Chapter 10
Removal of Blood Amyloid As a Therapeutic Strategy for Alzheimer’s Disease: The Influence of Smoking and Nicotine

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Abstract Accumulation of amyloid β protein (Aβ) in the brain causes cognitive impairment in Alzheimer’s disease (AD). The nature of the relationship between smoking and AD or dementia has been controversial. However, a recent meta-analysis revealed that smoking is a risk factor for AD. With regard to nicotinic acetylcholinergic receptors (nAChRs), both AD and control patients that smoke have been reported to show an increase in 3H-cytisine (an α4β4 nAChR agonist) binding in the temporal cortex. The α7 nAChR is also a key factor in AD pathology, particularly in relation to internalization of Aβ. Furthermore, there are many reports showing the neuroprotective effects of nicotine. The internalization of Aβ may lead to Aβ clearance in the brain.

We hypothesized that an extracorporeal system that rapidly removes Aβ from the blood may accelerate Aβ clearance from the brain. We have reported that (1) several medical materials including hemodialyzers can effectively remove blood Aβ, (2) the concentrations of blood Aβs decreased during hemodialysis, (3) removal of blood Aβ enhanced Aβ influx into the blood (ideally from the brain), resulting in maintenance or improvement of cognitive function, and (4) Aβ deposition in the brain of hemodialysis patients was significantly lower than in controls. Smoking affected blood Aβ removal efficiencies and brain atrophy. We believe this Extracorporeal Blood Aβ Removal Systems (E-BARS) may contribute as a therapy for AD.

Keywords Alzheimer’s disease · Amyloid β · Aβ · Blood purification · Hemodialysis · Dialyzer · HDC · E-BARS
10.1 Introduction: Amyloid β Protein in Alzheimer’s Disease

One of the major pathological changes associated with Alzheimer’s disease (AD) is the deposition of amyloid β protein (Aβ) as senile plaques and an increase in Aβ peptides in the brain (Kuo et al. 1996; Selkoe 2001). There are several Aβ species in the brain and plasma that are approximately 4 kDa in weight such as the 40-amino acid Aβ1–40 and the 42-amino acid Aβ1–42. Aβ1–42 aggregates more easily and is more toxic (Hung et al. 2008), forming soluble Aβ oligomers that can cause synapse loss and affect long-term potentiation in hippocampal neurons (Walsh et al. 2002). One mechanism proposed to underlie the increase in brain Aβ is reduced Aβ clearance rather than enhanced Aβ production, particularly in sporadic AD cases. Aβ production in the brains of AD patients was reported to be similar to that of normal subjects, yet Aβ clearance from AD brains was approximately 30% lower than in controls (Mawuenyega et al. 2010). In other words, it may be possible to treat AD by increasing Aβ clearance from the brain.

Recently, an anti-Ab monoclonal antibody that selectively targets aggregated forms of Aβ, aducanumab, was reported to be effective in improving cognitive function and reducing the brain Aβ burden, as measured by brain Aβ imaging (Sevigny et al. 2016). Similarly to anti-Aβ antibodies (Hock et al. 2003; Sevigny et al. 2016), peripheral administration of albumin, another Aβ-binding substance, was effective in improving cognitive function in AD patients in a Phase 2 study, and is currently undergoing a Phase 3 trial in AD patients (Boada et al. 2009, 2016).

We hypothesized that the rapid removal of Aβ from the blood by an extracorporeal system (E-BARS; extracorporeal blood Aβ removal system) may act as a peripheral Aβ sink from the brain, as shown in Fig. 10.1 (Kawaguchi et al. 2010). Smoking could affect the blood flow in the brain resulting in a change in the excretion of Aβ from the brain into the blood.

