Towards Theranostic Multicompartment Microcapsules: in-situ Diagnostics and Laser-induced Treatment

Ranhua Xiong, Stefaan J. Soenen, Kevin Braeckmans, Andre G. Skirtach

1. Lab. of General Biochemistry and Physical Pharmacy, Ghent University, Ghent 9000, Belgium;
2. Centre for Nano-Bio (NB)-photronics, Ghent University, Ghent 9000, Belgium;
3. Department of Molecular Biotechnology, Ghent University, Ghent 9000, Belgium;
4. Max-Planck Institute of Colloids and Interfaces, Potsdam 14424 Germany.

Corresponding author: Andre.Skirtach@UGent.be.

Abstract

Paving the way towards the application of polyelectrolyte multilayer capsules in theranostics, we describe diagnostic multi-functionality and drug delivery using multicompartment polymeric capsules which represent the next generation of drug delivery carriers. Their versatility is particularly important for potential applications in the area of theranostics wherein the carriers are endowed with the functionality for both diagnostics and therapy. Responsiveness towards external stimuli is attractive for providing controlled and on-demand release of encapsulated materials. An overview of external stimuli is presented with an emphasis on light as a physical stimulus which has been widely used for activation of microcapsules and release of their contents. In this article we also describe existing and new approaches to build multicompartment microcapsules as well as means available to achieve controlled and triggered release from their subcompartments, with a focus on applications in theranostics. Outlook for future directions in the area are highlighted.

Key words: theranostics, multicompartment, capsules, stimuli, laser, nanoplasmonics.

Introduction

Theranostics is an emerging area which combines multifunctionalization [1] for diagnostics and therapy, while nano and micromaterials are developed to enable both functionalities. Such a synergy, particularly if the materials allow to perform diagnosis and therapy simultaneously, will allow physicians not only to detect diseases, but ideally will also permit them to administer proper medicines at the appropriate time, location and dose in the body and to fight many diseases including cancer [2]. Laser light is particularly interesting for theranostics [3] largely due to its potential in phototherapy [4]. Application of light to theranostics opens extensive opportunities in pharmaceutical research to monitor effects of drugs, their distribution, release, and dose thus representing the next step [5] for nanomedicine [6].

A very interesting class of materials for theranostics is multicompartment polyelectrolyte multilayer capsules, which are responding to different stimuli [7]. Multicompartmentalization is an attractive strategy because it allows for assembling or encapsulating several types of molecules simultaneously in the same carrier. Such an approach is particularly suitable for theranostics as some compartments are intended for diagnostics, while other subcompartments are to be used for therapy. A generalization of various approaches to multicompartmentalization has been recently presented by Delcea et al [1]. The structure of multicompartment [8] delivery vehicles can be very diverse [9, 10] including a hybrid microcapsule-liposome system with liposomes [11-13] or self-assembled construction [14] including those...
based on polymeric systems [15]. Another desirable feature of a theranostic carrier is the induction of on-demand drug release [16, 17] to initiate a therapeutic treatment. Various stimuli are available for release [18] and these stimuli can be logically divided into physical, chemical and biological ones.

Endogenous biological stimuli are most often used for permeability [19] control as well as for performing a relatively slow release [20],[21] of pharmaceutical cargo based on enzymatic degradation [22, 23]. One example is enzyme-catalyzed carrier degradation, which can also be accomplished using sophisticated coupled mechanisms [24]. Another class of release mechanisms, at least for in-vitro experiments, is based on chemical reactions, i.e. chemical stimuli. A number of chemical stimuli such as differences in salt concentration, solvent types or pH levels [25] can be used for inducing cargo release. While chemical stimuli can be quite effective and quick, they are typically not suited for in-vivo applications. In this regard, physical stimuli such as light, magnetic fields and mechanical forces are more promising for application in a biological context since they have little or no effect on the surrounding medium and can be well-controlled due to possibilities to adjust the stimulating fields’ strength. Laser light has a special niche in this area as an external physical stimulus since it can be very well controlled spatially and temporally so that high resolution imaging is possible for diagnostics with highly localized controlled activation of the therapeutic action. In addition, also from a research perspective it is of interest since it can be easily combined with existing optical microscopes, thus allowing detailed investigation of light-carrier interaction in a biological environment.

