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| Citation       | He, Meian, Huan Guo, Xiaobo Yang, Li Zhou, Xiaomin Zhang, Longxian Cheng, Hesong Zeng, Frank B. Hu, Robert M. Tanguay, and Tangchun Wu. 2010. Genetic variations in HSPA8 gene associated with coronary heart disease risk in a Chinese population. PLoS ONE 5(3): e9684. |
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| Published Version | doi:10.1371/journal.pone.0009684                                                                                                                                                                                                                                         |
| Citable link    | http://nrs.harvard.edu/urn-3:HUL.InstRepos:8000917                                                                                                                                                                                                               |
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Genetic Variations in *HSPA8* Gene Associated with Coronary Heart Disease Risk in a Chinese Population

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**Abstract**

**Background:** There is ample evidence that Hsp70 takes part in the progress of coronary heart disease (CHD). This implies that genetic variants of Hsp70 genes such as *HSPA8* (HSC70) gene might contribute to the development of CHD. The present study aimed to investigate whether certain genetic variants of *HSPA8* gene are associated with CHD in Han Chinese people.

**Methodology/Principal Findings:** A total of 2006 subjects (1003 CHD cases and 1003 age- and sex- matched healthy controls) were recruited. Genetic variants in the *HSPA8* gene were identified by sequencing of the gene in 60 unrelated Chinese. Four tag single nucleotide polymorphisms (tagSNPs) (rs2236659, rs2276077, rs10892958, and rs1461496) were selected and genotyped. The function of the significant SNP was evaluated using luciferase reporter assays in two cell lines. By sequencing the promoter and all exons and introns of the *HSPA8* gene, 23 genetic variants were identified. One promoter SNP rs2236659 was associated with susceptibility to CHD. Carriers of the “C” allele of rs2236659 had decreased CHD risk with odds ratio (OR) of 0.78 (95% CI: 0.62, 0.98; P = 0.033) after adjustment for conventional risk factors. Haplotype analyses indicated that haplotype GGC, contributed to a lower CHD risk (OR = 0.78, 95% CI: 0.65, 0.93; P = 0.006) compared with the common haplotype AGGT. In a transfection assay, the C allele of rs2236659 showed a 37–40% increase in luciferase expression of the reporter gene luciferase in endothelial and non-endothelial cells compared with the T allele.

**Conclusions/Significance:** These findings suggest that genetic variants in *HSPA8* gene (especially promoter SNP rs2236659) contribute to the CHD susceptibility by affecting its expression level.

Citation: He M, Guo H, Yang X, Zhou L, Zhang X, et al. (2010) Genetic Variations in *HSPA8* Gene Associated with Coronary Heart Disease Risk in a Chinese Population. *PLoS ONE* 5(3): e9684. doi:10.1371/journal.pone.0009684

Editor: Florian Kronenberg, Innsbruck Medical University, Austria

Received: December 1, 2009; Accepted: February 20, 2010; Published: March 16, 2010

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Funding: This work was supported by grants from the National Natural Science Foundation (grant NSFC-30525031 and NSFC-30430590) and a China-Canada Joint Health Research Grant of the Canadian Institutes of Health Research (CIHR) and the National Natural Science Foundation (grant NSFC-3071120579) to RMT and TW. FBH’s work was supported in part by the American Heart Association Established Investigator Award. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

