The Application of a Split-Conjugated Anti-Brucellosis Vaccine as a “Provocing Factor” for Identification of Brucellosis in Animals

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**Abstract** | The split-conjugated vaccine against brucellosis of animals was tested as a provocateur in order to identify sick (latent) animals in both brucellosis safe and brucellosis poor farms. After administration of the test vaccine, antibodies remained in the blood of healthy animals for up to 2.5 months, then disappeared, and antibodies remained in the blood of sick animals after 3.5, 4.5 and 7.5 months and gradually disappearing over time. The split-conjugated animal brucellosis vaccine has a “provocative effect”, exacerbates the infectious process and thereby makes it possible to identify sick animals. We recommend the use of a split-conjugated animal brucellosis vaccine in brucellosis-poor households in order to detect latent animals and create humoral immunity in animals while maintaining antibodies in the blood of animals for 2.5 months. In the case of small detection of animals with brucellosis in the farm (from 1 to 5 animals), we recommend using the vaccine to detect animals with brucellosis and to create immunity in healthy animals twice with an interval of 3 months. If a large number of animals with brucellosis in the farm (from 6 to 20 and above goals) are detected, we recommend using the vaccine once after eliminating the sick livestock, followed by the use of the vaccine from the Brucella abortus 82 strain, which must be vaccinated after passing calving (May - July). The split-conjugated animal brucellosis vaccine can be used on both non-pregnant and pregnant cows, as it does not cause abortion.

**Keywords** | Animal brucellosis, Split-conjugated vaccine, Latent brucellosis, Exacerbation of the brucellosis infection process.

INTRODUCTION

Brucella as an optional intracellular pathogen establishes a close relationship with the host’s immune cells. Through the disruptive function of the immune system, the pathogen is able to support a chronic infection, which often makes treatment and diagnostics difficult. In recent decades, many studies have been conducted in an attempt to develop safe and effective animal brucellosis vaccines. There is no licensed vaccine for the prevention of human brucellosis. A human vaccine would be useful in protecting farmers, veterinarians, animal care workers, laboratory staff and the general population living in endemic areas of brucellosis (Dornless et al., 2015).

Studies aimed at developing the ideal vaccine against brucellosis in animals and humans have been being carried out since the beginning of the twentieth century (Blasco et al., 2016).

Since then, live as well as inactivated vaccines have been developed. Vaccination is the best economical measure to control brucellosis in endemic areas. Many countries have developed control measures to eradicate the disease in an-
Currently, vaccination programs are based on the control of brucellosis mainly caused by *B. melitensis* and *B. abortus* (Ductoroy et al., 2014).

In foreign countries, it is recommended to use only three live attenuated vaccines to fight infection caused by *B. abortus* in cattle: *B. abortus* 45/20, strain *B. abortus* 19 (S19) and *B. abortus* RB51 (Mansoori et al., 2016). *B. abortus* strain 45/20 was isolated after 20 passages (sequential reseeding of microorganism on various nutrient media or inoculation of pathogenic microbes from one infected animal to another) in guinea pigs. This R-strain form (rough strain) is used only as a thermally killed vaccine to avoid reversion to the virulent strain. This vaccine can be administered with an adjuvant to adult cattle. It does not interfere with serological diagnostics, and is safe for use in pregnant animals. A positive result in test mode was obtained in some countries (Khan and Zahoor, 2018; Li et al., 2017).

Another licensed live attenuated bovine brucellosis vaccine is *B. abortus* S19. This vaccine was isolated at the beginning of the twentieth century and subjected to attenuation by room temperature for one year [32]. *B. abortus* S19 is effective in controlling bovine brucellosis (prevents abortion) and also reduces the prevalence of *brucella* in herds. However, due to the characteristic features of strain S19 and a strong antibody response against the O-side chain, it does not distinguish between infected and vaccinated animals. Competitive ELISA and radial immunodiffusion tests were used to allow differentiation between vaccinated or infected animals (Godfroid, 2017).

