Spectrofluorimetric Determination of Oxamniquine in Dosage Forms and Spiked Human Plasma through Derivatization with 1-dimethylaminonaphthalene-5-sulphonyl chloride

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Abstract A sensitive, simple and selective spectrofluorimetric method was developed for the determination of oxamniquine (OXM) in pharmaceutical formulations and biological fluids. The method is based on the reaction between the drug and 1-dimethylaminonaphthalene-5-sulphonyl chloride (dansyl chloride) in presence of 0.5 M sodium carbonate (pH 10) to yield a highly fluorescent derivative that is measured at 445 nm after excitation at 335 nm. The different experimental parameters affecting the development and stability of the reaction product were carefully studied and optimized. The fluorescence concentration plot was rectilinear over the range of 0.02–0.2 μg ml⁻¹ with a lower detection limit (LOD) of 0.007 μg ml⁻¹ and limit of quantitation (LOQ) of 0.02 μg ml⁻¹. The proposed method was successfully applied to the analysis of commercial capsules. The results obtained were in good agreement with those obtained using the official spectrophotometric method. Furthermore, the method was applied for the determination of oxamniquine in spiked human plasma, the mean % recovery (n=4) is 97.77±1.19. A proposal of the reaction pathway was presented.

Keywords Oxamniquine · 1-dimethylaminonaphthalene-5-sulphonyl chloride · Dosage forms · Spiked human plasma

Introduction

Oxamniquine (OXM), 1,2,3,4-tetrahydro-2[(isopro- pylamino methyl]-7-nitro-6-quynoline methanol is an antischistosomal agent that is indicated for the treatment of schistosoma mansoni (intestinal schistosomiasis) infection (Fig. 1). It has been shown to inhibit DNA, RNA and protein synthesis in schistosomes. The oral bioavailability of oxamniquine is good and effective plasma levels are achieved in 1–1.5 h [1].

Oxamniquine is the subject of a monograph in the USP (XXIII) [2] whereby a spectrophotometric method is recommended for its determination, whether in its pure form or in capsules. Several methods have been published for the determination of OXM, either per se or in pharmaceutical preparations and biological fluids. These methods include: spectrophotometry [3–5], non-aqueous titration [6], gas chromatography [7], HPLC [8–13], capillary electrophoresis [11], polarography [14], flow injection analysis [15], cyclic voltammetry [16] and fluorimetry [17].

All these methods are either insufficiently sensitive or tedious and require highly sophisticated and dedicated instrumentation [8–13]. This led us to study the reaction of oxamniquine with dansyl chloride to develop simple and sensitive spectrofluorimetric method for its determination in pharmaceutical preparations and biological fluids. Dansyl chloride is a useful derivatizing agent for primary amines, secondary amines, imidazoles and phenols, etc. Several pharmaceutical compounds have been determined through this approach [18–24].
The reported spectrofluorimetric method for oxamniquine [17] is tedious and time consuming. It involves the use of 2-cyanoacetamide, a hazard reagent.

**Experimental**

**Apparatus**

The spectrofluorimetric measurements were recorded using ARF-1501 Shimadzu Spectrofluorometer, equipped with Xenon arc lamp.

**Materials and reagents**

All reagents and solvents were of analytical reagent grade

- Oxamniquine pure sample was kindly provided by Pfizer (Sandwich, UK).
- Capsules containing 250 mg of Oxamniquine each (Vansil capsules) were obtained from commercial sources in the local market.
- 1-Dimethyl aminonaphthalene-5-sulphonyl chloride (dansyl chloride), purchased from Sigma (St. Louis, USA). A stock solution containing 0.1% of dansyl chloride was freshly prepared in acetone and was further diluted with the same solvent to obtain 0.001% solution.
- Sodium carbonate (BDH, UK) 0.5 M aqueous solution (pH 10).
- Sodium hydroxide (BDH, UK) 1 M aqueous solution.
- Isobutylmethyl ketone (IBMK) (Merck, Darmstadt, Germany).
- Plasma was kindly provided by Mansoura University Hospital, and kept frozen until assay after gentle thawing.
- Methanol and diethyl ether (Merck, Darmstadt, Germany).

