Short Term Isocaloric Ketogenic Diet Modulates NLRP3 Inflammasome Via B-hydroxybutyrate and Fibroblast Growth Factor 21

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A ketogenic diet (KD) is known to have beneficial health effects. Various types of KD interventions have been applied to manage metabolic syndrome based on modification of diet parameters such as duration of intervention, macronutrient components, and total calories. Nevertheless, the beneficial health impact of isocaloric KD is largely unknown, especially in healthy subjects. The present study investigated the acute effects of a 3-day isocaloric KD. In this non-randomized intervention study, we recruited 15 healthy volunteers aged 24-38 years (7 men and 8 women) and placed them on an isocaloric KD restricting intake of carbohydrates but not energy (75% fat, 20% protein, 5% carbohydrate) for 3 days. Biochemical profiles and laboratory measurements were performed. Peripheral blood mononuclear cells were cultured, and measured cell stimulated cytokines. After short-term isocaloric KD, subjects lost body weight and serum free fatty acid levels were increased. These results accompanied elevated serum β-hydroxybutyrate (BHB) concentration and fibroblast growth factor 21 (FGF21) levels and improved insulin sensitivity. Regarding the direct effect of BHB on inflammasome activation, interleukin-1β (IL-1β) and tumor necrosis factor-α secretion in response to adenosine triphosphate or palmitate stimulation in human macrophages decreased significantly after isocaloric KD. In ex-vivo experiments with macrophages, both FGF21...
INTRODUCTION

Obesity and diabetes are global health concerns. A predominant clinical and pathological characteristic of these conditions is chronic low-grade inflammation, a key component in the pathogenesis of insulin resistance and metabolic syndrome (1). Although medical intervention to counteract obesity and diabetes is a common solution, dietary and lifestyle changes are also critical factors in resolving these problems (2). Ketogenic diet (KD) is known to have beneficial health effects. It has been utilized for nearly a century to treat epilepsy. Recent evidence suggests beneficial effects from KD to manage metabolic disorders by reducing body weight and fat accumulation and ameliorating liver steatosis and insulin resistance (2–4). The KD regimen has been shown to improve cardiomyopathy in a diabetic mouse model (db/db) and in humans by unloading mitochondrial burden (5, 6). Patients with obesity and type 2 diabetes mellitus gain better control of glucose levels when on a very low carbohydrate diet (7, 8). In addition, dietary effects were found to improve and prevent cognitive decline in human trials of dementia (9).

A KD is comprised of very low carbohydrate content (5%-10% of total daily calorie intake, or 20-50 g/day), which can induce ketosis via production of β-hydroxybutyrate (BHB), acetoacetate (AcAC), and acetone, mainly in the liver (10). Hepatocyte-derived AcAC in macrophages suppressed hepatic stellate cells, resulting in protection against tissue fibrosis (11). BHB has been exclusively studied among other ketone bodies. High levels of BHB ameliorate inflammation and increase lean mass in patients with multiple sclerosis (12). Lipopolysaccharide (LPS)-induced NLRP3 inflammasome activation and IL-1β production are suppressed in macrophages that protect against muscle loss, suggesting a possible therapeutic effect of KD to treat pathophysiology associated with inflammation (13). BHB also acts on neutrophils, which inhibit NLRP3 inflammasome activation and block IL-1β secretion in mice and humans (14, 15). Further, it acts as a signal molecule by binding to hydroxy-carboxylic acid receptor 2 (16, 17). There was a report that fibroblast growth factor 21 (FGF 21) increases insulin sensitivity (18) and the BHB signaling pathway (19). However, the precise mechanisms underlying these effects remain incompletely understood.

Various types of KD interventions have been applied to manage metabolic disorders depending on modification of diet parameters, such as duration of intervention, macronutrient components, and total calorie. The beneficial health impact of isocaloric KD is largely unknown, especially in healthy subjects. Thus, the underlying mechanisms that explain the effect of isocaloric KD are unclear.

Therefore, the present study aimed to investigate the acute effects of isocaloric KD on metabolic parameters in healthy subjects. Furthermore, we evaluated macrophages as mediators of isocaloric KD effects.

METHODS

Study Design: Isocaloric Ketogenic Diet in Healthy Subjects

This study was carried out at Severance Hospital. Eligible participants were healthy adults (19–44 years of age) with a BMI of at least 18 kg/m². Exclusion criteria included: any disease including diabetes, hypertension, and dyslipidemia; any current medication; or pregnant women. We recruited 15 healthy volunteers aged 24–38 years of age (7 men and 8 women) and placed them on an isocaloric KD restricting intake of carbohydrates but not energy (75% fat, 20% protein, 5% carbohydrate) for 3 days (Figure 1). Total energy (calories)/day were determined by the ‘Dietary Reference Intakes for Koreans’ (20): men, 10.88 MJ (2,600 kcal)/day for age under 30 and 10 MJ (2,400 kcal)/day for age 30 or older; women, 8.79 MJ (2,100 kcal)/day for age under 30 and 7.95 MJ (1,900 kcal)/day for age 30 or older. Participants’ body composition was analyzed by bioelectrical impedance (InBody 720, BioSpace Co., Seoul, Korea) before and after the KD intervention at the same time of day (early morning). This study was approved by the Institutional Review Board at Severance Hospital (4-2016-0795), is registered at clinicaltrials.gov (NCT02964572), and complied with the revised 2008 Declaration of Helsinki. All participants provided written informed consent prior to initiation of any study procedure.

