Clinical Study

Ankylosing Spondylitis and Rheumatoid Arthritis: Serum Levels of TNF-α and Its Soluble Receptors during the Course of Therapy with Etanercept and Infliximab

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Received 15 January 2014; Accepted 27 February 2014; Published 24 March 2014

Academic Editor: Lorenzo Cavagna

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The effects of the TNF-α blockers infliximab or etanercept on the levels of TNF-α, TNF-receptor 1 (TNF-R1), and TNF-receptor 2 (TNF-R2), as well as the levels of the inflammation markers CRP and IL-6, were measured in ankylosing spondylitis (AS) and rheumatoid arthritis (RA) patients receiving treatment with either compound. We found that RA patients tend to have higher levels of TNF-α than both healthy individuals and AS patients prior to treatment (P < 0.05). We measured greatly increased levels of TNF-α in both the AS and RA etanercept patient groups during the course of treatment, while in the infliximab treated patients, the amount of TNF-α measured remained unchanged. Elevated TNF-α in the etanercept treated patients does not appear to be a significant risk factor for the spontaneous development of further autoimmune diseases in our study group. Increased levels of TNF-R1 were determined in both AS (P < 0.05) and RA (P < 0.001) patients when compared to healthy controls. In AS patients, the levels of TNF-R1 dropped significantly when treated with either infliximab (P < 0.01) or etanercept (P < 0.001). In contrast, the levels of this receptor remained unchanged in RA patients treated with either compound.

1. Background

Proinflammatory signaling that is activated by TNF-α is an important aspect in the pathology of rheumatic autoimmune diseases such as ankylosing spondylitis (AS) and rheumatoid arthritis (RA). TNF-α exerts its activity through binding to the membrane bound TNF-α receptors TNF-R1 and TNF-R2. In addition to their cell-bound forms, these receptors also exist as soluble molecules (sTNF-Rs) that are a result of enzymatic cleavage of the extracellular portions of the receptors [1]. These soluble receptors are free-floating in the serum and can bind to and act as natural TNF-α antagonists. It is thought that sTNF-R1 and sTNF-R2 modulate and balance the activity of TNF-α in the course of inflammatory events [2, 3].

Anti-TNF-α biologicals that complex TNF-α and neutralize its disease driving activities are beneficial drugs in the treatment of chronic inflammatory disorders. Infliximab, a humanized mouse monoclonal antibody against TNF-α, and etanercept, a soluble fusion protein comprising an epitope derived from the human TNF-receptor 2 fused to the Fc portion of human IgG, represent two different types of biotechnical engineered molecules developed for therapeutic targeting of TNF-α in inflammatory diseases. Both drugs are approved for the treatment of AS and RA as well as for other autoimmune diseases [4].

Although different autoimmune disorders share TNF-α as a major player in disease pathology, there are likely to be alterations in TNF-α regulation that can distinguish between different disease manifestations and respond to different types of TNF-α blocking strategies. Furthermore, studies have reported elevated levels of TNF-α in patients treated with TNF-α blockers and have speculated about the possible consequences thereof [5–8]. In order to see whether we could measure any differences in TNF-α regulation in patients with AS and RA being treated with infliximab and etanercept, we determined the serum concentration of TNF-α as well as
the levels of the soluble TNF-receptors 1 and 2. The measurement of interleukin 6 and C-reactive protein in the serum samples was used to further assess the inflammatory activity in these patients. To follow the course of the disease, we calculated the Bath ankylosing spondylitis disease activity index (BASDAI) and the disease activity score 28 (DAS28) for the AS and RA patients, respectively, at the start of the study and after 12 weeks into treatment.

