Amino Acid Synthesis in a Supercritical Carbon Dioxide - Water System

Kouki Fujioka 1,2, Yasuhiro Futamura 1,*, Tomoo Shiohara 3, Akiyoshi Hoshino 1, Fumihide Kanaya 1, Yoshinobu Manome 2 and Kenji Yamamoto 1,*

1 International Clinical Research Centre, Research Institute, International Medical Centre of Japan, 1-21-1 Toyama, Shinjuku-ku, Tokyo, Japan; E-Mails: kfujio@ri.imcj.go.jp (K.F.); hoshinoa@nih.go.jp (A.H.)
2 Department of Molecular Cell Biology, Institute of DNA Medicine, Jikei University School of Medicine, 3-25-8 Nishi-shimbashi, Minato-ku, Tokyo, Japan; E-Mail: manome@jikei.ac.jp
3 Sekisui Chemical Co., Ltd., 2-1 Hyakuyama, Shimamoto-cho, Mishima-gun, Osaka, Japan; E-Mail: shiohara002@sekisui.jp

* Authors to whom correspondence should be addressed; E-Mails: backen@ri.imcj.go.jp (K.Y.); futamura@nih.go.jp (Y.F.); Tel. +81-3-3202-7181; Fax: +81-3-3202-7364

Received: 21 May 2009; in revised form: 5 June 2009 / Accepted: 8 June 2009 / Published: 15 June 2009

Abstract: Mars is a CO₂-abundant planet, whereas early Earth is thought to be also CO₂-abundant. In addition, water was also discovered on Mars in 2008. From the facts and theory, we assumed that soda fountains were present on both planets, and this affected amino acid synthesis. Here, using a supercritical CO₂/liquid H₂O (10:1) system which mimicked crust soda fountains, we demonstrate production of amino acids from hydroxylamine (nitrogen source) and keto acids (oxylic acid sources). In this research, several amino acids were detected with an amino acid analyzer. Moreover, alanine polymers were detected with LC-MS. Our research lights up a new pathway in the study of life’s origin.

Keywords: supercritical carbon oxide; amino acid; prebiotic chemistry; soda fountain; Mars
1. Introduction

Prebiotic amino acids syntheses have attracted the attention of scientists [1], since amino acids are one of the essential materials for chemical evolution in early Earth. The prebiotic synthesis by electric discharge in reducing gasses (CH$_4$, NH$_3$, H$_2$, and H$_2$O) through Miller’s experiment has given experimental support to the chemical evolution [2]. After Miller’s report, many prebiotic syntheses in various types of geological environments have been examined, including volcanoes [3], hydrothermal system [4-8], meteorites[9,10], and other planetary bodies [11].

In the recent research, however, the early Earth atmosphere is thought to have had a moderately oxidative state, being mostly composed of CO$_2$ and N$_2$[12]. There are few reports on the synthesis of organic molecules including amino acids under such a condition [13]. For this reason, the fundamental question about the origin of life – how organic molecules appeared in the early Earth – nowadays is still open. From the early Earth atmosphere, we focused on CO$_2$, which is also a component of the atmosphere of Mars, as a reaction medium. CO$_2$ is known to change directly from solid phase to gas phase through sublimation at $-78.51 \, ^\circ$C and normal pressure on Earth. Liquid carbon forms only above the triple point ($-56.6 \, ^\circ$C, 0.52 MPa). The critical point of CO$_2$ is known at 31.1 $^\circ$C, 7.38 MPa and supercritical CO$_2$ is reactive towards other molecules.

Considering the reactivity, we assumed that the supercritical CO$_2$ in soda fountains in the crusts on early Earth and Mars affected amino acid synthesis. For the existence of such soda fountains, there are two essential factors: (1) abundant CO$_2$ (carbonate) and H$_2$O and (2) late stage of volcanoes. Although McKay et al. reported the detection of polycyclic aromatic hydrocarbons in the Martian meteorite ALH84001 as a clue of organic synthesis [14], is there any evidence of soda fountains on Mars? Different from the Earth, on Mars the main component of the atmosphere is CO$_2$. Moreover, carbonate globules were also detected in the Martian meteorite. Additionally, like on Earth, H$_2$O was present abundantly on early Mars, since the vestiges of rivers were found on the over-3-Gyr ground [15]. On the other hand, there are thought to have been some active volcanoes 3.5–2.5 Gyr before, due to the volcanism simulation based on the model of Martian mantle convection [16]. Therefore soda fountains could existence on early Earth or Mars. To the best of our knowledge, no previous study has examined soda fountains in the crusts as a reaction medium for producing organic molecules necessary for the origin of life.

