High prevalence of chronic pituitary and target-organ hormone abnormalities after blast-related mild traumatic brain injury

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INTRODUCTION

Recent studies investigating chronic pituitary dysfunction resulting from TBI have reported a prevalence of posttraumatic hypopituitarism (PTHP) ranging from 5 to 95% with a median of 35%, the variation being primarily due to differences in screening criteria (Bavisetty et al., 2008; Srinivasan et al., 2009; Berg et al., 2010; Englander et al., 2010; High et al., 2010; Kokshoorn et al., 2010, 2011; Krahulik et al., 2010; Park et al., 2010; Pavlovic et al., 2010; Reimunde et al., 2011; Schneider et al., 2011). Pituitary hormone disorders are frequent among the immediate consequences of TBI; some resolve during the following months while a smaller proportion of new dysfunctions emerge (Agha et al., 2005; Aimaretii et al., 2005; Schneider et al., 2006, 2011; Tanriverdi et al., 2006, 2008b; Klose et al., 2007; Krahulik et al., 2010). By ~6 months subsequent to TBI, the pattern of pituitary deficits is considered to be relatively permanent.

Studies of traumatic brain injury from all causes have found evidence of chronic hypopituitarism, defined by deficient production of one or more pituitary hormones at least 1 year after injury, in 25–50% of cases. Most studies found the occurrence of posttraumatic hypopituitarism (PTHP) to be unrelated to injury severity. Growth hormone deficiency (GHD) and hypogonadism were reported most frequently. Hypopituitarism, and in particular adult GHD, is associated with symptoms that resemble those of PTSD, including fatigue, anxiety, depression, irritability, insomnia, sexual dysfunction, cognitive deficiencies, and decreased quality of life. However, the prevalence of PTHP after blast-related mild TBI (mTBI), an extremely common injury in modern military operations, has not been characterized. We measured concentrations of 12 pituitary and target-organ hormones in two groups of male US Veterans of combat in Iraq or Afghanistan. One group consisted of participants with blast-related mTBI whose last blast exposure was at least 1 year prior to the study. The other consisted of Veterans with similar military deployment histories but without blast exposure. Eleven of 26, or 42% of participants with blast concussions were found to have abnormal hormone levels in one or more pituitary axes, a prevalence similar to that found in other forms of TBI. Five members of the mTBI group were found with markedly low age-adjusted insulin-like growth factor-I (IGF-I) levels indicative of probable GHD, and three had testosterone and gonadotropin concentrations consistent with hypogonadism. If symptoms characteristic of both PTHP and PTSD can be linked to pituitary dysfunction, they may be amenable to treatment with hormone replacement. Routine screening for chronic hypopituitarism after blast concussion shows promise for appropriately directing diagnostic and therapeutic decisions that otherwise may remain unconsidered and for markedly facilitating recovery and rehabilitation.

Keywords: traumatic brain injury, hypopituitarism, blast, concussion, growth hormone, pituitary

The risk factors and the mechanisms, other than immediate trauma-induced tissue damage and subsequent edema, for chronic hypothalamo-pituitary dysfunction due to TBI are unclear. Roles for polymorphisms in apolipoprotein E genotype (APOE), inflammatory processes – both systemic and neural, and anti-hypothalamic (AHAs) and anti-pituitary antibodies (APAs) have been proposed, and each has empirical support.

There is evidence that the apolipoprotein E (APOE) ε3/ε3 genotype may be associated with a reduced risk of TBI-related hypopituitarism. APOE ε3 is the most common of the three alleles and is found in more than half of the general population. The ε2 and ε4 alleles have been associated with altered risks for Alzheimer’s disease, hyperlipoproteinemia, and atherosclerosis. Pituitary dysfunction in patients with TBI has been found to be significantly less prevalent in individuals with the APOE ε3/ε3 genotype (17.7%)
Evidence for the involvement of APAs and/or AHAs in the development of chronic PTHP comes from two studies. APAs were detected in 44.8% of patients who had completed a 3-year follow-up after TBI and in none of the healthy control subjects, and the prevalence of hypopituitarism was significantly higher in APA-positive (46.2%) than APA-negative TBI patients (12.5%; \( p = 0.04 \); Tanriverdi et al., 2008b). In another study of active and retired boxers, AHAs were detected in 21.3% and APAs in 22.9% of boxers, whereas no evidence of APAs or AHAs was found in control subjects (Tanriverdi et al., 2010a).

It is well established that TBI results in the acute induction of both neural and systemic inflammatory responses and consequent anti-inflammatory counter-responses (Lu et al., 2009; Ziebell and Morganti-Kossmann, 2010). In addition, animal studies provide evidence of the development of a chronic inflammatory state after TBI. Three months after moderate focal brain injury in rats, persistence of boxers, whereas no evidence of APAs or AHAs was found in control subjects (Tanriverdi et al., 2010a).

In general, the frequency of occurrence of pituitary hormone abnormalities has not been found to be related to the severity of the trauma (Lieberman et al., 2001; Agha et al., 2004a; Aimaretti et al., 2004, 2005; Bondanelli et al., 2004; Schneider et al., 2006; Park et al., 2011; Kokshoorn et al., 2011), although there have been reports of a positive relationship (Kelly et al., 2000; Klose et al., 2007). Of the traumatic brain injuries sustained by \( \sim 1.7 \) million Americans annually (Faull et al., 2010), 75% are considered mild TBI (mTBI; National Center for Injury Prevention and Control, 2003).

