| Title | Dissolved and Particulate Organic Matter Dynamics Relative to Sediment Resuspension Induced by the Tidal Cycle in Macrotidal Estuaries, Kyushu, Japan |
|-------|----------------------------------------------------------------------------------------------------------------------------------|
| Author(s) | Takasu, Hiroyuki; Uchino, Koji; Mori, Koichiro |
| Citation | Water, 12(9), art.no.2561; 2020 |
| Issue Date | 2020-09-13 |
| URL | http://hdl.handle.net/10069/40314 |
| Rights | ©2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/). |
Communication

Dissolved and Particulate Organic Matter Dynamics Relative to Sediment Resuspension Induced by the Tidal Cycle in Macrotidal Estuaries, Kyushu, Japan

Hiroyuki Takasu 1,2,* , Koji Uchino 3 and Koichiro Mori 3

1 Faculty of Environmental Science, Nagasaki University, 1-14 Bunkyo-machi, Nagasaki 852-8521, Japan
2 Division of Marine Energy Utilization, Organization for Marine Science and Technology, Nagasaki University, 1-14 Bunkyo-machi, Nagasaki 852-8521, Japan
3 Graduate School of Fisheries and Environmental Sciences, Nagasaki University, 1-14 Bunkyo-machi, Nagasaki 852-8521, Japan; bb53619005@ms.nagasaki-u.ac.jp (K.U.); bb53619025@ms.nagasaki-u.ac.jp (K.M.)

* Correspondence: takasu@nagasaki-u.ac.jp; Tel.: +81-95-819-2752

Received: 31 July 2020; Accepted: 12 September 2020; Published: 13 September 2020

Abstract: The tide-induced resuspension of sediments plays important roles in the efflux of particulate organic carbon (POC) from rivers to the coastal sea. Although dissolved organic carbon (DOC) comprises more than half of the riverine flux of organic carbon to the ocean, the influence of sediment resuspension induced by the tidal cycle on DOC dynamics is largely unknown. This study examined the dynamics of POC, particulate organic nitrogen (PON), DOC, humic-like and protein-like fluorescent dissolved organic matter (DOM) in relation to the tidal cycle in the Chikugo and Hayatsue River Estuaries, Kyushu, Japan. This study is the first to show both the particulate organic matter (POM) and DOM dynamics in relation to the tidal cycle in the macrotidal estuaries. We found that the turbidity changed with the tidal cycle and that there were significant positive correlations of turbidity with POC and PON in the Chikugo River mouth, suggesting that tide-induced sediment resuspension supplied POM to the water column. The DOC concentration, humic-like and protein-like fluorescent DOM were not correlated with turbidity but were negatively correlated with salinity. These results suggest that POM efflux, but not DOM efflux, can be accelerated by tide-induced sediment resuspension in the macrotidal estuaries.

Keywords: riverine organic matter; dissolved organic matter; tidal cycle; sediment resuspension; Ariake Sea; Chikugo River

1. Introduction

Estuaries, as transition zones between terrestrial and marine environments, are the most dynamic regions in the biosphere [1]. The tidal cycle may strongly affect organic matter dynamics, especially in macrotidal estuaries (tidal range > 4 m). A strong tidal current suspends sediments, which provide high concentrations of particulate organic carbon (POC) [2,3]. A previous study reported that a part of POC in the tidal-influenced turbidity water mass was bioavailable [4]. Thus, the tide-induced resuspension of sediments may play important roles in the efflux of labile POC from rivers to the coastal sea. Allochthonously input organic carbon fuels both pelagic and benthic food webs [5,6], whereas an excessive input of labile organic carbon often causes environmental problems, such as hypoxia, in coastal ecosystems [7]. Coastal hypoxia has been a global phenomenon since the late 1950s, as it causes the deterioration of marine ecosystems [8]. Thus, understanding the mechanisms controlling riverine organic matter delivery to the ocean is essential to maintain healthy coastal ecosystems.