The principal objective of this study was to determine the possibility of amino acid synthesis in the co-existence of supercritical CO$_2$ and liquid H$_2$O, mimicking crust soda fountains which represent more moderate conditions than supercritical H$_2$O. Our research does not contradict other hypotheses of the life’s origin and we assumed that simple organic molecules from other prebiotic syntheses enter to our synthesis system. Therefore we chose hydroxylamine and pyruvic acid as the primary reactants, which could be synthesized in other prebiotic synthesis [17,18]. In this paper, we demonstrated that the co-existence of supercritical CO$_2$ and liquid H$_2$O could produce some amino acids and polymers from the simple organic molecules.
2. Results and Discussion

2.1. Amino acid synthesis in supercritical carbon dioxide

We investigated the possibility that amino acids can be synthesized from hydroxylamine and pyruvic acid or glyoxylic acid, under supercritical CO₂ conditions (60 °C). Additionally, in order to compare the conditions, we repeated the same reaction under subcritical CO₂ conditions (31 °C) as a control. For the detection of amino acids, an amino acid analyzer was used (Figure 1). Since this analysis system is adjusted for detecting amino-acid monomers, the differences before and after the hydrolysis reaction indicates the amount of polymer hidden in the original products.

**Figure 1.** Chromatograms of amino-acid analysis. (a) Amino acid standards. (b) Pyruvic acid + Hydroxylamine at 31 °C. (c) Pyruvic acid + Hydroxylamine at 60 °C. (d) Glyoxylic acid + Hydroxylamine at 31 °C. (e) Glyoxylic acid + Hydroxylamine at 60 °C.
Using pyruvic acid as carbon sources, two kinds of amino acids, alanine and glycine, were detected at 60 °C and 31 °C (Table 1). The free alanine yield at 60 °C (60.7 µmol) was higher than that at 31 °C (4.1 µmol), whereas the free glycine yield at 60 °C (0.99 µmol) was lower than that at 31 °C (2.2 µmol). These results suggested that alanine synthesis was conducted more effectively at 60 °C.

After hydrolysis treatment with 6N HCl, 256.2 µmol and 70.2 µmol of alanine were detected at 60 °C and 31 °C, respectively (Table 1). Additionally, arginine (0.32 µmol) was detected at 60 °C. From the significant differences of concentrations between before and after hydrolysis reaction, alanine polymers were synthesized in larger quantity at 60 °C. However, the ratios of free alanine: alanine polymers, were 1:3.2 at 60 °C; 1:16.5 at 31 °C. Considering these results, alanine polymers may be more stable in lower temperature or degraded at high temperature.

In order to estimate the mechanism of alanine synthesis, we compared the reactions using glyoxylic acids as another carboxylic acid source. Different from the reaction using pyruvic acid, free alanine was not detected at both temperatures, whereas free glycine was detected at 60 °C (866.3 µmol) and 31 °C (183.3 µmol); free aspartic acid (2.5 µmol) was detected at 60 °C (Table 1). These results suggested that the methyl group of alanine synthesized from pyruvic acid were derived from the methyl group of pyruvic acid, and that the alanine was synthesized from hydrogenation of pyruvic acid oxime, since alanine was not detected in the glyoxylic acid reaction.

Some methods have been proposed for the synthesis of alanine from pyruvic acid oxime. Hamlin et al. reported a hydrogenation of the oxime with a palladium-charcoal catalyst [19], and Borszeky et al. reported that a hydrogenation of pyruvic acid oxime with palladium/alumina catalysts provided the high yields of racemic alanine [20]. Although these hydrogenation were conducted with catalysts, we speculated that oxime hydrogenation also occurred under our supercritical CO2 conditions.

After the hydrolysis of the product from glyoxylic acid, glycine (819.2 µmol) and aspartic acid (1.8 µmol) were detected at 60 °C, whereas glycine (209.2 µmol) was detected at 31 °C reaction (Table 1). Additionally, 1/2-cystine (1.2 µmol) and leucine (0.37 µmol) was detected at 60 °C and 31 °C, respectively (data not shown). The sulfur atoms of leucine and 1/2-cystine may be derived from impurities of the product of hydroxylamine hydrochloride (see Experimental Section).