Outcome Measures

The primary endpoint was change in secretion levels of IL-1β from supernatants of macrophages from baseline to end of the isocaloric KD regimen. Secondary endpoints included: change in

Abbreviations: AcAC, acetoacetate; ATP, adenosine triphosphate; BHB, β-hydroxybutyrate; DIO, diet-induced obese; FGF21, fibroblast growth factor 21; FFA, free fatty acid; HOMA-IR, homeostatic model assessment of insulin resistance; IL-1β, interleukin-1β; KD, ketogenic diet; LPS, lipopolysaccharide; N.S., non-significant; PA, palmitate; PBMC, peripheral blood mononuclear cell; QUICKI, quantitative insulin sensitivity check index; TNF-α, tumor necrosis factor-α.
secretion levels of tumor necrosis factor-α (TNF-α) from supernatants of macrophages; serum levels of IL-1β, BHB, insulin, FGF21, and other biochemical profiles [free fatty acid (FFA), glucose, and total cholesterol]; and change in body weight from baseline to end of the isocaloric KD regimen.

**Laboratory Measurements**

After overnight fasting, serum BHB was determined by an enzymatic assay using a commercial reagent from Nittobo Medical Co., LTD (Tokyo, Japan) and the Hitachi 7600 analyzer. Serum glucose, FFA, and total cholesterol were measured using a Hitachi 7600 automated chemistry analyzer (Hitachi High-Technologies Corporation, Tokyo, Japan). Fasting serum insulin was measured by electrochemiluminescence immunoassay using a Cobas e601 analyzer (Roche Diagnostics, GmbH, Germany). Insulin sensitivity was assessed using the following indices (21): homeostatic model assessment of insulin resistance (HOMA-IR) = [(fasting serum insulin [µU/mL] × fasting serum glucose [mmol/L])/22.5]; and quantitative insulin sensitivity check index (QUICKI) = [1/(log(fasting serum glucose [mg/dL]) + log(fasting serum insulin [µU/mL]))] (22). Serum IL-1β and FGF21 were measured with ELISA using human Quantikine HS ELISA kits (R&D Systems, MN, USA). Assay sensitivities (the minimum detectable levels) for serum IL-1β and FGF21 were 0.033 pg/mL and 8.69 pg/mL, respectively, determined by zero standard +2 standard deviation.

**Isolation and Culture of Peripheral Blood Mononuclear Cells**

The differentiation of macrophages from peripheral blood mononuclear cells (PBMCs) and cytokine assays were performed as described in our previous paper (23). Samples of whole blood were collected into acid citrate dextrose tubes. PBMCs were isolated from blood by density centrifugation (20 min at 1,600 × g (without brakes) at 18-20°C) using Ficoll Medium (Ficoll-Paque PLUS, GE Healthcare Life Science, Uppsala, Sweden). After removing the top layer of clear plasma, the PBMC-containing layer was aspirated, and the cells were washed with Dulbecco’s phosphate-buffered saline. Then the cells were re-suspended in RPMI-1640 supplemented with 1% penicillin, 1% streptomycin, and 10% fetal bovine serum. To generate human macrophages, cells were incubated at 1 × 10⁶ cells per ml in 24-well plates in RPMI medium plus 10% fetal bovine serum for 2 hours and then incubated with 20 ng/mL M-CSF for 3 days. After 3 days, the cells were then incubated with fresh RPMI medium plus 10% fetal bovine serum containing 10 ng/mL M-CSF and the medium was freshly replaced every 2 days (total 7 days) as previously described (23).

**Cell Stimulation and Cytokine Assays**

Human macrophages were incubated at 1 × 10⁶ cells per ml in 24-well plates in RPMI medium plus 10% fetal bovine serum with 0.1 µg/mL LPS (Sigma-Aldrich, St. Louis, MO, USA, L6529) for 4 hours (24). To stimulate the release of IL-1β, 2 mM adenosine triphosphate (ATP) (Sigma-Aldrich) or 0.2 mM palmitate (Sigma-Aldrich, P9767) was added for the last 1 or 12 hours of incubation, respectively. In addition, to evaluate the direct inhibitory or stimulating effects on NLRP3 inflammasome activation, BHB (1 mM), FGF21 (15, 30 or 100 nM), or baflomycin (10 nM) were pre-treated for 5 hours before adding LPS and ATP. Supernatants were collected, centrifuged to remove cells and debris, and stored at -80°C for later analysis. IL-1β and TNF-α was measured using ELISA (eBioscience, Waltham, MA, USA, human 88-7261-88 and human 88-7346-88, respectively). Results were normalized to cell number, as determined by the CyQuant cell proliferation assay (Invitrogen, Waltham, MA, USA). Experiments on the participants were repeated up to three times per sample. The examiner conducting these experiments remained blinded to the subject’s status throughout the study.

