Novel Serotype of Bluetongue Virus, Western North America

To the Editor: Bluetongue is an arboviral disease of domestic and wild ruminants characterized by vascular injury that produces widespread edema and tissue necrosis (1). Bluetongue virus (BTV), the causative agent of bluetongue, is the prototype virus of the genus Orbivirus in the family Reoviridae (2).

BTV occurs throughout temperate and tropical areas of the world coincident with the distribution of vector Culicoides spp. midges (3–5). Different midge species transmit different constellations of BTV serotypes in distinct global episystems (3,5). For example, C. sonorensis is the principal, if not exclusive, vector of BTV serotypes 10, 11, 13, and 17 in much of North America, whereas C. insignis is the major vector of multiple BTV serotypes (including BTV 1–4, 6, 8, 12, 17, 19, 20, and probably others) in the Caribbean basin, Central America, and South America. C. insignis is also found in the southeastern United States, and although this species might have recently expanded its range in the region, its distribution in North America remains poorly defined. Serotypes of BTV other than 10, 11, 13, and 17 are found in areas of the United States: BTV-2 was first reported in Florida in 1982. Since 1998, ten additional serotypes (BTV-1, 3, 5, 6, 9, 12, 14, 19, 22, and 24) have been identified in the southeastern United States (6).

Approximately 26 BTV serotypes have been described and the global distribution of BTV has recently been altered (2,4). Coincident with the invasion of novel BTV serotypes into the southeastern United States (6), likely by extension from the adjacent Caribbean basin, multiple BTV serotypes have spread throughout much of continental Europe and parts of the British Isles and Scandinavia, precipitating an economically devastating epidemic (7). Similarly, ongoing surveillance has identified novel BTV serotypes in regions to which it historically has been endemic (e.g., Australia and the Middle East) (2). Climate change may have contributed to this dramatic recent expansion in global distribution of BTV, most notably in Europe (8).

Bluetongue was first described in the late 19th century among sheep brought from Europe to South Africa, and later in North America in ≈1950 (4). Surveillance in western North America since that time has confirmed that only BTV-10, 11, 13 and 17 are present in this region, including our recent intensive surveillance of sentinel cattle on dairy farms throughout California, USA (9,10).

However, during investigation of an outbreak of acute coronitis and ulcerative stomatitis among cattle at a dairy farm in the northern Sacramento Valley in California in August 2010, a blood sample from a heifer was found by using described methods (10) to be positive for BTV by serogroup-specific quantitative reverse transcription PCR (qRT-PCR) but negative by serotype-specific qRT-PCRs for BTV-10, 11, 13, and 17.

Further analysis using additional serotype-specific qRT-PCRs identified virus in the blood sample as BTV-2. BTV was isolated in primary bovine endothelial cells from blood collected from the heifer. Sequence analysis of the serotype-specific L2 gene of the virus isolate confirmed it to be BTV-2 (2), and phylogenetic analyses showed it to most closely related to a strain of BTV-2 isolated in Florida in 1999 (Figure). However, sequence analysis of the entire genome of the virus from California indicated that it is a reassortant that includes genes from BTV-6 and BTV-2. Specifically, genes encoding the viral protein 1 polymerase and viral protein 3 major core protein segregate with those of the US prototype strain of BTV-6 (isolated in 2006), but other genes are derived from BTV-2. BTV-2 and BTV-6 have been isolated only in the southeastern United States, which
indicates translocation within the United States of reassortant BTV-2.

How this virus spread to California is not known, and its distribution in the United States is uncertain because there is no comprehensive national BTV surveillance program. However, BTV-2 was not detected previously in California, suggesting that this serotype was recently introduced into the region or that it is uncommon. Identification of this novel BTV serotype in western North America emphasizes the need for ongoing entomologic and livestock surveillance, particularly in light of recent changes in the global distribution and nature of BTV infection (4,6,8).

