28.1 Pulmonary Surfactant and Exogenous Surfactant Therapy in Neonates

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Educational Aims
- Understand the composition, metabolism, and physiology of pulmonary surfactant
- Understand the effect of surfactant in infants with surfactant deficiency or dysfunction
- Understand the optimal timing and methods of surfactant administration

28.1.1 Introduction

Exogenous pulmonary surfactant, widely used in neonatal care, is one of the best-studied treatments in neonatology, and its introduction in the 1990s led to a significant improvement in neonatal outcomes in preterm infants, including a decrease in mortality. This chapter provides an overview of surfactant composition and function in health and disease and summarizes the evidence for its clinical use.

28.1.2 Surfactant Composition, Metabolism, Physiology, and Pathophysiology

28.1.2.1 Surfactant Composition and Metabolism

The alveoli of all mature mammals are lined with pulmonary surfactant, a lipoprotein that reduces surface tension and prevents alveolar collapse. The constituents of surfactant are phospholipids (80%), neutral lipids (8%), and proteins (20%). Among the phospholipids, the predominant one (60%) is dipalmitoylphosphatidylcholine (DPPC), with lesser amounts of unsaturated phosphatidylcholine compounds, phosphatidylglycerol, and phosphatidylinositol. The proteins consist of four unique surfactant-associated apoproteins. Two of these, SP-A and SP-D, are hydrophilic proteins and belong to a subgroup of mammalian lectins called collectins. The other two, SP-B and SP-C, are hydrophobic proteins.

Surfactant is produced in the type II cells of the alveoli (Fig. 28.1), which differentiate between 24 and 34 weeks of gestation in the human. It is assembled and stored in the lamellar bodies, which are concentric or parallel lamellae of phospholipid bilayers. Lamellar bodies are extruded into the fluid layer lining the alveoli by exocytosis and form long stacked tubes composed mainly of...
phospholipid bilayers called tubular myelin. On cross section, tubular myelin has a lattice-like structure because the corners of the component tubes appear fused. Tubular myelin enters the air–water interface quickly by adsorption and greatly reduces the surface tension of that interface (Hallman 2004). It is the major source of the monolayer surface film lining the air–liquid interface in the alveoli. In this monolayer the hydrophobic fatty acyl groups of the phospholipids extend into the air, while the hydrophilic polar head groups bind water (Possmayer et al. 1984).

The total pulmonary surfactant content can be divided into an intra-alveolar and an intracellular pool (Nkadi et al. 2009). However, the total surfactant pool size is not equivalent to the amount
of active surfactant. Maintaining an adequate intra-alveolar surfactant pool is essential for lung function and is dependent on the dynamic cycle of surfactant metabolism (Nkadi et al. 2009). Surfactant pool size increases in late pregnancy, followed by a gradual decrease after birth to adult values (Zimmermann et al. 2005). During and shortly after birth, large amounts of surfactant are released into the alveolar space (Zimmermann et al. 2005). Among different species, humans have the smallest alveolar pool sizes, whereas the amount of saturated phosphatidyl choline in lung tissue is similar across species. The lower alveolar surfactant pool size makes the human lung particularly vulnerable to surfactant dysfunction in case of lung injury. The surfactant pool size in preterm infants with respiratory distress syndrome (RDS) is around 10 mg/kg or less, while healthy term infants have an estimated surfactant pool size of 100 mg/kg (Nkadi et al. 2009). The rate of synthesis of surfactant in preterm infants is also low (Zimmermann et al. 2005).

Alveolar surfactant can be cleared by different pathways. All the main components of surfactant (DPPC, PG, SP-A, SP-B, and SP-C) are recycled. The phospholipid from the monolayer eventually reenters the type II cells through endocytosis and forms multivesicular bodies, which are then either incorporated into lamellar bodies (“recycled”) or degraded in lysosomes. Degraded surfactant components are also used to synthesize new surfactant lipids or proteins. Finally, surfactant can be removed from the lung, either as intact molecules or as degraded products (Zimmermann et al. 2005; Jobe and Ikegami 1993).

Control of surfactant production: Surfactant secretion can be stimulated by a number of mechanisms. Type II cells have beta-adrenergic receptors and respond to beta-agonists with increased surfactant secretion (Nkadi et al. 2009). Purines, such as adenosine triphosphate are potent stimulators of surfactant secretion and may be important for its secretion at birth. Mechanical stretch, such as lung distension and hyperventilation, has also been found to be involved in stimulating surfactant secretion (Nkadi et al. 2009). Hormones also play a role in surfactant secretion. Thyroxine accelerates type II cell differentiation while acting synergistically with glucocorticoids to enhance lung compliance and DPPC synthesis. Finally, both prenatally administered maternal glucocorticoids and exogenous surfactant administration after birth stimulate endogenous surfactant synthesis.

28.1.2.2 Surfactant Physiology
28.1.2.2.1 In the Normal Lung
Pulmonary alveoli are bubble shaped, with a high degree of curvature. The attraction between the molecules in the alveolar fluid of the moist inner surface of the alveoli generates surface tension and tends to make the alveoli collapse. Unchecked, this tendency would result in lung collapse. Surfactant greatly reduces the surface tension at the air–liquid interface in the alveoli and distal bronchioles. This prevents the alveoli from collapsing during expiration and promotes lung expansion during inspiration.

The main component responsible for decreasing the surface tension is DPPC. However it adsorbs very slowly to air–liquid interfaces and therefore requires surfactant proteins or other lipids to facilitate its adsorption. SP-B and SP-C enhance spreading of phospholipid in the airspaces. SP-B promotes phospholipid adsorption and induces the insertion of phospholipids into the monolayer, thus enhancing the formation of a stable surface film (Creuwels et al. 1997). SP-C enhances phospholipid adsorption, stimulates the insertion of phospholipids out of the subphase into the air–liquid interface, and may increase the resistance of surfactant to inhibition by serum proteins or by edema fluid (Creuwels et al. 1997; Griese 1999).

As the alveolar surface expands during inspiration, surfactant components insert from the hypophase (epithelial lining fluid) into the monolayer. At expiration the alveolar surface reduces and the monolayer is compressed, thereby squeezing out some surfactant proteins, unsaturated PC, and other lipids. By this mechanism, the monolayer comprises mainly of
DPPC, the most important surface-tension-lowering component during compression (Zimmermann et al. 2005).

Surfactant also has a role in pulmonary host defense. SP-A and SP-D may play important roles in the defense against inhaled pathogens, and SP-A may have a regulatory function in the formation of the monolayer that lowers the surface tension (Creuwels et al. 1997).

28.1.2.2.2 In the Premature Lung

The preterm infant with respiratory distress syndrome (RDS) has immaturity of the lungs, especially of the type II cells, and decreased synthesis of surfactant. This results in low amounts of surfactant in the alveoli (surfactant pool size of 2–10 mg/kg) (Zimmermann et al. 2005) that contains a lower percent of disaturated phosphatidylcholine species, less phosphatidylglycerol, and less of all the surfactant proteins than surfactant from a mature lung. Minimal surface tensions are also higher for surfactant from preterm than term infants (Nkadi et al. 2009). However, preterm infants may have increased recycling of surfactant when compared to term infants (Zimmermann et al. 2005).

Shortly after birth, infants with RDS develop tachypnea, grunting, nasal flaring, use of accessory muscles of respiration, intercostal or subcostal retractions, cyanosis, poor feeding, and apnea. A chest radiograph typically shows a diffuse reticulogranular “ground glass” opacification (the result of diffuse alveolar atelectasis) with superimposed air bronchograms (Nkadi et al. 2009). The lungs of infants who die from RDS show alveolar atelectasis, alveolar and interstitial edema, and diffuse hyaline membranes in distorted small airways (Nkadi et al. 2009). RDS is one of the most common causes of death and morbidity in preterm neonates. It occurs worldwide with a slight male predominance (Nkadi et al. 2009). Prenatal corticosteroids significantly reduce the incidence, severity, and mortality associated with RDS.

28.1.2.2.3 In the Injured Neonatal Lung

Surfactant dysfunction can develop secondary to a variety of other conditions that result in lung injury in neonates, such as meconium aspiration syndrome, pulmonary hemorrhage, and pneumonia.

In meconium aspiration syndrome, the mechanisms underlying surfactant inactivation are not fully understood, but it has been shown that meconium destroys the fibrillar structure of surfactant and decreases its surface adsorption rate (Nkadi et al. 2009). In vitro studies (Moses et al. 1991; Clark et al. 1987) and animal studies (Sun et al. 1993; Davey et al. 1993) have demonstrated that meconium inhibits surfactant function and is likely to be partially responsible for alveolar collapse in meconium aspiration syndrome. Components of meconium that may contribute to altered surfactant function include cholesterol, free fatty acids, bile salts, bilirubin, and proteolytic enzymes (Moses et al. 1991; Clark et al. 1987; Sun et al. 1993; Lieberman 1966). In particular, phospholipase-A2 (PLA2) in meconium has been found to inhibit the activity of surfactant in vitro in a dose-dependent manner, through the competitive displacement of surfactant from the alveolar film (Nkadi et al. 2009). PLA2 is also known to induce hydrolysis of DPPC, releasing free fatty acids and lyso-PC which damage the alveolar–capillary membrane and induce intrapulmonary sequestration of neutrophils (Nkadi et al. 2009).

In pulmonary hemorrhage, capillary filtrate builds up in the interstitial space and can burst through the pulmonary epithelium into the airspaces. Neutrophils are released following endothelial damage and they, in turn, express proteases, oxygen free-radicals, and cytokines. These free oxygen molecules damage the type II cells that produce surfactant proteins, thus inhibiting production of the proteins (Nkadi et al. 2009). Elastase, one of these proteases, damages and degrades SP-A, thereby inhibiting SP-A-mediated surfactant lipid aggregation and adsorption in vitro (Nkadi et al. 2009).

Acute respiratory distress syndrome (ARDS) is a significant cause of morbidity and mortality in all age groups following infection (the most common cause), hemorrhage, or other forms of lung injury. It is defined as a severe form of acute lung injury (ALI) and a syndrome of acute
pulmonary inflammation. ALI/ARDS is characterized by sudden onset, impaired gas exchange, decreased static compliance, and a non-hydrostatic pulmonary edema (Nkadi et al. 2009). ARDS is characterized by an increase in the permeability of the alveolar–capillary barrier due to injury to the endothelium and/or alveolar lining cells. Damage to the alveolar type I cells leads to an influx of protein-rich edema fluid into the alveoli, as well as decreased fluid clearance from the alveolar space. Neutrophils are attracted into the airways by host bacterial and chemotactic factors and express enzymes and cytokines which further damage the alveolar epithelial cells (Nkadi et al. 2009). Type II epithelial cell injury leads to a decrease in surfactant production, with resultant alveolar collapse.

28.1.2.2.4 Effect of Mechanical Ventilation on Pulmonary Surfactant
Mechanical ventilation in itself can worsen lung disease and effect surfactant function. The damage caused by mechanical ventilation can cause fluid, protein, and blood to leak into the airways, alveoli, and the lung interstitium, interfering with lung mechanics, inhibiting surfactant function, and promoting lung inflammation (Clark et al. 2001). Even a short period of mechanical ventilation can cause a decrease in lung compliance that is associated with a large influx of proteins into the alveolar space and with alterations in the pulmonary surfactant system (Veldhuizen et al. 2000). The changes of surfactant in these experiments are different from those seen in acute lung injury, indicating that they may represent an initial response to mechanical ventilation.

In the adult lung, injuries due to mechanical ventilation occur primarily when the sum of the functional residual capacity (FRC) and tidal volume approach or exceed maximal lung volume (Dreyfuss and Saumon 1993). The preterm lung is particularly susceptible to injury by mechanical ventilation because of structural immaturity, and tidal volumes considered safe for the adult may approach the maximum lung volumes in the preterm lung (Wada et al. 1997). Initiation of ventilation in preterm lambs with high volumes causes lung injury and decreases the subsequent response to surfactant treatment (Wada et al. 1997).

In premature baboons, mechanical ventilation results in abnormal surfactant metabolism (Seidner et al. 1998). In the normal lung, large lipid arrays or large-aggregate forms of surfactant are the source of the surface film (Wright 1990). Small vesicles, primarily containing lipids, reenter the hypophase for recycling or catabolism. The amount of inactive vesicular forms increases with lung injury, and this increase is associated with a deterioration in lung mechanics.

Long-term ventilation of the lungs in immature preterm infants with respiratory distress leads to ventilator-induced lung injury and chronic lung disease. Ventilation of these infants not only interferes with alveolarization but also with the surfactant system. Ventilated preterm animals accumulated very large lipid pools in tissue, but alveolar pools stay relatively low indicating decreased secretion of newly synthesized saturated phosphatidylcholine and increased catabolism. These effects of both acute and long-term ventilation leave the preterm infant particularly susceptible to problems associated with surfactant deficiency and dysfunction.

28.1.3 Exogenous Surfactant Therapy
28.1.3.1 Types of Exogenous Surfactant
There are two broad categories of exogenous surfactant available for treatment: animal-derived surfactants and synthetic surfactants.

28.1.3.1.1 Animal-Derived Surfactants
These include the following:

Bovine surfactant obtained by lung mince: Beractant (Survanta) and Surfactant-TA (Surfacten) are lipid extracts of bovine lung mince with added DPPC, tripalmitoylglycerol, and palmitic acid.

Bovine surfactant obtained by lung lavage: Calf lung surfactant extract (CLSE, calfactant, Infasurf), SF-R11 (Alveofact), and bovine
lipid extract surfactant (BLES) are bovine lung washes subjected to chloroform–methanol extraction.

**Porcine surfactant obtained by lung mince:**
Poractant (Curosurf) is a porcine lung mince that has been subjected to chloroform–methanol extraction and further purified by liquid–gel chromatography. It consists of approximately 99% polar lipids (mainly phospholipids) and 1% hydrophobic, low molecular weight proteins (SP-B and SP-C) (Wiseman and Bryson 1994).

### 28.1.3.1.2 Synthetic Surfactants

These include the following:

**Protein-free synthetic surfactants:** There is one product in this category, colfosceril palmitate, cetyl alcohol, tyloxapol (Exosurf), and it consists of 85% dipalmitoylphosphatidylcholine (DPPC), 9% hexadecanol, and 6% tyloxapol (a spreading agent). Another product in this category, pumactant (also known as artificial lung expanding compound, ALEC) is no longer manufactured (Halliday 2006) and was a 7:3 mixture of DPPC and phosphatidyl glycerol. These synthetic surfactants lack many of the components of animal-derived surfactant, particularly the hydrophobic surfactant proteins B and C.

