Prolonged Mechanical Ventilation Induces Cell Cycle Arrest in Newborn Rat Lung

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Abstract

Rationale: The molecular mechanism(s) by which mechanical ventilation disrupts alveolar development, a hallmark of bronchopulmonary dysplasia, is unknown.

Objective: To determine the effect of 24 h of mechanical ventilation on lung cell cycle regulators, cell proliferation and alveolar formation in newborn rats.

Methods: Seven-day old rats were ventilated with room air for 8, 12 and 24 h using relatively moderate tidal volumes (8.5 mL.kg⁻¹).

Measurement and Main Results: Ventilation for 24 h (h) decreased the number of elastin-positive secondary crests and increased the mean linear intercept, indicating arrest of alveolar development. Proliferation (assessed by BrdU incorporation) was halved after 12 h of ventilation and completely arrested after 24 h. Cyclin D1 and E1 mRNA and protein levels were decreased after 8–24 h of ventilation, while that of p27Kip1 was significantly increased. Mechanical ventilation for 24 h also increased levels of p57Kip2, decreased that of p16INK4a, while the levels of p21Waf/Cip1 and p15INK4b were unchanged. Increased p27Kip1 expression coincided with reduced phosphorylation of p27Kip1 at Thr157, Thr187 and Thr198 (p<0.05), thereby promoting its nuclear localization. Similar -but more rapid- changes in cell cycle regulators were noted when 7-day rats were ventilated with high tidal volume (40 mL.kg⁻¹) and when fetal lung epithelial cells were subjected to a continuous (17% elongation) cyclic stretch.

Conclusion: This is the first demonstration that prolonged (24 h) of mechanical ventilation causes cell cycle arrest in newborn rat lungs; the arrest occurs in G₁ and is caused by increased expression and nuclear localization of Cdk inhibitor proteins (p27Kip1, p57Kip2) from the Kip family.

Introduction

Introduction of more gentle ventilation strategies -together with surfactant replacement and antenatal corticosteroids- has improved the survival rate of very premature infants. In parallel, the number of infants with ‘new’ bronchopulmonary dysplasia (BPD) [1] has also increased. Currently, infants born at ≤26 weeks of gestation are at the greatest risk of developing such ‘new’ BPD [2], a syndrome of arrested lung development with fewer and larger alveoli and dysmorphic vasculature [3]. BPD can no longer be considered only a pediatric disease because the substantial lung-function abnormalities -and significant symptoms- persist into adulthood [4,5,6]. The pathogenesis of BPD is incompletely understood and its treatment is empirical, but mechanical ventilation remains a major risk factor.

Lung development between 24–32 weeks of gestation is characterized by extensive vasculogenesis within the developing terminal saccules, followed by secondary crest formation as well as interstitial extracellular matrix loss and remodeling [7]. This tissue remodeling requires well-coordinated regulation of cell proliferation and apoptosis. Recent studies have shown that prolonged mechanical ventilation increases apoptosis and impairs alveolar septation in newborn mice [8], however the effect of mechanical ventilation on lung cell growth is mostly unknown. In vitro studies have demonstrated that mechanical stretch (5% elongation, 60 cycles per min, 15 min/h for 24 h) and oxygen (95%) can inhibit lung cell proliferation [9,10], but molecular mechanisms are yet to be determined. Cell proliferation is a precisely coordinated set of events regulated by interaction of gene products that activate or
suppress cell cycle progression. A series of cyclins and cyclin-dependent kinases (Cdk) act in concert to drive the cell forward through the G1, S and G2/M phases [11]. In mammalian cells, G1/S transition is an important checkpoint in the cell cycle. Entry into the cell cycle is initiated by mitogen-stimulated expression of D-type cyclins which activate Cdk4/6. Shortly thereafter, cyclin E expression is increased and cyclin E-Cdk2 complexes are formed, promoting entry into the S phase [12]. While cyclin-Cdk complexes positively drive progression of the cell cycle, Cdk inhibitors (CKI) negatively regulate progression by binding to and inactivating cyclin-Cdks [13]. There are two distinct CKI families in mammalian cells: INK4 proteins, which block the progression of the cell cycle by binding to either Cdk4 or Cdk6 and inhibiting the action of cyclin D; and, G1p/Kip proteins that inhibit a broader spectrum of cyclin-Cdk complexes [14,15,16].

