Removal or component reversal of local geomagnetic field affects foraging orientation preference in migratory insect brown planthopper *Nilaparvata lugens*

Yingchao Zhang¹ ², Weidong Pan¹ ²

¹ Beijing Key Laboratory of Bioelectromagnetics, Institute of Electrical Engineering, Chinese Academy of Sciences, Beijing 100190, China; ² University of Chinese Academy of Sciences, Beijing 100049, China

Correspondence: Weidong Pan, Beijing Key Laboratory of Bioelectromagnetics, Institute of Electrical Engineering, Chinese Academy of Sciences, Beijing 100190, China. E-mail: panwd@mail.iee.ac.cn
Abstract

Background. Migratory brown planthoppers, *Nilaparvata lugens* (*N. lugens*), annually migrate to Northeast Asia in spring and return to Southeast Asia in autumn. However, mechanisms for orientation and navigation during their flight remain largely unknown. The geomagnetic field (GMF) is an important source of directional information for animals (including *N. lugens*), yet the magnetic compass involved has not been fully identified.

Methods. Here we assessed the influences of GMF on the foraging orientation preference of *N. lugens* by removing or component reversal of local GMF. At the same time, we examined the role of iron-sulfur cluster assembly1 (*IscA1*), a putative component of magnetoreceptor, in the foraging orientation preference of *N. lugens* under the controlled magnetic fields by RNA silencing (RNAi).

Results. We found that the near-zero magnetic field (NZMF) or vertical reversal of GMF could lead to *N. lugens* losing the foraging orientation preference, suggesting that a normal level of GMF, in the way of either intensity or inclination, was essential for the foraging orientation of *N. lugens*. Moreover, the gene knockdown of IscA1, also affected the foraging orientation preference of *N. lugens*, pointing out a potential role of IscA1 in the insects’ sensing the varying GMF.

Discussion. These results suggested a foraging orientation preference is associated with the GMF and revealed new insights into the relationship between the IscA1 and magnetosensitivity mechanism in *N. lugens*.

Subjects: Agricultural Science, Entomology

Key words: *Nilaparvata lugens*; Foraging orientation; Iron-sulfur cluster assembly1; Magnetosensitivity
1. Introduction

The brown planthoppers, Nilaparvata lugens (N. lugens), are recognized as a major migratory rice pest and virus vector. Adults exhibit wing dimorphism with macropterous and brachypterous phenotypes. The macropterous insects have functional wings for long-distance migration and the brachypterous individuals are non-migratory (Guerra 2011). In East Asia, N. lugens adults overwinter in Vietnam and southern China. In order to find enough food and suitable living environment, they migrate to Northeast Asia in spring and return back to Southeast Asia in autumn (Kisimoto 1976; Cheng et al. 1979). N. lugens is a nocturnal species that usually takes flight sometime between sunset and sunrise (Kisimoto 1979). However, the mechanisms for orientation and navigation during their flight remain largely unknown.

Many insects utilize magnetic information as a compass for their orientation and navigation. For instance, migratory butterflies such as the sulphur butterflies Aphrissa statira or the monarch butterfly Danaus plexippus can orient with a sun compass, but are also observed migrating directionally under overcast skies. Accordingly, it has been confirmed that a magnetic compass was involved in both species (Srygley et al., 2006, Guerra et al., 2014). It is also the case for some migratory moths as Mythimna separata that maintain migratory direction in the night sky (Xu et al., 2017). The use of a magnetic compass has also been found in foraging insects, including honeybee and ants. For honeybees, altering the GMF causes misdirection in the waggle dance, which is performed in the foraging trip, while there is no misdirection when dance orient along magnetic field lines (Towne & Gould, 1985). Ant foragers can be trained to recognize the location of a food source in the magnetic field and their orientation will turn according to the artificial magnetic fields (Anderson & Meer, 1993; Camlitepe & Stradling, 1995). In this respect, the use of magnetic compass as a key mechanism involved in insect directional movements appears feasible. This perspective was further supported by the findings of magnetic particles in insect tissues (Gould et al., 1978;... 

