Review

Risk Factors and Pathogenesis of HIV-Associated Neurocognitive Disorder: The Role of Host Genetics

Ian Simon Olivier 1,*, Ramón Cacabelos 2 and Vinogran Naidoo 1,*

1 Division of Physiological Sciences, Department of Human Biology, Faculty of Health Sciences, University of Cape Town, Observatory 7925, South Africa; iansimonolivier@gmail.com
2 EuroEspes Biomedical Research Center, Institute of Medical Science and Genomic Medicine, 15165 Bergondo, Corunna, Spain; rcacabelos@euroespes.com
* Correspondence: vinogran.naidoo@uct.ac.za; Tel.: +27-21-406-6243

Received: 23 October 2018; Accepted: 12 November 2018; Published: 14 November 2018

Abstract: Neurocognitive impairments associated with human immunodeficiency virus (HIV) infection remain a considerable health issue for almost half the people living with HIV, despite progress in HIV treatment through combination antiretroviral therapy (cART). The pathogenesis and risk factors of HIV-associated neurocognitive disorder (HAND) are still incompletely understood. This is partly due to the complexity of HAND diagnostics, as phenotypes present with high variability and change over time. Our current understanding is that HIV enters the central nervous system (CNS) during infection, persisting and replicating in resident immune and supporting cells, with the subsequent host immune response and inflammation likely adding to the development of HAND. Differences in host (human) genetics determine, in part, the effectiveness of the immune response and other factors that increase the vulnerability to HAND. This review describes findings from studies investigating the role of human host genetics in the pathogenesis of HAND, including potential risk factors for developing HAND. The similarities and differences between HAND and Alzheimer’s disease are also discussed. While some specific variations in host genes regulating immune responses and neurotransmission have been associated with protection or risk of HAND development, the effects are generally small and findings poorly replicated. Nevertheless, a few specific gene variants appear to affect the risk for developing HAND and aid our understanding of HAND pathogenesis.

Keywords: HIV-associated neurocognitive disorders; HIV-associated dementia; HIV-encephalitis; neuroAIDS; host genetics; risk factors; HIV; cognitive impairment; polymorphisms

1. Introduction

The human immunodeficiency virus (HIV) epidemic is a huge public health issue worldwide, and is prominent in developing countries. According to UNAIDS (2018), approximately 36.9 million people were living with HIV in 2017. Eastern and southern Africa have the greatest proportion of cases by far, with an overall prevalence of 7.4% [1]. HIV is spread by bodily fluids (blood, semen, breast milk, rectal and vaginal fluids), with sexual transmission generally being the most common, although sharing of drug injection equipment is also a common mechanism. HIV is a retrovirus that causes progressive disease resulting in immune system destruction through CD4+ T-cell, macrophage and monocyte infection [2]. If left untreated, HIV infection ultimately leads to acquired immunodeficiency syndrome (AIDS), which is associated with characteristic opportunistic infections due to a weakened immune system. The advent of combination antiretroviral therapy (cART) has drastically decreased the rate of progression of AIDS and death, leading to a much improved quality of life and approximating...
a near-normal life expectancy. Treatment has now improved to such an extent that HIV is presently viewed as a chronic disease, rather than a terminal illness [3].

HIV infection, however, still presents with considerable neurological pathology in daily clinical settings, which can affect the everyday functioning of patients [2,3]. Fortunately, since the introduction of cART, the prevalence of more severe neurocognitive conditions caused by opportunistic infections and the virus itself, such as HIV dementia, have decreased profoundly. The characteristic “AIDS dementia complex” is now much rarer. Nonetheless, HIV causes a variety of milder, yet no less common forms of neurocognitive impairments (NCIs), collectively named HIV-associated neurocognitive disorders (HAND). The prevalence of HAND amongst HIV positive individuals in South Africa has been found to be between 23.5%, and is even higher in other settings and countries, with up to 60% presenting with HAND, depending on the setting [4,5]. HAND comprises several different types of cognitive impairments, predominantly presenting with impairments in attention, memory [6], processing speed and executive function [7]. Emotional manifestations, such as depression and apathy, and, to a lesser degree, motor deficits also occur [2,3]. Impairments of memory and executive functioning often affects workplace functioning and medication adherence, as does depression [3,6]. Importantly, failure of HIV therapy, which is a common problem in developing countries, is associated with higher risk of progression of neurocognitive impairments [3]. In fact, nearly 58% of the burden of dementia occurs in low- and middle-income countries [8]. Many of those affected do not receive the treatment they require and experience severe cognitive and psychiatric problems, thus decreasing the quality of life for those patients [7,8]. The prevalence of dementia in developing countries is increasing, and it is expected that within the next 20 years, these countries will have 71% of all people living with dementia [8]. This is in direct contrast to Western Europe and the United States, reporting a 22–40% decrease in prevalence [9], and an overall incidence of 9% [10], respectively, which is attributed to improved education, lifestyle and living conditions. In South Africa, which is grappling with the HIV/AIDS epidemic, there are about 187,000 documented people living with dementia [11], suggesting a significant problem for the African continent as a whole. In South Africa, and other African countries, inequalities in the distribution of health care resources remain. People from poor socio-economic backgrounds have limited access to health care, with the additional problem that proper medical treatment is hampered due to a lack of resources in the South African public health care system [12–14]. Most patients cannot afford to access private health care due to the cost of the required medications, medical consultations and evaluations [15]. With a vital need for better therapeutic avenues, therefore, it is important to find effective health care treatments against dementia-related neurodegeneration, which includes HAND.

2. Classification and Diagnosis of Human Immunodeficiency Virus-Associated Neurocognitive Disorder (HAND) Phenotypes

HAND is currently sub-classified into three forms, according to the Frascati criteria [16]: asymptomatic neurocognitive impairment (ANI), mild neurocognitive disorder (MND) and HIV-associated dementia (HAD) (Figure 1). This criteria has also been adapted by other countries on the African continent, such as South Africa [4]. All of those forms can present with or without HIV encephalitis (HIVE), the active infection of brain parenchyma by HIV, which has become less common in HAND with cART and is not a necessary feature of HAND pathogenesis [17]. HAD is the most severe, constituting a type of subcortical dementia, but has become relatively rare following cART [18]. HAD is diagnosed when at least two cognitive domains are impaired, and subtypes are differentiated by the presence and degree of impairment of everyday functioning [16]. As such, HAD and MND present with varying degrees of impairment in activities of daily living, while no such functional impairments are seen in ANI [16]. ANI is currently the most prevalent manifestation of HAND, including in developing countries like South Africa, while HAD is now relatively rare (less than 5%, depending on the setting) [5]. Importantly, the clinical usefulness or relevance of diagnosing the asymptomatic neurocognitive impairments remains controversial, and the Frascati criteria has been
accused of being overly inclusive, thus inflating the severity of impairments [19]. Although the rate and degree of neurocognitive decline in HAND is low, ANI often progresses to symptomatic neurocognitive impairment, even with a suppressed viral load [20]. Thus, the importance of ANI in a clinical setting should not be underestimated. There have also been criticisms that diagnoses based on self-reported measures of functional impairment are imprecise, as they are highly variable and influenced by social, educational and cultural factors [18]. This has posed significant challenges to studies addressing HAND, partly explaining the variable findings and the few replications of associations between risk factors (including genotypes) and HAND manifestations found in previous studies, where different neuropsychological measures were often used for diagnosis [21]. Furthermore, while rapid screening tests for HAD are often less sensitive, NCI for the Frascati criteria can be measured using different batteries of neuropsychological assessments and tests, thus enabling classification according to the Frascati criteria. One of these, which is relatively quick to use, is the international HIV dementia scale (IHDS), shown to be adequately sensitive in developed countries [22]. However, using the same range of neuropsychological tests in low-resource settings such as in sub-Saharan countries like South Africa is often less feasible or valid. This could be either due to the extensive expertise and time needed to administer these test [23] or simply because tests such as the IHDS have been found to be less sensitive and valid in such settings. This may be due to the prevalence of different HIV subtypes in such countries; for example, clade C is the dominant subtype by far in sub-Saharan Africa [5,24,25]. Therefore, additional but specific neuropsychological tests such as the Trail-Making Test and Digit Span Test can be used to assess NCI more appropriately within such settings [23]. In addition, there is a need to develop tests that are appropriate for developing countries which are normed according to those specific populations, rather than norms from studies in developed countries [4,5,24–26], such as the central nervous system (CNS) HIV Antiretroviral Therapy Effects Research (CHARTER) studies [7]. It is thus important to note that HAND consists of several, variable phenotypes that can be crudely classified, but not without limitations.

**Figure 1.** Categories and types of human immunodeficiency virus (HIV)-associated neurocognitive disorder (HAND), according to the Frascati criteria. When diagnosing patients with HAND, it is important to consider comorbidities, such as substance use and psychiatric illnesses, and to exclude other possible causes. Neurocognitive tests should evaluate at least the following: verbal/language skills, attention-information processing/working memory, memory (both learning and recall), motor skills, sensory perceptual, abstraction/executive function, and speed of information processing. Functional impairments, regarding mental acuity, social interactions, work, and other aspects, are usually self-reported, but witnesses are helpful [16,18]. HIVE, HIV encephalitis.
Although HAD and the final fully-blown subcortical dementia of the pre-cART era have become less frequent, the milder neurocognitive impairments are no less prevalent, with the CHARTER cohort indicating a high prevalence of HAND in over 50% of people living with HIV in developed countries like the USA [7]. The prevalence in developing countries like South Africa is similar, with a wide range from 15 to 60%, depending on the setting [4,5,24,25]. The causes of these cognitive impairments are likely to be multifactorial, and include factors such as cardiovascular and metabolic effects, drug use and depression [27]. This makes current treatment protocols that mainly focus on the HIV infection and control through antiretroviral medication (ARVs) less successful, and it is becoming increasingly important to consider additional contributing factors [2]. As such, while there is biological plausibility in using ARVs with greater CNS penetration to treat HAND, the effectiveness of this approach is still unclear and other types of intervention are also required.

3. Pathogenesis of HAND

The pathogenesis of HAND itself is complex and multidimensional, with current understanding still largely incomplete, although intensive investigations have uncovered several mechanisms involved. A detailed analysis of the pathogenic processes in HAND is beyond the scope of this review and described thoroughly elsewhere [17,28,29]. Two main processes are thought to cause neuronal injury that underlies HAND pathogenesis: direct neurotoxicity by HIV and/or its viral proteins, and, more importantly, indirect neuronal damage through neuroinflammation, the so-called ‘bystander effect’ [17,28–30]. Another key pathogenic component relates to resident microglia and astrocytes, in addition to some T cells, that act as latent HIV reservoirs despite cART. The importance of such reservoirs was highlighted when six patients had profound reductions in HIV levels and infection following allogeneic hematopoietic stem cell transplantation, although the exact mechanisms are as yet unknown [31]. HIV enters the CNS early in infection, primarily through infected lymphocytes and monocytes that cross the blood-brain barrier (BBB). Migration across the BBB and CNS tropism may be affected by HIV subtypes, resulting in differential susceptibility to HAND. HIV-1 subtype (or clade) C, for example, is predominant in India and sub-Saharan Africa, and may be associated with decreased CNS severity [32]. Subtype B, on the other hand, is more dominant in the USA and Europe, and was associated with higher pre-cART incidence levels [33]. Migration through the BBB by peripheral mononuclear cells in the blood is increased by astrocytes and microglia releasing chemokines—cytokines that induce chemotaxis, such as macrophage inflammatory protein-1 (MIP-1, or CCL3/CCL4) and monocyte chemoattractant protein (MCP-1, or CCL2) [34]. In the CNS, HIV productively infects macrophages and microglia, and, to some extent astrocytes, although they do not support viral replication [35]. Direct HIV infection of neurons does not occur especially with cART and, therefore, does not explain the cause of neuronal damage and/or apoptosis associated with neurocognitive impairment (NCI) [3,17]. Neurotoxic viral proteins, however, are shed by the virus and released by infected cells (macrophages, microglia, and, to a lesser degree, astrocytes), and can still cause direct neuronal injury [36–39]. These include viral surface glycoprotein (gp120), transactivator of transcription (Tat) and viral protein R (Vpr). Importantly, these viral proteins are also indirectly neurotoxic, contributing to neuroinflammation (the ‘bystander effect’) through activation of macrophages, microglia and astrocytes by binding to the α-chemokine receptor, C-X-C chemokine receptor (CXCR4) and C-C chemokine receptor 5 [17,28–30,39–41]. The most intensely studied and perhaps most important viral protein is glycoprotein120 (gp120), a neurotoxic surface protein that enables HIV entry into CD4 T cells, or T helper cells. It has been particularly implicated in the neuropathogenesis of HIV/AIDS, mostly in the presence of glial cells [29,36,41]. Gp120 can also cause excitotoxicity by binding to N-methyl-d-aspartate (NMDA)-coupled ion channel receptors (NMDARs) on neurons, resulting in excessive calcium influx, while Tat phosphorylates NMDARs which potentiate glutamate excitotoxicity [38]. Both Tat and Vpr have also been implicated in neuronal damage, with considerable neuronal apoptosis associated particularly with Tat [37–39]. Importantly, most subtype C viruses have a mutation in the gene encoding the Tat protein resulting
in a Cys31Ser polymorphism which disrupts the dicysteine motif making the protein homologous to β-chemokines [42,43]. This is possibly responsible for decreased neurotoxicity and monocyte chemotaxis, when compared to subtype B viruses [44]. This may be the factor reducing HAND severity in Indian populations [45]. In fact, NMDA-receptor activation by the mutated form of the Tat protein leads to reduced human neuronal cell death in clade C compared to clade B [43,46], data that correlate to elevated neurobehavioral deficits in clade C than clade B-infected mice [44]. This suggests that neurotoxicity may be clade-specific [43,47]. However, direct neurotoxicity by the virus is an infrequent occurrence in HAND pathogenesis, with neuroinflammation playing a far more significant role [17,29].

Neuroinflammatory processes in HAND are regulated by perivascular macrophages, microglia and astrocytes [29], which release neurotoxic substances and inflammatory cytokines upon immune activation or viral infection. This leads to impaired neuronal function, injury, and, to some extent, apoptosis [28]. Both infected and activated macrophages and microglia release inflammatory cytokines with direct neurotoxicity, such as tumour necrosis factor-alpha (TNF-α) [48,49], and indirect neurotoxicity, such as interleukin-1beta (IL-1β). Moreover, chemokines are also released by macrophages and microglia, as well as astrocytes, and play an important role in regulating neuronal injury. While α-chemokines (or CXC chemokines) can be neurotoxic [such as stromal-derived factor (SDF)-1], C–C-chemokines, particularly MIP-1 and RANTES, may have neuroprotective effects and protect against HIV/gp120 binding and neurotoxic processes [29,36,50,51]. Furthermore, infected/activated macrophages and microglia release neurotoxic substances, including ATP, arachidonate, and the excitatory amino acids, glutamate, quinolinate, and cysteine, while astrocytes also release glutamate and nitric oxide radicals [28,52]. Ultimately, this leads to disturbed bioenergetic homeostasis, neuronal injury and dendritic/synaptic pruning through both neuroinflammation and direct neurotoxicity [17]. The release of more inflammatory cytokines, particularly TNF-α, IL-1β, and SDF-1, further activate other macrophages and microglia, inducing the production of TNF-α [28]. Furthermore, gp120 and inflammatory cytokines TNF-α, IL-1β and interferon-gamma (IFN-γ) stimulate activation and proliferation of astrocytes (astrocytosis). Activated astrocytes release nitric oxide that further damages neurons directly through lipid peroxidation [50,52]. Both arachidonic acid and TNF-α released from infected and activated brain macrophages/microglia increase astrocytic glutamate release and impair its uptake and clearance [28,50]. Subsequently, excessive increases in the concentration of the neurotransmitter glutamate produce excitotoxicity [28]. Excitotoxicity occurs when excess glutamate activates neuronal NMDARs, resulting in abnormally high calcium influx, and further free radical (NO) formation, mitochondrial injury and production of reactive oxygen species (ROS) concomitant with lipid peroxidation and caspase activation that damage neurons [17,28,48]. Thus, HAND pathogenesis primarily involves the response of non-neuronal cells (perivascular macrophages, microglia and astrocytes) to HIV infection and specific viral proteins that results in a neurotoxic process. Although higher HIV levels in cerebrospinal fluid (CSF) are associated with HAND, they do not correlate well with the degree of cognitive impairment [6]. This points to the importance of host immune response and host factors in the pathophysiology of HAND, rather than only the direct neurotoxic effects of HIV [41]. Importantly, while subtype C is also predominant in sub-Saharan Africa, levels of HAND are much greater compared to India, and more similar to regions where subtype B predominates [45,53]. This is particularly due to HIV-1 subtype C Tat proteins having an intact dicysteine motif in sub-Saharan variants, unlike the south-east Asian subtype C variants which developed later on. The intact dicysteine motif in the sub-Saharan subtype C Tat protein, being more homologous to β-chemokines (like the subtype B Tat protein), induces a greater rate of monocyte chemotaxis and release of inflammatory cytokines and chemokines (such as CCL2, IL-4 and IL-10) from astrocytes and microglia than clade B Tat protein [44,46,53,54]. This points to the importance of geographical location, as well as viral genotypes and phenotypes in the pathogenesis of and risk factors for HAND.
4. Risk Factors for HAND

Several risk factors for HAND have been described, including lower levels of education, early immunosuppression, increasing age, and increased plasma concentration of TNF-α, MCP-1, and nadir CD4 T cell levels, in particular [6,17]. Moreover, cardiovascular factors such as blood pressure and hyperlipidaemia were found to worsen neurocognitive function in HAND [17]. Since many of these factors are controlled by the genetic make-up of the host, there is ongoing research into which genes may be associated with development and/or progression of HAND. Thus far, two genome-wide association studies (GWAS) have been conducted [55,56] and several candidate gene studies have investigated genes implicated in HAND and/or NCIs in HIV infected individuals (for an older, but thorough review, see Kellianpur and Levine (2014) and Levine, Parvos and Horvath (2014) [57,58]). Neither GWAS found any genes associated with HAD or milder neurocognitive impairments, as assessed by neuropsychological tests or global deficit score (GDS), that reached the genome-wide significance threshold. However, the latter GWAS by Jia et al. (2017), looking at GDS, found some biologically plausible gene variants previously associated with Alzheimer’s disease (SH3RF3 and CSMD1), as well as four single nucleotide polymorphisms (SNPs) at the T-cell receptor alpha locus, involved in neurogenesis, which did approach significance [55]. Both studies were, however, limited by sample size, indicating a need for further research to support and validate previous findings. In the present paper, we aim to provide a brief overview of some of the genes and the related mechanisms that have been found to be associated with HAND development and progression, and specifically those found in candidate gene studies.

5. Genes Involved in Neurotransmitter Systems

In the brain, the neurotransmitters dopamine and serotonin are associated with the control of executive functions, memory and attention. Several genetic polymorphisms of those neurotransmitters are associated with cognitive functions [59]. Although these were found to have small effects, they may have more significant impact with pre-existing cognitive deficits, such as in HAND [60]. In view of this, several studies have investigated the effect of genetic polymorphisms related to dopamine and serotonin neurotransmission on cognitive functions in HAND [60–62].

