Megakaryocyte progenitor cell function is enhanced upon aging despite the functional decline of aged hematopoietic stem cells

Donna M. Poscablo,1,2 Atesh K. Worthington,1,2 Stephanie Smith-Berdan,1 and E. Camilla Forsberg1,3,*

1Institute for the Biology of Stem Cells, University of California-Santa Cruz, Santa Cruz, CA, USA
2Program in Biomedical Sciences and Engineering, Department of Molecular, Cell, and Developmental Biology, University of California-Santa Cruz, Santa Cruz, CA 95064, USA
3Biomolecular Engineering, University of California-Santa Cruz, Santa Cruz, CA, USA
*Correspondence: cforsber@ucsc.edu
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SUMMARY

Age-related morbidity is associated with a decline in hematopoietic stem cell (HSC) function, but the mechanisms of HSC aging remain unclear. We performed heterochronic HSC transplants followed by quantitative analysis of cell reconstitution. Although young HSCs outperformed old HSCs in young recipients, young HSCs unexpectedly failed to outcompete the old HSCs of aged recipients. Interestingly, despite substantial enrichment of megakaryocyte progenitors (MkPs) in old mice in situ and reported platelet (Plt) priming with age, transplanted old HSCs were deficient in reconstitution of all lineages, including MkPs and Plts. We therefore performed functional analysis of young and old MkPs. Surprisingly, old MkPs displayed unmistakably greater regenerative capacity compared with young MkPs. Transcriptome analysis revealed putative molecular regulators of old MkP expansion. Collectively, these data demonstrated that aging affects HSCs and megakaryopoiesis in fundamentally different ways: whereas old HSCs functionally decline, MkPs gain expansion capacity upon aging.

INTRODUCTION

The elderly population is affected by numerous hematological abnormalities, including decreasing capacity to mount an immune response, increasing incidences of thrombotic cardiovascular disorders, and dramatically increased incidence of myelogenous diseases. Dysfunctions during chronological aging is paralleled by the declining functionality of hematopoietic stem cells (HSCs), leading to conclusions that the age-related pathologies of the blood manifests within the HSC compartment (Elias et al., 2017). HSCs self-renew and differentiate, giving rise to progenitor cells that are committed to either the myeloid or lymphoid lineage throughout the lifespan. However, there is convincing evidence that HSCs lose the full extent of these properties during aging. Evaluation of aged HSCs in transplantation studies has demonstrated reduced regenerative potential compared with young HSCs. Aged HSCs seem to be less effective at homing and engrafting in the host, suggesting that alterations to HSCs during aging persist (become intrinsic) on transplantation (Dykstra et al., 2011; Morrison et al., 1996; Rossi et al., 2005; Sudo et al., 2000). In those experiments, HSC multipotency has been traditionally defined by qualitative measurements of GM, B, and T cell donor-to-host chimerism; more quantitative measurements and direct tracking of erythroid cells and platelets (Plts) from hematopoietic stem and progenitor cells (HSPCs) were developed relatively recently to rectify limitations of transplantation experiments that focus on only white blood cell reconstitution (Boyer et al., 2019; Hamanaoka et al., 2013; Schaefer et al., 2001; Yamamoto et al., 2013). In addition, analysis of HSCs at the single cell level has identified an increased molecular Plt priming and functional Plt bias in the old HSC compartment (Grover et al., 2016; Sanjuan-Pla et al., 2013; Yamamoto et al., 2018). These alterations raise the need to understand potential alterations in megakaryocyte differentiation in aged mice and humans.

The varying challenges to the blood system during aging may lead to alterations to multiple levels of differentiation, resulting in the dysregulation of homeostatic control maintained by committed progenitors. Previous studies have noted that the bone marrow (BM) frequencies of committed lymphoid and myeloid progenitors are altered with age (De Haan and Van Zant, 1999; Miller and Allman, 2003; Morrison et al., 1996; Rossi et al., 2005), but the functional properties and roles of hematopoietic progenitor cells during aging remains an enigma. Here, we investigated young and old HSCs and the contribution of both intrinsic changes and the environment on their differentiation. In our analysis, we included quantitative measurements of their erythroid and Plt production, as well as assessment of progenitor cells at steady-state and upon heterochronic HSC transplantation. These findings prompted us to further pursue age-associated changes in the Plt lineage, leading to unexpected revelations on the effect of aging on megakaryopoiesis.