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**Introduction**

Coronary heart disease (CHD) is a complex disease with high morbidity and mortality. Very little is known about its genetic etiology. Heat shock protein 70 (HSP70), as a dominant chaperone in the HSPs families, can help in the assembly of newly synthesized proteins, in protein transport, and in the removal of damaged proteins [1]. In humans, the HSP70kDa family comprises 13 members, some of which show constitutive expression while others are stress inducible [2]. These isoforms have highly homogenous structure. They are all composed of a conserved ATPase domain, a peptide-binding domain, a middle region with protease sensitive sites, and a C-terminal domain [3,4]. For instance, HSPA8, previously referred to as HSP73 or HSC70, shares 86% amino acid homology to inducible HSP40 [3]. Consistent with their homogenous structure, these proteins have distinct but overlapping functions [3]. Thus both stress-inducible Hsp70 and constitutively expressed HSPA8 can perform similar functions and are capable of protecting cardiac muscle cells against injuries like an oxidative challenge [5,6]. There is much evidence indicating that Hsp70 can take part in the progress of CHD [7-9]. A previous study from our laboratory also demonstrated that genetic variants in the *HSP70* gene may be novel genetic risk markers for CHD [10]. Based on their high degree of structural homology and similar function in protecting against injuries in cardiac muscle cell, it is conceivable that the main constitutively-expressed member of the HSP70 family, HSPA8 might also be involved in the development of CHD and that single nucleotide polymorphisms (SNPs) and haplotypes of this gene may be associated with CHD and contribute to CHD susceptibility.

To test this hypothesis, we first sequenced and identified all SNPs in the *HSPA8* gene in 60 unrelated Han Chinese. We then...
selected 4 tagging SNPs (tagSNPs) to identify potential genetic markers of this gene for CHD susceptibility in a case-control study comprised of 1,003 CHD cases and 1,003 age- and sex-frequency matched controls in a Chinese population. We also examined the function of the SNPs associated with CHD susceptibility by performing a reporter gene luciferase activity assay in two types of cell lines.

Results

SNPs Identification in the HSPA8 Gene in Han Chinese

In total we found 23 genetic variants in HSPA8 gene with minor allele frequency (MAF) from 5.0% to 45.0% (Table 1). Among those, 6 novel SNPs had not been reported previously, including 301 c/g and 395 c/g (intron 1), 1255 t/c (intron 2), 1671 c/t (intron 3), 2692 gacctc/- and 2716 ca/- (intron 5); The remaining 17 SNPs have been reported in NCBI as shown in Table 1 (http://www.ncbi.nlm.nih.gov/genome). SNP rs11218942 (G>T> C) is triallelic and was not analyzed in further linkage disequilibrium analysis and tagSNPs selection.

Selection of tagSNPs in HSPA8 Gene

Based on the above sequencing data, linkage disequilibrium analysis results showed that all detected SNPs located in the same haplblock (Figure 1). The ltSNPer1.0 software was used to pick out the tagSNPs[11], and finally four SNPs were selected as tagSNPs, including rs2236660, rs2236658, rs10892958 and rs1461496 (Figure 2). Because the sequences around the SNPs rs2236660 and rs2236658 are rich in GC and not suitable to be detected by TaqMan SNP allelic discrimination assay, we selected the other two SNPs rs2236659 and rs2276077, which are in high linkage with rs2236660 and rs2236658 for further analysis.

General Characteristics of the Subjects

The general characteristics of the CHD cases and controls have been described in a previous study[12] and are summarized in Table 2. CHD patients had a higher prevalence of conventional vascular risk factors, including smoking, non-drinking, history of hypertension and diabetes mellitus, family history of CHD and higher level of FBG, whereas TC level in patients were surprisingly lower than in controls probably due to cholesterol-lowering treatment in the cases.

HSPA8 Genotypes and CHD Risk

The genotype frequencies of the four studied SNPs in HSPA8 polymorphisms are summarized in Table 3. The distributions of SNPs rs2236659, rs2276077, rs10892958 and rs1461496 did not depart from the Hardy-Weinberg equilibrium in control group (P=0.73, 0.62, 0.79 and 0.22 respectively). There was significant difference in genotype distribution of rs2236659 between CHD and controls. Adjustment for the conventional risk factors such as age, sex, pack-years of smoking, drinking, activity, hypertension, DM and family history of CHD did not appreciably alter the results. Compared with TT genotype of rs2236659, subjects with C allele had lower risk of CHD after adjustment for the conventional risk factors above (Crude odds ratio (OR) = 0.83, 95% CI: 0.69, 1.00; P=0.049 and adjusted OR = 0.78, 95% confidence interval (CI): 0.62, 0.98; P=0.033 respectively) (Table 3). Stratified analysis according to age (<60 years and >60 years), sex and smoking status indicated that subjects with C allele of rs2236659 in men, older (>60 years) or smokers subgroups had significant lower risk of CHD. However in females, younger or non-smokers subgroups there were no significant differences. Further analysis indicated that there were no interactions between SNP rs2236659 and above factors respectively (data not shown). There were no significant differences between CHD and control group in SNPs of rs2276077, rs10892958 and rs1461496 before or after adjusting for conventional risk factors (P >0.05).

