Should rescue ICSI be re-evaluated considering the deferred transfer of cryopreserved embryos in in-vitro fertilization cycles? A systematic review and meta-analysis

Alessio Paffoni1*, Marco Reschini2, Valerio Pisaturo2, Cristina Guarneri2, Simone Palini3 and Paola Viganò2

Abstract

Background: Total fertilization failure represents a particularly frustrating condition for couples undergoing in vitro fertilization. With the aim of reducing the occurrence of total fertilization failure, intracytoplasmic sperm injection (ICSI) has become the first choice over conventional in vitro fertilization (IVF) procedures although evidence of improved results is still debated and its use in couples without male factor infertility is not recommended. Among the strategies potentially useful to promote the use of conventional IVF, we herein call attention to the late rescue ICSI, which consists in performing ICSI after 18–24 h from conventional insemination on oocytes that show no signs of fertilization. This treatment has however been reported to be associated with a low success rate until recent observations that embryos derived from late rescue ICSI may be transferred after cryopreservation in a frozen-thawed cycle with improved results. The aim of the present study was to assess whether frozen embryos deriving from rescue ICSI performed about 24 h after conventional IVF may represent a valuable option for couples experiencing fertilization failure.

Methods: A systematic review on the efficacy of late rescue ICSI was performed consulting PUBMED and EMBASE.

Results: Including twenty-two original studies, we showed that clinical pregnancy rate per embryo transfer and implantation rate obtainable with fresh embryo transfers after rescue ICSI are not satisfactory being equal to 10 and 5%, respectively. The transfer of cryopreserved rescue ICSI embryos seems to offer a substantial improvement of success rates, with pregnancy rate per embryo transfer and implantation rate equal to 36 and 18%, respectively. Coupling rescue ICSI with frozen embryo transfer may ameliorate the clinical pregnancy rate for embryo transfer with an Odds Ratio = 4.7 (95% CI:2.6–8.6).

Conclusion: Results of the present review support the idea that r-ICSI coupled with frozen embryo transfer may overcome most of the technical and biological issues associated with fresh transfer after late r-ICSI, thus possibly representing an efficient procedure for couples experiencing fertilization failure following conventional IVF cycles.

Trial registration: Prospero registration ID: CRD42021239026.

Keywords: IVF, Total fertilization failure, Rescue ICSI, Delayed ICSI, Intracytoplasmic sperm injection
Background

Intracytoplasmic sperm injection (ICSI), initially developed to treat severe male infertility, was introduced in the early 1990s as one of the most dramatic technological breakthroughs in assisted reproductive technology (ART) [1]. The technique was rapidly integrated into the routine clinical practice and is presently considered the most widely used insemination method worldwide [2]. The reliability in achieving fertilization in cases of severe male factor infertility has led to the expansion of its use also for other infertility indications. In the United States, ICSI use increased from 36% in 1996 to 76% in 2012, with the largest relative increase among ART cycles without an indication of male factor infertility. According to Zagadaïlov et al. [3], state mandates for ART coverage can encourage more restrictive use of laboratory resources. From 2000 to 2016, absolute rates of ICSI use per clinic increased by 20% in both ART-mandated (42.5 to 62.5%) and non-mandated states (46.9 to 67.6%) with statistically significant lower ICSI utilization in insurance-mandate states. An increase in ICSI use has been reported in several countries worldwide, with ICSI rate close to 100% in the Middle East [4].

Reducing the occurrence of total fertilization failure (TFF) represents the plausible reason for this “indication creep” of ICSI over conventional in vitro fertilization (IVF) procedures. Total fertilization failure represents a particularly frustrating condition for couples undergoing ART and for professionals since it results in the premature termination of the cycle. Its incidence following conventional insemination is not infrequent, being estimated to range between 5 and 20% [5]. Notably, however, evidence of improved fertilization results with the use of ICSI is still debated and strongly related to the infertility indication considered [6–9]. Furthermore, clues in support for the need to limit widespread use of ICSI stem from inconclusive data on improved post-fertilization reproductive outcomes for non-male factor infertility diagnosis and from the significantly higher rate of de novo, chromosomal abnormalities and birth defects observed in children born after ICSI compared with the rate in the general population [10]. It is therefore, not unexpected that the Practice Committee of the American Society for Reproductive Medicine and the Society for Assisted Reproductive Technology, have declared that there is insufficient evidence to suggest ICSI use in couples without male factor infertility [10].

Among the strategies potentially useful to promote the use of conventional IVF, we herein call attention to the rescue ICSI (r-ICSI), which consists in performing ICSI after 4–24 h from conventional insemination on oocytes that show no signs of fertilization. This treatment, potentially valuable in rescuing cycles with total or partial fertilization failure, has however been reported to be associated with a low success rate when performed after 24 h (late r-ICSI) [11]. Reasons underlying this low rate may include the time-dependent deterioration in oocyte quality and the loss of synchronization between endometrial growth and embryo development. To limit these deleterious effects, a r-ICSI strategy to be carried out approximately 4–8 h after conventional insemination (early r-ICSI) has been proposed, allowing to obtain higher fertilization rates [12]. Unfortunately, given its difficult implementation, poorly compatible with the organization of a laboratory, the technique is currently quite unpopular [11, 13].

Recently, a step forward in this context has derived from the observation that embryos derived from late r-ICSI may be transferred after cryopreservation in a frozen-thawed cycle with improved results [14, 15]. The strategy of cryopreservation could overcome all the technical and biological issues associated with late r-ICSI, allowing the procedure to be more frequently used in limiting the risk of TFF associated with conventional IVF cycles. Therefore, in the present systematic review, we sought to verify whether r-ICSI coupled with frozen embryo transfer may favor ART success rate of couples experiencing TFF following conventional IVF cycles.

Methods

Studies were considered for inclusion in the systematic review following the PICOC framework as follows - patients/population: couples undergoing IVF cycles; intervention: rescue (delayed) ICSI performed on the day after oocyte retrieval and TFF following conventional IVF; comparison: when possible, r-ICSI coupled with frozen embryo transfer compared to r-ICSI with fresh embryo transfer; main Outcome: clinical pregnancy rate per cycle (clinical evidence of intrauterine foetal sac); additional outcomes: fertilization rate, implantation rate, ongoing pregnancy rate, delivery rate, malformation rate according to the International Classification of Diseases 11th Revision [16]; clinical outcomes were calculated separately for fresh and frozen embryo transfers; context: r-ICSI has been reported to be associated with a low efficacy and this may be explained by the asynchrony between embryo development and endometrial receptivity.

