Determination of Antioxidant Properties and Antimicrobial Activity of Vinyl Phenolic Compounds Extracted from *Saccharomyces cerevisiae* Against Uropathogenic Bacteria

MUAAZAM SHERIFF MAQBUL¹, AEJAZ A. KHAN², TASNEEM MOHAMMED², S. M. SHAKEEL IQUBAL²*, IBRAHIM AHMED SHAIKH³, UDAY M. MUDDAPUR⁴, GOUSE BASHA SHEIK⁵, S. K. SINGH⁶, MOHAMMED SHAHID HUSSAIN⁷ and MOHAMMED GAMAL⁸,⁹

¹Department of Microbiology and Immunology, Ibn Sina National College for Medical Studies, Jeddah, Kingdom of Saudi Arabia.
²Department of General Science, Ibn Sina National College for Medical Studies, Jeddah, Kingdom of Saudi Arabia.
³Department of Pharmacology, College of Pharmacy, Najran University, Najran, Saudi Arabia.
⁴Department of Biotechnology, KLE Technological University, BVB Campus, Hubballi, 580031, India.
⁵Department of Microbiology, College of Applied Medical Sciences, Ad-Dawadmi, Shaqra University, Saudi Arabia.
⁶Department of Chemistry, GGV (Central University), Bilaspur (C.G) – 495009, India.
⁷Department of Orthodontics and Dentofacial Orthopedics, M. A. Rangoonwala Dental College, Pune, Maharashtra, India.
⁸Pharmaceutical Chemistry Department, Pharmacy college, Jouf University, P.O. Box 2014, Sakaka, Aljouf, Kingdom of Saudi Arabia.
⁹Pharmaceutical Analytical Chemistry Department, Faculty of Pharmacy, Beni-Suef University, Alshaheed Shehata Ahmed Hegazy St., 62574, Beni-Suef, Egypt.

*Corresponding author E-mail: shakeeliqubal@gmail.com

http://dx.doi.org/10.13005/ojc/360104

(Received: November 25, 2019; Accepted: January 05, 2020)

**ABSTRACT**

The aim of research study of determining the vinyl phenolic compounds with antioxidant properties and antimicrobial activity using acetone and methanol extracts of *Saccharomyces cerevisiae* (*S. cerevisiae*). The HPLC-UV technique was employed for the identification of the vinyl phenolic compounds and Ferric and Ferrous reducing anti-oxidant power assay along with radical scavenging methods were applied to determine the anti-oxidant properties of the yeast extract. The biochemical tests showed the presence of alkaloids, reducing sugars, steroid, proteins, phenol, cardiac glycosides. Further antimicrobial properties of the yeast extract using Kirby-Bauer disc diffusion method indicates, *E.coli* exhibits the best susceptibility towards the yeast extract. The antimicrobial susceptibility was excellent for all the isolated uropathogens when compared with the standard antibiotics. The metabolites produced by the yeast exhibits vital pharmaceutical important substances such as analgesic, antipyretic, anti-proliferative and antimicrobial properties. This study is a small attempt towards a larger future to serve the mankind with natural remedies.

**Keywords:** Vinyl Phenolic compound, Antioxidant Properties, Antimicrobial Activity, *Saccharomyces cerevisiae*, Uropathogenic Bacteria.
INTRODUCTION

