The ameliorative effects of exogenously applied proline on physiological and biochemical parameters of wheat (Triticum aestivum L.) crop under copper stress condition

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ABSTRACT

The presence of significant amount of heavy metals in rivers and canals due to mixing of untreated industrial effluents is a common phenomenon, especially in developing countries. The agricultural crops are influenced by the presence of various pollutants in the sewage, being applied for irrigation purpose. The effluents containing copper affect the growth and development of crop species, thereby, ought to be mitigated by foliar spray of osmoprotectants, e.g., proline. A pot culture experiment was conducted at the University of Agriculture, Faisalabad-Pakistan during the crop season 2015–2016. The treatments consisted of (a) three wheat varieties (Punjab-96, MH-97, FSD-83), (b) two levels of copper (0, 400 µM) applied through rooting medium, and (c) two levels of proline (0, 80 mM) applied through foliar application. The treatments were arranged in a completely randomized design with four replications. The results showed that application of 400 µM copper caused a reduction in biomass accumulation, chlorophyll (‘a’ and ‘b’) contents, and eventually yield (100-grain weight). There were also significant decreases in gas exchange parameters (stomatal conductance, internal CO2 concentration), photosynthetic rate, water-use-efficiency, and transpiration rate in response to copper stress. Metal toxicity caused the maximum reduction in productivity of PSII, electron transport rate, and photochemical quenching, while higher values of non-photochemical quenching were recorded in the wheat varieties. The activities of antioxidant enzymes (superoxide dismutase, catalase, peroxidase), as well as quantities of proline, protein and calcium contents were accelerated in response to copper stress. The uptake of calcium, magnesium, and potassium constituents by plants was reduced, while assimilation of calcium was increased in plants under copper stress. However, the occurrence of negative effects on these parameters due to copper stress was mitigated by foliar spray of proline at the rate of 80 mM solution. The exogenous application of proline at the rate of 80 mM resulted in the reduction of generation of reactive oxygen species and enhanced accumulation of proline and protein contents in wheat varieties under copper stress environment.

Introduction

The production of agricultural crops is significantly influenced by the presence of various pollutants in the sewage, being applied for irrigation purposes (Yadav 2010). The accumulation of heavy metals in the surface soil and their uptake by the plants has caused serious concern related to food safety for humans and livestock (Rehman et al. 2011). The agricultural production areas contaminated with heavy metals (i.e., copper, cadmium, and nickel) have posed serious implications to sustain crop production (Singh et al. 2007). The farming communities in the vicinity of the industrial zones have resorted to use municipal and industrial waste as a source of irrigation water, considering it as a highly nutritious in nature for raising their crops (Jamal et al. 2002; Ahmad et al. 2013). On the other hand, these effluents are highly toxic for the growth of plants, because of containing excessive amounts of copper, lead, zinc, and nickel (Younas et al. 1998; Jamal et al. 2002). Among these heavy metals, copper is the most obnoxious metal which imposes health hazards to both humans and livestock through food and fodder chain (Houshm and Moraghebi 2011).

Copper (Cu) as a plant nutrient is involved in the maintenance of a number of enzymes (Hall and Williams 2003) and as a co-factor in protein and enzymes (Yurekli and Porgali 2006; Shar et al. 2011). However, its excessive concentration is a rhizosphere causes oxidative stress and reduces the activities, e.g., physiological, photosynthesis and growth of plants (Adrees et al. 2015). The higher content of Cu in the root zone causes reduction in seed germination (Ouzounidou et al. 1992); imbalance mineral concentration in plant system (Ke et al. 2007); photosynthesis (Nussbaum et al. 1988); Chlorophyll contents (Zengin and Kirbag 2007); non-proliferation of roots (Sheeldon and Menzies 2005); inhibition of plant growth (Rehman and Iqbal 2006); reduction in uptake of essential nutrients by roots (Michaud et al. 2008); and in certain cases, the death of plants. The higher concentration of Cu caused oxidative stress through generation of reactive oxygen species (ROS), such as hydroxyl radical, superoxide radicals and hydrogen peroxide, which disintegrated the cell membranes, and biological molecules through lipid peroxidation (Hall 2002; Mittler et al. 2004). The cumulative effects resulted in leaf chlorosis, reduced plant growth and yield (Chen et al. 2007). The negative effects...
could be augmented by improvements in production of antioxidants enzymes and proteins (Imlay 2003; Brahim and Mohamed 2011). The antioxidant defense systems, such as superoxide (SOD) and catalase (CAT), act as scavengers of toxic radicals and adapt to production of ROS. The SOD induces the exchange of superoxide anions to water and hydrogen peroxide, while CAT decomposes hydrogen peroxide (Xu et al. 2006; Frary et al. 2010).

The plants resort to defense mechanism through enhancing the process of osmoregulation (Szabados and Savoure 2010). The osmolytes maintain structure of proline and photosynthetic apparatus and detoxify ROS through cellular osmoregulation in response to abiotic stresses (Ashraf and Foolad 2007). The adverse effects caused by copper stress could be alleviated by foliar spray of osmoprotectants, particularly proline (El-Sherbeny and Silva 2013). The accumulation of greater quantum of proline in the plant system resulted in enhancing the photosynthetic system and soluble protein (Shahid et al. 2014). Therefore, the research studies were undertaken to quantify the effects of Cu stress on various physiological, biochemical, and chemical attributes and to determine the efficacy of proline to mitigate adverse effects on wheat crop.

Materials and methods
A pot culture experiment was conducted to quantify the effects of proline under copper stress conditions on wheat crop during crop season 2015–2016 at University of Agriculture, Faisalabad-Pakistan. The treatments consisted of three wheat varieties (‘Punjab-96g’, ‘MH-97’, ‘FSD-83’); two levels of copper (0, 400 µM, CuSO₄·5H₂O) applied through rooting medium and two levels of proline (0, 80 mM) applied exogenously and arranged in a completely randomized design with factorial arrangement with four replications.

