OPTIMIZATION OF CLAVULANIC ACID PRODUCTION BY STREPTOMYCES DAUFPE 3060 BY RESPONSE SURFACE METHODOLOGY

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ABSTRACT

Clavulanic acid (CA) is a β-lactam antibiotic which has a potent β-lactamase inhibiting activity. In order to optimize its production by the new isolate Streptomyces DAUFPE 3060, the influence of two independent variables, temperature and soybean flour concentration, on clavulanic acid and biomass concentrations was investigated in 250 mL-Erlenmeyers according to a $2^2$ central composite design. To this purpose, temperature and soybean flour (SF) concentration were varied in the ranges 26-34°C and 10-50 g/L, respectively, and the results evaluated utilizing the Response Surface Methodology. The experimental maximum production of clavulanic acid (629 mg/L) was obtained at 32°C and 40 g/L SF after 48 h, while the maximum biomass concentration (3.9 g/L) at 30°C and 50 g/L soybean flour, respectively. These values are satisfactorily close to those (640 mg/L and 3.75 g/L, respectively) predicted by the model, thereby demonstrating the validity of the mathematical approach adopted in this study.

Key words: Clavulanic acid, response surface methodology, production, optimization, streptomyces sp.

INTRODUCTION

Clavulanic acid (CA) is a β-lactamase inhibitor that is administered in combination with penicillin group antibiotics to overcome certain types of antibiotic resistance. Despite sharing the β-lactam ring typical of penicillins, CA has low intrinsic antimicrobial activity. However, such a similarity in chemical structure allows it acting as a competitive inhibitor of β-lactamases secreted by certain bacteria to confer resistance to β-lactam antibiotics (30). The combined action as β-lactamase inhibitor and antibacterial agent makes CA very important both clinically and economically (21).
The pharmacokinetic characteristics of CA supported the development of combined therapy regimens with amoxicillin and ticarcillin, and the therapeutic success of these combination drugs is well recognized. CA formulations have been used widely and effectively in the treatment of a broad range of clinical infections for nearly 20 years (11).

Nowadays, the β-lactam antibiotics, particularly penicillins and cephalosporins, represent the world’s major biotechnology products, with around 65 % of the total world market of antibiotics (10).

*Streptomyces* is the largest antibiotic-producing genus in the discovered microbial world. Species belonging to this genus still remain an important source of antibiotics. In recent years, screening of natural products, particularly microbial products, has fallen out. Nevertheless, it is becoming increasingly apparent that 99 % of the diverse bacterial species is still unexplored (10, 28, 37). Thus, the discovery of new *Streptomyces* species is a challenge for the improvement of CA production.

However, closer inspection of metabolite production patterns among other producers of CA, clavams and cephamycin C suggests that a strong selective pressure, rather than mere chance, has created actinomycetes that coproduce CA and a β-lactam antibiotic such as cephamycin C. Among these, several strains of *Streptomyces clavuligerus, Streptomyces jumonjinensis, Streptomyces katsurahamanus,* and an unclassified *Streptomyces* sp. are reported to produce CA (15).

The productivity of microbial metabolites is, in general, closely related to the fermentation process (6, 12, 18). In order to reduce the costs of a bioprocess, it is necessary to develop strains with increased productivity, use inexpensive raw materials, improve filtration properties, perform the process under favorable conditions (30), thus requiring its optimization. Most of previous studies on CA were devoted to the improvement of its production by *S. clavuligerus* (36), for which a final CA concentration of even 1384 mg/L has recently been reported (31).

However, there are only a few works that deal with high CA producing mutants of this species or other *Streptomyces* species (17, 19) as well as process optimization by statistical design (35).

Statistical design of experiments is a widely used tool for process optimization and control. By a factorial design, the significant factors and their effects can be studied using only a few experiments, thus saving time and reducing the operating costs (4). Response surface methodology (RSM), originally described by Box and Wilson (3), enables evaluation of the effects of many factors and their interactions on response variables. The main advantage of RSM is the reduced number of experimental runs needed to provide sufficient information for statistically acceptable results (9). Therefore, it is less laborious and time-consuming compared to full-factorial experimentation (32).