**Standard solutions**

A stock solution was prepared by dissolving 20.0 mg of OXM in 20 ml of acetone and 80 ml of 0.5 M sodium carbonate solution. This solution was further diluted with the same solvent mixture as appropriate. The standard solutions were stable for seven days when kept in the refrigerator.

**General procedure**

Aliquots of OXM standard solution were transferred into a series of 10 ml volumetric flasks. 0.7±0.1 ml of 0.001% of dansyl chloride reagent was added, followed by 0.4 ml of acetone and mixed well. The reaction mixture was left for 30 min, and then completed to the mark with IBMK. the reaction mixture was allowed to stand for 10 min. The fluorescence intensity of the reaction product was measured at 445 nm after excitation at 335 nm. Blank experiment was carried out simultaneously. The corrected fluorescence intensity was plotted vs the final drug concentration (μg ml⁻¹) to get the calibration graph. Alternatively, the corresponding regression equation was derived.

**Applications**

**Procedure for commercial capsules**

The content of ten capsules were emptied, and mixed well. A weighed quantity of the powder equivalent to 5.0 mg OXM was transferred into a small conical flask, and extracted with 2×10 ml of methanol. The extract was filtered into 25 ml volumetric flask. The conical flask was washed with several milliliters of methanol. The washing was passed into the same volumetric flask, and the combined extract was evaporated to dryness on a boiling water bath. The residue was dissolved and diluted to volume with a mixture of 5 ml of acetone and 20 ml of 0.5 M of sodium carbonate solution. Aliquots covering the working concentration range (cited in Table 1) was

| Parameter                | Proposed method |
|--------------------------|-----------------|
| Concentration range (μg ml⁻¹) | 0.02–0.2        |
| Minimum detection limit, LOD (μg ml⁻¹) | 0.007            |
| Limit of Quantification, LOQ(μg ml⁻¹) | 0.02            |
| Correlation coefficient (r) | 0.9999          |
| Slope                    | 3,473.114       |
| Intercept                | 0.574           |
| Sᵧ                        | 1.273           |
| Sₓ                        | 7.733           |
| Sᵧ                        | 8.965           |
| % Error                  | 0.28            |

Sᵧ = standard deviation of the residuals
Sₓ = standard deviation of the intercept of regression line
Sᵧ = standard deviation of the slope of regression line
% Error = RSD%√n

![Fig. 1](image-url)
transferred into 10 ml volumetric flasks. The “General procedure” was then applied. The nominal content of the capsules was determined either from the calibration curve or using the corresponding regression equation.

Procedure for spiked human plasma

A stock solution containing 20 μg ml⁻¹ of OXM was prepared. Control samples of plasma was spiked with different quantities of OXM to give a final drug concentration cited in Table 5. One molar NaOH (0.8 ml) was added to 1.0 ml of the spiked plasma and shaken gently. The solution was vortexed with 3×5 ml of diethylether for 2 min, then centrifuged at 2,500 rpm for 5 min. The resulting supernatant was evaporated to dryness under nitrogen at ambient temperature. The residue was dissolved and diluted to volume with a mixture of 5 ml of acetone and 20 ml of 0.5 M of sodium carbonate solution. Aliquots covering the working concentration range was transferred into 10 ml volumetric flasks. The recommended procedures were then applied. The nominal content of the drug was determined using the corresponding regression equation.

Results and discussion

Dansyl chloride was first introduced for the determination of some primary, secondary amines, imidazoles and phenols [18–20].

In recent reports. DNS was further used as a fluorogenic reagent for the determination of some pharmaceutical compounds [21–24].

In the present study, OXM was found to react with Dansyl chloride at pH 10.0 forming a highly yellow fluorescent derivative with λ maximum emission at 445 nm after excitation at 335 nm (Fig. 2).

