USE OF WIDAL TEST IN DIAGNOSIS OF TYPHOID FEVER IN NORTH INDIAN POPULATION BY ESTIMATING BASELINE TITER IN CONTROL GROUP

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ABSTRACT

Objective: This study was undertaken to establish a cutoff significant titer for Widal test using healthy volunteers as control group. Utilizing the baseline titer obtained from the control group, a diagnosis of typhoid fever was made in the test group comprising outpatients and inpatients.

Methods: Blood samples were collected from healthy volunteers and patients attending G.S Medical College and Hospital, Pilkhuwa, over a period of 6 months from September 2016 to March 2017. Antibodies to Salmonella typhi (TO, TH) and Paratyphi A (AH) and Paratyphi B (BH) are determined by this tube agglutination test. A total of 124 healthy controls and 303 patients having clinical suspicion of typhoid fever were subjected to Widal test.

Results: In healthy control group, titer TO ≥20 was observed in 43 (34.7%), TO ≥40 in 48 (38.7%), TO ≥80 in 25 (20.2%), and titer TO ≥160, TO ≥320 was observed in none of the control group. Titer TH ≥40 in 58 (46.8%), AH ≥40 in 7 (5.6%), and BH ≥40 in 13 (10.5%) were observed in the control group. Among the test group, 96 (31.7%) sera were positive out of 303 clinically suspected enteric fever by the Widal test. Among different age group studied, 34 (46.6%) patients belonged to the age group of 11-20 years which formed the highest followed by the age group of 21-30 years (33.3%).

Conclusion: Based on the study, a cutoff titer of ≥160 for anti-O and anti-H antibodies and titer of ≥80 for anti-AH and anti-BH antibodies be considered as significant titer in diagnosis of enteric fever in this region. The baseline titer helps in early recognition and treatment of this serious health problem.

Keywords: Titer, Widal, Antibodies, Typhoid fever, Agglutination.

INTRODUCTION

Enteric fever is endemic in the Indian subcontinent, and population-based surveillance studies in selected regions add considerably in estimating the global burden of enteric fever. The incidence rate of typhoid is more than 100 cases per 100000 people per year [1]. Enteric fever caused by Salmonella typhi is called as typhoid fever and Salmonella paratyphi causes the paratyphoid fever. Typhoid fever is characterized by malaise, fever, abdominal discomfort, relative bradycardia, transient rash, and organomegaly and may end up in complications such as intestinal perforation, hemorrhage, and perinephric abscess [2,3]. Human beings are the only known reservoir host for typhoid fever, and infected person or carrier transmits the organism in his feces. Ingestion of food or water contaminated by such feces is the source of infection [4].

Drug resistance has been posing major problems in the treatment of typhoid fever and continues to be a serious problem for public health authorities [5]. Studies are being carried out to use plant constituents as potential antimicrobial agents in future for treatment [6]. Newer vaccines are being developed to overcome poor immunogenicity of some available vaccines, and research work on better adjuvant delivery systems is being carried out [7].

The definitive diagnosis of enteric fever is by culture of blood, stool, urine, and intestinal secretions. However, in many countries, where injudicious use of antibiotics and inadequate culture facilities is common, Widal test is still the most convenient way for serological diagnosis of typhoid fever. Moreover, for effective diagnosis of typhoid fever, baseline titer of the population has to be assessed.

Widal test is a classical tube agglutination test which measures agglutinating antibodies against lipopolysaccharide O and protein flagellar H antigens of Salmonella. The diagnosis relies on demonstrating rising antibody titer in paired samples 10-14 days apart. However, this rise is not demonstrable even in blood culture-confirmed cases so prompting baseline titer estimation of population a necessity [8].

There are no studies about the baseline titer been estimated earlier in this region. In studies done previously in other parts of the country, the antibody titer was found to be unusually high though it can vary from region to region [9]. In the background of such reports, it is necessary to perform baseline titer in healthy control group before conducting the test on patient sera.

