Review

Lipoxygenase Inhibition by Plant Extracts

Melita Lončarić 1, Ivica Strelec 1, Tihomir Moslavac 1, Drago Šubarić 1, Valentina Pavić 2 and Maja Molnar 1,*

1 Faculty of Food Technology Osijek, Josip Juraj Strossmayer University of Osijek, Franje Kuhaca 18, 31000 Osijek, Croatia; melita.loncaric@ptfos.hr (M.L.); ivica.strelec@ptfos.hr (I.S.); tihomir.moslavac@ptfos.hr (T.M.); drago.subaric@ptfos.hr (D.Š.)
2 Department of Biology, Josip Juraj Strossmayer University of Osijek, Ulica cara Hadrijana 8/A, 31000 Osijek, Croatia; vpavic@biologija.unios.hr
* Correspondence: maja.molnar@ptfos.hr

Abstract: Lipoxygenases are widespread enzymes that catalyze oxidation of polyunsaturated fatty acids (linoleic, linolenic, and arachidonic acid) to produce hydroperoxides. Lipoxygenase reactions can be desirable, but also lipoxygenases can react in undesirable ways. Most of the products of lipoxygenase reactions are aromatic compounds that can affect food properties, especially during long-term storage. Lipoxygenase action on unsaturated fatty acids could result in off-flavor/off-odor development, causing food spoilage. In addition, lipoxygenases are present in the human body and play an important role in stimulation of inflammatory reactions. Inflammation is linked to many diseases, such as cancer, stroke, and cardiovascular and neurodegenerative diseases. This review summarized recent research on plant families and species that can inhibit lipoxygenase activity.

Keywords: lipoxygenase; plant extracts; polyphenols

1. Introduction

Lipoxygenase (LOX) was discovered as an enzyme that catalyzes oxidation of carotene destruction, as first reported by Bohn and Haas. The enzyme was named carotene oxidase. Throughout the years, LOX has been identified in a variety of ways. From 1932 to 1940, names such as unsaturated lipid oxidase, lipoxidase, fat oxidase, and lipid oxidase were proposed [1]. In 1940, it was recognized as the same enzyme. All previously used names were consolidated and the enzyme was officially named lipoxygenase [2].

Lipoxygenases (LOXs) are a family of monomeric proteins that catalyze oxidation of polyunsaturated fatty acids (PUFA) (linoleic, linolenic, and arachidonic acid) to produce hydroperoxides [3]. LOXs are widely spread in animal, plant, and fungi kingdoms, as well as in cyanobacteria [4]. Moreover, 5-LOX is ubiquitous in mammalians and oxygenates carbon-5 on arachidonic acid, while 9-LOX and 13-LOX are plant LOXs that catalyze oxygenation of linoleic and linolenic acid [4,5].

Lipoxygenases are present in the human body and play an important role in the stimulation of inflammatory reactions. Excessive amounts of reactive oxygen species can cause inflammation that stimulates a release of cytokines and subsequent activation of LOXs. Inflammation is linked to many diseases, such as cancer, stroke, and cardiovascular and neurodegenerative diseases. LOXs are involved in the synthesis of prostaglandins and leukotrienes. They are associated with disease development and their inhibition is considered as a crucial step in disease prevention [6].

LOXs can be abundantly found in cereals (wheat, corn, oat, rye, barley), legumes (mung beans, green beans, peas, navy beans, soybeans), and potato tubers [7,8]. Products generated by LOX pathways have various functions. LOX is used as storage protein during vegetative growth. In addition, LOX is related to the mobilization of storage lipids during germination [9]. LOX also plays an important role in the production of protective components (jasmonates, divinyl ethers, leaf aldehydes), which helps protect plants from...
insects and pathogens or during abiotic stress [10,11]. On the other hand, LOX reactions with unsaturated fatty acids can produce off-flavors/off-odors and cause food spoilage. Due to the undesired components of the LOX pathway, research was carried regarding lipoxygenase inhibition [3,12].

Plant phytochemicals play an important defensive role, which could be beneficial in the prevention of diseases caused by oxidative stress. Various plants have been used for thousands of years as medicine. Nowadays, many drugs are produced via isolation from natural sources and there is increasing interest in plant derived therapeutics [13].

The aim of this review was to summarize the plant extraction methods for the best inhibitory activity of lipoxygenase enzymes recently published.

2. Lipoxygenase Inhibition by Plant Extracts of Family Apiaceae

Apiaceae (Umbelliferae) is an angiosperm family that consists of 454 genera and roughly 3700 species. Family members are used as food, spices, as well as medicinal plants, since they are rich in polyphenols. They possess aromatic nature and wide range of chemical compounds, but some of them can also be toxic [14]. Many compounds of the Apiaceae family possess biological activities, such as antibacterial, vasorelaxant, hepatoprotective, cyclooxygenase inhibitory, antitumor activities, and many others [15]. A short overview of the extraction on different plant parts of the Apiaceae family, with the investigation of their inhibitory activity against LOX, is shown in Table 1.

Table 1. Lipoxygenase inhibition by extracts of plants of family Apiaceae.

| Plant Species                     | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC₅₀ µg/mL) | References |
|----------------------------------|---------------------|----------------------|------------------------|----------------------------|------------|
| Alepidea amatymbica Eckl. & Zeyh| Acetone             | Roots                | -                      | -                          | [16]       |
| Apium graveolens Linn.           | Methanol (60%)      | Seeds                | 0.34                   | -                          | [17]       |
| Petroselinum crispum Mill.       | Methanol            | Aerial parts         | 0.29                   | -                          | [18]       |
| Bupleurum marginatum Wall.       | Methanol            | Aerial parts         | -                      | 45.28                      | [19]       |
| Scandix pecten-veneris L.         | Methanol (85%, v/v) | Leaves               | 377.94                 | 641.57                     | [20]       |
| Pteruranthos chloranthus Benth.   | Water               | Aerial parts         | 1.36                   | 20.00                      | [21]       |
| Eryngium planum L.               | Ethanol (50%, v/v)  | Aerial parts         | 1.88                   | 31.30                      | [22]       |
| Levisticum officinale W.D.J. Koch| Ethanol:water:HCl (70:29:1 v/v/v) | Leaves | - | - | [23] |
| Anethum graveolens L.            | Water               | Aerial parts         | 67.22                  | 43.00                      |            |
| Foeniculum vulgare Mill.         | Water               | Aerial parts         | 35.76                  | 52.00                      | [23]       |
| Pimpinella anisum L.             | Water               | Aerial parts         | 29.08                  | 35.00                      |            |

1 results are expressed as mg gallic acid equivalent (GAE)/mL of extract. 2 results are expressed as mg gallic acid equivalent (GAE)/g of extract.

Muleya et al. isolated five diterpenoids from the roots of the plant Alepidea amatymbica [16]. Inhibition of 15-soybean lipoxygenase were performed and 14-acetoxo-12-oxokaur-16-en-19-oic acid showed high inhibitory activity (EC₅₀ = 19.10 µg/mL), slightly
lower than the control compound indomethacin (EC\textsubscript{50} = 17.22 \mu g/mL). Four other compounds showed poor-to-moderate LOX inhibition in EC\textsubscript{50} range 25.98–81.18 \mu g/mL.

Inhibition of 15-lipoxygenase with seed extracts of \textit{Apium graveolens} and \textit{Petroselinum crispum} was performed by Danciu et al. [17]. It was found that 86.15 mg of \textit{Apium graveolens} and 69.45 mg of \textit{Petroselinum crispum} extract is necessary to inhibit 50% of 15-lipoxygenase activity. Phytochemical compositions of obtained methanolic extracts were estimated and nine components were detected. Apigenin and apigenin glucoside were the most prevalent compounds in both extracts. Aerial parts of \textit{Bupleurum marginatum} were extracted in two different solvents, methanol and dichloromethane [18]. Both extracts showed 5-lipoxygenase inhibitory activity at concentration of 25 \mu g/mL. Methanolic extract showed higher IC\textsubscript{50} value (45.28 \mu g/mL) than the extract obtained in dichloromethane (25.92 \mu g/mL). Phytochemical studies showed that \textit{B. marginatum} extracts are rich in lignans, flavonoids, saikosaponins, and other terpenoids. Methanol extract possessed more flavonoids, while dichloromethane extract had more lignans and terpenoids. The potential ability of soybean lipoxygenase inhibition was also tested by the methanolic extract of \textit{Scandix pecten-veneris} [19]. Enzyme inhibition was measured on five different concentrations of plant extracts, of 100, 200, 300, 600, and 900 \mu g/mL, and obtained results showed 3.50, 16.42, 34.62, 44.56, and 77.49% of LOX inhibition, respectively. Total phenolic content of examined extract was 377.94 mg equivalent (GAE)/g extract. It was reported that coumarin umbelliprenin found in the plant family Apiaceae strongly inhibits lipoxygenase [24]. Water extract of \textit{Pituranthos chloranthus} showed strong lipoxygenase inhibition with IC\textsubscript{50} value of 0.02 mg/mL [20]. Total polyphenol content of the extract was 1.36 mg GAE/g. Faun et al. extracted aerial parts of \textit{Eryngium planum} in 50% ethanol with water (v/v) [21]. The obtained extract was subjected to lipoxygenase inhibition testing and showed high inhibitory activity with an IC\textsubscript{50} value of 31.30 \mu g/mL. The polyphenol profile of \textit{E. planum} showed a high concentration of flavonoids, especially isoquercetin (36.11 \mu g/mL) and rutin (290.52 \mu g/mL).

