Glioma SOX2 expression decreased after adjuvant therapy

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Abstract

**Background:** SOX2 is regarded as an important marker in stem cell. The change of SOX2 expression after adjuvant therapy in high grade glioma (HGG) remains unknown so far. Few patients with recurrent glioma have opportunity to undergo operation once again, so the recurrent glioma samples are scarce. This study tries to analyze SOX2 expression in paired primary and recurrent HGG, aims to better understand the transformation law of SOX2 after adjuvant therapy in HGG.

**Methods:** Twenty-four recurrent HGG patients who undergone a second resection were included. 16 patients received adjuvant therapy, the remaining 8 patients didn’t receive any adjuvant therapy at all. The protein expression of SOX2 in paired primary and recurrent HGG was tested by immunohistochemistry. The statistical analysis was conducted by IBM SPSS Statistics 19.0.

**Results:** In primary HGG, SOX2 expression of 3 +, 2 +, 1+ and 0+ were seen in 20 (83.3%), 1 (4.2%), 1 (4.2%) and 2 cases (8.3%), respectively. The expression of SOX2 was decreased in recurrent HGG compared to the paired primary sample (p = 0.001). The decrease of SOX2 was often seen in patients received chemotherapy, radiotherapy or both (p = 0.003). Patients with SOX2 high expression in primary glioma had a longer median PFS than those with SOX2 low expression with marginal statistic significance (12.7 vs. 5.4 months, p = 0.083). For cases with SOX2 high expression in the primary glioma, those had SOX2 low expression after recurrence seemed to have worse prognosis as compared to patients with stable SOX2 high expression (PFS: 10.4 vs. 14.9 months, p = 0.036; OS: 27.0 vs 49.5 months, p = 0.005).

**Conclusions:** This is the first study comparing the protein expression of SOX2 in recurrent HGG and its paired primary tumor. SOX2 high expression is common in brain HGG, a tendency of decreased SOX2 expression in recurrent gliomas was evidenced. Lower SOX2 expression was seen in those patients who received adjuvant chemotherapy and/or radiotherapy. Patients with low SOX2 expression in primary HGG usually have poorer prognosis, those with SOX2 expression decreased in recurrent HGG had worse outcome.

**Keywords:** Glioma, Recurrence, Prognosis, SOX2
Background

Glioma is the most common primary intracranial tumor, and it’s usually subdivided into WHO I to IV grades [1]. Grade I-II gliomas are considered as low grade gliomas (LGGs). The prognosis of LGGs is relatively better with a 5 to 10 years’ median overall survival (OS) [2–5]. However, 50–75% patients with LGGs would die of disease progression or deteriorate into higher grade gliomas [4]. Grade III-IV are called high grade gliomas (HGGs). HGGs are characterized by high recurrence rate and dismal survival. WHO IV grade glioma, named glioblastoma multiforme (GBM), occupy the highest proportion in glioma, with the highest malignant degree and poorest prognosis [1, 6]. Currently, standard care for newly diagnosed glioblastoma is maximum safe resection of the malignancy followed by radiotherapy (RT) and temozolomide (TMZ) chemotherapy. Grave median survival of GBM was reported as 14.6 months despite state-of-the-art treatment [7]. Almost all glioblastoma recurred inevitably.

SOX2 (SRY-related HMG-box 2) is a key member of transcription factor SOX family. It is mainly expressed in embryonic and adult stem cells, including neural progenitor cells, etc. It can also be expressed in tumor stem cells, such as glioma stem cells. SOX2 can stimulate somatic cell transform into pluripotent stem cell combined with other stem cell markers, such as NANOG and OCT, and also maintain the characteristics of cancer stem cells. It is a glioma stem cell marker [8]. The expression of SOX2 is associated with tumor formation, chemotherapeutic resistance and the tumor stem cell like phenotype [9, 10]. Schmitz and other studies have found that SOX2 was overexpressed in malignant gliomas and its expression in normal brain tissue was rare [11]. Gangemi et al. reported that silencing the SOX2 gene in tumor initiating cells of glioma can prevent cell proliferation, resulting in the loss of tumorigenicity in immunodeficient mice [12]. Ikushima et al. showed that inhibition of TGF-beta signaling can significantly reduce the tumorigenesis of glioma initiating cells by promoting differentiation, and this effect attenuated in SOX2 transduced tumor initiating cells [13]. Therefore, SOX2 gene is the key gene for the maintenance of the characteristics of glioma stem cells.

