Functiona studies of novel CYP21A2 mutations detected in Norwegian patients with congenital adrenal hyperplasia

Ingeborg Brønstad1, Lars Breivik1, Paal Methlie1,2, Anette S B Wolff1, Eirik Bratland1, Ingrid Nermoen3, Kristian Løva˚s1,2 and Eystein S Husebye1,2

1Department of Clinical Science, University of Bergen, 5021 Bergen, Norway
2Department of Medicine, Haukeland University Hospital, Bergen, Norway
3Division of Medicine, Akershus University Hospital, Lørenskog, Norway

Abstract

In about 95% of cases, congenital adrenal hyperplasia (CAH) is caused by mutations in CYP21A2 gene encoding steroid 21-hydroxylase (21OH). Recently, we have reported four novel CYP21A2 variants in the Norwegian population of patients with CAH, of which p.L388R and p.E140K were associated with salt wasting (SW), p.P45L with simple virilising (SV) and p.V211M + p.V281L with SV to non-classical (NC) phenotypes. We aimed to characterise the novel variants functionally utilising a newly designed in vitro assay of 21OH enzyme activity and structural simulations and compare the results with clinical phenotypes. CYP21A2 mutations and variants were expressed in vitro. Enzyme activity was assayed by assessing the conversion of 17-hydroxyprogesterone to 11-deoxycortisol by liquid chromatography tandem mass spectroscopy. PyMOL 1.3 was used for structural simulations, and PolyPhen2 and PROVEAN for predicting the severity of the mutants. The CYP21A2 mutants, p.L388R and p.E140K, exhibited 1.1 and 11.3% of wt 21OH enzyme activity, respectively, in vitro. We could not detect any functional deficiency of the p.P45L variant in vitro; although prediction tools suggest p.P45L to be pathogenic. p.V211M displayed enzyme activity equivalent to the wt in vitro, which was supported by in silico analyses. We found good correlations between phenotype and the in vitro enzyme activities of the SW mutants, but not for the SV p.P45L variant. p.V211M might have a synergistic effect together with p.V281L, explaining a phenotype between SV and NC CAH.

Key Words

- CYP21A2
- congenital adrenal hyperplasia
- functional studies
- novel mutations

Introduction

The CYP21A2 gene encodes the enzyme steroid 21-hydroxylase (21OH), which is essential for steroid synthesis in the adrenal cortex. Mutations in CYP21A2 are the main cause of the autosomal recessive disorder congenital adrenal hyperplasia (CAH) (1). The classical forms of CAH are the salt wasting (SW) and simple virilising (SV) phenotypes with adrenocorticotropic hormone-driven excess of androgens leading to ambiguous genitalia in female newborns. The SW phenotype has complete or almost complete loss of enzyme activity, leading to lack of cortisol and aldosterone, whereas the SV displays various degrees of reduced enzyme activity...
resulting in cortisol insufficiency (1, 2). The non-classical (NC) CAH is milder and characterised by signs of hyperandrogenism postnatally and in adulthood, and associated with minor mutations (3).

The most common mutations in the CYP21A2 gene are derived from a non-functional pseudogene CYP21A1P (4). Both CYP21A2 and CYP21A1P are located in the RCCX module of the HLA class III locus of chromosome 6 and share about 98% sequence homology. In general, there is a high correlation between the CYP21A2 genotype and the CAH phenotype (5, 6). In addition to the pseudogene mutations, more than 160 other pathological variants of CYP21A2 have been reported (www.cypalleles.ki.se/cyp21.htm).

In the present study, we used a new in vitro assay of 21OH enzyme activity and molecular computer modelling to predict the phenotypes of recently detected variants (p.L388R, p.E140K, p.P45L and p.V211M) in the Norwegian population (7). We also checked a novel variant (p.A159T), detected in Norwegian patients with autoimmune Addison’s disease (AAD) and in control populations.

Subjects and methods

Subjects

We detected four novel variants: p.L388R, p.E140K, p.P45L and p.V211M, in a cohort of 59 patients diagnosed with CAH (7). Four unrelated female patients, each had one of the missense variants listed above, except the subject with p.V211M who also had a coupled p.V281L mutation on the same allele. All patients exhibited a deletion in the other allele (7). Clinical characteristics of the patients are listed in Table 1. The fifth variant, p.A159T, was found in heterozygosity in three of 370 patients with AAD and in one out of 330 Norwegian healthy subjects.

Genetic analyses of CYP21A2

Protocols for sequencing and copy number analyses of the CYP21A2 gene were described previously (7, 8, 9, 10, 11).