To increase polyphenol production, Złotek and co-workers used jasmonic acid (1, 10, 100 \mu M) and yeast extract (0.01, 0.1, 1%) as elicitors [22]. The best results were observed in \textit{Levisticum officinale} leaves treated with 10 \mu M jasmonic acid, where total polyphenol content increased by 55% in comparison with the control sample. All extract showed the ability to inhibit lipoxygenase, but inhibition rate of elicited leaves did not differ significantly from the control. Similar research was conducted by Majdoub et al., who monitored the effect on 2 mM Zn on polyphenol amount in \textit{Anethum graveolens} (dill), \textit{Pimpinella anisum} (anise), and \textit{Foeniculum vulgare} (fennel) [23]. Total polyphenol content increased in dill and anise plants, while in fennel, a decrease of phenol content was observed. Treated and non-treated samples had the ability to inhibit 5-lipoxygenase. Anise showed strong lipoxygenase inhibitory activity, with IC\textsubscript{50} of 0.035 and 0.015 mg/mL for Zn non-treated and treated plants, respectively. In the case of dill, no significant difference was observed in LOX inhibition before (IC\textsubscript{50} = 0.043 mg/mL) and after (IC\textsubscript{50} = 0.041 mg/mL) treatment with Zn. Fennel showed higher IC\textsubscript{50} value in the sample treated (IC\textsubscript{50} = 0.062 mg/mL) with Zn, compared to non-treated (IC\textsubscript{50} = 0.052 mg/mL) one.

Among investigated extracts of the Apiaceae family, the best inhibitory activity of lipoxygenase was noticed in water extract of \textit{P. chloranthus}. Further, strong inhibitory activity (IC\textsubscript{50} = 25.92 \mu g/mL) of \textit{B. marginatum} extract could be related to phytochemicals identified in dichloromethane extract: isorhamnetin and its glycoside narcissin, quercetin and its glycoside rutin and isoquercitrin, (3,4-dimethoxybenzyl)-2-(3,4-methylenedioxybenzyl) butyro lactone and marginatoxin [25].

3. Lipoxygenase Inhibition by Plant Extracts of Family Asteraceae

Asteraceae is the biggest family of flowering plants and counts about 1100 genera and 20,000 species. Many plants from the Asteraceae family are used as traditional medicine due to the possession of numerous phytochemical components, such as polyphenols and diterpenoids. Antifungal, antibacterial, insecticide, anti-inflammatory, and antitumor
activities were investigated in the Asteraceae species [26]. Many plants from this family were assayed on the inhibitory activities against the lipoxygenase enzyme (Table 2).

Table 2. Lipoxygenase inhibition by extracts of plants of the family Asteraceae.

| Plant Species       | Extraction Solvents | Plant Part    | Total Phenolic Content | LOX Inhibition (IC\(_{50}\) µg/mL) | References |
|---------------------|---------------------|---------------|------------------------|------------------------------------|------------|
| *Matricaria chamomilla* L. | Methanol (60%) | Flowers | 458.10 \(^1\) | - | [17] |
| *Artemisia vulgaris* L. | Water | Aerial parts | 0.91 \(^2\) | 40.00 | [20] |
| *Cnicus benedictus* L. | Ethanol (50%) | Aerial parts | 2.65 \(^1\) | 52.7 | [21] |
| *Artemisia nilagirica* C.B. Clarke | Methanol | Leaves | - | 128.20 | [27] |
| *Chrysanthemum indicum* L. | Chloroform | Whole | 1.65 \(^3\) | - | [28] |
| *Tagetes erecta* L. | Chloroform | Whole | 6.97 \(^3\) | - | |
| *Vernonia oligocephala* DC. | Methanol (80%) | Roots | 113.11 \(^2\) | >500 | [29] |
| *Tanacetum ciliicum* Boiss. | Ethanol | Capitula | 87.01 \(^2\) | 156.00 | [30] |
| *Galatella tatarica* Less. | Methanol | Whole | 0.41 \(^1\) | - | [31] |
| *Galatella villosa* L. | Methanol | Whole | 0.39 \(^1\) | - | |
| *Cota fulvida* Holub. | Methanol | Aerial parts | 0.29 \(^1\) | - | [32] |

\(^1\) results are expressed as mg gallic acid equivalent (GAE)/mL extract; \(^2\) results are expressed as mg gallic acid equivalent (GAE)/g extract; \(^3\) results are expressed as g gallic acid equivalent (GAE)/100 g dry extract.

Bhat et al. extracted leaves of *Artemisia nilagirica* in three different solvents (hexane, methanol, and ethanol) [27]. All extracted inhibited lipoxygenase, but the most effective was the methanolic extract, with an IC\(_{50}\) value of 128 µg/mL, followed by hexane extract.
(196 µg/mL). Ethanolic extract showed poor inhibition of 21%. Burlec et al. extracted two plants from the Asteraceae family (*Chrysanthemum indicum* and *Tagetes erecta*), in four different organic solvents (chloroform, dichloromethane, hexane, and methanol) in order to investigate extract inhibition ability of 15-lipoxygenase [28]. The strongest enzyme inhibition of *Chrysanthemum indicum* was in chloroform extract with EC$_{50}$ value of 26.06 µg/mL while *Tagetes erecta* methanol extract showed similar inhibition (EC$_{50}$ = 25.85 µg/mL). The highest phenolic concentration was obtained in *Chrysanthemum indicum* chloroform extract, while dichloromethane extract of *C. indicum* showed the highest polyphenol content. Five different organic solvents (methanol, dichloromethane, ethyl acetate, n-hexane, and n-butanol) were employed in the extraction of *Vernonia oligocephala* roots [29]. All extracts were investigated for their lipoxygenase inhibition potential. Among all extracts, only butanol extract showed considerable LOX inhibition (IC$_{50}$ = 132.20 µg/mL). Phytochemical composition analysis of the roots showed the presence of saponins, terpenoids, flavonoids, glycosides, phenolic compounds, and steroids. Methanol extract had the highest flavonoid (97.35 mg quercetin equivalent (QE)/g of extract) and phenolic (113.11 mg GAE/g of extract) content. Yıldırım et al. extracted different parts of *Tanaacetum ciliicum* (capitula, leaves, and stem) in different solvents (ethanol, n-hexane, chloroform, and ethyl acetate) [30]. Extract of capitula in ethyl acetate showed significant LOX inhibition, with an IC$_{50}$ value of 9.44 µg/mL, which is even greater than the standard indomethacin (IC$_{50}$ = 22.39 µg/mL). Ethyl acetate was also proven to be the best choice for the leaves extraction, in respect to LOX inhibition (IC$_{50}$ = 85.20 µg/mL), while the best inhibition among stem extracts was observed for chloroform extracts (IC$_{50}$ = 112.40 µg/mL). Total polyphenol content in extracts varied 6.10–175.60 mg GAE/g of dry weight, while total flavonoid content was in the range of 6.13 to 30.48 mg QE/g of dry weight. Phenolic contents were the highest in all ethyl acetate extracts. Water extract of *Artemisia vulgaris* investigated by Nasr et al. had a high ability of lipoxygenase inhibition (IC$_{50}$ = 40.00 µg/mL) [20]. Total phenolic content of aqueous extract was 0.91 mg GAE/g of extract.

Ethanolic extract of *Cnicus benedictus* investigated by Paun et al. also showed very good ability of lipoxygenase inhibition (IC$_{50}$ = 52.70 µg/mL) [21]. Total flavonoid content in extract was 0.47 mg QE/mL and total phenolic content was 2.65 mg GAE/mL. High concentrations of phenolic acids were determined (especially sinapic acid and chlorogenic acid). Further, methanolic extract of *Matricaria chamomilla* showed good inhibition of lipoxygenase with EC$_{50}$ value of 166.32 µg/mL [17]. Total polyphenol content (458.10 µg GAE/mL of extract) and total flavonoids (342.40 µg/mL of extract) of extract were determined. Özek et al. investigated LOX inhibition ability of essential oils and methanolic extracts from *Galatella villosa* and *Galatella tatarica* [31]. The 5-LOX was inhibited with both extracts and essential oils. The oils proved to be better inhibitors with percentages of inhibition of 45.0 and 57.0% for *G. tatarica* and *G. villosa*, respectively. Both methanolic extracts showed poor 5-LOX inhibition of 29.0% and 19.0%, respectively. Content of total phenolic compounds did not differ significantly in the extracts. Similar research was conducted with the oil and extract of *Cota fulvida* and similar results were obtained [32]. The essential oil inhibited 5-LOX for 53.7% in comparison to poor inhibition of 23.9% of methanol extract.

