THE EFFECT OF ANGIOTENSIN II RECEPTOR BLOCKER VALSARTAN ON THE LEVEL OF CIRCULATING ENDOTHELIAL PROGENITOR CELLS IN DIABETIC PATIENTS WITH CHRONIC HEART FAILURE

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Key words: coronary heart disease; diabetes; heart failure; endothelial progenitor cells; valsartan

Reduction of circulating endothelial progenitor cells (EPCs) are considered as strong and robust biomarkers for prediction of cardiovascular outcomes in both chronic heart failure and diabetic populations. The aim of this study was to assess the effect of angiotensin II receptor blocker valsartan on the circulating level of endothelial progenitor cells in patients with ischemic chronic heart failure and type two diabetes mellitus (T2DM). The study population was structured retrospectively after determining coronary artery disease by multispiral computed tomography / contrast enhanced angiography in 126 subjects with chronic heart failure. Blood samples were taken at baseline before valsartan given and in 52 weeks after study entry. Endothelial progenitor cell populations were phenotyped by flow cytofluorimetry by means of monoclonal antibodies labeled with fluorochromes. Proangiogenic circulating EPCs were identified as CD45\textsuperscript{+}CD34\textsuperscript{+}CD309\textsuperscript{+} (VEGFR2), and Tie-2 antigens were also determined to identify subpopulations of EPCs coexpressing CD14 antigen. All subjects were distributed into two cohorts depending on daily doses of valsartan. Low (80-160 mg daily orally) and high doses (240-320 mg daily orally) of valsartan were used, and they were adjusted depending on achievement of the blood pressure level less than 140/80 mm Hg. The finding may suggest that the positive effect of valsartan on the endothelial function might be implemented by involving EPCs in reparative processes of vasculature in subjects with ischemic chronic heart failure with T2DM and thereby to improve the long-term prognosis in this patient population. The change from baseline in CD34\textsuperscript{+} subset of EPCs (frequencies and absolute values) was not significantly different between treatment cohorts. There was a significant increase of the circulating level of CD14\textsuperscript{+}CD309\textsuperscript{+} cells in two patient cohorts. But more prominent change of CD14\textsuperscript{+}CD309\textsuperscript{+} cells was observed in subjects who were given valsartan in high daily doses when compared with persons who were included into the cohort with low daily doses of the drug (P<0.05). Therefore, both frequencies and absolute values in CD14\textsuperscript{+}CD309\textsuperscript{+}Tie2\textsuperscript{+} were increased significantly in patients who were treated with high doses of valsartan only. A positive influence of angiotensin II receptor blocker valsartan in escalation doses on bone marrow-derived EPCs phenotyped as CD45\textsuperscript{+}CD309\textsuperscript{+} and CD14\textsuperscript{+}CD309\textsuperscript{+}Tie2\textsuperscript{+} in patients with ischemic chronic heart failure and T2DM has been found.

Type two diabetes mellitus (T2DM) is the most common cause of early manifestation of atherosclerosis and chronic heart failure that leads to increased cardiovascular morbidity and mortality [10]. It has been suggested that progression of atherosclerosis and myocardial dysfunction in T2DM patients may be related to endothelial dysfunction, that, probably, is mediated by suppression of the bone marrow-derived endothelial progenitor cells (EPCs) level in circulation, as well as decrease of the functional activity of EPCs [4, 15]. It has been previously found that EPC with the hematopoietic origin and the phenotype of CD34\textsuperscript{+}CD45\textsuperscript{+} are actively involved in angiogenesis, neo-vascularization, and tissue repair [13, 15]. Decrease in the population of circulating CD34\textsuperscript{+}CD45\textsuperscript{+} cells in patients with coronary heart disease (CHD), peripheral artery disease and dyslipidemia is associated with both severity of atherosclerotic lesions and severity of the arterial endothelial dysfunction [8, 12, 19]. However, circulating EPCs are considered as strong and robust biomarkers for prediction of cardiovascular outcomes in some populations, such as CAD and T2DM [4]. However, the most reliable prognostic indicators were not CD34\textsuperscript{+} EPA (CD45\textsuperscript{+}CD34\textsuperscript{+} and CD45\textsuperscript{+}CD45\textsuperscript{+}), but EPA with potentially proangiogenic qualities with phenotype CD14\textsuperscript{+}CD309\textsuperscript{+} and CD14\textsuperscript{+}CD309\textsuperscript{+}Tie2\textsuperscript{+} [1]. Thus, proangiogenic EPA can be considered as early markers of the beneficial effects of different modes of therapy in relation to short-term and long-term prognosis.