Mutagenesis

Stratagene’s QuikChange II Site Directed Mutagenesis Kit (Agilent Technologies, Santa Clara, CA, USA) was used to introduce point mutations in the pcDNA2 plasmid containing CYP21A2 cDNA (a kind gift from Prof. Holger Luthman, Department of Molecular Medicine, Karolinska Institute, Stockholm, Sweden (12)), corresponding to the

| Table 1 | Clinical description of the CAH patients with novel variants in CYP21A2. |
|---------|---------------------------------------------------|
| Variant | Clinical phenotype                                   | Age of diagnoses | Treatment                        |
| p.L388R | Salt wasting                                       | Newborn          | Mineralocorticoids               |
|         | Clitoris hypertrophy                               |                  | Glucocorticoids                 |
| p.E140K | Salt wasting                                       | 1 week old       | Mineralocorticoids               |
|         | Clitoris hypertrophy                               |                  | Glucocorticoids                 |
| p.P45L  | Simple virilising                                  | 4 years          | Mineralocorticoids               |
|         | Clitoris hypertrophy                               |                  | Glucocorticoids                 |
| p.V211M | Simple virilising/ non-classical CAH               | 21 years         | Glucocorticoids                 |
|         | Pubes growth and pronounced hirsutism from 7 years old |                  |                                 |

novel CYP21A2 variants (p.L388R, p.E140K, p.P45L, p.V211M and p.A159T). Three control variants of CYP21A2 were also generated: p.R102K (normal variant), p.I172N (SV phenotype) and p.G291R (SW phenotype). Primers used for mutagenesis are listed in Table 2. The mutagenesis was carried out according to the manufactures protocol, using 16 cycles with 5 min extension time. The mutagenesis was verified with direct sequencing, and plasmids from positive clones were purified by HiSpeed Plasmid Maxi Kit (Qiagen).

In vitro protein expression of 21OH mutants

Non-radioactive TNT Quick Coupled Transcription/Translation system (Promega) with Sp6 RNA polymerase was used for in vitro protein expression. Expression of all the 21OH variants was verified by western blot using a primary antibody against 21OH (c17, goat polyclonal IgG, Santa Cruz Biotechnology) and alkaline phosphatase-conjugated donkey anti-goat IgG secondary antibody (Santa Cruz Biotechnology).

Relative quantitation of 21OH from in vitro expression

ELISA plates (MaxiSorp, Nunc Thermo Scientific, Waltham, MA, USA) were coated over night with 10 μl of the in vitro expression product diluted in PBS, pH 7.4. Primary antibody against 21OH (Santa Cruz Biotechnology), together with the corresponding alkaline phosphatase-conjugated secondary antibody (Sigma) and the FAST pNPP detection system (Sigma), was used to determine the relative amount of 21OH in the reaction products.
The ELISA method was verified by selected reaction monitoring (SRM) against two signature peptides of 21OH (p.85–98, LQELDHELPGASSSR and p.317–333, WAD-FAGRPEPLTYK). Briefly, 5 μl of each version of 21OH reaction products were taken out for quantitative determination of 21OH. The lysates were purified on SDS–PAGE gel, and the bands corresponding to the molecular weight of wt 21OH (56 kDa) were cut out for trypsin treatment. Stable isotope-labelled synthetic peptides (N-terminal replacement of lysine ($^{13}$C$_6$, $^{15}$N$_2$) or arginine ($^{13}$C$_6$, $^{15}$N$_2$) respectively) were added in constant amount before purification, resulting in a mass difference of 8 or 10 Da to the corresponding endogenous peptide. A Q-Trap 5500 (AB SCIEX) connected to a Dionex Ultimate NCR-3500RS LC system was used for the SRM analyses. The SRM data were analysed using Skyline, version 2.1 (https://skyline.gs.washington.edu/), and the analyses. The SRM data were analysed using Skyline, version 2.1 (https://skyline.gs.washington.edu/), and the most abundant transition was used for quantification. The SRM analyses were carried out using standard procedures prepared by diluting stock solutions in a solution of 50% methanol and 0.1% formic acid in double-distilled (dd) H$_2$O (v/v). An internal standard (IS) solution (50 nM 17OHP-d$_8$ and 50 nM 11DOC-d$_2$ in methanol) was then added to the diluted samples in a 1:1 ratio. Working standards of 11DOC in the range of 0.61–150 nM were prepared by diluting stock solutions in a solution of 50% methanol and 0.1% formic acid in ddH$_2$O (v/v) and adding IS (1:1). Working standards and 11DOC produced from each enzymatic reaction were quantified by liquid chromatography coupled to tandem mass spectrometry (LC–MSMS) (14). The enzyme activity was calculated as the % ratio of 11DOC production between the 21OH mutants to the wt, adjusted for the quantity of 21OH wt and mutant in the enzymatic reactions.

