A note on noise suppression in cell-size control

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Abstract

Diverse cell types employ mechanisms to maintain size homeostasis and minimize aberrant fluctuations in cell size. It is well known that exponential cellular growth can drive unbounded intercellular variations in cell size, if the timing of cell division is size independent. Hence coupling of division timing to size is an essential feature of size control. We formulate a stochastic model, where exponential cellular growth is coupled with random cell division events, and the rate at which division events occur increases as a power function of cell size. Interestingly, in spite of nonlinearities in the stochastic dynamical model, statistical moments of the newborn cell size can be determined in closed form, providing fundamental limits to suppression of size fluctuations. In particular, formulas reveal that the magnitude of fluctuations in the newborn size is determined by the inverse of the size exponent in the division rate, and this relationship is independent of other model parameters, such as the growth rate. We further expand these results to consider randomness in the partitioning of mother cell size among daughters at the time of division. The sensitivity of newborn size fluctuations to partitioning noise is found to monotonically decrease, and approach a non-zero value, with increasing size exponent in the division rate. Finally, we discuss how our analytical results provide limits on noise control in commonly used models for cell size regulation.
Introduction

How cells regulate the timing of division to ensure size homeostasis and avoid getting abnormally large (or small) remains as one of the unsolved problems in biology [1]. Perhaps the simplest model of cell size regulation is one where the size of an individual cell grows exponentially over time, random cell division events occur at discrete times that partition size between two or more daughters cells. If the division timing is governed by a size-independent stochastic process (such as, a timer measuring the time elapsed since the last division event), then the statistical fluctuations in cell size grow unboundedly over time [2]. As a consequence, size homeostasis requires division timing to be regulated via a size-dependent process. Not surprisingly, uncovering mechanisms mediating size sensing and control remains a vigorous area of theoretical and experimental research in diverse organisms ranging from prokaryotes to microbial eukaryotes to plant and animal cells [3–27].

We formulate a simple stochastic model for size control, where the size grows exponentially over time throughout the cell cycle. Size dynamics is interspersed with random division events that divide size by approximately half (assuming two daughters). A key feature of the model is that the probability of division event increases as a power function of size. While such nonlinear stochastic dynamical systems are typically analytically intractable, we show that moments of the newborn size (i.e., size just after a division event) can be quantified in closed form. These results provide novel insights into how statistical variations in the newborn size are controlled by the division rate, and the randomness in size partitioning between daughters. Finally, we show how combining our analytical results with size distribution measurements allows inference of division rates.

Stochastic formulation of cell-size control

Let $v(t)$ denote the size of an individual cell at time $t$. Depending on the cell type, size can be quantified via different metrics, such as, cell length in rod-shaped bacteria, or cell volume/mass in animal cells. The size grows exponentially over time as

$$\frac{dv}{dt} = \alpha v,$$

where $\alpha > 0$ is referred to as the exponential growth coefficient. Cell division events occur stochastically with a division rate $f(v, \tau)$, where $\tau$ is a timer (cell-cycle clock) measuring the time elapsed from when the cell was born. More specifically, the probability of cell division occurring in the next
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Infinitesimal time interval \((t, t + dt]\) is given by \(f(v, \tau)dt\). Consistent with experimental observations \([5]\), the division rate is a non-decreasing function in both arguments, i.e., the likelihood of division increases with size and cell-cycle progression. Whenever a cell-division event occurs, the states are reset as

\[ v \rightarrow \frac{v}{2}, \quad \tau \rightarrow 0, \quad (2) \]

assuming that the mother cell divides into two identically-sized daughters.

It is well known that a size-independent division rate \((f \text{ only depends on timer } \tau)\) that yields a finite non-zero average cell size, leads to unbounded intercellular size variations \([2, 28]\), i.e.,

\[ \lim_{t \to \infty} \langle v^2 \rangle \to \infty, \quad (3) \]

where the symbol \(\langle \rangle\) is used to denote the expected value of a random variable. Hence, a size-dependent division rate is an essential feature for size homeostasis. We particularly restrict ourselves to a special class of division rates that take the form of power functions

\[ f(v, \tau) = \left(\frac{v}{\bar{v}}\right)^m, \quad (4) \]

where \(\bar{v}\) and \(m\) are positive constants. In the limit \(m \to \infty\), (4) corresponds to the well-known sizer paradigm, where the mother cell divides upon reaching a critical size threshold \(\bar{v}\) \([29–34]\). Given that sizer implementation is never perfect \((m\ is \ always \ finite)\), how does the extent of random fluctuation in cell size scale with the exponent \(m\)? Does this scaling depend on the exponential growth coefficient \(\alpha\) and \(\bar{v}\)? How does the scaling change if the two daughters are not identically-sized?

