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Haematopoietic Stem Cell Transplant for Norovirus-Induced Intestinal Failure in X-linked Agammaglobulinemia

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Abstract
Since the first clinical description in 1952, immunoglobulin replacement therapy remains the mainstay of treatment of patients with X-linked agammaglobulinemia (XLA). However, this therapy only replaces IgG isotype and does not compensate for the loss of Bruton tyrosine kinase in non-B-lymphocytes. Patients may still therefore develop complications despite current standard of care. Here, we describe an XLA patient with persistent chronic norovirus infection, refractory to treatment and causing intestinal failure. The patient underwent haematopoietic stem cell transplantation, curing XLA and allowed clearance of norovirus prior to humoral immunoreconstitution, suggesting non-humoral immunodeficiency in these patients.

Keywords X-linked agammaglobulinemia · haematopoietic stem cell transplantation · norovirus · inflammatory bowel disease

Introduction
X-linked agammaglobulinemia (XLA) is caused by defects in the gene encoding Bruton’s tyrosine kinase (BTK), characterised by the absence of peripheral circulating CD19 + B-lymphocytes and agammaglobulinemia. XLA typically presents with recurrent infections, usually by encapsulated bacteria after 6 months of age, as maternally transferred antibodies wane [1].

The mainstay of treatment is lifelong immunoglobulin replacement therapy (IgRT) and has been since XLA was first described by Colonel Ogden Bruton in 1952 [1]. There have been substantial improvements in manufacturing, tolerability, and flexibility in routes of administration of IgRT seen in recent decades [2, 3]. Unlike other primary immunodeficiencies, haematopoietic stem cell transplantation (HSCT) is not routinely offered for XLA patients, as IgRT is accepted as an effective therapy, and it is generally believed that the benefits of HSCT do not outweigh the risks.

However, as IgRT contains only the IgG isotype, XLA patients receiving treatment remain IgM and IgA deficient. In addition, BTK is expressed in all myeloid cells and may play important immune functions beyond B-lymphocytes [4]. This is supported by the presentation of XLA patients with infections other than encapsulated infection such as giardia and enterovirus [5, 6]. In addition, up to 26% of patients present with neutropenia and infections with pseudomonas [7]. Neutropenia and subsequent pseudomonal infection are rare once IgRT has been commenced, suggesting that the neutropenia is somehow IgG dependant, although the exact mechanism by which this occurs is unknown [8]. The failure to compensate for the loss of IgA,
IgM, and for the other functions of BTK implies that XLA patients may still be susceptible to complications.

This is illustrated by the increased risk of development of chronic lung disease (CLD) due to bacterial infections demonstrated by Lougaris et al., calculating the risk of developing CLD at 50 years of age of 47% [9].

Furthermore, there is gathering evidence of the impact from viral infections in XLA patients, and antibody deficiency more generally [6, 10]. Taken together, these findings suggest that the impact of the burden of bacterial and viral infection on morbidity and mortality has been significantly underestimated in XLA.

Chronic, treatment-refractory norovirus infection in common variable immunodeficiency (CVID) is well documented [11]. There is one report of chronic norovirus in an XLA patient, which was unresponsive to treatment and the patient continues with chronic norovirus infection [12]. We describe an XLA patient with persistent severe chronic norovirus-associated enteropathy resulting in intestinal failure and requiring parental nutrition (PN), for whom HSCT was performed to offer cure of XLA and clearance of norovirus infection.

Methods

Clinical data were collated from the printed and electronic medical record. Immunoglobulin levels (IgG, IgA, and IgM) were assayed by nephelometry (Siemens BN2 Nephelometer, Siemens), and serum electrophoresis (Sebia Capillaries 2; Sebia, Norcross, GA, USA). Where immunoglobulin was detectable, the lower limit of assay sensitivity (IgG, 1.34 g/L; IgA, 0.24 g/L; and IgM, 0.17 g/L) was used for data analysis. Specific antibody titres against haemophilus influenzae, tetanus, and pneumococcal capsular polysaccharide were determined by enzyme-linked immunosorbent assay (The Binding Site, Birmingham, UK). All testing was performed in the United Kingdom Accreditation Service accredited Immunology Laboratory at the University Hospital of Wales. Lymphocyte subsets were enumerated by flow cytometry. Donor chimerism was measured using short tandem repeats, in whole blood, or in separated myeloid, B-lymphocyte and T-lymphocyte lineages. All procedures were performed after informed consent was given by the parents, according to institutional protocols.

