INTRODUCTION

Neurofibromatosis (previously known as von Recklinghausen disease) are distinct clinical entities that represent numerous neurofibromas on the skin. Neurofibromatosis type 1 (NF1; OMIM:162200) is characterized by multiple café-au-lait macules (CALMs) and cutaneous neurofibromas (CNs). It frequently involves the head and neck region and exhibits a variety of symptoms, ranging from simple skin neurofibromas to devastating plexiform neurofibromas (PNs) that cause melting-skin disfiguration, blindness, nerve compression, and airway obstruction (Fig. 1) [1].

This article reviews the fundamental features of NF1 and current knowledge and individual strategies for disease management, particularly in the head and neck region.

HISTORY

NF1 was first described as a systemic disorder by Friedrich von Recklinghausen in 1882 [2]. Early studies focused on its clinical presentation and hereditary nature. This led to the development of a diagnostic criteria consensus in 1987 by the National Institutes of Health (NIH) [3]. Specialized clinical investigations have helped to recognize various neurodevelopmental symptoms other than the tumorigenic presentation of the disease [4].

Recently, the NF Clinical Trials Consortium was established to aid biologic experimental studies and the Response Evaluation in Neurofibromatosis and Schwannomatosis to collect clinical outcomes more uniformly [5-7].

There is an expansion in the knowledge of clinical disorders caused by germline mutations in genes encoding products of the Ras/mitogen-activated protein kinase (MAPK) pathway.
These are considered RASopathies, and now, NF1 is regarded as the first one identified [8].

**EPIDEMIOLOGY**

NF1 is the most common form of neurofibromatosis, with an estimated incidence of 1 in 2,700 and a prevalence of 1 in 4,500. NF1 represents monogenic disorders presenting a pattern of autosomal dominant inheritance, with a strikingly high rate of de novo mutations that were found to be 42%. These de novo mutations are believed to be responsible for the sporadic appearance of NF1 [9]. Penetrance is complete or, at least, nearly so before the age of 5 years, but expressivity differs greatly between affected individuals, even within the same consanguinity. There is no predilection for sex, race, or ethnicity [10,11].

The lifetime risk of malignancy in individuals with NF1 is estimated to be 59.6% [12]. This justifies the lifelong surveillance of patients with NF1 [13].

**GENETICS, MOLECULAR PATHOPHYSIOLOGY, AND GENOTYPE-PHENOTYPE CORRELATIONS**

NF1 occurs as a result of a heterozygous germline mutation in the **NF1** tumor suppressor gene (HGNC:7765). **NF1** is located on the long arm of chromosome 17 (17q11.2) and carries a genetic code for the production of neurofibromin [14,15]. Neurofibromin, a 220-kDa cytoplasmic protein, is an important negative regulator of Ras, a proto-oncogene that plays a key role as a signaling molecule in cell growth and differentiation. Ras mutation is associated with a wide variety of cancer types and has been identified in a series of clinical entities called RASopathies [16].

Neurofibromin is a ubiquitous protein produced by most cells, particularly at high levels in cells of the nervous system (neurons, oligodendrocytes, and non-myelinating Schwann cells). Astrocytes and myelinating Schwann cells do not produce neurofibromin. This finding may explain some of the clinical manifestations, such as nerve tumors and cognitive disabilities [17].

Every affected individual carries a defective **NF1** allele (one of paired arms). The other copy of **NF1** on the other allele is considered responsible for producing functional neurofibromin. Additional loss-of-function mutations in the functioning **NF1** copy may result in a wide variety of symptomatic presentations. These somatic mutations occur during embryonic development, resulting in mosaicism of the **NF1** copies. Mutations in the early stages cause more generalized symptoms resembling non-mosaic NF1 due to germline mutations. Later in development, mutations in more differentiated cells manifest as more localized symptoms, often described as segmental NF1 [18]. This “2-hit” theory was suggested to explain the development of several NF1-related tumors, including malignant peripheral nerve sheath tumors (MPNSTs). A phenotype is a result of the complex interplay between multiple factors. Links between genetic impairments and clinical presentation are still under investigation. More than 3,000 different pathogenic sequence variants have been identified; however, only four genotype-phenotype correlations have been clinically identified [7]. An in-frame deletion of codon 992 (p.Met992del) and a missense mutation in codon 1809 (p.Arg1809Cys) are associated with the absence of CNs or PNs [19-21]. Other missense mutations affecting one of codons 844–848 and some submicroscopic chromosomal deletions, so-called microdeletions, are associated with a more severe phenotype [22,23]. Current knowledge for genotype-phenotype correlations is limited because of the genetic heterogeneity and absence of mutational “hot spots” [7].

At present, NF1 is diagnosed using established diagnostic criteria. Genetic testing is reserved for tricky clinical presentations and reproductive decision-making.

**DIAGNOSIS**

The US NIH Consensus Development Conference proposed clinical diagnostic criteria for NF1. Seven criteria were suggested, of which at least two criteria are required to confirm the diagnosis (Table 1) [3].

