Research Article

Anti-Inflammatory Effects of Modified Buyang Huanwu Decoction

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Background/Aims. Buyang Huanwu decoction (BHD) is a traditional Chinese and Korean herbal medicine prescription that has been widely used to treat various diseases including cerebral ischemia, gynecological disease, and neurological disorders. BHD is commonly used as a variable modified combination for synergistic therapeutic effects. However, the mechanism by which modified BHD (mBHD) produces anti-inflammatory effects has not been elucidated yet. The purpose of this study was to develop mBHD with diminished potential side effects and verify its anti-inflammatory effects. Methods. A cytotoxicity assay for BHD was performed using the MTT assay. Following treatment with BHD, mBHD-1, and mBHD-2 in the presence of lipopolysaccharide (LPS), nitric oxide (NO) secretion was detected in cell supernatants using a NO detection kit. The expression of proinflammatory mediators was detected using RT-PCR and western blotting. To verify the mechanism of mBHD, specific inhibitors of JNK (SP600125) or p38 (SB203580) were used for co-treatment with mBHD, and then the changes in NO and nitric oxide synthase (iNOS) were measured. Results. Both mBHD-1 and mBHD-2 showed greater anti-inflammatory effects than BHD. Both mBHD-1 and mBHD-2 inhibited NO secretion and decreased the expression of IL-1β, IL-6, TNF-α, and iNOS. Treatment with a p38 inhibitor and a JNK inhibitor in mBHD-1- and mBHD-2-treated cells resulted in inhibition of NO and iNOS. Conclusion. We provided the first experimental evidence that mBHD may be a more useful anti-inflammatory than BHD. High concentrations or long-term use of BHD may be harmful to inflammatory status. Therefore, the length of treatment and concentration should be considered depending on the targeted disease.

1. Introduction

The inflammatory process is considered a major driving force in the ischemic process, brain repair, neuropathic pain, neuronal injury, and gynecological disease [1]. During the inflammatory response, diverse chemical mediators released by activated immune cells have effects on excitability and synaptic function, and these changes can contribute to the initiation and maintenance of persistent neuropathic pain [2, 3]. Cerebral ischemia is accompanied by a marked inflammatory reaction that is initiated by ischemia-induced expression of cytokines, adhesion molecules, and other inflammatory mediators, including prostanoids and nitric oxide [4, 5]. Thus, the control of the inflammatory process is a logical therapeutic target.

Nitric oxide (NO) has bifunctional roles as a free radical biological messenger in inflammatory responses. It improves the blood supply to injured tissue and is secreted as free radicals that are toxic to bacteria but can cause tissue damage by initiating apoptosis and killing otherwise healthy cells [6]. In rodents, NO can both promote [2, 7] and inhibit inflammatory pain [3, 7]. NO is synthesized by nitric oxide synthase (iNOS), which is mainly upregulated in activated immune cells such as macrophages and glial cells. Therefore,
NO and iNOS are major molecules that are generated by macrophages and other phagocytes in inflammation [8].

Traditional Chinese and Korean medicine has been prescribed for health protection and disease control for thousands of years in Eastern Asia. The Buyang Huanwu decoction (BHD) is a traditional medicine mainly used to treat circulatory disorders such as cerebral ischemia, angina, and myocardial infarction, as well as multiple neuritis and thrombophlebitis. According to experimental reports, BHD improves blood circulation, controls pain, regenerates neuronal cells [9–11], and protects against the neuronal damage caused by ischemic and oxidative stress [12, 13].

BHD consists of seven kinds of herbs: Radix Astragali (Astragalus), Angelicae gigantis radix, Paeoniae radix rubra, Cnidii rhizoma, Persicae semen, Carthami flos, and Lumbricus, all of which are recorded in the Chinese Pharmacopoeia. These ingredients have effects mainly upon improving blood circulation and protecting thrombopoeia. These ingredients have effects mainly upon improving blood circulation and protecting thrombophlebitis. According to experimental reports, BHD improves blood circulation, controls pain, regenerates neuronal cells [9–11], and protects against the neuronal damage caused by ischemic and oxidative stress [12, 13].

Considering those reports, we developed two types of mBHD and verified their efficacy according to comparative studies of the anti-inflammatory effect. One was manufactured by eliminating Lumbricus (renamed to mBHD-1). *Lumbricus* is a genus of earthworm, and among the seven ingredients of BHD is the only animal material. Thus, we omitted this animal-derived ingredient to avoid the possibility of adverse effects.