Deleted: which
Deleted: ..., and it...s a
Deleted: The adult insects
Deleted: …macropterous and brachypterous phenotypes.
The macropterous insects have functional wings for long-distance migration and the brachypterous individuals are non-migratory (Guerra 2011). In East Asia, N. lugens adults overwinter in Vietnam and southern China. In order to find enough food and suitable living environment, they migrate to Northeast Asia in spring and return back to Southeast Asia in autumn (Kisimoto 1976; Cheng et al. 1979). The... [2]
Deleted: insects
Deleted: as...a nocturnal species that usually takes off...light sometime between sunset and sunrise (Kisimoto 1979),...Hh...
Deleted: Actually,...my insects employ...lize the...
Deleted: The
Deleted: has been found in many insects
Deleted: theThe
Deleted: ..., such...such as the sulphur butterflies Aphrissa statira or the monarch butterfly Danaus plexippus or the sulphur butterflies Aphrissa statira
Deleted: ..., Accordingly, and...t has been confirmed that a magnetic compass was involved in A. statira Danaus plexippus and Aphrissa statira D. plexippus
Deleted: Guerra et al., 2014—Guerra et al, 2014Srygley et al. 2006... 
Deleted: the
Deleted: s
Deleted: Meanwhile,...e use of a magnetic compass has also been found in foraging of
Deleted: 
Deleted: the primary...key mechanism for...involved in...
Wajnberg, 1999; Chambarelli et al. 2008; Pan et al. 2016), which could become the substrate for the magnetic compass. However, until now no behavioral observations have been confirmed the presence of magnetic compass in N. lugens. So far there are two models which are most popular to explain how animals detect the magnetic field: the magnetite-based mechanisms (Beason 1986; Kirschvink & Gould 1981; Lohmann 2010) and the radical pair-based mechanisms (Ritz et al. 2000). Recently, a light-magnetism-coupled magnetosensitivity model has been proposed, in which the homolog of the bacterial iron-sulfur cluster assembly, IscA1, that forms a complex with cryptochromes is suggested to serve as a putative magnetic protein biocompass (Qin et al. 2016). The iron-sulfur cluster proteins are ancient macromolecules with highly conserved structures. They have many functions including iron homeostasis, electron transfer, metabolic catalysis, nitrogen fixation, regulation of gene expression and the detection of reactive oxygen species (Beinert et al. 1997; Beinert 2000). Qin et al. (2016) reported that the protein complex exhibited strong intrinsic magnetic polarity and rotated in synchrony with the external magnetic field. Previously we have found the IscA1 gene in N. lugens showed up-regulated mRNA expression during the period of migration (Xu et al. 2017). For the macropterous migratory N. lugens, compared with the GMF, the mRNA expression of the IscA1 gene and the cryptochrome1 gene were up-regulated under the magnetic fields of 0.5 millitesla (mT) and 1mT in strength. The findings revealed that the expression of IscA1 and cryptochromes in N. lugens exhibited coordinated responses to the magnetic field, suggesting the potential associations between IscA1 and the magnetic sensory system.

In this study, we demonstrated the effects of altered GMF, i.e., near-zero magnetic field (NZMF) or components reversal of GMF, on the foraging orientation in N. lugens. By using the RNA silencing (RNAi) on N. lugens, the functional role of IscA1 was investigated.
2. Materials and methods

2.1 Insect stock

Experiments were performed at Beijing Key Laboratory of Bioelectromagnetics, Institute of Electrical Engineering, Chinese Academy of Sciences, Beijing, China. The insects were reared in climate chambers at day/night temperatures of \((27\pm1)°C/(26\pm1)°C\) on susceptible Taichuang Native 1 (TN1) rice plant under 14:10 h light: dark cycle and 70±5% humidity (Wan et al. 2015) and the environmental conditions of the chambers was the same in the entire experiment. The TN1 rice plants were prepared in advance, and used as the food for the insects when they grew up to 10 cm height. The migratory macropterous female and male adults were selected from the same generation for the successive generations (Wan et al. 2016).

