**A role for dynamic contrast-enhanced magnetic resonance imaging in predicting tumour radiation response**

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**Background:** Dynamic contrast-enhanced (DCE) MRI may provide prognostic insights into tumour radiation response. This study examined quantitative DCE MRI parameters in rat tumours, as potential biomarkers of tumour growth delay following single high-dose irradiation.

**Methods:** Dunning R3327-AT1 prostate tumours were evaluated by DCE MRI following intravenous injection of Gd-DTPA. The next day tumours were irradiated (single dose of 30 Gy), while animals breathed air (n = 4) or oxygen (n = 4); two animals were non-irradiated controls. Growth was followed and tumour volume-quadrupling time (T₄) was compared with pre-irradiation DCE assessments.

**Results:** Irradiation caused significant tumour growth delay (T₄ ranged from 28 to 48 days for air-breathing rats, and 40 to 75 days for oxygen-breathing rats) compared with the controls (T₄ = 7 to 9 days). A strong correlation was observed between T₄ and extravascular-extracellular volume fraction (vₑ) irrespective of the gas inhaled during irradiation. There was also a correlation between T₄ and volume transfer constant (Kᵣ) for the air-breathing group alone.

**Conclusions:** The data provide rationale for expanded studies of other tumour sites, types and progressively patients, and are potentially significant, as many patients undergo contrast-enhanced MRI as part of treatment planning.

Tumours exhibit a range of response to therapy and there is an increasing desire to develop prognostic biomarkers to predict therapeutic efficacy. Proteomic and genomic screens may be related to tumour aggressiveness, but are often based on invasive biopsy specimens. Imaging potentially offers entirely non-invasive insight into tumour pathophysiology, which may influence tumour development and response to therapy.

Hypoxia is thought to influence radiation response and there have been many clinical trials aimed at exploiting or overcoming hypoxia (Wilson and Hay, 2011; Horsman et al, 2012). However, the meta-analysis of Overgaard (2007) suggested only a minor benefit for procedures designed to overcome hypoxia. There is general consensus that such trials are hampered by inability to identify hypoxic tumours and stratify patients for optimal therapy. This has led to extensive interest in developing biomarkers of hypoxia (Tatum et al, 2006). Perhaps the most effective, to date, has been the Eppendorf Histograph, which was successfully used to stratify human tumours at various disease sites based on various...
DCE MRI for predicting radiation response

BRITISH JOURNAL OF CANCER

oxygen-related criteria such as hypoxic fraction, median pO2 or ratio of tumour to local normal tissue (Falk et al, 1992; Fyles et al, 1998; Rosfjord et al, 2006; Vaupel and Mayer, 2007). However, electrode application is highly invasive and is seldom used today. Hypoxia may also be identified histologically based on bioreductive markers such as pimonidazole, and this strategy was successfully applied in head and neck cancer (Kaanders et al, 2004). Ideally, stratification would be achieved non-invasively and indeed multiple strategies are being developed using MRI, ESR, optical and radionuclide imaging (Tatum et al, 2006; Mason et al, 2010; Fleming et al, 2015), although to date, none has been approved for routine clinical use.

Several studies have suggested that DCE MRI may provide insights into tumour hypoxia and radiation response, as well as perfusion (Zahra et al, 2007; Horsman et al, 2012), and correlations have been shown with electrode measurements and radiobiological hypoxic fractions (Lyng et al, 2001; Loncaster et al, 2002; Wang et al, 2002; Egeland et al, 2012). Hitherto, such studies have generally fallen into two classes: (i) pre-clinical animal investigations with quantitative DCE analysis, but in many cases assessment of radiation response was limited to paired clonogenic survival assays (Gulliksrud et al, 2011; Egeland et al, 2012; Ovreo et al, 2012); (ii) clinical and large animal studies of response, generally relating semi-quantitative analysis of DCE based on relative signal enhancement, or initial area under the curve (Lyng et al, 2001; Loncaster et al, 2002; Mayr et al, 2009; Viglianti et al, 2009). Contrary results have been reported with more or less correlation between specific DCE parameters and hypoxia (Ovreo et al, 2012). As such, we decided to include DCE MRI as part of a larger MRI investigation of oxygen-modulated tumour response to radiation. We have published the oxygen-enhanced (OE) MRI previously (Hallac et al, 2014), and now present the more complex analysis of DCE MRI.

