Research Article

Synthesis, Characterization, and Antimicrobial Activity of Novel Sulfonated Copper-Triazine Complexes

Supun Katugampala,1 Inoka C. Perera, 2 Chandrika Nanayakkara, 3 and Theshini Perera 4

1Department of Chemistry, University of Sri Jayewardenepura, Nugegoda, Sri Lanka
2Department of Zoology and Environment Science, University of Colombo, Colombo, Sri Lanka
3Department of Plant Science, University of Colombo, Colombo, Sri Lanka

Correspondence should be addressed to Theshini Perera; theshi@sjp.ac.lk

Received 6 April 2018; Revised 5 June 2018; Accepted 8 July 2018; Published 29 August 2018

Academic Editor: Aurel Tabacaru

Copyright © 2018 Supun Katugampala et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Metallotriazine complexes possess interesting biological and medicinal properties, and the present study focuses on the synthesis, characterization, and antimicrobial activity of four novel copper-triazine derivatives in search of potent antibacterial and antifungal drug leads. In this study, 3-(2-pyridyl)-5,6-diphenyl-1,2,4-triazine-4,4′-disulfonic acid monosodium salt (L1, ferrozine) and 3-(2-pyridyl)-5,6-di(2-furyl)-1,2,4-triazine-5,5′-disulfonic acid disodium salt (L2, ferene) have been used as ligands to study the complexation towards copper(II). The synthesized complexes, \([\text{CuCl}_2(\text{ferrozine})]\cdot 7\text{H}_2\text{O} \cdot \text{MeOH} (1), [\text{CuCl}_2(\text{ferrozine})_2] \cdot 5\text{H}_2\text{O} \cdot \text{MeOH} (2), [\text{CuCl}_2(\text{ferene})] \cdot \text{H}_2\text{O} \cdot \text{MeOH} (3), \text{and} [\text{CuCl}_2(\text{ferene})_2] \cdot \text{H}_2\text{O} \cdot \text{MeOH} (4),\) have been characterized spectroscopically, and preliminary bioassays have been carried out. FTIR spectroscopic data have shown that \(\text{N} \equiv \text{N}\) and \(\text{C} \equiv \text{N}\) stretching frequencies of complexes have been shifted towards lower frequencies in comparison with that of the ligands, confirming new bond formation between Cu and N, which in turn lowers the strength of \(\text{N} \equiv \text{N}\) and \(\text{C} \equiv \text{N}\) bonds. In addition, a bathochromic shift has been observed for UV-visible spectra of complexes \((1), (2), (3), \text{and} (4).\) Furthermore, elemental analysis data have been useful to obtain empirical formulas of these complexes and to establish the purity of each complex. Complexes \((1) \text{and} (2)\) have shown antibacterial activity for both \(\text{S. aureus} (\text{ATCC}^\circledR 25923)\) and \(\text{E. coli} (\text{ATCC}^\circledR 25922) \) at 1 mg/disc concentration, and ferrozine has shown a larger inhibition zone against the clinical sample of \(\text{C. albicans} \) at 1 mg/disc concentration in comparison with the positive control, fluconazole.

1. Introduction

Transition metals have numerous and unique biological, chemical, and physical properties due to the availability of d electrons in valence shells. Much attention has been focused on copper complexes due to their various potential biological activities [1–4] out of which antimicrobial [5] and antiviral activities is paramount [6–15].

Since triazine is a well-known natural material which possesses many biological properties [16–21], it is not surprising that organometallic complexes of triazine with first row transition metals (Mn [22, 23], Co [24, 25], Ni [24, 25], Cu [22, 24–28], and Zn [25]), with second row transition metals (Ru [29], Pd [30], Ag [31], and Cd [32]), and with third row transition metals (Re [33] and Pt [34–36]) have been synthesized, and their activities explored as catalysts [37] and biological agents such as antibacterial [25], anticancer [29, 36], antifouling [24], antifungal [33], anti-HIV [35], antimicrobial [25], antiproliferative [26, 34], antiviral [28, 35], and DNA binding [26, 29, 30] agents.

