MicroRNA-296: a promising target in the pathogenesis of atherosclerosis?

Heng Li 1†, Xin-Ping Ouyang 2,3†, Ting Jiang 4, Xi-Long Zheng 5,8, Ping-Ping He 2,6* and Guo-Jun Zhao 5,7*

Abstract

Atherosclerosis has been recognized as an inflammatory disease involving the vascular wall. MicroRNAs are a group of small noncoding RNAs to regulate gene expression at the transcriptional level through mRNA degradation or translation repression. Recent studies suggest that miR-296 may play crucial roles in the regulation of angiogenesis, inflammatory response, cholesterol metabolism, hypertension, cellular proliferation and apoptosis. In this review, we primarily discussed the molecular targets of miR-296 involved in the development of atherosclerosis, which may provide a basis for future investigation and a better understanding of the biological functions of miR-296 in atherosclerosis.

Keywords: miR-296, Angiogenesis, Inflammation, Atherosclerosis

Introduction

The atherosclerotic cardiovascular disease is the main cause of death in developing and industrialized countries (Barquera et al. 2015). The pathogenesis and development of atherosclerosis are mechanistically associated with many pathophysiological factors, such as angiogenesis, inflammatory response, cholesterol metabolism, hypertension, cellular proliferation and apoptosis (Buckley and Ramji 2015; Camaré et al. 2017).

MicroRNAs (miRNAs) are endogenous small (20–22 nucleotides long) non-coding RNAs that can post-transcriptionally regulate metazoan gene expression. The underlying mechanisms involve either induction of mRNA degradation or direct inhibition of protein translation (Andreou et al. 2015; Selbach et al. 2008). MiRNA regulation is characterized by its active participation in and strict control of the negative feedback loop, which confers significant influences on a biological pathway (Ebert and Sharp 2012; Laffont and Rayner 2017). To date, accumulating experimental evidence indicates that miRNAs, which are involved in every stage of biological processes and the changes of their expression and/or function have been associated with the initiation and progression of atherosclerosis (Giral et al. 2016; Laffont and Rayner 2017).

MiR-296 is a family of microRNA precursors found in mammals, including humans, which is located in the chromosome 20q13.32 genomic locus. The miR-296 precursor generally gives rise to mature miR-296. It is called miRNA-296-5p if derived from the 5′ arm, and miRNA-296-3p if derived from the 3′ arm (Fig. 1). In most, but not all, tissues, the expression of miR-296-5p is higher than miR-296-3p (Robson et al. 2012). There has been a close association between miR-296 expression and cardiovascular diseases. MiR-296-5p is upregulated in patients with hypertrophic cardiomyopathy (Fang et al. 2015). Wei et al. also found that miR-296-5p is one of the five miRNAs that were significantly elevated in early plaque formation and did not change significantly in late plaques in apoE−/− mice fed high-cholesterol diet, suggesting that miR-296-5p may be a risk factor for atherosclerosis (Wei et al. 2013).

Studies have shown that miR-296 can be expressed in endothelial cells, macrophages and vascular smooth muscle cells, which are the major cell types involved in the development of atherosclerotic plaque (Feng et al. 2015; Meng et al. 2014; Reddy et al. 2016; Zheng et al. 2015). The atheromatous lesions showed distinct VEGF positivity of activated endothelial cells (Inoue et al. 1998) and VEGF alone is capable of increasing miR-296, (Würdinger et al. 2008) suggesting miR-296 may be expressed in endothelial cells in atherosclerotic lesions. Latent tuberculosis infection (LTBI) is associated with persistent chronic inflammation...
and has pro-atherogenic effect in human vessels (Huaman et al. 2015). Identification of LTBI infection-related miRNAs in human macrophages showed that miR-296-5p was significantly increased than the uninfected cells (Meng et al. 2014; Zheng et al. 2015). Type 1 and type 2 diabetes are associated with an enhanced inflammatory state and that inflammatory cells contribute to atherosclerotic lesion initiation and lesion disruption (Kanter et al. 2008). Reddy and colleagues found that miR-296 was upregulated in db/db VSMC compare to db/+ VSMC via small RNA-sequencing (Reddy et al. 2016). However, the function of miR-296, especially its role in atherosclerosis, has not been extensively studied.