2.2. Magnetic field devices setup

The GMF used in the experiment (total intensity: \(52487\pm841\) nT; declination \(5.30\pm0.59°\); inclination \(56.29\pm1.02°\)) were the local GMF at \(39°59′14″N, 116°19′21″E\). The artificial magnetic fields were produced using a Helmholtz coil system (Fig. 1). For NZMF, the Helmholtz coils were used to produce a near-zero magnetic field region with an average intensity of \(\sim500\) nT at a center spherical space (150mm in radius). For component reversal of GMF, the Helmholtz coils were used to generate a magnetic field with twice intensity and reversed direction to offset either the horizontal component or the vertical component of GMF, producing a reversed inclination with the same intensity, but reversed component of GMF. Routinely before and after each experiment, we measured the three components of GMF using a fluxgate magnetometer (Model 191A, Honor Top Magnetolectric Technology Co. Ltd., Qingdao, China; sensitivity: \(\pm1\) nT) to modulate the electric current of the coil pairs to produce the required intensity for NZMF.

2.3. Cross-tube choice system and foraging orientation experiments
The choice system consisted of a cross tube which was embedded in a plastic square. The length of each arm of the cross tube was 110mm. The width was 20mm and the depth was 30mm. The cross tube was covered with a plastic lid of same size. There were small holes at each arm end for air flowing through. A lamp (15W, $\lambda=320-680$ nm) was installed as the light source (there is faint light when the $N. lugens$ takes off at the sunset or sunrise) with 400 lumen of lumination intensity at the cross tube. The coils system was covered by a shade cloth during the experiment to shield the external environment (Fig. 1). During the experiment, the cross tube was placed horizontally inside the Helmholtz coils and the arms of the cross tube were oriented towards four cardinal points. The cardinal points used in the experiments were the same. Two cross tubes were used in the experiment, one containing food as a reward and the other without food. Ten fresh rice seedlings of susceptible variety of TN1 (Fu et al. 2001) was placed at one arm end as food reward.

The experiment was conducted in two parts: the first part of the trial was to provide food as a stimulus that the insects might associate with magnetic cues under the normal GMF, and the second part of the trial was to test for a disruption of their ability to exhibit this learned directional preference when the GMF was altered. Adult insects within 48h of emergence regardless of gender or mating status were collected from the rearing colony and introduced into the center of the cross tube using a self-made insect suction implement. The rice seedlings were placed 70mm away from the center (Fig. 2A). For the first part of the trial, the insects gathering around the rice seedlings (40±13 insects) were collected (Fig. 2B, using the same suction-implement into a vial and afterwards a new cross tube with no rice seedlings inside was placed horizontally in the Helmholtz coils. For the second part of the trial, the collected insects in the vial were replaced in the center of the new cross tube using the suction-implement (Fig. 2C) and the insects moving to each of the four arms of the new cross tube were recorded after 0.5h (Fig. 2D). The entire experiment was performed at room temperature (26±1°C) and
each magnetic field setup was performed individually for 12 replicates. The total number of insects tested was 511±69.

2.4. The effects of IscA1 gene silencing on the orientation of *N. lugens*

The IscA1 gene was previously cloned in *Nilaparvata lugens* and the results showed that the gene expression reached the peak at the third day after emergence (Xu *et al.* 2017), so adults of 1<sup>st</sup> day after emergence were selected for RNAi according to Liu *et al.* (2010). The primers (Table 1) were designed based on the fragment sequence that was searched from transcriptome of *N. lugens* by local BLAST search. The dsRNA of IscA1 gene was designed at two different regions: nearing the 3’end (dsRNA1) and nearing the 5’end (dsRNA2). Insects were anesthetized with CO<sub>2</sub> for 30 s at PCO<sub>2</sub> = 1 mPa and immobilized on a 1.5% agarose plate with abdomen upward. Each insect was injected with 250 ng (50 nl in volume) dsRNA. On the second day after injection (24-48 h), the injected insects were collected and placed inside the GMF for foraging orientation test with the cross-tube system. The cross-tube behavioral trials for the RNAi insects were conducted in the same manner as described in section 2.3 as part of a two-step trial. A total of 700 insects were used for the experiments. To ensure the silencing efficiency, the expression level of *IscA1* gene was investigated before and after the behavioral test using 3 pools of 6 insects for each group by fluorescence-based quantitative real-time PCR (q-PCR) (Bustin *et al.* 2009). The whole body of adult *N. lugens* was used for sampling and all the samples were collected during the same time period (19:00-20:00 hours). Total mRNA was extracted by TRIzol reagent (Invitrogen, USA). The quality of samples was determined by spectrophotometric optical density (OD) 260/280 and 2% agarose gel electrophoresis. The cDNA templates were synthesized with 1 μg of total RNA using PrimeScript<sup>®</sup> RT reagent Kit with gDNA Eraser (TaKaRa, Tokyo, Japan). Each cDNA product was diluted with sterilized double-distilled water. The house-keeping gene for the q-PCR was *actin1* (GenBank accession No. EU179846, and the PCR amplification efficiency was established by means of
calibration curves (Bustin et al., 2009). The optimized thermal program was designed according to the kit instructions. Quantification of the transcript level of genes was conducted according to the ∆∆Cq method (Livak and Schmittgen, 2001). RNA samples were analyzed independently for three times. The dsRNA of green fluorescent protein-GFP (GenBank accession No. U76561) was injected into the N. lugens as the control.