Although dissolved organic carbon (DOC) comprises more than half of the riverine flux of organic carbon to the ocean [9] and a part of riverine DOC is highly bioavailable [10,11], the influence of sediment
resuspension induced by the tidal cycle on DOC dynamics is largely unknown. One study reported the infusion of DOC in the water column via active sediment resuspension [12]. In addition, high DOC concentrations in pore water were found in estuarine sediments [13]. The rapid production of DOC from particle decomposition in near-surface bottom sediments was also reported [14]. These processes may affect DOC dynamics via the active resuspension induced by tidal currents in macrotidal estuaries.

The Chikugo River, in Kyushu, southwestern Japan, is the largest river discharging into the Ariake Sea. Most riverine materials move from the Chikugo River to the northern part of the Ariake Sea, a semienclosed embayment with the largest tidal range in Japan. The Chikugo River Estuary is classified as a macrotidal estuary with tidal amplitudes of up to 5 m at its mouth. The Ariake Sea receives large amounts of organic carbon from the Chikugo River [15]. Recently, severe bottom-water hypoxia has occurred every summer in the northern part of the Ariake Sea, resulting in a marked ecosystem deterioration [16]. Thus, an understanding of the mechanisms supplying organic matter from the Chikugo River estuary is essential to prevent hypoxia in the northern part of the Ariake Sea. Suzuki et al. [17] reported that a strong tidal current in a macrotidal estuary can result in a high POC concentration due to active sediment resuspension. Thus, the tidal current can accelerate a high POC outflow from the river into the northern part of the Ariake Sea. Although the importance of DOC as a potential driver of hypoxia in the northern part of the Ariake Sea is recognized [18], the influence of tide-induced sediment resuspension on DOC efflux from the Chikugo River is not known. Therefore, we examined the temporal variations in both POC and DOC concentrations in relation to the tidal cycle in the Chikugo River Estuary to understand the influence of tide-induced sediment resuspension on POC and DOC efflux from the estuary to the northern part of the Ariake Sea. Stable carbon isotope ratios of POC were measured to understand the source of particulate organic matter (POM). Humic substances make up one of the most refractory organic groups in seawater, whereas protein (amino acids) is one of the most labile organic groups in seawater [19]. Thus, humic-like and protein-like fluorescent dissolved organic matter (DOM), based on the fluorescence excitation-emission matrix (EEM), was also measured to understand the influence of tide-induced sediment resuspension on labile DOM efflux.

2. Materials and Methods

We conducted investigations in late June (early summer) because severe hypoxia occurs in the summer in the northern part of the Ariake Sea. In order to avoid extreme cases, samplings were performed midway through the lunar tidal cycle before the rainy season (July). One can expect that, in the river mouth, the near-bottom current, which is strong enough to facilitate resuspension of sediments, will be observable during a tidal cycle. Thus, water samples were collected at fixed stations in the Chikugo and Hayatsue (a branch of the Chikugo River) River mouths (Figure 1) from 6:00 (low tide) to 18:00 (low tide) on 22 and 23 June 2019 (middle tide). The sediment characteristics of the two river mouths were similar (e.g., mud content, >74%; total organic carbon content, approx. 18 mg g\(^{-1}\) dw) [20]. The current velocity and direction were measured continuously using an electromagnetic current meter (INFINITY-EM, JFE Alec, Kobe, Japan; sampling interval 0.1 s) at a 1 m depth. The salinity and turbidity were measured at the surface and 1 m above the bottom using a multiparameter water quality meter (WQC-24; DKK-TOA, Tokyo, Japan) at 90-minute intervals. Seawater samples were collected using a 3 L Van Dorn water sampler (RIGO, Tokyo, Japan) from the surface and 1 m above the bottom at each station at 90-minute intervals.

