In Vivo Comparison of Chlorine-Based Antiseptics versus Alcohol Antiseptic for Surgical Hand Antisepsis

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1. Introduction

During surgery, multiresistant bacterial flora of the hands of a surgical team is a potential source of pathogens that can be considered as a cause of surgical site infection (SSI). Skin antiseptics have been proven to rapidly and significantly reduce bacteria counts on the surgeons’ hands [1]. Among the most commonly used antiseptics for these purposes are alcohol-based ones, which are very effective in preventing infections in healthcare centres [2, 3]. However, being recognized as an active agent to prevent SSI in developed countries, alcohol-based antiseptics have the following serious limitations: (i) drying effect on the hands [4]; (ii) chronic hand eczema caused by alcohol-based antiseptics [5, 6]; (iii) liver cirrhosis, fetal alcohol syndrome and cancer caused by ingestion of alcohol [7]. Further, alcohol-based antiseptics are expensive [8] and must be stored away from flame due to high flammability [9].

Hypochlorous acid (HOCl) is an acceptable alternative to alcohol in terms of being explored as antiseptic and is effective against a wide range of microorganisms [10–12]. HOCl is produced by white blood cells, making it an endogenous substance that can be tolerated in low concentrations by humans [13]. Not only HOCl but also its salts can act as antiseptic; it was reported in [14] that 0.05% of sodium hypochlorite (NaOCl) can also be used as an antiseptic.

As demonstrated by Petrosyan [15], a solution of 0.2–0.3% NaOCl in combination with 3% hydrogen peroxide (vol. 9:1) buffered by monosodium phosphate (NaH2PO4) is an effective antiseptic for the treatment of the hands of surgeons and medical staff. The specified composition is very easy to be prepared for direct use by mixing...
the finished components (sodium hypochlorite, water, hydrogen peroxide, and monosodium phosphate). Further, it is possible to prepare a solution by electrolysis of available sodium chloride [10]. However, we are not aware of any published investigations evaluating this chlorine-based antiseptic in comparison with alcohol-based ones, following reputable test standards. In the present work, we compared the effectiveness of the indicated antiseptic with ethanol (70%) as well as with sodium hypochlorite solution. The study was designed as a noninferiority investigation. The ASTM E1115, Standard Test Method for Evaluation of Surgical Hand Scrub Formulation [16], and the in vivo method specified by the US Food and Drug Administration for evaluation of surgical hand scrubs were used for comparison of the specified composition and substances as antiseptics [17]. In vivo methods including hands testing for levels of skin microflora before and after antisepsis are comparatively easy to perform; however, it is subject to many uncontrollable factors [18, 19]. Nevertheless, such methods are widely used to test the capacity of a formulation to reduce the level of transient microflora on the hands without regards to the resident flora.

2. Materials and Methods

2.1. Test Products. One combination (NaOCl plus H2O plus NaH2PO4) and two substances (NaOCl and C2H5OH) were compared in terms of antimicrobial activity.

The composition was prepared from commercial sodium hypochlorite (CAS No. 7681-52-9, Hebei Qige Biotechnology Co., Ltd.), monosodium phosphate (≥ 99.0%, Sigma-Aldrich), and double-distilled water. The 0.25 wt% of NaOCl solution was prepared and 4 g of NaH2PO4 was added to 1 L of the solution. The resulting solution (further designated as Antiseptic I) was used for evaluation. The fresh solution was prepared every day to ensure the stability of the composition.

The 0.25 wt% of NaOCl solution and the 70% ethanol solution (Sigma-Aldrich) are further designated as Antiseptics II and III, accordingly.

2.2. Participants. A total of 89 volunteers were recruited for the study.

Practice nurses were responsible for recruiting participants. Nurses provided an information sheet to each potential participant, and with agreement with terms and conditions specified, participants recorded their written informed consent. All the volunteers had to meet the inclusion criteria: free of any underlying chronic diseases, at least 18 years of age, not pregnant, and had no dermatologic conditions or injuries on the hands. Candidates having taking antibiotics or with a history of sensitivity to alcohol, latex, and detergents were excluded from the number of potential participants.