2.7. Statistical analysis

All data were analyzed using SPSS 20.0 (IBM Inc., Armonk, U.S.A.). The chi-square test was used to analyze the ratio of the distribution of insects in four directions. If there was significant difference, Bonferroni correction was used to analyzed the difference between every two directions. One-way ANOVA was used to analyze the gene expression. Significant differences between dsGFP (control) and dsNl-IscA1 injection treatments were determined by one-way ANOVA at p < 0.05.

3. Results

3.1. The foraging orientation preference of N. lugens in the GMF vs NZMF

In the GMF, the N. lugens showed the highest preference for foraging orienting to the north direction with original food ($\chi^2=108.48$, p<0.001). The percentage of individuals orienting to the north was 39.06%, which was significantly higher than that to south 14% ($\chi^2=98.481$, p<0.001), west 25.40%($\chi^2=26.169$, p<0.001) and east 21.39% ($\chi^2=45.652$, p<0.001, Fig. 3A). In the NZMF, however, the N. lugens were relatively equally distributed and the percentage of individuals orienting to the north, south, west and east direction was 25.46%, 22.22%, 27.55% and 24.77%, respectively ($\chi^2=10.261$, p=0.083) (Fig. 3B).

3.2. The foraging orientation preference of N. lugens in the horizontal or vertical component reversal of GMF
In order to study how the GMF affects the foraging orientation ability of *N. lugens*, we conducted a behavior experiment in the horizontal or vertical component reversal of GMF. In the horizontal component reversal of GMF, most of the *N. lugens* were distributed in the north ($\chi^2=87.872, p<0.001$, Fig. 4A), similar to that in GMF. In the vertical component reversal of GMF, the percentage of insects orienting to the north, south, west and east direction observed as 25.18%, 21.74%, 29.53% and 23.55%, respectively ($\chi^2=9.371, p=0.102$) (Fig. 4B).

3.3. The IscA1 gene knockdown affected the foraging orientation preference of *N. lugens*

The q-PCR results showed that the mRNA expression of IscA1 was effectively downregulated after the gene silencing. Both of the silencing efficiencies of dsRNA1 and dsRNA2 were over 80% within 24h (before the behavioral experiment) and 48h (after the behavioral experiment) after microinjection (Fig. 5). Since the *N. lugens* preferred the north foraging direction in GMF or horizontal component reversal of GMF, we chose these conditions to investigate whether IscA1 gene silencing would affect the insects’ choice of direction. In these two magnetic fields, most of the *N. lugens* with IscA1 gene silencing distributed in the west, followed by the north, south and east (Fig. 6). Compared with the wild type, the percentage of IscA1 gene knockdown *N. lugens* distributed in the north direction significantly decreased from 39.06% to 28.82% in GMF ($\chi^2=13.183, p<0.001$, Fig. 6A) and to 28.57% in the horizontal component reversal of GMF ($\chi^2=10.151, p<0.001$, Fig. 6B).