The seed of wheat varieties was surface sterilized with 5.0 g L⁻¹ sodium hypochlorite solution for five minutes and air-dried at room temperature before dibbling in the experimental pots. Ten kilograms of washed river sand was filled in plastic pots measuring 24.5 × 28.0 cm² and having a drainage hole at the bottom. The impurities from the river sand were removed by a 10-hour leaching with 10% HCl followed by thorough washing with deionized water. Approximately, 15 seeds of wheat varieties were dibbled at 5 mm depth in each pot during November 2015. The pots were irrigated with modified half-strength Hoagland’s nutrient solution at every two days interval (Hoagland and Arnon 1950; Epstein 1972). The evaporated water was replenished with distilled water at other times. After complete germination, five healthy and uniform in size seedlings were retained in each pot. The seedlings were treated with various levels of copper nutrient solution through rooting medium, while foliar spray of proline was carried out at 12th day after complete germination. Proline was applied in combination with 0.1% (v/v) Tween-80 (poloxymethylene sorbiton monoleate) surfactant to maximize penetration into leaf tissues. Moreover, the untreated plants were also sprayed with distilled water. A constant volume of 20 ml proline and/or distilled water was sprayed per pot to ensure full foliage coverage.

**Morphological attributes**

**Biological yield and plant height**
The plants were harvested at day 35 after sowing. The material was washed with deionized water and blotted. The plants were divided into leaves and roots. The quantum of fresh weight of shoot and root was recorded. The material was oven dried at 70°C for 24 h till the constant weight was obtained.

**Gas exchange characteristics**
The measurements on various gas exchange characteristics were recorded by employing LCA-4 ADC portable gas analyzer (IRGA) by selecting flag leaf as the diagnostic leaf. The instrument was calibrated at 403.3 mmol m⁻² s⁻¹ for molar flow of air, 99.9 kPa atmospheric pressure, 6.0–8.9 mbar water vapor pressure, 1711 µmolm⁻² s⁻¹ PAR, 28.4–27.9°C leaf temperature and 352 µmol mol⁻¹ ambient CO₂ concentration.

**Chlorophyll contents**
Data for chlorophyll content were collected by acetone method. The 0.1 g of flag leaves were ground using pestle and mortar in 5 ml of 80% acetone. The centrifuged extract was then used for chlorophyll estimation. The readings were recorded by employing U-2001 Spectrophotometer and calculated by an equation:

\[
\text{Chl. } a (\text{mg ml}^{-1}) = \frac{12.7 (OD663) – 2.69(OD645)}{V/1000 \times W},
\]

\[
\text{Chl. } b (\text{mg ml}^{-1}) = \frac{22.9 (OD645) – 4.68(OD663)}{V/1000 \times W}.
\]

**Estimation of total soluble proteins**
Total soluble proteins of fresh leaf samples were recorded by method (Bradford 1976). For this assay, 0.5 g fresh leaves were ground using a tissue grinder in 5 ml of 50 mM cooled phosphate buffer (pH 7.8) placed in an ice bath. The homogenate was centrifuged at 15,000 rpm for 15 min at 4°C. The supernatant was used for protein determination. Each sample (0.1 ml) was taken in a test tube and mixed with 5 ml of Bradford reagent and incubated at 37°C for 10–15 min along with blank. Reading for absorbance was recorded at 595 nm employing spectrophotometer (IRMECO U2020).

**Proline determination**
Proline contents were determined by Bates et al. (1973). 0.5 g fresh healthy leaf tissue was homogenized in 10 ml of 30% sulpho-salicylic acid and filtered with filter paper. Two milliliters of acid ninhydrin (prepared by mixing 1.25 g ninhydrin in 30 ml glacial acetic acid) and 2 ml of glacial acetic acid were added to the filtrate. The filtrate was mixed and heated in water bath for 60 min at 100°C. The mixture was then cooled and 4 ml of toluene was added and mixed. The chromophore containing toluene was separated from the aqueous phase and its absorbance was recorded at 520 nm with a spectrophotometer (IRMECO U2020). Proline concentration was determinate by using following equation:

\[
\mu \text{ mol proline/fresh weight (g)} = \times (\mu g \text{ proline/ml } \times \text{ml of toluene}/115.5) / \text{g of sample}.
\]
**Determination of inorganic ions**

Sulfuric acid (H\(_2\)SO\(_4\)) and hydrogen peroxide (H\(_2\)O\(_2\)) were used in the process of digestion of plant material. 0.1 g dried plant material (leaf and root) from oven-dried samples and 2 ml of conc. H\(_2\)SO\(_4\) were put into the digestion flasks and incubated it overnight at 25°C temperature. 1 ml of H\(_2\)O\(_2\) was poured down through the sides of digestion flasks, mixed and flasks were placed onto the hot plate and warmed at 250°C until fume formation occurred. Heating was continuously done for 30 min and then digestion flasks were heated until fume formation occurred. Heating was continuously done for 30 min and then digestion flasks were placed onto the hot plate and warmed at 250°C until fume formation occurred. Heating was continuously done for 30 min and then digestion flasks were removed from the plate. Above process was repeated, with the addition of 1 ml H\(_2\)O\(_2\) when needed, until the material became colorless. Then the extract was put into the volumetric flasks and made the volume up to 50 ml, filtered and used for inorganic ion determination.