Haplotypes Associations with CHD Risk

All the pairwise LD measure D' of the four investigated tagSNPs in the HSPA8 gene ranged from 0.90 to 0.96 (data not shown). A total of 14 and 12 haplotypes were estimated in the CHD and control groups respectively by using PHASE 2.0 software to reconstruct haplotypes based on the observed genotypes[13]. Among these, 5 haplotypes of AGGT, GCCG, GGTG, and GGGT were >1.0% (from left to right the order of polymorphic bases in haplotype is rs1461496, rs10892958, rs2276077, and rs2236659). The associations between the common haplotypes (covering 96.95% and 98.16% of allelic variance in CHD and controls, respectively) encompassing HSPA8 polymorphisms and CHD risk were also examined. Compared with the highest-frequency haplotype of AGGT, the GCCG haplotype had 22% lower risk of CHD (OR = 0.78, 95% CI: 0.65, 0.93; P=0.006). However, haplotype GCCG, which is only different from GCCG in rs2236659, had no significant difference compared with AGGT (OR = 1.00, 95% CI: 0.85, 1.17; P=0.956), confirming the results.

Table 1. Variants discovered by sequencing all HSPA8 exons and introns and 1 Kb upstream of transcript start site in 60 unrelated Chinese.

| Variants | Position | Gene pos. | Alleles | MAF (%) | Heterozygozity |
|----------|----------|-----------|---------|---------|----------------|
| rs11218942 | −926 | promoter | G>T>C | 13.3 | 0.48 |
| rs2236660 | −703 | promoter | T>C | 14.2 | 0.24 |
| rs2236659 | −357 | promoter | T>C | 15.8 | 0.27 |
| rs2236658 | −308 | promoter | A>G | 35.8 | 0.46 |
| rs2276077 | 8 | 5’ UTR | G>A | 19.2 | 0.31 |
| rs2276075 | 23 | 5’ UTR | C>T | 19.2 | 0.31 |
| rs1136141 | 48 | 5’ UTR | C>T | 5.0 | 0.09 |
| rs2276074 | 49 | 5’ UTR | C>T | 30.0 | 0.42 |
| New1 | 301 | intron1 | C>G | 19.2 | 0.31 |
| New2 | 395 | intron1 | C>G | 17.5 | 0.29 |
| rs11218941 | 416 | intron1 | T>C | 37.5 | 0.47 |
| rs4935825 | 460 | intron1 | T>G | 1.7 | 0.03 |
| rs18823704 | 647 | intron1 | G>T | 18.3 | 0.30 |
| rs10892958 | 702 | intron1 | G>C | 40.4 | 0.48 |
| New3 | 1255 | intron2 | T>C | 20.0 | 0.32 |
| New4 | 1671 | intron3 | C>T | 12.5 | 0.22 |
| New5 | 2692 | intron5 | Ins GACTC | 12.5 | 0.22 |
| New6 | 2716 | intron5 | Ins CA | 5.8 | 0.11 |
| rs1461496 | 3201 | intron6 | C>T | 44.5 | 0.49 |
| rs1064585 | 3418 | exon7 | A>C | 5.8 | 0.11 |
| rs4935770 | 3950 | intron8 | A>G | 45.0 | 0.49 |
| rs3763897 | 4191 | intron8 | T>A | 11.7 | 0.21 |
| rs4802 | 4203 | exon9 | C>T | 45.0 | 0.49 |

*Reference the transcription start code as +1.

