THE INFLUENCE OF ANTI-HIV-1 SPECIFIC IgY IN INHIBITING HIV-1 INFECTION IN BINDING PHASE WITH SYNCTIUM EXAMINATION OF CD4 RECEPTOR DENSITY USING THE FLOWCYTOMETRY METHOD

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ABSTRACT

HIV/AIDS infections have increased and spread very quickly in the world, including in Indonesia. The absence of an effective vaccine and the fact that antiretroviral drugs can only suppress the progression of infection but cannot eradicate it lead to the efforts to find materials containing immunoglobulins that can replace the immune system which greatly declines in HIV/AIDS patients. The successful use of specific IgY in other studies opens up opportunities for the use of anti-HIV-1 specific IgY as passive immunotherapy. This type of research is true experimental research design with post-test only control group design. IgY was obtained from Lohmann Laying hens chicken eggs immunized with the inactivated HIV-1 virus. The concentration of IgY was determined using the Bradford method and then the characterization test was continued using the AGPT, ELISA, SDS-PAGE and Western blot tests which showed anti-HIV-1 specific IgY. The results of the test showed specific anti-HIV-1 IgY was effective in inhibiting the formation of syncytium in HIV-1 infection against CD4+ T lymphocytes in the binding phase (entry stage) in the treatment group p-value 0.047 (p <0.05). The results of CD4 receptor density tests using the Flowcytometry method showed that specific anti-HIV-1 IgY was effective in inhibiting HIV-1 infection against CD4+ T lymphocytes in the binding phase (entry stage) in the treatment group p-value 0.047 (p <0.05).

Keywords: Anti-HIV-1 Specific IgY; HIV-1; Syncytium and CD4 receptor

INTRODUCTION

There are 2 subtypes of HIV, namely HIV-1 (spreading worldwide) and HIV-2 (spreading in West Africa). In patients, HIV infection goes through four phases, namely acute, asymptomatic or latent phases, AIDS related complex (ARC) and Acquired Immune Deficiency Syndrome (AIDS). And, HIV-1 is a cytopathic virus classified in the family retroviridae, subfamily lentivirinae, genus lentivirus. HIV/AIDS infection can affect anyone with a means of transmission mainly through sexual contact, narcotics,
psychotropic and injectable addictive substances, blood transfusions and infections from mother to fetus. The HIV virus enters the body infecting cells that have the Cluster of Differentiation 4 (CD4 +) protein molecule (Kindt et al, 2007; Levinson, 2014; Merati, 2014).

Every year cases of HIV/AIDS have increased. In 2017 the number of HIV/AIDS cases in the world was 36.9 million, an increase compared to 2016 (36.7 million) (WHO, 2017, 2018). In Indonesia the region with the highest number of HIV infections was DKI Jakarta (55,099), followed by Java East (43,399), West Java (31,293), Papua (30,699) and Central Java (24,757) (DITJEN PP&PL, Dinas Kesehatan Provinsi Jawa Timur, 2017).

The high number of HIV/AIDS cases in Indonesia means a problem that must get concern from various parties, especially those in the medical field to give their real contribution both preventively and curatively in the response to HIV/AIDS. The fact that vaccines for HIV haven not been found and antiretroviral drugs that have been developed very fast growing recently are still at a level capable of suppressing the progression of the disease but have not been able to eradicate the HIV virus, substitution therapy is needed to replace the declining immune system using immunotherapy. The successful use of IgY as a specific immunoglobulin in other studies opens the opportunity for success in the application of using IgY as an anti-HIV-1 specific antibody (Nguyen et al, 2010; Rahman et al, 2013; Wen et al, 2012; Thu et al, 2017).

This study used syncytium formation and Flowcytometry tests to measure the density of CD4 receptors, so it could be known whether anti-HIV-1 specific IgY is effective in inhibiting HIV-1 infection against CD4+ T lymphocytes in the binding phase (entry stage) in the treatment group.

MATERIALS AND METHODS

Preparation of virus antigens

Sample of blood from HIV-1 patients was taken as much as 13 mL through a cubital vein for the making of vaccines and virus stocks then kept it in a vacutainer containing EDTA anti-coagulant, centrifuging the blood at 2000 rpm for 10 minutes. The plasma containing virus was collected and then activated using formaldehyde 1% (v/v) at 4°C for 5 days, then the virus that had been inactivated was injected intramuscularly into the body of a Lohmann laying hens chicken as an experimental animal.