**Statistical Analysis**

All statistical analyses were performed using Prism 8.3.0 (GraphPad Software, San Diego, CA, USA). A normality test was performed for all continuous variables. The effects of isocaloric KD on metabolic parameters and secretion of cytokines from macrophages and the effects of BHB, FGF21, and baflomycin treatment on secretion of cytokines from macrophages were assessed by two-sided paired t-test or Wilcoxon signed rank test. One-way ANOVA using Tukey’s test or a two-tailed Student’s t-test with the Bonferroni method for adjusting P-values for the number of comparisons being made were used to examine differences between treatments in ex-vivo experiments. All P-values <0.05 were considered statistically significant.
RESULTS

Short-Term Isocaloric Ketogenic Diet Changes Metabolic Parameters

After a short-term KD, fasting serum BHB concentration was significantly elevated (median [interquartile range]: 0.03 [0.02 - 0.06] to 0.49 [0.36 - 1.25] mM, \( P < 0.0001 \)) (Figure 2A). These results accompanied decrease in fasting serum insulin levels (7.00 ± 3.87 to 4.61 ± 3.54 µU/mL, \( P < 0.01 \)) (Figure 2B). Interestingly, the isocaloric KD produced no change in fasting serum glucose levels (Figures 2C). Thus, isocaloric KD significantly increases insulin sensitivity reflected by improvement in QUICKI and HOMA-IR (Figures 2D, E), indicating improved glucose metabolism (25, 26). Fasting serum levels of FFA and total cholesterol were increased after a 3-day isocaloric KD (504.6 ± 288.3 to 998.4 ± 411.1 µEq/L, \( P = 0.002 \); and 185.6 ± 31.8 to 208.4 ± 27.9 mg/dL, \( P < 0.001 \), respectively) (Figures 2F, G). Subjects lost body weight (mean ± standard deviation: -1.87 ± 0.74 kg, \( P < 0.0001 \), Figure 2H).

Short-Term Isocaloric Ketogenic Diet Suppresses Inflammasome Activation

To evaluate the effects of short-term ketogenic diet on inflammasome activation, a primary endpoint of our study, we compared and observed that IL-1β secretion in response to ATP or palmitate stimulation in human macrophages decreased significantly after KD (2,532 [1,882 - 5,645] to 1,719 [840 - 3,883] pg/mL, \( P < 0.001 \); and 1,637 [1,047 - 2,659] to 1,045 [470 - 1,441] pg/mL, \( P < 0.001 \), respectively) (Figures 3A, B). Furthermore, serum IL-1β was measured by ELISA and was found to have decreased significantly after the isocaloric KD regimen (0.2407 ± 0.0297 to 0.1423 ± 0.019 pg/mL, \( P < 0.01 \)) (Figure 3C). In terms of secondary endpoint, secretion of TNF-α

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**FIGURE 2** | Effects of isocaloric ketogenic diet on metabolic parameters. Fasting serum levels of BHB (A), insulin (B), glucose (C), QUICKI (D), HOMA-IR (E), FFA (F), total cholesterol (G) from baseline to day 3 of isocaloric KD (H). Body weight changes from baseline to day 3 of isocaloric KD. Two-sided paired t-test or Wilcoxon signed rank test; \*\( P < 0.05 \), \**\( P < 0.01 \), \***\( P < 0.001 \), and \****\( P < 0.0001 \) vs. baseline. BHB, β-hydroxybutyrate; FFA, free fatty acid; KD, ketogenic diet; QUICKI, quantitative insulin sensitivity check index; HOMA-IR, homeostatic model assessment of insulin resistance; N.S., non-significant.
in response to ATP or palmitate stimulation also decreased significantly after isocaloric KD (76 [57 - 212] to 64 [43 - 160] pg/mL, P = 0.02; and 65 [38 - 139] to 58 [24 - 93] pg/mL, P = 0.004, respectively) (Figures 3D, E).

