Brain Exposure to Piperacillin in Acute Hemorrhagic Stroke Patients Assessed by Cerebral Microdialysis and Population Pharmacokinetics

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Abstract

Background: The broad antibacterial spectrum of piperacillin/tazobactam makes the combination suitable for the treatment of nosocomial bacterial central nervous system (CNS) infections. As limited data are available regarding piperacillin CNS exposure in patients without or with low-grade inflammation, a clinical study was conducted (1) to quantify CNS exposure of piperacillin by cerebral microdialysis and (2) to evaluate different dosing regimens in order to improve probability of target attainment (PTA) in brain.

Methods: Ten acute hemorrhagic stroke patients (subarachnoid hemorrhage, n = 6; intracerebral hemorrhage, n = 4) undergoing multimodality neuromonitoring received 4 g piperacillin/0.5 g tazobactam every 8 h by 30-min infusions for the management of healthcare-associated pneumonia. Cerebral microdialysis was performed as part of the clinical neuromonitoring routine, and brain interstitial fluid samples were retrospectively analyzed for piperacillin concentrations after the first and after multiple doses for at least 5 days and quantified by high-performance liquid chromatography. Population pharmacokinetic modeling and Monte Carlo simulations with various doses and types of infusions were performed to predict exposure. A $T_{>\text{MIC}}$ of 50% was selected as pharmacokinetic/pharmacodynamic target parameter.

Results: Median peak concentrations of unbound piperacillin in brain interstitial space fluid were 1.16 (range 0.08–3.59) and 2.78 (range 0.47–7.53) mg/L after the first dose and multiple doses, respectively. A one-compartment model with a transit compartment and a lag time (for the first dose) between systemic and brain exposure was appropriate to describe the brain concentrations. Bootstrap median estimates of the parameters were: transfer rate from plasma to brain (0.32 h$^{-1}$), transfer rate from brain to plasma (7.31 h$^{-1}$), and lag time [2.70 h (coefficient of variation 19.7%)]. The simulations suggested that PTA would exceed 90% for minimum inhibitory concentrations (MICs) up to 0.5 mg/L and 1 mg/L at a dose of 12–16 and 24 g/day, respectively, regardless of type of infusion. For higher MICs, PTA dropped significantly.

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Introduction
The broad antibacterial spectrum of the piperacillin/tazobactam combination would possibly qualify the combination for the treatment of nosocomial bacterial central nervous system (CNS) infections [1]. Achievement of concentrations suitable for treatment has been reported in cerebrospinal fluid (CSF) of meningitis patients [2] at a daily dose of 324–436 mg/kg body weight. However, patients without generalized meningeal inflammation, e.g., those with hydrocephalus, achieved insufficient concentrations in CSF [1]. Though CSF concentrations may sometimes reflect brain target site concentrations well, drug distribution might be different between CSF and extracellular fluid (ECF) of the brain [3] and may differ manifold between lumbar, ventricular, and cisternal parts of the compartment [4, 5]. Conversely, ECF concentrations measured through microdialysis reflect target site concentrations well as it only measures free (thus biologically active) concentrations. In the last two decades, the use of microdialysis has evolved quickly and is now an integral part of individualized intensive care therapy of acute injury patients undergoing multimodal monitoring in several centers [6]. Limited data are available regarding CNS exposure and related target attainment of antibiotics in patients with acute brain injuries.

Like other beta-lactam antibiotics, piperacillin exhibits time-dependent bacterial killing, with T > MIC being the relevant pharmacokinetic/pharmacodynamic parameter [7]. Cefepime has been reported to show better exposure in terms of T > MIC in plasma as well as in CSF when administered as a continuous infusion in contrast to intermittent infusion in neurosurgical patients with postoperative intracranial infections [8]. Piperacillin has also been reported to achieve better plasma exposure with continuous infusion or extended infusion as compared to intermittent infusion [9–12]. However, it is not yet known whether prolonged infusion can improve CNS exposure of piperacillin in brain.

Therefore, the objectives of this study were to quantify brain ECF concentrations of piperacillin using cerebral microdialysis and to develop a model to describe the kinetics of its brain exposure in patients without general meningeal inflammation. Subsequently, this model was used to predict the probability of target attainment (PTA) of different doses, types of infusions, and minimum inhibitory concentration (MIC) values in order to assess the pharmacokinetic fundament for a potential use of piperacillin in such patients.