The highest capacity of LOX inhibition was achieved with ethyl acetate extract of *T. ciliicum* capitula. Among all extracts, this one showed high phenolic content (174.10 mg GAE/g of extract) as well as flavonoid concentration (26.94 mg QE/g of extract).

### 4. Lipoxygenase Inhibition by Plant Extracts of Family Clusiaceae

The Clusiaceae family has approximately 400 species widespread in Africa, Asia, Polynesia, and South America. *Garcinia* is the most numerous genus of Clusiaceae, and its plant parts, such as fruit, leaves, stems, and flowers, are commonly used for the treatment of inflammation and oxidative stress. Some plants also possess biological activities, such as antioxidant, anti-inflammatory, and antibacterial [33]. Potential lipoxygenase inhibition was investigated for extract of *Garcinia* plants (Table 3).
Table 3. Lipoxygenase inhibition by extracts of plants of family Clusiaceae.

| Plant Species              | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC$_{50}$ µg/mL) | References |
|----------------------------|---------------------|----------------------|------------------------|-----------------------------------|------------|
| *Garcinia porrecta* Laness.| Methanol Ethyl acetate n-Hexane | Stem bark | -                     | 0.23 0.52 4.87                  | [34]       |
| *Garcinia hombroniana* Pierre | Ethyl acetate n-Hexane | Leaves | -                     | 1.31 0.13 2.05                  | [35]       |
| *Garcinia lateriflora* Blume | Ethyl acetate n-Hexane | Leaves | -                     | 0.69 0.79 1.31                  | [36]       |
| *Garcinia kydia* Roxburgh  | Methanol Ethyl acetate n-Hexane | Leaves | -                     | 0.5 0.21 3.57                  | [37]       |

Sari and Elia performed LOX inhibition assay on the stem bark extracts of *Garcinia porrecta* Laness [34]. Extraction was conducted in different organic solvents (methanol, n-hexane, and ethyl acetate). LOX inhibition activity of extracts with IC$_{50}$ values were 0.23, 0.52, and 4.87 µg/mL for methanol, ethyl acetate, and n-hexane extracts, respectively. Total flavonoids of methanolic extract were equal to 5.66 mg QE/g. Extraction of *Garcinia hombroniana* Pierre leaves in n-hexane, methanol, and ethyl acetate was performed by Marlin and Elia [35]. It was proven that all leaf extracts are excellent LOX inhibitors with IC$_{50}$ values of 0.13, 1.31, and 2.05 µg/mL for ethyl acetate, methanol, and n-hexane extracts, respectively. Determined flavonoid content of the most active ethyl acetate extract was 42.00 mg QE/g sample. LOX inhibition potential of *Garcinia lateriflora* Blume leaf extracts was investigated [36]. IC$_{50}$ values of the LOX inhibition were 0.69, 0.79, and 1.32 µg/mL for methanol, ethyl acetate and n-hexane extracts, respectively. The most active extract was the methanolic one, with total flavonoid content of 6.29 mg QE/g of extract. Further, *Garcinia kydia* Roxburgh leaves were extracted with three previously mentioned solvents [37]. Obtained results on LOX inhibition had IC$_{50}$ values of 0.21, 0.55, and 3.57 µg/mL ethyl acetate, methanol, and n-hexane extracts, respectively. Total flavonoid content of 30.65 mg QE/g of extract was determined in the most active ethyl acetate extract.

Four mentioned *Garcinia* plants from the Clusiaceae family were extracted in the same solvents, and all showed strong inhibition of the LOX enzyme. It could be noticed that the best results are obtained with methanol and ethyl acetate extracts, while n-hexane extracts showed a slightly lower inhibition activity. The strongest LOX inhibitor was the ethyl acetate extract of *G. hombroniana* with the highest amount of total flavonoids among all investigated extracts.

5. Lipoxygenase Inhibition by Plant Extracts of Family Fabaceae

The Fabaceae family, also commonly known as Leguminosae, is the third largest plant family after Orchidaceae and Asteraceae, with 730 genera and more than 19,400 species. It includes shrubs, trees and herbaceous plants, which are easily recognized by their fruits (legume) and stipulated leaves. This plant family is mostly found in dry forests and in tropical rainforests [38]. Table 4 presents current knowledge on the extracts of plants from the family Fabaceae and their effect on LOX inhibition.

Souleymane et al. investigated 12-LOX inhibition by dichloromethane and ethyl acetate extracts of *Erythrina senegalensis* roots and stem bark [39]. Ethyl acetate extracts showed higher enzyme inhibition with IC$_{50}$ values of 4.95 and 7.21 µg/mL for the stem bark and roots, respectively. IC$_{50}$ values, concerning dichloromethane extracts, were
21.46 µg/mL for stem bark and 22.03 µg/mL for roots. Root extracts contained more phenolic acids, while stem bark contained more flavonoids, but the inhibition was found to be strongly dependent on the solvent used.

Yoo et al. conducted inhibition of 5-LOX with *Sophora tonkinensis* extract [40]. For the extraction, water ethanol solution (50%, v/v) was employed. *S. tonkinensis* extract showed strong LOX inhibition (IC$_{50}$ = 1.61 µg/mL). Methanol extract of *Cassia Alata* leaves showed LOX inhibition in the range from 28.3% (at 25.00 µg/mL) to 67.0% (at 100 µg/mL) [41]. Plant polyphenol profile affects the lipoxygenase (LOX)-dependent arachidonic acid metabolism, so the inhibition activity of the extracts could be explained by the polyphenol inhibiting the catalytic activities of LOX [42,43].

Ouédraogo and co-workers extracted the roots of *Pterocarpus erinaceus* in dichloromethane and methanol [44]. Individual components were isolated from dichloromethane (friedelin, 3α-hydroxyfriedelan-2-one, α-sophoradiol and stigmasterol) and methanol (maltol-6-O-apiofuranoside-glucopiranoside) extracts. Extracts, as well as individual components, were subjected to LOX inhibition assay. Dichloromethane and methanol extracts inhibited LOX activity for 54.2% and 45.1% at a concentration of 100 µg/mL, respectively. However, individual components were found more active than the extracts themselves. Maltol-6-O-apiofuranoside-glucopiranoside, friedelin and α-sophoradiol showed a significant LOX inhibition with IC$_{50}$ values of 23.11, 24.44, and 23.91 µg/mL, respectively.

*Crotalaria longipes* plant was extracted in ethanol [45]. LOX activity was checked at different extract concentrations (100–500 mg/mL). The highest inhibition activity (72%) was noticed at 500 mg/mL.

**Table 4.** Lipoxygenase inhibition by extracts of plants of family Fabaceae.

| Plant Species            | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC$_{50}$ µg/mL) | References |
|--------------------------|---------------------|----------------------|------------------------|------------------------------------|------------|
| *Erythrina senegalensis* DC. | Dichloromethane      | Stem bark            | -                      | 21.46                               | [39]       |
|                          | Ethyl acetate       |                      |                        | 4.95                                |            |
| *Sophora tonkinensis* Gahnep. | Dichloromethane     | Roots                | -                      | 22.03                               |            |
|                          | Ethyl acetate       |                      |                        | 7.21                                |            |
| *Cassia Alata*           | Methanol            | Leaves               | -                      | 1.61                                | [40]       |
| *Pterocarpus erinaceus* Poir. | Dichloromethane     | Root barks           | -                      | -                                   | [44]       |
|                          | Methanol            |                      |                        | -                                   |            |
| *Crotalaria longipes* Wight. | Ethanol             | Whole                | -                      | -                                   | [45]       |

6. Lipoxygenase Inhibition by Plant Extracts of Family Lamiaceae

Lamiaceae family is known as a mint family of flowering plants. It contains about 236 genera and 7200 species. The planta are usually aromatic, similar to some culinary herbs (mint, basil, sage, rosemary, oregano, lavender) [46].

Göger et al. investigated 5-LOX inhibition by *Marrubium stiosense* methanolic extract [47]. It was found that extract (at 1 mg/mL) shows poor inhibition of 18.7%. Individual polyphenols components were determined in methanolic extract, namely leucosепtose A, forsythoside B, verbascoside, martynoside, alyssonoside, as well as glucosides of quercetin, chrysoeriol, and apigenin, and coumaroylglucoside of apigenin.

