The application of methylene blue coloration technique in axillary lymph node dissection of breast cancer

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**Introduction**

In recent years, because of higher incidence, more attention has been given to breast cancer. It has been reported that breast cancer is the most common cancer among women in many Chinese cities (1). Surgery always plays an important role in the treatment of breast cancer. No matter how the method changes or the extent of the breast cancer surgery, ALND remains the standard surgical treatment for node-positive disease. It is performed to confirm the axillary...
lymph node status, which is important as a prognostic factor for breast cancer (2-4).

For classification of lymph node stations within the axilla, the description of three axillary levels, defined by the pectoralis minor muscle, prevails. In general, axillary lymph node metastasis obeys the laws of levels I to II to III; however, there are also some that do not follow this rule, called Skip Metastasis, of which the incidence is 1.5–19.2% (5). For the axillary lymph nodes of high areas (called high lymph nodes in our research), patients treated with clearing this area are at the greatest risk for developing impairments of the arm and shoulder; upper limb lymphedema, pain, and sensitivity disorders were reported most frequently (6). Although controversial, it is considered that the lymph nodes with higher positions also belong to the lymphatic drainage area of axilla and that there is a risk of skip metastasis, which makes its removal necessary (7). Therefore, it is of great significance to identify and remove the lymph nodes with higher positions.

Using the appropriate lymph tracer can improve the detection rate of sentinel lymph nodes as well as biopsy rates (8). Methylene blue (MB) can be used widely in sentinel node biopsies (SLNB), as it is characterized by cost effectiveness, avirulence, long residence times in lymph nodes; reports have shown that there is no significance in using MB alone in SLNB versus using a combined isotope tracer (9). Initial investigators used blue dye to identify the ARM nodes or lymphatics. The concept of ARM involves mapping the lymphatic drainage from the upper extremity to determine the anatomical lymphatic variation and thus provides a roadmap to preserving these nodes during ALND, thereby minimizing arm lymphedema (10,11).

Over the past decade, MB has been used widely in clinics, and it has a good effect on the SLNB tracer (9,12). However, researchers have been less enthusiastic about using MB in high lymph nodes in ALND. There is no consensus among researchers on clinical factors that affect the coloration of MB, and this missing conclusion has important guiding significance to clinical practice (13,14).

The present clinical trial was designed to analyze injection time, position and age, BMI, molecular typing, TNM staging, and neoadjuvant chemotherapies that affect the coloration of MB. The study was also designed to analyze the application of MB.

**Methods**

**Patients**

We studied 170 patients diagnosed with breast cancer from December 2016 to December 2017 in the Department of Second Breast Surgery of Yunnan Cancer Hospital.

A total of 136 cases were diagnosed as invasive ductal carcinoma, accounting for 80% of the cases. There were 8 cases of invasive lobular carcinoma and 15 cases of mucinous adenocarcinoma, accounting for 4.70% and 8.82%, respectively. The remaining 11 cases of breast cancer accounted for 6.47% of the enrolled patients. Basic demographic information, clinical diagnosis, treatment information and laboratory test information were collected by consulting medical records. Information collection strictly followed the principle of informed consent and the principle of patient information confidentiality. The ethics review of this study was completed by the medical ethics committee of Kunming Medical University (no: 2016-030-KJ-QT).

**Injection of MB injection position**

The positions of MB injection are the areola area (**Figure 1**) and the outer-upper quadrant of the breast (**Figure 2**). If patients were afflicted with tumors located in the outer-upper quadrant, with “local excision” or “biopsy”, injection of MB could be done outside of the selected area, away from the tumor, using 75% alcohol to disinfect the areola area and then inject at 3, 6, 9, and 12 o’clock sites of 0.5 mL MB. Injection of 2 mL MB into the outer-upper quadrant of breast was done to make sure the dot of injection is within the scope of the mastectomy.
Injection time

A total of 170 patients were divided into seven groups randomly; the groups were defined by minutes between the MB injection time and the ALND: 10, 20, 30, 40, 60, 120, and 240 minutes before ALND. After injection, the injection site was gently massaged with a medical cotton ball to facilitate the introduction of MB into the lymphatic vessels for drainage. Coloration of the axillary lymph nodes and lymphatics was observed.

Operation methods

After the anesthesia had taken effect, the patient was in a supine position, and her limbs in brackets, cleaning of the levels I and II lymph nodes was performed and then, cleaning of the lymph nodes of the upper axillary vein was performed. Using an ultrasonic knife, the upper axillary vein was fully exposed; having upward separation, we can see a “palisade” area, called the “upper axillary vein lymph node”, noticeably clearing and packing the tissue. It was necessary to pay more attention to preserve blood vessels and nerves. After surgery, relevant pathological data were collected.