Contrast-enhanced MRI is a routine clinical scan used to examine many tumour disease sites. It requires the administration of small molecular weight gadolinium (Gd) chelates that can aid in tumour delineation by virtue of the highly fenestrated leaky vasculature. Beyond the assessment of temporal and spatial variation in signal intensity following the administration of gadolinium, this technique has been widely used to evaluate tumour pathophysiology (Evelhoch, 1999; Tofts et al, 1999; Alonzi et al, 2007; Eyal and Degani, 2009). Sophisticated pharmacokinetic analysis has been developed initially by direct analogy with radiotracers methods (Tofts, 1997; Tofts et al, 1999). Several mathematical models, such as the modified Kety/Tofts model (Tofts et al, 1999), have been established to generate parametric images of volume transfer constant (Ktrans) and extravascular-extracellular volume fraction (ve).

In this study, we evaluated the potential use of DCE MRI for predicting the radiation response of the Dunning prostate R3327-AT1 rat tumour. The day following MR imaging, tumours were irradiated with a single high dose of 30 Gy and growth delay was compared with pre-irradiation DCE assessments. These tumours formed a subset of a larger cohort evaluated previously for OE-MRI. At the time, we were unable to evaluate the DCE MRI and therefore this study is presented separately here, in the context of the previous work (Hallac et al, 2014).

MATERIALS AND METHODS

Animal model. This study was approved by the UT Southwestern Institutional Animal Care and Use Committee in accordance with Federal, State, and Local laws and guidelines and consistent with Guidelines for the welfare and use of animals in cancer research (Workman et al, 2010). Ten male Copenhagen rats (weight ~200 g) were implanted with Dunning R3327-AT1 rat prostate tumours. The AT1 is an anaplastic tumour with a volume-doubling time of 5.2 days and a TCD50 (single radiation dose for 50% tumour control probability) of 75.7 Gy (Peschke et al, 1998, 2011). Tumours were originally provided by Dr J.T. Isaacs of Johns Hopkins University (Isaacs et al, 1986). Donor tissue was grown in the thigh of Copenhagen rats and fresh tumour tissue fragments were collected and surgically subcutaneously in the thigh via a small incision. Tumour volume was measured using a pair of calipers and calculated as V = (π/6) a × b × c, where a, b and c are the three orthogonal diameters. When tumours reached about 0.3 cm3 they were evaluated by MRI and irradiated.

MRI. MRI used a horizontal bore 4.7-T magnet system (Agilent, Palo Alto, CA, USA) with home-built 2 cm single-turn solenoid volume coil. Animals were anaesthetised with isoflurane (1.5%) in air (1 dm3 min–1) and kept warm (37 °C) using a circulating warm water blanket. High-resolution T2-weighted images were acquired using a Fast Spin-Echo (FSEMS) sequence: TR/TEeff = 2000/48 ms, echo train length (ETL) = 8, FOV = 40 × 40 mm, acquisition matrix 128 × 128, NEX = 8, 1 mm slice thickness without gap, and 15 slices acquired in the axial orientation. Interleaved BOLD (blood oxygen level dependent) and TOLD (tissue oxygen level dependent) (IBT) MRIs were acquired in an automated fashion using a home-written macro during air breathing followed by oxygen, as described in detail previously (Hallac et al, 2014). Following about 30 min oxygen breathing, quantitative R1 measurements were acquired with O2-breathing using a spin-echo sequence (SEMS): TE = 20 ms, TR = 0.1, 0.2, 0.3, 0.5, 0.7, 0.9, 1.5, 2.5, 3.5 s, FOV = 40 × 40 mm2 with 64 × 64 acquisition matrix. Time course DCE data were acquired continuously using a SEMS with TR/TE of 200/15 ms providing 11 s temporal resolution pre-and up to 30 min post-injection of 0.1 mmol kg–1 body weight Gd-DTPA (Magnevist) intravenously in the tail vein rapidly by hand. Images were acquired with the same spatial resolution as R1 maps. R1 maps were obtained before contrast injection in order to facilitate conversion of signal-intensity data to contrast agent concentrations.

