The soybean cultivar SCEL-1 shows potent anti-photoaging effects in a UV-induced three-dimensional human skin and hairless mouse model

Jin Woo Lee†, Lei Peng†, Hyun Jegal1, No-June Park1, Sim-Kyu Bong1, Joon Won Lee1, Jeong Joo Pyo1, Yongsso Choi1,2* and Su-Nam Kim1*

Abstract

Soybean (Glycine max) is one of the major sources of nutrition and is used as a raw material for food and as a source of feed for livestock. The efficacy of soybeans on skin health includes their ability to reduce wrinkles and pigmentation and increase skin elasticity and moisture content. Black soybean has been consumed worldwide for a long time, especially in Korea, and is used as a medicinal food against several disorders related to the skin. To evaluate whether its effect on the skin is different based on the cultivar of soybeans, three black soybean cultivars collected in Korea, Soybean Core collection Elite Line-1 (SCEL-1), Chung Ja-3 (CJ-3) and Won Heuk (WH), were selected to compare their effect on improving photoaging induced by ultraviolet rays (UVs). We found that SCEL-1 exhibited the best efficacy among the three cultivars tested, and treatment with this soybean extract significantly reduced the expression of matrix metalloproteinase-1 (MMP-1), preventing the degradation of collagen in a 3D human skin model. In addition, SCEL-1 application improved wrinkle- and photoaging-related symptoms, such as epidermal thickening, collagen deficiency and immune cell infiltration, in an animal model established by UV irradiation. Procyanidin B2 and epicatechin isolated from the SCEL-1 cultivar inhibited MMP-1 biosynthesis in UVB-irradiated human dermal fibroblasts, and these two major components are likely related to more significantly attenuated skin photoaging. Therefore, our results indicated that SCEL-1 exhibits good anti-wrinkle effects compared to the other two black soybean cultivars, suggesting that it represents an excellent agent for anti-photoaging.

Keywords: Glycine max, SCEL-1, Photoaging, MMP-1, 3D human skin model

Introduction

Soybean (Glycine max) is one of the major nutritional sources belonging to the Fabaceae family and is cultivated globally, primarily in temperate regions [1]. Soybean contains high-quality protein, healthy oil, and low carbohydrates and has been used as a raw material for food and as a source of feed for livestock using suitable cultivars [2]. Many studies conducted over the past 30 years have reported that soybeans have various health-related benefits, especially for the prevention and treatment of chronic diseases. For example, soybeans lower the risk of cardiovascular disease, improve bone health, lower the risk of breast and prostate cancer, ameliorate kidney disease, relieve menopausal symptoms, and improve cognitive function and mental health [3–10].

Soybean and its functional components have a good effect on skin health, and the effects of soybeans on the skin are known to reduce wrinkles and pigmentation and...
increase skin elasticity and moisture content [11–15]. Soybeans contain active constituents, such as phenolic acids, including syringic, ferulic, and sinapic acids; isoflavones that act as both phytoestrogens and selective estrogen receptor modifiers (SERMs); small proteins that act as protease inhibitors; and proanthocyanidins [1, 16]. These ingredients have been reported to have anti-inflammatory, depigmentation, elastin and collagen fiber production, UV protection, and prevention of lipid peroxidation effects on the skin [1].

**Glycine max**, a cultivated soybean, has a variety of shapes, colors, sizes, and components, depending on genetic diversity. Since soybean is an important crop with a very high utilization value in the food and livestock feed industries, data on the genetic resources of more than 20,000 kinds of soybeans exist, including elite lines and wild soybeans from various countries, such China, Japan, and Korea, and are preserved in the USDA Soybean Germplasm Collection in the United States [17]. In Korea, various soybean resources have also been collected and analyzed to establish a unique soybean core group. Black soybean, which features yellow or green cotyledons in black seed coats, has been consumed worldwide for a long time, especially in Korea, and is used as a medicinal food against several disorders [18].

In this study, to evaluate whether the effect on the skin is different depending on the cultivar of soybeans, 3 black soybean cultivars collected in Korea, Soybean Core collection Elite Line-1 (SCEL-1), Chung Ja-3 (CJ-3) and Won Heuk (WH), were selected to compare their effects on attenuating photoaging induced by UVB. As a result, it was confirmed that the SCEL-1 cultivar exhibited remarkable anti-photoaging efficacy compared to the other two species and contained procyanidin B2 and epicatechin, demonstrating potent efficacy for reducing expression of matrix metalloproteinase-1 (MMP-1).

