Brief Report

Reduced Loading of Intracellular Ca\(^{2+}\) Stores and Downregulation of Capacitative Ca\(^{2+}\) Influx In Bcl-2-Overexpressing Cells

Paolo Pinton, ‡ Davide Ferrari, * Paulo Magalhães, † Klaus Schulze-Osthoff, § Francesco D i Virgilio, * Tullio Pozzan, ‡ and Rosario Rizzuto *

*Department of Experimental and Diagnostic Medicine, Section of General Pathology, 44100 Ferrara, Italy; † Department of Biomedical Sciences and Consiglio Nazionale delle Ricerche (CNR) Center for the Study of Biomembranes, University of Padova, 35121 Padova, Italy; and § Eberhard-Karls University, 72076 Tübingen, Germany

Abstract. The mechanism of action of the oncogene bcl-2, a key regulator of the apoptotic process, is still debated. We have employed organelle-targeted chimeras of the Ca\(^{2+}\)-sensitive photoprotein, aequorin, to investigate in detail the effect of Bcl-2 overexpression on intracellular Ca\(^{2+}\) homeostasis. In the ER and the Golgi apparatus, Bcl-2 overexpression increases the Ca\(^{2+}\) leak (while leaving Ca\(^{2+}\) accumulation unaffected), hence reducing the steady-state [Ca\(^{2+}\)] levels. As a direct consequence, the [Ca\(^{2+}\)] increases caused by inositol 1,4,5 trisphosphate (IP3)-generating agonists were reduced in amplitude in both the cytosol and the mitochondria. Bcl-2 overexpression also reduced the rate of Ca\(^{2+}\) influx activated by Ca\(^{2+}\) store depletion, possibly by an adaptive downregulation of this pathway. By interfering with Ca\(^{2+}\)-dependent events at multiple intracellular sites, these effects of Bcl-2 on intracellular Ca\(^{2+}\) homeostasis may contribute to the protective role of this oncogene against programmed cell death.

Key words: calcium • organelles • oncogene • aequorin • cell signaling

Introduction

In the past years, Bcl-2, the mammalian homologue of CED-9, the antiapoptotic gene initially described in Caenorhabditis elegans, has emerged as a major checkpoint controlling the escape of cells from their death fate (Kroemer, 1997). In mammals, Bcl-2 is a member of a broad family of related gene products, with various degrees of sequence identity and different functions. While Bcl-2 and some members of the family (e.g., Bcl-x) protect the cells from apoptosis, others (e.g., Bax) have the opposite effect. However, while there is a general consensus on the role of Bcl-2, its mechanism of action remains elusive. On one hand, the various members of the family have been shown to dimerize and to interact with cofactors of caspases, key executors of the apoptotic machinery (Li and Yuan, 1999); on the other, the specific localization of the protein on the ER and outer mitochondrial membrane (Lithgow et al., 1994) suggests that these proteins may interfere with some specific functions localized in these organelles. In particular, Bcl-2 has been shown to prevent the release of cytochrome c and AIF from mitochondria (Kluck et al., 1997; Yang et al., 1997), events that are considered essential in triggering apoptosis, whereas expression of Bax has opposite effects (Jurgensmeier et al., 1998).

Among the various functions occurring on ER and mitochondrial membranes, ion fluxes are recognized to play major roles. Thus, the intriguing possibility has emerged that members of the Bcl-2 family affect the apoptotic process by modulating ion fluxes in these organelles (Minn et al., 1997; Schendel et al., 1998). In particular, recent work in cell clones stably expressing the oncogene suggests that Bcl-2 causes an overload of Ca\(^{2+}\) within the ER (Lam et al., 1994; He et al., 1997, Kuo et al., 1998). However, this effect appears in contrast with the observation that Bcl-2 insertion in artificial lipid bilayers results in the formation of ionic channels of low selectivity (Minn et al., 1997). Such property should in fact cause a reduction in the ER Ca\(^{2+}\) content (due to an increased leak) rather than excess accumulation. In this report, we have undertaken a detailed investigation of Ca\(^{2+}\) homeostasis at the
subcellular level using the recombinant aequorin (A E Q)\(^1\) approach developed in our laboratory (Rizzuto et al., 1992; De Giorgi et al., 1996). We conclude that Bcl-2 over-expression affects Ca\(^{2+}\) handling by reducing the state of filling of the intracellular Ca\(^{2+}\) stores. The implications of these results for the protective action of Bcl-2 on apoptosis are discussed.