The aim of this study was to assess the effect of angiotensin II receptor blocker valsartan on the circulating level of endothelial progenitor cells in patients with ischemic chronic heart failure in combination with type two diabetes mellitus (T2DM).

Materials and Methods

The study population was structured retrospectively after determining coronary artery disease by multispiral computed tomography / contrast enhanced angiography or X-ray contrast angiography in 126 subjects with CHF (I-II functional class, NYHA classification). All patients gave voluntary written informed consent to participate in the study. Exclusion criteria were left ventricular ejection fraction

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(LVEF) ≤ 40%; severe kidney and liver diseases, which may affect the clinical outcome; unstable angina, myocardial infarction, or intracranial hemorrhage within 30 days prior to the study; the level of plasma creatinine above 440 mmol/l; the glomerular filtration rate (GFR) < 35 mL/min/m²; the body mass index above 30 kg/m²; hemodynamically significant tachyarrhythmias; valvar heart disease; hyperthyroidism; all cardiovascular ischemic events occurred during three months preceding the beginning of the research; cancer; cardiac pacemakers; pregnancy.

**Contrast-Enhanced Spiral Computed Tomography Angiography**

In order to verify the nature of a coronary disease patients underwent multispiral computed tomography-angiography and/or angiography. The CHD diagnosis was considered to be determined with the presence of angiographic studies performed previously and not older than 6 months in the absence of new cardiovascular events during this period and availability of results of the procedure for quantitative analysis. The coronary artery wall structure, as well as geometric and compositional parameters of atheromas were measured by contrast spiral CT on a “Somatom Volume Zoom” scanner (Siemens, Erlangen, Germany) with 2 rows of detectors during breath holding at the end of exhalation. After a preliminary scanning introduction of non-ionic contrast “Omnipak” (Amersham Health, Ireland) used to obtain optimal images of the coronary arteries was performed. For the reconstruction of images axial tomographic slices with the width of 0.6 mm were used. Coronary artery calcification was quantified by calculating the Agatston index [3]. Besides, the presence of calcified atheromas, but not calcified atherosclerotic plaques of high and low density, respectively, was determined according to the traditional methods [2, 7].

**Echocardiography Examination**

According to recommendations of the American Society of Echocardiography, standard transthoracic echocardiography in B-mode was performed on an ACUSON S30 scanner (Siemens, Erlangen, Germany) using a transducer with a frequency of 2.5-5 MHz. End-diastolic and end-systolic LV volumes were obtained using a two-dimensional reference sector according to the Simpson method, and the LV ejection fraction (LVEF) was calculated according to the conventional methods [20].

**Evaluation of the glomerular filtration rate**

Evaluation of the glomerular filtration rate (eGFR) was calculated using the MDRD formula [15].

**Calculation of the cardiovascular risk value**

Calculation of the 10-year cardiovascular risk in the patients studied was performed according to the Framingham scale of the total cardiovascular risk (2008) using the on-line calculator.

**Measurement of circulating levels of biological markers**

Blood samples were taken at baseline, prior to application of valsartan and 52 weeks after the start of the study. All serum samples were placed in tubes containing EDTA for subsequent analysis by flow cytometry. After collection of the blood for analysis all samples were placed in chilled test tubes and immediately centrifuged at the rotation speed of 6000 rpm. After centrifugation the serum was encoded and stored in a refrigerator at -70°C until use.

The level of high-sensitivity C-reactive protein (hs-CRP) was measured using a “AU640” nephelometric technique analyzer (Olympus Diagnostic Systems Group, Japan). The content of uric acid in the blood serum was determined by the enzymatic method. The concentrations of total cholesterol (TC) and high density lipoprotein (HDL) cholesterol were determined by an “AU640” analyzer (Olympus Diagnostic Systems Group, Japan). Low-density lipoproteins (LDL) were calculated using the Friedewald formula [11]. Microalbuminuria was verified as the loss of albumin in the range of 30-300 mg/day.

