Abstract: Diabetic kidney disease develops in approximately 40% of diabetic patients and is a major cause of chronic kidney diseases (CKD) and end stage kidney disease (ESKD) worldwide. Hydrogen sulfide (H$_2$S), the third gasotransmitter after nitric oxide (NO) and carbon monoxide (CO), is synthesized in nearly all organs, including the kidney. Though studies on H$_2$S regulation of renal physiology and pathophysiology are still in its infancy, emerging evidence shows that H$_2$S production by renal cells is reduced under disease states and H$_2$S donors ameliorate kidney injury. Specifically, aberrant H$_2$S level is implicated in various renal pathological conditions including diabetic nephropathy. This review presents the roles of H$_2$S in diabetic renal disease and the underlying mechanisms for the protective effects of H$_2$S against diabetic renal damage. H$_2$S may serve as fundamental strategies to treat diabetic kidney disease. These H$_2$S treatment modalities include precursors for H$_2$S synthesis, H$_2$S donors, and natural plant-derived compounds. Despite accumulating evidence from experimental studies suggests the potential role of the H$_2$S signaling pathway in the treatment of diabetic nephropathy, these results need further clinical translation. Expanding understanding of H$_2$S in the kidney may be vital to translate H$_2$S to be a novel therapy for diabetic renal disease.

Keywords: hydrogen sulfide; renal physiology; oxidative stress; renin-angiotensin system; diabetic nephropathy

1. Introduction

Diabetic kidney injury is a leading cause of end-stage renal failure, predominantly due to the increase of diabetes and obesity [1,2]. Diabetic kidney disease is manifested by glomerular hypertrophy, deposition of extracellular matrix (ECM) proteins, expansion of mesangial matrix and glomerular basement membrane, tubulointerstitial fibrosis, excessive urinary loss of proteins, and loss of waste clearance function over time [3]. In fact, most of cardiovascular disease mortality for diabetic patients is related to diabetic kidney disease [4].

Despite optimal management, diabetic kidney disease is still a major contributor to morbidity and mortality of diabetic patients worldwide [5]. Accumulating evidence indicates that hyperglycemia mediates renal damage in diabetes via multiple molecular mechanisms including oxidative stress, proinflammatory cytokines, induction of transforming growth factor beta-1 (TGF-β1) expression, fibroblast and renin angiotensin system (RAS) activation, as well as depletion of adenosine triphosphate [6]. Though the great efforts to further understand the pathological mechanisms...
of diabetic kidney disease are continuous, the current therapies only retard the disease progression but cannot cure it. Thereby, there is a pressing demand to identify novel therapies or targets for the treatment of diabetic kidney disease.

Hydrogen sulfide (H₂S) has been identified to exert a wide coverage of biological functions similar to other gasotransmitters nitric oxide (NO) and carbon monoxide (CO) [7]. Biochemical studies have demonstrated that kidneys are a rich source for H₂S formation [3]. It is currently believed that cystathionine γ-lyase (CSE), cystathionine β-synthase (CBS) and 3-mercaptopyruvate sulfurtransferase (3-MST) are the dominated enzymes for H₂S production in the kidney [8]. As reviewed recently, the potential roles of H₂S in the regulation of glomerular filtration rate (GFR), sodium absorption, renin release and oxygen sensing in kidney system have been described in details [8,9]. Recent investigations support that H₂S generation from the renal cells is reduced in acute or chronic kidney disorders including diabetic nephropathy [8,10–12]. In this review, we will discuss recent progression on H₂S in both kidney physiology and diabetic kidney disease. Besides, we will discuss the recent experimental findings on the molecular mechanisms underlying the therapeutic effects of H₂S against diabetic kidney disease and its possibilities, challenges for clinical application in the future.

2. Pathophysiology of Diabetic Kidney Disease

Diabetic kidney disease exhibits destructive structural changes such as glomerular basement membrane expansion, loss of podocytes, thickening of mesangial matrix and fusion of foot processes [13]. Conventionally, it is accepted that renal hemodynamics changes, oxidative stress, inflammatory response, hypoxia and renin-angiotensin-aldosterone system (RAAS) are majorly responsible for the pathogenesis of diabetic kidney disease [14] (Figure 1).

Glomerular hyperfiltration participates in the occurrence of diabetic kidney disease, and increased local angiotensin II (Ang II) induces the constriction of efferent arteriole, thus causing changes of autoregulation and glomerular hypertension [15]. At the same time, hyperglycemia and compensatory hyperinsulinemia promote vascular endothelial dysfunction through reactive oxygen species (ROS) production, activation of protein kinase C (PKC) and advanced glycation end-products (AGEs)-mediated proinflammatory response [16]. Activation of endothelin cells is necessary for podocyte injury and renal fibrosis in the diabetic kidneys [17].

Glomerular and renovascular lesions in diabetic kidney disease reduce the oxygen supply, thus eliciting renal medulla hypoxia and tubular dysfunction [18]. Without adequate oxygen supply, hyperglycemia may impair the stability of hypoxia-inducible factor (HIF) and facilitate renal tissue fibrosis [19]. Activation of RAAS is observed in renal cells induced by hyperglycemia and AGES, this may be mediated by ROS and G protein-coupled metabolic receptor 91 (GPR91) [20,21]. Studies in animal models and clinical trials demonstrate that RAAS inhibition effectively slows the progression of diabetic kidney disease [22]. As an important member of RAAS, Ang II is critically involved in the process of renal tissue fibrosis and tubule dysfunction [23]. Aldosterone is also a key player in the pathophysiology of diabetic kidney disease via boosting macrophage infiltration and renal fibrosis [24].

Cellular and molecular experiments have shown that disturbed mitochondria function and endoplasmic reticulum stress, as well as abnormal activation of intracellular signaling pathways including nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB), Janus kinase-signal transducer and activator of transcription (JAK-STAT) pathway, have found to be integrally associated with diabetic kidney disease [14,25,26]. With further research, the novel fields of pathophysiology of diabetic kidney disease are gradually identified, such as genetic and epigenetic regulation as well as podocyte autophagy [14]. The completion of the molecular mechanism studies in these directions would improve our understanding of diabetic kidney disease, and also help us better screen treatment strategies for preventing the renal destruction associated with diabetes.
**Figure 1.** Conventional pathophysiology of diabetes kidney disease. Diabetic kidney disease is closely associated with renal hemodynamic changes, ischemia and glucose metabolism abnormalities, oxidative stress, inflammatory response and over-activated RAAS, which contributes to glomerular hypertension and sclerosis, tubulointerstitial fibrosis, tubular atrophy and mesangial cell expansion. RAAS, renin-angiotensin-aldosterone system; IGF-1, insulin-like growth factor 1; TGF-β, transforming growth factor β1; VEGF, vascular endothelial growth factor; PG, prostaglandin; Ang II, angiotensin II; ET-1, endothelin-1; SGLT2, sodium glucose co-transporters 2; ROS, reactive oxygen species; AGEs, advanced glycation end products; TNF-α, tumor necrosis factor α; NF-κB, nuclear factor kappa-light-chain-enhancer of activated B cells; HIF, hypoxia-inducible factor; ECM, extracellular matrix; JAK/STAT, Janus kinase-signal transducer and activator transcription factor; MCP-1, monocyte chemotactic protein 1.

