Identification of Polyphenolics from *Loranthus globosus* as Potential Inhibitors of Cholinesterase and Oxidative Stress for Alzheimer’s Disease Treatment

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Received 15 July 2021; Accepted 21 October 2021; Published 10 November 2021

Academic Editor: Juan Carlos Corona

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Mistletoes are considered to be the potential medicinal herbs due to their rich traditional uses. *Loranthus globosus* is a Bangladeshi mango mistletoe that has been reported as folk medicine for various ailments and diseases. In an attempt to explore its effectiveness in Alzheimer’s disease (AD), we investigated the antioxidant and acetylcholinesterase inhibitory activity of *L. globosus*. We report that the crude methanol extract (CME) of the plant contains a good amount of polyphenolics and possesses antioxidant and cholinesterase inhibitory activity. Fractionation of CME with solvents of varying polarity revealed the highest activity and polyphenolic content in the ethylacetate fraction (EAF). Correlation analysis revealed a significant (*P* < 0.05) association of polyphenolics with the antioxidant and cholinesterase inhibitory properties. Using column chromatography with diaion resin, the polyphenolics (EAF-PP) were isolated from the EAF that displayed the potent antioxidant and cholinesterase inhibitory activities. Kinetic analysis showed that EAF-PP exhibited a competitive type of inhibition. A total of thirty-six compounds including catechin and its different derivatives were identified in the EAF-PP by LC/MS analysis. Bioactivity-guided separation approach afforded the isolation of the two major active compounds catechin and catechin dimer from the EAF-PP. Hence, EAF-PP represents a potential source of antioxidants and cholinesterase inhibitors, which can be used in the management of AD.

1. Introduction

Among the neurodegenerative disorders, AD is the most devastative disorder of the aged people characterized by deficit of memory and cognition, psychobehavioral disturbances, and functional disabilities [1]. According to the recent estimate, approximately 36 million people are affected by AD around the world and it is projected that the number will be tripled by 2050 [2, 3]. Until now there is no effective treatment of this disorder. Therefore, exploration of new drugs for prevention of AD has become a priority now.

A common mechanism observed in the pathological process of neurodegenerative disorders including AD is oxidative stress, which originates due to disruption of balance between the oxidants and antioxidant system [4]. An increased production of reactive oxygen species (ROS), reactive nitrogen species (RNS), oxidation of lipid, protein, DNA and RNA, and glycoxidation have been observed in the
brains of individuals with AD [5, 6]. The increased oxidative stress in the brain is largely due to the abundance of polyunsaturated fatty acids which are exposed to oxidation by free radicals or ROS. It is increasingly evident that Abeta protein, which is overproduced in AD, can generate ROS and free radicals and leads to peroxidation of lipid in the neuronal membrane [7]. This in turn disrupts the membrane integrity of the neuronal cells and causes cellular dysfunction [8]. Antioxidant therapies have been found to be effective in ameliorating the oxidative damage and improve the memory and cognitive function in the experimental model of AD [9]. In recent times, natural antioxidants have attracted intense interest because they are safe and display a diverse biological activity.

Current treatment of AD is mainly based on the restitution of the normal concentration of acetylcholine in the synaptic cleft through inhibition of acetylcholinesterase. Acetylcholine is a neurotransmitter secreted from the cholinergic neuron that plays a role in memory and cognition, which become deficient in AD [10]. According to cholinergic hypothesis, the deficiency of acetylcholine is correlated well with the cognitive decline and severity of dementia in AD [11]. Cholinesterases, acetylcholinesterase, and butyrylcholinesterase are the enzymes that catalyze the degradation of acetylcholine. Hence, inhibitors of acetylcholinesterase can increase the endogenous level of acetylcholine in the brain and improve cholinergic neurotransmission. Currently, four drugs have been licensed to treat AD; three of them are acetylcholinesterase inhibitors: donepezil, rivastigmine, and galantamine [12]. These medications improve the symptoms for most patients with AD, but they are not able to completely stop or change the course of the illness [13]. Therefore, the researchers have been trying to develop an agent for treatment of AD that would target both the AChE and oxidative stress [14]. Medicinal plant has long been recognized as an alternative medicine and a rich source of novel drugs with potential therapeutic activity. Due to requirement of multitargeted agent for AD, plant has been recognized as an alternative medicine and a rich source of novel drugs with potential therapeutic activity. Due to the compounds responsible for activity.

2. Materials and Methods

2.1. Chemicals. 2,2-Diphenyl-1-picrylhydrazyl (DPPH), 5,5′-dithio-bis-(2-nitro) benzoic acid (DTNB), thiobarbituric acid (TBA), 2-deoxy-D-ribose, acetylthiocholine, S-butyrithiocholine, donepezil, and galantamine were procured from Sigma-Aldrich, Germany. silica gel GF254, silica gel 60-120, Folin-Ciocalteu reagent, Tris-HCl, aluminum chloride, potassium ferricyanide, ammonium molybdate, tri-chloroacetic acid (TCA), and triton X-100 were from Merck, Darmstadt, Germany. Catechin, gallic acid, and ascorbic acid were purchased from Wako Pure Chemical Company Ltd., Osaka, Japan. Petroleum ether, chloroform, ethylacetate, and methanol were obtained from Active Fine Chemicals Limited, Dhaka, Bangladesh. Unless specified, all other chemicals were of analytical grade.

2.2. Collection and Extraction of Plant Materials. The fresh barks of the plant L. globosus were collected from the campus of Rajshahi University in February 2017. The plant was authenticated by an expert of the Department of Botany, Rajshahi University, and a voucher specimen (accession no. 98) has been deposited at the herbarium of the department.

The plant material was washed with distilled water, cut into small pieces, shade dried, and ground to coarse powder by grinding machine. The powder (1.5 kg) was immersed in methanol for several days with occasional stirring. It was then filtered and concentrated in vacuo with a rotary evaporator to yield the semisolid mass. The crude methanolic extract (CME, 61.8 g) was then suspended in 10% methanol and successively partitioned with petroleum ether (PEF), chloroform (CLF), ethylacetate (EAf), and water (AQF) using separating funnel as described earlier [28]. The yield corresponding extracts were 5.3 g, 8.5 g, 28 g, and 20 g, respectively. They were stored at 4°C in a refrigerator until further use.

