Lactate-Dependent Regulation of Immune Responses by Dendritic Cells and Macrophages

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For decades, lactate has been considered an innocuous bystander metabolite of cellular metabolism. However, emerging studies show that lactate acts as a complex immunomodulatory molecule that controls innate and adaptive immune cells’ effector functions. Thus, recent advances point to lactate as an essential and novel signaling molecule that shapes innate and adaptive immune responses in the intestine and systemic sites. Here, we review these recent advances in the context of the pleiotropic effects of lactate in regulating diverse functions of immune cells in the tissue microenvironment and under pathological conditions.

**Keywords:** dendritic cells, macrophages, lactate signaling, GPR81/GPR132, regulatory and inflammatory responses, antitumor immunity, immune response to infections, inflammatory diseases

**INTRODUCTION**

For decades, lactate has been considered a waste product of cellular metabolism. New lines of investigations now recognize this glycolytic metabolite as an active immune modulator that shapes the functions of immune cells in the tissue microenvironment and under pathological conditions (1–3). Accumulation of lactate in the tissue microenvironment is an essential feature of both inflammatory diseases and cancer (1–3). Much is known about the production of lactic acid under various disease conditions, while the mechanisms by which it shapes the effector functions of immune cells and restores tissue homeostasis remain obscure. Recent evidence suggests an emerging role for lactate in the field of inflammation, autoimmunity, and cancer (1–3). Here, we review our current knowledge of the role of lactate in regulating inflammatory and regulatory responses in various tissue environments, highlight some unanswered questions, and discuss how this new information can be exploited in the rational design of therapies against various autoimmune disorders, infections, and cancers.

**DENDRITIC CELLS AND MACROPHAGES IN REGULATING INFLAMMATORY AND REGULATORY RESPONSES**

The immune system launches robust immunity against foreign antigens while maintaining a state of tolerance to self-antigens, food antigens, and commensal flora (4–8). Loss of tolerance to self-antigens,
food antigens, and commensal flora leads to immune cell-mediated inflammatory diseases and autoimmunity (4–9). Dendritic cells (DCs) and macrophages are a specialized subset of antigen-presenting cells (APCs) that form a critical link between innate and adaptive immune cells. These APCs represent a complex immunological system composed of several functionally distinct subsets distributed in different organs and microenvironments (4–9). A detailed discussion of DC and macrophage subsets and their influence on adaptive immunity is outside the scope of the present review and was reviewed extensively recently (4–9). DCs and macrophages express a wide array of pathogen recognition receptors (PRRs) which enables them to sense different pathogen-associated molecular patterns (PAMPs) and damage-associated molecular pattern molecules (DAMPs) (7, 10). Signaling through PRRs activate and program APCs to induce distinct innate responses that shape the type of T-helper (Th) responses (7, 10). In addition to inducing robust immune responses against infections, DCs and macrophages also play a critical role in suppressing inflammatory responses and maintaining tissue immune homeostasis. Furthermore, DCs and macrophages induce immune tolerance and contribute to the resolution of inflammation through several regulatory mechanisms (11). However, the cellular and molecular mechanisms underlying this phenomenon remain poorly understood. Emerging studies suggest a fundamental role for lactate in the tissue microenvironment in regulating immunity and immune tolerance by shaping the functions of DCs and macrophages. Here, we will discuss how lactate shapes the functions of DCs and macrophages under steady-state and inflammatory conditions.

**LACTATE METABOLISM AND TRANSPORT IN DENDRITIC CELLS AND MACROPHAGES**

Upon activation, DCs and macrophages undergo profound metabolic changes critical for biosynthesis and energy production (12). Lactate could also serve as a fuel source to produce energy by various cell types, including immune, cancer, and stromal cells (13, 14). Like other cell types, APCs produce lactate under hypoxic conditions or by aerobic glycolysis. A phenomenon similar to the Warburg effect in tumors is also observed in DCs and macrophages following TLR activation which induces a major metabolic reprogramming characterized by a switch from oxidative phosphorylation (OXPHOS) to glycolysis. This metabolic shift reprograms APCs from a regulatory state to an inflammatory state and intracellular and extracellular lactate levels play an essential role in this process.

Under homeostatic conditions, intracellular and extracellular lactate levels are tightly regulated. Lactate production occurs in the cytoplasm within the cell due to hypoxic or aerobic glycolysis and accumulates in the extracellular space. Lactate dehydrogenases (LDHs) are critical enzymes in glycolysis that reversibly catalyze the conversion of pyruvate to lactate or lactate to pyruvate (1, 13). LDH is a tetrameric enzyme composed of two types of subunits namely LDH-A and LDH-B. LDH-A has a higher affinity for pyruvate and preferentially catalyzes pyruvate to L-lactate, while LDH-B has a higher affinity for lactate and converts L-lactate to pyruvate, fueling oxidative metabolism. Immune cells, including DCs and macrophages, express both LDH-A and LDH-B subunits (1). However, proinflammatory DCs and macrophages express higher levels of LDH-A and show increased production of lactate due to the sustained glycolytic reprogramming induced by TLR ligands. In DCs this metabolic shift depends on activating transcription factors such as sterol regulatory element-binding protein (SREBP) and hypoxia-inducible factor (HIF)-1α (15). Furthermore, HIF-1α plays a crucial role in regulating the expression of LDH-A and several other genes involved in glycolysis (15). Evidence suggests that HIF1-α deficiency in DCs and macrophages leads to loss of GLUT1 (a facilitative glucose transporter) and LDHA (16–18). Besides, HIF-1α can be activated through a feedback mechanism by intracellular pyruvate or lactate (19, 20). However, whether this effect depends on SREBP or the direct control of inflammatory cytokine expression is unknown.

Lactate in the cell or extracellular space is transported across the plasma membrane by monocarboxylate transporters (MCTs) of the SLC16 solute carrier family, and they transport lactate by an H+-coupled transport mechanism (13, 14, 21, 22). MCTs, prevent intracellular accumulation of lactate by removing excess lactate produced due to increased glycolytic activity (23, 24). Dendritic cells and macrophages express MCT1, MCT2 and MCT4 (13, 14, 22). MCT1 and MCT2 have a higher affinity for lactate and are primarily responsible for transporting lactate into the cells. MCT4 has a lower affinity for lactate and is primarily responsible for the export of lactate. Interestingly, lactate also regulates the expression of MCT1 and MCT4 (25). In addition to MCTs, two other solute carrier family 5 members (SLC5), namely SLC5A8 (SMCT1, sodium-coupled monocarboxylate transporter 1) and SLC5A12 (SMCT2, sodium-coupled monocarboxylate transporter 2) can also mediate transmembrane transfer of lactate, and they transport lactate by a Na+-coupled transport mechanism. DCs, macrophages, and other immune cells express SLC5A8 and SLC5A12 (13, 14, 22). MCTs and SMCTs play a key role in lactate transport in APCs, yet their regulation and roles under steady-state and inflammatory conditions are incompletely understood.

**The Lactate-Mediated Signaling Pathway**

New lines of investigation now place lactate as an active signaling molecule that controls the differentiation and functions of immune cells under steady-state and inflammatory conditions. In addition, lactate exerts autocrine effects on the host cells and paracrine effects on other cell types in the tissue environment. Recent studies have revealed some of the signaling pathways by which lactate shapes the functions of DC and macrophage through receptor-dependent and receptor-independent mechanisms.