One approach to address these questions is to compute the statistical moments of \(v\), and investigate them as a function of model parameters. For the stochastic dynamical system (1)-(4), the following set of differential equations describes the time evolution of moments

\[
\frac{d\langle v^l \rangle}{dt} = l \alpha \langle v^l \rangle - \frac{\langle v^{l+m} \rangle}{\bar{v}^m} \left(1 - \frac{1}{2^l}\right), \quad l \in \{1, 2, \ldots\} \quad (5)
\]

\([35, 36]\). Note that for any \(m > 0\), (5) suffers from the problem of un-closed moment dynamics: the time evolution of any lower order moment depends on higher order moments. Hence, moments cannot be computed from (5), and approximate closure schemes are typically employed to solve such systems \([35, 37–48]\).
Statistical moments of newborn cell size

Note that considerable cell-to-cell variations in $v$ can simply be attributed to the fact that different cells are in different stages of their cell cycle. Taking this into account, perhaps a more meaningful quantity is the newborn cell size ($v$ conditioned on $\tau = 0$). We reformulate the size control model presented in the previous section into a discrete-time stochastic system that tracks the newborn size across generations.

Consider a single cell undergoing cycles of growth and division. Let random variable $v_i$ denote the newborn size during the start of the $i^{th}$ cell cycle. Then, given that size increases exponentially over time, and the division rate $f(v, \tau)$, the time $T_i$ from the start of the $i^{th}$ cell cycle when division is triggered follows the probability density function (pdf)

$$\text{Probability}\{T_i \in (\tau, \tau + d\tau)\} = f(v_i e^{\alpha \tau}, \tau) e^{-\int_{y=0}^{\tau} f(v_i e^{\alpha y}, y) dy} d\tau \quad (6)$$

Note that a constant function $f$ in (46) would correspond to exponentially distributed $T_i$. In our case, $f$ takes the following time-varying form based on (4)

$$f(v_i e^{\alpha \tau}, \tau) = \left(\frac{v_i}{\bar{v}}\right)^m e^{\alpha \tau} \quad (7)$$

that results in $T_i$ with lower statistical variations than an exponential random variable (i.e., memory in division timing), and dependent on $v_i$ (i.e., larger newborns divide earlier as compared to smaller newborns). Having determined the time to division, the newborn size in the next cycle is

$$v_{i+1} = \frac{v_i e^{\alpha T_i}}{2}, \quad \tau \in \{1, 2, \ldots\}. \quad (8)$$

This discrete-time stochastic system yields the following $l^{th}$ order moment of $v_{i+1}$ conditioned on $v_i$

$$2^l \left<v_{i+1}^l|v_i\right> = \int_{\tau=0}^{\infty} v_i^l e^{\alpha \tau} f(v_i e^{\alpha \tau}, \tau) e^{-\int_{y=0}^{\tau} f(v_i e^{\alpha y}, y) dy} d\tau \quad (9)$$

$$= v_i^l + l\alpha \int_{\tau=0}^{\infty} v_i^l e^{\alpha \tau} e^{-\int_{y=0}^{\tau} f(v_i e^{\alpha y}, y) dy} d\tau, \quad (10)$$

where the first integral is simplified via integration by parts. Next, we show how this recurrence equation can be solved in moments space.
Linear dependence of division rate on cell size

We first restrict ourselves to \( m = 1 \) in the division rate (4), which corresponds to the probability of division-event occurrence increasing linearly with size. In this case, using

\[
f(v_i e^{\alpha \tau}, \tau) = \frac{v_i}{\bar{v}} e^{\alpha \tau},
\]

\[(11)\]

\[(10)\] can be written as

\[
2^l \langle v_{l+1}^l | v_i \rangle = v_i^l + \bar{v} \alpha \int_{\tau=0}^{\infty} v_i^{l-1} e^{(l-1) \alpha \tau} f(v_i e^{\alpha y}, y) e^{-\int_{y=0}^{\tau} f(v_i e^{\alpha y}, y) dy} d\tau,
\]

\[
= v_i^l + 2^{l-1} \bar{v} \alpha \left( v_i^{l-1} | v_i \right), \quad l \in \{1, 2, \ldots \}.
\]

\[(12)\]

Now unconditioning on \( v_i \) and taking the limit \( i \to \infty \)

\[
\left(2^l - 1\right) \lim_{i \to \infty} \langle v_i^l \rangle = 2^{l-1} \bar{v} \alpha \lim_{i \to \infty} \langle v_i^{l-1} \rangle, \quad l \in \{1, 2, \ldots \}.
\]

\[(13)\]

The average newborn size at steady state is obtained by substituting \( l = 1 \)

\[
\lim_{i \to \infty} \langle v_i \rangle = \bar{v} \alpha.
\]

\[(14)\]

In general, \(13)\) can be solved iteratively to yield

\[
\lim_{i \to \infty} \langle v_i^l \rangle = \frac{\bar{v}^l \alpha^l}{2^l} \prod_{j=1}^{l} \frac{2^j}{2^j - 1}
\]

\[(15)\]

Thus, in spite of unclosed moment dynamics, the moments of newborn size can be computed exactly in closed form.

