Serum progranulin as an independent marker of liver fibrosis in patients with biopsy-proven nonalcoholic fatty liver disease

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Abstract. Background: Elevated progranulin levels are associated with visceral obesity, elevated plasma glucose, and dyslipidemia. Progranulin has not been previously investigated as a biomarker of nonalcoholic fatty liver disease (NAFLD). We sought to determine whether serum progranulin levels are altered in patients with biopsy-proven NAFLD and if they are associated with their clinical, biochemical, and histological characteristics.

Subjects and methods: We measured serum progranulin levels in 95 patients with biopsy-proven NAFLD and 80 age- and sex-matched controls. The potential associations between progranulin and the characteristics of NAFLD patients were examined by multiple linear regression analysis.

Results: Serum progranulin levels were significantly higher in NAFLD patients (34 ± 13 ng/mL) than in controls (28 ± 7 ng/mL, \(P < 0.001\)). In NAFLD patients, serum progranulin levels were associated with lipid levels and the degree of hepatic fibrosis. After adjustment for potential confounders, serum progranulin remained an independent predictor of the degree of hepatic fibrosis in NAFLD patients (\(\beta = 0.392; t = 2.226, P < 0.01\)).

Conclusions: Compared with controls, NAFLD patients have higher serum progranulin concentrations, which are closely associated with lipid values and the extent of hepatic fibrosis.

Keywords: Progranulin, nonalcoholic fatty liver disease, enzyme-linked immunosorbent assay, hepatic fibrosis

1. Introduction

Nonalcoholic fatty liver disease (NAFLD) is the most common form of metabolic liver disease in Western countries. It comprises a disease spectrum ranging from simple steatosis to steatohepatitis (NASH) [1]. NAFLD is now recognized as the hepatic manifestation of metabolic syndrome [2], and its prevalence has been linked to a number of different metabolic abnormalities (e.g., obesity, type 2 diabetes, hyperlipidemia, and hypertension) [3]. Due to the global epidemic of obesity and the potential progression to advanced liver disease, NAFLD has become an important emerging public health issue [4].

Progranulin is a widely expressed 593-amino acid glycoprotein encoded by a single gene on chromosome 17q21. Evidence suggests that progranulin plays a key role in a range of biological processes, including tumorigenesis, wound repair, neovascularization, inflammation, cell migration and mitosis, and cell survival [5, 6]. Interestingly, progranulin is not expressed in normal hepatocytes but its expression has been found to be increased in a mouse model of early diet-induced...
NASH [7]. In addition, the expression of progranulin has been identified in cultured primary and immortalized hepatic stellate cells [8]. Moreover, this molecule has been linked with the fibrosis response which follows tissue injury [9]. In a recent clinical study, Youn et al. [10] have reported that serum progranulin concentrations are significantly increased in individuals with type 2 diabetes and in obese subjects with predominant visceral fat accumulation. It is also noteworthy that progranulin levels correlate significantly with BMI, C-reactive protein levels, and total cholesterol [10]. Because the relationship between progranulin and NAFLD in human clinical studies remains unclear, we sought to examine the nature and the strength of the association between serum progranulin concentrations and the severity of liver histology in patients with biopsy-proven NAFLD.

2. Materials and methods

2.1. Study participants

A total of 95 patients with NAFLD (50 males and 45 females, mean age, 47 ± 8 years) and 80 healthy subjects (41 males and 39 females, mean age, 47 ± 9 years) were enrolled in this study. The two groups were comparable for age and sex distribution. All NAFLD patients were consecutively seen at the hospital-based specialized outpatient clinics of the Department of Gastroenterology, Marmara University School of Medicine, over the past 12 months. All patients described here were enrolled in a larger research project aiming at identifying novel biomarkers of NAFLD using a “knowledge-based” approach [11–13]. The “knowledge-based” approach relies on a direct understanding of the pathophysiological processes that underlie the development of NAFLD. It consists of biochemical assays aiming to assess attractive novel candidate markers informed by the biology of the disease process [14]. Inclusion criteria for this study comprised: (a) ultrasonographic evidence of steatosis grade 1 or higher; (b) absent-to-low alcohol consumption, i.e. < 20 g/day; (c) evidence of NAFLD on liver biopsy; (d) exclusion of viral B and C hepatitis, Wilson’s disease, α1-antitrypsin deficiency, autoimmune hepatitis, genetic hemochromatosis, and use of steatogenic drugs. Patients were excluded when they had major comorbidities, i.e. ischemic heart disease, cerebrovascular diseases, autoimmune disorders, malignancies, and chronic renal insufficiency. A total of 80 healthy age- and sex-matched volunteers served as controls. All subjects included in the control group were judged to be in good health, with normal results on liver function tests, and confirmed as having normal liver by ultrasound. Subjects with a consumption of alcohol > 20 g/day or who were taking any medication were not included in the control group. All patients and controls were of Turkish descent. The Ethics Committee of the Marmara University School of Medicine approved this study and all participants provided written informed consent.

