**ORIGINAL ARTICLE**

**HSPA1A gene polymorphism rs1008438 is associated with susceptibility to acute mountain sickness in Han Chinese individuals**

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**Abstract**

**Background:** Acute mountain sickness (AMS) usually occurs among non-acclimated individuals after rapid ascending to high-altitude environments (generally ≥2,500 m). However, the precise molecular mechanism of AMS remains unclear. Our study aimed to investigate the relationship between several single nucleotide polymorphisms (SNPs) and AMS susceptibility.

**Methods:** In this work, sequencing data were obtained from 69 AMS patients and 95 matched acclimated Han Chinese individuals from southwest China. Five SNPs (rs1008438, rs150877473, rs1799983, rs2153364, and rs3025039) were systematically investigated in all the participants.

**Results:** In our study, we found that allele frequencies of “A” (AMS 69.57% vs. non-AMS 54.74%) and “C” (AMS 30.43% vs. non-AMS 45.26%) in the HSPA1A gene rs1008438 were significantly different between the AMS and non-AMS groups (p = .01). Genotypes “CC” and “CA” of the HSPA1A gene (rs1008438) were associated with lower risk of developing AMS than the genotype “AA.” Comparing the genotypes “CC + CA” and “AA,” we also observed that the “CC + CA” genotype of rs1008438 was associated with lower AMS risk.

**Conclusions:** In our case-control study, there was a significant association between the rs1008348 polymorphism and AMS susceptibility, suggesting that this particular SNP might be a Han-specific risk factor for AMS. We believe that this study establishes a foundation for further elucidation of the genetic mechanisms underlying AMS.

**KEYWORDS**
acute mountain sickness, HSPA1A, single nucleotide polymorphism, susceptibility
1 | INTRODUCTION

Acute mountain sickness (AMS) is a medical condition resulting from acute exposure to high-altitude plateau, which develops mainly at above 2,500 m when the human body can no longer adapt to acute hypoxia (Koirala, Wolpin, & Peterson, 2018; Ogilvie, 2001; Wang, Chen, Li, Fu, & Yao, 2018). Many factors have been identified that contribute to AMS during rapid ascent to high altitudes (Koirala et al., 2018; Shen et al., 2013; Singh, 2017), including oxygen deficiency (Shen et al., 2013; Zhang et al., 2014), low temperature (Loeppky et al., 2003; Seys et al., 2013; Singh, 2017), and intense ultraviolet rays. Among these factors, oxygen deficiency plays a vital role in the development of AMS (Gonggalanzi et al., 2017). AMS can progress to life-threatening high-altitude pulmonary edema (HAPE) (Akunov et al., 2017; Du, Zhao, Su, Liu, & Yang, 2018) or to high-altitude cerebral edema (HACE) (Li, Zhang, & Zhang, 2018; West, 2015), which have a detrimental impact on an individual's health, thereby reducing the physical capability. As the modern community is increasingly undertaking activities in high plateau environments, AMS has become a public health problem. Generally, AMS is considered as a complication linked to hypoxia. However, the pathophysiology of AMS is not fully illustrated.

Some biochemical medium genetic mutations have been identified and characterized in different populations, which may affect the oxygen sensing mechanism, such as polymorphisms in the endothelial nitric oxide synthase (eNOS) (Altundag et al., 2014; Ding et al., 2011; Mansoor et al., 2005; Rossetti et al., 2017; Sun, Wang, Xi, & Hu, 2013; Tissot van Patot et al., 2005; Wang, Ha, Kidd, Koehle, & Rupert, 2010). eNOS plays a significant role in the production of nitric oxide (NO) gaseous hormone and is an essential regulator of physiological functions (Wang et al., 2013). Variations of several nitric oxide synthase 3 (NOS3) genes can affect the expression and activity of eNOS enzyme, as well as the levels of circulating and expired NO in the organism, which suggests that NOS3 variations may influence the high-altitude acclimatization (MacInnis, Wang, Koehle, & Rupert, 2011). The heat shock protein 70 (HSP70) gene family is associated with homeostatic adjustments to stress (Qi et al., 2009). One gene of the HSP70 superfamily that was described to be divergent between lowland and highland populations is HSPA1A (Dulin, Garcia-Barreno, & Guisasola, 2012). Endothelial PAS domain protein 1 (EPAS1) and egl nine homolog 1 (EGLN1) are the products of the genes involved in hypoxia adaptation. EPAS is a transcription factor and EGLN1 is a vital oxygen sensor, which plays a key role in the hypoxia-inducible factor pathway (Buroker et al., 2012; Guo et al., 2015; Hackinger et al., 2016; Yasukochi, Nishimura, Motoi, & Watanuki, 2018; Zhang et al., 2014). Additionally, vascular endothelial growth factor (VEGF) has been shown to be associated with the development of pulmonary edema under hypoxic conditions (Ding et al., 2012).