**LACTATE-GPR81 SIGNALING AXIS**

L-Lactate, a ubiquitous metabolite, functions as a natural ligand for GPR81 (HCA1, hydroxy-carboxylic acid receptor) (26, 27).
Lactate activates GPR81 in its physiological concentration range of 1–20 mM (17). GPR81 expression varies depending on the cell type and tissue microenvironment. For example, fat cells express high levels of GPR81 whereas secondary lymphoid tissues, gut, brain, kidney express low levels of GPR81 (26, 28, 29). Recently, several groups have reported that DCs and macrophages express GPR81, and its expression is regulated by the tissue microenvironment (29–32). Our recent work has shown that DCs and macrophages in the intestine and lung express higher levels of GPR81 compared with DCs and macrophages in the spleen (30). Likewise, DCs in the tumor microenvironment (TME) express high levels of GPR81 (13, 32). An important unresolved question is how the tissue microenvironment regulates GPR81 expression in these APCs. Adipocyte studies have shown that peroxisome proliferative–activated receptor γ (PPARγ) transcriptionally regulates GPR81 expression (27, 33). Lipids and their metabolites are potent activators of the PPAR family transcription factors in APCs (34–36). These ligands are widely present in the intestine and TME suggesting that PPAR-mediated signaling might regulate GPR81 expression in APCs (34–36). Besides, recent studies have shown that lactate can regulate GPR81 expression in tumor cells via the snail3/STAT3 (signal transducer and activator of transcription 3) pathway (37). Further studies are warranted to see whether the PPARγ and snail3/STAT3 pathways regulate GPR81 expression in DCs and macrophages. Recent studies have highlighted a protective role for GPR81 in minimizing tissue injury by controlling pathological inflammatory responses (31). Lactate-GPR81 mediated signaling in non-immune cells regulates several key signaling pathways such as the cyclic adenosine monophosphate (cAMP), protein kinase A (PKA), and extracellular signal-regulated kinase (ERK) pathways. However, the downstream signaling networks of GPR81 in DCs and macrophages are unknown. GPR81 suppresses inflammatory responses in monocytes and macrophages by limiting the activation of the β-arrestin/inflammasome pathway (31). In pDCs, GPR81 signaling regulates IFNα production by inducing intracellular Ca2+ mobilization and its downstream genes Ca2+/calmodulin dependent protein kinase II (CaMKII), and calcineurin (CaN) phosphatase (38). In addition to modulating these pathways, other signaling pathways, such as inhibition of nuclear factor-kappa B (NF-kB), play a role in the anti-inflammatory function of lactate in macrophages. GPR81 signaling in macrophages exerts suppressive effects on NF-kB and yes-associated protein (YAP) activation via activation of AMP-activated protein kinase (AMPK) and large tumor suppressor kinases (LATS), resulting in reduced proinflammatory cytokine production after exposure to LPS (39) (Figure 1). In contrast to its anti-inflammatory role, an in vitro study has shown that lactate augmented LPS-induced expression of inflammatory genes by enhancing NF-kB activation in human monocyte-derived macrophages and U937 cells (40). In the TME, GPR81-signaling plays an essential role in immune suppression against tumors by inducing regulatory APCs (32) and upregulating the expression of programmed

**FIGURE 1** | The Lactate-mediated receptor-dependent and receptor-independent signaling pathways. Lactate binds to GPR81 and GPR132 receptors and activates several downstream signaling pathways and transcription factors in DCs and macrophages. (A) Lactate binding to GPR81 and GPR132 results in the activation or suppression of several downstream pathways such as PI3K/AKT/CREB, PLC/IP3/Ca2+, β-arrestin/inflammasome, AMPK/LATS/YAP/NF-kB. This results in reduced expression of proinflammatory cytokine production and increased expression of immune regulatory factors (IL-10, IDO, RA, TGFβ) in response to TLR ligands. (B) Lactate can shape APC functions independent of surface receptors. MCTs transport extracellular lactate into the cells, and intracellular lactate can modulate APC functions by directly regulating the activation of multiple signaling pathways and transcription factors such as HIF-1α, MAPK, ERK, and NF-kB.
modulation of DC and macrophage functions by lactate

Robust immune responses against pathogens and tumors depend on several factors, such as the degree of maturation and activation of DCs, their ability to capture, process, and present exogenous antigens, them trafficking to secondary lymphoid organs and tissues type of factors they produce. Emerging studies have shown that lactate-mediated signaling is crucial in shaping immune responses by modulating DC and macrophage functions (Figure 2 and Table 1). Here, we will discuss how lactate shapes essential functions of DCs and macrophages that influence adaptive immune responses.

regulation of dc maturation and activation by lactate

DC maturation and activation are important in inducing a robust immune response against tumors and pathogens (11). Immature or tolerogenic DCs facilitate tolerance or immune regulatory responses, whereas immunogenic/inflammatory DCs facilitate robust inflammatory responses (7). Under homeostatic conditions, peripheral DCs typically display an immature phenotype characterized by low surface levels of MHC II and costimulatory molecules (CD80, CD86, and CD40) and induce robust T-cell activation and effector differentiation (7). However, certain stimuli induce the tolerogenic/regulatory DCs that express markedly lower costimulatory molecules and induce regulatory T cells and immune suppression. Several reports have shown that lactate-mediated signaling blocks DC differentiation, activation and antigen presentation (53, 60). Exposure to lactate conditions DCs to a regulatory or anti-inflammatory state. Earlier ex vivo studies have shown that DCs cultured in the presence of lactate exhibit regulatory functions (53–59). These regulatory DCs expressed low surface levels of MHC II and costimulatory molecules and produced markedly lower levels of IL-12 and higher levels of IL-10. Besides, a recent study has shown that human tolerogenic DCs produce high levels of lactate that shape T-cell responses toward tolerance and delayed graft-versus-host disease (91). Like DCs, lactate exposure polarizes macrophages to M2 phenotype with increased expression of CD163 and Arg1 and decreased expression of M1 markers such as CD86, iNOS, IL-1β, and IL-6 (41). This lactate effect on TAMs depends on Nrf2 (nuclear factor erythroid 2-related factor 2), HIF-1α, and MCT1 (20, 78, 79).

DCs and macrophages recognize diverse microbial structures through multiple receptors collectively known as PRRs (10). DCs and macrophages can also recognize damage-associated molecular patterns (DAMPs) and other endogenous ligands released from dying tumor cells through PPRs (92–94).
PRRs include Toll-like receptors (TLRs), C-type lectin-like receptors (CLRs), RIG-I-like receptors (RLRs), and Nod-like receptors (NLRs) (10). TLR ligands have gained significant interest in immunotherapy in recent years for their potential use as vaccine adjuvants (95, 96). In general, PRR engagement potently activates DCs by upregulating the surface expression of maturation markers such as MHCII, CD80, CD83, and CD86 (10). Even though PRR ligands are there in the TME and mucosal organs (92–94), DCs and macrophages present in these environments display markedly decreased expression of costimulatory molecules (97). Earlier ex vivo studies on human and murine DCs have shown that exposure to lactate markedly affected the maturation and activation in response to LPS (31, 53). Lactate also inhibited the LPS-mediated activation of bone marrow-derived macrophages and peritoneal macrophages. Lactate-conditioned macrophages failed to upregulate costimulatory molecules while expressing lower levels of proinflammatory cytokines and higher levels of IL-10 even in response to LPS ligands (29, 31, 39, 98). Further, mechanistic studies have shown that lactate signaling can negatively regulate the inflammatory pathways such as the NF-kB, NFAT (nuclear factor of activated T-cells), YAP, inflamasome, and MAPK (mitogen-activated protein kinases) pathways, critical for DC activation and expression of inflammatory factors (31, 59). Accordingly, DCs and macrophages that are deficient in GPR81 are hyper-responsive to TLR ligands (30). Also, other studies revealed the role of monocarboxylate transporters (MCTs) in mediating the lactate effect in macrophages (77). MCT4 inhibition significantly boosted lactate-induced M2 polarization, while blocking of MCT1/2 failed to reverse the immunosuppressive effect of lactate, correlating with the results from gene expression studies that showed lactate increasing MCT4 expression but downregulating the expression of MCT1/2 (59). Thus, the effects of lactate on the maturation and activation of DCs and macrophages involve GPR81 dependent and independent mechanisms via MCTs.

