Oral Carriage of *Streptococcus mutans* Harboring the *cnm* Gene Relates to an Increased Incidence of Cerebral Microbleeds

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**BACKGROUND AND PURPOSE:** Cerebral microbleeds (CMB) are associated with stroke and cognitive impairment. We previously reported a high prevalence of CMB in people with *Streptococcus mutans* expressing Cnm, a collagen-binding protein in the oral cavity. *S. mutans* is a major pathogen responsible for dental caries. Repeated challenge with *S. mutans* harboring the *cnm* gene encoding Cnm induced cerebral bleeding in stroke-prone spontaneously hypertensive rats. The purpose of this longitudinal study is to examine the relationship of *cnm*-positive *S. mutans* to the development of CMB.

**METHODS:** We retrospectively investigated patients with stroke receiving oral microbiological examination and head 3T magnetic resonance imaging evaluations twice in the period 2014 to 2019, allowing >180-day interval. Patients with *cnm*-positive *S. mutans* were compared with those without. Quasi-Poisson regression models were used to explore associations between *cnm*-positive *S. mutans* and the increase in number of CMB between the 2 magnetic resonance imaging scans.

**RESULTS:** A total of 111 patients were identified; 21 (19%) with *cnm*-positive *S. mutans* and 90 (81%) without. Clinical history, including blood pressure and the use of antithrombotic agents, were comparable between the 2 groups. New CMB were more commonly observed in patients with *cnm*-positive *S. mutans* (52% versus 23%; *P* = 0.008). The incidence of CMB was significantly higher in the group with *cnm*-positive *S. mutans*, especially in deep areas, (incidence rate ratios [95% CI], 5.1 [1.9–13.6] for CMB in any brain region; 15.0 [5.4–42.0] for deep CMB), which persisted after adjusting for age, sex, hypertension, and renal impairment (4.7 [1.8–11.9] for CMB in any brain region; 13.9 [4.3–44.5] for deep CMB).

**CONCLUSIONS:** This study demonstrates that *cnm*-positive *S. mutans* is associated with an increased incidence of CMB. Treatment for *cnm*-positive *S. mutans* infection may be a novel microbiota-based therapeutic approach for stroke and cognitive impairment.

**GRAPHIC ABSTRACT:** An online graphic abstract is available for this article.

**Key Words:** blood pressure ■ dental caries ■ hemorrhage ■ risk factor ■ *Streptococcus mutans*
S. mutans attaches to cerebrovascular basement membranes (BM)\textsuperscript{17–19} inducing local blood-brain barrier inflammation, resulting in ICH.\textsuperscript{19} Experimental intravenous administration of \textit{cnm}-positive \textit{S. mutans} in stroke-prone spontaneously hypertensive rats and a mouse model of cerebral hemorrhage exacerbates cerebral bleeds.\textsuperscript{19} Epidemiological studies from many countries have shown \textapprox 20\% to 30\% of patients with ICH\textsuperscript{15,22} and 7\% to 20\% of the general population\textsuperscript{14,23–25} have \textit{cnm}-positive \textit{S. mutans} in their oral cavity. Clarifying the effects of \textit{cnm}-positive \textit{S. mutans} on the cerebral vasculature is, therefore, both necessary and urgent.

In this study, we hypothesize that \textit{cnm}-positive \textit{S. mutans} contributes to the development of CMB. We investigated the association between \textit{cnm}-positive \textit{S. mutans} and incidence of CMB in a longitudinal retrospective study.

**METHODS**

**Data Availability Statement**

Raw data were generated and preserved at the National Cerebral and Cardiovascular Center. Derived data supporting the findings of this study are available from corresponding authors on request.