However, some clinical manifestations of NF1 are age-depen-
There (i.e., neurofibromas take time to become evident). Therefore, diagnosis is not easy in early childhood particularly for sporadic NF1 cases (because they have no parents with NF1). Approximately 46% of patients fail to meet the diagnostic criteria by the age of 1 year. Close persistent monitoring can reveal the hallmark symptoms of NF1, which are included in diagnostic criteria. In 97% of children suspected of having NF1, the criteria were met by the age of 8 years. Virtually all suspected cases are diagnosed by the age of 20 years. The order of appearance of the skin manifestations was CALMs, axillary freckling, Lisch nodules, and CNs or subcutaneous neurofibromas or PNs. Thus, CALM is often the first symptom that suggests the penetrance of NF1 [24]. CALMs tend to increase in size and number as the patient grows. By adulthood, approximately 95% of patients with NF1 will have light-brown skin hyperpigmentation lesions [25]. Genetic testing is not routinely recommended [7].

Recently, an international consensus group has suggested revised diagnostic criteria. Major changes included new separate diagnostic criteria for mosaic NF1 and new criteria for genetic diagnosis. The group recommended performing genetic analyses for patients with segmental symptoms, families with two or more affected siblings and unaffected parents, and children presenting only pigmented symptoms. Sphenoid wing deformation with an anatomically relevant PN is considered insufficient for sphenoid wing dysplasia [26].

### CLINICAL FEATURES AND MANAGEMENT

#### Cutaneous neurofibromas

CNs are the most common type of skin tumors in NF1. CNs present as well-defined cutaneous lesions that are localized but not encapsulated (Fig. 2) [27]. The prevalence of CN in patients with NF1 reached up to 99% [2]. They are benign tumors composed of atypical Schwann cells but also contain fibroblasts, mast cells, pericytes, and perineural cells [28-33]. CNs arise within the epidermal/dermal layer and increase in size and number after an appearance in late childhood [34]. CNs tend to grow more rapidly during puberty and pregnancy, but discrete evidence from prospective observations is lacking [35,36].

The shape of a CN is classified into the following five categories: nascent/latent, flat, sessile, globular, or pedunculated [37]. Approximately 20% of patients experience pruritus related to CNs [38]. The pathophysiology of pruritus in NF1 is still not clearly elucidated, but some studies suggest that mast cells may play a significant role [39-41].

Diffusely infiltrating PNs should be considered in the differential diagnosis since these PNs arise from deeper tissues and mimic CNs by dermal invasion [42]. Any atypia seen on a smear of a PN specimen has the possibility of early malignant transformation, whereas CN specimens often appear to show atypical cells reflecting reactive or degenerative changes [43]. Although CNs are benign and carry no risk of malignant transformation, they can still be troublesome because hundreds to thousands of CNs can cause significant disfigurement, particularly when they involve the head and neck region, causing emotional and physical discomfort [44].

The primary treatment for CNs is surgery. Excision is effective enough to remove one tumor; however, given that CNs are frequently numerous in number and size, excision is not feasible for the management of entire skin lesions [45,46].

#### Plexiform neurofibromas

PNs are a distinct feature of NF1 that causes severe clinical discomfort. They are histologically benign but can be devastating clinically and aesthetically. The prevalence of PNs among patients with NF1 is up to 50% [47,48]. PNs differ from CNs in that they can involve multiple fascicles.
and nerve branches [49]. They can cause significant discomfort to patients, such as pain, disfiguration, mass effect, and functional impairments of neurovascular structures and even the airway [50]. PNs are also believed to cause increased mortality owing to their risk of malignant transformation to MPNSTs throughout their lifespan [51,52]. The risk of malignant transformation of PNs to MPNSTs is increased 20-fold [53], reaching a 10%–15% lifetime risk.

PNs may be congenital, and their growth has been described as unpredictable [49]. Recent studies have found that the growth of PNs bursts out rapidly during childhood and adolescence, and then the rate diminishes over time to converge to zero growth in young adulthood [54]. Thus, unusual growth of PNs or any accompanying neurologic symptoms due to PNs in adulthood may be a warning sign of existing malignancies; therefore, active evaluation and careful monitoring are required. Magnetic resonance imaging (MRI) is the gold standard for the diagnosis and monitoring of PNs [47,54]. However, treatment decisions are based on clinical presentation. Thus, whether, when, and where to image targeted PNs are done as per the practitioners’ judgments [55].

Diffuse PNs appear to infiltrate tissues adjacent to the involved peripheral nerves; therefore, they are not well demarcated (Fig. 3). In contrast, nodular PNs show well-demarcated margins (Fig. 4) [56].

The only standard treatment for PNs is complete surgical removal, and indications for surgical removal are primarily symptomatic, including pain, severe disfiguration, and loss of neurologic and aesthetic function [13]. However, it is difficult to achieve an ideal result because of the large size, location, infiltration to adjacent tissue, and eventual recurrence of PNs. Furthermore, PNs, large enough to be symptomatic, are frequently accompanied by the laxity of the covering skin, risk of severe bleeding from impaired blood vessels, and poor separation from the nerve fibers [31,57]. The skin area involved with underlying PNs loses the elasticity, which results in the absence of normal viscoelastic properties of the skin, such as mechanical creep and stress relaxation [58].