Another formulation reduced the concentration of Astragalus (renamed to mBHD-2). Traditional BHD contains 120 g of Astragalus. Potentially adverse effects of Astragalus have been reported when used in high doses, despite the enhanced immune system, anti-inflammatory, and antioxidant effects. Astragalus can interact with prescription medications, including anti-hypertensives and immune suppressants and can decrease medication efficacies. It can cause bleeding when used with other anticoagulants, antithrombotic agents [16]. Some exporters have been concerned that Astragalus is a legume and thus can cause allergic reactions in people with legume allergies (such as beans and peanuts). For these reasons, we investigated the effects of BHD, mBHD-1, and mBHD-2 on NO secretion and inflammation-associated factors and the signaling pathways involved in the inflammatory responses mediated by lipopolysaccharide- (LPS-) exposed macrophages.

## 2. Materials and Methods

### 2.1. Chemicals and Reagents.

The composition of BHD and modified-BHD (named as mBHD-1 or mBHD-2) is described in Table 1. The mBHD-1 formulation omitted *Lumbricus* from the other seven herbs comprising BHD, and mBHD-2 reduced the amount of Astragali Radix used, compared with BHD. The mixture of dried ingredients was boiled in purified saline for 2 h and concentrated under vacuum pressure at 700 mm Hg for 15 h, after which it was freeze-dried. The lyophilized powder of BHD and mBHD were then stored at −20°C until use.

The LPS was purchased from Sigma-Aldrich (St. Louis, MO, USA). Primary antibodies against total extracellular signal-related kinase (ERK), phospho-ERK (p-ERK), total N-terminal kinase (JNK), phospho-JNK (p-JNK), p38 mitogen-activated protein kinase (p38-MAPK), phospho-p38 (p-p38), and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) were purchased from Cell Signaling Technology (Danvers, MA, USA). The antibodies against iNOS and Cox-2 were purchased from Abcam (Cambridge, MA, USA). Horseradish peroxidase- (HRP-) conjugated goat anti-rabbit IgG antibody was obtained from Santa Cruz Biotechnology (Santa Cruz, CA, USA).

### 2.2. Cell Line and Treatments.

The murine macrophage cell line RAW264.7 was acquired from the Korean Cell Line Bank (Seoul, Republic of Korea) and was cultured in high-glucose DMEM (Gibco BRL, Gaithersburg, MD, USA) supplemented with 10% fetal bovine serum (FBS, Gibco BRL) and maintained at 37°C in a humidified incubator with 5% CO₂. BHD, mBHD-1, and mBHD-2 were dissolved in phosphate-buffered saline and filtrated. RAW264.7 cells were pretreated with various concentrations of BHD, mBHD-1, and mBHD-2 or co-pretreated with the inhibitors p38 or JNK for 1 h and then were stimulated with LPS (100 ng/mL) for a period of 30 min to 24 h, depending on the experiment.

### 2.3. Cell Viability Assays.

The cytotoxic effect of BHD, mBHD-1, mBHD-2, and several inhibitors on RAW264.7 cells was examined using an EZ-cytox assay kit (Daeil Lab Serve Co, Seoul, Republic of Korea) according to the manufacturer’s instructions. Briefly, RAW264.7 cells (1 × 10⁵ cells per well) were seeded in 96-well plates and were treated with various concentrations of materials. After 24 h, 10 µL of EZ-cytox was added into each well and then incubated for another 2 h under standard culture conditions. Cell viability was quantified with a spectrophotometer at 450 nm.

### 2.4. Measurement of NO Production.

NO secreted from cells was detected using a NO detection kit (Intron Biotechnology, Seongnam, Republic of Korea). Briefly,

| Table 1: Raw materials of BHD, mBHD-1, and mBHD-2. |
|-----------------|-----------------|-----------------|
| BHD | mBHD-1 | mBHD-2 |
| Astragali Radix | 120 g | 120 g | 20 g |
| Angelicae gigantis radix | 6 g | 6 g | 6 g |
| Paeoniae radix | 4.5 g | 4.5 g | 4.5 g |
| Lumbricus | 3 g | — | 3 g |
| Cnidii rhizoma | 3 g | 3 g | 3 g |
| Persicae semen | 3 g | 3 g | 3 g |
| Carthami flos | 3 g | 3 g | 3 g |

BHD, Buyang Huanwu decoction; mBHD, modified Buyang Huanwu decoction.
RAW264.7 cells (1 × 10^5 per well) were pretreated with various concentrations of BHD, mBHD-1, and mBHD-2, or co-pretreated with SB203580 (a p38 inhibitor) or SP300125 (a JNK inhibitor) for 1 h and then stimulated with 100 ng/mL of LPS for 24 h. Then, 100 μL of conditioned media was mixed with an equal volume of Griess reagent and incubated at room temperature for 10 min. Absorbance values were measured at 560 nm using a plate reader. The nitrate concentration was calculated based on a nitrite standard curve.