**3. Expression of H$_2$S in Diabetic Kidney Disease**

H$_2$S is a colorless gas with a rotten egg smell at room temperature and ambient pressure [27]. Despite its toxic effects as an environmental hazard, recent studies have highlighted its modulatory roles in fundamental cellular processes in several tissues. Distinct from the other gaseous transmitters like NO and CO, H$_2$S is able to dissolve in water because of its weak acid property [28]. Analysis of physical and chemical properties has discovered that H$_2$S can freely penetrate into cell membrane of all cell types due to it is highly lipophilic [29], thereby allowing H$_2$S to exhibit a wide spectrum of phenomena including development, angiogenesis and carcinogenesis [7].

In mammalian cells, H$_2$S is primarily generated by the enzymes including CBS, and CSE in the cytosol through the trans-sulfuration pathway [30–32]. Another enzyme, 3-MST gives rise to the endogenous H$_2$S production in the mitochondria using 3-mercapto pyruvate (3-MP) as a substrate [33,34]. Additionally, homocysteine is condensed with serine by CBS to generate cystathionine, which is converted into l-cysteine by CSE. The presence of l-cysteine is served as a substrate for both CBS and CSE to produce H$_2$S. CSE also produces H$_2$S by catalyzing homocysteine into homolanthionine [30]. As reviewed recently, two main pathways are proposed to be involved in H$_2$S generation in the kidney,
such as the trans-sulfuration pathway and the D-cysteine pathway [3,35]. In the kidney, D-cysteine may generate more H\textsubscript{2}S relative to the L-cysteine pathway [36]. The mechanisms underlying the H\textsubscript{2}S-generated pathways in the kidney are summarized in Figure 2.

**Figure 2.** Endogenous synthesis of H\textsubscript{2}S in renal system. (A) CSE reacts with L-homocysteine to induce H\textsubscript{2}S generation accompanied by formation of \(\alpha\)-ketobutyrate and L-homolanthionine. CBS catalyzes L-homocysteine that leads to the production of L-cystathionine, which is converted into L-cysteine by CSE. The presence of L-cysteine serves as a substrate for generation of H\textsubscript{2}S by CBS and CSE. (B) L-Cysteine translocates to mitochondria, followed by conversion to 3-MP by CAT. 3-MST produces H\textsubscript{2}S generation from 3-MP. (C) Peroxisome-mediated generation of 3-MP from D-cysteine with the aid of DAO. 3-MP is then imported into the mitochondria and becomes a substrate for 3-MST to generate H\textsubscript{2}S.

CBS, cystathionine \(\beta\)-synthase; CSE, cystathionine g-lyase; 3-MST, 3-mercaptopyruvatesulfurtransferase; CAT, cysteine aminotransferase; DAO, D-amino acidoxidase; 3-mercapto pyruvate, 3-MP.

Similar to liver and lung, CSE, CBS and 3-MST are highly expressed in the kidney [37]. CBS is predominantly located in renal proximal tubules, while CSE is mainly expressed in renal glomeruli, proximal tubules, interstitium, and interlobular arteries [38,39]. Specifically, immunohistochemistry results have shown that CSE is present in glomerular endothelial and mesangial cells, podocytes, proximal and distal tubules as well as the peritubular capillaries and blood vessels [40]. Besides, 3-MST can be detected in proximal tubular epithelium in the kidney [37]. CBS and CSE are abundantly expressed in renal tissues that synergistically produce H\textsubscript{2}S in the kidney [41]. Under normal conditions, the expression of CSE protein in the kidney is 20-fold higher than CBS, thus, CSE may be the main H\textsubscript{2}S-forming enzyme in the kidneys [42,43].

Decrease in endogenous H\textsubscript{2}S is reported to participate in the pathogenesis of diabetic nephropathy [41,44]. Treatment with sodium hydrosulfide (NaHS, a H\textsubscript{2}S donor) ameliorates kidney lesions in type 2 diabetes via increasing glucose uptake in both myotubes and adipocytes [44].
Suppressed CSE-catalyzed endogenous H$_2$S production by hyperglycemia may play an important role in the pathogenesis of diabetic nephropathy [45]. In type 2 patients, the plasma H$_2$S level is lower than that in normoglycemic humans [46], and it is negatively correlated with the markers of adiposity [47]. Moreover, the diabetic patients with dialysis exhibit reduced level of plasma H$_2$S, and lower H$_2$S level may contribute to uremic atherosclerosis in chronic hemodialysis patients with diabetic nephropathy [48]. Recent documents have demonstrated that renal H$_2$S-producing enzymes CBS and CSE are downregulated in animals with type 1 or type 2 diabetes [45,49,50]. The above evidence indicates that H$_2$S deficiency contributes to the development of diabetic kidney injury.

4. H$_2$S Regulation of Renal Function

4.1. H$_2$S and Renal Excretory Function

It has been reviewed that H$_2$S participates in the regulation of renal functions [41]. In anesthetized Sprague-Dawley rats, intrarenal arterial infusion of H$_2$S donor NaHS results in increased renal blood flow, GFR, urinary sodium (U(Na) × V), and potassium (U(K) × V) excretion, this effect is mimicked by renal artery infusion of L-cysteine, a H$_2$S generating substrate [41], suggesting that H$_2$S plays a tonic role in the regulation of renal function under physiological condition. In that study, the authors propose that H$_2$S augments urinary excretion of sodium and potassium by the inhibition of Na$^+$/K$^+$-2Cl$^-$ cotransporter and Na$^+$/K$^+$-ATPase [41]. Mechanistic studies have shown that H$_2$S directly induces endocytosis and inhibition of Na$^+$/K$^+$-ATPase via phosphatidyl inositol 3 kinase (PI3K)/protein kinase B (Akt) pathway in renal tubular epithelial cells [51]. In vitro studies have identified that H$_2$S inhibits Na$^+$/H$^+$ exchanger-3 activity in tubular epithelial cells [51]. It may be important to examine whether H$_2$S directly affects the expressions of these transporters.

Patch clamp studies indicate that hydrogen peroxide augments PI3K activity to activate the epithelial sodium channel (ENaC), this is corrected by H$_2$S [52]. It is also demonstrated that H$_2$S prevents β-adrenergic agonist-induced ENaC-mediated Na transport in human lung epithelial cells [53]. The Cl$^-$/HCO$_3^-$ exchanger activity in aortic tissues and vascular smooth muscle cells is dampened by H$_2$S [54,55]. Unfortunately, the effect of H$_2$S on ENaC and Cl$^-$/HCO$_3^-$ exchanger activities in renal system is still undefined. Considering the indispensable role of ENaC and Cl$^-$/HCO$_3^-$ exchanger in the control of sodium reabsorption and homeostatic maintenance of physiological pH, it will be interesting to understand the action of H$_2$S on ENaC and Cl$^-$/HCO$_3^-$ exchanger activities as well as subsequent renal function.