METHODS

The study was conducted in the Department of Microbiology, G.S Medical College and Hospital, which is a tertiary care hospital with 350 beds situated in Pilkhuwa, Uttar Pradesh, for 6 months from September 2016 to March 2017. Consent and the approval of the Institutional Ethical Committee was obtained for carrying out this study. People with underlying respiratory infections, malaria, dengue, hepatitis, hematological, or other systemic disorders were excluded from the study.

Control group

The control group consisted of 124 healthy people who are free of signs and symptoms and not having significant ill health within the past 2 months or typhoid fever in the past 6 months. Those who gave history of vaccination in the preceding 3 years were excluded from the study. Among the control group, male constituted 41.13% and female 58.87% of vaccination in the preceding 3 years were excluded from the study.

Test group

The test population consisted of 303 clinically suspected typhoid fever cases attending the hospital as outpatients (O.P) or admitted as inpatients (I.P). Patients previously treated with antibiotics such as...
chloramphenicol or had recent typhoid illness or having chronic active liver disease were excluded from the study. Significant titer determined from control group was applied to make a clinical diagnosis of enteric fever.

A volume of 3-5 ml of venous blood was collected into a sterile test tube and centrifuged for 5 minutes to separate the serum from blood. Commercially available antigen containing Salmonella enterica serovar Typhi O and H antigens and Paratyphi AH and BH antigen were used (Arkay Healthcare Pvt., Ltd., Surat, India). The test used a two-fold serially diluted sera of control and test group, dilution being from 1:20 to 1:320. All serum samples were first diluted in a 1:20 ratio with isotonic normal saline in such a way that the final volume amounts to 1 ml. Dilutions of 1:40, 1:80, 1:160, and 1:320 each were made serially in 4 rows. Add a drop of the appropriate antigen to all the corresponding tubes in each row. A known positive control, negative control, and antigen control were also set up in each row. All the tubes are mixed well and incubated at 37°C for 16-20 hrs and examined for agglutination. The antibody titer is the highest dilution of serum showing distinct agglutination. The O somatic antigen being the somatic antigen gives compact, granular agglutination whereas H antigen, a flagellar antigen, gives large loose fluffy agglutination.

Data were entered and analyzed using SPSS (Statistical Package for Social Science) program version 24, and statistical significance was considered when p value was less than 0.05.

RESULTS

A total of 124 healthy controls and 303 patients having clinical suspicion of typhoid fever were subjected to Widal test. In healthy control group, titer TO ≥20 was observed in 43 (34.7%), TO ≥40 in 48 (38.7%), TO ≥80 in 25 (20.2%), and titer TO ≥160, TO ≥320 was observed in none of the control group. Titer TH ≥40 in 58 (46.8%), AH ≥40 in 59 (46.8%), and BH ≥40 in 51 (40.8%) were observed in the control group. Out of the 124 sera tested, only 8 sera (6.5%) and 19 sera (15.3%) showed positive agglutination for S. paratyphi AH and BH antibodies, respectively. Male constituted 41.13% (52 out of 124) of the control group.

Among the test group, 96 (31.7%) were positive by Widal test and 34 (46.6%) patients belonged to the age group of 11-20 years which formed the highest group whereas age group <10 years constituted the least (20.3%). Male constituted 50.49% and female 49.50% and O.P accounted for 56.74% and LP accounted for 41.25% in the test group (Tables 1-3).

The present study has found the paratyphi BH titer ≥40 which is found to be higher than other studies done on healthy volunteers (Table 4).

