Oxidative stress in cancer and fibrosis: Opportunity for therapeutic intervention with antioxidant compounds, enzymes, and nanoparticles

Jingga Morrya,1, Worapol Ngamcherdtrakulab,1, Wassana Yantaseea,b,*

a Department of Biomedical Engineering, Oregon Health and Science University, 3303 SW Bond Ave, Portland, OR 97239, USA
b PDX Pharmaceuticals, LLC, 3303 SW Bond Ave, Portland, OR 97239, USA

1 Authors with equal contribution.

E-mail address: yantasee@ohsu.edu (W. Yantasee).

⁎ Corresponding author at: Department of Biomedical Engineering, Oregon Health and Science University, 3303 SW Bond Ave, Portland, OR 97239, USA.

16–25

1. Introduction

The regulation of redox homeostasis is crucial for the maintenance of normal cellular growth, metabolism, and survival. Oxidative stress is defined as the imbalance between the production of reactive oxygen species (ROS) and the capability of the cell to elicit an effective antioxidant response. At lower concentrations, ROS are important signaling molecules involved in cellular proliferation, migration, and apoptosis [1,2]. Several sources of ROS in cells and tissue have been identified, including mitochondrial electron transfer chain [3] and NADPH oxidase (NOX) enzymes [4]. At higher concentrations, these molecules could be useful against pathogens, resulting in increased leukocyte and platelet activation, and increased leukocyte recruitment [5]. While this is true in the context of innate immunity and inflammatory signaling in the immune cells, most ROS are harmful to cells due to the accumulation of irreversible damages to proteins, lipids, and most importantly, to DNA leading to mutations and cell death [6,7].

ROS and oxidative stress have been implicated in a number of diseases, including fibrosis and cancer [8]. NOX-derived ROS have been identified as the main source of oxidative stress, which promotes key events in the development of fibrotic diseases (such as skin fibrosis [9], idiopathic pulmonary fibrosis [10], liver fibrosis [11], and kidney fibrosis [12]) as well as the initiation and progression of cancer [13]. To date, there is no cure for most of these diseases. Current approaches are limited to attempts on slowing down disease progression in fibrotic diseases (such as pirfenidone for pulmonary fibrosis). For cancer, there are several treatment approaches including, chemotherapy, surgery, radiation, immunotherapy and other novel targeted therapies. Cures can be achieved in some cases (e.g., when tumors are diagnosed early), but resistance and recurrence are common. Chemotherapy and radiation also generate ROS, which, at high levels, are toxic to cancer cells. Nevertheless, sub-lethal ROS generated by these treatments were also reported to promote cancer invasion and metastasis [14]. ROS are thus considered one of the mediators of drug resistance and metastasis in cancer [14–16].

In recent years, antioxidants have drawn much attention as potential therapeutic interventions due to their ability to fight oxidative stress (and thereby negate its role) in fibrosis and cancer development. The main function of antioxidants is to scavenge or neutralize free radical formation and to inhibit the deleterious downstream effects of ROS. However, most antioxidants, taken orally, have limited absorption profile, which leads to low bioavailability and insufficient concentrations at the target site [17,18]. To overcome this issue, research has been focused on developing nanoparticles with intrinsic antioxidant properties which can be functionalized to provide localized or targeted...
therapy. These nanoparticles are mostly made up of inorganic materials such as mesoporous silica, cerium oxide, and fullerene, which exhibit antioxidant activities, protecting cells against oxidative stress when evaluated in vitro and in animal models. The antioxidant capacities of these nanoparticles are thought to be contributed by their redox and catalytic properties, electronic configuration, oxygen vacancy defects, and high-surface-to-volume ratio [19,20]. Additionally, nanoparticles can be designed to be multi-functional, also serving as delivery platforms for other therapeutics.

2. Overview of reactive oxygen species (ROS)

Reactive species are broadly categorized into 4 groups: ROS, reactive nitrogen species (RNS), reactive sulfur species (RSS), and reactive chloride species (RCS) [21]. Among these groups, ROS are found to be most abundantly produced [21]. ROS are generally defined as oxygen-containing small species including superoxide anion radical \( \cdot O_2^- \), hydroxyl radical (\( \cdot OH \)), hydroxyl ion (\( \cdot O^- \)), hydrogen peroxide (\( H_2O_2 \)), singlet oxygen (\( ^1O_2 \)), and ozone (\( O_3 \)) [4,21]. ROS can be generated either by exogenous sources such as UV radiation, toxic chemicals and drugs, physiological changes such as aging or injury/inflammation [22], or by intracellular (endogenous) sources such as NOX enzymes on the plasma membrane [4], myeloperoxidases (MPO) in phagocytes [23], and as by-products of respiratory chain function in mitochondria [3]. As highlighted in Fig. 1, ROS generation is a cascade of reactions initiated by the production of \( O_2^- \) inside the cells, contributed by endogenous and exogenous cellular sources. Cellular defenses against these ROS molecules involve endogenous antioxidants, such as glutathione peroxidases (GPx), catalases (CAT), and superoxide dismutases (SOD) [24]. Under normal physiological conditions, the formation and elimination of ROS is tightly regulated through the help of the ROS-scavengers/endogenous antioxidants to maintain homeostasis and avoid the harmful effects of oxidative stress [24]. However, the elimination process can become saturated and the increased accumulation of ROS leads to permanent changes and/or damages to the DNA, lipids and proteins with detrimental effects, such as cell death, mutagenesis, carcinogenesis and fibrosis.

2.1. Roles of ROS in fibrosis

Fibrosis is a complex disease characterized by excessive synthesis and accumulation of extracellular matrices that occur as a result of activation and proliferation of fibroblasts and myofibroblasts. Fibrogenesis can be broadly categorized into four different stages: 1) initiation of tissue injury, 2) inflammation and activation of fibroblasts, 3) extracellular matrix (ECM) synthesis, and 4) deposition of ECM, which eventually leads to organ failure [25]. The causes of fibrosis vary greatly, but common contributing factors include i) physical or chemical injury, ii) autoimmune disease (e.g., systemic sclerosis) [26], iii) virus-induced (e.g., hepatitis C virus-induced liver fibrosis) [27], iv) alcohol-induced (e.g., liver fibrosis) [28], v) hypertension (e.g., hypertensive myocardial fibrosis), or vi) unknown (e.g., idiopathic pulmonary fibrosis) [26,29,30]. Notably, nearly 45% of all naturally-occurring deaths in the western world are attributed to some form of fibrotic disease [31].

