A Mac-2 Binding Protein Glycosylation Isomer-Based Risk Model Predicts Hepatocellular Carcinoma in HBV-Related Cirrhotic Patients on Antiviral Therapy

Chien-Hung Chen 1,* , Tsung-Hui Hu 1, Jing-Houng Wang 1, Hsueh-Chou Lai 2,3, Chao-Hung Hung 1, Sheng-Nan Lu 1 and Cheng-Yuan Peng 2,4,*

1 Division of Hepatogastroenterology, Department of Internal Medicine, Kaohsiung Chang Gung Memorial Hospital and College of Medicine, Chang Gung University, Kaohsiung 833253, Taiwan
2 Center for Digestive Medicine, Department of Internal Medicine, China Medical University Hospital, Taichung 40402, Taiwan
3 School of Chinese Medicine, China Medical University, Taichung 40402, Taiwan
4 School of Medicine, China Medical University, Taichung 40402, Taiwan
* Correspondence: e580306@ms31.hinet.net (C.-H.C.); cypeng@mail.cmuh.org.tw (C.-Y.P.); Tel.: +886-7-7317123 (C.-H.C.); +886-4-22052121 (ext. 2260) (C.-Y.P.); Fax: +886-7-7318762 (C.-H.C.); +886-4-22071600 (C.-Y.P.)

Simple Summary: Mac-2 binding protein glycosylation isomer (M2BPGi) has not been used in a risk score to predict hepatocellular carcinoma (HCC). We enrolled 1003 cirrhotic patients receiving entecavir or tenofovir monotherapy to construct an HCC risk score. The ASPAM-B score, based on age, sex, platelet count, AFP and M2BPGi at 12 months of treatment, was developed. The ASPAM-B scores accurately classified patients into low (0–3.5), medium (4–7) and high (>7) risk (p < 0.001). The values of AUROC for predicting 3-, 5- and 9-year risks of HCC were 0.742, 0.728 and 0.719, respectively. All AUROCs between the ASPAM-B and APA-B, PAGE-B, RWS-HCC and THRI scores at 3–9 years were significantly different. The M2BPGi-based risk model exhibited good discriminant function in predicting HCC in cirrhotic patients who received antiviral treatment.

Abstract: Mac-2 binding protein glycosylation isomer (M2BPGi) has not been used in a risk score to predict hepatocellular carcinoma (HCC). We enrolled 1003 patients with chronic hepatitis B and cirrhosis receiving entecavir or tenofovir therapy for more than 12 months to construct an HCC risk score. In the development cohort, Cox regression analysis identified male gender, age, platelet count, AFP and M2BPGi levels at 12 months of treatment as independent risk factors of HCC. We developed the HCC risk prediction model, the ASPAM-B score, based on age, sex, platelet count, AFP and M2BPGi levels at 12 months of treatment, with the total scores ranging from 0 to 11.5. This risk model accurately classified patients into low (0–3.5), medium (4–7), and high (>7) risk in the development and validation groups (p < 0.001). The areas under the receiver operating characteristic curve (AUROC) of 3-, 5- and 9-year risks of HCC were 0.742, 0.728 and 0.719, respectively, in the development cohort. All AUROCs between the ASPAM-B and APA-B, PAGE-B, RWS-HCC and THRI scores at 3–9 years were significantly different. The M2BPGi-based risk model exhibited good discriminant function in predicting HCC in cirrhotic patients who received long-term antiviral treatment.

Keywords: AFP; cirrhosis; hepatocellular carcinoma; M2BPGi; risk model

1. Introduction

Long-term treatment with nucleos(t)ide analogues (NA) could reduce rates of cirrhotic complications, hepatocellular carcinoma (HCC), and total or liver-related mortality [1,2]. Although NA treatment could reduce the rate of HCC, it does not eliminate its development, especially in cirrhotic patients [1,2]. In recent years, some risk prediction models of HCC in
patients with chronic hepatitis B (CHB) who received long-term NA treatment have been developed [3–9]. Notably, a recent study from the United States compared the predictive performance of 10 risk prediction models and demonstrated that APA-B, AASL-HCC, REAL-B and RWS-HCC exhibited an area under the receiver operating characteristic curve (AUROC) of >0.80 for predicting 3-year HCC risk [4,6,7,10–15]. However, these risk scores were developed from a mixed population with or without cirrhosis. Further validation of these risk models is warranted to determine their clinical utility in cirrhotic patients.

The *Wisteria floribunda* agglutinin (WFA)-positive Mac-2 binding protein glycosylation isomer (M2BPGi), a secreted glycoprotein from hepatic stellate cells (HSCs) in the serum and extracellular matrix, can induce the expression of Mac-2 protein in Kupffer cells, which in turn activates HSCs and increases alpha-smooth muscle actin expression [16]. In recent years, it has been demonstrated that serum M2BPGi levels correlate with the stage of liver fibrosis in patients with CHB and that serum M2BPGi level is a useful marker of HCC in CHB patients receiving NA therapy and a predictor of recurrence and prognosis in patients with HCC undergoing curative resection [17–22].

The aim of this study was to study the predictive role of serum M2BPGi for HCC occurrence and developed a new, M2BPGi-based risk model of HCC in a cohort of CHB patients with cirrhosis receiving entecavir or tenofovir disoproxil fumarate (TDF) treatment.

2. Materials and Methods

2.1. Patients

This study retrospectively enrolled a cohort of 689 CHB patients with cirrhosis who received entecavir treatment between 2008 and 2018, and 314 CHB patients with cirrhosis who received TDF treatment between 2011 and 2018. The patients were included from China Medical University Hospital (n = 337) and Kaohsiung Chang Gung Memorial Hospital (n = 666). In Taiwan, the costs of entecavir and TDF have been reimbursed for hepatitis B virus (HBV) treatment by Taiwan’s National Health Plan since 2008 and 2011, respectively. The inclusion criteria were: (1) age >18 years and hepatitis B surface antigen (HBsAg) was positive for more than 6 months before NA therapy; (2) entecavir or TDF monotherapy for at least 12 months before enrollment; (3) all patients fulfilled the diagnosis of cirrhosis either liver histology (n = 210) or cirrhosis was suggested by repeated ultrasounds and clinical features, such as gastroesophageal varices, splenomegaly, thrombocytopenia or ascites. The exclusion criteria were: (1) evidence of alcoholic liver disease, autoimmune hepatitis, or coinfection with hepatitis C virus (HCV), hepatitis D virus or human immunodeficiency virus; (2) HCC or liver transplantation at baseline or within the first year of NA therapy.

All enrolled patients were randomly assigned to the models of development or validation group in a 2:1 ratio to construct prediction model of HCC. The clinical parameters at baseline and 12 months of treatment were used to construct the HCC prediction model in the development cohort, and validation cohort were used to examine its predictive performance.