Even though many reports exist of metal complexes of triazine derivatives as detailed above, metal complexes containing the pyridyl-1,2,4-triazine core are relatively unexplored. Platinum(II) complexes of sulphonated 2-pyridyl-1,2,4-triazine have been reported to possess anti-HIV activity [35]. A copper(II) complex bearing 2,4,6-tris(2-pyridyl)-1,3,5-triazine ligand has been reported to bind DNA in a moderately strong way exhibiting significantly better anticancer activity against breast cancer in comparison with cisplatin [26]. An octahedral complex of rhenium(V), ML1L2L3L4
(where L1 = o xo, L2 = chloride, L3 = triphenylphosphine, and L4 = 3-hydravino-5,6-diphenyl-1,2,4-triazine), has shown comparable antifungal activity against *Alternaria alternata* and *Aspergillus niger* [33]. We ourselves have explored the potential of using rhenium complexes of ferene and ferrozone (Figure 1) as biological imaging agents [38]. In our most recent work, we have commented on the possible use of the scaffold of sulfonated pyridyl triazine complexes being utilized as serum albumin transporters [39]. As such, it seems prudent to now explore its binding towards copper.

Thus, the current study explores the synthesis of four novel water-soluble complexes of the type, MLnCl2 (Figure 2) where M = Cu²⁺, L = 3-(2-pyridyl)-5,6-diphenyl-1,2,4-triazine-4,4'-disulfonic acid sodium salt/3-(2-pyridyl)-5,6-di(2-furyl)-1,2,4-triazine-5,5'-disulfonic acid disodium salt, and n = 1/2, their chemical characterization, and preliminary tests to assess antimicrobial activity of above synthesized complexes as well as of the ligands.

2. Experimental

2.1. Materials Used. All chemicals and reagents used for the synthesis were commercially available and used without further purification. 3-(2-Pyridyl)-5,6-diphenyl-1,2,4-triazine-4,4'-disulfonic acid sodium salt (ferrozine), 3-(2-pyridyl)-5,6-di(2-furyl)-1,2,4-triazine-5,5'-disulfonic acid disodium salt (ferene), and methanol ACS reagent (assay ≥99.8%) were purchased from Sigma-Aldrich, and copper (II) chloride dihydrate was purchased from Research-Lab Fine Chem Industries. Mueller-Hinton agar was purchased from Hardy Diagnostics, USA. Sodium chloride, sodium hydroxide, and dextrose were purchased from HiMedia Laboratories. The bacteria were obtained by the Industrial Technology Institute, Colombo.

2.2. Instrumentation. Elemental analysis was carried out on PerkinElmer 2400 Series II CHNS/O Elemental Analyzer at Atlantic Microlabs, USA. IR spectra were recorded using Thermo Scientific NICOLET iS10 spectrophotometer in the spectral range 4000–650 cm⁻¹ for both ligands and complexes. Thermo Spectronic Helios alpha UV-Vis double-beam spectrophotometer was used to measure the absorbance in the range of 190–1100 nm, and baseline correction was performed using matched quartz cuvettes. High-resolution mass spectra were recorded on an Agilent 6210 ESI TOF LCMS mass spectrometer.

2.3. Synthesis

2.3.1. Preparation of [CuCl2(ferrozine)]·7H2O-MeOH (1). A solution of ferrozine (0.25 mmol, 0.1269 g) in methanol (8.0 cm³) was added to copper chloride dihydrate (0.25 mmol, 0.0435 g) in methanol (2.0 cm³). Then the resulting mixture was stirred for 2 hours at room temperature and progression of reaction checked using TLC. A light green colour crystalline precipitate was obtained after 2 days and collected by filtration (yield: 0.1264 g, 64%). IR (ATR; ν/cm⁻¹): 1596.84 (m) and 1498.22 (s), νC=O and νN=N. UV-Vis (MeOH; λmax [nm]): 205, 242, 298, and 327. Anal. Calc. for C20H13Cl2CuN4NaO6S2 · 7H2O: C, 32.12; H, 3.98; N, 7.14. Found: C: 31.68%, H: 3.80%, and N: 7.42%. ESI-MS (m/z): [M−H] calc’d for C20H13Cl2CuN4NaO6S2 · 7H2O, [M−H] calc’d found, 565.9179; found, 565.9188.