**Materials and methods**

**Plant materials**

Three black soybean cultivars, SCEL-1, CJ-3 and WH, were donated by the National Institute of Crop Science in Korea. The three soybeans were grown in a field in Paju, Korea, from May to October 2017. Soybeans were air-dried and stored at 10 °C in the dark until use. All cultivated plants were managed by essentially normal agricultural practices according to the standard protocol of the Rural Development Administration (http://www.nongsaro.go.kr/), including irrigation, fertilizer application, and pest control.

**Extraction**

Soybean extraction was performed according to a previous report with slight modification [19]. Dried soybeans were ground into fine powder, passed through a 60 mesh sieve, and 3 g of the resulting powder was mixed with 70% aqueous ethanol and sonicated for 15 min. Then, the solution containing soybean powder was retained at room temperature in the dark for approximately 14 h. Finally, the solution was sonicated again for 15 min. The extracted solution was passed through filter paper (Hyundai Micro, Size 300 mm, Grade No. 100, Seoul, South Korea), and ethanol in the extracted solution was evaporated using a SpeedVac evaporator (Thermo Fischer Scientific, Model SPD2010-220, Waltham, MA, USA). The remaining solution was completely dried using a freeze dryer machine (Operon, model FDCF-12003, Gimpo, Gyeonggi-do, South Korea). The final extracted powder was stored at 4 °C until use.

**Three dimensional human skin and cells**

A three-dimensional (3D) full thickness model of human skin (EpiDermFT™) was purchased from MatTek Corporation (Ashland, MA, USA). The skin equivalent was maintained in Dulbecco’s modified Eagle (DME)-based medium supplied by the manufacturer (EFT-300-MM) at 37 °C and 5% CO2. Normal human dermal fibroblasts (HDFs) were obtained from Lonza (Walkersville, MD, USA) and cultured in Dulbecco’s modified Eagle’s medium (DMEM; Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum (FBS; HyClone Laboratories Inc., Logan, UT, USA) and 1% penicillin/streptomycin (Invitrogen) at 37 °C and 5% CO2.

**UV irradiation**

A fluorescent UVB lamp (Sankyo Denki G15T8E, Sankyo Denki, Japan) with an emission spectrum between 280 and 360 nm at a peak of 312 nm was used as a UV light source. Human skin equivalents were treated with each reagent for 1 h, irradiated with UVB at 40 ml/cm2 and subsequently changed to reagents in serum-free medium for 72 h. Cells were seeded into 6-well plates, incubated for 24 h, washed with PBS, and serum-free medium was added. After 24 h, cells were pretreated with each reagent for 1 h before UV irradiation. The total energy dose of UVB irradiation was set to 20 ml/cm2 to optimize cell viability and MMP-1 stimulation. Thereafter, cells were irradiated with UVB and treated with reagents in serum-free medium for 48 h.

**Enzyme-linked immunosorbent assay (ELISA)**

Supernatants from skin equivalents were harvested and subjected to ELISA. MMP-1 and procollagen type-I secretion were quantified from supernatants using a human MMP-1 and procollagen type-I ELISA Kit (R&D Systems, Minneapolis, MN, USA). Relative levels of MMP-1 and procollagen type-I were normalized to
corresponding cell viability as measured by MTT assay (Ez-cytox, Dail Lab Service. Co., Seoul, Korea).

Animal experiments
SKH-1 hairless mice (female, seven weeks old) were purchased from Orient Bio Inc. (Seongnam, Korea). All experimental procedures involving animals complied with the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health (NIH Publication No. 85-23, 2011 revision). Procedures were approved by the Institutional Animal Care and Use Committee (IACUC) of Korea Institute of Science and Technology (Certification No. KIST-2016-011). SCEL-1, CJ-3, and WH (25, 50 or 100 mg/kg/day) in vehicle solution (0.5% carboxymethylcellulose) were orally administered to hairless mice for 8 weeks under UV exposure conditions. UV irradiation protocols were performed as follows. During the first week, a UV dose of 1 minimal erythema dose (MED) corresponding to 100 mJ/cm² was applied to the backs of hairless mice. The intensity was increased by 1 MED every 2 weeks for up to 8 weeks. At the end of the experiment, mice were irradiated with 4 MEDs.

