Pollen morphology of the African *Sclerosperma* (Arecaceae)

FRIDGEIR GRÍMSSON 1, JOHAN L.C.H. VAN VALKENBURG 2, JAN J. WIERINGA 2, ALEXANDROS XAFIS 1, BONNIE F. JACOBS 3 & REINHARD ZETTER 1

1Department of Palaeontology, University of Vienna, Vienna, Austria, 2Naturalis Biodiversity Center, National Herbarium of The Netherlands, Leiden, The Netherlands, 3Roy M. Huffington Department of Earth Sciences, Southern Methodist University, Dallas, TX, USA

**Abstract**

Three currently accepted *Sclerosperma* species appear to produce four different pollen morphologies. *Sclerosperma mannii* and *S. walkeri* pollen share the same distinct reticulate sculpture, but *S. profizianum* produces three different pollen types (microreticulate, fossulate, and perforate). The pollen morphology suggests that *S. mannii* and *S. walkeri* are sister taxa of the same intrageneric lineage. The pollen diversity observed in *S. profizianum* suggests (a) this taxon is unique regarding its pollen diversity despite being a non-heterostylous plant or (b) that circumscription of *S. profizianum* as a species may well be in the need of redefinition.

**Keywords:** light microscopy, palms, scanning electron microscopy, *Sclerosperma mannii*, *Sclerosperma profizianum*, *Sclerosperma walkeri*, swamp element, tropics

**Sclerosperma** G. Mann et H. Wendl. is a palm genus of only three species, *S. mannii* H. Wendl., *S. profizianum* Valk. et Sunderl. and *S. walkeri* A. Chev. (Arecoideae, Sclerospermeae; Dransfield et al. 2008), restricted to tropical central Africa, primarily in swampy habitats (Table I; Van Valkenburg et al. 2008). The first account of *Sclerosperma* pollen by Erdtman and Sing (1957) documented its unique morphology within the palm family (triangular, triporate, reticulate), features often discussed by M.M. Harley in the years 1991 to 2008 through her extensive work with colleagues on the pollen morphology of Arecaceae (Harley & Hall 1991; Harley 1996, 1999, 2004; Harley & Baker 2001; Harley & Dransfield 2003; Dransfield et al. 2008). Despite the number of publications containing pollen descriptions and micrographs of the extant genus, *Sclerosperma*, a detailed characterisation of the pollen morphology of the three species was needed. Also, the taxonomy of this genus was only recently revised (Van Valkenburg et al. 2008), and showed that previously published pollen material often originated in misidentified specimens.

Here we describe and illustrate pollen from each of the three currently accepted *Sclerosperma* species, compare their pollen, and highlight the diagnostic features that can be used to distinguish them from each other.

**Material and methods**

Flowers of *Sclerosperma* (see Table II) from the herbaria of the Botanic Garden Meise (BR), the Royal Botanic Gardens, Kew (K), and Naturalis (WAG; the National Herbarium of the Netherlands) were prepared following the protocol of Grímsson et al. (2017, 2018). Scanning electron microscopy (SEM) stubs with *Sclerosperma* pollen produced for this study are stored in the collection of the Department of Palaeontology, University of Vienna, Austria, under the accession numbers IPUW 7513/217 to IPUW 7513/222.

**Descriptive palynology**

The pollen terminology follows Punt et al. (2007; light microscopy [LM]) and Halbritter et al. (2018;
SEM). The classification and author names of extant species follow WCSF (2018). Classification above genus level follows Dransfield et al. (2008) and APG IV (2016). Herbarium materials were assigned to extant species according to Van Valkenburg et al. (2008). Pollen grains of each taxon are described individually. Pollen grains of the three *Sclerosperma* species are also compared with each other in Table III.

**Note regarding the following descriptions.** — According to Halbritter et al. (2018) an ulcus (pl. ulci) is a more or less circular aperture situated distally on the pollen. Ulci are confined to gymnosperms, magnoliid and monocot angiosperm taxa. Also, according to Halbritter et al. (2018) a porus (pl. pori) is a more or less circular aperture located at the equator or regularly spread over the pollen grain. Pori are confined to dicot angiosperm taxa. Still, in all the literature regarding *Sclerosperma* by M.M. Harley from the years 1991 to 2008 (Harley & Hall 1991; Harley 1996, 1999, 2004; Harley & Baker 2001; Harley & Dransfield 2003; Dransfield et al. 2008) the pollen of this genus was described as porate. In order to avoid confusion the apertures of *Sclerosperma* are here also termed pori and the pollen is regarded as triporate and not triloculate despite the distal position of the apertures.

