Computational modeling of the effects of auditory nerve dysmyelination

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INTRODUCTION

The rapid conduction of action potentials in both the central nervous system (CNS) and peripheral nervous system (PNS) depends on the myelin sheath around neuronal axons. Myelin plays a passive role by insulating axons. Myelin also plays an active role, allowing for saltatory conduction as action potentials regenerate via active, voltage dependent Na⁺ and K⁺ ion-specific conductances at the intermittent, non-myelinated nodes of Ranvier (Ranvier, 1871; Huxley and Stampfli, 1949; Rasband and Trimmer, 2001). The fundamental importance of myelin is highlighted in demyelinating diseases where the speed of conduction along axons is altered and sensory, motor and/or cognitive ability is severely compromised (Nave, 2010). The auditory nerve (AN) contains mainly myelinated axons projecting from type I spiral ganglion neurons to brainstem cochlear nuclei (Toesca, 1996). We recently showed that acoustic overexposure (AOE) leading to hearing loss also triggered notable morphological changes at AN myelin sub-domains such as nodes of Ranvier, paranodes and juxtaparanodes (Poliak and Peles, 2003) that were associated with the decreased conduction velocity (Tagoe et al., 2014). Specific changes involved a decreased number of lamella wraps and hence myelin thickness, a marked elongation of the nodes and juxtaparanodes, and a retraction of the paranodes (Tagoe et al., 2014). Precise localization of axonal ion channels is crucial for proper electrical and chemical functions of axons. It is unknown whether AOE-induced deficit in conduction velocity (Tagoe et al., 2014) is due to re-distribution of voltage-gated channels from specific myelin sub-domains. At the nodes of Ranvier, clusters of voltage-gated sodium channels facilitate saltatory conduction of action potentials (Rasband and Trimmer, 2001; Hedstrom and Rasband, 2006; Leterrier et al., 2010). Kv3.1 voltage-gated potassium (Kv) channels contributing to the ability of auditory neurons to fire at high frequencies (Kanemasa et al., 1995; Macica et al., 2003) are also clustered at the nodes (Devaux et al., 2003). Kv1.1 voltage-gated potassium (Kv) channels reduce action potential jitter (Gittelman and Tempel, 2006) and determine action potential threshold (Brew et al., 2003; Oertel et al., 2008), are clustered in the juxtaparanodal regions (Gu and Gu, 2011). Here we modeled changes in the AN conduction properties after AOE based on our recent data (Tagoe et al., 2014) and tested whether the decreased conduction velocity observed after AOE (Tagoe et al., 2014) could also be due to the redistribution of voltage gated Na⁺, Kv3.1 and Kv1.1 channels.

METHODS

COMPUTER SIMULATIONS

Simulations were carried out using Neuron 7.1 (Hines and Carnevale, 1997) and were based on previously published morphological data on the AN (Tagoe et al., 2014). Briefly, the AN was modeled as a single axon unsheathed by myelin lamellae in a manner similar to that described previously (Kolaric et al., 2013). The axon comprised narrow nodal regions separated by larger internodal regions (INR, Figure 1A). Passive electrical properties were based on data from corpus callosum axons and oligodendrocytes (Bakiri et al., 2011) and voltage dependent conductances were based on previous studies including an existing model of the AN (Kanemasa et al., 1995; Macica et al., 2003) (Figures 1A,B). Current densities were derived from values in Kanemasa et al. (1995) and Macica et al. (2003) where maximum current amplitude was given relative to a cell capacitance (Cm), based on a specific Cm of 1 μF.cm⁻². The simulated action potentials were computed using backward Euler integration with a time step of 10 ms. Our morphological measurements lack a value for the internodal length, thus we chose an intermediate value from two

Our previous study showed that exposure to loud sound leading to hearing loss elongated the auditory nerve (AN) nodes of Ranvier and triggered notable morphological changes at paranodes and juxtaparanodes. Here we used computational modeling to examine how theoretical redistribution of voltage gated Na⁺, Kv3.1, and Kv1.1 channels along the AN may be responsible for the alterations of conduction property following acoustic over-exposure. Our modeling study infers that changes related to Na⁺ channel density (rather than the redistribution of voltage gated Na⁺, Kv3.1, and Kv1.1 channels) is the likely cause of the decreased conduction velocity and the conduction block observed after acoustic overexposure (AOE).