Generation of replicas and image analysis
An image of the back skin of each mouse was acquired immediately after the experiment was over. Then, replicas of dorsal skin from sacrificed mice were obtained using SILFLO resin (Cuderm, Dallas, TX, U.S.A.). Image analysis of the replicas was performed by the New Drug Development Center of Daegu-Gyeongbuk Medical Innovation Foundation (Daegu, Korea). The degree of wrinkling was evaluated using shadow analysis by Visioline VL650 (Courage + Khazaka, Köln, Germany), which contained a camera, stage and LED light source shining at a 35° incident light angle to the stage. The degree of wrinkles is expressed as the mean from a factor defined as the mean ratio of width to length of all shadows considered wrinkles. The value 0 indicates a perfect circle, and 1 indicates a perfect line. Therefore, the closer a value is to 1, the more effective it is at preventing wrinkles [20].

Tissue preparation for microscopy and histology
Mouse dorsal skin samples were fixed in 3.7% formaldehyde solution and embedded in paraffin blocks. Serial sections were obtained using a microtome and stained with several dyes, such as hematoxylin and eosin (H&E), Masson trichrome (MT) and toluidine blue (TB). Histological appearance was examined using an Olympus CX31/BX51 microscope (Olympus Optical Co., Tokyo, Japan) and TE-2000U camera (Nikon Instruments Inc., Melville, NY, USA). Epidermal thickness was evaluated by measuring the length from the stratum corneum to the stratum basale using a ruler equipped on the microscope and the LAS v4.8 program (Leica Microsystems, Herbrugg, Switzerland).

Analysis of isoflavones and other flavonoids
The dried extract was dissolved in DMSO at 5 mg/ml, and quantitative analysis of 5 major isoflavones in each extract was performed using an Agilent 1200 HPLC system (Agilent Technologies, Santa Clara, CA, USA) equipped with a G1315A diode array detector (DAD) at 254 nm. Chromatographic separation was achieved using a Luna C18 reverse-phase column (4.6 × 150 mm, 5 µm, Phenomenex, Torrance, CA, USA) and two solvent systems, A (100% water containing 0.05% formic acid) and B (100% acetonitrile containing 0.05% formic acid), at a 1 ml/min flow rate. A gradient was used starting from 10 to 100% B for 30 min. Quantification of 5 major isoflavones was performed based on the UV peak area of the daidzin standard combined with the molar extinction coefficients, and eight-point calibration curves were prepared for daidzin at concentrations from 0.01 to 10 µg/ml.

The dried extract was dissolved in 1 mg/ml in DMSO containing 2 µg/ml phloridzin as an internal standard, and quantitative analysis of two flavonoids in each extract was performed using a Q-Exactive benchtop hybrid quadrupole–orbitrap mass spectrometer (Thermo Fisher Scientific, Waltham, MA, USA) equipped with Vanquish ultrahigh-performance liquid chromatography (Thermo Fisher Scientific, Waltham, MA, USA). Five microliters of injected sample were separated using a Kinetex C18 reverse-phase column (2.1 × 100 mm, 2.6 µm, 100 Å, Phenomenex, Torrance, CA, USA) and two solvent systems, A (95% water and 5% acetonitrile; 0.1% formic acid) and B (5% water and 95% acetonitrile; 0.1% formic acid), at a flow rate of 0.35 ml/min. A linear gradient was used as follows: 3% B for 1 min, 3 to 100% B for 10 min, and 100% B for 2 min. The column was maintained at room temperature and was re-equilibrated for at least 5 min between analyses. The separated sample was ionized by supplying +3.2 kV high voltage to a heated-electrospray ionization (HESI) source with sheath gas 42 and aux gas 10 for ionization stabilization. The ionized sample was moved through a transfer capillary tube at 320 °C, and all moved ions were detected using the Top 10-depend-ent acquisition (DDA) mode. The detailed parameters of DDA were as follows: In full MS, data were acquired at a resolution of 70,000, and ions were transferred by an autogain control (AGC) target of 1e6, max injection time (max IT) of 100 ms and scan range of 150–2000 m/z. MS2 data were acquired under a resolution of 17,500, AGC target of 2e5, max IT of 50 ms, isolation window and normalized collision energies (NCEs) of 2.0 and 30 m/z. Procyanidin B2 and epicatechin were quantified

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based on the calibration curve of ten different concentrations from 0.01 to 1 µg/ml using the MS peak area ratio of each compound to the internal standard.

