**Abstract**  The source of Parkinson disease-linked α-synuclein (αSyn) in human cerebrospinal fluid (CSF) remains unknown. We decided to measure the concentration of αSyn and its gradient in human CSF specimens and compared it with serum to explore its origin. We correlated αSyn concentrations in CSF versus serum (Q_αSyn) to the albumin quotient (Q_albumin) to evaluate its relation to blood–CSF barrier function. We also compared αSyn with several other CSF constituents of either central or peripheral sources (or both) including albumin, neuron-specific enolase, β-trace protein and total protein content. Finally, we examined whether αSyn is present within the structures of the choroid plexus (CP). We observed that Q_αSyn did not rise or fall with Q_albumin values, a relative measure of blood–CSF barrier integrity. In our CSF gradient analyses, αSyn levels decreased slightly from rostral to caudal fractions, in parallel to the recorded changes for neuron-specific enolase; the opposite trend was recorded for total protein, albumin and β-trace protein. The latter showed higher concentrations in caudal CSF fractions due to the diffusion-mediated transfer of proteins from blood and leptomeninges into CSF in the lower regions of the spine. In postmortem sections of human brain, we detected highly variable αSyn reactivity within the epithelial cell layer of CP in patients diagnosed with a range of neurological diseases; however, in sections of mice that express only human SNCA alleles (and in those without any Snca gene expression), we detected no αSyn signal in the epithelial cells of the CP. We conclude from these complementary results that despite its higher levels in peripheral blood products, neurons of the brain and spinal cord represent the principal source of αSyn in human CSF.

**Keywords**  α-Synuclein · Cerebrospinal fluid · Blood–CSF barrier · Biomarker · Choroid plexus

**Introduction**

The pathological hallmarks of Parkinson disease (PD) include progressive cell loss of selective nuclei in the nervous system and the formation of intracellular aggregates that contain insoluble, wild-type α-synuclein (αSyn) (Spillantini et al. 1997). The clinical phenotype of autosomal dominant PD parallels αSyn gene (SNCA) dosage with a more severe phenotype observed in the cases of locus multiplication (Singleton et al. 2003). The quantification of the 140 amino acid-long, 16–17 kDa αSyn protein in biological fluids has been proposed as a biomarker candidate (Mollenhauer et al. 2010). αSyn has been shown...
to be present in cerebrospinal fluid (CSF) in low picogram/microliter amounts; its levels in serum and plasma are up to tenfold and in whole blood up to 10,000-fold higher (Mollenhauer et al. 2010) due to the fact that aSyn is also abundantly expressed in the hematopoietic system (Scherzer et al. 2008).

The production and hydrodynamics of human CSF are complex and vary considerably depending on the level within the neuroaxis. The three main sources for CSF constituents comprise: one, the metabolism of neural cells in the central nervous system (CNS); two, diffusion of constituents from circulating blood into CSF [even in the absence of any trauma (Hong et al. 2010; Mollenhauer et al. 2011)]; and three, the secretion by the epithelial cells that reside within the brain’s choroid plexus (CP) (Chodobski and Szmydynger-Chodobska 2001). CSF volume and its continuous renewal are predominantly regulated by CP activity. An estimated 80% of all CSF proteins are blood-derived (with albumin being the most abundant); their concentrations in the nervous system increase in the subarachnoid space from rostral to caudal levels due to diffusion gradients that emanate from blood vessels intersecting with the flow of CSF (Reiber 2003). A rise from rostrum to cauda is also observed for certain constituents that are secreted by leptomeningeal structures (e.g., β-trace protein) (Reiber 2003). In the case of abnormally low CSF flow rates, which result in increasing concentrations of blood-derived proteins in lumbar CSF fractions, the concentration of brain-derived proteins [e.g., neuronspecific enolase (NSE)] usually remains constant (Reiber 2003). Because albumin is solely synthesized in the liver, the quotient of its CSF-to-serum concentrations (referred to as Q albumin) reflects an indirect measure of blood–CSF barrier function in vivo.

This study was designed to investigate whether CSF aSyn is mainly brain or blood derived. To this end, we compared aSyn concentrations in CSF versus serum (Q aSyn) with Q albumin in 42 patients with various neurological diseases. Next, we quantified aSyn levels in patients with normal blood–CSF barrier function and compared them to specific brain-derived (e.g., NSE), blood-derived (e.g., albumin), and leptomeninges-derived (e.g., β-trace) proteins in serially collected CSF samples, to examine their rostro-caudal concentration gradients. Furthermore, we also probed whether CP tissue contained aSyn reactivity.

