The betaine content of sweat from adolescent females

Shona S Craig¹, Stuart AS Craig²*, Matthew S Ganio³, Carl M Maresh³, Horrace Greg¹, Kerry-Ann da Costa⁴, Steven H Zeisel⁴

Abstract

Background: This study was developed to establish whether betaine was present in the sweat of females and to determine any correlations with other sweat components.

Methods: Sweat patches were placed on eight trained adolescent Highland dancers (age = 13.6 ± 2.3 yr), who then participated in a dance class for 2 hours. Patches were removed, and the sweat recovered via centrifugation. The sweat was subsequently analyzed for betaine, choline, sodium, potassium, chloride, lactate, glucose, urea and ammonia.

Results: Betaine was present in the sweat of all subjects (232 ± 84 μmol·L⁻¹), which is higher than typically found in plasma. The concentration of several sweat components were correlated, in particular betaine with most other measured components.

Conclusion: Betaine, an osmoprotectant and methyl donor, is a component of sweat that may be lost from the body in significant amounts.

Background

Betaine is a methylamine that is widely distributed in nature where it is found in microorganisms, plants and animals. It is a significant component of many foods, including whole grains (e.g. wheat, rye), spinach, shellfish and beets [1], and low levels of dietary intake may increase disease risk [2-5]. Betaine is a trimethyl derivative of glycine that functions as an organic osmolyte to protect cells under stress (e.g. dehydration, high concentrations of electrolytes, urea and ammonia) and as a source of methyl groups for use in many key pathways via the methionine cycle [2]. Betaine accumulates in most tissues (e.g. liver, kidney, intestine, skin, muscle, etc.) [6], is non-perturbing to cellular metabolism, highly compatible with enzyme function, and stabilizes cellular metabolic function [2,7-14]. Betaine plays an important role in several aspects of human health and nutrition and recent studies show that ingestion of betaine may improve athletic performance [15-17].

Betaine concentration has not been previously studied in sweat. Sweat can be considered a filtrate of plasma, cellular and interstitial fluid that contains electrolytes (e.g. potassium, sodium, and chloride), metabolic wastes (e.g. urea, ammonia and lactic acid), and various nutrients (e.g. vitamins and choline) [18-21]. The exact composition of sweat is dependent on several factors, including absorptive mechanisms in the sweat glands that may increase or decrease the concentration of solutes. We hypothesized that since betaine is a component of plasma and skin, it is also likely to be present in sweat. In addition, the above-mentioned protective role of betaine against electrolytes and metabolic wastes may extend to the sweat gland, duct and surrounding tissue. This study was conducted to determine whether betaine is a component of sweat that may be lost from the body during exercise.

Methods

Subjects

Eight trained female Scottish Highland dancers (10-17 yr) were recruited from the Stirling Highland Dance Company, Oakdale CT. The subjects trained regularly, and were actively competing in dance competitions. Subjects attended a briefing meeting before any
experimentation to ensure an understanding of the testing parameters and the benefits/risks of the study. The subjects and parents signed a written informed consent statement. The study was part of the Somers High School (SHS) Science Research Program and the protocol was approved by the SHS IRB.

Experimental Protocol
Sweat patches were prepared by placing two 2" × 2" gauze squares onto 4" × 4.5" adhesive film. Care was taken to minimize any cross-contamination. New disposable latex gloves were utilized for each subject. The skin on the lower back of the subjects was cleaned with gauze and distilled water, dried, and two patches were placed on both sides of the spine. The dancers then conducted a 2 hour class. The sweat patches were removed, placed in plastic 6-ml centrifuge tubes and stored on ice prior to centrifugation. The tubes were spun for 2 min at 1315 g in a benchtop centrifuge (Model 0151; Clay Adams, Parsippany, NJ). The patches were removed from the tubes, and the sweat (1-2 ml) at the bottom of the tubes was recovered. Each subject had two tubes from the two patches. The sweat from the two tubes was combined and stored frozen at -20°C prior to analysis.

Measurements
Betaine, choline, and choline metabolites were determined in duplicate by liquid chromatography/electrospray ionization-isotope dilution mass spectrometry [22]. Lactate and glucose were determined in duplicate by enzymatic techniques (YSI 2300 Stat Plus, Yellow Springs, OH). Sodium, potassium and chloride were measured in duplicate using ion selective electrodes (Medica Easy Electrolytes, Medica Corp., Bedford, MA). Urea and ammonia were measured using a COBAS Mira Plus Analyzer (Roche Diagnostics, Indianapolis, IN) and Pointe Scientific (Canton, MI) reagent sets and standards. Instruments were calibrated using NIST certified standards.

