Vaccinia virus (VACV) is a member of the genus Orthopoxvirus of the Poxviridae, a family of large double-stranded DNA viruses that replicate in the cytoplasm (Moss, 2001). The VACV genome encodes approximately 200 open reading frames (ORFs) (Goebel (Moss, 2001). The VACV genome encodes approximately 200 open reading frames (ORFs) (Goebel et al., 1990) with essential genes located mostly in the central, highly conserved region of the genome and non-essential genes in the variable terminal regions (Kotwal & Moss, 1988; Perkus et al., 1991).

Three genes in the VACV genome encode BTB/kelch proteins: A55R, C2L and F3L. These proteins contain an N-terminal BTB (broad-complex, tram-track and bric-a-brac) and a C-terminal kelch domain. The kelch motif sequence is 44–56 aa long, usually occurring as four to seven repeats. Together these form a tertiary structure known as the β-propeller, with each repeat unit forming a secondary structure of four anti-parallel β-sheets representing a single ‘blade’ of the structure (Adams et al., 2000). The BTB domain mediates protein–protein interactions (Bardwell & Treisman, 1994) and often serves to homodimerize the protein or heterodimerize with other BTB domains. A number of BTB/kelch proteins act as substrate adapters for the ubiquitination machinery, down-regulating their target protein and thereby influencing a number of critical cellular pathways (Zhang & Hannink, 2003; Zhang et al., 2004; Angers et al., 2006; Salinas et al., 2006).

Poxviruses are the only viruses known to encode kelch proteins, the number of which varies between species: cowpox virus contains six kelch proteins, ectromelia virus four and monkeypox virus just one. All the kelch proteins in variola virus are missing or fragmented (Shchelkunov et al., 2002). Of the three VACV BTB/kelch proteins, A55 has five kelch repeats, F3 has four and C2 has three.

Previous studies have investigated the phenotype of recombinant VACV strains that lack either the C2L or A55R genes (Pires de Miranda et al., 2003; Beard et al., 2006). With both these mutants the viral plaque morphology was altered, infected cells produced fewer cellular projections and the characteristic Ca2+-independent adhesion of VACV-infected cells was reduced. Murine intradermal infection with either vAC2 or vAA55 produced lesions significantly larger than those caused by infection with control viruses. A role for kelch proteins in poxvirus virulence was also suggested by the reports that the sequential deletion of multiple BTB/kelch genes from cowpox virus caused a reduction in virulence (Kochneva et al., 2005), and that one or more BTB/kelch proteins were disrupted in, or lost from, attenuated strains of sheeppox virus, goatpox virus and lumpy skin disease virus (Tulman et al., 2002; Kara et al., 2005).

The goals of this project were to characterize the F3 protein encoded by VACV Western Reserve (WR), specifically to determine its effect on virus growth in vitro and virulence in vivo.

A recombinant VACV lacking the entire ORF of the F3L gene (vAF3) was generated using transient dominant selection (Falkner & Moss, 1990). The primers pmb21 (5'-CT-TAAGTTATTGCATCCACCGAGTGA-3') and pmb29 (5'-AGTCAGTCUGCCGAGTAAATAAGGT-3') were used in a PCR to generate a 5' flanking region, and pmb24 (5'-AACGCTTATTGGAGCAGACCA-3') and pmb30 (5'-GCGTACTGCTTTTATCGAATAGGT-3') were used to generate a 3' flanking region. The two fragments were then joined by an overlap extension PCR (Horton et al., 1989) (the overlapping complementary DNA regions of primers pmb29 and 30 are shown above in bold) and the resulting fragment was

Vaccinia virus gene F3L encodes an intracellular protein that affects the innate immune response

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The Vaccinia virus BTB/kelch protein F3 has been characterized and its effects on virus replication in vitro and virus virulence in vivo have been determined. The loss of the F3L gene had no effect on virus growth, plaque phenotype or cytopathic effect in cell culture under the conditions tested. However, the virulence of a virus lacking F3L in an intradermal model was reduced compared with controls, and this was demonstrated by a significantly smaller lesion and alterations to the innate immune response to infection. The predicted molecular mass of the F3 protein is 56 kDa; however, immunoblotting of infected cell lysates using an antibody directed against recombinant F3 revealed two proteins of estimated sizes 37 and 25 kDa.
cloned into pSJH7 (Hughes et al., 1991) which contains the *Escherichia coli* guanine xanthine phosphoribosyltransferase (*Ecogpt*) gene as a selectable marker, to create plasmid pPB23. Plasmid pPB23 was transfected into VACV-infected cells, and a deletion and wild-type virus were isolated as described previously (Beard et al., 2006). A revertant virus was constructed using the same method. A 1290 bp PCR product was generated from VACV WR genomic DNA using primers pmb21 and pmb24, comprising full-length F3L with flanking regions. This was cloned into pSJH7 to create plasmid pPB25. This was transfected into cells infected with vΔF3 and plaques of the revertant virus (vF3-rev) containing the full-length F3L gene were isolated. The genomes of these viruses were analysed by PCR, restriction enzyme digestion and Southern blotting using a probe specific for the F3L gene and this confirmed that each virus had the predicted genome structure (data not shown).

