On the origin of Carcinoma

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The origin of cancer remains one of the most important enigmas in modern biology. The prevailing paradigm has failed to grasp a comprehensive view of the disease. Naturally, therapies developed under the current assumptions are inadequate and cancer is practically an incurable disease. Meanwhile, descriptive studies continuously extend the molecular complexity of cancer without an equivalent advancement in its understanding. Furthermore, they tend to accumulate inconsistencies inexplicable under the classical view. This paper presents a compelling theory of the origin of carcinomas. By hypothesis, a series of generic events in epithelial tissues promoted by cellular aging and inflammation enables the reactivation of developmental programs. The origin of carcinomas in vivo is described as the time-ordered cell state transitions undergone by epithelial cells in the hyperplasia due to replicative senescence and inflammation towards a mesenchymal undifferentiated endogenous cell state with cancerous behavior. In support of the theory, the molecular, cellular, and histopathological evidence is critically reviewed. A plausible model for the origin of carcinomas is presented to explain the mechanism underlying carcinogenesis from an evolutive and developmental perspective. The implications of the hypothesis in the current strategies for cancer prevention and treatment are discussed along with rational alternatives and some predictions for possible experimental validation.

Keywords: Senescence, Immortalization, Epithelial to Mesenchymal Transition, Carcinogenesis, Hypothesis

Introduction

Cancer is one of the leading causes of death worldwide. Every year 14 million people are diagnosed and 9 million will die from cancer. Despite the enormous amount of research and financial support during the last seven decades, no significant differences have been observed in mortality rates. Late diagnosis has been considered the main challenge in oncology since therapy is ineffective in advanced stages. Currently, over half of the patients are diagnosed with metastatic disease and 90% will experience therapy failure due to tumor resistance or unbearable side effects. The initial favorable response in the control of tumor growth is inexorably followed by the progression despite therapy in less than six months. Regarding toxicity, the mortality derived from cancer therapeutics itself may appear in 50% of the cases during the first month of application. The side effects are responsible for 20% of the suspension of the protocol since patients could experience intolerable mucositis, myelosuppression, pulmonary fibrosis, cardiotoxicity, hepatotoxicity, and nephrotoxicity. Strikingly, most antineoplastic agents and radiotherapy induce cancer in animals and humans, including the oncologic patients in which they are called secondary cancers. Despite over 50 years of research since the original protocols against cancer, the overall contribution of chemoirradiation to survival is less than 2% and there is no convincing evidence that the newer and expensive agents outperform his predecessors. In essence, the lack of understanding of cancer itself, how it develops, and progresses continues to halt the development of effective therapeutic strategies.
The prevailing paradigm for the origin of cancer is the somatic mutation theory. From this perspective, cancer is a genetic disease in which DNA mutations lead to a cancerous phenotype. Despite the lack of success, the rationale of therapy has been to kill cancer cells mostly with DNA poisons in combination with agents that target the molecular alterations conferred by the acquired genetic traits. According to the somatic mutation theory the genetic alterations allow cancer cells to proliferate uncontrollably, the ability to invade nearby and distant tissues, thus compromising systemic functions that eventually cause death. The processes that are considered altered in cancer cells are the proliferation signals, the evasion of growth suppression, the resistance of cell death, the restoration of limitless replicative capacity, changes in energy metabolism, the activation of invasion and migration. Furthermore, it assumes that cancer cells are constantly evolving due to genomic instability and by a Darwinian mechanism cells become resistant to therapy and able to escape the immune response. Additionally, mutations somehow confer tumors the capacity to reshape the tumor microenvironment to support its progression by the reactivation of processes such as angiogenesis and inflammation.

Despite the massive resources devoted to genomic sequencing of human cancers, in essence, it has not been possible to find an underlying genetic cause for carcinogenesis. Most oncogenes and tumor suppressors play crucial roles in the normal biology of cells. The somatic mutation theory is also challenged by several experimental inconsistencies, for example, many carcinogens lack from mutagenic effects, cancer cells show morphological and transcriptional convergence regardless of the initial cellular phenotype, and cancerous behavior can be adopted through processes of transdifferentiation. Notably, the reversion of the malignant phenotype is possible through the induction of differentiation using chemical agents, vitamins, transcription factors, and interactions with the extracellular matrix or the stroma. Those findings are incomprehensible with the current paradigm, and at the same time suggest an endogenous cellular state that can be accessed or discarded according to the nature of the stimulus.

In this paper, an alternative framework to understand the emergence and progression of carcinomas is proposed. In this hypothesis, it is considered that the functional, molecular, and developmental properties of epithelial cells, together with their organization as tissue promote their transformation. In addition, carcinomas share a pattern of histological progression and molecular events that suggest a generic process underlying carcinogenesis. Hence, one of the premises is that cancer is organized by conserved mechanisms operating behind the genetic alterations. It is proposed that hyperplasias enable the reactivation of developmental processes and that malignant cells arise from cellular senescence using time-ordered cell state transitions. In support of the idea, is provided a detailed overview of the molecular, cellular, and structural process that allows the reiteration of embryonic programs in epithelial hyperplasia. To accomplish an integrated hypothesis for the origin of carcinomas, this paper challenges the granted role of mutations, senescence, inflammation, fibroblasts, stem cells, and the epithelial to mesenchymal transition in carcinogenesis. Instead, the plasticity of epithelial cells induced by aging, inflammation, and tissue rearrangements are considered the most important factors for in vivo carcinogenesis. Cancer as the pathological outcome of the same mechanisms activated in development and during tissue regeneration.

The paper is organized into eight parts. The first presents the intrinsic plasticity of epithelial cells, the progressive nature of carcinomas, and the conserved histological and molecular events during their carcinogenesis. The second part covers the current role of senescent cells in carcinogenesis and their susceptibility to transformation. The third part focuses on the epithelial to mesenchymal transition and the mirroring of epithelial carcinogenesis with the spontaneous immortalization of epithelial cells in vitro. Part fourth presents the theory of the origin of carcinoma and the impact of aging, inflammation, and structural factors in tissues and their implication on the nature of metastasis. Part five covers aspects related to inflammation and cellular plasticity. Section six is devoted to uncovering the elusive nature of myofibroblasts. In part seven is discussed stemness as the cellular state with malignant behavior and cancer and chronic
diseases as part of development. The last section discusses the current strategies of cancer prevention and therapy in the light of the hypothesis and suggests a rational alternative and some predictions for possible experimental validation.

1. The epithelial cells and the transformation of the epithelium

1.1. The intrinsic plasticity of epithelial cells

Cancer that originates from epithelial tissue is called a carcinoma and is the most common type of neoplasia in humans. Up to 85% of cancer diagnosed in adults begins in the skin, breast, endometrium, prostate, colon, lung, pancreas, bladder, liver, or cervix. In general, epithelial tissues are constituted by two types of cells, the epithelial cells that line the surface of a tissue or organ and the mesenchymal cells located in the extracellular matrix. At the microscopic level, they are composed of epithelial cells attached to the basement membrane, which establishes an axis of apical-basal polarity and they communicate with each other through cell-cell interactions. Below the basement membrane is the stroma, which is formed by the three-dimensional extracellular matrix synthesized by the mesenchymal cells. Broadly, epithelia form specialized three-dimensional structures adapted to their physiological function, as is the case with secretory organs. These tissues secrete glandular products mainly from the apical surface and some also from the basal surface. Epithelial cells also have a polarized distribution of organelles with distinct apical and basolateral domains. Transmembrane proteins have cytoplasmic tails that bind to elements of the cytoskeleton, forming protein complexes necessary for their organization. These domains allow the polarity of the plasma membrane to be established, they provide an adhesion site for intercellular junctions and anchoring for the molecules of the intracellular signaling pathways responsible for the differentiation and survival of epithelial cells.

Transcription factors with a conserved ETS domain (epithelium-specific) are considered specific to epithelia and play a crucial role in their differentiation. This family of proteins has 26 members, which are important mediators in the morphogenesis and development of epithelial tissues by regulating their gene expression. The conserved ETS domain allows direct binding to promoter and enhancer regions of target genes crucial for the proliferation and formation of the cellular interactions of epithelial cells. The studies of mice in which the expression of ETS proteins was genetically eliminated revealed their importance in the correct differentiation of the structures of epithelial tissues. Some members of the family have been identified ubiquitously expressed, however, a subgroup called ESE proteins (ETS specific of epithelium) are reserved for epithelial tissues. These transcription factors are ESE-1, ESE-2, ESE-3, and the PDEF (Prostate-derived Ets factor).

The transcription factor ESE-1 is widely expressed in organs such as the lung, stomach, kidneys, colon, and skin. ESE-3 is constitutively expressed in the lung epithelium, prostate, pancreas, salivary glands, and trachea. PDEF is specifically expressed in the glandular epithelial cells of the prostate. Finally, the transcription factor ESE-2 is expressed from the extra-embryonic ectodermal lineage, it is essential for the survival of the embryo and in the formation of the placenta in mammals. This protein is expressed in the mammary gland, salivary gland, kidney, stomach, and skin. Consistently, it has been found that the silencing or downregulation of the ESE family of proteins is necessary for the loss of differentiation of the epithelial phenotype and the generation of carcinomas.

Epithelial cells are terminally differentiated; however, they maintain great cellular plasticity that is evident since the early stages of development. This intrinsic plasticity could be the origin of the susceptibility of epithelial tissues to malignant transformation. Interestingly, the first type of differentiated tissue to emerge during embryo development is the epithelia. From the eight-cell stage until the blastula, the blastomeres adhere using the binding protein epithelial cadherin (E-cadherin) and syndecan. This allows the aggregation of cells and the formation of a two-dimensional sheet. Differentiation of the epithelium is further promoted by the interaction of receptors with components of the basal lamina. The second phenotype in appearing is
mesenchymal, it arises from epithelial cells that undergo the process of epithelial to mesenchymal transition (EMT). The mesenchymal phenotype basically emerges from the suppression of the factors that induce epithelial differentiation \(^{23}\). The increased expression of the zinc-finger binding transcription factor (Snail), the Snail Family Transcriptional Repressor 2 (Slug), the twist family bHLH transcription factor 1 (Twist1), the Twist-related protein 2 (Twist2), and the Zinc finger E-box-binding homeobox protein 1 (ZEB1) and ZEB2 promote the downregulation of the ESE proteins. As a result, the E-cadherin and the epithelial phenotype are substituted by the neural cadherin (N-cadherin), vimentin, and the rest of the mesenchymal biomarkers and the eventual full acquisition of a fibroblastic morphology \(^{32}\). The process of EMT begins in the gastrulation with the breakdown of the basement membrane that underly the epiblast and is associated with the effects of fibroblast growth factor (FGF) on cells of the primitive streak and with the activation of the mesenchymal transcription factors such as Snail. The induction of EMT is completed with additional signals that lead to the migration of cells within the primitive streak. These cells undergo the reverse process called the mesenchymal to epithelial transition (MET) and originate the endoderm or remain differentiated in the mesenchyme and create the mesoderm. This migration of mesenchymal cells through the embryo is essential for proper morphogenesis \(^{33}\).

The intrinsic plasticity of epithelial cells displayed during embryo development resembles many of the observed phenomena in the carcinogenesis of epithelial tissues. Therefore, the events that result in the reactivation of such a process in the adult could be sufficient to initiate the malignant transformation of the epithelium and its dissemination, which outcome is the lethal metastatic disease.

1.2. The carcinogenesis of epithelial tissues

The incidence of carcinomas increases exponentially with age, especially in patients suffering from chronic inflammation. The relationship between aging, inflammation, and tumorigenesis is traditionally understood from the genetic point of view of cancer. Namely, aging and inflammation increase the chances of developing and accumulating random somatic mutations that eventually generate clones of epithelial cells with cancerous characteristics \(^{34}\). In contrast, a thoughtful examination of the histological progression of carcinomas in adults suggests a robust pattern of cellular and molecular events that suggest developmental processes rather than events generated by chance.

Histology is the study of the anatomy of cells and tissues and remains as the gold standard for cancer diagnosis. No molecular studies have overcome the careful examination of tissues in the microscope. The appearance of the epithelial tissue determines whether is normal, hyperplastic, or cancerous and its degree of malignancy. The use of complementary techniques provides a deeper insight into the molecular events that are taking place in the tissue by the identification of biomarkers. In this regard, current histological data support the progressive nature of carcinomas \(^{35-37}\). From this perspective, carcinomas are preceded by atypical hyperplasia that in turn progress from typical hyperplasia. The presence of atypia increases five times the risk of developing carcinomas. Therefore, atypical hyperplasia is considered the precursor of carcinoma in situ (CIS), which in turn increases 10 times the relative risk of developing invasive carcinoma. The carcinoma arising from the CIS then progresses from low to high histological grade (G1-G2-G3). The latter is a classification that describes the cellular and nuclear characteristics of cells and stages its degree of differentiation \(^{37}\). Furthermore, the histological progression of carcinomas in humans is also recapitulated in murine models of chemical carcinogenesis \(^{38}\) (Fig. 1.).

It seems very unlikely that the temporal order of histological events during epithelial carcinogenesis is the random result of genetic mutations and Darwinian selection. Instead, the progressive transformation of epithelial tissues suits better the premises of cancer as a developmental disease. From this perspective, cancer emerges by the reactivation of embryonic programs and not from novel traits earned by mutagenesis \(^{39}\).
Hyperplasia is considered the precursor lesion of cancer that originates in epithelial tissues. The diagnosis of atypical hyperplasias increases the risk of being subsequently diagnosed with invasive cancer \(^{40}\). In hyperplasias, are common the detection of biomarkers of cellular senescence such as short telomeres, expression of the protein p16, and the infiltration of immune cells \(^{41-43}\). As it progresses to carcinoma, the recovery in the length of the telomeres, telomerase enzyme activity is observed while decreasing the presence of senescent cells \(^{44, 45}\). In high-grade carcinomas, glandular histology and biomarkers of epithelial differentiation are lost \(^{46, 47}\), together with an increase in the expression of transcription factors of a mesenchymal phenotype, the presence of metastases \(^{48}\), and stem cell biomarkers \(^{49}\).

The following section describes the molecular events that would trigger epithelial carcinogenesis during aging, the role of inflammation, and the process that shapes the histological progression towards the gradual loss of differentiation.

2. The aging of epithelial tissues

2.1. The hyperplasia stains positive for senescence

The hyperplasias of the epithelial tissue are characterized by the presence of senescent cells, but its progression to carcinomas is accompanied by their lost \(^{41, 50}\). Therefore, the changes that hyperplasia undergoes in vivo are crucial for the understanding of epithelial carcinogenesis. The presence of senescent cells increases with age, mainly in tissues that contain mitotically competent cells \(^{51}\), which is consistent with the steady proliferation of epithelial tissues for their renewal and the maintenance of functions \(^{52}\). However, inherent to the biochemistry of the replication of genetic material, telomeric DNA base pairs are lost in each cell cycle. Telomeres are DNA sequences that, together with associated proteins, cover and stabilize the ends of chromosomes and prevent them from degradation or fusion \(^{53}\). Eventually, short telomeres activate a signal of genetic damage that originate cell cycle arrest and the acquisition of the phenotype known as replicative cellular senescence \(^{54}\). Senescent cells coexist with short telomeres in hyperplasias and trace the mitotic rate of the affected epithelial tissue \(^{55}\).

The DNA damage response (DDR) is activated by telomere erosion \(^{56}\). In senescent cells are detected structures in the telomeres known as the DNA repair foci, that are also formed during double-strand breaks \(^{57}\). One of the initial steps in the formation of those compartments is the phosphorylation of the histone family member X (H2AX) by the ATM protein kinase (Ataxia telangiectasia mutated) \(^{58}\), which also activates the proteins p53 and Chk2 (Checkpoint kinase 2) \(^{59}\). In turn, p53 activates its transcriptional targets, such as the
cyclin-dependent kinase inhibitor 1 (p21) 60 and PML (Promyelocytic leukemia protein) 61; both recognized biomarkers of senescence 62. The p21 protein inhibits the activity of the cyclin-CDK 2/4 complexes and the family of transcription factors E2F leading to cell cycle arrest 63. The PML protein is an essential component of the nuclear bodies that also accumulate in the senescent cells 64. These compartments of proteins belong to the nuclear matrix, a superstructure related to DNA replication, transcription, and epigenetic silencing 65. PML recruits the proteins cyclin-dependent kinase inhibitor 2A (p16), p53, and the pRb / E2F complex to the nuclear bodies and modulates the expression of genes associated with cellular senescence 66. The gene p16 is located within the locus of the inhibitors of kinase (Ink4b/ARF/Ink4a) on chromosome 9p21. The locus encodes three genes; p16 and using an alternative reading frame (ARF) is enabled the transcription of the ARF protein product of CDKN2A (p14), and the cyclin-dependent kinase inhibitor 2B (p15) 67. The induction of senescence recruits the polycomb proteins (PcG), the Junoniji domain-containing protein 3 (Jmd3), the enhancer of zeste homologue 2 (EZH2), and the Mixed lineage leukemia 1 protein (MLL1) to the locus Ink4b/ARF/Ink4a 68. The dissociation of the PcG proteins and the activity of the histone demethylases and methyltransferases result in the reconfiguration of chromatin and the transcriptional capability of cells 69. On the other hand, the proteins p16 and p15 inhibit the activity of the cyclin-dependent kinase (CDK) 4 and 6 preventing both, the phosphorylation of Rb protein and the actions of the E2F 70. Therefore, in senescence, the reorganization of chromatin coexists with the cell-cycle arrest 71. Besides, the p14 protein increases the activity of p53 by the negative regulation of its inhibitor the protein murine double minute 2 (Mdm2). Thus, in senescent cells, the cell cycle arrest is temporarily amplified 72, but the epigenetic changes will have some paradoxical consequences 73.

The cellular senescence was positioned as an irreversible arrest in the cell cycle which function in cancer as a tumor suppressor. Hence, precancerous lesions should overcome senescence to restore proliferation 74 by mutations or deregulation in the p16 and p53 signaling pathways 75. Later, the description of the secretory phenotype of the senescent cells, not only reinforces the potential role of inflammation in the transformation of the aged tissue but also offers a suitable explanation to overcome the molecular restrictions imposed in the cell cycle. Namely, the increased production of the mediators of inflammation and reactive oxygen species would lead to an increased chance of mutations that results in the abrogation of cellular senescence. The secretion of senescent cells includes cytokines, chemokines, growth factors, and proteases 76 which its expression was found to be coordinated by the transcription nuclear factor kappa B (NF-κB) 77. This protein itself portrays the process of inflammation, cancer development, and progression. The constitutive activation of NF-κB is frequent in tumors and its effects include the induction of proliferation, migration, and the inhibition of apoptosis 78. As consequence, the influence of the secretions of senescence cells on inflammation, proliferation, and tissue microenvironment plays a bigger role in the current understanding of the association of cellular aging and cancer. The premises are that in the initial phases of carcinogenesis, the immune system destroys senescent cells but during aging their accumulation surpass the clearance and the burden of inflammation became carcinogenic 79.

2.2. Overcoming the assumed role of cellular senescence

In this paper, a new role of senescent cells in epithelial carcinogenesis is considered. By hypothesis, the origin of carcinoma is the result of cellular transitions from senescent cells. To fully embrace the proposal is necessary to consider the replicative senescence as a cellular phenotype, and to explore the concept of cellular transitions and their interplay with the histological context. It is hypothesized that hyperplasia is the most plausible scenario to foster cellular transitions in vivo and cells arising from such events of transdifferentiation are primed with cancerous behavior. Therefore, aging and inflammation induce a deregulated but still endogenous cellular plasticity, which is the ultimate culprit for epithelial carcinogenesis.

The boundaries imposed in the concept of cellular senescence in carcinogenesis has been surpassed consistently in cell cultures. The epithelial cells overcome senescence in the absence of mutations in the locus
Ink4b/ARF/Ink4a, and far from being irreversible, the cellular state of senescence is now considered an unstable condition from which cells eventually emerge. Furthermore, cellular senescence can be viewed as a cell fate since any cell regardless of its original lineage activates the same conserved program. The senescence is accompanied by the loss of its initial cellular biomarkers, morphology, and transcriptome. Cells accumulate in the G1/G2 phase of the cell cycle despite nutrients and growth factors. The metabolism remains active, acquires a greater adhesion to the extracellular matrix, and the cell-cell contacts. The cells became secretory and more resistant to apoptosis. The structural rearrangements in the chromatin of senescent cells are so significant that can be viewed by light microscopy. The variations in the nuclear lamina are accompanied by the loss of heterochromatin domains and the gain of heterochromatic foci. Further molecular studies showed that the epigenome of replicative senescent cells undergo significant hypomethylation, and the result is the loss of repression of thousands of genes including those associated with cancer development.

The preceding is considered as a plausible endogenous event of epigenetic reprogramming in the epithelial hyperplasia. Cell reprogramming refers to the induction of the loss of cell differentiation in somatic cells towards an embryonic stem cell phenotype. In this context, the process of the epithelial cell undergoing replicative exhaustion is viewed as the first cellular transition, which means a lineage conversion among different cell phenotypes. The switch into another cell type of a different lineage through genetic reprogramming is also called transdifferentiation. Under this view, the replicative senescence predisposes the epithelia to carcinogenesis by epigenetic events. Once in the senescent state, the cell is prone to further cellular transitions and the mechanism that links the abrogation of cellular senescence along with subsequent events of dedifferentiation and carcinogenesis associates with inflammation. Observations of the transformation of hyperplasias and cultures of epithelial cells provide evidence of this assumption.

3. The transformation of epithelial tissues

3.1. Beyond cellular senescence

The concept of immunosurveillance considers the elimination of cancer cells as one of the most important functions of the immune system. For this reason, the infiltration of leukocytes in neoplasms was explained as an attempt of our body to destroy premalignant cells. Paradoxically, several studies have been shown that the infiltration of hyperplasias is positively correlated with their histological progression towards invasive carcinomas.

According to the essential thesis of this paper, the senescent cells in vivo undergo another cellular transition by the effect of the cytokines along with the structural changes fostered by the inflammation in the microenvironment of the hyperplasia. The infiltration of cells of the immune system into epithelial hyperplasias occurs due to the secretory phenotype of the senescent cells. The consequence is an inflammatory cellular response that increases the concentration of cytokines creating a positive feedback loop. Then, it is theorized that a certain threshold of inflammation is necessary before the induction of an EMT-like transition in senescent cells, the result is cells that arise with cancerous behavior (Fig. 2.).

