Abstract

A total of 518 fecal samples collected from 183 apparently healthy cattle, 180 pigs and 155 broilers throughout Japan in 1999 were examined to determine the prevalence and antimicrobial susceptibility of Salmonella. The isolation rates were 36.1% in broilers, 2.8% in pigs and 0.5% in cattle. S. enterica Infantis was the most frequent isolate, found in 22.6% of broiler fecal samples. Higher resistance rates were observed against oxytetracycline (82.0%), dihydrostreptomycin (77.9%), kanamycin (41.0%) and trimethoprim (35.2%). Resistance rates to ampicillin, ceftiofur, bicozamycin, chloramphenicol and nalidixic acid were <10%. CTX-M-2 β-lactamase producing S. enterica Senftenberg was found in the isolates obtained from one broiler fecal sample. This is the first report of cephalosporin-resistant Salmonella directly isolated from food animal in Japan.

Findings

Salmonella enterica is a causative agent of foodborne diseases in humans. In Japan, the Food Poisoning Statistics showed that bacterial food poisoning patients numbered at 10,331 in 2008. Of the patients, salmonellosis is a leading cause accounting for 24.7% (2,551 patients). S. enterica Typhimurium was the most common serovar isolated from human cases before 1988. After 1989, S. enterica Enteritidis became the predominant serovar, accounting for almost 50% of salmonellosis in humans [1]. Salmonella is sometimes isolated from apparently healthy food-producing animals. The subclinical Salmonella infected animals can act as a contamination source for meats and products.

The Japanese Veterinary Antimicrobial Resistance Monitoring System (JVARM) was formed in 1999 in response to international concern about antimicrobial resistance [2]. In 1999, JVARM preliminarily investigated the antimicrobial susceptibility of Escherichia coli [3], enterococci [4], Campylobacter [5], and Salmonella in apparently healthy products in 1998. On the remaining cases, animal meats and products are sources of human infections. In Japan, retail meats are contaminated with Salmonella at relatively high level in broiler meats, and low levels in pork and beef [1]. Salmonella is sometimes isolated from apparently healthy food-producing animals. The subclinical Salmonella infected animals can act as a contamination source for meats and products.
cattle, pigs and broilers on farms to establish entirely microbiological procedure for the national monitoring system. In the present study, we investigated the prevalence of Salmonella in apparently healthy food-producing animals, and demonstrated the presence of cefalosporin-resistant isolate of S. enterica Senftenberg in a broiler.

A total of 518 fecal samples were randomly collected from apparently healthy cattle (183 samples), pigs (180 samples) and broilers (155 samples) from all 47 prefectures of Japan from June to December 1999, as described previously [3-5]. In brief, four samples per animal species were collected from different farms in each prefecture. Each sample was collected from individual animals. The fecal samples were transported on ice in sterile plastic specimen tubes to our laboratory, and Salmonella was isolated within 3 days. One gram of each fecal sample was inoculated into 10 ml of Hajna tetrahionate broth (Eiken Chemical Co., Ltd., Japan), followed by incubation at 42°C for 18 h for first enrichment cultures, and an additional 5–7 days at room temperature for delayed secondary enrichment culture (DSEC). After incubation, each culture was streaked onto desoxycholate-hydrogen sulfate-lactose agar (Eiken Chemical Co., Ltd.) and brilliant green agar (Eiken Chemical Co., Ltd.) plates, each containing 20 μg/ml novobiocin (Wako Pure Chemical Industries, Ltd., Japan) and incubated at 37°C for 18 h. Candidate colonies were identified biochemically by triple sugar iron (TSI) agar (Eiken Chemical Co., Ltd.) and lysine indole motility (LIM) semisolid agar (Eiken Chemical Co., Ltd.). Identification of serovars was performed by slide and tube agglutination tests (Denka Seiken Co., Ltd., Japan), according to the Kauffmann-White scheme. For individual samples, two isolates were selected for the purpose of determining susceptibility. All of the isolates were stored in 10% skim milk at -80°C until use.

The minimum inhibitory concentration (MIC) of 122 Salmonella isolates was determined using a standardized agar dilution method, as described by the Japanese Society of Chemotherapy [6], using Mueller-Hinton agar (Becton, Dickinson and Company, USA). The following 15 antimicrobial agents, approved in Japan as veterinary medicines, were tested: ampicillin, cefotiofur, apramycin, dihydrostreptomycin, kanamycin, gentamicin, oxytetracycline, bicozamycin, chloramphenicol, colistin, nalidixic acid, enrofloxacin, ofloxacin, trimethoprim and sulfadimethoxine. E. coli NIHJ and Staphylococcus aureus 209P were used for quality control. MIC resistant breakpoints were defined microbiologically when the MIC distribution of antimicrobials was bimodal.

