Single gene enables plant pathogenic *Pectobacterium* to overcome host-specific chemical defence

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**Abstract**
Plants of the Brassicales order, including *Arabidopsis* and many common vegetables, produce toxic isothiocyanates to defend themselves against pathogens. Despite this defence, plant pathogenic microorganisms like *Pectobacterium* cause large yield losses in fields and during storage of crops. The bacterial gene saxA was previously found to encode isothiocyanate hydrolase that degrades isothiocyanates in vitro. Here we demonstrate in planta that saxA is a virulence factor that can overcome the chemical defence system of Brassicales plants. Analysis of the distribution of saxA genes in *Pectobacterium* suggests that saxA from three different phylogenetic origins are present within this genus. Deletion of saxA genes representing two of the most common classes from *P. odoriferum* and *P. versatile* resulted in significantly reduced virulence on *Arabidopsis thaliana* and *Brassica oleracea*. Furthermore, expressing saxA from a plasmid in a potato-specific *P. parmentieri* strain that does not naturally harbour this gene significantly increased the ability of the strain to macerate *Arabidopsis*. These findings suggest that a single gene may have a significant role in defining the host range of a plant pathogen.

**KEYWORDS**
*Arabidopsis*, isothiocyanate hydrolase, host-specific virulence, *Pectobacterium*, soft rot

1 | **INTRODUCTION**

With a growing human population, worldwide food security has become an international priority according to the United Nations’ Sustainable Development Goals. Phytopathogens infect crops on fields and in storage, and lead to the spoilage of considerable amounts of food. Soft rot is an important disease leading to crop losses and is caused by phytopathogens affiliated to the genera *Pectobacterium* and *Dickeya* (Czajkowski et al., 2015). Knowledge of the phylogeny and the genetic basis for the pathogenicity of *Pectobacterium* was recently greatly expanded, as 84 *Pectobacterium* genomes were screened for the presence of 159 genes that are known virulence factors (Li et al., 2018).

The characteristic virulence factor of the *Pectobacterium* and *Dickeya* genera are the plant cell wall-degrading enzymes (PCWDE), which are responsible for the rotting symptoms they may cause on their host. Other virulence factors include pilus assembly proteins, proteases, lipopolysaccharide biosynthesis proteins, regulators, biofilm formation, motility, and a number of other factors (Nykryi et al., 2012; Lee et al., 2013; Li et al., 2018). Many of these virulence factors are common among *Pectobacterium* strains as they appear indispensable in ensuring disease development. However, inconsistent
distribution of single genes or gene clusters suggests that some virulence factors only contribute to virulence depending on the context of the infection (Li et al., 2018).

Certain plant secondary metabolites are characteristic for specific groups of plants and are mostly known for their antiherbivorous properties. Much less is known about how secondary metabolites affect plant pathogenic bacteria and about ways in which these pathogens may overcome the host-specific defences. It was often recently shown that caffeine has antibacterial properties against multiple plant pathogenic bacteria, including Pectobacterium, Pseudomonas, and Dickeya (Sledz et al., 2015). Bacterial resistance to such plant antimicrobials can be considered a context-dependent virulence factor. One example of a context-dependent virulence factor in Pectobacterium is tolC, a gene that decreases the susceptibility of Pectobacterium brasiliense PCC21 to the antibacterial plant chemicals berberine, rhein, and genistein (Lee et al., 2013).

Most research about Pectobacterium and Dickeya has focused on potato pathogenesis as it is a staple food with high economic relevance (Motyka et al., 2017). Certain Pectobacterium spp., however, have a wide host range or are specific pathogens to other crops, which is also reflected in the high genomic variability of these bacteria (Li et al., 2018; Zoleadowska et al., 2018). Here we approached the identification of a novel virulence factor bottom-up by directly targeting a gene with a recently elucidated function that was hypothesized to aid specifically in the colonization of plants that produce the secondary metabolite isothiocyanate (ITC). Among the host plants of pectobacteria, those of the Brassicaceae family are some of the most produced vegetables worldwide (FAO, 2012). Plants of this family use a unique form of chemical defence to protect against herbivory and disease. Mustard oil glucosides (or glucosinolates, GSN) and the enzyme myrosinase are stored separately within plant compartments and toxic ITCs are produced when these two come into contact after tissue damage by herbivory or lesion caused during harvesting (Zhu et al., 2013). Coincidentally, soft rot commony establishes in crops after initial tissue damage. Phytopathogenic bacteria have been shown to be particularly susceptible to allylisothiocyanate (A-ITC), benzylisothiocyanate (B-ITC), 2-phenylethyliothiocyanate (2PE-ITC), and sulforaphane (SFN) in vitro (Aires et al., 2009). In nature, the composition of different ITCs that are produced in a plant varies greatly by host species, age, and a variety of other factors (Ván Dam et al., 2009). The model plant Arabidopsis thaliana, also a member of the Brassicaceae family, produces a range of mostly aliphatic ITCs (Hogge et al., 1988). Insects feeding on mutant Arabidopsis with abnormally low levels of GSN grow to larger sizes and do more damage to the leaves than those feeding on Arabidopsis with natural GSN concentrations (Beekwilder et al., 2008). A 2001 study determined that Pectobacterium carotovorum (then Erwinia carotovora) and Pseudomonas syringae were equally infectious to Arabidopsis gsm1-1 mutants that contain fewer GSNs as they were to wild-type plants (Tierens et al., 2001). Only the fungus Fusarium oxysporum was found to be significantly more aggressive on A. thaliana gsm1-1 than on wild-type plants.