MiR-296 has many bioinformatics targets that have been predicted and biologically validated in human and mouse (Table 1), suggesting that miR-296 has a wide range of potential biological functions. In this overview, we summarized the current knowledge regarding the potential roles of miR-296 in atherosclerosis (Table 2) and tried to provide a basis for future investigation and expand our insights into miR-296 functions in atherosclerosis.

**MiR-296 in angiogenesis**

Neovascularization in atherosclerotic lesions plays an important role in plaque growth and instability (Camaré et al. 2017). MiR-296 belongs to the family of “angiomirs” due to its functions associated with angiogenesis (Würdinger et al. 2008). MiR-296 and angiogenesis were significantly upregulated following cerebral ischemic injury (Feng et al. 2015). The formation of capillary-like structures of human umbilical vein endothelial cells (HUVEC) was markedly enhanced by adenosine-mediated overexpression of miR-296 (Feng et al. 2015). The sequence-specific inhibition of miR-296 by intravenous injection of cholesterol-conjugated antagonist can reduce neovascularization in a mouse tumor model (Würdinger et al. 2008).

The vascular endothelial growth factor (VEGF) and Notch signaling pathways are crucial in angiogenesis that is promoted by VEGF and inhibited by Notch under most developmental or pathological conditions (Kim et al. 2017; Mao and Meng 2017). Overexpression of miR-296 was observed to increase the expression levels of VEGF and vascular endothelial growth factor receptor 2 (VEGFR2) and simultaneously reduce the expression levels of delta-like ligand 4 (DLL4) and Notch1 (Feng et al. 2015). One of the miR-296 targets is hepatocyte growth factor-regulated tyrosine kinase substrate (HGS). MiR-296 can reduce the levels of HGS and HGS-mediated degradation of VEGFR2 and platelet-derived growth factor receptor-β (PDGFR-β) (Würdinger et al. 2008). VEGFR and PDGFR are the targets of anti-angiogenic therapies (Batchelor et al. 2007; Shih and Holland 2006). The Notch pathway lies downstream of the VEGF signaling, and the activation of VEGF signaling increases the expression of Notch ligands DLL4 (Liu et al. 2014; Lobov et al. 2007). This finding is contradictory to the research showing that miR-296 promotes VEGF signaling and inhibits the Notch pathway. It is possible that there is another target gene for miR-296, which downregulates the expression of DLL4 and Notch. Further research is needed to elucidate the mechanisms by which miR-296 regulates the angiogenesis.

**MiR-296 in inflammatory response**

MiR-296 regulates NF-κB pathway

In miR-296 overexpressed HCT116 cells, western blot analysis showed that nuclear transcription factor-κB (NF-κB) P65 protein was decreased, while the pP65 level was elevated with enhanced nuclear import. On the contrary, NF-κB P65 level was decreased in cells with low expression of miR-296 (Zhai et al. 2016). Therefore, these findings suggest that miR-296 plays an important role in regulating NF-κB signaling pathway.

In mammals, there is a Numb homolog called “Numb-like” or NumbL, that is a cytoplasm protein and has redundant functions in embryonic neurogenesis. The studies have shown that overexpression of NumbL can inhibit NF-κB...
activation induced by TNFα and IL-1β, and knockdown of NumbL promotes NF-κB activation, (Ma et al. 2008) suggesting that NumbL plays a negative effect on NF-κB activation. Transfection of A549 NSCLC (non-small cell lung cancer) cells with miR-296-5p was found to inhibit the mRNA and protein expression of NumbL. Two hypothetical miR-296-5p -responsive sites were predicted in the NumbL 3′ UTR. The combined mutagenesis of these two sites reversed miR-296-5p repression of NumbL regulatory sequences, suggesting that NumbL is a direct gene target of miR-296-5p (Vaira et al. 2013). Taken together, the current findings suggest that miR-296 may play an inflammatory role by targeted inhibition of NumbL and subsequent NF-κB activation.

