A case of Gitelman syndrome with membranous nephropathy

Xiafei Guo¹, Shanshen Yu¹, Jia Sun*¹ and Lijun Mou*²

Abstract

Background: Gitelman syndrome (GS) is a rare autosomal recessive inherited salt-losing tubulopathy (SLT). Here, we report, for the first time, a case of GS overlapping nephrotic syndrome (NS) related to PLA2R-associated membranous nephropathy (MN).

Case presentation: We described a male patient had a 4-year history of recurrent fatigue. Serum biochemistry revealed hypokalemia with renal potassium wasting, hypomagnesemia, metabolic alkalosis, hyperreninemia, hypocalciuria, as well as nephrotic-range proteinuria, hypoalbuminemia, and elevated serum anti-phospholipase A2 receptor (PLA2R) antibody. Gene sequencing identified compound heterozygous mutations in SLC12A3 [c.536T > A(p.V179D) and c.1456G > A(p.D486N)]. The unusual association of SLTs and nephrotic-range glomerular proteinuria prompted us to perform a renal biopsy. Renal biopsy showed idiopathic MN. Due to the potential to activate the sodium-chloride co-transporter (NCC) and cause hyperkalemia, tacrolimus was selected to treat NS. Following treatment with potassium chloride, magnesium oxide, low-dose glucocorticoid combined with tacrolimus, the fatigue significantly improved, and concurrently hypokalemia, hypomagnesemia were corrected and NS was remitted.

Conclusions: Renal biopsy should be warranted for GS patients with moderate to nephrotic-range proteinuria. Tacrolimus was preferred to the management of GS patients with NS.

Keywords: Gitelman syndrome, SLC12A3, Membranous nephropathy, Tacrolimus

Background

Gitelman syndrome (GS) is a rare autosomal recessive inherited salt-losing tubulopathy (SLT) because of the inactivating mutations in SLC12A3 gene, which encodes the sodium-chloride cotransporter of distal convoluted tubules. GS is characterized by chronic hypokalemia, metabolic alkalosis, hypomagnesemia and hypocalciuria. GS typically is not classically associated with proteinuria, especially nephrotic-range glomerular proteinuria [1, 2]. Therefore, when nephrotic-range proteinuria developed in patient with GS, overlapping glomerular diseases should be considered. PLA2R-associated membranous nephropathy (MN) is an auto-immune disease characterized by moderate to nephrotic-range proteinuria, a common cause of nephrotic syndrome (NS) in adults [3, 4]. Up to now, GS with concomitant MN has not been reported. We described one patient with GS overlapping MN. He was successfully treated with tacrolimus and glucocorticoid, potassium and magnesium supplementation.

Case presentation

A 24-year-old male patient was admitted to our hospital on Sep 1st, 2020 due to recurrent limb fatigue for four years and aggravating for one month. Four years ago, the patient presented limb weakness episode once sweating a lot in every summer, hypokalemia was revealed in a
local hospital. Following potassium chloride supplement during hospitalization, fatigue was relieved. However, he stopped taking potassium chloride after discharge, and fatigue recurred following sweating in every summer.

One month ago, the patient suffered from limb weakness again after tooth extraction. Serum biochemistry in local hospital showed that potassium was 2.8mmol/L and albumin was 22 g/L; urinalysis demonstrated 3+ proteinuria. Therefore, he was admitted to our hospital. The patient denied taking diuretics, laxatives, diet pills and Chinese herbal medicines; His parents are healthy and non-consanguineous marriage; his two sisters are healthy.

Blood pressure after admission was 108/77mmHg. While this patient had no oedema and other physical examination abnormality. Laboratory tests showed hypokalemia due to renal potassium wasting, hypomagnesemia, hypochloridemia, metabolic alkalosis, hyperreninemia, hypocalciuria (Table 1), which were suggestive of GS. Thiazide test was performed according to the protocol described in the previous study [5]. The difference value between the chloride excretion fraction before and after the use of hydrochlorothiazide was 0.468%, indicating that he was no response to hydrochlorothiazide, which could be functionally diagnosed of GS. Therefore, Next-Generation sequencing (NGS)-based panel was performed to identify the exact type of SLTs. The method of gene sequencing was performed as previously reported [6]. Two missense heterozygous mutations [c.536T > A(p.V179D) and c.1456G > A(p.D486N)] in SLC12A3 were revealed. His father carries one mutation (c.536T > A) and his mother carries the other mutation (c.1456G > A), which are compatible with compound heterozygosity in this patient (Fig. 1).