2. Methods

2.1. Study Participants. The study cohort consists of 45 healthy blood donors, 50 patients with ankylosing spondylitis (AS), and 48 patients with rheumatoid arthritis (RA). AS patients met the criteria of the European Spondyloarthropathy Study Group [9] and RA was diagnosed according to the classification criteria of the American College of Rheumatology [10]. None of the patients exhibited any comorbidities such as malignancies or psoriasis vulgaris, nor showed symptoms of any other chronic diseases. Patients that had been recently immunized, or had a history of viral infection such as HBV or HCV, were excluded from the study. All patients underwent anti-TNF-α therapy for the first time with either etanercept or infliximab. Twenty-two AS and 20 RA patients received etanercept 50 mg subcutaneously weekly during the study, and patients were treated with infliximab received IV infusions of 3 mg/kg body weight (28 RA patients) or 5 mg/kg body weight (28 AS patients) at 0, 2, and 6 and then every 8 weeks (RA patients) or 6 weeks (AS patients) into therapy. Patients treated with infliximab had been treated previously only with NSAIDs, whereas RA patients had received 2 or more DMARDs, including methotrexate, before infliximab or etanercept. RA patients treated with infliximab or etanercept received concomitant methotrexate combined with low-dose glucocorticoids with mean dosages of 15 mg/week and 5 mg/day, respectively. After obtaining written informed consent, serum samples were taken at different time points and stored at −80°C until assayed. Clinical and laboratory assessments were conducted prior to administration of the respective TNF-α blocker at baseline and at 2 and 12 weeks into therapy. The serum concentration of C-reactive protein (CRP) and the disease activity were determined. Assessment of the disease activity was performed before and after 12 weeks of therapy following the Bath ankylosing spondylitis disease activity index (BASDAI) for the AS patients and the disease activity score 28 (DAS28) was calculated for the RA patients.

The study was approved by the local ethics committee of the University of Rostock.

2.2. ELISA. Quantification of sTNF-R1, sTNF-R2, sTNF-α, and IL-6 in serum samples was performed using ELISA-kits (R&D Systems, Wiesbaden, Germany, Catalogue Numbers DY225, DY726, DY210, DY206, resp.) following the manufacturer’s protocol. The detection limit was 10 pg/mL for all determinations.

2.3. Statistical Analysis and Graphical Representation. Statistical analysis was performed using GraphPad Prism software (San Diego, USA). One way ANOVA was used to compare parameters measured in the different study groups and repeated measures ANOVA were used to compare values obtained from the same patients at the three time points. ANOVA analyses were followed by Tukey’s multiple comparison tests. Paired t-test was used to analyze changes in the disease activity scores during the study period. Comparisons among groups and among time points are depicted as box and whiskers plots. The boxes represent the range from the 25th to the 75th percentile and the whiskers represent the maximum and minimum values. The horizontal line through the box indicates the median.

3. Results and Discussion

Comparing the three groups analyzed in the present study, the age of the healthy controls and the AS patients is very similar with an age (mean ± SD) of 45 ± 12 and 45 ± 13 years in both groups. With a mean age of 56 ± 12 years in the RA group, these patients are significantly older (P < 0.001 for both comparisons). The gender distribution within the study cohort shows a higher proportion of females in the control group (25 of 45) and the RA patients (35 of 48), whereas females are underrepresented among the AS patients (21 of 50).

In the AS patient groups treated either with etanercept or infliximab, the anti-TNF-α therapy led to a reduction of the inflammatory activity as seen by a significant drop of the CRP concentration after 2 and 12 weeks into therapy (P < 0.01 for both treatments). The RA patients show a trend towards decreasing CRP levels during both treatments, but this effect does not reach statistical significance. The proinflammatory cytokine IL-6 also tends to decrease during therapy in all treatment groups, but only reached statistical significance in the AS patients treated with etanercept and infliximab at the 2 week time point (both P < 0.05). All serum parameters determined before and during anti-TNF-α therapy are listed in Table 1.

All four treatment groups showed a significant decrease in the disease activity scores (BASDAI for AS and DAS28 for RA) determined before the first administration of a TNF-α blocker and after 12 weeks into therapy (Figure 1). This response shows that 12 weeks of treatment with infliximab and etanercept leads to clinical improvement in both diseases in our study cohort.

3.1. Serum TNF-α. Prior to treatment, 32% (inflimabimagroup) and 20% (etanercept group) of RA patients were found to be TNF-α positive (>10 pg/mL). This is around twice as many as TNF-α positive AS patients prior to treatment (14% and 9%, infliximab and etanercept groups, respectively, Table 1). The mean TNF-α level prior to treatment of 8.7 ± 17.9 pg/mL in RA patients was significantly higher (P < 0.05) than in AS patients (2.4 ± 7.2 pg/mL) and in healthy participants (2.5 ± 7.9 pg/mL).

