The Effect of Different Routes of Injection of Bone Marrow Mesenchymal Stem Cells on Parotid Glands of Rats Receiving Cisplatin: A Comparative Study

Eman Hany¹, Mohammed A. Sobh², Mazen T. Abou ElKhier¹, Heba M. ElSabaa¹,³, Ahmed R. Zaher¹

¹Department of Oral Biology, Faculty of Dentistry, Mansoura University, Mansoura, Egypt
²Urology and Nephrology Center, Mansoura University, Mansoura, Egypt
³Department of Oral Biology, School of Dentistry, Badr University, Cairo, Egypt

Background and Objectives: Cisplatin is a powerful antitumor chemotherapeutic agent that is widely used in the treatment of many cancers but it has many side effects on many organs including salivary glands. Bone marrow is considered to be a rich environment that comprises many types of stem cells of which BMSCs (Bone marrow mesenchymal stem cells) are the most studied with potentiality to differentiate into many cell types. This study was conducted to evaluate the effect of different routes of injection of BMSCs on parotid glands of rats receiving cisplatin.

Methods and Results: Sprague-Dawley rats were divided into 3 groups: a negative control group receiving phosphate buffered saline, a positive control group receiving cisplatin, and an experimental group where rats received cisplatin and then received iron oxide-labeled BMSCs by either intravenous or intraparotid routes or both. Animals were sacrificed at periods of 3,6,10 and 15 days after cisplatin injection, then histological, ultrastructural and immunohistochemical studies were done. The experimental stem cell treated group showed better histological features and increased PCNA proliferation index when compared to the control. The systemic and combination groups showed better results than the local group. Iron oxide-labeled cells were detected with Prussian blue stain.

Conclusions: This study proved that BMSCs can improve cisplatin induced cytotoxicity in parotid glands. Systemic administration showed to have a better effect than local intraparotid administration and comparable effect to combined administration.

Keywords: Parotid gland, Chemotherapy, Bone marrow, Stem cells, Transmission electron microscopy, Immunohistochemistry

Introduction

The parotid gland is a purely serous gland that secretes about 25 to 35 percent of the whole daily salivary secretions (1). Cisplatin (cis-diammine dichloro platinum II, CDDP) is an effective, important antineoplastic drug which is applied for the managing of many types of tumors (2). It has general cytotoxic as well as many oral side effects like xerostomia, mucositis, bleeding tendency and dental infections (3). Cisplatin was also shown to affect submandibular as well as parotid salivary glands (4, 5).

Stem-cell therapy is a growing promising approach to treat many diseases and as a source of stem cells, adult
Bone marrow is considered to be a reliable, functional source of stem cells that can self-renew, proliferate and differentiate into many cell types (6). Bone marrow mesenchymal stem cells (BMSCs) were proven to retain a wide range of differentiation capabilities in vitro that is not restricted to mesodermal tissues but includes tissues normally derived from other embryonic germ layers like endoderm-derived hepatocytes (7) and ectoderm-derived neurons (8).

BMSCs were proven to enhance tissue regeneration after cisplatin-induced cytotoxicity in many organs like kidneys (9) and ovaries (10). They also showed the ability to increase the proliferation capacity of injured cells in different tissues (9, 11-13). This may be mediated by the paracrine release of cytokines and growth factors like IL-6 (Interleukin-6), HGF (Hepatocyte growth factor) and VEGF (Vascular endothelial growth factor), and anti-apoptotic factors like caspase-3 (11).

Almost all studies regarding bone marrow stem cell therapy for salivary glands were concerned only with the damaging effect of radiotherapy on the submandibular glands (12, 14, 15) while none studied its effect on chemotherapy-induced salivary gland injury and no available studies on parotid glands could be found so far, neither by systemic nor local routes. Also no studies detected the effect of combined treatment with both local and systemic routes together on salivary glands or any other organs. So, this work will be conducted to evaluate the effect of different routes of injection of bone marrow stem cells on parotid glands of rats receiving cisplatin histologically, ultrastructurally and immunohistochemically.

Materials and Methods

All experimental procedures were taken under a protocol approved by the Ethical Committee of Faculty of Dentistry, Mansoura University and according to the ARRIVE guidelines (Animal Research: Reporting In Vivo Experiments) for reporting animal research.