2.3. Phytochemical Analysis

2.3.1. Total Phenolic Content (TPC). The TPC of the extract and fractions from L. globosus was estimated by the Folin-Ciocalteu method as described by Singleton [29]. Briefly, the extract/fraction (0.5 ml) was added to 10% Folin-Ciocalteu reagent (2.5 ml) and 7.5% sodium carbonate solution (2.5 ml) and left in the dark for 20 minutes. The absorbance was read at 760 nm by a spectrophotometer. As the standard phenolics, gallic acid was used. The phenolic content was obtained from the standard curve for gallic acid.

2.3.2. Quantitation of Total Flavonoid Content (TFC). The TFC was determined by the aluminum chloride method
concentration.

2.4. Antioxidant Activity

2.4.1. Reducing Power. Reducing power of the extract/fraction was determined following the method of Oyazu [32]. Sample at various concentrations was added to 0.2 M potassium buffer (2.5 ml) and 1% potassium ferricyanide (2.5 ml) followed by incubation at 50°C for 20 minutes. Following incubation, TCA solution (10%, 2.5 ml) was added in the reaction mixture and centrifuged at 3000 rpm for 10 minutes. The resulting supernatant (2.5 ml) was mixed with equal volume of water (2.5 ml) and 0.1% ferric chloride solution (0.5 ml). The absorbance of the solution was read at 700 nm by a spectrophotometer. As a positive control, standard antioxidant catechin was used.

2.4.2. Total Antioxidant Capacity Assay. Total antioxidant activity was estimated according to the method of Prieto et al. [33]. Briefly, the extract/fractions were added to sodium phosphate (28 mM), sulphuric acid (0.6 M), and ammonium molybdate (4 mM) followed by heating at 95°C for 90 min. It was allowed to cool, and then, the absorbance was read at 695 nm. Catechin was used as a positive control.

2.4.3. DPPH Radical Scavenging Assay. DPPH radical scavenging activity of the extract/fractions was determined by the method of Choi et al. [34]. Briefly, sample in methanol at various concentrations (6.25-100 μg/ml) was added to DPPH (0.135 mM) and left in the dark at room temperature for 30 minutes. Then, the absorbance was read at 517 nm by a spectrophotometer. Catechin was used as a positive control. The following equation was used to calculate the percent scavenging:

$$\frac{(A_{control} - A_{sample})}{A_{control}} \times 100$$

where $A_{control}$ is the absorbance of control and $A_{sample}$ is the absorbance of sample. IC_{50} values were obtained from the plot of the percentage inhibition against the compound concentration.

2.4.4. Hydroxyl Radical Scavenging Activity. Hydroxyl radical scavenging activity of the extract/fractions was estimated by the method as described earlier [35]. Sample at various concentrations was added to 1 ml reaction mixture containing 2.8 mM 2-deoxy-2-ribose, 20 mM phosphate buffer pH 7.4, 100 μM EDTA, 1 mM H_{2}O_{2}, 100 μM FeCl_{3}, and 100 μM ascorbic acid and then incubated for 1 h at 37°C. To the reaction mixture (0.5 ml), TCA (1 ml, 2.8%) and TBA (1 ml, 1%) were added and heated at 90°C for 15 minutes followed by cooling. The absorbance of the solution was read at 532 nm by a spectrophotometer against an appropriate blank solution. Catechin was used as a positive control. Similar to DPPH assay, the percent scavenging of hydroxyl radical was calculated. IC_{50} values were obtained from the plot of the percentage inhibition against the compound concentration.

2.4.5. Lipid Peroxidation Inhibitory Activity. Lipid peroxidation inhibitory activity of the extract and fractions was determined by the TBA method as described by Liu et al. [36]. Brain homogenate was used as the source of lipid which was prepared from mice by the method as described earlier [37]. In brief, brain was homogenized in 50 mM phosphate buffer (pH 7.4) and centrifuged at 10000 g at 4°C for 20 min to yield the supernatant. Sample at various concentrations was added to the brain homogenates (0.5 ml), 10 μM hydrogen peroxide (100 μl), and 0.15 M KCl (1 ml) and incubated for half an hour at 37°C. Following incubation, 15% TCA, 0.38 TBA, and 5% BHT in 2 ml of HCl (0.25 N) were mixed with the reaction mixture and heated at 80°C for 60 minutes followed by cooling. The resulting mixture was centrifuged to separate the supernatant, and the absorbance was measured at 532 nm by a spectrophotometer. As positive control catechin was used. Similar to DPPH assay, the percent inhibition of lipid peroxidation was calculated. IC_{50} values were obtained from the plot of the percentage inhibition against the compound concentration.

2.5. Cholinesterase Inhibitory Activities. The widely used Ellman method was used to determine the ability of the extract/fraction to inhibit acetylcholinesterase (AChE) and butryrylcholinesterase (BChE) [38]. Mouse brain AChE enzyme and blood BChE enzyme were prepared by the method as described earlier [37]. For AChE inhibition, acetylthiocholine iodide (ATCI) was used as substrate, and for BChE inhibition, S-butyrylthiocholine iodide (BTCl) was used. The enzymatic hydrolysis of ATCI and BTCl was monitored by following the formation of yellow 5-thio-2-nitrobenzoate anion at 412 nm using a spectrophotometer. Sample at various concentrations was incubated with enzyme solution at 37°C for 15 min to allow for inhibition and then mixed with and DTNB (1 mM) in sodium phosphate buffer (50 mM, pH 8.0). The reaction was initiated by addition of ATCl (0.5 mM), and the absorbance of the mixture was determined against a control solution. The analyses were performed in triplicate. Donepezil was used as reference AChE inhibitor, and galantamine was used as reference BChE inhibitor. The following equation was used to
calculate the percent inhibition of cholinesterase activity:

$$\left[ \frac{A_{\text{control}} - A_{\text{sample}}}{A_{\text{control}}} \right] \times 100$$

where $A_{\text{control}}$ is the absorbance of control and $A_{\text{sample}}$ is the absorbance of extract or fractions. The dose response curve obtained by plotting the percent inhibition values against test concentrations was used to calculate IC$_{50}$ values of each extract/fraction and compounds.