**Short-Term Isocaloric Ketogenic Diet Suppresses Inflammasome Activation Mediated by FGF21**

To investigate whether FGF21 mediates the KD effect on ameliorating inflammasome activation, we measured serum FGF21 levels and found it increased after KD (mean ± standard error of the mean, 108.25 ± 34.1 to 167.28 ± 28.47 pg/mL, P<0.01) (Figure 4A). To further assess the direct effect of FGF21 on human macrophages, we treated FGF21 prior to assessing inflammasome activation with LPS and ATP. IL-1β secretion was significantly reduced in a dose dependent manner (Figure 4B). Both FGF21 and BHB further reduced IL-1β secretion levels compared to either BHB or FGF21 alone (Figure 4C). In addition, FGF21 is known to augment autophagy in islets (27). We further tested if higher FGF21 levels with isocaloric KD was also related to autophagy activation and found that the inhibitory effect of FGF21 on IL-1β secretion was blunted with bafilomycin treatment, which blocked autophagy flux (Figure 4D).

**DISCUSSION**

In this short-term isocaloric KD study, we showed that insulin sensitivity improved with low serum insulin levels, although there was no difference in blood glucose levels. The regimen reduced body weight and increased mobilization of body fat in healthy subjects. High BHB and low IL-1β levels were observed after isocaloric KD. Reduced IL-1β levels were also detected along with increased FGF21 levels in serum. In a test of the direct effect of FGF21 on macrophages, FGF21 was seen to reduce macrophage IL-1β secretion, and its inhibitory effect was blunted by an autophagy inhibitor (Figure 5).

It is important to note that there were no changes in glucose levels but insulin levels were low, suggesting that KD improved insulin sensitivity in a short period of time. BHB is involved in the regulation of glucose homeostasis. After 21 days of KD in diabetes patients, glucose and insulin levels significantly were reduced with high BHB levels in serum (28). A very low carbohydrate diet for 8 weeks in men and women with obesity improved HOMA-IR (29).

A recognized effect of KD is reduction in body weight (30, 31), which may explain most of its metabolic benefits. However, most studies were of long duration and investigated prolonged impact, making it difficult to differentiate between KD per se and weight reducing effect. We demonstrated that, even in a short period of time without reducing calorie intake or a large decrement in body weight, isocaloric KD can mobilize fat and change inflammasome activity in immune cells, a potential advantage for improving metabolic regulation (32).

Previously, Vandanaansgar et al. reported that NLRP3 and IL-1β are associated with insulin resistance in mice with obesity (33). In that study, insulin sensitivity improved in mice that were NLRP3 deficient. BHB deactivates NLRP3 and resolves inflammatory diseases (14, 15). In our study, inflammasome activation was significantly suppressed in ATP/LPS-induced...
human macrophages with isocaloric KD as well as with direct treatment of BHB. In line with these findings, we have previously reported that SGLT2 inhibitor, a glucose lowering drug, regulates NLRP3 inflammasome in macrophages by increasing serum BHB levels and lowering insulin levels in patients with type 2 diabetes mellitus (23). Along with the high levels of BHB after isocaloric KD, we also detected an increase in serum FGF21. FGF21 is a hormone widely expressed in multiple organs. It was first identified in the liver (34, 35). Hepatic FGF21 mRNA expression levels were induced in mice that were fed a KD for 30 days (36). Serum FGF21 levels increased up to four-fold with prolonged fasting in

![Figure 4](image-url)  
**FIGURE 4** | Effects of FGF21 as a mediator of ketogenic diet on NLRP3 inflammasome signaling activation in human macrophages. Changes in serum FGF21 levels from baseline to day 3 of isocaloric KD (A); ELISA assay measurement of IL-1β secretion from macrophages primed with 0.1 µg/ml LPS and stimulated with 2 mM ATP in the presence of various dose of FGF21 (15, 30, 100 nM) for 24 h (B); BHB (1 mM), FGF21 (100 nM), or both (C), bafilomycin (10 nM), FGF21 (30 nM), or both (D). Data are represented as mean ± SEM. One-way analysis of variance (ANOVA) and two-sided paired t-test or Wilcoxon signed rank test; *P<0.05, **P<0.01, ***P<0.001, and ****P<0.0001. ATP, adenosine triphosphate; LPS, lipopolysaccharide; FGF21, fibroblast growth factor 21; N.S., non-significant; KD, ketogenic diet; IL-1β, interleukin-1β; BHB, β-hydroxybutyrate.

![Figure 5](image-url)  
**FIGURE 5** | The summary of effects of the isocaloric ketogenic diet.
healthy subjects (25, 37). Exogenous FGF21 has been shown to reduce body weight gain and fat content in diet-induced obese (DIO) or ob/ob genetically obese mouse models. Insulin levels were significantly reduced with FGF21 treatment in a dose dependent manner in the DIO mouse model, indicating an improvement in insulin sensitivity (38). In addition, FGF21 controls triglyceride levels in rodents (39) and in humans (40).