2.3.2. Preparation of [CuCl2(ferene)]·5H2O-MeOH (2). A procedure similar to that given above was followed using copper chloride dihydrate (0.25 mmol, 0.0435 g) and ferrozine (0.50 mmol, 0.2538 g). The resulting mixture was stirred for 5 hours. A dark green colour crystalline precipitate was obtained after 2 days and collected by filtration (yield: 0.1937 g, 75%). IR (ATR; ν/cm⁻¹): 1595.66 (m) and 1498.50 (s), νC=O and νN=N. UV-Vis (MeOH; λmax [nm]): 213, 240, 301, and 334. Anal. Calc. for C40H26Cl2Cu2N4Na2O12S4 · 5H2O: C, 39.66; H, 3.25; N, 9.03. Found: C: 39.29%, H: 3.76%, N: 9.23%. ESI-MS (m/z): [M−H] calc’d for C40H26Cl2Cu2N4Na2O12S4 · 5H2O, [M−H] calc’d found, 999.9833; found, 999.9776.

2.3.3. Preparation of [CuCl2(ferene)]·7H2O-MeOH (3). A solution of ferene (0.25 mmol, 0.1269 g) in methanol (8.0 cm³) was added to copper chloride dihydrate (0.25 mmol, 0.0435 g) in methanol (2.0 cm³). Then the resulting mixture was stirred for 6 hours at room temperature and progression of reaction checked using TLC technique initially and at the end. A yellow colour crystalline precipitate was obtained after 1 day and collected by filtration (yield: 0.1183 g, 75%). IR (ATR; ν/cm⁻¹): 1567.49 (m) and 1499.15 (s), νC=O and νN=N. UV-Vis (MeOH; λmax [nm]): 202, 239, 338, and 371. Anal. Calc. for C16H8Cl2CuN4O6S2 · H2O·CH3OH: C, 32.16; H, 2.54; N, 8.82.
2.3.4. Preparation of \([\text{CuCl}_2(\text{ferene})_2]\)·H\(_2\)O·MeOH (4). A procedure similar to above was followed using copper chloride dihydrate (0.25 mmol, 0.0435 g) and ferene (0.50 mmol, 0.2472 g). The resulting mixture was stirred for 5 hours. A brown-yellow colour crystalline precipitate was obtained after 1 day and collected by filtration (yield: 0.1912 g, 65%). IR (ATR; \(\nu/\text{cm}^{-1}\)): 1569.82(m) and 1494.40(s), \(\nu_{\text{C-N}}\) and \(\nu_{\text{N-N}}\). UV-Vis (MeOH; \(\lambda_{\text{max}}\) [nm]): 208, 246, 338 and 371. Anal. Calc. for \(\text{C}_{32}\text{H}_{16}\text{Cl}_2\text{CuN}_4\text{O}_{16}\text{S}_4\cdot\text{H}_2\text{O}\cdot\text{CH}_3\text{OH}\): C, 33.78; H, 1.89; N, 9.56. Found: C: 33.76%, H: 2.42%, N: 9.58%.

2.4. Antimicrobial Assay. Compounds were tested against Gram-positive \textit{Staphylococcus aureus ATCC\textsuperscript{*} 25923} and Gram-negative \textit{Escherichia coli ATCC\textsuperscript{*} 25922} bacterial species and a clinical isolate of \textit{Candida albicans} as a fungal species. Antimicrobial assay was performed by a standard disk diffusion assay [40] where the inhibition zones were

