A Silent Public Health Threat: Emergence of Mayaro Virus and Co-infection with Dengue in Peru

Miguel Angel Aguilar-Luis  
UPC: Universidad Peruana de Ciencias Aplicadas  

Juana del Valle Mendoza  
University Peruana de Ciencias Aplicadas  https://orcid.org/0000-0002-6011-5040

Isabel Sandoval  
UPC: Universidad Peruana de Ciencias Aplicadas  

Wilmer Silva-Caso  
UPC: Universidad Peruana de Ciencias Aplicadas  

Fernando Mazulis  
UPC: Universidad Peruana de Ciencias Aplicadas  

Hugo Carrillo-Ng  
UPC: Universidad Peruana de Ciencias Aplicadas  

Yordi Tarazona-Castro  
IIN: Instituto de Investigacion Nutricional  

Johanna Martins-Luna  
UPC: Universidad Peruana de Ciencias Aplicadas  

Ronald Aquino-Ortega  
IIN: Instituto de Investigacion Nutricional  

Isaac Peña-Tuesta  
UPC: Universidad Peruana de Ciencias Aplicadas  

Angela Comejo-Tapia  
UPC: Universidad Peruana de Ciencias Aplicadas  

Luis J. del Valle  
Universitaria de Catalunya

Research note

Keywords: Arbovirus, Alphavirus, Mayaro virus, Dengue, PCR, Peru

DOI: https://doi.org/10.21203/rs.3.rs-101988/v1

License: © This work is licensed under a Creative Commons Attribution 4.0 International License.  
Read Full License
Abstract

Objective: To describe the prevalence and clinical characteristics of MAYV infection in Piura, as well as the association of this pathogen with DENV.

Results: A total of 86/496 (17.3%) cases of MAYV were detected, of which 54 were MAYV mono-infection and 32 were co-infection with DENV, accounting for 10.9% and 6.4%, respectively. When evaluating monoinfection by MAYV the main groups were 18-39 and 40 – 59 years old, with 25.9% and 20.4% respectively. Co-infections were more common in the age group 18-39 and those >60 years old, with 34.4% and 21.9%, respectively. The clinical presentation most frequent were headaches (94.4%, 51/54) followed by arthralgias (77.8%, 42/54). During the 8-month study period the most cases were identified in the months of May (29.1%) and June (50.0%).

Introduction

Mayaro virus (MAYV) infection is a neglected tropical disease that represents an important cause of acute febrile illness in endemic areas. In 2019, PAHO declared an epidemiological alert due to the recent increase in outbreaks and the potential of this disease to become a significant public health issue [1]. The virus was first isolated from 5 patients in Trinidad and Tobago during the outbreak in the county of Mayaro in 1954 [2]. Outbreaks throughout the South American amazon basin have been reported [3], as well as imported cases in Europe [4].

MAYV is a positive-sense single-stranded RNA virus of the Alphavirus genus [4]. This group is composed of seven viruses, which share common antigenic sites: Bebaru virus, Chikungunya virus, Getah virus, Semliki Forest virus, Ross River virus, O’nyong-nyong virus, and UNA virus. The antigenic similarities between these viruses have previously shown cross-reactivity in serological tests [5–7]; for this motive such as molecular assays are required for diagnosis [8].

The first case of MAYV in Peru was reported in 1999 by Tesh et al. [9], and multiple outbreaks have been reported in the Peruvian territory throughout the years [10–12]. Peru has the 2nd highest number of cases reported in Latin America according to academic literature with 230 confirmed cases, following Brazil with 495 cases [13]. However, it is important to note that previous studies have shown that the real disease burden may be underestimated due to underreporting and misdiagnosis [14].