Detection of the β-lactamase gene was carried out by polymerase chain reaction (PCR), as previously described by Kojima et al. [7]. Nucleotide sequences of both strands were determined, directly on PCR products. The DNA alignments and deduced amino acid sequences were examined using the BLAST program (National Center for Biotechnology Information, USA).

Statistical analysis was performed using the Chi-square test or Fisher's Exact test. Salmonella was isolated from 56 (36.1%, 95% Confidence Intervals [CI] 28.6–44.2%) of 155 broiler fecal samples, 5 (2.8%, CI 95% 0.9–6.4%) of 180 porcine fecal samples and 1 (0.5%, CI 95% 0–3.0%) of 183 bovine fecal samples. Isolation rates of Salmonella were significantly higher in broiler samples than porcine and bovine samples (P < 0.01). Although Salmonella was isolated from 39 samples by first enrichment, it was newly isolated from 23 samples by DSEC. Isolation rate of Salmonella increased from 7.7% by first enrichment to 12.0% by DSEC. In Okinawa, which is located in southern Japan, the isolation rates of Salmonella from rectal swab samples were 18.0% in 688 broiler chickens and 0% in 100 pigs between 1995 and 2004[8]. Recently, two large-scale surveillance studies of Salmonella infections in pigs were reported in Japan [9,10]. Kishima et al. [10] demonstrated that the prevalence of fecal carriage of Salmonella was 3.1% in 5393 pigs in 2003–2005. Futagawa-Saito et al. [9] showed that Salmonella prevalence was 2.2% and 3.3% in pig fecal samples in 1998–1999 (2980 pigs) and 2004–2005 (3791 pigs), respectively. The present results demonstrated similar Salmonella isolation rates to those previously reported in Japan. It is difficult to compare the results in other countries because there are variations in sampling methods and methods for isolation of Salmonella. Especially, the prevalence of Salmonella in the present study may be underestimated because one gram feces were used for the isolation. However, common procedure of Salmonella isolation from all animal species studied was used in the current study. As high Salmonella isolation rates were found in broiler chickens, further study should be first performed to clarify the actual status of Salmonella colonization in broiler chickens.

Salmonella isolates were classified into 14 serovars, including 12 serovars in broiler isolates, 2 serovars in porcine isolates and one serovar in bovine isolate (Table 1). S. enterica Infantis was the commonest serovar among broilers in this study, as well as in previous studies [8,11]. In Great Britain, serovars Ohio (22.0%), Kedougou (17.1%), Livingstone (12.2%) and Senftenberg (12.2%) were predominant in the isolates of broiler origin between 2005 and 2006 [12]. In Korea, serovars Enteritidis (21.9%), Typhimurium (23.4%) and Tennessee (20.3%) were frequently isolated from broilers in 2002–2003 [13]. Thus the predominant serovar of Salmonella found in broilers...
Table 1: *Salmonella* serotype distributions by animal origin

| Origin of the samples | Serotype | FE a | DSEC b | total |
|-----------------------|----------|------|--------|--------|
|                       |          | No. of Salmonella-positive samples (%) |       |        |
| Broilers (n = 155)    | Infantis | 19 (12.3) | 16† (10.3) | 35 (22.6)† |
|                       | Hadar    | 4 (2.6)  | 0 (0)   | 4 (2.6)  |
|                       | Haifa    | 1 (0.6)  | 1 (0.6) | 2 (1.3)  |
|                       | Montevideo | 2 (1.3)  | 0 (0)   | 2 (1.3)  |
|                       | Schwarzengrund | 2 (1.3)  | 0 (0)   | 2 (1.3)  |
|                       | Thompson | 2 (1.3)  | 0 (0)   | 2 (1.3)  |
|                       | Augustenborg | 1 (0.6)  | 0 (0)   | 1 (0.6)  |
|                       | Blockley | 0 (0)    | 1 (0.6) | 1 (0.6)  |
|                       | Istanbul | 0 (0)    | 1 (0.6) | 1 (0.6)  |
|                       | Newport  | 1† (0.6) | 0 (0)   | 1 (0.6)† |
|                       | Schleissheim | 1 (0.6)  | 0 (0)   | 1 (0.6)  |
|                       | Senftenberg | 1 (0.6)  | 0 (0)   | 1 (0.6)  |
|                       | untypeable | 2† (1.3) | 3 † (1.9) | 5 (3.2) † |
|                       | All serotypes | 35 (22.5) | 21 (13.5) | 56 (36.1) |
| Pigs (n = 180)        | Typhimurium | 2 (1.1)  | 0 (0)   | 2 (1.1)  |
|                       | Ohio     | 0 (0)    | 1 (0.6) | 1 (0.6)  |
|                       | untypeable | 1 (0.6)  | 1 (0.6) | 2 (1.1)  |
|                       | All serotypes | 3 (1.7)  | 2 (1.1) | 5 (2.8)  |
| Cattle (n = 183)      | Blockley | 1 (0.5)  | 0 (0)   | 1 (0.5)  |
|                       |          |        |        |        |
| Total                 |          | 39 (7.5) | 23 (4.4) | 62 (12.0) |

a FE: First enrichment culture  
b DSEC: delayed secondary enrichment culture  
† Among samples, 2 harbored 2 different serotypes (Infantis and untypeable; Newport and untypeable).

