Case Report: A Unique Case of Pediatric Central Nervous System Embryonal Tumor Harboring the CIC–LEUTX Fusion, Germline NBN Variant and Somatic TSC2 Mutation: Expanding the Spectrum of CIC-Rearranged Neoplasia

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Central nervous system (CNS) embryonal tumors (WHO grade IV) are a heterogeneous group of rare, poorly differentiated neuroepithelial malignant neoplasms that commonly occur in children, and they have a poor prognosis. The 2016 WHO (World Health Organization) classification of CNS tumors created a major shift in paradigm of the classification of embryonal tumors. However, some cases were still difficult to classify. Further integrative genomic analysis is needed to improve the precise classification, diagnosis and treatment of CNS embryonal tumors. Herein, we firstly report a case of CNS embryonal tumor harboring the pathogenic CIC–LEUTX gene fusion. A 2-year-old male infant presented with a solid cystic mass in the left temporal lobe-basal ganglia and left parietal lobe (maximum diameter, 75 mm) and underwent gross tumor resection. The tumor was classified as a poorly differentiated embryonal neoplasm of neuroectodermal origin that lacked specific features and rosettes. By immunohistochemistry, the tumor cells were strongly positive for synaptophysin, and the Ki67 proliferation index was high (>50%). FISH (Fluorescence in situ hybridization) results indicated no change in the copy number at the 19q13.42 C19MC locus. Next generation sequencing showed a CIC–LEUTX gene fusion, a somatic TSC2 c.G2714A mutation, and a heterozygous germline NBN c.C127T mutation. One month after surgery, there was recurrence of the intracranial tumor (maximum diameter, 55 mm) as well as spinal cord implantation metastasis. The patient received chemotherapy (CTX+CBP+VCR/DDP+VP-16), radiotherapy, and a drug targeting the TSC2 gene (everolimus). At the time of this writing, the patient is alive without evidence of disease for 11 months. This is the first report of the CIC–LEUTX gene fusion in a case of CNS embryonal tumor.

Keywords: CIC, LEUTX, next generation sequencing, central nervous system embryonal tumor, TSC2, NBN
INTRODUCTION

Central nervous system (CNS) embryonal tumors are rare, poorly differentiated neuroepithelial malignant neoplasms that commonly occur in children, including a wide range of aggressive malignancies. The most common clinical manifestations are symptoms and signs of increased intracranial pressure. However, the overlapping morphological features of these lesions present a diagnostic challenge and undermine the discovery of optimal treatment strategies. A lot of supratentorial CNS embryonal tumors have been histologically classified as primitive neuroectodermal tumor (PNET) in the past, and can have histological features overlapping those of other brain tumors. In recent years, with the advancement of genome technology, the molecular heterogeneity of these tumors is gradually being understood. Some of CNS embryonal tumors have been reclassified from this group through the identification of unique molecular biomarkers, such as embryonal tumors with abundant neurips and true rosettes, which exhibit alterations in the 19q13.42 C19MC locus, and they are now considered to be a distinct genetically-defined entity known as the embryonal tumor with multilayered rosettes (ETMR), C19MC-Altered. Another entity has also been identified; embryonal tumors with multilayered rosettes (ETMR), DICER1-altered, have mutually exclusive biallelic DICER1 mutations in which the first hit is typically inherited through the germline (1). However, several CNS embryonal tumors remain unclassified and are presently defined as CNS embryonal tumors, NOS (not otherwise specified)/NEC (not elsewhere classified). The pathogenesis of these tumors is still unclear. Here, we firstly report the presence of the CIC–LEUTX gene fusion in a case of CNS embryonal tumor.