**Determination of mineral nutrients**

The amount of dissolved mineral nutrients like potassium (K\(^+\)), calcium (Ca\(^{2+}\)) and magnesium (Mg\(^{2+}\)) was recorded by using flame photometer (Model PFPI-7, Jenway, UK), while copper (Cu\(^{2+}\)) determination was done by Atomic Absorption Spectrum.

**Antioxidant enzyme extraction**

For the determination of antioxidant enzymes, 0.5 g fresh leaf samples were ground in tissue grinder in 5 ml of 50 mM cooled phosphate buffer (pH 7.8) placed in an ice bath. The homogenate was centrifuged at 15,000 rpm for 15 min at 4°C. The supernatant was used for the determination of enzyme activities.

**Superoxide dismutase**

The activity of SOD was determined by measuring its ability to inhibit production of nitro blue tetrazolium (NBT) ( Gianopolitis and Ries 1977). The 3 ml reaction solution contained 50 µM NBT, 1.3 µM riboflavin, 13 mM methionine, 75 nM EDTA, 50 mM phosphate buffer (pH 7.8) and 20–50 µl enzyme extract. Test tubes containing the reaction solution were irradiated under light (15 fluorescent lamps) at 78 µmol m\(^{-2}\) s\(^{-1}\) for 15 min. The absorbance of irradiant solution was taken at 560 nm by spectrophotometer (IRMESCO U2020). One unit of SOD activity was defined as the amount of enzyme that inhibited 50% NBT photoreduction.

**Catalase and peroxidase**

The CAT reaction solution (3 ml) contained 50 mM phosphate buffer (pH 7.0), 5.9 mM H\(_2\)O\(_2\) and 0.1 mM enzyme extract. Change in the reaction solution at 240 nm was recorded after every 20 s. One unit CAT activity was defined as an absorbance range of 0.01 units per min. Peroxidase (POD) reaction solution (3 ml) contained 50 mM phosphate buffer (pH 5.0), 20 mM guaiacol, 40 mM H\(_2\)O\(_2\) and 0.1 ml enzyme extract. Change in the reaction solution at 470 nm was recorded after every 20 s. One unit POD activity was defined as an absorbance range of 0.01 units per min (Chance and Maehly 1955).

**Yield**

At maturity, plants were harvested and data for 100-grain weight from each pot were quantified.

**Statistical analysis**

The three-way analysis of variance of data for all the parameters was computed by using a COSTAT computer program (Cohort software Berkeley, California). The least significance differences between mean values were calculated according to (Steel et al. 1997).

**Results**

**Morphological attributes**

(a) Plant height

Data for plant height differed significantly (p < 0.05) in response to wheat varieties, proline and copper levels, and their interactive effects. The foliar spray of proline at the rate 80 mM resulted in increased in plant height by 19.41% in ‘Punjab-96’ over the unsprayed crop. The addition of copper at the rate of 400 µM caused a reduction in plant height by 29.38% in variety ‘FSD-83’ over untreated plants (Table 1).

(b) Shoot and root fresh and dry weight

Data for fresh and dry weight of shoots differed significantly (p < 0.05) due to different wheat varieties, levels of proline and copper and their interaction as well. The foliar spray of proline at the rate of 80 mM resulted in increased in shoot fresh and dry weight by 19.7% and 18.0% in variety ‘FSD-83,’ respectively over untreated plants. The addition of copper at the rate of 400 µM caused a reduction in shoot fresh and dry weight by 42.1% and 42.3% in variety MH-97, respectively over control plants. Similarly, the foliar spray of proline at the rate of 80 mM resulted in increased in root fresh and dry weight by 10.7% and 13.3% in variety ‘FSD-83,’ respectively over untreated plants. The addition of copper at the rate of 400 µM caused a reduction in root fresh and dry weight by 25.9% and 34.0% in variety ‘MH-97,’ respectively over control plants. Results showed that variety ‘FSD-83’ showed better accumulation of biomass when compared to other two wheat varieties (Table 1).

**Chlorophyll constituents**

Data for chlorophyll contents ‘a’ and ‘b’ differed significantly (p < 0.05) due to wheat varieties, proline and copper levels and their interactive effects. Averaged across proline and copper level, wheat variety ‘FSD-83’ maintained higher chlorophyll contents ‘a’ and ‘b’ by 17.8% and 10.1%, respectively, when compared to other varieties. While the application of copper decreased chlorophyll contents by 35.0% in variety Punjab-96 and 27.0% in variety ‘MH-97’ over untreated plants (Table 2).

**Total soluble protein and proline contents**

Data for proline content differed significantly (p < 0.05) in response to wheat varieties, proline and copper levels and their interaction too. The values of proline content in leaf
Table 1. Effect of exogenously applied proline on fresh and dry matter yield under copper stress condition.

