Efficient programmable gene silencing by Cascade

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ABSTRACT

Methods that permit controlled changes in the expression of genes are important tools for biological and medical research, and for biotechnological applications. Conventional methods are directed at individually changing each gene, its regulatory elements or its mRNA’s translation rate. We demonstrate that the CRISPR-associated DNA-binding Cascade complex can be used for efficient, long-lasting and programmable gene silencing. When Cascade is targeted to a promoter sequence the transcription of the downstream gene is inhibited, resulting in dramatically reduced expression. The specificity of Cascade binding is provided by the integral crRNA component, which is easily designed to target virtually any stretch of DNA. Cascade targeted to the ORF sequence of the gene can also silence expression, albeit at lower efficiency. The system can be used to silence plasmid and chromosome targets, simultaneously target several genes and is active in different bacterial species and strains. The findings described here are an addition to the expanding range of CRISPR-based technologies and may be adapted to additional organisms and cell systems.

INTRODUCTION

Rapid modulation of gene expression is an essential feature of bacterial adaptive responses to changing environments. Gene expression can be regulated on both transcriptional and post-transcriptional level, often in a multi-layer fashion. Regulation by activators and repressors that alter transcription initiation is a well-studied phenomenon. The activity of protein factors is often affected by their expression levels by modifications, such as phosphorylation, and by binding of various co-factors (1). Once formed, the mRNA is subject to a variety of post-transcriptional regulatory mechanisms including those that regulate premature transcription termination, translation initiation and mRNA stability. With the discovery of riboregulators and riboswitches it has become clear that RNA structures can regulate gene expression. It is, however, interesting to note that most instances of regulation involving RNA operate at the post-transcriptional level (2) and that there are no known natural examples of RNA-mediated control of promoter activity or transcription initiation in bacteria.

Genetic engineering has allowed exploitation of naturally occurring regulatory mechanisms for both applied use and basic science. Artificial transcription factors have also been created; by fusing DNA-binding protein domains with regulatory domains transcription of specific genes can be controlled (3). Other approaches have involved manipulation of the sequence specificity of a given DNA binding protein, such as transcription-activator-like-effector or zinc finger proteins to direct transcription regulation (4,5). However, engineering of sequence specificity of proteins is a complex and time-consuming process which reduces applicability for simultaneous targeting of multiple genes (6). A programmable, simple and generic method for transcription control is a desirable tool for analysis of any cell and organism. Its effect would be comparable to that of RNAi-based silencing systems in eukaryotes, but it should be noted that these act on the post-transcriptional level.

Promising candidates for easy and efficient gene regulation are the RNA-guided DNA-binding components of CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats) systems, an adaptive immune system in bacteria and archaea that provides protection from invading viruses and mobile elements. A CRISPR-Cas system is characterized by two features: the CRISPR locus and a set of CRISPR-associated (cas) genes. Both components need to be expressed for defense against invasive genetic elements. For an in-depth review, see, e.g. van der Oost et al. (7).

The CRISPR loci contain short repeated sequences separated by short unique spacers usually acquired from phages
or other mobile genetic elements (8–10). The CRISPR is transcribed and the RNA is processed by Cas proteins into CRISPR RNA (crRNA) (11–13); crRNAs use sequence complementarity to guide Cas proteins to target DNA for degradation (14–17), although other functions have also been described (18).

CRISPR-Cas systems are present in about 45% of bacteria and 83% of archaea, and they are currently divided into three major types (I, II and III) and further subtypes (19,20). We focus our work on the Escherichia coli (E. coli) type I-E system, where Cas proteins and crRNA form a complex called Cascade (11). Cascade is a surveillance unit that upon detection of target DNA recruits Cas3 to destroy the target. Targeting requires the presence of a cognate protospacer-adjacent motif (PAM) which is a short sequence (3 bp in the case of Cascade) flanking the protospacer (11,16,21). Complexes similar to Cascade are also found in other type I and type III CRISPR-Cas systems (7,22,23, and references therein).

Type II CRISPR-Cas systems have already been adapted for biotechnological applications, mainly as a tool for genome editing (24,25), but also for gene regulation where a catalytically inactive ‘dead’ Cas9 (dCas9) was used. The dCas9 containing a small guide RNA (sgRNA) was shown to reduce transcription when targeted to a cognate promoter or the open reading frame (ORF) in E. coli and other organisms (26,27). Transcription activation or repression can also be achieved by fusing dCas9 with either a transcriptional activation domain or a repressor (28–30).