**Structural simulations and *in silico* analyses**

The molecular graphic images were generated from a humanised version of bovine 21OH as template (15), kindly provided from Dr Shozeb Haider (Centre of Cancer Research and Cell Biology, Queen’s University of Belfast, Belfast, UK), using The PyMOL Molecular Graphics System, version 1.3 (Schrödinger, LLC, Portland, OR, USA). Variants were modelled using the Mutagenesis tool in PyMOL and distances between molecules measured using the Measurement tool. The severity of the variants was predicted by PolyPhen-2 v2.2.2r398 HumVar model (http://genetics.bwh.harvard.edu/pph2/) (16) and PROVEAN (http://provean.jcvi.org/index.php) (17) using default settings. Sequence alignments of 21OH from seven different species (GenBank: AAB59440.1 (*Homo sapiens*), AAA30487.1 (*Boa taurus*), ABC69211.1 (*Felis catus*), AAD05573.1 (*Rattus norvegicus*) and BAF63009.1 (*Gallus gallus*); NCBI reference sequence: XP_003311237.1 (*Pan troglodytes*) and NP_999598.1 (*Sus scrofa*)) were carried out by default settings using Clustal omega (http://www.ebi.ac.uk/Tools/msa/clustalo/) (18) in order to assess the evolutionary conservation of the mutated amino acids.

### Table 2

| Mutations | Sequences *(5′→3′)* |
|-----------|--------------------|
| p.P45L    | F: CAGGCCGCACCTCTGTGATCTATCTGTTGG |
|           | R: CAAAGCGCATGATCCAGGGGCTGGGCTGG |
| p.R102K   | F: CAAAGCTGTGCTTAAGACTACCCGCAGGS |
|           | R: GTCCGGTTAGTCTCTTTGACACCAAGCTG |
| p.E140K   | F: CAGGGAGCGCTGGTAAAAAGCATGGACCCAGG |
|           | R: CTCCTGGGTACGTTTCACTCAGGGTCCTAG |
| p.A159T   | F: GGACCCCTGTTGACATTGAGGAGAGGAG |
|           | R: CTCCTCAATGCGTGACAGGGGTGCC |
| p.I172N   | F: CACCTGCAAGCATCAACTGTACCTACCC |
|           | R: GGTGAGGTAACTGGATGCTGAGGGT |
| p.G291R   | F: CTCGAGTGGGACCTCTGTGACCAGGGCACTGAGACTGAGAC |
|           | R: GTTTGTCTGTTGCTACGTGGGCCCCGTAGCAGGAGTCCTCGAG |
| p.L388R   | F: GTCATCATCCAGAAGACAAAGGCCCCCACCTG |
|           | R: CAGGGGGGCTCGTTGCTCGGAATGATGAG |

**Enzyme activity assay**

The activity of the different 21OH constructs was estimated by quantitating 11-deoxycortisol (11DOC) using 17-hydroxyprogesterone (17OHP) as substrate. The enzyme reactions were set up mainly as described previously (13). Briefly, from each 21OH construct, 50 μl of the *in vitro* expression product were mixed with 15 pmol of recombinant human cytochrome P450 oxidoreductase (CYPOR, Sigma–Aldrich) and 25 μl of 300 μM 17OHP (in 10% methanol) on ice. Pre-warmed reaction buffer (50 mM potassium phosphate (pH 7.4), 20% (v/v) glycerol, 2 mM dithiothreitol, 0.1 mM EDTA and 0.0034% (v/v) Emulgen 913 (Karlan Research Products Corporation, Cottonwood, AZ, USA)) was then added to a total volume of 500 μl. After 60 s of pre-incubation at 37 °C, the enzymatic reaction was initiated by adding $5 \mu$l of 100 mM NADPH. The enzymatic reaction was carried out for 1 h at 37 °C, and then terminated by putting onto ice. Further, the enzyme reaction solution was diluted 1:10 or 1:100 in a solution of 50% methanol and 0.1% formic acid in double-distilled (dd) H$_2$O (v/v). An internal standard (IS) solution (50 nM 17OHP-d$_8$ and 50 nM 11DOC-d$_2$ in methanol) was then added to the diluted samples in a 1:1 ratio. Working standards of 11DOC in the range of 0.61–150 nM were prepared by diluting stock solutions in a solution of 50% methanol and 0.1% formic acid in ddH$_2$O (v/v) and adding IS (1:1). Working standards and 11DOC produced from each enzymatic reaction were quantified by liquid chromatography coupled to tandem mass spectrometry (LC–MSMS) (14). The enzyme activity was calculated as the % ratio of 11DOC production between the 21OH mutants to the wt, adjusted for the quantity of 21OH wt and mutant in the enzymatic reactions.
Statistical analyses

