A Unique Capsule Locus in the Newly Designated *Actinobacillus pleuropneumoniae* Serovar 16 and Development of a Diagnostic PCR Assay

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ABSTRACT *Actinobacillus pleuropneumoniae* causes pleuropneumonia, an economically significant lung disease of pigs. Recently, isolates of *A. pleuropneumoniae* that were serologically distinct from the previously characterized 15 serovars were described, and a proposal was put forward that they comprised a new serovar, serovar 16. Here we used whole-genome sequencing of the proposed serovar 16 reference strain A-85/14 to confirm the presence of a unique capsular polysaccharide biosynthetic locus. For molecular diagnostics, primers were designed from the capsule locus of strain A-85/14, and a PCR was formulated that differentiated serovar 16 isolates from all 15 known serovars and other common respiratory pathogenic/commensal bacteria of pigs. Analysis of the capsule locus of strain A-85/14 combined with the previous serological data show the existence of a sixteenth serovar—designated serovar 16—of *A. pleuropneumoniae*.

KEYWORDS *Actinobacillus pleuropneumoniae*, PCR, diagnostics, serovar 16

*Actinobacillus pleuropneumoniae* is an economically important pathogen in the swine industry, causing peracute to chronic pleuropneumonia (1, 2). Isolates can be classified into two biovars depending on their requirement for NAD, with the majority of isolates belonging to biovar I (require NAD for growth) and biovar II isolates (NAD independent) less frequently identified (3). Isolates can be further differentiated into serovars, based mainly on capsular polysaccharide (CPS) structure (3, 4). Until recently, there were 15 recognized serovars with different geographical distributions (3, 5). Knowing the serovar is important for epidemiological tracking as well as bacterin-based vaccine formulation pertinent to the geographical area (3). The complete CPS biosynthetic loci of these 15 serovars have been determined either specifically (6–8) or as part of whole-genome sequence analysis (9–11). In all 15 serovars except serovar 15, the genes of the CPS biosynthetic locus are divergently transcribed from those of the export locus, cpxDCBA (6, 10); in serovar 15, the CPS biosynthetic genes are in the same orientation as, and immediately precede, those of the export locus (7).

Xu et al. (10) compared the CPS loci for serovars 1 to 7 and 9 to 13 and showed that they could be grouped into three types (see Fig. 1 for representative loci for each CPS type), in agreement with previous structural studies (4). The loci for serovars 2, 3, 6, 7, 8, 9, 11, and 13 (type I) have a common core, with the first three genes (*cps2ABC*)...
sharing a high degree of identity and encoding teichoic acid transferase, glycerol transferase, and glycerol-3-phosphate cytidylyltransferase, respectively (8, 10). The type II CPS loci, initially identified in serovars 1, 4, and 12, share the first gene (cps1A) encoding a CPS phosphotransferase (10), which is also seen in the subsequently sequenced serovar 14 CPS locus (6). Although not highly conserved at the nucleotide level, cps15A (accession no. BAR72992), the first gene in the serovar 15 CPS locus, also encodes a CPS phosphotransferase that shares 62% identity with cps1A, suggesting that serovar 15 also produces a type II CPS (7).

In the type III capsule loci, seen in serovars 5 and 10, it is the final three genes that are common, encoding 3-deoxy-8-phosphooctulonate synthase, 3-deoxy-manno-octulosonate cytidylyltransferase, and arabinose-5-phosphate isomerase, respectively.

It was recently reported that five Hungarian isolates of *A. pleuropneumoniae* could not be assigned to any of the known 15 serovars but rather formed a distinct serological group, and it was proposed that they constituted a new serovar of *A. pleuropneumoniae*—serovar 16 (12). Here, we used whole-genome sequencing to determine the genetic organization of the CPS biosynthesis locus for strain A-85/14, the proposed reference strain for serovar 16. The sequencing data show that the serovar 16 CPS locus is distinct from that of the other known 15 serovars. Furthermore, on the basis of this unique sequence, we have designed primers for specific detection of *A. pleuropneumoniae* serovar 16.

