Neonatal oral myiasis caused by the larvae of *Sarcophaga ruficornis* (Diptera: Sarcophagidae): a case report

Minyu Zhou1†, Ke Cao4†, Hui Huang1, Xiaojuan Luo4, Ying Wang4, Weike Ma4 and Zhiyue Lv1,2,3*

Abstract

**Background:** Myiasis is caused by dipterous larvae, and rarely affects the mouth. Diagnosis by traditional means is easy to be confused with other similar species. Here, we report a case of oral myiasis, in a 5-month-old infant who was diagnosed by morphological examination and molecular biological methods.

**Case presentation:** A 5-month old infant with acute myeloid leukemia was admitted due to recurrent skin masses for more than 4 months. The infant had lip swelling, which prevented him from closing the mouth and membranes were present in his mouth and there were also oral ulcers and erosions. Ten maggots were found in the mouth and one in the ear canal with pus flowing out and were confirmed as the third stage larvae of *Sarcophaga ruficornis* by morphological examination and a comparison of sequence of cytochrome oxidase subunit 1 (*COX1*) gene. After removal of the maggots and chemotherapy, the infant’s condition was gradually improved.

**Conclusions:** To the best of our knowledge, this is the first neonatal oral myiasis case reported in China and its diagnosis requires a high index of suspicion. Microscopy combined with specific DNA sequence analysis is an effective technological tool to provide rapid diagnoses of the larva specimen and cases of rare diseases, as illustrated in the current case.

**Keywords:** Neonatal oral myiasis, Nosocomial, Molecular identification

Background

Myiasis is an infestation in living mammals caused by dipterous larvae [1, 2]. The female flies land on the hosts and deposit eggs and maggots, and the larvae then penetrate the tissues [3]. In humans, myiasis rarely occurs in the mouth [4, 5]. In recent years, many cases of neonatal myiasis have been reported globally, especially in developing countries, with a myriad of symptoms, such as dyspnea, hyperemic inflammation of the tympanic membrane, and even cardiac arrest in severe cases [7–9].

Here, we report a case of oral myiasis, in a 5-month-old infant who was diagnosed by morphological examination, biology techniques and bioinformatics.

Case presentation

A 5-month-old male infant was referred to Shenzhen Children’s Hospital on July 15, 2019 with recurrent skin swellings for more than 4 months and juvenile cells in the peripheral blood of the patient were detected 24 h ago. His birth history, feeding history, growth history, vaccination history and family history are normal.

The patient was diagnosed as acute myeloid leukemia (AML, M5) based on the clinical examination of blood routine and bone marrow cytology on July 18, 2019. Physician examination at admission showed numerous hard magenta nodules with an indistinct boundary and poor mobility.
in the head and neck, torso, and limbs, with a maximum diameter of about 6 cm. Skin mass pathology revealed juvenile yellow granulomas. His lips were swollen and not ruddy, especially the upper lip. The lips could not be closed, and he breathed through his mouth. There were numerous white membranes in his mouth. Three days after admission, oral mucosa erosion and oral ulcers appeared, and then 10 maggots were found in the mouth, each about 3 to 5 mm in length, as shown in an additional video file [see Additional file 1]. Meanwhile, a wriggling larva was found in his right ear canal with an outflow of pus. All of them were removed by using sterile tweezers. One of the larvae collected from the patient’s mouth was sent to the Diagnostic Section in the Department of Parasitology of Zhongshan School of Medicine, Sun Yat-sen University.

The larva (1 mg) was white and about 5 mm in length. The larva collected showed the morphological characteristics of the third instar larva of Brachycera (Fig. 1). Next, we extracted total DNA of the larva using a Hipure Insect DNA Kit (Magen, China) and amplified the cytochrome oxidase subunit 1 (COX1) gene by Polymerase Chain Reaction (PCR) using dipteran-specific primer pairs: LCO1490, 5’TAACTTCAGGGTGACCAA AATCA-3’ and HCO2198, 5’GGTCAAACATCAATA AAGATATGGA-3’ [10] (Fig. 2) followed by sequencing (Tsingke Co.). BLASTn search for homology in the National Center for Biotechnology Information (NCBI) database (http://blast.ncbi.nlm.nih.gov/Blast.cgi) showed that the sequence had 100% homology with the COX1 gene of Sarcophaga ruficornis. We constructed phylogenetic trees by MEGA 6.0 software, based on 1000 bootstrap replicates from sequences we obtained. Sequences of the homologous genes, with accession numbers of MH937749.1, JQ582053.1, JQ582085.1, KT272859.1, KF819910.1, JQ806820.1, JQ582089.1, KX094928.1, KM279656.1, JQ582078.1 and KF038009.1, of Genus Sarcophaga and Genus Neobellieria were downloaded from NCBI. The results of the similarity analyses on the basis of the above sequences revealed that the maggots were accurately identified as S. ruficornis with 100% sequence matching identities (Fig. 3).