---

**Figure 2: Synthetic routes for ML\(_1\)Cl\(_2\) (complex (1)) (i), M(L\(_1\))\(_2\)Cl\(_2\) (complex (2)) (ii), ML\(_2\)Cl\(_2\) (complex (3)) (iii), and M(L\(_2\))\(_2\)Cl\(_2\) (complex (4)) (iv) complexes. NB: L\(_1\) = 3-(2-pyridyl)-5,6-diphenyl-1,2,4-triazine-\(p\',p''\)-disulfonic acid monosodium salt; L\(_2\) = 3-(2-pyridyl)-5,6-di(2-furyl)-1,2,4-triazine-5,5'-disulfonic acid disodium salt. Solvent molecules in complexes (1)–(4) have been omitted for clarity. Molar ratios of reactants: (i) \(\text{CuCl}_2:\text{L}_1 = 1:1\), (ii) \(\text{CuCl}_2:\text{L}_1 = 1:2\), (iii) \(\text{CuCl}_2:\text{L}_2 = 1:1\), and (iv) \(\text{CuCl}_2:\text{L}_2 = 1:2\).**
measured and expressed as a mean of three replicates. Gentamycin and flucanazole were used as positive controls, and methanol was used as the negative control.

3. Results and Discussion

3.1. Synthesis. Copper chloride and the relevant ligands were used in 1:1 and 1:2 ratios to synthesize the desired metal complexes (Figure 2). Thin-layer chromatography (TLC) was initially used to monitor the progress of reaction, and visualization of spots was done using an iodine bath.

3.2. FTIR Analysis. FTIR data were recorded for dried crystals of ligands and complexes (1)–(4), and literature values were utilized where relevant [41]. The stretching frequency of the pyridine ring ($\nu_{\text{C-N}}$) and stretching frequency of the triazine ring ($\nu_{\text{N-N}}$) are considered mostly, because their values change upon formation of new bonds serving as good indicators of complex formation.

Stretching frequencies of $\text{N-N}$ and $\text{C-N}$ in complexes (1) and (2) have shifted to lower frequencies as expected, compared to those values of the free ferrozine ligand, due to $\sigma$ donation of N lone pair which lowers strength of $\text{N-N}$ and $\text{C-N}$ bonds (Table 1). Furthermore, a broad band around 3400–3300 cm$^{-1}$ was observed due to OH groups from methanol or water.

Similarly, stretching frequencies of $\text{N-N}$ and $\text{C-N}$ in complexes (3) and (4) were observed at lower frequencies in comparison with those of the free ferrozine ligand (Table 1), and a broad band was observed around 3400–3300 cm$^{-1}$ due to OH groups of solvent.

3.3. UV-Visible Spectroscopy. UV-Vis spectra of reactants and complexes (1, 2, 3, and 4) were recorded in methanol at room temperature (Figure 3, Table S1, Supplementary Materials). The absorption wavelengths of complexes (1)–(4) have shifted towards longer wavelengths (bathochromic shift) compared to the wavelengths of the reactants (copper, ferrozine, and ferene). Both ferrozine and ferene have aromatic ring systems, and $\pi-\pi^*$ transitions are thus possible [42]. These results are in agreement with those previously reported for zinc complexes of ferene and ferrozine [39] where a bathochromic shift was observed for both mono and bis complexes in comparison with that of the free ligand.

3.4. Elemental Analysis. Empirical formulas related to experimental values aided in obtaining the exact molecular formulas of all four complexes (Table 2). It can be seen that experimental values are within $\pm 0.4\%$ of expected values indicating purity of the synthesized complexes.

3.5. Antimicrobial Activity. All four complexes and ligands were studied in vitro for their antimicrobial activity against Gram-positive *Staphylococcus aureus* ATCC® 25923 and negative bacteria *Escherichia coli* ATCC® 25922 as well as the unicellular fungal species, *Candida albicans*. Inhibition zones were obtained by adding a concentration of 1 mg/disc, and the diameters of the zones are given in Table 3 for bacteria and Table 4 for fungi.