The release of ROS along with the secretion of chemokines and growth factors (such as platelet-derived growth factor (PDGF), transforming growth factor beta (TGF-\( \beta \)), connective tissue growth factor (CTGF), interleukin-6 (IL-6), and interleukin-13 (IL-13)) by immune cells during the inflammation phase is known to promote the activation of fibroblast and collagen deposition in fibrosis [32,33]. Among them, TGF-\( \beta \) is the most potent profibrogenic cytokine, which plays a vital role in regulating important biological processes such as cellular proliferation, extracellular matrix (ECM) production, and epithelial–mesenchymal transition (EMT) [22]. TGF-\( \beta \) mRNA and/or protein expression has been found to be elevated in most fibrotic diseases in patients [34–36] as well as experimental fibrosis models [37–39]. As shown in Fig. 2, the presence of ROS could activate TGF-\( \beta \) signaling pathways, which then signal through either SMAD-dependent or SMAD-independent pathways (e.g., phosphatidylinositol-3-kinase

![Fig. 1. Sources of ROS and key ROS molecules in signaling.](image-url)

**Fig. 1. Sources of ROS and key ROS molecules in signaling.** ROS generation is a cascade of reaction initiated by the production of \( O_2^- \) inside the cells, contributed by endogenous and exogenous cellular sources. Mucosal oxygen is reduced to superoxide anion (\( \cdot O_2^- \)) by enzymes such as NOX and nitric oxide synthases (NOS), or as by-products of redox reactions in mitochondrial respirations. \( O_2^- \), being cell-impermeant molecule, is then rapidly dismutated to \( H_2O_2 \) either spontaneously or enzymatically by antioxidant enzyme superoxide dismutases (SODs). The intracellular removal of \( H_2O_2 \) can be categorized into three different mechanisms: 1) by the action of catalase (CAT) and glutathione peroxidases (GPx) which reduces \( H_2O_2 \) to water, 2) through conversion of \( H_2O_2 \) into hypochlorous acid (HOCl) and \( \cdot O^- \) by the heme enzyme myeloperoxidase (MPO) the neutrophils, which results in antimicrobial activity, and 3) by Fenton reaction whereby \( H_2O_2 \) is converted to the highly reactive \( \cdot OH \) through oxidation of \( Fe^{2+} \) to \( Fe^{3+} \). The \( \cdot OH \) produced will then react with \( H_2O_2 \) to form \( O_2^- \), which, again, reacts with \( H_2O_2 \) to form \( OH^- \) and \( \cdot O_2^- \), as a part of Haber-Weiss reaction.
into myofibroblasts, and excessive ECM deposition leading to fibrosis.

Fig. 2. ROS contribute to the induction and persistence of TGF-β-mediated fibrosis. The presence of ROS induces the conversion of latent TGF-β complex to its active form, which binds to its receptor and triggers signaling pathways such as SMAD2/3, PI3K, and JNK. This in turn increases the transcriptional activity of various pro-fibrotic genes, such as NOX4, a-SMA, and COL I. Increase in NOX4 expression also results in ROS generation, which leads activation of other ROS-dependent signaling transduction pathways such as, NFκB and JNK. Elevated ROS also causes irreversible DNA damage, through oxidation of its bases. Together, enhanced ROS and activated TGF-β signaling contributes to proliferation and transdifferentiation of fibroblast cells into myofibroblasts, and excessive ECM deposition leading to fibrosis.

(Pi3K), c-Jun N-terminal kinases (JNK)) [40]. Increased TGF-β signaling also induces elevated production of NOX4-generated ROS [41], which further stimulates the transcriptional activities of pro-fibrotic genes such as collagen I (COL1), alpha smooth muscle actin (α-SMA), and NOX4. In addition, the presence of NOX4-generated ROS could activate signaling pathways such as JNK and nuclear factor kappa B (NFκB) [42,43], and trigger DNA oxidation as the initial step in a cascade of events which lead to myofibroblast differentiation and overaccumulation of collagen deposition into ECM, leading to fibrosis.

2.2. Roles of ROS in cancer

Cancer is the second leading cause of death in the United States and was responsible for 584,872 deaths in 2013 [44]. The number of new cancer cases is estimated to climb to 22 million worldwide within the next two decades [45]. Elevated ROS levels have been detected in most cancer cell lines [46] and have been implicated in malignant progression and resistance to treatment [47]. As highlighted in Fig. 3, ROS play a critical role in various signaling cascades relating to survival, proliferation, resistance to apoptosis, neovascularization, invasion, and extravasation and growth into a distant metastasis site [48,49]. The roles of ROS in cancer can be described as follows.

2.2.1. Effects of ROS on reox-mediated cellular mechanisms

ROS are capable of modifying numerous cellular pathways by altering the DNA binding sites of reox-sensitive transcription factors (such as hypoxia-inducible factor-1 alpha (HIF-1α), NFκB, activator protein-1 (AP-1), and p53), or by oxidizing the cysteine residues on these molecules [43]. At the post-translational level, ROS could also directly oxidize multiple forms of amino acids, such as methionine to sulfoxide and cysteine to sulfonic acids [50]. These oxidative modifications on the amino acids will lead to structural and conformational change of the tertiary protein structure, which might cause protein degradation by proteasomes or activation/inhibition of the protein activities. Direct protein carbonylation can also occur through oxidative attack on amino acids involved in catalysis such as lysine, arginine, proline, and threonine, which leads to enzyme inactivation [50].

2.2.2. Role of ROS in genomic instability

ROS generated from either the extracellular/intracellular sources could also lead to DNA damage, which in turn activates a number of stress response genes and DNA repair mechanisms. The reox-sensitive p53 protein is an active transcription factor that is involved in numerous cell processes including cell cycle arrest, senescence, and apoptosis [51]. In the presence of excess ROS, p53 plays a crucial role in preventing the propagation of DNA damage [52]. However, in cancer cells, TP53 (gene which encodes p53) is a commonly mutated gene varying from 10% occurrence in diseases, such as hematopoietic malignancies, to close to 100% in high-grade serous carcinoma of the ovary [53]. In these cancers, DNA damage will accumulate more readily due to inadequate DNA repair mechanisms, resulting in gene mutation and/or deletion. The genomic instability will further activate a number of oncogenes resulting in abnormal metabolic activity and decreased antioxidant production. All these events will eventually lead to an increase in intracellular ROS production in a positive-feedback manner [54].

2.2.3. Role of ROS in tumor hypoxia

As primary tumors continue to grow, the demand for nutrients and oxygen supply will increase in parallel. However, these demands are not always met in rapidly growing tumors and regions of the tumor will become deprived of oxygen. In order to support tumor growth and proliferation in these hypoxic microenvironments, cancer cells undergo several changes to adapt to this oxygen- and nutrient-deprived state, including genotype selections favouring survival (such as TP53 mutation [55]) and activation of hypoxia inducible factor-1 (HIF-1) transcription factor [56]. The HIF family regulates a broad array of genes in response to oxygen deprivation and has been comprehensively reviewed elsewhere [56,57]. In hypoxic conditions, the hydroxylation of HIF-1α is inhibited, preventing it from being degraded as is normoxic conditions. The HIF-1α then dimerizes with HIF-1β, which later binds to hypoxia response elements (HREs) on the DNA and stimulates the transcription of its target genes, such as vascular endothelial growth factor (VEGF), N-myc downstream-regulated gene (NDRG), and glucose transporter I [58]. These hypoxia-responsive genes are involved in glucose transport, glycolysis, and angiogenesis, allowing cancer cells to survive in such harsh environment. A hypoxic microenvironment also contributes to ROS formation through the release of superoxide, hydrogen peroxide, and hydroxyl radical from the mitochondrial electron transport chain, and ROS, in turn, also stabilize HIF-1α under both normoxic and hypoxic conditions [59–61].