The relationship of the EMT with the suppression of cellular senescence in epithelial cells has been discussed primarily in the context of cancer progression. Consistent with the suggested temporal order of cellular transitions, it has been documented in the cultures of cancer cell lines that undergo senescence the emergence of subpopulations of cells with high expression of the mesenchymal transcription factor Zeb. Furthermore, the induction of oncogene overexpression first induces the senescence of cells, and upon the ectopic overexpression of the mesenchymal transcription factor Twist, the cellular senescence is eliminated by the induction of EMT. The effect of many oncogenic signals on senescent cells is accompanied by EMT and the transformation from the flat cell phenotype to a fibroblast morphology and the upregulation of the mesenchymal biomarkers. The capacity of the soluble mediators of inflammation to induce EMT in cell lines derived from carcinoma has been reviewed extensively. The cytokines tumor necrosis factor alpha
(TNFα), the transforming growth factor beta (TGFβ), interleukin 1 beta (IL1β), and the interleukin 6 (IL6) converge in the activation of the transcription factor NF-κB 92-94, which constant stimulation upregulates the expression of Snail, Slug, Twist, Zeb and the Forkhead Box C2 (FOXC2). In turn, the mesenchymal transcription factors repress the locus Ink4b/ARF/Ink4a and the ETS transcription factors 95-98. The detection of biomarkers of EMT and inflammation correlates with poor clinical outcomes, and the connection of the mesenchymal proteins in surpassing senescence in vivo is supported by the inverse correlation of the biomarkers in carcinomas 99. It can be said that all the characteristics to became a cancer cell are enabled through a single process of transdifferentiation 100. Cells subjected to EMT acquire metastatic capacity, resistance to apoptosis, immune evasion, stem cell qualities, and resistance to anti-cancer therapies. Furthermore, the process of EMT can be induced also by changes in the extracellular matrix (ECM), such as mechanical forces, matrix metalloproteinases (MMPs), and cytoskeletal rearrangements, hypoxia, pH, chemotherapy; doxorubicin, gemcitabine, fluorouracil, bevacizumab, nivolumab and radiotherapy 101. The epithelial tissue affected with infiltrated hyperplasia possesses favorable conditions for the induction of the EMT in senescent cells from which emerge cells primed for carcinogenesis. The same order of cellular transitions is observed in the spontaneous immortalization of epithelial cells, and suggest a principle, a generic pattern that may be recreated during epithelial carcinogenesis in vivo.

The hypothesis considers that tissues exposed to cell proliferation eventually become enriched with senescent cells. The hyperplastic lesions will be infiltrated by immune cells that may induce EMT of senescent cells in vivo. The result is a cell that emerges with the ability to induce carcinomas. During the spontaneous immortalization of normal cells in vitro, the same series of cellular transitions is experienced by epithelial cells that arise from senescence with markers of basal lineage such as vimentin adopting a fibroblast morphology.

3.2. The order of cell immortalization
The study of primary cultures of normal human mammary epithelial cells has shown that cells can overcome senescence since the tenth doubling time. Those cells lose the expression of p16 and beta-galactosidase and resume proliferation. The overexpression of the proteins p53, p21, and the presence of chromosomal abnormalities are common findings. They include translocations, deletions, rearrangements, telomeric associations, polyplody, and aneuploidy 80. Those reports also describe the gradual loss of epithelial markers since the 15-30 doubling time and the transformation of the luminal phenotype (epithelial) to a fibroblast morphology (basal or myoepithelial) with vimentin expression. The spontaneous immortalization over the long-term passages is understood as the consequence of genetic damage that result in the loss of p16, and telomerase reactivation 102. Regardless of the cause, this phenomenon implies an especially illustrative connection between cellular senescence with EMT and supports the temporal order of cellular transitions suggested by the hypothesis (Fig. 2.). Thus, the spontaneous immortalization of epithelial cells in vitro may parallel the histological progression of carcinomas in vivo. The association of aging and inflammation with the increased chances for carcinoma development could be reduced to cellular transitions. However, as
outlined above the immortalization of normal epithelial cells, and the abrogation of cellular senescence by inflammatory cytokines culminates with the emergence of cells morphological and transcriptional more related to mesenchymal stem cells.

This hypothetical origin of carcinomas is incompatible with the notion that cancers from the epithelial tissue must correspond with epithelial cells that acquire malignant traits by mutations. Furthermore, the cancers that arise from mesenchymal tissues are sarcomas. To address these conceptual inconsistencies is necessary an overview of the staging and prognosis of carcinoma. The following demonstrates that the hypothesis is more compatible with the facts, although still controversial under current assumptions. For example, the five-year relative survival rate of most carcinomas depends on the stage at diagnosis. In the TNM staging system, this means a global stratification based on the size of the primary tumor, the affected lymph nodes, and the presence of metastases. In the histological staging, the cancerous tissue is classified according to the degree of differentiation. Low-grade carcinomas show characteristics of normal tissue and tend to grow slowly. In contrast, high-grade cancers are poorly differentiated, lose the characteristics of normal epithelium, tend to grow faster, and respond poorly to therapy. Histologic grade is a prognostic factor for overall survival despite any TNM status. Essentially, in both systems, the probability of patients surviving more than five years exceeds ninety percent unless they already present metastasis or high grade at diagnosis.

A notorious event during the progression towards a carcinoma of high grade is the inversion in the ratio epithelium to the stroma. In other words, the content of mesenchymal cells and the expression of their markers increase, such as vimentin, the alpha smooth muscle actin (α-SMA), N-cadherin, cadherin-11, and secreted protein acidic and rich in cysteine (SPARC) outnumebering the epithelial proteins. Additionally, the proportion of stromal content is an independent factor of poor prognosis. Overall, it might be considered that both, the immortalization of normal epithelial cells in vitro and the lethality of carcinomas in vivo requires the adoption of mesenchymal traits.

3.3. The unavoidable fibroblasts contamination

Fibroblasts are considered the main effectors of fibrosis in normal or pathological conditions. The conventional view of fibrosis during epithelial carcinogenesis depicts that cancer-associated fibroblasts are cells programmed for the tumor to foster its progression. These cells are the most abundant entity in the carcinoma microenvironment and compelling evidence demonstrated their abilities to promote all the so-called hallmarks of cancer. In contrast to normal fibroblasts, they possess an increased proliferation rate and autocrine signaling. Over the years the presumed source of the cancer-associated fibroblasts has been evolving with observations that indicate an origin in the resident fibroblasts, pericytes, bone marrow, and more recently from epithelial cancer cells that undergo EMT in vivo.

The investigation of the formation of fibrosis during inflammatory conditions clearly showed that normal epithelial cells can assume a mesenchymal phenotype and contribute to the generation of connective tissue associated with stromal proliferation and with scar formation. With the arrival of more sophisticated techniques, it was possible to gather conclusive evidence that not only does tumor epithelial cells in vivo give rise to cancer-associated fibroblasts but also that cells emerge with the expression profile of cancer stem cells. The mesenchymal impurity of the carcinomas is an endless anomaly in the study of epithelial cancers. The establishment of epithelial cell lines for the study of carcinogenesis has historically faced the phenomenon of the emergence of fibroblasts along with the loss of the proliferative potential of epithelial cells. Even current attempts for the ex vivo evaluation of the chemosensitivity of tumors deals with the contamination of the samples by mesenchymal cells. Interestingly, the fibroblasts are more resistant to drugs than epithelial cells derived from the same tumor. Despite the findings, at best, the fibroblasts remain relegated as secondary players in epithelial carcinogenesis.

If the hypothesis is correct, the low-grade epithelial tumors are benign tissues in their way of transformation. This implies that the real carcinoma begins once a certain threshold of restrictions is surpassed and the events
of transdifferentiation and the stabilization of the mesenchymal phenotype are favored. This stage would correspond with the high-grade carcinomas. These assumptions also explain the large difference in survival between the well-differentiated tumors versus the high-grade carcinomas. Therefore, it is theorized that carcinomas are lethal once the mesenchymal phenotype is reached and stabilized. The previous stages are part of the necessary process for the in vivo reprogramming. The proposal that the origin of carcinoma lies in the mesenchymal cells that arise from senescence in epithelial hyperplasia can solve another recognized paradox. In the presence of TGFβ, the normal epithelial cells arrest their cell cycle. Thus, this cytokine was considered a potential effective anticancer agent in the management of carcinomas. The studies showed that it restrains the growth in early-stage tumors, however, an opposite effect resulted in advanced carcinomas. It was logical to justify the apparent dual activity to the loss of the regulation in the TGFβ pathway during carcinoma progression, in which tumor cells shifted the signal into proliferation and invasion. According to the hypothesis it is expected that the TGFβ presents the ability to foster tumor progression in advanced stages if carcinomas are mesenchymal driven diseases. Observations from the immortalization of normal epithelial cells indicate that the ability to sustain growth in presence of TGFβ develops along with the adoption of a fibroblast-like morphology. This is also consistent with the carcinoma cell lines, in which the ability to induce tumorigenesis is reserved for the cells with fibroblastic morphology. It is also not surprising that the proliferation of mesenchymal stem cells can be stimulated by TGFβ signaling. The unavoidable progression of carcinomas towards independence from ligands explains the temporal responses observed using hormonal agents or targeted therapy. The process is naturally understandable if the mesenchymal undifferentiated cells are considered the phenotype responsible for carcinomas. Furthermore, is unnecessary to invoke complicated mutational molecular mechanisms to understand the process of progression as a whole, since the induction of cellular transitions allows a discrete change in the cellular phenotype along with the shift into autocrine signals, the lack of anoikis, and the dedifferentiation of the epithelium. Additionally, the mesenchymal cells that arise from senescence in epithelial hyperplasia emerge with stemness and the ability to induce cancer, but more importantly, they explain the cellular plasticity at play in the advanced stages of carcinoma. Based on the hypothesis, is next provided an attempt to understand the underlying order in the middle of complexity at play during the histological progression of carcinomas.

4. The origin of carcinoma

4.1. More than mesenchymal
The process of EMT was temporally viewed as a mechanism reactivated by tumor cells to acquire an invasive phenotype. However, an increased number of experiments found that EMT also explains the cancer stem cell phenotype, the resistance to chemotherapy, the immune system evasion, and the ability to induce tumors and their heterogeneity. Consistent with the hypothesis, mesenchymal cells that emerge from cellular transitions are not simple fibroblasts, the empirical evidence confirms that EMT generates cells with the expression of the set of pluripotent genes, such as, SRY-Box Transcription Factor 2 (SOX2), the homeobox pluripotency transcription factor (Nanog), the octamer-binding transcription factor 4 (Oct4), the RNA-binding protein that regulates mRNA translation in embryonic stem cells (Lin28B), and the neurogenic locus notch homolog protein 1 (Notch1). Despite being a small proportion of the tumor, the cancer stem cells are considered the main culprits of tumor relapse after cancer therapy. Their functional characterization to seed new tumors have been reviewed by the analysis of expression of cell surface markers, the formation of mammospheres, and using the xenotransplantation limiting dilution assay. The induction of EMT by inflammatory cytokines also generates cancer stem cells, and carcinomas show a correlation among the biomarkers of the process of inflammation, EMT, and stemness. The increased levels of proinflammatory cytokines in vivo has been linked with the induction of EMT and the accumulation of cancer stem cells in tumors. Notwithstanding extensive empirical evidence indicating that the EMT might be the source of most aspects relevant for cancer, its contribution to in vivo carcinogenesis remains at debate
Since high-grade carcinomas are not a homogenous mass of mesenchymal stem cells and the metastasis manifest an epithelial phenotype, to support the hypothesis it is necessary to explain the generation of heterogeneity and metastasis. In the case of tumor heterogeneity is almost self-explanatory since the process of EMT originates fibroblasts, cancer stem cells, vascular endothelial cells, and pericytes \( ^{109} \) (reviewed in section 6). On the other hand, the change in microenvironment enables the MET of tumor cells and offers a compelling origin for the epithelial phenotype of metastasis. In the hormone-independent breast cancer cells MDA-MB-231, which present \textit{in vitro} a mesenchymal phenotype, the metastatic lesions that generate in mice express the protein E-cadherin and display an epithelial phenotype \( ^{127, 128} \). Furthermore, the process of MET was triggered by the interaction of cells with the parenchyma of the lungs and bones. At the molecular level, the reversion of EMT involves the negative regulation of the mesenchymal transcription factors product of the downstream signals of the microenvironment while the epithelial proteins are activated \( ^{129, 130} \). As is now recognized, the induction of MET \textit{in vitro} can reprogram fibroblasts into induced pluripotent stem cells (iPSCs) that showed the expression of the transcription factors Oct4, the zinc finger-containing transcription factor Kriippel-like factor 4 (KLF4) along with the repression of the Snail \( ^{131} \).

In general, the induction of epigenetic reprogramming \textit{in vivo} as a result of the histological changes in the hyperplasia suggests a plausible sequence of events for the reactivation of developmental programs that may account for most observations during carcinogenesis. According to the hypothesis, at least two cellular transitions are required for the generation of cancerous cells, however, the tumoral progression towards high-grade carcinoma would need conditions in the tissue that foster the constant generation and maintenance of the dedifferentiated cell states.

### 4.2. The Tissue Matters

The accumulation of senescent cells over the years compromises the renewal, structure, and function of the tissues. Eventually, the burden of senescent cells in the hyperplasia would reach a threshold of inflammation in which the cellular transitions towards the mesenchymal stem phenotype are attained. However, the maintenance of the cancerous behavior and the increased rate of cellular transitions would need a permissive microenvironment for carcinogenesis. This stage, perhaps, requires an extra input of proinflammatory cytokines along with the irreversible loss of the extracellular matrix and cell-cell interactions. Hence, the effects of senescence and inflammation in tissue architecture may explain the increased susceptibility to carcinogenesis of the aged epithelial tissues.

The contribution of senescent cells to structural disruption include the secretion of epithelial growth factors and proinflammatory cytokines that enhance the mitosis of epithelial cells and its eventual entry to senescence \( ^{132, 133} \). The monocyte chemoattractant protein 1 (MCP1) \( ^{134} \) is also liberated by senescent cells and upregulates the migration and infiltration of monocytes, lymphocytes, and NK cells \( ^{135} \). According to the hypothesis, is possible that macrophage activation plays a major role in the progression of hyperplasia since its transformation is consistently associated with their infiltration \( ^{136} \). Macrophages are essential in normal development and repair via tissue remodeling. Unlike the common view that tumor-associated macrophages are a special type of cells reprogramed to sustain inflammation and tumor growth \( ^{137} \), here is implied that the positive feedback loop that emerges between senescent cells and the activated macrophages synergize for the pathological transformation of the tissue in part by their production of MMPs and the degradation of the structural constraints that normally impose a regulated cell behavior. The substrates of the MMPs include many proteins in the extracellular matrix, proteinase inhibitors, cell surface receptors, and cell-cell adhesion molecules \( ^{138} \). Both cell types express several MMPs that allow the cell detachment from the extracellular matrix, the basal membrane, and cells \( ^{134} \). In this regard, clear evidence of the capacity of mechanical forces to induce pluripotent stem cells has been presented \( ^{139} \). The cell phenotype is coupled to cellular morphology and stromal interactions through the signaling pathways that sense the mechanical stress of the cytoskeleton by the focal adhesion kinase (FAK) that activates the transcription factor Yes-associated protein (YAP) and the transcriptional co-activator with PDZ-binding motif (TAZ). These in turn are translocated to the nucleus and
promote the transcription of genes associated with cell growth, resistance to apoptosis, and carcinogenesis. Furthermore, the activation of the transcription factors YAP and TAZ is sufficient to induce cancer stem cells. Currently, it is considered that tumors also reprogram normal fibroblast into cancer-associated fibroblast since the only detected differences in them are the epigenetic pattern of methylation. The shift is associated with a constant activation of NF-κB and the secretion of TGFβ, IL6, MCP1, the vascular endothelial growth factor (VEGF), and MMPs. These secretions act autocrine, paracrine, and modify the organization and the physical properties of the tissue microenvironment to promote fibroblast proliferation, the infiltration and activation of macrophages and inflammation, the induction of EMT, metastasis, angiogenesis, and the enrichment of cancer stem cells in the tumor microenvironment. On contrary, according to the hypothesis, the cancer-associated fibroblasts correspond mostly with cells that arise from EMT, either from inflammation or as the result of the stromal disruption, and would explain the gradual loss of senescent cells and the increased stromal content during the progression to high-grade carcinoma. This stage implies the acquisition of tumor autonomy and perhaps marks the irreversible arrow of carcinoma progression towards dedifferentiation.

Overall, it is suggested that the first step in the origin of carcinomas involves the combination of an increased number of senescent cells with the secretory phenotype, chronic inflammation, and mechanical disruption in the hyperplasia. Then, its progression is allowed by the sustained levels of inflammation, the irreversible loss of normal tissue architecture, and the amplification of the mesenchymal phenotype. The higher levels of proinflammatory cytokines combined with relatively few possibilities of interactions with epithelial cells, and the normal stroma that otherwise would prevent the cancerous phenotype results in the inexorable progression to high-grade invasive carcinoma. In these tumoral conditions, the mesenchymal and the undifferentiated cell phenotypes are preferentially sustained. However, the interaction of these cells with microenvironment clues from the stroma, cells, parenchyma, and the oscillation of the endocrine milieu, such as the concentration of oxygen, nutrients, and growth factors or the reduction in the levels of inflammation explain the tumor heterogeneity and the formation of metastasis. On the other hand, it is likely that some dedifferentiated cells that emerge within the tumor migrate and may differentiate into functional cells in healthy tissues, and most importantly; the possibility that the underlying reason for the targeting of senescent cells by the immune system, is the attempt to regenerate the tissue. In this sense, the origin of carcinoma is the pathological consequence of a normal endogenous process of tissue regeneration.

5. Inflammation and cellular plasticity

5.1. Epithelial regeneration by cellular transitions

The notion of cancer as the result of overhealing is by no means new to pathology. As a result of the hypothesis, a more encompassing view of epithelial carcinogenesis emerges and suggests an endogenous mechanism by which the immune system undertakes the maintenance and rejuvenation of tissues by the epigenetic reprogramming of damaged cells. Inflammation has been linked to the pathophysiology of carcinomas and with almost every hallmark of the cancer cell. Tumor tissues exhibit high levels of expression of cytokines compared to normal samples, and their presence is also positively correlated with advanced tumor grade. Elevated serum levels of inflammation are associated with adverse prognosis an increased number of metastases. The subsequent discovery that cytokines can induce EMT and pluripotency in cancer cells renewed the interest in the relationship between inflammation and cancer from a different perspective. However, it is important to notice that normal epithelial cells exposed to interleukins experience the same effects. This implies an endogenous mechanism by which inflammation induces cellular plasticity. In such a scheme, the outcome of the epigenetic reprogramming in vivo of senescent cells might result in a recovery of the proliferative potential along with normal epithelial phenotype. In the youth epithelia, the presence of stromal and cell interactions
ensures the normal differentiation of the reprogrammed cells and the outcome is the clearance of the secretions of senescent cells and the resolution of inflammation. Circumstantial evidence is emerging regarding the role of senescence, EMT, and inflammation in the regeneration of tissues in vivo using epigenetic reprogramming. It turned out that the secretory phenotype is essential for tissue repair, fibrosis reduction, and wound resolution. Furthermore, the locus Ink4b/ARF/Ink4a and the production of IL6 were crucial for the induction of EMT and the expression of the transcription factors cellular Myc (c-Myc), Oct4, Sox2, Klf4 during cell reprogramming and wound regeneration in vivo.

The results can be interpreted as a mechanism of tissue renewal triggered by the senescent cells and inflammation to induce epigenetic plasticity for the recovery of injured cells. In this scenario, the senescent cells in the hyperplasia are cells with DNA damage that became inflamed, activate NF-xB, and release proinflammatory cytokines and chemokines. The secretory phenotype attracts and activates the immune cells that generate higher levels of tissue disruption and production of bioactive mediators. These situations would promote the EMT in senescent cells since intrinsically are prone to reprogramming due to epigenetic derepression. This event would allow access to an endogenous cell state with the abilities of self-repair, then, the interactions of the dedifferentiated cell state with the normal tissue enable the proper instructions for the attainment of the cell fate. Hence, stemness is transient, and the inflammation is resolved in the absence of DDR. On the other hand, the combination of senescent cells with extended molecular damage and aberrant structural microenvironment would sustain the generation of mesenchymal cells and the properties of cancer stem cells. The lack of resolution of this attempt of healing would become the unavoidable histological progression towards the lethal high-grade carcinoma. Hence, the origin of sporadic carcinomas is assumed as the consequence of the endogenous process of healing in aged and inflamed tissues. The argument presented above is based on common observations in carcinomas although from a different insight. In the next section it is compared with the prevalent paradigm, alternative proposals, and then is provided a unified view of epithelial carcinogenesis.

5.2. Cellular plasticity unifies the models of carcinogenesis
There are several features inherent to cancer, such as the genetic and chromatin damage, the disruption of tissue during the formation of tumors and metastasis, the multistep nature of carcinogenesis, an association with aging and inflammation, and the remarkable similitude with the process of wound healing and embryo development. Accordingly, the models that have been proposed in the history of cancer research adopted those observations into their premises. Through the years the dominant paradigms evolve, but essentially the new findings were customized into their original assumptions. The following provides a brief description of each, its premises, and their interpretations of the evidence.

The somatic mutation theory posits that the accumulation of mutations in the genetic material of epithelial cells is favored as a function of time and the increased levels of inflammation. The effect of telomere attrition, chemical exposure, infections, or radiation would be additive in the accumulation of mutations and the chances of acquisition of malignant traits. However, most of the malignant features arguably conferred by oncogenes and tumor suppressors play essential roles in the normal biology of cells, the random events of mutations and Darwinian selection hardly explain the transcriptional convergence of high-grade cancers, and carcinogenesis does not require mutagenesis. Additionally, the reversion of the malignant behavior and the acquisition of the cancer stem phenotype through events of transdifferentiation challenges entirely the logic of this theory. The whole cancer phenomenon is attributed to a series of fortuitous mutagenic events in different cell phenotypes which then are associated with a fixed purpose in carcinogenesis. Hence, it demands the creation of at least the cancer version of the normal stem cells, macrophages, fibroblasts, or the pathological variants of senescence, EMT, and inflammation. As a result of the inconsistencies, this paradigm is moving towards epigenetic mechanisms to sustain most of its original premises.
The importance of tissue architecture in cancer development was neglected by the somatic mutation theory although adapted to some extent by considering the effects of tumoral cells under its microenvironment\(^{150}\). The theory of the field of tissue organization addresses this, being the stroma the primary target of carcinogens. The tumorigenesis of epithelium is explained by considering that tissue disruption releases the inherent proliferative and motile behavior of normal cells. Hence, provides an explanation for the induction of differentiation when tumors are exposed to the normal stroma, as well as the opposite effect of the acquisition of malignant behavior of normal cells in response to abnormal stroma\(^{151}\). Although offers no integral explanation for the process of carcinogenesis, exposes the limitations of the somatic mutation theory, and offer some relevant premises, for example, that tissue integrity throughout aging diminishes, that mutations are unrelated to causality, that cancer is a developmental process, and the possibility of tumoral reversion.

