Research advances in molecular mechanisms underlying the pathogenesis of cystic fibrosis: From technical improvement to clinical applications (Review)

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Abbreviations: CF, cystic fibrosis; CFTR, cystic fibrosis transmembrane conductance regulator

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Abstract. Cystic fibrosis (CF) is a chronic disease causing severe impairment to the respiratory system and digestive tracts. Currently, CF is incurable. As an autosomal recessive disorder, the morbidity of CF is significantly higher among Caucasians of European descent, whereas it is less pervasive among African and Asian populations. The disease is caused by identical mutations (homozygosity) or different mutations (heterozygosity) of an autosomal recessive mutation at position 7q31.2-q31.1 of chromosome 7. Diagnostic criteria and guidelines work concurrently with laboratory detection to facilitate precise CF detection. With technological advances, the understanding of CF pathogenesis has reached an unprecedented level, allowing for increasingly precise carrier screening, more effective early stage CF intervention and improved prognostic outcomes. These advances significantly increase the life quality and expectancy of patients with CF. Given the numerous improvements in the field of CF, the current review summarized the technical advances in the study of the molecular mechanisms underlying CF, as well as how these improvements facilitate the clinical outcomes of CF. Furthermore, challenges and obstacles to overcome are discussed.

1. Introduction

Cystic fibrosis (CF) is an autosomal recessive disease that can be attributed to the disrupted function of the CF transmembrane conductance regulator (CFTR) gene (1,2). Although CF predominantly affects the lungs, it is a multiorgan disease (3), affecting the pancreas, liver, kidneys (4) and intestine (5). CFTR mutation is the cause of the pathogenesis of CF (6) and CF is generally a result of the deletion of the phenylalanine at the 508th position of CFTR, which is induced by the loss of three nucleotides (7). In vertebrates, CFTR serves as a membrane protein and participates in the functions of Cl- channels (8,9). Due to its important regulatory functions, CFTR is ubiquitous throughout the body and is expressed in epithelial cells in the kidney, pancreas, airway, intestine (4), sweat glands and the male reproductive tract, where it serves a fundamental role in the transepithelial fluid (10). The number of identified CF-associated mutations are increasing, with ~1,700 CFTR mutations being previously recognized to be CF-prone (11); however, this number was re-estimated at 383 in 2019 according to the Clinical and Functional Translation of CFTR website (www.cftr2.org; date of access: 05/08/20). These potential mutations were screened by a specific criteria that determines the mutations responsible for the onset of CF: Firstly, the mutation could cause changes in the amino acid sequence, affecting both the expression level and functions of CFTR (12); secondly, the mutation introduces premature signals and exhibits a novel amino acid sequence that is absent in the normal CFTR gene (13).
The prevalence of CF varies with ethnicity (14,15). The relatively high incidence of CF among Caucasians may be attributed to their increased number (>1,400) of CFTR mutations (16). Furthermore, while 1/3,000 of Caucasians will develop CF, the incidence is lowered to 1/15,000 among the African population, further decreasing in Asian populations to 1/30,000, compared to the aforementioned two ethnic groups (15). The ratio of CF incidence between male and female is 1:1; however, the mortality rate of CF-associated lung infections is higher among female patients as they are subjected to greater deterioration of pulmonary function at puberty. These gender/age gaps have been proposed to be a result of the elevation in the hormone secretion (including estrogen) in adults, which may disrupt airway ion transport in lungs (17).

There are two major molecular subtypes of CF: Classic CF and non-classic CF. Non-classic CF refers to CF with better prognostic outcome, as certain functions of the CFTR protein are preserved, providing advantages for survival. Non-classic patients with CF have ≥1 copy of a defect CFTR gene with partially conserved CFTR protein functions. Due to the partial preservation of pancreatic exocrine functions, the symptoms of digestion disorders are less common among patients suffering from this milder type of CF. In contrast, patients suffering from classic CF have completely lost their functional CFTR protein. This subtype is characterized by persistent bacterial infection in the airways and sinuses, disrupted fat digestion due to the lack of pancreatic exocrine, male dysgenesis due to infection in the airways and sinuses, disrupted fat digestion from classic CF have completely lost their functional CFTR digestion disorders are less common among patients suffering preservation of pancreatic exocrine functions, the symptoms of partially conserved CFTR protein functions. Due to the partial.

The original description of CF can be dated back to 1938 (21). Since then, progress in the understanding of CF has been made in a step-by-step manner and the following 50 years has witnessed remarkable improvement in life expectancy and life quality among patients with CF (22), which may be attributed to technological innovations. In the late 1950s, a stimulated sweat test to diagnose patients with CF through C1 or Na levels was developed (23) based on the recognition of the altered electrolyte composition in sweat (24). The preliminary works contributed markedly to the diagnosis of CF and the understanding of CF was further promoted nearly 30 years later due to the discovery of the CFTR gene, a key mediator of CF (9), which enabled the diagnosis of CF by directly identifying 2 mutated CFTR alleles (25). Aside from improved diagnostics, numerous therapies have been applied to treat CF, including antibiotics against infections, nutritional supplementation and/or lung transplantation, through which the life expectancy of patients with CF can be significantly prolonged (26).