**Methods**

We collected CSF and serum from patients with different neurological conditions. These 42 participants underwent lumbar puncture (LP) for routine diagnostic purposes (Table 1) to determine the presence (or absence) of acute inflammatory disease, acute hemorrhage and suspected Lyme disease (n = 31); to exclude chronic inflammatory diseases (n = 3); to determine carcinomatosis of the leptomeninges in the case of a known primary tumor (n = 4), and to carry out CSF volume reduction in patients with normal pressure hydrocephalus (NPH; n = 4). All study participants were enrolled at the Department of Neurology at Goettingen University, Germany. Samples were collected and processed as published (Mollenhauer et al. 2010). We further examined samples that featured: a normal (<5/μl) white blood cell count; the absence of intrathecal immunoglobulin production (as determined by the absence of oligoclonal bands); normal CSF tau protein, and no LP-induced contamination by blood (determined by a red blood cell count of <50/μl). CSF aSyn and serum aSyn (diluted at 1:15) concentrations were measured by enzyme-linked immunosorbent assay (ELISA) as published (Mollenhauer et al. 2011).

In a second set of study participants we collected CSF from five patients at the Paracelsus-Elena-Klinik, Kassel, Germany. All patients suffered from NPH as determined by magnetic resonance imaging and according to the established clinical criteria (Relkin et al. 2005) (Table 1). We performed volume reduction by LP on each patient and serially collected seven CSF fractions (5 ml each) that were processed as published (Mollenhauer et al. 2010). LP was carried out at the L2/L3 level in the sitting position, between 8 and 9 a.m. with the patient having fastened for 12 h. In each fraction we analyzed CSF aSyn (Mollenhauer et al. 2011), NSE (Schaarschmidt et al. 1994) and β-trace protein (Tumani et al. 1994) and β-trace protein (Tumani et al. 1998), as described. Total protein and albumin levels were measured by nephelometry (Dade Behring/Siemens Healthcare Diagnostics).

**Table 1** Demographic data of enrolled patients of both cohorts

|             | Set 1     | Set 2     |
|-------------|-----------|-----------|
| N           | 42        | 5         |
| Sex (male/total) | 0.5       | 0.6       |
| Age (years)  | 58 ± 21   | 76 ± 4    |
| Mean ± standard deviation (range) | (19–93)   | (69–79)   |

**Immunohistochemistry of postmortem** brain sections containing CP structures was carried out following formalin fixation and paraffin embedding with anti-aSyn antibodies, as published (Louis et al. 2005; Schlossmacher et al. 2002). In parallel, sections from mouse brain transgenic for four Ala53Thr-containing human SNCA alleles (but no mouse Snca) and Snca-null mice without any endogenous Snca and no transgene expression [engineered as described (Kuo et al. 2011)] were processed by the same protocol. Polyclonal (hSA5.1; Open Biosystems Inc) and...
monoclonal (12–1; Epitomics Inc) antibodies to wild-type, full-length human aSyn were raised in goat and rabbit, respectively, purified and characterized (Cullen et al. 2009; Mollenhauer et al. 2008).

The study was carried out in accordance with the Declaration of Helsinki and with informed written consent provided by all patients or their next of kin. The study was approved by the ethics committee at the University of Goettingen (Germany).

Analysis

Possible correlations between the concentrations of CSF/serum aSyn to CSF/serum albumin (i.e., Qalbumin) were analyzed according to Pearson. Results for aSyn, NSE, β-trace, albumin and total protein values in CSF are demonstrated in Table 2 and Fig. 1. Statistical analyses (non-parametric) of the data were performed to assess the characteristics in CSF vials 5–7 (cisternal fractions) and 1–4 (lumbar fractions) for each described variable using the trend test from Page (Page 1963). The Page’s L trend test examined whether the levels are ordered in a specific order/sequence. For all tests, the α-level of significance was set at 5 %. Furthermore, effect sizes (Hedge’s g) were calculated (Ray and Shadish 1996). A correction formula for small sample bias was also applied (Hedges and Olkin 1985).