Case Presentation

A 2-year-old male infant presented with vomiting and drowsiness lasting for 2 months. On examination, the muscle strength of the right upper limb was grade II and lips skewed. This patient's pregnancy was of normal length, and his delivery was uncomplicated; his birth weight was 2600g, height 50cm, and his Apgar score is 9. Brain magnetic resonance imaging (MRI) scans revealed a solid cystic mass with lesions in the left temporal lobe–basal ganglia-corona radiate, midline shifted to the right, and nodules in the left parietal lobe (maximum diameter, 75 mm) (Figures 1A, B). The patient underwent gross resection of the lesions. Microscopical analysis revealed that the tumor was a poorly differentiated embryonal neoplasm of neuroectodermal origin lacking specific histopathological features. The tumor was comprised of diffuse undifferentiated embryonal cells with neurips, but no true rosettes (Figures 1C, D). Some tumor cells were small, round, and poorly differentiated, whereas other cells presented with atypia as well as rough and dark chromatin. Mitotic figures and apoptotic bodies were easily identified. By immunohistochemistry, cells were negative for glial fibrillary acidic protein (GFAP) and strongly positive for synaptophysin (Syn) (Figure 1E). The Ki67 proliferation index was high (>50%) (Figure 1F), and fluorescence in situ hybridization (FISH) results showed no change in the copy number at the 19q13.42 C19MC locus (Figures 1G, H). The final diagnosis was CNS embryonal tumor, not otherwise classified (NEC), according to the guidelines of the revised 2016 World Health Organization Classification of Tumors of the Central Nervous System and cIMPACT-NOW workgroup (2).

In order to integrate histological and molecular diagnosis, and seek candidate treatments, comprehensive genomic profiling was performed using a 539 cancer related genes panel based on next generation sequencing (NGS) via DNA-based hybrid capture in formalin-fixed and paraffin-embedded primary tumor tissue. The panel was sequenced with an average number of mapping reads as 2 megabase and high median depth (> 4000×). We found a fusion of CIC–LEUTX genes with breakpoints located in exon 20 of the CIC gene and exon 3 of the LEUTX gene (Figures 2A, B). In addition, a somatic mutation in the TSC2 gene (Figure 2C) and a germline mutation in the NBN gene (Figure 2D) were also identified. Copy number variation (CNV) showed the CIC gene to be amplified 4.85-fold on chromosome 19 (Figures 2E, F). TSC2 encoded tuberin, which was a GTPase-activating protein and regulator of mTORC1 by modulating brain-enriched Rhed (3). The somatic variant c.G2714A located on tuberin-type domain of TSC2, a critical well-established functional domain, and evaluated as pathogenic mutation by public database ClinVar. The mutation resulted in loss of function of TSC2 led to the activation of mTORC1, which regulated the cell growth and survival (4). The patient may benefit from the mTOR inhibitor RAD001 (everolimus), which has been proved an ability to penetrate the blood-brain barrier (5). Further studies by Sanger sequencing revealed that the patient's father, and not his mother, was a carrier of the NBN heterozygous mutation (Figures 2G, H). This patient's parents and elder sister are healthy. However, his grandfather died of leukemia, and his grandfather diagnosed with bladder cancer.

The patient presented with weakness of the right upper limb five weeks after surgery. Plain and enhanced MRI scans of the brain revealed a partial deletion in the left parietal lobe, the presence of multiple nodules and lesions (maximum diameter, 55 mm) near the surgical site, and left basal ganglia and left basis craniai, indicative of disease recurrence. The left forehead, bilateral temporal region, suprasellar area, front portion of the pons, and pia mater of the bilateral cerebellar hemisphere were unevenly thickened, suggesting implantation metastasis. Plain and enhanced MRI scans of the cervical vertebra revealed lumbo-vertebral canal subdural nodules, slightly thickened terminale meninges, and suspicious lesions that appeared to have spread via the cerebrospinal fluid. The patient received chemotheraphy (cyclophosphamide+carboplatin +vincristine/cisplatin+etoposide), an mTOR inhibitor (everolimus), and radiotherapy. The tumor shrunken significantly.

