Properties of Milk Treated with High-Power Ultrasound and Bactofugation

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SUMMARY

Research background. Two methods of milk treatment were used, ultrasound (innovative method) and bactofugation, after which the physicochemical and sensory properties of the milk were examined, with the primary aim of achieving the quality and consistency of the pasteurized milk.

Experimental approach. Ultrasound power of 200 and 400 W and frequency of 24 kHz with constant wave cycle were used. Milk was treated for 2.5, 5, 7.5 and 10 min with sonification at 20 °C (room temperature) and thermosonification (ultrasound at temperature higher than room temperature) at 55 °C. The purpose of this study is to investigate the effect of high-power ultrasound combined with a slightly increased temperature on whole, skimmed and skimmed cow’s milk pretreated with bactofugation.

Results and conclusions. The best sensory quality was achieved when milk was treated with ultrasound power of 200 W at 20 °C for max. 7.5 min. This research shows the potential of the applications of high-power ultrasound in dairy industry combined with bactofugation as a pretreatment of milk at a slightly increased temperature (up to 55 °C).

Novelty and scientific contribution. The application of these two treatments requires milder processing conditions than pasteurization, it is economical and more environmentally friendly technological process that preserves better nutritional values of milk, which is preferred by consumers.

Key words: milk properties, high-power ultrasound, bactofugation, milk treatment

INTRODUCTION

Since milk is the most complete natural liquid because it contains all the substances necessary for the maintenance of health and normal functioning of the human body, it deserves special attention (1). According to Food and Agriculture Organization (FAO) estimates and considering the trend of further growth of the world population, gradual increase in living standards and improvement in dietary habits (2), it is expected that the demand and production of milk will increase at rates of 1.4 % (1990–2010) to a projected growth rate of milk production of 2 % by 2021. Foods available on the market are expected to have sensory properties as close as possible to the original unprocessed food in addition to the usual inevitable safety, high quality and nutritional value. Therefore, recent research has focused on the development of new food processing methods with the aim of obtaining high quality food products (3). Special attention is now paid to non-thermal processing methods, which include the application of high intensity ultrasound (4,5), treatment with high hydrostatic pressures, pulsed electric and magnetic fields (6) and non-thermal atmospheric plasma (7). In food treatment, high hydrostatic pressures are commercially applicable and ultrasound (US) is used in homogenization, emulsification and dispersion processes (8,9). Other applications and processes are still in the experimental phase, and in this respect further research on application of high and low ultrasound frequencies is important. The Food and Agriculture Organization (FAO) of the United Nations and the World Health Organization (WHO) define various thermal processes for the reduction and removal of microorganisms from milk (10,11). Bactofugation is used to improve
the bacterial quality of raw milk. This mechanical process is mainly used in the production of cheese to eliminate anaerobic spores that can affect the flavour and destroy the texture of cheese due to uncontrolled gas formation. It is increasingly used in the production of pasteurized milk and ultra-high temperature milk (UHT), for reducing the number of heat-resistant microorganisms prior to thermal processing, all with the goal of increasing the shelf life of the milk through a mild temperature regime. Bactofugation uses a centrifugal force to remove bacteria and spores from milk as a simple and cost-effective complement to regular pasteurization (12). Bactofugation of milk can remove 80–90 % bacteria and 90–95 % spores (1). During the process, centrifugation force is gradually accelerated to achieve gentle treatment. The optimum bactofugation temperature at which the best results are achieved is 55–60 °C (12,13).

The use of high-power ultrasound has demonstrated several advantages over thermal pasteurization, such as minimizing flavour loss in juices, greater homogeneity and significant energy savings (14). Ultrasound improves the inactivation of microorganisms and affects enzyme activity through the effect of cavitation and is applicable to dairy products, fruits and vegetables (15,16). The advantages of ultrasound over sterilization are minimal loss of flavour, better homogenization and significant energy savings (17). A combination of ultrasound with high pressure, heat or pH change has been shown to be an effective method of killing microorganisms due to the effect of the produced free radicals, generated heat and resulting shear forces (18). However, it is necessary to pay attention to the proper application of ultrasound, as too high ultrasonic power can cause drastic changes in milk fat composition, resulting in a bitter, tasteless liquid due to the oxidation of fat (18). Proper application of ultrasound requires the use of appropriate power, amplitude of the sound wave and ultrasound frequency, as well as the optimum treatment time at lower temperatures to avoid undesirable changes in the treated material. Therefore, the aim of this work is to investigate the possibility of processing raw whole milk, skimmed milk and previously bactofuged skimmed milk by applying high-power ultrasound in combination with a slightly elevated temperature in order to achieve the same microbiological acceptability as with pasteurization and to monitor the possible effects of the two treatments (ultrasound and bactofugation) on the chemical composition and sensory properties of the milk.

MATERIALS AND METHODS

Milk samples were collected aseptically from a Croatian dairy industry in sterile vials from the sampling valves before the separator (raw whole milk, A), after the separator (skimmed milk, B), after the bactofugate (skimmed bactofuged milk, C), and the final sample was the milk pasteurized by the classical high-temperature short-time method (72 °C/15 s) used as reference sample (D) for each experiment, as shown in Fig. S1. Samples from each batch were analyzed in triplicate.