The major objective of this work is to determine the vinyl phenolic compounds with antioxidant properties of this yeast by acetone and methanol extracts of S. cerevisiae and determine the antimicrobial activity. The HPLC-UV technique was employed for the identification of the vinyl phenolic compounds present in the yeast extract. The Ferri and Ferrous reducing anti-oxidant power assay along with radical scavenging method using 1,1-diphenyl-2-picryl hydrazyl were applied to determine the anti-oxidant properties of the yeast extract. The biochemical properties of the yeast extract were determined by different chemical methodologies. The antimicrobial properties of the yeast extract was determined by the standard antibiotic sensitivity test methodology designed by the Kirby and Bauer. The reactive oxygen species generally gets produced by the cellular metabolism of aerobic bacteria in which the molecular oxygen plays a vital role. They are contributors for the signal transduction of various cellular physiological processes for the human. The diseases such as cardiovascular, Alzheimer, Parkinson, Rheumatoid arthritis, etc. can be caused due to the excess production of reactive oxygen species beyond the threshold antioxidant capacity limit of our biological system which can lead to oxidative stress. The oxidative stress can be minimized by restoring the balance between the oxidant and antioxidant in the human system for the proper function of the physiological system. The administration of synthetic antioxidant substances can be a remedy but also results in harmful side effects which can affect the human organs due to its residual effect. In the recent studies the harmful effect of the synthetic chemical compounds on humans was once again debated and hence there is a need for the alternate solution. The alternative solutions should be able to restore the balance between the oxidant and antioxidant in the human system for the proper function of the physiological system and at the same time it should not cause any negative harmful side effects like that of the synthetic compounds. The best solution to overcome is to rediscover the nature blessed natural resources used by the ancestors to diagnose various dangerous diseases. The natural products should also be considered only if it is as effective as the synthetic chemical substances. The commonly available brewer’s yeast used in the brewing industries proved to be one of the best natural substance to overcome the synthetic chemical substances and is much more effective. This study is intended to study this yeast’s potential reactive oxygen capacity. The brewer’s yeast employed to our study is S. cerevisiae. This yeast possess vinyl phenol compounds 4-vinylphenol and 4-vinylguaiacol in 1:1 ratio which promotes it to be one of the best biopharmaceutical substance. The enzymatic activity of S. cerevisiae is excellent with substituted cinnamatecarboxy-lyase which converts coumaric and ferulic acids into vinylphenols by its capability of transforming non-oxidative decarboxylation of phenolic acid. This endo-cellular activity of this yeast is a potential source which can be used for the prophylaxis of oxidative stress related disorders. The metabolites produced by the yeast exhibits vital pharmaceutical important substances such as analgesic, antipyretic, anti-proliferative and cytotoxic antimicrobial properties. The antimicrobial property of the yeast extract was experimented against uropathogenic bacterial infections from isolated specimens collected, purified and identified from the urinary tract infected patients.

MATERIALS AND METHODS

Reagents

Chemicals like methanol, DPPH, acetone, FeCl₃, Ninhydrin reagent, Fehlings, Benedicts, Salkowski’s reagents were procured from Sigma-Aldrich chemicals (USA). Microbiological selective isolation culture media along with biochemical identification reagents and standard antimicrobial discs were employed. All other chemicals were analytical grade.

Yeast collection and Identification

A purified and isolated standard strain sample of the yeast S. cerevisiae from one of the ATCC (American Type Collection Centre) specimen collection centre in India and was used for this experiment. The confirmation identification for the pure culture of the yeast was verified by employing various in vitro fungal identification technique.

Preparation of the yeast extracts

10 g of the dried yeast samples were sequentially extracted with 250 mL of acetone and methanol at 37°C for 24 hours. A rotatory evaporator was employed to dry the concentrated
extract under reduced pressure with the temperature of 40°C and stored in the sterile test tubes at -20°C. The yield of respective yeast extracts were estimated as \( \text{Yield(\%)} = \left( \frac{\text{dry weight of extract}}{\text{dry weight of samples}} \right) \times 100 \)

**Identification of the vinyl phenolic compound in the yeast**

Based on Huneck and Yoshimura the identification of the vinyl phenolic compounds present in the yeast *S. cerevisiae* were performed by HPLC-UV technique analytical technique. The stored dried yeast extracts were dissolved in 500 µL of acetone and analyzed by using Agilant Technologies, 1200 Series HPLC instrument with C18 column 25cm x 4.6mm, 10µm with UV spectrophotometric detector.