**REGULATION OF DENDRITIC CELL MIGRATION BY LACTATE**

The migration of DCs to secondary lymphoid organs and tissues is essential for initiating adaptive immune responses, tumor immune surveillance, regulation of inflammation in the tissues, and selective elimination of infected cells (99, 100). DC migration involves its trafficking to tissues, capturing and endocytosing dead or infected cells, and transporting associated antigens to the draining lymph nodes (DLNs) where they prime and activate T cells to initiate adaptive immune responses (101–104). DC migration depends on the expression of specific chemokine receptors on DCs and its
cognate chemokine ligand within the tissues and DLNs. DC migration to DLNs requires chemokine receptor CCR7, whereas its recruitment to the tissues depends on chemokines such as CCL4, CCL5, and XCL1 (99). However, only a tiny fraction of DCs migrate to tumor tissue and subsequently to the draining lymph nodes. Glycolytic metabolism is essential for CCR7 oligomerization and DC migration (94). Blocking glycolysis impairs CCR7 oligomerization and impairs migration (63). Ex vivo studies have shown that high lactate levels (20 mM) inhibited the migration of monocytes and DCs (53, 64). Similarly, lactate regulates macrophage functions such as adhesion, migration, and tissue recruitment (41, 80, 81). Furthermore, lactate in the TME and inflamed tissues can regulate the migration of immune cells by regulating the expression of several key enzymes involved in glycolysis (105).

**REGULATION OF ANTIGEN DELIVERY AND PRESENTATION BY LACTATE**

Cross-presentation is critical for initiating immune responses against tumors and viral infections, where DCs present extracellular antigens on MHC I to activate CD8⁺ T cell-mediated cytotoxicity (10). Effective cross-presenting involves uptake of extracellular antigens, processing antigens into peptides, loading peptides onto MHC I, and trafficking of MHC I: peptide complex to the cell’s surface (106). Membrane trafficking proteins such as SNARE (soluble n-ethylmaleimide-sensitive factor attachment protein receptor) and VAMP3 (vesicle-associated membrane protein 3) play a critical role in cross-presentation. Loss of these membrane trafficking proteins in DCs leads to defective cross-presentation of tumor-associated antigens (57). DCs within the TME are less efficient in cross-priming CD8⁺ T cells (61, 62), and the TME contains high lactate levels. Emerging evidence has shown that lactate affects DCs’ function by regulating antigen presentation and cross-priming CD8⁺ T cells (53, 60). However, the underlying molecular mechanisms by which lactate affects cross-presentation are not known. In this context, a recent study has shown that lactate can affect cross-presentation by downregulating membrane trafficking proteins such as SNAREs and VAMP3 while accelerating antigen degradation in DCs (57). Furthermore, these proteins facilitate the secretion of cytokines from DCs upon activation. Further studies are warranted to see whether the

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**TABLE 1** | Evidence for involvement of the lactate in the microenvironment in shaping the functions of innate and adaptive immune cells.

| Observations | References |
|--------------|------------|
| **Lactate effects on DC function** | |
| Lactate suppresses DC differentiation and maturation. | (53–59) |
| Lactate suppresses the activation of DCs and the expression of proinflammatory factors in response to TLR ligands. | (31, 53) |
| Lactate inhibits antigen delivery and presentation by DCs. | (53, 65–62) |
| Lactate accelerates antigen degradation in DCs by downregulating membrane trafficking proteins. | (67) |
| Lactate-GPR81 signaling in intestinal DCs induces the expression of immune regulatory factors to induce Tregs and suppresses the differentiation of Th1/Th17 cells. | (53, 60–62) |
| Lactate in the TME conditions DCs to a regulatory state to suppress antitumor immune responses. | (53, 60–62) |
| Lactate signaling regulates the expression of chemokine receptors and chemokines that are critical for DC migration. | (53, 63–65) |
| Lactate signaling in DCs regulates metabolic pathways involving glycolysis and fatty acid oxidation (FAO). | (60, 66) |
| **Lactate effects on T cells** | |
| Lactate suppresses T cell proliferation, cytokine production and Th1 differentiation. | (67–69) |
| Lactate promotes Treg proliferation and functions. | (67, 70–72) |
| Under inflammatory conditions, lactate signaling in CD4⁺ T cells favors Th17 cell differentiation. | (73, 74) |
| Lactate suppresses the T cell migration and trafficking. | (73, 74) |
| Tumor-derived lactate limits the expansion of tumor-antigen specific CD8⁺ T cells, cytokine production, CTL activity. | (75, 76) |
| Lactate synergizes with IL-21 to promote stemness of CD8⁺ T cells and antitumor immunity. | (70) |
| **Lactate effects on macrophage** | |
| Lactate induces alternative polarization (M2) of macrophages. | (29, 41, 77) |
| Lactate signaling in macrophages attenuates TLR-induced proinflammatory cytokine production. | (31, 39, 59) |
| Lactate signaling in TAMs promotes tumor growth, migration, metastasis, and immunosuppression. | (20, 41, 78–80) |
| Lactate-GPR132 signaling in macrophages contributes to tumor cell invasiveness and tumor growth. | (65) |
| Lactate-GPR81 signaling impairs regulatory phenotype on intestinal macrophages and induces the expression of immune regulatory factors to induce Tregs. | (53, 60–62) |
| Lactate-GPR81 signaling in macrophages suppresses expression of inflammatory factors in response to LPS. | (31, 39) |
| **Lactate effects on MDSCs** | |
| Lactate promotes the development and accumulation of MDSCs in tumors. | (82) |
| Lactate-conditioned MDSCs inhibit the function of natural killer (NK) cells and T lymphocytes. | (83, 84) |
| **Lactate effects on NK cells** | |
| Tumor-derived lactic acid inhibits natural killer (NK) cell maturation and function. | (75) |
| **Lactate effects on other immune cells** | |
| Lactate regulates the functions of basophils, neutrophils, mast cells. | (85–88) |
| **Lactate effects on Tumor cells** | |
| Lactate promotes tumor growth, migration, and metastasis. | (13, 89, 90) |
regulation of membrane trafficking proteins by lactate is dependent on GPR81 and MCTs. These studies collectively suggest that the lactate-mediated signaling suppresses efficient capture of tumor-associated antigens by tumor DCs and cross-priming of CD8⁺ T cells.

REGULATION OF IMMUNE REGULATORY AND INFLAMMATORY FACTORS BY LACTATE

DCs dictate the fate of naïve CD4⁺ and CD8⁺ T cells through differential production of pro- and anti-inflammatory cytokines (11, 107). Recent studies have shown that lactate can shape the adaptive immune responses by regulating the expression of immune regulatory factors and inflammatory factors in DCs and macrophages (108–113). DCs and macrophages exposed to TLR-ligands produce markedly higher levels of proinflammatory cytokines and type-1 interferons (IFN). In contrast, lactate-conditioned DCs and macrophages do not release immunostimulatory cytokines; instead, they express higher levels of IL-10 in response to TLR ligands (53–55, 57, 58). The TME contains higher levels of immune regulatory factors such as IL-10, retinoic acid (RA), and TGF-β that actively suppress differentiation and expansion of tumor-specific effector T cells (114, 115). Lactate in the TME condition DCs and macrophages to a regulatory or anti-inflammatory state (1, 2). Accordingly, tumor DCs deficient in the lactate receptor GPR81 expressed markedly higher levels of IL-12 and IL-6 (32). Similarly, lactate-GPR81 signaling influences the pDC functions in tumors by attenuating IFNα production (38). Furthermore, blocking GPR81 signaling can restore the IFNα production by pDCs. Lactate in the TME conditions DCs and macrophages to express higher levels of IL-10 (1, 2). The effects of lactate on the expression of the regulatory and inflammatory cytokines in APCs also depend on MCTs. Furthermore, blocking the MCT in DCs or macrophages can reprogram them to an inflammatory state (59). These APCs produce high levels of inflammatory factors in response to TLR ligands (38). In the intestine, anti-inflammatory factors such as IL-10, TGF-β, IDO, and RA produced by DCs and macrophages are critical for maintaining immune tolerance to commensal flora (116, 117). These immune regulatory factors are also necessary to suppress inflammation and restore immune homeostasis in the intestine. A recent study has highlighted an essential role for the lactate-GPR81 signaling in intestinal DCs and macrophages in regulating the expression of immune regulatory factors such as IL-10, retinoic acid (RA), and IDO (30). Intestinal DCs and macrophages isolated from GPR81-deficient mice produced markedly higher levels of inflammatory cytokines and lower levels of anti-inflammatory factors under homeostatic and inflammatory conditions (30). Furthermore, GPR81-deficient intestinal APCs are hyper-responsive to microbial ligands and express higher levels of proinflammatory cytokines (30). Collectively, these studies demonstrate that lactate-mediated signaling imparts an anti-inflammatory phenotype to DCs and macrophages.