In what follows we will describe approaches for theranostics in which multicompartmentalization of microcapsules can be logically applied for diagnostics and treatment.

Diagnostics and Treatment

Diagnostics using materials encapsulated in some compartments and therapy performed by drugs encapsulated in other compartments is the cornerstone of theranostic applications. In order to conduct the diagnostics part effectively, it is essential to develop and apply appropriate imaging. Here one would clearly distinguish two types of imaging modalities: a) those used in cell culture work, and b) those applicable in-vivo. Microscope-based approaches prove to be the most effective ones for cell cultures. Here contrast agents are incorporated into some compartments of the carrier according to the intended imaging modality to be used. Cell labeling is achieved by fluorophores, which permit visualization of cytoskeleton, nucleus, and other subcompartments as well as visualization of individual molecules with super-resolution. An interesting application is label-free, Raman imaging [26]. A number of other approaches can be used for in-vivo imaging; these include: X-ray, ultrasound, computer tomography (x-ray based), positron emission tomography, magnetic resonance imaging, and their combination often referred to as multimodal imaging.

Diagnosing various parameters can be intended for prevention or treatment, the latter being an inherent part of theranostics. Many different parameters can be diagnosed including acidity (pH) and ionic contents [27], glucose and other biomarkers, products of enzyme-catalyzed reactions, host-pathogen and other biomolecular interactions, bacteria, etc. Some of these parameters, like pH, can provide very important information on functions of cells, organs, and eventually the body. And these parameters can be monitored by molecules encapsulated into different subcompartments of multicompartmental capsules. Also, with regard to therapeutic applications, some of the above methods can be used not only for direct visualization, but also for activation and/or release of active bio-molecules from diagnostic subcompartments of multicompartmental capsules. The treatment modality is administered based upon results obtained from the diagnostic assays.

The following sections describe various approaches describing different types of multicompartmental microcapsules and methods of remote control [28] for on-site specific release.

Multicompartment microcapsules

Multicompartment microcapsules have been designed almost specifically with theranostics in mind. Diversity of materials for encapsulation into different compartments and variety of multicompartment morphologies and structures make multicompartmentalization a very versatile approach. Various different approaches can be used for assembling multicompartment capsules, namely: a) assembly from previously pre-formed different subcompartments; b) direct synthesis of capsules (or templates on which capsules are assembled); c) a combination of synthesis and assembly from previously pre-formed subcompartments [1].

Here we will discuss four of the most important multicompartmentalization morphologies, as illustrated in Figure 1. First, concentric structures can be obtained in which multiple compartments are built (concentrically) around the first compartment, see Figure 1 (a) [1]. The first compartment can be built around a particle or a capsule, while subsequent concentric subcompartments are further built around this
first block. Synthetic routes (for example, calcium carbonate) are used in this case for fabrication. This process can be separated in several steps. First the inner core is synthesized, which can be used as a precursor for outer concentric shells that are synthesized consequently. During further synthesis of concentric subcompartments around the primary subcompartment (or the inner core), some single compartment cores can also appear [29]: then functionalization of concentric multicompartment microcapsules by magnetic nanoparticles can be used for separation purposes.

The second type of multicompartment capsules is the so-called pericentric type, Figure 1 (b) [1]. In this approach, multiple compartments are adsorbed or connected (possibly chemically) to the first and main carrier capsule. This is a versatile approach amenable for mass production since it is based on a relatively simple assembly of pre-fabricated particles/capsules. The interaction of subcompartments takes place due to electrostatic forces, steric entrapment or hydrogen bonding. To this end, subcompartments are typically modified by oppositely charged polymers or molecules capable of inducing the above interactions.

The third type of multicompartment capsules are innercentric, in which case various compartments are incorporated into the interior of the first and main compartment, see Figure 1 (c) [1]. The sub compartments can be synthesized during the assembly process of this type of multicompartment capsules. Although the feasibility was already demonstrated, the relative complexity of this approach is a potential obstacle for widespread use.