Results

The first male child of non-consanguineous Caucasian parents presented with varicella at age 8 weeks, followed by 2 episodes of zoster. Hypogammaglobulinaemia was noted on his second admission, associated with neutropenia. A diagnosis of XLA was made aged 8 months based on absent B-lymphocytes, and low monocyte BTK expression. T-lymphocyte subsets and PHA proliferation studies were normal. Sanger sequencing confirmed a previously reported c.1750 + 2 T > C pathogenic variant in BTK [13]. IgRT was commenced with intravenous immunoglobulin (IVIg) loading and maintenance subcutaneous infusions (SClG) at 1 year at 0.1–0.2 g/kg/week. In the first 2 years of life, he was well, other than mild, recurrent Haemophilus influenzae conjunctivitis. Between the ages of 3 and 6 years, he required 4 courses of intravenous broad-spectrum antibiotics for respiratory infections. His mean IgG trough level was 6.7 g/L. The intracellular pathogen, Mycobacterium abscessus, was repeatedly cultured on induced sputum samples, and bronchoalveolar lavage (BAL) isolated Haemophilus influenza. Thoracic computerised tomography (CT) showed no bronchiectasis. Treatment with 3 weeks of intravenous cefoxitin, imipenem, amikacin, with oral co-trimoxazole, and clarithromycin was successful in eradicating Mycobacterium abscessus. At the age of 5 and a half years of age, faltering growth was noted associated with increased stool frequency, urgency, and nocturnal disturbance. An empirical course of high-dose metronidazole failed to ameliorate symptoms. Coeliac risk-associated HLA DQ2 or DQ8 alleles were absent. Faecal calprotectin was elevated (226.3 and 583.6 μg/g; normal level < 100 μg/g) and abdominal ultrasound showed three separate areas of small bowel intussusception, despite remaining pain-free. At this time, no pathogens were isolated in the stool and the intussusception resolved without any intervention. Upper and lower gastrointestinal tract endoscopy showed no gross abnormalities. Duodenal biopsy showed evidence of chronic enteropathy with villous blunting and increased intraepithelial lymphocytes (Fig. 1).

Extended stool microbiology testing identified type 2 norovirus, but was negative for all other pathogens, including Giardia lamblia and Campylobacter jejuni (Fig. 2). During this time, his oral intake became restricted, he experienced severe nausea and bloating, and his quality of life significantly decreased. Despite dietary supplementation including nasogastric (NG) feeding, his growth faltered, falling from the 99.6th centile for weight and height aged 6.2 years to the 58th centile for weight and 41st for height by age 8.0 years (Fig. 3A, B). He subsequently failed to gain further weight, and serum albumin at this time was 27 g/L, associated with peripheral oedema (Fig. 3D). Serum trough IgG levels fell to 2.73 g/L despite escalating subcutaneous replacement doses in line with body weight. In order to maintain an adequate trough level exceeding 6 g/L, 3 weekly intravenous immunoglobulin infusions were instigated alongside weekly SClG, equating to a weekly IgRT dose of 0.5–0.7 g/kg/week (Fig. 3C). Bovine colostrum, oral ribavirin, and modified release oral budesonide were trialled; however,
norovirus and gastrointestinal symptoms persisted. Age 8 years, acute-on-chronic flares of nausea, vomiting, and diarrhoea prompted instigation of PN. This allowed modest, if intermittent, weight gain over the next 6 months. In view of the faltering growth and treatment-resistant intestinal failure associated with chronic norovirus infection in the setting of primary immunodeficiency, HSCT was offered.

Aged 8 years old, the patient underwent a 10/10 HLA-matched unrelated peripheral blood stem cell transplant (CD34 + dose $9.0 \times 10^6$kg). Conditioning consisted of fludarabine 150 mg/m$^2$, treosulfan 42 g/m$^2$, thiotepa 10 mg/kg, and alemtuzumab 1 mg/kg. Graft-versus-host disease (GvHD) prophylaxis consisted of mycophenolate mofetil until day +42 and ciclosporin.