2.2. Methods

All patients received NA therapy with a median duration of 72 (12–172) months. During NA therapy, all patients were followed up every 1 to 3 months. Serum HBV DNA and alanine aminotransferase (ALT) levels and were checked at baseline, every 3 to 6 months during NA treatment, and at the time of biochemical breakthrough. All enrolled patients were followed until discontinuation of entecavir or TDF treatment or the last visit. HCC surveillance was implemented using serum alpha-fetoprotein (AFP) and abdominal ultrasonography every 3 months. HCC was diagnosed according to the practice guidance of the American Association for the Study of Liver Diseases [23].

2.3. Definitions

Diabetes mellitus (DM) was diagnosed according to the previous guideline [24]. Patients were also considered diabetic according to their medical history or if they had
received insulin treatment or oral hypoglycemic agents. Hypertension was diagnosed according to the medical history or having received anti-hypertensive drugs. Cirrhotic events were defined as new developments of hepatic encephalopathy, variceal bleeding, or ascites in patients without hepatic decompensation at the initiation of NA treatment.

2.4. Measurement of WFA-Positive M2BPGi

Serum WFA-positive M2BP (M2BPGi) level was measured based on a lectin-antibody sandwich immunoassay using the fully automatic immunoanalyzer, HISCL-2000i (Sysmex, Hyogo, Japan) [25]. The values of M2BPGi were expressed as cut-off index (COI) [25].

2.5. Serology

Serum HBV DNA was quantified by the COBAS AmpliPrep/COBAS TaqMan HBV test with a detection limit of 20 IU/mL. Hepatitis B core-related antigen (HBcrAg) levels were quantified using the Chemiluminescent Enzyme Immunoassay (CLEIA) system on a Lumipulse CLEIA analyzer (Fujirebio Inc., Tokyo, Japan) following the manufacturer’s instructions [26]. The automated estimation range was from 3 to 7 log U/mL. HBcrAg levels below 3 log U/mL were taken as 3 log_{10} U/mL for statistical analysis.

2.6. Statistical Analysis

The cumulative incidences of HCC, cirrhotic events and liver-related mortality were calculated by Kaplan–Meier method with the log-rank test. The risk factors of HCC occurrence, cirrhotic events and mortality were determined by Cox proportional hazards regression model. Missing data were assumed to be missing at random and were replaced with substituted values by multiple imputation [27]. The HCC risk scoring system and HCC risk was established by Cox proportional hazards regression model and the method has been previously described [6,28]. The HCC risk was estimated with the equation: 1–P_0 \exp(\sum \beta_1 \times \text{score}–\sum \beta_i \times M_i). The model discrimination was assessed with area under AUROC curves. AUROCs were calculated by time-dependent ROC curves for accessing the performance of the risk models for each year and used C-statistic to assess the performance of the risk model. The model calibration was compared by Hosmer–Lemeshow goodness-of-fit test between expected and observed rates of HCC in the development group. The time-dependent ROC, C-statistics, and comparisons of these values between two risk scores were performed using the timeROC package [29,30]. A two-sided p value of < 0.05 was considered statistically significant.

3. Results

3.1. Comparison of Clinical Characteristics of All Patients with or without HCC Development

Table 1 compares the clinical features of patients with or without HCC development. We select the variables associated with HCC for analysis according to previous studies [4,6,7,10,11]. Patients with HCC development were older and higher percentages of them were male and more likely had hepatic decompensation and hypertension than those without HCC development. They also had lower albumin levels and platelet counts and higher M2BPGi and HBcrAg levels than those without HCC development.

Table 1. Baseline clinical characteristics of patients with HCC or without HCC development.

| Variables             | HCC n = 183 | No HCC n = 820 | p Value |
|-----------------------|-------------|----------------|---------|
| Age (year)            | 58.1 ± 10.0 | 53.0 ± 12.0    | <0.001  |
| Sex, male             | 147 (80.3%) | 599 (73.0%)    | 0.041   |
| Entecavir versus TDF  | 147 vs. 36  | 542 vs. 278    | <0.001  |
| HBeAg-positive status | 50 (27.3%)  | 197 (24.0%)    | 0.349   |
| Decompensation status | 49 (26.8%)  | 140 (17.1%)    | 0.002   |
| NA-naïve              | 153 (83.6%) | 696 (84.9%)    | 0.666   |
The clinical features of the development and validation cohorts were presented in Supplementary Table S1 and were similar between two cohorts. The rates of HCC development were 10% versus 9.1% at 3 years, 15% versus 14.3% at 5 years and 25.8% versus 26.1% at 10 years in the development (Figure 1).

### Table 1. Cont.

| Variables                  | HCC  | No HCC | p Value |
|----------------------------|------|--------|---------|
| Diabetes mellitus, yes     | 45 (24.6%) | 176 (21.5%) | 0.356   |
| Hypertension, yes          | 61 (33.3%) | 202 (24.6%) | 0.016   |
| HBV DNA, log10 IU/mL      | 5.53 ± 1.52 | 5.40 ± 1.51 | 0.318   |
| AST, U/L                  | 119.1 ± 205.4 | 130.5 ± 269.8 | 0.589   |
| ALT, U/L                  | 134.2 ± 259.2 | 162.3 ± 362.8 | 0.321   |
| Total bilirubin, mg/dL    | 2.09 ± 3.75 | 2.03 ± 3.78 | 0.854   |
| INR                       | 1.19 ± 0.22 | 1.19 ± 0.28 | 0.992   |
| Albumin, g/dL             | 3.80 ± 0.64 | 4.02 ± 0.63 | <0.001  |
| Platelet, ×10^3/μL        | 121.0 ± 56.0 | 138.7 ± 56.0 | <0.001  |
| AFP, ng/mL                | 32.2 ± 82.3 | 30.2 ± 123.9 | 0.840   |
| M2BPGi, COI               | 3.80 ± 3.98 | 2.77 ± 3.50 | <0.001  |
| HBeAg, log10 IU/mL        | 5.33 ± 1.49 | 5.07 ± 1.48 | 0.034   |

Abbreviations: AFP, alpha-fetoprotein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; COI, cut-off index; HBeAg, hepatitis B e antigen; HCC, hepatocellular carcinoma; INR, international normalized ratio; M2BPGi, Mac-2 binding protein glycosylation isomer; NA, nucleos(t)ide analogue.

### 3.2. HCC Risk Predictors and Prediction Model of the Development Group

In the entire cohort, 183 subjects developed HCC during a median follow-up duration of 72 (12–172) months (6153.73 person years). The cumulative HCC incidences at 3, 5, and 10 years were 9.5%, 14.8%, and 25.8%, respectively.

The clinical features of the development and validation cohorts were presented in Supplementary Table S1 and were similar between two cohorts. The rates of HCC development were 10% versus 9.1% at 3 years, 15% versus 14.3% at 5 years and 25.8% versus 26.1% at 10 years in the development (n = 668) and validation groups (n = 335), respectively (p = 0.845) (Figure 1).