In the fourth and last approach acentric structures are prepared and described [1, 14]. Here, the subcompartments can be either synthesized during the assembly process of the multicompartment capsules, or they might be already pre-formed. Many different implementations of such an approach are possible [30]. Its key attribute is non-uniformity or non-symmetry of the structure. Figure 1 (d) shows several structures wherein two different subcompartments are connected, thus forming an elongated two-compartment capsule.

**Figure 1.** Overview and the road-map for future directions of multicompartment microcapsules. Four different approaches are identified in the schematics: a) concentric, b) pericentric, c) innercentric, and d) acentric. The structure in the middle incorporates all four approaches. The corresponding confocal microscope images of the first steps in each direction are also presented. Scale bars correspond to 2 mm. [1] Reproduced by permission of Wiley-VHC.
Anisotropic multicompartment microcapsules

Anisotropy of drug delivery vehicles, as in the case for the acentric capsules discussed above, is an important characteristic for theranostics. Anisotropy of delivery carriers is a very desirable feature for pulmonary drug delivery [31] because elongated carriers possess the same aerodynamic diameter as an equivalent spherical carrier but allows for delivery of a larger amount of cargo. Porosity of carriers is another desirable feature that is of importance for optimizing the aerodynamic diameter as it allows tuning the effective density. In this regard, calcium carbonate particles or hydrogels[32] are attractive candidates.

Early attention to anisotropic particles was given by De Gennes providing a significant impetus for development. Particles in which one half is distinct from the other are called Janus particles [33]. Chemical synthesis and modification are two of the most common routes of controlling [34] the patchiness [35] of Janus particles. Using these routes, various anisotropic structures and approaches were reported using microfluidics [36], electroless deposition [37], micro-contact printing [38], grafting [39], and plasma polymerization [40].

Anisotropic multicompartment structures have been recently reported using a method of partially shielding a lower part of particles/capsules incorporated into soft films [30]. In such an approach, particles or capsules are sedimented onto the films leaving the upper part non-protected. In the subsequent step, (typically) smaller particles/capsules are adsorbed onto the non-protected part of embedded particles. Extraction of embedded particles is done by turning the films upside-down and adding a solvent (sodium hydroxide of higher pH for hyaluronic acid/poly-L-lysine films). The solvent loosens the interaction between the films and capsules/particles, thus allowing the latter to detach and be collected.

Controlling protrusion of microcapsules into the soft polymeric films and thus controlling the degree of modification or patchiness [41] can be achieved by making the films harder or denser [34], see Figure 2.

Figure 2. Fluorescence CLSM images showing an overview of a-f) half covered (4.8 μm SiO2)@QDs (quantum dots) and g-l) 2/3 covered (SiO2)@QDs. Dotted lines represent the centre line, while bold lines show the termination of particle modification. Images a) and g) show top view, while c-f) and i-l) depict side view of protruding SiO2 particles into FITC labeled (HA/PLL)12 films with AuNP concentration of a-f) 0 μl and g-l) 300 μL. Arrows in a) and g) point to location of particles; insets in a) and g) show magnification of the transmission images of protruding (SiO2) particles into (HA/PLL)12 films. Arrows in b) and h) indicate the (HA/PLL)12 film height. Scale bars correspond to 5 μm. [34] Reproduced by permission of Wiley-VHC.
This modification can be done by chemical cross-linking or by adding metal nanoparticles onto the surface of the films. While particles can still be embedded into the denser films, they penetrate over a smaller distance [34]. Thus, the masked part of the particles or the degree of patchiness can be controlled through the stiffness of films. This property has been measured by the AFM colloidal probe technique [42, 43]. Peculiarly, such a modification also promotes growth of cells (which grow more effectively on harder substrates) on these soft PLL/HA films as it was recently shown by Schmidt [44]. Anisotropic morphology of particles can be also used for direction-specific release of drug molecules. In such an approach if only a specific part of the microcapsules is functionalized by nanoparticles for triggered release, then release will take place only at the site of nanoparticles. A proof-of-principle of such an approach has been recently demonstrated for giant microgel templated beads [45].