On the other hand, the notion of cancer as a deregulated process of healing takes into account the association of chronic inflammation, fibrosis, and tissue disruption with carcinogenesis\(^{152}\). Wound healing shares many features with tumorigenesis, such as the presence of activated immune cells, cell migration, angiogenesis, fibroblast, myofibroblast, and cell proliferation, the generation of stroma, the induction of EMT, and the production of mediators for tissue remodeling. Despite the suggestions that hyperplasia may transform by overhealing, and that tissue regeneration and carcinogenesis are linked at the molecular level, findings regarding this posture remained mostly descriptive.

The view of cancer as a developmental disease highlights that embryo and tumor development are parallel processes since both originate from a single cell that undergoes accelerated proliferation and migration using the same mechanisms. Since the nineteenth century, pathologists suggest that tumors are a deregulated process of development, and using modern molecular techniques was confirmed the reactivation of the mechanisms normally associated with embryo morphogenesis\(^{16, 153, 154}\). However, the identification of the EMT, MET in the generation of cancer stem cells and metastasis, and the embryonic transcriptional profile of high-grade carcinomas was mainly relegated as the result of the promoting effects of the tumor microenvironment over the mutated cancerous cells.

Regarding the position of cancer as an embryonic or primitive preexistent cell-state, the view of cancer as an atavistic state posits that in case of adverse conditions, cells reanimate an ancestral genetic program that was suppressed during evolution. This cellular state would correspond with functions associated with survival as a unicellular form. Hence, mutations may reactivate an endogenous behavior already primed for carcinogenesis\(^{155}\). Likewise, the proposal of cancer attractors suggests that cancer is the result of cellular states closely related to embryonic programs, but unable to differentiate. In this context, mutations or microenvironment perturbations would allow access to the cancerous phenotype\(^{156}\). Both perspectives agree on the undifferentiated nature of the cancer cell, and that carcinogens would promote the activation of an endogenous cellular state.

The main hypotheses of carcinogenesis have been integrated or overlapped in various degrees through the years. Invariably, most of them coincide that the event of genesis is associated with chromosome instability or genetic damage. The importance of mutations was greatly influenced by the study of the mutagenic effects of viruses or chemicals that induce cancer. Hence, cancers should probably originate from mutations in genes of DNA repair. These ideas were then incorporated in the two-hit model, which established the need for a second key mutation for tumor progression especially in genes that control cell proliferation\(^{149}\). That model prevails and is used to understand the multi-step nature of carcinogenesis. In other words, that most solid cancers are preceded by focal proliferative lesions, in the case of carcinomas the hyperplasia.

The different positions can be contextualized in a unified view of epithelial carcinogenesis. Telomere attrition due to replicative exhaustion justifies the presence of DNA damage in cells that originate the tumor. Instead of considering that the cancerous phenotype is created by mutations, selective silencing, or activation of genes, is proposed that this event generates unspecific transcriptional derepression that in the first instance lead to a discrete change in the cellular phenotype from which emerge the senescent cell. Chromosomal anomalies accumulate in senescent cells, which number increase in aged tissues and raise the level of immune
cell infiltration due to the secretory phenotype. Those events explain the long relationship of aging and inflammation with the susceptibility of carcinoma development. Moreover, imply the multistep process of carcinogenesis since the hyperplasia provides the conditions for the first hit necessary for the transformation, but still is insufficient for the invasive and lethal behavior. Then, the higher burden of senescent cells and the infiltration of immune cells contribute to the structural changes in the stroma and the loss of cell interactions that allow deregulated in vivo reprogramming that essentially drives progression. This stage would correspond in a sense, with the second hit, the disappearance of senescent biomarkers, and the rise of proteins associated with inflammation, EMT, and cancer stem cells. In this setting, the proposed series of cellular transitions and the progression towards dedifferentiated carcinoma partially relate with some of the premises of the theory of the field of tissue organization and with the view of cancer as a developmental disease that is the consequence of endogenous programs and undifferentiated phenotypes.

The increased transcriptional derepression of somatic cells in response to endogenous molecular damage or extracellular interactions results in the emergence of a mesenchymal stem-like phenotype that would drive the genesis and progression of the sporadic carcinomas. In the setting of the high-grade lesions or large-sized tumors, the disruption of the tissue and the high levels of inflammation are enough to stabilize the cancerous phenotype and explain the lack of differentiation and the autonomy that epitomizes the advanced carcinoma. The hypothesis of the origin of carcinomas presented in this paper might be considered a model since is rooted in observations and proposes mechanisms. In the context of this model, carcinomas can be collapsed for simplicity into a series of cellular transitions in which the original cellular phenotype, the magnitude, and type of perturbation govern the level of cellular plasticity (Fig. 3.).

![Fig. 3. Inflammation and cellular plasticity.](image)

1) Studies in vitro have shown that cytokines induce mitosis of competent epithelial cells until they enter replicative cell senescence. 2) Cells can escape cell arrest and acquire fibroblast morphology using EMT. 3) Cells undergoing EMT can achieve the stem cell phenotype if they continue exposed to high concentrations of proinflammatory cytokines. 4) Exposure to high levels of inflammation can induce EMT in normal epithelial cells and result in the emergence of mesenchymal stem cells. 5) The epigenetic reprogramming of epithelial cells into induced stem cells is possible with inflammation. 6) Stromal interactions induce several grades of differentiation and explain tumor heterogeneity, 7) the formation of metastasis by induction of MET, 8) and the maintenance and healing of aged tissues. In the main text is provided empirical evidence of each cellular transition.
According to the hypothesis, the main driver in the discrete switch in phenotype associated with the cellular transitions underlying carcinogenesis is epigenetic regulation. In line with the premises of the theory, growing evidence points out that the transcription factor NF-κB integrates the stimulus from proinflammatory cytokines and the structural change in tissues or the chromatin damage with the players of the epigenetic regulation.

5.3. NF-κB links inflammation with cellular plasticity

Cancer cells are characterized by extensive epigenetic changes compared to their normal counterparts. Epigenetics broadly refers to mechanisms that mediate the differential access to the genetic material by the modification of the chromatin structure, namely, stable changes in gene expression that are independent of the nucleotide sequence. Transcription is regulated by the degree of compaction between histones and genes through the covalent modification of DNA or in the nucleosome and ATP-dependent chromatin remodeling. Hence, the epigenetic marks create several genetic profiles from the same genetic code that govern the singularity of each cell phenotype. The temporal regulation of the gene expression by the epigenetic process is central for the appropriated cell fate commitment and differentiation 157. The epigenetic control mediated by the enzymatic alteration of the structure of the chromatin is one of the most studied mechanisms in the network of transcriptional regulation. The activity of DNA methyltransferases (DNMTs) results in transcriptional repression due to chromatin compaction by the methylation of cytosines. The proteins that modify the histones include methyltransferases (HMTs) and acetyltransferases (HATs). Histone methylation occurs in lysine and arginine, whereas acetylation, on serine and threonine residues. The polycomb proteins also mediate gene silencing by the formation of the polycomb repressive complexes (PRC) PRC1 and PRC2 that change the configuration of the chromatin and prevent the transcription. In the PRC2 the histone lysine methyltransferase EZH2, acts together with the zinc finger protein SUZ12 (suppressor of zeste 12 homologue) to catalyze the trimethylation on K27 histone H3 (H3K27me3), which is then recognized by PRC1, with members such as the proto-oncogene Polycomb Ring Finger (Bmi1) protein that maintains chromatin silencing. Is considered that the more stable form of epigenetic repression and to maintain the cell identity and differentiation is the silencing of genes by H3K27me3 and PRCs. However, the epigenetic marks can be reversed by the activities of DNA demethylases (DNMDs), such as the Ten Eleven Translocation enzyme (TET), the Lysine-Specific Demethylase 1 (LSD1), and the Jumonji C-terminal domain (JmjC), both histone lysine demethylases (HDMs), and the histone deacetylases (HDACs), such as HDAC1 and HDAC2, and Trithorax proteins (TRXG) that counteract the repression by PRCs 157.

The molecular mechanisms necessary to explain the events of transdifferentiation begin to be understood. Interestingly, increasing reports suggest that NF-κB activation can erase cell identity through the induction of enzymes that reshape the chromatin and enables the epigenetic reprogramming that manifests as cellular plasticity that underlies carcinogenesis according to this manuscript.

Despite the activities of NF-κB are usually viewed as a process exclusive to the immune system or with tumor progression, it is ubiquitously expressed, and its inhibition is lethal to any cell type. NF-κB suppresses the pathways of cell death in normal and cancer cells and is essential in the regulation of normal epithelial homeostasis 158. Since the NF-κB pathway is induced in response to changes in the environment, including cytokines, radiation, oxidative stress, hormones, or growth factors, and by intracellular damage is comprehensible its hyperactivation in cancer 159. The NF-κB transcription factor family has five members: the transcription factor p65 subunit (RelA), the cellular homolog of v-Rel (c-Rel), the V-Rel Avian Reticuloendotheliosis Viral Oncogene Homolog B (Rel B), the DNA binding subunit of the NF-κB (p50), and the NF-κB p52 subunit (p52) which form various homo and heterodimers. In the resting state, NF-κB dimers are in the cytoplasm in an inactive form, through their binding to inhibitory proteins known as inhibitors of κB (IκBa, IκBβ, and IκBε). The majority of stimulus that leads to NF-κB activation converge on the IκB kinase (IKK) complex, which is responsible for IκB phosphorylation and allow the nuclear translocation of
NF-κB. Its transcriptional activities increase the expression of more than 500 genes, among them, the chemokines, lymphokines, interferons, acute phase proteins, and their receptors, the enzymes cyclooxygenase 2 (COX-2), lipoxygenase 5 (LOX-5), and the nitric oxide synthase (NOS). The antiapoptotic proteins such as the B-cell lymphoma 2 (Bcl-2), the Bcl extra-large (Bcl-xL), the Bcl-2-like protein 11 (Bim), the cellular FLICE-inhibitory protein (c-FLIP), the X-linked inhibitor of apoptosis protein (XIAP), the membrane proteins ATP-binding cassette transporters, the P-glycoproteins, the growth factors FGF8, the hepatocyte growth factor (HGF), the bone morphogenetic protein 2 (BMP-2), BMP-4, the nerve growth factor (NGF), VEGF, the placenta growth factor (PIGF), the epidermal growth factor receptor (EGF) and the human epidermal growth factor receptor 2 (HER2). The transcription factors c-myc, the homeodomain transcription factors (HOXs), the proto-oncogene AP-1 Transcription Factor Subunit (JunB), p53, the E2F transcription factor 3 (E2F3a), Twist, the E74 like ETS transcription factor 3 (Elf3), and the chromatin modifiers Bmi1 and jmjd3. Traditionally, the mechanism that underlies the regulation of distinct sets of genes by NF-κB is attributed to its different dimers and the association with several families of transcription factors specific to each cell phenotype 160.

The activation of the NF-κB signaling pathway in carcinoma cell lines is linked with demethylation that results in the upregulation of genes normally expressed in pluripotent embryonic stem cells 161. Furthermore, the epigenetic reprogramming with the cytokines TGFβ, TNFa, IL1, and IL6 increases the expression of the markers CD44, CD133, Nanog, Bmi1, Oct4, and vimentin 162, 163. The molecular regulatory networks that coordinate the events of transdifferentiation such as the EMT and the generation of cancer stem cells by inflammatory cytokines involve the activation of Snail, Twist, Zeb, and epigenetic regulators by NF-κB 164. For example, the induction of Jmjd3 depends on the direct binding of NF-κB to a cluster of three kB sites in its promoter. Then, Jmjd3 promotes site-specific H3K27me3 histone demethylation leading to transcriptional derepression and cellular plasticity 165. The process of EMT involves the repression mediated by the PRCs of epithelial genes through the initial recruitment of Snail, which binds to the E-box elements of the promoters. Snail recruits PRC2 and interacts with EZH2 and SUZ12 to catalyze the H3K27me3 and silencing of gene transcription by PRC1. Snail also mediates the recruitment of LSD1 to target genes that can trigger EMT and cancer progression, in conjunction with additional epigenetic modifications. The formation of LSD1-Snail complexes on gene promoters catalyzes the removal of methyl groups from the H3K4me3 activation mark leading to the loss of transcriptional activation of epithelial genes. LSD1 is highly expressed in several cancer types and correlates with poor survival 164. During EMT, Twist directly activates Bmi1 expression and confer the properties of cancer stem cells. The axis of NF-κB, Twist, and Bmi1 is essential for the repression of the epithelial phenotype, the Ink4b/ARF/Ink4a locus, and to sustain the mesenchymal stem cell transcriptional signature 166. Snail and Twist can repress gene activity through deacetylation of gene promoters by recruiting HDAC1 and HDAC2, which catalyze the removal of acetyl groups from lysine 9 and lysine 14 residues of histone H3. Snail also promote the proteosomal degradation of p53 by HDAC1 deacetylation and recruit the HMTs such as the euchromatic histone-lysine n-methyltransferase 2 (EHMT2) and the suppressor of variegation 3-9 homolog 1 (SUVR3H1), which act cooperatively to catalyze the trimethylation of H3K9 that is required for the recruitment of DNMTs. In turn, leads to the methylation of cytosine and guanine linked by phosphate in a repeated sequence (CpG islands) in gene promoters that block the transcriptional activity by the histone lysine methyltransferases 164. This mechanism was also found during the EMT induced by the TGFβ signaling to repress the epithelial genes. In mammary epithelial cells, TGFβ also showed the induction of Jmjd3, the removal of H3K27me3 marks, and the reactivation of the mesenchymal genes 167. The activation of NF-κB by IL6 triggers an autocrine loop sufficient for the epigenetic switch from immortalized breast cells to cancer stem cells 168. IL6 can change the pattern of DNA methylation in cancer cells by inducing the expression of DNMTs. The IL6 pathway is required for the CpG island methylation in promoters to repress tumor suppressors such as p53, and the epigenetic reprogramming to drive cancer cells towards the stem phenotype 164.
The mechanical disruption of tissues and the disorganization of cells also activate inflammation by coupling the signals of the integrin and Rac kinases with the NF-κB pathway through the activation of the IKK complex \[169\]. Further, the comparison of the breast cancer cells HMT-3522 that grow in clusters versus a variant of disorganized cells show the differential overexpression of 180 genes that share binding sites for NF-κB. Additionally, the suppression of the transcriptional activities of NF-κB restores the formation of clusters and the suppression of motility. The culture of HMT-3522 for 4 days in contact with an artificial ECM and the presence of an inhibitor of IKK results in the reversion of the malignant phenotype and the formation of structures with polarity and integrin expression \[170\]. Related findings showed that breast cancer cells depend on the sets of genes activated by NF-κB to sustain the basal phenotype and the invasiveness \[171\]. Hence NF-κB governs the malignant cell phenotype but is a reversible process in the presence of normal stromal signals and downregulation of inflammation.

The induction of inflammation also comes from intracellular inputs. For example, following irradiation or chemotherapeutic drugs that induce DNA breaks, the ATM kinase is activated and phosphorylates H2AX, leading to the recruitment of several other proteins involved in the DDR. The activation of ATM phosphorylates the NF-κB essential modulator (NEMO), which in turn, induces NF-κB activity \[171\]. Furthermore, the hormone-independent phenotype of breast cancer cells requires the activation of ATM in response to H2AX foci in the chromatin. The potential protumorigenic consequence of a constantly activated DDR has been observed in the mammary gland, in which the induction of the cytokine IL6 activates NF-κB and ignite a positive inflammatory signaling loop, that perpetuate the proliferative signaling and the acquisition of mesenchymal stem traits \[171\].

Overall, mutations appear inadequate to explain the complex behavior and cell heterogeneity observed during epithelial carcinogenesis. Some attempts to justify the acquisition of elaborated instructions for malignancy propose as the source the chromosome abnormalities such as breakages, fusions, and aneuploidy \[172\]. Aneuploidy can be induced by local microenvironmental changes, confers increased survival, and is associated with inflammation and metastasis \[173\]. Seems logical that large genomic rearrangements are more effective in inducing adaptation than single-gene mutations. Among the multiple factors that have been proposed to induce chromosomal instability and aneuploidy are telomere dysfunction, stress, and DNA damage that also increased during the histological progression of solid tumors. However, increasing evidence support that the activity of the chromatin modifiers underlies chromosomal instability and aneuploidy since neither aneuploidy nor mutations are exclusive of cancer cells. Thousands of mutations occur in differentiated cells of healthy individuals and appear to be essential in the physiology of normal cells. Regarding aneuploidy, although is associated with cellular senescence and telomere attrition, it also temporarily increases in response to a normal regenerative stimulus. For example, the number of aneuploides in hepatocytes rise in function of proliferation to restore the liver mass in vivo but is reestablished to normal \[174\]-\[176\].

Here, is proposed that the prominent presence of chromosomal aberrations in cancer is the result of chromatin rearrangements that lead to large-scale genetic changes that modify the expression of many genes at once and allow events of transdifferentiation according to environmental and intracellular inputs. The secretory phenotype of the senescence cells is ruled by NF-κB activation in response to telomere attrition since is sensed as DDR. In turn, intracellular inflammation in senescent cells within the hyperplasia loss transcriptional repression that makes them prone to epigenetic reprogramming. At the level of tissue, the change in morphology of senescent cells, the immune infiltration, and the combined secretions of cytokines and MMPs disrupt the interactions with the extracellular matrix and among cells that otherwise would restrict the cellular plasticity. The gradual overthrow of the constrains at both levels result in the hyperactivation of the NF-κB pathway and increased epigenetic plasticity that fuels the progression to high-grade carcinoma in which the mesenchymal stem states are self-sustained, amplified, and intrinsically motile. The selective
evolutive advantage of linking the signal of adversity with transcriptional derepression is clear since cellular dedifferentiation would provide the necessary mechanisms for self-renewal and survival. However, in aged tissues, the chances for carcinogenesis clearly overthrow the process of healing. Hence, the irreversible nature of carcinoma progression is towards dedifferentiation and metastasis. In line with the premises adopted in this paper, the endogenous genetic damage and the aberrant tumor microenvironment lead to transcriptional derepression mediated by cellular inflammation. NF-κB recruits and interacts with epigenetic regulators creating feedback molecular loops that regulate cellular plasticity and link inflammation with epigenetic reprogramming. The coupling of the extracellular and intracellular perturbations to epigenetic reprogramming by the degree of activity of NF-κB and the magnitude of plasticity results in the dynamic and reversible nature of cellular transitions that have been challenging the attempt to conceptualize a fixed purpose for each of the processes or phenotypes involved in carcinogenesis. Furthermore, explains the elusive nature of EMT, MET, and the cancer stem cells during the histological progression of the carcinomas in vivo.

6. The enigma of myofibroblasts

6.1. The emergence of myofibroblast

The origin and the mechanisms that regulate the emergence, clearance, or persistence of myofibroblasts have been elusive. Myofibroblasts exert traction forces in the extracellular matrix and synthesize its components during connective tissue remodeling and epithelialization in wound healing and disappear afterward. However, in organ fibrosis, these cells are persistent and predict its progression to carcinoma. The proliferation of myofibroblasts is also present in tumor desmoplasia and is associated with poor prognosis 177.

The hypothesis predicts that cells in transit of the second cellular transition in vivo would exhibit a temporal hybrid phenotype between senescent and the mesenchymal cells. Interestingly, the stromal cells that correspond with this profile are the myofibroblasts that display the markers of replicative senescence and mesenchymal stem cells. Consistently with the proposal presented in this manuscript, myofibroblasts are not only involved in fibrosis and cancer but also well known for their transient appearance in the process of tissue homeostasis and wound healing. According to the hypothesis, the phenomenon of myofibroblasts is related to aging and inflammation of the parenchyma, in which mostly the resident epithelial cells undergo senescence and initiate the process of healing. However, in conditions of severe tissue disruption, inflammation, or catastrophic chromosomal damage the events of dedifferentiation are persistent. In such conditions, the myofibroblasts are robustly produced from several cell types and novel discoveries in the field of fibrosis unveil its intrinsic plasticity and its protumorigenic potential.

Cells that have been involved in the proliferation of the stroma are the resident fibroblastic and mesenchymal stem cells along with cells from the bone marrow and perivascularulature. Using more recent technology, epithelial cells showed to account for most of the fibroblastic cells during organ fibrosis and in the desmoplasia associated with carcinoma 178. Hence, the process of EMT became the most studied cellular transition in cancer biology and fibrotic diseases associated with inflammation. However, the cellular plasticity endowed by inflammation not only underlies the process of transdifferentiation of epithelial cells into fibroblasts but also in endothelial cells, adipocytes, and pericytes. These cells can transdifferentiate by EMT in cells that can behave as fibroblasts, myofibroblasts, or mesenchymal stem cells. In the case of epithelial cells, the cuboidal shape along with the expression of E-cadherin and zonula occludens-1 (ZO-1) is lost and turn into a spindle-shaped cell that expresses the fibroblast-specific protein (FSP1) or the α-SMA in the case of the myofibroblast 178. Interestingly, the fibroblast detected in neoplastic lesions and chronic inflammatory conditions also express the markers of senescence 179. These cells are also present in the injured tissues limiting the accumulation of fibrosis and facilitated their resolution and healing. According to the expression of α-SMA these cells correspond with myofibroblasts, that exhibit cellular plasticity and multipotency 180. In line with the continuum of cellular transitions proposed in this paper, those events
correspond with the transition of senescent cells by EMT into mesenchymal stem cells with self-renewal and multipotency, hence, that intrinsically harbors properties for healing or carcinogenesis.

The endothelial cells, multipotent monocytes, fibrocytes, pericytes, adipocytes, and local fibroblasts, also dedifferentiate into myofibroblasts in response to external or internal stimuli. Increasing studies show that the adoption of the fibroblastic program confers the cells with the intrinsic potential to be activated for differentiation into subtypes of fibroblast-like cells and several non-mesenchymal lineages. Hence, the current view of fibroblasts is as not terminally differentiated cell types, that exhibit heterogeneity in response to differences in the endocrine milieu, the neighboring cell phenotypes, mechanical forces, and its position in the tissue. The ability to sense the changes in the ECM, the autocrine and paracrine signals influences the epigenetic regulation that translates into transcriptional changes and may explain the plasticity of fibroblasts.