RESULTS

Both the frequencies and numbers of hematopoietic stem and progenitor cell populations are altered during aging

Aging has been shown to alter the frequencies of HSCs in the BM (De Haan and Van Zant, 1999; Morrison et al.,
Aging hematopoietic system has been reported to include classically defined MkPs in old mice (Figures 1E and 1F). The frequency and number of MPPs remained consistent throughout life. The Myeloid Progenitor (MyPro) compartment increased in total cell numbers, but not frequency. Classically defined CMPs and GMPs (Akashi et al., 2000) were found at similar frequencies in young and old BM, while MEP frequencies were reduced with age (Figure 1B). Due to the overall increased cell numbers with age, however, absolute quantification revealed an increase in CMP and GMP numbers, but no change in MEPs, in old BM.

We also investigated erythromyeloid populations more recently defined by alternative markers within the MyPro compartment (Figure 1D) (Prönk et al., 2007). Few age-related alterations in frequencies and numbers were observed: CFU-E, pCFU-E, pGMP, and pMegE frequencies and numbers were not significantly altered (Figures 1E and 1F). Alternatively defined “GMPs” trended toward an increase in frequency and were significantly increased in absolute numbers (Figures 1E and 1F), consistent with the highly overlapping classical GMPs (Figures 1B and 1C). Intriguingly, we observed a dramatic increase in both the frequency and total cell numbers of phenotypically defined MkPs in old mice (Figures 1E and 1F). The aging hematopoietic system has been reported to include alterations to circulating mature cells (Valletta et al., 2020; Young et al., 2020). Consistent with previous reports, we also observed age-related increase in Plt frequencies and numbers, whereas erythroid cells decreased (Figures 1G and 1H). Our data also recapitulated reported myeloid bias among nucleated cells, primarily driven by a decrease in B and T cells (Figures 1G and 1H). These data revealed that the composition of HSPC populations selectively changes during aging. Of note was the dramatic expansion of old HSCs and old MkPs, accompanied by a significant increase in circulating Plts, raising the need to explore megakaryopoiesis during the aging process.

Old HSCs exhibit impaired reconstitution potential compared with young HSCs

To directly interrogate the differences in reconstitution potential between young and old HSCs, we purified HSCs from fluorescent young or old mice and transplanted them into young recipients (Figure 2A). These experiments are analogous to those performed previously by others (Erigen et al., 2012; Rossi et al., 2005), except we expanded aforementioned investigations by also including direct detection of donor-derived Plts and circulating erythroid cells, in addition to the more commonly assayed granulocytes/myelomonocytes (GMs), and B and T cells.

First, also as commonly done, we determined the reconstitution kinetics of HSCs by measuring donor chimerism in the peripheral blood (PB) for the duration of 18 weeks posttransplantation (Figure 2B). Consistent with previous reports, analysis of lineage potential between young and old HSCs revealed that old HSCs have a diminished capacity to reconstitute GM, B, and T cells. Similarly, erythroid and Plt chimerism were also lower from old versus young HSCs (Figure 2B).
Second, we removed the host cell variable by quantifying the absolute numbers of donor-derived mature cells in the PB of recipient mice (Boyer et al., 2019; Cool et al., 2020; Rajendiran et al., 2020a, 2020b) to more quantitatively assess the differences in regenerative activity between young and old HSCs. Old HSCs generated fewer erythroid, GM, B, T, and Plt cells compared with young HSCs (Figure 2C); this observation is consistent with host cell production presented as donor-derived chimerism (Figure 2B). Our data demonstrated a pan-hematopoietic reconstitution deficit of HSCs during aging, which is consistent with previous reports in white blood cell reconstitution. Taken together, these results also support the notion that key changes intrinsically maintained by HSCs drive their feeble performance upon aging.