**Novel SNP not reported in public database before.**

doi:10.1371/journal.pone.0009684.t001

PLoS ONE | www.plosone.org | 2 March 2010 | Volume 5 | Issue 3 | e9684
of single SNP analyses that subjects with C allele of rs2236659 had lower risk of CHD (Table 4).

**HSPA8 Promoter Region Carrying the −357C (rs2226659) Leads to Higher Expression in a Reporter Assay**

In order to understand the functional significance of the −357 T/C change, we used a reporter assay with luciferase. As shown in Figure 3, relative luciferase expression driven by the T-C-G containing promoter were 37–40% higher than that by the haplotype T-T-G containing promoter in the two types of cell lines (P = 0.008 for HBE and 0.046 for HUVEC). Because the two haplotypes of T-C-G and T-T-G are only different in −357T/C, these results suggested that the −357C variant (rs2236659) had a higher promoter activity than −357T allele. Similar results were obtained for the T-T-A haplotype when comparing with haplotypes T-T-G or C-C-G in HBE; however, there were no significant differences observed in HUVEC cells.

**Discussion**

Our study is the first one to examine the associations of variants of a constitutively expressed member of the HSP70 family, HSPA8 and CHD susceptibility. Subjects with the C allele of rs2236659 had lower risk of CHD independent of other conventional risk factors. Further functional study in a cell reporter assay suggested that this association might be due to the increased promoter activity of the C allele of rs2236659 which may result in higher levels of expression of this HSPA8 protein.

HSPA8 is constitutively expressed and only mildly induced during stress situations[14]. It plays an important role in folding protein during their synthesis, transporting protein across membranes, regulating stress response and it is also involved in cell survival[4,15]. In cardiac muscle cells, overexpression of HSPA8 attenuated oxidative injuries and enhanced cell survival[5]. This implies that HSPA8 might participate in the progress of CHD since it is believed that oxidative injuries are involved in the etiology of CHD[16]. Variants of HSPA8 gene could affect HSPA8 levels and/or function. HSPA8 might take part in the development of CHD by two ways. First, as mentioned above it is believed that reactive oxygen species (ROS) are involved in the etiology of CHD[16–18]. This Hsp could protect against endogenous or exogenously generated ROS[5] and thus contribute to the progression of CHD. Second, this protein has also been reported to protect against hypoxia-induced apoptosis in hypoxia-induced apoptosis-resistant macrophages[19] and in the control of apoptosis during embryogenesis[20]. Other studies have found
that HSPA8 protects cells against injuries by suppressing of apoptosis signaling and that its overexpression results in resistance against stress-induced caspase activation[4,21]. Although to our knowledge no previous studies have investigated the role of HSPA8 in endothelium cells apoptosis, it might protect endothelium cells against apoptosis, which is believed to be the initiating event of atherogenesis and plays a crucial role in the transition from a stable endothelialized plaque to plaque erosion and thrombosis[22].

SNPs in the promoter of the HSPA8 gene might affect its level of transcription and then lead to similar changes at the protein level. The SNP rs2236659 locates 357 bp upstream of transcriptional start code. In silico analysis using bioinformatics softwares of Alibaba2.0 (Niels Grabe, http://www.gene-regulation.com/pub/programs/alibaba2/) and TESS program (http://agave.humgen.upenn.edu/utess/tess), predicts that the C allele has stronger binding capacity with transcriptional factor sp1 when compared with the T allele. Consistent with the computation, our study confirmed that C allele of SNP rs2236659 leads to an increase in promoter activity and probably heightens synthesis level of the corresponding HSPA8 protein which could decreases the risk of CHD. By using global expression data available from human lymphoblastoid cell lines[23], we did not find any evidence that rs2236659 directly influences HSPA8 expression. However, further studies conducted in cardiac tissues, vascular smooth muscle and/or endothelial cells would be necessary to draw conclusions about the possible effects of the rs2236659 SNP on HSPA8 expression.