### Table 1. Products obtained from keto acids and hydroxylamine in supercritical conditions (60 °C) or subcritical condition (31 °C) before/after hydrolysis reaction.

| Primary Reactant | Temp. (°C) | Products (µmol) | Before hydrolysis reaction | After hydrolysis reaction |
|------------------|------------|-----------------|---------------------------|---------------------------|
|                  |            | Ala  | Gly  | Asp  | Arg | Ala  | Gly  | Asp  | Arg |
| Pyruvic acid     | 60         | 60.7 | 0.99 | —    | —   | 256.2| 1.5  | —    | 0.32|
|                  | 31         | 4.1  | 2.2  | —    | —   | 70.2 | 3.4  | —    | —   |
| Glyoxylic acid   | 60         | —    | 866.3| 2.5  | —   | —    | 819.2| 1.8  | —   |
|                  | 31         | —    | 183.3| —    | —   | —    | 209.2| —    | —   |

— indicates “Not Detected”.
The main detected amino acids both before/after hydrolysis conditions, alanine, glycine, and aspartic acid, are GNC code encoding amino acids (N means either of four bases). Ikehara et al. proposed in their hypothesis a [glycine, alanine, aspartic acid, and valine (GADV)]-protein world hypothesis on the origin of life, based on the GNC code encoding amino acids as the most primitive genetic code [21]. These GADV amino acids are known to be detected in the Miller discharge experiment [2] and the Miller volcanic spark discharge experiment using his apparatus performed after his death [22]. Although valine was not detected in our experiments using pyruvic acid or glyoxylic acid, as a hypothesis, if the carboxylic acid source is changed to α-ketoisovaleric acid (3-methyl-2-oxobutanoic acid), valine may be synthesized under supercritical CO₂ conditions. Our data in CO₂ were thus partially agreement with the GADV-protein world hypothesis.

Surprisingly, in the glyoxylic acid reaction, the concentrations of glycine before and after hydrolysis reaction did not change dramatically (after hydrolysis reaction: before hydrolysis reaction = 209.2 µmol:183.3 µmol at 31 ºC; 819.2:866.3 at 60 ºC). Comparing the alanine data in the pyruvic acid reaction, it was estimated that glycine polymers were more unstable under supercritical or subcritical CO₂ conditions, since the ratio of alanine polymers was higher than that of free alanine in the above experiments. In contrast with the results, our previous study showed that glycine polymers, including decaglycine, were obtained from glycine monomer under hydrothermal condition (270 ºC, 10 MPa, and 27-second reaction) with an adiabatic expansion cooling system [23]. The cooling system may be an essential condition to obtain unstable glycine polymers. We speculated that alanine polymers were stable due to the formation of helix forms [24,25], but glycine polymers were unstable due to the large flexibility of the residues under CO₂ conditions, like in the aqueous model [25].

2.2. Analysis of alanine polymer

In order to confirm alanine polymer synthesis under supercritical CO₂ conditions, we analyzed the sample from pyruvic acid at 60 ºC with a LC-MS. We obtained 56 peaks under the gradient LC conditions used (Figure 2). Among these peaks, the positive ions, [M+H]⁺, of 232.13 (alanine trimer) and 303.16 (alanine tetramer) were detected at 33.7 and 34.6 min retention time, respectively (Table 2). The alanine dimer and 5 – 7 mers were not detected. These results suggested that alanine trimer and tetramer were stable under CO₂ supercritical conditions. From the ESR spectroscopic data, several alanine-based forms can adopt the 3₁₀ helix formation in aqueous solution [26,27]. Also under these supercritical conditions, the tetramers could be stable by forming the 3₁₀ helix and were thus detected in the aqueous sample due to their stability. However, we could not found out the reason why the trimer was stable under CO₂ supercritical conditions. We speculate that the trimer was a metastable structure in the supercritical CO₂; therefore the trimer was synthesized directly from three alanine monomers, not through the dimer structure, and the tetramer was synthesized from a trimer + monomer reaction. The polymers higher then pentamer may be separated out as deposits from supercritical CO₂. This deposition polymerization in supercritical CO₂ is known in ethylene polymerizations [28,29]. At the present, it is outside the scope of our research to investigate the deposition, but it will be the subject in future studies.
Table 2. Polymers obtained from pyruvic acid and hydroxylamine in supercritical conditions.