Here, we exploit the DNA-binding ability of the Cascade protein-RNA complex to design a programmable system for targeted silencing of gene expression, adding to the emerging CRISPR-Cas toolbox for genetic modifications and gene regulation. We express the components of Cascade in the absence of the Cas3 nuclease, which allows stable target binding without inducing its cleavage. We show that co-expression of a minimal CRISPR array carrying spacers designed to target a promoter result in effective silencing of the downstream gene. Repression could also be achieved by targeting coding regions of a gene, although at lower efficiency. As the system can be made inducible and can be targeted to virtually any region of DNA, it holds the potential as a general silencing tool for any gene of interest in tractable bacteria, archaea and eukaryotes.

**MATERIALS AND METHODS**

**Strains, plasmids and culture media**

All experiments in *E. coli* were performed in Δcas3 genetic background, except BL21 strains that naturally lack cas3 genes. The kanamycin cassette from JW2731(2) Δcas3, a Keio collection strain (31), was removed by Flp recombinase expressed from a temperature-sensitive plasmid, pCP20, as previously described (32). P1 vir lysate grown on BL21AI was used, in a transduction protocol previously described (33), to introduce arabinose-inducible T7 RNA polymerase gene into this strain. This strain was designated MLS367. For routine growth, *E. coli* strains BL21DE3 (34), BL21AI (Invitrogen), MLS367 and *Salmonella typhimurium* LT2 strain DA6192 were cultured in Luria Bertani (LB) medium and grown with aeration at 37°C. When necessary, the media was supplemented with kanamycin (50 μg/ml), ampicillin (100 μg/ml), streptomycin (50 μg/ml), tetracycline (25 μg/ml) and chloramphenicol (15 μg/ml). Chromosomal markers were not selected for during growth in liquid cultures.

The plasmid pEH9 was used as silencing target and carries a Venus gfp gene preceded by a P1 tetO-1 promoter (35). It has a 158-bp polymerase chain reaction (PCR)-amplified DNA fragment containing the 5’ UTR and first 8 codons of the ompA gene inserted into the NsiI and NheI sites of the plasmid pXG10 (36). To compare targeting of a plasmid and targeting of the chromosome, the gfp gene, P1 tetO-1 promoter and resistance marker from pEH9 were amplified by PCR using primer LA085 and LA086 (see Supplementary Table S1) and introduced into the *E. coli* BW25113 chromosome in the Lambda attB site (position 807747–807769 on the BW25113 chromosome) using Lambda red recombination (37). After verification of insertion, the construct was moved into MLS367 by P1 transduction, this strain was designated MLS541.

To test the simultaneous silencing of multiple targets, the mTagBfp2 gene under the control of the constitutive promoter J23101 (Genbank KM018299) was inserted by P1 transduction in the galK locus of MLS367. The chloramphenicol selection marker was removed by Flp recombinase expressed from pCP20. To construct the final strain containing both reporters, the gfp construct (above) was moved to the chromosome of this strain by P1 transduction, this strain was designated MLS561.

Cascade was expressed from pWUR400 (11). For expression of crRNA, minimal CRISPR arrays containing the transcribed 53 bp of the leader sequence from the CRISPR1 array of *E. coli* K12 (38) and a spacer flanked by two repeats were synthesized and subcloned by Life technologies. The CRISPRs were cloned into the EcoRI and XbaI sites of pZE12Luc (35), thereby removing the luciferase gene and placing the CRISPR cassette downstream of the IPTG-inducible promoter. For demonstration of silencing in *S. typhimurium* LT2 strain DA6192 cultures were transformed with pWUR400, pEH9 and CRISPR expressing plasmids. For details of the target sequences, CRISPR arrays and PAMs used, see Supplementary Table S2.

**Measurement of bacterial growth and fluorescence**

For *E. coli* fluorescence measurements on population level, overnight cultures grown in LB were diluted 1:100 in 10 ml of LB medium supplemented with required antibiotics, and 0.2% arabinose and 0.1 mM IPTG to induce CRISPR and Cas protein expression. Cultures were grown in flasks with aeration at 37°C. OD<sub>600</sub> and fluorescence were acquired in exponential phase, 3 h after inoculation, or in stationary phase, 9 h after inoculation. Measurements were made on 100 µl of culture in black Corning flat-bottomed plates with an Infinite M200 Pro microplate reader (Tecan). *S. typhimurium* was analyzed as *E. coli*, except that modified M9-CA medium was used (1× M9 salts, 0.1 mM CaCl<sub>2</sub>, 2 mM MgSO<sub>4</sub>, 10 µg/ml thiamine hydrochloride and 1% (w/v) casamino acids, supplemented with 0.4% glycerol and antibiotics as required) and an Infinite 200 microplate reader (Tecan) was used for fluorescence and OD<sub>600</sub> measurements.
Note that 0.2% arabinose and 0.1 mM IPTG were added to the media to induce crRNA and Cas protein expression. For investigation of E. coli silencing over time, overnight cultures were diluted 1:100 in 100 μl of LB with inducers and required antibiotics and transferred to a 96-well plate. Fluorescence and OD<sub>600</sub> were measured in the microplate reader every 5 min. For fluorescence measurements the following wavelengths were used: 480 nm for Gfp excitation, 520 nm for Gfp emission, 399 nm for Bfp excitation and 456 nm for Bfp emission. Fluorescence readings were normalized to OD and corrected for background fluorescence by subtracting normalized fluorescence from a strain without fluorescent reporters. Relative fluorescence is calculated as the ratio between the tested cells and cells expressing non-targeting crRNA.

