ABSTRACT

Objective: Develop a simple isocratic reverse phase high performance liquid chromatographic method (RP-HPLC) and validate for the determination of lincomycin hydrochloride (LMH) in bulk and pharmaceutical preparations.

Methods: RP-HPLC quantification was carried out by using fine pack SIL RP C18 column. The mobile phase (methanol: water) was pumped at a flow rate of 1 ml/min in the ratio of 90:10 v/v and the eluents were monitored at 254 nm.

Results: The retention time of the drug was 3.73 min and produced at a linear response in the concentration range of 5-25µg/ml. The percentage RSD was found to be below 2%. The LOD and LOQ were found to be 0.854µg/ml and 0.258µg/ml respectively.

Conclusion: Validation of the method was performed for precision, accuracy, linearity, ruggedness, specificity and sensitivity to conform to ICH guidelines for validation methods for analytical applications.

Keywords: Lincomycin hydrochloride (LMH), RP-HPLC, Validation, Recovery, Tablet dosage

INTRODUCTION

Lincomycin hydrochloride (LMH) broadly used as a systemic antibiotic, belongs to the collection of lincosamide, which is effective against maximum common gram-positive bacteria. Lincomycin hydrochloride prevents cell growth and microbial protein synthesis, by interacting powerfully and precisely with the 50S ribosomal subunit, at mutually related sites (fig. 1) [1]. Chemically LMH is a methyl 6-amino-6,8-dideoxy-N-[[2S,4R)-1-methyl-4-propylprolyl]-1-thio-D-erythro-o-D-galacto-octopyranoside hydrochloride monohydrate, antimicrobial bodies produced by Streptomyces lincolnensis var. lincolnensis or by some other means [2,3]. It is approved in Indian pharmacopoeia [4, 5], British pharmacopoeia [6], the United States of pharmacopoeia and National formulary [7]. A few analytical method have been stated for its quantitative determination in pharmaceutical formulations like GC [4-7], UV spectrophotometry [8], colorimetry [10], and atomic absorption spectroscopy [11]. In view of the above fact, specific fast and sensitive analytical methods are in need for its quantitative determination.

The purpose of the study was to improve a simple, precise, rapid and accurate reverse phase HPLC method for the estimation of lincomycin hydrochloride in bulk and pharmaceutical dosage forms.

MATERIALS AND METHODS

Reagents and chemicals

Analytically pure lincomycin hydrochloride was attained as gift sample from Wallace Pharmaceuticals Pvt Ltd, Goa, India. Methanol and water of HPLC grade obtained from Hi-media chemicals, Mumbai were used for the preparation of mobile phase. As this drug has no marketed tablet preparations yet, we have formulated the tablets by changing the ratio of polymers by most usually used excipients by keeping the strength as constant (750 mg of LMH) and estimated the drug.

Instruments used

An isocratic high pressure liquid chromatographic system consisted of the following components: HPLC equipment (Jasco, Japan) consists of model PU-2080 HPLC pumps, UV-2075 model UV/Visible detector and AS-1559 model sampler. Chromatographic analysis was performed using the chrompass software on reverse phase fine park SIL C18 T-5 column with 250 x 4.6 mm i. d and particle size 5µm.

Preparation of mobile phase and stock solutions

The mobile phase consisting of methanol:water (90:10) v/v was selected. The solution was sonicated for 5 min. 750 mg of lincomycin hydrochloride raw material was weighed and transferred to 100 ml volumetric flask and dissolved in HPLC water to give 1000 µg/ml of LMH. Lincomycin hydrochloride solutions were further diluted with mobile phase to obtain a final concentration of 1.00 µg/ml.

Chromatographic conditions

Ideal composition of mobile phase comprising of methanol: water (90:10) v/v was selected. The solution was sonicated for 5 min. 750 mg of lincomycin hydrochloride raw material was weighed and transferred to 100 ml volumetric flask and dissolved in HPLC water to give 1000 µg/ml of LMH. Lincomycin hydrochloride solutions were further diluted with mobile phase to obtain a final concentration of 1.00 µg/ml.

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Linearity of data

Suitable aliquots of lincomycin hydrochloride stock solutions were pipetted into different 10 ml standard flasks and made up to the mark with mobile phase to attain a final concentration of 5, 10, 15, 20, 25 μg/ml respectively. Triplicate dilutions of all concentration were introduced into the HPLC in duplicate. The linearity of calibration graphs and obedience of the system to beer’s law was confirmed by the high significance of correlation coefficient. Solutions were injected using a 20 μl fixed loop system and chromatograms were noted. Calibration curves were made by plotting area versus concentration and regression equation was calculated for lincomycin hydrochloride.