The induction of EMT and the maintenance of the fibroblastic phenotype in adult tissues is associated with the stimulation of soluble growth factors, cytokines, and the influence of the extracellular matrix. In response to those events, the resident epithelial cells and fibroblasts can dedifferentiate into myofibroblasts. During fibrosis, the ligands of the Wnt, Notch, and Sonic hedgehog pathways, such as the TGFβ, BMP, EGF, HGF, and FGF are commonly involved in the regulation of myofibroblast differentiation. The onset of fibrosis has been associated with changes in the mechanical forces of the tissue that activate the fibroblasts and the production of TGFβ that in turn induces the proliferation of myofibroblasts and the accumulation of collagen.

In carcinomas, the differentiation of fibroblast to myofibroblast and its proliferation is also regulated by TGFβ. Regardless of their origin, myofibroblasts display a malignant phenotype resistant to apoptosis. In other chronic inflammatory diseases such as rheumatoid arthritis, the fibroblasts that proliferate in the affected joints also resemble immature malignant cells. They behave differently from normal fibroblasts, shown invasive potential and properties of mesenchymal stem cells. Furthermore, it has been suggested that resident stromal cells undergo dedifferentiation in vivo by the conditions of hypoxia and proinflammatory cytokines in the affected joints.

6.2. The multipotency of fibroblasts

The fibroblasts represent the heterogeneous mesenchymal cells which are extensively found in conjunction with almost every cell type as the primary system of interstitial support. Classically, is considered that the stromal connective tissue found in all organs is derived from mesothelial tissue and that the function of the fibroblasts is mainly to synthesize the ECM including the collagen, proteoglycans, fibronectin, laminins, glycosaminoglycans, and in the formation of scar tissue. However, current evidence supports their active role in restoring the function of tissues by transdifferentiation into several cell types.

Vimentin is the major structural component of the intermediate filaments used for fibroblast identification, which functions include contractility, migration, and proliferation. Other than vimentin and morphology, the biomarkers that precisely define the fibroblasts remain to be fully understood. The set of transcription factors that seem essential in the maintenance of the fibroblastic phenotype are the Paired related homeobox 1 (Prrx1), the paired-related homeobox protein SHOT (Shox2), the AP-1 Transcription Factor Subunit (c-fos), Slug, and Twist1. However, during fibrogenesis, the mesenchymal cells acquire traits of embryonic cells and become highly proliferative. Furthermore, the origin of myofibroblasts is also tightly linked to upregulation of genes expressed during early embryonic development. In this regard, the consensus is that mesenchymal stem cells and fibroblasts can be distinguished by the differential expression of genes associated with developmental programs and the stromal functions. Mesenchymal stem cells show ubiquitous activation of the germline programs, with the activity of transcription factors of ectoderm, endoderm, and mesoderm. The transcription factors that differentiate them from normal fibroblasts are the expression of the ETS variant transcription factor 1 (ETV1), the ETV5, the forkhead box protein P1 (FOXP1), the GATA-
binding factor 6 (GATA6), the high mobility group AT-Hook 2 (HMGA2), KLF12, the PR-domain zinc finger protein 16 (PRDM16), the single-minded family BHLH transcription factor 2 (SIM2) and SOX11 191. Moreover, the mesenchymal stem cells also possess higher grades of hypomethylation 180 and increased expression of the pluripotent marker Oct4 compared to fibroblast 192. Therefore, seems logical to employ fibroblasts to obtain mesenchymal stem cells. A task that has been successfully accomplished in vitro with the transcription factors SOX2, OCT4, KLF4, and c-MYC 193, or using a mixture of chemical inhibitors and growth factors 192. However, fibroblasts are highly plastic by endogenous mechanisms. Their multipotency was first described in studies that show its chondrogenic differentiation in response to mechanical stimulation. Consistently, fibroblasts show transcriptional diversity and heterogeneity relative to the mechanical forces that experience due to structural changes, position, and the endocrine milieu in different tissues 182. Furthermore, they share features that were considered exclusive of the mesenchymal stem cells. For example, they express the surface markers CD90, CD105, CD166, and upon appropriate conditions differentiate into adipocytes, chondrocytes, osteocytes, hepatocytes, neurons, myocytes, or pancreacytes 194,195. Hence, both are administered as a form of cell therapy for stroke, heart failure, COPD, liver damage, osteogenesis imperfecta, Duchenne, Muscular Dystrophy, and cosmetic procedures 183. Due to their inherent immunomodulatory properties are also used in inflammatory and autoimmune diseases and to fasten recovery 196. Their injection into affected tissues is followed by differentiation to functional cells 197. However, several studies show that mesenchymal stem cells also stimulate tumor development 198 and form teratomas in vivo 193, raising concerns about the safety of this type of therapy.

Fibroblasts are also the most used cells to obtain iPSCs. In turn, these cells can be cultured to produce induced mesenchymal stem cells (iMSC) using MSC medium, inhibitors of the kappaB kinase epsilon, the TGF-β/Activin type I receptor, BMP4, or growing the cells in precoated surfaces with gelatin, collagen, or synthetic polymers 199. Not surprisingly, an endogenous mechanism that induces the spontaneous transformation of fibroblasts is telomere attrition due to replicative senescence. As previously reviewed, the senescent program is essentially the same despite the original phenotype, and fibroblasts also remain in arrest while undergoing changes in morphology, gene expression, metabolism, and epigenetics. The early molecular events also involve the recruitment of proteins activated by DDR 200 and the induction of NF-kB 201. The careful examination of the effects of cell passage in fibroblasts founds a gradual decrease of methylation, differentiation, and the emergence of mesenchymal stem traits and tumor behavior. Fibroblasts undergoing cellular senescence arise with the molecular, structural, and contractile features of myofibroblasts. Additionally, the fibroblasts obtained from carcinomas are usually senescent and positive for the expression of the myofibroblast marker α-SMA 179, 180. Seems plausible that fibroblast dedifferentiation into mesenchymal stem cells with tumor behavior in vivo is driving by epigenetic rearrangements associated with cellular aging, inflammation, and structural cues. According to the hypothesis, in the setting of progression or the advanced stages of the disease, is expected that several cell types, including cells in the stromal compartment or derived from the bone marrow, can dedifferentiate to myofibroblasts with cancerous behavior by the same series of cellular transitions.

6.3. The EMT is linked to embryonic programs
The epithelial cells are the precursors of the endoderm and mesoderm using developmental pathways. During gastrulation, the epithelial cells acquire plasticity by EMT and adopt the fibroblastic phenotype and motility for normal embryo development. The activation of the mesenchymal transcription factors is mediated by the TGFβ, Wnt, and Notch pathways and their proper functioning avoid death during gastrulation or defects in morphogenesis 202. The coupling of EMT with an embryonic transcriptome involves the machinery of epigenetic remodeling. The overexpression of Snail, Zeb, or Twist on normal, immortalized, or cancerous epithelial cells result in the activation of Bmi1, stemness, and tumor-initiating capacity. The process of EMT in vivo is followed by Bmi1 induction and the enrichment of stem markers in carcinomas. Bmi1 overexpression represses the effectors of cellular senescence and increases self-renewal and multipotency in
adult and embryonic stem cells. During EMT induced by inflammation, NF-κB activates the mesenchymal transcriptional factors and the enzymes of epigenetic remodeling. The activation of Twist directly binds the Bmi1 promoter and confer cancer stem cell properties, represses the epithelial phenotype, the Ink4b/ARF/Ink4a locus, and preserves the mesenchymal stem phenotype 203.

According to the hypothesis, cellular plasticity is the result of epigenetic derepression, in which the original cell type and the magnitude of the stimulus are the main factors that limit the degree of cellular reprogramming. It is likely that cellular plasticity is increasing as the physiological levels of cytokines rise and the restrictions imposed by the stroma diminishes. Hence, carcinogenesis in vivo is self-organized and the tangible evidence is the typical histological progression of carcinomas. Essentially, at the core of the advanced carcinomas is deregulated cellular plasticity due to inflammation and the result is the stemness. Tumors and metastasis are the natural consequence of the subsequent partial differentiation of cancer cells in response to environmental changes that restrain chromatin derepression. A great body of current literature supports a bigger role of physical forces, tissue homeostasis, cellular aging, and inflammation on epigenetics, cell behavior, and plasticity that may ultimately result in carcinogenesis.

Consistent with the model, studies have shown that maintenance of the fibroblastic shape, the malignancy, and stemness in vitro requires high levels of proinflammatory cytokines that upon decrease, induce the reversal into an epithelioid morphology 187. The mesenchymal stem cells also become heterogeneous due to spontaneous differentiation or maturation associated with the production of ECM 204, 205. The experimentally silencing of Sox2 205, Twist1, or Slug decreases Bmi1 overexpression, abolishes stemness, the invasiveness, and enables the differentiation towards a luminal phenotype 203. Hence, the connection of inflammation, transcriptional derepression, and stemness is likely the molecular basis of the cellular plasticity driving carcinogenesis.

Recent evidence shows that full malignancy is associated with cells that exhibit a hybrid phenotype. Their multipotency is enough to explain the tumor growth and cell heterogeneity in vivo 206. The finding of the expression of markers from several lineages in the hybrid malignant cells is expected considering the current view of the molecular biology of stem cells. Furthermore, imply a more comprehensive and unified view of cancer as the result of development.

7. Cancer and development

7.1. Transcriptional derepression and stemness

The study of stem cells has revealed that they do not express a specific set of genes or signaling pathways to sustain their multipotency. Instead, it is suggested that stemness is a transient trait, characterized by having many potentials but no specialization. The transcriptional hyperactivity is considered the main responsible for the plasticity of the stem cells and implies that is a cell state produced as the result of repressing differentiation 207. Furthermore, the analysis of embryonic stem cells showed the overexpression of genes related to chromatin-remodeling, transcription and that differentiation correlates with a reduction of transcriptional activity 208. Consistently, the activity of the transcription factors Oct4, Nanog, Sox2, Klf4, and c-Myc collectively occupy 10% of the promoters in the human genome and repress genes implicated in differentiation. Moreover, the PcG, DNA methyltransferases, and stem transcription factors are associated with the upregulation of transcription in stem cells 209. Another interesting feature is the structural coupling of the cytoskeleton with the nucleus of stem cells that allow the sense of mechanical signals from the cell microenvironment and translate them into transcriptional multipotency 210. In general, the emergence of stem cells and their maintenance is strongly influenced by environmental factors and the dynamics of cell growth, inducing heterogeneity in cell-cell interactions, cytokines, oxygen, growth factors, geometry, and mechanical
forces\textsuperscript{211, 212}. Some niches induce stemness \textit{in vivo} by paracrine effects and mechanical interactions. Hence, the cell microenvironment plays a major role in stemness, and these cellular states cannot be defined in isolation. Conceptually, niches of stem cells can be viewed as domains in which a group of cells and their structural and endocrine interactions result in stemness. This is also consistent with their apparent elusive presence, its nature as a cell state that requires an induction, and its particular spatial location in tissues. Through the comparison of different stem cell systems, some traits suggest generic characteristics of the relationship between stem cells and their supporting niche \textit{in vivo}. It is proposed that the stemness is inducing by a combination of proximity to the basal lamina and other cell types that activate the JAK-STAT, TGFβ, and Wnt-signaling\textsuperscript{213}. The cultures of stem cells also indicate that stemness is a property induced by unspecific stressors. Among the conditions that induce and sustain stemness \textit{in vitro} include hypoxia\textsuperscript{214}, oxidative stress\textsuperscript{215}, radiation\textsuperscript{216}, chemotherapy agents\textsuperscript{217}, DNA damage\textsuperscript{218}, ECM degradation with proteases, and the four pluripotency factors (Oct4, c-Myc, KIf4, and Sox2). Stem cells need to grow in serum-free culture media enriched with growth factors such as the BMPs, TGFβ, TNFα, and under hypoxic conditions, otherwise, they start to differentiate\textsuperscript{204}. In response to adverse non-lethal conditions, any cell can undergo stemness that enables access into survival mechanisms and process of renewal that are normally silenced during differentiation to optimally perform highly specific tasks associated with the different cell phenotypes that are necessary for multicellularity. Hence, seem that stemness is a transient robust cell state that can be accessed for self-preservation, but once the conditions are suitable for survival, cells can differentiate according to environmental cues. Furthermore, the normal biology of the stem cells is implicated in every aspect of the so-called hallmarks of cancer (Fig. 4.).

Transcriptional derepression reconfigures several cellular processes largely associated with carcinogenesis. The metabolism of stem cells is characterized by the conversion of glucose to lactate despite the presence of enough oxygen to sustain the complete oxidation of glucose-derived carbons. These cells are heavily reliant on glucose and glutamine to proliferate\textsuperscript{219, 220}. Transcription factors that characterize stem cells downregulate p16 and p53 proteins preventing apoptosis\textsuperscript{221, 222}. Stem cells possess an increased mechanism of DNA repair, detoxifying enzymes, and overexpression of ABC transporters\textsuperscript{223}. In stem cells, telomerase is activated and maintains telomere length, and confers cellular immortality\textsuperscript{224}. Pluripotent stem cells showed chromosomal instability in long term cultures. Additionally, stem cells showed to acquire aneuploidy reminiscent of malignant cell transformation\textsuperscript{225}. Stem cells constitutively activate NF-κB (cellular inflammation) to sustain an undifferentiated cell phenotype\textsuperscript{162}. Stem cells lack anoikis and have an inherent ability to migrate, which is as important as their capacity for self-renewal and to differentiate, enabling them to maintain tissue homeostasis and mediate repair and regeneration\textsuperscript{226}. Additionally, stem cells acquire mechanisms for immune evasion\textsuperscript{227}.  

\begin{figure}[h]
\centering
\includegraphics[width=0.5\textwidth]{stemness.png}
\caption{Tumorigenic capabilities conferred by stemness.}
\end{figure}
The comparison of cancer stem cells and embryonic stem cells reveals an absence of differences at transcriptional 117, 228, 229, and functional level 117. Therefore, it has been suggested that the only difference between them is the microenvironment 230. Is tempting to speculate that in parallel to normal stem niches, the epithelial tissues during carcinogenesis create aberrant compartments that foster transcriptional derepression and a deregulated access to stemness.

Cells that undergo senescence experience changes in chromatin, nucleus, and PcG proteins associated with transcriptional derepression. It can be argued that senescent cells are a step closer to stemness. Those features might underlie the reported closer epigenetic relationship between the senescent and cancer cells 86. According to the fundamental thesis in this paper, entry to senescence and tissue disruption during aging, in essence, make cells prone to dedifferentiation. Cellular aging reshapes the epigenetic landscape and links inflammation with regeneration and cancer. The possible outcomes are already codified in the genome and govern the entire course of carcinogenesis implying that the degeneration of tissues that underlies chronic diseases are part of the developmental process.

7.2. An unexpected, unified view of chronic diseases

Embryos originate from a cell that undergoes accelerated proliferation and migration through processes that parallels carcinogenesis. Long before the genomic era, the use of a simple microscope was enough to propose that cancer may be the product of an aberrant developmental process. In 1859, the pathologist Rudolf Virchow suggested that tumors originate on the same principles that operate during embryonic development. Later on, in the 1960s the ectopic implantation of normal embryos resulted in teratocarcinoma 231 and the injection of these cells into the blastocyst induced their differentiation in normal tissues 12. The cells extracted from these tumors were incorporated into functional tissues and expressed genes associated with differentiated phenotypes in healthy mice. In contrast, the cells that were kept in vitro were unable to express the sets of genes observed in vivo. Overall, those experiments exhibit an interesting facet of cancer; in which embryonic stem cells generate neoplasms in absence of mutagenesis and that carcinogenesis can be reversed by differentiation. Unfortunately, those results were regarded as rarities exclusives of teratomas and is likely that most modern molecular biologists are unaware. Furthermore, few researchers are even concerned about the inconsistencies in the current cancer field and rarely question their conceptual framework. Despite the historical delay, the late twentieth century witnessed the resurgence of evidence indicating that embryonic programs are involved in almost every aspect associated with cancer 154. Furthermore, an increasing number of studies in carcinomas correlate the infiltration of immune cells in vivo with the enrichment of embryonic stem cell markers and progression to higher histological grades 232. A remarkable finding was the transcriptional convergence into embryonic stem cell programs of high-grade tumors irrespective of the germinal cell layer or original tissue 16. Additionally, stemness has been envisioned as the non-specific emergent product of the loss of the transcriptional repression 207. Altogether, the evidence is consistent with the notion presented in this paper that carcinogenesis is a generic process in which somatic cells flow through cellular transitions towards dedifferentiation and stemness.

During early embryo development, when the embryonic totipotent cells reach the eight cell-stage, the first differentiated cell type that appears is the epithelial, and just a few divisions afterward arise the second cell type, the mesenchyme 33. During embryogenesis, senescence act as a transient form of plasticity in which cells eventually resume proliferation and contribute to the morphogenesis of tissues 233. The process is broadly similar to the premises of the model, and suggests, that aging foster its reiteration by replicative senescence and chronic inflammation that may result in cancers. The robust emergence of the same basic cell phenotypes and the directionality of the observed cellular transitions suggests a genetic program that imposes the same functional and structural constraints to govern both, embryo development and epithelial carcinogenesis. Importantly, there is no reason to limit the argument to cancer since the emergence of most chronic diseases seems to follow the same order of cellular transitions.
Several tissues gradually undergo fibrosis with aging, including the skin, lung, liver, pancreas, and heart. Progressive fibrosis is a major cause of morbidity and mortality associated with repeated epithelial injuries and accumulation of myofibroblasts. The process of fibrosis is caused by the hyperproliferation of fibroblasts and the production of abnormal amounts of EMC that replace the normal structure of the parenchyma. The result is the functional impairment of organs and sometimes persistent fibrosis. The therapeutic options for organ fibrosis are handicapped by an incomplete understanding of the molecular mechanisms. Likewise, despite the current multitarget approaches to cope with cardiovascular diseases, diabetes mellitus, and cancers, they remain the principal causes of morbidity and mortality among adults in industrialized countries.

The model presented for the origin of carcinomas provides a mechanistic explanation of the underlying causes of fibrosis and steatosis accompanying the degeneration, the loss of function, and in some cases the transformation of tissues during aging from an evolutionary and molecular perspective. First, it is assumed that cellular senescence precedes the onset of most chronic diseases. Second, that inflammation and infiltration in the affected tissues accelerate their degeneration by increasing their susceptibility to cellular transitions. Finally, the cellular transition into mesenchymal phenotypes underlies fibrosis and steatosis. Being developmental biology the study of the mechanisms that govern the development of organisms from fertilization to senescence; epithelial carcinogenesis and chronic diseases seems to be part of the same processes that determine the loss of functions associated with aging. From an evolutionary perspective, the involution of the parenchyma makes sense since epithelial tissues are highly dependent on ligands produced by other specialized cells and are fine-tuned by structural interactions. In contrast, cells that undergo transdifferentiation into fibroblasts are more robust to adverse conditions. Thus, carcinogenesis could be viewed as an extreme consequence of the same conserved process that originates the rest of the chronic diseases, and the mesenchymal enrichment of tissues embraced as the logical consequence of the systemic conditions in aged patients.

The role of cells that undergo EMT or similar transitions during the fibrosis of vessels, hearth, liver, kidney, and pancreas is now increasingly recognized as an essential process for tissue degeneration in disease and aging. The hypothesis implies that terminally differentiated cells can transdifferentiate into fibroblasts after senescence and explain the myofibroblasts expressing beta-galactosidase in the fibrosis of aged tissues. Thus, unifying the process of aging and chronic diseases under the same theoretical framework proposed for carcinogenesis. In essence, the hypothesis suggests that the involution of tissues is mediated by dedifferentiation and transdifferentiation in vivo. Furthermore, implies that the organized deterioration of tissues is preceded and the onset of chronic diseases and their manifestations are byproducts of the process. Consistent with this view, even when fibrosis is inevitable, the likelihood of tumorigenesis decreases by the downregulation of inflammation. Hence, the dynamics of aging and the associated diseases are ruled by development and its interplay with lifestyle. Overall, the cellular plasticity gained in aged tissues, their functional deterioration, and the eventual death is mainly the result of the unidirectional nature of DNA replication. Telomere length plays a major role in the possible number of cell divisions before the onset of a DDR response that induces cellular inflammation, transcriptional derepression, and ultimately cellular plasticity.

Regarding leukemias, its onset is preceding by the myelodysplastic syndrome that is characterized by cells with chromosome alterations and biomarkers of senescence. Furthermore, the development of myeloproliferative disorders is associated with bone marrow fibrosis, and the leukemogens disrupt its stroma. Upon evolution from chronic to acute leukemia, the chromosomal abnormalities increase, and the markers of cell dedifferentiation diminish. In the blast crisis, which is the final stage of the disease, the cells become unresponsive to therapy and present stem cell gene signatures. The overall process is then remarkably similar with the premises of this manuscript and suggest a principle. Although the details provided were oversimplified, and the molecular biology of each chronic disease on its own can be extremely complex; it emerges an underlying and valuable simplicity. If the theory presented in this paper is correct, some strategies of cancer prevention and therapy should be reassessed.
8. Reshaping cancer prevention and therapy

8.1. Cancer and lifestyle

The contribution of genetic factors in cancer development represents only 5% of cases, while the rest is attributed to environmental factors. Therefore, sporadic cancers are considered lifestyle diseases where most cases are associated with unhealthy habits or infections. Consequently, the policy for cancer prevention focuses on reducing the use of tobacco and alcohol, avoid obesity, the proper management of infections, and dietary advice. Several investigations have addressed the potential mechanisms of lifestyle on cancer development. In this contribution, current knowledge was integrated to illustrate the global effect of lifestyle in epithelial carcinogenesis according to the hypothesis.

Diet is one of the most important factors for cancer development but also one of the most complex to understand. Even on a conceptual level, the term healthy diet is poorly defined. Nonetheless, several lines of evidence support that a diet based on fruits, vegetables, and whole grains is protective against cancer even in the presence of risk factors and reduces overall mortality among survivors. Importantly, the protective effect of a diet with an emphasis on plants extends to the rest of the chronic diseases. The concept of dietary pattern aid to define the type of diet associated with the protective effect in epidemiological studies. In the case of cancer, a healthy dietary pattern refers to a diet high in fruits, vegetables, poultry, fish, whole grains, and a low daily fat intake. In the opposite case, a diet with a high content of red meat, processed meats, refined sugars, sweet foods, and a high fat intake is considered unhealthy.

