Introduction

Exposure to spaceflight or simulated microgravity profoundly compromises the structure and function of a mammalian skeletal muscle, especially affecting muscular strength\textsuperscript{1,2}, motor tasks as well as performance control\textsuperscript{1}. Skeletal muscle changes are readily observed in simulated microgravity models such as dry immersion\textsuperscript{1-3}. Both gravitational forces and amount of cutaneous mechanosensitive pressure gradients of the floating body (head-out) in dry immersion (via interaction of the floating trunk and limbs covered by a thin waterproof fabric with water) are much smaller than, for example, in bed rest models (via interaction of head-down tilt body at supine position with mattress)\textsuperscript{4}. The rate at which muscle strength and transverse stiffness alter, and the pattern of recruiting of motor units in voluntary movements are also clearly different between these two body deconditioning models\textsuperscript{2}. Dry immersion therefore appears to be a model of choice for studying skeletal muscle responses on ground under conditions of whole body unloading with little mechanical support to the body surface which is very similar to real spaceflight.

Earlier, we did not observe any significant signs of atrophy (reduction in cross-sectional area of muscle fibers) in human soleus following 3 days of dry immersion (unpublished data). However, it has been previously shown that a 7-day exposure to dry immersion may lead to a series of changes in human postural muscle at the cellular level, specifically, 7-day immersion induced a decrease in diameter and maximum force of isometric tension in permeabilized soleus muscle fibers (MF), reduced calcium sensitivity and transverse stiffness of MF, decreased cross-sectional area of, predominantly, type I MF, as well as reduced relative content of titin, nebulin, desmin and alpha-actinin in soleus muscle\textsuperscript{5-7}. Thus, we can see that by the 7th day under conditions of support elimination, basic structural and a number of functional parameters in human soleus muscle are already changed. These changes developed against the background of the profound decline in the electrical activity of postural muscles\textsuperscript{8-9}. In earlier studies, it was shown that by the 3rd day of dry immersion slow type motor units had reduced activity or were “switched off”\textsuperscript{10}. In a rat experiment a decreased EMG activity of \textit{m. soleus} was observed already during the first
hours of support elimination. It is most likely that during the first hours/days under eliminated support slow motor units disabling would lead to the activation of the catabolic pathways, activating proteolytic processes and causing the destruction of cytoskeletal and contractile proteins. It also has been shown that during the first days of hindlimb suspension calcium ions concentration in myoplasm in rat soleus is increased, and activation of calpains is observed leading to degradation of a number of cytoskeletal proteins. The degradation of desmin, a typical substrate of μ-calpains, is observed in rat soleus after 3 days of hindlimb suspension. At the same time, one of the most powerful endogenous inhibitors of μ-calpains is nitric oxide (NO). NO is produced in MF by neuronal NO-synthase (nNOS) as well as by endothelial NO-synthase (eNOS). Interestingly, the activity of S-nitrosylated μ-calpain molecules is significantly lower than the activity of calpain molecules, which were not subjected to the action of NO. At the same time, in experiments with hindlimb-suspended rats and under space flight conditions as well as during head-down tilt bedrest in humans there was a decrease in nNOS content, and nNOS was translocated from sarcolemma to cytosol. A general reduction of NO-synthase under conditions of gravitational unloading (and in MF translocation to the cytoplasm, for example, due to disruption of chemical bonds with synaptophyins) could be associated with the action of μ-calpains. However, it is still unclear why unloading-induced proteolytic effect of calpains on NO-synthase is more powerful than inhibitory effect of NO on the activity of calpains. It is not excluded, that a decreased nNOS activity is followed by calpain-dependent breakdown of nNOS. It is obvious, that reduction of nNOS activity (as well as eNOS) as well as by endothelial NO-synthase (eNOS). It has been shown that inactivation of insulin/IGF-I-dependent protein kinases and via AMP-activated protein kinase (AMPK). It has been shown that nNOS phosphorylation can occur through insulin/IGF-I-dependent pathway under unloading is, primarily, caused by the degradation of the insulin receptor substrate - 1 (IRS-1) and via AMP-activated protein kinase (AMPK). It has been shown that nNOS phosphorylation in human soleus at the early stage of unloading may be of great scientific interest.

Thus, our study was aimed at the analysis of signaling processes that determine the initial development of proteolytic events in human soleus muscle during short-term dry-immersion. The results of the present study revealed that by the 3rd day of dry immersion there are signs of calpain-dependent proteolysis due to reduced total and phosphorylated nNOS content. We have also observed a significant decrease in AMPK phosphorylation.

