ = low risk; ? = unclear risk; - = high risk.

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DISCUSSION

The current evidence according to the PICF levels of biomarkers and enzymes that used to distinguish between healthy and inflamed implant sites and their diagnostic and prognostic potential for prediction of future peri-implantitis was assessed and results from 41 original were explored in the present review. It is obvious that a wide range of biomarkers and enzymes are reported to be involved in peri-implant inflammation (Table 1). The PICF levels of 13 different cytokines (IL-1β, IL-2, IL-4, IL-5, IL-6, IL-7, IL-8, IL-10, IL-12, IL-17, IFN-γ, PGE-2 and TNF-α) have been compared in different clinical peri-implant conditions. MMPs are endopeptidases capable of degrading various extracellular matrix proteins and can play a role in cell proliferation, differentiation, migration, and apoptosis [28]. Peri-implantitis has been shown to demonstrate a similar pattern of destruction as periodontitis, and MMP upregulation has been associated with irreversible peri-implant connective tissue destruction [28]. Six studies assessed collagens in PICF in different peri-implantitis lesions which are important around peri-implant tissues, are MMP-1, MMP-3, MMP-8, and MMP-13 and tissue inhibitors [21,29-33]. And according to all included studies, MMPs were reported to be positively correlated with clinical inflammatory conditions around implants [21,29-33]. IL-1β and TNF-α are the two most targeted biomarkers among the majority of included studies and take part in osteoclast formation and bone resorption [6,34-36]. IL-1β regulates the degradation of extracellular matrix components of the plasminogen system and the collagenase activity in inflammation and wound healing [35,37]. It has been shown that inhibition of IL-1β reduced tissue breakdown and the progression of tissue inflammation [26]. TNF-α induces fibroblast apoptosis and reduction of the repair capacity of the peri-implant tissue, but mechanical therapy seems to revert this situation [26]. Of the 19 studies assessed IL-1β, 5 of them showed no statistically significant differences between healthy and diseased groups [20,27,38-40]. One of them reported no significant change after de novo plaque accumulation [25]. Thirteen studies showed higher levels of IL-1β in PICF than healthy implant sites [7,21,22,30,35,36,41-47]. Of the 10 studies assessed TNF-α, 3 of them showed no relationship with this cytokine with peri-implant inflammation [7,25,43], while other 7 studies showed significant relationship with this cytokine [23,24,36,41,46-48]. These findings suggest that pro-inflammatory cytokines such as IL-1β and TNF-α are up to date, the most promising proteins to be used as markers in PICF for differentiation between peri-implantitis and healthy implants. Although ILs are the most of interest in PICF analyses followed by MMPs, there are other biomarkers and enzymes that their levels were evaluated to reflect the local inflammatory condition of implants. Myeloperoxidase (MPO) is an antimicrobial leukocyte-derived enzyme found in high concentrations in the primary granules of leukocytes that catalyzes the formation of a number of reactive oxidant species [48]. Three studies reported significantly higher amounts of MPO in PISF collected around implants with inflammatory lesions [50-52]. In the peri-implant region as well as the natural dentition, it has also been demonstrated that no metabolism is closely related to the status and degree of peri-implant inflammation [52]. Elastase is a major enzyme released from human leukocytes and contributes to tissue damage during inflammation, significantly higher amounts of alkaline phosphatase and elastase were found in PISF around implants with peri-implantitis compared with healthy controls [53]. Prostaglandin E2 (PGE2) is a vasodilator that increases vascular permeability at sites of inflammation and also has a role in bone resorption. One study reported that PGE2 showed positive correlations with gingival index and PD [29], 2 studies reported no difference between peri-implant health and disease condition according to PICF levels [27,45]. Another enzyme associated with bone resorption is cathepsin-K, which is a protease that is released during the inflammatory process after tissue injury. Yamalik et al. [54] showed that cathepsin-K activity was positively correlated with the volume of PISF where there was also inflammatory bone loss, indicating it could be a biomarker used to predict or assess peri-implant alveolar bone loss. Additional biomarkers for peri-implant bone loss have also been studied for peri-implant diseases. OPG/RANKL are both produced by osteoblasts. RANKL normally binds to RANK, which is originate on the surfaces of osteoclast precursors as well as mature osteoclasts, and this binding is necessary for their formation, function, and existence. OPG acts as a trap receptor for RANKL, and its binding prevents RANKL binding to RANK, thereby inhibiting the differentiation of an osteoclast precursor into a mature osteoclast. Arikani et al. [55] reported that sRANKL concentrations, OPG total amounts, and OPG concentrations were significantly lower in peri-implantitis group when compared to healthy group. In contrast, RANKL, RANK, and OPG concentrations were found...
to be significantly higher in peri-implantitis sites compared with healthy implant sites, but the ratio of OPG/RANKL was not different [56]. It was also demonstrated that the OPG/RANKL ratio improved with mechanical treatment of peri-implantitis sites [26]. Osteocalcin is a 5.4-kDa calcium-binding protein of bone and the most abundant non-collagenous protein of the mineralized bone tissue [57]. One study reported higher osteocalcin (OC) levels in PICF as possible biomarker to define the inflammatory conditions around implants [58] whereas two studies reported conflicting results for PICF OC levels [23,57].

Limitations

First limitation of the present systematic review is the wide range of different definitions regarding peri-implant mucositis and peri-implantitis that were employed in the included investigations. Another limitation is a lack of information on whether the inflammatory markers in PICF were matched for the clinical parameters of the respective collection sites, such as PD, clinical attachment loss, and BOP. This is a vital point due to a strong link between PICF biomarker levels and severity and extent of local inflammatory disease. The majority of studies reported the mean clinical parameters of all implant sites or did not clarify whether the clinical parameters presented were related to all implant sites or to sites selected for PICF sampling. Another limitation is; much evidence came from cross-sectional studies. Due to a cyclic progression of peri-implant diseases, the immune-inflammatory event biomarkers responsible for tissue breakdown may not always be active in cross-sectional studies with a single moment of fluid collection.

CONCLUSIONS

For better understanding of the immune inflammatory peri-implant diseases and for developing host-modulation therapies, biomarkers plays a crucial role to aid clinicians to elucidate the complex biologic process involved. Based on this systematic review, it was concluded that inflammatory mediators, such as interleukin-1 beta and plasma tumor necrosis factor-alpha, in crevicular fluid collected from peri-implant pockets are the most used biomarkers to assist in the early diagnosis of peri-implantitis. It is suggested that studies should be conducted to establish a standardized method to diagnose and classify the peri-implant diseases. Standardized investigations should be performed based on the criteria of subject selection, peri-implantitis diagnosis, as well as peri-implant crevicular fluid sampling method (e.g. number and severity of sampling sites, sampling time), sample handling and detection sensitivity/specificity of the used assay.

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Dursun E, Tözüm TF.

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