A low rate of abortion in livestock production and a significant increase in milk production during vaccination with *B. abortus* S19 have been reported (Figueredo et al., 2015). Another live vaccine from strain *B. abortus* RB 51 is a spontaneous “rough” mutant obtained by subculturing a virulent strain of *B. abortus* 2308 on a medium containing rifampicin and penicillin [31]. Unlike strain 45/20, *B. abortus* RB 51 is very stable and is currently used in many countries instead of *B. abortus* S19. Rough deformation of RB51 is less virulent and does not cause a positive response in a typical serological diagnostic test (Saez et al., 2014).

Vaccination of pregnant cows with strain RB 51 can cause low abortion rates (less than 0.2%); however, it is safe at lower doses during pregnancy. The vaccine strain RB 51 can infect humans, but it is less virulent than strain S 19 (Saez et al., 2014).

Another live vaccine *B. melitensis* Rev. 1 is used to control brucellosis in small ruminants. This strain was developed by Herzberg and Elberg in the mid-1950s. The strain retains the general characteristics of *Brucella* species; it is resistant to streptomycin 2.5 μg / ml and susceptible to penicillin G, which makes it possible to differentiate from field strains [18]. Subcutaneous or conjunctival immunization with *B. melitensis* Rev. 1 provides sufficient immunity in small ruminants. *B. melitensis* Rev. 1 induces a positive antibody response in serological tests in vaccinated animals. However, *B. melitensis* Rev. 1 can infect people. Vaccination is recommended before the first pregnancy in 3 to 7 months to avoid abortion in pregnant animals.

Since *B. melitensis* can be isolated from cattle, some scientists have suggested using the live attenuated vaccine *B. melitensis* Rev. 1 for controlling the disease in cattle (Lopez-Santiago et al., 2019).

Compared to extensive research efforts to develop new vaccines against *B. melitensis* or *B. abortus*, a small study was conducted to protect pigs against *B. suis*. Oral or intramuscular vaccination with strain RB 51 killed by *B. suis* or purified with O-polysaccharide has been reported to be effective in protecting pigs infected by boars (Ivanov NP, 2006).

However, other studies have shown very little protection with strain RB 51 in immunized pigs against infection with the field strain (Mansoori et al., 2016; Li et al., 2017).

For the first time, immunization of animals with a vaccine from *B. abortus* 19S strain in 1953 was included in the system of anti-brucellosis measures of the USSR. This vaccine was used until 1970. During the application of the vaccine many households, and even entire areas, were cured of the disease (Hosein et al., 2018). However, in regions with a wide spread of brucellosis, the effectiveness of recreational activities was insufficient. This was primarily due to the high agglutinogenicity of the vaccine. Agglutinins and complement-binding antibodies remain in the body of immunized animals up to 5-8 years, which greatly complicates the differentiation of such animals from patients with brucellosis.

Since 1974, in Russia, vaccines from *B. abortus* strains 82 and 19 have been used in the general complex of anti-brucellosis measures. Since 1997 vaccines from *B. abortus* 75/79-AB and 17/100 strains were also used (Sklyarov et al., 2011).

Ivanov AV et al. (2009) found that in the fight against cattle brucellosis, the most effective vaccine strains are in the SR form, with weak agglutinogenic and pronounced immunogenic properties. The vaccine from the *Brucella abortus* 82 strain possesses all these properties (Ivanov AV...
A vaccine from the strain of *Brucella abortus* KB 17/100 was successfully tested for immunogenicity, and its prototype from the strain of *Brucella abortus* 45/20 is widely used in the countries of the Near and Middle East, Asia and Africa. Its main drawback is pronounced reactogenicity, its advantages are inagglutinogenicity, pronounced immunogenicity and the fact that in animals with a latent form of brucellosis it provokes the synthesis of antibodies detected by serological studies with standard antigens, which allows animals to be removed from the herd in a short time (Sklyarov et al., 2011).

