Factors associated with acquisition of glycopeptide-resistant enterococci during a single-strain outbreak

S. Deboscker1,2, P. Schneider3, F. Séverac2,4, C. Ménard5, J. Gaudart6,7, T. Lavigne1,8 and N. Meyer2,4

1Service d’hygiène hospitalière, Hôpitaux universitaires de Strasbourg, Strasbourg, France; 2iCube, UMR7357, Université de Strasbourg, Strasbourg, France; 3Pharmacie, Hôpitaux universitaires de Strasbourg, Strasbourg, France; 4Groupe Méthode en Recherche Clinique (GMRC), Service de Santé Publique, Hôpitaux universitaires de Strasbourg, Strasbourg, France; 5Laboratoire de bactériologie, Hôpitaux universitaires de Strasbourg, Strasbourg, France; 6Hôpital La Timone, Service Biostatistique et Technologies de l’Information et de la Communication, APHM, Marseille, France; 7IRID, INSEER, SESSTIM UMR9312, Aix Marseille Univ, Marseille, France and 8EA7290, Virulence bactérienne précoce, Fédération de médecine translationnelle de Strasbourg, Université de Strasbourg, Strasbourg, France

Summary

The aim of our study was to describe and to investigate the factors associated with glycopeptide-resistant enterococci (GRE) acquisition during a single-strain outbreak which occurred in several wards of hospital from September 2013 to January 2014. We designed a case–control study. Analyses were performed using Bayesian methods. Univariate logistic regressions with informative priors from published studies were conducted. A multivariate model was built including variables with a probability of odd-ratio exceeding one (Pr) >85% or <15%. Thirteen cases and 52 controls were recruited. The description of this outbreak highlighted the importance to quickly detect patients at risk of GRE carriage in order to implement the isolation measures and to transfer to dedicated department if they are effectively carriers. Following multivariate analysis, antibiotics during hospitalisation (Pr = 0.968), number of hospitalisation days in the year (Pr = 0.964), antacids intake (Pr = 0.878) (with a risk increase), immunosuppression (Pr = 0.026) and isolation measures (Pr = 0.003) (both with protective effect) were associated with GRE acquisition. The use of Bayesian statistics was useful because of our study’s small population size and prior information availability.

Introduction

In French hospitals, the first outbreaks of glycopeptide-resistant enterococci (GRE) were reported in 2004–2005, then other reports followed [1–3]. Nevertheless, contrary to several European countries in which the spread of GRE is now endemic, their diffusion remains limited in France [4]. While several studies have shown an increase in GRE-related mortality and costs [5, 6], the risk is more ecological than infectious, by spread of the resistance gene to Staphylococcus aureus strains. In recent years, risk factors for acquiring GRE have been investigated in numerous studies [7–19], but only few have involved situations with strains having genomically identical profiles [7, 13, 15]. We have thus become interested in one GRE outbreak occurring at the Hôpitaux Universitaires de Strasbourg (HUS), France. Our study’s primary objective was to determine the factors associated with GRE acquisition by cross-transmission during a hospital outbreak. The secondary goal was to describe a hospital GRE outbreak.

Methods

Management of GRE

The study involved an outbreak of GRE faecium VanB (with genomically identical profiles, verified by the National Reference Centre (CNR) on resistance to antibiotics) that occurred from September 2013 to January 2014 in several wards of HUS, a university hospital with a capacity of approximately 2700 beds.

The outbreak was managed by the Infection Control Team (ICT), along with the help of healthcare workers (HCW). To control this outbreak, we applied rules reflecting current recommendations [20]: Standard precautions associated with the contact precautions (CP) for carrier patients (cases) and for patients having been managed by same nursing team as a case (contact patients). In 2013, at the HUS, the CP for cases involved managing the patient in a private room, dedicated equipment and regular use by the HCW and visitors of disposable
gloves, as well as gowns. For contact patients, CP consisted of using dedicated equipment and a disposable apron, but there was no requirement for a private room, gloves or gown. As soon as possible, case patients were transferred to a dedicated unit: for hospitalisation, the dedicated department was a separate unit with only GRE-carriers and dedicated HCW; for chronic haemodialysis, it was a sector with individual rooms but HCW could be shared with non-GRE-carriers. As advised by recommendations: case and contact patients were registered on the information system to be identified as carriers or contacts by the hospital’s admissions software; contact patients were screened and considered to be non-GRE carriers after three weekly GRE-negative screening (rectal and/or colostomy swab) following the end of exposure to a case; environmental sampling was recommended only in case of a not ending outbreak.

