Alpha Thalassemia Genotypes in Kuwait

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Abstract

Background: The frequency of the alpha thalassemia trait is approximately 40% in the Kuwaiti population, but there has been no comprehensive study of the prevalent alleles. This is a report of patients who were referred for molecular diagnosis over a 20-year period.

Methods: This is a retrospective study of the a-globin genotypes obtained in the Hemoglobin Research Laboratory of the Department of Pediatrics, Kuwait University from 1994 to 2015. Genotyping was performed by a combination of PCR, allele-specific oligonucleotide hybridization and reverse dot blot hybridization (Vienna Lab Strip Assay).

Results: Four hundred samples were characterized and analyzed from individuals aged <1 month to 80 years, with a median of 6 years from 283 unrelated families. Most (90.8%) were Kuwaiti nationals. The commonest genotype was homozygosity for the polyadenylation-1 mutation (αPA-1/αPA-1) in 33.3% of the samples, followed by heterozygosity (α/αPA-1) for the same mutation in 32.3%. PA-1 was therefore the most frequent allele (0.59). The frequency of the α0 (--MED) allele was 0.017. Rare alleles that were found in very low frequencies included α0 (--FIL) in a Filipino child, Hb Constant Spring, Hb Adana, and Hb Icaria.

Conclusion: There is a wide variety of alpha thalassemia alleles among Kuwaitis, but nondeletional PA-1 is by far the most common cause of the moderate to severe HbH (β4 tetramer) disease phenotype. The α0 (--MED) allele is also encountered, which has implications for premarital counseling, especially for the possibility of having babies with alpha thalassemia major (Barts hydrops fetalis).

Background

Alpha thalassemia is one of the most widespread genetic diseases worldwide, with frequencies of the carrier state reaching up to 80 – 90% in some areas [1]. The incidence is approximately 25 – 30% in tropical Africa [2], while in the Arabian Peninsula, the frequency varies from a low of close to zero in the desert areas to as high as 60% in the agricultural zones of Eastern Saudi Arabia [3, 4].

The normal individual has a complement of four α-globin genes (αα/αα), any one of which can be absent, producing varying degrees of alpha thalassemia [1]. In α0 alleles, both genes on a chromosome are deleted (–), while in α+, a variable portion of the α2 and/or α1 gene is deleted, resulting in a reduction in chain synthesis. The size of the deletion in α0 alleles is variable, and each is named for the part of the world where it is prevalent. Thus --MED, --FIL, and --SEA deletions are found in the Mediterranean, Philippines and South East Asia, respectively [5]. The α+ deletions are designated by their sizes, and the most common is the −α3.7 kb allele. One- or 2-gene deletions (−a/αα, −a/−a, −/αα) produce the a-thal trait, which is usually associated with mild microcytic and hypochromic anemia. The loss of 3 genes (−/−a) produces classical hemoglobin H (HbH, β4 tetramer) disease, which is characterized by moderate to severe anemia.
The loss of all 4 genes (--/--) causes Hb Barts (γ4) hydrops fetalis, which is incompatible with life in the absence of the early onset of chronic transfusion or stem cell transplantation [6].

Apart from deletions, there are a few point mutations that affect the transcriptional efficiency of the α-globin genes and are designated αT or ααT depending on whether the α2 or α1 gene is affected [7]. These nondeletional α-thalassemia alleles commonly affect the promoter, initiation codon (ATG), splicing signals (GT/AG), termination codon (TAA), and polyadenylation signal (AATAAA) of the affected gene. The most common of these are αIVSI(-5nt)α (in the Mediterranean deletion), polyadenylation site mutations α2AATAAG, α2AATGAA and α2AATA- (in the Mediterranean and Middle East deletions) [7-11], and termination codon mutations leading to elongated Hb variants, such as Hb Constant Spring (HbCS), Hb Icaria, Hb Koya Dora, Hb Seal Rock and Hb Pakse [12-14]. Heterozygotes for these alleles have a thalassemia minor phenotype, but homozygotes and compound heterozygotes often have HbH disease of varying severity and blood transfusion requirements.