![Gene analysis results. A, C. The patient and his father carry a c.536T>A (p.V179D) (arrow) heterozygous mutation in SLC12A3 gene, respectively and his mother has no mutation in SLC12A3 gene at this site. B, D. The patient and his mother carry a c.1456G>A (p.D486N) (arrow) heterozygous mutation in SLC12A3 gene, respectively and his father had no mutation in SLC12A3 gene at this site.](image-url)
The mutation of \textit{SLC12A3} (p.V179D) was predicted to be benign by PolyPhen-2, polymorphism by Mutation Taster, and damaging by SIFT. This mutation has been reported by Lee, et al. \cite{7}. The mutation of p.D486N was predicted to be probably-damaging by PolyPhen-2, disease-causing by Mutation Taster, and damaging by SIFT. This mutation has been reported by Simon, et al \cite{8}. Both mutations are classified as likely pathogenic according to American College of Medical Genetics (ACMG) guidelines. Eventually, GS was identified.

24 h proteinuria showed nephrotic-range proteinuria, he also has hypoalbuminemia. Results of ANA, ENA, anti-dsDNA, anti- thyroid antibodies, imaging of chest/abdomen were negative. Taken together, nephrotic syndrome (NS) was identified. Therefore, renal biopsy was performed. Light microscopy showed 18 glomeruli, including 3 sclerotic ones, with thickened basement membrane, spike-like structures and subepithelial deposition of erythrophilic proteins.; The immunofluorescence results of renal biopsy showed that IgG (3+), IgG1 (3+), IgG4 (3+), C3 (+) and PLA2R (2+) were granular deposition along capillaries; Electron microscopy showed irregular thickening of glomerular basement membrane with the thickest part of 1500 nm, diffuse foot process effacement, and deposition of electron dense substance in subepithelial and basement membrane. The serum anti-phospholipase A2 receptor (PLA2R) antibody was also positive. Eventually, diagnosis of PLA2R-associated MN was established (Fig. 2).

According to the patient’s history, laboratory tests and gene sequencing findings, the final diagnosis was GS concomitant with MN. Following immunosuppressant with methylprednisolone tablets (12 mg once daily) plus tacrolimus (1 mg twice daily), electric supplement with potassium chloride tablets (3.0 g per day), magnesium oxide (300 mg magnesium per day), the level of serum potassium and magnesium increased, NS was partially remitted, anti-PLA2R titre reduced after 6 months (Table 1).

### Discussion and conclusions

GS is a rare autosomal recessive inherited SLT. American doctor Gitelman first reported 3 cases of familial hypokalemia, hypomagnesemia, hypocalciuria and metabolic