Pre-transplant, *Streptococcus pneumoniae* was detected on routine bronchoalveolar lavage and treated with IV co-amoxiclav. *Campylobacter jejuni* was detected in stools and treated with azithromycin from day −4. His stools were negative for *Campylobacter jejuni* by day +3.

He developed moderate acute kidney injury (eGFR 54 ml/min/1.73 m$^2$), which self-resolved within the first 12 months post-transplant. There were no other HSCT-related complications. He achieved neutrophil engraftment on day +19 and platelet engraftment on day +17. He demonstrated no evidence of GvHD. Whole blood chimerism on day +19 was 100%. His latest chimerism, +843 days post-HSCT, demonstrates 78% CD15 donor chimerism, 93% B-lymphocyte donor chimerism, and 71% donor T-lymphocyte chimerism.
By day + 88 post-HSCT, the patient’s stooling frequents had reduced from 6 times day pre-HSCT to twice a day and to once daily by day + 108. Norovirus became undetected by PCR on day +147 post-HSCT, for the first time in over 19 months, and parenteral nutrition was discontinued. At this stage, his IgRT dose was reduced to 0.15 g/kg/week. IgRT dosing was stopped at day +224, and at last follow-up, his CD19 count is 660 cells/μL (normal range 200–1600), IgG 12.4 g/L (normal 4.9–16.1), and IgM 0.78 g/L (0.5–1.80). He demonstrated excellent vaccine antibody responses following post-transplant routine childhood vaccinations (total pneumococcal antibody titre 153 mg/L, HiB antibody > 9 mg/L, and tetanus 1.64 IU/mL). A normal enteral diet has been resumed without the need for supplementation, and the patient is thriving. Height has returned to the 75th centile for age and weight between the 91st and 99th centile. His lung function is normal (FEV1 103% predicted, FVC 102% predicted).

**Discussion**

Norovirus-associated enteropathy in primary and secondary immunodeficiency is recognised as a chronic and debilitating disorder for which no reliable treatment options exist [14]. Given the clinical and biochemical evidence of intestinal failure and the risks associated with long-term parenteral nutrition, we sought to correct the underlying immunological
defect. To our knowledge, this is the first reported case of chronic norovirus-associated enteropathy in X-linked agammaglobulinemia cured by HSCT, although patients with X-linked SCID, poor immune reconstitution, and chronic norovirus enteropathy have responded to autologous gene therapy [15]. It is likely that our patient had protein-losing enteropathy, likely associated with chronic infection, and a degree of inflammatory colitis is also likely, which may be associated with X-linked agammaglobulinemia. We also contribute to the growing evidence base that HSCT can correct XLA [16–24] (Table 1). Of 16 cases reported to date, there is a mix of indications including a diagnosis of XLA, in countries where patient access to IgRT is not assured, malignancy and chronic, unremitting infection. All patients reported to date are alive, and all, but one, have discontinued IgRT, even in the presence of mixed chimerism. No consistent conditioning regimen has been used, but a low toxicity myeloablative regimen without total body irradiation appears satisfactory.

The mechanism by which XLA patients are susceptible to chronic norovirus infection remains unclear, but likely reflects the multiple roles of BTK in immune homeostasis. Beyond production of immunoglobulins, B-lymphocytes are potent antigen-presenting cells [25]. BTK is expressed in other immune cells, e.g., NK cells, macrophages, monocytes, neutrophils, and dendritic cells [26], where it participates in Toll-like receptor and NLRP3 inflammasome signalling [26, 27]. Loss of B-lymphocyte co-stimulation in CD4+ T-lymphocyte differentiation and impairment of types I and III interferon pathways has also been demonstrated in XLA patients [28, 29]. This is consistent with the susceptibility to a range of pathogens seen in our patient and the emerging literature that demonstrates that XLA patients are more vulnerable to frequent and chronic infections across a range of pathogens (including norovirus and mycobacterium abscessus) than has been previously appreciated [6, 12, 30].