Flow cytometry

To investigate silencing in individual cells, cultures were diluted in modified M9-CA medium (above) and ampicillin (50 μg/ml), streptomycin (50 μg/ml) and chloramphenicol (25 μg/ml) when required. Samples were taken after 4 h of growth at 37°C; the cells were pelleted and resuspended in phosphate buffered saline before analysis in a BD LSR II flow cytometer using FACS Diva 6.0 software (BD Biosciences). Excitation wavelength was 488 nm and a 525/50 bandpass filter was used for detection; ~10 000 events were recorded per sample. For data handling FlowJo 7.6.5 software (FlowJo) was used.

Total RNA isolation and northern blots

Total RNA was extracted by hot phenol method (39) from cultures grown as described for flow cytometry (above). The RNA was DNase I-treated (Thermo Scientific) according to manufacturer’s instructions. 1 μg of total RNA was separated on a denaturing agarose gel (1.5% agarose, 1× MOPS, 25 mM guanidine thiocyanate) and transferred to a Hybond-N+ membrane (GE Healthcare Life Sciences). Excitation wavelength was 488 nm and a 525/50 bandpass filter was used for detection; ~10 000 events were recorded per sample. For data handling FlowJo 7.6.5 software (FlowJo) was used.

RESULTS

Design of silencing system

To test our hypothesis of Cascade-based transcription silencing (Figure 1), we used an E. coli cas3-knockout strain, where we overexpressed Cascade from an IPTG-inducible T7 promoter (11) together with a CRISPR array targeting a gfp reporter gene. The gfp target was encoded on the chromosome or on a low-copy plasmid (pEH9), constitutively expressed from a P<sub>LtetO-1</sub> promoter, which allowed quantitative fluorescence-based read-out of changes in gene expression. Minimal CRISPR arrays consisting of the transcribed leader (32) and two 29-bp repeats flanking a 32-bp spacer were designed to produce crRNAs that guide Cascade to various elements of the P<sub>LtetO-1</sub> promoter or gfp ORF. Pairs of spacers were designed to recognize the opposite strands of DNA at approximately the same location to investigate strand bias (Figure 2A). CRISPR arrays were expressed from plasmids using a P<sub>LtetO-1</sub> promoter. For analysis of simultaneous silencing of multiple targets we constructed a strain carrying constitutively expressed gfp and bfp genes on the chromosome. To test the system’s functionality in other species, we transferred the same Cascade, gfp target and CRISPR plasmids used in E. coli to S. typhimurium.

Cascade targeted to promoter silences expression

All CRISPR-expressing strains exhibited comparable growth but Gfp fluorescence was dramatically reduced in all strains that carried spacers against the P<sub>LtetO-1</sub> promoter compared to a control with non-targeting spacer (Figure 2B–E). Silencing was effective on both plasmid and chromosome targets, with similar efficiency (Figure 3A). Gfp repression was evident very early during growth, but we used a time point 3 h post-induction, in exponential growth phase, to quantify change in fluorescence (Figure 2E). With gfp on a plasmid, P2 and P3 crRNA targeting the -35 and -10 element, respectively, decreased the fluorescence the most, up to about 50-fold. Silencing required both Cascade and a targeting crRNA, demonstrating that silencing was not due to any activity of the crRNA on its own, or the overexpression of the Cas proteins (Supplementary Figure S1). Overall, these results show that Cascade binding to promoter elements results in highly effective transcription silencing. Analysis of individual cells expressing P1 and P2 crRNA by flow cytometry demonstrates symmetric fluorescence distribution, suggesting that all cells displayed approximately similar level of silencing (Figure 4A and B and Supplementary Figure S2). We hypothesize that Cascade stably bound to the promoter blocks RNA polymerase access, which in turn prevents transcription.