Optimization of experimental parameters

The spectrofluorimetric properties of the colored product as well as the different experimental parameters affecting the development of the reaction product and its stability were carefully studied and optimized. Such factors were changed individually while the others were kept constant. The factors include pH, concentration of the reagent, type of buffer, temperature, reaction time and dilution time.

Effect of pH

The influence of pH on the fluorescence intensity of the reaction product was studied. Maximum fluorescence intensity was obtained upon using mixture of acetone and 0.5 M sodium carbonate solution. The pH of the reaction mixture was found to be 10.0. pH 10 was found to be the optimum pH for dansylation because labeling of most amino acids, amines, imidazoles and phenols has been found to be optimal at pH 9.5–10.5 [25]. The rate of dansylation process was found to increase with increasing the pH value this is due to an increase in the rate of hydrolysis of dansyl chloride into dansyl hydroxide [25]. The latter shows strong fluorescence and hence interferes seriously in the determination. However, under the proposed chosen conditions and wavelengths used, there was no interference arising from any dansyl hydroxide formed, as indicated by the low fluorescence intensity of the reagent.

Effect of concentration of dansyl chloride

The influence of the concentration of dansyl chloride was studied using different volumes of 0.001% of the reagent solution. It was found that, the reaction of dansyl chloride with OXM started upon using 0.1 ml of the reagent in the presence of sodium carbonate (pH 10.0). Increasing the volume of the reagent, produces a proportional increase in the fluorescence intensity of the reaction product up to 0.6 ml and remains constant up to 0.8 ml. Therefore, 0.7±0.1 ml of 0.001% of dansyl chloride solution was chosen as the optimal volume of the reagent (Fig. 3).

Effect of temperature

Increasing the reaction temperature higher than the room temperature would result in a subsequent decrease in the fluorescence intensity of the reaction product.

Effect of reaction time

Different time intervals were tested to ascertain the time after which the solution attains its highest fluorescence intensity. It was found that after 30 min, the reaction...
Effect of diluting solvent

Different solvents were tried to dilute the reaction mixture throughout the study. It was observed that isobutyl methyl ketone gave the highest fluorescence intensity. Dilution with 0.5 M sodium carbonate solution, water, acetone–water produced almost very week fluorescence and did not reduce the blank fluorescence intensity. While upon using isobutyl methyl ketone, the fluorescence intensity attained its highest value, this was attributed to the low fluorescence value of the reagent.

Effect of dilution time

Dilution times were tested to ascertain the time after which the solution attains its highest fluorescence. It was found that dilution with IBMK after 10 min, the reaction product reaches its highest fluorescence intensity (Fig. 5).

Analytical performance

Validation of the proposed methods The validity of the method was tested regarding; linearity, specificity, accuracy, repeatability and precision according to ICH Q2B recommendations [26].

Linearity

By using the above procedure, linear regression equation was obtained. The regression plots showed that there was a linear dependence of the fluorescence intensity on the concentration of the drug over the ranges cited in Table 1. Linear regression analysis of the data gave the following equation:

\[ F = 0.574 + 3.473.114C \quad (r = 0.9999) \]

where \( F \) is the fluorescence intensity, \( C \) is the concentration of the drug in \( \mu \)g ml\(^{-1} \) and \( r \) is the correlation coefficient.

The limit of quantification (LOQ) was determined by establishing the lowest concentration that can be measured according to ICH Q2B [26]. The results are shown in Table 1. The limits of detection (LOD) were determined by establishing the minimum level at which the analyte can be reliably detected, and the results are also abridged in Table 1.

LOQ and LOD were calculated according to the following equation [26]:

\[ \text{LOQ} = 10\sigma/S \]

\[ \text{LOD} = 3.3\sigma/S \]

Where \( \sigma \): is the standard deviation of the intercept of regression line. \( S \): is the slope of the calibration curve.

The proposed methods were evaluated for the accuracy as percent relative error (% Er) and the precision as percent relative standard deviation (% RSD) (Tables 1 and 2).

