Directed Evolution of an Improved Rubisco; In Vitro Analyses to Decipher Fact from Fiction

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Received: 30 August 2019; Accepted: 4 October 2019; Published: 10 October 2019

Abstract: Inaccuracies in biochemically characterizing the amount and CO₂-fixing properties of the photosynthetic enzyme Ribulose-1,5-bisphosphate carboxylase/oxygenase continue to hamper an accurate evaluation of Rubisco mutants selected by directed evolution. Here, we outline an analytical pipeline for accurately quantifying Rubisco content and kinetics that averts the misinterpretation of directed evolution outcomes. Our study utilizes a new T7-promoter regulated Rubisco Dependent Escherichia coli (RDE3) screen to successfully select for the first Rhodobacter sphaeroides Rubisco (RsRubisco) mutant with improved CO₂-fixing properties. The RsRubisco contains four amino acid substitutions in the large subunit (RbcL) and an improved carboxylation rate (k_catC, up 27%), carboxylation efficiency (k_catC/K_m for CO₂, increased 17%), unchanged CO₂/O₂ specificity and a 40% lower holoenzyme biogenesis capacity. Biochemical analysis of RsRubisco chimeras coding one to three of the altered amino acids showed Lys-83-Gln and Arg-252-Leu substitutions (plant RbcL numbering) together, but not independently, impaired holoenzyme (L_8S_8) assembly. An N-terminal Val-11-Ile substitution did not affect RsRubisco catalysis or assembly, while a Tyr-345-Phe mutation alone conferred the improved kinetics without an effect on RsRubisco production. This study confirms the feasibility of improving Rubisco by directed evolution using an analytical pipeline that can identify false positives and reliably discriminate carboxylation enhancing amino acids changes from those influencing Rubisco biogenesis (solubility).

Keywords: photosynthesis; carbon fixation; synthetic biology; metabolic engineering

1. Introduction

Ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) is the initial, often rate-limiting, enzyme in the photosynthetic Calvin–Benson–Bassham (CBB) cycle that converts CO₂ into the pre-cursor organic molecules that underpin life in the biosphere [1]. Rubisco constitutes 20–50% of the protein in leaves and 5–25% of the protein in algae and photosynthetic prokaryotes, making it the most abundant protein on earth [2–4]. This abundance is necessary due to the slowness of the rate at which Rubisco fixes CO₂ to Ribulose-1,5-bisphosphate (RuBP) to produce duplicate 3-phosphoglycerate (3PGA) molecules—a reaction completing only 2–5 cycles per second in vascular plants. The carboxylation reaction rate is also compromised by the erroneous fixation of O₂ instead of CO₂ producing a 3PGA and 2-phosphoglycolate (2PG), a metabolically toxic product whose recycling back into 3PGA (from two 2PG) by photorespiration consumes energy and releases fixed CO₂ [5,6]. In crops such as wheat and rice, 25% or more of the CO₂ assimilated by photosynthesis is lost during photorespiration. Improving the carboxylation properties of plant Rubisco has therefore long been considered a valuable approach for enhancing crop photosynthesis and growth [7]. As more efficient Rubisco isoforms of an equivalent Form I structure exist naturally in some red algae, the feasibility of improving plant Rubisco appears tenable [8].
Form I Rubisco comprises a structurally conserved ~550 kD complex comprising a core of eight large (RbcL) subunits (arranged as a tetrad of RbcL2 dimers) capped by two tetrads of small (RbcS) subunits [9]. The eight catalytic sites in each L\textsubscript{8}S\textsubscript{8} complex are formed by conserved amino acids in each adjoining RbcL2 pair (Figure A1). While we have a relatively detailed understanding of the conserved, complex catalytic chemistry of Rubisco [10,11] the rationale design of a better Rubisco remains elusive. Compounding the challenge is the finding that natural Rubisco kinetic diversity has evolved from amino acid differences outside of the catalytic pocket, sometimes in the distantly-located RbcS [12–14].

Defining what constitutes a better Rubisco is dependent on the cellular physiology of the target organism. In organisms incorporating a CO\textsubscript{2}-concentrating mechanism (CCM) to hinder Rubisco oxygenation (e.g., in most algae, cyanobacteria and plants with crassulacean acid metabolism, CAM, or C4-physiologies), improving the CO\textsubscript{2} fixation rate (\(k_{\text{cat}}^{C}\)) would represent a better Rubisco [1]. In organisms lacking a CCM, such as C\textsubscript{3} plants, the photosynthetic models show that improving Rubisco specificity for CO\textsubscript{2} over O\textsubscript{2} (SC/O) and increasing carboxylation efficiency (i.e., \(k_{\text{cat}}^{C}/K_{c}^{21\%O_{2}}\)) are key to producing a “better” Rubisco [15]. Rubisco from some red algae possesses these desired kinetic improvements. For example the SC/O and \(k_{\text{cat}}^{C}/K_{c}^{21\%O_{2}}\) values for red algae Griffithsia monilis Rubisco are, respectively, two-fold and 1.3-fold higher than tobacco Rubisco [16]. While this kinetic improvement has the potential to improve plant growth by 30% [17], the assembly requirements of red algae Rubisco are unfortunately not met in tobacco chloroplasts [18,19]. A comparable molecular chaperone incompatibility also impairs, or prevents, the assembly of heterologous plant and cyanobacteria Rubisco in tobacco chloroplasts [20–23].

A successful approach to improving Rubisco kinetics is via the directed evolution of randomly mutated \textit{rbcL} (+/− \textit{rbcS}) libraries using high-throughput Rubisco-dependent \textit{Escherichia coli} (RDE) screens [22,24,25]. As summarized in Figure 1, a core requirement shared among RDE screens is the expression of a \textit{prk} gene coding phosphoribulose kinase (PRK). PRK catalyses the ATP-dependent phosphorylation of the ribulose 5-phosphate produced in the \textit{E. coli} pentose phosphate pathway into RuBP. For unknown reasons, the PRK reaction is toxic to many \textit{E. coli} strains unless Rubisco is introduced to catalyze the RuBP. As RDE selection is undertaken under elevated CO\textsubscript{2}, the RuBP is primarily carboxylated by Rubisco to produce the glycolytic intermediate 3PGA. Any 2PG produced via Rubisco oxygenase activity can also be metabolized by \textit{E. coli} [26].

The life–death dependency of RDE screens on Rubisco activity has been exploited using the L-arabinose inducible \textit{P\textsubscript{BAD}} promoter to modulate PRK expression (i.e., RuBP production) to select for cells producing higher levels of Rubisco activity. This improvement to RDE fitness can stem from mutations in Rubisco that either improve its biogenesis (i.e., increasing the solubility of Rubisco) or/and increase its catalytic rate. Failure to accurately determine which of these Rubisco biochemical properties improves RDE selection fitness has led to solubility-enhancing RbcL substitutions being erroneously reported as mutations that enhance carboxylase activity (Table 1) [25].
Influence of commonly selected cyanobacteria Rubisco mutants on Rubisco content and catalysis (relative to wild-type, WT)—deciphering the trait that improves fitness.