**Family Areaceae Bercht. et J. Presl**

**Genus Sclerosperma G. Mann et H. Wendl.**

**Species Sclerosperma mannii H. Wendl.** (van der Burgt, 1958 [K])

(Figures 1A–E, 2; Table III)

**Description.** — Pollen, monad, heteropolar, polar axis/equatorial diameter (*P/E*) ratio oblate, outline straight-triangular to slightly concave-triangular in polar view, bean-shaped in equatorial view (convex distal face versus concave proximal face); equatorial diameter 32–38 µm in LM, 27–34 µm in SEM, polar axis 9–15 µm in LM; triporate, pori positioned subapically on the distal polar face, pori elliptic, 4.5–6.0 µm in diameter, pori equipped with opercula; exine 1.7–2.5 µm thick in LM, nexine thinner than sexine; pollen wall semitectate; sculture reticulate in LM, reticulate to perforate in SEM; distal face reticulate with broad muri and elliptic to triangular to polygonal lumina, 18–25 lumina per 100 µm² at central distal face, 0–6 nanogemmatae free-standing columellae per lumina (SEM); proximal face reticulate to perforate, lumina/perforations elliptic to triangular to polygonal, 0–6 nanogemmatae free-standing columellae per lumina; central polar areas and interapertural areas reticulate, sculpture becoming microreticulate to perforate towards apices; opercula with nanoverrucate to granulate sublayer and distinct microreticulate supra-layer (SEM).

**Remarks.** — The first LM and SEM micrographs showing pollen of this taxon are by Harley and Hall (1991, plate 4, figures 32 [SEM] and 33 [LM]). The same SEM micrograph is shown in Harley (1999, plate 1, figure 12), and again along with two additional LM micrographs and two attached grains under SEM in Harley (1996, plate 16, figures C and F [SEM], and G and H [LM]). These are all repeated in Harley and Baker (2001, figures 77 and 82 [SEM], 80 and 81 [LM]). A new SEM detail is provided in Harley and Dransfield (2003, figure 11). In total, five or six grains were illustrated using either LM or SEM micrographs, but no TEM micrograph has been presented thus far. All the pollen grains of this taxon illustrated by Harley (1996), Harley and Baker (2001) and Harley and Dransfield (2003) originate from the same herbarium sample (Tuley, s.n. [K]). All other previously published micrographs showing alleged pollen of *Sclerosperma mannii* originated from misidentified specimens (see Table IV). The LM and SEM based pollen morphology of the Tuley s.n. (K) sample is similar to that now observed in the van der Burgt 1958 (K) sample. The only difference is that the free-standing columellae are more prominent and more frequent in the specimen collected by Tuley versus that collected by van der Burgt, a feature comparable to what is observed in the *S. walkeri* pollen from the Leonard 1614 (BR) sample (compare figure 11 in Harley and Dransfield [2003], with Figures 2E and 6E this study).

**Species Sclerosperma profisionianum Valk. et Sunderl.**

(Figures 1F–Y, 3–5; Table III)

**Note.** — We encountered three pollen morphologies when analysing pollen grains from different herbarium specimens assigned to this taxon. The pollen morphologies are here described individually as Type A, B and C.

**Type A** (Gillet, 279a [WAG]; holotype)

(Figures 1F–J, 3; Table III)