2.6. Isolation of Polyphenols from EAF and AQF. For isolation of polyphenolics, EAF and AQF were subjected to column chromatography separately with Dianion HP 20 resin as stationery phase followed by elution with methanol as the mobile phase according to the manufacturer’s instruction. The polyphenolics from EAF (EAF-PP) and AQF (AQF-PP) were freeze-dried and stored at 4°C until use.

2.7. Inhibition Kinetics of Cholinesterase Enzymes by EAF-PP. The kinetic mode of AChE and BChE inhibition by EAF-PP was determined by preparing a range of EAF-PP concentrations (100, 200, and 400 μg/mL) in which the concentration of the substrate (ATCI/BTCI) was varied (1.4, 0.7, 0.35, 0.175, and 0.0875 mM). With different concentration (S) of substrate (ATCI/BTCI), the velocity of the enzyme inhibition was different. The assay was carried in triplicate. Lineweaver-Burk graph was plotted from $S^{-1}$ vs. $V^{-1}$ to determine the type of inhibition [39]. From these data, $V_{\text{max}}$ (maximum reaction velocity) and $K_m$ (dissociation constant) were calculated.

2.8. LC-MS Analysis of EAF-PP. The mass spectrum of the compounds in the EAF-PP was analyzed by LC-MS (Shimadzu 8050, Shimadzu, Japan) system. For separation of compounds, the sample (10 ml) was subjected to a C18 column with silica gel 60 (Merck, Germany) as a stationary phase followed by elution with gradient system consisting of n-hexane, chloroform, and methanol. The fractions were monitored on TLC, visualized under UV light, and combined based on similar $R_f$ values. Seven major fractions (F1 to F7) were obtained, and among them, F1 and F2 showed high antioxidant and cholinesterase inhibitory activity. F1 appeared as a single spot representing a single compound C-1 (54 mg), whereas the compound 2 (18 mg) was purified from the F2 was purified by preparative thin layer chromatography on silica gel GF$_{254}$ with n-hexane-acetone (6:4) as the mobile phase.

The compound 1 was characterized as catechin by comparing its $R_f$ value with those of an authentic sample, while the compound 2 was characterized by nuclear magnetic resonance (NMR) spectroscopy. The compound was dissolved in deuterated methanol and subjected to a Jeol-Ex 400 MHz spectrometer for $^1$H- and FT-NMR 100 MHz spectrometer for $^{13}$C-NMR spectra. The structure of the compound 2 was confirmed by comparing its spectral data with the reported values in the literature [23, 41].

2.10. Statistical Analysis. All experiments were done in triplicate, and the results were reported as mean ± SD. Graph Pad Prism (version 8.0.1) and Microsoft Excel 2010 were used for the statistical and graphical evaluations. The statistical significance ($P$ value < 0.05) between the means was calculated using one-way analysis of variance (ANOVA). Correlation study was performed using Pearson correlation test.

3. Results

3.1. Phytochemical Analysis. Quantitative analysis of the CME of L. globosus and its fractions was carried out for the total content of phenolics, flavonoids, and proanthocyanidin, and the results have been presented in Table 1. The results demonstrated that the CME possesses a good amount of phenolics (336.989 ± 1.837 mg GAE/g dried extract), flavonoids (180.00 ± 2.06 mg GAE/g dried extract), and proanthocyanidin (291.00 ± 1.50 mg GAE/g dried extract). Following fractionation of the CME, the highest content was found in the EAF followed by AQF, PEF, and CLF. The total phenolic contents of EAF, AQF, PEF, and CLF were 270.466 ± 0.657, 240.932 ± 1.51, 64.803 ± 0.448, and

| Sample   | TPC mg GAE/gm dried sample | TFC mg CE/gm dried sample | TPAC mg CE/gm dried sample |
|----------|-----------------------------|---------------------------|---------------------------|
| CME      | 336.989 ± 1.837             | 180.00 ± 2.060            | 291.00 ± 1.500            |
| PEF      | 64.803 ± 0.448              | 49.714 ± 1.512            | 57.0 ± 1.500              |
| CLF      | 30.753 ± 1.55               | 5.267 ± 0.162             | 21.0 ± 1.500              |
| EAF      | 270.466 ± 0.657             | 281.715 ± 2.060           | 278.0 ± 2.291             |
| AQF      | 240.932 ± 1.510             | 60.191 ± 3.805            | 179.0 ± 0.866             |

CME: crude methanolic extract; PEF: petroleum ether fraction; CLF: chloroform fraction; EAF: ethylacetate fraction; AQF: aqueous fraction; TPC: total phenolic content; TFC: total flavonoid content; TPAC: total proanthocyanidin content; GAE: gallic acid equivalent; CE: catechin equivalent.
Figure 1: Antioxidant activities of the extract and fractions from *L. globosus*. (a) DPPH radical scavenging activity. $IC_{50}$ (µg/ml): CME, 4.156 ± 0.088; PEF, 11.223 ± 0.248; CLF, 24.617 ± 0.421; EAF, 3.130 ± 0.022; AQF, 7.975 ± 0.225; CAT, 3.41 ± 0.004. (b) Hydroxyl radical scavenging activity. $IC_{50}$ (µg/ml): CME, 15.60 ± 0.356; PEF, 26.617 ± 0.293; CLF, 31.697 ± 0.570; EAF, 12.623 ± 0.268; AQF, 22.687 ± 0.389; CAT, 11.333 ± 0.356. (c) Reducing power. At 100 µg/ml concentration, the absorbances are as follows: CME, 1.874 ± 0.014; PEF, 1.624 ± 0.036; CLF, 1.117 ± 0.116; EAF, 2.457 ± 0.034; AQF, 1.634 ± 0.006; CAT, 2.660 ± 0.032. (d) Total antioxidant capacity. At 100 µg/ml concentration, the absorbances are as follows: CME, 2.039 ± 0.129; PEF, 1.326 ± 0.009; CLF, 0.954 ± 0.025; EAF, 2.688 ± 0.008; AQF, 1.578 ± 0.098; CAT, 2.251 ± 0.151. Results are expressed as mean ± SD ($n = 3$). Means with different letters (a-f) differ significantly ($P < 0.05$). CME: crude methanolic extract; PEF: petroleum ether fraction; CLF: chloroform fraction; EAF: ethylacetate fraction; AQF: aqueous fraction; CAT: catechin.
capacity assays which re
tantioxidant catechin. DPPH scavenging activity of EAF than that of the standard hydroxyl scavenging, respectively. We have noted high absorbance of CME was increased in a dose-dependent manner (Figure 1(c)). The tron or proton. In reducing power assay, the crude extract estimated their capacity to donate elec-
several in vitro extract/fractions to scavenge the free radicals, and the results have been shown in Figures 1(a) and 1(b). In both assays, the CME showed marked activity as judged by their IC50 values. The values were 4.156 ± 0.088 μg/ml and 15.60 ± 0.375 μg/ml for DPPH and hydroxyl radical scavenging, respect-
Concentration (μg/ml) of 100 μg/ml. Among the fractions, EAF showed the highest activity and CLF, the lowest. The absorbance of EAF, AQF, PEF, and CLF was 2.457 ± 0.034, 1.634 ± 0.006, 1.624 ± 0.036, and 1.117 ± 0.116, respectively, at the same concentration. In the total antioxidant activity assay based on the capacity to reduce Mo (VI) to Mo (V), the CME exhibited good activity with absorbance of 2.039 ± 0.129 at 100 μg/ml concentration (Figure 1(d)). Similar to reducing power, the highest total antioxidant activity was found in the EAF followed by AQF, PEF, and CLF with the absorbance of 2.688 ± 0.008, 1.578 ± 0.098, 1.326 ± 0.009, and 0.954 ± 0.025, respectively. Importantly, in this assay, the EAF showed more activity than that of the antioxidant catechin.