Abbreviations: CNS, central nervous system; PNET, primitive neuroectodermal tumor; IHC, immunohistochemistry; FISH, fluorescence in situ hybridization; NGS, next generation sequencing; CNV, copy number variation; NEC, not elsewhere classified; ETMR, embryonal tumor with multilayered rosettes; MRI, magnetic resonance imaging; GFAP, glial fibrillary acidic protein; Syn, synaptophysin; FFPE, formalin-fixed and paraffin-embedded; GBM, glioblastoma; N/A, not applicable; GRT, gross total resection; RT, radiotherapy; TMZ, temozolomide; STR, subtotal resection; CSI, craniopinal irradiation; PD, progression disease; doxo, doxorubicin; ifos, ifosfamide; CTX, cyclophosphamide; CBP, carboplatin; VCR, vincristine; DDP, cisplatin; VP-16, etoposide; HAT, histone acetyltransferase.
FIGURE 1 | Pre-operative T1-weighted (A) and T2-weighted (B) magnetic resonance images showing a solid cystic mass with heterogeneous hyperintensity and surrounding edema. Histologically, the tumor was comprised of poorly differentiated/undifferentiated cells with neuropils and proliferated endothelial cells, but no typical true rosettes (C). Some tumor cells were small, round, and poorly-differentiated, whereas some cells presented with atypia as well as rough and dark chromatin. Mitotic figures and apoptotic bodies were easily identified under HPF ×400 (D). Immunohistochemically, the tumor cells were synaptophysin positive (E), and the Ki67 index was >50% (F). The C19MC locus showed no alteration (G) compared with the positive amplification positive control (H).
FIGURE 2 | Next generation sequencing (NGS) revealing (A) a CIC–LEUTX gene fusion in exon 20 of the CIC gene and exon 3 of the LEUTX gene, (B) a germline NBN c.C127T variant, and (C) a somatic TSC2 c.G2714A variant (D). (E) Copy number variation (CNV) of the CIC gene was highest in the whole genome. (F) The CIC gene was amplified 4.85-fold on chromosome 19. (G) The unaffected father of the patient carried the heterozygous NBN c.C127T mutation, (H) whereas the healthy mother did not.

TABLE 1 | Clinical, histological features and treatment of the tumor with the CIC–LEUTX gene fusion in central nervous system.

| No | Age | Sex | Histologic diagnosis | Molecular changes | Location | Therapy | Follow-up (Month) | Status |
|----|-----|-----|----------------------|-------------------|----------|---------|------------------|--------|
| 1(8) | 43 | F | angiosarcoma | CIC-LEUTX and TSC2 | left tentorial extension to the left midbrain and thalamus | N.A | N.A | N.A |
| 2(7) | 12 | F | anaplastic astrocytoma with epithelioid GBM features | CIC-LEUTX | Intraventricular | GTR + RT/TMZ | 3 | Alive, on therapy |
| 3(7) | 19 | F | anaplastic ganglioglioma | CIC-LEUTX | Frontoparietal | GTR + TMZ → STR + RT (CSI) + carboplatin → cytoxin/ doxa/fos/ etoposide | 10 | Progression |
| 4 | 2 | M | CNS embryonal tumor | CIC-LEUTX, TSC2 and NBN | left temporal lobe-basal ganglia and left parietal lobe | PR | 1 | Progression |

GBM, glioblastomas; CNS, central nervous system; NA, not applicable; GTR, gross total resection; RT, radiotherapy; TMZ, temozolomide; STR, subtotal resection; CSI, craniospinal irradiation; PD, progression disease; doxo, doxorubicin; fos, ifosfamide; CTX, cyclophosphamide; CBP, carboplatin; VCR, vincristine; DDP, cisplatin; VP-16, etoposide.
with clinical symptoms relief after therapy. At the time of this writing, the patient was still being treated without any clinical symptoms.

**DISCUSSION AND CONCLUSION**

CNS embryonal tumors are aggressive, poorly differentiated brain tumors with poor prognosis and limited effective therapies. Although new subtypes, such as embryonal tumors with multilayered rosettes (ETMR), C19MC altered, and embryonal tumors with multilayered rosettes (ETMR), Dicer1-altered, have been described, there are few reported cases of new molecular changes in CNS embryonal tumors. In 2016, Dominik Sturm et al. (6) identified several CNS embryonal tumors designated as CNS neuroblastoma with FOXR2 activation (CNS NB–FOXR2), CNS Ewing sarcoma family tumor with CIC alteration (CNS EFT–CIC), CNS high-grade neuroepithelial tumor with MN1 alteration (CNS HGNET–MN1), and CNS high-grade neuroepithelial tumor with BCOR alteration (CNS HGNET–BCOR). However, a CIC–NUTM1 gene fusion or a CIC frameshift deletion, not a CIC–LEUTX gene fusion and a CIC amplification, were detected in CNS EFT–CIC. In review of the literature, we found only three cases of the CIC–LEUTX gene fusion were reported in CNS, namely a case of CNS angiosarcoma, a case of anaplastic ganglioglioma, and a case of anaplastic astrocytoma with epithelioid GBM features (7, 8) (Table 1), and this is the first case of CIC–LEUTX gene fusion in a CNS embryonal tumor.