All peaks produced by HPLC were processed using ChemStation software (Rev. B.02.01-SR1, Agilent Technology, Santa Clara, CA, USA), and raw data produced by LC–MS/MS were processed by Qual Browser software provided by the Xcalibur package (Thermo Fischer Scientific, Waltham, MA, USA, ver.4.1.3). Standard chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA).

**Immunoblotting**

Whole supernatants from cells were precipitated with 10% trichloroacetic acid, and cell lysates were prepared in lysis buffer (Cell Signaling Technology, Danvers, MA, USA) containing a protease inhibitor (Roche, Penzberg, Germany) and a phosphatase inhibitor cocktail (Sigma-Aldrich). Then, the protein concentration of each sample was determined using the QuantiPro™ BCA assay kit (Sigma-Aldrich). Proteins in the supernatants and lysates were electrophoresed on 8% SDS-PAGE gels and transferred to polyvinylidene fluoride membranes (Millipore, Billerica, MA, USA). Then, membranes were probed with anti-MMP-1 (Calbiochem, MA, USA) primary antibody for 24 h at 4 °C and detected with corresponding horseradish peroxidase-conjugated secondary antibodies at room temperature for 1 h. Chemiluminescence was detected using SuperSignal® West Femto Maximum Sensitivity Substrate (Pierce, Rockford, IL, USA) and visualized using an iBright FL1500 (Invitrogen, CA, USA).

**Statistical analysis**

The data are expressed as the means ± SDs. Differences between the mean values in the two groups were analyzed using one-way analysis of variance (ANOVA) and Tukey’s multiple comparisons post hoc analysis. P < 0.05 was considered statistically significant.

**Results**

**SCEL-1 extract inhibits UV-induced expression of MMP-1 in a 3D human skin model**

MMP-1, also known as collagenase 1, is a major collagen digesting enzyme that causes collagen degradation in UV-exposed human skin [21]. To evaluate the inhibitory effects of SCEL-1, CJ-3 and WH extracts on collagenase, we measured expression levels of MMP-1 in a 3D human skin model. UVB irradiation resulted in a three-fold increase in levels of MMP-1 compared to the nonirradiated control group. However, levels of MMP-1 were significantly decreased in response to SCEL-1, CJ-3 and WH treatment in a dose-dependent manner compared to the UVB irradiation alone group. Furthermore, MMP-1 levels were decreased the most in response to application of SCEL-1 (Fig. 1a).

Type 1 collagen is a staple component of structural skin proteins and functions to maintain the integrity and tension of the dermis produced by fibroblasts [22]. Therefore, we examined the effects of SCEL-1, CJ-3 and WH on preventing collagen degradation in a 3D skin model. UVB irradiation reduced the amount of type I procollagen by 22% compared to the nonirradiated control group. However, levels of type I procollagen were significantly increased in response to SCEL-1, CJ-3 and WH treatment in a dose-dependent manner compared to the UVB-irradiated group, and once again, SCEL-1 was the most effective cultivar (Fig. 1b). These results indicate...
that SCEL-1 has the best efficacy among the three cultivars tested, and treatment with soybean extracts significantly reduces expression of MMP-1 induced by UVB, thus preventing the degradation of collagen.

**SCEL-1 extract prevents UV-induced wrinkle formation in hairless mice**

Next, we investigated whether SCEL-1, CJ-3, and WH can prevent wrinkle formation and skin changes in response to UV exposure in hairless mice. The 8-week schedule of UVB irradiation on the mouse was as follows (Fig. 2a). Hairless mice in the control group treated with vehicle alone did not show any significant changes in wrinkle formation over 8 weeks. However, repeated UV exposure on hairless mice increased deep and fine wrinkles on the back skin compared to the control group. In contrast, oral administration of the 3 black soybean extracts reduced wrinkle formation, and among them, the SCEL-1 extract exhibited the best anti-wrinkle effect (Fig. 2b). These results were further confirmed by image analysis using skin replicas. As a result of expressing the wrinkles as the mean form factor value, it was confirmed that the value decreased in the UV irradiated-group, but the value increased in the 3 black soybean extract-treated-groups, compared to the UV group. Consistent with previous observations, among the 3 black soybean treated groups, the value increased the most in the SCEL-1 treated group (Fig. 2c).