REGULATION OF IMMUNE CELL METABOLISM BY LACTATE

Cellular metabolic pathways play a critical role in modulating the functions of DCs and macrophages (12, 56), and emerging evidence support lactate as one of the essential molecules that links metabolism and immunity. DC and macrophage subsets have potential metabolic differences under homeostatic and inflammatory conditions (12, 56). Tolerogenic or regulatory DCs and macrophages show a catabolic metabolism marked by increased oxidative phosphorylation, fatty acid oxidation (FAO), and glutaminolysis (12, 56). In contrast, immunogenic or inflammatory DCs display an anabolic metabolism marked by increased glycolysis and lactate production (12, 56). Preliminary evidence suggests that lactate mediates immune cell-intrinsic effects on metabolism (73). Besides, extracellular lactate induces metabolic reprogramming of DCs and macrophages, resulting in reduced glycolysis and increased FAO (29, 60, 66). This metabolic reprogramming of APCs significantly changes cytokine production with predominantly anti-inflammatory effects, emphasizing the complex interplay between metabolism and APC functions (29, 60, 66). The effects of lactate on immune cell metabolism may serve as a negative feedback signal limiting inflammation (3). For example, lactate can modulate APC functions by regulating the expression of critical enzymes involved in glycolysis (3). These studies show that lactate imparts regulatory phenotype on APCs by metabolic reprogramming.

EFFECTS OF LACTATE IN MODULATING THE FUNCTIONS OF OTHER IMMUNE CELLS

Emerging studies are beginning to provide insights into the mechanisms by which lactate signaling cascade directly shapes the effector phenotypes of myeloid-derived suppressor cells (MDSCs), Tregs, CD4⁺ T cells, CD8⁺ T cells, and natural killer (NK) cells. Several excellent studies and reviews discuss extensively how extracellular lactate shapes the functions of other immune cells (12, 56) and will thus be discussed only briefly (Table 1). Lactate can exhibit a proinflammatory or anti-inflammatory effect depending on the microenvironment and immune cell type conditions and factors. For example, lactate exerts an immune-suppressive role in the TME, whereas lactate exerts an inflammatory role in chronic conditions like arthritis. Lactate in the TME promotes expansion and accumulation of MDSCs while suppressing the effector functions of NK cells, CD4⁺ T lymphocytes, CTLs and mast cells (83–85). On the other hand, under chronic inflammatory conditions, lactate manifests an inflammatory role on CD4⁺ T cells by promoting the differentiation of Th17 cells (73). Effect of lactate on Th17 cell differentiation is T cell-intrinsic, but the underlying molecular mechanism is unknown and requires further investigation. These studies collectively show that the multi-faceted effect of lactate on the immune response is dependent on cellular and environmental contexts.
REGULATION OF AUTOIMMUNITY AND ANTITUMOR IMMUNITY BY LACTATE

Accumulation of lactate in the tissue microenvironment is a feature of both inflammatory disease and cancer. Emerging evidence suggests that this is due to metabolic disturbances in immune cells. Lactate exhibits an inflammatory or anti-inflammatory role depending on its effects on immune cell type, disease type, and tissue environment (12, 56). This section will review recent developments in our understanding of the role of lactate-mediated signaling in regulating immune responses in pathological conditions.

INFLAMMATORY BOWEL DISEASE (IBD)

Loss of immune tolerance to intestinal commensal flora and oral antigens leads to chronic intestinal inflammation and inflammatory bowel disease (IBD). In the colon, lactate is one of the primary metabolites produced by bacterial fermentation of dietary products and gastrointestinal mucosa is exposed to high concentrations of lactate (66, 73, 117). Besides, intestinal epithelial cells and immune cells can produce lactate (118-120). Initial study on murine models of IBD showed that the intrarectal treatment with lactate prevents intestinal inflammation by downregulating proinflammatory response in epithelial cells (121). However, whether lactate regulates immune responses to gut commensal flora remains largely unknown. Our recent work has revealed an essential role for GPR81 in programming tolerogenic DCs and macrophages in the intestine (30). Mice deficient in GPR81 are highly susceptible to chemically-induced colitis and T cell-mediated colitis. Besides, genetic deletion of GPR81 in mice led to loss of immune homeostasis in the intestine, which enhanced susceptibility to colonic inflammation (30). Besides intestinal APCs, lactate plays a crucial role in intestinal stem-cell-mediated regeneration of the epithelial layer through the GPR81-Wnt signaling pathway (122). This observation is particularly relevant in the intestine, given the importance of Wnt signaling in intestinal DCs and macrophages in regulating immune tolerance and commensal homeostasis in the intestine (123, 124). It would be interesting to see how lactate and Wnt signaling pathways cross-regulate each other in establishing immune tolerance and commensal homeostasis in the gut. In a striking functional similarity with GPR81 knockout mice, genetic deficiency of GPR132 also resulted in significantly worsened chemically-induced colitis in mice (48, 125). However, GPR132 deficiency does not alter intestinal immune homeostasis under homeostatic conditions. GPR132-mediated signaling in myeloid and lymphoid cells limits intestinal inflammation in a mouse model of colitis induced by dextran sodium sulfate (125). Collectively, these studies have identified a new and essential role for lactate, GPR81, and GPR132 signaling pathways in regulating immune tolerance and colonic inflammation.

OTHER IMMUNE-MEDIATED INFLAMMATORY DISEASES

Lactate plays a protective role in murine models of immune hepatitis and pancreatitis (31). In this model, the lactate-mediated protective effect is dependent on GPR81 signaling that limits the expression of proinflammatory factors by macrophages. Mice deficient in GPR81 are highly susceptible to LPS-induced hepatitis and pancreatitis. Confirming this finding, pharmacological activation of GPR81 decreased LPS-induced activation of the caspase-1 and NF-κB pathways and production of proinflammatory factors by macrophages and reduced disease severity in mice (31). Lactate plays a similar anti-inflammatory role in Multiple sclerosis (MS). MS is a chronic inflammatory demyelinating neurological disease of the central nervous system (CNS). In the experimental autoimmune encephalomyelitis (EAE) model of MS, macrophages in the CNS display higher expression of LDHA and increased glycolysis (126). CNS macrophages also expressed higher levels of MCT4. siRNA-mediated knockdown of LDHA and MCT4 or blocking MCT4 reduced leukocyte infiltration and the clinical severity of EAE (126). However, the effects of lactate on the functions of DCs and other immune cells in this chronic inflammatory disease are not known.

In contrast to its regulatory and protective role, lactate significantly induces and promotes inflammation in rheumatoid arthritis (RA) (1). RA is a chronic inflammatory disease that affects joint linings causing pain, swelling, and deformity (127). The inflamed synovial tissue microenvironment includes an increased number of inflammatory DCs, macrophages and pathological effector T cells. Recent studies have revealed that lactate exacerbates disease severity by regulating migration of immune cells in the arthritic synovium (73, 74). Mainly, lactate inhibits T cell motility, which contributes to their entrapment in the inflammatory site. This depends on the lactate transporters SLC5A12 and SLC16A1 (MCT1) (73, 74). In addition, lactate also drives the differentiation of T helper 17 (Th17) cells that can exacerbate inflammation and disease severity. However, the biological effects of lactate on the APCs under inflammatory conditions are much less understood. Therefore, further investigation requires a more detailed understanding of the lactate effect on different subsets of immune cells under inflammatory versus steady-state conditions.