In this study we determined the effect of prolonged (24 h) mechanical ventilation on lung cell cycle regulators, proliferation and alveolar formation in a newborn rat model [17]. We hypothesized that continuous cyclic (over)stretching of the primitive air sacs would adversely affect proliferation of lung cells by influencing cell cycle regulators.

Methods

Ethics statement

The study was conducted according to the guidelines of the Canadian Council for Animal Care and with approval of the Animal Care Review Committee of the Hospital for Sick Children (protocol #7217).

Animal preparation

Timed pregnant Wistar rats (Charles River, Oakville, Quebec, Canada) were allowed to deliver and immediately afterwards litters were reduced to 10 pups. Newborn rat pups were anesthetized by i.p. injection of 30 mg kg\(^{-1}\) pentobarbital and a tracheotomy was performed. The trachea was cannulated with a 1 cm 19G cannula and connected to a rodent ventilator (FlexiVent Scireq, Montreal, PQ). Spontaneously breathing, non-ventilated, littermates served as sham controls. One pup per litter was ventilated and a littermate was used as non-ventilated control. Isoflurane was used as general anesthesia during the ventilation period and 0.9% saline was used as sham controls. One pup per litter was ventilated and a littermate was used as non-ventilated control. Isoflurane was used as general anesthesia during the ventilation period and 0.9% saline (100 ml kg\(^{-1}\)/24 h) was administered subcutaneously by continuous infusion with a 27G needle to prevent dehydration. First rat pups at postnatal days 6, 7, 8, 9, 10 and 14 were ventilated to assess lung cell proliferation. For all subsequent experiments 7-day old rat pups were used. Preliminary experiments were performed to determine ventilator settings [18]. Starting from a normal respiratory rate for newborn rats (150 bpm), tidal volume was adjusted to achieve normal blood gas values after the ventilation period. Animals were monitored by ECG. Rectal temperature was maintained at 37°C using a thermal blanket, lamp and plastic wrap. At the end of the ventilation period a blood sample from the carotid artery was taken for blood gas analysis prior to euthanasia.

Mechanical ventilation

Rat pups were ventilated with room air and moderate-V\(\text{T}\) (8.5 ml kg\(^{-1}\) RR 150 min\(^{-1}\), PEEP 2 cm H\(\text{O}\)) for 8, 12 and 24 h. In a few cases, pups were ventilated for 4 h with high-V\(\text{T}\) (40 ml kg\(^{-1}\) RR 30 min\(^{-1}\), PEEP 2 cm H\(\text{O}\)). The 7-day old pups weighed 15.5–18.6 g. Dynamic compliance was estimated every 4 h from data obtained during a single-frequency forced oscillation manoeuvre, using a mathematical model-fitting technique according to the specifications of Scireq Inc. (Montreal, PQ). Two h before completion of ventilation pups were injected ip with 50 mg/kg 5-bromo-2-deoxyuridine (BrdU). At completion of ventilation a blood sample was taken from the carotid artery for blood gas analysis and the animals killed by exsanguination. Lung tissues were processed for histology or fresh frozen for molecular/protein analyses.

Histology

After flushing whole lungs were infused in situ with 4% (w/v) paraformaldehyde (PFA) in PBS with a constant pressure of 20 cm H\(\text{O}\) over 5 minutes to have equalized filling pressure over the entire lung. Under these constant pressure conditions the cannula was removed and the trachea immediately ligated. The lungs were excised and immersed in 4% PFA in PBS overnight, dehydrated in an ethanol/xylene series and embedded in paraffin. Sections of 5 µm were stained with hematoxilin and eosin or stained for elastin using acustain artrazine solution (Sigma, St. Louis MO, USA).