Results

CSF α-synuclein in subjects with known blood–CSF barrier function

CSF specimens analyzed from 42 patients showed a mean Qalbumin of 8.90 ± 7.7 × 10−3 (mean ± standard deviation; SD) (range 1.8–37.2 × 10−3). CSF aSyn levels ranged from 1.29 to 2.97 pg/μl (mean ± SD; 1.99 ± 0.39). Serum aSyn levels ranged from 14.33 to 67.47 pg/μl (mean 29.54 ± 10.79), and the mean CSF/serum aSyn (QaSyn) was calculated as 0.08 ± 0.032. The coefficients of variation (CV) for aSyn values measured 19.6% in CSF, 36.5% in serum, and the CV for QaSyn was 40%. Importantly, when examining QaSyn together with Qalbumin values, we recorded no significant correlation (−0.109; p = 0.493; Fig. 2).

CSF α-synuclein and other protein values from rostro-caudal fractions

CSF aSyn levels across all fractions examined ranged from 1.17 to 2.64 pg/μl (mean 1.70 ± 0.36) in specimens from five NPH patients that were analyzed. Qalbumin in these cases ranged between 5.4 and 7.8 × 10−3. Across the 35 ml of CSF collected in our NPH patients, aSyn concentrations showed a slight increase that initially paralleled the proximity of the sample collected toward the brain (and as reflected by increasing fraction numbers in the first 20 ml collected), which represented a trend without statistical significance (L = 130; p > 0.05). The levels of exclusively neuron-derived NSE remained stable throughout (L = 134; p > 0.05; Table 2; Fig. 1). In contrast, blood-derived albumin (L = 145.5; p < 0.05) and total protein (L = 139; p < 0.05) decrease significantly in the rostral fractions (#5 to #7, i.e., in the last 15 ml of each LP volume collected; Table 2; Fig. 1) compared to caudal fractions (#1 to #4, i.e., corresponding to the first 20 ml collected), as expected from the published literature (Sullivan et al. 2006). Albeit to a lesser extent, leptomeninges-derived β-trace protein also showed a mild decrease in its concentration with progression from the lumbar region toward the cranial compartment (L = 138.5; p < 0.05). Figure 1 shows the median values for each fraction and for each protein.

Table 2 Quantification results including mean levels in seven fractions (F1 lower F7 upper position) for CSF α-synuclein (aSyn), total protein, albumin, neuron specific enolase (NSE) and β-trace protein

| Fractions | α-Synuclein (pg/μl) | Total protein (mg/l) | Albumin (mg/l) | NSE (mg/l) | β-Trace protein (mg/l) |
|-----------|---------------------|----------------------|----------------|------------|------------------------|
| F7        | 1.92                | 387                  | 224            | 9.49       | 12.66                  |
| F6        | 1.88                | 398                  | 226            | 9.21       | 12.25                  |
| F5        | 1.91                | 436                  | 255            | 9.48       | 12.86                  |
| F4        | 1.91                | 449                  | 266            | 9.30       | 13.68                  |
| F3        | 1.79                | 468                  | 290            | 8.89       | 14.20                  |
| F2        | 1.69                | 526                  | 321            | 9.09       | 13.94                  |
| F1        | 1.66                | 634                  | 401            | 8.41       | 14.02                  |
| Rostrocaudal ratio | 1.16             | 0.61                | 0.56           | 1.13       | 0.90                   |

Choroid plexus epithelium is an unlikely source of α-synuclein

The intra-ventricular network of the CP represents the predominant site for CSF volume production in vivo. To also explore CP as a possible source for CSF aSyn, we first examined postmortem human brain sections by routine
immunohistochemistry (IHC). We unexpectedly detected highly variable αSyn reactivity within the epithelial cell layer of human CP (Fig. 3a). When comparing sections from several diagnostic categories, we saw considerable differences in the staining intensity of adult CP epithelium among cases with synucleinopathy [PD, dementia with Lewy bodies (DLB)], progressive supranuclear palsy, Alzheimer’s and Huntington disease and aged control subjects (Fig. 3a; data not shown), but did not record a correlation between αSyn reactivity in CP epithelium and the final diagnosis. To determine whether the αSyn signal in postmortem CP was reflective of physiological metabolism in vivo, we decided to examine brain sections of “humanized” SNCA-transgenic mice (Kuo et al. 2011) versus animals deficient in murine (and human) αSyn using the same IHC protocol. There, we detected no αSyn reactivity in the CP epithelium-containing sections of SNCA-transgenic mice, wild-type animals and Snca-null mice (Fig. 3b–d; data not shown). In contrast, CNS structures including neurons and presynaptic terminals (as well as intravascular erythrocytes) showed strong αSyn immunoreactivity, as expected from the corresponding genotype (Fig. 3b, c; data not shown). We concluded from these findings that CP, a highly vascularized organ within the brain, represents an unlikely source for the constitutive release of αSyn into CSF in vivo; however, a possible uptake of αSyn from CSF by CP cells postmortem could not be excluded.