**Table 1** Bioinformatics targets of miR-296 that were predicted and biologically validated in human and mouse

| miRNA Name     | Target gene  | Gene Description                        | Bioinformatics prediction tool | Biological validation |
|----------------|--------------|-----------------------------------------|--------------------------------|-----------------------|
| hsa-miR-296-5p | HMGA1        | high mobility group AT-hook 1           | miRDB, TargetScan              | Yes                   |
| hsa-miR-296-5p | NUMBL        | numb homolog (Drosophila)-like          | miRDB, TargetScan              | Yes                   |
| hsa-miR-296-5p | BOK          | BCL2-related ovarian killer             | miRDB                          | Yes                   |
| hsa-miR-296-3p | CXCR1        | chemokine (C-X-C motif) receptor 1      | miRDB                          | Yes                   |
| mmu-miR-296-5p | Pin1         | protein (peptidyl-prolyl cis/trans isomerase) NIMA-interacting 1 | miRDB                          | Yes                   |
| hsa-miR-296-3p | PTEN         | phosphatase and tensin homologue        | TargetScan                     | Yes                   |
| hsa-miR-296-5p | NGFR         | nerve growth factor receptor            | TargetScan                     | Yes                   |
| hsa-miR-296-5p | FGFR1        | fibroblast growth factor receptor 1     | TargetScan                     | Yes                   |
| hsa-miR-296-3p | SOCS6        | suppressor of cytokine signaling 6      | miRDB, TargetScan              | No                    |
| hsa-miR-296-5p | RNF44        | ring finger protein 44                  | miRDB, TargetScan              | No                    |
| hsa-miR-296-5p | EPN1         | epsin 1                                 | miRDB, TargetScan              | No                    |
| hsa-miR-296-3p | ZBTB20       | zinc finger and BTB domain containing 20 | miRDB, TargetScan              | No                    |
| mmu-miR-296-3p | ZBTB20       | zinc finger and BTB domain containing 20 | miRDB, TargetScan              | No                    |
| mmu-miR-296-3p | STRN3        | striatin, calmodulin binding protein 3  | miRDB, TargetScan              | No                    |
| hsa-miR-296-5p | LYPLA2       | lysophospholipase 2                     | miRDB, TargetScan              | No                    |
| mmu-miR-296-5p | DYRK1B       | dual-specificity tyrosine-(Y)-phosphorylation regulated kinase 1b | miRDB, TargetScan              | No                    |
| mmu-miR-296-5p | LYPLA2       | lysophospholipase 2                     | miRDB, TargetScan              | No                    |
| mmu-miR-296-5p | Srf          | serum response factor                   | miRDB, TargetScan              | No                    |
| mmu-miR-296-5p | SRRM3        | serine/arginine repetitive matrix 3      | miRDB, TargetScan              | No                    |
| hsa-miR-296-3p | LMLN         | leishmanolysin-like (metalloproteinase M8 family) | miRDB                          | No                    |
| hsa-miR-296-3p | KLHL23       | kelch-like family member 23             | miRDB                          | No                    |
| hsa-miR-296-3p | ZNF117       | zinc finger protein 117                 | miRDB                          | No                    |
| hsa-miR-296-3p | CADPS        | Ca++-dependent secretion activator      | miRDB                          | No                    |
| hsa-miR-296-3p | NTF3         | neurotrophin 3                          | miRDB                          | No                    |
| hsa-miR-296-5p | TEAD3        | TEA domain family member 3              | TargetScan                     | No                    |
| hsa-miR-296-5p | BBC3         | BCL2 binding component 3                | TargetScan                     | No                    |
| hsa-miR-296-5p | ISCA2        | iron-sulfur cluster assembly 2          | TargetScan                     | No                    |
| hsa-miR-296-5p | RIMS4        | regulating synaptic membrane exocytosis 4 | TargetScan                     | No                    |
| mmu-miR-296-3p | SOCS6        | suppressor of cytokine signaling 6      | TargetScan                     | No                    |

