Quantitative Evaluation of Viability- and Apoptosis-Related Genes in Ascaris suum Eggs under Different Culture-Temperature Conditions

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Abstract: Ascaris suum eggs are inactivated by composting conditions; however, it is difficult to find functional changes in heat-treated A. suum eggs. Here, unembryonated A. suum eggs were incubated at 20˚C, 50˚C, and 70˚C in vitro, and the gene expression levels related to viability, such as eukaryotic translation initiation factor 4E (IF4E), phosphofructokinase 1 (PFK1), and thioredoxin 1 (TRX1), and to apoptosis, such as apoptosis-inducing factor 1 (AIF1) and cell death protein 6 (CDP6), were evaluated by real-time quantitative RT-PCR. No prominent morphological alterations were noted in the eggs at 20˚C until day 10. In contrast, the eggs developed rapidly, and embryonated eggs and hatched larvae began to die, starting on day 2 at 50˚C and day 1 at 70˚C. At 20˚C, IF4E, PFK1, and TRX1 mRNA expression was significantly increased from days 2-4; however, AIF1 and CDP6 mRNA expression was not changed significantly. IF4E, PFK1, and TRX1 mRNA expression was markedly decreased from day 2 at 50˚C and 70˚C, whereas AIF1 and CDP6 mRNA expression was significantly increased. The expressions of HSP70 and HSP90 were detected for 9-10 days at 20˚C, for 3-5 days at 50˚C, and for 2 days at 70˚C. Taken together, incremental heat increases were associated with the rapid development of A. suum eggs, decreased expression of genes related to viability, and earlier expression of apoptosis-related genes, and finally these changes of viability- and apoptosis-related genes of A. suum eggs were associated with survival of the eggs under temperature stress.

Key words: Ascaris suum, egg, temperature, viability, apoptosis, real-time quantitative RT-PCR
physiologic saline (0.9% NaCl), eggs were transferred to a Petri dish and incubated with physiologic saline at 20°C, 50°C, and 70°C for 40 days to examine the survival period of the eggs. About 200 eggs were examined in each case, and the development of the eggs was evaluated under a microscope on the basis of morphological changes in the germinal cells and the presence of viable larvae in the eggs or hatched larvae. We also evaluated the expression levels of genes or proteins related to viability, apoptosis, and heat stress for 10 days.

Real-time qRT-PCR analyses of IF4E, PFK1, TRX1, AIF1, and CD6P expressions were performed using the ABI StepOne Real-Time PCR System (Applied Biosystems, Foster City, California, USA). All assays used a similar amplification efficiency, with a ΔΔCt experimental design for relative quantification. The reactions were performed in triplicate in a 20-μl reaction volume containing 10 μl of the expression probe (Power SYBR Green PCR Master mix; Applied Biosystems), 1 μl of forward (F)-primer, 1 μl of reverse (R)-primer, 2 μl of cDNA, and 6 μl of distilled water. The resulting calibration graph (Ct vs. log unit of the standard template) correlates Ct values with the amount of template in the reaction. This calibration was performed separately for every amplicon. To standardize the amount of cDNA used, the housekeeping gene encoding elongation factor 1-α (EF1α) served as an endogenous control. The amplification program consisted of 10 min at 95°C followed by 40 cycles of 15 sec at 95°C and 1 min at 60°C. The oligonucleotides used were: EF1α-F, 5′-CCCTAGACTGCCTCCTCAGAAGC-3′; EF1α-R, 5′-GAGCCCGACCTACACCGGTTGA-3′; IF4E-F, 5′-GTATTTGACGGCGGAGTCAAA-3′; IF4E-R, 5′-GACCACCCCTTGACGTTT-3′; PFK1-F, 5′-CATCTCCCTTGTACTGACC-3′; PFK1-R, 5′-GGGTCTCCTGGAATGACTT-3′; CDP6-F, 5′-ACATCCGTGGATTG-3′; CDP6-R, 5′-CCACCCACTCAACGTTGA-3′; AIF1-F, 5′-ACCCGCTGACTGTCGGAG-3′; AIF1-R, 5′-CGGCCATACCACAAAATAGTCT-3′; TRX1-F, 5′-GACCTCGGAGGTAAAGGTCTT-3′; TRX1-R, 5′-CAGCATCAGCGTGATTGTG-3′; HSP70 and HSP90 expressions were analyzed by western blotting. Eggs were lysed in SDS-PAGE loading buffer (2% SDS, 62.5 mM Tris-HCl, 10% glycerol, 50 mM dithiothreitol, and 0.01% bromophenol blue; pH 6.8) and heated at 98°C for 5 min. Proteins (50 μg) were resolved by SDS-PAGE using 15% acrylamide and then transferred to PVDF membranes (BioRad, Hercules, California, USA) by a semidry transfer method. The membranes were then blocked overnight at 4°C with 1 × PBS/0.1% Tween 20 (TPBS) plus 5% (w/v) non-fat dry milk and incubated for 3 hr at room temperature with monoclonal anti-HSP70 and anti-HSP90 antibodies (1: 5,000 dilution; Sigma, St. Louis, Missouri, USA). After washing in TPBS, the membranes were incubated with HRP-conjugated rabbit anti-mouse (Sigma) secondary antibodies for 1 hr at room temperature and then washed 4 times in TPBS for 10 min each. The blots were developed using an enhanced chemiluminescence detection kit (Amersham, Buckinghamshire, UK) and visualized by autoradiography.

