Systematic Review on the Correlation of Quantitative PCR Cycle Threshold Values of Gastrointestinal Pathogens With Patient Clinical Presentation and Outcomes

Stéphane Bonacorsi †*, Benoit Visseaux2,3, Donia Bouzid2,4, Josep Pareja5, Sonia N. Rao6, Davide Manissero7, Glen Hansen8,9 and Jordi Vila10

1 Department of Microbiology, Robert Debré University Hospital, AP-HP, Paris, France, 2 Université de Paris, IAME, INSERM, Paris, France, 3 Université de Paris, Laboratoire de Virologie, Hôpital Bichat Claude Bernard, Assistance Publique-Hôpitaux de Paris, Paris, France, 4 Université de Paris, Service d’Accueil des Urgences, Hôpital Bichat Claude Bernard, Assistance Publique-Hôpitaux de Paris, Paris, France, 5 STAT-Dx Life, S.L. (a Qiagen Company), Medical Affairs, Barcelona, Spain, 6 Qiagen Inc., Medical Affairs, Germantown, MD, United States, 7 Qiagen Manchester Ltd, Medical Affairs, Manchester, United Kingdom, 8 Microbiology and Molecular Diagnostics, Hennepin County Medical Center, Department of Infectious Diseases, School of Medicine, University of Minnesota, Minneapolis, MN, United States, 9 Department of Pathology and Laboratory Medicine, School of Medicine, University of Minnesota, Minneapolis, MN, United States, 10 Biomedical Diagnostic Centre, Department of Clinical Microbiology, Institute of Global Health, School of Medicine, Hospital Clinic, University of Barcelona, Barcelona, Spain

Background: Quantitative (q) polymerase chain reaction (PCR) cycle threshold (Ct) values represent the number of amplification cycles required for a positive PCR result and are a proxy of pathogen quantity in the tested sample. The clinical utility of Ct values remains unclear for gastrointestinal infections.

Objectives: This systematic review assesses the global medical literature for associations between Ct values of gastrointestinal pathogens and patient presentation and clinical outcomes.

Data Sources: MEDLINE, EMBASE, Cochrane library databases: searched January 14–17, 2020.

Study Eligibility Criteria: Studies reporting on the presence or absence of an association between Ct values and clinical outcomes in adult and pediatric populations were included. Animal studies, reviews, meta-analyses, and non-English language studies were excluded.

Participants: Humans infected with gastrointestinal pathogens, detected with qPCR.

Interventions: Diagnostics assessing Ct values. Extracted data were reported narratively.

Results: Thirty-three eligible studies were identified; the most commonly studied pathogens were Clostridioides difficile (n = 15), norovirus (n = 10), and rotavirus (n = 9). Statistically significant associations between low C. difficile Ct values and increased symptom severity or poor outcome were reported in 4/8 (50%) studies, and increased risk of death in 1/2 (50%) studies; no significant associations were found between Ct value
INTRODUCTION

Gastrointestinal infections contribute significantly to the burden of illness from infectious diseases worldwide (1, 2). Rotavirus is the principal cause of diarrhea mortality, responsible for a high attributable fraction among all age groups (13.9%) (3). _Shigella_, the second most common cause of diarrhea mortality, is a key contributor to diarrheal death among children younger than 5 years (14.3%), mainly in low income countries (3).

Quantitative (q) polymerase chain reaction (PCR) is a robust and increasingly common methodology for rapid syndromic testing due to its sensitivity and specificity for identification of pathogens. In infectious diseases, qPCR cycle threshold (Ct) values represent the number of amplification cycles required for the fluorescent signal to exceed the basal threshold level. Ct values are inversely related to the number of copies of the target gene in a sample, meaning that lower Ct values correlate with higher pathogen loads. In infectious diseases, qPCR Ct values have potential utility in providing clinicians with information regarding genomic load that may help guide clinical and infection-control decisions. In addition, Ct values may help to clarify diagnostic uncertainty in cases where there is difficulty interpreting binary results, for example when distinguishing between causative infectious pathogen and asymptomatic carriage/colonization (4–6), particularly as identification of multiple pathogens is common (7, 8).

Notably, unprecedented challenges from the COVID-19 pandemic have raised the interest in clinical and diagnostic utility of Ct values (9, 10). However, in a recent systematic review of the utility of Ct values in respiratory infections (parallel to this study), no universal conclusions could be reached [In press: J Antimicrob Chemother 2021]. This systematic review assesses the global medical literature for associations between Ct values of gastrointestinal pathogens and patient or healthcare outcomes.

METHODS

This systematic review was undertaken according to the principles outlined in the Cochrane handbook and guidance published by the Center for Reviews and Dissemination. The original protocol was published in the PROSPERO database (CRD42020167239) and included broad search terms unrestricted by pathogen or disease type. This review focuses on gastrointestinal pathogens.

Eligibility Criteria

Literature searches of MEDLINE, EMBASE, and the Cochrane Library using search tools at ncbi.nlm.nih.gov/pubmed, embase.com and cochranelibrary.com were undertaken to identify studies reporting on the presence or absence of an association between qPCR Ct values and patient or healthcare outcomes (see Supplementary Table 1 for the PubMed search strategy). The search strategy comprised three concepts: (real-time [rt]-PCR OR qPCR) AND Ct values AND pathogen. Randomized-controlled, single-arm, non-randomized comparative and observational (retrospective or prospective) studies were included. Animal studies, systematic reviews, non-systematic reviews and meta-analyses were excluded; however, additional publications were identified by manual citation searching of appropriate reviews. Searches were limited to English language studies, for reasons of feasibility.

Study Selection and Data Extraction

Titles and abstracts were screened, based on eligibility criteria, for inclusion by two independent reviewers who then assessed the full texts of relevant studies; a third reviewer resolved conflicts. Key data from all included studies were captured by one reviewer, and subsequently verified by another reviewer. Outcomes were broadly divided into the following categories: mortality, symptomatic vs. asymptomatic, severity of symptoms, duration of symptoms, intensive care unit (ICU) admission, hospitalization and length of stay (LOS).

The quality and risk of bias of each study was assessed using a tool relevant for each study design (Newcastle Ottawa Scale for cross-sectional, cohort, and case-control studies (11)).
Records identified through database searching  
*n = 1,181*

Records added manually  
*n = 147*

Records after duplicates removed  
*n = 1,029*

Records excluded based on titles/abstract  
*n = 830*

Records excluded based on full text  
*n = 165*

*Not GI pathogens (n = 137)  
No relevant outcome or Ct values not reported (n = 26)  
Relevant outcomes but excluded (n=3)*

Pathogens reported (number of articles); some studies reported Ct value associations for more than one pathogen

**Clostridioides difficile**  
*n = 15*

**GI viruses**  
Norovirus  
*n = 10*  
Rotavirus  
*n = 9*  
Adenovirus  
*n = 4*  
Astrovirus  
*n = 4*  
 Sapovirus  
*n = 4*  

**Non-C. difficile bacteria**  
Shigella spp.  
*n = 6*  
Escherichia coli  
*n = 6*  
Campylobacter spp.  
*n = 5*  
Salmonella spp.  
*n = 5*  
Vibrio cholerae  
*n = 1*  
Aeromonas spp.  
*n = 1*  

**Parasites**  
Cryptosporidium spp.  
*n = 6*  
Giardia spp.  
*n = 3*  
Entamoeba spp.  
*n = 2*  
Cyclospora spp.  
*n = 1*  

**FIGURE 1** | PRISMA flow diagram. Ct, cycle threshold; GI, gastrointestinal. "Details of these publications are provided in the Supplementary Material."
**RESULTS**

**Overview of Studies Included**

Literature searches, conducted January 14–17, 2020, identified 1,029 unique records. Application of distinct screening and restriction parameters specific to gastrointestinal infections identified 33 eligible studies. Most studies reported Ct value association for more than one pathogen; the most commonly studied pathogens were *Clostridioides difficile* (n = 15), norovirus (n = 10), and rotavirus (n = 9) (Figure 1). All studies identified gastrointestinal pathogens from stool samples. In studies of *C. difficile*, the majority used genes encoding toxin A or B as PCR targets.

The majority of outcomes reported were related to symptoms, including symptom severity, symptomatic vs. asymptomatic and duration of symptoms. Mortality was assessed by three studies. No studies investigated associations between Ct values and hospitalization and/or ICU admission. The majority (84.8%; 28/33) of studies did not report normalized Ct values. Some (66.7%; 22/33) studies presented Ct value distributions.

