Nanostructured titanium dioxide for use in bone implants: a short review

(Dióxido de titânio nanoestruturado para uso em implantes ósseos: uma breve revisão)

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Abstract
Titanium dioxide (TiO2) based nanostructured materials have shown great potential for use in implants thanks to their excellent physicochemical properties, such as high specific surface area, ability to elicit positive cell response, and stability in body fluids. However, there are few studies in the literature that focus on the use of nanostructured TiO2 to support cell growth and bone regeneration. The purpose of this survey is to review the state of the art of TiO2 for use in bone implants, as well as provide insight into its characteristics and synthesis methods. Studies of the biological properties of nanostructured TiO2 are described, establishing its potential for the biomedical field.

Keywords: nanostructured materials, titanium dioxide, biomaterials.

INTRODUCTION
Materials for bone repair or regeneration are needed in many situations in orthopedics and orthodontics and are critical for people suffering from osteoporosis, bone cancer, joint and spinal disease, and disorders. As the world population ages and cases of osteoporosis and bone trauma increase sharply among the elderly, the number of individuals that need bone implants to boost bone regeneration has grown dramatically in recent decades [1, 2]. The use of autogenous and allogeneic bone grafts may help in the treatment of bone lesions. However, they have numerous disadvantages, such as the need for secondary surgery, limitation in the number of grafts, risk of infections, and immune responses, which may cause other severe health problems [3, 4]. In recent years, most orthopedic research has focused on nanostructured biomaterials due to their similarities to the natural physiological environment [5-11]. Nanoscale morphology has unique properties such as high surface area and a higher degree of biological plasticity than other microstructures [12-14]. Studies have shown that the surface or chemical properties of these microstructures are similar to those of native bone, affecting how cells adhere to their surface, their biochemical functions, and cell differentiation [15-17].

Several materials have been studied for the fabrication of nanostructures for biomedical applications in bone repair [18-24], including TiO2. The mechanical properties, biocompatibility, low cytotoxicity, stability in body fluids [25-30], and corrosion resistance of TiO2 give it a great potential for use in bone implants [31, 32]. Researches [15, 33-35] have shown that nanostructured TiO2 elicits a favorable molecular response and osseointegration, with better bone formation than non-nanostructured materials. However, despite advances in the development of nanostructured TiO2 systems for bone repair, review articles addressing this topic are still scanty. Therefore, this paper aims to review the characteristics and fabrication methods of nanostructured titanium dioxide and to focus on studies that evaluated its potential for bone implants.

CHARACTERISTICS OF TITANIUM DIOXIDE
Titanium dioxide is a white solid that is technologically
important due to its numerous properties [36, 37], such as low modulus of elasticity, good tensile strength, biocompatibility, and corrosion resistance, as well as its abundance [38-41]. TiO$_2$ has been used in a variety of applications, e.g., in biomedicine as catalyst support [37, 42], in water and air purification systems [17, 43], in pigments or opacifiers, in cosmetics [36, 37] and solar cells [44, 45], and in various biomedical applications [46, 47].

Under atmospheric pressure, TiO$_2$ exists in 3 polymorphic forms (Table I), known as rutile, anatase, and brookite [38, 39]. These polymorphs can be described by a Ti$^{4+}$ cation coordinated by 6 oxygen atoms, forming a distorted octahedron [45, 48, 49], whose polymorphic shapes are dictated by the way these octahedrons combine. The anatase form has a tetragonal structure and a unit cell with 4 titanium dioxide molecules (Fig. 1a). The rutile form also has a tetragonal structure and two titanium dioxide molecules per unit cell (Fig. 1b). Brookite has an orthorhombic structure and a unit cell containing 8 molecules (Fig. 1c). Compared to anatase and rutile, brookite is the least dense form of TiO$_2$ [42, 44, 50, 51]. The properties of TiO$_2$ vary according to the polymorphic phase (Table I). Rutile is the most stable phase, while anatase and brookite are metastable and can be irreversibly transformed into rutile at high temperatures. This phase transition and stability can be influenced by impurities, defects, grain size, reaction atmosphere, and synthesis conditions [49]. In addition, the stability of polymorphic phases depends on grain size. It has been reported that anatase is stable in crystallite sizes smaller than 11-45 nm due to the high surface free energy associated with this particle size [50, 52].

