Circular Single-Stranded Synthetic DNA Delivery Vectors for MicroRNA

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Abstract

Single-stranded (ss) circular oligodeoxynucleotides were previously found to undergo rolling circle transcription (RCT) by phage and bacterial RNA polymerases (RNAPs) into tandemly repetitive RNA multimers. Here, we redesign them to encode minimal primary miRNA mimics, with the long term aim of intracellular transcription followed by RNA processing and maturation via endogenous pathways. We describe an improved method for circularizing ss synthetic DNA for RCT by using a recently described thermostable RNA ligase, which does not require a split oligonucleotide to juxtapose the ligating ends. In vitro transcription of four templates demonstrates that the secondary structure inherent in miRNA-encoding vectors does not impair their RCT by RNAPs previously shown to carry out RCT. A typical primary-miRNA rolling circle transcript was accurately processed by a human Drosha immunoprecipitate, indicating that if human RNAPs prove to be capable of RCT, the resulting transcripts should enter the endogenous miRNA processing pathway in human cells. Circular oligonucleotides are therefore candidate vectors for small RNA delivery in human cells, which express RNAPs related to those tested here.

Introduction

Small RNAs such as miRNA and siRNA are now well established as sequence-specific regulators of gene expression at the mRNA level [1]. For experimental and potential therapeutic uses, small RNAs must either be synthesized and introduced into cells directly [2,3], or encoded by double stranded (ds) DNA vectors whose transcripts are made and processed within a cell [4,5,6]. While useful and effective, neither approach is without drawbacks. RNA is more expensive, more difficult to synthesize and less stable in storage than DNA, and ribonucleases from serum, cellular or other sources can degrade small RNAs before they reach their target. Plasmid and viral vectors encoding short hairpin transcripts (shRNA) carry the same information in a relatively stable form, but typically waste more than 99% of their nucleotide (nt) mass. They must also be biosynthesized in cells or by microorganisms, risking contamination in therapeutic applications, and can undergo genomic integration, potentially causing cancer [7]. These shortcomings in small RNA delivery methods have prompted us to investigate single stranded circular oligodeoxynucleotides (COLIGOs) as minimized relay molecules for shuttling small RNA sequence information into cells.

Single-stranded (ss) DNA circles were previously found to serve as in vitro templates for bacteriophage RNA polymerases (RNAPs) [8] and later E. coli RNAP [9,10,11]. Lacking a promoter, they undergo relatively infrequent transcription initiation but, once engaged, possibly by virtue of a resemblance to the transcription bubble, their circular topology permits transcription around the template many times, a process termed rolling circle transcription, or RCT (Fig. 1A). RCT can lead to the production of large multimeric transcripts before the RNAP terminates. Although their size (~20–200 nt) limits the amount of information they can carry, COLIGOs are well suited to encoding the amount of information found in small RNAs, as previously demonstrated for ribozyme sequences [9,11]. If the multimeric transcript can be made to undergo processing, for example self-processing in the ribozyme case, a large amplification of the DNA sequence into many small RNA copies per transcription event can result. This approach has been used recently as an in vitro alternative to run-off transcription in the production of siRNA using T7 RNAP [12].

We are investigating the possibility that COLIGOs might be useful as small RNA intracellular delivery vectors – chemically synthesized like siRNA but constructed in circular form from DNA, and thus more chemically stable. COLIGOs are resistant to exonuclease activity and easier to synthesize free of biologic contamination than siRNA-encoding plasmids and viruses. In addition, we envision the possibility that COLIGOs would give rise to a multimeric transcript that, by mimicking transcripts formed during the expression of endogenous miRNA clusters [13], would enter the natural miRNA maturation pathway, ultimately programming ribonucleoprotein complexes capable of miRNA and siRNA activity (Fig. 1A). To begin an assessment of this approach for producing small RNA in cells, it is necessary to test whether COLIGOs encoding primary (pri)-miRNA transcripts are accepted as transcription templates by RNAPs, and to learn whether RCT RNA multimers will be processed by Drosha/Microprocessor, the maturation pathway entry point leading to miRNA. In addition to describing an improved method for
circularizing the linear RCT template oligonucleotides, we show here that miRNA-encoding COLIGOs are substrates for RCT by bacterial and bacteriophage RNAPs. Our additional finding that miRNA rolling circle transcripts are accurately processed by the human Drosha/Microprocessor complex provides impetus for future studies assessing the RCT capabilities of human RNAPs.

