Research Article

A Comparison of Endoscopic Ultrasound-Guided Fine-Needle Aspiration and Fine-Needle Biopsy in the Diagnosis of Solid Pancreatic Lesions

Lachlan R. Ayres,1 Elizabeth K. Kmiotek,2 Eric Lam,3 and Jennifer J. Telford3

1Poole Hospital NHS Foundation Trust, Poole, UK
2Jagiellonian University Medical College, Krakow, Poland
3Division of Gastroenterology, St. Paul’s Hospital, University of British Columbia, Vancouver, BC, Canada

Correspondence should be addressed to Lachlan R. Ayres; lachlanayres@hotmail.com

Received 15 January 2018; Accepted 20 February 2018; Published 19 April 2018

Background and Aims. Endoscopic ultrasound (EUS) guided fine-needle aspiration (FNA) is the method of choice for sampling pancreatic lesions. This study compares the diagnostic accuracy and safety of FNB using a novel core needle to FNA in solid pancreatic lesions.

Methods. A retrospective review of patients in whom EUS FNA or FNB was performed for solid pancreatic lesions was conducted. Diagnostic performance was calculated based upon a dual classification system: classification 1, only malignant pathology considered a true positive, versus classification 2, atypical, suspicious, and malignant pathology considered a true positive.

Results. 43 patients underwent FNB compared with 51 FNA. Using classification 1, sensitivity was 74.0% versus 80.0%, specificity 100% versus 100%, and diagnostic accuracy 77.0% versus 80.0% for FNB versus FNA, respectively (all \( p > 0.05 \)). Using classification 2, sensitivity was 97% versus 94.0%, specificity 100% versus 100%, and diagnostic accuracy 98.0% versus 94.0% for FNB versus FNA, respectively (all \( p > 0.05 \)). FNB required significantly fewer needle passes (median = 2) compared to FNA (median = 3; \( p < 0.001 \)). Adverse events occurred in two (4.5%) FNB patients compared with none in the FNA group (\( p > 0.05 \)). Conclusion. FNA and FNB have comparable sensitivity and diagnostic accuracy. FNB required fewer passes.
pancreatic lesions by two experienced endosonographers at St. Paul’s Hospital, Vancouver, BC, using the Covidien Shark-Core platform (Shark Core®, Covidien, Dublin, Leinster, Ireland) using 19 G, 22 G, or 25 G needles. When the study was conceived 50 FNB had been performed on solid pancreatic lesions (November 2014 to July 2015). Thus a similar number of consecutive patients undergoing FNA using a 22 G or 25 G needle (Expect, Boston Scientific, Natick, MA, USA) of solid pancreatic lesions were taken for comparison (from October 2013 to October 2014). There was no onsite pathologist present for either cohort. The number of needle passes and needle throws was not standardized and was at the discretion of the endosonographer. Assessment of an adequate specimen was also at the discretion of the endosonographer generally using a crude visual assessment of the material expressed from the needle.

Patients were excluded when there was a predominantly cystic component to the mass or if adequate follow-up was not available: either surgical pathology or six months’ clinical follow-up. The study was approved by the University of British Columbia Ethics Board.

Electronic medical records were interrogated and demographic data recorded. The size of the lesion was documented based on the largest dimension reported in millimeters. The location of the lesion was categorized as falling within the head, uncinate, genu, body, or tail of pancreas. Technical failures, number of needle passes, and needle gauge were recorded.

The pathological diagnosis by EUS was categorized as nondiagnostic, benign, atypical, suspicious, or malignant (i.e., classes 1–5, resp.). Diagnostic accuracy, positive predictive value, negative predictive value, sensitivity, and specificity were calculated using a dual classification system employed in a meta-analysis of EUS FNA by Hewitt et al. 2012 [2]. Under this approach malignancy status was established as follows:

(i) Classification 1: nondiagnostic, benign, atypical, and suspicious are considered negative for malignancy. Only the designation “malignant,” that is, class 5, is considered a true positive.