10.2 Smoking, Nicotine, and AD

Determining the exact nature of the relationship between smoking and AD or dementia has been controversial. However, a recent meta-analysis revealed that smoking is a risk factor for AD, as described below. These controversial findings may be due to the mixed effects of smoke itself and components of tobacco such as nicotine.
10.2.1 Smoking and AD Prevalence

Sabia et al. (2008) reported that ex-smokers had a 30% lower risk of poor vocabulary and low verbal fluency. However, the correlation between smoking history and cognitive decline was inconsistent in longitudinal analysis. Despite this ameliorative effect of smoking on memory (Sabia et al. 2008), the risk of AD was reported to be unaffected by any measure of tobacco consumption (Garcia et al. 2010). Contrary to these favorable or neutral effects of smoking on dementia, there are many reports showing that smoking has a deleterious influence on AD risk. Lower AD risk was observed in alcohol drinkers of both genders who had never smoked (OR = 0.37, 95% CI: 0.21, 0.65), regardless of the presence of apolipoprotein E4 (APO\textsubscript{4}). Ott et al. (1998) showed that smokers had an increased risk of dementia (relative risk 2.2 [95% CI: 1.3–3.6]) and AD (relative risk 2.3 [95% CI: 1.3–4.1]) compared with never smokers, based on a study of 6870 people aged 55 years and older. Smoking was a strong risk factor for AD in individuals without the APO\textsubscript{4} allele (relative risk 4.6 [95% CI: 1.5–14.2]), but had no effect in participants with this allele (relative risk 0.6 [95% CI: 0.1–4.8]). By meta-analysis of 19 prospective studies with at least 12 months of follow-up, Anstey et al. (2007) concluded that elderly smokers had increased risks of dementia and cognitive decline. Current smokers at baseline, relative to never smokers, had risks of 1.79 (95% CI: 1.43, 2.23) for AD and 1.78 (95% CI: 1.28, 2.47) for vascular dementia. Compared to those who had never smoked, current smokers at baseline also showed greater
yearly declines in Mini-Mental State Examination scores over the follow-up period. Compared to former smokers, current smokers at baseline showed an increased risk of AD and an increased decline in cognitive ability (Anstey et al. 2007). Furthermore, Barnes and Yaffe (2011) reported that smoking was associated with a higher risk of AD (relative risk 1.59 [95% CI: 1.15, 2.20]), and that a 10% reduction in smoking prevalence could potentially lower AD prevalence by about 412,000 cases worldwide and by almost 51,000 cases in the USA, while a 25% reduction in smoking prevalence could potentially prevent more than 1 million AD cases worldwide and 130,000 cases in the USA.

**10.2.2 AD Pathology and Smoking**

Recently, an interesting animal study on AD pathology was reported that used cigarette smoke rather than administration of some components of tobacco such as nicotine. When APP/PS1 transgenic mice were exposed to smoke from cigarettes, AD pathology, such as Aβ deposition and the Iba1-labeled area indicating an inflammatory response, was enhanced in the cortex and hippocampus. This enhancement was observed in the high-dose smoking group but not in the low-dose group (Moreno-Gonzalez et al. 2013).

Contrary to the animal study, it has been reported that smoking reduces both soluble and insoluble Aβ1–40 and Aβ1–42 in the frontal cortex and Aβ1–40 in the temporal cortex and hippocampus in AD patients (Hellström-Lindahl et al. 2004).

**10.2.3 Nicotinic Acetylcholinergic Receptors and Aβs**

Regarding nicotinic acetylcholinergic receptors (nAChRs), both AD and control patients that smoked showed increased 3H-cytisine (an agonist of the α4β4 nAChR) binding in the temporal cortex (Hellström-Lindahl et al. 2004). Further, Aβ levels in the brain was reduced in this study. Therefore, these authors proposed that a selective nAChR agonist could be a novel protective therapy for AD.

The α7 nAChR is also a key factor in AD pathology, particularly in relation to internalization of Aβs. Soluble Aβ is known to bind to the α7 nAChR with high affinity (Wang et al. 2000). By in vitro experimentation with SH-SYSY cells, Yang et al. (2014) revealed that extracellular Aβ1–42 was internalized by the cells and accumulated in endosomes/lysosomes and mitochondria. This internalization was mediated through an α7 nAChR-dependent pathway related to the activation of p38 MAPK and ERK1/2. The authors proposed that blockade of the α7 nAChR may have a beneficial effect by limiting intracellular accumulation of amyloid in the AD brain, thereby representing a potential therapeutic target for AD.
However, there are many articles showing the neuroprotective effects of nicotine. The internalization of Aβ may lead to Aβ clearance from the brain. Akaike and Shimohama’s research group first demonstrated the neuroprotective effect of nicotine on Aβ toxicity (Kihara et al. 1997). Concomitant administration of nicotine with Aβ25–35 ameliorated the death of rat cortical neurons induced by Aβ toxicity. In addition, the selective α7 nAChR antagonist, α-bungarotoxin, blocked this neuroprotective effect of nicotine. This group also revealed that stimulation of the α7 nAChR protected neurons against Aβ-enhanced glutamate neurotoxicity via PI3K (Kihara et al. 2001). Shimohama’s research group reported that treatment of rat microglia with galantamine, an acetylcholinesterase inhibitor, significantly enhanced microglial Aβ phagocytosis via the nAChR pathway (Takata et al. 2010). This group also revealed early accumulation of CD68-positive microglia at Aβ deposition sites and gradual reduction of Aβ in an Aβ-injected AD mouse model, which indicates the importance of the α7 nAChR in microglia as a therapeutic target in AD (Matsumura et al. 2015).