Alphavirus infections are mainly characterized by a broad range of non-specific signs and symptoms including fever, headache, rash, myalgias, among others [19, 20]. However, Semliki group alphaviruses have been implicated in causing arthralgias [15] and persistent incapacitating joint pain due to MAYV infection has been previously reported in a series of imported cases [16–18]. Accurate clinical differentiation between arbovirus and MAYV is a challenge for clinicians in endemic areas [21]; however, without sensitive and specific diagnostic tools, a precise etiological diagnosis cannot be easily achieved [14, 22].
A topic of great interest is the association between MAYV infection with other arboviruses, particularly Dengue virus (DENV). Sporadic cases of co-infection between MAYV and DENV have been previously reported [23, 24, 19]. However, several issues remain unanswered regarding the clinical presentation of co-infections between these pathogens. For example, it has not yet been characterized if co-infections exhibit a more specific clinical presentation or more severe symptoms. Furthermore, it has not been determined if co-infections suggest evidence of a shared vector.

Non-human primates play an important role as reservoirs for MAYV, maintaining the zoonotic cycle in the rainforest. [25]. Even though enzootic cycles are not fully characterized, canopy dwelling mosquitoes of the genus haemagogus are considered the main vectors [26–28]. Izurieta et al. [29] proposed that *Haemagogus* spp. was responsible for the sylvatic cycle of virus transmission and people living or working in the peri-urban area or forest peripheries, would provide the bridge to the urban setting.

The degree to which MAYV is adapted to infect more anthropophilic urban mosquitoes such as *A. aegypti*, *Ae. albopictus* and *Ae. scapularis* has only been evidenced in laboratories [30, 31]. However, single amino acid mutations in alphaviruses have been shown to improve adaption to mosquito species that are not normally considered the primary vector [32]. Consequently, MAYV urbanization poses an important risk to become a significant public health issue with the potential for epidemics in the same way CHIKV evolved over the years [33].

Our aim is described the prevalence and clinical characteristics of MAYV infection in Piura, as well as the association of this pathogen with DENV.

**Methods**

**Study location**

A consecutive cross-sectional study was performed in six primary heath care centers between February and September of 2016 in the district of Morropon, Piura (Figure S1). The department of Piura is located in the northern coast of Peru and this region is recognized as an endemic region for different arboviral diseases including DENV, as well as other etiologies of AFI, such as leptospirosis.

**Study subjects**

Patients with a suspected acute febrile illness (AFI) who attended the healthcare facilities within 7 days since the onset of quantified or not quantified patient-reported fever, were included. For patients in whom fever could be corroborated, a temperature greater than 38 °C for < 7 days was required without an identifiable source of infection and associated with one or more of the following signs and symptoms: headache, myalgia, arthralgia, retro-ocular pain, lower back pain, rash, hyperoxia, odynophagia, nausea, emesis, abdominal pain, asthenia, syncope, hypothermia, jaundice, and others. The exclusion criteria were patients with an incomplete record of their medical data and patients with an identifiable source of
infection, such as acute upper respiratory tract infections, pneumonia, and urinary tract infections, among others.

**Ethics statement**

The study protocol was approved by the Research Ethics Board of the Hospital Regional Docente de Cajamarca, Peru. The samples were obtained in the context of the epidemiological/syndromic surveillance program according to the health directives of the National Center for Epidemiology, Disease Control and Prevention of the Ministry of Health of Peru. Therefore, it was exempt from informed consent.

**Samples**

A total of 496 samples were collected using Vacuette TUBE Serum Separator Clot Activator (Vacuette; Greiner Bio-One, Kremsmünster, Austria). All the samples were stored at -80 °C after collection for molecular assays.

**Real-time reverse transcriptase PCR amplification for the detection of MAYV and DENV**

RNA extraction was performed using the High Pure RNA Isolation Kit (Roche Applied Science, Mannheim, Germany) following the manufacturer’s instructions; 200 µl of the serum samples was used. The viral RNA obtained was stored at − 80 °C until use.