The old bone marrow environment impairs the reconstitution potential of HSCs

We next sought to determine how cues in the aging environment influence HSC reconstitution capacity. Because
young HSCs outperformed old HSCs in young hosts (Figure 2), we reasoned that young HSCs would engraft with higher efficiency in old compared with young recipients. To test this directly, we purified HSCs from young mice and transplanted them into young or old recipient mice (Figure 3A). We then analyzed mature cell production of transplanted HSCs to quantitatively assess the role of host age on expansion capacity and lineage potential. Young HSCs were capable of long-term, multilineage reconstitution (LTMR) in both young and old hosts. However, in contrast to the young hosts, the old hosts allowed for significantly lower reconstitution chimerism of HSC-derived GM, B, T, erythroid, and Plt cells (Figure 3B). Overall, the differences were more significant at earlier timepoints (~4–14 weeks) than at later timepoints.

In the same cohorts, we also determined the absolute number of donor-derived mature cells in the PB after transplantation. The donor HSCs in the young host produced greater numbers of donor-derived cells per microliter of PB compared with the old host (Figure 3C). Although this pattern was overall consistent with the patterns observed by donor chimerism, the differences in total cell

**Figure 3. Young HSCs exhibited lower reconstitution potential in old compared with young recipients**

(A) Schematic of transplantation of HSCs from young mice into separate cohorts of young or old mice.

(B) Old recipients displayed lower HSCs donor-host-chimerism compared with young recipients. Analysis of donor-derived mature cells (GMs, B cells, T cells, erythroid cells, and platelets) in PB of recipients presented as percent donor chimerism.

(C) Donor HSCs produced fewer total mature cells in old recipients compared to in young recipients. Reconstitution data from (B) re-plotted as absolute number of donor-derived GMs, B cells, T cells, erythroid cells, and platelets per microliter of PB.

YY, Young HSCs transplanted into young recipients; YO, Young HSCs transplanted into old recipients. Data are representative means ± SEM from nine young recipient mice (four independent experiments) and 10 old recipient mice (three independent experiments). p values were determined using unpaired two-tailed t test. *p < 0.05, **p < 0.005, ***p < 0.0005.
production was attenuated and, with the exception of T cells, was not sustained over time. This analysis revealed that the old host reduced donor chimerism by young HSCs. However, when we removed the host variable by measuring absolute mature cell production, our analysis revealed a less different reconstitution capacity between young HSCs placed in the young environment and the young HSCs in the old environment.

Because the isochronic Y-into-Y transplants led to higher chimerism than the two heterochronic (O-into-Y or Y-into-O) transplants (Figures 2 and 3), we hypothesized that an old environment would provide a more supportive environment for old HSCs. We therefore compared the reconstitution potential of young and old HSCs in an old host (Figure 4A) and assayed for donor chimerism (Figure 4B) and absolute number of donor-derived cells (Figure 4C). Surprisingly, transplantation of young HSCs led to higher reconstitution of old recipients compared with transplantation of old HSCs. We also examined the impact of reconstitution by old HSCs in a young or old environment; our analysis revealed that old HSCs exhibited poor reconstitution capacity in the old host compared with the young host (Figure S1). Therefore, while the aged BM environment impaired the reconstitution potential of transplanted young HSCs (Figure 3), HSCs from old mice performed even worse than young HSCs in an aged environment.
Figure 5. Both old HSCs and the old niche impaired reconstitution of the stem and progenitor cell compartment

(A) Schematic of HSC transplantation from young or old mice into young mice.

(A') The total number of BM cells was similar in young recipient mice >16 weeks after transplantation of either young or old HSCs.

(A'') Old HSCs contributed to significantly lower donor-to-host chimerism of BM stem and progenitor cells in young recipient mice compared with young HSCs. Percent donor chimerism of HSPCs and erythromyeloid progenitors in the recipient BM of young or old HSCs >16 weeks posttransplant.

(A'''') Old HSCs generated fewer total stem and progenitor cells than young HSCs in young recipient mice. Quantification of total donor-derived classical HSPCs and erythromyeloid progenitors in the BM of recipients >16 weeks posttransplant.

(B) Schematic of HSC transplantation from young mice into separate cohorts of young or old mice.