Discussion

To elucidate the source of αSyn in CSF, we measured its levels in CSF and serum from a cohort with variable Qalbumin values (Blennow et al. 1993; Reiber 2003) and in a second, small cohort of NPH patients, where we measured...
CSF αSyn together with several proteins from different CNS sources. Our collective results suggest that CSF αSyn is predominantly derived from structures and functions associated with the brain rather than from peripheral blood products.

CSF αSyn has been discussed as a possible biomarker for αSyn-related disorders such as PD, DLB and multiple system atrophy (MSA) (Mollenhauer et al. 2011) and reviewed in (Mollenhauer et al. 2010); its presence in normal human CSF has been previously confirmed by mass spectrometry (Mollenhauer et al. 2008). The majority (but not all) of the CSF αSyn quantification studies conducted over the last 6 years have shown a decrease of total αSyn levels in CSF from PD, DLB and MSA patients, as measured by ELISA and Luminex platforms [reviewed in: (Mollenhauer et al. 2010, 2011)]. These biomarker exploration efforts raised—among other issues—the question of the possible sources of CSF αSyn, which theoretically include the following: (1) physiological exocytosis by neural cells of the brain and spinal cord (Lee et al. 2005; Mollenhauer et al. 2008); (2) secretion by the epithelial cells of the CP; (3) release by cells of the ependymal lining of the brain’s ventricular system; (4) secretion by endothelial cells of the brain’s vasculature; (5) normal filtration of peripheral plasma through the brain’s capillary system (or following a compromise in the integrity of the blood–CSF barrier); (6) diffusion and/or entry of blood products in anatomically privileged sites without effective blood–CSF barrier; and lastly, (7) physiological attrition of neural cells as well as neuronal injury in the CNS.

Because αSyn is highly abundant in blood cells, especially in erythrocytes, discrepant findings across cross-sectional studies have raised the possibility of divergence due to artificial blood contamination, which occurs in 10–20% of routine LPs (Mollenhauer et al. 2010, 2011). However, in most (but not all) published CSF αSyn quantification studies, the possibility of blood contamination as the source of CSF αSyn has been addressed and largely excluded [reviewed in (Mollenhauer et al. 2010)].

In examining the influence of the blood–brain barrier on CSF and serum αSyn levels from neurological controls, we found no correlation between the concentration of CSF αSyn and Qalbumin. Therefore, we concluded that CSF αSyn levels were not controlled by the blood–CSF
barrier under physiological conditions. However, larger cohorts comprising patients with various inflammatory conditions will need to be examined (e.g., multiple sclerosis) to discern the effect of aSyn distribution under pathological blood–CSF barrier conditions.

Due to the protective role of the barrier function on CNS integrity, we also calculated the coefficients of variation (CV) as a reflection of biological variability. The CV value for aSyn in CSF was smaller than that for serum aSyn. The latter observation provided independent (albeit indirect) support for our conclusion that under normal conditions the majority of aSyn species detected in CSF was unlikely to be blood borne; if it would have been predominantly derived from blood, the CV of CSF aSyn values would have been expected to be at least equal—if not larger—than those of serum aSyn. Another possible explanation for the different CV values is that our assay could have captured CSF aSyn more readily than serum aSyn (for example due to protein complex formation and thus altered antibody accessibility), but this scenario seems unlikely given the significantly larger amounts of aSyn recorded by our assay in all serum samples than in CSF (Mollenhauer et al. 2010, 2011).