TiO$_2$ is an n-type semiconductor whose electronic and optical properties are influenced by the nature of the conduction band [71, 72]. The photocatalytic process is based on the absorption of UV light by titanium dioxide. When the level of photon energy is greater than or equal to that of the band gap, the photoexcited electrons jump from the valence band to the conduction band, leaving electron holes (Eq. A). In aqueous media, reactions may occur between excited TiO$_2$ and the medium. Electron holes interact with water or with the hydroxyl radical, forming hydrogen and hydroxyl ions (Eqs. B and C). Oxygen can also be adsorbed on titanium dioxide particles, reacting with electrons in the conduction band and generating superoxide (Eq. D) [37, 73-75]:

\[
\text{TiO}_2 + \text{hv}(\text{UV}) \leftrightarrow \text{h}^+ + \text{e}^-
\]  
(A)

\[
\text{h}^+ + \text{H}_2\text{O} \leftrightarrow \text{H}^+ + \cdot\text{OH}
\]  
(B)

\[
\text{h}^+ + \text{OH}^- \leftrightarrow \cdot\text{OH}
\]  
(C)

\[
\text{e}^- + \text{O}_2 \leftrightarrow \cdot\text{O}_2
\]  
(D)

where, $h^+$ and $e^-$ represent an unpaired electron, an electron

![Figure 1: Crystalline structure of: a) anatase; b) rutile; and c) brookite. Gray and white spheres represent O and Ti atoms, respectively.](image)

Table I - Properties of anatase, rutile, and brookite.

| Property                  | Anatase          | Rutile           | Brookite         | Ref.          |
|---------------------------|------------------|------------------|------------------|--------------|
| Structure                 | Tetragonal       | Tetragonal       | Orthorhombic     | [45, 53-56]  |
| Space group               | I4/amd           | P42/mnm          | Pbca             | [43, 44, 57-59] |
| Unit cell parameters (nm) |                  |                  |                  |              |
| a=0.3733-0.3785           | a=0.45936-0.4594 | a=0.917-0.9184   |                  |              |
| c=0.937-0.9515            | c=0.2953-0.29589 | b=0.5447-0.5456  |                  | [44, 45, 50, 57, 58, 60-62] |
| c=0.514-0.5154            |                  | c=0.514-0.5154   |                  |              |
| Cell volume (Å$^3$)        | 34.02-34.061     | 31.12-31.216     | 32.172-32.20     | [43, 45, 57-59] |
| Ti-O bond length (Å)       | 1.937-1.94(4)    | 1.946-1.95(4)    |                  |              |
| O-Ti-O bond angle          | 77.7-102°        | 81.1-98.9°       | 80.7-105°        | [45, 57, 58, 63] |
| Specific density (g/cm$^3$)| 3.79-3.83        | 4.13-4.25        | 3.99-4.170       | [44, 50, 54, 59, 62] |
| Band gap energy (eV)       | 3.20-3.30        | 3.01-3.02        | 3.13-3.26        | [41, 61, 64-70] |
hole in the valence band, and an electron in the conduction band, respectively, and hv is the photon energy [74]. Rutile and anatase are photoactive, and their electrons are excited in the range of ultraviolet radiation [72, 76-79]. The structure of anatase has higher photocatalytic activity than that of rutile [53, 80], because of its greater density of localized states and its lower electron-hole recombination rate than rutile [74].