Mild TBI is defined by the American Congress of Rehabilitation Medicine (ACRM) as a head trauma resulting in any one of the following: loss of consciousness (LOC) for 30 min or less, alteration of mental state for up to 24 h (being dazed, confused, disoriented, etc.), or loss of memory for events immediately before or after the trauma (American Congress of Rehabilitation Medicine, 1993). The terms mTBI and concussion are frequently used interchangeably (National Center for Injury Prevention and Control, 2003; Department of Veterans Affairs/Department of Defense, 2009).

Mild TBI-related chronic pituitary dysfunction has been reported in boxers and kick boxers subjected to repetitive head injuries. In a preliminary study, 45% of professional boxers were found with apparent growth hormone deficiency (GHD), but no other pituitary hormone deficiencies were observed (Kelestirm et al., 2004). In a larger study of active and retired boxers 18% had pituitary hormone deficiencies in one or more axes (Tanriverdi et al., 2008c). An investigation of pituitary dysfunction in amateur kick boxers revealed GH and/or adrenocorticotropic (ACTH) deficiencies in 27.3% of the athletes (Tanriverdi et al., 2007).

In 2010, the injuries in 80% of over 30,000 U.S. military service members medically diagnosed with TBI were classified as mTBI (Military Health System, 2011), and mTBI sustained from explosive blasts is one of the most common combat injuries resulting from deployment to Iraq or Afghanistan. About 10–20% of returnees report having experienced at least one blast concussion (Tanielian et al., 2008; Terrio et al., 2009).

The extensive documentation of the high prevalence of hypopituitarism after TBI from all causes and the absence of any published studies of the frequency of PTHP after blast-related mTBI provided the rationale for this investigation of hypopituitarism in U.S. Veterans of combat in Iraq and/or Afghanistan who have experienced at least one blast concussion.

**MATERIALS AND METHODS**

**PARTICIPANTS AND SAMPLE ACQUISITION**

The VA Puget Sound Health Care System (VAPSHCS) Institutional Review Board and the U.S. Army Medical Research and Materiel Command (USAMRMC) Office of Research Protections (ORP) Human Research Protection Office (HRPO) approved the subject protocol with a waiver of informed consent. All plasma and serum samples, demographic, and blast exposure data were obtained from an established biorepository entitled “Alzheimer’s Disease Research Center (ADRC) Participant Registry and Sample Repository.” All subjects whose samples were utilized had consented to have their samples and data used in future research of this type.

The mTBI Veteran participants (T group) whose samples were obtained from the repository were a convenience sample of 26 male Veterans recruited from VAPSHCS, all of whom had documented hazardous duty experience in Iraq and/or Afghanistan with the U.S. Armed Forces and had reported experiencing at least one blast exposure in the war zone that resulted in acute mTBI as defined by ACRM criteria (American Congress of Rehabilitation Medicine, 1993) except that Glasgow Coma Scale scores were not obtained in the combat setting. Samples from the repository were also collected from seven male Veterans who had been deployed to Iraq and/or Afghanistan but who had not been exposed to blast and had no history of TBI. These individuals made up the deployment control (DC) group.

Additional samples from the repository which were used to establish normal hormonal reference ranges had been collected from 59 cognitively normal male community volunteers recruited from the ADRC, all of whom were medically healthy and had Mini-Mental State Examination scores of 29.4 \( \pm 1.0 \) (mean \( \pm \) SEM; range 27–30); Clinical Dementia Rating scores of zero; no evidence or history of cognitive or functional decline; and no history of blast exposure or head injury. These samples were used only for the establishment of normative hormone concentrations with our assay methods. Resting blood samples had been collected from all participants between 9:00 and 10:00 a.m., at least 30 min after the insertion of an intravenous catheter in an antecubital vein.

None of the Veteran or community control participants had a history of blast exposure, head injury with LOC greater than 30 min; penetrating head wound; seizure disorder; insulin-dependent diabetes; current or past DSM-IV diagnoses of schizophrenia, other psychotic disorders, bipolar disorder, or dementia; or a DSM-IV diagnosis of alcohol or other substance abuse or dependence within the previous 3 months. Participants using medications likely to affect brain function, such as opioids,
benzodiazepines, or anti-depressants, were asked not to take those medications for 24 h prior to blood sampling.

BLAST EXPOSURE ASSESSMENT
Blast exposure and mTBI histories had been obtained from mTBI Veteran participants during a clinical interview in which specific inquiries were made regarding total number of blast exposures accompanied by acute symptoms of TBI and/or LOC in Iraq and/or Afghanistan and lifetime history of non-blast exposure head injuries accompanied by acute symptoms of TBI and/or LOC (e.g., sports or motor vehicle accident-related concussion).

NEUROLOGICAL ASSESSMENT
All subjects underwent a full neurological examination, including the Unified Parkinson’s Disease Rating Scale (UPDRS) motor section (Martínez-Martín et al., 1994). Olfactory function was assessed using the Brief Smell Identification Test (B-SIT; Doty et al., 1996).