The α-thalassemia frequency is approximately 30 – 40% in Kuwait, and HbH disease is mostly linked to the polyadenylation (polyA, αPA-1) mutation [15, 16]. However, there has been no comprehensive study of the spectrum of alpha thalassemia alleles in the country. The present study was carried out to document the frequencies of moderate to severe alpha thalassemia phenotypes among patients referred for molecular diagnosis. In particular, it aimed to investigate the presence of the α0 allele in the population and to determine the possibility of having a newborn with alpha thalassemia major (Hb Bart’s hydrops fetalis) [1, 17]. This will enable more purposeful counseling of patients and empower physicians in the identification of patients who require further investigations. To this end, we analyzed all cases of suspected α-thalassemia referred to the hemoglobin research laboratory at Kuwait University over a 20-year period.

Methods

The hemoglobin research laboratory in the Department of Pediatrics, Kuwait University received samples from the hematology clinics in Mubarak Al-Kabeer Hospital and some of the other government hospitals in Kuwait. The study was approved by the Human Research Ethics Committee of the Faculty of Medicine, Kuwait University and the Ministry of Health, Kuwait. Patients were suspected to have some form of α-thalassemia if they had persistent microcytic and hypochromic anemia in the absence of iron deficiency and normal hemoglobin electrophoresis. Some patients had H inclusions on cresyl blue staining of the peripheral erythrocytes or an H band on high-performance liquid chromatography (HPLC). Quite often, but not always, there was a record of the clinical presentation and the complete blood count (CBC). This study is a retrospective study of all such samples analyzed from 1994 to 2005.

Blood was obtained by venipuncture into EDTA tubes, and a complete blood count was performed using an ABX Pentra 120 cell counter (ABX France, Montpellier). Hemoglobin quantitation was determined using cation-exchange high-performance liquid chromatography (HPLC) (Shimadzu LC-20AT, Shimadzu Corporation, Kyoto, Japan). DNA was isolated from peripheral leucocytes using phenol extraction. All the
samples were screened for the α-thal-2 (-3.7 kb) deletion [18] and the α2-globin gene polyadenylation site \((AATAA@AATAAG)\) mutation using a PCR method and allele-specific oligonucleotide hybridization, respectively, as previously reported [18, 19]. All other samples that were not resolved with these methods were subjected to reverse dot blot hybridization using the Vienna Lab StripAssay technique (Vienna Lab Diagnostics, Vienna, Austria).

The ethnic origins of the patients were documented by their nationalities. The different α-globin genotypes were identified, and the allele frequencies were calculated. The mean Hb level and RBC indices were compared among patients with different genotypes. Clinical data were collected from the HbH disease patients, especially whether they were being followed in a hematology clinic and if they were on any medications or how often they were transfused with blood.

The disease severity was classified as silent carrier, α-thalassemia trait or HbH disease depending on the number, type and location of mutations detected, in addition to the clinical phenotype and presence of HbH inclusions or H bands on HPLC [1, 17, 20]

Data are presented as the means ± SD or as percentages, as appropriate. Statistical differences between mean values among the major groups were tested using Student’s t test. All descriptive analyses were performed with the International Business Machines Corporation Statistical Package for Social Sciences (IBM SPSS Statistics, New York, USA), version 25 software. The criterion for statistical significance was \(p<0.05\).

**Results**

Four hundred blood samples were received with the diagnosis of suspected moderate to severe alpha thalassemia or HbH disease over the period of the study from 1994 to 2015. There were 183 females and 217 males, with ages ranging from 1 to 80 years and a median of 6 years. Table 1 shows the nationalities of the patients in whom these were documented; most (90.8%) were Kuwaiti. The patients were from 283 unrelated families.

Table 2 shows the distribution of α-globin genotypes in the study. The most common (33.3%) was homozygosity for \(α^{PA-1} (c.*94A>G)\) nondeletional α-thal allele, which gives an HbH disease phenotype, followed by heterozygosity for the same allele (32.3%). The compound heterozygosity of \(-α^{-3.7}/α^{PA-1}\) was the next most common at 20.5%. None of the other frequencies reached 10%.