A multivariate analysis showed that sex and age, platelet count, AFP and M2BPGi levels at 12 months of treatment were the independent predictors associated with HCC in the development group (Table 2). The HCC risk prediction model was constructed, based on age, sex, platelet count, M2BPGi and AFP levels at 12 months of treatment, to develop the risk score, named as ASPAM-B (Table 3). We converted the regression coefficients of the independent risk factors to compute integer risk scores (Table 3). HCC predictive risk scores after 2–10 years of ETV or TDF therapy were listed in Supplementary Table S2. Figure 2 shows the nomograms of 3-, 5-, 7- and 9-year risk for hepatocellular carcinoma for this model.
Table 2. The risk predictors of hepatocellular carcinoma in the development cohort.

| Variables                             | Univariate Analysis | Multivariate Analysis |
|---------------------------------------|---------------------|-----------------------|
|                                      | HR (95% CI)         | p Value               |
| Baseline                              |                     |                       |
| Age (per year)                        | 1.034 (1.018–1.056) | <0.001                |
| Sex, male vs. female                  | 1.458 (0.927–2.295) | 0.103                 |
| HBV DNA, per log_{10} IU/mL           | 0.951 (0.705–1.566) | 0.808                 |
| Decompensation, yes vs. no            | 1.883 (1.263–2.810) | 0.002                 |
| NA-naïve, yes vs. no                  | 0.843 (0.530–1.340) | 0.470                 |
| TDF vs. entecavir                      | 0.634 (0.398–1.012) | 0.056                 |
| Diabetes mellitus, yes vs. no         | 1.662 (1.151–2.400) | 0.007                 |
| HBV DNA, per log_{10} IU/mL           | 0.999 (0.998–1.000) | 0.212                 |
| ALT, per U/L                          | 0.999 (0.999–1.000) | 0.120                 |
| Total bilirubin, per mg/dL            | 0.998 (0.955–1.044) | 0.944                 |
| Albumin, per g/L                      | 0.627 (0.484–0.812) | <0.001                |
| INR, per ratio                        | 0.901 (0.464–1.750) | 0.758                 |
| Platelet, per 10^{3}/µL               | 0.993 (0.990–0.997) | <0.001                |
| AFP, per ng/mL                        | 1.001 (0.999–1.003) | 0.375                 |
| M2BPGi, per COI                       | 1.054 (1.013–1.098) | 0.010                 |
| HBcrAg, per log_{10} U/mL             | 1.066 (0.942–1.206) | 0.312                 |

12 months of treatment

| Variables                             | HR (95% CI)         | p Value               |
|---------------------------------------|---------------------|-----------------------|
| Age (year)                            | 1.034 (1.018–1.050) | <0.001                |
| ALT < 40 U/L, per U/L                 | 0.685 (0.468–1.001) | 0.051                 |
| AFP, per ng/mL                        | 1.009 (1.004–1.015) | <0.001                |
| Platelet, per 10^{3}/µL               | 0.992 (0.988–0.995) | <0.001                |
| M2BPGi, per COI                       | 1.123 (1.069–1.180) | <0.001                |
| HBcrAg, per log_{10} U/mL             | 1.085 (0.941–1.252) | 0.262                 |

Figure 2. Nomograms for the prediction of the risk of hepatocellular carcinoma development.
Table 3. The risk scores of hepatocellular carcinoma in the development cohort.

| Variables                  | HR (95% CI)          | Parameter | p Value  | Risk Scores |
|----------------------------|----------------------|-----------|----------|-------------|
| Age at 12 months, years    |                      |           |          |             |
| <40                        | 1.532 (1.297–1.809)  | 0.4265    | <0.0001  |             |
| 40–49                      |                      |           |          |             |
| 50–59                      | 2.164 (1.356–3.452)  | 0.7718    | 0.0012   | 2           |
| 60–69                      |                      |           |          |             |
| ≥70                        |                      |           |          |             |
| Sex                       | 1.000                |           |          |             |
| Female                    |                      |           |          |             |
| Male                      | 2.164 (1.356–3.452)  | 0.7718    | 0.0012   | 2           |
| Platelet at 12 months, 10^3/µL |                |           |          |             |
| ≥80                        | 1.779 (1.170–2.706)  | 0.5760    | 0.0071   | 1.5         |
| <80                        |                      |           |          |             |
| AFP at 12 months, ng/mL    | 2.264 (1.406–3.645)  | 0.8170    | 0.0008   | 2           |
| <9                         | 1.000                | 0         |          |             |
| >9                         |                      |           |          |             |
| M2BPGi at 12 months, COI   |                      |           |          |             |
| <1.0                      | 1.000                |           | 0        |             |
| 1.0–2.6                   | 1.904 (1.222–2.967)  | 0.6441    | 0.0044   | 1.5         |
| >2.6                      | 2.163 (1.336–3.500)  | 0.7714    | 0.0017   | 2           |

AFP, alpha-fetoprotein; CI, confidence interval; COI, cut-off index; HR: hazard ratio; M2BPGi, Mac-2 binding protein glycosylation isomer.

In this model, the total risk scores ranged from 0 to 11.5. The C-statistic of the model was 0.716 (0.665–0.768). The calibration of the model revealed a good model fit (p = 0.5661).

We categorized the ASPAM-B score into three subgroups according to the HCC incidence: ≤3.5, 4–7 and >7, respectively. The cumulative HCC rates at 8 years of treatment in the three subgroups were 9.0%, 23.5% and 57.9%, respectively (p < 0.001, Figure 3A).

3.3. Validation of the HCC Risk Prediction Model

According to ASPAM-B score, validation cohort was also categorized into low (≤3.5), medium (4–7), and high (>7) risk. The cumulative HCC rates at 5 years of treatment in the three subgroups were 5.6%, 24.8% and 44.6%, respectively (p < 0.001, Figure 3B). The C-statistic of this risk model was 0.714 (0.647–0.782).

3.4. Comparisons of AUROC and C-Statistic between Different Prediction Models of HCC

In the development cohort, the AUROCs for predicting 3-, 5-, 7- and 9-year risks of HCC were 0.742, 0.728, 0.721 and 0.719, respectively, based on ASPAM-B score. The AUROCs for predicting 3-, 5-, 7- and 9-year risks of HCC for the APA-B score [6], PAGE-B score [5], RWS-HCC score [14], AASL-HCC score [10] and Toronto HCC risk index (THRI) [31] are shown in Table 4. All AUROCs between the ASPAM-B and the APA-B, PAGE-B, RWS-HCC and THRI scores at 3–9 years were significantly different (Supplementary Table S3).
Figure 3. Cumulative incidence rate of HCC according to the ASPAM-B risk score in the (A) development and (B) validation cohorts.