(B') The total number of BM cells was similar in young and old recipient mice >18 weeks after transplantation of young HSCs.

(B'') Old recipients displayed lower donor-to-host HSPC chimerism than young recipient mice transplanted with young HSCs. BM analysis of young or old mice transplanted with young HSCs. HSPCs were analyzed >18 weeks posttransplant and presented as donor chimerism.
Both old HSCs and the old niche impairs reconstitution of the stem and progenitor cell compartment

The reported Plt bias of old HSCs (Grover et al., 2016; Haas et al., 2015; Sanjuan-Pla et al., 2013; Yamamoto et al., 2013, 2018) had the potential to translate into maintained, or even increased, Plt reconstitution by old HSCs, possibly at the expense of the other lineages. However, Plt reconstitution by old HSCs in either young (Figure 2) or old (Figure 4) hosts, and by young HSCs in old hosts (Figure 3) was as impaired as reconstitution of erythroid cells and WBCs. To determine whether the observed reconstitution of the PB is reflected in the BM of the recipients, we also determined the capacity of young and old HSCs to regenerate stem and progenitor cells. In the cohorts analyzed for PB reconstitution in Figures 2 and 3, we determined donor chimerism in the BM stem and progenitor cell compartment >18 weeks posttransplantation (Figure 5A). All expected populations were detected, supporting the LTMR capability of HSCs. Quantitatively, young recipients of the expected populations were detected, supporting the LTMR capability of HSCs. Quantitatively, young recipients of the BM stem and progenitor cells. In the cohorts analyzed for PB reconstitution in Figures 2 and 3, we determined donor chimerism in the BM stem and progenitor cell compartment >18 weeks posttransplantation (Figure 5A). All expected populations were detected, supporting the LTMR capability of HSCs. Quantitatively, young recipients of the BM stem and progenitor cells. In the cohorts analyzed for PB reconstitution in Figures 2 and 3, we determined donor chimerism in the BM stem and progenitor cell compartment >18 weeks posttransplantation (Figure 5A). All expected populations were detected, supporting the LTMR capability of HSCs. Quantitatively, young recipients

Old MkPs were functionally enhanced compared with young MkPs

The most profound and consistent change in old mice was alterations in megakaryopoiesis, denoted by a dramatic increase in MkPs and Plts (Figure 1). However, this age-dependent increase in MkPs was not accompanied by an age-dependent functional bias of HSCs toward the megakaryocyte lineage: transplanted old HSCs were as deficient in MkP (Figures 5A″, 5B″, 5A‴, 5B‴, 6A″, 6B″, 6A‴″, and 6B‴″) and Plt (Figures 2B, 2C, 4B, 4C, 5B, and 5C) reconstitution as they were in generating other lineages. Thus, we hypothesized that the dramatic increase in the frequency and total numbers of old MkPs in Plts (Figures 1D–1H) may be manifested by MkP-intrinsic changes, possibly in addition to increased contribution from the HSC compartment.

To begin to test this hypothesis, we determined functional differences between young and old MkPs in vitro. Interestingly, evaluation of MkP proliferative capacity by short-term culture assays revealed a significant, 2.5-fold increase in expansion of old MkPs compared with young MkPs (Figures 7A and 7B). These experiments showed that old MkPs displayed a surprising increased functional capacity compared with young MkPs in vitro.

Old MkPs possessed greater platelet reconstitution potential compared with young MkPs

The notable increase in expansion capacity of old MkPs in vitro led us to predict potential functional differences in vivo. To determine the reconstitution potential of old MkPs in vivo, we transplanted young or old MkPs into young recipient mice (Figure 7C). We evaluated MkP multilineage reconstitution in the host by measuring their contribution to mature cells (Plt, erythroid cells, GMs, B, and T) in the PB for 16 weeks. Given that young MkPs have been shown to be strongly committed to the Plt lineage in in vitro assays (Nakorn et al., 2003; Pronk et al., 2007), we predicted that MkPs would primarily produce Plts. Indeed, young MkPs contributed to nominal donor-chimerism levels of GM, B, and T cells, with a more robust contribution to Plts, as well as, unexpectedly, a short burst of erythroid cells (Figure 7D). Surprisingly, relative to young MkPs, old MkPs exhibited a remarkable capacity to engraft and contribute to Plts in the recipient mouse (Figure 7D). Although young MkPs repopulated a more modest 7.4% of Plts, the same number of old MkPs repopulated on average 34% of Plts in the recipient mice. Interestingly,

(B‴″) Transplanted young HSCs produced fewer total HSPCs in the BM of old compared with young recipient mice. Quantification of total donor-derived HSPC numbers in the BM of recipients >18 weeks posttransplant.