Recently we found that the bacterial gene saxA encodes ITC hydrolase (ITCase) that is capable of hydrolysing ITCs in vitro (van den Bosch et al., 2018). The protein catalytic site activates one water molecule to subsequently hydrolyse the ITC, resulting in the release of the ITC’s corresponding amine and a molecule of carbonyl sulfide (Equation 1).

\[
\text{R} - \text{N} = \text{C} = \text{S} + \text{H}_2\text{O} \xrightarrow{\text{ITCase}} \text{R} - \text{NH}_2 + \text{O} = \text{C} = \text{S}
\]

(1)

The saxA gene is widely distributed among the Enterobacteriales and the saxA gene family could be categorized into five phylogenetic clusters. The saxA gene was originally identified in plant pathogenic P. syringae pv. tomato DC3000, where it was shown to be one of six genes increasing survival on Arabidopsis extracts (Fan et al., 2011). The lifestyle of plant pathogenic P. syringae differs markedly from that of soft rot pectobacteria in that it does not rely on tissue maceration in order to utilize nutrients from dead host cells (Davidsson et al., 2013). It is currently unknown whether ITCs are involved in defence against the latent phase of phytopathogenic infections, but it is clear that tissue damage, which the PCWDE arsenal of pectobacteria is known to cause, activates the production of ITCs.

In this study we investigated the effect of deleting the saxA genes in two Pectobacterium soft rot strains (P. odoriferum NCPPB3841 and P. versatilis SCC1) on their ability to infect Arabidopsis. A third strain, P. polariis NCPPB3395, also containing a genomic saxA gene, could not infect Arabidopsis leaves and was therefore tested in medium with Arabidopsis leaf extract. A fourth Pectobacterium strain, P. parmentieri SCC3193, is a known potato pathogen that naturally lacks a genomic saxA gene and we assessed how its virulence was affected by appending it with a saxA gene. Based on the known function of SaxA in vitro, we hypothesized that deleting the saxA gene in Pectobacterium strains results in reduced virulence on plants that use the GSN–myrosinase system as a defence, whereas adding it to a strain that naturally lacks saxA would achieve the opposite. We also compared the distribution of the saxA gene in most known Pectobacterium whole genomes to see whether presence of the saxA gene is restricted to strains isolated from Brassica plants. Besides the use of Arabidopsis and its mutants as model plants, we also conducted infection experiments with cabbages as a representative of common vegetable crops. Taken together, our results indicate that the deletion of saxA in pectobacteria can lead to greatly reduced virulence whereas the introduction of saxA to a potato-infecting Pectobacterium can transform it to successfully macerate Brassica plants. We therefore conclude that a single gene can enable phytopathogenic pectobacteria to overcome the chemical defence system of the host.

2 | RESULTS

2.1 | Deletion of single-copy saxA in P. odoriferum NCPPB3841 and P. versatilis SCC1 significantly reduces virulence on A. thaliana and cabbage leaves

We observed previously that Pectobacterium spp. isolated from Brassica plants contain one or several genes encoding SaxA, an ITC
hydrolase that may mitigate the plant defence of *Brassica* plants (van den Bosch et al., 2018). In order to assess whether *saxA* was required for infection in planta and would therefore be a new virulence factor, we deleted the *saxA* gene in the genomes of *P. odoriferum* (*Po*) NCPPB3841 and *P. versatil* (Pv) SCC1. The *saxA* genes of *Po* NCPPB3841 and *Pv* SCC1 are phylogenetically distinct (44.9% amino acid identity) and cluster in different subfamilies (CL2 and CL5, respectively, Figure 1).