Phenotyping of EPC populations was carried out by the flow cytometry using monoclonal antibodies labeled with fluorochromes, FITC (fluorescein isothiocyanate) or double-labeled FITC/PE (phycoerythrin) (BD Biosciences, USA), antigens CD45, CD34, CD14, Tie-2 and SD309 (VEGFR2) according to the HD-FACS methodology (High-Definition Fluorescence Activated Cell Sorter) with mandatory removal of erythrocytes with the lysing buffer according to the protocol of ISHAGE gating [21]. For each sample 500000 events were analyzed. Circulating EPC were defined as CD45+CD34+. To identify subpopulations of EPC co-expressing antigen CD14, antigens SD309 (VEGFR2) and Tie-2 were determined additionally. The results of scattergrams obtained in the longitudinal and transverse dispersal of the laser beam in the flow cytometer were analyzed using the Boolean principles for double or triple positive events. The total number of identified cells was standardized with respect to the concentration of circulating CD45+ mononuclear cells.

Researchers strictly adhered to all the requirements for clinical trials according to the Helsinki Declaration of Human Rights (1964), the Conference on Harmonization Good Clinical Practice (GCP-ICH), Council of Europe Convention on protection of human rights and dignity in relation to the application of achievements of biology and medicine, the Convention on human rights and bio-medicine, including the Additional Protocol to the Convention on biomedical research and legislation of Ukraine.

**Statistical Analysis**

All the statistical analyses were performed in SPSS for Windows v. 20.0 (SPSS Inc., Chicago, IL, USA,
receptor blocker valsartan, β-blocker bisoprolol in target doses (10 mg/day), aspirin or other antiplatelet agents, a loop diuretic, statins and metformin, if necessary. Among the patients selected there were no persons requiring additional prescription of insulin or other antidiabetic medicines, except metformin.

The patients were divided into two groups depending on the daily dose of valsartan. Relatively low (80-160 mg per day) and high (240-320 mg daily) doses of valsartan were adjusted according to the achievement of the blood pressure level less than 140/80 mm Hg as 63.0% and 67.5% of the patients in both groups had a mild hypertension. Furthermore, microalbuminuria was detected in 37.0% and 32.5% of the patients, respectively. Both groups under study were matched in the frequency of cardiovascular risk factors, the body mass index, systolic and diastolic blood pressure, left ventricular ejection fraction, the heart rate, serum creatinine, glucose, glycated hemoglobin (HbA1c), uric acid and GFR. Increase in the concentration of hs-CRP, as well as decrease of LDL cholesterol levels were found in the patients with chronic heart failure and type 2 diabetes receiving high doses of valsartan compared to the patients who received valsartan in lower doses. Medians of the 10-year total cardiovascular risk were 22% and 24% for both groups of the patients with low and high daily dose of valsartan, respectively. The number of coronary arteries with the atheromas documented, as well as characteristics of the latter were comparable for both groups of the patients. There were no significant differences between the groups of the patients with chronic heart failure and type 2 diabetes in relation to the dynamics of blood pressure. In all patients with arterial hypertension in both groups the target blood pressure (less than 140/80 mm Hg) was achieved. Diuretics were added to the treatment regimen in 10.9% and 10.0% of cases, respectively. The mean systolic and diastolic blood pressure in both groups of patients in week 52 after the start of the study was 132 ± 4/73 ± 3 mmHg and 134 ± 4/75 ± 4 mmHg (p = 0.66).

Analysis of the results showed no significant changes in the content of the EPC with the phenotypes of CD45+CD34+, and CD45 CD34+, CD14+CD309+ and CD14+CD309+Tie2- between groups of the patients at baseline prior to the prescription of valsartan (Table 2). The nature of the effect of treatment with valsartan in different daily doses on the level of circulating EPC is shown in Table 3. As seen from the Table, the change in the total number of CD34+ of EPC in two groups of the patients during the observation period has no statistically significant character (p > 0.2 for all cases). In contrast, there was a significant increase in the level of circulating CD14+CD309+ EPC by the end of week 52 of the therapy in two groups of patients. However, a more pronounced dynamics was observed in patients receiving valsartan in high daily doses (p < 0.05). Thus, the content of valsartan provided a distinct dose-dependent effect on circulating levels of pro-angiogenic EPC with the phenotypes of CD14+CD309+ and CD14+CD309+Tie2-, while CD45+ pool of EPC did not undergo significant changes within the whole observation period.

