Maternal mRNA expression levels of H19 are inversely associated with risk of macrosomia

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Abstract

Introduction: To investigate the associations between the mRNA levels of H19 in term placenta and risk of macrosomia.

Material and methods: Term placentas were collected from 37 macrosomia and 37 matched neonates with normal birth weight (controls) born in Changzhou Women and Children Health Hospital, Jiangsu province, P. R. China from March 1 to June 30, 2008. The mRNA levels of H19 in those placentas were measured by real-time polymerase chain reaction (PCR). Simple and multiple logistic regression models were used to explore the risk factors in the development of macrosomia. All analyses were performed using Stata 10.0 (StataCorp, College Station, Texas, USA).

Results: The average H19 mRNA level of the macrosomia group was 1.450 ±0.456 while in the control group it was 2.080 ±1.296. Based on the result of Student’s t test, there was a significant difference in H19 mRNA level between the macrosomia group and the control group (p = 0.008). After controlling for potential confounders, the multivariable adjusted odds ratio (OR) of macrosomia for those in the highest tertile of H19 mRNA level was 0.12 (95% CI: 0.02–0.59) when compared to those in the lowest tertile (p for linear trend = 0.009).

Conclusions: The term placental H19 mRNA levels were inversely related to the occurrence of macrosomia. Our findings suggest that the low expression of H19 mRNA may contribute to the development of macrosomia.

Key words: macrosomia, placenta, H19, insulin-like growth factor II, real-time polymerase chain reaction.

Introduction

Overweight and obesity have increased tremendously during the past 10–15 years and have become a global health problem. The prevalence of overweight and obesity in Chinese adults has increased two- to three-fold in the past 10–15 years [1, 2] and among Chinese children and adolescents it has increased steadily from 5.2% in 1991 to 13.2% in 2006 [3]. The prevalence of overweight and obesity increased in all
age groups, in both rural and urban areas, and in North and South China. The overall combined prevalence of overweight and obesity was 23.2% (17.6% for overweight and 5.6% for obesity) in 2002 [4].

Macrosomia has been suggested as one of the possible risk factors for obesity in many studies [5, 6]. Macrosomia was defined as a condition in which a neonate has a birth weight (BW) equal to or greater than 4000 g. The morbidity of macrosomia reaches 7–10% [7]. In 2006, the morbidity of macrosomia was 6.5% in China [8]. Socioeconomic advances and the improvement in women's living standards can predispose to the development of fetal macrosomia, which may result in childhood obesity [5, 6]. It is a vicious cycle.

The placenta, as the interface between mother and fetus, is central to prenatal growth control. The fetus is dependent upon the placenta for its supply of nutrients and oxygen from the mother. Our previous research found that the placental weights in the macrosomia group were significantly higher than those in the control group and placental weight was positively correlated with birth weight [9]. Recent evidence suggests that imprinted genes may regulate the growth and transport capacity of the placenta by controlling the supply of nutrients [10].

Imprinted genes are expressed in a parent-of-origin manner by epigenetic modifications that silence either the paternal or maternal allele. In general, paternally expressed genes enhance feto-placental growth while maternally expressed genes limit conceptus growth. Insulin-like growth factor II (IGF-II)/H19 is a pair of imprinted genes discovered early. They are similarly positioned, reciprocally imprinted, their expression being correlated with regulation. They co-regulate the growth and transport capacity of the placenta, thereby controlling the supply of nutrients, and are important for fine tuning of normal fetal growth.

Studying the risk factors for macrosomia is of particular public health importance for its primary prevention and therefore reducing the morbidity of obesity.

Previous research focused on whether loss of imprinting status of IGF-II and H19 existed in the placentas of congenital growth disorders such as Beckwith-Wiedemann (BWS) [11], Silver-Russell (SRS) syndromes [12], fetal growth restriction (FGR) [13] and pre-eclampsia [14]. But there has been little research concerning the relation between the expression of IGF-II and H19 and these diseases. However, imprinting status may not be associated with the RNA level [15]. So, it is inconclusive how IGF-II and H19 work: through altering the imprinting status? Changing the RNA level? Or both? Our previous research found that the levels of placental IGF-II were over-expressed in macrosomia [9]. Therefore, in this study we will further investigate the association between the expression of H19 in the placenta and the occurrence of macrosomia.

