Establishment of immortalized mesenchymal stem cells derived from the submandibular glands of tdTomato transgenic mice

SHINJI FURUKAWA1*, YUKINORI KUWAJIMA1*, NAOYUKI CHOSA2, KAZURO SATOH1, MASATO OHTSUKA3, HIROMI MIURA3, MINORU KIMURA3, HIDETOSHI INOKO3, AKIRA ISHISAKI5, AKIRA FUJIMURA4 and HIROYUKI MIURA1

1Division of Orthodontics, Department of Developmental Oral Health Science, Iwate Medical University School of Dentistry, Morioka, Iwate 020-8505; 2Division of Cellular Biosignal Sciences, Department of Biochemistry, Iwate Medical University, Yahaba, Iwate 028-3694; 3Department of Molecular Life Science, Division of Basic Medical Science and Molecular Medicine, Tokai University School of Medicine, Isehara, Kanagawa 259-1193; 4Division of Functional Morphology, Department of Anatomy, Iwate Medical University, Yahaba, Iwate 028-3694, Japan

Received September 13, 2014; Accepted July 29, 2015

DOI: 10.3892/etm.2015.2700

Abstract. Transgenic mice that overexpress the red fluorescent protein tdTomato (tdTomato mice) are well suited for use in regenerative medicine studies. Cultured cells from this murine model exhibit strong red fluorescence, enabling real-time in vivo imaging through the body surface of grafted animals. Mesenchymal stem cells (MSCs) have marked potential for use in cell therapy and regenerative medicine; however, the mechanisms that regulate their dynamics in vivo are poorly understood. In the present study, an MSC line was derived from the submandibular gland fibroblasts of tdTomato mice. The fluorescent signal from this cell line was observed in organs throughout the body, as well as in salivary glands. Primary culture cells derived from the submandibular gland were immortalized with SV40 large T antigen (GManSV cells); these cells exhibited increased migratory ability, as compared with those isolated from the sublingual gland. GManSV cells were tdTomato-positive and exhibited spindle-shaped fibroblastic morphology; they also robustly expressed mouse MSC markers: Stem cell antigen-1 (Sca-1), CD44, and CD90. This cell line retained multipotent stem cell characteristics, as evidenced by its ability to differentiate into both osteogenic and adipogenic lineages. These results indicate that Sca-1+/CD44+/CD90+-GManSV cells may be useful for kinetic studies of submandibular gland-derived MSCs in the context of in vitro co-culture with other types of salivary gland-derived cells. These cells may also be used for in vivo imaging studies, in order to identify novel cell therapy and regenerative medicine for the treatment of salivary gland diseases.

Introduction

Saliva, which is produced by the salivary glands, consists of mucus, various electrolytes, glycoproteins, enzymes, and antibacterial compounds. Therefore, dysfunction or disruption of saliva production presents a significant clinical concern. In particular, hyposalivation, which is a characteristic of xerostomia, may significantly reduce the quality of life of patients (1,2). Hyposalivation is most common in patients with Sjögren's Syndrome (3), ectodermal dysplasias (4-6), head and neck cancer following γ-irradiation therapy (7), or as a side effect of various medications. With regards to therapeutic strategies for the treatment of these diseases, previous studies have adopted regenerative medicine strategies using stem cell sources in order to engineer artificial salivary tissue that can mitigate the effects of xerostomia and hyposalivation (8,9). Removed via lateral parotidectomy, in vitro isolation and characterization of stem cells from the human parotid gland, has previously been achieved (10). Subsequent flow cytometric analysis demonstrated that these stem cells were strongly positive for the classic mesenchymal stem cell (MSC) markers: CD13, CD29, CD44, and CD90, and negative for the key hematopoietic stem cell (HSC) markers, CD34 and CD45. MSCs are multipotent stem cells capable of differentiating into numerous cell lineages, including chondrocytes, adipocytes, osteoblasts, acinar cells, and salivary epithelial cells (11-14). Therefore, MSCs have been highlighted as powerful candidates for experimental investigations (in vitro and in vivo) and clinical treatment due to their anti-inflammatory effects, low immunogenicity and potential to repair damaged tissues (11-15). In this regard, the salivary gland-derived stem cells exhibit MSC-like...
characteristics, as they can be differentiated into adipogenic, osteogenic, and chondrogenic lineages. Therefore, MSCs may exert useful effects for the regeneration and functional restoration of the salivary gland.

