Apical pressures generated by several canal irrigation methods: a laboratory study in a maxillary central incisor with an open apex

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Running title: NaOCl extrusion in immature teeth

Keywords: apical extrusion, endodontics, immature tooth, NaOCl accident, root canal treatment, sodium hypochlorite

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Abstract

Aim A laboratory study to determine the apical pressure generated by seven canal irrigation methods in an anterior tooth with an open apex.

Methodology Canal irrigation was performed on a 3D-printed central maxillary incisor with an open apex (maximum diameter of 2.1 mm). Ultrasonically Activated Irrigation (UAI), sonic activation (EDDY), negative pressure irrigation (EndoVac), the self adjusting file (SAF) and the XP-endo Finisher were employed at tooth length (TL), TL – 1 mm, TL – 2 mm and TL – 3 mm. UAI was tested at three intensity levels additionally. Hydrodynamic irrigation with RinsEndo was performed in the pulp chamber, at the canal orifice, the coronal third, the middle of the canal and at TL. Er:YAG laser-activation, at four frequency settings, was performed in the pulp chamber and at the orifice of the canal. The pressure of the fluid towards the canal terminus generated by activation was directly transferred to a pressure sensor with a range of 0 to 120 mmHg and a response time of ≤ 0.5 milliseconds. The critical threshold for apical extrusion of the irrigant was set at 5.73 mmHg (lower limit of the central venous pressure: 5.88 ± 0.15 mmHg). Each experiment was repeated ten times. The tests were followed by descriptive analyses (maximum, mean, standard deviation, 95% confidence interval).

Results EndoVac, the SAF, the XP-endo Finisher and UAI never exceeded the critical threshold at any insertion depth or intensity level. Er:YAG laser-activation exceeded the critical threshold exclusively at frequency settings varied from the manufacturer’s recommendation. EDDY at TL and RinsEndo at any insertion depth exceeded the critical threshold in 100% of the measurements. EDDY at TL - 1, - 2 and - 3 mm crossed the critical threshold in 30%, 10% and 20% of the measurements, respectively.

Conclusions In a simulated maxillary central incisor with an open apex, irrigation with EndoVac, Er:YAG laser-activation, UAI, the SAF and the XP-endo Finisher generated apical pressures below the critical threshold of 5.73 mmHg. By contrast, using EDDY and RinsEndo for irrigation produced higher apical pressures that exceeded the critical threshold.
Introduction

Irrigation is an essential part of the chemo-mechanical preparation of root canals aiming to remove dentine debris, pulp tissue and microbes (Gulabivala et al. 2005, Haapasalo et al. 2005). Sodium hypochlorite (NaOCl) is the most commonly used irrigant owing to its excellent chemical properties (Haapasalo et al. 2005, Dutner et al. 2012, Zaugg et al. 2019). It has the potential to dissolve organic tissue, to act effectively against a broad range of microorganisms and to neutralize their products (Buttler & Crawford 1982, Zehnder 2006).

On the other hand, NaOCl has toxic effects on vital tissues, potentially leading to necrosis, haemolysis and ulceration (Pashley et al. 1985). It is therefore important to avoid irrigant extrusion into the periapical region. The frequency of such an extrusion is unknown but it is considered to be rare (Spencer et al. 2007). Nonetheless, the extrusion of NaOCl followed by severe symptoms has been described in several case reports (Boutsioukis et al. 2013). Possible symptoms include severe pain, tissue necrosis, oedema, profuse interstitial bleeding with haemorrhage or bleeding from the root canal, anaesthesia or paraesthesia, irritation of the throat and secondary infection (Hülsmann & Hahn 2000).

