Cumulative meta-analysis and trial sequential analysis of correlation between hOGG1 Ser326Cys polymorphism and the risk of head and neck squamous cell carcinoma

Yan Yan1,*, Ai-Ping Deng2,*, Wen Chen3,*, Yu-Hua Ming1, Xian-Tao Zeng1 and Wei-Dong Leng1

1Department of Stomatology, Taihe Hospital, Hubei University of Medicine, Shiyan 442000, China
2Department of Neurosurgery, Taihe Hospital, Hubei University of Medicine, Shiyan 442000, China
3Department of Radiology, Taihe Hospital, Hubei University of Medicine, Shiyan 442000, China

*These authors contributed equally to this work

Correspondence to: Wei-Dong Leng, email: lengtaihe@163.com

Keywords: human 8-oxoguanine glycosylase 1, hOGG1, polymorphism, head and neck squamous cell carcinoma, meta-analysis

Received: March 27, 2017  Accepted: August 28, 2017  Published: January 06, 2018

ABSTRACT

Background: The human 8-oxoguanine glycosylase 1 (hOGG1) Ser326Cys polymorphism has been involved in the risk of head and neck squamous cell carcinoma (HNSCC), but the results of published studies on this topic still inconsistent.

Results: Finally 11 qualified publications with 13 independent case-control studies were yielded. Overall, we observed significant differences in CysCys vs. SerSer [odds ratio (OR) = 1.55, 95% confidence interval (95% CI) = 1.01–2.38] and CysCys vs. SerCys+SerSer (OR = 1.42, 95% CI = 1.005–1.99) genetic models. Sensitivity analyses showed the results were not robust, cumulative meta-analyses and trial sequential analysis indicated the results didn't not need more studies to identification. Subgroup analyses showed there was a significant association in Caucasian, laryngeal squamous cell carcinoma, studies agreement with Hardy-Weinberg equilibrium, and alcohol drinkers subgroups under the corresponding contrasts. In addition, the results of Egger’s test were contradictory.

Materials and Methods: All eligible studies were searched from the online databases including PubMed, Web of Science, China Knowledge Resource Integrated Database, and Wanfang databases up to February 10, 2017. After study selection and data extraction, the meta-analysis was performed using STATA 12.0 software and TSA software version 0.9 Beta.

Conclusions: Our meta-analysis results indicated that hOGG1 Ser326Cys polymorphism may be associated with increased risk of HNSCC, especially in Caucasians, alcohol drinkers and the patients with laryngeal squamous cell carcinoma.

INTRODUCTION

Head and neck squamous cell carcinoma (HNSCC), involving the oral cavity, pharynx, and larynx, constitutes 12% of all malignant neoplasms worldwide [1]. It is estimated that a total of 400,000 cases of the oral cavity and pharynx cancers and 160,000 cases of laryngeal cancer have been diagnosed and 300,000 people die of these diseases per year [2]. Hence, investigating and preventing the risk factors of HNSCC is a necessary and significant research field. In the past decades, many epidemiological researches have suggested that active and passive smoking, alcohol consumption, genetic factors, viral infection, gender, tooth loss, periodontal disease, and occupational exposure are the significant risk factors for HNSCC [3–12]. Of them, tobacco smoking and
alcohol are supposed to the most important risk factors [3, 5]. However, such markers could not comprehensively explain the etiology of HNSCC [13]. That indicates that the individual susceptibility may play a certain role in the carcinogenesis of HNSCC.

With the rapid development of molecular epidemiology, some meta-analyses have provided increasing evidence to support the hypothesis that some genetic polymorphisms play a significant role in determining individual susceptibility to HNSCC [14–19]. DNA damage is considered as a critical factor to carcinogenesis. DNA repair mechanisms play an important role in the integrity and stability of the genome. Base excision repair (BER), which is an important DNA repair pathway, plays a vital role in the repair of mutations generated by reactive oxygen species (ROS). The 8-hydroxy-2 deoxyguanine (8-OH-dG) is one of the most abundant oxidative products of high mutagenesis among numerous factors of oxidative DNA damage, because it has the tendency to mispair with adenine during DNA replication and finally result in GC to TA mutation [20]. The human oxoguanine glycosylase 1 (hOGG1) is a DNA glycosylase or AP lyase, which has been investigated to play a vital role in preventing carcinogenesis by repairing oxidative damage to DNA [21]. The glycosylase or AP lyase could efficiently catalyze and remove 8-OH-dG adducts produced by reactive free radicals, which is a major form of DNA damage. It has been hypothesized that the polymorphism in hOGG1 gene may affect the risk of developing HNSCC because of the critical roles in stabilizing genome integrity. The hOGG1 gene has a G1245C polymorphism in exon 7 making the codon 326 coding Ser or Cys (rs1052133) [22], and Cys326 has lower ability to prevent mutagenesis by 8-OH-dG than Ser326 in human cells in vivo [21]. Many studies have suggested that this mutation may be associated with increased risk of several cancers [23]. Several of them have focused on the association of hOGG1 Ser326Cys polymorphism with head and neck cancer risk. However, we observe that the results of the association between hOGG1 Ser326Cys polymorphism and HNSCC susceptibility remain controversial.