In the development cohort, the C-statistics of the models of ASPAM-B, APA-B, PAGE-B, RWS-HCC, AASL-HCC and THRI were 0.716 (95% confidence interval [CI]: 0.665–0.768), 0.659 (0.607–0.712), 0.671 (0.620–0.721), 0.618 (0.562–0.673), 0.651 (0.600–0.702), and 0.664 (0.613–0.715), respectively. The ASPAM-B had higher values of C-statistic than APA-B ($p = 0.011$), PAGE-B ($p = 0.050$), RWS-HCC ($p = 0.0014$), AASL-HCC ($p = 0.0059$) and THRI ($p = 0.020$).
Table 4. The values of AUROCs for predicting hepatocellular carcinoma according to different risk models.

| Development Cohort (n = 668) | ASPAM-B AUROC (95% CI) | APA-B AUROC (95% CI) | PAGE-B AUROC (95% CI) | RWS-HCC AUROC (95% CI) | AASL-HCC AUROC (95% CI) | THRI AUROC (95% CI) |
|-----------------------------|-------------------------|----------------------|-----------------------|------------------------|------------------------|----------------------|
| 3 years                     | 0.742 (0.672–0.811)     | 0.661 (0.588–0.734)  | 0.673 (0.601–0.746)   | 0.601 (0.525–0.677)    | 0.677 (0.611–0.744)    | 0.660 (0.590–0.731)   |
| 5 years                     | 0.728 (0.668–0.788)     | 0.669 (0.610–0.729)  | 0.676 (0.616–0.736)   | 0.604 (0.540–0.668)    | 0.654 (0.594–0.714)    | 0.663 (0.604–0.722)   |
| 7 years                     | 0.721 (0.665–0.777)     | 0.668 (0.612–0.724)  | 0.667 (0.611–0.723)   | 0.606 (0.546–0.665)    | 0.644 (0.588–0.701)    | 0.650 (0.593–0.706)   |
| 9 years                     | 0.719 (0.666–0.772)     | 0.667 (0.614–0.721)  | 0.671 (0.617–0.724)   | 0.614 (0.556–0.673)    | 0.651 (0.598–0.704)    | 0.656 (0.603–0.710)   |

AUROC, the area under the receiver operating characteristic curve; CI, confidence interval.

3.5. Incidences and Predictors of Cirrhotic Events

Among the 814 patients with compensated cirrhosis at baseline, 44 experienced cirrhotic events during treatment, of which 28, 22, and 5 developed ascites, variceal bleeding, and hepatic encephalopathy, respectively. The cumulative incidences of cirrhotic events at 3, 5, and 10 years were 2.9%, 5%, and 8.6%, respectively. A multivariate analysis revealed that lower albumin levels, lower platelet count and higher M2BPGi levels at 12 months of treatment were independent risk factors for cirrhotic events (Table 5). An M2BPGi level of 1.2 COI at 12 months of treatment was the optimal value for predicting cirrhotic events within 10 years (AUROC: 0.819) by time-dependent ROC curve. The 10-year cumulative incidence of cirrhotic events in patients with the M2BPGi level ≤ 1.2 and >1.2 COI were 3.3% and 18.5%, respectively (p < 0.001) (Figure 4A).

Table 5. Univariate and multivariate analyses of factors associated with hepatic events (new events of variceal bleeding, ascites and hepatic encephalopathy) in patients without decompensated cirrhosis at baseline.

| Variables                          | Baseline Hazard Ratio (95% CI) | p Value | Multivariate Analysis Hazard Ratio (95% CI) | p Value |
|------------------------------------|--------------------------------|---------|-------------------------------------------|---------|
| Age (year)                         | 1.005 (0.980–1.031)           | 0.680   |                                           |         |
| Sex, male vs. female               | 1.162 (0.574–2.353)           | 0.677   |                                           |         |
| HBeAg, yes vs. no                  | 1.087 (0.560–2.111)           | 0.805   |                                           |         |
| NA-naïve, yes vs. no               | 2.592 (0.802–8.372)           | 0.111   |                                           |         |
| TDF vs. entecavir                  | 1.030 (0.534–1.986)           | 0.930   |                                           |         |
| Diabetes mellitus, yes vs. no       | 1.101 (0.544–2.229)           | 0.789   |                                           |         |
| Hypertension, yes vs. no           | 1.284 (0.680–2.422)           | 0.441   |                                           |         |
| HBV DNA, per log_{10} IU/mL        | 0.965 (0.790–1.179)           | 0.729   |                                           |         |
| AST, per U/L                       | 1.000 (0.999–1.002)           | 0.614   |                                           |         |
| ALT, per U/L                       | 0.998 (0.994–1.001)           | 0.215   |                                           |         |
| Total bilirubin, per mg/dL         | 1.068 (0.844–1.353)           | 0.582   |                                           |         |
| Albumin, per g/L                   | 0.262 (0.154–0.448)           | <0.001  |                                           |         |
| INR, per ratio                     | 1.328 (0.412–4.288)           | 0.635   |                                           |         |
| Platelet, per 10^3/μL              | 0.983 (0.976–0.990)           | <0.001  |                                           |         |
| AFP at baseline, per ng/mL         | 0.996 (0.985–1.006)           | 0.412   |                                           |         |
| M2BPGi, per COI                    | 1.180 (1.082–1.288)           | <0.001  |                                           |         |
| HBcrAg, per log_{10} U/mL          | 0.987 (0.804–1.211)           | 0.900   |                                           |         |
| 12 months of treatment             |                                 |         |                                           |         |
| ALT < 40 U/L, per U/L              | 0.536 (0.292–0.984)           | 0.044   |                                           |         |
Table 5. Cont.

| Variables            | Univariate Analysis |                  |                  | Multivariate Analysis |                  |                  |
|----------------------|---------------------|------------------|------------------|-----------------------|------------------|------------------|
|                      | Hazard Ratio (95% CI) | p Value          | Hazard Ratio (95% CI) | p Value              | Hazard Ratio (95% CI) | p Value |
| AFP, per ng/mL       | 1.011 (1.002–1.020)  | 0.016            | 0.986 (0.979–0.994) | 0.001                | 0.986 (0.979–0.994) | 0.001 |
| Platelet, per 10^3/µL| 0.980 (0.973–0.987)  | <0.001           | 0.439 (0.283–0.679) | <0.001               | 0.986 (0.979–0.994) | 0.001 |
| Albumin, per g/L     | 0.280 (0.197–0.398)  | <0.001           | 0.439 (0.283–0.679) | <0.001               | 0.986 (0.979–0.994) | 0.001 |
| M2BPGi, per COI      | 1.349 (1.241–1.466)  | <0.001           | 1.135 (1.026–1.256) | 0.014                | 0.986 (0.979–0.994) | 0.001 |
| HBcrAg, per log_{10} U/mL | 1.078 (0.853–1.363) | 0.531            |                  |                      |                  |                  |

AFP, alpha-fetoprotein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; CI, confidence interval; COI, cut-off index; HBcrAg, hepatitis B core related antigen; HBeAg, hepatitis B e antigen; HBV, hepatitis B virus; INR, international normalized ratio; M2BPGi, Mac-2 binding protein glycosylation isomer; NA, nucleos(t)ide analogue; TDF, Tenofovir disoproxil fumarate.