| Wheat line | Proline (mM) | Copper level (µM) | 0 | 400 | Mean |
|------------|--------------|-------------------|---|-----|------|
| (a) Plant height (cm) |              |                   |   |     |      |
| FSD-83     | 0            |                   | 90.1 | 77.7 | 83.9 |
| 80         | 100.7        |                   | 87.5 | 94.1 |
| MH-97      | 0            |                   | 77.1 | 67.1 | 72.1 |
| 80         | 83.0         |                   | 75.6 | 79.3 |
| Punjab-96  | 0            |                   | 75.2 | 65.6 | 70.4 |
| 80         | 88.8         |                   | 75.1 | 81.9 |
| Mean       |              |                   | 85.8 | 74.7 |
| LSD (p < 0.05) = Line (L); 0.10** Proline (P); 0.19* Copper (C) 0.19**; LxPxC: 0.22ns. |
| (b) Shoot fresh weight (g) |            |                   | 0 |     |      |
| FSD-83     | 0            |                   | 9.7 | 7.7 | 8.7 |
| 80         | 11.0         |                   | 9.2 | 10.1 |
| MH-97      | 0            |                   | 7.5 | 5.5 | 6.5 |
| 80         | 8.6          |                   | 5.9 | 7.0 |
| Punjab-96  | 0            |                   | 7.6 | 5.4 | 6.5 |
| 80         | 8.6          |                   | 7.0 | 7.8 |
| Mean       |              |                   | 8.8 | 6.7 |
| LSD (p < 0.05) = Line (L); 0.11ns. Proline (P) 0.10** Copper (C) 0.10**; LxPxC: 0.22ns. |
| (c) Shoot dry weight (g) |            |                   | 0 |     |      |
| FSD-83     | 0            |                   | 3.5 | 3.0 | 3.3 |
| 80         | 3.6          |                   | 3.2 | 3.4 |
| MH-97      | 0            |                   | 1.5 | 1.2 | 1.4 |
| 80         | 2.6          |                   | 1.6 | 2.1 |
| Punjab-96  | 0            |                   | 2.1 | 1.2 | 1.7 |
| 80         | 2.4          |                   | 1.7 | 2.1 |
| Mean       |              |                   | 2.6 | 2.0 |
| LSD (p < 0.05) = Line (L); 0.03** Proline (P); 0.12** Copper (C) 0.10**; LxPxC: 0.22 ns. |
| (d) Root fresh weight (g) |            |                   | 0 |     |      |
| FSD-83     | 0            |                   | 2.8 | 2.1 | 2.5 |
| 80         | 3.1          |                   | 2.2 | 2.7 |
| MH-97      | 0            |                   | 2.7 | 2.0 | 2.4 |
| 80         | 2.9          |                   | 2.4 | 2.7 |
| Punjab-96  | 0            |                   | 2.0 | 1.4 | 1.7 |
| 80         | 2.2          |                   | 1.7 | 2.0 |
| Mean       |              |                   | 2.1 | 2.0 |
| LSD (p < 0.05) = Line (L); 0.02** Proline (P); 0.10** Copper (C) 0.09**; LxPxC 0.19 ns. |
| (e) Root dry weight (g) |            |                   | 0 |     |      |
| FSD-83     | 0            |                   | 0.5 | 0.3 | 0.4 |
| 80         | 0.5          |                   | 0.4 | 0.5 |
| MH-97      | 0            |                   | 0.4 | 0.2 | 0.3 |
| 80         | 0.4          |                   | 0.4 | 0.3 |
| Punjab-96  | 0            |                   | 0.5 | 0.3 | 0.4 |
| 80         | 0.6          |                   | 0.4 | 0.5 |
| Mean       |              |                   | 0.5 | 0.3 |
| LSD (p < 0.05) = Line (L); 0.01**Proline (P); 0.02** Copper (C) 0.03**; LxPxC 0.11ns. |

Table 2. Effect of exogenously applied proline on chlorophyll constituents.

| Wheat line | Proline (mM) | Copper level (µM) | 0 | 400 | Mean |
|------------|--------------|-------------------|---|-----|------|
| (a) Chlorophyll 'a' (mg g⁻¹Fw) |              |                   |   |     |      |
| FSD-83     | 0            |                   | 0.004 | 0.003 | 0.004 |
| 80         | 0.005        |                   | 0.004 | 0.005 |
| MH-97      | 0            |                   | 0.003 | 0.002 | 0.003 |
| 80         | 0.004        |                   | 0.003 | 0.004 |
| Punjab-96  | 0            |                   | 0.004 | 0.003 | 0.004 |
| 80         | 0.004        |                   | 0.003 | 0.004 |
| Mean       |              |                   | 0.004 | 0.002 |
| LSD (p < 0.05) = Line (L); 0.001**Proline (P); 0.001**Copper (C) 0.001**; LxPxC 0.003ns. |
| (b) Chlorophyll 'b'(mg g⁻¹Fw) |            |                   | 0 |     |      |
| FSD-83     | 0            |                   | 0.006 | 0.005 | 0.006 |
| 80         | 0.006        |                   | 0.005 | 0.006 |
| MH-97      | 0            |                   | 0.005 | 0.004 | 0.005 |
| 80         | 0.006        |                   | 0.005 | 0.006 |
| Punjab-96  | 0            |                   | 0.005 | 0.005 | 0.005 |
| 80         | 0.006        |                   | 0.005 | 0.006 |
| Mean       |              |                   | 0.006 | 0.005 |
| LSD (p < 0.05) = Line (L); 0.001**Proline (P); 0.001** Copper (C) 0.001**; LxPxC 0.002ns. |

tissues was increased significantly (72.9%) in variety 'FSD-83' due to copper presence in growing media. Similarly, total soluble protein contents were also increased significantly in all varieties especially in variety 'MH-97' (33.0%) due to copper presence in growing media (Figure 1).

Antioxidant enzymes

Data for antioxidant enzymes was significantly (p < 0.05) enhanced due to the imposition of copper in growing media in all wheat varieties. The foliar spray of proline produced a little effect on increasing the levels of SOD. However, the addition of copper at the rate of 400 µM caused an increase in SOD by 58.14% in Punjab-96 over the untreated crop. Amount of peroxidase (POD) differed significantly in response to various treatments. The wheat variety 'Punjab-96' maintained higher POD contents by 64.1% under 400 µM copper stress followed by 'FSD-83' and 'MH-97' varieties. Data for catalase (CAT) differed significantly due to the imposition of copper treatments by 25.4% in variety 'MH-97' when compared to untreated crop. Averaged across wheat varieties and spray of proline caused an increase in SOD, POD and CAT levels by 20.9%, 22.2%, and 9.1%, respectively (Figure 1).