Table 1. Influence of commonly selected cyanobacteria Rubisco mutants on Rubisco content and catalysis (relative to wild-type, WT)—deciphering the trait that improves fitness.

| Amino Acid Mutation | Selected Trait | Rubisco (Solubility) | \( k_{\text{cat}}/K_C \) | \( K_C \) | \( k_{\text{cat}}^{\text{air}}/K_C \) | \( k_{\text{cat}}^{\text{air}}/K_C \) | b Selection Host | Reference |
|---------------------|----------------|----------------------|----------------|-------|----------------|----------------|----------------|----------|
| F140I               | solubility    | ↑ 46%                 | ↓ 48%            | -     | ↑ 288%         | ↓ 9%           | MM1            | [27]     |
| V189I               | solubility    | ↑ 12%                 | ↓ 16%            | -     | ↑ 6%           | WT             | MM1            | [27]     |
| M262T               | solubility    | ↑ 13%                 | ↓ 13%            | -     | ↑ 29%          | WT             | RDE2           | [28]     |
| F345I               | solubility    | ↑ 14%                 | ↓ 15%            | ↑ 14% | ↓ 15%          | MM1            | [27]           |
| F345L               | solubility    | ↑ 17%                 | ↓ 22%            | -     | ↓ 32%          | MM1            | [27]           |
| F345V               | solubility    | ↑ 15%                 | ↓ 15%            | -     | ↓ 21%          | MM1            | [27]           |
| \( \text{F}145A \)  | solubility    | ↑ 16-fold             | ↑ 15%            | ↑ 15% | ↑ 18%          | R. capsulatus   | [28]           |
| \( \text{F}145A \)  | catalytic     | ↑ 15%                 | ↓ 4%             | -     | -              | MM1            | [27]           |
| \( \text{F}214S \)  | catalytic     | ↑ 14%                 | ↓ 14%            | -     | -              | MM1            | [27]           |

\( \text{ScRubisco (Synechococcus sp. PCC 6301)} \)—Expressed in \( \text{E. coli at -1\% (w/v) of the Cell Soluble Protein.} \)

| Host | Reference |
|------|-----------|
| \( \text{E. coli} / \text{w} \) | [27] |

\( \text{Ee-Rubisco (Thermosynechococcus elongatus BP-1)} \)—Expressed in \( \text{E. coli at -6\% (w/v) of the Soluble Protein.} \)

| Host | Reference |
|------|-----------|
| \( \text{E. coli} / \text{w} \) | [27] |
The common Rubisco substrate used in directed evolution studies is that from the cyanobacterium *Synechococcus elongatus* PCC6301 (SrRubisco, Table 1). When expressed in *E. coli*, the SrRubisco RbcL and RbcS are produced in high abundance, with >90% of the RbcL produced forming misfolded insoluble aggregates, thereby limiting SrRubisco expression to ~1% (w/w) of the soluble cellular protein [27,29,30,32]. Unassembled RbcS are less prone to insoluble aggregate formation, allowing them to rapidly bind (within <1 sec) to rbcL4 cores to form stable RbcL4RbcS8 (L4S8) complexes [33]. The limitations to GroELS chaperonin facilitating *E. coli* folding and assembly in *E. coli* have led to the repeated selection of particular amino acids changes (e.g., RbcL residues 140, 189, 262, 345; Figure A1). The governing biochemical feature that improves RDE fitness (i.e., cell growth rate). b A directed evolution screen is used to isolate the mutant (see Figure 1). The *R. capsulatus* screen is a low transformation frequency system that uses a Rubisco null mutant [31]. c Underestimates of Rubisco content likely explain the erroneous carboxylation rate and efficiency improvements. d Improvements in carboxylase properties may contribute to the improved fitness that is primarily imparted by the >4-fold increases in Rubisco biogenesis (solubility). Enhanced carboxylase activities are shown in bold-type. Arrows indicate whether the kinetic value is higher (↑) or lower (↓).

A recent, two-tiered directed evolution study used *Te*-Rubisco from *Thermosynechococcus elongatus* whose assembly requirements are better met in *E. coli* (produced at ~6% (w/w) of the soluble cellular protein [22]). Using the “false-positive free” RDE2 screen (Figure 1), *Te*-Rubisco was evolved in sequential stages, with the first identifying the catalytically neutral P415A mutation that improved *Te*-Rubisco assembly 1.6-fold. The second cycle of evolution then successfully selected additional mutations to P415A that were all co-located at the RbcL–RbcS interface of the holoenzyme that could enhance $k_{\text{cat}}^C$, $k_{\text{cat}}^C/K_C^{21\%\text{O}_2}$ and $S_{\text{CO}_2}$ (Table 1).

In this study, we trial a re-designed version of the RDE2 screen (here, named RDE3) to evolve Form I *RsRubisco* from *Rhodobacter sphaeroides*. Unlike SrRubisco and plant Rubisco (that are both in the “green-type” Form IB lineage), RsRubisco is in the “red-type” Form IC lineage and more related to the catalytically superior Form ID algae Rubisco from *G. monilis* [34,35]. Unlike the RDE2 screen, RsRubisco production in the RDE3 system is regulated by bacteriophage T7 transcription and undertaken in JM109(DE3), a lacUV5-regulated T7 RNA polymerase-expressing *E. coli* strain sensitive to PRK expression. Described is a model-directed evolution experimental pipeline using RDE3 that distinguished amino acid substitutions that impair RsRubisco biogenesis from one that significantly enhanced $k_{\text{cat}}^C$ and $k_{\text{cat}}^C/K_C$.

### Table 1. Cont.

| Amino Acid Mutation | Trait Selected | Rubisco (Solubility) | $k_{\text{cat}}^C$ | $K_C$ | $K_C^{21\%\text{O}_2}$ | $k_{\text{cat}}^C/K_C$ | $k_{\text{cat}}^C/K_C^{21\%\text{O}_2}$ | Selection Host | Reference |
|---------------------|----------------|----------------------|------------------|-------|----------------------|-----------------------|-----------------------------|--------------|----------|
| P415A A48V          | kinetic        | ↑ 1.5-fold            | ↑ 33%            | -     | WT                   | ↑ 35%                 | ↓ 10%                       | RDE2         | [22]     |
| P415A H47L          | kinetic        | ↑ 1.7-fold            | ↑ 27%            | -     | WT                   | ↑ 28%                 | ↓ 7%                        | RDE2         | [22]     |
| P415A Y36N/G112D    | kinetic        | ↑ 1.5-fold            | ↑ 41%            | -     | ↓ 12%                | ↑ 60%                 | ↓ 10%                       | RDE2         | [22]     |
| P415A L74M/D397N    | solubility/kinetic | ↑ 1.9-fold        | ↑ 24%            | -     | ↓ 14%                | ↑ 40%                 | = WT                        | RDE2         | [22]     |

Amino acid numbering relative to plant RbcL (Figure A1). Data summarized from the Rubisco content and kinetic listings in Table A1. a The common Rubisco substrate used in directed evolution studies is that from the cyanobacterium *Synechococcus elongatus* PCC6301 (SrRubisco, Table 1). When expressed in *E. coli*, the SrRubisco RbcL and RbcS are produced in high abundance, with >90% of the RbcL produced forming misfolded insoluble aggregates, thereby limiting SrRubisco expression to ~1% (w/w) of the soluble cellular protein [27,29,30,32]. Unassembled RbcS are less prone to insoluble aggregate formation, allowing them to rapidly bind (within <1 sec) to rbcL4 cores to form stable RbcL4RbcS8 (L4S8) complexes [33]. The limitations to GroELS chaperonin facilitating *E. coli* folding and assembly in *E. coli* have led to the repeated selection of particular amino acids changes (e.g., RbcL residues 140, 189, 262, 345; Figure A1) that improve SrRubisco biogenesis by >4-fold (Table 1). The impact of these mutations on catalysis is that they mostly impair—or at best provide only modest changes in—the carboxylation properties SrRubisco.

2. Results

2.1. Ambiguity in How Cyanobacteria Rubisco Mutations Improve RDE Fitness

Prior RDE-directed evolution studies have mistakenly reported SrRubisco amino acid substitutions which improve the folding and assembly (solubility) of SrRubisco as mutations that enhance catalysis. As summarized in Table 1, the commonly selected M262T SrRubisco substitution initially reported to enhance catalysis [36] was subsequently shown to enhance SrRubisco solubility in *E. coli* [29] and may also account for the same mutation enhancing fitness in *R. capsulatus* selection [28]. Similarly the
SeRbcL F140L and V189I mutations were initially characterized a catalytic enhancers of RDE fitness [27] but subsequently shown to improve SeRubisco solubility by nearly 5-fold in E. coli [22].

The erroneous assessment of L8S8 recombinant holoenzyme production in E. coli often stems from the inherent complexity of the enzyme assembly requirements that impair its synthesis in heterologous hosts [24,25,37]. This is particularly true for the cyanobacteria Rubisco, whose assembly requirements are either not met, or are poorly met, in E. coli [29,32]. For example, the SeRbcL and SeRbcS subunits are produced in abundance in E. coli, with SeRbcL comprising >10% (w/w) of the total cellular protein (Figure 2a). Compared to SeRbcS that is mostly soluble, more than 90% of the SeRbcL produced in E. coli forms misassembled insoluble protein aggregates (compare the lysate and soluble SeRbcL and SeRbcS immunoblot signals in Figure 2b). The remaining soluble SeRbcL are primarily found assembled in SeL8S8 complexes, with the unassembled soluble SeRbcS monomers able to bind instantaneously with any assembled RbcL8 cores via a chaperone-independent process [33]. By comparison, the assembly requirements of RsRubisco are more readily met in E. coli, with a greater proportion of soluble RsRbcL produced and assembled into RsL8S8 complexes (Figure 2c).