Results and Discussion

It has been determined that EPC derived from the bone marrow and less often from peripheral tissues maintain the integrity of the vascular endothelium, as well as participate in vascular repair, angiogenesis and revascularization [22]. The number of circulating EPC increases in response to ischaemia and usually decreases with asymptomatic atherosclerosis, stable angina, type 2 diabetes, dyslipidemia, obesity [5, 8, 17, 18]. According to the findings of the experimental studies, EPC with the phenotypes of CD14+CD309+ and
Table 1

| Parameters                        | T2DM patients with CHF                             | P value |
|-----------------------------------|---------------------------------------------------|---------|
|                                   | All patients (n=86)                  | Cohort with a low daily dose of valsartan (n=46) | Cohort with a high daily dose of valsartan (n=40) |
| Age, years                        | 58.3±9.60                            | 57.98±8.12                                | 59.32±7.55                           | 0.52   |
| Male, n (%)                       | 54 (62.8%)                           | 28 (60.9%)                                | 26 (65.0%)                            | 0.58   |
| Framingham General Cardiovascular Risk, % | 23 (16-27)         | 22 (17 – 25)                              | 24 (16 – 30)                         | 0.001  |
| Arterial hypertension, n (%)      | 56 (65.1%)                           | 29 (63.0%)                                | 27 (67.5%)                            | 0.48   |
| Dyslipidemia, n (%)               | 44 (51.2%)                           | 24 (52.2%)                                | 20 (50.0%)                            | 0.58   |
| Microalbuminuria, n (%)           | 30 (34.9%)                           | 17 (37.0%)                                | 13 (32.5%)                            | 0.52   |
| Premature CAD, n (%)              | 9 (10.5%)                            | 4 (8.7%)                                  | 5 (12.5%)                             | 0.048  |
| Smoking, n (%)                    | 16 (18.6%)                           | 10 (21.7%)                                | 6 (15.0%)                             | 0.042  |
| Body mass index, kg/m²            | 24.1 (95% CI = 21.6-26.1)            | 22.5 (95% CI = 20.3-26.0)                 | 25.2 (95% CI = 20.6-27.2)            | 0.055  |
| eGFR, ml/min/m²                   | 82.3 (95% CI = 68.7-102.6)           | 85.4 (95% CI = 70.1-98.2)                 | 82.1 (95% CI = 67.5-101.3)           | 0.36   |
| HbA1c, %                          | 6.8 (95% CI = 4.1-9.5)               | 6.69 (95% CI = 4.8-9.32)                  | 6.85 (95% CI = 4.2-9.5)              | 0.061  |
| Fasting glucose, mmol/L           | 5.20 (95% CI = 3.3-9.7)              | 5.16 (95% CI = 3.7-8.8)                   | 5.24 (95% CI = 3.2-9.6)              | 0.057  |
| Creatinine, μmol/L                | 72.3 (95% CI = 58.7-92.6)            | 75.7 (95% CI = 57.1-90.4)                 | 71.2 (95% CI = 56.4-98.9)            | 0.49   |
| SUA, mmol/L                       | 23.8 (95% CI = 15.8-31.3)            | 23.4 (95% CI = 15.6-30.1)                 | 23.9 (95% CI = 16.9-31.6)            | 0.48   |
| hs-CRP, mg/L                      | 4.95 (95% CI = 3.1-5.9)              | 4.87 (95% CI = 3.6-6.1)                   | 5.22 (95% CI = 3.1-9.9)              | 0.046  |
