Specifics of vitrification of in vitro-produced cattle embryos at various development stages

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Introduction

In 2020, the world’s population reached 7.7 B people, and is expected to increase up to 8.9 B in 2035 and 9.2 B in 2050 (Raney et al., 2009). This, in turn, would require significant increase in the provision of food supplies. Improvement of agricultural methods becomes increasingly important for food provision in the world, for animal protein is first of all necessary for the correct cognitive development of children. As a result of increase in human population and demand for limited resources, the cost of energy carriers and concentrates would increase. The elements most economically impactful on profitability of livestock breeding sphere are fodders, which account for over 40% of the production costs. The main contribution to the stability of the sphere is efficiency in fodder consumption, since it affects economic and ecological parameters (Siberski-Cooper & Koltes, 2021).

The global changes in the environmental conditions due to climate change (Petrescu-Mag et al., 2022) makes it necessary for us to pay more attention to new approaches to managing herds, especially aspects of optimizing the efficiency of using the generative resources. Cattle-holding industries would have to search for new methods of reducing emissions of greenhouse gases during the production cycle by increasing the effectiveness of the production and decreasing wastes, or seek for new technologies to minimize contamination. In this context, there is a need of new indicators that could have been used in management and breeding of herds (Egger-Danner et al., 2015). Therefore, in particular, increases in milk production per dairy cow have led to decrease in the need of energy per unit of produced milk, and therefore lessened CH4 emissions per unit of milk (Capper et al., 2009; Gerber et al., 2013). However, further progress is possible only by introducing new developments, including those that improve genetic traits of herd (Lovendahl et al., 2018). Inclusion of functional traits in the program of genetic improvement is important for long-term development of dairy populations. Genetic achievement is cumulative, so insignificant improvements build up over time and thus would provide stable economy to all farmers, especially those using selection indices to combine data from multiple various traits (Egger-Danner et al., 2015). Nonetheless, changes in certain traits resulting from selection breeding are occurring at relatively low rate (0.5–3.0% per year) (Jennifer et al., 2013). At the same time, reproductive biotechnology that would induce changes could be implemented at fast rates by intensifying the selection, and not only reproduction rates. Therefore, the technology of
Currently, embryo transplantation is an important technology used for genetic improvement and cattle breeding programs (Doichi, 2019). There are two methods of obtaining cattle embryos: in vivo and in vitro. Until 2016, embryos have been predominantly produced in vivo (Viana, 2020), though scientists noted that producing oocytes by ovum pick up (OPU) makes it possible to obtain larger amounts of in vitro-derived embryos per animal compared with multiple ovulation and embryo transfer (MOET), which is due to variability in donors, which hinders the diagnoses of the number of in vivo embryos (Boland et al., 1991; Hasher, et al., 2003; Ferré et al., 2020). Therefore, there has been a rapid increase in the number of in vitro-produced cattle embryos around the globe, up to 1 M in 2019 against the background of decrease in the number of embryos produced in vivo (378,769 in 2019) (Viana, 2020). In order to manage such an amount of embryos, cryoconservation is used, which is a perfect tool logistics-wise, since transplantsologists can optimally choose the time of transplanting the embryo with reproductive cycle to the recipient and transport it large distances. Unfortunately, the cryosurvival level of in vitro-produced embryo is lower compared with embryos obtained in in vivo conditions (Moore & Hasler, 2017; Ferré et al., 2020; Viana, 2020).