Fluorescent transgenic (Tg) mice have been extensively used for analyses of gene function, cellular dynamics, and bioimaging. In addition, Tg mice have been used as in vivo models for the study of numerous diseases (16). Fluorescent proteins (FPs) span the entire color spectrum, and may be used to color-code cells of a specific genotype or phenotype. For example, the behavior of one cell type labeled with green FP (GFP) can be compared with another cell type labeled with red FP (RFP) in vivo. Alternatively, host and donor cells can be differentially labeled with FPs; in this way, a Tg mouse constitutively expressing GFP can be the recipient of transplanted cells that express RFP, in order to visualize the interaction between the two cell types in real time. The current FP color palette includes modified proteins based on Aequorea GFP, as well as various FPs that have been cloned from other marine organisms and improved for live-cell imaging applications via genetic engineering (17,18). The mushroom anemone Discosoma striata was the source of an RFP known asDsRed (19); subsequently, a much brighter dimeric RFP, called tdTomato, was generated (20). This probe is useful for applications that require minimal exposure to excitation illumination to maintain cell viability. The authors of the present study previously generated a series of fluorescent Tg mouse lines using the pronuclear injection-based targeted transgenesis (PITT) method, and demonstrated that these mice exhibit a high level of transgene expression (21). In addition, unlike those developed by random-integration-based transgenesis, the Rosa26CAG::FP mice generated by the PITT method exhibited stable, reproducible and uniform FP expression in various organs, including the liver, kidney and intestine, indicating the potential of these mice for use in chimeric and transplantation analyses (22).

The present study established an MSC line, GManSV, using submandibular gland (Gman) fibroblast-like cells from Rosa26CAG::tdTomato Tg mice (tdTomato mice), which were immortalized by transfection with a plasmid expressing SV40 large T antigen. The aim was to evaluate the applicability of GManSV cells for use in kinetic studies of MSC in vitro and in vivo.

Materials and methods

Rosa26CAG::tdTomato Tg mice (tdTomato mice). The study was approved by the ethics committee of the Animal Studies Committee at Iwate Medical University (nos. 21-039 and 23-075; Center for In Vivo Science, Iwate Medical University, Yahaba, Iwate, Japan). tdTomato mice were maintained by crossbreeding homozygous mutant mice. C57BL/6 wild-type and tdTomato mice were maintained at the Iwate Medical University Center for In Vivo Science under standard conditions (23°C, 12-h light/dark cycle) with sawdust bedding and food (CE-2; CLEA Japan, Inc., Tokyo, Japan) and water ad libitum. The tdTomato mice were generated using the PITT method and the red fluorescence of tdTomato was observed in all organs of the transgenic mice, as described in a previous study (21).

Preparation and observation of tissue sections. The tdTomato mice were sacrificed by excessive inhalation of 90-100% CO2. Whole mice were subsequently cryo-embedded in 5% carboxymethyl cellulose in hexane cooled by liquid nitrogen without fixation and decalcification. The samples were placed in a CM3050S cryostat (cutting edge angle: 7.10°, CT: -22°C, OT: -22°C; Leica Microsystems GmbH, Wetzlar, Germany) and 10-µm frontal serial cryosections were cut using a TC-65 tungsten carbide blade (Leica Microsystems GmbH) according to Kawamoto’s film-transfer method (23,24) with Cryofilm TYPE I-B (Leica Microsystems GmbH). Neighboring serial sections were counterstained with hematoxylin-eosin (Wako Pure Chemical Industries, Inc., Osaka, Japan). The red fluorescence of tdTomato in the prepared sections was observed using a VIOREVO BZ-9000 fluorescence microscope (Keyence Corporation, Osaka, Japan) with a Texas Red fluorescence filter.

Isolation of primary culture cells from salivary glands. After the capsule covering the salivary gland of 1-week-old tdTomato mice was removed, the gland body was extracted. The GMan and sublingual gland (GLin) were segmented as a tissue mass using scissors. The glands were allowed to attach to the bottom of a 35 mm plastic cell culture dish, and the cells were cultured in Dulbecco's modified Eagle's medium (DMEM; Sigma-Aldrich, St. Louis, MO, USA) supplemented with 10% fetal bovine serum (FBS; HyClone, GE Healthcare Life Sciences, Logan, UT, USA) for 1 week at 37°C in a humidified atmosphere containing 5% CO2. Adherent cells that grew out from the tissue mass were then placed in 90 mm culture dishes and cultured in DMEM supplemented with 10% FBS. Once the culture reached 80% confluence, the cells were replated.