**RESULTS AND DISCUSSION**

The CPS biosynthetic locus for strain A-85/14 was identified as six genes, designated cps16A to cps16F, transcribed divergently from cpsD (Fig. 1). BLASTx revealed that, although similar sequences could be found in other bacterial species (see Table 1 and below), including other members of the *Pasteurellaceae* family, these six genes are not...
present in the CPS biosynthetic loci of any other serovar of *A. pleuropneumoniae*.

Furthermore, the arrangement of genes is not similar to that seen in any other bacterial species sequenced thus far.

The first gene, *cps16A*, encodes a 261-amino-acid (aa) predicted glycosyltransferase that shares greatest identity with the C-terminal half (82% identity for aa 198 to 458 of 465) of DcbB in *Actinobacillus ureae* (GenBank accession no. WP_005621190). Similar glycosyltransferase proteins are also found in *Pasteurella multocida* (accession no. WP_059246061), *Avibacterium paragallinarum* (GenBank accession no. ACY25522), and *Gallibacterium anatis* (accession no. WP_021461679), though their exact function is not known.

The next gene, *cps16B*, encodes a 565-aa protein with similarity to HyaE (34% identity and 51% positive substitutions over the entire length; GenBank accession no. WP_010906846) involved in synthesis of the hyaluronic acid capsule of *P. multocida* (13). Again, similar proteins are found in *A. paragallinarum* (accession no. WP_046097317), and *G. anatis* (accession no. WP_065232000); with smaller proteins (379 and 415 aa) found in strains of *A. ureae* (GenBank accession no. WP_044024188 and EFX92827).

The product of *cps16C* is a 678-aa glycosyltransferase that shows extensive similarity to a predicted glycosyltransferase in *Acinetobacter seohaensis* (GenBank accession no. SDC36622) and RfbG (dTDP-rhamnosyl transferase) from *Escherichia coli* (GenBank accession no. WP_061091032). There are two predicted rhamnosyl transferase genes (*rfbF* and *rfbN*) present in lipopolysaccharide (LPS) loci of *A. pleuropneumoniae* serovars 1, 9, and 11 (10), and L-rhamnose was shown to be a component of the LPS O chain in these serovars (4). Structural studies also showed the presence of L-rhamnose in the O chains of serovars 2, 4, 7, and 13 (4, 14), and genes encoding dTDP-4-dehydrorhamnose reductase and dTDP-4-dehydrorhamnose 3,5-epimerase have been identified in these

### TABLE 1 Homologs of proteins encoded by the predicted CPS biosynthesis locus of *A. pleuropneumoniae* serovar 16

| ApS16 protein* | Length (no. of aa) | Homologous protein | Length (no. of aa) | Bacterium | % ID* | Accession no. | Known or putative function |
|---------------|--------------------|--------------------|--------------------|-----------|------|---------------|---------------------------|
| Cps16A        | 261                | DcbB               | 465                | *A. ureae* | 82 (91) | WP_005621190 | Predicted glycosyltransferase |
|               |                    | DcbB               | 477                | *P. multocida* | 58 (76) | WP_059246061 | Predicted glycosyltransferase |
|               |                    | CcbB               | 309                | *A. paragallinarum* | 58 (73) | ACY25522 | Predicted glycosyltransferase |
|               |                    |                    | 479                | *G. anatis* | 57 (73) | WP_021461679 | Predicted glycosyltransferase |
| Cps16B        | 565                | HyaE               | 662                | *P. multocida* | 34 (51) | WP_010906846 | Synthesis of hyaluronic acid capsule |
|               |                    |                    | 542                | *G. anatis* | 34 (52) | WP_065232000 | Hypothetical protein |
|               |                    |                    | 553                | *A. paragallinarum* | 33 (51) | WP_046097317 | Hypothetical protein |
|               |                    |                    | 415                | *A. ureae* | 53 (71) | EFX92827 | Hypothetical protein |
|               |                    |                    | 379                | *A. ureae* | 50 (69) | WP_044024188 | Hypothetical protein |
| Cps16C        | 678                | RfbG               | 677                | *A. seohaensis* | 51 (69) | SDC36622 | Predicted glycosyltransferase |
|               |                    |                    | 664                | *E. coli* | 49 (68) | WP_061091032 | dTDP-rhamnosyl transferase |
| Cps16D        | 367                | Haemophilus*       | 382                | *A. pleuropneumoniae* | 71 (81) | WP_049357969 | UDP-galactopyranose mutase |
|               |                    |                    | 383                | *A. pleuropneumoniae* | 70 (81) | WP_005613015 | UDP-galactopyranose mutase |
| Cps16E        | 226                | E. coli           | 217                | *E. coli* | 57 (73) | WP_024190280 | Hypothetical protein |
|               |                    |                    | 206                | *H. sputorum* | 31 (47) | WP_007526031 | Hypothetical protein |
| Cps16F        | 389                | A. ureae          | 390                | *A. ureae* | 88 (94) | WP_005621201 | UDP-glucose 6-dehydrogenase |
|               |                    |                    | 390                | *G. anatis* | 67 (14) | WP_065232026 | UDP-glucose 6-dehydrogenase |
|               |                    |                    | 390                | *P. multocida* | 65 (14) | AAK02860 | UDP-glucose 6-dehydrogenase |