In type 2 diabetes mellitus, NLRP-IL-1β Inflammasome is a key factor in the development of insulin resistance (41). Since FGF21 agonists suppress the inflammasome after reduction of sequential inflammatory processes (42, 43), reduced serum IL-1β levels may be affected by not only BHB activity but also by FGF21 activity. Among human FGF21 receptors, FGF21 receptor 1 and β-klotho are expressed in multiple tissues, including PBMCs ([https://www.proteinatlas.org/ENSG00000134962-KLB/blood, https://www.proteinatlas.org/ENSG00000077782-FGFR1/blood]). After direct treatment of FGF21 on human macrophages, we observed reduced cytokine secretion related to the inflammasome. When patients with obesity and type 2 diabetes mellitus lose body weight, FGF21 levels also decrease (44). This may be due to improvement in FGF21 signaling sensitivity similar to the effect on insulin resistance, which is ameliorated by loss of body weight. Taken together, our findings suggest that isocaloric KD not only increases FGF21 levels but also improves FGF21 signaling pathways thereby lowering inflammatory activation.

Consistent with these findings, a recent study reported that serum FGF21 levels can predict NAFLD improvement in patients with a very low caloric KD (32); fatty liver is also improved with increased tissue FGF21 mRNA expression levels in intermittent KD [abstract: diabetes 2020 Jun; 69 (Supplement 1):1957-P]. FGF21 is also known to reduce hepatic glucose production (45); however, we did not observe any changes in glucose levels. This may possibly be due to the acute response levels of FGF21 to the isocaloric KD regimen. FGF21 global levels may be affected by not only BHB activity but also by FGF21 levels with a KD also mimics the effects of exercise (49). Therefore, our study suggests that isocaloric KD has positive effects in healthy subjects, even in the short term.

Multiple studies have explained that FGF21 augments and exerts its effects through autophagy. For example, fasting induced exogenous FGF21 significantly improved obese phenotypes of mice with increased autophagy-related proteins and autophagosomes (27, 50–52). However, the role of autophagy in isocaloric KD, especially in macrophages of healthy individuals, has not been thoroughly studied. The current study showed that the suppressive effect of FGF21 on inflammasome activation in human macrophages was at least partially mediated through the autophagy signaling pathway.

Since autophagy controls oxidative mediated inflammation and rescues macrophages from inflammatory processes (53), the effect of the isocaloric KD–FGF21–autophagy axis on macrophages provides a clear health benefit.

Our results suggest that low caloric intake or fasting was not necessary to induce ketosis. Isocaloric KD for 3 days is sufficient to mobilize fat depot with incremental rise in BHB levels and to improve glucose metabolism by increasing insulin sensitivity. This diet regimen suppresses inflammasome activity in macrophages by increasing FGF21 levels and inducing autophagy, resulting in promising health benefits related to improved metabolic and inflammatory status.

**DATA AVAILABILITY STATEMENT**

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

**ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by Institutional Review Board at Severance Hospital (4-2016-0795). The patients/participants provided their written informed consent to participate in this study.

**AUTHOR CONTRIBUTIONS**

EK, SK and S-GL wrote the manuscript, analyzed data, and performed the statistical analysis; SHK, JK, EC, and WC conducted the experiments and contributed to acquisition of data; JJ, J-HK, B-WL, ESK, B-SC, M-SL, J-WY, and JC provided critical review, advice, and consultation throughout. Y-HL contributed to the conception and design of the study, the interpretation of data, and the drafting of the manuscript. Y-HL is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis. All authors contributed to the article and approved the submitted version.

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REFERENCES

1. Schnurr TM, Jakupovic H, Carrasquilla GD, Angquist L, Grarup N, Sorensen TIA, et al. Obesity, Unfavorable Lifestyle and Genetic Risk of Type 2 Diabetes: A Case-Cohort Study. *Diabetologia* (2020) 63(7):1324–32. doi: 10.1007/s00125-020-05140-5

2. O’Neill BJ. Effect of Low-Carbohydrate Diets on Cardiometabolic Risk, Insulin Resistance, and Metabolic Syndrome. *Curr Opin Endocrinol Diabetes Obes* (2020) 27(5):301–7. doi: 10.1097/MED.0000000000000569

3. Yuan X, Wang J, Yang S, Gao M, Cao L, Li X, et al. Effect of the Ketogenic Diet on Glycemic Control, Insulin Resistance, and Lipid Metabolism in Patients With T2dm: A Systematic Review and Meta-Analysis. *Nutr Diabetes* (2020) 10(1):38. doi: 10.1214/jccta.2020.00142-z

4. Watanabe M, Tozzi R, Rasi R, Tuccinardi D, Mariani S, Basciani S, et al. Ketogenic Diet and Lifestyle Recommendations Versus a Plate Method Diet in Overweight Individuals With Type 2 Diabetes: A Randomized Controlled Trial. *J Med Internet Res* (2019) 17(9):e36. doi: 10.2196/2013.09.5010