**Description.** — Pollen, monad, heteropolar, *P/E* ratio oblate, outline straight-triangular to slightly concave-triangular in polar view, bean-shaped in equatorial view (convex distal face versus concave proximal face); equatorial diameter 35–40 µm in LM, 29–35 µm in SEM, polar axis 10–14 µm in LM; triporate, pori positioned sub-apically on the distal polar face, pori elliptic, 5.0–8.5 µm in diameter, pori equipped with opercula; exine 1.7–2.5 µm thick in LM, 2.1–2.5 µm thick in SEM, nexine thinner than sexine, nexine 0.5–0.8 µm thick...
Figure 1. LM micrographs of all *Sclerosperma* pollen types. **A–E.** *Sclerosperma mannii* (from Cameroon, coll. van der Burgt, 1958 [K]), same grain in polar and equatorial view. **F–J.** *Sclerosperma proflizianum* Type A (from DR Congo, coll. Gillet, 279a [WAG]), same grain in polar and equatorial view. **K–O.** *Sclerosperma proflizianum*, Type B (from Angola, coll. Grobbelaar, s.n. [K]), same grain in polar and equatorial view. **P–T.** *Sclerosperma proflizianum*, Type C (from Ghana, coll. Hall & Enti, GC 36150 [K]), same grain in polar and equatorial view. **U–Y.** *Sclerosperma proflizianum*, Type C (from R Congo, coll. Profizi, 841 [K]), same grain in polar and equatorial view. **Z–D’.** *Sclerosperma walkeri* (from DR Congo, coll. Leonard, 1614 [BR]), same grain in polar and equatorial view. **A’, B’, G, I, L, N, Q, S, V, X, A’, C’.** Polar view, optical cross section. **E, J, O, T, Y, D’.** Equatorial view, upper in high focus and lower in optical cross-section. Scale bars – 10 µm.
Figure 2. SEM micrographs of *Sclerosperma mannii* (from Cameroon, coll. van der Burgt, 1958 [K]). A, C, E, G. Pollen in polar view, distal side. B, D, F, H. Pollen in polar view, proximal side. C, D. Close-ups of apices (aperture on distal side). E, F. Close-ups of central polar areas (reticulum narrower on proximal side). G, H. Close-ups of interapertural areas. Scale bars – 10 µm (A–B), 1 µm (C–H).
Figure 3. SEM micrographs of *Sclerosperma profizianum*, Type A (from DR Congo, coll. Gillet, 279a [WAG]). A, C, E, G. Pollen in polar view, distal side. B, D, F, H. Pollen in polar view, proximal side. C, D. Close-ups of apices (aperture on distal side). E, F. Close-ups of central polar areas (reticulum narrower on proximal side). G, H. Close-ups of interapertural areas. Scale bars – 10 µm (A–B), 1 µm (C–H).
Figure 4. SEM micrographs of *Sclerosperma profizianum*, Type B (from Angola, coll. Grobbelaar, s.n. [K]). A, C, E, G. Pollen in polar view, distal side. B, D, F, H. Pollen in polar view, proximal side. C, D. Close-ups of apices (aperture on distal side). E, F. Close-ups of central polar areas. G, H. Close-ups of interapertural areas. Scale bars – 10 µm (A–B), 1 µm (C–H).
Figure 5. SEM micrographs of *Sclerosperma prozizianum*, Type C (from Ghana, coll. Hall & Enti, GC 36150 [K], [A, C, E, G]; from R Congo, coll. Profizi, 841c2 [K], [B, D, F, H]). A, C, E, G. Pollen in polar view, distal side. B, D, F, H. Pollen in polar view, proximal side. C, D. Close-ups of apices (aperture on distal side). E, F. Close-ups of central polar areas. G, H. Close-ups of interapertural areas. Scale bars – 10 µm (A–B), 1 µm (C–H).
Figure 6. SEM micrographs of *Sclerosperma walkeri* (from DR Congo, coll. Leonard, 1614 [BR]). **A, C, E, G.** Pollen in polar view, distal side. **B, D, F, H.** Pollen in polar view, proximal side. **C, D.** Close-ups of apices (aperture on distal side). **E, F.** Close-ups of central polar areas (reticulum narrower on proximal side). **G, H.** Close-ups of interapertural areas. Scale bars – 10 µm (A–B), 1 µm (C–H).
in SEM, sexine 1.2–1.8 µm thick in SEM; pollen wall semitectate; sculpture reticulate in LM, microreticulate to perforate in SEM; distal face microreticulate with broad muri and elliptic to triangular to polygonal lumina, 30–35 lumina per 100 µm² at central distal face (SEM); proximal face microreticulate to perforate, lumina/perforations elliptic to circular or slit-like; central polar areas and interapertural areas microreticulate, sculpture becoming nanoreticulate to perforate towards apices; opercula with nanoverrucate to granulate sublayer and a distinct reticulate supra-layer (SEM).

Remarks. — The first LM, SEM and transmission electron microscopy (TEM) micrographs showing pollen of this taxon are by Harley from the isotype (Gillett, 279a [K]) of the Kew herbarium (Harley & Dransfield 2003; figures 10, 16 and 17 [SEM], 13 and 14 [LM], and 19 [TEM]). The same SEM micrograph is shown in Harley (2004, figure 6F), and an additional SEM micrograph is provided in Dransfield et al. (2008, p. 391, figure b). All previously illustrated pollen grains that are here referred to S. prozianum (pollen Type A) were formerly assigned to S. mannii (see Table IV).