These genes have been proved to be related to the high-altitude adaptation or the development of AMS. Thus, these genes might play a key role in AMS pathogenesis. Although several previous findings emphasized the molecular mechanisms underlying gene polymorphisms associated with adaptation to high-altitude hypoxia, most investigations were carried out on European populations or non-Han Asian populations. However, China has a population of 1.3 billion, which is one-fifth of the world's population, and the vast majority of them are Han Chinese. Thus, previous studies could not thoroughly explain the occurrence of AMS in Han Chinese individuals.

To better understand the molecular mechanisms associated with AMS risk in Han Chinese, five single nucleotide polymorphisms (SNPs) of the above-mentioned genes, including HSPA1A (rs1008438), EPAS1 (rs150877473), NOS3 (rs1799983), EGLN1 (rs2153364), and VEGF (rs3025039), were selected to investigate their association with AMS susceptibility.

2 | MATERIALS AND METHODS

2.1 | Ethical statements

This study was approved by the Ethics Committee of the Southwest Hospital, Third Military Medical University (Army Medical University), Chongqing, China. Additionally, this work has been registered in the Chinese Clinical Trial Register (ChiCTR-PRC-16008441). All participants provided written informed consent to join this study, and the individual information was documented strictly confidential to protect the participants’ privacy rights.

2.2 | Data collection

For this study, we selected 250 eligible healthy Han Chinese males and aged <30 years. Meanwhile, those with high-altitude (>2,500 m) exposure history in the past year, or severe organic diseases, such as congenital heart disease, dysrhythmia, liver or kidney dysfunction, or psychological and neurological disorders; or other unsuitable conditions were excluded. Subjects were ascended to 4,000 m altitude (Litang County, Sichuan Province, China) from a low-altitude area (2,000 m, Malong County, Yunnan Province, China) by car. The AMS group consisted of 69 individuals detected by the Lake Louise scoring system (LLS) results. Meanwhile, 95 healthy control individuals without symptoms of AMS were randomly selected for the non-AMS group.
We collected the clinical characteristics of the AMS and non-AMS groups, including age, weight, height, body mass index (BMI), and pulse oxygen saturation (SpO₂). Blood pressure was detected using electronic sphygmomanometers (OMRON Healthcare Co. Ltd.). SpO₂ levels of participants were detected using pulse oximeters (Nonin Medical Co. Ltd.). Blood pressure and SpO₂ levels were detected three times after the participants rested in a quiet environment for above 15 min, respectively.

2.3 DNA extraction and index determination

Venous blood (4–5 ml) was drawn from all participants for further genetic analysis. Genomic DNA was extracted from whole blood cells of the participants using the Gentra Puregene Blood Kit (Qiagen Co. Ltd.) and the process was carried out according to the manufacturer’s protocol (Yang et al., 2018).

2.4 Genotyping

Single nucleotide polymorphisms genotyping was characterized by the single base extension detecting technology (iPLEX). The single-base extension primers and SNP loci-tested polymerase chain reaction (PCR) primers were designed by the Sequenom mass array assay design genotyping software (Sequenom). The purified extension products were added into the 384-element SpectroCHIP bioarray (Sequenom) by the mass array nanodispenser RS 1000 (Capital Bio Corporation). The data were analyzed using the TYPER 4.0 software (Sequenom).