**Study Design**

The current study was approved by the Ethical Committee of the National Cerebral and Cardiovascular Center (M23-073, M25-111 and M27-015) and included in the analysis: (1) subjects who developed acute IS, transient ischemic attack (TIA), or ICH from February 15, 2014 to April 8, 2018; (2) subjects who signed an informed consent form for the current research, including receiving oral bacterial assessments from February 15, 2014 to April 30, 2018; and (3) subjects receiving 3T-MRI scans for clinical purposes twice, with more than a 180-day interval between examinations, from February 15, 2014 to February 15, 2019. The first MRI scan was used for baseline evaluation and the second for follow-up. If >2× of 3T-MRI scans were performed, the oldest and the latest MRI data were selected. The observational period was defined as the period from baseline to follow-up MRI scans. We previously reported a role of systemic inflammation in CMB development.\textsuperscript{10} Higher circulating levels of high-sensitivity CRP (C-reactive protein), IL (interleukin)-6, and IL-18 are associated with CMB.\textsuperscript{10} Experimental models for CMB include mice subcutaneously injected with lipopolysaccharide.\textsuperscript{9,11} CMB are known to be induced by infective endocarditis\textsuperscript{12} or bacterial sepsis.\textsuperscript{6} Streptococcus mutans is a Gram-positive bacterium and a major pathogen responsible for dental caries.\textsuperscript{13} Several cross-sectional studies have shown that oral infection with \textit{S. mutans} expressing Cnm protein is associated with an increased prevalence of CMB.\textsuperscript{14,15} Cnm is a cell-surface 120-kDa collagen-binding protein of \textit{S. mutans}, and its coding gene is \textit{cnm}.\textsuperscript{16–19} \textit{S. mutans} resides on the surface of teeth and frequently induces bacteremia through brushing, flossing, or tooth extraction.\textsuperscript{20,21} Once in the bloodstream, \textit{cnm}-positive...
visits. Hypertension was defined as systolic blood pressure ≥140 mm Hg, diastolic blood pressure ≥90 mm Hg, or history of antihypertensive medication use. Diabetes was considered present through a history of anti-diabetic drug or insulin use, a fasting plasma glucose level of ≥126 mg/dL, or glycated hemoglobin A1c level of ≥6.5%. Dyslipidemia was defined as low-density lipoprotein cholesterol level ≥140 mg/dL, high-density lipoprotein cholesterol level ≤40 mg/dL, triglyceride level ≥150 mg/dL, or use of lipid-lowering drugs. Renal impairment was defined as <60 mL/min/1.73 m² of estimated glomerular filtration rate, according to previous reports.26,27 The presence of atrial fibrillation and current smoking pattern were also noted. Previous IS, TIA and ICH were defined according to the presence of each disease >3 months before the baseline MRI scan, whereas events within 3 months before the baseline MRI were described as recent IS, TIA, or ICH.

Detection of cnm-Positive S. mutans

Dental plaque specimens were collected and inoculated on Mitis-Salivarius medium with bacitracin (Sigma-Aldrich, St. Louis, MO) and 15% sucrose agar plates and anaerobically incubated at 37 °C for 48 hours. S. mutans strains were identified and isolated based on rough morphological features on agar plates, and all strains were cultured in brain heart infusion broth (Becton, Dickinson and Company, Franklin Lakes, NJ) at 37 °C for 24 hours. Bacterial genomic DNA of each strain was extracted, and S. mutans and cnm genes screened using polymerase chain reaction. MKD primer sets for S. mutans and cnm were used to identify cnm-positive and cnm-negative S. mutans.23 Experiments were conducted by researchers blind to clinical information.

MRI Evaluation

Fluid-attenuated inversion recovery and gradient-echo T2*-weighted images were obtained at baseline and follow-up MRI (3T, Magnetom Verio or Spectra; Siemens Medical Solutions, Erlangen, Germany). The presence of CMB on T2*-weighted images was noted according to the Brain Observer MicroBleed Scale.7 CMB were categorized into 3 groups: (1) deep CMB in the deep gray matter in the basal ganglia or thalamus, or white matter in the corpus callosum, internal, external, or extreme capsule, (2) lobar CMB in the cortical gray or subcortical white matter, and (3) subtentorial CMB in the cerebellum or brain stem. CMB in any brain region (any CMB) were also recorded. Newly developed CMB were recorded at follow-up, but not baseline, MRI. All slices were taken parallel to the orbitomeatal line from the base of the skull to the vault. The sequence parameters of T2*-weighted images were as follows: slice thickness, 4.0 mm; interslice gap, 2.0 mm; echo time, 12 ms; repetition time, 550 ms; and flip angle, 20 degrees.