(ii) Classification 2: nondiagnostic and benign are negative for malignancy. Atypical and suspicious are also considered positive for malignancy.

Diagnostic accuracy was compared to gold standard surgical pathology, subsequent EUS FNA/FNB, or six-month clinicoradiological follow-up. It was computed as the ratio between the sum of true positive and true negative values divided by the total number of lesions. Neuroendocrine tumors which have a range of malignant potential were all regarded as a “positive” diagnosis and grouped with adenocarcinoma and lymphoma for the purposes of the analysis. Adverse events, as determined by interrogating electronic medical records, were also recorded and compared.

2.1. Statistical Analysis. Statistical analysis was performed using the chi-square test, Fisher’s exact test, or Wilcoxon rank sum test as appropriate using SAS 9.4 and R 3.2.0. A p value of ≤0.05 was considered statistically significant.

3. Results

3.1. Patient Demographics. Patients ranged from 33 to 88 years of age (median = 66 years of age for both groups). There were 22 (43.1%) versus 25 (58.1%) males in the FNA and FNB groups, respectively. Demographic data of the study population is reported in Table 1.

3.2. Lesion Characteristics. There was no difference in location, size, or pathological class (all p > 0.05) between the two groups. Lesion characteristics are shown in Table 2. Most lesions were located in the head of pancreas (61.7%) followed by the body (13.8%), tail (10.6%), uncinate (7.4%), and genu (6.4%). The mean lesion size was 27 mm (± 12.2 (SD)), 72.3% were malignant, 10.6% suspicious, 6.4% atypical, 8.5% benign, and 2.1% nondiagnostic. The majority of lesions (68%) were pancreatic ductal adenocarcinoma with neuroendocrine lesions the second most common. The two groups differed significantly in terms of the final diagnosis (p = 0.018) in that there were more neuroendocrine tumors (NET) in the FNA group and more inflammatory lesions and lymphomas in the FNB group.

3.3. Diagnostic Accuracy: Classification 1. The diagnostic performance of FNA and FNB is presented in Table 3. In the FNA group 39/51 (77%) specimens were positive for malignancy compared to 29/43 (67%) in the FNB group. Of 12 samples negative for malignancy in the FNA group, ten were false negatives (14 and ten for FNB). This translates to a
Table 2: Lesion characteristics.

| Location of lesion in pancreas, n (%) | FNA (n = 51) | FNB (n = 43) | p   |
|--------------------------------------|-------------|-------------|-----|
| Head                                 | 27 (52.9)   | 31 (72.1)   | 0.275 |
| Uncinate                             | 5 (9.8)     | 2 (4.7)     |     |
| Genu                                 | 3 (5.9)     | 3 (7.0)     |     |
| Body                                 | 8 (15.7)    | 5 (11.6)    |     |
| Tail                                 | 8 (15.7)    | 2 (4.7)     |     |

| Lesion size (largest dimension, mm) | FNA         | FNB         | p   |
|-------------------------------------|-------------|-------------|-----|
| Missing, n (%)                      | 1 (2.0)     | 5 (11.6)    | 0.787 |
| Median (IQR)                        | 26.0 (18.0, 34.0) | 26.0 (18.0, 36.0) |     |
| Mean (SD)                           | 26.8 (12.8) | 27.4 (11.5) |     |
| Range                               | (5.0, 70.0) | (9.0, 55.0) |     |

| Cytologic/histologic diagnosis      |            |             |     |
|-------------------------------------|-------------|-------------|-----|
| Nondiagnostic                       | 2 (3.9)     | 0 (0.0)     |     |
| Benign                              | 3 (5.9)     | 5 (11.6)    |     |
| Atypical                            | 2 (3.9)     | 4 (9.3)     |     |
| Suspicious                          | 5 (9.8)     | 5 (11.6)    |     |
| Malignant                           | 39 (76.5)   | 29 (67.4)   |     |

| Final diagnosis                     |            |             |     |
|-------------------------------------|-------------|-------------|-----|
| Pancreatic adenocarcinoma           | 37 (72.5)   | 31 (72.1)   | 0.018 |
| Neuroendocrine tumour               | 12 (23.5)   | 4 (9.3)     |     |
| Inflammatory                        | 0 (0.0)     | 4 (9.3)     |     |
| Lymphoma                            | 0 (0.0)     | 1 (2.3)     |     |
| Benign/normal                       | 2 (3.9)     | 1 (2.3)     |     |
| Other                               | 0 (0.0)     | 2 (4.7)     |     |