### Table 1: FTIR data comparison chart of complexes (1)–(4) in comparison with those of free ligands.

| Ligand     | $\nu_{\text{C-N}}$ (cm$^{-1}$) | $\nu_{\text{N-N}}$ (cm$^{-1}$) |
|------------|-------------------------------|-------------------------------|
| Ferrozine  | 1608                          | 1503                          |
| Complex (1)| 1596                          | 1498                          |
| Complex (2)| 1595                          | 1498                          |
| Ferene     | 1589                          | 1507                          |
| Complex (3)| 1567                          | 1499                          |
| Complex (4)| 1570                          | 1494                          |

### Table 2: Elemental analysis data of complexes.

| Complex | Value   | C (%) | H (%) | N (%) |
|---------|---------|-------|-------|-------|
| (1)     | Calculated | 32.12 | 3.98  | 7.14  |
|         | Experimental | 31.68 | 3.80  | 7.42  |
| (2)     | Calculated | 39.66 | 3.25  | 9.03  |
|         | Experimental | 39.29 | 3.76  | 9.23  |
| (3)     | Calculated | 39.16 | 2.54  | 8.82  |
|         | Experimental | 39.12 | 2.76  | 9.29  |
| (4)     | Calculated | 33.78 | 1.89  | 9.55  |
|         | Experimental | 33.76 | 2.42  | 9.58  |

Figure 3: UV-visible spectra recorded in methanol of ferrozine, complexes (1) and (2) (a) and ferene, complexes (3) and (4) (b).
Analysis of the inhibition zone diameter revealed that only complex (1) and complex (2) show moderate antibacterial activity when compared to the positive control. It is interesting to see that ferrozine ligand demonstrates antifungal activity.

Antimicrobial activity reported here is of moderate value. Further studies are warranted to optimize this system for greater activity.

4. Conclusions

We have described the synthesis of four novel water-soluble copper complexes bearing sulfonated pyridyl triazine ligands. FTIR spectroscopic data have confirmed the existence of Cu-N bonds in all four complexes because stretching frequencies of N–N and C=N complexes have been shifted towards lower frequencies in comparison with that of the ligands. In UV-Vis spectra, a bathochromic shift has been observed for complexes (1)–(4). Furthermore, elemental analysis data have been useful to obtain empirical formulas of these complexes and to establish the purity of each complex.

Preliminary bioassays in antimicrobial activity showed moderate antibacterial activity with complexes (1) and (2) whereas ferrozine showed antifungal activity against Candida albicans. To the best of our knowledge, we are the first to report on the antifungal activity of ferrozine. These findings provide a potential lead for antimicrobial drug development.

Data Availability

The data used to support the findings of this study are included within the article and within the Supplementary Information file.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

Financial assistance from the University of Sri Jayewardenepura (Grant no. ASP/01/RE/SCI/2015/19) is gratefully acknowledged. Clinical sample of Candida albicans was provided by the Department of Microbiology, Faculty of Medicine, University of Colombo. An earlier version of this work with two of the complexes detailed here was presented as an abstract at Chemistry in Sri Lanka 2015 as found in the link http://www.ichemc.edu.lk/wp-content/uploads/2015/10/vol-31-no.-2.pdf. The authors also acknowledge the Centre for Advanced Material Research of the University of Sri Jayewardenepura.

Supplementary Materials

Table S1: comparison of UV-Vis data of ferrozine, ferene, and complexes (1)–(4) is presented in a tabulated form in Supplementary Materials. (Supplementary Materials)

