TP53 and EGFR Mutational Status in Thymoma: A Genetic Sequencing Study

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Abstract

Background and objective: Thymoma is a rare malignant tumor that usually with an indolent presentation, which was falsely assumed to be benign previously. The tumor suppressor P53 (TP53) and EGFR gene mutate most frequently in human cancers. We tried to investigate the presence of TP53 and EGFR mutations among thymoma patients referred to an Indonesian referral respiratory hospital and to discuss its potential role in thymoma management and prognosis.

Material and methods: Surgically resected tumor tissues were collected from thymoma patients and then underwent genomic analysis. PCR was performed on the extracted Paraffinized DNA to amplify exon 6 of TP53 and exons 18, 19, and 21 of EGFR. The evaluation of mutational status was done using direct sequencing and sequence analysis of purified PCR products. Results: Among 27 collected samples, TP53 exon 6 mutation, namely missense mutation and nonsense mutation, was observed in two samples (7.4%). EGFR exon 18 mutation, namely E709K and nonsense mutation, was found in 2 samples (7.4%). An intronic mutation in EGFR exon 19 (3.7%) and exon 21 (3.7%) was observed in one sample.

Conclusion: TP53 and EGFR mutations were not most frequent, so it seems that these genes are not involved in the pathogenesis of thymoma in Indonesian patients. Nevertheless, we found two samples with a significant mutation in p53 and EGFR genes, suggesting further research on thymoma prognostification and targeted therapy.

Keywords: EGFR- TP53- Thymoma

Introduction

Thymoma is a rare malignant tumor usually with an indolent presentation, which was falsely assumed to be benign previously. The actual incidence of thymoma is unknown; however, 47% of anterior mediastinum tumors are thymoma (Dettetback et al., 2013). From 1970 to 1990, thymoma accounted for 24% of 137 cases of surgically resected mediastinum tumor in Persahabatan National Respiratory Referral Hospital Indonesia (PDPI, 2003). Around one-third of thymoma patients suffer from myasthenia gravis (Dettetback et al., 2013).

Thymoma staging, histologic type, and complete eradication through any treatment modalities are significant to increase the survival of patients and reduce its recurrence rate. Prognosis of thymoma is worse at higher stages of the disease. The patients at the advanced stage of the thymoma have a 5-year median survival of 69%. Currently, the treatment choice in thymoma is surgery providing a better prognosis (Dettetback et al., 2013, NCCN, 2016). However, the role of molecular therapy in this tumor, such as targeted therapy, is unknown.

The tumor suppressor 53 (TP53) is the most frequently mutated gene in human cancer and known as “the guardian of the genome”. TP53 role is a gatekeeper of cell growth and division acting a central role in gene induction for cell cycle arrest and apoptosis when DNA is damaged (Giacomelli et al., 2018). Thus, this protein role is vital for suppressing the growth of the tumor and reducing cell malignant transformation. Alteration of this gene was observed in somatic mutation with a specific clinical phenotype for each cancer. TP53 is known as a marker that can predict therapeutic response and prognosis in squamous cell lung carcinoma (Feng Xu et al., 2020, Kobayashi et al., 2015). In addition, its higher level detection was associated with better prognosis in other cancers, namely, acute myeloid leukemia (Sadek, 2020).

Epidermal growth factor receptor (EGFR) is a receptor with tyrosine kinase activity, which is important in transducing signals from extracellular into the intracellular (De Luca, 2008). EGFR mutation has been reported in non-small cell lung carcinoma cases and is frequently
mutated in exon 18, 19, 20, and 21. Commonly, the mutation is missense. For instance, G719S, L858R, and L861Q (Lynch et al., 2004, Kobayashi et al., 2015) mutations are characterized by the short deletion in exon 19 and point mutations in exons 19 and 21. Those three mutations are associated with higher sensitivity to tyrosine kinase inhibitors (TKI). However, D761Y (Balak et al., 2006) and T790M (Lee et al., 2021) mutations are resistant to this treatment. The presence of EGFR mutation also better predicts overall survival (OS) (Okamoto et al., 2018), response to TKI therapy as well as progression-free survival (>9 months) in NSCLC (Pas-Arez et al., 2009, Lynch et al. 2004).