Other researchers used an adjuvant, a vaccine against cattle brucellosis from the strain Brucella abortus KB 17/100. In a controlled epizootic experiment, the high epizootic efficacy of inactivated non-agglutinogenic adjuvant, a vaccine from the Brucella abortus strain KB 17/100, was established during the recovery of farms with cattle brucellosis (Khairov et al., 2005).

Unfortunately, the drug has reactogenic properties, manifested by a short-term increase in body temperature, swelling at the injection site and slight depression with a partial loss of appetite. All these signs gradually disappear, without harming the health of animals and without affecting their productivity. The introduction of adjuvant vaccines causes specific sensitization of the body of immunized animals. Due to the prospects of using the adjuvant vaccine from the strain *Brucella abortus* KB 17/100 as part of complex measures to combat cattle brucellosis, an adjuvant vaccine was used to immunize all cows and 3-6 month old heifers in 1998 in 5 districts of the Saratov region.

It has long been known that many vaccines have not only an immunostimulating effect, forming immunity, but also excite the whole animal and human organism (Hull and Schumaker, 2018), as a result of which antibodies from latent animals are released into the blood. Such action of vaccines is able to detect and thereby quickly eliminate many infectious diseases, including animal brucellosis. It is called a provocative action in the fight against infectious diseases (Novitsky, 2016). The elimination of brucellosis actually takes a lot of time, and often stretches over years, especially in places where the fight against brucellosis has not been carried out for a long time (Ivanov NP, 2006). There were cases when a double negative result for brucellosis took several years, which made it impossible to use vaccines in the fight against brucellosis, and contributed to the further spread of the infection. In addition, a long procedure for the elimination of brucellosis requires certain financial costs, patience and labor from both the veterinary services and the owners of animals (The concept of controlling the risks of the occurrence and spread of epizootic foci of zoonanthroponosis: Method. Regulations, 2011).

So, the goal of research was to evaluate the “provocative” effect of the split-conjugated brucellosis vaccine when used in a brucellosis-poor farm.

**MATERIALS AND METHODS**

Serological studies of blood samples of vaccinated animals were carried out in the Republic of Kazakhstan, the Akto-be region, the Mugalzhar district in the farm «Kustabaev». 39 heads of cows aged 3 to 6 years were used for the experiment.

The split-conjugated animal brucellosis vaccine was administered subcutaneously in the middle third of the neck at a dose of 1.0 ml. Before use, the vaccine was mixed with a solvent (immunoprotector). The immunoprotector polypeptide-C was obtained from activated calf spleen cells, consisting of a mixture of peptide activators of B cells, and a technique was developed for its conjugation with brucella antigens. The injection site was trimmed and pretreated with a 70% alcohol solution.

Blood was examined in the district veterinary laboratory of the Mugalzhar region in the agglutination reaction (AR), rosbengal test (RBT) and complement fixation reaction (CFR). Assessment of the results of serological studies was carried out according to the «Instructions for the diagnosis of animal brucellosis” issued in September 29, 2003 No. 13–5–02 / 0850 [10].

Blood from vaccinated calves of 6 months of age, owned by Berezovskoye Ltd. in the Engels district of the Saratov region, was taken 15, 30 and 90 days after immunization (a brucellosis-free farm).

Blood tests for brucellosis were performed using the complement fixation reaction (CFR) and agglutination reaction (AR) in the Engels veterinary laboratory (Saratov region) for combating animal diseases.

Results were analyzed after 3,5, 4,5 and 7 months after vaccine injection.

Blood from vaccinated calves of 6 months of age, owned by Berezovskoye Ltd. in the Engels district of the Saratov region, was taken 15, 30 and 90 days after immunization (a brucellosis-free farm).

Before vaccination, all animals underwent serological studies for brucellosis in AR and CFR. The results of preliminary studies were negative.