2.2. Clinical and biochemical characterization

All subjects underwent a detailed physical examination, anthropometric measurements, and biochemical screening. Body mass index (BMI) was calculated from measurements of height and weight. Diabetes mellitus was diagnosed according to ADA criteria [15]. The metabolic syndrome was diagnosed using the ATP III criteria [16]. The estimate of insulin resistance was calculated using the HOMA-IR index, with the following formula: insulin resistance = fasting plasma insulin (in microunits per milliliter) × fasting plasma glucose (FPG) (in millimoles per liter)/22.5. Blood pressure was measured using a mercury sphygmomanometer in a quiet room after ≥ 10-min rest. Korotkoff 1 and 5 were taken for systolic blood pressure and diastolic blood pressure, respectively. Routine blood chemistry analyses were performed at the central laboratory of our hospital. Serum high-sensitivity C-reactive protein (hs-CRP) levels were measured in duplicate using a commercially available method (Dade Behring, Marburg, Germany). The intra-assay and the inter-assay coefficients of variation for hs-CRP were 4.6% and 6.1%, respectively. The lower detection limit was 0.19 mg/dL.

2.3. Liver histology

Ultrasonography-guided liver biopsies were performed under conscious sedation using a 16-gauge Hepafix needle (Braun Melsungen AG, Germany). All biopsy specimens were placed in formalin solution for fixation and embedded in paraffin blocks. Serial sections (sectioned at 4 mm intervals) were stained with hematoxylin–eosin, Masson’s trichrome. An experienced pathologist blinded to clinical data scored the liver biopsies according to the NIDDK NASH Clinical Research Network scoring system [17]. Steatosis was scored from 0 to 3 with a four grades scoring system.
from S0 to S3: S0: no steatosis or less than 5%, S1: 5–33%, S2: 33–66%, S3: >66%. Lobular inflammation was graded as follows: stage 0, no foci; stage 1: <2 foci per 200 × field; stage 2: 2–4 foci per 200 × field; stage 3: >4 foci per 200 × field. Ballooning degeneration of liver cells was evaluated as: grade 0, absent; grade 1, few cells; grade 2, many cells. The histological NASH score was defined as the unweighted sum of the scores for steatosis (0–3), lobular inflammation (0–3), and ballooning (0–2); thus ranging from 0 to 8. Cases with scores of 0–2 were considered as having simple steatosis; on the other hand, cases with scores of 5 or greater were diagnosed as definitive NASH. Cases with activity scores of 3 and 4 were considered as borderline NASH [17]. Fibrosis was staged as follows: stage 0, no fibrosis; stage 1: perisinusoidal or periportal fibrosis with 3 different patterns: 1A: mild, zone 3, perisinusoidal; 1B: moderate, zone 3, perisinusoidal fibrosis, and 1C portal/periportal fibrosis; stage 2: perisinusoidal and portal/periportal fibrosis; stage 3: bridging fibrosis; stage 4: cirrhosis.

2.4. Measurement of serum progranulin levels

Blood samples were collected from an antecubital vein between 8:00 and 9:00 AM after an overnight fast. Samples were centrifuged at 2500 g for 10 min, and serum aliquots were stored at −80°C until immediately before analysis. Serum progranulin levels were measured using a commercially available enzyme-linked immunosorbent assay (ELISA) (Progranulin ELISA kit E103, Mediagnost, Reutlingen, Germany) according to the manufacturer’s protocol. Absorbance at 450 nm was read by using a microtiter plate reader. The detection limit of the assay was 0.018 ng/mL. The intra- and inter-assay coefficients of variance were 4.2% and 5.8%, respectively.