In surgical cases, an intraoperative nerve stimulator and/or an operative microscope will help clearly distinguish the lesion.

Fig. 3. Diffuse plexiform neurofibromas infiltrate adjacent tissues and involve peripheral cervical nerve branches. They are not well demarcated.

Fig. 4. Nodular plexiform neurofibromas, originating from the trigeminal nerve (zygomaticotemporal nerve of V2), exhibit well-demarcated margins.

Fig. 5. Dissection and preservation of facial nerve branches using an intraoperative nerve stimulator while resecting a plexiform neurofibroma of the cheek.
from the surrounding tissue (Fig. 5). Preoperative angiography and/or percutaneous vascular embolization may be performed to prevent intraoperative bleeding from the surrounding tissue as in the treatment of vascular malformations [59]. However, the tumor microenvironment (TME) frequently contains weak blood vessels with dysplastic walls, which are prone to cause bleeding and difficult hemostasis. Since ectatic and dysplastic veins and capillaries are the main foci of bleeding, arterial embolization frequently has a limited effect.

In the head and neck, rich vascular collateralization and shallow positioning of major vessels make hemostasis even more difficult. The vessels in NF1 seem to lack functioning tunica media in the wall, so bleeding from the stretched vessels tends to be low-pressure bleeding, and electrocautery, in this case, does not work well as there is weak or no contraction of the muscle layer. Therefore, physical ligation of the vessels using sutures or external compression is required to achieve appropriate hemostasis. Surgeons should be aware of the risk of intractable bleeding as the resection plane becomes deeper and closer to the neck. Surgical removal of PNs is often performed as palliative rather than curative care, with the goal of symptomatic relief rather than margin-free extirpation. Therefore, physical ligation of the vessels using sutures or external compression is required to achieve appropriate hemostasis. Surgeons should be aware of the risk of intractable bleeding as the resection plane becomes deeper and closer to the neck. Surgical removal of PNs is often performed as palliative rather than curative care, with the goal of symptomatic relief rather than margin-free extirpation. Since ectatic and dysplastic veins and capillaries are the main foci of bleeding, arterial embolization frequently has a limited effect.

In the head and neck, rich vascular collateralization and shallow positioning of major vessels make hemostasis even more difficult. The vessels in NF1 seem to lack functioning tunica media in the wall, so bleeding from the stretched vessels tends to be low-pressure bleeding, and electrocautery, in this case, does not work well as there is weak or no contraction of the muscle layer. Therefore, physical ligation of the vessels using sutures or external compression is required to achieve appropriate hemostasis. Surgeons should be aware of the risk of intractable bleeding as the resection plane becomes deeper and closer to the neck. Surgical removal of PNs is often performed as palliative rather than curative care, with the goal of symptomatic relief rather than margin-free extirpation. Therefore, physical ligation of the vessels using sutures or external compression is required to achieve appropriate hemostasis. Surgeons should be aware of the risk of intractable bleeding as the resection plane becomes deeper and closer to the neck. Surgical removal of PNs is often performed as palliative rather than curative care, with the goal of symptomatic relief rather than margin-free extirpation. Therefore, physical ligation of the vessels using sutures or external compression is required to achieve appropriate hemostasis. Surgeons should be aware of the risk of intractable bleeding as the resection plane becomes deeper and closer to the neck. Surgical removal of PNs is often performed as palliative rather than curative care, with the goal of symptomatic relief rather than margin-free extirpation.
logic damage and a safe margin is achieved. However, this is often limited because MPNSTs frequently invade adjacent structures and involve vital neurovascular networks [75].

Nonetheless, complete resection and tumor-free margins are important indicators for better prognosis [76], whereas old age, metastatic lesions at the time of diagnosis, and only biopsy without resection were significant and independent predictors of poor outcomes [77].

Radiation therapy is generally avoided in patients with NF1 owing to their predisposition to cancer development; however, it can still be used for limited purposes, including local control, especially in cases of incomplete tumor resection. Radiation appears to delay recurrence without affecting mortality [63]. However, further investigation is needed since the association between radiation therapy and mortality, especially after oncologic resection and reconstruction for scalp lesions, has been reported [78].

The efficacy of chemotherapy remains controversial [79,80]; however, in advanced stages, a single-agent anthracycline or doxorubicin-ifosfamide dual regimen can be used for palliative purposes [81,82]. Recently targeted therapies are reviewed in the following section.

**Molecular Targeted Therapies**

Surgery is the current standard of care for both benign and malignant tumors in patients with NF1. However, because tumor recurrence after surgery is not rare, the need for adjuvant therapy has been proposed [88]. Early trials of general agents, including anti-inflammatory, antifibrotic, and antiangiogenic agents, failed to achieve therapeutic significance in the management of PNs [89-91]. A more recent trial with pegylated interferon showed that it elongates the time to progression. However, the application of interferon therapy is limited owing to side-

dle cranial fossa, herniated temporal lobe, and pulsating exophthalmos. Consequently, the malar and zygomatic arches are displaced downward [84].

