Effects of 90-Day Feeding of Transgenic Maize BT799 on the Reproductive System in Male Wistar Rats

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Abstract: BT799 is a genetically modified (GM) maize plant that expresses the Cry1Ac gene from Bacillus thuringiensis (Bt). The Cry1Ac gene was introduced into maize line Zhen58 to encode the Bt crystal protein and thus produce insect-resistant maize BT799. Expression of Bt protein in planta confers resistance to Lepidopteran pests and corn rootworms. The present study was designed to investigate any potential effects of BT799 on the reproductive system of male rats and evaluate the nutritional value of diets containing BT799 maize grain in a 90-day subchronic rodent feeding study. Male Wistar rats were fed with diets containing transgenic BT799, Zhen58 and the control. No significant differences in body weight, hematology and serum chemistry results were observed between rats fed with the diets containing transgenic BT799, Zhen58 and the control in this 13-week feeding study. Results of serum hormone levels, sperm parameters and relative organ/body weights indicated no treatment-related side effects on the reproductive system of male rats. In addition, no diet-related changes were found in necropsy and histopathology examinations. Based on results of the current study, we did not find any differences in the parameters tested in our study of the reproductive system of male rats between BT799 and Zhen58 or the control.

Keywords: transgenic maize; BT799; Bacillus thuringiensis; reproductive toxicity; sperm parameters

1. Introduction

Microbial Bacillus thuringiensis (Bt) products with toxicity to Lepidopteran species have been used as insecticides for more than 40 years. Ingestion of select Bt proteins lead to interactions between the Bt protein and receptors in the gut epithelial cells of targeted insect species (but not in mammals) that then result in pore formation and lysis of gut cells, which ultimately causes the insects to die [1–4]. Genetic engineering technology allows the application of Bacillus thuringiensis (Bt) crystalline (Cry) protein genes to crops (e.g., rice, maize, cotton, soy) to protect agricultural plants from insect infection. These crops are usually called transgenic Bt crops [5].

BT799 is a genetically modified Bt-containing maize currently under development in China, produced by insertion of the Cry1Ac gene into the parental maize Zhen58. Expression of the Cry1Ac
protein in BT799 provides farmers with an alternative way to decrease yield losses and reduce the use of chemical insecticides. However, there are still concerns about potential unintended effects following the insertion of exogenous gene in crops, especially in staple foods (e.g., rice, maize). In 2009, China’s Ministry of Agriculture released biosecurity certificates for two lines of transgenic Bt crops. This event revealed the commercialization of transgenic crops in China and put this topic on the public agenda [6,7]. In fact, heated disputes about the safety of transgenic crops has not stopped in China since then.

GM crops present challenges concerning food safety assessment because some scientists hold the opinion that unintended effects of transgenic crops due to the insertion of foreign genes are still possible [8–10]. The concept of substantial equivalence was proposed as the starting point for safety evaluation of GM crops [11,12]. A GM crop and its non-GM traditional version that has long been consumed in the human diet should be compared with each other in terms of their composition and agronomic performance. Several international organizations also have recommended subchronic rodent feeding studies to investigate any long-term effects of transgenic foods obtained from GM crops [13,14]. A large number of 90-day feeding studies to determine whether unintended adverse effects of GM crops will occur by evaluation of standard toxicology response variables have thus been reported. To date almost no harmful effects have been observed in these studies focusing on GM crops, including maize [15–19], soybean [20,21] and rice [22–25]. Despite the number of current studies on the safety of GM foods, there are still overwhelming doubts about the immature toxicological assessment procedures used for GM foods. Indeed, some studies have reported obviously harmful effects of GM foods. In the study led by Vecchio pregnant Swiss mice and male litters were fed on a standard diet containing 14% GM soybean. By means of immunoelectron microscopy at 2, 5 or 8 months of age, they focused on Sertoli cells, spermatogonia and spermatocytes. The results showed that GM-fed mice of all ages had larger number of perichromatin granules and lower nuclear pore density [26]. Meanwhile, enlargements of smooth endoplasmic reticulum was also observed in Sertoli cells of GM-fed mice. A three-generation study reported minimal histopathological changes in liver and kidney in F3 female offspring of rats fed a Bt maize [27]. Based on some of studies, Dona and Arvanitoyannis concluded that GM foods appeared to have toxic effect on the reproductive system, and might alter the normal hematological, biochemical, and immunologic parameters [10].

A multifaceted process that involves the combined action of testes, epididymides, accessory sex glands and associated hormones facilitates male reproduction [28]. The hypothalamic-pituitary-testicular axis strictly regulates the whole reproductive process, including the functions of testosterone (T), follicle-stimulating hormone (FSH) and luteinizing hormone (LH). The male reproductive system is usually considered at risk during the fetal development period, during puberty and even over the whole life span due to the sensitivity of the testes and accessory organs. Due to the high rate of cellular proliferation and distinctive differentiation in mammalian testis, molecular and cellular changes can be detected by this very sensitive organ when exposed to toxicants [28,29]. Thus, many scientists and research groups are concerned about possible negative effects that the GM food may exert on the male reproductive system and have conducted a number of studies in order to ensure the safety of GM foods by various methods using different species of animals [30]. Considering sperm maturation needs about 70 days in rats [31,32], a 90-day feeding period would put the male reproductive system at risk if unintended effects happen after exposure to transgenic crops, so studies of that length are often used.