**Materials and Methods**

**Cell Culture and Transfection**

Hela cells were grown in DMEM supplemented with 10% FCS in 75-cm\(^2\) Falcon flasks. Before transfection, cells were seeded onto 13-mm glass coverslips and allowed to grow to 50% confluence. At this stage, transfection with 4 \(\mu\)g of DNA (3 \(\mu\)g Bcl-2 + 1 \(\mu\)g A E Q) was carried out as previously described (Rizzuto et al., 1995a) and A E Q measurements or immunocytochemistry were performed 36 h after transfection. For fura-2 measurements, the cells were seeded onto 24-mm coverslips and transfection was carried out with 8 \(\mu\)g plasmid DNA (6 \(\mu\)g Bcl-2 + 2 \(\mu\)g mtGFP).

**Aequorin Measurements**

For cytosolic A E Q (cytAEQ) and mitochondrial A E Q (mtAEQ), the coverslip with the cells was incubated with 5 \(\mu\)M coelenterazine for 1-2 h in DMEM supplemented with 1% FCS, and then transferred to the perfusion chamber. For reconstituting with high efficiency, the A E Q chimeras targeted to the Golgi apparatus and the ER (GoAEQ and erAEQ, respectively), the luminal [Ca\(^{2+}\)] of these compartments first must be reduced. This was obtained by incubating the cells for 1 h at 4°C in KRB (Kreb-Ringer modified buffer: 125 mM NaCl, 5 mM KCl, 1 mM Na\(_2\)PO\(_4\), 1 mM MgSO\(_4\), 5.5 mM glucose, and 20 mM Hepes, pH 7.4, at 37°C) supplemented with coelenterazine 5 \(\mu\)M, the Ca\(^{2+}\) ionophore ionomycin, and 600 \(\mu\)M EGTA. After this incubation, the cells were extensively washed with KRB supplemented with 2% BSA and 1 mM EGTA, and then A E Q reconstitution was carried out as described.

All A E Q measurements were carried out in KRB. A gonits and other drugs were added to the same medium. In experiments with permeabilized cells, a buffer mimicking the cytosolic ionic composition (intracellular buffer, IB: 140 mM KCl, 10 mM NaCl, 1 mM KHPO\(_4\), 5.5 mM glucose, 2 mM MgSO\(_4\), 1 mM ATP, 2 mM sodium succinate, 20 mM Hepes, pH 7.05, at 37°C) was employed. This solution was supplemented with either 100 \(\mu\)M EGTA or a fixed [Ca\(^{2+}\)] (buffered with 2 mM EGTA). The experiments were terminated by lysing the cells with 100 \(\mu\)M digitonin in a hypotonic Ca\(^{2+}\)-rich solution (10 mM CaCl\(_2\), in H\(_2\)O), thus discharging the remaining A E Q pool. The light signal was collected and calibrated into [Ca\(^{2+}\)] values as previously described (Brini et al., 1995; Rizzuto et al., 1995a). In brief, a 13-mm round coverslip with the transfected cells was placed in a perfused, thermostatted chamber located in the close proximity of a low-noise photomultiplier, with built-in amplifier discriminator. The output of the discriminator was captured by a Thorn-EMI photon counting board and stored in an IBM-compatible computer for further analyses. The A E Q luminescence data were calibrated off-line into [Ca\(^{2+}\)] values, using a computer algorithm based on the Ca\(^{2+}\) response curve of wild-type and mutant A E Qs, as previously described (Brini et al., 1995; Barrero et al., 1997).

In some experiments using erAEQ, after reaching the plateau level, an artificial slow decrease in [Ca\(^{2+}\)] was observed. Such an artifact is due to a small fraction of missorted A E Q in a low [Ca\(^{2+}\)] compartment and a simple practical cure for it has been described by Macher et al. (1999).

