Primary paediatric epidural sarcomas: molecular exploration of three cases

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Abstract
Background: Primary paediatric epidural sarcomas are extremely rare. Overall, there remains a paucity of knowledge in paediatric epidural sarcomas owing to the infrequent number of cases. The Archer FusionPlex Sarcoma Kit (ArcherDX, Inc) is a next-generation sequencing assay that has been reported to be a useful technique to detect recurrent fusion in sarcomas. We report the molecular exploration of 3 primary paediatric epidural sarcomas—one in the cranium (mesenchymal chondrosarcoma) and 2 in the spine (mesenchymal chondrosarcoma and Ewing sarcoma respectively).

Case presentation: This is a study approved by the hospital ethics board. Clinico-pathological information from 3 consenting patients with primary epidural sarcomas was collected. These selected tumours are interrogated via Archer FusionPlex Sarcoma Kit (ArcherDX, Inc) for genomic aberrations. Results were validated with RT-PCR and Sanger sequencing. All findings are corroborated and discussed in concordance with current literature. Our findings show 2 variants of the HEY1-NCOA2 gene fusion: HEY1 (exon 4)-NCOA2 (exon 13) and HEY1 (exon 4)-NCOA2 (exon 14), in both mesenchymal chondrosarcoma patients. Next, the Ewing sarcoma tumour is found to have EWSR1 (exon 10)-FLI1 (exon 8) translocation based on NGS. This result is not detected via conventional fluorescence in situ testing.

Conclusions: This is a molecularly-centered study based on 3 unique primary paediatric epidural sarcomas. Our findings to add to the growing body of literature for these exceptionally rare and malignant neoplasms. The authors advocate global collaborative efforts and in-depth studies for targeted therapy to benefit affected children.

Keywords: Epidural sarcoma, Next-generation sequencing

Background
Primary paediatric epidural sarcomas are extremely rare and little is known about such tumours. Gene fusions are an important category of driver mutations in paediatric sarcomas [1]. While well-characterized and commonly occurring gene fusions can be identified by standard laboratory assays such as FISH and RT-PCR, an advanced technique such as next-generation sequencing (NGS) is often required to identify rare or novel gene fusions [2]. Gene fusion identification serves to confirm a pathological diagnosis, and is also important in relation to treatment as certain gene fusions have drug-targetable domains [3].

In this study, we report clinical, pathological and molecular features of three unique epidural sarcomas presenting with neurological compromise located in the cranium and spine. We describe the use of a next-generation sequencing-based assay (Archer FusionPlex Sarcoma assay (ArcherDX, Inc)) as a technique to identify gene fusions in these three primary paediatric epidural sarcomas [2, 4, 5].
Case presentation

Case 1: Cranial epidural mesenchymal chondrosarcoma
A previously well 11-year-old female presented with progressively worsening headaches associated with bilateral papilledema. An MRI brain reported a large, heterogeneously enhancing fronto-temporal extra-axial lesion supplied by the right middle meningeal artery. The lesion was resected. Histology showed a mesenchymal chondrosarcoma featuring crowded sheets of primitive spindle to round tumour cells admixed with interspersed islands of neoplastic cartilage demonstrating foci of hyalinization and secondary ossification. Tumour cells were immunoreactive for CD99 but negative for epithelial membrane antigen and progesterone receptor. Ki-67 proliferation index was 1 to 2%. (Fig. 1). The Archer™ FusionPlex Sarcoma Assay detected 2 gene fusion transcripts: HEY1 (exon 4)-NCOA2 (exon 13) and HEY1 (exon 4)-NCOA2 (exon 14) (Fig. 2a and 3).