Three (0.8%) patients were classified as silent carriers, 154 (38.5%) as α-thalassemia trait, and 243 (60.8%) had HbH disease. Table 3 shows the Hb and RBC indices in the 3 groups of patients. When values were compared between the patients with α-thal trait and those with HbH disease, all indices were significantly lower in the latter (\(p <0.05\)), although the mean Hb was not lower than 10 g/dl in all groups, showing that the α-thalassemia phenotypes were relatively mild in this population. None of the patients were on chronic transfusion therapy, but many had received sporadic transfusions. The patients with the most severe phenotypes in the study were 2 siblings with the \(\alpha^{MED}/α^{cd19}\) genotype. They both had poor
growth parameters and baseline Hb of <6 g/dl and are now transfused every 6 to 8 weeks. Interestingly, they have 2 other affected siblings (included in Tables 2 and 4) who do not require transfusions.

Table 4 shows the frequencies of the common individual alleles in the whole population and among the Kuwaiti individuals in the study. The PA-1 allele was the most frequent, at ~0.60 in both groups, followed by the -α -3.7 kb deletion at 0.153. Interestingly, α0-MED (0.019), -α -4.2, α2cd 142 (TàC) (Hb Constant Spring, c.427T>Cp.X143Gln), α2cd59 (GàA) (Hb Adana, c.179G>Ap.Gly60Asp), αPA-2 (c.*92A>G), and α2cd142 (TàA) (Hb Icaria, c.427T>Ap.X143Lys) alleles were all found only among Kuwaitis. When the frequencies were computed based on the total number of families in the study, again, the PA-1 allele was most frequent at 0.0552, followed by the -α -3.7 kb deletion at 0.18. This was approximately the same when only Kuwaiti families were considered.

Discussion

The molecular diagnosis of α-thalassemia is not straightforward and requires resources that are not readily available in many centers. Moreover, it is economically unviable to provide this service to all suspected patients in many countries with a high prevalence. It is therefore necessary to have a good understanding of the prevalent genotypes and phenotypes, which will facilitate genetic counseling and the identification of patients who would best benefit from molecular diagnosis. This report is limited to patients who were suspected to have moderate to severe α-thalassemia based on persistent microcytic and hypochromic anemia, no laboratory evidence of iron deficiency and normal hemoglobin electrophoresis or HPLC. It also included patients with H inclusion bodies and/or an H band on HPLC.

Given the selection criteria for screening patients in this study, it is not surprising that most patients had the HbH disease phenotype and that few had thalassemia trait or silent carrier status. A previous study from Kuwait showed that most alpha thalassemia silent carriers had Hb levels >10 g/dl. Therefore, individuals with mild microcytosis and hypochromia, but with Hb >10 g/dl and no HbH inclusions or bands, should not be considered for molecular diagnosis, but counseling should be offered after screening the family with CBC and HPLC to rule out α-thalassemia. Resources can then be channeled to individuals with a more severe outlook to identify those with HbH disease that need to be followed in the clinic.

This study revealed the extent of the diversity among individuals with alpha thalassemia in Kuwait. As expected, the vast majority of the patients had the αPA-1 nondeletional allele, which has been reported as the most common cause of HbH disease in Kuwait and in the region. The next most common allele was the –α -3.7 kb deletion, although it was usually present as a compound heterozygote with other alleles. Interestingly, the third most frequent allele in this study was α0 (–MED), which was found in 13 patients, in whom it mostly presented an HbH disease phenotype. The –MED deletion was in compound heterozygosity with α2cd19-G in 6 patients from 2 related families, with –α -3.7 kb in 2 patients and with αIVS1-5nt and αPA-1 in 1 patient each.
The αPA-1 allele was first described in Eastern Saudi Arabia [10], and previous studies from Kuwait [2, 16] highlighted its role in the etiology of HbH disease among our patients. It has also been reported as the most common cause of HbH disease in Saudi Arabia [21], Jordan, [22], Bahrain [23] and the UAE [24]. The allele is therefore widespread in the Arabian Gulf, especially in the countries adjoining Saudi Arabia. It is uniformly associated with a moderate thalassemia intermediate phenotype with only occasional blood transfusion requirements, usually with intercurrent infection [16]. HbH disease is mainly a disease of childhood in Kuwait. Most patients are referred to the clinic for investigation of persistent microcytic and hypochromic anemia not responding to iron therapy. Indeed, the anemia tends to improve as the child stops rapidly growing and enters puberty.