**Protein-containing synthetic surfactants:** These surfactants contain synthetic phospholipids along with proteins (produced through peptide synthesis and recombinant technology) that attempt to mimic the function of either SP-B or SP-C. Of these, lucinactant (Surfaxin) contains dipalmitoylphosphatidylcholine, palmitoyl oleoylphosphatidyl glycerol, and palmitic acid (Cochrane et al. 1996, 1998) combined with a mimic of SP-B called sinapultide or KL4 peptide. KL4 is a 21-residue peptide comprised of repeated units of four hydrophobic leucine (L) residues, bounded by basic polar lysine (K) residues arranged in the following order: KLLLLKLLLLLKLKLK. This structure resembles the repeating pattern of hydrophobic and hydrophilic residues in the C-terminal part of SP-B and stabilizes the phospholipid layer by interactions with the lipid heads and the acyl chains (Cochrane and Revak 1991). Another synthetic SP-B analog currently under testing is called dSP-B_{1-25}, which resembles the N-terminal segment of SP-B and when combined with synthetic phospholipids has shown some efficacy in animal studies.

Another type of synthetic protein-containing surfactant is called rSP-C surfactant or lusupultide (Venticute). It contains DPPC, palmitoyl oleoylphosphatidyl glycerol, palmitic acid, and calcium chloride (Hafner and Germann 2000; Spragg et al. 2000) combined with a recombinant SP-C analog (rSP-C), which is similar to the 34-amino acid human SP-C sequence, except that it contains cysteine (in place of phenylalanine) in positions 4 and 5 and contains isoleucine (instead of methionine) in position 32.

During surfactant replacement therapy, exogenous surfactants are given at doses between 10 and 20 times the usual pool sizes found in preterm infants with RDS, which approximates the pool size in term infants (Nkadi et al. 2009). The table summarizes the individual characteristics of each of these products, including the dosage, volume, and the recommended repeat dosing interval. All these products are administered intratracheally with each dose divided into multiple aliquots and should be administered according to the manufacturers’ recommendations (Table 28.1).

### 28.1.3.2 Administration of Exogenous Surfactant: Practical Issues

The dynamics of how exogenously administered surfactant might spread through the airways into the alveoli have been well studied in the laboratory and theoretical models created (Halpern et al. 1998, 2008). When a bolus of surfactant is instilled and propagates down the airways, it deposits a liquid layer that coats the airways. The bolus may rupture because it may not pick up as much fluid as it is depositing and therefore may not reach the terminal bronchioles and alveoli where it is needed. However, the deposited liquid layer that is left behind may still advance due to the effects of gravity and surface tension, especially as the liquid layer thins.

The potential magnitude of uneven distribution of exogenously administered surfactant has
been emphasized by Jobe (2006), who points out that there are 20 generations of airway branching from the trachea to the respiratory bronchioles and saccules, with 250,000 binary branch points and 500,000 distal airways leading to saccules. If the distribution is not proportionate to the number of saccules beyond each branch point, surfactant distribution will not be uniform. Any nonuniformity at a proximal branch point will be amplified at subsequent branch points. When exogenous surfactant is administered in clinical practice, its distribution is not ideal, but is often good enough because of the biophysical properties of surfactant and the small amount that is needed regionally in the lung for a treatment response (Jobe 2006).

Table 28.1 Products available for exogenous surfactant therapy

| Name of product (pharmaceutical name) | Source | Phospholipid (mg/ml) | Dose (volume) (ml/kg) | Dose (mg/kg of phospholipid) | Repeat dosing interval (h) |
|---------------------------------------|--------|----------------------|-----------------------|-----------------------------|---------------------------|
| **Animal-derived products**           |        |                      |                       |                             |                           |
| Beractant (Survanta)                  | Lipid extract of bovine lung mince with added DPPC, tripalmitoylglycerol and palmitic acid | 25 | 4 | 100 | 6 |
| Calfactant (Infasurf)                 | Bovine lung wash subjected to chloroform–methanol extraction | 35 | 3 | 100 | 6–12 |
| SF-R1I (bovactant, Alveofact)         | Bovine lung wash subjected to chloroform–methanol extraction | 50 | 1–2 | 50–100 | 8 |
| bLES (BLES)                           | Bovine lung wash subjected to chloroform–methanol extraction | 27 | 5 | 135 | 6 |
| Poractant (Curosurf)                  | Porcine lung mince that has been subjected to chloroform–methanol extraction and further purified by liquid–gel chromatography | 80 | 1.25–2.5 for initial dose, 1.25 for subsequent doses | 100–200 | 12 |
| **Protein-free synthetic surfactants**|        |                      |                       |                             |                           |
| Colfosceril palmitate, hexadecanol, tyloxapol | 85 % dipalmitoylphosphatidylcholine (85 %), hexadecanol (9 %), and tyloxapol (a spreading agent, 6 %) | 13.5 | 5 | 67.5 | 12 |
| **Protein-containing synthetic surfactants**|        |                      |                       |                             |                           |
| Lucinactant (Surfaxin) (Moya et al. 2005) | Synthetic phospholipids and proteins produced through peptide synthesis and recombinant technology. Contains a mimic of SP-B called sinapultide or KL4 peptide | 30 | 5.8 | 175 | 6 |
| Recombinant SP-C surfactant (Venticute) | Synthetic phospholipids and proteins produced through peptide synthesis and recombinant technology. Contains recombinant SP-C (rSP-C) combined with DPPC, palmitoyl oleoylphosphatidyl glycerol, palmitic acid, and calcium chloride | 50 | 1–2 | 50–100 | 4 |

*No neonatal studies were available at time of publication; the displayed details of dosing were obtained from studies on animals and in adult patients (Spragg et al. 2004; Hilgendorff et al. 2006)
There are several variables that contribute to the distribution of surfactant in the lungs (Jobe 2006): surface activity causes rapid adsorption and spreading, gravity contributes to the distribution of surfactant in large airways, a higher volume of surfactant and faster administration cause better distribution, positive pressure ventilation and positive end-expiratory pressure help clear the airways of fluid, and higher volumes of fetal lung fluid or edema fluid improve distribution. Therefore techniques to improve surfactant distribution include positioning the infant to minimize gravity, giving surfactant quickly in a reasonable volume, and giving the infant enough ventilator support to quickly clear the airways of fluid.

According to the manufacturers’ recommendations, beractant and poractant should be administered through a catheter inserted into the endotracheal tube, colfosceril should be administered through a side-port adapter attached to the endotracheal tube, and calf lung surfactant extract can be administered either through a feeding catheter or through a side-port adapter. Other methods of administration of surfactant have been tested in randomized trials as well and are described below.

**Administration Through Catheter, Side Port, or Suction Valve:** In a randomized trial, the administration of beractant through a catheter inserted into the endotracheal tube, colfosceril should be administered through a side-port adapter attached to the endotracheal tube, and calf lung surfactant extract can be administered either through a feeding catheter or through a side-port adapter. Other methods of administration of surfactant have been tested in randomized trials as well and are described below.

**Administration Through Dual-Lumen Endotracheal Tube:** The administration of poractant through a dual-lumen endotracheal tube without a change in position or interruption of mechanical ventilation was compared to bolus instillation in a randomized trial (Soler et al. 1998). The dual-lumen group had fewer episodes of dosing-related hypoxia, a smaller decrease in heart rate and SaO_2_, and a shorter total time in supplemental oxygen than the bolus group. The dual-lumen method has also been compared to the side-port method of administration of colfosceril in a randomized trial (Nelson et al. 1997). No difference was found between the two methods in dosing-related hypoxemia.

**Slow Infusion Versus Bolus Administration:** In one randomized clinical trial (Sitler et al. 1993), the slow infusion of colfosceril using a microinfusion syringe pump over 10–20 min was compared to manual instillation over 2 min. Pump administration resulted in fewer infants with loss of chest wall movement during dosing as well as a lesser increase in peak inspiratory pressure than with hand administration. In another small clinical trial of preterm infants (Zola et al. 1993b), there were no differences in clinical outcomes with administration by bolus versus slow infusion. However in animals, slow infusion of surfactant into the endotracheal tube results in nonhomogeneous distribution of surfactant in the lung (Ueda et al. 1994; Segerer et al. 1993). Because the evidence about the best method of administration is scant, and because bolus administration is likely to lead to better distribution, bolus administration of surfactant is preferred.

**Other Methods:** Other methods of administration such as nebulization or aerosolization (Berggren et al. 2000; Dijk et al. 1998; Ellyett et al. 1996; Fok et al. 1998; Jorch et al. 1997) and in utero administration to the human fetus (Cosmi et al. 1996; Petrikovsky et al. 1995) have also been reported. These methods require further clinical testing and are not currently recommended.

**Chest Position During Administration of Surfactant:** In a study in rabbits, pulmonary
distribution of intratracheally instilled surfactant was largely determined by gravity, and changing the chest position after instillation did not result in any redistribution of the surfactant. Therefore, for neonates receiving surfactant, keeping the chest in the horizontal position may result in the most even distribution of the surfactant in the two lungs (Broadbent et al. 1995).

In summary, based on available evidence, surfactant should be administered in the standard method of aliquots instilled into an endotracheal tube. There is evidence to suggest that the administration of surfactant using a dual-lumen endotracheal tube or through a catheter passed through a suction valve is effective and may cause less dosing-related adverse events than standard methods. The side-port method of administration and the catheter method of administration appear to be equivalent. More studies are required before firm conclusions can be drawn about the optimal method of administration of surfactant and whether the optimal method is different for different types of surfactant.

28.1.3.3 Clinical Use of Surfactant Therapy in Preterm Infants with RDS

Exogenous surfactant therapy is one of the best-studied therapies in neonatology and numerous randomized controlled trials have been performed comparing various treatment regimens and strategies. The findings from these trials, many of which are summarized in multiple systematic reviews in the Cochrane Database of Systematic Reviews (Sinclair et al. 2003), are described in the following sections. The results of the meta-analysis in these reviews are expressed as typical relative risk (RR) and typical absolute risk difference (ARD), with 95% confidence intervals (CI) for each of these.

It is useful to clarify the terminology used for various treatment strategies with surfactant.

Rescue (or selective) surfactant therapy: Administration of exogenous surfactant to an infant who has already developed clinical features of RDS.

Many clinical trials in the late 1980s and early 1990s studied the effects of rescue and prophylactic surfactant therapy compared to placebo or no therapy. Systematic reviews of these trials show that, compared to placebo or no therapy, surfactant treatment or prophylaxis (with either animal-derived or synthetic surfactant) decreases the risk of pneumothorax and of mortality. Estimates from the meta-analyses indicate that there is a 30–65% relative reduction in the risk of pneumothorax and up to a 40% relative reduction in the risk of mortality. There were no consistent effects on other clinical outcomes such as chronic lung disease, patent ductus arteriosus, and intraventricular hemorrhage.

Further evidence of the benefits of surfactant therapy is derived from studies demonstrating decreased mortality and morbidity in very low birth weight infants following the introduction of surfactant therapy into practice (Schwartz et al. 1994; Lee et al. 1999; Philip 1995; Doyle et al. 1999; Hamvas et al. 1996; Horbar et al. 1993a; Hoekstra et al. 1994).

28.1.3.3.1 Efficacy of Surfactant Therapy in Established RDS (Rescue Surfactant Therapy)

Many of the early surfactant trials studied the effects of surfactant treatment in preterm infants with clinical and/or radiologic features of RDS (rescue or treatment trials). Some of these studies used animal-derived surfactant and others used protein-free synthetic surfactant.

Rescue Therapy with Animal-Derived Surfactant: In a systematic review and meta-analysis of 13 randomized trials of animal-derived surfactant (Seger and Soll 2009), infants treated with surfactant had a rapid improvement in respiratory status (improved oxygenation and decreased need for ventilator support), as well as a significant decrease in the risk of (a) any air leak (typical RR 0.47, 95% CI 0.39–0.58; typical ARD −0.16, 95% CI −0.21 to −0.12), (b) pneumothorax (typical RR 0.42, 95% CI
0.34–0.52; typical ARD −0.17, 95 % CI −0.21 to −0.13), and (c) pulmonary interstitial emphysema (typical RR 0.45, 95 % CI 0.37–0.55; typical ARD −0.20, 95 % CI −0.25 to −0.15). There was also a significant decrease in the risk of (a) neonatal mortality (typical RR 0.68, 95 % CI 0.57–0.82; typical ARD −0.09, 95 % CI −0.13 to −0.05), (b) mortality prior to hospital discharge (typical RR 0.63, 95 % CI 0.44–0.90; typical ARD −0.10, 95 % CI −0.18 to −0.03), and (c) bronchopulmonary dysplasia (BPD) or death at 28 days of age (typical RR 0.83, 95 % CI 0.77–0.90; typical ARD −0.11, 95 % CI −0.16 to −0.06).

**Rescue Therapy with Protein-Free Synthetic Surfactant:** In a similar systematic review and meta-analysis of six randomized trials of protein-free synthetic surfactant treatment of established RDS (Soll 1998), surfactant therapy improved pulmonary gas exchange and decreased the requirement for ventilatory support. It also decreased the risk of (a) pneumothorax (typical RR 0.64, 95 % CI 0.55–0.76; typical ARD −0.09, 95 % CI −0.12 to −0.06), (b) pulmonary interstitial emphysema (typically RR 0.62, 95 % CI 0.54–0.71; typical ARD −0.12, 95 % CI −0.16 to −0.09), (c) patent ductus arteriosus (typical RR 0.90, 95 % CI 0.84–0.97; typical ARD −0.06, 95 % CI −0.10 to −0.02), (d) intraventricular hemorrhage (typical RR 0.88, 95 % CI 0.77–0.99; typical ARD −0.04, 95 % CI −0.08 to −0.00), (e) bronchopulmonary dysplasia (typical RR 0.75, 95 % CI 0.61–0.92; typical ARD −0.04, 95 % CI −0.06 to −0.01), (f) neonatal mortality (typical RR 0.73, 95 % CI 0.61–0.88; typical ARD −0.05, 95 % CI −0.07 to −0.02), (g) bronchopulmonary dysplasia or death at 28 days (typical RR 0.73, 95 % CI 0.65–0.83; typical ARD −0.06, 95 % CI −0.11 to −0.05), (h) mortality prior to hospital discharge (typical RR 0.79, 95 % CI 0.68–0.92; typical ARD −0.05, 95 % CI −0.07 to −0.02), and (i) mortality during the first year of life (typical RR 0.80, 95 % CI 0.69–0.94; typical ARD −0.04, 95 % CI −0.07 to −0.01). Treatment with synthetic surfactant increased the risk of apnea of prematurity (typical RR 1.20, 95 % CI 1.09–1.31; typical ARD 0.08, 95 % CI 0.04–0.12).

### 28.1.3.3.2 Efficacy of Surfactant Therapy in Infants at Risk for RDS (Prophylactic Surfactant)

Several of the early trials of surfactant therapy also studied the effects of prophylactic surfactant in preterm infants at risk for developing RDS (i.e., before they had overt clinical features of RDS). Some of these studies used animal-derived surfactant and others used protein-free synthetic surfactant.