We think CNS tumors harboring the CIC–LEUTX gene fusion may be a new entity. It should be noted that the histological features of these four cases were totally different; however, the gene fusion was identical. Interestingly, both our case and the case of CNS angiosarcoma also harbored the TSC2 gene mutation apart from CIC–LEUTX gene fusion, and our targeted therapy was effective. However, it is not known whether the other two cases had similar mutations. The TSC2 protein participates in the mTOR/ AKT pathway, and mutations in its gene commonly cause the TSCs.

In addition, this is the first report of the CIC–LEUTX gene fusion in CNS embryonal tumors, which expands the spectrum of CIC–rearranged neoplasia. CIC rearrangement predominantly occurs in small round cell sarcomas that present as Ewing like sarcomas. For instance, CIC–DUX4 gene fusion sarcomas, a new molecular entity recently described in the 2019 WHO Classification of Tumors of Soft Tissue and Bone, are different from Ewing sarcomas in that more aggressive treatment is needed to improve the overall survival. CIC–DUX4 gene fusion sarcomas account for 22–68% in none EWSR1–rearranged small round cell sarcomas (10, 11). In addition to CIC–DUX4 gene fusion Ewing-like sarcomas and CIC–LEUTX gene fusion in angiosarcomas, other genes, such as NUTM1, NUTM2A, and FOXO4, were also involved in CIC gene rearrangement sarcomas (Table S1). However, it is not known whether these genes can affect the prognosis of cancer patients harboring CIC fusions (12), and further studies are needed.

We speculate that one of the reasons for the CIC–LEUTX gene fusion in this patient was the fragmentation and instability of the CIC chromosome caused by the NBN gene mutation. The NBN gene is responsible for double-strand DNA damage repair, and heterozygous mutation carriers are at risk for several types of tumors such as CNS relapse of B-cell precursor acute lymphoblastic leukemia (13). Mutations in the NBN gene lead to DNA breakage, with abnormal meiosis causing the presence of several CNVs, including the CIC gene. Therefore, high-risk populations should be prenatally screened for pathogenic mutations.

Germline variant in NBN increases the risk of cancer, however, only one germline mutation in a single allele is not sufficient to induce the tumors (14). The CIC–LEUTX fusion appears to be the oncogenic alteration. CIC gene encodes a transcriptional repressor protein which regulates kinase signaling (10). The hypothesis of the molecular mechanism of oncogenic CIC–NUTM1 fusion in CNS sarcomas is well characterized, a n d we are involved in CIC–LEUTX gene fusion sarcomas account for 22–68% in none EWSR1–rearranged small round cell sarcomas (10, 11). In addition to CIC–DUX4 gene fusion Ewing-like sarcomas and CIC–LEUTX gene fusion in angiosarcomas, other genes, such as NUTM1, NUTM2A, and FOXO4, were also involved in CIC gene rearrangement sarcomas (Table S1). However, it is not known whether these genes can affect the prognosis of cancer patients harboring CIC fusions (12), and further studies are needed.

**DATA AVAILABILITY STATEMENT**

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.
ETHICS STATEMENT

Written informed consent was obtained from the individual(s), and minor(s)’ legal guardian/next of kin, for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

WH, LY, and JZ identified and diagnosed the reported case. XZ, YJ, and CS performed NGS analysis. and WH, JW, JZ, and XS wrote and revised the manuscript with input from all authors. All authors contributed to the article and approved the submitted version.

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Conflict of Interest: XZ, YJ, and CS were employed by Jiangsu Simcere Diagnostics Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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