Antioxidant and Antidiabetic Activities of Biologically Synthesized Silver Nanoparticles using Linum usitatissimum Extract

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ABSTRACT

This research work is mainly focused to study the anti-oxidant and anti-diabetic activities of biologically synthesis of silver nanoparticles (AgNPs) from the flaxseed extract of Linum usitatissimum. Qualitative tests identify the presence of phytochemicals in the flax seed extract and its results showed the presence of tannins, terpenoids, saponins, flavonoids, steroids, cardiac glycosides, anthraquinones, coumarins, xanthoproteins, alkaloids, emodins, and carbohydrate in it. Preliminarily AgNPs formation is confirmed by the UV-spectra and it showed maximum adsorption band at 438nm. FT-IR spectroscopic studies reveal the Phyto-constituents which are involved in the reduction of silver (Ag⁺) into silver nanoparticles (Ag⁰). The spherical shapes of AgNPs are observed with crystalline nature are found in the aid of SEM and XRD analysis. Synthesized AgNPs have the maximum percentage of a silver element which is examined by the EDX analysis. The in vitro antioxidant and antidiabetic activities of L. usitatissimum mediated AgNPs were analyzed by using the DPPH, alpha-amylase, and alpha glycosides assays respectively. The DPPH result shows that the AgNPs possess 59.01% of radical scavenging property and the standard ascorbic acid reveals 48.63% at 100 µg/mL concentration. Similarly in anti-diabetic activity, AgNPs shows the maximum inhibition of 79.84% in the alpha-amylase assay, and for alpha-glucosidase, AgNPs showed 58.86% at 100 µg/ml concentration.

Keywords: Linum usitatissimum, Silver nanoparticles, Anti-oxidant activity, Anti-diabetic activity.

INTRODUCTION

Nanoscience technology is a multidisciplinary subject and it is widely used in research areas of physics and chemistry. Various physical, chemical, and greener syntheses processes are available in nanoparticle (NPs) production. Chemical and physical methods are more labor-intensive and also possess hazardous than the biological (green method) synthesis. The greener process is non-toxic, reproducible, eco-friendly, one step, easy, and more efficient. In additionally the greener method consumes low energy, it produces harmless and stable products. In the greenway, the plant extract-based nanoparticles synthesize get more attention than the other green methods (fungus, algae, bacteria, etc.).
The most viable nanomaterial is silver nanoparticles (AgNPs) which are more commercialized nano-materials and nearly 500 tons of AgNPs were produced per year. Owing to high sensitivity in the detection of biomolecules, medicine, and catalysis silver nanoparticles were recognized as a strong pharmacophore. A wide range of medical applications are revealed by the silver nanoparticles (AgNPs) and are wound healing, dental material fills, bone repairing, anti-diabetics, vaccine adjuvants, bio-imaging, and biosensors. Flax or Linum usitatissimum is commonly known as the linseed which is the flowering plant, and it belongs to the Linaceae family. Traditionally it is used in the treatment of gastrointestinal infections and diarrhea. Flaxseed has macro and micronutrients along with that it has an excess of omega-3 fatty acids and vitamin B6. It has 54% of omega-3 fatty acids, 18% of oleic acid (omega-9 fatty acids), 6% of linoleic acid (omega-6 fatty acids), 9% of saturated fat, and 5% of palmitic acid. Diabetes mellitus is an endocrine disorder. Due to insulin resistance and deficiency which results from the high level of sugar in the blood for an extended period which is referred to as diabetes mellitus. This research work mainly aimed to study the In vitro of anti-oxidant and anti-diabetic activities of green synthesized silver nanoparticles mediated from Linum usitatissimum.

MATERIALS AND METHODS

Plant collection and extraction

The fresh flax seeds were collected from the Thiruvarur district, Tamilnadu, India. Disease-free seeds were washed thoroughly with the running water several times. Then again seeds were washed by double distilled water to remove other water-soluble impurities. After that seeds were dried in sunshade then made into a fine powder using the mechanical grinder. 100 g of seed powder was taken in a 500 mL dry beaker and to that 300 mL of ethanol was added. Then heated on the water bath, the final extract is filtered through the Whatman No.41 filter paper. Filtered extract was stored in the brown bottle which is used for further analysis.

