Culture-Expanded Autologous Adipose-Derived Mesenchymal Stem Cell Treatment for Osteonecrosis of the Femoral Head

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Background: Outcomes of traditional treatment for osteonecrosis of the femoral head (ONFH) are not always satisfactory. Hence, cell-supplementation therapy has been attempted to facilitate necrotic-tissue regeneration. Adipose-derived mesenchymal stem cell (ADMSC) transplantation is potentially advantageous over bone marrow-derived MSC implantation, but its outcomes for ONFH remain unclear. The aim of this study was to determine 2-year radiological and clinical outcomes of culture-expanded autologous ADMSC implantation for ONFH.

Methods: Eighteen hips with necrotic lesions involving ≥ 30% of the femoral head were included. ADMSCs were harvested by liposuction and culture expanded for 3 passages over 3 weeks. With a 6-mm single drilling, ADMSCs were implanted into the necrotic zone. All patients underwent magnetic resonance imaging (MRI), single-photon emission computed tomography/computed tomography (SPECT/CT) at screening and 6 months, 12 months, and 24 months postoperatively. The primary outcome was the change in the size of necrotic area on MRI. Secondary outcomes were changes in clinical scores and radioisotope uptake on SPECT/CT. Conversion total hip arthroplasty (THA) was defined as the endpoint.

Results: Preoperatively, the necrotic lesion extent was 63.0% (38.4%–96.7%) of the femoral head. The mean Harris hip score was 89.2, the University of California at Los Angeles (UCLA) score was 5.6, and Western Ontario and McMaster Universities Arthritis index (WOMAC) was 79.4. Three patients underwent THA and 1 patient died in an accident. Finally, 11 patients (14 hips) were available for ≥ 2-year follow-up. At the last follow-up, no surgery-related complications occurred, and 14 of 17 hips (82%) were able to perform daily activities without THA requirement. There was no significant decrease in lesion size between any 2 intervals on MRI. However, widening of high signal intensity bands on T2-weighted images inside the necrotic lesion was observed in 9 of 14 hips (64%); 11 of 14 hips (79%) showed increased vascularity on SPECT/CT at 2 years postoperatively. No significant differences were observed between preoperative and 24-month mean Harris hip score (89.2 vs. 88.6), WOMAC (79.4 vs. 75.7), and UCLA score (5.6 vs. 6.2).

Conclusions: Our outcomes suggest that culture-expanded ADMSC implantation is a viable option for ONFH treatment without adverse events.

Keywords: Osteonecrosis of the femoral head, Stem cell, Core decompression, Lesion size, Magnetic resonance imaging
Osteonecrosis of the femoral head (ONFH) is relatively common and leads to femoral head collapse and secondary hip osteoarthritis. Although ONFH's natural course varies with the location or size of the necrotic lesion, symptom development and disease progression are likely if > 30% of the femoral head is affected. If left untreated, it may require total hip arthroplasty (THA). Considering that ONFH occurs frequently in younger patients (age, 30–50 years), prevention or delay of femoral head collapse is the primary goal of surgical intervention.

Femoral head core decompression or multiple drilling had been widely used for ONFH treatment in the past. These procedures aim to preserve the femoral head and facilitate necrotic tissue regeneration by revascularization and provision of channels for osteogenic cell recruitment from the proximal femur. However, the reported success rate of isolated core decompression is 30%–86%, and the results are unsatisfactory for larger lesions.

One of the reasons for the unsatisfactory outcome of isolated core decompression might be that osteoprogenitor cell numbers in the femoral head and proximal femur are insufficient for complete ONFH reconstruction or repair. Thus, many attempts have been made to facilitate necrotic lesion repair by stem cell implantation, as several authors have reported favorable outcomes of autologous bone marrow grafting for ONFH.

Adipose-derived mesenchymal stem cell (ADMSC) implantation has recently become a promising method for stem cell therapy because these cells have multi-mesenchymal lineage potential and can differentiate into osteoprogenitor cells under appropriate conditions. Moreover, ADMSC harvesting is relatively simple and cell lines can be easily expanded for sufficient cell numbers. Thus, ADMSC implantation is promising for ONFH treatment. However, few studies have reported the efficacy of ADMSC implantation for ONFH treatment.