Nanostructured TiO$_2$ has attracted great attention in recent years because it can be used in various fields of science and technology [81-87]. At least one of the dimensions of nanostructured TiO$_2$ polymorphs ranges from 1 to 100 nm. Depending on the field of science and technology, materials that have one of its dimensions larger than 100 nm and smaller than 1000 nm are also referred to as nanometric or are considered submicrometric. Nanometric materials, which have a high surface area to volume ratio, comprise thin films, nanotubes, nanowires, nanostrips, nanofibers, and nanoparticles [72, 78, 88]. Nanostructured TiO$_2$ has been described as a promising non-resorbable material for use in bone implants [41, 89, 90]. Recent studies [16, 21, 91-94] indicate that nanoscale modification of the TiO$_2$ surface topology can positively influence cell behavior, improving implant osseointegration and aiding greater initial biointegration with surrounding tissue [95, 96]. Bone is a nanostructured compound of collagen and hydroxyapatite fibers [14, 17, 97, 98]. Hence, nanoscale surface topography mimics the topographic characteristics of the extracellular matrix of bone tissue, providing cellular support, and in some cases, adequate porosity for the process of cell adhesion, proliferation, and differentiation during the bone formation process [29, 99, 100].

TiO$_2$ can also be used as a coating for metal implants in tissue engineering applications. Ti osseointegration can be improved by changing the surface, such as depositing a thicker layer of TiO$_2$ on the metal surface [101-103]. It has been reported that TiO$_2$ coatings manufactured on the surface of a metal alloy improve corrosion resistance, biocompatibility, and promote good cell functionality and bone formation [104-107]. Composite materials also have satisfactory properties for biomaterials due to the good combination of mechanical properties and excellent biocompatibility. Studies have shown that hydroxyapatite and titanium dioxide (HAp-TiO$_2$) composites improve mechanical resistance, osseointegration, and cell fixation due to the similarity of hydroxyapatite to various calcified tissues of vertebrates. TiO$_2$ is able to improve the bond strength of the hydroxyapatite layer and Ti substrate, as well as corrosion resistance [108-111]. Some authors [112-116] have found that the presence of hydroxyapatite increases the resistance to corrosion, the integration of implants in bone tissue, and bone growth. In addition, the HAp-TiO$_2$ composite has excellent chemical and structural uniformity.

**FORMS OF SYNTHESIS USED IN THE PRODUCTION OF BIOMATERIALS**

Nanostructured TiO$_2$ can be synthesized by several methods, including anodization [117], sol-gel [118, 119], hydrothermal [89, 120], and electrospinning [121-123]. Among these techniques, anodization and sol-gel have attracted the attention of research groups for biomedical applications, due to their simplicity, low-cost [124, 125], ability to produce nanostructures with high surface area to volume ratios [124, 126], and their ability to generate highly organized and corrosion resistant structures [12, 127]. Table II describes the characteristics of nanostructured TiO$_2$ produced by anodization and sol-gel. Several properties of nanoscale TiO$_2$ can significantly improve the performance of biomaterials, which can be affected by the method employed for their fabrication [41, 128-132]. Therefore, depending on the experimental conditions, synthesized materials have different characteristics [31, 133, 134], including pore structure and interconnectivity, nanotopography, and mechanical properties [135-139].

Anodization is a process that involves the application of an electric field between the metal and an anode, which triggers ionic diffusion, resulting in the formation of ordered nanotubes on the anode surface. The process is performed when a two-electrode system is exposed to alternating voltage in an electrolyte under controlled conditions [90, 168]. TiO$_2$ nanotubes can be grown in aqueous hydrogen fluoride (HF) electrolytes containing acid [144, 150, 153] or in neutral mixtures with added fluoride salts [143, 146, 151]. The voltage applied in the anodization process affects the TiO$_2$ nanotube diameter. Several studies [142, 147-149, 152] reported that the TiO$_2$ nanotube diameter increases in response to increasing voltage (Fig. 2). In addition, as the applied voltage is increased, the surface morphology changed from particulate to tubular. Recently, other studies [105, 140, 141] also demonstrated that different TiO$_2$ nanotube diameters can be obtained by controlling the voltage applied in the anodization process. TiO$_2$ nanotube diameter can also be influenced by the type of electrolyte. In an early study [154], titanium metal sheet in phosphate and fluorine electrolytes was used to form nanotubes; the results indicated that the nanotube diameters were 50 and 100 nm using NH$_4$F and H$_3$PO$_4$ electrolytes, respectively. In another study [145], TiO$_2$ nanotubes were produced by anodizing titanium metal sheet in NH$_4$F and H$_3$PO$_4$ electrolyte; the authors also stated that changing the electrolyte from NH$_4$F to H$_3$PO$_4$ caused the nanotube diameter to increase from 174 nm to 235.3 nm.

