Genetics of systemic sclerosis: recent advances

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Purpose of review
Large-scale and follow-up genetic association studies in systemic sclerosis (SSc) have implicated over 40 regions in disease risk, 15 of which with robust associations. Nevertheless, the causal variants and the functional mechanisms underlying the genetic associations remain elusive, and the reasons for the higher disease burden in African Americans unknown. Incorporating tools from diverse fields is beginning to unveil the role of genetic diversity and regulatory variation in SSc susceptibility. This review will summarize recent advances in SSc genetics, including autoimmune disease overlap, evidence of natural selection, and current progress towards the dissection of the functional role of associated risk variants.

Recent findings
In the past year, multiple large-scale studies reported novel strong and suggestive SSc associations. These results, coupled with the regions shared with other autoimmune diseases, emphasize the role of dysregulation of immune pathways as a key causative factor in SSc pathogenesis. Strong evidence implicates natural selection as a mechanism contributing to the maintenance of some of these SSc alleles in the population. Studies integrating genomic, transcriptomic, and epigenomic datasets in specific cell types to identify causal autoimmune disease variants are emerging.

Summary
The identification and comprehensive understanding of the factors and mechanisms contributing to SSc will contribute to improved diagnosis and disease management.

Keywords
functional variation, genetic association, population genetics

INTRODUCTION
Systemic sclerosis (SSc, scleroderma) is a multisystem disease characterized by cutaneous and visceral fibrosis, immune dysregulation, and vasculopathy. SSc is a fibrosing disease preceded and accompanied by autoimmunity and vasculopathy [1]. SSc is very heterogeneous, with patients being commonly classified into two main subsets on the basis of the pattern of skin involvement: limited cutaneous SSc (lcSSc), which is typically associated with ant centromere antibodies (ACA), pulmonary hypertension, but generally an overall better prognosis, and diffuse cutaneous SSc (dcSSc), typically associated with antitopoisomerase I (Scl-70) antibodies (ATA), anti-RNA polymerase antibodies, propensity for severe pulmonary fibrosis, earlier age of onset, and a poorer prognosis [2,3].

There is great variation in reported incidence and prevalence estimates, with the latter varying between 31 and 659 cases per million [4]. Like other autoimmune diseases, SSc is associated with a strong sex bias of between four and nine affected women for one man [5–7]. African Americans are more likely to develop SSc than Caucasians [8,9], and experience greater morbidity, reduced survival, earlier age of onset, and worse pulmonary disease [8–13]. Nearly 70% of SSc patients show pulmonary involvement, which is the main cause of mortality in SSc [14]. The cause of SSc and the reasons for the ethnic disparities remain elusive.

Despite the progress in the identification of genetic risk factors for SSc, little progress has been made in the identification of the specific disease-predisposing functional variations. Since most functional variants lie in noncoding regions of the
Despite its limited genetic burden, multiple genetic risk loci for SSc continue to be identified, underscoring the role of immune system activation as an early and key event in SSc pathogenesis.

The majority of the SSc-associated loci are shared with at least one other autoimmune disease, mostly with rheumatoid arthritis, systemic lupus erythematosus, multiple sclerosis, inflammatory bowel disease, and primary biliary cirrhosis.

Several SSc risk regions show compelling evidence for natural selection, supporting the hypothesis that this population phenomenon has contributed to genetic variation that influences disease risk and ethnic and geographic health disparities.

Despite the disproportionate burden on African Americans, the dramatic under-representation of African Americans in research continues to exacerbate the health disparities gap.

The integration of genetic association with population genetic and functional annotation tools will help elucidate the role of recent human adaptation in ethnic disparities and the functional role of associated risk variants in relevant cell types, supporting future gene editing studies aimed at experimentally validating the disease mechanisms.