LACTATE IN REGULATING IMMUNE RESPONSES TO INFECTIONS

Emerging studies show that lactate modulates immune responses to infections. As discussed above, DCs and macrophages recognize different pathogens through PRRs, and signaling through these receptors leads to increased glycolysis and increased lactate production. Sepsis is a common and frequently fatal clinical condition characterized by an initial systemic inflammatory response to infection followed by an immunosuppressive phase (128). A recent study utilizing murine models of sepsis has shown that, lactate-GPR81-mediated signaling suppresses the expression of proinflammatory cytokines and induces alternative polarization of macrophages to M2 phenotype (39, 98). Similarly, lactate-induced activation of GPR109a improves survival in mice with sepsis (129). However, the effects of lactate on the functions of DCs and other immune cells in sepsis are unknown. RIG-I-like receptor (RLRs)-
mediated signaling is necessary for Type I interferon (IFN) production and this is critical for augmenting host immunity for viral clearance and cancer immune surveillance (10). A recent study has shown that lactate can affect IFN production by negatively regulating the RLR-MAVS-RIG-I pathway (130). Besides, blocking lactate production or metabolism increased type I IFN production with enhanced viral clearance (130). Several pathogens can modulate DC and macrophage function as a mechanism to evade host immune response, resulting in chronic infections such as TB, HIV, HCV, HBV, and SIV (10). However, the role of lactate in the regulation of innate and adaptive immune responses to chronic infections is unknown. In this context, a recent study has shown that in response to Mycobacterium tuberculosis (Mtb) infection, macrophages switch from pyruvate oxidation to reduction of pyruvate into lactate (131). Besides, Mtb utilizes intracellular lactate as an energy source for growth in macrophages (131). This metabolic switch in macrophages to Mtb infection also increases anti-inflammatory factors such as IL-10 (132). Anti-inflammatory factors produced by APCs play a significant role in establishing chronic infections (10). Collectively, these studies showed that lactate could modulate the immune responses to infections.

**LACTATE SIGNALING IN TUMOR-INDUCED IMMUNE TOLERANCE**

Tumors express self-antigens that actively suppress host antitumor immune responses (114, 133). Increased lactate levels positively correlate with tumor grade, progression, recurrence, metastasis, and poor prognosis in several types of cancer (13, 134, 135). As discussed above, lactate secreted by the tumor cells suppresses immune responses by modulating the phenotype and functions of DCs and macrophages in the TME (136, 137). Besides, high lactate levels in the TME impart an anti-inflammatory phenotype on APCs, contributing to immune suppression. Lactate also promotes tumor progression by inducing the prostaglandin E2 (PGE2) synthesis and cyclooxygenase 2 (COX2) upregulation in monocytes (65). PGE2 is a potent immunomodulator that exhibits both proinflammatory and anti-inflammatory effects on DCs and macrophages. Tumors exploit lactate-mediated signaling to effectively suppress host antitumor immune responses (1, 138). DCs and macrophages in the TME express lactate receptors GPR81 and GPR132 (32, 38). The importance of lactate-mediated signaling in controlling antitumor immune responses was demonstrated in a study using GPR81 knockout mice (32). Accordingly, GPR81-deficiency in mice resulted in delayed tumor growth and significantly reduced tumor burden in a syngeneic transplant model and a constitutive breast cancer model in mice (32). Tumor DCs from these mice displayed enhanced activation and increased expression of proinflammatory cytokines such as IL-6 and IL-12 (32). pDCs produce type I IFN and are critical for antitumor immunity. However, pDCs in the TME are dysfunctional and produce low levels of IFNs, which is partly due to lactate in the TME (38). The lactate effect on pDC dysfunction is dependent on GPR81 Signaling and MCTs. Besides, lactate signaling in pDCs induced regulatory T Cell induction by regulating the tryptophan metabolism. Like DCs, macrophages in the TME exert potent effects on cancer metastasis and antitumor immunity. Similar to GPR81, lactate signaling via GPR132 in macrophages promotes tumor growth and metastasis (41). Consistent with these observations, mice deficient in GPR132 displayed a significant reduction in tumor burden and breast cancer metastasis. However, the underlying molecular mechanisms are unknown. In addition to APCs, lactate can suppress antitumor immune responses by modulating the functions of other immune cells (67, 70, 71, 83, 84). In summary, these studies reveal an exciting and unappreciated role for lactate in contributing to immune suppression against tumors through different effector mechanisms.

**TARGETING THE LACTATE SIGNALING PATHWAY FOR IMMUNE MODULATION AND IMMUNOTHERAPY**

There is considerable interest in the lactate signaling pathway as a therapeutic target, especially as a treatment for inflammatory diseases and cancer. Studies involving human cancers and inflammatory diseases strongly suggest that targeting the lactate signaling pathway and lactate metabolism is a promising approach to overcome immune evasion by tumors and suppressing immune-mediated inflammatory diseases. These strategies include targeting signaling (GPR81/GPR132 antagonists), lactate transporters (MCT inhibitors), and lactate metabolism (LDH inhibitors). In addition, pharmacological activators and inhibitors of the lactate signaling pathway exist, and several of them are currently in clinical testing. Here, we will briefly discuss preclinical studies related to the effects of blocking the lactate signaling pathway and lactate metabolism on antitumor immunity and autoimmunity.

**TARGETING LACTATE-GPR81/GPR132 SIGNALING**

Lactate receptor expression is upregulated in several types of cancer and lactate signaling plays a vital role in tumor development, progression, and metastasis (13, 89, 90). As discussed above, Lactate receptor-mediated signaling in immune cells contributes to the suppression of antitumor immune responses. Thus, blocking specific lactate ligand with cognate GPR81/GPR132 receptors represents a potential strategy to restrain tumor cell proliferation while boosting the antitumor immunity. In this context, a recent study using a 4T1 breast cancer model has shown that intratumoral injection of a GPR81 inhibitor along with an MCT inhibitor resulted in a significant reduction in tumor burden in mice (38). Another critical study has demonstrated that blocking GPR132 signaling in macrophages markedly reduced tumor burden, progression, and breast cancer metastasis in mice (41).
Likewise, pharmacological inhibition of GPR132 signaling had a similar effect on tumor burden and antitumor immune responses (139).

On the other hand, activating the GPR81 pathway in APCs may help prevent and treat immune cell-mediated inflammatory diseases such as IBD, hepatitis, and pancreatitis. In this context, our previous study has shown that pharmacological activation of the GPR81 pathway suppressed intestinal inflammation by inducing Treg responses and limiting pathological Th1/Th17 responses. Preclinical studies have shown that lactate treatment suppresses inflammatory responses in the intestine and mitigates intestinal injury (121). Oral administration of lactate had a similar suppressive effect on inflammation-associated gastric injury (140).

Furthermore, intratumoral injection of AR-C155858 caused a significant reduction in 4T1 tumor burden in mice (38). Immune checkpoint inhibitors are currently used in cancer immune therapy and ameliorated the disease severity (73, 74, 153). CD147 (EMMPRIN) plays a crucial role in regulating MCT expression by stabilizing and localizing MCTs to the cell membrane. Therefore, disrupting the interaction between CD147 and MCT is also an attractive strategy to regulate immune responses in human diseases. Targeting CD147 has yielded encouraging results in preclinical models of inflammatory diseases (154, 155). Studies have shown that the loss of CD147 function decreases the levels of MCT1 and MCT4 proteins and reduces tumor growth (156–158).