Methods
Patient Selection and Ethical Approval
Ten comatose patients (median [range] age and body weight of 52 [32–72] years and 73 [60–95] kg) with acute hemorrhagic stroke (subarachnoid hemorrhage [SAH], n = 6; intracerebral hemorrhage [ICH], n = 4) admitted to the neurological intensive care unit (ICU) of the Department of Neurology at the Medical University of Innsbruck, Austria, requiring invasive multimodal neuromonitoring were recruited between October 2010 and November 2014 (Table 1). Patients were eligible if they developed healthcare-associated pneumonia and if antibacterial treatment with piperacillin/tazobactam was indicated. Sample size calculation was not performed prior to the study. The conduct of this study was approved by the Ethics Committee of the Medical University of Innsbruck (Approval Numbers AN3898 285/4.8, AM4091-292/4.6, UN3898 285/4.8) and registered with the institutional Clinical Trial Center (https://ctc.tirol-kliniken.at; study identifier 20131218-868). Additionally, the public at the research site was informed about the study by notice on the bulletin board at the neurological ICU. All provisions of the WMA Declaration of Helsinki in its applicable version were followed, and informed consent was obtained from all patients or legal representatives according to federal regulations. Clinical care of SAH and ICH patients strictly adhered to current international guidelines [13–15] with the exception of nimodipine being administered intravenously in SAH patients.

Data Collection, Neuromonitoring, and Sampling Procedures
Patient characteristics, medical complications, and outcome were prospectively recorded in the respective institutional SAH and ICH databases. In line with international consensus, patients underwent intracranial neuromonitoring including measurement of intracranial pressure, brain tissue oxygen tension, and cerebral metabolites based on clinical and radiological criteria [16]. A cerebral microdialysis catheter (71 High Cut-Off
Brain Microdialysis Catheter, M Dialysis AB, Stockholm, Sweden) was tunneled and inserted into the white matter either “perilesionally” (i.e., placement of the catheter gold tip within 1 cm to a focal brain lesion) or otherwise into “normal-appearing tissue.” Isotonic perfusion fluid (Perfusion Fluid CNS; M Dialysis AB) was pumped through the microdialysis system at a flow rate of 0.3 µL/min. Hourly samples were analyzed with ISCUSflex Point-of-Care Analyzer (M Dialysis AB) for interstitial glucose, pyruvate, lactate, and glutamate concentrations and frozen thereafter at −80 °C. During the neuromonitoring period, all patients were intubated and mechanically ventilated. To facilitate mechanical ventilation, patients received continuous infusions of midazolam plus sufentanil and/or S-ketamine.

Immediately after diagnosis of healthcare-associated pneumonia, treatment was initiated according to local clinical infectious diseases guidelines with a 30-min intravenous infusion of 4 g piperacillin/0.5 g tazobactam (Fresenius Kabi, Graz, Austria) every 8 h. As stated, microdialysis samples of brain interstitial fluid were obtained in 1 h intervals both after first dose and after multiple doses at steady state (median of seven samples each). Importantly, cerebral microdialysis was performed as part of the clinical neuromonitoring routine and the cerebral microdialysis catheter remained in situ for the total neuromonitoring period, usually exceeding the span of piperacillin/tazobactam administration.

**Determination of In Vitro Recovery**

Determination of recovery in vitro was performed using identical probes, flow rate, and perfusion fluid as in patients. In forward µD experiments, the microdialysis catheter was placed into the immersion solution containing piperacillin/tazobactam (1/0.125 [C1], 10/1.25 [C2], or 100/12.5 mg/L [C3]) and was constantly perfused with Perfusion Fluid CNS at a flow rate of 0.3 µL/min. In reverse µD experiments, immersion solutions contained plain Perfusion Fluid CNS, whereas the perfusion solution contained piperacillin/tazobactam. In both forward and reverse µD experiments, µD samples were collected over three consecutive sampling intervals of 60 min for two different probes.

