Circulating Endothelial Progenitor Cells in Cerebrovascular Disease

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Stroke is associated with high disability and mortality burdens worldwide, but there are few effective and widely available therapies. There is therefore a need to develop treatments that promote the repair and regeneration of ischemic brain tissue. In this regard, a population of adult stem cells-called endothelial progenitor cells (EPCs)-has been identified in peripheral blood that could provide novel approaches in regenerative medicine for curing patients with acute ischemic stroke. There is accumulating evidence that EPCs can repair damaged endothelia and attenuate the development and progression of atherosclerosis. Also, EPCs can be recruited in response to acute ischemic events and participate in reparative vasculogenesis. Most studies related to EPCs have involved patients with cardiovascular diseases, and there is emerging evidence that EPCs represent a risk marker and a potential therapeutic agent in cerebrovascular disease. Here we review the characteristics and biology of EPCs in cerebrovascular disease and discuss the challenges that must be addressed to clarify the role and therapeutic applicability of EPCs in cerebrovascular disease.

Key Words endothelial progenitor cells, cerebrovascular disease, stroke, atherosclerosis, regeneration.

Introduction

Stroke is an important cause of mortality and disability worldwide, with an associated high socioeconomic impact, but the only effective curative therapeutic approaches are thrombolytic treatments, which have narrow time windows and limited availabilities. Furthermore, stroke eventually leads to tissue necrosis and possibly to irreversible impairment of brain function. Therefore, repair processes after cerebral ischemia should be investigated in order to develop therapeutic strategies for promoting neurorecovery. Endothelial repair and neovascularization are possible in the adult brain recovering from ischemic stroke, and could include both angiogenesis and vasculogenesis-combining these two processes could be one of the most promising therapeutic strategies for stroke.

Endothelial progenitor cells (EPCs) are a type of adult stem cell that has been actively investigated. These immature hematopoietic endothelial cells circulate in peripheral blood (PB). EPCs counteract ongoing risk-factor-induced endothelial cell injury, and in response to acute hypoxia are mobilized from bone marrow (BM) to PB and participate in endothelial cell repair and regeneration and also in tissue neovascularization. Experimental and human studies have shown that EPCs participate in neovascularization processes in ischemic organs, and hence their regulation could have therapeutic applications in various vascular diseases.

Increased cardiovascular risk factors and the presence of atherosclerosis are associated with dysfunction and reduced numbers of EPCs. Moreover, a low number of EPCs is an independent risk factor for future cardiovascular events. Coronary artery disease and cerebrovascular disease (CVD) are two sides of the same coin with similar etiologies that result in endothelial damage and arteriosclerosis. Although EPCs have been studied in cardiac disease and might be surrogate markers of vascular function, there have been only a few observation studies on the contribution of EPCs to CVD. Biological assays of EPCs in stroke might reveal the specific mechanisms of ischemic lesions and predict their severities and outcomes. In this review, we discuss the cur-
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ent developments in EPC research with special emphasis on CVD, including in the risks of stroke, acute stroke, and chronic stroke. We also discuss the challenges of transplanting EPCs as a treatment for stroke.

**Biology**

BM contains a mobile pool of nonhematopoietic cells that express various markers of tissue-committed stem cells that could migrate to the peripheral circulation and exert beneficial effects on regeneration. Their mobilization, recruitment, and homing mechanisms are regulated by various chemokines and cytokines, and several physiological and pathological conditions can influence the number of circulating progenitor cells. EPCs constitute a population of purified cells that originate directly from the hemangioblast, a common precursor of hematopoietic and endothelial cells. Such EPCs are mobilized during ischemia or exogenously by stimuli such as vascular endothelial (VE)-cadherin and CD133, and whilst being extremely rare in PB they are markedly increased after vascular trauma. EPCs isolated from PB differentiate into a mature endothelial phenotype based on their microscopic appearance, functional characteristics (uptake of acetylated low-density lipoprotein (acLDL) and nitric oxide synthesis), and expression of cell-surface markers (E-selectin, von Willebrand factor (vWF), VE-cadherin, platelet-endothelial cell adhesion molecule, and c-kit) associated with loss of CD133 expression.