We measured the survival periods of A. suum eggs incubated at 20°C, 50°C, and 70°C for 40 days. As shown in Fig. 1, at 20°C, > 99% of the eggs survived until day 10. On day 20, the percentage of viable A. suum eggs was 89 ± 6%. On day 40, 72 ± 9% of the eggs were viable. However, at 50°C, about 67% of the eggs were viable on day 1, and 97% of the eggs or larvae were inactivated on day 2. Almost all eggs were devitalized by day 4. At 70°C, eggs started to die on day 1, and almost all eggs were dead on day 2. We also assessed the morphological characteristics of A. suum eggs during development at different temperatures for 10 days (Fig. 2). At 20°C, no morphological alterations compared to day 0 were noted until day 5. On day 10, 1% of the eggs were in the cleavage stage. During incubation at 50°C, about 35% of the eggs were embryonated on day 1. On day 2, the eggs were further developed; thus, 85 ± 4% of the eggs were observed to be in the larval stage. On day 4, the

![Fig. 1. Survival days of Ascaris suum eggs at 20°C, 50°C, and 70°C of incubation for 40 days. At each time, about 200 eggs were evaluated under a microscope on the basis of morphological changes in triplicate. Data are presented as the mean ± SD of 200 A. suum eggs and are representative of 2 separate experiments.](image-url)
eggs were in the larval stage or hatching naturally, and most of hatched larvae or embryonated eggs were dead. Among the eggs incubated at 70˚C, 62% were in the larval stage on day 1, and most of them were not viable. On day 2, all observed embryos were in the larval stage or hatching naturally, and almost all of the hatched larvae or larval-stage eggs were dead.

To quantify the functional changes in *A. suum* eggs under different heat stress conditions, mRNA expressions of viability- and apoptosis-related genes were evaluated for 10 days (Fig. 3). At 20˚C, IF4E mRNA expression was significantly increased on day 4 compared with day 0, and this level increased slowly until day 8, after which the level remained constant. PFK1 and TRX1 mRNA expressions were significantly increased on day 2 (*P* < 0.05), after which similar levels were maintained until day 10. However, AIF1 and CDP6 mRNA expression was not significantly changed during the experimental period. At 50˚C and 70˚C, IF4E, PFK1, and TRX1 mRNA expressions were markedly decreased on day 2 (*P* < 0.01), after which very low levels were maintained until day 10. However, AIF1 and CDP6 mRNA expressions were markedly decreased on days 2-6 at 50˚C or 70˚C (*P* < 0.01), and then increased from day 8 at 50˚C and day 4 at 70˚C.