Qualitative and Quantitative analysis of Aqueous Extract of Linum usitatissimum

The various phytochemical tests (qualitative) were performed to establish the profile of the Linum usitatissimum extract. Primary and secondary metabolites presence was tested by the screening tests against the metabolites such as tannins, carbohydrates, terpenoids, phenolics, anthocyanins, and flavonoids, etc., Quantification of these metabolites can be performed by the standard protocols which estimate the total amount of Flavonoids, Tannins, Saponins, Alkaloids, Phenol and Terpenoids in the seed extract.

Synthesis and Characterisation of AgNPs using Linum usitatissimum Extract

1mM concentration of silver nitrate is prepared in a 50 mL standard flask, and the flask was covered with aluminum foil to prevent photochemical reactions with sunlight. 50 mL of 1mM silver nitrate solution was taken in a 300 mL beaker to this 50 mL of seed extract is slowly added then kept on a stirrer for 5 hours. After that, the reaction mixture was kept at room temperature for 24 h to complete the reaction. Then formed nanoparticles were collected by centrifuging at 6000rpm. Collected AgNPs were washed with double distilled water and followed by ethanol several times to remove impurities from that. Subsequently, AgNPs were dried in a hot air oven for 6 to 12 h to achieve high purity of obtained AgNPS. The formation of any nanoparticles is initially confirmed by the UV-Visible spectral studies. The same AgNPs formation is identified by analyzing 2 mL of colloidal AgNPs on UV-Visible. A small quantity of dried AgNPs was taken for the FT-IR analysis in the range of 500 cm⁻¹ to 4000 cm⁻¹ to identify the functional groups present in the extract which involves in the AgNPs synthesis. Similarly, the morphology of synthesized AgNPs was analyzed by SEM (1-10nm), Elemental composition by EDX (1-10KeV), and Crystallinity by XRD (10°<2θ<80°) studies.

In vitro antioxidant activity of AgNPs by dPPH method

The 20, 40, 60, 80, and 100 µg/mL of AgNPs were taken in the five different test tubes then 1 mL of ethanol was added and mixed well. To each test tubes, 3 mL of 0.1mM DPPH was added and kept in a dark room for 30 minutes. After the reaction time interval, the absorbance of each mixture was recorded at 517nm using the spectrophotometer.
Blank served as without adding NPs. From the below equation the percentage of inhibition of DPPH scavenged is calculated.

\[
\text{DPPH activity (\% inhibition)} = \left(\frac{A - B}{A}\right) \times 100
\]

A and B represents the absorbance value for the test samples (NPs added DPPH) and blank sample. The percent inhibition versus concentration curve and 50% inhibition was determined from the concentration verse inhibition percentage graph.\(^{13}\)

**In vitro antidiabetic activity of AgNPs by Alpha-amylase inhibition assay**

The alpha-amylase inhibitory protocol was given by McCue and Shetty. Five different concentration of AgNPs (20, 40, 60, 80 and 100 µg/mL) was taken in a test tube and 1 mL of ethanol added to each tube. To this 250 µL of sodium phosphate buffer (pH 6.9) containing α-amylase solution (0.5mg/mL) is added and incubated for 10 min at room temperature. Over 250 µL of 1% starch in 0.02M sodium phosphate buffer (pH 6.9) was added again incubated for 10 minutes. To that 500 µL of dinitrosalicylic acid (DNS) reagent is added slowly and heated on a water bath for 5 min and their absorbance was measure at 540nm in a spectrophotometer by diluting and cooling the samples. The α-amylase inhibition percentage was calculated by the below equations.

\[
\%\text{Inhibition} = \left(\frac{\text{Abs control} - \text{Abs sample}}{\text{Abs control}}\right) \times 100
\]

Where Abs control and Abs of the sample represent the absorbance value for the test and blank sample respectively.