To highlight the factors associated with acquiring this GRE epidemic strain, we implemented a monocentric, case–control study with individual matching of four controls for one case. The choice to match four controls for one case was based on the possibilities to recruit controls, and on the information from literature showing that the gain in power is negligible beyond four controls for one case [21].

**Definition of cases and controls**

The cases were patients at the HUS with at least one positive sample containing the involved strain of GRE *faecium* VanB during this outbreak (infection or colonisation). Each case created contact patients. According to current French recommendations [20], these contact patients are defined as patients having been managed by same nursing team as a case during inpatient hospitalisation (possibly in several wards) or/and haemodialysis sessions. The search period for contact patients was defined based on the last negative screening of the case (or, otherwise, his admission to the HUS) until his management in the dedicated department or until the end of his management (death or discharge from the HUS). Among contact patients of each case, we drew randomly four controls. The controls were contact patients who never had any positive sample with the strain involved and who had at least three negative screenings after exposure to a case, within a minimum time period of 14 days after exposure (weekly screening, according to the recommendations [20]) and not exceeding 6 weeks. Contact patients without GRE screening or with screenings that were excessively late (more than 6 weeks) or incomplete (<three screenings) were excluded from participating to the study.

This study received an approval opinion from the Ethics Committee of the HUS and an approval from the CNIL (Commission Nationale de l'Informatique et des Libertés (National Commission for Information Technology and Civil Liberties)).

**Study parameters**

Parameters were selected based on both our experience and the literature (Medline database). A collection grid was completed for each patient (cases and controls), comprising the following data: demographic and clinical; use of antibiotics or antacids over the 3 previous months and during hospitalisation (from admission to screenings); length of hospitalisation and the number of dialysis sessions during the year and in the month preceding the screenings; exposure to a case (presence of diarrhoea, and isolation measures implemented for the case); cross-transmission risk (acute or chronic dialysis, diarrhoea, dependence defined by the need for assistance when washing one’s body, management of excretions defined by the use of anatomical protection, the use of a bedpan, existence of stool collection through an ostomy or the ability to go to the bathroom, physical therapy, isolation measures, i.e. protective isolation or CP implemented throughout the duration of exposure).

**Microbiology**

GRE screening was performed by swabbing (Copan ESwab) the carrier sites (rectum and/or ostomy), preferably in the morning, before toilette. The sample was cultured in a specific medium (bioMérieux chromID VRETM agar containing 8 mg/l of vancomycin) for 48 h. If suspicious colonies were isolated, an initial search for *vanA* and *vanB* resistance genes was performed using molecular biology directly on the sample (real-time PCR, ABI Prism 7500, Applied Biosystems, Foster City, CA, USA). If this search was negative, no additional tests were carried out. If the result was positive, additional tests were done, namely an identification (mass spectrometer – MALDI-TOF, Bruker) to differentiate the *Enterococcus faecium* and *E. faecalis* strains, along with an antibiogram (Vitek 2, bioMérieux) to assess sensitivity to vancomycin and teicoplanin. PCR was used to confirm the isolated strains. The bacteriology laboratory forwarded the results in the next 2–4 days. The laboratory reported positive results directly to the ICT, and a message was also attached to the result inviting the HCW to contact the ICT. Genomic comparison of the strains was performed by the CNR. This comparison was conducted using the DIVERSILAB® rep-PCR method (bioMérieux).