KEY POINTS
- Despite its limited genetic burden, multiple genetic risk loci for SSc continue to be identified, underscoring the role of immune system activation as an early and key event in SSc pathogenesis.
- The majority of the SSc-associated loci are shared with at least one other autoimmune disease, mostly with rheumatoid arthritis, systemic lupus erythematosus, multiple sclerosis, inflammatory bowel disease, and primary biliary cirrhosis.
- Several SSc risk regions show compelling evidence for natural selection, supporting the hypothesis that this population phenomenon has contributed to genetic variation that influences disease risk and ethnic and geographic health disparities.
- Despite the disproportionate burden on African Americans, the dramatic under-representation of African Americans in research continues to exacerbate the health disparities gap.
- The integration of genetic association with population genetic and functional annotation tools will help elucidate the role of recent human adaptation in ethnic disparities and the functional role of associated risk variants in relevant cell types, supporting future gene editing studies aimed at experimentally validating the disease mechanisms.

Genome and the majority of complex disease-associated variants are concentrated in regulatory DNA [15], functional studies of associated loci is an area of current focus. In addition, despite the disproportionate burden on African Americans, virtually all studies have been conducted in populations of European ancestry. The dramatic under-representation of African Americans in research continues to exacerbate the health disparities gap and is a major priority. A collaborative large-scale effort to identify common and low-frequency SSc variants in African Americans is underway, which, coupled with the increasing understanding of the regulatory genome, will undoubtedly contribute to the elucidation of the functional mechanisms that underlie SSc cause.

GENETIC RISK

Although multiple lines of indirect evidence support a modest genetic risk in SSc, including a sibling risk ratio (λs) of 15–19 [16–19], a low disease concordance in monozygotic twins (4.2%) [20], and genetic heritability (h^2 = 0.008) [20], the high concordance of antinuclear antibodies (ANAs) in monozygotic twins (90 and 71% for titers ≥1:40 and ≥1:80, respectively) [20] and the increased relative risk of autoimmune disease in first-degree relatives [21] support a higher genetic contribution to the development of autoimmunity in SSc. The modest genetic burden suggests a substantial role for epigenetic or environmental factors in SSc susceptibility.

Recently, more direct evidence for the role of genetic variation in the pathogenesis SSc has emerged. Until 2010, only a handful of candidate gene polymorphisms had been convincingly implicated is SSc risk. With the advent and application of high-throughput technologies and increased collaborations, four genome-wide association studies (GWAS) [22–25], two large-scale studies [26,27] using a high-density genotyping array of immune loci (Immunochip), and multiple GWAS and Immunochip follow-up studies [28–32] have been published since 2010. Currently, 45 genetic regions are implicated in SSc risk, 15 of which have compelling genetic associations at the genome-wide significance level (P < 5 × 10^-8) and/or multiple suggestive (5 × 10^-8 < P < 5 × 10^-6) independent replications. These regions are shown in Table 1, which, in order to avoid likely spurious associations, includes all genetic associations with minimum P value no larger than 5 × 10^-4 tested in a total sample size of at least 1000 individuals. In spite of the complex genetic architecture, these discoveries demonstrate that a broad array of pathways underlies the genetic heterogeneity of SSc, including innate immunity pathways (Toll-like receptor, interferon, DNA clearance during apoptosis), adaptive immunity pathways [interleukin (IL)-12], and fibrosis pathways.

The human leukocyte antigen (HLA) class II region remains the most significant genetic association with SSc, similar to all other autoimmune diseases, and has been extensively reviewed [70]. Non-HLA loci have also been reviewed [71,72]. In the past year, two Immunochip [26,27], an Immunochip and GWAS follow-up [31,32], and a candidate gene study [73] reported novel strong and suggestive associations with SSc. The findings from these studies are discussed below, some of which have already been included in recent reviews [74,75]. The majority of these regions overlap with those implicated in other autoimmune diseases, underscoring the role of immune system dysregulation as a primary event in SSc pathogenesis.

Epigenetic studies [76,77] have been recently reviewed elsewhere and are not discussed here.