4.2. H$_2$S and Oxygen Sensing

Despite the production of H$_2$S is independent of oxygen, the metabolism of H$_2$S is largely linked with the presence of oxygen [56]. The role of H$_2$S in oxygen sensing has been tested in the carotid body, which participates in ventilation, heart rate and blood pressure [57]. In addition, emerging physiological evidence for H$_2$S-mediated oxygen sensing has been clarified in cardiovascular system [58,59], respiratory system [60] and gastrointestinal tract [61]. In the kidney, partial pressure of oxygen declines from cortex to medulla in normal rats [62]. The recognition of H$_2$S as an oxygen sensor in the kidney, especially in medulla, has been identified [11]. Compared with the renal cortex, the oxygen content is lower in medullar, resulting in a higher level of H$_2$S in this area [11]. In the mitochondria, H$_2$S is regarded as an electron donor for adenosine triphosphate release [63,64]. It is still unclear whether H$_2$S can be a possible source of energy in renal medulla. At least, reduction of oxygen leads to H$_2$S generation, which is helpful to recovery oxygen supply through raising medullary blood flow and suppressing tubular transport [65]. However, the downstream effectors of H$_2$S-mediated oxygen sensor in renal system remain to be further elucidated. Notably, the excessive erythropoietin levels are synthesized in anemia and hypoxia kidney [66,67]. Given H$_2$S participates in oxygen sensing, whether H$_2$S exerts a regulatory role in erythropoietin synthesis in the kidney, needs in depth exploration.
5. Role of H$_2$S in Diabetic Kidney Disease

5.1. Renin–Angiotensin System (RAS) and H$_2$S in Diabetic Nephropathy

The RAS is a multistep enzymatic cascade that plays a role in the control of blood pressure and fluid homeostasis [68]. Angiotensinogen is a major substrate of this cascade, and the 10 amino acids from the N-terminus of angiotensinogen are specifically cleaved to form angiotensin I. The conversion of angiotensin I to Ang II by angiotensin-converting enzyme (ACE) is a central step in the classical RAS pathway [22]. The initial evidence for a role of the RAS in diabetic nephropathy is established in a rat model of diabetes induced by streptozotocin (STZ) [69]. It is suggested that the dysregulated glomerular hemodynamic profile in diabetic rats might be associated with activation of the RAS [70,71]. Further studies unveil that hyperglycemia stimulates the renal RAS, and renders an increase in glomerular hydrostatic pressure, proteinuria, and structural injury with sclerosis and fibrosis [22]. Blockade of the RAS has been shown to delay the progression of diabetic kidney disease [72]. Over-activation of intrarenal RAS aggravates diabetic nephropathy due to a fact that inhibition of RAS retards the progression of diabetic nephropathy in animal models and clinical trials [22,72]. It is likely that intrarenal RAS is activated in patients suffering from diabetes, thereby RAS blockers have the benefits for abnormal glomerular structures in patients with diabetic kidney disease [73–75]. Present researches recommend that RAS blockers can relieve the nephropathy for albuminuric patients with diabetes [14]. These above studies would offer great promise for RAS-inhibition-based therapies in diabetic nephropathy.

Renin is mainly produced from the so-called juxtaglomerular epithelioid cells, and this process is regulated by intracellular cyclic adenosine monophosphate (cAMP) level [76]. Actually, H$_2$S is capable of downregulating the intracellular cAMP level in various cell types [77,78], implying that H$_2$S may regulate renin release. It has been reported that H$_2$S treatment inhibited the upregulation of renin level in renovascular hypertensive rats accompanying with a reduction of intracellular cAMP level in primary cultures of renin-rich kidney cells [78,79]. We also find that H$_2$S inhibits forskolin-induced renin degranulation in mast cells by lowering intracellular cAMP level, thus protecting against isoproterenol (ISO)-induced heart failure [80]. Likewise, H$_2$S therapy protects multiple organs including the heart, kidney, and blood-vessels and improves exercise capacity, coupled with inactivation of RAS in a murine model of transverse aortic constriction-induced heart failure [81]. These existing results show that H$_2$S may modulate renal activity and release, which may be beneficial for the prevention of diabetic nephropathy. However, the researches regarding the direct effect of H$_2$S on renin activity and release in diabetic kidney disease are still rare.

Mounting evidence suggests that hyperglycemia acts synergistically with Ang II to promote renal cellular injury in diabetes [82]. In proximal renal tubules, increased Ang II synthesis induces interstitial fibrosis [83]. The two main Ang II receptors (AT1 and AT2) have been cloned and well characterized [83]. Most biological effects of Ang II are mediated by the AT1 receptors [84]. Accordingly, it has been demonstrated that Ang II receptor blockers have a slowing effect on the progression of diabetic nephropathy [85]. Within RAS, high glucose stimulation upregulates mRNA levels of angiotensinogen (AGT), angiotensin converting enzyme (ACE), Ang II type I receptor (AT1) mRNA levels when compared with that of normal glucose-treated cells, which are reversed by supplementation of H$_2$S [86]. The protein expressions of ACE and AT1 receptors as well as Ang II are significantly upregulated in the diabetic kidneys and downregulated after treatment with H$_2$S donor, NaHS [50]. These data suggested H$_2$S alleviated the development of diabetic nephropathy by partially attenuating RAS activity (Figure 3).
Figure 3. A proposed sketch showing the effect of H$_2$S on the renin-angiotensin-system (RAS). Angiotensinogen is cleaved by renin to produce angiotensin I, then angiotensin I is further converted to Ang II by ACE. The deleterious effects of Ang II are mediated by AT1 receptors, whereas Ang II acts on AT2 receptors to function as a negative modulator of AT1 receptor actions. The activated RAS in diabetic kidney disease was ameliorated by H$_2$S treatment via inhibition of angiotensinogen, ACE, Ang II and AT1 receptors. RAS, renin-angiotensin-system; ACE, angiotensin converting enzyme; Ang II, angiotensin II.

5.2. Oxidative Stress and H$_2$S in Diabetic Nephropathy

A plethora of molecules are implicated for ROS generation, such as, reduced nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, uncoupled nitric oxide synthase (NOS) and mitochondrial respiratory chain via oxidative phosphorylation [87,88]. Excessive accumulation of ROS activates PKC, MAPK, and numerous cytokines and transcription factors, which eventually lead to increased ECM production and progressive fibrosis in diabetic kidney disease [89]. High glucose induces substantial ROS generation in glomerular mesangial cells [90].

Antioxidants inhibit high glucose- and hydrogen peroxide-induced TGF-β1 and fibronectin upregulations in mesangial cells and tubular epithelial cells, thus providing evidence that ROS are necessary for high glucose-induced renal injury [89,91–95]. Redundant ROS trigger signal transduction cascade and transcription factors as well as their downstream molecules, which are implicated in glomerular mesangial expansion and tubulointerstitial fibrosis [89]. In the kidney, the excessive oxidative stress and its associated inflammation have been found to aggravate diabetic nephropathy in recent years [94]. These observations suggest that ROS may act as intracellular messengers in diabetic kidney disease, thus making it an attractive target for developing novel therapeutic strategies to ameliorate diabetic nephropathy.
Supplementation with H$_2$S attenuates high glucose-induced elevation in ROS production in renal mesangial cells and diabetic rat kidneys [86]. H$_2$S treatment protects the kidneys of type 1 diabetic rats, which may be related to the suppression of oxidative stress via incrementing the activities of superoxide dismutase (SOD) [95]. Exposure to high glucose promotes ROS generation in mesangial cells, and the effect is counteracted by NaHS [45]. High glucose increases NADPH oxidase 4 (NOX4) expression in renal proximal tubular epithelial cells, this change is inhibited by NaHS [49]. These above studies provide the direct evidence that H$_2$S plays an important role in the renal cellular response to high glucose-induced oxidative stress (Figure 4). Thus, application of H$_2$S donors seems to be a choice for the treatment of diabetic nephropathy.