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References

1. Maclachlan NJ, Drew CP, Darpel KE, Worwa G. The pathology and pathogenesis of bluetongue. J Comp Pathol. 2009;141:1–16. http://dx.doi.org/10.1016/j.jcmpath.2009.04.003
2. Maan S, Maan NS, Nomikou K, Batten C, Antony F, Belaganahalli MN, et al. Novel bluetongue virus serotype from Kuwait. Emerg Infect Dis. 2011;17:886–9. http://dx.doi.org/10.3201/eid1705.101742
3. Gibbs EP, Greiner EC. The epidemiology of bluetongue. Comp Immunol Microbiol Infect Dis. 1994;17:207–20. http://dx.doi.org/10.1016/0147-9571(94)90044-2
4. Maclachlan NJ. Bluetongue: History, global epidemiology, and pathogenesis. Prev Vet Med. 2011;102:107–11http://dx.doi.org/10.1016prevetmed.2011.04.005
5. Tabachnick WJ. Culicoides and the global epidemiology of bluetongue virus infection. Vet Ital. 2004;40:144–50.
6. Johnson DJ. Identification of new United States bluetongue types. Proceedings of the United States Animal Health Association. 2011;11:209–10.
7. Saegerman C, Berkvens D, Mellor PS. Bluetongue epidemiology in the European Union. Emerg Infect Dis. 2008;14:539–44. http://dx.doi.org/10.3201/eid1404.071441
8. Purse BV, Brown HE, Harrup L, Mertens PP, Rogers DJ. Invasion of bluetongue and other orbivirus infections into Europe: the role of biological and climatic processes. Rev Sci Tech. 2008;27:427–42.
9. Osburn BI, McGowan B, Heron B, Loomis E, Bushnell R, Scott JL, et al. Epizootiologic study of bluetongue: virologic and serologic results. Am J Vet Res. 1981;42:884–7.
10. Mayo CE, Barker CM, Mullens BA, Gerry AC, Mertens PP, Maan S, et al. The combination of abundance and infection rates of Culicoides sonorensis estimates risk of subsequent bluetongue virus infection of sentinel cattle on California dairy farms. Vet Parasitol. 2012;187:295–301. http://dx.doi.org/10.1016/j.vetpar.2012.01.004

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Hepatitis E Virus Genotype 3 Strains in Domestic Pigs, Cameroon

To the Editor: Hepatitis E virus (HEV) is a positive-stranded, non-enveloped RNA virus of the family Hepeviridae that is considered to be the main causative agent of enterically transmitted acute hepatitis (1). HEV is classified into 4 genotypes (1). HEV genotypes 1 and 2 cause large waterborne epidemics of acute hepatitis in developing countries, especially in Africa and Asia (1).

In contrast, HEV genotypes 3 and 4 are increasingly identified as causative agents of acute viral hepatitis in industrialized countries (1). Genotypes 1 and 2 are found only in humans, whereas genotypes 3 and 4 are associated with food-borne zoonotic transmission from domestic pigs, wild boar, and deer (1).

In addition to these 4 genotypes, HEV-related viruses were detected in avian, rodent, and bat hosts, which formed novel genera within the family Hepeviridae (2). In Africa, HEV genotype 1 and 2 strains have been identified during HEV epidemics (3–5). An HEV genotype 3 strain was detected in 1 of 40 fecal samples from domestic pigs in Kinshasa, Democratic Republic of the Congo, and it was suggested that this strain was imported from Belgium to the Democratic Republic of the Congo by animal trade (6). Therefore, we investigated whether HEV strains of genotype 3 or 4 are circulating among domestic pigs in Cameroon.

During February–March 2012, a total of 345 liver samples were collected from domestic pigs (age range 6 months–3 years) in abattoirs in Douala and Yaoundé, Cameroon, and in slaughter slaps (areas) in Bamenda, Cameroon. Pigs were mainly of the local breed. In addition, pigs originating from extensive cross-breeding (local X landrace and local X Duroc) were sampled. Liver samples were collected during post-mortem inspection.

Viral RNA was extracted from liver samples by using the RTP DNA/RNA Virus Mini Kit II (STRATEC Molecular, Berlin, Germany) according to the manufacturer’s instructions. Extracted RNA was analyzed for HEV RNA by using 2 nested reverse transcription PCRs (RT-PCRs) specific for open reading frame 1 (ORF 1) and ORF 2 of HEV (7,8). Nested RT-PCRs and direct sequencing of amplicons were performed as described (9). RNA of