In recent years, concerns about the dietary effects of GM crops on humans and animals have been raised. However, the previous studies cannot respond unequivocally to the public’s doubts or determine unintended effects on the male reproductive system. Considering maize is a widely consumed food in China and a serious debate on the risk assessment of transgenic crops continues, 90-day feeding studies are very necessary to evaluate the male reproductive toxicity of BT799 maize. In this study, we therefore aimed to perform such a study and investigate the toxicological
effects of BT799 maize on the growth performance and reproductive parameters of male Wistar rats. The present study was conducted in accordance to the domestic OECD 408 guidelines for Good Laboratory Practices [33].

2. Experimental Section

2.1. Plant Materials

Transgenic maize (BT799) and its none-transgenic near isoline control maize (Zhen58) were cultivated in the experimental field of China Agricultural University (Beijing, China) and provided by Chinese Center for Disease Control and Prevention (Beijing, China). Seeds of BT799 and Zhen58 were dried, hulled and stored in dry conditions for experimental use after harvesting.

2.2. Diet Formulation and Compositional Analysis of Maize

Samples for compositional analysis were randomly selected from both types of maize flour crushed from Zhen 58 and BT 799. Protein, fiber, fat, moisture and ash were determined in accordance with Chinese Standard methods [34–39]. Carbohydrate content was calculated using the following expression as described by Han et al. [40]:

\[
\text{Carbohydrate \%} = 100 - (\% \text{ protein} + \% \text{fat} + \% \text{ash} + \% \text{moisture})
\]

Maize flours from BT799 and Zhen58 were respectively added at 84.7% by mass to produce rodent diets. Carbohydrates in the BT799 formula were completely derived from transgenic maize. Casein was added to the feeds in order to make the protein content equivalent to the AIN-93G formula as the amount of protein provided by maize can’t meet the formulation requirements. Other macronutrients in the formulated feeds were added following the same principle as protein when the nutrient content from transgenic maize or non-transgenic maize was lower than in the standard AIN-93G formula. An additional AIN-93G diet was included as negative control. All diets were produced by Hua Fu Kang Feed Co. Ltd. (Beijing, China), following AIN-93G guidelines [41]. Diets formulation details are summarized in Table 1, and the nutritional composition of diets in the different groups is presented in Table 2.

Table 1. Composition of diets for rats.

| Ingredient (%) | AIN-93G | Zhen58 | BT799 |
|----------------|---------|--------|-------|
| Maize          | 0.00    | 84.68  | 84.68 |
| Casein         | 20.00   | 11.82  | 12.54 |
| Cane sugar     | 10.00   | 0.00   | 0.00  |
| Corn starch    | 52.95   | 0.00   | 0.00  |
| Soybean oil    | 7.00    | 5.12   | 4.07  |
| Cellulose      | 5.00    | 2.56   | 2.67  |
| Minerals       | 3.50    | 3.50   | 3.50  |
| Vitamins       | 1.00    | 1.00   | 1.00  |
| Methionine     | 0.18    | 0.18   | 0.18  |
| Choline chloride | 0.17 | 0.17   | 0.17  |

Table 2. Nutritional composition of diets.

| Components (%) | AIN93G | Zhen58 | BT799 |
|----------------|--------|--------|-------|
| Crude Protein  | 20     | 20     | 20    |
| Crude Fat      | 7.0    | 7.0    | 7.0   |
| Carbohydrate   | 62.95  | 63.99  | 62.95 |
| Cellulose      | 5.0    | 5.0    | 5.0   |
| Energy, kcal/g | 3.77   | 3.81   | 3.77  |
2.3. Animals and Housing

A total of 30 male Wistar rats (n = 10/group) purchased from Experimental Animal Division of Peking University Health Sciences Center (Beijing, China) were 4 weeks of age at the start of the study and housed in the specific pathogen-free animal facility in a climate controlled room with a 12-h on/off light cycle. This study was approved by the Medical Ethics Committee and performed in accordance with the Guidelines for Animal Experiments of the Peking University Health Science Center.

2.4. Experimental Design

Animals were randomly assigned into three groups according to body weight, fed on the respective diets and provided tap water ad libitum. Each group included 10 rats and mean body weight in each group at the beginning of the study did not exceed more than 20% variation. Formulated diets containing either 84.7% BT799 or Zhen58 was provided to the animals in two of the experimental groups. The remaining 10 rats were fed with standard AIN-93G diet as negative control. During the experimental period all animals were clinically observed twice daily for mortality, signs of morbidity or any other noteworthy signs. Blood samples for hematology examination were collected from the tail vein. Samples for serum chemistry and serum hormone evaluation were collected from the abdominal aorta under sodium pentobarbital anesthesia and the animals were fasted overnight to minimize fluctuations of the measured parameters. At the end of the study, animals were sacrificed under sodium pentobarbital anesthesia.