*ApS16 protein, protein encoded by the predicted CPS biosynthesis locus of *A. pleuropneumoniae* serovar 16.

Protein names given where available. The matches with highest percent identity are listed first for each protein.

% ID, percent identity over the matching span. Numbers in parentheses indicate the percentage of amino acids which are similar according to their physiochemical properties over the matching span.

*A. seohaensis, Acinetobacter seohaensis.*

Multiple species of *Haemophilus*. 
However, this is the first report of L-rhamnose as a predicted component of capsule in *A. pleuropneumoniae*. Cps16D is a predicted UDP-galactopyranose mutase that is common to many members of the *Pasteurellaceae*, including *A. pleuropneumoniae* serovars 5, 10, and 12 (GenBank accession no. WP_005613015), where it is encoded by the *glf1* gene found in the respective LPS loci (10). A different galactopyranose mutase gene, *glf*, is found in the LPS loci of *A. pleuropneumoniae* serovars 3, 6, and 8 (10, 15). Although similar in size, the proteins encoded by *glf* and *glf1* share only 43% identity. The enzyme catalyzes the conversion of UDP-galactopyranose to UDP-galactofuranose, which in turn is the precursor of galactofuranose (Gal\(_\text{f}\)), a five-membered ring form of galactose found in cell surface glycans in a variety of organisms except mammals (16). Structural studies confirmed the presence of Gal\(_\text{f}\) in the LPS O chains of *A. pleuropneumoniae* serovars 3, 6, 8, 10, 12, 14, and 15 (4, 17, 18), but not in any of the reported CPS structures.

The *cps16E* gene encodes a 226-aa hypothetical protein containing a domain indicative of coenzyme A (CoA)-dependent O-acetylation of a sugar substrate. The protein sequence is most similar (57% identity) to a hypothetical *E. coli* protein (GenBank accession no. WP_024190280), with the only detected match in the *Pasteurellaceae* being a protein in *Haemophilus sputorum* (accession no. WP_007526031) with 31% identity. Structural studies identified O-acetylated sugars in the CPS of serovars 1, 8, 9, 10, and 11 (4); however, the genes predicted to encode acetyltransferases in the CPS loci of serovars 1 (*cps1D*) and 10 (*cps10B*) (10) are distinct from *cps16E*, and no acetyltransferase genes have been identified in the other CPS loci.

The final gene in the locus, *cps16F*, encodes a UDP-glucose 6-dehydrogenase which is most similar (88% identity) to that found in *A. ureae* (GenBank accession no. WP_005621201), with proteins in *P. multocida* and *G. anatis* (accession no. AAK02860 and WP_065232026, respectively) sharing less than 70% identity. This enzyme catalyzes the conversion of UDP-glucose to UDP-glucuronic acid, the substrate for generation of various surface structures, including CPS, LPS, and exopolysaccharides in a variety of bacterial species (19).