---

**Table 1** Comparison of patients before and after treatment in laboratory tests

| Examination item                  | 2021.09.02 | 2021.03.07 | Reference value |
|-----------------------------------|------------|------------|-----------------|
| Serum biochemistry                |            |            |                 |
| potassium (mmol/L)                | 3.00       | 4.09       | 3.50 ~ 5.50     |
| magnesium (mmol/L)                | 0.57       | 0.65       | 0.73 ~ 1.06     |
| calcium (mmol/L)                  | 2.01       | 2.48       | 2.08 ~ 2.60     |
| sodium (mmol/L)                   | 144.8      | 143.3      | 135.0 ~ 145.0   |
| chloride (mmol/L)                 | 101.9      | 101.5      | 96.0 ~ 106.0    |
| creatinine (μmol/L)               | 63         | 93         | 40 ~ 106        |
| albumin (g/L)                     | 24.5       | 44.9       | 35.0 ~ 52.0     |
| cholesterol (mmol/L)              | 5.21       | 5.47       | 3.0 ~ 5.7       |
| triglycerides (mmol/L)            | 4.70       | 1.86       | < 1.7           |
| PLA2R (RU/mL)                     | 25.4       | 2.1        | < 2.00          |
| Upright plasma renin (UIU/mL)     | 294.1      | -          | 4.4 ~ 46.1      |
| Upright plasma aldosterone (pg/mL)| 108.0      | -          | 30.0 ~ 353.0    |
| Arterial blood gas analysis       |            |            |                 |
| PH value                          | 7.442      | -          | 7.350 ~ 7.450   |
| Base excess (mmol/L)              | 6.3        | -          | -3.0 ~ 3.0      |
| HCO3⁻ (mmol/L)                    | 30.9       | -          | 22.0 ~ 26.0     |
| PaCO2 (mmHg)                      | 46         | -          | 36.0 ~ 44.0     |
| Urine analysis                    |            |            |                 |
| proteinuria (g/L)                 | 2+         | 1+         | negative        |
| hematuria (number/ul)             | 5          | < 1        | < 12            |
| Albumin to creatinine ratio (mg/g.Cr) | 3083.3  | 356.3      | < 25.0          |
| 24 h proteinuria (mg/24 h)        | 4838.4     | -          | 22.0 ~ 132.0    |
| 24 h potassium (mmol/24 h)        | 57.60      | -          | 25 ~ 100        |
| Spot urinay calcium/creatinine (mmol/mmol) | 0.022 | -          | -               |
| Spot urinay potassium /creatinine (mmol/mmol) | 8     | -          | -               |
alkalosis in 1966. The cause of GS is the inactivating mutations of SLC12A3 gene, which encodes the sodium-chloride co-transporter of distal convoluted tubules [8]. GS are characterized by hypokalemia, hypomagnesemia, metabolic alkalosis, hypocalciuria, secondary renin-angiotensin-aldosterone (RAAS) activation, normal or low blood pressure [9]. GS patients may manifest fatigue, salt craving, polydipsia, polyuria, and paroxysmal tetany triggered by hypomagnesemia [10]. It is suggested GS patient intake potassium and magnesium supplements and sodium chloride-containing food for life [11], as GS is caused by the deficiency of sodium-chloride co-transporter.

However, nephrotic-range glomerular proteinuria in GS patients is rare generally. At present, there are 6 cases of GS accompanied by moderate to nephrotic-range proteinuria [12–17]. Two of the patients were female and four were male; Three patients presented with nephrotic syndrome, two of whom received glucocorticoid therapy; 5 patients underwent renal biopsy, including C1q nephropathy, minimal change disease (MCD), diabetic nephropathy and 2 cases of focal segmental glomerulosclerosis (FSGS) (Table 2).

Therefore, renal biopsy should be warranted for GS patients with moderate to nephrotic-range glomerular proteinuria. If there are primary glomerular diseases, glucocorticoid or other drugs may be added, in addition to potassium and magnesium supplements for GS [12, 17].

So is there a correlation between GS and glomerular proteinuria? It has been reported that the possible

| Case                  | Sex  | Age | Proteinuria | Serum albumin | Edema | Renal pathology | Use of glucocorticoids |
|-----------------------|------|-----|-------------|---------------|-------|------------------|------------------------|
| Hanevold C, et al. [13]| Female | 12  | 2.5 mg/mg   | 3.5 gm/dL    | No    | C1qN             | Not used               |
| Pandey DB, et al. [14]| Female | 19  | 1.445 gm/d  | 2.6 gm/dL    | Yes   | N/A              | Glucocorticoid         |
| Ceri M, et al. [15]   | Male  | 32  | 1094 mg/d   | N/A           | NO    | FSGS             | Not used               |
| Demoulin N, et al. [16]| Male  | 27  | 770 mg/d    | N/A           | N/A   | FSGS             | N/A                    |
| Chen Q, et al. [17]   | Male  | 47  | 10.20 g/d   | 22 g/L        | N/A   | MCD              | Glucocorticoid         |
| Chen Q, et al. [12]   | Male  | 40  | 2793 mg/d   | N/A           | N/A   | DN               | Not used               |