Figure 4. Cumulative incidences of (A) cirrhotic events and (B) liver-related mortality or liver transplantation according to M2BPGi levels at 12 months of treatment.
3.6. Incidences and Predictors of Liver-Related Mortality or Liver Transplantation

In the entire cohort, 62 patients developed liver-related mortality during treatment, including 19 patients who underwent liver transplantation. The cumulative incidences of liver-related mortality or liver transplantation at 3, 5, and 10 years were 1.8%, 5.2%, and 10.6%, respectively. A multivariate Cox regression analysis revealed that lower albumin levels, lower platelet count and higher M2BPGi levels at 12 months of treatment were independent predictors for liver-related mortality or liver transplantation (Table 6). AFP tumor biomarker or AST/ALT metabolism makers at baseline or 12 months of treatment were not independent factors of liver related mortality or liver transplantation.

Table 6. Univariate and multivariate analyses of factors associated with liver related mortality or liver transplantation.

| Variables             | Univariate Analysis | Multivariate Analysis |
|-----------------------|---------------------|-----------------------|
|                       | Hazard Ratio (95% CI) | p Value | Hazard Ratio (95% CI) | p Value |
| **Baseline**          |                     |         |                     |         |
| Age (year)            | 1.010 (0.988–1.031)  | 0.377    |                     |         |
| Sex, male vs. female  | 1.096 (0.613–1.960)  | 0.757    |                     |         |
| HBeAg, yes vs. no     | 1.084 (0.620–1.894)  | 0.778    |                     |         |
| Decompensation, yes vs. no | 5.519 (3.352–9.087) | <0.001  |                     |         |
| NA-naive, yes vs. no  | 1.550 (0.706–3.404)  | 0.275    |                     |         |
| TDF vs. entecavir     | 0.651 (0.346–1.228)  | 0.185    |                     |         |
| Diabetes mellitus, yes vs. no | 1.210 (0.676–2.164) | 0.521    |                     |         |
| Hypertension, yes vs. no | 0.958 (0.542–1.692) | 0.882    |                     |         |
| HBV DNA, per log_{10} IU/mL | 0.948 (0.806–1.117) | 0.525    |                     |         |
| AST, per U/L          | 0.999 (0.997–1.001)  | 0.190    |                     |         |
| ALT, per U/L          | 0.997 (0.994–1.000)  | 0.041    |                     |         |
| Total bilirubin, per mg/dL | 1.033 (0.983–1.085) | 0.204    |                     |         |
| Albumin, per g/L      | 0.303 (0.239–0.419)  | <0.001   |                     |         |
| INR, per ratio        | 1.983 (1.078–3.650)  | 0.028    |                     |         |
| Platelet, per 10^{3}/µL | 0.984 (0.978–0.989) | <0.001   |                     |         |
| AFP at baseline, per ng/mL | 0.999 (0.994–1.003) | 0.491    |                     |         |
| M2BPGi, per COI       | 1.137 (1.087–1.189)  | <0.001   |                     |         |
| HBcAg, per log_{10} U/mL | 0.987 (0.821–1.166) | 0.806    |                     |         |
| **12 months of treatment** |  |  |                     |         |
| ALT < 40 U/L, per U/L | 0.763 (0.441–1.321)  | 0.335    |                     |         |
| AFP, per ng/mL        | 1.011 (1.003–1.019)  | 0.011    |                     |         |
| Platelet, per 10^{3}/µL | 0.984 (0.978–0.989) | <0.001   | 0.992 (0.986–0.998) | 0.009   |
| Albumin, per g/L      | 0.280 (0.197–0.398)  | <0.001   | 0.350 (0.250–0.490) | <0.001  |
| M2BPGi, per COI       | 1.239 (1.175–1.306)  | <0.001   | 1.083 (1.012–1.159) | 0.021   |
| HBcAg, per log_{10} U/mL | 1.156 (0.944–1.416) | 0.160    |                     |         |

AFP, alpha-fetoprotein; ALT, alanine aminotransferase; AST, aspartate aminotransferase; CI, confidence interval; COI, cut-off index; HBcAg, hepatitis B core related antigen; HBeAg, hepatitis B e antigen; HBV, hepatitis B virus; INR, international normalized ratio; M2BPGi, Mac-2 binding protein glycosylation isomer; NA, nucleos(t)ide analogue; TDF, Tenofovir disoproxil fumarate.

We used an M2BPGi level of 1.2 COI as the optimal cutoff value. The 10-year cumulative incidences of liver-related mortality or liver transplantation in patients with the M2BPGi level ≤ 1.2 and >1.2 COI were 5.4% and 17.5%, respectively (p < 0.001) (Figure 4B).

4. Discussion

Our study demonstrated that the cumulative HCC rates at 3, 5, and 10 years were 9.5%, 14.8%, and 25.8%, respectively, among patients with CHB and cirrhosis undergoing entecavir or TDF treatment. In the development group, we constructed the ASPAM-B risk score on the basis of age, sex, platelet, AFP and M2BPGi levels at month 12 of treatment, with the total scores ranging of 0 to 11.5. The ASPAM-B score predicted HCC risk over 2
to 9 years with an overall C-statistic of 0.716, which was significantly higher than those of the APA-B, PAGE-B, RWS-HCC, AASL-HCC and THRI, and stratified patients into three subgroups with distinct HCC risks. Furthermore, the stratified risk scores could be verified accurately in the validation cohort (p < 0.001).

To date, no risk models were specifically developed to predict HCC in HBV-related patients with cirrhosis, particularly those who have been receiving long-term NA treatment. It would be desirable if one can develop an HBV-specific risk model which enables us to stratify the risk of HCC among such population.

To our knowledge, at least 10 risk models have been developed to predict the risk of HCC in CHB patients receiving NA therapy [5–15,31,32]. All of them were developed from a mixed population with or without cirrhosis (19.1–50.2% with cirrhosis). A recent cohort study from the United States demonstrated that three models (APA-B, REAL-B and AASL-HCC) developed from patients receiving NA treatment and one model (RWS-HCC) developed from predominantly treatment-naïve patients exhibited the highest AUROCs (all > 0.80) for predicting 3-year HCC risk [6,10,11,14,15]. Common parameters of these four risk models such as age, cirrhosis or platelet count and AFP are documented risk factors for HCC. A recent Korean study revealed that cirrhosis at baseline, platelet count and AFP at 12 months of NA treatment were the optimal predictive factors for HCC in CHB patients receiving entecavir or TDF treatment [13]. A study from Toronto enrolled all cirrhotic patients with mixed etiology showed that age, sex, etiology and platelets were associated with HCC [31]. In the current study, we found that age,sex, platelet count and AFP level at 12 months of treatment were independent predictors associated with HCC occurrence in the development cohort of cirrhotic patients, similar to what we demonstrated in a prior study cohort (APA-B) which comprised only 36% cirrhotic patients [6]. However, the discriminant performance of APA-B for HCC occurrence was less satisfactory in the cirrhotic cohort compared to the mixed population with or without cirrhosis (C-statistic: 0.659 versus 0.850) [6]. Further effort in identification of novel biomarkers into the APA-B model to improve its performance in cirrhotic patients is warranted.