| TC, mmol/L                        | 5.1 (95% CI = 3.9-6.1)               | 5.0 (95% CI = 3.7-6.0)                    | 5.3 (95% CI = 3.8-5.8)               | 0.12   |
| LDL cholesterol, mmol/L           | 3.23 (95% CI = 3.1-4.4)              | 3.31 (95% CI = 3.0-4.2)                   | 3.23 (95% CI = 3.1-4.4)              | 0.047  |
| HDL cholesterol, mmol/L           | 0.91 (95% CI = 0.89-1.12)            | 0.93 (95% CI = 0.90-1.10)                 | 0.90 (95% CI = 0.87-1.12)            | 0.18   |
| Mean systolic BP, mm Hg           | 144±8                                | 144±5                                    | 146±7                                | 0.44   |
| Mean diastolic BP, mm Hg          | 84±5                                 | 85±4                                     | 83±6                                 | 0.56   |
| Heart rate, beat per min          | 71±3                                 | 72±4                                     | 69±4                                 | 0.52   |
| LV EF, %                          | 48.10±0.76                           | 48.50±0.58                               | 48.80±0.34                           | 0.64   |
| CAP, n                            | 56 (95% CI = 31-76)                  | 52 (95% CI = 46-78)                      | 59 (95% CI = 42-79)                  | 0.44   |
| HD-NCP, n                         | 21 (95% CI = 11-45)                  | 19 (95% CI = 13-39)                      | 24 (95% CI = 11-48)                  | 0.63   |
| LD-NCP, n                         | 15 (95% CI = 13-21)                  | 16 (95% CI = 13-20)                      | 13 (95% CI = 11-17)                  | 0.38   |
| Agatston score index              | 586 (95% CI = 401-838)               | 579 (95% CI = 466-820)                   | 592 (95% CI = 412-844)               | 0.60   |
| Coronary arteries with plaques determined |                         |                                          |                                      |        |
| Single-vessel disease, n (%)      | 26 (30.3%)                           | 13 (28.3%)                               | 13 (32.5%)                           | 0.058  |
| Two-vessel disease, n (%)         | 22 (25.6%)                           | 12 (26.1%)                               | 10 (25.0%)                           | 0.62   |
| Multivessel disease, n (%)        | 38 (44.2%)                           | 21 (45.6%)                               | 17 (42.5%)                           | 0.54   |
| Aspirin, n (%)                    | 81 (94.1%)                           | 43 (93.5%)                               | 38 (95.0%)                           | 0.38   |
| Other antiagregants, n (%)        | 5 (5.8%)                             | 3 (6.5%)                                 | 2 (5.0%)                             | 0.36   |
| Statins, n (%)                    | 56 (65.1%)                           | 29 (63.0%)                               | 27 (67.5%)                           | 0.48   |
| Metformin, n (%)                  | 44 (51.2%)                           | 24 (52.2%)                               | 20 (50.0%)                           | 0.58   |
| Thiazide-like diuretic, n (%)     | 9 (10.5%)                            | 5 (10.9%)                                | 4 (10.0%)                            | 0.82   |

