Phase 1 Study of an Inactivated Vaccine against American Tegumentary Leishmaniasis in Normal Volunteers in Brazil

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A Phase 1 double-blind placebo-controlled study was performed to evaluate a vaccine against American tegumentary leishmaniasis in 61 healthy male volunteers. Side effects and the immune response to the vaccine were evaluated, with 1- and 2-dose schemes, with intervals of 7 or 21 days, each dose containing 1440 mg of protein N antigen of a single strain of Leishmania amazonensis (PH 8) diluted in merthiolated saline (1:10,000). Merthiolated saline and an inert substance were used as placebos. No significant clinical alterations were found following the respective injections in the vaccinated individuals as compared to the placebos, except for local pain, which was associated significantly with injection of the vaccine. The laboratory alterations we observed bore no association with the clinical findings and were unimportant. We observed no differences between the groups with regard to seroconversion or the Montenegro skin test. However, the group that received a single dose of the vaccine and the one that received two doses with a 21-day interval displayed cutaneous induration significantly larger than in the control group, with 100%, 100%, and 66% conversion in the skin test, respectively. We concluded that the vaccine does not present any major side effect that would contraindicate its use in healthy individuals.

Key words: Leishmania - vaccine - Phase I study - side effects - Montenegro skin test

American tegumentary leishmaniasis (ATL) is an endemic anthropozoonosis undergoing expansion on the American continent. Its importance is due to its broad range of clinical manifestations, varying from self-limiting localized cutaneous lesions to disfiguring lesions in the mucosal and diffuse forms, in addition to the difficulties encountered in treatment and control. Pentavalent antimonials, the main drugs currently used, are highly toxic and expensive and require prolonged treatment schemes, and some forms of the disease are resistant to this treatment (Bryceson 1987, Olliaro & Bryceson 1993). On-going research on alternative drugs is being done, and the action of immunotherapy is being assessed (Badaró & Johnson Jr 1993, Monjour et al. 1994). In Brazil the incidence coefficient varies from 93.07 cases per 100,000 inhabitants in the Northern region (Amazônia) to 3.91/100,000 inhabitants in the Southern region, with 31,713 cases notified in 1994 (Marzochi & Marzochi 1994).

In Brazil, there are at least six species of dermotropic Leishmania for humans in various ecological situations (Lainson et al. 1994). Control is thus complex, and there are different patterns of transmission (Marzochi & Marzochi 1994). In the Amazon region it implies human contact with forests through participation of sylvatic reservoirs and various sandfly vectors. In areas that have been settled longer, transmission is generally rural and peridomiciliary, and there is discussion concerning possible sources of infection such as canines, equids, rodents, and humans, as well as association with various species of sandflies which adapt well to altered environments (Desjeux 1992, Marzochi 1992, Lainson et al. 1994, Marzochi & Marzochi 1994). Subclinical infection is highly frequent (Marzochi et al. 1980, Isaza 1980, Guerra et al. 1985, Cordova-Uscanga et al. 1993) and probably underestimated. Current control methods oriented towards combating the vector are also debatable and limited to peridomiciliary transmission (Marzochi & Marzochi 1994, Oliveira Filho & Melo 1994).
Utilization of a vaccine against ATL in Brazil was a pioneering initiative; produced on the basis of cultures of killed parasites, it led to a reduction in the number of disease cases in the populations studied (Salles Gomes 1939, Pessoa & Pestana 1940, Pessoa 1941). However, these studies were abandoned, probably due to the decline in the incidence of ATL in old endemic areas. More recently, vaccination studies in new areas yield results that agreed with these previous studies, due to acquisition of protective immunity, assessed principally by post-vaccinal conversion of the Montenegro skin test (leishmanin) (Mayrink et al. 1979, 1985, Antunes et al. 1986, Castés et al. 1994).

In the Old World, vaccination was employed with live promastigote forms of *L. major* using intradermal inoculation (Nadim 1990), and considerable reduction was also achieved in the number of cases of the disease among vaccinated individuals, although the side effects were considerable, leading to interruption of the tests.

The evaluation of the *Leishmania* vaccine in this research stems from studies by Mayrink et al. (1979, 1985) producing a vaccine with killed promastigote forms from a pool of *Leishmania* strains (240 mg of protein nitrogen per dose) and immunizing a population from an endemic area for this disease, obtaining 78.4% of conversion for the Montenegro skin test in previously negative individuals; in a new test, using the same antigen in a population from a different area, some 85% of conversion for the Montenegro skin test and a significantly lower incidence rate for the disease were achieved. Factors such as the disappearance of ATL in the area studied in 1979 and a major population migration observed in 1985 hindranded evaluation of results.