Therefore, the aim of the current study was to determine the 2-year clinical and radiological outcomes of ADMSC implantation for ONFH treatment. The authors hypothesized that ADMSC implantation would (1) decrease the necrotic lesion size on magnetic resonance imaging (MRI), (2) improve clinical scores, and (3) increase necrotic lesion radioisotope uptake on single-photon emission computed tomography/computed tomography (SPECT/CT).

### METHODS

#### Study Design and Subjects

The phase I/IIa trial protocol was approved by the Korean Food and Drug Administration (KFDA). The study protocol was approved by the Institutional Review Board of Seoul National University Boramae Medical Center Institutional Review Board (IRB No. 06-2010-128). Written informed consent was obtained from all participants before enrollment in the study. This study is registered with the International Clinical Trials Registry (NCT01643655).

The phase I cohort included 3 patients receiving injections of $1 \times 10^8$ autologous ADMSCs in 3-mL saline. Adverse event occurrence was evaluated for 28 days after the injection. Adverse events were defined according to the National Cancer Institute common terminology criteria for adverse events (NCI-CTCAE) grade 3 (dose-limiting toxicity) as follows: (1) allergic reaction/hypersensitivity, (2) injection-site reaction, (3) injection-site infection, (4) fibrosis-cosmesis, and (5) muscular/skeletal hypoplasia. A safety review conducted on day 28 found no incidence of the abovementioned adverse events. Thus, the phase IIa study with 12 patients was commenced; each patient received phase I dosages. Thus, 15 patients (18 hips) were enrolled from July 2012 to January 2013.

Patients met the following eligibility criteria: (1) age 18–70 years; (2) nontraumatic ONFH; (3) hips without femoral head collapse or ≤ 2 mm femoral head depression (Steinberg stage I, II, and IIIA); and (4) necrotic lesion size ≥ 30% of the femoral head area. Necrotic lesion size was measured on MRI as described previously. Exclusion criteria were (1) femoral head collapse ≥ 2 mm; (2) previous history of core decompression/multiple drilling; (3) history of bisphosphonate or parathyroid hormone therapy; (4) current consumption of immunosuppressants, cytotoxic agents, or corticosteroids; (5) inability to undergo MRI examination; (6) pregnancy or breastfeeding; (7) active infectious disease including human immunodeficiency virus infection; (8) serious comorbidities that may affect the treatment process; (9) history of participation in another clinical trial ≤ 3 months; and (10) refusal to participate.

After providing informed consent, all patients underwent screening, including physical examination, blood and urine testing, chest X-ray, and electrocardiography. Anteroposterior and frog-leg hip X-rays, MRI, and SPECT/CT were also conducted at the initial visit. Liposuction was performed 1 week afterward. When patients were admitted for ADMSC implantation, baseline Harris hip score (HHS), Western Ontario and McMaster Universities Arthritis index (WOMAC), and University of California at Los Angeles (UCLA) scores were obtained. Hip X-ray, MRI, and SPECT/CT were performed preoperatively and at 6 months, 12 months, and 24 months postoperatively. HHS, WOMAC index, and UCLA score were also obtained at each postoperative visit.
Mesenchymal Stem Cell Preparation

Abdominal fat tissue samples were harvested by liposuction under local anesthesia for ADMSC isolation 1 week after the patient screening. ADMSC preparation was conducted under standard conditions as previously described. Harvested fat tissues were enzymatically digested and stromal vascular fraction cells were isolated and cultured in keratinocyte serum-free medium (Invitrogen Corp., Carlsbad, CA, USA) containing 0.2-mM ascorbic acid, 0.09-mM calcium, 5-ng/mL recombinant epidermal growth factor, and 5% fetal bovine serum. The cells were passaged until reaching 90% confluence and then culture-expanded for 3 passages over 3 weeks. Afterward, the cells were tested for cell number, viability, purity (CD31, CD34, CD45), identity (CD73, CD90), sterility, endotoxins, and mycoplasma. Finally, $1 \times 10^8$ ADMSCs in 3-mL saline were prepared in a syringe (Jointstem; R Bio, Seoul, Korea; http://www.rbio.co.kr/).