Immunohistochemistry

Following sectioning and antigen retrieval by heating in 10 mM sodium citrate pH 6.0, sections were washed in PBS and endogenous peroxidase was blocked in 3% (v/v) H\(\text{2}\)O\(_2\) in methanol. Blocking was done with 5% (w/v) normal goat serum (NGS) and 1% (w/v) bovine serum albumin (BSA) in PBS. Sections were then incubated overnight at 4°C with either 1:50 diluted mouse anti-BrdU (Boehringer Mannheim, Germany) or 1:400 diluted rabbit anti-phospho-histone H3 (Millipore, Billerica, MA, USA) antibodies (Lab Vision Corporation, Fremont, Canada). Biotinylated rabbit anti-mouse IgG or goat anti-rabbit IgG were used as secondary antibodies, respectively. Color detection was performed according to instruction in the Vectastain ABC and DAB kit (Vector Laboratories, Burlingame, CA, USA). All sections were counterstained with hematoxylin. For quantitative analysis, digital images were captured using a Leica digital imaging system at 20× magnification with random sampling of all tissue in an unbiased fashion. Images were captured randomly from 15 non-overlapping fields from each slide, with 3 slides per animal and 4 animals per group.

Morphometric analysis

Digital images were captured from either H&E- or elastin-stained slides with random sampling of all tissue in an unbiased fashion. Images were captured randomly from 15 non-overlapping fields/slide with 3 slides/animal and 4 animals/group. Tissue fraction was calculated from pixel counts of black/white images [19], mean linear intercepts (Ln) were measured and calculated [20] and the number of elastin-positive secondary septa determined.

Western blot analysis

Lung tissues were lysed, protein content measured [21] and aliquots (40 g protein) were subjected to 10% SDS-PAGE and transferred to PVDF membranes. After blocking with 5% (w/v) skim milk in TBST (20 mM Tris, 137 mM NaCl, 0.1% Tween 20) membranes were incubated with appropriate primary antibody overnight in 4°C. Because of decreased BrdU incorporation and cyclin D1 and E1 expression, we focused on CKIs inhibiting Cdk2, -4 and -6 [22]. Primary antibodies were rabbit anti-p15INK4B (dilution of 1:500), rabbit anti-p16INK4A (dilution of 1:1000), mouse anti-p21\(^{\text{Waf1/Cip1}}\) (dilution 1:500), mouse anti-p27\(^{\text{Kip1}}\) (dilution 1:500) and rabbit anti-p57\(^{\text{Kip2}}\) (dilution of 1:1000), rabbit anti-cyclin D1 (dilution of 1:1000) (all from Cell Signaling Technology, Danvers, USA) and rabbit anti-cyclin E1 (dilution of 1:1000) (Santa Cruz Biotechnology, Santa Cruz USA). Primary phosphorylated
p27\textsuperscript{Kip1} antibodies were rabbit anti-p27\textsuperscript{Kip1} (pThr198) (dilution of 1:400) and rabbit anti-p27\textsuperscript{Kip1} (pSer10)-R (dilution of 1:2000; both from Santa Cruz Biotechnology, Santa Cruz, USA), rabbit anti-p27\textsuperscript{Kip1} (pThr157) (dilution of 1:300; R&D Systems Inc, Burlington, Canada) and rabbit anti-p27\textsuperscript{Kip1} (pThr167) (dilution of 1:800; Novus Biologicals, Littleton, USA). The next day the membranes were washed TBST and incubated with either horseradish peroxidase-conjugated anti-rabbit or anti-mouse IgG (dilution of 1:1000; Cell Signaling Technology, Danvers, USA). After several washes with TBST, protein bands were visualized using an enhanced chemiluminescence detection kit (Amersham, Piscataway, NJ, USA). Equal protein loading was confirmed by immunoblotting for \textbeta-actin of same membrane.