Later, in a controlled study, with the same vaccine, on military personnel headquartered in an endemic area, Antunes et al. (1986) observed similar disease incidence rates among vaccinated and non-vaccinated individuals, although there was a significant predominance of conversion in the Montenegro skin test in the vaccinated group. The authors attributed the disease incidence rate in the vaccinated individuals to immunosuppression caused by immunization for yellow fever shortly after the vaccination for leishmaniasis.

Beginning in 1988 the ATL vaccine proposed by Mayrink et al. (1985) began commercial production with the name LEISHVACINO®, however, based on a recommendation by the World Health Organization (WHO), from there on it was composed of a single strain of *Leishmania*, and the reference PH8 strain of *L. amazonensis* (IFLA/BR/67/PH8) belonging to the initial pool was selected. Although no side effects have been observed in the utilization of this vaccine in Brazil, thus far no well-controlled assessment of this has been done in the various studies cited above. The objective of this Phase 1 study was to perform an analysis of possible clinical and laboratory side effects of LEISHVACINO®, constituted from a single strain of *Leishmania* and with a greater concentration of antigen (1440 mg of protein nitrogen per dose) in normal volunteers.

**MATERIALS AND METHODS**

Before started, the study was submitted and approved by an institutional human subjects committee responsible for monitoring the ethical conduct of research involving human subjects of the Oswaldo Cruz Foundation (Fiocruz).

**Volunteers and skin test** - The study initially included 206 male military volunteers ranging from 18 to 40 years of age, headquartered at a military unit in Rio de Janeiro, who were submitted to an initial Montenegro skin test corresponding to 0.1 ml containing 40 mg of protein nitrogen/ml of standardized antigen (Melo et al. 1977), produced with 5 strains of *Leishmania* (the same composition as that of the vaccine described by Mayrink et al. 1979), according to Good Manufacturing Processes (GMP), applied intradermally on the anterior surface of the forearm. The reading was performed and recorded after 48 hr, by stamping the papule on paper (Sokal 1975). Individuals with a negative response, with induration less than or equal to 5 mm along the longest diameter were accepted for the vaccination protocol, making a total of 61 volunteers. All of these individuals also had negative serology for HIV and *Trypanosoma cruzi*, had no signs or symptoms of infectious or chronic degenerative disease, and denied skin allergies. None of them had a previous or recent history or scars compatible with leishmaniasis. One individual, who was a chronic hepatitis C carrier, expressed his desire to participate as a volunteer, and signed a second, specific term of responsibility in addition to the protocol term.

**Vaccine** - The vaccine used in the study (BIOBRAS, S/A) was produced according to GMP and Mayrink et al. (1979), but only with the PH8 (*L. amazonensis*) strain, at a concentration of 1440 mg of protein nitrogen per dose, made up of 50% of whole promastigote forms of the parasite and 50% of macerated material, all diluted in merthiolated saline at 1:10,000.

**Placebo** - Two types of placebo were used, in the control group, consisting of the vaccine vehicle, 1:10,000 merthiolated saline, and an inert substance, physiological saline.

**Vaccinal scheme** - The vaccine or placebo was
applied IM (left deltoid region) in doses of 1.5 ml on days 0, 7, and 21 of the study, in double-blind fashion. The volunteers were divided into groups according to the scheme presented in Table I.

**Non-specific clinical and laboratory evaluation** - Clinical examination was performed before each injection and was repeated at 24 hr, 72 hr, and 7 days. Blood samples were taken at the same time (without prior fasting) for the following laboratory tests: blood - CBC, ESR, platelets; hepatic function - SGOT, SGPT (Celm), PT, bilirubin (Merck), LDH, alkaline phosphatase (Biotrol), GGT (Merck); and renal function - urea (Merck) and creatinine (Celm). We defined laboratory alteration as any deviation outside the standard limits of the test, regardless of clinical correlation.

**Evaluation of immune response** - This was measured 20 days after the final injection. The cellular immune response was assessed on the basis of positive conversion in the Montenegro skin test, with induration > 5 mm, using the same antigen and procedure as in the initial test. Diameters of induration were also observed in the different groups. The serological response was evaluated by indirect immunofluorescence (IIF) with *L. major*-like antigen (Marzochi et al. 1980) and ELISA (Voller et al. 1976), with antigen from the same parasite, and negative serology was considered that below dilution of 1:40 for both tests.