YY, Young HSCs transplanted into young recipients; OY, Old HSCs transplanted into young recipients; YO, Young HSCs transplanted into old recipients. Data are representative means ± SEM. PB analysis of the same recipient mice are presented in Figures 2A′ and 2A‴″ and Figure 3B′ and 3B‴″. P values were determined using unpaired two-tailed t test. *p ≤ 0.05, **p < 0.005, ***p < 0.0005, ****p < 0.0001.
Figure 6. Stem and progenitor cell reconstitution by old HSCs was impaired and exacerbated by the old niche

(A) Schematic of transplantation of HSCs from young or old mice into old mice.

(A') Quantification of total BM cells in recipient mice in (A) >16 weeks posttransplant.

(A'') Old HSCs contributed to significantly lower donor-to-host chimerism of BM stem and progenitor cells compared with young HSCs in an old host. Percent donor chimerism of HSPCs and erythromyeloid progenitors in the old recipient BM of transplanted young or old HSCs >16 weeks posttransplant.

(A'''') Old HSCs generated fewer total stem and progenitor cells than young HSCs in old recipient mice. Quantification of total donor-derived classical HSPCs and erythromyeloid progenitors in the BM of recipients >16 weeks posttransplant.

(B) Schematic of transplantation of HSCs from old mice into separate cohorts of young or old mice.

(B') Quantification of total BM cells in recipient mice in (B) >16 weeks posttransplant.

(B'''') Old recipients displayed lower donor-to-host HSPC chimerism than young recipient mice transplanted with old HSCs. BM analysis of young or old mice transplanted with old HSCs. HSPCs were analyzed >18 weeks posttransplant and presented as donor chimerism.

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old MkPs also contributed to significantly higher erythroid cell and GM chimerism in the recipient mice relative to young MkPs. It is also notable that unlike the long-term reconstitution capacity of HSCs (Figures 2, 3, 4, 5, and 6), the Plt, erythroid cell, and GM production from old MkPs was robust only in the first few weeks posttransplant and then declined over time.

These differences between young and old MkPs were reinforced by quantification of the absolute cell numbers from the same experiments (Figure 7E). The absolute quantification revealed that old MkPs engrafted and produced Plts, as well as erythroid cells and GMs, at a remarkably high efficiency compared with young MkPs. Taken together, our in vitro and in vivo results indicated that MkPs gain both expansion and reconstitution capacity during aging.

**RNA sequencing revealed different molecular patterns between young and old MkPs**

We were surprised by the remarkable difference in expansion and reconstitution capacity between young and old MkPs. To test how aging affects the molecular profile of MkPs, we evaluated the molecular regulation of old MkPs by performing RNA-sequencing (RNA-seq) analysis of young and old MkPs. We found that young and old MkPs had very similar gene expression profiles as expected of youthful and aging megakaryopoiesis. Importantly, our RNA-seq data revealed well-known genes implicated in the differentiation-specific (ETS) transcription factors (Banu et al., 2020; Jin et al., 2018; Levay and Slepak, 2007). We also observed differential expression of multiple genes associated with Plt regulation, production, and function: Coagulation Factor X, Dtnbp1, Gfi1b, Enpp4, Pafah1b3, Pafah1b1, Mmnn1, and Selpg. Mutations in Gfi1b and Dtnbp1 are associated with various Plt disorders and bleeding disorders (Ghiani and Dell’Angelica, 2011; Songdej and Rao, 2017). Interestingly, our analysis also revealed a small subset of upregulated genes previously implicated in acute myeloid leukemia (AML), including Pbx3, Lair1, and Mll3 (Kang et al., 2015; Li et al., 2013; Marschalek, 2010), and is possibly linked to increased risk of AML in the elderly (Almeida and Ramos, 2016).