References

[1] A. Valenti, M. Melnik, D. Hudecová, B. Dudová, R. Kivekás, and M. R. Sundberg, “Copper(II) salicylidenediiminate complexes as potential antimicrobial agents,” Inorganica Chimica Acta, vol. 340, pp. 15–20, 2002.
[2] M. Ibrahim, F. Wang, M. M. Lou et al., “Copper as an antibacterial agent for human pathogenic multidrug resistant Burkholderia cepacia complex bacteria,” Journal of Bioscience and Bioengineering, vol. 112, no. 6, pp. 570–576, 2011.
[3] A. Latif Abuhijleh and C. Woods, "Synthesis, characterization, and oxidase activities of copper(II) complexes of the anticonvulsant drug valproate," Journal of Inorganic Biochemistry, vol. 64, no. 1, pp. 55–67, 1996.
[4] B. S. Creaven, B. Duff, D. A. Egan et al., “Anticancer and antifungal activity of copper(II) complexes of quinolin-2-(H)-one-derived Schiff bases,” Inorganica Chimica Acta, vol. 363, no. 14, pp. 4048–4058, 2010.
[5] H. I. Cervantes, J. A. Alvarez, J. M. Munoz, V. Arreguin, J. L. Mosqueda, and A. E. Macias, “Antimicrobial activity of copper against organisms in aqueous solution: a case for copper-based water pipelines in hospitals?”, American Journal of Infection Control, vol. 41, no. 12, pp. e115–e118, 2013.
[6] N. Shionoiri, T. Sato, Y. Fujimori et al., “Investigation of the antiviral properties of copper iodide nanoparticles against feline calicivirus,” Journal of Bioscience and Bioengineering, vol. 113, no. 5, pp. 580–586, 2012.
[7] G. Betanzos-Cabrera, F. J. Ramirez, J. L. Munoz, B. L. Barron, and R. Maldonado, “Inactivation of HSV-2 by ascorbate-Cu (II) and its protecting evaluation in CF-1 mice against encephalitis,” Journal of Virological Methods, vol. 120, no. 2, pp. 161–165, 2004.
[8] J. L. Sagripanti, L. B. Routson, A. C. Bonifacino, and C. D. Lyle, “Mechanism of copper-mediated inactivation of herpes simplex virus,” Antimicrobial Agents and Chemotherapy, vol. 41, no. 4, pp. 812–817, 1997.
[9] L. A. White, C. Y. Freeman, R. D. Forrester, and W. A. Chappell, “In vitro effect of ascorbic acid on infectivity of herpesviruses and paramyxoviruses,” Journal of Clinical Microbiology, vol. 24, no. 4, pp. 527–531, 1986.
[10] G. Borkow, S. S. Zhou, T. Page, and J. Gabbay, “A novel antinfluenza copper oxide containing respiratory face mask,” PLoS One, vol. 5, no. 6, p. e11295, 2010.
[11] M. Horie, H. Ogawa, Y. Yoshida et al., "Inactivation and morphological changes of avian influenza virus by copper ions," Archives of Virology, vol. 153, no. 8, pp. 1467–1472, 2008.

[12] J. O. Noyce, H. Michels, and C. W. Keevil, "Inactivation of influenza A virus on copper versus stainless steel surfaces," Applied and Environmental Microbiology, vol. 73, no. 8, pp. 2748–2750, 2007.

[13] J. I. Nieto-Juarez, K. Pierzchla, A. Sienkiewicz, and T. Kohm, "Inactivation of MS2 coliphage in Fenton and Fenton-like systems: role of transition metals, hydrogen peroxide and sunlight," Environmental Science & Technology, vol. 44, no. 9, pp. 3351–3356, 2010.

[14] M. T. Yahya, T. M. Straub, and C. P. Gerba, "Inactivation of coliphage MS-2 and poliovirus by copper, silver, and chloride," Canadian Journal of Microbiology, vol. 38, no. 5, pp. 430–435, 1992.

[15] F. X. Abad, R. M. Pinto, J. M. Diez, and A. Bosch, "Disinfection of human enteric viruses in water by copper and silver in combination with low levels of chlorine," Applied and Environmental Microbiology, vol. 60, no. 7, pp. 2377–2383, 1994.

[16] R. Menicagli, S. Samaritani, G. Signore, F. Vaglini, and L. Dalla Via, "In vitro cytotoxic activities of 2-alkyl-4,6-diheteroaryl-1,3,5-triazines: new molecules in anticancer research," Journal of Medicinal Chemistry, vol. 47, no. 19, pp. 4649–4652, 2004.