**Results**

**Design and synthesis of circular DNA templates encoding miRNAs**

We designed DNA circles to encode shortened versions of two human pri-miRNAs, miR-19a and miR-122. We based our design on the endogenous human miRNA genomic template-strand sequences, and included DNA encoding ~3 helical turns of the pri-miRNA stem-loop sequence (Fig. 1B), the minimum stem length required for Drosha/Microprocessor processing [14,15]. In two of the COLIGOs we minimized the length of the sequences flanking the stem-loops to 4 nt (COLIGO 19a) and 6 nt (COLIGO 122) per repeat. In two other COLIGOS, 19am and 122m, we included more of the natural flanking sequences (taken in both cases from the miR19a genomic template-strand sequence), for a total of 22 and 18 flanking nt, respectively. Additional flanking nt are believed to enhance Drosha processing [15,16]. However, in a rolling circle transcript all but the terminal stem-loop repeats would be flanked by varying lengths of RNA, so it is not yet clear how much flanking
RNA should be encoded by a given COLIGO. The complete monomer transcripts for the four templates - two for each miRNA - used in this study are shown in Fig. 1B. Although the pri-miRNA stem-loop structure forms the basis of our COLIGO design, we note that promoterless transcription may initiate at multiple sites in the DNA circle, and that alternate folding patterns are possible for multimeric COLIGOs [12].

Some miRNAs have conserved loops that aid in their processing [17], but specific loop sequences are likely not critical to the biogenesis of most miRNAs [15,18]. Loop sequences at the DNA level in the COLIGO, however, may influence transcription efficiency, as shown for E. coli RNAP RCT [19]. We tested this variable, in combination with the longer flanking sequences, by swapping the native miRNA loops in pre-miR-19a and miR-122 for an unrelated RNA hairpin loop (from HIV TAR RNA), resulting in pre-miR19am and pre-miR122m (monomer transcripts are shown in Fig. 1B).

To choose the site where the linear oligonucleotide is closed to form the COLIGO we predicted the folding of the COLIGO using mFold [20] and selected the region having the least secondary structure, which in the linear form might prevent juxtaposition of the ends. This sequence typically coincided with the flanking nucleotides at the base of the stem of the transcripts. The DNA circles encoding the transcripts shown in Fig. 1B were therefore ligated between the 5' and 3' deoxynucleotides corresponding to the 3' and 5' terminal ribonucleotides, respectively, depicted in the monomer transcripts (Fig. 1B).

Rolling circle transcription templates have been circularized by three methods: chemical ligation within a DNA triple-helix [8]; T4 DNA ligase closure mediated by a splint oligonucleotide [21]; and T4 DNA ligase closure within a nicked DNA dumbbell structure, as for the shRNA-encoding templates [12]. Among these methods, splinted DNA ligase closure should be the most general approach for natural miRNA cDNAs, since their stem sequences include multiple mismatches, which might interfere with DNA ligase closure. However, the requirement for a splint to bring the 5' and 3' ends together increases the cost of DNA synthesis and requires an additional step to remove the splints after circularization. In preliminary experiments we also found that splint-mediated ligation resulted in unsatisfactory circularization yields and varying amounts of linear multimers.

We obtained better results using a newly discovered thermostable RNA ligase (Rnl) from bacteriophage TS2126 (Fig. 2A). TS2126 Rnl has been reported to efficiently circularize ss DNA without a splint [22]. In a typical COLIGO preparation using this enzyme, the entire 5' splint 

![Figure 2. Circularization of DNA templates (COLIGOs) for Rolling Circle Transcription.](image)

Figure 2. Circularization of DNA templates (COLIGOs) for Rolling Circle Transcription. A. Synthetic 5' phosphorylated linear DNA sequences were circularized using the thermostable TS2126 RNA ligase. B. Denaturing polyacrylamide gel electrophoresis (PAGE) at four stages during miR-19am DNA circle synthesis. Lane 1, crude DNA IDT Ultramer synthesis of COLIGO 19am. Lane 2, after preparative PAGE. Lane 3, crude circularization product. Lane 4, DNA circle template following Exonuclease I clean-up. Visualization using Stains-All. C. Verification of circular topology. Nicking of circular templates by S1 nuclease leads first to linear forms, which are then further digested to successively smaller linear forms. doi:10.1371/journal.pone.0016925.g002

and bacteriophage RNPs, COLIGOs encoding native pri-miRNAs have not been investigated. To determine whether native miRNA cDNA secondary structure might pose a general impediment to RCT, we exposed the four COLIGOs encoding miR-19a and miR-122 to T7 and E. coli RNAPs in vitro (Fig. 3A). Transcripts were visualized by [α-32P]-UTP incorporation. All four COLIGOs generated the large transcripts characteristic of RCT. For each template sequence, the circular topology was required to produce transcripts significantly longer than the COLIGOs’ circumference, and withholding one nucleotide triphosphate (ATP, Lane C-) led as expected to the loss of all but very short aborted transcripts. For all templates, more RNA was produced using E. coli RNAP compared to T7 RNAP under similar conditions (see Materials and Methods). Denaturing formaldehyde agarose gel electrophoresis showed that the maximum length of the transcripts typically reached 5 kilobases (5 kb, Fig. 3B). Thus, all four DNA circles served as RCT templates despite the extensive secondary structure predicted for COLIGOs encoding pri-miRNA stem loop structures.