**Quality and Bias**

Using Newcastle-Ottawa scales, all cross-sectional studies, cohort studies and case-control studies were classed as being of poor quality (Supplementary Tables 2–4, respectively). This was generally due to a lack of comparability between groups, insufficient or unjustified sample sizes, the use of non-representative samples (often hospitalized patients or age-specific populations) and a lack of detail regarding patient follow-up or non-response; ascertainment of exposure and outcome was usually appropriate. In an assessment of qPCR methodology, 14/29 (48%) full-length articles were considered to have some or many gaps in the reported methodology (Supplementary Table 5).

**Clostridioides difficile**

*C. difficile* was the most commonly reported gastrointestinal pathogen with respect to articles describing associations between Ct values and patient or healthcare outcomes (Table 1). Two studies investigated the association of Ct value with mortality, of which one (N = 1,013) reported no significant associations (12). The second study, conducted by Davies et al. at four UK hospitals, was the largest *C. difficile* assessment in this systematic review (N = 1,281). The authors reported significantly lower median Ct values for patients who died with *C. difficile* infection compared with those who survived [25.5 (n = 123) vs. 27.5 (n = 762), respectively; p = 0.021] (13).

Among 12 articles reporting associations between Ct values and symptoms, eight investigated severity of symptoms. Three studies reported significantly lower median Ct values in patients with severe or complicated disease vs. those with mild/moderate disease: De Francesco et al. (N = 421) severe 25.9 (n = 199) vs. mild/moderate 28.1 (n = 222), p = 0.00001; Jazmati et al. (N = 99) severe 26.5 vs. mild/moderate 31.2, p = 0.02; Kamboj et al. (N = 183) severe 24.5, complicated 22.5, and non-severe 28.0, p = 0.005 (14–16). Jazmati et al. further described lower Ct values as a predictor of severe disease [area under the receiver operating characteristic curve 0.77, 95% confidence interval (CI) 0.62–0.92; p = 0.013] (14). Reigadas et al. (n = 299) showed that Ct value was independently associated with poor outcome (p < 0.001) and classified patients into risk categories accordingly; high risk of poor outcome (median Ct <23.5); medium risk of poor outcome (median Ct 23.5–27.9); and low risk of poor outcome (median Ct ≥28.0) (17). A further three studies with numbers of PCR-positive patients ranging from 62 to 219, reported lower Ct values in patients with poorer outcomes or more severe disease; however, differences did not reach statistical significance (18–20).

Four studies investigated differences in Ct values in case vs. control subjects. In Crobach et al. (N = 208) mean quantification cycle (Cq) values were significantly lower (p < 0.001) in symptomatic patients who were toxin A/B-positive by enzyme immunoassay (24.4, 95% CI 23.5–25.3) than symptomatic patients who were toxin A/B-negative (30.4, 95% CI 29.5–31.3) and asymptomatic carriers (29.2, 95% CI 27.3–31.2) (5). Similar observations were reported in pediatric patients by Bub et al. (N = 13; median Ct 32 in symptomatic cases vs. 36 in controls, no significance reported) and Hecht et al. (N = 193; median Ct 23.8 in true infections vs. 30.5 in colonized, p = 0.03) (6, 21).

In a study (n = 85) by Brujinesteijn van Coppenraet et al., although no significant difference in Ct values were observed between controls and across all subjects, Ct values were significantly lower in cases vs. controls for age group 21–50 years (22).

Two studies (N ≤ 120) investigated association of Ct value with duration of symptoms; no significant associations were reported in either study (23, 24). One large study (N = 1,281) of diarrheal patients in the UK investigated Ct value and LOS; however, no significant associations were reported, except for patients with PCR-ribotype 027, where LOS was significantly increased in those with low vs. high Ct value (32.5 vs. 28 days; p = 0.018) (13).

**Gastrointestinal Viruses**

Associations between patient or healthcare outcomes and the Ct value of gastrointestinal viruses were investigated in 14 studies, with the most commonly studied viruses being norovirus and rotavirus (n = 10 and n = 9, respectively) (Table 2). The majority of studies (n = 10) investigated the difference in Ct values (or viral load derived from Ct values) between cases and controls (symptomatic and asymptomatic, or patients with or without diarrhea).

In general, norovirus, particularly norovirus genogroup II (GI), infections were found to have significantly lower median Ct values in infections vs. controls. Kabue et al. (N = 122) reported that lower median Ct values were observed in symptomatic pediatric patients compared with asymptomatic pediatric patients infected with norovirus GII (n = 104; 27.0 vs. 34.6; p = 0.0009) (25). Similar outcomes were reported in Kabayiza et al. (n = 51; 25.8 vs. 29.5; p = 0.04), Phillips et al. (n = 589; 34 vs. 37; p < 0.0001), Saito et al. (n = 467; 26.4 vs. 30.1; p = 0.0001), and Dung et al. (n = 138; 6.85 log copies/ml vs. 5.07 log copies/ml; p = 0.02) (4, 26–28).
### TABLE 1 | Summary of studies that assessed PCR Ct values for *C. difficile* infections against patient clinical presentation and outcomes.

| Outcome | Study | Number of PCR+ patients | Population | Outcome measure (significant associations bolded) |
|---------|-------|--------------------------|------------|--------------------------------------------------|
| Mortality | Davies et al. Plos One 2018 | 1,281 | UK, hospital | • Lower median Ct values for patients who died (*n* = 123) with infection compared with those who survived (*n* = 762; median Ct 25.5 and 27.5, respectively; *p* = 0.021)  
• Following optimal cut-off determination, low Ct was defined as ≤25 and was significantly associated with mortality (*p* = 0.032)  
• There was no association between Ct values and 30-day mortality [OR 1 (95% CI 0.93–1.08); *p* = 0.95] |
|          | Rao et al. CID 2015 | 1,013 | USA, hospital, ≥ 18 years | |
| Severity of symptoms | Rao et al. CID 2015 | 1,013 | USA, hospital, ≥ 18 years | • There was no association between Ct values and severe CDI [OR 1.01 (95% CI 0.93–1.09); *p* = 0.873]  
• Ct values ≤25 were significantly associated with severe disease vs. mild/moderate disease [97 (55%) vs. 79 (44%); *p* = 0.0075]  
• Ct values >25 were significantly associated with mild/moderate disease vs. severe disease [143 (58.3%) vs. 102 (41.6%); *p* = 0.004]  
• The mean Ct values of *tcdB* PCR in patients with mild/moderate disease were significantly higher (28.1; IQR 7.7) than in patients with severe disease (25.9; IQR 5.9) (*p* = 0.00001)  
• A Ct value ≤26 was significantly associated with patients with a severe disease  
|          | De Francesco et al. Anaerobe 2019 | 421 | Italy, hospital and community | • The mean PCR Ct was lower for patients with a poor outcome (24.9 ± 4.24 vs. 26.05 ± 4.47; *p* = 0.07) (“Poor outcome” was defined as the occurrence of a severe or severe-complicated first CDI episode and/or all-cause death within the first 8 weeks after the end of treatment)  
• The optimal cut-off Ct value was established as 27.55, yielding a sensitivity of 78.6% (95% CI 67.1–87.5), a specificity of 35.7% (95% CI 32.8–43.7), a PPV of 35.3% (95% CI 31.5–39.2), and an NPV of 78.9% (95% CI 69.5–85.9) |
|          | Origüen et al. JCM 2019 | 219 | Spain, hospital, ≥ 18 years | • Severe and complicated infections were associated with lower Ct values than non-severe infections [median Ct values for non-severe Ct = 26.0 (95% CI 168), severe Ct = 24.5 (95% CI 11), and complicated Ct = 22.5 (95% CI 4); *p* = 0.005]  
|          | Kamboj et al. J Infect 2018 | 183 | USA, tertiary care cancer hospital | Derivation cohort  
• Ct value was independently associated with poor outcome by multivariate analysis [OR 0.701 (95%CI 0.604–0.813); *p* < 0.001]  
• Patients were classified into risk categories; high risk of poor-outcome (Ct >23.5 cycles); medium risk of poor-outcome (Ct 23.5–27.9 cycles); and low risk of poor-outcome (Ct ≤28.0 cycles). The sensitivity of the rule was 46.5% (95% CI 32.5–61.1) and specificity was 98.8% (95% CI 93.7–99.8), the PPV was 95.2% (95% CI 86.1–100) and NPV was 78.7% (95% CI 70.9–86.4); the diagnostic accuracy was 81.4% (95% CI 74.7–88.1)  
• Patients with poor-outcome CDI episodes had lower median Ct values than those without poor-outcome CDI episodes (24.8 vs. 28.9; *p* < 0.001)  
|          | Reigadas et al. J Antimicrob Chemother 2016 | 129 (Validation cohort: 170) | Spain, hospital, ≥ 17 years | Validation cohort  
• Median Ct value was lower for episodes with poor outcome than favorable (21.9 vs. 27.0; *p* < 0.001)  
• Independent association between Ct value and poor outcome (*p* < 0.001) and the high-risk category (Ct < 23.5) and poor outcome (*p* < 0.001)  
• No difference in organism burden between groups with (*n* = 59) and without (*n* = 59) clinically significant diarrhea [median Ct, 26.9; IQR 23.9–32.2] vs. 27.1 (IQR 23.4–30.7); *p* = 0.25; mean Ct 27.9 vs. 27.4]  
• Patients with severe disease had significantly lower Ct values compared with non-severe infections [26.5 ± 4.8, (*n* = 9) vs. 31.2 ± 4.8 (*n* = 49); *p* = 0.022], describing lower Ct values as a predictor of severe disease (area under the receiver operating characteristic curve 0.77, 95% CI 0.62–0.92; *p* = 0.013)  
• The was no significant difference between Ct values of patient with and those without serious disease (27 ± 4 (*n* = 42) vs. 29 ± 4 (*n* = 20); respectively; *p* = 0.23)  
| Case vs. control | Crobach et al. J Clin Microbiol 2018 | 208 | Netherlands, hospital | • Comparable mean Ct values were observed for symptomatic patients with subsequent negative toxin A/B immunoassay results [30.4 (95% CI 29.5–31.3)] and asymptomatic carriers [29.2 (95% CI 27.3–31.2)], while symptomatic patients with positive toxin A/B results had significantly lower mean Ct values, according to ANOVA (24.4 (95% CI 23.5–25.3); *p* < 0.001) |

