Abstract: Implant periapical lesion (IPL) is an infectious-inflammatory alteration surrounding an implant apex. It is a multifactorial disease that may ultimately cause implant failure. The diagnosis of IPL is based on examination of clinical manifestations and apical radiolucency. Many etiologies have been attributed to IPL, including preexisting microbial pathology and surgical trauma. Moreover, many systems have been used to classify IPL based on different parameters. To date, non-surgical and surgical treatment, as well as removal of failed implants, have been considered to successfully manage IPL. However, prevention of IPL surpasses all modes of treatment. An increased number of IPL cases are expected as implants have become standard for tooth replacement in dentate arches. Therefore, it is necessary to understand IPL more comprehensively. Herein, an introduction to IPL, including its etiology, diagnosis, classification, treatment, and prevention, has been undertaken.

Keywords: implant periapical lesion; retrograde peri-implantitis; periapical implant pathology; endodontic-implant pathology; periapical implant lesion; retrograde peri-implant infection; apical peri-implantitis

1. Introduction

Implant periapical lesion (IPL) is an infectious-inflammatory alteration surrounding implant apex that ultimately leads to implant failure [1–4]. IPL was first reported in 1992, when McAllister et al. showed two submerged implants with periapical radiolucency [5]. Different nomenclatures were used in literature to refer to IPL, including apical implantitis, apical peri-implantitis, early peri-implantitis, endodontic-implant pathology, inflammatory implant periapical lesion, periapical implant lesion, periapical implant pathology, retrograde peri-implant infection, and retrograde peri-implantitis [6–9]; however, these terms were defined somewhat differently [7,10]. For example, retrograde peri-implantitis was defined as a clinically symptomatic radiolucency around the apical area of an osseointegrated implant, which differed from apical infection in an implant without osseointegration [11].

The prevalence of IPL varies from 0.26% to 9.9%, and its incidence may reach 7.8% if adjacent teeth receive endodontic treatment [6,7,9,12,13]. Additionally, recent studies have shown that IPL occurs with a frequency of 0.34–3.8% [14–16]. In the maxilla and mandible, the incidence reported were 0.28–5.8% and 0.42–2.7%, respectively [11,14,15]. In contrast, other studies demonstrated a high prevalence in the maxillary premolar [10,17].

The endodontic status of the tooth extracted from the implant site and adjacent teeth could affect the occurrence of IPL [18]. An increased presence of IPL has been noted when endodontic treatment fails in adjacent teeth or prior to implant placement [10]. Therefore, the incidence was low when implant therapy was in its initial stages as implants were mostly placed in edentulous ridges. Currently, more IPL cases are expected as implants are used to replace single/multiple teeth in partially edentulous ridges [7,19,20]. There are some differences between IPL and peri-implantitis based on the involved pathogens (i.e., endodontic pathogens in IPL and periodontal pathogens in peri-implantitis), rate of expansion, and pathway of infection [7]. Therefore, modes of treatment and prevention of IPL mildly differ from those of peri-implantitis.
The aim of this narrative review article was to introduce IPL, including etiology, diagnosis, classification, treatment, and prevention.

2. Materials and Methods

A literature search was performed on PubMed for English papers on IPL, published between January 1992 and June 2021. We found 60 articles, using different search terms focused on title, including apical implantitis, apical peri-implantitis, endodontic-implant pathology, implant periapical lesion, periapical implant lesion, periapical implant pathology, retrograde peri-implant infection, and retrograde peri-implantitis. Eight papers were excluded after reviewing their abstracts. Exclusion criteria were only involving peri-implantitis, endodontic implant and knowledge towards IPL. Then, seven retrospective studies, seven review articles and other case report/case series papers were included. In addition, 37 papers were searched from the reference list of the above 52 papers. Therefore, a total of 89 papers are cited in present review.

3. Results

3.1. Etiology

It is well known that IPL is a multifactorial disease [21]. However, there is no consensus on the etiology of IPL [22], and it remains unclear whether IPL is composed of healthy tissue, new tissue destruction, or activation of a preexisting condition [23]. Dental implants do not withstand any bacterial challenge during the first stage of osseointegration [24]. Thus, bacterial contamination appears to be the predominant cause, with several other contributing factors [17,25]. The possible etiologies of IPL may be divided into three categories: patient-associated factors, material-associated factors, and method-associated factors (Figure 1). However, local bone and intraoral factors (e.g., etiology of tooth extraction and the endodontic condition of adjacent teeth) were stronger than other factors associated with IPL [15].