Few patients have opportunity to undergo operation once again. The data used to compare the primary glioma and its paired recurrent tumor is scarce. SOX2 expression of recurrent gliomas remains unknown so far. In this study, we collected 24 cases with paired sample of primary HGGs and corresponding recurrent tumor after a second resection. We attempt to analyze the expression of SOX2 protein of primary HGG and its paired recurrent tumor in order to better understand the biological behaviour of HGG and the transformation law of SOX2 in primary and recurrent HGG, and eventually enrich our knowledge of HGG.

Methods

Patients

This study was consented by the Institutional Review Board of the Second Affiliated Hospital, Zhejiang University School of Medicine (SAHZU). Informed consent was obtained from all the patients before surgery regarding the data and samples to be used for research. Patients had undergone first operation for HGG at SAHZU from Jan. 2008 to Dec. 2014 were retrospectively analyzed, 24 recurrent patients with second surgery were included. Histopathology diagnosis were achieved after surgery and diagnosed by neuropathologists. The medical records were retrospectively reviewed.

Immunohistochemistry

Formalin-Fixed and Paraffin-Embedded (FFPE) blocks from the pathology department of SAHZU were cut into serial 4 um slices with a microtome. The paraffin sections were incubated in 60°C in incubator overnight. Then the sections were deparaffinized in xylene and rehydrated through graded alcohols (100, 95, 75%). The sections were put in boiled antigen retrieval solution (EDTA, pH = 9) in electric cooker for 10 min and heat preserved for 10 min. Then the sections were incubated with 100ul SOX2 primary antibody (ZSGB-BIO, China) at room temperature for 2.5 h. After removing the primary antibody, the sections were incubated with 100ul ready-to-use secondary antibody (Polink-1 HRP staining system (ZSGB-BIO)) at room temperature for 2.5 h. After removing the secondary antibody, the sections were incubated with chromagen 3, 3’-diaminobenzidine (DAB) at room temperature for 1.5 min. The sections were counterstained with hematoxylin for 3 min and dehydridation through graded alcohols (75, 95, 100%). Then the slides were covered slips with mounting medium.

Scoring system

Immunohistochemical staining evaluation of SOX2 protein was based on the range of staining. The proportion of immunopositive cells among the total number of tumor cells was subdivided into 4 categories, as follows: 0: < 10%; 1: 10–50%; 2: 50–90%; 3: > 90% [14]. Two pathologists read the pathological sections and were blinded to the clinical information and histological grades.

Statistical analysis

Chi-square test and Fisher’s exact test were used among subgroups to compare enumeration data such as sex, WHO grade, tumor location, adjuvant therapy, residue, and type of recurrence. P < 0.05 was considered significant. For the ranked data of paired primary and recurrent samples, Wilcoxon rank sum test was used to analyze the variation trend of SOX2 expression.
Correlation between SOX2 expression and WHO classification were tested by Spearman rank correlation test. Overall survival (OS) is defined as the time from the first operation of glioma to the death of the patient. Progression-free survival (PFS) is defined as the time from the first operation of glioma to the progression of the tumor. Kaplan Meier method and Cox proportion hazard regression model were used to estimate PFS and OS for univariate and multivariate analysis. P < 0.05 was statistically significant. IBM SPSS Statistics 19.0 was used for statistical analyses.

Results
Patient clinical characteristics
24 HGG patients with pathologically confirmed and complete data who received second operation when recurred after first operation were included. There were 16 males and 8 females. The average age at first operation was 45.4 years (range: 12–60 years old). Seventeen cases had gliomas in frontal lobe, 5 in temporal lobe, 1 in parietal lobe and 1 in insula lobe. At the first operation, 14 patients underwent subtotal resection and the other 10 cases total gross resection. After the first operation, 12 cases were diagnosed as grade III gliomas, 12 cases were glioblastomas. 22 cases were IDH1 wild type, 2 cases were IDH1 mutant type which were both SOX2 high expression, their PFS and OS were both longer than the medium level (PFS: 20 & 13.5 months; OS: 49.5 & 57.0 months). Twelve patients received radiotherapy and chemotherapy after the first operation, 3 patients only received radiotherapy, 1 patient only received chemotherapy, and the rest 8 patients had not adjuvant therapy at all.