Rat bone marrow mesenchymal stem cells harvesting
Bone marrow mesenchymal stem cells were obtained from dissected femur bones of adult Sprague-Dawley rats as described by Sumita et al. (12) where Dulbecco's Modified Eagle's Medium (DMEM) was used containing 10% fetal bovine serum (FBS) obtained from Lonza (Verviers, Belgium), with antibiotic-antimycotic reagent, L-glutamine and D-glucose at 37°C in a humidified atmosphere that contained 5% CO2. Cells were used for the experiment after the third passage. Mesenchymal stem cells features were demonstrated by typical spindle-shaped morphology and adherence to plastic walls. Cells were also characterized by fluorescence-activated cell sorting (FACS) analysis as described by Lotfy et al. (16). BMSCs were negative for the hematopoietic lineage marker CD45 and positive for CD29 and CD90. Fluorescent labeled antibodies were obtained from eBioscience (San Diego, CA, USA).

Stem cells labeling
Cells were labeled by Sacrofere (Iron sucrose) from Amoun Pharmaceutical Co. (El Obour city, Cairo, Egypt). For this, 20 μg/ml iron was added to the cell medium two hours before transplantation. Some Iron oxide-labeled cells were tested for positive reaction with prussian blue (PB) on histological slides before transplantation into rats. Almost all of the labeled cells showed positive staining.

Animals and study design
One hundred and forty male Sprague-Dawley rats of 200~250 g weight (5~6 months of age) were kept under standard housing conditions in cages of 6 rats each with normal access to food and water in the animal house in Mansoura Experimental Research Center (MERC). Animals were acclimatized for at least 2 weeks before starting the study and then they were randomly divided into 3 groups as follows:

**Group I (PBS group):** Sixty rats received Phosphate buffered saline (PBS) and considered as negative control. They were divided into 3 subgroups of twenty rats each as follows:
- **Subgroup A (Local transplantation group):** Received 0.5 ml of PBS injected into the parotid gland.
- **Subgroup B (Systemic administration group):** Received 0.5 ml of PBS injected into the tail vein.
- **Subgroup C (Combination group):** Received both intraparotid and systemic injections with the same doses as subgroups IA and IB.

**Group II (Cisplatin group):** Twenty rats received single intraperitoneal injection of Cisplatin drug (Hospira, UK Limited, Warwickshire) (10 mg/kg) each (17) and considered as positive control.

**Group III (Experimental group):** The rest of the animals (60 rats) received the same dose of Cisplatin and were divided into three subgroups each of twenty rats and received stem cell therapy as follows:
- **Subgroup A (Local transplantation group):** Received intraparotid transplantation of BMSCs (5×10^5 cells) (18) suspended in 0.5 ml of PBS at day 1 after cisplatin administration.
Subgroup B (Systemic administration group): Received intravenous injection of BMSCs (2×10⁶ cells) (19) suspended in 0.5 ml of PBS via the tail vein at day 1 after cisplatin administration.

Subgroup C (combination group): Animals of this group received both systemic and local stem cell therapy with the same doses as subgroups IIIA and IIIB.

Five rats of each of the three groups (I, II, III) were sacrificed at days 3, 6, 10 and 15 after Cisplatin or PBS administration.

Histological evaluation

Animals were euthanized with overdose of halothane at the specified periods then the parotid glands were excised and processed for:

1) Haematoxylin and Eosin staining as a routine stain.
2) Immunohistochemical staining: with anti PCNA (Proliferating cell nuclear antigen) antibody to detect cell proliferation.
3) Prussian blue staining: to detect the iron oxide labeled BMSCs in the tissues of the parotid gland.
4) Transmission electron microscopic study.

Tissue processing

For heamatoxylin and eosin, immunohistochemical and Prussian blue staining the parotid gland specimens were fixed in 10% formaldehyde diluted with phosphate buffered saline and processed with an automated tissue processor. Specimens were embedded in paraffin blocks and cut into sections of 4 μm. For each histological block, several sections were prepared for examination.

Heamatoxylin and eosin staining: Tissue sections were deparaffinized with xylol, rehydrated in descending grades of alcohol and stained with the basophilic heamatoxylin stain. Sections were washed under tap water and then stained with the acidophilic eosin stain.