HORMONE MEASUREMENT
Blood samples for the measurement of plasma hormone concentrations were collected between 9:00 and 10:00 a.m. in chilled tubes containing ethylenediaminetetraacetic acid (EDTA), placed on ice, and centrifuged at 4°C prior to removal of the plasma fraction. Blood samples for measurement of serum hormones were collected in serum-separator tubes, allowed to clot at room temperature for 10 min, and centrifuged to isolate serum. Serum and plasma samples were aliquoted and stored at −70°C. Twelve pituitary or target-organ hormones were measured in these samples. The type, source, and performance characteristics of the assay kits used for the measurement of hormone concentrations in serum and plasma are shown in Table 1. ACTH, cortisol, thyroid-stimulating hormone (TSH), oxytocin, and vasopressin concentrations were determined in plasma; free thyroxine, luteinizing hormone (LH), follicle-stimulating hormone (FSH), total testosterone, insulin-like growth factor-I (IGF-I), growth hormone, and prolactin were measured in serum.

CLINICAL LAB DATA
Measurements of plasma and urine osmolality were not available but urine specific gravity was measured and used as a criterion to determine functional vasopressin insufficiency.

STATISTICAL ANALYSIS AND CRITERIA FOR PITUITARY DEFICIENCIES
The criteria for PTHP, derived using hormone measurements from the 59 community control participants are shown in Table 2. For each hormone, age-adjusted percentiles based on the lognormal distribution from community control participants were estimated and dysfunction in each of seven hormonal axes was defined (R Development Core Team, 2011). Hypopituitarism was defined as a dysfunction in at least one of these seven axes. These criteria were

Table 1 | Sources and characteristics of hormone assay kits.

| Assay | Kit name | Manufacturer | Location |
|-------|----------|--------------|----------|
| ACTH  | ACTH Immunoradiometric (IRMA) Assay | Scantibodies Laboratory | Santee, CA, USA |
| Cortisol | GammaCoat™Cortisol 125I RIA | Diasorin | Stillwater, MN, USA |
| FSH | DELPHIA hFSH | Perkin Elmer | Walham, MA, USA |
| GH | hGH-ELISA, Ultra-Sensitive | DSL | Webster, TX, USA |
| IGF-I | IGF-I RIA | IBL America | Minneapolis, MN, USA |
| LH | ImmuChem™Coated Tube LH 125I RIA | MP Biomedicals | Costa Mesa, CA, USA |
| Oxytocin | Oxytocin EIA Kit – Extraction-free | Peninsula Labs/Bachem | San Carlos, CA, USA |
| Prolactin | ImmuChem™Coated Tube Prolactin 125I IRMA | MP Biomedicals | Costa Mesa, CA, USA |
| Testosterone | Total Testosterone | Siemens Diagnostics | Los Angeles, CA, USA |
| Thyroxine | Free Thyroxine (FT4) Microplate EIA | MP Biomedicals | Costa Mesa, CA, USA |
| TSH | ImmuChem™Coated Tube TSH 125I IRMA | MP Biomedicals | Costa Mesa, CA, USA |
| Vasopressin | Vasopressin Direct RIA | ALPCO | Salem, NH, USA |

| Hormones | Assay type | Sample type | Sample size | Sample size | Assay range | Sensitivity | Intra-assay CV | Inter-assay CV |
|-----------|------------|-------------|-------------|-------------|-------------|-------------|---------------|---------------|
| ACTH      | IRMA       | Plasma      | 100 Tubes   | 200 μl      | 9–1693 pg/ml | <1.0 pg/ml  | 4.05          | 6.66          |
| Cortisol  | RIA        | Plasma      | 100 Tubes   | 10 μl       | 1–60 μg/dl  | 0.21 μg/dl | 7.03          | 9.20          |
| FSH       | Fluoroimmunoassay | Serum | 96 Wells   | 25 μl       | 0.98–256 U/l | 0.05 U/l   | 2.33          | 1.87          |
| GH        | EIA        | Serum       | 96 Wells    | 100 μl      | 4.5–500 pg/ml | 0.66 pg/ml | 6.00          | 5.40          |
| IGF-I     | RIA        | Serum       | 100 Tubes   | 100 μl      | 0.16–10.0 ng/ml | 0.02 ng/ml | 2.97          | 10.30         |
| LH        | RIA        | Serum       | 100 Tubes   | 100 μl      | 2.5–2000 mU/ml | 1.5 mU/ml | 5.90          | 7.90          |
| Oxytocin  | EIA        | Plasma      | 96 Wells    | 50 μl       | 0–630 pg/ml | 6.5 pg/ml  | 9.36          | 13.67         |
| Prolactin | IRMA       | Serum       | 100 Tubes   | 25 μl       | 2.5–100 ng/ml | 2.5 ng/ml  | 5.13          | 8.08          |
| Testosterone | Solid-phase RIA | Serum | 100 Tubes | 50 μl       | 20–1600 ng/dl | 4 ng/dl   | 3.40          | 7.90          |
| Thyroxine | EIA        | Plasma      | 96 Wells    | 50 μl       | 0.45–76 ng/dl | 0.05 ng/dl | 6.83          | 6.47          |
| TSH       | IRMA       | Plasma      | 100 Tubes   | 200 μl      | 0.2–50 μU/ml | 0.04 μU/ml | 4.10          | 5.23          |
| Vasopressin | RIA       | Plasma      | 100 Tubes   | 400 μl      | 1.25–80 pg/ml | 0.1 pg/ml  | 6.00          | 9.90          |
Table 2 | Screening criteria for identifying abnormal circulating hormone levels.