**In vitro antidiabetic activity of AgNPs by Alpha glucosidase inhibition assay**

The α-glucosidase protocol was initiated by Saccharomyces cerevisiae. Five different concentration of AgNPs (20, 40, 60, 80 and 100 µg/mL) was taken in a test tube and 1 mL of ethanol added to each tube. 100 µL of α-glucosidase is added to each test tube and incubated for 10 minutes. To that 50 µL of 3mM P-nitro phenyl glucopyranoside was added followed by incubation for 20 min at 37°C. Two milliliters of 0.1M sodium carbonate were added then the absorbance of each mixture is spectrophotometrically measured at 405nm by spectrophotometer. Blank was performed similarly except for the addition of samples. The inhibition percentage of α-glucosidase was calculated by the below equation.

\[
\%\text{Inhibition} = \left(\frac{\text{Abs control} - \text{Abs aqueous}}{\text{Abs control}}\right) \times 100.
\]

Where A and B represent the absorbance value for the test and blank sample respectively.\(^{14,15}\)

**RESULTS AND DISCUSSION**

**Qualitative results of Linum usitatissimum seed extract**

The results of phytochemical screening of *Linum usitatissimum* seed extract on different qualitative tests are represented in Table 1 and Fig. 2. The screening results revealed that the seed extract shows the presence of tannins, saponins, flavonoids, steroids, terpenoids, cardiac glycosides, anthraquinones, coumarins, xanthoproteins, alkaloids, emodin, carbohydrates which are responsible for medicinal activities while leucoanthocyanin, phlobatannin, and phenol were absent.

**Table 1: Qualitative analysis of seed of Linum usitatissimum**

| S.No | Phytocompounds        | Observation      | Results |
|------|-----------------------|------------------|---------|
| 1    | Terpenoids            | No precipitate   | Absent  |
| 2    | Flavanoid             | Yellow color     | ++      |
| 3    | Saponin               | Forth form       | +++     |
| 4    | Tannin                | Brownish green   | +++     |
| 5    | Alkaloid              | Yellow color     | +++     |
| 6    | Steroids              | No precipitate   | Absent  |
| 7    | Glycosides            | No precipitate   | Absent  |
| 8    | Phlobatannins         | No precipitate   | Absent  |
| 9    | Proteins              | White precipitate| ++      |
| 10   | Coumarin              | Yellow color     | +++     |
| 11   | Emodin                | Red color        | +++     |
| 12   | Anthraquinone         | Red              | +++     |
| 13   | Carbohydrates         | Reddish violet color | +++     |
| 14   | Leucoanthocyanin      | No precipitate   | Absent  |
| 15   | Cardiac glycosides    | Brown ring       | ++      |
| 16   | Anthocyanin           | Bluish violet    | +++     |
| 17   | Xanthoproteins        | Reddish orange precipitate | +++ |
| 18   | Phenol                | Blue-black       | +++     |
Quantitative results of *Linum usitatissimum* seed extract

From the quantitative analysis results, the phytoconstituents composition in the Linum usitatissimum seed extract has been reported in different amounts. It reveals that alkaloids (0.193 mg/g) are present in higher percentage in the seed extract followed by saponins (0.089 mg/g), phenols (0.011 mg/g), tannin (0.008 mg/g), flavonoids (0.006 mg/g), and terpenoids (0.003 mg/g) which was given in Table 2 and Figure 3.

| S.No | Phytochemical Constituents | Samples(mg/g) |
|------|---------------------------|---------------|
| 1    | Flavonoids                | 0.006 mg/g    |
| 2    | Tannin                    | 0.008 mg/g    |
| 3    | Saponins                  | 0.089 mg/g    |
| 4    | Alkaloids                 | 0.193 mg/g    |
| 5    | Phenol                    | 0.011 mg/g    |
|      | Terpenoids                | 0.003 mg/g    |

Spectral characterization of AgNPs using *Linum usitatissimum* seed extract

The color changes are the main identification step in nanoparticles synthesis. For AgNPs formation after adding ethanol extract the colorless solution of silver nitrate turns into brown and it extended to dark brown. Similarly, the UV-Visible results of synthesized AgNPs showed maximum adsorption of 445nm which denotes the formation of silver nanoparticles in the resulted mixture i.e. the plasma resonance was observed in the range of 445nm which related AgNPs.