In the experiment of Fig. 4, cells transfected with erAEQ were used to determine the steady [Ca\(^{2+}\)]\(_i\) obtained by incubating the cells in low Ca\(^{2+}\) medium. Two protocols were adopted that gave identical results. In the first, cells incubated in normal medium were depleted according to the standard protocol, but the Ca\(^{2+}\) concentration during the refilling was reduced to 100 \(\mu\)M. In the second protocol, the cells were preincubated in 100 \(\mu\)M CaCl\(_2\) for 16 h, depleted, and finally refilled with an extracellular Ca\(^{2+}\) concentration of 100 \(\mu\)M.

**Fura-2 Measurements**

The coverslip with the cells was incubated with 5 \(\mu\)M fura-2 (added to DMEM + 1% FCS) at 37°C for 30 min, and after a brief washout was then transferred to the thermostated stage of a Zeiss Axiovert inverted microscope equipped with a Sutter filterwheel and 340/380 excitation filters. The fluorescence data were collected with a Princeton Instruments back-illuminated camera and expressed as emission ratios, using Metafluor software (Universal Imaging). Within the microscope field, transfected and untransfected cells were identified before carrying out the fura-2 monitoring by revealing mitochondrial-targeted green fluorescent protein (mtGFP) fluorescence (excitation 480 nm, emission 510 nm).

**Results and Discussion**

To directly investigate the role of Bcl-2 in Ca\(^{2+}\) homeostasis, we have selectively measured the [Ca\(^{2+}\)] in different cell compartments, i.e., the cytoplasm and the organelles as sources (ER and Golgi apparatus) or targets (mitochondria) of the Ca\(^{2+}\) signal. Targeted A E Q chimeras developed in our lab (Rizzuto et al., 1992; Brini et al., 1995; Montoro et al., 1995; Pinton et al., 1998) were employed. This approach has two major advantages. The first is the high subcellular specificity of the probes, which allows to clearly identify the effects of Bcl-2 expression on the signaling properties of the various compartments; the second is that the Ca\(^{2+}\) probe can be cotransfected with Bcl-2. We and others have previously shown that in transient cotransfection, the two recombinant proteins are expressed by the same subset of cells, thus avoiding the risks associated with the use of stable clones or single cells (Brini et al., 1995).

The effect of Bcl-2 expression on the cytosolic Ca\(^{2+}\) signal elicited by an agonist, A TP, acting on receptors coupled, through G\(_q\) proteins, to the production of inositol 1,4,5 trisphosphate (IP3) was first investigated. In the experiment shown in Fig. 1, Hela cells, either coexpressing Bcl-2 and cytAEQ (Bcl-2–transfected) or expressing only cytAEQ (controls) were challenged with ATP, which acts on a G\(_q\)-coupled P2Y receptor. Both in control and Bcl-2–transfected cells, A TP stimulation causes a rapid rise in cytoplasmic Ca\(^{2+}\) concentration ([Ca\(^{2+}\)]\(_i\)), followed by a gradually declining sustained plateau. In Bcl-2–transfected cells, the [Ca\(^{2+}\)] increases evoked by stimulation with A TP are significantly smaller than in controls (peak amplitude 1.53 ± 0.22 vs. 2.06 ± 0.13 \(\mu\)M, n = 10; Fig. 1A).

Since Bcl-2 is localized in the mitochondria also, and thus could, at least in principle, enhance Ca\(^{2+}\) uptake in this organelle, a simple explanation for this result would be that cytosolic Ca\(^{2+}\) is more rapidly cleared by mitochondria, acting as an immobile buffer, placed in closed proximity to IP3-gated channels. Fig. 1 B, however, shows that this is not the case. Indeed, Bcl-2 overexpression does not increase, but rather reduces, the mitochondrial [Ca\(^{2+}\)] rise induced by A TP (peak amplitude 6.56 ± 0.72 in controls vs. 4.42 ± 0.46 \(\mu\)M in Bcl-2–expressing cells; n = 10). Interestingly, the reduction caused by Bcl-2 overexpression appears larger in the mitochondria than in the cytosol, both in absolute terms and as percent (26 vs. 33% reduction, respectively). While not responsible for the alteration