**MiR-296 regulates IKBKE expression**

The inhibitor of κB kinase epsilon (IKBKE, IKKe, or IKKi) has been identified as an emerging modulator that limits chronic inflammation during metabolic disease and atherosclerosis. IKBKE is necessary to limit both the magnitude and temporal dynamics of Nod-like receptor protein 3 (NLRP3) inflammasome priming in macrophages. It has been reported that Ikbke−/− macrophages can secrete a large amount of IL-1β. Furthermore, IKBKE ablation in apoE−/− mice enhances NLRP3 inflammasome priming and promotes atherosclerosis (Patel et al. 2015). MiR-296-5p may play a negative role in the regulation of IKBKE by targeted inhibition. Robson et al. found that miR-296-5p transfection represses luciferase expression of the PGL3-
IKBKE luciferase vector, whereas anti-miR-296-5p transfection activates luciferase expression of the PGL3-IKBKE luciferase vector (Robson et al. 2012). In addition, it was reported that miR-296-5p does not have any remarkable effect on pGL3-IKBKE-3′UTR-mut construct with a mutated target site (Robson et al. 2012). Thus, it is possible that miR-296-5p may promote inflammation and atherosclerosis through directly targeting IKBKE.

MiR-296 regulates macrophage polarization

It has been widely acknowledged that M1-polarized macrophages play a pro-inflammatory role in atherosclerosis, whereas M2-polarized macrophages possess an anti-inflammatory effect (De Paoli et al. 2014; Tabas and Bornfeldt 2016). Suppressors of cytokine signaling (SOCS) may be a candidate for controlling leukocyte differentiation and helping shape inflammatory reactions. Recent studies have shown that SOCS facilitates macrophage development and polarization by regulating cytokine expression. SOCS-2 can direct the polarization toward M2 macrophages, and the absence of SOCS-2 resulted in increased polarization toward M1 macrophages (Spence et al. 2013). Further studies have demonstrated that miR-296 can significantly suppress Scrib protein expression by inhibiting Scrib 3′UTR activity, in which Scrib was validated as a target gene directly suppressed by miR-296. These results suggest that miR-296 promotes macrophage polarization toward M1 macrophages probably by repressing Scrib expression.

MiR-296 regulates adhesion molecules

Adhesion molecules play an important role in mediating the adhesion of leukocytes and monocytes to endothelial cells, which can accelerate atherosclerosis development. Intercellular adhesion molecule-1 (ICAM-1) is considered as one of the important risk factors involved in atherosclerosis progression. Liu et al. observed that exogenous