During the course of TNF-α blockade, no significant change in the level or in the percentage of TNF-α positive study subjects could be observed in the AS and the RA
to a previous study by Charles et al., in which a dose
and measured the amount of TNF-
not different when comparing the AS and RA patients during treatment (Table 1).

Our observation that the level of measured TNF-α does not increase in infliximab treated patients is in contrast to a previous study by Charles et al., in which a dose
dependent increase in serum TNF-α in RA patients being treated with infliximab (cA2) was found [11]. We therefore tested whether the amount of TNF-α that is measured with the ELISA kits that we employed can be affected by either the presence of etanercept or infliximab in the sample. In order to do so, we mixed and incubated (30 minutes at room temperature) various concentrations of TNF-α with physiologically relevant amounts of either TNF-α blocker, and measured the amount of TNF-α using our anti-TNF-α ELISA assay. We found that both agents reduced the measured amount of TNF-α, with the effect by infliximab being much larger than that of etanercept (Figure 2(c)). As such, we are not able to interpret our measurements of TNF-α made in the presence of infliximab, since our assay system is not capable of measuring most of the TNF-α that is bound by this compound. Although we cannot rule out the possibility

| Week | Infliximab | Etanercept | Infliximab | Etanercept |
|------|------------|------------|------------|------------|
| 0    | CRP        | N = 28     | CRP        | N = 22     |
|      | mg/mL      | ±21.2      | ±6.9       | ±9.5       | ±15.5      | ±5.0       | ±4.8       | ±27.5      | ±14.9      | ±24.6      | ±31.2      | ±23.3      | ±28.4      |
| 2    | 18.4       | 5.9        | 11.24      | 3.1        | 20.9       | 7.7        | 13.0       | 26.5       | 19.4       | 21.8       |           |           |           |
| 12   | 7.2        | 15.2       | 6.3        | 11.7       | 7.5        | 5.2        | 19.8       | 13.3       | 10.9       |           |           |           |
| IL-6 |           | ±0.4       | ±1.9       | ±18.3      | ±8.7       | ±10.8      | ±28.8      | ±21.4      | ±7.1       | ±25.7      | ±27.5      | ±20.4      |
| TNF-α|           | ±2.5       | ±10.7      | ±17.0      | ±23.4      | ±19.3      | ±13.0      | ±27.2      | ±15.7      | ±15.8      | ±28.7      |           |           |
| Patients|         | 14%        | 11%        | 9%         | 96%        | 82%        | 32%        | 18%        | 32%        | 20%        | 70%        | 65%        |
| TNF-R1| ng/mL      | ±0.74      | ±0.1       | ±1.9       | ±18.3      | ±10.8      | ±28.8      | ±21.4      | ±7.1       | ±25.7      | ±27.5      | ±20.4      |
| TNF-R2| ng/mL      | ±2.3       | ±1.6       | ±1.9       | ±18.3      | ±10.8      | ±28.8      | ±21.4      | ±7.1       | ±25.7      | ±27.5      | ±20.4      |
that autoantibodies to infliximab could also influence the quantitation of TNF-\(\alpha\), the fact that the presence of infliximab influences the amount of TNF-\(\alpha\) measured in our assay might explain the different results that we obtained when compared with the study of Charles et al., that used a different ELISA assay system to measure TNF-\(\alpha\) in the presence of infliximab.

In the past years, studies have emerged reporting increased serum concentrations of TNF-\(\alpha\) as measured using ELISA assays in individuals being treated with etanercept for various conditions including rheumatic autoimmune disease [12]. The elevated serum levels could be explained by both an increased half-life of TNF-\(\alpha\) when complexed with etanercept [13], and an upregulated expression and/or release of soluble TNF-\(\alpha\), that would be consistent with the observation that etanercept can enhance the capacity of immune cells to produce TNF-\(\alpha\) [14], or possibly by the different biochemical and pharmacokinetic properties of the two anti-TNF-\(\alpha\) agents.