BM cells circulating in the PB increase in number during tissue injury and are chemotactically to the ischemic tissues. It has been shown that BM progenitor cells can be released from BM/tissue niches, circulate, and finally be chemotactically to ischemic tissue in an SDF-1/CXCR4-dependent manner. In a manner characteristic of circulating BM cells, EPCs are mobilized during ischemia or exogenously by stimulation with cytokines, and contribute to neovascularization of ischemic tissues. An experimental study found that the number of CD34+ EPCs gradually increased for 7 days after a stroke and remained significantly above the prestroke baseline to day 14, returning to baseline by day 30.

EPCs can be isolated from PB in sufficient quantities for harvesting, and after their ex vivo expansion they can be administered systemically to enhance neovascularization. Thus, circulating EPCs have the potential for vascular repair after injury. Vascular progenitor cells have been shown to incorporate into areas of active vascular growth in animal models of hind-limb, myocardial, and cerebral ischemia, indicating their therapeutic potential either by providing endothelial cells for new vessel growth or through secretion of angiogenic growth factors that activate neighboring cells. The suggested regenerative potential of EPCs has prompted clinical studies of the following two hypotheses: (1) patients with lower numbers of EPCs have a higher risk of vascular diseases, and (2) patients with ischemic events would benefit from EPC transplantation.

**Characteristics and Measurement**

EPCs are maturing cells derived from immature stem cells, and hence they possess functional and structural characteristics of both stem cells and mature endothelial cells. During their development, EPCs gradually lose stem-cell characteristics and progressively gain endothelial-cell characteristics. EPCs should be quantified with caution due to their heterogeneity. Two quantification techniques have been reported. The first is flow cytometry, in which cells are labeled with fluorescent antibodies to EPC cell-surface antigens (see Table 1) and subsequently counted with a flow cytometer. The rarity of EPCs in PB makes it necessary to reduce the noise-to-signal ratio. Isolated BM-derived EPCs represent more

| Table 1. Characteristics of early endothelial progenitor cells (EPCs), endothelial outgrowth cells (EOCs), and neuronal outgrowth cells (NOCs) |
|-----------------------------------------------|
| **Morphology**                              | **Early EPCs** | **EOCs** | **NOCs** |
| CFU, spindle                                | Cobblestone    | Palisading |
| **Growth pattern in vitro**                 | Early growth   | Late outgrowth | Late outgrowth |
| **Antigen expression**                      | CD34, KDR, Tie-2, CD34, CD31, vWF, VE-cadherin, CD14, CD45 | KDR, CD31, CD36 | Nestin, Vimentin, Tuj-1, vWF, VE-cadherin, Tie-2 |
| acLDL uptake and lectin binding             | Positive       | Positive   | Negative |
| Incorporation into HUVECs                   | Good           | Better     | Possible |
| Tube formation                              | None           | Yes        | None     |
| Cytokine expression                         | High level     | Low level  | Low level |
| Population                                  | Heterogeneous  | Homogeneous | Homogeneous |
| Angiogenic potential                        | Good           | Good       | Possible |
| Neurogenic potential                        | None           | None       | Good     |

CFU: colony-forming units; HUVECs: human umbilical vein endothelial cells; vWF: von Willebrand factor; Tuj-1: beta III tubulin; Dcx: doublecortin; acLDL: acetylated low-density lipoprotein; VE-cadherin: vascular endothelial-cadherin.
immature cells, expressing the early hematopoietic marker CD133 and their phenotype is in general CD133+/CD34+/VEGFR-2+/VE-cadherin. In contrast, EPCs isolated from PB obviously lose CD133 and gradually start to express CD31, VE-cadherin, c-Kit, vWF, CD146, and CXCR4. A combination of CD34 and VEGFR2 cell-surface markers in a mononuclear population has recently been recommended as the best choice for EPC identification. The second technique is cell culture (see Fig. 1), in which PB-derived mononuclear cells (PBMNCs) are cultured for several days in conditions that selectively favor the growth of EPCs. These conditions include using gelatin-coated plates and adding endothelial growth factors to the culture medium. Since EPCs have to be viable and able to respond to the culture conditions, culture-based quantification of EPCs also depends on their function. EPCs forming colony-forming units (CFU) from the third day after plating are typically counted after 7 to 28 days. Further testing to confirm the endothelial phenotype of the cells involves the uptake of acLDL, binding of *Ulex europaeus* lectin, and binding of specific antibodies. In contrast to flow cytometry, culturing methods depend on EPC function, and indirectly measure the number of EPCs. This difference means that the results from the two widely used techniques cannot be directly compared, making some sort of standardization desirable.