In the sol-gel method, a solution transitions to a gel state that can solidify into an interconnected network of particles. The solution may be formed by means of hydrolysis and polymerization reactions [124, 126]. The firing temperature of gels affects the morphology of nanostructured TiO$_2$. Several studies [56, 155, 156, 158-161, 163-165, 167] reported that firing at up to 550 °C resulted in spherical particles, and only anatase phase was found. At temperatures above 600 °C, larger particles and development of the rutile phase were observed [56]. Peaks of the rutile phase observed after heating at 700 °C were attributed to anatase to rutile phase transition [157, 162, 163]. The morphology of
nanostructures can also be influenced by the ratio between quantities of reactants, notably the ratio of ceramic precursor to water. Nanocrystalline titanium dioxide powders were synthesized by the sol-gel method, varying the rate of hydrolysis [163]. The nanoparticles were synthesized by hydrolysis using as precursor 20 mL of titanium isopropoxide mixed with 100 mL of ethanol and different amounts of water to control the hydrolysis rate. The results indicated that powder prepared with 2 mL of water contained very fine anatase crystals and quasi-spherical nanoparticles. Upon increasing the amount of water to 5 and 10 mL, spherical and homogeneous structures were obtained in response to the increase in hydrolysis rate, which proves that increasing the amount of water facilitates particle formation [163]. TiO$_2$ nanoparticles were also produced by the sol-gel method using 0.5 g of titanium tetrabutoxide, 50 mL of diethylene