Sphenoid wing dysplasia is associated with an anatomically relevant space-occupying lesion, which is mostly a PN that causes local compression. Most patients do not require bone correction [85]. However, if there are symptomatic presentations, removal of the tumor is required. Flaps can be utilized to fill the resultant dead space, which is further accentuated by the dysplastic sphenoid bone [86].

Mandibular dysplasia can also be seen in patients with NF1 and is characterized by a pathognomonic deep sigmoid notch (Fig. 9) as well as a wide inferior alveolar canal, enlarged mandibular foramen, and notching of the inferior border (antegonial notch) [87]. The clinical significance and management strategies for mandibular dysplasia are not clear in the literature.

**Morphologic changes of facial bones**

The most distinctive bony abnormality of NF1 in the field of plastic surgery is sphenoid wing dysplasia, with a prevalence of 11.3% in the probands (Fig. 8) [83]. The sphenoid wing dysplasia typically occurs unilaterally and involves the affected-side orbital walls causing orbital fossa widening, expansion of middle cranial fossa, herniated temporal lobe, and pulsating exophthalmos. Consequently, the malar and zygomatic arches are displaced downward [84].

Sphenoid wing dysplasia is associated with an anatomically relevant space-occupying lesion, which is mostly a PN that causes local compression. Most patients do not require bone correction [85]. However, if there are symptomatic presentations, removal of the tumor is required. Flaps can be utilized to fill the resultant dead space, which is further accentuated by the dysplastic sphenoid bone [86].

Mandibular dysplasia can also be seen in patients with NF1 and is characterized by a pathognomonic deep sigmoid notch (Fig. 9) as well as a wide inferior alveolar canal, enlarged mandibular foramen, and notching of the inferior border (antegonial notch) [87]. The clinical significance and management strategies for mandibular dysplasia are not clear in the literature.

**Fig. 8.** A computed tomography scan shows sphenoid wing dysplasia and orbital wall defects.

**Fig. 9.** A 3-dimensional computed tomography scan shows a deep sigmoid notch, which is pathognomonic, as well as an increased antegonial notch size.
effects [92].

The cumulative knowledge of the molecular pathophysiology of PNs has enabled the development of molecular targeted therapies. The protein product of the \(NF1\) gene, neurofibromin, is a negative regulator of Ras protein, and defective \(NF1\) produces dysfunctional neurofibromins, resulting in constitutive activation of the Ras pathway and, thus, tumorigenesis [93]. Ras activates Raf kinase, MEK, and MAPK (ERK). MAPK regulates the transcription factors involved in the cell cycle [16].

Selumetinib, a selective MEK inhibitor, was the first medication approved by the US Food and Drug Administration for the treatment of intractable, symptomatic PNs in children aged > 2 years. It has been proven to decrease the volume of PNs for more than 1 year even with symptomatic relief [94,95]. Owing to the success of selumetinib, other MEK inhibitors, including trametinib, binimetinib, and mirdametinib, have been investigated, and, so far, the trials have shown remarkable success [96].

Other advancements in molecular targeted therapy of PNs were achieved by studies of cabozantinib, a tyrosine kinase inhibitor that plays a critical role in the regulation of the TME. In a phase II study, 42% of patients achieved partial response, which indicates a > 20% reduction in tumor volume [97].

Most clinical trials investigating targeted therapy regimens for MPNST are still ongoing.

**CONCLUSION**

\(NF1\) is the most common tumor predisposition syndrome inherited in an autosomal dominant (100% penetrance) fashion with a wide variety of expressivity. From the perspective of plastic surgery, the most significant clinical symptoms, including disfiguration, peripheral neurologic symptoms, and skeletal abnormalities, are caused by various tumors originating from the affected nerves. Surgical removal is the standard of care for these tumors. However, the outcome is frequently unsatisfactory, facilitating the search for additional therapeutic adjuvants. Current trials of molecularly targeted therapies are promising.

**NOTES**

Conflict of interest

No potential conflict of interest relevant to this article was reported.

Ethical approval

The patients provided written informed consent for the publication and the use of their images.