The effect of foods in blood biomarkers shows the integrated changes induced by dietary patterns in physiology and their potential impact on carcinogenesis. A healthy dietary pattern induces low levels of insulin, insulin-like growth factor (IGF-1), estrogen, testosterone, and proinflammatory cytokines such as TNFα, IL6, and C-reactive protein (CRP). In turn, chronic exposure to low levels of these ligands results in low activation of the insulin receptor (IR), the mTOR protein (mammalian Target of Rapamycin), the estrogen receptor (ER), the androgen receptor (AR), the leptin receptor (Ob-R), and cytokine receptors, as well as their signaling pathways. Genetic, pharmacological, or dietary manipulation of these pathways increases longevity and delays the onset of all chronic diseases, including cancer. Healthy diets, being rich in plants, are also characterized by a low content of calories and fat, they are rich in fiber, therefore, they have a low glycemic index and are considered incomplete sources of amino acids. Dietary interventions that involve those factors increase lifespan through mechanisms conserved in evolution. Examples include the caloric restriction, protein, or amino acid restriction (such as asparagine, glutamate, or methionine), or the ingestion of complex carbohydrates. The antiaging effects of these dietary strategies result in the negative modulation of the insulin / IGF-1 signaling pathway and that of mTOR, which are the most important regulators of lifespan in humans.

Healthy diets are also rich in bioactive phytochemicals such as terpenes and phenolic compounds capable of exerting direct effects on the extension of longevity by modulating insulin / IGF-1 signaling pathways, mTOR, activated protein kinase by adenine monophosphate (AMPK), NF-κB, the NAD-dependent protein sirtuin deacetylase (SIRT1), and the sex hormone receptors. The high fiber content exerts indirect effects through the proliferation of commensal bacteria that reduce systemic inflammation through their interactions with the mucosa-associated lymphoid tissue and the generation of regulatory T cells. Healthy diets are also associated with high consumption of monounsaturated and polyunsaturated fatty acids that, when metabolized, generate resolvins and other anti-inflammatory ligands of the receptor peroxisome proliferator activated gamma (PPAR-γ), Toll-like receptors (TLRs), G protein-coupled receptor 120 (GPR120), COXs, and LOXs with antagonism in the activation pathways of NF-κB. The ingestion of diets rich in complex carbohydrates and polyunsaturated fatty acids decreases the levels of insulin, IGF-1, estrogen, testosterone, and proinflammatory cytokines such as TNFα, IL6, and CRP. In addition, the low circulating levels of
Biomarkers of inflammation are associated with a delay in the onset of all chronic diseases. Adherence to healthy diets shows lower levels of biomarkers of cellular aging, hyperplasias, prevents EMT, and delays tumor progression. Overall, healthy dietary patterns modulate epithelial carcinogenesis by slowing the rate of cell growth, proliferation, metabolism, and systemic inflammation. In doing so, they comprehensively delay aging, the accumulation of senescent cells, tissue infiltration, and inflammatory levels, resulting in a low probability of cellular transitions.

In contrast, obesity and unhealthy diets promote higher circulating levels of proinflammatory cytokines, sex hormones, insulin, and growth factors. An unhealthy lifestyle is associated with obesity and the early onset of chronic diseases. Obesity and western diets promote accelerated aging. Moreover, obesogenic diets promote the accumulation of senescent cells, induce EMT and tumor progression. In the case of infections, it is proposed that the etiological agents age and induce inflammation in some specific tissues since the process is mediated by receptors. Therefore, only the affected tissues increase cellular turnover and immune responses because of infection. Thus, the human papillomavirus, Helicobacter pylori, and schistosomiasis generate carcinogenesis of the cervical, gastric, and bladder epithelia, respectively. In the case of smoking and alcoholism, a similar event occurs, since due to the location some tissues are more exposed, or due to their activities in processes of detoxification are also highly affected. This fundamentally explains the preferential carcinogenesis of the lungs and liver respectively, and the association of both with oropharyngeal cancer.

In this theoretical framework, the effect of lifestyle on carcinogenesis converges in the modulation of aging and inflammation. It proposes a mechanistic and integrative overview of the most important regulators of the process considering the premises of the model. In conclusion, seem that a healthy lifestyle delays cellular aging and downregulates systemic inflammation resulting in cancer prevention, but ultimately it increases lifespan by slowing the process of development. In the following section, this type of view is transferred to cancer therapy in an effort to explain its failure along with an alternative that arises from the hypothesis.
8.2. Cytotoxic therapy promotes cancer.

Surgery is the central component of the curative treatment for localized solid tumors.\(^{269}\) Cancer in situ is curable using surgical resection as the only therapeutic strategy.\(^{5-8}\) Regarding early-stage invasive carcinomas, the contribution of surgery to the relative survival rate at 10 years surpasses 90%.\(^{270}\) Therefore, several studies suggest that most of the beneficial effect of cancer therapy is attributed to the removal of tumor burden.\(^{269,271}\) Although the 5-year relative survival rate of patients diagnosed and treated with low-grade carcinomas is greater than 90%, they eventually progress to high-grade carcinomas and die from causes associated with cancer.\(^{37,272}\) The invasive metastatic cancers are responsible for 90% of cancer mortality.\(^{5-8}\) and the only therapeutic option at this stage, due to relapses or tumor progression during treatment is palliative systemic therapy. In other words, surgery is not part of the treatment, only chemotherapy, and radiation. The use of cytotoxic therapies at advanced stages for palliative purposes or to increase survival is at a debate since high-grade tumors and metastases do not respond. However, the use of aggressive antineoplastic treatments is frequent in terminal patients despite the lack of therapeutic response. The net result is an increased cost and a reduction in the quality of life without a clear benefit for patient survival.\(^{273}\) In contrast, a growing number of studies indicate that surgical resection of the primary tumor and larger metastases improve survival even in patients with high-grade metastatic cancers.\(^{269,274}\) This suggests that most of the benefits of cancer treatment are derived almost exclusively from the removal of the cancerous tissues.

Historically, the therapeutic failure in cancer has been attributed to late diagnosis. In other words, tumors are detected when they already acquired most of the molecular mechanisms necessary to resist the treatment. However, studies have shown that resistance, metastasis, and aggressive behavior arise relatively quickly.\(^{275}\) Lack of tumor response to treatment may occur from the first therapeutic regimen or after a temporal response.\(^{276}\) Therapy is usually a combination of cytotoxic drugs and radiation to avoid the chances of developing resistance. However, they fail to eradicate cancer cells, and patients often relapse within months to a year after treatment. Furthermore, recurrences generally manifest a greater degree of malignancy and do not respond to therapy.\(^{277,278}\) Tumor progression in the presence of therapy is traditionally understood by Darwinian mechanisms of positive selection mediated by the acquisition or modification of genetic traits. The hypothesis of cancer stem cells was recently added to explain treatment failure and tumor recurrence. This position considers that a small subset of cells in the tumor possess this cellular phenotype, the ability to self-renew and differentiate into tumor cells. Hence, they would be inherently resistant to cancer therapy and responsible for tumor recurrence.\(^{279}\) Like many other phenomena in cancer, the origin of cancer stem cells and their role in carcinogenesis is not fully accepted. However, the last decade has validated the ability of chemotherapy and radiotherapy to induce stem cells within tumors.\(^{2,280,282}\)

Most of the antineoplastics drugs, including the alkylating agents, antimetabolites, antimitotics, antibiotics, and epipodophyllotoxins are carcinogenic since they induce malignant tumors and secondary cancers.\(^{7,283}\) The generation of neoplasms or the progression towards aggressive behavior has also been documented for radiotherapy.\(^{216,284}\) and targeted therapies such as bevacizumab\(^ {285}\) and imatinib\(^ {286}\). Most of the tumors induced by cytotoxic therapy are acute myeloid leukemias, followed by solid tumors of the breast, bone, lung, prostate, ovary, bladder, and brain.\(^ {7,283,287}\) The use of neoadjuvant chemotherapy (applied before surgery) also seems to promote tumor aggressiveness and recurrence negatively affecting survival.\(^ {288}\) One of the responses of the tumor to therapy is an increased transcriptional signature of embryonic stem cells.\(^ {118,216}\) In this regard, recent evidence indicates a relationship between senescence, EMT, and increased expression of embryonic genes with events triggered by antitumor strategies.\(^ {118,289,290}\)

The hypothesis of epithelial carcinogenesis as the product of cellular transitions promoted by cellular aging and inflammation suggests a mechanism by which the current treatment is not only ineffective but accelerates cancer progression. Molecular responses to the non-lethal adverse conditions imposed by treatment promote...
transcriptional derepression, inflammation and enable the emergence of undifferentiated cell states endogenously resistant and associated with a negative prognosis. Under this view, current cytotoxic therapies are doomed to failure in their attempts to cure cancer. According to the postulates of the hypothesis, the detrimental effects of cytotoxic therapy are mediated by the increase in the systemic load of senescent cells that translates into inflammation and accelerated cellular aging. Consequently, the EMT and events of dedifferentiation are promoted, increasing organ fibrosis and the emergence of cells with malignant and metastatic behavior that are stabilized in inflammatory conditions.

Current cytotoxic therapy leads to systemic events that promote accelerated tumor progression. Most patients exhibit transient pancytopenia caused by extensive damage to the bone marrow components 291, 292. These histological changes provide the basis for the generation of leukemias in response to cytotoxic therapy. In the first phase, the decrease in tumor burden and proinflammatory cytokines generate the partial or complete, but always temporal responses associated with cancer therapy. However, in a second phase, the continuation of the therapeutic regimen increases cell damage, senescence, dysbiosis, fibrosis, and the functional loss of many tissues. The cytokines produced by all damaged somatic cells, would overcome the initial reduction that manifest as cachexia. From here, the systemic conditions would only favor accelerated progression.

The antineoplastic drugs cyclophosphamide, fluorouracil, cisplatin, methotrexate, adriamycin, and etoposide induce cachexia 293, 294. The drivers of cachexia are the cytokines TNFα, IL6, IL1, CRP, and TGFβ. The elevated levels of these cytokines generate insulin and IGF-1 resistance, which hinders the metabolism of glucose and amino acids in the skeletal muscle while favoring lipolysis in adipose tissue and the oxidation of fatty acids. The metabolic state is reminiscent of fasting but in feeding conditions. For this reason, the syndrome of cachexia is characterized by the wasting of the skeletal muscle and adipose tissue despite the presence of an adequate caloric intake. As cachexia progresses, cancer patients deplete the reserves in adipose and muscle tissue, generating weakness, motor difficulty, and cardiopulmonary dysfunction 295. Cachexia is found in 80% of late-stage cancer patients, frequently along with metastasis and unresponsive tumors. Furthermore, those patients usually cannot tolerate the side effects of cytotoxic therapy. Their involuntary weight loss is accompanied by immunosuppression, lack of wound repair, fibrosis, and systemic dysfunction, which decrease their quality of life and negatively affect the prognosis. It has been estimated that cachexia is responsible for 40% of cancer death 296. As expected, the use of parenteral nutrition or diets with calorie surplus is ineffective against the effects of cachexia. In the advanced stages, the secretions of the tumor masses are sufficient to generate cachexia. Proinflammatory cytokines reprogram the metabolism of the hosts for the mobilization of nutrients towards the tumor masses. Overall, those inflammatory and metabolic mechanisms increase tumor progression at expense of the host 295.

The physiological conditions by which tumor nutrition is favored remarkably resemble embryonic development. During the second half of gestation, the maternal reserves are mobilized to allow the fetal exponential growth. The production of glucose, amino acids, and fatty acids in the bloodstream is increased due to insulin resistance, gluconeogenesis, proteolysis, and lipolysis. These effects are mediated by insulin, IGFs, glucagon, catecholamines, cytokines, and leptin. Insulin resistance in the mother ensures proper sparing of nutrients and energy to the fetus to fulfill the anabolic pathways since its metabolism is relatively independent of the activity of insulin. Also, the fetus overexpresses proteins related to the transport and metabolism of nutrients, among which glucose, lactate, ketocids, amino acids, and fatty acids stand out. The anaerobic metabolism of these substrates is the main source of energy and growth 297. All of the mechanism described has been considered the metabolic characteristics of tumors 298. However, the reactivation of developmental events seems to naturally explain the metabolic process responsible for cancer cachexia.

Given the above arguments, the direct effect of cytotoxic therapy might result in a net increase in cellular transitions towards a mesenchymal stem cell phenotype. At the same time, therapy induces cachexia, which
indirectly favors the maintenance of the dedifferentiated phenotypes and their metabolic and migratory activities. Meanwhile, the nutritional and inflammatory status of the host is at disadvantage. The approach to cancer as a developmental disease provides a simple explanation for therapeutic failure in advanced stage carcinomas.

Since the 1970s, cytotoxic agents and radiation therapy have been an important part of the treatment of most low-grade carcinomas despite their little contribution to survival. Likewise, they are used in the palliative treatment of high-grade carcinomas regardless of their null effectiveness. The incorporation of drugs directed at molecular targets was irrelevant in survival since most signaling pathways are highly redundant and the complete blockade of essential elements is lethal for the host 299. Hence, for the most part, high-grade cancers, remain untreatable and incurable diseases 271. Furthermore, since the first requirement in the guideline for the search for new anticancer agents is the cytotoxicity at very low concentrations 299 is foresee that during the next years the new marketed molecules as antineoplastic drugs, will suffer from the same problems as the current agents, essentially, they will be very toxic and fail to cure cancer.

8.3. Paving the way for tumor reversion.
Countless experiments demonstrate the ability of tumor cells to differentiate. The first observations that tumor cells could originate a normal tissue come from studies carried out with teratomas in 1907 300, while the regression and differentiation of an epithelial tumor were possible in 1962 through its implantation in normal tissue 301. However, tumor differentiation with a pharmacological agent was possible until 1988. All-trans retinoic acid (ATRA) allowed complete remissions in cases of acute promyelocytic leukemia (APML) by inducing terminal differentiation of leukemic cells to granulocytes. With this type of treatment, more than 75% of patients diagnosed with APML are cured. The implementation of differentiation therapy in leukemias seems logical since many of them are characterized by the accumulation of immature lymphoid cells 302. However, in the rest of the neoplasms, the genocentric view of cancer conceptually weakens the therapeutic potential of induction of differentiation in at least two fundamental points. It does not consider that solid tumors are generated by immature precursor cells and postulates that cancer cells are unable to differentiate since possess an aberrant genetic code. The scarce events of cell differentiation in tumors were reconciled with the conventional genetic approach by attributing them to the activation of endogenous differentiation pathways that were unaltered during carcinogenesis.

On the contrary, in the hypothesis of epithelial carcinogenesis as a product of discrete cellular transitions, it is implicit that solid tumors are also originated from undifferentiated precursor cells. In addition, this approach allows the understanding of tumor reversion by inducing differentiation even in the presence of genomic damage. In the theoretical framework presented here tumor reversion is achieved through the induction of cell differentiation. In this view, the modulation of proliferation, senescence, and the inflammatory pathways with pharmacological agents modifies the epigenetic landscape to induce cell differentiation. The use of molecules from diverse chemical and structural nature can induce the attainment of a differentiated phenotype even in the absence of specific instructions (as would be the case of the stroma, of specific lineage growth factors, etc.) or conventional pharmacological interactions (Fig. 6.).

The objective of differentiation therapies in cancer is primarily to induce phenotypic maturation to decrease aggressive tumor behavior 303. Preclinical studies to assess the capacity of drugs to induce differentiation in solid tumors are scarce but are consistent with the premises of the model. For example, the use of troglitazone in patients with liposarcoma, demonstrated histological changes consistent with differentiation into adipocytes 304 and the use of ATRA in nasopharyngeal carcinoma increases the expression of epithelial markers 305. The molecular mechanism of both events of differentiation converges in the regulation of NF-κB and upregulation
of transcription factors that induce the normalization of tumor tissues towards organized histology that expresses markers of differentiated cells. The DNA acts as a robust network to structural changes, that is, most mutations do not affect the epigenetic landscape. Cells can differentiate into normal phenotypes despite mutations, and cancer can be induced in the absence of mutations. A) Mutations or genomic damage do not confer malignant traits, but rather activate and perpetuate the DDR and cellular inflammation (NF-κB), causing a significant loss of the mechanisms of transcriptional repression. B) This leads to the modification of the epigenetic landscape and allows the affected cells access to a multipotent state with a transcriptional profile of embryonic cells and the potential to induce cancer. Most carcinogens converge on these mechanisms, including cellular senescence. Telomere attrition facilitates the acquisition of chromosomal damage, which is established as DDR, cellular inflammation, and increased transcriptional activity. Proinflammatory cytokines and EMT allow access to a transcriptional state with a configuration similar to the mesenchymal stem cell. Therefore, nonspecific mutations allow access to the cell state that causes cancer. Similarly, ligands of diverse structural and chemical nature can lead to cell differentiation by altering the epigenetic landscape and essentially reducing cellular inflammation.

The induction of structural and functional changes in cancer cells related to phenotypic maturation is called cytoeducation. The study of the mechanism underlying the reversion of the malignant phenotype in cell cultures is growing and among the molecules with this type of effects is the ATRA, 1α, 25-dihydroxyvitamin D3, the coumarins daphnetin and 5-methoxy-6,7-methylenedioxy coumarin, the flavan-3-ols, such as catechin and epicatechin, and the flavonoids wogonin and apigetrin. The exposure of cancer cells to these compounds causes the inhibition of the cell cycle and the cancer-associated signaling pathways, including NF-κB. It is expected that the elucidation of the mechanisms by which some agents induce the differentiation of cancer cells eventually allows the establishment of more rational strategies for the treatment of cancer. This approach will lead to the development of less toxic agents to improve the organization of the affected tissues and the expression of epithelial markers. This would guarantee a delay in progression accompanied by an increase in survival. Based on the same rationale, the optimization of this type of therapy requires the surgical removal of tumor masses, the use of antiinflammatory and antiproliferative agents in conjunction with a healthy lifestyle to increase the chances of tumor reversal. We hope that the implementation of this type of strategy for cancer treatment will lead to an increased rate of remissions and eventually the chronic management of the disease.
9. Conclusion

Through the studying of epithelial carcinogenesis was developed an integrated theory that reconciles disperse results in an evolutive and developmental framework of the underlying causes of cancer. The rationale of the mechanistic association of aging and inflammation with sporadic carcinomas was uncovered and suggests the basis for the study of chronic diseases as part of the developmental process. The hypothesis of epithelial carcinogenesis as the result of cellular transitions promoted by cellular aging and inflammation considers cancer an endogenous cell state that matches the transcriptome of stem cells. Proposes that sporadic carcinomas are mainly the result of the reactivation of developmental processes in hyperplasias and implies that epithelial cells undergoing replicative senescence are susceptible to reprogramming in vivo using cellular transitions promoted by inflammation. These transitions are driven by transcriptional derepression that enables stemness, healing, and cancer. Although it might seem controversial, it provides a natural explanation of paradoxes accumulated in the field, and most importantly a rational basis for the therapeutic failure in cancer. The result of this theoretical endeavor offers a simple model to capture knowledge from the ever-increasing complexity in the molecular and cellular biology of cancers. Derived from the insights of the hypothesis the role of lifestyle and therapy was critically reviewed and some encouraging strategies for the management of carcinomas are provided. In conclusion, the underlying order in carcinogenesis suggests that is a disease resulting from the process of development and its interplay with lifestyle.

Acknowledgments

The author is grateful to Universidad Autónoma de Nuevo León, FASPYN and CONACYT for current support. He would like to thank colleagues for countless discussions during his tenure at the University of Cagliari, the Center of Complexity Sciences (C3, UNAM), the Biomedical Research Center of Northeast (IMSS), the Max Planck Institute for Biology of Ageing, and the School of Biological Sciences (UANL). The content is solely the responsibility of the author and does not necessarily represent the official views of the institutions.