Despite prognostic improvements of CF, the median survival of patients with CF is <50 years (22). As described above, as the molecular mechanisms in CF are associated with ethnic and sex differences in terms of incidence rate, they can also be used to determine phenotypes of CF. Therefore, the innovation of methods for the detection and identification of CF at the molecular level will be beneficial to the diagnosis and prognosis of CF. The current review aimed to summarize the recent research advances of CF, including technical improvement in the understanding of the molecular mechanism of CF. Additionally, the increasing number of molecular markers that have the potential to improve diagnostic and prognostic outcomes of CF are discussed. Briefly, a PubMed (pubmed.ncbi.nlm.nih.gov; date of access: 13/08/2020) search was conducted using the following key words: ‘Cystic fibrosis’, ‘molecular’, ‘diagnosis’, ‘prognosis’ and ‘therapy’. Examples were chosen as long as they fulfilled one of the following eligibility criteria: i) Provided genetic information regarding the pivotal role of CFTR in CF; ii) described the latest progresses in parsing the molecular mechanisms underlying CF using novel techniques; iii) demonstrated the association between CF and other bioactive molecule (molecular chaperone) and the potential clinical implementations, including CF diagnosis and treatment.

2. Molecular mechanism underlying the CFTR mutation in CF

Molecular structure of CFTR. The molecular weight and length of the CFTR protein are 1,480 amino acids and 168,173 Da, respectively (7,12,27). The length of its coding sequence, which encodes the amino acid sequence for protein products, is 4,443 bp (28). The intron-free sequence of the CFTR transcript is 6,129 bp in length (12,28), whereas the normal allelic variant for CFTR is ~250,000 bp in length and contains 27 exons (12,28). CFTR is comprised of 5 functional domains (12): Two domains (MSD1 and MSD2) controlling membrane-spanning, which constitute the ion channel for Cl- transportation; an R domain, which exerts regulatory roles; and two domains (NBD1 and NBD2) that bind and catalyze the hydrolysis of adenosine triphosphate.

Biological functions of CFTR. The CFTR protein is positioned in the cell membrane (29) and is associated with proteins involved in the active transportation of material through the cell membrane (12,30). Specifically, CFTR regulates the movement of Cl-. Therefore, defects in CFTR gene can render the CFTR protein absent or dysfunctional, thereby blocking the transportation of Cl- to the cell surface (29,30). Additionally, aside from Cl-, CFTR regulates the epithelial Na channel (31).

Abnormalities in the CFTR protein disrupt the balance between Na and Cl- ions (30,32), which leads to changes in mucous constituents and abnormal reabsorption of H2O. This produces a layer of thick, sticky mucus that cannot be removed by cilia, which eventually causes inadequate mucociliary function and chronic infections (33). This can be fatal. In the lungs, accumulated mucus can become infested with bacteria and the chronic inflammation leads to pneumonia, resulting in deterioration with life-threatening difficulties in breathing. Given the molecular mechanisms of the deficiency of CFTR, the common symptoms of CF include severe cough and shortness of breath; however, CF can also lead to abnormal bowel movements, difficulty in gaining weight and infertility (12,29).

Classification of CFTR mutations. Based on the effects on protein translation, cellular processing or channel gating of CFTR (28,30), several different classification systems (Fig. 1) have been proposed over the years. Generally, the Class 1 mutation results in severe disease, as this mutation prevents the CFTR protein from being generated. Patients with Class 1A mutation do not synthesize any CFTR mRNA. Furthermore, patients with Class 1B produce damaged CFTR mRNA,
which cannot be converted into protein (28). In Class 2 mutations, the CFTR protein is produced; however, it is misfolded. The misfolded protein will be prevented from migrating to the cell membrane. In Class 3 mutations, channels in the CFTR protein are not properly opened due to gate defect (29,32,34). For the Class 4 mutation, while the CFTR protein is responsive to cell signaling, it is misshapen, resulting in a limited flow of Cl⁻ ions. Furthermore, in Class 5 mutations, insufficient CFTR protein is produced, leading to a reduction in the number of CFTR protein channels at the cell membrane (35). In class 6 mutations, less stable protein is prematurely degraded after it reaches the cell surface. Relatively, this mutation is less severe compared with the other mutations and, therefore, is a milder subtype. Generally, the Class 1, 2 and 3 mutations are more common and responsible for insufficiency in the organs suffering from CF.

3. Technical advances and implementations in CF

**PCR analysis of CF.** PCR is a widely used laboratory technique that allows for the semi-quantification of mRNAs. As early as 1992, allelic specific-PCR was used for detecting F508del mutation in CFTR (36). In the following decades, expression of CFTR had been verified by various models by reverse transcription-quantitative PCR (RT-qPCR). Certain implementations of RT-qPCR in CF include the following: i) In CF cell IB3-1 transduced with CFTR vectors, CFTR mRNA expression was detected using RT-qPCR, whereby the efficiency of transduction was measured (37); and ii) multiplex fluorescent RT-qPCR was used for scanning the exons to detect large CFTR rearrangements (38).

Aside from the aforementioned utilizations in CFTR detection, PCR is frequently performed to examine bacterial infection in CF; for instance, PCR was used to detect *Aspergillus fumigatus* DNA, which commonly infects the airways of patients with CF (39), in samples collected from patients with CF. Preimplantation genetic testing (PGT) is an important method to detect CF before or at pre-embryonic stages (40). The updated version of the PGT guidelines regarding CF proposed that PCR analysis should be performed to detect the causative mutation(s), along with associated polymorphisms within or near to the *CFTR* gene (41). Nevertheless, PCR has its own limitations. For instance, during unequal allelic PCR amplification, allele dropout can hamper the

![Figure 1. Different types of CFTR mutations. Generally, intact CFTR mRNA can be generated from the cell nucleus and following correct folding, sufficient amount of normal CFTR protein is transported to the cell membrane to serve as a Cl⁻ channel. In contrast, different malfunctions in this multi-step process lead to different CFTR defects. CFTR, cystic fibrosis transmembrane conductance regulator; Cl⁻, chloride ion.](image-url)
detection of CFTR mutations, as the annealing of a primer to the matched allelic sequence is predominated as compared to its mismatched counterpart (42).

