LETTER TO EDITOR

SARS-CoV-2 infection inducing severe flare up of Deficiency of Interleukin Thirty-six (IL-36) Receptor Antagonist (DITRA) resulting from a mutation invalidating the activating cleavage site of the IL-36 receptor antagonist

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To the Editor,

The Deficiency of Interleukin Thirty-six Receptor Antagonist (IL36RA) (DITRA) is a rare autoinflammatory monogenic syndrome resulting from the interaction of autosomal-recessive or double composite heterozygous mutations of IL36RN, the gene encoding IL-36RA, with environmental triggers such as infection and pregnancy. This sporadic or familial syndrome is characterized by recurrent acute flares of generalized pustular psoriasis (GPP, OMIM 614,204), with life-threatening potential. Recently, we reported for the first time documented viral respiratory tract infections (RTI) as potent triggers in acutely flaring psoriasis of different subtypes, including GPP [1]. We herein report a case of acute, severe DITRA flare triggered by SARS-CoV-2 infection, and discuss the implications regarding disease pathogenesis and management in view of immunologic profiling studies.

The causal mutation in this pedigree has already been reported as a p.V2F aminoacid substitution in the IL-36RA protein resulting from a homozygous c.4G>T nucleotidic variant of IL36RN in 3 out of 4 siblings born from related parents of Algerian descent [2]. This mutation is unique in impairing the cleavage of the immature form of IL-36RA at aminoacid residue 2 by proteases, mainly by neutrophil-derived elastase, a mandatory step for IL36RA optimal biological activity [3]. There was no mutation of CARD14 or AP1S3 in any individual from the pedigree, and the parents and one sibling have been shown to be heterozygous carriers of the IL36RN c.4G>T mutation, and were free of any skin or mucosal lesions (see Fig. 1 for the pedigree).

A 16-year-old male patient (Fig. 1) suffering from early onset DITRA with frequently relapsing typical GPP with systemic symptoms and a geographic tongue, had been in complete stable clinical remission for several months after withdrawal from adalimumab treatment, which had been initiated following failures of acitretin and cyclosporine. In late september 2020, he reported a rapidly disseminating pustular eruption with superficial scaling and yellow crusts accompanied with asthenia and myalgia, occurring 2 days after the onset of the first COVID-19 symptoms consisting in cephalalgia, fever, pharyngitis with anosmia and ageusia. Three weeks later, he was clinically assessed for a GPP/DITRA eruption covering 60% of the body surface area. The 5 points scale Generalized Pustular Psoriasis Physician Global Assessment (GPPPGA) score performed by a trained physician (HB) was 3 (moderate) with presence of pustular lakes, while the generalized pustular psoriasis area and severity index (GPPASI) was 45.2 for a maximal value of 72 [4]. (For full details, see online additional data supplementary ESM1). Biological investigations revealed peripheral blood leukocytosis with neutrophilia (neutrophil count: 17,610/mm³ and lymphocyte count: 2460/mm³), while C-reactive protein (CRP) serum level was normal. Polymerase chain reaction (PCR)-based analysis of nasopharyngeal swabs, using a Filmarray multiplex test detecting sixteen respiratory viruses and four bacteria (Chlamydophila...
pneumonia, Mycoplasma pneumonia, Legionella pneumophila, Bordetella sp.) (Biomérieux, Macry-l’Etoile, France) confirmed the diagnosis of COVID-19 with the exclusive presence of SARS-CoV-2. Both E and N genes were detected with Cycle Threshold value at 38 (Xpert® Xpress SARS-CoV-2, Cepheid, Sunnyvale, CA). The skin rash partially resolved under topical steroid treatment with betamethasone once daily, while the SARS-CoV-2 PCR-based detection in nasopharyngeal swabs became negative one week later. The serological detection of anti-SARS-CoV-2 nucleocapsid (N) and spike (S) IgG antibodies using a chemiluminescent microparticle immunoassay (Architect, Abbott, Sligo, Ireland), performed 32 days after COVID-19 symptoms onset, was strongly positive (index 5.79), whereas PCR and serological detection of SARS-CoV-2 were negative in his heterozygous carrier mother (Fig. 1 patient I.1), in his 5-year-old sister (patient II.4) with mild and limited GPP/DITRA skin lesions (GPPPGA:2; GPPASI:3), as well as in his 18 year-old brother (patient II.1) who showed plicatuated tongue as the only DITRA symptom since birth, both homozygously mutated for \( IL36RN \). The SARS-CoV-2-induced GPP flare completely resolved within 3 months after restart of adalimumab treatment, without relapse over a 6 months follow-up period. To note, the patient’s medical records revealed one episode of GPP/DITRA acute flare already triggered by infection with \( Bordetella pertussis \).

As GPP has been associated with both neutrophilic and various degrees of type I interferon (IFN-I) transcriptomic signatures [5], IFN signature genes (ISG) and neutrophil signature genes (NSG) mRNA expression was quantitated using whole blood nanostring technology as previously described [6], at day 32 post-onset of COVID-19 symptoms in the proband and in his mildly affected sister. Results showed a strongly increased neutrophilic signature in the proband, but not in the mildly affected sister, while mRNA expression of ISG was in the normal range in both cases (Fig. 1).

