T-cell reduction is an important characteristic of coronavirus disease 2019 (COVID-19), and its immunopathology is a subject of debate. It may be due to the direct effect of the virus on T-cell exhaustion or indirectly due to T cells redistributing to the lungs. HIV/AIDS naturally served as a T-cell exhaustion disease model for recognizing how the immune system works in the course of COVID-19. In this study, we collected the clinical charts, T-lymphocyte analysis, and chest CT of HIV patients with laboratory-confirmed COVID-19 infection who were admitted to Jin Yin-tan Hospital (Wuhan, China). The median age of the 21 patients was 47 years [interquartile range (IQR) = 40–50 years] and the median CD4 T-cell count was 183 cells/μl (IQR = 96–289 cells/μl). Eleven HIV patients were in the non-AIDS stage and 10 were in the AIDS stage. Nine patients received antiretroviral treatment (ART) and 12 patients did not receive any treatment. Compared to the reported mortality rate (nearly 4%–10%) and severity rate (up to 20%–40%) among COVID-19 patients in hospital, a benign duration with 0% severity and mortality rates was shown by 21 HIV/AIDS patients. The severity rates of COVID-19 were comparable between non-AIDS (median CD4 = 287 cells/μl) and AIDS (median CD4 = 97 cells/μl) patients, despite some of the AIDS patients having baseline lung injury stimulated by HIV: 7 patients (33%) were mild (five in the non-AIDS group and two in the AIDS group) and 14 patients (67%) were moderate (six in the non-AIDS group and eight in the AIDS group). More importantly, we found that a reduction in T-cell number positively correlates with the serum levels of
INTRODUCTION

Lymphopenia is a common feature in patients with severe coronavirus disease 2019 (COVID-19), with drastically reduced numbers of T cells (Chen et al., 2020). A retrospective study of 522 patients with COVID-19 found that a clinical severity-dependent reduction in the number of T cells is inversely correlated with the serum level of interleukin 6 (IL-6) (Diao et al., 2020). This suggests that, as the disease severity progresses in patients with COVID-19, a concomitant rise in the inflammatory cytokine levels may drive the depletion of T-cell populations, which contributes to the host’s weaker defense and to worse recovery. Therefore, treatments boosting immunity are needed (Moon, 2020; Zheng et al., 2020).

However, the immune system is also affected by damaged lung cells through the release of cytokines, which can cause T cells to migrate to the lungs and lead to lymphopenia in the peripheral blood. Most patients with severe COVID-19 exhibit substantially elevated serum levels of pro-inflammatory cytokines such as IL-6 and IL-1β. Also, C-reactive protein (CRP) is found to be abnormally high (Jin et al., 2020). Pathological findings of COVID-19 patients showed a massive infiltration of lymphocytes in the lungs. Spleen atrophy was also observed (Cao, 2020). These findings indicate that aggressive lymphocyte redistribution in the lungs was induced and that treatments to regulate immunity are needed for COVID-19 infection (Huang et al., 2020; Mehta et al., 2020; Qin et al., 2020; Wan et al., 2020; Wu and Yang, 2020).

Defining the role of T-cell changes in COVID-19 patients provides potential targets for clinical management. HIV/AIDS naturally served as a T-cell exhaustion disease model for recognizing how the immune system works in the course of COVID-19 infection. Thereof, in this study, we conducted a comprehensive evaluation of the clinical and radiological features of COVID-19 patients with confirmed HIV infection to reveal the role of T lymphocytes in this disease.

MATERIALS AND METHODS

Patients

On March 4, 2020, 38 suspected cases with fever and dry cough were transferred to Jin Yin-tan Hospital, considering their shared history of exposure to patients with confirmed COVID-19. Of them, 24 have HIV/AIDS. Subsequently, 21 patients were confirmed to be infected with COVID-19, the diagnosis of which was based on clinical characteristics, chest imaging, test of COVID-19 virus, and the ruling out of common bacterial and viral pathogens that cause pneumonia.

Initial investigations included complete blood count, T lymphocyte count, coagulation profile, and serum biochemical test. Respiratory specimens were tested for common viruses using real-time RT-PCR assays approved by the China Food and Drug Administration. Routine bacterial and fungal examinations were also performed.