Immunohistochemical staining: Sections were mounted on silane-coated glass slides, deparaffinized with xylol and then rehydrated. Endogenous peroxidase was blocked using H₂O₂ at 3% and antigen retrieval was done at high temperature with citrate buffer 0.01M (pH 6.0) according to the manufacturer's instructions. Slides were then incubated with the primary monoclonal PCNA antibody (Abcam, Cambridge, UK) and the secondary biotinylated antibody followed by Streptavidin-Biotin Peroxidase complex. Development with chromogen substrate diaminobenzidine (DAB) was done and finally the slides were counterstained with Harris hematoxylin. Negative controls were obtained using non-immune serum instead of the primary antibody. Positive reaction was seen as brown staining of the nuclei.

Digital image analysis: Five different fields of each slide were photographed using Olympus digital camera installed on Olympus microscope with 1/2 X photo-adaptor, using 40 X objective. The images were analyzed on Intel Core i3 based computer using Video Test Morphology software with a specific built-in automated object. The positive immuno-stained nuclei were counted regardless of intensity and then the percentage of positive nuclei of the total number of nuclei was calculated.

Prussian blue (PB) staining: Slides were kept for 20 minutes in a mixed solution of 20% HCL and 10% potassium ferrocyanide of equal proportions. They were then washed for 3 times then fast red was used as a counter-stain for 5 minutes. On examination, the cytoplasm showed a pink color, the nuclei red and the areas containing the ferric iron were bright blue.

Transmission electron microscopic study: Specimens were fixed for 1 hour in buffered glutaraldehyde (2.5%) at 4°C temperature followed by 2 hours in osmium tetroxide (1%). Dehydration was then done by immersion in ethanol of increasing concentrations. Specimens were then placed in propylene oxide, and then resin Epon 812 was used for embedding. Samples were cut into ultrathin sections (60 nm), contrasted with 4% uranyl acetate and Reynold’s lead citrate and finally examined by a transmission electron microscope.

Statistical analysis

PCNA (Proliferating cell nuclear antigen) labeling index data was tabulated, coded then analyzed using the computer program Statistical Package for Social Science (SPSS, Version 17.0, Chicago, IL, USA) to obtain descriptive data which was calculated in the form of Mean±Standard deviation (SD). Two way Analysis of Variance (ANOVA) test was used to detect the effect of different groups and different time periods on the PCNA proliferation index. Statistical significance (S) was considered when p-value was ≤0.05.

Results

In the present study, some animals died during the study course and were replaced to keep the number of each group.

Histological and ultrastructural results (Fig. 1, 2)

Group I: The salivary glands of the rats receiving PBS with different routes showed normal structure with rounded serous acini, pyramidal cells bordering narrow lumens,
basal spherical nuclei and regular duct systems. Nuclei were euchromatic with regular outlines. Densely packed homogenous secretory granules and rough endoplasmic reticulum (rER) with parallel condensed cisternae were observed.

**Group II:** Histological sections in parotid glands treated with cisplatin showed different signs of degeneration at all periods. The secretory acini showed ill-defined outlines and multiple intracytoplasmic vacuoles. The connective tissue septa appeared widened with fibrosis and inflammatory infiltrate and eosinophilic material condensation was observed between the acini. Vacuolation was also noted in the lining of some excretory ducts. Moreover, complete degeneration of whole acini leaving empty spaces could be observed.

By electron microscopy, some acinar nuclei appeared irregular and fragmented with condensed heterochromatin. Secretory granules were also non-homogenous with increased electrolucency. In addition, dilated rER cisternae and some degenerated mitochondria could be noticed.

**Group III**

**Local transplantation group:** Showed some improvement in the gland tissues if compared to the cisplatin group. At days 3 and 6 the salivary acini and nuclei still had irregular outlines with intracytoplasmic vacuolation and non-homogenous secretory granules. However, the nuclei showed some mitotic figures and the ducts appeared normal unlike the positive control group. At days 10 and 15 the histological appearance of the gland was more improved showing more regular secretory acini and nuclear outlines.

**Systemic administration group:** At day 3, results similar to the local group at the same period were found with no much obvious improvement, but starting from day 6 better acinar, nuclear morphology and secretory granules appearance was noticed with increased number of nuclei showing
signs of mitosis and less degenerative vacuolations. At day 15 much improvement was observed where the parotid sections appeared nearly normal with well-arranged acini and nuclei, regularly arranged sacculles of rER and electrodense homogenous secretory granules which are similar in shape and size.

**Combination group:** The glandular tissue showed regular, almost normal histological features starting from day 6 through 10 and 15 days.