Regardless of production method, all embryos that had not been transplanted to recipients are cryoconserved. The technology of embryo cryoconservation is one of the most important achievements in transplantation of cattle embryos (Hasler, 2014). The study of technology of freezing cattle embryos began in 1973, when Wilmut and Rowson obtained calves from frozen-defrozen embryos for the first time (Wilmut & Rowson, 1973). Currently, there are two protocols: slow freezing and vitrification (Valente et al., 2022). Slow freezing was developed in the 1970s (Hasler, 2014), whereas vitrification was for the first time performed in 1985 (Rall & Fahy, 1985). Slow freezing using low concentration of permeating cryoprotectant is broadly used to store cattle embryos (Hasler, 2014; Junari Idrissi et al., 2021). Despite the reliability of protocols of cryoconservation of in vivo-derived embryos, they are less effective in the work with in vitro-produced embryos (Dobrinsky, 2002; Hasler, 2014). The vitrification methods are currently seen to be undergoing improvements (Hasler, 2014; Rierau et al., 2017; Arshad et al., 2021), and furthermore, this method of cryoconservation has several advantages specifically for in vitro-derived embryos. Secondly, rates of re-expansion, hatching of embryos produced by extracorporeal fertilization and the survival rate of vitrification-frozen embryos were higher than those of in vivo-produced methods (Nedambale et al., 2004; Mucci et al., 2006; Najafzadeh et al., 2021). Also, this technology reduces the cryosamage to oocytes and membranes of cattle (Vajta et al., 1998; Inaba et al., 2016; Junari Idrissi et al., 2021) and decreases the sensitivity of embryos to freezing (Mahmoudzadeh et al., 1994). Another peculiarity of vitrification is that it prevents the formation of ice crystals during the freezing process by increasing the rates of freezing and warming (Vajta et al., 1998; Estudillo et al., 2021). Thirdly, vitrification is cheaper than stepwise methods (Martinez-Rodero et al., 2021). Fourthly, it is very simple to perform, does not require complicated equipment, and the technology is easy to learn over a short practical course (Campos-Chillon et al., 2006). Moreover, frequency of pregnancy of recipient cows in the conditions of using vitrified in vitro-derived embryos is appropriate for commercial purposes (Liebermann et al., 2006; Arshad et al., 2021).

Vitrification of embryos at various stages of the development is being debated (Juanparich et al., 2018; Do, & Taylor-Robinson, 2020). Using cryoconservation of blastocysts, embryos can be transferred right after freezing, avoiding the re-cultivation of embryos in in vitro conditions (Acino et al., 2013). Aagast et al. (2012) found that 5–8-cellular embryos stopped developing, whereas 8–16-cellular morulas became blastocysts, which is explained by the influence of the development synchronicity. Shirazi et al. (2009) demonstrated that early blastocysts have higher cryotolerance than the expanded ones. However, Sommerfeld & Niemann (1999) indicated that expanded blastocysts survive after vitrification better than non-expanded – 42% and 12%, respectively. On the other hand, cryoconservation of embryos at early stage slows the in vitro production of embryos, since it is important that those embryos continue to be in vitro cultivated after thawing, developing to the blastocyst phase, when they can be transplanted to recipients (Do & Taylor-Robinson, 2020).

Despite the fact that many aspects of cryoconservation of cattle embryos have been described in the literature, the recent interest in this problem coming from both scientists and entrepreneurs has led to emergence of new commercial solutions that facilitate the cryoconservation procedure and increase the survival rate of the embryos.

Because of absence of scientific data on the effects of equilibration period of cattle embryos on their survival and further development after using commercial kit Cryotech Vitrification Kit for vitrifying human embryos (Cryotech, Japan), we conducted vitrification of cattle embryos at various development stages so as to determine the optimal equilibration time.

