Variability in the Chemical Composition of *Eugenia biflora* Essential Oils from the Brazilian Amazon

Pablo Luis B. Figueiredo¹², Henryck A. Fernandes³, Alberto Ray C. da Silva⁴, Nayara Sabrina F. Alves⁵, William N. Setzer⁶⁷, Joyce Kelly R. da Silva¹⁵, and José Guilherme S. Maia¹⁸

Abstract

*Eugenia* species are well known for their great economic potential as edible fruits. The leaves of 4 *Eugenia biflora* specimens (Ebi-1 to Ebi-4) were sampled in the Caratateua Island, Pará state, Brazilian Amazon. Then, the essential oils were hydrodistilled, analyzed by gas chromatography (GC) and GC mass spectroscopy, and their volatile compositions submitted to multivariate analysis (principal component analysis and hierarchical cluster analysis). Ebi-1 and Ebi-2 specimens were classified in the caryophyllene group, with significant content for the sesquiterpenes (E)-caryophyllene (16.8% and 11.4%) and caryophyllene oxide (28.6% and 20.5%). Ebi-3 specimen was grouped into the cadinane group, characterized by the presence of α-cadinol (14.7%), an oxygenated sesquiterpene. EBI-4 specimen was inserted into the aromadendrane group, with the predominance of the sesquiterpenes globulol (9.8%), germacrene B (7.9%), and γ-elemene (3.1%). Based on the results, a remarkable chemical variability was observed in the oils of *Eugenia biflora* with occurrence in Pará state, North Brazil. This work is presenting for the first time its caryophyllene, cadinane, and aromadendrane profiles.

Keywords

Myrtaceae, sesquiterpene compounds, essential oil, chemical composition, multivariate analysis

Received: August 27th, 2019; Accepted: October 23rd, 2019.

Myrtaceae consists of 144 genera and 6019 species distributed mainly in the Southern Hemisphere with particular emphasis in Australia and South America.¹² Among the genera that belong to this family, *Eugenia* has one of the most significant numbers of species, which corresponds to approximately 1000 species.³ Species of the genus *Eugenia* are well known for their great economic potential evidenced by the commercial exploitation of edible fruits, woods, and essential oils and by their use as ornamental plants. Besides this, many taxa are employed in the traditional medicine of tropical and subtropical regions around the world.⁴⁻⁷

Essential oils produced by some species of *Eugenia* from the Amazon have critical pharmacological applications because of their biological activities. For instance, *E. uniflora* has antioxidant, antimicrobial, and antitumor potential,⁸⁻⁹ and *E. egensis*, *E. flavescens*, *E. patricii*, and *E. polystachya* have cytotoxic effects.¹⁰

The species *Eugenia biflora* (L.) DC. (syn. *E. acuminatissima* Miq., *E. alfaroana* Standl., *E. amanuensis* Steyerm.), popularly known as “Vassourinha,” “Murta,” or “Pedra-hume-caa”,¹¹⁻¹² is distributed in North, Central, and South America. The plant is native and not endemic to Brazil, occurring in Northern (Acre, Amazonas, Amapá, Pará, Rondônia, Roraima, and Tocantins states), Northeastern (Maranhão, Piauí states), and Midwestern (Goiás, Mato Grosso do Sul, Mato Grosso states) regions.¹³

¹Programa de Pós-Graduação em Química, Universidade Federal do Pará, Belém, Brazil 
²Departamento de Ciências Naturais, Centro de Ciências Sociais e Educação, Universidade do Estado do Pará, Belém, Brazil 
³Programa Institucional de Bolsas de Iniciação Científica, Universidade Federal do Pará, Belém, Brazil 
⁴Faculdade Integrada Brasil Amazônia, Belém, Brazil 
⁵Programa de Pós-Graduação em Biologia, Universidade Federal do Pará, Belém, Brazil 
⁶Programa de Pós-Graduação em Química, Universidade Federal do Maranhão, São Luís, Brazil 
⁷Department of Chemistry, University of Alabama in Huntsville, USA 
⁸Aromatic Plant Research Center, Lehi, UT, USA 
⁹Programa de Pós-Graduação em Química, Universidade Federal do Maranhão, São Luís, Brazil 