![Figure 2](image_url)

**Figure 2.** SDS PAGE analysis of SeRubisco and RsRubisco production in BL21(DE3). Rubisco production was induced at 25 °C with 0.5 mM isopropyl-β-D-thiogalactosidase (IPTG) for 8 h, the cells were lysed and (a) 4 µg of soluble protein (Sol) and an equivalent volume of the total cellular protein (Lys, comprising soluble, membrane and insoluble inclusion body proteins) were separated by SDS PAGE and Coomassie stained. Duplicate PAGE gels loaded with lower amounts of each protein preparation (as indicated) were blotted onto nitrocellulose membranes and probed with antibodies (Ab) to (b) SeRubisco and (c) RsRubisco. Empty vector BL21(DE3)-pET28a(+) cells were used as a “no Rubisco” control. *, E. coli protein recognized by the SeRubisco antibody. The SeRbcL is produced in high abundance and mostly insoluble (lysate signal >> soluble signal), while SeRbcS, RsRbcL and RsRbcS are mostly soluble. SeRubisco and RsRubisco L8S8 contents comprised, respectively, 1.1% and 4.8% (w/w) of the soluble cellular protein (as quantified by 14C- carboxyarabinitol 1,5-bisphosphatase (CABP)-binding). M, protein markers (sizes shown).

The low level of SeRubisco produced in E. coli (comprising ~1% (w/w) of the E. coli-soluble protein, Table 1) hinders the viability of SDS PAGE, Western blotting and native PAGE methods to quantify Rubisco content. Even when analyzed alongside known amounts of SeRubisco, these methods can lack sufficient quantifiable resolution [38]. By contrast, the tight and stoichiometric binding properties of the Rubisco specific inhibitor carboxyarabinitol 1,5-bisphosphatase (CABP) provide an accurate method to quantify Rubisco in any soluble cellular protein sample [39]. The method, termed 14C-CABP binding, takes advantage of CABP’s capacity to tightly bind to each Rubisco catalytic site to form a highly stable complex that, upon separation from unbound 14C-CABP (e.g., by gel filtration), provides an accurate quantitative measure of Rubisco content [38].
2.2. An Analytical Pathway for the Directed Evolution of Rubisco Using an RDE Screen

Our routine Rubisco-directed evolution experimental protocol spanning RDE selection to in vitro biochemistry analytics is summarized in Figure 3 [22,40]. The experimental pipeline begins with the generation and transformation of an rbcL (+/- rbcS) mutant library into an RDE screen alongside RDE cells expressing the non-mutated rbcL-rbcS gene control (Figure 3a). Colonies showing improved RDE fitness (i.e., those which can grow on PRK-inducing arabinose and IPTG-inducing Rubisco concentrations that have been pre-determined to be non-permissive to the growth of cells expressing the parental Rubisco substrate) are individually selected, and the rbcL-rbcS plasmid is isolated, retransformed into RDE and the colony growth on increasing arabinose compared with the control cells to gauge the relative improvement in cell “fitness” (Figure 3b). The rbcL-rbcS plasmid is isolated and sequenced from the RDE cells with improved fitness and transformed into a suitable E. coli strain to quantify changes in Rubisco expression and catalysis (Figure 3c). SDS PAGE samples of the total and soluble cell protein are taken to qualitatively assess the proportion of soluble and insoluble RbcL and RbcS produced (Figure 2). The soluble protein concentration is measured, the Rubisco content is quantified by 14C-CABP binding, and 4 to 8 µg of protein is separated by native PAGE to verify the LsRb Rubisco content and aliquots used in 14CO2-fixation assays to quantify the CO2-fixation rate (kcatC) and the Km for CO2 under anaerobic conditions (Kc), ambient O2 levels (KC 21%O2), or over a range of O2 levels to quantify the Km for O2 (KO) [22]. To accurately measure the specificity for CO2 over O2 (S(O2)), the method of Kane et al. (1994) [41] is used, which requires Rubisco that has been purified from the E. coli using a rapid ion exchange and gel filtration process [42].

![Figure 3](https://example.com/figure3.png)

Figure 3. The analytical pipeline of an RDE experiment. An RDE-directed evolution project begins with (a) Rubisco gene library generation, transformation and RDE selection. The Rubisco gene plasmid (rbcL-rbcS) from cells with improved fitness is (b) transformed into fresh RDE cells and re-screened on PRK-inducing arabinose to confirm the improved growth phenotype before (c) sequencing to ascertain the amino acid substitutions. Each Rubisco mutant is then expressed in E. coli and the Rubisco subunit folding/assembly capacity evaluated by SDS PAGE (e.g., Figure 2); the LsRb holoenzyme content is then quantified (14C-CABP binding), confirmed (native PAGE) and (d) the kinetics (S(O2), kcatC, Kc and KC 21%O2) measured as described in [22].

2.3. Determination of a Suitable E. coli Strain for RDE3

As in any directed protein evolution experiment, the level of success is dependent on the fidelity and throughput of the selection system employed. For evolving Rubisco, the use of the photosynthetic R. capsulitis bacterial screen is hampered by low throughput (screening is limited to a few thousand mutants) and hindered in its ability to characterize mutants with alterations in solubility [28,31,43,44]. As summarized in Figure 1, while RDE screens have higher throughput, their fidelity can be compromised by false positives. These occur at a higher frequency (~0.5% of...
the colony forming units plated) in the original RDE screens than in the metabolically altered MM1 RDE strain [24]. The false positives produced in MM1 selection have primarily been attributed to transposon insertions into a pACYC<sub>RPK</sub> plasmid that silenced PRK production [25]. This has been avoided in the RDE2 screen by fusing NPTII onto the PRK C-terminus and including kanamycin in the selection media, meaning that false positives originating from transposon-induced PRK silencing cannot grow (Figure 1) [22].

In this study, we sought to adapt the RDE2 screen so that Rubisco production was regulated by bacteriophage T7 transcription rather than the previously used lac promoter in pTrcHisB. This necessitated that RDE selection be undertaken in a <i>lacUV5</i>-regulated T7 RNA polymerase-expressing <i>E. coli</i> strain. The plating of differing <i>E. coli</i> strains transformed with pACYC<sub>prk</sub>kan on media containing differing arabinose concentrations found differences in their susceptibility to PRK-NPTII production (Figure 4). The JM109(DE3), XL1-Blue and DH5α strains were sensitive to PRK-NPTII synthesis, while DB3.1, BL21(DE3) and BW25113 were not, making the latter three strains unsuitable for use in an RDE screen. The pACYC<sub>prk</sub>kan plasmids from DB3.1, BL21(DE3) and BW25113 colonies growing on 0.25% (w/v) arabinose were purified, restriction digested (Figure A2) and sequenced. There was no evidence that PRK-NPTII production was compromised, indicating that these three strains show an insensitivity to PRK production under the growth conditions trialed.

As JM109(DE3) is suited to T7-regulated recombinant expression, it was the strain used in our new RDE3 screen. Comparative expression analyses found that RsRubisco was expressed equally well in JM109(DE3) (~5% (w/w) of the soluble protein, see Table 2) as in BL21(DE3) (Figure 2).