TARGETING LACTATE TRANSPORTERS

The second strategy to augment antitumor immune responses involves blocking lactate transporters using MCT inhibitors (22, 144). MCTs are highly expressed in tumors and positively correlate with cancer patients’ poor outcomes (22, 144). Besides, MCTs promote migration and invasion processes in several cancer types, including lung and breast cancers (22, 144). Several studies have examined antitumor immune responses by blocking lactate transport into the APCs using clinically relevant murine tumor models. Blockade of cytosolic transport of lactate in pDCs using AR-C155858 (MCT inhibitor) restored the IFNα production and augmented the immune responses against 4T1 tumors in mice. Furthermore, intratumoral injection of AR-C155858 caused a significant reduction in 4T1 tumor burden in mice (38). Immune checkpoint inhibitors are currently used in cancer immune therapy to enhance immune responses. Another study has shown that silencing MCT1 and MCT4 can restore T cell-induced immune function and boost the immune response to immune checkpoint inhibitors in melanoma patients (75). Also, treatment of Raji xenograft-bearing severe combined immunodeficiency mice with AZD9365 led to inhibition of tumor growth with increased tumor immune cell infiltration involving DCs and natural killer cells (145). MCT1/2 inhibitors are currently in Phase I/II clinical trials to treat patients with advanced prostate cancer, gastric cancer, or diffuse large B-cell lymphoma (146–150). Collectively, these preclinical studies show that drugs that target MCTs alone or in combination with immune checkpoint inhibitors hold much promise as cancer treatments.

In certain autoimmune diseases such as arthritis and MS, MCTs play an inflammatory role. Accumulating evidence shows that blocking lactate efflux or influx has an immunosuppressive effect (151). MCT1/2/4 play a vital role in this, and pharmacological inhibitors of the transporters are attractive targets in these immune-cell mediated inflammatory diseases. Preclinical studies utilizing the murine model of MS have shown that silencing or blocking MCT4 reduced leukocyte infiltration into the CNS and the clinical severity of EAE. Similarly, the silencing of MCT4-inhibited proliferation of RA synovial fibroblast (RASFs) reduced the severity of arthritis in a mouse model of collagen-induced arthritis (152). Further studies utilizing murine models of peritonitis and arthritis have shown that blocking or silencing lactate transporter (SLC5A12) restored the T cell functions and ameliorated the disease severity (73, 74, 153).

TARGETING LACTATE METABOLISM

The third strategy to augment antitumor immune responses involves targeting lactate metabolism (23, 159). LDH-A increases the production of lactate in tumor cells and immune cells resulting in tumor immune escape by inhibiting the function of immune cells (71, 160). There is a strong correlation between elevated lactate dehydrogenase (LDH) and poor prognosis in cancer patients. Besides, cancer patients with high LDH levels respond poorly to immunotherapy and other anticancer therapies such as chemotherapy and targeted therapy (159, 161, 162). Thus, targeting the lactate metabolic pathway in immune cells can overcome immune cell dysfunction in the TME. For example, suppressing LDH activity in macrophages can reprogram M2 phenotype to M1 phenotype (163). Besides, deletion of LDH-A in myeloid cells triggers antitumor immunity in the K-Ras murine lung carcinoma model (164). Likewise, blocking LDH in CD8+ T cells enhances adoptive T cell therapy (70). Genetic disruption or silencing of LDHA and LDHB in tumor cells inhibits tumor growth (165). Thus, targeting lactate metabolism changes lactate levels in the tumor microenvironment and can enhance antitumor immune responses. These targeting strategies collectively provide attractive angles for immunotherapy but warrant a better understanding of the actions of lactate on immune cells under steady-state and inflammatory conditions.

SUMMARY

Although lactate was initially recognized as a waste product of cellular metabolism, research over the past decade has revealed a fundamental role for this metabolite in shaping the function of the immune cells. Besides, as evidenced from the discussion above,
lactate in the tissue microenvironment programs APCs and other immune cells to regulate the balance between regulatory and inflammatory responses. Though moderate inflammation is essential to mount normal immune responses, uncontrolled, chronic, and excessive inflammation leads to allergic and autoimmune diseases. Lactate exhibits an inflammatory or anti-inflammatory role depending on its effects on immune cell type and disease type. Furthermore, lactate signaling in immune cells could be a critical pathway that links metabolism and immunity. While it is clear that both extracellular and intracellular lactate can program DCs and macrophages to induce robust regulatory immune responses, several important questions remain. For example, how do the lactate signaling pathways regulate adaptive immune responses under homeostatic conditions, inflammation, and cancer?; What are the downstream mediators of the lactate-GPR81/GPR132 pathway?; What role do receptor-dependent and independent lactate signaling play in regulating immunity versus tolerance?; How do the lactate act in concert with other signaling pathways in shaping anti-inflammatory and inflammatory immune responses?; and finally, the question of whether persistent chronic infections such as HIV, HCV or TB exploit the lactate-mediated signaling pathways and, if so, whether blocking this pathway would enhance the immune response is unknown. Addressing these questions will guide the rational design of therapeutic vaccines to reprogram the innate and adaptive immune system towards autoimmune disease tolerance or enhance immune responses against cancer and chronic infections.

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IM, PP, MT and SM have performed bibliographic researches and drafted the manuscript. All authors contributed to the article and approved the submitted version.

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27. Wanders D, Graff EC, Judd RL. Effects of High Fat Diet on GPR109A and GPR81 Gene Expression. *Biochim Biophys Acta Mol Basis Dis* (2019) 1850:1423. 10.1016/j.bbadis.2019.05.001

28. Morland C, Lauritzen KH, Pouchades M, Holm-Hansen S, Andersson K, Ranganathan P, Shanmugam A, Swafford D, Suryawanshi A, Bhattacharjee P, Hussein MS, et al. GPR81, a Cell-Surface Receptor for Lactate, Regulates Intestinal Homeostasis and Protects Mice From Experimental Colitis. *J Immunol* (2018) 200:1781–9. 10.4049/jimmunol.1700604

29. Errea A, Cayet D, Marchetti P, Tang C, Kluza J, Offermanns S, et al. Lactate Sensing of Lactate Mediates Tumor-Macrophage Interplay to Promote Breast Cancer Metastasis. *Cancers (Basel)* (2019) 11:53190. 10.3390/cancers11053190

30. Ranganathan P, Shanmugam A, Ahmad S, et al. Homeostatic PPARalpha Signaling Limits Inflammatory Agents. *Br J Pharmacol* (2015) 173:1841–50. 10.1111/bjp.13246

31. Xie Q, Zhu Z, He Y, Zhang Z, Zhang Y, Wang Y, et al. A Lactate-Induced LPS-Suppression of Secretion by Human Plasmacytoid Dendritic Cells. *Cell Mol Bioeng* (2019) 12:265. 10.1007/s12195-018-0555-y

32. Brown TP, Bhattacharjee P, Ramachandran S, Sivaprakasam S, Ristic B, Gjedde A, et al. The Lactate Receptor, G-protein-coupled Receptor 81 (GPR81), is Expressed in Human Brain and Regulates Neuronal Function. *J Neurosci Res* (2013) 91:10455–55. 10.1002/jn.24587

33. Zeng Z, Mukherjee A, Varghese AP, Yang X, Chen S, Zhang H. Roles of G Protein-Coupled Receptors in Inflammatory Bowel Disease. *World J Gastroenterol* (2020) 26:1242–61. 10.3748/wjg.v26.i2.1242

34. Wahli W, Michalik L. Ppars at the Crossroads of Lipid Signaling and Genomic Transcription. *Nature* (2013) 504:214–22. 10.1038/nature12977

35. Manoharan I, Suryawanshi A, Hong Y, Ranganathan P, Shanmugam A, Ahmad S, et al. Homeostatic PPARalpha Signaling Limits Inflammation and Metabolic Reprogramming in Murine Macrophages in a GPR81-Independent Manner. *PloS One* (2016) 11: e0163694. 10.1371/journal.pone.0163694

36. Brown TP, Bhattacharjee P, Ramachandran S, Sivaprakasam S, Ristic B, Gjedde A, et al. The Lactate Receptor, G-protein-coupled Receptor 81 (GPR81), is Expressed in Human Brain and Regulates Neuronal Function. *J Neurosci Res* (2013) 91:10455–55. 10.1002/jn.24587

37. Chen J, Yao Y, Gong C, Yu F, Su S, Chen J, et al. CCL18 From Tumor-Associated Macrophages Promotes Breast Cancer Metastasis Via PITPNM3. *Cancer Cell* (2011) 19:54–55. 10.1016/j.ccr.2011.02.006