2.5. Reverse Transcription-Polymerase Chain Reaction (RT-PCR). The cells (5 × 10^5 per well) were pretreated with various concentrations of BHD, mBHD-1, or mBHD-2 for 1 h and then stimulated with 100 ng/mL of LPS for 4 h. Total RNA was isolated using TRizol reagent (Invitrogen, Carlsbad, CA, USA). Total RNA was reverse-transcribed to synthesize cDNA using the GoScript™RT system (Promega, Madison, WI, USA). For first-strand synthesis, purified total RNA (1 μg) was incubated with oligo dT (0.5 μg/μL) for 5 min at 70°C, followed by addition of buffer containing 4 μL of 5x reaction buffer, 1 μL of dNTP (10 mM each), 3.5 μL of 25 mM MgCl2, 2 μL of RNase inhibitor (40 U/μL) (Promega), and 1 μL of reverse transcriptase (200 U/μL). The reaction volume was 20 μL, and the mixture was incubated for 1 h at 42°C. Synthesized cDNA (500 ng) was amplified using iSTAR Taq polymerase (Intron Biotechnology, Seongnam, Republic of Korea). The primer information is described in Table 2. The amplified DNA was detected on an agarose gel containing Eco dye™ (BIOFACT, Daejeon, Republic of Korea). L#_he signals were detected by enhanced chemiluminescence (ECL, Amersham Biosciences, Buckinghamshire, UK). L#_he data were expressed as mean ± standard error of the mean (SEM). L#_he statistical differences between the groups were analyzed by one-way ANOVA followed by Tukey’s multiple comparisons test using GraphPad Prism 7.0 (GraphPad Software Inc., CA, USA). A p value <0.05 was considered statistically significant.

2.6. Western Blot Analysis. RAW264.7 cells (1 × 10^5 per well) were pretreated with various concentrations of BHD, mBHD-1, or mBHD-2 or co-pretreated with SB203580 (a p38 inhibitor) or SP300125 (a JNK inhibitor) for 1 h and then stimulated with 100 ng/mL of LPS for 30 min or 24 h, respectively. For preparation of total protein, cells were lysed in RIPA buffer (50 mM Tris (pH 8.0), 5 mM EDTA, 150 mM sodium chloride, 0.5% deoxycholic acid, 0.1% SDS, 1% NP-40, 1 mM PMSF, and 1 mg/mL protease inhibitor cocktail). The protein concentrations were determined using a Bio-Rad Protein Assay Kit (Bio-Rad, Hercules, CA, USA). Total protein (10 μg) was separated by 8–10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to PVDF membranes (Pall Corp., Hauppauge, NY, USA). The membranes were incubated for 1 h at room temperature with 5% nonfat dry milk, probed overnight at 4°C with each of the primary antibodies, and incubated with horseradish peroxidase-labeled goat anti-rabbit antibody. The signals were detected by enhanced chemiluminescence (ECL, Amersham Biosciences, Buckinghamshire, UK). The data correspond to three independent experiments.

2.7. Statistical Analysis. Data were expressed as mean ± standard error of the mean (SEM). The statistical differences between the groups were analyzed by one-way ANOVA followed by Tukey’s multiple comparisons test using GraphPad Prism 7.0 (GraphPad Software Inc., CA, USA). A p value <0.05 was considered statistically significant.

### Table 2: Primers used in this study.

| Target         | Primer sequence                  |
|----------------|----------------------------------|
| GAPDH          | F. caggatggagcatgtgacac          |
|                | R. ctctgcagactcaatcctca          |
| IL-1β          | F. caggatggagcatgtgacac          |
|                | R. ctctgcagactcaatcctca          |
| IL-6           | F. ggaattagggttgagggaga          |
|                | R. cggagagagactcagcagc           |
| TNF-α          | F. cagccttgctccctgaagagaga       |
|                | R. tactgaactctggttggatgtggc      |

GAPDH, glyceraldehyde 3-phosphate dehydrogenase; IL, interleukin; TNF, tumor necrosis factor.
Figure 1: Cytotoxic effects of BHD, mBHD-1, and mBHD-2 on murine macrophage RAW264.7 cells. Cells were treated with 10, 100, 1,000, 2,500, or 5,000 μg/mL of BHD (a), mBHD-1 (b), or mBHD-2 (c), and their cytotoxicity was detected using the MTT assay. Statistical differences were analyzed using one-way ANOVA followed by Tukey’s multiple comparisons test. Error bars indicate SEM. NS, not significant.