**Determination of anti-oxidant properties in the yeast**

1) Based on the Oyaizu Ferric and Ferrous Reducing-Antioxidant Power (FRAP) assay methodology the anti-oxidant properties of yeast extract of *S. cerevisiae* was determined by the reducing powers of the dried yeast extracts by using various concentrations of the extract ranging from 50 to 1000 µg/mL diluted with saline buffer 0.2 M, pH 6.6 & 2.5 mL of ferric chloride (1%) and ferrous solution (1%) respectively and incubated at 50°C for 30 minute. After incubation period 2.5 mL of 10% trichloroacetic acid (TCA) was added to the mixture to stop the reaction. The mixture was centrifugated at 3000 g for 10 min and 2.5 mL distillated water added to 25 mL of the supernatant with 0.5 mL FeCl₃ (0.1%) and then the optical density was observed at 700 nm with an American made UNICO spectrophotometer which determined that the higher absorbance of the reaction mixture which indicates greater reducing power of the yeast. Suitable positive and negative controls were used for the test. The test was repeated for the five times and the mean value was recorded for the accuracy of the results.

2) Based on Kosanic *et al.*, radical scavenging methodology using 1,1-diphenyl-2-picryl hydrazyl, the anti-oxidant properties of the yeast extract of *S. cerevisiae* to scavenge DPPH free radicals was estimated by the reduction of the reaction color between DPPH solution and sample extracts was determined by adding 2 mL of 0.12 mM solution DPPH in methanol to the 1 mL of various concentrations ranging from 50 - 1000 µg/mL of the yeast extract respectively. The mixture was incubated at 37°C for 30 min and the absorbance of the reaction mixture was measured at 517 nm with an American made UNICO spectrophotometer to determine the radical scavenging activity of the yeast extract. Suitable positive and negative controls were used for the test. The test was repeated for the five times and the mean value was recorded for the accuracy of the results.

**Bio-chemical analysis of the yeast extract**

The biochemical properties of the yeast extract of *S. cerevisiae* was determined by Mayer's test, Fehling's test, Salkowski's test and Ninhydrin test, Ferric Chloride test, Libermann Burchard's Test, Benedict's Test, Keller-kilani test and ammonia test.

**Antimicrobial Property of the yeast extract**

The antimicrobial activity of the yeast extract was performed against the clinically isolated uropathogens from the urinary tract infected persons.

**Isolation and Identification of uropathogenic Specimen**

The clinical urinary sample was collected from the suspected urinary tract infected patients and inoculated on the selective media Cystine Lactose Electrolyte Deficient Agar with pH indicator bromothymol blue. The inoculated plates were incubated at 37°C for 24 h and biochemical reaction test were done for all the isolated uropathogens from the isolated colonies by using IMVIC test and results were observed respectively.

**Antibiotic Susceptibility testing by Kirby-Bauer Disc Diffusion Method**

Based on the standard antibiotic sensitivity methodology developed by Kirby and Bauer the antimicrobial activity of the yeast extract were determined by incorporating the standard antibiotic discs coated with the yeast extract on the various Mueller Hinton agar plates inoculated with different uropathogenic samples such as *E.coli*, *Klebsiella Sp.*, *Pseudomonas Sp.* and *Proteus Sp.* isolated from the clinical urine specimens respectively. The inoculated plates impregnated
with the antibiotic disc of yeast extract were incubated for 24 h at 37°C. The zone formation around the disc indicates the susceptibility of the organism and no zone formation indicates the resistance of the organism towards the antibiotic disc of the yeast extract.

RESULTS AND DISCUSSION

The yeast extract results were exemplary for all the tests performed. Based on Huneck and Yoshimura the identification of the vinyl phenolic compounds present in the *S. cerevisiae* were performed by the HPLC-UV analytical technique analysis of acetone and methanol extracts of the yeast extract was used to identify their major vinyl phenolic compounds Figure 1.

By comparing the retention times (tR) yeast extract were identified. The main compounds in acetone extract of yeast were physodalic acid (tR = 3.295 ± 0.012 min), atranorin (tR = 8.642 ± 0.020 min) and chloratranorin (tR = 11.918 ± 0.026 min). The most abundant substance in the acetone yeast extract was physodalic acid where as in methanol yeast extract only small amount of physodalic acid was observed in the chromatogram. It is evidenced in the chromatograms of ferric and ferrous reducing power assay of oxidant/antioxidant based on Oyaizu technique as shown in Figure 2.