After removal of the maggots in the mouth and strengthening oral care, the oral health of the patients improved and no maggot infection recurred. Moreover, the patient received subsequent chemotherapy with the DAE regimen (cytarabine, daunorubicin, and VP-16), anti-infection and ventilator-assisted respiratory therapy, continuous renal replacement therapy (CRRT) support therapy, and correction of coagulation function. The patient gradually improved following 2 months of symptomatic treatment, and was discharged on September 24, 2019.
Discussion and conclusion
This is the first neonatal oral myiasis case reported in China. Oral myiasis is rare because the mouth cannot provide a conducive habitat and oral tissues are not directly exposed to the external environment [11, 12].

In this case, the infant could not close his mouth to breathe because of lip swelling and oral ulcers developed three days after admission. His immunity had been compromised by his AML and related treatment. Nocturnal mouth breathing and oral carcinoma are predisposing factors of oral myiasis [13], and the larvae may invade the necrotic tissues through wound. The infant was too young to articulate foreign body sensation or pain in the mouth, let alone the history of being disturbed by flies. Additionally, the summer is the peak season for flies owing to the warm and humid environment [14] and infants are more susceptible to being infected because they have no ability to defend against the flies. Thus, we presume that this is nosocomial according to the patient’s symptoms. Hospitals should pay more attention to sanitary conditions and take certain measures to control mosquitoes and flies, such as using screens and mosquito nets [15]. Emphasizing the oral hygiene of patients, especially infants, is also essential for prevention of myiasis.

We confirmed a myiasis caused by *S. ruficornis* in this case. As a result, clinicians could not have a deep understanding of this disease. Due to lack of personnel in parasite identification, relatively subjective morphological results, and clinicians’ lack of experience in myiasis, simple morphological examination may easily lead to missed or inaccurate diagnosis. Fortunately, more and more methods are being developed to diagnose myiasis. Recently, molecular biology techniques have been popular in diagnosing myiasis, and sequences analysis and phylogenetic analysis have been particularly widely applied [16–20]. Dermoscopy [21], ultrasound scanning [22], mineral oil and magnification [2], serology [23] and many other techniques are applied to diagnose myiasis. Because molecular biological methods can avoid misjudgment and lead to specific detection compared to the traditional methods, they increase accuracy and efficiency of species determination and diagnosis. Moreover, they can help to understand the development of species and explore the relationships between species in order to obtain better prevention and treatment. In summary, we combined morphological observation and molecular investigation to identify *S. ruficornis* as the cause of oral myiasis.

Abbreviations
AML: Acute myeloid leukemia; CRRT: Continuous renal replacement therapy; PCR: Polymerase chain reaction; COX1: Cytochrome oxidase subunit 1.

Supplementary Information
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Authors’ contributions
ZL conceived and designed the experiments and draft the manuscript; MZ and HH participated in drafting the manuscript; KC collected the samples;
MZ performed the experiments and analyzed the data; XL, YW and WM participated in technological guidance and coordination. All authors read and approved the final version of the manuscript.

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Availability of data and materials
All data generated or analysed during this study are included in this published paper.

Declarations

Ethics approval and consent to participate
All the clinical specimens were obtained with written consent and approved by the parents of the patient involved in this study.

Consent for publication
The parents of the patient have signed the consent form and agreed to publish this study and a copy of written consent is available for the journal.

Competing interests
The authors declare that they have no competing interests.

Author details
1 Key Laboratory of Tropical Disease Control (Sun Yat-Sen University), Ministry of Education, Guangzhou, China. 2 Key Laboratory of Tropical Translational Medicine of Ministry of Education, Hainan Medical University, Haikou, China. 3 NHC Key Laboratory of Control of Tropical Diseases, Hainan Medical University, Haikou, China. 4 Shenzhen Children’s Hospital, Shenzhen, China.