(Continued)
Additionally, Liu et al. reported a pathogen quantity-dependent association with diarrhea in children <5 years old (29). Elfvéng et al. also reported lower median Ct values in patients vs. controls, but the difference was not significant (25.1 vs. 26.9; p = 0.28) (30).

One study investigated Ct values of norovirus GII and fatal outcomes (n = 534) and found no association (31). One other study reported no significant associations between Ct values and symptom duration (n = 623) or infectiousness (n = 110) in patients with infections caused by norovirus (32).

Similar to norovirus, multiple studies showed significantly lower Ct values (or Cq) in cases of symptomatic rotavirus infection vs. controls. Phillips et al. (N = 153) reported lower median Ct values in rotovirus intestinal infections vs. controls (18 vs. 37; p < 0.0001) (33). Dung et al. (n = 113) reported significantly higher median viral loads in children with diarrhea compared with those without (10.6 log copies/ml vs. 8.33 log copies/ml; p < 0.001) (26), and one study in children <5 years by Liu et al. reported strong pathogen quantity-dependent associations with diarrhea (29). Supporting these observations, Kabayiza et al. (n = 325) reported that lower median Ct values were significantly associated with more severe symptoms, including vomiting, severe dehydration and intravenous fluid therapy, in patients with infections caused by rotavirus (27). Kang et al. (N = 91) also reported significant associations between severe diarrhea and low Ct values (reported as "crossing points") (34). Four further studies also reported lower median Ct values in patients vs. controls/asymptomatic patients, but differences did not reach statistical significance: Elfvéng et al. (n = 19; 24.4 vs. 26.0; p = 0.50); Kabayiza et al. (n = 238; 21.16 vs. 23.29; p = 0.24); Ramani et al. (n = 103; 26.26 vs. 27.34; p = 0.087) and Mukhopadhya et al. (n = 15; 17.21 vs. 30.98; p = 0.086) (27, 30, 35, 36). Notably, adjustment of an outlier in the study by Mukhopadhy et al. resulted in the difference reaching statistical significance (p = 0.007) (36).

**TABLE 1 | Continued**

| Outcome | Study | Number of PCR+ patients | Population | Outcome measure (significant associations bolded) |
|---------|-------|--------------------------|------------|--------------------------------------------------|
| Duration of symptoms | Feghaly et al. CID 2013a | 120 | USA, hospital, adult | When patients were segregated into quartiles based on their initial Ct values, there was no difference in the time to diarrhea resolution among patients |
| | Feghaly et al. J Ped 2013b | 74 | USA, hospital, pediatric | When patients were segregated into quartiles based on their initial Ct values, there was a paradoxical trend toward a longer interval to diarrhea resolution in children with a lower bacterial burden at diagnosis (p = 0.06) |
| | | | | Lower fecal bacterial burden at diagnosis (as calculated by Ct value) was associated with longer times to diarrhea resolution (HR: 0.93; 95% CI: 0.86–1; p = 0.068) |
| Recurrence | Orgüen et al. JCM 2019 | 219 | Spain, hospital, ≥18 years | The mean Ct value was lower in patients with recurrence compared with those without (24.00 ± 3.28 vs. 26.02 ± 4.54; p = 0.002) |
| Median LOS | Davies et al. Plos One 2018 | 1,281 | UK, hospital | Patients with low Ct values (≥25) had a numerically greater LOS compared with those with high Ct values (<25); however this difference was not significant (Ct ≥25: 25.7 days vs. Ct <25: 23 days; p = 0.77) |
| | | | | In patients with presence of PCR-ribotype 027, LOS was significantly increased in those with low vs. high Ct (32.5 days vs. 28 days; p = 0.018) |
| Other biomarker | Davies et al. Plos One 2018 | 1,281 | UK, hospital | Lower Ct values were associated with: |
| | | | | Higher mean white cell count (Ct ≥25: 12.1 × 10^9/L vs. Ct >25: 10.9 × 10^9/L; p = 0.3) |
| | | | | Higher baseline mean serum creatinine (Ct ≥25: 120.0 mg/dL vs. Ct >25: 110.7 mg/dL; p = 0.04) |
| | De Francesco et al. Anaerobe 2019 | 421 | Italy, hospital and community | Statistically significant correlation between low Ct values and leucocytosis (p < 0.001) but not with the alteration in baseline creatinine or serum albumin level |

Text in bold indicates a statistically significant association. ANOVA, analysis of variance; CDI, C. difficile infection; CI, confidence interval; Ct, cycle threshold; HR, hazard ratio; IQR, interquartile range; LOS, length of stay; NPV, negative predictive value; OR, odds ratio; PCR, polymerase chain reaction; PPV, positive predictive value; UK, United Kingdom; USA, United States of America.

*Likely setting although not confirmed in source material.
TABLE 2 | Summary of studies that assessed PCR Ct values for gastrointestinal viruses against patient clinical presentation and outcomes.

| Outcome                      | Study                                           | Pathogen(s)           | Number of PCR+ patients | Population                          | Outcome measure                                                                 |
|------------------------------|------------------------------------------------|-----------------------|-------------------------|-------------------------------------|---------------------------------------------------------------------------------|
| Mortality                    | Gustavsson et al. J Clin Virol 2015             | Norovirus             | 534*                    | Sweden, hospital, ≥60 years          | ▪ Ct values were not associated with fatal outcomes [46 patients deceased; HR 0.97 [95% CI 0.92–1.02] per Ct unit decrease; p = 0.17] |
| Severity of symptoms         | Kabayiza et al. Clin Microbiol and Infec 2014b| Norovirus GII         | 98                      | Rwanda, community and hospital, ≤5 years | ▪ No significant difference in Ct values was observed for norovirus GI or GII for clinical markers including vomiting, dehydraton or intravenous fluid use |

| Norovirus GII                | OR 1.80                                         | 0.33                  | 2.08                    | ▪ There was no difference in median Ct values between symptomatic and asymptomatic patients [28.06 vs. 27.58, respectively; p = 0.32] |
|                              | p = 0.35                                        | p = 0.77              | p = 0.13                | ▪ Significantly lower median Ct values were observed in symptomatic patients compared with asymptomatic patients [27.02 vs. 28.94; p = 0.06] |
|                              | Ct 30.2/31.7                                    | 34.0/29.4/31/0        | 30.2/28.3               | ▪ Significantly lower median Ct values were observed in symptomatic patients compared with asymptomatic patients [25.79 vs. 29.54; p = 0.04], but not for norovirus GII (29.25 vs. 28.94; p = 0.06) |
|                              | OR 0.84                                         | 0.69                  | 0.37                    | ▪ Significantly lower Ct values in patients vs. controls for norovirus GII (25.79 vs. 29.54; p = 0.04), but not for norovirus GII (29.25 vs. 28.94; p = 0.06) |
|                              | Ct 27.7/28.6                                    | 30.4/27.2/28.4        | 27.0/28.5               | ▪ There was no significant difference in median Ct values between patients and controls for infections caused by norovirus GII (25.1 vs. 26.9, respectively; p = 0.28) |