Processing of milk with ultrasonic processor

The ultrasonic processor used in this study was model UP 400S, Hielser Ultrasonics GmbH, Teltow, Germany, with effective output power 400 W, voltage 230 V, 10–100 % ultrasound cycle, ultrasonic frequency 24 kHz and amplitude 12–260 μm. We used a 7-mm diameter titanium probe immersed at a depth of 2 cm in each sample of milk.

Four experimental treatments were designed with different applied ultrasound power (at the same frequency 24 kHz) and temperature during the transduction, as follows: experiment A: \( P=200 \text{ W}, t=20 \text{ °C} \), experiment B: \( P=200 \text{ W}, t=55 \text{ °C} \), experiment C: \( P=400 \text{ W}, t=20 \text{ °C} \), and experiment D: \( P=400 \text{ W}, t=55 \text{ °C} \).

Design of ultrasound treatment

Within each of the four experimental treatments, ultrasonic processing was performed on three different samples of milk: on the raw whole milk, skimmed milk and skimmed bactofuged milk, while pasteurized samples were used as a reference (19). Each of the three different milk samples was treated during four time periods (2.5, 5, 7.5 and 10 min). Thus, 12 treatments (3 milk samples × 4 time periods) were performed in each experiment (A–D), including four control samples (raw whole milk, skimmed milk, skimmed bactofuged milk and pasteurized milk), hence a total of we analysed 64 samples (4×12 treated and 4×4 reference samples). All trials were performed as three independent measurements, and the results represent the mean value of all three measurements. In order to carry out the statistical analysis, the applied multivariate tools showed the influence of the main components and the correlation of the values of all the experiments in which the treatment (T) conditions are observed: treatments T1–T4 (\( P=200 \text{ or } 400 \text{ W}, v=24 \text{ kHz, } t=20 \text{ or } 55 \text{ °C} \)) are raw whole milk samples treated for 2.5 (T1), 5 (T2), 7.5 (T3) and 10 (T4) min, treatments T5–T8 (\( P=200 \text{ or } 400 \text{ W, } v=24 \text{ kHz, } t=20 \text{ or } 55 \text{ °C} \)) are skimmed milk samples treated for 2.5 (T5), 5 (T6), 7.5 (T7) and 10 (T8) min, and treatments T9–T12 (\( P=200 \text{ or } 400 \text{ W, } v=0 \text{ or } 24 \text{ kHz, } t=20 \text{ or } 55 \text{ °C} \)) are skimmed bactofuged milk samples treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min. The process parameters \( P, v, t \) and time varied in each conducted experiment (experiment A: \( P=200 \text{ W, } t=20 \text{ °C} \), experiment B: \( P=200 \text{ W, } t=55 \text{ °C} \), experiment C: \( P=400 \text{ W, } t=20 \text{ °C} \), and experiment D: \( P=400 \text{ W, } t=55 \text{ °C} \).

The acidity of milk

Physical quality of all milk samples before and immediately after the treatment was measured. Titratable acidity and active acidity of the samples was monitored. We used the standard Soxhlet-Henkel method for determining the titration acidity of milk (T). Reagents used for the analysis were 0.1 M NaOH solution, 2 % alcoholic solution of phenolphthalein, and 5 % cobalt sulfate solution (CoSO₄·7H₂O). Cobalt sulfate
mixed with 50 mL milk was the reference pink. Briefly, 1 mL 
of phenolphthalein indicator was added to 20 mL tempered milk sample at 20 °C, the mixture was stirred and quickly 
titrated with 0.1 M NaOH with continuous stirring until the col-
our changed to a light pink that was stable for one min and 
compared to the prepared standard pink colour (a mixture of 1 
ml of 5 % cobalt sulfate solution and 20 mL milk freshly pre-
pared, no longer than 3 h before titration). The milk sample 
was titrated with 0.1 M NaOH and milk acidity was calculated 
according to the formula:

\[ \text{Titratable acidity} = V_{\text{NaOH}} \cdot 5 \cdot c_{\text{NaOH}} / 1 \]

where \( V_{\text{NaOH}} \) is the volume of 0.1 M NaOH (in mL) used to neu-
tralize 20 mL of the test sample, \( S \) is a constant, and \( c_{\text{NaOH}} = 0.1 \) 
mol/L. The results are expressed in Soxhlet-Henkel degrees 
(°SH) as the arithmetic mean of three parallel analyses. The 
maximum permissible difference in triple milk analyses was 
0.2 °SH. If larger difference is encountered, the analysis was 
repeated.

The pH was measured using a pH meter model 225 (Met-
tler Toledo, Munich, Germany) with an integrated tempera-
ture compensation. The electrodes were immersed in a sam-
ples of milk that was lightly stirred until the value of the display 
remained stable. For the same sample, three measurements 
were performed with washing and wiping of the electrode 
between measurements, and the result was the arithmetic 
mean.

**Determination of the chemical composition of milk samples**

The composition of all milk samples before and immedi-
ately after the treatment was analyzed. The mass fractions (w/
g/100 g) of total solids, non-fat solids, milk fat, protein and 
lactose were determined using the IR spectroscopy (20) and 
MilkoScan 4000 (Type 71200; Foss Electric A/S, Copenhagen, 
Denmark) instrument, while calcium mass fraction was deter-
mined with the titrimetric method (21). A volume of 2.5 mL 
milk was supplemented with 100 mL distilled water. The solu-
tion was transferred to an Erlenmeyer flask in which 2 mL of 
10 M NaOH were added and the mixture was stirred on a 
metallic stirrer for 2–3 min. Two drops of indicator cal-
carbonic acid were added and the mixture was titrated with 
a 0.05 M EDTA solution with stirring until the light red 
colour changed to light blue.