N/A Not applicable, C1qN C1q Nephropathy, FSGS Focal segmental glomerulosclerosis, MCD Minimal change disease, DN Diabetes nephropathy
mechanisms between GS and glomerular proteinuria is the chronic activation of RAAS, leading to increased systemic and local levels of angiotensin-II and renin, may in turn cause podocyte lesions. Angiotensin II induced proteinuria through vascular endothelial growth factor and transforming growth factor-β1 (TGF-β1) [12, 16]. At present, abnormal heterogeneity of basement membrane thickness and disappearance of podocyte foot processes, as well as decreased expressions of nephrin and podocin, have been observed in transgenic mice overexpressing renin. TGF-β1 is considered the major pro-fibrotic agent in renal disease [18]. Nephrotic range proteinuria was also present in patients with Addison's disease, a disease associated with hyperreninemia, and renal biopsy showed FSGS, nodular deposition of IgM, and C3 [19]. Severe intraglomerular detachment of podocytes was described in another case of GS before, which was also associated with decreased renal nephrin expression [20]. Chronic hypokalemia may also cause proteinuria, Reungjui et al. detected mild proteinuria in hypokalemic rat models with or without hydrochlorothiazide. For the same degree of hypokalemia, renal injury was more obvious in the hydrochlorothiazide treated group, which was attributed to the secondary hyperaldosteronism due to long-term volume depletion [21]. According to the above studies, there is similar RAAS activation caused by blood volume depletion in GS patients. Therefore, there may be a correlation between GS and FSGS.

PLA2R-associated MN is an auto-immune disease, characterized by non-inflammation mediated subepithelial immune complex deposition with diffuse thickening of glomerular basement membrane. Approximately 70-80% of patients with primary MN have circulating PLA2R antibodies [22]. Renal pathology of this patient we described was MN, with increased titer of serum PLA2R antibody. Therefore, it was an immune related disease rather than a metabolic related disease. By above knowable, we consider that FSGS may be related to the pathophysiological changes of GS in GS patients with nephrotic-rangeproteinuria, while other pathological changes may be irrelevant to GS, including PLA2R-associated MN, which involves an autoimmune response. In addition, GS is commonly diagnosed in children and young adults, such as the patient described, MN is untypical at the age of the patient, which still suggest the association between GS and PLA2R-associated MN is probably coincidental. At this time, renal biopsy is supposed to be necessary.

In the treatment, we chose a low-dose glucocorticoid with tacrolimus and supplements of potassium chloride and magnesium oxide. 2017 Kidney Disease: Improving Global Results (KDIGO) guidelines recommended that the initial dose of potassium chloride supplement for adults should be 40 mmol (equivalent to 3.0 g) per day, and suggests that the serum potassium and magnesium levels of GS patients should be maintained at 3.0 mmol/L and 0.6 mmol/L at least, respectively [23]. We gave the patient an initial dose of 3.0 g potassium chloride per day, the serum potassium easily increased to 4.09mmol/L consequently. We found that serum potassium of the patient exceeded the target value of KDIGO, and the proteinuria was effectively controlled through treatment. It has been reported that tacrolimus has the potential to increase the activity of phosphorylated NCC and NCC regulated kinases WNK3, WNK4 and SPAK, resulting in over activation of NCC, leading to hyperkalemia, similar to Gordon syndrome or familial hyperkalemic hypertension [24, 25]. This may explain why the patient’s serum potassium is easy to surpass the standard by far.

In conclusion, we report a case of GS comorbid with MN. Renal biopsy should be warranted for GS patients with moderate to nephrotic-range glomerular proteinuria in order to guide treatment. In addition, due to the potential to activate NCC and cause hyperkalemia, tacrolimus may have more advantages in the treatment of GS patients with NS secondary to PLA2R-associated MN.