38. Wang S, Ciszewski WM, Kania KD, L- and D-lactate Enhance DNA Repair and Modulate the Resistance of Cervical Carcinoma Cells to Anticancer Drugs Via Histone Deacetylase Inhibition and Hydroxyacrylic Acid Receptor 1 Activation. *Br J Pharmacol* (2013) 168:1216–33. 10.1111/bjph.12144

39. Gottfried E, Kunz-Schughart LA, Ebner S, Mueller-Klieser W, Hoves S, Agathokleous TM, et al. Downregulation of Membrane Trafﬁcking Proteins and Lactate Converting Dehydrogenase Lactate Dehydrogenase Limits Lactate Conversion and Dendritic Cell Function. *Biochem Pharmacol* (2015) 91. doi: 10.1016/j.bcp.2014.11.008

40. Samuvel DJ, Sundararaj KP, Stafetta F, Guarnaccia C, Ruiz-Moreno JS, Opitz B, et al. Lactate Promotes PGE2 Synthesis of YAP and NF-kappaB Activation Via GPR81-Mediated Signaling. *J Biol Chem* (2009) 284:26385–93. 10.1074/jbc.M109.04741

41. Chen P, Zuo H, Xiong H, Kolar MJ, Chu Q, Saghatelian A, et al. Gpr132 Boosts TLR4 Signaling and NF-kappaB Pathway-Mediated Gene Transcription of YAP and NF-kappaB Activation Via GPR81-Mediated Signaling. *J Biol Chem* (2009) 284:26385–93. 10.1074/jbc.M109.04741

42. Bach K, Chevallier M, Kunz-Schughart LA, Ebner S, Mueller-Klieser W, Hoves S, et al. Downregulation of Membrane Trafﬁcking Proteins and Lactate Converting Dehydrogenase Lactate Dehydrogenase Limits Lactate Conversion and Dendritic Cell Function. *Biochem Pharmacol* (2015) 91. doi: 10.1016/j.bcp.2014.11.008

43. Manoharan I, Suryawanshi A, Ahmad S, et al. Homeostatic PPARalpha Signaling Limits Inflammatory Responses to Commensal Microbiota in the Intestine. *J Immunol* (2016) 196:4739–49. 10.4049/jimmunol.1501489

44. Chen J, Yao Y, Gong C, Yu F, Su S, Chen J, et al. CCL18 From Tumor-Associated Macrophages Promotes Breast Cancer Metastasis Via PITPNM3. *Cancer Cell* (2011) 19:54–55. 10.1016/j.ccr.2011.02.006

45. Radu CG, Yang LV, Riedinger M, Au M, Witte ON. T Cell Chemotaxis to Lysophosphatidylcholine Through the G2A Receptor. *Proc Natl Acad Sci USA* (2002) 99:10184–9. 10.1073/pnas.1614035114
Manoharan et al. Lactate Signaling in DC and Macrophages

Cytokine Production of Human Primary BMDMs and Monocytes. Front Immunol (2018) 9:2564. doi: 10.3389/fimmu.2018.02564
67. Comito G, Iscaro A, Sacci M, Morandi A, Ippolito L, Parri M, et al. Lactate Modulates CD4(+)-T Cell Polarization and Induces an Immunosuppressive Environment, Which Sustains Prostate Carcinoma Progression Via TLR8/miR21 Axis. Oncogene (2019) 38:3681–95. doi: 10.1038/s41388-019-0688-7
68. Quinn WJ3rd, Jiao J, TeSlaa T, Stadniklic J, Wang Z, Wang L, et al. Lactate Limits T Cell Proliferation Via the NAD(H) Redox State. Cell Rep (2020) 33:108500. doi: 10.1016/j.celrep.2020.108500
69. Wen J, Cheng S, Zhang Y, Wang R, Xu J, Ling Z, et al. Lactate Anions Participate in T Cell Cytokine Production and Function. Sci China Life Sci (2021) 64:534–47. doi: 10.1007/s11427-020-1887-7
70. Hermans D, Gautam S, Garcia-Canaveras JC, Gromer D, Mitra S, Spolski R, et al. Lactate Dehydrogenase Inhibition Synergizes With IL-21 to Promote CD8(+) T Stemness and Antitumor Immunity. Proc Natl Acad Sci USA (2020) 117:6047–55. doi: 10.1073/pnas.1920411117
71. Brand A, Singer K, Koehl GE, Kolitzus M, Schoenhammer G, Thiel A, et al. Ldh-A-Associated Lactic Acid Production Blunts Tumor Immunosurveillance by T and NK Cells. Cell Metab (2016) 24:657–71. doi: 10.1016/j.cmet.2016.08.011
72. Watson MJ, Vignali PDA, Mullett SJ, Overacre-Delgoffe AE, Peralta RM, Grebinski S, et al. Metabolic Support of Tumour-Infiltrating Regulatory T Cells by Lactic Acid. Nature (2021) 591:645–51. doi: 10.1038/s41586-020-03045-2
73. Haas R, Smith J, Rocher-Ros V, Nadkarni S, Montero-Melendez T, D’Acquisto F, et al. Lactate Metabolites Regulate Membrane and Pro-inflammatory Circuits in Control of T Cell Migration and Effector Functions. PloS Biol (2015) 13:e1002202. doi: 10.1371/journal.pbi.1002202
74. Pucino V, Certo M, Bulusu V, Cucchi D, Goldmann K, Pontarini E, et al. Lactate Buildup at the Site of Chronic Inflammation Promotes Disease by Inducing CD4(+) T Cell Metabolic Rewiring. Cell Metab (2019) 30:1055–1074 e8. doi: 10.1016/j.cmet.2019.10.004
75. Renner K, Bruss C, Schnell A, Koehl G, Becker HM, Fante M, et al. Restricting Glycolysis Preserves T Cell Effector Functions and Augments Checkpoint Therapy. Cell Rep (2019) 29:135–50. doi: 10.1016/j.celrep.2019.06.068
76. Fischer K, Hoffmann P, Voellk S, Meidenbauer N, Ammer J, Edinger M, et al. Inhibitory Effect of Tumor Cell-Derived Lactic Acid on Human T Cells. Blood (2007) 109:3812–9. doi: 10.1182/blood-2006-07-035972
77. Li B, Yang Q, Li Z, Xu Z, Sun S, Wu Q, et al. Expression of Monocarboxylate Transporter 1 in Immunosuppressive Macrophages Is Associated With the Poor Prognosis in Breast Cancer. Front Oncol (2020) 10:574787. doi: 10.3389/fonc.2020.574787
78. Zhang L, Li S. Lactic Acid Promotes Macrophage Polarization Through MCT-HIF1alpha Signaling in Gastric Cancer. Cell Death Dis (2020) 11:3888. doi: 10.1038/s41419-020-02658-7
79. Cheng B, Cai Y, Li X, Cheng J, Chen J, et al. Lactate Protects Macrophages From gp130-STAT3 Signaling via a Distinct Mechanism. Sci Immunol (2018) 3:eaav900. doi: 10.1126/sciimmunol.aav900
80. Wang L, He HW, Xing QZ, Tang B, Zhou X. Lactate Induces Alternative Polarization (M2) of Macrophages Under Lipo polysaccharide Stimulation In Vitro Through G-protein Coupled Receptor 81. (Engl) (2020) 133:1761–3. doi: 10.1097/CMA.0000000000000955
81. Bottcher JP, Reis ESC. The Role of Type 1 Conventional Dendritic Cells in Cancer Immunity. Trends Cancer (2018) 4:784–92. doi: 10.1016/j.trecan.2018.09.001
82. Cancil JC, Crozat K, Dalod M, Mattuiz R. Are Conventional Type 1 Dendritic Cells Critical for Protective Antitumor Immunity and How? Front Immunol (2019) 10:959. doi: 10.3389/fimmu.2019.00959
83. Gardner A, Ruffell B. Dendritic Cells and Cancer Immunity. Trends Immunol (2016) 37:855–65. doi: 10.1016/j.it.2016.09.006
84. Roberts EW, Broz ML, Binnewies M, Headley MB, Nelson AE, Wolf DM, et al. Critical Role for CD103(+)CD11b(+) Dendritic Cells Bearing CCR7 for Tumor Antigen Trafficking and Priming of T Cell Immunity in Melanoma. Cancer Cell (2016) 30:324–36. doi: 10.1016/j.ccell.2016.06.003
85. Salmon H, Idoyaga J, Rahman A, Leboeuf M, Remark R, Jordan S, et al. Expansion and Activation of CD103(+) Dendritic Cell Progenitors at the Tumor Site Enhances Tumor Responses to Therapeutic Pd-L1 and BRAF Inhibition. Immunity (2016) 44:924–38. doi: 10.1016/j.immuni.2016.03.012
86. Engelhardt JJ, Boldaiprer B, Beemiller P, Pandurangi P, Sorensen C, Werb Z, et al. Marginating Dendritic Cells of the Tumor Microenvironment Cross-Present Tumor Antigens and Stably Engage Tumor-Specific T Cells. Cancer Cell (2012) 21:402–17. doi: 10.1016/j.ccr.2012.01.008
144. Payen VL, Mina E, Van Hee VF, Porporato PE, Sonveaux P. Monocarboxylate Transporters in Cancer. Mol Metab (2020) 33:48–66.