2.2.4. Interplay between ROS and TGF-β signaling in cancer

Similar to fibrosis, the cross-talk between ROS and TGF-β signaling in cancer has been well-documented and comprehensively reviewed [62,63]. TGF-β1 is one of the most potent cytokines known to contribute to immunosuppression of immune cells and promoting angiogenesis and EMT in cancer cells [64]. TGF-β1 induces apoptosis in immune cells by directly suppressing the production of cytolytic factors in T-cells, inhibiting proliferation and differentiation of numerous immune cells, and decreasing the tumor surface immunogenicity through inhibition of major histocompatibility complex class II antigens [65]. Gorelik et al. showed that T-cell specific blockade of TGF-β signaling could enhance anti-tumor immunity by the generation of CD8+-mediated tumor-specific cytotoxic T-cells response [66].
Tumor angiogenesis is vital for tumor growth and can also facilitate the dissemination of tumor cells. TGF-β plays a critical role in promoting angiogenesis. The TGF-β SMAD-dependent signaling pathway has been shown to induce vascular endothelial growth factor (VEGF) expression. In addition, different levels of TGF-β expression show distinct effects on angiogenesis: at low levels, TGF-β upregulates angiogenic factors including VEGF, CTGF, and fibroblast growth factor (FGF), while at high levels, TGF-β stimulates smooth muscle cells recruitment and cell differentiation, while inhibiting endothelial cell growth [67].

TGF-β is a major inducer of EMT and cell migration through a combination of SMAD-dependent and -independent pathways (e.g., p38 MAPK) [68]. The downstream effects of the EMT response include transcriptional reprogramming which promotes inactivation of genes (such as E-cadherin) that encodes for epithelial markers and activation of genes for mesenchymal proteins such as N-cadherin and vimentin [69,70]. Downregulation of E-cadherin is a common feature in many cancers such as metastatic breast cancer [71] and non-small cell lung cancer (NSCLC) [72]. Studies have shown that forced expression of E-cadherin in cancer cells in vitro could suppress cellular migration and invasiveness [73], while forced expression of N-cadherin in cancer cells caused the opposite effects [74]. The shift in expression from E- to N-cadherin and their distinctive expression patterns reflects the EMT phenotype, which is associated with cancer malignancy and metastasis [69].

In addition, TGF-β has been identified as a major contributor of intracellular ROS production through NOX4 activation. NOX4-derived ROS have been implicated in the EMT phenotype in pancreatic cancer cells [62], increased cell survival in urothelial carcinoma [75], and increased cellular migration and invasiveness in breast cancer [76] and ovarian cells [77], respectively.

2.2.5. Role of ROS in metastases

Metastasis is the main cause of cancer-related mortality, which accounts for 90% of death in cancer patients [78]. The metastatic cascade, as shown in Fig. 3, is a complex process encompassing multiple steps, which lead to cancer cell dissemination, such as: 1) loss of cellular adhesion, 2) increased motility and invasiveness of cancer cells through ECM, 3) intravasation and entry into the circulation, 4) exit into a distant tissue (extravasation), and 5) colonization of a new foreign environment [78]. ROS can activate several pathways involved in metastasis. For example, ROS can activate matrix metalloproteinases (MMPs), which can degrade basement membranes and extracellular matrices, facilitating intravasation and extravasation of cancer cells [79]. Furthermore, ROS generated by the NOX family (to be discussed in the next section) were shown to be crucial for the formation of invadopodia, actin-rich membrane protrusions of cancer cells that facilitate pericellular proteolysis and invasive behavior [80]. Reduction of ROS using antioxidant such as N-acetylcysteine (NAC) or NOX inhibitor, DPI, in cancer cells has the ability to decrease cell viability [81], invasion and invadopodia formation [80], suggesting the role of antioxidant in mitigating metastasis.

3. NOX4 – the main source of ROS in fibrosis and cancer

3.1. NADPH oxidase (NOX) family and ROS

NOX family is comprised of seven members including NOX1-5 and dual oxidase DUOX1-2, which are among the best-characterized intracellular ROS-generating enzymes (as shown in Fig. 4) [4,82]. All are transmembrane flavoproteins, which generate ROS by transferring an electron to an oxygen molecule, resulting in superoxide anion (O$_2^-$), which is then either spontaneously (by low pH) or catalytically (by SOD) dismutated to H$_2$O$_2$. Most NOXes require additional subunits to be functional, specifically NOX 1-3, bind to the transmembrane protein p22phox, which further recruits cytosolic regulatory subunits, such as organizers (p47phox, p40phox, or NOXO1), activators (p67phox or NOXA1), and small GTPases (Rac1 or Rac2) [83,84]. NOX4, being the exception, only needs to bind to p22phox and does not require cytosolic subunits for maximal oxidase activity. NOX1-5 are mostly located at the plasma membrane of the cell, with NOX4 being additionally detected in the endoplasmic reticulum (ER), mitochondrial membrane, nuclear membrane, focal adhesions, and invadopodia [84]. Extensive details on the structure and activation of NOX isoforms have been reviewed elsewhere [4,82,85].

3.2. High NOX4 expression in fibrosis

NOX4 mRNA expression has been found to be upregulated in both pulmonary fibroblasts isolated from idiopathic pulmonary fibrosis (IPF) patients [86] and skin fibroblasts from scleroderma patients [87], as well as in a number of in vivo fibrosis models, including liver fibrosis [88], pulmonary fibrosis [89,90], diabetic neuropathy (kidney fibrosis associated with diabetes mellitus) [91].

As mentioned in the previous section (Section 2), TGF-β signaling is the major contributor to fibrogenesis. TGF-β upregulates NOX4...
3.3. High NOX4 expression in cancer

High expression of NOX4 has been detected in several cancer types including gliomas [96], melanoma [97], breast cancer [98], ovarian cancer [98], and pancreatic cancer [62]. In cancer cell lines, elevated levels of NOX4 are associated with PI3K/Akt-regulated cell proliferation and invasion [99], TGF-β/SMAD3-driven EMT and cell migration [95], as well as Tks5-dependent invadopodia formation [80]. Depletion of NOX4 with siRNA treatment significantly reduced tumor growth in the in vivo models of bladder cancer [75], renal cancer [100], and glioblastoma [57]. These results suggest that NOX4 is a potential target for pharmacological intervention for cancer treatment.

4. Strategies to suppress oxidative stress

Antioxidants have been commonly described as substances that can delay, prevent or remove oxidative damage to a target molecule [101]. Given that fibrotic and cancer cells are generally present with higher oxidative stress levels than normal cells, it is believed that patients who suffers from those diseases will benefit from antioxidant supplementation.

4.1. Dietary antioxidant supplements

Dietary antioxidants including vitamin C (ascorbic acid), vitamin E (tocopherol), vitamin A (β-carotene), and selenium have the ability to counteract oxidative damage and can be obtained through food components such as fruits and vegetables.

Vitamin C is a water-soluble and strong antioxidant. Vitamin C exists in two forms: L-ascorbic acid and the oxidized form, dehydro-L-ascorbic acid. It can directly react with hydroxyl and lipid peroxy radicals to form H₂O and lipid hydroperoxides. Vitamin C can also neutralize vitamin E and glutathione radicals, and regenerate these antioxidants [102].

Vitamin E exists in at least 8 different isoforms (α-, β-, γ-, δ-tocopherols, and α-, β-, γ-, δ-tocotrienols), which differ only in the number of methyl groups and in the side chains of their aliphatic tails [103]. Only α-tocopherols isomer is mostly retained in the body due to the preferential transfer of α-tocopherol to the lipid particles by a liver α-tocopherol transfer protein. The main role of vitamin E is to act as a chain-breaking antioxidant, which prevents the propagation of lipid peroxidation [104].

Vitamin A is a fat-soluble vitamin and usually found in the diet as preformed vitamin A from animal products such as meat and fish, and as pro-vitamin A from plant-based products such as fruits and vegetables. β-carotene has the highest provitamin A activity, which is further metabolized to retinoic acid and retinol, the active form of vitamin A. β-carotene can physically quench 1O₂ and protect organisms from oxidative damage [105].