The following search string was used in PUBMED and EMBASE on 23rd February 2021 and repeated on 19th April 2021:

(((“rescue ICSI” OR (R-ICSI)) OR (“rescue intracytoplasmic sperm injection”))) OR ("delayed ICSI") OR ("delayed intracytoplasmic sperm injection"). No restrictions were used at this stage, with the exception of the “article” publication type for EMBASE.

Two people independently screened records for inclusion and their decision was blinded to each other. A third author checked for disagreement between results.
and a decision was taken by three authors. This process was recorded through an excel spreadsheet. Reference lists cited in study reports included in the systematic review were examined in order to retrieve additional papers suitable for inclusion.

For data extraction, studies were included in the data synthesis if reporting: 1) clinical pregnancy rate per cycle after r-ICSI performed on the day after oocyte retrieval; 2) indication regarding fresh or frozen embryo transfer; 3) results published in full in English language. The following data were extracted: First Author, publication ID, year of publication, period of recruitment, study design, mean age of included women, inclusion of cases with total or partial fertilization failure, fertilization rate with conventional IVF, number of cycles included, time of r-ICSI after conventional insemination, use of sperm from previous day or freshly collected, number of oocytes treated with r-ICSI, r-ICSI fertilization rate and abnormal fertilization rate, number of embryos obtained, number of fresh embryos transferred, number of transferred frozen embryos, technique of cryopreservation, strategy of endometrial preparation, number of pregnancies, number of newborns, number of newborns with malformations. Those data were recorded in an excel spreadsheet and were used to calculate the main outcomes and to account for possible heterogeneity among studies. Studies including a comparison between r-ICSI cycles with fresh and frozen embryo transfer were also considered for quantitative evaluation. If not clearly indicated, the number of inseminated or fertilized oocytes and the number of transferred or obtained embryos were calculated using available data such as mean values or rates. Two reviewers collected data from each report working independently; disagreements between data collectors were resolved with the intervention of a third reviewer and collegial discussion.

Quality assessment of included papers was performed using the “JBI critical appraisal checklist for cross sectional studies” [17], an evaluation tool developed to evaluate representativeness and reliability of studies. Each of 8 criteria was assessed (Yes, No, Unclear, Not applicable) by two independent reviewers and disagreements were solved in a collegial discussion with a third reviewer after reconsidering the following aspects: criteria for inclusion in the sample, description of basal characteristics of patients/cycles, methodological definition of r-ICSI, identification of confounding factors and strategies to deal with them, definition of the outcomes including pregnancy rate, use of statistical analysis.

A narrative and tabular synthesis was used for presenting the outcomes. Confidence intervals of proportions for the narrative synthesis were obtained with a binomial exact calculation. Clinical outcomes (fertilization rate, pregnancy rate per embryo transfer/cycle, implantation rate) were synthesised with the inverse-variance method. Odds ratios (OR) were obtained for case/control studies comparing fresh and frozen cycles using the Mantel-Haenszel method. A visual synthesis of results was obtained with forest plots. Analysis and figures were done with R packages [18]. Studies with missing values were excluded from the synthesis of the specific outcome. With the main goal to estimate the mean effect in a range of studies, a random-effect model was selected to conduct the meta-analysis; in case of low inconsistency ($I^2 < 30\%$), results obtained with a fixed model were also included. $r^2$ was reported as a measure of heterogeneity among studies.

**Results**

Studies were identified and selected for inclusion in the review as reported in the flowchart (Fig. 1). Out of 89 initially retrieved studies through PUBMED/EMBASE search and reference lists, 22 were finally included [12, 14, 15, 19–37]. The key characteristics and results of the studies are summarized in Table 1. Eight case-reports were used for qualitative synthesis of data regarding late r-ICSI but were excluded from pregnancy and implantation rates calculation [21, 25–27, 29, 30, 34, 35]. Fourteen retrospective studies were included in the quantitative synthesis [12, 14, 15, 19, 20, 22–24, 28, 31–33, 36, 37] and three studies were also used to calculate the OR for pregnancy and implantation rates between frozen and fresh embryo transfer after r-ICSI [14, 15, 22]. Main results of selected studies including transfer of fresh and frozen rescue ICSI embryos are reported in Tables 2 and 3, respectively. An additional table shows the critical appraisal of included studies according to the Joanna Briggs Institute checklist (see Additional file 1).

As reported in Table 1, a total of $n = 1686$ late r-ICSI cycles with $n = 12,945$ inseminated oocytes were reported in 22 studies. Rescue ICSI was performed after 15–24 h from initial conventional IVF. The number of clinical pregnancies following r-ICSI were reported to be $n = 83$ in fresh cycles and $n = 149$ in frozen cycles with $n = 65$ and $n = 121$ ongoing pregnancies/deliveries, respectively. The rate of r-ICSI on the total of conventional IVF cycles was available in 7 retrospective studies [12, 14, 22, 24, 28, 31, 37] and was equal to 3.1% (95%CI: 3.0–3.3%).

In the eight case-reports, a total of nine r-ICSI cycles performed >18 h after the conventional IVF insemination using $n = 71$ oocytes was reported. Age of included women ranged between 28 and 42 years. The cumulative normal fertilization rate was 63.4% (95%CI: 52–74%). Eighteen fresh embryos were replaced in seven embryo transfers between day 3 and 6 after oocyte retrieval; ten embryos implanted in $n = 6$ patients with an implantation rate equal to 56% (95%CI: 34–75%). In one case-
report study, embryo transfer was not performed as the cytogenetic analysis demonstrated the absence of euploid embryos [27]. The delivery of at least one baby was reported in five studies [21, 25, 29, 34, 35]; the newborns were healthy and no malformations were reported. Two studies reported two successful frozen embryo transfers; one of them was performed with \( n = 4 \) slow-frozen embryos [26] and the other with one vitrified embryo [35]. Both pregnancies resulted in the delivery of a healthy baby.