| Method               | Calcination temperature | Morphology | Phase        | Size                                | Ref. |
|----------------------|-------------------------|------------|--------------|-------------------------------------|------|
| Anodization of Ti    | 530 °C                  | Nanotube   | Anatase      | Nanotube diameter: 70-110 nm        | [105]|
| Anodization of Ti    | 450 °C                  | Nanotube   | Anatase      | Nanotube diameter: 160-200 nm       | [140]|
| Anodization of Ti    | 420 °C                  | Nanotube   | Anatase      | Nanotube diameter: 45-60 nm         | [141]|
| Anodization of Ti    | 450 °C                  | Nanotube   | Anatase      | Nanotube diameter: 26-115 nm        | [142]|
| Anodization of Ti    | 450 °C                  | Nanotube   | Anatase      | Nanotube diameter: 65.7-87 nm       | [143]|
| Anodization of Ti    | 750 °C                  | Nanotube   | Anatase, rutile | Nanotube diameter: 50-70 nm   | [144]|
| Anodization of Ti    | -                       | Nanotube   | -            | Nanotube diameter: 174.2-235.3 nm    | [145]|
| Anodization of Ti    | 450 °C                  | Nanotube   | Anatase      | Nanotube diameter: 45 nm            | [146]|
| Anodization of Ti    | 500 °C                  | Nanotube   | Anatase      | Nanotube diameter: 30-90 nm         | [147]|
| Anodization of Ti    | 480 °C                  | Nanotube   | Anatase      | Nanotube diameter: 40-80 nm         | [148]|
| Anodization of Ti    | 450 °C                  | Nanotube   | Anatase      | Nanotube diameter: 20-120 nm        | [149]|
| Anodization of Ti    | 500 °C                  | Nanotube   | Anatase      | Nanotube diameter: 80-100 nm        | [150]|
| Anodization of Ti    | 580 °C                  | Nanotube   | Anatase      | Nanotube diameter: 165 nm           | [151]|
| Anodization of Ti    | -                       | Nanotube   | -            | Nanotube diameter: 15-120 nm        | [152]|
| Anodization of Ti    | 480 °C                  | Nanotube   | Anatase, rutile | Nanotube diameter: 40-110 nm      | [153]|
| Anodization of Ti    | -                       | Nanotube   | -            | Nanotube diameter: 50-100 nm        | [154]|
| Sol-gel              | 400 °C                  | Nanoparticle | Anatase    | Crystallite size: 11-22 nm          | [155]|
| Sol-gel              | 500 °C                  | Nanoparticle | Anatase    | Particle size: 51.98 nm             | [156]|
| Sol-gel              | 800 °C                  | Nanoparticle | Anatase, rutile | Particle size: 47 nm  | [157]|
| Sol-gel              | 400 °C                  | Nanoparticle | Anatase    | Crystallite size: 17.7 nm           | [158]|
| Sol-gel              | 400 °C                  | Nanoparticle | Anatase    | Crystallite size: 13.8 nm           | [159]|
| Sol-gel              | 450 °C                  | Nanoparticle | Anatase    | Crystallite size: 10 nm             | [160]|
| Sol-gel              | 500 °C                  | Nanoparticle | Anatase    | Crystallite size: 19.3 nm           | [161]|
| Sol-gel              | 550 °C                  | Nanoparticle | Anatase    | Particle size: 0.8 μm               | [162]|
| Sol-gel              | 950 °C                  | Nanoparticle | Rutile     | Particle size: 1.0 μm               | [162]|
| Sol-gel              | 700 °C                  | Nanoparticle | Anatase, rutile | Crystallite size: 80 nm        | [163]|
| Sol-gel              | 500 °C                  | Nanoparticle | Anatase    | Crystallite size: 20 nm             | [163]|
| Sol-gel              | 400 °C                  | Nanoparticle | Anatase    | Crystallite size: 3.9 nm            | [164]|
| Sol-gel              | 450 °C                  | Nanoparticle | Anatase    | Particle size: 50-100 nm            | [165]|
| Sol-gel              | 350 °C                  | Nanoparticle | Anatase    | Particle size: 30-90 nm             | [166]|
| Sol-gel              | 400 °C                  | Nanoparticle | Anatase    | Crystallite size: 27.4 nm           | [167]|
| Sol-gel              | 400 °C                  | Nanoparticle | Anatase    | Crystallite size: 13.7 nm           | [56] |
| Sol-gel              | 600 °C                  | Nanoparticle | Anatase, rutile | Crystallite size: 25.8 nm        | [56] |
glycol, and different amounts of water; it was found that, as the amount of water increased from 0.5 to 5 mL, the mean particle diameter decreased from 550 to 250 nm [56].

STUDIES OF THE BIOLOGICAL PROPERTIES OF NANOSTRUCTURED TiO₂ USED IN BIOMEDICINE

As mentioned earlier herein, nanostructured TiO₂ has attracted the attention of researchers for the development of biomaterials [169-173] and implant technology [6, 71, 174-178]. Studies have shown that the cellular response is affected by the material’s topography [179-184]. Cells respond positively to nanotopography, with changes in cell morphology, cytoskeleton organization, and proliferation. This means that nanostructuring plays an important role in cellular interaction [15, 31], Table III lists several studies on nanostructured TiO₂, revealing how nanoscale topography and crystal structure affect cell-surface interactions. It has been reported [31, 191, 204-208] that nanotubular surfaces produced by anodization are able to display cellular responses and simulate a bioactive matrix for cellular accommodation. This is due to increased surface wettability and selective adsorption of proteins in body fluids [12, 47]. TiO₂ nanotubes have good biocompatibility due to their low cytotoxicity, good stability, and cytocompatibility, including adhesion, proliferation, and differentiation of osteoblasts with high surface area to volume ratio [209]. In this context, the TiO₂ nanotube surface favors osteoblast adhesion and exhibits a strong ability to bind to bone [210]. Moreover, it facilitates fluid exchange, which is responsible for improving molecular signaling in bone remodeling and functioning characteristics [211].