This study investigated the presence of TP53 and EGFR mutations among thymoma patients in an tertiary respiratory referral hospital, Indonesia, and discussed its impact with respect to prognostification and therapeutic aspects on anti-EGFR therapy. Genetic sequencing using direct sequencing and sequence analysis in TP53 exon 6 and EGFR exon 18, 19, and 21 were also done.

**Materials and Methods**

**Study participants and DNA extraction**

This study was approved by the Ethic Committee of Persahabatan Hospital (No 139/KEPK-RSUPP/12/2020). Thirty deparaffinized histopathology samples were collected from thymoma patients diagnosed at the Persahabatan National Referral Respiratory Hospital, Indonesia. The samples were obtained from surgically resected tumor tissues and chosen to undergo genomic analysis. DNA was extracted from Formalin Fixed Paraffin Embedded using Quick-DNA FFPE mini prep (Zymo Research, USA) according to manufacturer’s protocols as follows. Approximately, FFPE were sliced using microtome in 1-2 slices and 4 µm thickness each. Slices were transferred to 1.5 ml centrifuge tubes. Deparaffinization, tissue digestion, and DNA purifications were carried out according to manufacturer’s protocols. The DNA was eluted in 50 µl of DNA elution buffer. DNA concentrations and purity were measured using a Biodrop Nano spectrophotometer. Further, a polymerase chain reaction (PCR) was performed for all DNA samples. The clinicopathological data of specimens, including patient’s age and sex, WHO subtype, and Masaoka staging were also collected.

**Mutation analysis**

PCR amplification was performed for all DNA samples for TP53 exon 6 and EGFR exon 18, 19, and 21 using MyTaq™ Hotstar RedMix PCR mix (Bioline, United Kingdom). The process was done based on the manufacturer’s protocol. The primers and their product size base pair are presented in Table 1. A PCR reaction consisted of PCR buffer, 0.2 mM dNTP, 0.3 µM forward and reverse primers, 1.25 Taq DNA polymerase, and 250 ng of DNA with a total volume of 50 µL. PCR cycling conditions included pre-denaturation step in 95°C for 1 min. The steps were followed by denaturation 95°C for 15 s, annealing in 65°C for 15 s, and extension 72°C in 15 s for 40 cycles. The steps were continued by final extension under 72°C for 10 minutes afterward.

The PCR product was then loaded and analyzed using 2% of agarose gel electrophoresis. Non-template control was done using PCR primer mix without any DNA to detect unwanted DNA contaminations within PCR mix. Samples were then run in 2% of agarose electrophoresis in Tris Buffer EDTA pH 8.0.

**DNA sequencing and sequence analysis**

DNA sequencing was performed by using Sanger Sequencing according to 1st BASE Asia standard Laboratory procedure (First base Asia, Malaysia). The sequencing results were imported, converted to reverse complement, and analyzed using ClustalW in BioEdit Sequence Alignment Editor 7.2.5 editions. The reverse sequence was also analyzed to human reference genome NG_007524.2 (LRG_433) using nBLAST NCBI and standard preferences. The BLASTn results were also aligned with corresponding coding sequences (CDS). Lastly, the sequence was analyzed using FinchTv 1.4 and reverse complement views.

**Results**

**Tumor characteristics**

Clinical characteristics of the entire samples are present in Table 2. The majority of collected samples were obtained from male patients with a median age of 55. The primary thymoma staging and WHO subtype was IIB and type A, respectively.

Genomic DNA was extracted from 30 surgically resected thymoma tissues. Three samples were failed to amplify for TP53 exon 6 and EGFR exon 18, 19, and 21 so they were discarded. Twenty seven samples were successfully underwent PCR, gene sequencing, nBLAST, and electropherogram analysis.

