Research Article

The Association between Platelet Glycocalcin and High Microsatellite Instability in Colorectal Cancer

Zeng-yao Liu,1,2 Qing-chun Jia,1 Wen Wang,1 Yu-xi Liu,1 Rui-tao Wang,1 and Jia-yu Li3

1Department of Internal Medicine, Harbin Medical University Cancer Hospital, Harbin Medical University, Harbin, Heilongjiang 150081, China
2Department of Interventional Medicine, The First Affiliated Hospital, Harbin Medical University, Harbin, Heilongjiang 150001, China
3Institute of Intensive Care Medicine, Heilongjiang Academy of Medical Science, Harbin, Heilongjiang 150081, China

Correspondence should be addressed to Rui-tao Wang; ruitaowang@126.com and Jia-yu Li; jiayu93@outlook.com

Received 15 January 2022; Accepted 23 March 2022; Published 7 April 2022

Academic Editor: Eiji Sakai

Copyright © 2022 Zeng-yao Liu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Elevated platelet volume is the risk factor for the development and poor overall survival of colorectal cancer (CRC) patients. Both microsatellite status and platelet glycoprotein Ibα (GPIbα) are related to platelet volume in CRC patients. This study aimed to investigate platelet GPIbα ectodomain (termed glycocalcin) levels among CRC patients and the association between the glycocalcin levels and microsatellite status in CRC. Methods. The clinical and laboratory data of 430 CRC patients between January 2018 and December 2018 in Harbin Medical University Cancer Hospital were collected. The participants were separated into high microsatellite instability (MSI-H) and microsatellite stable (MSS) groups according to microsatellite status. The glycocalcin levels were measured with an enzyme-linked immunosorbent assay, and the cut-off point was determined with the receiver-operating characteristics curve. The clinical and pathological characteristics were collected via electronic medical records. Logistic regression was used to explore the association between glycocalcin and microsatellite status. Results. Among the 430 CRC patients enrolled, 64 patients (14.9%) were identified as MSI-H and others as MSS CRC. Glycocalcin levels were significantly reduced in patients with MSI-H than those with MSS. After controlling for potential confounders, logistic regression analysis revealed that glycocalcin levels were independently associated with MSI-H CRC. Conclusions. Reduced glycocalcin levels are associated with the MSI-H subtype of CRC. Further research is needed to elucidate the mechanisms of the association between glycocalcin and MSI-H in CRC patients.

1. Introduction

The occurrence of colorectal cancer involves the gradual accumulation and malignant transformation of various genetic changes, which are conducive to the proliferation and growth of tumor cells [1]. Microsatellite instability (MSI) results from DNA mismatch repair system defects, characterized by a significant increase in the intragenic mutation rate of short tandem repeat DNA sequences called microsatellites [2]. CRC tumors can be classified into high MSI (MSI-H), low MSI (MSI-L), and microsatellite stable (MSS) according to their mutation patterns and the proportion of markers showing MSI [1]. Approximately 15% of CRC patients demonstrate MSI-H tumors [3]. Patients with MSI-H CRC generally have a better prognosis and a more effective immune response than patients with MSS CRC [4, 5]. Glycoprotein (GP) Ib is a complex formed by the linking of GPIbα with two GPIbβ subunits through a membrane-
proximal disulfide bond. The number of GPIb on the platelet surface is strongly associated with platelet volume [6]. GPIb receptors are more abundant in larger platelets [5]. As a specific platelet adhesion receptor, GPIbα is an essential platelet function regulator, regulating the survival and clearance of platelets [7, 8]. The amount of GPIbα could induce signals to enter platelets across the membrane by binding to the Willebrand factor (VWF), leading to platelet activation and subsequent platelet aggregation and thrombosis [9]. GPIbα releases glycocalicin (extracellular domain of GPIbα) into the plasma after binding with VWF [10–13].

Both MSI-H subtype and amount of GPIb are related to the platelet size. The levels of glycocalicin depend on the levels of GPIbα. However, there is limited evidence to show the association between glycocalicin and microsatellite status among CRC patients. This study aimed to examine glycocalicin levels in CRC patients and investigate the association between glycocalicin levels and MSI status.