In vivo and ex vivo studies as well as reviews of case reports have attempted to identify factors that determine irrigant extrusion (Hülsmann & Hahn 2000, Dutner et al. 2012, Boutsioukis et al. 2013). Although some factors remain unclear, technical aspects have been investigated fully. For example, wedging of the syringe tip can inhibit the coronal outflow of the irrigant and thus increase the pressure of the fluid towards the foramen (Shen et al. 2010). A larger apical preparation size or the presence of apical ramifications have been reported to result in less extrusion in vitro (Boutsioukis et al. 2010, Lorono et al. 2020). In addition, an open apex is considered to be a significant factor favoring extrusion of irrigants. In a survey among board-certified endodontists (diplomates of the American Board of Endodontics), an open apex was named the most common reason for a NaOCl accident (Kleier et al. 2008).

Severe NaOCl accidents such as extensive ecchymosis around the periorbital region and at the angle of the mouth and the neck are likely due to penetration of NaOCl into the facial venous system. Since anatomical variations in the venous system are not uncommon (Hollinshead 1968), drainage from the anterior teeth may occur into the facial vein via tributaries. More frequently, the drainage occurs into the pterygoid plexus. For NaOCl to enter the venous system, the central venous pressure must be exceeded (Lyons & Burwell 1938, Baumann et al. 2005, Zhu et al. 2013). This critical threshold corresponds to 5.73 mmHg (mean: 5.88 mmHg ± SD of 0.15 mmHg) and has been chosen in several studies investigating apical irrigant extrusion (Goode et al. 2013, Khan et al. 2013, Charara et al. 2016). The apical tissue pressure countering an extrusion is a
crucial factor, but it is considered to be individual and dependent on the clinical situation (Lorono et al. 2020).

Syringe irrigation is indispensable and remains the most commonly used method (Peters 2004, Zaugg et al. 2019). However, several irrigation methods aiming to improve irrigation are available. The ultrasonically activated irrigation (UAI) as well as sonic irrigation transmit acoustic energy from the file within the canal to the irrigant to enhance the efficacy of the irrigation process (van der Sluis et al. 2007, Neuhaus et al. 2016). UAI works at a frequency of 28 to 36 kHz and at an amplitude of 4 to 200 µm whereas sonic irrigation works at a frequency of up to 6.000 Hz and at an amplitude of 240 µm. The sonic irrigation device EDDY (VDW, Munich, Germany) is a flexible, non-cutting polyamide tip of 28 mm length with an ISO size of 20 and a .02 taper, that can be coupled to an air scaler (Neuhaus et al. 2016, Conde et al. 2017, Eggmann et al. 2020).

Laser-activation works via laser-pulses vaporizing the irrigant. The resulting bubbles collapse, creating shockwaves and generating cavitation effects (de Groot et al. 2009). Besides activation of the irrigant, novel methods for affecting fluid dynamics by suction have been developed. RinsEndo (Dürr Dental, Bietigheim-Bissingen, Germany) is a hydrodynamic irrigation method rinsing with alternate sequences of suction and pressure (Rodig et al. 2010). EndoVac (Kerr Dental, Orange, CA, USA) brings the irrigant into the pulp chamber by a master delivery tip. The irrigant reaches the apical regions because of negative pressure created by suction of a cannula that is placed within the main root canal close to the canal terminus (Nielsen & Baumgartner 2007). The macrocannula has an ISO size of 55, a taper of .02 and is used for irrigation of the pulp chamber and the coronal and middle segments of the canal during initial instrumentation. The microcannula has an ISO size of 32 and enables irrigation of the apical parts of the root canal system (Desai & Himel 2009).

In addition, coreless shaping instruments have been developed to perform more minimally invasive root canal preparations. The Self-Adjusting-File (SAF, ReDent NOVA, Berlin, Germany) adapts to the shape of the root canal and thus causes little alteration to its original form (Metzger et al. 2010, Lacerda et al. 2017). SAF works with simultaneous irrigation, the irrigant flows continuously through the hollow file, which produces favourable cleaning results (De-Deus et al. 2011). SAF ejects 4 mL of irrigant per minute and works at a speed of 5000 rpm. The XP-endo Finisher (FKG dentaire SA, La Chaux-de-Fonds, Switzerland) with its hook-shaped tip was designed for the final preparation and cleaning of complex root canal shapes (De-Deus et al. 2019).