Corresponding Author:
Pablo Luis B. Figueiredo, Programa de Pós-Graduação em Química, Universidade Federal do Pará, Belém, PA 66075-110, Brazil. Email: pablolbf@ufpa.br
E. biflora is a shrub that can grow to a height of 0.5-3.0 m. The species has elliptical, oval-lanceolate-shaped leaves with flat margins and grooved midribs on the adaxial surface, while its inflorescences are raceme type, solitary, or overlapping, axillary or terminal, with rachis of 4-27 mm. The flowers have a persistent nonshowy bract, and its fruits present a globular, ellipsoidal shape with a smooth surface.13 The plant is employed persistent nonshowy bract, and its fruits present a globular, ellipsoidal shape with a smooth surface.13 The plant is employed by traditional Amazonian communities to treat diarrhea, thrush, intestinal inflammation, and hemorrhages.12,14 The light petroleum extracts from leaves were rich in β-amyrin and eucalyptin,11 showing antimicrobial activity, which also emphasizes its pharmacological potential.12

There is only 1 study of leaf essential oils of E. biflora previously reported, which was collected in the municipality of Maracanã, State of Pará, showing a high content of α- and β-pinene and (E)-caryophyllene.15 Based on the species potential, the present study aimed to investigate the variability of the chemical composition of different specimens that occur in the Brazilian Amazon, to contribute to the knowledge of its chemotaxonomy.

E. biflora belongs to the group of plants known in the Amazon as “Pedra-hume-caá” (pedra-hume means alumina and caá means leaf in Tupi-Guarani language). The group comprises, in addition to E. biflora, the species Myrcia multiflora and Myrcia sylvatica. These 3 species have a broad pharmacological application and generate income in traditional communities because of their cultivation and commercialization.12

Four specimens of E. biflora were collected and showed distinct chemical composition for their essential oils (Table 1). These oils were light yellow with a woody odor and presented yields of 0.4% to Ebi-1, 0.3% to Ebi-2 and Ebi-4, and 0.2% to Ebi-3 samples (Table 1). The quantification and identification by gas chromatography (GC) and gas chromatography mass spectroscopy (GC-MS) presented 78 constituents in the oils of the specimens, representing an average of 92.1% of the total oils content (Table 2). In oils, oxygenated sesquiterpenes (48.0%-74.1%) and sesquiterpene hydrocarbons (19.0%-45.6%) predominated. Monoterpene was not identified in the oils. The compounds caryophyllene oxide (0.5%-28.6%), (E)-caryophyllene (1.8%-16.8%), α-cadinol (6.2%-14.7%), globulol (0.3%-9.8%), and epi-α-murolol (4.0%-8.4%) were the main constituents.

To classify the oils, a multivariate analysis (principal component analysis, PCA, and hierarchical cluster analysis) was applied using the chemical components (>3%) as variables (Figures 1 and 2). The PCA explained 92.7% of the total data variability. The component PC1 explained 56.0% and displayed positive correlations with the variables α-murolol, copaboreno, α-cadinol, epi-α-cadinol, spathulenol, viridiflorol, γ-elemene, germacrene B, and globulol. The component PC2 explained 36.7% and showed a positive correlation with the compounds caryophyllene oxide, 1-epi-cubenol, (E)-nerolidol, epi-α-murolol, α-murolol, copaboreno, α-cadinol, epi-α-cadino, and spathulenol.

The specimens Ebi-1 and Ebi-2 were grouped in the caryophyllene group due to its significant content of the sesquiterpenes, (E)-caryophyllene (16.8% and 11.4%), and caryophyllene oxide (28.6% and 20.5%). The specimen Ebi-3 was grouped into the cadinane group characterized by the oxygenated sesquiterpene α-cadinol (14.7%). The specimen Ebi-4 was grouped into the aromadendrene group with the predominance of the sesquiterpenes globulol (9.8%), germacrene B (7.9%), spathulenol (4.5%) and viridiflorol (3.5%) (Figure 2).