Table 2: Antimicrobial susceptibility of *Salmonella* from food-producing animals

| Antimicrobials       | Resistant breakpoint (μg/mL) | MIC range (μg/mL) | MIC50 (μg/mL) | MIC90 (μg/mL) | No. of resistant isolates (%) |
|----------------------|-----------------------------|-------------------|---------------|---------------|------------------------------|
|                      |                             |                  |               |               | Broiler (n = 111) | Pigs (n = 10) | Cattle (n = 1) | Total (n = 122) |
| Ampicillin           | 25                          | 0.39–100          | 1.56          | 3.13          | 4 (3.6) 5 (50) 0 (0) 0 (0) 9 (7.4) |
| Ceftiofur            | 6.25                        | 0.2–25            | 0.78          | 1.56          | 2 (1.8) 0 (0) 0 (0) 0 (0) 2 (1.6) |
| Apramycin            |                             | 0.39–12.5         | 1.56          | 3.13          | 88 (79.3) 6 (60) 1 (100) 95 (77.9) |
| Dihydrostreptomycin | 100                         | 6.25–100          | >100          | >100          | 49 (44.1) 0 (0) 1 (100) 50 (41.0) |
| Kanamycin            | 25                          | 0.39–100          | 1.56          | >100          | 93 (83.8) 6 (60) 1 (100) 100 (82.0) |
| Gentamicin           |                             | 0.1–3.13          | 0.39          | 0.78          | 25 (21.5) 25 (25) 5 (50) 32 (26.2) |
| Oxytetracycline      | 25                          | 3.13–100          | >100          | >100          | 25 (21.5) 25 (25) 5 (50) 32 (26.2) |
| Bicozamycin          | 100                         | 12.5–400          | 25            | 8 (7.2)       | 8 (6.6) |
| Chloramphenicol      | 50                          | 0.78–100          | 3.13          | 6.25          | 0 (0) 4 (40) 0 (0) 4 (3.3) |
| Colistin             |                             | 0.39–3.13         | 0.78          | 1.56          | 2 (1.6) 0 (0) 1 (100) 3 (2.4) |
| Nalidixic acid       | 25                          | 3.13–100          | 3.13          | 6.25          | 6 (5.4) 0 (0) 0 (0) 6 (4.9) |
| Enrofloxacin         | ≤0.05–0.39                  | ≤0.05            | 0.1           |              | 2 (1.8) 0 (0) 0 (0) 2 (1.6) |
| Ofloxacin            | ≤0.05–0.78                  | 0.1              |              |              | 6 (5.4) 0 (0) 0 (0) 6 (4.9) |
| Trimethoprim         | 3.13                        | 0.1–6.25          | 0.39          | 6.25          | 42 (37.8) 1 (10) 0 (0) 43 (35.2) |
| Sulfadimethoxine     | 100–1600                    | 100–1600          | >1600         | >1600         | 42 (37.8) 1 (10) 0 (0) 43 (35.2) |
varies between regions. In the Netherlands, the predominant serovar changed from Typhimurium in 1984–1989 to Enteritidis in 1996–2001 [14]. In Japan, S. enterica Infantis was likely to be the predominant serovar among broilers around 1997 [15]. S. enterica Infantis, with similar pulsed field gel electrophoresis profiles and resistance patterns, has been prevalent in Japanese broiler flocks for some time [16].

In several countries, S. enterica Typhimurium is the leading serovar in the isolates from pigs [17]. Futagawa-Saito et al. [9] showed that the predominant serovars were Agona (28.4%) and Typhimurium (17.9%) in 1998–1999 and Typhimurium (32.5%) and Anatum (17.9%) in 2004–2005. Kishima et al. [10] also showed that untypeable O4,12:d:- was most frequently found in 29.1% (50/172) of all isolates, followed by serovar Typhimurium (15.1%) in 2003–2005. Our previous study showed that S. enterica Typhimurium is the leading serovar in the isolates from diarrheic pigs in 1996–2001 [18]. Thus, S. enterica Typhimurium is likely to be predominant in Salmonella isolates from pigs in Japan.