| Axis                        | Criteria using lognormal distribution of community control reference sample |
|-----------------------------|-------------------------------------------------------------------------------|
| Adrenal insufficiency       | Cortisol < 10th percentile (6.7 μg/dl), and ACTH < 10th percentile (18 pg/ml) |
| Thyroid deficiency          | Free T-4 < 5th percentile (0.87 ng/dl), and TSH < 50th percentile (2.39 μIU/ml) |
| Hypogonadism                | Total testosterone < 5th percentile (330 ng/dl) and either LH or FSH < 10th percentile (2.3 mIU/ml, 1.3 U/l, respectively) OR total testosterone < 5th percentile and prolactin > 95th percentile (32 ng/ml) |
| Vasopressin abnormality     | Vasopressin > 95th percentile (9.46 pg/ml) OR vasopressin < 5th percentile (0.27 pg/ml) and urine specific gravity < 1.003 |
| Prolactin abnormality       | Prolactin > 95th percentile (32.0 ng/ml) OR prolactin < 5th percentile (6.7 ng/ml) |
| GH deficiency               | IGF-1 < age-adjusted 10th percentile (SDS < −1.4) |
| Oxytocin deficiency         | Oxytocin < 5th percentile (22.7 pg/ml) |
| Hypopituitarism             | Abnormalities in at least one of these 7 axes |

modeled after those used in published studies of hypopituitarism after TBI from all causes.

RESULTS

PLASMA/SERUM HORMONE SCREENING EVALUATIONS

Eleven of 26 mTBI subjects (T), or 42%, were found to have abnormal hormone values in at least one axis. As reported in earlier studies of PTHP, deficiencies in the growth hormone-IGF-I and pituitary–gonadal axes were observed most frequently (Bavisetty et al., 2008; Dusick et al., 2008; Schneider et al., 2008; Englander et al., 2010; Kokshoorn et al., 2010; Krahulik et al., 2010; Park et al., 2010; Pavlovic et al., 2010; van der Eerden et al., 2010).

Markedly low IGF-I levels are strong indicators of adult GHD (Juul et al., 1997; Hartman et al., 2002; Hadjadj et al., 2007; Ho, 2007; Prodam et al., 2008; Tanriverdi et al., 2011; Zgaljardic et al., 2011). The red line in Figure 1 represents the cutoff level used to define our criterion for subnormal IGF-I levels indicative of probable GHD. The cutoff level was defined to be an IGF-I concentration below the age-adjusted 10th percentile level [equivalent to an SD score (SDS) below −1.4] of the community control reference sample (Figure 1; Table 2). Five Veteran participants with mTBI (T-4, T-8, T-16, T-25, and T-28) were found to have serum IGF-I concentrations below this cutoff line. None of the Veteran participants in the DC group were found to have subnormal age-adjusted IGF-I levels (Figure 1).

Three participants with mTBI (T-4, T-13, and T-28) were found with abnormal hormonal profiles indicating probable hypogonadism. The criteria were a total testosterone concentration less than the 5th percentile of the reference sample together with an LH or FSH level below the 10th percentile reference level (Figure 2; Table 2). T-4 and T-28 also had the lowest IGF-I levels among the participants (T-4: 126 ng/ml, SDS = −2.325; T-28: 86 ng/ml, SDS = −2.989). Elevated prolactin levels in conjunction with low testosterone are also indicative of hypogonadism. A serum prolactin concentration markedly higher than the 95th percentile of the reference sample was found in serum from participant T-4. A subnormal prolactin concentration (<5th percentile), also associated with sexual dysfunction, was measured in serum from T-13.

None of the Veterans in the DC group were found to have hormone levels indicative of hypogonadism. One participant in the DC group was found with a total testosterone concentration below the 5th percentile reference standard and another had an LH concentration below the 10th LH percentile, but neither exhibited the combined gonadotropin and testosterone deficiencies consistent with hypogonadism.

None of the Veteran participants in either the T or DC group exhibited abnormalities in the hypothalamic-pituitary–adrenocortical or hypothalamic-pituitary–thryroid axis (Table 3). The corticotrophs and thyrotrophs are located in the protected median wedge of the anterior pituitary and are anatomically less vulnerable to injury than gonadotropin- and GH-secreting cells. This differential anatomical vulnerability correlates well with the frequency of chronic hormonal abnormalities observed after TBI (Bavisetty et al., 2008; Blair, 2010; Krahulik et al., 2010).

In addition to the findings of anterior pituitary hormone abnormalities in six Veteran participants with mTBI, eight instances of anomalous posterior pituitary hormone levels were
found in six Veterans in the mTBI group, one of whom, T-28, also had evidence of presumptive GHD and hypogonadism. The plasma oxytocin concentration was unmeasurably low in this individual (Table 3). None of the Veterans in the DC group were found to have abnormal posterior pituitary hormone values.

Three additional participants from the mTBI group (T-10, T-14, and T-22) also were found to have circulating oxytocin concentrations below the reference sample’s 5th percentile level. Two of these participants, T-10 and T-22, also met our criteria for arginine vasopressin (AVP) deficiency: plasma vasopressin concentration below the 5th percentile of the reference level in combination with urine specific gravity less than 1.003. In addition, plasma vasopressin concentrations in participants T-2 and T-12 were abnormally elevated above the 95th percentile of the reference group.