**Prophylaxis with Animal-Derived Surfactant:** A systematic review and meta-analysis of eight randomized trials (Soll and Özek 1997) found that prophylaxis with animal-derived surfactant lead to an initial improvement in respiratory status and a decrease in the risk of respiratory distress syndrome in infants. It also lead to a decrease in the risk of (a) pneumothorax (typical RR 0.35, 95 % CI 0.26–0.49; typical ARD −0.15, 95 % CI −0.20 to −0.11), (b) pulmonary interstitial emphysema (typical RR 0.46, 95 % CI 0.35–0.60; typical ARD −0.19, 95 % CI −0.25 to −0.13), (c) neonatal mortality (typical RR 0.60, 95 % CI 0.44–0.83; typical ARD −0.07, 95 % CI −0.12 to −0.03), and (d) bronchopulmonary dysplasia or death (typical RR 0.84, 95 % CI 0.75–0.93; typical ARD −0.10, 95 % CI −0.16 to −0.04).

**Prophylaxis with Protein-Free Synthetic Surfactant:** In a similar systematic review and meta-analysis of seven randomized trials (Soll and Özek 2010), protein-free synthetic surfactant prophylaxis leads to a variable improvement in the respiratory status and a decrease in respiratory distress syndrome in infants who receive prophylactic protein-free synthetic surfactant. It also leads to a decrease in the risk of (a) pneumothorax (typical RR 0.67, 95 % CI 0.50–0.90), (b) pulmonary interstitial emphysema (typical RR 0.68, 95 % CI 0.50–0.93), and (c) neonatal mortality (typical RR 0.70, 95 % CI 0.58–0.85). However, prophylactic protein-free synthetic surfactant administration was associated with an increase in the risk of patent ductus arteriosus (typical RR 1.11, 95 % CI 1.00–1.22) and an increase in the risk of pulmonary hemorrhage (typical RR 3.28, 95 % CI 1.50–7.16).
Many investigators believed that prophylactic administration of surfactant would be the most effective way to deliver surfactant based on the observation in animal studies that surfactant is distributed more uniformly and homogenously when it is administered into a fluid-filled lung (Jobe et al. 1984; Seidner et al. 1995) and the belief that administering surfactant into a previously unventilated or minimally ventilated lung will diminish acute lung injury. In animal models, even brief (15–30 min) periods of mechanical ventilation prior to surfactant administration have been shown to cause acute lung injury resulting in alveolar–capillary damage, leakage of proteinaceous fluid into the alveolar space, and release of inflammatory mediators (Ikegami et al. 1998; Jobe and Ikegami 1998a, b) and to decrease the subsequent response to surfactant replacement (Bjorklund et al. 1997; Rider et al. 1992). Surfactant-deficient animals who receive assisted ventilation develop necrosis and desquamation of the bronchiolar epithelium as early as 5 min after onset of ventilation (Nilsson et al. 1980).

Shortly after surfactant was approved for clinical use, eight randomized controlled trials compared the effects of prophylactic surfactant administration to surfactant treatment of established RDS (Bevilacqua et al. 1996, 1997; Dunn et al. 1991; Egberts et al. 1993; Kattwinkel et al. 1993; Kendig et al. 1991; Merritt et al. 1991; Walti et al. 1995). All these trials used animal-derived surfactant preparations. Trials varied whether surfactant was given before or after the onset of air breathing (pre- or post-ventilatory administration), but all administered surfactant before 15 min of age. The average time of administration of surfactant in the selective treatment groups ranged from 1.5 to 7.4 h. The results of the meta-analysis of the eight trials from a systematic review (Soll and Morley 2012) are summarized in Fig. 28.2.

Compared to surfactant treatment of established RDS, prophylactic administration of surfactant resulted in a decrease in the risk of pneumothorax (typical RR 0.62, 95 % CI 0.42–0.89; typical ARD –0.02, 95 % CI –0.04 to –0.01), a decrease in the risk of pulmonary interstitial emphysema (typical RR 0.54, 95 % CI 0.36–0.82; typical ARD –0.03, 95 % CI –0.04 to –0.01), a reduction in the risk of neonatal mortality (typical RR 0.61, 95 % CI 0.48–0.77; typical ARD –0.05, 95 % CI –0.07 to –0.02), and a trend towards a decrease in the risk of intraventricular hemorrhage (typical RR 0.92, 95 % CI 0.82–1.03; typical ARD –0.03, 95 % CI –0.06 to 0.01). Because of the greater risk of respiratory distress syndrome and mortality with decreasing gestational age, the benefits of

| Outcome (no. of trials) | Typical risk difference (95 % CI) |
|-------------------------|----------------------------------|
| Pneumothorax (6)        | −0.02 (−0.04, −0.01)             |
| Bronchopulmonary dysplasia (8) | −0.01 (−0.03, 0.02) |
| Mortality (7)           | −0.05 (−0.07, −0.02)             |
| BPD or death (8)        | −0.04 (−0.07, −0.01)             |

Fig. 28.2 Meta-analysis of eight randomized controlled trials comparing surfactant prophylaxis and treatment (Soll 2001)
prophylactic administration compared to selective administration were of greater magnitude. The meta-analysis demonstrates that compared to selective administration, prophylactic administration of animal-derived surfactant to infants less than 30 weeks gestation resulted in a greater reduction in neonatal mortality (typical RR 0.62, 95 % CI 0.49–0.78; typical ARD −0.06, 95 % CI −0.09 to −0.03) and a reduction in the combined outcome of bronchopulmonary dysplasia or death (typical RR 0.87, 95 % CI 0.77–0.97; typical ARD −0.05, 95 % CI −0.09 to −0.01).

Preventilatory Versus Post-ventilatory Prophylactic Surfactant Administration: The initial studies using prophylactic surfactant administered the drug as an immediate bolus after intubating the infants rapidly after birth (i.e., “before the first breath”). This approach delays the initiation of neonatal resuscitation, including positive pressure ventilation, and is associated with a risk for surfactant delivery into the right main stem bronchus or esophagus. A randomized trial demonstrated that prophylaxis may be administered in small aliquots soon after resuscitation and confirmation of endotracheal tube position, with equivalent or greater efficacy (Kendig et al. 1998). Based on this trial, prophylactic surfactant should be administered after initial resuscitation of the infant at birth and administration prior to the “first breath” is unnecessary.

The trials mentioned above comparing the use of prophylactic surfactant to surfactant treatment of established RDS were all performed in an era when the use of maternal antenatal glucocorticoids was not as high as in the current era, where 90 % or more of mothers who deliver prematurely receive antenatal glucocorticoids. Also, in these trials, surfactant in the “rescue” group was given relatively late, between 1.5 and 7.4 h, after birth. There are no trials comparing the effects of prophylactic intubation and surfactant administration shortly after birth to infants at high risk of RDS (with intubation primarily performed to administer surfactant) to very early selective administration (e.g., at 30–60 min of life) in intubated infants with early RDS or respiratory insufficiency. Therefore, in current practice, the benefits of prophylactic surfactant demonstrated in these trials might not be as large. A strategy of surfactant prophylaxis subjects infants without RDS to unnecessary intubation and surfactant administration and also to the risk of ventilator-induced lung injury (Clark et al. 2001) (although admittedly some of these infants without RDS may still require intubation and ventilation for respiratory failure not due to RDS). In recent years, prophylactic surfactant administration to preterm infants at risk of RDS has been compared with the use of nasal CPAP (and attempting to avoid mechanical ventilation) as the primary method of respiratory management at birth. These studies are discussed in the next section.

28.1.3.3.4 Initial Respiratory Management of Preterm Infants: CPAP Compared to Intubation Followed by Surfactant Administration

Four large multicenter trials have evaluated the use of prophylactic or early surfactant administration to immediate stabilization on continuous distending pressure (SUPPORT Study Group of the Eunice Kennedy Shriver NICHD Neonatal Research Network 2010; Vermont Oxford Network DRM Study Group et al. 2010; Sandri et al. 2010; Morley et al. 2008).

In the SUPPORT trial (SUPPORT Study Group of the Eunice Kennedy Shriver NICHD Neonatal Research Network 2010), infants 24–27 weeks gestation were randomly assigned to intubation and surfactant treatment (within 1 h after birth) or to CPAP treatment initiated in the delivery room, with subsequent use of a “protocol-driven limited ventilation strategy.” The primary outcome, death, or bronchopulmonary dysplasia (defined as supplemental oxygen requirement at 36 weeks post-menstrual age) did not differ significantly between the CPAP group and the surfactant group (47.8 and 51.0 %, respectively; RR with CPAP, 0.95; 95 % CI, 0.85–1.05) after adjustment for gestational age, center, and familial clustering. The results were similar when bronchopulmonary dysplasia was defined according to the need for any supplemental oxygen at 36 weeks (rates of primary outcome, 48.7 and 54.1 %, respectively; RR with CPAP, 0.91; 95 %
Infants who received CPAP treatment, as compared with infants who received surfactant treatment, less frequently required intubation or postnatal corticosteroids for bronchopulmonary dysplasia \((P<0.001)\), required fewer days of mechanical ventilation \((P=0.03)\), and were more likely to be alive and free from the need for mechanical ventilation by day 7 \((P=0.01)\). In secondary analyses, among infants 24 and 25 weeks gestation, there was a significant reduction in the risk of death in the CPAP group, as compared with the early-intubation group. The rate of death during hospitalization was 24% versus 32%, RR with CPAP, 0.74; 95% CI 0.57–0.98. The rate of death at 36 weeks was 20% versus 29%, RR, 0.68; 95% CI 0.5–0.92.

In the Delivery Room Management randomized trial conducted by the Vermont Oxford Network (Vermont Oxford Network DRM Study Group et al. 2010), three strategies were compared in infants at risk of RDS (preterm infants 26–29 weeks): prophylactic surfactant followed by a period of assisted ventilation, intubation with immediate surfactant treatment and rapid extubation to nasal CPAP (ISX), and early stabilization on nasal CPAP (NCPAP). The study was terminated prior to reaching the desired sample size. There were over 200 infants in each of the three study arms. No statistically significant differences were found between the three arms of the trial in the primary outcome of death or chronic lung disease (defined as supplemental oxygen requirement at 36 weeks post-menstrual age). The incidence of the primary outcome was 37% in the prophylactic surfactant group, 29% in the ISX group (RR with ISX compared to prophylactic surfactant 0.78, 95% CI 0.59–1.03), and 31% in the CPAP group (RR with CPAP compared to prophylactic surfactant 0.83, 95% CI 0.64–1.09).

In the CURPAP trial (Sandri et al. 2010), infants 25–28 weeks gestation who were not intubated at birth were randomly assigned to prophylactic surfactant or nasal CPAP within 30 min of birth. There were no statistically significant differences between the two groups in the outcomes studied – need for mechanical ventilation in the first 5 days of life, death at 28 days of life, death at 36 weeks post-menstrual age, and the main morbidities of prematurity.

In the COIN trial (Morley et al. 2008), infants 25–28 weeks gestation who were breathing spontaneously but had developed respiratory distress were randomly assigned to CPAP or intubation and ventilation at 5 min after birth. Infants intubated before randomization and those not requiring respiratory support or oxygen were excluded. The primary outcome of death or chronic lung disease (defined as the need for oxygen treatment at 36 weeks gestational age) was not statistically different in infants assigned to receive intubation (39%) compared with infants assigned to receive CPAP (34%, odds ratio favoring CPAP, 0.80; 95% CI 0.58–1.12). At 28 days, there was a lower risk of death or need for oxygen therapy in the CPAP group than in the intubation group (odds ratio, 0.63; 95% CI 0.46–0.88; \(P=0.006\)). There was little difference in overall mortality. In the CPAP group, 46% of infants were intubated during the first 5 days, and the use of surfactant was halved. However, the incidence of pneumothorax was 9% in the CPAP group, as compared with 3% in the intubation group \((P<0.001)\). There were no other serious adverse events. The CPAP group had fewer days of ventilation.

The results of these trials suggest that in preterm infants at high risk of RDS, instead of routinely intubating and administering prophylactic surfactant immediately after birth, it is reasonable to try and initially stabilize such infants on nasal CPAP. This approach can avoid unnecessary intubation, unnecessary surfactant administration, and minimize ventilator-induced lung injury, while reducing healthcare costs and resource utilization. However, clinicians should closely monitor infants initially stabilized on CPAP to identify infants who have progressive respiratory failure, so that infants who require surfactant can be intubated and given surfactant as early as possible. Clinicians should also use information available prior to the birth of the infant to identify infants who are not likely to do well with initial stabilization on nasal CPAP, such as infants born to mothers who have not received antenatal glucocorticoids.
28.1.3.3.5 The Preterm Infant with Respiratory Distress Syndrome Who Is Not on Mechanical Ventilation

When a preterm infant has respiratory distress syndrome that is initially managed with CPAP or with supplemental oxygen through a hood, is it better to intubate him early, administer surfactant and extubate within an hour after brief mechanical ventilation, or to wait and see if the infant develops significant respiratory insufficiency, and if he does, only then intubate, give surfactant, and wean him off the ventilator gradually? Six randomized controlled clinical trials addressed this question and are summarized in a systematic review (Stevens Timothy et al. 2007). The rapid (within one hour) extubation attempted in these trials contrasts with the traditional approach – developed when surfactant therapy was first used – of keeping an infant on mechanical ventilation after surfactant administration, weaning ventilator support gradually as the pulmonary status improved, and extubating the infant from low ventilator settings. This approach of rapid extubation followed immediately by the use of nasal CPAP has been called “INSURE” (INtubate, SURfactant, Extubate to CPAP) and is intended to prevent ventilator-induced lung injury than can result from even brief periods of mechanical ventilation (Donn and Sinha 2006; Schmolzer et al. 2008). The six randomized trials, all of which are trials of “rescue” surfactant administration, are summarized in a systematic review (Stevens Timothy et al. 2007). Most of these studies included infants with a gestation of 35 weeks and below and a birth weight of 2,500 g and below. Many of the infants in these studies were between 32 and 35 weeks (few were extremely premature). In these studies of infants with signs and symptoms of RDS, intubation and early surfactant therapy followed by extubation to nasal CPAP (NCPAP) compared with later selective surfactant administration was associated with a lower incidence of mechanical ventilation (typical RR 0.67, 95 % CI 0.57–0.79; typical ARD −0.19, 95 % CI −0.26 to −0.11), air leak syndromes (typical RR 0.52, 95 % CI 0.28–0.96; typical ARD −0.04, 95 % CI −0.08 to 0.00), and BPD (typical RR 0.51, 95 % CI 0.26–0.99; typical ARD −0.08, 95 % CI −0.15 to −0.01). A larger proportion of infants in the early surfactant group received surfactant than in the selective surfactant group (typical RR 1.62, 95 % CI 1.41–1.86; typical ARD 0.38, 95 % CI 0.30–0.47). The number of surfactant doses per patient was significantly greater among patients randomized to the early surfactant group (WMD 0.57 doses per patient, 95 % CI 0.44–0.69). In stratified analysis by FiO2 at study entry, a lower threshold for treatment (FiO2 ≤0.45) resulted in lower incidence of air leak (typical RR 0.46 and 95 % CI 0.23–0.93; typical ARD −0.05, 95 % CI −0.10 to −0.01) and BPD (typical RR 0.43, 95 % CI 0.20–0.92; typical ARD −0.10, 95 % CI −0.19 to −0.02). A higher treatment threshold (FiO2 >0.45) at study entry was associated with a higher incidence of patent ductus arteriosus requiring treatment (typical RR 2.15, 95 % CI 1.09–4.13; typical ARD 0.12, 95 % CI 0.02–0.21). In another recent randomized trial (Rojas et al. 2009), infants 27–31 weeks gestation with RDS who were randomly assigned within the first hour of life either to intubation, very early surfactant, extubation, and nasal continuous positive airway pressure required less ventilation and had a lower incidence of mortality and air leaks (pneumothorax and pulmonary interstitial emphysema) than infants assigned to nasal continuous airway pressure alone. These data suggest that for a preterm infant with RDS who is not on mechanical ventilation (and is being managed on CPAP or an oxygen hood), early intubation at a low FiO2 threshold (FiO2<0.45), surfactant administration, and rapid extubation to CPAP decreases the risk of needing mechanical ventilation, BPD, and air leak syndrome, although it results in more surfactant use. Whether the same INSURE approach should be followed when prophylactic surfactant therapy is used is not clear due to lack of evidence.