Keywords: myelin sheath, hearing loss, node of Ranvier, conduction velocity, conduction block, deafness, myelin domains, action potential
FIGURE 1 | Schematic model of the auditory nerve. (A) The auditory nerve fiber is divided into three distinct regions. The peripheral axonal region terminates in the organ of Corti. The soma resides in the spiral ganglion. The central axon projects to the cochlear nucleus, is myelinated with internodal myelinated regions modeled as 100 μm in length. (B) Equivalent circuit of the central portion of the auditory nerve, which has been adapted from an existing model of corpus callosum axon (Tagoe et al., 2014). The morphological and electrical values are contained in Tables 1, 2 respectively. The axon is divided into the nodal region and the internodal region. The internodal region is subdivided into the paranodal (PN) juxtaparanodal (JP) and axonal regions (axon). The nodal region expresses the voltage-dependent conductances of the INa (gNa) and IHT (gIHT) currents as well as a leak current (gL). GiLT is expressed in the JP. The axolemma of the internodal regions expresses a leak current (gL), as does the overlying myelin (gmyl). The dotted lines enclose the leak and capacitative properties of each INR component. Internal resistance (Ra) is constant throughout the model and external resistance (Re) is zero. The dark region represents the myelin, and only one PN and JP abutting the node are shown for clarity. gmyl and Cmyl are the passive conductance and capacitance across the myelin, respectively. gL is the passive conductance, gL(N) refers to the passive conductance at the node, Epas is the reversal potential for the passive conductance (Vl), and ENa and EK are the reversal potentials for the Na+ current and ILT and IHT respectively.

existing models of AN (Rattay et al., 2001; Smit et al., 2010) of 100 μm.

MORPHOLOGICAL VALUES
The morphological dimensions of the axonal compartments have recently been published (Tagoe et al., 2014). The model of the control axon comprised alternating nodal and internodal regions (INR). The INRs were subdivided into paranodal (PN), juxtaparanodal (JP) and axonal compartments (Figure 2A), such that each INR comprised two PNs, each abutting consecutive nodal regions, two JP regions located between the PN and a central axonal portion (Figure 2A). The dimensions of the compartments are contained in Table 1. AOE changed the nodal length and diameter from 1.3 to 6.15 μm and 0.8 to 1.28 μm, respectively; the PN length and diameter from 2.34 to 1.52 μm and 0.75 to 1.23 μm respectively and the JP length and diameter from 5.14 to 6.23 μm and 2.13 to 1.32 μm respectively (Tagoe et al., 2014). The model contained a fast sodium current (INa), a high threshold (IHT) Kv3.1 potassium current, a low threshold (ILT) Kv1.1 potassium current and a leak current (IL). We assumed that the current density for INa and IHT in the nodal region were 6.6 mS.cm⁻² and 1.98 mS.cm⁻², respectively, and the value for ILT in the PN was 2.13 mS.cm⁻² (Kanemasa et al., 1995).

We used these morphological changes to determine conduction properties under two conditions. Firstly, we modeled the effect of altering current distributions such that the compartments resulting from AOE treatment contained the same number of channels, but the density of these channels was altered to reflect an even distribution of the channels along the altered nodal, PN and JP length (AOE1 condition illustrated in Figure 3A). This condition named AOE1 resulted in morphological values contained in Table 1. Secondly, we modeled the axon as if the channels had remained in place and did not encroach on the AOE-induced changes in compartment size (AOE2 condition illustrated in Figure 3A). In this condition, the nodal compartment had the same altered dimensions as the nodal compartment in AOE1. However, the expression of INa and IHT remained the same as the control condition. In AOE2 the regions n1 contained no voltage dependent channels and were sized such that the combined values of the two n1 regions and the nodal region equaled the value of the nodal region in Table 1. Similar calculations were carried out for the PN and JP and considered the PN length decrease the JP length increase after AOE (Tagoe et al., 2014).
FIGURE 2 | Action potential conduction along the central portion of the auditory nerve. (A) Schematic model of the axonal compartments in the control auditory nerve model. (B) Evoked action potentials recorded at six successive nodes illustrating action potential conduction along an axon. Scale bar is 50 mV and the duration of the recording is 2 ms. (C) The conduction velocity decreases as internodal length (INL) is increased from the control value of 100 μm.