The sesquiterpenes (E)-caryophyllene and caryophyllene oxide are found in a significant number of plant species. These compounds have a significant anticancer activity, which affects the growth and proliferation of numerous tumor lineages, with low antioxidant activity.18 Two different chemical profiles of E. biflora, collected in Pará state, were described by Pereira and coworkers.15 The first one was characterized by the presence of the sesquiterpene hydrocarbon (E)-caryophyllene (15.36%), however, with the absence of caryophyllene oxide. The second profile was characterized by the presence of the monoterpenic hydrocarbons α- and β-pinene, with contents of 27.34% and 27.85%, respectively.

Species of Eugenia from the Amazon have shown significant infraspecific chemical variation. Leaf essential oils from E. patrida, collected from Northeastern Pará State, were rich in sesquiterpene hydrocarbons, such as germacrene D, β-elemene, δ-cadinene, or keto-phenolic ethers, as dimethylxanthoxylin.19 Similarly, oils of E. flavescens, from different regions of Pará State, were rich in sesquiterpenes with the germacrane skeleton (germacrene D and bicyclogermacrone), or the bisabolane skeleton (β-γ, (E)-iso-γ- and (E)-γ-bisabolene).10,15

Additionally, oils of E. patrida were characterized by (2E,6Z)-farnesol (23.2%) and (2E,6E)-farnesol (34.5%), or by a mixture of sesquiterpene hydrocarbons characterized by the presence of trans-cadina-1,4-diene (16.5%), trans-murola-3,5-diene (13.3%), (E)-caryophyllene (11.1%), and α-cubebene (9.8%), as its main compounds.10,15

### Table 1. Identification Data for the Specimens of Eugenia biflora.

| Samples | Herbarium number | Local coordinates |
|---------|------------------|-------------------|
| Ebi-1   | NR               | 1°14'55.97"S / 48°26'32.61"W |
| Ebi-2   | MG-229216        | 1°14'55.47"S / 48°26'32.31"W |
| Ebi-3   | MG-229219        | 1°14'26.79"S / 48°25'39.29"W |
| Ebi-4   | MG-229220        | 1°14'26.25"S / 48°25'40.16"W |

NR = not yet registered (in process). Botanical identification was made by comparison with authentic samples.

### Conclusion

At least 5 E. biflora profiles are occurring in the State of Pará, at Brazilian Amazon. In the oils of these profiles oxygenated sesquiterpenes and sesquiterpene hydrocarbons predominated.
Table 2. Essential Oil Yield and Composition of 4 Samples of *Eugenia biflora*.