5.1. Genes Related to Serotonin Neurotransmission

Two SNPs of serotonin-related genes have been associated with NCI in HIV-positive(+) adults, particularly in African American males [61,63]. A homozygous TT genotype in the SNP rs4570625 was associated with poorer executive functioning in HIV+ individuals, most of whom were with ANI, while a small proportion had MNI (HAD was not found in any participants), as assessed by Frascati criteria [61]. This SNP is located in the transcriptional region for the gene encoding tryptophan hydroxylase 2 (TPH2), the rate-limiting enzyme in serotonin biosynthesis, and affects the transcription rate of TPH2 [61]. The same study further found that the polymorphism GALM rs6741892 of the galactose mutarotase gene (GALM), which indirectly increases local serotonin release, was linked to impaired memory in the same participants [61]. The SLC6A4 5-HTTLPR polymorphism, associated with serotonin transport, had no significant effect. These results were in agreement with earlier findings of associations between the same SNPs and poorer executive function and memory [63], with some limitations due to the cross-sectional design and lack of a healthy control group.

5.2. Genes Related to Dopamine Neurotransmission

Other studies have looked at the effect of genetic polymorphisms related to the neurotransmitter dopamine, which is involved in executive function, working memory, and drugs-of-addiction and reward pathways in the CNS [60,64]. Dopamine is largely implicated in the fronto-striatal system, which consists of connections between the midbrain and prefrontal cortex, areas important for various cognitive functions. Studies have found that the fronto-striatal system is particularly affected in HAND
and HIVE, demonstrating both structural and metabolic abnormalities and pathology (for review, see Hauser and Knapp, 2014) [65]. Chronic HIV infection results in depleted levels of dopamine and a reduced quantity of dopamine receptors in the basal ganglia, with dopaminergic neurons being particular vulnerable to injury by Tat and gp120 [65]. In the context of HAND, the dopamine-related genes that have been studied include genes for: catechol-O-methyltransferase (COMT), an enzyme involved in dopamine metabolism; the dopamine transporter (DAT), the channel for the re-uptake of dopamine from the synaptic cleft; brain-derived neurotrophic factor (BDNF) which is involved in regulating dopaminergic and serotonergic activity; and dopamine beta-hydroxylase. Those genotypes have been found to affect cognitive functioning in HIV-negative (−) populations, but their role in HAND is still unclear due to the limited number of reported studies [60]. Nonetheless, the genes encoding COMT and dopamine receptors do seem to be associated with various measures of NCIs in HAND populations [64,66–68]. Regarding the COMT gene locus (rs4680), it was found that the Met/Met genotype (val158met) was associated with improved performance in various executive functioning tests in HIV-infected individuals with mild cognitive impairment, but not in those concurrently using methamphetamine [66]. This is due to this variant resulting in reduced COMT activity in metabolising dopamine, thus increasing levels of dopamine [66]. SNPs in the dopamine receptor gene DRD2 (rs6277), which affect D2 receptor density in the striatum, is associated with somewhat impaired cognitive flexibility and executive function in adults with HIV [64]. Variations in the DRD4 gene (48 base pair-variable number tandem repeats) were also linked with impaired executive function, which affect both D2 receptor density and binding affinity [64]. Similarly, DRD1 and DRD2 gene polymorphisms have main effects with substance use on neuropsychological performance in various cognitive domains in HIV+ individuals [68], with opposite effects in those who used substances compared to those who did not. Furthermore, a genetic polymorphism of DRD3 (rs6280TC)—resulting in a serine to glycine amino acid substitution and subsequent increase in dopamine binding affinity of the D2 receptor DRD3, also found in macrophages—was linked to considerable cognitive impairment in HIV-infected methamphetamine (meth) users [67]. That dopamine-related gene variants influence cognitive impairment in HAND also has biological plausibility considering a different mechanism: monocyte infection and chemotaxis is increased by extracellular dopamine concentrations [69]. The gene variants mentioned cause changes of extracellular dopamine and affect the binding affinity and density of DA receptors expressed in monocytes, such as DRD1, DRD5, and DRD4 [69]. The findings linking dopamine-related genes to cognitive impairment, however, may not be specific to HAND populations as they have not been adequately replicated in the context of HAND, and because similar trends have also been observed in HIV negative populations. In fact, the two GWAS studies [55,56] did not replicate those relationships with the COMT, DRD1, or DRD2 gene polymorphisms. Importantly, a longitudinal study [60] extending over 10 years did not find any effects of the genetic polymorphisms of COMT val158met, BDNF val66met, and the DAT genes on neurocognitive functioning (as assessed by comprehensive neuropsychological exams) in HIV+ populations, but did not include people with HIV dementia. Another extensive candidate gene study, which did assess HIV dementia scores, found no associations either [62].

6. Genes Affecting Integrity of Mitochondrial and Nuclear DNA in HAND

HAND pathogenesis, to a large degree, involves the production of ROS and oxidative damage which affect both nuclear and mitochondrial DNA structure and function, ultimately leading to neuronal apoptosis [17,70]. One important pathogenic mechanism in HAND includes Tat-induced double strand breaks in DNA which is lethal to cells if left unrepaired [71] (for review, see [71] for HIV-induced mitochondrial toxicity and [72] for mitochondrial DNA (mtDNA) haplogroups related to HIV infection and treatment). Only a few studies, however, have investigated both nuclear and mitochondrial oxidative DNA damage in neurocognition in HIV, and its role in neuronal apoptosis and HAND pathogenesis [73–75]. Higher levels of mtDNA damage are associated with poorer general neurocognitive function in HIV+ patients [71]. Moreover, a frequently found mtDNA deletion mutation

Int. J. Mol. Sci. 2018, 19, 3594 7 of 33
(known as the “common deletion”) was associated with NCI in HIV+ patients using methamphetamine, as assessed by a continuous GDS (with a GDS > 0.5 being the criteria for NCI) [70]. This mutation, consisting of a 4977 base pair deletion affecting genes involved in the respiratory chain, causes mitochondrial dysfunction and is more common in brain tissue in neurodegenerative diseases [70]. Furthermore, post-mortem tissues from the prefrontal cortex in HIV− and HIV+ patients with and without non-specific HAND (assessed by the International HIV Dementia Scale) were compared and modification of 8-hydroxydeoxyguanosine (8-oxoG), a marker for oxidative damage, examined [74]. Patients with HAND of any severity had particularly high levels of nuclear and mitochondrial DNA 8-oxoG damage, compared to HIV+ patients without HAND and HIV− controls. Furthermore, mtDNA in the HAND group was found to have a much greater number of non-coding displacement (D) loop mutations. Both of those features indicate that higher levels of ROS contribute to HAND pathogenesis [74]. Interestingly, oxidative stress and neuronal apoptosis induced by gp120 and Tat can be reduced by the delivery of antioxidant enzymes through gene transfer [73]. Another cross-sectional study, which looked at GDS results and Frascati criteria for HAND diagnosis, found that mtDNA haplogroup B is associated with less NCI and decreased prevalence of HAND of any severity in individuals with Hispanic ancestry [76]. No associations of mtDNA variations with HAND were found in individuals of European or African ancestry. Thus, the ‘common deletion’ may be a risk factor of HAND development, with mtDNA haplogroup variation having an ancestry-specific influence on neurocognitive impairment in HAND.

7. Telomere Length and Cell Age

Telomerase activity is closely related to oxidative damage. This enzyme extends the length of telomeres, structures of repetitive DNA sequences at the end of chromosomes that protect against the shortening of DNA during cell division. Telomere length also gives an indication of biological age, since they shorten with each cell division cycle depending on the level of telomerase activity. Telomerase activity is impaired by oxidative stress [57], and telomere length decreases with chronic psychological stress [77]. In the context of HAND, there was a positive correlation between longer telomere length in leukocytes and learning performance (as part of the International Neuropsychological Test Battery developed by the HIV Neurobehavioural Research Centre) in a group of HIV+ women in South Africa (HAND severity was not indicated) [77]. Shorter leukocyte telomere length may also indicate immune senescence, suggesting accelerated cellular aging in HIV disease [78]. However, these findings were not replicated; a different study found no associations between leukocyte telomere length and overall neurocognition (assessed by a comprehensive neuropsychological battery, with mild to moderate NCI) [79]. Comparing biological age to chronological age by measuring the levels of epigenetic DNA methylation can also indicate whether accelerated aging is a feature of HAND. In fact, epigenetic age is increased in HAND, but has not been associated with HAND severity [80]. Moreover, the protein HIV-1 Tat has been found to upregulate the expression of histone deacetylase-2 (HDAC2) in human neurons [81], resulting in a decreased number of acetyl groups and subsequent chromatin condensation with reduced transcription.

8. Genes Related to Cytokines and Associated Receptors of the Immune System

8.1. Gene Variants of CCR2 and Associated Ligand CCL2 (MCP-1)

MCP-1 has been associated with HAND more consistently than most other gene products. Increased levels and expression of MCP-1 were found in the CSF in HIV-infected patients with increased GDS (indicating greater cognitive impairment) [82] and in astrocytes in AIDS dementia [40]. MCP-1 plays an important role in neuroinflammation in HAND, as it draws immune cells, monocytes in particular, to the CNS across the BBB [83]. However, it has also been shown to protect neurons from apoptosis induced by NMDA or HIV Tat. Several studies have looked at genetic polymorphisms in the MCP-1 gene and Prep 1, a transcription factor binding to the MCP-1 promoter region, as well as the
MCP-1 target receptor CCR2 [40,55,56,84–91]. One SNP in the MCP-1 gene (rs1024611), resulting in the MCP-1-2578G allele which is more easily transcribed, increases CSF MCP-1 concentrations [86] and possibly monocyte infiltration [82]. Importantly, this allele has been associated with a highly increased risk of AIDS dementia [84] and greater GDS along with other CSF pro-inflammatory markers [82]. The longitudinal study by Levine et al. (2014) also found a lack of improvement in working memory compared to HIV+ participants (not including HAD patients) with the allele, albeit a small effect [60]. The most recent GWAS study showed that this same MCP-1 G allele was linked to a decreased risk of NCI as indicated by GDS, but only with nominal significance [55]. Other studies, however, did not find these associations in adults with AIDS dementia or neurocognitive impairment in HIV-positive children [87–89], nor did the other GWAS [56]. These inconsistent findings may be due to variations in the PREP-1 gene polymorphisms that affect transcription of MCP-1 [89]. Indeed, the heterozygous genotype for SNP rs2839619 in the PREP-1 gene was much less common in HAD cases compared to controls, suggesting a protective effect [89]. Several studies have also looked at polymorphisms of the C–C chemokine receptor type 2 (CCR-2), which is the natural ligand for MCP-1 and a minor HIV-1 co-receptor. Only a single study found associations between CCR-2 gene polymorphisms and HIV-associated neuropsychological impairment, where the CCR2-64I allele was associated with a greater rate of progression to NCI over time in adults [87]; several other studies found no effects with NCI or AIDS dementia in HIV+ adults [91,92], or impairment in various cognitive domains in children [85] with HIV.

8.2. Gene Variants of CCR5 and Associated Ligand CCL3 (MIP-1α)

MIP-1α (or CCL3) is a C–C chemokine secreted by microglia and astrocytes, and binds to its natural ligand CCR5, one of the most important HIV co-receptors [41]. Due to its competitive binding and protection from gp120-induced apoptosis in neurons, MIP-1α, and MIP-1β (CCL4), have shown neuroprotective roles in vitro [48]. The longitudinal study by Levine et al. [60] found that one SNP resulting in a homozygous or heterozygous A allele in the MIP-1α gene (rs1719134), instead of a GG allele, was associated with declining ability in learning and memory in adults with HIV, but not HAD. Similarly, an earlier study found that the AA allele at another SNP in the MIP1-α gene (rs1130371) increased the risk of HAD two-fold [88]; another study demonstrated no differences in HAD [89]. Other studies have looked at copy number variants (CNV) in the MIP-1α gene, particularly CCL3L1. An increased CCL3L1 copy number may hinder HIV binding and infection of cells by increasing MIP-1α production, outcompeting HIV for CCR5. While a below average CCL3L1 copy number was associated with higher susceptibility to HIV/AIDS progression [34,93], one study found no differences in CCL3L1 CNV between NCI, mild NCI, or HAD, while another found no differences between no NCI and NCI (as determined by GDS > 0.5) in a Chinese cohort [90,92]. MIP-1α may thus play a dual role in HAND: while it binds to neurons and prevents neurotoxicity from HIV/gp120, it is also a potent chemoattractant, and variations in its genotype (particularly SNP rs1719134), may increase monocyte CNS entry, exacerbating neuroinflammation [21].

The CCR5 receptor is a crucial co-receptor for HIV entry into monocytes and microglia, and mutated CCR5 affected HIV disease progression and neurocognitive impairment in the pre-CART era [94,95]. Importantly, a meta-analysis found that CCR5 locus variation, together with the human leukocyte antigen (HLA) locus explained the large majority of variation in set-point viremia; increased HIV disease progression was associated with variants (CCR5P1 haplotype) in the CCR5-promoter region [96]. The CCR5 Δ32 allele (rs333) arises from a deletion of 32 base pairs within the CCR5 gene, and in loss of CCR5 receptor expression and function [97]. This mutation protects against infection in those homozygous for the allele, while those heterozygous for the gene also had delayed disease progression [97]. Indeed, the only known patient in whom HIV-1 was completely eradicated (the “Berlin patient”) had received an allogeneic hematopoietic stem cell transplant, with the donor being homozygous for the CCR5A32 allele. This polymorphism likely prevented HIV infection of the graft, thus allowing eradication of the virus [98,99]. Examination of neurocognitive impairment in children
with HIV revealed that this mutation, but not other SNPs of the CCR5 gene, slowed disease progression and decreased neurocognitive impairment [85]. The CCR5 Δ32 allele is also found at a lower prevalence in people with AIDS dementia [91,100]. This finding, however, may be specific to the pre-cART era and different viral loads, since later studies with patients on cART did not find this association [87,89,92]. In fact, Bol et al. (2012) noted that this effect occurred only in those developing AIDS prior to 1991, before cART was introduced [89]. It may be helpful to investigate how CCR5 mutations correlate with specific features of neurocognitive function and the degree of impairment, rather than simply the diagnosis of HAD. Indeed, a 24-week treatment with a CCR5 antagonist (maraviroc) produced improvements in neurocognitive function in HIV+ adults on cART [101]. This highlights the potential role of CCR5 and its genotypes in treatment of HAND.

8.3. Gene Variants of TNF-α

TNF-α is a pro-inflammatory neurotoxic cytokine that plays a role in HAND by increasing BBB permeability and activating NMDA receptors, resulting in excessive, neurotoxic calcium increase in neurons [49]. The TNF-α-308 allele (rs1800269) is associated with increased TNF-α production, and is more common in patients with HAD, compared to HIV+ or HIV− controls [102,103]. Other studies, however, found no association between this TNF-α genotype and HAD [88,89] or any effect of TNF genotype on HIVE [104,105]. Increased gene expression related to the TNF-α signalling pathway [106] in HIV infected adults, and TNF receptors [107] in AIDS dementia, have been observed, but this is controversial as other studies did not find an association between HAND of any severity and neurocognition [108,109]. Interferon-α (IFN-α) is another inflammatory cytokine that may be more important due to its role in virus-specific immune responses and activation of astrocytes, and has been positively associated with HIV [106].

8.4. Gene Variants of the Chemokine Ligand CXCL12 (SDF-1)

CXCL12, or SDF-1, is an α chemokine (CXC chemokine group) that binds to CXCR4, an important chemokine receptor expressed on neurons, microglia and astrocytes [28]. It is also the main co-receptor for HIV through which T cells are infected [50]. SDF-1 prevents HIV binding to CXCR4 by competing for it and down-regulates CXCR4 expression [110], but has been implicated in direct neuronal toxicity. It is increasingly expressed in HIVE, and has been implicated in neuronal apoptosis [36,50] and astrocytes exposed to HIV-1 [107]. A gene variant of SDF1, consisting of a SNP with a G to A substitution in the 3′ untranslated region of the SDF-1 gene (SDF1-3′-A/A polymorphism; rs1801157) has been implicated in HIV disease progression [111]. Compared to the SDF1-3′-G/G wildtype, the SDF1-3′-A/A genotype increases production of SDF1 and reduces AIDS progression [111]. However, several other studies found no such association [91,112,113]. Still yet, these studies did not specifically analyze associations with HAND, considering only disease progression in general. In the context of HAND, children homozygous for the SDF1-3′-A/A genotype experienced a much higher rate of disease progression and had a greater prevalence of neurocognitive impairment compared to those with the SDF1-3′-G/G (wild type) genotype [85]. This replicated earlier findings which linked greater HIV disease progression to the SDF-13′-A allele, in children [114]. This, however, may be a finding specific in children, as other studies did not echo any associations with the SDF1-3′-A/A genotype and neurocognitive functioning in adults [60,88,92,115]. Nevertheless, one study did indicate that SDF1 gene expression in astrocytes was up-regulated in patients and possibly implicated in HAND [107].

8.5. Genetic Variation of Interleukins in HAND

Interleukins are a large group of cytokines that are secreted by numerous immune cells and are crucial to immune system regulation. Neuroinflammation in HAND is also regulated by several interleukins, such as IL-6 and IL-10 which have been linked to BBB crossing by monocytes, and IL-1β which is a neurotoxic inflammatory cytokine released by activated microglia [41]. With regard to HIV,
an IL-2 gene variant reduces HIV infection risk [116], and a SNP in the IL-1 gene differed in those with rapid disease progression from those with slow progression [117]. Furthermore, SNPs in the gene coding for IL-1α predicted HIV-1 replication and levels of viraemia [118]. A number of studies investigated associations between SNPs in interleukin genes and neurocognitive function in HIV, but none reported any significant findings. Regarding IL-1β gene variants, neither the IL1-1β*2 allele, nor the interleukin-1 receptor antagonist (IL1RN)*2 allele (which regulates IL-1β release), had an effect in HIV [105]. Furthermore, allele frequencies of IL-1α − 889 (rs3783525), IL-1β +3953, and IL-12 were not different in people with HAD compared to controls [103], and neither were alleles of IL-1α (rs17561) or IL-10 (rs1800872) [88]. Lastly, a polymorphism in IL-4 (IL4-589-C/T) was not associated with neurocognitive function in individuals with HIV [92]. Thus, it is unlikely that genetic variants of interleukins specifically have a significant effect on neurocognitive functioning in HAND, even though these cytokines play an important role in HAND pathogenesis, as indicated by gene expression studies [107].