The antimicrobial resistances patterns of the isolates are shown in Table 2. Higher resistance rates were observed against oxytetracycline (82.0%), dihydrostreptomycin (77.9%), kanamycin (41.0%) and trimethoprim (35.2%). Resistance rates to ampicillin, cefotiofur, bicozamycin, chloramphenicol and nalidixic acid were <10%. The MICs of apramycin, gentamicin, colistin, enrofloxacin, ofloxacin, and sulfadimethoxine were distributed unimodally. Two S. enterica Senftenberg isolates with MIC values higher than the breakpoint concentration (6.25 μg/ml) for cefotiofur were obtained from a broiler. The CTX-M-2 β-lactamase gene was detected in the cefotiofur-resistant isolates. In Japan, cephalosporins are not approved for the disease treatment in poultry. To date, extended spectrum β-lactamase (ESBL)-producing E. coli harboring the CTX-M-2 or CTX-M-18 β-lactamase has been obtained from broilers [7] and cattle in Japan [19]. ESBL-producing S. enterica Senftenberg obtained from river water was reported in Japan [20]. The β-lactamase gene type in these isolates was CTX-M-3 [20]. In addition, S. enterica Infantis strains resistant to cephalosporin were isolated from retail meats of domestic poultry in 2001–2003 [21] and in 2004–2005 [22]. Taguchi et al. [22] demonstrated that the cephalosporin-resistant S. enterica Infantis produced CMY-2 β-lactamase. The present study indicated that the CTX-M-2 β-lactamase producing S. enterica Senftenberg was prevalent in broiler chickens on the farm investigated before 1999. Thus, various types of β-lactamase producing Salmonella is found in the broiler chickens and the retail chicken meats in spite of non-approval for usage of cephalosporin antibiotics in chickens under the Japanese Pharmaceutical Affairs Law. We must continue to monitor the prevalence of cephalosporin-resistant Salmonella and should clarify the reasons why the resistant Salmonella has been prevalent in broiler industries.

Most S. enterica Infantis isolates (91.3%) exhibited resistance to two or more of the tested antimicrobials (Table 3). In this study, S. enterica Infantis was the most frequently found serovar in broiler isolates obtained by first enrichment and DSEC. Resistance to dihydrostreptomycin, kan-

### Table 3: Antimicrobial resistance patterns of S. enterica Infantis by isolation methods

| No. of antimicrobials | Antimicrobial resistance patterns a | Broilers | Total (%) |
|-----------------------|----------------------------------|----------|-----------|
|                       |                                  | FEb      | DSECc     |           |
| 0                     | Susceptible                       | 2 (5.3)  | 0 (0)     | 2 (2.9)   |
| 1                     | DSM                              | 2 (5.3)  | 0 (0)     | 2 (2.9)   |
|                       | OTC                              | 2 (5.3)  | 0 (0)     | 2 (2.9)   |
| 2                     | DSM, OTC                         | 9 (23.7) | 8 (25.8)  | 17 (24.6) |
| 3                     | DSM, KM, OTC                     | 7 (18.4) | 2 (6.5)   | 9 (13.0)  |
|                       | DSM, OTC, TMP                    | 4 (10.5) | 2 (6.5)   | 6 (8.7)   |
|                       | KM, OTC, TMP                     | 1 (2.6)  | 0 (0)     | 1 (1.4)   |
| 4                     | DSM, KM, OTC, TMP                | 11 (28.9)| 19 (61.3) | 30 (43.5) |
| Total                 |                                  | 38 (100) | 31 (100)  | 69 (100)  |

a DSM: dihydrostreptomycin, KM: kanamycin, OTC: oxytetracycline, TMP: trimethoprim
b FE: First enrichment culture
c DSEC: Delayed secondary enrichment culture
amycin, oxytetracycline and trimethoprim was found in 28.9% of the isolates by first enrichment, but in 61.3% by DSEC (Table 3). S. enterica Infantis isolates with similar resistance patterns were isolated using the two methods, although it is likely that there was a difference in the proportions of resistant isolates between the two methods. 

This study showed the prevalence of Salmonella in apparently healthy food-producing animals in Japan. In addition, CTX-M-2-beta-lactamase-producing S. enterica Senftenberg was isolated from broilers for the first time in 1999.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
KI provided data, discussed the results gained, and drafted. TT, AM, AK, KM, and TY provided data, discussed the results gained, and participated in revising the manuscript. TA discussed the results gained and revised the manuscript. All authors read and approved the final manuscript.

Acknowledgements
We thank the staff of the Livestock Hygiene Service Centers across Japan for sampling. This work was supported in part by grant-in-aid of Ministry of Health, Labor and Welfare (H21-Shokuhin-Ippan-013).

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