**2.4. Directed Evolution of RsRubisco in the RDE3 Screen**

In this pilot RDE3-directed evolution experiment, the Form I<sub>C</sub> RsRubisco was the chosen substrate as (1) its assembly requirements are well met in <i>E. coli</i> (Figure 2) including in JM109(DE3) (Table 2), (2) it is of the “red-type” Rubisco phylogenetic lineage that encompasses the superior Form ID Rubisco isoforms from red algae [35], and (3) it has not been used in a directed evolution study. Electro-competent JM109(DE3) containing pACYC<sub>prk</sub>kan [22] (here on termed RDE3 cells) were transformed with an Rs<sub>rbcL–rbcS</sub> mutant library cloned into plasmid pYZRsLS (Amp<sup>R</sup>, Figure 5a). Approximately 10<sup>5</sup> colony forming units (cfu) were plated on a 150 mm plate of selective LB media containing 75 μM IPTG, 0.25% (w/v) arabinose and antibiotics (Figure 5b). RDE3–pYZRsLS cells expressing non-mutated RsRubisco (2 x 10<sup>4</sup> cfu; wild-type (WT) control) were grown on a 90 mm plate of replica selection media. After five days at 25 °C in air supplemented with 2% (v/v) CO<sub>2</sub>, 37 colonies had grown on the mutant RsRubisco plate and eight on the WT RsRubisco control (Figure 5b).
The pYZRsLS from 33 mutant and all eight WT colonies were purified and restriction-digested to confirm they contained the Rs–rbcL operon before re-transforming into fresh RDE3 cells and plating again onto LB containing 0.25% (w/v) arabinose (Figure 5c). Only one mutant, called YZ1, enhanced the growth fitness of RDE3 under this selection pressure, with the remainder showing comparable arabinose sensitivities to the pYZRsLS(WT) control (Figure 5c).

Figure 5. RDE3 evolution of RsRubisco. (a) Wild-type (WT) and a mutated Rs–rbcL operon library cloned into pYZRsLS were transformed into RDE3 and (b) screened under strong arabinose selection as shown. The surviving colonies were grown (under non-selective conditions), the pYZRsLS purified and those containing the Rs–rbcL operon re-transformed into RDE3 and (c) re-screened on selection media (examples for five of the 41 RDE3 re-screens are shown). Only the YZ1 mutant improved RDE3 growth, and (d) the sequencing of its Rs–rbcL operon showed that it coded RbcL mutations V11L, K84Q, R253L and Y345F (plant RbcL numbering, Table A1). Nine YZ1 chimers (named YZ1a to YZ1i) coding differing combinations of these mutations were created to study how they influence Rs–rbcLS biogenesis (Figure 6) and catalysis (Table 2).

Table 2. Comparative E. coli biogenesis (solubility) and catalysis (at 25 °C) of wild-type, YZ1 and their RsRubisco chimeras.

| RsRubisco Isoform | RbcL Residue Number | Rubisco (%) CSP | kC (µM) | kcatC (s⁻¹) | kcatC/KC (mM⁻¹s⁻¹) | σC mol mol⁻¹ | KC (µM) | kcatC/KO (mM⁻¹s⁻¹) | σO µM µM⁻¹ |
|-------------------|---------------------|----------------|--------|-------------|--------------------|------------|--------|-------------------|------------|
| WT                | Val Lys Arg Tyr     | 4.9 ± 0.4 a    | 60 ± 1 a| 3.7 ± 0.2 a  | 60 ± 1             | 58 ± 1 a  | 1724 ± 280 | 1.8 ± 0.9 a      | 1.0 ± 0.1 a |
| YZ1               | Ile Glu Leu Phe     | 2.9 ± 0.1 b    | 66 ± 1 b| 4.7 ± 0.1 e  | 70 ± 1             | 57 ± 1 b  | 1296 ± 207 | 1.6 ± 1.3 a      | 1.3 ± 0.1 a |
| YZ1a              | Val Glu Leu Phe     | 3.3 ± 0.4 b    | 66 ± 1 b| 4.5 ± 0.1 e  | 68 ± 2             | -         | -       | -                 | -          |
| YZ1b              | Val Glu Leu Tyr     | 4.0 ± 0.1 b    | 61 ± 1 b| 3.8 ± 0.1 b  | 63 ± 1             | -         | -       | -                 | -          |
| YZ1c              | Val Glu Leu Tyr     | 3.6 ± 0.4 b    | 63 ± 1 b| 3.9 ± 0.1 b  | 63 ± 1             | -         | -       | -                 | -          |
| YZ1d              | Ile Glu Arg Phe     | 4.6 ± 0.1 a    | 62 ± 1 a| 4.6 ± 0.1 a  | 76 ± 1             | -         | -       | -                 | -          |
| YZ1e              | Ile Glu Leu Phe     | 4.5 ± 0.3 a    | 63 ± 1 a| 4.3 ± 0.1 a  | 70 ± 3             | -         | -       | -                 | -          |
| YZ1f              | Val Glu Arg Tyr     | 5.1 ± 0.4 a    | 61 ± 1 a| 3.9 ± 0.2 a  | 65 ± 1             | 57 ± 1 a  | 1220 ± 199 | 1.4 ± 1.1 a      | 1.1 ± 1.1 a |
| YZ1g              | Val Lys Leu Tyr     | 4.3 ± 0.2 a    | 62 ± 1 a| 3.5 ± 0.1 a  | 57 ± 3             | -         | -       | -                 | -          |
| YZ1h              | Ile Lys Arg Phe     | 4.6 ± 0.1 a    | 61 ± 1 a| 4.4 ± 0.1 a  | 73 ± 1             | -         | -       | -                 | -          |
| YZ1i              | Val Lys Leu Tyr     | 4.7 ± 0.2 a    | 64 ± 1 a| 4.6 ± 0.1 a  | 71 ± 2             | 59 ± 1 a  | 1551 ± 293 | 1.9 ± 1.2 a      | 1.2 ± 1.2 a |

Values in bold represent significant improvements in carboxylase activity. CSP (percentage (w/v) of the cell soluble protein), kcatC and KC data are the mean ± SE of N = 5 (and for S̄O, N = 2) biological samples, each assayed in duplicate. Letters show the ranking of the means using a post hoc Tukey test (different letters indicate statistical differences at the 5% level, p < 0.05) following a one-way ANOVA comparison relative to WT. kcatC, maximal oxygenation rate, calculated from S̄O = (kcatC/KC)µM/kcatC/KO. Mutated residues are shaded black.

2.5. The RbcL Substitution Y345F Enhances RsRubisco Carboxylation without Impairing Enzyme Biogenesis

The sequencing of the YZ1 pYZRsLS plasmid found it coded five Rs–rbcL nucleotide mutations, with four coding the RsbRbcL mutations V11L (GTT-ATT), K84Q (AAA-CAA), R253L (CGT-CTT) and Y345F (TAT-TTT) and one a synonymous CTG-CTC change to L391. Relative to tobacco Rubisco, numbering these mutations corresponded to K83Q, R252L and Y345F (Figure A1a). A kinetic analysis of YZ1 RsRubisco found that the RbcL mutations had no effect on the S̄O but significantly stimulated kcatC and carboxylation efficiency (kcatC/KC) by 27% and 17%, respectively (Table 2). The mutations also increased the affinity of YZ1-Rubisco for O2 (i.e., KO was 25% lower than RsRubisco), thereby...
reducing its CO₂ affinity under ambient O₂ (K_C^121%O₂ = 78.8 μM) relative to wild-type RsRubisco (K_C^21%O₂ = 68.8 μM). Nevertheless, the carboxylation efficiency of YZ1-Rubisco in air (k_cat/C/K_C^21%O₂ = 59.6 mM⁻¹.s⁻¹) still exceeded RsRubisco (53.8 mM⁻¹.s⁻¹) by 11%.

The analysis of YZ1-RsRubisco solubility by 14C-CABP binding and native PAGE showed its biogenesis in JM109(DE3) was ~40% lower than wild-type RsRubisco (Table 2, Figure 6). A series of nine YZ1 RsRubisco chimeras coding one to three RbcL amino acid substitutions was made (called YZ1a to YZ1i, Figure 5d) to examine whether the changes in biogenesis and kinetics could be uncoupled. As summarized in Table 2, all YZ1-chimers incorporating the Y345F mutation sustained the kinetics of RsRubisco, demonstrating that this mutation imparted the improved k_cat C and k_cat C/K_C kinetic phenotype. The assembly of RsRubisco was only impaired in the YZ1-chimers coding both K83Q and R252L together (YZ1, YZ1a, YZ1b, YZ1c, Table 2, Figure 6). On their own (YZ1f, Yz1g) or in combination (YZ1c), the K83Q and R252L substitutions had no significant influence on RsRubisco kinetics (Table 2). The separation of all the YZ1 RsRubisco chimeras incorporating K83Q through native PAGE was marginally faster, suggesting that the mutation altered the net charge of the enzyme or induced a structural conformational change that increased its migration rate (Figure 6).