FNA = fine needle aspiration; FNB = fine needle biopsy; IQR = interquartile range; SD = standard deviation.

Table 3: Diagnostic performance: classification 1.

|                | FNA         | FNB         | p   |
|----------------|-------------|-------------|-----|
| Sensitivity    | 0.80 (0.66, 0.90) | 0.74 (0.58, 0.87) | 0.615 |
| Specificity    | 1.00 (0.99, 1.00) | 1.00 (0.82, 1.00) | 1.000 |
| PPV            | 1.00 (0.87, 1.00) | 1.00 (0.83, 1.00) | 1.000 |
| NPV            | 0.17 (0.02, 0.48) | 0.29 (0.08, 0.58) | 0.652 |
| Accuracy       | 0.80 (0.67, 0.90) | 0.77 (0.61, 0.88) | 0.801 |

Values in brackets are 95% confidence interval. p value is based on Fisher’s exact test; FNA = fine needle aspiration; FNB = fine needle biopsy.

sensitivity of 80% versus 74%, specificity of 100% versus 100%, positive predictive value (PPV) of 100% versus 100%, negative predictive value (NPV) of 17% versus 29%, and an accuracy of 80% versus 77% for FNA and FNB, respectively (all p > 0.05).

3.4. Diagnostic Accuracy: Classification 2. Using the less stringent classification 2 (see Table 4), the sensitivity, NPV, and accuracy all increased as compared to classification 1, but there was still no statistically significant difference between FNA and FNB in any of these measures. In the FNA group 46/51 (90%) were malignant versus 38/43 (88%) in the FNB group. Of five samples negative for malignancy three were false negatives for FNA (compared to five and one for FNB). Thus sensitivity was 94% versus 97%, specificity 100% versus 100%, PPV 100% versus 100%, NPV 40% versus 80%, and accuracy 94% versus 98% for FNA and FNB, respectively.

3.5. Technical Outcomes. In the FNB group, 35 (81%) lesions were sampled using a 25 G needle, six (14%) lesions were sampled using 22 G, and one (2%) lesion was sampled using 19 G (needle gauge not reported in one case). Technical outcomes are reported in Table 5. Fewer needle passes were performed in the FNB group: median two (mean 2.1) compared to a median of three (mean 3.2) in the FNA group (p < 0.001). In the FNA group five (9.8%) patients required a repeat EUS compared to eight (18.6%) in the FNB group (p = 0.218). Two adverse events were reported (one gastrointestinal bleed, no blood transfusion or endoscopic therapy required, and one self-limiting episode of mild acute pancreatitis) in the FNB group compared to none in the FNA group (p > 0.05). No technical failures were reported in either group.

In the FNA group four (7.8%) specimens were paucicellular/inadequate, three of which required repeat EUS versus
Table 4: Diagnostic performance: classification 2.

|               | FNA                  | FNB                  | p    |
|---------------|----------------------|----------------------|------|
| Sensitivity   | 0.94 (0.83, 0.99)    | 0.97 (0.87, 1.00)    | 0.626|
| Specificity   | 1.00 (0.09, 1.00)    | 1.00 (0.28, 1.00)    | 1.000|
| PPV           | 1.00 (0.89, 1.00)    | 1.00 (0.87, 1.00)    | 1.000|
| NPV           | 0.40 (0.05, 0.85)    | 0.80 (0.28, 0.99)    | 0.524|
| Accuracy      | 0.94 (0.84, 0.99)    | 0.98 (0.88, 1.00)    | 0.623|

Values in brackets are 95% confidence interval. p value is based on Fisher’s exact test; FNA = fine needle aspiration; FNB = fine needle biopsy.

