**PAI-1 4G/5G Polymorphism Contributes to Cancer Susceptibility: Evidence from Meta-Analysis**

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**Abstract**

**Background:** The plasminogen activator inhibitor-1 (PAI-1) is expressed in many cancer cell types and allows the modulation of cancer growth, invasion and angiogenesis. To date, studies investigated the association between a functional polymorphism in PAI-1 (4G/5G) and risk of cancer have shown inclusive results.

**Methods:** A meta-analysis based on 25 case-control studies was performed to address this issue. Odds ratios (OR) with corresponding 95% confidence intervals (CIs) were used to assess the association. The statistical heterogeneity across studies was examined with I² test.

**Results:** Overall, a significant increased risk of cancer was associated with the PAI-1 4G/4G polymorphism for the allele contrast (4G vs. 5G: OR = 1.10, CI = 1.03–1.18, I² = 49.5%), the additive genetic model (4G/4G vs. 5G/5G: OR = 1.21, CI = 1.06–1.39, I² = 51.9%), the recessive genetic model (4G/4G vs. 5G/5G: OR = 1.11, CI = 1.04–1.18, I² = 20.8%). In the subgroup analysis by ethnicity, the results indicated that individuals with 4G/4G genotype had a significantly higher cancer risk among Caucasians (4G/4G vs. 5G/5G: OR = 1.31, 95%CI = 1.09–1.59, I² = 59.6%; 4G/4G vs. 5G/5G: OR = 1.12, 95%CI = 1.04–1.21, I² = 3.6%; recessive model: OR = 1.12, 95%CI = 1.05–1.21, I² = 25.3%).

**Conclusions:** The results of the present meta-analysis support an association between the PAI-1 4G/5G polymorphism and increasing cancer risk, especially among Caucasians, and those with 4G allele have a high risk to develop colorectal cancer and endometrial cancer.

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**Introduction**

The urokinase plasminogen activator inhibitor system is a serine protease family [1]. The included urokinase-type plasminogen activator (uPA) system provides the most substantial amount of activated plasminogen when tissues are being degraded and is involved in extracellular matrix (ECM) degradation [2,3], hence it has been involved in numerous pathophysiological processes requiring the remodeling of basement membranes (BM) and ECM. Metastasis and invasion of malignant cancers require proteolytic degradation of the ECM, BM and infiltration of cancer cells into the surrounding tissues, the blood stream, or the lymphatic vessels. Studies revealing the uPA system, universal to all cancers, is associated with the process of cancer metastasis and progression by participating in the degradation and regeneration of the BM and ECM [4,5,6].

The plasminogen activator inhibitor-1 (PAI-1), a 52 kDa glycoprotein belong to the serine proteinase inhibitor super family, is a multifaceted proteolytic factor. It is the principal inhibitor of tissue and urinary plasminogen activators, and therefore constitutes an important regulatory protein in fibrinolysis [7,8]. It is also involved in the regulation of cell adhesion, detachment and migration, playing an important role in cancer progression [9,10,11]. Indeed, PAI-1 is expressed in many types of cancer cell and allows the modulation of cancer growth, invasion and angiogenesis in a dose-dependent manner [12].

Genetic polymorphisms in the PAI-1 gene seem to contribute to the level of PAI-1 biosynthesis [13]. A single nucleotide insertion/deletion (4G/5G) polymorphism located at 675 base-pair (bp) upstream of the transcriptional start site in the PAI-1 promoter, is the most frequently studied variant because of its possible involvement in the regulation of PAI-1 transcription [14,15,16]. Based on the investigation by CDC (Centers for Disease Control and Prevention), the 4G/5G allele frequencies range in various populations from 26.7/73.3% to 52.5/47.5%, respectively.
The distribution of 4G/5G allele frequencies has been shown in Table S1. Homozygosity of the 4G allele is considered to be a risk factor for developing deep vein thrombosis, myocardial infarction and high rate of miscarriage during pregnancy [17,18,19]. Many molecular epidemiological studies have been conducted to investigate the association between 4G/5G polymorphism and cancer risk in humans [20–43]. However, the results from these studies are to some extent divergent, but nevertheless intriguing, which may be owe to limitations in individual studies. To address this issue, we performed a meta-analysis with subgroup analysis from all eligible studies, to obtain a more precise estimation of the relationship between PAI-1 4G/5G polymorphism and cancer risk.