At the second operation for recurrent tumors, 18 patients underwent subtotal resection and 6 cases total gross resection. In recurrent gliomas, 6 cases were grade III gliomas, 18 cases were glioblastomas. Six cases deteriorated from grade III gliomas to glioblastomas. After the operations for recurrent tumors, 4 cases received both radiotherapy and chemotherapy, 1 patient only received radiotherapy, 7 patients chemotherapy alone, and the rest 12 cases had no more adjuvant therapy.

The deadline for follow-up was August 18, 2017. The overall median PFS was 12.7 months (95% CI: 10.0–15.4 months): 14.9 months (95% CI: 9.0–20.8 months) for the primary grade III gliomas and 10.3 months (95% CI: 7.5–13.0 months) for the primary glioblastomas. At the end of the follow-up visit, 4 patients were alive. The median OS was 30 months (95% CI: 22.2–37.8): 31.0 months (95% CI: 26.6–35.4) for the primary grade III gliomas and 24.4 months (95% CI: 22.2–37.8 months) for the primary glioblastomas. The median OS after second operation was 18.1 months (95% CI: 13.2–23.0).

SOX2 expression in primary glioma
In primary HGGs, SOX2 expression of 3+, 2+, 1+ and 0+ were seen in 20 (83.3%), 1 (4.2%), 1 (4.2%) and 2 cases (8.3%), respectively. The corresponding data for grade III glioma were 9 (75.0%), 1 (8.3%), 1 (8.3%) and 1 cases (8.3%), and for glioblastoma were 11 (91.7%), 0 (0.0%), 0 (0.0%) and 1 cases (8.3%) (Fig. 1).
Decreased SOX2 protein expression in recurrent gliomas compared with their paired primary tumors

The overall expression of SOX2 decreased (p = 0.001). In recurrent HGGs, SOX2 expression of 3+, 2+, 1+ and 0+ were seen in 7 (29.2%), 7 (29.2%), 4 (16.7%) and 6 cases (25.0%), respectively. The corresponding data for grade III gliomas were 3 (50.0%), 2 (33.3%), 0 (0.0%) and 1 cases (16.7%), and for glioblastomas were 4 (22.2%), 5 (27.8%), 4 (22.2%) and 5 cases (27.8%) (Fig. 1).

Compared with primary high grade tumors, SOX2 expression increased in their paired recurrent gliomas in 0 cases (0.0%), while 13 cases (54.2%) decreased, 11 cases (45.8%) did not change. In patients with grade III in primary gliomas and developed into glioblastoma in recurrent gliomas, 1 cases (1/6, 16.7%) with SOX2 expression decreased and 5 cases (5/6, 83.3%) were stable. In patients with grade III in primary and paired recurrent gliomas, 2 cases with SOX2 expression decreased (2/5, 40.0%) with SOX2 expression decreased, 3 cases (3/5, 60.0%) were stable. In patients with glioblastoma in primary and paired recurrent gliomas, 10 cases (10/11, 90.9%) with SOX2 expression decreased, 1 cases (1/11, 9.1%) were stable (Fig. 2). Figure 3 show the expression of SOX2 in primary and its paired recurrent glioma in a typical patient.

Effect of postoperative adjuvant therapy on the expression of SOX2

Out of the 24 patients, 16 patients received adjuvant therapy including radiotherapy, chemotherapy and chemoradiotherapy after first surgery (adjuvant group), the other 8 patients received no adjuvant therapy at all (non-adjuvant group). Compared with primary gliomas, the overall expression of SOX2 decreased in adjuvant group (p = 0.003), but such tendency was largely not seen in non-adjuvant group (p = 0.317) (Table 1).