Materials and methods

Production and maturation of oocyte-cumulus complexes. The ovaries from clinically healthy cows were selected in the slaughter house and transported in a thermos at the temperature of 30–33 °C no more than 3 h after the extraction. In the laboratory, the ovaries were 4 times rinsed in sterile Dulbecco’s Phosphate Buffered Saline (Sigma, USA) with addition of 0.075 mg/cm³ of kanamycin sulfate (Sigma, USA) (the temperature of the solution was 37–38 °C). Removal of oocyte-cumulus complexes from the antral follicles (2–8 mm in size) of the ovaries of cows was performed in a laminar box by cutting the follicles with a safe blade in the oocyte collecting solution comprising 5 cm³ of TL HEPES (Minitube, Germany) and with addition of 30 mg of bull serum albumin (Sigma, USA). Removal of oocyte-cumulus complexes, their selection and maturation, insemination and further cultivation were also carried out in the conditions of sterile box. After evaluation under SZ51 stereo-microscope (Olympus, Japan), we selected 120–130 µm of oocyte-cumulus complexes, with entire dense cumulus, undamaged transparent membrane and homogenous non-vacuolated ooplasm of regular round shape, without noticeable morphological signs of atresia. The extracted oocyte-cumulus complexes were 6 times rinsed in the solution for collecting oocytes. The oocyte-cumulus complexes were selected and rinsed on a heating table at the temperature of 37 °C. The oocytes had been maturing in in vitro conditions for 22–24 h in 4-well dishes (Oosafe, USA). Into each well, we added 300 mm³ of the solution comprising 4.5 cm³ of the initial TCM 199 (Minitube, Germany) solution for maturation, 0.5 cm³ of estrous cow serum, 12.5 IU of follicle-stimulating hormone and 1.25 IU of luteotropic hormone (50 mm³ of Phutat (Laboratories Calier S.A., Spain) and 0.125 IU of follicle-stimulating hormone (FSG-super (Agrobiomed, Russian Federation) and 50 mm³ of anticymotic antibiotic (Sigma, USA), and then it was covered by mineral oil (Origio, Denmark), added 25 oocyte-cumulus complexes and cultivated in a CO₂-incubator at the temperature of 38.5 °C and 6% CO₂ and 5% O₂.

Preparing sperm to insemination. Preparation of bull’s sperm to insemination was performed using density gradient Origio Gradient Series (Origio, Denmark) and solution for capacitation of spermatozoids (Minitube, Germany). Constituents of gradient were heated up to the room temperature (20–25 °C), and all other reagents were equilibrated in a CO₂-incubator at the temperature of 38.5–39.0 °C and 5% CO₂ for no less than 2 h. The gradient was prepared by carefully layering 1 cm³ of Origio Gradient 80 on 1 cm³ of Origio Gradient 40, and then sperm that had been defrozen on a water bath was carefully introduced. The formed system was centrifuged at centrifugal force of 300 g for 20 min. The supernatant was removed and the sediment was transferred using a sterile nozzle to a test tube with 2 cm³ of the solution for preparation and capacitation of spermatozoids, consisting 5 cm³ of basic solution for capacitation (Minitube, Germany); 30 mg of bovine serum albumin (Sigma, USA), 0.55 mg of sodium pinitrate (Sigma, USA) and 50 mm³ of anticymotic antibiotic (Sigma, USA), and then was centrifugated at centrifugal force of 300 g for 5 min, and most of the supernatant was removed. The procedure was repeated twice. After rinsing, the sediment was transferred to the bed of the test tube with 1 cm³ of new portion of the solution for preparation and capacitation of spermatozoids. Obtaining mobile fraction of spermatozoids was performed using the swim-up method described by Parrish et al.
(1986). One-hour incubation was enough for mobile spermatozoids to elevate to the upper layers of the solution, while dead and pathological ones remained at the bottom of the test tube. Mobile spermatozoids were capacitated in solution for preparation and capacitation in the conditions of 4 h influence of heparin (Sigma, USA) in the concentration of 20 μg/cm³ in the CO₂-incubator at the temperature of 38.5 °C with 6% CO₂ and 5% O₂. After capacitation, the spermatozoids were centrifugated at centrifugal force of 200 g for 5 min. We removed the supernatant and added 1 cm³ of the solution for insemination and determined the concentration of spermatozoids in Goryaev’s chamber hemocytometer.

**Co-cultivation of oocytes and spermatozoids.** After maturation, the oocytes were co-cultured with spermatozoids. Insemination of oocytes was carried out in 4-well dishes (Oosafe, USA) in 300 mm³ of solution composed of 5 cm³ solution for fertilization (Minitube, Germany), 30 mg of bovine serum albumine, 0.11 μg of sodium pyruvate, 0.2 mg of heparin and 50 mm³ of antimycotic antibiotic (Sigma, USA). It was coated by mineral oil (Origio, Denmark) for 18 h after the addition of capacitated spermatozoids (in calculation of 1×10⁶ mobile spermatozoids/cm³).

The number of oocytes in well varied 5 to 10.