4. Discussion

Previous studies revealed that exposure of both small brown planthopper and brown planthopper to NZMF delayed egg and nymphal developmental durations and decreased adult weight and female fecundity of insects (Wan et al. 2014). In addition to growth and development, the NZMF also affected positive phototaxis and flight behavior of insects has also been investigated.
In this study, we found majority of the *N. lugens* insect initially tested preferred north foraging orientation in the GMF, which was consistent with the field observation that *N. lugens* migrate to Northeast Asia under spring/summer-like conditions (Kisimoto 1976; Cheng et al. 1979). In our experiment, the first part of the trial with rice seedlings to the north provided the opportunity for the insects to associate magnetic field information under the normal GMF with the presence of food in a particular direction. 

The second part was designed to test whether the changed GMF (either NZMF in Figure 3 or component reversal in Figure 4) affected their ability to exhibit this learned directional foraging preference. As *S. furcifera* and *N. lugens* are both migratory insect pests of rice crops, the reported effects of removal of GMF suggest a role of the GMF, in terms of energy regulation or flight orientation in their local scale foraging movement and also possibly their long-distance migration.

Generally, the inclination compass worked when the vertical component of the geomagnetic field was reversed, as it was shown that the mealworm beetle *Tenebrio molitor* significantly turned their preferred direction by 180° when the vertical component was reversed (Vácha et al., 2008). It has also been reported that birds could not distinguish between north and south by the polarity of the geomagnetic field, but could distinguish poleward and equatorward movement by the inclination of the field lines (Wiltschko and Wiltschko, 1996). In this study, when the vertical component of GMF was reversed, *N. lugens* individuals showed no significant foraging orientation. Thus, the foraging orientation of *N. lugens* in the vertical component reversed magnetic field was partially consistent to the inclination compass observed in monarch butterfly (Guerra et al. 2014). In this study, when the vertical component was reversed, the *N. lugens* didn’t distribute in the opposite direction as did the monarch butterfly. We speculate that this may be due to the existence of multiple compasses involved in insect
orientation. *N. lugens* migrates in the sunset or sunrise (Kisimoto 1979), so it’s likely that a light-based mechanism of magnetoreception is also involved. Here our results suggested that a magnetic compass aided the foraging orientation preference of the migratory insect *N. lugens*. *N. lugens* might also use other orientation cues.

As the homologue of bacterial iron-sulfur cluster assembly, the IscA1 has been found in most prokaryotic and eukaryotic organisms with highly conserved structures. The inhibition of IscA1 could disrupt circadian rhythms in the fruit fly (Mandilaras & Missirlis 2012). Moreover, it was found that knockdown of the IscA1 led to anemia in zebra fish (Nilsson et al. 2009). Currently, a protein complex formed by the IscA1 interacting with cryptochromes was proposed as a putative magnetoreceptor and the protein crystal was claimed to exhibit strong magnetic polarity in response to an external magnetic field (Qin et al. 2016). The findings, however, have raised considerable controversy due to the broad interpretation of its biological meaning as well as the limitation of in vitro experiments (Friis et al. 2017; Hochstoeger et al. 2016; Pang et al. 2017). Therefore, an independent investigation should be performed to clarify as far as possible whether the IscA1 is involved in specific processes of magnetosensitivity in terms of functional behaviors as navigation and orientation in long-distance migration of animals (Meister 2016; Nicholls 2016). In this study, our results showed that the foraging orientation preference of insects was affected by the IscA1 gene knockdown under varying GMF, providing direct evidence of IscA1 involved in magnetosensitivity of *N. lugens*. Meanwhile, biogenic magnetic particles were proposed to function as a hypothetic magnetoreceptor: the external magnetic field can affect the internal magnetic clusters leading to magnetic orientation loss (Davila et al. 2005). Previously we have detected magnetic particles in *N. lugens* (Pan et al. 2016) which also provides additional support for a magnetic sense in *N. lugens*. Whether the IscA1 protein is functionally linked to formation of magnetic particles and how these hypothetic magnetoreceptors work in synergism in vivo remains to be further elucidated.
5. Conclusion

This study provided behavioral evidence that the foraging orientation preference of the migratory insect, *N. lugens*, is affected by removal or component reversal of local GMF. When the vertical component of GMF was reversed, the insects showed no significant foraging orientation preference, suggesting the potential use of inclination compass-aided orientation in *N. lugens*. The foraging orientation preference of *N. lugens* was also affected by JsCa1 gene knockdown, providing a feasible mechanistic explanation for the insects’ sensing of variation in the GMF. Further work is needed to investigate the potential associations between the JsCa1 and magnetic particles in terms of the magnetosensitivity mechanism in *N. lugens*.