FT-IR spectrum of ethanolic extract of *L. usitatissimum*, which gives information about the functional groups of phytochemicals present in the raw extract. Observed peaks were shown in the Fig. 5 and their data was given in the Table 3.

![UV-Vis spectrum of synthesized silver nanoparticles](image)

**Table 3: FTIR band values of synthesized silver nanoparticles**

| The observed frequency of the synthesized AgNPs (cm⁻¹) | Assignment of functional group |
|-------------------------------------------------------|--------------------------------|
| Strong band at 3331                                   | O-H Stretching and H-bonded   |
| Strong band at 2358                                   | C-N in aliphatic amine         |
| Strong band at 1639                                   | C-O stretching in aliphatic ether |
| Strong band at 1462                                   | C-F stretching in fluoro compound |
| Broadband at 659                                     | -C=C- in alkenes.             |
| A medium band at 599                                  | Medium C=C stretching conjugated alkenes |
| Strong band at 557                                   | C-Br stretching in Halo compound |

![FT-IR spectrum of synthesized silver nanoparticles](image)

The SEM images of synthesized AgNPs were given in the Fig. 6. The average size of obtained particles was found to be 82.34nm, and it was in needle shapes with well dispersed and almost uniform in nature. Mostly in the greener method, spherical shapes of NPs were observed but for the first time, needle shapes of AgNPs were reported in the green method.

Through Energy dispersive X-ray (EDX) analysis the elemental composition of AgNPs was studied and their signals (from Table 4 and Fig. 7) confirms the presence of elemental silver in it. In this...
59% of silver was present in the formed AgNPs and similarly, it has 25% of oxygen, 12% of copper, and 3% of sulfur.

Table 4: Elemental composition of synthesized AgNPs

| Element | Wt%  | At%  |
|---------|------|------|
| O       | 24.82| 25.29|
| Cu      | 18.14| 12.25|
| S       | 11.23| 3.44 |
| Ag      | 45.81| 59.02|

In X-ray crystallography analysis of AgNPs was studied and its resulted spectra are shown in the Fig. 8. The results exhibit that AgNPs are in crystalline form with the intense diffraction peaks at 2Θ values of 38.35, 45.63, 67.14, and 78.28 which are correspond to the plane of (111), (200), (220), and (311) respectively. The observed AgNPs patterns were in face-centered cubic which has a similar pattern of JCPDS, File No. 04-0783. Few unassigned peaks were found in the XRD results which may be from ethanolic extract of *L. usitatissimum* (because of the presence of phytochemicals).

Antioxidant activity of synthesized AgNPs by DPPH assay

The result depicts that the synthesized AgNPs showed maximum potent antioxidant activities at high concentrations as compared with standard ascorbic acid Table 5. The synthesized silver nanoparticles showed 59.01% of inhibition at the concentration of 100 µg/mL while ascorbic acid has 48.63% at the same concentration which was shown in Fig. 9. AgNPs reduces more DPPH than the ascorbic acid which showed higher anti-oxidant activity than the standard.

Table 5: Antioxidant activity of synthesized silver nanoparticles using *Linum usitatissimum* seed by DPPH assay

| S. No | Concentrations | Scavenging Effect (%) |
|-------|----------------|-----------------------|
|       | Silver Nanoparticles | Ascorbic Acid |
| 1     | 20 (µg/mL)      | 37.81±0.91            | 12.78±0.34 |
| 2     | 40 (µg/mL)      | 40.57±0.41            | 16.56±0.21 |
| 3     | 60 (µg/mL)      | 48.13±0.78            | 23.08±0.45 |
| 4     | 80 (µg/mL)      | 57.23±0.45            | 30.12±0.28 |
| 5     | 100 (µg/mL)     | 59.01±0.12            | 48.63±0.34 |