Gas exchange characteristics

Data for photosynthetic rate differed significantly (p < 0.05) in response to wheat varieties, proline and copper levels, and their interactive effects. The foliar spray of proline at the rate of 80 mM caused an increase in photosynthetic rate by 19.0%, transpiration rate by 6.8%, sub-stomatal CO₂ concentration by 6.9%, stomatal conductance by 21.2%, and water-use-efficiency by 11.4% over untreated crop. The addition of copper at the rate of 400 µM caused a reduction in photosynthetic rate by 26.06%, transpiration rate by 25.0%, sub-stomatal CO₂ concentration by 16.2%, stomatal conductance by 21.2%, and water-use-efficiency by 20.9% over untreated crop. Based on gas exchange characteristics variety, 'FSD-83' showed better results as compared to varieties 'MH-97' and 'Punjab-96' (Figure 2).

Chlorophyll florescence

Data for non-photochemical quenching co-efficient differed significantly (p < 0.05) in response to wheat varieties, proline and copper levels as well as their interaction. The values of non-photochemical quenching co-efficient were decreased by 14.9% in FSD-83 due to spray of proline chemical as compared to non-sprayed plants. However, the addition of copper at the rate of 400 mM caused an increase in non-photochemical quenching co-efficient by 23.6% in Punjab-96 over control. Data for efficiency of photosystem-II differed significantly in response to treatments among varieties, the wheat variety 'FSD-83' maintained higher level of efficiency of photosystem-II by 12.3% followed by varieties 'MH-97' and 'Punjab-96' varieties, whereas, the imposition of copper decreased PII efficiency by 17.6% in variety 'MH-97' over unsprayed crop. Similarly, the values for electron transfer rate (ETR) were enhanced by foliar application of proline by 12.7% in variety 'FSD-83' while the addition of copper caused a reduction in ETR by 16.7% over untreated crop (Figure 3).
**Ionic concentration in leaf tissues**

Data for metal elements in leaf differed significantly ($p < 0.05$) in response to proline, copper levels and wheat varieties, as well as their interaction. Data for calcium content differed significantly in leaf organs due to differed treatments. The wheat variety 'FSD-83' maintained higher leaf Ca$^{2+}$ content followed by 'MH-97' and 'Punjab-96' varieties. The foliar spray of proline caused an increase in Ca$^{2+}$ content by 40.3% over untreated crop. The addition of copper caused a reduction in Ca$^{2+}$ content by 42.8% in variety 'Punjab-96' over untreated crop. Data for magnesium content in leaf organ differed greatly in response to the imposition of different treatments. The variety 'FSD-83' maintained higher Mg$^+$ content 11.4% compared to other varieties. Moreover, the addition of copper caused a reduction in Mg$^+$ content by 20.5% in Punjab-96 over the untreated crop. Data for potassium content differed significantly in response to treatments. The wheat variety 'FSD-83' maintained maximum K$^+$ content by 16.7% due to foliar application of proline as compared to

![Figure 1. Effect of foliar application of proline (80 mM) on protein, proline, SOD, POD, and CAT contents of wheat varieties grown under Cu stress.](image-url)
other two varieties. However, the addition of Cu caused a reduction in K⁺ content by 36.3% in Punjab-96 over the untreated crop. Data for copper content in leaf organ also enhanced significantly due to copper treatments. The addition of Cu²⁺ at the rate of 400 µM caused an increase in Cu²⁺ contents by 87.5% in leaf tissues of variety Punjab-96 over control plants (Table 3).

**Ionic concentration in root tissues**

Data for metal elements in root differed significantly (p < 0.05) in response to proline, copper levels, and wheat varieties, as well as their interaction. Data for Ca²⁺ content differed significantly in root organs due to differed treatments. The wheat variety ‘FSD-83’ maintained higher root Ca²⁺ content followed by ‘MH-97’ and ‘Punjab-96’ varieties. The foliar

![Figure 2. Effect of foliar application of proline (80 mM) on stomatal conductance, sub-stomatal CO₂ concentration, CO₂ assimilation rate, transpiration rate, and water-use-efficiency in wheat varieties grown under Cu stress.](image-url)
spray of proline caused increase Ca\textsuperscript{2+} content by 17.0% over untreated crop. The addition of copper caused a reduction in Ca\textsuperscript{2+} content by 26.6% in variety ‘MH-97’ over untreated crop. Data for magnesium content in root organ differed greatly in response to imposition of different treatments. The variety ‘FSD-83’ maintained higher Mg\textsuperscript{2+} content 10.4% compared to other varieties. Moreover, the addition of copper caused a reduction in Mg\textsuperscript{2+} content by 24.1% in variety MH-97 over untreated crop. Data for potassium content differed significantly in response to treatments. The wheat variety ‘FSD-83’ maintained maximum K\textsuperscript{+} content by 13.9% due to foliar application of proline when compared to other two varieties, while, the values of 100-grain weight were decreased by 18.75% in Punjab-96 due to the addition of copper compared to untreated check (Table 5).