**Enzymatic and ionic sensing**

The next step following assembly of the multicompartment capsules is the incorporation of sensing modalities for diagnosis. One can discriminate between two major approaches. The first is encapsulation of larger biomolecules which would serve as sensors for ions. Ions are very small entities which easily penetrate the polyelectrolyte polymeric network. It is this principle that was used for fabrication of polyelectrolyte multilayer capsules: ions penetrating inside the polymeric shell dissolve the template upon which microcapsules are built. Ions are therefore constantly present at equilibrium inside and outside capsules. Monitoring ions allows for instance to assess local pH, which is a very important physiological parameter. For monitoring ions, larger (typically above 1 kD) fluorescent sensor molecules are encapsulated into the interior of sensing subcompartments. These molecules change their fluorescence characteristics (intensity and/or spectrum) which can be easily observed using fluorescence microscopy [27].

An alternative route for sensing is that based on the detection of products of enzyme-catalyzed reactions. This approach has some similarities to that for the detection of ions: larger sensor molecules are encapsulated inside some subcompartments of multicompartment theranostic capsules. The polymeric mesh allows not only ions, but also small molecules to penetrate through the polymeric network. The larger enzymatic signal molecules change their (often fluorescent) properties which allows for detection of products of enzyme catalyzed reactions [46].

**Therapeutic treatment and stimuli for inducing release from capsules**

Therapeutic drugs, such as peptides, proteins and biomolecules are encapsulated into capsules with a polyelectrolyte polymeric mesh. This polymeric mesh permits ions and small molecules to go through, but protects encapsulated drugs which are to be used for treatment. These therapeutic molecules will need to be released [47, 48] if the diagnostic signal necessitates so.

Release of molecules can be performed using different physical stimuli, including electromagnetic fields, ultrasound, mechanical force and temperature, Figure 3. Some stimuli possess certain limitations. For example, temperature (global temperature in solution, cell culture or in the body) cannot be substantially changed, thus limiting its application range. Mechanical force is applicable only at specific conditions permitting exerting the force. On the other hand, exerting mechanical forces can be used for measuring the stiffness [49] of capsules and the threshold forces [20] necessary for release. This approach is ideally suited for bench tests of capsules, but it is difficult to exert pressure on capsules in-vivo.

Ultrasound [50] is an interesting physical stimulus for release and some initial work using ultrasound has been conducted to initiate enzyme-catalyzed reactions. Enzyme and substrate were assembled into the same multicompartment microcapsule: the enzyme was held in the interior compartment, while the substrate was incorporated into liposomes attached to the surface of the pericentric multicompartment microcapsules [51]. Upon application of ultrasound, the substrate leaked from the liposomes, penetrated into the interior of the inner particle, initiating the enzyme-catalyzed reaction, Figure 4. An alternative approach is using microcapsules with freely-floating liposomes [52] or multicompartment particles [9], Figure 5. As an example, this approach can be used to fight cancer. An interesting feature of this approach is that both the enzyme and substrate are encapsulated in the same carriers. Calcium carbonate (in the vaterite form) particles used to demonstrate the feasibility of such an approach are biocompatible, porous, and easy to make. Most importantly, it was observed that enzymes immobilized in the pores of particles, were still fully active [51]. Although some progress on reducing the intensity of ultrasound has been recently achieved, a significant hurdle here is to achieve ultrasound mediated release at a power density that is medically acceptable.
Figure 3. Schematics showing various methods of release. The three main categories: physical, biological and chemical, are further sub-divided and are shown with specific methods of release. [18] Reproduced by permission of The Royal Society of Chemistry.

Figure 4. a) Evolution of the enzymatic reaction inside porous microparticles. b–d) Confocal transmission (middle row) and fluorescence (bottom row) images of microparticles after 1 min (b), 10 min (c), and 15 min (d) of ultrasonication and addition of H$_2$O$_2$. The insets show fluorescence profiles drawn across the same microcontainer (the red lines in the bottom row). The red profiles in the insets correspond to emission 1 min after sonication, the yellow and white profiles to emission 10 and 15 min after sonication, respectively. [51] Reproduced by permission of Wiley-VHC.
Electromagnetic energy, such as laser light, can be used as a trigger signal as well. The wavelength used is an important parameter since absorption of laser light by biomolecules is wavelength dependent. UV wavelengths are not useful since penetration in tissue is too limited due to scattering and absorption by biomolecules. Visible light typically cannot be used either since it is already needed for exciting the fluorescent moieties used for sensing and diagnosis. Laser light in the near-infrared range (the so-called biologically “friendly” window [53]) can pass relatively well through water (or a biological medium) so that a reasonable penetration depth in tissue can be obtained. This allows noble metal nanoparticles, such as gold nanorods [54], gold nanoparticles [55] or even carbon nanotubes [56], to be used for exerting a therapeutic action as they can have a strong absorption in the near-infrared range due to the surface plasmon resonance effect. More information on laser-induced release from capsules can be found in the next section.