### Table 2 The target genes of miR-296 and its roles in atherosclerosis

| Target gene | Pathways | Function | Potential effect in As | Ref. |
|-------------|----------|----------|------------------------|------|
| HGS         | PDGFR-β,VEGFR2 | pro-angiogenesis | ↑ | (Würdinger et al. 2008) |
| ?           | DI4/Notch | pro-angiogenesis | ↑ | (Feng et al. 2015) |
| NUMBL       | NF-kB    | pro-inflammatory | ↑ | (Zhai et al. 2016; Vaira et al. 2013) |
| IKBKE       | NLRP3    | pro-inflammatory | ↑ | (Robson et al. 2012; Patel et al. 2015) |
| SOCS-2      | M1 macrophage polarization | pro-inflammatory | ↑ | (Spence et al. 2013; Lee et al. 2016) |
| Scrib       | M1 macrophage polarization | pro-inflammatory | ↑ | (Zheng et al. 2016; Vaira et al. 2012) |
| Scrib       | ROS      | pro-inflammatory | ↑ | (Zheng et al. 2016; Vaira et al. 2012) |
| ICAM-1      | directly | anti-adhesion | ↓ | (Liu et al. 2013) |
| CX3CR1      | directly | anti-adhesion | ↓ | (Wouters et al. 2017; Luo and Lin 2016) |
| MDR1        | directly | intestinal cholesterol reabsorption | ↑ | (Tous et al. 2005; Hong et al. 2010) |
| WNK4        | directly | hypertension | ↓ | (Takahashi et al. 2017; Mao et al. 2010; Li et al. 2011) |
| p21WAF1     | directly | proliferation | ↑ | (Khanna 2009; Yang et al. 1996; Yoon et al. 2011) |
| p53         | p21WAF1  | proliferation | ↑ | (Guevara et al. 1999; Yoon et al. 2011) |
| p27         | directly | proliferation | ↑ | (Hong et al. 2010; Diez-Juan and Andres 2001) |
| PTEN        | directly | proliferation | ↑ | (Yuan et al. 2012; Li et al. 2009; Chen et al. 2004; Huang and Kontos 2002; Kakizaki et al. 2017) |
| NGFR        | directly | anti-apoptosis | ↑ | (Chaldakov et al. 2001; Wang et al. 2000; Lee et al. 2016) |
| Caspase-8   | directly | anti-apoptosis | ↑ | (Lee et al. 2016) |

↑, stimulatory effect; ↓, inhibitory effect; As, atherosclerosis

miR-296-3p may promote inflammation and atherosclerosis through directly targeting IKBKE.
expression of miR-296-3p significantly inhibits the luciferase activity of the wild-type 3' UTR of the human ICAM-1 reporter, but does not affect the luciferase activity of the mutant reporter. Furthermore, ICAM-1 protein is down-regulated in P69-miR-296-3p cell lines and is reversely up-regulated in M12-anti-miR-296-3p cell lines, as analyzed by both flow cytometry and immunoblotting (Liu et al. 2013). These results demonstrate that miR-296-3p inhibits the expression of ICAM-1 and may be involved in the regression of atherosclerosis.

CX3C chemokine receptor 1 (CX3CR1) has been identified as an important adhesion molecule in migration, adhesion and recruitment of monocytes during the pathogenesis of atherosclerosis (Wouters et al. 2017) CX3CR1 was validated as one of the direct target genes of miR-296-3p. Dual reporter assays have shown that introduction of miR-296-3p in A549 cells inhibits the activity of a luciferase reporter fused to the wild-type 3' UTR of CX3CR1, but does not inhibit that of a reporter fused to a mutant version of the 3' UTR. The introduction of miR-296-3p in A549 and H157 cells reduces CX3CR1 expression at the mRNA and protein levels (Luo and Lin 2016). These results suggest that miR-296-3p may inhibit monocyte adhesion through targeting CX3CR1.

**MiR-296 in cholesterol metabolism**

Multidrug resistance protein 1 (MDR1), also known as ATP-binding cassette sub-family B member 1 (ABCB1) or P-glycoprotein 1, is an important protein of cell membrane that functions as a regulator of cholesterol metabolism. In vitro and in vivo studies have shown that MDR1 could redistribute cholesterol from the inner leaflet of the membrane to the outer leaflet (Garrigues et al. 2002) or from the membrane to the endoplasmic reticulum to enhance cholesterol esterification (Batetta et al. 2001). Furthermore, MDR1 could be involved in the intestinal cholesterol re-absorption and the susceptibility to atherosclerosis (Tous et al. 2005). The studies have shown that downregulation of miR-296 results in a significant decrease in MDR1 expression. In addition, cotransfection of the MDR1 reporter gene with an increasing amount of antagonirs of miR-296 results in a substantially linear decrease in MDR1 promoter activity, indicating that MDR1 may be a target gene for miR-296 (Hong et al. 2010). However, the mechanism for miR-296 to up-regulate MDR1 is not clear. It was assumed that the effects of miR-296 might depend on not only the promoter sequences but also the association of miR-296 with different cofactors. These studies suggest miR-296 may regulate cholesterol metabolism and promote the formation of atherosclerotic lesions.

