Chapter 14
Simulation of Preterm Neonatal Brain Metabolism During Functional Neuronal Activation Using a Computational Model

T. Hapuarachchi, F. Scholkmann, M. Caldwell, C. Hagmann, S. Kleiser, A.J. Metz, M. Pastewski, M. Wolf, and I. Tachtsidis

Abstract We present a computational model of metabolism in the preterm neonatal brain. The model has the capacity to mimic haemodynamic and metabolic changes during functional activation and simulate functional near-infrared spectroscopy (fNIRS) data. As an initial test of the model’s efficacy, we simulate data obtained from published studies investigating functional activity in preterm neonates. In addition we simulated recently collected data from preterm neonates during visual activation. The model is well able to predict the haemodynamic and metabolic changes from these observations. In particular, we found that changes in cerebral blood flow and blood pressure may account for the observed variability of the magnitude and sign of stimulus-evoked haemodynamic changes reported in preterm infants.

Keywords Mathematical model • fNIRS • Haemodynamics • Autoregulation • Stimulus – evoked functional response

This chapter was originally published under a CC BY-NC 4.0 license, but has now been made available under a CC BY 4.0 license. An erratum to this chapter can be found at DOI 10.1007/978-1-4939-3023-4_66.

T. Hapuarachchi
CoMPEX, University College London, London, UK
Department of Medical Physics and Bioengineering, University College London, London, UK
e-mail: t.hapuarachchi@ucl.ac.uk
F. Scholkmann • S. Kleiser • A.J. Metz • M. Pastewski • M. Wolf
Division of Neonatology, Biomedical Optics Research Laboratory, University Hospital Zurich, Zurich, Switzerland
M. Caldwell • I. Tachtsidis
Department of Medical Physics and Bioengineering, University College London, London, UK
C. Hagmann
Clinic of Neonatology, University Hospital Zurich, Zurich, Switzerland

© The Author(s) 2016
C.E. Elwell et al. (eds.), Oxygen Transport to Tissue XXXVII, Advances in Experimental Medicine and Biology 876, DOI 10.1007/978-1-4939-3023-4_14
1 Introduction

Our research focuses on the development of a family of computational models of cerebral metabolism, primarily to investigate the effects of stimuli and physiological insults, and to inform the clinical treatment of brain injury. This work has so far centred on human adult [1] and piglet cerebral activity [2]. We have recently extended our focus to the preterm neonatal brain.

A number of studies investigating functional activity in neonates using functional near infrared spectroscopy (fNIRS) have observed different haemodynamic responses. Inconsistent results have been reported in literature regarding the characteristics of stimulus-evoked changes (i.e. magnitude and sign) in oxyhaemoglobin (HbO₂) and deoxyhaemoglobin (HHb). In particular, some studies report a decrease in HHb (an adult-like response) while others report the opposite. In order to research the mechanisms of these responses we have adapted an existing model of adult cerebral metabolism (BrainSignals) [1] to the preterm neonatal brain. In this paper, we (1) present a model of metabolism and haemodynamics in the preterm brain, (2) use the model to simulate observations of two published preterm functional response studies and (3) use the model to predict recently collected data from a stimulus-evoked haemodynamic response study in preterm neonates.

2 Modelling Functional Activation in the Developing Preterm Brain

The original BrainSignals model simulates blood circulation and energy metabolism. It uses a combination of differential equations and algebraic relations to mimic biochemical reactions and processes in a brain cell and the immediate vasculature. The model predicts in particular responses to changes in arterial blood pressure, oxygenation, carbon dioxide levels and functional activation. Figure 14.1 illustrates a simple schematic of the model. This model was adapted to the human neonate by altering a number of physiological parameters known to be significantly different in the young (a method similar to that employed in developing the piglet model [2]). These parameters are listed in Table 14.1. In particular, the reduction of normal arterial blood pressure (BP) was seen to have a significant effect on the behaviour of the model. Figure 14.2a shows the autoregulation curve of the adult model and the neonatal model for preterm neonates, comparable to approximations found in
In order to simulate functional activation, the model uses a dimensionless parameter $u$ which represents demand. A change in $u$ produces a response in vascular smooth muscle and affects ATP production by influencing the driving force for complex V in mitochondria.