Acknowledgements

We thank Prof. Hongxia Hua of College of Plant Science and Technology of Huazhong Agricultural University providing the original laboratory stock of the *N. lugens*. We also thank Space Electromagnetic Environment Laboratory for providing geomagnetic data.

Funding

This research was supported by the National Natural Science Foundation of China (31670855, 31870367, 31470454, 31672019, 31701787), the National Department of Public Benefit Research Foundation (201403031), the Natural Science Foundation of Jiangsu Province Youth Fund (SBK2016043525) and the China Postdoctoral Science Foundation (2016M590470). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

National Natural Science Foundation of China (NSFC): 31670855, 31870367,
13
31470454, 31672019, 31701787.

National Department of Public Benefit Research Foundation: 201403031.

Natural Science Foundation of Jiangsu Province Youth Fund: SBK2016043525.

China Postdoctoral Science Foundation: 2016M590470.

**Competing Interests**

The authors declare that they have no competing interests.

**Author Contributions**

- Yingchao Zhang conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.

- Weidong Pan conceived and designed the experiments, approved the final draft.

**Data availability**

The following information was supplied regarding data availability:

The raw data are available in a Supplemental File.

**References**

Anderson JB, Meer RKV. 1993. Magnetic orientation in the fire ant, Solenopsis invicta. Naturwissenschaften, 80:568-570. DOI:10.1007/BF01149274.

Beason RC. 1986. Magnetite biomineralization and magnetoreception in organisms: a new magnetism. by Joseph L. Kirschvink; Douglas S. Jones; Bruce J. MacFadden. Quarterly Review of Biology, 61, 429-430. DOI:10.1016/0300-9629(87)90453-1.

Beinert H. 2000. Iron-sulfur proteins: ancient structures, still full of surprises. Journal of Biological Inorganic Chemistry, 5, 2-15. DOI:10.1007/s007750050002.

Beinert H, Holm RH, Münck E. 1997. Iron-sulfur clusters: nature's modular,
multipurpose structures. *Science*, 277, 653-659.

DOI:10.1126/science.277.5326.653.

Bustin SA, Benes V, Garson JA, Heltelmans J, Huggett J, Kubista M, Mueller R, Nolan T, Pfaffl MW, Shipley GL, Vandersompele J, Wittwer C. 2009. The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. *Clinical Chemistry*, 55, 611-622. DOI: 10.1373/clinchem.2008.112797.

Camilitepe Y, Stradling DJ. 1995. Wood Ants Orient to Magnetic Fields. *Proceedings of the Royal Society B: Biological Sciences*. 261, 37-41. DOI:10.2307/50044.

Chambarelli LL, Pinho MA, Abraçado LG, Esquivel DM, Wainberg E. 2008. Temporal and preparation effects in the magnetic nanoparticles of apis mellifera, body parts. *Journal of Magnetism & Magnetic Materials*, 320, 207-210. DOI:10.1016/j.jmmm.2008.02.049.

Cheng SN, Chen J, Si H, Yang LM, Chu TL, Wu CT, Chen JK, Yang CS. 1979. Studies on the migrations of brown planthopper Nilaparvata lugens Stal. *Acta Entomologica Sinica*, 22(1), 1-21.

Davila AF, Winklhofer M, Shcherbakov VP, Petersen N. 2005. Magnetic pulse affects a putative magnetoreceptor mechanism. *Biophysical Journal*, 89, 56-63. DOI: 10.1529/biophysj.104.049346.