Several strengths of this study should be acknowledged. First, our population is racially homogeneous (all Han Chinese), which weakens the possible biases from population stratification. In addition, the findings from case-control association study were confirmed by detailed functional assays in both endothelial and nonendothelial cell lines strengthened the association of the HSPA8 gene variations and CHD. However, three major limitations should also be mentioned. Firstly, like all case-control studies, selection bias (inclusion of patients surviving CHD) may exist and might influence interpretation of the results. Secondly, the controls selected without performing coronary angiography might include some false negative cases. However, because the prevalence of

Figure 2. Construction of haplotype and tagging SNPs selection in HSPA8 gene. A total of 22 single nucleotide polymorphisms (SNPs) were analyzed by htSNPer1.0 software to select tagSNPs. 1.0 indicated rs2236660 and 22.0, the last SNP rs4802. The results showed that rs2236660, rs2236658, rs10892958 and rs1461496 were selected for tagSNPs.

doi:10.1371/journal.pone.0009684.g002

Table 2. General characteristics of CHD cases and controls*.

| Variables                        | Cases (n = 1003) | Controls (n = 1003) | P Value |
|----------------------------------|-----------------|---------------------|---------|
| Sex (male/female)                | 638 (63.8)/365  | 637 (63.7)/366      | 0.94    |
| Age (y)                          | 59.7 (8.8)      | 59.5 (8.5)          | 0.31    |
| Systolic blood pressure (mmHg)   | 134.6 (24.6)    | 131.6 (21.1)        | 0.004   |
| Diastolic blood pressure (mmHg)  | 82.0 (14.4)     | 82.5 (11.1)         | 0.45    |
| Body mass index                  | 24.46 (3.51)    | 24.41 (3.37)        | 0.77    |
| FBG (mmol/L)                     | 6.66 (3.63)     | 5.32 (2.00)         | <0.01   |
| TC (mmol/L)                      | 4.50 (1.05)     | 5.08 (1.30)         | <0.01   |
| TG (mmol/L)                      | 1.73 (1.24)     | 1.69 (1.31)         | 0.45    |
| Smoking (no/past or current)     | 435 (43.5)/564  | 535/468             | <0.01   |
| Smoking index (pack-year)        | <0.01           |                     |         |
| 0                                | 435 (43.3)      | 535 (53.3)          |         |
| 0–31                             | 281 (28.1)      | 273 (27.2)          |         |
| 31+                              | 283 (28.4)      | 195 (19.5)          |         |
| Drinking (no/yes)                | 733 (73.4)/266  | 681/316             | 0.01    |
| Past history                     | 705 (70.4)      | 342 (34.9)          | <0.01   |
| Hypertension                     | 277 (27.8)      | 63 (6.5)            | <0.01   |
| Diabetes mellitus                | 111 (11.1)      | 31 (3.1)            | <0.01   |

*Data are means (standard deviation) for continuous variables and n (%) for categorical variables.

doi:10.1371/journal.pone.0009684.t002
CHD in China is still low [24], and our controls all had normal ECG and no clinical symptoms before enrollment, the false negative cases in the controls are likely to be rare. Finally, replication is the best way to validate an association, however, the present matched case-control study is well designed and had enough power (0.80%) to detect SNPs with risk ratios 1.35, 1.30, and 1.25, given an α of 0.05 and allele frequencies of 0.1, 0.3, and 0.5, respectively. In addition, our detailed functional assays conducted in both endothelial and nonendothelial cell lines confirmed and strengthened the associations of the HSPA8 gene variations and CHD. However, these associations still need to be validated in other ethnic groups.

In summary, our case-control study and reporter assays results suggest that variants in HSPA8 gene contribute to CHD susceptibility. Future studies are needed to validate these findings and further investigate potential mechanisms underlying the links between variations of HSPA8 gene and CHD risk.