In the 14 retrospective cohort studies, a total of \( n = 1677 \) r-ICSI cycles (range 3–625 cycles per study), performed in women with a mean age ranging from 31.1 to 36.7 years, were included [12, 14, 15, 20, 22–24, 28, 31, 33–37]. Rescue cycles were performed using \( n = 12,874 \) unfertilized oocytes (range 20–4824 per study) 15–24 h after conventional IVF resulting in total or partial fertilization failure. Normal fertilization rate (2 pronuclei) in individual studies ranged between 30 and 92% with a cumulative effect size equal to 54% (95% CI: 48–60%; \( I^2 = 95\% \), \( t^2 = 0.17 \)) estimated on a total of \( n = 8881 \) r-ICSI oocytes (the forest plot of fertilization rate is available in the Additional file 2). The rate of abnormal fertilization (1 or 3 pronuclei) was reported in five studies [12, 15, 19, 20, 23] and ranged between 5 and 9% of r-ICSI oocytes. In the majority of the studies, r-ICSI was performed using the sperm sample collected on the previous day; one study found a higher rate of normally fertilized oocytes using freshly collected sperm cells (51%) compared to 1-day-old-spermatozoa (36%) [18]. Twelve studies reported a total of \( n = 1031 \) women undergoing \( n = 879 \) fresh embryo transfers on day 3 or 4 after oocyte retrieval with \( n = 2024 \) embryos [12, 14, 15, 20, 22–24, 28, 31–33, 36]; the implantation rate ranged between 0 and 11% with an effect size equal to 5% (95% CI: 3–7%). Seventy-six clinical pregnancies were obtained corresponding to a clinical pregnancy rate per r-ICSI cycle ranging between 0 and 17% and an overall effect size equal to 10% (95% CI: 7–15%). A quantitative synthesis of implantation and clinical pregnancy rates per embryo transfer in fresh cycles is summarized in Fig. 2.

Fifty-nine ongoing/deliveries pregnancies were reported. Among fifty-three newborns from fresh transferred r-ICSI embryos, no malformations were reported; two terminations for trisomy 21 (ICD-11: LD40.0) and congenital eye abnormality (ICD-11: LA10) were
As reported in Table 2, viable embryos can not be obtained with late r-ICSI in a proportion of women up to 38%; the crude incidence of cases without viable embryos in retrospective studies was 14% (95%CI: 11–16%; n = 107/785). Five studies provided results on the transfer of cryopreserved embryos following r-ICSI; in three studies, supernumerary embryos were slow-freezed after fresh embryo transfer [14, 15, 22] and in the remaining two studies, cryopreservation was elective and performed

| First Author, year [ID] | Country | Years recruitment | Source Type | Type of study | total (TFF) or partial (PFF) fertilization failure | total number of IVF cycles | female age mean (±SD) | n° of r-ICSI cycles | n° of r-ICSI oocytes | timing of r-ICSI post IVF (h) | Sperm |
|--------------------------|---------|-------------------|-------------|--------------|-----------------------------------------------|---------------------------|----------------------|------------------|------------------|----------------------------|------|
| Lundin K, 1996 [19]      | Sweden  | < 1995            | Ref         | R            | TFF and PFF                                  | 57                        | 450                  | 20–22            | P,F              |                           |      |
| Morton PC, 1997 [20]     | Usa     | 1993–1996         | Ref         | R            | TFF                                           | 35.3 ± 4.3                | 54                   | 489              | 20–24            | P              |      |
| Bussen, 1997 [21]        | Germany | < 1997            | Ref         | CR           | PFF                                           | 32                        | 1                    | 6                | 20               | P              |      |
| Yuzpe AA, 2000 [22]      | Canada  | 1997–1999         | Pub/Emb     | R            | TFF and PFF                                  | 34.4 ± 4.0                | 32                   | 234              | 19–22            | P              |      |
| Park KS, 2000 [23]       | Korea   | < 2000            | Ref         | R            | TFF and PFF                                  | 31.7 ± 1.6                | 17                   | 68               | > 18             | P              |      |
| Kuczyński W, 2002 [24]   | Poland  | 1996–2000         | Pub/Emb     | R            | TFF                                           | 32.9 ± 5.0                | 120                  | 779              | 18–20            | P              |      |
| Chian RC, 2003 [25]      | Canada  | < 2003            | Pub/Emb     | CR           | TFF                                           | 29                        | 1                    | 4                | > 18             | P              |      |
| Lombardi E, 2003 [26]    | Argentina | 1998        | Pub/Emb     | CR           | PFF                                           | 36                        | 1                    | 12               | 20               | P              |      |
| Chen C, 2003 [12]        | Singapore | 1997–1998   | Pub/Emb     | R            | TFF                                           | 35.2 ± 4.1                | 20                   | 182              | 22               |                |      |
| Pehlivan T, 2004 [27]    | Spain   | < 2003            | Pub/Emb     | CR           | PFF                                           | 35                        | 1                    | 11               | 21               |                |      |
| Amarin ZO, 2005 [28]     | Saudi Arabia | 1995–2001  | Pub/Emb     | R            | TFF                                           | 32.6                      | 78                   | 616              | > 18–24          | P              |      |
| DelUgarte CM, 2006 [29]  | USA     | < 2005            | Pub/Emb     | CR           | TFF                                           | 42                        | 1                    | 7                | 20               | P              |      |
| Esfandiar N, 2008 [30]   | Canada  | 2007              | Pub/Emb     | CR           | PFF                                           | 28                        | 1                    | 8                | 19               |                |      |
| Sermondade N, 2010 [15]  | France  | 2004–2009         | Pub/Emb     | R            | TFF                                           | 35.5 ± 3.6                | 17                   | 127              | > 18             | P              |      |
| Shalom-paz E, 2011 [31]  | Canada  | 1999–2008         | Pub/Emb     | R            | TFF                                           | 35.5 ± 4.5                | 92                   | 883              | 16–18            | P              |      |
| Xiong S, 2011 [32]       | China   | 2009              | Pub/Emb     | R            | TFF                                           | 35.3 ± 3.2                | 3                    | 20               | 20               | P              |      |
| Zhu L, 2011 [33]         | China   | 2007–2009         | Emb         | R            | TFF                                           | 31.3 ± 5.3                | 16                   | 98               | 20–22            | P              |      |
| Ming L, 2012 [14]        | China   | 2006–2011         | Pub/Emb     | R            | TFF                                           | 31.1–33.2 ± 4.3           | 534                  | 4824             | > 16–18          | P              |      |
| Singh N, 2013 [34]       | India   | < 2010            | Pub/Emb     | CR           | TFF                                           | 32                        | 1                    | 4                | > 18             | P              |      |
| Moon JH, 2015 [35]       | Canada  | < 2014            | Pub/Emb     | CR           | TFF                                           | 28.5                      | 2                    | 19               | 21               |                |      |
| Sachdev NM, 2016 [36]    | USA     | 2003–2015         | Pub/Emb     | R            | TFF and PFF                                  | 36.7 ± 4.3                | 12                   | 111              | 15–18            |                |      |
| Li M, 2021 [37]          | China   | 2013–2016         | Pub/Emb     | R            | TFF                                           | 31.4–32.4 ± 4.7           | 625                  | 3993             | > 16–19          | P              |      |