CT Visual Quantitative Evaluation

Two radiologists blinded to the clinical information reviewed all the images independently. The distribution and the density of lesions in each lobe, as well as the overall lung “total severity score” (TSS), were recorded. Each of the five lung lobes was assessed for percentage of lobar involvement and classified into none (%), mild (<50%), or severe (>50%), with corresponding scores of 0, 1, and 2, respectively. The density in each lobe was classified as having ground-glass opacities or with mixed ground-glass opacities or consolidation, with corresponding scores of 1 and 2, respectively. The TSS was reached by summing the five lobe distribution scores multiplied by the density score (ranging from 0 to 20).

Statistical Analysis

For variables that were non-normally distributed or are non-parametric, differences were assessed using the Mann–Whitney or Wilcoxon signed-rank test. Categorical variables were compared for the groups using the chi-square test (Fisher’s exact test when the expected value is <5). Statistical analysis was performed with SPSS 25 (IBM Corp., Armonk, NY, USA).

RESULTS

Demographic and Clinical Characteristics

The median age of the 21 patients was 47 years [interquartile range (IQR) = 40–50 years]; there was a prevalence of females (2 males and 19 females). The HIV infection stages of these patients according to the WHO classification are as follows: 1 patient had no adverse event (CD4 > 500), 2 patients were mild (350 < CD4 < 499), 7 patients were advanced (200 < CD4 < 349), and 10 patients were severe (CD4 < 200) (Table 1). Considering the clinical features, 11 patients were non-AIDS and 10 were AIDS presenters according to the Centers for Disease Control and Prevention (CDC) classification. Nine patients received antiretroviral treatment (ART) and 12 patients did not receive any ART. During hospitalization for COVID-19, antiretroviral therapy with lamivudine and tenofovir was administered to 20
### TABLE 1 | Baseline, treatments, and outcomes of COVID-19 patients with HIV/AIDS.

| Age (years) | All (N = 21) | Non-AIDS (N = 11) | AIDS (N = 10) | p-value |
|-------------|--------------|-------------------|---------------|---------|
| 35–40       | 6 (29)       | 4 (36)            | 2 (20)        | 0.338   |
| >40         | 13 (62)      | 6 (55)            | 7 (70)        |         |
| CD4+ T-cell count | <0.001 | | | |
| >500        | 1 (5)        | 1 (9)             | 0 (0)         |         |
| 351–500     | 2 (10)       | 2 (18)            | 0 (0)         |         |
| 201–350     | 7 (33)       | 7 (64)            | 0 (0)         |         |
| 100–200     | 5 (24)       | 0 (0)             | 5 (50)        |         |
| <100        | 5 (24)       | 0 (0)             | 5 (50)        |         |
| ART         | 9 (43)       | 6 (55)            | 3 (30)        | 0.466   |
| Current smoking | 0.063 |       | | |
| Any comorbidity |        |       | | |
| Hypertension | 4 (19)      | 1 (9)             | 3 (30)        | 0.310   |
| Diabetes    | 0 (0)        | 0 (0)             | 0 (0)         | 1.000   |
| Cardiovascular disease | 0.000 |       | | |
| Chronic obstructive | 0 (0)      | 0 (0)             | 0 (0)         | 1.000   |
| Malignancy  | 0 (0)        | 0 (0)             | 0 (0)         | 1.000   |
| Pulmonary disease | 2 (10)     | 1 (9)             | 1 (10)        | 1.000   |
| Chronic liver disease | 2 (10)     | 2 (18)            | 0 (0)         | 1.000   |
| COVID-19 type |            |                   |              | 0.362   |
| Mild        | 7 (33)       | 5 (45)            | 2 (20)        |         |
| Moderate    | 14 (67)      | 6 (55)            | 8 (80)        |         |
| Severe—critical | 0 (0)      | 0 (0)             | 0 (0)         |         |
| Signs and symptoms |      |                   |              |         |
| Highest temperature (°C) | 1.000 |       | | |
| >39         | 0 (0)        | 0 (0)             | 0 (0)         |         |
| 38–39       | 0 (0)        | 0 (0)             | 0 (0)         |         |
| 37.3–37.9   | 5 (24)       | 3 (27)            | 2 (20)        |         |
| Cough       | 8 (38)       | 4 (36)            | 4 (40)        | 1.000   |
| Diarrhea    | 0 (0)        | 0 (0)             | 0 (0)         | 1.000   |
| Dyspnea     | 3 (14)       | 1 (9)             | 2 (20)        | 0.086   |
| White blood cell count, 10^9/L |             |                   |              |         |
| <4          | 10 (48)      | 3 (27)            | 7 (70)        |         |
| 4–10        | 11 (52)      | 8 (73)            | 3 (30)        |         |
| >10         | 0 (0)        | 0 (0)             | 0 (0)         |         |
| Lymphocyte count, 10^9/L | 0.149 |       | | |
| >1.0        | 16 (76)      | 10 (91)           | 6 (60)        |         |
| <1.0        | 5 (24)       | 1 (9)             | 4 (40)        |         |
| Hemoglobin (g/L) | 0.659 |       | | |
| >110        | 14 (67)      | 8 (73)            | 6 (60)        |         |
| <110        | 7 (33)       | 3 (27)            | 4 (40)        |         |
| Platelet count, 10^9/L | 0.476 |       | | |
| >100        | 20 (95)      | 11 (100)          | 9 (90)        |         |
| <100        | 1 (5)        | 0 (0)             | 1 (10)        |         |
| Aspartate   |              |                   |              | 1.000   |
| aminotransferase (U/L) |             |                   |              |         |
| >40         | 7 (33)       | 3 (27)            | 4 (40)        |         |
| <40         | 14 (67)      | 8 (73)            | 6 (60)        |         |
| Creatine kinase (U/L) | 1.000 |       | | |
| >185        | 0 (0)        | 0 (0)             | 0 (0)         |         |
| <185        | 21 (100)     | 11 (100)          | 10 (100)      |         |
| Procalcitonin (ng/ml) |             |                   |              |         |
| <0.1        | 20 (95)      | 10 (91)           | 10 (100)      | 1.000   |
| 0.1–0.5     | 1 (5)        | 1 (9)             | 0 (0)         |         |
| >0.5        | 0 (0)        | 0 (0)             | 0 (0)         |         |
| CRP (mg/L)  |             |                   |              | 0.403   |
| <2          | 16 (76)      | 9 (82)            | 7 (70)        |         |
| 2–10        | 4 (19)       | 2 (18)            | 2 (20)        |         |
| >10         | 1 (5)        | 0 (0)             | 1 (10)        |         |