While TNF-\(\alpha\) is thought to be biologically inactive when complexed with the receptor fusion protein etanercept, the idea has been put forward that increased serum levels of TNF-\(\alpha\) resulting from etanercept treatment can trigger autoimmune reactions such as psoriasis and Crohn’s disease. The possible consequences thereof have been the subject of some debate, and it has been suggested that routine monitoring of serum TNF-\(\alpha\) should be performed in all patients being treated with anti-TNF-\(\alpha\) biologicals for this reason—even though the interpretation of such measurements remains unclear [5–8]. Although TNF-\(\alpha\) is increased in the vast majority of the patients treated with etanercept in this study, we had not observed any new autoimmune symptoms such as psoriasis in these patients. Although we did measure differences in the levels of TNF-\(\alpha\) in individual patients during the course of therapy with etanercept, the reasons for these differences are unclear, and we have no evidence to support the routine measurement of TNF-\(\alpha\) in patients being treated with these compounds.

### 3.2. Soluble TNF-\(\alpha\) Receptor 1 and 2

Serum TNF-RI showed elevated concentrations in both the AS and RA patient groups prior to anti-TNF-\(\alpha\) treatment when compared to the controls (Figure 3). The mean concentration in the healthy blood donors was 0.66 ± 0.17 ng/mL compared to 0.89 ± 0.39 ng/mL in the AS patients and 1.22 ± 0.49 ng/mL in the RA patients. The differences among the groups reached statistical significance with \(P < 0.05\) (AS versus control) and \(P < 0.001\) (RA versus control and RA versus AS). No significant
differences in the serum TNF-R2 concentration could be detected when comparing the healthy control subjects with the two patient groups at the 0 week time point. The mean concentrations were 2.86 ± 1.80 ng/mL in the healthy subjects and 2.79 ± 1.33 and 3.22 ± 1.49 ng/mL in the AS and RA patient group, respectively. These results are in partial agreement with previously published data on RA patients, which showed elevated levels of both TNF-α receptors when compared with healthy controls [15, 16].

After 2 weeks into therapy, the AS patients showed a significant decrease in TNF-R1 levels when treated with either infliximab or etanercept. This effect was maintained at the 12 week time point, resulting in a 15 and 23% reduction of TNF-R1 in this group, the fact that the levels of TNF-R1 are not affected by either TNF-α blocker in RA patients might be explained by a distinct regulation of TNF-α signaling in AS compared to RA patients. Our study suggests that soluble TNF-R1 might be a good marker of inflammation in AS patients. It is an easily measured parameter that could be accurately quantified in every patient, whereas TNF-α, CRP, and IL-6 are often below the detection limits in routinely used assays.

No significant change in TNF-R2 was observed during infliximab treatment of the AS and RA patients. In the presence of etanercept, the measurement of serum TNF-R2 was interfered with by the TNF-R2 portion of the drug, and this resulted in values that were out of the range of the assay. We analyzed serial dilutions of etanercept using our TNF-R2 ELISA and found that the reactivity of etanercept in this immunoassay was about 10 to 15% when compared to the recombinant TNF-R2 standard calibrators (Figure 4(a)). We therefore diluted etanercept treated patient sera 1000-fold and reassayed them in the TNF-R2 ELISA. The ELISA determinations were about 100-fold higher as compared to endogenous TNF-R2 levels prior to the start of treatment, with mean concentrations corresponding to 570 ± 348 and 280 ± 198 ng/mL in AS and RA patient sera, respectively. Although etanercept is only partially recognized in the TNF-R2 ELISA, the assay can still be used to compare the relative concentration of etanercept in the patient sera. At the 12 week time point, when an established level of etanercept can be expected, the concentrations detected as well as the variation of the levels found in the group of AS patients were significantly higher (P < 0.001) when compared to the RA patients (Figure 4(b)). To test the possibility that high levels of TNF-α might interfere in the TNF-R2 ELISA, we mixed etanercept with varying concentrations of TNF-α and assayed these samples using ELISA. We found no effect of TNF-α up to a concentration of 1000 pg/mL (data not shown).