Lacunar infarcts and white matter hyperintensities (WMH) were evaluated by fluid-attenuated inversion recovery images. Lacunar infarcts were defined as supratentorial hypointense lesions of 3 to 15 mm in diameter with a hyperintense rim. Periventricular hyperintensities (PVH) and deep WMH (DWMH) were scored by the Fazekas scale.28 Sequence parameters of fluid-attenuated inversion recovery images were as follows: slice thickness, 5.0 mm; interslice gap, 1.0 mm; echo time, 94 to 114 ms; and repetition time, 12 000 ms.

Severity of SVD

Total severity of SVD was rated as described previously.29 Briefly, 1 point was added if each SVD feature was present: ≥1 of any CMB, ≥1 of lacunar infarcts, irregular PVH extending into deep white matter (Fazekas score 3), and confluent DWMH (Fazekas score 2 or 3). Sum of ratings was used as a total SVD severity (range, 0–4).29

Ratings

SVD markers were independently rated by 2 neurologists. Interrater correlation coefficients were 0.87 for any CMB, 0.94 for deep CMB, 0.94 for lobar CMB, 0.93 for subtentorial CMB, 0.79 for lacunar infarcts, 0.70 for DWMH, and 0.91 for PVH.

Statistical Analyses

Variables were presented as median and interquartile range or numbers and percentages. Mann-Whitney U or Kruskal-Wallis test for continuous data and χ² or Fisher exacts test for categorical data was used. Quasi-Poisson regression models were applied for associations between cnm-positive S. mutans and number of newly developed CMB during the observational period. The incidence rate ratios (IRR) and their 95% CI were estimated. Based on previous reports,15,27,30 age, sex, hypertension, and renal impairment were set as adjustment factors. We estimated hazard ratios by applying Cox proportional hazard models for associations between cnm-positive S. mutans and symptomatic ICH, IS, and TIA incidence. A P<0.05 (2-tailed) was considered statistically significant. Statistical analysis was conducted using SPSS version 26 (IBM, Armonk, NY) and SAS version 9.4 (SAS Institute, Cary, NC).

RESULTS

Patient Characteristics

From the 3782 patients with acute stroke, 404 patients (11%) received oral bacterial examination (Figure I in the Data Supplement). The clinical profiles of subjects with and without bacterial assessment were similar apart from age, the National Institutes of Health Stroke Scale, and modified Rankin Scale (Table I in the Data Supplement).

We identified 111 subjects fulfilling all the criteria and found that cnm-positive S. mutans was present in 21 (19%), and absent in 90 (81%), patients. Among the 90 patients in the cnm (−) group, cnm-negative S. mutans was detected in 69 and no S. mutans in 21. Characteristics of subjects at baseline MRI are described in Table 1. Age, sex, blood pressure, and vascular risk factors were similar between cnm (+) and cnm (−) groups (systolic blood pressure: 126 mm Hg [116–134] versus 130 mm Hg [118–147], P=0.267; diastolic blood pressure: 76 mm Hg [64–85] versus 74 mm Hg [65–84], P=0.946). The 2 groups also exhibited equivalent blood pressure at follow-up evaluation (systolic blood pressure: 125 mm Hg [117–135] versus 122 mm Hg [115–135]; diastolic blood pressure: 75 mm Hg [70–80] versus 70 mm Hg [64–80]). The cnm (+) group showed higher, but
not significant, levels of CRP and fibrinogen than the cnm(−) group (CRP: 0.15 mg/dL [0.04–0.79] versus 0.08 mg/dL [0.04–0.23], P=0.250; fibrinogen: 344 mg/dL [274–415] versus 308 mg/dL [273–358], P=0.185).

CMB were detected in 12 (57%) of the cnm(+), and 38 (42%) of the cnm(−), group. Lacunar infarcts, PVH, and DWMH were commonly observed in the cnm(+) group (lacunar infarcts: 62% versus 31%, P=0.008; PVH: 38% versus 14%, P=0.026; DWMH: 76% versus 54%, P=0.069). Consequently, total SVD severity was significantly more advanced in the cnm(+) than cnm(−) group (3.0 [1.0–3.0] versus 1.0 [0–2.0], P=0.004).