Surgical Technique and Rehabilitation

Under spinal anesthesia, the patient was placed supine on a fracture table. A longitudinal 5-cm skin incision was made just below the greater trochanter. Under fluoroscopic visualization, a guide pin was directed to the necrotic lesion center, which had been preoperatively identified on MRI. A 6-mm reamer was introduced along the guide pin, and reaming was performed just under the subchondral bone of the lesion (Fig. 1). Before ADMSC injection, the fracture table was tilted so that the ipsilateral greater trochanter faces upwards to prevent backing out of the injected cells. The syringe filled with ADMSCs was then connected to an 18-G spinal needle and $1 \times 10^8$ ADMSCs in 3-mL saline were injected under fluoroscopic guidance.

After injection, the lateral cortex reamed channel opening was sealed with a collagen bone plug (TERUPLUG; Olympus Terumo Biomaterials Co., Tokyo, Japan). Ipsilateral, partial, crutch/cane-assisted weight-bearing was allowed for the first 6 weeks postoperatively. Afterward, total weight-bearing was gradually permitted as tolerated.

Outcome Measures

The primary outcome was the change in the necrotic lesion size on MRI as previously reported. Measurements were taken by 2 independent, blinded radiologists. Secondary outcomes were changes in the clinical scores and SPECT/CT radioisotope uptake. Clinical assessments were performed using HHS. Patient activity level was assessed using WOMAC index and UCLA score at the last follow-up. In the original WOMAC scoring system, a higher score indicates a poorer status. For consistency with other scoring tools, the WOMAC index was reversed so that a higher score indicates better functional status. Radioisotope uptake degree on SPECT/CT was evaluated qualitatively by an independent nuclear medicine expert and increased uptake was determined in a dichotomized fashion. All adverse events were recorded according to the NCI-CTCAE criteria version 4.0. Conversion to THA was defined as the endpoint and no further evaluation was made if the patient underwent THA.

Statistical Analysis

The change in the necrotic lesion size was determined by one-way analysis of variance with the Bonferroni post-hoc test. HHS, WOMAC index, and UCLA scores were compared preoperatively and 24 months postoperatively using the paired t-test. Statistical analyses were conducted using IBM SPSS ver. 20.0 (IBM Corp., Armonk, NY, USA). A p-value < 0.05 was considered statistically significant.

RESULTS

Demographic and Preoperative Characteristics of Patients

From July 2012 to January 2013, 15 patients (13 men and 2 women; 18 hips) underwent ADMSC implantation and were followed up for 24 months. The mean age and body mass index were 43.8 years (range, 20–59 years) and 25.5 ± 3.3 kg/m$^2$, respectively. Seven patients had a history of heavy alcohol consumption, 2 received steroid therapy, 1 had caisson disease, and 6 were diagnosed with idiopathic ONFH. Fourteen hips were classified as Steinberg stage IIC and 4 as stage IIIA. Six hips were associated with pain, whereas 12 were asymptomatic and contralateral hip ONFH was incidentally found on diagnostic imaging.
Six hips showed subchondral fractures with bone marrow edema patterns outside the necrotic area on preoperative MRI. The mean necrosis extent of the 18 lesions was 63.0% (38.4%–96.7%) of the total femoral head area. The mean HHS was 89.2/100, the mean UCLA score was 5.6/10, and the mean WOMAC index was 79/96. During the follow-up period, 3 hips (18%) in 3 patients underwent THA at 13–16 months after the initial surgery because of femoral head collapse and pain aggravation, which disrupted daily activities. At 12 months after initial surgery, the mean HHS, UCLA score, and WOMAC index of 3 hips were 75/100, 5/10, and 61/96, respectively. One patient with unilateral ONFH died in an unrelated accident during follow-up. Therefore, 24-month outcome data were available for 11 patients (14 hips).