The chromatogram showed evernic acid, (tR = 3.045 ± 0.029 min), usnic acid (tR = 7.854 ± 0.025 min), atranorin (tR = 8.236 ± 0.028 min) and chloratranorin (tR = 11.554 ± 0.028 min) are present in the acetone extracts. The most abundant compound in acetone extract is evernic acid while in methanol extract also evernic acid and usnic acid detected but the methyl lecanorate (tR = 1.515 ± 0.009 min) is the most abundant compound.

Based on Kosanic et al., radical scavenging methodology using 1,1-diphenyl-2-picryl hydrazyl, the anti-oxidant properties of the yeast extract of *S. cerevisiae* to scavenge DPPH free radicals was estimated by the reduction of the reaction color between DPPH solution and sample extracts was determined results showed a statistically significant data as the antioxidant activity was elevated from 2.01 ± 0.009% to 86.14 ± 0.013% in accordance with the increase of the concentration of the extracts from 50 to 1000 µg/mL of yeast extract Figure 3.

The acetone yeast extract with 1000 µg/mL showed largest DPPH radical scavenging activity: 86.14 ± 0.013%. According to test results the antioxidant activities obtained from the methanol yeast extracts were obtained but lower than that measured for the acetone yeast extract which showed the highest DPPH radical scavenging activity with an IC₅₀ = 240.220 ±15.165 µg/mL.

The biochemical properties analysis of the yeast *S. cerevisiae* extract showed the presence of alkaloids, reducing sugar, steroids, protein, phenol, glycosides, cardiac glycosides and amino acids as shown in Table 1.
Table 1: Biochemical properties of yeast

| Test                     | Observation                        | Result | Interpretation            |
|--------------------------|------------------------------------|--------|---------------------------|
| Mayer's test             | Formation of a creamy substance    | Positive | Presence of alkaloids.    |
| Fehling's test           | Brick red color formation          | Positive | Presence of reducing sugar|
| Salkowski’s test         | Reddish brown color formation      | Positive | Presence of steroids      |
| Ninhydrin test           | Violet color formation             | Positive | Presence of protein       |
| Ferric chloride test     | Blue black coloration              | Positive | Presence of Phenol        |
| Libermann Burchard’s test| Violet to bluish green color formation | Positive | Presence of Glycosides    |
| Benedict’s test          | Formation of orange red precipitate| Positive | Presence of reducing sugar|
| Keller-kilani test       | Brown ring at the interface        | Positive | Presence of Cardiac glycosides|
| Ammonia test             | Yellowish color formation          | Positive | Presence of Amino Acids   |

Further, collected specimens from the urinary tract infected persons were isolated by culturing on a selective media Cystine Lactose Electrolyte Deficient Agar with pH indicator Bromothymol blue showed Yellow lactose fermenting colonies for the *Klebsiella sp.* and *E.coli* whereas blue non lactose fermenting colonies for the *Proteus sp.* and *Pseudomonas sp.* The isolated organisms were confirmed using biochemical reactions of IMVIC and the result observed were tabulated (Table 2) and confirmed the identification of the pathogens.

Table 2: biochemical assessment of the isolated uropathogens

| Organism       | Indole   | Methyl red | Voges proskauer | Citrate | Urease | Oxidase |
|----------------|----------|------------|-----------------|---------|--------|---------|
| E.Coli         | pink ring| red color  | yellow color    | green color | yellow color | no purple color |
| Positive       | positive | negative   | positive        | negative| negative| negative |
| Negative       | negative | positive   | negative        | positive| positive| negative |
| Klebsiella sp. | yellow ring | yellow color | yellow color | blue color | pink color | no purple color |
| Positive       | negative | negative   | positive        | positive| positive| negative |
| Negative       | positive | negative   | positive        | positive| positive| positive |
| Pseudomonas sp.| yellow ring | yellow color | yellow color | blue color | yellow color | purple color |
| Positive       | negative | negative   | positive        | positive| negative| positive |
| Negative       | positive | negative   | positive        | negative| positive| negative |
| Proteus sp.    | yellow ring | red color  | yellow color    | blue color | yellow color | no purple color |
| Positive       | positive | negative   | positive        | positive| negative| negative |
| Negative       | negative | positive   | positive        | negative| positive| negative |