To determine the accuracy of the titration, 0.25 mL of 0.05 M 
CaCl\(_2\) was added to the titrated solution and the solution 
turned red again, then it was titrated once again with 0.05 M 
EDTA solution until the colour changed to light blue. The cal-
cium mass (in mg) in 100 g milk was calculated according to 
the following formula:

\[ m(Ca^{2+}) = 80.16 \cdot (V_1 + V_2 - 0.25) / 2 \]

where 80.16 and 0.25 are constants, \( V_1 \) is the volume of EDTA 
solution used for the first titration (mL), and \( V_2 \) is the volume 
of EDTA solution used for the second titration (mL).

**Sensory evaluation of milk samples**

Sensory examination included an assessment of the ap-
pearance (max. 3 points), colour (max. 2 points), odour (max. 
3 points) and taste (max. 12 points) of milk. The analysis of 
appearance included texture, consistency and possible dep-
osition, and the taste included standard flavours and aromas
based on the IDF Standards (22).

The sensory evaluation was conducted by a committee 
consisting of five certified sensory analysts for milk and dairy 
products (mean age 35.7, two men and three women). All 
samples were analyzed at 20 °C in labelled glass cups. The 
odour was evaluated first. When evaluating the colour of 
the samples, the reference samples of the original colour of 
whole and skimmed milk were used, in fully transparent 
glasses. The precipitate and the consistency of milk were con-
trolled by pouring the milk into another glass cup down the 
wall to make it easy to observe possible flakes and bruises 
created by the grain.

For the evaluation of the analysis, the scoring method 
was used with a maximum of 20 points (22). In the cases when 
the grade of individual sensory properties in the examined 
sample decreased by ±1 point, the mean was taken, and 
when it deviated by more than 1 point the analysis was re-
peated. The disqualified sample on the sensor was the one 
that received 0 points for any property.

**Statistical data processing**

Descriptive statistics was used to obtain mean values, 
standard errors and minimum and maximum values for each 
treatment and separately for each experiment (23). In the ex-
amination of similarities or differences in the data for each 
observed characteristic (chemical, physical parameter and 
sensory evaluation), t-test was used for different treatments 
and experiments with the chosen level of significance (risk) 
0.05 (95 %). For the purposes of linking, i.e. establishing simi-
larities and/or differences in a large set of data for each ob-
served feature (treatment by experiment), multivariate statis-
tical methods were applied on the same set of data as well as 
on the data not belonging to the same set (24).

Factor analysis is the generic name given to the class of 
multivariate statistical methods whose primary purpose is to 
define the basic structure in the data matrix. In general, it 
deals with the problem of analyzing the correlation structure 
between a large number of variables (e.g. sensory test, phys-
icochemical measurements, etc.) by defining a set of common 
dimensions known as factors (24). In factor analysis, factors 
are formed by maximizing their explanation of a whole set of 
variables.

We applied the principal component analysis (PCA) to 
confirm the grouping, and the analysis of the main compo-
nents also provided a pictorial representation to determine 
the reasons why groups were formed in a particular manner. 
Multivariate analysis (chemometrics) proved to be a powerful 
tool for identifying the data in the experimental part (25,26).
Among the chemometric methods, the principal component analysis served to identify the experimental data and group them based on their similarity and variety. First, the data were organized in the matrix with the treatments set in rows and experiments located in the columns (27). Each variable vector was automatically scaled with the corresponding wavelength of the sample and displayed as:

\[ X_{ij} \leftarrow \frac{X_{ij} - \bar{X}_j}{\sigma_j(X_i)} \]

The scaled matrix of data \( X \) is approximated with projections in the subsystem of the main components \( P \):

\[ T = X \cdot P \]

Data processing software STATISTICA v. 8 (28) was used for data processing.

**RESULTS AND DISCUSSION**

The research was carried out on milk sampled from the production line at various processing stages. Raw whole milk, skimmed milk and skimmed milk after bactofugation were used. The effect of pretreatment of milk by bactofugation has been investigated, as we assumed that microbiological results of sonification of previously bactofuged milk would be more significant and that this combination might extend the shelf life of milk. The centrifugal separator of bacteria (bactofuge) can isolate about 80–90% of the bacteria and 90–95% of the spores from the milk, so it is to be expected that such a reduced number of remaining bacteria in the bactofuged milk would be successfully inactivated by ultrasound. The nominal power used on 200 mL of milk was 200 and 400 W at 24 kHz. The treated samples were measured after 2.5, 5, 7.5 or 10 min, and untreated reference samples were used for comparison between the same batch and milk group (raw whole milk, skimmed milk and skimmed bactofuged milk).

The milk treated with ultrasound for too long had an unpleasant taste, which can be explained by the study of Zabbia et al. (29) where the effect of treatment of milk with high intensity ultrasound resulted in the formation of certain volatile components causing a change of taste and the appearance of unpleasant odor. Furthermore, it has been demonstrated that ultrasonic processing of milk at temperatures higher than 60 °C resulted in denaturation of whey proteins (30, 31).