Abbreviations used in this paper: A E Q, aequorin; cytAEQ, cytosolic A E Q; erAEQ, A E Q chimera targeted to the ER; G F P, green fluorescent protein; GoAEQ, A E Q chimera targeted to the Golgi apparatus; IB, intracellular buffer; IP3, inositol 1,4,5 trisphosphate; K RB, K reb-Ringer modified buffer; mtAEQ, mitochondrial A E Q; mtGFP, mitochondrial targeted GFP; S E R C A, sarcoendoplasmic reticulum Ca\(^{2+}\) ATPases; T B u B H Q, 2,5-di-(tet-butyl)-1,4-benzohydroquinone.

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regulated by the agonist-sensitive Ca$^{2+}$ permeability transition pore, PTP (Bernardi, 1999), are possibly the opening of the mitochondrial matrix, such as the stimulation of ATP production (Jouaville et al., 1999) and/or the opening of the mitochondrial Ca$^{2+}$ release channel (LCM) (Montero et al., 1995). In addition, since we recently have demonstrated that the Golgi apparatus shares many of the Ca$^{2+}$ homeostatic properties of the ER (Pinton et al., 1998), this latter organelle was also investigated. Fig. 2 shows the calibrated [Ca$^{2+}$]$_{i}$ values in the two compartments. To obtain reliable quantitative estimates of the [Ca$^{2+}$]$_{i}$ in the lumen of these two organelles, their [Ca$^{2+}$]$_{i}$ needs to be decreased during both the reconstitution of A E Q with coelenterazine and the subsequent initial phase of perfusion with K R B /E G T A in the luminometer (see Materials and Methods). Under those conditions, the [Ca$^{2+}$]$_{i}$ was <10 µM in both organelles. When the [Ca$^{2+}$]$_{i}$ in the perfusion medium was switched to 1 mM, the [Ca$^{2+}$]$_{i}$ in the lumen of the two compartments gradually increased, reaching in control cells plateau values of 452 ± 46 µM (n = 7) in the ER and 262 ± 47 µM (n = 7) in the Golgi apparatus, respectively. In Bcl-2 transfected cells, lower steady state values were attained in both compartments (304 ± 38 µM , n = 7 for the ER, and 186 ± 34 µM , n = 7 for the Golgi apparatus, respectively). When the cells were treated with A TP, rapid decreases in the [Ca$^{2+}$]$_{i}$ of the two compartments were observed, both in control and Bcl-2-expressing cells. In both organelles, the decrease of [Ca$^{2+}$]$_{i}$ was larger and faster in controls compared with Bcl-2-expressing cells, reflecting the higher filling state and thus the more rapid flow through the IP3-gated channels.

The data reported above differ substantially from those reported in WEHI 7.2 clones, where Bcl-2 overexpression results in an augmentation of the ER Ca$^{2+}$ content (Lam et al., 1994; He et al., 1997). This discrepancy may be due to the use of transient vs. stable transfections, in which the results might reflect unique properties of the selected clone and/or the long-term expression of a foreign gene may result in adaptation phenomena that are only indirect consequences of the expressed protein. This interpretation might also explain the results obtained by Kuo and coworkers in clones of MCF10A cells overexpressing...
Bcl-2, in which an increase in SERCA2 (sarcoplasmic reticulum Ca2+ ATPases 2) protein leads to accelerated Ca2+ uptake and enhanced Ca2+ loading (Kuo et al., 1998). On the other hand, it is possible that, although the duration of these experiments is compatible with the time course of the apoptotic process, the events observed in transient experiments could include only some of the relevant effects of Bcl-2 on the mechanisms controlling programmed cell death.

But how does Bcl-2 affect ER and Golgi Ca2+ handling? Given that the steady-state [Ca2+] in both organelles depends on the equilibrium between active accumulation and passive leak, the effect of Bcl-2 could be on either process. On one hand, Bcl-2 could reduce Ca2+ uptake by the SERCA, either by a direct effect on the pump or by reducing the resting cytosolic [Ca2+]. On the other, it could increase the passive diffusion of Ca2+ from the ER. To clarify this critical issue, we investigated these possibilities independently.