Measurement of cell migration in salivary gland-derived primary cells. Single cell migration was evaluated by measuring the distance moved by the cell in a time-lapse period using a VIOREVO BZ-9000 fluorescence microscope (Keyence Corporation) with a Texas Red fluorescence filter, in addition to an IX70 phase contrast microscope (Olympus Corporation, Tokyo, Japan). Images were captured from ~75 primary culture cells derived from GLin or GMan 200 times every 10 min for up to 33 h. Two-dimensional moving distances of the cells were measured using BZ-II image analysis software (Keyence Corporation).

Establishment of immortalized GMan cells. The expanded cells (1.0x10^5) derived from the GMan of tdTomato mice were transfected with a pBABE-puro-SV40LT plasmid containing a puromycin resistance gene (Addgene Inc., Cambridge, MA, USA) using Lipofectamine LTX (Invitrogen Life Technologies, Carlsbad, CA, USA) for 6 h at 37°C in 5% CO2, according to the manufacturer's instructions. The cells were exposed to DMEM supplemented with 10% FBS and 1 μg/ml puromycin (Invitrogen Life Technologies) for 12-15 days. The surviving cells were trypsinized (Invitrogen Life Technologies) and allowed to grow in 90 mm culture dishes.

Detection of MSC markers by flow cytometry. Following selection with puromycin, GMan-derived cells (1.0x10^5) were suspended in phosphate-buffered saline supplemented
with 0.5% FBS and 2 mM EDTA. The cells were incubated with fluorescein isothiocyanate (FITC)-conjugated anti-mouse Sca-1 (1:10; 130-102-297), anti-mouse CD44 (1:10; 130-102-511), or anti-mouse CD90 (1:10; 130-102-452) antibodies for 1 h at 4°C in the dark. All FITC-conjugated antibodies were purchased from Miltenyi Biotec GmbH (Bergisch Gladbach, Germany). Image acquisition was performed with an EPICS XL ADC system (Beckman Coulter, Inc., Brea, CA, USA).

Osteogenic and adipogenic differentiation. The in vitro differentiation method used here was reported in our previous study (25). Briefly, to investigate osteogenic differentiation, bone matrix mineralization was evaluated using 1% Alizarin Red S (Sigma-Aldrich) staining. To investigate adipogenic differentiation, lipid droplets were stained with 0.18% Oil Red O (Sigma-Aldrich).

Statistical analysis. The experiments were repeated at least three times and representative images or data are presented. Statistical data are presented as the mean ± standard deviation. Differences between samples were statistically analyzed using paired two-tailed Student's t-tests. P<0.05 was considered to indicate a statistically significant difference.

Results

Detection of tdTomato fluorescence in salivary glands. The red fluorescence of tdTomato was detected in tissue sections prepared from tdTomato mice. As shown in Fig. 1, tdTomato fluorescence was detected ubiquitously in all organs (Fig. 1A). In addition, fluorescence was detected in the cells that constitute the salivary gland tissues (Fig. 1B). There was no difference in fluorescence intensity between the GLin and GMan glands.

Evaluation of the migratory ability of salivary gland-derived primary cells. The present study subsequently isolated primary cultured GLin and GMan cells from the tdTomato mice. As shown in Fig. 2, the cells that grew out from the tissue masses possessed a fibroblast-like morphology. Notably, although the morphology of GLin and GMan cells was similar (Fig. 2A), the migratory ability of GMan cells was significantly higher, as compared with that of GLin cells (Fig. 2B). Usually, MSCs...
maintain a high migratory ability (26-28). Therefore, the GMan cells were selected for the establishment of the MSC line.

Establishment of an MSC line from GMan cells. GMan cells were immortalized using SV40 large T antigen (SV40LT) in order to produce GManSV cells. As shown in Fig. 3,
these cells had a spindle-shaped fibroblastic morphology (Fig. 3A) and exhibited a strong expression of tdTomato, as determined by fluorescence imaging (Fig. 3B). Furthermore, the expression of mouse MSC markers and the differentiation potential of the cell line were determined. As shown in Fig. 4, Sca-1, one of the most functionally critical MSC markers in mice, was strongly expressed in GManSV cells (Fig. 4A). In addition, CD90 and CD44 were also highly expressed (Fig. 4B and C). In order to determine whether the cell line possessed multipotent properties, the osteogenic and adipogenic differentiation potentials of the GManSV cells were evaluated. Alizarin Red S staining demonstrated that GManSV cells could differentiate into osteoblasts (Fig. 5A). In addition, Oil Red O staining demonstrated the cells could differentiate into adipocytes (Fig. 5B). These results strongly suggest that GManSV cells retain MSC-like multipotency.