Statistics
Grubbs’ test http://graphpad.com/quickcalcs/Grubbs1.cfm was used to determine outliers in data sets (alpha = 0.05). Pearson’s correlation test (SigmaPlot v11, Systat Software Inc, San Jose, CA) and Passing-Bablok regression analysis (MedCalc, Mariakerke, Belgium) were conducted to compare data sets.

Results
The measures of sweat composition are shown in Table 1. Phosphatidylcholine and sphingomyelin were also measured, but were not detected (data not shown). The mean betaine content was 232 ± 84 μmol·L⁻¹. The other components of sweat were found at levels similar to that of previous studies [18,19,21]. Four data points were identified (*) as outliers via Grubb’s test. Pearson’s correlations test and Passing-Bablok regression analysis were conducted for the data in Table 1. Table 2 shows the Pearson’s correlation coefficients between sweat components - values that show statistical significance (p < 0.05) or trends (p < 0.10) are in bold. Betaine is correlated with all components except sodium and chloride (Fig. 1). The non-parametric regression analysis (Passing-Bablok) gave similar results (not shown). None of the Pearson’s correlations for potassium remain after removal of a data point (19.3 mmol·L⁻¹) that is an outlier via Grubb’s test (Table 1). Table 3 compares the content of sweat measured in this study with typical fasting levels published for plasma [18,23-26].

We observed that betaine levels can drop if kept at room temperature for prolonged periods; therefore, it is important when collecting sweat samples to keep them in crushed ice until frozen. We speculate that enzyme or bacterial action might reduce betaine levels, but this requires further study. Also, preliminary results (not shown) suggest that betaine levels in sweat are higher after ingestion of betaine. Future work on the relationship between plasma and sweat levels is warranted.

Discussion
This is the first study to determine the betaine content of sweat, and the average concentration in adolescent female sweat (232 ± 84 μmol·L⁻¹) was found to be about 7 times higher than that typically found in female plasma (34 ± 11 μmol·L⁻¹) [25]. The majority of constituents in sweat, such as sodium, chloride, glucose and choline, are more dilute than in the blood plasma or interstitial fluid [20]. However, some constituents are more concentrated in sweat, such as lactate, urea, ammonia, and potassium to a small extent. There are studies that support the concept of higher betaine concentrations in sweat versus plasma. Firstly, betaine is actively accumulated as an osmolyte in skin cells under osmotic and oxidative stress [12,27]. Also, there are higher betaine concentrations (expressed as μmol·L⁻¹ tissue water) in rat skin (males 412 ± 185 μmol·L⁻¹; females 305 ± 153 μmol·L⁻¹) compared to rat plasma (males 186 ± 43 μmol·L⁻¹; females 101 ± 37 μmol·L⁻¹) [6].

Mean dietary intake of betaine was recently estimated to be 100-200 mg/d [28,29]. Loss via urine averages about 10 mg/d [30]. Sweat rates are variable, but daily fluid requirements for sedentary to very active persons range from 2-4 L/d in temperate climates and from 4-10 L/d in hot climates [31]. Therefore, a range of 2-10 L/d sweat loss translates to a betaine loss of approximately 50-270 mg/d from the regional sweat data. These results suggest that betaine loss through sweat is greater than that lost through urine and may even exceed dietary intake in some cases. Collection of sweat using
regional patches is convenient and useful for relative comparisons, but the concentration of sweat constituents tends to be higher compared to values using whole body washdown [32,33]. Therefore further work is required to accurately determine total body loss, perhaps under varied exercise conditions. In addition, it would be valuable to determine any correlation between dietary intakes, serum concentrations, sweat concentrations and level of physical activity.