Figure 4. Effects of H$_2$S on oxidative stress in diabetes kidney disease. H$_2$S is found to reduce high glucose-induced oxidative stress by activating the Nrf2 antioxidant pathway and two downstream targets of Nrf2, HO-1 and NQO1, as well as enhancing the SOD and glutathione peroxidase activities in diabetes kidney disease. Nrf2, nuclear factor erythroid-2 related factor 2; HO-1, heme oxygenase-1; NQO1, NADPH: quinone oxidoreductase-1; SOD, superoxide dismutase.

Nuclear factor erythroid-2 related factor 2 (Nrf2) is a transcription factor with a basic leucine zipper motif [96]. This transcription factor is highly conserved with Nrf2-ECH homology domains, Neh1 to Neh7 [97,98]. Among which, Neh1 facilitates the nuclear translocation of Nrf2 [99], while Neh2 enables the coupling of Nrf2 with Kelch-like ECH-associated protein 1 (Keap1) [100,101]. In addition, the lysine residues and a serine residue within Neh2 domain are inhibitory for proteasome-mediated Nrf2 degradation [102]. The Neh3–Neh5 domains are necessary for Nrf2 transcriptional activity [103], and Neh6 domain is responsible for Nrf2 degradation [104]. With respect to Neh7 domain, it is identified to suppress Nrf2 downstream gene expressions via binding to the retinoic acid receptor $\alpha$ [105]. Under normal status, Keap1 functions as an endogenous inhibitor of Nrf2 by interacting with Nrf2 to form a complex [106,107]. During oxidative stress, the modification of three important cysteine residues of Keap1 leads to Nrf2 liberalization, followed by Nrf2 nuclear translocation [108,109]. After that, in the nucleus, Nrf2 promotes the induction of phase II detoxification enzymes and antioxidants [110]. The activity of Nrf2 plays a central role in cellular resistance to oxidative stress [111]. In fact, Nrf2-related signaling pathway is demonstrated to be crucial in maintaining the balance between oxidation and reduction in the kidney, thus activation of Nrf2 pathway appears to be an effective method for the treatment of diabetic kidney disease [112–114]. In regard this, it may be interesting to know whether H$_2$S activates Nrf2 pathway to prevent oxidative stress in disease. As expected, H$_2$S could activate the Nrf2 signaling pathway together with the upregulations of antioxidant proteins haem oxygenase-1 (HO-1) and NAD(P)H: quinone oxidoreductase 1 (NQO1), thus alleviating the development of diabetic
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5.2. Cardiomyopathy and H_{2}S in Diabetic Kidney Disease

Cardiomyopathy via attenuation of oxidative stress in the heart [115]. Upon exposure to high glucose plus oxidized low density lipoprotein (ox-LDL), the superoxide anions and adhesion molecule levels are elevated in endothelial cells [116]. However, administration of H_{2}S rectifies these dysregulated changes via increasing S-sulfhydration of Keap1, interfering with the interaction between Keap1 and Nrf2, as well as stimulating Nrf2 nuclear translocation [116]. S-propargyl-cysteine, a novel molecule that upregulates endogenous H_{2}S production, is reported to attenuate the generation of ROS and apoptosis in diabetic cardiomyocytes, this effect may be dependent on increased stability and nuclear translocation of Nrf2 and the dissociation of Nrf2 from Keap1 [117]. In cisplatin-induced nephrotoxicity, we demonstrate that hydrogen polysulfide, a novel H_{2}S-derived signaling molecule, leads to the nucleus translocation of Nrf2 in renal proximal tubular cells [118]. As a result, hydrogen polysulfide mitigates cisplatin-induced renal cell oxidative stress and apoptosis [118]. Similar to these observations, H_{2}S is found to activate the Nrf2 antioxidant pathways, thereby decreasing malondialdehyde (MDA) levels and restoring SOD and glutathione peroxidase activities in diabetic kidney [50]. To be noticed, further studies are required to elucidate the precise behavior of H_{2}S-mediated suppression of oxidative stress via Nrf2 pathway in diabetic kidney disease. Anyway, activation of Nrf2 pathway by H_{2}S might serve as a promising strategy for the treatment of oxidative stress-associated kidney damage under diabetic condition.

5.3. Inflammation and H_{2}S in Diabetic Kidney Disease

The intimate mechanisms contributing to the progression of diabetic renal injury are not well elucidated, but current knowledge indicates that immune and inflammation responses appear to be relevant factors [119]. Growing evidence suggests that pathogenesis of diabetes mellitus is widely linked with the innate immune system activation and a chronic low-grade inflammatory state [120,121]. It has been accepted that pro-inflammatory signaling pathways and their downstream effectors are emerging as promising therapeutic targets for diabetic nephropathy [122]. In the kidney, many intrinsic renal cells including glomerular, endothelial, tubular, and mesangial cells are capable of synthesizing inflammatory cytokines, and these substances are gradually upregulated as diabetic nephropathy progresses, suggesting a role of inflammation in the pathogenesis of diabetic kidney disease [123].

In the present time, inflammatory cytokines activate diverse transduction pathways in the pathogenesis of diabetic kidney, such as oxidative stress and transcription factors including NF-κB and JAK/STAT pathways [124,125]. In addition, macrophages and T lymphocytes within kidneys also play determinant roles in diabetic renal damage, and their accumulation is positively related with the severity of diabetic nephropathy experimental models [1]. Certainly, inflammation response is an important contributor to the pathogenesis of diabetic nephropathy, this participation involves increased chemokine and pro-inflammatory cytokine production, infiltration of inflammatory cells to the kidney, and subsequent renal damage. Such in-depth exploration of inflammatory response may identify novel anti-inflammatory approaches for the treatment of diabetic nephropathy.

Recently, it is widely accepted that H_{2}S inhibits inflammatory cytokine production, and suppresses activation of key transcriptional factors [126]. In renal system, administration of H_{2}S donors ameliorates renal ischemia/reperfusion injury accompanied by reductions in oxidative stress and inflammatory response [3]. In unilateral obstruction kidney injury, the renal H_{2}S generation is declined, and application of NaHS obviously retards the kidney injury, including inflammatory response [38]. It can be concluded that H_{2}S metabolism is dysregulated in inflammatory diseases including diabetic nephropathy.

In STZ-induced diabetic rats, H_{2}S donor NaHS exerts an-inflammatory actions through inhibiting NF-κB signaling in rat glomerular mesangial cells [50], thus alleviating the development of diabetic nephropathy. MMP-9 is a zinc-dependent endopeptidase, which can be activated by ROS [127]. As an inflammatory cytokine, inflammatory cell-derived MMP-9 leads to ECM degradation and renal vascular remodeling [128,129]. Hyperglycemic Akita mice exhibit higher level of MMP-9 and lower production of H_{2}S, and H_{2}S treatment reverses the altered diabetic renal remodeling induced by...
MMP-9 [130]. Therefore, H$_2$S has therapeutic potential to ameliorate diabetic renal remodeling in association with suppression of inflammation response (Figure 5). However, the regulation of H$_2$S in the diabetic kidney inflammation still warrants more investigation.