In order to assess the virulence, *A. thaliana* Col-0 plant leaves were infected with *Po* NCPPB3841 wild-type and *saxA* deletion mutant (Δ*saxA*) (Figure 2a,b) or with *Pv* SCC1 wild-type and Δ*saxA* (Figure 2d,e), and incubated for 24 hr before the area of maceration was measured. The macerated area of plants infected with Δ*saxA* was significantly smaller than the area of maceration in plants infected with the wild-type (*Po* = 3.0e−06, 7.4e−07, 7.4e−07, and *Pv* = 7.4e−07, 7.4e−07, 7.4e−07, respectively, for three independent experiments, Wilcoxon rank sum test). In order to verify that the reduction of virulence in the mutant is indeed due to the loss of *saxA*, a complementation of the Δ*saxA* strains was performed (Figure 2b,e). Transforming *Po* NCPPB3841 Δ*saxA* with a plasmid containing the deleted gene (pMW119-NCPPB3841_saxA) resulted in significantly more maceration than transformation with the empty plasmid (*p* = 3.0e−06, 7.4−07, and 7.4e−07, respectively, for three independent experiments, Wilcoxon rank sum test). Furthermore, a plasmid containing the *saxA* gene from *Pv* SCC1 was also able to restore the virulence of *Po* NCPPB3841 Δ*saxA* (*p* = 1.0e−04, 7.4e−07, and 7.4e−07, respectively, for three independent experiments, Wilcoxon rank sum test). Similarly, complementing *Pv* SCC1 Δ*saxA* using either pMW119 with the *saxA* gene from SCC1 (*p* = 7.4e−07 for all three independent experiments, Wilcoxon rank sum test) or NCPPB3841 (p = 7.4e−07 for all three independent experiments, Wilcoxon rank sum test) restored the virulence phenotype (Figure 2e). In order to show that the effect of *saxA* on virulence is dependent on plant-produced ITCs, *A. thaliana* myb28 myb29 was used, which is impaired in aliphatic GSN production and therefore does not release the derived ITCs (Sønderby et al., 2007). Figure 2c,f shows that

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**FIGURE 1** Phylogenetic tree adapted from Li et al. (2018) with arbitrary branch lengths. The presence of a *saxA* gene in the strain genome is overlaid in branch colours. The colour of the strain identifier corresponds with isolation sources from either potato (*Solanum tuberosum*) or *Brassica* vegetables.
**FIGURE 2** Isothiocyanate hydrolase SaxA is required for full virulence of *Pectobacterium* on *Arabidopsis thaliana*. *Arabidopsis* leaves were inoculated locally with approximately $10^4$ cfu and maceration symptoms were documented 24 hr after inoculation. Representative images of symptoms on *Arabidopsis* Col-0 caused by *P. odoriferum* NCPPB3841 (a) or *P. versatile* SCC1 (d). Quantification of *Arabidopsis* Col-0 leaf maceration caused by *P. odoriferum* NCPPB3841 (b) or *P. versatile* SCC1 (e), the respective saxA mutants, and the saxA mutants complemented with two saxA genes of different origins. Quantification of *Arabidopsis* Col-0 leaf maceration caused by *P. odoriferum* NCPPB3841 (c) or *P. versatile* SCC1 (f) and the respective saxA mutants on wild-type and myb28 myb29 double mutants with impaired aliphatic glucosinolate accumulation. Three leaves per plant were inoculated and one dot represents the combined area of maceration per plant. Different symbols represent experimental replicates on different dates. For each of the three experiments, $N = 12$ in (b) and (e), and $N = 10$ in (c) and (f). Each individual experiment was analysed separately and asterisks indicating statistically significant differences according to the Wilcoxon rank sum test are shown for the largest $p$ value obtained (in black, ***$p < .001$ in all three experiments, ns: $p > .05$ in all three experiments) or for each experiment separately in cases where differing results were obtained (in grey, circles/squares/triangles; **$p < .01$, *$p < .05$, ns: $p > .05$)
Po NCPPB3841 ΔsaxA and P. SCC1 ΔsaxA were both significantly more virulent on myb28 myb29 plants than on wild-type Col-0 plants ($p_{Po} = 2.2e^{-05}$, 2.1e−04, 2.1e−04 and $p_{Po} = 1.1e^{-05}$, 1.1e−05, 2.2e−05, respectively, for three independent experiments, Wilcoxon rank sum test) whereas the wild-type strains macerated myb28 myb29 plants only slightly better than or equally well as wild-type plants ($p_{Po} = .17, .015, .80$ and $p_{Po} = .052, .0052, .11$, respectively, for three independent experiments, Wilcoxon rank sum test). The effect of saxA on virulence of Po NCPPB3841 and P. SCC1 was further assessed using discs of cabbage leaves (Brassica oleracea var. capitata) as a model for the common crop Brassicaceae. The disease progressed markedly differently between wild-type and ΔsaxA. Infection with ΔsaxA resulted in a significantly smaller area of maceration 48 hr after inoculation (Figure 3b, $p_{Po} = 1.5e^{-06}$, 2.0e−04, 7.4e−07 and $p_{Po} = 1.4e^{-05}$, 2.7e−04, 3.0e−06, respectively, for three independent experiments, Wilcoxon rank sum test). The macerations caused by ΔsaxA also seemed to turn dark and often even stopped spreading altogether (Figures 3a and S2a,b).