Materials and Methods

Identification and Eligibility of Relevant Studies

All case-control studies on the association between PAI-1 polymorphisms and cancer risk published up to July 31, 2012 were identified through comprehensive searches using the PubMed and EMBASE database with the following terms and keywords: “plasminogen activator inhibitor-1”, “PAI-1” and “polymorphism”, “variation”, “mutation” and in combination with “cancer”, “tumor” and “carcinoma”. The search was limited to human studies and English language papers.

Inclusion Criteria

For inclusion in the meta-analysis, the identified articles have to meet the following criteria: (a) there is information on the evaluation of the PAI-1 4G/5G polymorphism and cancer risk, (b) using a case-control design, and (c) containing complete information about all genotype frequency. The exclusion criteria are as follows: (a) not for cancer research, (b) review articles, (c) reports without usable data and (d) duplicate publications.

Data Extraction

Information was carefully extracted from all the eligible publications independently by two researchers (SQ Wang and Q Cao) according to the inclusion criteria listed above. For conflicting evaluation, a consensus was reached by discussion. The following information was extracted from each included study using a standardized data collection protocol (File S1): the first
Table 1. Characteristics of studies included in the meta-analysis.

| First author | Ethnicity | Country | Cancer | Genotyping | Source of Controls | Sample size | HWE |
|--------------|-----------|---------|--------|------------|-------------------|-------------|-----|
| Turkmen 1997 | Caucasian | Germany | Ovarian cancer | PCR-RFLP | HB | 22 | 23 | Y |
| Smolaz 1999 | Caucasian | Poland | Breast cancer | Allele-specific PCR | HB | 37 | 53 | Y |
| Blasiak 2000 | Caucasian | Poland | Breast cancer | Allele-specific PCR | HB | 100 | 106 | Y |
| Loktionov 2003 | Caucasian | UK | Colorectal | PCR-RFLP | HB | 206 | 355 | Y |
| Castello 2006 | Caucasian | Spain | Breast cancer | Allele-specific PCR | HB | 104 | 104 | Y |
| Eroglu 2006 | Caucasian | Turkey | Breast cancer | PCR-RFLP | HB | 34 | 90 | Y |
| Sternlicht 2006 | Caucasian | UK | Breast cancer | PCR-RFLP | PB | 2539 | 1832 | Y |
| Eroglu 2007 | Caucasian | Turkey | Others | PCR-RFLP | HB | 125 | 180 | Y |
| Forstl 2007 | Caucasian | Sweden | Colorectal cancer | Taqman | PB | 304 | 581 | Y |
| Jorgenson 2007 | Mixed | USA | Prostate cancer | PCR-RFLP | PB | 638 | 478 | Y |
| Minisini 2007 | Caucasian | Italy | Breast cancer | Allele-specific PCR | HB | 193 | 142 | Y |
| Woo 2007 | Asian | Korea | Colorectal cancer | PCR-RFLP | HB | 185 | 304 | Y |
| Lei 2008 | Caucasian | Sweden | Breast cancer | Taqman | PB | 956 | 943 | Y |
| Bentov 2009 | Mixed | Canada | Ovarian cancer | MassARRAY | PB | 772 | 889 | Y |
| Palmirota 2009 | Caucasian | Italy | Breast cancer | PCR-RFLP | HB | 99 | 50 | Y |
| Vairaktaris 2009 | Caucasian | Greece | Oral cancer | PCR-RFLP | HB | 104 | 106 | Y |
| Ju 2010 | Asian | Korea | Gastric cancer | MassARRAY | PB | 252 | 406 | Y |
| Weng 2010 | Asian | Taiwan | Hepatocellular cancer | PCR-RFLP | HB | 102 | 344 | Y |
| Gilbart-Estelles 2011 | Caucasian | Spain | Endometrial cancer | Allele-specific PCR | HB | 212 | 211 | Y |
| Su 2011 | Asian | Taiwan | Endometrial | PCR-RFLP | HB | 134 | 302 | Y |
| Vossen 2011 | Caucasian | Germany | Colon cancer | Taqman | PB | 1059 | 1799 | Y |
| Vossen 2011 | Caucasian | Germany | Rectal cancer | Taqman | PB | 672 | 1799 | Y |
| Weng 2011 | Asian | Taiwan | Oral cancer | PCR-RFLP | HB | 253 | 344 | Y |
| Onur 2012 | Caucasian | Turkey | Others | Two parallel PCR | HB | 28 | 50 | Y |
| Tee 2012 | Asian | Taiwan | Cervical cancer | PCR-RFLP | HB | 75 | 336 | Y |