Quantitative RT-PCR

Total RNA was extracted from lung tissues and reverse transcribed [14]. Complementary DNA was amplified for target genes cyclin D1, cyclin E1 and p27 as previously described [17,19]. For relative quantification, polymerase chain reaction signals were compared between groups after normalization using 18S as internal reference. Fold change was calculated [23].

Stretch of epithelial cells isolated from fetal rat lungs

Distal fetal lung epithelial cells (day 19 of gestation) were isolated as previously described [24]. Cells were cultured on type-1 collagen-coated BioFlex plates and subjected various durations of cyclic continuous 17% stretch using a FX-4000 Flexercell Strain Unit (Flexercell Int., NC, USA) [25]. Neither cell viability (trypan blue exclusion) nor cell attachment was affected by the duration of applied stretch regimen. Cell lysates were processed for Western Blotting.

Statistical analysis

Stated otherwise all data are presented as mean ± SD. Data was analyzed using SPSS software version 15 (SPSS Inc, Chicago, IL). Statistical significance (p<0.05) was determined by using one-way ANOVA or Kruskal-Wallis test. Post hoc analysis was performed using Duncan’s multiple-range test (data presented as mean ± SD) or Mann-Whitney test (data presented as median and interquartile range).

Results

Physiologic data

Blood gases were in the normal range after 8, 12 and 24 h of ventilation (Table 1). Mean airway pressures, peak pressures and delivered VT remained constant up to 8 h of ventilation [18], but altered slightly after 12 h of ventilation compared to baseline (Table 1). Dynamic compliance of the respiratory system was constant up to 8 h of ventilation [18] decreased after 12 h and remained stable afterwards (Fig. 1). These results are indicative of the impact of 8 h of ventilation that did not subsequently worsen.

Morphometric analyses

Seven-day old rat pups ventilated for 12 and 24 h had fewer and larger alveoli when compared to the lungs of non-ventilated 8 day-old pups (Fig. 2A). The tissue-to-air ratio corroborated these findings; it decreased after 12 h of ventilation and declined further during the next 12 h of ventilation (Fig. 2B). To quantify alveolar development, we calculated the number of elastin-positive secondary crests per unit area (Fig. 2D). The number of secondary crests – indicating alveolar formation– increased significantly between the 7\textsuperscript{th} and 8\textsuperscript{th} postnatal days in non-ventilated rat pups. The number of secondary crests increased after 12 h of ventilation when compared to day 7 controls. In contrast, the number of secondary crests was significantly lower in pups ventilated for 24 h vs. non-ventilated day 8 control pups, even when corrected for tissue fraction. To further evaluate alveolar development, we measured the mean linear intercept (Lm; Fig. 2C). Ventilation increased the Lm after 12 h, and more so after 24 h.

Together the data suggest that during the first 12 h of ventilation alveolar space increases because of hyperinflation while a further increase of alveolar space during the next 12 h of ventilation is in part due to arrest in alveolar development as well as hyperinflation.

Lung cell proliferation

Lung cell proliferation was assessed in non-ventilated vs. ventilated rat pups at postnatal days 6, 7, 8, 9, 10 and 14. In non-ventilated rats, the number of proliferating lung cells was

| Table 1. Blood gas analysis and airway pressures after 8, 12 and 24 h of ventilation. |
|---------------------------------|-----------------|-----------------|-----------------|
|                                 | 8 hrs | 12 hrs | 24 hrs |
| pH                              | 7.39±0.07 | 7.29±0.05 | 7.30±0.05 |
| pCO\textsubscript{2} (mmHg)      | 44.3±6.4 | 35.8±6.2 | 39.4±4.5 |
| pO\textsubscript{2} (mmHg)       | 83.9±8.4 | 87.4±11.2 | 73.5±11.7 |
| BE (mmol/L)                     | 4.2±2.3  | 3.9±1.6  | 5.2±2.1  |
| Ppeak (cm H\textsubscript{2}O)   | 10.9±1.1 | 12.7±1.1* | 13.1±1.3* |
| Pmean (cm H\textsubscript{2}O)   | 6.3±0.5  | 7.2±0.5*  | 7.5±0.6*  |
| PEEP (cm H\textsubscript{2}O)    | 2 2 2   |          |          |
| Frequency (breaths/min)         | 150   | 150   | 150   |
| Delivered VT (ml/kg)            | 8.9±0.2 | 8.5±0.4* | 8.4±0.2* |