**Statistical analyses**

Statistical analyses were performed using a Bayesian approach [22, 23] with the R software, Version 3.2.2 (copyright The R Foundation for Statistical Computing, Vienna, Austria) and with JAGS software, Version 3.4.0. Univariate analysis was carried out using a mixed logistic regression model including a cluster random effect to account for individual matching between the cases and controls. Parameters were estimated using MCMC (Markov chain Monte Carlo). After a warm-up period of 5000 iterations, 100 000 new iterations were produced to establish the parameter’s posterior distribution. Convergence of the algorithm to a stationary distribution was assessed graphically. Results were presented as odd ratio (OR), with its respective 95% credibility interval (CI), and the probability that the OR would be higher than one was calculated based on the posterior distribution. Variables presenting a probability of being associated with the case’s status (Prob OR>1, here in after abbreviated ‘Pr’) >0.850, indicating an increased risk of being a case, or lower than 0.150, meaning a reduced risk of being a case (or a protective effect), were included in a multivariate model. It should be noted that these probabilities (Pr) must not be confused with the P-value of classical statistical analyses. The Pr value close to either 0 or 1 is suggested of an effect. Bounds for categorisation of Pr were defined at 85% (or 15%), 95% (or 5%) and 99% (or 1%), corresponding with moderate, strong and very strong evidence for an association with the status of the patient.

When available, results from previous studies were used to build informative prior distributions on the log (OR) (Table 1 and appendix). The prior was built by using a Gaussian distribution with average $\mu$ and standard deviation $\sigma$, noted $N(\mu, \sigma)$. The average corresponded to the log (OR) originating from the
The outbreak

Overall, we were able to identify 13 patients carrying genetically identical *GRE faecium* VanB between September 2013 and January 2014. These 13 cases were distributed among several hospital wards, mostly nephrology (54%) and infectious diseases (23%) (Table 2). Seven cases were identified fortuitously (Table 3). Indeed they were not screened in the follow-up of contact patients of this outbreak (because they were not identified as a contact, i.e. as a patient managed by same nursing team as a case of this outbreak). But four of these cases (cases #2, #3, #4, #5) had previously been hospitalised together before their screening was found positive, precisely in dialysis and nephrology wards. In total, 10 cases have been transferred in the dedicated department after their identification. Figures 1 and 2 show chronology of events and links between cases.

Table 1. Informative priors

| Reference          | Country | Context            | Number of cases and controls | Parameters: OR (95% CI)                          |
|--------------------|---------|--------------------|------------------------------|--------------------------------------------------|
| Servais A, 2009 [13] | France  | Epidemic* (identical strains) | 14/125 | Gender (male): 9.0 (1.3–388.0)  
Diabetes: 1.7 (0.5–5.9)  
Glycopeptides the previous month: 11.1 (1.3–90.0)  
Fluoroquinolone the previous month: 5.0 (0.4–38.9)  
Dialysis: 9.6 (2.0–90.9)  
Diarrhoea: 2.3 (0.1–25.8) |
| Macintyre CR, 2001 [11] | Australia | Epidemic* | 19/66 | Vancomycin during hospitalisation: 1.1 (0.4–3.4)  
Metronidazole during hospitalisation: 4.4 (1.5–13.4)  
Dialysis: 1.0 (0.2–5.1) |
| McEvoy SP, 2006 [10] | Australia | Epidemic* | 107/107 | Gender (male): 0.86 (0.50–1.47)  
Diabetes: 3.98 (1.97–8.04)  
Chronic renal insufficiency: 4.82 (2.00–11.64)  
C3G during hospitalisation: 3.43 (1.83–6.44)  
Vancomycin during hospitalisation: 2.88 (0.99–8.40)  
Metronidazole during hospitalisation: 2.93 (1.43–6.01)  
Fluoroquinolone during hospitalisation: 2.36 (0.86–6.45)  
Age: 1.03 (1.01–1.05) |
| Karki S, 2012 [15] | Australia | Prevalence* | 58/116 | Gender (male): 0.92 (0.46–1.88)  
Antibiotics during hospitalisation: 3.83 (1.79–8.54)  
Metronidazole during hospitalisation: 2.93 (0.13–14.3)  
Dialysis: 2.54 (0.94–6.85) |
| Hoshuyama T, 2008 [7] | Japan | Epidemic* (identical strains) | 14/45 | Chemotherapy: 1.7 (0.4–5.7)  
Prolonged bed rest: 4.1 (1.07–15.5) |

*Enterococcus faecium* vanB epidemic.