Interpreting the results from the ever-increasing number of reports on EPC biology has been hampered by a lack of corroborated methods for precisely identifying EPCs. Some investigators have used flow cytometry to estimate EPC numbers, whereas others refer to the number of CFU. Since the number of circulating EPCs represents a dynamic balance between their production induced by chronic or acute stimuli and their consumption in damaged areas, performing flow cytometry at a small number of time points during acute ischemia is of limited usefulness. There is emerging evidence that EPC functional properties are better represented by the number of CFU than by the number of EPCs. Whereas EPC function is critical for endothelial maintenance and repair, the number of CFU might indicate the cumulative vascular risk in ischemia.

Since Asahara et al. first reported the existence of EPCs in PB, several studies have found significant heterogeneity among EPC populations in *in vitro* cultures. At least three types of EPC have been described (Table 1). PBMNCs contain cells termed “early EPCs” that share some endothelial but also monocytic characteristics and a restricted capacity for expansion. Recent studies have also shown the existence of a more promising population that originate from BM, circulate in PB, and whose morphology and proliferation pattern differs from the EPCs reported by Asahara et al. So-called endothelial outgrowth cells appear after 2 to 3 weeks of culture, rapidly replicate from multiple cells, and form into monolayers with a cobblestone-like morphology and a high proliferation capacity. Current data suggest that this popu-

![Fig. 1. Serial images of human peripheral-blood mononuclear cells (PBMNCs) cultured in endothelial growth media (EGM) to produce various types of progenitor cells. Phase-contrast images show the maturation of PBMNCs obtained from stroke patients during the culture immediately (A) and 7 days (B and C) after plating. Long-term cultures produced a heterogeneous population of cells (D) and led to a cobblestone (E) or palisading (E) outgrowth of cells. Scale bars: 30 μm (C), 60 μm (A and B), and 150 μm (D-F).](image_url)
lation of outgrowing cells is a subset of true EPCs deriving from BM that exhibit the potential for vascular repair after injury. More recently, we have also isolated and cultured two types of outgrowing cells obtained from the PB of patients with acute stroke.19 These cells were named endothelial or neuronal outgrowth cells (NOCs), according to their morphological characteristics and protein or gene expression profiles. Both types of outgrowing cells maintained their proliferative capacities during a culture period of 3 months. Although numerous reports have described the clinical significance of circulating EPCs, there are few data supporting their stem or progenitor status—namely, the ability to give rise to proliferating, functional endothelial cells such as outgrowing cells. We have performed studies validating the number of outgrowth cells as a potential marker representing the number of circulating progenitor cells.19,20

**EPC Profiles in Patients at Risk of CD**

Circulating EPCs could be a marker of endothelial function and cardiovascular risk. EPC numbers are significantly decreased in subjects with elevated serum cholesterol, hypertension, and diabetes, and in smokers.37,38 Measurements of flow-mediated brachial-artery reactivity also revealed a significant relation between endothelial function and the number of EPCs, supporting a role for EPCs in the maintenance of endothelial integrity.2 Endothelial cell injury and endothelial dysfunction are predictors of the risk of vascular events, providing stimuli for the development of atherosclerotic plaques.8

Consistent with this hypothesis, cardiovascular risk factors—such as smoking, arterial, and cerebral atherosclerosis—have been associated with low EPC numbers,4,8,11,15,39 which has also been shown to represent an independent risk factor for future cardiovascular events.12,13 Moreover, depletion of CD34+/KDR+ EPCs was found to be an independent predictor of early subclinical atherosclerosis in healthy subjects.39

There have been a few observation studies on the contribution of EPCs to cerebral atherosclerosis and stroke. Table 2 summarizes data from previous studies related to circulating EPCs in CVD. In patients with prior stroke, the number of circulating CD34+/KDR+ EPCs is negatively correlated with carotid intima-media thickness, and is an independent risk factor for increased carotid intima-media thickness and the presence of carotid plaques.16 Furthermore, stroke patients with lower than normal numbers of CD34+/KDR+ EPCs had a significantly greater carotid intima-media thickness and a significantly higher prevalence of carotid plaques. In addition, there was a strong negative correlation between