28.1.3.3.6 Early Versus Late Treatment of Established RDS

Preterm infants who do not receive prophylaxis and subsequently develop RDS should be treated with surfactant as soon as possible. This strategy
is supported by many of the same arguments that support prophylactic surfactant administration as well as by clinical trials. Four randomized controlled trials (European Exosurf Study Group 1992; Gortner et al. 1998; Konishi et al. 1992; The OSIRIS Collaborative Group 1992), including the largest randomized trial conducted in neonatology (the OSIRIS trial), have evaluated early versus delayed selective surfactant administration. The results of these trials are summarized in a systematic review (Yost and Soll 2000). In these trials, early administration of surfactant consisted of administration of the first dose within the first 30 min to the first 2 h of life. Two of these studies used animal-derived surfactants and two used protein-free synthetic surfactant. The results of the meta-analysis of these studies are summarized in Fig. 28.3.

Early selective treatment resulted in a decrease in the risk of pneumothorax (typical RR 0.70, 95 % CI 0.59–0.82; typical ARD −0.05, 95 % CI −0.08 to −0.03), a decrease in the risk of pulmonary interstitial emphysema (typical RR 0.63, 95 % CI 0.43–0.93; typical ARD −0.06, 95 % CI −0.10 to −0.01), a decrease in the risk of chronic lung disease (requirement for supplemental oxygen at 36 weeks gestation, typical RR 0.70, 95 % CI 0.55–0.88; typical ARD −0.03, 95 % CI −0.05 to −0.01) and a decrease in the risk of neonatal mortality (typical RR 0.87, 95 % CI 0.77–0.99; typical ARD −0.03, 95 % CI −0.06 to 0.00). Therefore preterm infants who do not receive prophylactic surfactant and subsequently develop clinical features of RDS should receive the first dose of surfactant as early as possible. Outborn infants are at highest risk of delayed administration. Tertiary referral units accepting outborn infants should attempt to develop systems to ensure that surfactant is administered as early as possible to these infants, either by the transporting team or, if appropriate, by the referring hospital. In inborn infants, delays in administration of surfactant occur if other admission procedures such as line placement, radiographs, and nursing procedures are allowed to take precedence over surfactant dosing soon after birth. Surfactant administration should be given priority over such admission procedures.

### 28.1.3.3.7 Single Versus Multiple Surfactant Doses

Many of the initial trials of surfactant therapy tested a single dose of surfactant. However, surfactant may become rapidly metabolized and functional inactivation of surfactant can result from the action of soluble proteins and other factors in the small airways and alveoli (Jobe and Ikegami 1993). Administering repeat doses of surfactant can overcome such inactivation. The results of two randomized controlled trials

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**Fig. 28.3** Meta-analysis of four randomized controlled trials comparing early and delayed surfactant treatment (Yost and Soll 2000)
that compared multiple dosing regimens to single-dose regimens of animal-derived surfactant extract for treatment of established respiratory distress syndrome (Dunn et al. 1990; Speer et al. 1992) have been evaluated in a systematic review (Soll and Özek 2009). In one study (Dunn et al. 1990), after the initial dose of bovine lipid extract surfactant, infants assigned to the multiple-dose group could receive up to three additional doses during the first 72 h of life if they had a respiratory deterioration, provided they had shown a positive response to the first dose and a pneumothorax had been eliminated as the cause of the respiratory deterioration. In the other study (Speer et al. 1992), infants in the multiple-dose group received additional doses of poractant at 12 and 24 h after the initial dose if they still needed supplemental oxygen and mechanical ventilation. Approximately 70% of the infants randomized to the multiple-dose regimen received multiple doses.

The meta-analysis supports a decreased risk of pneumothorax associated with multiple dose surfactant therapy (typical RR 0.51, 95% CI 0.30–0.88; typical ARD −0.09, 95% CI −0.15 to −0.02). There was also a trend towards decreased mortality (typical RR 0.63, 95% CI 0.39–1.02; typical ARD −0.07, 95% CI −0.14 to 0.00). No differences were detected in other clinical outcomes. No complications associated with multiple-dose treatment were reported in these trials. In a third study, in which protein-free synthetic surfactant was used in a prophylactic manner, the use of two doses of surfactant in addition to a prophylactic dose lead to a decrease in mortality, respiratory support, necrotizing enterocolitis, and other outcomes when compared to a single prophylactic dose (Corbet et al. 1995). In the OSIRIS trial, which used protein-free synthetic surfactant, a two-dose treatment schedule was found to be equivalent to a treatment schedule permitting up to four doses of surfactant.

28.1.3.3.9 Comparisons Between Surfactant Products

Comparison of Animal-Derived and Protein-Free Synthetic Surfactants: Although both protein-free synthetic and animal-derived surfactants are effective, their composition differs. Animal-derived surfactant extracts contain surfactant-specific proteins that aid in surfactant adsorption and resist surfactant inactivation (Kuroki and Voelker 1994; Possmayer 1990). Eleven randomized trials have compared the effects of animal-derived and protein-free synthetic surfactants in the treatment or prevention of RDS (Ainsworth et al. 2000; Alvarado et al. 1993; da Costa et al. 1999; Horbar et al. 1993b; Hudak et al. 1996, 1997; Kukkonen et al. 2000;
Modanlou et al. 1997; Pearlman et al. 1993; Sehgal et al. 1994; Neonatal 1996). A total of over 4,500 infants were studied in these trials. A systematic review of these trials is available (Soll and Blanco 2001). The results of the meta-analysis are summarized in Fig. 28.4.

Compared to protein-free synthetic surfactant, treatment with animal-derived surfactant extracts resulted in a significant reduction in the risk of pneumothorax (typical RR 0.63, 95 % CI 0.53–0.75; typical ARD −0.04, 95 % CI −0.06 to −0.03) and the risk of mortality (typical RR 0.87, 95 % CI 0.76–0.98; typical ARD −0.02, 95 % CI −0.05 to 0.00). Natural surfactant extract is associated with a marginal increase in the risk of intraventricular hemorrhage (typical RR 1.09, 95 % CI 1.00–1.19; typical ARD 0.03, 95 % CI 0.00–0.06), but no increase in grade 3 to 4 intraventricular hemorrhage (typical RR 1.08, 95 % CI 0.92–1.28; typical ARD 0.01, 95 % CI −0.01 to 0.03). The meta-analysis also supports a marginal decrease in the risk of bronchopulmonary dysplasia or mortality associated with the use of natural surfactant preparations (typical RR 0.95, 95 % CI 0.90–1.01; typical ARD −0.03, 95 % CI −0.06 to 0.00).

In addition to these benefits, animal-derived surfactants have a more rapid onset of action, allowing ventilator settings and inspired oxygen concentrations to be lowered more quickly than with protein-free synthetic surfactant (Horbar et al. 1993b; Hudak et al. 1996; Modanlou et al. 1997; Rollins et al. 1993; Choukroun et al. 1994).

A comparison of physical properties and the results of animal studies also suggest that animal-derived surfactants have advantages over protein-free synthetic surfactants (Halliday 1996). These properties are attributed to the presence of the surfactant proteins SP-B and SP-C in animal-derived surfactants (Hall et al. 1992a).

The use of animal-derived surfactant preparations should be favored in most clinical situations, as their use results in greater clinical benefits than protein-free synthetic surfactants. However, all animal-derived surfactants have to be refrigerated for storage. The protein-free synthetic surfactant colfosceril is available as a lyophilized powder that is to be stored at below 30 °C in a dry place (not to be frozen) and reconstituted with sterile water before use. Therefore in situations where refrigeration is a problem (as in developing countries), it may be more practical to use colfosceril than animal-derived surfactants.

**Comparison of Animal-Derived and Protein-Containing Synthetic Surfactants:** Clinical trials have compared the effects of synthetic surfactants containing peptides to animal-derived surfactant preparations. These synthetic surfactants do not have the theoretical concerns of animal-derived surfactants, but they do not provide the same clinical benefits.
associated with animal-derived surfactants, namely, transmission of microorganisms, exposure to animal proteins and inflammatory mediators, susceptibility to inactivation, and inconsistent content (Engle and Committee on Fetus and Newborn 2008). Lucinactant, the synthetic surfactant containing an analog of SP-B, sinapultide, was compared with beractant in SELECT, a multicenter masked randomized trial of surfactant prophylaxis in infants 24–32 weeks gestation (Moya et al. 2005). Lucinactant was also compared with poractant in STAR, a multicenter randomized trial of surfactant prophylaxis in infants 24–28 weeks gestation that was structured as a non-inferiority trial (Sinha et al. 2005). A meta-analysis of these two studies (Pfister et al. 2007) found no significant differences in outcomes between lucinactant and the comparison animal-derived surfactant in mortality at 36 weeks post-menstrual age (typical RR 0.81, 95 % CI 0.64–1.03), chronic lung disease at 36 weeks post-menstrual age (typical RR 0.99, 95 % CI 0.84–1.18), the composite outcome of mortality or chronic lung disease at 36 weeks post-menstrual age (typical RR 0.96, 95 % CI 0.82–1.12), or in other respiratory outcomes. A decreased risk of necrotizing enterocolitis, a secondary outcome, was noted in infants receiving lucinactant (typical RR 0.60, 95 % CI 0.42–0.86; typical RD −0.06, 95 % CI −0.10 to −0.01).

However, both trials of lucinactant described above had multiple methodologic problems (Kattwinkel 2005) that undermine their validity, and at present there is no clear evidence of the equivalence or superiority of lucinactant over any animal-derived surfactant product (Halliday 2006). While these newer surfactants show promise, further research is required to elucidate their role in the prevention or treatment of RDS.

Comparison of Protein-Containing Versus Protein-Free Synthetic Surfactants: In the SELECT trial (Moya et al. 2005; Pfister Robert et al. 2009), the randomized trial of lucinactant mentioned above where it was compared with beractant, it was also compared to colfosceril. Compared to infants receiving colfosceril, infants receiving lucinactant had less RDS (39 % vs 47 %) and less RDS-related mortality (4.7 % vs 9.4 %, RR 0.50, 95 % CI 0.32–0.80). All-cause mortality at 36 weeks post-menstrual age was not significantly different (21 % for lucinactant vs 24 % for colfosceril). A trend towards a reduction in BPD or death at 36 weeks post-menstrual age was associated with lucinactant treatment when compared to colfosceril (RR 0.88, 95 % CI 0.77–1.01).

Comparison of Different Types of Bovine Surfactants: Two randomized trials, both from the same group of investigators, have compared the efficacy and adverse effects of different bovine surfactant products. In a comparison of beractant (Survanta) and calf lung surfactant extract (Infasurf) (Bloom et al. 1997), there were no differences detected between the two groups in the frequency of air leaks, complications associated with dosing, complications of prematurity, mortality, or survival without chronic lung disease. However, some differences were noted among subgroups of infants. Among infants treated for established respiratory distress syndrome, those who received calf lung surfactant extract had a significantly longer interval between doses, a lower inspired oxygen concentration, and a lower mean airway pressure in the first 48 h of life than infants treated with beractant. Among infants in whom these surfactants were administered in a preventive manner, mortality in infants with a birth weight <600 g was significantly higher with calf lung surfactant extract than with beractant. In a second report (Bloom et al. 2005) that included two separate trials – a prophylaxis trial and a treatment trial – the trials were halted prematurely due to recruitment problems and hence had inconclusive results with no demonstrated differences in outcomes between the two products. Thus, there is no evidence of the superiority of one bovine preparation over the other.

Comparison of Porcine and Bovine Surfactants: Five studies comparing surfactant treatment of established moderate to severe RDS with poractant versus beractant have been published (Baroutis et al. 2003; Speer et al. 1995; Malloy et al. 2005; Halahakoon 1999; Ramanathan et al. 2004).

A meta-analysis of these studies (Halliday 2005) found that compared to beractant,
poractant treatment led to a significant reduction in neonatal mortality (typical RR 0.57, 95% CI 0.34–0.96). The dose of beractant was uniformly 100 mg/kg across all five studies. When only studies that used a 100 mg/kg dose of poractant were considered, the reduction in mortality was not statistically significant (typical RR 0.82, 95% CI 0.44–1.58), emphasizing the fact that the most significant effect on mortality was seen with a 200 mg/kg dose of poractant (typical RR 0.29, 95% CI 0.10–0.79). Two of these five studies (Speer et al. 1995; Ramanathan et al. 2004) also reported more rapid improvement in oxygenation with poractant compared to beractant. The difference in outcomes described above between poractant and beractant may well be related to the dose of phospholipids and not to other characteristics of the products. There are no studies to determine whether poractant, especially in a 100 mg/kg dose, is superior to beractant when surfactant is dosed for prophylaxis.

28.1.3.3.10 Factors Affecting the Response to Surfactant Therapy

Several factors have been reported by various authors to be associated with a poor response to surfactant therapy, either in terms of immediate pulmonary response or in terms of later morbidity and mortality. These factors include high total fluid and colloid intake in the first days of life (Hallman et al. 1993), a low mean airway pressure relative to the FiO₂ (Hallman et al. 1993), the presence of an additional pulmonary disorder such as infection (Segerer et al. 1991) and perinatal asphyxia, other complications of prematurity (Konishi et al. 1992), high fraction of inspired oxygen requirement at entry (had a negative impact on a/APO₂ and 24 h after treatment), lower birth weight, male sex, outborn status, and high airway pressure requirement at entry (Collaborative European Multicentre Study Group 1991). Low birth weight, low Apgar score, and initial disease severity were associated with an increased mortality (Herting et al. 1992).