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**Fig. 3** Effect of volume of dansyl chloride (0.001%) on the fluorescence intensity of the reaction product of oxamniquine (0.2 \( \mu \)g ml\(^{-1} \)) at pH 10.0

**Fig. 4** Effect of reaction time of on the fluorescence intensity of the reaction product of oxamniquine (0.2 \( \mu \)g ml\(^{-1} \)) with dansyl chloride

**Fig. 5** Effect of dilution time on the fluorescence intensity of the reaction product of oxamniquine (0.2 \( \mu \)g ml\(^{-1} \)) with dansyl chloride
Accuracy

To test the validity of the proposed method it was applied to the determination of authentic sample of OXM over the working concentration range. The results obtained were in good agreement with those obtained using official method. Using Student’s t test and variance ratio F test, [27] revealed no significant difference between the performance of the two methods regarding the accuracy and precision, respectively (Table 2).

The validity of the methods was proved by statistical evaluation of the regression lines, using the standard deviation of the residuals (S_y), the standard deviation of the intercept (S_b) and standard deviation of the slope (S_b). The results are abridged in Table 1. The small values of the figures point out to the low scattering of the points around the calibration line and high precision.

**Table 2** Application of the proposed and official methods to the determination of Oxamniquine in pure form

| Parameters                      | Spectrofluorimetric method | Official method [2] |
|--------------------------------|----------------------------|---------------------|
| No. of experiments             | 6                          | 3                   |
| Mean found (%)                 | 100.13                     | 100.09              |
| ± SD                           | 0.68                       | 0.87                |
| % RSD                          | 0.68                       | 0.87                |
| Variance                       | 0.462                      | 0.76                |
| Student’s t value              | 0.081 (2.31)               |                     |
| Variance ratio F test          | 1.65 (5.41)                |                     |

Figures between parentheses are the tabulated t and F values respectively, at p=0.05 [27]

**Table 3** Validation of the proposed method for the determination of oxamniquine in pure form

| Sample concentration (μg ml⁻¹) | % Recovery (repeatability) | % Recovery intermediate precision |
|--------------------------------|----------------------------|----------------------------------|
| 0.08                           | 99.64                      | 101.08                           |
|                                | 100.72                     | 100.89                           |
| X’                              | 101.44                     | 100.42                           |
| ± SD                           | 0.91                       | 0.99                             |
| % RSD                          | 0.91                       | 0.99                             |
| % Error                       | 0.53                       | 0.57                             |
| 0.2                             | 99.74                      | 101.05                           |
| X’                              | 100.44                     | 100.53                           |
| ± SD                           | 0.53                       | 0.57                             |
| % Error                       | 0.91                       | 0.99                             |

Precision

a. Repeatability

The repeatability was performed by applying the proposed methods for the determination of two concentrations of OXM in pure form on three successive times, and the results are listed in Table 3.

b. Intermediate precision

It was performed through repeated analysis of OXM in pure form, using the concentrations showed in Table 3 for a period of three successive days. The results are summarized in Table 3.

Robustness of the method

The robustness of the method adopted is demonstrated by the constancy of the fluorescence intensity with the deliberated

| Sample                      | Amount added (μg ml⁻¹) | Amount found (μg ml⁻¹) | % Recovery |
|-----------------------------|------------------------|------------------------|------------|
| 1-a-Plasma (inter-day precision) | 0.02                   | 0.01957                | 97.85      |
|                             | 0.04                   | 0.03856                | 96.40      |
|                             | 0.10                   | 0.09754                | 97.54      |
|                             | 0.20                   | 0.19856                | 99.28      |
| Mean                       |                        | 97.77                  | 1.19       |
| 1-b-Plasma (inter-day precision) | 0.20                   | 0.1914                 | 95.70      |
|                             | 0.20                   | 0.1969                 | 98.45      |
|                             | 0.20                   | 0.1948                 | 97.40      |
|                             | 0.20                   | 0.1955                 | 97.75      |
|                             | 0.20                   | 0.1979                 | 98.95      |
| Mean                       |                        | 97.58                  | 1.13       |

Each result is the average of three separate determinations
minor changes in the experimental parameters such as change in the volume of dansyl chloride (0.001%), 0.7 ± 0.1 ml, the change in reaction time 35 ± 5 min and the change in dilution time 15 ± 5 min. These minor changes that may take place during the experimental operation didn’t affect the fluorescence intensity of the reaction product.