Serum M2BPGi levels correlate with the liver fibrosis stage in CHB patients and could predict HCC occurrence in CHB patients receiving NA treatment [17–22]. Given that platelet count shows an inverse correlation with the hepatic venous pressure gradient. It remains to be elucidated whether M2BPGi can serve as a biomarker of HCC risk independently of platelet in cirrhotic patients receiving NA treatment [33,34]. We demonstrated that both M2BPGi level and platelet count at 12 months of NA treatment, rather than at baseline, were independent predictors of HCC occurrence. Incorporation of sex and M2BPGi into the APA-B risk model generated the ASPAM-B model, which improved the discriminative performance for HCC occurrence and yielded significantly higher AUROC values for predicting 3-, 5-, 7- and 9-year HCC risks than did the APA-B model (all p < 0.05). The ASPAM-B model also outperformed the PAGE-B, RWS-HCC, AASL-HCC and THRI models with a significantly higher C-statistic for HCC risk prediction. The current demonstration of its predictive role for HCC occurrence independent of platelet in cirrhotic patients suggests that M2BPGi may act in additional unrecognized pathways related to hepatocarcinogenesis, rather than simply serve as a surrogate marker of liver fibrosis. Moreover, in addition to the liver fibrosis stage, serum M2BPGi levels correlate with the degree of hepatic inflammation in patients with CHB [17]. M2BPGi levels at 12 months of treatment may reliably reflect the actual liver fibrosis stage. A previous study also revealed that M2BPGi levels at 12 months of treatment correlated better with future HCC risk compared to the baseline measurement [20]. Thus, we propose that 12 months after NA treatment may represent an ideal time point for future refinement of the optimal risk model to predict HCC occurrence in patients with CHB receiving long-term NA treatment.

Serum HBcrAg reflects the levels of intrahepatic covalently closed circular DNA transcripational activity [26]. Baseline or on-treatment HBcrAg levels were reported to predict HCC occurrence in CHB patients receiving NA treatment [35]. In the current study,
serum HBcAg level at baseline or 12 months of NA treatment was not a risk factor of HCC occurrence in the entire cohort.

In addition to HCC, serum M2BPGi level and platelet count at 12 months of treatment also predicted cirrhotic events, liver-related mortality or liver transplantation in patients receiving NA treatment. This finding could be reconciled by the fact that M2BPGi and platelet may reflect the severity of underlying cirrhosis or hepatocyte dysfunction.

The current study has some limitations to note. First, the diagnosis of cirrhosis was confirmed by histology only in 210 patients of the entire cohort. Patients with early cirrhosis might have been excluded from this study. Second, the present model was developed from Asian patients who acquired HBV genotype B or C infection during neonatal period. External validation of the risk model with patients of different ethnicities or HBV genotypes are required to further verify its discriminant performance for HCC occurrence. Third, because we only measured serum M2BPGi levels at 12 months of NA treatment, we could only address the predictive role of serum M2BPGi level at 12 months of treatment in the subsequent risk of HCC in this study. We will explore the predictive role of serum M2BPGi levels at later time points during NA treatment in the subsequent risk of HCC in future studies. Fourth, despite that ASPAM-B exhibited the highest C-statistic for HCC occurrence in cirrhotic patients among all available risk models, its performance was moderate. Future efforts should be directed toward implementing novel biomarkers to facilitate early prediction and diagnosis of HCC in cirrhotic patients.

5. Conclusions

The M2BPGi-based ASPAM-B risk model exhibited good discriminant function in predicting HCC occurrence and stratified HCC risk in cirrhotic patients who received long-term NA treatment. Risk stratification of HCC occurrence in these patients may assist clinicians with individualization of HCC surveillance program in the antiviral therapy era. In addition to HCC, M2BPGi level at 12 months of treatment was a useful marker to predict cirrhotic events and liver-related mortality in cirrhotic patients receiving NA treatment.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/cancers14205063/s1, Table S1: Baseline characteristics in the development and validation groups; Table S2: Total risk scores predict the rates of 2–10-year hepatocellular carcinoma in the development cohort; Table S3: P values of AUROC comparisons between the ASPAM-B score and each risk score in the development group.

Author Contributions: Conceptualization, C.-H.C. and C.-Y.P.; Methodology, C.-H.C. and C.-Y.P.; Validation, C.-H.C. and C.-Y.P.; Formal Analysis, C.-H.C. and C.-Y.P.; Investigation, C.-H.C. and C.-Y.P.; Resources, C.-H.C. and C.-Y.P.; Data Curation, C.-H.C., T.-H.H., J.-H.W., H.-C.L., C.-H.H., S.-N.L., and C.-Y.P.; Writing—Original Draft Preparation, C.-H.C. and C.-Y.P.; Writing—Review and Editing, C.-H.C. and C.-Y.P.; Supervision, C.-H.C. and C.-Y.P.; Project Administration, C.-H.C.; Funding Acquisition, C.-H.C. and C.-Y.P. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by grants CMRPG8K1101 from Chang Gung Memorial Hospital, Taiwan (C.-H.C.), and MOST 109-2314-B-039-020 from Ministry of Science and Technology, Taiwan (C.-Y.P.).

Institutional Review Board Statement: The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of Chang Gung Memorial Hospital (IRB No.: 202000445B0) and China Medical University Hospital (IRB No.: CMUH102-REC1-113).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

Data Availability Statement: All data generated or analyzed in the study are included in the article or its online Supplementary Material files. Further inquiries can be directed to the corresponding author.

Acknowledgments: The authors would like to thank Chia-Hsin Lin for statistical analysis.
Conflicts of Interest: Cheng-Yuan Peng has served as an advisory committee member for AbbVie, Bristol-Myers Squibb, Gilead, Merck Sharp & Dohme, and Roche. All other coauthors have no conflicts of interest to declare.