Aiming at reducing the cost of the process, a new good producer (*Streptomyces* DAUFPE 3060) is proposed in this study for the production of CA, using a minimal medium composed only of soybean flour and glycerol. In previous study, the influence of the most important process variables on CA production by this strain was screened using a $2^{5-2}$ fractional factorial design, and temperature and soybean flour concentration resulted to be the most significant ones (34). Although the new isolate was not yet characterized, its high performance in CA production with respect to common *S. clavuligerus* strains constituted the main novelty of that study. On the basis of this ground, a $2^2$ central composite design combined with RSM was used in this work to select the best values of these two variables able to optimize CA production by the same strain.

**MATERIALS AND METHODS**

**Reagents**

Potassium clavulanate from *Streptomyces clavuligerus* was
used to obtain the calibration curve needed to determine CA concentration. The imidazole used in the CA determination was provided by Sigma Aldrich (São Paulo, São Paulo, Brazil). The salts, glycerol, bacto-peptone, malt extract, and yeast extract used to prepare the media were analytical grade reagents.

Microorganism

The *Streptomyces* spp. DAUFPE 3060 strain was kindly provided by the Microorganism Collection of the Department of Antibiotics of the Federal University of Pernambuco, Recife-PE, Brazil. The microorganism, which is still at the stage of characterization, was stored in cryotubes (glycerol 10 %, v/v) at -70 °C and used throughout the present work.

Culture media

The seed medium had the following composition (in g/L distilled water): glycerol, 15; bacto-peptone, 10; malt extract, 10; yeast extract, 1.0; K₂HPO₄, 2.5; MgSO₄·7H₂O, 0.75; MnCl₂·4H₂O, 0.001; FeSO₄·7H₂O, 0.001; and ZnSO₄·7H₂O, 0.001. The pH of the medium was adjusted to 6.8 with NaOH 5.0 M before autoclaving at 121 °C for 15 min. The inoculum medium used in the cultivations, based on that proposed by Maranesi et al. (22), had the following composition (in g/L distilled water): glycerol, 10; soybean flour (SF), 20; K₂HPO₄, 1.2; MnCl₂·4H₂O, 0.001; FeSO₄·7H₂O, 0.001; ZnSO₄·7H₂O, 0.001, pH 6.8. The composition of the production medium was similar to that used for the inoculum, except for the concentration of glycerol (5.0 g/L) and the SF concentration that was varied according to the selected experimental design.

Fermentation conditions

Spores of *Streptomyces* spp. DAUFPE 3060 were obtained from colonies grown on solid culture medium in Petri plates. These colonies were withdrawn and grown in liquid culture medium for 96 h. After determination of initial biomass concentration by dry weight, the spores were lyophilized and stored in glycerol 10 % v/v. The seed culture was prepared by adding the lyophilized spores (6.6 g/L dry weight) contained in a cryotube with glycerol to 25 mL of seed medium in 250 mL-Erlenmeyer flasks that were incubated in an orbital shaker at 28 °C and 200 rpm for 24 h. Afterward, 250 mL-Erlenmeyer flasks containing 45 mL of inoculum medium were inoculated with 5.0 mL of the seed culture and incubated in an orbital shaker under the same conditions. Then, 5.0 mL aliquots of this suspension with cells at the exponential growth phase (24 h) were transferred to 250 mL-Erlenmeyer flasks containing 45 mL of the production medium. The exponential phase was obtained after 24 h. The 14 production runs, which lasted 168 h, were performed in an orbital shaker at 150 rpm at different temperatures, according to the experimental design described later.

Analytical methods

The fermentation broth was centrifuged at 5500 x g for 20 min at 4 °C, and the cell pellet was washed twice with distilled water and dried to constant weight at 80 °C. The CA concentration in the fermented broth was determined spectrophotometrically by its reaction with imidazole (2). For this purpose, the increase in the optical density at 311 nm consequent to the formation of the product [1-(8-hydroxy-6-oxo-4-azooct-2-enol)-imidazole] was determined using a spectrophotometer Ultrospec 3000 pro UV/Visible (GE Healthcare, Life Sciences, Uppsala, Sweden). Glycerol concentration was determined according to Hae Bok and Demain (13). All the analyses were performed in triplicate.

Experimental design and data analysis

In order to quantify the influence of the selected independent variables, specifically soybean flour concentration (v₁) and temperature (v₂), on the responses (concentrations of CA and biomass), a 2² central composite design with threecoded levels leading to 14 experiments was used, which contained a factorial or fractional factorial matrix with center
points and star points to allow estimation of the curvature (7).