(Continued)
### TABLE 2 | Continued

| Outcome | Study | Pathogen(s) | Number of PCR+ patients | Population | Outcome measure |
|---------|-------|-------------|-------------------------|------------|-----------------|
| Duration of symptoms | Partridge et al. J Hosp Infect 2012 | Norovirus | 623\(^1\) | UK, hospital | No significant correlation was identified between duration of symptoms from time of sampling and Ct value of the sample (Spearman rank correlation coefficient: \(-0.077; p > 0.2\)) |
| Infectiousness Partridge et al. J Hosp Infect 2012 | Norovirus | 110 | UK, hospital | There was no significant difference in initial Ct value between onward transmitters and non-transmitters (24.98 vs. 26.56; \(p = 0.19\)) |

**ROTAVIRUS**

#### Severity of symptoms

| Study | Pathogen(s) | Number of patients | Population | Outcome measure |
|-------|-------------|-------------------|------------|-----------------|
| Kabayiza et al. Clinc Microbiol and Infec 2014b | Rotavirus | 325 | Rwanda, community and hospital, ≤5 years | Lower Ct values for rotavirus were significantly associated with multiple clinical markers (vomiting, more severe dehydration and intravenous fluid therapy) in univariate and multivariate analyses |
| Kang et al. J Med Virol 2004 | Rotavirus A | 91 | India, hospital and community, pediatric | There was a significant negative correlation (\(r = -0.80; p < 0.001\)) between symptom severity and the crossing point (Ct value) on the assay, indicating that children with more severe diarrhea have higher viral loads than children with less severe disease |
| Liu et al. Lancet 2016 | Rotavirus | 5,304\(^1\) | Bangladesh, India, Pakistan, The Gambia, Kenya, Mal and Mozambique, community, <5 years | Rotavirus had strong Ct-dependent associations with diarrhea |
| Phillips et al. J Clin Virol 2009b | Rotavirus A | 153 cases | England, community/primary care | The median rt-PCR Ct value was significantly lower in infectious intestinal disease cases vs. control, both in all ages and when the analysis was restricted to children aged <5 years |
| Kabayiza et al. Pediatr Infect Dis J 2014a | Rotavirus | 238 | Rwanda, community and hospital, ≤5 years | No significant difference in median Ct values between patients vs. controls (21.16 vs. 22.39; \(p = 0.24\)) |

(Continued)
### TABLE 2 | Continued

| Outcome | Study | Pathogen(s) | Number of PCR+ patients | Population | Outcome measure |
|---------|-------|-------------|-------------------------|------------|-----------------|
| Dung et al. J Virol Methods 2012 | Rotavirus A | 113 | Vietnam, hospital, <60 months | • Results are presented in log of target RNA copy number per mL; however, it is noted that these were converted from Cp values using a standard curve • The viral load of rotavirus A was significantly higher in samples from children with diarrhea (10.6 log/RNA copies/mL; 5.56–12.49) than from those without (8.33 log/RNA copies/mL; 5.43–10.52) (p < 0.001) |
| Raman et al. J Med Virol | Rotavirus | 103 | India, hospital, neonatal | • The mean Ct value was 26.26 (SD 3.06) for symptomatic neonates and 27.34 (SD 2.73) for asymptomatic neonates • There was no significant difference in viral load between symptomatic and asymptomatic neonates (p = 0.087) • Neonates with feed intolerance and abdominal distension had significantly lower Ct values than those with other gastrointestinal symptoms (p = 0.02) |
| Elfving et al. JCM 2014 | Rotavirus | 19 | Zanzibar, community, 2 months−5 years | • There was no significant difference in median Ct values between patients and controls for infections caused by rotavirus (24.4 vs. 26.0, respectively; p = 0.5) • By multivariate logistic regression analysis (accounting for age and gender), a cut-off Ct value of 45 was associated with disease (OR 5.8; CI 1.7–20.3; p < 0.003) |
| Mukhopadhya et al. J Med Virol 2013 | Rotavirus | 15: 10 symptomatic and 5 asymptomatic | India, hospital and community, <5 years | • The median Cq at presentation in symptomatic children was 17.21 (IQR 14.36–23.96) compared with 30.98 (IQR 29.38–31.50) in asymptomatic children (p = 0.086) • Once removing an outlier in the asymptomatic group, the difference between the initial shedding between symptomatic and asymptomatic samples was found to be statistically significant (p = 0.007) |

### ALL OTHER GASTROINTESTINAL VIRUSES

| Severity of symptoms | Kabayiza et al. Clin Microbiol and Infec 2014b | Rwanda, community and hospital, ≤5 years | • No significant associations between Ct values and clinical markers were observed for adenovirus, astrovirus or sapovirus |
|-----------------------|-----------------------------------------------|-------------------------------------------|--------------------------------------------------|
| Vomiting (Y/N) Dehydration (Severe/moderate/mild) IV fluid (Y/N) | Adenovirus | 216 | OR | 0.77 | 0.59 | 0.70 |
| | | | p | 0.18 | 0.014 | 0.049 |
| | Astrovirus | 36 | OR | 0.69 | 1.13 | 1.67 |
| | | | p | 0.34 | 0.85 | 0.17 |
| | | Cq | 26.7/24.7 | 26.0/26.7/24.6 | 26.7/24.9 |
| | Sapovirus | 33 | OR | 0.68 | 0.44 | 0.75 |
| | | | p | 0.33 | 0.32 | 0.48 |
| | | Cq | 29.4/29.2 | 39.1/28.6/26.4 | 30.6/26.4 |

### Symptomatic vs. asymptomatic (or case versus control)

| Liu et al. Lancet 2016 | Adenovirus, Sapovirus, Astrovirus | Bangladesh, India, Pakistan, The Gambia, Kenya, Mal and Mozambique, community, <5 years | • Adenovirus had strong Ct-dependent associations with diarrhea. Cq values <35.0 were defined as “diarrhea-associated” as the 95% CI or the OR was >1. Cq values <22.7 were defined as “highly diarrhea-associated” as the 95% CI or the OR was >2. The ROC cut-off maximally discriminating case-control status was a Cq value of 30.2 (Youden Index 0.08) • Astrovirus showed associations with diarrhea. Cq values <25.5 were defined as “diarrhea-associated” and <22.2 were defined as “highly diarrhea-associated” (ROC cut-off 28.1; Youden index 0.18) • Sapovirus was only moderately associated with diarrhea. A Cq values <31.6 were defined as “diarrhea-associated” (ROC cut-off 34.1; Youden Index 0.02) |
Three studies investigated gastrointestinal viruses other than norovirus and rotavirus. In one study that investigated pathogen quantity and diarrhea in children <5 years old, associations between Ct value and diarrhea were reported for cases of adenovirus and astroviruses (29). No other associations between Ct values and cases vs. controls were identified (27, 30).

### Non-**C. difficile** Bacterial and Parasitic Pathogens

Associations between patient clinical outcomes and the Ct value of non-**C. difficile** bacterial and parasitic pathogens were investigated in nine studies (Table 3).

Among bacterial studies, the majority investigated associations between quantitative PCR-derived bacterial loads and cases vs. controls (symptomatic vs. asymptomatic, or patients with vs. without diarrhea), and most studies found significant associations. Among five studies reporting differences in Ct values between cases vs. controls, significantly lower median Ct values were reported in cases of enterotoxigenic *Escherichia coli* (ETEC), enteropathogenic *E. coli* (EPEC), *Campylobacter* spp., enteroinvasive *E. coli* (EIEC)/*Shigella* spp., and *Salmonella* spp. (22, 27, 29, 30, 37). However, associations were not consistent across studies, including two reports (n = 9 and n = 46) of no significant difference in cases vs. controls for *Salmonella* spp. (27, 30). In one study, associations were notably weaker for *Campylobacter* spp. and typical EPEC (29). In a study (n = 143) of patients with EPEC, a 29% increase in risk of diarrhea was observed for each log_{10} unit increase (calculated by Ct value) in bacterial load (OR 1.29; 95% CI 1.08–1.53) (37).