Table 1 | Morphological parameters of the auditory nerve in control and in the two simulated AOE conditions.

|        | Node | n1  | PN  | JP  | j1  | Axon |
|--------|------|-----|-----|-----|-----|------|
| CONTROL| 1.3  | –   | 2.34| 5.14| –   | 10.46|
| d (μm) | 0.8  | –   | 0.75| 2.13| –   | 1.25 |
| AOE1   | 6.15 | –   | 1.52| 6.23| –   | 9.79 |
| d (μm) | 1.28 | –   | 1.23| 1.32| –   | 1.25 |
| AOE2   | 1.3  | 2.425| 2.34| 5.14| 1.09| 9.79 |
| d (μm) | 1.28 | 1.28| 1.23| 2.13| 1.16| 1.25 |

Morphological dimensions of the various compartments illustrated in Figures 2A, 3A, where l and d refer to length and diameter and p/j/a refer to paranodal, juxtaparanodal and axonal compartments. Axon dimensions are taken from previously published data Kolaric et al. (2013). The length indicates the compartments length of the axon used in the model with 20 compartments per internodal length.

Each INR was divided into compartments such that the length of each compartment was less than 0.1 λ (length constant), to ensure each compartment was isopotential, the generally accepted practice in such simulations (Carnevale and Hones, 2006; Sterratt et al., 2011). The length constant (λ) was calculated as:

$$\lambda = \sqrt{\frac{\text{rad} \times R_m}{2 \times R_a}}$$

where rad is the axon radius, Ra is the axoplasmic resistance and Rm is the membrane resistance.

Table 2 | Passive properties of the auditory nerve in control and after AOE.

|        | Control | AOE  |
|--------|---------|------|
| Cm p/j/a (μF.cm⁻²) | 0.0184 | 0.0290 |
| Cm node (pF) | 0.0327 | 0.247 |
| Cm INR (pF) | 0.136 | 0.167 |
| gL node (mS.cm⁻²) | 10.89 | 20.3 |
| gL p/j/a (μS.cm⁻²) | 0.2 | 0.2 |

Passive properties of nodal and INR compartments in control and AOE treated nerves. p/j/a refers to paranodal, juxtaparanodal and axonal regions, all of which express the same values. G refers to conductance and Cm to capacitance. The capacitance and leak current were calculated for each individual INR and node demonstrating that AOE causes an increased INR capacitance due to decreased myelin wraps and increased nodal capacitance and leak current.

PASSIVE MEMBRANE PROPERTIES

The basic passive properties and the methods used to calculate these properties have previously been described in detail (Kolaric et al., 2013). The value of axoplasmic resistance (Ra) was 70 Ω.cm.
throughout. Cm was assumed to be 1 μF.cm⁻² and nodal gL was 0.2 mS.cm⁻². In the model the control axon had 23.7 lamella wraps of myelin, whereas AOE decreased the number to 15.8 (Tagoe et al., 2014). The values were rounded-up to the nearest whole number to ease computations.

**VOLTAGE-DEPENDENT CONDUCTANCES**

The model described in Hodgkin and Huxley (1952a,b,c) contained a fast sodium current (INa), a high threshold (IHT) Kv3.1 potassium current, a low threshold (ILT) Kv1.1 potassium current and a small leak current (IL). INa and IHT were located at the nodal regions and ILT was located at the JPN (7).