Cascade targeted to ORF also silences expression

We then designed spacers that targeted different regions within the gfp ORF to test whether Cascade binding can in-
Figure 1. Model of Cascade-mediated silencing system. A crRNA co-expressed with Cascade proteins form the Cascade RNA-protein complex. Efficient binding of Cascade to DNA is directed by base pairing of crRNA to a cognate sequence in the target DNA, which also requires an adjacent PAM motif. (A) Aided by the sigma factor, the RNA polymerase initiates transcription of a gene and elongation produces the mRNA. (B) When Cascade is targeted to the promoter it may interfere with transcription initiation. (C) When Cascade is targeted to a region within the ORF, it may affect transcription elongation.

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terfere with an elongating RNA polymerase. Again, pairs of ORF-targeting spacers were designed to bind the same location, on either the template or non-template strand (Figure 2A), and Gfp fluorescence was quantified 3 h post-induction (Figure 2F). With a plasmid target, the most efficient silencing when targeting the ORF was achieved with the T1 and T2 crRNA which both gave about 5-fold reduction of fluorescence. T1 and T2 target the template strand 269–301 and 437–469 bp, respectively, downstream of the transcription start site. NT1 gave only slight reduction of fluorescence and NT2 crRNA had no silencing effect (Figure 2F). Like promoter targets, ORF targets on chromosome and plasmid were silenced to same degree (Figure 3A). Our results indicate that Cascade targeted to the template strand of the ORF gives better silencing than when targeted to the non-template strand (Figure 2F), but that neither is as efficient as promoter targeting. Flow cytometry analysis revealed that cells expressing Cascade and NT1 or T1 crRNA displayed reduced fluorescence from both plasmid- and chromosome-encoded Gfp compared to non-targeting control, but again not to the same degree as promoter-targeting spacers (Figure 4A and B and Supplementary Figure S2). In contrast to when the promoter was targeted, the fluorescence distribution was not symmetrical when the ORF was targeted, particularly for plasmid-encoded Gfp (Figure 4A and Supplementary Figure S2A). The level of silencing thus appears to be different in individual cells, but it should be noted that very few cells displayed fluorescence equal to the non-silenced control.

Cascade-mediated silencing is long lasting

Since the in vivo nature of Cascade-DNA interaction is not known, we investigated the efficiency of our silencing system over time in the presence of inducers. Measurement of relative fluorescence from 3 to 23 h after induction with promoter-targeting crRNA demonstrated robust gfp silencing over time (Figure 5). For cultures expressing NT1, T1 or T2 crRNAs targeting the ORF gfp remained silenced over time (Supplementary Figure S3), while NT2 expressing cells did not show any silencing, even with prolonged induction.

Silencing of multiple targets

To determine if it is possible to use Cascade to silence several targets simultaneously, we constructed a strain carrying both gfp and bfp on the chromosome in a cas3 knockout background. In this strain we expressed plasmid-encoded Cascade and CRISPRs containing spacers targeting the bfp and/or the gfp promoter. The bfp spacer pair was designed to target both strands of the promoter (Figure 3B). Spacers targeting gfp or bfp promoters were only active on their intended targets and, when encoded in same CRISPR array, gfp and bfp expression were silenced simultaneously. Silencing was effective independent of spacer order in the CRISPR array (Figure 3C). A time point in stationary phase (9 h after inoculation) was chosen for quantification as small fluctuations in the measurements during exponential phase had large impact on normalized Bfp fluorescence values. However, the time course experiments show stable silencing over time (Supplementary Figure S4).

Cascade-based silencing is effective in different strains and species

To demonstrate the applicability of Cascade-mediated silencing in strains other than E. coli K-12, we tested our system in BL21AI and BL21DE3, two E. coli B strains that are distinctly different from K-12 (40). Silencing of Gfp expression was demonstrated in BL21AI with P1 (12-fold at
Figure 2. Cascade-mediated control of transcription. (A) Schematic drawing showing the positions of protospacers in the PLtetO-1 promoter and the gfp ORF (not drawn to scale). (B) E. coli cells expressing Cascade with P3 crRNA display Gfp silencing (left), compared to cells expressing Cascade with non-targeting crRNA (right). (C) Optical density analysis of cultures grown in 96-well plates demonstrates little difference in growth between strains expressing Cascade targeted to different regions of the PLtetO-1 promoter and strain expressing non-targeting Cascade (indicated by 'C'). (D) Gfp fluorescence of cultures grown in 96-well plates expressing Cascade guided to different regions of the PLtetO-1 promoter is dramatically lower than of cultures expressing non-targeting Cascade, demonstrating efficient silencing of gfp. (E) Relative Gfp fluorescence of cultures expressing Cascade guided to different parts of the promoter 3 h after induction compared to cultures expressing Cascade and non-targeting crRNA. (F) Relative Gfp fluorescence of cultures expressing Cascade guided to different part of the gfp ORF 3 h after induction compared to cultures expressing Cascade and non-targeting crRNA. Target is plasmid encoded in all experiments. Error bars represent SD.
Figure 3. Transcription silencing of chromosomal, plasmid and multiple targets. (A) Comparison of relative fluorescence 3 h after induction when target is on a plasmid or on the chromosome. (B) Schematic drawing showing the positions of protospacers in the J23101 promoter (not drawn to scale). (C) Relative Bfp and Gfp fluorescence when expressing different CRISPRs targeting \textit{bfp}, \textit{gfp} or both 9 h after induction. Order of spacers in double-spacer CRISPRs is indicated. Error bars represent SD. ‘C’ indicates strain expressing Cascade and non-targeting CRISPR.