10.3 Our Hypothesis of a Therapeutic System for AD by Removal of Blood Aβ

As described earlier, one mechanism proposed to underlie increased brain Aβ in AD is reduced Aβ clearance rather than an increase in Aβ production, particularly in sporadic AD cases. Therefore, it may be possible to treat AD by enhancing Aβ clearance from the brain. There are several known Aβ transporters such as those involved in the Aβ influx pathway from the brain into the blood; e.g., LRP1 or APOE (Donahue et al. 2006; Bell et al. 2007), and RAGE (Silverberg et al. 2010), which is also known to mediate an Aβ influx pathway into the brain. In addition, perivascular elimination of Aβ in brain capillaries has been proposed (e.g., Morris et al. 2014).

Aβ concentrations in the cerebrospinal fluid (CSF) of AD patients are almost 100 times higher than those in plasma. Aβ concentrations in the CSF in cases of AD are reported to be 7.4–42.7 ng/ml for Aβ1–40 and 0.12–0.67 ng/ml for Aβ1–42 (Schoonenboom et al. 2005). Concentrations in the plasma of AD patients are reported to be 190.1 ± 61.7 pg/ml for Aβ1–40 and 23.0 ± 15.5 pg/ml for Aβ1–42 (Lopez et al. 2008). In brief, there are large gradients with respect to Aβ concentrations between the brain and plasma. Therefore, removing Aβ from the blood could accelerate Aβ transfer from the brain, thereby reducing the Aβ burden in the brain.

Peripheral administration of Aβ-binding substances, such as anti-Aβ antibodies, non-immunogenic substances, and albumin, can reduce the Aβ burden in the brain. However, attempts to use Aβ-binding substances in the blood in a therapeutic context resulted in the formation of Aβ complexes with the binding substances inside the body, which were sometimes retained in the plasma for a long period of time (DeMattos et al. 2001). Aβ antibodies generated by passive immunization or by active immunization using synthetic Aβ peptides reduced the occurrence of senile...
plaques and somewhat improved cognitive impairment in AD patients (Schenk et al. 1999; Hock et al. 2003). Furthermore, non-immunogenic Aβ-binding substances, such as GM1 ganglioside or gelsolin, also decreased the Aβ burden in the brain when they were peripherally injected into mouse models of AD (Matsuoka et al. 2003). Currently, a clinical trial is in progress where AD patients are being treated using intravenous administration of albumin, an Aβ-binding substance (Boada et al. 2009). In this Phase 2 trial, plasma exchange (discard) removes the plasma of AD patients, which contains Aβ–albumin complexes, and a new albumin solution is introduced into the blood as a replacement solution; the results thus far suggest that this therapy has improved cognitive function in AD subjects. The Phase 3 trial is now also underway (Boada et al. 2016).

Based on these observations, the removal of Aβ from the blood could act as peripheral drainage and an Aβ sink from the brain. We proposed that the E-BARS, which transfers Aβ out of the body, may be useful as a therapy for AD (Kawaguchi et al. 2010) (Fig. 10.1). The rapid reduction of Aβ concentrations in the blood could act as a trigger to enhance Aβ excretion from the brain, resulting in cognitive improvement.