These results suggest that SCEL-1 treatment exhibits very strong anti-wrinkle efficacy in an animal model of wrinkle formation induced by UV irradiation.

**SCEL-1 extract inhibits UV-induced epidermal thickening, decreased dermal collagen and immune cell infiltration in hairless mice**

We next investigated the effect of the 3 black soybean extracts on epidermal thickness in a UVB-exposed mouse model with increasing doses of UV (1 to 4 MED) for 8 weeks. After the experiment was finished, skin samples were obtained from the dorsal skin of mice, and serially sectioned samples were stained with hematoxylin and eosin (H&E). Epidermal thickness was increased in the UV-irradiated group by 3.5-fold compared to the control group, but oral administration of SCEL-1 decreased the epidermal thickness to 16 and 34% in response to 25 and 100 mg/kg/day, respectively, in a dose-dependent manner compared to the UV-irradiated group. CJ-3 and WH treatment caused an approximately 20% decrease in epidermal thickness only in the 100 mg/kg/day administered group compared with the UVB-irradiated group, and their 25, 50 mg/kg/day application did not exert any significant effects on epidermal changes (Fig. 3a, d).

Collagen deficiency occurs from repeated exposure to UV rays, and the decrease in collagen content in the dermis has been thought to be the cause of wrinkles on photoaged skin. Therefore, we examined whether the 3 kinds of black soybean extracts prevented decreased collagen content induced by UV exposure in skin by Masson’s trichrome (MT) staining. As shown in MT-stained tissue, collagen fibers colored blue were reduced in the UV-irradiated group compared to the control group. However, collagen fibers were increased in the SCEL-1-, CJ-3-, or WH-treated groups compared to the UV-irradiated group, and the SCEL-1-treated group exhibited a higher collagen boosting effect than the other two groups (Fig. 3b, e).

UV exposure to skin increases the number of infiltrating mast cells in the dermis, the release of inflammatory substances from the skin. The blue dot in the dermis is toluidine blue-stained mast cells, and this blue spot increased in the UV-irradiated group compared to the control group, while the oral application of the SCEL-1, CJ-3 and WH extracts for 8 weeks decreased the infiltrated mast cell number compared to UV irradiation. Furthermore, SCEL-1 treatment showed better efficacy in reducing mast cell infiltration (Fig. 3c, f).

**Procyanidin B2 and epicatechin isolated from the SCEL-1 cultivar inhibit MMP-1 biosynthesis in UVB-irradiated human dermal fibroblasts**

Through animal experiments, we confirmed the beneficial effect of black soybean extract on wrinkle formation and unwanted skin changes. Therefore, we next determined which components in the SCEL-1 cultivar improved the status of UV-irradiated skin. It is well known that isoflavones and other flavonoids in soybeans have antioxidant and anti-inflammatory properties in vivo and vitro [1]. For the analysis of isoflavones, five major isoflavones, daidzin, 6'-O-malonyldaidzin, 6'-malonylgenistin, daidzein, and genistin, were quantified using LC-DAD at 254 nm based on the UV peak area of the daidzin standard combined with the molar extinction coefficients, and the quantitative results in three different black soybean cultivars are summarized in Table 1. The results showed that there was no significant difference in the five major isoflavones among the three different cultivars used in this study (Fig. 4a). However, a significant difference in the contents of two flavonoids, proanthocyanidin B2 and epicatechin, measured by LC–MS was clearly observed among the three different cultivars. SCEL-1 contained 17- to 30-fold higher levels of proanthocyanidin B2 and epicatechin than the other soybean cultivars, CJ-3 and WH (Fig. 4b, c). From the quantitative analysis of isoflavones and two major antioxidants, proanthocyanidin B2
Fig. 2 SCEL-1 extract prevents UVB-induced wrinkle formation in hairless mice. 

**a** The dorsal skin of hairless mice was irradiated with UVB three times per week for the first 4 weeks and two times per week for the subsequent 4 weeks. The UV dose was increased every 2 weeks by 1 MED (1 MED = 100 mJ/cm²) up to 4 MEDs. 

**b** Indicated soybean extracts (25, 50 or 100 mg/kg/day) were applied to the backs of hairless mice 2 h before UV irradiation. After 8 weeks of repeated UV treatment, UV-exposed dorsal areas (bottom panel) and skin replicas (top panel) were imaged. 