DISCUSSION

The baseline titer in Widal test among control group for TO, TH antibodies was found to be ≥80 and AH and BH to be ≥40 each. The high titers noticed against the salmonella antigens in the study may be due to the exposure to the cross-reacting organisms in the region. Based on this, the significant titer was determined as a cutoff titer of ≥160 for anti-O and anti-H antibodies and titer of ≥80 for anti-TH and anti-BH antibodies to make a diagnosis of enteric fever [9]. A study done in determining baseline titer in Garbhwal region of Uttarakhand has recommended the baseline titer of ≥40 for TO antibodies and ≥80 for TH antibodies. Positive agglutinins for paratyphi AH and BH were found in only 3.04% and 1.3%, respectively. In this study, 8 sera (6.5%) and 19 sera (15.3%) showed positive agglutination titer for AH and BH antibodies. The baseline titer for paratyphi AH and BH groups was found to be ≥20 each in their study [10]. Similar baseline titer was also noticed in Thrivananthapuram in South Kerala [11]. The baseline titer determined was 40 for anti-O and anti-H antibodies and <20 for anti-AH and anti-BH antibodies in another study done in North Kerala where public health facilities and hygienic conditions are better [12]. Another study done in Raichur recommended TO and TH titer of ≥20 as diagnostic of typhoid fever, and for AH and BH titers, it is ≥40 and ≥160, respectively [13]. Baseline Widal titer study done in rural Pondicherry recommended ≥160 for O and H agglutinins while for AH to be ≥80 and BH to be ≥40 to be considered a significant titer in diagnosing typhoid [14]. A study done in Dehradun on apparently healthy population found agglutination titer for TO ≥20 in 28%, TO ≥40 in 24.6%, TO ≥80 in 10.3%, TH ≥80 in 7.6%, AH ≥20 in 6.6%, and BH ≥20 in 4.6% [15].

In this study, TO antibodies (93.5%) agglutinated more compared with TH (87.1%), AH (6.5%), or BH (15.3%). Even infected patients' sera agglutinated more with somatic antigen of S. typhi than the flagellar antigen. These findings have been noted in several studies [16]. Furthermore, several studies highlight the increasing cases of enteric fever due to paratyphi, but this study differs from such observation. Widal test helps the physician in making a rapid diagnosis instead of relying on blood or stool culture which is time-consuming. Added to this is self-medication by the patients and injudicious prescription of antibiotics by local doctors make culture tests less reliable.

Among the patients who attended the hospital with clinical suspicion of typhoid, 31.7% showed significant titer to S. typhi antibodies and none to paratyphi. Age group of 11-20 years showed highest prevalence (46.6%) followed by the age group of 21-30 years (33.3%). A similar baseline titer determined was ≥20 each in their study [10]. Similar baseline titer was also noticed in Thrivananthapuram in South Kerala [11]. The baseline titer determined was 40 for anti-O and anti-H antibodies and <20 for anti-AH and anti-BH antibodies in another study done in North Kerala where public health facilities and hygienic conditions are better [12]. Another study done in Raichur recommended TO and TH titer of ≥20 as diagnostic of typhoid fever, and for AH and BH titers, it is ≥40 and ≥160, respectively [13]. Baseline Widal titer study done in rural Pondicherry recommended ≥160 for O and H agglutinins while for AH to be ≥80 and BH to be ≥40 to be considered a significant titer in diagnosing typhoid [14]. A study done in Dehradun on apparently healthy population found agglutination titer for TO ≥20 in 28%, TO ≥40 in 24.6%, TO ≥80 in 10.3%, TH ≥80 in 7.6%, AH ≥20 in 6.6%, and BH ≥20 in 4.6% [15].

| Antibody titer | N (%) | ≥20 N (%) | ≥40 N (%) | ≥80 N (%) | ≥160 N (%) | ≥320 N (%) |
|---------------|-------|-----------|-----------|-----------|-----------|-----------|
| S. typhi O    | 116   | 43 (34.7) | 48 (38.7) | 25 (20.2) | 0 (0)     | 0 (0)     |
| S. typhi H    | 108   | 39 (31.5) | 59 (46.8) | 11 (8.9)  | 0 (0)     | 0 (0)     |
| S. paratyphi AH | 34   | 01 (0.8)  | 07 (5.6)  | 00 (0)    | 0 (0)     | 0 (0)     |
| S. paratyphi BH| 21    | 06 (4.6)  | 13 (10.5) | 00 (0)    | 0 (0)     | 0 (0)     |