References

1. Jemal, A.; Bray, F.; Center, M. M.; Ferlay, J.; Ward, E.; Forman, D., Global cancer statistics. CA a Cancer Journal for Clinicians 2011, 61 (2), 69-90.
2. Manders, D. B.; Keohoe, S. M.; Miller, D. S.; Lea, J. S.; Richardson, D. L., Third-line salvage chemotherapy for recurrent carcinoma of the cervix is associated with minimal response rate and high toxicity. American journal of clinical oncology 2017.
3. Holohan, C.; Van Schaeybroeck, S.; Longley, D. B.; Johnston, P. G., Cancer drug resistance: an evolving paradigm. Nature Reviews Cancer 2013, 13 (10), 714-726.
4. Rivera, E.; Gomez, H. In Chemotherapy resistance in metastatic breast cancer: the evolving role of ixabepilone, Breast Cancer Research, BioMed Central: 2010; p S2.
5. Wallington, M.; Saxon, E. B.; Bomb, M.; Smittenaar, R.; Wickenden, M.; McPhail, S.; Rashbass, J.; Chao, D.; Dewar, J.; Dodwell, D. 30-day mortality after systemic anticancer treatment for breast and lung cancer in England: a population-based, observational study. The Lancet Oncology 2016, 17 (9), 1203-1216.
6. Neugut, A. I.; Hillyer, G. C.; Kushi, L. H.; Lamatero, L.; Buono, D. L.; Nathanson, S. D.; Bovbjerg, D. H.; Mandelblatt, J. S.; Tsai, W.Y.; Jacobson. A prospective cohort study of early discontinuation of adjuvant chemotherapy in women with breast cancer: the breast cancer quality of care study (BQUAL). Breast cancer research and treatment, 2016, 158 (1), 127-138.
7. Boffetta, P.; Kaldor, J. M., Secondary malignancies following cancer chemotherapy. Acta Oncologica 1994, 33 (6), 591-598.
8. Morgan, G.; Ward, R.; Barton. The contribution of cytotoxic chemotherapy to 5-year survival in adult malignancies. Clinical oncology 2004, 16 (8), 549-560.
9. Zhu, W.Y.; Fang, K.; He, J.; Cui, R.; Zhang, Y.; Le, H. A prediction rule for overall survival in non-small-cell lung cancer patients with a pathological tumor size less than 30 mm. Disease Markers 2019.
10. Ufen, M.P.; Köhne, C.; Wischnewsky, M.; Wolters, R.; Novopashenny, I.; Fischer, J.; Constantinidou, M.; Possinger, K.; Regierer, A. Metastatic breast cancer: are we treating the same patients as in the past? Annals of oncology, 2014, 25 (1), 95-100.
11. Sonnenschein, C.; Soto, A. M., The aging of the 2000 and 2011 Hallmarks of Cancer reviews: a critique. Journal of biosciences, 2013, 38 (3), 651-63.
12. Baker, S. G.; Kramer, B. S., Paradoxes in carcinogenesis: new opportunities for research directions. BMC Cancer 2007, 7, 151.
13. Hanahan, D.; Weinberg, R. A., Hallmarks of cancer . Cell 2000, 100 (1) 57-70.
14. Hanahan, D.; Weinberg, R. A., Hallmarks of cancer: the next generation. Cell 2011, 144 (5), 646-674.
15. Weinberg, R. Coming full circle—from endless complexity to simplicity and back again. Cell 2014, 157 (1), 267-271.
16. Ben-Porath, I.; Thomson, M. W.; arev, V. J.; Ge, R.; Bell, G. W.; Regev, A.; Weinberg, R. A., An embryonic stem cell-like gene expression signature in poorly differentiated aggressive human tumors. Nature Genetics 2008, 40 (5), 499-507.
17. Mani, S. A.; Guo, W.; Liao, M. J.; Eaton, E. N.; Ayyanan, A.; Zhou, A. Y.; Brooks, M.; Reinhard, F.; Zhang, C. C.; Shipitsin, M.; Campbell, L. L.; Polyak, K.; Brinken, C.; Yang, J.; Weinberg, R. A., The epithelial-mesenchymal transition generates cells with properties of stem cells. Cell 2008, 133 (4), 704-15.
18. Lin, T.; Vala, M.; Barber, J.; Karp, J.; Smith, B.; Matsui, W.; Jones, R. Induction of acute lymphocytic leukemia differentiation by maintenance therapy. Leukemia 2007, 21 (9), 1915.
19. Gocek, E.; Studzinski, G. P., Vitamin D and differentiation in cancer. Critical reviews in clinical laboratory sciences 2009, 46 (4), 190-209.
20. Mahalingam, D.; Kong, C. M.; Lai, J.; Tay, L. L.; Yang, H.; Wang, X. J. Reversal of aberrant cancer methylome and transcriptome upon direct reprogramming of lung cancer cells. Scientific reports, 2012, 2, 592.
21. Willhauck, M. J.; Mirancea, N.; Vosseler, S.; Pavesio, A.; Boukamp, P.; Mueller, M. M.; Fusenig, N. E.; Stark, H. J. Reversion of tumor phenotype in surface transplants of skin SCC cells by scaffold-induced stroma modulation. Carcinogenesis 2007, 28 (3), 595-610.
22. Siegel, R. L.; Miller, K. D.; Jemal, A. Cancer statistics, 2016. CA: A Cancer journal for clinicians 2016, 66 (1), 7-30.
23. Tyler, S. J. Epithelium, the primary building block for metazoan complexity. Integrative and comparative biology 2003, 43 (1), 55-63.
24. Sementchenko, V. I.; Watson, D. K., ETS target genes: past, present and future. Oncogene 2000, 19 (55), 6533-48.
25. Hollenhorst, P. C.; Jones, D. A.; Graves, B. I., Expression profiles frame the promoter specificity dilemma of the ETS family of transcription factors. Nucleic Acids Research 2004, 32 (18), 5693-702.
26. Tymms, M. J.; Ng, A. Y.; Thomas, R. S.; Schutte, B. C.; Zhou, J.; Eyre, H. J.; Sutherland, G. R.; Seth, A.; Rosenberg, M.; Papas, T.; Debouck, C.; Kola, I., A novel epithelial-expressed ETS gene, ELF3: human and murine cDNA sequences, murine genomic organization, human mapping to 1q32.2 and expression in tissues and cancer. Oncogene 1997, 15 (20), 2449-62.
27. Andreoli, J. M.; Jung, S. I.; Chung, E.; Coticchia, C. M.; Steinert, P. M.; Markova, N. G. The expression of a novel, epithelium-specific ets transcription factor is restricted to the most differentiated layers in the epidermis. Nucleic Acids Research 1997, 25 (21), 4287-95.
28. Silverman, E. S.; Baron, R. M.; Palmer, L. J.; Le, L.; Hallock, A.; Subramanian, V.; Riese, R. J.; McKenna, M. D.; Gu, X.; Libermann, T. A.; Tugores, A.; Haley, K. J.; Shore, S.; Drazen, J. M.; Weiss, S. T., Constitutive and cytokine-induced expression of the ETS transcription factor ESE-3 in the lung. American Journal of Respiratory Cell and Molecular Biology 2002, 27 (6), 697-704.
29. Oetjgen, P.; Finger, E.; Sun, Z.; Akbarali, Y.; Thamrongaks, U.; Bolton, J.; Grall, F.; Dube, A.; Weiss, A.; Brown, L.; Quinn, G.; Kas, K.; Endress, G.; Kunisch, C.; Libermann, T. A., PDEF, a novel prostate epithelium-specific ETS transcription factor, interacts with the androgen receptor and activates prostate-specific antigen gene expression. Journal of Biological Chemistry 2000, 275 (2), 1216-25.
30. Tummala, R.; Sinha, S., Differentiation-specific transcriptional regulation of the ESE-2 gene by a novel keratinocyte-restricted factor. Journal of cellular biochemistry, 2006, 97 (4), 766-81.
31. Donnison, M.; Beaton, A.; Davey, H. W.; Broadhurst, R.; L’Huillier, P.; Pfeffer, P. L., Loss of the extraembryonic ectoderm in Elf5 mutants leads to defects in embryonic patterning. Development 2005, 132 (10), 2299-308.
32. Choi, Y. S.; Sinha, S., Determination of the consensus DNA-binding sequence and a transcriptional activation domain for ESE-2. Biochemical Journal 2006, 398 (3), 497-507.
33. Micalizzi, D. S.; Farabaugh, S. M.; Ford, H. L., Epithelial-mesenchymal transition in cancer: parallels between normal development and tumor progression. Journal of mammary biology and neoplasia 2010, 15 (2), 117-34.
34. Ahmad, A.; Banerjee, S.; Wang, Z.; Kong, D.; Majumdar, A. P.; Sarkar, F. H., Aging and inflammation: etiological culprits of cancer. Current aging science, 2009, 2 (3), 174-86.
35. Méndez-López, I. F.; Zapata-Benavides, P.; Zavala-Pompa, A.; Aguado-Barrera, M. E.; Pacheco-Calleros, J.; Rodríguez-Padilla, C.; Cerda-Flores, R. M.; Cortés-Gutiérrez, E. I.; Dávila-Rodríguez, M. I., Immunohistochemical analysis of prostate apoptosis response-4 (Par-4) in Mexican women with breast cancer: a preliminary study. Archives of medical research 2010, 41 (4), 261-268.
36. Méndez-López, I. F.; Zavala-Pompa, A.; Cortés-Gutiérrez, E. I.; Cerda-Flores, R. M.; Dávila-Rodríguez. Leptin receptor expression during the progression of endometrial carcinoma is correlated with estrogen and progesterone receptors. Archives of Medical Science 2017, 13 (1), 228.
37. Cserni, G., Tumour histological grade may progress between primary and recurrent invasive mammary carcinoma. Journal of clinical pathology, 2002, 55 (4), 293-7.
38. Shilkaitis, A.; Green, A.; Steele, V.; Lubet, R.; Kelloff, G.; Christov, K. J., Neoplastic transformation of mammary epithelial cells in rats is associated with decreased apoptotic cell death. Carcinogenesis 2000, 21 (2), 227-233.
39. Mintz, B., Gene expression in neoplasia and differentiation. Harvey Society Lectures 1978, Series 71.
40. Dupont, W. D.; Parl, F. F.; Hartmann, W. H.; Brinton, L. A.; Winfield, A. C.; Worrell, J. A.; Schuyler, P. A.; Plummer, W. D., Breast cancer risk associated with proliferative breast disease and atypical hyperplasia. Cancer 1993, 71, 1258-1258.
41. Radisky, D. C.; Sanistieban, M.; Berman, H. K.; Gauthier, M. L.; Frost, M. H.; Reynolds, C. A.; Vierkant, R. A.; Pankratz, V. S.; Visscher, D. W.; Tlsty, T. D., p16INK4a expression and breast cancer risk in women with atypical hyperplasia. Cancer prevention research, 2011, 4 (12), 1953-1960.
42. DeNardo, D. G.; Coussens, L. M., Inflammation and breast cancer. Balancing immune response: crosstalk between adaptive and innate immune cells during breast cancer progression. Breast Cancer Research 2007, 9 (4), 1.
43. Pare, R.; Soon, P. S.; Shah, A.; Lee, C. S. Differential expression of senescence tumour markers and its implications on survival outcomes of breast cancer patients. PloS one 2019, 14 (4).
44. Umbricht, C. B.; Sherman, M. E.; Dome, J.; Carey, L.; Marks, J.; Kim, N.; Sukumar, S. J. Telomerase activity in ductal carcinoma in situ and invasive breast cancer. Oncogene 1999, 18 (22), 3407-3414.
45. Meeker, A. K.; Hicks, J. L.; Gabrielson, E.; Strauss, W. M.; De Marzo, A. M.; Argani, P. J. Telomere shortening occurs in subsets of normal breast epithelium as well as in situ and invasive carcinoma. The American journal of pathology 2004, 164 (3), 925-935.
46. Younis, L. K.; El Sahka, H.; Haque, I. The prognostic value of E-cadherin expression in breast cancer. Journal of health sciences 2010, 1 (1).
47. Méndez-López, L. F.; Dávila-Rodríguez, M. I., Zavala-Pompa, A., Torres-López, E., González-Martínez, B. E., López-Cabanillas-Lomeli, M. Expression of leptin receptor in endometrial biopsies of endometrial and ovarian cancer patients. Biomedical reports, 2013 1(4), 659-663.
48. Chen, Y.C.; Hsu, H.S.; Chen, Y.W.; Tsai, T. H.; How, C. K.; Wang, C. Y.; Hung, S.C.; Chang, Y.-L.; Tsai, M.L.; Lee, Y. Oct-4 expression maintained cancer stem-like properties in lung cancer-derived CD133-positive cells. PloS one 2008, 3 (7).
49. Lakhataka, R.; Aljarrah, A.; Furnikh, M.; Ganguly, S. Epithelial mesenchymal transition (EMT) in metastatic breast Cancer in Omani women. Cancer Microenvironment 2017, 10 (1-3), 25-37.
50. Chen, Z.; Trotman, L. C.; Shaffer, D.; Lin, H. K.; Dotan, Z. A.; Niki, M.; Koutcher, J. A.; Scher, H. I.; Ludwig, T.; Gerald, W.; Cordon-Cardo, C.; Pudlof, P. Crucial role of p53-dependent cellular senescence in suppression of Pten deficient tumorigenesis. Nature 2005, 436 (7051), 725-30.
51. Campisi, J. Aging, cellular senescence, and cancer. Annual Review of Physiology 2013, 75, 685-705.
52. de Lange, T.; Cell biology. Telomere capping one strand fits all. Science 2001, 292 (5519), 1075-6.
53. Jacobs, J.; de Lange, T. Significant role for p16INK4a in p53-independent telomere-directed senescence. Current Biology 2004, 14 (24), 2302-8.
54. Karlseifer, J.; Smogorzewska, A.; de Lange, T., Senescence induced by altered telomere state, not telomere loss. Science 2002, 295 (5564), 2446-9.
55. Graham, M. K.; Meeker, A. J. Telomerases and telomerase in prostate cancer development and therapy. Nature Reviews Urology 2017, 14 (10), 607.
56. von Zglinicki, T.; Saretzki, G.; Ladhoff, J.; d’Adda di Fagagna, F.; Jackson, S. P. Human cell senescence as a DNA damage response. Mechanisms of ageing and development 2005, 126 (1), 111-7.
57. Rogakou, E. P.; Pilch, D. R.; Orr, A. H.; Ivanova, V. S.; Bonner, W. M. DNA double-stranded breaks induce histone H2AX phosphorylation on serine 139. Journal of biological chemistry 1998, 273 (10), 5858-68.
58. Burma, S.; Chen, B. P.; Murphy, M.; Kuriyama, A.; Chen, D. J. ATM phosphorylates histone H2AX in response to DNA double-strand breaks. Journal of Biological Chemistry 2001, 276 (45), 42462-7.
59. Herbig, U.; Jobling, W. A.; Chen, B. P.; Chen, D. J.; Sedivy, J. M. Telomere shortening triggers senescence of human cells through a pathway involving ATM, p53, and p21(CIP1), but not p16(INK4a). Molecular cell 2004, 14 (4), 501-13.
60. el-Deiry, W. S.; Kern, S. E.; Pietenpol, J. A.; Kinzler, K. W. Vogelstein, B. Definition of a consensus binding site for p53. Nature genetics 1992, 1 (1), 45-9.
61. de Stanchina, E.; Querido, E.; Narita, M.; Davuluri, R. V.; Pandolfo, P. P.; Ferbeyre, G.; Lowe, S. W. PML is a direct p53 target that modulates p53 effectors function. Molecular cell 2004, 13 (4), 523-35.
62. Afshari, C. A.; Nichols, M. A.; Xiong, Y.; Mudryj, M. A role for the p21-E2F interaction during senescence arrest of normal human fibroblasts. Cell Growth and Differentiation 1996, 7 (8), 979-88.
63. Waga, S.; Hannon, G. J.; Beach, D.; Stillman, B. The p21 inhibitor of cyclin-dependent kinases controls DNA replication by interaction with PCNA. Nature 1994, 369 (6481), 574-8.
64. Ferbeyre, G.; de Stanchina, E.; Querido, E.; Baptiste, N.; Prives, C.; Lowe, S. W. PML is induced by oncogenic ras and promotes premature senescence. Genes & development 2000, 14 (16), 2015-27.
65. Stuurman, N.; Meijne, A. M.; van der Pol, A. J.; de Jong, L.; van Driel, R.; van Renswoude, J. The nuclear matrix from cells of different origin. Evidence for a common set of matrix proteins. Journal of biological chemistry 1990, 265 (10), 5460-5.
66. Vernier, M.; Bourdeau, V.; Gaumont-Leclerc, M. F.; Moiseeva, O.; Begin, V.; Saad, F.; Mes-Masson, A. M.; Ferbeyre, G. Regulation of E2Fs and senescence by PML nuclear bodies. Genes & development 2011, 25 (1), 41-50.
67. Krishnamurthy, J.; Torrice, C.; Ramsey, M. R.; Koval, G. I.; Al-Regaiey, K.; Su, L.; Sharpless, N. E. Ink4a/Arf expression is a biomarker of aging. The Journal of clinical investigation 2004, 114 (9), 1299-307.
68. Bracken, A. P.; Kleine-Kohlbrecher, D.; Dietrich, N.; Pasini, D.; Gargiulo, G.; Beekman, C.; Theilgaard-Mönch, K.; Minucci, S.; Porse, B. T.; Marine, J. C. The Polycomb group proteins bind throughout the INK4A-ARF locus and are disassociated in senescent cells. Genes & development 2007, 21 (5), 525-530.
69. Sparmann, A.; van Lohuizen, M. Polycomb silencers control cell fate, development and cancer. Nature Reviews Cancer 2006, 6 (11), 846-56.
70. Iaquina, P. J.; Lees, J. A. Life and death decisions by the E2F transcription factors. Current opinion in cell biology 2007, 19 (6), 649-57.
71. Di Micco, R.; Sulli, G.; Dobревa, M.; Liontos, M.; Botrugno, O. A.; Gargiulo, G.; dal Zaffo, R.; Matti, V.; d’Ario, G.; Montani, E.; Mercurio, C.; Hahn, W. C.; Gorgoulis, V.; Minucci, S.; d’Adda di Fagagna, F. Interplay between oncogene-induced DNA damage response and heterochromatin in senescence and cancer. Nature Cell Biology 13 (3), 292-302.
99. Ansieau, S.; Bastid, J.; Doreau, A.; Morel, A.P.; Bouchet, B. P.; Thomas, C.; Fauvet, F.; Puisieux, I.; Doglioni, C.; Piccinin, S. Induction of EMT by twist proteins as a collateral effect of tumor-promoting inactivation of premature senescence. Cancer cell 2008, 14 (1), 79-89.

100. Huang, S., Tumor progression: chance and necessity in Darwinian and Lamarckian somatic (mutationless) evolution. Progress in biophysics and molecular biology 2012, 110 (1), 69-86.

101. Sistigu, A.; Di Modugno, F.; Manic, G.; Nistico, P. Deciphering the loop of epithelial-mesenchymal transition, inflammatory cytokines and cancer immunoediting. Cytokine & growth factor reviews 2017, 36, 67-77.

102. Stampfer, M. R.; LaBarge, M. A.; Garbe, J. C. An integrated human mammary epithelial cell culture system for studying carcinogenesis and aging. In Cell and Molecular Biology of Breast Cancer, Springer: 2013; pp 323-361.

103. Henson, D. E.; Ries, L.; Freedman, L. S.; Carriaga, M. Relationship among outcome, stage of disease, and histologic grade for 22,616 cases of breast cancer. The basis for a prognostic index. Cancer 1991, 68 (10), 2142-2149.

104. Ugnat, A.; Xie, L.; Morris, J.; Semenciw, R.; Mao, Y. Survival of women with breast cancer in Ottawa, Canada: variation with age, stage, histology, grade and treatment. British Journal of cancer 2004, 90 (6), 1138.

105. Chen, Y.; Zhang, L.; Liu, W.; Liu, X. Prognostic significance of the tumor-stroma ratio in epithelial ovarian cancer. BioMed research international 2015.

106. Sarrió, D.; Rodríguez-Pinilla, S. M.; Hardisson, D.; Cano, A.; Moreno-Bueno, G.; Palacios, J. Epithelial-mesenchymal transition in breast cancer relates to the basal-like phenotype. Cancer research 2008, 68 (4), 989-997.

107. Finak, G.; Bertos, N.; Pepin, F.; Sadekova, S.; Souleimanova, M.; Zhao, H.; Chen, H.; Omeroglu, G.; Meterissian, S.; Omeroglu, A., Stromal gene expression predicts clinical outcome in breast cancer. Nature medicine 2008, 14 (5), 518-527.

108. Chander, C.; Liu, T.; Buckanovich, R.; Coffman, L. The double edge sword of fibrosis in cancer. Translational Research 2019.

109. Bartoschek, M.; Oskolkov, N.; Bocci, M.; Lövrot, J.; Larsson, C.; Sommarin, M.; Madsen, C. D.; Lindgren, D.; Pekar, G.; Karlsson, G. Spatially and functionally distinct subclasses of breast cancer-associated fibroblasts revealed by single cell RNA sequencing. Nature communications 2018, 9 (1), 5150.

110. Kaimori, A.; Potter, J.; Kaimori, I.; Wang, C.; Mezey, E.; Koteish, A. Transforming growth factor-β1 induces an epithelial-to-mesenchymal transition state in mouse hepatocytes in vitro. Journal of Biological Chemistry 2007, 282 (30), 22089-22101.

111. Chopra, D. P.; Yeh, K. Long-term culture of epithelial cells from the normal rat colon. In vitro 1981, 17 (5), 441-449.

112. Dollner, R., Granzow, C., Helmke, B. M., Ruess, A., Schad, A., Dietz, A. The impact of stromal cell contamination on chemosensitivity testing of head and neck carcinoma. Anticancer research 2004, 24 (1), 325-332.

113. Tian, M.; Schiernmann, W. P. The TGF-β paradox in human cancer: an update. Future Medicine 2009, 259-271.

114. Wu, X.; Gong, S.; Roy-Burman, P.; Lee, P.; Culig, Z. Current mouse and cell models in prostate cancer research. Endocrine-related cancer 2013, 20 (4), 155-170.

115. Xu, X.; Zheng, L.; Yuan, Q.; Zhen, G.; Crane, J. L.; Zhou, X.; Cao, X. Transforming growth factor-β in stem cells and tissue homeostasis. Bone research 2018, 6 (1), 2.

116. Biswas, D. K.; Cruz, A. P.; Gansberger, E.; Pardee, A. Epidermal growth factor-induced nuclear factor κB activation: a major pathway of cell-cycle progression in estrogen-receptor negative breast cancer cells. Proceedings of the National Academy of Sciences 2000, 97 (15), 8542-8547.

117. Floor, S.; van Staveren, W. C.; Larsimont, D.; Dumont, J. E.; Maenhaut, C. Cancer cells in epithelial-to-mesenchymal transition and tumor-propagating-cancer stem cells: distinct, overlapping or same populations. Oncogene 2011, 30 (46), 4609-21.

118. Li, Q. Q.; Xu, J. D.; Wang, W. J.; Cao, X.; Chen, Q.; Tang, F.; Chen, Z. Q.; Lui, X. P.; Xu, Z. D. Twist-1-mediated adriamycin-induced epithelial-mesenchymal transition relates to multidrug resistance and invasive potential in breast cancer cells. Cancer Research 2009, 15 (8), 2657-65.

119. Hiscox, S.; Jiang, W. G.; Obermeier, K.; Taylor, K.; Morgan, L.; Burmi, R.; Barrow, D.; Nicholson, R. I., Tamoxifen resistance in MCF7 cells promotes EMT-like behavior and involves modulation of beta-catenin phosphorylation. International Journal of Cancer 2006, 118 (2), 290-301.

120. Terry, S.; Savagner, P.; Ortiz-Cuaran, S.; Mahjoubi, L.; Saintigny, P.; Thiery, J. P.; Chouaib, S. New insights into the role of EMT in tumor immune escape. Molecular oncology 2017, 11 (7), 824-846.

121. Xie, G.; Ji, A.; Yuan, Q.; Jin, Z.; Yuan, Y.; Ren, C.; Guo, Z.; Yao, Q.; Yang, K.; Lin, X. J. Tumour-initiating capacity is independent of epithelial-mesenchymal transition status in breast cancer cell lines. British journal of cancer 2014, 110 (10), 2514.

122. Reya, T.; Morrison, S. J.; Clarke, M. F.; Weissman, I. L. Stem cells, cancer, and cancer stem cells. Nature 2001, 414 (6859), 105-11.

123. Creighton, C. J.; Chang, J. C.; Rosen, J. M. Epithelial-mesenchymal transition (EMT) in tumor-initiating cells and its clinical implications in breast cancer. Journal of Mammary Gland Biology and Neoplasia 2010, 15 (2), 253-60.

124. Gao, X.; Liu, X.; Lu, Y.; Wang, Y.; Cao, W.; Liu, X.; Hu, H.; Wang, H. PIM1 is responsible for IL-6-induced breast cancer cell EMT and stemness via c-myc activation. Breast Cancer 2019, 1-9.