To verify the data obtained Student t-test was used, and the statistical significance level of cow parameters was
Experimental research, maintenance, care and euthanasia were carried out according to the requirements of the «European Convention for the Protection of Vertebrates used for experiments and other scientific purposes» (1986).

RESULTS

The change in the number of animals that respond positively to brucellosis in AR and CFR during a triple study of blood samples after vaccination in a brucellosis-friendly farm is shown in Table 1.

| Table 1: Change in the number of animals that respond positively to brucellosis in AR and CFR during a triple study of blood samples after vaccination |
|---|---|---|---|
| After 14 days | After 30 days | After 2.5 months |
| AR | CFR | RBT |
| 200 | 100 | 0 |

A month after vaccination a blood test by serological reactions revealed that almost all blood samples in CFRs lack complement-binding antibodies. Agglutinins were present in the vast majority of cases in a titer of 1:100, sometimes 1:50, which is considered a dubious reaction. In only one sample in the CFR, questionable results were found (1/5). This indicates that complement-binding antibodies in the blood circulate for a short time, and after a month disappear from the blood.

When examining blood samples 2.5 months after vaccination, it was found that negative results for brucellosis were obtained in both AR and CFR. Only one test using CFR was doubtful (1/5), all the rest were negative (Veselovsky et al., 2018).

The results of serological studies of blood samples of cattle after vaccination with a test vaccine in a dysfunctional farm are presented in Table 2.

Table 2: Serological studies of blood samples of cattle after vaccination with a test vaccine in a dysfunctional farm

| Results after vaccination | Animals examined, heads | Positive reaction - heads |
|---|---|---|
| AR | CFR | RBT |
| After 3.5 months | 39 | 31/79.5 | 22/56.4 | 29/74.4 |
| After 4.5 months | 32 | 20/62.5 | 16/50 | 18/56.3 |
| After 7 months | 33 | 17/51.5 | 18/54.5 | 15/45.5 |

Table 3: Dynamics of a positive reaction to cattle brucellosis after vaccination in the study of blood samples by various serological reactions %

Table 3 shows the dynamics of changes in the number of animals with brucellosis using various serological reactions. In CFR, there was a fluctuation in animals that respond positively to brucellosis (56.4%), then a decrease in such animals to 50%, and then again an increase to 54.6% in
RBT, a gradual decrease in animals responding positively to brucellosis is from 74.4% to 45.5%. In AR, the number of brucellosis-infected animals gradually decreased from 79.5% to 51.6% over time.

**DISCUSSION**

Dzhupina S.I. determined that in farms poor for brucellosis, many sick animals are not detected by generally accepted serological reactions. Such animals are involved in the spread of infection and the maintenance of epizootic situation, both of the farm itself and of the farms located nearby (Dzhupina et al., 2013). We agree with the opinion of Dzhupina S.I., since latent forms of brucellosis can release the causative agent of the disease into the environment and thereby infect new animals.

Other researchers recognized this feature as well. Having analyzed research works on small doses vaccination of cattle with the vaccine from strain 19, P.S. Ulasevich and V.A. Romakhov (Ulasevich and Romakhov, 1983) noted that this method allowed to identify hidden carriers of the causative agent of brucellosis using serological studies carried out after vaccination of animals. In our case, small doses of the vaccine provoked the appearance of antibodies in the blood of animals and thereby found latent animals.

A.S. Mangazeeva together with I.A. Kosilov, as well as our experiment, prove that in herds with a natural chronic course of brucellosis, the secondary administration of antigens to hidden carriers of the infection pathogen provokes the synthesis of antibodies captured by antigens in AR and CFR (Kosilov et al, 1999; Mangazeva, 1976) The same results were obtained by V.I. Kudla (Kudla, 1987) during the study of ram infectious epididymitis. Double administration of the vaccine we tested 3 months after the first injection will also allow to identify more animals with brucellosis, since repeated administration of the vaccine additionally stimulates the release of the pathogen of brucellosis into the bloodstream and, as a result, the formation of anti-brucellosis antibodies. Also, adjuvants, such as calcium hydroxyapatite, together with the vaccine, extended the immunity in animals, as shown in our previous experiments (Agoltsov et al., 2019).