**Discussion**

The productivity of crop species is affected to a greater proportion due to biotic (diseases, insect pests) and abiotic (eco-edaphic factors, salinity, drought, temperature, and heavy metal pollutants) in the agricultural production system (Lawlor and Cornic 2002). In the recent years, the rapid industrialization, the outflow of effluents from different industries has caused havoc in polluting the irrigation water and surface soils of agricultural areas. These pollutants contain heavy toxic metals, which not only affect the crop yield but also add poison to the food chain for causing abnormalities on the healthiness of humans and livestock. Among these heavy metals, the copper metal plays a vital role in growth and development of crops as it is the main constituent of many proteins and enzymes processes. On the other hand, high concentration results in toxicity in plants, thereby growth is inhibited (Hall 2002). Various researchers (Hall and Williams 2003; Shar et al. 2011) reported that excessive concentration of copper caused disruption in protein and nitrogen metabolism, by disrupting the photosynthesis and
Table 3. Effect of exogenously applied proline on nutrient concentration in shoot organ.

| Wheat line | Proline (mM) | Copper level (µM) |
|------------|--------------|-------------------|
|            | 0            | 400               | Mean |
| (a) Calcium (Ca+2) (mg g−1 dw) |
| FSD-83     | 0            | 4.3               | 3.0   | 3.7   |
|            | 80           | 6.0               | 4.7   | 5.4   |
| MH-97      | 0            | 4.0               | 2.3   | 3.2   |
|            | 80           | 6.0               | 4.0   | 5.0   |
| Punjab-96  | 0            | 4.7               | 2.7   | 3.7   |
|            | 80           | 5.7               | 3.3   | 4.5   |
| Mean       |              | 5.1               | 3.3   |       |
| LSD (p < 0.05) = Line (L); 0.08* Proline (P); 0.07** Copper (C) 0.06*; LxPxC 0.12ns |
| LSD (p < 0.05) = Line (L); 0.10** Proline (P); 0.11** Copper (C) 0.11**; LxPxC 0.15ns |
| LSD (p < 0.05) = Line (L); 0.08* Proline (P); 0.06** Copper (C) 0.04*; LxPxC 0.13ns |
| (b) Magnesium (Mg+2) (mg g−1 dw) |
| FSD-83     | 0            | 8.0               | 6.7   | 7.4   |
|            | 80           | 8.7               | 7.2   | 8.0   |
| MH-97      | 0            | 8.0               | 6.7   | 7.4   |
|            | 80           | 8.8               | 6.9   | 7.9   |
| Punjab-96  | 0            | 8.6               | 6.8   | 7.7   |
|            | 80           | 8.9               | 6.7   | 7.8   |
| Mean       |              | 8.5               | 6.8   |       |
| LSD (p < 0.05) = Line (L); 0.15** Proline (P); 0.21* Copper (C) 0.22**; LxPxC 0.78ns |
| LSD (p < 0.05) = Line (L); 0.04* Proline (P); 0.008** Copper (C) 0.09*; LxPxC 0.04ns |
| LSD (p < 0.05) = Line (L); 0.03** Proline (P); 0.004* Copper (C) 0.03**; LxPxC 0.12ns |

Table 4. Effect of exogenously applied proline on nutrient concentration in root organ.

| Wheat line | Proline (mM) | Copper level (µM) |
|------------|--------------|-------------------|
|            | 0            | 400               | Mean |
| (a) Calcium (Ca+2) (mg g−1 dw) |
| FSD-83     | 0            | 15.7              | 12.0  | 13.9  |
|            | 80           | 16.0              | 13.0  | 14.5  |
| MH-97      | 0            | 15.0              | 12.3  | 13.7  |
|            | 80           | 15.0              | 13.3  | 14.2  |
| Punjab-96  | 0            | 14.0              | 12.3  | 13.2  |
|            | 80           | 16.0              | 14.3  | 15.2  |
| Mean       |              | 15.3              | 12.8  |       |
| LSD (p < 0.05) = Line (L); 0.07** Proline (P); 0.07** Copper (C) 0.004*; LxPxC 0.16ns |
| LSD (p < 0.05) = Line (L); 0.15** Proline (P); 0.21* Copper (C) 0.22**; LxPxC 0.78ns |

Table 5. Effect of exogenously applied proline on 100-grain weight under copper stress.

| Wheat line | Proline (mM) | Copper level (µM) |
|------------|--------------|-------------------|
|            | 0            | 400               | Mean |
| (a) 100-grain weight (g) |
| FSD-83     | 0            | 4.4               | 3.2   | 3.8   |
|            | 80           | 4.6               | 3.8   | 4.2   |
| MH-97      | 0            | 3.9               | 3.2   | 3.6   |
|            | 80           | 4.1               | 3.3   | 3.7   |
| Punjab-96  | 0            | 3.5               | 2.8   | 3.2   |
|            | 80           | 4.0               | 3.3   | 3.7   |
| Mean       |              | 4.1               | 3.3   |       |
| LSD (p < 0.05) = Line (L); 0.04* Proline (P); 0.008** Copper (C) 0.09*; LxPxC 0.13ns |

respiration processes. The occurrence of chlorosis of leaves caused inhibition of plant growth (Fariduddin et al. 2009). Janas et al. (2010) reported that metabolic processes such as leaf and root growth, photosynthetic rate, production of biomass and pigment contents were hindered. The growth and development of crop species is an outcome of various morphological parameters i.e. plant height, number of leaves per plant and flag leaf area in wheat. The results of the study indicated reduced plant height in response to a higher content of copper in the rhizosphere. The findings corroborated with those of Houshm and Moraghebi (2011) that plant height was reduced due to the presence of higher content of copper in the root zone. The spray of proline caused an increase in apical meristem and cell division, which improved the plant height. These results agree with those of Ali et al. (2013).

The copper stress caused significant reduction in fresh and dry weights of shoots and root organs. Various researchers (El-Tayeb et al. 2006; Sommez et al. 2006; Muslu and Ergun 2013) also reported that biological yield was depressed in response to copper stress. Furthermore, the foliar spray of proline caused improvement in shoot and root weights. The stress due to higher quantity of copper caused nutrients imbalance in the plant system, which caused reduction in seedlings growth and germination rate. Various researchers (Imlay 2003; Ouzounidou 1995; Yadav 2010; Sethy and Ghosh 2013; Kalai et al. 2014) reported that osmotic stress impacted negatively on the growth of seedlings in response to specific ion toxicity.
2011; Muslu and Ergun 2013; Sharma and Singh 2013) reported that increase in antioxidant enzymes (SOD, POD, CAT) and protein contents provided protection to avoid possible destruction in important enzymes. Furthermore, accumulation of proline protected the plants against damage of ROS caused by copper stress (Matsysik et al. 2002; Verma et al. 2011; Azzoø et al. 2012).