**Cultivation of embryos.** After the co-cultivation, oocytes of cattle which had swollen expanded cumulus were freed from cumulus cells using the method of soft pipetting in 0.1% of hyaluronidase solution (Sigma, USA). Then, the oocytes were rinsed from the enzyme in 5-6 drops of TL HEPES (Minitube, Germany) and transferred to the cultivation solution, which was composed of 5 cm³ of growth medium with pyruvate (Minitube, Germany), 0.5 cm³ of estrous cow serum, 200 mm³ of essential amino acids (Sigma, USA), 50 mm³ of non-essential amino acids and 50 mm³ of antimycotic antibiotic (Sigma, USA). Embryos were cultivated in 65 mm³ of microdrops (3 embryos in one drop) under the layer of mineral oil (Origio, Denmark) in cultivation dishes (Oosafe, USA) in CO₂-incubation in the temperature of 38.5 °C with 6% CO₂ and 5% O₂. Inseminated oocytes were determined according to the division of 2-8-cellular stages 48 h after the contact with spermatozoids. Embryos reached the morula stage on the 3rd day of cultivation, and blastocysts on the 6–8th days.

**Vitrification.** Vitrification was performed for embryos of two development stages, according to the classification proposed by Bö & Mapleton (2013), recommended by the International Embryo Technology Society (IETS) (Rocha et al., 2016), morula – cycle day 4, stage code 3 (cellular mass comprised of no less than 16 cells, some blastomeres were hard to distinguish one from another, cellular mass of embryo accounted for most of the perivitelline space), quality code 1 (embryo had symmetric and spherical mass, which comprised separate blastomeres of the same size, colour and density, it corresponded to the expected development stage; irregularities were relatively insignificant, and at least 85% of the cellular material had undamaged vital embryonic mass; the zona pellucida was smooth and had no concavities or flat surfaces, Fig. 1) and blastocyst – cycle day 7, stage code 6 (we observed a notable differentiation of trophoblast – outer layer of cells and embryoblast – compact inner mass, blastocoeel was strongly expressed, embryo occupied most of the perivitelline space), quality code 1, Fig. 2).

Quality of embryos was evaluated under inverted microscope Zeiss Axio Observer A (Carl Zeiss, Germany), photos and measurements were taken using Octux NaviLase laser system (OCTAX Microscience GmbH, Germany).

In order to cryoconservate cattle embryos, we used Vitrification Kit 101 (Cryotech, Japan), which comprises equilibration solutions (ES) and vitrification solution (VS), 3-well dishes Vitr, Cryotech. Prior to the use, reagents were held for at least 1 h in the room temperature. Taking into account the data of Cryotech protocol, where maximal exposure time in the solution for equilibration was 15 min for oocytes and blastocysts and 12 min for other stages of embryonal development, we decided to compare the effects of 10, 12 and 15 min equilibration time on the further development of cattle embryos.

Prior to the start of cryoconservation procedure, the container for freezing was filled with liquid nitrogen and marked Cryotech. For the procedures with embryos, we used Stripper micropipette (CooperSurgical, USA) with flexible polycarbonate nozzles Flexipet (Cook, USA), 170 or 300 μm, depending on the size of embryo. The first well of the dish was filled with 300 μL of the equilibration solution (ES); the second and the third – 300 μL of the vitrification solution (VS). Using the pipette, we distributed the embryos on the surface of the equilibration solution. Because the objective of our study was determining the influence the equilibration time takes on the effectiveness of freezing, the morules (overall number – 120) and blastocysts (total of 120) were divided into 4 groups, with different time of embryo contacting ES – 10, 12 and 15 min, and embryos that were not subjected to vitrification (the control group). To confirm the significance of the obtained data, the experiment was repeated three times.

**Fig. 1.** Morula of cattle prior to vitrification (cycle day – 4, stage code – 3, quality code – 1): zona pellucida (a), compact cell mass (b)

**Fig. 2.** Blastocyste of cattle prior to vitrification (cycle day – 7, stage code – 6, quality code – 1): zona pellucida (a), blastocyst cavity (b), inner cell mass (c), trophoblast (d)

The next steps lasted over 25 s, but no longer than 90 s. After the equilibration, embryo aspiration was performed with a small amount of equilibration solution and transferred to another well (vitrification solution), descending the embryo on the middle depth of the solution. The pipette was rinsed from the solution. The embryo in this solution surfaced, and therefore it was 5–6 times suctioned with pipette and put onto the bottom of the well.