Several properties can affect cellular responses and protein adsorption in biological systems [47, 212]. Hydrophilicity and surface roughness [101, 213-216], associated with the high surface area to volume ratio characteristic of nanostructured materials, have been shown to promote high cell adhesion, proliferation, and differentiation [217-220]. Numerous studies [16, 17, 174, 185, 189-191, 195] have shown that nanoscale TiO₂ nanotube surfaces present uniform structure, roughness, and hydrophilicity, with contact angles smaller than 55°. Porosity is essential for bone implants, being responsible for the formation of bone tissue, proliferation of osteoblasts, vascularization, and bioactivity. In addition, a porous structure provides good mechanical stability due to the better mechanical interlock between the biomaterial implant and the natural bone [203, 221, 222]. Some studies [223-227] report that a porous structure significantly accelerates cell adhesion and bone growth capacity. The pores must be interconnected with dimensions between 200 to 500 μm to meet the migration and transport requirements of the cell, providing adequate space and permeability for viable bone formation in nanostructured TiO₂.

Nanoscale topography has been shown to be the ideal size for the best osteoblast function [194, 228, 229]. According to [198], osteoblast adhesion and propagation was improved by the topography of TiO₂ nanotubes, forming an interconnected cell structure. The nanoscale structure hastened the growth rate of MC3T3-E1 osteoblastic cells by up to 300% to 400%. Several studies [149, 174, 185, 187, 192, 194, 195] also found that cells attached to the surface of nanotubes became increasingly elongated and formed small filopodia, suggesting that they could be used for orthopedic purposes. Over the years, several researchers [144, 188, 190, 192, 193, 196] have found that the surfaces of TiO₂ nanotubes immersed in simulated body fluid (SBF) favor the formation of osteoblast-like hydroxyapatite, which is critical for osseointegration, with strong adhesion and binding [230, 231]. As noted by several authors [16, 35, 198, 232, 233], the anatase and rutile phases provide adequate atomic arrangements for the formation of hydroxyapatite. Anatase provides the best effects for cell adhesion, proliferation, and differentiation because it is more chemically reactive. Nevertheless, the rutile phase is also interesting due to its higher modulus of elasticity, hardness, and adhesive strength than anatase. Hence, the anatase phase and the mixed anatase-rutile phase are beneficial for cell growth, due to their hydroxyapatite mineralization and increased corrosion resistance. It has been reported that the crystal structure of anatase facilitated osteoblast growth and exhibited structural stability in physiological medium [149, 174, 186, 188, 190, 192, 193, 196, 198]. Moreover, it has been suggested that osteoblasts disseminate better in anatase-rutile mixtures [186, 193, 195, 234].

Alkaline phosphatase (ALP) activity serves to analyze the functionality of cells on a surface. TiO₂ nanotubes were produced through the anodization process and evaluated for bone cell interactions using ALP [195]. During each day of analysis, the cells showed a filamentous network-like structure scattered over the entire surface. It was reported that after the 5th day of cultivation, the ALP level exceeded the titanium substrate [195]. It has been pointed out that ALP in osteoblasts gradually increased on the surface of TiO₂ nanotubes as the number of days of cultivation increased,
Table III - Studies of the biological properties of nanostructured TiO$_2$.