Implementation of next-generation sequencing (NGS) in CF. The revolutionary innovation of sequencing technologies, including NGS platforms, allows for the detection of a broader spectrum of potential mutations in CF, particularly single-nucleotide polymorphism (43), which are hypothesized to be a cause of CF (44). NGS outperformed whole genome sequencing in terms of cost-efficiency and its accuracy is guaranteed by stringent thresholds during data processing (43). Therefore, NGS has been widely used in carrier (at-risk individual) screening, including CFTR mutation screening, to improve genetic counseling and reduce the incidence of CF among carriers' (at-risk-couples) descendants (43). In another study of methodology establishment, NGS-based expanded carrier screening, which determines variants through hybridization capture gene enrichment, identified several genetic alterations, including copy-number variants in the CFTR gene. The combination of NGS and variant interpretation achieved higher accuracy in identifying CF-associated phenotypes compared with the traditional method (23 variants screening) (45). Furthermore, by retrospectively performing NGS assays on patients with single CF mutated screened by sweat tests, all CFTR mutations were correctly detected, indicating that NGS assays were completely concordant with traditional methods (46). These reports demonstrated the effective implementation of NGS in CF detection, particularly at the early stage of the disease.

Gene editing for CF. Clustered regularly interspersed short palindromic repeats (CRISPR)/CRISPR-associated protein 9 (Cas9) is an emerging technology by which Cas9 proteins work in conjunction with guide RNA molecules and locate the site of target DNA sequence prior to cutting it out (47), following which the gap can be filled with the corrected gene sequence through the endogenous cellular regeneration. Therefore, this technology can be implemented in different single-gene-driven heritable deficiencies. With the advances in such gene editing technology, a promising future has been demonstrated in regard to the replacement of defective CFTR gene at the DNA level, through which normal CFTR function could be fundamentally restored (48). Although gene editing is still at its infant stage due to the relatively high off-target rate (49), it has demonstrated greater potential when compared to traditional CF therapies targeting (instead of editing) DNA, RNA or proteins. For instance, the functional repair of CFTR has been successfully performed in an in vitro model derived from stem cells of patients with CF, namely intestinal organoids (50). Another approach, Zinc finger nucleases-mediated gene editing, was used to correct defective CFTR in induced pluripotent stem cells (51). Notably, a mutation which could cause β thalassemia was corrected in human embryos using CRISPR/Cas9, indicating the capability of embryonic gene editing with this technique (52). Thus, we hypothesized that CRISPR/Cas9-induced gene editing in embryos threatened with potential mutations is a promising for the future treatment of CF.

4. Molecular regulators in CF

Non-coding RNAs (ncRNAs) in CF. Although CF is monogenic, the phenotypes of patients with CF are heterogeneous, which may be attributed to multiple regulators that contribute to CF pathogenesis (53). Non-coding (nc)RNAs are a type of RNA molecule that do not translate into a protein. Instead, ncRNAs exerts regulatory roles in multiple biological processes, such as translation (54), RNA splicing (55) and gene regulation (56), among which microRNA (miRNA or miR), long non-coding RNA (lncRNA) and circular RNA (circRNA) have been extensively studied. The current review discusses several ncRNAs that participate in CF pathogenesis.

miRNA and CF. Previous studies have demonstrated that miR-145, miR-494 and miR-101 directly or indirectly target and regulate CFTR (53-55). The inhibition of miR-145 through peptide nucleic acids was reported to promote the expression of CFTR, as miR-145 binds to the 3′-untranslated region (3′UTR) of the CFTR gene (56). Additionally, the interaction of miR-494/miR-101 and CFTR was verified and confirmed through luciferase reporter assays (57). Considering evidence that has demonstrated the inhibitory effects of these miRNAs on CFTR, the exacerbated pulmonary condition caused by air pollution or cigarette smoke was studied and attributed to elevated miR-101 and miR-144 (58). Although most miRNAs that directly target CFTR serve as suppressors, certain miRNAs exert indirect regulatory roles on CFTR to promote its expression (Fig. 2). For instance, in primary epithelial cells derived from CF bronchia (CFTR defect phenotype), miR-138 was reported to downregulate the expression of SIN3 transcription regulator family member A (SIN3A), a negative transcriptional regulator of CFTR (61). In a more recent study, miR-138 promoted the expression of CFTR through alleviating the repression that SIN3A imposed on CFTR.

Furthermore, miRNAs participate in other biological processes. In CF lung epithelial cells, miR-155 promoted inflammation through the inositol 5′-phosphatase 1-P13K/Akt cascade. Moreover, as chronic bacterial lung infection is a major cause of CF morbidity, exhaled breath condensate was used for miRNA profile analysis of patients with CF with microbial infection. The results demonstrated that 6 miRNAs (has-miRNA-432-5p, hsa-miRNA-3170, hsa-miR449c, hsa-miR-1276, hsa-miR-1247 and hsa-miR-548) were identified as potential biomarkers for patients with CF and chronic Pseudomonas infection (62). These reports indicated that miRNAs serve crucial functions in the pathogenesis of CF and are key diagnosis and prognosis markers for CF.

lncRNA and CF. Dysregulation of IncRNAs have been reported to be associated with chronic pulmonary infection, adaptive immune responses and inflammation in patients with CF (53). By working concurrently with several proteins, IncRNA BGas (a novel long noncoding RNA located in the intron 11 of the CFTR gene) regulated CFTR by regulating its local chromatin and DNA structure (53). Microarray profiling of IncRNAs revealed one upregulated (X-inactive specific transcript) and several downregulated (HOX Transcript Antisense RNA, Metastasis Associated Lung Adenocarcinoma Transcript 1 and Toll Like Receptor 8 Antisense RNA 1) IncRNAs in
The majority of studies have focused on the direction of miRNAs in the regulation of CFTR, whereby miRNAs lead to silence or degradation of CFTR mRNA by binding to its 3'UTR. Additionally, miRNAs inhibit the expression of certain CFTR suppressors, through which they promote the expression of CFTR mRNA. miRNA, microRNA; CFTR, cystic fibrosis transmembrane conductance regulator; UTR, untranslated region.