When the embryo stopped surfacing, it was transferred to the third well on the middle depth of the vitrification solution. It was rinsed with fresh vitrification solution and the mixture around the embryo was stirred, expecting its complete sedimentation on the bottom of the well. We suc-
tioned the embryo with vitrification solution on the pipette end and put it near the black marking on the Cryotec in minimum volume of liquid (less than 0.1 µL). We removed the excessive amount of the vitrification solution by putting the pipette end in the lower part of the large drop of the vitrification solution, moving the pipette in horizontal direction, thereby expanding the drop and carried out the aspiration of the excess of the liquid by decreasing the drop as much as we could. Under a stereomicroscope, we checked that embryo is on the Cryotec with minimal volume of vitrification solution and quickly submerged it into liquid nitrogen. Having not taking it out of nitrogen, we covered the Cryotec by a protective lid and transferred it into cryogenic storage dewar for maintenance.

Thawing. To thaw the cattle embryos, we used Warming Kit 102 (Cryotech, Japan), comprising solutions for thawing (TS), diluent (DS) and washing (WS), 4-well Warm Plate. Prior to using DS, WS, we held it for at least 1 h in the conditions of room temperature. Prior to use, TS was heated in a thermostat at 37 °C for 3 h. Into well No. 2, we introduced 300 µL of DS, into wells No. 3 and 4 – 300 µL of WS into each. We prepared Cryotec to thawing (we took the protective lid off using tweezers in the conditions of liquid nitrogen). We introduced TS into well No. 1 right before the thawing and quickly (in 1 s) submerged the Cryotec into well No. 1 and waited 1 min. Then, using Stripper micropipette (CooperSurgical, USA) with a flexible 170 or 300 µm polycarbonate nozzle, regardless of the embryo size, we transferred it to well No. 2 and left it there for 3 min. After the indicated period, the embryo was transferred to well No. 3 for 5 min, then – to well No. 4 for 1 min. The final stage of thawing was transferring the embryo into cultivation solution. After thawing, we evaluated the development status of embryo every 24 h.

The obtained results were statistically analyzed in ANOVA software, the data in tables are presented as x ± SE (mean ± standard error). To compare the difference between the parameters of the control and experimental groups, we used the Tukey test, where the differences were considered statistically significant at P < 0.05 for all the data.

Results

According to the results of the studies, we determined that the period of contact with equilibration solution in the vitrification conditions caused various levels of survival of the morules. Analysis of the parameters of blastulation and hatching of embryos revealed some differences in the development of embryos of the experimental groups 2 and 4, compared with the control (without freezing, Table 1).

Table 1

| Period of equilibration, min | Blastocysts on the 24th h after thawing, % | Blastocysts on the 48th h after thawing, % | Blastocysts that did not hatch, % |
|-----------------------------|---------------------------------------------|---------------------------------------------|-------------------------------|
| Control (without freezing)  | 43.3 ± 3.3a                                | 96.7 ± 3.3a                                 | 90.0 ± 5.7b                   |
| 10                          | 36.7 ± 3.3a                                | 90.0 ± 5.7b                                 | 83.3 ± 3.3c                   |
| 12                          | 46.7 ± 3.3a                                | 96.7 ± 3.3a                                 | 90.0 ± 0.0d                   |
| 15                          | 33.3 ± 3.3b                                | 83.3 ± 3.3b                                 | 73.3 ± 3.3b                   |

Note: letters indicate significant differences between the groups within a column (P < 0.05) according to the results of the Tukey test.