The data showed several statistically significant correlations between sweat metabolites. Not surprisingly, the strongest correlation was between sodium and chloride. Betaine was correlated with all components except sodium and chloride (somewhat surprising given the known relationship between betaine accumulation and salt tolerance). The correlation between lactate and potassium agrees with the correlation found (+0.78) in a previous study [33] in males. Muscle contractions cause lactic acidosis and loss of intracellular potassium with accumulation of extracellular potassium [34]. Lactic acidification has been shown to counteract the effects of elevated potassium associated with muscle fatigue [35]. This may form the basis of a correlation. Betaine, lactate and glucose were all correlated with each other. Lactate and glucose are closely related via anaerobic metabolism. Also, a study showed that ingestion of betaine led to elevated serum lactate [15]. Although the current study was not designed to determine causation, betaine may accumulate to protect the sweat gland, duct and surrounding tissue from the deleterious effects of elevated concentrations of inorganic ions, urea, ammonia and possibly lactate - which are known to perturb cellular metabolism. Betaine protects the kidney from high concentrations of electrolytes and urea [2,36,37], prevents myosin structural change due to urea [9], and protects against ammonia toxicity of neurons [14]. This may relate to the correlations between betaine, ammonia, urea, lactate and potassium found here in sweat. Further research on the significance and reproducibility of these correlations is warranted.

In conclusion, betaine is a component of sweat. Betaine is an osmoprotectant, and we speculate that it protects the sweat gland against the deleterious effects of other sweat components. Further research is warranted, such as evaluation of male and/or older athletes, sweat collection via total body washdown [38], and

### Table 1 Sweat composition of subjects

| Subject | Betaine (μmol·L⁻¹) | Choline (μmol·L⁻¹) | Lactate (mmol·L⁻¹) | Glucose (mmol·L⁻¹) | Sodium (mmol·L⁻¹) | Potassium (mmol·L⁻¹) | Chloride (mmol·L⁻¹) | Ammonia (mmol·L⁻¹) | Urea (mmol·L⁻¹) |
|---------|--------------------|--------------------|--------------------|-------------------|------------------|---------------------|---------------------|-------------------|---------------|
| 1       | 363                | 2.77               | 27.6               | 582               | 37.9             | 19.3*               | 29.1                | 11.73*            | 19.68         |
| 2       | 160                | 1.38               | 15.7               | 302               | 46.7             | 8.62                | 34.6                | 4.31              | 7.69          |
| 3       | 332                | 5.75*              | 27.2               | 447               | 46.6             | 8.73                | 35.2                | 6.75              | 13.77         |
| 4       | 277                | 0.98               | 18.7               | 415               | 52.4             | 9.06                | 37.7                | 5.41              | 6.75          |
| 5       | 140                | 1.17               | 13.8               | 272               | 52.0             | 6.20                | 36.5                | 3.01              | 7.67          |
| 6       | 157                | 1.61               | 23.1               | 491               | 40.9             | 9.11                | 26.5                | 6.40              | 12.61         |
| 7       | 196                | 1.01               | 18.5               | 411               | 36.3             | 8.03                | 24.9                | 5.57              | 9.17          |
| 8       | 229                | 2.28               | 18.0               | 356               | 81.7*            | 8.59                | 57.6*               | 3.34              | 8.59          |
| Average | 232                | 2.12               | 20.4               | 410               | 49.3             | 9.7                 | 35.3                | 5.81              | 10.74         |
| SD      | 84                 | 1.60               | 5.1                | 101               | 14.4             | 4.0                 | 10.2                | 2.74              | 4.38          |

* Outlier via Grubb’s Test (p < 0.05)

### Table 2 Pearson’s correlations (r) for sweat components

|        | Betaine | Choline | Lactate | Glucose | Sodium | Potassium | Chloride | Ammonia | Urea |
|--------|---------|---------|---------|---------|--------|-----------|----------|---------|------|
| Betaine| x       | +0.65*  | +0.78*  | +0.69*  | -0.08  | +0.70*    | +0.03    | +0.73*  | +0.67*|
| Choline| x       |         | +0.72*  | +0.36   | +0.02  | +0.21     | +0.10    | +0.36   | +0.55|
| Lactate| x       | +0.90*  |         | -0.36   | +0.67* | -0.31     | -0.85*   | +0.89*  |      |
| Glucose| x       | -0.45   | +0.79*  |         | -0.43  | +0.92*    | +0.86*   |         |      |
| Sodium | x       | -0.31   | +0.99*  | -0.57   |         | -0.43     |          |         |      |
| Potassium| x     | -0.23   | +0.92*  | +0.85*  |         |          |          |         |      |
| Chloride| x      | -0.50   | -0.37   |         |         |          |          |         |      |
| Ammonia| x       |         |         | +0.92*  |         |          |          |         |      |
| Urea   | x       |         |         |         |         |          |          |         |      |