### 2.2 P. polaris NCPPB3395 contains two functionally redundant copies of saxA that are linked to survival on A. thaliana extract

P. polaris NCPPB3395 harbours two distinct copies of saxA in its genome, one of which is closely related to the saxA gene found in many P. carotovorum and P. odoriferum strains, including P. odoriferum NCPPB3841 (CL2), and the other which is closely related to the saxA commonly present in P. brasiliense, P. atrosepticum and P. versatile, including strain SCC1 (CL5) (van den Bosch et al., 2018). Three deletion strains of P. polaris NCPPB3395 were constructed: P. polaris ΔsaxA1, P. polaris ΔsaxA2, and P. polaris ΔsaxA1 + ΔsaxA2. P. polaris does not cause symptoms on A. thaliana, and trial assays on slices of turnip (Brassica rapa subsp. rapa) and slices of cabbage (B. oleracea var. capitata, Figure S2d) led us to conclude that P. polaris NCPPB3395 is not fully virulent on these three hosts and thus a possible in planta effect of saxA could not be tested in these systems. In order to better assess the role of saxA genes in P. polaris NCPPB3395 we conducted growth experiments of P. polaris NCPPB3395 wild type and three different deletion mutants with medium containing A. thaliana extract (Figure 4a). The single deletion mutants P. polaris ΔsaxA1 and P. polaris ΔsaxA2 did not show growth inhibition on medium containing A. thaliana Col-0 extract as compared to the wild-type whereas P. polaris ΔsaxA1 + ΔsaxA2 showed severely impaired growth. Medium containing myb28 myb29 extract devoid of aliphatic GSN/ITCs enabled all deletion strains to grow equally well as the wild-type. This suggests that the two copies of saxA in P. polaris are either functionally redundant in detoxifying a blend of naturally occurring ITCs in Arabidopsis or that the differential function of saxA genes within different phylogenetic clusters remains to be elucidated.

### 2.3 In trans expression of saxA enables potato pathogen P. parmentieri SCC3193 to cause maceration on Arabidopsis

P. parmentieri SCC3193 was isolated from diseased potato stem and does not naturally contain a saxA gene in its genome. A. thaliana Col-0 extract has been shown to be toxic to SCC3193 in a

![Figure 3](image)

**FIGURE 3** Isothiocyanate hydrolase SaxA is required for full virulence of Pectobacterium on cabbage. Brassica oleracea var. capitata leaves were inoculated locally with approximately $5 \times 10^6$ cfu of P. odoriferum NCPPB3841 or P. versatile SCC1 and the respective saxA mutants, and maceration symptoms were documented 48 hr after inoculation. Representative images of symptoms (a) and quantification of maceration (b). One dot represents one cabbage leaf disc. Different symbols represent experimental replicates on different dates. N = 12 for each experiment. Each individual experiment was analysed separately and asterisks indicating statistically significant differences according to Wilcoxon rank sum test are shown for the largest $p$ value obtained (**p < .001 in all three experiments).
myrosinase-dependent manner (Brader et al., 2001) and SCC3193 does not readily macerate Col-0 leaves. We observed that even an inoculum 100 times the one used with Po NCPPB3841 and Pv SCC1 rarely resulted in initiation of maceration. Ten out of 30 plants showed maceration on one out of five inoculated leaves after 24 hr. Furthermore, the maceration caused by wild-type SCC3193 never seemed to spread significantly (Figure S1). As a result of our previous experiments, we wondered whether transforming this strain with the previously constructed saxA-containing vector would impart it with the ability to tolerate ITC-rich Col-0 extract and macerate Col-0 leaves. In fact, the P. parmentieri SCC3193 with the saxA gene sourced from the genome of P. versatile SCC1 was able to grow on Col-0 extract, unlike the wild-type strain (Figure 4b). Furthermore, it caused maceration in all five inoculated leaves in 12 out of 30 tested plants after 24 hr. Thirteen plants showed maceration in four out of five leaves, four plants showed maceration in three out of five leaves and one plant showed maceration in two out of five inoculated leaves. The maceration was often able to spread to cover large parts of the leaf (Figures 4c and S1). These results indicate that saxA could increase the virulence of some Pectobacterium strains on non-host brassica plants. However, only a high inoculum led to maceration even when SCC3193 was amended with saxA, suggesting that there are also other factors contributing to the poor ability of SCC3193 to macerate Col-0. This conclusion is consistent with the result of cabbage inoculations where neither wild-type SCC3193 nor SCC3193 with saxA were able to cause spreading disease symptoms (Figure S2).

