In situ expression and significance of B7 costimulatory molecules within tissues of human gastric carcinoma

Xiao-Li Chen, Xu-Dong Cao, An-Jing Kang, Kang-Min Wang, Bao-Shan Su, Yi-Li Wang

AIM: To explore the role and significance of costimulatory molecules B7H1, B7H2 and ICOS within tissues of human gastric carcinoma and the possible mechanisms in tumor escape.

METHODS: mRNA expressions of costimulatory molecules including B7H1, B7H2, ICOS and B7-1 in tissues of human gastric carcinoma were investigated by in situ hybridization using digoxigenin-labeled oligonucleotide-probes. The tissue of chronic gastric ulcer was used as a control. All data were analyzed by SPSS statistic software.

RESULTS: At the site of gastric carcinoma, mRNA expression levels of B7H1, B7H2 and ICOS were much higher than that of B7-1. Their mRNA positive expression indexes were 0.512±0.333, 0.812±0.454, 0.702±0.359 and 0.293±0.253, respectively. The positively stained cells were mainly tumor infiltrating lymphocytes (TILs), and some tumor cells. The difference between them was greatly significant P<0.005. The mRNA expression levels of four molecules were not correlated to the pathological grade and metastasis of gastric carcinoma.

CONCLUSION: ICOS-B7H costimulatory pathway may be predominant at the site of gastric carcinoma. B7-1 mRNA might be the basis of ICOS-B7H interaction. ICOS-B7H interaction induces the production of IL-10 which inhibits the antitumor immune responses. Therefore, it is supposed that ICOS-B7H costimulatory pathway may be involved in the negative regulation of cell-mediated immune responses.

INTRODUCTION
Tumor immunity is primarily cell-mediated. The tumor antigen leads to anti-tumor immune response of the host. A body of evidences have shown that the specific activation of lymphocytes requires two signals; one is provided by T-cell receptor complex coupled to CD3/MHC peptide antigen, and the other is costimulatory signal delivered by interaction of costimulatory molecules with their ligands expressed on antigen presenting cells (APCs). In the absence of costimulatory signals, antigen-MHC complex interaction may lead to T-cell clonal anergy or deletion, and thus, the effective cellular immune response can not be induced. Furthermore, there are evidences that activation of lymphocyte subpopulation with different function needs unique costimulation factors. Therefore, investigation of costimulators on infiltrating lymphocytes and tumor cells at the tumor site and of their significance will be of great help in developing new methods of anti-tumor immunotherapy. Cell-mediated immunity is the main mechanism of anti-tumor immunity of the host. With presentation of antigen recognition and deepening of the understanding of activated T-cells, it is believed that in the presence of costimulatory signals, effective anti-tumor immune response presents tumor antigen multipeptide to T cells and stimulates T-cell immune response. B7 is an important costimulatory molecule. Most tumors lack or have a low expression of B7-1, so they do not induce effective anti-tumor immune response. The tumor cells escape from immune system of the host and continue to grow[1-15]. Although many in vitro experiments support this idea, in situ studies on human tumors have shown that most tumor tissues express the B7-1 or B7-2 molecules. So the idea meets challenges. An in vitro study on new members of B7 family - B7H1, B7H2 and ICOS suggested that the role of ICOS-B7H costimulatory pathway stimulated IL-10 production and induced secretion of Th2 type cytokines. The present study was to explore the role and biological significance of ICOS-B7H costimulatory signals and the possible mechanisms in tumor escape, and to provide the theoretical foundation for designing effective schemes of immunotherapy of carcinoma.

MATERIALS AND METHODS
Patients
17 patients with gastric carcinoma (10 metastatic, and 7 non-metastatic) and 6 patients with gastric ulcer were identified by pathological diagnosis at the Department of Pathology, Second Hospital of Xi’an Jiaotong University. The fresh tissue samples were instantly fixed in 10% formalin, and embedded in paraffin according to routine procedures. Tonsil tissue served as a positive control.