*Ajuga reptans* and *Ajuga genevensis* are used as traditional medicine for their diuretic, tonic, and astringent activity [48]. Their ethanolic extracts were investigated as potential LOX inhibitors. Obtained results showed that extract of *Ajuga genevensis* is a better 15-LOX inhibitor, but both extracts were found as weak LOX inhibitors with inhibition rates lower than 30.0% at a concentration of 5 mg/mL. *A. genevensis* extract had a higher phenolic
and flavonoid content than *A. reptans* extract. Six individual phenolic components (caffeic acid, coumaric acid, rosmarinic acid, luteolin-β-glucoside, luteolin, and apigenin) were identified in both extracts. The most abundant compound was luteolin-β-glucoside in concentrations of 83.25 and 63.21 µg/g of dry weight for *A. reptans* and *A. genevensis*, respectively.

*Clerodendrum* leaves are used for the treatment of diseases, such as asthma and rheumatism. Phosrithong and Nuchtavorn extracted *Clerodendrum disparifolium* and *Clerodendrum laevifolium* leaves in ethanol and hexane in order to investigate their ability to inhibit 5-LOX activity [49]. Both plant extracts showed great potential in the inhibition of LOX activity. IC$_{50}$ values were more promising for ethanol extract, 24.39 and 14.12 µg/mL, while IC$_{50}$ values in hexane extract were slightly higher, 30.56 and 30.70 µg/mL for *C. disparifolium* and *C. laevifolium*, respectively (Table 5). Total phenolic and flavonoid content were determined in all extracts. Total polyphenols were in the range of 1167.20 to 3344.50 mg GAE/g of dry weight, while flavonoid amount was in the range 9.05–59.91 mg QE/g of dry extract. Ethanol extract with the highest polyphenol content also showed the strongest LOX inhibition.

Table 5. Lipoxygenase inhibition by extracts of plants of family Lamiaceae.

| Lamiaceae Plants | Plant Species | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC$_{50}$ µg/mL) | References |
|------------------|---------------|---------------------|----------------------|------------------------|----------------------------------|------------|
| *Marrubium sivasense* Aytaç, Akgül & Ekici | Methanol | Aerial parts | - | - | [47] |
| *Ajuga genevensis* L. *Ajuga reptans* L. | Ethanol (70%) | Whole | 354.70 ² | - | 296.50 ² | [48] |
| *Clerodendrum disparifolium* Blume | Ethanol | Leaves | 1167.20 ¹ | 24.39 | 1272.40 ¹ | [49] |
| *Clerodendrum laevifolium* Blume | Ethanol | Leaves | 3344.50 ¹ | 14.12 | 1611.00 ¹ | [49] |

¹ results are expressed as mg gallic acid equivalent (GAE)/g dry extract; ² results are expressed in mg gallic acid %.

7. Lipoxygenase Inhibition with Plant Extracts of Family Melastomataceae

Plants *Memecylon talbotianum*, *Memecylon umbellatum*, and *Memecylon malabaricum* belong to the family Melastomataceae. Some of the *Memecylon* species are traditionally used as a cure for skin problems [50].

Leaves of mentioned plants are dried and extracted in methanol in order to examine the 15-LOX inhibition [51]. Obtained IC$_{50}$ values were 29.87, 39.19, and 54.60 µg/mL for *M. malabaricum*, *M. umbellatum*, and *M. talbotianum*, respectively (Table 6). Estimated amount of phenolic content were 1.12, 1.08, and 1.96 g GAE/100 g dry weight for *M. malabaricum*, *M. umbellatum*, and *M. malabaricum*, respectively. Among the tested extracts, the one with the best LOX inhibitory activity also showed the highest polyphenol content. Afagnigni et al. extracted leaves of *Dissotis multiflora* from the family Melastomataceae in ethanol [52]. Inhibition of 15-LOX was examined at different concentrations of the extract. Obtained result showed that inhibition of 15-LOX was lower than 50% at the concentration of 100 µg/mL.
Table 6. Lipoxygenase inhibition by extracts of plants of family Melastomataceae.

| Melastomataceae Plants | Plant Species               | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC₅₀ µg/mL) | References |
|------------------------|-----------------------------|---------------------|----------------------|------------------------|-----------------------------|------------|
|                        | *Memecylon talbotianum* Brandis | Methanol            | Leaves               | 1.12 ¹                  | 34.60                       |            |
|                        | *Memecylon malabaricum* Cogn. | Methanol            | Leaves               | 1.96 ¹                  | 29.87                       | [51]       |
|                        | *Memecylon umbellatum* Burm. f. | Methanol            | Leaves               | 1.08 ¹                  | 39.19                       |            |
|                        | *Dissotis multiflora* Sm.    | Ethanol             | Leaves               | -                      | -                           | [52]       |

¹ results are expressed as g gallic acid equivalent (GAE)/100 g dry weight.

8. Lipoxygenase Inhibition by Plant Extracts of Family Ericaceae

*Gaultheria* plants belong to the Ericaceae family and there is more than 1700 species of this plant. *Gaultheria trichophylla* has blue berries and pink to red flowers. It also known as Himalayan Snowberry. It is commonly used as a folk medicine for the treatment of arthritis. Methanolic and chloroform extracts of *Gaultheria trichophylla* were found to inhibit 5-LOX with 90.5% and 66.9% inhibition at 0.5 mg/mL and IC₅₀ values of 277.30 µg/mL and 379.50 µg/mL, respectively (Table 7). n-Hexane extract showed lower inhibition of 57.0%. The highest flavonoids (41.30 mg QE equivalent/g) and phenolic (17.50 mg GAE equivalent/g) concentrations were found in methanolic extract [53].

Table 7. Lipoxygenase inhibition by extracts of plants of family Ericaceae.

| Ericaceae Plants | Plant Species               | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC₅₀ µg/mL) | References |
|-----------------|-----------------------------|---------------------|----------------------|------------------------|-----------------------------|------------|
|                 | *Gaultheria trichophylla* Royle | Methanol            | Whole                | 17.50 ¹                 | 277.30                      | [53]       |
|                 |                             | Chloroform          | Whole                | 5.00 ¹                  | 379.50                      |            |
|                 |                             | n-Hexane            | Whole                | 3.20 ¹                  | 448.00                      |            |
|                 | *Gaultheria procumbens* L.  | Petroleum ether     | Leaves               | -                      | 738.49                      | [54]       |
|                 |                             | Chloroform          | Leaves               | -                      | 899.97                      |            |

¹ results are expressed as mg gallic acid equivalent (GAE)/g of extract.

Leaves of *Gaultheria procumbens* L. were extracted in petroleum ether and chloroform [54]. Lipophilic extracts of the wintergreen leaves showed inhibitory effects on LOX with IC₅₀ values of 738.50 µg/mL and 899.97 µg/mL in the case of petroleum ether and chloroform extract, respectively. The performed GC-MS analysis of petroleum ether extracts revealed the presence of four major compound groups (aliphatic hydrocarbons, alcohols, carboxylic acids, and terpenoids). The docanose and octacosane were present in the highest amounts. Simple phenols and terpenoids were also present in significant percentage (methyl benzoate, ursolic acid, α-amirin, β-sitosterol, methyl salicylate, and oleanolic acid). Chloroform extract contained a lower amount of alcohol, simple aliphatic hydrocarbons, and carboxylic acid compared with petroleum ether extract. Dominant compounds in chloroform extract were ursolic acid, methyl benzoate, methyl salicylate, and oleanolic acid.

It could be noticed that all examined extracts from the family Ericaceae had poor LOX inhibition with high IC₅₀ values.
9. Lipoxygenase Inhibition by Plant Extracts of Family Menispermaceae

Plant *Cyclea Barbarata* belong to the plant family of Menispermaceae. Leaves of this plant contain many secondary metabolites, such as flavonoids, steroids, tannins, and saponins. It was reported that flavonoids have many benefits, such as antibacterial, anti-inflammatory, anticancer, antioxidant, hepatoprotective, and antiviral properties [55,56]. Handayani et al. reported inhibition of LOX with leaf extracts of *C. Barbarata* [57]. Three various solvents (ethyl acetate, *n*-hexane, and methanol) were employed for extraction. Among the investigated extracts, only ethyl acetate extract showed significant inhibition of LOX with an IC$_{50}$ value of 0.26 µg/mL (Table 8). Total flavonoids content of ethyl acetate extracts was 21.62 mg QE/g.

| Menispermaceae Plants | Plant Species | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC$_{50}$ µg/mL) | References |
|-----------------------|---------------|---------------------|----------------------|------------------------|---------------------------------|------------|
| *Cyclea Barbarata* Miers. | Ethyl acetate *n*-Hexane Methanol | Leaves | - | 0.26 | | [57] |
| *Sphenocentrum jollyanum* Pierre. | Ethanol (70%) | Roots Stem | - | 541.00 426.00 | | [58] |

LOX inhibition activity of ethanolic extracts of *Sphenocentrum jollyanum* was investigated by Sinbad et al. [58]. *S. jollyanum* possesses many biological and pharmaceutical properties, including antimalarial, antioxidant, anti-inflammatory, antidiabetic, and anti-angiogenic. Roots and stems were extracted and both extracts showed poor inhibition of LOX enzyme with IC$_{50}$ values of 541.00 and 426.00 µg/mL, respectively.