**Discussion**

The present study established an MSC line from murine GMan cells overexpressing the RFP tdTomato, by immortalization of the cells with SV40LT. The SV40LT protein was selected based on its role in the infection of both permissive and non-permissive cells, leading to the production of progeny virions and malignant transformation, respectively (29,30). In addition, SV40LT has been used in cancer research for immortalization and transformation of cells (31), as primary cells are non-permissive for SV40LT, and infection with wild-type SV40LT leads to immortalization and transformation of a small percentage of infected cells. Such transformation approaches usually involve the overexpression of oncogenes and/or inactivation of tumor suppressor genes. Commonly used oncogenes include K-Ras, c-myc, cyclin-dependent kinase 4, cyclin D1, Bmi-1, and human papillomavirus 16 E6/E7, and frequently
inactivated tumor suppressor genes include p53, retinoblastoma, and p16INK. In contrast to the previously mentioned oncogenes, SV40LT expression generally leads to immortalization, but not transformation (32-35). Therefore, SV40 is often used in order to avoid the excessive cellular changes that are associated with full-blown transformation.

The salivary glands of the majority of mammals consist of three main cell types: Serous-producing acinar cells, mucus-producing acinar cells, and myoepithelial cells (36). Serous-producing acinar cells possess a pyramidal morphology and join together to form spheroidal shapes; whereas mucus-producing acinar cells are cuboidal in shape and group together to form tubules. Myoepithelial cells are located near ductal openings and are associated with the contraction of ducts, in order to facilitate salivary secretion (37). Furthermore, in normal salivary glands, the mesenchymal tissue between the acinar-ductal epithelial structures (38) may also contain stem cells of mesenchymal origin, as detected in various other organs, including the bone marrow (39). Since adult stem cells are generally restricted to cell lineages of the body part of origin, previous studies have aimed to use salivary gland-derived stem cells, in order to reduce hyposalivation and restore natural function (37,40,41). Stem cells have been isolated and characterized from major salivary glands of humans and animals (42-44). The injection of these cells into the major salivary gland parenchyma has been proposed as an ideal therapeutic strategy for the restoration of irreversibly damaged gland tissue in patients with head and neck cancer who have received radiotherapy (45). To date, numerous approaches have been taken; some groups have harvested cells from the parotid gland (10) and GMan (45), whereas others have harvested cells from a combination of both glands (46), and by co-culture (47). In particular, stem cells isolated from a combination of the human parotid gland and GMan were demonstrated to partially restore salivary gland function in radiation-damaged rat salivary glands in vivo (46). Furthermore, previous in vitro studies have demonstrated that salivary gland-derived stem cells differentiate into MSC lineages, express MSC markers (CD44, CD49f, CD90, and CD105) in lieu of HSC markers (CD34 and CD45), and can differentiate into amylase-expressing cells (48-50).

Successful isolation of MSCs from various organs can be determined by a combination of criteria including morphology, surface marker phenotype, and differentiation potential (51,52). A defined panel of markers has been suggested in humans, however there are currently no standard criteria that have been proposed in mice. However, stable identification and isolation of murine MSCs using a lineage-, and Sca-1+ phenotype have been reported (53,54). CD90 may function as an activator of stem cell differentiation (55), and CD44 is a marker that is commonly detected in human MSCs (56). In the present study, the MSC line derived from GManSV cells were shown to express Sca-1, CD44, and CD90 on the cell surface. Furthermore, the GManSV cells retained the multipotent characteristics of MSCs, as they could differentiate into both osteogenic and adipogenic lineages. Therefore, these findings suggested that GManSV cells may be used as a GMan-derived MSC line for research focused on various regenerative medicine strategies.

To the best of our knowledge, the present study is the first to report a stable cell line that has been derived from tdTomato mice. The tdTomato probe is useful for applications that require minimal exposure to excitation illumination, in order to maintain cell viability. By taking advantage of this attribute, GManSV cells may be adopted as useful tools for kinetic studies of GMan-derived MSCs for in vitro co-culture systems with other types of salivary gland-derived cells. These cells may be well suited for in vivo imaging studies of cell therapy and regenerative medicine focused on salivary gland diseases.