125. Xie, G.; Yao, Q.; Liu, Y.; Du, S.; Liu, A.; Guo, Z.; Sun, A.; Ruan, J.; Chen, L.; Ye, C. IL-6-induced epithelial-mesenchymal transition promotes the generation of breast cancer stem-like cells analogous to mammosphere cultures. International journal of oncology 2012, 40 (4), 1171-1179.

126. Huang, Z.; Wu, T.; Liu, A. Y.; Ouyang, G. Differentiation and transdifferentiation potentials of cancer stem cells. Oncotarget 2015, 6 (37), 39550.

127. Oltean, S.; Febbo, P. G.; Garcia-Blanco, M. Dunning rat prostate adenocarcinomas and alternative splicing reporters: powerful tools to study epithelial plasticity in prostate tumors in vivo. Clinical & experimental metastasis 2008, 25 (6), 611-619.
128. Chao, Y. L.; Shepard, C. R.; Wells, A. Breast carcinoma cells re-express E-cadherin during mesenchymal to epithelial reverting transition. Molecular cancer 2010, 9 (1), 179.

129. Chaffer, C. L.; Brennan, J. P.; Slavin, J. L.; Blick, T.; Thompson, E. W.; Williams, E. Mesenchymal-to-epithelial transition facilitates bladder cancer metastasis: role of fibroblast growth factor receptor-2. Cancer research 2006, 66 (23), 11271-11278.

130. Yao, D., Dai, C., Peng, S. Mechanism of the mesenchymal-epithelial transition and its relationship with metastatic tumor formation. Molecular cancer research 2011, 9 (12), 1608-1620.

131. Liao, T. T.; Yang, M. Revisiting epithelial-mesenchymal transition in cancer metastasis: the connection between epithelial plasticity and stemness. Molecular oncology 2017, 11 (7), 792-804.

132. Nelson, G.; Wordsworth, J.; Wang, C.; Jurk, D.; Lawless, C.; Martin-Ruiz, C.; von Zglinicki, T. A senescent cell bystander effect: senescence-induced senescence. Aging Cell 2012, 11 (2), 345-9.

133. Krotolina, A.; Campisi, J. Integrating cancer, aging stroma and cellular senescence. Advances in gerontology 2003, 11, 109-16.

134. Lin, C. Y.; Tsai, P. H.; Kandaswami, C. C.; Lee, P. P.; Huang, C. J.; Hwang, J. J.; Lee, M. T. Matrix metalloproteinase-9 cooperates with transcription factor Snail to induce epithelial-mesenchymal transition. Cancer science 2011, 102 (4), 815-27.

135. Wan, S., Zhao, E., Kryczek, I., Vatan, L., Sadowskaya, A., Ludema, G., Welling, T. H. Tumor-associated macrophages produce interleukin 6 and signal via STAT3 to promote expansion of human hepatocellular carcinoma stem cells. Gastroenterology 2014, 147 (6), 1393-1404.

136. Houghton, A. N., Uchi, H., Wolchok, J. D. The role of the immune system in early epithelial carcinogenesis: B-ware the double-edged sword. Cancer cell. 2005, 7(5), 403-405.

137. Joyce, J. A., Pollard, J. W. Microenvironmental regulation of metastasis. Nature reviews cancer. 2009, 9(4), 239-252.

138. Liu, D., Hornsby, P. J. Senescent human fibroblasts increase the early growth of xenograft tumors via matrix metalloproteinase secretion. Cancer research 2007, 67 (7), 3117-3126.

139. Roy, B.; Venkatachalapathy, S.; Ratna, P.; Wang, Y.; Jokhun, D. S.; Nagarajan, M.; Shivashankar, G. Laterally confined growth of cells induces nuclear reprogramming in the absence of exogenous biochemical factors. Proceedings of the National Academy of Sciences, 2018, 115 (21), 4741-4750.

140. Cordononsi, M.; Zanconato, F.; Azzolin, L.; Forcato, M.; Rosato, A.; Frasson, C.; Inui, M.; Montagner, M.; Parenti, A. R.; Poletti, A. The Hippo transducer TAZ confers cancer stem cell-related traits on breast cancer cells. Cell 2011, 147 (4), 759-772.

141. Mishra, R., Haldar, S., Suchard, S., Bhowmick, N. A. Epigenetic changes in fibroblasts drive cancer metabolism and differentiation. Endocrine-related cancer 2019, 1 (aop).

142. Farhood, B., Najafi, M., Mortezaei, K. Cancer-associated fibroblasts: Secretions, interactions, and therapy. Journal of cellular biochemistry 2019, 120 (3), 2791-2800.

143. Sullivan, N.; Sassar, A.; Axel, A. E.; Vesuna, F.; Raman, V.; Ramirez, N.; Oberszyn, T.; Hall, B. Interleukin-6 induces an epithelial mesenchymal transition phenotype in human breast cancer cells. Oncogene 2009, 28 (33), 2940.

144. Hackett, T. L., Warner, S. M., Stefanowicz, D., Shaheen, F., Pechkovsky, D. V., Murray, L. A., Knight, D. A. Induction of epithelial–mesenchymal transition in primary airway epithelial cells from patients with asthma by transforming growth factor-β1. American journal of respiratory and critical care medicine. 2009, 180(2), 122-133.

145. Lee, J. M.; Dedhar, S.; Kalluri, R.; Thompson, E. W. The epithelial-mesenchymal transition: new insights in signaling, development, and disease. Journal of cell Biology. 2006, 172 (6), 793-811.

146. Demaria, M.; Ohtani, N.; Youssef, S. A.; Rodier, F.; Toussaint, W.; Mitchell, J. R.; Laberge, R. M.; Vijg, J.; Van Steeg, H.; Dolé, M. An essential role for senescent cells in optimal wound healing through secretion of PDGF-AA. Developmental cell 2014, 31 (6), 722-733.

147. Mosteiro, L.; Pantajo, C.; de Martino, A.; Serrano, M. J. Senescence promotes in vivo reprogramming through p16 INK4a and IL-6. Aging cell 2018, 17 (2), e12711.

148. Vineis, P., Schatzkin, A., Potter, J. D. Models of carcinogenesis: an overview. Carcinogenesis, 2010, 31(10), 1703-1709.

149. Vogelstein, B., Kinzler, K. W. The multistep nature of cancer. Trends in genetics, 1993, 9(4), 138-141.

150. Place, A. E., Huh, S. J., Polyak, K. The microenvironment in breast cancer progression: biology and implications for treatment. Breast cancer research 2011, 13(6), 1-11.

151. Sonnenschein, C.; Soto, A. M. Somatic mutation theory of carcinogenesis: why it should be dropped and replaced. Molecular carcinogenesis 2000, 29 (4), 205-11.

152. Haddow, A. Molecular repair, wound healing, and carcinogenesis: tumor production a possible overhealing?. Advances in cancer research 1973, 16, 181-234.

153. Cofre, J.; Abdelhay, E. Cancer is to embryoology as mutation is to genetics: hypothesis of the cancer as embryological phenomenon. The Scientific World Journal 2017.

154. Battula, V. L.; Evans, K. W.; Hollier, B. G.; Shi, Y.; Marini, F. C.; Ayyanan, A.; Wang, R. Y.; Brisken, C.; Guerra, R.; Andreeff, M.; Mani, S. A. Epithelial-mesenchymal transition-derived cells exhibit multilinage differentiation potential similar to mesenchymal stem cells. Stem Cells 2010, 28 (8), 1435-45.

155. Davies, P.; Lineaweaver, C., Cancer tumors as Metazoa 1.0: tapping genes of ancient ancestors. Physical biology 2011, 8 (1), 015001.

156. Huang, S.; Ernberg, I.; Kauffman, S. Cancer attractors: a systems view of tumors from a gene network dynamics and developmental perspective. Seminars in cell & developmental biology 2009, 20 (7), 869-76.

157. Lu, D. Epigenetic modification enzymes: catalytic mechanisms and inhibitors. Acta Pharmacologica Sinica B 2013, 3(3), 141-149.
158. Wullaert, A.; Bonnet, M. C.; Pasparakis, M. NF-xB in the regulation of epithelial homeostasis and inflammation. Cell research 2011, 21 (1), 146-158.

159. Hoesel, B.; Schmid, J. A., The complexity of NF-kappaB signaling in inflammation and cancer. Molecular Cancer 2013, 12, 86.

160. Loop, T.; Pahl, H. Activators and target genes of Rel/NF-xB transcription factors. In Nuclear Factor xB, Springer 2003; pp 1-48.

161. Nakshatri, H., Appaiah, H. N., Anjanappa, M., Gilley, D., Tanaka, H., Badve, S., Bhut-Nakshatri, P. NF-xB dependent and independent epigenetic modulation using the novel anti-cancer agent DMAPT. Cell death & disease 2015, 6(1), e1608-e1608.

162. Takase, O., Yoshikawa, M., Idei, M., Hirahashi, J., Fujita, T., Takato, T., Hishikawa, K. The role of NF-xB signaling in the maintenance of pluripotency of human induced pluripotent stem cells. PLoS One, 2013, 8(2), e56399.

163. Li, C. W., Xia, W., Huo, L., Lim, S. O., Wu, Y., Hsu, J. L., Hung, M. C. Epithelial-mesenchymal transition induced by TNF-α requires NF-xB mediated transcriptional upregulation of Twist1. Cancer research 2012, 72(5), 1290-1300.

164. Markopoulos, G. S., Roupakia, E., Marcu, K. B., Kolettas, E. Epigenetic regulation of inflammatory cytokine-induced epithelial-to-mesenchymal cell transition and cancer stem cell generation. Cell 2019, 8(10), 1143.

165. De Santa, F.; Totaro, M. G.; Prosperini, E.; Notarbartolo, S.; Testa, G.; Natoli, G. The histone H3 lysine-27 demethylase Jmjd3 links inflammation to inhibition of polycomb-mediated gene silencing. Cell 2007, 130(6), 1083-1094.

166. Wu, K. J. Direct activation of Bmi1 by Twist1: implications in cancer stemness, epithelial-mesenchymal transition, and clinical significance. Chang Gung Med J 2011 34(3), 229-238.

167. Ramadoss, S., Chen, X., Wang, C. Y. Histone demethylase KDM6B promotes epithelial-mesenchymal transition. Journal of Biological Chemistry 2012 287(53), 44508-44517.

168. Iliopoulos, D., Hirsch, H. A., Struhl, K. An epigenetic switch involving NF-xB, Lin28, Let-7 MicroRNA, and IL6 links inflammation to cell transformation. Cell 2009, 139(4), 693-706.

169. Tong, L., Tergaonkar, V. Rho protein GTPases and their interactions with NF-xB: crossroads of inflammation and matrix biology. Bioscience reports, 2014, 34(3).

170. Becker-Weimann, S., Xiong, G., Furuta, S., Han, J., Kuhn, I., Akavia, U. D., Xu, R. NF-xB disrupts tissue polarity in 3D by preventing integration of microenvironmental signals. Oncotarget 2013, 4(11), 2010.

171. Sau, A., Cabrita, M. A., Pratt, M. C. NF-xB at the Crossroads of Normal Mammary Gland Biology and the Pathogenesis and Prevention of BRCA1-Mutated Breast Cancer. Cancer Prevention Research 2018, 11(2), 69-80.

172. Christine, J. Y., Regan, S., Liu, G., Alemana, S., Heng, H. H. Understanding aneuploidy in cancer through the lens of system inheritance, fuzzy inheritance and emergence of new genome systems. Molecular cytogenetics, 2018, 11(1), 31.

173. Tijhuis, A. E., Johnson, S. C., McClelland, S. E. The emerging links between chromosomal instability (CIN), metastasis, inflammation and tumour immuno. Molecular cytogenetics 2019, 12(1), 17.

174. Herrera, L. A., Prada, D., Andonegui, M. A., Dueñas-González, A. The epigenetic origin of aneuploidy. Current genomics 2008, 9(1), 43-50.

175. Duncan, A. W. In Aneuploidy, polyploidy and ploidy reversal in the liver. Seminars in cell & developmental biology, Elsevier: 2013; pp 347-356.

176. Lodato, M. A., Woodworth, M. B., Lee, S., Evrony, G. D., Mehta, B. K., Karger, A., Walsh, C. A. Somatic mutation in single human neurons tracks developmental and transcriptional history. Science 2015, 350(6256), 94-98.

177. Radisky, D. C., Kenny, P. A., Bissell, M. J. Fibrosis and cancer: do myofibroblasts come also from epithelial cells via EMT? Journal of cellular biochemistry 2007, 101(4), 830-839.

178. Willis, B. C., duBois, R. M., Borok, Z. Epithelial origin of myofibroblasts during fibrosis in the lung. Proceedings of the American Thoracic Society 2006, 3(4), 377-382.

179. Willis, B. C., duBois, R. M., Borok, Z. Epithelial origin of myofibroblasts during fibrosis in the lung. Proceedings of the American Thoracic Society 2006, 3(4), 377-382.

180. Koch, C. M., Suschek, C. V., Lin, Q., Bork, S., Goergens, M., Joussen, S., Wagner, W. Specific age-associated DNA methylation changes in human dermal fibroblasts. PloS one 2011, 6(2), e16679.

181. El Agha, E., Kramann, R., Schneider, R. K., Li, X., Seeger, W., Humphreys, B. D., Belluscio, S. Mesenchymal stem cells in fibrotic disease. Cell stem cell 2017, 21(2), 166-177.

182. Foote, A. G., Wang, Z., Kendzierski, C., & Thibeault, S. L. Tissue specific human fibroblast differential expression based on RNA sequencing analysis. BMC genomics 2019, 20(1), 308.

183. Ichim, T. E., O’Heeron, P., Kesari, S. Fibroblasts as a practical alternative to mesenchymal stem cells. Journal of translational medicine 2018, 16(1), 121.

184. Kendall, R. T., Feghali-Bostwick, C. A. Fibroblasts in fibrosis: novel roles and mediators. Frontiers in pharmacology 2014, 5, 123.

185. Kim, W., Barron, D. A., Rowley, D. R. RUNX1 is essential for mesenchymal stem cell proliferation and myofibroblast differentiation. Proceedings of the National Academy of Sciences 2014, 111(46), 16389-16394.

186. Hinz, B., Phan, S. H., Thannickal, Gabbiani, G. Recent developments in myofibroblast biology: paradigms for connective tissue remodeling. The American journal of pathology 2012, 180(4), 1340-1355.

187. Choi, H. S., Ryu, C. J., Choi, H. M., Kim, K. S. Effects of the pro-inflammatory milieu on the dedifferentiation of cultured fibroblast-like synoviocytes. Molecular medicine reports 2012, 5(4), 1023-1026.

188. Ray, T., Shah, A., Bulla, G. A., Ray, P. S. Gatekeeper transcription factors regulate switch between lineage preservation and cell plasticity. bioRxiv 2020.

189. Selman, M., Pardo, A., Kaminski, N. Idiopathic pulmonary fibrosis: aberrant recapitulation of developmental programs? PLoS Med, 2008, 5(3), e62.
190. Xie, T., Liang, J., Liu, N., Huan, C., Noble, P. W. Transcription factor TBX4 regulates myofibroblast accumulation and lung fibrosis. *The Journal of clinical investigation* 2016, 126(8), 3063-3079.

191. Kubo, H., Shimizu, M., Taya, Y., Kato, Y. Identification of mesenchymal stem cell transcription factors by microarray and knockdown analyses, and signature molecule markers of MSC in bone marrow by immunohistochemistry. *Genes to cells* 2009, 14(3), 407-424.

192. Lai, P. L., Lin, H., Chen, S. F., Lu, J. Efficient generation of chemically induced mesenchymal stem cells from human dermal fibroblasts. *Scientific reports* 2017, 7(1), 1-13.

193. Chen, F., Zhang, G., Yu, L., Pradhan, S. High-efficiency generation of induced pluripotent mesenchymal stem cells from human dermal fibroblasts using recombinant proteins. *Stem cell research & therapy* 2016, 7(1), 99.

194. Huang, H. I., Chen, S. K., Ling, Q. D., Chan, S. H. Multilineage differentiation potential of fibroblast-like stromal cells derived from human skin. *Tissue Engineering Part A* 2010, 16(5), 1491-1501.

195. Song, L. N., Tuan, R. S. Transdifferentiation stem cell potential of human mesenchymal stem cells derived from bone marrow. *The FASEB Journal* 2004, 18(9), 980-982.

196. Ugrulu, B., Karaoz, E. Comparison of similar cells: Mesenchymal stromal cells and fibroblasts. *Acta histochemical* 2020, 122(8), 151634.

197. Zipori, D. The nature of stem cells: state rather than entity. *Nature Reviews Genetics* 2004, 5 (11), 873.

198. Lee, H. Y., Hong, I. S. Double-edged sword of mesenchymal stem cells: cancer-promoting versus therapeutic potential. *Cancer science* 2017, 108(10), 1939-1946.

199. Xu, M., Shaw, G., Murphy, M., Barry, F. Induced Pluripotent Stem Cell-Derived Mesenchymal Stromal Cells Are Functionally and Genetically Different From Bone Marrow-Derived Mesenchymal Stromal Cells. *Stem cells* 2019, 37(6), 754-765.

200. d’Adda di Fagagna, F.; Reaper, P. M.; Clay-Farrace, L.; Fiegler, H.; Carr, P.; Von Zglinicki, T.; Saretzki, G.; Carter, N. P.; Jackson, S. P. A DNA damage checkpoint response in telomere-initiated senescence. *Nature* 2003, 426 (6963), 194-8.

201. Vaughan, S., Jat, P. S. Deciphering the role of nuclear factor-kB in cellular senescence. *Aging* 2011, 3(10), 913

202. Francou, A., Anderson, K. V. The Epithelial-to-Mesenchymal Transition in Development and Cancer. *Annual Review of Cancer Biology* 2020, 4, 197-220.

203. Wilson, M. M., Weinberg, R. A., Lees, G., Venu, J. Emerging Mechanisms by which EMT Programs Control Stemness. *Trends in Cancer* 2020.

204. Van Der Sanden, B.; Dhobb, M.; Berger, F.; Wion, D. Optimizing stem cell culture. *Journal of cellular biochemistry* 2010, 111 (4), 801-807.

205. Abedian, Z., Akhavan Niaki, H., Zabibi, Mostafazadeh, A. Spontaneous mesenchymal to epithelial like tissue transition (MET) in a long term human skin culture. *International Biological and Biomedical Journal* 2017, 3(3), 112-118.

206. Lu, W., Kang, Y. Epithelial-mesenchymal plasticity in cancer progression and metastasis. *Developmental cell* 2019, 49(3), 361-374.

207. Casanova, J. Stemness as a cell default state. *EMBO reports* 2012, 13 (5), 396-7.

208. Efroni, S.; Duttagupta, R.; Cheng, J.; Delghani, H.; McKay, R. D.; Buetow, K. Global transcription in pluripotent embryonic stem cells. *Cell stem cell* 2008, 2 (5), 437-447.

209. Kashyap, V.; Rezende, N. C.; Scotland, K. B.; Shaffer, S. M.; Persson, J. L.; Mongan, N. Regulation of stem cell pluripotency and differentiation involves a mutual regulatory circuit of the NANOG, OCT4, and SOX2 pluripotency transcription factors with polycomb repressive complexes and stem cell microRNAs. *Stem cells and development* 2009, 18 (7), 1093-1108.

210. Lozoya, O. A.; Gilchrist, C. I.; Guilak, F. Universally conserved relationships between nuclear shape and cytoplasmic mechanical properties in human stem cells. *Scientific reports* 2016, 6, 23047.

211. MacLean, A. L.; Kirk, P. D.; Stumpf, M. P. Cellular population dynamics control the robustness of the stem cell niche. *Biology open* 2015, 4 (11), 1420-1426.

212. Hoffmann, M.; Chang, H. H.; Huang, S.; Ingber, D. E.; Loeffler, M.; Galle, J. Noise-driven stem cell and progenitor population dynamics. *PLoS One* 2008, 3 (8), e2922.

213. Xin, T.; Greco, V.; Myung, P. Hardwiring stem cell communication through tissue structure. *Cell* 2016, 164 (6), 1212-1225.

214. Kang, N.; Choi, S. Y.; Kim, B. N.; Yeo, C. D.; Park, C. K.; Park, J. Hypoxia-induced cancer stemness acquisition is associated with CXCR4 activation by its aberrant promoter demethylation. *BMC cancer* 2019, 19 (1), 148.

215. Gopal, K.; Gupta, N.; Zhang, H.; Alshareef, A.; Alqahtani, H.; Bigras, G.; Lewis, J.; Douglas, D.; Kneteman, N.; Lavasanifar, A. Oxidative stress induces the acquisition of cancer stem-like phenotype in breast cancer detectable by using a Sox2 regulatory region-2 (SRR2) reporter. *Oncotarget* 2016, 7 (3), 31111.

216. Zhang, L.; Shi, H.; Chen, H.; Xu, X.; You, T.; Fan, X.; Wang, D. Dedifferentiation process driven by radiotherapy-induced HMGBl/TLR2/YAP/HIF-1 signaling enhances pancreatic cancer stemness. *Cell death & disease* 2019, 10 (10), 1-16.

217. Martins-Neves, S. R.; Cleeton-Jansen, A. M.; Gomes, C. Therapy-induced enrichment of cancer stem-like cells in solid human tumors: Where do we stand? *Pharmaceutical research* 2018, 137, 193-204.

218. Liu, X.; Li, F.; Huang, Q.; Zhang, Z.; Zhou, L.; Deng, Y.; Zhou, M.; Fleenor, D. E.; Wang, H.; Kastan, M. Self-inflicted DNA double-strand breaks sustain tumorigenicity and stemness of cancer cells. *Cell research* 2017, 27 (6), 764-783.

219. Intlekofer, A. M.; Finley, L. W. Metabolic signatures of cancer cells and stem cells. *Nature metabolism* 2019, 1 (2), 177.

220. Liu, Y.; Muñoz, N.; Tsiakas, A. C.; Logan, T. M.; Ma, T. Metabolic reconfiguration supports reacquisition of primitive properties in human mesenchymal stem cell aggregates. *Stem cells* 2017, 35 (2), 398-410.

221. Meng, L.; Hu, H.; Zhi, H.; Liu, Y.; Shi, F.; Zhang, L.; Zhou, Y.; Lin, A. OCT4B regulates p53 and p16 pathway genes to prevent apoptosis of breast cancer cells. *Oncology letters* 2018, 16 (1), 522-528.
222. Jain, A. K.; Barton, M. p53: emerging roles in stem cells, development and beyond. Development 2018, 145 (8), dev158360.