Free radicals are reported to directly attack lipid, resulting in lipid peroxidation [42]. We evaluated the potential of the CME and its fractions to inhibit the peroxidation of lipid by the TBA method, and the results have been shown in Figure 2. An increased peroxidation of lipid was observed in the brain homogenates in the presence of hydrogen peroxide which was indicated by formation of pink color, and the CME showed considerable inhibition of lipid peroxidation with IC50 value of 56.073 ± 1.176 μg/ml. EAF exhibited the highest activity among the fractions followed by AQF, PEF, and CLF with IC50 values of 25.997 ± 0.246, 38.087 ± 0.417 μg/ml, 66.003 ± 1.754, and 85.863 ± 0.246 μg/ml, respectively. The high activity of EAF and AQF indicated that they might be effective in the inhibition of lipid caused by free radicals.

3.2. Antioxidant Activity. We evaluated the antioxidative property of the CME of L. globosus and its fractions using several in vitro models. DPPH and hydroxyl free radicals scavenging models were used to assess the capacity of the extract/fractions to scavenge the free radicals, and the results have been shown in Figures 1(a) and 1(b). In both assays, the CME showed marked activity as judged by their IC50 values. The values were 4.156 ± 0.088 μg/ml and 15.60 ± 0.375 μg/ml for DPPH and hydroxyl radical scavenging, respectively. Under the same condition, the IC50 values of the reference standard catechin were 3.41 ± 0.004 μg/ml and 11.333 ± 0.356 μg/ml, respectively. When CME was fractionated, the activity was found to be distributed in all the fractions. However, the activity was high in the EAF followed by AQF, PEF, and CLF. Their IC50 values were 3.130 ± 0.022, 7.975 ± 0.225, 11.223 ± 0.248, and 24.617 ± 0.421 μg/ml for DPPH radical scavenging and 12.625 ± 0.268, 22.687 ± 0.389, 26.617 ± 0.293, and 31.697 ± 0.570 μg/ml for hydroxyl scavenging, respectively. We have noted high DPPH scavenging activity of EAF than that of the standard antioxidant catechin.

The antioxidant activity of the extract/fractions was further evaluated by reducing power and total antioxidant capacity assays which reflected their capacity to donate electron or proton. In reducing power assay, the crude extract exhibited good reducing activity and the activity was increased in a dose-dependent manner (Figure 1(c)). The absorbance of CME was 1.874 ± 0.014 at a concentration of 100 μg/ml. Among the fractions, EAF showed the highest activity and CLF, the lowest. The absorbance of EAF, AQF, PEF, and CLF was 2.457 ± 0.034, 1.634 ± 0.006, 1.624 ± 0.036, and 1.117 ± 0.116, respectively, at the same concentration. In the total antioxidant activity assay based on the capacity to reduce Mo (VI) to Mo (V), the CME exhibited good activity with absorbance of 2.039 ± 0.129 at 100 μg/ml concentration (Figure 1(d)). Similar to reducing power, the highest total antioxidant activity was found in the EAF followed by AQF, PEF, and CLF with the absorbance of 2.688 ± 0.008, 1.578 ± 0.098, 1.326 ± 0.009, and 0.954 ± 0.025, respectively. Importantly, in this assay, the EAF showed more activity than that of the antioxidant catechin.

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3.3. Cholinesterase Inhibitory Activity. Inhibition of acetylcholinesterase (AChE) and butyrylcholinesterase (BChE) by the CME and its fractions were evaluated by the Ellman’s method [38], and the results have been shown in Figure 3. The CME exerted inhibition of both AChE and BChE in a dose-dependent manner, with IC50 values of 153.767 ± 2.409 and 155.733 ± 0.907 μg/ml, respectively (Figures 3(a)
the reference AChE inhibitor donepezil showed an IC50 of 129 μg/ml. Cholinesterase inhibitory activities of the extract and fractions from L. globosus. (a) Inhibition of AChE. IC50 (μg/ml): CME, 153.76 ± 2.409; PEF, 123.367 ± 0.306; CLF, 171.533 ± 5.478; EAF, 64.987 ± 0.669; AQF, 87.417 ± 0.610; DON, 8.351 ± 0.076. (b) Inhibition of BChE. IC50 (μg/ml): CME, 155.733 ± 0.907; PEF, 353.633 ± 3.408; CLF, 391.633 ± 4.561; EAF, 85.270 ± 0.982; AQF, 129.267 ± 1.002; GAL, 8.208 ± 0.105. Results are expressed as mean ± SD (n = 3). Means with different letters (a-f) differ significantly (P < 0.05). CME: crude methanolic extract; PEF: petroleum ether fraction; CLF: chloroform fraction; EAF: ethylacetate fraction; AQF: aqueous fraction; AChE: acetylcholinesterase; BChE: butyrylcholinesterase; DON: donepezil; GAL: galantamine.