Type B (Grobblea, s.n. [K])
(Figures 1K–O, 4; Table III)

Description. — Pollen, monad, heteropolar, PE ratio oblate, outline straight-triangular to slightly concave-triangular in polar view, bean-shaped in equatorial view (convex distal face versus concave proximal face); equatorial diameter 32–38 µm in LM, 30–34 µm in SEM, polar axis 10–12.5 µm in LM; triporate, pori positioned sub-apically on the distal polar face, pori circular to elliptic, 4.0–5.5 µm in diameter, pori equipped with opercula; exine 1.7–2.5 µm thick in LM, nexine thinner than sexine; pollen wall tectate; sculpture rugulate in LM, fossulate, rugulate/verrucate and perforate in SEM; distal face fossulate with tiny circular to slit-like perforations aligned within the fossulae, sinuous fossulae outlining irregular shaped rugulae/verrucae (SEM); proximal face fossulate with tiny circular to slit-like perforations aligned within the fossulae, sinuous fossulae outlining irregular shaped rugulae/verrucae, sculpture becoming microrugulate to nanorugulate/verruculate and perforate towards apices; opercula with nanoverrucate to granulate sublayer and perforate supra-layer (SEM).

Remarks. — Pollen of this type was originally noted as Sclerosperma gilletii by Harley and Dransfield (2003, figure 12 [SEM]) and Harley (2004, figures 6C [SEM], 6D and 6E [LM]). The same micrographs were provided in Dransfield et al. (2008, p. 391, figures a [SEM], c and d [LM]), but then affiliated to S. prozianum following description of that new species in the publication of Van Valkenburg et al. (2008). It is interesting that our study of the same herbarium material (Profizi, 841 [K]), but different flower, did not give the same pollen Type B as that figured by Harley and Dransfield (2003), Harley (2004) and Dransfield et al. (2008), but pollen Type C (Table IV). This suggests that S. prozia-
|                      | *S. mannii* | *S. pnfzianum* (Type A) | *S. pnfzianum* (Type B) | *S. pnfzianum* (Type C) | *S. walkeri* |
|----------------------|-------------|--------------------------|--------------------------|--------------------------|-------------|
| Outline polar view   | Straight-triangular to slightly concave-triangular | straight-triangular to slightly concave-triangular | straight-triangular to slightly convex-triangular | straight-triangular to slightly concave-triangular | straight-triangular to slightly concave-triangular |
| Outline equatorial view | Bean-shaped | Bean-shaped | Bean-shaped | Bean-shaped | Bean-shaped |
| Equatorial diameter (µm in LM) | 32–38 | 35–40 | 32–38 | 37–42 | 35–40 |
| Equatorial diameter (µm in SEM) | 27–34 | 29–35 | 30–34 | 31–39 | 30–35 |
| Polar axis (µm in LM) | 9–15 | 10–14 | 10–13 | 10–16 | 15–19 |
| Aperture type position | Triporate | Triporate | Triporate | Triporate | Triporate |
| Aperture outline diameter (µm in SEM) | Elliptic | Circular to elliptic | 4.0–5.5 | Circular to elliptic | 4.0–6.5 |
| Exine thickness (µm in SEM) | 1.7–2.5 | 1.7–2.5 | 1.7–2.5 | 1.7–2.5 | 1.7–2.5 |
| Pollen wall SEM | Semitectate | Semitectate | Tectate | Tectate | Semitectate |
| Sculpture (LM) | Reticulate | Reticulate | Rugulate | Scabrate | Reticulate |
| Sculpture (SEM) | Reticulate to perforate | Microreticulate to perforate | Fossulate, rugulate/verrucate, perforate | Perforate, rugulate/verrucate and fossulate | Reticulate to perforate |
| Sculpture distal face (SEM) | Reticulate with broad muri and elliptic to triangular to polygonal lumina, 0–6 nanogemmæ free-standing columellæ per lumina | Microreticulate with broad muri and elliptic to triangular to polygonal lumina | Fossulate with tiny circular to slit-like perforations aligned within the fossulae, sinuous fossulae outlining irregular shaped rugulae/verrucae | Perforate, perforations elliptic to slit-like, perforations often aligned in sinuous rows, rows of perforations outlining irregular shaped rugulae/verrucae | Reticulate with broad muri and elliptic to triangular to polygonal lumina, 0–6 nanogemmæ free-standing columellæ per lumina |

