Sequential Astrocytic 5-HT$_{2B}$ Receptor Stimulation, [Ca$^{2+}$]$_{i}$ Regulation, Glycogenolysis, Glutamate Synthesis, and K$^{+}$ Homeostasis are Similar but Not Identical in Learning and Mood Regulation

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INTERRELATION BETWEEN ANXIETY AND MEMORY

A close interrelation between anxiety and memory was first suggested by Kalueff and Nutt (1996), reviewing effects of $\gamma$-aminobutyrate (GABA) on both conditions. Wall and Messier (2000) showed subsequently that pretreatment with an opioid kappa receptor antagonist was anxiogenic and disrupted working memory. Additional papers supporting interactions between anxiety and memory were cited by Kalueff (2007). They included demonstration of memory improvement by serotonin, whereas a decreased ability to increase serotonin is a model of anxiety.

SEROTONIN AND THE 5-HT$_{2B}$ RECEPTOR

Serotonin acts on many different receptors. The present paper specifically deals with the 5-HT$_{2B}$ receptor, which is expressed in human brain (Schmuck et al., 1994; Bonhaus et al., 1995). Its mRNA expression is two times higher in freshly isolated (Lovatt et al., 2007) mouse astrocytes than in neurons (Li et al., 2012). It is necessary for consolidation of one-trial aversive learning in day-old chickens (Gibbs and Hertz, 2014) at an early stage of memory consolidation. It is also required for the therapeutic effect of serotonin-specific reuptake inhibitors (SSRIs) in major depression (Diaz et al., 2012; Li et al., 2012; Hertz et al., 2015b), a disease often accompanied by anxiety. The 5-HT$_{2B}$ receptor in cultured astrocytes is stimulated by fluoxetine (Li et al., 2008; Qiao et al., 2015) and all other SSRIs (Zhang et al., 2010). Chronic treatment of mice with fluoxetine for 14 days up-regulates the astrocytic, but not the neuronal 5-HT$_{2B}$ receptor, although this receptor is expressed in both cell types (Li et al., 2012; Hertz et al., 2015b). Decrease of its astrocytic gene expression parallels development of a depressive phenotype in a mouse model of Parkinson’s disease (Zhang et al., 2015), and Pitychoutis et al. (2015) reported schizophrenia-like symptoms in mice lacking the 5-HT$_{2B}$ receptor gene or treated with a receptor inhibitor. Schizophrenia is often associated with depressed mood (Fortunati et al., 2015).
Inhibition of learning by a 5-HT\textsubscript{2B/C} receptor antagonist (SB221284) and equipotent rescue of impaired learning by the 5-HT\textsubscript{2B} receptor agonists fluoxetine and paroxetine (Gibbs and Hertz, 2014) injected intracerebrally at specific times shows the importance of this receptor for establishment of memory soon after training. The similar potency of the two drugs is important, because they have widely different affinities for SERT (Wong and Bymaster, 1995) whereas all SSRIs have similar affinity for the 5-HT\textsubscript{2B} receptor (Zhang et al., 2010). Another SSRI, citalopram, counteracts spatial memory deficits (Ren et al., 2015). Mice lacking the 5-HT\textsubscript{2B} receptor gene show learning disabilities (Pitychoutsis et al., 2015).