![Figure 6](image-url)

**Figure 6.** Native PAGE analysis of RsRubisco and the YZ1 chimeras’ relative expression in JM109(DE3) after 0.5 mM IPTG induction for 6 h at 28 °C. The separated soluble cellular protein (4 μg, including an empty vector (EV) JM109(DE3)-pET28a(+) control) was Coomassie stained, with the RsRubisco band intensity correlating with that quantified by 14C-CABP binding (indicated at base of gel). RsRubisco biogenesis for the YZ1 chimers similar to wild-type (WT) are shaded green, while those produced in amounts equivalent to YZ1-RsRubisco are shaded red.

3. Discussion

A common challenge encountered with Rubisco-directed evolution studies using RDE selection is accurately measuring the biochemical properties of the mutant enzymes to correctly interpret the basis of how they improved RDE fitness [25]. Inaccuracies primarily arise from the erroneous evaluation of how amino acid substitutions affect Rubisco assembly (solubility) and kinetics (Table 1). Here, we demonstrate an experimental pipeline that spans the “selection” to the “analysis” phases of RDE-directed evolution (Figure 3). Using the pipeline, we successfully identified amino acids in Form IC “red-type” RsRubisco that influence either solubility or catalysis (Table 2). Our success in improving the carboxylation properties of RsRubisco extends to the comparable achievements made using MM1 and RDE2 in identifying amino acid substitutions that improve the carboxylase activities of the archaeon Methanococcales burtonii Form III Rubisco decamer (rbcL_L0/40) and the Form IB “green-type” LsS5 TrRubisco [22]. The chimeric dissection of the selected YZ1 RsRubisco mutant identified Y345F as a mutation that increases carboxylase activity and K84Q and R253L as substitutions that, in combination, impair LsS5 biogenesis (Table 2). Our findings also show that sensitivity to PRK production varies between E. coli strains (Figure 4) and that RDE false positive selection can arise independently from prk silencing (Figure 5b,c and Figure A2). This raises questions...
regarding the validity of the underpinning basis for RDE selection; i.e., that cell fitness is dependent on Rubisco expression to alleviate the inexplicable cellular toxicity of PRK-dependent RuBP production (Figure 1, [24]), a metabolic impediment that does not appear universal among all E. coli strains.

3.1. Mutating RbcL Amino Acid 345 Has Alternate Effects on Form I Rubisco Biogenesis and Catalysis

A common outcome of RDE screens is the selection of Rubisco mutants coding amino acid substitutions at residue 345 (numbered according to plant Rubisco, Table 1). Amino acid 345 is positioned in α-helix 6 of the six-stranded α/β-barrel C-domain of RbcL (as structurally defined by [45]). In the L₄S₈ structure residue, 345 is embedded beneath the enzymes’ outer surface, away from subunit interfaces and ~5.6 Å from Arg-295, a conserved catalytic site residue involved in binding the P₅ phosphate group of RuBP (Figure 7). Adjoining α-helix 6 is the catalytic loop 6 (residues 331AVGKLE336, Figure A1). This conserved, flexible, “lid-like” sequence interacts with the C-terminal tail to close over the catalytic pocket upon RuBP binding, then opening upon catalysis to allow product release [10]. Among Form IB and ID Rubisco isoforms, position 345 is phenylalanine (Phe, F), and in cyanobacteria Form IA Rubisco and RsRubisco Form IC, it is a tyrosine (Tyr, Y) (Figure A1). A mutagenic analysis of Form IB SeRubisco found changes to α-helix 6 amino acids located closer to loop 6 dramatically impaired carboxylase activity [46]. Not included in this analysis were changes to the other α-helix 6 C³⁴⁵FVDL residues that are conserved among Form IB Rubisco. From this analysis, it was hypothesized that structural changes in α-helix 6 can influence loop 6 positioning and movement [46]. Such movements are thought to alter how K334 (the catalytically critical loop 6 apex residue) ligands with the RuBP enediolate transition state intermediate and influences its catalysis with CO₂ or O₂.

Figure 7. Mapping the location of the RDE3-selected YZ1 mutations in R. sphaeroides Rubisco. Pymol-assembled RsRubisco (PDB 5NV3) structures of (a) a single RsRbcL subunit, (b) L₄S₈ holoenzyme (side view) and (c) L₄S₂ sub-complex (cross-section view). RbcL pairs are colored with differing shades of grey, and RbcS is alternately colored pink/salmon. Shown are the locations of the YZ1-selected RbcL amino acid substitutions K83Q (green, strand C), R252L (yellow, α-helix 3) and Y345F (blue, α-helix 6) relative to the conserved catalytic pocket (CP) residues (dark pink), including R295 (green), K201 (light blue) loop 6 (red) and CABP (black, RuBP intermediary analog). E109 (orange) forms a salt bridge with a conserved R253 in the adjacent RbcL. The R252L substitution is located on the surface of the central solvent channel (CSC) that traverses the inner core of the L₄S₈ holoenzyme (dashed arrow, panel c). Residues are numbered relative to plant Rubisco using the alignment in Figure A1.
The existing data suggest that changes to residue 345 have a pervasive influence on Rubisco catalysis. For example, replacing Phe 345 with Ile, Leu or Val impairs the carboxylation properties of SrRubisco \([27,30]\) and TeRubisco \([22]\) (Table 1). It is difficult to structurally ascertain whether these kinetic changes arise from the mutations altering the positioning and movement of loop 6 or by altering RuBP ligand binding with Arg-295 in the catalytic pocket. Inexplicably, mutations to F345 also somehow impart two to 11-fold improvements in TeRubisco and SrRubisco production (Table 1). It is presumed that these changes enhance SeRubisco and TeRubisco folding and/or assembly in \textit{E. coli} \([22,24,27]\). By contrast, the Y345F substitution in RsRubisco had no effect on RsRubisco biogenesi in \textit{E. coli} and improved its carboxylation rate and efficiency by 27 and 17%, respectively (Table 2). This incongruence in how changes to amino acid 345 differentially influence the catalysis and assembly of Form IB and IC Rubiscos underpins the challenge of using rationale design approaches to identify “all-purpose” amino acid substitutions that can benefit catalysis between, and within, differing Rubisco lineages \([1,13,22,47]\).

3.2. How Do the K83Q and R252L Substitutions Function Cooperatively to Reduce RsRubisco Biogenesis?

K83 and R252 in RsRubisco are located, respectively, in strand C of the RbcL N-domain and \(\alpha\)-helix 3 of the \(\alpha/\beta\)-barrel (Figure 7a). Their distant locations from the catalytic sites are in accordance with our findings that the mutations K83Q and R252L, both singly (RsRubisco chimer YZ1f and YZ1g) and in combination (YZ1c), have no significant effect on catalysis (Table 2). RsRubisco biogenesis was not affected in YZ1 chimer containing only one of these mutations (chimers YZ1d to g), but was reduced by \(\sim 40\%\) when both were present (YZ1 and YZ1a to c, Table 2). How the K83Q and R252L substitutions work in combination, but not in isolation, to reduce the level of RsRubisco is unclear. It is possible that the mutations reduce RsRibcL compatibility with one or more components of the \textit{E. coli} protein folding machinery that impairs holoenzyme assembly or alters the stability of the \(L_8S_8\) complex. As summarized in Table 1, a number of single amino acid mutations are known to enhance cyanobacteria Rubisco biogenesis in \textit{E. coli}. For TeRubisco, the solubility-enhancing F345I and P415A mutations were able to function in a cumulative fashion to further augment solubility (Table 1). It is proposed that these improvements arose by improving the compatibility of RbcL with components of the \textit{E. coli} protein folding machinery \([22]\). It is known that chaperonin (GroELS) availability, but not DnaK, DnaJ, GrpE or Trigger Factor, has a pervasive influence on SrRubisco biogenesis in \textit{E. coli} \([29,32]\). Sequence complementarity with \textit{E. coli} GroELS has been used to explain the vast difference is the assembly capabilities of differing cyanobacteria Rubisco (e.g., PCC7002 Rubisco is produced at \(<0.5\%\) CSP \([32]\), SeRubisco at \(\sim 1.3\%\) CSP and TeRubisco at \(\sim 7\%\) CSP \([22,48]\)) and with RsRubisco (5% CSP, Table 2). It is therefore possible that the K84Q and R253L mutations in combination reduce RsRibcL compatibility with GroELS to impair RsRubisco biogenesis.