HB, hospital based; PB, population based; HWE, Hardy–Weinberg equilibrium. doi:10.1371/journal.pone.0056797.t001

author’s name, the year of publication, ethnicity, country of origin, cancer type, genotyping method and source of control groups (population- or hospital-based controls) and deviation from Hardy-Weinberg Equilibrium (HWE) of the control group. Different ethnic descents were categorized as African, Asian, European, or Mixed (composed of different ethnic groups). Meanwhile, different case-control groups in one study were considered as independent studies.

Statistical Methods

The strength of the association between the PAI-1 4G/5G polymorphism and cancer risk was measured by odds ratios (ORs) with corresponding 95% confidence intervals (CIs). The percentage weight determined by the precision of its estimate of effect and, in the statistical software in STATA and SAS, is equal to the inverse of the variance. The risks (ORs) of cancer associated with the PAI-1 4G/5G polymorphism were estimated for each study. In our study, the 5G allele was considered the reference genotype. The pooled ORs were performed for additive genetic model (4G/4G vs. 5G/5G and 4G/4G vs. 4G/5G), dominant model (4G/4G +4G/5G vs. 5G/5G) and recessive model (4G/4G vs. 4G/5G +4G/5G), respectively. Stratified analyses were also performed by cancer types (if one cancer type contained less than two individual studies, it was classified as other cancers group), ethnicity, source of controls and sample size (subjects $\geq$500 in both case and control groups or not). In consideration of the possibility of heterogeneity across the studies, a statistical test for heterogeneity was performed by a $I^2$ test. A $I^2$ smaller than 31% indicates lack of heterogeneity among the studies, and then the fixed-effects model (the Mantel-Haenszel method) was used to calculate the summary OR estimate of each study. Otherwise, the random effects model (DerSimonian and Laird method) was used. For each study, we examined whether the genotype distribution of controls was consistent with HWE using the $x^2$ test. One-way sensitivity analysis was performed to assess the stability of the results, namely, a single study in the meta-analysis was deleted each time to reflect the influence of the individual data set to the pooled OR. An estimate of potential publication bias was carried out by the funnel plot, in which the standard error of log (OR) of each study was plotted against its log (OR). An asymmetric plot suggests a possible publication bias. Funnel plot asymmetry was assessed by the method of Egger’s linear regression test, a linear regression approach to measure funnel plot asymmetry on the natural logarithm scale of the OR. All statistical analyses were performed with the Stata software (version 12.1; StataCorp LP, College Station, TX, USA) and SAS software (Version 9.2; SAS Institute Cary, NC, USA), using two-sided $P$-values.
Results

Characteristics of Studies

There were 25 studies retrieved on the basis of the search criteria (Fig. 1). Totally, 9,205 cases and 11,827 controls were included in the meta-analysis. Study characteristics were summarized in Table S1. Among the 25 case-control studies, there were 17 studies of Caucasians, 6 studies of Asians, and 2 studies of mixed descendents. There were 8 breast cancer studies, 5 colorectal cancer studies, 2 endometrial cancer studies, 2 oral cancer studies, and the others were categorized into the “other cancer” group. Cancers were confirmed histologically or pathologically in most studies. Controls were mainly matched on sex and age, of which 17 were hospital based [21,22,23,24,29,30,31,32,33,35,36,37,38,40,41,42,43], 8 were population based [20,25,26,27,28,34,39]. Furthermore, 10 studies were conducted with subjects ≥500 in both case and control groups [20,25,26,27,28,29,34,39,40]. Diverse genotyping methods were used, including PCR-RFLP, TaqMan, allele-specific PCR, MassARRAY, and two parallel PCR. The distribution of genotypes in the controls of all studies was consistent with HWE.