*p < 0.05

*p < 0.10
Figure 1: Correlations between betaine and other components of sweat.
Table 3 Solute contents of sweat compared with published fasting values for plasma [18,23-26]

|                           | Sweat (S) | Plasma (P) |
|---------------------------|-----------|------------|
| Betaine (µmol L⁻¹)        | 232       | 340        |
| Choline (µmol L⁻¹)        | 2.1       | 14.5       |
| Lactate (mmol L⁻¹)        | 20.4      | 0.7        |
| Glucose (mmol L⁻¹)        | 0.41      | 4.9        |
| Sodium (mmol L⁻¹)         | 49.3      | 141        |
| Chloride (mmol L⁻¹)       | 35.3      | 105        |
| Ammonia (mmol L⁻¹)        | 5.81      | 0.07       |
| Urea (mmol L⁻¹)           | 10.74     | 5.7        |

acknowledgements

we would like to thank dr. lawrence armstrong (university of connecticut) for his valuable comments regarding the manuscript and dr. qing shi (university of north carolina) for conducting some of the analyses. current address of shona s. craig is ithaca college; ithaca ny. current address of matt ganio is texas health resources presbyterian hospital, dallas tx. some funding was provided by danisco a/s.

authors' contributions

ssc was the primary investigator, study design, data collection/analysis and manuscript draft. sasc conceived the study, supervised, statistical analysis, manuscript preparation. mgs, kac supervised and sweat analysis. cmm, gh, shz participated in concept, design, coordination and helped draft the manuscript. all authors read and approved the final manuscript.

competing interests

stuart craig is employed by danisco a/s, a manufacturer of betaine.

received: 21 december 2009

accepted: 22 january 2010 published: 22 january 2010

references

1. zeisel sh, mar mh, howe jc, holden jm. concentrations of choline-containing compounds and betaine in common foods. j nutr 2003, 133:1302-1307.
2. craig sas. betaine in human nutrition. am j clin nutr 2004, 80:539-549.
3. konstantinova sv, tell gs, vollset se, nygard o, bleie o, ueland pm. divergent associations of plasma choline and betaine with components of metabolic syndrome in middle age and elderly men and women. j nutr 2008, 138:914-920.
4. cho e, willett wc, collette ga, fuchs cs, wu k, chan at, zeisel sh, giovannucci el. dietary choline and betaine and the risk of distal colorectal adenoma in women. j natl cancer inst 2007, 102:124-131.
5. shaw gm, carmichael sl, yang w, selvin s, schaffer dm. periconceptional dietary intake of choline and betaine and neural tube defects in offspring. am j epidemiol 2004, 160:102-109.
6. slow a, lever m, chambers st, george fm. plasma dependent and independent accumulation of betaine in male and female rat tissues. physiol res 2009, 58:403-410.
7. yancey ph, clark me, hand sc, bowlus rd, somero gn. living with water stress: evolution of osmolyte systems. science 1982, 217:1214-1222.
8. olsen sn, ramlov h, westh p. effects of osmolytes on hexokinase kinetics combined with macromolecular crowding testing of the osmolyte compatibility hypothesis towards crowded systems. comp biochem physiol a mol integr physiol 2007, 148:339-345.
9. ortiz-costa s, sorenson mm, sola-penna m. betaine protects urea-induced denaturation of myosin subfragment-1. feds journal 2008, 13:3388-3396.
10. vrain s, willie d. the relationship between plasma urea levels and some muscle trimethylamine levels in xenopus laevis: a 31p and 1h nuclear magnetic resonance study. j exp biol 1995, 198:575-578.
11. vernet c, bride j, morel b, roeulx c, hembert p. glycine betaine stimulates human skin fibroblasts growth and collagen production in culture. j invest dermatol 2002, 118:1099.
12. warksulat u, reinein a, greither-beck s, kruftmann j, haussinger d. the osmolyte strategy of normal human keratinocytes in maintaining cell homeostasis. j invest dermatol 2004, 123:516-521.
13. coelho-sampaio t, feerreira st, castro ej, junior v, veyra a. betaine counteracts urea-induced conformational changes and uncoupling of the human erythrocyte ca2+ pump. eur j biochem 1994, 221:1103-1110.
14. minana m, hermenegildo c, lhanolasi m, montoliu c, grisoila s, felipo v. carntline and choline derivatives containing a trimethylamine group prevent ammonia toxicity in mice and glutamate toxicity in primary cultures of neurons. j pharmacol exp ther 1996, 279:194-199.
15. armstrong le, caja dj, roti mw, lee ec, craig sa, sutherland jw, fiala ka, maresch cm. influence of betaine consumption on strenuous running and sprinting in a hot environment. j strength cond res 2008, 22:851-860.
16. maresch cm, fellarj mj, kraemer wi, yamamoto lm, lee ec, armstrong le, hatfield dl, sokmen b, diao jc, speiring ba, et al. the effects of betaine supplementation on strength and power performance. med sci sports exerc 2007, 39:5101.
17. hoffman j, ratamess n, kang j, rashit s, faigenbaum a. effect of betaine supplementation on power performance and fatigue. journal of the international society of sports nutrition 2009, 6:7-11.
18. koc h, mar mh, ranasinghe a, swenberg ja, zeisel sh. quantitation of choline and its metabolites in tissues and foods by liquid chromatography/electrospray ionization-isotope dilution mass spectrometry. anal chem 2002, 74:4734-4740.
19. fnb. dietary reference intakes for water, potassium, sodium, chloride, and sulfate washington dc. the national academies press 2004.
20. dirosh a, shai i, tekes-manova d, israeli e, pered d, shochar t, kochba i, rudich a, the israeli diabetes research group. normal fasting plasma glucose levels and type 2 diabetes in young men. n engl j med 2005, 353:1494-1462.
21. lever m, szeland pc, basom lm, hayman cm, chambers st. glycine betaine and proline betaine in human blood and urine. biochim biophys acta 1994, 1200:259-264.
22. von allorwen hn, horn s, kahl j, feldheim w. the influence of lecithin on plasma choline concentrations in triathletes and adolescent runners during exercise. eur j appl physiol occup physiol 1993, 67:87-91.
23. warkulat u, brookman s, reinen a, haussinger d. ultraviolet b radiation induces cell shrinkage and increases osmolyte transporter mrna expression and osmolyte uptake in hacat keratinocytes. biochem biophys res commun 2000, 274:135-140.
24. bidulescu a, chambless l, siega-riz am, zeisel s, heiss g. repeatability and measurement error in the assessment of choline and betaine dietary intake: the atherosclerosis risk in communities (arc) study. nutrition journal 2009, 8:14.
25. cho e, zeisel sh, jacques p, selhub j, dougerty l, collette ga, willett wc. dietary choline and betaine assessed by food-frequency questionnaire
in relation to plasma total homocysteine concentration in the Framingham Offspring Study. *Am J Clin Nutr* 2006, 83:905-911.