**Analytical Assay**

Due to the low flow rate (0.3 µL/min) used during clinical microdialysis, a sample volume of 18 µL for each hourly sample was obtained out of which approximately 6 µL was required for diagnostic purposes. The remaining sample volume was not sufficient for two analyses, and thus, tazobactam could not be quantified. Piperacillin concentrations were determined by an isocratic high-performance liquid chromatography (HPLC) method with ultraviolet detection at 225 nm which has been validated according to the U.S. Food and Drug Administration (FDA) [17] and European Medicines Agency (EMA) [18] recommendations. Quality control samples of appropriate concentrations prepared in Ringer’s solution were analyzed with each assay. The coefficient of variation in intra- and inter-assay precision and accuracy was < 3% based on in-process quality controls (80, 8, and 0.8 mg/L). For liquid chromatography, an XBridge C18 BEH 2.5µ, 50 × 3 mm column with an Xcelery RP18 2.7µ, 4 × 3 mm guard column was used as the stationary phase. Isocratic elution was carried out with 0.1 M H₃PO₄, pH 2.7/acetonitrile 75:25 (v/v) using a flow rate of 0.4 mL/min. The lower limit of quantification (LLOQ) was 0.05 mg/L. Undiluted cerebral microdialysis samples were injected directly into the HPLC system, with a defined injection volume of 1–3 µL selected according to the expected concentration.

**Population Pharmacokinetic Analysis**

The NONMEM software (version 7.4.3) [19] was used to develop a population pharmacokinetic (PopPK) model of piperacillin. Estimations were performed using first-order conditional estimation with interaction. Different models were assessed based on improvement in objective function value (drop of ≥ 3.84 corresponding to p < 0.05 with one degree of freedom, assuming a Chi-squared distribution), goodness-of-fit plots, and precision of parameter estimates from nonparametric bootstrap analysis of 1000 samples [20]. Visual predictive checks [21] were performed to assess the predictive performance of the models. Different models with one, two, and three compartments with or without lag time (Tlag) and various number of transit compartments were tested to fit the brain ECF data.

Because plasma piperacillin concentrations were not available, a plasma model with similar study design and demographics was selected from the literature [10] to drive the brain concentrations in our model. Fixed effects parameters as well as inter-individual and intra-individual variability parameters were fixed in the model.

Dead space of the catheter (distance between semi-permeable membrane of the probe and catheter outlet) was 5.1 µL, and thus, it would take approximately 0.28 h for the fluid to reach the outlet based on the flow rate (0.3 µL/min) used in the study. To account for the dead space, this time was subtracted from the end time of the microdialysis time (1-h interval) beforehand in the dataset. Microdialysis recovery was assumed to be 100% in the model based on the results of in vitro experiments. Piperacillin concentrations below LLOQ were retained in the dataset and evaluated with M3 and M5 methods [22].
The integrated approach [23] (numerical integration to calculate average concentrations during the corresponding microdialysis intervals) was used to model the brain data. Plasma protein binding was considered to be linear and was fixed to 30% according to the plasma pharmacokinetic model [10]. Inter-individual variability was estimated while assuming a log-normal distribution of parameters. Additive, proportional, and combined error models of residual unexplained variability (RUV) were evaluated on the model.

Monte Carlo Simulations and Probability of Target Attainment (PTA)

Monte Carlo Simulations (MCS) (5000 simulated subjects) were performed based on the final PopPK model of piperacillin at various MIC levels (up to 16 mg/L), for three doses (12, 16, and 24 g/day) and three types of infusion (intermittent infusion over 30 min, extended infusion over 3 or 4 h, and continuous infusion). Based on plasma data, a pharmacokinetic/pharmacodynamic (PK/PD) index of $f_{T>MIC}$ of 50% is considered to be essential for the optimal activity of piperacillin [24]. Therefore, $f_{T>MIC}$ of 50% was selected as PK/PD index for the assessment of probability of target attainment against various pathogens according to the MIC distribution suggested by the European Committee on Antimicrobial Susceptibility Testing (EUCAST) for susceptible pathogens [25]. Minimum inhibitory concentration required to inhibit the growth of 50% (MIC$_{50}$) and 90% (MIC$_{90}$) of bacteria commonly involved in CNS infections was derived from MIC distribution of wild-type microorganisms on EUCAST Web site (Table 2) [25].

Sensitivity Analysis

An additional evaluation (sensitivity analysis) was performed to avoid that the selected plasma model would bias simulated PTA in brain. To this end, the plasma model used was replaced by two other published plasma models [26, 27]. New parameters for the brain data were estimated, and MCS were done to assess PTA in brain as described above. These PTAs were then compared with those predicted in the previous simulations.