| Hospital department | Number of cases (%) |
|---------------------|---------------------|
| Nephrology          | 7 (53.8)            |
| On chronic dialysis | 3 (23.1)            |
| Infectious diseases | 3 (23.1)            |
| Surgical intensive care | 1 (7.7)        |
| Medical intensive care | 1 (7.7)        |
| Orthopaedic surgery | 1 (7.7)            |
| TOTAL               | 13                  |

Population characteristics and statistical analyses

Sixty-five patients were included in the study (13 cases and 52 controls). The two principal reasons for hospitalisation were kidney disease (19/65, or 29%) and infection (15/65, or 23%). Most patients came from their homes (91%). Table 4 shows the population characteristics and results from univariate analyses. In univariate analyses, many parameters have indicated a probability of being associated with the case’s status >0.850 (including or without informative priors): diabetes, chronic renal insufficiency, antibiotic usage during the last 3 months, antacids intake during hospitalisation, antibiotherapy during hospitalisation, acute or chronic dialysis session, management of excretions, age and number of hospitalisation days in the year (particularly in nephrology). Several factors have shown a probability of being associated with the case’s status lower than 0.150: corticosteroid therapy, immunosuppressive treatment, metronidazole intake during the last 3 months and isolation measures.

Following multivariate analysis including informative priors when the latter were available (Table 5), the intake of antibiotics during hospitalisation (OR = 3.52 (0.94–13.62) with a probability
Moreover, we succeeded in collecting other specific cross-transmission parameters. Some were found in the literature, such as the use of dialysis sessions, dependence, presence of diarrhoea and management of excretions [7, 11, 13, 15]. Others were less specific, such as physical therapy and presence of diarrhoea in index cases. These different parameters did not show any impact on univariate or multivariate analyses, although they were instrumental in increasing the frequency of contacts among patients and personnel, as well as environmental contamination. Finally, it would have been interesting to collect information on the sharing of rooms, nursing staff or medical equipment, though this information proved difficult to collect a posteriori. It would thus be more appropriate to collect specific cross-transmission parameters in a prospective manner.

**Table 3. Detection circumstances of carriers during GRE faecium VanB outbreak and sampling types**

| Circumstances of case detection | Sampling type                                      | Number of cases (%) | Identification of cases |
|---------------------------------|---------------------------------------------------|---------------------|-------------------------|
| Follow-up of contact patients   | Screening of digestive GRE carriers               | 6 (46.2)            | #6, #7, #9, #10, #11, #12 |
| Fortuitous discovery            | Screening of digestive carriers of another GRE strain | 4 (30.8)            | #1, #2, #3, #13          |
|                                 | Systematic screening at intensive care unit       | 2 (15.4)            | #4, #8                  |
| Clinical sampling               |                                                   | 1 (7.7)             | #5                      |
| TOTAL                           |                                                   |                     | 13                      |
Fig. 2. Epidemic curve. New cases of glycopeptide-resistant enterococci (single-strain) per week from September 2013 to February 2014 and wards in which the cases were discovered (wards that took sample).