Fig. 14.1 A simple schematic of the model. Model inputs are blood pressure, arterial oxygenation saturation, partial pressure of arterial carbon dioxide and functional activation.
Table 14.1 BrainSignals parameters modified to represent the preterm neonatal brain

| Parameter      | Description                                                      | Units               | BrainSignals | Preterm neonate | Source |
|----------------|------------------------------------------------------------------|---------------------|--------------|-----------------|--------|
| \( \text{CBF}_n \) | Normal cerebral blood flow (CBF)                                 | ml 100 g\(^{-1}\) min\(^{-1}\) | 49           | 19.8            | [3]    |
| \( [\text{CCO}]_{tiss} \) | Total concentration of cytochrome-c-oxidase (CCO) in tissue     | \( \mu \text{M} \) | 5.5          | 2.2             | [4, 5] |
| \( \text{Cu}_{A, \text{frac}, n} \) | Normal fraction of oxidised CCO                                  |                     | 0.8          | 0.67            | [5]    |
| \( \text{CMRO}_2,n \) | Normal cerebral metabolic rate of oxygen consumption (CMRO\(_2\)) | \( \mu \text{mol} 100 \text{ g}\(^{-1}\) min\(^{-1}\) | 155          | 40.865          | [6]    |
| \( P_a \) and \( P_{a,n} \) | Mean arterial blood pressure                                    | mmHg                | 100          | 30              | [7]    |
| \( [\text{Hbtot}] \) and \( [\text{Hbtot}]_n \) | Total concentration of haemoglobin in blood                     | mM                   | 9.1          | 9.75            | [8]    |
| \( V_{\text{blood}, n} \) | Normal brain blood fraction                                      |                     | 0.04         | 0.0233          | [9]    |
| \( P_{ic} \) and \( P_{ic,n} \) | Intracranial pressure                                           | mmHg                | 9.5          | 5.1             | [10]   |

Fig. 14.2 (a) Autoregulation curve—cerebral blood flow (CBF) against blood pressure—for the adult BrainSignals model and the preterm neonate model. (b) Demand as a model input, using a haemodynamic response function, to simulate functional activation

3 Model Simulations

Kozberg et al. [12] conducted a study in postnatal rats age-equivalent to human newborns. Although our model is focused on the human neonate, we were interested in replicating the qualitative response observed in the animals as they were subjected to a somatosensory stimulus. The rats which exhibited a rise in BP in response to the stimulus also showed an increase in HbO\(_2\) and total haemoglobin
(HbT) and a decrease in HHb, where the increase in HbO₂ was greater than the decrease in HHb (functional hyperemia). However, some rats showed a slight decrease or no change in BP. In these animals, the opposite results were observed—a decrease in HbO₂ and HbT and a rise in HHb. These conflicting results were attributed to a lack of functional hyperemia and an overarching effect of arterial vasoconstriction in the latter group. In order to model these results, we increased the model’s demand parameter $u$ to simulate functional activation with the shape of a steep rising haemodynamic response function (HRF). The demand was calculated as $u = 1.0 + \alpha \text{ HRF}$ where $\alpha$ is a real number (Fig. 14.2b). We simulated BP also using the HRF ($P_a = P_{a,n} - \beta \text{ HRF}$). The values for $\alpha$ and $\beta$ were optimised to achieve the best fit of our model simulations to the observed results. Arterial oxygen saturation ($\text{SaO}_2$) and partial pressure of carbon dioxide ($\text{PaCO}_2$) were assumed to remain constant. The model is able to predict changes in haemodynamics ($\Delta\text{HbO}_2$, $\Delta\text{HHb}$, $\Delta\text{HbT}$) which we compare against experimental data. The model is also capable of simulating cerebral blood flow (CBF) and the cerebral metabolic rate of oxygen consumption (CMRO₂). For the first group of animals who showed an increase in blood pressure, we used $\alpha = 2$ and $\beta = 1.5$. Although we attempted to include vasoconstriction as observed in the study, we found that preventing the dilation of the vessels reversed our results. For the second group we reduced the arterial radius by 1% ($r = r_n \times (1 - 0.01 \text{ HRF})$) and used $\beta = 0.18$. We did not need to increase the demand $u$, suggesting that an increase in oxygen consumption was not required to produce this response. Figures 14.3 and 14.4 illustrate these results, which are very closely comparable to the original data obtained by Kozberg et al. [12].

The second study we investigated was conducted in preterm human neonates by Roche-Labarbe et al. [13]. They observed a decrease in HHb and an increase in HbO₂, CBF, CMRO₂ and cerebral blood volume (CBV) in response to a somatosensory stimulus. We were able to replicate these results relatively well by simply increasing the model’s demand ($\alpha = 0.5$) (Fig. 14.5). However, our predicted CBF and CMRO₂ was higher and HHb slightly lower than that observed.

![Fig. 14.3](image-url) Model simulated and observed haemodynamic response of the Kozberg et al. study [12], investigating functional response in rats, with an increase in demand and blood pressure (BP). (a) Changes in deoxy- and oxy-haemoglobin (HHb, HbO2) concentrations. (b) Changes in BP and total haemoglobin (HbT)
We also simulated data from a functional near infrared spectroscopy (fNIRS) study conducted at University Hospital Zurich (USZ) which investigates the functional response in the preterm brain evoked by a visual stimulus. A blinking pocket LCD display was used as the stimulus and changes in tissue oxygenation and haemodynamics were measured over the prefrontal cortex using a novel spatially-resolved NIRS device (OxyPrem). Measurements of changes in HbO2, HHb and HbT were averaged over repeated stimuli. Characteristics of two preterm subjects are detailed in Table 14.2.