At first, the possibility that Bcl-2 affected primarily cytosolic [Ca2+] (and hence [Ca2+]ER only indirectly) was verified. Given that AEQ is not accurate enough to reveal small differences of [Ca2+], the resting [Ca2+] was measured with the fluorescent indicator fura-2 (Gryniewicz et al., 1985). In this experiment, to identify Bcl-2-overexpressing cells in single-cell imaging experiments, the cells were cotransfected with mitochondrially targeted GFP, mtGFP(S65T) (Rizzuto et al., 1995b). GFP-positive cells were distinguished from controls by the typical fluorescence emitted upon illumination with blue light. No difference in the resting cytoplasmic [Ca2+] between Bcl-2-transfected and control cells was observed (data not shown).

Then, we investigated whether Bcl-2 overexpression had any direct effect on the activity of the SERCA pump. For this purpose, the kinetics of Ca2+ accumulation were studied in permeabilized cells (Fig. 3 A). A fura reconstituting the photoprotein as in the experiment of Fig. 2, Bcl-2-transfected and control cells were transferred to the luminometer chamber and perfused with a buffer (IB, see Materials and Methods) mimicking intracellular ionic conditions, supplemented with 100 μM EGTA (IB/EGTA). A fura permeabilization with 100 μM digitonin added to the same medium and wash in IB/EGTA, the medium was switched to IB, containing 200 nM [Ca2+] (IB/Ca2+). Under those conditions, [Ca2+] EGTA mutated AEQ (control). 36 h after transfection, depletion of Ca2+ stores and AEQ were carried out as in Fig. 2. The coverslip with the cells was then transferred to the luminometer chamber. The perfusion medium was changed to IB/EGTA, and permeabilization was carried out by perfusing 100 μM digitonin (added to the same medium) for 1 min. After a 2-min wash with IB/EGTA, Ca2+ accumulation in the ER was initiated by switching the medium to IB containing a buffered Ca2+ concentration of 0.2 μM (see text for details). AEQ calibration was carried out as described in Materials and Methods. B, Dependence on [Ca2+] of the Ca2+ leak rate from the ER. Transfection, depletion of Ca2+ stores, and AEQ reconstitution were carried out as in A, then the coverslip with the cells was transferred to the luminometer and perfused with KRB/Ca2+ until the steady-state [Ca2+] was reached. Ca2+ release was initiated by treating the cells with 50 μM tBuBHQ. Based on the experimental trace, the maximal rates of Ca2+ release (measured from the first derivative) at different values of [Ca2+] were calculated and plotted for Bcl-2-transfected and control cells. The plot contains data obtained from ten independent experiments. In each experiment, duplicate samples of control and Bcl-2-overexpressing cells were measured. Due to the mixing time in the luminometer chamber, the kinetics of [Ca2+] decrease are sigmoidal and the maximal rate is obtained 2-3 s after addition of tBuBHQ. Accordingly, we considered the maximal rates the best approximation for the initial rate of [Ca2+] decrease. The fitting of the curve shown in B was performed using Microsoft Excel software.

