### Supplementary Text

#### Tables

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| A                  | Strains used in this study   |
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| C. albicans strain name | Parent | Genotype                                                                 | Strain background /construction                                                                 | Reference   |
|------------------------|--------|--------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------|-------------|
| SC5314                 |        | Wild type                                                                |                                                                                                | [1]         |
| SN95                   |        | arg4Δ/arg4Δ his1Δ/his1Δ IRO1/iro1Δ::λimm34 URA3/ura3Δ::λimm34             |                                                                                                | [2]         |
| JKC917                 | SN95   | hisΔ1/his1Δ:: telR-FRT arg4/arg4 IRO1/iro1Δ::λimm34 URA3/ura3Δ::λimm34    | JKC917 transformed with PCR product of fjk1184 & rjk1186 using Candida cDNA library as template   | [3]         |
| JKC1361                | JKC917 | hisΔ1/his1Δ:: telR-FRT arg4/ARG4 IRO1/iro1Δ::λimm34 URA3/ura3Δ::λimm34    | JKC917 transformed with PCR product of fjk1184 & rjk1186 using Candida cDNA library as template   | [3]         |
| JKC1713                | JKC917 | hisΔ1/his1Δ:: telR-FRT arg4/ARG4 IRO1/iro1Δ::λimm34 URA3/ura3Δ::λimm34    | JKC917 transformed with PCR product of fjk1184 & rjk1186 using Candida cDNA library as template   | This work   |
| JKC1347                | JKC917 | tor1::ARG4/TOR1                                                         | tor1/TOR1 heterozygote constructed like JKC1347, independent isolate.                           | This work   |
| JKC1346                | JKC917 | tor1::ARG4/TOR1                                                         | tor1/TOR1 heterozygote constructed like JKC1347, independent isolate.                           | This work   |
| JKC1441                | JKC1347| tor1::ARG4/tetO::FRT-tetO-TOR1-De1381                                    | JKC1347 transformed with Sull/Ncol digested pJK1189 to have TOR1-De1381 under tetO (OFF) promoter, after inducing FLP | [3]         |
| JKC1442                | JKC1345| tor1::ARG4/tetO::FRT-tetO-TOR1-De1381                                    | tor1/tetO-TOR1-De1381 constructed like JKC1441, distinct lineage from JKC1345.                  | This work   |
| JKC1445                | JKC1346| tor1::ARG4/tetO::FRT-tetO-TOR1-De1381                                    | tor1/tetO-TOR1-De1381 constructed like JKC1441, distinct lineage from JKC1346.                  | This work   |
| JKC1549                | JKC1347| tor1::ARG4/tetO::FRT-tetO-TOR1-FL                                       | JKC1347 transformed with Sull/Ncol digested pJK1236 to have TOR1-FL under tetO (OFF) promoter, after inducing FLP | [3]         |
| JKC1543                | JKC1345| tor1::ARG4/tetO::FRT-tetO-TOR1-FL                                       | tor1/tetO-TOR1-FL constructed like JKC1549, distinct lineage from JKC1345.                      | This work   |
| JKC1546                | JKC1346| tor1::ARG4/tetO::FRT-tetO-TOR1-FL                                       | tor1/tetO-TOR1-FL constructed like JKC1549, distinct lineage from JKC1346.                      | This work   |
| TETG25B                | CAI4   | ADH1/adh1::tetO(ON)-GFP                                                 | CAI4 transformed with pTET25 tetracycline inducible GFP cassette                                 | [4]         |
| JKC2616                | JKC1713| hisΔ1/his1Δ:: telR-FRT arg4/ARG4 MAL2/pMAL2-GFP-FRT-pMAL2-MAL2          | JKC1713 transformed with BsrGI digested pJK1489 to have GFP under pMAL2 promoter, after inducing FLP | This work   |
| JKC2620                | JKC1347| tor1::ARG4/tetO::FRT-tetO-TOR1-De1381                                    | JKC1347 transformed with BsrGI digested pJK1489 to have GFP under pMAL2 promoter, after inducing FLP | This work   |
| JKC2624                | JKC1441| tor1::ARG4/tetO::FRT-tetO-TOR1-De1381                                    | JKC1441 transformed with BsrGI digested pJK1489 to have GFP under pMAL2 promoter, after inducing FLP | This work   |
| JKC2628                | JKC1549| tor1::ARG4/tetO::FRT-tetO-TOR1-FL                                       | JKC1549 transformed with BsrGI digested pJK1489 to have GFP under pMAL2 promoter, after inducing FLP | This work   |
Table B. Plasmids used in this study.