**Materials and Methods**

**Screening for SNPs in HSPA8 Gene**

DNA samples extracted from whole blood of 60 randomly selected healthy Han Chinese (28 males and 32 females) were used to identify SNPs in HSPA8 gene (GenBank accession NM_006597.3). Resequencing region included all HSPA8 exons and introns and 1 Kb upstream of the transcript start site. Genomic DNA was amplified and then purified using the ethanol/NaAc method. The PCR products were used as templates for sequencing reactions with the BigDye Terminator kit v3.1 (Applied Biosystems, Foster City, CA, USA). Purified sequencing reactions were run on an ABI 3100 genetic analyzer. Sequence analysis, SNP detection, and genotype were performed using Sequencing Analysis 5.1.1 and DNAStar software. All primers and reaction conditions are displayed in Table S1.

**Human Subjects**

The study design for this investigation has been described earlier [12]. Briefly, the study population was composed of 1,003

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**Table 3. Analysis of associations between the HSPA8 polymorphisms and risk of CHD.**

| Genotype | Cases N (%) | Controls N (%) | Crude OR (95% CI) | Adjusted OR (95% CI)* |
|----------|-------------|----------------|-------------------|----------------------|
| rs1461496 |             |                |                   |                      |
| GG       | 321 (33.3)  | 341 (34.4)     | 1.00              | 1.00                 |
| AG       | 435 (45.1)  | 464 (46.9)     | 1.00(0.82, 1.22)  | 1.01(0.79, 1.29)     |
| AA       | 209 (21.7)  | 185 (18.7)     | 1.20(0.94, 1.54)  | 1.19(0.88, 1.62)     |
| AG+AA    | 644 (66.8)  | 649 (65.6)     | 1.05(0.87,1.27)   | 1.06(0.84, 1.34)     |
| rs10892958 |            |                |                   |                      |
| GG       | 336 (34.6)  | 305 (30.7)     | 1.00              | 1.00                 |
| GC       | 453 (46.7)  | 495 (49.8)     | 0.83(0.68, 1.02)  | 0.79(0.62, 1.02)     |
| CC       | 181 (18.7)  | 194 (19.5)     | 0.85(0.66, 1.09)  | 0.82(0.60, 1.13)     |
| GC+CC    | 634 (65.4)  | 689 (69.3)     | 0.84(0.69, 1.01)  | 0.80(0.64, 1.01)     |
| rs2276077 |            |                |                   |                      |
| GG       | 721 (74.6)  | 749 (75.5)     | 1.00              | 1.00                 |
| AG       | 228 (23.6)  | 228 (23.0)     | 1.04(0.84, 1.28)  | 0.99(0.76, 1.28)     |
| AA       | 18 (1.9)    | 15 (1.5)       | 1.25(0.62, 2.49)  | 2.00(0.81, 4.99)     |
| AG+AA    | 246 (25.5)  | 243 (24.5)     | 1.05(0.86,1.29)   | 1.03(0.81,1.33)      |
| rs2236659 |            |                |                   |                      |
| TT       | 652 (68.0)  | 624 (63.7)     | 1.00              | 1.00                 |
| CT       | 267 (27.8)  | 313 (32.0)     | 0.82(0.67, 0.99)  | 0.76(0.59, 0.97)     |
| CC       | 40 (4.2)    | 42 (4.3)       | 0.91(0.58, 1.43)  | 0.93(0.54, 1.61)     |
| CT+CC    | 307 (32.0)  | 355 (36.3)     | 0.83(0.69, 1.00)  | 0.78(0.62, 0.98)     |

*Adjusted for age, sex, pack-years of smoking, drinking, activity, hypertension, diabetes mellitus and family history of CHD.