2. Material and Methods

2.1. Study Population. This study reviewed 430 CRC patients at the Harbin Medical University Cancer Hospital from January 2018 to December 2018 via an online medical system. CRC was confirmed with a pathological diagnosis. Patients were excluded under the following conditions: (1) with infection, autoimmune disease, hematological disorders, hypertension, and diabetes mellitus; (2) with a history of cancer other than CRC; (3) with a history of chemotherapy or radiotherapy or medical treatment with acetylic salicylic acid; and (4) with insufficient information.

2.2. Clinical Examination and Biochemical Measurements. Clinicopathological information was collected via the medical records: age, gender, body mass index (BMI), carcioembyronic antigen (CEA), smoking status, drinking status, albumin, creatinine, white blood cells (WBC), hemoglobin, platelet count, tumor location, tumor size, histological type, histological grade, venous invasion, perineural invasion, T classification, lymph node metastasis, and distant metastasis. Two pathologists independently evaluated the pathological tumor stage based on the 8th edition of the American Joint Committee on Cancer (AJCC) cancer staging system. Another pathological expert in CRC was invited to decide if the diagnosis was inconsistent between these two pathologists. Venous blood samples were collected from all participants with sodium citrate tubes under fasting conditions before any treatment. Each blood sample was centrifuged at 2,500 rpm for 10 minutes, and the supernatant was kept frozen at -80°C until assayed. Routine blood tests were conducted in the hospital’s clinical laboratory. Glycocalicin was measured with an enzyme-linked immunosorbent assay (ELISA) kit (CUSABIO, Wuhan, China) based on the manufacturer’s instructions. Each sample was tested in duplicate. The intra- and inter-assay variations were below 8%.

2.3. MSI Analysis. MSI was evaluated with DNA obtained from freshly frozen tumor tissue samples, using polymerase chain reaction and amplified microsatellite-labeled primers, including BAT25, BAT26, NR-21, NR-24, and NR-27. The MSI-H was defined as at least three of the five markers being unstable, or it was described as MSS when the unstable markers were fewer than three. No sample in this study had only two unstable markers.

2.4. Statistical Analysis. The data were presented as the mean ± standard deviation (SD) for normally distributed data, the median (interquartile range) for not normally distributed data, and frequencies for categorical data. Normally distributed continuous variables in the two groups were compared with Student’s t test and skewed-distributed with the Mann-Whitney U test. The chi-square test was used for categorical variables. Binary logistic regression was used to assess clinicopathological factors associated with MSI-H status, reporting an adjusted odds ratio with 95% confidence interval (AOR, 95% CI). The statistical analyses were conducted using SPSS Statistics version 25.0 (SPSS Inc., Chicago, IL, USA). Receiver-operating characteristics (ROC) curve analysis was performed to calculate the area under the curve (AUC) and evaluate the optimal cut-off point of glycocalicin using MedCalc version 15.0. Differences were considered significant when \( p < 0.05 \).

3. Results

The median value of glycocalicin among all patients was 26.5 (range, 11.7-51.5). According to the MSI-H, the optimal cut-off value of glycocalicin was determined with ROC analysis (Figure 1). Specificity was 45.3% and sensitivity was 92.6% (AUC = 0.708, 95% CI: 0.632-0.783, \( p < 0.001 \)). According to the cut-off value (20.5 ng/mL), there were 374 patients (87.0%) in the group with glycocalicin levels >20.5 ng/mL and 56 patients (13.0%) in the group with glycocalicin levels ≤20.5 ng/mL. Baseline characteristics were compared between the two groups, and only the MSI status was significantly different with \( p < 0.001 \) (Table 1).
Table 1: Baseline characteristics of CRC patients according to glycocalcin levels.