2.5 Statistical analysis

Allele frequencies and genotype distribution were calculated in the AMS and non-AMS groups and analyzed for an association with the development of AMS via the Chi-square test. Odds ratios (ORs) and 95% confidence intervals (CI) were calculated to test the association between AMS risk and genotype. Statistical analysis was conducted using SPSS 17.0 (SPSS Co. Ltd.) and significance was set at p < .05.

3 RESULTS

3.1 Participant characteristics

The characteristics of participants are summarized in Table 1. Comparing the AMS and non-AMS groups, our results indicated no significant differences in terms of age, height, weight, BMI, and SpO₂ at 2,000 m among the study subjects. The SNP selection information is presented in Table 2.

3.2 Genotyping of target SNPs associated with AMS risk

In the assay, dominant, codominant, and recessive models were applied to assess the association of SNPs within HSPA1A, EPAS1, NOS3, EGLN1, and VEGF with the risk of AMS. For HSPA1A (rs1008438), the distribution of the allele and overall genotype frequency was significantly different between the AMS and non-AMS groups (p = .01). However, there was no significant difference in the distribution of both the variant genotypes (p > .05) and alleles (p > .05) for the other four genes between the AMS and non-AMS groups (Table 3).

The allele frequencies of “A” (AMS 69.57% vs. non-AMS 54.74%) and “C” (AMS 30.43% vs. non-AMS 45.26%) of HSPA1A rs1008438 were significantly different between the AMS and non-AMS groups (p = .01). However, there was no significant difference in the distribution of both the variant genotypes (p > .05) and alleles (p > .05) for the other four genes between the AMS and non-AMS groups (Table 3).

The allele frequencies of “AA” (AMS 69.57% vs. non-AMS 54.74%) and “CC” (AMS 30.43% vs. non-AMS 45.26%) of HSPA1A rs1008438 were significantly different between the AMS and non-AMS groups (p = .01). The frequency of the “AA” genotype was significantly higher among the AMS group (52.17%) than the non-AMS group (30.53%) (p = .01). The genotype frequencies of HSPA1A rs1008438 “AA,” “CA,” and “CC” were 52.17% (n = 36), 34.78% (n = 24), and 13.04% (n = 9) in the AMS group, respectively, and 30.53% (n = 29), 48.42% (n = 46), and 21.05% (n = 20) in the non-AMS group, respectively. Additionally, genotypes “CC” (OR, 0.36; 95% CI, 0.14–0.92) and “CA” (OR, 0.42; 95% CI, 0.21–0.84) of rs1008438 were associated with a significantly lower AMS risk than genotypes “AA” (p = .03 and p = .01, respectively; Figure 1-a). Comparing the genotype

| Characteristics | AMS (n = 69) | Non-AMS (n = 95) | p-Value |
|-----------------|-------------|-----------------|---------|
| Age (mean ± SD, years) | 28.20 ± 0.80 | 27.00 ± 1.20 | .805 |
| Weight (mean ± SD, kg) | 65.48 ± 8.14 | 65.05 ± 7.17 | .723 |
| Height (mean ± SD, cm) | 172.00 ± 4.00 | 173.00 ± 5.00 | .687 |
| BMI (kg/m²) | 22.19 ± 2.16 | 21.96 ± 1.99 | .502 |
| SpO₂ (%) | 96.78 ± 1.04 | 96.71 ± 1.24 | .707 |

Abbreviations: AMS, acute mountain sickness; BMI, body mass index; SpO₂, pulse oxygen saturation.
of “CC + CA” with “AA” (Figure 1-b), the results illustrated that the “CC + CA” genotype of rs1008438 was associated with a lower AMS risk (OR, 0.40; 95% CI, 0.21–0.77; p = .01). However, genotype “CC” (OR, 0.56; 95% CI, 0.24–1.33) of rs1008438 presents no significant differences when compared with genotypes “CA + AA” (p = .19; Figure 1-c). These results illustrated that rs1008438 “AA” homozygous genotype was significantly associated with an elevated morbidity due to AMS compared with the homozygous “CC” and heterozygous “CA” genotypes. Taken together, HSPA1A rs1008438 was associated with AMS risk.

