Letter to the Editor

Fatal Cerebral Haemorrhage in a Thrombolysed Patient with Ischaemic Stroke Who Developed Interval Thrombocytopaenia from Acute Dengue Infection

Dear Editor,

Intravenous thrombolysis is an evidence-based treatment in acute ischaemic stroke. Symptomatic intracranial haemorrhage (sICH) is a known complication of intravenous thrombolysis with a prevalence rate of 5–6%.

The known predictors of sICH include high blood pressure during thrombolysis, large volume of ischaemic change on imaging studies, extensive cerebral microbleeds and extensive leukoariosis.

We report a rare case of devastating sICH following thrombolysis for ischaemic stroke with underlying thrombocytopaenia that was attributed to dengue fever.

Case Presentation

A 73-year-old man with a history of hypertension and hyperlipidaemia—managed with Niften (slow-release nifedipine 20 mg and atenolol 50 mg) once daily and lovastatin 20 mg nocte—presented with symptoms of left-sided weakness and slurring of speech. He did not have a history of stroke and antithrombotic medication-taking. Physical examination revealed left ataxic hemiparesis with a score of 4 on the National Institutes of Health Stroke Scale (NIHSS). He was afebrile and his blood pressure was 160/95 mmHg. Computed tomography (CT) of his brain (Fig. 1) did not show early ischaemic changes over the right hemisphere and the Alberta Stroke Program Early CT Score was 10. Hypodensity was seen over the left occipital pole in the absence of corresponding signs or symptoms (Fig. 1A). Full blood count and coagulation profile were normal, and there were no contraindications for thrombolysis.

Fig. 1. Computed tomography of brain at (A) pre-thrombolysis, (B) 8 hours post-thrombolysis, (C) day 9 and (D) day 16 of admission.
He was treated with standard intravenous alteplase dose of 0.9 mg/kg which was initiated 4 hours after symptom onset. Over the next 8 hours, his blood pressure ranged between 150/80–160/90 mmHg. At 2 hours post-thrombolysis, an isolated reading of 190/120 mmHg fell to 158/93 mmHg after 30 minutes (Fig. 2), the signs and symptoms resolved and NIHSS score was nil.

At 8 hours post-thrombolysis, he developed headache. Physical examination revealed right homonymous hemianopia. Brain CT showed left parieto-occipital parenchymal haemorrhage 1 (PH1) (Fig. 1B) according to the definition of Safe Implementation of Treatments in Stroke. A new area of hypodensity was seen over the right thalamic region that was suggestive of an evolving infarct, and there was a spike in blood pressure to 199/96 mmHg (Fig. 2). Transdermal glyceryl trinitrate patch 2.5 mg and oral amlodipine 2.5 mg were administered intermittently to manage blood pressure. Laboratory tests showed normal full blood count (haemoglobin 15.5g/dL, total white blood cell [WBC] count 12 × 10^9/L and platelet 377 × 10^9/L), coagulation profile (activated partial thromboplastin time of 30 seconds and thromboplastin time of 16.2 seconds), serum fibrinogen level (0.46 g/L) and liver function. He was treated with 6 units of cryoprecipitate concentration.

On day 2, he was started on oral amlodipine 5 mg once daily and repeat brain CT showed stable size of left parieto-occipital haemorrhage. On day 3, magnetic resonance image (MRI) of the brain showed multiple foci of acute infarction over both cerebellar lobes and established infarction of right thalamus (Fig. 3). Magnetic resonance angiography did not show significant intracranial large artery stenosis, and signs of aneurysm or cerebral microbleeds were absent. The diagnosis was acute right thalamic infarction and haemorrhagic transformation of silent left occipital infarct of grade PH1. Echocardiography and electrocardiogram monitoring did not detect the source of cardioembolism.

Between day 4 and day 8, he had daily temperature spikes (Fig. 4) and his blood pressure ranged between 150/80–170/100 mmHg. His neurological status was stable with no new physical signs and his headache improved with analgesia. Chest radiograph, blood and urine culture findings were normal, and he was treated as presumptive viral fever. WBC count ranged between 8 × 10^9/L–10 × 10^9/L (Fig. 5). In view of intracranial haemorrhage, antiplatelet therapy was not started.

