The Effect of Administration of Extract from Areca Nut Seeds (Areca Catechu L) on the Estradiol and Estrus Cycle Balb/C Female Rats

T Rina Aritonang1*, R Natzir2, A Wardihan Sinrang3, M Nasrum Massi4, M Hatta4, and Kamelia5

1 Midwifery Program, Sekolah Tinggi Ilmu Kesehatan Medistra Indonesia, Indonesia.
2 Department of Biochemistry, Faculty of Medicine, Universitas Hasanudin, Makassar 90245, Indonesia.
3 Department of Physiology, Faculty of Medicine, Universitas Hasanudin, Makassar 90245, Indonesia.
4 Molecular Biology and Immunology Laboratory for Infections Diseases, Faculty of Medicine, Universitas Hasanudin, Makassar 90245. Indonesia.
5 Department of Dental Nursing, Poltekkes Kemenkes Tasikmalaya, Tasikmalaya, Indonesia

*tetty.rina.2109@gmail.com

Abstract. Population control is very important for individual and national welfare. Although various synthetic contraceptive agents are available, their use is associated with severe side effects. Therefore, an approach is taken to identify new antifertility agents from natural sources. This study was conducted to determine the effect of areca nut extract on estradiol levels and the estrus cycle of female balb/c mice. The experimental animal models used were fifteen adult female balb/c mice, aged 8-12 weeks, weight 20-40 g and induced for 1 week with water extract of areca nut with a dose of 1 g/200 g body weight of mice (K1), 2 g/200 g of body weight of mice (K2) and distilled water in the control group (K0). The estrus cycle was identified by a daily assessment of the relative ratio of nucleated epithelial cells, cornified squamous epithelial cells and leukocytes found at the time of vaginal swabs for 15 days continuously. At the proestrus stage blood is taken from the tail. Serum estradiol levels were determined by ELISA technique. There was a decrease in serum estradiol levels in the treatment group 1 (K1) 1.9 times compared to the control group (K0) and decreased estradiol levels in the treatment group 2 (K2) 5.29 times compared to the control group (K0). Changes in the duration of the estrus cycle occurred in treatment group 1 (K1) to be elongated and treatment group 2 (K2) became non-estrus, while the control group (K0) did not change the duration of the estrus cycle (normal). The decrease in serum estradiol levels affects the estrus cycle of mice (P = 0.000). The administration of areca nut extract has the effect of reducing serum estradiol levels and disrupting the duration of the estrous cycle of female balb/c mice. This finding has important implications for the development of contraception in women.

1. Introduction
The explosion in population caused a number of adverse effects, such as environmental damage, global warming, famine, and disease development [1]. In 2025 there are estimated numbers in
Indonesia the population reached 273.65 million [2]. Overcoming this problem has been many modern contraceptive methods are used, but users of modern contraceptive methods worry about side effects that can affect their daily lives [3]. The development of science in the processing of natural ingredients has experienced a rapid increase. Nearly 80% of the world’s population depends on traditional medicines for primary health care, most of which involve the use of plant extracts. [4]. Herbal preparations have also been used since time immemorial for their effects on reproductive health in particular to suppress fertility, regulate the menstrual cycle, eliminate dysmenorrhea, treat prostate enlargement, menopausal symptoms, breast pain and pain during and after childbirth [5]. Wrong one ingredient that is widely studied is betel nut (Areca catechu) [6]. The main component contained in the areca nut is Arecoline [7]. There are four alkaloids playing in areca nut, namely arecoline (7.5 mg / g), arecaidine (1.5 mg / g), guvacoline (2.0 mg / g) and guvacine (2.9 mg / g) [8]. Giving areca can cause cytotoxicity [9], apoptosis [10], hepatoprotective, [11] hypoglycemic, [12] astringent, vermifugal, sialogogue, [13] antibacterial, antioxidant, antiseptic, bronchostimulant, euphorient, and wound healing properties. [14] In addition, it has abortifacient, anti-implantation and antifertility activities [15]. Research has revealed that betel nut causes morphological changes such as hormonogenesis stimulation and spermatogenesis disorders. [16] This certainly opens wider opportunities to study the effects of areca nut on estradiol levels and the estrus cycle of female mice in an effort to unmask the potential of areca nut as a candidate for antifertility in women.