most) and P2 (15-fold at most) targeting \textit{P}_{LtetO-1} promoter (Figure 2A and Supplementary Figure S5). Similar results were obtained with BL21DE3 (data not shown). Transfer of Cascade, CRISPR and target plasmid to \textit{S. typhimurium} LT2 demonstrate on average 8-fold silencing with P1 and 12-fold with P2 after 4 h of induction (Supplementary Figure S6) but longer induction resulted in more silencing (data not shown). The results demonstrate that Cascade-based transcription silencing is applicable in different strains and species but that the efficiency of silencing may differ.

Figure 4. Single-cell analysis of silencing. Flow cytometry analysis of Gfp fluorescence in individual cells when \textit{gfp} is on plasmid (A) or chromosome (B) and targeted by P1 or NT1 crRNA. Fluorescence of cells with non-targeting crRNA and cells lacking \textit{gfp} are shown for comparison.
Cascade silencing is caused by transcription interference

To assess if silencing is, as hypothesized, caused by transcription interference (and not by post-transcriptional effects), we measured \( gfp \) mRNA levels using northern blots and a probe binding downstream of the crRNA target sites, thereby detecting full-length \( gfp \) transcripts. Plasmid-derived \( gfp \) transcripts were non-detectable in cells expressing P1-4 crRNA, in a Cascade-dependent manner, but observed in cells expressing non-targeting crRNA (Figure 6A). T1, T2 or NT1 crRNA caused a reduction in \( gfp \) mRNA levels, but not as efficiently as P1–4 (Figure 6B), and NT2 did not reduce \( gfp \) transcripts. All results were consistent with fluorescence data (Figure 2E and F). A probe upstream of ORF target sites revealed that T1, T2 and NT1 crRNA gave rise to partial \( gfp \) transcripts of sizes corresponding to transcription termination at the position of the protospacer (Supplementary Figure S7), further indicating interference with transcription.

DISCUSSION

In this report, we used the sequence-specific DNA binding activity by the crRNA-guided Cascade complex as a simple and efficient method for targeted transcription silencing. The crRNA component guides Cascade to complementary target DNA containing an adjacent PAM motif. By changing the spacer sequence in the crRNA, Cascade can be directed to bind virtually any chosen stretch of DNA. Binding of Cascade to target DNA is strong, specific and virtually permanent, with very low \( K_d \) values (13 ± 1.4 nM, (16); 8 ± 4 nM, (41)). Cascade remains bound to DNA until released by Cas3-mediated DNA degradation (16). In absence of functional Cas3, Cascade could therefore be used to bind regulatory DNA elements, such as promoters, enhancers or operators, to prevent access of RNA polymerase, activators or repressors, respectively, thereby causing increase or decrease in transcript levels. In extension, any DNA-interacting element could potentially be affected by Cascade binding to its target site. Using \( E. coli \) and \( S. typhimurium \) as model systems, we show that Cascade targeted to promoter elements gives highly efficient expression silencing, determined both on transcript and protein level. Cascade silencing is specific and efficient, the silencing effect is stable over time and independent of growth phase. Cascade can be used to silence both chromosome and plasmid targets, as well as to silence several targets simultaneously. Silencing is also achieved when Cascade is targeted to the ORF albeit, for the spacers tested here, at lower efficiency than when targeting the promoter. Flow cytometry analysis of individual cells reveal that when targeting the promoter,
all cells are uniformly silenced to a high degree, but when targeting the ORF, cells display some variation in silencing, especially when the target is on a multi-copy plasmid. We speculate that actively transcribing RNA polymerase occasionally dislodges or by-passes DNA-bound Cascade and that this explain the cell-to-cell variation in expression. It is also possible that an RNA polymerase stalls as it occasionally dislodges or by-passes DNA-bound Cascade or silenced. The larger size of Cascade compared to Cas9, it is conceivable that Cascade produces less off-target effects than dCas9.