The isolation of vΔF3 shows that F3L is not essential for virus replication. The growth properties of vΔF3 were analysed and compared with vF3 and vF3-rev by both one-step (m.o.i. of 10) and multi-step (m.o.i. of 0.02) growth curves and no statistical difference was found (data not shown). There was no discernable difference in the morphology of plaques formed by vΔF3 on confluent BS-C-1 cells when compared to vF3 or vF3-rev (Fig. 1a, b) and the size of the plaques generated was not significantly different on RK-13, BS-C-1, TK-143 or CV-1 cell lines (data not shown). vF3, vΔF3 and vF3-rev were used to characterize the effect of F3 on various aspects of VACV-induced cytopathic effect, including the number of VACV-induced cellular projections and relative increase in cell motility (Sanderson et al., 1998), Ca2+-dependent adhesion to the extracellular matrix (ECM) (Sanderson & Smith, 1998) and the number of actin tails produced from the cell surface. In each case there was no difference found between vF3, vΔF3 and vF3-rev (data not shown).

The virulence of vΔF3 was examined in both an intranasal (Alcami & Smith, 1992) and intradermal (Tscharke et al., 2002) model of VACV infection. There was no significant difference in weight loss caused by intranasal infection with vΔF3 when compared to vF3 or vF3-rev (5 × 105 p.f.u. per mouse, data not shown); however, in the intradermal infection model (Fig. 2) vΔF3 produced significantly smaller lesions (P<0.05) than both vF3 and vF3-rev from days 6 to 9 post-infection (p.i.) (Fig. 2a). To determine the basis for this difference in viral virulence, the immune cell populations present in the ears during infection were analysed using flow cytometry as described previously (Jacobs et al., 2006). NK cells were defined as the NK1.1+CD3− population using fluorescein isothiocyanate–anti-NK1.1 monoclonal antibody (mAb) (BD Pharmingen) and phycoerythrin (PE)–anti-CD3 mAb (BD Pharmingen). TCRγδ cells were labelled with PE–anti-TCRγδ mAb (BD Pharmingen).

A rabbit polyclonal antibody to F3 was generated for protein characterization. The C-terminal 310 aa of the F3 protein (including the kelch repeats central region) was amplified from VACV WR by PCR using primers pmb39 (5′-GAATT CATGGATGAGTTATG-3′) and pmb17 (5′-AAGCTTATTTATCCATCCATA-3′), generating an *EcoR*I site (underlined) and start site (bold) at the 5′ end of the gene and a *Hind*III site at the 3′ end (italics). This product was cloned into pET28(a) (EMD Biosciences) to introduce DNA encoding a his-tag at the 5′ end of the

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**Fig. 1.** Plaque phenotypes of VACVs lacking each BTB/kelch protein. (a) Plaques produced on confluent BS-C-1 cells by vF3, vΔF3, vF3-rev infection (top row) vA55, vΔA55, vA55-rev (middle row) and vC2, vΔC2 and vC2-rev (bottom row). Infected cells were overlaid with DMEM/2.5 % fetal bovine serum/1.5 % carboxymethylcellulose for 2 days at 37 °C before being stained with crystal violet. (b) Higher magnification detail of plaque edges under phase-contrast microscopy.
ORF, creating plasmid pPB27. This plasmid was transformed into Rosetta E. coli cells (EMD Biosciences) and cultured in Luria–Bertani medium at 37°C to an OD600 of 0.6 before protein expression was induced by the addition of 1 mM isopropyl β-D-thiogalactoside for 4 h at 30°C. The his-tagged recombinant protein was purified from the insoluble fraction of transformed E. coli by denaturation of the inclusion body in 6 M guanidine hydrochloride, application of the denatured protein to Ni-NTA beads (Qiagen) and elution of the protein in 0.5 M imidazole. Purified protein was used for rabbit polyclonal antibody production (Harlan Seralabs).

The IgG fraction of the resulting polyclonal serum was used to identify the F3 protein in infected cell lysates (Fig. 3). Confluent BS-C-1 cells were infected at 5 p.f.u. per cell with vF3, vΔF3, vF3-rev or mock-infected either in the presence or absence of the proteasome inhibitor MG132 (10 μM) and lysates were analysed by immunoblotting with (a, b) anti-F3 IgG (1:1000). (b) Intentionally overdeveloped image of (a), to reveal the 25 kDa band more clearly. (c) Immunoblot of lysates from cells infected with vF3, vΔF3 or vF3-rev in the presence of MG132. (d) Anti-F13 mAb p37 (1:1000). (e) AraC (40 μg ml⁻¹) was added at time 0 as indicated and the blot was probed with anti-F3 antibody (1:1000). (f) Intentionally overexposed image of (e) to reveal the presence of the 25 kDa band.