**Immunohistochemical results (Fig. 3)**

**PBS group:** Positive immuno-labeled cells were randomly distributed in the gland.

**Cisplatin group:** The number of PCNA positive cells obviously decreased in all periods.

**Experimental group:** Positive acinar and ductal cells could be detected but the PCNA proliferation index was higher in the systemic and combination groups than the local or the cisplatin groups.

**Prussian blue staining results (Fig. 4)**

For the experimental groups only, some positive cells appeared in the parotid glands at all periods as granular blue or purple deposits.

**Statistical results**

Two way ANOVA (Analysis of Variance) test was used to detect the effect of different groups and different time periods on the PCNA proliferation index. Statistical significance (S) was considered when p value was ≤0.05.

The summary table (Table 1) shows a statistically significant interaction between the effects of groups and time on the proliferation index. The analysis also showed significance of the main effect of groups and the main effect of time.

The marginal means of different groups, as presented in Table 2, show that the cisplatin receiving group had much depressed proliferation index than the negative control PBS groups indicating the suppressive effect of cisplatin on the cell cycle. The local group showed higher proliferation than the cisplatin group but not reaching the
normal control levels. Systemic and combination groups had elevated proliferation indices than PBS groups may be to compensate the damage that occurred to the gland but their values were insignificantly different from each other. This shows that systemic and combination stem cell therapy had better effects in enhancing proliferation of the affected glands more than local therapy.

The marginal means of different time periods over groups as presented in Table 3 show that cellular proliferation in the glands increased through the experiment starting from day 3 to compensate for the cisplatin induced cellular damage where proliferation at day 6 and 10 was significantly higher than day 3 reaching its peak at day 10 then dropped back at day 15 to be insignificant from day 3.

The line graph in Fig. 5 shows the course of each group through the experiment at each time period. The PBS groups appeared to be stable at all periods. Cisplatin group had a very much lowered index that was almost static through different periods. Local administration group was below the normal PBS line but showed increased proliferation. Systemic and combination groups showed higher proliferation indices than the control PBS group from the beginning, reaching a peak and then coming down to approach its level.

Discussion

Cisplatin generates its antitumour effect by inducing cytotoxic changes leading to apoptosis or programmed cell death (20) by binding to cellular DNA (21), or mitochondrial DNA (22). It also induces reactive oxygen species production leading to increased lipid peroxidation and Ca\(^{2+}\) influx which can mediate apoptosis (23).

A known complication of cisplatin drug is acute kidney injury where administration of 5 ml/kg cisplatin in the abdominal cavity is associated with development of acute renal failure in rats within 72 hours (24). This may count for the death of some animals during the study course.

In the cisplatin control group, the histological and ul-
trastructural examinations showed many changes in the gland structure. These results agreed with the study by Kitashima who detected irregularity and swelling of the acinar cells with abnormal nuclei, large fused secretory granules with light and dark areas, enlarged fragmented rER, and swollen mitochondria in submandibular glands of rats receiving cisplatin (4).

Changes in the nuclear outlines and chromatin distribution as seen in the current study may be related to the mechanism of action of cisplatin where it affects the nucleus causing DNA platination and cytotoxic changes that may end with apoptosis. Typical morphological changes are characteristic for apoptosis as cell shrinkage, DNA fragmentation and chromatin condensation (25) similar to the changes observed in the present study. So, these changes might be signs of degeneration or a pre-apoptosis stage of acinar cells.

Intracytoplasmic vacuoles may be attributed to the damage of some intracellular structures and according to Kitashima (4), membranes of adjacent secretory granules can fuse forming intracytoplasmic vacuoles. Electronlucent secretory granules as those seen in the parotid gland sections in the present study are usually encountered in cases of sialedenosis and are thought to be signs of changes in the salivary protein component (26).

According to McInnes (27), eosinophilic materials, as those seen in the cisplatin control group, can be seen as an abnormal change in mice mucosal epithelium, submucosal glands and ducts, and are thought to be accumulation of proteinaceous materials in the tissues.

The proposed mechanisms by which bone marrow cells improve organ functions have been investigated over a long period. Initial reports proposed the ability of BM cells to (trans) differentiate into cells of a non-mar row/non-hematopoietic lineage (28). Then reports on fusion of BM cells with cells of other tissues were documented (29). The third proposed mechanism was vasculogenesis from endothelial progenitor cells contained within BM (30). Lastly, the most recent mechanism is that BM cells provide a local paracrine effect on endogenous cells that may be achieved by secreting anti-apoptotic, anti-inflammatory and proliferation promoting factors (31).