| Variables                          | Total | Glycocalcin ≤20.5 ng/mL | Glycocalcin >20.5 ng/mL | p value |
|------------------------------------|-------|-------------------------|-------------------------|---------|
| Age (years)                        |       |                         |                         |         |
| ≤65                                | 309 (71.9) | 42 (75.0)               | 267 (71.4)              | 0.575   |
| >65                                | 121 (28.1) | 14 (25.0)               | 107 (28.6)              |         |
| Gender (%)                         |       |                         |                         | 0.581   |
| Male                               | 245 (57.0) | 30 (53.6)               | 215 (57.5)              |         |
| Female                             | 185 (43.0) | 26 (46.4)               | 159 (42.5)              |         |
| BMI (kg/m²)                        | 23.4 ± 3.2 | 23.9 ± 3.6              | 23.3 ± 3.2              | 0.199   |
| Current smoker (%)                 |       |                         |                         | 0.809   |
| Yes                                | 183 (42.6) | 23 (41.1)               | 160 (42.8)              |         |
| No                                 | 247 (57.4) | 33 (58.9)               | 214 (57.2)              |         |
| Drinker (%)                        |       |                         |                         | 0.375   |
| Yes                                | 139 (32.3) | 21 (37.5)               | 118 (31.6)              |         |
| No                                 | 291 (67.7) | 35 (62.5)               | 256 (68.4)              |         |
| WBC (×10⁹/L)                      | 7.09 ± 2.44 | 7.23 ± 2.43            | 7.07 ± 2.45             | 0.657   |
| Hemoglobin (g/L)                   | 132.9 ± 23.4 | 128.0 ± 27.0           | 133.6 ± 22.8            | 0.138   |
| Platelet count (×10⁹/L)            | 269.7 ± 87.8 | 278.5 ± 96.1         | 268.3 ± 86.6            | 0.417   |
| Creatinine (µmol/L)                | 81.1 ± 18.6 | 80.4 ± 12.5           | 81.2 ± 19.3             | 0.790   |
| CEA (ng/mL)                        | 4.37 (2.03-11.31) | 3.49 (1.86-12.04) | 4.52 (2.12-11.07)      | 0.499   |
| Tumor size (cm)                    |       |                         |                         | 0.208   |
| <5.0                               | 278 (64.7) | 32 (57.1)               | 246 (65.8)              |         |
| ≥5.0                               | 152 (35.3) | 24 (42.9)               | 128 (34.2)              |         |
| Tumor location (%)                 |       |                         |                         | 0.051   |
| Proximal                           | 157 (36.5) | 27 (48.2)               | 130 (34.8)              |         |
| Distal                             | 273 (63.5) | 29 (51.8)               | 244 (65.2)              |         |
| Histological type (%)              |       |                         |                         | 0.756   |
| Non-mucinous                       | 307 (71.4) | 39 (69.6)               | 268-71.7                |         |
| Mucinous                           | 123 (28.6) | 17 (30.4)               | 106 (28.3)              |         |
| Histological grade (%)             |       |                         |                         | 0.147   |
| Well/moderately differentiated     | 359 (83.5) | 43 (76.8)               | 316 (84.5)              |         |
| Poorly differentiated              | 71 (16.5)  | 13 (23.2)               | 58 (15.5)               |         |
| Lymphatic invasion (%)             |       |                         |                         | 0.185   |
| Absent                             | 331 (77.0) | 47 (83.9)               | 284 (75.9)              |         |
| Present                            | 99 (23.0)  | 9 (16.1)                | 90 (24.1)               |         |
| Perineural invasion (%)            |       |                         |                         | 0.545   |
| Absent                             | 365 (84.9) | 48 (85.7)               | 317 (84.8)              |         |
| Present                            | 65 (15.1)  | 8 (14.3)                | 57 (15.2)               |         |
| T classification (%)               |       |                         |                         | 0.372   |
| T1+T2                              | 63 (14.7)  | 6 (10.7)                | 57 (15.2)               |         |
| T3+T4                              | 367 (85.3) | 50 (89.3)               | 317 (84.8)              |         |
| Lymph node metastasis (%)          |       |                         |                         | 0.511   |
| Absence                            | 267 (62.1) | 37 (66.1)               | 230 (61.5)              |         |
| Presence                           | 163 (37.9) | 19 (33.9)               | 144 (38.5)              |         |
| Distant metastasis (%)             |       |                         |                         | 0.426   |
| Absence                            | 382 (88.8) | 48 (85.7)               | 334 (89.3)              |         |
| Presence                           | 48 (11.2)  | 8 (14.3)                | 40 (10.7)               |         |
| Stage (%)                          |       |                         |                         | 0.682   |
| I-II                               | 258 (60.0) | 35 (62.5)               | 223 (59.6)              |         |
| III-IV                             | 172 (40.0) | 21 (37.5)               | 151 (40.4)              |         |
All CRC patients were classified into quartiles according to their glycocalicin levels, including Q1 ≤ 23.2 ng/mL, 23.2 ng/mL < Q2 ≤ 26.3 ng/mL, 26.3 ng/mL < Q3 ≤ 29.6 ng/mL, and Q4 > 29.6 ng/mL (Figure 2). The percentages of patients with MSI-H in each group were 29.0%, 14.5%, 9.4%, and 6.5%, respectively. The results showed that elevated glycocalicin levels were associated with MSI-H (p < 0.001).

