ASSOCIATION OF ENAM C2452T POLYMORPHISM WITH HIGH RATES OF CARIES OCCURRENCE IN AN INDONESIAN POPULATION

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ABSTRACT

Objective: Tooth decay or the caries process is a common dental problem that affects millions of people worldwide. Many risk factors are modifiable, while others are not (e.g., genetic factors). Polymorphism of the enamelin (ENAM) gene, which is required to ensure production of an essential protein for enamel development, may pose as a risk factor for the caries process. This study sought to investigate the possibility of ENAM C2452T polymorphism acting as a risk factor in the caries process.

Methods: The polymerase chain reaction–restriction fragment length polymorphism method was employed to evaluate DNA samples taken from 95 subjects with a high caries prevalence and 89 control subjects for ENAM C2452T polymorphism.

Results: Based on Chi-squared tests, there were significant genotype and allele distribution differences between the group with a high caries prevalence and the control group (p=0.005 and p=0.007). Polymorphism in this context may, therefore, serve as a risk factor for caries onset and progression (OR: 3.62).

Conclusion: ENAM C2452T polymorphism is related to the caries process and may constitute a risk factor.

Keywords: Caries, Enamelin, Polymorphism, Indonesia.

INTRODUCTION

According to the United States (US) National Institute of Dental and Craniofacial Research, roughly 92% of adults aged 20–64 years show tooth decay on their permanent teeth [1]. Further, among this population, 26% do not receive regular dental care. In Indonesia, approximately 65 million residents have dental problems including tooth decay, with only 31% receiving dental care [2].

Tooth decay or dental caries is a dynamic process that affects the outer layers of the tooth when the demineralization and remineralization processes are not balanced with one another [3,4]. The caries process is initiated by dental plaque or biofilm, made up of a community of microorganisms that adhere to the dental surfaces and which can demonstrate cariogenic properties if left untreated. Microorganisms can be characterized as cariogenic if they produce polysaccharides, convert sugar to fatty acids (acidogenic), or tolerate a highly acidic environment (aciduric). Cariogenic microorganisms act to disrupt the regular remineralization–demineralization process, as they promote an increase in the demineralization side of the process [4]. Caries itself can be categorized as either occurring at a high rate or a low rate using the decayed-missing-filled (DMFT) index (for permanent teeth) or decayed-extracted-filled teeth (def-t) index (for deciduous teeth). Both indices consider the presence of decayed, missing, and filled teeth, with a score of 10 points for DMFT or 4.5 points for def-t being the cutoff values for high caries occurrence versus low occurrence [5].

There are currently several known risk factors that contribute to caries onset and progression. However, among these, certain risk factors (e.g., age and genetic factors) cannot be changed. According to a recent study, individuals boasting genetic polymorphism of certain genes such as enamelin (ENAM), mannose-binding lectin-2, and amelogenin X are more prone to experiencing this process [2,6,7]. Genetic polymorphism can be described as gene sequence variations between individuals that may or may not affect one’s health. A genetic variation is considered a polymorphism when it affects a specific allele in more than 1% of a population group [8]. Single-nucleotide polymorphisms (SNPs) are the most common kind of genetic polymorphism, where a specific nucleotide base changes into another, affecting one in every 300 nucleotide bases and which may act as a biological marker for certain health concerns. While most SNPs do not have any discernible effects on health, some have been proven to be associated with certain diseases [9].

The ENAM gene, located on the q arm of the fourth chromosome on 13.3 (4q13.3), provides instructions for constructing ENAM proteins that are required in normal tooth development, especially including the formation of the enamel, the outermost layer of a tooth structure. Until now, its specific behavior has not been fully recognized [10]. Several polymorphisms of this gene have been suggested previously to be associated with the caries process [6,7,11-14].

MATERIALS AND METHODS

Study subjects and ethical approval

This study used 184 DNA samples extracted from consenting patients' peripheral blood, with the study population consisting of 95 individuals with high caries occurrence (study group) and 89 individuals with low or no caries occurrence (control group). The extracted DNA samples were stored in the Oral Biology Laboratory, Faculty of Dentistry, Universitas Indonesia, at −20°C. This study and its methods were approved by the Universitas Indonesia, Faculty of Dentistry’s Ethical Committee (Number: 92/Ethical Approval/FKGUI/VI/2017).
DNA isolation and genotyping

To identify ENAM C2452T polymorphism, we performed DNA isolation as described by Auerkari et al. [15-17] and genotyping procedures as described by Ouryouji et al. [14]. Each 20-μL polymerase chain reaction (PCR) mixture contained 10 μL of ready-to-use master mix (BioLine, London, UK), 0.4 μL of forward (5'-AGG ATT TTT ATT ACA GTG AAT TTT ACC CAT GGG GCC-3') and reverse primers (5'-GCT TAA GGA GCC TGG TGT (CT ATC TTC-3'), and 20 μg of genomic DNA. PCR was performed in a T100™ thermal cycle (Bio-Rad Laboratories, Hercules, CA, USA) with the following temperature profiles: Initial denaturation at 96°C for 2 min, followed by 37 cycles of denaturation at 94°C for 1 min, annealing at 58.2°C for 1.5 min, and extensions at 72°C for 1 min, with a final extension occurring at 74°C for 7 min.