**Materials and methods**

*Dry immersion model, subjects, ethics*

In dry immersion, the subject is immersed in a thermostatically controlled water bath while being protected from water with a thin waterproof fabric much larger than the area of the bath container. Thus, the subject’s body (with head-out) almost freely floats in warm water without getting a wet skin. Hydrostatic pressure is exerted equally on all body surfaces, nearly balancing the gravity force and thereby creating almost non-reaction-force conditions (Figure 1). The study group consisted of 6 healthy voluntary males with mean age of 22 (±2.23) and mean weight of 67.7 (±6.12) kg. Using microbiopsy technique samples of soleus muscle were taken before (pre-immersion) and after 3 days of dry immersion (post-immersion). The muscle samples were quickly blotted with gauze to remove superficial blood, frozen in liquid N2 for 20s, and then stored at -80°C until analysis. After the protocol of the experiment had been explained to volunteers, all the participants provided signed informed consent. The study protocol and consent form were approved by Biomedicine Ethics Committee of the Institute of Biomedical Problems, Russian Academy of Sciences (protocol no. 302, 25.07.2012) and all procedures conformed to the Declaration of Helsinki, International and Russian Law.

*Immunoblot analysis*

Skeletal muscle tissue (30 mg) was homogenized in ice-cold RIPA lysis buffer: 50 mM Tris (pH 7.4), 150 mM NaCl, 1% Nonidet P-40, 0.5% sodium deoxycholate, 0.1% SDS, 0.004% sodium azide, and 5 mM EDTA, supplemented with 1 mM DTT, 1 mM PMSF, and 5 μl/ml pepstatin (Sigma-Aldrich, St. Louis, MO, USA), mammalian protease inhibitor cocktail (Amresco, Solon, OH, USA), and phosphatase inhibitor cocktail B (Santa Cruz Biotechnology, Santa Cruz, CA, USA). The total protein concentration of the lysates was determined by incubation for 20 min at 4°C and centrifugation for 10 min at 12,000 g. Protein content of supernatants was quantified using an assay based on a modification of the Lowry protocol (RC DC Protein Assay; Bio-Rad Laboratories, Hercules, CA, USA). Bovine serum albumin was used as a standard. The samples were diluted in Laemmli buffer. Total protein (20-50 μg) was subjected to SDS-PAGE, and the proteins were then stained with Coomassie Blue R250.
transferred to nitrocellulose membrane (Bio-Rad Laboratories, CA, USA). Membranes were blocked for 1 h at room temperature with blocking buffer (4% nonfat milk powder; TBS, pH 7.4; and 0.1% Tween 20) and incubated overnight at 4°C with primary monoclonal antibodies against IRS-1 (diluted 1:900; sc-559, Santa Cruz Biotechnology, CA, USA), desmin (diluted 1:1000, Santa Cruz Biotechnology, CA, USA), anti-phospho-AMPKα1/2 (Thr172) (diluted 1:10000; Millipore Chemicals, MA, USA), nNOS (diluted 1:10000; BD Transduction Laboratories, NJ, USA) and glyceraldehyde-3-phosphate dehydrogenase (GAPDH; diluted 1:4000; Abcam, MO, USA). Antibodies were diluted in the blocking buffer. Three 10-min washes with TBS-Tween (TBS and 0.1% Tween 20) were then performed, after which membranes were incubated for 1 h at room temperature with horse-radish peroxidase-conjugated secondary antibodies to rabbit or mouse immunoglobulins (diluted 1:200,000; Bio-Rad Lab-
oratories, CA, USA). The membranes were washed again in TBS-Tween 3 times for 10 min, incubated in Immun-Star HRP Chemiluminescent system (Bio-Rad Laboratories, CA, USA), and exposed to X-ray film (Eastman Kodak, Rochester, NY, USA); the images were scanned then. The protein bands were quantified using densitometry scanning (GS-800; Quantity-One software, Bio-Rad Laboratories, CA, USA). Protein density was normalized to GAPDH. Western blots were performed at least 3 times.

**Statistical analysis**

The Wilcoxon signed rank test was used to compare proteins content before and after dry-immersion. The level of significance was set at p<0.05. The general content of the studied proteins was expressed as the median and interquartile range (25-75 percentiles).

**Results**

Three-day dry immersion resulted in 10% significant decrease (p<0.05) in cytoskeletal protein desmin general content in soleus muscle as compared to baseline values (Figure 2A). And in four of the six subjects decreased desmin concentrations were ranged from 12 to 24% (Figure 2B-C).

We did not find any significant changes in total nNOS content in soleus muscle after 3-day immersion (Figure 3A-C). However there was a 43% significant decrease (p<0.05) in the content of phosphorylated nNOS (Figure 4A-C).

Since a decrease in the content of phosphorylated nNOS can be caused by reduced protein-kinase activity of IGF-I/IRS-1/Akt pathway, an important factor in this case is IRS-1 content. In general, the content of IRS-1 in soleus muscle after 3-day dry immersion did not differ from the pre-immersion level, except for two subjects, which revealed an increase in the content of this protein (Figure 5A-C).

Another factor contributing to the reduction of nNOS phosphorylation could be the level of AMPK phosphorylation. Three days of dry immersion resulted in significantly reduced (by 36%) content of phosphorylated AMPK (Figure 6A) as compared to pre-immersion values (p<0.05). In five of the six subjects decreased phospho-AMPK content was in the range from 26 to 84%, and one subject revealed an increased level of this protein (Figure 6B-C).