Radiation treatment. Radiation was delivered using a dedicated small animal X-ray irradiator (XRAD 225Cx, Precision X-Ray, Inc., North Brantford, CT, USA). The XRAD 225Cx features an X-ray tube mounted on a c-arm gantry, which rotates about a motorised x–y–z (3D) translational stage (Parker 404XE, Parker–Hannifin Corp., Irwin, PA, USA). Absolute-dose calibration was performed in accordance with the recommendations of the AAPM TG-61 protocol (Ma et al, 2001). The XRAD 225Cx was operated at 225 kV and 13 mA, producing a dose rate of ~3.5 Gy min–1. A circular aperture of 20 mm in diameter was used to limit the radiation beam only to the tumour. An image-guidance system utilising a digital-imaging panel for digital radiography, fluoroscopy and cone-beam CT was used to ensure accurate target localisation.

Rats were divided into three groups: Group 1 control (n = 2, no irradiation); Group 2 (n = 4) breathed air during irradiation and Group 3 (n = 4) breathed oxygen 15 min before and during irradiation. Half the radiation dose (15 Gy) was delivered Dorsal–Ventral, which was followed immediately by another dose of 15 Gy from the opposite side (Figure 1). Following irradiation tumour volume (V), measured with a pair of calipers, was normalised to the day of irradiation (V0) and response was evaluated as the time for the tumour to quadruple in volume (T4). Rats were killed within 80 days of irradiation.

Data analysis. DCE data were analysed using a recent reference tissue model (Yankeelov et al, 2005) avoiding the need to directly measure the arterial input function (AIF). The model compares the contrast agent curves in a tissue of interest (tumour) to that of a
reference region (muscle). Using reported values for the volume transfer constant of muscle, \( K_{\text{trans}} \), (0.1 min\(^{-1}\)) and the extravascular-extracellular volume fraction, \( v_e \), (0.1) in the muscle (Padhani, 2002, Yankeelov et al, 2005), it is possible to extract the \( K_{\text{trans}} \) and \( v_e \) values for the tumour without knowledge of the AIF. In this model, the contrast concentration curve, \( C(t) \), in tumour tissue is given by (Yankeelov et al, 2005):

\[
C(t) = R \times C_0(t) + R \times \left[ \frac{K_{\text{trans}, \text{M}}}{v_e} - \frac{K_{\text{trans}, \text{M}}}{v_e} \right] \times \int_0^t \left( C_A(t') \times \exp\left( -
\frac{K_{\text{trans}, \text{M}}}{v_e} \times (t-t') \right) \right) dt'
\]

where, \( R = (K_{\text{trans}, \text{M}}/K_{\text{trans}, \text{M}}) \) and \( C_0(t) \) is the contrast agent concentration curve in muscle tissue. Analysis of BOLD and TOLD data (both semi-quantitative changes in signal intensity (ΔSI) and quantitative \( R_1 \) and \( R_2^* \)) was described in detail previously (Hallac et al, 2014), and the results are compared with the new DCE data.

Statistics. Pearson correlation analysis was performed between the DCE-derived parameters and tumour growth delay. The significance of differences between Groups was established using analysis of variance. The statistical analysis was carried out by using Excel 14 (Microsoft, Redmond, WA, USA) software.

RESULTS

A baseline T\(_2\)-weighted MR image slice is shown for a representative tumour (Figure 2A) together with heterogeneous contrast following administration of Gd-DTPA (Figure 2B). The \( K_{\text{trans}} \) and \( v_e \) maps (Figure 2C and D) were heterogeneous with most high values in the periphery. Mean \( K_{\text{trans}} = 0.11 \pm 0.05 \) min\(^{-1}\) and mean \( v_e = 0.19 \pm 0.09 \) (8 treated tumours).