Some of the gas exchange characteristics (i.e. stomatal conductance (gs), photosynthetic rate (A), internal CO₂ concentration (Ci), transpiration rate (E) and water-use-efficiency) were greatly reduced due to copper stress. However, the negative effects were ameliorated in response to spray of proline. The similar results have been reported by Ali et al. (2007) and Ben Ahmad et al. (2010). Kaymakanova et al. (2008) reported that stomatal conductance was reduced by abiotic stress while CO₂ concentration and biochemical capability was reduced (Khan and Panda 2008). Janas et al. (2010) reported that photosynthetic activity was reduced due to inhibition of Rubisco activity. Chlorophyll fluorescence parameters were a little affected due to copper stress. Jamil et al. (2007) reported that values of electron transfer (ETR), efficiency of photosystem-II (Fv/Fm), photochemical quenching co-efficient (qP) and non-photochemical quenching co-efficient (qN) were slightly reduced, while, the activity of non-photochemical quenching (NPQ) were slightly enhanced in response to copper stress. The reduction in values of Fv/Fm ratio was mainly due to the reduction in Fm value. The reduction resulted due to increase in energy dissipation, disintegration of accessory pigments for capturing light from photosystem-II (Maxwell and Johnson 2000).

The uptake of nutrients was greatly impacted in response to copper stress. The uptake of calcium, magnesium, and potassium ions by root, shoot, and leaf organs was reduced due to copper stress. On the other hand, copper content substantially increased in root organs compared to shoot and leaf organs under copper stress environment. The maintenance of higher content of copper by roots inhibited the uptake of other nutrients. The results agree with those of Michaud et al. (2008) and Karimi et al. (2012) that presence of copper in excessive amount in roots inhibited the other nutrients for translocation from roots to above ground parts. Contrary to it, the foliar spray of proline caused an increase in uptake of Ca²⁺, Mg²⁺, Na⁺, and K⁺ by root, shoot and leaf organs, while the uptake of copper was also reduced by all parts. Ashraf and Foolad (2007) reported that uptake of ions by plants was regulated by proline spray under stress environment. The copper stress caused a reduction in 100-grain weight. Different researchers (Athar and Ahmad 2002; Ahmad et al. 2013; Gang et al. 2013) also reported that yield and yield components of wheat were reduced under copper stress.

The results of the study indicated that presence of higher copper ion in the rhizosphere resulted in reduction of plant growth, physiological and biochemical parameters. However, the stress could be mitigated by foliar spray of proline at the rate of 80 mM under copper stress conditions. In the present study wheat variety ‘FSD-83’ produced showed better bio-mass accumulation and biochemical attributes when compared to varieties ‘MH-97’ and ‘Punjab-96.’

Disclosure statement

No potential conflict of interest was reported by the authors.

References

Adrees M, Ali S, Rizwan M, Ibrahim M, Abbas F, Farid M, Zia-ur-Rehman I, Irshad MK, Bhawana SA. 2015. The effect of excess copper on growth and physiology of important food crops: a review. Environ Sci Pollut Res. 22:8148–8162.

Ahmad K, Khan ZI, Ashraf S, Ejaar A, Shaheen M, Raza SH, Abbas F, Tahir HM. 2013. Effect of sewage water irrigation on the uptake of some essential minerals in canola (Brassica napus L.): A potential forge crop for ruminants. Pak J Life Sci Soc. 11:42–47.

Ahmed A, Hasnain A, Akhtar S, Hussain A, Yasin G, Wahid A, Mahmood S. 2010. Antioxidant enzymes as bio-markers for copper tolerance in safflower (Carthamus tinctorius L.). Afr J Biotechnol. 9:5441–5444.

Ali Q, Ashraf M, Athar H-U-R. 2007. Exogenously applied proline at different growth stages enhances growth of two maize cultivars grown under water deficit conditions. Pak J Bot. 39:1133–1144.

Ali HM, Siddiqui MH, Al-Wahaib MH, Basalam MA, El-Zaidy M. 2013. Effect of proline and abscisic acid on the growth and physiological performance of faba bean under water stress. Pak J Bot. 45:933–940.

Ashraf M, Foolad M. 2007. Roles of glycine betaine and proline in improving plant abiotic stress resistance. Environ Exp Bot. 59:206–216.

Athar R, Ahmad M. 2002. Heavy metal toxicity: effect on plant growth and metal uptake by wheat, and on free living Azotobacter. Water, Air, Soil Pollut. 138:165–180.

Azzoø MM, Abou-Elhamd MF, Al-Fredan MA. 2012. Biphasic effect of copper on growth, proline, lipid peroxidation and antioxidant enzyme activities of wheat (Triticum aestivum cv. Hasawali) at early growing stage. Aust J Crop Sci. 6:688.

Bates L, Waldren R, Teare I. 1973. Rapid determination of free proline for water-stress studies. Plant Soil. 39:205–207.

Ben Ahmed C, Ben Rouina B, Sensoy S, Boukhriss M, Ben Abdullah F. 2010. Exogenous proline effects on photosynthetic performance and antioxidant defense system of young olive tree. J Agric Food Chem. 58:4216–4222.

Bradford MM. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem. 72:248–254.

Brahim I, Mohamed M. 2011. Effects of copper stress on antioxidative enzymes, chlorophyll and protein content in Atriplex halimus. Afr J Biotechnol. 10:10143–10148.