This study is the first to report the presence of the α0 (–MED) allele among Kuwaiti patients. It presented as compound heterozygotes with other α-thal alleles; however, the phenotype was mild with moderate anemia. The exceptions were 2 patients with the –MED/αcd19α genotype, who presented with stunted growth and are on regular transfusion because of severe anemia. The other rare alleles found in the study included α0 (–FIL) in a Filipino child, HbCS (α2Cd 142 TAA → CAA), Hb Adana (αcd59G/A), αPA-2 (AATAAA®AATGAA), and Hb Icaria (αcd 142 TAA → AAA).

Conclusions

This study demonstrated the diversity of alpha thalassemia alleles among Kuwaitis. While the deletional allele is quite widespread, nondeletional αPA-1 is responsible for most moderate to severe HbH disease. Interestingly, the α0 (–MED) allele is also encountered, with implications for premarital genetic counseling to prevent the birth of babies with alpha thalassemia major (Bart’s hydrops fetalis).

List Of Abbreviations

| Abbreviation | Definition                  |
|--------------|-----------------------------|
| PCR          | Polymerase chain reaction   |
| PA           | Polyadenylation             |
| MED          | Mediterranean               |
| FIL          | Filipino                    |
| SEA          | Southeast Asia              |
| Kb           | Kilobases                   |
| Hb           | Hemoglobin                  |
| CS           | Constant Spring             |
| CBC          | Complete blood count        |
Declarations

Ethics Approval and Consent to Participate

The study was approved by the Human Research Ethics Committees of the Faculty of Medicine, Kuwait University and the Ministry of Health, Kuwait. No administrative permissions or licenses were required to access any of the data used in this research. The patients and/or their parents/guardians (as appropriate) gave written consent to participate in this study.

Consent for Publication

Not applicable.

Competing Interests

There are no other competing financial or nonfinancial interests.

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Authors’ Contributions

1. AA was the lead physician who planned the study and carried out the data analysis and wrote the first draft.
2. MH was the head of the laboratory and carried out the molecular characterization. He contributed to the writing.
3. JS, DT, and TDS all contributed to sample and data collection/analysis.

All authors have read and approved the manuscript.

Availability of Data and Materials

The dataset used and/or analyzed during the current study is available from the corresponding author on reasonable request.
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References