SOX2 expression correlates with survival of Glioma

Patients were grouped into SOX2 high expression group (2+ and 3+, n = 21) and SOX2 low expression group (0+ and 1+, n = 3) according to the expression of SOX2 in primary gliomas. The comparison of baseline clinical characteristics between those 2 groups was seen in Table 2. Univariate analysis showed that the median PFS was longer in SOX2 high expression group than in the SOX2 low expression group (12.7 vs. 5.4 months, p = 0.083, but p value had no significant difference (Fig. 4a). The median OS was also longer in SOX2 high expression group than in the SOX2 low expression group with significant difference 33.6 vs. 18.3 months, p < 0.001) (Fig. 4b). In patients with SOX2 high expression in primary glioma, 13 cases (54.2%) changed to SOX2 low expression after recurrence, the prognosis of these patients seemed worse than patients with stable SOX2 expression (PFS: 10.4 vs. 14.9 months, p = 0.036; OS: 18.5 vs 32.8 months, p = 0.249) (Fig. 4c, d). In patients with IDH1 wild type, the median PFS and OS was also longer in SOX2 high expression group than in the SOX2 low expression group (PFS: 12.5 vs 5.4, p = 0.098; OS: 32.8 vs 18.3, p = 0.001) (Fig. 4e, f). Multivariate analysis showed that SOX2 was an independent prognostic factor (Table 3).
For the small sample size to analyze the prognostic value of SOX2, we further searched The Cancer Genome Atlas (TCGA) database and found SOX2 mRNA expression in 153 glioma cases (Additional file 1: Table S1). SOX2 low expression also predicted poor survival (Fig. 5).

Discussion
This is the first study comparing the protein expression of SOX2 in recurrent HGG and its paired primary tumor. SOX2 high expression is common in brain gliomas, a tendency of decreased SOX2 expression in recurrent HGG was evidenced. Lower SOX2 expression was seen in those patients who received adjuvant chemotherapy and/or radiotherapy. Patients with low SOX2 expression in primary HGG usually have poorer prognosis, those with SOX2 expression decreased in recurrent glioma had worse outcome.

In our case series, about 83.3% of the primary HGG cases were SOX2 high expression. Elsir et al. [14] and Ballester et al. [15] reported a SOX2 high expression rate of 97.8 and 43.5% in primary HGGs. In the study by Guo et al. [16], western blot and RT-PCR were performed to evaluate the expression of SOX2, 95% of the gliomas expressed SOX2 at both the mRNA and protein

Table 1 Effect of postoperative adjuvant therapy on the expression of SOX2

| Group(n)                      | Primary (%) | Recurrence (%) |
|-------------------------------|-------------|----------------|
|                               | 0+ | 1+ | 2+ | 3+ | 0+ | 1+ | 2+ | 3+ | P*  |
| No adjuvant therapy n = 8     | 1  | 0  | 1  | 6  | 1  | 2  | 4  | 0.317 |
| Adjuvant therapy n = 15       | 1  | 1  | 0  | 14 | 3  | 5  | 3  | 0.003 |
| Chemo-radiotherapy n = 12     | 1  | 1  | 0  | 10 | 5  | 2  | 3  | 2   |
| Radiotherapy n = 3            | 0  | 0  | 3  | 0  | 1  | 1  | 1  |     |
| Chemotherapy n = 1            | 0  | 0  | 1  | 0  | 0  | 1  | 0  |     |

*Wilcoxon rank sum test was used to analyze the change of SOX2 expression between paired primary and recurrent samples

Table 2 Comparison of baseline clinical characteristics between SOX2 low expression and high expression group in primary glioma