2.3. Pretreatment Phase. According to the informed consent, all the participants agreed not to expose their unprotected hands or forearms to any chemicals throughout one week before sample collecting. During the 2 hours before any sample collecting, participants refrained from washing their hands. At the end of the 1 week, baseline samples were collected from both hands of the participants, by using the glove juice technique [16], on Monday, Wednesday, and Friday.

Before sample collection, all the participants washed their hands and lower two-thirds of the forearms by using nonmedicated soap. Hands were exposed under running tap water for 30 seconds. For all the participants, average baseline counts were >5 log10 colony-forming units per hand.

2.4. Treatment Procedure. All participants were randomly divided into three groups, and each group was assigned to apply one of the test antiseptics (I, II, or III).

Before application of a test product, participants clipped fingernails to ~1 mm free edge if necessary, removed all jewelry from hands and arms, and cleaned under fingernails with a nail cleaner.

During the 5 days, antiseptics were applied via the following regimen: once on days 1 and 5 and three times on days 2, 3, and 4. Samples were collected 4 times, on days 1 and 5: immediately after the antiseptic had air-dried and between 6 and 6.5 hours after drying.

2.5. Counting the Number of Bacteria. Powder-free, sterile latex gloves were placed on both of the participant’s hands. Within 1 minute of donning the gloves, 75 ml of sterile stripping fluid was instilled into the glove. The wrist was secured, and an attendant massaged the hand through the glove in a standardized manner for 1 minute ± 5 seconds. A 5 ml aliquot of the glove juice was removed and diluted in 5 ml of Butterfield’s phosphate buffer solution with product neutralizers (109 dilutions). The 109 dilution was then serially diluted in Butterfield’s phosphate buffer solution with product neutralizers.

Duplicate spread plates were prepared from each of these dilutions on tryptic soy agar with 0.07% lecithin and 0.5% Tween 80 and incubated at 30°C ± 2°C for 48–72 h. Bacteria were counted by using the standard plate-counting procedure. Raw data (colony-forming unit (CFU)/mL) were converted to log10 CFU/hand.

Butterfield’s phosphate-buffered solution and plating medium were evaluated following ASTM E-1054 [20] to ensure that the active components of the test products were effectively neutralized.

2.6. Statistics. Descriptive statistics were calculated using Minitab Statistical software. Comparison of the effectiveness of antiseptics was carried out following the ±20 rule [21]; antiseptics II and III were the comparators. A single trial for comparison of the antiseptic I versus antiseptics II and III was performed.
3. Results and Discussion

3.1. Participants Involved in the Study. 89 volunteers met inclusion criteria, 6 of whom declined to participate, and 1 was allergic to alcohol; thus, 82 participants were randomly divided into 3 groups for testing procedures. Antiseptics I, II, and III were applied to 28, 27, and 27 subjects, accordingly. Because of protocol violation as well as losing to follow-up by 8 participants during the 5-days testing period, only 74 participants were included in the analysis for the primary outcome (Figure 1).

The number of participants excluded from the primary outcome analysis: 1, 4, 5–1; 2, 3–3.

3.2. Bacteria Count. The values of mean, standard deviation (SD), and standard error of the mean (SEM), calculated on the \( \log_{10} \) microbial recoveries, are presented in Table 2. The

![Flowchart of study participants]
log_{10} values for samples collected in each of 3 days on baseline week were averaged before including in the Table. Analysis of variance (ANOVA) used for comparing 3 baseline means; the \( P \) value was calculated to be 0.375, indicating that baselines did not significantly differ from each other.

3.3. Noninferiority and Superiority Trials. The log_{10} reduction from the baseline at each point specified in Table 1 compared between the Antiseptic I and Antiseptic II, as well as between the antiseptic I and antiseptic III, by using the Student’s \( t \) test.

To determine whether the Antiseptic I is no less effective than antiseptics II and III, a confidence interval (CI) approach was used. A two-sided 95% confidence interval was applied for the noninferiority test as this approach preserves consistency between significance testing and subsequent estimation [22].

Figure 2 demonstrates a graphical representation for the noninferiority test.

It is indicated that the CIs for all comparisons (Antiseptic I minus Antiseptic II and Antiseptic I minus Antiseptic III for all 8 cases) lie above zero. \( P \) values were calculated for all cases as seen in Figure 2. As shown, Antiseptic I is noninferior to both Antiseptics II and III and in fact demonstrated superiority to both reference antiseptics in day 5 (\( P < 0.05 \) for each comparative trial in this day).