doi:10.1371/journal.pone.0009684.t003

**Table 4. Haplotype distribution of HSPA8 in CHD and control group.**

| Haplotype | CHD N (%) | Controls N (%) | OR (95% CI) | P value |
|-----------|-----------|----------------|-------------|---------|
| AGGT      | 850 (42.37) | 819 (40.83) | 1.00 | - |
| GCGT      | 495 (24.68) | 479 (23.88) | 1.00(0.85, 1.17) | 0.958 |
| GGCC      | 317 (15.80) | 392 (19.54) | 0.78(0.65, 0.93) | 0.006 |
| GGAT      | 245 (12.21) | 251 (12.51) | 0.94(0.77, 1.15) | 0.549 |
| GGGT      | 38 (1.89) | 28 (1.40) | 1.31(0.80, 2.15) | 0.291 |
| Others †  | 61 (3.04) | 37 (1.84) | 1.59(1.04, 2.42) | 0.031 |
| Total     | 2006 100 | 2006 100 | - | - |

*Polymeric bases were in 3’ to 5’ order and from left to right the order is rs1461496, rs10892958, rs2276077 and rs2236659. Other haplotype includes ACAT, GGGC, AGGC, AGAT, GCAT, ACGC, ACC, AGAC, GGAC, ACAC, and ACAT with frequency <1.0%.  

doi:10.1371/journal.pone.0009684.t004
Transcript luciferase reporter gene expression assays with constructs containing different haplotypes of HSPA8 promoter in human bronchial epithelial cells (HBE) and human umbilical vein endothelial cells (HUVEC). Upper: schematic representation of different reporter gene constructs. Variants of -703C/T, -357C/T, and -308A/G represent rs2236659, rs2236658 and rs2236656 respectively; lower: luciferase expression of these constructs. All constructs were cotransfected with pRL-SV40 to standardize the transfection efficiency. Fold increase of luciferase activity was calculated by defining the activity of the empty pGL3-Basic vector as 1. All experiments were performed in triplicates at least in three independent transfection experiments and each value represents mean (standard deviation). * and † indicated that P<0.05, compared with each of the construct.

Figure 3. Effects of haplotypes in promoter of HSPA8 on luciferase activity. Transient luciferase reporter gene expression assays with constructs containing different haplotypes of HSPA8 promoter in human bronchial epithelial cells (HBE) and human umbilical vein endothelial cells (HUVEC). Upper: schematic representation of different reporter gene constructs. Variants of -703C/T, -357C/T, and -308A/G represent rs2236659, rs2236658, and rs2236656 respectively; lower: luciferase expression of these constructs. All constructs were cotransfected with pRL-SV40 to standardize the transfection efficiency. Fold increase of luciferase activity was calculated by defining the activity of the empty pGL3-Basic vector as 1. All experiments were performed in triplicates at least in three independent transfection experiments and each value represents mean (standard deviation). * and † indicated that P<0.05, compared with each of the construct.

doi:10.1371/journal.pone.0009684.g003

Genotyping of HSPA8 Polymorphisms

Fasting venous blood was collected in 5 ml heparin tubes, and genomic DNA was isolated with a Puregene kit (Gentra Systems, Inc., Minneapolis, MN, USA). Genotyping was performed with TaqMan SNP allelic discrimination method on an ABI 7900HT real-time quantitative polymerase chain reaction (PCR) system (Applied Biosystems), in 304-well format. PCR reactions were carried out in reaction volume of 5 μl containing 5 ng DNA, 2.5 μl 2× TaqMan universal PCR Master Mix, No AmpErase UNG (Applied Biosystems), 0.125 μl 40× Assay Mix (Applied Biosystems). PCR conditions included 95°C for 10 min followed by 40 cycles of 15 s at 92°C and 1 min at 60°C. Two blank controls (DNA hydration solution) and two replicate quality control samples were included in each 384-well format, and two replicate samples were genotyped with 100% concordance. The intensity of each SNP met the criteria of three clear clusters in two scales generated by SDS software (Applied Biosystems). The TaqMan primers and probes are displayed in Table S2. Finally, genotyping failed in 15 (1.30%), 9 (0.90%), 11 (1.10%) and 24 (2.40%) controls and 30 (3.00%), 35 (3.3 0%), 36(3.60%) and 44(4.40%) cases in rs1461496, rs10892958, rs2276077 and rs2236659 locus respectively owing to DNA quantity or quality.