Statistical analyses were performed by the GraphPad Prism, version 5.02 Software (La Jolla, CA, USA). Correlations between the ELISA and SRM methods for quantifying the 21OH proteins produced were calculated by Pearson’s correlation test. Deviation of enzyme activity of the 21OH mutants compared with the wt 21OH was calculated by ANOVA combined with Dunnett’s multiple comparison tests using a significance level of P<0.05.

Results

Verification and relative quantitation of 21OH by ELISA and SRM methods

Western blot analysis of the samples from in vitro expression showed a 56-kDa band corresponding to the molecular weight of 21OH for all constructs (data not shown).

The ELISA method for quantitation of the mutated 21OH proteins correlated very well with the SRM method (P<0.0001, Pearson’s r=0.9206, and 95% CI (0.8016, 0.9695), n=19). Thus, the ELISA method was used for further quantitation of 21OH.

In vitro 21OH activity studies

The assay showed no significant reduction in the enzyme activity of the p.R102K normal variant relative to the wt (119.7%), whereas the p.I172N (SV phenotype) and p.G291R (SW phenotype) mutations showed significant (P<0.05) decline (1.6 and 0%, respectively, relative to wt (Table 3)). The novel CAH mutations, p.L388R and p.E140K, displayed significantly (P<0.05) reduced activity of 1.1 and 11.3% compared with wt respectively (Table 3). The other two novel CAH variants, p.P45L and p.V211M, were similar to the wt (105 and 99.5% respectively) as was p.A159T (126.6%) (Table 3).

Structural simulations

The structures of 21OH and the novel variants based on the humanised model (15) are illustrated in Fig. 1. For p.L388R, the shift from the hydrophobic leucine to a negatively charged arginine in position 388 will probably lead to a disruption in a hydrophobic pocket surrounding the heme group (Fig. 1a). This is a highly conserved region (Fig. 2), and the p.L388R mutation was predicted as pathologic (Table 3).

The p.E140 residue forms a salt bridge with the R444 residue, and the shift from negatively charged glutamic acid to positively charged lysine breaks the salt bridge (Fig. 1b). A negatively charged amino acid in this residue is highly conserved (Fig. 2); however, PolyPhen2 predicted this mutation as possible damaging, while PROVEAN predicted it as neutral (Table 3) in line with significant residual enzyme activity (see above).

p.P45 is located early in the A helix (Fig. 1c), which is a highly conserved region (Fig. 2). The proline-to-leucine shift in position 45 was predicted as pathologic by both PolyPhen2 and PROVEAN (Table 3), but in vitro enzyme activity was normal.

The p.V211 residue (Fig. 1d) may interact with L65 through hydrophobic interactions and is located in a semiconservative region (Fig. 2). The shift from valine to methionine was predicted as benign/neutral (Table 3) in line with normal enzyme activity.

The variant p.A159T (Fig. 1e) is located on the surface of the 21OH protein. Sequence alignment shows that the

| Mutant/variant | % Average enzyme activity (s.d.) | Phenotype | Predicted phenotype (score) |
|----------------|----------------------------------|-----------|-----------------------------|
| p.K102R        | 119.7 (22.5)                     | Normal    | Benign (0.001)               |
| p.A159T        | 126.6 (29.9)                     | Unknown   | Benign (0.023)               |
| p.V211M        | 99.5 (32.4)                      | SV/NC     | Benign (0.029)               |
| p.I172N        | 1.6 (0.8)                        | SV        | Probably damaging (0.998)    |
| p.P45L         | 105.0 (10.6)                     | SV        | Probably damaging (1.0)      |
| p.G291R        | 11.3 (2.4)                       | SW        | Possibly damaging (0.49)     |
| p.E140K        | 1.1 (0.6)                        | SW        | Probably damaging (0.986)    |
| p.L388R        | 0.0 (0.0)                        | SW        |                            |
| p.V211M        | 0.0 (0.0)                        | SW        |                            |
| p.E140K        | 11.3 (2.4)                       | SW        |                            |
| p.L388R        | 1.1 (0.6)                        | SW        |                            |
| p.A159T        | 126.6 (29.9)                     | Unknown   | Benign (0.023)               |
| p.V211M        | 0.0 (0.0)                        | SW        | Benign (0.029)               |
| p.I172N        | 1.6 (0.8)                        | SV        | Probably damaging (0.998)    |
| p.P45L         | 105.0 (10.6)                     | SV        | Probably damaging (1.0)      |
| p.G291R        | 11.3 (2.4)                       | SW        | Possibly damaging (0.49)     |
| p.E140K        | 1.1 (0.6)                        | SW        | Probably damaging (0.986)    |

NC, non-classical CAH; SV, simple virilising; SW, salt wasting.