Immunization of chicken with HIV-1 Virus and IgY characterization

Lohmann Laying chickens were intramuscularly immunized with the HIV-1 virus which has been inactivated using 1% (v/v) formaldehyde at 32oC for 5 days with a dose of 80 E1g and mixed with 0.25 E1L of Freund's Complete Adjuvant (FCA). Immunizations were repeated twice at two-week intervals with a dose of 80 E1g and mixed with 0.25 E1L Incomplete Freund's Adjuvant (IFA). Then purification is done using the PEG 6000 precipitation method. The concentration of the purified IgY was determined by Bradford method. Then IgY was characterized using AGPT, ELISA, SDS-PAGE and Western blot test which showed that the IgY tested was anti-HIV-1 specific IgY, then stored at -20°C.

Syncytium formation test

Syncytium formation test begins by measuring the MOLT4 cells. If there are 106 cells/50 E1L, then it can be used. Then HIV-1/MT4 cells are measured, and, if there are 105 cells/50 E1L, then the neutralization activity test is ready to be done. Next, antibodies (IgY) are dissolved with RPMI media according to the desired concentration, that is, the concentration obtained from the results of the Bradford test characterization by the spectrophotometer method, and a preliminary (optimization) test is carried out until the IgY concentration obtained does not cause cell death. The next step is putting 100 E1L RPMI in all wells both control and treatment groups, putting 200 E1L IgY in the first row of wells continued with outting 100 E1L RPMI in wells in the first, second and so on up to the 8th rows, then making a serial dilution by taking 100 E1L from the first row and adding it to the second row, then pipetting it taking 100 E1L from the second row and adding it to the third row. The process is repeated until it goes to the 8th row, so the concentration obtained reaches half of the previous concentration. The next step is putting 50 μL of HIV-1/MT4 cells into each of the wells in a plate already containing IgY. The plates were then incubated for 30 minutes in CO2 incubators (to see whether IgY and HIV-1/MT4 cells were bound or not) and then put 50 E1L MOLT4 cells into each well in a plate containing IgY and incubated in CO2 incubators for 24 hours. At the same time, the same process was done but without the administration of IgY as a control group. Syncytium measurement was done from the results formed under a microscope thus evaluating the anti-syncytium forming activity of anti-HIV-1 specific IgY. At the same time the same process was done to the group without treatment. Culture fluids from the results of the syncytium formation test can be used to analyze the density of CD4 receptors.
**CD4 receptor density using the flowcytometry method**

In preparing the sample for the flowcytometry test it began by preparing 1 conicle for 1 type of treatment, then labelling the conicles in each treatment group, taking a culture fluid (the results of the anti-HIV-1 specific IgY activity test using the Syncytium formation test method) and then pipetting it. The culture liquid was then centrifuged at 1500 rpm for 5 minutes, the supernatant was kept in a freezer at -80°C, whereas the pellet was used for the flowcytometry test. Cell pellets were washed with staining buffer cells 1 time and then centrifuged at 2500 rpm for 3 minutes at 40°C.

The supernatant was removed and the cell pellets formed were ready for staining with CD4 surface marker cell antibodies (5–µL per sample 1:10 diluted in staining buffer cells). The diluted antibodies were then taken as much as 50 –µL and mixed with cell pellets and homogenized. Cell pellets that had been given antibodies were incubated for 20 minutes in the dark at room temperature. After incubation the cell pellets were then added with 500 –µL staining buffer cells, the addition of staining buffer cells at this stage was a buffer without surface marker antibodies (buffer only). This stage aims to color the target bound by the marker, and then homogenized again then transferred to the cuvette and then measured with flowcytometer.

**Research method**

This was true experimental research to prove the anti-HIV-1 specific IgY neutralization activity with the syncytium formation test method and the examination of CD4 receptor density using the Flowcytometry method. The study used a post-test only control group design.

**Technique of statistical analysis**

The data obtained was then analyzed descriptively and inferentially. Data analyzed descriptively were used to determine the mean values of syncytium formation and CD4 receptor density, while the inferential analysis used were t and ANOVA Kruskal Wallis tests using the SPSS 21 application.

**RESULTS**

**Syncytium formation test**

Based on syncytium formation test for the group added with anti-HIV-1 specific IgY (as a treatment group) and the group not added with anti-HIV-1 specific IgY (as a control group) at the 24-hour observation, the number of syncytium formed turned to be more in the control group (28.95) than that in the treatment group (14.9) with p-value of 0.000 (p <0.05), meaning there were significant differences (there was a significant effect) of giving anti-HIV-1 specific IgY to the formation of Syncytium (Table 1).