Abbreviations
GS: Gitelman syndrome; SLT: salt-losing tubulopathy; NS: nephrotic syndrome; MN: membranous nephropathy; PLA2R anti-phospholipase A2 receptor; NCC: sodium-chloride co-transporter; NGS: next-generation sequencing; ACMG: American College of Medical Genetics; RAAS: renin-angiotensin-aldosterone; MCD: minimal change disease; FSGS: focal segmental glomerulosclerosis; TGF-β1: transforming growth factor-β1; KDIGO: Kidney Disease:Improving Global Results.

Acknowledgements
Not applicable.

Authors’ contributions
XG and SY were responsible for designing and data analysis, XG contributed to write the first draft of manuscript, and SY assisted in translating the manuscript into English and editing. JS analyzed pathological and mutational data. LM participated in the treatment as primary physician, contributed to the collection patient's data, drafted the manuscript; JS and LM were responsible for analyzing patient's data. All authors have read and approved the manuscript.

Funding
This research was funded by the Key Research and Development Program of Ningxia Hui Autonomous Region (2018BFG02010). The funding body only provided the financial means, and played no role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript.

Availability of data and materials
Data regarding this study were obtained from clinical charts stored in the physician office records of Second Affiliated Hospital of Zhejiang University School of Medicine, therefore, cannot be shared. Any reasonable request to access the data must be approved before the data can be released. The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.
Declarations

Ethics approval and consent to participate
Ethical approval was obtained from the ethics committee of Second Affiliated Hospital of Zhejiang University School of Medicine. All procedures in this study involving consent to participate were performed in accordance with the ethical standards of the institutional ethics and research committee. Written informed consent was obtained from the patient and his family members for participation in this study.

Consent for publication
Written informed consent was obtained from the patient and his family members who participated in our study for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor of this journal.

Competing interests
The authors declared that they have no competing interests.