Empty cells = missing values; r-ICSI rescue-ICSI; Pub Pubmed; Emb Embase, Ref cited references; R retrospective/cross-sectional; CR case-report; P previous day; F fresh

recorded [14, 31].
| First Author, year [ID] | 2PN(%) | n° patients | Patients without viable embryos | n° ET | n° of obtained embryos | n° of fresh embryos transferred | developmental stage | n° clinical pregnancies | n° ongoing pregnancies or deliveries | n° embryos impacted | n° malformations/newborns |
|-------------------------|--------|-------------|---------------------------------|-------|------------------------|-------------------------------|-------------------|----------------------|-------------------------------|----------------------|-----------------------------|
| Lundin K, 1996 [19]     | 46.9   | 48          | 18                              | 29    | 57                     | cleavage                      | 2                 | 2                    | 2                             | 0                    | Q’2                         |
| Morton PC, 1997 [20]    | 44.0   | 54          | 5                               | 48    | 164                    | 140                           | cleavage                      | 8                   | 8                   | 15                           | 0                    | Q’15                        |
| Bussen, 1997 [21]       | 50.0   | 1           | 0                               | 1     | 2                     | cleavage                      | 1                 | 1                    | 1                             | 0                    |                             |
| Yuzpe AA, 2000 [22]     | 60.2   | 32          | 3                               | 27    | 89                     | cleavage                      | 5                 | 3                    | 6                             | 0                    | Q’4                         |
| Park KS, 2000 [23]      | 47.1   | 17          |                                 | 1     | 4                     | 4                             | cleavage                      | 0                   | 0                    | 0                             | 0                    |                             |
| Kuczyński W, 2002 [24] | 30.4   | 120         |                                 | 100   | 166                    | cleavage                      | 0                 | 0                    | 0                             | 0                    |                             |
| Chian RC, 2003 [29]     | 100    | 1           | 0                               | 1     | 4                     | 3                             | cleavage                      | 1                   | 1                    | 2                             | 0                    |                             |
| Chen C, 2003 [12]       | 91.7   | 20          | 0                               | 20    | 73                     | 58                            | cleavage                      | 1                   | 1                    | 1                             | 0                    | Q’1                         |
| Pehlivan T, 2004 [27]   | 36.4   | 1           | 1                               | 0     | 4 (a)                  | 0                             | cleavage                      | 0                   | 0                    | 0                             | 0                    |                             |
| Amarín ZO, 2005 [28]    | 50.9   | 78          | 14                              | 64    | 208                    | 174                           | cleavage                      | 4                   | 3                    | 4                             | 0                    | Q’3                         |
| DeUgarte CM, 2006 [29]  | 42.9   | 1           | 0                               | 1     | 3                     | 1                             | blastocyst                    | 1                   | 1                    | 1                             | 0                    |                             |
| Esfandiari N, 2008 [30] | 87.5   | 1           | 0                               | 1     | 6                     | 6                             | cleavage                      | 1                   | 1                    | 3                             | 0                    |                             |
| Sermondade N, 2010 [15] | 60.9   | 17          |                                 | 15    | 55                     | 35                            | cleavage                      | 1                   | 1                    | 1                             | 0                    |                             |
| Shalom-Paz E, 2011 [31] | 56.2   | 92          | 0                               | 92    | 278                    | cleavage                      | 15                              | 10                  | 17                             | 0                    | Q’12 (b)                    |
| Xiong S, 2011 [32]      | 87.7   | 3           | 1                               | 2     | 3                     | cleavage                      | 0                              | 0                   | 0                             | 0                    |                             |
| Zhu L, 2011 [33]        | 52.0   | 16          | 4                               | 12    | 21                     | 20                            | cleavage                      | 0                   | 0                    | 0                             | 0                    |                             |
| Ming L, 2012 [14]       | 45.0   | 534         | 62                              | 469   | 1416                   | 1000                          | cleavage                      | 40                  | 31                   | 45                           | 0                    | Q’16 (c)                    |
| Singh N, 2013 [14]      | 75     | 1           | 0                               | 1     | 3                     | 3                             | cleavage                      | 1                   | 1                    | 1                             | 0                    |                             |
| Moon JH, 2015 [13]      | 63.2   | 2           | 0                               | 2     | 7                     | 2                             | blastocyst                    | 2                   | 1                    | 2                             | 0                    |                             |

Empty cells = missing values; 2PN = 2 pronuclei; ET = Embryo Transfer
a) no suitable embryos after genetic screening for aneuploidies; b) +1 termination for Down syndrome; c) +1 termination for eye defect
with vitrification [36, 37]. The cumulative percentage of cycles with cryopreservation of supernumerary embryos was $n = 415/1220$ (34%, 95% CI: 32–37%). In the study with elective embryo vitrification coupled with preimplantation genetic testing (PGT) [36], the rate of patients receiving euploid embryos was $n = 3/12$ (25%, 95% CI: 9–53%); in the second study with elective embryo vitrification, the rate of patients obtaining viable embryos was $n = 406/625$ (65%, 95% CI: 61–69%).