The evaluation of the RsRubisco \(L_8S_8\) crystal structure (Figure 7b) showed that R252 is located near the surface of the inner solvent channel that traverses the \(L_8S_8\) structure and is situated at the edge of the interface of adjoining large subunits that are arranged head to tail in each \(L_2\) unit (Figure 7c). The adjacent R253 residue is conserved among Form I Rubisco and forms a conserved salt bridge with E109 in the adjoining RbcL of each \(L_2\) (Figure 7c). While this intradimer interaction is hypothesized to maintain \(L_2\)'s structural integrity \([49]\), our findings suggest the R252L substitution does not disturb this Arg–Glu salt bridge as RsRubisco synthesis and stability seems unaffected (Table 2). In contrast to R252, K83 is a solvent-exposed outer surface residue and independent of any inter-subunit interactions (Figure 7b). Curiously, the YZ1 chimeras incorporating K83Q showed a faster mobility through native PAGE, independent of other amino acid substitutions (Figure 6), suggesting the mutation alters the net surface charge of the protein or influences RsRubisco quaternary structure, but without an effect on catalysis (Table 2). The disparate locations of the K83R and R252L mutations in the RsRubisco do not support the hypothesis that they jointly function to reduce RsRubisco content in \textit{E. coli} through destabilization of the \(L_8S_8\) complex.
3.3. Understanding the Basis of RDE Selection

The adage that “you get what you screen for” highlights how crucial it is to ensure any directed evolution screen can select for a desired fitness trait with high fidelity [50,51]. The production of false positives commonly compromises the fidelity of RDE screens, especially those using PRK expressing wild-type E. coli (Figure 1a). In the MM1 gapA− mutant RDE screen, the impaired cell viability associated with blocking glycolysis/gluconeogenesis by deleting glyceraldehyde-3-phosphate dehydrogenase (Figure 1b) reduces the frequency of false positives [24,26] which primarily occur from the transposon-interrupted expression of PRK [25]. The selection of these false positives in the XL1-Blue-based RDE2 screen was prevented using PRK:NPTII fusion so that PRK production and kanamycin resistance are linked, thereby preventing the selection of PRK-silenced mutants (Figure 1c). While initial tests indicated the suitability of the JM109(DE3) strain for RDE3 selection (Figure 4), a false positive production rate of approximately 0.04% was found when more cells were grown on non-permissive, high arabinose LB media (Figure 5b). Curiously, no sequence changes were evident in the purified pACYCprk:kan or the pYZRsLS from the eight WT control colonies that grew. A comparable frequency of false positives was also identified in the RDE3-mutant RsRubisco screen (Figure 5b), in which again the sequence integrity of pACYCprk:kan was maintained, and only one (YZ1) of the 33 mutant pYZRsLS plasmids re-transformed into RDE3 survived re-selection on high arabinose. The growth of the false positives in the initial RDE3 screens therefore arose independently of changes to either RPK:NPTII or RsRubisco expression. This suggests the false positives may result from chromosomal mutations that alter JM109(DE3) cellular metabolism to dampen its sensitivity to PRK production. This hypothesis is consistent with our finding that other E. coli strains (e.g., DB3.1, BL21(DE3) and BW25113) show resistance to PRK:NPTII production (Figure 4). Current research is underway to identify the genetic and/or metabolic basis for the contrasting sensitivities of E. coli strains to PRK production as a means to improve the fidelity of RDE screens.

3.4. Pathways for Evolving RsRubisco Further—Why and How?

Our interest in continuing to explore the evolutionary potential of RsRubisco stems from its relatedness to the Form ID “red-Rubisco” lineage that comprises red algae Rubisco isoforms whose high $S_{\text{CO}}$ and carboxylation efficiencies dramatically exceed those of crop plants [16,18]. While optimizing the fidelity of the RDE3 to avoid false positive selection is desirable, we found it is not critical if suitable care is taken to re-screen selected RsRubisco mutants (i.e., confirming that Rubisco is improving RDE3 fitness, Figure 5c) before pursuing Rubisco biochemical analyses. As the carboxylase improved YZ1i (Y345F) and biogenesis-impaired YZ1c (R83Q, R252L) mutants offer alternative starting points in the RsRubisco evolutionary landscape, separately evolving them using RDE3 and comparing the biochemical fitness traits selected (solubility and/or kinetic) would provide valuable strategical guidance to successive rounds of evolution.

Incorporating the R. sphaeroides chaperonin folding machinery components into RDE3 would alter—and possibly benefit—the RsRubisco evolutionary fitness landscape that can be surveyed. A similar approach taken with SeRubisco found that the inclusion of the Rubisco-specific post-chaperonin assembly chaperone RbcX altered the fitness landscape of the MM1 screen, with different mutants selected compared with a screen lacking RbcX [27]. This supported an earlier hypothesis that Rubisco evolution is influenced—and possibly even constrained—by the need to maintain complementarity with the protein folding/assembly machinery of a cell [24]. This appears particularly apparent for Form IB plant Rubiscos, whose assembly in E. coli requires five compatible molecular partners (i.e., the Rubisco-specific chaperones Raf1, RbcX, Raf2, BSD2 and Cpn60xβ20 chaperonin cages [37,52,53]). Thus far, maintaining sequence compatibility between Raf1 and RbcL appears to be the most important factor influencing plant Rubisco biogenesis [54]. Our findings that the R83Q and R252L substitutions inexplicably impair RsRubisco biogenesis in E. coli suggest sequence compatibility between RbcL and that the bacterial GroELS protein folding machinery also impacts Rubisco biogenesis potential.
4. Materials and Methods

4.1. Comparing the PRK Sensitivity of Different E. coli Strains

The pACYC<sup>prk:kan</sup> plasmid [22] was transformed into CaCl<sub>2</sub>-competent JM109(DE3), XL1-B, DH5α, DB3.1, BL21(DE3), and BW25113 E. coli cells, and a colony of each was grown in LB-Chlor (30 µg·mL<sup>−1</sup>) at 25 ºC. At an absorbance (A<sub>600</sub>) of 0.6, the cultures were diluted by 10<sup>−4</sup> and 20 µL was plated onto LB-Chlor plates containing 0, 0.05%, 0.1%, 0.25% (w/v) L-arginine with/without 100 µg·mL<sup>−1</sup> kanamycin. Colony growth at 25 ºC in air containing 2% (v/v) CO<sub>2</sub> was monitored up to 5 days. Electro-competent RDE3 cells (i.e., pACYC<sup>prk:kan</sup> transformed JM109(DE3)) in 10% (v/v) glycerol were N<sub>2</sub> frozen and stored at −80 ºC.

4.2. Library Construction and Rubisco Selection Using the RDE Screening

Figure 3 provides a general summary of an RDE-directed evolution experiment. The pilot RDE3 RsRubisco evolution experiment undertaken in this study is summarized in Figure 5. The 1965-bp synthetic Rs-rbcL operon in which the codon use was modified to match the tobacco rbcL gene (Genbank Accession KM464722) was PCR-amplified using primer 1355 (5′-ATCGAGGCTCGCCATA-3′, BsaI site underlined) and primer 1356 (5′-GCATGGTCTCCAAGCA-3′, BsaI site underlined) and cloned into a modified pTriEx-2 Amp plasmid (Novagen, Addgene, Cambridge, Massachusetts, USA) to give plasmid pYZrsLS (Genbank Accession MN541175, Figure 5a). The 13 N-terminal amino acids of the T7 promoter and adjacent to the T7 terminator. A mutant Rs-rbcL operon, constructed by restriction cloning so that each chimer only coded one to three of each mutation (Figure 5d). The amplified Rs-rbcL operon was ligated into pYZrsLS, containing the wild-type (WT) control. The cells were added to 1 mL LB, grown for 1 h at 37 ºC, 5 µL diluted and plated on non-selective media (LB-chlor-amp plates containing 125 µg/mL chloramphenicol and 100 µg/mL ampicillin) to determine the colony forming unit (cfu) size of the WT and mutant RsRubisco library. The mutant pYZrsLS plasmids from 10 colonies grown on permissive media were transformed into RDE3 and the transfection fitness was re-screened to determine the colony forming unit (cfu) size of the WT and mutant RsRubisco library. The mutant pYZrsLS plasmids from 10 colonies grown on permissive media were transformed into XL1-Blue (LB-amp selected), purified and fully sequenced. The Rs-rbcL mutualization frequency was 2.4 mutations per kb.