| Characteristics      | SOX2 high expression (n = 21) | SOX2 low expression (n = 3) | P value |
|----------------------|-------------------------------|-----------------------------|---------|
| Age (year)           |                               |                             |         |
| Mean (range)         | 45(22–60)                     | 49(43–53)                   | 0.373   |
| Sex                  |                               |                             |         |
| Male                 | 14                            | 2                           |         |
| Female               | 7                             | 1                           | 1.000   |
| WHO grade            |                               |                             |         |
| III                  | 10                            | 2                           |         |
| IV                   | 11                            | 1                           | 0.546   |
| IDH1 status          |                               |                             |         |
| Wild type            | 19                            | 3                           |         |
| Mutated              | 2                             | 0                           | 1.000   |
| Resection type       |                               |                             |         |
| Total gross resection| 8                             | 2                           |         |
| Subtotal resection   | 13                            | 1                           | 0.55    |
| Adjuvant therapy     |                               |                             |         |
| after first surgery  |                               |                             |         |
| Radiotherapy         | 3                             | 0                           |         |
| Chemotherapy         | 1                             | 0                           |         |
| Chemoradiotherapy    | 10                            | 2                           |         |
| No adjuvant therapy  | 7                             | 1                           | 0.266   |
| Type of recurrence   |                               |                             |         |
| Local recurrence     | 19                            | 1                           |         |
| Distant recurrence   | 2                             | 1                           | 0.343   |
| Median PFS (month)   | 12.7                          | 5.4                         | 0.083   |
| Median OS (month)    | 33.6                          | 18.3                        | < 0.001 |
Fig. 4 Correlation between the expression of SOX2 and prognosis. 

a) Correlation between the expression of SOX2 and PFS.

b) Correlation between the expression of SOX2 and OS.

c) Correlation between the decreasing expression of SOX2 and PFS in patients with SOX2 high expression in primary HGG.

d) Correlation between the decreasing expression of SOX2 and OS in patients with SOX2 high expression in primary HGG.

e) Correlation between the expression of SOX2 and PFS in primary glioma with IDH1 wild type.

f) Correlation between the expression of SOX2 and OS in primary glioma with IDH1 wild type.
levels. The results in our study are inconsistent with the previous reports.

The protein expression of SOX2 in recurrent glioma has not been reported. For the first time, we presented that SOX2 expression decreased in recurrent glioma as compared to the corresponding primary glioma. It has been known that among proneural, mesenchymal and proliferative subtypes, the prognosis of the proneural subtype is better than the other two subtypes [17]. Verhaak et al. found that SOX2 expression was mainly in the proneural subtype and was rarely expressed in the mesenchymal and proliferative subtypes [18]. While the recurrent glioma tended to transform into the mesenchymal subtype [19, 20]. Wang et al. found that low

Table 3 Results of multivariate analyses with the Cox proportional hazard models

| Variable                        | Overall survival | Progression-free survival |
|---------------------------------|------------------|----------------------------|
|                                 | HR(95% CI)       | P  | HR(95% CI)     | P  |
| Age                             | 1.060            | 0.08 | 1.000          | 1.000 |
| Sex                             | 0.641            | 0.438 | 3.070          | 0.122 |
| Adjuvant therapy after first surgery | 1.121            | 0.852 | 1.459          | 0.563 |
| Resection type                  | 1.171            | 0.799 | 0.460          | 0.244 |
| SOX2                            | 0.215            | 0.076 | 0.160          | 0.045 |
| WHO grade                       | 1.722            | 0.427 | 11.606         | 0.001 |
| Type of recurrence              | 2.301            | 0.377 | 1.166          | 0.866 |

Fig. 5 Correlation between the mRNA expression of SOX2 and OS in TCGA database
miR-21/high SOX2 group tended to express in proneuronal and classical genotypes, while most of the high miR-21/low SOX2 group belongs to mesenchymal phenotype [21]. Therefore, decreased SOX2 expression in recurrent glioma might probably due to the tumor transformation from the proneural subtype into the mesenchymal subtype.

To study the influence of adjuvant therapy on the expression of SOX2, we further conducted subgroup analysis according to the adjuvant therapy patients received after first operation. A novel finding was that patients received adjuvant chemotherapy/radiotherapy usually had decreased SOX2 expression in the recurrent tumor. While those without adjuvant therapy had largely the same SOX2 expression in the recurrence as compared to the primary tumor.

This study indicated that low expression of SOX2 in primary HGG is a negative prognosticator although with marginal statistic significance. The prognostic value of SOX2 in HGG remained controversial so far. Sathyan et al. found that glioblastoma patients with low miR-21/high SOX2 expression had longer overall survival than those with high miR-21/low SOX2 (15.3 vs. 16.7 months, \( p = 0.0088 \)), which was in accordance with our results. While Wang et al. reported that expression of SOX2 predicted poor prognosis [22]. It has been reported that expression of SOX2 related to promoter methylation [23–29]. This might allow the transposition of SOX2 protein testing to DNA methylation testing, which could be performed in cerebral spinal fluid, in less invasive mode, using cell free circulating DNA. Though testing SOX2 methylation status from cerebral spinal fluid, we can predicted the prognosis of the patients and choose the appropriate treatment strategy.