223. Turdo, A.; Veschi, V.; Gaggianesi, M.; Chinnici, A.; Bianca, P.; Todaro, M.; Stassi, G. Meeting the challenge of targeting cancer stem cells. Frontiers in cell and developmental biology 2019, 7, 16.

224. Hiyama, E.; Hiyama, K. Telomere and telomerase in stem cells. British journal of cancer 2007, 96 (7), 1020-1024.

225. Na, J.; Baker, D.; Zhang, J.; Andrews, P. W.; Barbaric, I. Aneuploidy in pluripotent stem cells and implications for cancerous transformation. Protein & cell 2014, 5 (8), 569-579.

226. de Lucas, B.; Pérez, L. M.; Gálvez, B. G. Importance and regulation of adult stem cell migration. Journal of cellular and molecular medicine 2018, 22 (2), 746-754.

227. Agudo, J.; Park, E. S.; Rose, S. A.; Alibo, E.; Sweeney, R.; Dhainaut, M.; Kobayashi, K. S.; Sachidanandam, R.; Baccarini, A.; Merad, M. Quiescent tissue stem cells evade immune surveillance. Immunity 2018, 48 (2), 271-285, e5.

228. Neph, S.; Stergachis, A. B.; Reynolds, A.; Sandstrom, R.; Borenstein, E.; Stamatoyannopoulos, J. Circuitry and dynamics of human transcription factor regulatory networks. Cell 2012, 150 (6), 1274-86.

229. Karsten, U.; Goletz, S. What makes cancer stem cell markers different? SpringerPlus 2013, 2 (1), 1-8.

230. Li, L.; Navees, W. B. Normal stem cells and cancer stem cells: the niche matters. Cancer Research 2006, 66 (9), 4553-7.

231. Pierce Jr, G.; Dixon Jr, F.; Verney, E. Teratocarcinogenic and tissue-forming potentials of the cell types comprising neoplastic embryoid bodies. Journal of technical methods and pathology 1960, 9, 583.

232. Malta, T. M.; Sokolov, A.; Gentles, A. J.; Burzykowski, T.; Poisson, L.; Weinstein, J. N.; Kamińska, B.; Huelskens, J.; Omer, L.; Gevaert, O. Machine learning identifies stemness features associated with oncogenic dedifferentiation. Cell 2018, 173 (2), 338-354, e15.

233. Li, Y.; Zhao, H.; Huang, X.; Zhou, B. Embryonic senescent cells re-enter cell cycle and contribute to tissues after birth. Cell research 2018, 28 (7), 775-778.

234. Santos, F.; Moreira, C.; Nóbrega-Pereira, S.; Bernardes de Jesus, B. New Insights into the Role of Epithelial-Mesenchymal Transition during Aging. International journal of molecular sciences 2019, 20 (4), 891.

235. Tian, L.; Lu, Z. P.; Cai, B. B.; Jiang, K. R. Activation of pancreatic stellate cells involves an EMT-like process. International Journal of Oncology 2016, 48 (2), 783-792.

236. Wang, Y. Y.; Cen, J. N.; He, J.; Shen, H. J.; Chen, Z. X. Accelerated cellular senescence in myelodysplastic syndrome. Experimental hematology 2009, 37 (11), 1310-1317.

237. Greim, H.; Kaden, D. A.; Larson, R. A.; Snyder, R. (2014). The bone marrow niche, stem cells, and leukemia: impact of drugs, chemicals, and the environment. Annals of the New York Academy of Sciences, 1310 (1), 7.

238. Cortes, J.; O'Dwyer, M. E. Clonal evolution in chronic myelogenous leukemia. Hematology/oncology clinics of North America 2004, 18 (3), 671.

239. Riether, C.; Schürch, C. M.; Bührer, E. D.; Hinterbrandner, Ochsenbein, A. F. CD70/CD27 signaling promotes blast stemness and is a viable therapeutic target in acute myeloid leukemia. Journal of Experimental Medicine 2017, 214 (2), 359-380.

240. Anand, P.; Kunnunakkara, A. B.; Sundaram, C.; Harikumar, K. B.; Tharakan, S. T.; Lai, O. S.; Sung, B.; Aggarwal, B. B. Cancer is a preventable disease that requires major lifestyle changes. Pharmaceutical research 2008, 25 (9), 2097-116.

241. Eyre, H.; Kahn, R.; Robertson, R. M.; Clark, N. G.; Doyle, C.; Hong, Y.; Gansler, T.; Glynn, T.; Smith, R. A.; Taubert, K.; Thun, M. J. Preventing cancer, cardiovascular disease, and diabetes: a common agenda for the American Cancer Society. Stroke 2004, 35 (8), 1999-2010.

242. Schwingshackl, L.; Schwedhelm, C.; Galbete, C.; Hoffmann, G. Adherence to Mediterranean diet and risk of cancer: an updated systematic review and meta-analysis. Nutrients 2017, 9 (10), 1063.

243. Kubik, A.; Zlatoukal, P.; Tomasek, L.; Pauk, N.; Havel, L.; Doležal, J.; Plesko, I. Interactions between smoking and other exposures associated with lung cancer risk in women: diet and physical activity. Neoplasma 2007, 54 (1), 83-88.

244. Ruiz, R. B.; Hernández, P. Diet and cancer: risk factors and epidemiological evidence. Maturitas 2014, 77 (3), 202-208.

245. Klement, R.; Fink, M. Dietary and pharmacological modification of the insulin/IGF-1 system: exploiting the full repertoire against cancer. Oncogenesis 2016, 5 (2), e193.

246. Patterson, R. E.; Rock, C. L.; Kerr, J.; Natarajan, L.; Marshall, S. J.; Pakiz, B.; Cadmus-Bertram, L. A. Dietetics, Metabolism and breast cancer risk: frontiers in research and practice. Journal of the Academy of Nutrition and Dietetics 2013, 113 (2), 288-296.

247. Galland, L. Diet and inflammation. Nutrition in Clinical Practice 2010, 25 (6), 634-640.

248. Heymach, J. V.; Shackelford, T. J.; Tran, H. T.; Yoo, S. Y.; Do, K. A.; Wergin, M.; Polascik, T. J.; Snyder, D. Effect of Low-Fat Diets on Plasma Levels of NF-κB Regulated Inflammatory Cytokines and Angiogenic Factors in Men with Prostate Cancer. Cancer Prevention Research 2011, 4 (10), 1509-1509.

249. Vitale, G.; Pellegrino, G.; Vollery, M.; Hofland, L. Role of IGF-1 system in the modulation of longevity: controversies and new insights from a centenarians’ perspective. Frontiers in endocrinology 10: 27. 2019.

250. Fontana, L.; Partridge, L. Promoting health and longevity through diet: from model organisms to humans. Cell 2015, 161 (1), 106-118.

251. Betteli, L.; Foukas, L. Growth factor, energy and nutrient sensing signalling pathways in metabolic ageing. Biogerontology 2017, 18 (6), 913-929.

252. Seidemann, S. B.; Claggett, B.; Cheng, S.; Henglin, M.; Shah, A.; Steffen, L. M.; Folsom, A. R.; Rimml, E. B.; Willett, W. C.; Solomon, S. Dietary carbohydrate intake and mortality: a prospective cohort study and meta-analysis. The Lancet Public Health 2018, 3 (9), e419-e428.

253. McCarty, M. F.; Barroso-Aranda, J.; Contreras, F. The low-methionine content of vegan diets may make methionine restriction feasible as a life extension strategy. Medical hypotheses 2009, 72 (2), 125-128.
254. Redman, L. M.; Ravussin, E. Caloric restriction in humans: impact on physiological, psychological, and behavioral outcomes. *Antioxidants & redox signaling* 2011, 14 (2), 275-287.

255. Hever, J.; Cronise, R. Plant-based nutrition for healthcare professionals: implementing diet as a primary modality in the prevention and treatment of chronic disease. *Journal of geriatric cardiology* 2017, 14 (5), 355.

256. Vitale, G.; Pellegrino, G.; Vollery, M.; Hofland, L. Role of IGF-1 system in the modulation of longevity: controversies and new insights from a centenarians' perspective. *Frontiers in endocrinology* 2019, 10, 27.

257. Leonov, A.; Afia-Ciommo, A.; Piano, A.; Svishtova, V.; Lutchman, V.; Medkour, Y.; Titorenko, V. Longevity extension by phytochemicals. *Molecules* 2015, 20 (4), 6544-6572.

258. Singh, R. K.; Chang, H.W.; Yan, D.; Lee, K. M.; Zhu, T. Influence of diet on the gut microbiome and implications for human health. *Journal of translational medicine* 2017, 15 (1), 73.

259. Edwards, I. J.; O’Flaherty, J. T.; Omega-3 Fatty Acids and PPARgamma in Cancer. *PPAR Research* 2008, 358052.

260. Kummumakkara, A. B.; Sailo, B. L.; Banik, K.; Harsha, C.; Prasad, S.; Gupta, S. C.; Bharti, A. C.; Aggarwal, B. B. Chronic diseases, inflammation, and spices: how are they linked? *Journal of translational medicine* 2018, 16 (1), 14.

261. Boccardi, V.; Esposito, A.; Rizzo, M. R.; Marfella, R.; Barbieri, M.; Paolisson, G. Mediterranean diet, telomere maintenance and health status among elderly. *PloS one* 2013, 8 (4), e62781.

262. Webb, P. M.; Byrne, C.; Schnitt, S. J.; Connolly, N. L.; Jacobs, T. W.; Baer, H. J.; Willett, W. C.; Colditz, G. A. A prospective study of diet and benign breast disease. *Cancer Epidemiology and Prevention Biomarkers* 2004, 13 (7), 1106-1113.

263. Dunlap, S. M.; Chiao, L. J.; Nogueira, L.; Usary, J.; Perou, C. M.; Varticovski, L.; Hursting, S. Dietary energy balance modulates epithelial-to-mesenchymal transition and tumor progression in murine claudin-low and basal-like mammary tumor models. *Cancer prevention research* 2012, 5 (7), 930-942.

264. Rojas, J.; Chávez, M.; Oliviar, L.; Rojas, M.; Morillo, J.; Mejías, J.; Calvo, M.; Bermúdez, V. Polycystic ovary syndrome, insulin resistance, and obesity: navigating the pathophysiological labyrinth. *International journal of reproductive medicine* 2014, 2014.

265. Fontana, L. Modulating human aging and age-associated diseases. *Biochimica et Biophysica Acta* 2009, 1790 (10), 1133-1138.

266. Pérez, L. M.; Pareja-Galeano, H.; Sanchis-Gomar, F.; Emanuele, E.; Lucia, A.; Gálvez, B. 'Adipaging': ageing and obesity share biological hallmarks related to a dysfunctional adipose tissue. *The Journal of physiology* 2016, 594 (12), 3187-3207.

267. Ogrodnik, M.; Zhu, Y.; Langhi, L. G.; Tchkinson, T.; Ruswhandi, R. A.; Giorgiadze, N.; Pirskhalava, T. Obesity-induced cellular senescence drives anxiety and impairments neurogenesis. *The Journal of physiology* 2019, 29 (5), 1061-1077, 88.

268. Dunlap, S. M.; Chiao, L. J.; Nogueira, L.; Usary, J.; Perou, C. M.; Varticovski, L.; Hursting, S. Dietary energy balance modulates epithelial-to-mesenchymal transition and tumor progression in murine claudin-low and basal-like mammary tumor models. *Cancer Prevention Research* 2012, 5 (7), 930-942.

269. Ruiterkamp, J.; Ernst, M. The role of surgery in metastatic breast cancer. *European journal of cancer* 2011, 47, S6-S22.

270. Fisher, S.; Gao, H.; Yasui, Y.; Dabbs, K.; Winget, M. Survival in stage I-IIIB breast cancer patients by surgical treatment in a publicly funded health care system. *Annals of Oncology* 2015, 26 (6), 1161-1169.

271. Morgan, G.; Ward, R.; Barton, M. The contribution of cytotoxic chemotherapy to 5-year survival in adult malignancies. *Clinical oncology* 2004, 16 (8), 549-560.

272. Simon, M.; Bosset, P.O.; Benhamou, S.; Lebret, T. Multiple recurrences and risk of disease progression in patients with primary low-grade (TaG1) non muscle-invasive bladder cancer and with low and intermediate EORTC-risk score. *PloS one* 2019, 14 (2).

273. Earle, C. C.; Landrum, M. B.; Souza, J. M.; Neville, B. A.; Weeks, J. C.; Ayanian, J. Aggressiveness of cancer care near the end of life: is it a quality-of-care issue? *Journal of clinical oncology* 2008, 26 (23), 3860.

274. Gnerlich, J.; Jeffe, D. B.; Deshpande, A. D.; Beers, C.; Zander, C.; Margenthaler, J. Surgical removal of the primary tumor increases overall survival in patients with cancer: analysis of the 1988–2003 SEER data. *Annals of surgical oncology* 2007, 14 (8), 2187-2194.

275. Gray, J. W. Evidence emerges for early metastasis and parallel evolution of primary and metastatic tumors. *Cancer Cell* 2003, 4 (1), 4-6.

276. Lippert, T. H.; Ruoff, H.J.; Volm, M. J. A. Intrinsic and acquired drug resistance in malignant tumors. *Arzneimittel Forschung* 2008, 58 (06), 261-264.

Transdifferentiation potential of human mesenchymal stem cells derived from bone marrow.

277. Pattison, S.; Mitchell, C.; Lade, S.; Leong, T.; Busuttil, R. A.; Boussiotas, A. Early relapses after adjuvant chemotherapy suggests primary chemoresistance in diffuse gastric cancer. *PloS One* 2017, 12 (9).

278. Geurs, Y.; Witteveen, A.; Breetveld, R.; Poortmans, P.; Sonke, G. S.; Strobbe, L.; Siesling, S. Patterns and predictors of first and subsequent recurrence in women with early breast cancer. *Breast cancer research and treatment* 2017, 165 (3), 709-720.

279. Housman, G.; Byler, S.; Heerboth, S.; Lapinska, K.; Longacre, M.; Snyder, N.; Sarkar, D. Drug resistance in cancer: an overview. *Cancers* 2014, 6 (3), 1769-1792.

280. Saif, M. W.; Shahrakni, A.; Cornfeld, D. Gemcitabine-induced liver fibrosis in a patient with pancreatic cancer. *JOP* 2007, 8 (4), 460-281. Vyas, D.; Laput, G.; Vyas, A. Chemotherapy-enhanced inflammation may lead to the failure of therapy and metastasis. *OncoTargets and therapy* 2014, 7, 1015.

281. Li, Q.; Chen, Z. Q.; Cao, X.; Xu, J. D.; Xu, J. W.; Chen, Y.; Xu, Z. D. Involvement of NF-kappaB/miR-448 regulatory feedback loop in chemotherapy-induced epithelial-mesenchymal transition of breast cancer cells. *Cell Death and Differentiation* 2011, 18 (1), 16-25.

282. Relling, M. V.; Rubnitz, J. E.; Rivera, G. K.; Boyett, J. M.; Hancock, M. L.; Felix, C. A.; Kun, L. E.; Walter, A. W.; Evans, W. E.; Pui, C. H. High incidence of secondary brain tumours after radiotherapy and antimetabolites. *The Lancet* 1999, 354 (9172), 34-39.
284. Schneider, U. Modeling the risk of secondary malignancies after radiotherapy. *Genes* 2011, 2 (4), 1033-1049.
285. Ueda, S.; Saeki, T.; Osaki, A.; Yamane, T.; Kuji, I. Bevacizumab induces acute hypoxia and cancer progression in patients with refractory breast cancer: multimodal functional imaging and multiplex cytokine analysis. *Clinical Cancer Research* 2017, 23 (19), 5769-5778.
286. Duman, B.; Paydas, S.; Disel, U.; Besen, A.; Gürkan, E. Secondary malignancy after imatinib therapy: eight cases and review of the literature. *Leukemia & lymphoma* 2012, 53 (9), 1706-1708.
287. Josting, A.; Wiedemann, S.; Franklin, J.; Diehl, V. Secondary myeloid leukemia and myelodysplastic syndromes in patients treated for Hodgkin’s disease. *Journal of Clinical Oncology* 2003, 21 (18), 3440-3446.
288. Tajima, H.; Ohta, T.; Kitagawa, H.; Okamoto, K.; Sakai, S.; Kinoshita, J.; Makino, I.; Furukawa, H.; Hayashi, H.; Nakamura, K. Neoadjuvant chemotherapy with gemcitabine for pancreatic cancer increases in situ expression of the apoptosis marker M30 and stem cell marker CD44. *Oncology Letters* 2012, 3 (6), 1186-1190.
289. Sidi, R.; Pasello, G.; Opitz, I.; Soltermann, A.; Tutic, M.; Rehrauer, H.; Weder, W.; Stahel, R. A.; Felly-Bosco, E. Induction of senescence markers after neo-adjuvant chemotherapy of malignant pleural mesothelioma and association with clinical outcome: an exploratory analysis. *European Journal of Cancer* 2011, 47 (2), 326-332.
290. Tato-Costa, J.; Casimiro, S.; Pacheco, T.; Pires, R.; Fernandes, A.; Alho, I.; Pereira, P.; Costa, P.; Castelo, H. B.; Ferreira, J. Therapy-induced cellular senescence induces epithelial-to-mesenchymal transition and increases invasiveness in rectal cancer. *Clinical colorectal cancer* 2016, 15 (2), 170-178. e3.
291. Fraiser, L. H.; Kanekal, S.; Kehrer, J. Cyclophosphamide toxicity. *Drugs* 1991, 42 (5), 781-795.
292. Testa, N. G.; Hendry, J. H.; Molineux, G. Long-term bone marrow damage in experimental systems and in patients after radiation or chemotherapy. *Anticancer research* 1985, 5 (1), 101-110.
293. Damrauer, J. S.; Studler, M. E.; Acharya, S.; Baldwin, A. S.; Couch, M. E.; Guttridge, D. Chemotherapy-induced muscle wasting: association with NF-κB and cancer cachexia. *European journal of translational myology* 2018, 28 (2).
294. Pin, F.; Barreto, R.; Couch, M. E.; Bonetto, A.; O’Connell, T. Cachexia induced by cancer and chemotherapy yield distinct perturbations to energy metabolism. *Journal of cachexia, sarcopenia and muscle* 2019, 10 (1), 140-154.
295. Porpora, P. Understanding cachexia as a cancer metabolism syndrome. *Oncoogenesis* 2016, 5 (2), e200-e200.
296. Tisdale, M. Cachexia in cancer patients. *Nature Reviews Cancer* 2002, 2 (11), 862-871.
297. Rao, P. S.; Shashidhar, A.; Ashok, C. In utero fuel homeostasis: Lessons for a clinician. *Indian journal of endocrinology and metabolism* 2013, 17 (1), 60.
298. Sturmay, R.; Smith, D. G.; Sturmay, R. Parallels between embryo and cancer cell metabolism. *Biochemical Society Transactions* 2013, 41 (2).
299. Grever, M. R.; Schepartz, S. A.; Chabner, B. A. In The National Cancer Institute: cancer drug discovery and development program, *Seminars in oncology, Elsevier* 1992; pp 622-638.
300. Askanazy, M. Die Teratome nach ihrem Bau, ihrem Verlauf, ihrer Genese und im Vergleich zum experimentellen Teratoid. *Verhandlungen der Deutschen Pathologischen Gesellschaft* 1907, 11, 39-82.
301. Seiler-Amfang, F.; Kratochwil, K. Induction and differentiation of an epithelial tumour in the newt (Triturus cristatus). *Development* 1962, 10 (3), 337-356.
302. Wang, Z.; Sun, G.; Shen, Z.; Chen, S.; Chen, Z. Differentiation therapy for acute promyelocytic leukemia with all-trans retinoic acid: 10-year experience of its clinical application. *Chinese medical journal* 1999, 112 (11), 963-967.
303. Leszczynecka, M.; Roberts, T.; Dent, P.; Grant, S.; Fisher, P. Differentiation therapy of human cancer: basic science and clinical applications. *Pharmacology & therapeutics* 2001, 90 (2-3), 105-156.
304. Demetri, G. D.; Fletcher, C. D.; Mueller, E.; Sarraf, P.; Naujoks, R.; Campbell, N.; Spiegelman, B. M.; Singer, S. Induction of solid tumor differentiation by the peroxisome proliferator-activated receptor-γ ligand troglitazone in patients with liposarcoma. *Proceedings of the National Academy of Sciences* 1999, 96 (7), 3951-3956.
305. Yan, M.; Zhang, Y.; He, B.; Xiang, J.; Wang, Z.; Wen, H. IKKβ restoration via EZH2 suppression induces nasopharyngeal carcinoma differentiation. *Nature communications* 2014, 5 (1), 1-15.
306. Zhang, Y. Cancers with Stem-Like Attractors and “Loss Of Differentiation” Novel Hallmark: Does the “Cyto-Education” with Stem Cell Therapy Help. *Journal of Genetic Syndromes and Gene Therapy* p2014, 4 (130), 6.
307. Warrell Jr, R. P.; Frankel, S. R.; Miller Jr, W. H.; Scheinberg, D. A.; Itri, L. M.; Hittelman, W. N.; Vyas, R.; Andreeff, M.; Tafur, A.; Jakubowski, A. Differentiation therapy of acute promyelocytic leukemia with retinoic (all-trans-retinoic acid). *New England Journal of Medicine* 1991, 324 (20), 1385-1393.
308. Tallman, M. S.; Andersen, J. W.; Ogden, A.; Weinstein, H.; Shepherd, L.; Willman, C. All-trans retinoic acid in acute promyelocytic leukemia: long-term outcome and prognostic factor analysis from the North American Intergroup protocol. *Blood* 2002, 100 (13), 4298-4302.
309. Finn, G. J.; Creaven, B. S.; Egan, D. A., Daphnetin induced differentiation of human renal carcinoma cells and its mediation by p38 mitogen-activated protein kinase. *Biochemical pharmacology* 2004, 67 (9), 1779-1788.
310. Riveiro, M. E.; Shayo, C.; Monczor, F.; Rossi, J.; Debenedetti, S.; Davio, C. Induction of cell differentiation in human leukemia U-937 cells by 5-oxygenated-6, 7-methylenedioxycoumarins from Pterocaulon polystachyum. *Cancer letters* 2004, 210 (2), 179-188.
311. Agarwal, C.; Sharma, Y.; Zhao, J.; Agarwal, R. A polyphenolic fraction from grape seeds causes irreversible growth inhibition of breast carcinoma MDA-MB-468 cells by inhibiting mitogen-activated protein kinases activation and inducing G1 arrest and differentiation. *Clinical Cancer Research* 2000, 6 (7), 2921-2930.