### 10.4 Definition of Aβ Removal Activities of the Devices

The Aβ removal activities assessed in our study were: (1) the removal rate for batch analysis in vitro, (2–1) the removal efficiency based on the concentration change at pre-/post-application of the Aβ removal device, (2–2) the reduction rate of Aβ in the whole blood circulation, and (2–3) the filtration rate. The definitions were as follows:

1. **Batch analysis in vitro:**

   Adsorptive materials were mixed with Aβ solutions or plasma and shaken for the designated time.

   $$\text{Removal Rate (\%) } = 100 \times \left(1 - \frac{A\beta \text{ concentration with materials at the designated time}}{A\beta \text{ concentration without adsorbents at the same time}}\right)$$

2. **Flow analysis in vitro and the hemodialysis session**

   2-1 The Aβ removal efficiency of a dialyzer was defined as follows:

   $$\text{Removal efficiency (\%) } = 100 \times \left\{1 - \frac{\text{concentration of } A\beta \text{ after leaving the dialyzer (device) at a designated time}}{\text{concentration of } A\beta \text{ before entering the dialyzer (device) at that time}}\right\}$$
2-2 The Aβ reduction rate for the experimental pool solution or the whole blood circulation was defined as follows:

\[
\text{Reduction rate (\%) = } 100 \times \left\{ 1 - \frac{A\beta \text{ concentration in the pool solution or whole blood circulation at a designated time}}{\text{Initial A\beta concentration in the pool solution or whole blood circulation}} \right\}
\]

2-3 The Aβ filtration rate of a dialyzer was defined as follows:

\[
\text{Filtration rate (\%) = } 100 \times \left\{ \frac{\text{concentration of filtrated A\beta solution at the designated time}}{\text{concentration of A\beta before the dialyzer at the same time}} \right\}
\]

10.5 Adsorption Devices for Blood Aβ Removal

To obtain suitable materials for the removal of blood Aβ, we firstly investigated adsorptive materials for therapeutic blood purification (apheresis). We employed six materials: hexadecyl-alkylated cellulose particles (HDC), used to remove β2-microglobulin in carpal tunnel syndrome; cellulose particles ligated with dextran sulfate (CLD); charcoal (CHA), which is commonly used therapeutically, for example, in hepatic failure; tryptophan-ligated polyvinyl alcohol gel (TRV), used in Guillain–Barré syndrome; and cellulose acetate particles and non-woven polyethylene terephthalate filter, used in ulcerative colitis. Among these materials, HDC and CHA demonstrated a removal rate of almost 99% for both Aβ1–40 and Aβ1–42 in batch analysis using synthetic Aβ peptides (Fig. 10.2) (Kawaguchi et al. 2010).

HDC is used in cases where there are complications associated with hemodialysis and, therefore, we were able to investigate Aβ concentrations before (pre, inlet of) and after (post, outlet of) HDC column in hemodialysis sessions. The high removal efficiency of HDC was maintained at approximately 50% for both Aβ1–40 and Aβ1–42 during a 4-h hemodialysis session, as shown in Table 10.1.

10.6 Blood Aβ Removal by Hemodialyzers in Hemodialysis

We previously reported that hemodialyzers showed high Aβ removal activity based on analyses of hemodialysis patients (Kitaguchi et al. 2011, 2015; Kato et al. 2012). Measurements of Aβ concentrations at pre (inlet of) and post (outlet of) dialyzers during hemodialysis sessions revealed that the hemodialyzers effectively removed both Aβ1–40 and Aβ1–42 from the plasma of non-diabetic patients. Figure 10.3 shows the Aβ concentrations at the inlet of the dialyzers (Pre) and the outlet of the dialyzers (Post) for each dialysis session (n = 57). The average removal efficiencies for
Aβ1–40 were 66.0% at the 1-h point and 52.0% at the 4-h point of the hemodialysis sessions. Those for Aβ1–42 were 61.1% and 49.2%, as shown in Fig. 10.3. The removal efficiency for Aβ1–40 was significantly higher than for Aβ1–42 both at 1 h and at 4 h of each dialysis session (p < 0.0001 for both time points). Each dialyzer maintained its removal efficiency during the entire dialysis session. This indicates that the dialyzers had sufficient capacity for Aβ removal during the 4-h treatment.