Note: CI = confidence interval, T2DM = type 2 diabetes mellitus, eGFR = estimated glomerular filtration ratio, TC = total cholesterol, HbA1c = glycated hemoglobin, LDL = low-density cholesterol, HDL = high-density cholesterol, SUA = serum uric acid, BP = blood pressure, hs-CRP = high sensitive C-reactive protein, LV EF = left ventricular ejection fraction, HD-NCP = high-density noncalcified atherosclerotic plaque, LD-NCP = low-density noncalcified atherosclerotic plaque, CAP = calcified atherosclerotic plaques.

CD14+CD309+ Tie2+ contribute to the restoration of the endothelium of the arteries, and it is the necessary condition for repair of blood vessels [12]. It has been found that reduction in CD14+CD309+ and CD14+CD309+ Tie2+ of EPC is closely associated with cardiovascular risk factors and has a predictive value for occurrence of adverse cardiovascular outcomes [4, 12]. The present study has tested the hypothesis that in patients with ischemic CHF in combination with the accompanying type 2 diabetes the level of recovery of circulating EPC can be achieved after prolonged use of angiotensin II recep-
Frequencies and absolute values of circulating mononuclear cells in the study patient population

| Parameters                        | T2DM patients with the known CAD                                                                 |
|----------------------------------|--------------------------------------------------------------------------------------------------|
|                                  | All patients (n = 86)                                                                                |
|                                  | Cohort with a low daily dose of valsartan (n=46)                                                    |
|                                  | Cohort with a high daily dose of valsartan (n=40)                                                   |
|                                  | P value                                                                                           |
| CD45^+CD34^+%, %                 | 2.19 (IQR = 1.76-2.613)                                                                            |
| CD45^+CD34^+, cells×10^3/µL      | 0.007 (IQR = 0.003-0.006)                                                                           |
| CD45^+CD34^+, cells×10^4/µL      | 0.507 (IQR = 0.005-0.007)                                                                           |
| CD14^+CD309^+×10^4, %            | 57.00 (IQR = 43.20-81.50)                                                                           |
| CD14^+CD309^+, cells×10^4/µL     | 1.09 (IQR = 1.00-1.348)                                                                            |
| CD14^+CD309^+Tie2^+×10^4, %      | 5.50 (IQR = 3.05-8.15)                                                                             |
| CD14^+CD309^+Tie2^+, cells×10^4/µL | 0.270 (IQR = 0.241-0.411)                                                                          |

Note: The values correspond to medians and a interquartile range (IQR) of (25-75)%. Statistical comparisons are made using Mann-Whitney test with significance levels of 0.05 and 0.01 (for 2-tailed).

Follow-up frequencies and absolute values of circulating mononuclear cells in treatment cohorts

| Parameters                        | Cohort with a low daily dose of valsartan (n=46)                                                    |
|----------------------------------|--------------------------------------------------------------------------------------------------|
|                                  | Cohort with a high daily dose of valsartan (n=40)                                                   |
| CD45^+CD34^+ (%) at baseline     | 2.14 (IQR = 1.79-2.50)                                                                            |
| CD45^+CD34^+ (%) follow-up       | 2.12 (IQR = 1.82-2.54)                                                                            |
| CD45^+CD34^+ (cells×10^4/µL) at baseline | 0.114 (IQR = 0.095-0.120)                                                                           |
| CD45^+CD34^+ (cells×10^4/µL) follow up | 0.118 (IQR = 0.098-0.128)                                                                          |
| CD45^+CD34^+×10^4 (%) at baseline | 1.07 (IQR = 0.99-1.35)                                                                            |
| CD45^+CD34^+×10^4 (%) follow-up  | 1.06 (IQR = 0.97-1.36)                                                                            |
| CD45^+CD34^+ (cells×10^4/µL) at baseline | 0.055 (IQR = 0.052-0.067)                                                                           |
| CD45^+CD34^+ (cells×10^4/µL) follow up | 0.059 (IQR = 0.055-0.066)                                                                          |
| CD14^+CD309^+×10^4 (%) at baseline | 57.00 (IQR = 41.50-76.90)                                                                            |
| CD14^+CD309^+×10^4 (%) follow up  | 73.00 (IQR = 58.20-84.50)*                                                                         |
| CD14^+CD309^+ (cells×10^4/µL) at baseline | 2.96 (IQR = 2.30-4.24)                                                                            |
| CD14^+CD309^+ (cells×10^4/µL) follow up | 4.92 (IQR = 3.20-6.71)*                                                                           |
| CD14^+CD309^+Tie2^+×10^4 (%) at baseline | 5.53 (IQR = 3.20-6.71)*                                                                           |
| CD14^+CD309^+Tie2^+×10^4 (%) follow up  | 5.66 (IQR = 3.35-8.80)                                                                            |
| CD14^+CD309^+Tie2^+ (cells×10^4/µL) at baseline | 0.265 (IQR = 0.243-0.410)                                                                          |
| CD14^+CD309^+Tie2^+ (cells×10^4/µL) follow up | 0.321 (IQR = 0.265-0.476)                                                                          |

Note: The values correspond to medians and a interquartile range (IQR) of (25-75)%. Statistical comparisons are made for each cohort between the values at baseline and follow-up. * – statistically significant changes (P<0.05).
creasing the level of potentially proangiogenic EPCs using valsartan, as well as its ability to provide a dose-dependent effect on circulating levels of the latter may have a serious clinical significance. Probably, it is necessary to conduct a new research in this direction aiming at detailing the nature of the relationship between the use of valsartan and the functional activity of EPCs, as well as the ability of the drug to improve clinical outcomes in a population of patients with heart failure and type 2 diabetes mellitus.

CONCLUSIONS

1. In the cohort of patients with heart failure and type 2 diabetes mellitus valsartan has no significant influence on the content of endothelial progenitor cells with CD45\(^+\)CD34\(^+\) and CD45\(^-\)CD34\(^+\) phenotypes.