NEW STUDIES

Beyond the initial wave of GWAS studies, gene identification studies have since aimed to capture as much genomic variation as possible in each region to maximize the odds of identifying the
| Gene region | Band location | Approach | Phenotype | References | AD overlap | Natural selection | Selective pressure |
|-------------|---------------|----------|-----------|------------|------------|------------------|-------------------|
| KIAA0319L  | 1p34.3        | GWA      | lcSSc     | [24]       |            |                  |                   |
| IL12RB2    | 1p31.3        | GFU      |           | [28]       | BeD, PBC   |                  |                   |
| IL23R      | 1p31.3        | CG       |           | [33,34]    | AS, BeD, CD, IB, PS, UC | Y                | Protozoa          |
| VCAM1      | 1p21.2        | IC       |           | [27*]      | MS         |                  |                   |
| PTPN22     | 1p13.2        | CG       |           | [35–37]    | CD, MG, RA, T1D, VT | Y                | Protozoa          |
| CD247*     | 1q24.2        | CG, GWA  |           | [22,23,25,38] | GeD, RA    |                  |                   |
| TNFSF4*    | 1q25.1        | CG, GWA  | ACA       | [24,39–41] | MS, RA, SLE | Y                |                   |
| NMNAT2     | 1q25.3        | GWA      | ACA       | [24]       |            |                  |                   |
| RHOB       | 2p24.1        | GWA      |           | [22]       |            |                  |                   |
| STAT4*     | 2q32.3        | CG, GWA  |           | [22–25,27*,42–45] | BeD, IB, PBC, RA, SLE, SS, CelD + RA |               |                   |
| PPARG      | 3p25          | GFU      |           | [32]       |            |                  |                   |
| PLC12      | 3p24.3        | CG       |           | [73]       | MS, PBC, RA |                  |                   |
| DNASE1L3-PKX* | 3p14.1     | IC       | ACA       | [24,26*,27*] | RA, SLE   |                  |                   |
| IL12A*     | 3q25.33       | IC       | lcSSc     | [26*]      | BeD, CelD, MS, PBC | Y                |                   |
| TLR2       | 4q32          | CG       | ATA       | [46]       |            |                  |                   |
| BANK1-NFkB1| 4q24          | CG, GFU  | dcSSc, ATA| [29,73,47,48] | CD, PBC, MS, SLE, UC |                  |                   |
| TNIP1*     | 5q33.1        | GFU, GWA |           | [22,24,30] | IB, MG, PS, SLE | Y                |                   |
| HLA region* | 6p21.33       | IC, GWA  | SSc, dcSSc and ATA | [24,49–51] | CelD, IB, MS, PS, RA, SLE, SS, UC |               |                   |
| UHRF1BP1   | 6p21.31       | GWA      |           | [24]       | SLE        | Y                | M. tuberculosis   |
| ATG5*      | 6q21          | IC, GWA  |           | [24,26*]   | RA, SLE    |                  |                   |
| TNFAIP3*   | 6q23.3        | CG, GWA  | SSc, dcSSc and ATA | [24,49–51] | CelD, IB, MS, PS, RA, SLE, SS, UC |               |                   |
| COR6       | 6q27          | CG       | ATA       | [52]       | RA         |                  |                   |
| JAZF1      | 7p15.1        | GWA      |           | [24]       | CD, RA, T1D |                  |                   |
| GRB10      | 7p12.1        | GWA      | lcSSc     | [23]       |            |                  |                   |
| IRF5*      | 7q32.1        | CG, GWA  |           | [22–25,27*,53–55] | PBC, RA, SLE, UC | Y                |                   |
| BLK*       | 8p23.1        | CG, GWA  |           | [24,56–58] | KA, RA, SLE | Y                |                   |
| PSD3       | 8p21.3        | GFU      |           | [29]       |            |                  |                   |
| IL2RA      | 10p15.1       | CG       | ACA       | [59]       | AA, CD, IB, MS, RA, T1D |            |                   |
| FAS         | 10q24.1       | CG       |           | [60,61]    |            |                  |                   |
| RF7         | 11p15.5       | CG, GWA  | ACA       | [24,62]    | SLE        |                  |                   |