Material and methods

This is a case-control study of the association between mRNA levels of H19 in the placenta and the occurrence of macrosomia. The study was approved by the human research ethics committees of the Changzhou Women and Children's Health Hospital.

Subjects

Thirty-seven macrosomia cases were chosen randomly from 116 cases found among children born in Changzhou Women and Children Health Hospital from March 1 to June 30, 2008. Macrosomia was defined as a condition in which neonates had a BW equal to or greater than 4000 g. Thirty-seven neonates with normal BW in the corresponding period were chosen as controls. Mothers of both groups had no diabetes or other complications during pregnancy, confirmed by negative results of a 75 g oral glucose tolerance test in the 24th–28th week of pregnancy.

Biomarker detection

A piece of villus tissue 1 cm × 1 cm × 1 cm in size was collected from the center of the placenta under aseptic conditions within 5 min after the delivery of the placenta. Then the specimen was cleansed with normal saline (NS) and preserved in liquid nitrogen with Frozen Pipe.

The primers and probes were designed by Primer Premier 5.0 according to the sequence of GAPDH, IGF-II and H19 genes (GenBank accession number NM_002046, J03242 and AK092486, respectively) and synthesized at Shanghai Sangon Biological Engineering Technology and Services Co., LTD. The standard samples of each gene were ordered from Shanghai Shenyou Biological Technology Co., LTD. The sequences of primers and probes are displayed in Table I.

Tissue general RNA was gathered according to steps described in the manual of general RNA extraction kit. 2 µg of general RNA was gathered from every tissue. Reverse transcription reaction was done in the qualitative PCR device. All real-time PCR reactions were done on the ABI 7300 real-time PCR device. Complex control was done on every specimen. A standard sample was involved in every reaction. The PCR reaction system was composed of: 10 × PBS 2.5 µl, 25 mmol/l MgCl2, 1.5 µl, 10 mmol/l 4 × dNTPs 0.5 µl, 100 µmol/l Jus-
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Covariates
Information about pregnant women's age, height, education level, weight before pregnancy, weight gain during pregnancy, duration of pregnancy, and time of delivery were collected using an interviewer-administered questionnaire.

Statistical analysis
Group comparisons of basic characteristics were performed using Student’s t test or Wilcoxon’s z test as appropriate. Pearson’s or Spearman’s rank correlation coefficients between indicators were calculated depending on the distributions of the variables. H19 expression levels were divided into three groups according to their tertiles, with the lowest tertile as the reference. Multiple logistic regression models were used to explore the association between H19 levels and the occurrence of macrosomia.

All analyses were performed using Stata 10.0 (StataCorp, College Station, Texas, USA). Value of \( p \leq 0.05 \) was considered statistically significant.

Results

General characteristics of the two groups
The average BW of the macrosomia group was 4175.8 ±178.7 g while the control group BW was 3230.0 ±303.5 g. The macrosomia group had a higher BMI before pregnancy, higher maternal weight before pregnancy and higher weight gain during pregnancy. There were no significant differences between cases and controls in maternal age, maternal height, paternal height, paternal weight, paternal BMI, pregnant days or gestational weeks between the two groups (Table II). No significant differences were found in mothers’ education level (Wilcoxon’s \( z = –1.161, p = 0.246 \)), family income (\( z = 0.763, p = 0.445 \)), or gender of the baby (\( z = –0.256, p = 0.798 \)).

Levels of IGF-II and H19 in the placenta
The level of IGF-II showed a positively skewed distribution; therefore, median and inter-quartile ranges (Q1-Q3) were used to describe the data. The level of H19 was normally distributed; therefore, mean and standard deviation were used to describe the data. The levels of IGF-II and H19 expression in placenta of macrosomia and control groups are shown in Table III. Based on the results of Wilcoxon’s \( z \) test and Student’s \( t \) test, there were significant differences in IGF-II mRNA and H19 mRNA level between the macrosomia group and the control group.