Table 1. Summary table of 2-way ANOVA analysis

| Source       | F       | P     |
|--------------|---------|-------|
| Groups       | 243.496 | <0.001*|
| Time         | 7.003   | <0.001*|
| Groups x Time| 5.679   | <0.001*|

P: probability.
*Significance ≤ 0.05.

Table 2. Marginal means of different groups over time and statistical relation between them.

|                  | Local PBS | Systemic PBS | Combined PBS | Cisplatin | Cisplatin+Local BMSCs | Cisplatin+Systemic BMSCs | Cisplatin+Combined BMSCs |
|------------------|-----------|--------------|--------------|-----------|-----------------------|--------------------------|--------------------------|
| Mean             | 14.41     | 14.51        | 14.64        | 1.64      | 9.67                  | 16.80                    | 17.47                    |
| ±SD              | 1.75      | 1.87         | 1.68         | 0.52      | 2.80                  | 3.37                     | 4.34                     |
| P1               | 1.00      | 1.00         | <0.001*      |           | <0.001*               | <0.001*                  | <0.001*                  |
| P2               | 1.00      | <0.001*      | <0.001*      |           | <0.001*               | <0.001*                  | <0.001*                  |
| P3               |           | <0.001*      | <0.001*      |           | <0.001*               | <0.001*                  | <0.001*                  |
| P4               |           | <0.001*      | <0.001*      |           | <0.001*               | <0.001*                  | <0.001*                  |
| P5               |           | <0.001*      | <0.001*      |           | <0.001*               | <0.001*                  | <0.001*                  |
| P6               |           |              |              |           |                      |                          | 1.00                     |

P1: significance relative to local PBS group, P2: significance relative to systemic PBS group, P3: significance relative to combined PBS group, P4: significance relative to cisplatin group, P5: significance relative to local BMSCs group, P6: significance relative to systemic BMSCs group. *Significance ≤ 0.05.
Table 3. Marginal means of time periods over groups and statistical relation between them

|          | 3 days | 6 days | 10 days | 15 days |
|----------|--------|--------|---------|---------|
| Mean     | 11.89  | 13.02  | 13.53   | 12.49   |
| ±SD      | 6.02   | 5.86   | 6.05    | 4.82    |
| P1       | 0.018* | <0.001*| 0.67    |
| P2       | 1.00   | 0.95   |         |
| P3       | 0.036* |        |         |

P1: significance relative to 3 days group, P2: significance relative to 6 days group, P3: significance relative to 10 days group.

Previous studies on the effect of local BMSCs transplantation into the submandibular glands of irradiated mice showed preserved acinar cells and gland morphologies along with improved gland functions as salivary flow rate and glandular weight (13, 14).

Similar studies were conducted by Tran et al. (15) and Sumita et al. (12) who injected BMSCs in tail veins of mice and rats with irradiated submandibular glands respectively. These studies detected better submandibular gland functions, increased proliferation, decreased apoptosis and increased vascularity as well.

In this study, it was noticed that parotid gland tissues started to show signs of recovery in the experimental local group at day 10 whereas in the systemic group starting from day 6. Moreover, at the end of the study; at day 15, the final histological picture of the gland sections treated with systemic BMSCs was more improved than the local intraglandular transplantation group. This may point to a better effect of systemic BMSCs administration than local transplantation route.

In consistency with our results, a study on the effect of different routes of BMSCs administration on a rat model of liver fibrosis compared between intraperitoneal, intrahepatic and intravenous routes and the results of the study showed the intravenous route to be the most effective in regeneration of the fibrosed liver functions and histology. The authors justified their results by detecting elevated levels of IL-10 in the intravenous group than the other 2 groups where they suggested that the presence of BMSCs in the venous blood stimulates IL-10 release which can modulate the host immune response and enhances the healing process (32). Also in accordance with our results, a study on the effect of BMSCs on persistent orofacial pain rat model detected improvement in both intravenous and local injection routes but the results of the systemic group lasted for about 5 months while the local lasted only for 1 month (33) which gives an advantage to the systemic over the local route.