(Continued)
| Gene region | Band location | Approach  | Phenotype      | References | AD overlap          | Natural selection | Selective pressure |
|-------------|---------------|-----------|----------------|------------|---------------------|-------------------|-------------------|
| EHF         | 11p13         | GWA       | lcSSc          | [24]       | SLE                 |                   |                   |
| TREH-DDX6   | 11q23.3       | IC        |                | [26\*]     | SLE, CelD + RA      |                   |                   |
| KCNA5       | 12p13         | CG        | SScPAH         | [63]       |                     |                   |                   |
| SOX5        | 12p12.1       | GWA       | ACA            | [23]       |                     |                   |                   |
| RPL41-ESYT1 | 12q13.2       | GWA       | dcSSc          | [23]       | T1D                 |                   |                   |
| PLD4        | 14q32.33      | CG        |                | [50]       | RA                  |                   |                   |
| CSK\*       | 15q24.2       | GFU, GWA  |                | [24,29]    |                     |                   |                   |
| IRF8\*      | 16q24.1       | GWA       | lcSSc          | [23,24,73] | IBD, MS, PBC, RA, UC|                   |                   |
| NRP1        | 17p13.2       | CG        |                | [64]       | Y                   |                   |                   |
| IKZF3       | 17q12         | GWA       | ACA            | [24]       | IBD, PBC            |                   |                   |
| TBX21       | 17q21.32      | CG        |                | [64]       | AS, MS              |                   |                   |
| CD226       | 18q22.2       | CG        |                | [65]       | IBD, RA, T1D        |                   |                   |
| IL12RB1\*  | 19p13.1       | IC FU     |                | [31]       | MS                  |                   |                   |
| MIF         | 22q11.23      | CG        |                | [66,67]    |                     |                   |                   |
| IRAK1       | Xq28          | CG        |                | [68,69]    | RA                  |                   |                   |

Disease abbreviations: AA, alopecia areata; AD, autoimmune disease; AS, ankylosing spondylitis; BeD, Behçet’s disease; CelD, celiac disease; CD, Crohn’s disease; ID, inflammatory bowel disease; KA, Kawasaki disease; MG, myasthenia gravis; MS, multiple sclerosis; PBC, primary biliary cirrhosis; PS, psoriasis; RA, rheumatoid arthritis; HLA: human leukocyte antigen; SLE, systemic lupus erythematosus; SS, Sjögren’s syndrome; T1D, type 1 diabetes; UC, ulcerative colitis; VT, vitiligo. No overlap was observed for Addison’s disease, autoimmune thyroid disease, Graves’ disease, juvenile idiopathic arthritis, primary sclerosing cholangitis, psoriatic arthritis, and sarcoidosis. Genetic regions with strong evidence of selection attributed to host–pathogen coevolution were compiled from a recent review [66]. Established SSC loci (genome-wide significance or two independent replications with \( P < 5 \times 10^{-8} \)) are denoted by an asterisk. Approaches used include genome-wide association (GWA), candidate gene (CG), Immunochip (IC), GWAS follow-up (GFU), and IC follow-up (IC FU) studies. SSC phenotypes with stronger association than SSC are shown (lcSSC: limited cutaneous SSC; dcSSC: diffuse cutaneous SSC; AFA: antifilaggrin antibody; ACA: anticytokeratin antibody; SScPAH: SSc-associated pulmonary arterial hypertension). Autoimmune disease (AD) sharing based on genetic regions reported in the National Human Genome Research Institute’s Catalog of Published Genome-Wide Association Studies (http://www.genome.gov/gwastudies) accessed on 27 March 2015 [80].
potential causal variants. Approaches include fine mapping, resequencing, and imputation of variants not directly genotyped. Since the first GWAS and, more recently, Immunochip studies, a plethora of manuscripts following up individual candidate genes in independent cohorts have been published, sometimes with statistical significance below suggestive evidence for association. Hence, caution must be used when interpreting association results.