9. Mannose Binding Lectin (MBL-2)

Variations of the MBL-2 allele have been associated with HIV infection and AIDS progression [119], likely due to the role of MBL-2 in macrophage-related innate and adaptive immunity. For example, one study found that in children, homozygous for the B, C and D alleles (rs1800450, rs1800451 and rs5030737, respectively), experienced greater cognitive decline [120]. Those findings were supported by a study of a cohort of Chinese adults, where the same genotypes were associated with mild NCI and HAD [92]. Levine et al. (2014), however, did not replicate these findings in HIV positive adults with NCI (excluding HAD), and only the HIV-control group showed an association between the ‘O’ MBL-2 genotype and NCI. This suggested that this relationship was not associated with HIV comorbidity [60]. MBL-2 expression is increased in neurons in HIV [121], but this relationship has not been correlated with neurocognitive functioning. Although associations of SNPs of MBL-2 were specifically evaluated in each GWAS, no significant findings were reported [55,56]. It is therefore unlikely that MBL-2 has a specific role in neurocognitive impairment in the context of HIV, although earlier findings that did indicate associations warrant further research.

10. Human Leukocyte Antigen Class I and II (HLA) Genes

HLA is the protein machinery that presents antigens to immune cells on the cell surface. There are two classes of HLA, class I and class II, both encoded for by highly polymorphic genes. HLA class I genes (HLA-A, HLA-B, HLA-C) code for cell surface proteins that ‘present’ antigens produced by the host cell itself (which can include viral proteins if a cell is infected). HLA class II genes (HLA-DR, HLA-DP, HLA-DQ) code for molecules that present viral antigens engulfed by immune cells [122]. HLA I is important in viral immunity and implicated in HIV disease progression, as infected cells present viral antigens via HLA I to cytotoxic T lymphocytes (CTL), also known as CD8 T cells, in order to enable CTLs to recognize and kill infected cells [122]. For an effective CD8 T cell response, however, T helper cells (CD4 T cells) are required, which are activated by HLA II [123]. HLA alleles, particularly HLA class I alleles, affect HIV disease progression (for review, see Naranbhai and Carrington, 2017 [122]). Some of the same alleles (HLA-B*27, -B*57, -B*58:01) have also been associated with reduced HIV-associated neurocognitive functioning in adults (with and without AIDS dementia) [123] and children [124] with HIV. A study in China found that increased NCI (determined by GDS), as well as a greater rate of neurocognitive decline (over 12 months), occurred in adults carrying the HLA-DR*04 allele (an HLA class II allele which codes for a low CD4 T cell response) [123]. Furthermore, several HLA class I alleles (specifically, HLA-A*03, -A*33, HLA-B*27, -B*57 and -B*58:01) had protective effects, with less neurocognitive impairment at baseline and lower rates of decline. Those protective HLA class I alleles are associated with CD8 T cell responses to more conserved regions of HIV [123]. The HLA-DR*04 allele reduced the protective effect of those HLA class I alleles. This highlights the importance of the additional role of CD4 T cells in the control of HIV by CD8 T cells. Taken together,
these findings support an earlier study of an American cohort, where the HLA-DR*04 allele was associated with greater risk of HAND [123]. With regard to children, the HLA-B*27 genotype, a gene variant of HLA class I, was also found to protect against neurocognitive impairment and the delay onset of CNS impairment [124]. In addition, further protective effects were associated with the Cw-2 allele and HLA class II DQB1-2 allele [124]. In contrast, the HLA-A*24 genotype was associated with greater CNS disease progression. Both these studies correlate well with previous findings relating HLA alleles to HIV disease progression [122]. The expression of genes encoding HLA class 1 HLA-A, HLA-B, HLA-C, HLA-G, and HLA-F are upregulated in response to interferon type I and II in HIV [106]. Similarly, HLA-E was highly upregulated in patients with HAND (severity not specified) and HIV [108], as was HLA-B [125]. In a more recent study, the HLA-DR genotype did not correlate significantly with neurocognitive functions (assessed as a global clinical rating; GCR) or histopathological markers in HIV-infected adults [21]. Research investigating the role of HLA genes have possibly been limited possibly due to the large sample sizes that are required for HLA genotyping. Given the clear role of HLA alleles on HIV disease progression and potential associations with cognitive impairment, more studies are required to investigate the impact of HLA gene variants in the context of HAND [122].

11. Matrix Metalloproteinase (MMP) Genes

Matrix metalloproteinases (MMPs) are enzymes involved in remodelling of the extracellular matrix, and, together with the associated natural tissue inhibitors of MMPs (TIMPs), play a crucial role in chronic inflammation and HAND pathogenesis [73]. Increased expression and activity of MMPs and imbalances with TIMPs, cause injury and subsequent increase in BBB permeability by breaking down the tight junctions of endothelial cells and the extracellular matrix [126]. This results in increased migration of infected and uninfected immune cells into the CNS and may thus lead to development of HAND. The genetic polymorphisms of some of these have been implicated in HAND pathogenesis, although there are few studies and no replications of findings. For example, a case control study found that the MMP-21 572 C/T genotype was associated with increased risk of HAND development, as were polymorphisms of MMP-1 (-1607 2G/1G) and MMP-3 (MMP-3-1612 6A/5A allele) [127]. Other MMP-3 alleles (MMP-3-1612 5A allele and MMP-3-1612 5A/5A allele) presented a lower risk of HAND [128]. A polymorphism of the MMP-2 gene (MMP-2-735 C>T) has been implicated in risk of HAND development and severity, both alone and synergistically with a MMP-9 variation (MMP-9-1562 C>T) [129]. A SNP of the MMP-7 gene, resulting in MMP-7-181 A or G genotypes, was not associated with HAND, even though MMP-7 is upregulated by the Tat protein [130]. Similarly, MMP-8 gene polymorphisms (−799C/T and +17C/G) were not significantly associated with a risk of HAND [131]. All these case control studies included HAND of any severity, diagnosed by having an International HIV-associated Dementia Score (IHDS) of less than 9.5. Thus far, these findings have not been replicated, and further research is required to support the reported associations between MMP polymorphisms and HAND. Nonetheless, gene expression of various MMPs is upregulated in HAND, and play a significant role in HAND pathogenesis [73].

12. Contribution of Host Genes in Alzheimer’s Disease (AD) and HAND

The most common cause of dementia is the degenerative disease AD. Both HAND/HAD and AD share common histopathological, pathophysiological and genetic features. Synaptic damage in particular appears to be common to both disorders. A study in the South African context reported the prevalence of dementia in a low-income rural community to be 8–12% for those aged 60 years or more, and 11% for those above 65 years of age [132]. Therefore, the expected burden of dementia in South Africa is greater than expected. That study did not, however, seek to investigate other risk factors for dementia such as the apoE4 genotype linked to Alzheimer’s disease (AD). AD appears to occur as a consequence of a sustained neuroinflammatory process caused by a plethora of different factors such as trauma, oxidative agents, infection, and oligomers of tau and Aβ [133,134]. Histologically, features of AD include extensive cerebral atrophy coupled to neuronal loss, formation of
characteristic neuritic plaques comprising an extracellular core with Aβ amyloid and entangled neuritic processes, appearance of neurofibrillary tangles with hyperphosphorylated tau protein, and synaptic loss [134–136]. In approximately 98% of AD cases, the typical form of the disease is a sporadic or a late onset syndrome, mostly affecting patients older than 65 years of age. Of the 695 genes associated with AD, the most important is that encoding the lipid transporter apolipoprotein E (apoE) found on chromosome 19 (19q13.2) [137]. ApoE, produced primarily by astrocytes and activated microglia, is also responsible for the modulation of synaptic plasticity as well as facilitation of brain repair pathways in response to injury [138–140]. Among the three apoE alleles (apoE2, apoE3, and apoE4), homozygosity for the apoE4 allele is a strong genetic risk factor in those patients for developing late-onset AD. It should be noted that individuals with the apoE4 genotype generally experience cognitive decline twice as fast as non-carriers of the apoE4 allele [141,142]. Compared to apoE2 and apoE3, the apoE4 allele increases, at a high rate, the conversion of amyloid-β precursor protein (APP) into β-amyloid via a non-canonical mitogen-activated protein (MAP) kinase signaling cascade. This involves phosphorylation of MAP Kinase Kinase 7 (MKK7) by the dual leucine-zipper kinase DLK which then phosphorylates extracellular signal-regulated kinase (ERK)1/2. Phosphorylation of c-fos by ERK1/2 stimulates the transcription factor activator protein 1 (AP-1) inducing APP gene transcription, thus enhancing production and accumulation of Aβ that injures neurons [143].

HIV-infection accelerates and accentuates premature aging of the nervous system by potentiating, at least in part, immunological senescence, measured by the accumulation of CD28+/CD57+ T cells [78,144] and activation of the monocyte markers CD163 and CXCL10 [145]. The neurocognitive performance of young (<40 years of age) HIV+ patients was found to be equivalent to those of older (≥50 years of age) seronegative individuals [146,147]. Several studies have reported a positive correlation between HAND of any severity and AIDS dementia and β-amyloid deposition in the CNS in patients with the apoE4 genotype [148–150]. Such reports have provided glimpses into the interaction between apoE4 and HIV infection [92,147,151,152]. Sections of temporal lobe/hippocampus, entorhinal cortex and frontal lobes from the brains of HIV-1 infected patients obtained at autopsy were immunohistochemically stained with antibodies against Aβ and phosphorylated-tau (p-tau) [149,153]. In contrast to the neuritic Aβ plaques observed in the AD brain, diffusely distributed Aβ plaques and sparsely-scattered neurofibrillary lesions were detected in both HIV cases and non-HIV controls [144]. Those features correlated with aging rather than symptomatic AD. ApoE4 increases somatodendritic tau accumulation and neuroinflammation in mice expressing the human apoE4 isoform [154–156]. In a gp120-transgenic mouse model of HAND, levels of total and p-tau were elevated in hippocampal dentate gyrus granule cells and CA3 pyramidal cells, which is important given the role of the hippocampus in learning and memory [157]. In postmortem brain samples from ART-treated HIV-infected patients, p-tau levels were increased compared to age-matched non-HIV subjects [158–160]. An association between the apoE4 genotype and tau pathology in HAND is not clear.

Interestingly, the abundance of cortical Aβ deposition and synaptic loss was associated with HAND of any severity only in those individuals carrying the apoE4 allele [149], a feature that correlated with poor cognitive function and performance [92,161]. In fact, there are strong associations between apoE4 and the rate of HIV-1 disease progression (enhanced viral attachment and fusion) [151], and between apoE4 and HAD in older (≥50 years) HIV-1 infected patients. This is despite the fact that those same patients do not show an increased risk for HAD [147,162]. Moreover, in an 11-patient HIV cohort with and without the apoE4 allele, patients with the apoE4 genotype were found to exhibit very mild/moderate dementia [152]. The human apoE4 isoform is also the least effective in Aβ clearance compared to apoE2 or apoE3 [163]. Structurally, the N-terminal domain (amino acid residues 1–191) of apoE contains the receptor-binding region, while the C-terminal domain (residues 206–299) consists of amphipathic α-helices and the high-affinity lipid-binding region (residues 244–272) [164,165]. The apoE4 isoform preferentially binds to triglyceride-rich very low density lipoproteins (VLDL) [166], with evidence indicating that the amphipathic α-helix domain of apoE4 may be important in facilitating HIV-1 fusion by binding to gp41 [167]. This contrasts
with apoE3 which inhibits HIV infection [168]. Importantly, a truncated form of apoE4 (residues 1–244) has been found in AD brains in which the lipid binding C-terminal domain induces CNS toxicity [169]. HIV+ patients with symptomatic neurocognitive impairment expressing the apoE4 allele also exhibited enhanced glial metabolite myo-inositol:total creatine ratios [170] and reduced neuronal metabolite N-acetyl aspartate:total creatine ratios, indicating cellular injury or aging [171]. Persistent glial (astrocytes and microglia) cell activation was detected in the basal ganglia, frontal white matter and parietal cortex [170]. Those individuals also performed poorly in cognitive tests [51,170], suggesting susceptibility to HAND. The introduction of cART has led to dramatic reductions in morbidity and mortality in HIV-infected patients. Even with cART, chronic HIV-infection continues to drive systemic immune activation and an inflammatory response (HIVE) [172,173]. In HIVE, the BBB barrier is disrupted and microglia and astrocytes are persistently activated [30,35,174]. cART drugs also disrupt microglial function, increasing neuronal synthesis of Aβ [175] but reducing clearance of β-amyloid [176]. The net result is progressive worsening of inflammation and brain damage. Therefore, it is likely that a neuroinflammatory response may favor Aβ plaque formation. Neuroinflammation is in fact a critical element in the brain’s response to injury and a common feature when analyzing the progression of HAD and AD.

Gene expression microarray analysis on HIV-1-infected cultured human neuronal progenitor (hNP1) cells treated with recombinant human apoE4, revealed that from the 85% of genes downregulated, 22% of those [including early B-cell factor 3 (EBF3), myelin transcription factor 1, forkhead box C1, neurogenin 2, basic helix-loop-helix family member e22, doublecortin, neurofilament medium polypeptide and internexin neuronal intermediate filament protein alpha], are involved in neurogenesis [177]. Those data, together with earlier evidence implicating apoE4 as a transcriptional repressor [178] indicates that apoE4 may be detrimental to neuronal functions, thus suggesting a further mechanism of apoE4’s effect on neurocognition in HAND. The hippocampus, being one of the first brain regions to be affected in AD, is particularly vulnerable to injury in this disease [179]. In the dentate gyrus in 10-week-old C57BL/6] mice expressing the human apoE4 allele, nestin-positive newborn neurons had lower dendritic spine densities associated with fewer dendritic branches [180]. This is significant when considering the potential impact on the learning and memory circuitry and cognitive decline.

Excitotoxicity is associated with pathogenesis in several neurological injury states including AD and involves excessive stimulation of glutamate receptors with consequent neuronal degeneration. Over activation of glutamate receptors increases nitric oxide and free radical release, and Bax protein activation through elevated levels of intracellular calcium [181]. Tat and gp120 trigger excitotoxicity in the CNS via persistent activation of NMDA receptors causing neuronal apoptosis [37–39]. Experimental models further reveal that Tat protein also stimulates Aβ production [182], prevents its degradation by inhibiting the amyloid-degrading enzyme neprilysin [183,184], and promotes the size and numbers of the β-amyloid plaques by potentiating β-site APP cleaving enzyme 1 (BACE1) (or β-secretase) activity [52]. Recently, Stern and others (2018) found increased Aβ oligomer and BACE1 protein expression in the hippocampus and dorsolateral prefrontal cortex of HIV-infected patients diagnosed with HANDs of any severity [185]. The same authors further treated Sprague–Dawley rat cortical neuron cultures with the supernatant of HIV-1-infected human monocyte-derived macrophages and discovered NMDA-receptor-dependent elevation of BACE1 levels. Taken together, those data suggest that BACE1 inhibition may provide a therapeutic option in patients with HAND. Glutathione S-transferase pull-down assays in cultured human fetal neurons showed that apoE4 and Tat are competitive ligands for low-density lipoprotein receptor-related protein 1 (LRP1) [186]. However, according to Liu et al. (2000), Tat uptake/endocytosis into neurons through LRP1, mediated by initial, rapid, and high-affinity binding to the protein heparan sulfate proteoglycan, prevents apoE4 uptake/clearance [186]. This increases the half-life of apoE4 in the cerebral circulation, and as apoE4 microglia are neurotoxic, more so than apoE3 [187], brain injury may be augmented [186].
Diffusion tensor imaging (DTI) is routinely used to detect changes in cerebral white matter integrity [188,189]. A characteristic neuropathological feature in the brain of AD and HIV-infected patients is damage to the corpus callosum; the medial temporal lobe is also vulnerable to injury in individuals with AD and both types of damage correlate to cognitive impairment [190–196]. The white matter of the body of the corpus callosum is damaged in HIV- patients homozygous for apoE4, but not in non-HIV-infected patients negative for apoE4 [195,197]. Positron emission tomography (PET) scans with the radiotracer $^{18}$F Florbetaben have been used as a screening tool for the presence of β-amyloid plaques. It is worth noting that several studies found no evidence of amyloid burden in HIV-infected patients with the apoE4 genotype [198–200]. Therefore, it is possible that apoE4 does not affect HAND. However, as Cysique et al. (2015) suggest, the accumulation of β-amyloid is a gradual process. In that study [200], a possibility is that affected patients with HAND were still relatively young and that the association between the apoE4 allele in HIV+ individuals may thus be age-dependent. This is supported by several studies which found that apoE4 genotypes had differential effects across age groups (so-called “antagonistic pleiotropy”), with more pathological implications on neurocognitive function in older patients, (see review, Geffin and McCarthy, 2018). CNS HIV infection may not preclude Aβ/β-amyloid deposition [201]. Here, a 71-year-old HIV-infected male on cART, whose apoE genotype remains unknown, was diagnosed with mixed HAND and (possibly) AD syndromes [201]. $^{18}$F Florbetaben was localized to the frontal, temporal, and parietal lobes bilaterally including the posterior cingulate/precuneus, reported by the investigators as consistent with AD [201]. Chang et al. (2013) using magnetic resonance imaging (MRI), found that older (≥50 years) HIV+ and seronegative apoE4-carriers had smaller global brain volumes compared to younger (<50 years old) seronegative apoE4 carriers; when age-matched (<50 years old) to younger HIV-infected non-apoE4 carriers, younger HIV-infected apoE4 carriers displayed striatal and general white matter atrophy [202], indicating a role for apoE4 as a possible risk factor for premature aging. Despite reports that HIV-infected patients carrying the apoE4 allele have progressively reduced cognitive functions associated with a loss of white matter integrity and cerebral atrophy [203], susceptibility to cognitive impairment conferred by the apoE4 allele in patients with HAND remains controversial. Several studies [105,198,204–207] did not found any correlation between apoE4 and HIV dementia in infected individuals. These discrepancies may be attributed to an age-dependent effect of the apoE4 allele.