Novitsky A.A. studied the antiepizootic efficacy of the combined use of brucellosis chemical vaccine (BCV) and vaccine from strain B. abortus 82 when stopping the focus of brucellosis infection. He proved that the provocative property of BCV allowed to additionally isolate animals positively responding to brucellosis on 21 day after its use. Further studies of the livestock after 2 and 3 months showed a marked decrease in epizootic tension. Three months later, animals with brucellosis infections were no longer recorded. Thus, as well as in our experiment, it was possible in a shorter time to heal the farm from cattle brucellosis (Novitsky, 2016).

**CONCLUSIONS**

1. After administration of the test vaccine, antibodies remain in the blood of healthy animals for up to 2.5 months, and then disappear, and antibodies are detected in the blood of animals that respond positively to brucellosis after 3.5 months and gradually their number decreases.

2. The split-conjugated animal brucellosis vaccine has a “provocative effect”, exacerbates the infectious process, and thereby makes it possible to identify sick animals.

Recommendations.

The use of a split-conjugated animal brucellosis vaccine in farms with dysfunctional brucellosis is recommended in order to:

1. Detect latent (patients with a latent form of brucellosis) animals for immediate vaccine administration after the identification (for cattle). 14 days after the introduction of the vaccine, blood should be examined for brucellosis by all of the above mentioned reactions. If positively reacting animals are detected, record the inventory numbers and antibody titers of such animals and again examine them with serological reactions after 30 days. Compare the results of serological blood tests with previous ones; record the inventory numbers of animals and antibody titers. Examine the blood of all animals in the farm after 2.5 months for brucellosis by serological reactions, including RBP. Consider animals positively responding to brucellosis 2.5 months after vaccination as sick and eliminate them within 15 days. If necessary and if suspected of still having brucellosis-infected animals on the farm, re-vaccination with a split-conjugated animal brucellosis vaccine should be carried out and a triple blood test of all farm animals for brucellosis should be repeated.

2. To create the humoral immunity of the animal organism to the administered vaccine.

In the case of small detection of animals with brucellosis in the farm (from 1 to 5 animals), we recommend using the vaccine both to identify animals with brucellosis and to create immunity for healthy animals. The scheme is as follows: after isolation of positively reacting animals, they should be eliminated within 15 days, and the entire cattle stock of the farm should be vaccinated with a split-conjugated animal brucellosis vaccine. 2.5 months after vaccination, re-examine the blood of healthy animals for brucellosis with serological reactions. Consider positively reacting animals as patients with brucellosis and eliminate them within 15 days. Re-examine the blood of farm animals after 14 days. Upon receipt of a negative result for brucellosis, re-vaccinate all animals with a vaccine. After 3
months, again examine the blood of animals for brucellosis twice with an interval of 14 days. By this time, animals with brucellosis should not remain on the farm and the vaccine, when applied twice, will form a humoral immunity for brucellosis in animals, associated with the formation of specific antibodies and the creation of immune memory in animals. In order to prevent brucellosis, the vaccine should be used every 6 months.

In case of large detection of sick animals (from 6 to 20 and above) in the farm, the vaccine is recommended to use once after eliminating the sick livestock.

The scheme is as follows: 2.5 months after vaccination, re-examine the blood of healthy animals for brucellosis with serological reactions. Consider positively reacting animals as patients with brucellosis and eliminate them within 15 days. Re-examine the blood of animals in the farm after 14 days and, if necessary, every 2 weeks until a double negative result for brucellosis is obtained. The purpose of the vaccine: to detect and eliminate the sick (latent) animals in the shortest possible time. After the identification and elimination of all animals suffering from brucellosis, the whole cattle of the farm should be inoculated with the vaccine from the strain *Brucella abortus* 82 from the moment of calving (May - July). Examine blood according to the instructions for brucellosis after 10 months. The split-conjugated animal brucellosis vaccine can be administered to animals at any time of the year because it does not have abortogenic properties.