On day 8, the fever settled. Full blood count taken on the same day revealed thrombocytopenia of 11 × 10^9/L (Fig. 5), a marked drop from the normal platelet level seen on day 4. There was a concurrent raise in total WBC count of 20 × 10^9/L (Fig. 6) with monocytosis (1.2 × 10^9/L) and serum transaminases.
Fig. 4. Temperature (degree Celsius) readings from day 4 to day 10.

Fig. 5. Platelet count ($\times 10^9/L$) from day 1 to day 15.

Fig. 6. White blood cell count ($\times 10^9/L$) from day 1 to day 15.
(alanine aminotransferase 178 U/L and aspartate aminotransferase 233 U/L). Coagulation profile and serum fibrinogen level remained normal. Reverse-transcription polymerase chain reaction tested positive for dengue virus ribonucleic acid, but dengue serotyping was not performed. Physical examination did not show ascites, hepatosplenomegaly, petechial rash or pleural effusion. One unit of platelet concentrate was transfused; at 10 hours post-transfusion, repeat platelet count showed a reading of 27 × 10^9/L. Patient remained clinically stable and afebrile.

On day 9, repeat brain CT showed extension of left cortical haemorrhage, but no associated mass effect (Fig. 1D). Platelet count was 48 × 10^9/L. Another unit of platelet concentrate was administered and it was gradually increased from 29 × 10^9/L to 48 × 10^9/L, 82 × 10^9/L and 129 × 10^9/L (Fig. 4). However, right hemiparesis and aphasia worsened progressively. On day 16, brain CT showed progression from PH1 to PH2 (Fig. 1E). The level of consciousness dropped rapidly, and he died on the same day.

Discussion
This is the first reported case of sICH following stroke thrombolysis with dengue fever. Dengue is a vector-borne disease caused by a flavivirus transmitted by the Aedes aegypti mosquito. Previous case reports on dengue that contributed to ischaemic stroke had reported onset of ischaemic stroke during the febrile stage of dengue which ranged between day 1–15 of fever. A few reports suggested that thrombotic risk in dengue fever can be heightened by factors such as increased levels of immunoglobulin M against phospholipids and lupus anticoagulants, increased plasminogen activator inhibitor-1 plasma levels, low concentrations of plasma anticoagulant proteins C and S and antithrombin III, and disseminated intravascular coagulopathy. It was proposed that inflammation could be attributed to infections that cause hypercoagulable state.

In our patient, we postulate that dengue did not contribute to ischaemic stroke since clinical features of dengue or thrombocytopenia at initial stroke presentation were absent. The fever started 4 days after the onset of ischaemic stroke, but the aetiology of ischaemic stroke could not be determined. Findings from initial investigations did not show cardioembolism, large vessel obstruction and infective endocarditis. Coagulation profile screen was also normal.

Primary intracranial haemorrhage with dengue fever has been reported in a few case studies that cited coagulopathy, platelet dysfunction, thrombocytopenia and vasculopathy as possible causes. Bleeding from dengue infection is also attributed to direct fibrinolysis effect by dengue virus.

Our patient had parenchymal haemorrhage (PH) of asymptomatic infarct due to thrombolysis. He remained stable for 4 days after onset of PH, but subsequently deteriorated with worsening hemiparesis and progressive drowsiness. CT findings also showed progression from PH1 to PH2 with associated mass effect. PH progression may be explained by severe thrombocytopenia associated with dengue haemorrhagic fever and, possibly, endothelial leakage which is known to occur with dengue infection.

During the febrile phase of dengue fever, an immune-mediated process involving inflammatory cytokines—such as monocyte chemo-attractive protein-1—has been observed to cause alteration of tight junction of vascular endothelium leading to plasma leakage. Other cytokines—such as certain interleukins and platelet-derived growth factors—may lead to platelet destruction and limit platelet aggregation that may precede the defervescence phase.

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