**Detection of mutation**

We observed several mutations in different exons of our samples, and the findings are summarized in Table 3. TP53 mutations were detected in two samples. These samples showed different base pairs illustrating a point mutation. A missense mutation was observed in one sample, at positions 155 from G>A in exon 6. Another sample had substitution from 19319 T>G (silent mutation), sample, at positions 155 from G>A in exon 6. Another sample had substitution from 19268 T>G (silent mutation).

| Gene and Exon | Forward Primer | Reverse Primer | PCR Product Size (bp) |
|---------------|----------------|----------------|----------------------|
| TP53 Exon 6   | CAGGTCCTCCCACAGGCGAC | GCAAGCAGAGGCTGGGACAC | 219 |
| EGFR Exon 18  | ATGCTGGGCAGCTGTTCCA | CACAGGGCCACTAGTTACT | 392 |
| EGFR Exon 19  | TCACCTGGGCACGATGGGCA | CAGCTGGCCAGCATCGGAAA | 241 |
| EGFR Exon 21  | ATTCGGATGCAGACTCTT | CCTGGTGTCCAGGAAAATGCT | 265 |

**Table 1. Primer Sequences**

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was found in one sample at g.177642 C>T.

**Discussion**

Based on the findings of this study, there were several mutations, including missense and silent mutation, in TP53 exon 6, and missense (E709K) mutation in EGFR exon 18. In addition, it was found that two samples had intronic substitution in exons 20 and 21. In a previous study, p53 mutation was found in thymoma, but with lower frequencies or in zero prevalence (Franz et al., 2017, Oberndorfer et al., 2019, Peric et al., 2019, Girard et al., 2009, Lei Yu et al., 2019, Li et al., 2019). The aforementioned study showed that 7.14% of p53 genes were mutated in the samples. In general, a missense mutation in p53 gene is the most common finding in human cancers (73.16%) (Bouaoun et al., 2016), which is in line with our study.

**P53** is known as the most commonly mutated gene in malignant tumors. Thus, p53 mutation could be a predictor of tumor aggressivity since its appearance is common in thymic carcinoma. Moreira et al., (2015) observed that not a single sample of B3 thymoma had p53 mutation; whereas, thymic carcinoma showed chromosomal...
gains and losses. Moreover, thymic carcinoma had p53 chromosomal gain or losses, which are responsible for its aggressive behaviour. Other recent studies also confirmed that p53 and EGFR were frequently spotted in thymic carcinoma (Sakane et al., 2021; Zhan et al., 2018; Prays et al., 2021).

Previous studies reported contrasting findings on the presence of EGFR mutation in thymoma. For instance, Suzuki et al., found that direct sequencing in did not reveal any EGFR missense mutation in 38 samples obtained from Japanese patients diagnosed with thymoma (Suzuki et al., 2006). On the other hand, Yoh et al., (2008) depict two of 20 patients with thymoma. They also observed that all of the EGFR mutations found were missense mutations in exon 21. Further, in another study, Sakane et al., performed direct sequencing in exon 19 of EGFR and indicated no mutation in EGFR gene (Sakane et al., 2019). Based on these studies and our results, it seems that EGFR mutation in thymoma is rare and potentially might not be related to thymoma carcinogenesis.

To best of our knowledge, few studies discussed intronic mutation in thymoma. The intronic mutation is also known as a cancer-related mutation, especially in mis-splicing more than 20 base pairs (Jung et al., 2021). However, Meister et al., (2007) reported that some thymoma patients had a single base pair variation in an intron and also a non-tumor cell. However, a recent meta-analysis reported that polymorphism in TP53 codon 72 G>C might be associated with increased risk of thyroid cancer (Nedooshan, 2016). Hence, further studies are needed to reveal its clinical implication, disease progression, targeted therapy, and prognosis.