### Table 10.1  Removal efficiencies of HDC columns in hemodialysis

| Time points during a hemodialysis session | Aβ1–40 | Aβ1–42 |
|------------------------------------------|-------|-------|
| 1 h (n = 5)                              | 51.1 ± 6.6% | 44.9 ± 5.0% |
| 4 h (n = 4)                              | 46.1 ± 6.6% | 38.2 ± 5.8% |

Taken from Kawaguchi et al. (2010)

Aβ1–40 were 66.0% at the 1-h point and 52.0% at the 4-h point of the hemodialysis sessions. Those for Aβ1–42 were 61.1% and 49.2%, as shown in Fig. 10.3. The removal efficiency in for Aβ1–40 was significantly higher than for Aβ1–42 both at 1 h and at 4 h of each dialysis session (p < 0.0001 for both time points). Each dialyzer maintained its removal efficiency during the entire dialysis session. This indicates that the dialyzers had sufficient capacity for Aβ removal during the 4-h treatment.

### 10.7 Removal of Blood Aβs Evoked Influx of Aβs into the Blood

Due to the effective removal activity of the dialyzers during the hemodialysis sessions (Fig. 10.3), the concentrations of blood Aβs after 4-h hemodialysis would have been approximately 10% of the concentrations at the starting point if there had been no Aβ influx into the blood (“Calcd” in Fig. 10.4). However, observed
concentrations of blood Aβs (“Obsd” in Fig. 10.4) were not decreased compared to “Calcd.” The differences between “Obsd” and “Calcd” were attributed to Aβ influx into the blood. We calculated the influx based on the differential equation described previously (Kitaguchi et al. 2011). The results of this simulation of 37 non-diabetic hemodialysis patients are shown in Fig. 10.4.

Table 10.2 shows more detailed results of the simulation of Aβ influx with 30 non-diabetic hemodialysis patients (Kitaguchi et al. 2015). The average removal efficiencies at the 1-hr point of the hemodialysis sessions were 67.3% and 51.3% for Aβ_{1-40} and Aβ_{1-42}, respectively. Aβ influxes during 4-hr hemodialysis were calcu-
lated as 9243 ng and 719 ng for\( A_\beta^{1-40} \) and\( A_\beta^{1-42} \), respectively, which were around five times the level of pre-existing\( A_\beta \)s in the blood, that is, 1952 ng and 165 ng, just before hemodialysis.

A similar\( A_\beta \) influx into the blood was also observed in a rat study using HDC.

**10.8 Are the Influxes of\( A_\beta \)s into the Blood from the Brain?**

Recently, we reported that\( A_\beta \) accumulation in the brains of hemodialysis (HD) patients was significantly lower than that in age-matched non-hemodialysis controls, as assessed by histopathological studies (Sakai et al. 2016). Senile plaques stained with anti-\( A_\beta \) antibodies were observed more frequently in non-HD subjects and were either sparse or not seen at all in HD patients (Fig. 10.5). Regarding the ratio of senile plaques (plaque-positive/-negative subjects), there were significantly fewer neuritic and cored plaques in HD patients; only 5 of 17 HD patients showed neuritic plaques stained with 4G8 anti-\( A_\beta \) antibody, whereas 12 out of 16 non-HD subjects exhibited these plaques. These findings suggest that the brain may be one origin of the\( A_\beta \) influx during the hemodialysis sessions.
Table 10.2  Average $\beta$ influx into the blood during the hemodialysis sessions

| Time point of HD session | $\beta_{1-40}$ | $\beta_{1-42}$ |
|-------------------------|----------------|----------------|
| 0 h                     | 750.7          | 63.3           |
| 1 h                     | 517.7          | 50.0           |
| 4 h                     | 361.8          | 41.5           |

| Removal Efficiency (%) of Pre/Post dialyzers | 67.3 | 51.3 |

| A$\beta$ removed by dialyzers (ng) | (0–1 h) | (1–4 h) | Total removed A$\beta$ (0–4 h) (a) | (0–1 h) | (1–4 h) | Total removed A$\beta$ (0–4 h) (a) |
|-----------------------------------|--------|---------|-----------------------------------|--------|---------|-----------------------------------|
|                                   | 3329   | 6925    | 10,254                            | 227    | 549     | 776                               |

| Change of A$\beta$s in the blood (ng) | 1952 | 941 | Decreased A$\beta$ (0–4 h) (b) | 165 | 108 | Decreased A$\beta$ (0–4 h) (b) |
|--------------------------------------|-----|----|--------------------------------|----|----|--------------------------------|
|                                      | 1011| 57 |                                |    |    |                                  |

| A$\beta$ influx into the blood during hemodialysis sessions (ng) (a–b) | 9243 | 719 |