2.4 | Distribution of saxA virulence factor in the Pectobacterium genus

The saxA gene is widely distributed among Enterobacterales, and a phylogenetic analysis of a subset of enterobacterial saxA genes showed that the saxA gene family can be divided into five separate phylogenetic clusters, numbered CL1 to CL5 (van den Bosch et al., 2018). We analysed the distribution of saxA genes in 84 Pectobacterium and three Dickeya genomes that were previously analysed by Li et al. (2018) regarding virulence factors. The results of the BLAST analysis of the remaining 10 strains are discussed separately as the exact phylogeny is currently insufficiently supported. saxA from CL5 is widely distributed in the Pectobacterium genus, as it is represented in most strains of P. brasilienne, P. versatilis, P. polaris, and P. atrosepticum (Figure 1). Notable exceptions are P. polaris SS28 and a large branch of P. brasilienne that has lost this gene. saxA from CL2 seems to have been acquired only once in the Pectobacterium genus.
by a common ancestor of both *P. odoriferum* and *P. carotovorum* but not *P. brasiilense*. Five out of 97 genomes contained a saxA gene from both CL2 and CL5. These are distributed among the *P. carotovorum*, *P. odoriferum*, and *P. polaris* species. *Pectobacterium* saxA genes are restricted to phylogenetic clusters CL2 and CL5, with the exception of *P. wasabiae*. The *P. wasabiae* species is unique in that all three genomes (CP015750.1, JQHP00000000.1, and JQOH00000000.1), besides a CL2- and a CL5-related saxA gene, each contain a third copy of a saxA gene that is related to CL4. Neither of the two *P. beta-vascularulorum* strains (NCPPB2795 and NCPPB2793) contains saxA. All four of the *P. peruvianum* strains (IFBS232, IFBS229, A97-S13-F16, and A350-S18-N16) contain saxA from CL5, as well as *P. punjabense* SS92. No whole-genome sequences are known for *P. cacticida* and *P. aroidearum*, the two remaining known *Pectobacterium* species. Out of the 26 *Pectobacterium* strains that were isolated from *Brassica* hosts, 23 contained at least one saxA gene. The other three, *P. odoriferum* strains S6, S6–2, and BC57, were isolated from Chinese cabbage but did not contain a saxA gene, suggesting that saxA may not be required for infection of certain Brassica plants. These strains may possibly cope with the ITC content of their host not by degradation but by the use of efflux systems like saxD/F/G (Fan et al., 2011), or by other still unknown mechanisms. Lastly, 26 out of 39 known potato-isolated *Pectobacterium* strains contained a saxA gene from CL5, whereas *P. carotovorum* WPP14 is the only isolate from a potato host that contains a saxA gene from CL2, indicating either that those pectobacteria have a broader host range or that SaxA has a secondary function.

### 3 | DISCUSSION

A range of virulence factors and conditions are required for a pathogenic bacterium to successfully colonize and cause disease in a host plant. Many factors are important for virulence irrespective of host-specific defence mechanisms. In this study we focused on the chemical plant defence mechanism of the Brassicales order. We investigated whether a single enzyme that impairs this specific chemical host defence contributes to the virulence of several *Pectobacterium* spp. As a model for this, we chose *A. thaliana*. Brassicales plants employ the GSN–myrosinase defence system, which on tissue damage, releases a blend of toxic volatiles, of which ITCs are particularly toxic to insects and bacteria (Wittstock et al., 2003; Dufour et al., 2015). We have shown previously that the bacterial saxA gene encodes a hydrolyase that can detoxify ITCs in vitro and that the gene is widespread among Enterobacteriales (Welte et al., 2016; van den Bosch et al., 2018). In this study, we demonstrated that deletion of two different saxA gene variants results in greatly reduced virulence of *Pectobacterium* spp. on *A. thaliana* and *B. oleracea* var. *capitata*, thereby demonstrating that saxA is a vital virulence factor of *Pectobacterium* spp. when colonizing certain Brassicales plants. Moreover, the host range of a natural potato pathogen, *P. parmentieri* SCC3193, was broadened to include ITC-producing *Arabidopsis* by the introduction of a vector containing the saxA gene. The broadening of the host range after inclusion of saxA appears limited as SCC3193 did not readily infect cabbage leaves. Potato pathogen *P. polaris* NCPPB3395, containing two genomic saxA genes, was avirulent on *Arabidopsis* and only showed limited virulence on cabbage leaves. These results suggest that other factors are at play during infection of Brassicales. The evidence presented here shows that saxA specifically aids in overcoming the GSN–myrosinase system of the host, as saxA deletion mutants were only fully virulent in hosts with defective GSN biosynthesis pathways. The two model pathogens with a single genomic saxA copy (*P. odoriferum* NCPPB3841 and *P. versatile* SCC1) responded similarly to the deletion of their respective saxA genes, although the phylogenetic origins of those genes differ. It remains unclear what the functional differences are between saxA genes of different phylogenetic origins. These observations may be the effect of convergent evolution, although differential effectiveness against particular variations of GSN/ITC profiles of hosts cannot be excluded. The presence of two saxA genes in the genome of *P. polaris* NCPPB3395 and the presence of three saxA genes with unique origins in strains of *P. wasabiae* (Welte et al., 2016) suggest that there may be added benefit to diversifying genomic saxA gene content when infecting specific hosts. The analysis of the distribution of saxA genes in the *Pectobacterium* genus showed that most *Pectobacterium* strains isolated from *Brassica* host contain one or more saxA genes in their genome, with the exception of one group of *P. odoriferum* strains (S6, S6–2, and BC57). Assuming that the last common ancestor of *Pectobacterium* species contained a saxA gene from CL5, we may conclude that this gene was lost in this genus on multiple occasions, mostly in branches of the family tree that are associated with non-Brassicales hosts. For example, *P. parmentieri* and *P. actinidiae* have only been found on potatoes and kiwi fruits, respectively, and all strains in both species have lost saxA. Furthermore, *P. brasiilense* contains a branch of strains without saxA, all of which were isolated from potatoes (with the exception of PcbHPI01, which was isolated from cucumber, another non-brassica plant). Genes of the saxA family are not exclusively found within plant-pathogenic microbes but are relatively widespread among the Enterobacteriales, further suggesting that SaxA may serve additional purposes beside detoxifying plant ITCs. Whereas the clustering of saxA representatives from all Enterobacteriales results in five distinct phylogenetic groups, the saxA genes found exclusively in *Pectobacterium* cluster in three of these groups and appear to have at least three different evolutionary origins. However, the catalytic properties of SaxA are preserved in all five phylogenetic clusters, not only in those found in plant-pathogenic microbes, as demonstrated in our previous work (van den Bosch et al., 2018). Horizontal gene transfer between other members of the Enterobacteriales seems the most likely method of acquisition. Many bacteria encode the saxA gene on a mobile genetic element enabling mobilization and conjugation within a bacterial community (Welte et al., 2016), suggesting that this method for overcoming host plant resistance could spread quickly in a microbial community. The data presented here may help us understand why *Pectobacterium* is capable of infecting a broad spectrum of hosts.
The trial experiments on important cabbage crops indicate that our findings can be extrapolated to real-life scenarios, therefore potentially aiding research to improve food security.