Case 2: Lumbar intradural extramedullary mesenchymal chondrosarcoma
A 12-year-old female complained of persistent lower back pain associated with bilateral lower limb radicular symptoms over a 4-month duration. The MRI lumbar spine demonstrated an enhancing intradural, extramedullary lesion with adjacent dura thickening at the level of L2. Laminectomy and excision of the lesion was performed. Histology showed a mesenchymal chondrosarcoma featuring round to spindle cells with interspersed cartilage and bone formation. Tumour cells showed diffuse CD99 immunoreactivity and negative staining for epithelial membrane antigen, STAT6 and glial fibrillary acid protein. The Ki-67 index was about 30%. (Fig. 4). Similar to the previous case, the Archer™ FusionPlex Sarcoma Assay detected 2 gene fusion transcripts: HEY1 (exon 4)-NCOA2 (exon 13) and HEY1 (exon 4)-NCOA2 (exon 14) (Fig. 2b).

Case 3: Thoracic intradural extramedullary Ewing sarcoma
An 11-year-old female presented with a 3-day history of acutely worsening lower limb weakness, numbness, urinary retention and lax anal tone. There was no prior history of associated trauma or injury, and no complaints of back pain. MRI spine revealed an extramedullary extradural soft tissue mass spanning T6 to T9 and causing moderate to severe spinal canal stenosis. This mass was heterogeneous enhancing with suggestion of a dural tail. She underwent emergency T7 to T9 laminectomy and excision of tumour. Histopathology reported a malignant round cell neoplasm with CD99 immunopositivity consistent with Ewing sarcoma (Fig. 5). Fluorescence in situ (FISH) with an EWSR1 break-apart probe was unexpectedly negative. This FISH test was performed twice with different sections of the tumour. However, the Archer™ FusionPlex Sarcoma Assay reported a EWSR1 (exon 10)-FLI1(exon 8) translocation (Fig. 6).

Study outline and experimental details
This was a single-institution, retrospective study approved by our hospital ethics review board. All patients’ legal guardians (as they are below 21 years old) provided signed informed consent for the research use of their medical data and pathological material obtained as part of their routine clinical data, and publication. The project is exploratory in design, and included 3 patients less than 18 years of age with a histopathological diagnosis of epidural sarcoma.

Total RNA was extracted from macro-dissected tissue sections using the Promega ReliaPrep™ FFPE Total RNA Miniprep System (Promega, USA) as per manufacturer’s protocol. The quantity of extracted RNA was measured using the QuantiFluor™ RNA System (Promega, USA). Archer™ FusionPlex Sarcoma Assay is an RNA-based targeted sequencing assay that can identify fusions involving any of 26 sarcoma-related genes (ALK, CAMTA1, CCNB3, CIC, EPC, EWSR1, FKHR, FUS, GLI1, HMGA2, JAZF1, MEAF6, MKL2, NCOA2, NTRK3, PDGFB, PLAG1, ROSI, SS18, STAT6, TAF15, TCF12, TFE3, TFG, USP6, YWHAE) without prior knowledge of fusion partners or breakpoints. 150 ng of RNA was used for library preparation utilizing the Archer™ FusionPlex Sarcoma Panel kit (AK00328) following the manufacturer’s protocol (ArcherDX, USA). The prepared library was sequenced using an Ion Torrent PGM™ next-generation sequencer. The Hi-Q™ sequencing kit and Hi-Q™ View CHEF kit were used according to the manufacturer’s protocol (Life Technologies, USA). Data was analyzed by the Archer Data Analysis (version 5.1.0) portal (ArcherDX, USA). Confirmatory RT-PCR and Sanger Sequencing on the amplicon product were performed to confirm the fusion variant. Two separate confirmatory RT-PCR using primers for HEY1/NCOA2 (4–13) – 5’ CGAG ATCCTGCAGATGACC 3’ (forward), 5’ GAGG TATCACTGAGTGGGACTA (reverse) and HEY1/ NCOA2 (4–14) – 5’ CGAGATCCTGACATGACC 3’ (forward), 5’ CTGCTGGTCTCCATCAT (reverse) flanking the breakpoint, and Sanger Sequencing on the amplicon products were performed to confirm the fusion variant. For the thoracic epidural tumour, confirmatory RT-PCR using primers – 5’ GAGC GAGTGGGTCTCAAATA 3’ (forward), 5’ GTTT GGCTAGGCGACTG (reverse) flanking the breakpoint, and Sanger Sequencing on the amplicon product was performed to confirm the fusion variant.
Discussion
Childhood sarcomas occurring in the CNS children are rare and poorly understood. To begin with, mesenchymal chondrosarcoma (MCS) is an infrequent member of the heterogeneous sarcoma family of tumours [6]. Presently, its developmental mechanisms remain poorly understood. Clinical experience with CNS-related MCS is also extremely limited; nonetheless, local recurrence and distant metastasis has been reported [7]. Presently, CNS-based MCS has only been reflected in surgical case reports and small series in the literature [7–15]. Recently, the HEY1 (exon 4)-NCOA2 (exon 13) gene fusion is reported as a recurrent event unique to all MCS [16, 17]. In this study, we report 2 variant gene fusions HEY1 (exon 4)-NCOA2 (exon 13) and HEY1 (exon 4)-NCOA2 (exon 14) in two of our MCS patients. With reference to current literature, only HEY1 (exon 4)-NCOA2 (exon 13) gene fusion has been previously reported as a recurrent event unique to MCS [16, 17]. However, the concurrent HEY1 (exon 4)-NCOA2 (exon 14) fusion found in both of our CNS cases has not been previously described.