**Epigenetic modifications in CF.** Epigenetics is a mechanism that alters gene expression without changing the fundamental DNA sequence. The two mostly known mechanisms underlying epigenetic modifications include histone modifications and DNA methylation (69), both of which are involved in regulation of CFTR. It had been hypothesized that aside from ncRNAs, epigenetics is a contributing factor in the disease variability of CF (70). Additionally, epigenetic mechanisms have been proposed to be an activator of host defenses that induce a robust immune response (71). The association between immunity and epigenetics has demonstrated that DNA methylation at numerous gene loci in lung macrophages was responsible for the malfunction of innate immune cells in lungs with CF (72). Additionally, differentially altered DNA methylation at CpG sites was associated with lung function and their over-expression was demonstrated in numerous regulatory genes responsible for cell adhesion (for example, ETS homologous factor) and inflammatory responses (for example, baculoviral IAP repeat-containing protein 1) (73) in nasal epithelial samples from patients with CF (74). Furthermore, acetylation has been proposed to be associated with CF. For instance, the inhibition of histone deacetylase (HDAC) 7 was demonstrated to restore the function of F508del (75) and HDAC2 was reported to be responsive to defective CFTR function (76). A previous study demonstrated that microtubule deacetylase regulated cholesterol accumulation and NF-κB activation in CF cells through the HDAC6-Ac-tub cascade, which corroborated with the findings that HDAC6 may be a therapeutic site for various CF phenotypes (77). Collectively, therapeutic approaches for CF that target epigenetic mechanisms have been considered promising, as epigenetic alterations are dynamic and reversible. However, epigenetic therapy of CF disease is still at its infant stage (78).

5. Clinical applications of CF-associated molecules

**Molecular diagnosis of CF.** The association between clinical presentations and residual CFTR function has been established (79). Congenital absence of the vas deferens is established when the proportion of normal CFTR function is <10%, as this number decreases (<5%), positive sweat test results could be supported and patients might suffer from pulmonary infection when CFTR function further drops to <4.5%. The worst cases (CFTR function <1%) lead to pancreatic insufficiency, aside from the aforementioned symptoms (80). As varying molecular subtypes are associated with different phenotypes, experts from the ‘Cystic Fibrosis Foundation’ convened a panel of criteria for diagnosing CF in 1996 (81,82). Several tests, including the sweat test (83,84), nasal potential difference (NPD) (85-87), DNA screening (88,89) and a ciliary test, were recommended. The current review discussed traditional (regular) approaches (sweat test and NPD measurement) and novel methods.

The sweat test is an effective method for detecting CF, covering all age ranges (83,84). However, the application of creams and lotions within 1 day prior to sweat collection can disrupt the precision of diagnosis. The criteria for determining CF varies based on different ages. In infants up to 6 months of age, a CF diagnosis is very unlikely if the level of Cl is not >29 mmol/l. However, the possibility of establishing a
CF diagnosis increases when Cl⁻ levels range 30-59 mmol/l. Generally, the diagnosis of CF can be confirmed when this level is >60 mmol/l. The criteria vary slightly in patients aged >6 months. CF cannot be diagnosed when the Cl⁻ concentration is <39 mmol/l. When levels range 40-59 mmol/l, a higher probability of CF is expected. The diagnosis of CF can be established if the Cl⁻ levels are >60 mmol/l. Collection of a sufficient volume of sweat is required for laboratory assays, through which Na and Cl⁻ concentrations are determined. Incorrect results occur due to contamination, evaporation, insufficient sample and technical errors (83,84).

NPD measurement is used to follow-up patients with CF (85-87). NPD is generally used to evaluate the voltage between the reference electrode and the exploring electrode, which is sensitive and specific (85‑87). In vivo, NPD provides data about incorrect ion transport due to CFTR protein dysfunction in the nasal epithelial cells of patients. Ancillary test is used to verify the phenotype of patients and identify ion channel abnormality. However, specific skills are required to perform the test and interpret the results.

DNA screening can detect severe mutations, including F508del and minor mutations such as the 5T variant, and is particularly useful to detect CF in patients who are unable to perform the sweat test (88,89). This method can provide a general idea associated with the severity of the illness and can detect less severe CF variants, including azaospermia and congenital bilateral absence of the vas deferens (90). Previously, children suffering from CF‑associated metabolic disorders were classified into non‑typical or moderate type of CF. However, these indistinct categories lack stringent criteria. Therefore, this resulted in ambiguities in subtype stratification. Currently, DNA screening is widely used in newborn screening for improved stratification of the different subtypes of CF. Nevertheless, regular evaluation remains important (42). The most significant benefit of newborn screening and early diagnosis of CF is the possibility to treat disease‑prone patients prior to the occurrence of serious symptoms (91).

One of the consequences of developing CF is the chronic pulmonary infection caused by colonized bacteria at an early age. Phenotypic features associated with CF diagnosis provides information about chronic sinopulmonary disease manifestation due to many microorganism, including Staphylococcus aureus, nontypeable Haemophilus influenzae, mucoid/non‑mucoid Stenotrophomonas maltophilia, Pseudomonas aeruginosa and Burkholderia cepacia (92). These pathogenic bacteria can provoke gastrointestinal dysfunction responsible for intestinal, pancreatic, hepatic and nutritional troubles. Identification of the microorganism in patients with CF guides the path for subsequent antibiotic therapy (93). As mentioned in previous sections, microorganism detection can be performed by analyzing the expression profile of miRNAs (62) or other novel biomarkers (63). Therefore, traditional PCR analysis, microarray methods and NGS are capable of biomarkers profiling.