Some studies detected equal results clinically and histologically as well as equal incorporation of BMSCs into the target tissue by both local and systemic ways (34, 35). On the other hand, some studies reported better results with local transplantation than systemic BMSCs administration in oral mucosal ulcers as well as bone defect injuries (19, 36). Also, the regenerative effect of intravenous administration of BMSCs or BM soup on irradiated mice submandibular glands was found to be comparable to the local intraglandular route but with four times the required dose (15).

Local administration of BMSCs has shown efficiency in healing different injuries in many tissues as fore-mentioned. However, local infusion is likely not clinically feasible in many cases due to its potentially high degree of invasiveness (e.g., into the heart or brain), and locally administered cells may die before significantly contributing to the healing response due to diffusion limitations of nutrients and oxygen (37). Moreover, according to many studies it was reported that the main problem facing the efficacy of local direct stem cell injections is the ability of the injected cells to reside in the injury site and that is why different types of carriers like injectable hydrogel carriers, tissue grafts and cell-sheets are used to enable stem cell localization (38). In addition to that, it was noted that in previous studies of local BMSCs administration on submandibular glands, the glands were surgically exposed first then injected directly (13, 15) but in this study for the parotid glands, the authors managed to assess the ability of the externally in-
jected BMSCs to reach the gland and contribute to its regeneration. Also, the number of injected stem cells in the local administration group was less than some of the previous studies. These findings may justify the less histological improvement noted in the local transplantation group in comparison to the other experimental groups.

As far as we know, no available studies using a combination protocol for stem cell therapy could be found but the results of the combination group are close to those of the systemic intravenous group which gives no advantage for using both delivery routes together, besides the risk of increasing the stem cells dose and increasing manipulation procedures for the recipient.

In the experimental local intraparotid BMSCs transplantation group, the PCNA proliferation index was increased compared to the control group. This is in accordance with the results by Lim et al. (13) who found increased proliferation index as marked by PCNA and decreased apoptosis index as marked by TUNEL in the irradiated submandibular gland tissues treated with intraglandular BMSCs.

In the systemic intravenous BMSCs administration group, the average number of positive immunolabeled PCNA cells was significantly increased indicating the better effect of intravenous than the intraglandular BMSCs delivery routes. Similarly, in the study by Sumita et al. (12), intravenous BMSCs administration led to increased PCNA proliferation marker in irradiated submandibular glands.

No previous studies detecting the delivery of stem cells to the parotid glands were found but other studies confirmed the ability of BMSCs to home to submandibular glands after local as well as systemic administration with different labeling methods (14, 39).

Iron oxide labeled stem cells were detected in all experimental groups at all periods. As far as we know, only one in vivo study was found using iron sucrose for labeling of MSCs which were injected into lapine intravertebral discs and were detected after sacrifice by prussian blue staining technique (40).

Lastly it was concluded that BMSCs can reach the parotid glands by local as well as systemic routes and can enhance the healing potential and the proliferative capacity of the parotid gland cells after cisplatin administration. These results can be subjected for further experimental trials before clinical application on patients receiving chemotherapy to ameliorate their salivary gland functions during treatment.

Acknowledgment
The authors thank Dr. Mohammed E. Helal and Dr. Mohammed E. Grawish, professors of Oral Biology, Faculty of Dentistry, Mansoura University for their assistance in editing and revising this manuscript.

Potential conflict of interest
The authors have no conflicting financial interests with respect to the authorship or publication of this article.