Received: 18 January 2022 Accepted: 3 July 2022
Published online: 26 July 2022

References

1. Nakhoul F, Nakhoul N, Dorman E, Berger L, Skorecki K, Magen D. Gitelman’s syndrome: a pathophysiologic and clinical update. Endocrine. 2012;41(1):53–7.
2. Matsunoshita N, Nozu K, Shono A, Nozu Y, Fu X, Morisada N, et al. Differential-diagnosis of Bartter syndrome, Gitelman syndrome, and pseudo-Bartter/Gitelman syndrome based on clinical characteristics. Genet medicine official J Am Coll Med Genet. 2016;18(2):180–8.
3. Sethi S, Madden B, Debiec H, Morelle J, Charlesworth M, Gross L, et al. Protecachelin 7-Associated Membranous Nephropathy. J Am Soc Nephrol: JASN. 2021;32(5):1249–61.
4. von Groote T, Williams G, Au E, Chen Y, Mathew A, Hodson E, et al. Immunosuppressive treatment for primary membranous nephropathy in adults with nephrotic syndrome. Cochrane Database Syst Rev. 2021;11:CD004293.
5. Colussi G, Bettinelli A, Tedeschi S, De Ferrari ME, Syren ML, Borsa N, et al. A thiazide test for the diagnosis of renal tubular hypokalemic disorders. Clin J Am Soc Nephrol. 2007;2(3):454–60.
6. Mou L, Wu F. Simultaneous Homozygous Mutations in SLC12A3 and CLCNKB in an Inbred Chinese Pedigree. Genes. 2021;12(3):369.
7. Lee J, Lee J, Heo N, Cheong H, Han J. Mutations in SLC12A3 and CLCNKB and Their Correlation with Clinical Phenotype in Patients with Gitelman and Gitelman-like Syndrome. J Korean Med Sci. 2016;31(1):47–54.
8. Simon DB, Nelson-Williams C, Bia MJ, Ellison D, Karet FE, Molina AM, et al. Gitelman’s variant of Bartter’s syndrome, inherited hypokalaemic alkalosis, is caused by mutations in the thiazide-sensitive Na–Cl cotransporter. Nat Genet. 1996;12(1):24–30.
9. Group GSCS. Expert consensus for the diagnosis and treatment of patients with Gitelman syndrome. Chin J Intern Med. 2017;56(9):712–6.
10. Uzunulu M, Dumanoğlu B. Gitelman syndrome presenting with hypomagnesemia, hypokalemia and hypocalciuria - a case report. Medeniyet Med J. 2019;34(3):214–7.
11. Francini F, Gobbi L, Ravarotto V, Tonazio S, Nalesso F, Spinella P, et al. The Dietary Approach to the Treatment of the Rare Genetic Tubulopathies Gitelman’s and Bartter’s Syndromes. Nutrients. 2021;13(9):2960.
12. Chen Q, Wang X, Min J, Wang L, Mou L. Kidney stones and moderate proteinuria as the rare manifestations of Gitelman syndrome. BMC Nephrol. 2021;22(1):12.
13. Hanevold C, Mian A, Dalton R. Clcq nephropathy in association with Gitelman syndrome: a case report. Pediatr Nephrol. 2006;21(12):1904–8.
14. Pandey D, Pednekar S, Chavan S, Korvi D, Shah A, Kulkarni U. A case of nephrotic syndrome with Gitelman’s syndrome. J Assoc Phys India. 2010;58:324–5.
15. Cenci M, Unverdi S, Altay M, Unverdi H, Kurultak I, Yilmaz R, et al. Focal segmental glomerulosclerosis in association with Gitelman syndrome. Int Urol Nephrol. 2011;43(3):905–7.
16. Demoulin N, Aydin S, Cosyns J, Dahan K, Corret N, Auburger I, et al. Gitelman syndrome and glomerular proteinuria: a link between loss of sodium-chloride cotransporter and podocyte dysfunction? Nephrol Dial Transplant. 2014;29 Suppl 11:172–120.
17. Chen Q, Wu Y, Zhao L, Jia Y, Wang W. A case of hypokalemia and proteinuria with a new mutation in the SLC12A3 Gene. BMC Nephrol. 2018;19(1):275.
18. Huby A, Rastaldi M, Caron K, Smithies O, Dussaule J, Chatziantoniou C. Restoration of podocyte structure and improvement of chronic renal disease in transgenic mice overexpressing renin. PLoS ONE. 2009;4(8):e6721.
19. Arizabalaga P, Bergadá E, Solé M, Halperin J, Botej A. Focal segmental glomerulosclerosis in two patients with Addison’s disease: any more than fortuitous development of glomerular disease? Am J Nephrol. 2002;22(4):389–93.
20. Takahashi N, Kimura H, Mizuno S, Hara M, Hirayama Y, Kurosawa H, et al. Severe intra-glomerular detachment of podocytes in a Gitelman syndrome patient. Clin Exp Nephrol. 2012;16(3):495–500.
21. Reungsi S, Hu H, Mu W, Roncal-C, Croker BP, Patel JM, et al. Thiazide-induced subtle renal injury not observed in states of equivalent hypokalemia. Kidney Int. 2007;72(12):1483–92.
22. Bobart S, Han N, Tehrani S, De Vriese A, Roman J, Sethi S, et al. Non-invasive Diagnosis of PLA2R-Associated Membranous Nephropathy: A Validation Study. Clinical journal of the American Society of Nephrology: JASN. 2021.
23. Blanchard A, Bockenhauer D, Bolignano D, Calò LA, Cosyns E, Devuyst O, et al. Gitelman syndrome: consensus and guidance from a kidney disease: Improving Global Outcomes (KDIGO) Controversy Conference. Kidney Int. 2017;91(1):24–33.
24. Hoom E, Walsh S, McCormick J, Furstenberg A, Yang C, Roeschel T, et al. The calcineurin inhibitor tacrolimus activates the renal sodium chloride cotransporter to cause hypertension. Nat Med. 2011;17(10):1304–9.
25. Farouk S, Rein J. The Many Faces of Calcineurin Inhibitor Toxicity-What the FK? Adv Chronic Kidney Dis. 2020;27(1):56–66.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.