Fluoxetine increases free cytosolic Ca\textsuperscript{2+} ([Ca\textsuperscript{2+}]\textsubscript{i}) and stimulates glycogenolysis (Chen et al., 1995) with similar potency by stimulation of 5-HT\textsubscript{2B} receptors (Kong et al., 2002; Figure 1A). [Ca\textsuperscript{2+}]\textsubscript{i} regulates many astrocytic functions, including gliotransmission and glycogenolysis (Gucek et al., 2012; Hertz et al., 2015a). Inhibition of glycogenolysis with DAB (1,4-dideoxy-1,4-imino-D-arabinitol) prevents 5-HT\textsubscript{2B}-receptor-mediated memory enhancement by serotonin or fluoxetine during the early part of memory formation after one-trial averse learning in the day-old chicken, a precocious animal (Gibbs and Hertz, 2014). In brain both glycogen and its degrading enzyme glycogen phosphorylase are virtually confined to astrocytes (Ibrahim, 1975; Pfeiffer-Guglielmi et al., 2003). Induction of glycogenolysis by fluoxetine occurs both in our cultured astrocytes, differentiated by dibutyryl cyclic AMP and in astrocytes grown in the absence of this agent (Allaman et al., 2011). The association with increased [Ca\textsuperscript{2+}]\textsubscript{i} (Chen et al., 1995) is important because increased [Ca\textsuperscript{2+}]\textsubscript{i} is a requirement for stimulation of glycogenolysis in astrocytes (Xu et al., 2014a; Hertz et al., 2015a) as in muscle (Ozawa, 2011). In rat brain 5-HT\textsubscript{2} receptor stimulation similarly induces glycogenolysis (Darvesh and Gudelsky, 2003). The enhanced glycogenolysis is accompanied by an increased lactate release (Allaman et al., 2011). This might affect neurons either by use of lactate as an additional metabolic fuel, as suggested by Suzuki et al. (2011) and Newman et al. (2011), or by lactate signaling (Tang et al., 2014; Bergrersen, 2015). The signaling mechanism established by Tang et al. (2014) is, like memory (Gibbs et al., 2006; Newman et al., 2011; Suzuki et al., 2011; Gibbs and Hutchinson, 2012; Duran et al., 2013), glycogenolysis-dependent, and its signaling is specifically directed to neurons releasing noradrenaline. Noradrenaline has effects on both neurons and astrocytes (O’Donnell et al., 2012).

Glycogenolysis is also required for formation of glutamate, and its metabolite GABA (Figure 1B) in the brain in vivo (Gibbs et al., 2007, 2008) at a time when glutamate production must be evoked by 5-HT\textsubscript{2B} stimulation (Gibbs and Hertz, 2014). It also increases uptake of glutamate into cultured astrocytes and neurons as well as release of lactate from astrocytes (Sickmann et al., 2009). Glutamate is synthesized intracerebrally from glucose. This can only occur in astrocytes, because neurons lack an enzyme, pyruvate carboxylase, needed for its synthesis (reviewed by Gibbs et al., 2008; Hertz, 2013). Glutamate is subsequently converted to glutamine and carried to neurons (green arrow in Figure 1B) in an extremely active glutamine-glutamate/GABA cycle, which also returns released transmitter glutamate to neurons after its initial astrocytic accumulation (reviewed by Hertz, 2013; Hertz and Rothman, in press). The importance of glutamate receptor activity for memory is beyond doubt (Riedel et al., 2003), and interruption of the glutamine-glutamate/GABA cycle by inhibition of either glutamine synthetase (Kant et al., 2014) or astrocytic glutamate uptake (Gibbs et al., 2004) abolishes learning. GABA is also important for learning (Kaluueff and Nutt, 1996; Gibbs and Bowser, 2009), and besides its neuronal effects stimulates glycogenolysis in cultured astrocytes and brain slices (Xu et al., 2014a).
A major role of glutamate (Figure 1B) is stimulation of postsynaptic glutamate receptors, leading to increases in brain metabolism (Howarth et al., 2012) and in extracellular K\(^+\) concentration (Hertz et al., 2015c and references therein). Cellular re-accumulation of K\(^+\) includes an initial uptake mediated by the astrocytic Na\(^+\),K\(^+\)-ATPase (MacAulay and Zeuthen, 2012; Hertz et al., 2015c), release of astrocytically accumulated K\(^+\) by Kir4.1 channels (Bay and Butt, 2012) and neuronal reuptake. The astrocytic Na\(^+\),K\(^+\)-ATPase is important for learning (Moseley et al., 2007; Schaefer et al., 2011; Hertz et al., 2013; Tadi et al., 2015). Extracellular K\(^+\) concentrations high enough to stimulate the Na\(^+\), K\(^+\), 2 Cl\(^-\) co-transporter NKCC1 (>10–12 mM) also causes release of gliotransmitters (Song et al., 2014; Xu et al., 2014b; Liu et al., 2015). Both astrocytic release of glutamate (Lee et al., 2014) and ATP (Gibbs et al., 2011; Stehberg et al., 2012) are crucial for learning. Facilitation of learning by K\(^+\)-mediated depolarization of oligodendrocytes and increased myelination at high extracellular K\(^+\) concentration (Roitbak, 1984) and attributed to increased ability of the myelinated axon to carry out rapid impulse conduction has recently been confirmed and characterized by Yamazaki et al. (2014).