Figure 3. Kinetics of ER Ca2+ uptake and release in control and Bcl-2-transfected cells. A, ER Ca2+ refilling in permeabilized cells. Parallel batches of HeLa cells were either cotransfected with eAEQ mut and Bcl-2 (Bcl-2), or transfected with eAEQ mut alone (control). 36 h after transfection, depletion of Ca2+ stores and eAEQ were carried out as in Fig. 2. The coverslip with the cells was then transferred to the luminometer chamber. The perfusion medium was changed to IB/EGTA, and permeabilization was carried out by perfusing 100 μM digitonin (added to the same medium) for 1 min. After a 2-min wash with IB/EGTA, Ca2+ accumulation in the ER was initiated by switching the medium to IB containing a buffered Ca2+ concentration of 0.2 μM (see text for details). AEQ calibration was carried out as described in Materials and Methods. B, Dependence on [Ca2+] of the Ca2+ leak rate from the ER. Transfection, depletion of Ca2+ stores, and AEQ reconstitution were carried out as in A, then the coverslip with the cells was transferred to the luminometer and perfused with KRB/Ca2+ until the steady-state [Ca2+] was reached. Ca2+ release was initiated by treating the cells with 50 μM tBuBHQ. Based on the experimental trace, the maximal rates of Ca2+ release (measured from the first derivative) at different values of [Ca2+] were calculated and plotted for Bcl-2-transfected and control cells. The plot contains data obtained from ten independent experiments. In each experiment, duplicate samples of control and Bcl-2-overexpressing cells were measured. Due to the mixing time in the luminometer chamber, the kinetics of [Ca2+] decrease are sigmoidal and the maximal rate is obtained 2-3 s after addition of tBuBHQ. Accordingly, we considered the maximal rates the best approximation for the initial rate of [Ca2+] decrease. The fitting of the curve shown in B was performed using Microsoft Excel software.
then added that initiates the release of stored Ca$^{2+}$. Given that, by definition, the rates of Ca$^{2+}$ uptake and leak in steady state are equal, the rate of [Ca$^{2+}$]$_{se}$ decrease upon blockade of the SERCA must reflect the rate of Ca$^{2+}$ cycling across the ER membrane and, thus, of the leak rate at any given steady-state [Ca$^{2+}$]$_{se}$. Fig. 3 B shows the relationship between [Ca$^{2+}$]$_{se}$ and leak rate, which can be fitted by a power equation. In Bcl-2-overexpressing cells, the data are more scattered, but for each [Ca$^{2+}$]$_{se}$ value, the rate of Ca$^{2+}$ efflux is faster than in control cells. This result is consistent with the evidence that Bcl-2 can induce cation channels of low selectivity in artificial lipid bilayers (Minn et al., 1997).