**MiR-296 in hypertension**

With-no-lysine kinase 4 (WNK4) regulates electrolyte homeostasis and blood pressure. WNK4 is a hypertension causing gene involved in the regulation of salt-sensitive hypertension (Takahashi et al. 2017). Human WNK4 (hWNK4) 3' UTR acts as the enhancer through distant crosstalk with the hWNK4 promoter in a cell-specific manner. Meanwhile, the hWNK4 3' UTR contains a miR-296 binding site as predicted by bioinformatics algorithms and confirmed by a reporter assay. MiR-296 was able to down-regulate the expression of hWNK4 at the posttranscriptional level in cell-specific pattern as shown by real-time quantitative PCR and western blot assays (Mao et al. 2010). These data demonstrate that miR-296 may regulate blood pressure by suppressing hWNK4 expression through targeting on its 3' UTR. In addition, Li et al. analyzed that the expression levels of some selected miRNAs, including miR-296-5p, in circulating endothelial cells and endothelial progenitor cells and found that the expression of miR-296-5p is lower in hypertensive patients than healthy control subjects, (Li et al. 2011) suggesting its potential role in the regulation of blood pressure.

**MiR-296 in cellular proliferation and apoptosis**

MiR-296 regulates cell proliferation

It has been shown that overexpression of miR-296-5p significantly promotes gastric cancer growth, (Li et al. 2014) suggesting that miR-296-5p may have a role in cell proliferation. It is known that induction of endogenous p53, (Guevara et al. 1999) p21WAF1 (Khanna 2009; Yang et al. 1996) or p27(Diez-Juan and Andres 2001) limits the hyperplastic growth of vascular smooth muscle cells (VSMCs) and macrophages and protects against diet-induced atherosclerosis. In addition, p53/p21 axis plays a central role in the DNA damage response and cell cycle control (Pelletier et al. 2012). Yoon and his colleagues reported that synthetic pre-miR-296-transfected cells show the decreased levels of p53 and p21WAF1 transcripts, and anti-miR-296 increases p53 and p21WAF1 transcript levels. By monitoring the activities of a luciferase reporter connected to p53 and p21WAF1 untranslated regions, those authors demonstrated that miR-296 has a weak but significant effect on the luciferase-p53–3’ UTR, and it interacts with the p21WAF1 HU region in 3’ UTR and decreases p21WAF1 mRNA level (Yoon et al. 2011). MiR-296 also regulates the expression of p27. In miR-296 antagonim-treated cells, both the protein and mRNA levels of p27 were decreased, suggesting both transcriptional and translational regulation of p27 by miR-296 (Hong et al. 2010). miR-296 may promote proliferation of VSMCs and macrophages through inhibition of p53, p21WAF1 and p27.

The phosphatase and tensin homologue (PTEN) gene is protective in the progression of atherosclerosis (Li et al. 2009; Yuan et al. 2012). Overexpression of PTEN was found to inhibit PDGF-induced phosphorylation of protein kinase B (Akt), p70-S6 kinase, glycogen synthase kinase 3
(GSK3), and thus results in decreased cell proliferation and migration in VSMCs (Chen et al. 2004; Huang and Kontos 2002; Yuan et al. 2012). More recently, Kakizaki and his colleagues found that PTEN is a direct target of miR-296-3p (Kakizaki et al. 2017). The 3′ UTR of PTEN harbors a putative miRNA binding site, which has been shown to specifically bind to miR-296-3p. Following transfection with the miR-296-3p, the protein levels of PTEN was markedly decreased. Further analysis showed that miR-296-3p can regulate the phosphoinositide 3-kinase (PI3K)/Akt signaling pathway by targeting PTEN (Kakizaki et al. 2017). These studies suggest that miR-296-3p has an important role in regulating proliferation and migration of VSMCs by directly targeting the PTEN 3′ UTR for repression.