In vitro transcription by bacteriophage and bacterial RNA polymerases

While COLIGOs encoding shRNAs [12] and ribozymes [9,10,11] have been shown to undergo RCT in vitro by bacterial and bacteriophage RNPs, COLIGOs encoding native pri-miRNAs have not been investigated. To determine whether native miRNA cDNA secondary structure might pose a general impediment to RCT, we exposed the four COLIGOs encoding miR-19a and miR-122 to T7 and E. coli RNAPs in vitro (Fig. 3A). Transcripts were visualized by [α-32P]-UTP incorporation. All four COLIGOs generated the large transcripts characteristic of RCT. For each template sequence, the circular topology was required to produce transcripts significantly longer than the COLIGOs’ circumference, and withholding one nucleotide triphosphate (ATP, Lane C-) led as expected to the loss of all but very short aborted transcripts. For all templates, more RNA was produced using E. coli RNAP compared to T7 RNAP under similar conditions (see Materials and Methods). Denaturing formaldehyde agarose gel electrophoresis showed that the maximum length of the transcripts typically reached 5 kilobases (5 kb, Fig. 3B). Thus, all four DNA circles served as RCT templates despite the extensive secondary structure predicted for COLIGOs encoding pri-miRNA stem loop structures.

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The relative amount of RNA produced in these experiments varied with the DNA sequence outside of the miRNA stem sequence. The template encoding the pre-miR-19a with the natural loop and short flanking sequence was noticeably poor (Fig. 3A), but it became a more efficient template when its wild-
type (wt) loop was replaced with the TAR RNA loop and the flanking sequences were lengthened (compare 19a and 19am). In contrast, the same loop and flanking sequences, when placed on the miR122 stem, decreased transcription efficiency slightly (compare 122 and 122m). These comparisons demonstrate that the non-essential sequence regions of a pri-miRNA-encoding RCT template can influence the extent to which a COLIGO is transcribed.

Drosha/Microprocessor processing of rolling circle transcripts

Primary miRNA transcripts are first processed in their natural stem-loop context by Microprocessor, a complex that in mammalian cells includes Drosha and DGCR8 [23,24]. We tested whether pre-miRNA hairpins could be processed from a multimeric rolling circle transcript. Treatment of RCT transcripts (made in vitro with uniform 32P-labeling using E. coli RNAP) with HEK293T whole cell extract (WCE) led to the release of low levels of the pre-miRNA hairpin (71 nt predicted) and the intervening flanking region (46 nt predicted) (Fig. 4A, lane 2). In order to test whether Drosha was responsible for this processing, we immunoprecipitated (IP’d) FLAG-tagged Drosha from transfected HEK293T cells [16]. The activity of IP’d Drosha (and co-IP’d proteins) was verified on an in vitro transcribed human miRNA cluster whose processing is predicted to produce three pre-miRNAs of 57-nt (pre-miR-23a), 59-nt (pre-miR-24-2) and 62-nt (pre-miR-27a) [13] (Fig. 4B, lane 10, ~60 nt RNA was produced). When the miR-19a RCT transcript was treated with IP’d Drosha, the same processed RNAs produced by WCE were released from the transcript, but in higher amounts (Fig. 4A, lane 4). The processed transcripts accumulated over the standard in vitro Drosha processing time period of 90 minutes (Fig. 4B, lanes 5, 6, 7), making it unlikely that they resulted from random degradation. These results demonstrated that pre-miRNA hairpins can be accurately processed by human Drosha from RCT transcripts. If RCT were to take place in the nucleus of human cells, then some of the RNA should be capable of entering the natural processing pathway.

Discussion

Naturally occurring small RNAs have emerged in recent years as important regulators of cellular biochemistry. Small RNAs of designed sequence are regularly used to investigate gene function, and hold great promise for therapeutic intervention. However, the problem of delivering the RNA to the inside of a human cell remains an obstacle to their therapeutic exploitation. We are attempting to recast this problem into a potentially more tractable one, wherein a small chemically synthesized DNA sequence, in circular form, is used to carry the information for making the small RNA into a cell. Our approach could result in a distinct alternative to siRNA and shRNA vectors.