1. Piel FB, Weatherall DJ: The alpha-thalassemias. *The New England journal of medicine* 2014, 371(20):1908-1916.
2. Adekile AD, Liu JC, Sulzer AJ, Huisman TH: Frequency of the alpha-thalassemia-2 gene among Nigerian SS patients and its influence on malaria antibody titers. *Hemoglobin* 1993, 17(1):73-79.
3. White JM, Byrne M, Richards R, Buchanan T, Katsoulis E, Weerasingh K: Red cell genetic abnormalities in Peninsular Arabs: sickle haemoglobin, G6PD deficiency, and alpha and beta thalassaemia. *Journal of medical genetics* 1986, 23(3):245-251.
4. White JM, Byrne M, Richards R, Buchanan T, Sheik-Yousouf I, White YS, Frost B: Thalassaemia genes in Peninsular Arabs. *British journal of haematology* 1985, 60(2):269-278.
5. Fucharoen S, Viprakasit V: Hb H disease: clinical course and disease modifiers. *Hematology American Society of Hematology Education Program* 2009:26-34.
6. Vichinsky EP: Clinical manifestations of alpha-thalassemia. *Cold Spring Harbor perspectives in medicine* 2013, 3(5):a011742.
7. Harteveld CL, Losekoot M, Haak H, Heister GA, Giordano PC, Bernini LF: A novel polyadenylation signal mutation in the alpha 2-globin gene causing alpha thalassaemia. *British journal of haematology* 1994, 87(1):139-143.
8. Harteveld CL, Higgs DR: Alpha-thalassemia. *Orphanet journal of rare diseases* 2010, 5:13.
9. Hall GW, Higgs DR, Murphy P, Villegas A, de Miguel A: A mutation in the polyadenylation signal of the alpha 2 globin gene (AATAAA→AATA–) as a cause of alpha thalassaemia in Asian indians. *British journal of haematology* 1994, 88(1):225-227.
10. Higgs DR, Goodbourn SE, Lamb J, Clegg JB, Weatherall DJ, Proudfoot NJ: Alpha-thalassaemia caused by a polyadenylation signal mutation. *Nature* 1983, 306(5941):398-400.
11. Yuregir GT, Aksoy K, Curuk MA, Dikmen N, Fei YJ, Baysal E, Huisman TH: Hb H disease in a Turkish family resulting from the interaction of a deletional alpha-thalassaemia-1 and a newly discovered poly A mutation. *British journal of haematology* 1992, 80(4):527-532.
12. Clegg JB, Weatherall DJ, Contopolou-Griva I, Caroutsos K, Pounouras P, Tsevrenis H: Haemoglobin Icaria, a new chain-termination mutant with causes alpha thalassaemia. *Nature* 1974, 251(5472):245-247.
13. Clegg JB, Weatherall DJ, Milner PF: Haemoglobin Constant Spring—a chain termination mutant? *Nature* 1971, 234(5328):337-340.
14. De Jong WW, Meera Khan P, Bernini LF: Hemoglobin Koya Dora: high frequency of a chain termination mutant. *American journal of human genetics* 1975, 27(1):81-90.
15. Haider M, Adekile A: Alpha-2-globin gene polyadenylation (AATAAA->AATAAG) mutation in hemoglobin H disease among Kuwaitis. Med Princ Pract 2005, 14 Suppl 1:73-76.

16. Adekile AD, Azab AF, Al-Sharida SI, Al-Nafisi BA, Akbulut N, Marouf RA, Mustafa NY: Clinical and Molecular Characteristics of Non-Transfusion-Dependent Thalassemia in Kuwait. Hemoglobin 2015, 39(5):320-326.

17. Viprakasit V, Ekwattanakit S: Clinical Classification, Screening and Diagnosis for Thalassemia. Hematology/oncology clinics of North America 2018, 32(2):193-211.

18. Baysal E, Huisman T: Detection of common deletional alpha-thalassemia-2 determinants by PCR. Am J Hematol 1994, 46:208-213.

19. Adekile AD, Haider MZ: Morbidity, beta S haplotype and alpha-globin gene patterns among sickle cell anemia patients in Kuwait. Acta Haematol 1996, 96(3):150-154.

20. Gilad O, Shemer OS, Dgany O, Krasnov T, Nevo M, Noy-Lotan S, Rabinowicz R, Amitai N, Ben-Dor S, Yaniv I et al: Molecular diagnosis of alpha-thalassemia in a multiethnic population. European journal of haematology 2017, 98(6):553-562.

21. Al-Awamy BH: Thalassemia syndromes in Saudi Arabia. Meta-analysis of local studies. Saudi medical journal 2000, 21(1):8-17.

22. Abu-Ghoush M: Subtypes of alpha thalassemia diagnosed at a medical center in Jordan. TAF Prev Med Bull 2008, 7(5):373-376.

23. Jassim N, Al-Arrayed S, Gerard N, Al-Mukharraq H, Al-Ajami A, Ducrocoq R, Nagel R, R K: Molecular Basis of α-Thalassemia in Bahrain Bahrain Medical Bulletin, 2001, 23(1):3-7.