5-HT\(_{2B}\) RECEPTOR, [Ca\(^{2+}\)]\(_i\), GLYCOGENOLYSIS, GLUTAMATE, K\(^+\), AND MOOD DISORDERS

Fluoxetine is better known for its antidepressant effect, which in contrast to the acute stimulation of the 5-HT\(_{2B}\) receptor during learning takes several weeks to materialize. During this time many changes occur in gene expression and editing, as shown in mice chronically treated with fluoxetine. Studies in neuronal and astrocytic cell cultures freshly obtained from these mice (Lovatt et al., 2007) showed that most of these alterations occurred in astrocytes, although some neuronal changes also took place (Li et al., 2012; Peng et al., 2014; Hertz et al., 2015b). This finding suggests that astrocytes play a major role in the antidepressant effects of SSRIs (Li et al., 2012; Hertz et al., 2015b), a conclusion in agreement with results by many other authors (e.g., Ongür et al., 1998; Kugaya and Sanacora, 2005; Ongür et al., 2007; Valentine and Sanacora, 2009; Krysta et al., 2015). Both astrocytic release of glutamate (Lee et al., 2014) and ATP (Gibbs et al., 2011; Stehberg et al., 2012) are crucial for learning. Facilitation of learning by K\(^+\)-mediated depolarization of oligodendrocytes and increased myelination at high extracellular K\(^+\) concentration (Roitbak, 1984) and attributed to increased ability of the myelinated axon to carry out rapid impulse conduction has recently been confirmed and characterized by Yamazaki et al. (2014).

RELATED ASTROCYTIC MECHANISMS IN SCHIZOPHRENIA

Schizophrenia is treatable both by the dopamine antagonist haloperidol and atypical antipsychotics like clozapine, which is an antagonist at the 5-HT\(_{2B}\) receptor in the fundus of the stomach (Villazón et al., 2003). Again, acute stimulation of the 5-HT\(_{2B}\) receptor is likely to increase [Ca\(^{2+}\)]\(_i\), glycogenolysis and glutamate formation (Figure 1B). An increase in [Ca\(^{2+}\)]\(_i\) by stimulation of astrocytic dopamine receptors is reduced by exposure to clozapine (Reuss and Unsicker, 2001), and this seems also to be the case after clozapine activation of 5-HT\(_{2B}\) receptors. A resulting reduced production of glutamate (Figure 1B) in mice lacking 5-HT\(_{2B}\) receptors may explain a decreased content of glutamate in some brain areas (Pitychoutis et al., 2015), which may contribute to the impairment of learning.

CONCLUDING REMARKS

Activation of the astrocytic 5-HT\(_{2B}\) receptor stimulates an increase in [Ca\(^{2+}\)]\(_i\), glycogenolysis, glutamate formation, and the effect of glutamate on extracellular K\(^+\), all of which are involved in learning (Figure 1B). However, Sibille et al. (2015) found that acute inhibition of Ca\(^{2+}\) signaling in astrocytes by [Ca\(^{2+}\)]\(_i\) chelation potentiates excitatory synaptic transmission. This apparent contradiction may be explained by the complexity of astrocytic [Ca\(^{2+}\)]\(_i\) regulation (Volterra et al., 2014). An important difference between Gibbs and Hertz (2014) and Sibille et al. (2015) is that the latter authors elicited astrocytic increase in [Ca\(^{2+}\)]\(_i\) in response to adjacent neuronal activity.
during GABA_A receptor inhibition, whereas the former described transmitter-induced, glycolgenolytic (and thus Ca^{2+}-dependent) effects on learning without GABA_A receptor inhibition.

Drugs used for treatment of symptoms of major depression (fluoxetine) and of schizophrenia (clozapine), which included memory impairment, interfered with 5-HT_2B receptor-activated functions, but in different manners: the SSRI fluoxetine edited and thereby reduced some normal effects of this receptor, whereas clozapine caused a decrease in [Ca^{2+}],. This effect is consistent with the enhancement of excitatory synaptic transmission described by Sibille et al. (2015). Disposition to both major depression and schizophrenia is probably inborn, and perhaps these patients display quantitative and/or qualitative abnormalities in 5-HT_2B-mediated signaling, which might also affect learning processes. In agreement with this notion 5-HT_2B receptors play a major role during brain development (Lauder et al., 2000).

**AUTHOR CONTRIBUTIONS**

All authors planned or carried out reviewed experiments. YC and LH wrote the paper and MEG edited it.

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