| Plasmid  | Description                                                                 | Source (Reference) |
|----------|-----------------------------------------------------------------------------|--------------------|
| pJK1000  | *FLP*-NAT1 tetO-PES1 construct, vector backbone is pLitmus28 (New England Biolabs) | [5]                |
| pJK1027  | pUA34 with URA3 disrupted by Ag promoter TEF1-NAT1-TEF1                      | [6,7]              |
| pJK1189  | *FLP*-NAT1 tetO-TOR1-Del381 construct, derived from pJK1000.                  | [3]                |
| pJK1236  | *FLP*-NAT1 tetO-TOR1-FL construct, derived from pJK1000.                      | [3]                |
| pAU15    | pMAL2 expression vector                                                     | [6]                |
| pGFP-HIS1| GFPHIS1 fusion construct                                                    | [8]                |
| pJK1482  | pMAL2-GFP construct, derived from pAU15. Product of fjk2036 & rjk1633 using pGFP-HIS1 as template was ligated into pAU15 using SalI/XmaI sites. | This work          |
| pJK1489  | *FLP*-NAT1 pMAL2-GFP construct, derived from pJK1482. URA3 marker in pJK1482 was replaced with the 'FLP-NAT1' cassette by blunt cloning. | This work          |

Table C. Oligonucleotides used in this study.

| Primer name | Purpose                                                                 | Sequence 5' to 3' (lower cases - restriction enzyme recognition sites) |
|-------------|-------------------------------------------------------------------------|------------------------------------------------------------------------|
| fjk2062     | Forward primer for TOR1 quantitative real-time PCR                      | GCTTAGTTTTATCAGGCAAGGGA                                                |
| rjk2063     | Reverse primer for TOR1 quantitative real-time PCR                       | ACTCATCCCCGTGTCTCTTAG                                                  |
| fjk1400     | Forward primer for ACT1 quantitative real-time PCR                       | TGGTGATGTTGTTAACTCAG                                                  |
| rjk1401     | Reverse primer for ACT1 quantitative real-time PCR                       | GACAATTCTCTTTTACGAC                                                  |
| fjk1184     | Forward primer to amplify the ARG4 marker                              | GAATCCACAATCGTATATGAAC                                                 |
| rjk1186     | Reverse primer to amplify the ARG4 marker                              | GAATATAGTGATGAGGGAT                                                  |
| fjk1185     | Forward primer to confirm the 5'end integration of ARG4 marker in Candida strains. | GACATATGGACGACATAATTC                                              |
| rjk1187     | Reverse primer to confirm the 5'end integration of ARG4 marker in Candida strains. | GTCGTTTACCCGGTTGCCACTG                                             |
| fjk1188     | Forward primer to confirm the 3'end integration of ARG4 marker in Candida strains. | CAGTACCACAATAGCATCTC                                                |
| rjk1199     | Reverse primer to confirm the 3'end integration of ARG4 marker in Candida strains. | GTAGTCTCCGATATTGATCTC                                             |
| fjk2036     | Forward primer to amplify GFP sequence.                                 | CCTGCTgtgacATGTCTAAAGGTGAAGAATTAT                                     |
| rjk1633     | Reverse primer to amplify GFP sequence and to confirm the 5'end integration of pMAL2-GFP in Candida strains | GCAGCTccgggTTATTTTGAATAATTCCATCCATCCATGG                             |
| fjk2045     | Forward primer to confirm the 3'end integration of pMAL2-GFP in Candida strains. | CATTTGTTGAGCTGCGACT                                             |
| fjk1517     | Forward primer to confirm the 3'end integration of pMAL2-GFP in Candida strains. | GGAATTTGAGCGGATAC                                             |
| rjk2046     | Reverse primer to confirm the 3'end integration of pMAL2-GFP in Candida strains. | CAAGGTCCCGTATTTGCTGT                                                 |
Table D. Antibodies used in this study.