Statistical analysis

Analysis of clinical factors

To analyze the clinical factors that affected the coloration of MB, first, a single factor analysis was carried out using a $\chi^2$ test. In addition to this single factor analysis of $P<0.1$, the reports of MB color influence factors (13-16) were fed into a Logistic regression model for multivariate analysis; we set MB color as the dependent variable ($1=$ not color, $2=$ color).

To analyze the application of MB in the lymph node upper axillary vein

We evaluated the tracer effect of MB, compared with the biopsy results. We calculated sensitivity (the positive percentage of cases MB colored in axillary lymph node metastasis), specificity (the percentage of cases MB uncolored in axillary lymph node without metastasis), false negative rate (1-sensitivity) and false positive rate (1-specificity) to evaluate its accuracy. We calculated the kappa coefficient to evaluate its reliability: $K=1$ for both completely consistent, $0.4<K<0.75$ for both moderately consistent, $K\geq0.75$ for both excellently consistent, $K\leq0.4$ for both poorly consistent, when $K<0.4$; we did the $\chi^2$ test. SPSS 19.0 software was used for statistical analysis, and the alpha was set to $\alpha=0.05$.

Results

Clinical factors that affect the coloration of MB patients

A total of 170 patients were enrolled in the study; their mean age was 50.58±10.35 years, the minimum age was 27, and the maximum age was 85. The mean body mass index (BMI) was 23.55±3.32. In the clinical stage, there were 16 patients in stage I (9.41%), 118 patients in stage II (69.41%), 24 patients in stage III (14.12%) and 6 patients in stage IV (3.53%); 6 patients’ (3.53%) stages were not confirmed. Concerning the molecular typing of breast cancer, there were 40 patients of Luminal A (23.53%), 79 patients of Luminal B (46.47%), 24 patients of ERBB2+ (14.12%), and 27 patients of Basal-like (15.88%). A total of 138 patients with axillary lymph nodes and lymphatic vessels showed blue (colored group, 81.18%), 32 cases did not show blue (uncolored group, 18.82%) (Figures 3-5).

The results of single factor analysis

The results of single factor analysis showed that BMI ($\chi^2=24.074$, $P<0.0001$) and injection time ($\chi^2=41.207$, $P<0.0001$) were the clinical factors that affected the coloration of MB. Age, BMI, molecular typing, TNM staging, neoadjuvant chemotherapy and the injection position had no significant influence on the coloration of MB ($P>0.05$). BMI ($\chi^2=24.074$, $P<0.0001$) (Table 1).
The results of multivariate analysis

The results of multivariate analysis showed that injection time ($P=0.016$) was the clinical factor that affected the coloration of MB. Injection of MB more than 60 minutes before surgery resulted in 0.088 times higher than 10 minutes before surgery ($P=0.010$, 95% CI, 0.014, 0.554) (Table 2).

The application of MB in the lymph nodes of the upper axillary vein

In this study, 84 cases were selected for observation of the lymph nodes of the upper axillary vein. In some cases, MB is seen crossing the axillary vein along the lymphatic duct (Figure 5).

The results showed that 40 patients had no axillary lymph node metastasis, and 44 patients had axillary lymph node metastasis. Among the 84 patients, 69 cases of MB showed blue coloration. A total of 62 cases had no lymph node metastasis in the upper axillary vein, while 7 cases had lymph node metastasis upper axillary vein (Table 3).

MB was used to track the lymph nodes upper axillary vein with a sensitivity of 12.5%, a specificity of 100%, a false negative rate of 87.50%, a false positive rate of 0, and a kappa coefficient of 0.051 ($\chi^2=0.679$, $P=0.404$).

Discussion

An excellent stain, such as MB, to be used for analysis of the lymphatic system and lymph nodes should be characterized by cost effectiveness, avirulence, long residence times in the tissues and color clarity, among others. A large number of experiments have confirmed the accuracy and reliability of MB, and it is widely used in clinics.