Previously, we have demonstrated that a reduction of $\sim$30% in steady-state [Ca$^{2+}$]$_{se}$ results in a substantial activation (>50%) of the so-called capacitative Ca$^{2+}$ influx (Hofer et al., 1998). The reduction in steady-state [Ca$^{2+}$]$_{se}$ caused by Bcl-2 overexpression is thus expected to cause an activation of this pathway. However, indirect evidence (namely, the observation that the resting cytosolic Ca$^{2+}$ level is indistinguishable from controls) suggests that this is not the case. To further investigate the problem, we measured the capacitative Ca$^{2+}$ influx in Bcl-2-transfected and control HeLa cells expressing cytAEQ (Fig. 4). The cells were treated with the SERCA blocker (10 $\mu$M tBuBHQ) while perfused with Ca$^{2+}$-free medium (KRB/EGTA). This procedure first evoked a small transient increase in [Ca$^{2+}$], due to release of Ca$^{2+}$ from the stores. When the release of stored Ca$^{2+}$ was complete ($\sim$3 min), Ca$^{2+}$ was readded to the medium. This maneuver evoked a second, larger [Ca$^{2+}$] increase via the capacitative pathway, which was markedly lower in Bcl-2-transfected than in controls cells (1.19 $\pm$ 0.11 in controls vs. 0.68 $\pm$ 0.13 $\mu$M in Bcl-2-expressing cells, n = 6). The question then arises as to the mechanism(s) through which Bcl-2 downregulates capacitative Ca$^{2+}$ influx. Two possibilities were considered: one was that the reduction of this Ca$^{2+}$ influx pathway is an adaptive mechanism to the long lasting reduction in steady-state [Ca$^{2+}$]$_{se}$; the second was that the reduction is a direct consequence of Bcl-2 overexpression. To distinguish between these two possibilities, we verified whether a long-term reduction of [Ca$^{2+}$]$_{se}$, obtained independently of Bcl-2, could cause a comparable downregulation of the capacitative Ca$^{2+}$ influx. Capacitative Ca$^{2+}$ influx was thus compared in controls and in cells whose [Ca$^{2+}$]$_{se}$ was reduced to values similar to those of Bcl-2-overexpressing cells by prolonged incubation with a low (100 $\mu$M) extracellular [Ca$^{2+}$]. In cells that had been maintained at low external [Ca$^{2+}$], and thus experienced a long-term reduction in the resting [Ca$^{2+}$]$_{se}$ levels, the [Ca$^{2+}$]$_{se}$ peak evoked by activation of the capacitative Ca$^{2+}$ influx was markedly lower than in controls (0.59 $\pm$ 0.11 M, n = 5). Of interest, the level of steady-state [Ca$^{2+}$]$_{se}$ in cells incubated in low extracellular Ca$^{2+}$ was very similar (269 $\pm$ 46 $\mu$M, n = 4) to that measured in Bcl-2-overexpressing cells incubated in normal (1 mM) extracellular Ca$^{2+}$ concentration. The simplest explanation of these results is that the decrease in capacitative Ca$^{2+}$ influx is an adaptive consequence to the prolonged reduction in steady-state [Ca$^{2+}$]$_{se}$, though, at the moment, it cannot be excluded that other factors (e.g., a reduction in plasma membrane potential) contribute to this effect. This reduction in capacitative Ca$^{2+}$ influx, on one hand, appears critical to maintain a resting [Ca$^{2+}$], indistinguishable from that of control cells, despite a major reduction of [Ca$^{2+}$] in the ER; on the other hand, it could account, at least in part, for a higher resistance of cells expressing Bcl-2 to apoptotic cell death, as the reduction of this ubiquitous Ca$^{2+}$ influx pathway may prevent the overload of Ca$^{2+}$, considered one of the key events in apoptosis. In agreement with this view, Ma and coworkers recently demonstrated that SERCA overexpression, by causing ER Ca$^{2+}$ overload, increases spontaneous apoptosis (Ma et al., 1999). Finally, our data are not in contrast with previous results showing that the SERCA inhibitor, thapsigargin, activates programmed cell death (Bian et al., 1997). In this latter case, in fact, the Ca$^{2+}$ depletion is acute (1–2 min) and complete, whereas in Bcl-2-transfected cells, the drop in [Ca$^{2+}$]$_{se}$ is modest and develops slowly. Thus, thapsigargin not only causes a massive activation of the capacitative Ca$^{2+}$ influx pathway, but also such a drastic reduction in the level of [Ca$^{2+}$]$_{se}$ to interfere with the basic activity of ER chaperonins.

We have demonstrated that overexpression of Bcl-2 causes a reduction of the steady-state [Ca$^{2+}$] levels, both in the ER and in the Golgi apparatus, the two main agonist-sensitive Ca$^{2+}$ stores. We also showed that this alteration affects the Ca$^{2+}$ signal to which the effector systems are exposed, in particular those located in local environments highly dependent on the filling state of the intracellular stores (such as the mitochondria). The chronic reduction in [Ca$^{2+}$] of the ER results in a downregulation of the ca-

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The alterations in Ca\(^{2+}\) handling provoked by Bcl-2 overexpression could be part of the mechanisms through which Bcl-2 exerts its role in the control of cell growth. Targets of Bcl-2 effects on Ca\(^{2+}\) homeostasis could be: the ER itself, where sorting and trafficking of proteins, e.g., enzymes or membrane receptors, are modulated by luminal [Ca\(^{2+}\)]; (Booth and Koch, 1989); the mitochondria, where Ca\(^{2+}\) accumulation regulates the production of ATP (Robb-Gaspers et al., 1998); the concentration of which is a key determinant of the apoptosis vs. necrosis choice (Leist et al., 1997), and, possibly, the release of the apoptotic cofactor cytochrome c (Krajewski et al., 1999); or the cytoplasm, where bulk or local [Ca\(^{2+}\)] increases activate enzymatic effectors, such as the Ca\(^{2+}\)-dependent protease, calpain (Carafoli and Molineari, 1998; Squier et al., 1999), or the phosphatase, calcineurin (Wang et al., 1999). Whether, and how, these routes interact, and how they are affected by the alteration in Ca\(^{2+}\) homeostasis caused by Bcl-2, awaits further investigation.

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