Caliper measurement of tumour growth was obtained up to four times the initial tumour size or 80 days following irradiation. Untreated controls had the fastest growth rate with a T\(_4\) of 40–44 days; P < 0.4 vs air. Others showed significantly enhanced tumour growth delay (T\(_4\) = 70–75 days (n = 2); P < 0.005 vs air and P < 0.01 vs the unresponsive tumours).

A strong correlation was observed between \( v_e \) and \( T_4 \) for those tumours breathing air during irradiation (Group 2; \( r^2 > 0.93; P < 0.05 \); Figure 4). Intriguingly a strong correlation remained if all tumours, irrespective of inhaled gas were combined (Groups 2 + 3; \( r^2 > 0.72; P < 0.001 \)). For \( K_{\text{trans}} \) there was strong correlation with \( T_4 \) for Group 2 tumours (\( r^2 > 0.99; P < 0.005 \)), but this failed upon inclusion of Group 3 tumours (\( r^2 > 0.3 \)). There was a significant correlation between \( v_e \) and \( K_{\text{trans}} \) for the eight irradiated tumours (\( r^2 > 0.53; P < 0.05 \)).

The OE-MRI was analysed and published previously showing strong correlation between ΔSI TOLD, \( AR_1, \) \( AT_1 \), and \( T_4 \) (Hallac et al, 2014). No such correlations were observed between BOLD and \( T_4 \). We have now examined relationships between OE-MRI and DCE, but in general, there was lack of correlation, although TOLD (%ΔSI) vs \( v_e \) showed \( r^2 > 0.54 \) (P < 0.05) and \( R_1 \) vs \( K_{\text{trans}} \) showed \( r^2 > 0.59 \) (P < 0.05).
Distinct trends were observed between DCE MRI parameters and radiation treatment. Tumours exhibiting greater $v_e$ values had longer tumour growth delay, irrespective of the inhaled gas. There was also a strong correlation between $K^{\text{trans}}$ and tumour growth delay for the air-breathing rats alone.

In an extensive series of studies, Roñstad et al. (2000) examined relationships between quantitative DCE parameters, hypoxia and radiation response (Gulliksrud et al., 2011; Egeland et al., 2012; Ovrebo et al., 2012). For cohorts of eight human melanoma lines, exhibiting varying degrees of radiobiological hypoxia when growing as xenografts in nude mice, Egeland et al. (2012) reported robust correlations between $v_e$ or $K^{\text{trans}}$ and radiobiological hypoxic fractions. Ovrebo et al. (2012) found that $K^{\text{trans}}$ was significantly higher for individual radiation sensitive tumours, but $v_e$ did not differ. Several of these studies examined tumour heterogeneity and sought to discard voxels corresponding to necrotic regions (Gulliksrud et al., 2011; Egeland et al., 2012). We examined tumour images of whole cross sections with no attempt to eliminate potential necrosis. None of the previous studies examined tumour growth delay, but rather the more traditional paired clonogenic survival assays for cells from tumours on anaesthetised mice irradiated, while breathing air or shortly after killing to induce hypoxia (Egeland et al., 2012; Ovrebo et al., 2012). Our approach was somewhat different using air or oxygen breathing to modulate hypoxia and assessing tumour growth delay over several weeks.

The magnitudes of $K^{\text{trans}}$ and $v_e$ observed in this study lie in the ranges reported for various tumours such as human melanoma xenografts in mice (Gulliksrud et al., 2011; Ovrebo et al., 2012), and Dunning prostate R3327-AT6.1 tumours in rats (Fan et al., 2010). We observed a significant correlation between $K^{\text{trans}}$ and $v_e$ ($r^2 > 0.53$). We used the reference tissue approach for analysis thereby avoiding the need for a direct measurement of AIF. Also, attempts to measure AIF directly may be confounded by blood vessel motion, flow and partial volume (Faranesh and Yankeelov, 2008). Validity of the reference region model is subject to inter- and intra-subject variability, but the method is widely used and accepted. A reference region model appears particularly suitable in this study, as the reference tissue (thigh muscle) is immediately adjacent to the tumour in the leg and should have similar blood supply. The reference tissue method avoids the need for a large supply blood vessel within the field of view and relaxes the need for extremely high temporal resolution. The search for prognostic biomarkers is complex and consensus may be difficult to attain, as reports consider many different disease sites, tumour sizes and therapies. Here, we examined response to single high-dose irradiation, essentially stereotactic ablation radiotherapy. The AT1 is reported to be quite radiation resistant and 30 Gy represents slightly less than half the TCD50 (Peschke et al., 1998, 2011). Tumour growth delay following single-dose irradiation should match the paired survival assays following single dose. Much irradiation follows a fractionated radiation therapy protocol and it was reported that the response of hepatocellular carcinoma could be predicted based on DCE characteristics after 2 weeks of therapy, but not baseline (Liang et al., 2007).