One observation from \(15)\) is that if we define a normalized size (size scaled by its average)

\[
\hat{v}_i = \frac{v_i}{\bar{v} \alpha}.
\]

\[(16)\]

then all moments of \( \hat{v}_i \) become invariant of \( \alpha \) and the mean cell size. This was dramatically seen in experiments with \( E. coli \), where the newborn size distributions across different growth conditions collapse on top of each other once scaled by their respective means \(51\). The noise in newborn size, as quantified by its Coefficient of Variation (CV) squared, is give by

\[
CV^2 := \lim_{i \to \infty} \langle v_i^2 \rangle \lim_{i \to \infty} \langle v_i \rangle^2 - 1 = \frac{1}{3}
\]

\[(17)\]
and is completely independent of model parameters $\bar{v}$ and $\alpha$. $CV^2$ computed in (17) implies a standard deviation that is $\approx 58\%$ of the average size. An interesting point to note is

$$
\lim_{i \to \infty} \langle v_i^3 \rangle = \frac{27}{28} \lim_{i \to \infty} \langle v_i^2 \rangle^3.
$$

Since for any lognormally distributed random variable $Y$,

$$
\langle Y^3 \rangle = \frac{\langle Y^2 \rangle^3 \langle Y \rangle}{\langle Y \rangle^3},
$$

(18) suggests that as far as the third-order moment is concerned (i.e., skewness), the newborn size at equilibrium is approximately lognormal. Next, we consider the general case of any exponent $m$ in (4).

**Division rate as a power function of cell size**

Before considering the general case it is worthwhile to note that as $m \to \infty$, the size just before, and after division, approaches $\bar{v}$ and $\bar{v}/2$ with probability one, respectively. Thus, one expects the noise in newborn size $CV^2 \to 0$ as $m \to \infty$, but the question remains as to how fast it goes to zero.

Our strategy in this case relies on making the division rate linear through the following transformations

$$
z_i := \left( \frac{v_i}{\bar{v}} \right)^m, \quad z := \left( \frac{v}{\bar{v}} \right)^m.
$$

Given that $v(t)$ increases exponentially between division events, $z(t)$ follows similar dynamics, but with a different exponential growth coefficient

$$
\frac{dz}{dt} = \alpha m z,
$$

with resets

$$
z \to \frac{z}{2^m}, \quad \tau \to 0,
$$

that are activated at the time of division. This transformation leads to the following discrete-time system

$$
z_{i+1} = z_i e^{\alpha T_i}, \quad i \in \{1, 2, \ldots\},
$$

where $T_i$ is given by (46) with division rate $f = z$. Following steps similar to the previous section yield the following moments for $z_i$

$$
\lim_{i \to \infty} \langle z_i^l \rangle = \frac{\alpha^l m^l}{2^l m} \prod_{j=1}^{l} \frac{j^{2m_j}}{2^{m_j} - 1}.
$$
The case of exponent $m \leq 1$

To obtain the moments of the newborn size using (24) we first consider the case $m = 1/n \leq 1$, where $n$ is a positive integer. In this case,

$$v_i = \bar{v} z_i^n \implies \lim_{i \to \infty} \langle v_i \rangle = \bar{v} \lim_{i \to \infty} \langle z_i^{nl} \rangle = \left( \frac{\bar{v}}{2} \right)^l \left( \frac{\alpha}{n} \right)^{nl} \prod_{j=1}^{nl} \frac{j2^{\frac{j}{2}}}{2^{\frac{j}{2}} - 1},$$ (25)

which yields the following noise in newborn size

$$CV^2 = \frac{\lim_{i \to \infty} \langle v_i^2 \rangle}{\lim_{i \to \infty} \langle v_i \rangle^2} - 1 = \frac{\lim_{i \to \infty} \langle z_i^{2n} \rangle}{\lim_{i \to \infty} \langle z_i^n \rangle^2} - 1 = \frac{\prod_{j=n+1}^{2n} \frac{j2^{\frac{j}{2}}}{2^{\frac{j}{2}} - 1}}{\prod_{j=1}^{n} \frac{j2^{\frac{j}{2}}}{2^{\frac{j}{2}} - 1}} - 1 \geq \frac{1}{3}$$ (26)

As expected from (18), $CV^2 = 1/3$ for $m = 1$ ($n = 1$). Moreover, $CV^2$ increases with decreasing $m$, and $CV^2 \to \infty$ as $m \to 0$ ($n \to \infty$). This latter result is consistent with the fact that a size-independent division rate yields infinite variance in newborn size [2, 52].