The remaining transformed RDE cells were pelleted (6000× g, 1 min), suspended in 0.2 mL LB and plated onto LB-arabinose selection (LB-chlor-amp plates containing 125 µg/mL kanamycin, 75 µM IPTG and 0.25% (w/v) arabinose, Figure 5b). The RDE3-WT and RDE3-mutant RsRubisco library plates were grown at 25 ºC for 3–7 days in air containing 2% (v/v) CO<sub>2</sub>. All colonies from both plates were colony picked and grown in 1 mL LB-chlor-amp, their plasmids were purified and the pYZrsLS re-transformed into XL1-Blue (LB-Amp selection), and then purified for NdeI mapping (expect 3822, 2177 and 1020-bp products) before transforming into RDE3 for the re-screening of selection fitness.

Colonies of the RDE3-rescreened cells were grown to an OD<sub>600</sub> of 0.6 in 1 mL LB-chlor-amp, diluted 10<sup>−4</sup> with LB and 20 µL of cells plated onto LB-arabinose selection (Figure 5c). Of the 41 plasmids re-screened (including eight from the WT screen, Figure 5b) only one, the pYZ1 mutant, improved RDE3 growth on the LB-arabinose (0.25% w/v, Figure 5c). The Rs-rbcL operon in pYZ1 contained five nucleotide substitutions that coded four RbcL amino acid changes, from which nine YZ1 RsRubisco derivatives were generated by restriction cloning so that each chimer only coded one to three of each mutation (Figure 5d).

4.3. Rubisco Content and Catalysis

JM109(DE3) transformed with pYZrsLS and each pYZ1 derivative were grown at 28 ºC and RsRubisco expression induced for 6 h with 0.5 mM IPTG before harvesting by centrifugation (10 min,
4 °C, 6200×g) and replica cell pellet aliquots N2 frozen and stored at −80 °C (Figure 3c). The soluble and total (lysate) proteins in cell pellets from 10 mL of cultured cells were analyzed by SDS-PAGE, native PAGE and immunoblot analysis as described [55]. The Rubisco content in the E. coli soluble protein was quantified by 14C-CABP binding as described [38]. The same soluble protein sample was used to measure $k_{\text{cat}}^C$ and $K_C$ using 14CO2-fixation assays as described [22] using 0–170 µM 14CO2 in assays equilibrated with 0, 20%, 35% or 50% O2 (v/v) in N2. The increase in $K_C$ with increasing [O2–] was plotted and the slope of the linear fit used to derive $K_O$. $S_{\text{CO2}}$ was quantified using the [1-3H]-RuBP consumption assay as described [41] using RsRubisco purified from ~40 mL of cultured cells (Figure 3c) by rapid ion exchange and size exclusion (Superdex 200, GE Life Science, Chicago, IL, USA) as described [42]. Soluble protein concentration was measured using the dye-binding Coomassie Plus Kit (Thermo Fisher Scientific, Waltham, MA, USA) against bovine serum albumin.

5. Conclusions

This study provides another demonstration of how directed evolution can improve the carboxylase properties of Rubisco. As shown here, key challenges with RDE screens include differentiating the Rubisco mutants that enhance cell fitness from false positives and then distinguishing carboxylation enhancing mutations from those that improve Rubisco biogenesis. Success is also dependent on tailoring selection screen conditions to the substrate Rubisco being evolved; that is, determining the optimal arabinose and IPTG concentrations for PRK and Rubisco production, keeping in mind that Rubisco production itself impairs E. coli viability [25]. An ongoing consideration is the extent to which the red-type RsRubisco might serve as a suitable surrogate to sequentially evolve its properties towards the high $S_{\text{CO2}}$, high carboxylation efficiency properties of the related “better” red algae Rubisco. One possible objective might be to replace plant Rubisco with a kinetically improved RsRubisco, although this will depend on whether its folding and assembly requirements are met by plant chloroplasts. Current evidence suggests the extent of prokaryotic Rubisco expression in E. coli is a poor indicator of its biogenesis potential in tobacco [20–22]. An alternative to evolving RsRubisco might be to evolve plant Rubisco itself using an RDE screen incorporating the chloroplast protein folding machinery [53,56]. As the components of the plant Rubisco expression systems are primarily T7-promoter regulated [37,57], our JM109(DE3)-based RDE3 screen appears to be a suitable selection host for such a directed evolution endeavor.

Author Contributions: Y.Z. and S.W. contributed equally to the conceptualization, formal analysis, and writing of the article. Funding was awarded to S.W.

Funding: This research was supported by the Australian Government through the Australian Research Council Centre of Excellence for Translational Photosynthesis CE140100015.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

Abbreviations

- Raf1: Rubisco accumulation factor 1
- Raf2: Rubisco accumulation factor 2
- BSD2: Bundle Sheath Defective II Rubisco chaperone
- RbcX: Rubisco specific chaperone
- cfu: Colony forming units
- RuBP: Ribulose-1,5-bisphosphate
- $S_{\text{CO2}}$: Specificity for CO2 over O2
- $k_{\text{cat}}^C$: Substrate saturated carboxylation rate
- $k_{\text{cat}}^O$: Substrate saturated oxygenation rate
- $K_C$: Apparent $K_m$ for CO2
- $K_{C21\%O2}$: Apparent $K_m$ for CO2 under ambient (20.6% (v/v)) O2
- $K_O$: Apparent $K_m$ for O2
- PRK: Phosphoribulokinase
Appendix A

(a) Rbcl alignment

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\begin{array}{l}
1 \quad - \quad M \quad S \quad P \quad Q \quad T \quad E \quad T \quad K \quad A \quad S \quad V \quad G \quad F \quad K \quad A \quad G \quad V \quad K \quad E \quad Y \quad K \quad L \quad T \quad - \quad Y \quad T \quad P \quad E \quad Y \quad Q \quad T \quad D \quad D \quad L \quad L \quad A \quad \text{tob-L} \\
1 \quad - \quad P \quad K \quad T \quad Q \quad S \quad - \quad - \quad A \quad Y \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \quad . \ quartet. 
\end{array}
\]

Figure A1. Cont.
Alignments of (a) RbcL and (b) RbcS for Rubisco from Nicotiana tabacum (tob-L, Z00044; tob-S, XP_016440853), Synechococcus elongatus PCC6301 (SeRbcL, P00880.2; SeRbcS P04716), Thermosynechococcus elongatus BP1 (TeRbcL, NP_682296.1; TeRbcS NP_682294.1) and Rhodobacter sphaeroides (RsRbcL, WP_002721829.1; RsRbcS WP_002721826.1). The first 39 amino acids of the altered codon modified (cm) RsRbcL N-terminus coded in pYZRsLS are shown in blue text. Sequences were Clustal W-aligned, with only residues differing from the tob sequences shown. Dashes were included to improve the alignment of any missing residues. Amino acid numbering is relative to tob-L and tob-S. Coloring is as indicated in the legend under panel (b).

Figure A1. Alignments of (a) RbcL and (b) RbcS for Rubisco from Nicotiana tabacum (tob-L, Z00044; tob-S, XP_016440853), Synechococcus elongatus PCC6301 (SeRbcL, P00880.2; SeRbcS P04716), Thermosynechococcus elongatus BP1 (TeRbcL, NP_682296.1; TeRbcS NP_682294.1) and Rhodobacter sphaeroides (RsRbcL, WP_002721829.1; RsRbcS WP_002721826.1). The first 39 amino acids of the altered codon modified (cm) RsRbcL N-terminus coded in pYZRsLS are shown in blue text. Sequences were Clustal W-aligned, with only residues differing from the tob sequences shown. Dashes were included to improve the alignment of any missing residues. Amino acid numbering is relative to tob-L and tob-S. Coloring is as indicated in the legend under panel (b).