13. Drug Metabolism/Transporter Genes

Understanding the pharmacogenomic outcome of HAND that takes into account inter-individual genetic variations in drug metabolism is key to improving anti-retroviral drug efficacy and safety. Critical to this issue are genes that encode both drug-metabolizing enzymes (such as cytochrome P enzymes), and transporter proteins at the BBB. Polymorphisms in those genes can alter the pharmacokinetic profile of a drug, and classify people into extensive, intermediate or slow metabolizers for a particular drug. The widely-used non-nucleoside reverse transcriptase inhibitor (NNRTI). Efavirenz is often the antiretroviral drug of choice for first-line treatment of HIV [208,209], but also one that produces long-term neuropsychiatric side effects including psychosis, depression, hallucinations or dysphoria [210,211]. Efavirenz plasma levels above 4 µg/mL have been linked to high levels of CNS toxicity [212–214]. Efavirenz is oxidized mainly by hepatic cytochrome P450, family 2, sub-family B, polypeptide 6 (CYP2B6) isozyme. The CYP2B6 gene is highly polymorphic [215] and the CYP2B6 c.516 G>T variant appears to strongly predict the pharmacokinetics of Efavirenz [216]. That is, this 516G>T change, specifically in HIV-infected individuals of African-ethnicity, genetically predisposes these patients (“slow Efavirenz metabolizers”) to develop adverse CNS reactions with a frequency of 34–50% [215,217]. This has important consequences for the African continent as it may dramatically increase the burden of psychiatric illness in HIV-infected patients [217,218]. In those individuals, compared to 15–20% of Caucasian and Asian populations [213,219], CYP2B6 enzyme activity is reduced, impairing the metabolism of Efavirenz and increasing plasma levels of the drug [215,217,220]. Furthermore, the effect of the CYP2B6 G516T polymorphism on neurocognitive
performance was evaluated in non-HIV-infected individuals who were administered Efavirenz [221]. Elevated plasma levels of Efavirenz did not correlate with worse neurocognitive performance in those patients. Sandkovsky et al. (2017) examined the effect of CYP2B6 polymorphisms on neurocognitive function in older adults infected with HIV [222]. Higher levels of the Efavirenz metabolite 8-OH-Efavirenz correlated with better neuropsychological functioning while higher Efavirenz levels did not, in contrast to the findings by Johnson et al. (2013) [221]. Furthermore, the CYP2B6 G516T genotype was associated with higher Efavirenz levels in plasma, but this did not correlate with neuropsychological functioning. While these findings are perplexing, and the study was limited by its small sample size, it may point to increased metabolism of Efavirenz having a protective effect as its metabolite does not cross the BBB as easily. Whether cognitive function in HAND-positive individuals with the CYP2B6 G516T polymorphism is indeed impaired, remains undetermined. In HIV-infected persons, methamphetamine (meth) use and addiction are common and increase the rate of neurocognitive impairment [223–225]. Polymorphisms in CYP2D6 enhance the risk of neurocognitive impairment in a subset of meth users, termed “extensive metabolizers”, and may predispose such individuals who may also be HIV+, to HAND [226].

The two major groups of drug transporters for the movement of molecules across the BBB are the mostly primary active ATP-binding cassette (ABC) transporters, and secondary active solute carrier (SLC) transporters. In the CNS, ABCB1 (MDR1) encodes the important efflux transporter protein P-glycoprotein (P-gp) [227]. P-gp is known to associate with more than 1200 drugs [137] and upregulation of ABCB1 occurs in HAND [215,228,229]. Although the polymorphism 3435C>T in ABCB1 has been linked to low plasma levels of Efavirenz [230], P-gp, interestingly, has no influence on Efavirenz uptake at the BBB [231]. Johnson et al. (2013), in a cohort of non-HIV-infected adults administered with the P-gp substrate raltegravir, found that the concentrations of this integrase inhibitor in patients’ cerebrospinal fluid were unaffected by the 3435C>T variant [232]. Further studies, with increased statistical power are needed to assess the genomics of drug transporters as related to the pathogenesis and progression of HAND.

14. Summary

HAND consists of a spectrum of neurocognitive disorders and, for affected individuals, is associated with a poorer quality of life and medication adherence, decreased ability to perform complex daily tasks, and has an enhanced prevalence with other age-related diseases. Overall, patients on cART show a decline in the more severe form of HAND, but the frequency of less severe forms of cognitive impairment has increased. The neuropathogenesis is multifactorial, leading to neuronal damage or dysfunction, with CNS inflammation at the core of the disease (Figure 2). HIV crosses the BBB and accesses the CNS via a “Trojan horse” mechanism relatively early (within weeks) following exposure. Here, infected monocytes and lymphocytes enter the brain and activate and infect microglia, astrocytes and oligodendrocytes; the viral particles induce a cascade of events that are toxic to neuronal survival. HIV DNA recovered from brain tissue indicates the presence of latent cellular reservoirs (predominantly microglia/CD16+ macrophages) within the CNS for the virus. In the direct model of HIV-mediated CNS pathogenesis, viral proteins (Tat, gp120, Vpr) released from infected cells cause neuronal death. According to the indirect model, neuronal death is mediated predominantly by inflammation against HIV infection. TNF-α and IL-1β are quite likely the initiators of the inflammatory pathways that cause neuronal apoptosis. In the CNS, HIV-infected microglia and macrophages secrete a large amount of chemokines/cytokines that cause apoptosis, degeneration of the BBB, and glutamate reuptake; thus, chronic inflammation is important in the progression of HAND. However, little is known about potential risk factors of HAND, with some people being more likely to develop HAND than others, indicating the role of host factors in disease pathogenesis. There are multiple host–virus interactions that contribute to HAND, including genetic components (e.g., apoE4), metabolic disorders (e.g., insulin resistance), aging (e.g., β-amyloid and p-tau protein) (Figure 3), and vascular disease. Concerning the role of human host genetics in the pathogenesis of HAND, numerous host genes
are differentially expressed over the course of HAND, as discussed in this review. Analyzing the contribution of these host genes to neurotropic and/or neurovirulent CNS pathogenesis allows us to understand the neural effects in response to HIV over the course of HAND. The role of host genetic factors in HIV neuropathogenesis and cognitive impairment has been clouded by the critical issue of a lack of reproducibility among various studies. This is particularly due to heterogeneity and ambiguity in measuring disease outcomes (HAND phenotypes and/or measures of cognitive impairment), largely resulting from the variability of HAND presentations, subtypes, and severity. Nonetheless, polymorphisms in a number of host genes that affect common pathogenic pathways implicated in HAND, such as neurotransmitter function, nuclear and mitochondrial DNA integrity, telomere length, cytokine and chemokine activity and chronic inflammation, have differential effects on HAND susceptibility and progression, although effect sizes tended to be small. For example, the CCL2 (MCP-1) gene variant, MCP-1\(^{-2578}G\) (rs1024611) in HIV+ patients, has been associated with accelerated progression of HIV, accompanied with a 4.5-fold higher risk of developing HAD. Data from other groups, however, have yielded contradictory results, reporting no effect on neurocognitive function in HAND. HIV entry involves an initial interaction between gp120 and the host CD4 receptor, following which gp120 binds to the CCR5 co-receptor on the host cell. HIV infection of macrophages and microglia are mediated by CCR5. A mutation in the CCR5 receptor produces a truncated CCR5 receptor, the CCR5A32 variant; heterozygosity for CCR5A32 is associated with a reduced risk of impaired cognitive function in patients with HAND. Variants of HLA class I and class II gene have an important influence on the HIV-specific immune response by CD8 and CD4 T cells. Specific polymorphisms of HLA class I have been found to slow HIV disease progression as well as neurocognitive impairment, mainly by encoding a CD8 T cell response that targets more conserved regions of HIV. Some class II gene variants responsible for low CD4 T cell response to HIV may also play a role. More research is needed to establish the effects HLA gene variants have on HAND development, considering the large sample sizes needed for HLA genotyping. Furthermore, BBB disruption occurs through various inflammatory factors, including TNF-\(\alpha\) and IL-1\(\beta\). These two proinflammatory cytokines induce the expression of adhesion molecules allowing for an interaction between microvascular endothelial cells and HIV-infected cells. The host gene variant TNF-308A promotes elevated levels of TNF-\(\alpha\) and has been linked to neurotoxicity and progressive neurocognitive severity in the HIV-infected CNS; this has been found particularly in patients with HAD. A polymorphism in the IL-1\(\beta\) 3953T allele is associated with enhanced IL-1\(\beta\) expression. High levels of expression of IL-1\(\beta\) in microglia and macrophages were found in HIV-1-positive patients with HAD. Subsequent studies examining patients with the IL-1\(\beta\) 3953T, however, found no difference between patients with HAD versus HIV+ and HIV\(\neg\) individuals. The same trend was observed from analysing HIV+ patients expressing gene variants of IL-4 (IL4-589-C/T) and IL-10 (rs1800872). Taken together, the current data indicate that gene polymorphisms of interleukins may not be significant contributors to aberrant cognitive functioning in HAND. Data from studies linking the apoE4 allele to HAND have been inconsistent across cohorts, possibly due to age being an independent risk factor for HAD, and apoE4 playing a role in older HIV-infected populations, rather than younger adults. Accordingly, HAND pathogenesis has also been associated with accelerated and attenuated aging, linked to mitochondrial DNA mutations, oxidative stress, and related mitochondrial DNA damage (studies on shortened telomere length in HAND are still contradictory). Other genes implicated in the development of HAND are those coding for the various MMPs, but again, their roles have not yet been resolved. Generally, previous studies and reproducibility have been limited in several ways: relatively small sample sizes; lack of variety or inclusivity of certain study populations, with disproportionate focus on Caucasian populations and young adults, with lack of some other ethnic groups and pediatric patients; lack of specificity and clarity regarding disease outcome measures and diagnoses (for example, disease outcomes were often defined by measures of global cognitive impairment or HAND, instead of investigating domain-specific neurocognitive impairments, or HAND subtypes). It would be advisable for future studies to have larger sample sizes with different population groups and more specific outcome...
measures; the influence on particular cognitive processes and associated impairment and specific HAND subtypes, including ones that are less severe but more common, like ANI. Moreover, since HAND pathogenesis is multifactorial, it is unlikely that single gene variants are responsible or would have large impacts by themselves. Rather, the disease process is influenced by a number of variants that involve several common mechanisms, as shown in this review (such as neurotransmitter systems, immune regulation, aging, and oxidative stress). Thus, studies should investigate common pathways in HAND pathogenesis, combining these with epigenetic, transcriptomic and proteomic investigations as well histopathology and neuroimaging.

Understanding the influences of the host (and virus) genetic components that contribute to the development of HAND will help clarify the complex process of neuropsychological dysfunction in HIV+ individuals with this disorder. This would aid the development of therapeutically beneficial strategies such as pharmacogenetics procedures promoting neuroprotection against HAND.

Figure 2. Pathogenic mechanisms and possible genetic influences that lead to development of HIV-associated neurocognitive disorders. HIV and its proteins (gp120, Tat, Vpr) enter the central nervous system (CNS) mainly through infected monocytes and T cells, but also via transcytosis or paracellularly. Infected and uninfected microglia, perivascular macrophages and astrocytes release neurotoxic substances (including viral proteins, if infected), inflammatory cytokines (particularly TNF-α and IL-β), and various chemokines that further activate the same uninfected cells. Neurotoxic substances include gp120 and Tat, quinolinic acid, glutamate, arachidonic acid, and free radicals such as NO, some inflammatory cytokines (TNF-α), and chemokines and cytokines. The resulting neuronal injury is due to functional damage, such as disruption of bioenergetic homeostasis, oxidative stress, damage to synapses and dendrites, and, to a lesser extent, apoptosis. Chemokines may be protective (some C-chemokines, such as MCP-1/CCL2, MIP-1α/CCL3 and MIP-1β/CCL4) or harmful (some CXC or α chemokines, such as SDF-1). Many bind to the chemokine receptor CCR5 expressed by

Key
macrophages and many astrocytes and neurons, and importantly, CCR5 also interacts as a coreceptor with gp120, perhaps directly neurotoxic. Increased glutamate is released by neurons leading to excitotoxicity; ATP and cytokines released by macrophages, and decreased glutamate reuptake by astrocytes (as macrophages release arachidonate). This increases intracellular calcium through N-methyl-D-aspartate (NMDA)-coupled ion channel receptor (NMDAR)-coupled ion channels activated by excessive glutamate and excitotoxic substances. Finally, this results in neuronal injury through free radicals, oxidative damage, and apoptotic pathways. Potential gene variants may influence pathogenic processes by altering expression of cytokines, chemokines, MMPs, HLA types. Neurotransmitter levels (affecting cognitive processes), β-amyloid deposition, susceptibility to oxidative damage and stress, as well as aging, are further affected by genotypes. Gene variants are marked by small ‘chromosomes’ on the side of text, where green indicates protective and red harmful/neurotoxic effects. (TNF-α, tumor necrosis factor-alpha; IL-, interleukin; Tat, transactivator of transcription; gp120, glycoprotein120; NO, nitric oxide; MCP-1, monocyte chemoattractant protein; MIP-1, macrophage inflammatory protein-1; SDF-1, stromal-derived factor; NMDA, N-methyl-D-aspartate; MMP, matrix metalloproteinase; HLA, human leukocyte antigen.)

**Figure 3.** Common pathways in AD and HAND pathogenesis. In AD and HAND, there are several lines of attack that lead to neurodegeneration. β-amyloid aggregates not only have direct cytotoxic effects on neurons, but also indirectly through the generation and progression of a local neuroinflammation via activated and dystrophic resident glial cells in the vicinity of neurons. The latter, reactive gliosis, brings with its secretion of cytokines and chemokines that are released from microglia predominantly, and reactive astrocytes. The interaction between activated microglia and reactive astrocytes is an important component of HAND pathogenesis. Secreted factors from microglia include members of the complement system (C1q, C3b, C4d, C5b-9), and pro-inflammatory cytokines TNF-α, and IL-1β, IL-6, IL-10, whereas astrocytes release mainly TNF-α and IL-1β. Enhanced secretion of TGF-β from reactive astrocytes favors activation of microglia. Further, TREM2 is highly expressed by microglia and alleviates pro-inflammatory factor-induced AD pathology by mediating phagocytosis of cell debris [233]. Aβ binds to and reduces TREM2 activity; dysregulation of TREM2 signaling contributes to the pathogenesis of both AD and HAND [234]. One interesting aspect is the release of reactive oxygen species (ROS) which can have multiple effects in this inflammatory process. For example, ROS can induce neuronal protein and lipid oxidation, DNA damage, and impair mitochondrial function by disturbing the oxidative respiratory chain, affecting neuronal transport processes and the
ubiquitin-proteasome system. ROS, released from Aβ-stimulated microglia can also induce astrocytes to adopt a neurotoxic phenotype. The net result is the worsening of inflammation and brain damage. The HIV proteins Tat and gp120 activate NMDA receptors triggering excessive glutamate release and excitotoxicity. Tat also activates BACE1 promoting the formation of B-amyloid plaques, and enters neurons via LRP1 to prevent apoE4 uptake, amplifying neurotoxicity. Tat can also be synthesized and released in HIV-infected astrocytes; one source of infection is derived from activated microglia. ApoE4 is known to enhance HIV infection [151]. Neurons and microglia also secrete FGF and BDNF in an attempt to promote neuronal survival. Disease progression, however, results in the failure to repair the injured neurons. (AD, Alzheimer’s Disease; HAND, HIV-associated neurocognitive disorder; TNF-α, tumor necrosis factor-alpha; IL-, interleukin; TGF-β, transforming growth factor beta; TREM2, membrane-bound triggering receptor expressed on myeloid cells 2; ROS, reactive oxygen species; Tat, transactivator of transcription; gp120, glycoprotein120; NMDA, N-methyl-D-aspartate; BACE1, beta-site amyloid-beta precursor protein cleaving enzyme 1; LRP1, low-density lipoprotein receptor-related protein 1; FGF, basic fibroblast growth factor; BDNF, brain-derived neurotrophic factor.)

Funding: This research received no external funding.
Conflicts of Interest: The authors declare no conflicts of interest.

References
1. Danforth, K.; Granich, R.; Wiedeman, D.; Baxi, S.; Padian, N. Global mortality and morbidity of HIV/AIDS. In Major Infectious Diseases; Holmes, K.K., Bertozzi, S., Bloom, B.R., Jha, P., Eds.; World Health Organization: Washington, DC, USA, 2017.
2. Clifford, D.B. HIV-associated neurocognitive disorder. Curr. Opin. Infect. Dis. 2017, 30, 117–122. [PubMed]
3. Arendt, G.; Grauer, O.; Hahn, K.; Maschke, M.; Obermann, M.; Hussstedt, I. Neues bei HIV und neuro-AIDS. Aktuelle Neurol. 2015, 42, 445–455. [CrossRef]
4. Joska, J.A.; Fincham, D.S.; Stein, D.J.; Paul, R.H.; Seedat, S. Clinical correlates of HIV-associated neurocognitive disorders in south Africa. AIDS Behav. 2010, 14, 371–378. [CrossRef] [PubMed]
5. Mogambery, J.C.; Dawood, H.; Wilson, D.; Moodley, A. HIV-associated neurocognitive disorder in a kwazulu-natal HIV clinic: A prospective study. S. Afr. J. HIV Med. 2017, 18, 732. [CrossRef] [PubMed]
6. Eggers, C.; Arendt, G.; Hahn, K.; Hussstedt, I.W.; Maschke, M.; Neuen-Jacob, E.; Obermann, M.; Rosenkranz, T.; Schielke, E.; Straube, E.; et al. HIV-1-associated neurocognitive disorder: Epidemiology, pathogenesis, diagnosis, and treatment. J. Neurol. 2017, 264, 1715–1727. [CrossRef] [PubMed]
7. Heaton, R.K.; Franklin, D.R.; Ellis, R.J.; McCutchan, J.A.; Letendre, S.L.; Leblanc, S.; Corkran, S.H.; Duarte, N.A.; Clifford, D.B.; Woods, S.P.; et al. HIV-associated neurocognitive disorders before and during the era of combination antiretroviral therapy: Differences in rates, nature, and predictors. J. Neurovirol. 2011, 17, 3–16. [CrossRef] [PubMed]
8. Ferri, C.P.; Prince, M.; Brayne, C.; Brodaty, H.; Fratiglioni, L.; Ganguli, M.; Hall, K.; Hasegawa, K.; Hendrie, H.; Huang, Y.; et al. Global prevalence of dementia: A delphi consensus study. Lancet 2005, 366, 2112–2117. [CrossRef]
9. Wu, Y.T.; Fratiglioni, L.; Matthews, F.E.; Lobo, A.; Breteler, M.M.; Skoog, I.; Brayne, C. Dementia in western europe: Epidemiological evidence and implications for policy making. Lancet Neurol. 2016, 15, 116–124. [CrossRef]
10. Langa, K.M.; Larson, E.B.; Crimmins, E.M.; Faul, J.D.; Levine, D.A.; Kabeto, M.U.; Weir, D.R. A comparison of the prevalence of dementia in the united states in 2000 and 2012. JAMA Intern. Med. 2017, 177, 51–58. [CrossRef] [PubMed]
11. Alzheimer’s Disease International. World Alzheimer Report, Improving Healthcare for People Living with Dementia; Alzheimer’s Disease International: London, UK, 2016.
12. Wilmshurst, J.M.; Badoe, E.; Wammanda, R.D.; Mallewa, M.; Kakooza-Mwesige, A.; Venter, A.; Newton, C.R. Child neurology services in Africa. J. Child. Neurol. 2011, 26, 1555–1563. [CrossRef] [PubMed]
13. Kalula, S.; Petros, G. Responses to dementia in less developed countries with a focus on south Africa. Glob. Aging 2011, 7, 31–40.
14. Kalula, S.Z.; Ferreira, M.; Thomas, K.G.; de Villiers, L.; Joska, J.A.; Geffen, L.N. Profile and management of patients at a memory clinic. *S. Afr. Med. J.* 2010, 100, 449–451. [CrossRef] [PubMed]

15. Wilmshurst, J.M.; Kakooza-Mwesige, A.; Newton, C.R. The challenges of managing children with epilepsy in Africa. *Semin. Pediatr. Neurol.* 2014, 21, 36–41. [CrossRef] [PubMed]