The case of exponent $m \geq 1$

Experimental quantification of size distributions in both *E. coli* and mammalian cells report $CV^2$ values considerably smaller than $1/3$ [5, 51, 53–55]. Since $CV^2 \geq 1/3$ for $m \leq 1$, and $CV^2 \to 0$ as $m \to \infty$, suggests that the physiological values of $m$ are much larger than one. When $m \geq 1$

$$v_i = \bar{v} z_i^\frac{1}{m},$$ (27)

where all steady-state moments of $z_i$ are given by (24). While exact analytical formulas for the moments of $v_i$ are not available in this case, we show how simple approximate formulas can be derived.

The first approximation relies on Taylor series. Assuming fluctuations in $z_i$ are small, expanding the right-hand-side of (27) around the steady-state average $\lim_{i \to \infty} \langle z_i \rangle$

$$\lim_{i \to \infty} \langle v_i \rangle \approx \bar{v} \lim_{i \to \infty} \langle z_i \rangle \frac{1}{m}$$ (28)

$$\frac{v_i}{\lim_{i \to \infty} \langle v_i \rangle} - 1 \approx \frac{1}{m} \left( \frac{z_i}{\lim_{i \to \infty} \langle z_i \rangle} - 1 \right).$$ (29)
Squaring both sides of (29) and taking the expected value yields

\[ CV^2 = \lim_{i \to \infty} \left( \frac{\langle v_i^2 \rangle}{\langle v_i \rangle^2} \right) - 1 \approx \frac{1}{m^2} \left( \lim_{i \to \infty} \langle z_i^2 \rangle \frac{1}{\lim_{i \to \infty} \langle z_i \rangle^2} - 1 \right), \quad (30) \]

which using (24) yields the following approximation

\[ CV^2 \approx \frac{2^{1+m}}{m^2} - 1. \quad (31) \]

While this approximation is exact for \( m = 1 \ (CV^2 = 1/3) \), it predicts noise in newborn size to monotonically decrease with increasing \( m \) with the asymptote

\[ CV^2 \approx \frac{1}{m^2}, \quad m \gg 1. \quad (32) \]

The second approximation works for large values of \( m \). Assuming \( m \gg 1 \), then from (20) and (24)

\[ \lim_{i \to \infty} \langle v_i^m \rangle = \left( \frac{\bar{v}}{2} \right)^m \alpha^m \prod_{j=1}^{l} \frac{j^{2mj}}{2^{mj} - 1} \approx \left( \frac{\bar{v}}{2} \right)^m \alpha^m \prod_{j=1}^{l} j = \lambda^m \ell!, \quad \lambda = \frac{\bar{v} (\alpha m)^{\frac{1}{m}}}{2}. \quad (33) \]

The moments in (34) are consistent with the newborn size at equilibrium following a Weibull distribution with scale parameter \( \lambda \) and shape parameter \( m \) [33]. For such a Weibull distribution the coefficient of variation squared is given by

\[ CV^2 \approx \frac{\Gamma \left( 1 + \frac{2}{m} \right)}{\Gamma \left( 1 + \frac{1}{m} \right)^2} - 1 \approx \frac{1.65}{m^2}, \quad m \gg 1, \quad (35) \]

where \( \Gamma \) denotes the Gamma function. Both the above approximations are plotted in Fig. 1. While (31) is much more accurate for \( 1 \leq m \leq 5 \), (35) is better for \( m \geq 6 \). Note that both approximations imply a \( 1/m \) scaling of the newborn size \( CV \).
Figure 1: Noise in newborn cell size, as quantified by its steady-state coefficient of variation squared, is plotted as a function of the size exponent $m$ in the division rate (4). For each $m$, the exact noise level is determined by running $10^7$ Monte Carlo simulations of the discrete-time system (23). While the approximation (31) is more accurate for small $m$, approximation (35) performs better for large $m$.

**Incorporating partitioning errors**

Our analysis up till now has assumed perfect symmetric division of a parent cell into two daughters. Next we consider more realistic partitioning scenarios that allow for stochastic differences between daughter cell size. In particular, at the end of $i^{th}$ cell cycle the size is reset as

$$v \rightarrow \beta_i v,$$

(37)

where $\beta_i$ are independent and identically distributed random variables drawn from a size-independent beta distribution with mean $\langle \beta_i \rangle = 1/2$. Noise in the partitioning process is quantified via the coefficient of variation squared
of β, CV\textsuperscript{2}β. Experimentally, CV\textsuperscript{2}β is quantified by tracking sizes of both daughters via

\[ CV\textsuperscript{2}β = \left\langle \left( \frac{v_{i,1} - v_{i,2}}{v_{i,1} + v_{i,2}} \right)^2 \right\rangle \] (38)

where \(v_{i,1}\) and \(v_{i,2}\) denote the two daughter cell sizes just after division. The value of CV\textsuperscript{2}β is estimated to be 10\textsuperscript{-2} for mammalian cells [5], and 10\textsuperscript{-3} – 10\textsuperscript{-4} for \textit{E. coli} [53]. The low partitioning noise in \textit{E. coli} is due to the precise positioning of the septal ring at the cell midpoint using the Min protein system [56, 57].

