mild and the patient responded well to antimicrobial drug therapy, albeit without catheter removal. This case emphasizes that environmental bacteria can be an emerging threat for hemodialysis patients, who are at risk of acquiring opportunistic infection. In addition, this report demonstrates the usefulness of molecular methods for identifying uncommon isolates.

Acknowledgment

We thank Patrick Murray for helpful comments on this article.

Pattarachai Kiratisin,*
Premwadee Kowwigkai,*
Supanit Pattanachaiwit,*
Anucha Apisarnthanarak,† and Amornrut Leelaporn*

*Mahidol University, Bangkok, Thailand; and †Thammasat University Hospital, Prathumthani, Thailand

References

1. Taylor G, Gravel D, Johnston L, Embil J, Holton D, Paton S, et al. Incidence of bloodstream infection in multicenter infection cohorts of hemodialysis patients. Am J Infect Control. 2004;32:155–60.
2. Tokars JI, Miller ER, Stein G. New national surveillance system for hemodialysis-associated infections: initial results. Am J Infect Control. 2002;30:288–95.
3. Maraj S, Jacobs LE, Maraj R, Kotler MN. Bacteremia and infective endocarditis in patients on hemodialysis. Am J Med Sci. 2004;327:242–9.
4. Bambauer R, Schauer M, Jung WK, Vienken J, Daum V. Contamination of dialysis water and dialysate: a survey of 30 centers. ASAIO J. 1994;40:1012–6.
5. Kiratisin P, Li L, Murray PR, Fischer SH. Use of 16S rRNA gene sequencing to identify uncommon bacteria in a clinical laboratory. Eur J Clin Microbiol Infect Dis. 2003;22:628–31.
6. Xie C-H, Yokota A. Dyella japonica gen. nov., sp. nov., a β-proteobacterium isolated from soil. Int J Syst Evol Microbiol. 2005;55:753–6.
7. Stackebrandt E, Goebel BM. Taxonomic note: a place for DNA-DNA reassociation and 16S rRNA sequence analysis in the present species definition in bacteriology. Int J Syst Bacteriol. 1994;44:846–9.
8. An DS, Im WT, Yang HC, Yang DC, Lee ST. Dyella koreensis sp. nov., a β-glucosidase-producing bacterium. Int J Syst Evol Microbiol. 2005;55:1625–8.
9. Kim BY, Weon HY, Lee KH, Seok SJ, Kwon SW, Go SJ, et al. Dyella yeojunensis sp. nov., isolated from greenhouse soil in Korea. Int J Syst Evol Microbiol. 2006;56:2079–82.

Address for correspondence: Amornrut Leelaporn, Department of Microbiology, Faculty of Medicine, Siriraj Hospital, Mahidol University, 2 Prannok Rd, Bangkoknoi, Bangkok 10700, Thailand; email: siale@mucc.mahidol.ac.th

Mycobacterium cosmeticum, Ohio and Venezuela

To the Editor: Mycobacterium cosmeticum is a rapidly growing nontuberculous mycobacteria species that was first described in November 2004. The first strains were obtained from cultures of a sink drain in a nail salon in Atlanta, Georgia, USA, and from a granulomatous lesion of a female mesothery patient in Venezuela (1).

Among 3 additional isolates of M. cosmeticum obtained from July 2003 through November 2004, one was obtained from a 77-year-old man who was admitted to Ohio hospital A on September 22, 2004, with fever, exacerbation of chronic obstructive pulmonary disease, and urosepsis. Underlying medical conditions included diabetes, discitis, hyperlipidemia, coronary artery disease, and coal worker’s pneumoconiosis. He had received intravenous antimicrobial agents (rifampin and daptomycin) through a Groshong catheter that had been inserted to treat discitis. A routine blood culture was performed according to standard methods (2), and the catheter was removed. A diagnosis of catheter-associated bacteremia (CAB) was made, but the patient’s overall condition improved without antibacterial drug therapy, and he was discharged 4 days after admission. The culture specimen yielded only mycobacteria and was sent on to ARUP Laboratories, where it was identified as M. cosmeticum by 16S rDNA sequence analysis. The isolate was then sent to the Centers for Disease Control and Prevention (CDC) Mycobacteriology Laboratory Branch (Atlanta, GA, USA) and designated OH1.