Figure 2: Effects of BHD, mBHD-1, and mBHD-2 on NO production in LPS-exposed RAW264.7 cells. Cells were pretreated with different concentrations of BHD (a), mBHD-1 (b), and mBHD-2 (c) for 1 h and then were stimulated with 100 ng/mL LPS. After 24 h, secreted NO was measured in conditioned media using a NO detection kit. *** p < 0.001 vs. LPS. Statistical differences were analyzed using one-way ANOVA followed by Tukey’s multiple comparisons test. Error bars indicate SEM.
proinflammatory cytokines in LPS-exposed RAW264.7 cells. As shown in Figure 4(a), cells pretreated with BHD showed decreased expression of interleukin- (IL-) \(\beta\) at 2.5 mg/mL \((p < 0.05)\), and IL-6 at 1, 2.5, and 5 mg/mL \((p < 0.001)\) compared with that of cells stimulated only with LPS. The cells pretreated with mBHD-1 showed decreased expression of IL-1\(\beta\) at 2.5 \((p < 0.05)\) and 5 mg/mL \((p < 0.01)\) and IL-6 at 1, 2.5, and 5 mg/mL \((p < 0.01)\) compared to the LPS-treated cells. On the other hand, the cells pretreated with mBHD-2 showed decreased expression of IL-1\(\beta\) at 2.5 and 5 mg/mL \((p < 0.01)\) and IL-6 at 1 \((p < 0.05)\), 2.5, and 5 mg/mL \((p < 0.001)\) compared to the LPS-treated cells. mBHD-2 tended to decrease expression of tissue necrosis factor- (TNF-) \(\alpha\) at 5 mg/mL in a dose-dependent manner, but there was no statistical significance.

Cyclooxygenase- (Cox-) 2 and inducible iNOS are induced by proinflammatory cytokines in an inflammatory environment [17]. We examined the expression of Cox-2 and iNOS using western blotting. Compared to the LPS-treated cells, the expression of iNOS was significantly decreased in the cells pretreated with mBHD-1 and mBHD-2 at every dose \((p < 0.001)\), while it was highly induced in the BHD-pretreated cells \((p < 0.001)\). The expression of Cox-2 was increased in the BHD-pretreated cells at 1 \((p < 0.05)\) and 5 mg/mL \((p < 0.001)\) and decreased in the mBHD-2-pretreated cells at 2.5 mg/mL \((p < 0.01)\) compared to LPS-treated cells (Figure 4(b)).

3.5. Effect of mBHD-1 and mBHD-2 on Activation of p38, JNK, and ERK1/2 Pathways in LPS-Exposed RAW264.7 Cells

p38, JNK, and ERK play an important role in inflammatory responses such as iNOS expression. To elucidate the molecular mechanism of the anti-inflammatory effects of mBHD-1 and mBHD-2, we examined the activities of ERK, p38, and JNK using western blotting. The activation of p38 and JNK was decreased in mBHD-1 or mBHD-2 pretreated cells, compared with LPS-stimulated cells. However, mBHD-1 and mBHD-2 did not affect the activation of the ERK pathway in LPS-stimulated cells (Figure 5).