[17] S. Melato, D. Prosperi, P. Coghi, N. Basilico, and D. Monti, "A combinatorial approach to 2,4,6-trisubstituted triazines with potent antimalarial activity: combining conventional synthesis and microwave-assistance," ChemMedChem, vol. 3, no. 6, pp. 873–876, 2008.

[18] C. Zhou, J. Min, Z. Liu et al., "Synthesis and biological evaluation of novel 1,3,5-triazine derivatives as antimicrobial agents," Bioorganic & Medicinal Chemistry Letters, vol. 18, no. 4, pp. 1308–1311, 2008.

[19] K. Srinivas, U. Srinivas, K. Bhanuprakash, K. Harakishore, U. S. Murthy, and V. J. Rao, "Synthesis and antibacterial activity of various substituted s-triazines," European Journal of Medicinal Chemistry, vol. 41, no. 11, pp. 1240–1246, 2006.

[20] A. Baliani, G. J. Bueno, M. L. Stewart et al., "Design and synthesis of a series of melamine-based nitroheterocycles with activity against trypanosomatid parasites," Journal of Medicinal Chemistry, vol. 48, no. 17, pp. 5570–5579, 2005.

[21] Y. Z. Xiong, F. E. Chen, J. Balzarini, E. De Clercq, and C. Pannecoque, "Non-nucleoside HIV-1 reverse transcriptase inhibitors. Part 11: structural modifications of diaryl-triazines with potent anti-HIV activity," European Journal of Medicinal Chemistry, vol. 43, no. 6, pp. 1230–1236, 2008.

[22] J. G. Malecki, B. Machura, and A. Szwilicka, "X-ray studies, spectroscopic characterisation and DFT calculations for Mn(II), Ni(II) and Cu(II) complexes with 5,6-diphenyl-3-(2-pyridyl)1,2,4-triazine," Structural Chemistry, vol. 22, no. 1, pp. 77–87, 2010.

[23] M. M. Najafoor, D. M. Boghaei, and V. McKee, "Synthesis, characterization, crystal structure and oxygen-evolution activity of a manganese(II) complex with 2,4,6-tris (2-pyridyl)-1,3,5-triazine," Polychron, vol. 29, no. 17, pp. 3246–3250, 2010.

[24] H. A. E. Hemaia, A. A. E. Dissouky, and S. M. Sadek, "Potential antifouling agents: copper, cobalt, and nickel complexes of 3-(2-acetyl pyridylidene) hydrazino-5,6-diphenyl-1,2,4-triazine," Egyptian Journal of Aquatic Research, vol. 31, pp. 45–56, 2005.

[25] K. Singh, Y. Kumar, P. Puri, C. Sharma, and K. R. Aneja, "Antimicrobial, spectral and thermal studies of divalent cobalt, nickel, copper and zinc complexes with triazole Schiff bases," Arabian Journal of Chemistry, vol. 10, pp. S978–S987, 2017.

[26] K. Abdi, H. Hadadzadeh, M. Salimi, J. Simpson, and A. D. Khalaji, "A mononuclear copper(II) complex based on the poly(dimethylglyoximato) ligand 2,4,6-tris(2-pyridyl)-1,3,5-triazine (tpzt), [Cu(tpzt)2]2+: X-ray crystal structure, DNA binding and in vitro cell cytotoxicity," Polyhedron, vol. 44, no. 1, pp. 101–112, 2012.

[27] B. Machura, A. Skwolka, R. Kruszynski, J. Mrzoinski, J. Klak, and J. Kusz, "Coordination studies of 5,6-diphenyl-3-(2-pyridyl)-1,2,4-triazine towards Cu2+ cation. X-ray studies, spectroscopic characterization and DFT calculations," Polyhedron, vol. 27, no. 13, pp. 2959–2967, 2008.

[28] C. G. Palivan, H. M. N. Palivan, B. A. Goodman, and C. Cristescu, "ESR study of some asymmetric-triazine copper (II) complexes having high antiviral activity," Applied Magnetic Resonance, vol. 15, no. 3–4, pp. 477–488, 1998.