*Threshold as deleterious set to c<−2.5.

*bPhenotype of p.V211M+ p.V281L/deletion genotype.

cProduction of 11DOC concentration under the detection limit of LC-MSMS method.

Table 3  Enzyme activity (% of WT) from four to five different experiments and in silico prediction of CYP21A2 mutations and variants.
alanine-to-threonine shift in position 159 is equal to the sequences of *B. taurus* and *S. scrofa* 21OH (Fig. 2), and was predicted as harmless (Table 3), again in line with a normal enzyme activity.

**Discussion**

In the present study, we introduced a novel method for functional studies of *CYP21A2* mutants. Using the coupled transcription/translation system for protein expression, we were able to measure the 21OH enzyme activity without the need of cell culture facilities and radiolabelling substrates, which is the current standard method (19, 20, 21, 22). We then characterised five novel and three known variants of the *CYP21A2* gene found in patients with CAH (7) and AAD. Overall, the measured enzyme activities and structural simulation corresponded to the phenotype with some exceptions.

The three polymorphisms used for verification of the *in vitro* enzyme assay behaved as expected; p.R102K (23) showed no reduction in activity, whereas the well-known SV mutation p.I172N had <2% activity and the G291R showed no activity at all, in accordance with previously reports (24, 25, 26). The novel p.L388R mutation showed almost complete abolishment of enzyme activity *in vitro*, which is in total agreement with the SW phenotype and the structural simulation. p.L388 is a crucial residue as it interacts with the heme prosthetic group through hydrophobic interactions. When the leucine at this position is replaced by the positively charged and more space-demanding arginine, the heme group is restricted from its normal binding to 21OH, which would have deleterious consequences for the enzymatic activity as well as the three-dimensional folding of 21OH.

The patient with the p.E140K mutation was diagnosed as SW, but even if *in vitro* enzyme assay showed a significant reduction in 21OH activity (11%), it was still much higher than expected for the SW phenotype. In addition, the prediction tools did not foresee a severe damaging effect, and was more in line with a NC phenotype. However, structural analysis shows that the p.E140K disrupts a salt bridge with p.R444, which is crucial for maintaining the tertiary structure (15) that could explain the SW phenotype. Thus, prediction tools alone are not always précis (27). On the other hand, phenotype and genotype divergence in classical CAH has been described, and the same mutation can give different phenotypes even in siblings (28).

The variant p.P45L was detected in a SV patient, but the *in vitro* enzyme activity did not correlate to such a severe
phenotype. However, this variant was predicted as probably damaging/deleterious by computer modelling. The p.P45L residue is located in the N-terminal region of the enzyme close to the hydrophobic domain that anchors 21OH to the endoplasmic reticulum (ER) membrane (15). The importance of the far N-terminal region has been experimentally determined earlier with regards to membrane targeting and insertion, as well as \textit{in vivo} protein stability (29). It has also been demonstrated that the first 20 amino acids are necessary for membrane integration but that enzymatic activity \textit{in vitro} can still be retained when these residues are deleted (30). Moreover, the p.P45L residue marks the start of an alpha helix that bends the direction of the polypeptide chain back towards the membrane. It is possible that this bending, for which proline in the 45 position is mandatory, is crucial for the correct orientation of 21OH in the ER membrane. If p.P45 is replaced by another amino acid, the overall stability and \textit{in vivo} enzymatic activity of the protein may be severely decreased. Another possibility is that the removal of the proline residue at position 45 alters the enzymes orientation to the ER membrane and somehow disturbs the electron transfer mediated \textit{in vivo} by the membrane-bound CYPOR. As we added exogenous CYPOR in our \textit{in vitro} assay, this effect could be lost. However, it is still possible that the rest of the enzyme maintains its structural integrity and that the intrinsic enzymatic function is intact. Similar effects of mutated proline residues have been described for other proteins, such as cubilin (the \textit{intrinsic factor-B12 receptor}) or the guanylate cyclase-activating protein 1 (involved in central vision) (31, 32).