Two studies also investigated associations between Ct values and bacterial disease severity. In cases of EIEC/*Shigella* spp., lower Ct values were significantly associated with higher vs. lower categories of disease severity (n = 286; Ct value 25.3 vs. 36.6), dehydration (n = 154; OR 3.89; p = 0.02), and requirement for intravenous fluids (n = 154; OR 2.29; p = 0.01) (27, 38). Lower Ct values for ETEC-estA were significantly associated with vomiting (n = 167; OR 1.74; p = 0.02) and with intravenous fluids (n = 167; OR 1.81; p = 0.004), and *Campylobacter* spp. with vomiting (n = 147; OR 2.21; p = 0.03) (27).

One study (n = 143) investigated the effect of EPEC bacterial load on the duration of symptoms; however, no significant association was observed (37).

All studies of parasites investigated associations between Ct values (or Cq) and cases vs. controls (symptomatic vs. asymptomatic, or patients with vs. without diarrhea). In studies including *Cryptosporidium* spp., two reported significantly lower Ct values in cases vs. controls, including Elfving et al. (n = 67; median Ct 32.1 vs. 36.8; p = 0.0009) (22, 30). One further study also reported lower Ct values in cases vs. controls (n = 23), but did not reach statistical significance (27). Furthermore, and contrary to expected results, Haque et al. reported higher mean Ct values in *Cryptosporidium parvum* and *Cryptosporidium hominis* cases than controls, although the differences were not significant (p = 0.127 and 0.098) (39). In a study in children <5 years (n = N/A), strong pathogen quantity-dependent associations
### TABLE 3 | Summary of studies that assessed PCR Ct values for non-\textit{C. difficile} bacterial and parasitic pathogens against patient clinical presentation and outcomes.

| Outcome                  | Study                         | Pathogen(s)          | Number of PCR+ patients | Population                      | Outcome measure |
|--------------------------|-------------------------------|-----------------------|--------------------------|---------------------------------|-----------------|
| **BACTERIA**             |                               |                       |                          |                                 |                 |
| Severity of symptoms     | Kabayiza et al. Clin Microbiol and Infec 2014b | 
Non-\textit{C. difficile} bacterial and parasitic pathogens | Rwanda, community and hospital, ≤5 years | • Higher pathogen load (lower Ct value) of ETEC-\textit{estA}, \textit{Shigella} spp and \textit{Campylobacter} spp was significantly associated with multiple clinical markers (vomiting, dehydration and intravenous fluid therapy) by age-adjusted multivariate analysis |
|                          |                               |                       |                          |                                 |                 |
| ETEC-\textit{estA}       | 167                           |                       |                          | Vomiting:                       |                 |
|                          |                               |                       |                          | Yes/No                          | 1.74            |
|                          |                               |                       |                          | OR (CI)                          | (1.08–2.84)     |
|                          |                               |                       |                          | \( p = 0.004 \)                 |                 |
| Shigella spp             | 154                           |                       |                          | Dehydration:                     |                 |
|                          |                               |                       |                          | Yes/No                          | 0.97 (0.59–1.60) |
|                          |                               |                       |                          | OR (CI)                          | (1.20–2.75)     |
| Campylobacter spp.       | 147                           |                       |                          | IV Fluids:                       |                 |
|                          |                               |                       |                          | Yes/No                          | 1.81            |
|                          |                               |                       |                          | OR (CI)                          | (1.21–4.55)     |
|                          |                               |                       |                          | \( p = 0.01 \)                  |                 |
| **OTHERS**               |                               |                       |                          | By univariate analysis, lower Ct values were associated with worse symptoms for \textit{Campylobacter} spp., ETEC-\textit{eltB}, ETEC-\textit{estA}, \textit{EPEC bfpA} and \textit{Shigella} |
|                          |                               |                       |                          |                                 |                 |
| Campylobacter spp.       | 147                           |                       |                          | Vomiting:                       | 2.21            |
|                          |                               |                       |                          | No/Yes                          | 1.90 (0.82–4.66) |
|                          |                               |                       |                          | OR (CI)                          | (0.90–3.02)     |
|                          |                               |                       |                          | \( p = 0.11 \)                  |                 |
| ETEC-\textit{eltB}       | 275                           |                       |                          | Dehydration:                     |                 |
|                          |                               |                       |                          | Yes/No                          | 1.49            |
|                          |                               |                       |                          | OR (CI)                          | (1.21–4.55)     |
|                          |                               |                       |                          | \( p = 0.01 \)                  |                 |
| ETEC-\textit{estA}       | 167                           |                       |                          | IV Fluids:                       |                 |
|                          |                               |                       |                          | Yes/No                          | 1.71            |
|                          |                               |                       |                          | OR (CI)                          | (1.21–4.55)     |
|                          |                               |                       |                          | \( p = 0.04 \)                  |                 |
| EPEC \textit{bfpA}       | 125                           |                       |                          | Vomiting:                       |                 |
|                          |                               |                       |                          | No/Yes                          | 1.50            |
|                          |                               |                       |                          | OR (CI)                          | (1.21–4.55)     |
|                          |                               |                       |                          | \( p = 0.04 \)                  |                 |
| EPEC \textit{eae}        | 222                           |                       |                          | Dehydration:                     |                 |
|                          |                               |                       |                          | Yes/No                          | 1.50            |
|                          |                               |                       |                          | OR (CI)                          | (1.21–4.55)     |
|                          |                               |                       |                          | \( p = 0.04 \)                  |                 |
| Salmonella               | 58                            |                       |                          | IV Fluids:                       |                 |
|                          |                               |                       |                          | Yes/No                          | 1.49            |
|                          |                               |                       |                          | OR (CI)                          | (1.21–4.55)     |
|                          |                               |                       |                          | \( p = 0.04 \)                  |                 |
| \textit{Shigella}        | 154                           |                       |                          | Vomiting:                       |                 |
|                          |                               |                       |                          | No/Yes                          | 1.49            |
|                          |                               |                       |                          | OR (CI)                          | (1.21–4.55)     |
|                          |                               |                       |                          | \( p = 0.04 \)                  |                 |