The range and levels of the variables investigated in this study are given in Table 1. Six tests at the central point were performed to estimate the pure error needed for the analysis of variance, to examine the presence of curvature in the response surface, and to investigate the suitability of the proposed models.

The concentrations of CA and biomass were determined after 48 h, because previous experiments demonstrated that after this time CA degradation begun. The quadratic model to predict the optimal point was expressed by the equation:

\[
y = b_0 + \sum b_i x_i + \sum b_{ij} x_i x_j + \sum b_{ii} x_i^2
\]  

[1]

where \( y \) represents the predicted value of the response variables, \( b_0 \) is the intercept coefficient, \( b_i \) are the linear coefficients, \( b_{ii} \) are the quadratic coefficients and \( b_{ij} \) are the interaction ones.

### Table 1. Levels of factors used in the 2\(^2\) central composite design.

| Coded values | Low | Central | High | Axial | Axial |
|--------------|-----|---------|------|-------|-------|
| \( v_1 \) - SF concentration (g/L) | 20  | 30  | 40  | 10  | 50  |
| \( v_2 \) - Temperature (°C)      | 28  | 30  | 32  | 26  | 34  |

The statistical significance of the regression coefficients was determined by the Student’s t-test, and the second-order model equation was determined by the Fischer’s test. The effects of unexplained variability in the observed response due to extraneous factors were minimized by randomizing the order of experiments (20).

### RESULTS AND DISCUSSION

#### Experimental design and optimization by RSM

The results of a first attempt to point out the main variables influencing the CA production by *Streptomyces* spp. DAUFPE 3060 were already discussed in previous work (34). For this purpose, the effects of temperature, agitation intensity, pH, glycerol and soybean flour (SF) concentrations were investigated using a 2\(^{5-2}\) fractional factorial design. The regression analysis showed that both SF concentration and temperature were significant for CA production at probability levels of 95 and 97%, respectively, thereby proving the mostly influencing independent variables. However, the simultaneous variations of five variables in that effort did not enable us to predict their optimum levels, but only suggested ranges of temperature and SF concentration within which subsequent optimization should be focused. This is the objective of the present optimization work.

The main results obtained by *Streptomyces* DAUFPE 3060 fermentations according to the 2\(^2\) central composite design selected for this study are listed in Table 2. Experiments were planned so as to obtain quadratic models able to describe the CA and biomass concentrations as simultaneous functions of temperature and SF levels.

The maximum CA production (629 mg/L) was achieved at 32 °C and SF = 40 g/L (run 4). Similarly, the maximum biomass concentration (3.9 g/L) was observed at SF = 50 g/L and 30 °C (run 6).
Table 2. Level combinations of the two independent variables (temperature and SF concentration) used in the $2^2$ central composite design, and results of the selected responses (clavulanic acid and biomass concentrations).

| Run | Temperature ($^\circ$C) | SF (g/L) | Clavulanic acid (mg/L) | Biomass (g/L) |
|-----|-------------------------|----------|------------------------|---------------|
| 14 (C) | 30                      | 30       | 567                    | 3.4           |
| 7    | 26                      | 30       | 501                    | 2.4           |
| 12 (C) | 30                      | 30       | 566                    | 3.3           |
| 10 (C) | 30                      | 30       | 567                    | 3.1           |
| 5    | 30                      | 10       | 221                    | 1.8           |
| 6    | 30                      | 50       | 582                    | 3.9           |
| 2    | 32                      | 20       | 377                    | 2.0           |
| 13 (C) | 30                      | 30       | 567                    | 3.5           |
| 11 (C) | 30                      | 30       | 568                    | 3.4           |
| 4    | 32                      | 40       | 629                    | 3.3           |
| 9 (C) | 30                      | 30       | 565                    | 3.1           |
| 8    | 34                      | 30       | 595                    | 2.9           |
| 3    | 28                      | 40       | 562                    | 3.0           |
| 1    | 28                      | 20       | 306                    | 1.3           |

*All values were statistically significant at 95% confidence level.