Correlation between levels of IGF-II and H19 in the placenta
Spearman rank correlation analysis suggested that the level of IGF-II had a linear upward trend along with the increase of the H19 in the normal birth weight group (\( r = 0.34, p = 0.04 \)).

Table I. Sequences of PCR primers and specific probe (5′-3′) for the detection of GAPDH, IGF-II and H19 by real-time PCR

| Primer | Sequence |
|--------|----------|
| GAPDH Sense primer | GGA AGG TGA AGG TCT GAG TC |
| Antisense primer | CGT TCT CAG CCT TGA CGG T |
| Probe | FAM-TTT GGT CTT ATT GGG CGC CTG-TAMRA |
| IGF-II Sense primer | CGG CTT CTA CTT CAG CAG GC |
| Antisense primer | TGG CGG GGG TAG CAG ACT |
| Probe | FAM-CAA GCC GTG TGA GCC GTG GCA-TAMRA |
| H19 Sense primer | CAG AAC CCA CAA CAT GAA AGA AAT |
| Antisense primer | TCA GGT AGT GCA GTG GTT GTA AAG T |
| Probe | FAM-AAG CCT GGG CCT TTG AAT CCG GAC A-TAMRA |
there was no correlation in the macrosomia group ($r = 0.19, p = 0.25$).

**Association between H19 levels and risk of macrosomia**

We categorized exposures of interest into tertiles. The initial analysis (model 1) was an unadjusted model. In model 2, we adjusted for maternal height (continuous), maternal weight before pregnancy (continuous) and weight gain during pregnancy (continuous), and in model 3 we additionally adjusted for IGF-II (tertiles). Continuous variables were used for linear trend tests. As compared with those in the lowest tertile of H19 mRNA level, the multivariable adjusted odds ratio (OR) of macrosomia was $0.22$ (95% CI: $0.05$–$0.89$) ($p$ for linear trend $= 0.033$) for those in the highest tertile (model 2). When further adjusted for IGF-II levels (tertiles), the inverse association was a little strengthened (OR $= 0.12$ (95% CI: $0.02$–$0.59$); $p$ for linear trend $= 0.009$) (Table IV).

**Discussion**

Our research showed that the level of H19 in the placenta in macrosomia was significantly lower than that of control group. There was a positive correlation between the levels of IGF-II and H19 in the placenta in normal birth weight. The H19 mRNA level, the multivariable adjusted odds ratio (OR) of macrosomia was $0.22$ (95% CI: $0.05$–$0.89$) ($p$ for linear trend $= 0.033$) for those in the highest tertile (model 2). When further adjusted for IGF-II levels (tertiles), the inverse association was a little strengthened (OR $= 0.12$ (95% CI: $0.02$–$0.59$); $p$ for linear trend $= 0.009$) (Table IV).

**Table II. Basic characteristics of pregnant women**

| Parameter                  | Macrosomia† | Normal birth weight | Value of $p$‡ |
|----------------------------|-------------|---------------------|---------------|
| No. of participants       | 37          | 37                  |               |
| Maternal age [years]      | $27.95 \pm 3.3$ | $27.22 \pm 3.04$ | $0.33$         |
| Maternal height           | 162.65      | 161.22              | $0.18$        |
| Maternal weight before pregnancy | 56.20      | 52.30               | $0.01$        |
| BMI before pregnancy      | $21.22 \pm 2.36$ | $20.15 \pm 1.58$ | $0.03$        |
| Weight gain during pregnancy [kg] | $20.50 \pm 4.34$ | $17.20 \pm 4.78$ | $< 0.01$      |
| Paternal height           | 174.08      | 172.89              | $0.28$        |
| Paternal weight           | 70.62       | 69.97               | $0.79$        |
| Paternal BMI              | 23.07       | 23.37               | $0.71$        |
| Pregnant days             | $278.86 \pm 6.36$ | $276.35 \pm 4.79$ | $0.06$        |