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References

1. Hope FW. On insects and their larvae occasionally found in the human body. Trans R Entomol Soc Lond. 1840;2:256–71.
2. Noutsis C, Millikan LE. Myiasis. Dermatol Clin. 1994;12(4):729–36.
3. Vazir S, DiB B, Yalcin S, Demirtas F, Yaman O, Ozturk M, Sahin I. Nosocomial oral myiasis by Sarcophaga sp. in turkey. Yonsei Med J. 2005;46(3):431–4.
4. Reddy MH, Das N, Vivekananda MR. Oral myiasis in children. Contemp Clin Dent. 2012;3(Suppl 1):19–22.
5. Sharma J, Mamatha GP, Acharya R. Primary oral myiasis: a case report. Med Oral Patol Oral Cir Bucal. 2008;13(11):E714–6.
6. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Mol Mar Biol Biotechnol. 1994;3(5):294–9.
7. Martinez-Rojano H, Noguez JC, Huerta H. Nosocomial myiasis caused by Lucilia sericata (Diptera: Calliphoridae) and neonatal myiasis by Sarcophaga spp. (Diptera: Sarcophagidae) in Mexico. Case Rep Infect Dis. 2018. https://doi.org/10.1155/2018/5067569.
8. Ata N, Guzelkara F. Aural myiasis in an infant. J Craniofac Surg. 2017;28(1):e89–90.
9. Singh AK, Nag SS, Mitra P, Roy A. Neonatal umbilical myiasis. Indian Dermatol Online J. 2015;6(4):312–3.
10. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. Mol Mar Biol Biotechnol. 1994;3(5):294–9.
11. Vinit GB, Jayavelu P, Shrutha SP. Oral myiasis in a maxillofacial trauma patient. J Pharm Bioallied Sci. 2013;5(Suppl 2):195–7.
12. Sharma A. Oral myiasis is a potential risk in patients with special health care needs. J Glob Infect Dis. 2012;4(1):60–1.
13. Reddy MH, Das N, Vivekananda MR. Oral myiasis in children. Contemp Clin Dent. 2012;3(Suppl 1):19–22.
14. Dutto M, Bertero M. Cutaneous superficial myiasis: report of a rare nosophomarial parasitic disease caused by Sarcophaga spp. (Diptera, sarcophagidae). Gent Eur J Pub Health. 2011;19(4):232–4.
15. Francesconi F, Lupi O. Myiasis. Clin Microbiol Rev. 2012;25(1):79–105.
16. Boehme P, Amendt J, Disney RH, Zehner R. Molecular identification of carrion-breeding scuttle flies (Diptera: Phoridae) using COI barcodes. Int J Legal Med. 2010;124(6):577–81.
17. Hall MJ, Adams ZJ, Napy WT, Testa JM, Edge W, Nikoluas M, Farkas R, Ready PD. Morphological and mitochondrial DNA characters for identification and phylogenetic analysis of the myiasis-causing flesh fly Wolhafertia magnifico and its relatives, with a description of Wolhafertia monegrosensis sp. n. Wyatt & Hall. Med Vet Entomol. 2009;23(Suppl 1):59–71.
18. Sun X, Wang LF, Feng Y, Xie H, Zheng XY, He A, Karim MR, Lv ZY, Wu ZD. A case report: a rare case of infant gastrointestinal canthariasis caused by larvae of Lasioderma serricorne (Fabricius, 1792) (Coleoptera: Anobiidae). Infect Dis Poverty. 2016;5:34–9.
19. Zhang B, Wang L, Liu J, Xu L, Song L, Wu X, Sun X, Wu Z. Case report: a rare case of urinary myiasis induced by the fourth instar larvae of Telmatoscopus altipunctatus. PLoS Negl Trop Dis. 2017;11(12):e0006016(7).
20. Solgö R, Djaddid ND, Eslamifar A, Raz A, Zakeri S. Morphological and molecular characteristic of Megaselia scalaris (Diptera: Phoridae) larvae as the cause of urinary myiasis. J Med Entomol. 2017;54(3):781–4.
21. Giangaspero A, Marangi M, Balotta A, Venturrelli C, Szpila K, Di Palma A. Wound myiasis caused by Sarcophaga (Loppyga) Angyrostoma (robineau-desvoidyi) (Diptera: Sarcophagidae): additional evidences of the morphological identification dilemma and molecular investigation. Scientific World J. 2017;2017:90645311(1):906453(9).
22. Gontijo JRV, Bittencourt FV. Wound myiasis: the role of entodermoscopy. An Bras Dermatol. 2018;93(5):745–6.
23. Szczerko C, Dompmartin A, Moreau A, Bellory B, Remond B, Leroy D. Ultra-sonography of furuncular cutaneous myiasis: detection of Dermatobia hominis larvae and treatment. Int J Dermatol. 1994;33(4):282–3.
24. Puente S, Otranto D, Panadero R, Herrera MD, Rivas P, Ramirez-Olivia G, Mariscal C Jr, Perteuguer MJ, Diez-Banos P, Garate T. First diagnosis of an imported human myiasis caused by Hypoderma simense (Diptera: Oestridae), detected in a European traveler returning from India. J Travel Med. 2010;17(6):419–23.

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