| IR(\(_C\)) | IR(\(_L\)) | Constituents               | Ebi-1 | Ebi-2 | Ebi-3 | Ebi-4 |
|------------|------------|----------------------------|-------|-------|-------|-------|
| 858        | 857\(^b\)  | Ethyl-benzene              | 0.3   | 0.1   |       |       |
| 904        | 907\(^a\)  | Butyl propanoate           | 0.9   | 0.2   | 0.1   | 0.1   |
| 999        | 1000\(^a\) | \(n\)-Decane               | 0.2   |       |       |       |
| 1338       | 1335\(^a\) | \(\delta\)-Elemene         | 0.2   | 0.1   | 0.3   |       |
| 1352       | 1345\(^b\) | \(\alpha\)-Cubebene        | 0.1   | 0.1   | 0.1   |       |
| 1373       | 1373\(^a\) | \(\alpha\)-Ylangene        | 0.1   | 0.1   | 0.1   |       |
| 1374\(^a\) |            | \(\alpha\)-Copaene         | 1.7   | 3.1   | 1.0   | 0.9   |
| 1381       | 1378\(^a\) | (3Z)-Hexenyl hexenoate     |       |       |       | 0.1   |
| 1387       | 1387\(^a\) | \(\beta\)-Bourbonene       | 0.5   | 0.1   | 0.1   |       |
| 1391       | 1387\(^a\) | \(\beta\)-Cubebene         |       | 0.2   |       |       |
| 1394       | 1389\(^b\) | \(\beta\)-Elemene          | 0.7   | 1.8   | 0.9   | 2.6   |
| 1411       | 1409\(^a\) | \(\gamma\)-Gurjunene       |       |       |       | 0.1   |
| 1422       | 1417\(^a\) | \((\beta\)-Caryophyllene)  | 16.8  | 11.4  | 1.8   | 9.8   |
| 1431       | 1430\(^a\) | \(\beta\)-Copaene          | 0.4   | 0.4   | 0.7   | 0.5   |
| 1433       | 1432\(^b\) | \(\gamma\)-Elemene         | 0.5   | 0.3   | 3.1   |       |
| 1436       | 1432\(^b\) | \(trans\)-\(\alpha\)-Bergamotene | 0.2   |       |       |       |
| 1440       | 1437\(^a\) | \(\alpha\)-Guaiane         | 0.4   | 0.8   |       |       |
| 1442       | 1439\(^a\) | Aromadendrene              |       | 0.2   |       |       |
| 1445       | 1442\(^a\) | 6,9-Guaiadiene             |       |       | 0.4   |       |
| 1451       | 1447\(^b\) | Isogermacrene D            |       |       | 0.1   |       |
| 1454       | 1452\(^a\) | \(\alpha\)-Humulene        | 1.6   | 1.0   | 0.4   | 1.1   |
| 1462       | 1458\(^a\) | \(allo\)-Aromadendrene    |       |       |       | 0.2   |
| 1462       | 1464\(^a\) | 9-\(\varepsilon\)-(\(E\))-Caryophyllene | 0.2   |       | 0.5   |       |
| 1477       | 1478\(^a\) | \(\gamma\)-Murolene        | 0.7   | 0.8   | 1.6   | 1.1   |
| 1481       | 1483\(^b\) | \(\alpha\)-Amorphene       |       |       | 0.4   |       |
| 1484       | 1476\(^b\) | Selina-4,11-diene\(^c\)    |       |       |       | 0.1   |
| 1484       | 1484\(^a\) | Germacrene D               | 0.7   | 1.7   | 2.2   |       |
| 1487       | 1492\(^a\) | \(\beta\)-Selinene         | 0.4   | 0.6   | 1.0   |       |
| 1494       | 1493\(^b\) | \(trans\)-Murola-4(14),5-diene | 0.2   |       |       |       |
| 1494       | 1493\(^c\) | \(\varepsilon\)-Cabebol     | 1.8   |       |       |       |
| 1495       | 1496\(^c\) | Valencene                  |       | 1.4   |       |       |
| 1496       | 1496\(^a\) | Viridiflorene              |       |       | 2.1   |       |
| 1498       | 1500\(^a\) | Bicyclogermacrene          |       |       | 0.2   | 2.1   |
| 1501       | 1500\(^a\) | \(\alpha\)-Murolene        | 0.7   | 0.6   | 1.4   | 0.8   |
| 1506       | 1505\(^b\) | \(\alpha\)-Bulnesene       | 0.1   | 0.2   |       |       |
| 1509       | 1508\(^b\) | \(\beta\)-Bisabolene       | 0.1   | 0.2   |       |       |
| 1515       | 1513\(^b\) | \(\gamma\)-Cadinene        | 2.9   | 0.8   | 2.2   | 0.9   |
| 1519       | 1520\(^b\) | 7-\(\varepsilon\)-\(\alpha\)-Selinene | 0.4   | 0.7   |       |       |
| 1524       | 1522\(^a\) | \(\delta\)-Cadinene        |       |       |       | 2.1   |
| 1527       | 1521\(^a\) | \(trans\)-Calamene         | 2.1   | 1.5   | 1.5   |       |
| 1524       | 1522\(^a\) | \(\delta\)-Cadinene        |       |       |       | 2.1   |
| 1536       | 1540\(^b\) | Selina-4(15),7(11)-diene   |       |       | 0.5   |       |
| 1538       | 1532\(^a\) | \(\gamma\)-Cuprenene       | 1.5   |       |       |       |
| 1540       | 1537\(^a\) | \(\alpha\)-Cadinene        | 0.4   | 0.4   |       |       |
| 1542       | 1545\(^b\) | Selina-3,7(11)-diene       |       | 0.5   |       |       |
| 1549       | 1548\(^a\) | Elemol                     | 0.3   | 1.8   | 0.7   |       |
| 1558       | 1559\(^a\) | Isoelemol                  | 1.1   | 0.7   | 7.9   |       |
| 1564       | 1561\(^c\) | (\(\varepsilon\))-Nerolidol | 1.4   | 3.6   | 1.6   | 0.5   |
| 1569       | 1567\(^a\) | Palustrol                  |       |       | 1.4   |       |
| 1579       | 1577\(^a\) | Spathulenol                | 2.2   | 2.4   | 4.9   | 4.5   |
| 1583       | 1584\(^a\) | \textbf{Caryophyllene oxide} | 28.6  | 20.5  | 7.5   | 0.5   |
| 1588       | 1590\(^a\) | Germol                     | 0.3   |       |       | 9.8   |