**c** To quantitatively assess skin wrinkling, a visiometer was used to analyze replica surfaces. Results are expressed as the mean ± SD (n = 7) for the parameter mean form factor. Statistical significance for the results is given as *p < 0.05 vs. control (CON) and #p < 0.05 vs. the UVB-irradiated group.
Fig. 3  SCEL-1 extract inhibits UV-induced epidermal thickening, dermal collagen deficiency and immune cell infiltration in hairless mice. Hairless mice were orally treated with the indicated concentrations (25, 50 or 100 mg/kg/day) of SCEL-1, CJ-3 and WH extracts as described in “Materials and methods”. After 8 weeks of repeated UV irradiation, animals were sacrificed, and dorsal areas were biopsied. Serial sections (4 μm) were mounted onto silane-coated slides and stained with H&E (a), MT (b) and TB (c). Epidermal thickness (d), collagen density (e) and mast cell number (f) are expressed as the mean ± SD (n = 7) compared to vehicle controls. Statistical significance for the results is given as *p < 0.05 vs. control (CON or C) and #p < 0.05 vs. the UVB-irradiated group.
and epicatechin, it could be hypothesized that two major antioxidants, proanthocyanidin B2 and epicatechin, are likely related to more significantly attenuated skin photoaging in response to the SCEL-1 cultivar.

To investigate the effects of two major compounds in SCEL-1 on collagenase, the major skin degradation enzyme, we measured MMP-1 protein levels in human dermal fibroblasts (HDFs). Results showed that proanthocyanidin B2 (Fig. 5a, c) or epicatechin (Fig. 5b, d) significantly inhibited MMP-1 biosynthesis in UVB-irradiated HDFs. Taken together, our data indicate that procyanidin B2 and epicatechin, which are specific components found in the SCEL-1 cultivar, decrease the expression of MMP-1, showing efficacy in attenuating wrinkles.

**Discussion**

Photoaging is a noticeable premature skin aging condition caused by prolonged and repeated exposure to UV light, which is linked to a series of physiological and pathological processes and is characterized by fine and coarse wrinkles, along with a combination of skin lags, tissue changes, roughness, dryness, and impaired skin barrier function [23–25].

Black soybeans exert positive effects on skin care. In this study, the effect of three black soybean cultivar extracts on wrinkle improvement was confirmed using in vivo and in vitro experiments. Among the 3 soybean extracts, SCEL-1 exerted the best anti-wrinkle effect, which was demonstrated by reducing expression of MMP-1 and inhibiting degradation of collagen in the 3D human skin model (Fig. 1a–d). In addition, the fact that the SCEL-1 extract was superior in wrinkle improvement compared to the other two cultivars was also confirmed in the skin of a hairless mouse model irradiated with UV (Fig. 2b, c). Considering the staining results of the skin tissue, it was confirmed that administration of SCEL-1 exhibited the best improvement in epidermal thickness, collagen density, and the number of infiltrating mast cells among extracts of the three cultivars (Fig. 3a–d).

Taken together, the above results show that the effects of SCEL-1 have the best wrinkle improvement among the three black soybean cultivars. Analysis of the three black soybean cultivars showed that they contained similar amounts of isoflavones but different amounts of procyanidin B2 and epicatechin (Table 1, Fig. 4a, b). These two ingredients were 17 to 30 times higher in SCEL-1 than in the other two cultivars, and the difference in the content of these two components was predicted to mediate the difference in wrinkle improvement efficacy. Therefore, we investigated the wrinkle improvement effect of these compounds, which was in accordance with MMP-1 production in a UV-irradiated HDF cells. The application of procyanidin B2 or epicatechin decreased the expression of MMP-1 in UV-exposed HDFs (Fig. 5a–d), indicating that it suppresses the formation of wrinkles by preventing the breakdown of collagen molecules. Collagen loss in the dermal extracellular matrix due to decreased synthesis or irregular degradation that changes the amount and quality of the skin constituents is a major cause of natural aging- and photoaging-driven wrinkles [26, 27]. Oxidative stress is another important factor that induces skin photoaging [28, 29]. ROS lead to cross-linking, inhibition of biosynthesis and damage to collagen fibers [30]. The two components identified as the primary components of SCEL-1, procyanidin B2 and epicatechin, were reported to have a cell protective effect against UVB by inhibiting oxidative stress and preventing collagen decomposition by reducing expression of MMP [31]. These reports support our results that SCEL-1 is effective at attenuating wrinkle photoaging.