HSPs are induced by a variety of stressors, including heat, free radicals, and heavy metals. We assessed the expression levels of HSP70 and HSP90 to evaluate the heat stress response of *A. suum* eggs (Fig. 4). At 20˚C, HSP70 derived from *A. suum* eggs was detected for 10 days, with high levels during days 1-3, after which a decrease was observed. Similar to HSP70 expression, HSP90 levels were high until day 3 and then tapered slowly. HSP70 and HSP90 were detected during days 1-5 at 50˚C and days 1-3 at 70˚C. At 50˚C, the expression levels of HSP70 and HSP90 were the highest on day 1, and then decreased. At 70˚C, HSP70 and HSP90 protein expression was detected on days 2 and 1, respectively, and their expression levels were much lower than those observed in eggs incubated at 20 or 50˚C.

Pigs are the main host for *A. suum*, and it is estimated that approximately 10-50% of pigs are infected by *A. suum* worldwide [13-15]. Following the ingestion of *A. suum* egg-contaminated food and water by humans or other mammals, larvae reach the liver and lung alveoli via the bloodstream, resulting in eosinophilic pneumonia, liver lesions, myelitis, and visceral larva migrans, characterized by neurological symptoms, including encephalomyeloradiculoneuropathy [5,6]. Thus, the inacti-
vation of helminth eggs by fermentation or composting is critical for minimizing risks to the environment and public health.

Our results demonstrate that the devitalization of *A. suum* eggs occurred in a temperature-dependent manner. Almost all *A. suum* eggs were alive for 20 days at 20˚C, whereas inactivation started prominently from days 2 and 1 at 50˚C and 70˚C, respectively. The complete devitalization of unembryonated *A. suum* eggs during the composting of organic wastes has been reported at temperatures ranging from 49˚C to 64˚C [8]. Ham-er [3] also suggested that temperatures between 60˚C and 72˚C were sufficient for pathogen inactivation. Compared with the report of Cruz et al. [16], who incubated unembryonated *A. suum* eggs at 28˚C, our timeline of egg development at 20˚C was somewhat delayed. These data suggest that temperature is critically important in the devitalization of *Ascaris* eggs, although pH, ammonia concentration, and anaerobic conditions are also involved in the composting process.

Here, we evaluated the expression levels of genes related to viability and apoptosis by real-time qRT-PCR. The expression periods of these viability-related genes were decreased in a temperature-dependent manner. At 20˚C, IF4E, PFK1, and TRX1 mRNA levels were significantly increased, which may present the development of *A. suum* eggs. However, *A. suum* eggs started to die at 50˚C and 70˚C, thus the mRNA levels of IF4E, PFK1, and TRX1 were sharply decreased. In addition, the expression of these apoptosis-related genes appeared early with incremental temperature increases. In particular, AIF1 and CDP6 mRNA levels were significantly high from day 8.
and 4 at 50°C and 70°C, respectively. These results may be reflected as the signals of heat-treated eggs or the remains of dead eggs.

HSPs are involved in the transport, folding, and assembly of proteins and serve to maintain cell homeostasis by functioning as molecular chaperones [17]. In this study, we found that the proteins HSP70 and HSP90 were expressed at higher levels for a longer period in eggs incubated at 20°C than in those incubated at 50°C or 70°C. This expression pattern is similar to that for viability-related gene expression. Interestingly, HSP70 protein was expressed for longer periods than HSP90 at all temperatures analyzed. These differences may be due to the different regulatory systems between HSP70 and HSP90. Based on the quantitative evaluation of genes or proteins related to viability, apoptosis, and heat stress, the expression periods and levels of these genes or proteins were concomitant with the survival periods of the eggs according to incubation temperature.

In this study, we quantified the functional changes in genes or proteins related to viability, apoptosis, and heat stress in A. suum eggs at composting temperatures. The expression periods of IF4E, PFK1, and TRX1 were shortened with incremental temperature increases, whereas AIF1 and CDP6 were expressed earlier at increased temperatures. The protein expression levels and periods of HSP70 and HSP90 were decreased with incremental temperature increases. The functional changes in these genes or proteins were correlated with survival of the eggs under heat stress conditions. Our results confirmed those of previous studies showing that the inactivation of A. suum eggs is strongly dependent on temperature.