In 2011, Wei et al performed a meta-analysis based on 6 case-control studies indicating that hOGG1 Ser326Cys polymorphism was significantly associated with HNSCC risk only under CysCys vs. SerSer model [24]. However, another meta-analysis of 6 case-control studies by Wang et al in 2012 [23] showed that there was a significant association under all five genetic models. Obviously, the results of these meta-analyses were inconsistent (Supplementary Table 1), which might be attributed to the small effect of the Ser326Cys variation on HNSCC risk or the relatively low statistical power of published papers. At present, eleven eligible articles [25–35] on this issue have been published. Hence, in order to more systematically investigate the association between hOGG1 Ser326Cys polymorphism and risk of HNSCC, we included these eleven articles [25–35] and performed present updated meta-analysis through combing the different studies which a quantitative approach. For inspecting whether sample size influenced the overall results and judging whether more relevant studies would be worthwhile, we conducted not only cumulative analysis by cumulating the single study according to the publication year [36, 37] but also trial sequential analysis (TSA) to explore whether further studies are needed or not [38–40]. Additionally, we also preformed subgroup analyses according to ethnicity, tumor site, source of control, smoking, alcohol patients, genotyping method, and controls agreement with Hardy-Weinberg equilibrium (HWE), respectively.

RESULTS

Study identification and characteristics

We initially yielded 73 papers and finally 11 publications with 13 case-control studies [25–35] were included in the meta-analysis, involving a total of 3,875 patients and 4,696 healthy controls. Figure 1 presents the selection process.

Of them, the study by Hall et al [29] focused on many different tumor sites and it was considered as three independent trials. The published language of one study was in Chinese [30], and others were in English [25–29, 31–35]. The sample size of cases varied greatly, ranging from 29 to 706, and the number of controls ranged from 30 to 1196. Four studies involving six trials did not satisfy the HWE for control group [29, 33–35]. The main characteristics and quality of the included studies are shown in Table 1.

Meta-analysis

The estimation of the association between hOGG1 Ser326Cys polymorphism and HNSCC risk is presented in Table 2. In consequence of the high degree of heterogeneity under all the genetic models, random-effects model was applied. Overall, the pooled analysis showed that a statistical significance under Cys/Cys vs. Ser/Ser [odds ratio (OR) = 1.55, 95% confidence interval (95% CI) = 1.01–2.38, \( F = 79.6\% \), Figure 2] and CysCys vs. (CysSer + SerSer) (OR = 1.42, 95% CI = 1.00–1.99, \( F = 74.3\% \)) genetic models, but nonsignificant association under the other three genetic models [Cys vs. Ser: OR = 1.16, 95% CI = 0.98–1.37, \( F = 81.5\% \); CysSer vs. SerSer: OR = 1.11, 95% CI = 0.93–1.33, \( F = 67.2\% \); (CysCys + CysSer) vs. SerSer: OR = 1.16, 95% CI = 0.95–1.42, \( F = 77.2\% \)]. TSA showed that the overall results might be conclusive, as their cumulative z-curves crossed both the conventional boundary and the trial sequential monitoring boundary providing firm evidence (Figure 3).
Table 1: Characteristics of all included studies in the meta-analysis