**DEMOGRAPHICS, DEPLOYMENT HISTORY, BLAST EXPOSURE, AND MEDICATION USE**

After completion of hormone measurement and identification of Veterans with apparent hypopituitarism, participants in the T group were divided into two subgroups, based on the presence or absence of hormone abnormalities, with demographic, deployment history, blast exposure, and medication use data with each other and with the DC group. The three groups of Veteran participants did not differ in age, education, or body mass index at the time of enrollment, and the two mTBI subgroups did not differ significantly from one another on any of the measures of deployment history or blast exposure (Table 4).

**CONCURRENT MEDICATIONS**

Medications with potential neuroendocrine effects taken by mTBI subjects found to have indications of hypopituitarism were opiates (2/11), prazosin (2/11), selective serotonin reuptake inhibitors (SSRIs; 4/11), serotonin and norepinephrine reuptake inhibitors (SNRIs; 2/11), hypnotics (2/11), atypical antipsychotics (1/11), calcium channel blockers for migraine (1/11), benzodiazepines (1/11), and mirtazapine (1/11). Five subjects in this group were not taking any neuroactive medications. Medications with potential neuroendocrine effects taken by mTBI subjects found to have hormone levels within normal ranges were opiates (1/15), prazosin (4/15), SSRIs (3/15), SNRIs (2/15), mirtazapine (1/15), trazodone (1/15), benzodiazepines (1/15), and disulfiram (1/15). Nine subjects in this group were not taking any neuroactive medications.

**DISCUSSION**

Our findings in this preliminary study support the hypothesis that blast mTBI carries a risk of PTHP similar to that found in several previous studies of hypopituitarism in the general population after TBI from all causes. We have found that blood samples from 11 of 26, or 42% of Veterans of combat in Iraq or Afghanistan had abnormal circulating hormone concentrations consistent with PTHP. Five participants with blast mTBI exhibited evidence of anterior pituitary dysfunction, five additional subjects had anomalous posterior pituitary hormone levels, and the eleventh was found to have both anterior and posterior pituitary hormonal abnormalities. In contrast, none of the seven Veterans of deployment to Iraq and/or Afghanistan in the study who did not experience blast trauma – the DC group – were found to have evidence of pituitary dysfunction. As Kokshoorn et al. (2010) pointed out in their review of 14 investigations of PTHP conducted between 2000 and 2009, these early studies used a broad variety of screening criteria that were sometimes described in general terms rather than with specifically

**FIGURE 2 | Serum LH (left) and testosterone (right) in the deployment control (DC, triangles) and mTBI (T, circles) groups.** Screening criteria for hypogonadism: LH (or FSH) levels below the 10th percentile of the control range together with testosterone below the 5th percentile (red lines). Green circles mark data from 3 T subjects falling below both cutoffs. No DC subjects met both criteria.
It should be cautioned that the determinations of basal hormone concentrations, such as those made in this study, are meant to be screening tools, and are not intended to be, nor should they be interpreted to be, diagnostic in the absence of clinical assessment. Measurement of basal circulating hormone concentrations is generally considered an appropriate screening tool for provisional identification of deficient thyroid function, hypogonadism, and prolactin and oxytocin deficiencies. Diagnosis of significant abnormalities of vasopressin secretion normally require confirmation by measures of plasma and/or urine osmolality, urine specific gravity (UGS), and/or the administration of a water deprivation test. Although provocative testing is generally considered necessary for diagnosis of SAI and GHD, measurement of basal cortisol and IGF-I concentrations remain valuable screening tools to identify individuals most likely to benefit from additional testing and clinical referral. Evaluation of clinical signs and symptoms are essential for definitive diagnoses in all cases.

Previous studies have found GHD to be the most prevalent chronic endocrine consequence of TBI, and it carries with it a potentially large range of symptoms. Provocative testing is considered to be a requisite for the reliable diagnosis of GHD because serum GH concentrations measured in the morning are not valid indicators of daily secretion or somatotroph function. GH secretion occurs predominantly during sleep, and morning levels are generally very low but punctuated unpredictably by short secretory bursts (Van Cauter et al., 1992). However, GH stimulates hepatic production of IGF-I that provides a useful index of somatotroph function. IGF-I concentrations have low diagnostic sensitivity for identifying GHD but are highly specific. The presence of normal IGF-I values cannot be used to exclude GDH because it is often diagnosed in individuals with normal or even elevated IGF-I levels. However, markedly low age-adjusted levels of IGF-I are strongly indicative of GHD (Juul et al., 1997; Hadjadj et al., 2007; Ho, 2007; Prodam et al., 2008; Tanriverdi et al., 2011; Zgaljardic et al., 2011).