Fu Q, Zhang ZT, Hu C, Lai FX, Sun ZX. 2001. A chemically defined diet enables continuous rearing of the brownplanthopper, Nilaparvata lugens (stal) (homoptera : delphacidae). *Applied Entomology & Zoology*, 36, 111-116. DOI:10.1016/S0007-8506(07)60148-6.

Friis I, Sjulstok E, Solov'Yov IA. 2017. Computational reconstruction reveals a candidate magnetic biocompass to be likely irrelevant for magnetoreception. *Scientific Reports*, 7, 13908. DOI:10.1038/s41598-017-13258-7.

Gould JL, Kirschvink JL, Deffeyes KS. 1978. Bees have magnetic remanence.
Science, 201, 1026-1028. DOI: 10.1126/science.201.4360.1026.

Guerra PA. 2011. Evaluating the life-history trade-off between dispersal capability and reproduction in wing dimorphic insects: a meta-analysis. Biological Reviews, 86, 813-835. DOI:10.1111/j.1469-185X.2010.00172.X.

Guerra PA, Gegear RJ, Reppert SM. 2014. A magnetic compass aids monarch butterfly migration. Nature Communications. 5, 4164-4164. DOI: 10.1038/ncomms5164.

Hochsteoeg T, Nimpf S, Keays D, 2016. ISCA1 and CRY4: An improbable proposition. BioRxiv, 094458. DOI:10.1101/094458.

Kirschvink JL, Gould JL. 1981. Biogenic magnetite as a basis for magnetic field detection in animals. Biosystems, 13, 181-201. DOI: 10.1016/0303-2647(81)90060-5.

Kisimoto R. 1976. Synoptic weather conditions inducing long-distance immigration of planthoppers, Sogatella furcifera Horvath and Nilaparvata lugens Stal. Ecological Entomology, 1, 95-109. DOI:10.1111/j.1365-2311.1976.tb01210.x.

Kisimoto R. 1979. Brown planthopper migration. Brown Planthopper Threat to Rice Production in Asia. pp:113-124. International Rice Research Institute, Los Banos.

Lohman KJ. 2010. Animal behaviour: magnetic-field perception. Nature, 464, 1140-1142. DOI:10.1038/4641140a.

Liu SH, Ding ZP, Zhang CW, Yang BJ, Liu ZW. 2010. Gene knockdown by intro-thoracic injection of double-stranded rna in the brown planthopper, Nilaparvata lugens. Insect Biochemistry & Molecular Biology, 40, 666-671. DOI:10.1016/j.ibmb.2010.06007.

Mandilaras K, Missirlis F. 2012. Genes for iron metabolism influence circadian rhythms in Drosophila melanogaster. Metallomics, 4, 928-936. DOI:10.1039/c2mt20065a.
Meister M. 2016. Physical limits to magnetogenetics. *Elife*, 5, e17210. DOI:10.7554/eLife.17210.

Nilsson R, Schultz IJ, Pierce EL. 2009. Discovery of genes essential for heme biosynthesis through large-scale gene expression analysis. *Cell Metabolism*, 10, 119-130. DOI:10.1016/j.cmet.2009.06.012.

Pan WD, Wan GJ, Xu JJ, Li XM, Liu YX, Qi LP, Chen FJ. 2016. Evidence for the presence of biogenic magnetic particles in the nocturnal migratory brown planthopper, Nilaparvata lugens. *Scientific Reports*, 6, 18771. DOI:10.1038/srep18771.

Pang K, You H, Chen Y, Chu P, Hu M, Shen J, Guo W, Xie C, Lu B. 2017. MagR alone is insufficient to confer cellular calcium responses to magnetic stimulation. *Frontiers in Neural Circuits*, 11, 11. DOI:10.3389/fncir.2017.00011.

Qin SY, Yin H, Yang CL, Dou YF, Liu ZM, Zhang P, Yu H, Huang YL, Feng J, Hao J, Deng LZ, Yan XY, Dong XL, Zhao ZX, Jiang TJ, Wang HW. Luo SJ, Xie C. 2016. A magnetic protein biocompass. *Nature Materials*, 15, 217-226. DOI:10.1038/nmat4484.

Ritz T, Adem S, Schulten K. 2000. A model for photoreceptor-based magnetoreception in birds. *Biophysical Journal*, 78, 707-718. DOI:10.1016/S0006-3495(00)76629-X.