The results of the RNA-seq analysis corroborates the behaviors observed in our functional analyses of MkPs. Together, these data illuminate the fundamental difference between youthful and aging megakaryopoiesis. Importantly, our RNA-seq data revealed well-known genes implicated in the molecular mechanisms of megakaryopoiesis during aging, as well as a number of novel genes that can be pursued to understand, and possibly mitigate, the age-related changes that occur in megakaryopoiesis.

**DISCUSSION**

**Aging alters the reconstitution potential of HSCs**

As previously reported by others, our characterization of the old BM populations revealed an expansion of HSCs, accompanied by alterations to descendant mature cells in the PB (Figure 1). These observations raised the need understand how HSC differentiation is modulated by the balance of both intrinsic changes to HSCs and extrinsic influence on HSCs during aging. Young HSCs have been suggested to have superior engraftment and reconstitution capacity compared with old HSCs (Dykstra et al., 2011; Morrison et al., 1996; Rossi et al., 2005; Sudo et al., 2000). Indeed,

(B^−) Transplanted old HSCs produced fewer total HSPCs in the BM of old compared with young recipient mice. Quantification of total donor-derived HSPC numbers in the BM of recipients >18 weeks posttransplant.

YO, Young HSCs transplanted into old recipients; OO, Old HSCs transplanted into old recipients; OY, Old HSCs transplanted into young recipients. Data are representative means ± SEM. PB analysis of the same recipient mice are presented in Figures 4A and 4A’ and 5B and 5B’. p values were determined using unpaired two-tailed t test. *p ≤ 0.05, **p < 0.005, ***p < 0.0005, ****p < 0.0001.
Figure 7. Old MkPs had enhanced reconstitution potential and altered gene expression signatures compared with young MkPs (A and B) Old MkPs have enhanced proliferative capacity compared to young MkPs in vitro. (A) Representative images of Y and O MkPs grown in vitro. To evaluate proliferative capacity, 2,000 MkPs were FACS sorted from young and old mice and grown in liquid culture. Three days after seeding, MkPs were microscopically evaluated for expansion efficiency. (B) MkP expansion in vitro. Flow cytometry was used to quantify the total fold expansion of cultured young and old MkPs from (A). n = 3 from three independent experiments. (C) Schematic of transplantation of young or old MkP into young mice.

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our experiments transplanting young or old HSCs indicated that old HSCs exhibited a diminished reconstitution capacity (Figures 2 and 5). To address whether the reduced reconstitution activity of HSCs during aging is influenced by the aged environment, we transplanted young HSCs into young or old hosts (Figures 3 and 5). Our data revealed that the old environment was less supportive of reconstitution by transplanted HSCs compared with the young environment. Therefore, transplanted young HSCs did not have an obvious competitive advantage over the resident old HSCs in the aged host. Notably, transplanted old HSCs had even lower capacity to reconstitute the old host compared with young HSCs (Figures 4 and 6), demonstrating that the poor performance of old HSCs in young recipients (Figures 2 and 5) was not due to the age-mismatch (heterochronic) between donor and host.

Because both young and old HSCs perform worse in old compared with young recipients, it is clear that age-related changes in the host impact donor HSC performance. This could be related to the reported increased proliferation of old HSCs (Kirschner et al., 2017), age-related changes to the distal and local microenvironment, or both. Our collective iso- and heterochronic transplantation data show that both HSC-intrinsic and extrinsic mechanisms during aging affect reconstitution efficiency (SanMiguel et al., 2020). Interestingly, we did not detect a selective retention of MKP or Plt reconstitution capacity of transplanted old HSCs, despite the increased frequencies and numbers of HSCs, MKPs, and Plts in unmanipulated aged mice (Figure 1). It remains possible that HSCs contribute to the greater production of MKPs in situ (Carrelha et al., 2018), with participation of the aging BM microenvironment (Ho et al., 2019). Together with the observed expansion of HSCs in situ (Figure 1), our transplantation results suggest that the impaired reconstitution performance of aged HSCs reflects reduced engraftment capacity rather than functional decline in situ. A better understanding of how young and old BM cells respond to niche-clearing methods could help to develop optimal systems for investigating reconstitution differences between young and old HSCs, and between young and old recipients.