| References   | Country (Ethnicity) | Tumor site    | Cases/ Controls | Source of control | Genotyping method | HWE  |
|--------------|--------------------|---------------|-----------------|-------------------|-------------------|------|
| Elahi 2002   | USA (Caucasian)    | Oropharynx    | 169/338         | PB                | PCR-RFLP          | Yes  |
| Cho 2003     | China (Asian)      | Nasopharynx   | 334/283         | PB                | PCR-RFLP          | Yes  |
| Zhang 2004   | USA (Caucasian)    | HNSCC         | 706/1196        | HB                | PCR-RFLP          | Yes  |
| Gorgens 2007 | Germany (Caucasian)| HNSCC         | 29/30           | PB                | PCR               | Yes  |
| Hall 1 2007  | EE (Caucasian)     | Oral cavity   | 160/754         | HB                | Taqman            | No   |
| Hall 2 2007  | EE (Caucasian)     | Pharynx       | 107/554         | HB                | Taqman            | No   |
| Hall 3 2007  | EE (Caucasian)     | Larynx        | 312/754         | HB                | Taqman            | No   |
| Yang 2008    | China (Asian)      | Larynx        | 72/72           | HB                | PCR-RFLP          | Yes  |
| Pawlowska 2009 | Poland (Caucasian)| Larynx      | 253/253         | HB                | PCR-RFLP          | Yes  |
| Laantri 2011 | African (North African) | Nasopharynx | 598/545        | PB                | TaqMan            | Yes  |
| Mitra 2011   | India (Caucasian)  | HNSCC         | 250/325         | PB                | PCR-RFLP          | No   |
| Sliwinski 2011 | Poland (Caucasian)| Larynx      | 265/280         | PB                | PCR-RFLP          | No   |
| Tsai 2012    | China (Asian)      | Oral cavity   | 620/620         | HB                | PCR-RFLP          | No   |

Notes: EE, Eastern European (Romania, Poland, Russia, Slovakia, and Czech Republic); African, Morocco, Algeria and Tunisia; HNSCC, head and neck squamous cell carcinoma; N., sample size; HWE, Hardy-Weinberg equilibrium.

Figure 1: Flowchart of study section in the meta-analysis.
Table 2: The results of overall and subgroup analysis of the all genetic models

|                      | N. | Cys vs. Ser | CysCys vs. SerSer | CysSer vs. SerSer | (CysCys+ CysSer) vs. SerSer | CysCys vs. (CysSer + SerSer) |
|----------------------|----|-------------|--------------------|------------------|-----------------------------|----------------------------|
|                      |    | OR (95% CI) | F (%)              | OR (95% CI)      | F (%)                       | OR (95% CI)                |
| Overall              | 13 | 1.16 (0.98–1.37) | 81.5               | 1.55 (1.01–2.38) | 79.6                        | 1.11 (0.93–1.33)           |
|                      |    |             |                    | 1.11 (0.93–1.33) | 67.2                        | 1.16 (0.95–1.42)           |
|                      |    |             |                    |                  | 77.2                        | 1.42 (1.005–1.99)          |
|                      |    |             |                    |                  |                            |                            |
| Ethnicity            |    |             |                    |                  | 81.5                        | 1.55 (1.01–2.38)           |
| Caucasian            | 9  | 1.19 (0.95–1.49) | 82.0               | 1.79 (0.97–3.30) | 79.3                        | 1.09 (0.88–1.34)           |
|                      |    |             |                    |                  | 65.7                        | 1.15 (0.90–1.47)           |
|                      |    |             |                    |                  | 77.2                        | 1.73 (1.01–2.97)           |
| Asian                | 3  | 1.14 (0.75–1.75) | 86.5               | 1.15 (0.49–2.71) | 80.5                        | 1.38 (0.68–2.80)           |
|                      |    |             |                    |                  | 84.7                        | 1.36 (0.63–2.93)           |
|                      |    |             |                    |                  | 88.2                        | 0.87 (0.65–1.17)           |
| North African        | 1  | 1.06 (0.87–1.28) | NA                 | 1.22 (0.77–1.91) | NA                          | 0.99 (0.77–1.28)           |
|                      |    |             |                    |                  | NA                          | 1.03 (0.81–1.31)           |
|                      |    |             |                    |                  | NA                          | 1.22 (0.79–1.89)           |
| Tumor site           |    |             |                    |                  |                            |                            |
| Larynx               | 4  | 1.49 (1.04–2.13) | 80.6               | 2.65 (1.27–5.54) | 53.1                        | 1.40 (0.97–1.33)           |
|                      |    |             |                    |                  | 72.0                        | 1.53 (1.01–2.33)           |
|                      |    |             |                    |                  | 79.5                        | 2.29 (1.29–4.08)           |
| Oral cavity          | 2  | 0.98 (0.63–1.52) | 85.8               | 1.36 (0.30–6.21) | 91.7                        | 0.86 (0.65–1.15)           |
|                      |    |             |                    |                  | 31.2                        | 0.89 (0.55–1.42)           |
|                      |    |             |                    |                  | 77.3                        | 1.47 (0.39–5.55)           |
| Nasopharynx          | 2  | 1.07 (0.92–1.24) | 0                  | 1.31 (0.94–1.84) | 0                           | 1.26 (0.73–2.16)           |
|                      |    |             |                    |                  | 74.2                        | 1.22 (0.80–1.87)           |
|                      |    |             |                    |                  | 62.8                        | 1.03 (0.79–1.34)           |
| Source of control    |    |             |                    |                  |                            |                            |
| Population-based     | 6  | 1.17 (0.86–1.60) | 85.4               | 1.50 (0.72–3.12) | 81.2                        | 1.21 (0.85–1.72)           |
|                      |    |             |                    |                  | 77.0                        | 1.24 (0.84–1.83)           |
|                      |    |             |                    |                  | 83.0                        | 1.30 (0.72–2.36)           |
| Hospital-based       | 7  | 1.14 (0.92–1.40) | 78.3               | 1.60 (0.91–2.82) | 79.4                        | 1.02 (0.85–1.23)           |
|                      |    |             |                    |                  | 50.9                        | 1.09 (0.87–1.36)           |
|                      |    |             |                    |                  | 70.0                        | 1.56 (0.96–2.55)           |
| Smokers              | 4  | 1.40 (0.78–2.50) | 90.3               | 1.73 (0.54–5.56) | 84.2                        | 1.40 (0.78–2.51)           |
|                      |    |             |                    |                  | 80.4                        | 1.34 (0.69–2.62)           |
|                      |    |             |                    |                  | 86.9                        | 1.64 (0.67–3.98)           |
| Alcohol drinkers     | 2  | 1.62 (1.23–2.13) | 0                  | 2.54 (0.72–8.95) | 58.5                        | 1.67 (1.19–2.35)           |
|                      |    |             |                    |                  | 0                           | 1.75 (1.26–2.42)           |
|                      |    |             |                    |                  | 2.15 (0.58–8.02)           |                            |
| HWE (P > 0.05)       | 7  | 1.26 (1.03–1.55) | 71.8               | 1.53 (1.04–2.24) | 47.0                        | 1.29 (1.00–1.65)           |
|                      |    |             |                    |                  | 63.2                        | 1.35 (1.04–1.76)           |
|                      |    |             |                    |                  | 70.0                        | 1.28 (0.92–1.78)           |
| PCR-RFLP             | 8  | 1.20 (0.93–1.56) | 88.7               | 1.42 (0.79–2.56) | 84.9                        | 1.23 (0.92–1.63)           |
|                      |    |             |                    |                  | 80.0                        | 1.26 (0.91–1.75)           |
|                      |    |             |                    |                  | 86.3                        | 1.22 (0.80–1.86)           |
|                      |    |             |                    |                  | 77.8                        |                            |