145. Beloueche Babari M, Casals Galobart T, Delgado-Goni T, Wantuch S, Parkes HG, Tandy D, et al. Monocarboxylate Transporter 1 blockade with AZD3965 inhibits lipid biosynthesis and increases tumour immune cell infiltration. Br J Cancer (2020) 122:895–903. doi: 10.1038/s41416-019-0717-x

146. Fisel P, Schaefele E, Schwab M. Clinical and functional relevance of the monocarboxylate transporter family in disease pathophysiology and drug therapy. Clin Transl Sci (2018) 11:352–64. doi: 10.1111/cts.12551

147. Noble RA, Bell N, Blair H, Sikka A, Thomas H, Phillips N, et al. Inhibition of monocarboxylate transporter 1 by AZD3965 as a novel therapeutic approach for diffuse large B-cell lymphoma and Burkitt lymphoma. Haematologica (2017) 102:1247–57. doi: 10.3324/haematol.2016.163030

148. Benjamin D, Robay D, Hindupur SK, Pohlmann J, Colombi M, El-Shemerly A, et al. Inhibition of the lactate transporters MCT1 and MCT4 is synthetic lethal with metformin due to NAD+ depletion in cancer cells. Cell Rep (2018) 25:3047–58.e4. doi: 10.1016/j.celrep.2018.11.043

149. Lee JY, Lee I, Chang WJ, Ahn SM, Lim SH, Kim HS, et al. MCT4 as a potential therapeutic target for metastatic gastric cancer with peritoneal carcinomatosis. Oncotarget (2016) 7:43489–503. doi: 10.18632/oncotarget.9523

150. Guan X, Bryniarski MA, Morris ME. In vitro and in vivo efficacy of the monocarboxylate transporter 1 inhibitor Ar-C155858 in the murine 4T1 breast cancer tumor model. AAPS J (2018) 21:3. doi: 10.1208/s12248-018-0261-2

151. Murray CM, Hutchinson R, Bantick JR, Belfield GP, Benjamin AD, Brazena D, et al. Monocarboxylate transporter MCT1 is a target for immunosuppression. Nat Chem Biol (2005) 1:371–6. doi: 10.1038/nchembio74

152. Fuji W, Kawahito Y, Nagahara H, Kukida Y, Seno T, Yamamoto A, et al. Monocarboxylate transporter 4, associated with the acidification of synovial fluid, is a novel therapeutic target for inflammatory arthritis. Arthritis Rheumatol (2015) 67:2888–96. doi: 10.1002/art.39270

153. Pucino V, Cucchi D, Mauro C. Monocarboxylate Transporters in Cancer. Expert Opin Ther Targets (2018) 22:735–43. doi: 10.1080/14728228.2018.151796

154. Yurchenko V, Constant S, Eisenmesser E, Bukrinskyy M. Cyclophilin CD147 Interactions: A New Target for Anti-Inflammatory Therapeutics. Clin Exp Immunol (2010) 160:305–17. doi: 10.1111/j.1365-2249.2010.04115.x

155. Zhu X, Song Z, Zhang S, Nanda A, Li G. CD147: A Novel Modulator of Inflammatory and Immune Disorders. Curr Med Chem (2014) 21:2138–45. doi: 10.2174/0929867321666131227163352

156. Le Froch R, Chiche J, Marchig I, Naiken T, Ilc K, Murray CM, et al. CD147 subunit of Lactate/H+ symporters MCT1 and Hypoxia-inducible MCT4 is critical for energetics and growth of glycolytic tumors. Proc Natl Acad Sci USA (2011) 108:16663–8. doi: 10.1073/pnas.1016213108

157. Baba M, Inoue M, Itoh K, Nishizawa Y. Blocking CD147 Induces Cell Death in Cancer Cells Through Impairment of Glycolytic Energy Metabolism. Biochem Biophys Res Commun (2008) 374:111–6. doi: 10.1016/j.bbrc.2008.06.122

158. Schneiderhan W, Scheler M, Holzmann KH, Marx M, Gschwend JE, Buchholz M, et al. CD147 silencing inhibits lactate transport and reduces malignant potential of pancreatic cancer cells in vitro and in vivo. Gut (2009) 58:1391–8. doi: 10.1136/gut.2009.181412

159. Van Wilpe S, Koornstra R, Den Brok M, De Groot JW, Blank C, De Vries J, et al. Lactate dehydrogenase: A marker of diminished antitumor immunity. Oncoimmunology (2020) 9:1731942. doi: 10.1080/2162402X.2020.1731942

160. Husain Z, Huang Y, Seth P, Sukhmatte VP. Tumor-Derived Lactate Modifies Antitumor Immune Response: Effect on Myeloid-Derived Suppressor Cells and NK Cells. J Immunol (2013) 191:1486–95. doi: 10.4049/jimmunol.1202702

161. Hodi FS, Chariot-Sileni V, Gonzalez R, Grob JJ, Rutkowski P, Cowey CL, et al. Nivolumab Plus Ipilimumab or Nivolumab Alone Versus Ipilimumab Alone in Advanced Melanoma (CheckMate 067): 4-year outcomes of a multicentre, randomised, phase 3 trial. Lancet Oncol (2018) 19:1480–92. doi: 10.1016/S1470-2045(18)30700-9

162. Long GV, Grob JJ, Nathan GP, Benjamin AD, Brazma D, et al. Factors Predictive of Response, Disease Progression, and Overall Survival After Dabrafenib and Trametinib Combination Treatment: A Pooled Analysis of Individual Patient Data From Randomised Trials. Lancet Oncol (2016) 17:1743–54. doi: 10.1016/S1470-2045(16)30578-2

163. Song TJ, Kim A, Kim GT, Yu HY, Lee ES, Park MJ, et al. Inhibition of Lactate Dehydrogenase A Suppresses Inflammatory Response in RAW 264.7 Macrophages Mol Med Rep (2019) 19:629–37. doi: 10.3892/mmr.2018.9678

164. Seth P, Czuzmadia E, Hedblom A, Vuerich M, Xie H, Li M, et al. Deletion of Lactate Dehydrogenase-a in Myeloid Cells Triggers Antitumor Immunity. Cancer Res (2017) 77:3632–43. doi: 10.1158/0008-5472.CAN-16-2938

165. Le A, Cooper CR, Gouw AM, Dinavahi R, Maitra A, Deck LM, et al. Inhibition of Lactate Dehydrogenase A Induces Oxidative Stress and Inhibits Tumor Progression. Proc Natl Acad Sci USA (2010) 107:2037–42. doi: 10.1073/pnas.0914433107

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