Quantitative Synthesis

The relationship between the 4G/5G polymorphism in PAI-1 and the risk of different kinds of cancer are summarized in Table 2. Overall, a significantly increased risk of cancer was associated with the PAI-1 4G polymorphism for the allele contrast (4G vs. 5G: OR = 1.10, CI = 1.03–1.18, \( I^2 = 49.5\% \)), the additive genetic model (4G/4G vs. 5G/5G: OR = 1.21 CI = 1.06–1.39, (Fig. 2); 4G/4G vs. 4G/5G: OR = 1.10 CI = 1.03–1.18), the recessive genetic model (4G/4G vs. 4G/5G+5G/5G OR = 1.11 CI = 1.04–1.18). In the subgroup analysis by ethnicity, the results indicated that individuals with 4G/4G genotype had a significantly higher cancer risks among Caucasians (4G/4G vs. 5G/5G: OR = 1.31, 95%CI = 1.09–1.59; 4G/4G vs. 4G/5G: OR = 1.12, 95%CI = 1.04–1.21; recessive model: OR = 1.38, 95%CI = 1.10–1.74; recessive model: OR = 1.45, 95%CI = 1.14–1.80). In the stratified analysis by cancer types, significant associations were found for Endometrial cancer (4G/4G vs. 5G/5G: OR = 2.23, 95%CI = 1.45–3.42; 4G/4G vs. 4G/5G: OR = 1.45, 95%CI = 1.04–2.04; dominant model: OR = 1.74, 95%CI = 1.20–2.51).