and 3(b)). Of the fractions, highest activity was found in the EAF followed by AQF, CLF, and PEF with IC50 values of 64.987 ± 0.669, 87.417 ± 0.610, 171.533 ± 5.478, and 123.367 ± 0.306 μg/ml for AChE and 85.270 ± 0.982, 129.267 ± 1.002, 391.633 ± 4.561, and 353.633 ± 3.408 μg/ml for BChE, respectively. Under the same condition, the reference AChE inhibitor donepezil showed an IC50 of 8.351 ± 0.076 μg/ml and the reference BChE inhibitor galantamine had an IC50 of 8.208 ± 0.105 μg/ml. Taken together, the EAF and AQF display appreciable inhibition against both AChE and BChE enzymes.

3.4. Correlation between Total Phenolic, Flavonoid, and Proanthocyanidin Contents and the Antioxidant and Cholinesterase Inhibitory Activities. Since the antioxidant and cholinesterase inhibitory activity was found to be high in the fractions which are rich in polyphenolics, we hypothesized that the polyphenolics might be associated with the activity. To investigate the correlation of phenolics, flavonoids, and proanthocyanidins with the antioxidant and cholinesterase inhibitory activities, the correlation studies were performed and the results have been given in Table 2. It has been observed that the total flavonoid content is significantly correlated with DPPH radical scavenging activity ($R^2 = 0.6021$, $P < 0.05$), hydroxyl radical scavenging ($R^2 = 0.8771$, $P < 0.01$), reducing activity ($R^2 = 0.881$, $P < 0.01$), total antioxidant activity ($R^2 = 0.9267$, $P < 0.001$), lipid peroxidation inhibition ($R^2 = 0.693$, $P < 0.05$), AChE ($R^2 = 0.6274$, $P < 0.05$), and BChE ($R^2 = 0.741$, $P < 0.05$) inhibition. The phenolic content showed a significant
correlation with DPPH radical scavenging activity ($R^2 = 0.7689, P < 0.01$), hydroxy radical scavenging ($R^2 = 0.8692, P < 0.01$), reducing activity ($R^2 = 0.6179, P < 0.05$), total antioxidant activity ($R^2 = 0.7653, P < 0.01$), lipid peroxidation inhibition ($R^2 = 0.6976, P < 0.05$), and BChE ($R^2 = 0.9056, P < 0.001$) inhibition. The total proanthocyanidin content was correlated significantly with DPPH radical scavenging activity ($R^2 = 0.6612, P < 0.05$), hydroxy radical scavenging ($R^2 = 0.9015, P < 0.01$), total antioxidant activity ($R^2 = 0.8665, P < 0.01$), lipid peroxidation inhibition ($R^2 = 0.681, P < 0.05$), and BChE ($R^2 = 0.8274, P < 0.01$) inhibitory activities. The significant correlation found between polyphenolics and antioxidant and cholinesterase inhibitory activities indicated that the polyphenolics might be involved in the bioactivities.

### 3.5. Isolation of Polyphenolics from EAF and AQF and Determination of Their Activity

To further understand the role of polyphenolics, the polyphenolics EAF-PP and AQF-PP were isolated from the EAF and AQF separately by using column chromatography with diaion resin and assessed for their activities. As shown in Figure 4, EAF-PP and AQF-PP were found to possess more phenolics (366.380 ± 3.119 and 364.731 ± 3.983 mg GAE/g extract), flavonoids (357.143 ± 11.999 and 365.143 ± 10.286 mg CE/g extract), and proanthocyanidins (499.5 ± 1.5 and 328.5 ± 1.5 mg CE/g extract) and exhibited potent antioxidant and cholinesterase inhibitory activity. The $IC_{50}$ values of EAF-PP and AQF-PP were 37.617 ± 0.323 and 54.283 ± 0.289 μg/ml for AChE inhibition and 28.42 ± 0.404 μg/ml and 2.843 ± 0.123 μg/ml for BChE inhibition, respectively, while the values were 2.747 ± 0.026 and 3.777 ± 0.055 μg/ml, for DPPH free radical scavenging, and 9.736 ± 0.116 and 13.483 ± 0.150 μg/ml for hydroxyl free radical scavenging, respectively. Comparing with AQF-PP, EAF-PP was more potent in terms of antioxidant and cholinesterase inhibitory activities.

### 3.6. Analysis of Mode of Inhibition of EAF-PP

Since EAF-PP showed strong inhibition against both the AChE and BuChE, we investigated further to determine the modes of enzyme inhibition of this fraction using Lineweaver-Burke plots. Plots of AChE and BuChE inhibition by EAF-PP were linear and intersected at a point on x-axis (Figure 5). These results indicated that the EAF-PP is a noncompetitive inhibitor for both the AChE and BChE enzymes.

### 3.7. LC-MS Analysis of the EAF-PP

LC/MS is an important technique for qualitative analysis of the phytochemicals in the extract. To identify the polyphenolic compounds that are contributing to the bioactivity, the EAF-PP was analyzed by LC-MS and the compounds’ profile has been shown in Table 3 [24, 43–53]. Thirty-six compounds were tentatively assigned on the basis of m/z comprising of phenolic acids, flavonoids, and proanthocyanidins. The identified phenolic acids were gallic acid and its derivatives (ethyl gallate, gallic acid 3-O-gallate, galloyl glucose, and octa-O-galloyl glucose), p-coumaric acid and its derivatives (p-coumaroyl tartaric acid and p-coumaroyl-4-O-glucoside), quinic acid, ferulic acid, coniferin, p-amoeno benzoate, syringetin-7-O-hexoside, rosammarinic acid, sinapic acid hexoside, and linamarin gulate. The flavonoids that were detected include catechin and its derivatives (catechin-3-O-gallate, epicatechin 3,5,7-gallate, epigallocatechin 3-O-gallate, epigallocatechin dimer, and O-methylated (+) catechin gallate), kaempferol and its derivatives (kaempferol-3-O-acetylgallate and kaempferol-7-O-rhamnoside), apigenin and its derivative apigenin-7-O-glucoside, quercetin and its derivative 3,7-Di-O-methyl quercetin, naringenin and naringin-7-O-glucoside, rhamnetin, and myricetin rhamnogluicoside. Catechin dimer and catechin tetrater glucoside appeared to be the proanthocyanidin.