A high pulmonary resistance prior to therapy was associated with a poor response to therapy at 24 and 48 h (Wallenbrock et al. 1992). In addition, the immediate response to surfactant therapy itself has been reported to be a significant prognostic indicator for mortality and morbidity (Kuint et al. 1994). In animal studies, poor response to surfactant has been associated with delayed administration (Seidner et al. 1995) and the leakage of proteinaceous fluid into the alveolar spaces. Within some multicenter trials, significant differences in outcomes of surfactant-treated infants have been noted between participating hospitals (Collaborative European Multicentre Study Group 1991; Herting et al. 1992), suggesting that variations in patient care practices have an important influence on the outcomes of surfactant-treated infants.

As noted earlier, observational studies have demonstrated a decrease in mortality and morbidity for such infants after the introduction of surfactant therapy. However, racial differences in this decline in mortality have been reported. In one study, the overall neonatal mortality for black very low birth weight infants did not change after the introduction of surfactant therapy (Hamvas et al. 1996), and in another study, declines in neonatal mortality risks caused by respiratory distress syndrome and all respiratory causes were greater for non-Hispanic white VLBW infants than for black VLBW infants (Ranganathan et al. 2000). While such racial differences have been noted at a population level, the role of racial factors in the response pattern of individual infants with respiratory distress syndrome to exogenous surfactant therapy is unknown.

28.1.3.4 Adverse Effects of Surfactant Therapy

Transient hypoxia and bradycardia can occur due to acute airway obstruction immediately following surfactant instillation (Zola et al. 1993a; Liechty et al. 1991). Other acute adverse effects of surfactant administration include reflux of surfactant into the pharynx from the endotracheal tube, increase in transcutaneous carbon dioxide tension, tachycardia, gagging, and mucous plugging of the endotracheal tube. These complications of surfactant administration generally respond to a slower rate of surfactant administration or to an increased airway pressure or FiO₂.
during administration. Rapid improvement in oxygenation after surfactant administration necessitates close monitoring and appropriate reduction of ventilatory parameters.

Several authors have reported a transient decrease in blood pressure (Hellstrom-Westas et al. 1992; Skov et al. 1992a, b), a transient decrease in cerebral blood flow velocity (Cowan et al. 1991; Murdoch and Kempley 1998; Edwards et al. 1992), a transient decrease in cerebral oxyhemoglobin concentration (Edwards et al. 1992), and a transient decrease in cerebral activity on amplitude-integrated electroencephalography (Hellstrom-Westas et al. 1992) immediately after surfactant administration. The EEG depression observed after surfactant instillation is not caused by cerebral ischemia (Bell et al. 1994), and the EEG suppression is not directly related to alterations in blood gases or systemic circulation (Lundstrom and Greisen 1996). The clinical significance of these findings is uncertain. One study (Horbar et al. 1990) reported an increase in the incidence of intraventricular hemorrhage and a case report documents a temporal association between the development of intraventricular hemorrhage and the administration of surfactant-TA to improve respiratory failure caused by pulmonary hemorrhage (Funato et al. 1992). However, the meta-analyses of multiple trials do not show an increase in the risk of intraventricular hemorrhage with surfactant therapy compared to placebo (Seger and Soll 2009; Soll 1998; Soll and Özek 1997, 2010).

There is well-described increase in the risk of pulmonary hemorrhage with surfactant therapy (Raju and Langenberg 1993; Tomaszewska et al. 1999). Although trials in which animal-derived surfactants were used reported a higher incidence (5–6 %) of pulmonary hemorrhage than trials of protein-free synthetic surfactant (1–3 %), direct comparison demonstrates no difference in the risk of pulmonary hemorrhage. The overall incidence of pulmonary hemorrhage was low and the absolute magnitude of the increased risk is small (Raju and Langenberg 1993). However, moderate and severe pulmonary hemorrhage is associated with an increased risk of death and short-term morbidity. It is not associated with increased long-term morbidity (Pandit et al. 1999). The occurrence of pulmonary hemorrhage may be related to the presence of a hemodynamically significant patent ductus arteriosus (Garland et al. 1994). Seppanen et al. studied the association of neonatal complications with the Doppler-derived aortopulmonary pressure gradient (APPG) across the ductus arteriosus, which reflects pulmonary artery pressure during the first day of life. Infants in whom the APPG decreased after birth had a lower frequency of patent ductus arteriosus and pulmonary hemorrhage than those whose APPG remained low (Seppanen et al. 1995). Another mechanism for the pulmonary hemorrhage may be a direct cytotoxicity, which has been demonstrated in in vitro studies and appears to be different for different surfactants and different dosages (Findlay et al. 1995).

When surfactant initially became available for clinical testing, there was concern that the introduction of foreign proteins from animal-based lung surfactants into the lungs of preterm infants could lead to immunological responses. Two studies did not find antibodies specific to surfactant protein in the sera of preterm infants treated with bovine surfactant (Bartmann et al. 1992; Whitsett et al. 1991). In other studies, immune complexes or antibodies to the protein in exogenous porcine, bovine, or human surfactant have been identified in the sera of neonates with respiratory distress syndrome. However similar immune complexes or antibodies were also noted in control infants who did not receive surfactant, and no significant differences were noted between surfactant-treated and control infants (Chida et al. 1991; Strayer et al. 1989; Robertson et al. 1992). The presence of antibodies in control infants may be the result of leakage of surfactant proteins into the circulation (Chida et al. 1991).

With animal-derived surfactants, there is a theoretical risk of the transmission of infectious agents, including bovine spongiform encephalitis with surfactants derived from bovine sources and other viral infections in swine. Organic solvent processing of phospholipids, terminal sterilization techniques, and screening of animal sources have been used to minimize this risk.
28.1.3.5 Long-Term Outcomes After Surfactant Therapy

Long-term outcomes after surfactant therapy have been well studied for protein-free synthetic surfactant. Follow-up studies of long-term outcomes after animal-derived surfactant therapy have consisted of small numbers of patients, with a variable proportion of survivors being tested. For both protein-free synthetic and animal-derived surfactant, the “long-term” outcomes reported consist of outcomes predominantly in the first 3 years of life, with very few reports of outcomes at school age or higher. Given these limitations, the evidence suggests that not only do more infants survive from surfactant therapy but they are also at no selective disadvantage for neurodevelopmental sequelae due to the surfactant therapy. Most comparisons of long-term outcomes have been between infants treated with surfactant and placebo. There are few or no comparisons of long-term outcomes between infants treated with different types of surfactant or different regimens of the same surfactant. The following sections mainly address comparisons between infants treated with surfactant and placebo.

28.1.3.5.1 Neurodevelopmental Outcomes

No significant differences have been reported in the long-term neurodevelopmental outcomes of infants treated with surfactant compared to those treated with placebo, either with protein-free synthetic surfactant (Corbet et al. 1995; Morley and Morley 1990; Courtney et al. 1995) or animal-derived surfactant (Hoekstra et al. 1994; Robertson et al. 1992; Dunn et al. 1988; Ferrara et al. 1991; Vaucher et al. 1988; Wagner et al. 1995; Ware et al. 1990).

28.1.3.5.2 Long-Term Respiratory Outcomes

Compared to infants treated with placebo, infants treated with surfactant in the neonatal period have been reported to have either improved (Abbasi et al. 1993; Pelkonen et al. 1998; Yuksel et al. 1993) or equivalent (Couser et al. 1993; Gappa et al. 1999; Walti et al. 1992) results on pulmonary function testing. Some studies have reported a lower frequency of subsequent clinical respiratory disorders in surfactant-treated infants compared to placebo (Vaucher et al. 1988; Sell et al. 1995), while others have reported no difference (Robertson et al. 1992; Morley and Morley 1990; Dunn et al. 1988; Abbasi et al. 1993) or a trend towards an increase in allergic manifestations (Ware et al. 1990).

28.1.3.5.3 Physical Growth

No significant differences have been reported in weight or height outcomes between surfactant- and placebo-treated infants on follow-up (Corbet et al. 1995; Robertson et al. 1992; Courtney et al. 1995; Ware et al. 1990; Abbasi et al. 1993; Pelkonen et al. 1998; Gappa et al. 1999; Sell et al. 1995).

28.1.3.5.4 Outcomes of Prophylactic Versus Rescue Treatment Strategies

Two studies compared the long-term outcomes of infants treated with prophylactic surfactant to those treated with a “rescue” strategy. In one, there were no differences at school age in neurodevelopmental outcome or in the results of pulmonary function testing between the two groups, though infants who had received prophylactic surfactant showed fewer clinical pulmonary problems than those that received rescue treatment (Sinkin et al. 1998). In another study, in which there was significant loss of infants to follow-up (and therefore a high likelihood of attrition bias), the mean scores on the Bayley scales of infant development at 12 months adjusted age were higher in the rescue group than in the prophylactic group (Vaucher et al. 1993).

28.1.3.6 Exogenous Surfactant Therapy for Conditions Other than RDS

28.1.3.6.1 Meconium Aspiration Syndrome

In non-controlled studies of human infants with meconium aspiration syndrome, improved oxygenation has been reported with exogenous surfactant therapy (Auten et al. 1991; Halliday et al. 1996; Khammash et al. 1993). A randomized
trial in infants greater than 34 weeks gestation (including infants with MAS) with severe respiratory failure on extracorporeal membrane oxygenation (ECMO) showed that infants treated with beractant had improved lung function, a shorter duration of ECMO, and fewer complications after ECMO (Lotze et al. 1993).

Four randomized trials (Lotze et al. 1998; Findlay et al. 1996; Chinese Collaborative Study Group for Neonatal Respiratory Diseases 2005; Maturana et al. 2005) have studied the effect of animal-derived surfactant in term infants with meconium aspiration syndrome and are included in a systematic review. In these trials, surfactant therapy was administered as a continuous infusion over 20 min (Findlay et al. 1996) or as a bolus. The meta-analysis of these four trials (El Shahed et al. 2007) showed a decreased need for extracorporeal membrane oxygenation with surfactant therapy (typical RR 0.64, 95% CI 0.46–0.91; typical ARD −0.17, 95% CI −0.30 to −0.04). One trial reported a reduction in the length of hospital stay (mean difference −8 days (95% CI −14 to −3 days)). There were no statistically significant effects on mortality (typical RR 0.98 (95% CI 0.41–2.39), typical ARD 0.00 (95% CI −0.05 to 0.05)) or other outcomes (duration of assisted ventilation, duration of supplemental oxygen, pneumothorax, pulmonary interstitial emphysema, air leaks, chronic lung disease, need for oxygen at discharge, or intraventricular hemorrhage).

In summary, infants with severe meconium aspiration syndrome are likely to benefit from treatment with animal-derived surfactants. Multiple doses are usually required in such infants. Only animal-derived surfactants have been tested in human clinical trials in this setting. Each dose should be administered cautiously, with close cardiac, respiratory, and oxygen saturation monitoring, because surfactant can aggravate preexisting airway obstruction from meconium and transient oxygen desaturation and endotracheal tube obstruction have been reported with bolus administration in nearly one-third of infants (Lotze et al. 1998).

Investigators have also attempted to treat MAS by lavaging the airways with diluted surfactant solutions in order to wash out residual meconium (Dargaville et al. 2007; Wiswell et al. 2002; Hung et al. 2006; Lista et al. 2006; Gadzinowski et al. 2008). When this approach was tested in a randomized trial with a small number of babies, there were no statistically significant differences in clinical outcomes, although there was a trend towards reduction in the combined outcome of death or need for ECMO (Dargaville et al. 2010).

28.1.3.6.2 Acute Respiratory Distress Syndrome

Surfactant dysfunction is well described in acute lung injury (Willson et al. 2008). Therefore, surfactant replacement has been proposed as a treatment for patients with acute lung injury and the acute respiratory distress syndrome (ARDS), which, although more common in adults and older children, can occur in term neonates (Faix et al. 1989; Pfenninger et al. 1991). Exogenous surfactant therapy has been attempted in ARDS in adults but the results of clinical trials have not been promising (Anzueto et al. 1996; Gregory et al. 1997a). There are no randomized trials of exogenous surfactant therapy specifically for ARDS in neonates, but in older children with acute respiratory failure, surfactant use decreased mortality and duration of ventilation (Willson et al. 1999; Duffett et al. 2007). Because of this, and based on the pathophysiologic, clinical, and radiologic similarities between RDS and ARDS, it is reasonable to provide exogenous surfactant therapy to term infants with clinical and radiologic features of ARDS (severe respiratory failure with pulmonary opacification and air bronchograms on chest radiographs). The use of surfactant in ARDS is discussed in detail in Sect. 28.2.

28.1.3.6.3 Other Conditions

There are reports (single-case reports or case series) of the use of exogenous surfactant therapy in human infants for the management of pulmonary hemorrhage (Pandit et al. 1995; Amizuka et al. 2003) and neonatal pneumonia (Auten et al. 1991; Robertson 1996; Herting et al. 2000; Fetter et al. 1995). However, the efficacy of surfactant in these conditions is uncertain and its routine use in these conditions cannot be recommended. Surfactant therapy for infants with congenital diaphragmatic hernia has also been attempted
(Bos et al. 1991; Lotze et al. 1994; Glick et al. 1992; Lally et al. 2004; Van Meurs and Congenital Diaphragmatic Hernia Study Group 2004) but actually resulted in worse outcomes and therefore is not recommended.

28.1.4 Future Developments

Future research in neonatal surfactant therapy will likely attempt to understand the functioning and clinical effects of protein-containing synthetic surfactants. Emerging research is also addressing different methods of surfactant delivery to the lungs.

Surfactant administration through a laryngeal mask airway is noninvasive, avoids endotracheal intubation, and has been reported in a series of eight preterm infants with RDS managed with nasal CPAP (Trevisanuto et al. 2005). The mean arterial-to-alveolar oxygen tension ratio improved significantly after the treatment and no complications were reported. This method of administration is promising, as it potentially avoids the complications associated with intubation, but requires testing in a large randomized trial before it can be recommended.

Another noninvasive method of surfactant administration is instillation of surfactant into the nasopharynx during or immediately after delivery and before the first breath. Such instillation is thought to cause the surfactant to be aspirated into the fluid-filled airway as an air–fluid interface is established. A case series (Kattwinkel et al. 2004) of 23 preterm infants 27–30 weeks receiving such intrapartum nasopharyngeal instillation of surfactant followed by placement on CPAP immediately after birth (mask CPAP initially followed by nasal CPAP) demonstrated the feasibility of such administration. However, more evidence is required to prove the efficacy of this approach before it can be used or recommended.

The administration of surfactant to spontaneously breathing infants on nasal CPAP by passing a thin catheter into the trachea has been reported, initially from a single center (Kribs et al. 2008) and subsequently from a nonrandomized multicenter study (Kribs et al. 2010). While this method is promising, it requires rigorous evaluation in randomized trials. It is now being evaluated in a multicenter study called the AMV (Avoid Mechanical Ventilation) trial.

Finally, future research is likely to (or should) address methods of better identifying infants with respiratory disorders who would benefit from surfactant therapy. Current methods of selecting infants for surfactant therapy are primarily demographic (e.g., infants below a certain gestation), clinical (signs of respiratory distress with oxygen requirement), or radiographic (radiographic features of RDS). The development of more sophisticated and highly accurate tests to identify infants who will benefit from surfactant and those in whom surfactant therapy is not required will be very useful to clinicians.