Only a few studies have analysed irrigant extrusion using new irrigation devices. EndoVac appears to reduce extrusion compared to conventional irrigation in extracted mature teeth (Romualdo et al. 2017). By contrast, the results in terms of extrusion for other devices such as...
RinsEndo, UAI, laser activation, SAF and the XP-endo Finisher are inconclusive (Desai & Himel 2009, Mitchell et al. 2011, Boutsikouikis et al. 2013, Iriboz et al. 2015, Azim et al. 2018). Evidence on NaOCl extrusion in teeth with open apices is limited (dos Reis et al. 2020). The open apex situation is clinically relevant not only in immature teeth but also in resected teeth, teeth with root fractures, in teeth with extensive apical resorption and in teeth with an iatrogenically overprepared root canal beyond the apex when these teeth have to be root canal treated or retreated (Boutsikouikis et al. 2013).

Therefore, the aim of this laboratory study was to evaluate the impact of seven current methods of irrigant agitation on the apical pressure of sodium hypochlorite generated in teeth with an open apex. The research hypothesis was that all tested irrigation methods would never produce maximum pressure values exceeding the critical threshold of the central venous pressure (5.73 mmHg).

Materials and methods
Experimental setup
Experiments were performed on a 3D-printed central maxillary incisor. The printed tooth was based on micro computed tomography imaging data of an immature tooth 21. The cross section of the canal terminus had a minimal diameter of 1.57 mm, a maximal diameter of 2.11 mm, a circumference of 5.84 mm and an area of 2.49 mm². The total distance from the incisal edge to the canal terminus, henceforth referred to as tooth length (TL), was 17 millimetres. A 3D rendered (itk-SNAPE, University of Pennsylvania, PA, USA) image of the micro computed tomography data and a photography of the immature tooth is shown in Figure 1.

The printed tooth was mounted in a threaded cylinder using C-silicone putty material (Coltoflax; Coltène/Whaledent AG, Altstätten, Switzerland). The cylinder was placed on a pressure sensor, the enclosed chamber between apex and the membrane of the sensor-was filled completely with water. The generated pressures during irrigation were thus transferred directly and without any alteration to the sensor owing to the lack of compressibility of water under the conditions in this laboratory setup. The pressure sensor (IMP 320, ICS Schneider Messtechnik GmbH, Höhen Neuendorf, Germany) had a measurement range of 0 to 120 mmHg. At an analogue output signal of 20 mA, the sensor registered with a precision of 0.1 percent and had a response time of ≤ 0.5 ms. Measurement data were transformed by an analogue-digital converter (12bit, RedLab 1208FL USB Mini-Messlabor; Meilhaus Electronic GmbH, Alling, Germany) and transferred via USB cable to a personal computer. Data were processed with a laboratory software (LabVIEW; International Instruments AG, Austin, TX, USA) with a customised written protocol to read and save the obtained data. Figure 2 shows the experimental setup.
Irrigation protocols

Irrigation was performed with seven irrigation devices. The activation time of each measurement was 10 s since preliminary tests had shown maximum pressure values to be reached within this time period. The measurement per group was repeated ten times. Before irrigation, the root canal and the pulp chamber were completely filled with 0.1 mL 1% NaOCl (Hänseler AG, Herisau, Switzerland). During the irrigation period of 10 s, no additional NaOCl was added to the root canal system except for the irrigation with EndoVac. The hand pieces were fixed in a solid and adjustable clamp.

1. UAI was performed using smooth wire irrigation instruments, Irri S files, and the VDW.ULTRA unit (both VDW, Munich, Germany). Files with a size 25 and a length of 25 mm were used. Intensity levels of 10, 20 and 30 were employed (28 - 36 kHz).