Taken from Kitaguchi et al. (2015)

10.9  Effects of Hemodialysis, One of the Blood A$\beta$ Removal Methods, on Cognitive Function

Renal failure is well known to cause cognitive decline. In our cross-sectional study, cognitive function as measured by the MMSE was impaired in renal failure patients who did not receive hemodialysis compared to age-matched healthy controls. However, MMSE scores of hemodialysis patients were similar to those of controls (Fig. 10.6) (Kato et al. 2012).

Figure 10.7 shows the relationship between plasma A$\beta$ concentrations, cognitive function, renal function, and hemodialysis vintage (the duration of hemodialysis) before and after initiation of hemodialysis. Before initiation of hemodialysis, plasma concentrations of both A$\beta_{1-40}$ and A$\beta_{1-42}$ increased along with a concomitant decline in renal function. However, when patients were introduced to hemodialysis (after initiation of hemodialysis), an increase in plasma A$\beta$ concentrations was no longer apparent, but there was instead a slight tendency toward a decrease. Although the cognitive function declined along with the decline in renal function, this was maintained following initiation of hemodialysis (bottom of Fig. 10.7).

In the prospective study with 18 and 36 months follow-up, average MMSE scores did not significantly change, as shown in Fig. 10.8a, b. However, analysis of the change in individual subjects revealed that most hemodialysis patients maintained...
Fig. 10.5 Comparison of senile plaques in patients who had undergone hemodialysis (HD) with those who had not undergone HD (non-HD). (a) Stained with the anti-Aβ_{17-24} antibody 4G8; (b) stained with the anti-Aβ_{1-16} antibody DE2. The numbers of all types of Aβ deposition (diffuse, cored, and neuritic plaques) were significantly lower in HD patients. HD, n = 17; non-HD, n = 16. (Taken from Sakai et al. 2016 and modified)
or improved their cognitive function, with the exception of patients that showed white matter ischemia at baseline (Fig. 10.8c). This suggests that hemodialysis, with Aβ removal from the blood three times a week, may have a positive effect on cognitive function but has almost no influence on the cognitive effects of brain ischemia.

Furthermore, using a database of over 200,000 hemodialysis patients in Japan, the risk of dementia was revealed to be significantly lower in the patient subgroup with a longer duration of hemodialysis in subjects without diabetes (Nakai et al. 2018).

10.10 Effects of Smoking on Removal of Blood Aβ

We then investigated the effects of smoking on Aβ removal efficiencies in hemodialysis. Subjects were non-diabetic hemodialysis patients; n = 57, 29 male and 28 female; age, 69.4 ± 3.8 years old (59–76 years old); duration of hemodialysis, 13.9 ± 9.4 years (1–37 years); 28 smokers and 29 non-smokers, with “smoker” defined as a patient who had ever smoked (former smokers and current smokers). Information regarding the duration of smoking, the number of cigarettes per day, and the brands of cigarettes were obtained by interview with each patient. The product of the duration and the number of cigarettes per day was also used for analysis.

Interestingly, removal efficiencies for both Aβ<sub>1-40</sub> and Aβ<sub>1-42</sub> in smokers significantly decreased during the 4-h hemodialysis sessions (Table 10.3). The efficiencies
for non-smokers showed a tendency to increase, which was insignificant, rather than a decrease. The reason for this difference is unclear at present. One possibility is that Aβ species in the blood of smokers may have certain characteristics that cause saturation of Aβ adsorption or clogging of the inner surface of dialyzer membranes. A second possibility is that Aβ species flowing into the blood during hemodialysis may be more difficult to remove using a dialyzer in smokers than in non-smokers.

However, there is a limitation regarding this speculation on the effects of smoking. The ratio of male/female subjects was higher in smokers than in non-smokers. Therefore, the differences between smokers and non-smokers could be partially attributable to gender.