3.4. Adverse Reactions. No adverse reactions attributable to the antiseptic application were detected.

3.5. Possible Reason for Antiseptic \( \text{NaOCl + H}_{2}\text{O}_{2} + \text{NaH}_{2}\text{PO}_{4} \) Effectiveness. The combined presence of sodium hypochlorite and hydrogen peroxide in the solution leads to the formation of free radicals and/or active oxygen [23]. The possibility of the formation of singlet oxygen by the following reactions was shown in [24]:

\[
\text{OCI}^- + \text{H}^+ \longrightarrow \text{HOCl} \quad (1)
\]

\[
\text{HOCl} + \text{H}_2\text{O}_2 \longrightarrow \text{H}^+ + \text{Cl}^- + \text{H}_2\text{O} + \text{O}_2 (\Delta \uparrow) \quad (2)
\]

Singlet oxygen is a strong bactericidal agent and acts as an effective antiseptic against multiple pathogens, including microbes, virus, and also cancer cells. Besides, singlet oxygen has a high-penetrating ability into the deeper layers of the
H2O2 may be a promising alternative to traditional alcohol presentsevidencethatanantisepticconsistingofNaOCLand
the statistical significance of 5% level. Together, this study
strated noninferiority and on day 5 superiority toward 70%
hand antiseptics.

Data Availability
The data supporting the results can be made available from
the corresponding author upon request.

Conflicts of Interest
The authors declare no conflicts of interest.

Authors’ Contributions
Zh. M. and G. K. were involved in the conceptualization; G. K. was responsible for the methodology and wrote, reviewed, and edited the article; R. N. and G. K validated the study; A. M. was involved in the formal analysis and data curation; G. K and R. N investigated the study; S. O. was involved in the statistical calculations and visualized the study; R. N wrote and prepared the original draft; Zh. M was involved in the supervision, project administration, and funding acquisition. All the authors have read and agreed to the published version of the manuscript.

Acknowledgments
This research was funded by the Science Committee of the Ministry of Education and Science of the Republic of Kazakhstan (Grant no. AP05132799).

References

[1] A. Maier, J. L. Ovesen, C. L. Allen et al., “Safety assessment for ethanol-based topical antiseptic use by health care workers: evaluation of developmental toxicity potential,” Regulatory Toxicology and Pharmacology, vol. 73, no. 1, pp. 248–264, 2015.
[2] A. Kanwar, J. A. Kumar, Y. K. Ng-Wong et al., “Evaluation of an alcohol-based antiseptic for nasal decolonization of methicillin-resistant Staphylococcus aureus in colonized pa-
tients,” Infection Control & Hospital Epidemiology, vol. 40, no. 12, pp. 1436-1437, 2019.
[3] A. Malani, K. Trimble, V. Parekh, C. Chenoweth, S. Kaufman, and S. Saint, “Review of clinical trials of skin antiseptic agents used to reduce blood culture contamination,” Infection Control & Hospital Epidemiology, vol. 28, no. 7, pp. 892–895, 2007.
[4] S. E. Czerwinski, J. Cozean, and C. Cozean, “Novel water-based antiseptic lotion demonstrates rapid, broad-spectrum kill compared with alcohol antiseptic,” Journal of Infection and Public Health, vol. 7, no. 3, pp. 199–204, 2014.
[5] J. P. Cimiotti, E. S. Marmur, M. Nesin, P. Hamlin-Cook, and E. L. Larson, “Adverse reactions associated with an alcohol-based hand antiseptic among nurses in a neonatal intensive care unit,” American Journal of Infection Control, vol. 31, no. 1, pp. 43–48, 2003.
[6] G. Kampf and H. Löfler, “Dermatological aspects of a suc-