Table 4. Results of univariate analysis

| Characteristics                        | Cases (n = 13) | Controls (n = 52) | OR  (95% CI) | Prob OR>1* |
|----------------------------------------|---------------|------------------|--------------|------------|
| **Qualitative variables (n (%))**      |               |                  |              |            |
| Gender (male)                          | 8 (61.5)      | 29 (55.8)        | 0.72 (0.20–2.43) | 0.308      |
| With informative priors               |               |                  |              |            |
|                                        |               |                  | 0.91 (0.34–2.38) | 0.421      |
| **Antecedents - setting**             |               |                  |              |            |
| Ongoing cancer treatment              | 1 (7.7)       | 2 (3.8)          | 1.64 (0.10–17.42) | 0.650      |
| With informative priors               |               |                  |              |            |
|                                        |               |                  | 1.85 (0.26–11.63) | 0.738      |
| Neutropenia (neutrophiles <2G/L)      | 0 (0.0)       | 1 (1.92)         | 0.35 (0.00–12.65) | 0.299      |
| With informative priors               |               |                  |              |            |
|                                        |               |                  | 0.92 (0.26–3.15) | 0.385      |
| Corticosteroid therapy                | 1 (7.7)       | 14 (26.9)        | 0.20 (0.02–1.15) | 0.655      |
| Immunosuppressive treatment           | 1 (7.7)       | 12 (23.1)        | 0.25 (0.02–1.40) | 0.663      |
| HIV                                    | 0 (0.0)       | 2 (3.8)          | 0.24 (0.00–5.32) | 0.203      |
| Chronic alcoholism or drug addiction  | 1 (7.7)       | 8 (15.4)         | 0.41 (0.03–2.47) | 0.180      |
| Diabetes (types 1 or 2)               | 7 (53.8)      | 19 (36.5)        | 1.86 (0.54–6.27) | 0.844      |
| With informative priors               |               |                  |              |            |
|                                        |               |                  | 2.32 (0.65–6.48) | 0.950      |
| Chronic renal insufficiency           | 9 (69.2)      | 25 (48.1)        | 2.25 (0.64–8.43) | 0.896      |
| With informative priors               |               |                  |              |            |
|                                        |               |                  | 3.45 (1.17–11.05) | 0.986      |
| Antibiotherapy during the last 3 months | 8 (61.5) | 15/51 (29.4) | 3.44 (1.02–12.32) | 0.976      |
| Third-generation cephalosporines      | 4 (30.8)      | 4/51 (7.8)       | 4.81 (0.97–23.72) | 0.972      |
| Carbapenems                            | 2 (15.4)      | 1/51 (2.0)       | 6.87 (0.66–83.53) | 0.946      |
| Glycopeptides                          | 5 (38.5)      | 1/51 (2.0)       | 24.16 (3.74–242.48) | >0.999      |
| With informative priors               |               |                  |              |            |
|                                        |               |                  | 27.59 (5.13–196.74) | >0.999      |
| Fluoroquinolones                      | 4 (30.8)      | 7/51 (13.7)      | 2.59 (0.59–10.79) | 0.893      |
| With informative priors               |               |                  |              |            |
|                                        |               |                  | 3.09 (0.71–13.01) | 0.934      |
| Metronidazole                          | 0 (0.0)       | 3/51 (5.9)       | 0.17 (0.00–3.21) | 0.138      |

(Continued)
Our study was able to identify protective factors, namely, implementation of isolation measures and immunosuppression (intake of corticosteroids or immunosuppressive treatment). Isolation measures included CP and protective isolation, which appears quite consistent, given that these measures are said to be barriers against the transmission of microorganisms. The

| Characteristics                          | Cases (n = 13) | Controls (n = 52) | OR (95% CI) | Prob OR>1* |
|-----------------------------------------|---------------|-----------------|-------------|------------|
| Others                                  | 5 (38.5)      | 10/51 (19.6)    | 2.37 (0.60–8.79) | 0.897      |
| **During hospitalisation**              |               |                 |             |            |
| Carrier of another multidrug resistant bacterium | 5 (38.5) | 17 (32.7) | 1.18 (0.32–4.14) | 0.602      |
| Antacids intake                         | 12 (92.3)     | 36 (69.2)       | 3.12 (0.76–17.15) | 0.939      |
| Antibiotherapy                          | 13 (100.0)    | 37 (71.2)       | 5.63 (1.10–44.02) | 0.982      |
| With informative priors                 |               |                 |             |            |
| Third-generation cephalosporines        | 7 (53.9)      | 16 (30.8)       | 2.39 (0.70–8.14) | 0.919      |
| With informative priors                 |               |                 |             |            |
| Carbapenems                             | 4 (30.8)      | 9 (17.3)        | 1.94 (0.45–7.71) | 0.817      |
| Glycopeptides                           | 8 (61.5)      | 20 (38.5)       | 2.34 (0.70–8.16) | 0.916      |
| With informative priors                 |               |                 |             |            |
| Fluoroquinolones                        | 6 (46.2)      | 18 (34.6)       | 1.50 (0.42–5.19) | 0.741      |
| With informative priors                 |               |                 |             |            |
| Metronidazole                           | 1 (7.7)       | 5 (9.6)         | 0.67 (0.05–4.47) | 0.351      |
| With informative priors                 |               |                 |             |            |
| Others                                  | 10 (76.9)     | 25 (48.1)       | 3.18 (0.88–13.28) | 0.961      |
| **Exposure**                            |               |                 |             |            |
| Dialysis (acute or chronic)             | 8 (61.5)      | 22 (42.3)       | 2.07 (0.59–7.48) | 0.872      |
| With informative priors                 |               |                 |             |            |
| Diarrhoea                               | 3 (23.1)      | 14 (26.9)       | 0.74 (0.15–2.80) | 0.334      |
| With informative priors                 |               |                 |             |            |
| Dependence                              | 8 (61.5)      | 27 (51.9)       | 1.36 (0.40–4.91) | 0.688      |
| With informative priors                 |               |                 |             |            |
| Pelvis/ostomy/protection                | 8 (61.5)      | 22 (42.3)       | 2.00 (0.58–8.88) | 0.864      |
| Physiotherapy sessions                  | 4 (30.8)      | 9 (17.3)        | 1.96 (0.45–7.60) | 0.825      |
| Isolation measures                      | 0 (0.0)       | 13 (25.0)       | 0.06 (0.00–0.59) | 0.006      |
| Diarrhoea present in cases              | 4/8 (50.0)    | 22 (42.3)       | 1.16 (0.25–5.31) | 0.579      |
| Isolation measures for cases            | 1/8 (12.5)    | 17 (32.7)       | 0.29 (0.02–1.83) | 0.100      |