Figure 5. A proposed model of diabetic kidney inflammation mediated by H$_2$S. The anti-inflammation mechanisms of H$_2$S may involve its inhibition of macrophages infiltration, as well as its blockade of NF-$\kappa$B and MAPK signaling in renal system. NF-$\kappa$B, nuclear factor kappa-light-chain-enhancer of activated B cells; MAPK, mitogen-activated protein kinase, TNF-$\alpha$, tumor necrosis factor $\alpha$; IL-1$\beta$, interleukin-1$\beta$; VCAM-1, vascular cell adhesion molecule-1; ICAM-1, intercellular adhesion molecule-1; MCP-1, monocyte chemotactic protein-1; MMP-9, matrix metalloproteinase-9.

6. Renal Fibrosis and H$_2$S in Diabetic Kidney Disease

Renal fibrogenesis is a complex process involving a host of pathological scarring processes such as dysregulated ECM assembly, anchoring or degradation in glomerular basement membranes and the tubulointerstitium, as well as activated fibroblasts [131–133]. Moreover, epithelial mesenchymal transition (EMT) and endothelial mesenchymal transition (EndMT) programs are also considered as alternative mechanisms for renal fibrosis [134]. EndMT is responsible for the deposition of activated fibroblasts and myofibroblasts in diabetic kidney fibrosis [135]. It is well known that even after control of blood glucose level, diabetic patients may continue to develop to glomerular and tubulointerstitial fibrosis, eventually resulting in renal failure [136]. These observations suggest that renal fibrosis may play a significant role in the pathobiology of diabetic kidney disease.

6.1. EndMT and H$_2$S in Diabetic Renal Fibrosis

In such fibrotic process, renal fibroblasts play prominent roles in the development of diabetic renal fibrosis [137]. Activated fibroblasts may be from resident quiescent fibroblasts via the process of EndMT [135]. It has been shown that activated fibroblasts are correlated with the excess deposition of interstitial ECM in diabetic kidney disease [138]. The emergence of EndMT is responsible for the accumulation of activated fibroblasts in renal fibrosis, suggesting that targeting EndMT might have therapeutic potential for diabetic renal fibrosis.
Vascular endothelial cells may be transformed to fibroblasts by undergoing a phenotypic transition similar to EMT, referred to as EndMT [139]. The process of EndMT involves the upregulated expressions of mesenchymal proteins including α-smooth muscle actin (α-SMA) and the loss of endothelial markers including endothelial cadherin (E-cadherin) and CD31 [140]. The vital role of EndMT in renal fibrosis is recently discussed and reviewed [23,141]. The first evidence for the EndMT in renal fibrosis is established by Zeisberg and colleagues, their results demonstrate that a large proportion of myofibroblasts coexpress the endothelium marker CD31 in three mouse models, unilateral ureteral obstruction (UUO), genetic modification, STZ-induced diabetic nephropathy [135], suggesting that these fibroblasts are likely derived from endothelial cells and EndMT may substantially contribute to the development and progression of renal fibrosis. Another group also confirms that EndMT occurs and leads to the formation of myofibroblasts in early diabetic renal fibrosis [142]. Interestingly, under endoplasmic reticulum stress, H\textsubscript{2}S blocks the EndMT process in human umbilical vein endothelial cells (HUVECs) via downregulating the mesenchymal marker expressions, and upregulating the endothelial marker expressions [143]. In cultured kidney fibroblasts, H\textsubscript{2}S inhibited the excessive cell proliferation. Furthermore, the differentiation of renal fibroblasts to myofibroblasts is antagonized by H\textsubscript{2}S donor, which is associated with inhibition of TGF-β1-Smad and MAPK signaling pathways [38]. However, no studies are conducted to determine whether H\textsubscript{2}S directly modulates the EndMT process in diabetic renal fibrosis.

6.2. EMT and H\textsubscript{2}S in Diabetic Renal Fibrosis

EMT is a pathological process in which epithelial cells lose epithelial characteristics and acquire properties of mesenchymal cells [131]. EMT has been divided into three subtypes on the basis of their functional consequences and biomarker context [144]. Type 1 EMT is a key modulator in the formation of diverse cell types without organ fibrosis, whereas type 2 EMT is related with the transition of epithelial cells to tissue fibroblasts in the process of organ fibrosis [144]. Meanwhile, type 2 EMT would continue in response to inflammation, resulting in persistent organ fibrosis and tissue destruction [145]. Type 3 EMT is detected in carcinoma cells, this process is a prominent contributor to tumor invasion, migration, and metastatic outgrowth [146]. It is highlighted that type 2 EMT is a direct contributor to the generation of myofibroblast population in the kidney, thus inducing the development of diabetic renal fibrosis [147]. In general, it is recognized that the transformation of tubular epithelial cells into mesenchymal cells is the most possible mechanism that underlies diabetic kidney fibrosis [148]. The existence of EMT could be initiated by a myriad of molecules, in which TGF-β1 seems to be a primary player [149–151].

Following UUO male Lewis rats, the uncontrollable expressions of EMT markers are mitigated upon H\textsubscript{2}S treatment [152]. Daily treatment with the slow-releasing H\textsubscript{2}S donor GYY4137 attenuates renal injury by regulating the TGF-β1-mediated EMT pathway in UUO model [153,154]. Consistently, H\textsubscript{2}S counteracts Ang II- and TGF-β1-induced EMT via inactive TGF-β1 monomer formation in renal tubular epithelial cells [155]. Under diabetic condition, H\textsubscript{2}S levels in the plasma and renal cortex are evidently reduced, while the levels of TGF-β1 and collagen IV are enhanced in STZ-induced diabetic rats, which are reversed by administration of NaHS [45]. H\textsubscript{2}S inhibits TGF-β1-induced EMT process in renal tubular epithelial cells, as evidenced by downregulated expressions of α-SMA and fibronectin, and upregulated expression of E-cadherin, and blockade of ERK- and β-catenin-dependent pathways may account for the protective effect of H\textsubscript{2}S [156]. In the process of EMT, future studies elucidating other targeted molecules induced by H\textsubscript{2}S in diabetic renal fibrosis will allow a better understanding of comprehensive cellular responses to H\textsubscript{2}S.

Upon high glucose challenge, the mTORC1 signaling is activated due to the inhibition of AMPK activity in renal epithelial cells, thus stimulating matrix protein synthesis and renal hypertrophy [157]. NaHS dose-dependently stimulates AMPK phosphorylation, inhibition of AMPK with Compound C abolishes the negative effect of NaHS on global matrix protein synthesis induced by high glucose [157], implying the necessary role of AMPK in H\textsubscript{2}S-mediated effects on diabetic renal disease. Intriguingly,
induction of inducible nitric oxide synthase (iNOS), not endothelial nitric oxide synthase (eNOS), is required for H₂S to block high glucose-induced oxidative stress and matrix protein overproduction in renal proximal tubular epithelial cells, suggesting a role of NO in H₂S-mediated beneficial effects against diabetic kidney disease [49]. These two gasotransmitters H₂S and NO may be considered as therapeutic targets in diabetic nephropathy (Figure 6).