Results

A total of 130 piperacillin brain concentrations from ten acute hemorrhagic stroke patients were available for the development of the PopPK model (median 0.95 mg/L [range 0.0–7.53]). Median peak concentrations of unbound piperacillin in brain interstitial space fluid after the first dose were 1.16 (range 0.08–3.59) mg/L. After

| Parameter | Our study | Published plasma model used to develop the brain PK model$^{[1]}$ |
|-----------|-----------|---------------------------------------------------------------|
| Gender (male/female) (n) | 2/8 | 5/3 |
| Weight (kg) | 73 (57–88) | 80 (74–86) |
| Age (years) | 52 (32–71) | 41 (22–65) |
| Height (cm) | 171 (166–176) | 174 (172–180) |
| Piperacillin dose (mg day/kg) | 166 | 229 (204–254) |
| Creatinine clearance (Cockcroft–Gault) (mL/min) | 101.5 (63.5–139.5) | 88.3 (53.3–101.0) |
| APACHE II score on day 1 | 26 (24–31) | 24 (18–26) |
| Stay in intensive care unit (days) | 34 (18–50) | – |

All values are given as median (inter-quartile range) except gender.

APACHE Acute physiology and chronic health evaluation

| Pathogen | MIC$_{50}$ | MIC$_{90}$ |
|-----------|------------|------------|
| Haemophilus influenzae | 0.016 | 0.125 |
| Streptococcus agalactiae | 0.5 | 0.5 |
| Streptococcus pneumoniae | 0.016 | 0.5 |
| Neisseria meningitidis | 0.5 | 0.5 |
| Listeria monocytogenes | 2 | 4 |
| Staphylococcus aureus | 1 | 4 |
| Escherichia coli | 2 | 8 |
| Staphylococcus epidermidis | 2 | 16 |
| Pseudomonas aeruginosa | 8 | 128 |

MIC$_{50}$ and MIC$_{90}$ mean minimum inhibitory concentration required to inhibit the growth of 50% and 90% of organisms, respectively. These values have been derived from the wild-type distributions of bacteria listed on the EUCAST Web site (European Committee on Antimicrobial Susceptibility Testing. Data from the EUCAST MIC distribution Web site; Available from: https://mic.eucast.org/Eucast/t2/SeachController/search.jsp?action=performSearch&BeginIndex=0&Micdistribution=mic&NumberIndex=50&Antib=251&Specie=-1).
multiple doses, median peak concentrations at steady state increased to 2.78 (range 0.47–7.53) mg/L. Mean recovery of piperacillin was $98.3 \pm 12.9\%$ (mean/SD).

**Brain ECF Pharmacokinetic Model**

Brain concentrations of piperacillin were best described by a one-compartment model with first-order elimination. Based on the observed delay in the initial rise of brain concentrations (Fig. 1), a lag time ($T_{\text{lag}}$) and a transit compartment between plasma and brain were added (Fig. 2). Neither $T_{\text{lag}}$ nor the transit compartment alone was sufficient to describe the data. Multiple transit compartments did not improve the model further. Separate population estimates were generated for transfer rate from brain to plasma ($K_{\text{bp}}$) and $T_{\text{lag}}$ after the first dose and at steady state. At steady state, $T_{\text{lag}}$ was not significantly different from zero; therefore, it was fixed to zero. A combined error model best described RUV in the model. Median estimates of the parameters with respective 95% confidence intervals (CI) are given in Table 3 from bootstrap analysis. In case of below quantification limit (BQL) concentrations, the M5 method performed similarly to the gold standard M3 method in terms of similar parameter estimates and goodness of fits. M5 was preferred for the final model because of numerical difficulties related to the M3 method.

**Model Evaluation**

Goodness-of-fit plots (Fig. 3) showed reasonable agreement between brain microdialysate observations and individual as well as population predictions for the final PopPK model. Conditional weighted residuals were uniformly distributed around zero; however, slight
under-prediction at lower concentrations was visible. The M3 method did not improve the observed under-prediction at low concentrations either. Nonetheless, the model adequately described the central tendency and dispersion of the observed data appropriately (Fig S1 in supplementary materials). Concentration–time plots of individual patients in brain also indicated that model had appropriately described the data (Fig S2). Bootstrap results (Table 3) indicated that PK parameter estimates were stable. Both eta and epsilon shrinkages were below 20% on all parameters of the final model. The condition number representing model identifiability was 11.23.