Fig. 3. Characterization of the F3 protein. Cells were infected at 5 p.f.u. per cell or mock-infected, with or without MG132 (10 μM), and lysates were analysed by immunoblotting with (a, b) anti-F3 IgG (1:1000). (b) Intentionally overdeveloped image of (a), to reveal the 25 kDa band more clearly. (c) Immunoblot of lysates from cells infected with vF3, vΔF3 or vF3-rev in the presence of MG132. (d) Anti-F13 mAb p37 (1:1000). (e) AraC (40 μg ml⁻¹) was added at time 0 as indicated and the blot was probed with anti-F3 antibody (1:1000). (f) Intentionally overexposed image of (e) to reveal the presence of the 25 kDa band.

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Under the conditions tested, immunoblotting did not detect the F3 protein in virions purified by sucrose density-gradient centrifugation (data not shown).

All three BTB/kelch proteins encoded by VACV have now been characterized in vitro and in vivo. Previous investigations indicated strong similarities between the viruses lacking C2L or A55R, however, vΔF3 appears to be quite different. Deletion of either C2L or A55R caused an alteration in viral plaque morphology in which the edges of the plaque appear less distinct than in wild-type and revertant controls (Pires de Miranda et al., 2003; Beard et al., 2006). However, the morphology of vΔF3 plaques is indistinguishable from that of vF3 and vF3-rev (Fig. 1a, b). Loss of C2L and A55R each reduced significantly the number of cells that produce projections late during infection and reduced the switch from Ca2+-dependent to Ca2+-independent ECM interaction. In contrast, vΔF3 has no significant effect on either of these processes. These results provide the first indication that the function of F3 protein during viral infection is distinct from that of either C2 or A55.

The mild attenuation demonstrated by vΔF3 in the early stages of the intradermal infection model (Fig. 2a) is different from both vΔC2 and vΔA55, which both exhibit an increased lesion size late in infection (Pires de Miranda et al., 2003; Beard et al., 2006). The flow cytometry data presented here indicates a link between F3 and the innate immune response to virus infection. When F3L is deleted the percentage of NK cells present in the lesion is increased 4 days p.i. and the percentage of TCRγδ cells decreased 2 days later (Fig. 2b). Both these cell types function as part of the innate immune system (Hamerman et al., 2005; Born et al., 2006) and as such provide a ‘first-line’ defence against viral infection. An increase in the proportion of NK cells early during infection could be responsible for an accelerated immune response and hence earlier decline in the proportion of TCRγδ cells observed at day 6 p.i. Notably, the levels of TCRγδ cells decline after day 4 in this model (Jacobs et al., 2006). The innate immune response of the skin to VACV infection is of particular interest as intradermal or subcutaneous inoculation is the most commonly used route for administering poxvirus-based vaccines. This environment contains a number of unique immunological features such as specialized γδ-expressing T cells known as dendritic epidermal T cells (Hayday & Tigelaar, 2003) and the distinctive antigen presenting cells, Langerhans cells and dermal dendritic cells (Romani et al., 2006). Vaccinia virus is known to express many proteins involved in modulation of innate immune responses of the host, including the production of secreted, soluble decay receptors and interruption to the intracellular signalling pathways that activate the transcription factor nuclear factor (NF)-κB (Haga & Bowie, 2005). Consequently, the role of F3 in the recruitment and activation of NK cells and TCRγδ cells in the skin in response to VACV infection is the subject of further study.

Immunoblotting of infected cell lysates revealed 37 and 25 kDa proteins specific to vF3 and vF3-rev lysates (Fig. 3a), but the absence of any band attributable to full-length F3 protein (predicted Mr, 56). Further work is under way to identify the provenance of the two bands; the rabbit polyclonal antibody used was raised to a recombinant F3 protein lacking only the N-terminal 20 kDa, and therefore cannot aid in further identification of the fragments. The two polypeptides seen may be derived from a common precursor, or, less likely, might be translated from different RNAs.

Both F3-specific bands are still apparent, at a relatively reduced level, in the presence of AraC, indicating that they are both products of early gene expression. It is also notable that the relative intensity of both bands is increased in the presence of the proteasomal inhibitor MG132, indicating that either they, or factors that regulate the putative F3 cleavage, are targets for proteasomal degradation. Previous work showed that the A55R gene encoded a protein of predicted size (Beard et al., 2006) and the level of expression of A55 is not affected by the presence of MG132, emphasizing the differences between these structurally related proteins.

This investigation has shown that the role of the F3 protein in VACV infection is notably different from that of C2 or A55, the other two BTB/kelch proteins encoded by the virus. F3 has no detectable effect on the cytoskeletal organization of the virus-infected cell but does affect the innate immune response to intradermal infection of mice. The mechanisms behind this phenomenon are the subject of future investigations.

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