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CONFLICT OF INTEREST

The authors have not declared any conflict of interests.

AUTHORS CONTRIBUTION

Valery Alexandrovich Agoltsov, Olga Mikhailovna Popova designed the experiment. Stepan Yurievich Veselovskiy conducted the statistical analysis, as well as conducting the experimental measurement, Davud Abdusamedovich Devrishov and Nataliya Victorovna Solotova contributed by writing the article and reviewing it thoroughly before submission.

REFERENCES

- Agoltsov VA, Popova OM, Veselovsky SY, Chastov AA, Semivolos AM, Solotova NV (2019). Results of Pre-Clinical and Clinical Tests of Organic Hydroxyapatite as Adjuvant of Bacterial Vaccine. Adv. Anim. Vet. Sci. 7(7): 583-592. https://doi.org/10.17582/journal.avs/2019/7.5.583.592
- Blasco JM, Moreno E., Moriyón I. (2016) Brucellosis vaccines and vaccine candidates. Veterinary Vaccines for Developing Countries. Chapter 5I. – Rome. – 35p.
- Caminiti A, Pelone F, Battisti S, Colafrancesco R, Sala M, La Torre G, Della Marta U, Scaramozzino P (2016). Tuberculosis, Brucellosis and Leucosis in Cattle: A Cost Description of Eradication Programmes in the Region of Lazio, Italy. Transbound. Emerg. Dis. - 206. https://doi.org/10.1111/tbed.12540
- Carvalho TF, Haddad JPA, Santos RL (2016) Meta-Analysis and Advancement of Brucellosis Vaccinology. PLoS ONE.11 (11): 1-28. https://doi.org/10.1371/journal.pone.0166582
- Dorneles EM, Sranganganathan N, Lage AP (2015). Recent advances in Brucella abortus vaccines. Vet. Res. 46(6). https://doi.org/10.1186/s13567-015-0199-7
- Ducrototy MJ, Berti WJ, Ocholi RA, Gusi AM, Bryssinckws K, Welburn S, Moriyón I (2014). Brucellosis as a new threat in developing countries: lessons from Nigeria. Plos. Negl. Trop. Tesis. 8: 3008. https://doi.org/10.1371/journal.pntd.0003008
- Dzhupina SI (2013). The epizootic process of bovine brucellosis and prospects for the devastation of its pathogen. Vet. Pathol. - No. 4: 97-105.
- Figueiredo de P, Ficht TA, Rice-Ficht A, Rossetti CA, Adams LG (2015). Pathogenesis and immunobiology of brucellosis: review of Brucella-host interactions. Am. J. Pathol. 185: 1505–1517. https://doi.org/10.1016/j.ajpath.2015.03.003
- Godfroid J (2017). Brucellosis in livestock and wildlife: zoonotic diseases without pandemic potential in need of innovative one health approaches. Arch Pub. Health. 75(34). https://doi.org/10.1186/s13690-017-0207-7
- Hosein HI, Zaki HM, Safwat NM, Menshawy AMS, Rouby S, Mahrous A, Madkour BE (2018). Evaluation of the General Organization of Veterinary Services control program of animal brucellosis in Egypt: An outbreak investigation of brucellosis in buffalo. Vet. World. 11(6): 748-757. https://doi.org/10.14202/vetworld.2018.748-757
- Hull NC, Schumaker BA (2018). Comparisons of brucellosis between human and veterinary medicine. Infect. Ecol. Epidemiol. 8: 12. https://doi.org/10.1080/20008686.2018.1500846
- Ivanov AV, Salmakov KM, Fomin AM, Chernov AN (2009). The state and prospects of finding a new generation of vaccines for animal brucellosis. Veterinarian. 6: 9-12.
- Ivanov AV, Yusupov RK, Salmakov KM (2009). The search for the most effective live and gamma-inactivated anti-brucellosis vaccines for small cattle. Vet. Doc. 4:19-22.
- Ivanov AV (2009). Immunological properties of vaccine strains of Brucella abortus. Vet. Med. 3: 27 - 29.
- Ivanov NP (2006). Brucellosis of animals: methods and means of dealing with it. - Almaty. Pp 506.
- Khairov SR, Yusupov OY, Shumilov KV, Klimanov AI (2005). The reaction of indirect hemagglutination in cattle brucellosis. Vet. Med. 5: 25-26.
- Khan MZ, Zahoor M (2018). An Overview of Brucellosis in...
Cattle and Humans, and its Serological and Molecular Diagnosis in Control Strategies. Trop. Med. Infect. Dis. 3(65): 14. https://doi.org/10.3390/tropicalmed3020065