Samples for chlorophyll a (Chl. a) and POC analyses were filtered on board using precombusted 0.7-µm GF/F filters (General Electric Company, Coventry, UK), and the filters were stored at −20 °C until further analysis. Samples for the DOM analysis were filtered on board using precombusted 0.7-µm GF/F filters (General Electric Company) and were stored in acid-washed 125-mL high-density polyethylene bottles at −20 °C until further analysis. Plastic gloves were worn during the sample collection and processing to avoid contamination.
10 samples, and the average pooled peak area of the Milli-Q water analysis over the entire day was
were fumed with HCl to remove carbonate salts, neutralized and dried in a desiccator, and were

Water 2020, 12, x FOR PEER REVIEW 3 of 10

Figure 1. The study area and sampling locations.

To determine the Chl. a concentration, the samples collected on GF/F filters were extracted with
N,N-dimethylformamide and analyzed by fluorometry (FP-8100; JASCO, Tokyo, Japan), according to
the method of Welschmeyer [21]. To determine the POC and particulate organic nitrogen (PON)
concentrations, and the stable carbon isotope ratios of POC, the samples collected on GF/F filters
were fumed with HCl to remove carbonate salts, neutralized and dried in a desiccator, and were
then analyzed using an elemental analyzer equipped with a stable isotope ratio mass spectrometer
(Flash 2000/Conflo IV/DELTA V Advantage, ThermoFisher Scientific, Bremen, Germany). Samples
and standards were individually wrapped in tin foil (Tin foil Squares, Säntis Analytical AG, Teufen,
Switzerland) before analysis. Stable isotope ratios are described as a per mil (‰) deviation from the
international standard, using the following equation: \( \delta X = \left( \frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right) \times 1000 \), where \( X \) and \( R \) represent the \(^{13}\text{C} \) and \(^{13}\text{C}/^{12}\text{C} \) ratio, respectively. PeeDee Belemnite and DL-alanine were used as the reference materials. The DOC concentrations were determined using a total carbon analyzer
(FOC-V; Shimadzu, Kyoto, Japan). A calibration curve was obtained by analyzing four distinct
centers of a standard solution prepared from potassium hydrogen phthalate. As a procedural
blank, ultrapure water (Milli-Q, Direct-Q UV3; Merck Millipore, MA, USA) was analyzed every
10 samples, and the average pooled peak area of the Milli-Q water analysis over the entire day was
subtracted from the seawater sample peak areas.

Excitation (Ex)/emission (Em) matrices of fluorescence were measured for DOC samples using a
fluorometer (FP-8100; JASCO, Tokyo, Japan). Bandwidths were set to 5 nm for excitation and 5 nm for
emission. A series of emission scans (200–500 nm) were collected over excitation wavelengths ranging
from 200–500 nm by 5 nm increments. To remove the Rayleigh scatter and the Raman signal, a Rayleigh-
and Raman-normalized Milli-Q Ex-Em matrix was subtracted from the sample data. The fluorescent
intensities were normalized with the quinine sulfate units (QSU), where 1 QSU is equivalent to the
fluorescence emission at an Ex/Em = 350/450 nm wavelength of 1 µg L\(^{-1} \) quinine sulfate in a 0.1 M
H$_2$SO$_4$ solution. According to [22], the peaks from Ex/Em = 220/290 nm and Ex/Em = 240/420 nm were determined as protein-like and humic-like components, respectively.

Pearson’s correlation analysis was used to analyze the data. A p-value of less than 0.05 was considered statistically significant. All statistical analyses were performed using R ver. 3.3.3 [23].