*Plus-minus values are mean ± SD. †Macrosomia is defined as neonates with birth weight $\geq$ 4000 g. ‡Difference between groups was tested by Student’s t test. §BMI – body mass index, which is the weight in kilograms divided by the square of the height in meters

**Table III. Levels of insulin growth factor-II (IGF-II) and H19 in maternal placenta between groups**

| Group                   | No. of participants | IGF-II mRNA | H19 mRNA |
|-------------------------|---------------------|-------------|-----------|
|                         | Median Q1–Q3*       | Mean SD     |           |
| Macrosomia†             | 37                  | 2.53        | 2.18–4.30 | 1.45 | 0.46 |
| Normal birth weight     | 37                  | 1.70        | 0.87–3.55 | 2.08 | 1.30 |

Value of $p$* 0.03* < 0.01*

*Q1–Q3 denotes inter-quartile range. †Macrosomia is defined as neonates with birth weight $\geq$ 4000 g. ‡Difference between groups was tested by Wilcoxon’s rank sum test and Student’s t test

**Table IV. Multivariable-adjusted ORs (95% CIs) of risk of macrosomia by tertiles of H19 levels**

| H19 mRNA | Macrosomia† (n = 37) | Normal BW (n = 37) | Model I† | Model II‡ | Model III§ |
|----------|----------------------|--------------------|----------|-----------|-----------|
| Tertile 1 | 15                   | 10                 | 1.00 (Referent) | 1.00 (Referent) | 1.00 (Referent) |
| Tertile 2 | 15                   | 10                 | 1.00 (0.32–3.10) | 1.12 (0.31–4.02) | 1.29 (0.31–5.38) |
| Tertile 3 | 7                    | 17                 | 0.27 (0.08–0.90) | 0.22 (0.05–0.89) | 0.12 (0.02–0.59) |

$P$ for linear trend† 0.033 0.033 0.009

*All the models were constructed by using logistic regression models. Macrosomia was defined as neonates with birth weight $\geq$ 4000 g. †Model I: unadjusted model. ‡Model II: Adjusted for maternal height (continuous), maternal weight before pregnancy and weight gain during pregnancy (continuous). §Model III: additionally adjusted for IGF-II (tertiles). †Continuous variables were used for the linear trend tests
mRNA in the placenta was inversely associated with the occurrence of macrosomia.

Factors influencing the level of H19 in the placenta

Expression of imprinted genes is classically associated with differential methylation of specific CpG-rich DNA regions (DMR). A methylated DMR on the paternal chromosome inactivates adjacent H19 and an unmethylated DMR on the maternal chromosome insulates IGF-II from the enhancers.

In a pathophysiological study of preeclamptic, data from methylation-sensitive high resolution melting demonstrated hypermethylation of the promoter region of the H19 gene, and results of real-time PCR showed decreased mRNA expression of the H19 gene in early-onset preeclamptic placentas as compared with the normal controls [23].

However, in a pathophysiological study of fetal growth restriction (FGR), placentas from growth-restricted pregnancies had lower methylation levels compared to normals, in the H19 promoter region. Placental expression levels of H19 were significantly increased in the FGR group [19]. The hypomethylation of the H19 promoters is compatible with the aberrant expression.

The over-expression of IGF-II could be caused by deletion or knockout of H19 [24], while loss of imprinting and hence biallelic expression could downregulate the expression of IGF-II [25]. It is still unclear whether alteration of the H19 level could regulate directly the expression of IGF-II mRNA and whether the level of H19 mRNA in the macrosomia placenta is involved in regulation of its methylation, which needs to be further investigated.

The mechanisms of these genes’ effects on macrosomia will be the focus of our continual study. In view of the factors that may affect the expression of imprinted genes, the methylation of DMR of imprinted genes is most critical. The key of our next study project will be to determine the methylation status of certain CpG islands in the control region on the genomic DNA.