2. Research Methods
The experimental procedure was carried out at the Laboratory of Molecular Biology and Immunology, Faculty of Medicine, Hasanuddin University, Makassar, Indonesia. This research is an in vivo post-design experimental research conducted in the period from December 2016 to April 2017. The experiment was approved by the PT Health and Health Research Ethics Committee Faculty of Medicine Hasanuddin University Makassar Indonesia (Number: 1625 / H4.8.4.5.31 / PP36-COMETIC / 2016).

2.1. Ingredients for Areca Palm Seeds (Are cacatechu L).
The areca nuts that are used come from areca nuts that have all yellow skin. This type of areca nut is known to contain lower levels of arecoline than young areca nut. Areca nuts are selected in good condition, fresh, not rotten or moldy.

How to make water fraction of areca nut extract. Areca nut shelled, crushed and mashed using a hammer. Then weighed as much as 100 g and added 200 mL of distilled water, heated to the remaining 100 mL of areca nut extract, so that every 1 mL contains 2 g of areca nut extract [7].

2.2. Experimental Animals and Treatment
The experimental animals used in this study were adult female balb / c mice. Age 8 - 12 weeks and weight of experimental animals 20-40 g. The chosen strain is balb / c. The total number of Balb / C mice used was 15 divided into 3 groups, each group consisted of 5 mice. Mice are adapted for 1 week in the laboratory and given standard feed. Mice are grounded at ordinary ambient temperatures with a cycle of 12 hours a day and 12 hours a night and are given food and drink ad libitum (acidified water). The administration of areca nut extract is done by force-feeding directly to the stomach using a 5 ml syringe that has been modified. Determination of the dosage of areca nut extract according to the results of the study [17] Group: K0 = given distilled water, K1 = given areca nut extract 1 g / 200g body weight of mice, K2 = given areca nut extract 2 g / 200gr body weight of mice, each given for 7 days.

2.3. How to take blood
Blood sampling in the tail vein was 0.1 mL using microhemactocrit syringe. The time for taking balb / c blood serum is done when the estrus cycle is at the proestrus stage. Proestrus stage determination is done by examining vaginal cytology. Blood is collected and centrifuged to get serum. Serum is stored
in a sterile tube and placed in a refrigerator temperature of -20 ° C. Pemeriksaan Estradiol levels using Elisa Kit.

2.4. Vaginal cytology method
Examination of vaginal reviews is done to examine the epithelial cell image in the vagina of the mouse so that it can determine each stage of the estrus cycle. Vaginal cells were collected using a pipette filled with 0.2 ml of physiological salt water and inserted into the vagina of the mouse and sucked back (2-3 x). Vaginal fluid is transferred to a dry glass slide by dripping and making 2 cultures in one slide. The slides were dried with bunsen fire and air, then stained with giemsa liquid in 45 seconds. The slide is rinsed with physiological fluid, covered with coverslip, and viewed directly at 400 magnification under bright field lighting. Identification of each stage of the estrus cycle from the daily assessment of the relative ratio of nucleated epithelial cells, cornified squamous epithelial cells and leukocytes found at the time of vaginal swab. The proestrus stage is determined by finding a group of round cells, nucleated epithelial cells [18]. Ulas vagina is carried out every day from 9:00 a.m. to 10:00 a.m. for 15 days continuously. Vaginal cytology examination starting from day 1 (first) after intervention treatment of areca nut fraction was completed.