### Table 2. Summary of previous studies on circulating EPCs in cerebrovascular disease

| Reference          | Parameter | Stroke risk | Prognosis                                      | Risk or burden                                      | Other                |
|--------------------|-----------|-------------|------------------------------------------------|----------------------------------------------------|----------------------|
| Chu et al., 2008   | CFU       | Acute stroke < healthy controls | ND                                               | Few EPCs: many infarctions                          |                     |
| Yib et al., 2008   | CD34+/KDR+| Acute stroke < chronic stroke | Stroke with few EPCs: atherosclerosis progression | Few EPCs: high carotid IMT and plaque formation     |                     |
| Lau et al., 2007   | ND        | ND          | Large increase in EPC numbers: good outcome,   | ND EPC change during 1 week after stroke            |                     |
| Sobrino et al., 2007 | ND       | ND          | reduced infarct growth, improvement of NIHSS    | EPC change during 1 week after stroke               |                     |
| Taguchi et al., 2004 | CD34+ cells | Serial changes in acute stroke (peak, D7: baseline, D30) | Many EPCs: CBF increase                           | Few EPCs: DM, being older                           |                     |

#### References

- Chu et al., 2008
- Lau et al., 2007
- Sobrino et al., 2007
- Taguchi et al., 2004

**ND:** not determined. **IMT:** intima-media thickness. **NIHSS:** National Institutes of Health Stroke Scale. **LAA:** large-artery atherosclerosis. **OC:** outgrowth cells. **CBF:** cerebral blood flow. **DM:** diabetes mellitus.
the numbers of circulating CD34- and CD133-positive cells and the presence of old infarction. Analysis of patients with cerebral artery occlusion revealed a significant positive correlation between circulating CD34- and CD133-positive cells and regional blood flow in areas of chronic hyperperfusion, suggesting that EPCs contribute to the homeostasis and repair of the cerebral circulation and maintenance of brain metabolism. The reduction in the number of EPCs appears to result from a decreased production and an enhanced degeneration of EPCs during the atherosclerotic process. Moreover, risk factors for atherosclerosis might directly influence the mobilization and survival of EPCs by impairing the bioavailability of nitric oxide. These findings support the notions that EPCs play an important part in the pathogenesis of atherosclerotic disease and that the measurement of EPCs could improve risk stratification.

**EPC Profiles in Acute Stroke**

EPCs are mobilized from BM during acute ischemia and contribute to the neovascularization of ischemic tissues. Acute myocardial infarction is associated with mobilization and a rapid increase in circulating EPCs. In the same line of acute cardiovascular events, vascular trauma such as coronary bypass grafting or burn injury induces a rapid but transient mobilization of VEGFR2+/AC133+ EPCs. Clinical trials assessing the therapeutic potential of BM-derived mononuclear cells (a rich source of immature cells including EPCs) in hind-limb and cardiac ischemia have yielded promising results. BM-derived immature cells have also been shown to participate in neovascularization of ischemic brain after experimentally induced stroke. Immature cells, including CD34- cells, have been shown to contribute to vasculature maintenance, not only as a pool of EPCs but also as the source of growth/angiogenesis factors. Stroke involves a complicated cascade of events involving cerebral ischemia: altered blood flow; disruption, inflammation, neuronal necrosis, and apoptosis of the blood-brain barrier; and neurological dysfunction. Which of these mechanisms are involved in the mobilization of EPCs is not clear, but vascular trauma and tissue ischemia appear to facilitate EPC mobilization to the peripheral pool, in part by the release of cytokines and vascular endothelial growth factor.

In initial observation studies, Taguchi et al. measured CD34+ cells by flow cytometry in 25 patients with an ischemic stroke. They found that the values peaked after 7 days and returned to baseline after 30 days. Ghani et al. reported that the number of clusters of rapidly adhering cells was decreased after stroke and in “stable CVD” compared to in controls free of vascular disease. They found that being older and the presence of CVD are generally independently related to a lower number of EPCs, with no significant increase in EPC numbers being observed in the weeks after acute or stable stroke. These conflicting results are probably due to a lack of corroborated methods that can precisely identify EPCs.