3. Results and Discussion

The water level reached a maximum at high tide (Figure 2a). The current velocity showed negative values from 7:30 to 11:30, indicating that seawater intrusion started and continued until high tide (Figure 2b). Then, the current velocity turned into positive values at 11:30, and this continued until 17:00. This means that river water kept outflowing until low tide. The salinity increased with time, reached a maximum at high tide, and then decreased until low tide (Figure 3a). The turbidity was higher at the bottom than at the surface in both rivers (Figure 3b). In both rivers, the turbidity tended to decrease from low to high tide and then increased until the next low tide. Although both ebb and flood tides induced sediment resuspension, it was greater during the ebb tide. A previous study reported that the transport of salinity and the river discharge in the Hayatsue River are weaker than those in the Chikugo River due to the thalweg topography of the Hayatsue River [24]. Consistent with the previous report, we found that the Chikugo River mouth was strongly influenced by the tidal current when compared to the Hayatsue River mouth (Figure 2b). A relatively high turbidity in the Chikugo River mouth (Figure 3b) can be caused by a high tidal current.

![Figure 2](image1.png)

**Figure 2.** Variations in the (a) water level and (b) current velocity during the sampling period. A positive value of the current velocity means a southward velocity (outflow from rivers).

The Chl. a concentration, a proxy for the phytoplankton biomass, changed in a similar manner as the salinity (Figure 3c). The Chl. a concentration increased with time, reached a maximum at high tide, and then decreased until low tide. This indicates that the flood tide transports marine-phytoplankton-derived POM from the Ariake Sea into the Chikugo and Hayatsue River Estuaries. However, the POC and PON concentrations changed in a direction opposite to that of the Chl. a concentration (Figures 3c and 4a,b). The POC and PON concentrations tended to decrease toward high tide and then to increase toward low tide. In macrotidal estuaries, a water mass with a high POC concentration results from the active resuspension of sediments by the strong tidal current, and the ebb tide accelerates the suspended POC discharge from the river to the coastal sea [25]. In this study, we found turbidity changes in relation to the tidal cycle (Figure 3a,b) and significant positive correlations of the turbidity with POC and PON in the Chikugo River mouth (Table 1), suggesting that the tidal current induced the resuspension of sediments, supplying POC to the water column. However,
such a relationship was not found in the Hayatsue River mouth (Table 2). A relatively weak tidal current in the Hayatsue River might not be enough to supply POC to the water column from sediments. The C/N ratios of POM changed in the opposite manner to the salinity (Figures 3a and 4c) and exhibited high values throughout the study (Figure 4c). Conversely, the δ13C values of POM changed in a similar manner to the salinity (Figures 3a and 4d) and exhibited low values. The δ13C values of POM are presented in relation to the C:N ratios in Figure 5. Previous studies have reported that each of the major sources of estuarine POM possesses distinctive δ13C values and C:N ratios: −30 to −24‰ and 5–10 for river-estuarine phytoplankton, −24 to −18‰ and 5–10 for marine phytoplankton, and −32 to −23‰ and higher than 20 for terrestrial C3 plants, respectively [26–31]. POM in both river mouths exhibited depleted δ13C values and high C:N ratios, and most of the plots were scattered within the range of terrestrial plants (Figure 5). Thus, resuspended POM is probably poorly bioavailable, since it is mainly composed of refractory terrestrial material [32]. Alternatively, depleted δ13C values and high C:N ratios might be a reflection of the presence of phytoplankton-derived detritus in POM [25]. A previous study reported a depletion of δ13C values and elevation of C:N ratios associated with the bacterial decomposition of phytoplankton [33]. This notion is partly supported by the results of Saliot et al. [34], who found that the concentrations of bacterial biomarkers in POM were generally high in the maximum turbidity zone in a macrotidal estuary.

**Figure 3.** Variations in the (a) salinity, (b) turbidity, and (c) chlorophyll a concentration during the sampling period.