With a large number of experimental data collected (physical quality, chemical composition and analysis of sensory properties of treated and untreated milk samples), it is very difficult to determine the changes in different milk samples under different conditions of treatment (power, frequency, temperature and time). In such complex systems, multivariate analysis of variance (MANOVA) is used to determine differences in the observed set of data (26).

Table 1 shows the results of the MANOVA analysis how treatment variations affect the titratable acidity and pH values. The multivariate analysis showed that the power of 400 W influenced the change in mean values of titratable acidity (6.78 °SH) and pH (6.680) at a significance level of \( \alpha=0.05 \). The frequency change did not affect the value of titratable acidity, but the frequency of 24 kHz used with a power of 400 W affected the pH change (6.692). Multivariate analysis suggests that the temperature change is significant for the samples heated to 55 and 72 °C for titratable acidity and pH. The processing of milk by high-power ultrasound does not lead to significant pH changes (32). Shanmugam et al. (33) came to the same conclusions using 12-mm diameter ultrasonic probes, 450 W and 20 kHz frequency, skimmed milk with 1.5% fat and time of 15, 30, 45 and 60 min. Walstra et al. (34) found that ultrasonic processing may lower the pH value of milk due to the hydrolysis of phosphoric esters because ultrasound mediates in enzymatic reactions of some enzymes and cavitation can accelerate some chemical reactions causing a fall in the pH. Therefore, this may be the cause of the pH drop at 400 W, 24 kHz and 20 °C (Table 1). The average protein mass fraction in milk is 3.4 g/100 g and ranges from 2.9 to 5.0 (14). Under different experimental conditions (200 or 400 W, frequency of 24 kHz, temperature 20 and 55 °C) and different treatment times (2.5, 5, 7.5 and 10 min), the influence of ultrasound on the change of the total solids, non-fat solids, fat and protein mass fractions was not observed, which is consistent with the conclusions reached by Cameron et al. (35). After the ultrasound treatment of milk, Chemat et al. (36) noted an increase in fat content as a result of the breakdown of fat-aggregate membranes under the influence of ultrasound.

**Table 1. Multivariate analysis (MANOVA) of the effect of experimental treatments (ultrasound power, frequency, temperature and time) on the changes in physical properties of milk samples.**

| Condition | Titratable acidity/°SH | pH       |
|-----------|------------------------|----------|
| P/W       |                        |          |
| 0         | (6.50±0.08) a          | (6.71±0.01) a |
| 200       | (6.63±0.07) a          | (6.70±0.01) a |
| 400       | (6.78±0.07) a          | (6.68±0.01) a |
| v/kHz     |                        |          |
| 0         | (6.5±0.08) a           | (6.71±0.01) a |
| 24        | (6.70±0.05) a          | (6.69±0.01) a |
| t/°C      |                        |          |
| 20        | (6.81±0.05) a          | (6.71±0.01) a |
| 55        | (6.58±0.06) a          | (6.69±0.01) a |
| 72        | (6.40±0.14) a          | (6.72±0.02) a |
| Time/min  |                        |          |
| 0         | (6.61±0.08) a          | (6.71±0.01) a |
| 2.5       | (6.59±0.08) a          | (6.71±0.01) a |
| 5         | (6.67±0.08) a          | (6.70±0.01) a |
| 7.5       | (6.74±0.08) a          | (6.70±0.01) a |
| 10        | (6.80±0.08) a          | (6.69±0.01) a |

Data are presented as mean value±standard error (SE), N=3. Values with different letters in superscript in the same column differ at a significance level of 5%.

The aforementioned effect, however, exceptionally low increase in fat content, can be observed in the present study when the whole milk is processed by ultrasound at 200 W (experiment A) and 400 W (experiments C and D). Average values of the milk composition and physical characteristics are presented in Table 2.
Table 2. Average values of composition and physical characteristics of differently treated milk

| Treatment | Titratable acidity/SH | pH | TS | SNF | Fat | Protein | Lactose | Ca |
|-----------|----------------------|----|----|-----|-----|---------|---------|----|
| SM        | 6.4                  | 6.7| 13.0| 8.7 | 4.3 | 3.4     | 4.6     | 121.8 |
| T1        | 6.2 *                | 6.7| 12.9| 8.6 | 4.3 | 3.4     | 4.5     | 122.2 |
| T2        | 6.2 *                | 6.8| 12.8| 8.7 | 4.3 | 3.4     | 4.6     | 122.2 |
| T3        | 6.2 *                | 6.7| 13.0| 8.7 | 4.3 | 3.4     | 4.6     | 122.9 |
| T4        | 7.0                  | 6.7| 13.0| 8.7 | 4.3 | 3.5     | 4.6     | 122.9 |
| RM        | 6.8                  | 6.8| 9.7 *| 9.4 | 0.4 *| 3.9     | 4.8     | 132.3 *|
| T5        | 7.0                  | 6.8| 9.7 *| 9.3 | 0.4 *| 3.9     | 4.8     | 132.2 *|
| T6        | 6.6                  | 6.8| 9.7 *| 9.3 | 0.5 *| 3.8     | 4.8     | 133.1 *|
| T7        | 6.8                  | 6.8| 9.7 *| 9.4 | 0.4 *| 3.9     | 4.8     | 133.4 *|
| T8        | 6.8                  | 6.8| 9.7 *| 9.3 | 0.5 *| 3.8     | 4.8     | 133.4 *|
| BF        | 6.8                  | 6.8| 9.4 *| 9.0 | 0.5 *| 3.6     | 4.8     | 122.6 |
| T9        | 6.8                  | 6.8| 9.4 *| 9.0 | 0.5 *| 3.6     | 4.8     | 123.0 |
| T10       | 6.8                  | 6.8| 9.5 *| 9.0 | 0.5 *| 3.6     | 4.8     | 122.9 |
| T11       | 6.8                  | 6.8| 9.4 *| 9.0 | 0.5 *| 3.6     | 4.8     | 123.2 |
| T12       | 6.8                  | 6.8| 9.5 *| 9.0 | 0.5 *| 3.6     | 4.8     | 124.1 |
| P         | 6.6                  | 6.6| 8.8  |10.5 *| 9.6 | 0.9 *| 3.7     | 5.0     | 120.2 |