2.5. Body Weight Gain and Food Utilization

Body weights were obtained once a week and feed intakes were also measured weekly. Mean feed utilization was determined according to the following calculation:

\[
\text{Mean weekly feed utilization (\%) = \frac{\text{weekly body weight gain}}{\text{weekly feed consumption}} \times 100\%}
\]

2.6. Hematology

Red blood cell count (RBC), white blood corpuscle (WBC), hemoglobin (HGB), hematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), blood platelet count (PLT), red cell distribution width (RDW), were measured by a MEK-7222K cell counter (Nihon Kohden, Tokyo, Japan).

2.7. Serum Chemistry

Serum total protein (TP), alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), and concentrations of albumin (ALB), cholesterol (CHOL), creatine (CREA), Glucose (GLUC), blood urea nitrogen (BUN), triglycerides (TG) and lactate dehydrogenase (LDH) were measured by using an Olympus AU400 Clinical Chemistry Analyzer (Olympus, Shizuoka, Japan).

2.8. Serum Sex Hormone Levels

Rats in this study were fasted for 12 h and blood samples were collected from the abdominal aorta under anesthesia. Samples were centrifuged at 3000 rpm for 10 min at the temperature of 4 °C to separate serum. Serum level of follicle-stimulating hormone (FSH), luteinizing hormone (LH) and testosterone (T) were measured by a radioimmunoassay (RIA) method using hormone specific kits purchased from the Beijing North Institute of Biological Technology (Beijing, China).
2.9. Sperm Motility and Count

Right cauda epididymides of all animals were quickly collected after rats were sacrificed according to Adedara and Farombi [42]. Then epididymides were cut with surgical blades into pieces and incubated in 5 mL pre-warmed DMEM at 37 °C for 10 min to permit the release of sperm. The sperm suspensions for evaluation of sperm motility, head counts and morphology were prepared following the procedure described by Tannenbaum et al., [43]. One drop of the sperm suspension was placed on a sterile clean microscope slide. At least 10 microscopic fields were observed at 400 × magnification using a phase-contrast microscope (Eclipse E200, Nikon, Tokyo, Japan) and the percentage of motile sperm was recorded [44]. The sperm suspension for sperm head count (0.5 mL) was diluted with physiological saline (9.5 mL), then diluted sperm suspension (10 µL) was transferred to each counting chamber and counted with a hemocytometer. Data were expressed as sperm/mL.

2.10. Sperm Morphology

The sperm suspension was smeared out on histological slides and stained with 1% Eosin Y for morphologic examination. A total of two hundred sperm from each animal were used to determine morphological abnormalities [44–46].

2.11. Organ Weights and Histopathology

A complete gross necropsy was conducted by visual inspection on all the animals following sacrifice. Brain, heart, liver, lung, spleen, kidney, testes weight were immediately examined. Relative weight of each organ (or paired organ weights) was calculated along with final individual body weights. Tissue sections from liver, kidney, intestine, testes and were fixed with 4% buffered formalin and embedded in paraffin, then sectioned to 5 µm and stained with hematoxylin and eosin. Histopathological examination of tissue sections was conducted at the Beijing Key Laboratory of Toxicological Research and Risk Assessment for Food Safety (Beijing, China).

2.12. Statistical Analysis

Statistical comparisons were designed to determine whether there were statistically significant differences attributable to consumption of BT799 compared to Zhen58 or the AIN-93G diet. Preliminary tests were applied to test homogeneity for all continuous data (Levenes’s test; and Shapiro-Wilks test). If both of the preliminary tests were not significant, then a one-way ANOVA was conducted. Otherwise, a least squared differences model or a Dennett’s multiple comparison should be conducted to determine whether there were significant differences between groups. Response variables from BT799 and Zhen58 groups were first compared individually with those of the control group, and then the values from the BT799 group were compared with those in the Zhen58 group. A p-value of less than 0.05 was considered significant in all analyses.

3. Results

3.1. Compositional Analysis

Nutritional composition analysis results are listed in Table 3. Contents of macro-nutrients (protein, fat, carbohydrate and dietary fiber) in BT799 maize are similar to that of Zhen58 maize according to the test results and within the normal ranges reported for maize [47,48].
3. Results

3.1. Compositional Analysis

Nutritional composition analysis results are listed in Table 3. Contents of macro-nutrients (protein, fat, carbohydrate and dietary fiber) in BT799 maize are similar to that of Zhen58 maize according to Table 3.

| Nutrients               | Zhen58      | BT799      |
|-------------------------|-------------|------------|
| Protein                 | 9.71 ± 0.15 | 8.96 ± 0.17|
| Fat                     | 2.72 ± 0.49 | 3.57 ± 0.36|
| Moisture                | 9.16 ± 0.22 | 9.55 ± 0.32|
| Ash                     | 1.35 ± 0.35 | 1.31 ± 0.04|
| Total dietary fiber     | 3.02 ± 0.22 | 2.78 ± 0.01|
| Carbohydrate            | 73.27 ± 1.21| 74.72 ± 2.00|

Values are presented as mean ± SD (n = 3).

3.2. Body Weight Gain and Food Consumption

During the feeding trial, there were no significant differences in weekly body weights (Figure 1) and mean food consumption (Figure 2) between the different groups.

Figure 1. Mean body weights. Values are presented as mean ± SD, analysed by ANOVA, and followed by least-significant difference or Dennett’s multiple comparisons for post-hoc comparison between multiple groups.