**REFERENCES**

1. Latham K, Buchanan EP, Suver D, Gruss JS. Neurofibromatosis of the head and neck: classification and surgical management. Plast Reconstr Surg 2015;135:845-55.
2. Huson SM, Harper PS, Compston DA. Von Recklinghausen neurofibromatosis: a clinical and population study in south-east Wales. Brain 1988;111(Pt 6):1355-81.
3. Neurofibromatosis. Natl Inst Health Consens Dev Consens Statement 1987;6:1-7.
4. CNossen MH, de Goede-Bolder A, van den Broek KM, Waasdorp CM, Oranje AP, Stroink H, et al. A prospective 10 year follow up study of patients with neurofibromatosis type 1. Arch Dis Child 1998;78:408-12.
5. Cimino PJ, Gutmann DH. Neurofibromatosis type 1. Handb Clin Neurol 2018;148:799-811.
6. Packer RJ, Fisher MJ, Cutter G, Cole-Plourde K, Korf BR. Neurofibromatosis clinical trial consortium. J Child Neurol 2018;33:82-91.
7. Bettgowda C, Upadhyaya M, Evans DG, Kim A, Mathios D, Hanemann CO, et al. Genotype-phenotype correlations in neurofibromatosis and their potential clinical use. Neurology 2021;97(7 Suppl 1):S91-8.
8. Rauen KA, Huson SM, Burkitt-Wright E, Evans DG, Farschtschi S, Ferner RE, et al. Recent developments in neurofibromatoses and RASopathies: management, diagnosis and current and future therapeutic avenues. Am J Med Genet A 2015;167A:1-10.
9. Evans DG, Howard E, Giblin C, Clancy T, Spencer H, Huson SM, et al. Birth incidence and prevalence of tumor-prone syndromes: estimates from a UK family genetic register service. Am J Med Genet A 2010;152A:327-32.
10. Friedman JM. Epidemiology of neurofibromatosis type 1. Am J Med Genet 1999;89:1-6.
11. Antonio JR, Goloni-Bertollo EM, Tridico LA. Neurofibromatosis: chronological history and current issues. An Bras Dermatol 2013;88:329-43.
12. Usitalo E, Rantanen M, Kallionpaa RA, Poyhonen M, Leppävirta J, Yla-Outinen H, et al. Distinctive cancer associations in patients with neurofibromatosis type 1. J Clin Oncol 2016;34:1978-86.
13. Ly KI, Blakeley JO. The diagnosis and management of neurofibromatosis type 1. Med Clin North Am 2019;103:1035-54.
14. Skuse GR, Kosciolk BA, Rowley PT. Molecular genetic analysis of tumors in von Recklinghausen neurofibromatosis: loss of heterozygosity for chromosome 17. Genes Chromosomes Cancer 1989;1:36-41.
15. Messiaen LM, Callens T, Mortier G, Bensin D, Vandenbroucke I, van Roy N, et al. Exhaustive mutation analysis of the NF1 gene allows identification of 95% of mutations and reveals a high frequency of unusual splicing defects. Hum Mutat 2000;15:541-55.
16. Weiss B, Bollag G, Shannon K. Hyperactive Ras as a therapeutic target in neurofibromatosis type 1. Am J Med Genet 1999;89:14-22.
17. Daston MM, Scorable H, Nordlund M, Sturbaum AK, Nissen et al. The clinical and diagnostic implications of mosaicism in the neurofibromatoses. Neurology 2001;56:1433-43.
18. Upadhyaya M, Huson SM, Davies M, Thomas N, Chuzhanova N, Giovannini S, et al. An absence of cutaneous neurofibromas associated with a 3-bp inframe deletion in exon 17 of the NF1 gene (c.2970-2972 delAAT): evidence of a clinically significant NF1 genotype-phenotype correlation. Am J Hum Genet 2007;80:140-51.
19. Pinna V, Lanari V, Daniele P, Consoli F, Agolino E, Margiotti K, et al. p.Arg1809Cys substitution in neurofibromin is associated with a distinctive NF1 phenotype without neurofibromas. Eur J Hum Genet 2015;23:1068-71.
20. Koczkowska M, Chen Y, Callens T, Gomes A, Sharp A, Johnson S, et al. Genotype-phenotype correlation in NF1: evidence for a more severe phenotype associated with missense mutations affecting NF1 codons 844-848. Am J Hum Genet 2018;102:69-87.
21. Kehrer-Sawatzki H, Mautner VF, Cooper DN. Emerging genotype-phenotype relationships in patients with large NF1 deletions. Hum Genet 2017;136:349-76.
22. DeBella K, Szudek J, Friedman JM. Use of the national institutes of health criteria for diagnosis of neurofibromatosis 1 in children. Pediatrics 2000;105(3 Pt 1):608-14.
23. Hirsch NP, Murphy A, Radcliffe JJ. Neurofibromatosis: clinical presentations and anaesthetic implications. Br J Anaesth 2001;86:555-64.
24. Legius E, Messiaen L, Wolkenstein P, Pancza P, Avery RA, Berman Y, et al. Revised diagnostic criteria for neurofibromatosis type 1 and Legius syndrome: an international consensus recommendation. Genet Med 2021;23:1506-13.