References

1. Chang, T.-T.; Liaw, Y.-F.; Wu, S.-S.; Schiff, E.; Han, K.-H.; Lai, C.-L.; Safadi, R.; Lee, S.S.; Halota, W.; Goodman, Z.; et al. Long-term entecavir therapy results in the reversal of fibrosis/cirrhosis and continued histological improvement in patients with chronic hepatitis B. *Hepatology* 2010, 52, 886–893. [CrossRef] [PubMed]

2. Marcellin, P.; Gane, E.; Buti, M.; Afdhal, N.; Sievert, W.; Jacobson, I.M.; Washington, M.K.; Germanidis, G.; Flaherty, J.F.; Schall, R.A.; et al. Regression of cirrhosis during treatment with tenofovir disoproxil fumarate for chronic hepatitis B: A 5-year open-label follow-up study. *Lancet* 2013, 381, 468–475. [CrossRef]

3. Su, T.-H.; Hu, T.-H.; Chen, C.-Y.; Huang, Y.-W.; Chuang, W.-L.; Lin, C.-C.; Wang, C.-C.; Su, W.-W.; Chen, M.-Y.; Peng, C.-Y.; et al. Four-year entecavir therapy reduces hepatocellular carcinoma, cirrhotic events and mortality in chronic hepatitis B patients. *Liver Int.* 2016, 36, 1755–1764. [CrossRef] [PubMed]

4. Nguyen, M.H.; Yang, H.-I.; Le, A.; Henry, L.; Nguyen, N.; Lee, M.-H.; Zhang, J.; Wong, C.; Wong, C.; Trinh, H. Reduced Incidence of Hepatocellular Carcinoma in Cirrhotic and Noncirrhotic Patients with Chronic Hepatitis B Treated With Tenofovir—A Propensity Score–Matched Study. *J. Infect. Dis.* 2019, 219, 10–18. [CrossRef] [PubMed]

5. Papatheodoridis, G.V.; Dalekos, G.N.; Sypsa, V.; Yurdaydin, C.; Buti, M.; Goulis, J.; Calleja, J.L.; Chi, H.; Manolakopoulos, S.; Mangia, G.; et al. PAGE-B predicts the risk of developing hepatocellular carcinoma in Caucasians with chronic hepatitis B on 5-year antiviral therapy. *J. Hepatol.* 2016, 64, 800–806. [CrossRef]

6. Chen, C.H.; Lee, C.M.; Lai, H.C.; Hu, T.H.; Su, W.P.; Lu, S.N.; Lin, C.H.; Hung, C.H.; Wang, J.H.; Lee, M.H.; et al. Prediction model of hepatocellular carcinoma risk in Asian patients with chronic hepatitis B treated with entecavir. *Oncotarget* 2017, 8, 92431–92441. [CrossRef]

7. Sohn, W.; Cho, J.-Y.; Kim, J.H.; Lee, J.I.; Kim, H.J.; Woo, M.-A.; Jung, S.-H.; Paik, Y.-H. Risk score model for the development of hepatocellular carcinoma in treatment-naïve patients receiving oral antiviral treatment for chronic hepatitis B. *Clin. Mol. Hepatol.* 2017, 23, 170–178. [CrossRef]

8. Hsu, Y.-C.; Yip, T.C.-F.; Ho, H.J.; Wong, V.W.-S.; Huang, Y.-T.; El-Serag, H.B.; Lee, T.-Y.; Wu, M.-S.; Lin, J.-T.; Wong, G.L.-H.; et al. Development of a scoring system to predict hepatocellular carcinoma in Asians on antivirals for chronic hepatitis B. *J. Hepatol.* 2018, 69, 278–285. [CrossRef]

9. Kim, J.H.; Lee, M.; Jun, B.G.; Kim, T.S.; Suk, K.T.; Kang, S.H.; Kim, M.Y.; Cheon, G.J.; Kim, D.J.; et al. Modified PAGE-B score predicts the risk of developing hepatocellular carcinoma in Asians with chronic hepatitis B on antiviral therapy. *J. Hepatol.* 2018, 69, 1066–1073. [CrossRef]

10. Yu, J.H.; Suh, Y.J.; Jin, Y.-J.; Heo, N.-Y.; Jang, J.W.; You, C.R.; An, H.Y.; Lee, J.-W. Prediction model for hepatocellular carcinoma risk in treatment-naive chronic hepatitis B patients receiving oral antiviral treatment. *Eur. J. Gastroenterol. Hepatol.* 2019, 31, 865–872. [CrossRef]

11. Yang, H.-I.; Yeh, M.-L.; Wong, G.L.-H.; Peng, C.-Y.; Chen, C.-H.; Trinh, H.N.; Cheung, K-S.; Xie, Q.; Su, T.-H.; Kozuka, R.; et al. Real-World Effectiveness from the Asia Pacific Rim Liver Consortium for HBV Risk Score for the Prediction of Hepatocellular Carcinoma in Chronic Hepatitis B Patients Treated with Oral Antiviral Therapy. *J. Infect. Dis.* 2020, 221, 389–399. [CrossRef] [PubMed]

12. Fan, R.; Papatheodoridis, G.; Sun, J.; Innes, H.; Toyoda, H.; Xie, Q.; Mo, S.; Sypsa, V.; Guha, I.N.; Kumada, T.; et al. aMAP risk score predicts hepatocellular carcinoma development in patients with chronic hepatitis B. *J. Hepatol.* 2020, 73, 1368–1378. [CrossRef] [PubMed]

13. Ahn, S.B.; Choi, J.; Jun, D.W.; Oh, H.; Yoon, E.L.; Kim, H.S.; Jeong, S.W.; Kim, S.E.; Shim, J.; Cho, Y.K.; et al. Twelve-month post-treatment parameters are superior in predicting hepatocellular carcinoma in patients with chronic hepatitis B. *Liver Int.* 2021, 41, 1652–1661. [CrossRef] [PubMed]

14. Poh, Z.; Shen, L.; Yang, H.-I.; Seto, W.-K.; Wong, V.W.; Lin, C.Y.; Goh, B.-G.; Chang, P.-E.; Chan, H.L.-Y.; Yuen, M.-F.; et al. Real-world risk score for hepatocellular carcinoma (RWS-HCC): A clinically practical risk predictor for HCC in chronic hepatitis B. *Gut* 2016, 65, 887–888. [CrossRef] [PubMed]

15. Kim, H.-S.; Yu, X.; Kramer, J.; Thrift, A.P.; Richardson, P.; Hsu, Y.-C.; Flores, A.; El-Serag, H.B.; Kanwal, F. Comparative performance of risk prediction models for hepatitis B-related hepatocellular carcinoma in the United States. *J. Hepatol.* 2020, 79, 294–301. [CrossRef] [PubMed]

16. Tamaki, N.; Kurosaki, M.; Loomba, R.; Izumi, A.N. Clinical Utility of Mac-2 Binding Protein Glycosylation Isoemer in Chronic Liver Diseases. *Ann. Lab. Med.* 2021, 41, 16–24. [CrossRef]

17. Ishii, A.; Nishikawa, H.; Enomoto, H.; IWata, Y.; Kishino, K.; Shimono, Y.; Hasegawa, K.; Nakano, C.; Takata, R.; Nishimura, T.; et al. Clinical implications of serum Wisteria floribunda agglutinin-positive Mac-2-binding protein in treatment-naive chronic hepatitis B. *Hepatol. Res.* 2017, 47, 204–215. [CrossRef]