Amplification by Real-time RT-PCR assay for the detection of MAYV was carried out using the primers and PCR conditions described by Aguilar-Luis et al. [34]. Amplification by Real-time RT-PCR assay for DENV was described by Leparc-Goffart et al. [35], and the PCR conditions were described by Alva-Urcia et al. [14].

**Statistical analysis**

Qualitative variables were reported as frequencies and percentages. All analyses were processed with the IBM Statistical Package for the Social Sciences (SPSS) software version 21.0 (SPSS, Chicago, IL, USA).

**Results**

Between February and September of 2016, 496 patients with suspected AFI were included in this study. Blood samples were confirmed with RT-PCR amplification of nsP1 for MAYV RNA, followed by confirmatory DNA sequencing. A total of 86/496 (17.3%) cases of MAYV were detected by this assay and among these: 54 were MAYV mono-infection and 32 were co-infection with DENV, accounting for 10.9% and 6.4%, respectively.

Table 1 summarizes the demographic characteristics of all the patients included in this study. The main age groups diagnosed with Mayaro virus were the patients between 18–39 and 40–59 years old, with 29.1% and 18.6% respectively. When evaluating monoinfection by MAYV, the main groups were 18–39 and 40–59 years old, with 25.9% and 20.4% respectively. Co-infections were more common in the age group 18–39 and those > 60 years old, with 34.4% and 21.9%, respectively.
### Table 1
Demographics in patients with arboviral acute febrile illness by MAYV and DENV.

| Characteristics | Total cases | MAYV | Co-infections |
|-----------------|-------------|------|---------------|
|                 | n = 496 (%) | n = 86 (%) | Only MAYV n = 54 (%) | DENV & MAYV n = 32 (%) |
| **Age (años)**  |             |      |               |
| < 5             | 39 (7.9)    | 8 (9.3) | 5 (9.3) | 3 (9.4) |
| 5 a 11          | 68 (13.7)   | 12 (14.0) | 8 (14.8) | 4 (12.5) |
| 12–17           | 56 (11.3)   | 10 (11.6) | 8 (14.8) | 2 (6.3) |
| 18–39           | 146 (29.4)  | 25 (29.1) | 14 (25.9) | 11 (34.4) |
| 40–59           | 109 (22.0)  | 16 (18.6) | 11 (20.4) | 5 (15.6) |
| ≥ 60            | 78 (15.7)   | 15 (17.4) | 8 (14.8) | 7 (21.9) |
| **Gender**      |             |      |               |
| Male            | 226 (45.6)  | 43 (50.0) | 30 (55.6) | 13 (40.6) |
| Female          | 270 (54.4)  | 43 (50.0) | 24 (44.4) | 19 (59.4) |

In regard to the clinical presentation, the most frequent symptom in MAYV infection were headaches (94.4%, 51/54) followed by arthralgias (77.8%, 42/54) (Table 2).
Table 2
Clinical symptoms in patients with MAYV and DENV infection confirmed by PCR.