Molecular therapy for CF. Molecular therapy serves a crucial role in CF treatment. The current review discussed several alternatives, which are summarized in Fig. 3. In 1993, a gene therapy clinical trial was performed. The first trial focused on the nasal epithelium and adenovirus vectors containing the CFTR gene was used in an attempt to restore CFTR function (1,94‑97). The rationale behind this method was to restore the dysfunctional gene or to supplement the patient with the corrected version of the protein prior to irreversible damage (95,98,99). For this technique, the DNA has to penetrate the nucleus to be transcribed, which is the major barrier in gene therapy. In practice, gene therapy entails inhalation of a spray which delivers therapeutic DNA to the lungs. During the therapeutic process, either viral vectors (including adenovirus, lentivirus and herpes virus) or non‑viral vectors (such as plasmids) were used. The best therapeutic outcome would be the successful replacement of the defective gene in the lungs to cure CF fundamentally. In other outcomes, CF symptoms are alleviated by decelerated disease progression; specifically, to clear aberrant and excessive secretions, combat pulmonary infections and to prevent intestinal obstruction (99). Additionally, gene therapy is the first and most advanced vector system using recombinant retroviruses ex vivo. In vivo gene therapy uses vectors based on the recombinant form of adenovirus. The recent virus‑based system is an adeno‑associated system and numerous vector systems have been validated in clinical trials involving human participants. Among them, adenoviruses and adeno‑related viruses have been widely used (37,100). Aside from virus vectors, cationic lipids‑based vectors are also popular (99).

Transcript supplementation therapy using the correct version of CFTR mRNA transfected or transduced into the respective target cells has been documented (95). In this therapy, mRNA is actively producing CFTR in the cytoplasm, thereby circumventing the nuclear membrane. However, protein delivery is often ineffective and it is difficult to include natural posttranscriptional protein modifications. Additionally, RNA antisense therapy is taken into account in CF treatment. The hypothesis is to use inhaled RNA antisense to produce functional CFTR protein by inducing RNA to work more efficiently (53). Notably, nanotechnology using package miRNAs to treat CF was proven to be safe and effective. However, more research is required before applying this model to other diseases (66).

Another alternative for CFTR treatment includes modulator therapies (101), which can be categorized into two groups: Potentiators and correctors. The potentiators act on the CFTR ion channels. Therefore, these modulators are geared toward the class III subject group (gate defect), among which ivacaftor prolonged the opening of the CFTR channel, thereby facilitating Cl⁻ ion flow (102). In January 2012, the U.S. Food and Drug Association approved ivacaftor use and, currently, ivacaftor is the only licensed CFTR potentiator (103). Observational data based on clinical and in vitro studies have indicated that ivacaftor is efficient for several mutations within classes III, IV and V in rat thyroid cell lines (102‑104).

The correctors serve a key role in the transportation of nascent proteins (104). For instance, corrector lumacaftor is considered a stabilizer that increases the stability of mutated CFTR proteins, through which these proteins could be transported to the cell membrane more effectively and remain there for an extended period of time (105). These stable substrates could be further enhanced by potentiators (106). Monotherapy with lumacaftor, as a corrector, failed to demonstrate significant results in homozygous patients (106,107). Furthermore, another type of corrector, tezacaftor, demonstrated great
Figure 3. Schematic of several alternatives of molecular therapy. Comparisons were made between prior to and post-treatment. In supplement therapy, the corrected version of the CFTR protein (protein supplementation) or mRNA (RNA antisense therapy) were delivered to the cell. These exogenous molecules exerted their regulatory roles mainly in the cytoplasm. In gene therapy, packaged lentiviruses correcting the CFTR gene are directly inserted into the cell nucleus, thereby facilitating the normal transcription of CFTR mRNA. Furthermore, in modulator therapy, potentiators enhance the gating properties of malfunctioned CFTR, correctors induce correct CFTR protein folding/trafficking and amplifiers increase the production of immature CFTR protein, providing sufficient substrate for the corrector and potentiator. CFTR, cystic fibrosis transmembrane conductance regulator.
result, improving the processing and trafficking of mutated CFTR, and promoting chloride transportation in bronchial epithelial cells derived from F508del/F508del donors, which were achieved without the problems associated with lumacaftor (for example, pulmonary exacerbation and increment in weight) (108). The underlying mechanism and propriety of tezacaftor are very close to those of lumacaftor (107), as tezacaftor therapy increases Cl− transport. When combined with ivacaftor, tezacaftor is efficient in transporting the CFTR protein to its correct position on cell surfaces (109-111). Therefore, potentiators and correctors are often combined to treat patients with CF. Specifically, CFTR potentiators increase the activity of CFTR on epithelial surfaces, whereas CFTR correctors promote processing and trafficking of mutated protein. In order to restore the availability and functionality of CFTR protein in the epithelium, CFTR modulator drugs are taken orally (106).

Furthermore, the third type of modulator, which is still in development, is termed the amplifier. These modulators selectively promote cellular immature CFTR protein production, supplying correctors and potentiator with sufficient substrate (112). For instance, patients with CF and F508del mutations received gentamicin nasal drops for 14 days, which led to a 22% increase in their wild-type CFTR function (113). Additionally, curcumin was used to treat CF by potentiating the activation of CFTR (114). Currently, triple combination therapy (elixacaftor, tezacaftor and ivacaftor) outperformed dual combination therapy (elixacaftor and tezacaftor) as it promoted the Cl− and fluid transportation, thereby further increasing the beat frequency of cilia, as manifested by in vitro efficacy in F508Del/F508Del human bronchial epithelial cells (115).