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REFERENCES

1. Leles D, Gardner SL, Reinhard K, Iñiguez A, Araujo A. Are Ascaris lumbricoides and Ascaris suum a single species? Parasit Vectors 2012; 5: 42.
2. Collick AS, Inglis S, Wright P, Steenhuis TS, Bowman DD. Inactivation of Ascaris suum in a biodrying compost system. J Environ Qual 2007; 36: 1528-1533.
3. Hamer G. Solid waste treatment and disposal: effects on public health and environmental safety. Biotechnol Adv 2003; 22: 71-79.
4. Pinelli E, Willers SM, Hoek D, Smit HA, Kortbeek LM, Hoelstra M, de Jongste J, van Knapen E, Postma D, Kerkmans M, Aalberse R, van der Giessen JW, Brunekreef B. Prevalence of antibodies against Ascaris suum and its association with allergic manifestations in 4-year-old children in the Netherlands: The PIAMA birth cohort study. Eur J Clin Microbiol Infect Dis 2009; 28: 1327-1334.
5. Inatomi Y, Murakami T, Tokuinaga M, Ishiwata K, Nawa Y, Uchino M. Encephalopathy caused by visceral larva migrans due to Ascaris suum. J Neurol Sci 1999; 164: 195-199.
6. Sakakihara A, Baba K, Niwa S, Yagi T, Wakayama H, Yoshida K, Kobayashi T, Yokoi T, Hara K, Itoh M, Kimura E. Visceral larva migrans due to Ascaris suum which presented with eosinophil pneumonia and multiple intra-hepatic lesions with severe eosinophil infiltration-outbreak in a Japanese area other than Kyushu. Intern Med 2002; 41: 574-579.
7. Vinnerås B. Comparison of composting, storage and urea treatment for sanitising of faecal matter and manure. Bioresour Technol 2007; 98: 3317-3321.
8. Szabová E, Juris P, Papajová I. Sanitation composting process in different seasons. Ascaris suum as model. Waste Manag 2010; 30: 426-432.
9. Persson BM, Barrios JA, Jiménez BE, Nelson KL. The effects of temperature, pH, and ammonia concentration on the inactivation of Ascaris eggs in sewage sludge. Water Res 2007; 41: 2893-2902.
10. Goodfellow IG, Roberts LO. Eukaryotic initiation factor 4E. Int J Biochem Cell Biol 2008; 40: 2675-2680.
11. Mort I, Cheung EC, Vousden KH. Control of glycolysis through regulation of PFK1: Old friends and recent additions. Cold Spring Harb Symp Quant Biol 2011; 76: 211-216.
12. Watanabe R, Nakamura H, Masutani H, Yodoi J. Anti-oxidative, anti-cancer and anti-inflammatory actions by thioredoxin 1 and thioredoxin-binding protein-2. Pharmacol Ther 2010; 127: 261-270.
13. Nissen S, Poulsen IH, Nejsund P, Olsen A, Roepstorff A, Rubaire-Akiiki C, Thamsborg SM. Prevalence of gastrointestinal nematodes in growing pigs in Kabale District in Uganda. Trop Anim Health Prod 2011; 43: 567-572.
14. Ismail HA, Jeon HK, Yu YM, Do C, Lee YH. Intestinal parasite infections in pigs and beef cattle in rural areas of Chungcheongnam-do, Korea. Korean J Parasitol 2010; 48: 347-349.
15. Lai M, Zhou RQ, Huang HC, Hu SJ. Prevalence and risk factors associated with intestinal parasites in pigs in Chongqing, China. Res Vet Sci 2011; 91: e121-e124.
16. Cruz LM, Allanson M, Kwa B, Azizan A, Izurieta R. Morphological changes of Ascaris spp. eggs during their development outside the host. J Parasitol 2012; 98: 63-68.
17. Rupik W, Jasik K, Bembrinek J, Widlak W. The expression patterns of heat shock genes and proteins and their role during vertebrate’s development. Comp Biochem Physiol A Mol Integr Physiol 2011; 159: 349-366.