(Continued)
lacked data on this. All HIV patients showed typical features of compromised immunity: the mean CD4 T-cell count (226 cells/μl) and the mean CD4/CD8 ratio (0.5) were reduced below the mean limits of the reported patients with either severe (CD4 = 263 cells/μl, CD4/CD8 = 1.5) or moderate (CD4 = 451 cells/μl, CD4/CD8 = 1.7) COVID-19 (Qin et al., 2020), and these were reduced more profoundly in AIDS than in non-HIV-infected COVID-19 cases (CD4 = 347 vs. 104 cells/μl, p < 0.001; CD4/CD8 = 0.7 vs. 0.2 cells/μl, p < 0.001, respectively) (Figures 2A–D).

The mean levels of CRP (2.0 mg/L) and IL-6 (7.0 pg/ml) in all HIV patients were markedly lower than the mean limits of the reported patients with either severe (CRP = 57.9 mg/L, IL-6 = 25.2 pg/ml) or moderate (CRP = 33.2 mg/L, IL-6 = 13.3 pg/ml) COVID-19 (Qin et al., 2020); the levels of CRP (1.2 vs. 3.1 mg/L, p = 0.230) and IL-6 (7.7 vs. 6.5 pg/ml, p = 0.971) among non-AIDS patients and those with AIDS were comparable (Figures 2E, F).

The lack of T cells in HIV patients had no impact on long-term virus control, and the discharge time of these patients was not prolonged. After 20 days of hospitalization, with clinical improvements, all patients had increased CD4 T cells and lymphocytes (p < 0.05; Figures 2G–I), which may have been caused by the antiretroviral therapy for HIV or immune activity in COVID-19 (Dessie et al., 2020).

**DISCUSSION**

T cells particularly play a significant antiviral role in combating pathogens. CD4 T cells have an important role in promoting the production of virus-specific antibodies by activating T-dependent B cells and enhancing CD8 T-cell toxicity in order to kill virus-infected cells. It has been reported that COVID-19 is more likely to infect older men with comorbidities (Huang et al., 2020), who have weaker immune functions. Several studies have shown that decreased CD4 T-cell and lymphocyte counts are associated with severe clinical types, which supports having weaker immune functions with T-cell exhaustion as contributing to the severity of COVID-19 (Moon, 2020; Zheng et al., 2020). However, in this study, we investigated 21 HIV/AIDS patients and found that all of them had a benign clinical form of COVID-19 and had better prognosis. Regardless of the CD4 cell counts and whether ARTs have been administered (Table 1), none of the patients...
progressed to the severe form or to death, despite some of the AIDS patients presenting with baseline lung injury stimulated by HIV (Figure 1).