Early in 2014, the results of the first Immunochip study [26] were published, unveiling strong SSc associations in the IL12A, ATG5, DNASE1L3-PXK, and Treh-Ddx6 regions. At the same time, a GWAS follow-up study [32] reported a suggestive association with PPARG. Later in the year, a second Immunochip study confirmed the association in the DNASE1L3-PXK region and reported a novel suggestive association with VCAM1 [27]. More recently, an Immunochip follow-up study [31] reported a strong association with IL12RB1, and a candidate gene study [73] reported suggestive evidence of association between PLC2L2 and SSc.

As with the vast majority of all reported associations, the causal variants remain unknown. Only the signal in the DNASE1L3-PXK region is most likely to be driven by a missense variant in DNASE1L3 [26,27]. Consistently with the association of ACA antibody status with lcSSc, this association is stronger in ACA-positive as well as lcSSc cases. Given the role of DNASE1L3 in DNA degradation in apoptosis, loss of function of the protein and consequent defects in DNA clearance might contribute to the production of ACA [26,27].

Since, by design, the Immunochip array targets immune-related genes, it is not surprising that most of the implicated genes participate in immune processes. IL12A and ATG5 are involved in interferon signaling, and ATG5 has roles in autophagy. IL12RB1 is a component of the IL12 pathway. VCAM1 mediates leukocyte–endothelial cell adhesion and signal transduction. PLC2L2 participates in B-cell receptor signaling and B-cell proliferation. Nevertheless, one of these recently reported genes is involved in fibrosis: PPARG is an antifibrotic effector whose expression may influence the uncontrolled progression of fibrosis in SSc [32]. Collectively, these associations corroborate the role for defects in DNA clearance, autophagy, interferon signaling, IL12 signaling, B-cell signaling, cell adhesion, and fibrotic processes in disease susceptibility.

Clearly, although fibroblast activation is the hallmark of SSc, most of the genetic factors associated with SSc lie in immune-related genes [71,75], and the reasons underlying the characteristic excess deposition of extracellular matrix (ECM) proteins remain elusive. The distinct genetic architecture of pulmonary fibrosis in SSc-associated interstitial lung disease compared with idiopathic pulmonary fibrosis is noteworthy, supporting a distinct genetic risk of these two forms of lung fibrosis and suggesting that fibrosis might be the result of different pathogenic mechanisms, with a more immune-driven cause in SSc [78]. In a recent bioinformatic analysis of gene expression data from skin biopsies integrated with SSc-associated genetic variation, Mahoney et al. [79] corroborated that the genetic risk in SSc was overwhelmingly related to immune abnormalities. Their data further support the hypothesis that immune system activation is an early and key event in SSc pathogenesis likely involving interferon activation and macrophage recruitment, which may influence or drive ECM remodeling and cell proliferation in the skin [79].

**SHARED LOCI WITH OTHER AUTOIMMUNE DISEASES**

The genetic regions currently reported in the National Human Genome Research Institute (NHGRI) GWAS catalog [80] as being associated with an autoimmune disease are included in Table 1. Two-thirds (30/45) of the SSc-associated loci are also associated with at least one other autoimmune disease, 46% (21/45) with at least two diseases, 37% (16/45) with three, 24% (11/45) with four, 17% (8/45) with five, and 11% (5/45) with at least six autoimmune diseases. Since we are only including loci reported in the NHGRI GWAS catalog, which does not include all the regions with established associations with autoimmune diseases, these numbers are conservative. The non-HLA loci that SSc shares with most autoimmune diseases include TNFAIP3 (eight other autoimmune diseases), IL23R (six autoimmune diseases), STAT4 (six autoimmune diseases), and IL2RA (six autoimmune diseases). Interestingly, despite having been identified in GWAS of six different autoimmune diseases, the IL21 region has been reported as modestly associated with SSc [81], and hence did not meet the criteria to be included in Table 1. The diseases with which SSc shares the most pleiotropic effects include rheumatoid arthritis (16 genetic regions), systemic lupus erythematosus (SLE) (13 regions), multiple sclerosis (9 regions), inflammatory bowel disease (8 regions), and primary biliary cirrhosis (PBC) (8 regions). A recent clustering of autoimmune diseases based on the number of shared genetic loci using a more stringent genome-wide significance level showed SSc to be highly correlated with PBC and SLE [82]. Given the known clinical associations of SSc...
with PBC [83] or SLE [84], these findings are not surprising.