Essentials to Remember

- Exogenous surfactant therapy is now a standard therapy for respiratory distress syndrome. Numerous randomized controlled trials have proved its efficacy and compared different products and treatment strategies.
- In infants with RDS, surfactant should be administered as early as possible after its need is identified.
- Animal-derived surfactant products are superior to protein-free synthetic surfactants. A new class of protein-containing synthetic surfactants is being tested for efficacy and might prove to have an important role in exogenous surfactant therapy in the future.
- While prophylactic surfactant administration to preterm infants at risk of RDS was previously the preferred practice compared to “rescue” surfactant administration, emerging new evidence suggests that initial stabilization of preterm infants on nasal continuous positive airway pressure might be equally effective, requires fewer resources, and avoids ventilator-induced lung injury.
- Exogenous surfactant therapy has potential benefits in term infants with meconium aspiration syndrome.
28.2  Exogenous Surfactant in the Pediatric Patient

Douglas Willson

**Educational Aims**
- To briefly review the history of surfactant therapy in neonates and in older children and adults
- To review the physiology underlying the potential value of exogenous surfactant therapy in ALI/ARDS
- To describe the different types of pharmaceutical surfactants
- To detail the animal and human studies examining the effects of exogenous surfactant in ALI/ARDS
- To speculate on the future role of exogenous surfactant as a component of therapy for ALI/ARDS

28.2.1 Introduction

With the identification of surfactant deficiency as the putative cause of infantile respiratory distress syndrome (IRDS) by Avery and Mead in 1959 (Avery and Mead 1959) and the closely following recognition of dipalmitoyl phosphatidylcholine (DPPC) as a major surfactant lipid component (Brown 1962, 1964), it was not long before treatment with this compound as a substitute “lung surfactant” was attempted. The studies of both Robillard et al. in 1964 (1964) and Chu et al. in 1967 (1967) tried unsuccessfully to treat infants with established IRDS (called hyaline membrane disease at the time) using aerosolized DPPC. The failure of this initial “surfactant replacement therapy,” together with a small but apparently positive physiological response to pulmonary vasodilatation, led to the erroneous conclusion that IRDS was due to ischemia and that surfactant deficiency was the result and not the cause of this disease (Chu et al. 1965, 1967). This misinterpretation was widely accepted until Enhorning and colleagues (Enhorning et al. 1973a, b) in 1973 demonstrated that whole surfactant isolated from adult rabbits could induce near-normal gas exchange when instilled into the trachea of premature rabbit pups, documenting that their respiratory failure was caused by surfactant deficiency. Moreover, it was not until 1981 that Fujiwara et al. (Fujiwara 1981) reported positive responses in respiratory function in human infants treated with a bovine-derived exogenous lung surfactant. We now understand that pulmonary surfactant is more than just DPPC, and that multiple lipid and peptide components are required for full activity. It is also now appreciated that the attempts of Robillard et al. (1964) and Chu et al. (1967) to aerosolize DPPC were not effective in delivering adequate amounts of material to the alveoli. Intratracheal instillation of exogenous surfactants in aqueous suspension is the current standard of care for the treatment and prevention of IRDS.

The ongoing development of surfactant therapy for clinical acute lung injury (ALI) and the acute (formerly “adult”) respiratory distress syndrome (ARDS) has been subject to some of the same issues that occurred during early attempts at this intervention in premature infants with IRDS. It has been known for some time that endogenous surfactant becomes dysfunctional in many forms of acute inflammatory lung injury, providing a direct conceptual rationale for exogenous surfactant supplementation. Despite this rationale, initial controlled studies attempting to treat adults with ARDS with exogenous surfactants were unsuccessful (Anzueto et al. 1996; Gregory et al. 1997b) and prompted calls for abandoning further clinical trials (Matthay 1996). However, the exogenous surfactants used in these studies (Anzueto et al. 1996; Gregory et al. 1997b) lacked one or more highly active components found in native surfactant, and the patients had sepsis-induced ARDS that involved substantial extrapulmonary pathology. Other studies of exogenous surfactant therapy in patients with ALI/ARDS give reason for cautious optimism about the use of this intervention, particularly in the case of direct pulmonary forms of these syndromes. This chapter briefly reviews the history and rationale for surfactant therapy in ALI/ARDS.
**28.2.2 Evidence of Surfactant Dysfunction in ALI/ARDS**

Unlike IRDS, surfactant deficiency is not a major factor in most forms of ALI/ARDS. Instead, surfactant dysfunction (inactivation, inhibition) induced by inflammatory lung injury is an important contributor to pathophysiology. In ALI/ARDS, an initially functional pulmonary surfactant system becomes “collateral damage” during whatever primary process injures the lung. Surfactant dysfunction can occur whether the cause of lung injury originates on the alveolar side (“direct” lung injury) or from the vascular side (“indirect” or “extrapulmonary” lung injury). However, both direct and indirect etiologies of injury induce pulmonary inflammation, alveolocapillary membrane injury, permeability edema, and reactive vasoconstriction or related vascular dysfunction. Surfactant dysfunction can occur by multiple mechanisms in the injured lungs, as shown in the figure and described in the text. The resulting loss of surface-active function contributes to acute respiratory failure with decreased lung volumes, decreased compliance, and severe ventilation/perfusion mismatching (Adapted from Wang et al. (2005a))

**Fig. 28.5** Schematic diagram illustrating pathways that can contribute to surfactant dysfunction in acute inflammatory pulmonary injury. Initiators of lung injury can act initially either from the alveolar side (“direct” lung injury) or from the vascular side (“indirect” or “extrapulmonary” lung injury). However, both direct and indirect etiologies of injury induce pulmonary inflammation, alveolocapillary membrane injury, permeability edema, and reactive vasoconstriction or related vascular dysfunction. Surfactant dysfunction can occur by multiple mechanisms in the injured lungs, as shown in the figure and described in the text. The resulting loss of surface-active function contributes to acute respiratory failure with decreased lung volumes, decreased compliance, and severe ventilation/perfusion mismatching (Adapted from Wang et al. (2005a))

1. Inhibition of surfactant biophysical function by plasma proteins (Holm et al. 1985; Seeger et al. 1985a, b, 1993; Holm and Notter 1987; Holm et al. 1988; Fuchimukai et al. 1987; Keough et al. 1989; Wang and Notter 1998; ) or other blood components such as fatty acids (Wang and Notter 1998; Seeger et al. 1985b; Hall et al. 1990, 1992b, 1994; Holm et al. 1999) that leak into the alveolar space as a result of alveolocapillary membrane injury and decreased barrier integrity
2. Alterations in alveolar surfactant aggregates whereby the most active “large aggregate” forms of surfactant are reduced in activity and/or percent content, while less active “small aggregate” forms of surfactant become more prevalent (Wang et al. 2005a; Hall et al. 1994; Davidson et al. 2005; Russo et al. 2002, 2007; Wright et al. 2001; Hickman-Davis et al. 2007; Raghavendran et al. 2008a; Lewis
et al. 1990; Günther et al. 1996; Veldhuizen et al. 1995)

3. Inhibition and/or chemical alteration of components in the alveolar surfactant film induced by cell membrane lipids (Holm and Notter 1987; Wang and Notter 1998; Holm et al. 1999; Cockshutt and Possmayer 1991; Wang et al. 2005b, 2008), meconium (Moses et al. 1991), or other substances present during the innate pulmonary inflammatory response such as proteases (Pison et al. 1989a), phospholipases (Holm et al. 1991; Enhorning et al. 1992; Wang et al. 2007), or reactive oxygen/nitrogen species (Seeger et al. 1985b; Hickman-Davis et al. 2001; Haddad et al. 1993; Amirkhanian and Merritt 1998)

4. An altered synthesis, secretion, or composition of active surfactant due to injury-induced changes in alveolar type II pneumocytes, which are stem cells for the alveolar epithelium in addition to being the primary cells of lung surfactant metabolism (Finkelstein 1990; Finkelstein et al. 1992; Mason et al. 1977; Mason and Williams 1997)

Abnormalities in surfactant composition and/or activity have been well documented in bronchoalveolar lavage (BAL) from patients with many forms of ALI/ARDS e.g., (Günther et al. 1996; Veldhuizen et al. 1995; Seeger et al. 1990; Pison et al. 1989b; Gregory et al. 1991; Griese 1999; Schmidt et al. 2007; Greene et al. 1999). Regardless of etiology, the practical consequences of surfactant dysfunction in ALI/ARDS are not dissimilar to those found in IRDS. As a result of decreased surfactant activity, the lungs become less compliant, with a progressive loss of aerated volume and an increased mismatching of ventilation with perfusion. Hypoxia, respiratory failure, and the need for respiratory support then ensue. In addition to surfactant dysfunction, ALI/ARDS also involves pulmonary inflammation and vascular dysfunction that can significantly impact overall patient outcomes and responses to therapy. Moreover, systemic inflammation and multiorgan (extrapulmonary) pathology are prominent in “indirect” forms of ALI/ARDS. The complex pathophysiology of ALI/ARDS is reviewed elsewhere (e.g., Bernard et al. 1994; Ware and Matthay 2000; Knight and Rotta 2005; Artigas et al. 1998; Krafft et al. 1996; Rubenfeld et al. 2005; Raghavendran et al. 2008b; DeBruin et al. 1992; Flori et al. 2005).

In considering surfactant dysfunction and therapy in ALI/ARDS, it is also important to have the perspective that lung injury is not a static phenomenon. Surfactant dysfunction, and hence surfactant replacement, is most important mechanistically in the acute exudative phase of ALI/ARDS. However, lung injury can progress to more chronic fibroproliferative and fibrotic stages, including superimposed effects from iatrogenic barotrauma, volutrauma, or atelectrauma induced by mechanical ventilation (“ventilator induced lung injury,” VILI) (Dos Santos and Slutsky 2000; Ricard et al. 2001; The Acute Respiratory Distress Syndrome Network 2000; Pelosi and Negrini 2008). The presence of evolving lung injury, together with inflammation and other non-surfactant factors in the pathophysiology of ALI/ARDS, complicates assessments of the long-term efficacy of exogenous surfactant therapy. Even if acute surfactant dysfunction is reversed effectively, long-term therapeutic benefits may not be apparent due to the presence of other aspects of disease. The long-term efficacy of surfactant therapy also depends on the specific activity of the exogenous surfactant used, the method of surfactant delivery, and almost certainly other variables that remain to be elucidated.

28.2.3 In Vitro Studies of Surfactant Inhibition

As noted above, multiple biophysical studies in vitro have shown that surfactant activity is reduced by exposure to albumin, hemoglobin, lysophospholipids, fatty acids, and other chemicals associated with lung injury ((Notter and Wang 1997; Wang et al. 2005a; Notter 2000) for detailed review). Importantly, these studies also document that inhibitor-induced inactivation can be overcome by raising surfactant concentration. The ability of exogenous surfactant supplementation to overcome inhibition in vitro is
not only a function of surfactant concentration but also depends on the content of essential apoproteins, particularly surfactant protein (SP)-B. Exogenous surfactants with higher contents of SP-B generally have greater activity and better inhibition resistance than those with little or no SP-B (Fig. 28.6). This is particularly relevant clinically, because the two major surfactant drugs studied unsuccessfully in the 1990s in controlled trials in adults with ARDS contained either no SP-B (Exosurf®) (Anzueto et al. 1996) or minimal levels of SP-B (Survanta®) (Gregory et al. 1997b).

28.2.4 Animal Studies of Exogenous Surfactant Therapy In Vivo

The effects of exogenous surfactant replacement have been studied in a large number of animal models of acute pulmonary injury (ALI/ARDS). Animal studies of acute injury most commonly examine responses to therapy over a timescale of hours, although some (e.g., hyperoxia) can address effects over longer times. Animal studies of acute pulmonary injury offer limited insight into long-term efficacy and the mitigation of chronic pathology, but are essential in providing a direct measure of the effectiveness of exogenous surfactants in mitigating injury-induced surfactant dysfunction and associated acute respiratory failure. Data from animal experiments on surfactant therapy have given insights into optimal preparations and delivery methods and are reassuring with regard to safety. Selected findings relating to exogenous surfactants and their efficacy in animal models of lung injury include the following (see Refs (Wang et al. 2005a; Notter 2000) for further review of animal studies of surfactant therapy in ALI/ARDS):

Type of lung injury: “Direct” lung injuries in animal models such as saline lavage (Lachmann et al. 1983; Kobayashi et al. 1984; Berggren et al. 1986; Lewis et al. 1996; Walther et al. 1997, 1998), acid aspiration (Kobayashi et al. 1990; Zucker et al. 1992; Schlag and Strohmaier 1993), viral infection (van Daal et al. 1991, 1992), or hyperoxic injury (Matalon et al. 1987, 1988; Loewen et al. 1989; Engstrom et al. 1989; Novotny et al. 1995) have been shown to respond well to the instillation of active exogenous surfactants. In contrast, significant benefits from surfactant replacement have not been documented in animal models of “indirect” lung injuries from systemic sepsis or intravenous oleic acid administration.

Fig. 28.6 Resistance of different clinical surfactants to inhibition by blood proteins. The graph shows the minimum surface tension reached by different clinical surfactants after 5 min of pulsation in a bubble surfactometer (37 °C, 20 cycles/min, 50 % area compression) in the presence of different concentrations of inhibitory blood proteins (fibrinogen) (a) and hemoglobin (b). Exogenous surfactants that most closely mimic natural surfactant (CLSE/Infasurf® and Alveofact®) are best able to resist inhibition and reach low surface tensions despite high levels of inhibitory proteins. Clinical exogenous surfactants are described and categorized in more detail in a subsequent section. Surfactant concentration for all preparations shown was uniform at 2 mg/ml (Data are from Seeger et al. (1993) as adapted by Notter (2000))
**Type of surfactant:** Surfactant preparations containing high levels of the hydrophobic surfactant proteins SP-B and SP-C are more effective physiologically than protein-free synthetic surfactants. Additionally, SP-B is known to have greater efficacy than SP-C in enhancing biophysical and physiological activity in exogenous surfactant mixtures (Curstedt et al. 1987; Oosterlaken-Dijksterhuis et al. 1991a, b, 1992; Revak et al. 1988; Seeger et al. 1992; Wang et al. 1996, 2002; Yu and Possmayer 1988; Notter et al. 2002), and supplementation with purified SP-B or synthetic SP-B peptides increases the activity of surfactants that contain SP-C in animal models of surfactant replacement (Walther et al. 1997; Notter et al. 2002; Mizuno et al. 1995).