**Fig. 14.4** Model simulated and observed haemodynamic response of the Kozberg et al. study [12], investigating functional response in rats, with a slight decrease in arterial radius and blood pressure (BP). (a) Changes in deoxy- and oxy- haemoglobin (HHb, HbO2) concentrations. (b) Changes in BP and total haemoglobin (HbT)

**Fig. 14.5** Model simulated and observed haemodynamic response of the Roche-Labarbe et al. study [13], investigating functional response in human preterm neonates, with an increase in demand. (a) Relative changes in oxy- and deoxy- haemoglobin (rHbO2, rHHb), (b) cerebral blood volume (rCBV) and cerebral blood flow (rCBF) and (c) cerebral metabolic rate of oxygen consumption (rCMRO2)

We also simulated data from a functional near infrared spectroscopy (fNIRS) study conducted at University Hospital Zurich (USZ) which investigates the functional response in the preterm brain evoked by a visual stimulus. A blinking pocket LCD display was used as the stimulus and changes in tissue oxygenation and haemodynamics were measured over the prefrontal cortex using a novel spatially-resolved NIRS device (OxyPrem). Measurements of changes in HbO2, HHb and HbT were averaged over repeated stimuli. Characteristics of two preterm subjects are detailed in Table 14.2.
|                | Gestational age (weeks) | Actual age (weeks) | Weight (g) | Haematocrit (%) | Haemoglobin (g/dL) | FiO2 (%) | Baseline SpO2 (%) | Baseline heart rate (BPM) |
|----------------|-------------------------|--------------------|------------|-----------------|-------------------|----------|------------------|--------------------------|
| Neonate 1      | 33.3                    | 33.4               | 2380       | 52.5            | 17.08             | 21.0     | 95               | 148                      |
| Neonate 2      | 25.9                    | 39.0               | 3460       | 30.0            | 9.70              | 28.0     | 92–95            | 165                      |
We assumed that the \(\text{SaO}_2\), \(\text{PaCO}_2\) and BP remain constant in the first instance. In Neonate 1, the measurements showed an increase in \(\Delta\text{HbO}_2\) and \(\Delta\text{HbT}\) during the stimulus, and a decrease in \(\Delta\text{HHb}\) (Fig. 14.6). We were able to simulate this response in our model by a simple increase in demand (\(\alpha = 0.7\)). Modelled CBF and CMRO\(_2\) showed a corresponding rise during the stimulus.

In Neonate 2, a similar increase in CMRO\(_2\), \(\Delta\text{HbO}_2\) and \(\Delta\text{HbT}\) was observed during the stimulus. However, an increase in \(\Delta\text{HHb}\) was also observed (Fig. 14.7). We were able to simulate this response in \(\Delta\text{HHb}\) by an increase in demand (\(\alpha = 0.7\)).

![Fig. 14.6](image1)

**Fig. 14.6** Model simulation of Neonate 1 of the USZ study. Simulated haemodynamic response with (a) an increase in demand \(u\). Vertical lines indicate stimulus period. Measured and simulated changes in (b) oxy- and deoxy- haemoglobin (\(\Delta\text{HbO}_2\), \(\Delta\text{HHb}\)) and (c) total haemoglobin (\(\Delta\text{HbT}\)).

![Fig. 14.7](image2)

**Fig. 14.7** Model simulation of Neonate 2 of the USZ study. Simulated haemodynamic response with (a) an increase in demand \(u\) and CBF maintained constant. Vertical lines indicate stimulus period. Measured and simulated changes in (b) oxy- and deoxy- haemoglobin (\(\Delta\text{HbO}_2\), \(\Delta\text{HHb}\)) and (c) total haemoglobin (\(\Delta\text{HbT}\)). (d) Simulated cerebral metabolic rate of oxygen consumption (CMRO\(_2\)) cerebral blood flow (CBF).
while maintaining CBF constant at its normal value throughout the stimulus. We were able to better predict $\Delta HbO_2$ and $\Delta HbT$ by adding a decrease in blood pressure during the stimulus ($P_a=P_{a,n} - \beta HRF$ with $\beta = 7$) to match the magnitude of $\Delta HbO_2$ and $\Delta HbT$. However this decrease was too large to be physiologically likely within this timeframe ($-7\text{ mmHg}$).