In our patient, diarrhoea began to improve with T-lymphocyte reconstitution before any significant humoral reconstitution, although the initial reduction in stooling may have been related, at least in part to the conditioning chemotherapy, which may have effectively treated any co-existing inflammatory bowel disease. Prior to HSCT, the patient was passing 5–6 Bristol-type 6/7 stools a day. At +47 days post-HSCT, he was passing 3 stools a day, when his CD3 count was 47 cells/μL, serum IgA was <0.05 g/L, and serum IgM was 0.06 g/L. By day +88 post-HSCT, his stooling frequency was twice a day, and his CD3 count was 137 cells/μL, serum IgA was <0.05 g/L, and serum IgM was 0.35 g/L. This timing of symptom improvement with T-lymphocyte reconstitution rather than significant production of IgM/IgA lends further credence to an undefined deficit in cellular immunity pre-HSCT predisposing him to chronic norovirus infection, rather than agammaglobulinemia (noting also that prior IgRT likely contained anti-norovirus antibodies and did not clear infection).

The immunological defects associated with BTK deficiency are likely only partially corrected by current approaches to treatment and prevention of infection, which differs little from Bruton’s original prescription of immunoglobulin replacement and antibiotics. It is important to note that current IgRT products typically replace only IgG, and do not include IgA and IgM. Consistent with an incomplete protection, we have recently highlighted viral persistence in XLA and other patients with predominant antibody deficiency despite standard of care [30].

There are a number of XLA patients who experience an inflammatory bowel disease-like phenotype, some of whom are given a formal diagnosis of Crohn’s disease [31]. The pathogenesis behind this inflammatory phenotype in XLA is poorly understood. Whilst this might be a direct consequence of BTK deficiency, it is possible that this inflammation is a post-viral infection phenomenon.

Patients with XLA are at elevated risk of developing malignancy and inflammatory bowel disease, risks which are unlikely to be altered by IgRT [16, 31]. Whilst HSCT is not currently routinely considered for patients with XLA, given the availability and safety of IgRT, we believe a paradigm shift of thought may be required. For instance, Ikegame et al. have recently reported successful HSCT for an adult with XLA experiencing recurrent infections and increasing risk of end-organ damage [19]. HSCT for XLA may be appropriate as standard of care in health care settings where reliable access to IgRT is not possible, particularly in resource-poor health care settings where HSCT is more cost effective than lifelong IgRT. Across all health care settings, the cost of lifelong IgRT is considerable, whilst access remains an ongoing concern, and the potential use of immunoglobulin therapy for COVID-19 has added to those pressures [32]. Although there are no cost-effectiveness analyses available, HSCT may be more cost effective than IgRT even in developed health care settings.

The decision regarding offering HSCT for any PID is complex and the potential benefit of complete cure and improved quality of life needs to be carefully balanced against the significant complications seen in HSCT, namely infection and GvHD which are associated with significant morbidity and mortality. Whilst limited data suggest that XLA patients will experience disease-related complications despite best current therapy, this has to be confirmed in larger cohorts, and so HSCT for XLA remains controversial and novel. However, for other PIDs, such as chronic granulomatous disease, HSCT was historically considered experimental, but is now offered as routine care due to improving transplant outcomes and increased awareness of the disease complications, particularly in adults [33], and improvement in quality of life [34]. Perhaps, given the safety of HSCT
### Table 1: Summary of published data of haematopoietic stem cell transplantation for X-linked agammaglobulinemia