A total of $n = 406$ embryo transfers with frozen/thawed embryos were performed in a cohort of $n = 415$ patients. The number of embryos transferred was $n = 781$ (mean number of embryos per transfer = 1.9) and the implantation rate, excluding the study with PGT showing 100% implantation rate with 3 transferred embryos [36], ranged between 13.3 and 23.7%. The clinical pregnancy rate per started frozen cycle ranged between 25.0 and 50.0%. The quantitative synthesis showed an implantation rate per embryo transfer equal to 36% (95% CI: 30–40) and the clinical pregnancy rate for r-ICSI embryos at the cleavage stage are below the competency values proposed by the ESHRE Consensus for ICSI cycles (60 and 25%, respectively) [38] and that the overall success rate is in the range of ‘futility’ or ‘very poor prognosis’ according to the Ethics Committee of the American Society for Reproductive Medicine [39]. Conversely, results deriving from cryopreservation of embryos obtainable by r-ICSI deserve some attention. Indeed, according to the present results, the change in the procedure allows ameliorate the clinical pregnancy rate for embryo transfer with an OR = 4.7 (95% CI: 2.6–8.6) and the implantation rate with an OR = 3.3 (95% CI: 2.0–5.6). Undoubtedly, it has to be recognized that only five studies produced data on r-ICSI coupled with frozen embryo transfer and all of them were retrospective observational studies. No randomized clinical trial is currently available. However, all the studies consistently reported acceptable success rates and the demonstration of an effect size greater than 3 or 4, as for implantation and clinical pregnancy rates, respectively, may be considered worthwhile, taking into account that observational studies are often not able to assess weak associations [40].

Even considering the lower limits of confidence intervals and therefore the statistical variability linked to the characteristics of the studies, results remain of interest with clinical pregnancy rates per embryo transfer of about 30%. Nevertheless, given the retrospective nature of the studies, we cannot exclude the presence of possible selection biases and residual confounding factors that may have led to incorrect interpretation of causal associations.

Similar considerations may be applied to the three studies reporting results of the comparison between fresh and frozen embryo transfers after r-ICSI. On the other hand, it has to be noted that, in these studies, transfers with cryopreserved embryos were carried out with residual embryos following a fresh embryo transfer from the same r-ICSI cycles. Therefore, even though the experimental design is not based on a randomization, it is still an intra-patient model of some clinical interest.

Another possible limit of the studies considered is that they might have included a very selected cohort of patients and/or embryos. Indeed, it is possible that unsuccessful events were not published and that patients who achieved pregnancies with frozen r-ICSI cycles may be overrepresented. Similarly, although speculative, embryos that succeeded in implantation might have derived from a very selected cohort of oocytes with exceptionally high quality and developmental potential therefore only marginally affected by in vitro ageing. There are at least three relevant observations to consider in this regard. First, embryos judged to have the highest probability of implantation are generally transferred in fresh cycles and it is therefore plausible that the cryopreserved supernumerary embryos were not the top quality embryos of the reported cycle cohort. Despite this, they showed a
Table 3 Main results of selected studies including transfer of cryopreserved rescue-ICSI embryos

| First Author, year [ID] | Embryo cryopreservation, indication | Technique of cryopreservation | n° of patients | n° Endometrial preparation | n° ET | n° of frozen embryos transferred | Developmental stage | n° Embryos implanted | n° clinical pregnancies | n° ongoing pregnancies or deliveries | n° malformations/newborns |
|-------------------------|----------------------------------|-------------------------------|----------------|---------------------------|------|-------------------------------|-------------------|-------------------|-------------------------|-------------------------------|--------------------------|
| Yuzpe AA, 2000 [22]     | Supernumerary                     | Slow freezing                 | 2              | P                         | 2    | 7                             | cleavage          | 1                 | 1                       | 1                             | 0/1                      |
| Lombardi E, 2003 [26]   | Supernumerary                     | Slow freezing                 | 1              | P                         | 1    | 5                             | cleavage          | 1                 | 1                       | 1                             | 0                        |
| Sermondaide N, 2010 [15]| Supernumerary                     | Slow freezing                 | 5              |                           | 5    | 12                            | cleavage          | 2                 | 2                       | 1                             | 0/1                      |
| Ming L, 2012 [14]       | Supernumerary                     | Slow freezing                 | 64             | N/P                       | 64   | 165                           | cleavage          | 22                | 19                      | 15                            | 0/12                     |
| Moon JH, 2015 [35]      | Supernumerary                     | Vitrification                 | 1              | P                         | 1    | 1                             | blastocyst        | 1                 | 1                       | 1                             | 0                        |
| Sachdev NM, 2016 [36]   | Elective PGT                      | Vitrification                 | 12             | N/P                       | 3    | 3                             | blastocyst        | 3                 | 3                       | 3                             | 0/3                      |
| Li M, 2021 [37]         | Elective                          | Vitrification                 | 332            | N/P                       | 332  | 594                           | cleavage/blastocyst| 141               | 122                     | 99                            | 1/109                     |

Empty cells missing values; ET embryo transfer; N Natural; P Programmed, PGT pre implantation genetic testing
higher implantation potential compared to the fresh counterparts. Second, based on studies with elective cryopreservation of embryos [36, 37], we can estimate in higher than 50% the proportion of women who actually succeeded in obtaining viable embryos for cryopreservation following r-ICSI. This data could have probably been influenced by the in vitro selection of embryos achievable through the culture up to the blastocyst stage; in fact, it has been reported that frozen r-ICSI embryos transferred at the blastocyst stage have a statistically significant higher implantation potential compared to the cleavage stage (41% versus 12%, respectively) [37]. Third,

Fig. 2 Quantitative synthesis of the studies reporting clinical pregnancy (upper panel) and implantation rate (lower panel) per embryo transfer in fresh cycles following rescue-ICSI performed 15–24 h after conventional IVF resulting in total or partial fertilization failure.