Table 3 | Plasma or serum hormone concentration for each participant.

| Subject | Age | #BE | ACTH | Cort | LH | FSH | tTest | PRL | TSH | fT-4 | IGF-I | GH | AVP | OT |
|---------|-----|-----|------|------|----|-----|-------|-----|-----|------|-------|-----|------|----|
| T-1     | 24  | 11  | 24   | 6.6  | 2.58 | 0.46 | 473   | 12.5 | 1.70 | 1.29 | 190   | 58  | 3.4  | 64 |
| T-2     | 26  | 6   | 20   | 11.9 | 2.03 | –    | 669   | 9.6  | 1.92 | 1.67 | 185   | 71  | 12.3 | 181|
| T-3     | 27  | 10  | 22   | 10.2 | 5.15 | 2.05 | 557   | 13.0 | 1.59 | 1.59 | 164   | 50  | 4.0  | 166|
| T-4     | 38  | 102 | 19   | 7.5  | 2.03 | 2.06 | 252   | 54.9 | 1.17 | 1.22 | 110   | 11  | 8.0  | 88 |
| T-5     | 46  | 5   | 29   | 12.1 | 1.95 | 1.41 | 402   | 11.1 | 1.08 | 1.09 | 143   | 152 | 0.7  | 61 |
| T-6     | 26  | 20  | 28   | 14.1 | 3.34 | 1.63 | 559   | 12.0 | 2.26 | 1.29 | 172   | 223 | 0.0  | 39 |
| T-7     | 25  | 21  | 35   | 11.3 | 2.72 | 4.02 | 401   | 11.9 | 1.13 | 1.55 | 141   | 294 | 0.2  | 44 |
| T-8     | 24  | 102 | 39   | 12.0 | 4.81 | 3.28 | 730   | 14.4 | 1.16 | 1.05 | 251   | 1288| 8.0  | 52 |
| T-9     | 26  | 20  | 30   | 9.6  | 1.97 | 2.43 | 520   | 12.3 | 2.41 | 1.39 | 230   | 68  | 0.2  | 22 |
| T-10    | 28  | 9   | 31   | 10.2 | 1.64 | 2.38 | 382   | 9.2  | 1.30 | 1.22 | 182   | 0   | 1.1  | 36 |
| T-11    | 27  | 15  | 27   | 9.9  | 7.27 | 5.70 | 715   | 13.0 | 1.57 | 1.38 | 198   | 60  | 12.0 | 55 |
| T-12    | 31  | 7   | 40   | 1.92 | 1.18 | 253  | 6.3  | 2.24 | 1.14 | 187   | 0   | 6.4  | 50 |
| T-13    | 30  | 6   | 36   | 8.8  | 2.66 | 2.51 | 390   | 12.0 | 3.75 | 1.13 | 151   | 310 | 0.5  | 19 |
| T-14    | 25  | 11  | 16   | 7.4  | 2.27 | 3.56 | 377   | 10.1 | 1.40 | 1.14 | 168   | 42  | 4.4  | 85 |
| T-15    | 28  | 66  | 25   | 11.7 | 2.64 | 4.01 | 380   | 21.5 | 0.65 | 1.52 | 126   | 0   | 0.9  | 190|
| T-16    | 25  | 18  | 26   | 11.2 | 9.32 | 3.00 | 465   | 17.1 | 1.26 | 1.25 | 197   | 0   | 4.8  | 151|
| T-17    | 24  | 7   | 30   | 14.3 | 3.65 | 2.21 | 556   | 12.2 | 1.27 | 1.30 | 179   | 60  | 4.3  | 32 |
| T-18    | 30  | 1   | 32   | 18.6 | 5.44 | 3.01 | 457   | 11.7 | 1.55 | 1.13 | 190   | 66  | 0.0  | 46 |
| T-19    | 28  | 3   | 17   | 25.0 | 6.65 | 3.91 | 309   | 10.4 | 0.79 | 1.31 | 210   | 1696| 7.9  | 519|
| T-20    | 24  | 4   | 21   | 17.6 | 4.00 | 4.48 | 588   | 12.8 | 1.09 | 1.24 | 227   | 110 | 0.0  | 21 |
| T-21    | 29  | 52  | 11   | 8.7  | 2.52 | 2.55 | 554   | 8.9  | 1.34 | 1.35 | 166   | 95  | 4.0  | 199|
| T-22    | 25  | 12  | 8    | 7.7  | 2.24 | 4.34 | 463   | 72   | 1.42 | 1.23 | 146   | 813 | 2.1  | 25 |
| T-23    | 26  | 11  | 9    | 6.8  | 5.94 | 1.14 | 488   | 8.4  | 0.81 | 1.18 | 185   | 42  | 0.9  | 457|
| T-24    | 43  | 5   | 29   | 7.5  | 2.52 | 3.89 | 263   | 7.1  | 0.60 | 1.04 | 126   | 13  | 2.6  | 50 |
| T-25    | 41  | 12  | 15   | 11.1 | 2.11 | 2.14 | 264   | 15.3 | 1.22 | 1.11 | 86   | 375 | 8.4  | 0  |

Shaded values indicate hormone axis abnormalities as defined in Table 2. #BE, number of self-reported blast exposures meeting ACRM criteria for mTBI during military career; ACTH, adrenocorticotropic; Cort, cortisol; LH, luteinizing hormone; FSH, follicle-stimulating hormone; tTest, total testosterone; PRL, prolactin; TSH, thyroid-stimulating hormone; fT-4, free thyroxine; IGF-I, insulin-like growth factor-I; GH, growth hormone; AVP, vasopressin; OT, oxytocin.
Studies using receiver operating characteristic (ROC) analysis to compare the diagnostic accuracy of IGF-I relative to diagnosis of GHD based on provocative testing of GH secretion have reported a diagnostic specificity of 100% with IGF-I SDS cutoffs of −1.3 (Corneli et al., 2007) or −1.7 (Maghnie et al., 2005).