(Continued)
This work reports the caryophyllene, cadinane, and aromadendrane profiles for the first time.

Materials and Methods

Plant Material

Leaves of 4 specimens of *E. biflora* were collected in Caratateua Island, State of Pará, Brazil. Plant vouchers were identified by Professor Marcos Sobral, a Brazilian Myrtaceae specialist, and deposited in the Herbarium of the Museu Emilio Goeldi, city of Belém, State of Pará, Brazil. The coordinates for each collected specimen and the voucher number are listed in Table 1. About 150 g of leaves from each plant were collected and dried for 2 days at room temperature. Dried leaves (60 g) were then subjected to hydrodistillation.

Table 2. Continued

| IR(C) | IR(L) | Constituents          | Ebi-1 | Ebi-2 | Ebi-3 | Ebi-4 |
|-------|-------|-----------------------|-------|-------|-------|-------|
| 1592  | 1592  | Viridiflorol          | 2.0   | 1.6   | 2.9   | 3.5   |
| 1594  | 1595  | Cubeban-11-ol         | 0.4   | 1.8   | 1.4   | 1.5   |
| 1602  | 1600  | Rostifolios           | 1.0   |       |       |       |
| 1605  | 1602  | Ledol                 | 1.8   | 2.2   | 1.3   |
| 1609  | 1608  | Humulene epoxide II   | 1.1   | 1.1   | 0.4   | 0.4   |
| 1615  | 1618  | 1,10-di-epi-Cubenol   | 2.0   | 0.9   | 0.3   |
| 1620  | 1618  | Junenol               | 0.4   | 1.8   | 1.4   |
| 1624  | 1624  | *epi*-Eudesmol        | 1.8   | 2.2   | 1.3   |
| 1627  | 1627  | *1-epi*-Cubenol       | 6.4   | 6.6   | 3.3   | 1.6   |
| 1633  | 1630  | *γ*-Eudesmol          | 1.8   | 0.8   |       |
| 1637  | 1639  | Caryophylla-4(12),8(13)-dien-5β-ol | 2.6   |       |
| 1641  | 1638  | *epi*-α-Cadinol (τ-cadinol) | 4.9   | 1.9   |
| 1642  | 1640  | *epi*-α-Murrolol (τ-murrolol) | 8.4   | 7.6   | 8.3   | 4.0   |
| 1647  | 1644  | α-Murrolol (Torreyol)  | 2.4   | 3.0   | 4.8   | 2.1   |
| 1651  | 1649  | β-Eudesmol            | 1.6   | 0.7   |       |
| 1653  | 1652  | Himachalol            | 0.9   |       |       |
| 1655  | 1652  | α-Cadinol             | 6.2   | 6.3   | 14.7  | 7.5   |
| 1657  | 1661  | Allohimachalol        | 0.9   |       |       |
| 1659  | 1660  | *cis*-Calamenen-10-ol  | 0.5   |       |       |
| 1666  | 1665  | Intermedeol           | 0.5   |       |
| 1669  | 1668  | trans-Calamenen-10-ol  | 0.3   | 0.2   | 0.2   |
| 1671  | 1668  | 14-Hydroxy-9-*epi*-(E)-caryophyllene | 1.1   | 0.8   | 0.5   |
| 1686  | 1685  | Germacre-4(15),5,10(14)-trien-1-α-ol | 1.4   | 1.0   | 0.3   |
| 1696  | 1696  | Juniper camphor       | 0.4   | 0.5   | 1.4   |
| 1701  | 1702  | 10-*nor*-Calamenen-10-one | 0.1   |       |       |