4 | EXPERIMENTAL PROCEDURES

4.1 | Bacterial strains

*P. odoriferum* NCPPB3841 (isolated from *Cichorium intybus* in France, 1979) and *P. polaris* NCPPB3395 (Dees et al., 2017) were obtained from the National Collection of Plant Pathogenic Bacteria (NCPPB), United Kingdom. In addition, *P. versatile* SCC1 (CFBP8537) (Niemi et al., 2017; Portier et al., 2019) and *P. parmentieri* SCC3193 (CFBP8536) (Nykyri et al., 2012; Khayi et al., 2016) stored at the University of Helsinki were used. The strains were grown overnight at 28 °C in liquid Luria–Bertani (LB) medium (1% tryptone, 0.5% yeast extract, 0.5% NaCl) with shaking (200 rpm) or on LB plates solidified with 1.5% agar unless otherwise stated. Ampicillin (100 μg/ml), tetracycline (10 μg/ml), chloramphenicol (10 μg/ml), or kanamycin (25 μg/ml) was added to the medium when appropriate.

4.2 | Mutant generation

*saxA* deletion mutants of *P. polaris* NCPPB3395, *P. odoriferum* NCPPB3841, and *P. versatile* SCC1 were generated by φ. Red recombination as described by Datsenko and Wanner (2000) with a protocol optimized for *Pectobacterium* (Nykyri et al., 2012). Overnight cultures were diluted 1:50 in fresh medium and grown to an optical density (OD<sub>600</sub>) value of approximately 0.5 before being chilled on ice for at least 30 min. Cells were then collected from 2 ml of culture in microcentrifuge tubes by centrifugation at 7,740 g for 1 min at 4 °C, washed once with ice-cold sterile water and resuspended in 40 μl of ice-cold 10% glycerol. The cells were transformed in 0.2 cm cuvettes with 100 ng pKD46 plasmid by electroporation at 2.5 kV and then allowed a recovery period of 2 hr in super optimal broth with 1.5% agar unless otherwise stated. Ampicillin (100 μg/ml), tetracycline (10 μg/ml), chloramphenicol (10 μg/ml), or kanamycin (25 μg/ml) was added to the medium when appropriate.