### 4 | DISCUSSION

Acute mountain sickness is considered a multifaceted disease whose occurrence and development are influenced by genetic and environmental factors (Koirala et al., 2018; Singh, 2017). An individual’s susceptibility to AMS under the specific environmental conditions may be related to its genetic makeup. Currently, 23 genes have been reported as putative genetic susceptibility factors for AMS. Among them, the most frequently reported are NOS3, HSPA1A, EPAS1, EGLN1, and VEGF.

NO is an effective vasodilator and plays an important role in acclimatization and adaptation to hypoxia. In the vasculature, NO acts locally via relaxing smooth muscles and thereby causing vasodilatation and is produced by NO synthases (NOSs) encoded by NOS3 (Altundag et al., 2014; MacInnis et al., 2011; Mansoor et al., 2005; Wang et al., 2010, 2013). Heat shock protein 70 (HSP70) family is a critical component of cytoprotective gene products (Qi et al., 2009). HSPA1A, one of the polymorphic genes of the HSP70 family, has been widely considered in association with a variety of disease symptoms. EPAS1 encodes a transcription factor that is involved in the hypoxia-inducible factor pathway, which is the key regulator of responses to hypoxia (Buroker et al., 2012; Dulin et al., 2012; Guo et al., 2015). The EGLN1 gene encodes a vital oxygen sensor that negatively regulates hypoxia-inducible factor-1 alpha (HIF-1A) activity (Yasukochi et al., 2018; Zhang et al., 2014). The inactivation of EGLN1 caused by hypoxia leads to an increase of HIF activity, which induces the expression of adaptive responses genes. This pathway suggests that EGLN1 may be an important factor for high-altitude adaptation in the population.

Under hypoxic conditions, VEGF expression is significantly upregulated and the high levels of VEGF in the lungs can cause pulmonary vascular permeability increase, leading to the occurrence of pulmonary edema (Ding et al., 2012). In most studies, it has been reported that VEGF is a key factor in the pathogenesis of high-altitude adaptation and sickness (Bian et al., 2016; Buroker et al., 2013; Yu et al., 2016). Thus, we selected 5 SNPs (rs1008438, rs150877473, rs17999983, rs2153364, and rs3025039) in these genes to investigate their association with AMS.

Recent genomic surveys of SNPs among humans in high-altitude environments have uncovered evidence for a history of positive selection of these genes. The most reliable signals include several genes with known roles in oxygen transport and regulation. One of these genes, HSPA1A was proved to be divergent between lowland and highland populations. Within the superfamily of HSP, HSPA1A has anti-inflammatory and anti-apoptotic functions. Two studies have associated variants of HSP gene polymorphisms with AMS in Chinese populations. Li et al. detected polymorphisms in the HSP70-1 (b1/b2; +190G/C) and HSP70-2 (A/B; +1267A/G) genes in their study; however, no association was found between the HSP70-1 polymorphism and AMS, whereas the HSP70-2 B/B genotype was over-represented in individuals with AMS (Li et al., 2004). Additionally, Zhou et al. also detected the same polymorphisms in HSP70-1 and HSP70-2, and another polymorphism (A/B; 2437G/C) in the HSP70-hom gene and their study confirmed the findings of Li et al. (Zhou et al., 2005).

Although five SNPs were investigated in the present study, the results demonstrated that only the HSPA1A polymorphism (rs1008438) might be a potential factor for the development of AMS. In our study, genotypes “CC” and “CA” of rs1008438 were associated with lower AMS risk than the genotypes “AA” (p = .03 and p = .01, respectively). Comparing with “AA” genotypes, the results also indicated that the “CC + CA” genotype of rs1008438 was associated with lower AMS risk (p = .01). Our analysis demonstrated that the “A” allele carriers were at a 2.29-fold higher risk of developing AMS than the “C” allele carriers. Thus, the “A” allele of the HSPA1A (rs1008348) polymorphism was associated with an increased risk of AMS, having implications for Han populations in particular.