2. Sonic irrigant activation was performed with EDDY, coupled to an air scaler handpiece, the SONICflex 2003 L handpiece (Kavo, Biberach, Germany), which was connected to a dental unit (Teneo; Dentsply Sirona, York, PA, USA). The air scaler handpiece used to drive the EDDY tips was employed at its highest power setting at all times (Eggmann et al. 2020).

3. EndoVac was tested with the macrocannula and the microcannula. Since EndoVac creates negative pressure by suction, NaOCl was added during the irrigation process. Following the manufacturer’s instructions, this was accomplished by a syringe. To monitor the force applied on the syringe plunger and to keep it as constant as feasible, the plunger of the syringe was equipped with a force sensor (KM25; Transmetra GmbH, Flurlingen, Switzerland). The force sensor had a measurement range of 0 to 100 N. Data were converted, processed, read and saved as described above. The force applied on the syringe plunger never exceeded 20 N throughout the experiment. Irrigation with the macrocannula required 2.5 mL additional NaOCl during the 10 s irrigation period, the microcannula required 0.3 mL additional NaOCl.

4. XP-endo Finisher files with a length of 25 mm and size 25 were used. A torque of 1 Ncm and the rotation speed of 1000 rpm were chosen according to the manufacturer’s recommendation.

5. The SAF was tested with a file of 21 mm in length and a diameter of 1.5 mm.

The five irrigation methods described above were tested at the following insertion depths: TL (i.e., flush to the canal terminus), TL – 1 mm, TL – 2 mm and TL – 3 mm.

6. LiteTouch Induced Photomechanical Irrigation (LT-IPI) was performed by using the Orcos medical litetouch III laser (Orcos medical AG, Küsnacht, Switzerland), an Er:YAG laser working at a wavelength of 2940 nm. Four different modes were tested, namely 20 mJ / 10 Hz, 20 mJ / 20 Hz, 20 mJ / 50 Hz and 40 mJ / 10 Hz. Laser activation was done with a bent handpiece and a tip with a length of 17 mm and a diameter of 0.4 mm. Each mode was

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performed at an insertion depth of 7 mm, which correlated with a position inside the coronal pulp chamber, and 11 mm, which correlated with the cementum-enamel junction / the root canal orifice.

7. RinsEndo, a hydrodynamic irrigation system working at a frequency of 1.6 Hz and ejecting 105 µL of irrigant per second, was used at an insertion depth of 7 mm, 11 mm, 12.5 mm (the coronal third of the root canal), 14 mm (the middle of the root canal) and at TL.

The workflow of the experiments is summarized in Figure 3.

Data for irrigation by syringe were obtained from another study where irrigation by syringe was performed in the same experimental setup as described above (Jäggi et al. 2021). Irrigation by syringe was performed with a 30G side-vented cannula, an applied pressure on the syringe plunger of 40 N and at an insertion depth of TL – 1 mm.

**Critical value and statistics**

A descriptive statistical analysis was performed. The maximum pressure values of each measurement and the 95% confidence intervals were subjected to the analysis. To gauge the risk of apical NaOCl extrusion, the lower limit of the mean central venous pressure of 5.73 mmHg (mean: 5.88 mmHg ± SD of 0.15 mmHg) was defined as the threshold. Pressure values exceeding this critical threshold were considered indicative of potential apical NaOCl extrusion.

**Results**

Irrespective of the insertion depth, EndoVac, the SAF, the XP-endo Finisher and UAI, across all its intensity levels (28 – 36 kHz), never exceeded the threshold value of 5.73 mmHg.

LT-IPI did not exceed the threshold value at any insertion depth when used at frequencies ≤20 Hz (the manufacturer recommends a frequency of 10 to 15 Hz for irrigant activation). Irrigant activation at an intensity level of 20 mJ / 50 Hz and an insertion depth to the pulp chamber resulted in the threshold value being exceeded in 10% of the measurements. At 20 mJ / 50 Hz and an insertion depth to the root canal orifice, 50% of the measurements were above the threshold value.