| Síntomas Clínicos          | Casos totales n = 496 (%) | MAYV n = 86 (%) | Co-infections |
|----------------------------|---------------------------|----------------|---------------|
|                            |                           | Only MAYV n = 54 (%) | DENV & MAYV n = 32 (%) |
| Headache                   | 404 (81.5)                | 77 (89.5)       | 51 (94.4)     | 26 (81.3)   |
| Arthralgia                 | 357 (72.0)                | 68 (79.1)       | 42 (77.8)     | 26 (81.3)   |
| Myalgia                    | 378 (76.2)                | 68 (79.1)       | 41 (75.9)     | 27 (84.4)   |
| Retro-ocular pain          | 306 (61.7)                | 62 (72.1)       | 41 (75.9)     | 21 (65.6)   |
| Hyporexia                  | 305 (61.5)                | 56 (65.1)       | 36 (66.7)     | 20 (62.5)   |
| Lumbar pain                | 246 (49.6)                | 45 (52.3)       | 29 (53.7)     | 16 (50)     |
| Nausea / Emesis            | 226 (45.6)                | 41 (47.7)       | 25 (46.3)     | 16 (50)     |
| Odinophagia                | 171 (34.5)                | 31 (36.0)       | 21 (38.9)     | 10 (31.3)   |
| Cutaneous eruption         | 81 (16.3)                 | 11 (12.8)       | 6 (11.1)      | 5 (15.6)    |
| Chest pain or dyspnea      | 5 (1.0)                   | 3 (3.5)         | 2 (3.7)       | 1 (3.1)     |
| Platelet decrease          | 6 (1.2)                   | 2 (2.3)         | 1 (1.9)       | 1 (3.1)     |
| Thoracic pain / dyspnea    | 20 (4.0)                  | 2 (2.3)         | 2 (3.7)       | 0 (0)       |
| Epistaxis                  | 8 (1.6)                   | 2 (2.3)         | 2 (3.7)       | 0 (0)       |
| Gingivorrhagia             | 3 (0.6)                   | 2 (2.3)         | 1 (1.9)       | 1 (3.1)     |
| Hematocrit increase        | 5 (1.0)                   | 2 (2.3)         | 1 (1.9)       | 1 (3.1)     |
| Petechiae                  | 11 (2.2)                  | 2 (2.3)         | 0 (0)         | 2 (6.3)     |
| Excessive decay or fainting| 3 (0.6)                   | 1 (1.2)         | 1 (1.9)       | 0 (0)       |
| Sudden decrease in T ° or hypothermia | 3 (0.6) | 1 (1.2) | 1 (1.9) | 0 (0) |
| Cold Extremities / cyanosis| 2 (0.4)                   | 1 (1.2)         | 1 (1.9)       | 0 (0)       |
| Hypotension                | 1 (0.2)                   | 1 (1.2)         | 1 (1.9)       | 0 (0)       |
| Conjunctival injection     | 3 (0.6)                   | 1 (1.2)         | 0 (0)         | 1 (3.1)     |
| Fast and weak pulse        | 1 (0.2)                   | 1 (1.2)         | 1 (1.9)       | 0 (0)       |
| Cough                      | 1 (0.2)                   | 1 (1.2)         | 0 (0)         | 1 (3.1)     |
| Persistent vomiting        | 5 (1.0)                   | 1 (1.2)         | 1 (1.9)       | 0 (0)       |
| BP differential < 20 MMHg  | 1 (0.2)                   | 0 (0.0)         | 0 (0)         | 0 (0)       |
Finally, an assessment of the monthly distribution of the MAYV cases detected during the 8-month period of study was performed (Fig. 1). During the 8-month study period the most cases were identified in the months of May (29.1%) and June (50.0%).

### Discussion

An important number of cases of MAYV were detected in patients with AFI that attended outpatient health-centers during this study. Approximately, a fifth of the total cases were diagnosed with MAYV, demonstrating that this pathogen may be circulating during this region and that peri-urban transmission may be ongoing. Moreover, approximately 6.4% of the total cases were co-infection between MAYV and DENV.

The association between MAYV and DENV is a topic of great interest for the scientific community, as both pathogens share common characteristics and can co-exist in a specific region. However, only two studies have previously reported co-infections of DENV and MAYV in the scientific literature, a child with an unspecified febrile illness, serologically diagnosed with MAYV, [24], and during an outbreak in Brazil [23]. Our study reports the first clinical characterization of patients co-infected with MAYV and DENV during surveillance of AFI near the Peruvian Amazon basin for eight months. The frequency of unspecific symptoms among patients with mono-infection and co-infection were similar, including fever, myalgia, arthralgia, retroorbital pain, and headache. This finding further confirms the majority of symptoms amongst arboviral infections are non-specific, even when presenting as co-infections. [9, 24, 36, 37].