At this point in time, it is hard to postulate if this result is unique to CNS-based MCS patients, or to other MCS tumours found in the rest of the body. In addition, the concurrent occurrence of these 2 transcripts is intriguing. Firstly, there is a possibility that a splicing event has given rise to the two variant gene fusions.
Nonetheless, as the breakpoints involve different exons, this is not a usual alternative splicing phenomenon. Next, this phenomenon may be biallelic in that there are two variants of the gene fusion occurring in each of the two copies of the genes. Functional validation at of these findings is required to elucidate their biological meaning. Furthermore, in the context of our patients, there is a role to explore if having 2 aberrant gene fusion variants implies increased oncogenicity in these rare tumours.

Following in the footsteps of scarcity, primary epidural Ewing sarcoma (EWS) is too, very rare in the EWS family of solid tumours. For our third patient, the Archer™ FusionPlex Sarcoma Assay detected EWSR1 (exon 10)-FLI1(exon 8) translocation. EWS/FLI has at least 10 different isoforms [18]; The most common are: type 1 [EWSR1 (exon 7)-FLI1(exon 6)], type 2 [EWSR1 (exon 7)-FLI1(exon 5)], type 3 [EWSR1 (exon 10)-FLI1(exon 6)] and type 4 [EWSR1 (exon 7)-FLI1(exon 7)] translocations [18]. Here, our result represents an uncommon variant in the cohort of EWSR1-FLI1 translocation breakpoints.

Broadly speaking, non-osseous forms of EWS, particularly those that originate in the epidural space, make up to approximately 20 case reports in the literature [19–21]. Next, EWS is notorious for multiple fusion combinations involving several partner genes [22]. To complicate matters, the specific breakpoint in each partner gene can be variable, resulting in a variety of exon-exon fusion combinations at the transcript level [23]. At this point, we remain uncertain if individual fusion isoforms portend different patient outcomes [24, 25]. However, most are in agreement that detection of translocations at the exon level will have implications for diagnosis, prognosis and treatment of EWS patients [18]. In the context of our patient, the gene fusion EWSR1 (exon 10)-FLI1(exon 8) was investigated using a EWSR1 break-apart probe that could detect a range of EWSR1 gene disruptions. The probes included the translocation region of interest. However, this particular gene fusion was only detectable using NGS methods, and not via FISH. Despite its proven reliability as

![Fig. 3](image-url)
Fig. 5 Representative post-contrast T1-weighted MRI images in sagittal (a) and axial (b) demonstrating a heterogeneously enhancing T6 to T9 extramedullary, extradural soft tissue mass with suggestion of a dural tail. Haematoxylin and eosin stain slide (× 100) illustrating a malignant round cell neoplasm. Immunohistochemistry slide (× 100) shows positivity for CD99.