Qi et al. (26) achieved the best repression with dCas9 that targeted the non-template strand of the ORF close to the promoter, and repression efficiency decreased dramatically with increasing distance of the protosparer from the ORF start site. This suggests that distance of the protosparer from the start site is an influential parameter for dCas9. Further, when targeting a promoter on the chromosome, not all spacers gave repression (27). These observations suggest that dCas9-mediated gene silencing is affected by several parameters, not all of which are fully understood. Cascade directed to the template strand of the ORF gave slightly better silencing than when targeted to the non-template strand, but due to the limited number of spacers tested we cannot rule out that the differences are caused by the efficiency of the individual spacers. Targeting by Cascade proceeds via formation of an R-loop (45). A stronger inhibition of RNA polymerase when Cascade is bound to the template strand (compared to when bound to the non-template strand) can be rationalized as RNA polymerase primarily needs access to the template strand for elongation.

Our system uses an unperturbed Cascade complex which, unlike systems based on engineered dCas9, allows for the use of endogenous cellular components for efficient programmable silencing in cells where Cas3 is absent, inactive or silenced. The larger size of Cascade compared to Cas9 may also be an additional advantage when interfering with RNA polymerase-DNA interaction. Cascade also uses a different set of PAMs than Cas9 and may have a different range of species and strains where it can be used. However, in-depth comparative studies are required to exactly determine the advantages and disadvantages of different programmable silencing methods.

Design of new CRISPRs is simple and cost-effective compared to constructing gene knockouts and, by using inducible CRISPR-Cas systems, essential genes can be targeted, which obviates the need for time-consuming con-
E.coli ingusingCascade. We have used activator genes, as a silencing switch. Tuning of target expression may get DNA. The Cascade-mediated transcriptional silencing demonstrated in this work is a powerful new addition to the rapidly emerging CRISPR-Cas toolbox for genetic modifications and gene regulation, expanding the versatility and applicability of CRISPR-Cas tools.

SUPPLEMENTARY DATA
Supplementary Data are available at NAR Online.