**Statistical analysis** - The data were analyzed utilizing the following tests: Fischer’s exact and Kruskal-Wallis.

**RESULTS**

**Clinical evaluation** - Table II shows the percentage of clinical alterations found by group, per injection. All manifestations reported by volunteers were catalogued, as well as alterations observed at the injection site and systemic alterations. Mild or moderate local pain was the most frequently reported effect in all of the groups and was more frequently associated with injections of LEISHVACINÒ. Vaccinated individuals had a significantly greater frequency than the placebo group while the group, with the autoclaved double vaccine (G V) had the lowest frequency among the vaccinated groups, mainly compared to the group submitted to the same scheme, with a non-auto-claved vaccine (G IV). The local pain never extended beyond 24 hr, was not accompanied by other local inflammatory signs or regional lymph node enlargement, nor did it require specific treatment except in the case of two volunteers who self-mediated, and it did not jeopardize physical activity in the barracks among the volunteer who did present this side effect. In one case only, the second dose of the vaccine set off a erythematous-pruriginous reaction at the site of application. Other reactions as described in Table I had similar frequencies among the various groups, including the placebo group, and it was not possible to associate them with any specific type of injection, nor did they require treatment. At no point in the study did the volunteers have to interrupt their normal physical and intellectual activities, nor was there any report of harm to their activities following application of the injections.

**Laboratory evaluation** - Table III shows the frequency of alternations in laboratory tests as found before the trial began and after injections of placebos and vaccine in the various groups. At these various moments the proportions of altered tests were similar.

No case to case correlation was observed between the clinical and laboratory alterations for any volunteer, whether he was from the placebo group or the vaccinated group after receiving vaccine or placebo.

**TABLE I**

| Groups | Schemes | Injection, day 0 | Injection, day 7 | Injection, day 21 | No. of volunteers |
|--------|---------|-----------------|-----------------|------------------|------------------|
| GI     | Placebo | Merthiolated saline | Physiological saline | Merthiolated saline | 12               |
| GII    | Vaccine, single dose | LEISHVACIN® | Physiological saline | Merthiolated saline | 12               |
| GIII   | Vaccine, double dose | LEISHVACIN® | LEISHVACIN® | Merthiolated saline | 13               |
| GIV    | Vaccine, double dose | LEISHVACIN® | Physiological saline | LEISHVACIN® | 12               |
| GV     | Autoclaved vaccine, double dose | LEISHVACIN® autoclaved | physiological saline | LEISHVACIN® autoclaved | 12               |
**DISCUSSION**

The results obtained in this study corroborate those of previous studies which, although not having the objective of this study - did not demonstrate the presence of side effects in the utilization of ALVAC vaccine produced with killed promastigotes, lower antigen concentration, and pool of strains (Pessoa & Pestana 1940, Pessoa 1941, Mayrink et al. 1979, 1985, Antunes et al. 1986). In the clinical and laboratory sense, the vaccine utilized in this study may be considered innocuous, since it was not possible to associate it with any parallel event of a serious nature (Mallory 1987), and there have been even non-specific side effects that can be associated with a variable reaction to the physical discomfort that is common to deep intramuscular injections. The frequency of pain in the placebo group, which was higher when the merthiolated saline was used (in the same concentration as that employed in the vaccine), merely indicates the occurrence of non-specific side effects that can be associated with Thiomersal. This dilutant, used in domestic and professional routine as a topical antiseptic and also included in numerous other vaccines, has been implicated in reactions of an allergic nature (Mallory 1987), and there have even been suggestions that it be removed from the composition of any group or injection.

**TABLE II**

Frequency of clinical alterations in volunteers participating in Phase I LEISHVACINÒ study, by volunteer group and injection

| Groups a (No.) | Local pains b | Headache | Asthenia | Other | Local pains c | Headache | Asthenia | Other | Local pains d | Headache | Asthenia | Other |
|----------------|---------------|----------|----------|-------|---------------|----------|----------|-------|---------------|----------|----------|-------|
| GI (12)        | 33.3          | 0.0      | 8.3      | 0.0   | 16.6          | 8.3      | 0.0      | 16.6  | 33.3          | 0.0      | 0.0      | 0.0   |
| GII (12)       | 75.0          | 8.3      | 8.3      | 8.3   | 73.9          | 8.3      | 0.0      | 15.3  | 7.6           | 7.6      | 0.0      | 0.0   |
| GIII (13)      | 69.2          | 0.0      | 0.0      | 0.0   | 0.0           | 0.0      | 0.0      | 0.0   | 81.8          | 0.0      | 0.0      | 9.0   |
| GIV (12)       | 91.6          | 0.0      | 8.3      | 0.0   | 0.0           | 0.0      | 0.0      | 0.0   | 33.3          | 0.0      | 0.0      | 16.0  |
| GV (12)        | 41.6          | 8.3      | 8.3      | 0.0   | 0.0           | 0.0      | 0.0      | 0.0   | 0.0           | 0.0      | 0.0      | 0.0   |