| Treatment | Titratable acidity/SH | pH | TS | SNF | Fat | Protein | Lactose | Ca |
|-----------|----------------------|----|----|-----|-----|---------|---------|----|
| SM        | 6.8                  | 6.7| 12.8| 8.7 | 4.3 | 3.4     | 4.6     | 119.4 |
| T1        | 6.6                  | 6.6| 12.9| 8.7 | 4.3 | 3.4     | 4.6     | 120.2 |
| T2        | 7.8 *                | 6.5| 13.0| 8.8 | 4.2 | 3.5     | 4.6     | 119.2 |
| T3        | 8.4 *                | 6.5| 13.0| 8.9 | 4.3 | 3.6     | 4.6     | 119.4 |
| T4        | 8.6 *                | 6.5| 13.2| 8.9 | 4.4 | 3.6     | 4.6     | 120.2 |
| RM        | 6.6                  | 6.7| 12.8| 8.7 | 0.5 *| 3.8     | 4.8     | 121.0 |
| T5        | 6.6                  | 6.7| 12.9| 8.7 | 0.5 *| 3.7     | 4.7     | 122.4 |
| T6        | 6.7                  | 6.7| 13.0| 8.8 | 0.5 *| 3.8     | 4.8     | 121.2 |
| T7        | 7.2                  | 6.7| 13.0| 8.9 | 0.4 *| 3.8     | 4.8     | 122.6 |
| T8        | 7.4                  | 6.7| 13.2| 8.9 | 0.4 *| 3.8     | 4.8     | 121.8 |
| BF        | 6.8                  | 6.7| 9.5 *| 9.0 | 0.5 *| 3.5     | 4.8     | 123.4 |
| T9        | 6.8                  | 6.6| 9.7 *| 9.0 | 0.5 *| 3.6     | 4.8     | 125.0 |
| T10       | 6.8                  | 6.6| 9.5 *| 9.0 | 0.5 *| 3.6     | 4.8     | 123.4 |
| T11       | 6.8                  | 6.6| 9.4 *| 9.0 | 0.41 *| 3.5     | 4.8     | 120.2 |
| T12       | 6.6                  | 6.7| 9.4 *| 9.0 | 0.5 *| 3.5     | 4.8     | 126.8 |
| P         | 6.6                  | 6.6| 9.7 *| 9.2 | 0.5 *| 3.7     | 5.0     | 122.6 |

* and * for the same experiment (C or D): significantly different characteristics (significance level of 5 %) of differently treated milk (RM=raw whole milk, SM=skimmed milk, BF=skimmed bactofuged milk and P=pasteurized milk), TS=total solids, SNF=non-fat solids, T1–T4=raw whole milk, T5–T8=skimmed milk treated for 2.5 (T5), 5 (T6), 7.5 (T7) and 10 (T8) min and T9–T12=skimmed bactofuged milk treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min.

The multivariate analysis of the chemical composition of milk samples (Table 3) treated with different intensities of ultrasound showed very low standard error (SE) for lactose. The difference in the lactose content was significant at 5 % significance level in pasteurized samples (a temperature of 72 °C and a time of 15 s) at a power of 200 W and a frequency of 24 kHz.

A study of the effect of ultrasound on the chemical composition of milk concluded that the ultrasound does not have a significant influence on the lactose content (35). However, using a multivariate analysis, lower lactose content was determined under experimental conditions of 200 W, 24 kHz and 55 °C (experiment B), as a result of the quality of the milk used for a particular batch. The composition of milk is variable, the average lactose mass fraction in milk is 4.8 g/100 g, ranging from 3.6 to 5.5 (14,37). Samples treated at 200 W, 24 kHz and 55 °C showed a mild change of odour in the sense that freshness of whole milk was affected by 7.5 min of processing and of skimmed bactofuged milk by 2.5 and 10 min of processing. These parameters (200 W, 24 kHz, 55 °C) influenced the taste of all whole milk samples in which a change in the characteristic taste of milk with mild taste was burnt, while in the sample of bactofuged skimmed milk treated for 10 min the loss of freshness was observable. Treatment of
whole milk at 400 W and 24 kHz at room temperature for 5 and 10 min caused a change of odour due to the loss of freshness. Zabbia et al. (29) concluded that prolonged treatment leads to a appearance of a large number of volatile compounds and results in the change of taste. The results in this paper (Table 4 and Table 5) are consistent with those findings.