Fig. 4 Sanger sequencing of the RT-PCR amplicon products for Patient 1 confirm the presence of both variants (a) and (b) of HEY1-NCOA2 gene fusion.
diagnostic method, FISH is reputed to have a very small risk of false negative results [26]. This technical limitation has been reported to be especially exemplified by EWS [26, 27], as the FISH technique relies on the identification of gene rearrangements to which probes are very specifically directed [18]. For our patient’s case, the consideration may be for multi-color FISH with more than 2 different probes [28]. Under such circumstances, obscure translocations that lead EWSR1 insertion into partner genes may be more readily detectable. However, this particular type of FISH testing is not readily available at our institution. Hence, this case highlights the utility of novel genomic advancements as a clinical tool in challenging cases. It should too, be emphasized that selected patients may proceed to be studied by more than one diagnostic modality, especially if the initial test is negative and the clinical features point to a high probability of genetic aberrations [29].

In general, paediatric cancers appear to be the consequence of chromosomal rearrangements, rather than mutation events [3]. Detection and characterization of gene fusions has been of great importance for clinical purposes, as well as for understanding tumorigenesis [30]. Furthermore, it has been reported that sarcoma patients whose tumours were interrogated by NGS as part of their clinical management received the option of access to targeted agents in their treatment [31]. This is because a difference in clinical outcome may be predicted by one or more of the following: firstly, incidence of a specific fusion; next, the presence of one or more variants of a fusion; or finally, the presence of one of a heterogeneous group of unrelated fusions [32, 33]. More significantly, as the cases in discussion are rare paediatric tumours, an isolated fusion found in a single patient can be important, especially for future therapeutic targeting [3]. Identification of specific fusion transcripts, including unusual variants of gene-related splices may help in the development of novel therapeutics to block their aberrant activity in cancer cells [34].

Study critique and future directions
The authors acknowledge that there are limitations that should be highlighted in this study. First and foremost, this is a retrospective study with a small number of cases. However, it should be emphasized that primary paediatric epidural sarcomas are extremely rare. Hence, given the infrequency of such patients, every case should be regarded as significant. Moving forward, the role of functional studies to determine the
The authors declare that they have no competing interests.

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A copy of the publication consent form is available for clinical images was obtained by the respective patients and/or their parents/legal guardians. Written informed consent for publication of their clinical details and/or their clinical material. Written, informed consent was obtained by the Singhealth CIRB: 2014/2079) protocol for patient enrolment and study of this is a study approved by the hospital ethics board (Study Reference: VIVA-KKH Paediatric Brain and Solid Tumour Programme.

Ethics approval and consent to participate

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Consent for publication

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Competing interests

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Abbreviations

CNS: Central nervous system; EWS: Ewing sarcoma; MCS: Mesenchymal chondrosarcoma; MRI: Magnetic resonance imaging; NGS: Next-generation sequencing

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

Conceptualization: SL. Data curation: SL, GT, WT, DL. Formal analysis: CH, HC, KM. Investigation: SL, WY, CH, NB. Methodology: SL, KC, CK. Project administration: SL, LP. Resources: KC, DL, ET, SY. Validation: CH, NB, HC. Writing – original draft: SL. Writing – review & editing: SL, KC. All authors read and approved the final manuscript.