Main reagents
Digoxigenin (DIG)-labeling and detection kits were purchased from Boehringer Mannheim Company in Germany. Solution for in situ hybridization was mixed with diethyl pyrocarbonate (DEPC) water. All apparatuses were baked at 180 °C for four hours and DEPC water was digested with RNase.

Specimens
Sections (4-5 μm) cut from the tissue block were de-paraffined in 65 °C oven for 24 hours and then stored at -70 °C until being used.
Labeling of oligonucleotide probes and detection of their sensitivities

Labeling On the ice, 4 µl reaction buffer (vial 1), 4 µl CoCl2 solution (vial 2), 1 µl probe (100 pmol), 1 µl DIG-dUTP solution (vial 3), 1 µl dATP solution (vial 4), 1 µl terminal transferase (vial 5) and 8 µl DEPC water, were consecutively added into an eppendorf tube, and carefully mixed and incubated at 37 °C for 15 minutes. 2 µl stopping solution (prepared by mixing 1 µl glycogen solution (vial 9) with 200 µl 0.2 mol/L EDTA solution) was added into the tube to stop the labeling reaction. Then, 2.5 µl 4mol/L CrCl2 and 75 µl pre-cold pure alcohol were added to deposit the labeled. After the supernatants were drained out, the probe was washed with 50 µl 70 % pre-cold alcohol and dried with a frozen dry machine, and stored at -20 °C until being used.

Detection of sensitivity Sensitivity of each probe was measured by the detection kit following the manufacturer’s instructions. The lowest concentration for positive staining was 1.25 pmol/ml. The oligonucleotide probe sequences were as follow: B7-1 5'-CAT GAA GCT GTG GTT GGT TG-3'; B7H1 5'-TGC TTG TCC AGG TGA CTT CG-3'; B7H2 5'-CCA TCG CTC TGA CCT TC-3'; and ICOS 5'-TTC AGC TGG CAA CAA AGT TG-3'.

Hybridization Sections stored at -70 °C were recovered at room temperature, deparaffined in dimethylbenzene, and hydrated in gradient alcohol. They were digested in 0.5 µg/ml fresh proteinase K solution at room temperature for 20 minutes, and fixed in 4 % poly-formaldehyde for 20 minutes after digestion was stopped in 0.2 % glycine, and then repeatedly rinsed with 1×PBS for 10 minutes. The sections were treated in 0.2N HCl solution at room temperature for 10 minutes, rinsed with DEPC water for 3 minutes; and dehydrated in gradient alcohol. At 42 °C in a bio-hybridization oven, all sections were pre-hybridized for 2 hours, and then hybridized for 20 hours. After hybridization, they were rinsed with 2×SSC for two times, 15 minutes each, rinsed with 1×SSC for 30 minutes and finally with buffer I solution (pH 7.5) for 6 minutes.

Detection 50 µl blocking reagent (containing 8 % normal goat serum, 0.3 % Triton X-100 buffer I) was instilled onto each section. The sections were incubated at 37 °C for one hour and a half, and then rinsed with 50 µl anti-DIG-AP conjugate (1:500) was added onto each section, and all sections were incubated at 37 °C for two hours and rinsed with buffer I two times, 15 minutes each, then rinsed with buffer III (100 mmol/L Tris-HCl, 100 mmol/L NaCl and 50 mmol/L MgCl2, pH 9.5) for 5 minutes, and stained with NBT-BCIP at 20 °C for 12 hours. Finally, the reaction was stopped with buffer IV solution (10 mmol/L Tris-HCl, and 1 mmol/L EDTA, pH 8.0), and the sections were washed in running water, counterstained with methyl green, dehydrated, cleaned in xylene and coverslipped.

Evaluation of results Under a light microscope, positively hybridized signals were mainly located in the infiltrating lymphocytes and colored purple blue. Reactivity was scored using a semi-quantitative method. First, each section was randomly scored one thousand cells. The positive cell ratio in each section was calculated. Then, according to the staining intensity of positive cells, results were graded as following: intense 3+, moderate 2+, weak 1+ and negative, a score of 3, 2, 1, 0 was assigned respectively. Finally, the positive cell ratio multiplied with staining intensity score of the positive cells was regarded as positive index of mRNA expression of costimulatory molecules.