16. Antinori, A.; Arendt, G.; Becker, J.T.; Brew, B.J.; Byrd, D.A.; Cherner, M.; Clifford, D.B.; Cinque, P.; Epstein, L.G.; Goodkin, K.; et al. Updated research nosology for HIV-associated neurocognitive disorders. *Neurology* 2007, 69, 1789–1799. [CrossRef] [PubMed]

17. Saylor, D.; Dickens, A.M.; Sacktor, N.; Haughey, N.; Slusher, B.; Pletnikov, M.; Mankowski, J.L.; Brown, A.; Volsky, D.J.; McArthur, J.C. HIV-associated neurocognitive disorder–pathogenesis and prospects for treatment. *Nat. Rev. Neurol.* 2016, 12, 234–248. [CrossRef] [PubMed]

18. Levine, A.J.; Soontornniyomkij, V.; Achim, C.L.; Masliah, E.; Gelman, B.B.; Sinsheimer, J.S.; Singer, E.J.; Grant, I.; Franklin, D.R., Jr.; Deutsch, R.; Woods, S.P.; Vaida, F.; Ellis, R.J.; Letendre, S.L.; Marcotte, T.D.; Kaul, M.; Garden, G.A.; Lipton, S.A. Pathways to neuronal injury and apoptosis in HIV-associated dementia. *Lancet Infect. Dis.* 2013, 13, 976–986. [CrossRef]

19. Gisslen, M.; Price, R.W.; Nilsson, S. The definition of HIV-associated neurocognitive disorders: Are we overestimating the real prevalence? *BMC Infect. Dis.* 2011, 11, 356. [CrossRef] [PubMed]

20. Grant, I.; Franklin, D.R., Jr.; Deutsch, R.; Woods, S.P.; Vaida, F.; Ellis, R.J.; Letendre, S.L.; Marcotte, T.D.; Atkinson, J.H.; Collier, A.C.; et al. Asymptomatic HIV-associated neurocognitive impairment increases risk for symptomatic decline. *Neurology* 2014, 82, 2055–2062. [CrossRef] [PubMed]

21. Levine, A.J.; Soontornniyomkij, V.; Achim, C.L.; Masliah, E.; Gelman, B.B.; Sinzheimer, J.S.; Singer, E.J.; Moore, D.J. Multilevel analysis of neuropathogenesis of neurocognitive impairment in HIV. *J. Neurovirol.* 2016, 22, 431–441. [CrossRef] [PubMed]

22. Milanini, B.; Paul, R.; Bahemana, E.; Adamu, Y.; Kiweewa, F.; Langat, R.; Owuoth, J.; Allen, E.; Polyak, C.; Ake, J.; et al. Limitations of the international HIV dementia scale in the current era. *AIDS (Lond. Engl.)* 2018, 32, 2477–2483. [CrossRef] [PubMed]

23. Singh, D.; Joska, J.A.; Goodkin, K.; Lopez, E.; Myer, L.; Paul, R.H.; John, S.; Sunpath, H. Normative scores for a brief neuropsychological battery for the detection of HIV-associated neurocognitive disorder (hand) among South Africans. *BMC Res. Notes* 2010, 3, 28. [CrossRef] [PubMed]

24. Goodkin, K.; Hardy, D.J.; Singh, D.; Lopez, E. Diagnostic utility of the international HIV dementia scale for HIV-associated neurocognitive impairment and disorder in south Africa. *J. Neuropsychiatry Clin. Neurosci.* 2014, 26, 352–358. [CrossRef] [PubMed]

25. Joska, J.A.; Westgarth-Taylor, J.; Hoare, J.; Thomas, K.G.; Paul, R.; Myer, L.; Stein, D.J. Validity of the international HIV dementia scale in south Africa. *AIDS Patient Care STDs* 2011, 25, 95–101. [CrossRef] [PubMed]

26. Cross, H.M.; Combrinck, M.I.; Joska, J.A. HIV-associated neurocognitive disorders: Antiretroviral regimen, central nervous system penetration effectiveness, and cognitive outcomes. *S. Afr. Med. J.* 2013, 103, 758–762. [CrossRef] [PubMed]

27. Schouten, J.; Su, T.; Wit, F.W.; Kootstra, N.A.; Caan, M.W.; Geurtsen, G.J.; Schmand, B.A.; Stolte, I.G.; Prins, M.; Majoie, C.B.; et al. Determinants of reduced cognitive performance in HIV-1-infected middle-aged men on combination antiretroviral therapy. *AIDS 2016*, 30, 1027–1038. [CrossRef] [PubMed]

28. Kaul, M.; Garden, G.A.; Lipton, S.A. Pathways to neuronal injury and apoptosis in HIV-associated dementia. *Nature 2001*, 410, 988–994. [CrossRef] [PubMed]

29. Gonzalez-Scarano, F.; Martín-García, J. The neuropathogenesis of AIDS. *Nat. Rev. Immunol.* 2005, 5, 69–81. [CrossRef] [PubMed]

30. Gendelman, H.E.; Persidsky, Y.; Ghorpade, A.; Limoges, J.; Stins, M.; Fiala, M.; Morrisett, R. The neuropathogenesis of the AIDS dementia complex. *AIDS (Lond. Engl.)* 1997, 11 (Suppl. A), S35–S45.

31. Salgado, M.; Kwon, M.; Galvez, C.; Badiola, J.; Nijhuis, M.; Bandera, A.; Balsalobre, P.; Miralles, P.; Buno, I.; Martinez-Laperche, C.; et al. Mechanisms that contribute to a profound reduction of the HIV-1 reservoir after allogeneic stem cell transplant. *Ann. Intern. Med.* 2018. [CrossRef] [PubMed]

32. Satishchandra, P.; Nalini, A.; Gourie-Devi, M.; Khanna, N.; Santosh, V.; Ravi, V.; Desai, A.; Chandramuki, A.; Jayakumar, P.N.; Shankar, S.K. Profile of neurologic disorders associated with HIV/AIDS from Bangalore, South India (1989–1996). *Indian J. Med. Res.* 2000, 111, 14–23. [PubMed]

33. Sacktor, N.; Lyles, R.H.; Skolasky, R.; Kleeberger, C.; Selnes, O.A.; Miller, E.N.; Becker, J.T.; Cohen, B.; McArthur, J.C.; Multicenter, A.C.S. HIV-associated neurologic disease incidence changes: Multicenter AIDS cohort study, 1990–1998. *Neurology* 2001, 56, 257–260. [CrossRef] [PubMed]
34. Dolan, M.J.; Kulkarni, H.; Camargo, J.F.; He, W.; Smith, A.; Anaya, J.M.; Miura, T.; Hecht, F.M.; Mamtni, M.; Pereyra, F.; et al. CCL3L1 and CCR5 influence cell-mediated immunity and affect HIV-AIDS pathogenesis via viral entry-independent mechanisms. *Nat. Immunol.* 2007, 8, 1324–1336. [CrossRef] [PubMed]

35. Kramer-Hammerle, S.; Rothenaigner, I.; Wolff, H.; Bell, J.E.; Brack-Werner, R. Cells of the central nervous system as targets and reservoirs of the human immunodeficiency virus. *Virus Res.* 2005, 111, 194–213. [CrossRef] [PubMed]

36. Zheng, J.; Thylin, M.R.; Ghopade, A.; Xiong, H.; Persidsky, Y.; Cotter, R.; Niemann, D.; Che, M.; Zeng, Y.C.; Gelbard, H.A.; et al. Intracellular CXCR4 signaling, neuronal apoptosis and neuropathogenic mechanisms of HIV-1-associated dementia. *J. Neuroimmunol.* 1999, 98, 185–200. [CrossRef]

37. Kruman, I.I.; Nath, A.; Mattson, M.P. HIV-1 protein tat induces apoptosis of hippocampal neurons by a mechanism involving caspase activation, calcium overload, and oxidative stress. *Exp. Neurol.* 1998, 154, 276–288. [CrossRef] [PubMed]

38. Haughey, N.J.; Nath, A.; Mattson, M.P.; Slevin, J.T.; Geiger, J.D. HIV-1 tat through phosphorylation of nmda receptors potentiates glutamate excitotoxicity. *J. Neurochem.* 2001, 78, 457–467. [CrossRef] [PubMed]

39. King, J.E.; Eugenin, E.A.; Buckner, C.M.; Berman, J.W. HIV tat and neurotoxicity. *Microbes Infect.* 2006, 8, 1347–1357. [CrossRef] [PubMed]

40. Conant, K.; Garzino-Demo, A.; Nath, A.; McArthur, J.C.; Halliday, W.; Power, C.; Gallo, R.C.; Major, E.O. Induction of monocyte chemoattractant protein-1 in HIV-1 tat-stimulated astrocytes and elevation in AIDS dementia. *Proc. Natl. Acad. Sci. USA* 1998, 95, 3117–3121. [CrossRef] [PubMed]

41. Hong, S.; Banks, W.A. Role of the immune system in HIV-associated neuroinflammation and neurocognitive implications. *Brain Behav. Immun.* 2015, 45, 1–12. [CrossRef] [PubMed]

42. Shin, A.H.; Kim, H.J.; Thayer, S.A. Subtype selective nmda receptor antagonists induce recovery of synapses lost following exposure to HIV-1 tat. *Br. J. Pharmacol.* 2012, 166, 1002–1017. [CrossRef] [PubMed]

43. Li, W.; Huang, Y.; Reid, R.; Steiner, J.; Malpica-Llanos, T.; Darden, T.A.; Shankar, S.K.; Mahadevan, A.; Satchishandra, P.; Nath, A. Nmda receptor activation by HIV-tat protein is clade dependent. *J. Neurosci.* 2008, 28, 12190–12198. [CrossRef] [PubMed]

44. Rao, V.R.; Sas, A.R.; Eugenin, E.A.; Siddappa, N.B.; Bimonte-Nelson, H.; Berman, J.W.; Ranga, U.; Tyor, W.R.; Prasad, V.R. HIV-1 clade-specific differences in the induction of neuropathogenesis. *J. Neurosci.* 2008, 28, 10010–10016. [CrossRef] [PubMed]

45. Rao, V.R.; Ruiz, A.P.; Prasad, V.R. Viral and cellular factors underlying neuropathogenesis in HIV-associated neurocognitive disorders (hand). *AIDS Res. Ther.* 2014, 11, 13. [CrossRef] [PubMed]

46. Mishra, M.; Vetrivel, S.; Siddappa, N.B.; Ranga, U.; Seth, P. Clade-specific differences in neurotoxicity of human immunodeficiency virus-1 b and c tat of human neurons: Significance of dicysteine C30C31 motif. *Ann. Neurol.* 2008, 63, 366–376. [CrossRef] [PubMed]

47. Kumar, S.; Maurya, V.K.; Danu, H.R.; Bhatt, M.L.; Saxena, S.K. Global perspective of novel therapeutic strategies for the management of neuroAIDS. *Biomol. Concepts* 2018, 9, 33–42. [CrossRef] [PubMed]

48. Zhou, L.; Sakshena, N.K. HIV associated neurocognitive disorders. *Infect. Dis. Rep.* 2013, 5, e8. [CrossRef] [PubMed]

49. Brabers, N.A.; Nottet, H.S. Role of the pro-inflammatory cytokines TNF-alpha and il-1beta in HIV-associated dementia. *Eur. J. Clin. Invest.* 2006, 36, 447–458. [CrossRef] [PubMed]

50. Kaul, M.; Lipton, S.A. Experimental and potential future therapeutic approaches for HIV-1 associated dementia targeting receptors for chemokines, glutamate and erythropoietin. *Neurotox. Res.* 2005, 8, 167–186. [CrossRef] [PubMed]

51. Letendre, S.L.; Zheng, J.C.; Kaul, M.; Yiannoutsos, C.T.; Ellis, R.J.; Taylor, M.J.; Marquie-Beck, J.; Navia, B. Chemokines in cerebrospinal fluid correlate with cerebral metabolite patterns in HIV-infected individuals. *J. Neurovirol.* 2011, 17, 63–69. [CrossRef] [PubMed]

52. Cho, Y.E.; Lee, M.H.; Song, B.J. Neuronal cell death and degeneration through increased nitroxidative stress and tau phosphorylation in HIV-1 transgenic rats. *PLoS ONE* 2017, 12, e0169945. [CrossRef] [PubMed]

53. Rao, V.R.; Neogi, U.; Talboom, J.S.; Padilla, L.; Rahman, M.; Fritz-French, C.; Gonzalez-Ramirez, S.; Verma, A.; Wood, C.; Ruprecht, R.M.; et al. Clade C HIV-1 isolates circulating in southern Africa exhibit a greater frequency of dicysteine motif-containing tat variants than those in southeast asia and cause increased neurovirulence. *Retrovirology* 2013, 10, 61. [CrossRef] [PubMed]
54. Gandhi, N.; Saiyed, Z.; Thangavel, S.; Rodriguez, J.; Rao, K.V.; Nair, M.P. Differential effects of HIV type 1 clade b and clade c tat protein on expression of proinflammatory and antiinflammatory cytokines by primary monocytes. *AIDS Res. Hum. Retrovir.* 2009, 25, 691–699. [CrossRef] [PubMed]

55. Jia, P.; Zhao, Z.; Hulgan, T.; Bush, W.S.; Samuels, D.C.; Blos, C.S.; Heaton, R.K.; Ellis, R.J.; Schork, N.; Marra, C.M.; et al. Genome-wide association study of HIV-associated neurocognitive disorder (hand): A charter group study. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 2017, 174, 413–426. [CrossRef] [PubMed]

56. Levine, A.J.; Service, S.; Miller, E.N.; Reynolds, S.M.; Singer, E.J.; Shapshak, P.; Martin, E.M.; Sacktor, N.; Becker, J.T.; Jacobson, L.P.; et al. Genome-wide association study of neurocognitive impairment and dementia in HIV-infected adults. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* 2012, 159B, 669–683. [CrossRef] [PubMed]

57. Kallianpur, A.R.; Levine, A.J. Host genetic factors predisposing to HIV-associated neurocognitive disorder. *Curr. HIV/AIDS Rep.* 2014, 11, 336–352. [CrossRef] [PubMed]

58. Levine, A.J.; Panos, S.E.; Horvath, S. Genetic, transcriptomic, and epigenetic studies of HIV-associated neurocognitive disorder. *J. Acquir. Immune Defic. Syndr.* 2014, 65, 481–503. [CrossRef] [PubMed]

59. Malhotra, A.K.; Kestler, L.J.; Mazzanti, C.; Bates, J.A.; Goldberg, T.; Goldman, D. A functional polymorphism in the comt gene and performance on a test of prefrontal cognition. *Am. J. Psychiatry* 2002, 159, 652–654. [CrossRef] [PubMed]

60. Levine, A.J.; Reynolds, S.; Cox, C.; Miller, E.N.; Sinzheimer, J.S.; Becker, J.T.; Martin, E.; Sacktor, N. Neuropsychology Working Group of the Multicenter, A.C.S. The longitudinal and interactive effects of HIV status, stimulant use, and host genotype upon neurocognitive functioning. *J. Neurovirol.* 2014, 20, 243–257. [CrossRef] [PubMed]

61. Villalba, K.; Devieux, J.G.; Rosenberg, R.; Cadet, J.L. Serotonin-related gene polymorphisms and asymptomatic neurocognitive impairment in HIV-infected alcohol abusers. *Genet. Res. Int.* 2016, 2016, 7169172. [CrossRef] [PubMed]

62. Horn, A.; Scheller, C.; du Plessis, S.; Burger, R.; Arendt, G.; Husstedt, I.W.; et al. The dopamine-related polymorphisms BDNF, COMT, DRD2, DRD3, and DRD4 are not linked with changes in CSF dopamine levels and frequency of HIV infection. *J. Neural. Transm. (Vienna)* 2017, 124, 501–509. [CrossRef] [PubMed]

63. Enge, S.; Fleischhauer, M.; Lesch, K.P.; Reif, A.; Strobel, A. Serotonergic modulation in executive functioning: Linking genetic variations to working memory performance. *Neuropsychologia* 2011, 49, 3776–3785. [CrossRef] [PubMed]

64. Villalba, K.; Devieux, J.G.; Rosenberg, R.; Cadet, J.L. DRD2 and DRD4 genes related to cognitive deficits in HIV-infected adults who abuse alcohol. *Behav. Brain Funct.* 2015, 11, 25. [CrossRef] [PubMed]

65. Hauser, K.F.; Knapp, P.E. Interactions of HIV and drugs of abuse: The importance of glia, neural progenitors, and substance-independent populations. *J. Neurovirol.* 2013, 19, 231–313. [PubMed]

66. Husstedt, I.W.; et al. The dopamine-related polymorphisms BDNF, COMT, DRD2, DRD3, and DRD4 are not linked with changes in CSF dopamine levels and frequency of HIV infection. *J. Neural. Transm. (Vienna)* 2017, 124, 501–509. [CrossRef] [PubMed]

67. Gupta, S.; Bousman, C.A.; Chana, G.; Cherner, M.; Heaton, R.K.; Deutsch, R.; Ellis, R.J.; Grant, I.; Everall, I.P. Impact of comt val158met on executive functioning in the context of HIV and methamphetamine. *Neurobehav. HIV Med.* 2015, 6, e0117450. [CrossRef] [PubMed]

68. Jacobs, M.M.; Murray, J.; Byrd, D.A.; Hurt, Y.L.; Morgello, S. HIV-related cognitive impairment shows bi-directional association with dopamine receptor DRD1 and DRD2 polymorphisms in substance-dependent and substance-independent populations. *J. Neurovirol.* 2013, 19, 495–504. [CrossRef] [PubMed]

69. Villalba, K.; Devieux, J.G.; Rosenberg, R.; Cadet, J.L. DRD2 and DRD4 genes related to cognitive deficits in HIV-infected alcohol abusers. *Behav. Brain Funct.* 2015, 11, 25. [CrossRef] [PubMed]

70. Var, S.R.; Day, T.R.; Vitomirov, A.; Smith, D.M.; Soontornniyomkij, V.; Moore, D.J.; Achim, C.L.; Mehta, S.R.; Perez-Santiago, J. Mitochondrial injury and cognitive function in HIV infection and methamphetamine use. *AIDS* 2016, 30, 839–848. [CrossRef] [PubMed]

71. RoZZI, S.J.; Avdoshina, V.; Fields, J.A.; Trejo, M.; Ton, H.T.; Ahern, G.P.; Mocchetti, I. Human immunodeficiency virus promotes mitochondrial toxicity. *Neurotox. Res.* 2017, 32, 723–733. [CrossRef] [PubMed]
72. Hart, A.B.; Samuels, D.C.; Hulgan, T. The other genome: A systematic review of studies of mitochondrial DNA haplogroups and outcomes of HIV infection and antiretroviral therapy. *AIDS Rev.* 2013, 15, 213–220. [PubMed]

73. Louboutin, J.P.; Strayer, D. Role of oxidative stress in HIV-1-associated neurocognitive disorder and protection by gene delivery of antioxidant enzymes. *Antioxidants (Basel)* 2014, 3, 770–797. [CrossRef] [PubMed]