6. Challenges and perspectives

Despite the fact that considerable data have been obtained in regard to the molecular mechanisms of CF, challenges still remain. Further research is required concerning the following aspects: i) Although 3-base-pair deletion and >100 related variants have been reported to account for CF pathogenesis, phenotypes of other variants, particularly those with single amino acid alterations, remain to be elucidated (41); ii) interpretation regarding molecular and genetic results of CFTR (whether specific variation should be defined as ‘disease-prone’ or ‘neutral’) has remained controversial, mainly due to the one-to-many association between CF genotypes to phenotypes (116), which result in difficulties in associating genetic information with clinical traits; iii) while gene therapies (gene editing) exhibit potential in CF treatment, the efficiency is decreased by high off-target effects (117); iv) another defect due to technical restriction is that prior to being intracelluarly de-packedaged, the transferred gene can be severely damaged by multiple natural barriers, including mucus and the immune response (118); and v) the spectrum of treatable mutations should be extended (119).

7. Conclusions

The current review summarizes the advances in the understanding of the molecular mechanisms underlying CF, the corresponding molecular regulators and their clinical implementations. Emerging technology, including NGS analysis and gene therapy, will improve the understanding of the underlying molecular mechanisms. Increasing numbers of novel molecular regulators, such as miRNAs and lncRNAs, have been reported, some of which displayed potential to be biomarkers of CF. CF diagnosis was improved by carrier screening, while newborn screening facilitates the prognostic outcome via the timely intervention of CF at the early stage. The developed understanding of molecular variants (genotypes) of CF defects have enabled the development of increasingly precise and customized CF treatments, which significantly prolonged the survival of patients with CF with novel therapies, including gene, supplementation and modulator therapies. These have demonstrated promising future for CF treatment. Although rapid progresses have been reported in the understanding and treatment of CF, improvements are required and challenges remain to be overcome.

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TW, HS and YS wrote and revised the manuscript. WC, YL, ZH and QJ contributed in drafting the manuscript. CX designed the work. All authors read and approved the final manuscript.

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The authors declare that they have no competing interests.

References

1. Klimova B, Kuc K, Novotny M and Maresova P: Cystic fibrosis revisited - a review study. Med Chem 13: 102-109, 2017.
2. Brawington JJ, Filbrandt ET, LaRosa FJ III, Ostmann AJ, Streeker LM, Szczesniak RD and Clancy JP: Detection of CFTR function and modulation in primary human nasal cell spheroids. J Cyst Fibros 17: 26-33, 2018.
3. Keiser NW, Birket SE, Evans IA, Tyler SR, Crooke AK, Sun X, Zhan B, Nellis JR, Streebele EK, Chu KK et al: Defective innate immunity and hyperinflammation in newborn cystic fibrosis transmembrane conductance regulator-knockout ferret lungs. Am J Respir Cell Mol Biol 52: 683-694, 2015.
4. Santoro D, Postorino A, Lucanto C, Costa S, Cristadoro S, Pellegrino S, Conti G, Buemi M, Magazzù G and Bellinghieri G: Cystic fibrosis: A risk condition for renal disease. J Ren Nutr 27: 470-473, 2017.
5. Dekkers JF, Wieringer CK, de Jonge HR, Bronsveld I, Janssens HM, de Winter-de Groot KM, Brandsma AM, Janssens HM, de Winter-de Groot KM, Brandsma AM, Burgel PR, Tullis E, Castaños C, Castellani C, Keeling KM and Bedwell DM: Identification of the amino acids required for Smad4 binding of the C5aR1 G protein-coupled receptor. J Biotechnol 231: 61-71, 2016.
6. Harutyunyan M, Huang Y, Yun KS, Yang F, Arora K and Naren AP: Personalized medicine in CF. From moderator development to therapy for patients with rare CFTR mutations. Am J Physiol Lung Cell Mol Physiol 314: L529-L543, 2018.
7. Pankow S, Bamberger C, Calzolari D, Martínez-Bartolomé S, Lavallée-Adam M, Balch WE and Yates JR III: ∆F508 CFTR interactome remodelling promotes rescue of cystic fibrosis. Nature 528: 510-516, 2015.
8. Liu F, Zhang Z, Csanády L, Gadsby DC and Chen J: Atomic structure of the cystic fibrosis transmembrane conductance regulator. Cell 169: 85-95.e8, 2017.
9. Rengaraju B, Thana K, La A, Pavithra K, Durairaj V, Mudge J, Langley RJ, Zhang L, Lee CC, Schilkey FD et al: Three births after preimplantation genetic diagnosis for cystic fibrosis with sequential first and second polar body analysis. J Mol Diagn 18: 3-14, 2016.
10. Van den Berghe WL, Blomme DM, de Winter-de Groot KM, Brandsma AM, Nijhof AM, Keeling KM and Bedwell DM: Identification of the amino acids required for Smad4 binding of the C5aR1 G protein-coupled receptor. J Biotechnol 231: 61-71, 2016.
11. Goetz D and Ren CL: Review of cystic fibrosis. Pediatr Ann 48: 155-161, 2019.
12. Costa C, Pruliere-Escabasse V, de Becdelievre A, Gameiro C, Golmard L, Guittard C, Bassinet L, Bienvenu T, Georges MD, Schwarz C: Aspergillus bronchitis in patients with cystic fibrosis. Mycopathologia 183: 61-69, 2018.
13. Brulé C, Rochmel J, Ménard C, Vanholle V, Melichar V, Niemann N and Schwartz C: Aquagenic wrinkling of the palms: A diagnostic clue to cystic fibrosis. J Cyst Fibros 10: 475-482, 2011.
14. Bell CJ, Dinwiddie DL, Miller NA, Hateley SL, Ganusova EE, Rengaraju B, Thana K, La A, Pavithra K, Durairaj V, Mudge J, Langley RJ, Zhang L, Lee CC, Schilkey FD et al: Three births after preimplantation genetic diagnosis for cystic fibrosis with sequential first and second polar body analysis. J Mol Diagn 18: 3-14, 2016.
15. Brennan ML and Schrijver I: Cystic fibrosis: A review of associated phenotypes, use of molecular diagnostic approaches, genetic characteristics, progress, and dilemmas. J Mol Diagn 18: 3-14, 2016.
16. Brennan ML and Schrijver I: Cystic fibrosis: A review of associated phenotypes, use of molecular diagnostic approaches, genetic characteristics, progress, and dilemmas. J Mol Diagn 18: 3-14, 2016.
17. Buck JL, Virant V, Plaza S, Dainia G, De Rycke M, Des Georges M, Fiorentino F, Harton G, Ishmukhametova A, Navarro J, et al: The improvement of the best practice guidelines for preimplantation genetic diagnosis of cystic fibrosis: Toward an international consensus. Eur J Hum Genot 27: 1298-1306, 2019.
18. Brennan ML and Schrijver I: Cystic fibrosis: A review of associated phenotypes, use of molecular diagnostic approaches, genetic characteristics, progress, and dilemmas. J Mol Diagn 18: 3-14, 2016.
19. Bell CJ, Dunwiddie DL, Miller NA, Hatesley SL, van der Aalst F, Mudge J, Langley RJ, Zhang L, Lee CC, Schilkey FD et al: Carrier testing for severe childhood recessive diseases by next generation sequencing. J Mol Diagn 18: 382-394, 2016.
20. Rengaraju B, Thana K, La A, Pavithra K, Durairaj V, Chappallali SH and Das A: Inquest of the best practice guidelines for preimplantation genetic diagnosis of cystic fibrosis: Toward a high level of consensus. Eur J Hum Genot 26: 1298-1306, 2018.
21. Beaukamp KA, Johansen Taber KA, Grauman PV, Spurka L, Lin J, Farnol SH, Svenson A, Goldberg JD and Muzzey D: Sequencing as a first-line methodology for cystic fibrosis carrier screening. Med Genet 21: 2569-2576, 2019.
48. Baker MW, Atkins AE, Cordovado SK, Hendrix M, Earley MC and Farrell PM: Improving newborn screening for cystic fibrosis using next-generation sequencing technology: A technical feasibility study. Mol Med 18: 231-238, 2016.