| 4.3 | Complementation of mutants

*P. odoriferum* NCPPB3841 Δ*saxA*, *P. versatile* SCC1 Δ*saxA*, and *P. parmentieri* SCC3193 were complemented by producing SaxA in trans from the low-copy-number plasmid pMW119 (Nippon Gene, GenBank: AB005476.2). Selection of pMW119 is based on ampicillin, to which *P. versatile* SCC1 is naturally resistant. Thus, the *npt* gene conferring resistance to kanamycin was amplified by PCR from Entranceposon (Kan<sup>R</sup>-3) F-779 (Thermo Scientific) using primers Kan(SacI)_F and Kan(EcoRI)_R (Table S1). The resulting fragment was digested with SacI and EcoRI and ligated into SacI- and EcoRI-digested pMW119. Subsequently, the saxA gene was inserted into the multiple cloning site of pMW119-npt in the same orientation as lacZ in the plasmid. saxA genes were amplified by...
PCR from genomic DNA of *P. odoriferum* NCPPB3841 and *P. versatile* SCC1 using the primer pairs Pco_3841_US/Pco_3841_DS and SCC1_saxA(BamHl)/F(SCC1_saxA(SacI)_R) (Table S1), respectively. The *P. odoriferum* NCPPB3841 saxA fragment was blunt-end cloned into Smal-digested cloning vector pUC19, excised from pUC19 by digestion with SacI taking advantage of a naturally occurring SacI site located 320 bp upstream of the start codon within the saxA fragment in addition to the SacI site located within the multiple cloning site of pUC19, gel purified and ligated into SacI-digested pMW119-npt. The *P. versatile* SCC1 saxA fragment was digested with BamHl and SacI and ligated into BamHl- and SacI-digested pMW119-npt. Correctness of insert sequences and orientations was verified by PCR and sequencing (Table S1). The complementation plasmids and pMW119-npt without an insert were electroporated into competent cells of *P. odoriferum* NCPPB3841 ΔsaxA, *P. versatile* SCC1 ΔsaxA as well as *P. parmentieri* SCC3193 as described above for plasmid pKD46 and plated on LB plates containing 25 μg/ml kanamycin.

### 4.4 | Arabidopsis leaf maceration assay

*A. thaliana* ecotype Col-0 wild-type and myb28-1 myb29-1 (Sønderby et al., 2007) seeds were germinated in 2:1 peat:vermiculite mixture and individual plants were repotted in 7 × 7 cm pots after 1 week. Plants were kept under 12 hr day/night cycles with a temperature of 23 °C during the day and 19 °C during the night, and used in infection experiments at the age of approximately 30 days. Infection was carried out approximately 1–2 hr before the start of the night. Overnight-grown *P. odoriferum* NCPPB3841 and *P. versatile* SCC1 cells were washed with 10 mM MgSO₄ and diluted to an OD₆₀₀ of 0.002 corresponding to approximately 2 × 10⁶ cfu/ml. Three leaves per plant (usually leaves 8 to 10) were wounded with forceps, after which 5 μl drops of bacterial suspension were pipetted on top of the wounds. Water was added to the bottom of the plant tray and a lid was placed on top to keep the plants in high atmospheric humidity. After approximately 24 hr, inoculated leaves were detached from the rosette, gently dried with tissue paper, and placed between transparent films for scanning with resolution of 600 dots per inch (dpi). The macerated leaf area was manually measured from the scanned images using Fiji ImageJ v. 1.51. All experiments were conducted so that bacterial suspensions and plant lines were colour-coded and the person performing the infections and processing the images was unaware of the identities of the bacteria or plants until after all data were gathered. Each experiment was conducted three times. Leaves inoculated with *P. polaris* NCPPB3395 did not show maceration even when a higher inoculum of OD₆₀₀ of 0.2 was used. The conditions for the *Arabidopsis* leaf maceration experiment using *P. parmentieri* SCC3193 were slightly altered as described here. The cells were diluted to an OD₆₀₀ of 0.2 instead of 0.002 and not three but five leaves per plant (leaves 5 to 9) were wounded. The maceration of a leaf after 24 hr was qualified manually to be either positive or negative in four experiments totalling 30 plants for *P. parmentieri* SCC3193 transformed with empty pMW119 vector and *P. parmentieri* SCC3193 transformed with pMW119 + SCC1_saxA.

### 4.5 | *B. oleracea* var. *capitata* leaf maceration assay

Leaves of store-bought cabbages (*B. oleracea* var. *capitata*) were surface-sterilized with 70% ethanol and discs 3 cm in diameter were cut and placed on top of wet tissue paper on six-well plates. Overnight-grown *P. odoriferum* NCPPB3841 and *P. versatile* SCC1 cells were washed with 10 mM MgSO₄ and diluted to an OD₆₀₀ of 0.01 corresponding to approximately 10⁷ cfu/ml. A small hole was punctured with a pipette tip at the centre of each disc and a 5 μl drop of bacterial suspension was pipetted on top of the hole. The plates were sealed with Parafilm, placed in a closed container, and incubated in the dark at room temperature. After 48 hr, the discs were gently dried with tissue paper and placed between transparent film for scanning with resolution of 600 dpi. The macerated leaf area was manually measured from the scanned images using Fiji ImageJ v. 1.51. Twelve discs were inoculated with wild-type bacteria and 12 discs with the saxA mutant in each experiment. The experiment was conducted three times and one cabbage was used per experiment.