2.5. Statistical Analysis
The data obtained were tested for normality with the Kolmogorv-Smirnov test and continued with a homogeneity test. The test results show that all data are normally distributed and homogeneous. The data was then tested using parametric analysis, namely paired T test (Pair T - Test). To see the estradiol levels, Anova was tested

3. Result and Discussion
Research on the effect of giving Pinang fruit seed extract (Areca catechu. L) to estradiol (E2) and estrous cycles in In Vitro results obtained as follows

| Table 1. Effect of areca nut extract on estradiol levels |
| --- | --- | --- | --- |
| Variable | Group | Mean | Elementary school |
| Estradiol levels | K0 (n = 5) | 626,691 | 31,833 |
| | K1 (n = 5) | 329,421 | 46,469 |
| | K2 (n = 5) | 118,475 | 49,764 |

Value is expressed in the mean, SD = Std. Deviation, sig = significant, n = 5 each group; P = 0.000 compared to control

Analysis
In this study there was a decrease in serum estradiol (E2) levels in the treatment group 1 (K1) 1.9 times compared to the control group (K0) and decreased estradiol levels in the treatment group 2 (K2) 5.29 times compared to the control group (K0) after being given areca nut extract for 7 days. Giving betel extract has an effect on decreasing serum estradiol hormone levels

| Table 2. Effect of Pinang seed extract on the duration of the estrus cycle |
| --- | --- | --- | --- |
| Group | Estrus cycle | Mean | Elementary school |
Table 3. Effect of Estradiol hormones on the estrus cycle

|              | Sum of Squares | df | Mean Square | F     | Sig. |
|--------------|----------------|----|-------------|-------|------|
| E2pgpermL    |                |    |             |       |      |
| Between Groups | 546579,837    | 2  | 25,634      | .000  |      |
| Within Groups| 127933,508    | 12 | 10661.126   |       |      |
| Total        | 674513,344    | 14 |             |       |      |

n = 5 each group, sig 0.000

Analysis

Changes in the estrus cycle occurred in treatment group 1 (K1) to be elongated and treatment group 2 (K2) became non-estrus after giving betel seed extract for 7 days, while the control group (K0) did not experience a change in the estrus cycle. The length of observation is 15 continuously.

Table 3. Effect of Estradiol hormones on the estrus cycle

|                  | Sum of Squares | df | Mean Square | F     | Sig. |
|------------------|----------------|----|-------------|-------|------|
| K0 (n = 5)       |                |    |             |       |      |
| K1 (n = 5)       |                |    |             |       |      |
| K2 (n = 5)       |                |    |             |       |      |

Value is expressed in the mean, SD = Std. Deviation, n = 5 each group.

Analysis

Decrease in serum estradiol hormone levels has a significant effect (sig 0.000) on changes in the duration of the estrus mice cycle. Where estrus status of treatment group 1 (K1) is elongated and treatment group 2 (K2) is not found in estrus phase.