(Continued)
### TABLE 3 | Continued

| Outcome | Study | Pathogen(s) | Number of PCR+ patients | Population | Outcome measure |
|---------|-------|-------------|--------------------------|------------|----------------|
| Vu DT et al. J Clin Microbiol 2004 | Shigella | 286 | Vietnam, community | • The trend between increasing number of rt-PCR cycles (Ct values) and decreasing disease severity was highly significant ($p < 0.001$)  
• The number of PCR cycles required to detect a PCR product was highest for patients $\geq 5$ years with culture-negative, non-bloody diarrheal specimens (36.6) and was lowest for children (<5 years) with culture-positive, bloody diarrheal specimens (25.3) ($p < 0.001$) |
| Liu et al. Lancet 2016 | Shigella spp., EIEC, ETEC, Campylobacter jejuni or C. coli, EPEC, Vibrio cholerae, Salmonella spp, EAEC, Aeromonas spp | 5,304* | Bangladesh, India, Pakistan, The Gambia, Kenya, Mali, and Mozambique, community, <5 years | • Shigella spp. or EIEC, and heat-stable ETEC had strong quantity-dependent associations with diarrhea  
• For Shigella/EIEC, Cq values $<33.1$ were defined as “diarrhea-associated” and $<27.9$ were defined as “highly diarrhea-associated” (ROC cut-off 26.1; Youden index 0.18)  
• For heat-stable ETEC, Cq values $<26.2$ were defined as “diarrhea-associated” and $<22.8$ were defined as “highly diarrhea-associated” (ROC cut-off 25.4; Youden index 0.25)  
• Campylobacter jejuni or C. coli and EPEC were only moderately associated with diarrhea  
• For Campylobacter spp., Cq values $<19.7$ were defined as “diarrhea-associated” and $<15.4$ were defined as “highly diarrhea-associated” (ROC cut-off 19.9; Youden index 0.07)  
• Vibrio cholerae and Salmonella spp. showed associations with diarrhea  
• For Vibrio cholerae, Cq values $<34.9$ were defined as “diarrhea-associated” and $<33.8$ were defined as “highly diarrhea-associated” (ROC cut-off 29.3; Youden index 0.55)  
• For Salmonella spp, Cq values $<32.4$ were defined as “diarrhea-associated” and $<30.7$ were defined as “highly diarrhea-associated” (ROC cut-off 28.7; Youden index 0.29)  
• EAEC and Aeromonas spp were associated with diarrhea in specific study sites or age strata  
• Significantly higher relative loads were observed for Campylobacter spp. ($p < 0.005$), Salmonella spp. ($p < 0.005$), ETEC ($p < 0.05$) and typical EPEC ($p < 0.005$)  
• Ct values were significantly higher for STEC cases vs. controls ($p < 0.05$)  
• No significant difference in Ct values between cases and controls were observed for EAEC or atypical EPEC |
| Bruijnestein et al. Clin Microbiol Infect 2015 | Campylobacter spp. | 187 | Netherlands, primary Care | • Significantly higher relative loads were observed for Campylobacter spp. ($p < 0.005$), Salmonella spp. ($p < 0.005$), ETEC ($p < 0.05$) and typical EPEC ($p < 0.005$)  
• Ct values were significantly higher for STEC cases vs. controls ($p < 0.05$)  
• No significant difference in Ct values between cases and controls were observed for EAEC or atypical EPEC |
|  | Salmonella spp | 32 |  |  |
|  | E. coli | 487 |  |  |
|  | ETEC | 56 |  |  |
|  | Typical EPEC | 20 |  |  |
|  | Atypical EPEC | 227 |  |  |
|  | STEC | 37 |  |  |
|  | EAEC | 127 |  |  |
|  | Shigella/EIEC | 14 |  |  |
| Elfving et al. JCM 2014 | Zanzibar, community, 2 months–5 years |  |  |  |
|  | Campylobacter | 112 | 31.8 | 0.12 |
|  | ETEC-eltB | 148 | 31.3 | 0.002 |
|  | ETEC-estA | 94 | 32.6 | 0.0001 |

*Continued*
### TABLE 3 | Continued

| Outcome | Study | Pathogen(s) | Number of PCR+ patients | Population | Outcome measure |
|---------|-------|-------------|--------------------------|------------|-----------------|
|    |       | Salmonella  | 13                       | Rwanda, community and hospital, ≤5 years | 42.2 40.6 0.22 |
|    |       | Shigella    | 113                      | Rwanda, community and hospital, ≤5 years | 29.2 34.5 < 0.0001 |
|    | Kabayiza et al. Pediatr Infect Dis J 2014a | Campylobacter   | 121                      | Rwanda, community and hospital, ≤5 years | Median Ct for patients 29.75 Median Ct for controls 33.02 p-value (PCR+) 0.007 |
|    |       | ETEC-eltB   | 213                      | Rwanda, community and hospital, ≤5 years | 33.91 34.15 0.90 |
|    |       | ETEC-estA   | 130                      | Rwanda, community and hospital, ≤5 years | 24.75 34.37 0.04 |
|    |       | EPEC-bfpA   | 68                       | Rwanda, community and hospital, ≤5 years | 33.74 33.00 0.52 |
|    |       | EPEC-eae    | 167                      | Rwanda, community and hospital, ≤5 years | 34.84 35.95 0.05 |
|    |       | Salmonella  | 48                       | Rwanda, community and hospital, ≤5 years | 41.41 40.70 0.23 |
|    |       | Shigella    | 90                       | Rwanda, community and hospital, ≤5 years | 30.35 33.99 0.10 |
|    | Barletta et al. CID 2011 | EPEC | 143 | Peru, community, ≤2 years | Median Ct for patients 33.02 Median Ct for controls 35.94 p-value (PCR+) 0.044 |
|    |       | Campylobacter spp. (ETEC) | 143 | Peru, community, ≤2 years | 29.75 33.02 0.007 |
|    |       | ETEC-eltB   | 213                      | Peru, community, ≤2 years | 33.91 34.15 0.90 |
|    |       | ETEC-estA   | 130                      | Peru, community, ≤2 years | 24.75 34.37 0.04 |
|    |       | EPEC-bfpA   | 68                       | Peru, community, ≤2 years | 33.74 33.00 0.52 |
|    |       | EPEC-eae    | 167                      | Peru, community, ≤2 years | 34.84 35.95 0.05 |
|    | PARASITES | Cryptosporidium | 69 | Rwanda, community and hospital, ≤5 years | 30.56 34.58 0.05 |
|    | Severity of symptoms | Kayabiza et al. Clin Microbiol and Infect 2014b | Cryptosporidium | 69 | Rwanda, community and hospital, ≤5 years | 30.56 34.58 0.05 |
|    | Barletta et al. CID 2011 | EPEC | 143 | Peru, community, ≤2 years | 30.35 33.99 0.10 |
|    | Duration of symptoms | Barletta et al. CID 2011 | EPEC | 143 | Peru, community, ≤2 years | 30.35 33.99 0.10 |
|    | Liu et al. Lancet 2016 | Cryptosporidium spp., Cyclospora cayetanensis, Entamoeba histolytica | 5,304 | Bangladesh, India, Pakistan, The Gambia, Kenya, Mal and Mozambique, community, ≤5 years | 29.66 34.58 0.05 |
|    | Bruijnehesten et al. Clin Microbiol Infect 2015 | C. parvum/ hominis | 56 | Netherlands, primary Care | 30.35 33.99 0.10 |
|    | Giardia lamblia | 118 | 30.35 33.99 0.10 |

#### Parasites

- **Cryptosporidium** spp., had strong quantity-dependent associations with diarrhea:
  - Cq values <29.1 were defined as “diarrhea-associated” and <24.0 were defined as “highly diarrhea-associated” (ROC cut-off 27.5; Youden index 0.17)
  - Cyclospora cayetanensis and Entamoeba histolytica showed associations with diarrhea:
    - For Cyclospora cayetanensis, Cq values <29.6 were defined as “highly diarrhea-associated” (ROC cut-off 26.9; Youden index 0.48)
  - For Entamoeba histolytica, Cq values <34.8 were defined as “diarrhea-associated” and <32.6 were defined as “highly diarrhea-associated” (ROC cut-off 26.9; Youden index 0.48)

- **C. parvum/hominis** had significantly higher relative loads were observed for C. parvum/hominis (p < 0.05)

- Higher loads were observed for G. lamblia, although statistical significance was not reached; p = 0.084

(Continued)
TABLE 3  | Continued

| Outcome | Study | Pathogen(s) | Number of PCR+ patients | Population | Outcome measure |
|---------|-------|-------------|-------------------------|------------|----------------|
| D. fragilis | Haque et al. Clin Infect Dis 2009 | 832 | Bangladesh, hospital | Patients: Mean Ct 41.0 (95% CI 37.5–44.5) | Case vs. control (p < 0.05) |
| Cryptosporidium parvum | | 20 | | Controls: Mean Ct 36.3 (29.4–43.1) | |
| Cryptosporidium hominis | | 61 | | Median Ct 43.2 | |
| Entamoeba histolytica D | | 83 | | Mean Ct 33.6 (95% CI 32.0–35.3) | |
| Giardia lamblia A | Forsell et al. Parasites and Vectors 2016 | 42 | Zanzibar, outpatients | Mean Ct 35.4 (95% CI 34.3–36.4) | |
| Giardia lamblia B | Elfving et al. JCM 2014 | 333 | Zanzibar, community, 2 months–5 years | Median Ct 35.8 | |
| Cryptosporidium | Kabayiza et al. Pediatr Infect Dis J 2014a | 23 | Rwanda, community and hospital, ≤5 years | Mean Ct 34.9 (95% CI 33.9–36.0) | |

CI, confidence interval; Cp, crossing point; Cq, quantification cycle; Ct, cycle threshold; EAEC, enteroaggregative E. coli; EIEC, enteroinvasive E. coli; EPEC, enteropathogenic E. coli; ETEC, enterotoxigenic E. coli; IV, intravenous; OR, odds ratio; PCR, polymerase chain reaction; qPCR, quantitative polymerase chain reaction; ROC, receiver operating characteristic; STEC, Shiga toxin-producing E. coli; UK, United Kingdom.