**Discussion**

Exposure of humans or animals to short-term simulated microgravity is usually accompanied by enhanced proteolytic activity of calpains that can be indirectly assessed in vivo by determination of content of some calpains substrates, cytoskeletal proteins, such as desmin^16. Previous studies in our laboratory have shown that 7-day dry immersion leads to a decrease in desmin content in human soleus by about 30%^7. In the present study after 3 days of dry immersion there was a little, but significant decrease in desmin content (by 10%). So, it can be suggested that degradation of cytoskeletal proteins, which are μ-calpains substrates, takes place already at the early phase of gravitational unloading. We also observed a reduced nNOS phosphorylation during early period of unloading in human soleus muscle. Since NO is an inhibitor of calpain activities, the reduced nNOS phosphorylation may lead to possible activation of calpain-dependent processes in muscle cell. The decrease in the content of NO-synthase under unloading conditions was previously observed in leg muscles of both hu-
humans and laboratory animals\textsuperscript{22-24}. However, these observations were carried out during the longer periods of exposure to unloading conditions. So, we were the first who found a decrease in nNOS phosphorylation in soleus muscle during short-term exposure of human to dry immersion.

We know that NO is one of the effective μ-calpains inhibitors\textsuperscript{19,32}. However NO-synthase at the initial stage of the unloading seems to be unable to prevent an enhanced calpains activity. Obviously, at this period the activity of NOS is reduced due to changes in its phosphorylation which was shown in the previous reports\textsuperscript{15,33}. Indeed, in the present study the content of the phosphorylated nNOS in soleus muscle is significantly reduced (by more than 40%). It is known that a similar effect (a decrease in NOS phosphorylation level and NO production) can be induced by inactivation of AMPK\textsuperscript{28}, protein kinase D\textsuperscript{34} as well as by inactivation of IGF-I/insulin/Akt/mTOR signaling pathway\textsuperscript{29}. In the present study we did not observe any significant changes in IRS-1 content in soleus of the volunteers after 3 days of dry immersion. In
the study of Nakao et al.\textsuperscript{30} it was previously found that 5 and 10 days of gravitational unloading (rat hindlimb suspension) resulted in IRS-I breakdown due to activity of ubiquitin-ligase Cbl-b. Reduced IRS-I content in rat soleus was also observed after 14 days of hindlimb suspension\textsuperscript{35}. However, to date, any studies concerning the content of IGF-1/mTOR cascade key elements in a human postural muscle during early stages of the gravitational unloading have not been carried out yet. The fact that we did not observe any changes in IRS-1 content after 3 days of gravitational unloading in humans means not only that the trigger mechanism of E3 ubiquitin-ligases up-regulation (which may be associated with IRS-1 breakdown) is not typical for the early stages of unloading, but it also may indirectly show that the decrease in nNOS phosphorylation level is likely not caused by the changes in the activity of the canonical cascade kinases.

At the same time, in our study we observed a significant decrease in the content of phosphorylated AMPK (Thr 172) in human soleus muscle. Earlier, an altered level of AMPK phosphorylation on this site in rat soleus was registered in the two studies with two-week hindlimb-suspension\textsuperscript{35-36}. In a report by Han et al.\textsuperscript{35} a decreased AMPK phosphorylation was shown but Hilder and co-workers\textsuperscript{36}, on the contrary, observed an increased phosphorylation level of AMPK. In any case, we did not find any reports about changes of AMPK phosphorylation in animals or humans during short-term unloading. It is known that the level of AMPK phosphorylation is regulated by both systemic factors (e.g., interleukin-6\textsuperscript{37}) and intracellular mechanisms, first of all, the ratio between phosphorylated and non-phosphorylated high energy nucleotides (ATP/ADP/AMP)\textsuperscript{38}. Wakatsuki et al. (1994)\textsuperscript{39} showed that 10-day hindlimb suspension brought about the slight but significant increase in the basal CrP levels in rat soleus muscle. And this increase (although very small) was found in spite of the fact that the longer periods of the unloading were usually accompanied by the slowly increasing EMG activity (i.e. contractile activity)\textsuperscript{40}. So, one could expect the accumulation of the high-energy phosphates at the early stages of unloading, when no EMG activities were recorded. This accumulation might induce the reduction in the AMPK phosphorylation level, which was found in the present study. The AMPK phosphorylation is known to be a main up-stream mechanism of the PGC1α expression\textsuperscript{41}, therefore, the reduced level of phospho-AMPK during unloading might be followed by the reduced PGC1α expression which was recently found by Cannavino et al. (2015)\textsuperscript{42}. The lowered level of AMPK phosphorylation may also lead to a decrease in nNOS (as well as eNOS) phosphorylation level and reduced NO production\textsuperscript{43}.

Thus, short-term (3 days) exposure of human to dry immersion (gravitational unloading) resulted in the activation of proteolytic processes, accompanied by substantial inactivation of nNOS, one of the most important components of the proteolytic negative control mechanisms. Three-day dry-immersion also revealed a reduction in AMPK phosphorylation, which could serve as a trigger event for the development of primary atrophic changes in skeletal muscle.

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