We recently analyzed the characteristics of EPCs in acute stroke patients, focusing on their differences according to the pathogenetic mechanism. We found that the number of CFU was lower in acute stroke patients than in control subjects, and much lower in patients with large-artery atherosclerosis than in those with cardioembolism. Our data are consistent with a previous report that EPC numbers differed significantly among acute stroke, stable stroke, and control subjects, and extend those observations by suggesting that CFU analysis can improve the understanding of stroke pathophysiology. We also demonstrated a relationship between the known surrogate markers for chronic vasculopathy, with the HbA1c level being an independent predictor for fewer CFU in a multivariate analysis. A previous study found that the number of EPCs was significantly related to the HbA1c and blood sugar levels in diabetics, and that improving glycemic control can significantly increase the number of EPCs. Our data showing that the degree of glycemic dysregulation affects EPC function are in line with this previous report. On the other hand, the outgrowing cells appeared in most of the stroke patients but only rarely in control subjects. It is possible that immediate endothelial or neural damage induced by stroke induces a compensatory BM overproduction of progenitor cells for damage repair. It should also be emphasized that indexes for neurological damage, such as the NIHSS (National Institutes of Health Stroke Scale) score and infarct volume, were associated with the number of outgrowth cells. Our results indicated that the numbers of CFU and outgrowing cells represent markers of the accumulated vascular risk and response to ongoing tissue damage, respectively.

**EPC as a Marker of Stroke Prognosis**

But do higher EPC numbers during the acute stage reverse the consequences of ischemia and improve prognosis? In observational studies involving patients with myocardial infarction, higher numbers of EPCs indeed relate to a better prognosis, more myocardial salvage, viability, and perfusion, and more collaterals in the ischemic zone. Human and animal model studies have shown that EPCs play a major role in angiogenesis and regeneration of ischemic brain tissue. However, the impact of the number of circulating EPCs on clinical outcomes after stroke remains uncertain. A recent clinical study found that the number of circulating EPCs was
significantly higher in patients with acute ischemic stroke than in at-risk control subjects, and that the magnitude of this difference is directly related to the functional outcome. One prospective study involving 48 stroke patients showed that the EPC increase during the first week was independently associated with a good outcome at 3 months. This favorable effect on the primary variable was supported by the reduction of infarct growth and neurological improvement at days 7 and 90. These findings are in line with experimental and human studies suggesting that EPCs mediate endothelial cell regeneration and neovascularization, and that EPCs participate in the cerebral neovascularization processes present in the adult brain after ischemia. Further prospective investigations of patients with CVD to establish a prognostic value of EPCs in ischemic stroke would be very useful.

EPC as a Potential Source for Cell Therapy

Mature nervous tissue has been considered incapable of cell renewal and structural remodeling for a long time, especially in mammals. However, stem cells are likely to replenish cells that are lost by physiological turnover, as well as in pathological conditions. It has recently been shown that appropriate \textit{in vitro} and \textit{in vivo} stimuli can induce adult somatic stem cells obtained from BM and PB to differentiate into neural-like cells. There have been intensive efforts over the past decade to develop therapeutic strategies for promoting revascularization of ischemic tissues. Since arteriogenesis and angiogenesis can be triggered by ischemia as native compensatory mechanisms, the discovery that EPCs are present in human PB raises the possibility of EPCs-mediated therapeutic neovascularization as a novel option for the treatment of ischemic diseases. The isolation and expansion of EPCs might be specifically useful for identifying therapeutic approaches targeting the progression and recurrence of stroke. Asahara et al. found transplanted EPCs in the endothelium of newly formed vessels in previously ischemic animal limbs. In other animal experiments, EPC administration also resulted in increased blood flow in ischemic zones and a decrease in limb loss. In experimentally induced cardiac ischemia, administration of progenitor cells resulted in neovascularization and a smaller infarcted area, although the involved mechanisms remained unclear.

Brain repair after stroke is a formidable task, because the lesions are often large and involve cells of all types, resulting in the loss of many complex synaptic connections. EPC transplantation has been used to minimize the effects of ischemic stroke in animal models. Taguchi et al. demonstrated that the systemic administration of human CD34-positive cells accelerated neovascularization in the cerebral ischemic zone 48 h after stroke in a mouse model. This treatment also increased neurogenesis and improved functional indexes in these animals. Similarly, human cord blood administered intravenously to rats after stroke was able to enter the brain and dispersed in the ischemic brain microenvironment, resulting in improved functional recovery compared to controls. BM-derived cells have also been shown to participate in neovascularization processes in the adult brain of mice following ischemia. Neurological functions after chronic cerebral ischemia were considerably better in rats intracerebrally transplanted with PB stem cells than in vehicle-treated control rats. PB stem cells transplanted into rats were observed to migrate toward infarcted cerebral zones and to differentiate into neurons, glial cells, and endothelial cells, thus enhancing neuroplasticity in the ischemic brain. The intravenous injection of PB stem cells into rats early after focal cerebral ischemia reduced lesion volumes and improved regional cerebral blood flow and cognitive functions. The use of cells derived from PB or BM, including EPCs, provides two main advantages over the use of other cell types: (1) it avoids ethical limitations because there is no need to work with fetal or embryonic tissue, and (2) there is a host of experience on the use of hematopoietic progenitor cells in hemato-oncology research, and therefore there is considerable knowledge of treatment tolerability and side effects. Furthermore, neovascularization and neurogenesis might be tightly linked in the brain. A rich vascular environment, along with the generation of other nurturing neuronal mediators by EPCs (e.g., VEGF, FGF2, and IGF-1) enhances subsequent neuronal progenitor migration to the damaged area, followed by their maturation and survival when EPCs have stimulated the formation of increased vascular channels.