25. Jouhilahit EM, Peltonen S, Callens T, Jokinen E, Heape AM, Messiaen L, et al. The development of cutaneous neurofibromas. Am J Pathol 2011;178:500-5.
26. Wu M, Wallace MR, Muir D. Tumorigenic properties of neurofibromin-deficient Schwann cells in culture and as syngrafts in NF1 knockout mice. J Neurosci Res 2005;82:357-67.
27. Rutkowski JG, Wu K, Gutmann DH, Boyer PJ, Legius E. Genetic and cellular defects contributing to benign tumor formation in neurofibromatosis type 1. Hum Mol Genet 2000;9:1059-66.
28. Le IQ, Parada LF. Tumor microenvironment and neurofibromatosis type I: connecting the GAPs. Oncogene 2007;26:4609-16.
29. Munchhoff AM, Li F, White HA, Mead LE, Krier TR, Fenoglio A, et al. Neurofibroma-associated growth factors activate a distinct signaling network to alter the function of neurofibromin-deficient endothelial cells. Hum Mol Genet 2006;15:1858-69.
30. Yang FC, Ingram DA, Chen S, Hingtgen CM, Ratner N, Monk KR, et al. Neurofibromin-deficient Schwann cells secrete a potent migratory stimulus for NF1+/- mast cells. J Clin Invest 2003;112:1851-61.
31. Yang FC, Ingram DA, Chen S, Zhu Y, Yuan J, Li X, et al. NF1-dependent tumors require a microenvironment containing NF1+/- and c-kit-dependent bone marrow. Cell 2008;135:437-48.
32. Williams VC, Lucas J, Babcock MA, Gutmann DH, Korf B, Maria BL. Neurofibromatosis type 1 revisited. Pediatrics 2009;123:124-33.
33. Huson SM, Compston DA, Harper PS. A genetic study of von Recklinghausen neurofibromatosis in south east Wales. II. Guidelines for genetic counselling. J Med Genet 1989;26:712-21.
34. Roth TM, Petry EM, Barald KE. The role of steroid hormones in the NF1 phenotype: focus on pregnancy. Am J Med Genet 2008;146A:1624-33.
35. Riccardi VM. An overview of NF-1: dysplasia and neoplasia. In: Friedman JM, Gutmann DH, MacCollin M, Riccardi VM, editors. Neurofibromatosis: phenotype, natural history, and
pathogenesis. Baltimore: The Johns Hopkins University Press; 1992. p. 28-33.
38. Brenet E, Nizery-Guermeur C, Audebert-Bellanger S, Ferkal S, Wolkenstein P, Misery L, et al. Clinical characteristics of pruritus in neurofibromatosis 1. Acta Derm Venereol 2016;96:398-9.
39. Aponte-Lopez A, Munoz-Cruz S. Mast cells in the tumor microenvironment. Adv Exp Med Biol 2020;1273:159-73.
40. Kamide R, Nomura N, Niimura M. Characterization of mast cells residing in cutaneous neurofibromas. Dermatologica 1989;179 Suppl 1:124.
41. Riccardi VM. Cutaneous manifestation of neurofibromatosis: cellular interaction, pigmentation, and mast cells. Birth Defects Orig Artic Ser 1981;17:129-45.
42. Ortonne N, Wolkenstein P, Blakeley JO, Korf B, Plotkin SR, Riccardi VM, et al. Cutaneous neurofibromas: current clinical and pathologic issues. Neurology 2018;91(2 Suppl 1):S5-13.
43. Beert E, Brems H, Daniels B, De Wever I, Van Calenbergh F, Schoenaers J, et al. Atypical neurofibromas in neurofibromatosis type 1 are premalignant tumors. Genes Chromosomes Cancer 2011;50:1021-32.
44. Wolkenstein P, Zeller J, Revuz J, Ecosse E, Leplege A. Quality-of-life impairment in neurofibromatosis type 1: a cross-sectional study of 128 cases. Arch Dermatol 2001;137:1421-5.
45. Verma SK, Riccardi VM, Plotkin SR, Weinberg H, Anderson RR, Blakeley JO, et al. Considerations for development of therapies for cutaneous neurofibroma. Neurology 2018;91(2 Suppl 1):S21-30.
46. Nguyen R, Dombi E, Widemann BC, Solomon J, Fuensterer C, Kluwe L, et al. Loss of NF1 expression and altered vascular morphogenesis in human endothelial cells promotes autonomous proliferation of peripheral nerve sheath tumors in neurofibromatosis type 1. PLoS One 2012;7:e49222.
47. Park H, Lee Y, Yeo H, Park H. Surgical refinement of the purse-string suture for skin and soft tissue defects of the head and neck. Arch Craniofac Surg 2021;22:183-92.
48. Ryu JY, Eo PS, Lee JS, Lee JW, Lee SJ, Lee JM, et al. Surgical approach for venous malformation in the head and neck. Arch Craniofac Surg 2019;20:304-9.
49. Lee JH, Kim NG, Lee KS, Kim JS. Malignant peripheral nerve sheath tumor in frontal sinus, orbital cavity and ethmoid cavity. Arch Craniofac Surg 2014;15:125-8.
50. Peltonen S, Kallionpaa RA, Rantanen M, Uusitalo E, Lahteenpera S, Mowinckel P, et al. Pediatric malignancies in neurofibromatosis type 1: a population-based cohort study. Int J Cancer 2019;145:2926-32.