Figure 2. Forest plot of cancer risk associated with the PAI-1 4G/5G polymorphism (4G/4G vs. 5G/5G). The squares and horizontal lines correspond to the study-specific OR and 95% CI. The area of the squares reflects the weight (inverse of the variance). The diamond represents the summary OR and 95% CI.
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| Variables | Sample size | 4Gvs5G | 4G/4Gsv5G/5G | 4G/4Gsv4G/5G | 4G/4Gsv4G/5G+5G/5G |
|-----------|------------|--------|--------------|-------------|-------------------|
| **N** | case | control | OR (95% CI) | I² (%) | OR (95% CI) | I² (%) | OR (95% CI) | I² (%) | OR (95% CI) | I² (%) |
| Total | 25 | 9205 | 11827 | 1.10 (1.03–1.18) | 49.5<sup>c</sup> | 1.21 (1.06–1.39) | 51.9<sup>c</sup> | 1.10 (1.03–1.18) | 0<sup>c</sup> | 1.11 (1.04–1.20) | 20.8<sup>c</sup> |
| **Tumor type** | | | | | | | | | | |
| Breast cancer | 8 | 4062 | 3320 | 1.14 (1.00–1.29) | 48.3 | 1.30 (0.99–1.70) | 48.8 | 1.05 (0.94–1.16) | 6 | 1.07 (0.97–1.18) | 22.8 |
| Colorectal cancer | 5 | 2426 | 4838 | 1.03 (0.96–1.11) | 0 | 1.04 (0.90–1.19) | 0 | 1.19 (1.06–1.33) | 0 | 1.14 (1.03–1.27) | 0 |
| Ovarian cancer | 2 | 794 | 912 | 0.98 (0.86–1.13) | 0 | 0.97 (0.74–1.27) | 0 | 1.01 (0.81–1.26) | 55.1 | 1.00 (0.81–1.23) | 22.9 |
| Endometrial cancer | 2 | 346 | 513 | 1.45 (1.19–1.77) | 0 | 2.23 (1.45–3.42) | 0 | 1.45 (1.04–2.04) | 0 | 1.64 (1.19–2.27) | 0 |
| Oral cancer | 2 | 357 | 450 | 1.39 (0.73–2.63) | 87.3 | 1.94 (0.54–6.91) | 86.5 | 1.07 (0.77–1.49) | 0 | 1.20 (0.88–1.64) | 67.7 |
| Others | 6 | 1220 | 1794 | 1.08 (0.90–1.30) | 57.8 | 1.18 (0.79–1.78) | 63.2 | 1.07 (0.89–1.28) | 0 | 1.08 (0.91–1.28) | 24.6 |
| **Ethnicity** | | | | | | | | | | |
| Caucasian | 17 | 6794 | 8424 | 1.14 (1.04–1.25) | 56.8 | 1.31 (1.09–1.59) | 59.6 | 1.12 (1.04–1.21) | 3.6 | 1.12 (1.05–1.21) | 25.3 |
| Asian | 6 | 1001 | 2036 | 1.07 (0.92–1.25) | 45.9 | 1.14 (0.84–1.56) | 44.8 | 1.07 (0.90–1.28) | 17.3 | 1.08 (0.91–1.27) | 37.8 |
| Mixed | 2 | 1410 | 1367 | 1.02 (0.92–1.13) | 0 | 1.03 (0.84–1.27) | 0 | 1.06 (0.89–1.27) | 0 | 1.05 (0.89–1.24) | 0 |
| **Control source** | | | | | | | | | | |
| Hospital based | 17 | 2013 | 3100 | 1.25 (1.11–1.40) | 43 | 1.59 (1.24–2.05) | 48.1 | 1.22 (1.07–1.40) | 0 | 1.30 (1.14–1.48) | 0 |
| Population based | 8 | 7192 | 8727 | 1.02 (0.97–1.07) | 0 | 1.03 (0.94–1.13) | 0 | 1.07 (0.99–1.15) | 8.4 | 1.06 (0.99–1.13) | 0 |
| **Sample size (both cases and controls)** | | | | | | | | | | |
| <500 | 15 | 1554 | 2401 | 1.30 (1.14–1.48) | 40 | 1.73 (1.31–2.31) | 46.5 | 1.24 (1.06–1.45) | 0 | 1.36 (1.17–1.57) | 0 |
| ≥500<sup>d</sup> | 10 | 7651 | 9426 | 1.02 (0.98–1.07) | 0 | 1.03 (0.94–1.13) | 0 | 1.07 (0.97–1.18) | 5 | 1.06 (0.99–1.14) | 0 |

<sup>a</sup>Number of studies.
<sup>b</sup>The value of heterogeneity test.
<sup>c</sup>Fix-effects model was used when I² value for heterogeneity test <31%; otherwise, random-effects model was used.
<sup>d</sup>Stratified according to subjects ≥500 in both case and control groups or not.
<sup>e</sup>The exact value is 1.077 (1.002–1.156).

**Sensitivity Analysis**

The sensitivity analysis was conducted by leaving out certain studies, such as the study that did not conform to HWE. The omission of individual studies did not materially alter the results, although on some occasions, the $I^2$ value for heterogeneity was reduced. The sensitivity analysis thus confirmed that the results of this meta-analysis were statistically robust. This procedure proved that our results were reliable and stable. Furthermore, when excluding the studies that were not in HWE, the estimated pool OR still did not change at all.

**Publication Bias**

Begg’s funnel plot and Egger’s test were performed to assess the publication bias of literatures. As shown in the Fig. 4, the shapes of the funnel plots seems symmetrical in the recessive genetic model (4G/4G vs. 4G/5G+5G/5G), but not for homozygote model (4G/4G vs. 5G/5G). Thus, the Egger’s test was used to provide statistical evidence of funnel plot symmetry. For recessive genetic model, the results did not show any evidence of publication bias ($t = 1.96, P = 0.097$ for 4G/4G vs. 4G/5G+5G/5G). However, the homozygote model showed significant publication bias ($t = 2.99, P = 0.014$). To adjust for this bias, a trim-and-fill method developed by Duval and Tweedie [44] was used to both identify and correct for funnel plot asymmetry arising from publication bias. We trimmed off the asymmetric outlying part of the funnel after estimating how many studies were in the asymmetric part. The sensitivity analysis thus confirmed that the results of this meta-analysis were statistically robust. This procedure proved that our results were reliable and stable. Furthermore, when excluding the studies that were not in HWE, the estimated pool OR still did not change at all.