Since p53 and EGFR are possibly not the founders of carcinogenesis in this tumor, another somatic mutation, rather than p53 and EGFR, should be considered as the possible driven carcinogenesis gene in thymoma. Using a high-throughput approach, one study in Serbia revealed that ten genes other than EGFR, including FBXW7, FGFR3, GNAQ, GNA11, HNF1A, KIT, MET, PIK3CA, PTEN, and RB1, harbored 40 variants of nonsense, missense, and frameshift mutations (Peric et al., 2019). Franz et al., (2017) using genetic sequencing of 50 mutational hotspots, found five mutations in 37 samples of thymoma, HRAS mutation in 3 cases of type A thymoma, and SMARCB1 and STK11 genes in one case of type B3 thymoma. GTF2I is also considered a potential driven gene in thymoma since its correlation with myasthenia gravis is supported (Radovich et al., 2018). In the present study, direct sequencing and sequence analysis of TP53 and EGFR genes were only performed; thus, the primary gene inducing carcinogenesis could not be further investigated in our samples.

Few studies discussed genetic variant among histological spectrum of thymoma. A number of studies investigated other molecular profiles of TP53 and EGFR mutation in terms of thymoma type (Girard et al., 2009, Oberndorfer et al., 2019; Suzuki et al., 2006). Girard et al., reported no p53 and EGFR mutation in thymoma type A, B2, and B3 (Girard et al., 2009). The proportion of p53 mutation was lower in thymoma (thymoma type A, 4.2%) than thymic carcinoma (21.7%) according to a previous study (Oberndorfer et al., 2019). A cohort study on forty Chinese patients with thymic epithelial tumor reported no p53 and EGFR mutation in thymoma patients (Wang et al., 2021). This research findings were in line with the abovementioned studies, revealing that the predominant subtype was thymoma type A. EGFR and p53 mutations were frequently spotted in advanced stage of the disease (Sakane, 2021).

In brief, it seems that thymoma type does not affect the genetic alterations. However, further studies on alteration in gene expression profiles of thymoma are needed. A recent meta-analysis suggested that some biomarkers in thymoma correlated with patients’ prognostic (Zeng H, 2020). However, the evidence showing that TP53 and EGFR mutations are the prognostic marker remains inconclusive. However, some past study depicted TP53
as an indicator of worse prognosis in thymoma. Using International Agency for Research on Cancer (IARC) TP53 database, Victor et al., concluded that TP53 mutation presence could significantly worsen median disease-free survival time (9.72 months vs. 115.05 months, with and without TP53 mutation, respectively) (Victor et al., 2018). This finding suggested that p53 and EGFR could play an essential role in determining patients’ prognosis and further urgency to be investigated.

Currently, the cornerstone of thymoma therapy is surgical resection. Moreover, chemotherapy and radiotherapy show promising results for stages III and IV (Detterback et al., 2013). Nevertheless, the role of targeted therapy, especially in TKI in thymoma, is unknown. Even though E709K is a rare mutation known in lung carcinoma (Frega et al., 2016), its detection has potentially become a good candidate for treatment using the 2nd generation TKI, namely, afatinib and neratinib (Kobayashi et al., 2015). Hopefully, the presence of this mutated gene, as displayed in this study, will have the same effect on thymoma patients.

The main limitation of our study was the relatively small sample size. In addition, we did not identify the role of p53 and EGFR in predicting patients’ prognosis.

In conclusion, our study found that the mutational frequency of exon 6 of p53 and exons 18, 19, and 21 of EGFR genes was low in thymoma. These genes are probably less critical as the primary oncogenic driver in thymoma Indonesian patients. Nevertheless, this study showed two samples with a significant mutation in p53 and EGFR genes, indicating the need for further research on thymoma prognostication and targeted therapy.

**Author Contribution Statement**

ES, JZ, DRN conceived, designed and performed the analysis; JZ, RS, RB, DRN, MRF collected the data and did experimental tools; ES, JZ, DRN, MRF wrote the paper. All authors read and approved the final version of this manuscript.

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**Ethical Approval**

This study was reviewed and approved by Persahabatan Hospital Ethic Committee, Ref no : 139/KEPK-RSUPP/12/2020.

**Availability of data**

The primary data could be obtained upon request and should be approved by Ethical Committee.

**Conflict of Interest**

Authors affirm no conflict of interest.

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