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REFERENCES
1. Browning,D.F. and Busby,S.J. (2004) The regulation of bacterial transcription initiation. Nat. Rev. Microbiol., 2, 57–65.
2. Winkler,W.C. and Breaker,R.R. (2005) Regulation of bacterial gene expression by riboswitches. Annu. Rev. Microbiol., 59, 487–517.
3. Ausländer,S. and Fussenegger,M. (2013) From gene switches to mammalian designer cells: present and future prospects. Trends Biotechnol., 31, 155–168.
4. Beerli,R.R., Segal,D.J., Dreier,B. and Barbas,C.F. 3rd. (1998) Toward controlling gene expression at will: specific regulation of the erbB-2/HER-2 promoter by using polydactyl zinc finger proteins constructed from modular building blocks. Proc. Natl. Acad. Sci. U.S.A., 95, 14628–14633.
5. Cong,L., Zhou,R., Kuo,Y.C., Cunniff,M. and Zhang,F. (2012) Comprehensive interrogation of natural TALE DNA-binding modules and transcriptional repressor domains. Nat. Commun., 3, 968.
6. Klug,A. (2010) The discovery of zinc fingers and their applications in gene regulation and genome manipulation. Annu. Rev. Biochem., 7, 213–231.
7. Van der Oost,J., Westra,E.R., Jackson,R.N. and Wiedenheft,B. (2014) Unravelling the structural and mechanistic basis of CRISPR-Cas systems. Nat. Rev. Microbiol., 12, 479–492.
8. Bolotin,A., Quinquis,B., Sorokin,A. and Ehrlich,S.D. (2005) Clustered regularly interspaced short palindromic repeats (CRISPRs) have spacers of extrachromosomal origin. Microbiology, 151, 2551–2561.
9. Mojica,F.J., Diez-Villaseñor,C., García-Martínez,J. and Soria,E. (2005) InterViewing sequences of regularly spaced prokaryotic repeats derived from foreign genetic elements. J. Mol. Evol., 60, 174–182.
10. Pourcel,C., Salvignol,G. and Vergnaud,G. (2005) CRISPR elements in Yersinia pestis acquire new repeats by preferential uptake of bacteriophage DNA, and provide additional tools for evolutionary studies. Microbiology, 151, 653–663.
11. Brouns,S.J., Jore,M.M., Lundgren,M., Westra,E.R., Slijkhuis,R.J., Snijders,A.P., Dickman,M.J., Makarova,K.S., Koonin,E.V. and van der Oost,J. (2008) Small CRISPR RNAs guide antiviral defense in prokaryotes. Science, 321, 960–964.
12. Carte,J., Wang,R., Li,H., Terns,R.M. and Terns,M.P. (2008) Cas6 is an endoribonuclease that generates guide RNAs for invader defense in prokaryotes. Genes Dev., 22, 3489–3496.
13. Haurwitz,B.E., Jinke,M., Wiedenheft,B., Zhou,K. and Doudna,J.A. (2010) Sequence-structure and structure-specific RNA processing by a CRISPR endonuclease. Science, 329, 1355–1358.
14. Gasiunas,G., Barrangou,R., Horvath,P. and Siksny s,V. (2012) Cas9-crRNA ribonucleoprotein complex mediates specific DNA cleavage for adaptive immunity in bacteria. Proc. Natl Acad. Sci. U.S.A., 109, 2579–2586.
15. Jinke,M., Chylinski,K., Fonfara,I., Hauer,M., Doudna,J.A. and Charpentier,E. (2012) A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. Science, 337, 816–821.
16. Westra,E.R., van Erp,P.B., Kunne,T., Wong,S.P., Staals,R.H., Seegers,C.L., Bol len,S., Jore,M.M., Sem enova,E., Severinov,K. et al. (2012) CRISPR immunity relies on the consecutive binding and degradation of negatively supercoiled invader DNA by Cascade and Cas9. Mol. Cell, 46, 595–605.
17. Garneau,J.E., Dupuis,M.E., Villion,M., Romero,D.A., Barrangou,R., Boyaval,P., Fremaux,C., Horvath,P., Magadan,A.H. and Moineau,S. (2010). The CRISPR/Cas bacterial immune system cleans bacteriophage and plasmid DNA. Nature, 468, 67–71.
18. Sampson,T.R., Saroj,S.D., Llewellyn,A.C., Tzeng,Y.L. and Weiss,D.S. (2013) A CRISPR/Cas system mediates bacterial innate immune evasion and virulence. Nature, 497, 254–257.
19. Makarova,K.S., Haft,D.H., Barrangou,R., Brouns,S.J., Charpentier,E., Horvath,P., Moineau,S., Mojica,F.J., Wolf,Y.I., Yakunin,A.F. et al. (2011) Evolution and classification of the CRISPR-Cas systems. Nat. Rev. Microbiol., 9, 467–477.
20. Crisola,J., Vergnaud,G. and Pourcel,C. (2007) The CRISPRdb database and tools to display CRISPRs and to generate dictionaries of spacers and repeats. BMC Bioinformatics, 8, 172.
21. Mojica,F.J., Diez-Villaseñor,C., García-Martínez,J. and Almendros,C. (2009) Short motif sequences determine the targets of the prokaryotic CRISPR defence system. Microbiology, 155, 733–740.
22. Wiedenheft,B. van Duin,E., Bultema,J.B., Waghmare,S.P., Zhou,K., Barendregt,A., Westphal,W., Heck,A.J., Boekema,E.J., Dickman,M.J. et al. (2011) RNA-guided complex from a bacterial immune system enhances target recognition through seed sequence interactions. Proc. Natl Acad. Sci. U.S.A., 108, 10092–10097.
23. Heidrich,N. and Vogel,J. (2013) Same same but different: new structural insight into CRISPR-Cas complexes. Mol. Cell, 52, 4–7.
24. Cong,L., Ran,F.A., Cox,D., Lin,S., Barretto,R., Habib,N., Hsu,P.D., Wu,X., Jiang,W., Marraffini,L.A., etal. (2013) Multiplex genome engineering using CRISPR/Cas systems. Science, 339, 819–823.
25. Mali,P., Yang,L., Esvelt,K.M., Aach,J., Guell,M., DiCarlo,J.E., Norville,J.E. and Church,G.M. (2013) RNA-guided human genome engineering via Cas9. Science, 339, 823–826.
26. Qi,L.S., Larson,M.H., Gilbert,L.A., Doudna,J.A., Weissman, J.S., Arkin,A.P. and Lim,W.A. (2013) Repurposing CRISPR as an RNA-guided platform for sequence-specific control of gene expression. Cell, 152, 1173–1183.
27. Bikard, D., Jiang, W., Samai, P., Hochschild, A., Zhang, F. and Marraffini, L.A. (2013) Programmable repression and activation of bacterial gene expression using an engineered CRISPR-Cas system. *Nucleic Acids Res.*, 41, 7429–7437.

28. Cheng, A.W., Wang, H., Yang, H., Shi, L., Katz, Y., Theunissen, T.W., Rangarajan, S., Shivalila, C.S., Dadon, D.B. and Jenisch, R. (2013) Multiplexed activation of endogenous genes by CRISPR-on, an RNA-guided transcriptional activator system. *Cell Res.*, 23, 1163–1171.