**Timing of administration:** Early as opposed to late administration of exogenous surfactant has the most potential efficacy in ALI/ARDS for several reasons: (a) a better distribution of exogenous surfactant is possible because lung injury is more homogeneous early on; (b) early surfactant administration can help to moderate subsequent ventilator-induced lung injury; and (c) surfactant deficiency/dysfunction is most pronounced, and can be most specifically targeted, early in the course of lung injury. The benefits of early exogenous surfactant therapy have previously been documented clinically in premature infants (Bevilacqua et al. 1996; Kattwinkel et al. 1993; Kendig et al. 1998, 1991; OSIRIS Collaborative Group 1992).

**Method of delivery:** Delivery of exogenous surfactants by direct airway instillation has been found to be more effective than aerosolization (Notter 2000; Lewis et al. 1991, 1993a, b). Despite the theoretical advantages of aerosolization, it has proven difficult to deliver adequate amounts of surfactant to the alveoli using current technology. Also, aerosolized surfactant tends to go where inspired gas goes and consequently may not reach and recruit poorly aerated lung regions effectively (Lewis et al. 1993a, b).

**Other variables:** The efficacy of exogenous surfactants can also be affected by the method or mode of mechanical ventilation (Van Kaam et al. 2004), dosage amount and concentration, surfactant viscosity (King et al. 2002), rate of delivery, and other physical variables. Further animal studies, however, are necessary in order to study these variables systematically.

Ultimately, although animal studies have been (and are) indispensable in assessing exogenous surfactants and their physiological effects and activity, therapeutic efficacy must be determined in clinical studies where important outcome variables are directly assessed in patients.

### Table 28.2 Approximate biochemical composition of endogenous pulmonary surfactant

| Component                  | Composition          |
|----------------------------|----------------------|
| 85–90 % phospholipids      |                      |
| 80 % phosphatidylcholine   | (PC)                 |
| 40–50 % DPPC               |                      |
| 10–15 % other disaturated  |                      |
| 35–45 % unsaturated PCs    |                      |
| 15 % anionic phospholipids | (PG, PI, PS)         |
| 5 % other phospholipid     | classes (PE, Sph)    |
| 7–10 % apoproteins         | SP-A, SP-B, SP-C,    |
|                            | SP-D (not involved   |
|                            | in biophysical function) |
| 4–7 % neutral lipids       | Cholesterol,         |
|                            | Cholesterol esters,  |
|                            | Glycerides           |

Adapted from the research text of Notter (2000)

Values are representative averages in weight percent for surfactant lavaged (washed) from the lungs of normal animals of different species and ages

**Abbreviations:** PC phosphatidylcholine, PG phosphatidyglycerol, PI phosphatidylinositol, PS phosphatidylserine, PE phosphatidylethanolamine, Sph sphingomyelin, SP surfactant protein

### 28.2.5 Pharmaceutical Surfactants

Pharmaceutical surfactants and native pulmonary surfactant are not identical. Endogenous lung surfactant is a complex mixture of lipids (primarily phospholipids) and specific apoproteins that is highly conserved across mammalian species (Table 28.2). The degree of resemblance of native...
surfactant to pharmaceutical surfactants is highly variable, and the latter can be divided into three functionally relevant groups (Notter 2000; Notter and Wang 2008; Chess et al. 2005):

I. Organic solvent extracts of lavaged lung surfactant from animals (Alveofact®, BLES®, Infasurf®)

II. Organic solvent extracts of processed animal lung tissue with or without additional synthetic additives (Curosurf®, Survanta® or Surfactant-TA®)

III. Synthetic preparations not containing surfactant material from animal lungs (ALEC; Exosurf®, Surfaxin®; Venticute®)

Surfactants in Categories I and II are sometimes classified together as “animal-derived” or “natural” surfactant preparations. However, clinical surfactants in Category I have the closest compositional analogy to endogenous surfactant because they are obtained directly from recovered alveolar lavage fluid. Category I surfactant preparations in principle contain all the surfactant phospholipids plus the two hydrophobic surfactant proteins SP-B and SP-C in close approximation to the natural ratio (the hydrophilic surfactant proteins SP-A and SP-D are removed by organic solvent extraction in all Category I and Category II surfactants). Surfactant preparations in Category II also contain surfactant phospholipids and one or both of the hydrophobic surfactant proteins, but in addition they potentially contain constituents such as cellular lipids and/or hydrophobic fragments of cellular proteins because they are derived from processed lung tissue.

In addition to animal-derived surfactants, there is significant current interest in developing improved synthetic surfactant drugs (Category III surfactants above). Two early protein-free synthetic surfactants (ALEC and Exosurf®) are now no longer used because their activity is significantly less than existing animal-derived surfactants. Two newer synthetic surfactants are Surfaxin® (KL4) and Venticute® (recombinant SP-C surfactant). However, the 21 amino acid KL4 peptide in Surfaxin® has only very approximate molecular analogy to native SP-B, and Venticute® contains no SP-B peptide. Thus, the development of more optimal fully synthetic surfactants with a highly active SP-B peptide component with or without added novel lipids that can resist phospholipase-induced degradation in lung injury is an active area of investigation (Notter 2000; Wang et al. 2003, 2007; Notter and Wang 2008; Notter et al. 2007). Synthetic lung surfactants have significant potential advantages in purity, manufacturing, quality control, scale-up, and cost relative to animal-derived preparations (Notter and Wang 2008; Notter et al. 2007). Synthetic surfactants are also free from animal pathogens like prions, and they are not subject to cultural or religious issues that can affect bovine- or porcine-derived preparations. Simple dosage calculations illustrate the potential practical importance of synthetic surfactants in ALI/ARDS, since drug amounts 50–100 times larger than those used in premature infants are required in adults and older children to achieve an equivalent dose normalized by body weight. The dollar expense of surfactant therapy in adult patients will be extremely high unless drug costs per unit weight can be reduced substantially from the current price of 100–200 mg infant vials of animal-derived surfactants. Synthetic surfactant production involves no animal costs, and chemical manufacturing methods can theoretically be scaled-up and quality controlled much more efficiently to allow more cost-effective therapy of ALI/ARDS.

Regardless of category (animal-derived or synthetic), the requirements for an effective therapeutic surfactant in ALI/ARDS are more stringent than in the case of IRDS. To treat severe acute inflammatory lung injury, exogenous surfactants must have the greatest possible activity and resistance to inhibition or inactivation. Specific differences in composition among clinical exogenous lung surfactants are a major factor in predicting and interpreting their clinical efficacy. For example, the protein-free exogenous surfactant Exosurf® has some activity in treating IRDS (Bose et al. 1990; Corbet et al. 1991; Long et al. 1991a, b), but it is significantly less efficacious in premature infants than apoprotein-containing animal surfactants like Infasurf® (Hudak et al. 1996, 1997; Notter 2000), and it has
no clinical benefits in adults with ARDS (Anzueto et al. 1996). Similarly, Survanta® has been found to have minimal benefits in adults with sepsis-induced ARDS (Gregory et al. 1997b), which correlates with its very low content of active SP-B (Seeger et al. 1993; Mizuno et al. 1995; Hamvas et al. 1994). At the same time, Survanta® has significant activity due to its content of SP-C (Curstedt et al. 1995; Findlay et al. 1997a; Hamvas et al. 1996). Meconium aspiration syndrome (MAS) (Auten et al. 1991) also found significant reductions in the incidence of pneumothorax, duration of mechanical ventilation and oxygen therapy, time of hospitalization, and requirements for ECMO in 20 term infants with MAS treated with Survanta® compared to a similar number of

SP-B, the larger and most biophysically active (Curstedt et al. 1987; Oosterlaken-Dijksterhuis et al. 1991a, b, 1992; Revak et al. 1988; Seeger et al. 1992; Wang et al. 1996, 2002; Yu and Possmayer 1988; Notter et al. 2002) of the two hydrophobic surfactant proteins, is a particularly essential component for optimal surfactant function in vivo. Knock-out mice with isolated SP-B deficiency die shortly after birth of respiratory failure (Clark et al. 1995), and human infants with SP-B mutations do not survive beyond the first days of life without surfactant replacement (and ultimately lung transplant) (Hamvas et al. 1994, 1995, 1997; Whitsett et al. 1995). An elegant series of experiments by Ikegami et al. (2005) using a conditional knock-out mouse model demonstrated that adult mice rendered acutely deficient in SP-B develop severe respiratory distress with evidence of surfactant dysfunction and pulmonary inflammation. Mice left SP-B deficient died with pathology resembling ARDS, but the abnormalities were reversed and the mice survived if SP-B synthesis was restored. Interestingly, these mice maintained normal levels of the SP-C protein during study (Ikegami et al. 2005).

28.2.6 Human Studies of Surfactant Therapy for ALI/ARDS

Multiple clinical studies have reported measurable benefits following the instillation of active exogenous surfactants to term newborns, children, or adults with ALI/ARDS or lung injury-related acute respiratory failure (Auten et al. 1991; Khammash et al. 1993; Lotze et al. 1993, 1998; Findlay et al. 1996; Willson et al. 1996, 1999, 2005; Gunther et al. 2002; Walmrath et al. 1996, 2002; Spragg et al. 1994; Wiswell et al. 1999; Lopez-Herce et al. 1999; Hermon et al. 2002; Herting et al. 2002; Luchetti et al. 1998, 2002; Moller et al. 2003; Amital et al. 2008) (Table 28.3). However, while a number of these have been controlled trials, many have been uncontrolled pilot studies or case series that reported significant improvements primarily in acute lung function (arterial oxygenation). Moreover, as noted earlier, two large controlled trials not included in the positive studies in Table 28.3 either showed no benefit (Anzueto et al. 1996) or minimal benefits (Gregory et al. 1997b) from surfactant therapy in adults with sepsis-induced surfactant therapy. A more detailed summary of the clinical experience with surfactant therapy in term infants, children and adults with ALI/ARDS follows below.

Perhaps the best-studied application of surfactant therapy for an indication other than IRDS is in full-term infants with meconium aspiration syndrome (MAS) (Auten et al. 1991; Khammash et al. 1993; Lotze et al. 1993, 1998; Findlay et al. 1996). Meconium is a thick, tarry mixture of bile acids and mucous glycoproteins that fills the fetal colon during gestation, and prenatal defecation associated with maternal/fetal stress can lead to meconium aspiration at birth. Meconium mechanically obstructs airways, causes inflammation (Holopainen et al. 1999), and inhibits the biophysical activity of lung surfactant (Moses et al. 1991; Clark et al. 1987). Auten et al. (1991), Khammash et al. (1993), and Findlay et al. (1996) have all reported significant lung functional improvements following exogenous surfactant administration (Infasurf®, Survanta®) to infants with MAS. The randomized study of Findlay et al. (1996) also found significant reductions in the incidence of pneumothorax, duration of mechanical ventilation and oxygen therapy, time of hospitalization, and requirements for ECMO in 20 term infants with MAS treated with Survanta® compared to a similar number of
controls. Lotze et al. (1993, 1998) have also reported favorable results using Survanta® in a controlled trial in term infants referred for ECMO due to severe respiratory failure (meconium aspiration was a prevalent diagnosis). Twenty-eight infants treated with four doses of Survanta® (150 mg/kg) had improved pulmonary mechanics, decreased duration of ECMO treatment, and a lower overall incidence of complications after ECMO compared to control infants (Lotze et al. 1993). A subsequent larger multicenter controlled trial in 328 term infants similarly reported significant improvements in respiratory status and the need for ECMO following surfactant treatment (Lotze et al. 1998). Exogenous surfactant is now used in many institutions to treat neonates with MAS. Surfactant therapy is also frequently used in intensive care nurseries to treat neonates with respiratory failure from pneumonia, although controlled studies in the NICU setting have not been done.

Experience with clinical surfactant therapy in adults with ALI/ARDS is much less positive than in infants. Several small uncontrolled studies in adults with ALI/ARDS have reported acute lung functional improvements following surfactant

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**Table 28.3** Selected controlled and uncontrolled clinical studies reporting benefits of exogenous surfactant therapy in acute respiratory failure (ALI/ARDS)

| Study           | Patients (N) | Disease               | Surfactant     | Outcomes                                     |
|-----------------|--------------|-----------------------|----------------|----------------------------------------------|
| Günther et al.  | Adults (27)  | ARDS                  | Alveofact®     | Improved surfactant function                 |
| Walmrath et al. | Adults (10)  | ARDS, sepsis          | Alveofact®     | Improved oxygenation                         |
| Spragg et al.   | Adults (6)   | ARDS, multiple causes | Curosurf®      | Improved oxygenation and biophysical function|
| Wiswell et al.  | Adults (12)  | ARDS, multiple causes | Surfaxin®      | Improved oxygenation                         |
| Amital et al.   | Adults (42)  | Lung transplant       | Infasurf®      | Improved oxygenation, better graft function  |
| Willson et al.  | Children (29 & 42) | ARDS, multiple causes | Infasurf®      | Improved oxygenation                         |
| Willson et al.  | Children (152) | ARDS, multiple causes | Infasurf®      | Improved survival and improved ventilation   |
| Lopez-Herce et al. | Children (20) | ARDS + post-op cardiac | Curosurf®      | Improved oxygenation                         |
| Hermon et al.   | Children (19) | ARDS + post-op cardiac | Curosurf® or Alveofact® | Improved oxygenation |
| Herting et al.  | Children (8) | Pneumonia             | Curosurf®      | Improved oxygenation                         |
| Moller et al.   | Children (35) | ARDS, multiple causes | Alveofact      | Improved oxygenation                         |
| Auten et al.    | Infants (14) | MAS or pneumonia      | Infasurf® (CLSE) | Improved oxygenation |
| Lotze et al.    | Infants (28 & 328) | ECMO, multiple indications | Survanta®     | Improved oxygenation, decreased ECMO         |
| Khammash et al. | Infants (20) | MAS                   | bLES®          | Improved oxygenation in 75% of patients      |
| Findlay et al.  | Infants (40) | MAS                   | Survanta®      | Improved oxygenation, decreased pneumothorax, and mechanical ventilation |
| Luchetti et al. | Infants (20 & 40) | RSV bronchiolitis    | Curosurf®      | Improved oxygenation                         |

The tabulated studies of Willson et al. (1999; 2005), Findlay et al. (1996), Moller et al. (2003), Lotze et al. (1993, 1998), Luchetti et al. (1998, 2002), and Amital et al. (2008) were controlled trials, while the remaining studies were uncontrolled pilot trials as detailed in the text.

MAS meconium aspiration syndrome, RSV respiratory syncytial virus

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Pediatric and Neonatal Mechanical Ventilation
treatment (Gunther et al. 2002; Walmrath et al. 1996, 2002; Spragg et al. 1994; Wiswell et al. 1999) (Table 28.3). In contrast, the influential large controlled trial of Anzueto et al. (1996) administered nebulized Exosurf® versus placebo in 725 adults with ARDS secondary to sepsis and found no improvement in any measure of oxygenation and no effect on morbidity or mortality. As noted earlier, this study is now recognized as having several flaws: (1) the synthetic protein-free surfactant Exosurf® has significantly lower activity than animal-derived surfactants; (2) current aerosol technology has not been found to be as effective as airway instillation in administering surfactant (the surfactant dose delivered was estimated to be 5 mg/kg/day as opposed to the standard 100 mg/kg instilled dose used in infants with IRDS); and (3) recent data indicate that surfactant is less effective in treating ALI/ARDS from sepsis or other “indirect” causes of lung injury compared to “direct” pulmonary ALI/ARDS.