The DOC concentration changed in the opposite manner to the salinity (Figures 3a and 6a) and was less variable than the POC concentration (Figure 4a,d) in both river mouths. Both protein-like and
humic-like components changed in a similar manner as DOC (Figure 6). A previous study suggested that DOC is supplied from the active resuspension of sediments induced by tidal currents through the infusion of DOC in pore waters and DOC release from suspended POM [35]. Nevertheless, in this study, the DOC concentration, protein-like and humic-like components were not correlated with turbidity but were negatively correlated with salinity (Tables 1 and 2). This conservative behavior of DOM suggests that sediment resuspension is not always a major determinant of DOM dynamics, including both labile and refractory fractions in macrotidal estuaries. Despite large differences in turbidity and DOM between the Chikugo and Hayatsue Rivers, consistent results were obtained. In addition, we did not find a significant positive correlation between turbidity and DOM around the Chikugo River mouth throughout seasonal monitoring (authors unpublished data). Those facts give us some added confidence about the robustness of our conclusion. Our conclusion may also be supported by the results of Kang et al. [36], who found that DOC fluxes were, in general, controlled by the water discharge in estuary systems.

**Figure 4.** Variations in the (a) particulate organic carbon (POC) concentration, (b) particulate organic nitrogen (PON) concentration, (c) POC/PON, and (d) δ13C values of POC during the sampling period.
Figure 5. Scatter diagrams of stable carbon isotope ratios ($\delta^{13}C$) against organic carbon-to-nitrogen atomic ratios (C:N). The $\delta^{13}C$ and C:N ranges for river-estuarine and marine phytoplankton, and for terrestrial plants, are derived from references [26–31].

Figure 6. Variations in the (a) dissolved organic carbon (DOC) concentration, (b) protein-like component, and (c) humic-like component during the sampling period.
Table 1. Pearson’s correlation coefficients among salinity (Sal), chlorophyll a (Chl.a), turbidity (Turb), particulate organic carbon (POC), particulate organic nitrogen (PON), δ¹³C values of POC, dissolved organic carbon (DOC), protein-like component (Protein), and humic-like component (Humic) in the Chikugo River. *, p < 0.05; **, p < 0.01; ***, p < 0.001.

| Variables | Sal  | Chl.a | Turb | POC  | PON  | δ¹³C | DOC   | Protein | Humic |
|-----------|------|-------|------|------|------|------|-------|---------|-------|
| Sal       | 0.67 | ***   |      |      |      |      |       |         |       |
| Chl.a     | -0.25|       | -0.25|      |      |      |       |         |       |
| Turb      | 0.01 | -0.08 | 0.58 | ***  |      |      |       |         |       |
| POC       | 0.13 | 0.02  | 0.47 | *    | 0.98 | ***  |       |         |       |
| PON       | 0.76 | ***   | 0.76 | ***  | -0.26| 0.11 | -0.18 |         |       |
| δ¹³C      | -0.55|       | -0.46| 0.31 | 0.17 | 0.05 | -0.71 | ***     |       |
| DOC       | -0.39|       | 0.21 | 0.19 | 0.24 | 0.20 | -0.38 | 0.61    | **    |
| Protein   | -0.84| ***   | -0.62| **   | 0.41 | 0.24 | 0.14  | -0.87   | ***   |
| Humic     | -0.54|       |       |      |      |     |       | -0.54   |       |

Table 2. Pearson’s correlation coefficients among salinity (Sal), chlorophyll a (Chl.a), turbidity (Turb), particulate organic carbon (POC), particulate organic nitrogen (PON), δ¹³C values of POC, dissolved organic carbon (DOC), protein-like component (Protein), and humic-like component (Humic) in the Hayatsue River. *, p < 0.05; **, p < 0.01; ***, p < 0.001.