Viglianti et al. (2009) found that pretreatment semi-quantitative DCE parameters were predictive of therapeutic response of canine soft tissue sarcoma to thermoradiotherapy. In many cases, semi-quantitative approaches have been used to characterise tumours based on simple changes in $T_1$ - or $T_2$-weighted signal intensity, time to reach maximum signal or rates of signal change (Alonzi et al., 2007). The most widely used measurement is the initial area under the curve (IAUC) following infusion of contrast agent and we previously observed a wide range of values in the AT1 tumour, which correlated with BOLD signal response in some tumours (jiang et al., 2004). We did find that semi-quantitative assessments were indicative of response to metronomic chemotherapy in AT1 tumours (Zhao et al., 2005).

The correlation between $v_e$ and $T_d$ might be explained by the larger extracellular extravascular space in the tumour, and therefore, less cellular density and oxygen consumption. Several factors affect the development of hypoxia in tumours including vascularity of the surrounding normal tissue, angiogenic factor secretion and the oxygen consumption of tumour cells (Moulder and Rockwell, 1984; Gulledge and Dewhirst, 1996). The AT1 tumour line is highly radiation resistant, anaplastic undifferentiated and variably hypoxic (Zhao et al., 2003; Hallac et al., 2014). In general large AT1 tumours are found to exhibit extensive hypoxia, whereas small tumours (<2 cm$^3$) may be more or less well oxygenated (Zhao et al., 2003; Hallac et al., 2014). Moreover, AT1 tumours, which are hypoxic, tend not to respond to oxygen-breathing gas (Hunjan et al., 2001; Bourke et al., 2007; Hallac et al., 2014).

The tumour growth delay observed here is in line with previous reports for this subline, notably $T_d = 11.6 \pm 1.3$ days and $56.0 \pm 8.3$ days for control and 30 Gy, respectively (Peschke et al., 1998). That study made no attempt to assess or modulate tumour oxygenation and anaesthetised rats breathed 33% O$_2$ during irradiation. In another study, we reported that $T_d$ (the time to double in volume) was a function of tumour pO$_2$ based on $^{19}$F MRI relaxometry (Bourke et al., 2007). We attempted to modify levels of oxygenation and compared response with irradiation. There was no significant difference in $T_d$ between tumours irradiated, while breathing air or oxygen and the study was terminated before $T_d$. However, it was noted that those tumours in the air-breathing group continued growing monotonically, albeit somewhat more slowly, whereas...
those irradiated while breathing oxygen showed a peak volume around 10–15 days post irradiation and then tended to shrink. Those studies required direct injection of hexafluorobenzene reporter molecule, which is unlikely to become practical in patients and is inconvenient in pre-clinical studies.

In a more recent study, we had explored OE-MRI, as well as the DCE MRI presented here. Although various reports have indicated that BOLD response to oxygen-breathing challenge is related to changes in pO2 (Al-Hallaq et al, 1998; Baudelet and Gallez, 2002; Zhao et al, 2009), we found no significant correlation between R2* or T2W-weighted signal response and tumour growth delay. We did however determine a correlation between TOLD (Al-Hallaq et al, 2009), found no specific correlation between R1 or T1Ws signal responses) and T4 (Hallac et al, 2014). It was possible to stratify oxygen-breathing animals into responders and non-responders, as might be expected based on the dependence of R1 on pO2 (Edelman et al, 1996; Matsumoto et al, 2006; Beeman et al, 2015).