Cerebral Microbleeds

The numbers of CMB are summarized in Table 2. The cnm(+) group showed a marginally increased number of CMB versus the cnm(−) group at baseline, especially in the deep region, but comparable in lobar and subtentorial regions (any CMB: 2.0 [0–10.5] versus 1.0 [0–5.3], P=0.094; deep CMB: 1.0 [0–7.5] versus 0 [0–2.0], P=0.091) and follow-up (any CMB: 4.0 [0.5–13.5] versus 1.0 [0–6.0], P=0.067; deep CMB: 2.0 [0–10.0] versus 0 [0–2.0], P=0.039).

We assessed the development of new CMB from baseline to follow-up MRI. The observational period was similar between the cnm(+) and cnm(−) group (509 [279–584] versus 482 [364–732] days, P=0.405). CMB development was significantly higher in the cnm(+) than cnm(−) group (52% versus 23%, P=0.008; Table 3). In particular, newly developed CMB were more frequent in deep regions (48% versus 9%, P<0.001) in the cnm(+) than cnm(−) group. Mean numbers of new CMB in the cnm(+) and cnm(−) groups were 2.2 versus 0.5 for any

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|       | cnm (+) group (n=21) | cnm (−) group (n=90) | P value |
|-------|----------------------|----------------------|---------|
| Age, y | 73.0 (63.0–78.0)     | 71.5 (64.0–81.0)     | 0.564   |
| Male, n (%) | 14 (67)            | 54 (60)              | 0.572   |
| Hypertension, n (%) | 18 (86)         | 72 (80)              | 0.759   |
| SBP, mm Hg | 126 (116–134)    | 130 (118–147)        | 0.267   |
| DBP, mm Hg | 76 (64–85)        | 74 (65–84)           | 0.946   |
| Diabetes, n (%) | 3 (14)            | 22 (24)              | 0.396   |
| Dyslipidemia, n (%) | 13 (62)          | 52 (58)              | 0.730   |
| Renal impairment, n (%) | 10 (48)         | 41 (46)              | 0.864   |
| Atrial fibrillation, n (%) | 2 (10)         | 13 (14)              | 0.732   |
| ATA use, n (%) | 15 (71)           | 73 (81)              | 0.372   |
| Antiplatelet agents, n (%) | 12 (57)         | 54 (60)              | 0.810   |
| Anticoagulants, n (%) | 5 (24)           | 23 (26)              | 0.868   |
| ≥2 ATA use, n (%) | 2 (10)           | 15 (17)              | 0.520   |
| Recent IS, n (%) | 7 (33)            | 31 (34)              | 1.000   |
| Recent TIA, n (%) | 1 (5)             | 10 (11)              | 0.687   |
| Recent ICH, n (%) | 1 (5)             | 7 (8)                | 1.000   |
| Previous IS, n (%) | 13 (62)           | 42 (47)              | 0.234   |
| Previous TIA, n (%) | 0 (0)             | 5 (8)                | 0.581   |
| Previous ICH, n (%) | 5 (24)            | 10 (11)              | 0.155   |
| Smoking, n (%) | 10 (48)           | 42 (47)              | 0.937   |
| mRS | 1.0 (0–3.5)        | 1.0 (0–3.0)          | 0.542   |
| CRP, mg/dL* | 0.15 (0.04–0.79)  | 0.08 (0.04–0.23)     | 0.250   |
| Fibrinogen, mg/dL† | 344 (274–415)    | 308 (273–358)        | 0.185   |
| CMB, n (%) | 12 (57)           | 38 (42)              | 0.216   |
| Lacunar infarcts, n (%) | 13 (62)          | 28 (31)              | 0.008   |
| PVH=3, n (%) | 8 (38)            | 13 (14)              | 0.026   |
| DWMH ≥2, n (%) | 16 (76)           | 49 (54)              | 0.069   |

Table 1. Clinical Characteristics at the Baseline Evaluation

Data represent median (interquartile range) or number (percent). ATA indicates antithrombotic agents; CMB, cerebral microbleeds; CRP, C-reactive protein; DBP, diastolic blood pressure; DWMH, deep white matter hyperintensities; ICH, intracerebral hemorrhage; IS, ischemic stroke; mRS, modified Rankin Scale; PVH, periventricular hyperintensities; SBP, systolic blood pressure; SVD, small vessel disease; and TIA, transient ischemic attack.