- Kosilov IA, Arakelyan PA, Dimov SK, Khlystunov AG (1999). Brucellosis of farm animals. Monograph. Ed. I.A. Kosilov. - Novosibirsk. Pp 343.

- Kudla VI (1987). Antiepizootic effectiveness of various means and methods for diagnosing infectious sheep epididymitis. Abstract of thesis. - Novosibirsk. - 19p.

- Li MT, Sun GQ, Zhang WY, Jin Z (2017). Model-Based Evaluation of Strategies to Control Brucellosis in China. Int J Environ Res Public Health. 14(3): 295. https://doi.org/10.3390/ijerph14030295

- López-Santiago R, Sánchez-Argáez AB, De Alba-Núñez LG, Baltierra-Urbel SL and Moreno-Lafont MC (2019). Immune Response to Mucosal Brucella Infection. Front Immunol. 10. Article 1759. https://doi.org/10.3389/fimmu.2019.01759

- Mangazeyeva AS (1976). The secondary immune response in cattle with the use of certain anti-brucellosis vaccines. Abstract of thesis. Frunze city, Pp 20.

- Mansoori N, Pourmand MR (2016). Vaccines and Vaccine Candidates against Brucellosis. Infect. Epidemiol. Med. 2(4): 32-36. https://doi.org/10.18869/acadpub.iem.2.4.32

- Manual on the diagnosis of animal brucellosis of September 29, 2003 No. 13-5-02 / 0850.

- Novitsky AA (2016). Using the provocative properties of a chemical vaccine to detect latent forms of brucellosis. Modern problems and scientific support for the development of animal husbandry. Proceedings of the international scientific conference at Omsk State Agrarian University named after P.A. Stolypin. 153-159.

- Saenz JL, Sanz C, Durán M, García P, Fernandez F, Mínguez O, Carbajo L, Mardones F, Perez A, Gonzalez S, Domínguez L, Alvarez J (2014). Comparison of depopulation and S19-RB51 vaccination strategies for control of bovine brucellosis in high prevalence areas. Vet. Rec. https://doi.org/10.1136/ vtr.101979

- Sklyarov OD, Shumilov KV, Klimanov AI, Zinova AA, Bukova NK, Loginov IA. (2011). Ways to solve problems that determine the relevance of brucellosis. Vet. Med. - No.1: 34-38.

- The concept of controlling the risks of the occurrence and spread of epizootic foci of zooanthroponosis: Method. Regulations. (2011). SB Russian Agricultural Academy. - Novosibirsk, - 21 p.

- Ulasevich PS, Romakhov VA (1983). Prospects for vaccination of cattle against brucellosis with small doses of vaccines. Agric. Abroad. 6:47 - 50.

- Veselovsky SY, Agol'sov AV, Popova OM, Devrishov DA (2018). The experimental use of the split-conjugated animal brucellosis vaccine using the polyoxidonium immunomodulatory. Scient. Life - Moscow. 2: 89 - 100.