### Test for Heterogeneity

There was significant heterogeneity for allele contrast (4G vs. 5G: $I^2 = 49.5\%$), homozygote comparison (4G/4G vs. 5G/5G: $I^2 = 51.9\%$), heterozygote comparison (4G/5G vs. 5G/5G: $I^2 = 48.7\%$), dominant model comparison (4G/4G+4G/5G vs. 5G/5G: $I^2 = 53.9\%$), recessive model comparison (4G/4G vs. 4G/5G+5G/5G: $I^2 = 20.8\%$). Then, we used a meta-regression analysis to explore the source of heterogeneity for homozygote comparison (4G/4G vs. 5G/5G) by Ethnicity, cancer types, source of controls and sample size. We found that the sample size ($\tau^2 = 0, P = 0.001$) contributed to substantial altered heterogeneity, which could account for 100% source of heterogeneity. Also, control source ($\tau^2 = 0, P = 0.005$) contributed to 100% source of heterogeneity. However, we did not find cancer types ($\tau^2 = 0.074, P = 0.615$), or ethnicity ($\tau^2 = 0.075, P = 0.947$) contributed to source of heterogeneity.
trimmed study and its missing counterpart around the center. The final estimate of the true mean, and also its 95% CI, were then based on the filled funnel plot. The OR estimates and 95% CI in fixed-effect model before and after trim-and-fill were 1.119, (1.032–1.213) and 1.115, (1.029–1.209). Also, for random-effect model, the results were 1.214, (1.057–1.394) and 1.204, (1.049–1.392). Meta-analysis with or without the trim-and-fill method did not draw different conclusions, indicating that our results were statistically robust.

Discussion

The present meta-analysis, including 9,205 cases and 11,827 controls from 25 case-control studies, explored the association between the PAI-1 4G/5G polymorphism and cancer risk. Our results indicated that the variant 4G/4G genotype was associated with an increased risk of cancers, especially of colorectal cancer and endometrial cancer.

In recent years, many studies have been conducted to investigate the associations between the PAI-1 4G/5G polymorphisms and disease risk across different countries. The results remain inconclusive. A new manuscript in Blood by Huang et al. [45] just published validated the role of the 4G/5G polymorphisms in circulating PAI-1 levels using GWAS data. However, they revealed no association between PAI-1 4G/5G and type 2 diabetes (T2D) and coronary artery disease (CAD), despite enormous sample sizes.

Many researchers investigated the relationship between PAI-1 blood concentrations and diseases risk. Palmirotta et al. [32]
reported plasma PAI-1 levels in breast cancer patients were approximately two-fold higher than those observed in control subjects and were strongly dependent on cancer size, suggesting that cancer-related factors might be responsible for PAI-1 up-regulation. However, the exact concentration of PAI-1 in these breast cancer patients’ blood stayed 27.2 ng/ml (16.5–35.0), and Huang et al. [45] revealed cumulative effect of all common alleles explained extremely low blood levels of PAI-1 in their GWAS data. How would such a seemingly small influence of genotype on PAI-1 levels be expected to modify cancer risk? Given the important roles of PAI-1 in multiple biological functions, such as regulation of cell adhesion, detachment and migration, it is biologically plausible that the PAI-1 4G/5G polymorphism may modulate the risk of cancers. Functional studies on this polymorphism have shown that the 4G allele binds only an activator, while the 5G allele binds a repressor as well as an activator, therefore results in reduced transcription of PAI-1 [46]. It suggests that the 4G allele is associated with reduced inhibition of the plasminogen activators and, consequently, increased plasminogen conversion to plasmin, increased activation of MMPs and decreased adhesive strength of...
cells for their substratum [17,18,46]. Consistent with these observations, our meta-analysis showed that individuals carrying 4G/4G genotype were associated with a higher cancer risk than subjects carrying at least one 5G allele.