**Figure 6.** Effect of H₂S on renal fibrosis in diabetic kidneys. Activated fibroblasts may be from resident quiescent fibroblasts via the process of EndMT. H₂S treatment prevents the differentiation of quiescent renal fibroblasts to myofibroblasts and myofibroblasts proliferation via inhibition of the TGF-β1/Smad and MAPK signaling pathways. Blockade of ERK- and β-catenin-dependent pathways may be involved in the protective effect of H₂S on the formation of EMT in renal tubular epithelial cells. In addition, H₂S dose-dependently stimulates AMPK phosphorylation and induces its subsequent inhibition of mTORC1 activity. Induction of iNOS, is required for H₂S to inhibit high glucose-induced oxidative stress and matrix protein generation in renal proximal tubular epithelial cells. EndMT, endothelial-mesenchymal transition; EMT, epithelial mesenchymal transition; TGF-β1, transforming growth factor-β1; MAPK, mitogen-activated protein kinase; mTORC1, mammalian target of rapamycin complex 1; ERK, extracellular regulated protein kinases; AMPK, adenosine 5'-monophosphate (AMP)-activated protein kinase; iNOS, inducible nitric oxide synthase.

### 7. Glomerular Expansion and H₂S in Diabetic Kidney Disease

Glomerular mesangial cells are vital components that maintain the normal morphology and functions of glomeruli [158]. Under diabetic conditions, both cell proliferation and ECM production of glomerular mesangial cells are increased, and those aberrant changes will ultimately lead to the thickening of glomerular basal membrane, and glomerular expansion [5,159,160]. Diabetic nephropathy is reflected by the abnormal ECM production and the phenotypic change of glomerular mesangial cells [161]. Data suggest that numerous glomerular diseases are associated with glomerular expansion, which may eventually produce glomerular scarring, especially in diabetes [162].

Over-activation of intrarenal RAS is known as a crucial step in the pathogenesis of diabetic nephropathy [163]. Within the RAS, Ang II is one of the biologically active peptides of RAAS, and it is also regarded as an essential mediator for diabetic nephropathy [164,165]. Activation of intrarenal RAS has been implicated in glomerular enlargement and secondary glomerulosclerosis,
interstitial fibroblast proliferation and ECM deposition [163,166]. In glomerulus, activated RAS may result in the proliferation of glomerular mesangial cells [167]. It has been revealed that high glucose stimulates ROS production and promotes fibronectin and collagen IV synthesis in cultured renal mesangial cells [168]. Exposure to high glucose raises the proliferation of rat renal glomerular mesangial cells, along with decreased the CSE expression and H$_2$S level, whereas supplement of H$_2$S prevents high glucose-triggered proliferation and ECM secretion in rat mesangial cells [45]. Mechanistically, inactivation of intrarenal RAS by H$_2$S is requisite for inhibition of cell proliferation rate and TGF-β1 and of collagen IV productions in renal mesangial cells [86]. This effect may be dependent on attenuation of ROS generation because administration of NADPH oxidase inhibitor is capable of reversing the hyperglycemia-induced RAS activation and the following cell proliferation as well as collagen synthesis of renal mesangial cells [86]. After treatment with the H$_2$S donors in human mesangial cells, the induction of antioxidant enzyme HO-1 may be a potential mechanism whereby H$_2$S exerts its protective effects in the context of high glucose [169]. Additionally, stimulation of high glucose facilitates the proliferation of mesangial cells, which is coupled with reduced endogenous H$_2$S synthesis. Exogenous H$_2$S markedly mitigates high glucose-induced over-proliferation of mouse mesangial cells via inhibition of toll-like receptor 4 (TLR4) and PI3K/Akt pathway [170]. Collectively, the decreased H$_2$S level responses to high glucose results in mesangial cell proliferation, which, in turn, contributes to the pathogenesis of diabetic glomerular hypertrophy and renal dysfunction (Figure 7).

Figure 7. Effect of H$_2$S on glomerular hypertrophy and podocyte injury in diabetic kidneys. (A) Activation of intrarenal renin-angiotensin system and NADPH-derived ROS contribute to the proliferation and ECM secretion in high glucose-incubated renal mesangial cells, this phenomenon is attenuated by H$_2$S, dependent on HO-1 induction and inhibition of TLR4 and PI3K/Akt pathway. (B) Further studies reveal that endoplasmic reticulum stress, dysregulation of autophagy and mTORC1 activation in podocyte promote the development of diabetic nephropathy. H$_2$S may induce AMPK phosphorylation and HO-1, and suppress the Wnt/β-catenin pathway to mitigate podocyte injury induced by hyperglycemia. NADPH, reduced nicotinamide adenine dinucleotide phosphate; ROS, reactive oxygen species; ECM, extracellular matrix; HO-1, heme oxygenase-1; TLR4, toll-like receptor 4; mTORC1, mammalian target of rapamycin (mTOR) complex 1; AMPK, adenosine 5’-monophosphate (AMP)-activated protein kinase.

8. Podocyte Injury and H$_2$S in Diabetic Kidney Disease

Over the past decade, overwhelming evidence has pointed to the podocyte as a key target of renal injury during diabetic kidney disease progression [171–174]. Once the RAS is activated in diabetic kidneys, it may induce the generation of growth factors or cytokines such as TGF-β, vascular endothelial growth factor (VEGF), and monocyte chemotactic protein 1 (MCP-1), which may directly or indirectly lead to renal fibrosis, oxidative stress and podocyte apoptosis [175]. Studies from diabetic patients and
animal models have revealed that the onset of albuminuria is closely related with podocyte injury, such as podocyte hypertrophy, detachment, apoptosis and foot process effacement [173]. Experimental data support that podocytes become nephrin-absent, effaced, and apoptotic, these events are correlated with the emergence of albuminuria [176]. Generally, the effacement and loss of podocytes are key events in the early progression of diabetic kidney disease [177]. AGEs dose- and time-dependently induce the apoptosis in murine podocytes via endoplasmic reticulum stress-mediated apoptotic pathway [178]. Further studies establish that dysregulation of autophagy or mTORC1 activation in podocytes participates in the development of diabetic nephropathy [179–181]. According to previous studies, the contribution of podocyte injury to diabetic nephropathy is a subject of great interest.

High glucose significantly reduced CSE expression in cultured mouse podocytes, and this podocyte injury responses to high glucose is alleviated by exogenous H$_2$S possibly through Zona occludens-2 (ZO-2) upregulation and the subsequent suppression of Wnt/β-catenin pathway [182]. Moreover, it is demonstrated a significant increase in HO-1 expression after incubation with the H$_2$S donors in both mesangial and podocyte cells [169], hinting that the ability to promote antioxidant enzyme HO-1 expression might be a potential mechanism by which H$_2$S exerts its protective effects (Figure 7). Additional studies are needed to better recognize the molecular mechanisms of H$_2$S in diabetic podocyte injury, and these studies could lead to novel therapeutic strategies for diabetic nephropathy.