The baseline characteristics of 430 participants were compared between the MSI-H and MSS groups in Table 2. Patients in the MSI-H group were younger (p < 0.05) and had higher BMI (p < 0.005) and levels of WBC (p < 0.005) than those in the MSS group. Lower levels of glycocalicin levels (p < 0.001), hemoglobin (p < 0.05), and CEA (p < 0.05) were found in the MSI-H group. Tumor characteristics, including tumor location, tumor size, lymphatic invasion, lymph node metastasis, clinical stage, and histological type, were significantly different between the MSI-H and MSS groups (p < 0.05). The final logistic regression model included all significant variables in the univariate analysis. Patients with a higher BMI, higher WBC levels, tumor size 5.0 cm, proximal CRC, and mucinous type were more likely to have MSI-H. Notably, decreased glycocalicin levels were associated with MSI-H after adjusting for other confounding variables (AOR, 0.854; 95% CI, 0.801-0.910; p < 0.001) (Table 3).

4. Discussion

Glycocalicin levels were significantly reduced in MSI-H CRC patients compared with MSS CRC patients. Glycocalicin levels were strongly associated with MSI-H status in CRC patients after controlling for potential confounders.

MSI status is a critical factor in the pathogenesis of CRC. It could also be used as an essential molecular marker for the prognosis and adjuvant therapy of CRC [14, 15]. MSI-H tumors are more likely to be mucinous and poorly differentiated [1]. Regardless of the depth of tumor invasion, CRC with the MSI-H phenotype is unlikely to spread to regional lymph nodes or distant organs [16, 17]. MSI-H CRC responds well to immunotherapy [18]. Age under 50 is a strong predictor of MSI, and younger patients are more likely to have MSI-H CRC [19]. In our study, patients in the MSI-H group had a younger age than those in the MSS group. In our study, patients with proximal CRC and mucinous type CRC tend to be MSI-H type, which also consists of the current evidence. MSI-H is more common in proximal colon cancer than in distal colon cancer [20, 21].

The exact mechanisms of decreased glycocalicin in MSI-H CRC are still unclear. Mean platelet volume (MPV) refers to platelet average size and indicates platelet activation [4, 5]. Activated platelets can cause tumor proliferation, angiogenesis, and increase tumor adhesion, promoting tumor development and metastasis [22]. Lower MPV levels are independently associated with better overall survival of CRC [23]. Moreover, the decreased MPV levels are associated with MSI-H CRC, and MSI-H CRC patients exhibited a better prognosis than MSI-L CRC patients [4, 24]. The evidence about the role of GPIb or glycocalicin levels in the occurrence or prognosis of CRC is limited. GPIb receptors are expressed in lower numbers in smaller platelets [5, 6].

---

**Table 1: Continued.**

| Variables | Total | Glycocalicin ≤20.5 ng/mL | Glycocalicin >20.5 ng/mL | p value |
|-----------|-------|--------------------------|--------------------------|---------|
| MSI status (%) |       |                          |                          | < 0.001 |
| MSS       | 366 (85.1) | 27 (48.2) | 339 (90.6) |         |
| MSI-H     | 64 (14.9)  | 29 (51.8)  | 35 (9.4)    |         |

Data are presented as means (standard deviation), median (interquartile range), or number (percentage). BMI: body mass index; WBC: white blood cells; CEA: carcinoembryonic antigen; MSS: microsatellite stable; MSI: microsatellite instability; MSI-H: high MSI.