EDDY used at TL exceeded the upper limit of the central venous pressure of 6.03 mmHg in 100% of the measurements. Pressure values above the threshold value of 5.73 mmHg were observed in 30%, 10% and 20% of the measurements when the EDDY tip was placed at TL – 1 mm, TL - 2 mm and TL – 3 mm, respectively.

100% of the measurements with RinsEndo lay over the the upper limit of the central venous pressure of 6.03 mmHg, irrespective of the insertion depth.
Table 1 shows the detailed results of all irrigation methods in relation to the lower and upper limit of the central venous pressure (5.73 mmHg / 6.03 mmHg). Also, data for irrigation by syringe is shown in Table 1.

Figure 4 shows a representative graph for each irrigation method outlining the changes in the apical pressure through the irrigation period of 10 s.

Discussion

This laboratory study evaluated the apical pressure of NaOCl generated during irrigation with seven irrigation methods in a simulated tooth with an open apex. EndoVac, the SAF, the XP-endo Finisher and UAI produced pressures below the critical threshold. Laser-activation (LT-IPI) produced pressures below the critical threshold as long as the intensity level is chosen according to the manufacturer’s recommendation. Irrigation with RinsEndo, by contrast, produced pressures far above the critical threshold. Likewise, sonic activation with EDDY created pressures exceeding the threshold. Thus, irrigant activation using RinsEndo and EDDY may lead to irrigant reaching the periapical tissue, potentially causing NaOCl extrusion accidents in teeth with open apices. Consequently, irrigant activation using RinsEndo and EDDY should be used with caution in immature teeth.

It remains elusive which apical pressure threshold needs to be exceeded for irrigant extrusion to occur to an extent that is clinically relevant (Park et al. 2013). The present investigation, in accordance with previous studies, chose the central venous pressures as a reference. It is, however, important to bear in mind that different threshold values may be applicable. Threshold values used in other studies are 25 mmHg representing the capillary pressure, 20-30 mmHg representing the interstitial pressure and 30 mmHg representing the intraosseous blood pressure (Goode et al. 2013, Khan et al. 2013, Park et al. 2013, Zhu et al. 2013, Charara et al. 2016, Lorono et al. 2020). The lowest reference value reported in the literature was chosen in the present study in order to err on the side of caution when providing recommendations. The results differ when shifting the threshold to other values mentioned in the literature (20 – 30 mmHg). Sonic activation with EDDY, in particular, generated maximal values that seem to be acceptable when considering critical values that are above the central venous pressure.

Theories concerning the conditions enabling serious ecchymotic NaOCl accidents have been discussed in several studies. As a necessary condition, the apical foramen has to be patent for the irrigant to extrude (Zhu et al. 2013). The setup in the present study, using a simulated tooth with a wide open apex, replicated this condition. Furthermore, due consideration must be given to individual conditions that may affect the periapical pressure. In maxillary teeth as well as in females in general, the counterpressure may be affected by lower bone density (Boutsioukis et al. 2016).
The periapical pressure may thus likely be lower. In addition, the location of the root apex in the maxillary sinus or towards other soft tissues may favour an extrusion. The pulp and periapical status prior to the extrusion have been reported to have an influence. Presumably, situations with a periapical pathosis and situations with healthy periapical tissues are dissimilar (Kleier et al. 2008, Boutsioukis et al. 2013).

Other studies examining irrigant extrusion neglected the presence of periapical tissues, for instance by surrounding the apical foramen only by atmospheric air (Boutsioukis et al. 2013). In the present study, extrusion was quantified as pressure in mmHg. This may be methodologically advantageous because the reference point can be adjusted if and when new evidence on periapical pressures emerges.