This study enrolled 170 cases, and 138 cases were...
Table 1 The results of single factor analysis of clinical factors that affect the coloration of methylene blue

| Clinical factors                  | Colored group, N=138 (%) | Uncolored group, N=32 (%) | Total, N=170 (%) | \( \chi^2 \) | P   |
|----------------------------------|--------------------------|---------------------------|------------------|------------|-----|
| Ages (yr)                        |                          |                           |                  |            |     |
| \( \leq 40 \)                    | 26 (18.84)               | 2 (6.25)                  | 28 (16.47)       | 4.652      | 0.199|
| 41–50                            | 46 (33.33)               | 14 (43.75)                | 60 (35.29)       |            |     |
| 51–60                            | 44 (31.88)               | 13 (40.63)                | 57 (33.53)       |            |     |
| \( \geq 60 \)                    | 22 (15.94)               | 3 (9.38)                  | 25 (14.71)       |            |     |
| BMI                              |                          |                           |                  | 24.074     | <0.0001|
| \( < 18.5 \)                     | 10 (7.25)                | 0 (0.0)                   | 10 (5.88)        |            |     |
| 18.5–24.9                        | 91 (65.94)               | 9 (28.13)                 | 100 (58.82)      |            |     |
| 25.0–29.9                        | 35 (25.36)               | 19 (59.38)                | 54 (31.76)       |            |     |
| 30.0–34.9                        | 2 (1.45)                 | 4 (12.50)                 | 6 (3.53)         |            |     |
| Molecular subtype                |                          |                           |                  | 2.86       | 0.414|
| Luminal A                        | 32 (23.19)               | 8 (25.0)                  | 40 (23.53)       |            |     |
| Luminal B                        | 61 (44.20)               | 18 (56.25)                | 79 (46.47)       |            |     |
| ERBB2+                           | 22 (15.94)               | 2 (6.25)                  | 24 (14.12)       |            |     |
| Basal-like                       | 23 (16.67)               | 4 (12.50)                 | 27 (15.88)       |            |     |
| TNM stage                        |                          |                           |                  | 1.875      | 0.759|
| Stage I                          | 12 (8.70)                | 4 (12.50)                 | 16 (9.41)        |            |     |
| Stage II                         | 96 (69.57)               | 22 (68.75)                | 118 (69.41)      |            |     |
| Stage III                        | 21 (15.22)               | 3 (9.38)                  | 24 (14.18)       |            |     |
| Stage IV                         | 5 (3.62)                 | 1 (3.13)                  | 6 (3.53)         |            |     |
| Stages were not confirmed        | 4 (2.90)                 | 2 (6.25)                  | 6 (3.53)         |            |     |
| Neoadjuvant chemotherapy         |                          |                           |                  | 0.249      | 0.618|
| Yes                              | 58 (42.03)               | 15 (46.88)                | 73 (42.94)       |            |     |
| No                               | 80 (57.97)               | 17 (53.13)                | 97 (57.06)       |            |     |
| Injection time (min)             |                          |                           |                  | 41.207     | <0.0001|
| 10                               | 28 (20.29)               | 2 (6.25)                  | 30 (17.65)       |            |     |
| 20                               | 30 (21.74)               | 0 (0.00)                  | 30 (17.65)       |            |     |
| 30                               | 28 (20.29)               | 2 (6.25)                  | 30 (17.65)       |            |     |
| 40                               | 26 (18.84)               | 4 (12.50)                 | 30 (17.65)       |            |     |
| \( \geq 60 \)                    | 26 (18.84)               | 24 (75.00)                | 50 (29.41)       |            |     |
| Injection position               |                          |                           |                  | 1.079      | 0.299|
| Areola area                      | 95 (68.84)               | 25 (78.13)                | 120 (70.59)      |            |     |
| The outer upper quadrant of breast | 43 (31.16)           | 7 (21.88)                 | 50 (29.41)       |            |     |
colored, accounting for 81.17%. Single factor analysis showed that BMI ($\chi^2=24.074$, $P<0.0001$) and injection time ($\chi^2=41.207$, $P<0.0001$) are clinical factors that affect the coloration of MB. Age, BMI, molecular typing, TNM staging, neoadjuvant chemotherapy and injection position had no significant influences on the coloration of MB ($P>0.05$). The results of the multivariate analysis showed that injection time ($P=0.016$) is a clinical factor that affects the coloration of MB. Injection of MB more than 60 minutes before surgery resulted in 0.088 times higher the possibility of MB colored than 10 minutes before surgery ($P=0.010$, 95% CI, 0.014, 0.554). We injected MB