a: GI: placebo; first and third injections: merthiolated saline; second: physiological saline; GII: first injection: LEISHVACINÒ; second and third: physiological saline; GIII: first and second injections: LEISHVACINÒ; third: physiological saline; GIV: first and third injections: LEISHVACINÒ; second: physiological saline; GV: same as GIV with autoclaved vaccine; b: p< 1, 6%; Fischer's exact test; c: p< 0, 1%; Fischer's exact test; d: p< 0, 3%; Fischer's exact test.
routine vaccines (Scarpa et al. 1992, De La Quadra 1993, Handley et al. 1993).

Pain at the site of application, associated significantly with injections of vaccine, was mild in nature and did not interfere with the volunteers’ normal activities, even those requiring physical effort. It was compatible with the pain caused by other vaccines traditionally in use, and was described by some volunteers as “milder than that caused by the tetanus toxoid (TT)”. The autoclaved vaccine presented a lower frequency of local pain as compared to the other vaccinated groups, after both the first injection and the second one after 21 days. The autoclave process may have fractionated the whole promastigotes, facilitating their absorption and reducing pain.

As a general rule, the laboratory tests used in this study should be evaluated in light of the individual’s clinical status, and never in isolation. Nonetheless, there were no significant differences as to the frequency of results outside the limits of the standard for normality, either before or after applications of vaccine and placebo. However, among the hematological alterations, only eosinophilia was more frequent both after injection of placebo and that of vaccine as compared to day zero (Table II), although this was not statistically significant. This rise may be explained by the presence of merthiolate in the vaccine and in almost all of the placebo injections (except for one group of 12 volunteers who received saline without merthiolate, once). In the general population, allergy to merthiolate is reported as 51% (Moriearty et al. 1978).

### Table III

| Date          | Injection(\(^{a}\)) | Leucocytes \(^{c}\) | Eosinohils \(^{c}\) | Neutrophils \(^{c}\) | Lymphocytes \(^{c}\) | Urea \(^{d}\) | PT \(^{e}\) |
|---------------|----------------------|---------------------|---------------------|---------------------|---------------------|-------------|----------|
| None          |                      | 0.08                | 0.13                | 0.21                | 0.29                | 0.45        | 0.08     |
| Day 0 (No.)   | Vaccine N (37)        | 0.16                | 0.16                | 0.33                | 0.41                | 0.45        | 0.08     |
|               | Vaccine A (12)        | 0.08                | 0.16                | 0.33                | 0.33                | 0.33        | 0.08     |
| Day 7 (No.)   | Placebo I (37)        | 0.02                | 0.08                | 0.32                | 0.45                | 0.18        | 0.00     |
|               | Vaccine N (12)        | 0.08                | 0.33                | 0.41                | 0.41                | 0.25        | 0.08     |
|               | Placebo I (25)        | 0.16                | 0.24                | 0.28                | 0.28                | 0.24        | 0.16     |
| Day 21 (No.)  | Placebo M (12)        | 0.00                | 0.16                | 0.25                | 0.41                | 0.41        | 0.16     |
|               | Vaccine N (12)        | 0.16                | 0.25                | 0.50                | 0.50                | 0.50        | 0.33     |
|               | Vaccine A (12)        | 0.33                | 0.41                | 0.41                | 0.66                | 0.08        | 0.16     |

Types of injection a: Vaccine N: non-autoclaved vaccine; Vaccine A: autoclaved vaccine; Placebo M: merthiolated saline (at 1:10000); Placebo I: physiological saline (\(^{b}\)): number of tests; \(^{c}\): frequency of alterations in number of cells; \(^{d}\): frequency of alterations in blood urea levels; \(^{e}\): frequency of alterations in prothrombin time.

### Table IV

Response of volunteers of LEISHVACIN\(^{\circ}\) phase I study to the second intradermal test, 47 days after first injection

| Montenegro skin test (% of positive volunteers/group, according to enduration diameter) |
|---------------------------------------------------------------|
| Groups \(^{a}\) (No.) | < 5 mm | €5mm<10 mm | €10 mm | Total | Mean induration |
| GI (12)           | 4 (33.40) | 2 (16.6) | 6 (50.00) | 8 (66.6) | 8.58 |
| GII (12)          | 0 (0.00) | 1 (8.33) | 11 (91.66) | 12 (100.0) | 13.66 |
| GIII (13)         | 1 (7.70) | 2 (16.6) | 10 (76.90) | 12 (95.6) | 10.53 |
| GIV (12)          | 0 (0.00) | 2 (16.6) | 10 (83.30) | 12 (100.0) | 14.25 |
| GV (12)           | 2 (16.70) | 1 (8.33) | 9 (73.00) | 10 (83.33) | 11.58 |
| Total (61)        | 7 (11.47) | 8 (13.11) | 46 (75.40) | 54 (88.51) | - |