The experimental data of Table 2 were then used to make regression analyses fitting both responses. The following equations, where the variables take their coded values, express the best models for CA and biomass concentrations, respectively:

\[ \hat{y}_1 = 566.67 + 127.32 x_1 - 84.15 x_1^2 + 33.87 x_2 - 10.89 x_2^2 - 1.00 x_1 x_2 \] \[ 3 \]

\[ \hat{y}_2 = 3.30 + 0.75 x_1 - 0.31 x_1^2 + 0.21 x_2 - 0.41 x_2^2 - 0.10 x_1 x_2 \] \[ 4 \]

where $\hat{y}_1, \hat{y}_2$ and $x_i$ are CA, biomass and SF concentrations, while $x_2$ is temperature.

The simultaneous effects of temperature and SF concentration and their interactions on the CA and biomass productions are better visualized in three-dimensional (3-D) graph projections (Figures 1 and 2) using the Response Surface Methodology (RSM).
Figure 2. Three-dimensional response surface contour plot showing the simultaneous effects of soybean flour concentration and temperature on biomass concentration.

Although the statistical analysis revealed positive main linear effects of both independent variables (significant at 95% confidence level) either on CA or biomass concentrations, the effect of SF level was the stronger. However, this behavior was not linear, since the quadratic effects of these variables were also significant. This means that the values of both responses increased up to a certain level when both variables were raised. As a result, there are two regions of temperature (30-34°C) and SF concentration (35-45 g/L) where both responses reach maximum values. Although the interaction term ($x_1x_2$) was not significant, it was kept in the model to allow for the error determination. Simultaneous regression by eqs. [3] and [4] provided maximum predicted values of CA concentration (640 mg/L) at 33.0 °C and SF = 37.5 g/L, and of biomass concentration (3.75 g/L) at 30.2 °C and SF = 41.77 g/L, respectively. These predicted values are only 1.7 % higher and 3.8% lower than the experimental ones, thus demonstrating the validity of the models employed.

The statistical significance of second-order model equations was confirmed by the F-test analysis of variance (ANOVA) and multiple regression (Tables 3 and 4). In fact, the computed F-values (27.08 for $\hat{y}_2$ and 3499 for $\hat{y}_1$) were much greater than the tabulated ones [$F_{(2,11)} = 11.63$ and 14.54, respectively]. Whereas the lack of fit was insignificant for $\hat{y}_2$ [$F = 3.04 < F_{(2,11)}$], that for $\hat{y}_1$, although significant ($F = 25.7$), showed a very low pure error (1.1). In addition, the high values of the determination coefficients ($R^2 = 0.97$ and 0.99 for biomass and CA concentrations, respectively) indicate that the models explained 97-99% of the variability data, and both regressions were statistically significant ($p < 0.0001$) at 95% confidence level.

Table 3. Analysis of variance applied to the regression model used for the CA concentration ($R^2 = 0.99$).

| Source    | Sum of squares | Degrees of freedom | Mean of square | F-value | Prob>F | Remarks      |
|-----------|---------------|--------------------|---------------|---------|--------|--------------|
| Model     | 191287.8      | 5                  | 38257.6       | 3499.7  | 0.000000 Significant |
| $^a b_1$  | 129675.7      | 1                  | 129675.7      | 121571.0| 0.000000 Significant |
| $b_11$    | 52286.9       | 1                  | 52286.9       | 49019.0 | 0.000000 Significant |
| $^a b_2$  | 9175.8        | 1                  | 9175.8        | 8602.3  | 0.000000 Significant |
| $b_{22}$  | 876.7         | 1                  | 876.7         | 821.9   | 0.000001 Significant |
| $b_{12}$  | 4.0           | 1                  | 4.0           | 3.7     | 0.110567 Not Significant |
| Residual  | 87.5          | 8                  | 10.9          |         |        |              |
| Lack of fit | 82.1      | 3                  | 27.4          | 25.7    | 0.001830 Significant |
| Pure error | 5.3        | 5                  | 1.1           |         |        |              |
| Total     | 191375.2      | 13                 |               |         |        |              |

Independent variables: $^1$ = SF concentration; $^2$ = Temperature.
Table 4. Analysis of variance applied to the regression model used for the biomass concentration ($R^2 = 0.97$).