![Figure 1. Possible etiologies of implant periapical lesion (IPL): subject-associated factors, material-associated factors, and method-associated factors.](image-url)
3.1.1. Patient-Associated Factors

Patient-Associated Factors include the presence of pre-existing/adjacent bone pathology (e.g., residual cyst), residual root fragments or foreign bodies; implant placed into infected sinus or poor quality bone, and patient under long-term oral bisphosphonates or those with a smoking habit [8,17,23,26–36]. Wang showed implant periapical radiolucency lesions in one patient taking long-term oral bisphosphonates [33].

Bacteria from endodontic-failed tooth or residual lesion might be reactivated by implant site osteotomy and subsequent colonization on implant apex [16,37]. There was a significant difference observed between the presence and absence of IPL in terms of endodontic failure at implant site and/or adjacent location [10]. The apical region of endodontically treated teeth often reveals histological signs of inflammation or persistent micro-organisms, even if optimal bone healing is seen on radiographs [38,39]. Additionally, residual encapsulated bacteria maybe remain in bone even after debridement of extraction socket, leading to IPL initiation [5,11,24,27]. It is difficult to detect other pathologies in radiographs, even if cone beam computed tomography (CBCT) is used [11]. Therefore, the endodontic status of tooth extracted from the implant site has a major influence on IPL [11,40]. An IPL presented in 31.5% of implants with previous endodontic treatment, but only in 2.84% of implants without previous endodontic treatment [10]. The odds ratio for IPL was 7.2 for an extracted tooth with an endodontic treatment history when compared with a pathology free tooth. Moreover, the odds ratio for IPL was 8 for adjacent teeth with an endodontic treatment history when compared with pathology-free teeth [18]. Moreover, the odds of developing early IPL in implant sites with apical lesion was up to 26.3-fold [14]. However, recent retrospective study including 25 patients showed that previous endodontic surgery in implant site did not increase risk of IPL [41].

Residual root fragments at the implant site, commonly seen in maxillary premolars, is another possible etiology [17,42]. Pre-mature loading usually results in early implant failure; however, it is controversial whether early loading is an etiology of IPL [3,11]. Other than local factors associated with implant site or adjacent tooth, Wang et al. reported apical radiolucency around three implants in a patient under long-term use of oral bisphosphonates [33]. Thus, the medical status of patients should be evaluated before dental implant therapy.

3.1.2. Material-Associated Factors

Material-associated factors include implant surface design and biocompatibility, specific techniques of implant surgery, and implant surface contamination during production and insertion [23,28]. Further, external contamination of the implants might result in implant periapical lesions [23].

In addition, the incidence of IPL is higher for TiUnite implant surface than for machined implant surface. The machined implants usually show higher failure rate when contacting with endodontic pathology; however, TiUnite implants have coronal osseointegration before fibrous capsulation reached this area. Thus, there was a greater incidence of apical lesion and less failure rate in TiUnite implants than machined implants [11]. Nevertheless, the material factors associated with IPL are fewer than patient-associated factors and method-associated factors [7].

3.1.3. Method-Associated Factors

Method-associated factors include overheating of bone during implant site preparation, compression of bone due to excessive tightening, vascular impairment/ischemia, apical bone fenestration, implant close to adjacent teeth, and surgical bed contamination (bacteria, epithelial cells or starch of gloves) [2,23,29,30,43,44]. In addition, one IPL case associated with the “socket shield technique” was reported recently [45].

When an implant fixature is placed close to the adjacent tooth, the blood supply of the adjacent tooth may be compromised due to osteotomy and flap elevation [24]. Moreover, immediate implant treatment (odds: 21-fold) or an apical mesial distance of
≤1.5 mm between implant and tooth are risk factors (5.12-fold) associated with early IPL [14]. Overheating of bone during implant site preparation and compression of bone during tightening could result in bone necrosis, which is also associated with IPL [5,28]. The occurrence of IPL may be associated with traumatic implant placement, making it preventable [46].