Pinheiro et al. [37] previously reported that the rash associated with MAYV infection appears on the fifth day of illness, suggesting its association with of humoral antibody appearance. In contrast, we found

| Síntomas Clínicos                  | Casos totales n = 496 (%) | MAYV n = 86 (%) | Co-infections | Only MAYV n = 54 (%) | DENV & MAYV n = 32 (%) |
|-----------------------------------|---------------------------|----------------|---------------|---------------------|------------------------|
| Ecchymosis                        | 2 (0.4)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
| Shaking chills                    | 3 (0.6)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
| Hemoptoic sputum                  | 1 (0.2)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
| Altered mental state (drowsiness) | 1 (0.2)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
| Gynecorrhage                      | 1 (0.2)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
| Hepatomegaly or jaundice          | 1 (0.2)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
| Dizziness                         | 1 (0.2)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
| Mane                              | 2 (0.4)                   | 0 (0.0)        | 0 (0)         | 0 (0)               | 0 (0)                  |
patients with a rash on the 3rd day of illness for MAYV, second day for co-infected patients, and the first day for DENV mono-infected patients.

Serologic evaluations for the detection of alphaviruses have shown cross-reactivity [8] as they belong to the Semliki complex serologic group [6, 7], denoting the importance of sensitive and specific molecular diagnostic methods such as RT-PCR. Previous studies report that viremia is detectable for up to 5 days post-infection [1]. However, we evidenced RT-PCR detection of MAYV up to 9 days in mono-infected patients and up to 7 days in co-infected patients. Our findings evidence that the window of detection for MAYV by RT-PCR could be longer than reported previously. Altogether, these characteristics make the RT-PCR an excellent diagnostic tool for the detection of MAYV during outbreaks.

Furthermore, our studied population showed no significant difference of positive cases between genders. A previous study determined that being male poses a risk for arboviral infections, given the higher occupational exposure [38]. Given that spillover zoonosis is considered the main source of recent arboviral outbreaks, this could have led to the peri-urban transmission of the disease [39]. Similarly, to our study, another outbreak in Brazil caused by peri-urban transmission of MAYV, showed that both genders were affected equally [23].

Additionally, we found a greater number of cases of both DENV and MAYV in May and July. This could be explained by some meteorological factors that could influence the vector expansion, behavior and biology. According to the meteorological national service (SENAMHI) the temperature and rainfall peak during the months of March and April; however, are still high during May and June, with a further decline in the later months. These factors altogether could enable a more easily transmission of the disease. Considering previous studies on the adaptability of alphaviruses to novel vectors [32], evidence of effective MAYV transmission by more urban anthropophilic mosquitoes in laboratory studies [30, 31, 35], and high aedic index reported in the studied region (aedic index 1–4%) [40], these findings may suggest that a common vector could be responsible for the transmission of both viruses during this outbreak. Further demographic and on-site vector studies are necessary to determine if the urbanization of MAYV is ongoing.

In conclusion, this study provides the first clinical characterization of patients co-infected with MAYV and DENV and also reports the first outbreak of MAYV-DENV co-infections, confirmed by molecular diagnostic methods. Our findings also provide further evidence that symptoms in co-infected patients are non-specific and that disease severity may not be associated with co-infections.

**Limitations**

Some limitations of the study are that a majority of AFI cases could not be diagnosed with a precise etiology. Another limitation is that because of the design of the study, follow up could not be performed and long-term symptoms were not evaluated. Further studies are required to better characterize the complications of these two pathogens.
Abbreviations

MAYV: Mayaro virus; DENV: Dengue virus; RT-PCR: reverse transcription polymerase chain reaction; RNA: ribonucleic acid; bp: base pairs.

Declarations

Ethics approval and consent to participate

This study was approved by the Research Ethics Board of the Hospital Regional de Cajamarca, Peru. The samples were collected within the framework of the epidemiological surveillance program of febrile syndrome in the Cajamarca Region, so according to international ethical guidelines for research related to human health prepared by CIOMS and WHO, review is not required. ethics or informed consent.