The generated pressures were recorded at an interval of 10 ms by a high-precision pressure sensor with a metering precision of 0.1 percent and a response time of \( \leq 0.5 \) milliseconds within a measuring range of 0 – 120 mmHg. Negative pressures were recorded (irrigation using EndoVac), too. Negative pressure readings must, however, be treated with caution as the pressure sensor was not calibrated to register negative pressures.

As the recommendations of the manufacturers concerning the insertion depths of the irrigation device into the root canal system varied and are related to the so-called working length in teeth with fully formed apices, several insertion depths were tested. The respective manufacturers recommend to insert EDDY one millimetre short of full working length (WL), to insert EndoVac, the XP-endo Finisher and SAF to working length, to insert the special cannula of RinsEndo to the coronal third of the root canal and to insert the tip of the LT-IPI into the coronal pulp chamber. The insertion depths in the present study were on one hand chosen to comply with the manufacturer’s recommendations but also to induce potential outliers (e.g., insertion of RinsEndo to TL). Owing to these considerations, LT-IPI was not only tested at 10 and 15 Hz in accordance with the manufacturer’s recommendation but also at 20 and 50 Hz. Mounting the irrigation devices into a solid and adjustable clamp provided constant defined and reproducible insertion depths.

There are only a few previous studies investigating irrigant delivery and activation with different irrigation devices in immature teeth. The results of the current study are in agreement with the results of previous studies. dos Reis et al. (2020) reported higher extrusion values when sonic agitation with the EDDY instrument was performed. On the contrary, the XP-endo Finisher produced less irrigant extrusion than EDDY, comparable to the extrusion when irrigating by syringe (dos Reis et al. 2020).

Irrigation by syringe was tested in the same setting in another study (Jäggi et al. 2021). The apical pressures were dependent on the diameter of the cannula (G25 versus G30), the needle
tip design (open-ended versus side-vented), the insertion depth (TL, TL – 1 mm, TL – 2 mm, TL – 4 mm) and the applied pressure on the syringe plunger (10 N, 20 N, 40 N, 80 N). When considering conditions realistic to daily practice (applied pressures on the syringe plunger: 40 N, irrigation with an G30 side-vented cannula, insertion depth: TL – 1 mm) the maximum value amounted 2.64 mmHg and the mean 1.71 mmHg. Conventional irrigation performed as mentioned results in higher apical pressures than EndoVac, UAI, the SAF and the XP-Endo finisher at an insertion depth of TL – 1 mm. Nevertheless, the critical value of 5.73 mmHg was never exceeded when performing conventional irrigation as mentioned above.

Although severe NaOCl accidents are considered to be rare, the issue of irrigant extrusion is of crucial importance. NaOCl accidents can have grave consequences, first and foremost for the patients who are affected, but also for the dentists involved (Zhu et al. 2013). In a survey, almost half of the endodontists in the US reported to have experienced at least one accident in their practice (Kleier et al. 2008). The present study may provide information to help to minimise the risk for such accidents.

The present study indicates that, in immature teeth and in other cases with open apices, the various irrigant methods tested have little risk for apical irrigant extrusion. These laboratory data suggest that NaOCl, with its numerous advantageous properties, can be used as irrigant in teeth with open apices when one of the recommended methods (EndoVac, UAI, SAF, XP-Endo finisher) is employed for improving disinfection and cleaning of the root canal without risk of apical extrusion when used cautiously.

Conclusion
In a maxillary central incisor with an open apex, irrigation with LT-IPI respecting the manufacturer’s recommendation, EndoVac, UAI, the SAF, and the XP-endo Finisher generate periapical pressures that were below the critical threshold of the central venous pressure (5.73 mmHg).

Performing irrigation with RinsEndo or EDDY in teeth with an open apex produced pressures higher than the critical threshold. The safety of these devices in immature teeth therefore warrants further investigation.

Conflict of Interest statement
The authors have stated explicitly that there are no conflicts of interest in connection with this article.
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Figure legends

Figure 1 (a) 3D printed immature tooth 21 and (b) 3D rendered image of the micro computed tomography data in different aspects.