Notes: OR, odd ratio; CI, confidence interval; N., number of case-control studies; HWE, Hardy-Weinberg equilibrium; NA, not available.

Figure 2: Forest plot of overall population based on CysCys vs. SerSer genetic model.
We performed sensitivity analysis for testing the robustness of the pooled estimations, and the results showed that our findings were not robust under all the contrasts (Figure 4). The cumulative meta-analysis demonstrated the overall results from significant to nonsignificant to significant; the trend was not stable when the studies were accumulated (Figure 5).

To evaluate whether there was a different genotype effect in predefined subgroups of studies, we conducted stratified analyses in relation to ethnicity, tumor site, source of control, smoking, alcohol drinking, controls agreement with HWE, and genotyping method. Of them, we observed increased risks of Ser326Cys polymorphism for Caucasians, laryngeal squamous cell carcinoma (LSCC), alcohol drinkers and controls agreement with HWE under corresponding models. We did not find any significant association in other subgroups by control source, smokers, and genotyping method. Table 2 presented the results of subgroup analyses.

### Publication bias

Funnel plots based on the trim and fill method indicated obvious publication bias (Figure 6). Egger’s test also provided that funnel plots were asymmetry in the Cys vs. Ser \((p = 0.048)\), CysCys vs. SerSer \((p = 0.04)\), and CysCys vs. (SerCys+SerSer) \((p = 0.02)\) genetic models; however, the publication bias was not detected in the CysSer vs. SerSer \((p = 0.15)\) and (CysCys+ CysSer) vs. SerSer \((p = 0.14)\) genetic models.

### DISCUSSION

The present meta-analysis including 3,875 HNSCC cases and 4,696 controls indicated that \(hOGG1\) Ser326Cys polymorphism might be associated with increased risk of HNSCC, and the TSA results also provided firm evidence. However, the results changed into nonsignificant after adjusting the publication bias by trim and fill method;