10. Lipoxygenase Inhibition by Plant Extracts of Family Rosacea

*Cydonia oblonga* is a plant from the Rosacea family and it is a rich source of antioxidants. Phytochemical screening revealed the presence of cardiac glycosides, alkaloids, terpenoids, tannins, flavonoids, saponins, and steroids [59]. Berkoz determined lipoxygenase inhibition and total phenolic content in the crude extract, aqueous and organic fractions (chloroform, ethyl acetate, butanol) of dry fruit of *Cydonia oblonga* [60]. The best LOX inhibition was achieved by methanol extract (IC$_{50}$ = 99.30 µg/mL), which showed the highest phenolic content of 367.00 mg GAE/g of extract. Other extracts also appeared to be the prominent inhibitors of LOX with IC$_{50}$ values in the range of 101.80 to 227.30 µg/mL (Table 9).

| Rosacea Plants | Plant Species | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC$_{50}$ µg/mL) | References |
|----------------|---------------|---------------------|----------------------|------------------------|---------------------------------|------------|
| *Cydonia oblonga* Mill. | Methanol (70%) Water Ethyl acetate Chloroform Butanol | Fruit | 367.00 $^1$ 115.30 $^1$ 65.00 $^1$ 244.90 $^1$ 123.40 $^1$ | 99.30 101.80 227.30 102.40 207.00 | | [60] |
| *Prunus spinosa* L. | Methanol (70%) Diethyl ether Ethyl acetate *n*-Butanol Water | Flower | 206.07 $^2$ 464.57 $^2$ 584.07 $^2$ 296.57 $^2$ 64.60 $^2$ | 327.36 150.36 135.36 171.10 479.50 | | [61] |

$^1$ results are expressed as mg gallic acid equivalent (GAE)/g of extract. $^2$ results are expressed as mg gallic acid equivalent (GAE)/g of dry weight.
Similar research was conducted by Marchelak and co-workers [61]. *Prunus Spinosa* flowers are known as traditional medicine for treatment of inflammation, urinary tract disorders, and cardiovascular diseases. Flowers of *P. spinosa* were subjected to the extraction in five different solvents. Up to 57 constituents, mostly phenolic acids, flavonoids, and proanthocyanidins were identified in these extracts. Ethyl acetate extract showed the strongest inhibition of LOX with IC₅₀ value of 135.36 µg/mL and had high polyphenol content of 584.07 mg GAE/g of dry weight. Total flavonoid amounts of extracts were in the range of 1.88 to 490.63 mg/g of dry weight.

11. Lipoxygenase Inhibition by Plant Extracts of Family Sapindaceae

*Paullinia pinnata* is a plant from family Sapindaceae, whose phytochemical profile showed the presence of numerous secondary metabolites [52]. Different plant parts showed various activities, such as antioxidant, antibacterial, anti diarrheal, anti-inflammatory, antityphoid, and anxiolytic properties. Ethanolic extracts (50% ethanol) of *P. pinnata* showed poor 15-lipoxygenase inhibitory potential.

Ferreres et al. extracted leaves and stem bark of *Allophylus africanus* with water [62]. Both extracts showed good LOX inhibitory activity with IC₅₀ values of 41.28 and 107.77 µg/mL for leaves and stem bark, respectively (Table 10). In aqueous leaf extracts, 30 flavones were identified, with the dominant ones being apigenin derivatives and luteolin derivatives. Stem bark was characterized by mono-C-glycosides-O-glycosylated and apigenin di-C-glycosides.

| Plant Species | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC₅₀ µg/mL) | References |
|---------------|---------------------|----------------------|------------------------|-----------------------------|------------|
| *Paullinia pinnata* L. | Ethanol (95%) | Leaves | - | - | [52] |
| *Allophylus africanus* P. Beauv. | Water | Leaves | - | 41.28 | [62] |
| | | Stem bark | - | 107.77 | |

12. Lipoxygenase Inhibition by Plant Extracts of Other Plant Families

Many authors reported inhibition of lipoxygenase enzymes by plant extracts from various plant families, as listed in Table 11. Plant extracts varied from poor to excellent LOX inhibitors.

Hexan seed extract of *Cannabis sativa* L. (Cannabaceae) showed poor LOX inhibition, lower than 50.0% [63]. Salleh et al. reported inhibition of LOX enzyme by *Beilschmiedia penangiana* (Lauraceae) leaves and stem bark extracts [64]. Plant parts were extracted in hexane, ethyl acetate, and methanol. Leaf extracts inhibited lipoxygenase for 39.8%, 45.8%, and 32.4% in hexane, ethyl acetate, and methanol, respectively. Stem bark also showed moderate inhibition of 24.2%, 41.5%, and 46.3%, respectively. Shafiri-Rad et al. reported LOX inhibition of 55.3% (methanolic extract of *Veronica persica* (Plantaginaceae) at concentration of 1 mg/mL [65]. Leaf extracts of *Kigelia Africana* (Bignoniaceae) showed a lipoxygenase inhibition at a concentration of 500 µg/mL [66]. Ethanolic, aqueous, butanol, ethyl acetate, and hexane fractions showed LOX inhibition of 91.7%, 85.8%, 84.8%, 91.6%, and 86.1%, respectively. Furthermore, methanol seed extract of *Nigella sativa* (Ranunculaceae) and ethanol extract of *Petiveria alliaceae* (Phytolaccaceae) showed LOX inhibition of 58.4% and 65.0%, at a concentration of 500 µg/mL, respectively [67,68]. *Archidium ohioense* (Archidiaceae) plant was extracted in four organic solvents and water [69]. Ethyl acetate and dichloromethane extract were the most active and showed inhibition against LOX of 75.0% and 71.6% at a concentration of 50 µg/mL, respectively.
### Table 11. Lipoxygenase inhibition by extracts of different plant families and species.