Selenium is an essential trace element, which can be acquired from the diet by the consumptions of nuts, meats, and fish. It is co-translationally incorporated into amino acids, such as selenocysteine and selenomethionine [106]. The selenium-containing amino acids act as antioxidants by scavenging free radicals and repairing oxidized selenium species.

A few clinical trials (summarized in Table 1) have been conducted, mainly using the synthetic form of these antioxidants on healthy and at-risk populations [107–117]. These observational studies were designed to provide evidence on the benefit of antioxidant supplementation for reducing or lowering the risk of patients developing or dying from cancer. However, most of the data were inconclusive, with the majority showing no protection or exhibiting harmful effects in the patient cohort. It is possible that this lack of benefit is due to: 1) the difference in the chemical composition of antioxidants found in food compared to those in supplements, 2) the disease-specificity of certain antioxidants (i.e., some antioxidants are more effective than the others in protecting against certain types of diseases), or 3) due to the low bioavailability, these supplements could not reach sufficient intracellular levels to be effective [118]. Therefore, a more potent antioxidant that can be delivered to a specific diseased tissue and with improved bioavailability is thought to be more beneficial than these antioxidant supplements.
Table 1
Antioxidant supplements in clinical trials.

| Name of trials                                | Type of antioxidants                                                                 | Target population                                                                 | Length of study | Conclusion of the study                                                                 | Ref. |
|-----------------------------------------------|--------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------|-----------------|----------------------------------------------------------------------------------------|------|
| Linxian general population nutrition intervention trial | 15 mg beta-carotene, 30 mg alpha-tocopherol, and 50 μg selenium daily                | 29,584 healthy Chinese men and women in North China at increased risk of developing esophageal cancer and gastric cancer were recruited | 5 years         | reduction in cancer mortality associated with gastric cancer, but not esophageal cancer | [107]|
| Alpha-Tocopherol/Beta-Carotene Cancer Prevention Study (ATBC) | alpha-tocopherol (50 mg/day) or beta-carotene (20 mg/day) or both                   | 29,133 male smokers in Finland                                                   | 5–8 years       | no overall reduction in the incidence of lung cancer or in mortality in all treatment groups | [112]|
| Carotene and Retinol Efficacy Trial (CARET)    | 30 mg of β-carotene plus 25,000 IU of retinyl palmitate daily                        | 816 men with substantial occupational exposures to asbestos and 1029 men and women who were either current or former cigarette smokers in United States | 6–12 years      | beta-carotene supplementation was associated with increased lung cancer incidence and all caused mortality which persisted up to 6 years after the supplementation was ended | [113]|
| Physicians' Health Study I (PHS I)            | 50 mg β-carotene every other day                                                    | 22,071 male physicians between age of 40–84 years in the United States            | 12 years        | supplementation did not reduce the incidence of prostate cancer or other cancers, including lymphoma, leukemia, melanoma, and cancers of the lung, bladder, pancreas, and colon and rectum | [114]|
| Physicians' Health Study II (PHS II)          | 400 IU vitamin E every other day, 500 mg vitamin C every day, 50 mg β-carotene or in combination | 14,642 male physicians older than 50 years old in the United States               | 8 years         | daily multivitamin use was associated with a reduction in total cancer among 1312 men with a baseline history of cancer, but did not differ significantly from that among 13,329 men initially without cancer | [109]|
| Women's Health Study (WHS)                    | 50 mg β-carotene every other day, vitamin E supplementation (600 IU every other day), and aspirin (300 mg every other day) | 39,876 women aged 45 years or older                                              | 2 years         | no benefit or harm associated with 2 years of beta-carotene supplementation           | [115]|
| Selenium and Vitamin E Cancer Prevention Trial (SELECT) | daily supplementation with selenium (200 μg), vitamin E (400 IU), or both          | 35,533 men from 427 participating sites in the United States, Canada, and Puerto Rico | 7 years         | the use of supplements did not reduce the incidence of prostate or other cancers after 1.5 years post supplementation, the follow-up study found 17% increase in prostate cancer incidence among men taking vitamin E alone than among men taking a placebo | [109]|
|                                               |                                                                                      |                                                                                   | 8.5 years       |                                                                                         | [108]|
4.2. Enzyme-related antioxidants

Glutathione (GSH), N-acetylcysteine (NAC), and superoxide dismutase (SOD) are enzyme-related antioxidants that act as the first-line defense against cellular oxidants. The effects of these molecules have also been investigated in several clinical trials (see Table 2), and the results from these studies will be discussed in details below.

4.2.1. GSH and NAC

GSH is the main non-protein thiol in cells, which acts as a reducing agent and is essential in regulating cellular redox status. GSH is involved in cell protection against free radicals and many other cellular functions [119]. GSH is also critical for the regeneration of other antioxidants, such as tocopherols and ascorbate [120]. NAC is a cysteine precursor that replenishes the intracellular levels of GSH [121]. A few clinical trials have been conducted with either GSH or NAC as interventions for fibrotic diseases, such as liver fibrosis [122,123] and lung fibrosis [124,125], as well as cancers, such as head and neck cancer or lung cancer [111]. However, the data have been largely disappointing, with most of them showing no beneficial effects. This is mostly contributed by the low bioavailability of GSH and NAC. GSH is known to be poorly absorbed when ingested due to the action of GSH peroxidase [121,122,123] and lung fibrosis [124,125], as well as cancers, such as head and neck cancer or lung cancer [111]. However, the data have been largely disappointing, with most of them showing no beneficial effects. This is mostly contributed by the low bioavailability of GSH and NAC. GSH is known to be poorly absorbed when ingested due to the action of GSH peroxidase [121,122,123].

NAC is usually prescribed at a much lower dose of only 150 mg/kg loading dose (within the first 60 min) and 2250 mg/kg maintenance dose (for the next 4 h) are needed to reach 10 mM concentration in blood based on the pharmacokinetic data of NAC in human volunteers [127], but NAC is usually prescribed at a much lower dose of only 150 mg/kg loading dose and 30 mg/kg maintenance dose (i.v.) (NAC, Acetadote®, package insert) or 600-mg oral dose (three times daily) for pulmonary fibrosis patients in the PANTHER-IPF trial [128].

SODs are metal-containing proteins that catalyze the conversion of superoxide to hydrogen peroxide. Three isoforms have been identified, namely, cytosolic Cu-Zn SOD (SOD1), mitochondrial MnSOD (SOD2), and extracellular SOD (SOD3). The cytosolic and mitochondrial SODs have been indicated in multiple studies as tumor suppressor genes. Overexpression of MnSOD suppressed the malignancy of human breast cancer cells [129,130], glioma cells [131], and melanoma cells [132]. In contrast, depletion of MnSOD resulted in increased cell proliferation in vitro and contributed to more aggressive tumor growth in vivo [133]. Likewise, overexpression of Cu-Zn-SOD decreased tumor growth in multiple cancer types [130,134].