Then using the Kirby-Bauer disc diffusion method the antimicrobial susceptibility of the prepared yeast extract antibiotic disc were tested against the isolated uropathogens and the results were exemplary. *E.coli* shown the best susceptibility towards the yeast extract while the *Pseudomonas sp.* show the least susceptibility. The *Klebsiella sp.* and *Proteus sp.* shown a moderate susceptibility with the yeast extract antibiotic disc. The antimicrobial susceptibility was excellent for all the isolated uropathogens when compared with the standard antibiotics (Table 3).

Table 3: Antimicrobial susceptibility comparison of yeast extract with standard antibiotics for urinary tract infections

| Antimbiotics      | *E.coli* | *Klebsiella sp.* | *Pseudomonas sp.* | *Proteus sp.* |
|-------------------|----------|-----------------|------------------|--------------|
| Yeast Extract     | Sensitive 25mm | Sensitive 23mm | Sensitive 19mm | Sensitive 21mm |
| Trimethoprim      | Sensitive 21mm | Intermediate 15mm | Resistant 2mm | Intermediate 14mm |
| Sulfamethoxazole  | Sensitive 18mm | Sensitive 24mm | Resistant 4mm | Sensitive 19mm |
| Fosfomycin        | Sensitive 21mm | Sensitive 22mm | Sensitive 21mm | Sensitive 18mm |
| Nitrofurantoin    | Sensitive 26mm | Sensitive 21mm | Sensitive 20mm | Sensitive 18mm |
| Azithromycin      | Sensitive 24mm | Sensitive 19mm | Sensitive 22mm | Sensitive 22mm |
| Ceftriaxone       | Sensitive 19mm | Sensitive 24mm | Sensitive 20mm | Sensitive 24mm |
| Cephalexin        | Sensitive 20mm | Sensitive 21mm | Sensitive 18mm | Sensitive 21mm |
| Levofloxacin      | Intermediate 16mm | Resistant 8mm | Resistant 2mm | Resistant 6mm |
| Ciprofloxacin     | Sensitive 22mm | Sensitive 19mm | Intermediate 14mm | Intermediate 13mm |
CONCLUSION

The antioxidant properties along with the vinyl phenolic compounds of the yeast *Saccharomyces cerevisiae* shows that this solated yeast extract can be used as a potential anti synthetic substance for the treatment of many diseases not only just as an antibiotic but can also help to balance between the oxidant and antioxidant level in the body for the proper function of the physiological system in maintaining the proper body functions as it contains many essential biochemical nutrients required for our body to produce reactive oxygen species to eradicate many diseases. This study is a small attempt towards a larger future to serve the mankind with natural remedies.

ACKNOWLEDGEMENT

We would like to Acknowledge our family members for their constant support and encouragement.

Conflicts of interest

No conflict of interest.