Figure A2. Restriction mapping of pACYC PRK:Kan. (a) The plasmid map and (b) DraI restriction mapping of the source pACYC PRK:Kan plasmid (control) and that purified from three separate colonies of the E. coli strains indicate that they survived growth on selective media containing 0.25% (w/v) L-Arabinose (Figure 4).
Table A1. Summary of directed evolution Rubisco mutants and the content and catalytic properties selected using *E. coli* and *Ralstonia eutrophus* screens.

| Rubisco Origin | Mutation(s) | Plant Rcbl. Numbering | K_T (mM) | K_P ^2kT/2oz (µM) | k_cat (s^-1) | k_cat (µM^-1) | k_cat/O2 (s^-1) | S_c (µM) | K_m (µM) | Rubisco (%)/CSP | RDE Strain |
|----------------|-------------|------------------------|----------|-------------------|--------------|--------------|----------------|----------|----------|----------------|-------------|
| *Synechococcus* | WT | - | 273 | - | 11.4 | 42 | - | - | - | 44.0 | 64 | 1.0 | **|** |
| PCC6301 (LpS) | M259T | 262 | 257 | - | 12.8 | 54 | - | - | - | 43.0 | 67 | 6.0 | E. coli | Ref [22] |
| | A83M/M259T | 10/262 | 216 | - | 11.4 | 51 | - | - | - | 41.0 | 48 | 5.5 | **|** |
| | F342S/M259T | 345/262 | 207 | - | 10.0 | 48 | - | - | - | 38.0 | 34 | 3.5 | **|** |
| *Synechococcus* | WT | - | 200 | 242 | 11.8 | 59 | 48.7 | 1191 | 1.7 | 1.42 | 41.5 | 44 | 0.2 | **|** |
| PCC6301 (LpS) | F345I | 345 | 208 | 266 | 10.1 | 49 | 38.0 | 908 | 1.2 | 1.33 | 36.6 | 31 | 3.4 | **|** |
| | F345I | 345 | 245 | - | 9.8 | 40 | - | - | - | 40.0 | 93 | 1.9 | **|** |
| | Q212L | 212 | 370 | 444 | 9.9 | 27 | 22.3 | 1252 | 0.9 | 0.72 | 37.4 | 31 | 1.7 | **|** |
| | L548M | 348 | 287 | - | 11.3 | 39 | - | - | - | 35.1 | 84 | - | **|** |
| | N115S | 115 | 191 | - | 9.0 | 47 | - | - | - | 43.5 | 44 | - | **|** |
| | M262T | 262 | - | - | - | - | - | - | - | - | - | - | **|** |
| | M169L | 169 | - | - | - | - | - | - | - | - | - | - | **|** |
| | L161M | 161 | - | - | - | - | - | - | - | - | - | - | **|** |
| *Synch-P7002* | WT | - | 132 | - | 6.7 | 51 | - | - | - | 52 | - | - | BL2I/DEU | Ref [58] |
| | E49V/D92C | - | 200 | 14.6 | 74 | - | - | - | 50 | - | - | - | **|** |
| | E51V/D96C | - | 200 | 14.6 | 74 | - | - | - | 50 | - | - | - | **|** |
| *Synechococcus* | WT | - | 248 | - | 12.9 | 52 | - | - | - | 0.9 | 56.1 | - | - | **|** |
| PCC6301 (LpS) | F140I | 140 | 129 | - | 19.4 | 150 | - | - | - | 2.9 | 51.3 | - | - | **|** |
| | C172G | 172 | 493 | - | 12.8 | 26 | - | - | - | 0.4 | 59.6 | - | - | **|** |
| | V189I | 189 | 160 | - | 14.3 | 89 | - | - | - | 1.8 | 49.2 | - | - | **|** |
| | V221H | 221 | 181 | - | 2.5 | 14 | - | - | - | 0.3 | 57.9 | - | - | **|** |
| | V346L | 346 | 256 | - | 4.3 | 17 | - | - | - | 0.3 | 56.9 | - | - | **|** |
| | E49V/D92C | 345 | 274 | - | 7.2 | 26 | - | - | - | 0.5 | 54.4 | - | - | **|** |
| | S99C | 398 | 322 | - | 18.6 | 58 | - | - | - | 1.0 | 56.2 | - | - | **|** |
| | I46V | 465 | 298 | - | 14.3 | 48 | - | - | - | 0.9 | 53.7 | - | - | **|** |
| | F345I | 345 | 274 | - | 11.2 | 41 | - | - | - | 0.8 | 52.1 | - | - | **|** |
| | F140I/F345I | 140/345 | 218 | - | 15.6 | 72 | - | - | - | 1.2 | 59.3 | - | - | **|** |
| *Thermotoga* | WT | - | 107 | 6.6 | 6.7 | 70 | - | - | - | 53.3 | 36 | 6.6 | **|** |
| elongatus (LpS) | F345I | - | 101 | 6.9 | - | 47 | - | - | - | 48.0 | - | 14.6 | **|** |
| | P415A | 415 | - | 111 | 6.9 | - | 62 | - | - | 52.6 | - | 10.4 | **|** |
| | V300A | 300 | - | 135 | 6.6 | - | 49 | - | - | 51.9 | - | 7.1 | **|** |
| | F345/P415A | 345/415 | - | 102 | 4.8 | - | 47 | - | - | 49.2 | - | 17.5 | **|** |
| | P415A/V88M | 415/88M | - | 93.5 | 95 | 100 | - | 56.3 | - | 20.4 | **|** |
| | P415A/A48V | 415/A48V | - | 105 | 8.8 | - | 84 | - | - | 48.0 | - | 9.7 | RDE2 | **|** |
| | P415A/H37L | 415/H37L | - | 100 | 8.4 | - | 84 | - | - | 49.7 | - | 11.1 | XL1-B | **|** |
| | P415A/Y306K/C312D | 415/Y306K/C312D | - | 94 | 9.3 | - | 99 | - | - | 47.7 | - | 9.7 | **|** |
| | P415A/Y36N/R51H | 415/Y36N/R51H | - | 120 | 8.0 | - | 68 | - | - | 51.8 | - | 13.6 | **|** |
| | P415A/A398T/A48V | 415/A398T/A48V | - | 105 | 8.1 | - | 75 | - | - | 45.5 | - | 13.3 | **|** |
| | P415A/L474M/L397N | 415/L474M/L397N | - | 92 | 8.2 | - | 90 | - | - | 51.8 | - | 12.5 | **|** |
| | P415A/A44T | 415/A44T | - | 102 | 8.0 | - | 78 | - | - | 55.8 | - | 13.9 | **|** |
| | P415A/Z93M | 415/Z93M | - | 90 | 7.8 | - | 87 | - | - | 48.1 | - | 16.6 | **|** |
| | P415A/L93T | 415/L93T | - | 110 | 6.4 | - | 59 | - | - | 50.5 | - | 16.6 | **|** |
| *Synechococcus* | WT | - | 231 | 12.6 | 50 | - | - | - | 46.0 | 44 | 1.3 | **|** |
| PCC6301 (LpS) | F345I | 345 | - | 285 | 10.3 | - | - | - | 42.0 | - | 14.4 | **|** |
| | F140I | 140 | - | 211 | 11.1 | - | - | - | 46.0 | - | 6.3 | **|** |
| | V189I | 189 | - | 186 | 10.6 | - | - | - | 39.0 | NA | 6.2 | **|** |
Table A1. Cont.

| Rubisco Origin          | Mutation(s) | Plant RbcL Numbering | \(K_c\) (µM) | \(K \text{cat}^{1/2}\text{O}_2\) (s\(^{-1}\)) | \(k_o\) (s\(^{-1}\) mM\(^{-1}\)) | \(k_o / K_c\) (s\(^{-1}\)) | \(K_o\) (µM) | \(k_o / K_o\) (s\(^{-1}\) mM\(^{-1}\)) | \(S_{CO}\) (µM) | \(K_o\text{spar}\) (µM) | Rubisco (%CSP) | RDE Strain |
|-------------------------|-------------|----------------------|--------------|-----------------------------------------------|-------------------------------|-----------------|----------|-----------------------------------------------|-----------------|-----------------|-----------------|------------|
| *Rhodospirillum Rubrum* |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| (L\(_2\))              |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| Rhodospirillum Rubrum   |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| (L\(_2\))              |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| WT                     |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| MM2                   |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| E138V                  |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| K32E                   |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| T421A                  |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| H471N                  |             |                      |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| G327S                  |             | 331/337/332          |              |                                               |                               |                 |          |                                               |                 |                 |                 |            |
| %CSP, Percentage (w/w) cell soluble protein; Red text—kinetic values corrected from those published in Mueller-cajar et al., 2008; (-) value not measured.
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