Fig. 3 Quantitative synthesis of the studies reporting clinical pregnancy (upper panel) and implantation rate (lower panel) per embryo transfer in frozen cycles following rescue-ICSI performed 15–24 h after conventional IVF resulting in total or partial fertilization failure. Cases with preimplantation genetic testing were excluded.
in the included studies, age and other variables as potential confounding factors have been controlled by intra-patient comparisons and should not have strongly impacted.

The available data are also limited by the lack of relevant clinical information since obstetric and perinatal findings were often not reported. Less than 180 births have been described so far deriving from both fresh and frozen cycles using r-ICSI; two miscarriages due to malformations and no relevant health problems in newborns with the exception of one case of microtia were reported. Therefore, although the cohort of babies born from this procedure is limited, present results do not suggest an increase of adverse outcomes following its application, including malformation rates.

Collectively, even considering the reported limits of the considered studies, the present findings highlight the consistent improvement in the success rate using frozen-thawed embryo transfer after late r-ICSI cycles. This observation has important implications for clinical embryologists. The opportunity to rely on a rescue procedure with satisfactory chances of success could entice the operators to a greater use of the conventional IVF technique. An excessive use of ICSI, aiming at preventing cases of TFF, is thought to have negative consequences both on the overall probability of pregnancy and on the safety of the procedures with higher costs and increased laboratory workload. According to the results of a recent systematic review [6], TFF risk is significantly increased after conventional IVF insemination compared to ICSI (relative risk = 2.63, 95%CI: 1.29–5.35) in couples with non-male factor infertility; on the contrary, overall fertilization rates are not significantly improved with the use of ICSI and clinical pregnancy rates are even higher using conventional IVF. Similarly, a previous Cochrane review confirmed that conventional IVF gives better fertilization results than ICSI in couples with male factor subfertility and also suggested that pregnancy, miscarriage or live-birth rates after conventional IVF and ICSI are comparable for couples with non-male subfertility. If anything, ICSI does not improve the success rate in these couples [41]. Even if results from a recent meta-analysis favour the use of ICSI to increase fertilization rates and decrease the risk of TFF in couples with well defined unexplained infertility, no data on the impact on clinical pregnancy and live birth rates have been provided in this publication [8]. A recent randomised clinical trial failed to demonstrate an advantage of ICSI compared to conventional IVF in couples without male factor indication in terms of total fertilization failure, live birth and implantation rates [7]. Since age has been correlated with zona pellucida thickening, ICSI has been proposed for improving ART outcomes in older patients [42]; however, a recent prospective randomized controlled trial comparing conventional insemination versus ICSI on sibling oocytes in advanced maternal age patients showed similar fertilization rate, average number of cleavage stage and average top-quality embryos between the two groups (9). Given this scenario, it is essential to illustrate any beneficial role of r-ICSI in order to increase the confidence of embryologists in proceeding with conventional IVF.

Late r-ICSI can be easily implemented in ART laboratories as it can be carried out the day after oocyte retrieval and it is not difficult to fit it timely in the context of the laboratory process while this is often the case for early r-ICSI. Rescue ICSI performed as early as 6 h after in vitro insemination has been similarly proposed as an interesting treatment option to avoid a complete failure of conventional IVF [12, 13]. Overall, according to the
review by Beck-Fruchter et al., a pregnancy rate of 44% can be achieved following the application of this procedure in cases of TFF [11]. Despite these encouraging results, early r-ICSI is still sporadically used and generally in laboratories located in China. The underlying plausible reason for this unpopularity relies on the organization problems that may arise in implementing this procedure in the context of the routine activities of standard IVF laboratories. It is therefore possible that late r-ICSI may find a greater consensus among operators if it proves to be equally efficient, even though it entails the need to apply the elective cryopreservation of the embryos. Cryopreservation procedures are currently well integrated among IVF laboratory treatments and can be routinely organized without very strict time requirements. Elective embryo freezing also allows conducting genetic testing in order to verify whether there is a concrete risk of genetic anomalies linked to the extension of the culture time between oocyte retrieval and insemination, as suggested in very preliminary case-reports [27, 29].

Of utmost interest, benefits of cryopreservation and transfer in subsequent cycles may explain the discrepancy between results derived from fresh or frozen/thawed embryo transfers. Indeed, some degree of asynchrony between embryo developmental stage and endometrial receptivity window may occur following fresh transfer of cleavage stage embryos derived from late r-ICSI [43]. Most of the reports indeed described results from transferring embryos at the cleavage stage [11]. Although endometrial receptivity is thought to have an extraordinary plasticity so that embryos could implant regardless of their precise phase of development (e.g., a cleavage-stage embryo could implant in an endometrium theoretically set to receive a blastocyst), we cannot exclude that small perturbations at the opening of the window of implantation may have a detrimental role.

Some critical variables may potentially affect the efficacy of late r-ICSI but, unfortunately, data are currently poorly available in this context. Among those variables, it is worth citing the following: 1) the use of 1-day old or freshly prepared spermatozoa; only one study compared the two types of ejaculate, suggesting that fresh spermatozoa are associated with higher fertilization rates. In all other reports, sperm used was collected on the day before. Since sperm quality and aging could explain, at least in part, observed fertilization rates which were found to be lower compared to standard ICSI cycles, this issue remains to be clarified through reliable information on the genetic and metabolic quality of spermatozoa after a 24-h incubation; 2) the rate of immature or nearly mature oocytes available at the time of conventional IVF. It is well known that the evaluation of nuclear maturity of oocytes still in their cumulus cells can be demanding and that metaphase I oocytes could benefit from in vitro culture until the day after retrieval in order to gain competence to undergo fertilization. For this reason, we cannot exclude that some positive results of late r-ICSI may be due to oocyte maturation rather than to the fertilization technique; 3) timing of oocyte retrieval after ovulation triggering and exposure of oocytes to sperm cells during conventional IVF insemination; both these aspects may in fact influence the rate of mature oocytes during the insemination window; 4) the specific freezing procedure. The vast majority of results using r-ICSI coupled with embryo freezing were obtained using the slow freezing procedure. Since vitrification is acquiring increasing popularity worldwide as a more efficient technique [44], we may assume that, in the near future, r-ICSI data will be positively influenced by the employment of vitrification. Finally, it has to be mentioned that some laboratories prefer to extend the culture of apparently unfertilized oocytes to the next day, in order not to discard viable embryos deriving from misclassified zygotes. The employment of r-ICSI should not imply some changes in this approach since, at 24 h after insemination, some evidence of fertilization should already be present. The use of time-lapse may represent a valid option [45].