(Continued)
Table III. (Continued).

|                  | S. mammii | S. proezianum (Type A) | S. proezianum (Type B) | S. proezianum (Type C) | S. walkeri |
|------------------|-----------|------------------------|------------------------|------------------------|------------|
| Number of lumina/ perforations at central distal face (SEM) | 18–25 per 100 µm² | 30–35 per 100 µm² | Not applicable | 45–55 per 100 µm² | 16–25 per 100 µm² |
| Sculpture proximal face (SEM) | **Reticulate** central polar area and mesosporium with elliptic to triangular to polygonal lumina, 0–6 nanogemmae free-standing columellae per lumina; becoming **micoreticular** to perforate towards apices | **Micoreticular** central polar area and mesosporium with elliptic to circular or slit-like lumina; becoming **nanoreticular** to perforate towards apices | **Fossulate** central polar area and mesosporium with tiny circular to slit-like perforations aligned within the fossulae, sinuous fossulae outlining irregular shaped rugulae/verrucae; becoming **microrugulate to nanorugulate/verru cate** and perforate towards apices | **Perforate** and fossulate central polar area and mesosporium, perforations elliptic to slit-like, perforations often aligned in sinuous rows, rows of perforations and fossulae outlining irregular shaped rugulae/verrucae; becoming **microrugulate to nanorugulate/verru cate** and perforate towards apices | **Reticulate** central polar area and mesosporium with elliptic to triangular to polygonal lumina, 0–6 nanogemmae free-standing columellae per lumina; becoming **micoreticular** to perforate towards apices |
| Opercula (SEM)   | Nanoverrucate to granulate sublayer and **micoreticular** supra-layer | Nanoverrucate to granulate sublayer and **reticulate** supra-layer | Nanoverrucate to granulate sublayer and **perforate** supra-layer | Nanoverrucate to granulate sublayer and **perforate** supra-layer | Nanoverrucate to granulate sublayer and **micoreticular** supra-layer |

Note: All measurements include only those from this study and are given in micrometres. Most diagnostic features appear in bold font.
Table IV. Affiliation of previously illustrated *Sclerosperma* pollen.

| Taxon (Type)            | Sample (herbarium) | Figured in                                | Noted as                        | Micrograph |
|-------------------------|--------------------|------------------------------------------|---------------------------------|------------|
| *Sclerosperma mannii*   | Tuley, s.n. (K)    | Harley and Dransfield (2003)             | *Sclerosperma mannii*           | Figure 11  |
|                         | Tuley, s.n. (K)    | Harley and Baker (2001)                  | *Sclerosperma mannii*           | Figures 77 |
|                         | Tuley, s.n. (K)    | Harley (1999)                            | *Sclerosperma mannii*           | Plates 1  |
|                         | Tuley, s.n. (K)    | Harley (1996)                            | *Sclerosperma mannii*           | Plate 16  |
|                         | Tuley, s.n. (K)    | Harley and Hall (1991)                   | *Sclerosperma mannii*           | Plate 4   |
| *Sclerosperma prozianum*| Gillett, 279a (K)  | Dransfield et al. (2008)                 | *Sclerosperma mannii*           | Page 391  |
|                         | Gillett, 279a (K)  | Harley (2004)                            | *Sclerosperma mannii*           | Figure 6F |
|                         | Gillett, 279a (K)  | Harley (2004)                            | *Sclerosperma prozianum*        | Page 391  |
| *Sclerosperma gilletii* | Profizi, 841 (K)   | Dransfield et al. (2008)                 | *Sclerosperma gilletii*         | Page 391  |
|                         | Profizi, 841 (K)   | Harley (2004)                            | *Sclerosperma gilletii*         | Figure 6C |
|                         | Profizi, 841 (K)   | Harley and Dransfield (2003)             | *Sclerosperma gilletii*         | Figure 12 |
| *Sclerosperma prozianum*| Hall & Enti, GC36150 (K) | Harley and Dransfield (2003) | *Sclerosperma mannii*         | Figure 18  |
|                         | Hall & Enti, GC36150 (K) | Harley and Baker (2001)              | *Sclerosperma mannii*           | Figures 78 |
|                         | Hall & Enti, GC36150 (K) | Harley (1996)                          | *Sclerosperma mannii*           | Plate 16   |
| *Sclerosperma walkeri*  | Leonard, 1614 (?)  | Sowunmi (1972)                           | *Sclerosperma mannii*           | Plate 3   |

Note: Species affiliation according to Van Valkenburg et al. (2008). The drawings presented by Erdtman and Singh (1957) are not included in this table since they are not affiliated to any of the three *Sclerosperma* samples used in their study.
ziánun Type B pollen and S. profizianum Type C pollen are not just very close in morphology, but might be produced by the same individual plant, or there is some sort of sampling error.