In conclusion, the significant positive associations between the level of placental IGF-II and H19 and macrosomia were explored. Our findings suggest that a potential risk effect of the value of the expression of IGF-II mRNA and H19 mRNA in the placenta may exist during the development of macrosomia. Further studies are warranted to confirm our findings and clarify the potential mechanisms.

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References

1. Wang H, Du S, Zhai F, Popkin BM. Trends in the distribution of body mass index among Chinese adults, aged 20-45 years (1989-2000) Int J Obes (Lond) 2007; 31: 272-8.

2. Wildman RP, Gu D, Muntner P, et al. Trends in overweight and obesity in Chinese adults: between 1991 and 1999-2000. Obesity (Silver Spring) 2008; 16: 1448-53.

3. Cui Z, Huxley R, Yu Y, Dibley MJ. Temporal trends in overweight and obesity of children and adolescents from nine Provinces in China from 1991-2006. Int J Pediatr Obes 2010; 5: 365-74.

4. Wu YF, Ma GS, Hu YH, et al. The current prevalence status of body overweight and obesity in China: data from the China National Nutrition and Health Survey. Zhonghua Yu Fang Yi Xue Za Zhi 2005; 39: 316-20.

5. Van Eerden P. Obesity in pregnancy. S D Med 2011; Spec No: 46-50.

6. Satpathy HK, Fleming A, Frey D, Barsoom M, Satpathy C, Khandalavala J. Maternal obesity and pregnancy. Postgrad Med 2008; 120: E01-9.

7. Stotland NE, Caughey AB, Breed EM, Escobar GJ. Risk factors and obstetric complications associated with macrosomia. Int J Gynaecol Obstet 2004; 87: 220-6.

8. Yu DM, Zhai FY, Zao LY, et al. Incidence of fetal macrosomia and influencing factors in China in 2006. CJCHC 2008; 16: 11-3.

9. Hua Jiang, Pengcheng Xun, Guanghua Luo, et al. Levels of insulin-like growth factors and their receptors in placenta in relation to macrosomia. Asia Pacif J Clin Nutrition 2009; 18: 171-8.

10. Frost JM, Moore GE. The importance of imprinting in the human placenta. PLoS Genet 2010; 6: 100-15.

11. Cooper WN, Luharia A, Evans GA, et al. Molecular subtypes and phenotypic expression of Beckwith-Wiedemann syndrome. Eur J Hum Genet 2005; 13: 1025-32.

12. Gilciquel C, Rossignol S, Cabrol S, et al. Epimutation of the telomeric imprinting center region on chromosome 11p15 in Silver-Russell syndrome. Nat Genet 2005; 37: 1003-7.

13. Ying W, Jingli F, Wei SW, et al. Genomic imprinting status of IGF-II and H19 in placenta of fetal growth restriction patients. J Genet 2010; 89: 213-6.

14. Zhao D, Li J, Yu L, et al. H19 expression in placenta with pre-eclampsia. Zhonghua Fu Chan Ke Za Zhi 2009; 44: 87-90.

15. Yu L, Chen M, Zhao D, et al. The H19 gene imprinting in normal pregnancy and pre-eclampsia. Placenta 2009; 30: 443-7.

16. Tycko B. Imprinted genes in placental growth and obstetric disorders. Cytogenet Genome Res 2006; 113: 271-8.

17. Diplas A, Lambertini L, Lee MJL, et al. Differential expression of imprinted genes in normal and IUGR human placentas. Epigenetics 2009; 4: 235-40.

18. Vambergue A, Fajardy I, Dufour P, et al. No loss of genomic imprinting of IGF-II and H19 in placentas of diabetic pregnancies. Growth Horm IGF Res 2007; 17: 130-6.

19. Koukoura O, Sifakis S, Zaravinos A, et al. Hypomethylation along with increased H19 expression in placentas from pregnancies complicated with fetal growth restriction. Placenta 2011; 32: 51-7.

20. Reik W, Constancia M, Fowden A, et al. Regulation of supply and demand for maternal nutrients in mammals by imprinted genes. J Physiol 2003; 547: 35-44.