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References

1. Mentens F, Johansson B, Fioresi T, Mitelman F. The emerging complexity of gene fusions in cancer. Nat Rev Cancer. 2015;15(6):371–81.
2. Szurian K, Kashofer K, Liegl-Atzwanger B. Role of next-generation sequencing as a diagnostic tool for the evaluation of bone and soft-tissue sarcomas. Pathobiology. 2017;84(6):323–38.
3. Dupain C, Harttrampf AC, Urbiniati G, Georger B, Massaad-Massade L. Relevance of fusion genes in pediatric cancers: toward precision medicine. Mol Ther Nucleic Acids. 2017;6:315–26.
4. Mok Y, Pang YH, Sanjeev JS, Kuick CH, Chang KT. Primary renal hybrid low-grade Fibromyxoid sarcoma-Sclerosing epithelioid Fibrosarcoma: an unusual pediatric case with EWSR1-CREB3L1 fusion. Pediatr Dev Pathol. 2018;21(6):574–9.
5. Lam SW, Cleston-Jansen AM, Cleven AHG, Ruano D, van Wezel T, Suhail K, Boeve J. Molecular analysis of gene fusions in bone and soft tissue tumors by anchored multiplex PCR-based next-generation sequencing. J Mol Diagn. 2018;20(5):653–62.
6. Dantonello TM, Int-Veen C, Leuscher I, Schuck A, Furtwaengler R, Claveliez A, Schneider DT, Klingebiel T, Bielack SS, Koscielniak E, et al. Mesenchymal chondrosarcoma of soft tissues and bone in children, adolescents, and young adults: experiences of the CWS and COSS study groups. Cancer. 2008;112(11):2424–31.
7. Lin L, Varki A, Baxter W, Ng T, Diagnostic pitfalls in the diagnosis of mesenchymal chondrosarcoma arising in the central nervous system. Neuropathology. 2012;32(1):82–90.
8. Agravail P, Gupta A, Juneja S, Gupta V. Infracranial Mesenchymal Chondrosarcoma: A Case Report. Indian J Neurosurg. 2015;4:3–14.
9. Andersson C, Österlund G, Enlund F, Kindblom LG, Hansson M. Primary spinal intradural mesenchymal chondrosarcoma with detection of fusion gene HEY1-NCOA2: a paediatric case report and review of the literature. Oncol Lett. 2014;8(4):1608–12.
10. Anvari K, Gharib M, Sanburi A, Javadina SA. Intracranial mesenchymal chondrosarcoma: case report and review of literature. GMJ. 2016;5(4):219–24.
11. Di Lorenzo N, Palatinsky E, Artico M, Palma L. Dural mesenchymal chondrosarcoma of the lumbar spine. Case report. Surg Neurol. 1989;31(6):470–2.
12. Zucker DK, Horoupian DS. Dural mesenchymal chondrosarcoma. Case report. J Neurosurg. 1978;48(5):829–33.
13. Zibis AH, Wade Shader M, Segal LS. Case report: mesenchymal chondrosarcoma of the lumbar spine in a child. Clin Orthop Relat Res. 2010; 468(8):2288–94.
14. Scheithauer BW, Rubinstein LJ. Meningeal mesenchymal chondrosarcoma: report of 8 cases with review of the literature. Cancer. 1978;42(3):2644–52.
15. Sajjad EA, Sikora K, Paciejewski T, Garbicz F, Paskal W, Sachtz M, Graftowski W, Woźniarski PK. Intraparenchymal mesenchymal chondrosarcoma of the frontal lobe—a case report and molecular detection of specific gene fusions from archival FFPE sample. Clin Neuropathol. 2015;34(5):288–95.
16. Wang L, Motio T, Khanin R, Olthen A, Mentens F, Bridge J, Dal Cin P, Antonescu CR, Singer S, Hameed M, et al. Identification of a novel, recurrent HEY1-NCOA2 fusion in mesenchymal chondrosarcoma based on a genome-wide screen of exon-level expression data. Genes Chromosomes Cancer. 2012;51(2):127–39.
17. Fitchie KJ, Jin I, Ruano A, Oliveira AM, Rubin BP. Are meningial hemangiopericytoma and mesenchymal chondrosarcoma the same?: a study of HEY1-NCOA2 fusion. Am J Clin Pathol. 2013;140(3):670–4.