**MiR-296 regulates apoptosis**

MiR-296 may also suppress apoptosis through nerve growth factor receptor (NGFR) and caspase-8. NGFR, also known as p75NTR, is a neurotrophin receptor that is involved in the pathogenesis of arterial diseases, which is highly expressed in the atherosclerotic adventitia (Chaldakov et al. 2001). NGFR is expressed exclusively in neointimal VSMCs, and the binding of the neurotrophins to NGFR induces cellular apoptosis (Wang et al. 2000). Lee et al. found that the 3′ UTR of NGFR harbors a putative miRNA binding site, which specifically binds to miR-296-5p as validated using a luciferase reporter assay. Moreover, the protein and mRNA expression of NGFR is downregulated by miR-296-5p, suggesting that NGFR may be a novel target of miR-296-5p. (Lee et al. 2016). Furthermore, caspase-8, which plays an obligatory role in apoptosis initiation, is also a target of miR-296-5p. MiR-296-5p can directly bind to the 3′ UTR of caspase-8 and downregulate its protein and mRNA expression (Lee et al. 2016). In addition, upregulation of miR-296-3p could reduce the resistance of αTC1–6 cells to apoptosis induced by cytokines, include IFN-γ, IL-1β and TNF-α. MiR-298-5p, which is located in the same chromosome locus, as miR-296-3p, also has similar effects on apoptosis (Barbagallo et al. 2013).

**Conclusions**

MiR-296 can play a critical role in the initiation and progression of atherosclerosis through regulating its target gene expression. These target genes of miR-296 are involved in the modulation of angiogenesis, inflammatory response, cholesterol metabolism, hypertension, cellular proliferation and apoptosis (Fig. 2).

The immunomodulatory role of miR-296 appears complicated. The role of miR-296 as potent regulators
of angiogenic phenotype has been widely accepted. Overexpression of miR-296 can increase the expression levels of VEGF, and simultaneously inhibit the Notch pathway (Feng et al. 2015). It does not conform to the fact that activation of VEGF signaling increases the expression of Notch ligand DLL4, suggesting that miR-296 may regulate multiple target genes to promote angiogenesis. However, miR-296-3p has the potential to inhibit atherosclerosis by regulating adhesion molecules, including ICAM-1 and CX3CR1 (Liu et al. 2013; Luo and Lin 2016). Furthermore, miR-296 also exhibits an anti- hypertensive effect by suppressing hWNK4 expression, (Mao et al. 2010) because there is a low expression of miR-296-5p in hypertensive patients compared with healthy control subjects (Li et al. 2011). In addition, miR-296-3p can reduce the resistance of αTC1–6 cells to apoptosis induced by cytokines, such as IFN-γ, IL-1β and TNF-α, suggesting its involvement in the regulation of Type 2 diabetes (Barbagallo et al. 2013; Dunning and Gerich 2007). Taken together, increasing evidence supports the hypothesis that miR-296 may be served as a potential indicator of atherosclerosis.

**Future perspectives and challenges**

In the future, ongoing studies of miR-296 will be necessary to clarify its role in atherosclerosis and animal models of various diseases. There are some significant and insightful questions that need to be answered: 1) what regulates miR-296 levels in the biological process of atherosclerosis? 2) How does a single miR-296 strictly regulate the expression of multi-target genes in vivo and in vitro? 3) Are there more miR-296 target genes that can affect the development and progression of atherosclerosis? 4) Does miR-296 influence the progress of atherosclerosis by targeting different signaling pathways in different cell types and tissues? Finally, it remains to be investigated whether miR-296 can be served as a diagnostic and prognostic marker for atherosclerosis based on the identified roles. Given that the functions of miR-296, in particular, its roles in atherosclerosis, have not been researched extensively, understanding the molecular mechanisms and cellular pathways controlled by miR-296 will certainly promote promising miR-296 research in atherosclerosis.