Sesquiterpene hydrocarbons 28.2 27.7 19.0 45.6
Oxygenated sesquiterpenes 62.2 61.7 74.1 48.0
Others 1.4 0.3 0.1 0.2
Total (%) 91.8 89.7 93.2 93.8
Oil yield (%) 0.4 0.3 0.2 0.3

RI(C) : calculated retention time; RI(L) : literature retention time; *16,17*; Bold: main constituents (above 5%).
*a*Adams (2007).*17
*b*Mondello (2011).*16
**Correct stereoisomer was not determined.

Extraction and Oil Composition Analysis

Essential oils were hydrodistilled using a Clevenger-type apparatus (3 hours), according to Gottlieb and Magalhães.*20 Then, the oils were dried over anhydrous sodium sulfate, and the total oil yields are expressed as mL/100 g of dried material. The oil composition analysis was performed by GC-MS, using a Shimadzu instrument Model QP 2010 ultra (Shimadzu, Tokyo, Japan) equipped with an Rtx-5MS (30 m × 0.25 mm; 0.25 µm film thickness) silica capillary column (Restek, Bellefonte, PA, USA). Helium was used as the carrier gas, with a linear velocity of 1.0 mL/min at 57.5 kPa; split injection (split ratio 1:20) of 1 µL of a hexane solution (5 µL: 500 µL hexane); injector and interface temperatures were 250°C. The programmed temperature was 60 to 240°C (3°C/min), followed by an isotherm of 10 minutes. Electron impact mass spectrometry: electron energy was 70 eV,
and the ion source temperature was 200°C. The mass spectra were obtained by automatic scanning at every 0.3 second, with mass fragments in the range of 35-400 m/z. The oil constituents were identified by comparison of their mass spectra and retention indices with those available in commercial libraries, such as Adams and FFNSC 2/Mondello. Retention indices were calculated using n-alkane standard solutions (C8-C40, Sigma-Aldrich, St. Louis, MO, USA) in the same chromatographic conditions. GC flame ionization detector (FID) analysis was carried out on a Shimadzu QP-2010 instrument equipped with an FID detector in the same conditions, except that hydrogen was used as the carrier gas. The oil constituents percentual was computed from GC-FID peak areas.

**Statistical Analysis**

The multivariate analysis was performed by using as variables the constituents of oils with content above 3.0%. The data
matrix was standardized by subtracting the mean and then dividing it by the SD. For hierarchical cluster analysis, the complete linkage method and the Euclidean distance were used. All analyses were performed using the software Minitab (free 390 version, Minitab Inc., State College, PA, USA).

Acknowledgments

We are grateful to CNPq and Capes, Brazilian government funding agencies, for scientific research support.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

ORCID ID

Pablo Luis B. Figueiredo  https://orcid.org/0000-0003-1365-3513
William N. Setzer  https://orcid.org/0000-0002-3639-0528