Figure 2. Mean feed consumption. Values are presented as mean ± SD, analysed by ANOVA followed by least-significant difference or Dennett’s multiple comparisons for post-hoc comparison between multiple groups.

3.3. Hematology

No significant differences (p > 0.05) observed in the hematology response variables among any of the groups after consuming corresponding formulating diets for 13 weeks (Table 4).

Table 4. Nutritional composition of two kinds of maize flour in g/100g.

| Nutrients       | Zhen58       | BT799       |
|-----------------|--------------|-------------|
| Moisture        | 9.16 ± 0.22  | 9.55 ± 0.32 |
| Fat             | 2.72 ± 0.49  | 3.57 ± 0.36 |
| Carbohydrate    | 73.27 ± 1.21 | 74.72 ± 2.00|
| Ash             | 1.35 ± 0.35  | 1.31 ± 0.04 |
| Total dietary fiber | 3.02 ± 0.22 | 2.78 ± 0.01 |

Values are presented as mean ± SD (n = 3).
Table 4. Hematology and coagulation.

| Items          | BT799      | Zhen58     | AIN-93G    |
|----------------|------------|------------|------------|
| WBC (×10⁹/µL) | 9.58 ± 2.13 | 8.62 ± 1.55 | 8.42 ± 2.47 |
| RBC (×10⁹/µL) | 1.9 ± 0.54  | 8.65 ± 0.45 | 8.04 ± 0.85 |
| HGB (g/L)     | 184.13 ± 8.87 | 167.30 ± 9.01 | 151.11 ± 23.09 |
| HCT (%)       | 47.86 ± 2.72 | 47.22 ± 3.22 | 42.21 ± 4.14 |
| MCV (FL)      | 18.19 ± 0.63 | 19.35 ± 1.05 | 18.79 ± 2.26 |
| MCH (pg)      | 343.12 ± 6.32 | 355.10 ± 14.89 | 357.35 ± 36.99 |
| RDW (%)       | 11.92 ± 0.38 | 11.97 ± 0.35 | 12.09 ± 0.27 |
| PLT (×10³/µL) | 1018.38 ± 28.04 | 930.10 ± 70.09 | 1034.00 ± 33.87 |

Values are presented as mean ± SD, analysed by ANOVA followed by least-significant difference or Dennett’s multiple comparisons for post-hoc comparison between multiple groups.

3.4. Serum Chemistry

There were no statistically significant changes (p > 0.05) observed in any serum chemistry response variables among different groups (Table 5).

Table 5. Serum chemistry.

| Items          | BT799      | Zhen58     | AIN-93G    |
|----------------|------------|------------|------------|
| TP (g/L)      | 74.81 ± 2.60 | 77.39 ± 4.38 | 71.85 ± 3.27 |
| ALB (g/L)     | 37.92 ± 1.10 | 39.14 ± 1.76 | 36.78 ± 1.06 |
| GLB (g/L)     | 36.89 ± 1.71 | 38.25 ± 2.74 | 35.07 ± 2.45 |
| A/G           | 1.03 ± 0.03 | 1.02 ± 0.04 | 1.05 ± 0.06 |
| ALT (U/L)     | 48.00 ± 14.41 | 47.70 ± 12.79 | 49.30 ± 8.65 |
| ALP (U/L)     | 157.74 ± 19.28 | 121.28 ± 36.88 | 113.19 ± 23.64 |
| BUN (mmol/L)  | 6.15 ± 0.87  | 6.82 ± 1.02  | 7.71 ± 0.96  |
| CHO (mmol/L)  | 2.19 ± 0.42  | 2.46 ± 0.50  | 2.44 ± 0.41  |
| TG (mmol/L)   | 1.17 ± 0.33  | 2.11 ± 1.58  | 1.64 ± 1.37  |
| GLU (mmol/L)  | 8.16 ± 1.88  | 6.99 ± 0.74  | 6.23 ± 0.39  |
| AST (U/L)     | 158.90 ± 87.22 | 144.90 ± 55.28 | 139.80 ± 21.60 |
| LDH (U/L)     | 1294.56 ± 386.24 | 1095.81 ± 320.08 | 1412.06 ± 580.79 |

Values are presented as mean ± SD, analysed by ANOVA, followed by least-significant difference or Dennett’s multiple comparisons for post-hoc comparison between multiple groups.

# p < 0.05 compared with Zhen58 group.

3.5. Serum Sex Hormone

Serum levels of FSH and T were not significantly different among the test groups (Table 6). LH value of BT799 group is higher than that of the Zhen58 group with a significant difference (p = 0.013 < 0.05). However, no significant differences were observed between the BT799 group and AIN-93G group.

Table 6. Serum hormones concentration.

| Items          | BT799      | Zhen58     | AIN-93G    |
|----------------|------------|------------|------------|
| FSH (UI/L)     | 4.84 ± 0.53 | 4.97 ± 0.29 | 5.19 ± 0.31 |
| LH (ng/L)      | 21.45 ± 1.68* | 19.53 ± 1.20 | 20.03 ± 1.45 |
| T (nmol/L)     | 87.90 ± 5.35 | 83.36 ± 2.74 | 92.70 ± 2.99 |

Values are presented as mean ± SD, analysed by ANOVA followed by least-significant difference or Dennett’s multiple comparisons for post-hoc comparison between multiple groups. * p < 0.05 compared with Zhen58 group.