### Probability of Target Attainment

MCS indicated that the probability of target attainment was more than 90% for MICs up to 0.5 mg/L for all three simulated doses (12, 16, and 24 g per day). For a MIC of 1 mg/L, PTA was more than 90% only for the doses of 24 g per 24 h. All three types of infusion performed similarly with respect to PTA over the dose range of 12–24 g per day (Fig. 4). These results were not significantly different when other plasma models were used (Fig. 4 and Fig S3 in supplementary materials).

### Discussion

This study quantified the concentrations of piperacillin in the interstitial fluid of brain and generated a PopPK model to describe its pharmacokinetics. MCS suggests that similar CNS exposures are attained in brain ECF with different types of infusion. Furthermore, our results infer that only highly susceptible pathogen can be empirically treated successfully by piperacillin in brain in patients without meningeal inflammation.

The similar piperacillin exposure and related PTA in brain with different types of infusion are an interesting finding (Fig. 4) because many studies based on plasma data have shown superiority of continuous and prolonged infusion over intermittent infusions with regard to PTA in various cohorts of patients including critically ill patients [9, 10]. This result is readily explained by the observation that fluctuations of plasma concentrations are dampened considerably by the transport of the drug to and from the brain, thus mimicking kind of a continuous infusion for the brain. Therefore, continuous infusion would not provide any pharmacokinetic benefit over intermittent infusion to achieve appropriate piperacillin brain concentrations in patients without general meningeal inflammation. This may also apply for other, mainly hydrophilic, anti-bacterials with slow transport into and from the brain and for other tissues with low transfer rates from and to the plasma.

Piperacillin is an anti-psudomonal penicillin derivative mainly used to treat nosocomial pathogens such as *Pseudomonas aeruginosa*. However, based on a daily dose of 12 g of piperacillin, maximum observed brain concentrations after multiple doses (i.e., about 7.5 mg/L) were far below the concentration expected to be successful in treating *P. aeruginosa* (Table 2). For the same daily dose, MCS showed that the pharmacodynamic target in brain is only achieved for bacteria up to a MIC of 0.5 mg/L, and thus, infections involving only highly susceptible pathogens (Table 2) could be treated with this commonly used dosing regimen. Using higher doses (16–24 g/day) only achieved an acceptable PTA up to a MIC of 1 mg/L. This shows that increasing the dose up to the maximum recommended doses might still not be sufficient to treat CNS infections caused by pathogens with higher MICs (Table 2). Thus, piperacillin might not be an option for prophylaxis for invasive neurosurgical procedures, despite of some previous evidence supporting its use [28]. As an additional problem for its potential use, high variability in piperacillin brain pharmacokinetics was found (Fig S1) which is similar to plasma data where a high variability has also been reported in critically ill patients of various populations [10, 27, 29–31]. Having said this, it is important to mention that brain exposure of piperacillin in meningitis patients is expected to be higher than in hemorrhagic stroke patients because only a small part of the brain is affected as compared to meningitis where the blood–brain barrier is disrupted based on the generalized inflammation [32].

An important consideration for the interpretation of our results is that tazobactam concentrations were not

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**Table 3 Pharmacokinetic parameters of the brain model of piperacillin**

| Parameter                      | Bootstrap estimates (samples = 1000) (median [95% CI]) |
|-------------------------------|--------------------------------------------------------|
| Fixed effects                 |                                                        |
| $K_{pb,f}$ (h$^{-1}$)         | 0.32 (0.16–0.39)                                      |
| $K_{op,f}$ (h$^{-1}$)         | 7.31 (3.47–12.9)                                     |
| $K_{op,md}$ (h$^{-1}$)        | 4.39 (1.35–10.6)                                     |
| $T_{lag,fd}$ (h)              | 2.70 (2.21–3.52)                                     |
| $T_{lag,md}$ (h)              | 0 FIXED                                              |
| Inter-individual variability (CV %) | 19.7 (5.0–33.6)                                 |
| Additive error                | 0.03 (0.02–0.03)                                      |
| Proportional error            | 0.35 (0.23–0.44)                                      |

$k_{pb}, k_{op,fd}, k_{op,md}$ represent rate of transfer from plasma to brain, transfer rate from brain to plasma after first dose, and transfer rate from brain to plasma after multiple doses, respectively. $T_{lag,fd}$ and $T_{lag,md}$ represent lag time after first and multiple doses, respectively. CI, confidence interval; CV, coefficient of variation; SD, standard deviation.
Fig. 3 Combined goodness-of-fit plots of the piperacillin brain concentrations. Individual predictions versus observed concentrations (upper left), population predictions vs. observed concentration (upper right), population predictions versus conditional weighted residuals (lower left), and time after dose versus conditional weighted residuals (lower right).