It is thought that comparison with PBC, an autoimmune liver fibrotic disorder, might help uncover the genetic risks underlying excessive fibrosis. A total of 25 loci from four GWAS are currently reported in the NHGRI GWAS catalog as being associated with PBC. As shown in Table 1, all the genetic regions shared between SSc and PBC harbor genes whose primary functions are immune-related (IRF5, IRF8, IKZF3, STAT4, IL12A, IL12RB2, BANK1-NFKB, PLCL2), providing further support for the main role of dysregulated immune pathways in these fibrotic disorders.

**POPULATION GENETIC TOOLS**

The genetic basis of disease is influenced by individual and population variation. The reasons for the geographic and ethnic disparities in the prevalence of SSc and the relative high frequency of SSc risk alleles in the population are not fully understood. Population genetic factors such as natural selection alter allele frequencies over generations and may help explain the persistence of such common risk variants in the population and the differential risk of SSc. For example, in an analysis of correlations between genetic risk of multiple diseases and worldwide migration trajectories, the authors report that variants associated with, among other, SSc, PBC, and SLE, have undergone genetic risk differentiation associated with migration [85].

As we have recently reviewed [86], immune responses can be particularly sensitive to environmental change, and immune function genes and pathways are consistently reported in tests for natural selection. Since infectious organisms are strong agents of natural selection, it is thought that the adaptation to pathogen pressure through functional variation in immune-related genes conferred a specific selective advantage for host survival, including protection from pathogens and tolerance to microbiota [87]. Interestingly, since fibrosis has a beneficial role in limiting pathogen invasion and promoting wound healing in response to injury, it has been hypothesized that fibrosis is also an evolutionarily conserved adaptive process [88]. Given the key role of immune and inflammatory processes in the cause of SSc, it is thus important to understand how population genetic factors such as natural selection have contributed to genetic variation that influences disease risk and ethnic and geographic health disparities in individuals and populations.

We have recently summarized the evidence for autoimmune disease-associated loci under selection and the candidate selective pressures [86*]. As shown in Table 1, 10 SSc risk regions show strong evidence for selection, including the HLA region, PTPN22, IL23R, TNFSF4, IL12A, TNIP1, UHRF1BP1, BLK, IRF8, and NLRP1 [86*]. Evidence of selection that has been attributed to host–pathogen coevolution is also shown in Table 1. In four of the SSc-associated regions with evidence for selection, known pathogens have been implicated as the selective pressure. These include, for example, variation in the HLA as a protective factor against bacterial infection, and resistance to protozoa as the selective pressure for PTPN22. It is noteworthy that natural selection helps explain the pleiotropy and complex genetic patterns observed at loci like PTPN22, where the same nonsynonymous variant increases the risk of some, but is protective against other autoimmune diseases [86*].

Regardless of the population phenomenon and the reasons for the emergence of both common and rare SSc-causing alleles, genetic variation is the basis for clinically relevant traits at both the individual and population levels, and thus incorporating population genetics to understand human genetic diversity will lead to a better understanding of the causes of health disparities, identification of functional variants and discovery of cellular mechanisms, and contribute to the development of new therapies.

**EMERGING STUDIES**

As mentioned above, large-scale studies have greatly enhanced our understanding of the complex cause of SSc, but a full understanding of its genetic etiology is unclear. Since these studies were designed to detect common variants, the role of rare variation remains unknown. Also, these studies were conducted in populations of European ancestry; hence variation in other ethnic groups remains elusive. The National Human Genome Institute [National Institute of Health (NIH)/NHGRI] is leading a large collaborative project entitled ‘Genome Research in African American Scleroderma Patients’ (GRASP Study) with the goal of discovering common and low-frequency variants associated with SSc susceptibility in African Americans.