74. Zhang, Y.; Wang, M.; Li, H.; Zhang, H.; Shi, Y.; Wei, F.; Liu, D.; Liu, K.; Chen, D. Accumulation of nuclear and mitochondrial DNA damage in the frontal cortex cells of patients with HIV-associated neurocognitive disorders. *Brain Res.* 2012, 1458, 11–1. [CrossRef] [PubMed]

75. Perez-Santiago, J.; Schrier, R.D.; de Oliveira, M.F.; G Ianella, S.; Var, S.R.; Day, T.R.; Ramirez-Gaona, M.; Suben, J.D.; Murrell, B.; Massanella, M.; et al. Cell-free mitochondrial DNA in CSF is associated with early viral rebound, inflammation, and severity of neurocognitive deficits in HIV infection. *J. Neurovirol.* 2016, 22, 191–200. [CrossRef] [PubMed]

76. Hulgan, T.; Samuels, D.C.; Bush, W.; Ellis, R.J.; Letendre, S.L.; Heaton, R.K.; Franklin, D.R.; Straub, P.; Murdoch, D.G.; Clifford, D.B.; et al. Mitochondrial DNA haplogroups and neurocognitive impairment during HIV infection. *Clin. Infect. Dis.* 2015, 61, 1476–1484. [CrossRef] [PubMed]

77. Malan-Muller, S.; Hemmings, S.M.; Spies, G.; Kidd, M.; Fennema-Notestine, C.; Seedat, S. Shorter telomere length—A potential susceptibility factor for HIV-associated neurocognitive impairments in south African women [corrected]. *PLoS ONE* 2013, 8, e88351. [CrossRef] [PubMed]

78. Appay, V.; Sauce, D. Assessing immune aging in HIV-infected patients. *Virulence* 2017, 8, 529–538. [CrossRef] [PubMed]

79. Giesbrecht, C.J.; Thornton, A.E.; Hall-Patch, C.; Maan, E.J.; Cote, H.C.; Money, D.M.; Murray, M.; Pick, N. Select neurocognitive impairment in HIV-infected women: Associations with HIV viral load, hepatitis C virus, and depression, but not leukocyte telomere length. *PLoS ONE* 2014, 9, e89556. [CrossRef] [PubMed]

80. Levine, A.J.; Quach, A.; Moore, D.J.; Achim, C.L.; Soontornniyomkij, V.; Masliah, E.; Singer, E.J.; Gelman, B.; Nemanin, N.; Horvath, S. Accelerated epigenetic aging in brain is associated with pre-mortem HIV-associated neurocognitive disorders. *J. Neurovirol.* 2016, 22, 366–375. [CrossRef] [PubMed]

81. Saiyed, Z.M.; Gandhi, N.; Agudelo, M.; Napuri, J.; Samikkannu, T.; Reddy, P.V.; Khatavkar, P.; Yndart, A.; Saxena, S.K.; Nair, M.P. HIV-1 tat upregulates expression of histone deacetylase-2 (HDAC2) in human neurons: Implication for HIV-associated neurocognitive disorder (hand). *Neurochem. Int.* 2011, 58, 656–664. [CrossRef] [PubMed]

82. Thames, A.D.; Briones, M.S.; Magpantay, L.I.; Martinez-Maza, O.; Singer, E.J.; Hinkin, C.H.; Morgello, S.; Gelman, B.B.; Moore, D.J.; Heizerling, K.; et al. The role of chemokine C–C motif ligand 2 genotype and cerebrospinal fluid chemokine c-c motif ligand 2 in neurocognition among HIV-infected patients. *AIDS* 2015, 29, 1483–1491. [CrossRef] [PubMed]

83. Eugenin, E.A.; Osiecki, K.; Lopez, L.; Goldstein, H.; Calderon, T.M.; Berman, J.W. CCL2/monocyte chemoattractant protein-1 mediates enhanced transmigration of human immunodeficiency virus (HIV)-infected leukocytes across the blood-brain barrier: A potential mechanism of HIV-CNS invasion and neuroAIDS. *J. Neurosci.* 2006, 26, 1098–1106. [CrossRef] [PubMed]

84. Gonzalez, E.; Rovin, B.H.; Sen, L.; Cooke, G.; Dhanda, R.; Mummidii, S.; Kulkarni, H.; Bamshad, M.J.; Telles, V.; Anderson, S.A.; et al. HIV-1 infection and AIDS dementia are influenced by a mutant mcp-1 allele linked to increased monocyte infiltration of tissues and mcpr-1 levels. *Proc. Natl. Acad. Sci. USA* 2002, 99, 13795–13800. [CrossRef] [PubMed]

85. Singh, K.K.; Barroga, C.F.; Hughes, M.D.; Chen, J.; Raskino, C.; McKinney, R.E.; Spector, S.A. Genetic influence of CCR5, CCR2, and SDF1 variants on human immunodeficiency virus 1 (HIV-1)-related disease progression and neurocognitive impairment, in children with symptomatic HIV-1 infection. *J. Infect. Dis.* 2003, 188, 1461–1472. [CrossRef] [PubMed]

86. Letendre, S.; Marquie-Beck, J.; Singh, K.K.; de Almeida, S.; Zimmerman, J.; Spector, S.A.; Grant, I.; Ellis, R.; Group, H. The monocyte chemotactic protein-1–2578 g allele is associated with elevated mcpr-1 concentrations in cerebrospinal fluid. *J. Neuroimmunol.* 2004, 157, 193–196. [CrossRef] [PubMed]

87. Singh, K.K.; Ellis, R.J.; Marquie-Beck, J.; Letendre, S.; Heaton, R.K.; Grant, I.; Spector, S.A. CCR2 polymorphisms affect neuropsychological impairment in HIV-1-infected adults. *J. Neuroimmunol.* 2004, 157, 185–192. [CrossRef] [PubMed]
88. Levine, A.J.; Singer, E.J.; Sinheimer, J.S.; Hinkin, C.H.; Papp, J.; Dandekar, S.; Giovannelli, A.; Shapshak, P. Ccl3 genotype and current depression increase risk of HIV-associated dementia. *Neurobehav. HIV Med.* 2009, 1, 1–7. [CrossRef] [PubMed]

89. Spector, S.A.; Singh, K.K.; Gupta, S.; Cystique, L.A.; Jin, H.; Letendre, S.; Schrier, R.; Wu, Z.; Hong, K.X.; Yu, X.; et al. Apo e epsilon4 and MBL-2 O/O genotypes are associated with neurocognitive impairment in HIV-infected plasma donors. *AIDS* 2010, 24, 1471–1479. [CrossRef] [PubMed]

90. Bol, S.M.; Booman, T.; van Manen, D.; Bunnik, E.M.; van Sighem, A.I.; Sieberer, M.; Boeser-Nunnink, B.; de Wolf, F.; Schuitemaker, H.; Portegies, P.; et al. Single nucleotide polymorphism in gene encoding transcription factor prep1 is associated with HIV-1-associated dementia. *PLoS ONE* 2012, 7, e30990. [CrossRef] [PubMed]

91. Brown, A.; Sacktor, N.; Marder, K.; Cohen, B.; Schifitto, G.; Skolasky, R.L.; Creighton, J.; Guo, L.; McArthur, J.C. CCL3L1 gene copy number in individuals with and without HIV-associated neurocognitive disorder. *Curr. Biomark. Find.* 2012, 2012, 1–6. [CrossRef] [PubMed]

92. van Rij, R.P.; Portegies, P.; Hallaby, T.; Lange, J.M.; Visser, J.; de Roda Husman, A.M.; van’t Wout, A.B.; Schuitemaker, H. Reduced prevalence of the CCR5 delta32 heterozygous genotype in human immunodeficiency virus-infected individuals with AIDS dementia complex. *J. Infect. Dis.* 1999, 180, 854–857. [CrossRef] [PubMed]

93. Gonzalez, E.; Kulkarni, H.; Bolivar, H.; Mangano, A.; Sanchez, R.; Catano, G.; Nibbs, R.J.; Freedman, B.L.; Quinones, M.P.; Bamshad, M.J.; et al. The influence of CCL3L1 gene-containing segmental duplications on HIV-1/AIDS susceptibility. *Science* 2005, 307, 1434–1440. [CrossRef] [PubMed]

94. Liu, H.; Chao, D.; Nakayama, E.E.; Taguchi, H.; Goto, M.; Xin, X.; Takamatsu, J.K.; Saito, H.; Ishikawa, Y.; Akaza, T.; et al. Polymorphism in rantes chemokine promoter affects HIV-1 disease progression. *Proc. Natl. Acad. Sci. USA* 1999, 96, 4581–4585. [CrossRef] [PubMed]

95. McDermott, D.H.; Beecroft, M.J.; Kleeberger, C.A.; Al-Sharif, F.M.; Ollier, W.E.; Zimmerman, P.A.; Boatin, B.A.; Leitman, S.F.; Detels, R.; Hajeer, A.H.; et al. Chemokine rantes promoter polymorphism affects risk of both HIV infection and disease progression in the multicenter AIDS cohort study. *AIDS* 2000, 14, 2671–2678. [CrossRef] [PubMed]

96. McLaren, P.J.; Coulonges, C.; Bartha, I.; Lenz, T.L.; Deutsch, A.J.; Bashirova, A.; Buchbinder, S.; Carrington, M.N.; Cossarizza, A.; Dalmau, J.; et al. Polymorphisms of large effect explain the majority of the host genetic contribution to variation of HIV-1 virus load. *Proc. Natl. Acad. Sci. USA* 2015, 112, 14658–14663. [CrossRef] [PubMed]

97. Samson, M.; Libert, F.; Doranz, B.J.; Rucker, J.; Liesnard, C.; Farber, C.M.; Saragosti, S.; Lapoumeroulie, C.; Cognaux, J.; Forceille, C.; et al. Resistance to HIV-1 infection in caucasian individuals bearing mutant alleles of the ccr-5 chemokine receptor gene. *Nature* 1996, 382, 722–725. [CrossRef] [PubMed]

98. Duarte, R.F.; Salgado, M.; Sanchez-Ortega, I.; Arnan, M.; Canals, C.; Domingo-Domenech, E.; Fernandez-de-Sevilla, A.; Gonzalez-Barca, E.; Moron-Lopez, S.; Nogues, N.; et al. CCR5 delta32 homozygous cord blood allogeneic transplantation in a patient with HIV: A case report. *Lancet HIV* 2015, 2, e236–e242. [CrossRef] [PubMed]

99. Hutter, G.; Nowak, D.; Mose, M.; Ganepola, S.; Mussig, A.; Allers, K.; Schneider, T.; Hofmann, J.; Kuchler, C.; Blau, O.; et al. Long-term control of HIV by CCR5 delta32/delta32 stem-cell transplantation. *N. Engl. J. Med.* 2009, 360, 692–698. [CrossRef] [PubMed]

100. Boven, L.A.; van der Bruggen, T.; van Asbeck, B.S.; Marx, J.J.; Nottet, H.S. Potential role of CCR5 polymorphism in the development of AIDS dementia complex. *FEBS Immunol. Med. Microbiol.* 1999, 26, 243–247. [CrossRef] [PubMed]

101. Ndhlovu, L.C.; Umaki, T.; Chew, G.M.; Chow, D.C.; Agsalda, M.; Kallianpur, K.J.; Paul, R.; Zhang, G.; Ho, E.; Hanks, N.; et al. Treatment intensification with maraviroc (CCR5 antagonist) leads to declines in CD16-expressing monocytes in cart-suppressed chronic HIV-infected subjects and is associated with improvements in neurocognitive test performance: Implications for HIV-associated neurocognitive disease (hand). *J. Neurovirol.* 2014, 20, 571–582. [PubMed]

102. Quasney, M.W.; Zhang, Q.; Sargent, S.; Mynatt, M.; Glass, J.; McArthur, J. Increased frequency of the tumor necrosis factor-alpha-308 allele in adults with human immunodeficiency virus dementia. *Ann. Neurol.* 2001, 50, 157–162. [CrossRef] [PubMed]
103. Pemberton, L.A.; Stone, E.; Price, P.; van Bockxmeer, F.; Brew, B.J. The relationship between apoe, tnfα, il1α, il1β and il12β genes and HIV-1-associated dementia. HIV Med. 2008, 9, 677–680. [CrossRef] [PubMed]

104. Sato-Matsumura, K.C.; Berger, J.; Hainfellner, J.A.; Mazal, P.; Budka, H. Development of HIV encephalitis in AIDS and tnfα regulatory elements. J. Neuroimmunol. 1998, 91, 89–92. [CrossRef]

105. Diaz-Arrastia, R.; Gong, Y.; Kelly, C.J.; Gelman, B.B. Host genetic polymorphisms in human immunodeficiency virus-related neurologic disease. J. Neurovirol. 2004, 10 (Suppl. 1), 67–73. [CrossRef]

106. Sanna, P.P.; Repunte-Canonigo, V.; Masliah, E.; Lefebvre, C. Gene expression patterns associated with neurological disease in human HIV infection. PLoS ONE 2017, 12, e0175316. [CrossRef] [PubMed]

107. Borjabad, A.; Brooks, A.I.; Volsky, D.J. Gene expression profiles of HIV-1-infected glia and brain: Toward better understanding of the role of astrocytes in HIV-1-associated neurocognitive disorders. J. Neuroimmun Pharmacol. 2010, 5, 44–62. [CrossRef] [PubMed]

108. Siangphoe, U.; Archer, K.J. Gene expression in HIV-associated neurocognitive disorders: A meta-analysis. J. Acquir. Immune Defic. Syndr. 2015, 70, 479–488. [CrossRef] [PubMed]

109. Do, H.; Vasilescu, A.; Diop, G.; Hirtzig, T.; Coulonges, C.; Labib, T.; Heath, S.C.; Spadoni, J.L.; Therwath, A.; Tresoldi, E.; Romiti, M.L.; Boniotto, M.; Crovella, S.; Salvatori, F.; Palomba, E.; Pastore, A.; Cancrini, C.; Ioannidis, J.P.; Rosenberg, P.S.; Goedert, J.J.; Ashton, L.J.; Benfield, T.L.; Buchbinder, S.P.; Coutinho, R.A.; Eugen-Olsen, J.; Gallart, T.; Katzenstein, T.L.; et al. Genetic restriction of AIDS pathogenesis by an SDF-1 chemokine gene variant. Alive study, hemophilia growth and development study (HGDS), multicenter AIDS cohort study (Macs), multicenter hemophilia cohort study (MHCS), San Francisco city cohort (SFCC). Science 1998, 279, 389–393. [CrossRef] [PubMed]

110. Meyer, L.; Magierowska, M.; Hubert, J.B.; Theodorou, I.; van Rij, R.; Prins, M.; de Roda Husman, A.M.; Coutinho, R.; Schuitemaker, H. CC-chemokine receptor variants, SDF-1 polymorphism, and disease progression in 720 HIV-infected patients. Seroco cohort. Amsterdam cohort studies on AIDS. J. Acquir. Immune Defic. Syndr. 1999, 13, 624–626. [CrossRef]

111. Pemberton, L.A.; Stone, E.; Price, P.; van Bockxmeer, F.; Brew, B.J. The relationship between apoe, tnfα, il1α, il1β and il12β genes and HIV-1-associated dementia. HIV Med. 2008, 9, 677–680. [CrossRef] [PubMed]

112. Agace, W.W.; Amara, A.; Roberts, A.I.; Pablos, J.L.; Thelen, S.; Ugucioni, M.; Li, X.Y.; Marsal, J.; Arenzana-Seisdedos, F.; Delaunay, T.; et al. Constitutive expression of stromal derived factor-1 by mucosal epithelia and its role in HIV transmission and propagation. Curr. Biol. 2000, 10, 325–328. [CrossRef]

113. Quach, A.; Horvath, S.; Nemanim, N.; Vatakis, D.; Witt, M.D.; Miller, E.N.; Detels, R.; Langfelder, P.; Shapshak, P.; Singer, E.J.; et al. No reliable gene expression biomarkers of current or impending neurocognitive impairment in peripheral blood monocytes of persons living with HIV. J. Neurovirol. 2018, 24, 350–361. [CrossRef] [PubMed]

114. Winkler, C.; Modi, W.; Price, P.; van Bockxmeer, F.; Brew, B.J. The relationship between apoe, tnfα, il1α, il1β and il12β genes and HIV-1-associated dementia. HIV Med. 2008, 9, 677–680. [CrossRef] [PubMed]

115. Ioannidis, J.P.; Rosenberg, P.S.; Goedert, J.J.; Ashton, L.J.; Benfield, T.L.; Buchbinder, S.P.; Coutinho, R.A.; Eugen-Olsen, J.; Gallart, T.; Katzenstein, T.L.; et al. Effects of CCR5-delta32, CCR2-64I, and SDF-1 3′a alleles on HIV-1 disease progression: An international meta-analysis of individual-patient data. Ann. Intern. Med. 2001, 135, 782–795. [CrossRef] [PubMed]

116. Shrestha, S.; Strathdee, S.A.; Galai, N.; Oleskyt, T.; Fallin, M.D.; Mehta, S.; Schaid, D.; Vlahov, D.; O’Brien, S.J.; Smith, M.W. Behavioral risk exposure and host genetics of susceptibility to HIV-1 infection. J. Infect. Dis. 2002, 185, 696–700. [CrossRef] [PubMed]

117. Levine, A.J.; Sinsheimer, J.S.; Bilder, R.; Shapshak, P.; Singer, E.J. Functional polymorphisms in dopamine-related genes: Effect on neurocognitive functioning in HIV+ adults. J. Clin. Exp. Neuropsychol. 2012, 34, 78–91. [CrossRef] [PubMed]

118. Shrestha, S.; Strathdee, S.A.; Galai, N.; Oleskyt, T.; Fallin, M.D.; Mehta, S.; Schaid, D.; Vlahov, D.; O’Brien, S.J.; Smith, M.W. Behavioral risk exposure and host genetics of susceptibility to HIV-1 infection. J. Infect. Dis. 2006, 193, 16–26. [CrossRef] [PubMed]

119. Do, H.; Vasilescu, A.; Diop, G.; Hirtzig, T.; Coulonges, C.; Labib, T.; Heath, S.C.; Spadoni, J.L.; Therwath, A.; Lathrop, M.; et al. Associations of the IL2Ra, IL4Ra, IL10Ra, and IFN (gamma) r1 cytokine receptor genes with AIDS progression in a french AIDS cohort. Immunogenetics 2006, 58, 89–98. [CrossRef] [PubMed]

120. Price, P.; James, I.; Fernandez, S.; French, M.A. Alleles of the gene encoding IL-1α may predict control of plasma viraemia in HIV-1 patients on highly active antiretroviral therapy. AIDS 2004, 18, 1495–1501. [CrossRef] [PubMed]

121. Garred, P.; Madsen, H.O.; Balslev, U.; Hofmann, B.; Pedersen, C.; Gerstoft, J.; Svejgaard, A. Susceptibility to HIV infection and progression of AIDS in relation to variant alleles of mannose-binding lectin. Lancet (Lond. Engl.) 1997, 349, 236–240. [CrossRef]
120. Singh, K.K.; Lieser, A.; Ruan, P.K.; Fenton, T.; Spector, S.A. An age-dependent association of mannose-binding
lectin-2 genetic variants on HIV-1-related disease in children. *J. Allergy Clin. Immunol.* 2008, 122, 173–180. [CrossRef] [PubMed]