Type C (Hall & Enti, GC36150 [K]; Profizi, 841 [KJ]) (Figures 1P–Y, 5; Table III)

Description. — Pollen, monad, heteropolar, P/E ratio oblate, outline straight-triangular to slightly concave-triangular in polar view, bean-shaped in equatorial view (convex distal face versus concave proximal face); equatorial diameter 37–42 µm in LM, 31–39 µm in SEM, polar axis 10–16 µm in LM; triporate, pori positioned sub-apically on the distal polar face, pori circular to elliptic, 4.0–6.5 µm in diameter, pori equipped with opercula; exine 1.7–2.5 µm thick in LM, nexine thinner than sexine; pollen wall tectate; sculpture scabrate in LM, perforate, rugulate/verrucate and fossulate in SEM; distal face perforate, perforations elliptic to slit-like, perforations often aligned in sinuous rows, rows of perforations outlining irregular shaped rugulae/verrucae (SEM); proximal face perforate and fossulate, perforations elliptic to slit-like, perforations often aligned in sinuous rows, rows of perforations and fossulae outlining irregular shaped rugulae/verrucae, sculpture becoming microrugulate to nanorugulate supra-layer (SEM).

Remarks. — Two LM micrographs showing pollen of this taxon are provided by Sowunmi (1972, plate 3, figure 8 and plate 4, figure 1), but assigned to S. manni (Table IV).

Discussion

Differentiating Sclerosperma pollen

Based on the pollen morphology of Sclerosperma presented herein it is clear that there are at least four different pollen morphologies produced by the three extant taxa. Sclerosperma manni and S. walkeri share similar pollen morphology, and are difficult to distinguish from each other in both LM and SEM. Sclerosperma profizianum produced three different pollen morphologies (Types A, B, and C), distinguishable from each other and from S. manni and S. walkeri, particularly in SEM.

Using LM only, Sclerosperma pollen can be divided into reticulate (including S. manni, S. profizianum Type A, and S. walkeri) and non-reticulate (including S. profizianum Type B and C; Table III). The reticulate pollen are further divided into coarsely reticulate (including S. manni and S. walkeri) versus finely reticulate (S. profizianum Type A; compare Figure 1A and 1Z with Figure 1F). Our measurements indicate the coarsely reticulate pollen of S. manni and S. walkeri can be set apart using the length of their polar axis, which is longer in the pollen of S. walkeri (15–19 µm) than in S. manni (9–15 µm). The non-reticulate Sclerosperma pollen grains are distinguishable by having rugulate (S. profizianum Type B) or scabrate (S. profizianum Type C) sculpture. The rugulate S. profizianum Type B pollen is usually smaller than that of S. profizianum Type C (Table III), and the outline of the pollen in polar view is more convex-triangular in S. profizianum Type C versus concave-triangular in S. profizianum Type B.

Applying additional SEM, there are a number of details separating the three different Sclerosperma pro-
S. profizianum pollen types from each other and from the pollen of S. mannii and S. walkeri. The magnification obtained using SEM shows that S. mannii and S. walkeri pollen is more or less identical (Table III). The only noticeable sculpture difference, so far, is that the free-standing columellae in S. walkeri pollen are more frequent and conspicuous than in S. mannii pollen (compare Figure 2E and 2F with Figure 6E and 6F). Still, the S. mannii and S. walkeri pollen are easily distinguished from the three S. profizianum pollen types. The S. mannii and S. walkeri pollen is reticulate with 16–25 lumina per 100 µm² at the central distal face versus microreticulate with 30–35 lumina per 100 µm² in S. profizianum Type A (compare Figures 2E and 6E with Figure 5E). The S. profizianum Type C pollen is perforate and rugulate/verrucate with 45–55 perforations per 100 µm² at the central polar face, and the S. profizianum Type B pollen is fossulate and rugulate/verrucate with the perforations hidden in the fossulae (compare Figure 5E with Figure 4E; Table III). Furthermore, the S. profizianum Type A pollen has regularly distributed elliptic to circular or slit-like lumina versus perforations aligned in sinuous rows in S. profizianum Type C versus tiny perforations aligned (hidden) within the fossulae in S. profizianum Type B pollen. Also, the operculum in S. profizianum Type A pollen has a clear reticulate supra-layer versus perforate in both S. profizianum Types B and C pollen (compare Figure 3C with Figures 4C and 5C).