**Quantitative variables: median (first and third quartiles)**

| Variable                                | Median (first and third quartiles) | (95% CI)       | Prob       |
|-----------------------------------------|-----------------------------------|----------------|------------|
| Age                                     | 71 (63–79)                        | 1.01 (1.00–1.04) | 0.669      |
| Number of hospitalisation days in the year | 68 (57–149)                   | 1.02 (1.01–1.03) | 0.999      |
| Nephrology                              | 0 (0–49)                          | 1.02 (1.00–1.04) | 0.980      |
| Number of hospitalisation days in the month | 28 (16–31)                   | 1.01 (0.96–1.07) | 0.704      |
| Nephrology                              | 2 (0–9)                           | 1.02 (0.96–1.08) | 0.725      |
| Number of haemodialysis sessions in the year | 0 (0–9)                         | 1.00 (0.92–1.07) | 0.486      |
| Number of haemodialysis sessions in the month | 0 (0–5)                        | 1.00 (0.90–1.10)** | 0.509      |

*Probability that the OR would be higher than one.

**Risk ratio (RR).
significant protective effect of CP was also found by Fossi Djembi et al. [17] but they have not made a genomic comparison of their strains. This observation should still increase our confidence and compliance regarding CP for controlling GRE outbreaks. Immunosuppression as a protective factor is a surprising result. Perhaps there could have been a sampling bias associated with the ward where controls came from. However, the controls were drawn randomly among contact patients of each case during the hospital path. Thus, these controls did not come from only one ward. Consequently an alternative hypothesis, pertaining to isolation measures (protective isolation or contact protection), must be evoked. It is worth mentioning that among the 15 immunosuppressed controls, only six of them had isolation measures. The comparison with the only immunosuppressed case (which did not have isolation measure) is thus not very relevant. Nevertheless, isolation measures do not seem to be an explanation of the protective effect. Lastly, to evaluate the stability of our results due to the small number of immunosuppressed patients, we ran a sensitivity analysis by using different alternative prior distributions for the OR. Surprisingly, the use of either an optimistic informative prior, a pessimistic informative prior or a lowly informative prior had only little impact on the result, while the addition of a single event among the cases (2/13 instead of 1/13), as part of this sensitivity analysis, led to a disappearance of the effect, suggesting maybe that we lack data to ascertain this result.

Certain parameters showed an effect in the univariate analysis, according to our model’s construction algorithm (Pr ≥ 0.850 or Pr ≤ 0.150), but were not included in the multivariate analysis. This is the case for parameters linked to nephrology, already documented in the literature and in the description of this outbreak as risk factors for carrying GRE. We made this choice in order to limit the number of parameters in our multivariate analysis, considering our sample size.