Group 3 showed no overall benefit from oxygen breathing compared with Group 2. However, a wide range of T4 (28–75 days) was observed and this was strongly correlated with ve for all tumours. A similar correlation was seen for T4 and Ktrans, although for Group 2 (air breathing) tumours only. Such observations could be of value in treatment planning. Moreover, those tumours which benefited significantly from oxygen breathing were identified based on a large TOLD response (Hallac et al, 2014) These results imply a role for combined OE- and DCE MRI.

Although this study is small, we believe it provides significant new data commensurate with previous reports. Notably, we now find that the relationships between Ktrans and ve vs tumour growth delay in syngeneic rat tumours are consistent with clonogenic paired-cell surviving fractions in studies of multiple human melanoma xenografts in mice. Clearly, it will be important to develop this observation in additional tumour types and with respect to other therapy paradigms. DCE is more commonly used to identify and delineate tumours (Alonzi et al, 2007) and the potential for predicting radiation response represents a bonus.

CONCLUSION

A strong correlation was observed between ve and tumour growth delay, irrespective of the inhaled gas during irradiation. No correlations were observed between OE-MRI and tumour growth delay and there were minimal correlations between OE-MRI and DCE parameters. Although contrast is part of most radiological examinations, it suggests that high temporal resolution DCE MRI could readily be implemented to provide predictive insight into response to radiation. It should be noted that this study used a single high-dose irradiation and this is becoming increasingly relevant as radiation oncology practice moves towards hypofractionated regimens such as stereotactic ablative radiotherapy (SABR).

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

Al-Hallaq HA, River JN, Zamora M, Okawa H, Karczmar GS (1998) Correlation of magnetic resonance and oxygen microelectrode measurements of carbogen-induced changes in tumor oxygenation. Int J Radiat Oncol Biol Phys 41: 151–159.

Alonzi R, Padhani AR, Allen C (2007) Dynamic contrast enhanced MRI in prostate cancer. Eur J Radiol 63: 335–350.

Baudelet C, Gallez B (2002) How does blood oxygen level-dependent (BOLD) contrast correlate with oxygen partial pressure (pO2) inside tumors? Magn Reson Med 48: 980–986.

Beeman SC, Shui Y-B, Perez-torres CJ, Engelbach JA, Ackerman JJH, Garbow JR (2015) O2-sensitive MRI distinguishes brain tumor vs radiation necrosis in murine models. Magn Reson Med; e-pub ahead of print 14 July 2015; doi:10.1002/mrm.25821.

Bourke VA, Zhao D, Gilio J, Chang C-H, Jiang L, Hahn EW, Mason RP (2007) Correlation of radiation response with tumor oxygenation in the dunning prostate R3327-AT1 tumour. Int J Radiat Oncol Biol Phys 67: 1179–1186.

Edelman RR, Hatibu H, Tadamura E, Li W, Prasad PV (1996) Noninvasive assessment of regional ventilation in the human lung using oxygen-enhanced magnetic resonance imaging. Nat Med 2: 1236–1239.

Egeland TAM, Gulliksrud K, Gaustad JV, Mathiesen B, Rosfjord EK (2012) Dynamic contrast-enhanced-MRI of tumor hypoxia. Magn Reson Med 67: 519–530.

Evelloch JL (1999) Key factors in the acquisition of contrast kinetic data for oncology. J Magn Reson Imaging 10: 254–259.

Eyal E, Degani H (2009) Model-based and model-free parametric analysis of breast dynamic-contrast-enhanced MRI. NMR Biomed 22: 40–53.

Falk S, Ward R, Bleehen N (1992) The influence of carbogen breathing on tumor tissue oxygenation in man evaluated by computerized pO2-histography. Br J Cancer 66: 919–924.

Fan X, Mustafi D, Zamora M, River JN, Foxley S, Karczmar GS (2010) Can DCE-MRI assess the effect of green tea on the angiogenic properties of rodent prostate tumors? Physica Medica 26: 111–116.