The voltage dependence of the IHT was described by:

\[ I_{HT} = g_k n^3 (1 - \gamma + \gamma p) (V_m - V_{HT}) \]

where \( \gamma > 1 \), and the variables \( p \) and \( n \) are described by \( d j/dt = \alpha_j (1 - j) - \beta_j \) where \( j = n \) or \( p \), and is calculated by \( \alpha_j = k_{\alpha j} \exp(\eta_{\alpha j} V) \) and \( \beta_j = k_{\beta j} \exp(\eta_{\beta j} V) \).

\( k_{\alpha j}, k_{\beta j}, \eta_{\alpha j} \) and \( \eta_{\beta j} \) determine the rate and voltage dependence respectively of current activation with the values required to calculate the rate constants contained in Table 3.

The voltage dependent currents INa and ILT were described by:

\[ I_{Na} = g_{Na} N^3 (V_m - V_{Na}) \]
\[ I_{LT} = g_L I_R (V_m - V_{LT}) \]

where \( I \) is the current per unit area and \( g \) is the voltage dependent conductance where \( g_{Na} = 0.05 \text{ S.cm}^{-2} \), \( g_{HT} = 0.015 \text{ S.cm}^{-2} \) and \( g_{LT} = 0.002 \text{ S.cm}^{-2} \). \( V_m \) is the membrane potential and \( V \) refers to the reversal potential whereby \( V_{HT} = V_{LT} = -80 \text{ mV} \) and \( V_{Na} = 50 \text{ mV} \).

The leak current was calculated as \( I_L = g_L (V_m - V_L) \), where \( g_L = 0.2 \text{ mS.cm}^{-2} \) and \( V_L = -63 \text{ mV} \).

The variables \( m, h, l \) and \( p \) are associated with channel activation (\( m, l \)) and inactivation (\( h, r \)) and are calculated as \( dj/dt = \alpha_j (1 - j) - \beta_j \) where \( j = l, r, m \) or \( h \). The rate and voltage dependence respectively of INa and ILT are calculated in a similar manner to IHT with the parameters for calculating ILT and INa contained in Table 3.

**RESULTS**

**ACTION POTENTIAL CONDUCTION**

The model comprising 7 alternating nodal and internodal regions and conduction velocity was calculated between the first (node 0) and last node (node 6). When the conduction velocity was reported in function of the number of INRs affected (Figure 3D), the conduction velocity at node 0 was equivalent to the control condition.

**AOE-INDUCED DECREASE IN CONDUCTION VELOCITY**

We adjusted the model to take into account the morphological changes that occur because of AOE. In our first simulation of the effects of AOE (AOE1), we kept the current density that would result from altered distribution of channels along the node and JPN (see Methods for details). We found that there was a significant effect on action potential propagation (Figure 3B) compared to control (Figure 2B) resulting in a decreased conduction velocity (Figures 3D,E). The results illustrated in Figure 3D derive from simulations where we imposed the AOE-induced dysmyelination on increasing numbers of adjacent INRs in the model. Incrementally increasing the number of INRs affected by AOE dysmyelination showed a non-linear decrease in conduction velocity that steeply decreased with increasing number of INRs affected, as previously described in the corpus callosum model (Kolaric et al., 2013).

In the second simulation we modeled unchanged current density and channel distribution as a result of AOE (AOE2, see Methods). Similarly to AOE1, we found that there was a significant effect on action potential propagation (Figures 3B,C) compared to control condition (Figure 2B). Conduction velocity was similarly decreased in AOE2 and AOE1 (Figure 3E) and the decreased conduction velocity was more apparent as the number of adjacent INRs affected increased (Figure 3D).

**ABSENCE OF EFFECT OF AOE ON FIRING FREQUENCY**

Repetitive firing was induced in the control model by injecting a current pulse of 0.35 nA for a duration of 200 ms (Figure 4A).
The resulting action potentials showed regular firing pattern, i.e., a lack of adaptation. Increasing the stimulus intensity resulted in increased frequency of firing, although at higher stimulus intensities the action potentials became much smaller. Imposing the same stimulus on the AOE1 and AOE2 models had a negligible effect of firing frequency (Figure 4B).