To further examine aSyn levels along the rostro-caudal flow of CSF, we collected a total of 35 ml in seven serial fractions in all NPH patients. The lumbosacral CSF volume is highly variable among individual patients and is estimated to contain 35.8 ± 10.9 ml with a total range from 10.6 to 61.3 ml in adult humans (Sullivan et al. 2006). With the LP performed at the L2/L3 level in all study participants, we inferred that fractions #1–4 contained CSF from the caudal subarachnoidal space where it supports spinal cord suspension (and function), whereas fractions #5–7 represented CSF derived from space closer to (and above) the craniovertebral juncture following direct contact with the brain. Our analysis revealed the expected rostro-caudal increase of plasma-derived proteins (i.e., total protein and albumin), which is thought to reflect a longer transit time of CSF to the lumbar spine region and a higher proportion of diffusion-mediated transfer of proteins from blood to CSF (Reiber 2003). In our study, β-trace protein concentrations also increased in a rostro-caudal gradient, as expected from studies that simultaneously analyzed ventricular and lumbar CSF samples (Reiber 2001, 2003); β-trace protein is mainly released by leptomeningeal structures. In contrast, the neuron-derived protein NSE showed the expected rostro-caudal stability in our hands, possibly even a slight reduction for the concentrations recorded in lumbar fractions #1–4. These collective findings strongly supported the conclusions that these seven fractions indeed represented different levels of CSF collected from the lumbar as well as cisternal regions of subarachnoid space. The concentration behavior of our analyte of interest, total aSyn, behaved most closely to that of NSE en route from the rostrum of the brain to the lumbosacral space. This decreasing rostro-caudal gradient is, therefore, in support of a concept that sees the brain as the most important source of CSF aSyn followed by nervous system structures below the medulla.

A decreasing rostro-caudal gradient for CSF aSyn concentrations was also reported by Hong and colleagues, who analyzed three fractions in 45 control subjects showing a trend for decreased CSF aSyn in lower # fractions (Hong et al. 2010). Although the Hong et al. CSF samples were not centrifuged after their collection, therefore, raising the theoretical possibility of blood and epithelial cell contamination (which could have influenced their findings), our results confirm and expand the conclusions reached by these and other authors exploring the source of extracellular aSyn in human brain (Emmanouilidou et al. 2011).

Our study on CSF aSyn is limited by its small sample size and a selection bias towards patients with non-neurodegenerative conditions. In future studies, patients with neurodegenerative and inflammatory illnesses as well as medically healthy participants have to be analyzed. Nevertheless, our results support the concept of a source of CSF aSyn that is predominantly (but not exclusively) associated with nervous system structure and function under physiological conditions, and is less likely to be blood borne. Our results do not exclude the possibility that this distribution is markedly altered in diseases associated with significant neuronal lysis (e.g., due to prion disease) (Mollenhauer et al. 2008).

Our finding of variable immunoreactivities for aSyn within the epithelium of CP in human brain (but not mouse brain) sections could indicate a relevant site for aSyn uptake postmortem from circulating CSF [as reported for example for amyloid-β-protein (Crossgrove et al. 2005) (Wolburg and Paulus 2010)] in some patients. However, our findings in genetically engineered mice make de novo gene transcription in CP and secretion (or filtration of aSyn from plasma into CSF by CP epithelial cells) very unlikely. Alternatively, technical reasons (such as those related to possible antibody cross-reactivity with unknown antigens) when processing human versus mouse brain could also explain the observed species difference. Lastly, in future studies we will revisit whether this variable immunoreactivity for aSyn in human CP may be related to the final diagnosis of the patient. To date, we have not yet observed such a correlation.

Understanding the sources of aSyn, delineating the molecular events underlying its presumed exocytosis by cells into the interstitial, ventricular and subarachnoid spaces of the CNS, and cataloging its modified variants represent pivotal tasks for our field in the future. They are important in the identification of the unknown function of
aSyn in CSF, its main degradation pathways in intra- and extracellular compartments (including in CSF), and its possible uptake by neural, ependymal and epithelial cells from circulating CSF. These related topics have major implications for the pathogenesis, the diagnosis and possibly, the progression of several, currently incurable synucleinopathy disorders of the brain including PD, MSA and DLB (Desplats et al. 2009; Hong et al. 2010; Kramer and Schulz-Schaeffer 2007; Mollenhauer et al. 2011; Volpicelli-Daley et al. 2011).

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