3.6. Sperm Motility, Sperm Head Counts and Morphology

After the 90-day feeding trial, sperm parameter test results of the three groups exhibited no significant differences (p > 0.05), including sperm motility, sperm head counts and morphology of epididymides sperm (Table 7).
Table 7. Sperm parameters.

| Items                  | BT799        | Zhen58       | AIN-93G      |
|------------------------|--------------|--------------|--------------|
| Sperm heads (×10⁶/mL)  | 32.35 ± 1.79 | 30.50 ± 2.62 | 31.69 ± 2.33 |
| Sperm motility (%)     | 87.84 ± 4.80 | 85.91 ± 6.02 | 84.06 ± 5.90 |
| Sperm abnormalities (%)| 9.44 ± 0.98  | 9.00 ± 0.93  | 9.38 ± 0.69  |

Values are presented as mean ± SD (n = 8), analysed by ANOVA followed by least-significant difference or Dennett’s multiple comparisons for post-hoc comparison between multiple groups.

3.7. Organ Weights and Pathology

There were no statistically significant differences (p > 0.05) in the relative organ weights among any test groups (Table 8).

Table 8. Organ/body weight.

| Organs      | BT799        | Zhen58       | AIN-93G      |
|-------------|--------------|--------------|--------------|
| Adrenals    | 0.013 ± 0.004| 0.013 ± 0.004| 0.010 ± 0.005|
| Brain       | 0.328 ± 0.031| 0.318 ± 0.025| 0.320 ± 0.040|
| Epididymides| 0.220 ± 0.043| 0.209 ± 0.028| 0.217 ± 0.035|
| Heart       | 0.242 ± 0.046| 0.250 ± 0.040| 0.237 ± 0.027|
| Kidneys     | 0.471 ± 0.079| 0.506 ± 0.087| 0.458 ± 0.059|
| Liver       | 2.440 ± 0.391| 2.421 ± 0.358| 2.155 ± 0.279|
| Spleen      | 0.148 ± 0.037| 0.147 ± 0.029| 0.137 ± 0.022|
| Testes      | 0.574 ± 0.096| 0.355 ± 0.081| 0.579 ± 0.036|
| Prostate    | 0.300 ± 0.096| 0.290 ± 0.089| 0.299 ± 0.092|

Values are presented as mean ± SD, analysed by ANOVA followed by least-significant difference or Dennett’s multiple comparisons for post-hoc comparison between multiple groups.

No evidence of altered incidence or severity of pathologic changes was observed in the organs of rats with different diet treatments. Gross and microscopic observations indicated no diet-related changes in the tissues of heart, brain, liver (Figure 3a–c), spleen, kidneys (Figure 3g–i), seminal vesicle, prostate, epididymides and testes (Figure 3d–f) in BT799, Zhen58 and AIN-93G control group.

![Figure 3. Photomicrographs of liver (a–c); kidney (d–f) and testis (g–i) tissues of male rats stained with H&E × 200. The rows (a–g), (b–h) and (c–i) were tissues from the AIN-93G control group, Zhen58 group and BT799 group, respectively.](image-url)
4. Discussion

Guidelines for the safety assessment of genetically modified crops and foods have been developed to assess whether they are “as safe as” conventional non-GM crops [11,12,49–52]. According to the principle of “substantial equivalence”, the compositional analyses of nutrients in GM crops and comparison between GM crops and conventional non-GM crops are a necessary step for the safety assessment. Rodent feeding studies are also recommended to assess any unintended side effects of transgenic crops [13,14].

In this study, nutritional composition results of transgenic maize and non-transgenic maize indicated that the concentrations of primary nutrients in BT799 were consistent with its near-isogenic maize strain. These results are also consistent with previous studies on GM maize with inserted transgenic genes conducted by other scientists [15,17]. Based on nutrient compositional analysis, transgenic maize flours were added into the diet formulation as the only carbohydrate source and the final BT799 formulation had equal nutritional composition to AIN-93G. A non-transgenic maize diet was formulated under the same equal nutritional principle and it had the same concentration of maize flour (84.7%) in the formulation as BT799 diets. Thus the diet formulations in the different groups had similar energy contents and the basic nutrients demands were also fulfilled.

Most reported subchronic feeding trials have focused on the comparison of nutritional performance between transgenic crops and non-transgenic crops in rodents using diverse responses such as body weight gain, feeding consumption, hematology, serum chemistry. Although a few studies have suggested the consumption of GM crops led to changes in the reproductive parameters of animals, scientific literature concerning the potential effects of GM crops on specific organs or systems are still limited. As a transgenic insect-resistant maize line cultivated in China, BT799 is undisputedly expected to resolve the restricted agricultural production of staple crops. However, it is of great importance to conduct well-designed safety assessments and toxicity studies before a transgenic crop is available on the market. At present, there are no related studies concerning the possible effects on the reproductive system of BT799 maize and the study presented herein definitely fills that gap.