Table 2 The results of multivariate analysis of clinical factors that affect the coloration of methylene blue

| Clinical factors          | Partial regression coefficients | Standard error | Wald value | P    | OR   | 95% CI         |
|--------------------------|--------------------------------|----------------|------------|------|------|----------------|
| Ages(\text{yr})          |                                |                |            |      |      |                |
| ≤40*                     |                                |                |            |      |      |                |
| 41–50                    | -2.046                         | 1.043          | 3.849      | 0.05 | 0.129| (0.017, 0.998) |
| 51–60                    | -1.291                         | 1.033          | 1.561      | 0.212| 0.275| (0.036, 2.084) |
| ≥60                      | -1.001                         | 1.193          | 0.703      | 0.402| 0.368| (0.035, 3.811) |
| BMI                      |                                |                |            |      |      |                |
| <18.5*                   |                                |                |            |      |      |                |
| 18.5–24.9                | -19.296                        | 10,934.018     | 0.0001     | 0.999| –    | –              |
| 25.0–29.9                | -20.747                        | 10,934.018     | 0.0001     | 0.999| –    | –              |
| 30.0–34.9                | -39.088                        | 12,306.456     | 0.0001     | 0.999| –    | –              |
| Molecular subtype        |                                |                |            |      |      |                |
| Luminal A*               |                                |                |            |      |      |                |
| Luminal B                | -0.515                         | 0.667          | 0.58       | 0.446| 0.597| (0.159, 2.250) |
| ERBB2+                   | 1.189                          | 1.132          | 1.103      | 0.294| 3.283| (0.357, 30.178)|
| Basal-like               | 0.294                          | 0.956          | 0.095      | 0.758| 1.342| (0.206, 8.747) |
| TNM stage                |                                |                |            |      |      |                |
| Stage I*                 |                                |                |            |      |      |                |
| Stage II                 | -0.084                         | 0.896          | 0.009      | 0.925| 0.92 | (0.159, 5.321) |
| Stage III                | 0.282                          | 1.126          | 0.063      | 0.802| 1.326| (0.146, 12.052)|
| Stage IV                 | -0.812                         | 1.66           | 0.239      | 0.625| 0.444| (0.017, 11.487)|
| Stages were not confirmed| -1.563                         | 1.652          | 0.895      | 0.344| 0.21 | (0.008, 5.339) |
| Neoadjuvant chemotherapy | 0.347                          | 0.616          | 0.318      | 0.573| 1.416| (0.042, 4.739) |
| Injection time (mins)**  |                                |                |            |      |      |                |
| 10*                      |                                |                |            |      |      |                |
| 20                       | 32.471                         | 8,048.385      | 0.0001     | 0.997| –    | –              |
| 30                       | 0.425                          | 1.205          | 0.125      | 0.724| 1.53 | (0.144, 16.225)|
| 40                       | -0.864                         | 1.046          | 0.681      | 0.409| 0.422| (0.054, 3.276) |
| ≥60                      | -2.433                         | 0.94           | 6.699      | 0.010| 0.088| (0.014, 0.554) |
| Injection position       | 1.315                          | 0.794          | 2.743      | 0.098| 3.726| (0.786, 17.666)|

*, comparable group; **, $P<0.05$. 

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in the areola area or the outer-upper quadrant of the breast; injection site was not shown to be a factor that affects the coloration of MB. The results are similar to some published studies, which investigated the use of MB in SLNB (15-20). MB can be a significant guide when identifying clear lymphatic vessels and lymph nodes in ALND.

Several researchers used blue dye approximately five minutes before SLNB (21,22). Sakurai et al. (23) performed surgery immediately after the injection of blue dye in ARM. In this study, the fastest time of separating skin and subcutaneous tissue in ALND was 10 minutes. The injection time of 10, 20 and 30 minutes can guarantee a MB color rate of above 90%. As time goes on, the color effect decreases. Injection of MB more than 60 minutes before surgery resulted in 0.088 times higher the possibility of MB colored than 10 minutes before surgery. Therefore, the injection time should be less than 60 minutes when using MB to guide ALND.

In a meta-analysis published by Pesek et al. (24), there were seven categories based on location of injection, reported by 302 articles. Four categories consisted of a single location site, while three categories represented a combination of locations. There were no statistically significant variations between these seven location categories (P=0.95). Intra-tumor injections had the lowest numerical FNR at 2.5%. Similarly, Ahmed et al. (25) also considered that there was no significant difference between superficial and deep injections of radioactive tracer for axillary SLN identification on lymphoscintigraphy. In our research, there was no statistical significance between areola and outer-upper quadrant for the injection of MB in ALND. If the clinical lesions were located in the outer area, a biopsy may be the cause of local tissue edema or lymphatic drainage structure change, which could affect the path of MB tracer. It is suggested that increasing injection quantity in the outer area or changing the injection site can improve the detection rate of lymph nodes. Nevertheless, it is a problem that the injection position diffusion zone with the axillary nodes is often overlapped with tumors located in the upper outer quadrant and axillary tail of the breast, or patients have undergone biopsies in the upper-outer quadrant of the breast. Thus, injecting MB in the subareolar area is a better choice (26).