49. Marangi M and Pistritto G: Innovative therapeutic strategies for cystic fibrosis: Moving forward to CRISPR technique. Front Pharmacol 9: 396, 2018.

50. Hodes CA and Conlon RA: Delivering on the promise of gene editing for cystic fibrosis. Genes Dis 6: 97-108, 2019.

51. Park S and others: Off-target editing by CRISPR-guided DNA base editors. Biochemistry 58: 3727-3734, 2019.

52. Schwank G, Koo BK, Sasselli V, Dekkers JF, Heo I, Demircan T, Sasaki N, Boymans S, Cuppen E, van der Ent CK, et al: Functional repair of CFTR by CRISPR/Cas9 in intestinal stem cell organoids from cystic fibrosis patients. Cell Stem Cell 15: 653-658, 2019.

53. Crane AM, Kramer P, Bui JH, Chung WJ, Li XS, Gonzalez-Garay ML, Hawkins F, Liao W, Mora D, Choi S, et al: Targeted correction and restored function of the CFTR gene in cystic fibrosis induced pluripotent stem cells. Stem Cell Reports 4: 569-577, 2015.

54. Liang P, Xu Y, Zhang X, Huang R, Zhang Z, Lv J, Xie X, Chen Y, Li Y, et al: CRISPR/Cas9-mediated gene editing in human tripronuclear zygotes. Protein Cell 6: 363-372, 2015.

55. Saayman SM, Ackley A, Burdach J, Clemson M, Gruenert DC, Fabbri E, Tamanini A, Jakova T, Gasparello J, Manicardi A, Ramachandran S, Karp PH, Jiang P, Ostedgaard LS, Walz AE, Hassan F, Nuovo GJ, Crawford M, Boyaka PN, Kirkby S, Balloy V, Koshy R, Perra L, Corvol H, Chignard M, Guillot L, et al: Improved newborn screening for cystic fibrosis: A computer modelling study. NPJ Genom Med 4: 21, 2019.

56. Cutting GR: Cystic fibrosis genetics: From molecular understanding to clinical application. Nat Rev Genet 16: 45-56, 2015.

57. Davis PB, Drumm M and Konstan MW: Cystic fibrosis. Am J Respir Crit Care Med 154: 1229-1256, 1996.

58. De Boeck K, Vermeulen F and Dupont L: The diagnosis of cystic fibrosis. Presse Med 46: 201, 2017.

59. Schwarzenberg SJ, Hempestad SE, McDonald CM, Powers SW, Wooldridge J, Blair S, Freedman S, Harrington E, Murphy PJ, Palmer L, et al: Trans-epithelial ion transport in mice. J Vis Exp: 57934, 2018.

60. Anderson MC, Mistry I, Shi W, Ouyang M, Ye L, Liu D, Lee W, Chan I, Jackson S, et al: Predictive value of genomic screening: Cross-sectional study of cystic fibrosis in 50,788 electronic health records. NPJ Genom Med 4: 21, 2019.
90. Ferlin A and Stuppi A: Diagnostics of CFTR-negative patients with congenital bilateral absence of vas deferens: Which mutations are of most interest? Expert Rev Mol Diagn 20: 265-267, 2020.