**Surprising reconstitution and differentiation potential of old MkPs**

Our data provided evidence about the changes to the homeostatic control of blood cell production at the MKP level of hematopoietic differentiation during aging. Given the reconstitution deficit displayed by old HSCs (Figures 2, 4, 5A, and 5A”), one might have expected that old MKPs would also display functional deficiencies. We were surprised to find from our in vitro experiments that old MKPs displayed greater proliferative potential (Figures 7A and 7B). Strikingly, we also observed from transplanting young and old MKPs that old MKPs harbored a remarkable capacity to engraft, expand, and reconstitute Plts (Figures 7C–7E). This age-associated elevation of MKP potential is reminiscent of the higher prevalence of thrombosis and other cardiovascular problems in the elderly. We hypothesize that age-related alterations made to MKP populations contribute to dysregulation and increased risk for age-related morbidities (Davizon-Castillo et al., 2019).

Our transplantation studies of old and young MKPs also revealed somewhat surprising evidence about the differentiation programs of immature progenitors during aging (Figures 7C–7E). Previous reports suggested that young MKPs are exclusively associated with megakaryocyte and Plt generation (Nakorn et al., 2003; Pronk et al., 2007). However, those conclusions were primarily based on
transcription profiles and in vitro differentiation capacity, with no direct assessment of differentiation potential in vivo. We recently reported that classically (Akashi et al., 2000) and alternatively (Pronk et al., 2007) defined GMPs produce erythroid cells in vivo. This fits a model where erythropoiesis is a default HSC fate (Boyer et al., 2019). Under that model, it is not surprising that our experiments revealed that both young and old MkPs harbor in vivo erythroid capacity. Surprisingly, old MkPs also demonstrated greater capacity for the GM lineage compared with young MkPs. It is possible that our observations may be a result of poorly defined cellular phenotypes in old mice, raising the question of how aging might affect the immunophenotype of MkPs. Given that several markers are shared between MkPs and HSCs (Zhu et al., 2019), with a gain of CD41 expression as HSCs age (Gekas and Graf, 2013), the possibility that the MkP compartment in old BM contains significant numbers of HSCs was considered. However, old MkPs lacked B and T cell generation and the enhanced production of Plts, erythroid cells, and GMs did not persist long-term, unlike that of multipotent and self-renewing HSCs. Additionally, we did not observe any age-related differences in expression of megakaryopoiesis master regulators such as Fli1, Gabpa, Mpl, Pf4, and Runx1 (Bruns et al., 2014; De Sauvage et al., 1994; Doré and Crispino, 2011; Elagib et al., 2003; Kaushansky et al., 1994; Klimchenko et al., 2009; Zhu et al., 2018) (Figure 7). A high similarity between these genes suggest high purity of the MkP populations. Together, these findings indicate that the high expansion and oligopotency of old MkPs by transient production of Plts, erythroid cells, and GMs cannot be explained by contaminating HSCs, but rather due to enhanced potential in the aged MkP compartment.

Aging is associated with changes in MkP genome-wide expression signatures

Our transcriptome data provide important clues about the intrinsic molecular regulation of megakaryopoiesis during aging (Figure 7). RNA-seq analysis of young and old HSCs has provided a paradigm of molecular switches during ontogeny (Cabezas-Wallscheid et al., 2014; Grover et al., 2016; Kowalczyk et al., 2015; Sanjuan-Pla et al., 2013; Sun et al., 2014). Similarly, our data showed that young and old MkPs have different gene expression programs, reflecting divergence in the molecular control of Plt differentiation during aging. For example, differential expression of genes involved in proliferation reinforced the differences in proliferative capacity between young and old MkPs. The expansion of MkPs with age also suggest that MkPs themselves may serve as a reservoir for mutations promoting clonal progression to cardiovascular diseases and cancer (Pardali et al., 2020; Steensma and Ebert, 2020). In line with this model, we observed age-related increase in expression of genes involved in AML. Moreover, different transcriptional networks involving Plt function were observed between young and old MkPs, supporting a model in which MkPs propagate events toward age-related dysregulation of Plt biology. An important next step will be to identify key molecular regulators that cause or are a consequence of age-related changes to megakaryopoiesis.