In the present study, sex hormone levels and sperm parameters were given significant attention and examined as the most important indicators of male reproductive function health. The study results showed that a BT799 diet did not affect the nutritional and growth performance of rats. No evident histopathology changes or sperm parameter abnormalities were observed after consumption of BT799 maize. BT799 maize also exhibited similar effects on serum sex hormone levels as the control AIN-93G diet. A higher serum LH level in the BT799 group than the Zhen58 group can be explained as the result of spontaneous alterations (p = 0.036 < 0.05). Considering no significant difference was observed between BT799 group and AIN-93G group, the higher LH mean value is not associated with the exposure to transgenic maize.

Transgenic crops, such as rice and maize, are usually consumed by a large number of people worldwide as daily staple foods. Exposure to increased amounts of the expressed protein is quite possible after long-term and multigenerational feeding and this could lead to an adverse effect on the reproductive system if this new protein is a potential toxin [30]. Any long term health impacts after dietary intake of BT799 should be verified before putting this crop on the market. Thus, further safety assessment of BT799 should focus on multigenerational reproductive feeding studies and long-term feeding trials with dietary consumption of the transgenic crop.

5. Conclusions

In this study, we observed that the primary nutrient concentrations in BT799 were consistent with those of its near-isogenic maize strain. No significant differences in body weight, hematology and serum chemistry results were observed between rats fed a diet containing transgenic BT799, Zhen58 or the control in this 13-week feeding study. Results of serum hormone levels, sperm parameters and relative organ/body weights indicated no treatment-related side effect on the
reproductive system of male rats. In addition, no diet-related changes were found in necropsy and histopathology examinations. Based on results of the current study, we did not find any difference in the parameters tested in our study of the reproductive system of male rats between BT799 and Zhen58 or the control.

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References
1. Betz, F.S.; Hammond, B.G.; Fuchs, R.L. Safety and advantages of Bacillus thuringiensis protected plants to control insect pests. Regul. Toxicol. Pharmacol. 2000, 32, 156–173. [CrossRef] [PubMed]
2. Schnepf, E.; Crickmore, N.; van Rie, J.; Lereclus, D.; Baum, J.; Feitelson, J.; Zeigler, D.R.; Dean, D.H. Bacillus thuringiensis and its pesticidal crystal proteins. Mol. Biol. Rev. 1998, 62, 775–806.
3. IPCS (International Programme on Chemical Safety). Environmental Health Criteria 217: Microbial Pest Control Agent Bacillus Thuringiensis; World Health Organization: Geneva, the Switzerland, 1999.
4. McClintock, J.T.; Schaffer, C.R.; Sjoblad, R.D. A comparative review of the mammalian toxicity of Bacillus thuringiensis-based pesticides. Pestic. Sci. 1995, 45, 95–105. [CrossRef]
5. Carman, J.A.; Vlieger, H.R.; Ver Steeg, L.J.; Sneller, V.E.; Robinson, G.W.; Clinch-Jones, C.A.; Edwards, J.W. A long-term toxicology study on pigs fed a combined genetically modified (GM) soy and GM maize diet. J. Org. Syst. 2013, 8, 8–38.
6. Lu, C. The first approved transgenic rice in China. GM Crop. 2010, 1, 113–115. [CrossRef] [PubMed]
7. Manimaran, P.; Ramkumar, G.; Mangrauthia, S.K.; Padmakumari, A.P.; Muthuraman, P.; Bentur, J.S.; Viraktamath, B.C. Bt rice evaluation and deployment strategies. GM Crop. 2010, 2, 1–3. [CrossRef] [PubMed]
8. Janczyk1, P.; Wolf, C.; Hartmann, A.; Junghans, H.; Schwerin, M.; Souffrant, W.B. Modification of potatoes, by either recombinant DNA technology or conventional breeding, affects their nutritional value for the rat. Arch. Zootec. 2007, 10, 12–25.
9. Schroeder, M.; Poulsen, M.; Wiks, A.; Kroghsbo, S.; Miller, A.; Frenzel, T.; Danier, J.; Rychlik, M.; Emami, K.; Gatehouse, A.; et al. A 90-day safety study of genetically modified rice expressing Cry1Ab protein in Wistar rats. Food Chem. Toxicol. 2007, 45, 339–349. [CrossRef] [PubMed]
10. Dona, A.; Arvanitoyannis, I.S. Health Risks of Genetically Modified Foods. Crit. Rev. Food Sci. Nutr. 2009, 49, 164–175. [CrossRef] [PubMed]
11. Organization for Economic Cooperation and Development. Safety Evaluation of Foods Derived by Modern Biotechnology, Concepts and Principles; Organization for Economic Cooperation and Development: Paris, France, 1993.
12. FAO. Biotechnology and Food Safety; Report of a Joint FAO/WHO Consultation, Food and Nutrition Paper 61; FAO: Rome, Italy, 1996.
13. FAO/WHO. Safety Aspects of Genetically Modified Foods of Plant Origin; Report of a Joint FAO/WHO Expert Consultation on Foods Derived from Biotechnology; Food and Agriculture Organization of the United Nations: Rome, Italy, 2000.
14. EFSA. Safety and nutritional assessment of GM plants and derived food and feed: the role of animal feeding trials. Food Chem. Toxicol. 2008, 46, S2–S70.
15. He, X.Y.; Huang, K.L.; Li, X.; Qin, W.; Delaney, B.; Luo, Y.B. Comparison of grain from corn rootworm resistant transgenic DAS-59122-7 maize with non-transgenic maize grain in a 90-day feeding study in Sprague-Dawley rats. Food Chem. Toxicol. 2008, 46, 1994–2002. [CrossRef] [PubMed]
16. He, X.Y.; Tang, M.Z.; Li, X.; Cao, S.S.; Yu, J.J.; Delaney, B.; Huang, K.L. A 90-day toxicity study of transgenic lysine-rich maize grain (Y642) in Sprague-Dawley rats. *Food Chem. Toxicol.* 2009, 47, 425–432. [CrossRef] [PubMed]