The work presented here demonstrates a new enzymatic method for circularizing synthetic DNA oligonucleotides for use in RCT. In addition, we find that the secondary structure necessarily present in a COLIGO encoding pri-miRNAs does not impair RCT by bacterial and bacteriophage RNAP. Both of these RNAPs are related to eukaryotic RNAPs. The T3 and T7 bacteriophage RNAPs have significant sequence homology to the mitochondrial (mt) RNAP [25], a splice variant of which was recently found to be expressed in the nucleus of human cells [26]. The three major eukaryotic nuclear RNAPs – I, II and III – contain subunits with sequence and structural homology to the bacterial enzyme’s α and β subunits [27]. Coupled with our finding that the human Drosha/Microprocessor complex processes rolling circle transcripts, this homology justifies future, though more technically difficult, testing of human RNAPs for RCT on COLIGOs designed according to the strategy described here.

Materials and Methods

Materials, reagents and general procedures

Synthetic DNA was made by Integrated DNA Technologies (Coralville, IA) and chemically 5’ phosphorylated. Preparative and analytical denaturing polyacrylamide gel electrophoresis (DPAGE) and denaturing agarose electrophoresis were done according to standard procedures [28]. DNA was located on preparative gels using UV shadowing over fluorescent silica gel plates (EMD Chemicals 5715-7). DNA was recovered by electroelution in

Figure 3. In vitro transcription of miRNA-encoding COLIGOs by E. coli and bacteriophage T7 RNA polymerases. A. Linear (L) and circular (C) templates 122, 122m, 19a, 19am, encoding either pre-miR-122 or pre-miR-19a, were transcribed in the presence of all NTPs or all except ATP (C-). 9% DPAGE. The Phosphorimager sensitivity setting was increased 5-fold for the weaker T7 RCT reactions. B. Denaturing 1% agarose gel showing the high molecular weight size range of the transcripts. No adjustment in the Phosphorimager sensitivity setting was made. doi:10.1371/journal.pone.0016925.g003
1xTBE within a sealed dialysis membrane tube with a molecular weight cut off of 1000 (Spectrum Laboratories). Eluted DNA was phenol-chloroform extracted and ethanol precipitated with sodium acetate. In some cases, the DNA gel slice was eluted in 0.3 M sodium-acetate at 37 °C overnight and ethanol precipitated. The 1% agarose gel was prepared using Ambion’s 10X denaturing gel buffer, blotted overnight onto positively charged nitrocellulose (Ambion), baked for 20 min at 55 °C, UV crosslinked and exposed to a PhosphorImager® screen (Molecular Dynamics).

E. coli RNAP was purchased from USB. T7 RNAP and DNA ligase were purchased from New England Biolabs. RNA markers were purchased from Sigma (R4142) or Ambion (Decade marker, AM7778) and prepared according to the manufacturer’s instructions. For the agarose gel (Fig. 3B), sizes were determined by running 8 μg of total cellular RNA extracted from HEK293T cells on the same gel and staining for ribosomal RNA (18S = 1.9 kb and 28S = 5 kb) with methylene blue after blotting.

Synthesis of linear DNA templates

Linear DNA templates encoding shortened forms of pri-miR-19a, -miR-122 and -miR19am (e.g. Fig. 2B) were synthesized by IDT as single 5’ phosphorylated Ultramer sequences. The linear templates for miR122m and miR19am were also made from two half-length oligonucleotides (e.g. 19am-1 and 19am-2) according to the standard splint mediated T4 DNA ligase procedure [21]. The following sequences were used, including splint oligonucleotides where applicable:

19am (117 nt; when the linear form was synthesized as one sequence, it began as in 19am-1 and continued through 19am-2):

19am-1 (63 nt): 5’-GAAGGAAATAGCAGGCCACCATCAGTTTTGCATAGATTTGCACAACGGCAGAGAGCTCCCAG-

19am-2 (54 nt): 5’-CTCAGATCTGCCTGCAACTATGCAAAACTAACAGAGGACTGCAAACAAAAACTA, splint 19am (30 nt): 5’-CAGGCAGATCTGAGCTGGAGCTCCTCTTGCC;

122m (124 nt): 122m-1 (65 nt): 5’-AGGAAATAGGGCATAGCTATTTAGTGTGATAATGGCGTTTGATAGG-

122m-2 (59 nt): 5’-GCTCAGA-TCTGCCACAAAACACCATTGTCACACTCCACAGCTCTGCTAAGGAAACAAAA, splint 122m (31 nt): 5’-TGAGGAGAGCTCCTCCAG;

19a (86 nt): 5’-TAGCAGGCGACACATCGTATTTCGTTATGCTGGAGCTCCTCTGCC;

122 (89 nt): 5’-CTGAGGACGCAAAGAGCTGCTTGTTGTTGATGCTGGAGCTCCTCTGCC.