The PCR products were then digested with one unit of Apal restriction enzyme (Genetika Science, Indonesia) for 20 min at 25°C. Next, the digested products were separated in 2% agarose gel stained with GelRed (Biotium Inc., Hayward, CA, USA) and recorded with GelDoc 200 (Bio-Rad Laboratories, Hercules, CA, USA).

Apal digestion yielded fragments of 36 bp and 144 bp (homozygote G/G); 36 bp, 144 bp, and 180 bp (heterozygote G/A); and 180 bp (homozygote C/C), respectively.

Statistical analysis

Genetic data were entered and processed using the Statistical Package for the Social Sciences (SPSS) version 22.0 software program (IBM Corp., Armonk, NY, USA) and examined using a descriptive analysis approach. Associations between high caries incidence and ENAM genotype were evaluated with the Chi-squared test and binary logistic regression analysis.

RESULTS

This study involved 96 samples DNA of individuals with high caries occurrence and 89 samples of DNA of individuals with low caries occurrence (Table 1). Amplified PCR products were confirmed as ENAM C2452T by electrophoresis as shown in Figs. 1-3. Fig. 1 also presents examples of digested PCR products, where the CT genotype (sample no. 4) was indicated by two bands of 180 bp and 144 bp; sample no. 5 and 6 showed a single band of 144 bp, representing the CC genotype; and sample no. 7 showed a single band of 180 bp, representing the TT genotype. No instance of a 36-bp band was seen as such is located within the primer and was not able to be discriminated by electrophoresis.

The results of PCR-restriction fragment length polymorphism (RFLP) evaluation of ENAM C2452T revealed that genotype CC and allele C were dominant in the general population. A total of 151 samples carried the wild-type homozygote C/C, 23 samples carried the polymorphic heterozygote C/T, and 10 samples carried the polymorphic homozygote T/T (Table 2). Among 368 alleles from 184 samples, only 43 demonstrated polymorphic alleles. Moreover, Table 3 describes a comparison of the distribution of ENAM C2452T polymorphism between Indonesian, Japanese, and Polish patients with high caries occurrence and low caries occurrence.

DISCUSSION

As a tooth-specific protein matrix, ENAM is produced by the ENAM gene. This protein matrix is secreted in an ameloblast secretory stage and is a necessary element in the enamel mineralization phase [11,14]. Research indicates a nucleic acid base change or SNPs of this gene can result in hereditary enamel hypoplasia and may alter individual enamel thickness as well as the teeth’s magnesium and calcium concentrations, impacting the demineralization process and resulting in a progression of the caries process [7].

The present study sought to identify the difference in the distribution of ENAM C2452T polymorphism among individuals with high caries activity and low caries activity in an Indonesian population. According to the US National Institute of Dental and Craniofacial Research, nine of 10 adults aged 20-64 years old have at least one active caries lesion on their permanent teeth [1]. In Indonesia, 65 million individuals have dental problems including caries [2].

Gene polymorphism changes the sequence of various genes including ENAM, which can lead either to no health effect or to susceptibility to certain conditions or diseases such as a vulnerability to the caries process. The polymorphism of the ENAM gene is also proven to be related with one type of amelogenesis imperfecta, a disorder of tooth development that results in teeth that appear discolored, pitted or grooved, unusually small, and inclined to wear and breakage, which is essentially an example of hypoplasia [14,18]. Thereby, a change in the nucleic acid base of this gene may adversely affect the capacity of the enamel structure.

In a previous study that focused on the influence of ENAM C2452T polymorphism on the caries process by comparing individuals with high caries occurrence and low caries occurrence across various races, a significant difference was not reported. However, the conversion of the cytosine base (C) to thymine base (T) in the +2452 position in the ENAM gene is also proven to be associated with one type of amelogenesis imperfecta, a disorder of tooth development that results in teeth that appear discolored, pitted or grooved, unusually small, and inclined to wear and breakage, which is essentially an example of hypoplasia [14,18]. Thereby, a change in the nucleic acid base of this gene may adversely affect the capacity of the enamel structure.