Fig. 1 On the left, a Family pedigree. b Acute flare up of generalized pustular psoriasis of the proband (patient II.2). On the right, Interferon Signature Genes (ISG) and neutrophil signature genes mRNA quantitative assessment in whole blood from patient II.2 (proband) and patient II.4 (proband’s sister). c Median fold expression of interferon-stimulated genes (ISG)*. d Median fold expression of neutrophil signature genes**. *ISG signature assessment: Total RNA was extracted from whole blood using the PAXgene RNA isolation kit (PreAnalytix, Hombrechtikon, Switzerland). The ISG mRNA expression was assessed using the nanoString (NanoString, Seattle, WA, USA) procedure; 200 ng of RNA was hybridized to the probes (a reporter probe and a capture probe) at 67 °C for 16–21 h. Samples were then inserted into the nCounter Prep Station for the removal of excessive probes, purification, and immobilization onto the internal surface of a sample cartridge for 2–3 h. Finally, the sample cartridge was transferred to the nCounter Digital Analyzer, where color codes were counted and tabulated for each target molecule. Count numbers obtained for the ISGs were normalized by the geometric mean of three housekeeping genes count numbers (β-actin, HPRT1 [hypoxanthine phosphoribosyltransferase 1] and POLR2A [RNA polymerase II subunit A]) as well as the negative and positive control values using the nSolver software. The median fold change of the ISGs compared to the median of the healthy controls. ** Neutrophil signature genes include CEACAM6, CRISP3, DEFA4, LCN2, LTF, and MMP8
To our knowledge, this is the first case of DITRA flare-up induced by SARS-CoV-2 infection. Indeed, the complete, stable remission status in patient 1 several months before the COVID-19 infection onset, the personal prior history of post-infectious DITRA flare-up, and previous reports of GPP flares occurring concomitantly or immediately after RTI including a recent case of post-SARS-CoV-2 new onset acral PP without evidence of DITRA, all support the impact of infectious triggers, especially of viral RTI, on the evolutive profile of this autoinflammatory syndrome [1, 7]. The molecular and cellular interactions linking RTI- and specifically SARS-CoV-2-induced bronchial inflammation, and skin and systemic inflammation in GPP, remain unknown to date. Although IFN-I, which has been showed to be instrumental in an effective anti-SARS-CoV-2 immune response [8] and to be upregulated in a subset of patients with GPP, might be involved in this inflammatory cascade, we did not find evidence for its deregulation, in discrepancy with previous reports in GPP without Covid19 [5]. This might be due either to the late sampling after the infection onset in our patient, or to alternative innate immune mechanisms operating in some patients. Likewise, the strong transcriptomic neutrophilic blood signature is in keeping with previous data in lesional skin of GPP patients showing upregulation of IL-36- and of neutrophil-attracting and/or activating chemokines such as CXCL-1, CXCL-2 and CXCL-8, among others [9]. Moreover, this case emphasizes the key role of elastase-driven catalytic cleavage of immature, biologically inactive form of IL-36RA for an optimal function of the antagonist, which is mandatory to prevent acute skin neutrophilic inflammation following environmental triggering events including SARS-CoV-2 infection [3]. Our observation has some limitations: first, the blood transcriptome assessment was performed in the proband 32 days after symptoms onset. Second, these experiments do not address the signatures in lesional skin or in lung tissues.

Finally, and due to the orphan natures of GPP and of DITRA, it is still unknown if inborn errors leading to upregulation of the IL-36 pathway and to GPP are protective or deleterious with regard to SARS-CoV-2 infection outcomes. In this respect, it is interesting to notice that COVID-19 infection was benign but symptomatic in our patient, despite the absence of upregulated IFN-I response. Given the yet equivocal prognostic value of a strong neutrophilic signature in COVID-19, it would be of interest to investigate the clinical outcomes and biological correlates of SARS-CoV-2 infection in neutrophilic systemic diseases such as GPP and DITRA on a larger scale, as currently only data in patients with psoriasis vulgaris are available at the population level. Finally, the growing evidence for a role of the IL-36 pathway in bronchial inflammation and the present case pave the way for mechanistic studies aiming to decipher the molecular and cellular links between RTI and psoriasis flare-ups.

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Author Contribution AB and HB wrote the manuscript. FA and HB performed the experiments and arranged the figures. AB and EB ongoing clinical care for the patient. HB supervised the study. All authors edited and approved the final version.

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Data Availability All data and materials can be obtained by contacting the corresponding authors.

Declarations

Ethical Statements The study was approved by the Leeds (East) Research Ethics Committee (reference 10/H1307/132) and by the Comité de Protection des Personnes (ID-RCB/EUDRACT: 2014-A01017-40 and 2011-A00820-41). Written informed consent was obtained from all patients before inclusion.

Consent to Participate The patient and the patient’s mother have consented to the submission of the case report to the journal.

Consent to Publish The patient and the patient’s mother have consented to the submission of the case report to the journal.

Conflicts of Interest Alizée Bozonnat, Florence Assan, Jérôme Le Goff and Emmanuelle Bourrat declare no conflict of interests nor any competing interests. Hervé Bachelez had paid consulting activities for AbbVie, Almirall, Boehringer-Ingelheim, Janssen, Kyowa-Kirin, Leo Pharma, Novartis, UCB, and Pfizer.
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