9. Phytopharmaceuticals/Agents-Mediated H$_2$S Induction in Diabetic Kidney Disease

As discussed above, H$_2$S donors such as NaHS or GYY4137 might be a good therapeutic strategy for the treatment of diabetic kidney disease (Tables 1 and 2). However, recent findings have disclosed that some phytopharmaceuticals or agents that participate with H$_2$S may exhibit renal protective effects in diabetes (Table 3). A novel H$_2$S-releasing compound, S-propargyl-cysteine, attenuates inflammation in diabetic kidneys via suppressing the phosphorylation of extracellular regulated protein kinases (ERK), p38 protein [183]. Tadalafil abrogates the global protein synthesis and matrix protein laminin γ1 in kidney podocytes induced by high glucose, this may be relied on induction of H$_2$S [184]. Induction of CSE-derived H$_2$S by tadalafil accelerates AMPK phosphorylation by stimulating calcium-calmodulin kinase kinase β, followed by inhibition of mTORC1 activity and mRNA translation in high glucose-treated kidney podocytes [184]. Metformin, an anti-diabetic agent, is widely prescribed to treat diabetic kidney disorder through saving the podocytes [185], this effect may be related with progressively increased H$_2$S concentration in the kidney [186]. Together with these results, it is reasonable to recommend that the induction of H$_2$S by natural plant-derived compounds might be potential therapeutic candidates to ameliorate diabetic nephropathy. However, more investigation on this subject is necessary.
Table 1. Beneficial effect of H$_2$S on experimental diabetic nephropathy in vitro.

| H$_2$S Donors | Cell Type | Main Findings | Ref. |
|---------------|-----------|---------------|------|
| NaHS          | Renal fibroblasts | H$_2$S inhibits the proliferation of renal fibroblasts. Furthermore, the differentiation of quiescent renal fibroblasts to myofibroblasts is prevented by H$_2$S, which involves the inhibition of TGF-β1-Smad and MAPK signaling pathways. | [38] |
| NaHS          | Glomerular mesangial cells | NaHS inhibits the ROS generation and cell proliferation, and downregulates the expressions of TGF-β1 and collagen IV in high glucose-incubated cells. | [45] |
| NaHS          | Renal tubular epithelial cells. | The activation of AMPK by H$_2$S prevents high glucose-induced NOX4 expression in epithelial cells. NaHS augments the expression of iNOS, this effect is involved in the protective effect of H$_2$S against high glucose-induced NOX4 expression, ROS generation, and matrix laminin expression. | [49] |
| NaHS          | Glomerular mesangial cells | H$_2$S activates the Nrf2 signaling pathway to restrain high glucose-induced oxidative stress. H$_2$S exerts anti-inflammatory effects by blocking NF-κB signaling. Additionally, the cell proliferation induced by high glucose is mediated by MAPK signaling pathways, which is impeded by H$_2$S. Supplementation of H$_2$S represses the cell proliferation, inhibits TGF-β1 and collagen IV expressions, and attenuates the elevation of ROS in high glucose-treated cells. | [50] |
| NaHS          | Glomerular mesangial cells | Meanwhile, AGT, ACE and AT1 receptor mRNA levels and Ang II concentration are upregulated in high glucose-challenged cells, which are diminished by H$_2$S. | [86] |
| NaHS          | Renal tubular epithelial cells | H$_2$S on TGF-β1-induced renal EMT in renal tubular epithelial cells, as evidenced by upregulated levels of E-cadherin, along with downregulated expressions of α-SMA and fibronectin. The activation of mTORC1 and inactivation of AMPK are involved in global matrix protein synthesis, and these events are all reversed by NaHS. Importantly, NaHS stimulates AMPK phosphorylation and restores AMPK phosphorylation induced by high glucose. | [156] |
| NaHS          | Renal tubular epithelial cells | H$_2$S upregulates the expression of HO-1 in both mesangial and podocyte cells. H$_2$S might have the ability to upregulate this antioxidant enzyme, which may be a potential mechanism by which H$_2$S exerts its protective effects. | [157] |
| NaHS          | Glomerular mesangial cells, podocytes | Exogenous H$_2$S treatment mitigates the proliferation of mesangial cells. Furthermore, H$_2$S supplementation remarkably inhibits TLR4 expression and curbs the mesangial cell proliferation. High glucose stimulation significantly reduces nephrin, ZO-2, and CSE expression levels, and elevates β-catenin production in mouse podocytes. Supplementation of NaHS rectifies these changes. Exogenous H$_2$S may alleviate high glucose-induced podocyte injury possibly through ZO-2 upregulation and the subsequent suppression of Wnt/β-catenin pathway. | [169] |
| AP39, AP106, AP72, AP67, GYY4134 | Glomerular mesangial cells | GYY4137 upregulates miR-194 level to mitigate ROS production under high glucose condition. | [170] |
| NaHS          | Glomerular endothelial cells | GYY4137 upregulates miR-194 level to mitigate ROS production under high glucose condition. | [170] |
Table 2. Beneficial effect of H₂S on experimental diabetic nephropathy in vivo.

| H₂S Donors | Animal Models | Main Findings                                                                                                                                                                                                 | Ref.  |
|------------|---------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-------|
| NaHS       | STZ-induced diabetic rats | Administration of NaHS reverses the increases in TGF-β1 and collagen IV in diabetic rats. H₂S attenuates glomerular basement membrane thickening, mesangial matrix deposition, and renal interstitial fibrosis, thereby improving renal function in diabetic rats. The protein expressions of ACE and AT1 receptors as well as Ang II are significantly up-regulated in diabetic kidneys and down-regulated after treatment with H₂S. | [45]  |
| NaHS       | STZ-induced diabetic rats | In STZ-induced diabetic rats, the changes in RAS are reversed by H₂S supplementation without affecting blood glucose concentration. The increased 24 h urinary protein, fasting blood glucose (FBG), blood urea nitrogen (BUN), serum creatinine (Scr) and renal index, as well as the elevated amount of glomerular mesangial matrix in diabetic rats are all ameliorated by H₂S treatment. In addition, the diabetic kidney shows the increased MDA content, caspase-3 activity and Bax expression, but decreased SOD activity and Bcl-2 expression, which are normalized by administration of H₂S. | [50]  |
| NaHS       | STZ-induced diabetic rats | The increased 24 h urinary protein, fasting blood glucose (FBG), blood urea nitrogen (BUN), serum creatinine (Scr) and renal index, as well as the elevated amount of glomerular mesangial matrix in diabetic rats are all ameliorated by H₂S treatment. In addition, the diabetic kidney shows the increased MDA content, caspase-3 activity and Bax expression, but decreased SOD activity and Bcl-2 expression, which are normalized by administration of H₂S. | [86]  |
| NaHS       | Mice with type 1 diabetes or type 2 diabetes | Renal cortical contents of CBS and CSE are significantly reduced, alongside with renal hypertrophy and matrix accumulation in mice with type 1 diabetes or type 2 diabetes. | [157] |
| GYY4137    | Diabetic Akita mouse | GYY4137 prevents collagen deposition and realignment and renal fibrosis in mice. The increased expressions of MMP-9, MMP-13 and MMP-14, and reduced vascular density in diabetic kidney are reversed by GYY4137. | [187] |

Table 3. Beneficial effect of H₂S-releasing compounds on experimental diabetic nephropathy.