The T-cell counts were significantly reduced in COVID-19 patients—the surviving T cells in severe COVID-19 patients with CD4 T-cell counts were lower than 400 cells/μl (Moon, 2020; Zheng et al., 2020)—but which were still higher than those of patients with HIV/AIDS in this study (226 cells/μl). COVID-19 caused a respiratory disease that led to a mortality rate of nearly 4%–10%: up to 20%–40% of patients required ventilatory support and even intensive care. However, the severity and mortality rates in this study were the lowest compared to previous reports, which indicates that a compromised immune system with a lower CD4 T-cell count might waive the clinical symptoms (Zhou F. et al., 2020).

HIV, as a T-cell exhaustion disease, leads to having a suboptimal immune function for defense against multiple viral infections. The lungs are one of the main target organs for virus-associated disease, and almost 70% of HIV patients suffer from at least one respiratory complication during the course of their illness (Rosen et al., 2001). In HIV patients, there is an enhanced prevalence of bronchial hyperresponsiveness and dysfunction of the small airways (Poirier et al., 2001). Surprisingly, when HIV patients are infected with COVID-19, they show mild symptoms. These data imply that the immune system actively participates in the severe pathogenesis of COVID-19. This immune activation

**FIGURE 2** | Lymphocyte subset counts and C-reactive protein (CRP) and interleukin 6 (IL-6) levels compared between AIDS and non-AIDS patients (A–F). One patient had no lymphocyte subset data on admission and 12 patients lacked lymphocyte subset data at follow-up (FL) (G–I). The Mann-Whitney test was performed for different group comparisons and Wilcoxon signed-rank test was performed for follow-up analysis. Mean values are shown in green for this study. Orange and blue dotted lines indicate the mean values reported by others for common patients or severe COVID-19 patients without HIV/AIDS (Qin et al., 2020).
may cause amplification of the inflammatory cascade and lead to a massive influx of lymphocytes to the lungs from the periphery (lymphocyte redistribution). Data reported on three aspects by others support this opinion: 1) accumulating evidence suggests that a subgroup of patients with severe COVID-19 might have higher serum levels of pro-inflammatory cytokines (TNF-α, IL-1, and IL-6) that activate T cells (Mehta et al., 2020; Qin et al., 2020; Wan et al., 2020; Wu and Yang, 2020); 2) pathological findings on the lungs of COVID-19 patients showed massive infiltration of lymphocytes in the lungs (Yang X et al., 2020); and 3) some chronic immune-inflammatory disease patients on immunosuppressants have no increased risk of COVID-19 (Brownlee et al., 2020; Monteleone and Ardzzone, 2020; Price et al., 2020).

Although immune-inflammatory treatment is not routinely recommended for COVID-19, according to the finding of this study and other findings about cytokine storm syndrome, a timely and appropriate use of immune modulators with antiretroviral therapy support should be considered for mild patients in order to prevent development of the severe form (Zhou Y. et al., 2020; Zhou et al., 2021). The sphingosine-1-phosphate receptor regulator fingolimod (FTY720) is an effective immunology modulator that targets CD4 T cells, which has been widely used in multiple sclerosis with a proven safety profile. Fingolimod acts as an immunology modulator that inhibits the egress of lymphocytes from the lymph nodes and limits their migration, especially CD4 T cells (Kappos L et al., 2010). With the hypothesis of reducing focal pulmonary inflammatory responses using fingolimod, this drug could be potentially considered for use in clinical trials.