|                | Alanine monomer | Dimer | Trimer | Tetramer | Pentamer | Hexamer | Heptamer |
|----------------|-----------------|-------|--------|----------|----------|---------|---------|
| **Calculated** |                 |       |        |          |          |         |         |
| **[M+H]**      | 90.05           | 161.09| 232.13 | 303.16   | 374.20   | 445.24  | 516.28  |
| **Retention**  | suspended        |       | 33.7   | 34.6     |          |         |         |
| **time(min)    |                 |       |        |          |          |         |         |

— indicates "Not Detected". The retention time for alanine monomer in the sample could not be specified with the LC-MS system used.

Figure 2. Total ion chromatogram (TIC) and MS chromatograms of alanine monomer and polymers.
3. Experimental Section

3.1. CO₂ reaction conditions

The reactor was made of stainless steel and was connected to a CO₂ inlet and outlet that have valves and a pressure gauge (Figure 3). Twenty-six mmol of hydroxylamine hydrochloride (>97%; Tokyo Chemical Industry Co. Ltd.) and 26 mmol of pyruvic acid (98%; Alfa Aesar) or glyoxylic acid monohydrate (98%; Alfa Aesar) were mixed with 3 mL of Milli-Q (Millipore) water gently in a glass tube. The hydroxylamine hydrochloride could contain the following impurities: sulfate: max. 0.005%; copper: max. 2 ppm; lead: max. 2 ppm; iron: max. 5 ppm. After the glass tube was inserted into the stainless tube, the stainless tube was capped and CO₂ liquid was injected to purge the air inside. Then 20 g of CO₂ liquid was injected through the inlet of the reactor contained in an ice box. In order to compare the reaction conditions, we conducted reactions under two sets of conditions: supercritical condition (60 °C, over 7.4 MPa) and subcritical condition (31 °C, less than 7.2 MPa). This subcritical condition was chosen on the basis of our hypothesis that subcritical CO₂ conditions near the supercritical conditions are most effective for the reactions. The reactor was placed in an oil bath for two h in each condition. After the reactions, the reactors were rapidly moved into an ice box and the CO₂ gas released. One mL of Milli-Q water was added into the glass tube, and all the samples were transferred to a new glass bottle.

Figure 3. Diagram of the autoclave reactor for the supercritical CO₂ reaction.
3.2. Amino-acid analysis

The aqueous samples (10 μL) were diluted with purified water (200-fold dilution) and filtered with 0.22-μm filters. On the other hand, the samples for hydrolysis studies were evaporated to dryness under reduced pressure. The dried residues were treated with 100 μL of 6N HCl. After vacuum sealing, the samples were hydrolyzed for 22 hr at 110 °C. After the hydrolysis, the hydrolyzed residues were evaporated to dryness under reduced pressure. The dried residues were dissolved with 200 μL of purified water and filtered with 0.22-μm filters. The filtrate samples were diluted with purified water (50-fold dilution). Both the aqueous samples and hydrolyzed samples were analyzed with an amino-acid analyzer, L-8500 (Hitachi High Technologies Co.), based on the absorbencies of 570 nm and 440 nm after ninhydrin reaction.

3.3. LC-MS analysis

LC-MS analysis of the 60 °C sample using pyruvic acid was performed using LCMS-IT-TOF (Shimadzu Co.). The LC-MS components included a LC-20AD pump, a CBM-20A system controller, SPD-20A UV detector. The column used for reversed-phase LC analysis was a TSKgel ODS-80Ts (Tosoh Co.) packed with 5 μm particles (2.0 x 250 mm) and it was incubated at 40 °C.

The LC analysis consisted of the following conditions: Mobile phase A: 100% H2O (0.1% formic acid); Mobile phase B: 80% ACN + 20% H2O (0.1% formic acid); Flow rate: 0.200 mL/min; Gradient: 0 min 1% B, 10 min 1% B, 40 min 65% B, 40.01 min 100% B, 50 min 100% B, 50.01 min 1% B, and 60 min 1% B. Stop time: 60 min. A 100 μL of injection volume was used. The LCMS-IT-TOF was operated under the following conditions: ESI in positive mode; Detection voltage: 1.53 kV; MS scan range: 80 – 800 m/z. Both amino-acid analyses and LC-MS analyses were conducted at Toray Research Center. Inc.