5. Deberes E, Maragnes P, Penniello-Valette MJ, Allouche S, Joubert M. Ketogenic Diet and NLRP3 Inflammasome Activity Via Balancing Mitochondrial Dynamics and Inhibiting Apoptosis in Type 2 Diabetic Mice. *Aging Dis* (2020) 11(2):229–40. doi: 10.15343/ade.2019.0510

6. Abbasi J. Interest in the Ketogenic Diet Grows for Weight Loss and Type 2 Diabetes. *JAMA* (2018) 319(3):215–7. doi: 10.1001/jama.2017.20639

7. Davis JJ, Fournakis N, Ellison J. Ketogenic Diet for the Treatment and Prevention of Dementia: A Review. *J Geriatr Psychiatry Neurol* (2011) 34 (1):3–10. doi: 10.1177/0891988710391985

8. Pinney SD, Bistrian BR, Evans WJ, Gervino E, Blackburn GL. The Human Metabolic Response to Chronic Ketosis Without Caloric Restriction: Kinetics and Signaling During LPS-Induced Inflammation in Humans. *J Clin Invest* (2019) 2019(12):4592–60. doi: 10.1172/JCI83260

9. Kim et al. Ketogenic Diet and NLRP3 Inflammasome Deactivates Neutrophil Nlrp3 Inflammases to Relieve Gout Flares. *Cell Rep* (2017) 18(9):2077–87. doi: 10.1016/j.celrep.2017.02.004

10. Yoon YH, Nguyen KY, Grant RW, Goldberg EL, Bodogai M, Kim D, et al. The Ketone Metabolite Beta-Hydroxybutyrate Blocks Nlrp3 Inflammases-Mediated Inflammatory Disease. *Nat Med* (2015) 21(3):263–9. doi: 10.1038/nn.3804

11. Blad CC, Tang C, Offermanns S. G Protein-Coupled Receptors for Energy Metabolites as New Therapeutic Targets. *Nat Rev Drug Discov* (2012) 11(8):603–19. doi: 10.1038/nrd3777

12. Chen Y, Ouyang X, Hoque R, Garcia-Martinez I, Yousaf MN, Tonack S, et al. Beta-Hydroxybutyrate Protects From Alcohol-Induced Liver Injury Via a Hcar2-Camp Dependent Pathway. *J Hepatol* (2018) 69(3):687–96. doi: 10.1016/j.jhep.2018.04.004

13. Li H, Wu G, Fang Q, Zhang M, Hui X, Sheng B, et al. Fibroblast Growth Factor 21 Increases Insulin Sensitivity Through Specific Expansion of Subcutaneous Fat. *Nat Commun* (2018) 9(1):272. doi: 10.1038/s41467-017-02677-9

14. Kim JC, Verdin E. Ketone Bodies as Signaling Metabolites. *Trends Endocrinol Metab* (2014) 25(1):42–52. doi: 10.1016/j.tem.2013.09.002

15. Ministry of HW. Dietary Reference Intakes for Koreans 2015. In: Ministry of Health & Welfare. Sejong: The Korean Nutrition Society (2015).

16. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis Model Assessment: Insulin Resistance and Beta-Cell Function From Fasting Plasma Glucose and Insulin Concentrations in Man. *Diabetologia* (1985) 28(7):412–9. doi: 10.1007/BF00280883

17. Katz A, Nambi SS, Mather K, Baron AD, Follmann DA, Sullivan G, et al. Quantitative Insulin Sensitivity Check Index: A Simple, Accurate Method for Assessing Insulin Sensitivity in Humans. *J Clin Endocrinol Metab* (2000) 85(7):2402–10. doi: 10.1210/jcem.85.7.6661

18. Kim SR, Lee SG, Kim SH, Kim JH, Choi E, Cho W, et al. Sglt2 Inhibition Modulates Nlrp3 Inflammases Activity Via Ketones and Insulin in Diabetes With Cardiovascular Disease. *Nat Commun* (2020) 11(1):2127. doi: 10.1038/s41467-020-15983-6

19. Traba I, Kwarteng-Saw M, Okoli TC, Li J, Huffstutler RD, Bray A, et al. Fasting and Refeeding Differentially Regulate Nlrp3 Inflammases Activation in Human Subjects. *J Clin Invest* (2015) 125(12):4592–600. doi: 10.1172/JCI83260

20. Ministry of HW. Dietary Reference Intakes for Koreans 2015. In: Ministry of Health & Welfare. Sejong: The Korean Nutrition Society (2015).