| Variables | Sal  | Chl.a | Turb | POC  | PON  | δ¹³C | DOC   | Protein | Humic |
|-----------|------|-------|------|------|------|------|-------|---------|-------|
| Sal       | 0.67 | ***   |      |      |      |      |       |         |       |
| Chl.a     | 0.01 | 0.09  |      |      |      |      |       |         |       |
| Turb      | -0.56|       | 0.42 | 0.02 |      |      |       |         |       |
| POC       | -0.21|       | 0.24 | 0.24 | 0.82 | ***  |       |         |       |
| PON       | 0.74 | ***   | 0.91 | ***  | -0.12| -0.54| *     | -0.34   |       |
| δ¹³C      | -0.81| ***   | -0.62| **   | -0.07| 0.48 | *     | 0.14    | -0.71 | *** |
| DOC       | -0.41|       | -0.34|      | -0.01| 0.26 | 0.23  | -0.47   | *     | 0.38 |
| Protein   | -0.95| ***   | -0.71| ***  | -0.14| 0.54 | *     | -0.80   | ***   | 0.91 | *** |
| Humic     | 0.40 |       |       |      |      |      |       |         |       |

4. Conclusions

In conclusion, this study is the first to show both POM and DOM dynamics in relation to the tidal cycle in macrotidal estuaries. In these estuaries, the POM dynamics are strongly affected by the resuspension of sediment organic matter induced by the tidal cycle, whereas the resuspension of sediment has a minor effect on both the refractory and labile DOM dynamics. The POM efflux can be accelerated by tide-induced sediment resuspension in the Chikugo River mouth. However, sediment resuspension is not always a major determinant of DOM dynamics in macrotidal estuaries. Future studies should assess the effects of variable tides on POM and DOM dynamics in macrotidal estuaries to improve our understanding of the controlling mechanisms of organic matter discharge from rivers.

Author Contributions: H.T., K.M. and K.U. designed the experiment and conducted the field sampling together. K.M. and K.U. conducted chemical analyses. H.T. analyzed data and wrote the paper. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported in part by JSPS KAKENHI grants no. 18K14511 to HT.

Acknowledgments: We thank Toshihiro Miyajima and Nobue Saotome (The University of Tokyo) for their assistance with the POM analysis. We are grateful to the anonymous reviewers as their comments helped us to greatly improve the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.
References

1. Gattuso, J.P.; Frankignoulle, M.; Wollast, R. Carbon and carbonate metabolism in coastal aquatic ecosystems. Annu. Rev. Ecol. Syst. 1998, 29, 405–434. [CrossRef]

2. Findlay, S.; Pace, M.; Lints, D. Variability and transport of suspended sediment, particulate and dissolved carbon in the tidal freshwater Hudson River. Biogeochimistry 1991, 12, 149–169. [CrossRef]

3. Bianchi, T.S. Biogeochemistry of Estuaries; Oxford University Press: New York, NY, USA, 2007; p. 706.

4. Etcheber, H.; Taillez, A.; Abril, G.; Garnier, J.; Servais, P.; Moatar, F.; Commarieu, M.V. Particulate organic carbon in the estuarine turbidity maxima of the Gironde, Loire and Seine estuaries: Origin and lability. Hydrobiologia 2007, 588, 245–259. [CrossRef]

5. Sandberg, J.; Andersson, A.; Johansson, S.; Wikner, J. Pelagic food web structure and carbon budget in the northern Baltic Sea: Potential importance of terrigenous carbon. Mar. Ecol. Prog. Ser. 2004, 268, 13–29. [CrossRef]

6. Bartels, P.; Ask, J.; Andersson, A.; Karlsson, J.; Giesler, R. Allochthonous organic matter supports benthic but not pelagic food webs in shallow coastal ecosystems. Ecosystems 2018, 21, 1459–1470. [CrossRef]

7. Paerl, H.W.; Pinckney, J.L.; Fear, J.M.; Peierls, B.L. Ecosystem responses to internal and watershed organic matter loading: Consequences for hypoxia in the eutrophying Neuse River Estuary, North Carolina, USA. Mar. Ecol. Prog. Ser. 1998, 166, 17–25. [CrossRef]

8. Diaz, R.J.; Rosenberg, R. Spreading dead zones and consequences for marine ecosystems. Science 2008, 321, 926–929. [CrossRef]