29. Gilbert, L.A., Larson, M.H., Morsut, L., Liu, Z., Brar, G.A., Torres, S.E., Stern-Ginossar, N., Brandman, O., Whitehead, E.H., Doudna, J.A. et al. (2013) CRISPR-mediated modular RNA-guided regulation of transcription in eukaryotes. *Cell*, 154, 442–451.

30. Farzadfard, F., Perli, S.D. and Lu, T.K. (2013) Tunable and multifunctional eukaryotic transcription factors based on CRISPR/Cas. *ACSSynth.Biol.*, 2, 604–613.

31. Baba, T., Ara, T., Hasegawa, M., Takai, Y., Okumura, Y., Baba, M., Datsenko, K.A., Tomita, M., Wanner, B.L. and Mori, H. (2006) Construction of *Escherichia coli* K-12 in-frame, single-gene knockout mutants: the Keio collection. *Mol.Syst.Biol.*, 2, 2006.0008.

32. Cherepanov, P.P. and Wackernagel, W. (1995) Gene disruption in *Escherichia coli*: TcR and KmR cassettes with the option of Flp-catalyzed excision of the antibiotic-resistance determinant. *Gene*, 158, 9–14.

33. Yosef, I., Goren, M.G., Kiro, R., Edgar, R. and Qimron, U. (2011) High-temperature protein G is essential for activity of the *Escherichia coli* clustered regularly interspaced short palindromic repeats (CRISPR)/Cas system. *Proc. Natl Acad. Sci. U.S.A.*, 108, 20136–20141.

34. Studier, F.W. and Moffatt, B.A. (1986) Use of bacteriophage T7 RNA polymerase to direct selective high-level expression of cloned genes. *J. Mol. Biol.*, 189, 113–130.

35. Lutz, R. and Bujard, H. (1997) Independent and tight regulation of transcriptional units in *Escherichia coli* via the LacR/O, the TetR/O and AraC/I1-I2 regulatory elements. *Nucleic Acids Res.*, 25, 1203–1210.

36. Urban, J.H. and Vogel, J. (2007) Translational control and target recognition by *Escherichia coli* small RNAs in vivo. *Nucleic Acids Res.*, 35, 1018–1037.

37. Datsenko, K.A. and Wanner, B.L. (2000) One-step inactivation of chromosomal genes in *Escherichia coli* K-12 using PCR products. *Proc. Natl Acad. Sci. U.S.A.*, 97, 6640–6645.

38. Pougach, K., Semenova, E., Bogdanova, E., Datsenko, K.A., Djordjevic, M., Wanner, B.L. and Severinov, K. (2010) Transcription, processing and function of CRISPR cassettes in *Escherichia coli*. *Mol. Micro.*, 77, 1367–1379.

39. Blombberg, P., Wagner, E.G. and Nordström, K. (1990) Control of replication of plasmid R1: the duplex between the antisense RNA, CopA, and its target, CopT, is processed specifically in vivo and in vitro by RNase III. *EMBO J.*, 9, 2331–2340.

40. Studier, F.W., Daegelen, P., Lenski, R.E., Maslov, S. and Kim, J.F. (2009) Understanding the differences between *Escherichia coli* B strains REL606 and BL21(DE3) and comparison of the *E. coli* B and K-12 genomes. *J. Mol. Biol.*, 394, 653–680.

41. Sashital, D.G., Wiedenheft, B. and Doudna, J.A. (2012) Mechanism of foreign DNA selection in a bacterial adaptive immune system. *Mol. Cell.*, 46, 606–615.

42. Fu, Y., Foden, J.A., Khayter, C., Maeder, M.L., Reyon, D., Joung, J.K. and Sander, J.D. (2013) High-frequency off-target mutagenesis induced by CRISPR-Cas nucleases in human cells. *Nat. Biotechnol.*, 31, 822–826.

43. Kasas, C., Arola, S., Singh, R., Thorpe, J. and Adli, M. (2014) Genome-wide analysis reveals characteristics of off-target sites bound by the Cas9 endonuclease. *Nat. Biotechnol.*, 32, 677–683.

44. Semenova, E., Jore, M.M., Datsenko, K.A., Semenova, A., Westra, E.R., Wanner, B., van der Oost, J., Brouns, S.J. and Severinov, K. (2011) Interference by clustered regularly interspaced short palindromic repeat (CRISPR) RNA is governed by a seed sequence. *Proc. Natl Acad. Sci. U.S.A.*, 108, 10098–10103.

45. Jore, M.M., Lundgren, M., van Duijn, E., Bulthman, J.B., Westra, E.R., Waghmare, S.P., Wiedenheft, B., Pul, U., Wurm, R., Wagner, R. et al. (2011) Structural basis for CRISPR RNA-guided DNA recognition by Cascade. *Nat. Struct. Mol. Biol.*, 18, 529–536.