**Acknowledgements**

Not applicable

**Funding**

This work was supported in part by the National Natural Sciences Foundation of China (81660082) and the Guangxi Natural Sciences Foundation (AA139209).

**Availability of data and materials**

Not applicable

**Authors’ contributions**

Conception and design: G-JZ and P-PH. Manuscript writing: HL and X-PO. Collection and assembly of data: TJ and X-LZ. Final approval of manuscript: All authors.

**Ethics approval and consent to participate**

Not applicable

**Consent for publication**

Not applicable

**Competing interests**

The authors declare that they have no competing interests.

**Publisher’s Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**References**

Andreu I, Sun X, Stone PH, et al. miRNAs in atherosclerotic plaque initiation, progression, and rupture[J]. Trends Mol Med. 2015;21(5):307–18.

Barbagallo D, Piro S, Condorelli AG, et al. miR-296-3p, miR-298-5p and their downstream networks are causally involved in the higher resistance of mammalian pancreatic alpha cells to cytokine-induced apoptosis as compared to beta cells[J]. BMC Genomics. 2013:1462.

Barquera S, Pedroza-Tobias A, Medina C, et al. Global overview of the epidemiology of atherosclerotic cardiovascular disease[J]. Arch Med Res. 2015;46(5):328–38.

Batchelor TT, Sorensen AG, di Tomaso E, et al. AZD2171, a pan-VEGF receptor tyrosine kinase inhibitor, normalizes tumor vasculature and alleviates edema in glioblastoma patients[J]. Cancer Cell. 2007;11(1):93–95.

Batetta B, Mulà MF, Petruzzo P, et al. Opposite pattern of MDR1 and caveolin-1 gene expression in human atherosclerotic lesions and proliferating human smooth muscle cells[J]. Cell Mol Life Sci. 2001;58(8):1113–20.

Buckley ML, Ramji DP. The influence of dysfunctional signaling and lipid homeostasis in mediating the inflammatory responses during atherosclerosis[J]. Biochim Biophys Acta. 2015;1852(7):1498–510.

Camarè C, Pucelle M, Nègre-Salvayre A, et al. Angiogenesis in the atherogenic plaque[J]. Redox Biol. 2017;12:18–34.

Chaldakov GN, Stankulov IS, Fiore M, et al. Nerve growth factor levels and mast cell distribution in human coronary atherosclerosis[J]. Atherosclerosis. 2001;159(1):57–66.

Chen WJ, Lin KH, Lai YJ, et al. Protective effect of propylthiouracil independent of its hypothyroid effect on atherogenesis in cholesterol-fed rabbits: PTEN induction and inhibition of vascular smooth muscle cell proliferation and migration[J]. Circulation. 2004;110(10):1313–9.

De Paoli F, Staels B, Chiniatti-Bagaud G. Macrophage phenotypes and their modulation in atherosclerosis[J]. Circ J. 2014;78(8):1775–81.

Diez-Juan A, Andres V. The growth suppressor p27(Kip1) protects against diet-induced atherosclerosis[J]. FASEB J. 2001;15(11):1589–95.

Dunning BE, Gerich JE. The role of alpha-cell dysregulation in fasting and postprandial hyperglycemia in type 2 diabetes and therapeutic implication[J]. Endocr Rev. 2007;28(3):253–83.

Ebert MS, Shapir PA. Roles for microRNAs in conferring robustness to biological processes[J]. Cell. 2012;149(3):515–24.