Table 5: Technical outcomes.

|                | FNA (n = 51) | FNB (n = 43) | p     |
|----------------|--------------|--------------|-------|
| Passes Unknown| 1            | 0            | <0.001|
| 1              | 1 (2.0)      | 0 (0.0)      |       |
| 2              | 5 (10.0)     | 36 (83.7)    |       |
| 3              | 27 (54.0)    | 6 (14.0)     |       |
| 4              | 16 (32.0)    | 1 (2.3)      |       |
| 5              | 1 (2.0)      | 0 (0.0)      |       |
| Required a repeat EUS/FNA, n (%) |              |              | 0.218 |
| No             | 46 (90.2)    | 35 (81.4)    |       |
| Yes            | 5 (9.8)      | 8 (18.6)     |       |
| Adverse events, n (%) | 0 (0)        | 2 (4.65)     | 0.207 |

FNA = fine needle aspiration; FNB = fine needle biopsy; EUS = endoscopic ultrasound.

4. Discussion

In this study comparing FNA to FNB, both needles demonstrate similar diagnostic performance, with FNB requiring significantly fewer needle passes to obtain sufficient diagnostic material regardless of the classification system used. To our knowledge this is the largest study comparing SharkCore FNB to conventional FNA in the diagnosis of solid pancreatic masses.

The dual classification system used in this investigation was adapted from Hewitt et al. 2012 who conducted a meta-analysis of EUS FNA in the diagnosis of pancreatic masses. Their pooled findings for sensitivity, specificity, PPV, and NPV were 85%, 98%, 99%, and 64% under classification I and 91%, 94%, 98%, and 72% under classification 2 [2]. These data are comparable to the findings presented here.

Our experience with SharkCore is similar to the findings described in a North American multicenter study using SharkCore for the diagnosis of various solid lesions. Of 250 lesions sampled, 88% were diagnostic with a median of two passes. Subgroup analysis showed that for pancreatic masses 86% were diagnostic [19]. They found a trend towards superior pathologic yield when compared to cytologic yield (87% versus 68%) but this was not statistically significant. Similarly, the Newcastle group (UK) found that SharkCore had a 90.1% sensitivity compared to ProCore™ 71.1% in a cohort of 201 patients with solid pancreatic lesions [20].

Various studies demonstrate that ProCore FNB requires fewer passes to obtain a pathologic diagnosis [13, 15–17, 19, 21]. Similarly, a recent pilot study by Adler et al. found that the SharkCore FNB required fewer passes compared to standard FNA (1.5 passes versus three passes, resp.) in a 30-patient cohort [18]. This finding supports the results of this study where a median number of two passes was needed for FNB versus three for FNA.

Of the studies investigating ProCore FNB, most did not identify a significant difference in diagnostic sensitivity between FNA and FNB for solid pancreatic masses [13–16, 19, 22]. Furthermore, when other lesions were analyzed in conjunction with pancreatic masses such as lymph nodes and gastrointestinal mass lesions, both needles still exhibited comparable levels of accuracy [17, 21, 23]. FNA sensitivity ranged within 72–92% versus 90–97.8% for FNB, and specificity ranged within 80%–100% versus 100%, respectively [13, 16, 17, 21]. This is similar to our own findings where FNA sensitivity was 80%–94% compared to 74%–97% for FNB in classification systems 1 and 2, respectively. Two studies found FNB (ProCore) to be inferior to FNA in the diagnosis of pancreatic masses [24, 25]. However both studies were small and one used a different number of passes for FNA and FNB.

Accuracy ranged within 90–94.8% for FNA and 84.6–98.3% for FNB in previously published comparative studies [15–17, 21, 24]. Not all investigations examined in this paper specified the criteria establishing malignancy; thus stringent comparisons using the two classification systems are not possible. As with most studies of EUS FNA/FNB false
5. Conclusions

In summary, the study found FNA and FNB to have comparable diagnostic accuracy and safety, with FNB requiring fewer passes. A prospective, randomized trial is warranted to establish whether FNB has an advantage over FNA.