Values represent means ± SD, n = 12 animals per group. *p<0.05 versus values at 0 hrs. Ppeak, peak pressure; Pmean, mean pressure; PEEP, positive-end expiratory pressure. doi:10.1371/journal.pone.0016910.t001

Figure 1. Dynamic compliance during 24 h of mechanical ventilation. Dynamic compliance decreased during first 12 h of ventilation with room air and low/moderate VT but remained stable during the next 12 h. Data are mean ± SD, n = 12 rat pups per time group. *p<0.05 prolonged versus 1-min ventilation. doi:10.1371/journal.pone.0016910.g001
greatest at postnatal day 6 (BrdU labelling index: ~12%), which declined gradually to almost undetectable at day 15 (Fig. 3). Ventilation for 24 h clearly inhibited lung cell proliferation in pups of all studied ages (days 6-14). Next, 7-day old rat pups were ventilated for all subsequent experiments. Proliferation was not affected by 8 h of ventilation (data not shown) but longer durations of ventilation significantly decreased the number of proliferating cells (Fig. 4A, B). The ratio of proliferating mesenchymal and epithelial cells did not significantly differ between non-ventilated pups and pups ventilated for 8 and 12 h, respectively (0.73±0.05 vs. 0.65±0.03 and 0.67±0.1). Since a 12-h ventilation decreased the total number of proliferating cells (Fig. 4B) the unchanged ratio suggest that cell proliferation of both tissue layers was equally affected by mechanical ventilation. Hardly any proliferating cells were seen after 24 h of ventilation; in agreement with a reduction in cell proliferation in both tissue layers. The almost total arrest in lung cell proliferation by prolonged (24 h) ventilation was confirmed by anti-PH3 immunochemistry (PH3-positive cells decreased from 8 to 1% of total).

**Cell cycle regulators**

mRNA levels of lung cyclin D1 and E1 were significantly down-regulated after 8, 12 and 24 h of ventilation while that of p27Kip1 was increased (Fig. 5A). Immunoblot (i.e. protein) analysis of lungs ventilated for 24 h confirmed these mRNA changes of cyclin D1, E1 and p27Kip1 (Figs. 5B, 5C, 6A). The amount of p27Kip1 was 1.5-fold increased after 12 h of ventilation (not shown). Other members of the Cip/Kip family of CKIs were either increased (p57Kip2, Fig. 6B) or unchanged (p21Waf/Cip1, not shown) by 24 h of ventilation. In contrast, CKIs belonging to the INK4 family

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**Figure 2. Ventilation inhibits alveolar growth.** (A) Histology after mechanical ventilation: (A1) non-ventilated 7-day old rat (A2) 7-day old rat ventilated for 12 h, (A3) 7-day old rat ventilated for 24 h, (A4) non-ventilated 8-day old rat. (B) Mechanical ventilation for 12 and 24 h significantly increased alveolar airspace reduction in tissue-to-air ratio (A) as well as increase in mean linear intercept (D) but decreased number of elastin-positive secondary septa (C). Medians with 25th and 75th quartiles are shown, bars are 5th and 95th percentiles, n = 12 rat pups per time group. MV, mechanical ventilation. * p<0.05.
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were either reduced [p16^{INK4a}] or not affected [p15^{INK4b}] by 24 h of ventilation (Fig. 6C, D). p27Kip1 can be phosphorylated at different sites, which influences its localization and activity [26]. A 12 h ventilation decreased phosphorylation of p27Kip1 at Thr^{157} (Fig. 7A) but did not affect phosphorylation of Thr^{198} (not shown). However, mechanical ventilation for 24 h decreased p27Kip1 phosphorylation at Thr^{157}, Thr^{197} and Thr^{198}, thereby promoting stability and nuclear localization (Fig. 7B–D). Similar—but more rapid—changes in cell cycle regulators were noted when 7-day newborn rats were ventilated with high V_{f}. Although β-actin can be responsive to stretch, no significant differences in β-actin expression was noted between ventilated animals and controls (not shown).