9. Hedges, J.I.; Keil, R.G.; Benner, R. What happens to terrestrial organic matter in the ocean? Org. Geochem. 1997, 27, 195–212. [CrossRef]

10. Rochelle-Newall, E.J.; Pizay, M.-D.; Middelburg, J.J.; Boschker, H.T.S.; Gattuso, J.-P. Degradation of riverine dissolved organic matter by seawater bacteria. Aquat. Microb. Ecol. 2004, 37, 9–22. [CrossRef]

11. Wu, K.; Lu, K.; Dai, M.; Liu, Z. The bioavailability of riverine dissolved organic matter in coastal marine waters of southern Texas. Estuar. Coast. Shelf Sci. 2019, 231, 106477. [CrossRef]

12. Gerringa, L.J.A. Aerobic degradation of organic matter and the mobility of Cu, Ni, Pb, Zn, Fe and Mn in marine sediment slurries. Mar. Chem. 1990, 29, 355–374. [CrossRef]

13. Burdige, D.J.; Zheng, S. The biogeochemical cycling of dissolved organic nitrogen in estuarine sediments. Limnol. Oceanogr. 1998, 43, 1796–1813. [CrossRef]

14. Cauwet, G.; Mackenzie, F.T. Carbon inputs and distribution in estuaries of turbid rivers: The Yang Tze and Yellow rivers (China). Mar. Chem. 1993, 43, 246–253. [CrossRef]

15. Suzuki, K.W.; Kasai, A.; Nakayama, K.; Tanaka, M. Year-round accumulation of particulate organic matter in the estuarine turbidity maximum: Comparative observations in three macrotidal estuaries (Chikugo, Midori, and Kuma Rivers), southwestern Japan. J. Oceanogr. 2012, 68, 453–471. [CrossRef]

16. Yoshino, K.; Hamada, T.; Yamamoto, K.; Hayami, Y.; Yamaguchi, S.; Ohgushi, K. Effects of hypoxia and organic enrichment on estuarine macrofauna in the inner part of Ariake Bay. Hydrobiologia 2010, 652, 23–38. [CrossRef]

17. Suzuki, K.W.; Sugimoto, R.; Kasai, A.; Nakayama, K.; Tanaka, M. Dynamics of particulate organic matter in the estuarine turbidity maximum of the Chikugo River estuary, Ariake Sea, in summer: Influence of the fluctuation of freshwater discharge. Bull. Jpn. Soc. Fish. Oceanogr. 2009, 73, 149–160. (In Japanese with English Abstract)

18. Uchino, K.; Inomata, H.; Tahara, S.; Takasu, H. Contribution of organic matter decomposition in water column to oxygen consumption in the inner part of the Ariake Sea. J. Jpn. Soc. Water Environ. 2019, 42, 195–200. (In Japanese with English Abstract) [CrossRef]

19. Ogawa, H.; Tanoue, E. Dissolved organic matter in oceanic waters. J. Oceanogr. 2003, 59, 129–147. [CrossRef]

20. Yokoyama, K.; Yamamoto, K.; Kawano, S. Morphodynamics and transport of sediment phosphorus in the northeast part of the Ariake Sea and the estuarine channel of the Chikugogawa River. J. Jpn. Soc. Civ. Eng. B 2008, 64, 366–370. (In Japanese with English Abstract)

21. Welschmeyer, N.A. Fluorometric analysis of chlorophyll-a in the presence of chlorophyll-b and pheopigments. Limnol. Oceanogr. 1994, 39, 1985–1992. [CrossRef]

22. Mayer, L.M.; Schick, L.L.; Loder, T.C., III. Dissolved protein fluorescence in two Maine estuaries. Mar. Chem. 1999, 64, 171–179. [CrossRef]
23. R Development Core Team. *R: A Language and Environment for Statistical Computing;* R Foundation for Statistical Computing: Vienna, Austria, 2017.