Control experiment Expression of ICOS mRNA of the chronic tonsillitis lymphocytes was regarded as a positive control. The results were negative control when no probe was used.

Statistical analysis All data were expressed as t±s and analyzed by SPSS statistic software. Differences in values were considered significant if \( P<0.05 \).

RESULTS mRNA expression of costimulatory molecules within tissues of human gastric carcinoma

At the site of gastric carcinoma, positive cells for B7H1, B7H2, ICOS were mainly tumor infiltrating lymphocytes (Figure 1). Some positive cells were tumor cells. Their mRNA positive expression indexes were 0.512±0.333, 0.812±0.454 and 0.702±0.359, respectively. B7-1 mRNA was expressed on tumor cells and TILs (Figure 2), with a positive expression index being 0.293±0.253. The mRNA expression levels of B7H1, B7H2 and ICOS were significantly higher than that of B7-1 (all \( P<0.05 \)).

Figure 1 B7H1 mRNA expression of tumor infiltrating lymphocytes (TILs) within tissues of human gastric carcinoma (in situ hybridization, NBT-BCIP Staining, counterstained with methyl green, ×1 000).

Figure 2 B7-1 mRNA expression of tumor infiltrating lymphocytes (TILs) within tissues of human gastric carcinoma (in situ hybridization, NBT-BCIP Staining, counterstained with methyl green, ×1 000).

Expression of costimulatory molecules in relation to metastasis of gastric carcinoma

The expression levels of B7-1, B7H1, B7H2 and ICOS had no statistical significance between metastasis and non-metastasis.
of gastric carcinoma (P > 0.05), which suggested that expressions of costimulatory molecules at tumor site were not correlated to the metastasis of tumor.

Expression of costimulatory molecules in relation to differentiation of gastric carcinoma
mRNA expression levels of B7-1, B7H1 and ICOS on infiltrating lymphocytes at the tumor site was not significantly different between well-differentiated and poorly-differentiated. However, the expression level of B7H2 mRNA was significantly higher in well-differentiated than in poorly-differentiated (1.040±0.400 vs 0.651±0.436, P = 0.047). mRNA expression of B7-1 was greatly lower than that of B7H1, B7H2 and ICOS in both well-differentiated and poorly-differentiated gastric carcinoma which might indicate that function of B7-1 was different from other costimulatory molecules in gastric carcinoma.

Expression of costimulatory molecules in comparison between gastric carcinoma and gastric ulcer
In the tissue of gastric ulcer, the expression levels of B7-1, B7H1, B7H2 and ICOS on infiltrating lymphocytes were higher than those in gastric carcinoma. But there was no difference in the expression of B7H1, B7H2 and ICOS between the tissues of gastric carcinoma and gastric ulcer except B7-1. The effect of costimulators in the gastric ulcer was not clear.

DISCUSSION
Cell-mediated immunity is the main mechanism of antitumor immunity of the host. With presentation of antigen identification and deepening of the understanding of T-cells activated, it is believed that in the presence of costimulatory signals, effective antitumor immune response presents tumor antigen multipeptide to T cells and stimulates T-cell immune response. B7 is an important costimulatory molecule. Most tumors lack or have a low expression of B7-1, so they do not induce effective antitumor immune response. The tumor cells escape from immune system of the host and continue to grow. Though many in vitro experiments support this idea, in situ studies on human tumors have shown that most tumor tissues express the B7-1 or B7-2 molecules. So the idea meets challenges.