Disclosure

This data was presented in abstract format at DDW 2016 (http://www.giejournal.org/article/S0016-5107(16)01175-5/full-text).

Conflicts of Interest

Drs. Elizabeth K. Kmiotek, Lachlan R. Ayres, Eric Lam, and Jennifer J. Telford have no conflicts of interest or financial ties to disclose.

References

[1] H. Uehara, K. Ikezawa, N. Kawada et al., "Diagnostic accuracy of endoscopic ultrasound-guided fine needle aspiration for suspected pancreatic malignancy in relation to the size of lesions," Journal of Gastroenterology and Hepatology, vol. 26, no. 8, pp. 1256–1261, 2011.
[2] M. J. Hewitt, M. J. W. McPhail, L. Possamai, A. Dhar, P. Vlavianos, and K. J. Monahan, "EUS-guided FNA for diagnosis of solid pancreatic neoplasms: a meta-analysis," Gastrointestinal Endoscopy, vol. 75, no. 2, pp. 319–331, 2012.
[3] B. R. Weston and M. S. Bhutani, "Optimizing diagnostic yield for EUS-guided sampling of solid pancreatic lesions: A technical review," Journal of Gastroenterology and Hepatology, vol. 9, no. 6, pp. 352–363, 2013.
[4] K. Hanada, A. Okazaki, N. Hirano et al., "Diagnostic strategies for early pancreatic cancer," Journal of Gastroenterology, vol. 50, no. 2, pp. 147–154, 2014.
[5] M. A. Eloubeidi, D. Jhala, D. C. Chhieng et al., "Yield of endoscopic ultrasound-guided fine-needle aspiration biopsy in patients with suspected pancreatic carcinoma: emphasis on atypical, suspicious, and false-negative aspirates," Cancer, vol. 99, no. 5, pp. 285–292, 2003.
[6] S. Yoshinaga, H. Suzuki, I. Oda, and Y. Saito, "Role of endoscopic ultrasound-guided fine needle aspiration (EUS-FNA) for diagnosis of solid pancreatic masses," Digestive Endoscopy, vol. 23, no. 1, pp. 29–33, 2011.
[7] M. S. Bhutani, P. Koduru, V. Joshi et al., "The role of endoscopic ultrasound in pancreatic cancer screening," Endoscopic Ultrasound, vol. 5, no. 1, pp. 8–16, 2016.
[8] P. Thomas Cherian, P. Mohan, A. Douiri, P. Taniere, R. K. Hejmadi, and B. S. Mahon, "Role of endoscopic ultrasound-guided fine-needle aspiration in the diagnosis of solid pancreatic and peripancreatic lesions: Is onsite cytopathology necessary?" HPB, vol. 12, no. 6, pp. 389–395, 2010.
[9] A. R. Schneider, A. Nerlich, T. Topalidis, and W. Schepp, "Specialized clinical cytology may improve the results of EUS (endoscopic ultrasound)-guided fine-needle aspiration (FNA) from pancreatic tumors," Endoscopy International Open, vol. 03, no. 02, pp. E134–E137, 2015.
[10] C. Fabbri, A. M. Polifemo, C. Luigiano et al., "Endoscopic ultrasound-guided fine needle aspiration with 22- and 25-gauge needles in solid pancreatic masses: A prospective comparative study with randomisation of needle sequence," Digestive and Liver Disease, vol. 43, no. 8, pp. 647–652, 2011.
[11] M. F. Madhoun, S. B. Wani, A. Rastogi et al., “The diagnostic accuracy of 22-gauge and 25-gauge needles in endoscopic ultrasound-guided fine needle aspiration of solid pancreatic lesions: A meta-analysis,” *Endoscopy*, vol. 45, no. 2, pp. 86–92, 2013.