### 3.2. Examination and Diagnosis

The findings of IPL are summarized in Figure 2. The symptoms and signs (S/S) of IPLs include: swelling, erythema, pain, tenderness, suppuration, sinus tract, fistula, abscess, and apical radiolucency [7]. Paraesthesia was also reported in one case with a large IPL [12]. Apical radiolucency is the prominent and commonest finding in IPL; however, the clinical signs may be different in each case, e.g., inactive IPL without infectious and/or inflammatory S/S clinically [30]. The sinus tract is the most common clinical expression; however, severe pain is uncommon and deep pockets are rare [17]. The time of diagnosis varies from 7 days to 4 years after implant surgery in reports, and it is commonly made within the first 6 months after implant placement [7,8,47]. The extent of active IPLs tends to increase with time, and thus, diagnosing IPL as early as possible is important to limit damage and initiate aggressive treatment if required [17]. Radiographic examination is valuable in early detection of IPL, even before sinus tract formation. Periapical radiographs are recommended immediately after implant placement as a baseline and at the third month of healing for detection of chronic IPL [5,17]. However, periapical radiographs have a high risk of false-negative results [8]. The lesion might not be evident on periapical radiographs of the mandible; therefore, occlusal view radiographs or CBCT (cone beam computed tomography) are used for confirmation of mandibular IPL [26].

**Figure 2.** Different examination findings of implant periapical lesion.

Histologic examination of IPL has revealed necrotic bone, inflammatory infiltrate, and/or cyst, which may be related to vascular impairment and bone fracture in the apical area [8,29,30]. These findings are analogous with those of granuloma or abscess at the root apex of nonvital teeth [48]. In addition, gram-positive rods, gram-negative rods, and coccal and filamentous microbes were found in pathologic examination of IPL [5]. It is difficult to obtain a bacterial culture from IPL and there is a high risk of contamination during sampling of IPL [8]. There are higher counts of anaerobic bacteria than that of aerobic bacteria because of intrabony lesions [18]. *Staphylococcus aureus, Porphyromonas gingivalis, Corynebacterium*,
Klebsiella pneumoniae, Prevotella intermedia, Enterococcus faecalis, actinomycotic colonies, and various anaerobic and facultative bacteria were found in IPL; Porphyromonas gingivalis was the prominent species and Candida albicans was found occasionally [7,8,18,49,50]. Thus, the microbiological findings must be taken into consideration for antibiotic selection [18]. Klebsiella pneumonia, group D streptococci, Corynebacterium species, and alpha-hemolytic streptococci were discovered in the apical lesions of adjacent teeth [19]. The relationship between an extra-radicular infection and endodontic failure remains controversial [8]. Some authors suggested that rare extra-radicular infections are caused by Actinomyces and Propionibacterium. Different studies have shown that extra-radicular infections are common and involve Candida, Staphylococcus, Enterococcus, and Enterobacter [8]. In addition, Verdugo reported that periodontopathogen and Epstein-Barr virus (EB virus) associated periapical lesions may also induce IPL. It is suggested that saliva polymerase chain reaction for EB virus could be used to predict granulation tissue infection and as reference for antimicrobial therapy [51]. Saliva EBV-positive patients might have granulation tissue infection with Tannerella forsythia and Treponema denticola [51].

3.3. Classification

Different systems have been used to classify IPL (Figure 3). According to the activity of the infection, IPLs are classified into active/infective and inactive lesions [28]. An inactive lesion may be an apical scar resulting from placing an implant shorter than the prepared length. Active lesions are infected lesions with S/S, which include pain, tenderness, swelling and/or the presence of a fistulous tract [11]. Lesions are also classified according to their stage of progression into acute non-suppurative phase with inflammatory infiltrate, acute supplicative phase with periapical granuloma, and chronic phase with periapical abscess [1]. There were acute symptoms in non-suppurative stage, but no radiographic change was observed [6]. The categories in the classification system were modified as “acute non-suppurated,” “acute suppurred,” and “subacute or suppurred-fistulized” apical peri-implantitis [52]. Based on the endodontic-implant infection pathway, two different infection patterns were “from implant to tooth (type I)” and “from tooth to implant (type II).” [53] Devitalization of adjacent tooth pulp occurs during osteotomy preparation in the first pathway while IPL occurs shortly after placement of the implant when adjacent tooth develops a periapical pathology in the second pathway [53]. To treat lesions based on the classification of IPL, the “PIST” system and the retrograde peri-implantitis (PRI) system were developed in recent years. The PIST system classifies the different pathological conditions around dental implants into “primarily periodontitis (P-class; P-1, P-2, P-3),” “primarily peri-implantitis (I-class; I-1, I-2, I-3),” “separately (S-class; S-1, S-2, S-3),” and traumatic lesions (T-class; T-0, T-1)” to determine the relationship between the periodontal, periapical, and peri-implant complications [54]. The “symptomatic T-1 lesion” is Sussman’s “type I infection pattern.” The PRI classification system was used in treatment planning and determination of prognosis in IPL based on the “bone loss/implant length” (in percentage) on periapical radiographs, which includes the following classes: mild—Class I (bone loss < 25%), moderate—Class II (bone loss 25% to 50%), and advanced—Class III (bone loss > 50%) [25,55]. Classes I and II lesions resolve completely; however, the prognosis of Class III lesions can range from fair to poor because the >50% bone loss may cause inadequate osseointegration [17,25]. Sarmast provided another classification in 2017 to aid decision-making for IPL: Class 1, implant placement induces devitalization of an adjacent previously vital tooth; Class 2, implant apex is infected by a peri-apical lesion on adjacent tooth/implant; Class 3, implant apex is outside the bone housing; and Class 4, implant apex lesion develops from residual infection of implant site. The endodontic treatment of involved tooth is suggested for Class 1 and Class 2 lesions per this classification. Endodontic evaluation of adjacent teeth before implant placement is helpful to prevent IPL [56].
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Figure 3. Classification systems of implant periapical lesion based on various parameters.