**Laser-induced remotely activated release of encapsulated materials**

Light energy can be converted into heat [57, 58] by nanoparticles [59, 60]. The amount of heat generated can be carefully controlled by tuning the laser intensity, illumination time and the surrounding environment [60]. This effect has been used to induce release from microcapsules [57, 61-63], Figure 6. For example, when irradiated by a continuous wave laser, the nanoparticles will act as local heaters, causing a heat gradient in the surrounding tissue. Here it is important to distinguish between a “global” versus “local” temperature rise. As it was mentioned above it is undesirable to raise the global temperature of biological matter. However, under appropriate illumination conditions, heating can be locally induced without affecting the cell or tissue. It is this localized temperature rise that has been used for inducing transient permeability [64] of the polymeric shell of capsules, thus achieving controlled release of the pharmaceutical cargo. Increasing the concentration of nanoparticles or augmenting the laser power density can lead to increased heat accumulation and therefore to explosive release or activation of microcapsules. It can also be used for hypothermia wherein extensive accumulation of heat is desirable for the killing of cancer cells [65].

Localized, mild temperature rise can be used to obtain a non-destructive permeability change of the capsule walls [64]. An advanced feature here is the possibility of a transient permeability change when the polyelectrolytes of the capsule walls are in the so-called “glassy” state. Localized heating slightly exceeding the “glass” transition temperature renders the polymeric complex flexible. The osmotic pressure of encapsulated materials is enough to push some
molecules through the polymeric shell. Upon turning off the laser beam, the polymeric complex goes back into the “glassy” state sealing the polymeric shell anew. Thus, the permeability of microcapsules can be regulated transiently upon switching laser on and off.

Using laser induced permeability control, release of encapsulated materials has been accomplished inside living cells by microcapsules [53] and directly from particles [55]. Using intracellular release inside living cells, the immune response has been investigated wherein peptides released from the interior of microcapsules were found to form the complex with MHC Class I polymers eliciting movement to the cell surface and the surface presentation [66]. This process is one of the key principles of immune system defense. Cell viability following capsule uptake and triggered release has been verified either by observing cell division or by specific essays, thus proving the feasibility and potential of such an approach for future in vivo use. It can be noted that either stabilized nanoparticles [67] or direct synthesis [68] of nanoparticles on the surfaces can serve to achieve desirable distribution of nanoparticles.

Vapor nanobubble induced by laser nanoparticle interaction in water and biological media have recently proven to offer exciting possibilities, such as the highly specific intracellular delivery [69] and processing [70] of therapeutics. Vapor nanobubbles can be induced by pulsed [71] or continuous wave [72] laser irradiation: the former case is certainly a very promising approach for drug delivery, hypothermia treatment or material modification. Short laser pulses possess specific advantages [73]: high laser peak power and better dissipation of heat. Generating bubbles by continuous wave laser irradiation has been shown applicable for affective soft, polymeric films [72]. A short pulse laser (typically in the order of nanoseconds [74] or less) illuminating metal nanoparticles such as gold nanoparticles leads to extreme localized heating of these nanoparticles due to the excitation of surface plasmon resonance. As a consequence, the spinodal temperature of the surrounding medium, such as water, can be easily surpassed and causes the nucleation of vapor at the nanoparticle surface and the formation of transient nanobubbles. When the thermal energy in consumed, the vapor nanobubble will collapse, causing local pressure waves that can damage nearby tissue. The size of the vapour nanobubbles can be precisely controlled through the laser pulse fluence. It was shown that vapor nanobubbles can aid in drug delivery, cancer treatment and gene therapy.