In summary, we confirmed that SCEL-1 exhibited the best efficacy among the three cultivars tested, and treatment with this soybean extract significantly reduced the expression of matrix metalloproteinase-1 (MMP-1), preventing the degradation of collagen in a 3D human skin model. In addition, SCEL-1 application improved wrinkle- and photoaging-related symptoms, such as epidermal thickening, collagen deficiency and immune cell infiltration, in an animal model established by UV

| Cultivars | Contents (mg/g) |
|-----------|-----------------|
|          | Pro-anthocyanidins | Flavanols | Isoflavones |
|          | Procyanidin B2<sup>a</sup> | Epicatechin<sup>a</sup> | Daidzin<sup>b</sup> | 6-Malonyldaidzin<sup>b</sup> | 6-Malonylgenistin<sup>b</sup> | Daidzein<sup>b</sup> | Genistin<sup>b</sup> |
| SCEL-1   | 4.77 ± 0.46 | 3.98 ± 0.51 | 1.31 ± 0.078 | 6.06 ± 0.45 | 3.58 ± 0.18 | 0.4 ± 0.18 | 0.08 ± 0.11 |
| CJ-3     | 0.27 ± 0.083 | 0.239 ± 0.059 | 0.835 ± 0.15 | 4.31 ± 0.18 | 4.39 ± 0.18 | 0.23 ± 0.21 | 0.235 ± 0.21 |
| WH       | 0.286 ± 0.059 | 0.126 ± 0.061 | 0.935 ± 0.12 | 4.46 ± 0.28 | 4.01 ± 0.23 | 0.37 ± 0.18 | 0.31 ± 0.19 |

<sup>a</sup> Contents were analyzed by mass spectrophotometry analysis
<sup>b</sup> Contents were analyzed by high performance liquid chromatography
Fig. 4 Chromatograms of major isoflavones, procyanidin B2 and epicatechin and chemical structures. 

**a** Three kinds of soybean extracts were analyzed using high-performance liquid chromatography. Five total peaks were identified by references. The identified isoflavones were daidzin (1), 6″-O-malonyldaidzin (2), 6″-malonylgenistin (3), daidzein (4), and genistin (5).

**b** Three kinds of soybean extracts were analyzed using high-resolution mass spectrometry. Procyanidin B2 (m/z 449.12) and epicatechin (m/z 291.09) contained in SCEL-1, CJ-3 or WH are expressed by extracted chromatogram (XIC). The intensity of XIC was fixed at 4e6, and the XIC window was ±0.01 Da.

**c** Chemical structures of procyanidin B2 and epicatechin
irradiation. Procyanidin B2 and epicatechin isolated from the SCEL-1 cultivar inhibited MMP-1 biosynthesis in UVB-irradiated human dermal fibroblasts, and these two major components are likely related to more significantly attenuated skin photoaging. Therefore, our results showed that SCEL-1 exhibits stronger anti-wrinkle effects compared to the other two black soybean cultivars, suggesting that it represents an excellent anti-photoaging agent.

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Authors’ contributions
JWL, LP and SNK designed the study. JWL, JWL, HJ, NJP and SKB performed the experiments. JWL, YC and SNK drafted the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials
Not applicable.

Declarations
Competing interests
All authors declare no conflict of interest.

Author details
1Natural Products Research Institute, Korea Institute of Science and Technology (KIST), 679 Saimdang-ro, Gangneung, Gangwon-do 25451, Republic of Korea. 2Division of Bio-Medical Science and Technology, KIST School, University of Science and Technology, Seoul 02792, Republic of Korea.

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Fig. 5 Procyanidin B2 and epicatechin isolated from SCEL-1 inhibited UV-induced expression of MMP-1 in HDFs. HDFs were treated with procyanidin B2 (1, 3 or 10 μM) [a, c] or epicatechin (10, 30 or 100 μM) [b, d] in combination with UVB (20 mJ/cm²) for 48 h to measure MMP-1 expression. Relative MMP-1 protein expression was determined by western blot analysis. Statistical significance for the results is given as *p* < 0.05 vs. control (CON); #p < 0.05 vs. the UVB-irradiated group.
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