Two other controlled clinical trials reporting disappointing results in adults with ALI/ARDS are those of Gregory et al. (1997b) with Survanta® and Spragg et al. (2003) with recombinant SP-C surfactant (Venticute®). These results reinforce the importance of including highly active SP-B in exogenous surfactants used to treat ALI/ARDS in adults, since Survanta® contains only minimal levels of this apoprotein (Seeger et al. 1993; Mizuno et al. 1995; Hamvas et al. 1994) and Venticute® contains none. The trial of Gregory et al. (1997b) found small benefits in oxygenation for patients with sepsis-induced ALI/ARDS who received intermediate-sized doses of Survanta® (100 mg/kg), but no benefits in other surfactant dosage groups. There were no overall long-term benefits from surfactant therapy as measured by length of mechanical ventilation or survival in the total of 43 surfactant-treated patients studied (Gregory et al. 1997b). The study of Spragg et al. (2003) showed acute increases in oxygenation following instillation of Venticute® in adults with ARDS, but no longer-term improvements (duration of mechanical ventilation, length of stay, or mortality). A post hoc analysis suggested possible benefits from surfactant therapy in the subgroup of patients with ARDS from “direct” lung injury. However, a follow-up randomized trial of Venticute® limited to patients with direct lung injury was recently stopped after 900 patients because of a failure to demonstrate clinical benefits (Spragg R, 2009, personal communication). Controlled studies of surfactant therapy in children with ALI/ARDS have been much more encouraging compared to studies in adults. Luchetti et al. (1998, 2002) have reported two small controlled studies showing that treatment with porcine surfactant (Curosurf®, 50 mg/kg) improved gas exchange plus led to a reduced time on mechanical ventilation and in the pediatric intensive care unit (PICU) for infants with RSV bronchiolitis. An eight center randomized unblinded trial by Willson et al. (1999) in 42 children in the PICU with ALI/ARDS showed that those receiving Infasurf® (70 mg/kg) had immediate improvements in oxygenation and fewer ventilator days and days in intensive care. This trial followed an initial uncontrolled treatment study by the same group indicating improved oxygenation in 24 children (0.1–16 years) with ALI/ARDS treated with instilled Infasurf® (Willson et al. 1996). A study by Moller et al. (2003) has also reported that children with ARDS showed immediate improvement in oxygenation and less need for rescue therapy following treatment with Survanta®, but was underpowered to assess more definitive longer-term outcomes.

A larger and more recent blinded controlled study in 2005 by Willson et al. (2005) in PICU patients with ALI/ARDS showed that treatment with Infasurf® (calfactant) relative to placebo was associated both with immediate benefits to oxygenation as well as a significant survival advantage (Table 28.4). In a post hoc analysis, all of the benefits of surfactant therapy were in the 98-patient subgroup with “direct” pulmonary forms of ALI/ARDS (Table 28.5). The 54-patient subgroup with “indirect” forms of ALI/ARDS showed no acute or long-term benefits from calfactant. The significant benefits of calfactant in treating direct pulmonary ALI/ARDS in children found by Willson et al. (2005) have prompted an even larger prospective trial in both adults and children with direct lung injury (CARDS, Calfactant in ARDS trial, NCT00682500).
This ongoing study is a prospective, masked randomized controlled trial of calfactant versus placebo in children and adults with ALI/ARDS from “direct” lung injury in more than 30 centers in the USA, Canada, Korea, Israel, Australia, and New Zealand.

Exogenous surfactant has also recently been used successfully after lung transplant in both adults and children. A randomized controlled trial by Amital et al. (2008) in 42 adult patients after lung transplant showed that calfactant surfactant-treated patients had better oxygenation, less post-graft dysfunction, shorter ICU stay, and improved lung function at 1 month. This study followed multiple case reports showing clinical improvements from exogenous surfactant therapy after lung transplantation (Kermeen et al. 2007; Della Rocca et al. 2002; Struber et al. 1999, 2007).

None of the studies of surfactant therapy in ALI/ARDS described in this chapter have identified any adverse long-term side effects in patients, particularly those with direct pulmonary forms of lung injury. The transient hypoxia and hemodynamic instability that typically surround surfactant instillation in severely ill patients have previously been well described in newborns with IRDS and have repeatedly been shown not to adversely impact long-term outcomes. The risk of transmission of infectious agents such as prions from the use of animal-derived surfactants cannot completely be ruled out, although it is greatly reduced by the organic solvent extraction methods used to prepare such drugs (synthetic surfactants have no risk of prion transmission).

No meaningful systemic toxicities or side effects of animal or synthetic exogenous surfactants have been identified in a quarter century of use in newborn infants. Exogenous surfactant chemical components (phospholipids and proteins) are largely recycled in the lungs, and it is unlikely that the current FDA risk status of “no contraindications” for the newborn use of surfactant drugs will need to change for applications in older patients with ALI/ARDS.

In the absence of definitive data from controlled clinical trials, exogenous surfactant therapy is often used in practice as a “rescue” intervention when conventional therapies fail. However, experience from studies of surfactant therapy in neonates indicates that rescue therapy will be less helpful than treatment earlier in the

| Table 28.4 | Clinical outcomes from the randomized controlled trial of exogenous surfactant (calfactant) therapy in pediatric acute lung injury of Willson et al. (2005) |
|-------------------------------|---------------------|---------------------|---------------|
|                              | Calfactant (n=77)   | Placebo (n=75)      | P value       |
| **Mortality**                 |                     |                     |               |
| Died (in hospital)            | 15 (19 %)           | 27 (36 %)           | 0.03          |
| Died w/o extubation           | 12 (16 %)           | 24 (32 %)           | 0.02          |
| Failed CMV*                   | 13 (21 %)           | 26 (42 %)           | 0.02          |
| ECMO                          | 3                   | 3                   | n.s.          |
| Use of nitric oxide           | 9                   | 10                  | 0.80          |
| HFOV after entry              | 7                   | 15                  | 0.07          |
| **Secondary outcomes**        |                     |                     |               |
| PICU LOS                      | 15.2 ± 13.3         | 13.6 ± 11.6         | 0.85          |
| Hospital LOS                  | 26.8 ± 26           | 25.3 ± 32.2         | 0.91          |
| Days of O2 therapy            | 17.3 ± 16           | 18.5 ± 31           | 0.93          |
| Hospital charges              | $205 ± 220          | $213 ± 226          | 0.83          |
| Hospital charges/day          | $ 7.5 ± 7.6         | $ 7.9 ± 7.5         | 0.74          |

Data from Willson et al. (2005)

Calfactant Infasurf®, CMV conventional mechanical ventilation, ECMO extracorporeal membrane oxygenation, HFOV high-frequency oscillatory ventilation, NO nitric oxide therapy, PICU pediatric intensive care unit, LOS length of stay in days

*Failed CMV was defined as requiring a non-conventional therapy such as ECMO, NO, or HFOV (note that some patients had more than one nonconventional therapy)

Secondary outcomes are mean ± S.D.

| Table 28.5 | Post hoc analysis of patient outcomes as a function of direct versus indirect lung injury in the Infasurf® (calfactant) study of Willson et al. (2005) in children with ALI/ARDS |
|-------------------------------|---------------------|---------------------|---------------|
|                              | Placebo             | Calfactant          | P value       |
| **Direct lung injury**        |                     |                     |               |
| (# patients)                  | 48                  | 50                  |               |
| OI ↓ 25 %                    | 31 %                | 66 %                | 0.0006        |
| Ventilator days              | 17 ± 10             | 13 ± 9              | 0.05          |
| Died                          | 38 %                | 8 %                 | 0.0005        |
| **Indirect lung injury**      |                     |                     |               |
| (# patients)                  | 27                  | 27                  |               |
| OI ↓ 25 % +                  | 41 %                | 37 %                | 0.79          |
| Ventilator days              | 17 ± 10             | 18 ± 10             | 0.75          |
| Died                          | 33 %                | 41 %                | 0.65          |

Data from Lopez-Herce et al. (1999)

OI ↓ 25 % = a decrease of 25 % or more in oxygenation index as a measure of improvement in the severity of respiratory failure; ventilator days = days on mechanical ventilation
clinical course (e.g., Bevilacqua et al. 1996; Kattwinkel et al. 1993; Kendig et al. 1991, 1998; OSIRIS Collaborative Group 1992). A major conceptual benefit of surfactant therapy in ALI/ARDS is to enhance lung function and reduce the need for positive pressure mechanical ventilation that can otherwise worsen inflammation and cause iatrogenic lung injury. Administering surfactant late in the course of respiratory failure after prolonged mechanical ventilation has already occurred precludes this potential benefit. Evolving evidence documenting benefits for the most active exogenous surfactants in pediatric and/or adult patients with direct pulmonary ALI/ARDS will hopefully allow earlier and better-targeted use of this intervention in the future.

Finally, a major issue regarding the most optimal use of surfactant therapy in ALI/ARDS involves its combination with added agents or interventions that target other aspects of the complex pathophysiology of acute inflammatory lung injury. Combination therapy approaches may be particularly important in adults with ALI/ARDS, where responses to exogenous surfactant have so far been disappointing in terms of improving long-term outcomes. Combination therapies designed to exploit potential mechanistic synergy between surfactant and agents directed at different facets of lung injury may have a more substantial impact on long-term outcomes in patients with ALI/ARDS. The rationale for the use of exogenous surfactant therapy in specific combined-modality interventions for ALI/ARDS is described in more detail elsewhere (Raghavendran et al. 2008b; Notter et al. 2000; Pryhuber et al. 2005). Examples of agents that might be synergistic with exogenous surfactant in ALI/ARDS include vasoactive drugs such as inhaled nitric oxide, which has the potential to selectively enhance perfusion in newly ventilated lung regions recruited by exogenous surfactant (Raghavendran et al. 2008b; Notter et al. 2000; Pryhuber et al. 2005). Other drugs for possible use in combination with exogenous surfactant include anti-inflammatory antibodies or receptor antagonists, antioxidants, and antioxidant enzymes (Raghavendran et al. 2008b; Notter et al. 2000; Pryhuber et al. 2005). Specific ventilator modalities or ventilation strategies that reduce iatrogenic lung injury may be equally important to consider in conjunction with surfactant therapy (Raghavendran et al. 2008b; Notter et al. 2000; Pryhuber et al. 2005).

### Conclusions

Although exogenous surfactant replacement in premature infants with IRDS (hyaline membrane disease) was first attempted in the mid-1960s (Robillard et al. 1964; Chu et al. 1967) soon after the putative role of surfactant deficiency in this disease was identified, it took a generation longer for this therapy to actually be approved by the FDA for use in premature infants. Today lifesaving surfactant replacement therapy in premature infants is a crucial staple of neonatal intensive care. The development of surfactant replacement therapy for acute respiratory failure associated with lung injury (ALI/ARDS) has shared some of the same growing pains found in the case of IRDS, including the use of nonoptimal exogenous surfactants and delivery methods in initial controlled clinical trials. In addition, the complex multifaceted pathology of ALI/ARDS clearly presents more of a challenge for surfactant therapy compared to surfactant-deficient IRDS. It remains to be determined if the reversal of surfactant dysfunction in patients with ALI/ARDS will prove similarly lifesaving, despite the fact that a firm basic science rationale for exogenous surfactant therapy exists.

A functioning active lung surfactant film is required for successful respiration in humans and other air-breathing animals. Evidence is clear that endogenous surfactant becomes dysfunctional in direct forms of acute inflammatory lung injury, and exogenous surfactant therapy has been shown to mitigate lung injury severity in multiple animal models of ALI/ARDS. Treatment of humans with ALI/ARDS with exogenous surfactants in clinical studies has been associated with improved oxygenation and few side effects and has led to improved longer-term outcomes in several patient populations. In particular, surfactant
therapy has been documented to be beneficial in term infants with meconium aspiration lung injury, and it is also used in treating acute respiratory failure in neonatal pneumonia. Studies on surfactant therapy in children with direct pulmonary ALI/ARDS are also promising, with improvements found not only in lung function but also in longer-term variables (including survival in a recent study (Willson et al. 2005)). A substantial number of studies in adults with ALI/ARDS have also reported acute improvements in lung function (oxygenation) from surfactant therapy (Table 28.3). Lung functional improvements in principle have the potential to lower required ventilator distending pressures and reduce ventilator-induced lung injury and inflammatory cytokine production that can worsen patient outcomes. Significant statistical correlations have been demonstrated between acute improvements in lung function and improvements in clinically significant long-term outcomes in surfactant-treated infants with IRDS (Segerer et al. 1991; Kuint et al. 1994).

However, demonstrating improved long-term outcomes from exogenous surfactant therapy in patients with ALI/ARDS is complicated by several factors. These lung injury syndromes have a complex multifaceted pathophysiology that includes not only surfactant dysfunction but also prominent elements of inflammation, vascular dysfunction, and oxidant injury. In addition, progressive fibroproliferative lung injury may be present, as well as multiorgan pathology, particularly in patients with “indirect” or “extrapulmonary” ALI/ARDS. This multifaceted pathology reduces the resolving power of clinical studies to demonstrate beneficial effects on long-term outcomes from surfactant therapy, even if the intervention itself is effective in mitigating its targeted aspect of lung injury (i.e., surfactant dysfunction). Controlled trials in adults with ALI/ARDS have not yet shown that surfactant therapy can substantially improve long-term outcomes, although exogenous surfactants with the greatest content of highly active SP-B have not been tested in detail in adults. Another major area that has received relatively little study in clinical trials to date is the use of exogenous surfactant therapy in combination with potentially synergistic agents or interventions that target additional aspects of the complex pathophysiology of acute inflammatory lung injury. This latter approach may ultimately be particularly important in obtaining the most substantial reductions in mortality and long-term morbidity for pediatric and adult patients with severe ALI/ARDS-related acute respiratory failure.

**Essentials to Remember**

- Lung surfactant is a complex mixture of phospholipids and proteins. Pharmaceutical surfactants deficient in any of these components, particularly in the surfactant-associated proteins B and C, are unlikely to be effective in ALI/ARDS.
- Animal and human data support that exogenous surfactant improves immediate lung function in ALI/ARDS but proof of longer-term efficacy requires further study.
- Surfactant dysfunction and inhibition occurs early in ALI/ARDS and much of subsequent injury is likely consequent to positive pressure ventilation of the surfactant-deficient lung. Consequently, as in treatment of infantile respiratory distress syndrome, if treatment is to be effective, it should be administered early.
- Side effects of surfactant therapy are limited to transient hypoxia and hypotension associated with instillation, as the drug is not systemically absorbed. Better methods of aerosol delivery in the future may obviate these side effects as well as allow administration in the non-intubated patient.
- At present, treatment of ALI/ARDS with exogenous surfactant remains experimental.
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