---

**Figure 2:** The association between high microsatellite instability and glycocalicin levels among colorectal cancer patients.
As mentioned before, glycocalicin is the extracellular domain released by GPIbα. Hence, glycocalicin levels fluctuate based on GPIbα levels. This consistent evidence suggests that platelet volume might play a role in the association between MSI-H and glycocalicin in CRC patients.

| Variables                   | Total   | MSI-H   | MSS     | p value |
|-----------------------------|---------|---------|---------|---------|
| Number                      | 430     | 64      | 366     |         |
| Age (years)                 | 59.4±10.0 | 56.4±11.8 | 59.9±9.6 | 0.029   |
| Gender (female, %)          | 181(42.1) | 33(51.6)  | 148(40.4) | 0.096   |
| BMI (kg/m²)                 | 23.4±3.2 | 24.5±3.3 | 23.2±3.2 | 0.004   |
| Current smoker (%)          | 183(42.6) | 25(39.1)  | 158(43.2) | 0.540   |
| Drinker (n, %)              | 139(32.3) | 17(26.6)  | 122(33.3) | 0.285   |
| Creatinine (µmol/L)         | 81.1±18.6 | 80.9±19.7 | 81.1±18.4 | 0.929   |
| CEA (ng/mL)                 | 3.14 (1.57-7.98) | 3.14 (1.57-7.98) | 4.86 (2.15-12.12) | 0.030   |
| WBC (×10⁹/L)                | 7.09±2.44 | 8.07±2.98 | 6.92±2.30 | 0.004   |
| Hemoglobin (g/L)            | 132.9±23.4 | 124.6±27.3 | 134.3±22.4 | 0.009   |
| Platelet count (×10⁹/L)     | 269.7±87.8 | 286.2±112.6 | 266.8±82.6 | 0.103   |
| Glycocalcin (ng/mL)         | 26.5±6.0  | 22.6±6.3  | 27.2±5.6  | <0.001  |

Data are presented as means (standard deviation), median (interquartile range), or number (percentage). Abbreviations see Table 1.
followed by the increased glycocalicin content in plasma [7]. Increased VWF is a risk factor for venous thromboembolism and long-term venous complications [27]. Microsatellite has been reported as a part of the VWF promoter region and promotes glucocorticoid-induced VWF levels [28]. Thus, VWF might be associated with our current findings. However, further study is required to investigate the association between MSI status and VWF in CRC patients.

Several limitations in this study should be considered. Our results were sourced from a single hospital. Only Chinese participants were included, so the results need to be confirmed in other ethnic populations. Further study with a potential mechanistic explanation is required.

5. Conclusions

Reduced glycocalicin is associated with the MSI-H subtype of CRC. Further mechanistic research is needed to explain the findings, which might help guide management strategies in CRC patients.

Abbreviations

| Abbreviation | Definition |
|--------------|------------|
| MPV:         | Mean platelet volume |
| CRC:         | Colorectal cancer |
| GPIbα:       | Glycoprotein Ibα |
| MSI:         | Microsatellite instability |
| MSI-H:       | High MSI |
| MSI-L:       | Low MSI |
| MSS:         | Microsatellite stable |
| VWF:         | Willebrand factor |
| BMI:         | Body mass index |
| CEA:         | Carcinoembryonic antigen |
| WBC:         | White blood cells |
| AJCC:        | American Joint Committee on Cancer |
| SD:          | Standard deviation |
| AOR:         | Adjusted odds ratio |
| CI:          | Confidence interval |
| AUC:         | Area under the curve |

Data Availability

The datasets generated and analyzed during the current study are not publicly available due to data protection regulations concerning patient information, but are available from the corresponding author upon reasonable request.

Ethical Approval

The present study protocol was reviewed and approved by the Institutional Review Board of Harbin Medical University Cancer Hospital (approval no. 2017-0103).