51. Farid M, Demicco EG, Garcia R, Ahn L, Merola PR, Cioffi A, et al. Malignant peripheral nerve sheath tumors. Oncologist 2014;19:193-201.
52. Hirbe AC, Gutmann DH. Neurofibromatosis type 1: a multidisciplinary approach to care. Lancet Neurol 2014;13:384-43.
53. Evans DG, Salvador H, Chang VY, Erez A, Voss SD, Schneider KW, et al. Cancer and central nervous system tumor surveillance in pediatric neurofibromatosis 1. Clin Cancer Res 2017;23:e46-53.
54. Mautner VF, Asuagbor FA, Dombi E, Funsterer C, Kluwe L, Wenzel R, et al. Assessment of benign tumor burden by whole-body MRI in patients with neurofibromatosis 1. Neuro Oncol 2010;12:593-8.
55. Miller DT, Freedenberg D, Schorry E, Ulrich NJ, Viskochil D, Korf BR, et al. Health supervision for children with neurofibromatosis type 1. Pediatrics 2019;143:e20190660.
56. Mautner VF, Asuagbor FA, Dombi E, Funsterer C, Kluwe L, Wenzel R, et al. Assessment of benign tumor burden by whole-body MRI in patients with neurofibromatosis 1. Neuro Oncol 2008;10:593-8.
Neurooncol 2014;116:307-13.
67. King AA, Debaun MR, Riccardi VM, Gutmann DH. Malignant peripheral nerve sheath tumors in neurofibromatosis 1. Am J Med Genet 2000;93:388-92.
68. Reilly KM, Kim A, Blakely J, Ferner RE, Gutmann DH, Legius E, et al. Neurofibromatosis type 1-associated MPNST state of the science: outlining a research agenda for the future. J Natl Cancer Inst 2017;109:jdx124.
69. Malbari F, Spira M, Knight P, Zhu C, Roth M, Gill J, et al. Elevated risk for MPNST in NF1 microdeletion patients. Am J Hum Genet 2003;72:1288-92.
70. Woodruff JM. Pathology of tumors of the peripheral nerve sheath in type 1 neurofibromatosis. Am J Med Genet 1999;89:23-30.
71. Wasa J, Nishida Y, Tsukushi S, Shido Y, Sugihara H, Nakashima H, et al. MRI features in the differentiation of malignant peripheral nerve sheath tumors and neurofibromas. AJR Am J Roentgenol 2010;194:1568-74.
72. Aslawat S, Fayad LM, Khan MS, Bredella MA, Harris GJ, Evans DG, et al. Current whole-body MRI applications in the neurofibromatoses: NF1, NF2, and schwannomatosis. Neurology 2016;87(7 Suppl 1):S31-9.
73. Ferner RE, Golding JF, Smith M, Calonje E, Jan W, Sanjayana al nerve sheath tumors in patients with neurofibromatosis type 1. J Neurosurg Pediatr 2013;11:e359-63.
74. Martin E, Pendleton C, Verhoef C, Spinner RJ, Coert JH; MONACO collaborators. Morbidity and function loss after resection of malignant peripheral nerve sheath tumors. Neurosurgery 2022;90:354-64.
75. Dunn GP, Spiliopoulos K, Plotkin SR, Hornicek FJ, Harmon DC, Delaney TF, et al. Role of resection of malignant peripheral nerve sheath tumors in patients with neurofibromatosis type 1. J Neurosurg 2013;118:142-8.
76. Lu VM, Wang S, Daniels DJ, Spinner RJ, Levi AD, Niazi TN. The clinical course and role of surgery in pediatric malignant peripheral nerve sheath tumors: a database study. J Neurosurg Pediatr 2021;29:92-9.
77. Tecce MG, Othman S, Mauch JT, Nathan S, Tilahun E, Broach RB, et al. Complex oncologic resection and reconstruction of the scalp: Predictors of morbidity and mortality. Arch Craniofac Surg 2020;21:229-36.
78. Frustraci S, Gherlinzoni F, De Paoli A, Bonetti M, Azzarelli A, Comandone A, et al. Adjuvant chemotherapy for adult soft tissue sarcomas of the extremities and girdles: results of the Italian randomized cooperative trial. J Clin Oncol 2001;19:1238-47.
79. Sarcoma Meta-analysis Collaboration (SMAC). Adjuvant chemotherapy for localised resectable soft tissue sarcoma in adults. Cochrane Database Syst Rev 2009;(4):CD001419.
80. Farmer RE, Gutmann DH. International consensus statement on malignant peripheral nerve sheath tumors in neurofibromatosis. Cancer Res 2002;62:1573-7.
81. Kroep JR, Ouali M, Gelderblom H, Le Cesne A, Dekker TJ, Van Gabbeke M, et al. First-line chemotherapy for malignant peripheral nerve sheath tumor (MPNST) versus other histological soft tissue sarcoma subtypes and as a prognostic factor for MPNST: an EORTC soft tissue and bone sarcoma group study. Ann Oncol 2011;22:207-14.
82. Friedman JM, Birch PH. Type 1 neurofibromatosis: a descriptive analysis of the disorder in 1,728 patients. Am J Med Genet 1997;70:138-43.