30. Lever M, Atkinson W, Sizeland PC, Chambers ST, George PM: Inter- and intra-individual variations in normal urinary glycine betaine excretion. *Clin Biochem* 2007, 40:447-453.

31. Sawka MN, Montain SJ: Fluid and electrolyte supplementation for exercise heat stress. *Am J Clin Nutr* 2000, 72:564S-572.

32. Palacios C, Wigeritz K, Weaver CM: Comparison of 24 hour whole body versus patch tests for estimating body surface electrolyte losses. *Int J Sport Nutr Exerc Metab* 2003, 13:479-488.

33. Patterson MJ, Galloway SD, Nimmo MA: Variations in regional sweat composition in normal human males. *Exp Physiol* 2000, 85:869-875.

34. Brooks GA: Lactate doesn’t necessarily cause fatigue: why are we surprised?. *J Physiol* 2001, 536.1.

35. Nielsen OB, de Paoli F, Overgaard K: Protective effects of lactic acid on force production in rat skeletal muscle. *J Physiol* 2001, 536:161-166.

36. Horio M, Ito A, Matsuoka Y, Moriyama T, Orita Y, Takenaka M, Imai E: Apoptosis induced by hypertonicity in Madin Darley canine kidney cells: protective effect of betaine. *Nephrol Dial Transplant* 2001, 16:483-490.

37. Yancey PH, Burg MB: Counteracting effects of urea and betaine in mammalian cells in culture. *Am J Physiol* 1990, 258:R198-204.

38. Armstrong LE, Casa DJ: Methods to Evaluate Electrolyte and Water Turnover of Athletes. *Athletic Training & Sports Health Care* 2009, 1:169-179.

doi:10.1186/1550-2783-7-3

Cite this article as: Craig et al. The betaine content of sweat from adolescent females. *Journal of the International Society of Sports Nutrition* 2010 7:3.