References

1. Wilson PG. Myrtaceae. In: Kubitzki K, ed. The Families and Genera of Vascular Plants: Flowering Plants Eudicots Sapindales, Cucurbitales, Myrtaceae. 1st ed. New Delhi: Springer-Verlag Berlin Heidelberg; 2011:436.
2. WSCP. World Checklist of Selected Plant Families. Facilitated by the Royal Botanic Gardens. Kew; 2012.
3. Van der Merwe MM, Van Wyk AE, Botha AM. Molecular phylogenetic analysis of Eugenia L. (Myrtaceae), with emphasis on southern African taxa. Plant Syst Evol. 2005;251(1):21-34.
4. Queiroz JMG, Suzuki MCM, Motra APR, Nogueira JMR, de CEM. Aspectos populares e científicos do uso de espécies de Eugenia como fitoterápico. Rev Fitoter. 2015;9(2):87-100.
5. Donadio LC, Moro FV. Potential of Brazilian Eugenia myrtaceae-as ornamental and as a fruit crop. In: Acta Horticulturae. Leuven, Belgium: International Society for Horticultural Science (ISHS); 2004:65-68.
6. Mitra SK, Irenaeus TKS, Gurung MR, Pathak PK. Taxonomy and importance of myrtaceae. In: Acta Horticulturae. Leuven, Belgium: International Society for Horticultural Science (ISHS); 2012:23-34.
7. dos Santos JFS, Rocha JE, Bezerra CF, et al. Chemical composition, antifungal activity and potential anti-virulence evaluation of the Eugenia uniflora essential oil against Candida spp. Food Chem. 2018;261:233-239.
8. Victoria FN, Lenardão EF, Savegnago L, et al. Essential oil of the leaves of Eugenia uniflora L.: antioxidant and antimicrobial properties. Food Chem Toxicol. 2012;50(8):2668-2674.
9. Figueiredo PLB, Pinto LC, da Costa JS, et al. Composition, antioxidant capacity and cytotoxic activity of Eugenia uniflora L. chemotype-oils from the Amazon. J Ethnopharmacol. 2019;232:30-38.
10. da Silva J, Andrade E, Barreto L, et al. Chemical composition of four essential oils of Eugenia from the brazilian amazon and their cytotoxic and antioxidant activity. Medicines. 2017;4(3):51.
11. Gottlieb OR, da Silva ML, Maia JGS. Eucalyptin from Eugenia and Myrcia species. Phytochemistry. 1972;11(3):1185.
12. da Silva FKS, dos Rosário AS, Secco RS, Zoghbi MGB. A survey of the species known as pedra-ume-caá (Myrtaceae) with emphasis with emphasis on those species recognized and commercialized in the city of Belém, Pará, Brazil. Biot. Amaz. 2015;5(1):7-15.
13. Sobral M, Proença C, Souza M, Mazine F, Lucas E. Myrtaceae in lista de espécies da flora do Brasil. Jard Bot Rio de Janeiro. 2012;50(8):2668-2674.
14. Tropicos. org. Eugenia biflora (L.) DC. Missouri Botanical. https://www.tropicos.org/Name/22101722. Accessed October 2019.
15. Pereira RA, Zoghbi MdasGB, Bastos MdelNdOC. Essential oils of twelve species of Myrtaceae growing wild in the sandbank of the Resex Maracanã, state of Pará, Brazil. Journal of Essential Oil Bearing Plants. 2010;13(4):440-450.
16. Mondello L. FFNSC 2: Flavors and Fragrances of Natural and Synthetic Compounds, Mass Spectral Database. New york: John Wiley & Sons Inc; 2011.
17. Adams RP. Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry. Carol Stream, Illinois, USA: Allured Publishing Corporation; 2007.
18. Fidży K, Fiedorowicz A, Strządała L, Szumny A. β-caryophyllene and β-caryophyllene oxide-natural compounds of anticancer and analgesic properties. Cancer Med. 2016;5(10):3007-3017.
19. Zoghbi MGB, Guilhon GSP, Sarges FN, Pereira RA, Oliveira J. Chemical variability of the volatiles from the leaves of Eugenia protenta McVaugh (Myrtaceae) growing wild in the North of Brazil. Biochem Syst Ecol. 2011;39(4-6):660-665.
20. Gottlieb OR, Magalhães MT. Modified distillation trap. Chemist Analyst. 1960;49(4):114.
21. van Den Dool H, Dec. Kratz P. A generalization of the retention index system including linear temperature programmed gas—liquid partition chromatography. J Chromatogr A. 1963;11:463-471.