Acknowledgements

The present study was supported in part by JSPS KAKENHI (grant no. 25463053 to Dr Naoyuki Chosa and grant no. 23592896 to Professor Akira Ishisaki); and a Grant-in-aid for Strategic Medical Science Research Centre from the Ministry of Education, Culture, Sports, Science and Technology of Japan, 2010-2014.

References

1. Thomson WM, Lawrence HP, Broadert JM and Poulton R: The impact of xerostomia on oral-health-related quality of life among younger adults. Health Qual Life Outcomes 4: 86, 2006.
2. Berk L: Systemic pilocarpine for treatment of xerostomia. Expert Opin Drug Metab Toxicol 4: 1333-1340, 2008.
3. Philipp SR, Matteson EL, Jacobsson LT, Martens PB, Melton LJ III, O’Fallon WM and Fox PC: Incidence of physician-diagnosed primary Sjögren syndrome in residents of Olmsted County, Minnesota. Mayo Clin Proc 76: 593-599, 2001.
4. Pinheiro M and Freire-Maia N: Ectodermal dysplasias: A clinical classification and a causal review. Am J Med Genet 53: 153-162, 1994.
5. Nordgarden H, Storhaug K, Lyngstadfaa SP and Jensen JL: Salivary gland function in persons with ectodermal dysplasias. Eur J Oral Sci 111: 371-376, 2003.
6. Clauw F, Manière MC, Obry F, Wallmann E, Hadj-Rabia S, Bodemer C, Alembyk Y, Lesot H and Schmittbuhl M: Dento-craniofacial phenotypes and underlying molecular mechanisms in hypohidrotic ectodermal dysplasia (HED): A review. J Dent Res 87: 1089-1099, 2008.
7. Jensen SB, Pedersen AM, Vissink A, Andersen E, Brown CG, Davies AN, Dutilb J, Fulton JS, Jankovic L, Lopes NN, et al: Salivary Gland Hypofunction/Xerostomia Section, Oral Care Study Group, Multinational Association of Supportive Care in Cancer (MASCOC)/International Society of Oral Oncology (ISOO): A systematic review of salivary gland hypofunction and xerostomia induced by cancer therapies: Prevalence, severity and impact on quality of life. Support Care Cancer 18: 1039-1060, 2010.
8. Nelson J, Manzella K and Baker OF: Current cell models for bioengineering a salivary gland: A mini-review of emerging technologies. Oral Dis 19: 236-244, 2013.
9. Ogawa M, Oshima M, Imamura A, Sekin Y, Ishida K, Yamashita K, Nakajima K, Hirayama M, Tachikawa T and Tsuki T: Functional salivary gland regeneration by transplantation of a bioengineered organ. Nat Commun 4: 2498, 2013.
10. Rotter N, Oder J, Schlenke P, Lindner U, Bühnens F, Kramer J, Rohwedel J, Huss K, Brandau S, Wollenberg B and Lang S: Isolation and characterization of adult stem cells from human salivary glands. Stem Cells Dev 17: 509-518, 2008.
11. Xu J, Wang D, Liu D, Fan Z, Zhang H, Liu O, Ding G, Gao R, Zhang C, Ding Y, et al: Allogeneic mesenchymal stem cell treatment alleviates experimental and clinical Sjögren syndrome. Blood 120: 3142-3151, 2012.
12. Sumita Y, Liu Y, Khalili S, Maria OM, Xia D, Key S, Cotrip AM, Mezey E and Tran SD: Bone marrow-derived cells rescue salivary gland function in mice with head and neck irradiation. Int J Biochem Cell Biol 43: 80-87, 2011.
13. Liu CY, Chang FH, Chen CY, Huang CY, Hu FC, Huang WK, Ju SS and Chen MH: Cell therapy for salivary gland regeneration. J Dent Res 90: 341-346, 2011.
Nitta M, Katabuchi H, Ohtake H, Tashiro H, Yamaizumi M and SV40-transformed human and monkey cells in immunodeficient mice. Jpn J Cancer Res 91: 1346‑1353, 2000.

Khaliti S, Liu Y, Kornete M, Roescher N, Kodama S, Peterson A, Piccirillo CA and Tran SD: Mesenchymal stem cells improve survival and reduce lymphocytic infiltrates in mice with Sjögren's‑like disease. PLoS One 7: e38615, 2012.