| Plant Family/Species               | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC\textsubscript{50} µg/mL) | References |
|-----------------------------------|---------------------|----------------------|------------------------|-------------------------------------------|------------|
| Cannabaceae/Cannabis sativa L.    | n-Hexane            | Seeds                |                        | -                                         | [63]       |
| Lauraceae/Beilschmiedia penangiana Gamble | n-Hexane | Leaves             | 101.50 \textsuperscript{3} | 66.80 \textsuperscript{3} | [64] |
|                                   | Ethyl acetate       |                      |                        | -                                         |            |
|                                   | Methanol            |                      |                        | 59.60 \textsuperscript{3}               |            |
|                                   | n-Hexane            | Stem bark            | 47.70 \textsuperscript{3} | -                                         |            |
|                                   | Ethyl acetate       |                      | 46.30 \textsuperscript{3} | -                                         |            |
|                                   | Methanol            |                      | 176.80 \textsuperscript{3} | -                                         |            |
| Plantaginaceae/Veronica persica Poir. | Methanol (70%)     | Leaves               |                        | -                                         | [65]       |
| Bignoniaceae/Kigelia Africana (Lam.) Benth. | Ethanol (70%) | Leaves             | 7.84 \textsuperscript{3} | -                                         | [66]       |
|                                   | n-Hexane            |                      | 5.73 \textsuperscript{3} | -                                         |            |
|                                   | Ethyl acetate       |                      | 22.55 \textsuperscript{3} | -                                         |            |
|                                   | n-Butanol           |                      | 18.77 \textsuperscript{3} | -                                         |            |
|                                   | Water               |                      | 12.67 \textsuperscript{3} | -                                         |            |
| Ranunculaceae/Nigella sativa L.   | Methanol            | Seeds                | -                      | -                                         | [67]       |
| Phytolaccaceae/Petiveria alliacea L. | Ethanol            | Whole                | -                      | -                                         | [68]       |
| Archidiaceae/Archidium ohioense Schimp. Ex Müll. Hal. | Dichloromethane | Whole                | -                      | -                                         | [69]       |
|                                   | Ethyl acetate       |                      | -                      | -                                         |            |
| Combretaceae/Terminalia chebula Retz. | Petroleum ether    | Leaves               | 107.00 \textsuperscript{4} | 51.00 \textsuperscript{4} | [70]       |
|                                   | Chloroform          |                      | 95.00 \textsuperscript{4} | 141.00 \textsuperscript{4}               |            |
|                                   | Water               |                      | -                      | -                                         |            |
| Moraceae/Ficus curtipes Corner.   | Methanol            | Leaves               | 200.80                 | 10.75                                     | [71]       |
| Oleaceae Fraxinus rhynchophylla Roxb. | Ethanol            | Leaves               | -                      | -                                         | [72]       |
|                                   |                     | Outer bark           | -                      | -                                         |            |
|                                   |                     | Endodermis           | -                      | -                                         |            |
| Rutaceae/Zanthoxylum armatum DC.  | Methanol            | Leaves               | 13.10 \textsuperscript{3} | -                                         | [73]       |
|                                   |                     | Bark                 | 15.50 \textsuperscript{3} | 90.50                                     |            |
|                                   |                     | Fruit                | 25.60 \textsuperscript{3} | 70.30                                     |            |
| Rhizophoraceae/Bruguiera cylindrica L. | Methanol           | Leaves               | -                      | -                                         | [74]       |
|                                   | Dichloromethane     |                      | -                      | 45.70                                     |            |
|                                   | Methanol            |                      | -                      | 71.00                                     |            |
|                                   | Dichloromethane     |                      | -                      | 64.00                                     |            |
| Areecaeae/Arenga Pinnata (Wurmb.) Merr. | Ethanol (95%)     | Endosperm            | -                      | 71.37                                     | [75]       |
| Myricaceae/Myrica rubra Siebold & Zucc. | Water              | Leaves               | 209.83                 | 50.57                                     | [76]       |
|                                   | n-Hexane            |                      | 18.41                  | -                                         |            |
|                                   | Ethyl acetate       |                      | 192.39                 | 8.30                                      |            |
|                                   | tert-Butylmethylether |                      | 379.50                | 32.32                                     |            |
| Oxalidaceae/Averrhoa carambola L. | 70% Ethanol         | Leaves               | -                      | 37.00 \textsuperscript{1}                | [77]       |
|                                   | Ethyl acetate       |                      | -                      | 7.84 \textsuperscript{1}                 |            |
|                                   | n-Hexane            |                      | -                      | 107.71 \textsuperscript{1}               |            |
|                                   | Water               |                      | -                      | 64.05 \textsuperscript{1}                |            |
| Piperaceae/Piper stylosum Miq.    | Methanol            | Aerial parts         | -                      | 80.50 \textsuperscript{2}                | [78]       |
|                                   | Ethyl acetate       |                      | -                      | 82.80 \textsuperscript{2}                |            |
|                                   | n-Hexane            |                      | -                      | 85.20 \textsuperscript{2}                |            |
Table 11. Cont.

| Plant Family/Species | Extraction Solvents | Plant Part Extracted | Total Phenolic Content | LOX Inhibition (IC50 µg/mL) | References |
|----------------------|----------------------|----------------------|------------------------|-----------------------------|------------|
| Asparagaceae/Polygonatum verticillatum All. | Methanol  
n-Hexane  
Chloroform  
Ethyl acetate  
n-Butanol  
Water | Aerial parts | - | 125.00 | [79] |
| Burseraceae/Boswellia dalzielii Hutch | Dichloromethane  
Ethyl acetate  
Methanol  
Cyclohexane | Leaves | 48.23 4 | 58.01 | [80] |
|  |  |  | 148.81 4 | 45.10 |  |
|  |  |  | 315.97 4 | 28.01 |  |
|  |  |  | 3.58 4 | - |  |
| Euphorbiaceae/Jatropha gossypifolia L. | Methanol  
n-Hexane  
Dichloromethane  
Ethyl acetate  
Butanol | Leaves | 162.50 | - | [81] |
|  |  |  | 57.10 | 58.50 |  |
|  |  |  | 59.40 |  |
| Malvaceae/Lavatera cretica L. | Water | Leaves | 254.62 4 | 0.01 | [82] |
|  |  |  | 191.38 4 | 0.01 |  |
|  |  |  | 189.60 4 | 0.15 |  |
|  |  |  | 43.69 4 | - |  |

1 results are expressed in ppm; 2 results expressed in µM; 3 results are expressed as mg gallic acid equivalent (GAE)/g of extract; 4 results are expressed as mg gallic acid equivalent (GAE)/g of dry mass.

Eshwarppa et al. employed four solvents (petroleum ether, chloroform, ethanol, and water) in the extraction of Terminalia chebula (Combretaceae) leaves [70]. Minimum inhibitory concentrations (MIC) were 560.0, 673.0, 726.0, and 847.0 µg/mL for ethanol, water, chloroform, and petroleum ether, respectively. Total phenolic content was in the range of 51.00 to 141.00 mg GAE/g of dry weight and total flavonoids in the range of 39.00 to 125.00 mg QE/g of dry weight.

Extraction of leaves and stem bark of Ficus curtipes (Moraceae) was carried out in methanol [71]. Stem extract showed strong inhibition against LOX with an IC50 value of 10.75 µg/mL, while leaf extract had significantly poorer IC50 value (IC50 = 62.60 µg/mL). Huh et al. extracted Fraxinus rhynchophylla (Oleaceae) leaves, outer bark, and endodermis in ethanol [72]. Only outer bark had LOX inhibitory activity with an IC50 value of 62.60 µg/mL. Furthermore, Alam et al. carried out extractions of Zanthoxylum armatum (Rutaceae) leaves, bark, and fruit in methanol [73]. Bark and fruit extracts showed very good inhibition of LOX, with IC50 values of 90.50 and 70.30 µg/mL. Fruit extracts had the higher phenolic amount (26.50 mg GAE/g of extract) than bark extracts (15.50 mg GAE/g of extract). Leaves and roots of Bruguiera cylindrica (L.) (Rhizophoraceae) were extracted in two organic solvents (methanol and dichloromethane) [74]. All extract showed LOX inhibition, except for the methanolic extracts of leaves. The best inhibition was accomplished in dichloromethane leaf extract (IC50 = 45.70 µg/mL), followed by dichloromethane root extracts (IC50 = 64.00 µg/mL) and methanol root extracts (IC50 = 71.0 µg/mL). Endosperm extracts of Arenga pinnata (Arecaceae) showed LOX inhibition of 71.37 µg/mL (IC50) [75].

Many authors reported extraction of plant materials in different organic solvents (ethanol, methanol, chloroform, dichloromethane, hexane, cyclohexane, ethyl acetate, butyl methyl ether) and water, which are tested on LOX inhibitory potential [76–81]. Langhansova et al. employed four solvents in the extraction of Myrica rubra (Myricaceae) leaves [76]. The best inhibition of lipoxygenase was achieved with the ethyl acetate extract (IC50 = 8.30 µg/mL). Extracts of a Averrhoa carambola L. (Oxalidaceae) and Polygonatum verticillatum (Asparagaceae) also showed the strongest activities against LOX in ethyl acetate with IC50 values of 7.84 and 97.0 µg/mL, respectively [77,79]. On the other hand, methano-
lic extracts of *Piper stylosum* (Piperaceae) and *Boswellia dalzielii* (Burseraceae) showed the best LOX inhibition with IC\textsubscript{50} values of 80.50 and 28.10 µg/mL, respectively [78,80]. Dichloromethane was proven the best extraction solvent in the case of *Jatropha gossypifolia* (Euphorbiaceae) [81]. With dichloromethane extracts, IC\textsubscript{50} value of 57.10 µg/mL was obtained for LOX inhibition.

LOX inhibition of water extract of *Lavatera cretica* L. (Malvaceae) should also be mentioned [82]. Leaf and flower extracts showed very strong inhibition of LOX, with IC\textsubscript{50} values of 0.01 µg/mL. Extracts had high polyphenolic content of 254.62 and 191.38 mg GAE/g dry weight.

13. Conclusions

This paper gives an overview of the effect of various plant extracts of various plant families on LOX inhibition. It turned out that soybean lipoxygenases, both 5-LOX and 15-LOX (found in mammals), were successfully inhibited by plant extracts. Ethanol extract of *Cnicus benedictus* showed great inhibition of 15-LOX with an IC\textsubscript{50} value of 52.7 µg/mL. The best inhibitor of 5-LOX was the ethanolic extract of *Sophora tokinensis* with an IC\textsubscript{50} value of 1.61 µg/mL. Soybean lipoxygenase was successfully inhibited by *Lavatera cretica* leaf and flower water extracts with an IC\textsubscript{50} value of 0.01 µg/mL.