| Purpose        | Antigen recognized | Species | Source or Reference                  |
|----------------|--------------------|---------|--------------------------------------|
| primary        | P-Mkc1             | rabbit  | Cell Signaling Technology, #4370P   |
| primary        | P-S6               | rabbit  | Cell Signaling Technology, #9611L   |
| primary        | S6                 | sheep   | R&D Systems, #AF5436                |
| primary        | P-Hog1             | rabbit  | Cell Signaling Technology, #4511S   |
| primary        | P-eIF2a            | rabbit  | Cell Signaling Technology, #3597S   |
| primary        | GFP                | mouse   | Roche, #11814460001                |
| loading control| PSTAIRE (Cdc2)     | rabbit  | Santa Cruz Biotechnology, #sc-53    |
| loading control| Tubulin            | rat     | Abcam, #ab6161                     |
| secondary      | Rabbit IgG         | goat    | Cell Signaling Technology, #7074S   |
| secondary      | Sheep IgG          | donkey  | Santa Cruz Biotechnology, #sc-2473  |
| secondary      | Mouse IgG          | horse   | Cell Signaling Technology, #7076S   |
| secondary      | Rat IgG            | goat    | Abcam, #ab97057                    |

Reference:
1. Fonzi WQ & Irwin MY. Isogenic strain construction and gene mapping in Candida albicans. Genetics. 1993;134(3):717-728.
2. Noble SM & Johnson AD. Strains and strategies for large-scale gene deletion studies of the diploid human fungal pathogen Candida albicans. Eukaryotic Cell. 2005;4(2):298-309.
3. Liu NN, Flanagan PR, Zeng J, Jani NM, Cardenas ME, Moran GP, et al. Phosphate is the third nutrient monitored by TOR in Candida albicans and provides a target for fungal-specific indirect TOR inhibition. Proc Natl Acad Sci U S A. 2017;114(24):6346-51. doi: 10.1073/pnas.1617799114. PubMed PMID: 28566496; PubMed Central PMCID: PMCPMC5474788.
4. Park YN & Morschhäuser J. Tetracycline-inducible gene expression and gene deletion in Candida albicans. Eukaryotic Cell. 2005;4(8):1328-1342.
5. Shen J, Cowen LE, Griffin AM, Chan L, Köhler JR. The Candida albicans pescadillo homolog is required for normal hypha-to-yeast morphogenesis and yeast proliferation. Proc Natl Acad Sci U S A. 2008;105(52):20918-23. Epub 2008/12/17. doi: 10.1073/pnas.0809147105. PubMed PMID: 19075239; PubMed Central PMCID: PMC2634893.
6. Uhl MA & Johnson AD. Development of Streptococcus thermophilus lacZ as a reporter gene for Candida albicans. Microbiology. 2001;147(Pt 5):1189-1195.
7. Patenaude C, Zhang Y, Cormack B, Köhler J, Rao R. Essential role for vacuolar acidification in Candida albicans virulence. The Journal of Biological Chemistry. 2013;288(36):26256-26264.
8. Gerami-Nejad M, Berman J, Gale CA. Cassettes for PCR-mediated construction of green, yellow, and cyan fluorescent protein fusions in Candida albicans. Yeast. 2001;18(9):859-864.