18. Ichikawa, Y.; Joshita, S.; Umemura, T.; Shobugawa, Y.; Usami, Y.; Shibata, S.; Yamazaki, T.; Fujimori, N.; Komatsu, M.; Matsumoto, A.; et al. Serum Wisteria floribunda agglutinin-positive human Mac-2 binding protein may predict liver fibrosis and progression to hepatocellular carcinoma in patients with chronic hepatitis B virus infection. *Hepatol. Res.* 2017, 47, 226–233. [CrossRef]
19. Kawaguchi, K.; Honda, M.; Ohta, H.; Terashima, T.; Shimakami, T.; Arai, K.; Yamashita, T.; Sakai, Y.; Yamashita, T.; Mizukoshi, E.; et al. Serum Wisteria floribunda agglutinin-positive Mac-2 binding protein predicts hepatocellular carcinoma incidence and recurrence in nucleos(t)ide analogue therapy for chronic hepatitis B. J. Gastroenterol. 2018, 53, 740–751. [CrossRef]

20. Shinkai, N.; Nojima, M.; Iio, E.; Matsunami, K.; Toyoda, H.; Murakami, S.; Inoue, T.; Ogawa, S.; Kumada, T.; Tanaka, Y. High levels of serum Mac-2-binding protein glycosylation isomer (M2BPGi) predict the development of hepatocellular carcinoma in hepatitis B patients treated with nucleos(t)ide analogues. J. Gastroenterol. 2018, 53, 883–889. [CrossRef]

21. Toyoda, H.; Kumada, T.; Tada, T.; Kaneoka, Y.; Maeda, A.; Korenaga, M.; Mizokami, M.; Narimatsu, H. Serum WFA+ -M2BP levels as a prognostic factor in patients with early hepatocellular carcinoma undergoing curative resection. Liver Int. 2016, 36, 293–301. [CrossRef] [PubMed]

22. Kim, H.S.; Kim, S.U.; Kim, B.K.; Park, J.Y.; Kim, D.Y.; Ahn, S.H.; Han, K.H.; Park, Y.N.; Han, D.H.; Kim, K.S.; et al. Serum Wisteria floribunda agglutinin-positive human Mac-2 binding protein level predicts recurrence of hepatitis B virus-related hepatocellular carcinoma after curative resection. Clin. Mol. Hepatol. 2020, 26, 33–44. [CrossRef] [PubMed]

23. Marrero, J.A.; Kulik, L.M.; Sirlin, C.B.; Zhu, A.X.; Finn, R.S.; Abecassis, M.M.; Roberts, L.R.; Heimbach, J.K. Diagnosis, Staging, and Management of Hepatocellular Carcinoma: 2018 Practice Guidance by the American Association for the Study of Liver Diseases. Hepatology 2018, 68, 723–750. [CrossRef] [PubMed]

24. Handelsman, Y.; Bloomgarden, Z.T.; Grunberger, G.; Umpierrez, G.; Zimmerman, R.S.; Bailey, T.S.; Blonde, L.; Bray, G.A.; Cohen, A.J.; Dagogo-Jack, S.; et al. American Association of Clinical Endocrinologists and American College of Endocrinology—clinical practice guidelines for developing a diabetes mellitus comprehensive care plan- 2015. Endocr. Pract. 2015, 21 (Suppl. 1), 1–87. [CrossRef] [PubMed]

25. Kuno, A.; Ikehara, Y.; Tanaka, Y.; Ito, K.; Matsuda, A.; Sekiya, S.; Hige, S.; Sakamoto, M.; Kage, M.; Mizokami, M.; et al. A serum “sweet-doughnut” protein facilitates fibrosis evaluation and therapy assessment in patients with viral hepatitis. Sci. Rep. 2013, 3, 1065. [CrossRef] [PubMed]

26. Seto, W.-K.; Wong, D.K.-H.; Fung, J.; Huang, F.-Y.; Liu, K.S.-H.; Lai, C.-L.; Yuen, M.-F. Linearized hepatitis B surface antigen and hepatitis B core-related antigen in the natural history of chronic hepatitis B. Clin. Microbiol. Infect. 2014, 20, 1173–1180. [CrossRef] [PubMed]

27. Rubin, D.B.; Schenker, N. Multiple imputation in health-care databases: An overview and some applications. Stat. Med. 1991, 10, 585–598. [CrossRef]

28. Sullivan, L.M.; Massaro, J.M.; D’Agostino, R.B., Sr. Presentation of multivariate data for clinical use: The Framingham Study risk score functions. Stat. Med. 2004, 23, 1631–1660. [CrossRef]

29. Uno, H.; Cai, T.; Pencina, M.J.; D’Agostino, R.B.; Wei, L.J. On the C-statistics for evaluating overall adequacy of risk prediction procedures with censored survival data. Stat. Med. 2011, 30, 1105–1117. [CrossRef]

30. Blanche, P.; Dartigues, J.F.; Jacqmin-Gadda, H. Estimating and comparing time-dependent areas under receiver operating characteristic curves for censored event times with competing risks. Stat. Med. 2013, 32, 5381–5397. [CrossRef]

31. Sharma, S.A.; Kowgier, M.; Hansen, B.E.; Brouwer, W.P.; Maan, R.; Wong, D.; Shah, H.; Khalili, K.; Yim, C.; Heathcote, E.J.; et al. Toronto HCC risk index: A validated scoring system to predict 10-year risk of HCC in patients with cirrhosis. J. Hepatol. 2018, 68, 92–99. [CrossRef] [PubMed]

32. Kim, H.Y.; Lampertico, P.; Nam, J.Y.; Lee, H.-C.; Kim, S.U.; Sinn, D.H.; See, Y.S.; Lee, H.A.; Park, S.Y.; Lim, Y.-S.; et al. An artificial intelligence model to predict hepatocellular carcinoma risk in Korean and Caucasian patients with chronic hepatitis B. J. Hepatol. 2022, 76, 311–318. [CrossRef] [PubMed]

33. Qamar, A.A.; Grace, N.D.; Grossmann, R.J.; Garcia-Tsao, G.; Bosch, J.; Burroughs, A.K.; Maurer, R.; Planas, R.; Escorsell, A.; Garcia-Pagan, J.C.; et al. Platelet count is not a predictor of the presence or development of gastroesophageal varices in cirrhosis. Hepatology 2008, 47, 153–159. [CrossRef] [PubMed]

34. Ripoll, C.; Groszmann, R.J.; Garcia-Tsao, G.; Bosch, J.; Grace, N.; Burroughs, A.; Planas, R.; Escorsell, A.; Garcia-Pagan, J.C.; Makuch, R.; et al. Hepatic venous pressure gradient predicts development of hepatocellular carcinoma independently of severity of cirrhosis. J. Hepatol. 2009, 50, 923–928. [CrossRef] [PubMed]

35. Hosaka, T.; Suzuki, F.; Kobayashi, M.; Fujiyama, S.; Kawamura, Y.; Sezaki, H.; Akuta, N.; Suzuki, Y.; Saitoh, S.; Arase, Y.; et al. Impact of hepatitis B core-related antigen on the incidence of hepatocellular carcinoma in patients treated with nucleos(t)ide analogues. Aliment. Pharmacol. Ther. 2019, 49, 457–471. [CrossRef]