Assay of formulations

Twenty tablets comprising 750 mg of LMH were weighed and finely powdered. An exactly weighed powder sample equivalent to 100 mg of LMH was dissolved in HPLC water. The above solution was sonicated for 30 min and then filtered over a 0.45 μm membrane filter. An appropriate aliquot of this solution was pipetted into 100 ml standard flask, and the solution was finally made up to the mark with mobile phase. From the above solution, two concentrations were introduced and chromatograms were noted and matched with the standard.

Validation of HPLC method

The proposed HPLC method was confirmed as per ICH guidelines [12-15].

The precision of the method was analysed by performing recovery studies at 25, 50 and 75 % of the test concentrations. The precision of the method was validated by interday and intraday studies. To calculate the robustness of the established RP-HPLC method, small deliberate deviations in the optimized method parameters like the effect of a change in flow rate, mobile phase ratio and column temperature on the retention time, tailing factor, area count and percentage content of LMH were studied. The results of validation and system suitability studies are given in table 2.

RESULTS AND DISCUSSION

Method validation

Linearity

The linearity of lincomycin hydrochloride employing RP-HPLC method was constructed by plotting area of lincomycin hydrochloride against concentration. The recommended method was found to be modest and true in the concentration range of 5-25 μg/ml for lincomycin hydrochloride. It was established to be true with a correlation coefficient 0.9998 and the data is shown table 1.

Limit of detection (LOD) and limit of quantification (LOQ)

LOD is calculated by using the formula 3.3 *σ/s, where “σ” is the standard deviation of intercept and “s” is the slope obtained for calibration curve and limit of quantification was found to be 0.854 μg/ml and 0.258 μg/ml respectively. Similarly, LOQ is calculated by using the formula 10 times σ/s. The data are depicted in table 1.

Intermediate precision

The analytical method was determined by performing method precision on the three successive days by different analyst under same experimental condition by injecting six replicate standard solutions of LMH. The % RSD was found to be 0.329.

Intermediate precision

Table 2: Analysis of tablet formulation and recovery studies

| Formulation | Amount present (mg) | Amount found*(%) | Standard deviation*(SD) | % RSD* | Standard error* | Mean % recovery# |
|-------------|---------------------|------------------|-------------------------|--------|----------------|-----------------|
| Tablet 1    | 744.87              | 99.3             | 0.2651                  | 0.5017 | 0.2041         | 99.57           |
| Tablet 2    | 749.91              | 99.9             | 0.5718                  | 0.6609 | 0.1952         | 99.73           |

*Denotes average of three determinations at each level of recovery.

Table 1: Validation parameters determination during method validation for the detection and quantification of lincomycin hydrochloride

| Validation parameter* | Lincomycin hydrochloride |
|-----------------------|--------------------------|
| Linearity range (μg/ml) | 5-25 μg/ml               |
| Coefficient of correlation(r²) | 0.99986                  |
| LOD(μg/ml)±(%RSD) | 0.258                    |
| LOQ (μg/ml)±(%RSD) | 0.854                    |
| Retention time (min) | 3.73                     |
| Accuracy (%) | 99.6±0.82                |
| Precision (%RSD) | 0.352                    |
| Intraday (n=6) | 0.229                    |
| Tailing factor | 1.2                      |
| Recovery (%)(±%RSD) | 99.8                     |
| Ruggedness (% CV) | 0.24                     |
| Column I | 0.07                     |
| Column II | 0.04                     |
| Robustness | 0.04                     |
| Repeatability (RSD*,n = 6) | 0.14                     |

*The validation parameters were determined from the analysis of eight standard solutions (n=6), %RSD: Percentage relative standard deviation. (% CV): coefficient of variation. LOD: Limit of detection, LOQ: Limit of quantification.
The outcomes of robustness reading as 0.04 also showed that the method is robust and is unaffected by minor differences in the chromatographic conditions.

System suitability
The developed method was validated according to regulatory guidelines [16-19]. Results of validation and system suitability testing are summed up in table 1. The results complied with the acceptance criteria laid down under regulatory guidelines [20] with % RSD values, tailing factor, asymmetry factor and height equivalent to theoretical plates being well below the acceptable limits and correlation coefficient, and a number of theoretical plates were above the minimum acceptance criterion. System suitability parameters such as capacity factor, tailing factor and number of theoretical plates was found to be 2.8, 1.2, 4025, respectively. The retention time of lincomycin hydrochloride was found to be 3.73 min and depicted in fig. 1.

Fig. 1: A typical chromatogram of LMH (3.73 min) in tablet formulation

CONCLUSION
In the present study, LMH is determined by RP-HPLC, good linearity obtained for the drugs 5-25 µg/ml with a regression coefficient of 0.9998. The results for precision, recovery and ruggedness were within the limits. Hence, the established RP-HPLC method is simple, correct, precise and robust and can be employed effectively for the routine analysis of LMH in bulk and pharmaceutical dosage preparations.

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CONFLICTS OF INTERESTS
Declare none

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