[29] N. Busto, J. Valladolid, M. Martinez-Alonso et al., "Anticancer activity and DNA binding of a bifunctional Ru(II) arene aqua-complex with the 2,4-diamino-6-(2-pyridyl)-1,3,5-triazine ligand," Inorganic Chemistry, vol. 52, no. 17, pp. 9962–9974, 2013.

[30] W. H. Al-Assy and M. M. Mostafa, "Comparative studies and modeling structures of two new isomers containing binuclear PdII complexes derived from 2,4,6-tri-(2-pyridyl)-1,3,5-triazine (TPTZ)," Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, vol. 120, pp. 568–573, 2014.

[31] M. M. Najafoor, M. Holyiska, M. Amini, S. H. Kazemi, T. Lis, and M. Bagherzadeh, "Two new silver(I) complexes with 2,4,6-tris(2-pyridyl)-1,3,5-triazine (tpzt): preparation, characterization, crystal structure and alcohol oxidation activity in the presence of oxone," Polyhedron, vol. 29, no. 14, pp. 2837–2843, 2010.

[32] F. Marandi, M. Jangholi, M. Hakimi, H. A. Rudbari, and G. Bruno, "Synthesis and crystal structures of the first cadmium complexes of 3,5,6-tris-(2-pyridyl)-1,2,4-triazine ligand," Journal of Molecular Structure, vol. 1036, pp. 71–77, 2013.

[33] M. M. Mashaly, H. F. El-Shafiy, S. B. El-Maraghy, and H. A. Habib, "Synthesis, properties and thermal studies of oxorhenium(V) complexes with 3-hydrazino-5,6-diphenyl-1,2,4-triazine, benzimidazolothione and 2-hydrazinobenzimidazole. Mixed ligand complexes, pyrolytical products and biological activity," Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, vol. 61, no. 8, pp. 1853–1869, 2005.

[34] I. Lakomska, B. Golankiewicz, J. Wietrzyk et al., "Synthesis, spectroscopical characterization and the biological activity in vitro of new platinum(II) complexes with imidazo[1,5-a]-1,3,5-triazine derivatives and dimethylsulfoxide," Inorganic Chemica Acta, vol. 338, no. 6, pp. 1911–1917, 2005.

[35] A. N. Vzorov, D. Bhattacharyya, L. G. Marzilli, and D. G. Truhlar, "Density functional theory for biomolecular and biomolecular spectroscopy," Computers in Chemistry, vol. 29, no. 1, pp. 130–131, 2005.

[36] C. J. Cramer and D. G. Truhlar, "Density functional theory for transition metals and transition metal chemistry," Physical Chemistry Chemical Physics, vol. 11, no. 46, pp. 10757–10816, 2009.
[38] K. Ranasinghe, S. Handunnetti, I. C. Perera, and T. Perera, "Synthesis and characterization of novel rhenium(I) complexes towards potential biological imaging applications," *Chemistry Central Journal*, vol. 10, no. 1, p. 71, 2016.

[39] N. Abeydeera, I. C. Perera, and T. Perera, "Synthesis, characterization, and BSA-binding studies of novel sulfonated zinc-triazine complexes," *Bioinorganic Chemistry and Applications*, vol. 2018, Article ID 7563820, 7 pages, 2018.

[40] Institute CaLS, "Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard—seventh edition," in *M7–A7*, Clinical and Laboratory Standards Institute, Wayne, PA, USA, 2006.

[41] V. Béreau and J. Marrot, " Coordination studies of 5,6-diphenyl-3-(2-pyridyl)-1,2,4-triazine towards Zn$^{2+}$ cation. Synthesis and characterization by X-ray diffraction and spectroscopic methods," *Comptes Rendus Chimie*, vol. 8, no. 6-7, pp. 1087–1092, 2005.

[42] D. Eastwood, R. L. Lidberg, and M. S. Dresselhaus, "Ultraviolet-visible fluorescence spectroscopy of selected polyaromatic hydrocarbons and organometallics on hexagonal graphite and boron nitride," *Chemistry of Materials*, vol. 6, no. 2, pp. 211–215, 1994.