Consent

All patients gave their informed consent for participation in the study.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors' Contributions

Zeng-yao Liu and Qing-chun Jia proposed and designed the study; Wen Wang and Yu-xi Liu collected the data; Rui-tao Wang and Jia-yu Li analyzed the data, interpreted the results, and drafted the article. Every author reviewed the draft and approved the final version of the manuscript for publishing. Zeng-yao Liu and Qing-Chun Jia contributed equally to this work.

Acknowledgments

I wish to extend my special thanks to all the pathologists in the Department of Pathology, Harbin Medical University Cancer Hospital. This work was supported by grants from the National Cancer Center Climbing Foundation (NCC201908B095).
References

[1] M. Kreidieh, D. Mukherji, S. Temraz, and A. Shamseddine, “Expanding the scope of immunotherapy in colorectal cancer: current clinical approaches and future directions,” *BioMed Research International*, vol. 2020, Article ID 9037217, 24 pages, 2020.

[2] R. Ward, A. Meagher, I. Tomlinson et al., “Microsatellite instability and the clinicopathological features of sporadic colorectal cancer,” *Gut*, vol. 48, no. 6, pp. 821–829, 2001.

[3] Z. Gatalica, S. Vranic, J. Xiu, J. Swensen, and S. Reddy, “High microsatellite instability (MSI-H) colorectal carcinoma: a brief review of predictive biomarkers in the era of personalized medicine,” *Familial Cancer*, vol. 15, no. 3, pp. 405–412, 2016.

[4] W. Wang, G. Wang, S. Fu, B. Zhang, Z. Liu, and R. Wang, “Decreased mean platelet volume is associated with microsatellite instability in colorectal cancer: a propensity score-matched analysis,” *Cancer Biomarkers*, vol. 31, no. 4, pp. 351–359, 2021.

[5] E. Varol, M. Ozaydin, Y. Turker, and S. Alaca, “Mean platelet volume, an indicator of platelet activation, is increased in patients with mitral stenosis and sinus rhythm,” *Scandinavian Journal of Clinical and Laboratory Investigation*, vol. 69, no. 6, pp. 708–712, 2009.

[6] S. G. Khaspekova, I. T. Zyuryaev, V. V. Yakushkin et al., “Relationships of glycoproteins Ibα–IIIα and Ib content with mean platelet volume and their genetic polymorphisms,” *Blood Coagulation & Fibrinolysis*, vol. 25, no. 2, pp. 128–134, 2014.

[7] Y. Cadroy, S. R. Hanson, A. B. Kelly et al., “Relative antithrombotic effects of monoclonal antibodies targeting different platelet glycoprotein-adhesive molecule interactions in nonhuman primates,” *Blood*, vol. 83, no. 11, pp. 3218–3224, 1994.

[8] K. M. Hoffmeister, T. W. Felbinger, H. Falet et al., “The clearance mechanism of chilled blood platelets,” *Cell*, vol. 112, no. 1, pp. 87–97, 2003.

[9] Y. Tao, X. Zhang, X. Liang, J. Zang, X. Mo, and R. Li, “Structural basis for the specific inhibition of glycoprotein Ibα shedding by an inhibitory antibody,” *Scientific Reports*, vol. 6, no. 1, p. 24789, 2016.

[10] E. E. Gardiner, D. Karunakaran, Y. Shen, J. F. Arthur, R. K. Andrews, and M. C. Berndt, “Controlled shedding of platelet glycoprotein (GP)VII and GPIb–IX–V by ADAM family metallopeptidases,” *Journal of Thrombosis and Haemostasis*, vol. 5, no. 7, pp. 1530–1537, 2007.

[11] F. Denorme, K. Vanhoorebeke, and S. F. De Meyer, “von Willebrand factor and platelet glycoprotein Ib: a thromboinflammatory axis in stroke,” *Frontiers in Immunology*, vol. 10, p. 2884, 2019.

[12] R. Yan, X. Mo, A. M. Paredes et al., “Reconstitution of the platelet glycoprotein Ib-IX complex in phospholipid bilayer Nanodiscs,” *Biochemistry*, vol. 50, no. 49, pp. 10598–10606, 2011.