Equilibration period lasting 12 min appeared to be optimal for vitrification of cattle morules. We obtained 46.7 ± 3.3% of blastocysts in the 24th h after thawing and 96.7 ± 3.3% in the 48th h, which corresponded to the parameters of the control, equaling 43.3 ± 3.3% and 96.7 ± 3.3%, respectively. Hatching rate of blastocysts in this experimental group did not differ from those we observed for fresh non-vitrified embryos (90.0 ± 0.0% against 90.0 ± 5.7%, respectively). The obtained parameters indicate the high efficiency of vitrification of cattle morules using the Cryotec kit in the conditions of 12 min exposure to equilibration solution.

Two minute decrease in the equilibration period led to decrease in the parameters of the embryos’ development. Therefore, the percentage of blastulation was 10% lower on the 24th h after thawing, 6.7% lower on the 48th h, compared with group of embryos where the equilibration lasted 12 min. The percentage of hatched blastocysts was also 6.7% lower. At the same time, in the group where the embryos had been subjected to the action of equilibration solution for 15 min, we observed significantly lower percentage of formation of blastocysts in the 24th and 48th h after thawing – 33.3 ± 3.3% and 83.3 ± 3.3%, as well as hatching of blastocysts – 73.3 ± 3.3%, compared with the control. The data we obtained suggest that increase in the time of impact of equilibration solution on the monole had negative consequences on the development of embryos.

When determining the optimal period of equilibration for blastocysts of cattle, we analyzed the percentage of re-expanded blastocysts and hatching of embryos. We determined some differences in the development of embryos after 10 and 12 min contacts with the equilibration solution, compared with the control (without freezing, Table 2), whereas during 15 min equilibration, the parameters corresponded to the control: percentage of hatched blastocysts accounted for 93.3 ± 6.7% and 93.3 ± 3.3%, respectively. Optimal parameters indicate high efficiency of vitrification of cattle blastocysts using the Cryotec kit in the conditions of 15 min exposure to equilibration solution.

Table 2

| Equilibration period, min | Blastocysts with re-expanded blastocysts on the 24th h after thawing | Hatched blastocysts, % |
|---------------------------|-----------------------------------------------------------------------|------------------------|
| Control (without freezing)| -                                                                     | 93.3 ± 6.7b            |
| 10                        | 86.7 ± 8.8b                                                           | 76.7 ± 3.3b            |
| 12                        | 93.3 ± 6.7b                                                           | 83.3 ± 6.7b            |
| 15                        | 96.7 ± 3.3b                                                           | 93.3 ± 3.3b            |

Note: “–” – impossible to calculate, since re-expansion of the embryo is possible only in the conditions of its freezing; letters indicate significant differences between the groups within the column (P < 0.05) according to the results of the Tukey test.

Three minutes decrease in the equilibration period led to 3.4% decrease in the percentage of re-expansion of blastocysts (Fig. 3 and 4) and 10% decrease in hatching, compared with the 15 min equilibration. Ten minutes equilibration was less effective. The percentage of blastocysts with re-expanded blastocysts was 10% lower compared with the group where the equilibration lasted 15 min. At the same time, the percentage of blastocyst hatching in this experimental group was significantly lower than such observed for fresh non-vitrified embryos (76.7 ± 3.3% against 93.3 ± 6.7%, respectively). The obtained data suggest decrease in the efficiency of vitrification when shortening the action of equilibration toward blastocyst.

Discussion

In order to effectively introduce the vitrification technology into practice, the protocols used for thawing and transfer of cryoconserved embryos of cattle should be as simple as possible to implement and be efficient at the same time. Currently, slow freezing of embryos is becoming less popular in spite of vitrification. Stinshoff et al. (2011) in their studies indicate that vitrification of in vitro-obtained embryo is more effective than the slow freezing, since in the studies, the percentage of expanded blastocysts after thawing equaled 81.1 ± 20.3% and 79.4 ± 16.5%, respectively. Gupta et al. (2017) also demonstrated advantages of vitrification, for the percentage of formation of blastocysts after thawing of vitrified morules accounted for 77% compared with 34% after slow freezing.