Fig. 4 Plots showing probability of target attainment in brain at various doses and MIC levels for different forms of infusion using plasma model from Roberts et al. [10].
quantifiable. MICs of piperacillin are typically higher than MICs of piperacillin/tazobactam for several pathogens, especially for beta-lactamase producers [33]. In the worst case, tazobactam would not reach the brain and the higher MICs of piperacillin rather than MICs of piperacillin/tazobactam would apply. It is generally believed that piperacillin and tazobactam exhibit similar pharmacokinetics [34] and tazobactam does not affect the pharmacokinetic behavior of piperacillin [35]. However, piperacillin inhibits the cumulative urinary excretion of tazobactam (mediated through OATs transporters) thereby increasing its plasma concentrations [36]. In addition, a recent study reported a high variability in the ratio of piperacillin over tazobactam (ranging from 1 to 10) despite of their high correlation (0.93) [37]. We speculate that this variability might be even higher in brain as compared to the plasma data.

Because microdialysis in vivo recovery experiments could not be carried out due to retrospective nature of the study, brain data were fitted on the basis of in vitro recovery of piperacillin. However, there is some evidence that in vivo recovery might slightly differ from in vitro, usually with lower values in vivo [38]. However, at low flow rates for hydrophilic molecules with relatively low molecular weight like piperacillin, high recovery can be expected. Therefore, the low flow rate used in our study (0.3 μL/min) would explain why our in vitro recovery values are higher as compared to in vivo values (8–40%) reported in the literature [39–41] where flow rates of 1.5–2 μL/min were used. Thus, PTA values estimated in the present study with the assumption of a recovery of 100% are a realistic, albeit conservative estimate. Similarly, we could not estimate brain penetration from area under the concentration time curve (AUC) ratio between brain and plasma (AUCbrain/AUCplasma) of piperacillin due to the lack of plasma data. However, the unavailability of plasma concentrations is not relevant for PTA estimations as also supported by the sensitivity analysis (Fig. S3). Additionally, our simulation results are based on pharmacodynamic targets validated in plasma as brain targets were not available. Small sample size and the presence of BQL values were among the other limitations of our study.

**Conclusion**

In conclusion, piperacillin exposure to brain is delayed after initial intravenous infusion, and concentration profiles would be expected to remain similar to different durations of infusion. The results suggest that piperacillin would not be appropriate in most CNS infections, in particular, in pseudomonas CNS infections.

**Electronic supplementary material**

The online version of this article (https://doi.org/10.1007/s12028-020-00947-x) contains supplementary material, which is available to authorized users.

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**Author Contributions**

RH, RB, M1, and M2 conceived, designed, and conducted the clinical study. CD and AK performed quantification and in vitro recovery experiments. UF and MT designed population pharmacokinetic analysis plan. SU and UD developed population pharmacokinetic model and performed simulations. SU wrote initial draft of the manuscript. All authors contributed to analysis and interpretation of results and agreed upon the final version of the manuscript.

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**Conflict of interest**

The authors declare that they have no conflict of interest.

**Ethical Approval**

This study was approved by the Ethics Committee of the Medical University of Innsbruck (Approval Numbers AN3898 285/4.8, AM4091-292/4.6, UN3898 285/4.8).

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**References**

1. Nau R, Kinzig-Schippers M, Sörgel F, et al. Kinetics of piperacillin and tazobactam in ventricular cerebrospinal fluid of hydrocephalic patients. Antimicrob Agents Chemother. 1997;41(5):987–91.
2. Dickinson GM, Droller DG, Greenman RL, Hoffman TA. Clinical evaluation of piperacillin with observations on penetrability into cerebrospinal fluid. Antimicrob Agents Chemother. 1981;20(4):481–6.

3. Westerhout J, Danhof M, De Lange ECM. Preclinical prediction of human brain target site concentrations: considerations in extrapolating to the clinical setting. J Pharm Sci. 2011;100(9):3577–93.

4. Blaney SM, Daniel MJ, Harker AJ, Godwin K, Balis FM. Pharmacokinetics of lamivudine and BCH-189 in plasma and cerebrospinal fluid of nonhuman primates. Antimicrob Agents Chemother. 1995;39(12):2779–82.