Srygley RB, Dudley R, Oliveira EG, Riveros AJ. 2006. Experimental evidence for a magnetic sense in Neotropical migrating butterflies (Lepidoptera: Pieridae). *Animal behaviour*, 71, 183-191. DOI:10.1016/j.anbehav.2005.04.013.

Towne WF, Gould JL. 1985. Magnetic field sensitivity in honeybees. *Magnete Biomineralization and Magnetoreception in Organisms*. pp385-406. New York: Plenum. DOI: 10.1007/978-1-4613-0313-8-18.

Vácha M, Dršt ková D, Pů řová T. 2008. *Tenebrio* beetles use magnetic inclination compass. *Naturwissenschaften*, 95, 761-765. DOI: 10.1007/s00114-008-0377-9.
Wajnberg E, Linhares MP. 1999. Evidence for magnetic material in the fire ant solenopsis sp. by electron paramagnetic resonance measurements. Naturwissenschaften, 86, 30-32. DOI: 10.1007/s001140050564.

Wan GJ, Jiang SL, Zhao ZC, Xu JJ, Tao XR, Sword GA, Gao YB, Pan WD, Chen FJ. 2014. Bio-effects of near-zero magnetic fields on the growth, development and reproduction of small brown planthopper, Laodelphax striatellus and brown planthopper, Nilaparvata lugens. Journal of Insect Physiology, 68, 7-15. DOI: 10.1016/j.jimsphys.2014.06.016.

Wan GJ, Wang WJ, Xu JJ, Yang QF, Dai MJ, Zhang FJ, Pan WD, Chen FJ. 2015. Cryptochromes and hormone signal transduction under near-Zero magnetic fields: new clues to magnetic field effects in a rice planthopper. Plos One, 10, e0132966. DOI: 10.1371/journal.pone.0132966.

Wan GJ, Yuan R, Wang WJ, Fu KY, Zhao JY, Jiang SL, Pan WD, Sword GA, Chen FJ. 2016. Reduced geomagnetic field may affect positive phototaxis and flight capacity of a migratory rice planthopper. Animal Behaviour, 121, 107-116. DOI:10.1016/j.anbehav.2016.08.024.

Wan GJ, Liu RY, Li CX, He JL, Pan WD, Sword GA, Hu G, Chen FJ. 2020. Change in geomagnetic field intensity alters migration-associated traits in a migratory insect. Biology Letters, 16 (4). DOI: 10.1098/rsbl.2019.0940

Wan GJ, Jiang SL, Zhang M, Zhao JY, Zhang YC, Pan WD, Sword GA, Chen FJ. 2021. Geomagnetic field absence reduces adult body weight of migratory insect by disrupting feeding behavior and appetite regulation. Insect Science, 28(1):251-260. DOI: 10.1111/1744-7917.12765.

Wiltschko W, Wiltschko R. 1996. Magnetic orientation in birds. Journal of Experimental Biology, 199, 29-38. DOI: 10.1007/978-1-4615-6787-5_2

Xu JJ, Pan W, Zhang YC, Li Y, Wan GJ, Chen FJ, Sword GA, Pan WD. 2017. Behavioral evidence for a magnetic sense in the oriental armyworm, Mythimna...
separate. *Biology Open*, DOI: 10.1242/bio.022954.

Xu JJ, Zhang YC, Wu JQ, Wang WH, Li Y, Wan GJ, Chen FJ, Sword GA, Pan WD. 2019. Molecular characterization, spatial-temporal expression and magnetic response patterns of the iron-sulfur cluster assembly1 (IscA1) in the rice planthopper, *Nilaparvata lugens*. *Insect Science*, 26, 413-423. DOI:10.1111/1744-7917.12546.

Zhang YC, Wan GJ, Wang WH, Li Y, Yu Y, Zhang YX, Chen FJ, Pan WD. 2019. Enhancement of the geomagnetic field reduces the phototaxis of rice brown planthopper *Nilaparvata lugens* associated with frataxin down-regulation. *Insect Science*, DOI: 10.1111/1744-7917.12714.