For diagnostic purposes, we designed primers to amplify a unique 212-bp fragment spanning the 3′ end of *cps16C* and the 5′ end of *cps16D* (Fig. 1). When used in conjunction with previously designed *apxIV* primers for detection of all serovars of *A. pleuropneumoniae* (8), the serovar 16 band was successfully amplified (along with *apxIV*) from the original five isolates (12) as well as two new Hungarian isolates, but the reference strains of *A. pleuropneumoniae* serovars 1 to 15 produced only the *apxIV* amplicon (Fig. 2). Further testing of clinical isolates of *A. pleuropneumoniae* and other bacteria associated with pigs, including publicly available genomes, confirmed the specificity of the primers for serovar 16. Although currently identified only in Hungary, the serovar 16-specific primers designed here will be important for wider epidemiological monitoring of the distribution of this newly identified serovar. The size of the
amplicon generated should allow these primers to be incorporated into existing multiplex PCRs (8, 20). We did not evaluate the use of the serovar 16 primers in quantitative PCR (qPCR).

In conclusion, we have identified the predicted capsule locus of *A. pleuropneumoniae* serovar 16 and confirmed that it is unlike that in any other serovar. Sharing no common genes with any of the type I to III CPS loci previously identified in *A. pleuropneumoniae*, it represents a new type IV locus. The combination of DNA sequence data (this study) and serology (12) show the existence of a sixteenth serovar—designated serovar 16—of *A. pleuropneumoniae*. Structural analysis of the serovar 16 capsule is under way.

**MATERIALS AND METHODS**

*Actinobacillus pleuropneumoniae* strains used in this study were as follows: the recently proposed serovar 16 reference strain A-85/14 plus the four other strains described by Sárközi et al. (12) and two further 2016 Hungarian isolates (151/16 and 228/16) identified as serovar 16 by serology; serovar 1 to 15 reference strains, i.e., 4074T, 1536, S1421, M62, L20, Femø, W83, 405, CVJ13261, D13039, 56153, 8328, N-273, 3906, and HS143, respectively (see also reference 21 for further details); and a collection of 68 clinical isolates (5 isolates each of serovars 1 to 8, 9/11, 10, 12, 14, and 15 and 3 isolates of serovar 13). In addition, we tested 31 strains of other bacterial species associated with pigs that have been described as clinical isolates (5 isolates each of serovars 1 to 8, 9/11, 10, 12, 14, and 15 and 3 isolates of serovar 13).

Serovar 16-specific primers, AP16F (F stands for forward) (TTACTCACTTGGGCTAGGGATAG) and AP16R (R stands for reverse) (ACCGCAATATGATTACGCCC) were designed to amplify a 212-bp fragment as shown in Fig. 1. These primers were initially tested for specificity using genomic DNA from the 16 serovar strains (including strain A-85/14). Subsequently, the primers were combined with apxIVA1 (TTATCCGAACTTTGGTTTAGCC) and apxIVA3 (CATATTTGATAAAACCATCCGTC) that amplify a 418-bp fragment of the species-specific *apxIV* gene of *A. pleuropneumoniae* (8) and further evaluated for specificity using the other 101 isolates, including the negative controls. Amplification was performed using the Qiagen Fast Cycling PCR kit, according to the manufacturer’s instructions (Qiagen). Each reaction mixture contained 10 μl Qiagen Fast Cycling PCR master mix, 2 μl CoralLoad dye, 1 μl genomic DNA, each primer at a final concentration of 0.5 μM, and water to a final volume of 20 μl. Cycling conditions included an initial 5-min activation at 95°C, followed by 30 cycles of PCR, with 1 cycle consisting of denaturation at 96°C for 5 s, annealing at 60°C for 5 s, and extension at 68°C for 10 s, followed by a final extension at 72°C for 1 min.

**Accession number(s).** The sequence of the complete capsule locus for serovar 16 (strain A-85/14) has been deposited in GenBank accession number KX007602.

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**REFERENCES**

1. Bossé JT, Janson H, Sheehan BJ, Beddek AJ, Rycroft AN, Kroll JS, Langford PR. 2002. *Actinobacillus pleuropneumoniae*: pathobiology and pathogenesis of infection. Microbes Infect 4:225–235. https://doi.org/10.1016/S1286-4579(01)01534-9.