The effective anti-tumor immune response requires three factors, including immunogenicity, costimulatory signals and Th1 type cytokines. In the past, it was emphasized that cytokines or/antigen essence determined the classification of immune responses. Generally, weak immunogenicity of tumor cells primarily induced Th2 response. In recent years, studies on costimulatory signals in vitro have shown that different costimulatory signals play a determinant role in immune response type. There might be Th2 predominant costimulatory signals in human tumor site. The present study has detected mRNA expression of B7H1, B7H2 and ICOS within the tissue of human gastric carcinoma, using in situ hybridization with DIG-labeled oligonucleotide probes. The results suggest that tumor infiltrating lymphocytes in tissue of gastric carcinoma express high levels of recently discovered members of B7 costimulatory family, B7H1, B7H2 and ICOS. Compared with mRNA expression level of B7-1, difference was dramatically significant. But mRNA expression levels of B7H1, B7H2 and ICOS were not correlated to the pathological grade and metastasis of gastric carcinoma. The experiments in vitro have verified that the interaction between B7h and ICOS stimulates the proliferation of T-cells and production of Th2 cytokines, preferentially secretion of IL-10. This is probably the main reason that Th2 cytokines may predominate at the tumor immune- microenvironment of gastric carcinoma, and thus the type of immune response shifts from Th1 to Th2. There was an increase of Th2 cytokine-IL-10 in tumor immune- microenvironment of gastric carcinoma.

In this study, we have also observed the expression of B7-1 in the tissue of gastric carcinoma, and found that tumor infiltrating lymphocytes (TIL) and tumor cells expressed low gastric carcinoma levels of B7-1 mRNA. Now that there were costimulatory molecules in the tissues of gastric carcinoma, why did not the host produce the effective antitumor immune response? With the further study of immunology and tumor immunology, a new concept related to interaction between TIL and tumor cells was introduced. That is the tumor microenvironment. This theory supposes that immune system of the host cannot eliminate tumors because there are many inhibitors of tumor immunity at the site of tumor microenvironment. For example, tumor cells produce TGF-β, PGE2, and so on, which are accumulated at tumor site and interfere with the activity of recruited immunocytes. Activated immunosuppressives further augment the immunodepression mechanism in tumor microenvironment. In addition, a recent study has shown that B7-1 costimulation also enhances the up-regulation effects of ICOS costimulation which results in the increase of Th2 cytokines, preferentially inducing the secretion of IL-10.

The present experimental results suggest that the tumor infiltrating lymphocytes in the tissues of gastric carcinoma express high levels of B7H1, B7H2 and ICOS mRNA. Some tumor cells also express them but the expression levels are low. This indicates that there are costimulatory signal pathways in the site of gastric carcinoma. Since the interaction between B7H1, B7H2 and ICOS stimulates proliferation of T cells by various ways and induces secretion of IL-10. IL-10 is known to be an immune inhibitor, which down-regulates function of Th1 type cells, and inhibits antitumor immune response of the body, and thus the body is in the state of immune inhibition. Therefore, it is supposed that although interaction between B7H1, B7H2 and ICOS delivers costimulatory signals to T-cells, promotes their activation, the effective antitumor immunity can not be induced. Tumor cells can escape from the immunosurveillance of the host and continue to grow. This is due to the fact that many inhibitors at tumor site have blocked the role of specifically activated T cells. As far as most tumors are concerned, the host’s immunoreaction can not eliminate the tumor because Th2 type response is stimulated, or it can be recognized by immune system of the body. Maybe there are other regulation mechanisms. The present study indicates that costimulatory pathways might exist at the tumor site with different functions. ICOS-B7h pathway is predominant and plays an important role in negative regulation of cell-mediated immune response. This finding might provide a new approach for designing effective immune-therapy of carcinoma.

REFERENCES
1 EllisJH, Burden MN, Vinogradov DV, LingeC, CroweJS. Interactions of CD80 and CD86 with CD28 and CTLA4. J Immunol 1996; 156: 2700-2709
2 GuinanEC, GribbenJG, BoussiotisVA, FreemanGJ, NadlerLM. Pivotal role of the B7:CD28 pathway in trans-plantation tolerant immunity. Blood 1994; 84: 3261-3262
3 IkemizuS, GilbertRJ, FennellyJA, CollinsAV, HarlosK, JonesEY, StuartDI, DavisSJ. Structure and dimerization of a soluble form of B7-1. J Immunol 1998; 158: 2367-2369
4 HungK, HayashiR, Lafond-WalkerA, LowensteinC, PardollD, LevitskyH. The central role of CD4+ T cells in the antitumor immune response. J Exp Med 1995; 188: 2347-2366
5 GansR, HanahanD. Tumor microenvironment can restrict the effectiveness of activated antitumor lymphocytes. Cancer Res 1998; 58: 4673-4681
6 DenfeldRW, DietrichA, WuttigC, TanczosE, WeissJM,
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Vanscheidt W, Schopf E, Simon JC. In situ expression of B7 and CD28 receptor families in human malignant melanoma: relevance for T cell-mediated anti-tumor immunity. Int J Cancer 1995; 62: 259-265