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To visualize the genotype of the gene for each DNA sample, amplification was performed on the specific gene position using PCR through denaturation, annealing, and elongation phases. Following successful amplification, all amplified DNA samples were digested with Apal restriction enzyme by way of an RFLP technique. Restricted or digested products were visualized with GelDoc. The visualization results were subsequently analyzed using the SPSS software program and showed that genotype and allele distribution differences were present between individuals with high caries occurrence and low caries occurrence, respectively. These findings may indicate that certain genetic factors have an affinity toward caries occurrence.
Ouryouji et al. in 2008 reported that there was no statistically significant difference between ENAM C2452T polymorphism and high caries occurrence in a Japanese population (p=0.143 and p>0.05). Their study, which included 57 individuals as control subjects (individuals with low caries occurrence or DMFT <10 points/def-t <4.5 points) and 80 individuals as caries subjects (individuals with high caries occurrence or DMFT ≥10 points/def-t ≥4.5 points) stated that CC genotype (wild-type or nonpolymorphic) dominated among both control (88.1%) and caries (96%) subjects [14].

Another study, conducted by Olszowski et al. in 2012, also reported that no statistically significant difference was present between ENAM C2452T polymorphism and high caries occurrence in a Polish population (p=0.67, p=0.18, and p>0.05). Their research included 84 individuals as control subjects (individuals with low caries occurrence or DMFT <10 points/def-t <4.5 points) and 95 individuals as caries subjects (individuals with high caries occurrence or DMFT ≥10 points/def-t ≥4.5 points) and stated that the C allele (wild-type or nonpolymorphic) dominated in both study populations (73.16% of caries subjects and 77.38% of control subjects) [6].

In contrast, this research, which focused on synonymous polymorphism, reported a different outcome. Among 95 caries subjects (individuals with high caries occurrence or DMFT ≥10 points/def-t ≥4.5 points), it was found that 25 individuals (26.3%) presented a mutant-type or polymorphic genotype (CT or TT genotype), while eight individuals (9%) among 89 control subjects (individuals with low caries occurrence or DMFT <10 points/def-t <4.5 points) displayed a mutant-type or polymorphic genotype (CT or TT genotype).

Table 2: Genotype frequency among individuals with high caries occurrence versus control subjects

| Polymorphism | High caries (DMFT≥10 points or def-t≥4.5 points) | Control (DMFT<10 points or def-t<4.5 points) | Pearson’s Chi-squared test |
|--------------|-----------------------------------------------|-----------------------------------------------|----------------------------|
| CC           | 70 (46.36%)                                   | 81 (53.64%)                                   | p=0.005                    |
| CT           | 19 (82.61%)                                   | 4 (17.39%)                                    |                            |
| TT           | 6 (60%)                                       | 4 (40%)                                       |                            |
| C            | 159 (48.92%)                                  | 166 (51.08%)                                  | p=0.007                    |
| T            | 31 (72.09%)                                   | 12 (27.91%)                                   |                            |
Table 3: Comparison of the distribution of ENAM C2452T polymorphism between Indonesian, Japanese, and Polish patients with high caries occurrence and low caries occurrence

| Study                  | Population | Caries (genotype/allele) (%) | Control (genotype/allele) (%) |
|------------------------|------------|------------------------------|-----------------------------|
| Present study          | Indonesian | CC (46.36)                   | CC (53.64)                  |
|                        |            | CT (82.61)                   | CT (17.39)                  |
|                        |            | TT (60)                      | TT (40)                     |
|                        |            | Callele (48.92)              | Callele (51.08)             |
|                        |            | T allele (72.09)             | T allele (27.91)            |
| Ouryouji et al. (2008) | Japanese   | CC (48.12)                   | CC (51.88)                  |
|                        |            | CT (70.41)                   | CT (29.59)                  |
|                        |            | TT (0)                       | TT (0)                      |
|                        |            | Callele (52.03)              | Callele (47.97)             |
|                        |            | T allele (31.04)             | T allele (68.96)            |
| Olszowski et al. (2012)| Polish     | Callele (50.93)              | Callele (49.07)             |
|                        |            | T allele (46.07)             | T allele (53.93)            |

Despite the fact that both a wild-type or nonpolymorphic allele (C) and genotype (CC) were found more often than the mutant-type or polymorphic allele (T) and genotype (CT or TT) in both subject groups, a continuity correction calculation revealed that there was a statistically significant difference between the distribution of wild-type or nonpolymorphic alleles and mutant-type or polymorphic alleles with p=0.007 (p<0.05). Further, the continuity correction calculation of the genotype showed p=0.005 (p<0.05), which means that there is a statistically significant difference between the distribution of the wild-type or nonpolymorphic genotype and mutant-type or polymorphic genotype. The Hardy–Weinberg equilibrium analysis result for this research was 28.61, which means that the genotype distribution in this research followed the Hardy–Weinberg equilibrium theorem, whereas the genotype and allele polymorphism distribution among the broader Indonesian population is likely to constant generation to generation.

CONCLUSION

This study found that ENAM C2452T polymorphism might represent a risk factor for the caries process. However, more substantial and well-designed studies in the future with larger population samples and which consider other correlating risk factors are still needed to further explore the role of ENAM gene polymorphism in the caries process.

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CONFLICTS OF INTEREST

The author has no conflicts of interest to declare.

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