In conclusion, our data revealed that profound perturbations in homeostatic control underlie megakaryopoiesis during the aging process. Given that HSCs gradually lose their regenerative potential during aging, it is reasonable to focus research on the HSC contribution to aging. There remains, however, a conceptual gap in the role of altered progenitor cells as potential perpetuators of aging. Our findings shed light on the mechanisms of aging within the Plt lineage that may dictate the etiology of Plt-related disorders, and other myeloid diseases that may be operating through committed progenitors rather than HSCs. Our study sets the stage for further exploration of mechanisms governing youthful and aging megakaryopoiesis and suggests that progenitor cell involvement should be considered when investigating normal and pathophysiological changes in the hematopoietic system during aging.

EXPERIMENTAL PROCEDURES

Mouse lines

All animals were housed and bred in the Association for Assessment and Accreditation of Laboratory Animal Care–accredited vivarium at UC Santa Cruz and maintained under approved Institutional Animal Care and Use Committee guidelines. Young adult mice were used between 8 and 16 weeks of age and old adult mice were 20+ months of age. Old recipient mice in transplantation experiments were 18+ months of age. All mice were randomized based on sex.

HSC and MkP isolation and analysis by flow cytometry

BM stem and progenitor cell populations and mature cell subsets were prepared and stained as previously described (Beaudin et al., 2014, 2016; Leung et al., 2019; Martin et al., 2021; Smith-Berdan et al, 2015, 2019; Ugarte et al., 2015). HSCs or MkPs from young or old mice were prospectively isolated using a fluorescence-activated cell sorting (FACS) Aria III (Becton Dickinson, San Jose, CA) as previously described (Boyer et al., 2019; Leung et al., 2019; Martin et al., 2021).

Transplantation reconstitution assays

Reconstitution assays were performed by transplanting double-sorted HSCs (200 per recipient) from young or old Ubc-GFP+ whole BM and transplanting into congenic C57BL/6 mice via retro-orbital intravenous transplant. We also transplanted double-sorted MkPs (22,000 per recipient) from C57Bl6 into Ubc-GFP+ hosts. Hosts were preconditioned with sub-lethal radiation (~750 rads) using a Faxitron CP160 X-ray instrument (Precision Instruments).
RNA-seq

The RNA-seq libraries were generated by sorting MkPs from young or old C57BL/6 mice using FACS Aria III (Becton Dickinson). Cells were stored in Trizol (Invitrogen) at −80°C until RNA isolation. RNA-seq libraries were generated using Nextera Library Prep, as we have previously done (Beaudin et al., 2016; Byrne et al., 2017). Libraries were validated using the Bioanalyzer (Agilent 2100), validated RNA-seq libraries were sequenced using Illumina HiSeq 4000 as Paired-end run reads at the QB3-Berkeley Genomics at University of California Berkeley, and DESeq analysis was done with the help of Dr. Sol Katzman at the UCSC Bioinformatics Core. Heatmap was generated using Morpheus, https://software.broadinstitute.org/morpheus. Go Term analysis was run using Panther.

Quantification and statistical analysis

Number of experiments, n, and what n represents can be found in the legend for each figure. Statistical significance was determined by two-tailed unpaired Student’s t test. All data are shown as mean ± standard error of the mean (SEM) representing at least three independent experiments.

Data and code availability

The datasets generated in the current study are available in the Gene Expression Omnibus, accession number GEO: GSE166704.

SUPPLEMENTAL INFORMATION

Supplemental information can be found online at https://doi.org/10.1016/j.stemcr.2021.04.016.

AUTHOR CONTRIBUTIONS

D.M.P. and E.C.F conceived of the project, designed experiments, and co-wrote the manuscript. D.M.P., A.K.W., and S.S.-B. performed experiments, analyzed data, and edited the manuscript. The authors have no competing financial interests.

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