A 43-year-old woman with a diagnosis of non-Hodgkin lymphoma, who had received regular central venous catheterizations, was admitted to Ohio hospital B on August 20, 2004. A left subclavian catheter was inserted, and

---

Table. Biochemical characteristics of patient’s isolate RB28 and type strains of Dyella species*

| Characteristics | Patient’s isolate RB28 | D. japonica XD53 | D. koreensis BB4 |
|-----------------|------------------------|------------------|-----------------|
| Oxidase         | +                      | +                | +               |
| Catalase        | +                      | +                | w               |
| Motility        | +                      | +                | +               |
| Acid from       |                        |                  |                 |
| L-arabinose     | –                      | –                | –               |
| D-galactose     | –                      | –                | –               |
| D-glucose       | +                      | +                | +               |
| D-mannose       | +                      | +                | –               |
| D-ribose        | –                      | –                | +               |
| D-sucrose       | –                      | –                | –               |
| D-xylene        | –                      | –                | –               |
| Caprate         | –                      | –                | –               |
| Citrate         | –                      | –                | –               |
| α-galactosidase | –                      | –                | +               |
| β-N-acetyl-glucosaminidase | – | w | + |
| α-glucosidase   | –                      | w                | +               |

*Data from references (6) and (8); T, type strain.
A routine blood specimen for culture was subsequently obtained on the day of admission. Before admission, the woman had been receiving acyclovir and cefepime. She received chemotherapeutic agent injections, platelet infusion, and an autologous stem cell transplant 6 days after admission. The blood culture was positive only for rapidly growing mycobacteria, and the final diagnosis was CAB. However, no symptoms of infection were observed, and no antimycobacterial drug therapy was administered. She was discharged without complications after the transplant was received and the catheter removed. The bacterial isolate was forwarded to CDC’s Special Bacteriology Reference Laboratory, where it was identified as *M. cosmeticum* by 16S rDNA sequence analysis, sent on to the CDC Mycobacteriology Laboratory Branch, and designated OH2.

A 36-year-old man with AIDS was admitted to hospital C in Caracas, Venezuela, in June 2003 with dyspnea, fever, and expectoration. A sputum sample was positive by acid-fast bacillus smear and culture, yielding both *M. cosmeticum* (designated VZ1) and *M. scrofulaceum* on Middlebrook 7H10 agar (Remel Co., Lenexa, KS, USA). At the time the sputum was obtained, the patient was receiving only trimethoprim-sulfamethoxazole, but he experienced respiratory arrest and died ≈6 weeks later.

The 3 isolates were confirmed to be *M. cosmeticum* by high-performance liquid chromatography mycolate analyses and by PCR restriction analysis of a 440-bp segment of *hsp65* (1). The relationship of these isolates to the only documented strains of *M. cosmeticum* was evaluated by analysis of large restriction fragments with pulsed-field gel electrophoresis (1) and by repetitive element PCR (3). Banding patterns for isolates OH1 and OH2 were different from one another as well as from isolate VZ1 and the 2 control strains. Typing patterns for isolate VZ1, however, matched the control strain from Venezuela (ATCC BAA-878), which indicates that these 2 isolates are likely a common strain (Figure).

Of the >125 recognized *Mycobacterium* species, ≈50 are etiologic agents of human disease (4). The type strain of *M. cosmeticum* (ATCC BAA-878) was associated with a soft-tissue infection in which the source was postulated to be environmental contamination of an unknown substance administered to the patient by injection as part of a weight loss regimen. This strain and isolate VZ1 were isolated from clinics in Caracas, Venezuela; both were found to be a common strain, but no other factors suggest that these represent an epidemic cluster. Although the Venezuelan patient from whom isolate VZ1 was obtained exhibited symptoms consistent with mycobacterial pulmonary disease, *M. cosmeticum* involvement cannot be proven because an additional *Mycobacterium* species, *M. scrofulaceum*, was isolated from the patient’s sputum. Because each of these organisms is found in the aqueous environment, they may represent colonization or may have been transiently present in the patient. Nonetheless, additional nontuberculous mycobacteria species have been reported to cause pulmonary disease, and the involvement of *M. cosmeticum* in this case cannot be excluded.

Successful treatment of CAB infections caused by rapidly growing mycobacteria has most often been achieved by removing the catheter with or without the use of antimicrobial drug therapy (4). Criteria to support a true bloodstream infection were met by one of the patients in Ohio. These criteria include the absence of a source for bacteremia alternative to *M. cosmeticum* OH1 and the resolution of the febrile syndrome after removal of the device. The second patient had no symptoms when the blood culture was obtained; thus, the clinical significance of *M. cosmeticum* in this case is unclear.
When all identified strains of \( M. \) cosmecutum are considered, this species is clearly present in diverse geographic regions and in healthcare institutions. These findings suggest that it may be widely distributed in the environment and should be regarded, along with other rapidly growing mycobacteria species, as a potential pathogen.