High V_{f} reduced the amount of D1 and D2 cyclins within 1 hour, while that of Cdk inhibitors p27Kip1 and p57Kip2 increased (Fig. 8A); in contrast, p16^{INK4a} content was decreased by high-V_{f} ventilation.

We do not know in which particular tissue layer (epithelium, mesenchyme) these changes occurred in vivo, but they at least occur in epithelial cells as subjecting ex vivo fetal lung epithelial cells to cyclic continuous 17% stretch resulted in similar patterns of alteration in cell cycle regulators (Fig. 8B).

Discussion

The hallmark of ‘new’ BPD is arrested alveolarization [1], but the molecular and cellular basis of the alveolar arrest remains mostly unknown. Alveolarization occurs as the immature saccules, which form the lung parenchyma at birth, are subdivided into smaller units by the formation and extension of secondary septa; new tissue ridges are lifted off the existing primary septa and grow in a centripetal direction into the airspaces. This process, called septation, is mainly postnatal (human: 36 weeks–infancy; rat: Pnd5–Pnd21) [7,27]. Before septation of the air spaces starts, the lung expands for a short period of time, and the cells of the inter-airway walls actively proliferate, peaking at day 5 in rats and steadily declining thereafter [28]. This active cell proliferation takes place just at the beginning of the septation of the distal airways. With the use of a newborn rat model [18] we demonstrate here that mechanical ventilation for 24 h with room air and moderate V_{f} results in cell cycle arrest, and reduced alveolar septation. This ventilation strategy (room air and moderate VT) was chosen to avoid/minimize lung injury.

In rats, lungs at birth have a saccular appearance and alveolarization is an exclusively postnatal (between P4 and P21) event, which makes this model relevant to the infant population developing BPD. However, major differences exist between mechanically ventilated newborn rats and premature born infants. Newborn rats have immature lung architecture at birth, but they do not need mechanical ventilation to survive (likely due to differences in airway structure, with large airways extending almost to the lung periphery and quickly reducing in diameter to the alveoli) and they do not lack surfactant. Infants with BPD demonstrate interstitial thickening that may partly be due to fibroproliferation while in rat pups mechanical ventilation of 24 h caused cell arrest in both mesenchymal and epithelial cell layers. Despite these differences, our results suggest that the observed cell cycle arrest is due to increased expression of two CKIs (i.e., p27Kip1 and p57Kip2) that are members of the Cip/Kip family; the other member, p21Waf1/Cip1, was not affected by 24 h of mechanical ventilation.