91. Wagener JS, Sontag MK and Accurso FJ: Newborn screening for cystic fibrosis. Curr Opin Pediatr 15: 309-315, 2003.

92. O’Brien TJ and Welch M: Recapitulation of polymicrobial communities associated with cystic fibrosis airway infections: A perspective. Future Microbiol 14: 1437-1450, 2019.

93. Lyczak JB, Cannon CL and Pier GB: Lung infections associated with cystic fibrosis. Clin Microbiol Rev 15: 194-222, 2002.

94. Savant AP and McColley SA: Cystic fibrosis year in review 2016. Pediatr Pulmonol 52: 1092-1102, 2017.

95. Wilson J: Treating genes and patients. Gene Ther 27: 109-110, 2020.

96. Bessonova L, Volkova N, Higgins M, Bengtsson L, Tian S, Simard C, Konstan MW, Sawicki GS, Sewall A, Ohlmann C, Mainguy C and Reix P: Viral respiratory tract infections in young children with cystic fibrosis: A prospective full-year seasonal study. Virol J 16: 111, 2019.

97. Moss RB, Flume PA, Elborn JS, Cooke J, Rowe SM, McColley SA, Rubenstein RC and Higgins M; VX11-770-110 (KONDUCt) Study Group: Efficacy and safety of ivacaftor in patients with cystic fibrosis who have an Arg117His-CFTR mutation: a double-blind, randomised controlled trial. Lancet Respir Med 3: 524-533, 2015.

98. Arjmand B, Larijani B, Sheikh Hosseini M, Payab M, Gilany K, Goodarzi P, Parhizkar Roudsari P, Amanollahi Baharvand M and Hoseini Mohammadi NS: The horizon of gene therapy in modern medicine: Advances and challenges. Adv Exp Med Biol 1247: 33-64, 2020.

99. Yang Q, Soltis AR, Sukumar G, Zhang X, Caohuy H, Freedy J, Krainer G, Schenkel M, Hartmann A, Ravamehr-Lake D, Wainwright CE, Elborn JS and Ramsey BW: Lumacaftor-ivacaftor combination lumacaftor and ivacaftor therapy in patients with cystic fibrosis homozygous for the F508del-CFTR mutation (PROGRESS): A phase 3, extension study. Lancet Respir Med 5: 107-118, 2017.

100. Sala MA and Jain M: Tezacaftor for the treatment of cystic fibrosis. Expert Rev Respir Med 12: 725-732, 2018.

101. Tümmler B: Treatment of cystic fibrosis with CFTR modulators. Therapeutic targets and future approaches. J Transl Med 15: 84, 2017.

102. Faruqi S, Shiferaw D and Morice AH: Effect of ivacaftor on therapeutic activity. Biochemistry 54: 1558-1566, 2015.

103. Raynal C, Baux D, Theze C, Bareil C, Taulan M, Roux AF, Bastos C, Cullen MD, Hauck S, Tait BD, Munoz B, and Hoseini Mohammadi NS: The horizon of gene therapy in modern medicine: Advances and challenges. Adv Exp Med Biol 1247: 33-64, 2020.

104. McColley SA, Rubenstein RC and Higgins M; VX11-770-110 (KONDUCt) Study Group : Efficacy and safety of ivacaftor or F508del/G551D-CFTR or F508del/G551D-CFTR. Am J Respir Crit Care Med 197: 214-224, 2018.

105. Taylor-Cousar JL, Munck A, McKone EF, van der Ent CK, Moeller A, Simard C, Wang LT, Ingenito EP, McKee C, Lu Y, et al: Tezacaftor-Ivacaftor in Patients with Cystic Fibrosis Homozygous for Phe508del. N Engl J Med 377: 2013-2023, 2017.

106. Giuliano KA, Wachi S, Drew L, Dukowski D, Green O, Bastos C, Cullen MD, Hauck S, Tait BD, Munoz B, et al: Use of a high-throughput phenotypic screening strategy to identify amplifiers, a novel pharmacological class of small molecules that exhibit functional synergy with potentiators and correctors. SLAS Discov 23: 111-121, 2018.

107. Gambari R, Breviglieri G, Salvatori F, Finotti A and Borgatti M: Therapy for cystic fibrosis caused by nonsense mutations. Cystic Fibrosis in the Light of New Research Ch. 13, 2015.

108. Wang G: Interplay between inhibitory ferric and stimulatory curcumin regulates phosphorylation-dependent human cystic fibrosis transmembrane conductance regulator and DeltaF508 activity. Biochemistry 54: 1558-1566, 2015.

109. Chaudary N: Triplet CFTR modulators: Future prospects for treatment of cystic fibrosis. Ther Clin Risk Manag 14: 2375-2383, 2018.

110. Raynal C, Baux D, Theze C, Bareil C, Taulan M, Roux AF, Claustres M, Tuffery-Giraud S and des Georges M: A classification model relative to splicing for variants of unknown clinical significance: Application to the CFTR gene. Hum Mutat 34: 774-784, 2013.

111. Mention K, Santos L and Harrison PT: Gene and base editing as a therapeutic option for cystic fibrosis-learning from other diseases. Genes (Basel) 10: 387, 2019.

112. Osman G, Rodriguez J, Chan SY, Chisholm J, Duncan G, Kim N, Tatler AL, Shakesheff KM, Hanes J, Suk JS and Dixon JE: PE-Gylated enhanced cell penetrating peptide nanoparticles for lung gene therapy. J Control Release 283: 35-45, 2018.

113. Condren ME and Bradshaw MD: Ivacaftor: A novel gene-based therapeutic approach for cystic fibrosis. J Pediatr Pharmacol Ther 18: 8-13, 2013.

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