In conclusion, the results of this review support the idea that r-ICSI coupled with frozen embryo transfer may represent an efficient procedure for couples experiencing TFF following conventional IVF cycles. The strategy of embryo cryopreservation seems to overcome most of the technical and biological issues associated with a fresh transfer after late r-ICSI. Data derived from embryo vitrification instead of slow freezing will provide a definitive answer on this topic.

Abbreviations
ART: Assisted reproductive technologies; ICSI: Intracytoplasmic sperm injection; IVF: In-vitro fertilization; OR: Odds Ratio; PFF: Partial fertilization failure; r-ICSI: rescue-ICSI; TFF: Total fertilization failure.

Supplementary Information
The online version contains supplementary material available at https://doi.org/10.1186/s12958-021-00784-3.

Additional file 1. JBI critical appraisal checklist for analytical cross sectional studies. The evaluation of included studies made according to the Joanna Briggs Institute.

Additional file 2. Fertilization rate in late r-ICSI cycles. Effect size of fertilization rate (2 pronuclei) in included studies.

Acknowledgements
Not applicable.

Authors’ contributions
A.P. and P.V. designed the study and drafted the manuscript. V.P., S.P., C.G. participated in the collection of the literature. All Authors participated in the critical evaluation of the literature. A.P. and M.R. performed statistical

Paffoni et al. Reproductive Biology and Endocrinology (2021) 19:121
analyses. A.P., P.V. and V.P. finished the manuscript. All authors read and approved the final manuscript.

**Authors’ information**

All authors are involved in clinical embryology activities.

**Funding**

No specific financial or non-financial support was received for the review.

**Availability of data and materials**

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

**Declarations**

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Competing interests**

All authors declare no competing interest.

**Author details**

1Infertility Unit, ASST Lariana, Cantù, Como 22063, Italy. 2Infertility Unit, Fondazione IRCCS Ca’ Granda Ospedale Maggiore Policlinico, Milan 20122, Italy. 3IVF Unit, AUSL Romagna Cattolica Hospital, 47841 Cattolica, Italy. 49(6):101706. https://doi.org/10.1016/j.jogoh.2020.101706.

**References**

1. Rubino P, Viganò P, Luddi A, Piomboni P. The ICSI procedure from past to future: a systematic review of the more controversial aspects. Hum Reprod Update. 2016;22(6):650. https://doi.org/10.1093/humupd/dmw050.

2. Haddad M, Stewart J, Xie P, Cheung S, Trout A, Keating D, et al. Thoughts on the popularity of ICSI. J Assist Reprod Genet. 2021; https://doi.org/10.1007/s10815-020-01987-0.

3. Zagallov P, Seifer DB, Shan H, Zarek SM, Hsu AL. Do state insurance mandates alter ICSI utilization? Reprod Biol Endocrinol. 2020;18(1):33. https://doi.org/10.1186/s12958-020-00589-w.

4. Boulet SL, Mehta A, Kazemis EM, Kawwass JF, Jamieson DJ. Trends and analyses. A.P., P.V. and V.P. finished the manuscript. All authors read and approved the final manuscript.

5. Authors' information

All authors are involved in clinical embryology activities.

6. Funding

No specific financial or non-financial support was received for the review.

7. Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

8. Declarations

**Ethics approval and consent to participate**

Not applicable.

9. Consent for publication

Not applicable.

10. Competing interests

All authors declare no competing interest.

11. Author details

1Infertility Unit, ASST Lariana, Cantù, Como 22063, Italy. 2Infertility Unit, Fondazione IRCCS Ca’ Granda Ospedale Maggiore Policlinico, Milan 20122, Italy. 3IVF Unit, AUSL Romagna Cattolica Hospital, 47841 Cattolica, Italy. 49(6):101706. https://doi.org/10.1016/j.jogoh.2020.101706.

**References**

1. Rubino P, Viganò P, Luddi A, Piomboni P. The ICSI procedure from past to future: a systematic review of the more controversial aspects. Hum Reprod Update. 2016;22(6):650. https://doi.org/10.1093/humupd/dmw050.

2. Haddad M, Stewart J, Xie P, Cheung S, Trout A, Keating D, et al. Thoughts on the popularity of ICSI. J Assist Reprod Genet. 2021; https://doi.org/10.1007/s10815-020-01987-0.

3. Zagallov P, Seifer DB, Shan H, Zarek SM, Hsu AL. Do state insurance mandates alter ICSI utilization? Reprod Biol Endocrinol. 2020;18(1):33. https://doi.org/10.1186/s12958-020-00589-w.

4. Boulet SL, Mehta A, Kazemis EM, Kawwass JF, Jamieson DJ. Trends and analyses. A.P., P.V. and V.P. finished the manuscript. All authors read and approved the final manuscript.

5. Authors' information

All authors are involved in clinical embryology activities.

6. Funding

No specific financial or non-financial support was received for the review.

7. Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

8. Declarations

**Ethics approval and consent to participate**

Not applicable.

9. Consent for publication

Not applicable.

10. Competing interests

All authors declare no competing interest.

11. Author details

1Infertility Unit, ASST Lariana, Cantù, Como 22063, Italy. 2Infertility Unit, Fondazione IRCCS Ca’ Granda Ospedale Maggiore Policlinico, Milan 20122, Italy. 3IVF Unit, AUSL Romagna Cattolica Hospital, 47841 Cattolica, Italy. 49(6):101706. https://doi.org/10.1016/j.jogoh.2020.101706.

**References**

1. Rubino P, Viganò P, Luddi A, Piomboni P. The ICSI procedure from past to future: a systematic review of the more controversial aspects. Hum Reprod Update. 2016;22(6):650. https://doi.org/10.1093/humupd/dmw050.