Outcomes
At the last follow-up, there were no surgery-related complications. MRI showed no significant lesion size changes between any 2 intervals (Table 1, Fig. 2). However, 1 patient (case 6 and 7) showed definite lesion size decrease by 9.3% (case 6) and 12.2% (case 7) on MRI at 24 months after surgery. His plain radiographs also showed healed subchondral fracture and increased radiodensity of the femoral head (Fig. 3). Interestingly, widening of high signal intensity band on T2-weighted images inside the necrotic lesion was observed in 9/14 hips (64%), which might indicate increased vascularity and tissue regeneration (Fig. 4). This finding was not observed until the last follow-up in 3 hips after THA (Fig. 5). There were no significant differences between preoperative and 24-month postoperative HHS, WOMAC index, and UCLA score in 11 patients (14 hips) (Table 2, Fig. 2). SPECT/CT at 24

Table 1. Change in the Size of Necrotic Lesion over 2-Year Follow-up

| Case | Sex | Age (yr) | Stage* | Preoperative | 6 mo | 12 mo | 24 mo | Change † | p-value ‡ |
|------|-----|----------|--------|--------------|------|-------|-------|----------|-----------|
| 1    | Male | 44       | II     | 67.0         | 64.2 | 64.2  | 64.2  | –4.2     | 0.974     |
| 2    | Male | 54       | II     | 51.5         | 50.9 | 50.7  |       |          |           |
| 3    | Male | 57       | II     | 46.5         | 45.9 | 45.9  | 45.9  | –1.3     |           |
| 4    | Male | 57       | II     | 49.9         | 49.7 | 49.3  | 49.3  | –1.2     |           |
| 5    | Female | 50     | II     | 84.3         | 84.3 | 83.9  |       |          |           |
| 6    | Male | 20       | III    | 81.6         | 76.4 | 74.0  | 74.0  | –9.3     |           |
| 7    | Male | 20       | II     | 66.3         | 64.7 | 58.2  | 58.2  | –12.2    |           |
| 8    | Female | 40    | II     | 76.8         | 74.5 | 73.7  | 74.5  | –3.0     |           |
| 9    | Female | 40    | III    | 96.7         | 95.6 | 94.6  | 91.0  | –5.9     |           |
| 10   | Male | 50       | II     | 53.0         | 53.4 | 53.5  | 53.5  | 0.9      |           |
| 11   | Male | 59       | II     | 41.8         | 41.9 | 41.9  |       |          |           |
| 12   | Male | 55       | II     | 50.9         | 50.0 | 50.0  | 50.0  | –1.8     |           |
| 13   | Male | 40       | II     | 60.7         | 60.7 | 60.7  | 60.0  | –1.2     |           |
| 14   | Male | 30       | II     | 66.9         | 70.9 | 71.0  | 69.6  | 4.0      |           |
| 15   | Male | 40       | II     | 53.8         | 53.3 | 53.3  | 52.0  | –3.3     |           |
| 16   | Male | 58       | II     | 76.8         | 73.1 | 73.8  |       |          |           |
| 17   | Male | 41       | III    | 70.9         | 69.7 | 69.7  | 68.5  | –3.5     |           |
| 18   | Male | 34       | III    | 38.4         | 37.7 | 37.4  | 36.3  | –5.5     |           |
| Mean |     | 63.0     | 62.0   | 61.4         | 60.5 | –3.4  | 0.974 |           |           |

Values are presented as percentages.
*Preoperative Association Research Circulation Osseous (ARCO) stage.
†Change in lesion size was calculated as: 100 × (size at 24 months – preoperative size)/preoperative size. The p-value was calculated by repeated measures analysis of variance.
months showed increased radioisotope uptake around the necrotic margin in 79% hips (11/14) (Fig. 6). Radiographs and MRI evaluations of the 3 hips (18%) indicated symptomatic subchondral fracture with femoral head collapse < 2 mm at 24 months of follow-up.
DISCUSSION

For ONFH treatment, several methods have been attempted for femoral head preservation before collapse. Core decompression and multiple drilling are attractive, relatively easy surgical options with acceptable outcomes. However, several cases result in failure, especially those with large necrotic lesions. There has been growing interest in cell supplementation along with core decompression. Since the first report by Hernigou and Beaujean, several studies have reported the outcomes of concentrated bone marrow aspirate or bone marrow-derived mesenchymal stem cell (BMMSC) implantation for ONFH treatment. However, only 1 study reported the safety and efficacy of culture-expanded BMMSCs for ONFH treatment. To the best of our knowledge, the current study is the first to evaluate the safety and outcome of culture-expanded autologous AD-MSC implantation for ONFH treatment in humans.