3.4. Treatment

Though many materials and methods are used for treating IPL, there is no evidence on which is the best [22,28,47,57–59]. If the IPL is inactive, such as a scar, regular follow-up is suggested. Elimination of infection is the only point of concensus in the treatment protocol for active IPL [60]. Requirement of endodontic therapy/surgery for teeth adjacent to implants must be examined [61,62]. In addition, medications, such as bisphosphonates, used by the patient should be reevaluated. The prognosis of IPL may be based on the extent of radiolucency [25]. Herein, treatment for active IPL is divided into three major modes (Figure 4).
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Figure 4. Treatment flowchart of implant periapical lesion. GBR, guided bone regeneration; S/S, symptoms and signs.

3.4.1. Nonsurgical Treatment

Antibiotics are usually used initially in active IPL to reduce the level of infection [3,17]. Systemic antibiotics might be used as adjuvant therapy, and definitive surgical intervention should be performed as early as possible to limit disease progression [3,17]. Antibiotic use alone is unlikely to be successful due to difficulties in eradicating bacterial colonies in IPL [19,21,59]. However, very few implants were treated successfully by non-surgical treatment alone, including one of the three implants with apical radiolucency in the patient under bisphosphonate medication [18,33,63,64]. Antibiotics are selected according to the etiology, presence or absence of pain and abscess, time of onset, and whether the endodontic lesion is open or closed [17]. Penicillin, amoxicillin, amoxicillin-clavulanate, metronidazole, and clindamycin are usually used to treat IPL; other drugs include azithromycin, cephalosporin, and tetracycline [6,7]. The combination of amoxicillin and metronidazole has also been used [5].

3.4.2. Surgical Treatment

This includes surgical debridement, regenerative treatment, resection of implant apex, and a combination of different approaches.

A surgical approach is chosen based on implant stability and the extension of IPL [1,17,47,55]. Surgical debridement alone or combined with regenerative treatment has been suggested for bone loss <50% of implant length [47]. Resection of the implant apex may be needed in cases with >50% bone loss and with difficulty in accessing the IPL [17,52,65]. However, maintenance of sufficient length be used after resection and keeping the implant unloaded for at least 6 months were found important for successful implant apicoectomy [20,66,67], and high success rates of implant apicoectomy have been reported [20,67,68].

The detoxication of implant surfaces is performed by chemical and physical methods. The universal implant Deplaquer (Straumann, USA) and an air-abrasive device with amino acid glycine powder were used for physical detoxication [69–71]. The chemical agents include chlorhexidine, tetracycline, citric acid, ethylenediaminetetraacetic acid (EDTA), and so on [65,72,73]. Moreover, calcium hydroxide is a better bacterial inhibitor than
Chlorhexidine for endodontic infections [27]. The use of lasers, such as Er,Cr:YSGG laser, is advantageous because of its bactericidal effect on oral pathogens and the ability to reach anatomically complex areas [71,74]. Moreover, laser supports rapid blood clot formation and graft containment in ridge preservation [75]. Therefore, laser may have the potential to detoxify the implant surface and apical lesion, and to improve the outcome of regenerative treatment in IPL [18]. The modes of bone regeneration used in the treatment of IPL and peri-implantitis are similar [76], and in critical size bone defects, the use of bone regeneration membrane or platelet rich fibrin (PRF) has been suggested [17,69,71,77]. However, it is difficult to gain re-osseointegration on a previously affected implant even with various regenerative materials and methods [3,17].