Knock-in mouse models have shown that p27Kip1 and p57Kip2 are interchangeable in vivo [29], suggesting similar mechanisms of regulation. Mechanical strain has been recognized as playing an important role in the regulation of fetal lung cell proliferation. Indeed the stimulatory effect of mechanical stretch (i.e., increased intratracheal pressure) on fetal lung growth has been extensively studied in tracheal occlusion (TO) models [30,31], where the number of proliferating alveolar type II cells significantly increased. Fetal sheep, exposed to TO during the alveolar stage of lung development, showed an increase in alveolar type II cells between days 2–4 after TO [31]. This proliferative response of fetal lung cells to strain has also been demonstrated in vivo. Intermittent cyclic 5% stretching (simulating normal fetal breathing movements) of distal fetal rat lung cells (epithelial cells and fibroblasts) increased cell proliferation [32]. However, a continuous cyclic 17% stretch (simulating mechanical ventilation [33]) for 24 h inhibited fetal lung cell proliferation (unpublished results), in agreement with our in vivo findings of a proliferative arrest after 24 h of mechanical ventilation. In the present study, continuous cyclic 17% stretch of fetal lung epithelial cells caused similar alternations in cell cycle regulators as observed in mechanically ventilated newborn lungs in vivo, namely increased levels of p27Kip1 and a decrease in the amount of cyclin D1. CKI members of the Cip/Kip family [p21^{WAF1/Cip1}, p27Kip1 and p57Kip2] preferentially inhibit cyclin-Cdk2 complexes [16]. How mechanical stretch influences CKIs is unknown. In many cancers, the ras/raf/mitogen activated protein kinase (MAPK) pathway increases p27Kip1 and p57Kip2, whereas in vivo, but they at least occur in epithelial cells as subjecting ex vivo fetal lung epithelial cells to cyclic continuous 17% stretch resulted in similar patterns of alteration in cell cycle regulators (Fig. 8B).
The amount of p27Kip1 is regulated at the level of its synthesis (transcription, translation), degradation and localization [39]. During the G0 phase, it accumulates in the nucleus and inhibits cyclin-Cdk complexes. In response to growth stimuli, p27Kip1 translocates from the nucleus to the cytoplasm during G1 phase and is degraded by the proteasome after ubiquitination [39]. permitting
the cell cycle to progress to S phase. Several signaling pathways that alter p27Kip1 phosphorylation influence its subcellular localization and function. For example, phosphorylation of the following essential sites regulate important functions: Thr^{157} prevents nuclear import; Ser^{10} mediates nuclear export; Thr^{198} promotes cytoplasmic translocation and increases p27-dependent motility, which may be important to prepare cells for shape changes in later phases of the cell cycle; and, Thr^{187} results in proteolysis [26,39]. In the present study, mechanical ventilation of 24 h increased the transcription of p27Kip1 and altered its phosphorylation: less phosphorylation of Thr^{157} (increasing nuclear import), Thr^{198} (decreasing nuclear export) and Thr^{187} (reduced proteolysis). No significant changes in

Figure 5. Impact on cyclin D, cyclin E and p27Kip1 expression. Mechanical ventilation for 24 h significantly decreased cyclin D1 and E1 mRNA (A) and protein (B and C) levels in lungs of 7-day old rats. In contrast, p27Kip1 mRNA increased (A). Inserts in (B) and (C) show cyclin D1 (B) and cyclin E1 (C) immunoblots of lung tissue of 7-day old rats ventilated for 24 h and non-ventilated 8-day old rats (controls). Blots were reprobed with β-actin for equal loading and transfer. Medians with 25th and 75th quartiles are shown, bars are 5th and 95th percentiles; qPCR, n = 6 rat pups per time group; immunoblot, n = 3 rat pups per time group. MV, mechanical ventilation. *p<0.05 versus non-ventilated group, ^p<0.05 versus 24 h ventilation. doi:10.1371/journal.pone.0016910.g005
Ser\textsuperscript{10} phosphorylation were noted (not shown). Together, these alterations in p27\textsuperscript{Kip1} phosphorylation favour its nuclear localization and stability. In addition, the reduced phosphorylation of p27\textsuperscript{Kip1} at Thr\textsuperscript{157} and Thr\textsuperscript{198} negatively affects the assembly function of p27\textsuperscript{Kip1} for cyclinD1-Cdk4, thereby negatively affecting cell cycle progression [26].

The second family of CKIs are INK4 proteins (p16\textsuperscript{INK4a}, p15\textsuperscript{INK4b}, p18\textsuperscript{INK4c} and p19\textsuperscript{INK4d}); they inhibit cyclin D-dependent kinases Cdk-4 and -6 [14,15] and, are thus specific for early G\textsubscript{1} phase. In the present study, we found a significant reduction in p16\textsuperscript{INK4a} protein after ventilation with low, moderate or high VT. In addition to Cdk inhibition and G\textsubscript{1} growth arrest, p16\textsuperscript{INK4a} plays a role in regulating apoptosis. It has been shown that p16\textsuperscript{INK4a}-deficiency increases apoptosis in osteosarcoma U2OS and mouse embryonic fibroblast (MEF) cells exposed to ultraviolet (UV) light [40], because of down-regulation of the anti-apoptotic protein Bcl-2. In contrast, the pro-apoptotic protein Bax was down-regulated in p16\textsuperscript{INK4a} expressing cells [40]. Thus, p16\textsuperscript{INK4a} appears to control apoptosis through the intrinsic mitochondrial death pathway.