Namely, the total obtained scores of the sensory properties of the observed milk are lower than the reference untreated milk samples and their values decrease with time although there are no differences in the colour parameters and appearance among the samples. Total sensory evaluation of milk samples (Table 4) is the sum of all evaluated parameters (appearance, colour, odour and taste). In experiment A (200 W, 24 kHz, 20 °C), 81% of the samples received ≥15 points, and only 19% received <15 points. The results show that the best process parameters to obtain acceptable organoleptic properties are power output of 200 W at 24 kHz and room temperature, while the lowest organoleptic properties were obtained using ultrasound power of 400 W at 24 kHz and room temperature. Since the frequency and temperature in both experiments were the same, the organoleptic characteristics were most affected by the power of ultrasound. The detailed analysis showed that treatment time is also one of the key parameters for good organoleptic parameters (37). The appearance and colour did not change significantly under the influence of ultrasound power, frequency, temperature and time. The odour changed considerably at higher ultrasound power (400 W), pasteurization at 72 °C and ultrasound treatment time of 7.5 and 10 min. Taste and total sensory evaluation of milk did not change significantly only under the following treatment conditions:

### Table 3. Results of multivariate analysis (MANOVA) of the effect of experimental treatments (ultrasound power, frequency, temperature and time) on the changes in chemical composition of milk samples. Results are compared with untreated samples (P, ν, t and time=0)

| Condition | TS | SNF | Fat | Protein | Lactose | Ca |
|-----------|----|-----|-----|---------|---------|----|
| P/W       | 0  | (10.5±0.4)a | (9.0±0.1)a | (1.2±0.5)a | (3.49±0.05)a | (4.8±0.0)a | (123.7±1.4)a |
| 200       | (10.4±0.4)a | (8.8±0.1)a | (1.6±0.4)a | (3.49±0.04)a | (4.6±0.0)a | (124.9±1.2)a |
| 400       | (11.0±0.4)a | (8.8±0.1)a | (1.5±0.4)a | (3.51±0.04)a | (4.7±0.0)a | (121.8±1.2)a |
| v/kHz     | 0  | (10.5±0.4)a | (9.0±0.1)a | (1.2±0.5)a | (3.49±0.05)a | (4.8±0.0)a | (123.7±1.4)a |
| 24        | (10.7±0.3)a | (8.8±0.0)a | (1.6±0.3)a | (3.50±0.03)a | (4.6±0.0)a | (123.3±0.9)a |
| t/°C      | 20 | (10.9±0.28)a | (8.9±0.0)a | (1.7±0.3)a | (3.54±0.03)a | (4.7±0.0)a | (127.3±0.9)a |
| 55        | (10.2±0.3)a | (8.7±0.1)a | (1.4±0.3)a | (3.44±0.03)a | (4.6±0.0)a | (123.1±0.1)a |
| 72        | (10.4±0.7)a | (9.2±0.1)a | (0.9±0.8)a | (3.49±0.08)a | (4.9±0.0)a | (122.9±2.5)a |
| Time/min  | 0  | (10.6±0.4)a | (8.8±0.1)a | (1.6±0.5)a | (3.49±0.05)a | (4.6±0.0)a | (124.6±1.4)a |
| 2.5       | (10.6±0.4)a | (8.8±0.1)a | (1.5±0.5)a | (3.48±0.05)a | (4.6±0.0)a | (125.1±1.4)a |
| 5         | (10.6±0.4)a | (8.8±0.1)a | (1.5±0.5)a | (3.49±0.05)a | (4.6±0.0)a | (125.1±1.4)a |
| 7.5       | (10.6±0.4)a | (8.8±0.1)a | (1.5±0.5)a | (3.50±0.05)a | (4.6±0.0)a | (125.8±1.4)a |
| 10        | (10.6±0.4)a | (8.8±0.1)a | (1.4±0.5)a | (3.49±0.05)a | (4.6±0.0)a | (126.2±1.4)a |

Data are presented as mean values±standard error (SE), N=3. Values with different letters in superscript in the same column differ at a significance level of 5 %. TS=total solids, SNF=non-fat solids, Ca=calcium

### Table 4. Average values of sensory properties in the experiments A, B, C and D for differently treated milk samples

| Sensory property | SM | T1 | T2 | T3 | T4 | RM | T5 | T6 | T7 | T8 | BF | T9 | T10 | T11 | T12 | P |
|------------------|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| Odour (N=3)      | A  | 3a | 3a | 3a | 3a | 2.5a | 3a | 3a | 3a | 3a | 3a | 3a | 3a | 3a | 3a | 1.5a |
| Taste (N=12)     | A  | 12b | 11.5a | 12b | 9.5a | 6a | 11a | 11a | 7.5a | 8a | 7.5ab | 11a | 11a | 7a | 9a | 5.5a |
| Total (N=20)     | A  | 20a | 19.5a | 20a | 17.5a | 13.5a | 19a | 19a | 15.5a | 16ab | 15.5a | 19a | 19a | 17a | 12a | 19a |