References
1. Alcamo IE. Anatomy coloring workbook. 2nd ed. New York: Random House; 2003. 222
2. Wang G, Reed E, Li QQ. Molecular basis of cellular response to cisplatin chemotherapy in non-small cell lung cancer (Review). Oncol Rep 2004;12:955-965
3. López BC, Esteve CG, Pérez GS. Dental treatment considerations in the chemotherapy patient. J Clin Exp Dent 2011;3:e31-e42
4. Kitashima S. Morphological alterations of submandibular glands caused by cisplatin in the rat. Kurume Med J 2005;52:29-38
5. Hey J, Setz J, Gerlach R, Vordermark D, Gernhardt CR, Kuhnt T. Effect of Cisplatin on parotid gland function in concomitant radiochemotherapy. Int J Radiat Oncol Biol Phys 2009;75:1475-1480
6. Weider SA, Donaldson C, Denning-Kendall P, Rice C, Bradley B, Hows JM. Adult bone marrow is a rich source of human mesenchymal 'stem' cells but umbilical cord and mobilized adult blood are not. Br J Haematol 2003;121:368-374
7. Yen MH, Wu YY, Liu YS, Rimando M, Ho JH, Lee OK. Efficient generation of hepatic cells from mesenchymal stromal cells by an innovative bio-microfluidic cell culture device. Stem Cell Res Ther 2016;7:120
8. Woodbury D, Schwarz EJ, Prockop DJ, Black IB. Adult rat and human bone marrow stromal cells differentiate into neurons. J Neurosci Res 2000;61:364-370
9. Qi S, Wu D. Bone marrow-derived mesenchymal stem cells protect against cisplatin-induced acute kidney injury in rats by inhibiting cell apoptosis. Int J Mol Med 2013;32:1262-1272
10. Liu J, Zhang H, Zhang Y, Li N, Wen Y, Cao F, Ai H, Xue X. Homing and restorative effects of bone marrow-derived mesenchymal stem cells on cisplatin injured ovaries in rats. Mol Cells 2014;37:865-872
11. Weil BR, Markel TA, Herrmann JL, Abarbanell AM, Meldrum DR. Mesenchymal stem cells enhance the viability and proliferation of human fetal intestinal epithelial cells following hypoxic injury via paracrine mechanisms. Surgery 2009;146:190-197
12. Sumita Y, Liu Y, Khalili S, Maria OM, Xia D, Key S, Cotrim AP, Mezey E, Tran SD. Bone marrow-derived cells rescue salivary gland function in mice with head and neck
irradiation. Int J Biochem Cell Biol 2011;43:80-87

13. Lim JY, Yi T, Choi JS, Jang YH, Lee S, Kim HJ, Song SU, Kim YM. Intraglandular transplantation of bone marrow-derived clonal mesenchymal stem cells for amelioration of post-irradiation salivary gland damage. Oral Oncol 2013;49:136-143

14. Lin CY, Chang FH, Chen CY, Huang CY, Hu FC, Huang WK, Ju SS, Chen MH. Cell therapy for salivary gland regeneration. J Dent Res 2011;90:341-346

15. Tran SD, Liu Y, Xia D, Maria OM, Khalili S, Wang RW, Quan VH, Hu S, Seuntjens J. Paracrine effects of bone marrow soup restore organ function, regeneration, and repair in salivary glands damaged by irradiation. PLoS One 2013;8:e61632

16. Lotfy A, Salama M, Zahran F, Jones E, Badawy A, Sobh M. Characterization of mesenchymal stem cells derived from rat bone marrow and adipose tissue: a comparative study. Int J Stem Cells 2014;7:135-142

17. Roldán-Fidalgo A, Martín Saldaña S, Trinidad A, Olmedilla-Alonso B, Rodríguez-Valiente A, García-Berrocal JR, Ramírez-Camacho R. In vitro and in vivo effects of lutein against cisplatin-induced ototoxicity. Exp Toxicol Pathol 2016;68:197-204

18. Jeong J, Baek H, Kim YJ, Choi Y, Lee H, Lee E, Kim ES, Hah JH, Kwon TK, Choi IJ, Kwon H. Human salivary gland stem cells ameliorate hyposalivation of radiation-damaged rat salivary glands. Exp Mol Med 2013;45:e58

19. Huang S, Xu L, Zhang Y, Sun Y, Li G. Systemic and local administration of allogeneic bone marrow-derived mesenchymal stem cells promotes fracture healing in rats. Cell Transplant 2015;24:2643-2655

20. Li XB, Schlesener IJ. Therapeutic effects of cisplatin on rat experimental autoimmune encephalomyelitis. Arch Immunol Ther Exp (Warsz) 2006;54:51-53

21. Cepeda V, Fuertes MA, Castilla J, Alonso C, Quevedo C, Pérez JM. Biochemical mechanisms of cisplatin cytotoxicity. Anticancer Agents Med Chem 2007;7:3-18

22. Fuertes MA, Castilla J, Alonso C, Pérez JM. Cisplatin biochemical mechanism of action: from cytotoxicity to induction of cell death through interconnections between apoptotic and necrotic pathways. Curr Med Chem 2003;10:257-266

23. Casares C, Ramírez-Camacho R, Trinidad A, Roldán A, Jorge E, García-Berrocal JR. Reactive oxygen species in apoptosis induced by cisplatin: review of physiopathological mechanisms in animal models. Eur Arch Otorhinolaryngol 2012;269:2455-2459.