4. Conclusions

In this research, we have demonstrated that four kinds of amino acid including alanine, glycine, aspartic acid, and arginine were obtained from hydroxylamine hydrochloride and pyruvic acid or glyoxylic acid under supercritical CO2 condition, mimicking a soda fountain (Figure 4). Additionally, trimer and tetramer alanine polymers from pyruvic acid were abundantly detected under supercritical CO2 conditions, but glycine polymers were almost not detected. From these data, we conclude that even at low temperatures less than 60 °C, supercritical CO2 and liquid H2O conditions could provide a variety of amino acids and short polymers when the formations are stable. Although the reaction mechanism was not revealed clearly (e.g. which factor is more important, temperature or pressure?), our research opens a new pathway in the study of the origin of life.
Figure 4. Reaction overview of the supercritical CO\textsubscript{2}/liquid H\textsubscript{2}O reactions. (a) Hydroxylamine + Pyruvic acid (Pyruvic acid oxime) (b) Hydroxylamine + Glyoxylic acid (Glyoxylic acid oxime).

\[
\text{HONH}_2\cdot\text{HCl} + \text{RCO}_2\text{H} \rightarrow \text{RC}=\text{NOH} \nonumber
\]

(a) \[ \text{HONH}_2\cdot\text{HCl} \text{ (Hydroxylamine hydrochloride)} \quad \text{RCO}_2\text{H} \text{ (Pyruvic or Glyoxylic acid)} \]

(b) \[ \text{HONH}_2\cdot\text{HCl} \text{ (Pyruvic acid oxime)} \]

Acknowledgements

We thank Mr. Tsuneo Shimazu, Mr. Yasuhiro Kagawa, Ms. Yasuko Mizuno, and Ms. Satoko Ota (Toray Research Center), for analyzing our products. We also thank Mr. Akira Nakasuga, Mr. Motoi Nagano, Mr. Yasuhiro Nakatani, Mr. Yohei Nishimura, and Ms. Hiroko Minamino (Sekisui Chemical Co., Ltd.) for great advices and managements of our research.
References and Notes

1. Simoneit, B.R.T. Prebiotic organic synthesis under hydrothermal conditions: an overview. *Adv. Space Res.* 2004, 33, 88-94.

2. Miller, S.L. A production of amino acids under possible primitive Earth conditions. *Science* 1953, 117, 528-529.

3. Markhinin, E.K.; Podkletnov, N.E. The phenomenon of formation of prebiological compounds in volcanic processes. *Orig. Life* 1977, 3, 225-235.

4. Charlou, J.; Donval, J. Hydrothermal methane venting between 12°N and 6°N along the Mid-Atlantic Ridge. *J. Geophys. Res.* 1993, 98, 9625-9642.

5. Gunter, B.D. C₁-C₄ hydrocarbons in hydrothermal gases. *Geochim. Cosmochim. Acta* 1978, 42, 137-139.

6. Simoneit, B.R.T.; Kawka, O.E.; Brault, M. Origin of gasses and condensates in the Guaymas Basin hydrothermal system (Gulf of California). *Chem. Geol.* 1988, 71, 169-182.

7. Welhan, J.A. Origins of methane in hydrothermal systems. *Chem. Geol.* 1988, 71, 183-198.

8. Welhan, J.A.; Lupton, J.E. Light hydrocarbon gasses in Guaymas Basin hydrothermal fluids: Themogenic versus abiogenic origin. *Am. Assoc. Petrol. Geol. Bull.* 1987, 71, 215-223.

9. Furukawa, Y.; Siekine, T.; Oba, M.; Kakegawa, T.; Nakazawa, H. Biomolecule formation by oceanic impacts on early Earth. *Nature Geosci.* 2009, 2, 62-66.

10. Gelpi, E.; Han, L.; Nooner, D.W.; Oro, J. Closed system Fischer Tropsch synthesis over meteoric iron, iron ore and nickel iron alloy. *Geochim. Cosmochim. Acta* 1970, 34, 965-979.

11. Studier, M.H.; Hayatsu, R.; Anders, E. Origin of organic matter in early solar system (I. Hydrocarbons). *Geochim. Cosmochim. Acta* 1968, 32, 151-173.