18. Luo W, Milash B, Dalley B, Smith R, Zhou H, Dutrow N, Cairns BR, Lessnick SL. Antibody detection of translocations in Ewing sarcoma. EMBO Mol Med. 2012;4(6):453–61.
19. Hsieh CT, Chiaw YH, Tsai WC, Sheu LF, Liu MY. Primary spinal epidural Ewing sarcoma: a case report and review of the literature. Turk J Pediatr. 2008;50(3):282–6.
20. Gopalakrishnan CV, Shrivastava A, Easwer HV, Nair S. Primary Ewing’s sarcoma of the spine presenting as acute paraplegia. J Pediatr Neurosci. 2012;7(1):64–6.
21. Dogan S, Lekovic GP, Theodore N, Horn EM, Eschbacher J, Rekate HL. Primary thoracolumbar Ewing’s sarcoma presenting as isolated epidural mass. Spine J. 2009;9(1):e9–14.
22. Barr FG, Womer RB. Molecular diagnosis of Ewing family tumors: too many fusions... J Mol Diagn : JMD. 2007;9(4):437–40.
23. Chang KTE, Goytain A, Tucker T, Karsan A, Lee CH, Nielsen TO, Ng TL. Development and evaluation of a pan-sarcoma fusion gene detection assay using the NanoString nCounter platform. J Mol Diagn : JMD. 2018;20(1):63–77.
24. van Doorninck JA, Ji L, Schaub B, Shimada H, Wing MR, Kioido MD, Lesnick SL, Marina N, Triche TJ, Spostro R, et al. Current treatment protocols have eliminated the prognostic advantage of type 1 fusions in Ewing sarcoma: a report from the Children’s oncology group. J Clin Oncol. 2010;28(12):1089–94.
25. Le Deley MC, Delattre O, Schaefer KL, Burchill SA, Koehler G, Hogendoorn PC, Lion T, Poremba C, Marandet J, Ballet S, et al. Impact of EWS-ETS fusion type on disease progression in Ewing’s sarcoma/peripheral primitive neuroectodermal tumor: prospective results from the cooperative euro-E.W.I.N.G. 99 trial. J Clin Oncol. 2010;28(12):1982–8.
26. Noujaim J, Jones RL, Swansbury J, Gonzalez D, Benson C, Judson I, Fisher C, Thway K. The spectrum of EWSR1-rearranged neoplasms at a tertiary sarcoma Centre; assessing 772 tumour specimens and the value of current ancillary molecular diagnostic modalities. Br J Cancer. 2017;116(2):669–78.
27. Newby R, Rowe D, Paterson L, Farquharson MA, MacDuff E, Coupe A, Hale J, Dilday P, Bown N. Cryptic EWSR1-FLI1 fusions in Ewing sarcoma: potential pitfalls in the diagnostic use of fluorescence in situ hybridization probes. Cancer Genet Cytogenet. 2010;200(1):60–4.
28. Eastmond DA, Schuler M, Rupa DS. Advantages and limitations of using fluorescence in situ hybridization for the detection of aneuploidy in interphase human cells. Mutat Res. 1995;348(4):153–62.
29. Niu X, Chuang JC, Berry GJ, Wakelee HA. Anaplastic lymphoma kinase testing: IHC vs. FISH vs. NGS. Curr Treat Options in Oncol. 2017;18(12):71.
30. Mitelman F, Johansson B, Mertens F. The impact of translocations and gene fusions on cancer causation. Nat Rev Cancer. 2007;7(4):233–45.
31. Groisberg R, Roszik J, Conley A, Patel SR, Subbiah V. The role of next-generation sequencing in sarcomas: evolution from light microscope to molecular microscopic. Curr Oncol Rep. 2017;19(12):78.
32. Nakada S, Minato H, Nojima T. Clinicopathological differences between variants of the NAB2-STAT6 fusion gene in solitary fibrous tumors of the meninges and extra-central nervous system. Brain Tumor Pathol. 2016;33(3):169–74.
33. Lee CH, Nucci MR. Endometrial stromal sarcoma—the new genetic paradigm. Histopathology. 2015;67(1):1–19.
34. Cantile M, Maara L, Franco R, Asciero P, Liquori G, De Chiara A, Botti G. Molecular detection and targeting of EWSR1 fusion transcripts in soft tissue tumors. Med Oncol. 2013;30(1):412.