Chance B, Maehly A. 1955. Assay of catalase and peroxidase. Methods Enzymol. 2:764–775.

Chen W, Chang AC, Wu L. 2007. Assessing long-term environmental risks of trace elements in phosphate fertilizers. Environ Toxicol Environ Saf. 67:48–58.

El-Sherbeny MR, Silva JATd. 2013. Foliar treatment with proline and tyrosine affect the growth and yield of beetroot and some pigments in beetroot leaves. J Hortic Res. 21:95–99.

El-Tayeb M, El-Enany A, Ahmed N. 2006. Salicylic acid-induced adaptive response to copper stress in sunflower (Helianthus annuus L.). Plant Growth Regul. 50:191–199.

Epstein F. 1972. Mineral nutrition of plants: principles and prospectives. New York (NY): Wiley.

Fariduddin Q, Yusuf M, Hayat S, Ahmad A. 2009. Effect of 28-homobrassinolide on antioxidant capacity and photosynthesis in Brassica juncea plants exposed to different levels of copper. Environ Exp Bot. 66:418–424.

Frary A, Göl D, Ökmen B, Pınar H, Şığlu A, Doğanlar S. 2010. Salt tolerance in Solanum penellii: antioxidant response and related QTL. BMC Plant Biol. 10:01–16.

Gang A, Vyas A, Vyas H. 2013. Effect of 28-homobrassinolide on antioxidant capacity and photosynthesis in Brassica juncea plants exposed to different levels of copper. Environ Exp Bot. 66:418–424.

Gang A, Vyas A, Vyas H. 2013. Toxic effect of heavy metals on germination and seedling growth of wheat. J Environ Dev. 8:206.

Giannopolitis CN, Ries SK. 1977. Superoxide dismutases I. Occurrence in higher plants. Plant Physiol. 58:309–314.

Hall J. 2002. Cellular mechanisms for heavy metal detoxification and tolerance. J Exp Bot. 53:1–11.

Hall J, Williams LE. 2003. Transition metal transporters in plants. J Exp Bot. 54:2601–2613.

Hoagland DR, Arnon DI. 1950. The water-culture method for growing plants without soil. Circular, California Agricultural Experiment Station, 347.

Houshm A, Moragheibi F. 2011. Effect of mixed cadmium, copper, nickel and zinc on seed germination and seedling growth of safflower. Afr J Agric Res. 6:1463–1468.
Imlay JA. 2003. Pathways of oxidative damage. Annu Rev Microbiol. 57:395–418.

Jamal A, Ayub N, Usman M, Khan AG. 2002. Arbuscular mycorrhizal fungi enhance zinc and nickel uptake from contaminated soil by soybean and lentil. J Int Phytoremediation. 4:205–221.

Jamil M, Rehman S, Lee KJ, Kim HS, Rha ES. 2007. Salinity reduced growth PS2 Photochemistry and chlorophyll content in radish. Sci Agric. 64:111–118.

Kaymakanova M, Stoeva N, Mincheva T. 2008. Salinity and its effects on the physiological response of bean (Phaseolus vulgaris L.). J Cent Eur Agric. 9:749–755.

Ke W, Xiong Z, Xie M, Luo Q. 2007. Accumulation, subcellular localization and ecophysiological responses to copper stress in two Daucus carota L. populations. Plant Soil. 292:291–304.

Khan M, Panda S. 2008. Alterations in root lipid peroxidation and anti-oxidative responses in two rice cultivars under NaCl-salinity stress. Curr Sci. 82:525–532.

Lawlor DW, Cornic G. 2002. Photosynthetic carbon assimilation and associated metabolism in relation to water deficits in higher plants. Plant Cell Environ. 25:275–294.

Matysik J, Alia, Bhalu B, Mohanty. 2002. The impact of copper ions on growth, lipid peroxidation, and phenolic compound accumulation and localization in lentil (Lens culinaris Medic.) seedlings. J Plant Physiol. 167:270–276.

Kalai T, Khamassi K, Silva JATd, Gouia H, Ben-Kaab LB. 2014. Molecular mechanisms of structural effects of copper in Cruciferae. Plant Soil. 310:151–165.

Kaya H, Uz I. 2006. High level of copper application to soil and leaves reduce the growth and yield of tomato plants. Scientia Agricola. 63:213–218.

Steel RGD, Torree JH, Dickey DA. 1997. Principles and procedures of statistics: a biometrical approach. 3rd ed. New York (NY): McGraw Hill.

Szafran L, Savouré A. 2010. Proline: a multifunctional amino acid. Trends Plant Sci. 15:89–97.

Verma JP, Singh V, Yadav J. 2011. Effect of copper sulphate on seed germination, plant growth and peroxidase activity of Mung bean (Vigna radiata). Int J Bot. 7:200–204.

Xu J, Yang L, Wang Z, Dong G, Huang J, Wang Y. 2006. Toxicity of copper on rice growth and accumulation of copper in rice grain in copper contaminated soil. Chemosphere. 62:602–607.

Yadav S. 2010. Heavy metals toxicity in plants: an overview on the role of glutathione and phytochelatins in heavy metal stress tolerance of plants. S Afr J Bot. 76:167–179.

Younas M, Shahzad F, Afzal S, Khan MI, Ali K. 1998. Assessment of Cd, Ni, Cu, and Pb pollution in Lahore, Pakistan. Environ Int. 24:761–766.

Yurechk F, Porgali ZB. 2006. The effects of excessive exposure to copper in bean plants. Acta Biol Cracov Ser Bot. 48:7–13.

Zengin FK, Kirbag S. 2007. Effects of copper on chlorophyll, protein, protein and abscisic acid level of sunflower (Helianthus annuus L.) seedlings. J Environ Biol. 28:561–566.