17. MacKenzie, S.A.; Lamb, I.; Schmidt, J.; Deo, L.; Morrisey, M.J.; Harper, M.; Layton, R.J.; Prochaska, L.M.; Sanders, C.; Locke, M.; et al. Thirteen week feeding study with transgenic maize grain containing event DAS-Ø15Ø7-1 in Sprague-Dawley rats. *Food Chem. Toxicol.* 2009, 47, 425–432. [CrossRef] [PubMed]

18. Juberg, D.R.; Herman, R.A.; Thomas, J.; Brooks, K.J.; Delaney, B. Acute and repeated dose mouse oral toxicity studies with Cry34Ab1 and Cry35Ab1 Bt proteins used in coleopteran resistant DAS-59122-7 corn. *Regul. Toxicol. Pharmacol.* 2009, 54, 154–163. [CrossRef] [PubMed]

19. Appenzeller, L.M.; Hoban, D.; Sykes, G.P.; Malley, L.A.; Delaney, B. Subchronic feeding study with genetically modified stacked trait lepidopteran and coleopteran resistant (DAS-Ø15Ø7-1x DAS-59122-7) maize grain in Sprague-Dawley rats. *Food Chem. Toxicol.* 2009, 7, 1512–1520. [CrossRef] [PubMed]

20. Appenzeller, L.M.; Munley, S.M.; Hoban, D.; Sykes, G.P.; Sanders, C.; Delaney, B. Subchronic feeding study of herbicide-tolerant soybean DP-356Ø43-5 in Sprague-Dawley rats. *Food Chem. Toxicol.* 2008, 46, 2201–2213. [CrossRef] [PubMed]

21. Delaney, B.; Appenzeller, L.M.; Munley, S.M.; Hoban, D.; Sykes, G.P.; Sanders, C. Subchronic feeding study of high oleic acid soybeans (Event DP-3Ø5423-1) in Sprague-Dawley rats. *Food Chem. Toxicol.* 2008, 46, 3808–3817. [CrossRef] [PubMed]

22. Cao, S.S.; He, X.Y.; Xu, W.T.; Ran, W.J.; Liang, L.X.; Luo, Y.B.; Yuan, Y.F.; Zhang, N.; Zhou, X.; Huang, K.L. Metabonomics study of transgenic Bacillus thuringiensis rice (T2A-1) meal in a 90-day dietary toxicity study in rats. *Mol. Biosyst.* 2011, 7, 2304–2310. [CrossRef] [PubMed]

23. Tang, M.; Xie, T.; Cheng, W.; Qian, L.; Yang, S.; Yang, D.; Cui, W.; Li, K. A 90-day safety study of genetically modified rice expressing rhIGF-1 protein in C57BL/6J rats. *Transgenic Res.* 2012, 21, 499–510. [CrossRef] [PubMed]

24. Wang, Z.H.; Wang, Y.; Cui, H.R.; Xia, Y.W.; Altosaar, L.; Shu, Q.Y. Toxicological evaluation of transgenic rice flour with a synthetic cry1Ab gene from Bacillus thuringiensis. *J. Sci. Food Agric.* 2002, 82, 738–744. [CrossRef]

25. Zhou, X.H.; Dong, Y.; Xiao, X.; Wang, Y.; Xu, Y.; Xu, B.; Shi, W.D.; Zhang, Y.; Zhu, L.J.; Liu, L.Q. A 90-day toxicity study of high-amylose transgenic rice grain in Sprague-Dawley rats. *Food Chem. Toxicol.* 2011, 49, 3112–3118. [CrossRef] [PubMed]

26. Vecchio, L.; Cisterna, B.; Malatesta, M.; Martin, T.E.; Biggiogera, M. Ultrastructural analysis of testes from mice fed on genetically modified soybean. *Eur. J. Histochem.* 2004, 48, 449–454. [CrossRef]

27. Kilic, A.; Akay, M.T. A three-generation study with genetically modified Bt corn in rats: Biochemical and histopathological investigation. *Food Chem. Toxicol.* 2008, 46, 1164–1170. [CrossRef] [PubMed]

28. D'Cruz, S.C.; Vaithinathan, S.; Jubendradass, R.; Mathur, P.P. Effects of plants and plant products on the testis. *Asian J. Androl.* 2010, 12, 468–479. [CrossRef] [PubMed]

29. Evenson, D.P.; Darzynkiewicz, Z.; Melamed, M.R. Comparison of human and mouse sperm chromatin structure by flow cytometry. *Chromosoma* 1980, 78, 225–238. [CrossRef] [PubMed]

30. Zhang, W.; Shi, F. Do genetically modified crops affect animal reproduction? A review of the ongoing debate. *Animal* 2011, 5, 1048–1059. [CrossRef] [PubMed]

31. OECD. *OECD Guidelines for the Testing of Chemicals; No. 416: Two-Generation Reproduction Toxicity Study;* Organization for Economic Cooperation and Development: Paris, France, 2001.