---

**Figure 3:** Trial sequential analysis of CysCys vs. SerSer genetic model. CC, CysCys; GG, SerSer; a low risk diversity adjusted information size of 3490 patients was calculated using \(\alpha = 0.05\) (two sided), \(\beta = 0.20\) (power 80%); TSA adjusted 95% confidence interval for a odds ratio of 1.55 is 1.01 to 2.38 based on random-effects model. TSA illustrated that the cumulative z curve crossed the conventional boundary and the trial sequential monitoring boundary for increased risk, and that the required information size was achieved, showing that currently cumulative evidence might be true positive.
sensitivity analysis and cumulative meta-analysis suggested the pooled ORs and corresponding 95% CIs were not robust. Minelli et al. [41] thought that papers that appear in which the controls deviated from HWE should not be excluded unless there were other factors affecting the quality of the study; hence, we performed a subgroup analysis according to the controls agreement with HWE. The results reached statistical significance in allele model, co-dominant model, and dominant model. Based on stratified analysis by tumor site, we observed that the hOGG1 Ser326Cys polymorphism was significantly associated with LSCC. However, only four studies focused on LSCC, that might reduce the reliability of the results. As we know, smoking and alcohol consumption have been identified as major risks for HNSCC [3, 5], hence, we preformed subgroup analyses according to smoking and alcohol drinking. There were four and two studies involving hOGG1 Ser326Cys polymorphism and HNSCC in smokers and alcohol drinkers, respectively. Interestingly, we found no significant association between hOGG1 Ser326Cys polymorphism and HNSCC in smokers but statistically significant relationship in alcohol drinkers. By carefully reading relevant papers, we found that the smokers were not stratified by smoking magnitude; nevertheless, it’s known that the power of influence of heavy smoking must be different from that of occasionally smoking. These results suggest that when designing a case-control study of genetic polymorphism and the risk of cancer relating to other factors, such as smoking- or alcohol-related cancer, researchers should stratify the smokers and drinkers based on magnitude, for giving a more reliable conclusion.

Comparing with the previous meta-analyses [23, 24], our meta-analysis has four advantages. First, the number of included studies and total sample sizes were larger. The number of included studies was six for both previous meta-analyses [23, 24], and the relevant results were contradictory. Of them, Wang et al [23] conducted a subgroup analysis focused on HNSCC containing six studies, which showed that hOGG1 Ser326Cys polymorphism increased HNSCC risk under all comparisons. The meta-analysis from Wei et al [24], included six studies and indicated a significant association between hOGG1 Ser326Cys polymorphism and HNSCC under CysCys vs. SerSer model. Our meta-analysis incorporated 13 studies, which showed that hOGG1 Ser326Cys polymorphism was associated with the elevated risk of HNSCC under CysCys vs. SerSer and CysCys vs. (CysSer + SerSer) contrasts. Moreover, we extracted the data of oral cavity, larynx, and nasopharynx cancers as well as the data of smokers and alcohol drinkers to performe relevant subgroup analyses.

Figure 4: Forest plot of sensitivity analysis based on CysCys vs. SerSer model by omitting each study in turn.
Figure 5: Forest plot of cumulative meta-analysis based CysCys vs. SerSer model by accumulating each study according to the publication year.

Figure 6: Funnel plots based on the trim and fill method of CysCys vs. SerSer model.
Relatively speaking, our paper was more scientific than previous studies on this topic, because it was the first one containing subgroup analyses based on the tumor sites, smokers, and alcohol drinkers. Third, we used cumulative meta-analysis and TSA to explore the results and found the relevant original studies didn’t need to be conducted continuously. Fourth, we assessed the methodological quality of included studies.

Meta-analysis is a retrospective analysis [42]; therefore, some limitations should be taken into consideration. First, the studies included in the present meta-analysis were medium-sized case-control studies, with little power to detect the final aggregated ORs. The results of cumulative analysis also provided that the sample size was not enough. Second, four included studies in our meta-analysis were not in HWE and a significant association was obtained under three genetic models based on HWE conformed studies. This indicated that the overall results might be not reliable. Third, large publication bias existed in this meta-analysis, which might affect the precision of our estimates. Furthermore, sensitivity analysis also showed the overall results were not robust. Fourth, the heterogeneity was moderate to high degree and it was failed to be explained by subgroup analyses. This might be induced by other internal validity, so we could not ignore the influence of heterogeneity. Fifth, although the result of methodological assessment of included studies got high scores, none of them reached the top score, indicating that the reliable of our results was influenced. Finally, like other genetic association meta-analyses [17, 23, 43], this meta-analysis was based on unadjusted data due to lack of detailed information stratified by main confounding variables from primary studies. Hence, gene-gene and gene-environment interactions results remain unclear.

In summary, our meta-analysis suggests that hOGG1 Ser326Cys polymorphism may contribute to the occurrence of HNSCC, especially in Caucasians, alcohol drinkers and the patients with LSCC. However, due to the aforementioned limitations, the results of current meta-analysis should be treated with caution.

MATERIALS AND METHODS

This meta-analysis was reported in accordance with the Meta-Analysis of Observational Studies in Epidemiology (MOOSE) guideline [44].