121. Singh, K.K.; Nathamu, S.; Adame, A.; Alire, T.U.; Dumaop, W.; Gouaux, B.; Moore, D.J.; Masliah, E. Expression of mannose binding lectin in HIV-1-infected brain: Implications for HIV-related neuronal damage and neuroAIDS. *Neurobiol. HIV Med.* 2011, 3, 41–52. [CrossRef] [PubMed]

122. Naranbhai, V.; Carrington, M. Host genetic variation and HIV disease: From mapping to mechanism. *Immunogenetics* 2017, 69, 498–498. [CrossRef] [PubMed]

123. Schrier, R.D.; Gupta, S.; Riggs, P.; Cysique, L.A.; Letendre, S.; Jin, H.; Spector, S.A.; Singh, K.K.; Wolfson, T.; Wu, Z.; et al. The influence of HLA on HIV-associated neurocognitive impairment in Anhui, China. *PLoS ONE* 2012, 7, e32303. [CrossRef] [PubMed]

124. Singh, K.K.; Gray, P.K.; Wang, Y.; Fenton, T.; Trout, R.N.; Spector, S.A. Hla alleles are associated with altered
risk for disease progression and central nervous system impairment of HIV-infected children. *J. Acquir. Immune Defic. Syndr.* 2011, 57, 32–39. [CrossRef] [PubMed]

125. Sagar, V.; Pilakka-Kanthikeel, S.; Martinez, P.C.; Atluri, V.S.R.; Nair, M. Common gene-network signature of
different neurological disorders and their potential implications to neuroAIDS. *PLoS ONE* 2017, 12, e0181642. [CrossRef] [PubMed]

126. Xing, Y.; Shepherd, N.; Lan, J.; Li, W.; Rane, S.; Gupta, S.K.; Zhang, S.; Dong, J.; Yu, Q. MMP9/timp
imbalances in the peripheral blood and cerebrospinal fluid are associated with the pathogenesis of
HIV-1-associated neurocognitive disorders. *Brain Behav. Immun.* 2017, 65, 161–172. [CrossRef] [PubMed]

127. Singh, H.; Samani, D.; Nambiar, N.; Ghate, M.V.; Gangakhedkar, R.R. Effect of matrix metalloproteinase-21
(572C/T) polymorphism on HIV-associated neurocognitive disorder. *APMIS* 2018, 126, 329–336. [CrossRef] [PubMed]

128. Singh, H.O.; Marathe, S.D.; Nain, S.; Samani, D.; Nema, V.; Ghate, M.V.; Gangakhedkar, R.R. Promoter
polymorphism MMP-1 (-1607 2G/1G) and MMP-3 (-1612 5A/6A) in development of hand and modulation
of pathogenesis of hand. *J. Biosci.* 2017, 42, 481–490. [CrossRef] [PubMed]

129. Singh, H.; Marathe, S.D.; Nain, S.; Nana, V.; Ghate, M.V.; Gangakhedkar, R.R. Genetic variation of MMP-2(-735 c>t)
and MMP-9(-1562 c>t) gene in risk of development of hand and severity of hand. *J. Gene Med.* 2016, 18, 250–257. [CrossRef] [PubMed]

130. Singh, H.; Marathe, S.D.; Nain, S.; Nema, V.; Angadi, M.; Bapat, S.S.; Ghate, M.V.; Gangakhedkar, R.R. Impact
of variants of mmp-7(-181a>g) gene in susceptibility to the development of HIV-associated neurocognitive
disorder and its severity. *APMIS* 2016, 124, 966–972. [CrossRef] [PubMed]

131. Singh, H.; Samani, D.; Nambiar, N.; Ghate, M.V.; Gangakhedkar, R.R. Prevalence of mmp-8 gene
polymorphisms in HIV-infected individuals and its association with HIV-associated neurocognitive disorder. *Gene* 2018, 646, 83–90. [CrossRef] [PubMed]

132. De Jager, C.A.; Msemburi, W.; Pepper, K.; Combrinck, M.I. Dementia prevalence in a rural region of south
Africa: A cross-sectional community study. *J. Alzheimer's Dis. JAD* 2017, 60, 1087–1096. [CrossRef] [PubMed]

133. Zott, B.; Busche, M.A.; Sperling, R.A.; Konnerth, A. What happens with the circuit in Alzheimer’s disease in
mice and humans? *Annu. Rev. Neurosci.* 2018, 41, 277–297. [CrossRef] [PubMed]

134. Serrano-Pozo, A.; Frosch, M.P.; Masliah, E.; Hyman, B.T. Neuropathological alterations in Alzheimer disease.
*Cold Spring Harb. Perspect. Med.* 2011, 1, a006189. [CrossRef] [PubMed]

135. Mucke, L.; Selkoe, D.J. Neurotoxicity of amyloid beta-protein: Synaptic and network dysfunction. *Cold Spring
Harb. Perspect. Med.* 2012, 2, a006338. [CrossRef] [PubMed]

136. Pozueta, J.; Lefort, R.; Shelanski, M.L. Synaptic changes in Alzheimer’s disease and its models. *Neuroscience* 2013, 251, 51–65. [CrossRef] [PubMed]

137. Cacabelos, R. Pharmacogenomics of Alzheimer’s and parkinson’s diseases. *Neurosci. Lett.* 2018. [CrossRef] [PubMed]

138. Weeber, E.J.; Beffert, U.; Jones, C.; Christian, J.M.; Forster, E.; Sweatt, J.D.; Herz, J. Reelin and apoE receptors
cooperate to enhance hippocampal synaptic plasticity and learning. *J. Biol. Chem.* 2002, 277, 39944–39952. [CrossRef] [PubMed]

139. Lane-Donovan, C.; Herz, J. Apoe, apo receptors, and the synapse in Alzheimer’s disease. *Trends Endocrinol. Metab.* 2017, 28, 273–284. [CrossRef] [PubMed]
140. Bu, G. Apolipoprotein e and its receptors in Alzheimer’s disease: Pathways, pathogenesis and therapy. Nat. Rev. Neurosci. 2009, 10, 333–344. [CrossRef] [PubMed]

141. Brecht, W.J.; Harris, F.M.; Chang, S.; Tesseur, I.; Yu, G.Q.; Xu, Q.; Mahley, R.W.; Dolan, M.J.; et al. Apolipoprotein (apo) E4 enhances HIV-1 cell entry in vitro, and the apoe epsilon4/epsilon4 genotype accelerates HIV disease progression. Proc. Natl. Acad. Sci. USA 2008, 105, 8718–8723. [CrossRef] [PubMed]

142. Corder, E.H.; Robertson, K.; Lannfelt, L.; Bogdanovic, N.; Eggertsen, G.; Wilkins, J.; Hall, C. HIV-infected subjects with the E4 allele for apoe have excess dementia and peripheral neuropathy. AIDS (Lond. Engl.) 2005, 19, 407–411. [CrossRef] [PubMed]

143. Huang, Y.A.; Zhou, B.; Wernig, M.; Sudhof, T.C. Apoe2, apoe3, and apoe4 differentially stimulate app transcription and abeta secretion. Cell 2017, 168, 427–441. [CrossRef] [PubMed]

144. Esiri, M.M.; Biddolph, S.C.; Morris, C.S. Prevalence of alzheimer plaques in AIDS. J. Neurovirol. 2018. [CrossRef] [PubMed]

145. Hearps, A.C.; Martin, G.E.; Angelovich, T.A.; Cheng, W.J.; Maisa, A.; Landay, A.L.; Jaworowski, A.; Crowe, S.M. Aging is associated with chronic innate immune activation and dysregulation of monocyte phenotype and function. Aging Cell 2012, 11, 867–875. [CrossRef] [PubMed]

146. Kang, Y.J.; Digicaylioglu, M.; Russo, R.; Saul, M.; Achim, C.L.; Fletcher, L.; Masliah, E.; Lipton, S.A. Erythropoietin plus insulin-like growth factor-i protects against neuronal damage in a murine model of tauopathy. Nature 2017, 549, 523–527. [CrossRef] [PubMed]

147. Valcour, V.; Shikuma, C.; Shiramizu, B.; Watters, M.; Poff, P.; Selnes, O.A.; Grove, J.; Liu, Y.; Abdul-Majid, K.B.; Gartner, S.; et al. Age, apolipoprotein E4, and the risk of HIV dementia: The Hawaii aging with HIV cohort. J. Neuroimmunol. 2004, 157, 197–202. [CrossRef] [PubMed]

148. Soontornniyomkij, V.; Moore, D.J.; Gouaux, B.; Soontornniyomkij, B.; Tatro, E.T.; Umlauf, A.; Masliah, E.; Fletcher, L.; et al. ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. Nature 2017, 549, 523–527. [CrossRef] [PubMed]

149. Hearps, A.C.; Martin, G.E.; Angelovich, T.A.; Cheng, W.J.; Maisa, A.; Landay, A.L.; Jaworowski, A.; Crowe, S.M. Aging is associated with chronic innate immune activation and dysregulation of monocyte phenotype and function. Aging Cell 2012, 11, 867–875. [CrossRef] [PubMed]

150. Bien-Ly, N.; Gillespie, A.K.; Walker, D.; Yoon, S.Y.; Huang, Y. Reducing human apolipoprotein e levels attenuates age-dependent abeta accumulation in mutant human amyloid precursor protein transgenic mice. J. Neurosci. 2012, 32, 4803–4811. [CrossRef] [PubMed]

151. Burt, T.D.; Agan, B.K.; Marconi, V.C.; He, W.; Kulkarni, M.; Huang, Y.; Mahley, R.W.; Dolan, M.J.; et al. Apolipoprotein E4 transgenic mice is associated with activation of extracellular signal-regulated kinase: Modulation by zinc. J. Biol. Chem. 2004, 279, 44795–44801. [CrossRef] [PubMed]

152. Brodaty, H.; Singer, E.J.; Vinters, H.V.; et al. Cerebral beta-amyloid deposition predicts HIV-associated neurocognitive disorders in apoe epsilon4 carriers. AIDS (Lond. Engl.) 2012, 26, 2327–2335. [CrossRef] [PubMed]

153. Green, D.A.; Masliah, E.; Vinters, H.V.; Beizai, P.; Moore, D.J.; Achim, C.L. Brain deposition of beta-amyloid is a common pathologic feature in HIV positive patients. AIDS (Lond. Engl.) 2005, 19, 407–411. [CrossRef] [PubMed]

154. Shi, Y.; Yamada, K.; Liddelow, S.A.; Smith, S.T.; Zhao, L.; Luo, W.; Tsai, R.M.; Spina, S.; Ginberg, L.T.; Rojas, J.C.; et al. ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. Nature 2017, 549, 523–527. [CrossRef] [PubMed]

155. Harris, F.M.; Brecht, W.J.; Xu, Q.; Mahley, R.W.; Huang, Y. Increased tau phosphorylation in apolipoprotein e4 transgenic mice is associated with activation of extracellular signal-regulated kinase: Modulation by zinc. J. Biol. Chem. 2004, 279, 44795–44801. [CrossRef] [PubMed]

156. Brecht, W.J.; Harris, F.M.; Chang, S.; Tesseur, I.; Yu, G.Q.; Xu, Q.; Dee Fish, J.; Wyss-Coray, T.; Buttini, M.; Mucke, L.; et al. Neuron-specific apolipoprotein E4 proteolysis is associated with increased tau phosphorylation in brains of transgenic mice. J. Neurosci. 2004, 24, 2527–2534. [CrossRef] [PubMed]

157. Kang, Y.J.; Digicaylioglu, M.; Russo, R.; Saul, M.; Achim, C.L.; Fletcher, L.; Masliah, E.; Lipton, S.A. Erythropoietin plus insulin-like growth factor-i protects against neuronal damage in a murine model of human immunodeficiency virus-associated neurocognitive disorders. Ann. Neurol. 2010, 68, 342–352. [CrossRef] [PubMed]

158. Brew, B.J.; Pemberton, L.; Blennow, K.; Wallin, A.; Hagberg, L. CSF amyloid beta42 and tau levels correlate with AIDS dementia complex. Neurology 2005, 65, 1490–1492. [CrossRef] [PubMed]
159. Anthony, I.C.; Ramage, S.N.; Carnie, F.W.; Simmonds, P.; Bell, J.E. Accelerated tau deposition in the brains of individuals infected with human immunodeficiency virus-1 before and after the advent of highly active anti-retroviral therapy. *Acta Neuropathol.* 2006, 111, 529–538. [CrossRef] [PubMed]

160. Soontornniyomkij, V.; Umlauf, A.; Soontornniyomkij, B.; Gouaux, B.; Ellis, R.J.; Levine, A.J.; Moore, D.J.; Letendre, S.L. Association of antiretroviral therapy with brain aging changes among HIV-infected adults. *AIDS (Lond. Engl.)* 2018, 32, 2005–2015. [CrossRef] [PubMed]

161. Kantarci, K.; Lowe, V.; Przybelski, S.A.; Weigand, S.D.; Senjem, M.L.; Ivnik, R.J.; Preboske, G.M.; Roberts, R.; Geda, Y.E.; Boeve, B.F.; et al. ApoE modifies the association between abeta load and cognition in cognitively normal older adults. *Neurology* 2012, 78, 232–240. [CrossRef] [PubMed]

162. Nir, T.M.; Jahanshad, N.; Busovaca, E.; Wendelken, L.; Nicolas, K.; Thompson, P.M.; Valcour, V.G. Mapping white matter integrity in elderly people with HIV. *Hum. Brain Mapp.* 2014, 35, 975–992. [CrossRef] [PubMed]

163. Piconi, S.; Trabattoni, D.; Gori, A.; Parisotto, S.; Magni, C.; Meraviglia, P.; Bandera, A.; Capetti, A.; French, M.A.; King, M.S.; Tschampa, J.M.; da Silva, B.A.; Landay, A.L. Serum immune activation markers of individuals infected with human immunodeficiency virus-1 before and after the advent of highly active anti-retroviral therapy. *Acta Neuropathol.* 2006, 111, 529–538. [CrossRef] [PubMed]

164. Castellano, J.M.; Kim, J.; Stewart, F.R.; Jiang, H.; DeMattos, R.B.; Patterson, B.W.; Fagan, A.M.; Morris, J.C.; Mawuenyega, K.G.; Cruchaga, C.; et al. Human apoE isoforms differentially regulate brain amyloid-beta peptide clearance. *Sci. Transl. Med.* 2011, 3, 89ra57. [CrossRef] [PubMed]

165. Bharat, I.; Weisgraber, K.H.; Huang, Y. Apolipoprotein E—A multifunctional protein with implications in various pathologies as a result of its structural features. *Comput. Struct. Biotechnol. J.* 2017, 15, 359–365. [CrossRef] [PubMed]

166. Mahley, R.W.; Weisgraber, K.H.; Huang, Y. Apolipoprotein E: Structure determines function, from atherosclerosis to Alzheimer’s disease to AIDS. *J. Lipid Res.* 2009, 50, S183–S188. [CrossRef] [PubMed]

167. Chang, L.; Mahley, R.W. Apolipoprotein e: Structure and function in lipid metabolism, neurobiology, and Alzheimer’s diseases. *Neurobiol. Dis.* 2014, 72 Pt A, 3–12. [CrossRef]

168. Martin, I.; Dubois, M.C.; Saermark, T.; Ruysschaert, J.M. Apolipoprotein a-1 interacts with the n-terminal fusogenic domains of HIV (human immunodeficiency virus) gp41: Implications in viral entry. *Biochem. Biophys. Res. Commun.* 1992, 186, 95–101. [CrossRef]

169. Briz, V.; Poveda, E.; Soriano, V. Human apoE isoforms differentially regulate brain amyloid-beta peptide clearance. *Sci. Transl. Med.* 2011, 3, 89ra57. [CrossRef] [PubMed]

170. Nir, T.M.; Jahanshad, N.; Busovaca, E.; Wendelken, L.; Nicolas, K.; Thompson, P.M.; Valcour, V.G. Mapping white matter integrity in elderly people with HIV. *Hum. Brain Mapp.* 2014, 35, 975–992. [CrossRef] [PubMed]

171. Nir, T.M.; Jahanshad, N.; Busovaca, E.; Wendelken, L.; Nicolas, K.; Thompson, P.M.; Valcour, V.G. Mapping white matter integrity in elderly people with HIV. *Hum. Brain Mapp.* 2014, 35, 975–992. [CrossRef] [PubMed]

172. Buckner, C.M.; Luers, A.J.; Calderon, T.M.; Eugenin, E.A.; Berman, J.W. Neuroimmunity and the blood-brain barrier: Molecular regulation of leukocyte transmigration and viral entry into the nervous system with a focus on neuroAIDS. *J. Neuroimmune Pharmacol.* 2006, 1, 160–181. [CrossRef] [PubMed]

173. Giunta, B.; Ehrhart, J.; Obregon, D.F.; Lam, L.; Le, L.; Jin, J.; Fernandez, F.; Tan, J.; Shytle, R.D. Antiretroviral medications disrupt microglial phagocytosis of beta-amyloid and increase its production by neurons: Implications for HIV-associated neurocognitive disorders. *Mol. Brain* 2011, 4, 23. [CrossRef] [PubMed]

174. Lan, X.; Kiyota, T.; Hanamsagar, R.; Huang, Y.; Andrews, S.; Peng, H.; Zheng, C.; Swindells, S.; Carlson, G.A.; Ikezu, T. The effect of HIV protease inhibitors on amyloid-beta peptide degradation and synthesis in human cells and Alzheimer’s disease animal model. *J. Neuroimmune Pharmacol.* 2012, 7, 412–423. [CrossRef] [PubMed]
177. Geffen, R.; Martinez, R.; de Las Pozas, A.; Issac, B.; McCarthy, M. Apolipoprotein e4 suppresses neuronal-specific gene expression in maturing neuronal progenitor cells exposed to HIV. *J. Neuroimmune Pharmacol.* 2017, 12, 462–483. [CrossRef] [PubMed]

178. Theendakara, V.; Peters-Libeu, C.A.; Spilman, P.; Poksay, K.S.; Bredesen, D.E.; Rao, R.V. Direct transcriptional effects of apolipoprotein E. *J. Neurosci.* 2016, 36, 685–700. [CrossRef] [PubMed]

179. Mu, Y.; Gage, F.H. Adult hippocampal neurogenesis and its role in Alzheimer’s disease. *Mol. Neurodegener.* 2011, 6, 85. [CrossRef] [PubMed]

180. Tensaouti, Y.; Stephanz, E.P.; Yu, T.S.; Kernie, S.G. ApoE regulates the development of adult newborn hippocampal neurons. *eNeuro* 2018, 5. [CrossRef] [PubMed]

181. Wang, R.; Reddy, P.H. Role of glutamate and NMDA receptors in Alzheimer’s disease. *J. Alzheimer’s Dis. JAD* 2017, 57, 1041–1048. [CrossRef] [PubMed]