Based on the abovementioned, it becomes clear that the selection of appropriate solvents for phytochemical extractions, as well as the corresponding phytochemical profiles, play important roles in LOX inhibition. While some of the extracts show excellent inhibitory activities, others seem poorly effective in LOX inhibition. Moreover, several reports shows greater LOX inhibitory activity of isolated compounds themselves, than extracts on the whole. Therefore, it seems that there is lot of space to fill in the gaps in regards to finding the most novel, prominent LOX inhibitors among the multitude of plant phytochemicals.

Author Contributions: Conceptualization, I.S., M.L. and M.M.; visualization, M.L., V.P. and M.M.; writing—original draft preparation, M.L., M.M. and I.S.; writing—review and editing, T.M., D.Š., M.M. and V.P.; supervision, M.M.; project administration, M.M.; funding acquisition, M.M. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported in part by the Croatian Science Foundation under the project “Green Technologies in Synthesis of Heterocyclic Compounds” (UIP-2017-05-6593).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is not applicable to this article.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Shi, Y.; Mandal, R.; Singh, A.; Pratap Singh, A. Legume lipoxygenase: Strategies for application in food industry. *Legume Sci.* 2020, 2, e44. [CrossRef]
2. Hayward, S.; Cilliers, T.; Swart, P. Lipoxygenases: From isolation to application. *Compr. Rev. Food Sci. Food Saf.* 2017, 16, 199–211. [CrossRef]
3. Chedea, V.S.; Jisaka, M. Lipoxygenase and carotenoids: A co-oxidation story. *Afr. J. Biotechnol.* 2013, 12, 2786–2791. [CrossRef]
4. Newcomer, M.E.; Brash, A.R. The structural basis for specificity in lipoxygenase catalysis. *Protein Sci.* 2015, 24, 298–309. [CrossRef]
5. Pallavi, P.C.; Singh, A.K.; Singh, S.; Singh, N.K. In Silico Structural and Functional Insights into the Lipoxygenase Enzyme of Legume Cajanus Cajan. *Int. J. Recent Innov. Trends Comput. Commun.* 2014, 5, 87–91.
6. Srivastava, P.; Vyas, V.K.; Variya, B.; Patel, P.; Qureshi, G.; Ghate, M. Synthesis, anti-inflammatory, analgesic, 5-lipoxygenase (5-LOX) inhibition activities, and molecular docking study of 7-substituted coumarin derivatives. *Bioorg. Chem.* 2016, 67, 130–138. [CrossRef]
7. Baysal, T.; Demirdöven, A. Lipoxygenase in fruits and vegetables: A review. *Enzym. Microb. Technol.* 2007, 40, 491–496. [CrossRef]
8. Lampi, A.M.; Yang, Z.; Mustonen, O.; Piironen, V. Potential of faba bean lipase and lipoxygenase to promote formation of volatile lipid oxidation products in food models. *Food Chem.* 2020, 311, 125982. [CrossRef]
35. Marlin, S.; Elya, B. Antioxidant Activity and Lipoxygenase Enzyme Inhibition Assay with Total Flavonoid Content from Garcinia hombroniiana Pierre Leaves. Pharmacogn. J. 2017, 9. [CrossRef]

36. Avelia, S.; Mauldina, M.G.; Elya, B. Antioxidant activity and lipoxygenase inhibitory assay with total flavonoid content from Garcinia lateriflora Blume leaves extract. Asian J. Pharm. Clin. Res. 2017, 10, 163–165. [CrossRef]

37. Putri, N.L.; Elya, B.; Purisitasari, N. Antioxidant activity and lipoxygenase inhibition test with total flavonoid content from Garcinia kydia Roxburgh leaves extract. Pharmacogn. J. 2017, 9. [CrossRef]

38. Rahman, A.H.; Parvin, M.I. Study of medicinal uses on Fabaceae family at Rajshahi, Bangladesh. Res. Plant Sci. 2014, 2, 6–8.

39. Souleymane, F.; Charlemagne, G.; Moussa, O.; Eloi, P.; Baptiste, N.J.; Pierre, G.I.; Jacques, S. DPPH radical scavenging and lipoxygenase inhibitory effects in extracts from Erythrina senegalensis (Fabaceae) DC. Afr. J. Pharm. Pharmacol. 2016, 10, 185–191.

40. Yoo, H.; Kang, M.; Pyo, S.; Chae, H.S.; Ryu, K.H.; Kim, J.; Chin, Y.W. SKI3301, a purified herbal extract from Sophora tonkinensis, Cydonia oblonga. [CrossRef]

41. Babatunde, J.O.; Kayode, O.K. Inhibitory action of dried leaf of Cassia alata (Linn.) Roxb against lipoxygenase activity and nitric oxide generation. Sci. Agric. 2019, 10, 185–190.

42. Hussain, T.; Gupta, S.; Adhami, V.M.; Mukhtar, H. Green tea constituent epigallocatechin- 3-gallate selectively inhibits COX-2 without affecting COX-1 expression in human prostate carcinoma cells. Int. J. Cancer 2005, 113, 660–669. [CrossRef]

43. Singh, B.; Nadkarni, J.R.; Vishwakarma, R.A.; Bharate, S.B.; Nivsarkar, M.; Anandjiwala, S. The hydroalcoholic extract of Cassia alata (Linn.) leaves and its major compound rhein exhibits anti-allergic activity via mast cell stabilization and lipoxygenase inhibition. J. Ethnopharmacol. 2012, 141, 469–473. [CrossRef]

44. Ouédraogo, N.; Hay, A.E.; Ouédraogo, J.C.W.; Sawadogo, W.R.; Tibiri, A.; Lombo, M.; Guissou, I.P. Biological and phytochemical investigations of extracts from Pterocarpus erinaceus Poir (Fabaceae) root barks. Afr. J. Tradit. Complement. Altern. Med. 2017, 14, 187–195.

45. Rajesh, A.; Doss, A.; Tresina, P.S.; Mohan, V.R. In-vitro anti-inflammatory activity of ethanol extract of Crotalaria longipes. J. Med. Plants Res. 2013, 7, 281–285. [CrossRef]

46. Raja, R.R. Medicinally potential plants of Labiatae (Lamiaceae) family: An overview. J. Ethnopharmacol. 2017, 192, 22–28. [CrossRef]

47. Göger, F.; Özek, G.; Tekin, M.; Yur, S.; Özek, T. Phytochemical profiling and evaluation of marrubium sivasense aytaç, akgül & ekici for antioxidant activity and inhibition effects on?-amylase, lipoxygenase, xanthine oxidase and tyrosinase enzymes. J. Turk. Chem. Soc. Sect. Chem. 2019, 8, 281–292.

48. Păduraru, A.F.; Cioancă, O.; Mircea, C.; Trifan, A.; Aprotoasaie, A.C.; Miron, A.; Hâncianu, M. Bioactive Extracts from Cultivated Ajuga Genevensis L. And A.; Reptans, L.: In Vitro/In Vivo Pharmacological Effects. Farmaciu 2019, 67, 603–609. [CrossRef]

49. Phosriithong, N.; Nuchtavnorn, N. Antioxidant and anti-inflammatory actives of Clerodendrum leaf extracts collected in Thailand. Eur. J. Integr. Med. 2016, 8, 281–285. [CrossRef]

50. Karuppawamy, S. Medicinal plants used by Paliyan tribes of Sirumalai hills of Southern India. Indian Nat. Prod. Res. 2007, 6, 436–442.

51. Sekhar, S.; Sampath Kumar, K.K.; Niranjan, S.R.; Prakash, H.S. In vitro antioxidant activity, lipoxygenase, cyclooxygenase-2 inhibition and DNA protection properties of Memecylon species. Int. J. Pharm. Pharm. Sci. 2013, 5, 257–262.

52. Afagnigni, A.D.; Nyegue, M.A.; Djova, S.V.; Etoa, F.X. LC-MS Analysis, 15-Lipoxygenase Inhibition, Cytotoxicity, and Genotoxicity of Dissotis multiflora (Sm) Triana (Melastomataceae) and Paullinia pinnata Linn (Sapindaceae). Int. J. Pharm. Pharm. Sci. 2017, 9, 436–442. [CrossRef]

53. Alam, F.; us Saqib, Q.N.; Ashraf, M. Gaultheria trichophylla (Royle): A source of minerals and biologically active molecules, its antioxidant and anti-lipoxygenase activities. BMC Complement. Altern. Med. 2017, 17, 3. [CrossRef] [PubMed]

54. Michel, P.; Owczarek, A.; Matczak, M.; Kosno, M.; Szymański, P.; Mickiucuk-Olasik, E.; Kilanowicz, A.; Wesolowski, W.; Olszewska, M.A. Metabolite profiling of eastern teaberry (Gaultheria procumbens L.) lipophilic leaf extracts with hyaluronidase and lipoxygenase inhibitory activity. Molecules 2017, 22, 412. [CrossRef] [PubMed]

55. Kumar, S.; Pandey, A.K. Chemistry and biological activities of flavonoids: An overview. Sci. World J. 2013, 2013. [CrossRef]

56. Choironi, N.A. Activity Inhibition of Lipoxygenase (LOX) by Tribe Zingiberaceae Plant Extracts and Isolation of Selected Plant Secondary Metabolites. Master’s Thesis of Pharmacy, Bandung Institute of Technology, Bandung, Indonesia, 2014. Unpublished.