Weigert R, Port‑Shliom N and Amornphimoltham P: Imaging cell‑bioluminescence in live animals: Ready for prime time. J Cell Biol 201: 969‑979, 2013.

Shaner NC, Patterson GH and Davidson MW: Advances in fluorescence protein technology. J Cell Sci 120: 4247‑4260, 2007.

Labas YA, Gurskaya NG, Yanushevich YG, Fradkov AF, Lukyanov KA, Lukyanov SA and Matz MV: Diversity and evolution of the green fluorescence protein family. Proc Natl Acad Sci USA 99: 4256‑4261, 2002.

Matz MV, Fradkov AF, Labas YA, Savitsky AP, Zaraisky AG, Markelov ML and Lukyanov SA: Fluorescent proteins from nonbacterioluminescent Anthozoa species. Nat Biotechnol 17: 969‑973, 1999.

Day RN and Schauefele F: Fluorescent protein tools for studying protein dynamics in living cells: A review. J Biomed Opt 13: 031202, 2008.

Ohtsuka M, Ogawa S, Miura H, Mizutani A, Warita T, Sato A, Hiro K, Hozumi K, Sato T, Tanaka M, et al: Pronuclear injection‑based mouse targeted transgenesis for reproducible and highly efficient transgene expression. Nucleic Acids Res 38: e198, 2010.

Ohtsuka M, Miura H, Gurumurthy CB, Kimura M, Inoko H, Yoshimura S and Sato M: Fluorescent transgenic mice suitable for multi‑color aggregation chimera studies. Cell Tissue Res 350: 251‑260, 2012.

Kawamoto T: Light microscopic autofluorography for study of early changes in the distribution of water‑soluble materials. Histochem Cytochem 38: 1805‑1814, 1990.

Kawamoto T and Shimizu M: A method for preparing 2 to 50‑micron‑thick fresh‑frozen sections of large samples and undecalciﬁed hard tissues. Histochem Cell Biol 113: 331‑339, 2000.

Aomatsu E, Takahashi N, Sawada S, Okubo N, Hasegawa T, Taira M, Miura H, Ishisaki A and Chosa N: Novel SCRG1/BST1 axis regulates self‑renewal, migration, and osteogenic differentiation potential in mesenchymal stem cells. Sci Rep 4: 3652, 2014.

Chamberlain G, Fox J, Ashton B and Middleton J: Concise review: Mesenchymal stem cell‑mediated therapy: Their phenotype, differentiation capacity, immunological features, and potential for homeostatic tissue repair ‑ current views. Stem Cells 25: 2537‑2545, 2007.

Belema‑Bedada F, Uchida S, Martire A, Kostin S and Braun T: Efficient homing of multipotent adult mesenchymal stem cells depends on FROUNT‑mediated clustering of CCR2. Cell Stem Cell 2: 566‑575, 2008.

Sohni A and Pelle JC: Mesenchymal stem cells migration homing and tracking. Stem Cells Int 2013: 130736, 2013.

Borowicz JA, Dean FB, Bullock PA and Hurwitz J: Binding and unwinding ‑ how T antigen engages the SV40 origin of DNA replication. Cell 60: 181‑184, 1990.

Prives C: The replication functions of SV40 T antigen are regulated by phosphorylation. Cell 61: 735‑738, 1990.

Arlt VM, Stiborová M, vom Brocke J, Simões ML, Lord GM, Prives C: The replication functions of SV40 T antigen are essential for efficient homing of multipotent adult mesenchymal stromal cells. Cell Stem Cell 9: 176‑184, 2011.

Lombaert IM, Brunsting JF, Wierringa PK, Faber H, Stokman MA, Kok T, Verfaillie CM: Mesenchymal stem cell‑mediated salivary gland repair: A review: Mesenchymal stem cell‑mediated salivary gland repair. BMJ Open 4: e004579, 2014.

Lim JY, Yi T, Lee S, Kim J, Kim SN, Song SU and Kim YM: Establishment and characterization of mesenchymal stem‑cell‑derived salivary gland cells differentiated from mouse early ES cells in vitro. Hum Cell 26: 80‑90, 2013.

Lin JY, Yi T, Lee S, Kim J, Kim SN, Song SU and Kim YM: Establishment and characterization of mesenchymal stem cell‑derived salivary gland cells differentiated from mouse early ES cells in vitro. Hum Cell 26: 80‑90, 2013.