Fararsh AZ, Yankeelov TE (2008) Incorporating a vascular term into a reference region model for the analysis of DCE-MRI data: a simulation study. Phys Med Biol 53: 2617–2631.

Fleming IN, Manavaki R, Blower PJ, West C, Williams KJ, Harris AL, Domarkas J, Lord S, Baldry C, Gilbert FJ (2015) Imaging tumor hypoxia with positron emission tomography. Br J Cancer 112: 238–250.

Fyles AW, Milosevic M, Wong R, Kavanagh MC, Pintilie M, Sun A, Chapman W, Levin W, Manchul L, Keane TJ, Hill RP (1998) Oxygenation predicts radiation response and survival in patients with cervix cancer. Radiother Oncol 48: 149–156.

Gulledge CJ, Dewhirst MW (1996) Tumor oxygenation: a matter of supply and demand. Anticancer Res 16: 741–749.

Gulliksrud O, Ovrebo KM, Mathiesen B, Rosfjord EK (2011) Differentiation between hypoxic and non-hypoxic experimental tumors by dynamic contrast-enhanced magnetic resonance imaging. Radiother Oncol 98: 360–364.

Hallac RR, Zhou H, Picditik R, Song K, Stojadinovic S, Zhao D, Solberg T, Peckhe S, Mason RP (2014) Correlations of noninvasive BOLD and TOLD MRI with pO2, and relevance to tumor radiation response. Magn Reson Med 71: 1863–1873.

Horsman MR, Mortensen LS, Petersen JB, Busk M, Overgaard J (2012) Imaging hypoxia to improve radiotherapy outcome. Nat Rev Clin Oncol 9: 674–687.

Hunjan S, Zhao D, Constantinescu A, Hahn EW, Antich PP, Mason RP (2001) Tumor Oximetry: demonstration of an enhanced dynamic mapping procedure using fluorescein-19 echo planar magnetic resonance imaging in the Dunning prostate R3327-AT1 rat tumor. Int J Radiat Oncol Biol Phys 49: 1097–1108.

Isaacs JT, Isaac WB, Feitz WFJ, Scheres J (1986) Establishment and characterization of 7 Dunning prostate cancer cell lines and their use in developing methods for predicting metastatic ability of prostate cancer. Prostate 9: 261–281.

Jiang L, Zhao D, Constantinescu A, Mason RP (2004) Comparison of BOLD contrast and Cd-DTPA dynamic contrast enhanced imaging in rat prostate tumor. Magn Reson Med 51: 953–960.

Kaaners JHAM, Bussink J, Van Der Kogel AJ (2004) Clinical studies of hypoxia modification in radiotherapy. Sem Radiat Oncol 14: 233–240.
DCE MRI for predicting radiation response

Le QT, Chen E, Salim A, Cao HB, Kong CS, Whyte R, Dunington J, Cannon W, Wakele H, Tishirani R, Mitchell JD, Richardson D, O’Byrne KJ, Koong AC, Giaccia AJ (2006) An evaluation of tumor oxygenation and gene expression in patients with early stage non-small cell lung cancers. *Clin Cancer Res* 12: 1507–1514.

Liang PC, Ch’ang HJ, Hsu C, Tseng SS, Shih TTF, Wu Liu T (2007) Dynamic MRI signals in the second week of radiotherapy relate to treatment outcomes of hepatocellular carcinoma: a preliminary result. *Liver Int* 27: 516–528.

Loncaster JA, Carrington BM, Sykes JR, Jones AP, Todd SM, Cooper R, Le QT, Chen E, Salim A, Cao HB, Kong CS, Whyte R, Dunington J, Donington J, DCE MRI for predicting radiation response. *Int J Radiat Biol Oncol Phys* 54: 759–767.

Lyng H, Vorren AO, Sundfor K, Taksdal I, Lien HH, Kaalhus O, Rofstad EK (2002) Prediction of radiotherapy outcome using dynamic contrast enhanced MRI of carcinoma of the cervix. *Int J Radiat Oncol Biol Phys* 54: 759–767.