24. Matsumura, K.; Morimura, Y.; Shintani, T.; Yokoyama, K. Three-dimensional numerical simulation of saline intrusion in Chikugo River estuary. *J. Ipn. Soc. Civ. Eng.* **B1 2017**, *73*, I_1039–I_1044. (In Japanese with English Abstract)

25. Hayami, Y.; Wada, M.; Umezawa, Y.; Fujii, N.; Nakamura, A.; Mori, F. Hypoxic water mass in the highly turbid well-mixed macrotidal Rokkaku River Estuary, Ariake Sea, Japan. *Estuar. Coast. Shelf Sci.* **2019**, *219*, 210–222. [CrossRef]

26. Redfield, A.C.; Ketchum, B.H.; Richards, F.A. The influence of organisms on the composition of seawater. In *The Sea: Ideas and Observations on Progress in the Study of the Seas*; Hill, M.N., Ed.; Wiley: New York, NY, USA, 1963; Volume 2, pp. 26–77.

27. Deines, P. The isotopic composition of reduced organic carbon. In *Handbook of Environmental Isotope Geochemistry;* The Terrestrial Environment; Fritz, P., Fontes, J.C., Eds.; Elsevier: Amsterdam, The Netherlands, 1980; Volume 1, pp. 329–406.

28. Fry, B.; Sherr, E.B. $\delta^{13}$C measurements as indicators of carbon flow in marine and freshwater ecosystems. *Contrib. Mar. Sci.* **1984**, *27*, 13–47.

29. Fontugne, M.R.; Jouanneau, J.M. Modulation of the particulate organic carbon flux to the ocean by a macrotidal estuary: Evidence from measurements of carbon isotopes in organic matter from the Gironde system. *Estuar. Coast. Shelf Sci.* **1987**, *24*, 377–387. [CrossRef]

30. Meyers, P.A. Preservation of elemental and isotopic source identification of sedimentary organic matter. *Chem. Geol.* **1994**, *114*, 289–302. [CrossRef]

31. Fuji, M.; Murashige, S.; Ohnishi, Y.; Yuzawa, A.; Miyasaka, H.; Suzuki, Y.; Komiyama, H. Decomposition of phytoplankton in seawater. Part I. Kinetic analysis of the effect of organic matter concentration. *J. Oceanogr.* **2002**, *58*, 433–438. [CrossRef]

32. Savoye, N.; David, V.; Morisseau, F.; Etcheber, H.; Abril, G.; Billy, I.; Charlier, K.; Oggyan, G.; Derrienic, H.; Sautour, B. Origin and composition of particulate organic matter in a macrotidal turbid estuary: The Gironde Estuary, France. *Estuar. Coast. Shelf Sci.* **2012**, *108*, 16–28. [CrossRef]

33. Lehmann, M.F.; Bernasconi, S.M.; Barbieri, A.; McKenzie, J.A. Preservation of organic matter and alteration of its carbon and nitrogen isotope composition during simulated and in situ early sedimentary diagenesis. *Geochim. Cosmochim. Acta* **2002**, *66*, 3573–3584. [CrossRef]

34. Saliot, A.; Tronczynski, J.; Scribe, P.; Letolle, R. The application of isotopic and biogeochemical markers to the study of the biochemistry of organic matter in a macrotidal estuary, the Loire, France. *Estuar. Coast. Shelf Sci.* **1988**, *27*, 645–669. [CrossRef]

35. Middelburg, J.J.; Herman, P.M.J. Organic matter processing in tidal estuaries. *Mar. Chem.* **2007**, *106*, 127–147. [CrossRef]

36. Kang, S.; Kim, J.H.; Kim, D.; Song, H.; Ryu, J.S.; Ock, G.; Shin, K.H. Temporal variation in riverine organic carbon concentrations and fluxes in two contrasting estuary systems: Geum and Seomjin, South Korea. *Environ. Int.* **2019**, *133*, 105126. [CrossRef] [PubMed]

© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).