5. Shapiro WR, Young DF, Mehta BM. Methotrexate: distribution in cerebrospinal fluid after intravenous, ventricular and lumbar injections. N Engl J Med. 1975;293(4):164–1.

6. Carteron L, Bouzat P, Oddo M. Cerebral microdialysis monitoring to improve individualized neurointensive care therapy: An update of recent clinical data. Front. Neurol. 2017;8:1–10. Available from: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5693841/pdf/natu.00601.pdf. Accessed 21 Dec 2019.

7. Drusano GL. Antimicrobial pharmacodynamics: critical interactions of “bug and drug”. Nat Rev Microbiol. 2004;2(4):289–300.

8. Huang H, Huang S, Zhu P, Xi X. Continuous versus intermittent infusion of cefepime in neurological patients with post-operative intracranial infections. Int J Antimicrob Agents. 2014;43(1):68–72.

9. Abdul-Aziz MH, Lipman J, Akova M, et al. Is prolonged infusion of piperacillin/tazobactam and meropenem in critically ill patients associated with improved pharmacokinetic/pharmacodynamic and patient outcomes? An observation from the Defining Antibiotic Levels in Intensive care unit patients (DALI) cohort. J Antimicrob Chemother. 2016;71(1):196–207.

10. Roberts JA, Kirkpatrick CMJ, Roberts MS, Dalley AJ, Lipman J. First-dose and steady-state population pharmacokinetics and pharmacodynamics of piperacillin by continuous or intermittent dosing in critically ill patients with sepsis. Int J Antimicrob Agents. 2010;35(2):156–63.

11. Teo J, Liew Y, Lee W, Kwa AL-H. Prolonged infusion versus intermittent boluses of β-lactam antibiotics for treatment of acute infections: a meta-analysis. Int J Antimicrob Agents. 2014;43(5):403–11.

12. Rhodes NJ, Liu J, O’Donnell JN, et al. Prolonged infusion piperacillin-tazobactam decreases mortality and improves outcomes in severely ill patients. Results of a systematic review and meta-analysis. Crit Care Med. 2018;46(2):236–43.

13. Morgenstern LB, Hemphill JC, Anderson C, et al. Guidelines for the management of spontaneous intracerebral hemorrhage: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. Stroke. 2010;41(9):2108–29.

14. Connolly ES, Rabinstein AA, Carhuapoma JR, et al. Guidelines for the management of aneurysmal subarachnoid hemorrhage: a guideline for healthcare professionals from the Neurocritical Care Society and the European Society of Intensive Care Medicine. Intensive Care Med. 2014;40(9):1189–209.

15. US Department of Health and Human Services Food and Drug Administration. (2013) FDA guidance for industry: bioanalytical method validation. Available from: http://www.fda.gov/Drugs/GuidanceComplianceReguloryInformation/Guidances/default.htm. Accessed 14 Oct 2018.

16. European Medicines Agency (2011) EMA guideline on bioanalytical method validation. Available from: http://www.ema.europa.eu. Accessed 25 May 2016.

17. Beal SL, Sheiner LB, Boeckmann AJ, Bauer RJ. NONMEM 7.4. 3 Users Guides (1989-2018). Hanover, MD, USA: ICON Development Solutions. 2018.

18. Krekels EH, van Hasselt JGC, Tibboel D, Danhof M, Knibbe CAJ. Systematic evaluation of the descriptive and predictive performance of paediatric morphine population models. Pharm Res. 2011;28(4):797–811.

19. Karlsson MO, Holford N (2008) A tutorial on visual predictive checks. Abstr17th PAGE, Abstr1434. Available from: https://www.page-meeting.org/?abstract=1434. Accessed 15 May 2019.

20. Beal SL. Ways to fix a PK model with some data below the quantification limit. J Pharmacochemical Pharmacodynam. 2001;2(5):481–504.

21. Tunkład B, Hammarlund-Udenaes M, Jonsson EN. An integrated model for the analysis of pharmacokinetic data from microdialysis experiments. Pharm Res. 2004;21(9):1698–707.

22. Lodise TP, Lomaestro B, Rodovld KA, Danziger LH, Drusano GL. Pharmacodynamic profiling of piperacillin in the presence of tazobactam in patients through the use of population pharmacokinetic models and Monte Carlo simulation. Antimicrob Agents Chemother. 2004;48(12):4718–24.