2.5. Data analysis

The Kolmogorov-Smirnov test was performed in all continuous variables to define the presence of normality. Gaussian variables are expressed as mean ± standard deviation (SD), skewed data as medians and interquartile ranges, and categorical variables as counts. Skewed variables were logarithmically transformed to improve normality before statistical analysis and then back-transformed to their natural units for presentation in the text and tables. The Student’s t-test was used to evaluate the differences between the two study groups in normally distributed continuous variables. When normality was not confirmed, the Mann–Whitney U test was used. Correlations among variables were tested using the Spearman’s correlation coefficient. Multivariable stepwise linear regression analyses were performed to identify independent predictors of the histological features of NAFLD (i.e., steatosis, lobular inflammation, hepatocyte ballooning, liver fibrosis); the covariates included were progranulin and all variables listed in Table 1. We determined the sensitivity and specificity of serum progranulin concentration in predicting NAFLD at different cutoff values by the construction of a ROC curve (sensitivity plotted against 1-specificity at different concentrations of progranulin). All calculations were performed using SPSS version 14.0 (SPSS, Inc., Chicago, IL, USA) and MedCalc 7.2 (Mariakerke, Belgium). A value of \( P < 0.05 \) (two-sided) was considered statistically significant.

3. Results

Table 1 shows the general characteristics of subjects with and without NAFLD. The two study groups did not differ in terms of age, sex, and HDL cholesterol. Body mass index, HOMA-IR, AST, ALT, systolic and diastolic blood pressure, total cholesterol, LDL cholesterol, and tryglicerides of NAFLD patients differed significantly from the control group. The prevalence of diabetes and the metabolic syndrome was higher in NAFLD patients than in controls. Table 2 summarizes the severity of histological features of NAFLD patients. Serum progranulin levels were significantly higher in NAFLD patients (34 ± 13 ng/mL) than in controls (28 ± 7 ng/mL, Student’s t test, \( P < 0.001 \), Fig. 1). The ROC curve analysis (Fig. 2) suggested that the most useful cutoff value of serum progranulin concentration was 30 ng/mL, where the sum of sensitivity (59%) and specificity (70%) was the highest.

We then analyzed the associations between serum progranulin levels and the clinical, biochemical, and histological characteristics of the study participants. In
Table 1
General characteristics of the study participants

|                                | NAFLD patients \((n = 95)\) | Control group \((n = 80)\) | \(P\) value |
|--------------------------------|-----------------------------|-----------------------------|-------------|
| Sex (males/females)            | 50/45                       | 41/39                       | NS          |
| Age (years)                    | 47 ± 8                      | 47 ± 9                      | NS          |
| Body mass index (kg/m\(^2\))   | 30.4 ± 4.4                  | 27.6 ± 4.1                  | < 0.01      |
| Diabetes mellitus (yes/no)     | 30/65                       | 0/80                        | < 0.001     |
| Metabolic syndrome (yes/no)    | 61/34                       | 0/80                        | < 0.001     |
| HOMA-IR                        | 3.8 (2.4–5.2)               | 1.5 (0.6–2.4)               | < 0.001     |
| Systolic blood pressure (mmHg) | 134 ± 23                    | 122 ± 18                    | < 0.01      |
| Diastolic blood pressure (mmHg)| 84 ± 12                     | 77 ± 10                     | < 0.01      |
| AST (U/L)                      | 46 ± 16                     | 23 ± 9                      | < 0.001     |
| ALT (U/L)                      | 69 ± 34                     | 20 ± 9                      | < 0.001     |
| Total cholesterol (mg/dL)      | 223 ± 52                    | 192 ± 45                    | < 0.01      |
| HDL cholesterol (mg/dL)        | 44 ± 12                     | 43 ± 17                     | NS          |
| LDL cholesterol (mg/dL)        | 151 ± 55                    | 124 ± 19                    | < 0.001     |
| Triglycerides (mg/dL)          | 181 ± 83                    | 143 ± 69                    | < 0.001     |
| hs-CRP (mg/dL)                 | 3.2 (2.6–4.4)               | —                           | —           |

HOMA-IR: homeostasis model of insulin resistance; AST: aspartate aminotransferase; ALT: alanine aminotransferase; HDL: high-density lipoprotein; LDL: low-density lipoprotein; hs-CRP: high-sensitivity C-reactive protein; NASH: nonalcoholic steatohepatitis. Data are given as means and SD, counts, or medians and interquartile ranges, as appropriate.

Patients with NAFLD, serum progranulin levels were positively associated with total cholesterol \((r = 0.38, P < 0.001)\), LDL cholesterol \((r = 0.32, P < 0.001)\), and the degree of hepatocyte fibrosis \((r = 0.22, P < 0.01)\). A serum progranulin level > 32 ng/ml had a positive predictive value for a fibrosis score \(\geq 2\) of 93% and a negative predictive value of 56%. There was no association between serum progranulin levels and the NASH score (data not shown).