### 4.6 | Statistical analysis of maceration assays

Statistical analysis was performed in the R programming environment and figures were prepared using the ggplot2 package in R (Wickham, 2016; R Core Team, 2018). Due to non-normal distributions and/or unequal variances, the Kruskal–Wallis rank sum test followed by pairwise Wilcoxon rank sum test was used to address the statistical significance of differences between groups. *p* values were not adjusted to correct for multiple comparisons.

### 4.7 | *P. polaris* and *P. parmentieri* growth on *Arabidopsis* extracts

Plant extracts were prepared from 6-week-old *A. thaliana* ecotype Col-0 wild-type and myb28-1 myb29-1 (Sønderby et al., 2007) plants with a protocol modified from Fan et al. (2011). Rosette leaves were collected and 2 ml M9 salt solution (40 mM Na₂HPO₄, 22 mM KH₂PO₄, 8.5 mM NaCl, 18.7 mM NH₄Cl) was added for every 1 g of plant tissue followed by homogenization with a Polytron PT 2100 homogenizer (Kinematica AG). Cell debris was pelleted by centrifugation at 3,230 g for 15 min and the supernatant was first filtered through a 0.45 μm filter followed by filtering through a 0.20 μm filter to produce the final extract. A 96-well plate growth assay was used to assess the effect of saxA on the ability of *P. polaris* NCPPB3395 and *P. parmentieri* SCC3193 to grow in the presence of plant extract. Fresh colonies of *P. polaris* NCPPB3395 wild-type, ΔsaxA1, ΔsaxA2, and the ΔsaxA1 + ΔsaxA2 double mutant, as well as *P. parmentieri* SCC3193 and *P. parmentieri* SCC3193 transformed with pMW119 + SCC1_saxA were inoculated into the wells of 48-well plates containing 750 μl of liquid LB medium. Six inoculations were performed per strain and in addition six control wells with medium were left uninoculated. The
plate was incubated overnight at 28 °C with shaking (120 rpm). The contents of the wells were then diluted 1:5 in M9 salt solution supplemented with 10 mM sucrose on a 96-well plate and attenuation of light at 600 nm was measured using an Enspire 2300 plate reader (Perkin Elmer) to ensure approximately equal overnight growth of all cultures. Then 20 μl of the dilutions were used to inoculate wells of another 96-well plate containing 180 μl of M9 salt solution supplemented with 10 mM sucrose or 100 μl of plant extract (wild-type or myb28 myb29) and 80 μl of M9 salt solution supplemented with 10 mM sucrose. Attenuation at 600 nm was measured immediately after inoculation and subsequently every 2 hr for a period of 10 hr to detect bacterial growth. Between measurements the 96-well plate was incubated with a lid on at 28 °C with shaking (120 rpm).

4.8 | saxA distribution in the Pectobacterium genus

In order to understand the evolutionary history of saxA genes in the Pectobacterium genus, we screened 97 near-complete and complete genomes for the presence of saxA. Five saxA genes, one representative of each cluster (CDS accessions: ALG88671.1, AA055377.1, ANJ99341.1, QAA04876.1, AFR03517.1; van den Bosch et al., 2018) were used as query in NCBI’s discontinuous megablast using the most recent contigs of Pectobacterium whole-genome shotgun sequencing projects as the subjects. The BLAST subjects included 84 Pectobacterium and three Dickeya genus sequences analysed by Li et al., complemented with the genomes of 10 strains of P. wasabiae, P. betavasculorum, P. pueruliense, and the recently described species P. punjabense. The presence of a saxA gene was manually verified and the phylogenetic cluster of a positive saxA gene was determined by the representative with the closest hit.

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**SUPPORTING INFORMATION**

Additional supporting information may be found online in the Supporting Information section.

**Figure S1** In trans expression of saxA in Pectobacterium parmentieri SCC3193 increases virulence on Arabidopsis thaliana Col-0 plants. Symptoms caused by strain SCC3193 transformed with empty pMW119 vector and SCC3193 transformed with pMW119 carrying saxA from P. versatilis SCC1 24 hr (a) and 48 hr (b) after local inoculation with approximately 10⁶ cfu.

**Figure S2** Progression of symptoms caused by different Pectobacterium strains on cabbage (Brassica oleracea var. capitata) leaf discs after local inoculation. (a) P. odoriferum NCPPB3841 wild-type (wt) and saxA mutant (dsaxA), inoculum of approximately 5 × 10⁶ cfu. (b) P. versatilis SCC1 wild-type (wt) and saxA mutant (dsaxA), inoculum of approximately 5 × 10⁴ cfu. (c) Mock inoculation for (a) and (b) with 10 mM MgSO₄. (d) P. parmentieri SCC3193 wild-type (wt) and saxA1 saxA2 double mutant (dsaxA), inoculum of approximately 10⁵ cfu. (e) P. parmentieri SCC3193 transformed with empty pMW119 vector (wt) and SCC3193 transformed with pMW119 carrying saxA from P. versatilis SCC1 (+saxA), inoculum of approximately 10⁶ cfu.

**Table S1** List of primers used in the study

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