To confirm whether mBHD-1 or mBHD-2 regulate NO and iNOS through stimulate the JNK and p38 signaling, we co-treated cells with a JNK inhibitor (SP600125) or a p38 inhibitor (SB203580) with mBHDs. After treatment of LPS-stimulated cells with SP600125 alone, NO was reduced by approximately 20%. When cells were co-treated with LPS and mBHD-1 (or mBHD-2), NO increased by approximately 15% compared with that in cells treated with...
**Figure 4:** Effect of BHD, mBHD-1, and mBHD-2 on the expression of proinflammatory cytokines in LPS-exposed RAW264.7 cells. Cells were pretreated with 1, 2.5, or 5 mg/mL of each drug for 1 h and then were stimulated with LPS for 4 h. Transcriptional levels of inflammatory cytokines of IL-1β, IL-6, and TNF-α were detected using RT-PCR (a) and the protein expression of iNOS and Cox-2 was detected using western blotting (b). IL: interleukin; TNF: tumor necrosis factor; iNOS: inducible nitric oxide; Cox-2: cyclooxygenase-2; GAPDH: glyceraldehyde 3-phosphate dehydrogenase; LPS: lipopolysaccharide. C, control; *p < 0.05; **p < 0.01; ***p < 0.001 vs. control; #p < 0.05; ##p < 0.01; ###p < 0.001 vs. LPS. Statistical differences were analyzed using one-way ANOVA followed by Tukey’s multiple comparisons test. Error bars indicate SEM.
SP600125 alone ($p < 0.001$). In cells treated with SB203580 alone, NO was reduced by about 40%, and the NO reduction between the SB203580 treatment alone and the SB203580+mBHD-1 (or mBHD-2) treatment showed similar results, compared with that in cells treated with LPS alone (Figure 6(a)). The expression of iNOS was significantly decreased in both SP600125- and SB203580-treated cells, compared with that of the control group (Figure 6(b)). These results indicated that mBHD-1 and mBHD-2 regulate NO and iNOS expression through the p38 MAPK and JNK pathway (Figure 7).

### 4. Discussion

Traditional BHD consists of seven natural herbal components that function to improve the circulatory system and immunity in humans [18–20] and protect neural activity and regeneration [21, 22]. Thus, this traditional medicine has been prescribed mainly to patients with autoimmune encephalomyelitis [23] or ischemic brain disorders [12, 24] for more than 300 years. Recent studies on the use of various mBHD, which have reduced BHD components or are combined with other effective herbs, have provided reports regarding their therapeutic advantage. In a rat model of Alzheimer’s disease, mBHD with a reduced quantity of all components improved memory deficiency and decreased the cerebral infarction in the hippocampus [25, 26]. Another version of mBHD with added Polygalae radix and Acori Graminei Rhizoma showed anti-inflammatory and antioxidant effects on microglial cells [20]. Two studies using modified BHD with Achyranthis Radix, Acori Graminei Rhizoma, Polygalae Radix, Cinnamomi Ramulus, and Salviae Miltiorrhizae Radix reported reduction of the cerebral infarction size and neurological deficit in ischemia/reperfusion models [14] and hippocampal inflammatory responses in vascular dementia models [15].

A BHD preparation including Lycopi Herba, Typae pollen, Trogopterori feces, and Corydalis tubers showed antithrombotic activities and can be applied to diverse female diseases caused by thrombosis and inflammation such as endometriosis, myoma, and chronic pelvic inflammatory disease [27]. According to a clinical report in 2011, BHD without lumbricus improved symptoms in patients with hyperhidrosis resulting from sequel of spinal cord injuries. Tritic Fructus Levii is utilized mainly for inhibiting spontaneous perspiration and night sweats by securing the exterior.

Astragalus not only has various therapeutic effects, including enhanced immune system function as well as anti-inflammatory and antioxidant effects, but also poses potential risk when used at a high dose [28–31]. Lumbricus is an earthworm genus considered to possess therapeutic effects in epilepsy and stroke [32]. Until now, the side effects of Lumbricus have not been reported, but out of the seven ingredients composing BHD, this represents the only animal-derived component, whereas the other components of BHD are plant-derived.

Based on these reports, we developed two types of mBHD, the mBHD-1 preparation (without Lumbricus) and the mBHD-2 preparation (reduced amount of Astragalus) and verified the improvement of anti-inflammatory effects. We compared the anti-inflammatory effects of BHD and mBHDs in LPS-stimulated murine macrophage RAW264.7 cells. The activity of the iNOS and NO system is mediated by macrophages and other phagocytes during inflammatory episodes [8]. In immune-defense status, iNOS is activated by proinflammatory cytokines such as IL-1β,
Figure 6: Both mBHD-1 and mBHD-2 inhibit NO secretion by regulating the p38 signaling pathway in LPS-exposed RAW264.7 cells. Cells were pretreated with SP600125 (a JNK inhibitor) or SB203580 (a p38 inhibitor) with or without BHD, mBHD-1, or mBHD-2 for 1h and then were stimulated with LPS (100 ng/mL). After 24h, NO was detected in conditioned media using a NO detection kit (a), and expression of iNOS and Cox-2 was measured in total protein using western blotting (b). LPS + SP: LPS and SP600125 treatment; LPS + SB: LPS and SB203580 treatment. **p < 0.01; ***p < 0.001 vs. LPS; ###p < 0.001 vs. LPS + SP. Statistical differences were analyzed using one-way ANOVA followed by Tukey’s multiple comparisons test. Error bars indicate SEM.