[13] A. Amelirad, K. Shamsasesanj, P. Akbarzadehlahle, and S. D. Pashoutan, “Signaling pathways of receptors involved in platelet activation and shedding of these receptors in stored platelets,” *Adv Pharm Bull.*, vol. 9, no. 1, pp. 38–47, 2019.

[14] S. Hasan, P. Renz, R. E. Wegner et al., “Microsatellite instability (MSI) as an independent predictor of pathologic complete response (PCR) in locally advanced rectal cancer: a National Cancer Database (NCDB) analysis,” *Annals of Surgery*, vol. 271, no. 4, pp. 716–723, 2020.

[15] M. Hewish, C. J. Lord, S. A. Martin, D. Cunningham, and A. Ashworth, “Mismatch repair deficient colorectal cancer in the era of personalized treatment,” *Nature Reviews. Clinical Oncology*, vol. 7, no. 4, pp. 197–208, 2010.

[16] A. Malesci, L. Laghi, P. Bianchi et al., “Reduced likelihood of metastases in patients with microsatellite-unstable colorectal cancer,” *Clinical Cancer Research*, vol. 13, no. 13, pp. 3831–3839, 2007.

[17] R. Gryfe, H. Kim, E. T. Hsieh et al., “Tumor microsatellite instability and clinical outcome in young patients with colorectal cancer,” *The New England Journal of Medicine*, vol. 342, no. 2, pp. 69–77, 2000.

[18] G. Golshani and Y. Zhang, “Advances in immunotherapy for colorectal cancer: a review,” *Therapeutic Advances in Gastroenterology*, vol. 13, p. 175628482091752, 2020.

[19] J. K. Greenson, S. C. Huang, C. Herron et al., “Pathologic predictors of microsatellite instability in colorectal cancer,” *The American Journal of Surgical Pathology*, vol. 33, no. 1, pp. 126–133, 2009.

[20] F. A. Sinicrope, R. L. Rego, K. C. Halling et al., “Prognostic impact of microsatellite instability and DNA ploidy in human colon carcinoma patients,” *Gastroenterology*, vol. 131, no. 3, pp. 729–737, 2006.

[21] M. Yamauchi, T. Morikawa, A. Kuchiba et al., “Assessment of colorectal cancer molecular features along bowel subsites challenges the conception of distinct dichotomy of proximal versus distal colorectum,” *Gut*, vol. 61, no. 6, pp. 847–854, 2012.

[22] H. A. Goubran, T. Burnouf, M. Radosevic, and M. El-Ekiaby, “The platelet-cancer loop,” *European Journal of Internal Medicine*, vol. 24, no. 5, pp. 393–400, 2013.

[23] J. Y. Li, Y. Li, Z. Jiang, R. T. Wang, and X. S. Wang, “Elevated mean platelet volume is associated with presence of colon cancer,” *Asian Pacific Journal of Cancer Prevention*, vol. 15, no. 23, pp. 10501–10504, 2014.

[24] S. Kang, Y. Na, S. Y. Joung, S. I. Lee, S. C. Oh, and B. W. Min, “The significance of microsatellite instability in colorectal cancer after controlling for clinicopathological factors,” *Medicine (Baltimore)*, vol. 97, no. 9, article e0019, 2018.

[25] Y. Qi, W. Chen, X. Liang et al., “Novel antibodies against GPIbα inhibit pulmonary metastasis by affecting vWF-GPIbα interaction,” *Journal of Hematology & Oncology*, vol. 11, no. 1, p. 117, 2018.

[26] M. Malehmir, D. Pfister, S. Gallage et al., “Platelet GPIbα is a mediator and potential interventional target for NASH and subsequent liver cancer,” *Nature Medicine*, vol. 25, no. 4, pp. 641–655, 2019.

[27] A. Michels, D. Lillicrap, and M. Yacob, “Role of von Willebrand factor in venous thromboembolic disease,” *JVS Vasc Sci.*, vol. 3, pp. 17–29, 2022.

[28] V. Daidone, E. Pontara, C. Romualdi et al., “Microsatellite (GT)n is part of the von Willebrand factor (VWF) promoter region that influences the glucocorticoid-induced increase in VWF in Cushing’s syndrome,” *Thrombos Research*, vol. 125, no. 6, pp. e275–e280, 2010.