### Table 2: Correlation of total phenolic, total flavonoid, and total proanthocyanidin contents with antioxidant and cholinesterase inhibitory activities.

| Assays                      | Correlation coefficient ($R^2$) | TPC       | TFC       | TPAC       |
|-----------------------------|--------------------------------|-----------|-----------|-----------|
| DPPH radical scavenging     | 0.7689                         | 0.6021    | 0.6612    |
| Lipid peroxidation inhibition| 0.6976                         | 0.6930    | 0.6810    |
| Acetycholinesterase inhibition| 0.4389                         | 0.6274    | 0.5528    |
| Butyrycholinesterase inhibition| 0.9056                         | 0.7410    | 0.8274    |
| Hydroxyl radical scavenging | 0.8692                         | 0.8771    | 0.9015    |
| Reducing power assay        | 0.6179                         | 0.8810    | 0.5680    |
| Total antioxidant activity  | 0.7653                         | 0.9267    | 0.8665    |

TPC: total phenolic content; TFC: total flavonoid content; TPAC: total proanthocyanidin content.

### 3.8. Activity-Guided Isolation of Active Compound from EAF-PP

Initial identity of the compounds present in the EAF-PP prompted us to define the role of compounds in the bioactivity. We followed the activity-guided chromatographic separation approach that afforded the isolation of two major compounds 1 and 2 from the EAF-PP (Figure 6). The compound 1 was identified as catechin by direct comparison of its $R_f$ value (0.6; chloroform: ethylacetate, 1:2.5) with that of the authentic sample as well as the previously reported compound from this plant (Figure 6(a)) [18], while the compound 2 was characterized as catechin dimer by comparison of its $^1$H- and $^{13}$C-NMR spectral data with the values published in the literature (Figures 6(b)–6(d)) [22, 41]. Both the compounds 1 and 2 displayed inhibition of AChE ($IC_{50}$: 32.697 ± 0.340 and 29.06 ± 0.453 μg/ml) and BChE ($IC_{50}$: 17.510 ± 0.101 and 22.767 ± 0.162 μg/ml) enzymes as well as antioxidant activity ($IC_{50}$ for DPPH scavenging: 3.477 ± 0.084 and 2.580 ± 0.038 μg/ml) (Figure 7). Comparison of activities revealed the compound 2 to be more potent than the compound 1 in respect of acetycholinesterase inhibition and antioxidant activity. Additional works are warranted for isolation and evaluation of the remaining active compounds in the EAF-PP.

### 4. Discussion

Discovering new drugs for AD is a major challenge of the present moment. Extensive research on AD has elucidated...
Figure 4: Quantitative analysis of the EAF-PP and AQF-PP and assessment of their activities. (a) Total phenolic, total flavonoid, and total proanthocyanidin contents. (b) DPPH scavenging activity. (c) Hydroxyl scavenging activity. (d) AChE inhibitory activity. (e) BChE inhibitory activity. Activities were expressed as IC$_{50}$. Data represent as mean ± SD (n = 3). Means with different letters (a-f) differ significantly (P < 0.05). EAF: ethylacetate fraction; AQF: aqueous fraction; CAT: catechin; TPC: total phenolic content; TFC: total flavonoid content; TPAC: total proanthocyanidin content; GAE: gallic acid equivalent; CE: catechin equivalent; AChE: acetylcholinesterase; BChE: butyrylcholinesterase; DON: donepezil; GAL: galantamine; EAF-PP: polyphenols from EAF; AQF-PP: polyphenols from AQF.
numerous mechanisms of pathogenesis that offered the targets for development of therapeutics [54]. Among the targets, cholinesterase has attracted much interest due to the best therapeutic outcome. In addition, oxidative stress, which is associated with the progression of AD, has been emerged as another target to prevent and halt the progression of AD [55]. The multifactorial nature of AD necessitates the development of drug that will target both the cholinesterases and oxidative stress. Polyphenolics from natural products have received intense interest not only due to their safety and antioxidant activity, but also for other biological activities including inhibition of cholinesterase [56, 57]. Individual polyphenolic components such as resveratrol, curcumin, catechin, quercetin, and luteolin, as well as the polyphenolic mixtures from grape seed and lotus, have been reported to exert protective effects against oxidative stress-induced damage and cholinergic dysfunction in AD [20]. Loranthus globosus (Bengali name—Dhara), is a Bangladeshi mistletoe that grows on the host mango (Mangifera indica) tree and distributed throughout the country. The plant has been indicated in traditional medicine for different disorders [16, 17]. In this study, we report that L. globosus is a rich source of polyphenolics and possesses potential antioxidant and cholinesterase inhibitory activity.

Medicinal plants exhibit the biological activity due to presence of diverse secondary metabolites. Polyphenols are a major class of natural products that can be exploited as neuroprotective agents. These compounds contain hydroxyl groups that have the ability to display the antioxidant activity [58, 59]. Polyphenols exist in the plant as nonflavonoids such as phenolic acids and as flavonoids such as flavones. More often, catechin has been found to occur in the plant in association with epicatechin, which is known as proanthocyanidin [60]. In this study, we carried out a quantitative analysis of polyphenolics in the crude extract and fractions from L. globosus. We found a high content of phenolics, flavonoids, and proanthocyanidin in the crude methanolic extract (CME) (Table 1). Among the fractions of CME, the highest content appeared to be present in the EAF followed by AQF, CLF, and PEF. Several reports have shown earlier the polyphenolic constituents and the antioxidant activity of the different species of the genus Loranthus [21–27]. These results report for the first time that L. globosus has large amount of polyphenolics that might exert potential antioxidant activity.