24. El-Kalla S, Baysal E: alpha-thalassemia in the United Arab Emirates. Acta haematologica 1998, 100(1):49-53.

Tables

Table 1

Nationalities of the Patients in the study

| Nationality                | n   | %   |
|----------------------------|-----|-----|
| Kuwaiti                    | 317 | 90.8|
| Iraqi                      | 10  | 2.9 |
| Egyptian                   | 6   | 1.7 |
| Lebanese                   | 4   | 1.1 |
| Non Kuwaiti Bedouin        | 3   | 0.9 |
| Syrian                     | 3   | 0.9 |
| Others                     | 6   | 1.7 |
| Total                      | 349*| 100.0 |
These are all the patients in whom nationality was documented

Table 2
Distribution of Alpha Globin Genotypes of the Patients in the Study

| Genotype              | Number | %  |
|-----------------------|--------|----|
| α^PA-1α/α^PA-1α       | 133    | 33.3|
| α^PA-1α/aa            | 129    | 32.3|
| −α^3.7/α^PA-1α        | 81     | 20.5|
| −α^3.7/−α^3.7         | 20     | 5.0 |
| _MED/α^CD19α          | 6      | 1.5 |
| α^CD59α/α            | 4      | 1.0 |
| −α^3.7/αaa            | 3      | 0.8 |
| _MED/αaa              | 3      | 0.8 |
| α^5ntα/α^5ntα         | 3      | 0.8 |
| −α^3.7/α^5ntα         | 2      | 0.5 |
| −α^3.7/−MED           | 2      | 0.5 |
| aa/α^CD19α           | 2      | 0.5 |
| α^5ntα/α^PA-1α        | 1      | 0.3 |
| _MED/α^5ntα           | 1      | 0.3 |
| _MED/α^PA-1α          | 1      | 0.3 |
| α^CSα/α^CD142α        | 1      | 0.3 |
| α^PA-2α/αaa           | 1      | 0.3 |
| α^5ntα/aa             | 1      | 0.3 |
| −α^4.2/−α^4.2         | 1      | 0.3 |
| α^CD19α/α^CD19α       | 1      | 0.3 |
| α^PA1α/α^PA2α         | 1      | 0.3 |
| α^CD59α/αaa           | 1      | 0.3 |
| α^CD59α/α^PA-1α       | 1      | 0.3 |
| α^CD59α/α^5ntα        | 1      | 0.3 |
| _SEA/−α^3.7           | 1      | 0.3 |
| Total                 | 400    | 100 |

Table 3
Red Blood Cell Indices in Patients
| Group             | Hb g/dl   | MCV fl   | MCH pg  |
|------------------|-----------|----------|---------|
| Silent Carrier   | 10.4 ± 0.8| 60.7 ± 9.5| 19.8 ± 2.5|
| a-Thalassemia Trait | 11.3 ± 2.0| 64.4 ± 11.0| 21.1 ± 3.5|
| HbH              | 10.0 ± 1.6| 59.2 ± 8.7| 18.3 ± 3.1|
| Total            | 10.5 ± 1.9| 61.2 ± 10.0| 19.4 ± 3.5|

**Table 4**

Frequency of the Common Alpha Thalassemia Alleles in the Study

| Allele                                      | All population | Frequency | Kuwaiti | Frequency |
|---------------------------------------------|----------------|-----------|---------|-----------|
| AATAAAA\>AATAAG (PA-1)                      | 412            | 0.590     | 386     | 0.608     |
| -α-3.7 single gene deletion                 | 107            | 0.153     | 91      | 0.143     |
| --MED double gene deletion                  | 12             | 0.017     | 12      | 0.019     |
| α2 cd19 (-G)                                | 10             | 0.0143    | 9       | 0.0141    |
| α2 IVS-1 (-5nt)                             | 8              | 0.011     | 5       | 0.008     |
| α2 cd59 (G\>A)                              | 4              | 0.006     | 4       | 0.006     |
| -α-4.2 single gene deletion                 | 2              | 0.003     | 2       | 0.003     |
| AATAAA\>AATGAA (PA-2)                      | 2              | 0.003     | 2       | 0.003     |
| α2 cd 142 (T\>C) Hb Constant Spring        | 2              | 0.003     | 2       | 0.003     |