2. Gottschalk M. 2012. Actinobacillosis, p 653–669. In Karriker L, Ramirez A, Schwartz K, Stevenson G, Zimmerman J (ed), Diseases of swine, 10th ed. Wiley, Hoboken, NJ.

3. Gottschalk M. 2015. The challenge of detecting herds sub-clinically infected with *Actinobacillus pleuropneumoniae*. Vet J 206:30–38. https://doi.org/10.1016/j.tvjl.2015.06.016.
4. Perry MB, Altman E, Brisson J-R, Beynon LM, Richards JC. 1990. Structural characteristics of the antigenic capsular polysaccharides and lipopolysaccharides involved in the serological classification of Actinobacillus (Haemophilus) pleuropneumoniae strains. Serodiag Immunother Infect Dis 4:299–308. https://doi.org/10.1016/0888-0786(90)90018-J.

5. Blackall PJ, Klaasen HLBIM, van den Bosch H, Kuhnert P, Frey J. 2002. Proposal of a new serovar of Actinobacillus pleuropneumoniae: serovar 15. Vet Microbiol 84:47–52. https://doi.org/10.1016/S0378-1135(01)00428-X.

6. Ito H. 2015. The genetic organization of the capsular polysaccharide biosynthesis region of Actinobacillus pleuropneumoniae serotype 14. J Vet Med Sci 77:583–586. https://doi.org/10.1292/jvms.14-0174.

7. Ito H, Sueyoshi M. 2015. The genetic organization of the capsular polysaccharide biosynthesis region of Actinobacillus pleuropneumoniae serotype 15. J Vet Med Sci 77:483–486. https://doi.org/10.1292/jvms.14-0203.

8. Bossé JT, Li Y, Angen Ø, Weinert LA, Chaudhuri RR, Holden MT, Williamson SM, Maskell DJ, Tucker AW, Wren BW, Rycoft AN, Langford PR, BRaDPT1 consortium. 2014. Multiplex PCR for unequivocal differentiation of Actinobacillus pleuropneumoniae serovars 1 to 3, 5 to 8, 10, and 12. J Clin Microbiol 52:2380–2385. https://doi.org/10.1128/JCM.00665-14.

9. Foote SJ, Bossé JT, Bouvietch AB, Langford PR, Young NM, Nash JHE. 2008. The complete genome sequence of Actinobacillus pleuropneumoniae L20 (serotype 5b). J Bacteriol 190:1495–1496. https://doi.org/10.1128/JB.01845-07.

10. Xu Z, Chen X, Li L, Li T, Wang S, Chen H, Zhou R. 2010. Comparative genomic characterization of Actinobacillus pleuropneumoniae. J Bacteriol 192:5625–5636. https://doi.org/10.1128/JB.00535-10.

11. Xu Z, Zhou Y, Li L, Zhou R, Xiao S, Wan Y, Zhang S, Wang K, Li W, Li L, Jin H, Kang M, Dalai B, Li T, Liu L, Cheng Y, Zhang L, Xu T, Zheng H, Pu S, Wang B, Gu W, Zhang X-L, Zhu G-F, Wang S, Zhao G-P, Chen H. 2008. Genome biology of Actinobacillus pleuropneumoniae JLO3, an isolate of serotype 3 prevalent in China. PLoS One 3:e1450. https://doi.org/10.1371/journal.pone.0001450.

12. Sárközi R, Makrai L, Fodor L. 2015. Identification of a proposed new serovar of Actinobacillus pleuropneumoniae: serovar 16. Acta Vet Hung 63:444–450. https://doi.org/10.1556/175.2015.041.

13. Chung JY, Zhang Y, Adler B. 1998. The capsule biosynthetic locus of Pasteurella multocida A1. FEMS Microbiol Lett 166:289–296. https://doi.org/10.1111/j.1574-6968.1998.tb13903.x.

14. MacLean LL, Perry MB, Vinogradov E. 2004. Characterization of the antigenic lipopolysaccharide O chain and the capsular polysaccharide produced by Actinobacillus pleuropneumoniae serotype 13. Infect Immun 72:5925–5930. https://doi.org/10.1128/IAI.72.10.5925-5930.2004.