7 Salvadori S, Martineili G, Zier K. Resection of solid tumors reverses T cell defects and restores protective immunity. J Immunol 2000; 164: 2214-2220

8 Yu X, Fournier S, Allison JP, Sharpe AH, Hodges RJ. The role of B7 costimulation in CD4/CD8 T cell homeostasis. J Immunol 2000; 164: 3543-3553

9 Martín-Fontecha A, Moro M, Crosti MC, Veglia F, Casorati G, Delabonna P. Vaccination with mouse mammary adenocarcinoma cells coexpressing B7-1(CD80) and B7-2(CD86) discloses the dominant effect of B7-1 in the induction of antitumor immunity. J Immunol 2000; 164: 698-704

10 Kinoshita K, Tesch G, Schwarting A, Maron R, Sharpe AH, Kelley VR. Costimulation by B7-1 and B7-2 is repaired for autoimmune disease in MRL-Fas mice. J Immunol 2000; 164: 6046-6056

11 Geldhof AB, Raes G, Bakkus M, Devos S, Thielemans K, De Baetselier P. Expression of B7-1 by highly metastatic mouse T lymphomas induces optimal natural killer cell-mediated cytotoxicity. Cancer Res 1995; 55: 2730-2733

12 Huang M, Wang J, Lee P, Sharma S, Mao JT, Meissner H, Uyemura K, Modlin R, Wollman J, Dubinett SM. Human non-small cell lung cancer cells express a type 2 cytokine pattern. Cancer Res 1995; 55: 3847-3853

13 Romagnani S. The Th1/Th2 paradigm. Immunol Today 1997; 18: 263-266

14 Fujii H, Inobe M, Kimura F, Murata J, Murakami M, Onishi Y, Azuma I, Uede T, Saiki I. Vaccination of tumor cells transfected with the B7-1(CD80)gene induces the anti metastatic effect and tumor immunity in mice. Int J Cancer 1996; 66: 219-224

15 Wu TC, Huang SY, Jaffe EM, Levitsky HI, Pardoll DM. A reassessment of the role of B7-1 expression in tumor rejection. J Exp Med 1995; 182: 1415-1421

16 Judge TA, Tang A, Turkla LA. Immunosuppression through blockade of CD28/B7-mediated costimulatory signals. Immunol Res 1996; 15: 38-49

17 Hu JY, Wang S, Zhu JG, Zhou GH, Sun QB. Expression of B7 costimulatory molecules by colorectal cancer cells induces tumorigenesis and induces anti-tumor immunity. World J Gastroenterol 1999; 5: 147-151

18 Ren XF, Luo KK. Expression of B7 and liver diseases. Shijie Huanren Xiuha 2000; 1999; 7: 415-416

19 Guo J, Wang Y, Liu J, Wang Y, Yang Z. An in situ study on immunostimulatory molecules in cancer cells within the cervical carcinoma tissues. Zhongh Guox Xiuha Z 2000; 80: 342-345

20 Song YJ, Yang ZY, Dong JH. Th1/Th2 cells and immune tolerance of transplantation. Shijie Hua Ren Xiuha Z 2001; 9: 794-798

21 Dong H, Zhu G, Tamada K, Chen L. B7-H1, a third member of the B7 family, co-stimulates T cell proliferation and interleukin-10 secretion. Nat Med 1999; 5: 1365-1369

22 Wang S, Zhu G, Chapoval AI, Dong H, Tamada K, Ni J, Chen L. Costimulation of T cells by B7-H2, a B7-like molecule that binds ICOS. Blood 2000; 96: 2808-2813