**EFFECTS OF gNa EXPRESSION ON CONDUCTION VELOCITY**

Decreasing the density of Na\(^+\) channel expression by attenuating the value of gNa caused a non-linear decrease in conduction velocity until conduction block occurred at a value of about 35% of control (Figure 5A). The passive leak conductance across the node (gL) under control and both AOE conditions are superimposed to show there are slight effects on gL as a result of AOE but these differences are minimal (Figure 5B) and do not indicate increased shunting of current via the node.

**DISCUSSION**

The present computer simulations are based on our previous morphological data on the AN (Tagoe et al., 2014). We have shown that the conduction velocity was dependent on the INR length as previously described in the corpus callosum model (Kolaric et al., 2013) and this over a range of between 0.05 and 0.3 mm. Our estimated conduction velocities of the AN in the range of 2–12 m.s\(^{-1}\) were similar to values (6–14 m.s\(^{-1}\)) reported for the AN (Poma et al., 2008; Imennov and Rubinstein, 2009). Huxley and Stampfli (1949) suggested that the conduction velocity in myelinated nerve fibers should reach a maximum at a particular internode distance, and that the maximum should be relatively flat (Huxley and Stampfli, 1949). We also found that the conduction velocity in myelinated AN fibers did not increase linearly with increasing internodal length and reached a plateau at an internodal length of 0.3 mm. Our previous field potential recordings of the AN estimated its conduction velocity at 3 m.s\(^{-1}\) (Tagoe et al., 2014). Although this lower range of values could be due to an AN internodal length of around 200 \(\mu\)m, it is likely to be due to the recruitment of lower conduction velocity fibers while recording compound action potentials.

Our previous study showed that dysmyelination of the AN resulted in the decrease of the myelin thickness, a marked elongation of nodes of Ranvier and juxtaparanodes, and a retraction of paranodes, slowing the AN conduction velocity by about 3-fold (Tagoe et al., 2014). Using the same morphological values as in Tagoe et al. (2014) and keeping the internodal length of 100 \(\mu\)m (Rattay et al., 2001; Smit et al., 2010) constant, we have reproduced here a 3-fold slowing of conduction velocity. This result was anticipated considering that the elongation of the node would increase its capacitance and decrease its resistance, and therefore...
increase the time to reach the threshold contributing to lower conduction velocity (Hartline and Colman, 2007). The decreased conduction velocity was more apparent with the number of INR affected, as the increased nodal surface area resulted in a decreased current density, increased nodal and INR capacitances (Table 2). As the conduction velocity decreases with increasing length of dysmyelinated axon, we would anticipate conduction block for distances exceeding 700 μm (Figure 3D) as a significant length of axon would be affected. Conduction block has indeed been observed for distances exceeding 1 mm (Tagoe et al., 2014).

Nodal clusters of voltage-gated Na+ channels are lost after demyelination, but reappear after remyelination (Rasband et al., 1998). Deficits of myelin subdomains along the AN could therefore alter spatial segregation of voltage-gated channels and impair the action potential propagation (Peles and Salzer, 2000; Hossain et al., 2005). However we found that spatial segregation of nodal Na+ and Kv3.1 channels or juxtaparanodal Kv1.1 channels had a minimal impact on AN conduction velocity. Axonal targeting of Kv3.1 channels is critical to enable neurons to fire action potentials at their maximal frequency (Gu et al., 2012). Our study further suggests that the re-localization of Kv3.1 channels is unlikely to affect action potential firing frequency during AN dys-myelination. Axonal demyelination has been shown to decrease nodal expression of INa (Craner et al., 2004). Our computational analysis indicates that decreasing the Na+ channel density has a negative impact on the conduction velocity and that decreasing the Na+ channel density by 65% relative to control values can fully account for the conduction block observed after AOE.

In conclusion the AOE induced morphological alterations of myelin sub-domains and changes of density of Na+ channels are the likely cause of the decreased conduction velocity of the AN and the conduction block observed for distances exceeding 1 mm (Tagoe et al., 2014).

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