**CD4 receptor density test using the flowcytometry method**

The average percentage of CD4 receptor density formed in the control group was smaller (9.69) than that in the treatment group (11.8) with a p-value of 0.047 (p<0.05), so that it could be concluded that administration of anti-HIV-1 specific IgY was effective in inhibiting HIV-1 infection in CD4+ T lymphocytes in the binding phase (entry stage) in the treatment group at 24-hour observation (Table 2).

| Table 1. Frequency distribution of formation of Syncytium in the treatment and control groups |
|---------------------------------------------------------------|
| **Group** | **N** | **Mean** | **Minimum** | **Maximum** | **p-value** |
| Control | 40 | 28.9 | 10 | 82 | 0.000 |
| Treatment | 40 | 14.9 | 4 | 45 | |

| Table 2. Frequency distribution of CD4 receptor density in the treatment and control groups |
|---------------------------------------------------------------|
| **Group** | **N** | **Mean** | **Minimum** | **Maximum** | **p-value** |
| Control | 5 | 9.69 | 8.65 | 12.502 | 0.047 |
| Treatment | 5 | 11.8 | 10.70 | 13.08 | |
DISCUSSION

The advantages of using IgY rather than mammalian IgG are that because IgY is: 1) not toxic, because eggs are common daily food consumed, 2) Non allergic, as allergic reactions generally occur due to consuming large amounts of egg whites, but IgY is soluble in water purified from egg yolks (without lipids) and it usually does not cause allergies, 3) No serological cross reactivity, i.e. IgY does not react with rheumafactors because the FC epitopes (factor of crystallin) of IgY are not recognized by the antibody binding site of rheumafactor, IgY does not activate mammalian complement, and IgY is a functional analog with mammalian IgGs so it does not cause effects beside serum sickness and shock anaphylactic, and 4) Effective alternative to antibiotics: for example, IgY can be an alternative treatment for antibiotic resistance to enteral pathogens, because it has been proven that IgY can prevent interactions between microorganisms and target cells. Therefore, the use of IgY as an anti-HIV-1 specific antibody may be applied (Larsson et al, 1991; Larsson et al, 1992; Warr et al, 1995; Carlander, 2002; Mine & Kovacs-Nolan, 2002; Karlsson et al, 2004; Schade et al, 2005; Wang et al, 2011; Rahman et al 2013; Zoerriehzahra et al, 2016).

The results of the syncytium formation test showed that anti-HIV-1 specific IgY was effective in inhibiting the formation of Syncytium in HIV-1 infection against CD4+ T lymphocytes in the binding phase (entry stage) at 24-hour observation (p <0.05). The results of this study supported by Delves et al (2011) stating that the first time the HIV-1 virus infected CD4+ T lymphocytes mediated by spike interactions (which are sharply rising deflections) of the gp120 envelope on the surface of the virus with CD4 receptors on the surface of T lymphocytes CD4+, but with the provision of anti-HIV-1 specific IgY, the antibody molecule would approach the spike and bind to it, so that no HIV-1 infection process would occur. This process is a function of the neutralizing antibody ability (inhibits the ability of the virus to infect by inhibiting viral attachment, penetration, or the release of viral envelopes, or all three processes at once). These neutralization antibody processes depend on: the number of glycoprotein molecules (spikes) from the envelope surface which are generally around 7 to 14 envelope knobs per virion, but the amount can vary, the ability of antibodies to cover and shed/release envelope protein gp120 from virion (Shedding), all envelope proteins in the virion must be targeted by at least one antibody to get enough neutralization, and the ability of

Fig. 1. Syncytium test in control group at 24 hours observation (A); Syncytium test in treatment group at 24 hours observation (B).
antibodies to penetrate hidden CD4 binding sites at gp120 (Levy 2007).

CONCLUSION
Anti-HIV-1 specific IgY is effective in inhibiting the syncytium formation in HIV-1 infection against CD4+ T lymphocytes in the binding phase (entry stage) in the treatment group. The results of the examination of CD4 receptor density using Flowcymetry method showed that anti-HIV-1 specific IgY is effective in inhibiting HIV-1 infection of CD4+ T lymphocytes in the binding phase (entry stage) in the treatment group.

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