Previous studies have reported that several genetic variants of EPAS1 are associated with high-altitude adaptation in Tibetans. Among these variants, the derived G allele of rs150877473 was 78% of a higher frequency in Tibetans than
TABLE 3  SNPs genetic models and analyses of the association between SNPs and AMS

| SNP No.   | Genotype | Total    | Non-AMS (%) | AMS (%) | B     | SE     | $\chi^2$ | p       | OR (95% CI) |
|-----------|----------|----------|-------------|---------|-------|--------|---------|---------|-------------|
| rs1008438 | C        | 128 (39.02) | 86 (45.26) | 42 (30.45) |       |        |         | .01*    | 0.36 (0.14, 0.92) |
|           | A        | 200 (60.98) | 104 (54.74) | 96 (69.57) | 7.39  | 0.47   | 4.61    | .03     |             |
|           | CC       | 29 (17.68)  | 20 (13.04)  | 9 (13.04)  | -1.01 | 0.47   | 4.61    | .03     | 0.36 (0.14, 0.92) |
|           | CA       | 70 (42.68)  | 46 (34.78)  | 24 (34.78) | -0.87 | 0.35   | 5.98    | .01     | 0.42 (0.21, 0.84) |
|           | AA       | 65 (39.63)  | 29 (52.17)  | 36 (52.17) |       |        |         |         |             |
|           | CC + CA  | 99 (60.37)  | 66 (47.83)  | 33 (47.83) | -0.91 | 0.33   | 7.68    | .01     | 0.40 (0.21, 0.77) |
|           | AA       | 65 (39.63)  | 29 (52.17)  | 36 (52.17) |       |        |         |         |             |
|           | CC       | 29 (17.68)  | 20 (13.04)  | 9 (13.04)  | 0.58  | 0.35   | 1.73    | .19     | 0.56 (0.24, 1.33) |
|           | CC + CA  | 135 (82.32) | 75 (86.96)  | 60 (86.96) |       |        |         |         |             |
| rs150877473 | G       | 9 (2.74)   | 5 (2.63)    | 4 (2.90)   | 0.02  | .88    |         |         |             |
|           | C        | 319 (97.26) | 185 (97.37) | 134 (97.10)|       |        |         |         |             |
|           | GG       | 0 (0.00)   | 0           | 0         |       |        |         |         |             |
|           | GC       | 9 (5.49)   | 5 (5.26)    | 4 (5.80)   | 0.10  | 0.69   | 0.02    | .88     | 1.11 (0.29, 4.29) |
|           | CC       | 155 (94.51) | 90 (94.74)  | 65 (94.20) |       |        |         |         |             |
|           | GG + GC  | 9 (5.49)   | 5 (5.26)    | 4 (5.80)   | 0.10  | 0.69   | 0.02    | .88     | 1.11 (0.29, 4.29) |
|           | CC       | 155 (94.51) | 90 (94.74)  | 65 (94.20) |       |        |         |         |             |
|           | GG       | 0 (0.00)   | 0           | 0         |       |        |         |         |             |
|           | GC + CC  | 164 (100.0)| 95 (100.00) | 69 (100.00)|       |        |         |         |             |
| rs1799983 | T        | 38 (11.59) | 21 (11.05)  | 17 (12.32) |       |        |         |         |             |
|           | G        | 290 (88.41) | 169 (88.95) | 121 (87.68)| 0.13  | .72    |         |         |             |
|           | TT       | 3 (1.83)   | 1 (1.05)    | 2 (2.90)   | 1.02  | 1.24   | 0.68    | .41     | 2.78 (0.25, 31.42) |
|           | GT       | 32 (19.51) | 19 (20.00)  | 13 (18.84) | -0.05 | 0.40   | 0.02    | .90     | 0.95 (0.43, 2.09) |