4 Discussion

The autoregulatory capacity of the preterm neonatal brain remains unclear. However, as our model simulation (Fig. 14.2a) suggests, preterm neonates may be able to maintain constant blood flow only within a very narrow range of blood pressure values. Studies have shown that the response of HHb to a functional stimulus is sometimes ‘inverted’ in preterm neonates as compared to adults. Our efforts to simulate the haemodynamic responses observed in studies performed by Kozberg et al. [12] and Roche-Labarbe et al. [13] show that the preterm model is capable of simulating the varied functional responses observed. We observed that the model predicted an HHb decrease in response to the stimulus unless we imposed vasoconstriction (as observed by Kozberg et al. [12]). Decreasing the radius of the blood vessel resulted in the ‘inverted’ response. The model was also able to simulate fNIRS data from the USZ study relatively well. In Neonate 1 we observed a similar response of hyperemia as observed in the Roche-Labarbe et al. study. In Neonate 2, the observed rise in $\Delta HHb$ was simulated here by a constant CBF (Fig. 14.7). However, the magnitude of $\Delta HbO_2$ and $\Delta HbT$ response was not sufficiently simulated. Neonates 1 and 2 are markedly different in both gestational and actual age (Table 14.2). The former, being older, is more likely to have a developed autoregulatory capacity although we note that both subjects showed an increased HbT response. Their differences in haematocrit and haemoglobin are also notable. Indeed it has been suggested previously that HbT may have an effect on the haemodynamic response in newborns [14]. However, by changing baseline HbT, we did not observe an effect on the magnitude or shape of the model’s simulations.

In adapting the model to the human neonate we did not alter the normal radius of the blood vessel and other similar parameters, such as the thickness and muscular tension of the vessel wall. Our current work suggests that these baseline values do not have a significant effect on the results. However, these changes will be made in a forthcoming version of the model.

We aim to further investigate the functional response in neonates using our model. Our initial results suggest that the interaction of many variables affect this response including CBF, BP and the varied stages of development. This makes it very difficult to define a ‘normal’ functional response for all neonates.

Acknowledgments This work was supported by a UCL-UZH neuroscience collaborative grant. TH is supported by the doctoral training centre CoMPLEX at UCL and IT by the Wellcome Trust (088429/Z/09/Z).
References

1. Banaji M, Mallet A, Elwell CE et al (2008) A model of brain circulation and metabolism: NIRS signal changes during physiological challenges. PLoS Comput Biol 4(11):e1000212
2. Moroz T, Banaji M, Robertson NJ et al (2012) Computational modelling of the piglet brain to simulate near-infrared spectroscopy and magnetic resonance spectroscopy data collected during oxygen deprivation. J R Soc Interface 9(72):1499–1509
3. Greisen G (1986) Cerebral blood flow in preterm infants during the first week of life. Acta Paediatr Scand 75(1):43–51
4. Cooper CE, Springett R (1997) Measurement of cytochrome oxidase and mitochondrial energetics by near-infrared spectroscopy. Phil Trans R Soc Lond B 352:669–676
5. Springett R, Newman J, Cope M et al (2000) Oxygen dependency and precision of cytochrome oxidase signal from full spectral NIRS of the piglet brain. Am J Physiol Heart Circ Physiol 279:H2202–H2209
6. Elwell CE, Henty JR, Leung TS et al (2005) Measurement of CMRO2 in neonates undergoing intensive care using near infrared spectroscopy. Adv Exp Med Biol 566:263–268
7. Polin RA, Fox WW, Abman SH (2011) Fetal and neonatal physiology, 4th edn. Saunders, Philadelphia
8. Jopling J, Henry E, Wiedmeier SE et al (2009) Reference ranges for hematocrit and blood hemoglobin concentration during the neonatal period: data from a multihospital health care system. Pediatrics 123(2):e333–e337
9. Wyatt JS, Cope M, Delpy DT et al (1990) Quantitation of cerebral blood volume in human infants by near-infrared spectroscopy. J Appl Physiol (1985) 68(3):1086–1091
10. Easa D, Tran A, Bingham W (1983) Noninvasive intracranial pressure measurement in the newborn: an alternate method. Am J Dis Child 137(4):332–335
11. Volpe JJ (2008) Neurology of the newborn, 5th edn. Elsevier, Philadelphia
12. Kozberg MG, Chen BR, Deleo SE et al (2013) Resolving the transition from negative to positive blood oxygen level-dependent responses in the developing brain. Proc Natl Acad Sci U S A 110(11):4380–4385
13. Roche-Labarbe N, Fenoglio A, Radhakrishnan H et al (2014) Somatosensory evoked changes in cerebral oxygen consumption measured non-invasively in premature neonates. Neuroimage 85(Pt 1):279–286
14. Zimmermann BB, Roche-Labarbe N, Surova A et al (2012) The confounding effect of systemic physiology on the hemodynamic response in newborns. Adv Exp Med Biol 737:103–109

Open Access This chapter is licensed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license and indicate if changes were made.

The images or other third party material in this chapter are included in the chapter’s Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the chapter’s Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder.