Cav2.2-mediated NMDA receptor signaling in short-term memory

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Abstract
It has been reported that Cav2.1-mediated N-methyl-D-aspartate (NMDA) receptor signaling is critical pathway in hippocampus and nucleus accumbens for spatial short-term memory. Neuronal voltage-dependent Cav2.1 and Cav2.2 have predominantly expressions at presynaptic neuronal terminals and mediate glutamate release in the central nervous systems. Recently, although Cav2.2 in the hippocampus and nucleus accumbens is also critical for spatial cognition, it remains unknown that functional Cav2.2-mediated NMDA receptor signaling in cognitive performance at the system level. This study examined whether Cav2.2-mediated NMDA receptor signaling mediates spatial short-term memory using the Y-maze test via a combined subthreshold doses of pharmacological approach. In previous our studies, Mice received systemic injection of NMDA receptor blocker (+/–)-3-(2-carboxypiperazin-4-yl)-propyl-1-phosphonic acid (CPP, 5mg/kg), mice received intra-hippocampal injection of Cav2.2 blocker (ω-conotoxin GVIA, 1pg/side), or mice received intra-accumbens injection of Cav2.1 blocker (ω-conotoxin GVIA, 1pg/side), or mice received intra-accumbens injection of Cav2.2 blocker (ω-conotoxin GVIA, 1pg/side) were ineffective for the spatial short-term memory. However, a combination of subthreshold doses of 5mg/kg CPP systemic injection and 1pg/side ω-conotoxin GVIA intra-hippocampal injection triggered a spatial short-term memory deficit. Furthermore, a combination of subthreshold doses of 5mg/kg CPP systemic injection and 1pg/side ω-conotoxin GVIA intra-accumbens injection also showed impaired spontaneous alternation patterns. These results indicate that Cav2.2-mediated NMDA receptor signaling is critical in the hippocampus and nucleus accumbens for spatial short-term memory. This study examined whether Cav2.2-mediated NMDA receptor signaling in cognitive performance at the system level. This study examined whether Cav2.2-mediated NMDA receptor signaling in cognitive performance at the system level.

Introduction
Neuronal voltage-dependent Ca2+ channel (VDCC), including Cav2.1 (P/Q-type), Cav2.2 (N-type), and Cav2.3 (R-type) channels, mediate the presynaptic machinery involved in the vesicular release of neurotransmitters [1-3]. A VDCC is a molecular complex comprising α1, α2-δ, β, and γ subunits [3]. The α1 subunit is essential for channel functioning and determines fundamental channel properties [3]. The ~190kDa pore-forming transmembrane α1 subunit (~2800 amino acids) is organized in four homologous domains (I–IV), comprising six transmembrane α helices (S1–S6) and the pore-forming P loop between S5 and S6.

It has been reported that N-methyl-D-aspartate (NMDA)-dependent processes are involved in the mechanisms of memory [4] and that glutamatergic system is one of the neurotransmitter systems regulated by Cav2.1 and Cav2.2 [5,6]. It has been also reported that glutamatergic afferents from the hippocampus provide the main source of information to the nucleus accumbens (NAc) during cognitive activity [7-11]. Our previous reports have showed that Cav2.1-regulated glutamatergic signaling in the hippocampus [12] and NAc [13] is important in short-term spatial learning. We have also demonstrated the importance of Cav2.2-regulated signaling in the hippocampus [14] and NAc [15] for spatial short-term memory. However, the role of Cav2.2-regulated glutamatergic signaling in the neural circuits underlying spatial short-term memory has not been studied.

In previous study, subthreshold doses of cannabinoid CB1 receptor antagonist and selective serotonin reuptake inhibitors, which separately had no effect on antidepressant behavioral tests, showed a clear effect in combination without the side effects [16], suggesting that combinations of compounds with different molecular targets are useful tools to study the neuronal signaling mechanisms.

In this study, to examine the importance of Cav2.2-mediated glutamatergic transmission in the hippocampus and NAc for spatial short-term memory, we administered the Y-maze test for mice treated with ω-conotoxin GVIA, Cav2.1 inhibitor and (+/–)-3-(2-carboxypiperazin-4-yl)-propyl-1-phosphonic acid (CPP), a NMDA receptor antagonist.

Materials and methods

Animals
All animal procedures were approved by the Animal Experiments Committee of Shanghai Jiao Tong University and RIKEN. The C57BL/6J mice were provided by Charles River Japan (Kanagawa, Japan). The mice were given free access to water and food pellets (CRF-1; Oriental Yeast Co. Ltd., Tokyo, Japan) and were housed under a 12/12-h light/dark cycle (lights on from 08:00 to 20:00) at 23 ± 1°C and 50± 10% relative humidity, on a 12-h light/dark cycle (lights on from 08:00 to 20:00) at 23 ± 1°C and 50± 10% relative humidity.

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55 ± 5% humidity. Testing was performed during the light phase of the cycle. We used separate groups of male 2-month-old mice for each of the behavioral tests. All experiments were conducted blind to the treatment condition of the mice.

Drugs

CPP (5mg/kg, Sigma-Aldrich) was dissolved in 0.9% NaCl (vehicle) and injected intraperitoneally 20min before behavioral testing. Cav2.2 blocker, ω-conotoxin GVIA (1, 10, or 50 pg/μL, Peptide Institute, Osaka, Japan) were dissolved in 0.9% NaCl (vehicle). Mice that were not treated with drugs received an equivalent volume of vehicle.

Infusion

Under anesthesia and using standard stereotaxic procedures, stainless-steel guide cannulae (22-gauge) were implanted into the dorsal hippocampus (posterior to bregma, -2.0mm; lateral to midline, ±2.0mm; ventral from the dura, -2.0mm) or nucleus accumbens (+1.7mm, ±1.0 mm, +2.3 mm). Mice were allowed to recover for at least 1 week following surgery. The mice were briefly anesthetized with isoflurane to facilitate insertion of the injection cannula (26-gauge). Infusions into the hippocampus (0.1μL/side) or nucleus accumbens (0.1μL/side) are accomplished at a rate of 0.05μL/min 30min before behavioral testing. The injection cannula was left in place for 2min following the infusion.

Y maze tests

All behavioral tests were conducted between 10:00 and 16:00 by a trained experimenter who was blind to the mouse strains. We used separate groups of male mice for each of the behavioral tests. The mice were moved into the behavioral testing room at least 2h before testing. The Y-maze test was performed using the reported procedure with slight modifications [12]. The experiments were performed at 35 lux. Before behavioral testing, the mice were placed in one of the compartments and allowed to move freely on the one of the arms for 10min. Each mouse performed one trial. An arm entry was defined as any three legs entering one of the arms, and the sequence of entries was recorded manually using videotapes. An alteration was defined as entry into all three arms with consecutive choices. The percentage of spontaneous alteration was calculated as (actual alteration/maximum alteration) × 100.

Histology

Histological verification of the cannula locations was performed at the end of behavioral testing. Mice were perfused transcardially with 4% PFA. After extraction from the skull, the brains were postfixed in 4% PFA and then transferred to a 30% sucrose solution until sectioning. Coronal sections (40μm thick, taken every 120μm) were cut on a cryostat (–16°C) and mounted on glass microscope slides. After drying, the sections were stained with cresyl violet. Mice with injection needle placements outside of the boundaries of targeted areas were excluded from behavioral analysis.

Data analysis

Data are presented as means ± standard error on the mean (SEM). Statistical analyses for the behavioral tests were conducted using Excel Statistics 2006 (SSRI, Tokyo, Japan). Data were analyzed using repeated measures ANOVA with Tukey’s test.

Results

The drug doses were determined according to previous report [14,15,17]. In our previous study, we examined four groups of mice, including four groups each of mice that were given systemic injections of 0, 2.5, 5, or 10mg/kg CPP. Although the groups did not differ significantly in the total number of arm entries, spontaneous alterations were significantly different [17]. The mice given 10mg/kg CPP showed fewer spontaneous alteration behaviors than the mice given 5mg/kg CPP. We used 5mg/kg as a subthreshold dose of CPP in this study. On the other hand, among mice received intra-hippocampal or intra-acccumbens injection of 0, 1, 5, or 10pg/side ω-conotoxin GVIA, mice received 5 or 10pg/side ω-conotoxin GVIA in both regions triggered a spatial short-term memory deficit [14,15]. To examine the effect of subthreshold doses of ω-conotoxin GVIA on Y-maze test, we injected 0, 0.1, 1, or 5pg/side ω-conotoxin GVIA into hippocampus or NAC.

Effects of subthreshold doses of CPP plus intra-hippocampal injection of ω-conotoxin GVIA on Y-maze test

We examined four groups of mice (n=10 each), including group each of mice that were given intra-hippocampal injections of 0, 0.1, 1, or 5pg/side ω-conotoxin GVIA plus 5mg/kg CPP, respectively. The groups did not differ significantly in arm entries [F(3,36)=0.8, P>0.05] (data not shown) (Figure 1A), while spontaneous alterations were significantly different [F(3,36)=122.2, P<0.01] (Figure 1B). The mice given 1pg/side ω-conotoxin GVIA plus 5mg/kg CPP showed fewer spontaneous alteration behaviors than the mice given 0pg/side ω-conotoxin GVIA plus 5mg/kg CPP.

Effects of subthreshold doses of CPP plus intra-acccumbens injection of ω-conotoxin GVIA on Y-maze test

We examined four groups of mice (n=10 each), including group each of mice that were given intra-acccumbens injections of 0, 0.1, 1, or 5pg/side ω-conotoxin GVIA plus 5mg/kg CPP, respectively. The groups did not differ significantly in arm entries [F(3,36)=0.7, P>0.05] (Figure 2A), while spontaneous alterations were significantly different [F(3,36)=112.2, P<0.01] (Figure 2B). The mice given 1pg/side ω-conotoxin GVIA plus 5mg/kg CPP showed fewer spontaneous alteration behaviors than the mice given 0pg/side ω-conotoxin GVIA plus 5mg/kg CPP.

Discussion

In this study, to evaluate whether Ca2,2-mediated NMDA receptor signaling has an important role in memory, we administered the Y-maze tests for spatial memory to mice treated with a combination...
of subthreshold doses of systemic injection of NMDA receptor CPP and local injection of Cav2.2 blocker, ω-conotoxin GVIA. The Y-maze test is based on the spontaneous tendency of rodents to enter a novel arm more often than the other arms due to the spatial short-term memory [18]. Behavioral stimuli such as electrical stimulation in a fear-conditioning test are used to examine memory formation in rodents [4]. Since CPP and ω-conotoxin GVIA are thought to be effect on analgesia [19,20], cognitive tests without stimulations would be more appropriate for this study. The administration of different drugs at a lower dose induced the effects of phenotypes and identified functional signaling pathways [16], because the precise neuronal circuit systems play an important role in phenotypes at behavioral levels.

Pharmacological studies have showed that NMDA receptor antagonists impair the cognitive functions [21-23] and that NMDA receptor agonists enhance memory tasks [24]. Previous studies have demonstrated that systemic injection of CPP dose-dependently inhibited spatial short-term memory in the Y maze test [17]. Since Cav2.2 and NMDA receptor are present at a variety of synapses [25,26], we need to examine which sites are important for spatial short-term memory formation. In this regard, studies using local injection into specific region are a useful tool to specifically understand the relevant neuronal circuits. Indeed, local injections have showed that Cav2.1-regulated glutamatergic signaling in the hippocampus [12] and NAc [13] is important in short-term spatial learning.

We showed that a subthreshold dose of CPP significantly decreased the spontaneous alteration behaviors when combined with a subthreshold dose of ω-conotoxin GVIA, although administration of CPP or ω-conotoxin GVIA alone was ineffective in the Y maze test [14,15,17]. The precise regulation of Ca²⁺ signaling through Cav2.2 plays an important role in neuronal circuits [3]. Therefore, because Cav2.2 on the presynapse is important in glutamate release [3] and NMDA receptor on the postsynapse is important in signaling leading to cognitive function [4], the combination of subthreshold doses induced impairment of glutamatergic circuit system and deficit of spatial short-term memory. Furthermore, these results were detected in a combination of subthreshold doses of CPP systemic injection plus ω-conotoxin GVIA intra-hippocampal injection and a combination of subthreshold doses of CPP systemic injection plus ω-conotoxin GVIA intra-accumbens injection. It has been reported that glutamatergic transmission within the NAC plays a central role in the transfer of different types of information from cortical and limbic regions including hippocampus [7-11]. Our results indicate that spatial short-term memory formation is impaired when either Cav2.2-mediated NMDA receptor signaling in hippocampus or NAc is inhibited. On the other hand, previous reports have showed that Cav2.1-regulated glutamatergic signaling in the hippocampus [12] and NAc [13] is important in short-term spatial learning. We need a future study to understand the role of different VDCCs in the neural circuits underlying spatial short-term memory.

In conclusion, we found that subthreshold doses of ωα-conotoxin GVIA and CPP in the hippocampus and NAc, which are effective when administered separately at threshold doses, have a combined effect on spatial short-term memory. These results indicate that Cav2.2-mediated NMDA receptor signaling is critical in the hippocampus and nucleus accumbens for spatial short-term memory. The subthreshold pharmacological approach would be useful to analyze specifically functional signaling cascade in neuronal circuit. Our results also indicate that the detailed comparison of combinations of different pharmacological agents, concentrations and infusion sites would be helpful in understanding the relationships among the synaptic signaling, neuronal circuit and observed behavior.

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Authors’ contributions

WL and ET designed and supervised the research, and wrote the manuscript. YZ and KN performed the surgeries and behavioral experiments. All authors read and approved the final version of the manuscript.

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