Figure 7: Summary of anti-inflammatory effects of mBHD-1 and 2.
TNF-α, IFN-γ, and LPS, resulting in large amounts of NO release.

On the basis of those reports, we examined the effect of BHD, mBHD-1, and mBHD-2 on changes in NO secretion and expression of iNOS and proinflammatory cytokines in LPS-stimulated RAW264.7 cells. According to our results, the cells treated with mBHD-1 and mBHD-2 showed significantly and dose-dependently decreased NO secretion and expression of IL1-β, TNF-α, Cox-2, and iNOS. The results of mBHD-1 and mBHD-2 treatments showed similar results. However, BHD-treated cells showed opposite results. BHD increased NO secretion and the expression of these factors. As mentioned above, BHD is composed of ingredients that are useful in blood circulation.

NO has bifunctional roles as a free radical biological messenger and a modulator of inflammatory responses; these roles can be separated as defensive mechanisms and tissue damage mechanisms. Furthermore, NO improve the blood supply to injured tissue and is secreted as a free radical that is toxic to bacteria and modulates the actions of macrophage-derived cytokines on target cells. However, NO can cause tissue damage by inducing apoptosis [33]. In an oxidative environment, high levels of NO released by iNOS have the opportunity to react with superoxide, leading to peroxynitrite formation and cell toxicity. These properties may define the roles of iNOS in host immunity, enabling its participation in antimicrobial and antitumor activities as part of the oxidative burst produced by macrophages [34]. Proinflammatory cytokines (TNF-α, IL-6, and IL-1β) are important mediators in the inflammatory process and are mainly produced by activated macrophages [35]. Excessive production of these proinflammatory cytokines will lead to clinical problems associated with autoimmune disorders (rheumatoid arthritis) and inflammatory response syndrome (septic shock) [36].

To further investigate the mechanism of the anti-inflammatory effects of mBHD, we examined major signaling molecules associated with the inflammatory process. LPS-stimulated macrophages have been widely reported to activate three different groups of MAP kinases: ERK, JNK, and p38 [37]. Our results showed that BHD, mBHD-1, and mBHD-2 significantly reduced p38 and JNK activated by LPS but did not affect ERK1/2 phosphorylation. Activation of p38 and JNK inhibited by mBHD-1 and mBHD-2 was stably maintained 24 h later but that inhibited by BHD was recovered in 24 h.

Finally, we confirmed whether mBHD-1 and mBHD-2 control NO secretion and iNOS expression through the regulation of the p38 and JNK pathway using the inhibitors SB203580 for p38 and SP600125 for JNK in LPS-induced RAW264.7 cells. SB203580 is a selective inhibitor of p38 [38], and SP600125 is a potent, selective, and reversible inhibitor of JNK [39]. As our results demonstrate, the cells co-treated with mBHD and SB203580 showed significantly reduced NO and iNOS expression in the presence of LPS, compared with those exposed to LPS alone. The inhibitory effects on NO secretion produced by SB203580 were higher than those produced by SP600125.

5. Conclusion

In conclusion, mBHD-1 and mBHD-2 were more effective in suppressing the NO/iNOS system following activation by LPS through the regulation of the p38 and JNK pathway in LPS-exposed RAW264.7 cells. Both mBHD-1 and mBHD-2 showed greater anti-inflammatory activity than BHD. These results may indicate that the prescribed concentration of Astragalus in BHD should be determined depending on the target disease. This study provided the first experimental evidence of an anti-inflammatory effect of mBHD. However, further studies are still needed to elucidate the fundamental mechanism of mBHD in various types of in vitro and in vivo disease models. Moreover, further research should continue to develop various combinations of BHD and develop specific prescriptions for each targeted disease.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflicts of interest.

Authors’ Contributions

HR performed in vitro experiment and data analysis and drafted the manuscript. JK performed in vitro experiment. UN contributed to the conception of the work and assisted with protocol development. SK performed data analysis and writing of the article. JP designed the study, coordinated the acquisition of all study data, and contributed to the interpretation of results and writing of the article. All authors were involved in critical revision of initial drafts.

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