Values in the same column marked with different letters in superscript differ at a significance level of 5 %. RM=raw whole milk, SM=skimmed milk, BF=skimmed bactofuged milk and P= Pasteurized milk, T1–T4= raw whole milk treated for 2.5 (T1), 5 (T2), 7.5 (T3) and 10 (T4) min, T5–T8= skimmed milk treated for 2.5 (T5), 5 (T6), 7.5 (T7) and 10 (T8) min and T9–T12=skimmed bactofuged milk treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min. Experimental treatment: A (P=200 W, ν=20 °C); B (P=200 W, ν=55 °C); C (P=400 W, ν=20 °C) and D (P=400 W, ν=55 °C)
and T9–T12=skimmed bactofuged milk treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min. Experimental treatment: A (T4=raw whole milk treated for 2.5 (T1), 5 (T2), 7.5 (T3) and 10 (T4) min, T5–T8=skimmed milk treated for 2.5 (T5), 5 (T6), 7.5 (T7) and 10 (T8) min and T9–T12=skimmed bactofuged milk treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min. Experimental treatment: A (P=200 W, t=20 °C), B (P=55 °C), C (P=400 W, t=20 °C) and D (P=400 W, t=55 °C)

Table 5. Results of multivariate analysis (MANOVA) on the sensory properties based on the effect of experimental conditions (ultrasound power, frequency, temperature and time) on changes in sensory properties of milk samples. Results are compared with untreated samples (P, ν, t and time=0)

| Condition | Appearance | Colour | Odour | Taste | Total |
|-----------|------------|--------|-------|-------|-------|
| P/W       | (3.00±0.03)* | (2.00±0.00)* | (3.0±0.1)* | (11.4±0.4)* a | (19.4±0.5)* a |
| 200       | (2.98±0.02)* a | (2.00±0.00)* a | (2.9±0.1)* a | (8.8±0.3)* b | (16.7±0.4)* b |
| 400       | (2.94±0.02)* b | (2.00±0.00)* b | (2.5±0.1)* b | (7.9±0.3)* c | (15.4±0.4)* c |
| ν/kHz 0   | (3.00±0.03)* c | (2.00±0.00)* c | (3.0±0.1)* c | (11.3±0.4)* a a | (19.3±0.5)* a a |
| 24        | (2.96±0.02)* b | (2.00±0.00)* b | (2.7±0.1)* b | (8.4±0.2)* a b | (16.0±0.3)* a b |
| 20        | (2.97±0.02)* a | (2.00±0.00)* a | (2.7±0.1)* a | (8.5±0.2)* c a | (16.1±0.3)* c a |
| 55        | (2.97±0.02)* a | (2.00±0.00)* a | (2.5±0.1)* a | (9.2±0.3)* b a | (16.8±0.3)* b a |
| 72        | (3.00±0.03)* a | (2.00±0.00)* a | (3.0±0.2)* a | (11.2±0.7)* b a | (19.2±0.8)* b a |
| t/°C 0    | (3.00±0.03)* a | (2.00±0.00)* a | (3.0±0.1)* a | (11.4±0.4)* a a | (19.4±0.5)* a a |
| 2.5       | (3.00±0.03)* a | (2.00±0.00)* a | (2.9±0.1)* a | (10.3±0.4)* b a | (18.1±0.5)* b a |
| 5         | (3.00±0.03)* a | (2.00±0.00)* a | (2.8±0.1)* a | (9.1±0.4)* b a | (16.9±0.5)* b a |
| 7.5       | (3.00±0.03)* a | (2.00±0.00)* a | (2.6±0.1)* a | (8.3±0.4)* b a | (15.8±0.5)* b a |
| 10        | (2.89±0.03)* b | (2.00±0.00)* b | (2.3±0.1)* b | (7.3±0.4)* a a | (14.4±0.5)* a a |

Data are presented as mean value± standard error (SE), N=3. Values with different letters in superscript differ at a significance level of 5 %. T1–T4=raw whole milk treated for 2.5 (T1), 5 (T2), 7.5 (T3) and 10 (T4) min, T5–T8=skimmed milk treated for 2.5 (T5), 5 (T6), 7.5 (T7) and 10 (T8) min and T9–T12=skimmed bactofuged milk treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min. Experimental treatment: A (P=200 W, t=20 °C), B (P=55 °C), C (P=400 W, t=20 °C) and D (P=400 W, t=55 °C)

![Fig. 1](image)

Fig. 1. Principal component analysis for total sensory evaluation of samples (T1–T12). RM=raw whole milk, SM=skimmed milk, BF=bactofuged milk and P=pasteurized, T1–T4=raw whole milk treated for 2.5 (T1), 5 (T2), 7.5 (T3) and 10 (T4) min, T5–T8=skimmed milk treated for 2.5 (T5), 5 (T6), 7.5 (T7) and 10 (T8) min and T9–T12=skimmed bactofuged milk treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min. Experimental treatment: A (P=200 W, t=20 °C), B (P=55 °C), C (P=400 W, t=20 °C) and D (P=400 W, t=55 °C)
In order to test the relationship between the composition of milk samples from all experiments and their sensory properties, the results of Pearson’s matrix correlation are presented separately for each experiment in Tables S1–S4. Although the results of experiment A suggest that there is no correlation among the sensory properties, milk composition and physical characteristics, they exist but are not significant (significance level 0.05). The protein mass fraction is in a positive correlation with titratable acidity, non-fat solids, lactose and Ca. Total solids are in a negative correlation with most parameters, with the exception of fat, which is understandable since the milk fat is part of the total solids.