The individuals classified here as having a high probability of GHD all had values less than −1.4 SDS below the age-adjusted means of the reference sample. The high specificity of IGF-I measurements at this level assures a very low likelihood of false positives in diagnosing GHD. However, in light of the low sensitivity of IGF-I concentrations in predicting GHD, it is probable that some Veteran participants with normal IGF-I levels may be growth hormone deficient.

The long-term sequelae of GHD in adults for health, quality of life (QoL), and morbidity are multifaceted and complex. Low GH secretion has been associated with behavioral symptoms and deficits in several cognitive domains (Popovic et al., 2004; Fall et al., 2006; Pavlovic et al., 2010). GHD also has significant deleterious effects on body composition and cardiovascular function. Adult GHD is associated with lipidemia, reduced lean body mass, and increased adiposity. Even partial GHD in adult patients is associated with adverse lipid profiles and early atherosclerosis (Colao et al., 2006a, b; Colao, 2008). Impairment in QoL is also a prominent feature of adult GHD, especially in the areas of energy and vitality (McGauhey, 1989; Kelly et al., 2006; Bushnik et al., 2007; Svensson et al., 2007; Bavisetty et al., 2008). Adult GHD is also associated with reductions in muscle volume and strength, decreased physical mobility, fatigue, sleep impairment, social isolation, depression, lowered metabolic rate, low sexual drive, and reduced aerobic capacity (Rosén et al., 1994; Mossberg et al., 2008).

Many of the symptoms of GHD can be successfully ameliorated or reversed by growth hormone replacement therapy. Five retrospective studies have shown that the risk of premature death from cardiovascular disease is elevated in patients with GHD (Svensson et al., 2004a). The increased risk factors such as adverse lipid profiles, increased blood pressure, abnormal body composition, increased body weight, increased coagulability, and increased markers of inflammation have all been shown to improve with GH replacement (Svensson et al., 2004a, 2007; Götherström et al., 2007a; Verhelst and Abs, 2009). GH replacement has been found to be effective in reversing cognitive impairments in several domains including simple motor speed, information processing speed, episodic memory, mental flexibility, verbal memory, and executive functioning in patients after TBI (High et al., 2010; Reimunde et al., 2011). GH replacement also normalizes muscle strength and increases bone mineral density (Götherström et al., 2007b, 2009), improves psychiatric functioning by ameliorating depression, intensity of interpersonal sensitivity, hostility, paranoid ideation, and anxiety (Maric et al., 2010), and improves QoL (Svensson et al., 2004b, 2007; Kreitschmann-Andermahr et al., 2008).

Table 4 | Mean ± SEM and (range) for demographic, deployment, and blast exposure data for each group of participants.

| DC (n = 7) | mTBI without PTHP (n = 15) | mTBI with PTHP (n = 11) |
|------------|-----------------------------|--------------------------|
| A. DEMOGRAPHICS | | |
| Age (years) | 31.1 ± 3.3 (23–47) | 29.7 ± 1.8 (24–46) | 28.8 ± 1.5 (25–41) |
| Education (years) | 14.0 ± 0.7 (12–17) | 13.3 ± 0.4 (11–16) | 13.6 ± 0.5 (12–16) |
| Marital status | 3/7 Married, 4/7 single | 7/15 Married, 4/15 single, 2/15 divorced, 2/15 unknown | 7/11 Married, 1/1 single, 1/11 separated, 2/11 unknown |
| Body mass index (BMI) | 28.5 ± 2.1 (n = 5)† | 27.9 ± 1.3 (n = 14)† | 29.0 ± 1.3 (n = 10)† |
| B. DEPLOYMENT HISTORY | | |
| Number of deployments | 1.7 ± 0.4 (1–3) | 1.9 ± 0.2 (1–4) | 2.1 ± 0.3 (1–3) |
| Time between first and second deployments (months) | 14.3 ± 7.0 (3.5–27.5) (n = 3) | 15.9 ± 3.1 (4.0–39.5) (n = 10) | 15.4 ± 2.4 (7.5–30.0) (n = 8) |
| Time between second and third deployments (months) | 6.0 ± 1.0 (5.0–70) (n = 2) | 8.0 ± 2.0 (6–12) (n = 3) | 76 ± 2.0 (3.0–12.5) (n = 4) |
| Time between third and forth deployments (months) | 8.0 ± 0.0 (n = 1) | | |
| Total deployment time (months) | 13.0 ± 1.8 (7–21) | 18.7 ± 2.2 (7–37) | 18.2 ± 1.7 (11–27) |
| C. BLAST EXPOSURE | | |
| Deployment blast exposures meeting ACRM criteria for mTBI | 0 | 11.1 ± 3.3 (1–52) | 14.6 ± 5.4 (4–66) |
| Blast exposures meeting ACRM criteria during military career | 0.3 ± 0.3 (0–2) | 24.5 ± 8.7 (1–102) | 16.7 ± 5.2 (4–66) |
| Blast exposures with LOC | 0 | 1.3 ± 0.3 (0–4) | 0.6 ± 0.2 (0–2) |
| Lifetime events with LOC | 0.1 ± 0.1 (0–1) | 3.1 ± 0.7 (0–11) | 1.3 ± 0.4 (0–3) |
| Time since last blast exposure (months) | 45.2 ± 4.2 (14–66) | 47.4 ± 4.3 (20–67) | |

The Veterans with blast mTBI (T group) were divided into two subgroups based upon the presence or absence of abnormal hormonal profiles suggesting PTHP. BMIs were not obtained for all participants.
Three of the Veteran participants in the T group met our criteria for hypogonadism: a total testosterone concentration less than the 5th percentile of the reference sample together with an LH or FSH level below the 10th percentile reference level. In our very small sample, the occurrence of hypogonadism was found to be next highest in frequency to that of GH and GH-secretoma, on the other hand, are anatomically more vulnerable to damage because of their location in the pituitary's exposed lateral wings and their primary dependence on vascular input from the portal system alone. Gonadotrophs are distributed throughout the anterior pituitary, and the cells in the lateral wings are relatively vulnerable.