In the primary SOX2 high expression gliomas, 2 cases had IDH1 mutation, their PFS and OS were both longer than the medium level (PFS: 20 & 13.5 months; OS: 49.5 & 57.0 months). While the primary SOX2 low expression cases were IDH1 wild. It has been reported that IDH1 mutation predicted better prognosis in glioma [30, 31]. In these two cases, IDH1 mutation status and SOX2 expression were consistent to predict better prognosis. To eliminate the influence of IDH1 to the prognosis, we further analyzed the the correlation between SOX2 expression and prognosis in IDH1 wild type group. Finally, we found SOX2 low expression still had worse PFS and OS than SOX2 high expression group.

Twenty four HGG cases with both primary tumor samples and recurrent samples were analyzed in the present study. It is not a large patient series. However, few patients with recurrent glioma have opportunity to undergo operation once again, it is not easy to get the paired samples. Our study provides precious knowledge about SOX2 status in the recurrent gliomas, which further enrich our understanding to glioma.

**Conclusions**

This study firstly compared the protein expression of SOX2 in primary and its paired recurrent HGG. The expression of SOX2 decreased in recurrent gliomas compared with the primary gliomas. Lower SOX2 expression was seen in those patients who received adjuvant therapy. Low expression of SOX2 in primary HGG predicts poor prognosis. Those patients with SOX2 expression decreased in recurrent glioma often had worse prognosis.

**Supplementary information**

Supplementary information accompanies this paper at https://doi.org/10.1186/s12885-019-6292-y.

**Abbreviations**

GBM: Glioblastoma multiforme; HGG: High grade glioma; LGG: Low grade glioma; OS: Overall survival; RT: Radiotherapy; SAHZU: Zhejiang University School of Medicine; SOX2: SRY-related HMG-box 2; TCGA: The Cancer Genome Atlas; TMZ: Temozolomide

**Acknowledgments**

The authors would like to thank Professor Shu Zheng for organizing the logical flow of the manuscript and Dr. Yang Tang for help with experiments.

**Authors’ contributions**

Wy participated in the design of the study, did the immunohistochemistry experiment, carried out the clinical data analysis, and wrote the manuscript; XR participate in the immunohistochemistry experiment; CH interpreted the experimental and clinical data and wrote the manuscript. All authors read and approved the final manuscript.

**Funding**

This work was supported by the National Natural Science Foundation of China (81572952). The funding had no role in the design of the study and collection, analysis, and interpretation of data and in the writing of the manuscript.

**Availability of data and materials**

The datasets used and analysed during the current study are available from the corresponding author on reasonable request.

**Ethics approval and consent to participate**

This study was approved by the Ethics Committee of SAHZU, and was carried out in accordance with the Declaration of Helsinki. Written informed consent was obtained from all patients.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

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Received: 20 August 2019 Accepted: 25 October 2019
Published online: 12 November 2019