However, the advantage of the principal component analysis (Fig. 2) is that it simultaneously shows the changes in sensory and physical characteristics of milk. Thus, it is possible to observe the potential grouping or separation of the treated milk samples against the bactofuged milk samples. Fig. 2 shows the principal component analysis for four different process conditions (A–D), where we observed the physical and sensory characteristics of the milk and the bactofuged samples grouped together in all experiments. Bactofuged samples are independent of the experimental data spread in different squares, but still make a group clearly separated from the other treatments. Under experiment conditions: 400 W, 24 kHz and 20 °C, the bactofuged

**Fig. 2.** Biplot of the first two main components (D1 and D2) of the principal component analysis for physical and sensory properties of the treated milk samples: a) $P_{\text{ultrasound}}=200$ W, $\nu=24$ kHz, $t=20$ °C, b) $P_{\text{ultrasound}}=200$ W, $\nu=24$ kHz, $t=55$ °C, c) $P_{\text{ultrasound}}=400$ W, $\nu=24$ kHz, $t=20$ °C, and d) $P_{\text{ultrasound}}=400$ W, $\nu=24$ kHz, $t=55$ °C. RM=raw whole milk, SM=skimmed milk, BF=skimmed bactofuged milk and P=pasteurized milk. T1–T4=raw whole milk treated for 2.5 (T1), 5 (T2), 7.5 (T3) and 10 (T4) min, T5–T8=skimmed milk treated for 2.5 (T5), 5 (T6), 7.5 (T7) and 10 (T8) min and T9–T12=skimmed bactofuged milk treated for 2.5 (T9), 5 (T10), 7.5 (T11) and 10 (T12) min. Experimental treatment: A ($P=200$ W, $t=20$ °C), B ($P=200$ W, $t=55$ °C), C ($P=400$ W, $t=20$ °C) and D ($P=400$ W, $t=55$ °C)
samples form a very narrow group and are arranged exclusively in the first quadrant (Fig. 2c).

All variations in the observed data set ranged from 69.64% under experimental conditions of 200 W, 24 kHz and 55 °C (Fig. 2b) to 77.14% under the conditions of 200 W, 24 kHz and 20 °C (Fig. 2a). The higher percentage in all experiments belongs to the first major component (D1) dominated by the milk composition and physical characteristics observed in the different samples of milk. In Fig. 2b and Fig. 2c, the bactofuged samples are very close to the reference bactofuged sample. The proximity of treated milk sample and bactofuged milk sample indicates similarity in physico-chemical composition and/or sensory evaluation. At 200 W, 24 kHz and 55 °C, total solids, non-fat solids, fat, proteins, lactose and Ca affected the taste (Fig. 2b). The average overall impression of bactofuged samples under these conditions received 12.43 points. Under experimental conditions of 400 W, 24 kHz and 20 °C, a high contribution of the titratable acidity, pH, odour, taste and total impression (Fig. 2c) was observed.

The worst total sensory score (Fig. 1 and Fig. 2) was found in bactofuged samples (score <15 points for 43.75% samples). Bactofuged milk samples received the worst overall sensory score (<15 points: 43.75% of the samples; Fig. 1 and Fig. 2). The reason for this can be seen from a comparative presentation of the results of the analysis of the main components (Fig. 2), where the samples of bactofuged milk were treated for a longer time than the reference untreated bactofuged sample.

CONCLUSIONS

The results of the performed experiments and the multivariate statistical analysis revealed that the applied ultrasound power of 200 W, frequency of 24 kHz and the treatment time of the milk samples had no significant effect on the titratable acidity and pH of the milk. All the bactofuged samples of skimmed milk had lower values of total solids, non-fat solids, protein and calcium than the samples of the same batch and milk fat. Different conditions of high-power ultrasonic treatment did not affect the changes in total solids, non-fat solids, milk fat and protein. Temperature (20 and 55 °C) and shorter treatment time with ultrasound up to 5 min had no significant effect on the overall sensory evaluation of milk, while ultrasound power of 400 W had a significant effect on the change of taste of the whole milk samples treated for longer than 5 min. The best sensory characteristics of the bactofuged skimmed milk samples were obtained with an ultrasound power of 400 W for up to 7.5 min regardless of the temperature. For all milk types (whole milk, skimmed and bactofuged milkfat), the highest sensory evaluation was obtained with ultrasound power of 200 W, frequency of 24 kHz, temperature of 20 °C and treatment time of 7.5 min. Ultrasonic treatment of milk is a relatively inexpensive technology that rounds out flavour and aroma at much lower temperatures than pasteurization. Automated ultrasound systems that can be safely integrated into food production are still being developed.

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SUPPLEMENTARY MATERIALS

All supplementary materials are available at: www.ftb.hr.

AUTHORS’ CONTRIBUTION

Z. Herceg and E. Juraga designed the work. E. Juraga and T. Juraga collected data, while T. Juraga and J. Gajdoš Kljusurić were in charge of data analysis and interpretation. J. Gajdoš Kljusurić and T. Vukušić Pavić performed the data analyses using appropriate tools. T. Vukušić Pavić and Z. Herceg prepared the draft of the article. E. Juraga, M. Brnčić and Z. Herceg conducted the critical revision of the paper, which was sent to all authors for their final approval.

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