The results of this study indicate that after 7 days the administration of areca nut extract has a reduced effect on estradiol levels (E2) at the proestrus stage. Serum estradiol levels decrease with increasing doses of areca nut extract. Ovarian hormones are produced by various types of ovarian cells such as granulosa cells from follicle mature and corpus luteum. [19] The lack of hormone imbalance causing irregularity of ovarian function and the duration of the estrous cycle [20]. This study is different from what is conveyed by research of sebelimnya wherein at proestrus phase serum estradiol concentration is higher than all other estrus phase [21]. Decreased serum estradiol levels after administration of the host seed extract (arecacatechu L.) in animals caused by arecoline content contained in the betel nut causes hypoglycemic, hipolipidemia and the onset of cytotoxicity that spur apoptosis of germ cells [22]-[24]. Supplementation of arecanut extract significantly reduced triglyceride absorption and plasma lipid concentration [25]. Absorption of free cholesterol in the intestine and small intestine pCEase activity is significantly reduced when supplementing areca nut extract [26]. Cholesterol is a precursor for steroidogenesis of ovarian endocrine tissues, estrogens, progestins, and androgens. [27]. Most species specifically use LDL cholesterol as a precursor to ovarian steroid synthesis. A da positive relationship between HDL content of apolipoprotein E and the importance of HDL cholesterol as a precursor for steroidogenesis. Cholesterol used for steroid synthesis by ovarian tissue can originate from cellular uptake of lipoprotein cholesterol [25]. In addition arecoline found in Areca catechu has hypoglycemic activity and becomes alloxanize, [12], has potent α-glucosidase for inhibitors and is effective in suppressing rat blood glucose elevation [28]. Changes in the use of glucose by steroid-sensitive reproductive tract tissues can affect the disruption of reproductive ability, [29]. Insulin is the key that opens the door to tissue cells, inserts sugar in the cell and closes the door again. Insulin stimulates ovarian androgen production, [30] There was a significant correlation between basal levels of plasma insulin and androstenedione found between regions of plasma insulin response. The results of the study show that hydrogenation is correlated with hyperinsulimism [31]. Cell type changes during the estrus cycle are indicative of endocrine events in mice [18] Histologic
changes and cytology of vaginal mucosa depend on sex steroid hormones. The main hormone that induces changes in the vaginal mucosa is estradiol [32]. During proestrus, vaginal papules contain many nucleated epithelial cells and several leukocytes [33]. In this study hormone levels decreased significantly (0.000 sig) to estrous mouse cycles, where the duration of the estrus cycle treatment group 1 (K1) became elongated (> 5 days) and treatment group 2 (K2) was not found estrus phase (no experiencing estrus stage) while the duration of the control group is normal (4-5 days). This condition occurs because during the observation of 15 consecutive days found leukocytes that dominated the field of view (diestrus) which lasted more than 4 days in the treatment group. If the majority of cells are leukocytes, then it is labeled as a diestrus phase. [34] Previous studies that revealed the estrous cycle in mice lasted for 5 days [35]. Each phase in the cycle is determined based on the shape of the epithelial cell there is a vaginal cytology observation. Diestrus is the longest stage that lasts more than 2 days [36]. In the diestrus phase the vaginal contents are consistently deficient in cornified cells and leukocytes predominate in the smear. The frequency of cornified epithelial cells is reduced and nucleated epithelial cells begin to be detected only before the transition to proestrus [33]. The estrus cycle consists of 4 phases, namely proestrus, estrus, metestrus and diestrus [36]. This proves that the decreased estradiol level after giving areca nut extract results in a change in the duration of the estrus cycle. Lengthening of the diestrus phase indicates the absence of mature graafian follicles or nonsecondary follicular maturation. Estrogen positive feedback that stimulates de novo neuroprogesterone synthesis to trigger important luteinizing surge hormone for ovulation [37]. At the stage of diestrus, the serum estradiol concentration is lower than proestrus [21]. In this study, the pathway from estradiol cannot be ascertained due to the administration of areca nut water extract

4. Conclusion
The parameters in this study indicate that areca nut extract (A. Catechu) has a reduced effect on estradiol levels and disruption in the duration of the estrus cycle. Serum estradiol levels decrease with increasing doses of areca nut extract. Decreased estradiol levels are associated with changes in the duration of the estrus cycle. This finding has important implications for the development of natural contraception in women.

Acknowledgment
The research was supported by the ministry of research, technology and higher education Indonesia, College of Health Sciences M edistra Indonesia Bekasi and laboratory Molecular Biology and Immunology, Faculty of Medicine Hasanuddin University Makassar Indonesia.

References
[1] P. ST, Amory JK, Bremner WJ. Advances in male contraception. Endocrine Revies. 29 (4), 465-493, 2008
[2] Fauzi A, Lucianawaty M, Hanifah L, Bernadette N, Indonesian Population 2025 Will Reach 273.65 Million Souls. http: //site.kesrepro.info/info/agu/2005/ info02.htm, 2005.
[3] Effie K Chipeta, Wanangwa Chimwaza, and Linda Kalilani-Phiri (2010), Contraceptive Knowledge, Beliefs and Attitudes in Rural Malawi: Misinformation, Misbeliefs and Misperceptions, Malawi Med J. 2, 38-41.
[4] Sandhya B, Thomas S, Isabel W, Shenbangarathai R, Ethnomedicinal plants used by Valaiyan community of Piramalai Hills (Reserved Forest), Tamil Nadu, India. A pilot study. Afr. J. Traditional Alter Complement 1, 101-14, 2006
[5] Williamson EM, Okpako DT, Evans FJ, Pharmacological methods in phytotherapy research. Selection, preparation and pharmacological evaluation of plant material. John Wiley and Sons, London 1, 191-212, 1996
[6] Syadida, F., SS Karyono, and HM Mahdi. The use of areca nut seedlings (Areca catechu) as miotikum in rabbit eyes (Lepus sp.). Medical Journal Brawijaya. XXI (3), 7-13, 2005
[7] Shyi-Wu, Guey-Shyang WH, Te-Jung C, Wang PS, Effects of arecoline on testosterone release
rat. *Am J Physiol Endocrinil Metab.* 295, E497-E504, 2008