57. Handayani, E.; Nuraini, P. Cyclea barbata leaf extract: Lipoxygenase inhibitory activity and phytochemical screening. Int. J. Pharm. Pharm. 2018, 10. [CrossRef]

58. Sinbad, O.O.; Adewale, A.; Adedoyin, O. Evaluation of membrane stabilizing, proteinase and lipoxygenase inhibitory activities of ethanol extract of root and stem of Sphenocentrum jollyanum Pierre. J. Adv. Biol. Biotechnol. 2017, 13, 1–8. [CrossRef]

59. Erturk, A.G.; Erturk, O.; Ayyaz, M.C.; Erturk, E.Y. Screening of Phytochemical, Antimicrobial and Antioxidant Activities in Extracts of Some Fruits and Vegetables Consumed in Turkey. CBU J. Sci. 2018, 14, 81–92. [CrossRef]

60. Berkoz, M. Antioxidant and anti-lipoxygenase activities of Cydonia oblonga. Medicine 2020, 9, 251–254. [CrossRef]

61. Marchelak, A.; Owczarek, A.; Matczak, M.; Pawlak, A.; Kołodziejezyk-Czepas, J.; Nowak, P.; Olszewska, M.A. Bioactivity potential of Prunus spinosa L. flower extracts: Phytochemical profiling, cellular safety, pro-inflammatory enzymes inhibition and protective effects against oxidative stress in vitro. Front. Pharmacol. 2017, 8, 680. [CrossRef]
62. Ferreres, F.; Gomes, N.G.; Valentão, P.; Pereira, D.M.; Gil-Izquierdo, A.; Araújo, L.; Silva, T.C.; Andrade, P.B. Leaves and stem bark from Allophlyllus africânus P. Beauv.: An approach to anti-inflammatory properties and characterization of their flavonoid profile. Food Chem. Toxicol. 2018, 118, 430–438. [CrossRef] [PubMed]

63. Jin, S.; Lee, M.Y. The ameliorative effect of hemp seed hexane extracts on the Propionibacterium acnes-induced inflammation and lipogenesis in sebocytes. PLoS ONE 2018, 13, e020933. [CrossRef]

64. Wan Salleh, W.M.N.H.; Ahmad, F.; Abdul Azziz, S.S.; Ahmad, M.S. In Vitro Pharmacological Evaluation of the Leaves and Stem Bark Extracts of Beilschmiedia penangalana Gamble. Jordan J. Pharm. Sci. 2019, 12, 47–57.

65. Sharifi-Rad, J.; Tayyebon, G.S.; Niknam, F.; Sharifi-Rad, M.; Mohajeri, M.; Salehi, B.; Iriti, M.; Sharifi-Rad, M. Veronica persica Poir. extract-antibacterial, antifungal and scabicialdical activities, and inhibitory potential on acetylcholinesterase, tyrosinase, lipoygenase and xanthine oxidase. Cell. Mol. Biol. 2018, 64, 50–56. [CrossRef] [PubMed]

66. Apalowo, O.E.; Adekola, M.B.; Asaolu, F.T.; Oryomi, O.V.; Ogunleye, G.S.; Areola, J.O.; Babaloba, O.O. GC-MS analysis and inhibitory effect of Kigelia africana Leaf extract and fractions on 5-lipoxygenase. J. Pharmacogn. Phytochem. 2019, 8, 625–628.

67. Ansari, M.I. A comparative anti-inflammatory and antioxidative potent of Nigella sativa seeds extract and its oil. Int. J. Green Pharm. 2019, 13. [CrossRef]

68. Rajesh, A.; Doss, A.; Tresina, P.S.; Mohan, V.R. Assessment of In vitro anti-inflammatory activity of ethanol extract of Petiveria allistae L. (Phytolaccaceae). Int. J. Bio-Pharma Res. 2019, 8, 2569–2574.

69. Akinpelu, B.A.; Godwin, A.; Aderogba, M.A.; Makinde, A.M.; Azeez, S.O.; Oziegbe, M. Evaluation of anti-inflammatory and genotoxicity potentials of the fractions of Archidium ohiense (Schimp. ex Mull) extract. Ije J. Sci. 2018, 20, 487–496. [CrossRef]

70. Eshwarappa, R.S.B.; Ramachandra, G.S.; Niknam, F.; Sharifi-Rad, M.; Mohajeri, M.; Salehi, B.; Iriti, M.; Sharifi-Rad, M. Veronica persica Poir. extract–antibacterial, antifungal and scabicialdical activities, and inhibitory potential on acetylcholinesterase, tyrosinase, lipoygenase and xanthine oxidase. Cell. Mol. Biol. 2018, 64, 50–56. [CrossRef] [PubMed]

71. Andrade, C.; Ferreres, F.; Gomes, N.G.; Duangsrissai, S.; Srisombat, N.; Vajrodaya, S.; Pereira, D.M.; Gil-Izquierdo, A.; Andrade, A.B.; Valentão, P. Phenolic Profiling and Biological Potential of Ficus curtipes Corner Leaves and Stem Bark: 5-Lipoxygenase Inhibition and Interference with NO Levels in LPS-Stimulated RAW 264.7 Macrophages. Biomolecules 2019, 9, 400. [CrossRef]

72. Huh, M.K.; Cho, K.S.; Jeon, S.J. Inhibitory effect of lipoygenase and dpph radical scavenging activity of Fraxinus rhynchophylla. Eur. J. Adv. Res. Biol. Life Sci. 2015, 3, 10–16.

73. Alam, F.; Ashraf, M. Phenolic contents, elemental analysis, antioxidant and lipoygenase inhibitory activities of Zanthoxylum armatum DC fruit, leaves and bark extracts. Pak. J. Pharm. Sci. 2019, 32, 1703–1708. [PubMed]

74. Eldeen, I.M.; Ringe, J.; Ismail, N. Inhibition of Pro-inflammatory Enzymes and Growth of an Induced Rheumatoid Arthritis Synovial Fibroblast by Bruguiera cylindrica. Pharmacogn. Mag. 2019, 15, 916–925. [CrossRef]

75. El-Fadil, M.; Elsabban, M.; El-Belamy, B.; Suleiman, M. The effect of d-α-tocopherol on 5-lipoxygenase activity of human neutrophils. Cell. Mol. Toxicol. 2006, 22, 67–72. [PubMed]

76. Langhansova, L.; Landa, P.; Kutil, Z.; Tauchen, J.; Marsik, P.; Rezek, J.; Lou, J.D.; Yun, Z.L.; Vanek, T. Myrica rubra leaves as a potential source of a dual 5-LOX/COX inhibitor. Food Sci. Biotechnol. 2018, 27, 101–109. [PubMed]

77. Nabilah Elya, B.; Djajadisastra, J. Lipoxygenase inhibitory assay of averrhoa carambola L. leaves extract. Int. J. Chemtech. Res. 2019, 10, 342–347.

78. Salleh, W.M.N.H.W.; Hashim, N.A.; Khamis, S. Chemical constituents and lipoygenase inhibitory activity of Piper stylosum Miq. Bull. Chem. Soc. Ethiop. 2019, 33, 587–592. [CrossRef]

79. Khan, H.; Saeed, M.; Muhammad, N.; Gaffar, R.; Gul, F.; Raziq, N. Lipoygenase and urease inhibition of the aerial parts of the Polygonatum verticillatum. Toxicol. Ind. Health 2013, 31, 758–763. [CrossRef]

80. Kohoude, M.J.; Gbaguidi, F.; Agbani, P.; Ayedoun, M.A.; Cazaux, S.; Bouajila, J. Chemical composition and biological activities of extracts and essential oil of Boswellia dalzielii leaves. Pharmacogn. Mag. 2017, 55, 33–42. [CrossRef]

81. Saleem, H.; Ahmad, I.; Gill, M.S.A. Evaluation of lipoygenase inhibition of Jatropha gossypifolia, a medicinal plant from Pakistan. Bangladesh J. Pharmacol. 2016, 11, 319–320. [CrossRef]

82. Ben-Nasr, S.; Aazza, S.; Mnif, W.; Miguel, M.D. Antioxidant and anti-lipoxygenase activities of extracts from different parts of Lavatera cretica L. grown in Algarve (Portugal). Pharmacogn. Mag. 2015, 11, 48. [PubMed]