Pharmaceutical applications

The proposed methods were then applied to the determination of OXM in its capsules. The methods were tested for linearity, specificity, accuracy, repeatability and precision according to ICH Q2B recommendations.

Specificity

The specificity of the method was investigated by observing any interference encountered from the common capsule excipients, such as talc, lactose, starch, avisil, gelatine, and magnesium stearate. These excipients did not interfere with the proposed method.

Accuracy

The results of the proposed methods were statistically compared with those obtained using the official method [2]. Statistical analysis [27] of the results, using Student’s t test and variance ratio F test revealed no significant difference between the performance of the proposed and reference methods regarding the accuracy and precision, respectively (Table 4).

Analysis of biological fluid

The high sensitivity of the proposed method allowed the determination of OXM in spiked human plasma. Oxamniquine is readily absorbed following oral ingestion, and a peak concentration in plasma occurs within about 3 h. The presence of food significantly delays absorption and limits the concentration achieved in plasma during the first several hours after administration. Urinary excretion is the major route of elimination in man [28]. Oxamniquine is given orally in a dose of 250 mg three times daily; this leads to a final blood level concentration of about 5 μg ml⁻¹ i.e. higher than the upper limit of the working concentration range of the proposed method. The high sensitivity of the proposed method allowed the determination of OXM in spiked human plasma. The results are shown in Table 5.

The extraction procedure described by Woolhouse and Wood [7] was adopted here. The results are satisfactorily accurate and precise.

Scheme 1 Proposed reaction pathway between dansyl chloride and oxamniquine
Precision The within-day precision was evaluated through replicate analysis of Plasma samples spiked with different concentrations of the drug. The percentage recoveries based on the average of four separate determinations were 97.77 ± 1.19, thus indicating the high precision of the method (Table 5).

The inter-day precision was also evaluated through replicate analysis of plasma samples spiked with 0.2 μg ml⁻¹ of drug on four successive days. The percentage recoveries based on the average of four separate determinations were 97.58 ± 1.13. The results are shown in Table 5.

Mechanism of the reaction

The stoichiometry of the reaction was studied adopting the limiting logarithmic method [29]. The fluorescence intensity of the reaction product was alternatively measured in the presence of excess of either dansyl chloride or OXM. A plot of log fluorescence versus log [dansyl chloride] and log [OXM] gave straight lines, the values of the slopes are 0.901 and 0.984 respectively (Fig. 6). Hence, it is concluded that, the molar reactivity of the reaction is 0.901/0.984, i.e. the reaction proceeds in the ratio of 1:1. Based on the observed molar reactivity of the reaction, and depending on the presence of secondary amino group and by analogy to similar reports dealing with the reaction of dansyl chloride with compounds containing secondary amino group, the reaction pathway proposed in Scheme 1 is presented.

Conclusion

The proposed method has the advantage of being simple, sensitive and suitable for routine analysis in quality control laboratory. Also, it is suitable for the determination of oxamniquine in spiked human plasma with minimum detection limit lower than the reported value. In addition, it could be applied to the determination of OXM in its pharmaceutical preparation.