| Method         | Phase   | Type of cell | Type of test                                                                 | Cell response                                                                                         | Ref.       |
|----------------|---------|--------------|-------------------------------------------------------------------------------|--------------------------------------------------------------------------------------------------------|------------|
| Anodization    | -       | MG-63        | Contact angle, atomic force microscopy, *in vitro* assay                     | Hydrophilicity; cells exhibit rounded morphology with small filopodia on the surface; high adhesion of MG-63 cells to TiO$_2$ nanotubes | [185]      |
| Anodization    | Anatase | MC3T3        | Contact angle, alkaline phosphatase (ALP) activity                          | Hydrophilicity; after 7 days of culture, ALP of MC3T3 grown in different samples exceeded the Ti substrate | [177]      |
| Anodization    | Anatase | G8080        | Contact angle, cell proliferation                                           | Hydrophilicity, with cells favoring cell proliferation                                               | [186]      |
| Anodization    | Anatase + rutile | G8080 | Contact angle, cell proliferation                                           | TiO$_2$ nanotubes less hydrophilic, with more stimulated proliferation, compared to that containing only anatase phase | [186]      |
| Anodization    | -       | MC3T3-E1     | Contact angle, atomic force microscopy, *in vivo* assay                     | Hydrophilicity; surface grown osteoblastic cells exhibit a well-distributed cytoskeleton organization; *in vivo* tests indicate that the anodized surface of the implant promotes osseointegration and greater bone binding strength than pure Ti substrate | [187]      |
| Anodization    | Anatase | hBMSCs       | Contact angle, immersion in SBF, *in vitro* assay                          | Hydrophilicity; formation of hydroxyapatite after 21 days in simulated body fluid (SBF)               | [188]      |
| Anodization    | Anatase | -            | Contact angle                                                              | Hydrophilicity                                                                                         | [189]      |
| Anodization    | Anatase | MC3T3-E1     | Contact angle, atomic force microscopy, immersion in SBF                    | Hydrophilicity; formation of hydroxyapatite after 12 days in SBF                                       | [190]      |
| Anodization    | Anatase + rutile | SAOS-2 | Contact angle, cell proliferation, fluorescence microscopy                | Hydrophilicity; cells adhere to all the surfaces; seeded osteoblasts promote adhesion                | [191]      |
| Anodization    | Anatase + rutile | -    | Immersion in SBF                                                           | Formation of hydroxyapatite after 7 and 14 days in SBF                                               | [144]      |
| Anodization    | Anatase | G-292        | Scanning electron microscopy, immersion in SBF                             | Cells attached to the surface of nanotubes became increasingly elongated and formed filopodia; formation of hydroxyapatite after 5 and 10 days in SBF | [192]      |
| Anodization    | Anatase | MC3T3-E1     | Immersion in SBF, *in vitro* assay                                         | Formation of hydroxyapatite after 3 days in SBF, with size increasing as a function of immersion time | [193]      |
| Anodization    | Anatase + rutile | MC3T3-E1 | Immersion in SBF, *in vitro* assay                                         | Formation of hydroxyapatite after 3 days in SBF; a greater amount of hydroxyapatite was formed in the synthesized sample containing the anatase and rutile mixture than in the sample containing only anatase | [193]      |
| Anodization    | Anatase + rutile | MC3T3-E1 | Atomic force microscopy, confocal laser scanning microscopy, Alizarin Red staining protocol | Cells grown on nanotubes disperse better and put forth more filopodia than smooth layers; cytoskeleton of osteoblasts grown on anatase display a regular arrangement | [149]      |
| Anodization    | -       | HSCs         | Scanning electron microscopy, cell proliferation                           | Cells normally disperse on nanotubes, forming several filopodia                                       | [194]      |
| Anodization    | Anatase | MC3T3-E1     | Atomic force microscopy, contact angle, alkaline phosphatase (ALP) activity | Osteoblasts grown on the surface of TiO$_2$ nanotubes showed several filopodia extending along the main edges; hydrophilicity; extremely elongated cell morphology and high levels of alkaline phosphatase | [174]      |
| Anodization    | Anatase + rutile | OPC1   | Atomic force microscopy, contact angle, alkaline phosphatase (ALP) activity | Hydrophilicity; filamentous network-like structure with excellent cell to cell connection; ALP in osteoblasts was high after the 5th day of culture | [195]      |
| Anodization    | Anatase | -            | Immersion in SBF                                                           | Formation of hydroxyapatite after 2 days in SBF                                                       | [196]      |
| Anodization    | -       | G8080        | Alkaline phosphatase activity, *in vitro* assays                           | Approximately 50% increase in ALP levels on nanotube surfaces after 3 weeks of culture; TiO$_2$ nanotubes did not cause chronic inflammation or fibrosis under *in vivo* conditions | [197]      |
| Anodization    | Anatase | MC3T3-E1     | Alkaline phosphatase activity, *in vivo* microscopy                        | Cells grown on TiO$_2$ nanotubes presented filopodia; cell growth accelerated by up to 300-400%    | [198]      |
| Sol-gel        | Anatase | -            | Atomic force microscopy                                                   | Ca-doped TiO$_2$ gels showed good adhesion                                                            | [199]      |
| Sol-gel        | Anatase | -            | Immersion in SBF                                                           | Formation of hydroxyapatite after 28 days in SBF                                                     | [200]      |
| Sol-gel        | Anatase | -            | Immersion in SBF                                                           | Formation of hydroxyapatite after 2 days in SBF                                                       | [201]      |
| Sol-gel        | Anatase | -            | Immersion in SBF                                                           | Amount of hydroxyapatite formed after 14 days in SBF was greater in the gels containing only anatase than in those containing anatase and rutile mixture | [202]      |
| Sol-gel        | Anatase + rutile | -    | Immersion in SBF                                                           | Formation of hydroxyapatite after 14 days in SBF                                                      | [202]      |
| Sol-gel        | -       | -            | Immersion in SBF                                                           | Formation of hydroxyapatite after 3 to 6 days in SBF                                                  | [203]      |
showing excellent cell-to-cell binding [15, 17, 197, 198]. TiO$_2$ gels produced by the sol-gel technique can potentially be used in biomedical applications. The formation of hydroxyapatite immersed in SBF in amorphous and crystalline TiO$_2$ gels was investigated [202, 203]. TiO$_2$ gels with an amorphous structure did not induce the formation of hydroxyapatite on their surfaces in simulated body fluid, while gels with anatase or mixed anatase-rutile induced hydroxyapatite formation on their surfaces. Some authors [199-201] have discussed the influence of doping elements via the sol-gel method. They found that doping with calcium ions stimulates bioactivity, improving the formation of hydroxyapatite on the surface of nanostructured TiO$_2$.