In the limit \(m \to \infty\), i.e., division occurs upon attainment of a critical size \(\bar{v}\), the newborn size is given by

\[ v_i = \beta v \implies CV^2 = CV\textsuperscript{2}β, \] (39)

and hence, newborn size fluctuations are completely determined by the randomness in the partitioning process. Does \(CV^2\) become more or less sensitive to \(CV\textsuperscript{2}β\) for lower values of \(m\)? To address this question, we consider a linear division rate \((m = 1)\), where moments can be determined exactly. With inclusion of random partitioning, the stochastic dynamics of the newborn size is described by the discrete-time system

\[ v_{i+1} = \beta v_i e^{\alpha T_i}, \quad i \in \{1, 2, \ldots\}, \] (40)

which yields the following moments of \(v_{i+1}\) conditioned on \(v_i\)

\[ \left\langle v_{i+1}^l | v_i \right\rangle = \langle \beta^l \rangle v^l + \frac{\langle \beta^l \rangle \bar{v} \alpha}{\langle \beta^{l-1} \rangle} \left\langle v_{i+1}^{l-1} | v_i \right\rangle, \quad l \in \{1, 2, \ldots\}. \] (41)

Unconditioning on \(v_i\) and taking the limit \(i \to \infty\)

\[ \frac{1 - \langle \beta^l \rangle}{\langle \beta^l_i \rangle} \lim_{i \to \infty} \left\langle v_i^l \right\rangle = \frac{\bar{v} \alpha}{\langle \beta^{l-1} \rangle} \lim_{i \to \infty} \left\langle v_{i+1}^{l-1} \right\rangle, \quad l \in \{1, 2, \ldots\}. \] (42)

Solving this recurrence equation leads to the following moments

\[ \lim_{i \to \infty} \left\langle v_i^l \right\rangle = \bar{v}^l \alpha^l \prod_{j=1}^{l} \frac{\langle \beta^j \rangle}{\langle \beta_{i}^{j-1} \rangle} \frac{\langle \beta^j \rangle}{1 - \langle \beta^j \rangle} \] (43)

Finally, substituting \(l = 1, 2\) and using \(\langle \beta^2 \rangle = (1 + CV^2\textsuperscript{2}β)/4 \) results in

\[ \lim_{i \to \infty} \langle v_i \rangle = \bar{v} \alpha, \quad CV^2 = \frac{\lim_{i \to \infty} \left\langle v_i^2 \right\rangle}{\lim_{i \to \infty} \left\langle v_i \right\rangle^2} - 1 = \frac{1 + 5CV^2\textsuperscript{2}β}{3 - CV^2\textsuperscript{2}β}. \] (44)
Our results show that for \( m = 1 \), \( CV^2 \) increases monotonically with \( CV^2_\beta \), but unlike (40) the dependence is nonlinear. For small values of \( CV^2_\beta \), \( CV^2 \) can be decomposed as

\[
CV^2 \approx \frac{1}{3} + \frac{16}{9} CV^2_\beta, \quad CV^2_\beta \ll 1,
\]

where the terms on the right represent noise contributions from random division events and stochastic partitioning, respectively. This result implies that the sensitivity of newborn size fluctuations to random partitioning is \( 16/9 \approx 1.78 \) for \( m = 1 \), and higher than the sensitivity for \( m = \infty \) (40).

**Conclusion**

In summary, we have analyzed a stochastic model for size control, where the likelihood of division events takes the form of power functions. Exact and approximate formulas quantifying the magnitude of fluctuations in the newborn size were derived and reveal the following key insights:

- In the absence of partitioning noise (\( CV^2_\beta = 0 \)), the extent of newborn size fluctuations is predicted to scale inversely with the exponent \( m \) in the division rate.

- Newborn size follows a lognormal distribution for values of \( m \) close to one, and a Weibull distribution for \( m \gg 1 \).

- Consistent with experimental observations [51], the normalized size defined by (16) has moments that are independent of the exponential growth coefficient and the mean cell size.

- The inclusion of random partitioning (\( CV^2_\beta \neq 0 \)) creates an additional noise term that increases nonlinearly with \( CV^2_\beta \). The sensitivity of this noise term to \( CV^2_\beta \) decreases with increasing \( m \), and approaches a value of one for \( m \to \infty \).