2. Valsartan has the dose-dependent ability to increase the circulating levels of potential proangiogenic endothelial progenitor cells with the phenotypes of CD14\(^+\)CD309\(^+\) and CD14\(^+\)CD309\(^+\)Tie2.

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52 тяжелые. Змена загальны й кількість CD34+ ЕКП у двох групах пацієнтів протягом періоду спостереження не носила статистично значущого характеру. Спостерігалася суттєве підвищення рівня циркулюючих CD14+CD309+ ЕКП до кінця 52 тижня терапії у двох групах хворих. Однак більш виражена динаміка відзначалась у пацієнтів, які отримували валсартан у високих дозових дозах. Таким чином, яміс валсартану чинить виразний дозозалежний ефект щодо циркулюючого рівня проангіогенних ЕКП з фенотипами CD14+CD309+ і CD14+CD309+Tie2+, тоді як пул CD34+ ЕКП не зазнав істотних змін протягом всего періоду спостереження. Отже, блокатор рецепторів ангіотензину II валсартан чинить дозозалежний позитивний вплив на кількість циркулюючих ендотеліальних прогеніторних клітин кістковомозкового походження, фенотипованих як CD14+CD309+ і CD14+CD309+Tie2+ у хворих з ХСН ішемічного генезу в поєднанні з ІД 2 типу.

ВЛИЯНИЕ БЛОКАТОРА РЕЦЕПТОРОВ АНГИОТЕНЗИНА II ВАЛСАРТАНА НА УРОВЕНЬ ЦИРКУЛИРУЮЩИХ ЭНДОТЕЛИАЛЬНЫХ ПРОГЕНИТОРНЫХ КЛЕТОК У ПАЦИЕНТОВ С ХРОНИЧЕСКОЙ СЕРДЕЧНОЙ НЕДОСТАТОЧНОСТЬЮ ИШЕМИЧЕСКОГО ГЕНЕЗА В СОЧЕТАННИИ С САХАРНЫМ ДИАБЕТОМ 2 ТИПА

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Ключевые слова: ишемическая болезнь сердца; сердечная недостаточность; сахарный диабет 2 типа; циркулирующие эндотелиальные прогениторные клетки; валсартан

Снижение циркулирующих эндотелиальных клеток-предшественников (ЭКП) рассматривается в качестве прогностического биомаркера сердечно-сосудистых исходов у пациентов с хронической сердечной недостаточностью и сахарным диабетом 2 типа. Целью данного исследования была оценка эффекта блокатора рецепторов ангиотензина II валсартана в отношении циркулирующего уровня ЭКП у больных с хронической сердечной недостаточностью ишемического генеза в сочетании с сахарным диабетом 2 типа. После подтверждения ИБС с помощью мультиспектральной компьютерной томографии/ангиографии с контрастиным усилением или рентгеноконтрастной коронарографии в исследование было ретроспективно включено в 126 пациентов с ХСН в сочетании с сахарным диабетом 2 типа. Все пациенты были распределены на две когорты в зависимости от суточной дозы валсартана. Были использованы низкие (80-160 мг в день) и высокие (240-320 мг в день) дозы валсартана, скорректированные в зависимости от достижения уровня АД менее 140/80 мм рт. Период наблюдения составил 52 недели. Изменение общего количества CD34+ ЕКП в двух группах пациентов на протяжении периода наблюдения не носило статистически значимого характера. Изменило место существенное повышение уровня циркулирующих CD14+CD309+ ЕКП к концу 52 недели терапии в двух группах больных. Однако более выраженная динамика отмечалась у пациентов, получающих валсартан в высоких суточных дозах. Таким образом, валсартан оказывал отчётливый дозозависимый эффект в отношении циркулирующего уровня проангиогенных ЭКП с фенотипами CD14+CD309+ і CD14+CD309+Tie2+, тоді як пул CD34+ ЕКП не претерпевал существенных изменений на протяжении всего периода наблюдения. Таким образом, блокатор рецепторов ангиотензин-II валсартан оказывает дозозависимое позитивное влияние на количество циркулирующих эндотелиальных прогениторных клеток костномозкового происхождения, фенотипованных как CD14+CD309+ і CD14+CD309+Tie2+ у больных с ХСН ішемичного генезу в сочетании с СД 2 типа.

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