CONCLUSIONS

There are some caveats to the present study, such as the small sample size, its retrospective design, and the lack of detection of TCR zeta-chain expression (Baniyash, 2004). However, only a few clinical studies have, so far, tested the hypothesis that modulation of inflammation and immune response can ameliorate the severity of COVID-19. HIV/AIDS naturally served as a T-cell exhaustion disease model for us to recognize how the immune system works in the course of COVID-19. This study demonstrated that lower CD4 counts might waive the clinical symptoms and inflammatory responses in COVID-19, which would advance our understanding of the virus–immune system interaction in clinical settings. In addition, this study suggests that patients with chronic immune-inflammatory diseases (e.g., multiple sclerosis and rheumatic arthritis) should stay on immunosuppressant or immune modulator medications. On the other hand, it is noteworthy that suppression of the effector CD4 T lymphocyte could be beneficial for mitigating the progress of COVID-19.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Jin Yin-tan Hospital Ethics Committee (KY-2020-51.01). The patients provided written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

YF, and C-LH had full access to all of the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. YF and C-LH contributed to the study concept and design. All authors acquired, analyzed, and interpreted the data. YF and X-HL drafted the manuscript. NW, W-JC and C-LH critically reviewed the manuscript for important intellectual content. YF did the statistical analysis, obtained funding, and provided administrative, technical, or material support. YF and C-LH supervised the study. All authors contributed to the article and approved the submitted version.

FUNDING

This work has been supported by grant 81771279 (to YF) from the National Natural Science Foundation of China. This work also has been supported by grant No.2021MP03 (H-SL) from the Nursery Fund Project of the Second Affiliated Hospital of Fujian Medical University.

ACKNOWLEDGMENTS

We thank our patients for their participation in this study and the medical team of Jin Yin-tan Hospital for the treatment and prevention of COVID-19.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fcimb.2021.564938/full#supplementary-material

REFERENCES

Baniyash, M. (2004). TCR Zeta-Chain Downregulation: Curtailing an Excessive Inflammatory Immune Response. Nat. Rev. Immunol. 4, 675–687. doi: 10.1038/nri1434

Brownlee, W., Bourdette, D., Broadley, S., Killestein, J., and Ciccarelli, O. (2020). Treating Multiple Sclerosis and Neuromyelitis Optica Spectrum Disorder During the COVID-19 Pandemic. Neurology 94, 949–952. doi: 10.1212/WNL.0000000000009507

Cao, X. (2020). COVID-19: Immunopathology and Its Implications for Therapy. Nat. Rev. Immunol. 20, 269–270. doi: 10.1038/s41577-020-0308-3

Chen, G., Wu, D., Guo, W., Cao, Y., Huang, D., Wang, H., et al. (2020). Clinical and Immunologic Features in Severe and Moderate Coronavirus Disease 2019. J. Clin. Invest. 130, 2620–2629. doi: 10.1101/2020.02.16.20023903
Dessie, Z. G., Zewotir, T., Mwambu, H., and North, D. (2020). Modelling of Viral Load Dynamics and CD4 Cell Count Progression in an Antiretroviral Naive Cohort: Using a Joint Linear Mixed and Multistate Markov Model. BMC Infect. Dis. 20, 246. doi: 10.1186/s12879-020-04972-1

Diao, B., Wang, C., Tan, Y., Chen, X., Liu, Y., Ning, L., et al. (2020). Reduction and Functional Exhaustion of T Cells in Patients With Coronavirus Disease 2019 (COVID-19). Front. Immunol. 11, 827. doi: 10.3389/fimmu.2020.00827

Huang, C., Wang, Y., Li, X., Ren, L., Zhao, J., Hu, Y., et al. (2020). Clinical Features of Patients Infected With 2019 Novel Coronavirus in Wuhan, China. Lancet 395, 497–506. doi: 10.1016/S0140-6736(20)30183-5

Lin et al. HIV Meets COVID-19

Zhou, Y., Fu, B., Zheng, X., Wang, D., Zhao, C., Qi, Y., et al. (2020). Pathogenic T-Cells and Inflammatory Monocytes Incite Inflammatory Storms in Severe COVID-19 Patients. Cell. Mol. Immunol. 17, 541–543. doi: 10.1038/s41423-020-0401-3

Zhou, Y., Zhang, J., Wang, D., Guan, W., Qin, J., et al. (2021). Pro inflamatory Response in Patients With Coronavirus 2019 (COVID-19) in Wuhan, China: A Retrospective Cohort Study. Lancet 395, 1054–1062. doi: 10.1016/S0140-6736(20)30566-3

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Lin, Lin, Wang, Wen, Xiang, Fan, Li, Wu, Lin, Lin, Sun, Chen, Lian, Xiao, Lin, Xie, Li, Peng, Wang, Lin, Chen, Huang and Fu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.