**Fig. 10.7** Summary of cross-sectional study of renal failure patients before/after initiation of hemodialysis (HD). The central box indicates initiation of hemodialysis. Left of the central box, data from renal failure patients without hemodialysis (non-HDRF) are shown. Right of the central box, data from hemodialysis patients (with-HDRF) are shown. Vertical axis: upper, plasma Aβ\(_{1-40}\) concentrations; middle, plasma Aβ\(_{1-42}\) concentrations; lower, the Mini-Mental State Examination (MMSE) score (30 indicates no mistakes). Plasma for measuring Aβ concentrations after the initiation of hemodialysis was sampled at the beginning of each hemodialysis session. Horizontal axis: before initiation of hemodialysis, plasma creatinine concentrations (CRN), which indicate decline of renal function; after initiation of hemodialysis, the vintage (duration) of hemodialysis. (Data from Kato et al. 2012)
Figure 10.9 indicates that there appears to be no clear difference between the smoker and non-smoker cognitive function, as measured by the MMSE, in our study with a small sample size. The MMSE scores of smokers were similar to those of non-smokers in all three groups; age-matched healthy controls (AMC, seven smokers, ten non-smokers), renal failure patients who did not need hemodialysis (non-HDRF, Figs. 10.8 and Table 10.3). Patients whose MMSE declined by −4 and −5 showed white matter ischemia at baseline. (Taken from Kitaguchi et al. 2015 and modified)

Table 10.3 Effects of smoking; comparison of Aβ removal efficiencies at pre-/post dialyzers in hemodialysis sessions

| Removal efficiencies % | 1 h                  | 4 h          |
|-------------------------|----------------------|--------------|
| Aβ1–40                  | Smoker               | 70.0 ± 9.6   | 60.0 ± 8.6 |
|                         | p=0.0016             |              |
|                         | Non-smoker           | 65.4 ± 9.9   | 70.4 ± 20.3 |
| Aβ1–42                  | Smoker               | 56.8 ± 9.1   | 53.7 ± 6.2 |
|                         | p=0.049              |              |
|                         | Non-smoker           | 50.2 ± 11.4  | 55.3 ± 8.5  |

10.11 Effects of Smoking on Cognitive Function and Brain Atrophy in Renal Failure Patients

Figure 10.9 indicates that there appears to be no clear difference between the smoker and non-smoker cognitive function, as measured by the MMSE, in our study with a small sample size. The MMSE scores of smokers were similar to those of non-smokers in all three groups; age-matched healthy controls (AMC, seven smokers, ten non-smokers), renal failure patients who did not need hemodialysis (non-HDRF,
seven smokers, seven non-smokers), and severe renal failure patients who received hemodialysis three times a week (HDRF, 28 smokers, 29 non-smokers).

However, brain CT scans revealed that there were differences in brain atrophy between smokers and non-smokers. Frontal/temporal and temporal/parietal atrophies were more severe in smokers than in non-smokers, as detected by brain CT scans (p = 0.0465 and p = 0.0062, respectively, by the \( \chi^2 \) test). This suggests that the effects of smoking on the brain may not be sufficiently serious to affect cognitive function in our study, or that hemodialysis including A\( \beta \) removal from the blood three times a week may maintain cognitive function despite the presence of more severe atrophies in smokers.

**Fig. 10.9** The cognitive function of smokers and non-smokers was similar in our study. The patients were the same as those represented in Fig. 10.6 except that smoking history was obtained from only 16 non-HDRF patients. AMC age-matched healthy controls (seven smokers, ten non-smokers), non-HDRF renal failure patients without hemodialysis (seven smokers, nine non-smokers), HDRF severe renal failure patients who received hemodialysis three times a week (28 smokers, 29 non-smokers). MMSE Mini-Mental State Examination

**Fig. 10.10** Brain atrophy in smokers and non-smokers. Frontal/temporal atrophy and temporal/parietal atrophy was more severe in smokers than in non-smokers, as detected by brain CT scans (p = 0.0465 and p = 0.0062, respectively, by the \( \chi^2 \) test). (Taken from Kitaguchi et al. 2015)
10.12 Closing

As described above, removal of blood Aβ may enhance Aβ influx into the blood from the brain, resulting in maintenance or improvement of cognitive function. We believe that the E-BARS could contribute as a therapy for Alzheimer’s disease. With respect to smoking, the patient’s history in this regard may have some effect on brain atrophy and on the forms of Aβs existing in the blood. Additional study will be necessary in the future to further clarify this.

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