SOD overexpression has been shown to confer protection against radiation and display chemopreventive effects in in vivo cancer models. Preclinical studies in mouse models have shown that intraoral delivery of MnSOD2 plasmid/liposomes (MnSOD-PL) to mice decreased radiation-induced mucosal ulceration [135] as well as esophagitis [136]. Based on the results from these pre-clinical studies, the chemoprotective effects of MnSOD-PL were investigated in radiation/chemotherapy-induced esophagitis in NSCLC patients (see Table 2). Overall, in the phase I clinical trial study, the response rate for the chemoradiation regimen was satisfying at 70% and the treatment was safe and well-tolerated [137]. Unfortunately, the Phase II study was later suspended (reason unknown). In another study [138], topical delivery of Cu-Zn SOD1 (APN 201) was tested on 44 female breast cancer patients for its ability to prevent radiation-induced dermatitis in the patients who undergo radiotherapy after surgery. The topical treatment of Cu-Zn SOD1 reduced pain in the fibrotic region in more than 90% of the cases and decreased the fibrotic size by half in 30% of the patients. In a separate study, topical delivery of liposomal human recombinant Cu-Zn SOD1 (APN 201) was safe and well tolerated in 20 female breast cancer patients who received radiation therapy after breast-preserving surgery [139].

Overall, these studies show some promising results on the chemoprotective effects of the SODs. That said, no clinical trial has been
4.3. Inhibiting NOX enzymes

Currently, a few nonspecific NOX inhibitors have been identified (which target more than one NOX isoforms) [88,142–147]. Some issues with specificity, potency, and toxicity of these inhibitors have limited the use of these compounds in clinical studies [148]. Gene silencing of NOXes has been carried out with shRNA and siRNA [75,99,149] but primarily as a proof-of-concept in in vivo studies without using suitable delivery agents such as liposomes or nanoparticles to safely and efficiently deliver these molecules to the right target. The different types of NOX inhibitors as well as NOX gene therapeutics, and their potential applications from the in-vivo studies are summarized in Table 3.

*Diphenyleneiodonium (DPI)* is a nonspecific inhibitor for NOX enzymes. It inhibits a number of flavoproteins including eNOS, xanthine oxidase, and cholinesterases and the internal calcium pump [150]. Although at least two pre-clinical studies using DPI have shown positive results on lung cancer inhibition [142] as well as skin fibrosis reduction [143], the off-target effects of DPI against other flavoproteins will prevent its translation into clinical use.

**Fulvene-5** is a water-soluble small molecule inhibitor for NOX2 and NOX4 enzymes. It has an aromatic structure, which allows electron delocalization. Treatment of Fulvene-5 on bEnd.3 endothelioma cells NOX4 enzymes. It inhibits a number of flavoproteins including eNOS, xanthine oxidase, and cholinesterases and the internal calcium pump [150]. Although at least two pre-clinical studies using DPI have shown positive results on lung cancer inhibition [142] as well as skin fibrosis reduction [143], the off-target effects of DPI against other flavoproteins will prevent its translation into clinical use.

**Imipramine blue** is an organic triphenylmethane blue dye that is a derivative for the antidepressant drug, imipramine. It has been shown to target NOX4; however, there was insufficient characterization data provided regarding its selectivity on other NOX isoforms. Treatment with imipramine blue in vivo resulted in tumor inhibition of human glioma cells as well as enhancing treatment efficacy in combination with chemotherapy, doxorubicin [152]. In another study, imipramine blue treatment in head and neck squamous cell carcinoma (HNSCC) tumor in mice inhibited tumor invasion and metastatic colonization in the lungs [153].

**GKT136901** and **GKT137831** are both small molecule inhibitors developed by a French biotech company, Genkyotex. Both compounds have high specificity to NOX1 and 4 (and less for NOX2 and 5). Both compounds show therapeutic potential in reducing liver fibrosis and tumor growth in mice [88,147,154]. GKT137831 is currently a lead compound undergoing Phase II clinical trials in diabetic kidney disease (NCT02010242). The preliminary results from this clinical trial showed that patients receiving treatment for 12 weeks had few adverse effects than placebo. However, the primary efficacy endpoint was not achieved due to negligible change in albuminuria (Genkyotex press release, September 2015).

4.3.1. NOX4 shRNA/siRNA

NOX4 gene silencing using siRNA or shRNA has shown good results in terms of tumor reduction in multiple cancer types including NSCLC, hepatocarcinoma, and bladder cancer [75,99,149] as well as conducted so far to investigate the effectiveness of SODs in combatting cancer growth in humans; however, several SODs mimetics have been shown to be beneficial in the pre-clinical models of prostate [140] and breast cancer [141].
reduction in pulmonary fibrosis [90]. However, delivering siRNA into the target cells requires a delivery agent, such as atelocollagen [75], since siRNA is unstable under physiological conditions.

4.4. Nanoparticles with intrinsic antioxidant properties

Nanotechnology has become a main focus of biomedical research in recent years. Several types of inorganic nanoparticles possess intrinsic antioxidant properties by scavenging free radicals and decreasing ROS concentrations. In this section, these inorganic nanoparticles will be discussed and summarized in Table 4.

4.4.1. Cerium oxide (CeO2)

Cerium is a rare-earth element, which belongs to the lanthanide series in the periodic table. CeO2 can exist in both Ce3+ and Ce4+ valence states, which give it the unique ability as an antioxidant [155]. The ROS-scavenging capability of CeO2 is based on redox cycling between Ce3+ and Ce4+ on the nanoparticle surface. Giri et al. reported that treatment of CeO2 nanoparticles attenuated tumor growth in A2780 ovarian cancer mice and significantly inhibited the metastasis of these cancer cells into the lungs of mice [156]. They also found that angiogenesis in the tumors was reduced in treated mice as indicated by less CD31-positive staining. The same group also reported that surface functionalization of CeO2 nanoparticle with folic acid and its co-delivery with cisplatin further decreased the tumor burden and angiogenesis [157]. In other studies, CoO2 nanoparticle administration to carbon-tetrachloride (CCL4)-induced liver fibrosis mice was shown to inhibit oxidative [158,159] and endoplasmic reticulum stress signaling pathways as well as reduction in inflammatory cytokines such as TNF-α and IL-1β [158].

4.4.2. Fullerene (C60)

Fullerene is a sphere-like molecule composed of 60 carbon atoms, which are arranged in a hexagonal formation to form a hollow spherical structure. It is a powerful antioxidant due to the delocalization of the π-electrons over the carbon atoms, which can readily react with free radicals [160]. Fullerene is also capable of inactivating hydroxyl radicals via attachment to its double bonds [161]. Due to these attractive antioxidant properties, fullerenes and its derivatives have been studied in numerous biomedical applications including fibrosis and cancer. Prophylactic treatment of fullerene in CCl4-induced hepatic injury rat model prevented damage in the liver and kidney of the rats [162]. Another study has also shown that co-treatment of Doxorubicin chemotherapy drug with fullerene resulted in inhibition of tumor growth and metastasis, and increased survival in LLC tumor-bearing mice [163].

4.4.3. Platinum nanoparticles (PtNPs)

PtNPs are known to possess ROS scavenging ability due to the catalytic activity contributed by its high ratio of electrons to particle surface [164]. PtNPs have demonstrated its therapeutic potential in several pre-clinical applications such as treating aging-related skin disease in SOD1 knockout mice [165], protection against UV-induced apoptosis in HaCaT keratinocytes [166], and prevention of hepatic injury from hepatic ischemia/reperfusion injury in mice [164].