Figure 2 Experimental setup: (a) Irrigation device in an adjustable clamp. (b) 3D-printed central incisor on the pressure sensor. (c) analogue-digital converter. (d) computer with laboratory software. (e) 3D-printed central incisor embedded in a threaded cylinder by silicone. (f) Pressure sensor with (g) pressure membrane.

Figure 3 Workflow: setting and insertion depth of the seven irrigation methods. (a) Irrigation method and setting. (b) Insertion depth. (c) Irrigation. (d) Measurement. (e) Data processing.

Figure 4 Representative graph for each irrigation method. The pressure values are not true to scale for a better understanding of the pressure gradients.
Table 1 Results: Maximum values, mean, standard deviation (SD), 95% confidence interval (CI), percentage of measurements exceeding 5.73 mmHg per irrigation method; maximum value above 6.03 mmHg (red), maximum value above 5.73 mmHg (orange). Data for syringe irrigation was obtained from a previous study (Jäggi et al. 2021).

| Irrigation | Setting | Insertion depth | Maximum (mmHg) | Mean (mmHg) | SD (mmHg) | 95%-CI | Percentage > 5.73 mmHg (%) |
|------------|---------|----------------|----------------|-------------|-----------|--------|----------------------------|
| UAI        | 10      | TL             | 0.47           | 0.28        | 0.13      | 0.19 – 0.38 | 0 |
|            | (intensity) | TL – 1 mm     | 0.41           | 0.19        | 0.10      | 0.11 – 0.26 | 0 |
|            |         | TL – 2 mm      | 0.23           | 0.08        | 0.06      | 0.04 – 0.13 | 0 |
|            |         | TL – 3 mm      | 0.29           | 0.17        | 0.08      | 0.12 – 0.23 | 0 |
| 20         | TL      | (intensity)    | 1.17           | 0.77        | 0.32      | 0.54 – 0.98 | 0 |
|            | TL – 1 mm | 0.77           | 0.42           | 0.17        | 0.08      | 0.30 – 0.54 | 0 |
|            | TL – 2 mm | 0.35           | 0.17           | 0.08        | 0.12 – 0.23 | 0 |
|            | TL – 3 mm | 0.41           | 0.17           | 0.18        | 0.03 – 0.23 | 0 |
| 30         | TL      | (intensity)    | 1.23           | 0.91        | 0.26      | 0.73 – 1.13 | 0 |
|            | TL – 1 mm | 1.00           | 0.67           | 0.23        | 0.50 – 0.83 | 0 |
|            | TL – 2 mm | 0.47           | -0.01          | 0.40        | -0.29 – 0.28 | 0 |
|            | TL – 3 mm | 0.65           | 0.09           | 0.50        | -0.26 – 0.44 | 0 |
| EDDY       | TL      | 9.79           | 8.48           | 0.90        | 7.80 – 9.15 | 100 |
|            | TL – 1 mm | 6.56           | 4.82           | 1.28        | 3.90 – 5.78 | 30 |
|            | TL – 2 mm | 6.39           | 3.77           | 1.37        | 2.78 – 4.73 | 10 |
|            | TL – 3 mm | 6.44           | 4.99           | 0.92        | 4.28 – 5.55 | 20 |
| EndoVac    | Macrocannula | TL             | 0.12           | -0.14       | 0.13      | -0.23 – 0.05 | 0 |
|            | TL – 1 mm | 0.06           | -0.16          | 0.11        | -0.24 – 0.08 | 0 |
|            | TL – 2 mm | 0.00           | -0.12          | 0.08        | -0.17 – 0.06 | 0 |
|            | TL – 3 mm | 0.00           | -0.17          | 0.08        | -0.23 – 0.07 | 0 |
|            | Microcannula | TL             | 1.88           | 1.45        | 0.20      | 1.28 – 1.58 | 0 |