83. Jackson IT, Carbonnel A, Potparic Z, Shaw K. Orbitotemporal neurofibromatosis: classification and treatment. Plast Reconstr Surg 1993;92:1-11.
84. Arrington DK, Danehy AR, Peleggi A, Proctor MR, Irons MB, Ullrich NJ. Calvarial defects and skeletal dysplasia in patients with neurofibromatosis type 1. J Neurosurg Pediatr 2013;11:410-6.
85. Kim SH, Lee WJ, Chang JH, Moon JH, Kang SG, Kim CH, et al. Anterior skull base reconstruction using an anterolateral thigh free flap. Arch Craniofac Surg 2021;22:232-8.
86. Lee L, Yan YH, Pharoah MJ, Radiographic features of the mandible in neurofibromatosis: a report of 10 cases and review of the literature. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1996;81:361-7.
87. Sanchez LD, Bui A, Klesse LJ. Targeted therapies for the neurofibromatoses. Cancers (Basel) 2021;13:6032.
88. Richter V, Spiliopoulos K, Plotkin SR, Hornicek FJ, Harmon DC, Delaney TF, et al. Role of resection of malignant peripheral nerve sheath tumors in patients with neurofibromatosis type 1. J Neurosurg Pediatr 2013;11:18:142-8.
89. Lu VM, Wang S, Daniels DJ, Spinner RJ, Levi AD, Niazi TN. The clinical course and role of surgery in pediatric malignant peripheral nerve sheath tumors: a database study. J Neurosurg Pediatr 2021;29:92-9.
90. Wasa J, Nishida Y, Tsukushi S, Shido Y, Sugihara H, Nakashima H, et al. MRI features in the differentiation of malignant peripheral nerve sheath tumors and neurofibromas. AJR Am J Roentgenol 2010;194:1568-74.
91. Ahlawat S, Fayad LM, Khan MS, Bredella MA, Harris GJ, Evans DG, et al. Current whole-body MRI applications in the neurofibromatoses: NF1, NF2, and schwannomatosis. Neurology 2016;87(7 Suppl 1):S31-9.
92. Martin E, Pendleton C, Verhoef C, Spinner RJ, Coert JH; MONACO collaborators. Morbidity and function loss after resection of malignant peripheral nerve sheath tumors. Neurosurgery 2022;90:354-64.
93. Dunn GP, Spiliopoulos K, Plotkin SR, Hornicek FJ, Harmon DC, Delaney TF, et al. Role of resection of malignant peripheral nerve sheath tumors in patients with neurofibromatosis type 1. J Neurosurg 2013;118:142-8.
94. Lee L, Yan YH, Pharoah MJ, Radiographic features of the mandible in neurofibromatosis: a report of 10 cases and review of the literature. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1996;81:361-7.
95. Sanchez LD, Bui A, Klesse LJ. Targeted therapies for the neurofibromatoses. Cancers (Basel) 2021;13:6032.
96. Richter V, Spiliopoulos K, Plotkin SR, Hornicek FJ, Harmon DC, Delaney TF, et al. Role of resection of malignant peripheral nerve sheath tumors in patients with neurofibromatosis type 1. J Neurosurg Pediatr 2013;11:18:142-8.
97. Lu VM, Wang S, Daniels DJ, Spinner RJ, Levi AD, Niazi TN. The clinical course and role of surgery in pediatric malignant peripheral nerve sheath tumors: a database study. J Neurosurg Pediatr 2021;29:92-9.
98. Wasa J, Nishida Y, Tsukushi S, Shido Y, Sugihara H, Nakashima H, et al. MRI features in the differentiation of malignant peripheral nerve sheath tumors and neurofibromas. AJR Am J Roentgenol 2010;194:1568-74.
99. Ahlawat S, Fayad LM, Khan MS, Bredella MA, Harris GJ, Evans DG, et al. Current whole-body MRI applications in the neurofibromatoses: NF1, NF2, and schwannomatosis. Neurology 2016;87(7 Suppl 1):S31-9.
Ullrich NJ, et al. Phase II trial of pegylated interferon alfa-2b in young patients with neurofibromatosis type 1 and unresectable plexiform neurofibromas. Neuro Oncol 2017;19:289-97.

93. Cichowski K, Jacks T. NF1 tumor suppressor gene function: narrowing the GAP. Cell 2001;104:593-604.

94. Dombi E, Baldwin A, Marcus LJ, Fisher MJ, Weiss B, Kim A, et al. Activity of selumetinib in neurofibromatosis type 1-related plexiform neurofibromas. N Engl J Med 2016;375:2550-60.

95. Gross AM, Wolters PL, Dombi E, Baldwin A, Whitcomb P, Fisher MJ, et al. Selumetinib in children with inoperable plexiform neurofibromas. N Engl J Med 2020;382:1430-42.

96. Gross AM, Dombi E, Widemann BC. Current status of MEK inhibitors in the treatment of plexiform neurofibromas. Childs Nerv Syst 2020;36:2443-52.

97. Fisher MJ, Shih CS, Rhodes SD, Armstrong AE, Wolters PL, Dombi E, et al. Cabozantinib for neurofibromatosis type 1-related plexiform neurofibromas: a phase 2 trial. Nat Med 2021;27:165-73.