Eligible criteria

Each study included in the present meta-analysis was required to meet all the following criteria: (i) the study focused on the association between hOGG1 Ser326Cys polymorphism and risk of HNSCC; (ii) the study design was a cohort or case-control study; and (iii) the data of genotype distributions in case and control groups were available. The language was restricted to English and Chinese.

Search strategy

A comprehensive electronic search was performed in PubMed, Web of Science, CNKI (Chinese National Knowledge Infrastructure) and Wanfang database up to February 10, 2017. The following search terms were used: (“human oxoguanine glycosylase 1” OR “hOGG1” OR “human 8-oxoguanine glycosylase 1”) AND (“head and neck” OR “oral” OR “pharyngeal” OR “oropharyngeal” OR “laryngeal” OR “tongue” OR “mouth”) AND (“cancer” OR “carcinoma”) AND (“polymorphism” OR “variant”). We also conducted a manual search of references cited in include studies and previously meta-analyses [24, 45] for additional pertinent studies.

Data extraction

Two authors independently selected included studies and extracted essential data was extracted from all eligible studies, which contained surname of the first author, year of publication, country of study performed, ethnicity of descent, source control, HWE of control, tumor site, sample size, number of genotype distribution, genotyping method, and relevant methodological information. A cross-check over these data was performed by these two authors; any disagreements would be settled through discussion until reaching a final consensus.

Data analysis

All the data analyses were applied using the STATA 12.0 software. HWE test of control group was performed by chi-square test. Heterogeneity among the included studies was tested by the $I^2$ value [46]. If $I^2 < 25\%$ indicating the homogeneity was good, the fixed effect model was used; otherwise, random-effects model was applied. The OR and its 95% CI were calculated to estimate the overall results. Subgroup analysis was used to explore and explain the diversity of the results in different researches using the following factor as stratifying variables: ethnicity, tumor site, source of control, smokers, alcohol drinkers, agreement with HWE in control group, and genotyping method. TSA was performed using diversity-adjusted information size based on $\alpha = 0.05$, $\beta = 0.20$ (power at 80%), information size with estimate, and case and control event proportion calculated from meta-analysis, through TSA software version 0.9 Beta [38–40].

The sensitivity analysis was conducted by deleting every included study each time to investigate the influence of overall results [43]. The cumulative analysis was performed to assess the change trend with the sample size accumulated according to the publication year [36]. The funnel plot and Egger’s test were used to evaluate
the underlying publication bias, and trim and fill method was applied if significant publication bias existed in the meta-analysis.

ACKNOWLEDGMENTS AND GRANT SUPPORT

This research was supported (in part) by the Nature Science Foundation of Hubei Province (2012FFB03902) and the Foundation of Evidence-based Medicine Nursery Fund of Taihe Hospital (EBM2013002 and EBM20140067), without commercial or not-for-profit sectors. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. No additional external funding received for this study. We also wish to thank relevant editors and peer-reviewers for their hard work and suggestions.