[12] M. J. Levy, “Endoscopic ultrasound-guided trucut biopsy of the pancreas: Prospects and problems,” *Pancreatology*, vol. 7, no. 2-3, pp. 163–166, 2007.

[13] A. Alatawi, F. Beuvon, S. Grabar et al., “Comparison of 22G reverse-beveled versus standard needle for endoscopic ultrasound-guided sampling of solid pancreatic lesions,” *United European Gastroenterology Journal*, vol. 3, no. 4, pp. 343–352, 2015.

[14] M. J. Yang, H. Yim, J. C. Hwang et al., “Endoscopic ultrasound-guided sampling of solid pancreatic masses: 22-gauge aspiration versus 25-gauge biopsy needles,” *BMC Gastroenterology*, vol. 15, pp. 1–8, 2015.

[15] Y. N. Lee, J. H. Moon, H. K. Kim et al., “Core biopsy needle versus standard aspiration needle for endoscopic ultrasound-guided sampling of solid pancreatic masses: A randomized parallel-group study,” *Endoscopy*, vol. 46, no. 12, pp. 1056–1062, 2014.

[16] R. L. Ganc, A. P. Carbonari, R. Colaiacovo et al., “Mo1414 EUS-FNA of Solid Pancreatic Lesions: A Prospective, Randomized, Single Blinded, Comparative Study Using the 22-Gauge EchoTip® ProCore™ HD and the 22-Gauge EchoTip® Ultra HD Endoscopic Ultrasound Needles,” *Gastrointestinal Endoscopy*, vol. 79, no. 5, pp. AB427–AB428, 2014.

[17] T. Hucl, E. Wee, S. Anuradha et al., “Feasibility and efficiency of a new 22G core needle: A prospective comparison study,” *Endoscopy*, vol. 45, no. 10, pp. 792–798, 2013.

[18] D. G. Adler, B. Witt, B. Chadwick et al., “Pathologic evaluation of a new endoscopic ultrasound needle designed to obtain core tissue samples: A pilot study,” *Endoscopic Ultrasound*, vol. 5, no. 3, pp. 178–183, 2016.

[19] B. L. Witt, D. G. Adler, K. Hilden, and L. J. Layfield, “A comparative needle study: EUS-FNA procedures using the HD ProCore™ and EchoTip® 22-gauge needle types,” *Diagnostic Cytopathology*, vol. 41, no. 12, pp. 1069–1074, 2013.

[20] M. K. Nayar, B. Paranandi, M. F. Dawwas et al., “Comparison of the diagnostic performance of 2 core biopsy needles for EUS-guided tissue acquisition from solid pancreatic lesions,” *Gastrointestinal Endoscopy*, vol. 85, no. 5, pp. 1017–1024, 2017.

[21] M. Lin, C. D. Hair, L. K. Green et al., “Endoscopic ultrasound-guided fine-needle aspiration with on-site cytopathology versus core biopsy: a comparison of both techniques performed at the same endoscopic session,” *Endoscopy International Open*, vol. 02, no. 04, pp. E220–E223, 2014.

[22] J. Y. Bang, S. Hebert-Magee, J. Trevino, J. Ramesh, and S. Varadaraju, “Randomized trial comparing the 22-gauge aspiration and 22-gauge biopsy needles for EUS-guided sampling of solid pancreatic mass lesions,” *Gastrointestinal Endoscopy*, vol. 76, no. 2, pp. 321–327, 2012.

[23] G. Mavrogenis, B. Weynard, A. Sibille et al., “25-gauge histology needle versus 22-gauge cytology needle in endoscopic ultrasonography-guided sampling of pancreatic lesions and lymphadenopathy,” *Endoscopy International Open*, vol. 03, no. 01, pp. E63–E68, 2015.

[24] G. Vanbiervliet, B. Napoléon, M. C. Saint Paul et al., “Core needle versus standard needle for endoscopic ultrasound-guided biopsy of solid pancreatic masses: A randomized crossover study,” *Endoscopy*, vol. 46, no. 12, pp. 1063–1070, 2014.