Can the results presented here be generalized to other forms of division rates? Recent size measurements in prokaryotes have uncovered the adder strategy for size homeostasis, where division is triggered after the cell accumulates a constant size from birth [51, 53, 58–60]. In our framework, the adder can be implemented via

\[
f(v, \tau) = \left( \frac{v (1 - e^{-\alpha \tau})}{\bar{v}} \right)^m,
\]
and a large enough $m$ corresponds to division occurring after adding a size $\bar{v}$ to the newborn size [61]. Note that (46) has timer control in addition to size control that prevents a cell to divide just after birth (i.e., $f(v, 0) = 0$). This added timer control ensures that for perfect size partitioning, the noise in newborn size is always lesser than the sizer strategy (4) for the same value of $m$ [61]. This result has important implication for estimation of $m$ from data. For example, [51] showed that the $CV$ of $E. coli$ newborn was $\approx 15\%$ across growth conditions. Using a $CV^2 = .023$, Fig. 1 provides a value of $m \approx 7$ for the division rate (4), and this implies a $m < 7$ for the adder strategy. With the increasing focus of measuring size at a single cell resolution, our theoretical results provide an important basis to understand origins of size fluctuations, and in turn use these noise measurements to uncover size control principles.

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**References**

1. Wikipedia, “List of unsolved problems in biology,”

2. C. A. Vargas-Garcia, S. Mohammad, and A. Singh, “Conditions for cell size homeostasis: A stochastic hybrid systems approach,” *arXiv:1606.00535v1 [q-bio.CB]*, 2016. https://arxiv.org/abs/1606.00535.

3. Y. Tanouchi, A. Pai, H. Park, S. Huang, R. Stamatov, N. E. Buchler, and L. You, “A noisy linear map underlies oscillations in cell size and gene expression in bacteria,” *Nature*, vol. 523, pp. 357–360, 2015.

4. A. C. Lloyd, “The Regulation of Cell Size,” *Cell*, vol. 154, pp. 1194–1205, 2013.

5. A. Tzur, R. Kafri, V. S. LeBleu, G. Lahav, and M. W. Kirschner, “Cell Growth and Size Homeostasis in Proliferating Animal Cells,” *Science*, vol. 325, pp. 167–171, 2009.
6. D. J. Kiviet, P. Nghe, N. Walker, S. Boulineau, V. Sunderlikova, and S. J. Tans, “Stochasticity of metabolism and growth at the single-cell level,” Nature, vol. 514, pp. 376–379, 2014.

7. L. Robert, M. Hoffmann, N. Krell, S. Aymerich, J. Robert, and M. Doumic, “Division in Escherichia coli is triggered by a size-sensing rather than a timing mechanism,” BMC Biology, vol. 12, p. 17, 2014.

8. T. P. Miettinen and M. Björklund, “Mevalonate Pathway Regulates Cell Size Homeostasis and Proteostasis through Autophagy,” Cell Reports, vol. 13, pp. 2610–2620, 2015.

9. K. M. Schmoller and J. M. Skotheim, “The Biosynthetic Basis of Cell Size Control,” Trends in Cell Biology, vol. 25, pp. 793–802, 2015.

10. S. Jun and S. Taheri-Araghi, “Cell-size maintenance: universal strategy revealed,” Trends in Microbiology, vol. 23, pp. 4–6, 2015.

11. A. Zaritsky and C. L. Woldringh, “Chromosome replication, cell growth, division and shape: a personal perspective,” Microbial Physiology and Metabolism, vol. 6, p. 756, 2015.

12. A. Zaritsky, “Cell-Shape Homeostasis in Escherichia coli Is Driven by Growth, Division, and Nucleoid Complexity,” Biophysical Journal, vol. 109, pp. 178–181, 2015.

13. Y. Zegman, D. Bonazzi, and N. Minc, “Measurement and manipulation of cell size parameters in fission yeast,” Methods in Cell Biology, vol. 125, pp. 423–436, 2015.

14. S. Banerjee, K. Lo, T. Kuntz, M. K. Daddysman, A. R. Dinner, and N. F. Scherer, “Crossover in the dynamics of cell wall growth controls bacterial division times,” bioRxiv 047589, 2016. http://biorxiv.org/content/early/2016/04/07/047589.

15. A. Sveiczer, B. Novak, and J. Mitchison, “The size control of fission yeast revisited,” Journal of Cell Science, vol. 109, pp. 2947–2957, 1996.

16. D. Barik, D. A. Ball, J. Peccoud, and J. J. Tyson, “A stochastic model of the yeast cell cycle reveals roles for feedback regulation in limiting cellular variability,” PLOS Computational Biology, vol. 12, pp. e1005230–, 12 2016.
17. I. Soifer, L. Robert, and A. Amir, “Single-cell analysis of growth in budding yeast and bacteria reveals a common size regulation strategy,” Current Biology, vol. 26, pp. 356–361, 2016.

18. H. Schmidt-Glenewinkel and N. Barkai, “Loss of growth homeostasis by genetic decoupling of cell division from biomass growth: implication for size control mechanisms,” Molecular Systems Biology, vol. 10, no. 12, 2014.