| H₂S-Releasing Compounds | Animal Models/Cell Type | Main Finding                                                                                                                                                                                                                                                                                                                                 | Ref.  |
|-------------------------|-------------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-------|
| S-propargyl-cysteine    | STZ-induced diabetic rat/mesangial cells | S-propargyl-cysteine, a H₂S-releasing compound, reduces the level of creatinine, kidney to body weight ratio and 24-h urine microalbuminuria excretion in STZ-induced diabetic kidney injury. The renal fibrosis, inflammation, and hypertrophy are suppressed by this compound. The renal protective effects of this compound may be mediated by inhibition of TGF-β1/Smad3 pathway and blockade of MAPK signaling pathway. Tadalafil, increases the expression and activity of the H₂S-generating enzyme CSE by accelerating its translation. It can effectively abrogate high glucose-induced global protein synthesis in podocytes. Tadalafil activates AMPK by stimulating calcium-calmodulin kinase β, thus attenuating the activation of mTOR induced by high glucose. Furthermore, in tadalafil-treated podocytes, the iNOS expression is rapidly upregulated. Knockdown or inhibition of iNOS abolished the effect of tadalafil on CSE expression and AMPK phosphorylation in podocytes. | [183]  |
| Tadalafil              | Podocytes               | Tadalafil activates AMPK by stimulating calcium-calmodulin kinase β, thus attenuating the activation of mTOR induced by high glucose. Furthermore, in tadalafil-treated podocytes, the iNOS expression is rapidly upregulated. Knockdown or inhibition of iNOS abolished the effect of tadalafil on CSE expression and AMPK phosphorylation in podocytes. | [184]  |
10. Current Molecular Mechanisms of H\textsubscript{2}S in Diabetic Kidney Disease

H\textsubscript{2}S has recently gained enormous attention in diabetic nephropathy since its critical role in the progression of diabetic nephropathy \[7,188\]. Circulating H\textsubscript{2}S levels appear to be reduced in type 2 diabetic subjects \[46\], and are inversely correlated with adiposity parameters \[47\]. Animal studies have highlighted the crucial role of H\textsubscript{2}S dysregulation in diabetic kidney disease. Sen and colleagues have revealed that induction of matrix metalloproteinase-9 (MMP-9) elicits the decreased production of H\textsubscript{2}S-synthesizing enzymes CBS and CSE in the diabetic kidney \[130\]. The same group further demonstrates that H\textsubscript{2}S donor GYY4137 ameliorates diabetic oxidative injury and renal fibrosis through miR-194-dependent pathway \[187\]. The downregulated miR-194 is restored by H\textsubscript{2}S, followed by diminished ROS production, and normalized MMP-9, MMP-13 and MMP-14 in glomerular endothelial cells under high glucose condition \[187\]. TGF-\textbeta is critically involved in the synthesis of matrix proteins and diabetic renal fibrosis \[189,190\]. The increased expressions of TGF-\textbeta and matrix proteins as well as urinary albumin loss in diabetic mice are reversed by H\textsubscript{2}S administration \[45,50\]. Additionally, the enhanced matrix protein synthesis is a contributor for renal hypertrophy \[191\]. It is well documented that PI3K-Akt-mammalian target of rapamycin (mTOR) signaling pathway plays a key role in matrix protein synthesis during diabetic kidney disease \[7\], since blockade of this pathway ameliorates renal injury in diabetic rodent models \[192\]. An additional signaling change in diabetic kidney is reduced activity of 5'-monophosphate (AMP)-activated protein kinase (AMPK), which is indicated to abrogate diabetic kidney injury \[193,194\]. In high glucose-treated renal epithelial cells, H\textsubscript{2}S donor NaHS stimulates AMPK activity, reduces mTOR complex-1 (mTORC1) activity and prevents matrix protein deposition \[157\]. NaHS also retards high glucose-induced mesangial proliferation through inhibition of mitogen-activated protein kinases (MAPK) activity \[50\]. The existing data demonstrate that the important signaling pathways recruited by H\textsubscript{2}S are sufficient to ameliorate diabetic nephropathy. However, more research is required to elucidate additional mechanisms by which H\textsubscript{2}S exerts a protective effect in diabetic kidney injury. Next, in detail, we will outline the current studies about the interactions between dysregulated H\textsubscript{2}S and other renal lesions in diabetic kidney disease.

11. Conclusions and Perspectives

The novel and constructive insights into progressive diabetic nephropathy become urgent due to the continuously increasing incidence of diabetes and obesity. Continuous experimental studies have been performed to identify the possible molecular mediators in diabetic kidney disease. As the third gaseous mediator after NO and CO, the H\textsubscript{2}S signaling pathway may provide potential therapeutic target for the treatment of diabetic kidney disease. Targeting the H\textsubscript{2}S signaling pathway may be postulated to be a reprogramming strategy against diabetic kidney disease. However, many questions remain regarding which renal cells are most affected by decreased H\textsubscript{2}S level and why the renal H\textsubscript{2}S level is downregulated in response to hyperglycemia? As of yet, few studies evaluate the temporal patterns of H\textsubscript{2}S activation and it is probable that timing will have a profound effect on outcomes. Focused studies investigating the roles of H\textsubscript{2}S in humans or rodent models will broaden our understanding of the H\textsubscript{2}S signaling pathway in diabetic kidney disease.

Based on the underlying mechanisms whereby H\textsubscript{2}S exerts protective effects against diabetic kidney disease, the precursors for H\textsubscript{2}S synthesis, H\textsubscript{2}S donors, and natural plant-derived compounds for H\textsubscript{2}S generation are indicated to ameliorate tubular lesions, and reverse renal injury in diabetes. However, the challenge still exists, the regression of renal lesions and recovery of renal structure should be the focus in future novel therapeutic strategies. Similarly, it should be mentioned that the possible molecular mechanisms of H\textsubscript{2}S in diabetic nephropathy are not fully understood. Thereafter, in-depth understanding of interaction between H\textsubscript{2}S and its downstream targeted genes will undoubtedly help to propel the development of H\textsubscript{2}S-medaited novel therapies that can halt or reverse diabetic renal disease. Moreover, more research is required to understand how H\textsubscript{2}S dysfunction interacted with other pathogenic factors in diabetic kidney disease.
A wide range of impressive studies focusing the role of H$_2$S in diabetic renal damage are growing rapidly in recent years. Accordingly, the expressions of H$_2$S-producing enzymes and levels of its precursors and metabolites are abnormal in diabetic renal pathology. With this in mind, it is interesting to know whether H$_2$S or its metabolites are thought to be biomarkers for renal disease severity in diabetes. Nevertheless, only a few studies have addressed this uncertainty. To answer this possibility, large population studies investigating the clinical value of H$_2$S metabolites as predictors of diabetic kidney disease are underway.

The emerging approach that is highly relevant to the H$_2$S signaling pathway is a flourishing field because of the increasing epidemic of diabetic kidney disease. However, these results await further clinical translation. N-acetylcysteine (NAC) is a derivate of cysteine and it is the main substrate for H$_2$S production [195]. Therefore, NAC may be able to promote H$_2$S production in humans. For this reason, a clinical trial had investigated the effect of NAC on the production of H$_2$S in patients with CKD several years ago (ClinicalTrials.gov Identifier: NCT01232257). However, to date, it is still unknown whether NAC could increase H$_2$S levels in CKD patients or dialysis patients. As such, more large-scale clinical studies are needed to further determine the protective role of H$_2$S in kidney diseases including diabetic nephropathy.

Taken together, since the impaired H$_2$S signaling pathway appears to be one of the denominators for diabetic renal injury, it may be a suitable target to ameliorate renovascular complications of diabetes. With more and more excited discoveries regarding H$_2$S functions in renal physiology and disease, we expect multiple innovative H$_2$S applications to evolve in the near future.

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