Acknowledgment

We thank Lena Fischer of the Ohio Department of Health for technical contributions.

Robert C. Cooksey,* Jacobus H. de Waard,† Mitchell A. Yakrus,* Sean R. Toney,* Omaira Da Mata,‡ Scott Nowicki,‡ Kevin Sohner,‡ Elizabeth Koch,‡ Cathy A. Petti,§ Roger E. Morey,* and Arjun Srinivasan*

*Centers for Disease Control and Prevention, Atlanta, Georgia, USA; †Instituto de Biomedicina, Caracas, Venezuela; ‡Ohio Department of Health, Columbus, Ohio, USA; and §ARUP Laboratories, Salt Lake City, Utah, USA

References

1. Cooksey RC, de Waard JH, Yakrus MA, Rivera I, Chopite M, Toney SR, et al. Mycobacterium cosmecutum sp. nov., a novel rapidly growing species isolated from a cosmetic infection and from a nail salon. Int J Syst Evol Microbiol. 2004;54:2385–91.

2. Pfiffer GE, Brown-Elliott BA, Wallace RJ Jr. Mycobacterium: general characteristics, isolation, and staining procedures. In: Murray PR, Baron EJ, Pfiffer MA, Jorgensen JH, Yolken RH, editors. Manual of clinical microbiology. Washington: ASM Press; 2003. p. 532–59.

3. Dombek PE, Johnson LK, Zimmerley ST, Sadowsky MJ. Use of repetitive DNA sequences and the PCR to differentiate \( E. coli \) isolates from human and animal sources. Appl Environ Microbiol. 2000;66:2572–7.

4. Wagner D, Young LS. Nontuberculous mycobacterial infections: a clinical review. Infection. 2004;32:257–70.

Ecoregional Dominance in Spatial Distribution of Avian Influenza (H5N1) Outbreaks

To the Editor: Recent articles in Emerging Infectious Diseases (1,2) and elsewhere (3,4) have highlighted the role of Anatidae migration in dispersal of the H5N1 subtype of highly pathogenic avian influenza (HPAI) virus. Although these articles point out that identifying the geographic origin of migrating waterfowl is needed to understand and predict pathogen dispersal, study analyses have been limited to pathways with nominal reference to climatic and vegetation patterns that control spatiotemporal patterns of this migration.

We propose that a better understanding of the threat of future spread can be obtained by identifying specific climatic and vegetation zones that are important in the life cycle of Anatidae, and which account for a disproportionately large number of HPAI outbreaks. The concept of ecoregions (5,6), i.e., distinct assemblages of natural communities determined by climate, geology, and evolution, is a useful zonal classification for evaluating HPAI outbreaks. A World Wildlife Fund classification delineating 825 terrestrial ecoregions (7), combined with a Google Earth map of 3,133 avian influenza outbreaks from November 24, 2003, to November 21, 2006 (8), provided the basis for this analysis.

All files were converted to shapefiles (Environmental Systems Research Institute, Redlands, CA, USA), and overlay analysis was performed by using ArcGIS software (Environmental Systems Research Institute).

The online Appendix Figure (available from www.cdc.gov/EID/content/13/8/1269-appG.htm) shows a choropleth map (display of quantitative or qualitative information about subentities in terms of symbols or colors) of ecoregions with numbers of avian influenza cases (each spatially and temporally isolated set of individual events, regardless of number of deaths, is recorded as a case). Panels A, B, and C of this figure show enlargements of specific ecoregions with large numbers of known cases in regions of Eurasia, Southeast Asia, and Africa, respectively. Twenty-five ecoregions, representing 8.8% of the terrestrial surface area, accounted for 2,407 (76.8%) cases. A total of 132 of 825 ecoregional classifications had ≥1 recorded case of an avian influenza outbreak, but most (83) had <10 cases each.

Regionally, Southeast Asia has 12 ecoregions that collectively account for 1,651 cases (online Appendix Figure, panel B) that have occurred consistently, albeit cyclically, since 2003. Among these ecoregions, the freshwater wetlands of the Chao Phraya, Tonle Sap, and Red Rivers are known migratory waterfowl wintering habitats in which 719 cases were located. Recent phylogenetic evidence suggests that this area is a local hotspot for an endemic strain of avian influenza (H5N1) that demonstrates bidirectional dispersal among localities within the region (9).

In the Eurasian region (online Appendix Figure, panel A), 12 ecoregions accounted for 712 cases. The easternmost ecoregions, the Kazakh forest steppe (location of Lake Chany, an Anatidae habitat and breeding area) and the Kazakh Steppe, accounted for 132 cases, with the first case recorded