Structural analysis shows that p.V211 is a part of the substrate cavity site and that it probably interacts with L65 through hydrophobic interactions (15). The conversion of valine into methionine at this position may alter this interaction slightly, but probably not to a damaging degree. PolyPhen2 and PROVEAN predicted p.V211M as benign/neutral, but with a score close to the threshold. It is unclear whether this mutation contributes to the intermediate form of SV/NC CAH due to a synergistic effect, similarly that has been seen in other studies were mild NC variants together have given SV (20, 21, 22, 34, 35) or SW phenotypes (36).

Finally, the p.A159T variant showed no reduction in the enzyme activity \textit{in vitro}. This variant was found in heterozygosity in three patients with AAD and in one healthy subject, and was predicted as harmless. The p.A159 residue is located on the surface of the protein in a region outside the secondary structure motifs and away from the active site, and occurs naturally in other mammals such as pigs and cows (Fig. 2). The substitution of alanine with threonine is also common in homologous proteins from related species.

In conclusion, the \textit{in vitro} enzyme assay and structural simulations revealed good correlations to the most severe phenotypes. For the other variants, a heterogeneous picture was observed. Nevertheless, \textit{in vitro} enzyme assays...
and structural simulations are valuable supplements for evaluating the severity of novel CYP21A2 variants.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

Funding

The study was supported by grants from the Regional Health Authorities of Western Norway, Bergen Medical Research Foundation, The Norwegian Funding

Acknowledgements

The authors thank Mrs Hilde Kristin Garberg at the Proteomics Unit at the University of Bergen (grant number 201167). I Brønstad is a PhD student supported by University of Bergen.