Accumulation of evidences implicates oxidative stress in the neurotoxicity of AD [5, 6]. Oxidative stress resulting from excessive generation of Aβ-induced free radicals in neuronal cell can cause serious damage to the cell and cell death [7, 8]. Antioxidants from plant sources that counteract the free radicals by scavenging have shown effectiveness in the reduction of oxidative stress as well as oxidative stress-induced cell damage and death [9]. In this work, we evaluated the antioxidant activity of the CME and its fractions in several in vitro assays/models. The radical scavenging assay using DPPH is a rapid method for evaluation of the antioxidant activity. Our results demonstrated a strong radical scavenging activity of the CME (IC50 4.156 ± 0.088 μg/ml). Although the activity was found to be distributed in all the fractions of the CME, EAF showed the highest activity (IC50 3.130 ± 0.022 μg/ml) which was even greater than that of the standard antioxidant catechin (IC50 3.41 ± 0.004 μg/ml) used in this study (Figure 1(a)). Among the free radicals produced in the biological system, hydroxyl radical is the most reactive that affects almost all biomolecules of the cell. Our results revealed the hydroxyl radical scavenging activity of the CME and its fractions. Similar to DPPH radical scavenging, the CME and its fraction EAF exhibited good activity with IC50 of 15.600 ± 0.375 and 12.623 ± 0.268 μg/ml, respectively (Figure 1(b)). Likewise, the CME and its fractions were found to display the hydrogen and proton donating abilities as revealed from the reducing power and total antioxidant activity assays. A marked activity was observed in the EAF which was higher

![Figure 5: Lineweaver-Burk plot for inhibition of (a) AChE and (b) BChE by different concentrations of EAF-PP. Results represent the average values (n = 3). AChE: acetylcholinesterase; BChE: butyrylcholinesterase.](image-url)
than of the standard antioxidant catechin used as positive control (Figures 1(c) and 1(d)). Free radicals are well known to attack on lipids in neuronal membrane resulting in lipid peroxidation. It is extensive in AD and considered as a biomarker of oxidative stress [6, 8]. The results (Figure 2) demonstrated the potential of CME and its fractions toward the reduction of lipid peroxidation. EAF exhibited the highest activity among the extract and fractions with an IC<sub>50</sub> value of 25.997 ± 0.246 μg/ml. The antioxidative property of the EAF had considerably high when compared with the other species of Loranthus [21, 24, 26]. Taken together, EAF possesses potential antioxidant activity which might be effective in the prevention of oxidative damage in AD.

Cholinesterase inhibitors are used as the first line pharmacotherapeutics in AD. Cholinesterase inhibitors can elevate the level and activity of acetylcholine in the brain and improve memory and cognition [12]. Plant and plant-derived phytochemicals, which are used as alternative

| Sl no. | Proposed compounds                        | Mode of ionization | Observed mass (m/z) | References |
|-------|------------------------------------------|--------------------|---------------------|------------|
| 1     | Gallic acid                              | [M-H]<sup>+</sup>  | 171.0               | [41–43]    |
| 2     | Ethyl gallate                            | [M-H]<sup>+</sup>  | 221.1               | [41, 42]   |
| 3     | Gallic acid 3-O-gallate                  | [M-H]<sup>+</sup>  | 322.5               | [43]       |
| 4     | Gallloyl glucose                         | [M-H]<sup>+</sup>  | 333.0               | [43]       |
| 5     | Octa-O-gallloyl glucose                  | [M-H]<sup>+</sup>  | 1396.5              | [44]       |
| 6     | Quinic acid                              | [M-H]<sup>+</sup>  | 190.7               | [42]       |
| 7     | Ferulic acid                             | [M-H]<sup>+</sup>  | 195.1               | [42]       |
| 8     | p-Coumaric acid                          | [M-H]<sup>+</sup>  | 165.0               | [42]       |
| 9     | p-Coumaroyl tartaric acid               | [M-H]<sup>+</sup>  | 297.5               | [43]       |
| 10    | p-Coumaroyl-4-O-glucoside               | [M-H]<sup>+</sup>  | 326.4               | [43]       |
| 11    | 3-p-Coumaroylquinic acid                | [M-H]<sup>+</sup>  | 339.1               | [43]       |
| 12    | Coniferin                                | [M-H]<sup>+</sup>  | 340.6               | [42]       |
| 13    | p-Amino benzoate                         | [M-H]<sup>+</sup>  | 137.7               | [42, 43]   |
| 14    | Syringetin-7-O-hexoside                  | [M-H]<sup>+</sup>  | 509.6               | [42]       |
| 15    | Rosamarinic acid                         | [M-H]<sup>+</sup>  | 359.2               | [45]       |
| 16    | Sinapic acid hexoside                    | [M-H]<sup>+</sup>  | 384.9               | [46]       |
| 17    | Linamarin gallate                       | [M-Na]<sup>+</sup> | 421.3               | [22]       |
| 18    | Catechin                                | [M-Na]<sup>+</sup> | 311.3               | [47]       |
| 19    | (+) Catechin 3-O-gallate                 | [M-H]<sup>+</sup>  | 441.2               | [42, 43]   |
| 20    | (+) Epicatechin 3,5,7-trigallate         | [M-H]<sup>+</sup>  | 747.2               | [48]       |
| 21    | Epigallocatechin-3-O-gallate             | [M-H]<sup>+</sup>  | 459.8               | [42, 49]   |
| 22    | Epigallocatechin dimer                   | [M-Na]<sup>+</sup> | 627.5               | [49]       |
| 23    | O-methylated(+) catechin gallate         | [M-H]<sup>+</sup>  | 456.7               | [50]       |
| 24    | Kaemferol                                | [M-Na]<sup>+</sup> | 310.4               | [47]       |
| 25    | Kaemferol-3-O-acetylglicoside            | [M-H]<sup>+</sup>  | 488.5               | [42]       |
| 26    | Kaemferol-7-O-rhamnoside                 | [M-H]<sup>+</sup>  | 434.8               | [42]       |
| 27    | Quercetin                                | [M-Na]<sup>+</sup> | 325.05              | [47]       |
| 28    | 3,7-Di-O-methyl quercetin                | [M-H]<sup>+</sup>  | 329.0               | [42]       |
| 29    | Rhamnetin                                | [M-H]<sup>+</sup>  | 317.0               | [43]       |
| 30    | Apigenin                                 | [M-H]<sup>+</sup>  | 270.6               | [42]       |
| 31    | Apigenin-7-O-glucoside                   | [M-H]<sup>+</sup>  | 433.9               | [51]       |
| 32    | Naringenin                               | [M-H]<sup>+</sup>  | 272.9               | [42, 46]   |
| 33    | Naringin-7-O-glucoside                   | [M-H]<sup>+</sup>  | 434.8               | [42]       |
| 34    | Myrecetin rhamnohexoside                 | [M-H]<sup>+</sup>  | 627.6               | [46]       |
| 35    | Catechin dimer                           | [M-H]<sup>+</sup>  | 577.3               | [42, 50]   |
| 36    | Catechin tertramer glucose               | [M-H]<sup>+</sup>  | 1379                | [47]       |