Pollen morphology and taxonomic resolution

The ‘identical’ pollen of Sclerosperma mannii and S. walkeri are from sites near the centre of distribution for the genus (see map 1 in Van Valkenburg et al. 2008). Pollen of S. profizianum Type C is from a disjunct population in Ghana and is also found in a more centrally located population in Republic of the Congo. The two other S. profizianum pollen Types, A and B, are from the southern edge of the distribution of that species/genus near the border between Democratic Republic of the Congo and Angola.

There are many examples of clades in which species can be clearly separated on the basis of plant body and reproductive parts, but the pollen produced by them are similar or identical to each other morphologically (termed stenopalynous taxa, see Halbritter et al. 2018). However, it is not common for a single species to produce two or more distinct pollen morphologies (in sculpture and/or size), unless the plants are heterostylos (for a list of such genera see table 1 in Ganders 1979). There are no hints in the literature that heterostyly occurs in Sclerosperma.

We did not discover more than a single pollen type from a particular or several anthers out of an individual Sclerosperma flower; the pollen morphology observed within an anther or anthers were distinct and consistent within an individual or between flowers from the same herbarium sample. The pollen morphology of S. profizianum Types B and C suggests that they are very close, and based on previous work by Harley on some of the same herbarium material [Profizi, 841 (K)] it is even possible that they were produced by the same plant. Still, M.M. Harley only figured a single pollen grain in SEM (Harley & Dransfield 2003; Harley 2004; Dransfield et al. 2008) and therefore a sampling error or contamination cannot be excluded. The material studied might also have been assigned to the wrong collector information. Disregarding all that, and assuming that the Types B and C pollen originate from the same plant or taxon it is clear that S. profizianum still seems to produce two undoubtedly different pollen types: (1) the microreticulate pollen Type A and (2) the fossulate/perforate Type B/C pollen.

The reason Sclerosperma profizianum seems to produce different pollen types is unclear currently, but we can think of two possible explanations for this situation. First, S. profizianum is a unique taxon that produces different pollen types without being heterostylous. This seems very unlikely, but cannot be excluded. Second, the currently accepted species definitions in Sclerosperma do not reflect its actual biological diversity. Sclerosperma profizianum may be composed of more than a single natural species, or at least it may be in the process of genetic diversification related to its disjunct distribution and marginal occurrences. This could explain observed variations in pollen morphology, including the intermediate sculpture features of S. profizianum pollen Type A (microreticulate versus reticulate in S. mannii S. walkeri, and perforate in S. profizianum Type B/C). Whatever the explanation, it cannot be resolved from the data presented here.

Conclusion and outlook

Combined LM and SEM analyses demonstrate that there are four different pollen morphologies produced by Sclerosperma. Sclerosperma mannii and S. walkeri share similar pollen morphologies, but S. profizianum produces three different pollen types. Despite the detailed pollen work presented here and all the available literature regarding Arecaceae taxonomy, pollen morphology and phylogeny (e.g. Dransfield et al. 2008), there is still much that needs to be studied. For Sclerosperma, it would be vital to explore intrageneric relationships and conduct a
molecular phylogenetic study using several representatives from each alleged species to see how they align in a phylogenetic tree. In this sense it would be interesting to see if the S. mannii samples group together and appear as sister taxon to the S. walkerii samples, and if all the S. profizianum fall into one branch or are not clearly resolved suggesting some sort of species differentiation. When conducting such a study it would be highly informative to analyse pollen morphology from the same plants used for the molecular phylogeny and plot the pollen on the resulting tree. In such a case the evolution of pollen morphology in Sclerosperma could be resolved. Also, the comprehensive LM and SEM based pollen morphology presented here will now allow for a better determination of fossil Sclerosperma pollen grains and the re-analyses of fossil material previously affiliated to this genus.