In this study, 84 cases were selected for observation of the lymph nodes in the upper axillary vein. A total of 69 cases of MB showed blue. A total of 62 cases had no lymph node metastasis in the upper axillary vein, and 7 cases had lymph node metastasis in the upper axillary vein. MB was used to track the lymph nodes in the upper axillary vein and showed a sensitivity of 12.5%, a specificity of 100%, a false negative rate of 87.50%, a false positive rate of 0, and a kappa coefficient of 0.051 (χ²=0.679, P=0.404). The results showed that methylene blue trace has a high false negative rate, low sensitivity, and poor consistency. We suggest that MB can serve as an excellent guide for identifying and clearing lymphatic vessels and lymph nodes in ALND; however, it is still prudent to use MB for high lymph node dissection.

It is reported that the detection rate of SLN was 85–95%, the accuracy rate was 71–98%, and the false negative rate (FNR) was 0–29% when MB was used in SLNB (9,27). In ALND, MB is used to perform axillary reverse mapping (ARM); different identification rates (DR) are reported for ARM nodes ranging from 65% to 82% by different investigators (11,28). In the meta-analysis published by Kim et al. (29), the mean identification rate was 83.1% with blue dye alone and 96.4% with dye and a tracer. There was no difference between tracer-only and dye-only (P=0.370), or tracer-only and dye-and-tracer (P=0.178). The lymph node upper axillary vein almost covers the high lymph nodes of axilla, which are rich in blood supply and of great significance. It has been confirmed (30,31) that blood vessels and tumor angiogenesis are generally associated with tumor growth and poor clinical outcome in cancer patients. Angiogenesis, the formation of neo-vessels, is one of the most important hallmarks of cancer. Therefore, the lymph

| Methylene blue tracer | Biopsy          | Total |
|-----------------------|-----------------|-------|
|                       | Metastasis      |       |
| Positive              | 7               | 7     |
| Negative              | 49              | 62    |
| Total                 | 56              | 69    |

Table 3: The diagnosis results that axillary lymph node metastasis upper axillary vein by methylene blue tracer
nodes’ upper axillary veins were taken as target areas in this study. Axillary lymph nodes of levels I, II, III are artificial groups; three levels are in the same station on transfer behavior, and three levels may be the first station for breast cancer metastatic lymph nodes. It is controversial to clean level III axillary lymph nodes. The lymph node upper axillary vein in our study, as well as axillary level III lymph nodes on the anatomy, is higher in the area of axilla. Some scholars believe that the higher area upper axillary vein belongs to the brachial plexus lymphatic drainage area. Lymphedema, dysfunction, bleeding risk and even destruction of the axillary vein scabbard are connected with cleaning the lymph node upper axillary vein. Other scholars argue that failing to clean the lymph node upper axillary vein may downstage inadequate treatment similar to level-III axillary lymph node (7,32). We used MB to guide the cleaning of the lymph node upper axillary vein. Because of the high position, MB color was required to move along the lymphatic longer, and MB required a longer time to color, which may have affected the coloration. Therefore, we should pay more attention when cleaning the high lymph node with MB tracer.

One of the main limitations of the current study was the use of MB. MB has some major disadvantages (33), such as allergic reactions, long time staining at the injection site and that proper use requires a certain training period. This method may be preferable for all clinicians if its accuracy is demonstrated to be comparable to that of the combination method. In addition, we analyze the clinical factors that affect the coloration of MB. BMI was found to be influential in the single-factor analysis, while no factors of influence were found in the multiple-factor analysis of our research. Proposed explanations for this are different measuring times and that we did not require patients to take off their shoes or remove clothing. These factors should be improved upon in subsequent studies.

Our study has shown that MB can be used in ALND and that it has great guiding value to novices of this surgery and primary hospitals. However, it is still prudent to use MB for high lymph node dissection. In breast cancer, MB has a great deal of advantages for SLNB. We believe that MB also has wide application prospects in ALND.

Conclusions

Among the clinical factors, MB coloration was worse more than 60 minutes before ALND. Using the technology of MB color, lymph nodes can be clearly identified. It has great guiding value for the doctors who learn the operation initially. However, it is still prudent to use MB for high lymph node dissection.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/tcr.2019.10.42). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). All patients were informed about the study objectives, and they were assured of confidentiality. They were asked to indicate their agreement and understanding with a signed informed consent form before the investigation. The study was approved by the ethics research committee of Kunming Medical University, China (Reference number: 2016-026-KJ-QT).

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