As both AS and RA are treated by self-administration of etanercept, both at the same dosage and time interval, our measurements indicate that distinct differences in drug metabolism between our patient groups exist. This is in contrast to a study where no significant change in etanercept pharmacokinetics could be detected when comparing AS patients with RA patients [20]. It is unlikely that the difference in the composition of our patient groups might account for the different etanercept levels we observe between AS and RA patients, since it has been shown that etanercept disposition does not differ between males and females and does not vary with age in adult patients [21, 22]. The fact that two patients with AS and two patients with RA displayed TNF-R2 concentrations at the 12 week time points similar to those found prior to treatment begins could be explained by noncompliance in these patients. This could also explain why elevated levels of TNF-α were not detected in these patients. Interestingly, a recent study found that the levels of etanercept at 3 months into treatment can predict response to therapy at 6 months, suggesting that the monitoring of etanercept during treatment might be of prognostic value [23].

4. Conclusions

The increase in measurable TNF-α frequently observed upon etanercept treatment likely reflects biochemical and physiological properties in the regulation of TNF-α and was not identified as being a risk factor for triggering further inflammatory reactions in our study. In AS patients, the measurement of TNF-α receptor 1 might be useful as a biological indicator of TNF-α blocker activity. The determination
of serum etanercept might be useful in the monitoring of medication compliance and to allow for a quantification of the steady state concentration of these compounds in individual patients.

Abbreviations

ACR: American College of Rheumatology
AS: Ankylosing spondylitis
BASDAI: Bath ankylosing spondylitis disease activity index
CRP: C-reactive protein
DAS28: Disease activity score 28
IL-6: Interleukin-6
RA: Rheumatoid arthritis
sTNF-α: Soluble tumor necrosis factor α
sTNF-R1/2: Soluble tumor necrosis factor receptor 1/2.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Authors’ Contribution

Martin Schulz, Helmut Dotzlaw, and Gunther Neeck participated in the design of the study. Gunther Neeck selected and performed clinical evaluation of study subjects. Helmut Dotzlaw performed ELISA analyses, and Martin Schulz performed statistical analyses. Martin Schulz, Helmut Dotzlaw, and Gunther Neeck discussed and interpreted all data. Martin Schulz drafted the paper, and Helmut Dotzlaw and Gunther Neeck critically read and revised the paper. All authors read and approved the final submitted paper.

References

[1] C. A. Smith, T. Farrah, and R. G. Goodwin, “The TNF receptor superfamily of cellular and viral proteins: activation, costimulation, and death,” Cell, vol. 76, no. 6, pp. 959–962, 1994.
[2] K. J. Tracey and A. Cerami, “Tumor necrosis factor: a pleiotropic cytokine and therapeutic target,” Annual Review of Medicine, vol. 45, pp. 491–503, 1994.
[3] S. Xanthoulea, M. Pasparakis, S. Kouteni et al., “Tumor necrosis factor (TNF) receptor shedding controls thresholds of innate immune activation that balance opposing TNF functions in infectious and inflammatory diseases,” Journal of Experimental Medicine, vol. 200, no. 3, pp. 367–376, 2004.
[4] P. P. Sfikakis, “The first decade of biologic TNF antagonists in clinical practice: lessons learned, unresolved issues and future directions,” Current Directions in Autoimmunity, vol. 11, pp. 180–210, 2010.
[5] A. Bhatia and R. E. Kast, “Tumor necrosis factor (TNF) can paradoxically increase on etanercept treatment, occasionally contributing to TNF-mediated disease,” The Journal of Rheumatology, vol. 34, no. 2, pp. 447–449, 2007.
[6] R. E. Kast and E. L. Altschuler, “Both etanercept and infliximab can elevate tumor necrosis factor (TNF)-α and be the cause of treatment related new onset disease: the need to measure circulating TNF-α,” The Journal of Rheumatology, vol. 35, no. 8, p. 1679, 2008.
[7] I. Sari, S. Akar, M. Birlik, B. Sis, F. Onen, and N. Akkoc, “Anti-tumor necrosis factor-α-induced psoriasis,” The Journal of Rheumatology, vol. 33, no. 7, pp. 1411–1414, 2006.
[8] H. S. In, H. Appel, H. Haibel et al., “New onset of Crohn’s disease during treatment of active ankylosing spondylitis with etanercept,” The Journal of Rheumatology, vol. 35, no. 3, pp. 532–536, 2008.
[9] M. Dougados, S. van der Linden, R. Juhlin et al., “The European Spondylarthropathy Study Group preliminary criteria for the classification of spondylarthropathy,” Arthritis and Rheumatism, vol. 34, no. 10, pp. 1218–1227, 1991.
[10] D. Aletaha, T. Neogi, A. J. Silman et al., “2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative,” *Arthritis and Rheumatism*, vol. 62, no. 9, pp. 2569–2581, 2010.