32. Šimić, B.; Teuta Murati, I.K.; Kniewald, J. Effects of lindane on reproductive parameters in male rats. *Vet. Arh.* 2012, 82, 211–220.

33. OECD. *OECD Guidelines for Testing of Chemicals; No. 408: Repeated Dose 90-Day Oral Toxicity Study in Rodents;* OECD: Paris, France, 1998.

34. Detection of Genetically Modified Plants and Derived Products: Qualitative PCR Method for Insect-Resistant Rice TT51-1 and Its Derivates; *Ministry of Agriculture of the People’s Republic of China: Beijing, China, 2009.*

35. National Food Safety Standard: Determination of Protein in Foods; *Ministry of Agriculture of the People’s Republic of China: Beijing, China, 2010.*

36. Determination of Fat in Foods: Chinese Standard GB/T 5009.6; Ministry of Agriculture of the People’s Republic of China: Beijing, China, 2003.
37. National Food Safety Standard: Determination of Moisture in Foods: Chinese Standard GB 5009.3; Ministry of Agriculture of the People’s Republic of China: Beijing, China, 2010.

38. National Food Safety Standard: Determination of Ash in Foods: Chinese Standard GB 5009.4; Ministry of Agriculture of the People’s Republic of China: Beijing, China, 2010.

39. Determination of Dietary Fiber in Foods: Chinese Standard GB/T 5009.88; Ministry of Agriculture of the People’s Republic of China: Beijing, China, 2008.

40. Han, J.H.; Yang, Y.X.; Chen, S.Y.; Wang, Z.; Yang, X. L.; Wang, G.D.; Men, J.H. Comparision of nutrient composition of parental rice and rice genetically modified with cowpea trypsin inhibitor in China. J. Food Sci. 2005, 18, 297–302.

41. Reeves, P.G.; Nielsen, F.H.; Fahey, G.C. AIN-93 purified diets for laboratory rodents: Final report of the American Institute of Nutrition ad hoc writing committee on the reformulation of the AIN-76A rodent diet. J. Nutr. 1993, 123, 1939–1951. [PubMed]

42. Adedara, I.A.; Farombi, E.O. Chemoprotection of ethylene glycol monoethyl ether-induced reproductive toxicity in male rats by kolaviron, isolated biflavonoid from Garcinia kola seed. Hum. Exp. Toxicol. 2012, 31, 506–517. [CrossRef] [PubMed]

43. Tannenbaum, L.V.; Thran, B.H.; Williams, K.J. Testing the Limits of Rodent Sperm Analysis: Azoospermia in an Otherwise Healthy Wild Rodent Population. Arch. Environ. Contam. Toxicol. 2009, 56, 157–164. [CrossRef] [PubMed]

44. Terpsidis, K.I.; Papazahariadou, M.G.; Taitzoglou, L.A.; Papaioannou, N.G.; Georgiadis, M.P.; Theodoridis, I.T. Toxoplasma gondii: Reproductive parameters in experimentally infected male rats. Exp. Parasitol. 2009, 121, 238–241. [CrossRef] [PubMed]

45. Toledo, F.C.; Perobelli, J.E.; Pedrosa, F.P.; Anselmo-Franci, J.A.; Kempinas, W.D. In utero protein restriction causes growth delay and alters sperm parameters in adult male rats. Reprod. Biol. Endocrinol. 2011. [CrossRef] [PubMed]

46. Kato, M.; Makino, S.; Kimura, H.; Ota, T.; Furuhashi, T.; Nagamura, Y. Sperm motion analysis in rats treated with Adriamycin and its applicability to male reproductive toxicity studies. J. Toxicol. Sci. 2001, 26, 51–59. [CrossRef] [PubMed]

47. OECD. Consensus Document on the Compositional Considerations for New Varieties of Maize (Zea Mays): Key Food and Feed Nutrients, Antinutrients, and Secondary Plant Metabolites; Organization for Economic Cooperation and Development: Paris, France, 2002.

48. ILSI (International Life Sciences Institute). International Life Sciences Institute Crop Composition Database; International Life Sciences Institute: Washington DC, USA, 2006.

49. Codex. Joint FAO/WHO Food Standards Programme; Codex Alimentarius Commission: Rome, Italy, 2003.

50. OECD. OECD Documents: Report of the OECD Workshop on the Toxicological and Nutritional Testing of Novel Foods; Organization of Economic Cooperation and Development: Paris, France, 1997.

51. WHO. Strategies for Assessing the Safety of Foods Produced by Biotechnology; World Health Organization: Geneva, The Switzerland, 1991.

52. WHO. Application of the Principles of Substantial Equivalence to the Safety Evaluation of Foods and Food Components from Plants Derived from Modern Biotechnology; World Health Organization: Geneva, The Switzerland, 1995.

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