Consent to publish

All authors have given their authorization for the publication of the manuscript.

Availability of data and materials

Abstraction format used in the study and dataset are available and accessible from the corresponding author upon request in the link: https://figshare.com/s/03fd58ac3f5cc806629f

Conflicts of interest

On behalf of all authors, the corresponding author states that there are no conflicts of interest or funding related to this study

Funding

This work was supported by Incentives for Research of the Universidad Peruana de Ciencias Aplicadas, grant Nº UPC-A-058-2020, Lima-Peru. This research was supported by the Bio & Medical Technology Development Program of the National Research Foundation (NRF) funded by the Korean government (MSIT) (No. 2015M3A9B6073666).

Authors’ contributions

JdVM, LJdV, WSC and MAAL designed the study protocol. JdVM, MAAL, CPR. CTV, YTC, JML, IPT and ACT performed the PCR. JdVM, LJdV and MAAL were responsible for obtaining funding and laboratory
work supervision. MAAL, IS, FM, HCN and RAO was responsible for the clinical assessment, samples collection and database completion. JdVM, WSC and LJdV were responsible to draft the manuscript. All authors critically revised the manuscript for intellectual content. All authors read and approved the final manuscript.

Acknowledgments

We thank the staff of the health network from la DIRESA Piura, Peru.

References

1. org. 2019. Alerta Epidemiológica Fiebre De Mayaro. [online] Available at: <https://www.paho.org/sites/default/files/2019-06/2019-mayo-01-phe-alerta-epidemiologica-mayaro.pdf> [Accessed 13 July 2020].
2. R. Anderson, W.G. Downs, G.H. Wattley, N.W. Ahin, A.A. Reese Mayaro virus: a new human disease agent. II. Isolation from blood of patients in Trinidad, B.W.I Am J Trop Med Hyg, 6 (6) (1957), pp. 1012-1016
3. Patiño-Barbosa, A., Bedoya-Arias, J., Cardona-Ospina, J. and Rodriguez-Morales, A., 2016. Bibliometric assessment of the scientific production of literature regarding Mayaro. Journal of Infection and Public Health, 9(4), pp.532-534
4. Ganjian, N. and Riviere-Cinnamond, A., 2020. Mayaro virus in Latin America and the Caribbean. Revista Panamericana de Salud Pública, 44, p.1
5. Acosta-Ampudia, D.M. Monsalve, Y. Rodriguez, Y. Pacheco, J.M. Anaya, C. Ramirez-Santana Mayaro: an emerging viral threat? Emerg Microbes Infect, 7 (1) (2018), p. 163
6. Hassing RJ, Leparc-Goffart I, Tolou H, van Doornum G, van Genderen PJ. Cross-reactivity of antibodies to viruses belonging to the Semliki forest serocomplex. Euro Surveill. 2010;15(23):19588. Published 2010 Jun 10.
7. Prat C, Flusin O, Panella A, Tenebray B, Lanciotti R, Leparc-Goffart I. Evaluation of Commercially Available Serologic Diagnostic Tests for Chikungunya Virus. Emerging Infectious Diseases. 2014;20(12):2129-2132
8. Fischer C, Bozza F, Merino Merino X, Pedroso C, de Oliveira Filho E, Moreira-Soto A et al. Robustness of Serologic Investigations for Chikungunya and Mayaro Viruses following Coemergence. mSphere. 2020;5(1).
9. B. Tesh, D.M. Watts, K.L. Russell, C. Damodaran, C. Calampa, C. Cabezas, et al. Mayaro virus disease: an emerging mosquito-borne zoonosis in tropical South America Clin Infect Dis, 28 (1) (1999), pp. 67-73
10. Estudio interinstitucional desarrollado por las instituciones del Ministerio de Salud del Perú, en colaboración con el Instituto de Investigación de Enfermedades Tropicales de la Marina de los
Estados Unidos, la Universidad Nacional Mayor de San Marcos, la Universidad Peruana Cayetano Heredia. Perfil etiológico del síndrome febril en áreas de alto riesgo de transmisión de enfermedades infecciosas de impacto en salud pública en el Perú, 2000-2001. Rev. perú. med. exp. salud publica [Internet]. 2005 Jul [citado 2020 Jul 13]; 22(3):165-174