The vitrification protocol is usually used for human embryos and is now being approved in animal husbandry. Vitrification is characteristic of both increase in rates of freezing and increase in cryoprotectant concentration. However, it has to be noted that most cryoprotectants have a number of negative effects such as toxicity and osmotic damages. Nonetheless, there is no unanimous opinion on toxicity of permeating cryoprotectants. Their toxicity was observed to increase over time with temperature and concentration (Wang et al., 2007; Martinez-Rodero et al., 2021). The toxicity of cryoprotectants is actually considered the single most limiting factor for the development of successful protocols of cryoconservation of cells and the tissues (Szurak & Eroglu, 2011). Therefore, any vitrification protocol must have adequate balance between the production of high level of dehydration, high viscosity and minimal toxicity. The first step of vitrification is the stage of equilibration in solution with relatively low concentration of cryoprotectant, the second – ultrashort exposure (30-90 s) to
vitrification solution with low concentration of cryoprotectant (usually double initial concentration) and the third – dehydration (usually, disaccharides are used, for example sucrose). Equilibration period varies broadly: it can be short (1 min) (Vieira et al., 2007; Sanches et al., 2016), last for 3 min (Gómez et al., 2020; Oliveira et al., 2020) or long (10–15 min) (Morató et al., 2010; Gutnisky et al., 2013).

Authors of many reports indicate the further development of the embryo as being dependent on the stage of its development during vitrification. Pagh et al. (2000) report equilibration of the embryos for 10 min in 10% ethylene glycol at room temperature, vitrified using one in 40% ethylene glycol, two in 25% ethylene glycol and 25% dimethyl sulphoxide or three in 20% ethylene glycol, 20% dimethyl sulphoxide and 10% L-lysine. Vitrification of cattle embryos is in the scope of studies by a large number of scientists, who propose different variants of cryoprotectant solutions to produce commercially appropriate results. Therefore, in the conditions of vitrification in TCM-199 solution with addition of Hepes/bicarbonate buffer, 20% calf serum, and also dimethyl sulphoxide and ethylene glycol as cryoprotectant and after thawing in 1 M solution of sucrose and further in vitro cultivation, Vajta et al. (1996) determined that 79% of blastocysts re-expanded and 59% hatched. They emphasize that the rates of re-expansion and hatching were different from blastocysts that had been vitrified on the 7th and 8th days (84 and 69% and 70 and 41%, respectively). Souza et al. (2018) consider the combination of 25% ethylene glycol, 25% dimethyl sulphoxide and 1.0 M sucrose optimal for vitrification of cattle blastocysts. It allowed them to obtain 75% of re-expanded blastocysts and 70% hatching. By using the new method of application of hollow fibre vitrification for cryopreservation of morules, Uchikura et al. (2016) observed blastocysts of vitrified embryos forming at rates (66.1–92.5%) similar to those of non-vitrified embryos (74.5–87.5%). By using straw vitrification dilution, Inaba et al. (2011) obtained 97.3% of hatched blastocysts on the 48th day of the cultivation.

As with the equilibration period, the protocols of commercial kits recommend it being dependent on recovery of embryo form, which could be evaluated by visualization under stereomicroscope. However, in practice, during embryo vitrification at the blastocyst stage, as well as morule, it is often hard to evaluate whether decrease or expansion has occurred, and additionally some bablastocysts do not decrease in the vitrification solution. Study of this stage of vitrification is important, for it provides adequate survival of the embryo, hatching of blastocysts and level of nutrition (Martínez-Rodero et al., 2021), because prolonged action of equilibration solution can be harmful to the potential of development of embryo, while insufficient action may influence the permeation of cryoprotectants into blastomeres. Vitrification of cattle embryos is in the scope of studies by a large number of scientists, who propose different variants of cryoprotectant solutions to produce commercially appropriate results. Therefore, in the conditions of vitrification in TCM-199 solution with addition of Hepes/bicarbonate buffer, 20% calf serum, and also dimethyl sulphoxide and ethylene glycol as cryoprotectant and after thawing in 1 M solution of sucrose and further in vitro cultivation, Vajta et al. (1996) determined that 79% of blastocysts re-expanded and 59% hatched. They emphasize that the rates of re-expansion and hatching were different from blastocysts that had been vitrified on the 7th and 8th days (84 and 69% and 70 and 41%, respectively). Souza et al. (2018) consider the combination of 25% ethylene glycol, 25% dimethyl sulphoxide and 1.0 M sucrose optimal for vitrification of cattle blastocysts. It allowed them to obtain 75% of re-expanded blastocysts and 70% hatching. By using the new method of application of hollow fibre vitrification for cryopreservation of morules, Uchikura et al. (2016) observed blastocysts of vitrified embryos forming at rates (66.1–92.5%) similar to those of non-vitrified embryos (74.5–87.5%). By using straw vitrification dilution, Inaba et al. (2011) obtained 97.3% of hatched blastocysts on the 48th day of the cultivation.