23. European Committee on Antimicrobial Susceptibility Testing. Data from the EUCAST MIC distribution website. Available from: https://mic.eucast.org/Eucast2/SearchController/search.jsp?action=performSearch&BeginIndex=0&MidOfMicroset&OrderByIndex=50&An=25&Species=1.

24. Abdul-Aziz MH, Lipman J, Akova M, et al. Population pharmacokinetics of piperacillin in nonobese, obese, and morbidly obese critically ill patients. Antimicrob Agents Chemother. 2017;61(3):e01276.

25. Obirink-Hansen K, Juul RV, Storgaard M, et al. Population pharmacokinetics of piperacillin in the early phase of septic shock: Does standard dosage result in therapeutic plasma concentrations? Antimicrob Agents Chemother. 2015;59(1):7018–26.

26. Bullock R, van Dellen JR, Ketebel W, Reinach SG. A double-blind placebo-controlled trial of perioperative prophylactic antibiotics for elective neurosurgery. J Neurosurg. 1988;69(5):687–91.

27. Taccone FS, Latere P-F, Dugernier T, et al. Insufficient β-lactam concentrations in the early phase of severe sepsis and septic shock. Crit Care. 2010;14(4):R126.

28. Roberts JA, Paul SK, Akova M, et al. DALI defining antibiotic levels in intensive care unit patients: Are current β-lactam antibiotic doses sufficient for critically ill patients? Clin Infect Dis. 2014;58(8):1072–83.

29. Carlier M, Carrette S, Stove V, Verstraete AG, De Waele JJ. Does consistent piperacillin dosing result in consistent therapeutic concentrations in critically ill patients? A longitudinal study over an entire antibiotic course. Int J Antimicrob Agents. 2014;43(5):470–3.

30. Rodriguez-Cerrato V, McCooq CG, Michelow IC, et al. Pharmacodynamics and bactericidal activity of moxifloxacin in experimental Escherichia coli meningitis. Antimicrob Agents Chemother. 2001;45(11):3092–7.

31. Hayashi Y, Roberts JA, Paterson DL, Lipman J. Pharmacokinetic evaluation of piperacillin–tazobactam. Expert Opin Drug Metab Toxicol. 2010;6(8):1017–31.

32. Occhipinti DJ, Pendland SL, Schoonover LL, Rypins EB, Danziger LH, Rodvold KA. Pharmacokinetics and pharmacodynamics of two multiple-dose piperacillin-tazobactam regimens. Antimicrob Agents Chemother. 1997;41(11):2511–7.

33. Sörgel F, Kinzig M. The chemistry, pharmacokinetics and tissue distribution of piperacillin/tazobactam. J Antimicrob Chemother. 1993;31(Suppl A):39–60.

34. Rodriguez-Cerrato V, McCooq CG, Michelow IC, et al. Pharmacodynamics and bactericidal activity of moxifloxacin in experimental Escherichia coli meningitis. Antimicrob Agents Chemother. 2001;45(11):3092–7.

35. Sörgel F, Kinzig M. The chemistry, pharmacokinetics and tissue distribution of piperacillin/tazobactam. J Antimicrob Chemother. 1993;31(Suppl A):39–60.

36. Wen S, Wang C, Duan Y, et al. OAT1 and OAT3 also mediate the drug–drug interaction between piperacillin and tazobactam. Int J Pharm. 2018;537(1–2):172–8.

37. Zander J, Dobbelger G, Nagel D, et al. Variability of piperacillin concentrations in relation to tazobactam concentrations in critically ill patients. Int J Antimicrob Agents. 2016;48(4):135–9.

38. Matznerell P, Österreicher Z, Reiter B, Lackner E, Stimpf M, Zeitlinger M. Tissue pharmacokinetics of telavancin in healthy volunteers: a microdialysis study. J Antimicrob Chemother. 2016;71(1):3179–84.

39. Roberts JA, Roberts MS, Robertson TA, Dalley AJ, Lipman J. Piperacillin penetration into tissue of critically ill patients with sepsis—Bolus versus continuous administration. Crit Care Med. 2009;37(3):926–33.

40. Dalla Costa T, Nolting A, Kovař A, Derendorf H. Determination of free interstitial concentrations of piperacillin–tazobactam combinations by microdialysis. J Antimicrob Chemother. 1998;42(6):769–78.

41. Nolting A, Costa TD, Vistele R, Rand KH, Derendorf H. Determination of free extracellular concentrations of piperacillin by microdialysis. J Pharm Sci. 1996;85(4):369–72.