2. Haddad M, Stewart J, Xie P, Cheung S, Trout A, Keating D, et al. Thoughts on the popularity of ICSI. J Assist Reprod Genet. 2021; https://doi.org/10.1007/s10815-020-01987-0.

3. Zagallov P, Seifer DB, Shan H, Zarek SM, Hsu AL. Do state insurance mandates alter ICSI utilization? Reprod Biol Endocrinol. 2020;18(1):33. https://doi.org/10.1186/s12958-020-00589-w.

4. Boulet SL, Mehta A, Kazemis EM, Kawwass JF, Jamieson DJ. Trends and analyses. A.P., P.V. and V.P. finished the manuscript. All authors read and approved the final manuscript.

5. Authors' information

All authors are involved in clinical embryology activities.

6. Funding

No specific financial or non-financial support was received for the review.

7. Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

8. Declarations

**Ethics approval and consent to participate**

Not applicable.

9. Consent for publication

Not applicable.

10. Competing interests

All authors declare no competing interest.

11. Author details

1Infertility Unit, ASST Lariana, Cantù, Como 22063, Italy. 2Infertility Unit, Fondazione IRCCS Ca’ Granda Ospedale Maggiore Policlinico, Milan 20122, Italy. 3IVF Unit, AUSL Romagna Cattolica Hospital, 47841 Cattolica, Italy. 49(6):101706. https://doi.org/10.1016/j.jogoh.2020.101706.
30. Esfandiari N, Claessens EA, Burjaq H, Gotlieb L, Casper RF. Ongoing twin pregnancy after rescue intracytoplasmic sperm injection of unfertilized abnormal oocytes. Fertil Steril. 2008;90(1):199.e5–7. https://doi.org/10.1016/j.fertnstert.2007.07.1299.

31. Shalom-paz E, Alshalati J, Shehata F, Jimenez L, Son WY, Holzer H, et al. Clinical and economic analysis of rescue intracytoplasmic sperm injection cycles. Gynecol Endocrinol. 2011;27(12):993–6. https://doi.org/10.3109/09513 590.2011.579655.

32. Xiong S, Han W, Liu JX, Zhang XD, Liu WW, Liu H, et al. Effects of cumulus cells removal after 6 h co-incubation of gametes on the outcomes of human IVF. J Assist Reprod Genet. 2011;28(12):1205–11. https://doi.org/10.1007/s10815-011-9630-9.

33. Zhu L, Ren X, Wu L, Hu J, Li Y, Zhang H, et al. Rescue ICSI: choose the optimal rescue window before oocyte aging. Reprod Contracept. 2011;22(1):29–36. https://doi.org/10.1016/S1001-7844(12)60004-2.

34. Singh N, Malhotra N, Shinde U, Tiwari A. Successful live birth after rescue ICSI following failed fertilization. J Hum Reprod Sci. 2013;6(1):77–8. https://doi.org/10.4103/0974-1208.112388.

35. Moon JH, Henderson S, Garcia-Cerrudo E, Mahfoudh A, Reinblatt S, Son WY. Successful live birth after transfer of blastocyst and frozen blastocyst from rescue ICSI with application of polarized light microscopy for spindle examination on unfertilized eggs. J Ovarian Res. 2015;8(1):22. https://doi.org/10.1186/s13048-015-0150-6.

36. Sachdev NM, Grifo JA, Licciardi F. Delayed intracytoplasmic sperm injection (ICSI) with trophectoderm biopsy and preimplantation genetic screening (PGS) show increased aneuploidy rates but can lead to live births with single thawed euploid embryo transfer (STEET). J Assist Reprod Genet. 2016;33(11):1501–5. https://doi.org/10.1007/s10815-016-0743-z.

37. Chen J, Li Q, Wang Y, Huang J, Liu P. Blastocyst cryopreservation and cryopreservation-warming transfer is an effective embryo transfer strategy for day 1 rescue intracytoplasmic sperm injection cycles. Sci Rep. 2021;11(1). https://doi.org/10.1038/s41598-021-87693-y.

38. ESHRE Special Interest Group of Embryology and Alpha Scientists in Reproductive Medicine. The Vienna consensus: report of an expert meeting on the development of ART laboratory performance indicators. Reprod Biomed Online. 2017;35(5):494–510. https://doi.org/10.1016/j.rbmo.2017.06.015.

39. Ethics Committee of the American Society for Reproductive Medicine. Fertility treatment when the prognosis is very poor or futile: an Ethics Committee opinion. Fertil Steril. 2019. https://doi.org/10.1016/j.fertnstert.2019.01.033.

40. Grimes DA, Schulz KF. False alarms and pseudo-epidemics: the limitations of observational epidemiology. Obstet Gynecol. 2012;120(4):920–7. https://doi.org/10.1097/AOG.0b013e31826a61a.

41. van Rumste MM, Evers JL, Farquhar CM. Intra-cytoplasmic sperm injection versus conventional techniques for oocyte insemination during in vitro fertilisation in patients with non-male subfertility. Cochrane Database Syst Rev. 2003. https://doi.org/10.1002/14651858.CD001301.

42. Kilani SS, Cooke S, Kan AK, Chapman MG. Do age and extended culture affect the architecture of the zona pellucida of human oocytes and embryos? Zygote. 2006;14(1):39–44. https://doi.org/10.1017/S0967199406003 625.

43. Viganò P, Alteri A, Busellini A, Vanni VS, Somigliana E. Frozen IVF cycles to circumvent the hormonal storm on endometrium. Trends Endocrinol Metab. 2020;31(4):296–307. https://doi.org/10.1016/j.tem.2020.01.009.

44. Rienzi L, Gracia C, Maggiulli R, LaBarbera AR, Kaiser DJ, Ubaldi FM, et al. Oocyte, embryo and blastocyst cryopreservation in ART: systematic review and meta-analysis comparing slow-freezing versus vitrification to produce evidence for the development of global guidance. Hum Reprod Update. 2017. https://doi.org/10.1093/humupd/dmw038.

45. ESHRE Working group on Time-lapse technology. Apter S, Ebner T, Freour T, Guns Y, Kovacic B, et al. Good practice recommendations for the use of time-lapse technology. Hum Reprod Open. 2020;2020(2). https://doi.org/10.1093/hropen/hoa008.