4.4.4. Mesoporous silica nanoparticles (MSNPs)

MSNPs have been reported to have reactive oxygen species (ROS), hydroxyl radical, and free radical scavenging capability [167–169]. ROS scavenging by MSNP also attenuates NOX4 mRNA expression in melanoma cells in vitro [168]. Thus, MSNPs have great potential in the treatment of oxidative-induced pathological conditions such as fibrosis and cancer. Among all classes of antioxidant nanoparticles discussed herein, MSNP is most widely used for delivery of drugs, genes, and imaging agents. MSNP delivery carriers have many favorable attributes, such as tailorable mesoporous structures, high surface areas, large pore volumes, ease of controlling size, and high scalability [170]. MSNP is soluble in physiological pH to non-toxic silicic acid, which can be cleared by the kidneys [171]. Silica is endogenous in the human body unlike cerium and platinum. Numerous studies have also shown great in vitro [172,173] and in vivo [174–176] biocompatibility of silica nanoparticles. These features are desirable since it will prevent long-term toxicity issues of the nanomaterials in vivo. Silica nanoparticles with PET tracers have also recently entered a clinical trial with a favorable safety profile (Cornell dots) [177]. Amorphous silica nano-particles have been widely used in cosmetics, food and pill additives as anti-caking.

Our group reported successful development of MSNP-based siRNA delivery platform, which is a 50-nm MSNP core coated with cross-linked PEI, PEG, and conjugated with an antibody for targeted systemic delivery for breast cancer treatment [173]. We also showed that the same platform is effective for skin fibrosis treatment by simultaneously utilizing therapeutic siRNAs and inherent antioxidant property of MSNP [169]. Treatment of the polymer-coated MSNP reduced ROS in murine dermal fibroblast cells under TGF-β stimulated conditions. Localized intradermal injection of the material in scleroderma-like (skin fibrosis) mice resulted in reduction of pro-fibrotic gene expressions and skin thickness (Fig. 5) [169]. In brief, we established a skin fibrosis mouse model by repeated intradermal bleomycin injections, resulting in an increase in skin thickness (Fig. 5A–C). The antioxidant MSNP based nanoparticle clearly suppressed the NOX4 expression to the baseline level of mice not receiving bleomycin (Fig. 5E). The anti-fibrotic effect was enhanced by incorporating siRNA against heat shock protein 47 (siHSP47) on the nanopar-
The antioxidant property of MSNP resulted in significant decrease in skin thickness (Fig. 5B–C) and pro-fibrotic markers, α-SMA (Fig. 5F) and COL I-positive area of skin sections harvested upon sacrifice. Immunofluorescence images can be found in ref [169]. Reproduced with modification (permission from Elsevier [169]).

5. Conclusions

Reactive oxygen species (ROS) have prominent roles in pathogenesis of fibrosis and cancer. Many researchers thus have studied antioxidant compounds, enzymes, and NOX inhibitors for treating such diseases, some of which have also been evaluated in clinical trials. However, the results to date have been suboptimal mainly due to their low systemic bioavailability and insufficient levels at the target sites. Antioxidant nanoparticles present a unique opportunity because they can be made larger than a kidney filtration cut-off size (~10 nm), hence prolonging circulation compared to small molecules. They can also be designed further to avoid rapid phagocytic clearance or to target specific sites and organs. Nanoparticles are also widely studied as delivery platforms for genes, oligonucleotides, drugs, and imaging agents, providing numerous opportunities in developing theranostic agents or co-delivering synergistic therapeutic agents within the same nanoparticle. For instance, we have recently reviewed different classes of nanoparticle candidates for siRNA delivery and the progress in clinical trials for systemic cancer treatment [179]. Nanoparticles are typically viewed as merely passive delivery carriers. Inherent antioxidant property of nanoparticles is thus one beneficial attribute that will set certain nanoparticle classes apart from others.

Conflict of interest disclosure

The authors declare the following competing financial interest(s): OHSU, J.M., W.N., and W.Y. have a significant financial interest in PDX Pharmaceuticals, LLC, a company that may have a commercial interest in the results of this research and technology. This potential personal and institutional conflict of interest has been reviewed and managed by OHSU.

Acknowledgements

We are grateful to Dr. Xiaolin Nan for independently reviewing this manuscript. This work was supported by National Cancer Institute of NIH under a contract HHSN261201300078C, the Prospect Creek Foundation, and OHSU’s Office of the Vice President for Research.

References

[1] D.I. Brown, K.K. Griengl, Regulation of signal transduction by reactive oxygen species in the cardiovascular system, Circ. Res. 116 (3) (2015) 531–549.
[2] G.-Y. Liou, P. Storz, Reactive oxygen species in cancer, Free Radic. Res. 44 (5) (2010). http://dx.doi.org/10.3109/10715761003667554.
[3] Y. Lin, G. Fiskum, D. Schubert, Generation of reactive oxygen species by the mitochondrial electron transport chain, J. Neurochem. 80 (5) (2002) 780–787.
[4] K. Bedard, K.H. Krause, The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology, Physiol. Rev. 87 (1) (2007) 245–313.
[5] F.C. Fang, Antimicrobial actions of reactive oxygen species, mBio 2 (5) (2011).
[6] L.A. Rowe, N. Degtyareva, P.W. Doetsch, D.N.A. Damage-induced, Reactive oxygen species (ROS) stress response in Saccharomyces cerevisiae, Free Radic. Biol. Med. 45 (8) (2008) 1167–1177.
[7] M.Schieber, N.Chandel, ROS function in redox signaling and oxidative stress, Curr. Biol. 24(10) , pp. R453–R462.
[8] G. Waris, H. Ahsan, Reactive oxygen species: role in the development of cancer and various chronic conditions, J. Carcinog. 5 (2006) 14-14.
[9] O. Babalola, A. Mamalis, H. Lev-Tov, J. Jagdeo, NADPH oxidase enzymes in skin fibrosis: molecular targets and therapeutic agents, Arch. Dermatol. Res. 306 (4) (2014) 313–330.
[10] L. Hecker, J. Cheng, V.J. Thanrickkal, Targeting, NOX enzymes in pulmonary fibrosis, Cell Mol. Life Sci. 69 (14) (2012) 2365–2371.
[11] S. De Minicis, D.A. Brenner, NOX in liver fibrosis, Arch. Biochem. Biophys. 462 (2) (2007) 266–272.
[12] C.E. Holtermann, N.C. Read, C.R. Kennedy, Nox and renal disease, Clin. Sci. (London) 128 (8) (2015) 465–481.
[13] K. Roy, Y. Wu, J.L. Meitlzer, A. Juhasz, H. Liu, G. Jiang, J. Lu, S. Antony, J.H. Doroshow, NADPH oxidases and cancer, Clinical Sci. (London) 128 (12) (2015) 863–875.
[14] M. Quintavalle, L. Elia, J.H. Price, S. Heynen-Genel, S.A. Courtneidge, A cell-based high-content screening assay reveals activators and inhibitors of cancer cell invasion, Sci. Signal. 4 (183) (2011) ra49.
[15] I.S. Okon, M.-H. Zou, Mitochondrial, ROS and cancer drug resistance: implica-
tions for therapy, Pharmacol. Res. 100 (2015) 170–174.

[16] M. Nishikawa, Reactive oxygen species in tumor metastasis, Cancer Lett. 266 (1) (2008) 53–59.

[17] B. Olsson, M. Johansson, J. Gabrielson, P. Bolme, Pharmacokinetics and bioavailability of reduced and oxidized N-acetylcysteine, Eur. J. Clin. Pharmacol. 34 (1) (1988) 77–82.

[18] E.B. Souto, P. Severino, R. Basso, M.H. Santana, Encapsulation of antioxidants in gastroprotectin-based vesicular nanoparticulate carriers, Methods Mol. Biol. (Clifton, N.J.) 1028 (2013) 37–46.