References
1. Zhang AS, Ostrom QT, Kruchko C, Rogers L, Peerboom DM, Barnholtz-Sloan JS. Complete prevalence of malignant primary brain tumors registry data in the United States compared with other common cancers, 2010. Neuro-Oncology. 2017;19(5):726–35.
2. Piepmeier J, Baehring JM. Surgical resection for patients with benign primary brain tumors and low grade gliomas. J Neuro-Oncol. 2004;69(1–3):55–65.
3. Leighton C, Fisher B, Bauman G, Deppieri S, Stitt L, Macdonald D, et al. Supratentorial low-grade glioma in adults: an analysis of prognostic factors and timing of radiation. J Clin Oncol. 1997;15(4):294–301.
4. Keles GE, Lamborn KR, Berger MS. Low-grade hemispheric gliomas in adults: a critical review of extent of resection as a factor influencing outcome. J Neurosurg. 2003;99(5):735–45.
5. Gannett DE, Wisbeck WM, Silbergeld DL, Berger MS. The role of postoperative irradiation in the treatment of oligodendroglioma. Int J Radiat Oncol Biol Phys. 1994;30(3):567–73.
6. Ostrom QT, Gittleman H, Kruchko C, Louis DN, Brat DJ, Gilbert MR, et al. Surveillance, epidemiology, and end results (SEER) 18 database (2004–2012, varying). J Neuro-Oncol. 2016;130(1):31–42.
7. Stupp R, Mason WP, van den Bent MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma: a randomized phase III study of the EORTC Brain Tumor Cooperative Group. N Engl J Med. 2005;352(10):987–96.
8. Rodda DJ, Chew JL, Lim LH, Yang B, Ng HH, et al. Transcriptional regulation of nanoG by OCT4 and SOX2. J Biol Chem. 2005;280(26):24731–7.
9. Singh S, Trevino J, Bora-Singhal N, Coppola D, Haura E, Altiok S, et al. EGFR/NF1. Cancer Cell. 2010;17(1):98–108.
10. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. Cell. 2006;126(4):663–76.
11. Schmitz M, Temme A, Senner V, Ebner R, Schwind S, Stevanovic S, et al. Genetic and epigenetic modifications of Sox2 contribute to the pattern of disease progression, and resemble stages in neurogenesis. J Neuropathol Exp Neurol. 2016;75(10):925–35.
12. Satyanayanan, Zinn PO, Mariessy AL, Liu B, Kamal MM, Singh SK, et al. Mir-21-Sox2 Axis delineates Glioblastoma subtypes with prognostic impact. J Neurosci. 2015;35(45):15097–112.
13. Wang B, Li M, Wu Z, Li X, Li YU, Shi X, et al. Associations between SOX2 and miR-200b expression with the clinicopathological characteristics and prognosis of patients with glioma. Exp Ther Med. 2015;10(1):58–64.
14. Russo MV, Esposito S, Tupone MG, Manzoli L, Airolidi I, Pompa P, et al. SOX2 boosts major tumor progression genes in prostate cancer and is a functional biomarker of lymph node metastasis. Oncotarget. 2016;7(11):12372–85.
15. Maehara R, Fujikura K, Takeuchi K, Akita M, Abe-Suzuki S, Karbanova J, et al. SOX2-silenced squamous cell carcinoma: a highly malignant form of esophageal cancer with SOX2 promoter hypermethylation. Mod Pathol. 2018;31(1):83–92.
16. Fang L, Zhang L, Wei W, Jin X, Wang P, Tong Y, et al. A methylation-phosphorylation switch determines Sox2 stability and function in ESC maintenance or differentiation. Mol Cell. 2014;55(4):537–51.
17. Alonso MM, Díez-Valle R, Manterola L, Rubio A, Liu D, Cortés-Sánchez N, et al. Genetic and epigenetic modifications of Sox2 contribute to the invasive phenotype of malignant gliomas. PLoS One. 2011;6(11)e26740.
18. Wong OG, Huo Z, Siu MK, Zhang H, Jiang L, Wong ES, et al. Hypermethylation of SOX2 promoter in endometrial carcinoma. Obstet Gynecol Int. 2010;2010:62504.
19. Otsubo T, Akiyama Y, Yanagihara K, Yusa Y. SOX2 is frequently downregulated in gastric cancers and inhibits cell growth through cell-cycle arrest and apoptosis. Br J Cancer. 2008;98(4):824–31.
20. Stolzenburg S, Rots MG, Beltran AS, Rivenbark AG, Yuan X, Qian H, et al. Targeted silencing of the oncogenic transcription factor SOX2 in breast cancer. Nucleic Acids Res. 2012;40(14):6725–40.
21. Slodzenburg S, Rots MG, Beltran AS, Rivenbark AG, Yuan X, Qian H, et al. Id2a and Id2b mutations predict longer survival and response to temozolomide in low-grade gliomas. Neurology. 2010;75(17):1560–6.
22. SongTao Q, Lei Y, Si G, YanQing D, HuiXia H, XueLin Z, et al. IDH1 or IDH2 mutations predict longer survival and response to temozolomide in secondary glioblastoma. Cancer Sci. 2012;103(2):269–73.

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