In addition, our results showed that the 4G allele may be a risk factor for colorectal cancer and endometrial cancer but not for breast cancer, ovarian cancer, oral cancer, or hepatocellular cancer. One factor that would contribute to the discrepancy among different studies is that this polymorphism might play a different role in different cancer sites. However, even at the same cancer site, considering the possible small effect size of this genetic polymorphism to cancer risk and the relatively small sample size in some studies, the discrepancy will become apparent since some of these studies may be underpowered to detect a small but real association. For endometrial cancer, there were only two studies included in the analysis with limited sample sizes, therefore, the results should be interpreted with caution.

In the subgroup analysis by ethnicity, an increased risk in 4G carriers was found among Caucasians but not Asians or Mixed. One explanation for this result may be that the studies using Mixed ethnicity participants enrolled them from various countries with diverse cultural, environmental and genetic characteristics. It is expected that these factors affected the synthesis results. On the other hand, the sample size and numbers of studies in Asian group were not adequate to evaluate the association. Other factors such as selection bias and different matching criteria may also play a role.

The genetic models were summarised in Table 2 including allele contrast model, homozygote model, heterozygote model and recessive model. Because of the strong heterogeneity in allele contrast and homozygote model, though, the results of these two shows significantly different, we do not suggest any one of these two as the best-fit model to represent the whole genetic models. There is a relatively low heterogeneity ($\chi^2 = 20.8\%$) in recessive model, the OR value and the confidence interval shows significantly different. As a result, the recessive model might be the best-fit model in this meta-analysis to reflect the whole results.

Furthermore, despite the overall robust statistical evidence generated through this analysis, some methodological limitations have been identified. Firstly, the relatively high heterogeneity and small sample size are the major defect in this meta-analysis. In the subgroup analyses by ethnicity and cancer type, the sample size of studies among Asians and among several cancer types is small and limited. As a result, the sample size accounted for most of the source of heterogeneity. Also, lacking the original data of the studies among Asians and among several cancer types is small and sample size are the major defect in this meta-analysis. In the analysis tools: MT JZF JQ. Wrote the paper: SQW XXW WQY. Analyzed the data: SQW QC BJL. Contributed reagents/materials/experiments: QC CQ WZ. Performed the experiments: SQW QC XXW. Provided software analysis: BJL SQW. Conceived and designed the experiments: QC CQ WZ. Performed the experiments: SQW QC XXW. Made substantial contributions to the design of the study: MT JZF JQ. Wrote the paper: SQW XXW WQY.

### Conclusions

In conclusion, the evidence of the results from the present meta-analysis support an association between the PAI-1 4G/5G polymorphism and increasing cancer risk, especially among Caucasians, and those with colorectal cancer and endometrial cancer or cancers identified in the other cancers group, though significant heterogeneity from included studies existed. To advance an understanding of this relationship, the following recommendations have been made: (1) Large studies using standardized unbiased methods, enrolling precisely defined cancer patients and well matched controls, with more detailed individual data is needed. (2) Studies conducted with ethnic groups other than Caucasians are required to gain a more comprehensive and generalizable conclusion. (3) More and larger studies, especially studies stratified for gene-environmental interaction, should be performed to clarify the possible roles of the PAI-1 4G/5G polymorphisms in the etiology of cancer.

### Supporting Information

Table S1 The genotype frequencies on each studies. A generalized distribution of genotype frequencies on each included studies are listed. (DOC)

Table S2 Stratification analyses of the $F$ and 95% confidence interval. If $F = 0$, the one-sided 97.5% CI is presented. Otherwise, a two-sided 95% CI is performed. (DOC)

File S1 PRISMA 2009 Checklist. (DOC)

### Author Contributions

Provided software analysis: BJL SQW. Conceived and designed the experiments: QC CQ WZ. Performed the experiments: SQW QC XXW. Made substantial contributions to the design of the study: MT JZF JQ. Wrote the paper: SQW XXW WQY.

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