*Total number of matched pairs; individual pathogen PCR+ve n values were not provided, 2,254 samples were positive for one diarrhea-associated pathogen, and 2,063 samples were positive for ≥2.

Bold indicates a statistically significant association.

Discussion

The objective of this systematic review was to assess the global medical literature for any correlation between Ct values and clinical outcomes of patients with gastrointestinal infections. Lower Ct values correspond with greater quantities of detectible target gene and therefore a higher pathogen load, which may correspond with less favorable clinical outcomes. Here we report outcomes from studies identified that report on gastrointestinal pathogens only. This review gathers data from 33 studies, with the largest number of studies for C. difficile (n = 15). The most common outcomes reported were related to symptoms, including case vs. control, with vs. without diarrhea, and severity of symptoms.

Evidence in this review suggests associations between Ct values and symptomatic C. difficile infections. Four out of eight studies reporting the association between lower Ct values and increased disease severity found the association to be significant,
including two studies that reported lower Ct values as a predictor of poor outcome (14, 17). Furthermore, 2/4 case vs. control studies reported significantly lower Ct values in symptomatic cases. Most of the C. difficile studies reported genes encoding toxin A/B as the target for PCR diagnostics, which when detected by other methods, is generally inferred as marker of disease severity (5).

All studies of norovirus and rotavirus reported lower Ct values in cases vs. controls; the majority for norovirus GII and ~50% for rotavirus reported significant differences. Furthermore, two studies of rotavirus infections reported significant associations between lower Ct values and severity of symptoms, including vomiting, severe dehydration and administering intravenous fluids (27, 34). Notably, the association of Ct values and symptom severity was more pronounced for norovirus GII than norovirus GI (25, 27, 28). One possible explanation for this is the increased virulence observed with GII infection compared with other norovirus genogroups (41), although more investigation is necessary to draw firm conclusions.

This review found less evidence for the clinical utility of Ct values in non-C. difficile bacterial and parasitic infections compared with C. difficile and gastrointestinal viruses. Multiple studies reported significant associations between bacterial loads and symptomatic cases, particularly for Shigella (29, 30). Two studies reported Shigella association with symptom severity (27, 38). Inconsistencies were found in studies of parasitic infections; some studies indicated an association between low Ct values and symptomatic infection in patients with Cryptosporidium spp., however, evidence is limited (22, 29, 30). There is insufficient evidence to draw conclusions for other parasitic infections.

Among the studies included in this review, evidence suggests that Ct values may have utility in defining symptomatic causality, particularly in cases of polymicrobial infection. In one study of norovirus-positive samples, coinfection with rotavirus was observed in 3.7 and 7.4% of asymptomatic and diarrheal samples, respectively; probable etiology was determined based on relative Ct values, highlighting their utility for defining causative organisms in this setting (28). Ct values may also aid causative diagnosis in patients with C. difficile infection, where asymptomatic colonization (5, 6), and coinfections have been reported (42). C. difficile fecal load is already considered to be of diagnostic utility in distinguishing between infection and colonization (43, 44). However, it is essential to consider Ct values within the context of clinical presentation rather than utilize Ct values as an independent marker of disease.

Despite multiple studies reporting significant associations between high genomic load (low Ct values) and symptomatic infections, particularly for C. difficile, norovirus and rotavirus, statistically significant evidence was inconsistent across studies despite similar trends. A possible explanation for this is the diversity of populations investigated across each study (e.g., hospital vs. community setting, pediatric vs. adult populations); adjusting for similar settings may uncover stronger trends toward Ct value and patient outcomes. Further assessments of associations between Ct values and LOS, hospital/ICU admission, for example, could also aid in understanding the utility of Ct values in the diagnosis of gastrointestinal infections.

When interpreting the studies in this systematic review, consideration must be given to the settings and populations in which they were conducted. Studies for some pathogens, such as norovirus, were conducted primarily in pediatric populations and as such their conclusions may not apply to adult populations. All but one of the studies investigating non-C. difficile bacterial pathogens and parasites were performed in non-industrialized countries; therefore, the clinical impact of Ct values for these pathogens in industrialized countries remains to be determined. Of the seven studies that detected parasites, five investigated a large list of GI pathogens and multiple pathogens were detected for 8–72% patients (22, 27, 29, 30). These studies highlight the utility of syndromic testing in gastrointestinal infection, where multiplex testing is able to detect more pathogens and co-infections than conventional methods (42). It should also be noted that multiplex PCR for GI pathogens does not currently provide a picture of the microbiome, whereas culture-based techniques are able to provide an understanding of dysbiosis resulting from GI infections.

Differences in study methodology and qPCR workflow are likely to impact Ct values, including: specimen source, collection method, transport media type and volume, stability, quality of the sample, time of sampling vs. onset of infection, master mix components, type and concentration of passive reference dye, reaction efficiency, inter- and intravariability in assay platforms, and whether they were single or multiplex systems. Methodologies varied widely between studies and many (39%) had some or many gaps in reporting defined standardized methodologies. Therefore, within-study variability may have limited the ability to detect associations. The majority (84.8%; 28/33) of studies did not report normalized Ct values, which would have provided more accurate estimations of genomic load for each sample. Although outside the scope of our review, we noted not all (66.7%; 22/33) studies presented Ct value distributions. Further studies to understand the distribution of Ct values in relation to patient outcomes across the populations would be necessary if Ct values are to be utilized in clinical decision-making. After data analysis had been completed, we became aware of the Minimum Information for Publication of Quantitative rt-PCR Experiments (MIQE) guidelines (45), which should be applied to laboratory-developed tests. Some of the studies utilized in this review use commercially available assays and, therefore, when implemented in clinical diagnostic routines, applicable validation, and verification using external controls are necessary. Due to the late discovery it was not possible to re-assess the studies using laboratory-developed assays with the MIQE guidelines in mind; however, we believe that assessment of study methodology using these guidelines would not significantly alter the findings of this systematic review.

There were a number of limitations to this systematic review. The protocol restricted articles referring to Ct values as a measure of genomic load, therefore studies which reported genomic load in measures other than Ct value were not picked up in the database searches or excluded from during
screening. Furthermore, articles describing Ct values but with no mention of Ct values in the title, abstract or keywords, were not retrieved based on the search parameters used in the database searches. In addition, late in the review we became aware of alternative wording for Ct values, including Cq and “crossing point” [discussed in detail in (45)]; while we have added articles with these terms manually, it is possible that some may have been missed. Another limitation to this review was the assessment of all included studies as poor quality for bias by the Newcastle-Ottawa scale. This is due to the majority of studies reporting Ct values as secondary outputs, as opposed to seeking to compare clinical outcomes against Ct values. Consequently, the studies did not fully align with the risk and bias assessment. There was considerable variability between studies. Given the high heterogeneity between studies, it was not possible to conduct aggregated/meta-analyses, a key limitation in the scope of this review. A number of studies only made comparative analysis between symptomatic and asymptomatic cases, which limits the clinical utility of these studies in defining Ct values as a measure of disease severity. However, Ct values of asymptomatic patients still hold clinical value in order to discriminate between infection and colonization, an observation reported in multiple studies (4–6, 25, 27). A single reviewer conducted the data extraction and a second reviewer checked all the data points. Whilst an acceptable approach, the methodology could have been optimized by double independent reviewer data extraction with a third reviewer for discrepancy resolution. Due to the large number of studies identified as potential data sources for this review, the single-reviewer extraction method ensured that the review remained feasible. Despite these limitations, we believe this review provides insights into the potential clinical utility of gastrointestinal pathogen Ct values. In summary, there is evidence to support relationships between Ct values and clinical outcomes in gastrointestinal infections. Considered alongside clinical presentation, Ct values could help to guide treatment decisions, particularly in cases of C. difficile, where treatment is guided by severity of disease and asymptomatic colonization.
in COVID-19. 

11. Wells GA, Shea B, O’Connell D, Peterson J, Welch V, Losos M, Tugwell P. The Newcastle Ottawa Scale (NOS) for Assessing the Quality of Nonrandomised Studies in Meta-Analyses. Available online at: http://www.ohri.ca/programs/clinical_epidemiology/oxford (accessed September 03, 2021).