References

1. Delgado JN, Remers WA (Eds) (1991) Wilson and Gisvold’s text book of organic medicinal and pharmaceutical chemistry, 9th edn, p 184
2. The USP Pharmacopoeia XXIII (1995) NF 18, The US Pharmaceutical Convention, Rockville, p.1120.
3. Hassan SM, Belal F, Sharif-El-Din M, Sultan M (1988) Spectrophotometric determination of some pharmaceutically nitro compounds in their dosage forms. Analyst 113:1807–1809
4. Bebawy LI, El-Kelani K, Abdel Fattah L, Ahmed AK (1997) Study of 7,7′,8,8′-tetracyanoquinodimethane charge transfer complexes with some lone-pair-donating drugs. J Pharm Sci 86:1030–1033
5. Rizk M, Belal F, Ibrahim F, Ahmed SM, El-Enany NM (2000) A simple kinetic spectrophotometric method for the determination of oxamniquine in formulations and spiked biological fluids. J Pharm Biomed Anal 23:503–513
6. Korolkovas A, Haraguchi T (1980) Rev. Farm. Bioquim Univ. Sao Paulo, 16, 12. Tho. Chem. Abstr. vol 94, pp 7846e, 1981
7. Woolhouse NM, Wood PR (1977) Determination of oxamniquine in serum. J Pharm Sci 66:429–430
8. Jun HW, Radwan MA (1985) Anal Lett 18:1345–1355
9. Pierr E, Almeida AE, Gremiao MP (2001) Determination of oxamniquine in capsules by HPLC. J Pharm Biomed Anal 26:675–679
10. Masimirembwa CM, Hasler JA, Johansson I (1995), Inhibitory effects of antiparasitic drugs on cytochrome P450 2D6. Eur J Clin Pharmacol 48:35–38
11. Abushooffa MA, Clark BJ (1995) Resolution of the enantiomers of oxamniquine by capillary electrophoresis and high-performance liquid chromatography with cyclodextrins and heparin as chiral selectors. J Chromatogr 700:51–58
12. Noctor TA, Fell AF, Kaye B (1990), High-performance liquid chromatographic resolution of oxamniquine enantiomers: application to in vitro metabolism studies. Chirality 2:269–274
13. Fell AF, Noctor TA, Mama JE, Clark BJ (1988) Computer-aided optimisation of drug enantiomer separation in chiral high-performance liquid chromatography. J Chromatogr 434:377–384
14. Belal F, Aly FA (1995) Polarographic behavior and determination of oxamniquine in dosage forms. Electroanalysis 7:483–487
15. Mohamed MY, El-Gendy AE, El-Bardicy MG, Tawakol MS, Ahmed AKS (1996) Spectrosc Lett 29:299–319
16. Radi A, Belal F (1998) Reaction of electrogenerated oxamniquine radical anion with glutathione. J Electroanal Chem 441:39–42
17. Rizk M, Belal F, Ibrahim F, Ahmed SM, El-Enany NM (1999) Fluorimetric determination of oxamniquine in biological fluids. II Farmaco 54:47–50
18. Ayad MM (1984) Spectrofluorimetric microdetermination of imidazoline derivatives using 1-dimethylaminonaphthaline-5-sulphonylchloride. Analyst 109:431–1434
19. Putter J (1979) A fluorometric method for the determination of praziquantel in blood–plasma and urine. Eur J Drug Metab Pharmacokinet 4:143–148
20. Frei-Hausler M, Frei RW (1973) An investigation of fluorogenic labelling of chlorophenols with dansyl chloride. J Chromatogr 84:214–220
21. Lucca A, Gentilini G, Lopez-Silva S, Soldanini A (2000) Simultaneous determination of human plasma levels of four selective serotonin reuptake inhibitors By HPLC. Ther Drug Monit 22:271–276
22. Dennis G, Mear J, Charles B, El-Din M, Sultan M (1988) Spectrofluorimetric determination of some pharmaceutically nitro compounds in their dosage forms. Analyst 113:1807–1809
23. Bebawy LI, El-Kelani K, Abdel Fattah L, Ahmed AK (1997) Study of 7,7′,8,8′-tetracyanoquinodimethane charge transfer complexes with some lone-pair-donating drugs. J Pharm Sci 86:1030–1033
24. Bagdonaitė K, Viklund G, Skog K, Kovic M (2006) Analysis of 3 Amino-propionamide: a potential precursor of acrylamide. J Biochem Biophys Methods 69:421–423
25. Rose J (1964) Advanced physico-chemical experiments. Pitman, London