Acknowledgements

This study was funded by the Austrian Science Fund (FWF) with a grant to FG, project number P29501-B25. The authors thank the herbaria and staff members of the Botanic Garden Meise (BR), the Royal Botanic Gardens, Kew (K), and Naturalis (WAG; the National Herbarium of the Netherlands) for providing flower material for this study.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

This work was supported by the Austrian Science Fund [P29501-B25].

ORCID

Frödeir Grimsson (http://orcid.org/0000-0002-1874-6412)
Johan L.C.H. van Valkenburg (http://orcid.org/0000-0001-7281-7819)
Jan F. Wieringen (http://orcid.org/0000-0003-0566-372X)
Alexandros Xafis (http://orcid.org/0000-0001-5528-3106)
Bonnie F. Jacobs (http://orcid.org/0000-0002-4770-0181)
Reinhard Zetter (http://orcid.org/0000-0002-0220-6921)

References

APG IV. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. Botanical Journal of the Linnean Society 181: 1–20. doi:10.1111/bot.2016.181.issue-1.

Bouro bou Bourobou PH, Niangadouma R, Issembe Y, Couvreur TLP. 2016. Two new records of palm species for Gabon: Sclerosperma profizianum Valk. & Sunder. and Eremospatha quique-costulata Becc. Biodiversity Data Journal 4: e10187. doi:10.3897/BDJ.4.e10187.

Dransfield J, Uhl NW, Asmussen CB, Baker WJ, Harley MM, Lewis CE. 2008. Genera Palmarum. The evolution and classification of palms. Kew: Kew Publishing.

Erdman G, Sing G. 1957. On the pollen morphology in Sclerosperma mannii. Bulletin du Jardin botanique de l’État a Bruxelles 27: 217–220. doi:10.2307/3666958.

Ganders FR. 1979. The biology of heterostyly. New Zealand Journal of Botany 17: 607–635. doi:10.1080/0028825X.1979.10432574.

Grimsson F, Grimm GW, Zetter R. 2013. Tiny pollen grains: First evidence of Saururaceae from the Late Cretaceous of western North America. PeerJ 5: e3434. doi:10.7717/peerj.3434.

Grimsson F, Grimm GW, Zetter R. 2018. Evolution of pollen morphology in Loranthaceae. Grana 57: 16–116. doi:10.1080/00173130.2016.1261939.

Halbritter H, Ulrich S, Grimsson F, Weber M, Zetter R, Hesse M, Buchner R, Svojtka M, Frosch-Radivo A. 2018. Illustrated pollen terminology. Second edition. Vienna: Springer.

Harley MM. 1996. Palm pollen and the fossil record. PhD Thesis, University of East London, London, UK.

Harley MM. 1999. Tetrad variation: Its influence on pollen form and systematics in the Palmae. In: Kurmann MH, Hemsley AR, eds. Evolution of plants architecture, 289–304. Kew: Kew Publishing.

Harley MM. 2004. Triarperate pollen in the monocotyledons: Configurations and conjectures. Plant Systematics and Evolution 247: 75–122. doi:10.1007/s00606-003-0107-x.

Harley MM, Baker WJ. 2001. Pollen aperture morphology in Arecaceae: Application within phylogenetic analyses, and a summary of the fossil record of palm-like pollen. Grana 40: 45–77. doi:10.1080/00173130152591877.

Harley MM, Dransfield J. 2003. Triporate pollen in the Arecaceae. Grana 42: 3–19. doi:10.1080/00346667(04)0008535.

Harley MM, Hall DH. 1991. Pollen morphology of the African palms. Palaeoecology of Africa and the surrounding islands 22: 11–25.

Punt W, Hoen PP, Blackmore S, Nilsson S, Le Thomas A. 2007. Glossary of pollen and spore terminology. Review of Palaeobotany and Palynology 143: 1–81. doi:10.1016/j.revpalbo.2006.06.008.

Sowummi MA. 1972. Pollen morphology of the Palmae and its bearing on taxonomy. Review of Palaeobotany and Palynology 13: 1–80. doi:10.1016/0034-6667(72)90044-9.

Van Valkenburg JLCH, Sunderland TCH, Couvreur TLP. 2008. A revision of the genus Sclerosperma (Arecaceae). Kew Bulletin 63: 75–86. doi:10.1007/s12225-007-9002-x.

WCSP. 2018. World Checklist of Selected Plant Families. Facilitated by the Royal Botanic Gardens, Kew. Published on the Internet, http://wcsp.science.kew.org/; accessed 6 February 2018.