TS2126 RNA Ligase I

A synthetic gene (Top Gene Technologies Inc., Canada) encoding the first 393 amino acids of TS2126 RNA ligase (Rnl) I [22] followed by a C-terminal hexahistidine tag was inserted into the EcoRI/SmaI sites of pBluescriptSK(+) plasmid (Stratagene). The synthetic DNA contained codons optimized for expression in E. coli. The entire coding sequence of this plasmid, pTS2126H, was verified by DNA sequencing. E. coli BL21-CodonPlus(DE3)-RIL (Stratagene) was transformed with pTS2126H and grown in 2 liters of culture. At OD_{595} ~ 0.5 the culture was induced to a final concentration of 1 mM IPTG and grown for another 3 hrs at...
37°C. The cells were pelleted at 3000 g for 15 min at 4°C, resuspended in 30 ml native lysis buffer (50 mM NaH2PO4, 300 mM NaCl, 10 mM imidazole, pH 8 with NaOH), and disrupted using a French press. After centrifugation to remove debris, the lysate was incubated with 1 ml Ni-NTA agarose (Qiagen) with agitation for 2 hrs at 4°C. The beads were washed three times with 5 ml native lysis buffer each time, once with elution buffer without imidazole (20 mM Tris-HCl pH 8, 100 mM KCl, 2 mM DTT) and finally eluted with 500 μl portions of elution buffer containing 250 mM imidazole. The pooled fractions containing the protein were dialyzed (3 times, 100 mM KCl, 0.1 mM EDTA and 1 mM DTT, and stored at 4°C for later use. For all in vitro RNAP reactions, radiolabeled transcripts were incubated for 1 hr at 37°C, after which time the RNA was extracted with 150 μl TriReagent (Ambion), isopropanol precipitated and separated on a 9% denaturing polyacrylamide gel, dried and exposed to a PhosphorImager® screen.

The pCK-Drosha-FLAG plasmid (kindly provided by V. N. Kim) was expressed and purified as previously described [16]. Briefly, 8 μg of plasmid was transfected per 6 cm dish of HEK293T cells using 20 μl Lipofectamine 2000 (Invitrogen). For 14 hrs, the cells were lysed (900 μl Sigma FLAG® lysis buffer/6 cm dish) and Flag-IP was carried out using 40 μg of a 50% anti-FLAG bead slurry (Sigma FLAG® Tagged Protein Immunoprecipitation Kit) for at least 4 hrs at 4°C. The beads were washed 4 times with 1X Wash Buffer and once in processing buffer (20 mM Hepes-KOH, pH 7.9, 100 mM KCl, 0.2 mM EDTA, 0.5 mM DTT, 0.2 mM PMSE, 5% glycerol). A parallel mock-IP was carried out using Lipofectamine without the Drosha-encoding plasmid. The activity of the immunoprecipitated protein was verified using the miRNA-23a~27a~24-2 cluster transcribed in vitro from plasmid pGEM-T-easy_pri-miR-23, 27, 24-2+ (also provided by V. N. Kim). For HEK293T WCE as the Drosha source, cells were lysed as described above; the crude lysate was adjusted to 4°C for 30 min, centrifuged for 30 min at 13000 rpm and the supernatant collected and adjusted to 20% glycerol. For in vitro processing of rolling circle transcripts from COLIGO 19am, 1/10 of an in vitro transcription reaction using E. coli RNA polymerase (described above) or 1/4 of the eluted pri-miR-23a~27a~24-2+ transcript was used in a total volume of 30 μl containing 15 μl beads from Drosha-Flag-IP (or mock-IP, or processing buffer including 6.4 mM final MgCl2 and 0.1 u/μl RNase inhibitor). The processing reaction was carried out for 90 min at 37°C (Fig. 4A), or for the times indicated in Fig. 4B. RNA was extracted with TriReagent (Ambion), isopropanol precipitated and separated on a 10% denaturing polyacrylamide gel, dried and exposed to a PhosphorImager® screen.

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Author Contributions

Conceived and designed the experiments: KR. Performed the experiments: CIS. Analyzed the data: KR CIS. Contributed reagents/materials/analysis tools: CIS. Wrote the paper:KR.
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