15. Bossé JT, Chaudhuri RR, Li Y, Leans LE, Fernandez Crespo R, Coupland P, Holden MTG, Bazzoli DM, Maskell DJ, Tucker AW, Wren BW, Rycoft AN, Langford PR, BRaDPT1 consortium. 2016. Complete genome sequence of MIDG2331, a genetically tractable serovar 8 clinical isolate of Actinobacillus pleuropneumoniae. Genome Announc 4:e01667-15. https://doi.org/10.1128/genomeA.01667-15.

16. Heiss C, Skowyra ML, Liu H, Klufts JS, Wang Z, Williams M, Srikanta D, Beverley SM, Azadi P, Doering TL. 2013. Unusual galactofuranose modification of a capsule polysaccharide in the pathogenic yeast Cryptococcus neoformans. J Biol Chem 288:10994–11003. https://doi.org/10.1074/jbc.M112.411998.

17. Perry MB, MacLean LL. 2004. Structural characterization of the antigenic O-polysaccharide in the lipopolysaccharide produced by Actinobacillus pleuropneumoniae serotype 14. Carbohydr Res 339:1399–1402. https://doi.org/10.1016/j.carres.2004.03.011.

18. Perry MB, MacLean LL, Vinogradov E. 2005. Structural characterization of the antigenic capsular polysaccharide and lipopolysaccharide O-chain produced by Actinobacillus pleuropneumoniae serotype 15. Biochim Cell Biol 83:61–69. https://doi.org/10.1113/bio.04.0112287.

19. Mainprize IL, Bean JD, Bouwman C, Kimber MS, Whitfield C. 2013. The genomic characterization of Actinobacillus pleuropneumoniae serovars 1 to 3, 5 to 8, 10, and 12. J Clin Microbiol 46:800–803. https://doi.org/10.1128/JCM.01787-07.

20. Turni C, Singh R, Schembri MA, Blackall PJ. 2014. Evaluation of a multiplex PCR to identify and serotype Actinobacillus pleuropneumoniae serovars 1, 5, 7, 12 and 15. Lett Appl Microbiol 59:362–369. https://doi.org/10.10111/lam.12287.

21. Zhou L, Jones SCP, Angen Ø, Bossé JT, Nash JHE, Frey J, Zhou R, Chen HC, Kroll JS, Rycoft AN, Langford PR. 2008. Multiplex PCR that can distinguish between immunologically cross-reactive serovar 3, 6, and 8 Actinobacillus pleuropneumoniae strains. J Clin Microbiol 46:800–803. https://doi.org/10.1128/JCM.00685-14.

22. Chalker VJ, Toomey C, Opperman S, Brooks HW, Ibuoye MA, Brownlie J, Rycoft AN. 2008. Respiratory disease in kennelled dogs: serological responses to Bordetella bronchiseptica lipopolysaccharide do not correlate with bacterial isolation or clinical respiratory symptoms. Clin Diagn Lab Immunol 10:352–356. https://doi.org/10.1128/CDLI.10.3.352-356.2003.

23. Baumeister AK, Runge M, Ganter M, Doering TL. 2013. Unusual galactofuranose modification of a capsule polysaccharide in the pathogenic yeast Cryptococcus neoformans. J Biol Chem 288:10994–11003. https://doi.org/10.1074/jbc.M112.411998.

24. Hotchkiss EJ, Hodgson JC, Lainson FA, Zadoks RN. 2011. Multilocus sequence typing of Pasteurella multocida isolates from cattle and other host species demonstrates niche association. BMC Microbiol 11:115. https://doi.org/10.1186/1471-2180-11-115.

25. Howell KJ, Peters SE, Wang J, Hernandez-Garcia J, Weinert LA, Luan S-L, Chaudhuri RR, Angen Ø, Aragon V, Williamson SM, Parkhill J, Langford PR, Rycoft AN, Wren BW, Maskell DJ, Tucker AW. 2015. Development of a multiplex PCR assay for rapid molecular serotyping of Haemophilus parasuis. J Clin Microbiol 53:3812–3821. https://doi.org/10.1128/JCM.01991-15.