For in vivo evaluations, it is essential for nanostructured biomaterials to be biocompatible and prevent inflammatory response. Such studies can be performed using animal models that can mimic the growth environment and bone infection. Several animals such as rats and pigs can be used for in vivo experiments, depending on research needs, proper bone size, ease of handling, and low costs [16, 235-238]. Popat et al. [197] synthesized TiO$_2$ nanotubes by anodization and used the G8080 cell line. They assessed in vivo biocompatibility on surfaces of subcutaneous implants in male Lewis rats and made a histological analysis of the tissue around the implants after 4 weeks. The results indicated that TiO$_2$ nanotubes were uniform and caused no inflammation, and that the tissue was normal and healthy. Li et al. [187] evaluated the behavior of MC3T3-E1 osteoblasts in experiments with Sprague Dawley rats. After 4 weeks they observed that the anodized implant surface promoted osseointegration. Their study demonstrated the good biological performance of TiO$_2$ nanotubes, which are able to withstand the forces they undergo during and after insertion into the bone.

CONCLUSIONS

The studies presented in this review of nanostructured TiO$_2$ demonstrate a promising perspective for tissue engineering, given the possible use of this nanomaterial in bone reconstruction. Biological responses can be controlled by modifying fabrication procedures, e.g., by applying variations in voltage, electrolytes, and using heat-treatments. Studies have shown that nanostructured TiO$_2$ immersed in simulated body fluid favors the formation of osteoblast-like hydroxyapatite. Equally important, in vivo assays have confirmed the biocompatibility and absence of inflammatory response of TiO$_2$ nanotubes, which is a material with great potential for application in bone regeneration.

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