*CRP data was missing in 1 patient in the cnm(−) group.
†Fibrinogen was obtained 18 subjects in the cnm(+) and 70 in the cnm(−) group.
CMB, 1.4 versus 0.1 for deep, 0.4 versus 0.4 for lobar, and 0.4 versus 0.1 for subtentorial.

We estimated the IRR considering newly developed CMB and observational period. IRR for CMB in deep and subtentorial, but not lobar, regions were significantly higher using unadjusted analysis (any CMB: IRR, 5.1 [95% CI, 1.9–13.6], \( P=0.001 \); deep CMB: IRR, 15.0 [95% CI, 5.4–42.0], \( P<0.001 \); subtentorial CMB: IRR, 6.4 [95% CI, 1.3–30.9], \( P=0.020 \); lobar CMB: IRR, 1.3 [95% CI 0.2–7.7], \( P=0.808 \)). Statistical significance for any and deep CMB was confirmed after adjusting for age, sex, hypertension, and renal impairment (any CMB: IRR, 4.7 [95% CI, 1.8–11.9], \( P=0.001 \); deep CMB: IRR, 13.9 [95% CI, 4.3–44.5], \( P<0.001 \); Table 4). Representative images showing the increase in deep CMB are illustrated in Figure II in the Data Supplement.

### Table 3. The Frequency of New CMB Development

| CMB Type                      | cnm (+) Group | cnm (−) Group | \( P \) Value |
|-------------------------------|---------------|---------------|---------------|
| Any CMB, n (%)                | (n=121)       | (n=90)        | 0.008         |
| Deep CMB, n (%)               | 11 (52)       | 21 (23)       | <0.001        |
| Lobar CMB, n (%)              | 10 (48)       | 8 (9)         | 0.736         |
| Subtentorial CMB, n (%)       | 4 (19)        | 13 (14)       | 0.064         |

Table 3. The Frequency of New CMB Development

Progression of Other SVD Markers

We next evaluated progression of SVD features other than CMB. Frequency of lacunar infarcts (cnm [+] versus cnm [−]: 67% versus 36%), PVH (38% versus 17%), and DWMH (76% versus 57%) on follow-up MRI was subtly increased from baseline. The change in frequency of each SVD feature other than CMB during the observation period was equivalent between cnm (+) and cnm (−) groups (lacunar infarcts: 5% versus 4%, \( P=1.000 \); PVH: 0% versus 2%, \( P=1.000 \); DWMH: 0% versus 2%, \( P=1.000 \)).

Stroke and TIA

Symptomatic stroke and TIA frequency during the observation period was investigated. ICH, IS, and TIA incidence was similar in cnm (+) and cnm (−) groups (ICH: 2 [10%] versus 3 [3%], hazard ratios, 1.7 [95% CI, 0.6–3.4]; TIA, 1 [5%] versus 3 [3%], hazard ratios, 5.3 [95% CI, 0.7–38.8]; IS: 6 [29%] versus 23 [26%], hazard ratios, 1.4 [95% CI, 0.6–3.4]; TIA, 1 [5%] versus 3 [3%], hazard ratios, 1.7 [95% CI, 0.2–16.8]).

Comparison Between the 3 Groups

S. mutans, whether cnm positive or not, may contribute to mycotic aneurysms and cerebral hemorrhage. We, therefore, compared the 3 groups: (1) subjects with cnm-positive S. mutans, (2) those with cnm-negative S. mutans, and (3) those without S. mutans. Background profiles were similar among the 3 groups, except for some imaging markers of SVD, such as lacunar infarcts, WMH, and total SVD severity (Table II in the Data Supplement). Development of CMB was most prominent in cnm-positive S. mutans subjects (Table III in the Data Supplement). No significant difference was observed between subjects with cnm-negative S. mutans and those without S. mutans.