[11] P. Charles, M. J. Elliott, D. Davis et al., “Regulation of cytokines, cytokine inhibitors, and acute-phase proteins following anti-TNF-α therapy in rheumatoid arthritis,” *The Journal of Immunology*, vol. 163, no. 3, pp. 1521–1528, 1999.

[12] J. X. Zou, J. Braun, and J. Sieper, “Immunological basis for the use of TNFα-blocking agents in ankylosing spondylitis and immunological changes during treatment,” *Clinical and Experimental Rheumatology*, vol. 20, no. 6, supplement 28, pp. S34–S37, 2002.

[13] M. L. Nowlan, E. Drewe, H. Bulsara et al., “Systemic cytokine levels and the effects of etanercept in TNF receptor-associated periodic syndrome (TRAPS) involving a C33Y mutation in TNFRSF1A,” *Rheumatology*, vol. 45, no. 1, pp. 31–37, 2006.

[14] J. Zou, M. Rudwaleit, J. Brandt, A. Thiel, J. Braun, and J. Sieper, “Up regulation of the production of tumour necrosis factor α and interferon γ by T cells in ankylosing spondylitis during treatment with etanercept,” *Annals of the Rheumatic Diseases*, vol. 62, no. 6, pp. 561–564, 2003.

[15] T. Robak, A. Gidlaska, and H. Stepień, “The tumour necrosis factor family of receptors/ligands in the serum of patients with rheumatoid arthritis,” *European Cytokine Network*, vol. 9, no. 2, pp. 145–154, 1998.

[16] Y. Valle, I. Y. Ledezma-Lozano, N. Torres-Carrillo et al., “Circulating TNFRI and TNFRII levels correlated with the disease activity score (DAS28) in rheumatoid arthritis,” *Scandinavian Journal of Rheumatology*, vol. 38, no. 5, pp. 332–335, 2009.

[17] F. M. Coelho, H. J. Reis, R. Nicolato et al., “Increased serum levels of inflammatory markers in chronic institutionalized patients with schizophrenia,” *NeuroImmunoModulation*, vol. 15, no. 2, pp. 140–144, 2008.

[18] D. L. Mattey, J. R. Glossop, N. B. Nixon, and P. T. Dawes, “Circulating levels of tumor necrosis factor receptors are highly predictive of mortality in patients with rheumatoid arthritis,” *Arthritis and Rheumatism*, vol. 56, no. 12, pp. 3940–3948, 2007.

[19] P. Scalzo, A. Kümmér, P. Cardoso, and A. L. Teixeira, “Increased serum levels of soluble tumor necrosis factor-α receptor-1 in patients with Parkinson’s disease,” *Journal of Neuroimmunology*, vol. 216, no. 1-2, pp. 122–125, 2009.

[20] H. Zhou, M. Buckwalter, J. Boni et al., “Population-based pharmacokinetics of the soluble TNFα etanercept: a clinical study in 43 patients with ankylosing spondylitis compared with post hoc data from patients with rheumatoid arthritis,” *International Journal of Clinical Pharmacology and Therapeutics*, vol. 42, no. 5, pp. 267–276, 2004.

[21] “Enbrel (etanercept) Prescribing Information,” http://www.enbrel.com.

[22] D. Tracey, L. Klareskog, E. H. Sasso, J. G. Salfeld, and P. P. Tak, “Tumor necrosis factor antagonist mechanisms of action: a comprehensive review,” *Pharmacology and Therapeutics*, vol. 117, no. 2, pp. 244–279, 2008.

[23] C. I. Daïen, V. Daïen, E. Parussini, A. M. Dupuy, B. Combe, and J. Morel, “Etanercept concentration in patients with rheumatoid arthritis and its potential influence on treatment decision: a pilot study,” *The Journal of Rheumatology*, vol. 39, no. 8, pp. 1533–1538, 2012.