REFERENCES

1. Kapoor SL., Kapoor LD. On the botany and distribution of ’pashanbhed’. *Sachitra Ayurved*, 1976, 28(12), 769–791.
2. Singh AP. A Short review on Didymocarpus-pedicellata: The Lithntriptic Ethnomedicine, *Ethnobotanical Leaflets*, 2007, 11, 73–75.
3. Mojab F., Kamalinejad M., Naysanch G., Hamid R. Phytochemical screening of some species of Iranian plants. *Iranian Journal of Pharmaceutical Research*, 2003, 2(2), 77–82.
4. Awika JM., Rooney LW., Wu X., Prior RL., Cisneros-Zevallos L. Screening methods to measure antioxidant activity of sorghum (Sorghum bicolor) and sorghum products. *Journal of Agricultural and Food Chemistry*, 2003, 51, 6657-6662.
5. Huneck S., Yoshimura I. Identification of Lichen Substances, 1st ed, Berlin, Springer–Verlag., 1996.
6. Oyaizu M. Studies of products browning reaction: antioxidative activity of products of browning reaction prepared from glucosamine. *Jpn J Nutr.*, 1986, 44, 307.
7. Muazzam SM., AlHasel HMB., Majid DH., Momen TN., AlHazmi HAM., AlJeddani FMS., AlMalaki RTW., Khan AA., Iqbal SMS. Chemical Analysis (GC-FID-MS) and Antimicrobial Activity of *Parmotrema perlatum* Essential Oil Against Clinical Specimens. *Orient.J.Chem.*, 2019, 35(6), in press.
8. Gouse BS., Muazzam SM, Gokul SS, Ranjith MS. Isolation and characterization of actinomycetes from soil of ad-dawadmi, Saudi Arabia and screening their antibacterial activities. *Int J of Pharm and Pharmaceutical Sc.*, 2017, 9(10),267-279.
9. Huang D., Ou B., Prior RL. The chemistry behind antioxidant capacity assays. *Journal of Agricultural and Food Chemistry*, 2005, 53, 1841-1856.
10. Ben Salah M., Abderraba M., Tarhouni MR., Abdelmelek H. Effects of ultraviolet radiation on the kinetics of in vitro percutaneous absorption of lavender oil. *Int J Pharm*, 2009, 382, 33–38.
11. Poopal AC., Laxman RS. Studies on the biological reduction of chromate by Streptomyces griseous. *J Haz Mat.*, 2009, 169, 539-545.
12. Adams RP. Identification of Essential Oil Compone- nts by Gas Chromatography/ Mass Spectroscopy, 4th edition. Allured Publishing Corporation, Carol Stream, IL., 2007.
13. Kokate CK., Purohit AP., Gokhale SB. Pharmacognosy., 20th Edn, Nirali prakashan, Pune., 2002, 108-109.
14. Kaur G., Lone IA., Athar M. Protective effect of Didymocarpuspedicellata on ferric nitrolotriacetate (FeNTA) induced renal oxidative stress and hyperproliferative response, *Chemico biological interaction.*, 2007, 165(1), 33–44.
15. Rathore JS., Garg SK., Gupta SR. A chalcone and flavones from Didymocarpuspedicellata. *Phytochemistry.*, 1981, 20, 1755-1756.
16. Bisht CMS., Iqubal SMS., Khan AA., Tasneem M., Dawoud A., Gamal M., Singh SK., Asghar BH. Natural Products in Drug Discovery: Antibacterial and Antifungal Activity of Essential Oil of Compound Isolated from Senecio royleanus. J Pure Appl Microbio., 2019, 13(3), 1611-1617.

17. Arpin C., Quentin C., Grobost F., Cambau E., Robert J., Dubois V., Coulanje L., Andre C. Nationwide survey of extended-spectrum \( \beta \) lactamase producing Enterobacteriaceae in the French community setting. J Antimicrob Chemother., 2009, 63, 1205–1214.

18. Zhu M., Cui Y. Determination of 4-vinylgaiaicol and 4-vinylphenol in top-fermented wheat beers by isocratic high performance liquid chromatography with ultraviolet detector. Braz arch biol technol., 2013, 56(6), 1018-1023.

19. Dey TB., Kuhad RC. Enhanced production and extraction of phenolic compounds from wheat by solid-state fermentation with Rhizopus oryzae RCK2012. Biotechnol Rep., 2014, 4, 120-127.

20. Bagewadi ZK., Muddapur UM., Madiwal SS., Mulla SI., Khan AA. Biochemical and enzyme inhibitory attributes of methanolic leaf extract of Datura inoxia Mill. Environmental Sustainability., 2019, 2, 75–87.

21. Bahl CP., Seshadri TR. Pashanbhedi: drugs for urinary calculus, Eds., K.N. Udupa., 1978 7 7–98.