| Author, number of patients | Patient age (years) | Indication | Conditioning | Stem cell source | GvHD | Donor chimism | Discontinued IgRT | Outcome (FU–years) |
|---------------------------|---------------------|------------|--------------|-----------------|------|---------------|-------------------|-------------------|
| Huang et al., 2           | N/A                 | XLA        | N/A          | MSD BM          | No   | N/A           | Y                 | A 2               |
| Wan et al., 1             | 14                  | XLA        | Anti-CD3 Bu12/Cy200 | MUD UCB          | No   | 100%          | Y                 | A 1               |
| Abur-Arja et al., 1       | 14                  | AML        | TBI (1.2 Gy) Cy120 Etop40 | MUD PBSC         | aGvHD II | 100%      | Y                 | A 2               |
| Ikegame et al., 1         | 28                  | Infectious complications | ATG (2.5 mg) Flu180 Cy100 | MSD PBSC         | aGvHD I | 100%    | Y                 | A 1               |
| Van Zelm et al., 1        | 25                  | Pre-B ALL | TBI (3 Gy) Etop60 | MSD PBSC         | Limited cGvHD | 100% | Y                 | A 1               |
| Nie et al., 2             | 4                   | Infectious complications | ATG6 Flu150 Bu12 Ara-C3g/m² Cy50 | mMUD-PTCy | Limited cGvHD | 100% | Y                 | A 2               |
|                          |                     |            | Haplo-id sib-PTCy | No               | 100% | Y             | A 2               |
| Swaminathan et al., 2     | 1.42                | XLA        | TT8 Treo42 Flu160 | MSD BM          | cGvHD—skin | 100% | Y                 | A 3               |
|                          |                     |            | Bu12.8 Flu160 | No               | Rejected | N          | A                 |
|                          |                     | 1.67 (1st) | XLA        | MSD BM          | No       | Rejected | N          | A                 |
|                          |                     | 2.83 (2nd) | XLA        | MSD PBSC        | No       | 100%     | Y                 | A 1               |
| Rawat et al., 4           | N/A                 | XLA        | TT,Treo,Flu* | MSD | aGvHD | 100% | Y                 | A N/A             |
|                          | N/A                 | XLA        | TT,Treo,Flu* | MSD | No     | 100% | Y                 | A N/A             |
|                          | N/A                 | XLA        | Flu,Treo*    | Haplo-id parent | No | mixed | Y                 | A N/A             |
|                          | N/A                 | XLA (1st)  | Flu,Bu*     | MSD | No     | Rejected | N                  |
|                          | XLA (2nd)           | Flu,Treo,Flu* | MSD | No | 100% | Y                 | A N/A             |
| Bucciol et al., 1         | 14                  | Chronic Aichivirus | Treo42 Flu120 | MSD PBSC | aGvHD II | 100% | N                 | A 2               |
|                          |                     |            | Haplo-id parent | No | Mixed | Y                 | A N/A             |
| Shillitoe et al., 1       | 8                   | Chronic Norovirus | Alemtuzumab TT10 Treo42 Flu150 | MUD PBSC | N/A | 78% CD15 93% CD19 71% CD3 | Y | A 2               |

**Notes:** ALL, acute lymphoid leukaemia; AML, acute myeloid leukaemia; Ara-C, cytarabine; ATG, anti-thymocyte globulin; BM, bone marrow; Bu, busulfan; Gy, cyclophosphamide; Etop, etoposide; Flu, fludarabine; a/cGvHD, acute/chronic graft-versus-host disease; Gy, gray; MSD, matched sibling donor; (m)MUD, (mis)matched unrelated donor; N/A, not available; PBSC, peripheral blood stem cells; PTCy, post-transplant cyclophosphamide; TBI, total body irradiation; Treo, treosulfan; TT, thiopeta; UCB, umbilical cord blood; XLA, X-linked agammaglobulinemia; *doses not available.
in the current era, and the morbidity and decreased quality of life documented in XLA patients on lifelong antibody treatment [35], HSCT should be considered more often, particularly in younger patients, for whom transplant results are most successful.

In conclusion, this case demonstrates the curative role of HSCT for XLA alongside the complication of norovirus-associated enteropathy. It illustrates the importance of correction of cellular immune responses in this disease, and the limitations of conventional therapeutic approaches to prevent or treat illness. Continued improvement in the safety profile and outcomes for HSCT and related approaches, such as gene therapy, are likely to widen the offer of a similar curative approach to other patients with XLA in the future.

**Author Contribution** ARG and SJ conceived the study. BMJS and MP collated and analysed the data and wrote the manuscript. MAS, JE, SS, MC, and ID provided clinical data. All authors contributed to writing of the manuscript and approved the final version.

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**Data Availability** For further information, please contact ARG: a.r.gennery@ncl.ac.uk.

**Declarations**

**Ethics Approval** Not applicable.

**Consent to Participate** The family gave fully informed consent to treatment according to institutional protocols.

**Consent to Publish** The family gave fully informed consent to publication.

**Competing Interests** The authors declare no competing interests.

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