The role of nonadditive inter-relationships between loci and the effect of environmental exposures in modulating these allelic effects remain unexplored. The role for epigenetic effects is now beginning to be pursued. Research on the role of the microbiome in SSc is still lacking. Variation in microbiome structure has been associated with other autoimmune diseases, and there is an increasing appreciation for the impact of the host genotype.
on the microbiome, and the impact of the microbiome on host. Finally, experimental validation of these genetic associations is lacking. Although functional causal alleles can be protein-altering, the majority of genetic changes associated with complex diseases are concentrated in regulatory DNA [15]. Despite this central importance of noncoding variants, the lack of experimentally validated regulatory mechanisms is hampering the understanding of the regulatory mechanisms underlying observed genotype–phenotype associations. Due to the advancements of sequencing technologies in the past few years, functional genomics, epigenomics, and regulatory genomics datasets are beginning to allow the dissection of the most likely regulatory variants, so that these can be experimentally validated.

For example, Farh et al. [82**] integrated genetic and epigenetic data, including transcription and cis-regulatory element annotations in primary immune cells, to identify causal autoimmune disease variants, including SSc. They found that approximately 90% of causal variants were noncoding, 60% mapped to immune-cell enhancer elements, with causal variants often occurring near binding sites for master regulators of immune differentiation and stimulus-dependent gene activation. The authors report that the enhancers and promoters mapping to SSc-associated variants did not reveal strong enrichment of specific cell subsets, but a modest enrichment of T-cell types, in a pattern more similar to vitiligo or primary sclerosing cholangitis [82**]. In contrast, both SLE and PBC variants preferentially mapped to enhancers and promoters active in B cells [82**]. Since this study only included a limited number of SSc-associated regions that met genome-wide significance level in the NHGRI GWAS catalog (13 regions) and did not include any nonimmune cell or tissue that might be relevant in SSc (e.g. skin, lung, vasculature), the results might be a consequence of the limited number of SSc-associated loci, and/or cell types analyzed. The authors concluded that the noncoding variants might cause subtle key differences in transcription or epigenetics that influence autoimmunity.

The identification of the specific cell types most relevant to disease biology is an area of major interest. This year, the NIH Roadmap Epigenomics Consortium published an integrative analysis of 111 reference human epigenomes profiled for histone modification patterns, DNA accessibility, DNA methylation, and RNA expression [89**]. SSc was not included; however, the authors reported that genetic variants associated with SLE were the most enriched in H3 lysine 4 monomethylation (H3K4me1) peaks, a marker of enhancer regions, in memory T-helper and B cells, whereas genetic variants associated with PBC were the most enriched in this enhancer mark in B cells and lymphocytes [89**]. Looking at the relevant SSc tissues, fibroblasts were the most enriched with enhancer marks for height-associated variants, and vascular tissue was enriched with aortic root size variants. These results illustrate that tissue-specific epigenomic annotations can help to identify biologically relevant cell types and to interpret genetic changes associated with disease.

CONCLUSION

In spite of the limited genetic burden, multiple genetic risk loci for SSc continue to be identified, underscoring the role of immune pathways in its genetic risk. Nevertheless, the identification of the specific disease-predisposing functional variations and reasons underlying the ethnic disparities remain unknown. African Americans suffer worse health outcomes in part because they are severely underrepresented in research and, consequently, may be omitted from developments in new genetic technologies and clinical advancements. This disengagement in research participation can only serve to exacerbate the health disparities gap as healthcare moves towards precision medicine, and must imperatively be addressed.

The integration of genetic association with population genetic and functional annotation tools will help elucidate the role of recent human adaptation in ethnic disparities and the functional role of associated risk variants in relevant cell types. This knowledge will guide gene editing studies that will uncover the disease mechanisms, which has direct implications for personalized medicine approaches.

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Conflicts of interest
There are no conflicts of interest.
Raynaud phenomenon, scleroderma, overlap syndromes and other fibrosing syndromes

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