In addition to the six participants with hormonal levels consistent with hypogonadism and/or GH, six of the Veterans with mTBI (including one with anterior pituitary hormonal abnormalities) exhibited abnormal plasma vasopressin and/or oxytocin concentrations. Oxytocin concentrations below the 5th percentile value of the community control group were observed in four of the mTBI participants. Two of the four also exhibited indications of vasopressin deficiency as defined by vasopressin levels below the 5th percentile of the community control group together with urine specific gravity less than 1.003. The occurrence of deficits of both vasopressin and oxytocin in two participants suggests the possibility of disruption of the pituitary stalk or hypothalamic damage in these individuals. In addition, elevated plasma vasopressin concentrations above the reference 95th percentile were measured in two subjects.

In several studies, elevated cerebrospinal fluid (CSF) or peripheral vasopressin concentrations have been associated with PTSD, depression, schizophrenia, and other psychiatric disorders, but a causal relationship has not been established (Purba et al., 1996; van Londen et al., 1997; Coccaro et al., 1998; Merali et al., 2006; de Kloet et al., 2009). In contrast, there is evidence from both animal and human studies for the positive association of oxytocin levels with social bonding, attenuation of stress responses in socially relevant challenges, mediation of social support, and positive social interactions (Heinrichs et al., 2009; Campbell, 2010). Our finding of a high frequency of abnormal peripheral hormone levels after blast mTBI in this preliminary study is consistent with the investigations cited above, in which the prevalence of pituitary dysfunction fell in the 30–60% range in 11 of 22 reports. However, in general, those studies focused exclusively on anterior pituitary dysfunction. Although few studies have investigated the prevalence of chronic posterior pituitary hormonal abnormalities after TBI, most (Agha et al., 2004b, 2005; Krahulik et al., 2010), but not all (Bondanelli et al., 2004), found significant evidence of damage in that lobe as well. In this study we found significant anterior pituitary dysfunction in 23.1% of Veterans with mTBI and abnormal posterior pituitary hormone levels in 23.1% of this group as well. In contrast, the prevalence of hypopituitarism in the general adult population ranges between 290 and 455 cases per million (Regal et al., 2001).

The only other ongoing study of hypopituitarism after blast mTBI of which we are aware recently reported preliminary results based on two retrospective chart reviews. Of 147 Marines with blast-related mTBI screened approximately 1 year or more after injury, 25% were found to have abnormal levels of one or more anterior pituitary hormones (Stokes and Gallagher, 2011). The Veteran groups in this study are highly similar in demographic characteristics and share the common experience of
To our knowledge, this is the first published study of basal hormonal evaluation of combat Veterans with blast mTBI, and our preliminary findings of PTHP in 42% of study participants supports the advisability of routine screening for hypopituitarism after brain injury in both military and civilian populations. Routine provocative testing for sAL and for GHD, the most frequently observed chronic endocrine disorder resulting from TBI, is neither economically nor practically viable. However, measurement of basal concentrations of a panel of pituitary and target-organ hormones similar to those performed here may provide valuable preliminary indications of those individuals most likely to benefit from further clinical evaluation. Similar preliminary screening for pituitary dysfunction after blast concussion shows promise for appropriately directing diagnostic and therapeutic decisions that otherwise may remain unconsidered and for markedly facilitating recovery and rehabilitation.

ACKNOWLEDGMENTS

We gratefully acknowledge the skillful assistance of Carl Sikkema for laboratory work, Carol Xiang and Daniel Morelli for data management, Molly Chinn and Alka Goyal for administrative guidance, Natalia Czajkiewicz for graphic design, and Dr. Leanne Madre for significant contributions in shaping the manuscript into its final form. The work was supported by U.S. Department of Defense Congressionally Directed Medical Research Program Concept Award PT090753; the Gericatric Research, Education and Clinical Center, and the Research and Development Service of the VA Puget Sound Health Care System; the VA Northwest Network Mental Illness Research, Education and Clinical Center; the Seattle Institute for Biomedical and Clinical Research; the University of Washington Alzheimer’s Disease Research Center NIA AG05136; and an anonymous foundation.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 02 December 2011; paper pending published: 27 December 2011; accepted: 17 January 2012; published online: 07 February 2012.

Citation: Wilkinson CW, Pagulayan KF, Petrie EC, Mayer CL, Colasurdo EA, Shofer JB, Hart KL, Hoff D, Tarabochia MA and Peskind ER (2012) High prevalence of chronic pituitary and target-organ hormone abnormalities after blast-related mild traumatic brain injury. Front. Neurol. 3:11. doi: 10.3389/fneur.2012.00011

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