[8] Wang CK, Lee WH, Peng CH, Contents of phenolics and alkaloids in Areca catechu Linn. during maturation. *J Agric Food Chem.* 45,1185-1188, 1997.

[9] Jeng JH, Chang MC, and Hahn LJ, quid-associated chemical carcinogenesis, Current awareness and future perspectives. *Oral. Oncol. Pathol. Med* 28, 64-71, 2001

[10] Aulanni'am, Akmal, M, Rosmaidar, Anfertility Effect of Areca catechu as an Apoptotic Agent in Rattus norvegicus Testicular Tissue Cells. *Journal of Veterinary Media.* 23 (3), 179-183, 2007

[11] Pimolpan P, Saruth N, Rapepol B, Hepatoprotective potential of extracts from seeds of Areca catechu and Nutgalls of Quercus infectoria. *Molecules.* 14,4987-5000, 2009. [PubMed]

[12] Chempakam B, Hypoglycemic activity of arecoline in betel nut Areca catechu Indian L. *J Exp Biol* 31, 474-5, 1993. [PubMed]

[13] Nadkarni KM. 2nd ed. Bombay, Popular Book Department, Indian, Materia Medica, pp. 219-21, 1954

[14] Azeez S, Amudhan S, Adiga S, Vasanthakumar A, Rao N, Udupa AL, Sanath H, Wound healing profiles of Areca catechu extracts on different wound models in Wistar rats. *Kuwait Med J.*, 39, 48-52, 2007.

[15] Duke JA, Handbook of Medicinal Herbs, Florida: CRC Press, 2002, editor. 2nd ed.

[16] Azeez S, Amudhan MS, Central Plantation Crops Research Institute. Pharmacological uses of areca nut; pp. 40-2, 2006

[17] Akmal M, Chaniif Mahdi, Aulanni'am, Increased Testosterone Concentration in Mice Due to Exposure to Areca Seed Water Extract, Syiah Kuala University, Banda Aceh, December *Veterinary Journal*, Vol. 11 No. 4, 244-250, 2010

[18] McLean, A C Nicolas Valenzuela, Stephen Fai, Steffany AL Bennett, Vaginal Lavage Performing, Crystal Violet Staining, and Vaginal Cytological Evaluation for Mouse Estrous Cycle Staging Identification ... URL: http://www.jove.com/video/ 4389. DOI: doi: 10.3791 / 4389, 2012

[19] SS teacher, Fundamentals and Biomedical Implications, New Delhi, Narosa Publishing House, pp. 10-346, 1998

[20] Cincotta C, Sanogo R, Occhiuto procera on estrous cycle and on estrogenic functionality in rats. *Farmacuco* 56, 373-8, 2001 [PubMed]

[21] Gal Arnon, Lin Po-Ching, Barger Anne M, MacNeill Amy L, and KoCheMyong, Vaginal fold histology reduces the variability introduced by vaginal exfoliative cytology in the classification of mouse cycle stages Published in final edited form as, Toxicol Pathol. 42, 8, 1212-1220, 2014 (doi: 10.1177 / 0192623314526321)

[22] Jeng JH, Chang MC, and Hahn LJ, Quid-associated chemical carcinogenesis, Current awareness and future perspectives, Oral, Oncol, Pathol, Med, 28, 64-71, 2001.