Figure 6. Confocal microscope images demonstrating remote release of encapsulated rhodamine-labeled PSS polymers from a polyelectrolyte multilayer capsule containing gold sulfide core/gold shell nanoparticles in its walls. Fluorescence intensity profiles along the line through the capsule show that it is filled with fluorescent polymers before (a) and empty after (b) laser illumination. After the release of encapsulated polymers, the leftover fluorescent intensity is observed only in the walls of the capsule, (b). Insets show black and white transmission microscope images of the same capsule. Incident intensity of laser diode operating at 830 nm was set at 50 mW. Reproduced with permission from [57]. Copyright (2005) American Chemical Society.
Bright perspectives for multicomartment capsules for theranostics

It is envisioned that theranostics will take a unique place in the nanomedicine field in the near future. Precise diagnostics and unique custom initiated, on-demand treatments are the hallmarks of personalized medicine so vigorously being developed now. The use of multicompartment capsules would furthermore allow driving the development of theranostics to another level, Figure 7. A lot of effort is being put in exploring the concept of multimodality, where either different imaging or different therapeutic modalities can be exploited in a combined manner so that any intrinsic shortcomings of a single modality can be compensated for by the strengths of another. Using multicompartment capsules would allow the same entity to be bestowed with several types of imaging contrast agents together with several types of drugs. The different trigger systems also allow the incorporation of various types of therapeutics which can then be selectively released, depending on the opportune time and conditions. As an example, the multicompartment capsules could simultaneously or in a specific order release several anti-HIV drugs to form a more potent cocktail than a single entity. Also, when the same drug would be incorporated in different shells, the use of various stimuli at different time points would allow a repeated release of high levels of the drug at the target site, which could offer many advantages over a single bolus injection or repeated injections of the drug. Further research and developmental work, including in vivo studies, will surely contribute to its wide applicability in biomedicine.

Further assembly of microcompartment capsules can be based not only on synthetic carriers, but also based on red blood cells [75, 76] from which release has also been recently demonstrated [77].

Conclusions

Theranostics is an emerging area which combines diagnostics and therapy. It will allow physicians not only to detect diseases, but will also permit them to adequately administer medicines, thus paving the way to personalized medicine. Multicompartment microcapsules are ideal candidates for application in theranostics owing to the multifunctionality by the constituting subcompartments. Indeed, therapeutic compounds can be incorporated inside some subcompartments, while other subcompartments can be filled with sensing agents, which are to be used for sensing and diagnosis. A particular attractive property of such elaborate drug delivery carriers is that they allow for diagnostics and subsequent treatment by implementing release of drugs at a desired site and rate. Laser light, in combination with noble metal nanoparticles, has proven to be a particularly attractive for achieving localized triggered release. Further developments of light sensitive drug carriers, and capsules in general [78], are expected to impact the area of theranostics and nanomedicine in the future.

Figure 7. Schematics of implementation of theranostic multicompartmen capsules: diagnostics is achieved by monitoring pH, glucose, lactose, etc., while therapy is attained by release of encapsulated drugs/medicine.
Acknowledgments

We thank FWO (Fonds Wetenschappelijk Onderzoek), the Belgian Research Council, for support of this work.

Competing Interests

The authors have declared that no competing interest exists.

References

[1] Delcea M, Yashchenok A, Videnova A, Kreft O, Mohwald H, Skirtach AG. Multicompartimental micro- and nanocapsules: hierarchy and applications in biosciences. Macromol Biosci. 2010; 10: 465-474.
[2] Berg MC, Zhai L, Cohen RE, Rubner MF. Controlled drug release from porous polyelectrolyte multilayers. Biomacromol. 2006; 7: 357-364.
[3] Chow AHL, Tong HHY, Chattopadhyay P, Shekunov BY. Particle engineering for pulmonary drug delivery. Pharm Res. 2007; 24: 1134-1138.
[4] Cui JQ, Kretzschmar I. Surface-anisotropic polystyrene spheres by multilayer assembly. Langmuir. 2008; 24: 11276-11276.
[5] Delcea M, Madaboosi N, Delcea M, Schmied S, De Geest BG, Mohwald H, Skirtach AG. Multicompartmentalized multilayer microparticles: hierarchy and applications in biosciences. Macromol Biosci. 2010; 10: 465-474.