Table 3: LC/MS analysis of the EAF-PP.
**Figure 6**: Characterization of the compounds isolated from the polyphenolic EAF-PP. (a) TLC profile of the compound 1 and authentic catechin. (b) $^1$H NMR spectroscopic data of the compound 2. (c) $^1$C NMR spectroscopic of the compound 2. (d) Chemical structures of the compounds 1 and 2.
Such as phenolic acids and flavonoids compounds fall into two major categories, non-flavonoids, a significant amount of the polyphenols, a separate column chromatography using diaion resin from EAF and AQF (termed as EAF-PP and AQF-PP) polyphenols in the activity, we have isolated the polyphenol-rich fractions. Therefore, to explore the association of the polyphenolics with antioxidant and cholinesterase inhibitory activities (Table 2). To ascertain the role of polyphenols in the activity, we have isolated the polyphenolics from EAF and AQF (termed as EAF-PP and AQF-PP) separately by column chromatography using diatomeous earth resin and evaluated their activities similarly. With the increase of polyphenols, a significant increase of antioxidant property and cholinesterase inhibitory activities of the EAF-PP and AQF-PP was observed, suggesting further the role of polyphenolics in the bioactivity (Figure 4). EAF-PP was found to be more potent than AQF-polyphenolics in respect of both antioxidant and cholinesterase inhibitory activities. Kinetic studies of EAF-PP revealed a noncompetitive antagonism against both the acetylcholinesterase and butyrylcholinesterase (Figure 5).

To gain insights into the compounds responsible for bioactivity, we performed LC-MS analysis of the EAF-PP, which tentatively identified 36 compounds (Table 3). These compounds fall into two major categories, nonflavonoids such as phenolic acids and flavonoids such as flavonols, flavones, flavonols, flavonones, and proanthocyanidin. Gallic acid and its derivatives, p-coumaric acid and its derivatives, quinic acid, ferulic acid, quinic acid, rosamic acid, coniferin, and linamarin gallocate were the important phenolic acids detected in the EAF-PP. The occurrence of these phenolic acids is common in fruits and vegetables, and the antioxidant activity of these compounds is well established [43–54, 67, 68]. The flavonoid gallocatechin has been found to be the major constituent of tea, grapes, lotus, and other fruits [67–69]. In these plants, catechin also occurs in combination with gallic acid such as catechin gallate, gallochatechin, and gallochatechin gallate. In addition, few other common flavonoids such as quercetin, kaempferol, apigenin, and naringenin also occur in these plants. Interestingly, all these flavonoids and their derivatives have been found in the EAF-PP. Extensive information on the antioxidant and acetylcholinesterase potential of these compounds has been documented, which suggests that EAF-PP may be used in the treatment of AD. Recently, Okello et al. [70] reported that catechin, epicatechin, and epicatechin gallate, while in a mixture, exhibited synergistic activity in terms of inhibition of cholinesterases. This can explain, in part, the strong cholinesterase inhibitory activity of the EAF-PP.

Proanthocyanidins are the oligomeric and polymeric compounds formed from catechin and epicatechin and abundant in grapes, lotus, mango, and many other fruits [60]. Proanthocyanidins have received much attention due to their potential neuroprotective properties. Proanthocyanidins from grape seed and lotus have shown significant improvement in memory and cognition through multiple mechanisms including modulating oxidative stress and cholinergic neuron function [60, 71]. In this study, we have detected proanthocyanidin molecules catechin dimer and catechin tetramer glucose in EAF-PP by LC/MS. Through bioactivity-guided approach, the two major compounds catechin and catechin dimer have been isolated and characterized from EAF-PP (Figure 6). In an earlier investigation, we reported the isolation of catechin from the same ethylacetate fraction of this plant [18] and Wong et al. characterized a catechin trimer in Loranthus parasiticus [23]. The antioxidant activity of catechin and its polymer have been studied extensively and showed that the activity is proportional with the degree of polymerization [72], which is consistent with our result (Figure 7). In the current study, we show that catechin dimer has cholinesterase inhibitory activity which shows high specificity for acetylcholinesterase than that of catechin. So far, this report appears to be the first describing the isolation of catechin and catechin dimer as the active molecules relevant to AD treatment from this species. More studies are warranted for isolation and evaluation of the remaining active compounds in the EAF-PP.
5. Conclusions

This study revealed the antioxidant and cholinesterase inhibitory activity of the crude methanolic extract of *L. globus*. The crude extract was found to contain a good amount of polyphenolics. When fractionated by solvents of different polarity, highest activity and polyphenolic content were observed in the EAF amongst the fractions. Correlation analysis showed a significant association of polyphenolic content with the antioxidant and cholinesterase inhibitory activities. The polyphenolics (EAF-PP) were isolated from the EAF that exhibited the most potent antioxidant and cholinesterase inhibitory activity. Thirty-six polyphenolic compounds were tentatively identified in the EAF-PP which were reported to have antioxidant and neuroprotective activity. Finally, two compounds catechin and catechin dimer were identified in the EAF-PP as the major active compounds. Hence, EAF-PP represents a source of potential antioxidants and cholinesterase inhibitors that may be used in the prevention and treatment of AD. Additional research in animal model of AD will be required to justify the therapeutic potential of EAF-PP.

Data Availability

The data presented in this study are available on request from the corresponding author.

Conflicts of Interest

The authors declare that there is no conflict of interests.

Acknowledgments

The research was supported in part by the Faculty of Science, University of Rajshahi, Bangladesh (grant number 425-5/52/RU/Science-5/12-13). The authors gratefully acknowledge the research facilities provided by the Faculty of Science, Rajshahi University, Bangladesh.

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