We have previously described the \textit{in vitro} expansion and characterization of two types of progenitor cells from the PB of acute stroke patients. These cells are referred to as NOCs, in accordance with their morphological characteristics and protein or gene expression profiles. If neural progenitors could be isolated from human PB in sufficient quantities to permit harvesting, theoretically they would offer unlimited therapeutic possibilities. NOCs express immature neuronal markers over at least five passages, while maintaining rates of expansion characteristic of progenitor cells. Furthermore, the expressions of these markers are maintained in subclones obtained from these cells, and these subclones can be induced to undergo uniform differentiation into neurons in a chemically defined medium. Since PB-derived neural progenitor cells can be isolated without risk or potential side effects, their storage, expansion, and differentiation abilities make them valuable candidates for transplantation therapy.
Moreover, we found that intracerebral grafts of purified unmodified neural progenitor cells survived, migrated, and differentiated into neuronal phenotypes in ischemic rat brains. The exploitation of adult stem cells with neuronal characteristics—preferably obtained from easily accessible tissues—represents the best way of approaching cell therapy in stroke patients.

Future Challenges of EPC Research in CD

In future studies on the role of EPCs in stroke, several caveats should be borne in mind. First, we consider that the greatest current challenge in the field of EPC research is the lack of appropriate cell-surface markers specific for the identification of EPCs. Second, the laboratory technique of EPC quantification should be standardized as much as possible, and the timing of blood sampling (directly after the stroke or in the stable phase) should be considered, since the values obtained could differ between the acute and chronic phases. Third, the role of EPCs in the pathophysiology of CVD is no more than speculative at present. Therefore, it would be wise to further investigate the role of EPCs in different forms of stroke before planning clinical trials involving EPC injections. We also believe that the remarkable healing potential already demonstrated by EPC transplantation in experimentally induced stroke will become a clinical reality. However, we consider that clinical trials at this stage are premature and could be counterproductive. We suggest that this issue would be best progressed by standardizing certain aspects of basic research, especially the outcome measures, to facilitate comparisons between studies. In addition, long-term studies are required to determine whether cell-enhanced recovery is sustained, and to elucidate the tumorigenic potentials of cells. Furthermore, we must learn how to influence the pathological tissue environment and how this is related to repair. Other critical challenges include ensuring adequate characterization, manufacture, and quality control of cells, standardizing cell preparation protocols, and developing more definitive markers for stem cells. Identification of cell-surface markers for true stem cells would allow stem cell populations to be enriched or purified directly from heterogeneous cells.

EPCs or stem cells isolated from elderly stroke patients might retain dysfunctional characteristics, and therefore have reduced abilities to augment therapeutic regeneration. The expansion of cells in culture is an attractive strategy because it induces the potentiation of dysfunctional stem cells. Aside from these issues concerning ex vivo manipulation, considerable time is required to culture sufficient adult stem cells ex vivo to meet therapeutic requirements, and thus there is a need for an off-the-shelf supply of adult stem cells in the form of a cell bank that would allow physicians and surgeons to use stem cells directly at the point of care, rather than limiting their use to elective procedures.

Conclusion

EPCs hold great promise in CVD research, both as a marker for an increased risk of stroke and as a therapeutic agent after stroke. EPCs could change pathophysiological and therapeutic concepts, which will hopefully improve clinical treatments in vascular neurology. Recent work has provided a better understanding of these cells compared to what was known a decade ago. However, despite the promising results from various investigations (including our own), larger prospective studies should investigate this possible causation in order to confirm their role and applicability. Future studies should focus on establishing definitive markers of EPCs, better isolation methods, and the in-depth mechanisms underlying their beneficial effects.

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