21. Saslow LR, Mason AE, Kim S, Goldman V, Ploutz-Snyder R, Bayandorian H, et al. Online Intervention Comparing a Very Low-Carbohydrate Ketogenic Diet and Lifestyle Recommendations Versus a Plate Method Diet in Overweight Individuals With Type 2 Diabetes: A Randomized Controlled Trial. *J Med Internet Res* (2019) 17(9):e26. doi: 10.2196/2013.09.5010

22. Balancing Mitochondrial Dynamics and Signaling During LPS-Induced Inflammation in Humans. *J Clin Invest* (2019) 2019(12):4592–60. doi: 10.1172/JCI83260

23. Galman C, Lundansen T, Kharitonenkov A, Bina HA, Eriksson M, Häfström I, et al. The Circulating Metabolic Regulator FgF21 Is Induced by Prolonged Fasting and Pparalpha Activation in Man. *Cell Metab* (2008) 8(2):169–74. doi: 10.1016/j.cmet.2008.06.014

24. Sharman MJ, Kraemer WJ, Love DM, Aver NG, Gomez AL, Scheett TP, et al. A Ketogenic Diet Favorably Affects Serum Biomarkers for Cardiovascular Disease in Normal-Weight Men. *J Nutr Metab* (2012) 2012(7):1879–85. doi: 10.1093/jnut/1j2.7.1879

25. Cheng STW, Li SYT, Leung PS. Fibroblast Growth Factor 21 Stimulates Pancreatic Islet Autoaphagy Via Inhibition of Ampk-Mtor Signaling. *Int J Mol Sci* (2019) 20(10):2517. doi: 10.3390/ijms20102517

26. Boden G, Sargrad K, Homko C, Mozzi M, Stein TP. Effect of a Low-Carbohydrate Diet on Appetite, Blood Glucose Levels, and Insulin Resistance in Obese Patients With Type 2 Diabetes. *Ann Intern Med* (2005) 142(6):403–11. doi: 10.7326/0003-4819-142-6-200503150-00006

27. Goss AM, Gower B, Soleymani T, Stewart M, Pendergrass M, Lockhart M, et al. Effects of Weight Loss During a Very Low Carbohydrate Diet on Specific Adipose Tissue Deposits and Insulin Sensitivity in Older Adults With Obesity: A Randomized Clinical Trial. *Nutr Metab (Lond)* (2020) 17:64. doi: 10.1186/s12986-020-00481-9

28. Kinzig KP, Honors MA, Hargrave SL. Insulin Sensitivity and Glucose Tolerance Are Altered by Maintenance on a Ketogenic Diet. *Endocrinology* (2010) 151(7):3105–14. doi: 10.1210/en.2010-0175

29. Kennedy AR, Pissios P, Otu H, Roberson R, Xue B, Asakura K, et al. A High-Fat, Ketogenic Diet Induces a Unique Metabolic State in Mice. *Am J Physiol Endocrinol Metab* (2007) 292(6):E1724–39. doi: 10.1152/ajpendo.00717.2006

30. Watanabe M, Rui R, Camajani E, Contini S, Persichetti A, Tuccinardi D, et al. Baseline Homa Ir and Circulating FgF21 Levels Predict Nafld Improvement in Patients Undergoing a Low Carbohydrate Dietary Intervention for Weight
Lose: A Prospective Observational Pilot Study. *Nutrients* (2020) 12(7):2141. doi: 10.3390/nu12072141

35. Verpulzen L, Ferrari F, Bina-H, Ravussin A, Galgani JE, Stadler K, Mynatt RL, et al. The Nlrp3 Inflamasome Instigates Obesity-Induced Inflammation and Insulin Resistance. *Nat Med* (2011) 17(2):179–88. doi: 10.1038/nm.2279

36. Nishimura T, Nakatake Y, Konishi M, Itoh N. Identification of a Novel Fgf, Fgf-21, Preferentially Expressed in the Liver. *Biochim Biophys Acta* (2000) 1492(1):203–6. doi: 10.1016/s0167-4781(00)00067-1

37. Fazeli PK, Lun M, Kim SM, Bredella MA, Wright S, Zhang Y, et al. Fgf21 and Fibrinolysis: A Link Between Fibroblast Growth Factor 21 and Hemostasis. *Arterioscler Thromb Vasc Biol* (2010) 30(10):2039–46. doi: 10.1161/ATVBAHA.110.207588

38. Schlein C, Talukdar S, Heine M, Fischer AW, Krott LM, Nilsson SK, et al. Hepatic Fibroblast Growth Factor 21 Is Regulated by Pparalpha and Is a Key Mediator of Hepatic Lipid Metabolism in Ketotic States. *Cell Metab* (2007) 5(6):426–37. doi: 10.1016/j.cmet.2007.05.002

39. Fazeli PK, Lun M, Kim SM, Breedella MA, Wright S, Zhang Y, et al. Fgf21 and the Late Adaptive Response to Starvation in Humans. *J Clin Invest* (2015) 125(12):4601–11. doi: 10.1172/JCI83349

40. Coskun T, Bina HA, Schneider MA, Dunbar JD, Hu CC, Chen Y, et al. Fibroblast Growth Factor 21 Corrects Obesity in Mice. *Endocrinology* (2008) 149(12):6018–27. doi: 10.1210/en.2008-0816