Prolonged mechanical ventilation has been shown to significantly increase lung cell apoptosis in newborn mice lungs [8]. Although p16\textsuperscript{INK4a} levels were decreased in the present study, it remains yet to be determined whether it plays a role in ventilator-induced apoptosis.

**Figure 6. Different effects on Kip and INK proteins.** Mechanical ventilation for 24 h significantly increased p27\textsuperscript{Kip1} (A) and p57\textsuperscript{Kip2} (B) protein levels. In contrast, p16\textsuperscript{INK4a} protein (D) was decreased by ventilation while p15\textsuperscript{INK4b} (C) was unchanged. Inserts show immunoblots of lung tissue of 7-day old rats ventilated for 24 h and non-ventilated 8-day old rats (controls). Blots were reprobed with \(\beta\)-actin for equal loading and transfer. Medians with 25\textsuperscript{th} and 75\textsuperscript{th} quartiles are shown, bars are 5\textsuperscript{th} and 95\textsuperscript{th} percentiles; \(n = 3\) rat pups per time group. *\(p<0.05\) versus non-ventilated group.

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Another risk factor for BPD is oxygen [1]. Hyperoxia has been shown to interfere with cell-cycle progression in vitro [36,41,42] and hyperoxia-induced G1 arrest appears to be mediated by p21Waf1/Cip1 [43,44]. The hyperoxic induction of p21Waf1/Cip1 is p53-dependent [44] and its increase promotes survival of cells exposed to continuous oxidative stress by maintaining anti-apoptotic Bcl-2X(L) expression [45]. Hyperoxic-ventilated premature baboons delivered at 125 and 140 days of gestation have increased p53 and p21Waf1/Cip1 expression [46,47]. In the present study, we did not assess p53 but the absence of p21Waf1/Cip1 induction by 24 h of mechanical ventilation with room air suggests that p53 is likely not involved in ventilation-induced cell cycle arrest in these studies.

The increase of p27Kip1 and p57Kip2 by mechanical stretch in vitro and in vivo coincided with a reduced expression of cyclins D1 and E1, both of which are essential for cell cycle progression through G1 and entry in S phase. D-type cyclins are induced by mitogenic stimuli in quiescent cells. After association with Cdk4/6 and activation by Cdk activating kinase, they promote entry into the G1 phase, thereby triggering cyclin E expression. Cyclin E binds to Cdk2 and facilitates transition from G1 to S phase [22]. Both p27Kip1 and p57Kip2 are potent inhibitors of cyclin E-dependent suppression.
Ventilation Induces p27 and p57 Kip Proteins

In summary, we conclude that mechanical ventilation for 24 h using moderate V<sub>T</sub> without supplemental O<sub>2</sub> causes G<sub>1</sub> cell cycle arrest of lung cells in newborn rats due to increased transcription and altered phosphorylation (in favour of nuclear localization) of Kip CKIs, and down-regulation of cyclins D<sub>1</sub> and E. This proliferative arrest may cause a reduction in alveolarization, resulting in alveolar simplification. Such identification of ventilation-induced CKIs may have therapeutic potential for the prevention or treatment of arrested alveolarization in ventilated premature infants.

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Author Contributions

Conceived and designed the experiments: AAK MP. Performed the experiments: AAK JW MK. Analyzed the data: AAK BK JBvG MP. Wrote the manuscript: AAK BK JBvG MP. Provided technical assistance with the experiments: ZH.

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