11. Forshey B, Guevara C, Laguna-Torres V, Cespedes M, Vargas J, Gianella A et al. Arboviral etiologies of acute febrile illnesses in western south america, 2000–2007. PLoS Neglected Tropical Diseases. 2010;4(8):e787

12. Halsey ES, Siles C, Guevara C, et al. Mayaro virus infection, Amazon Basin region, Peru, 2010-2013. Emerg Infect Dis. 2013;19(11):1839-1842.

13. Ganjian N, Riviere-Cinnamond A. Mayaro virus in Latin America and the Caribbean. Revista Panamericana de Salud Pública. 2020;44:1

14. Alva-Urcia, M.A. Aguilar-Luis, C. Palomares-Reyes, W. Silva-Caso, L. Suarez-Ognio, P. Weilg, et al. Emerging and reemerging arboviruses: a new threat in Eastern Peru PLoS One, 12 (11) (2017), Article e0187897

15. Levi L, Vignuzzi M. Arthritogenic Alphaviruses: A Worldwide Emerging Threat?. Microorganisms. 2019;7(5):133.

16. de Thoisy B, Gardon J, Salas R, Morvan J, Kazanji M. Mayaro Virus in Wild Mammals, French Guiana. Emerging Infectious Diseases. 2003;9(10):1326-1329

17. Karbaat j, Jonkers ah, Spence l. Arbovirus FInfections in Dutch Military Personnel Stationed in Surinam: a Preliminary Study. Trop Geogr Med. 1964;16:370-376

18. Hassing R, Leparc-Goffart I, Blank S, Thevarayan S, Tolou H, van Doormum G et al. Imported Mayaro virus infection in the Netherlands. Journal of Infection. 2010;61(4):343-345

19. Acosta-Ampudia, D.M. Monsalve, Y. Rodriguez, Y. Pacheco, J.M. Anaya, C. Ramirez-Santana Mayaro: an emerging viral threat? Emerg Microbes Infect, 7 (1) (2018), p. 163

20. L.A. Esposito, B. Fonseca Will Mayaro virus be responsible for the next outbreak of an arthropod-borne virus in Brazil? Braz J Infect Dis, 21 (5) (2017), pp. 540-544

21. Paniz-Mondolfi A, Rodriguez-Morales A, Blohm G, Marquez M, Villamil-Gomez W. ChikDenMaZika Syndrome: the challenge of diagnosing arboviral infections in the midst of concurrent epidemics. Annals of Clinical Microbiology and Antimicrobials. 2016;15(1).

22. Silva-Caso, M.A. Aguilar-Luis, C. Palomares-Reyes, F. Mazulis, C. Weilg, L.J. Del Valle, et al. First outbreak of Oropouche fever reported in a non-endemic western region of the Peruvian Amazon: molecular diagnosis and clinical characteristics Int J Infect Dis, 83 (June) (2019), pp. 139-144

23. Zuchi N, Heinen L, Santos M, Pereira F, Slhessarenko R. Molecular detection of Mayaro virus during a dengue outbreak in the state of Mato Grosso, Central-West Brazil. Memórias do Instituto Oswaldo Cruz. 2014;109(6):820-823.

24. Lednicky J, De Rochars V, Elbadry M, Loeb J, Telisma T, Chavannes S et al. Mayaro Virus in Child with Acute Febrile Illness, Haiti, 2015. Emerging Infectious Diseases. 2016;22(11):2000-2002.
25. de Thoisy B, Gardon J, Salas R, Morvan J, Kazanji M. Mayaro Virus in Wild Mammals, French Guiana. Emerging Infectious Diseases. 2003;9(10):1326-1329.