*In vitro* alpha-amylase inhibitory assay

In this study, *In vitro* alpha-amylase, inhibitory activities of *L. usitatissimum* seed extract were investigated and their results are given in Fig. 10. The results showed that a dose-dependent activity occurs which increases the percentage of inhibitory activity against the alpha-amylase enzyme. The synthesized AgNPs showed the inhibitory
activity from 61.03% to 79.84% at concentrations of 20-100 µg/mL (Table 6 and Fig. 10). Acarbose is a standard drug for α-amylase inhibitor which showed α-amylase inhibitory activity from 40.12 to 57.01% at the same concentrations. While comparing the results synthesized AgNPs reveal a greater inhibitory effect than the standard.

Table 6: *In vitro* α-amylase activity of synthesized silver nanoparticles vs standard acarbose

| S.No | Concentrations | Synthesized Silver Nanoparticles | Acarbose       |
|------|----------------|----------------------------------|----------------|
| 1    | 20 (µg/mL)     | 61.03±0.03                       | 40.12±0.98     |
| 2    | 40 (µg/mL)     | 66.08±0.17                       | 43.56±0.95     |
| 3    | 60 (µg/mL)     | 68.01±0.23                       | 48.21±0.55     |
| 4    | 80 (µg/mL)     | 74.09±0.11                       | 52.32±0.67     |
| 5    | 100 (µg/mL)    | 79.84±0.14                       | 57.01±0.33     |

Fig. 10. Antidiabetic activity of synthesized silver nanoparticles using *Linum usitatissimum* plant extracts by an alpha-amylase method

**In vitro α-glucosidase inhibitory assay**

The results of antidiabetic activity using α-glucosidase inhibitory assay of synthesized AgNPs using a *Linum usitatissimum* seeds extract are shown in Table 7 and Fig. 11. The percentage inhibition at 20-100 µg/mL concentration of AgNPs showed a dose-dependent activity with the inhibition varied from 43.34% to 58.86% from highest concentration to the lowest concentration. While comparing the activity of AgNPs with the acarbose at higher concentration (100 µg/mL) AgNPs showed 58.86% of inhibition but the standard reveals 44.06% of inhibition against the α-glucosidase inhibitory assay. Thus, the inhibitory of α-glucosidase by AgNPs would delay the degradation of carbohydrates, which causes a decrease in the absorption of glucose, as a result, the reduction of postprandial blood glucose level elevation.

Table 7: *In vitro* alpha-glucosidase activity of synthesized silver nanoparticles vs standard acarbose

| S. No | Concentrations | Synthesized Silver Nanoparticles | Acarbose       |
|-------|----------------|----------------------------------|----------------|
| 1     | 20 (µg/mL)     | 43.34±0.45                       | 31.12±0.44     |
| 2     | 40 (µg/mL)     | 47.78±0.18                       | 36.34±0.56     |
| 3     | 60 (µg/mL)     | 49.90±0.11                       | 38.67±0.74     |
| 4     | 80 (µg/mL)     | 54.01±0.45                       | 41.02±0.09     |
| 5     | 100 (µg/mL)    | 58.86±0.90                       | 44.06±0.05     |

Fig. 11. Antidiabetic activity of synthesized silver nanoparticles using *Linum usitatissimum* plant extracts by an alpha-glucosidase method

**CONCLUSION**

In this study, silver nanoparticles were successfully performed by the green method using silver nitrate and *L. usitatissimum* extract used as the reducing agent. The aqueous extract of *L. usitatissimum* showed the great capability to synthesize the silver nanoparticles. The present study for the first time reports the needle shapes of silver nanoparticles in a greener way. The synthesized nanoparticle has 59% of elemental silver with noticeable activities of anti-oxidant and anti-diabetic. This study suggests that *L. usitatissimum* mediated silver nanoparticles can be alternate antioxidants and anti-diabetic agents for the treatments which is less toxic and more effective than the existing one.

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**Conflicts of interest**

The authors have declared no conflicts of interest.

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