References

1 Speiser PW, Aziz R, Baskin LS, Ghizzoni L, Hensle TW, Merke DP, Meyer-Bahlburg HF, Miller WL, Montori VM, Oberfield SE et al. Congenital adrenal hyperplasia due to steroid 21-hydroxylase deficiency: an Endocrine Society clinical practice guideline. Journal of Clinical Endocrinology and Metabolism 2010 95 4131–4160. (doi:10.1210/jc.2009-2631)
2 Wedell A. An update on the molecular genetics of congenital adrenal hyperplasia: diagnostic and therapeutic aspects. Journal of Pediatric Endocrinology & Metabolism 1998 11 581–589. (doi:10.1515/JPEM.1998.11.5.581)
3 New MI. Extensive clinical experience: nonclassical 21-hydroxylase deficiency. Journal of Clinical Endocrinology and Metabolism 2006 91 4205–4214. (doi:10.1210/jc.2006-1645)
4 White PC, New MI & Dupont B. Structure of human steroid 21-hydroxylase genes. PNAS 1986 83 5111–5115. (doi:10.1073/pnas.83.14.5111)
5 Wedell A, Ritzen EM, Haglund-Stengler B & Luthman H. Steroid 21-hydroxylase deficiency: three additional mutated alleles and establishment of phenotype-genotype relationships of common mutations. PNAS 1992 89 7232–7236. (doi:10.1073/pnas.89.15.7232)
6 Krone N, Braun A, Roscher AA, Knorr D & Schwarz HP. Predicting phenotype in steroid 21-hydroxylase deficiency? Comprehensive genotyping in 153 unrelated, well defined patients from southern Germany Journal of Clinical Endocrinology and Metabolism 2000 85 1059–1065. (doi:10.1210/jcem.85.3.6441)
7 Nermoen I, Brønstad I, Fougner KJ, Svartberg J, Oknes M, Husebye ES & Lovas K. Genetic, anthropometric and metabolic features of adult Norwegian patients with 21-hydroxylase deficiency. European Journal of Endocrinology 2012 167 507–516. (doi:10.1530/EJE-12-0196)
8 Wedell A & Luthman H. Steroid 21-hydroxylase deficiency: two additional mutations in salt-wasting disease and rapid screening of disease-causing mutations. Human Molecular Genetics 1993 2 499–504. (doi:10.1093/hmg/2.5.499)
9 Wilson RC, Wei JQ, Cheng KC, Mercado AB & New MI. Rapid deoxyribonucleic acid analysis by allele-specific polymerase chain reaction for detection of mutations in the steroid 21-hydroxylase gene. Journal of Clinical Endocrinology and Metabolism 1995 80 1635–1640. (doi:10.1210/jcem.80.5.7745011)
10 Lau IF, Soardi FC, Lemos-Marini SH, Guerra G Jr, Baptista MT & De Mello MP. H285-C insertion in the CYP21 gene: a novel frameshift mutation in a Brazilian patient with the classical form of 21-hydroxylase deficiency. Journal of Clinical Endocrinology and Metabolism 2001 86 5877–5880. (doi:10.1210/jcem.86.12.8113)
11 Parajes S, Quintero C, Dominguez F & Loidi L. A simple and robust quantitative PCR assay to determine CYP21A2 gene dose in the diagnosis of 21-hydroxylase deficiency. Clinical Chemistry 2007 53 1577–1584. (doi:10.1373/clinchem.2007.078761)
12 Falorni A, Nikoshkov A, Laureti S, Ghenbach E, Hulting AL, Casucci G, Santeusanio F, Brunetti P, Luthman H & Lenmark A. High diagnostic accuracy for idiopathic Addison’s disease with a sensitive radiobinding assay for autoantibodies against recombinant human 21-hydroxylase. Journal of Clinical and Endocrine Metabolism 1995 80 2752–2755. (doi:10.1210/jcem.80.9.7673419)
13 Brantland E, Bredholt G, Meilgreen G, Knappskog PM, Mozes E & Husebye ES. The purification and application of biologically active recombinant steroid cytochrome P450 21-hydroxylase: the major autoantigen in autoimmune Addison’s disease. Journal of Autoimmunity 2009 33 58–67. (doi:10.1016/j.jaut.2009.02.018)
14 Mathieu P, Hassel SD, Fellmann R, Almas E, Erichsen MM, Husebye E & Lovas K. Multistereoid LC-MS/MS assay for glucocorticoids and androgens, and its application in Addison’s disease. Endocrine Connections 2013 2 125–136. (doi:10.1530/edc.13.0023)
15 Haider S, Islam B, D’Atri V, Sgboa M, Poojari C, Sun L, Yuen T, Zaidi M & New MI. Structure-phenotype correlations of human CYP21A2 mutations in congenital adrenal hyperplasia. PNAS 2013 110 2605–2610. (doi:10.1073/pnas.1221133110)
16 Adzhubei IA, Schmidt M, Peshkin L, Ramsken VE, Gerassimova A, Bork P, Kondrashov AS & Sunyaev SR. A method and server for predicting damaging missense mutations. Nature Methods 2010 7 248–249. (doi:10.1038/nmeth0410-248)
17 Choi Y, Sims GE, Murphy S, Miller JR & Chan AP. Predicting the functional effect of amino acid substitutions and indels. PLoS ONE 2012 7 e46688. (doi:10.1371/journal.pone.0046688)
18 Sievers F, Wilm A, Dineen D, Gibson TJ, Karplus K, Li W, Lopez R, McWilliam H, Remmert M, Soding J et al. Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. Molecular Systems Biology 2011 7 539. (doi:10.1038/msb.2011.75)
19 Barbaro M, Baldazzi L, Balsamo A, Lajic S, Robins T, Barp L, Pirazzoli P, Ciaciari E, Cicognani A & Wedell A. Functional studies of two novel and two rare mutations in the 21-hydroxylase gene. Journal of Molecular Medicine 2006 84 521–528. (doi:10.1007/s00109-006-0043-7)
20 Barbaro M, Lajic S, Baldazzi L, Balsamo A, Pirazzoli P, Cicognani A, Wedell A & Caciari E. Functional analysis of two recurrent amino acid substitutions in the CYP21 gene from Italian patients with congenital adrenal hyperplasia. Journal of Clinical and Endocrinology and Metabolism 2004 89 2402–2407. (doi:10.1210/jcem.2003-031630)
21 Nikoshkov A, Lajic S, Holst M, Wedell A & Luthman H. Synergistic effect of partially inactivating mutations in steroid 21-hydroxylase deficiency. Journal of Clinical and Endocrinology and Metabolism 1997 82 194–199. (doi:10.1210/jcem.82.1.167)
22 Tardy V, Menassa R, Sulmont V, Lienhardt-Roussie A, Lecointre C, Brauner R, David M & Morel Y. Phenotype-genotype correlations of 13 rare CYP21A2 mutations detected in 46 patients affected with 21-hydroxylase deficiency and in one carrier. Journal of Clinical Endocrinology and Metabolism 2010 95 1288–1300. (doi:10.1210/jcem.2009-1202)
23 Rodrigues NR, Dunham I, Yu CY, Carroll MC, Porter RR & Campbell RD. Molecular characterization of the HLA-linked steroid 21-hydroxylase B gene from an individual with congenital adrenal hyperplasia. EMBO Journal 1987 6 1653–1661.
24 Amor M, Parker KL, Globerman H, New MI & White PC. Mutation in the CYP21B gene (Ile-172→Asm) causes steroid 21-hydroxylase deficiency. PNAS 1988 85 1600–1604. (doi:10.1073/pnas.85.5.1600)
25 Barbaro M, Soardi FC, de Mello MP, Wedell A & Lajic S. Functional studies of CYP21A2 mutants complement structural and clinical predictions of disease severity in CAH. *Clinical Endocrinology* 2012 **76** 766–768. (doi:10.1111/j.1365-2265.2011.04275.x)