23 McAdam AJ, Chang TT, Lumelsky AE, Greenfield EA, Boussiotis VA, Duke-Cohan JS, Chernova T, Malenkovich N, Jabs C, Kuchroo VK, Ling V, Collins M, Sharpe AH, Freeman GJ. Mouse inducible costimulatory molecule (ICOS) expression is enhanced by CD28 costimulation and regulate differentiation of CD4+ T cells. J Immunol 2000; 155: 5035-5040

24 Dong C, Juedes AE, Temann UA, Shresta S, Allison JP, Ruddie NH, Flavell RA. ICOS costimulatory receptor is essential for T cell activation and function. Nature 2001; 409: 97-101

25 McAdam AJ, Greenwald RJ, Levin MA, Chernova T, Malenkovich N, Ling V, Freeman GJ. ICOS is critical for CD40-mediated antibody class switching. Nature 2001; 409: 102-105

26 Tafuri A, Shahinian A, Blad F, Yoshinaga SK, Jordan M, Wakeham A, Boucher LM, Bouchard D, Chan VS, Duncan G, Odermatt B, Ho A, Itie A, Horan T, Whorliskey JS, Pawson T, Penninger JM, Ohashi PS, Mak TW. ICOS is essential for effective T helper-cell responses. Nature 2001; 409: 102-105

27 Hultoff A, Dittrich AM, Beier KC, Eljaschewitsch B, Kraft R, Anagnostopoulos I, Krocze RA. ICOS is an inducible T-cell costimulatory structurally and functionally related to CD28. Nature 1999; 397: 263-266

28 Liu X, Bai XF, Wen J, Gao XJ, Liu J, Lu P, Wang Y, Zheng P, Liu Y. B7-H costimulates clonal expansion of, and cognate destruction of tumor cells by, CD8+ T lymphocytes in vivo. J Exp Med 2001; 194: 1339-1348

29 Guo J, Stolina M, Bready JV, Yin S, Horan T, Yoshinaga SK, Senior G. Stimulatory effects of B7-related protein-1 on cellular and humoral immune responses in mice. J Immunol 2001; 166: 5578-5584

30 Riley JL, Blair P, Musser JT, Abe R, Tuszka K, Tsuji T, June CH. ICOS costimulation requires IL-2 and can be prevented by CTLA-4 engagement. J Immunol 2001; 166: 4943-4948

31 Yoshinaga SK, Whorlsie JS, Khare SD, Sarmiento U, Guo J, Horan T, Shi G, Zhang M, Caccia MA, Kohno T, Tafuri-Bladt A, Brandow K, Campbell P, Chang D, Chiu L, Dai T, Duncan G, Elliott GS, Hui A, McCabe SM, Scully S, Shahinian A, Shackle CL, Van G, Mak TW, T-cell co-stimulation through B7RP-1 and ICOS. Nature 1999; 420: 827-832

32 Tamatani T, Tuszka K, Hanazawa-Higuchi N. AILIM/ICOS: a novel lymphocyte adhesion molecule. Int J Immunol 2000; 12: 51-55

33 Mages HWW, Hultoff A, Heuck C, Buchner K, Himmelbauer H, Oliveri F, Krocze RA. Molecular cloning and characterization of murine ICOS and identification of B7H as ICOS ligand. Eur J Immunol 2000; 30: 1040-1047

34 Tamura H, Dong H, Zhu G, Sica GL, Flies DB, Tamada K, Chen L. B7-H1 co-stimulation preferentially enhances CD28-independent T-helper cell function. Blood 2001; 97: 1809-1816

35 Beier KC, Hultoff A, Dittrich AM, Heuck C, Rauch A, Buchner K, Ludewig B, Ochs HD, Mages HWW, Krocze RA. Induction, binding specificity and function of human ICOS. Eur J Immunol 2000; 30: 3707-3717

36 Liu P, Si LS, Li R, Lai BC, Wang YL. Dynamic change of the local immune environment of human gastric carcinoma during the progress of this disease. X' an Yi ke Da xue Xue bao 2001; 22: 408-410

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