24. Izuwa Y, Kusaba J, Horiuichi M, Aiba T, Kawasaki H, Kurosaki Y. Comparative study of increased plasma quinidine concentration in rats with glycerol- and cisplatin-induced acute renal failure. Drug Metab Pharmacokinet 2009;24:451-457.

25. Lee JH, Nakagawa T, Kita T, Kim TS, Iguchi F, Endo T, Shiga A, Lee SH, Ito J. Mechanisms of apoptosis induced by cisplatin in marginal cells in mouse stria vascularis. ORL J Otorhinolaryngol Relat Spec 2004;66:111-118

26. Satoh M, Yoshihara T. Clinical and ultrastructural investigation of sialadenosis. Acta Otolaryngol Suppl 2004; (553):122-127

27. McInnes EF. Background lesions in laboratory animals: a color atlas. China: Saunders Elsevier; 2012. 49

28. Grove JE, Bruscia E, Krause DS. Plasticity of bone marrow-derived stem cells. Stem Cells 2004;22:487-500

29. Quintana-Bustamante O, Alvarez-Barrientos A, Kofman AV, Fabregat I, Bueren JA, Theise ND, Segovia JC. Hematopoietic mobilization in mice increases the presence of bone marrow-derived hepatocytes via in vivo cell fusion. Hepatology 2006;43:108-116

30. Rajantie I, Ilmonen M, Alminiaite A, Ozerdem U, Alitalo K, Salven P. Adult bone marrow-derived cells recruited during angiogenesis comprise precursors for periendothelial vascular mural cells. Blood 2004;104:2084-2086

31. Zarjou A, Kim J, Traylor AM, Sanders PW, Balla J, Agarwal A, Curtis LM. Paracrine effects of mesenchymal stem cells in cisplatin-induced renal injury require heme oxygenase-1. Am J Physiol Renal Physiol 2011;300:F254-F262

32. Zhao W, Li JJ, Cao DY, Li X, Zhang LY, He Y, Yue SQ, Wang DS, Dou KF. Intravenous injection of mesenchymal stem cells is effective in treating liver fibrosis. World J Gastroenterol 2012;18:1048-1058

33. Guo W, Wang H, Zou S, Gu M, Watanabe M, Wei F, Dubner R, Huang GT, Ren K. Bone marrow stromal cells produce long-term pain relief in rat models of persistent pain. Stem Cells 2011;29:1294-1303

34. McFarlin K, Gao X, Liu YB, Dulchavsky DS, Kwon D, Arbab AS, Bansal M, Li Y, Chopp M, Dulchavsky SA, Gautham SC. Bone marrow-derived mesenchymal stromal cells accelerate wound healing in the rat. Wound Repair Regen 2006;14:471-478

35. Kwon DS, Gao X, Liu YB, Dulchavsky DS, Danyluk AL, Bansal M, Chopp M, McIntosh K, Arbab AS, Dulchavsky SA, Gautham SC. Treatment with bone marrow-derived stromal cells accelerates wound healing in diabetic rats. Int Wound J 2008;5:453-463

36. Aly LA, El-Menoufy H, Sadeq HS, Ragae A, Sabry D. Efficiency of systemic versus intraleisional bone marrow-derived stem cells in regeneration of oral mucosa following stem cell infusion. J Bone J Surg Am 2004;86-A:1541-1558

37. Abbas SA, Spanoudes K, O’Brien T, Pandit A, Zeoglisis DI. Assessment of stem cell carriers for tendon tissue engineering in pre-clinical models. Stem Cell Res Ther 2014;5:38

38. Schwarzer S, Huss R, Schulz-Siegmund M, Vogel B, Brandau WK, Ju SS, Chen MH. Cell therapy for salivary gland damage. Int J Stem Cells 2014;7:135-142

39. Zarjou A, Kim J, Traylor AM, Sanders PW, Balla J, Agarwal A, Curtis LM. Paracrine effects of mesenchymal stem cells in cisplatin-induced renal injury require heme oxygenase-1. Am J Physiol Renal Physiol 2011;300:F254-F262

40. Papadimitriou N, Li S, Barreto Henriksson H. Iron sucrose-labeled human mesenchymal stem cells: in vitro multilineage capability and in vivo traceability in a lapine xenotransplantation model. Stem Cells Dev 2015;24:2403-2412