Our results indicated that femoral head AD-MSC implantation is safe without serious complications. As this was the first study to utilize culture-expanded AD-MSCs for ONFH treatment, no direct comparisons with other studies were possible. Most previous studies utilized BMMSCs for ONFH treatment. BMMSCs have both

| Outcome score | Preoperative | 6 mo | 12 mo | 24 mo | p-value* |
|---------------|--------------|------|-------|-------|----------|
| Harris hip score | 89.2 ± 8.2 | 88.9 ± 8.7 | 88.0 ± 12.5 | 88.6 ± 11.8 | 0.976 |
| WOMAC score | 79.4 ± 14.7 | 79.9 ± 13.3 | 77.6 ± 15.0 | 75.7 ± 15.5 | 0.810 |
| UCLA score | 5.6 ± 2.4 | 4.1 ± 1.3 | 5.5 ± 1.8 | 6.2 ± 14.2 | 0.123 |

*Values are presented as mean ± standard deviation.

**Table 2. Changes in the Clinical Scores over 2-Year Follow-up**

Fig. 4. Coronal T2 magnetic resonance imaging scans showing widening of the high signal intensity band inside the necrotic lesion (arrows). (A) Preoperative. (B) Six months after surgery. (C) Twelve months after surgery. (D) Twenty-four months after surgery.

Fig. 5. Coronal T2 magnetic resonance imaging (MRI) scans (case 5) showing no definite widening of the high signal intensity band inside the necrotic lesion during follow-up. (A) Preoperative. (B) Six months after surgery. (C) Twelve months after surgery. The MRI shows a subchondral fracture (arrow) and bone marrow edema (asterisk).
angiogenic and osteogenic potential, and long-lasting reconstruction repair can be expected with BMMSC implantation. However, several limitations associated with its use for ONFH treatment exist. First, an invasive procedure under general or spinal anesthesia is required to obtain bone-marrow cells and available quantities are limited. Second, bone marrow mesenchymal stem cell isolation yield decreases with age and the potential of stem cells for differentiation and proliferation also decrease with patient age. On the other hand, large quantities of ADMSCs can be easily obtained using a less invasive procedure, and cell numbers obtainable from a given amount of adipose tissue do not decrease with age. Some studies reported that ADMSCs can induce paracrine and autocrine responses, which enhance osteogenic potential. The cells transduced by these mechanisms excrete bone morphogenetic protein, which may further enhance their osteogenic activity. A recent study reported that the proliferation capacity of ADMSCs is 4 times that of BMMSCs; ADMSCs also show better phenotypical characteristics, which indicates better bone differentiation potential. Thus, ADMSCs, instead of BMMSCs, are a more promising option for ONFH treatment. In this context, future studies are warranted to further reveal ADMSCs’ role for ONFH treatment despite our limited success.

Although lesion sizes remained almost the same 2 years postoperatively, 64% of the hips (11/17) survived without symptomatic femoral head collapse, which would require THA. Including 3 hips with subchondral fracture and femoral head collapse < 2 mm without significant associated symptoms, 82% hips did not require THA at the 2-year follow-up. This finding is especially important because only patients with > 30% femoral head lesion size were included. The mean preoperative lesion size was 63.0% in this population and the smallest lesion size was 38.4%. The decision to include only patients with large lesions was according to the findings of a previous study. At a 5-year follow-up of the natural course of ONFH, only 1 of 21 patients developed pain when the lesion size was < 30%, whereas 11 of 24 patients with a lesion size 30%-50% and 50 of 60 patients with a lesion size > 50% developed pain. Necrotic lesion size is one of the most important factors in determining the prognosis of ONFH. Hungerford and Jones reported that small lesions (< 15% of the femoral head) were unlikely to progress and large lesions (> 30% of the femoral head) could not be successfully treated with core decompression. Indeed, many previous studies reported high failure rates of core decompression for large lesions (Table 3). Although the present study did not include a control group, the 82% survival rate among hips with large necrotic lesions implies some contribution of implanted ADMSCs, in addition to 6-mm drilling, to prevention of disease progression.