Ovrebo KM, Hompland T, Mathiesen B, Rofstad EK (2012) Assessment of tumor oxygenation in human cervical carcinoma by use of dynamic Gd-DTPA-enhanced MR imaging. *J Magn Reson Imaging* 34: 259–280.

Matsumoto K, Bernardo M, Subramanian S, Choyke P, Mitchell JB, Krishna MC, Lizak MJ (2006) MR assessment of changes of tumor in response to hyperbaric oxygen treatment. *Magn Reson Med* 56: 240–246.

Mayr NA, Wang IZ, Zhang D, Montebello JF, Grecula JC, Lo SS, Fowler JM, Yuh WTC (2009) Synergistic effects of hemoglobin and tumor perfusion on tumor control and survival in cervical cancer. *Int J Radiat Oncol Biol Phys* 74: 1513–1521.

Moulder J, Rockwell S (1984) Hypoxic fractions of solid tumors, experimental technique, methods of analysis and survey of existing data. *Int J Radiat Oncol Biol Phys* 10: 695–712.

Mowas B, Chapman JD, Hanlon AL, Horwitz EM, Greenberg RE, Stobbe C, Hanks GE, Pollack A (2002) Hypoxic prostate/muscle Po-2 ratio predicts for biochemical failure in patients with prostate cancer: Preliminary findings. *Urology* 60: 634–639.

Overgaard J (2007) Hypoxic radiosensitization: adored and ignored. *J Clin Oncol* 25: 4066–4074.

Ovrebø KM, Hompland T, Mathiesen B, Rofstad EK (2012) Assessment of hypoxia and radiation response in intramuscular experimental tumors by dynamic contrast-enhanced magnetic resonance imaging. *Radiother Oncol* 102: 429–435.

Pashley AR (2002) Dynamic contrast-enhanced MRI in clinical oncology: current status and future directions. *J Magn Reson Imaging* 16: 407–422.

Peschke P, Hahn EW, Wenz F, Lohr F, Braunschweig F, Wolber G, Zuna I, Hao G, Thorpe P, Hahn EW, Peschke P (2010) Multimodality imaging of hypoxia in preclinical settings. *QF Nucl Med Mol Imaging* 54: 259–280.

Wang Z, Su M-Y, Nalcioglu O (2002) Applications of dynamic contrast enhanced mri in oncology: measurement of tumor oxygen tension. *Technol Cancer Res Treat* 1: 29–38.

Wilson WR, Hay MP (2011) Targeting hypoxia in cancer therapy. *Nat Rev Cancer* 11: 393–410.

Workman P, Aboagye EO, Balkwill F, Balmain A, Bruder G, Chaplin DJ, Double JA, Everitt J, Farningham DAH, Glennie MJ, Kelland LR, Robinson V, Stratford II, Tozer GM, Watson S, Wedge SR, Eccles SA, Navaratnam V, Ryder S. Committee of the National Cancer Research Institute (2010) Guidelines for the welfare and use of animals in cancer research. *Br J Cancer* 102: 1555–1577.

Yankeelov TE, Luci JJ, Lepage M, Li R, Debusk L, Lin PC, Price RR, Gore JC (2005) Quantitative pharmacokinetic analysis of DCE-MRI data without an arterial input function: a reference region model. *Magn Reson Imaging* 23: 519–529.

Zafar MA, Hollingsworth KG, Sala E, Lomas DJ, Tan LT (2007) Dynamic contrast-enhanced MRI as a predictor of tumour response to radiotherapy. *Lancet Oncol* 8: 63–74.

Zha D., Jiang L., Hahn EW, Mason RP (2005) Continuous low-dose (Metronomic) chemotherapy on rat prostate tumors evaluated using MRI in vivo and comparison with histology. *Neoplasia* 7: 678–687.

Zhao D., Jiang L., Hahn EW, Mason RP (2009) Comparison of 1H blood oxygen level-dependent (BOLD) and 19F MRI to investigate tumor oxygenation. *Magn Reson Med* 62: 357–364.

Zhao D., Ran S., Constantinescu A, Hahn EW, Mason RP (2003) Tumor oxygen dynamics: correlation of in vivo MRI with histological findings. *Neoplasia* 5: 308–318.

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