26 Tusie-Luna MT, Traktman P & White PC. Determination of functional effects of mutations in the steroid 21-hydroxylase gene (CYP21) using recombinant vaccinia virus. *Journal of Biological Chemistry* 1990 **265** 20916–20922.

27 Chan AO. Performance of in silico analysis in predicting the effect of non-synonymous variants in inherited steroid metabolic diseases. *Stemoids* 2013 **78** 726–730. (doi:10.1016/j.stemoids.2013.04.002)

28 Chin D, Speiser PW, Imperato-McGinley J, Dixit N, Uli N, David R & Oberfield SE. Study of a kindred with classic congenital adrenal hyperplasia: diagnostic challenge due to phenotypic variance. *Journal of Clinical Endocrinology and Metabolism* 1998 **83** 1940–1945. (doi:10.1210/jcem.83.6.4887)

29 Hsu LC, Hu MC, Cheng HC, Lu JC & Chung BC. The N-terminal hydrophobic domain of P450c21 is required for membrane insertion and enzyme stability. *Journal of Biological Chemistry* 1993 **268** 14682–14686.

30 Lajic S, Nikoshkov A, Holst M & Wedell A. Effects of missense mutations and deletions on membrane anchoring and enzyme function of human steroid 21-hydroxylase (P450c21). *Biochemical and Biophysical Research Communications* 1999 **257** 384–390. (doi:10.1006/bbrc.1999.0482)

31 Aminoff M, Carter JE, Chadwick RB, Johnson C, Grasbeck R, Abdelal MA, Broch H, Jenner LB, Verroust PJ, Møesstrup SK et al. Mutations in CUBN, encoding the intrinsic factor–vitamin B12 receptor, cubilin, cause hereditary megaloblastic anaemia 1. *Nature Genetics* 1999 **21** 309–313. (doi:10.1038/6831)

32 Newbold RJ, Deery EC, Walker CE, Wilkinson SE, Srinivasan N, Hunt DM, Bhattacharya SS & Warren MJ. The destabilization of human GCAP1 by a proline to leucine mutation might cause cone-rod dystrophy. *Human Molecular Genetics* 2001 **10** 47–54. (doi:10.1093/hmg/10.1.47)

33 Speiser PW, New MI & White PC. Molecular genetic analysis of nonclassic steroid 21-hydroxylase deficiency associated with HLA-B14,DR1. *New England Journal of Medicine* 1988 **319** 19–23. (doi:10.1056/NEJM198807073190104)

34 Menassa R, Tardy V, Despert F, Bouvattier-Morel C, Brossier JP, Cartigny M & Morel Y. p.H62L, a rare mutation of the CYP21 gene identified in two forms of 21-hydroxylase deficiency. *Journal of Clinical Endocrinology and Metabolism* 2008 **93** 1901–1908. (doi:10.1210/jc.2007-2701)

35 Soardi FC, Barbaro M, Lau IF, Lemos-Marini SH, Baptista MT, Guerra-Junior G, Wedell A, Lajic S & de Mello MP. Inhibition of CYP21A2 enzyme activity caused by novel missense mutations identified in Brazilian and Scandinavian patients. *Journal of Clinical Endocrinology and Metabolism* 2008 **93** 2416–2420. (doi:10.1210/jc.2007-2594)

36 Dolzan V, Solyom J, Fekete G, Kovacs J, Rakosnikova V, Votava F, Lebl J, Pribilincova Z, Baumgartner-Parzer SM, Riedl S et al. Mutational spectrum of steroid 21-hydroxylase and the genotype–phenotype association in Middle European patients with congenital adrenal hyperplasia. *European Journal of Endocrinology* 2005 **153** 99–106. (doi:10.1530/eje.1.01944)

Received in final form 24 March 2014
Accepted 26 March 2014

http://www.endocrineconnections.org
DOI: 10.1530/EC-14-0032
© 2014 The authors
Published by Bioscientifica Ltd

This work is licensed under a Creative Commons Attribution 3.0 Unported License.