**DISCUSSION**

We found harboring cnm-positive S. mutans was closely related to an increased incidence of CMB, especially in the deep area, together with a high prevalence of lacunar infarcts and WMH.
The strong linkage of cnm-positive S. mutans and deep CMB aligns with previous cross-sectional studies.\(^{14,15}\) Estimated IRR for deep CMB was high in comparison with other known risk factors in previous reports.\(^{30,32,33}\) Deep CMB were considered as biomarkers for hypertensive arteriopathy,\(^{3,34}\) but their pathogenesis cannot be fully explained by hypertension, as they are occasionally found in subjects without high blood pressure.\(^{14,32}\)

An important hallmark of S. mutans expressing Cnm protein\(^{13}\) is its binding activity to components of vascular BM, such as collagen-IV\(^{17}\) and laminin.\(^{18}\) Collagen-binding activity is positively correlated with cnm mRNA expression in S. mutans.\(^{23}\) Conversely, neither cnm-negative S. mutans nor cnm knockout strains of S. mutans can attach to soft tissues such as vessel walls.\(^{17}\) Aging and hypertension induce endothelial injury and BM thickening, resulting in collagen-IV and laminin exposure in cerebral small arteries.\(^{35,36}\) Once cnm-positive S. mutans adheres to BM, infiltration of neutrophils may activate local inflammation, increasing blood-brain barrier permeability, and production of enzymes, such as matrix metalloproteinase-9,\(^{19}\) inducing ICH or CMB (Figure). Endothelial injury related to aging and hypertension is prominent in the deep cerebral vessels,\(^{37}\) likely contributing to an increase in the deep CMB rather than lobar CMB, by cnm-positive S. mutans.

Furthermore, unlike cnm-negative S. mutans, the cnm-positive S. mutans can suppress collagen-induced platelet aggregation.\(^{19}\) All S. mutans, whether cnm-positive or not, have negative zeta potential values, an indicator of cell-surface charge, although cnm-positive S. mutans possesses lower zeta potential values.\(^{19}\) Since platelets also possess negative potentials, cnm-positive S. mutans may inhibit platelet adhesion and aggregation.

**Figure.** Hypothetical model of the mechanism contributing to develop cerebral microbleeds (CMB) by the infection of cnm-positive Streptococcus mutans (S. mutans).

A. Normal vessel. Cerebral bleeding may occur at the level of arterioles and capillaries. B. Aging and hypertension results in endothelial damage and thickened basement membranes (BM). C. Bacteremia of S. mutans are induced by brushing, flossing or tooth extraction. Unlike cnm-negative S. mutans, cnm-positive S. mutans can attach to the BM. D. Once cnm-positive S. mutans binds to the vessel wall, infiltration of neutrophils results in local inflammation. The negative charges on the surface of cnm-positive S. mutans inhibit aggregation of platelets, which also possess negative charges on the surfaces. CMB are eventually induced.
accelerating thus cerebral bleeding. Zeta potential values differ among strains of cnm-positive S. mutans and lower zeta potential values significantly correlate with decreased collagen-induced platelet aggregation.19

We previously reported cnm-positive S. mutans is significantly associated with severe dental caries.23 S. mutans expressing collagen-binding protein can strongly bind to the type-I collagen-composed dentin tooth layer, accelerating development of carious lesions.17 The increased predisposition of cnm-positive S. mutans to invade dental caries provides opportunities for S. mutans to enter the bloodstream and cerebral circulation. Poor oral health could facilitate dental bacteremia and cerebrovascular health.1234 The collagen-binding activity of cnm-positive S. mutans to type-I collagen in teeth and type-IV collagen in cerebrovascular BM facilitates CMB.

S. mutans, including cnm-positive, are commonly transmitted by vertical infection, colonizing mouths of infants at around 2 years.2339 Mothers and caretakers of children are the major sources of S. mutans,40 which generally remain after colonization16 but are not easily implanted again in adulthood.4142 Therefore, preventing vertical cnm-positive S. mutans infection could represent a major preventative factor in SVD and CMB.