[19] E. Sharpe, D. Andreassae, S. Andreassae, Artificial Nanoparticle Antioxidants, Oxidative Stress: Diagnostic, Prevention, and Therapy, American Chemical Society, Washington, DC, 2011, pp. 235–253 http://pubs.acs.org/doi/abs/10.1021/bk-2011-1128.ch033.

[20] R. Sandhir, A. Yadav, A. Sunkaria, N. Singhal, Nano-antioxidants: an emerging strategy for intervention against neurodegenerative conditions, Neurochem. Int. 89 (2015) 209–225.

[21] V. Sosa, T. Moliné, R. Paciucci, H. Kondoh, M.E. Lleonart, Oxidative stress and cancer: an overview, Ageing Res. Rev. 12 (1) (2013) 376–390.

[22] K. Richter, A. Konzack, T. Philipovich, R. Heljasvaara, T. Kietzmann, Redox-fibrosis: impact of TGFβ1α on ROS generators, mediators and functional consequences, Redox Biol. 6 (2015) 344–352.

[23] J.M. Robinson, Reactive oxygen species in phagocytic leukocytes, Histochem. Cell Biol. 130 (2) (2008) 281–297.

[24] C. Michiels, M. Raes, O. Toussaint, J. Remacle, Importance of SE-glutathione disulfide, J. Clin. Investig., 122(8), pp. 2756–2766.

[25] G. Sebastiani, K. Gkouvatsos, K. Pantopoulos, Chronic hepatitis C and liver fibrosis, Alcool. Clin. Exp. Res. 29 (11 Suppl) (2005) 1025–1096.

[26] T.K. Cohan, J.I. Chana, S. Czur, F. Czarny, T.K. Chen, Y. Tan, M.S. Tsai, T.-N. Lin, S.-K. Shyue, Stabilization of hypoxia-inducible factor-1α by prostatycin under prolonged hypoxia via reducing reactive oxygen species level in endothelial cells, J. Biol. Chem. 280 (45) (2005) 36567–36574.

[27] V.L. Semenza, Hypoxia-inducible factors: mediators of cancer progression and targets for cancer therapy, Trends Pharmacol. Sci. 33 (4) (2012) 207–214.

[28] D. Trachootham, J. Alexander, P. Huang, Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach?, Nat. Rev. Drug Discov. 8 (7) (2009) 591–607.

[29] T.G. Graeber, C. Osmanian, T. Jacks, D.E. Housman, C.J. Koch, S.W. Lowe, A.J. Giaccia, Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumors, Nature 374 (1995) 88–91.

[30] J.L. Semenza, Hypoxia-inducible factors in cancer progression, Science 316 (5824) (2007) 866–477.

[31] N. Rivlin, R. Brosh, M. Oren, V. Rotter, Mutations in the p53 Tumor Suppressor Gene: important Milestones at the Various Steps of Tumorigenesis, Genes Cancer 1 (4) (2010) 466–477.

[32] P.A. Grimsmo, H. Xie, T.J. Griffin, D.A. Bernholc, Oxidative stress and covalent modification of protein with bioactive aldehydes, J. Biol. Chem. 283 (32) (2008) 21787–21814.

[33] B. Liu, Y. Chen, D.K. St Clair, ROS and p53: versatile partner, Free Radic. Biol. Med. 44 (8) (2008) 1529–1535.

[34] Y.P. Lin, T.-T. Lin, Y.L. Chan, A.C. Song, B.H. Yeo, B. Vojtesek, D. Coomber, G. Rajagopal, D. Lamas, The p53 database: an integrated information resource for p53 research, Oncogene 26 (11) (2007) 1517–1521.

[35] D. Pohlers, J. Brenmoehl, I. Löf, J. Michiels, M. Raes, O. Toussaint, J. Remacle, Importance of SE-glutathione disulfide, J. Biol. Chem. 271 (50) (1996) 2555–2559.

[36] T.-C. Chang, J.-H. Chang, D. Tregay, C. Tan, S.-F. Chen, K.T. Tan, M.-S. Tsai, T.-N. Lin, S.-K. Shyue, Stabilization of hypoxia-inducible factor-1α by prostatycin under prolonged hypoxia via reducing reactive oxygen species level in endothelial cells, J. Biol. Chem. 280 (45) (2005) 36567–36574.

[37] D. Trachootham, J. Alexander, P. Huang, Targeting cancer cells by ROS-mediated mechanisms: a radical therapeutic approach?, Nat. Rev. Drug Discov. 8 (7) (2009) 591–607.

[38] T.-C. Chang, C.-H. Chang, J.-H. Chang, D. Tregay, C. Tan, S.-F. Chen, K.T. Tan, M.-S. Tsai, T.-N. Lin, S.-K. Shyue, Stabilization of hypoxia-inducible factor-1α by prostatycin under prolonged hypoxia via reducing reactive oxygen species level in endothelial cells, J. Biol. Chem. 280 (45) (2005) 36567–36574.

[39] T. Kato, Y. Sato, K. Shimizu, M. Kato, Y. Yamada, Oxidative stress: a new player in cancer chemotherapy, Curr. Med. Chem. 12 (21) (2005) 2651–2662.

[40] T. Sakurai, M. Kudo, Signaling pathways governing tumor angiogenesis, Oncology 67 (6) (2004) 364–369.

[41] D. Pohlers, J. Brenmoehl, I. Löf, J. Michiels, M. Raes, O. Toussaint, J. Remacle, Importance of SE-glutathione disulfide, J. Biol. Chem. 271 (50) (1996) 2555–2559.

[42] R.B. Hazan, G.R. Phillips, R.F. Qiao, L. Norton, S.A. Aaronson, Exogenous exposure to reactive oxygen species enhances the metastatic potential of ovarian cancer cells through signal transducer and activator of transcription 5-dependent regulation of wound healing in a solid organ, Nat. Rev. Immunol. 13 (5) (2013) 730–737.

[43] B.W. Stewart, C.P. Wild, World Cancer Report 2014, 2014.

[44] B.W. Stewart, C.P. Wild, World Cancer Report 2014, 2014.

[45] B.W. Stewart, C.P. Wild, World Cancer Report 2014, 2014.

[46] B.W. Stewart, C.P. Wild, World Cancer Report 2014, 2014.

[47] B.W. Stewart, C.P. Wild, World Cancer Report 2014, 2014.

[48] B.W. Stewart, C.P. Wild, World Cancer Report 2014, 2014.

[49] B.W. Stewart, C.P. Wild, World Cancer Report 2014, 2014.
[134] Y. Zhang, W. Zhao, H.J. Zhang, F.E. Domann, L.W. Oberley, Overexpression of manganese superoxide dismutase inhibits breast cancer growth, Free Radic. Biol. Med. 41 (2006) 520.

[135] R.P. Brandes, S. Devaraj, N.J. Torok, Liver distribution and in vivo antioxidant effects of cerium oxide nanoparticles in mice, PLoS One 9 (10) (2014) e109288.

[136] S. Garrido-Urbani, S. Jemelin, C. De Morel, K. Kusamori, M. Oyama, M. Sano, T. Sakane, A. Yamamoto, Pharmacokinetics and distribution of di-iodoanthracene-4,7-dione-modified mesoporous silica nanoparticles for treating human malignant melanoma growth by mesoporous silica nanoparticles through the NADPH oxidase pathway, J. Pharmacol. Exp. Ther. 334 (2) (2010) 489–498.

[137] S. Koulaloglou, Topical superoxide dismutase reduces post-irradiation breast inflammation, Exp. Dermatol. 19 (11) (2010) 829–836.