41. Khartitonova A, Shiyanova TL, Koester A, Ford AM, Micanovic R, Galbeath EJ, et al. Fgf-21 as a Novel Metabolic Regulator. *J Clin Invest* (2005) 115(6):1627–35. doi: 10.1172/JCI23606

42. Schlein C, Talukdar S, Heine M, Fischer AW, Krott LM, Nilsson SK, et al. Fgf21 Lowers Plasma Triglycerides by Accelerating Lipoprotein Catabolism in White and Brown Adipose Tissues. *Cell Metab* (2016) 23(3):441–53. doi: 10.1016/j.cmet.2016.01.006

43. Stienstra R, van Diepen JA, Tack CJ, Zaki MH, van de Veerdonk FL, Perera D, et al. Inflamasome Is a Central Player in the Induction of Obesity and Insulin Resistance. *Proc Natl Acad Sci USA* (2011) 108(37):15324–9. doi: 10.1073/pnas.1102681108

44. Liu MH. Fgf-21 Alleviates Diabetes-Associated Vascular Complications: Inhibiting Nf-kappaB/Nlrp3 Inflamasome-Mediated Inflammation? *Int J Cardio* (2015) 185:320–1. doi: 10.1016/j.ijcard.2015.03.165

45. Wei W, Li XX, Xu M. Inhibition of Vascular Neointima Hyperplasia by Fgf21 Associated With Fgf1/Syk/Nlrp3 Inflamasome Pathway in Diabetic Mice. *Atherosclerosis* (2019) 289:132–42. doi: 10.1016/j.atherosclerosis.2019.08.017

46. Christodoulides C, Dyson P, Sprecher D, Tsintzas K, Karpe F. Circulating Fibroblast Growth Factor 21 Is Induced by Peroxisome Proliferator-Activated Receptor Agonists But Not Ketosis in Man. *J Clin Endocrinol Metab* (2009) 94(9):3594–601. doi: 10.1210/jc.2009-0111

47. Berglund ED, Li CY, Bina HA, Lynn SE, Michael MD, Shanafelt AB, et al. Fibroblast Growth Factor 21 Controls Glycemia Via Regulation of Hepatic Glucose Flux and Insulin Sensitivity. *Endocrinology* (2009) 150(9):4084–93. doi: 10.1210/en.2009-0221

48. Yilmaz Y, Eren F, Yonal O, Kurt R, Aktas B, Celikel CA, et al. Increased Serum Fgf21 Levels in Patients With Nonalcoholic Fatty Liver Disease. *Eur J Clin Invest* (2010) 40(10):887–92. doi: 10.1111/j.1365-2362.2010.02338.x

49. Alhubaishi AA, Abdel-Kader MI. Phyllosphere and Phylloplane Fungi of Qat in Sana’a, Yemen Arab Republic. *J Basic Microbiol* (1991) 31(2):83–9. doi: 10.1002/jobm.3620310202

50. Inagaki T, Dutchak P, Zhao G, Ding X, Gautron L, Parameswara V, et al. Endocrine Regulation of the Fasting Response by Pparalpha-Mediated Induction of Fibroblast Growth Factor 21. *Cell Metab* (2007) 5(6):415–25. doi: 10.1016/j.cmet.2007.05.003

51. Kim KH, Kim SH, Min YK, Yang HM, Lee JB, Lee MS. Acute Exercise Induces Fgf21 Expression in Mice and in Healthy Humans. *PloS One* (2013) 8(5):e63517. doi: 10.1371/journal.pone.0063517

52. Byun S, Seok S, Kim YC, Zhang Y, Yau P, Iwamori N, et al. Fasting-Induced Fgf21 Signaling Activates Hepatic Autophagy and Lipid Degradation Via Jmd3 Histone Demethylase. *Nat Commun* (2020) 11(1):807. doi: 10.1038/s41467-020-14384-z

53. Zhu S, Wu Y, Ye X, Ma L, Qi J, Yu D, et al. Fgf21 Ameliorates Nonalcoholic Fatty Liver Disease by Inducing Autophagy. *Mol Cell Biochem* (2016) 420(1-2):107–19. doi: 10.1007/s11010-016-2774-2

54. Leung PS. Fgf21 Activation-Mediated Islet Autophagy in Type 2 Diabetes With Pharmacotherapeutic Potential. *Future Med Chem* (2019) 11(7):641–3. doi: 10.4155/fmc-2018-0491

55. Chatterjee T, Pattanayak R, Uch A, Chowdhury S, Bhattacharyya M. Autophagy Protects Peripheral Blood Mononuclear Cells Against Inflammation, Oxidative and Nitrosative Stress in Diabetic Dyslipidemia. *Free Radic Biol Med* (2019) 143:309–23. doi: 10.1016/j.freeradbiomed.2019.07.034

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