Here, overall prevalence of CMB was 45%, higher than previous IS cohorts,3443 and equivalent to IS and IS mixed stroke cohorts.45 The current study included about 20% of subjects with a history of ICH. Additionally, high magnetic field strength of MRI may have affected CMB frequency. Only patients receiving 3T-MRI scans were included, which is suitable for CMB detection and superior to 1.5T-MRI.5

Although CMB may predict future ICH,3444 the incidence of symptomatic ICH in the cnm (+) group was similar to the cnm (−) group. Circulating inflammatory marker level was increased, but nonsignificantly, in the cnm (+) group, which may be a consequence of a small sample size. Thus, to definitively establish an association between cnm-positive S. mutans, symptomatic ICH, and inflammatory marker levels, a large-scale prospective investigation is warranted. This study leads to new hypotheses and provides useful data to guide power calculations and effect sizes for future larger-scale investigations.

There are some limitations to this study. First, it involved Japanese subjects only, making predictions for other countries uncertain and demonstrating the need for multinational validation studies. Second, this was a retrospective study, posing potential risk of selection bias. Only 11% of the total stroke patients had oral bacterial evaluation due to age and factors, leading to difficulty providing informed consent, such as impaired consciousness, cognitive impairments, and advanced frailty. This resulted in the lower age and scores of National Institutes of Health Stroke Scale and modified Rankin Scale in patients receiving bacterial assessments (Table I in the Data Supplement). Finally, all patients had a history of stroke and the effect of cnm-positive S. mutans on CMB development should be examined in a population-based cohort in any future study.

In conclusion, cnm-positive S. mutans was associated with increased CMB incidence. Though the results should be verified by large-scale prospective studies, a close association between cnm-positive S. mutans and CMB development suggests treatments targeting cnm-positive S. mutans may act as novel therapeutic approaches for dementia and stroke.

ARTICLE INFORMATION
Received April 1, 2020; final revision received August 31, 2020; accepted Oc-
tober 2, 2020.

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Acknowledgments
We indebted to Yoko Kiyama and Natsuki Hanada for technical assistance and Dr Ahmad Khundakar for editorial assistance and helpful comments.

Sources of Funding
This study was funded by Grant-in-Aid for Japan Society for the Promotion of Science Fellows to Dr Saito (19J00106), Grant-in-Aid for Challenging Exploratory Research to Dr Ihara (16K14573, 19K22610), Mitsui Sumitomo Insurance Welfare Foundation to Dr Ihara, SENSISHIN Medical Research Foundation to Dr Ihara, Invitational Fellowships for Research in Japan to Dr Friedland, and the Jewish Heritage Fund for Excellence to Dr Friedland.

Disclosures
Dr Yoshimoto reports other support from Takeda Pharmaceutical Company Limited during the conduct of the study. Dr Nakahara reports grants from Boehringer Ingelheim, grants and personal fees from Daiichi Sankyo, grants and personal fees from Eisai, grants and personal fees from Otsuka, grants from Pfizer, and grants and personal fees from Sanofi outside the submitted work. Dr Koga reports honoraria from Otsuka, Takeda, Bayer, Pfizer, Bristol-Myers Squibb, Daiichi Sankyo, Ono, Mitsubishi Tanabe Pharma Corporation, and Boehringer Ingelheim. Dr Toyoda reports lecture honoraria from Bayer, Boehringer Ingelheim, Bristol-Myers Squibb, and Daiichi Sankyo. Dr Ihara reports research support not attributed in the artic from Shimadzu Corporation and Otsuka Pharmaceutical. The other authors report no conflicts.