[23] Meiyanto E. Ratna Asmah Susidarti, Sri Handayani, and Fitria Rahmi, Ethanolic Extract of Areca catechu L. fruit seeds can inhibit proliferation and promote apoptosis of MCF-7 cells. *Indonesian Pharmacy Magazine*, 19, 1, 12-19, 2008

[24] Sinha, A. and AR Rao, Induction of shape abnormality and unscheduled DNA syntesis by arecoline in the germ cells of mice, *Muta. Res*, 158,189-192, 1985.

[25] Byun SJ, Kim HS, Jeon SM, Park YB and MS Choi, Supplementation of Areca catechu L. extract alters triglyceride absorption and cholesterol metabolism in rats. *Ann Nutr Metab*, 45, 6, 279-84, 2001

[26] YB Park, SM Jeon, SJ Byun, HS Kim and MS Choi, Cholesterol-free Cholesterol Absorption was lowered by supplementation of Areca catechu L. extract in mice. *Life Sci.* 70, 16, 1849-59, 2002.

[27] Shivalingappa H, Satyanarayana ND, Purohit MG, Sharanabasappa A, Patil SB., Effect of ethanol extract of Rivea hypocrateriformis on the estrous cycle of the rat. *J Ethnopharmacol*, 82, 11-7, 2002. [PubMed]
[28] M Senthil Amudhan, and V Hazeena Begum, Protective effects of Areca catechu extract on gastric mucosa induced by ethanol lesions in online pharmacological rats, 1, 97-106, 2008

[29] David R Garris, Douglas L Coleman and Carl Morgan Morgan, Age-and Diabetes-related Changes in Tissue Glucose Uptake and Estradiol Accumulation in the C57BL / KsJ Mouse. Diabetes, 34, 1, 47-52, 1985. https://doi.org/10.2337/diab.34.1.47

[30] Callum LIVINGSTONE, Mary COLLISON, Sex steroids and insulin resistance. Clinical Science, 102, 2, 151-166; 2002, DOI: 10.1042 / cs1020151

[31] George A. Burghen James R. Givens Abbas E, Kitabchi Correlation of Hyperandrogenism with Hyperinsulinism in Polycystic Ovarian Disease Endocrinology & Metabolism, Volume 50, Issue 1, Pages 113-116, 1980 https://doi.org/10.1210/jcem-50-1-113

[32] Gupta PD, Vijayasaradhi S, Reddy AG, Keratinization of vaginal vaginal epithelium. III. Effect of estradiol on keratinization. Biology of the cell / under the auspices of the European Cell Biology Organization. 65, 281-289, 1989 [PubMed: 2473820]

[33] Marcondes, FK, FJ Bianchi, & AP Tanno, Determination of the estrous cyclephase of rats: some helpful considerations. *Journal of Brazilian Archives of Biology and Technology*, 4A, 600-614, 2002.

[34] Shrestha J., Tara Shanbhag, Smita Shenoy, Arul Amuthan, Krishnananda Prabhu, Stuti Sharma, Samik Banerjee, and Sajala Kafle, Antiovulatory and abortifacient effects of Areca catechu (betel nut) in female rats, Indian J Pharmacol, 42, 5, 306-311, 2010. doi: 10.4103 / 0253-7613.70350

[35] Champbell, AN, JB Reece, & LG Mitchell. 5th edition of Biology. Volume 3. Bro. from Biology 5th ed., by Manalu, W. Erlangga, Jakarta: xxii, 2004, 476 p.

[36] Byers SL Michael V. Wiles, Sadie L. Dunn1, Robert A. Taft, Mouse Estrous Cycle Identification Tool and Images. Bar Harbor, Maine, United States of America. Volume 7, Issue 4, e35538, 2012. (PLoS ONE | www.plosone.org) doi: 10.1371 / journal.pone.0035538

[37] Sinchak K. and Edward J. Wagner, Estradiol Signaling in the Regulation of Reproduction and Energy Balance, Front Neuroendocrinol, 33, 4, 342-363, 2012. doi: 10.1016 / j.yfne.2012.08.004