After stepwise linear regression analysis adjusting for age, sex, body mass index, diabetes mellitus, the metabolic syndrome, HOMA-IR, blood pressure values, liver enzymes, lipid variables, and hs-CRP, serum progranulin levels retained their independent significance as a predictor of hepatocyte fibrosis in NAFLD patients \((β = 0.392; t = 2.226, P < 0.01)\).

4. Discussion

The role of progranulin in the pathogenesis of metabolic liver diseases is unclear due to the lack of the systematic observations. To our knowledge, this is the first study to examine the potential alterations
Fig. 2. ROC curve of serum progranulin concentration for the diagnosis of NAFLD. The area under the curve was 0.64 (standard error: 0.042; 95% confidence interval 0.57–0.71).

of serum progranulin levels in patients with biopsy-proven NAFLD. The results from this study indicate that (i) NAFLD patients have significantly higher serum progranulin concentrations than controls; (ii) progranulin concentrations are positively associated with serum lipid levels in NAFLD patients; and (iii) progranulin levels are independently associated with the degree of hepatic fibrosis among NAFLD patients after adjustment for a broad spectrum of potential confounders.

The selection of progranulin as a novel candidate biomarker for NAFLD is supported by the hyperexpression of this molecule in a mouse model of this condition [7], its expression in primary hepatic stellate cells [8], and its association with fibrosis following experimental injury [9]. In our study, serum progranulin levels were positively associated with total and LDL cholesterol. This relationship is intriguing as dyslipidemia is one of the core features of the metabolic syndrome [16]. The results of the present study confirm and expand those of Youn et al. [10] who showed in a pilot study that serum progranulin levels are positively associated with total cholesterol. Another observation is that serum progranulin was independently associated with the extent of liver fibrosis in NAFLD patients. Of note, this relation was independent from serum lipid values in multivariable regression analysis. Although this was an observational clinical study and no direct inference can be made on the mechanisms linking altered levels of progranulin with hepatic fibrogenesis, we speculate that this can occur via the regulation of the hepatocyte growth factor (HGF)/mesenchymal-epithelial transition factor (c-Met) system signaling pathway. In this regard, a very recent study has shown that progranulin plays a key role in the regulation of this biochemical mechanism in vivo [18], which in turn has been shown to be linked with chronic liver injury and fibrosis progression in both humans and experimental models [19]. However, the precise mechanisms underlying the increase of progranulin in patients with NAFLD need further investigation. Since both the cellular source of serum progranulin and its mechanisms of secretion are multiple [8,9], it is unclear whether the significant elevation of serum progranulin levels in NAFLD reflects a higher production or a reduced clearance. In this study, serum progranulin levels were related to the degree of fibrosis but not to the presence of NASH. This observation seems to suggest that progranulin can affect hepatic fibrosis but not the degree of steatosis or necroinflammation. We propose that the relationship between progranulin and fibrosis reflects a specific pathogenic role of progranulin in hepatic fibrogenesis and/or a failure of antifibrogenic pathways.

Several limitations of this study merit comment. First, our study has a cross-sectional nature and does not elucidate the causal relationship between serum progranulin levels and the presence of liver fibrosis. Therefore, we do not know whether serum progranulin levels could be mechanistically related to hepatic fibrosis by reflecting tissue progranulin expression in the liver. Eventually, a longitudinal study is needed to clarify the causal relationship between progranulin levels and liver injury. Second, the relatively small sample size limits the generalizability of our conclusions. Third, our sample included subjects of Turkish nationality, so that results cannot be extrapolated to populations with different ethnic background. These limitations notwithstanding, this is the first clinical study to demonstrate a relationship between progranulin and liver injury in patients with NAFLD. In addition, all patients with NAFLD underwent liver biopsy, the best standard for diagnosing this condition [20,21]. Finally, it is noteworthy that NAFLD patients with significant comorbidities were excluded from this study. Therefore, the confounding effects of concomitant diseases on the observed associations should not be so influential.

In conclusion, the results from this study demonstrate that patients with biopsy-proven NAFLD have
significantly higher serum progranulin concentrations (vs. matched controls) that independently predict the extent of liver fibrosis. Further studies using larger populations will be needed to confirm our observations and to validate the current findings.

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