26. Neumayr A, Gabriel M, Fritz J, Günther S, Hatz C, Schmidt-Chanasit J et al. Mayaro Virus Infection in Traveler Returning from Amazon Basin, Northern Peru. Emerging Infectious Diseases. 2012;18(4):695-696

27. Powers a, Chandler I, Tesh r, Russell k, Watts d, da Rosa a et al. Genetic relationships among mayaro and una viruses suggest distinct patterns of transmission. The American Journal of Tropical Medicine and Hygiene. 2006;75(3):461-469

28. Hoch A, LeDuc J, Pinheiro F, Peterson N. An Outbreak of Mayaro Virus Disease in Belterra, Brazil. The American Journal of Tropical Medicine and Hygiene. 1981;30(3):689-698.

29. Izurieta R, DeLacure D, Izurieta A, Hoare I, Reina Ortiz M. Mayaro virus: the jungle flu. Virus Adaptation and Treatment. 2018;Volume 10:9-17.

30. Moore C. Aedes albopictus in the United States: Ten-Year Presence and Public Health Implications. Emerging Infectious Diseases. 1997;3(3):329-334.

31. Long K, Tesh R, Higgs S, Haußer N, Thangamani S, Kochel T et al. Experimental Transmission of Mayaro Virus by Aedes aegypti. The American Journal of Tropical Medicine and Hygiene. 2011;85(4):750-757.

32. Tsetsarkin K, Vanlandingham D, McGee C, Higgs S. A Single Mutation in Chikungunya Virus Affects Vector Specificity and Epidemic Potential. PLoS Pathogens. 2007;3(12):e201.

33. Mackay I, Arden K. Mayaro virus: a forest virus primed for a trip to the city?. Microbes and Infection. 2016;18(12):724-734.

34. Leparc-Goffart I, Baragatti M, Temmam S, Tuiskunen A, et al. Development and validation of real-time one-step reverse transcription-PCR for the detection and typing of dengue viruses. J Clin Virol. 2009 May;45(1):61-6.

35. Miguel Angel Aguilar-Luis, Juana del Valle-Mendoza, Wilmer Silva-Caso, Tamara Gil-Ramirez, Saul Levy-Blitchtein, Jorge Bazán-Mayra, Victor Zavaleta-Gavidia, Daniel Cornejo-Pacherres, Carlos Palomares-Reyes, Luis J. del Valle. An emerging public health threat: Mayaro virus increases its distribution in Peru, International Journal of Infectious Diseases, Volume 92, 2020, Pages 253-258.

36. Tesh RB Arthritides caused by mosquito-borne viruses. Annu Rev Med 1982;33:31–40 10.1146/annurev.me.33.020182.000335

37. Pinheiro F, Freitas R, da Rosa J, LeDuc J, Gabbay Y, Mello W. An Outbreak of Mayaro Virus Disease in Belterra, Brazil. The American Journal of Tropical Medicine and Hygiene. 1981;30(3):674-681.

38. Forshey BM, Guevara C, Laguna-Torres VA, Cespedes M, Vargas J, Gianella A, et al. Arboviral etiologies of acute febrile illnesses in Western South America, 2000–2007. PLoS Negl Trop Dis. 2010;4:e787. 10.1371/journal.pntd.0000787

39. Kreuder Johnson C, Hitchens P, Smiley Evans T, Goldstein T, Thomas K, Clements A et al. Spillover and pandemic properties of zoonotic viruses with high host plasticity. Scientific Reports. 2015;5(1).
40. Sala de Situación de Salud – Semana Epidemiológica N° 11 2017 (Internet). Lima: Centro Nacional de Epidemiología, Prevención y Control de Enfermedades - Ministerio de Salud; 2017 (cited on 19th July 2020) Available at: http://www.dge.gob.pe/portal/docs/vigilancia/sala/2016/salaSE52.pdf