Reportor Plasmids Construction

Because rs2236659 (-357T/C) was associated with CHD risk and located in the core promoter region of HSPA8, we evaluated whether this variant had allele-specific effect on its transcriptional activity. We constructed plasmid containing 3 promoter SNPs (rs2236660 -703T/C, rs2236659 -357T/C, and rs2236658 -308A/G). Firstly, we amplified the -1 to -780 promoter region of the HSPA8 gene, and then inserted it into Kpn I/Hind III enzyme sites of pGL3-Basic (Promega, Madison, Wisconsin, USA). The first constructed plasmid contained the T-T-A haplotype (from left to right: -703T/C, 357T/C, and -308A/G) of HSPA8 (Figure 3). Primer pairs of amplification and site-specific mutagenesis are listed in Table S3. The direction and sequence authenticity of the above constructs were validated by restriction analysis and direct sequencing.

Transient Transfection and Luciferase Reporter Assays

Human umbilical vein endothelial cells (HUVEC) and human bronchial epithelial cells (HBE) with density of 6×10³ were seeded into 48-well plates and transfected with 100 ng pGL3-Basic plasmids or its constructs (defined as -703T/-357T/-308A, -703T/-357T/-308G, -703T/-357C/-308G, and -703C/-357C/-308G) and 1 ng pRL-SV40 (Promega) using Lipofectamine 2000 (Invitrogen, Carlsbad, California, USA) when grown to 70% confluence. Luciferase activity was measured at 24-hr using Dual-Luciferase Reporter Assay System (Promega) on a TD-20/20n luminometer (Turner Design, Promega). Each construct was tested in triplicates and the transfection experiments were performed three times independently. The results are denoted as relative luciferase activity (RLA) since the luciferase activity was normalized by Renilla luciferase activity and the empty pGL3-Basic vector.

Biological Variables Determination

Fasting blood glucose (FBG), total cholesterol (TC), and triglyceride (TG) were assayed using standard laboratory procedures
at the Department of Clinical Laboratory at the Wuhan Union Hospital.

**Statistical Analysis**

A chi-square test was applied to compare categorical variables and the Hardy-Weinberg equilibrium of the polymorphisms. A multiple logistic regression analysis was used to evaluate the association between SNPs and CHD with appropriate adjustment of cardiovascular risk factors. The ANOVA test was used to examine the differences in luciferase reporter gene expression. The linkage relationship between the four SNPs in HSPA8 gene was measured by the linkage disequilibrium (LD) coefficient (D’), calculating by JLIN[25] and LDA program[26]. The htSNPer1.0 software was used to select tagSNPs in HSPA8gene[11]. All genotype data for each sample were taken to construct the haplotypes by using the PHASE 2.0 program[13]. P<0.05 was considered statistically significant. All data analyses were carried out using SPSS 12.0 software (SPSS Inc., Chicago, Illinois, USA). Power calculations were performed using Quanto 1.2.3 (available from http://hydra.usc.edu/gxe).

**Supporting Information**

**Table S1**

| Primers and PCR conditions for sequencing HSPA8 gene. |

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**Acknowledgments**

We would like to thank all individuals who voluntarily participated in the present study and the medical assistants of Tongji Hospital, Union Hospital and Wugang Hospital in Wuhan city, Hubei Province, China.

**Author Contributions**

Conceived and designed the experiments: MH RMT TW. Performed the experiments: MH HG XY LZ XZ LC HZ. Analyzed the data: MH HG XY LZ XZ HF FBH TW. Contributed reagents/materials/analysis tools: LC TC. Wrote the paper: MH HG FBH RMT TW.

Found at: doi:10.1371/journal.pone.0009684.s001 (0.06 MB DOC)

**Table S2**

| Primers and probes for genotyping 4 TagSNPs in HSPA8 gene. |

Found at: doi:10.1371/journal.pone.0009684.s002 (0.05 MB DOC)

**Table S3**

| Primer sequences used in amplification and reporter plasmids construction. |

Found at: doi:10.1371/journal.pone.0009684.s003 (0.05 MB DOC)