12. Rao K, Micic D, Natarajan M, Winters S, Kiel MJ, Walk ST, et al. Clostridium difficile ribotype 027: relationship to age, detectability of toxins A or B in stool with rapid testing, severe infection, and mortality. Clin Infect Dis. (2015) 61:233–41. doi: 10.1093/cid/civ254

13. Davies KA, Planche T, Wilcox MH. The predictive value of quantitative nucleic acid amplification detection of Clostridium difficile toxin gene for faecal sample toxin status and patient outcome. PLoS ONE. (2018) 15:e0205941. doi: 10.1371/journal.pone.0205941

14. Jazmati N, Hellmich M, Licanin B, Plum G, Kaasch AJ. PCR cycle threshold and severe infection. J Antimicrob Chemother. (2016) 72:170–7. doi: 10.1093/jac/dkv497

15. Anikst VE, Gaur RL, Schrofer LF, Banai N. Organism burden, toxin concentration, and lactoferrin concentration do not distinguish between clinically significant and nonsignificant diarrhea in patients with Clostridium difficile. Diagn Microbiol Infect Dis. (2016) 84:343–6. doi: 10.1016/j.diagmicrobio.2015.11.022

16. Reigadas E, Alcala L, Valerio M, Marin M, Martin A, Bouza E. Toxin B PCR cycle threshold as a predictor of poor outcome of Clostridium difficile infection: a derivation and validation cohort study. J Antimicrob Chemother. (2016) 71:1380–5. doi: 10.1093/jac/dkv497

17. Origue J, Orellana MA, Fernandez-Ruiz M, Corbella L, San Juan R, Ruiz-Ruigomez M, et al. Toxin B PCR amplification cycle threshold adds little to clinical variables for predicting outcomes in Clostridium difficile infection: a retrospective cohort study. J Clin Microbiol. (2019) 57:e01125–18. doi: 10.1128/JCM.01125-18

18. Bab F, Chatigre JK, Yao JA, Coulibaly JT, Von Mueller L, Polman M, et al. Markers of intestinal inflammation, not bacterial burden, correlate with clinical outcomes in Clostridium difficile infection. Clin Infect Dis. (2013) 56:1713–21. doi: 10.1093/cid/cit147

19. El Feghaly RE, Stauber JL, Tarr PI, Haslam DB. Norovirus prevalence and estimated viral load in symptomatic and asymptomatic children from rural communities of Vhembe district, South Africa. J Clin Virol. (2016) 84:122–8. doi: 10.1016/j.jcv.2016.09.005

20. Dung TT, Phat VV, Nga TV, My PV, Duy PT, Campbell JL, et al. The validation and utility of a quantitative one-step multiplex RT real-time PCR targeting rotavirus A and norovirus. J Virol Methods. (2013) 187:138–43. doi: 10.1016/j.viromet.2012.09.021

21. Kabayiza JC, Andersson ME, Nilsson S, Bergstrom T, Muhirwa G, Lindh M. Real-time PCR identification of agents causing diarrhea in Rwandan children less than 5 years of age. Pediatr Infect Dis J. (2014) 33:1037–42. doi: 10.1097/INF.0000000000000448

22. Saito M, Goel-Apaza S, Espetia S, Velasquez D, Cabrera L, Loli S, et al. Multiple norovirus infections in a birth cohort in a Peruvian Peruvian community. Clin Infect Dis. (2014) 58:883–91. doi: 10.1093/cid/cit763

23. Liu J, Platts-Mills JA, Juma I, Kabir F, Nkeze J, Okoi C, et al. Use of quantitative molecular diagnostic methods to identify causes of diarrhoea in children: a reanalysis of the GEMS case-control study. Lancet. (2016) 388:1291–301. doi: 10.1016/S0140-6736(16)31529-X

24. Elving K, Andersson M, Msellem MI, Welinder-Olsson C, Petzold M, Bjorkman A, et al. Real-time PCR threshold cycle cutoffs help to identify agents causing acute childhood diarrhea in Zanzibar. J Clin Microbiol. (2014) 52:916–23. doi: 10.1128/JCM.02697-13

25. Gustavsson L, Westin J, Lindh M, Andersson LM. Faecal viral load does not predict short-term mortality in norovirus infection. J Clin Virol. (2015) 70:557. doi: 10.1016/j.jcv.2015.07.135

26. Partridge DG, Evans CM, Raia M, Kudesia G, Parsons HK. Lessons from a large norovirus outbreak: impact of viral load, patient age and ward design on duration of symptoms and shedding and likelihood of transmission. J Hosp Infect. (2012) 81:25–30. doi: 10.1016/j.jhin.2012.02.002

27. Phillips G, Lopman B, Tam CC, Iturriza-Gomara M, Brown D, Gray J. Diagnosing rotavirus A associated ID: Using ELISA to identify a cut-off for real time RT-PCR. J Clin Virol. (2009) 44:242–5. doi: 10.1016/j.jcv.2008.12.001

28. Keng G, Iturriza-Gomara M, Wheeler JG, Crystal P, Monica B, Ramani S, et al. Quantitation of group A rotavirus by real-time reverse-transcription-polymerase chain reaction: correlation with clinical severity in children in South India. J Med Virol. (2004) 73:118–22. doi: 10.1002/jmv.20053

29. Ramani S, Sankaran P, Arumugam R, Sarkar R, Banerjee I, Mohanty I, et al. Comparison of viral load and duration of virus shedding in symptomatic and asymptomatic neonatal rotavirus infections. J Med Virol. (2010) 82:1803–7. doi: 10.1002/jmv.21872

30. Mukhopadhyay I, Sarkar R, Menon VK, Babji S, Paul A, Rajendran P, et al. Rotavirus shedding in symptomatic and asymptomatic children using reverse transcription-quantitative PCR. J Med Virol. (2013) 85:1661–8. doi: 10.1002/jmv.23641

31. Barletta F, Ochoa TJ, Mercado E, Ruiz J, Ecker L, Lopez G, et al. Quantitative real-time polymerase chain reaction for enteropathogenic Escherichia coli: a tool for investigation of asymptomatic versus symptomatic infections. Clin Infect Dis. (2011) 53:1223–9. doi: 10.1093/cid/cir730

32. Vu DT, Sethabutr O, Von Seidlein L, Tran VT, Do GC, Bui TC, et al. Detection of Shigella by a PCR assay targeting the ipaH gene suggests increased prevalence of shigellosis in Nha Trang, Vietnam. J Clin Microbiol. (2004) 42:2031–5. doi: 10.1128/JCM.42.10.2031-2035.2004

33. Haque R, Mondal D, Karim A, Molla IH, Rahim A, Faruque AS, et al. Prospective case-control study of the association between common enteric protozoal parasites and diarrhoea in Bangladesh. Clin Infect Dis. (2009) 48:1191–7. doi: 10.1086/597580

34. Forsell J, Granlund M, Samelsson L, Koskimetsi S, Edebro H, Evengard B. High occurrence of Blastocystis sp. subtypes 1-3 and Giardia intestinalis assemblage B among patients in Zanzibar. Tanzania Parasit Vectors. (2016) 9:370. doi: 10.1186/s13071-016-1637-8

35. Huhti L, Szakal ED, Puustinen L, Salminen M, Huhtala H, Valve O, et al. Real-time PCR identification of agents causing diarrhea in Zanzibar. J Clin Virol. (2013) 52:916–23. doi: 10.1016/j.jcv.2012.09.021
immunoassay and cell culture cytotoxicity assay. J Clin Microbiol. (2013) 51:3624–30. doi: 10.1128/JCM.01444-13

45. Bustin SA, Benes V, Garson JA, Hellemans J, Huggett J, Kubista M, et al. The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. Clin Chem. (2009) 55:611–22. doi: 10.1373/clinchem.2008.112797

46. Public Health England. Updated Guidance on the Management and Treatment of Clostridium difficile Infection. (2013). Available online at: https://www.gov.uk/government/publications/clostridium-difficile-infection-guidance-on-management-and-treatment (accessed March 31, 2021).

Conflict of Interest: JP, SR, and DM are employed by Qiagen. BV reports grants, personal fees and non-financial support from Qiagen, personal fees and non-financial support from BioMérieux, personal fees from Hologic, personal fees from Gilead, outside the submitted work. DB reports personal fees from Qiagen, outside the submitted work.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The authors declare that this study received funding from Qiagen Manchester Ltd. The funder had the following involvement in the study: article processing fees and provision of medical writing support. The funder was not involved in the study design, collection, analysis, interpretation of data, the writing of this article or the decision to submit it for publication.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Bonacorsi, Visseaux, Bouzid, Pareja, Rao, Manissero, Hansen and Vila. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.