\(^{a}\): GI : placebo; first and third injections: merthiolated saline; second: physiological saline; GII: first injection: LEISHVACIN\(^{\circ}\); second and third: physiological saline; GIII: first and second injections: LEISHVACIN\(^{\circ}\); third: physiological saline; GIV: first and third injections: LEISHVACIN\(^{\circ}\); second: physiological saline; GV: same as GIV with autoclaved vaccine; \(^{b}\): 0.14 vs placebo group, Kruskall-Wallis test.
As for the biochemical tests, there was also no difference in the frequency of alterations observed before and after injections for the tests as a whole. The increases in waste nitrogen in some tests, regardless of whether before or after injections of vaccine or placebo, are associated with the absence of fasting among the volunteers.

In relation to evaluation of the immune response, several factors may have interfered in the results (Table III), requiring further assessment.

The conversion rate observed for the Montenegro skin test, was the highest as compared to previous studies, even considering a cut-off of 10 mm of induration, based on a recommendation by the World Health Organization (Castés et al. 1994). Nevertheless, we were unable to associate it definitively with injections of the vaccine, since we found a significant conversion rate for the skin test in the placebo group. We attempted to correlate this rate to exposure of volunteers to an endemic leishmaniasis area, where they stayed for two days during the study period and during which some received a third dose of TT. However, when we compare the response to the Montenegro skin test among volunteers who experienced this exposure to those who were not exposed (data not shown), we failed to find statistically significant differences. Neither did the third dose of TT display interference in this response.

The dilutant and the method used for evaluation in this study deserve further consideration. The dilutant used originally was phenol (Salles Gomes 1939, Pessoa & Pestana 1940), later replaced by merthiolate (Mayrink et al. 1979). However, allergic reactions have been described for Merthiolate that are similar to the cutaneous reactions to the Montenegro test, due to the mechanisms involved in the morphology of the reaction (type IV cellular hypersensitivity reaction) (Mallory 1987). Therefore, we cannot rule out the possibility that a portion of the reactions observed in this skin test are associated with an allergic response to the vaccine dilutant rather than a manifestation of protective immunity to the Leishmania antigen.

The choice of the Montenegro skin test as a method for evaluating the cellular immune response to LEISHVACIN® was based on characteristics of this response in individuals with ATL, as the presence of a developed and lasting cellular response in cured patients (with and without treatment), and a weak one (anergia) in individuals that are resistant to treatment (Salles Gomes 1939, Manson-Bahr 1987, Convit et al. 1993). The Montenegro test is considered highly specific and is used routinely in diagnosing the disease, although it is associated with recent and past infections with Leishmania, and some authors believe it can reflect, main positive throughout the individual’s lifetime (Montenegro 1926, Manson-Bahr 1987, Nascimento et al. 1993). Populations from endemic areas present relatively high frequencies of Montenegro-positive individuals unrelated to the clinical disease, indicating possible prior exposure to the parasite with asymptomatic infection (Guerra et al. 1985, Kadaro et al. 1993, Stolf et al. 1993). A “false-positive” response in individuals with other infections may occur in a few cases (Furtado 1980). Some authors also admit the possibility of an initial test altering the response to a second test, through sensitization (Nascimento et al. 1993).

Despite such drawbacks, the Montenegro skin test is considered the safest method for evaluating the immune response to Leishmania infection (Alimohammadian et al. 1993, Convit et al. 1993, Zijlstra & El-Hassan 1993). However, there is no doubt that a reassessment is required concerning its actual specificity when applied in populations from endemic areas for ATL, the correlation with the in vitro cellular immune response, alteration of the individual’s initial response in different areas after one or more tests, and other aspects. Sensitization induced by the test itself, or the test acting as a booster in individuals previously exposed to a common antigen, may be another hypothesis to explain the rate of positive response to the final test found in the placebo group. We performed a concurrent evaluation (search for volunteers) among non-vaccinated military submitted to the Montenegro skin test and retested 65 days later, showing a conversion rate equal to that in our placebo group (data not shown).

We conclude that in terms of safety, LEISHVACIN® can be used in human populations, even with two doses of non-autoclaved vaccine, at a concentration of 1440 mg of protein nitrogen per dose, without causing side effects capable of jeopardizing normal daily activities in young male adults.

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