Supplemental Materials
Tables I–III
Figures I–II

REFERENCES
1. Ihara M, Yamamoto Y. Emerging evidence for pathogenesis of sporadic cerebral small vessel disease. Stroke. 2016;47:554–560. doi: 10.1161/STROKEAHA.115.009627
2. Akoudad S, Wolters FJ, Viswanathan A, de Brujin RF, van der Lugt A, van der Lugt A, Hofman A, Koudstaal PJ, Ikram MA, Vernooij MW. Association of cerebral microbleeds with cognitive decline and dementia. JAMA Neurol. 2016;73:934–943. doi: 10.1001/jamaneurol.2016.1017
3. Païli M, Cordonnier C. Clinical relevance of cerebral small vessel diseases. Stroke. 2020;51:47–53. doi: 10.1161/STROKEAHA.119.024148
4. Debette S, Schilling S, Duperon MG, Larsson SC, Markus HS. Clinical signi-
ficance of magnetic resonance imaging markers of vascular brain injury;
31. Akoudad S, Aarts N, Noordam R, Ikram MA, Tiemeier H, Hofman A, Stricker BH, Vroomen MW, Visser LE. Antidepressant use is associated with an increased risk of developing microbleeds. Stroke. 2016;47:251–254. doi: 10.1161/STROKEAHA.115.011574

32. Liu W, Liu R, Sun W, Peng Q, Zhang W, Xu E, Cheng Y, Ding M, Li Y, Hong Z, et al. CASiSP Study Group. Different impacts of blood pressure variability on the progression of cerebral microbleeds and white matter lesions. Stroke. 2016;47:2916–2922. doi: 10.1161/STROKEAHA.116.010747

33. Farrall AJ, Wardlaw JM. Blood-brain barrier: ageing and microvascular disease. Aging Cell. 2016;15:104–111. doi: 10.1111/acy.12345

34. Vernooij MW, van der Lugt A, Ikram MA, Wielopolski PA, Niessen WJ, Alituusua S, Ooshima T. Molecular and clinical analyses of the gene encoding collagen binding activity is a risk factor for cerebral microbleeds associated with IgA nephropathy. Neurology. 2016;86:867–871. doi: 10.1212/01.wnl.00005002419

35. Horn K, Schwind S, Dvornikowa AK, Paganini-Hill A, Kim R, Cribbs DH, Fisher MJ. A murine model of inflammation-induced cerebral microbleeds. Stroke. 2011;42:3202–3206. doi: 10.1161/STROKEAHA.111.621193

36. Meurman JH, Hämäläinen P. Oral health and morbidity–implications of oral inflammation. J Dent. 2000;28:337–341. doi: 10.1016/S0300-5712(00)00153-4

37. Lammie GA. Pathology of small vessel stroke. Stroke. 2003;34:2035–2042. doi: 10.1161/01.STR.0000081030.68031.69

38. Meurman JH, Pelkonen J, Hartikka J, Hallikainen-Miettinen T, Suominen J, Hämäläinen P. Associations of dental caries and periodontal disease with systemic disease. J Periodontol. 2005;76:9–17. doi: 10.1902/jop.2005.040418

39. Caufield PW, Cutter GR, Dasanayake AP. Initial acquisition of mutans streptococci in the human oral cavity. Arch Oral Biol. 1967;12:231–236. doi: 10.1016/0003-9969(67)90139-8

40. Lapirattanakul J, Nakano K. Mother-to-child transmission of mutans streptococci. Future Microbiol. 2006;1:17–26. doi: 10.4155/fmb.1.2

41. Watanabe I, Kuriyama Y, Miyataki F, Nomura R, Saka K, Ibara M, Iwai K, Matsuji D, Ozaki E, et al. Oral cmn-positive Streptococcus mutans expressing collagen binding activity is a risk factor for cerebral microbleeds and cognitive impairment. Sci Rep. 2016;6:38561.

42. Nomura R, Ogaya Y, Nakano K. Contribution of the collagen-binding proteins of Streptococcus mutans to bacterial colonization of inflamed dental pulp. PLoS One. 2011;6:e19563. doi: 10.1371/journal.pone.0019563

43. Nomura R, Nakano K, Momoide H, Otsugu M, Nakamura S, Otsugu T, Nakano K. Potential high virulence for infective endocarditis in Streptococcus mutans expressing collagen binding activity. Mol Oral Microbiol. 2016;6:20074. doi: 10.1038/srep20074

44. Wilson D, Charidimou A, Ambler G, Fox GV, Gregoire S, Rayson P, Imaizumi T, Fluri F, Nakano H, Horstmann S, et al. Recurrent stroke risk and cerebral microbleed burden in ischemic stroke and TIA: A meta-analysis. Neurology. 2016;87:1501–1510. doi: 10.1212/WNL.000000000003183