3.4.3. Removal of Failed Implants

In a recent review, Wiedemann concluded that mobile implants and/or more than 50% bone loss require removal of implant combined with immediate or delayed implant placement [59]. Removal of mobile teeth and new wider implant placement is only performed at non-infected sites. Bone regeneration after implant extraction and staged implant therapy is suggested at infected sites [17,46]. However, the exact etiology of implant failure should be confirmed and corrected before new implant placement, otherwise, implant failure may occur again [5,17].

4. Discussions

IPL is a preventable disease [56]. Hence, a detailed treatment plan, careful and sterile surgery, and regular follow-up would limit the incidence and extent of infection in active IPL, thereby reducing implant removal rate.

4.1. Detailed Treatment Plan

A thorough medical history, focusing on the possible influences on implant therapy, is paramount [8,33]. It is important to thoroughly evaluate the endodontic and periodontal condition of the extracted and adjacent teeth by clinical and radiographic examination [61]. However, it is difficult to detect histologic inflammation and infection in radiographs, even with CBCT [8,11]. Most of the infectious oral bacteria, such as streptococci, are susceptible to penicillin, and bacteria in periapical lesions are susceptible to metronidazole, cefoxitin, and clindamycin [27]. Therefore, antibiotic prophylaxis and postoperative use for 5–7 days may be considered in implant therapy, especially with a history of failed endodontic treatment [27,78]. Additionally, one way to reduce incidence of IPL could be through increasing the distance between implant and adjacent tooth as well as extending follow-up time after endodontic treatment of adjacent tooth [13]. Immediate implant placement in periapical infected sites remains controversial, as there is no consensus on appropriate surgical protocol and antibiotic prophylaxis [79–82]. Meticulous cleaning, thorough debridement/ curettage, and chemical irrigation (such as 0.12% chlorhexidine) prior to implant placement could improve implant survival rate at infected sites [18,83,84]. Currently, lasers, including Er-Cr:YSGG and Nd:YAG Laser, are used as adjuncts for the debridement after extraction, especially during immediate implant placement [85]. Laser has been demonstrated to be superior to chemical treatment [85]. To alleviate concerns of IPL, it may be helpful to decontaminate the socket after tooth extraction and to sterilize the site before implant placement with laser.

4.2. Careful and Sterile Surgery

This involves prevention of method-associated etiologic factors of IPL, by preventing contamination through sterile surgery, avoiding overheating of bone by adequate cooling and gentle drilling, avoiding compression of bone chip by optimal tightening, and accurate implant placement using a well-designed surgical stent [7]. The distance between the teeth and implants is crucial in preventing IPL [6]. In most cases, placement of implants did not affect the pulpal and apical condition of the adjacent teeth [86]. Less irrigation in the
apical region and intermittent drilling (pumping) for proper cooling, every 15 to 20 s, is suggested during deep preparation [24,28]. Nonetheless, method-associated factors may be more easily handled than patient-associated factors.

4.3. Regular Follow-Up

Avoiding premature loading after implant surgery has been suggested to prevent IPL [11,87]. Radiolucency surrounding implant apex usually appears between 7 and 16 days after implant placement; however, in some cases, radiolucency has appeared even up to 3 months post-surgery [6,9]. The mean period for the appearance of clinical symptoms of IPL was 51.83 days [16]. Moreover, a mean period of 21.7 days was reported for the diagnosis of early IPL after dental implant placement [14]. A periapical X-ray is suggested at implant placement and post-surgery 6–8 weeks before prosthetic treatment [16]. However, periapical radiography within one month after implant surgery may be suggested in some cases.

The survival rate in IPL varies between 73.2% to 97.4% [6]. The cumulative survival rate of 78.3% with a mean survival time of 85.4 months was recently reported [88]. Early detection of active IPL could increase implant survival rate by over 91% [6,9,89]. Therefore, it is very crucial to detect IPL as early as possible. In addition, the diagnostic stage of IPL and the previous periapical lesion at the implant site significantly affect the survival of implants after surgical treatment [88].

5. Conclusions

Implant periapical lesion, with a multifactorial etiology, is an infectious-inflammatory lesion surrounding the implant apex. To date, many methods are used for treating IPL, including non-surgical and surgical techniques. Detection of IPL as early as possible by regular follow-up is necessary to avoid implant failure and removal. However, prevention is the best treatment, and the incidence of IPL could be reduced by detailed examination before dental implant therapy and careful surgical technique. Furthermore, a comprehensive understanding of IPL, including its etiology and treatment, is necessary.

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