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

REFERENCES

1. Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. CA Cancer J Clin. 2014; 64:9–29. https://doi.org/10.3322/caac.21208.
2. Wozniak A, Szyfter K, Szyfter W, Florek E. [Head and neck cancer-history]. [Article in Polish]. Przegl Lek. 2012; 69:1079–83.
3. Conway DI, Hashibe M, Boffetta P, Wunsch-Filho V, Muscat J, La Vecchia C, Winn DM, INHANCE consortium. Enhancing epidemiologic research on head and neck cancer: INHANCE - The international head and neck cancer epidemiology consortium. Oral Oncol. 2009; 45:743–6. https://doi.org/10.1016/j.oraloncology.2009.02.007.
4. Lee YC, Boffetta P, Sturgis EM, Wei Q, Zhang ZF, Muscat J, Lazarus P, Matos E, Hayes RB, Winn DM, Zaridze D, Wunsch-Filho V, Eluf-Neto J, et al. Involuntary smoking and head and neck cancer risk: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. Cancer Epidemiol Biomarkers Prev. 2008; 17:1974–81. https://doi.org/10.1158/1055-9965.EPI-08-0047.
5. Mehanna H, Paleri V, West CM, Nutting C. Head and neck cancer—Part 1: Epidemiology, presentation, and prevention. BMJ. 2010; 341:c4684. https://doi.org/10.1136/bmj.c4684.
6. Kim MM, Califano JA. Molecular pathology of head-and-neck cancer. Int J Cancer. 2004; 112:545–53. https://doi.org/10.1002/ijc.20379.
7. Hunter K, Parkinson EK, Thakker N. An overview of the molecular pathology of head and neck cancer, and its clinical implications. Periodontol 2000. 2011; 57:132–49. https://doi.org/10.1111/j.1600-0757.2011.00387.x.
8. Hashibe M, Brennan P, Benhamou S, Castellsague X, Chen C, Curado MP, Dal Maso L, Dawd AW, Fabianova E, Fernandez L, Wunsch-Filho V, Franceschi S, Hayes RB, et al. Alcohol drinking in never users of tobacco, cigarette smoking in never drinkers, and the risk of head and neck cancer: pooled analysis in the International Head and Neck Cancer Epidemiology Consortium. J Natl Cancer Inst. 2007; 99:777–89. https://doi.org/10.1093/jnci/djk179.
9. Gandini S, Botteri E, Iodice S, Boni M, Lowenfels AB, Maisonneuve P, Boyle P. Tobacco smoking and cancer: a meta-analysis. Int J Cancer. 2008; 122:155–64. https://doi.org/10.1002/ijc.23033.
10. Zeng XT, Deng AP, Li C, Xia LY, Niu YM, Leng WD. Periodontal disease and risk of head and neck cancer: a meta-analysis of observational studies. PLoS One. 2013; 8:e79017. https://doi.org/10.1371/journal.pone.0079017.
11. Zeng XT, Luo W, Huang W, Wang Q, Guo Y, Leng WD. Tooth loss and head and neck cancer: a meta-analysis of observational studies. PLoS One. 2013; 8:e79074. https://doi.org/10.1371/journal.pone.0079074.
12. Zeng XT, Leng WD, Zhang C, Liu J, Cao SY, Huang W. Meta-analysis on the association between toothbrushing and head and neck cancer. Oral Oncol. 2015; 51:446–51. https://doi.org/10.1016/j.oraloncology.2015.02.095.
13. Foulkes WD, Brunet JS, Sieh W, Black MJ, Shenouda G, Narod SA. Familial risks of squamous cell carcinoma of the head and neck: retrospective case-control study. BMJ. 1996; 313:716–21.
14. Liu J, Zheng Y, Lei D, Liu D, Xu F, Jin T, Cao X, Zhao X, Yu X, Pan X. MDM2 309T>G polymorphism and risk of squamous cell carcinomas of head and neck: a meta-analysis, Asian Pac J Cancer Prev. 2011; 12:1899–903.
15. Zhang Y, Ni Y, Zhang H, Pan Y, Ma J, Wang L. Association between GSTM1 and GSTT1 allelic variants and head and neck squamous cell carcinoma. PLoS One. 2012; 7:e47579. https://doi.org/10.1371/journal.pone.0047579.
16. Wang Y, Chu X, Meng X, Zou F. Association of X-ray repair cross complementing group 1 Arg399Gln polymorphisms with the risk of squamous cell carcinoma of the head and neck: evidence from an updated meta-analysis. PLoS One. 2013; 8:e77898. https://doi.org/10.1371/journal.pone.0077898.
17. Leng WD, He MN, Chen QL, Gong H, Zhang L, Zeng XT. Vascular endothelial growth factor (VEGF) gene polymorphisms and risk of head and neck cancer: a meta-analysis involving 2,444 individuals. Mol Biol Rep. 2013; 40:5987–92. https://doi.org/10.1007/s11033-013-2708-y.
18. Zeng XT, Luo W, Geng PL, Guo Y, Niu YM, Leng WD. Association between the TP53 codon 72 polymorphism and risk of oral squamous cell carcinoma in Asians: a meta-analysis. BMC Cancer. 2014; 14:469. https://doi.org/10.1186/1471-2407-14-469.
19. Leng WD, Wen XJ, Kwong JS, Huang W, Chen JG, Zeng XT. COX-2 rs689466, rs5275, and rs20417 polymorphisms and
risk of head and neck squamous cell carcinoma: a meta-analysis of adjusted and unadjusted data. BMC Cancer. 2016; 16:457. https://doi.org/10.1186/s12885-016-2535-3.

20. Shihutani S, Takeda M, Grollman AP. Insertion of specific bases during DNA synthesis past the oxidation-damaged base 8-oxodG. Nature. 1991; 349:431–4. https://doi.org/10.1038/349431a0.

21. Yamane A, Kohno T, Ito K, Sunaga N, Aoki K, Yoshimura K, Murakami H, Nojima Y, Yokota J. Differential ability of polymorphic OGG1 proteins to suppress mutagenesis induced by 8-hydroxyguanine in human cell in vivo. Carcinogenesis. 2004; 25:1689–94. https://doi.org/10.1093/carcin/bgh166.