|            | TL – 1 mm | 2.05           | 1.64           | 0.32        | 1.35 – 1.80 | 0 |
|            | TL – 2 mm | 2.64           | 1.82           | 0.50        | 1.43 – 2.18 | 0 |
|            | TL – 3 mm | 2.46           | 2.21           | 0.38        | 1.65 – 2.25 | 0 |
| RinsEndo   | TL      | 107.65         | 96.71          | 11.66       | 88.36 – 105.08 | 100 |
|            | Middle of root canal | 107.30       | 96.94          | 10.72       | 89.26 – 104.63 | 100 |
|            | Coronal third of root canal | 64.46     | 56.19          | 4.28        | 53.10 – 59.25 | 100 |
|            | Root canal orifice | 71.90      | 57.10          | 7.65        | 51.60 – 62.56 | 100 |
|            | Coronal pulp chamber | 46.59   | 37.77          | 7.23        | 32.55 – 42.83 | 100 |
| LT-IPI     | 20mJ/10Hz | Root canal orifice | 1.94           | 1.52        | 0.24      | 1.35 – 1.65 | 0 |
|            | Pulp chamber | 1.17          | 0.91           | 0.14        | 0.83 – 0.98 | 0 |
|            | 20mJ/20Hz | Root canal orifice | 4.04           | 3.31        | 0.44      | 3.00 – 3.60 | 0 |
|                | Pulp chamber | Root canal orifice | Pulp chamber | Root canal orifice | Pulp chamber |
|----------------|--------------|-------------------|--------------|-------------------|--------------|
| **20mJ/50Hz**  | 2.81         | 6.62              | 5.80         | 4.34              | 2.64         |
|                | 2.13         | 5.66              | 4.35         | 3.83              | 2.17         |
|                | 0.47         | 0.68              | 0.84         | 0.29              | 0.34         |
|                | 1.80 – 2.48  | 5.18 – 6.15       | 3.75 – 4.95  | 3.60 – 4.05       | 1.95 – 2.40  |
| **40mJ/10Hz**  | 50           | 10                | 0            | 0                 | 0            |
| **SAF**        | 0            | 0                 | 0            | 0                 | 0            |
| **XP-endo**    | 0            | 0                 | 0            | 0                 | 0            |
| **Finisher**   | 0            | 0                 | 0            | 0                 | 0            |
| **Syringe**    | 0            | 0                 | 0            | 0                 | 0            |
| **TL**         | 1.23         | 0.82              | 1.00         | 0.71              | 1.00         |
| **TL - 1mm**   | 0.88         | 0.69              | 1.00         | 0.70              | 1.00         |
| **TL - 2mm**   | 1.00         | 0.70              | 1.00         | 0.71              | 1.00         |
| **TL - 3mm**   | 1.00         | 0.70              | 1.00         | 0.71              | 1.00         |
| **TL - 1mm**   | 2.64         | 2.17              | 0.51         | 0.11              | 0.51         |
| **TL - 2mm**   | 0.12         | 0.08              | 2.64         | 1.10              | 0.12         |
| **TL - 3mm**   | 0.17         | 0.11              | 0.12         | 0.09              | 0.17         |
| **40 N (force)** | 0.09 – 0.32  | 0.05 – 0.10       | 0.07 – 0.11  | 0.08 – 0.14       | 0.09 – 0.32  |
| **TL - 1mm**   | 0.09 – 0.32  | 0.05 – 0.10       | 0.07 – 0.11  | 0.08 – 0.14       | 0.09 – 0.32  |
| **TL - 2mm**   | 0.07 – 0.11  | 0.04              | 0.08 – 0.14  | 0.08 – 0.14       | 0.07 – 0.11  |
| **TL - 3mm**   | 0.07 – 0.11  | 0.04              | 0.08 – 0.14  | 0.08 – 0.14       | 0.07 – 0.11  |