[138] J. Morry, W. Ngamcherdtrakul, M.-K. Kim, J.-K. Lee, J. Jeong, S.-W. Kim, K.-J. Park, S.-M. Goodyear, T. Sangvanich, In vivo biodistribution and urinary excretion effects of two polymorphic variants of manganese superoxide dismutase on human breast MCF-7 carcinoma cells, Drug Deliv. 21 (4) (2014) 291–296.

[139] H. Schmidt, K. Wingler, Comparative pharmacology of chemically distinct mesoporous silica nanoparticles: applications and prospects in nanomedicine, Nanomedicine (Lond. Eng.) 8 (9) (2013) 1483–1508.

[140] S. Patel, K. Kato, I. Watanabe, J. Kubo, T. Yamashita, Pharmacokinetics and bioavailability of cerium oxide nanoparticles in mice, PLoS One 9 (8) (2014) e105626.

[141] K. Rong, S. Zheng, X. Xiao, Conservative management of neonatal hepatic hemangiomata: a report from one institute, Pediatr. Surg. Int. 25 (6) (2009) 491–498.

[142] J.M. Munson, L. Fried, S.A. Rowson, M.Y. Bonner, L. Karumabiah, B. Diaz, S.A. Courteille, U.G. Knaus, D.J. Brat, J.L. Arisher, R.V. Bellackonda, Anti-inflammatory adjuvant therapy with imipramine blue enhances chemotherapeutic activity, Cancer Biol. Med. 4 (127) (2012) 127x236-127x46.

[143] W.H. Yang, Y.H. Su, W.H. Hsu, C.C. Wang, J.L. Arisher, M.H. Yang, Imipramine blue halts head and neck cancer invasion through promoting F-box and leucine-rich repeat protein 14-mediated Tweak1 degradation, Oncogene 35 (18) (2016) 2287–2298.

[144] J.-A. Lee, M.-K. Kim, H.-J. Paek, Y.-R. Kim, J.-K. Lee, J. Jeong, S.-W. Kim, K.-J. Park, S.-M. Goodyear, T. Sangvanich, Inhibition of NADPH oxidase protects against metastasis of human lung cancer by inhibiting breast cancer growth, Free Radic. Biol. Med. 41 (2) (2006) 226–237.

[145] S. Shibuya, Y. Ozawa, K. Watanabe, N. Ito, T. Doi, Y. Koko, T. Shimizu, Palladium and platinum nanoparticles attenuate aging-like skin atrophy via antioxidant activity in mice, PLoS One 9 (10) (2014) e109288.

[146] Y. Yoshihisa, A. Honda, Q.L. Zhao, T. Makino, R. Abe, K. Matsui, H. Shimizu, Y. Matsugotou, T. Kondo, T. Shimizu, Protective effects of platinum nanoparticles against UV-light-induced epidermal inflammation, Exp. Dermatol. 19 (11) (2010) 1000–1006.

[147] S. Hori, R. Yasuda, T. Fujita, T. Sato, T. Komagome, S. Miyahara, K. Noguchi, N. Ohtani, K. Ito, T. Fujita, T. Sato, Anti-inflammatory activity of zinc superoxide dismutase and manganese superoxide dismutase in the lung, J. Cell. Mol. Med. 8 (1) (2004) 109–117.

[148] K. Morry, W. Ngamcherdtrakul, S. Gu, S.M. Goodyear, D.J. Castro, J.A. Nieweg, Y. Zhao, V. Lin, Biocompatible mesoporous silica nanoparticles: applications and prospects in nanomedicine, Biomaterials 36 (15) (2014) 3435–3442.

[149] D. Tarn, C.E. Ashley, M. Xue, E.C. Carnes, J.I. Zink, C.J. Brinker, Mesoporous silica nanoparticles with di-iodoanthracene-4,7-dione-modified mesoporous silica nanoparticles for treating human malignant melanoma growth by mesoporous silica nanoparticles through the NADPH oxidase pathway, J. Pharmacol. Exp. Ther. 334 (2) (2010) 489–498.

[150] S. Shih, A. Aras, H. L.mac, M. Tsang, K. Kusamori, M. Sano, T. Sakane, A. Yamamoto, Pharmacokinetics and preventive effects of platinum nanoparticles as reactive oxygen species scavengers on hepatic ischemia/reperfusion injury in mice, Metallo Biomarkers 6 (5) (2014) 1050–1056.

[151] Q. He, Z. Zhang, F. Gao, Y. Li, J. Shi, In vivo biodistribution and urinary excretion effects of cerium oxide nanoparticles in mice, Environ. Toxicol. 28 (2) (2013) 107–118.

[152] J. Grebowski, A. Krokosz, Fullerenes in radiobiology, Post. Biochem. 56 (4) (2010) 341–348.

[153] G.V. Andrievsky, V.I. Bruskov, A.A. Tykhomyrov, S.V. Gudkov, Peculiarities of the antioxidant and radioprotective effects of hydrated C60 fullerene nanostuctures in vitro and in vivo, Free Radic. Biol. Med. 46 (6) (2009) 786–793.

[154] S. Garrido-Urbani, S. Jemelin, C. De Morel, K. Kusamori, M. Oyama, M. Sano, T. Sakane, A. Yamamoto, Pharmacokinetics and preventive effects of platinum nanoparticles as reactive oxygen species scavengers on hepatic ischemia/reperfusion injury in mice, Metallo Biomarkers 6 (5) (2014) 1050–1056.

[155] S. Das, J.M. Dowding, K.F. McMinnis, W. Self, C.M. Reilly, Bioavailability and biodistribution of SiO2 nanoparticles: applications and prospects in nanomedicine, Nanomedicine (Lond. Eng.) 8 (9) (2013) 1483–1508.

[156] S. Patel, K. Kato, I. Watanabe, J. Kubo, T. Yamashita, Pharmacokinetics and distribution of di-iodoanthracene-4,7-dione-modified mesoporous silica nanoparticles for treating human malignant melanoma growth by mesoporous silica nanoparticles through the NADPH oxidase pathway, J. Pharmacol. Exp. Ther. 334 (2) (2010) 489–499.

[157] J.V. Tschudin, D. Tarn, C.E. Ashley, M. Xue, E.C. Carnes, J.I. Zink, C.J. Brinker, Mesoporous silica nanoparticles: applications and prospects in nanomedicine, Nanomedicine (Lond. Eng.) 8 (9) (2013) 1483–1508.
J. Choi, Tissue distribution and excretion kinetics of orally administered silica nanoparticles in rats, Int. J. Nanomed. 9 (Suppl. 2) (2014) 251–260.

[177] E. Phillips, O. Penate-Medina, P.B. Zanzonico, R.D. Carvajal, P. Mohan, Y. Ye, J. Humm, M. Gönen, H. Kalaigian, H. Schöder, H.W. Strauss, S.M. Larson, U. Wiesner, M.S. Bradbury, Clinical translation of an ultrasmall inorganic optical-PET imaging nanoparticle probe, Sci. Transl. Med. 6 (260) (2014) 260ra149.

[178] Y. Ishida, K. Nagata, Hsp47 as a collagen-specific molecular chaperone, Methods Enzymol. 499 (2011) 167–182.

[179] W. Ngamcherdtrakul, D.J. Castro, S. Gu, J. Morry, M. Reda, J.W. Gray, W. Yantasee, Current development of targeted oligonucleotide-based cancer therapies: perspective on HER2-positive breast cancer treatment, Cancer Treat. Rev. 45 (2016) 19–29.