22. Poplawski T, Arabski M, Koziorowska D, Blasinska-Morawiec M, Morawiec Z, Morawiec-Bajda A, Klupinska G, Jeziorski A, Chojnacki J, Blasiak J. DNA damage and repair in gastric cancer—a correlation with the hOGG1 and RAD51 genes polymorphisms. Mutat Res. 2006; 601:83–91. https://doi.org/S0027-5107(06)00195-3.

23. Wang W, Wang M, Chen Y, Zhang Z, Wang S, Xu M, Wang B, Zhao Q. The hOGG1 Ser326Cys polymorphism contributes to cancer susceptibility: evidence from 83 case-control studies. Mutagenesis. 2012; 27:329–36. https://doi.org/10.1038/59369.

24. Wei B, Zhou Y, Xu Z, Xi B, Cheng H, Ruan J, Zhu M, Hu Q, Wang Q, Wang Z, Yan Z, Jin K, Zhou D, et al. The effect of hOGG1 Ser326Cys polymorphism on cancer risk: evidence from a meta-analysis. PLoS One. 2011; 6:e27545. https://doi.org/10.1371/journal.pone.0027545.

25. Elahi A, Zheng Z, Park J, Eyring K, McCaffrey T, Lazarus P. The human OGG1 DNA repair enzyme and its association with orolaryngeal cancer risk. Carcinogenesis. 2002; 23:1229–34.

26. Cho EY, Hildesheim A, Chen CJ, Blasinska-Morawiec M, Morawiec Z, Morawiec-Bajda A, Klupinska G, Jeziorski A, Chojnacki J, Blasiak J. DNA damage and repair in gastric cancer—a correlation with the hOGG1 and RAD51 genes polymorphisms. Mutat Res. 2006; 601:83–91. https://doi.org/S0027-5107(06)00195-3.

27. Zhang Z, Shi Q, Wang LE, Sturgis EM, Spitz MR, El-Naggar AK, Hong WK, Wei Q. No association between hOGG1 Ser326Cys polymorphism and risk of squamous cell carcinoma of the head and neck. Cancer Epidemiol Biomarkers Prev. 2004; 13:1081–3. https://doi.org/10.1093/carcin/bgl160.

28. Lu JT, Deng AP, Song J, Zhang L, Luo J. Reappraisal of XRCC1 Arg194Trp polymorphism and glioma risk: a cumulative meta-analysis. Oncotarget. 2017; 8:21599–21608. https://doi.org/10.18632/oncotarget.15376.

29. Weng H, Li JG, Mao Z, Zeng XT. Randomised trials of vitamin D3 for critically ill patients in adults: systematic review and meta-analysis with trial sequential analysis. Intensive Care Med. 2017; 43:277–8. https://doi.org/10.1007/s00134-016-4591-1.
40. Weng H, Li S, Zeng XT. Prevention of statins against AKI and mortality following cardiac surgery - Do we know enough? A trial sequential analysis. Int J Cardiol. 2017; 228:812–3. https://doi.org/10.1016/j.ijcard.2016.11.250.

41. Minelli C, Thompson JR, Abrams KR, Thakkinstian A, Attia J. How should we use information about HWE in the meta-analyses of genetic association studies? Int J Epidemiol. 2008; 37:136–46. https://doi.org/10.1093/ije/dym234.

42. Zeng X, Zhang Y, Kwong JS, Zhang C, Li S, Sun F, Niu Y, Du L. The methodological quality assessment tools for preclinical and clinical studies, systematic review and meta-analysis, and clinical practice guideline: a systematic review. J Evid Based Med. 2015; 8:2–10. https://doi.org/10.1111/jebm.12141.

43. Zhang J, Zeng XT, Lei JR, Tang YJ, Yang J. No association between XRCC1 gene Arg194Trp polymorphism and risk of lung cancer: evidence based on an updated cumulative meta-analysis. Tumour Biol. 2014; 35:5629–35. https://doi.org/10.1007/s13277-014-1745-z.

44. Stroup DF, Berlin JA, Morton SC, Olkin I, Williamson GD, Rennie D, Moher D, Becker BJ, Sipe TA, Thacker SB. Meta-analysis of observational studies in epidemiology: a proposal for reporting. Meta-analysis Of Observational Studies in Epidemiology (MOOSE) group. JAMA. 2000; 283:2008–12. https://doi.org/jst00003.

45. Liu J, Cao XL, Zhang Z, Lei DP, Jin T, Yu XM, Pan XL. HOGG1 Ser326Cys polymorphism and susceptibility to head and neck cancer: a meta-analysis. Asian Pac J Cancer Prev. 2011; 12:2491–4.

46. Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. BMJ. 2003; 327:557–60. https://doi.org/10.1136/bmj.327.7414.557.