One of the reasons this study failed to show lesion size decrease may be the large initial lesion size in the study population. Although no firm conclusions can be drawn about the reasons for no significant lesion size decrease, we believe that the implanted cells failed to migrate throughout the entire necrotic region for effective repair with only a single drilling. We chose a 6-mm single drilling method for ADMSC implantation to minimize core decompression or multiple drilling effect because KFDA had only approved it in order to prove the effect of ADMSC. Increased radioisotope uptake might indicate increased vascular activity, but more effective methods to deliver the implanted cells to the entire necrotic tissue are needed. Previous studies reported favorable outcomes with bone marrow or BMMSC implantation, but they included only patients with relatively small lesions or did not report...
Therefore, direct comparisons between these studies and the present study are difficult. A previous study of bone marrow grafting reported that cases with lesions > 25% of the femoral head had significantly worse outcomes. In a study of a rabbit ONFH model, the group treated with adipose-derived stem cells showed higher bone trabecular volume on micro-CT, which suggests necrotic bone regeneration in the lesion. The outcomes of this study suggest that ADMSCs can also facilitate regeneration in human ONFH. However, further studies of ADMSC implantation in patients with smaller lesions are needed to clarify this issue.

In the current study, no significant changes in any of the clinical scores were observed before and 24 months postoperatively. This is probably because many hips were asymptomatic preoperatively. Pain was only noted in 6 of 18 hips, and 12 were incidentally diagnosed. The presence or absence of pain was not considered as inclusion or exclusion criteria in the current study, because it was unethical to exclude patients with large necrotic lesions regardless of pain as hips with large necrotic lesions are likely to collapse. Preoperative clinical scores of asymptomatic hips were already high; therefore, no room for further improvement existed because of a ceiling effect. There was some improvement in the scores of hips that were initially painful, but the difference was not statistically significant because of the small sample size. Further studies with larger sample sizes would enable subgroup analysis of hips with initial pain.

Our study has some limitations. First, there was no control group, which limits interpretation of outcomes. Hence, future studies with control groups are required to further elucidate the efficacy of ADMSC implantation. Second, the sample size was too small to conduct robust statistical analysis, which was inevitable considering that this was a phase I/IIa trial and only a small number of patients were enrolled. Because there were no procedure-related serious adverse events in this study, future studies with larger sample sizes are encouraged. Third, the fate of implanted ADMSCs was not examined. Although there was increased radioisotope uptake on SPECT/CT in many cases, it is unclear whether this change was induced by the implanted ADMSCs. In our cohort, the femoral head was drilled with a 6-mm reamer, which is smaller than that used for standard core decompression to minimize adverse effects. Nonetheless, it was not possible to verify whether the implanted cells functioned as expected.

At the last follow-up, there was progression of disease in 6 out of 17 hips (36%), including 3 hips with symptomatic subchondral fractures and femoral head collapse < 2 mm and 3 hips with THA. Our data may be insufficient to guarantee the successful treatment of ONFH; however, they can suggest that culture-expanded ADMSC implantation has a favorable safety profile with a low risk of systemic or surgical adverse events. Further comparative studies with larger sample sizes and longer follow-up are needed to establish the efficacy and benefit of culture-expanded ADMSC implantation for ONFH treatment over other joint preserving procedures.

**CONFLICT OF INTEREST**

One of the institutions (Seoul National University Borame Hospital, Seoul, Korea) received funding for this study from R Bio Co., Ltd., Seoul, Korea. No other potential conflict of interest relevant to this article was reported.
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