N: Total number of positive sera in the control group; N: Number of positive sera against different antibody titer

Table 1: Antibodies titer against Salmonella antigens in the control group (N=124)

Table 2: Prevalence of typhoid fever in test group after estimating baseline titer

| Age group (years) | N | M | F | O.P | LP | N (%) |
|-------------------|---|---|---|-----|----|-------|
| <10               | 59| 31| 28| 34  | 25 | 12 (20.3) |
| 11-20             | 73| 40| 33| 35  | 38 | 34 (46.6) |
| 21-30             | 60| 31| 29| 41  | 19 | 20 (33.3) |
| 31-40             | 46| 21| 25| 29  | 17 | 14 (22.2) |
| 41-50             | 30| 08| 22| 20  | 10 | 08 (26.7) |
| >50               | 35| 22| 13| 19  | 16 | 08 (22.9) |
| Total             | 303| 153|150|178 |125 | 96 (31.7) |

N: Total number of patients examined in different age groups (N=303); M: Male; F: Female; O.P: Outpatient; I.P: Inpatient; N: Number of patients positive by Widal test by utilizing baseline titer
Table 4: Comparative analysis of baseline titer of O and H agglutinins in different parts of India

| Author                  | Place     | Year | Titer TO | Titer TH | Titer AH | Titer BH |
|-------------------------|-----------|------|----------|----------|----------|----------|
| Shekar Pal et al.       | Garhwal   | 2013 | ≥40      | ≥80      | ≥20      | ≥20      |
| Kataria et al.          | Dehradun  | 2013 | ≥80      | ≥80      | ≥20      | ≥20      |
| Bijapur et al.          | Kannur    | 2014 | ≥40      | ≥40      | <20      | <20      |
| Jeyakumari et al.       | Puducherry| 2015 | ≥80      | ≥80      | ≥40      | ≥20      |
| Present study           | Pilkhuwa  | 2017 | ≥80      | ≥80      | ≥40      | ≥40      |

There was no statistical significance between gender and age in this study (p>0.05). Similarly, no significant result was seen in another study done in Yemen, but the prevalence was highest in the age group >20 years. The study found that the main symptom in majority was fever followed by diarrhea and abdominal pain [19]. A study conducted in Owerri, Nigeria, has found that the typhoid fever was high among youths who consume unsafe drinking water and food from outside source which may be one of the reasons in this region too [20]. Serological tests such as Widal performed in the laboratory help in differentiating from other causes of febrile illness such as dengue [21].

Although blood culture is gold standard for diagnosis of typhoid fever, excessive antibiotic use has reduced this isolation rate. A study in Varanasi has found a significant increase in the overall seropositivity rates from 1998 to 2011. A rising prevalence of non-typhoidal salmonellae has contributed to such change. Furthermore, a fact that typhoid fever is primarily a disease of childhood and early adolescents may not hold true against a background of changing epidemiology due to childhood vaccination policies [22]. Relative frequencies and age distributions of invasive non-typhoidal salmonella and typhoid are found to be contrasting [23]. When the level of agglutinins was correlated with age in another study, it was found that the titre increased with duration of illness in adults but not in children [24].

CONCLUSIONS

Widal test still remains an important diagnostic tool since it is more convenient, reliable, cheaper and faster than culture, molecular, and other serological tests. Although new rapid serological tests are available, they need to be carefully validated before being used. However, the results of Widal test need to be analyzed based on endemicity and cross-reaction of antigen with other salmonella and non-salmonella species. Thus, it can be concluded that in hospitals, where facilities are limited, a presumptive diagnosis of typhoid fever can be comfortably made after estimating baseline titre of the region.

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