scessful introduction and continuation of alcohol-based hand rubs for hygienic hand disinfection,” Journal of Hospital Infection, vol. 55, no. 1, p. 7, 2003.
[7] V. Bessonneau, M. Clément, and O. Thomas, “Can intensive
use of alcohol-based hand rubs lead to passive alcohol-
ization?” International Journal of Environmental Research and Public Health, vol. 7, no. 8, pp. 3038–3050, 2010.
[8] J. Boyce, “Antiseptic technology: access, affordability, and acceptance,” Emerging Infectious Diseases, vol. 7, no. 2, pp. 231–233, 2001.
[9] M. K. Wolfe, K. Gallandat, K. Daniels, A. M. Desmarais, P. Scheinman, and D. Lantagne, “Handwashing and E. bula

virus disease outbreaks: a randomized comparison of soap, hand sanitizer, and 0.05% chlorine solutions on the inacti-
vation and removal of model organisms Phi6 and E. coli from hands and persistence in rinse water,” PLoS one, vol. 12, no. 2, Article ID e0172734, 2017.
[10] K. A. Mourad and S. Hobro, “Developing chloride-based antiseptic by electrolysis,” Science of The Total Environment, vol. 709, Article ID 136108, 2020.
[11] A. G. Anagnostopoulos, A. Rong, D. Miller et al., “0.01% hypochlorous acid as an alternative skin antiseptic: an in vitro comparison,” Dermatologic Surgery, vol. 44, no. 12, pp. 1489–1493, 2018.
[12] S. Sakarya, N. Gunay, M. Karakulak, B. Oztürk, and B. Ertugrul, “Hypochlorous acid: an ideal wound care agent with powerful microbicidal, antibiofilm, and wound healing potency,” Wounds: A Compendium of Clinical Research and Practice, vol. 26, no. 12, pp. 342–350, 2014.
[13] L. Wang, M. Bassiri, R. Najafi et al., “Hypochlorous acid as a potential wound care agent: part I. stabilized hypochlorous acid: a component of the inorganic armamentarium of innate immunity,” Journal of Burns and Wounds, vol. 11, no. 6, p. e5, 2007.
[14] M. Ciccia, R. Chakrokh, D. Molinazzi, A. Zanni, P. Farruggia, and F. Sandri, “Skin antisepsis with 0.05% sodium hypo-

chlorite before central venous catheter insertion in neonates: a 2-year single-center experience,” Journal of Burn Care & Research, vol. 31, no. 1, pp. 1, 2010.
[15] A. Malani, K. Trimble, V. Parekh, C. Chenoweth, S. Kaufman, and S. Saint, “Review of clinical trials of skin antiseptic agents used to reduce blood culture contamination,” Infection Control & Hospital Epidemiology, vol. 28, no. 7, pp. 892–895, 2007.
hand-rub and hand-wash agents,” *Journal of Hospital Infection*, vol. 73, no. 3, pp. 191–199, 2009.

[20] ASTM International, *E1054–08: Standard Test Methods for Evaluation of Inactivators of Antimicrobial Agents*, ASTM International, West Conshohocken, PA, USA, 2008.

[21] J. E. De Muth, *Basic Statistics and Pharmaceutical Statistical Applications*, CRC Press, Boca Raton, FL, USA, 2014.

[22] L. K. M. Olson, D. J. Morse, C. Duley, and B. K. Savell, “Prospective, randomized in vivo comparison of a dual-active waterless antiseptic versus two alcohol-only waterless antiseptics for surgical hand antisepsis,” *American Journal of Infection Control*, vol. 40, no. 2, pp. 155–159, 2012.

[23] M. Kashimatanaka, Y. Tsujimoto, K. Kawamoto, N. Senda, K. Ito, and M. Yamazaki, “Generation of free radicals and/or active oxygen by light or laser irradiation of hydrogen peroxide or sodium hypochlorite,” *Journal of Endodontics*, vol. 29, no. 2, pp. 141–143, 2003.

[24] R. R. Cui, L. Z. Deng, W. B. Shi, H. P. Yang, G. H. Sha, and C. H. Zhang, “Liquid-liquid reaction of hydrogen peroxide and sodium hypochlorite for the production of singlet oxygen in a centrifugal flow singlet oxygen generator,” *Quantum Electronics*, vol. 41, no. 2, p. 139, 2011.

[25] T. A. Dahl, W. R. Midden, and P. E. Hartman, “Comparison of killing of gram-negative and gram-positive bacteria by pure singlet oxygen,” *Journal of Bacteriology*, vol. 171, no. 4, pp. 2188–2194, 1989.