Authors of many reports indicate the further development of the embryo as being dependent on the stage of its development during vitrification. Pagh et al. (2000) report equilibration of the embryos for 10 min in 10% ethylene glycol at room temperature, vitrified using one in 40% ethylene glycol, two in 25% ethylene glycol and 25% dimethyl sulphoxide or three in 20% ethylene glycol, 20% dimethyl sulphoxide and 10% L-lysine. Vitrification of cattle embryos is in the scope of studies by a large number of scientists, who propose different variants of cryoprotectant solutions to produce commercially appropriate results. Therefore, in the conditions of vitrification in TCM-199 solution with addition of Hepes/bicarbonate buffer, 20% calf serum, and also dimethyl sulphoxide and ethylene glycol as cryoprotectant and after thawing in 1 M solution of sucrose and further in vitro cultivation, Vajta et al. (1996) determined that 79% of blastocysts re-expanded and 59% hatched. They emphasize that the rates of re-expansion and hatching were different from blastocysts that had been vitrified on the 7th and 8th days (84 and 69% and 70 and 41%, respectively). Souza et al. (2018) consider the combination of 25% ethylene glycol, 25% dimethyl sulphoxide and 1.0 M sucrose optimal for vitrification of cattle blastocysts. It allowed them to obtain 75% of re-expanded blastocysts and 70% hatching. By using the new method of application of hollow fibre vitrification for cryopreservation of morules, Uchikura et al. (2016) observed blastocysts of vitrified embryos forming at rates (66.1–92.5%) similar to those of non-vitrified embryos (74.5–87.5%). By using straw vitrification dilution, Inaba et al. (2011) obtained 97.3% of hatched blastocysts on the 48th day of the cultivation.

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Analysis of the described data may suggest that using Cryotec commercial solution for vitrification and thawing is more effective and simpler than schemes proposed by other scientists. In addition, our results of experimental studies are coherent with the data of Gutnisky et al. (2013), who – by using vitrification of blastocysts with Vitrification Kit 101 – achieved 100% survival of in vitro-derived cow embryos on the 7th day of the development.

**Conclusion**

Therefore, the study of vitrification of 4-day morules of cattle produced in in vitro conditions using Cryotec Vitrification Kit with 12 min...
exposure to equilibration solution revealed the results after thawing which were similar to each fresh non-vitrified morulas. Blastulation of 46.7 ± 3.3% in the 24th h after thawing, 96.7 ± 3.3% – in the 48th h and 90.0 ± 0.0% of hatched blastocysts corresponded to the parameters in intact group – 43.3 ± 3.3%, 96.7 ± 3.3% and 90.0 ± 5.7%, respectively. The best result of survival of in vitro-derived cattle blastocysts (7th day of cultivation) after vitrification using CryoTech Vitrification Kit was seen after equilibration period extended to 15 min. We obtained 96.7 ± 3.3% of blastocysts that recovered blastocoels after thawing on the 24th day and 93.3 ± 3.3% hatching, which corresponded to that in the intact group of blastocysts (93.3 ± 6.7%).

Thus, the obtained results indicate the high efficiency of CryoTech Vitrification Kit, which allowed the hatching of greater amount of embryos during the cultivation after thawing, and survival of vitrified embryos did not differ from the parameters of embryos without vitrification. The presented protocols of vitrification may have important consequences for commercial programs of transplantation of in vitro embryos, because they decrease the period that is necessary for vitrification of each individual embryo, and facilitate the use of this technique in production conditions.

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