19. L. Willis, Y. Refahi, R. Wightman, B. Landrein, J. Teles, K. C. Huang, E. M. Meyerowitz, and H. Jönsson, “Cell size and growth regulation in the Arabidopsis thaliana apical stem cell niche,” Proceedings of the National Academy of Sciences, vol. 113, pp. E8238–E8246, 2016.

20. Y. Li, D. Liu, C. López-Paz, B. J. Olson, and J. G. Umen, “A new class of cyclin dependent kinase in chlamydomonas is required for coupling cell size to cell division,” eLife, vol. 5, p. e10767, 2016.

21. T. P. Miettinen and M. Björklund, “Cellular allometry of mitochondrial functionality establishes the optimal cell size,” Developmental Cell, vol. 39, pp. 370–382, 2016.

22. Á. Sveiczer and A. Horváth, “How do fission yeast cells grow and connect growth to the mitotic cycle?,” Current Genetics, pp. 1–9, 2016.

23. S. Kunche, H. Yan, A. Calof, J. Lowengrub, and A. Lander, “Feedback, lineages and self-organizing morphogenesis,” PLoS Comput Biol, vol. 12, p. e1004814, 2016.

24. E. Wood and P. Nurse, “Sizing up to divide: Mitotic cell-size control in fission yeast,” Annual Review of Cell and Developmental Biology, vol. 31, pp. 11–29, 2017/01/01 2015.

25. A. Horváth, A. Rácz-Mónus, P. Buchwald, and Á. Sveiczer, “Cell length growth patterns in fission yeast reveal a novel size control mechanism operating in late g2 phase,” Biology of the Cell, vol. 108, no. 9, pp. 259–277, 2016.

26. M. S. Bush, N. Crowe, T. Zheng, and J. H. Doonan, “The rna helicase, eif4a-1, is required for ovule development and cell size homeostasis in Arabidopsis,” The Plant Journal, vol. 84, no. 5, pp. 989–1004, 2015.

27. B. R. Karamched and P. C. Bressloff, “Delayed feedback model of axonal length sensing,” Biophysical Journal, vol. 108, pp. 2408–2419, 5 2015.
28. A. Amir, “Cell size regulation in bacteria,” *Physical Review Letters*, vol. 112, p. 208102, 2014.

29. J. J. Tyson and O. Diekmann, “Sloppy size control of the cell division cycle,” *Journal of Theoretical Biology*, vol. 118, pp. 405–426, 1986.

30. J. J. Turner, J. C. Ewald, and J. M. Skotheim, “Cell Size Control in Yeast,” *Current Biology*, vol. 22, pp. R350–R359, 2012.

31. K. Z. Pan, T. E. Saunders, I. Flor-Parra, M. Howard, and F. Chang, “Cortical regulation of cell size by a sizer cdr2p,” *eLife*, vol. 3, p. e02040, 2014.

32. K. M. Schmoller, J. J. Turner, M. Koivomagi, and J. M. Skotheim, “Dilution of the cell cycle inhibitor Whi5 controls budding-yeast cell size,” *Nature*, vol. 526, pp. 268–272, 2015.

33. M. Osella, E. Nugent, and M. C. Lagomarsino, “Concerted control of *Escherichia coli* cell division,” *Proceedings of the National Academy of Sciences*, vol. 111, pp. 3431–3435, 2014.

34. K. R. Ghusinga, C. A. Vargas-Garcia, and A. Singh, “A mechanistic stochastic framework for regulating bacterial cell division,” *Scientific Reports*, vol. 6, p. 30229, 2016.

35. A. Singh and J. P. Hespanha, “Approximate moment dynamics for chemically reacting systems,” *IEEE Transactions on Automatic Control*, vol. 56, pp. 414–418, 2011.

36. A. Singh and J. P. Hespanha, “Stochastic hybrid systems for studying biochemical processes,” *Philosophical Transactions of the Royal Society A*, vol. 368, pp. 4995–5011, 2010.

37. A. Singh and J. P. Hespanha, “Lognormal moment closures for biochemical reactions,” in *Proc. of the 45th Conf. on Decision and Control, San Diego*, 2006.

38. M. Soltani, C. Vargas, and A. Singh, “Conditional moment closure schemes for studying stochastic dynamics of genetic circuits,” *IEEE Transactions on Biomedical Systems and Circuits*, vol. 9, pp. 518–526, 2015.

39. A. Singh and J. P. Hespanha, “Stochastic analysis of gene regulatory networks using moment closure,” in *Proc. of the 2007 Amer. Control Conference, New York, NY*, 2006.
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40. E. Lakatos, A. Ale, P. D. W. Kirk, and M. P. H. Stumpf, “Multivariate moment closure techniques for stochastic kinetic models,” *The Journal of Chemical Physics*, vol. 143, p. 094107, 2015.

41. P. Milner, C. S. Gillespie, and D. J. Wilkinson, “Moment closure approximations for stochastic kinetic models with rational rate laws,” *Mathematical Biosciences*, vol. 231, pp. 99–104, 2011.