| Source | Sum of squares | Degrees of freedom | Mean of square | F-value | Prob>F | Remarks |
|--------|----------------|--------------------|----------------|---------|--------|---------|
| Model  | 6.70           | 5                  | 1.34           | 27.09   | 0.000082 | Significant |
| $b_1$  | 4.45           | 1                  | 4.45           | 159.10  | 0.000056 | Significant |
| $b_{11}$ | 0.72         | 1                  | 0.72           | 25.76   | 0.003851 | Significant |
| $b_2$  | 0.36           | 1                  | 0.36           | 13.01   | 0.015430 | Significant |
| $b_{22}$ | 1.26         | 1                  | 1.26           | 44.87   | 0.001121 | Significant |
| $b_{12}$ | 0.04         | 1                  | 0.04           | 1.43    | 0.285591 | Not Significant |
| Residual | 0.39         | 8                  | 0.05           |         |         | Significant |
| Lack of fit | 0.26        | 3                  | 0.08           | 3.04    | 0.130860 | Not Significant |
| Pure error | 0.14        | 5                  | 0.03           |         |         |         |
| Total   | 7.10           | 13                 |                |         |         |         |

Independent variables: $^1 = $ Soybean flour concentration; $^2 = $ Temperature.

Relationship between CA production and biomass growth

As expected by the above trends, an almost linear relationship seems to hold between CA and biomass levels (Figure 3), which demonstrates that this strain is able to produce CA during growth. These results agree with those of Mayer and Deckwer (23), who observed that growth and production occurred simultaneously in the trophophase. CA production associated to cell growth was already detected utilizing soybean derivatives as nitrogen sources (21, 16). On the other hand, other authors observed that CA synthesis occurred during the trophophase, but the highest CA concentration was obtained during the stationary one (1, 33).

These results are of particular concern, when compared with those of Neves et al. (24), who utilized as industrial inoculum cells at the exponential growth phase after the same time (24 h) and obtained a maximum CA production of about 100 mg/L after a significantly longer time (about 70 h). CA production comparable to that of the present study (672-681 mg/L) was observed with S. clavuligerus using 38 g/L soy meal powder, 0.39 g/L FeSO$_4$$\cdot$7H$_2$O and 1.18 g/L ornithine (35).

As suggested by Butterworth (5), soybean proteins are important nutrients for CA production. The effectiveness of SF as a nitrogen source is in accordance with the results of Ortiz et al. (25), who observed CA productions in Erlenmeyer flasks increasing from 338 to 698 mg/L when 20 g/L SF substituted 10.9 g/L soybean protein isolate as a nitrogen source. Higher CA levels (up to 1384 mg/L) were obtained in medium containing glycerol and sucrose as carbon sources supplemented with amino acids (31), and in a glycerol-based medium with protein extract of soybean (1178 mg/L) (12).

As far as the effect of temperature is concerned, it is well known that an increase in this parameter stimulates CA production (34), even if it is very difficult to evaluate it, because of the simultaneous occurrence of temperature-dependent CA degradation. In fact, it has recently been demonstrated, in experiments with commercial CA, that an increase in temperature from 15 to 45 °C leads to a corresponding increase in the first-order degradation constant from 0.005 to 0.012 h$^{-1}$ (29).
Fermentation under the best conditions

Figure 4 shows the time behavior of CA production, biomass growth and glycerol consumption during the fermentation that provided the best results (run 4). It should be noticed that the highest cell concentration (5.06 g/L) and CA production (629 mg/L) were both obtained before the complete consumption of glycerol, although after different times (72 and 48 h, respectively). But the most evident result is that the CA concentration rapidly fell after 48 h, as the likely result of higher degradation rate when compared to its production.
Similar CA production simultaneous to growth, achievement of maximum CA concentration and subsequent quick degradation were observed by several authors (8, 23, 26). In agreement with these findings, Lynch and Yang (21) observed that the rate of CA degradation during fermentation was comparable to that of its production and proposed that the degradation products could have been used by the microorganism for further production of the antibiotic. It was proposed that such degradation could be the result of the action of metabolites or enzymes produced by the strain (16), of compounds in the culture medium such as amino acids and inorganic materials (27) or of the autolysis resulting from the rising pH, microbial decomposition, and re-metabolization (14).

CONCLUSIONS

These results on the whole suggest that a rigorous control of the residence time would be a fundamental requisite in scaling-up an industrial process for continuous clavulanic acid production by the new isolate Streptomyces DAUFPE 3060. The next effort in this direction will deal with the attempt to carry out this process in a bench-scale fermenter.

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