|           | GG       | 129 (78.66) | 75 (78.95)  | 54 (78.26) |       |        |         |         |             |
|           | TT + GT  | 35 (21.34) | 20 (21.05)  | 15 (21.74) | 0.04  | 0.39   | 0.01    | .92     | 1.04 (0.49, 2.22) |
|           | GG       | 129 (78.66) | 75 (78.95)  | 54 (78.26) |       |        |         |         |             |
|           | TT       | 3 (1.83)   | 1 (1.05)    | 2 (2.90)   | -1.03 | 1.24   | 0.70    | .40     | 2.81 (0.25, 31.58) |
|           | GT + GG  | 161 (98.17) | 94 (98.95)  | 67 (97.10) |       |        |         |         |             |
| SNP No. | Genotype | Total | Non-AMS (%) | AMS (%) | B     | SE  | χ² | p   | OR (95% CI) |
|---------|----------|-------|-------------|---------|-------|-----|----|-----|-------------|
| rs2153364 | A       | 169 (51.84) | 104 (55.32) | 65 (47.10) | −0.73 | 0.48 | 2.15 | .14 | 0.48 (0.19, 1.24) |
|         | G       | 157 (48.16) | 84 (44.68)  | 73 (52.90)  | 2.15 | .14 | —  | —  | —           |
|         | AA      | 40 (24.39)  | 27 (28.42)  | 13 (18.84)  | −0.73 | 0.48 | 2.15 | .14 | 0.48 (0.19, 1.24) |
|         | AG      | 89 (54.27)  | 50 (52.63)  | 39 (56.52)  | −0.25 | 0.40 | 0.38 | .54 | 0.78 (0.35, 1.72) |
|         | GG      | 35 (21.34)  | 18 (18.95)  | 17 (24.64)  | —    | —   | —   | —   | —           |
|         | AA + AG | 129 (78.66) | 77 (81.05)  | 52 (75.36)  | 1.03 | .31 | —  | —  | —           |
| rs3025039 | T       | 52 (15.95)  | 29 (15.43)  | 23 (16.67)  | —    | —   | —   | —   | —           |
|         | C       | 274 (84.05) | 159 (84.57) | 115 (83.33) | 0.09 | .76 | —  | —  | —           |
|         | TT      | 6 (3.66)    | 4 (4.21)    | 2 (2.90)    | 0.89 | 0.14 | .71 | .71 | 0.72 (0.13, 4.08) |
|         | CT      | 40 (24.39)  | 21 (22.11)  | 19 (27.54)  | 0.89 | 0.14 | .71 | .71 | 0.72 (0.13, 4.08) |
|         | CC      | 118 (71.95) | 70 (73.68)  | 48 (69.57)  | 0.26 | 0.37 | 0.51 | .48 | 1.30 (0.63, 2.68) |
|         | TT + CT | 46 (28.05)  | 25 (26.32)  | 21 (30.43)  | 0.19 | 0.35 | 0.59 | .59 | 1.21 (0.61, 2.40) |
|         | CC      | 118 (71.95) | 70 (73.68)  | 48 (69.57)  | —    | —   | —   | —   | —           |
|         | TT      | 6 (3.68)    | 4 (4.21)    | 2 (2.90)    | —    | —   | —   | —   | —           |
|         | CT + CC | 158 (96.34) | 91 (95.79)  | 67 (97.10)  | —    | —   | —   | —   | —           |

Abbreviations: AMS, acute mountain sickness; CI, confidence intervals; OR, Odds ratio; SNP, single nucleotide polymorphism.

*Indicates statistically significant after the false discovery rate (FDR) correction.
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CONFLICT OF INTEREST

All authors declared that there was no conflict of interest.

AUTHOR CONTRIBUTIONS

Xiaomei Wang and Chunyan Yao performed the experiment; Zhicheng Liu and Ting Xu analyzed the data and written manuscript; Hong Chen and Chunyan Yao designed the research and revised the manuscript. All authors have approved the final manuscript to submission.

DATA AVAILABILITY STATEMENT

The data were not shared to protect participants’ privacy.

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