42. C. H. Lee, K. Kim, and P. Kim, “A moment closure method for stochastic reaction networks,” *Journal of Chemical Physics*, vol. 130, p. 134107, 2009.

43. J. Zhang, L. DeVille, S. Dhople, and A. Dominguez-Garcia, “A maximum entropy approach to the moment closure problem for stochastic hybrid systems at equilibrium,” in *IEEE Conference on Decision and Control*, pp. 747–752, 2014.

44. P. Smadbeck and Y. N. Kaznessis, “A closure scheme for chemical master equations,” *Proceedings of the National Academy of Sciences*, vol. 110, pp. 14261–14265, 2013.

45. P. Azunre, C. A. Gomez-Uribe, and G. Verghese, “Mass fluctuation kinetics: analysis and computation of equilibria and local dynamics,” *IET Systems Biology*, vol. 5, pp. 325–335, 2011.

46. M. Ullah and O. Wolkenhauer, “Investigating the two-moment characterisation of subcellular biochemical networks,” *Journal of Theoretical Biology*, vol. 260, pp. 340–352, 2009.

47. C. Schilling, S. Bogomolov, T. A. Henzinger, A. Podelski, and J. Ruess, “Adaptive moment closure for parameter inference of biochemical reaction networks,” *Biosystems*, vol. 149, pp. 15–25, 2016.

48. A. Singh and J. P. Hespanha, “Models for multi-specie chemical reactions using polynomial stochastic hybrid systems,” in *Proceedings of the 44th IEEE Conference on Decision and Control*, pp. 2969–2974, Dec 2005.

49. S. M. Ross, *Introduction to Probability Models*. Academic Press, tenth ed., 2010.

50. M. Evans, N. Hastings, and B. Peacock, *Statistical Distributions*. Wiley, 3rd ed., 2000.
51. S. Taheri-Araghi, S. Bradde, J. T. Sauls, N. S. Hill, P. A. Levin, J. Paulson, M. Vergassola, and S. Jun, “Cell-size control and homeostasis in bacteria,” *Current Biology*, vol. 25, pp. 385–391, 2015.

52. S. K. Modi, C. A. Vargas-Garcia, K. R. Ghusinga, and A. Singh, “Analysis of noise mechanisms in cell size control,” *bioRxiv*, 2016.

53. M. Campos, I. V. Surovtsev, S. Kato, A. Paintdakhi, B. Beltran, S. E. Ebmeier, and C. Jacobs-Wagner, “A constant size extension drives bacterial cell size homeostasis,” *Cell*, vol. 159, pp. 1433–1446, 2014.

54. O. Padovan-Merhar, G. P. Nair, A. G. Biaesch, A. Mayer, S. Scarfone, S. W. Foley, A. R. Wu, L. S. Churchman, A. Singh, and A. Raj, “Single mammalian cells compensate for differences in cellular volume and DNA copy number through independent global transcriptional mechanisms,” *Molecular Cell*, vol. 58, pp. 339–352, 2015.

55. H. Kempe, A. Shwabe, F. Cremazy, P. J. Verschure, and F. J. Brugge-man, “The volumes and transcript counts of single cells reveal concentration homeostasis and capture biological noise,” *Molecular Biology of the Cell*, vol. 26, pp. 797–804, 2014.

56. J. Mannik, F. Wu, F. Hol, P. Bisicchia, D. Sherratt, J. Keymer, and C. Dekker, “Robustness and accuracy of cell division in escherichia coli in diverse cell shapes,” *Proceedings of the National Academy of Sciences*, vol. 109, pp. 6957–6962, 2012.

57. A. G. Marr, R. J. Harvey, and W. C. Trentini, “Growth and division of escherichia coli,” *Journal of Bacteriology*, vol. 91, pp. 2388–2389, 1966.

58. J. T. Sauls, D. Li, and S. Jun, “Adder and a coarse-grained approach to cell size homeostasis in bacteria,” *Current Opinion in Cell Biology*, vol. 38, pp. 38–44, 2016.

59. M. Deforet, D. van Ditmarsch, and J. B. Xavier, “Cell-size homeostasis and the incremental rule in a bacterial pathogen,” *Biophysical Journal*, vol. 109, pp. 521–528, 2015.

60. A. Fievet, A. Ducret, T. Mignot, O. Valette, L. Robert, R. Pardoux, A. R. Dolla, and C. Aubert, “Single-cell analysis of growth and cell division of the anaerobe Desulfovibrio vulgaris Hildenborough,” *Frontiers in Microbiology*, vol. 6, p. 1378, 2015.
61. C. A. Vargas-Garcia, M. Soltani, and A. Singh, “Stochastic hybrid systems approach to modeling dynamics of cell size,” in *2016 IEEE 55th Conference on Decision and Control (CDC)*, pp. 5863–5868, 2016.