182. Chen, X.; Hui, L.; Geiger, N.H.; Haughey, N.J.; Geiger, J.D. Endolysosome involvement in HIV-1 transactivator protein-induced neuronal amyloid beta production. *Neurobiol. Aging* 2013, 34, 2370–2378. [CrossRef] [PubMed]

183. Daily, A.; Nath, A.; Hersh, L.B. Tat peptides inhibit neprilysin. *J. Neurovirol.* 2006, 12, 153–160. [CrossRef] [PubMed]

184. Kim, J.; Yoon, J.H.; Kim, Y.S. HIV-1 tat interacts with and regulates the localization and processing of amyloid precursor protein. *PLoS ONE* 2013, 8, e77972. [CrossRef] [PubMed]

185. Stern, A.L.; Ghura, S.; Gannon, P.J.; Akay-Espinoza, C.; Phan, J.M.; Yee, A.C.; Vassar, R.; Gelman, B.B.; Kolson, D.L.; Jordan-Sciutto, K.L. Bace1 mediates HIV-associated and excitotoxic neuronal damage through an app-dependent mechanism. *J. Neurosci.* 2018, 38, 4288–4300. [CrossRef] [PubMed]

186. Liu, Y.; Jones, M.; Hingtgen, C.M.; Bu, G.; Lanibee, N.; Tanzi, R.E.; Nath, A.; He, J.J. Uptake of HIV-1 tat protein mediated by low-density lipoprotein receptor-related protein disrupts the neuronal metabolic balance of the receptor ligands. *Nat. Med.* 2000, 6, 1380–1387. [CrossRef] [PubMed]

187. Maezawa, I.; Nivison, M.; Montine, K.S.; Maeda, N.; Montine, T.J. Neurotoxicity from innate immune response is greatest with targeted replacement of E4 allele of apolipoprotein e gene and is mediated by microglial P38MAPK. *FASEB J.* 2006, 20, 797–799. [CrossRef] [PubMed]

188. Salat, D.H.; Tuch, D.S.; Greve, D.N.; van der Kouwe, A.J.; Hevelone, N.D.; Zaleta, A.K.; Rosen, B.R.; Fischl, B.; Corkin, S.; Rosas, H.D.; et al. Age-related alterations in white matter microstructure measured by diffusion tensor imaging. *Neurobiol. Aging* 2005, 26, 1215–1227. [CrossRef] [PubMed]

189. Salat, D.H.; Tuch, D.S.; van der Kouwe, A.J.; Greve, D.N.; Pappu, V.; Lee, S.Y.; Hevelone, N.D.; Zaleta, A.K.; Growdon, J.H.; Corkin, S.; et al. White matter pathology isolates the hippocampal formation in Alzheimer’s disease. *Neurobiol. Aging* 2010, 31, 244–256. [CrossRef] [PubMed]

190. Wu, Y.; Storey, P.; Cohen, B.A.; Epstein, L.G.; Edelman, R.R.; Ragin, A.B. Diffusion alterations in corpus callosum of patients with HIV. *AJNR Am. J. Neuroradiol.* 2006, 27, 656–660. [PubMed]

191. Chang, L.; Wong, V.; Nakama, H.; Watters, M.; Ramones, D.; Miller, E.N.; Cloak, C.; Ernst, T. Greater than age-related changes in brain diffusion of HIV patients after 1 year. *J. Neuroimmune Pharmacol.* 2008, 3, 265–274. [CrossRef] [PubMed]

192. Thurnher, M.M.; Castillo, M.; Stadler, A.; Rieger, A.; Schmid, B.; Sundgren, P.C. Diffusion-tensor MR imaging of the brain in human immunodeficiency virus-positive patients. *AJNR Am. J. Neuroradiol.* 2005, 26, 2275–2281. [PubMed]

193. Taoka, T.; Iwasaki, S.; Sakamoto, M.; Nakagawa, H.; Fukusumi, A.; Myochin, K.; Hirohashi, S.; Hoshida, T.; Kichikawa, K. Diffusion anisotropy and diffusivity of white matter tracts within the temporal stem in Alzheimer disease: Evaluation of the “tract of interest” by diffusion tensor tractography. *AJNR Am. J. Neuroradiol.* 2006, 27, 1040–1045. [PubMed]

194. Persson, J.; Lind, J.; Larsson, A.; Ingvar, M.; Cruts, M.; Van Broeckhoven, C.; Adolsson, R.; Nilsson, L.G.; Nyberg, L. Altered brain white matter integrity in healthy carriers of the apoe epsilon4 allele: A risk for AD? *Neurology* 2006, 66, 1029–1033. [CrossRef] [PubMed]

195. Hoare, J.; Westgarth-Taylor, J.; Fouche, J.P.; Combrinck, M.; Spottiswoode, B.; Stein, D.J.; Joska, J.A. Relationship between apolipoprotein E4 genotype and white matter integrity in HIV-positive young adults in south Africa. *Eur. Arch. Psychiatry Clin. Neurosci.* 2013, 263, 189–195. [CrossRef] [PubMed]
196. Underwood, J.; Cole, J.H.; Caan, M.; De Francesco, D.; Leech, R.; van Zoest, R.A.; Su, T.; Geurtsen, G.J.; Schmand, B.A.; Portegies, P.; et al. Gray and white matter abnormalities in treated human immunodeficiency virus disease and their relationship to cognitive function. *Clin. Infect. Dis.* 2017, 65, 422–432. [CrossRef] [PubMed]

197. Wendelken, L.A.; Jahanshad, N.; Rosen, H.J.; Busovacca, E.; Allen, I.; Coppola, G.; Adams, C.; Rankin, K.P.; Milanini, B.; Clifford, K.; et al. ApoE epsilon4 is associated with cognition, brain integrity, and atrophy in HIV over age 60. *J. Acquir. Immune Defic. Syndr.* 2016, 73, 426–432. [CrossRef] [PubMed]

198. Morgan, E.E.; Woods, S.P.; Letendre, S.L.; Franklin, D.R.; Bloss, C.; Goate, A.; Heaton, R.K.; Collier, A.C.; Marra, C.M.; Gelman, B.B.; et al. Apolipoprotein E4 genotype does not increase risk of HIV-associated neurocognitive disorders. *J. Neuropathol. 2013*, 19, 150–156. [CrossRef] [PubMed]

199. Hanson, O.; Zetterberg, H.; Buchhave, P.; Londos, E.; Blennow, K.; Minthon, L. Association between CSF biomarkers and incipient Alzheimer’s disease in patients with mild cognitive impairment: A follow-up study. *Lancet Neurol.* 2006, 5, 228–234. [CrossRef]

200. Cysique, L.A.; Hewitt, T.; Croitoru-Lamoury, J.; Taddei, K.; Martins, R.N.; Chew, C.S.; Davies, N.N.; Price, P.; Brew, B.J. ApoE epsilon4 moderates abnormal CSF-abetta-42 levels, while neurocognitive impairment is associated with abnormal CSF tau levels in HIV+ individuals—A cross-sectional observational study. *BMC Neurol.* 2015, 15, 51. [CrossRef] [PubMed]

201. Turner, R.S.; Chadwick, M.; Horton, W.A.; Simon, G.L.; Jiang, X.; Esposito, G. An individual with human immunodeficiency virus, dementia, and central nervous system amyloid deposition. *Alzheimers Dement. (Amst)* 2016, 4, 1–5. [CrossRef] [PubMed]

202. Chang, L.; Andres, M.; Sadino, J.; Jiang, C.S.; Nakama, H.; Miller, E.; Ernst, T. Impact of apolipoprotein e epsilon4 and HIV on cognition and brain atrophy: Antagonistic pleiotropy and premature brain aging. *Neuroimage* 2011, 58, 1017–1027. [CrossRef] [PubMed]

203. Milanini, B.; Valcour, V. Differentiating HIV-associated neurocognitive disorders from Alzheimer’s disease: An emerging issue in geriatric neuroHIV. *Curr. HIV/AIDS Rep.* 2017, 14, 123–132. [CrossRef] [PubMed]

204. Dunlop, O.; Goplen, A.K.; Liestol, K.; Myrvang, B.; Rootwelt, H.; Christophersen, B.; Kvittingen, E.A.; Maehlen, J. HIV dementia and apolipoprotein e. *Acta Neurol. Scand.* 1997, 95, 315–318. [CrossRef] [PubMed]

205. Joska, J.A.; Combrinck, M.; Valcour, V.G.; Hoare, J.; Leisegang, F.; Mahne, A.C.; Myer, L.; Stein, D.J. Association between apolipoprotein e4 genotype and human immunodeficiency virus-associated dementia in younger adults starting antiretroviral therapy in South Africa. *J. Neuropathol.* 2010, 16, 377–383. [CrossRef] [PubMed]

206. Panos, S.E.; Hinkin, C.H.; Singer, E.J.; Thames, A.D.; Patel, S.M.; Sinsheimer, J.S.; Del Re, A.C.; Gelman, B.B.; Morgello, S.; Moore, D.J.; et al. Apolipoprotein-e genotype and human immunodeficiency virus-associated neurocognitive disorder: The modulating effects of age and disease severity. *Neurobehavioral HIV Mol.* 2013, 5, 11–22. [CrossRef] [PubMed]

207. Becker, J.T.; Martinson, J.; Penugonda, S.; Kingsley, L.; Molsberry, S.; Reynolds, S.; Arnow, A.; Goodkin, K.; Levine, A.; Martin, E.; et al. No association between apoepsilon4 alleles, HIV infection, age, neuropsychological outcome, or death. *J. Neuropathol.* 2015, 21, 24–31. [CrossRef] [PubMed]

208. WHO. Who guidelines approved by the guidelines review committee. In *Consolidated Guidelines on the Use of Antiretroviral Drugs for Treating and Preventing HIV Infection: Recommendations for a Public Health Approach* (Available at Pmid: 24716260); WHO: Geneva, Switzerland, 2013.

209. WHO. Consolidated Guidelines on HIV Prevention, Diagnosis, Treatment and Care for Key Populations. 2016. Available online: [https://www.Ncbi.Nlm.Nih.Gov/books/nbk379694](https://www.Ncbi.Nlm.Nih.Gov/books/nbk379694) (accessed on 14 November 2018).

210. Gaida, R.; Truter, I.; Grobler, C.; Kotze, T.; Godman, B. A review of trials investigating efavirenz-induced neuropsychiatric side effects and the implications. *Expert Rev. Anti-Infect. Ther.* 2016, 14, 377–388. [CrossRef] [PubMed]

211. Ma, Q.; Vaida, F.; Wong, J.; Sanders, C.A.; Kao, Y.T.; Croteau, D.B.; Collier, A.C.; Gelman, B.B.; Marra, C.M.; et al. Long-term efavirenz use is associated with worse neurocognitive functioning in HIV-infected patients. *J. Neuropathol.* 2016, 22, 170–178. [CrossRef] [PubMed]

212. Apostolova, N.; Funes, H.A.; Blas-Garcia, A.; Galindo, M.J.; Alvarez, A.; Esplugues, J.V. Efavirenz and the CNS: What we already know and questions that need to be answered. *J. Antimicrob. Chemother.* 2015, 70, 2693–2708. [CrossRef] [PubMed]
213. Swart, M.; Skelton, M.; Ren, Y.; Smith, P.; Takuva, S.; Dandara, C. High predictive value of CYP2B6 SNPS for steady-state plasma efavirenz levels in south African HIV/AIDS patients. *Pharmacogenet. Genom.* 2013, 23, 415–427. [CrossRef] [PubMed]

214. Haas, D.W.; Ribaudo, H.J.; Kim, R.B.; Tierney, C.; Wilkinson, G.R.; Gulick, R.M.; Cliford, D.B.; Hulgan, T.; Marzolini, C.; Acosta, E.P. Pharmacogenetics of efavirenz and central nervous system side effects: An adult AIDS clinical trials group study. *AIDS (Lond. Engl.)* 2004, 18, 2391–2400. [CrossRef]

215. EA, B.; Rodríguez, S.N. The pharmacogenetics of HIV treatment: A practical clinical approach. *J. Pharmacogenom. Pharmacoproteomics* 2013, 4, 116.

216. Gallien, S.; Journot, V.; Loriot, M.A.; Sauvageon, H.; Morlat, P.; Reynes, J.; Reliquet, V.; Chene, G.; Molina, J.M. Cytochrome 2B6 polymorphism and efavirenz-induced central nervous system symptoms: A substudy of the ANRS Alize trial. *HIV Med.* 2017, 18, 537–545. [CrossRef] [PubMed]

217. Masimirembwa, C.; Dandara, C.; Leutsker, P.D. Rolling out efavirenz for HIV precision medicine in Africa: Are we ready for pharmacovigilance and tackling neuropsychiatric adverse effects? *Oncom* 2016, 20, 575–580. [CrossRef] [PubMed]

218. Mukonzo, J.K.; Owen, J.S.; Ogwal-Okeng, J.; Kuteesa, R.B.; Nanzigu, S.; Sewankambo, N.; Thabane, L.; Gustafsson, B.; Ross, C.; Akillu, E. Pharmacogenetic-based efavirenz dose modification: Suggestions for an African population and the different CYP2B6 genotypes. *PloS ONE* 2014, 9, e86919. [CrossRef] [PubMed]

219. Pinillos, F.; Dandara, C.; Swart, M.; Strehalu, R.; Kuhn, L.; Patel, F.; Coovadia, A.; Abrams, E. Case report: Severe central nervous system manifestations associated with aberrant efavirenz metabolism in children: The role of CYP2B6 genetic variation. *BMC Infect. Dis.* 2016, 16, 56. [CrossRef] [PubMed]

220. Sinxadi, P.Z.; Leger, P.D.; McIlerson, H.M.; Smith, P.J.; Dave, J.A.; Levitt, N.S.; Maartens, G.; Haas, D.W. Pharmacogenetics of plasma efavirenz exposure in HIV-infected adults and children in south Africa. *Br. J. Clin. Pharmacol.* 2015, 80, 146–156. [CrossRef] [PubMed]

221. Johnson, D.H.; Gebretsadik, T.; Shintani, A.; Mayo, G.; Acosta, E.P.; Stein, C.M.; Haas, D.W. Neuropsychometric correlates of efavirenz pharmacokinetics and pharmacogenetics following a single oral dose. *Br. J. Clin. Pharmacol.* 2013, 75, 997–1006. [CrossRef] [PubMed]

222. Sandkovsky, U.; Podany, A.T.; Fletcher, C.V.; Owen, A.; Felton-Coleman, A.; Winchester, L.C.; Robertson, K.; Swindells, S. Impact of efavirenz pharmacokinetics and pharmacogenomics on neuropsychological performance in older HIV-infected patients. *J. Antimicrob. Chemother.* 2017, 72, 200–204. [CrossRef] [PubMed]

223. Rippeth, J.D.; Heaton, R.K.; Carey, C.L.; Marcotte, T.D.; Moore, D.J.; Gonzalez, R.; Wolfs, T.; Grant, I. Methamphetamine dependence increases risk of neuropsychological impairment in HIV infected persons. *J. Int. Neuropsychol. Soc. JINS* 2004, 10, 1–14. [CrossRef] [PubMed]

224. Weber, E.; Morgan, E.E.; Judicello, J.E.; Blackstone, K.; Grant, I.; Ellis, R.J.; Letendre, S.L.; Little, S.; Morris, S.; Smith, D.M.; et al. Substance use is a risk factor for neurocognitive deficits and neuropsychiatric distress in acute and early HIV infection. *J. Neurovirol.* 2013, 19, 65–74. [CrossRef] [PubMed]

225. Passaro, R.C.; Pandhare, J.; Qian, H.Z.; Dash, C. The complex interaction between methamphetamine abuse and HIV-1 pathogenesis. *J. Neuroimmune Pharmacol.* 2015, 10, 477–486. [CrossRef] [PubMed]

226. Chermer, M.; Bousman, C.; Everall, I.; Barron, D.; Letendre, S.; Vaida, F.; Atkinson, J.H.; Heaton, R.; Grant, I. Cytochrome p450-2d6 extensive metabolizers are more vulnerable to methamphetamine-associated neurocognitive impairment: Preliminary findings. *J. Int. Neuropsychol. Soc. JINS* 2010, 16, 890–901. [CrossRef] [PubMed]

227. Fellay, J.; Marzolini, C.; Meaden, E.R.; Back, D.J.; Buclin, T.; Chave, J.P.; Decosterd, L.A.; Furrer, H.; Opravil, M.; Pantaleo, G.; et al. Response to antiretroviral treatment in HIV-1-infected individuals with allelic variants of the multidrug resistance transporter 1: A pharmacogenetics study. *Lancet* 2002, 359, 30–36. [CrossRef]

228. Vautier, S.; Fernandez, C. Abcb1: The role in parkinson’s disease and pharmacokinetics of antiparkinsonian drugs. *Expert Opin. Drug Metab. Toxicol.* 2009, 5, 1349–1358. [CrossRef] [PubMed]

229. Sankatsing, S.U.; Beijnen, J.H.; Schinkel, A.H.; Lange, J.M.; Prins, J.M. P glycoprotein in human immunodeficiency virus type 1 infection and therapy. *Antimicrob. Agents Chemother.* 2004, 48, 1073–1081. [CrossRef] [PubMed]

230. Wang, D.; Johnson, A.D.; Papp, A.C.; Kroetz, D.L.; Sadee, W. Multidrug resistance polypeptide 1 (MDR1, ABCB1) variant 3435C>T affects mRNA stability. *Pharmacogenet. Genom.* 2005, 15, 693–704. [CrossRef] [PubMed]
231. Dirson, G.; Fernandez, C.; Hindlet, P.; Roux, F.; German-Fattal, M.; Gimenez, F.; Farinotti, R. Efavirenz does not interact with the abcb1 transporter at the blood-brain barrier. Pharm. Res. 2006, 23, 1525–1532. [CrossRef] [PubMed]

232. Johnson, D.H.; Sutherland, D.; Acosta, E.P.; Erdem, H.; Richardson, D.; Haas, D.W. Genetic and non-genetic determinants of raltegravir penetration into cerebrospinal fluid: A single arm pharmacokinetic study. PLoS ONE 2013, 8, e82672. [CrossRef] [PubMed]

233. Lee, C.Y.D.; Daggett, A.; Gu, X.; Jiang, L.L.; Langfelder, P.; Li, X.; Wang, N.; Zhao, Y.; Park, C.S.; Cooper, Y.; et al. Elevated TREM2 gene dosage reprograms microglia responsivity and ameliorates pathological phenotypes in Alzheimer’s disease models. Neuron 2018, 97, 1032–1048. [CrossRef] [PubMed]

234. Fields, J.A.; Spencer, B.; Swinton, M.; Qvale, E.M.; Marquine, M.J.; Alexeeva, A.; Gough, S.; Soontornniyomkij, B.; Valera, E.; Masliah, E.; et al. Alterations in brain TREM2 and amyloid-beta levels are associated with neurocognitive impairment in HIV-infected persons on antiretroviral therapy. J. Neurochem. 2018. [CrossRef] [PubMed]

© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).