12. Kasting, J.F. Bolide impacts and the oxidation of carbon in the Earth's early atmosphere. *Orig. Life Evol. Biosphere* 1989, 20, 199-231.

13. Kobayashi, K.; Tsuchiya, M.; Oshima, T.; Yanagawa, H. Abiotic synthesis of amino acids and imidazol by proton irradiation of simulated primitive Earth's early atmosphere. *Orig. Life Evol. Biosphere* 1990, 20, 99-109.

14. McKay, D.S.; Gibson, E.K.; Thomas-Keprta, K.L.; Vali, H.; Romanek, C.S.; Clemett, S.J.; Chilier, X.D.F.; Maechling, C.R.; Zare, R.N. Search for Past Life on Mars: Possible Relic Biogenic Activity in Martian Meteorite ALH84001. *Science* 1996, 273, 924-930.

15. Sagan, C.; Toon, O.B.; Gierasch, P.J. Climatic Change on Mars. *Science* 1973, 181, 1045-1049.

16. Harder, H.; Christensen, U.R. A one-plume model of martian mantle convection. *Nature* 1996, 380, 507-509.

17. Cody, G.D.; Boctor, N.Z.; Filley, T.R.; Hazen, R.M.; Scott, J.H.; Sharma, A.; Yoder, H.S., Jr. Primordial synthesis of carbonylated iron-sulfur clusters and the synthesis of pyruvate. *Science* 2000, 289, 1337-1340.

18. Sokolskaya, A. Glycine and alanine synthesis from formaldehyde and hydroxylamine in the field of ultrasound waves. *Orig. Life* 1976, 7, 183-185.

19. Hamlin, K.E., Jr.; Hartung, W.H. The synthesis of α-amino acids from substituted acetoacetic esters. *J. Biol. Chem.* 1942, 145, 349-357.
20. Borszeky, K.; Mallat, T.; Aeschiman, R.; Schweizer, W.B.; Baiker, A. Enantioselective Hydrogenation of Pyruvic Acid Oxime to Alanine on Pd/Alumina. *J. Catalysis* 1996, 161, 451-458.

21. Ikehara, K. Origins of gene, genetic code, protein, and life. *Viva Origino* 2001, 29, 66-85.

22. Johnson, A.P.; Cleaves, H.J.; Dwokin, J.P.; Glavin, D.P.; Lazcano, A.; Bada, J.L. The miller volcanic spark discharge experiment. *Science* 2008, 322, 404.

23. Goto, T.; Futamura, Y.; Yamaguchi, Y.; Yamamoto, K. Condensation reaction of amino acids under hydrothermal conditions with adiabatic expansion cooling. *J. Chem. Eng. Jpn.* 2005, 38, 295-299.

24. Bixon, M.; Scheraga, H.A.; Lifson, S. Effect of hydrophobic binding on the stability of poly-L-alanine helices in water. *Biopolymers* 1963, 1, 419-429.

25. Aleman, C.; Roca, R.; Luque, F.J.; Orozco, M. Helical Preferences of alanine, glycine, and aminoisobutyric homopeptides. *Proteins* 1997, 28, 83-93.

26. Miick, S.M.; Martinez, G.V.; Fiori, W.R.; Todd, A.P.; Millhouser, G.L. Short alanine-based peptides may form 3(10)-helices and not alpha-helices in aqueous solution. *Nature* 1992, 359, 653-655.

27. Fiori, W.R.; Miick, S.M.; Millhouser, G.L. Increasing sequence length favors alpha-helix over 3(10)-helix in alanine-based peptides: Evidence for a length dependent structural transition. *Biochemistry* 1993, 32, 11957-11962.

28. Hagiwara, M.; Mitsui, H.; Machi, S.; Kagiya, T. Liquid carbon dioxide as a solvent for the radiation polymerization of ethylene. *J. Polym. Sci. A-Polym. Chem.* 1968, 6, 603-608.

29. Hagiwara, M.; Mitsui, H.; Machi, S.; Kagiya, T. Kinetics of the γ-radiation-induced polymerization of ethylene in liquid carbon dioxide. *J. Polym. Sci. A-Polym. Chem.* 1968, 6, 609-621.

© 2009 by the authors; licensee Molecular Diversity Preservation International, Basel, Switzerland. This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).