It is possible that our study lacks evidence, as it did not allow us to reveal an association between our variable of interest and all of the study’s parameters that were selected due to their probable role in GRE transmission. We used a Bayesian approach in order to limit this bias. In fact, Bayesian analysis enables the use of prior information for the parameter of interest, built for instance on previous publications results. In the absence of information, we used a Gaussian vague prior with parameter centred on the value 0 with a wide standard deviation, N (0; 2.6). We used data from the literature to be included in the models. These data should be comparable and collected from the literature in a similar way as the data from the study in question. It was thus not always possible to find informative priors for each parameter.

Finally, the description of this outbreak highlighted the importance to detect quickly patients at risk of GRE carriage to implement CP as soon as possible. Indeed none of our cases were in isolation measures before their discovery and many cross-transmissions may have occurred before the knowledge of the carriage. During this period, the patients may have visited different wards, particularly Nephrology wards. After the GRE detection, the rapid transfer to a dedicated department (separated unit with dedicated HCW) no longer has allowed the cross-transmission what has led to the extinction of the outbreak. In France, patients defined at risk of GRE carriage are contact patients and those hospitalised abroad in the previous year. Upon admission, CP are implemented and patients are screened. Others patients do not have preventive measure with regard to the GRE [20].

In our study, we have shown that a history of hospitalisation and the intake of antibiotics and antacids during hospitalisation favour the acquisition of a hospital epidemic strain of GRE while isolation measures are logically protective. The Bayesian approach was particularly useful considering our limited sample size and given that some informative data were available in the literature. Early detection of GRE carriers in order to implement isolation measures and to transfer to dedicated department is an important factor for the management of an outbreak. Systematic search of factors associated with acquisition of GRE could lead to faster implementation of these measures.

**Supplementary material.** The supplementary material for this article can be found at https://doi.org/10.1017/S0950268818003655

**Author ORCIDs.** D. S. Debocker, 0000-0002-6225-9785

**Acknowledgements.** Thanks to healthcare personnel, bacteriology laboratory and National Reference Centre. This study received an approval opinion from the Ethics Committee of the hospital.

**Financial support.** This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

**Conflict of interest.** None.

**Ethical standards.** The authors assert that all procedures contributing to this work comply with the ethical standards of the relevant national and institutional committees on human experimentation and with the Helsinki Declaration of 1975, as revised in 2008.

**References**

1. Leclercq R and Coignard B (2006) Les entérocoques résistants aux glycopeptides: situation en France en 2005. *Bulletin Epidémiologique Hebdomadaire* 13, 85–87.
2. Henard S et al. (2011) Control of a regional outbreak of vanA glycopeptide-resistant *Enterococcus faecium*, Eastern France, 2004–2009. *International Journal of Hygiene and Environmental Health* 214, 265–270.
3. Bourdon N, Fines M and Leclercq R (2008) Caractéristiques des souches d’entérocoques résistants aux glycopeptides isolées en France, 2006–2008. *Bulletin Epidémiologique Hebdomadaire* 41–42, 391–394.
4. Stockholm: ECDC (2015) European Centre for Disease Prevention and Control. Antimicrobial resistance surveillance in Europe 2014. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net).

5. Hautemanière A et al. (2009) A prospective study of the impact of colonization following hospital admission by glycopeptide-resistant enterococci on mortality during a hospital outbreak. American Journal of Infection Control 37, 746–752.

6. Salgado CD and Farr BM (2003) Outcomes associated with vancomycin-resistant enterococci: a meta-analysis. Infection Control and Hospital Epidemiology 24, 690–698.

7. Hoshuyama T et al. (2008) Vancomycin-resistant enterococci (VRE) outbreak at a university hospital in Kitakyushu, Japan: case-control studies. Journal of Infection and Chemotherapy 14, 354–360.

8. Correa AAF et al. (2015) Small hospitals matter: insights from the emergence and spread of vancomycin-resistant enterococci in 2 public hospitals in inner Brazil. Diagnostic Microbiology and Infectious Disease 82, 227–233.

9. Martínez JA et al. (2003) Role of environmental contamination as a risk factor for acquisition of vancomycin-resistant enterococci in patients treated in a medical intensive care unit. Archives of Internal Medicine 163, 1905–1912.

10. McEvoy SP et al. (2006) Risk factors for the acquisition of vancomycin-resistant enterococci during a single-strain outbreak at a major Australian teaching hospital. Journal of Hospital Infection 62, 256–258.

11. MacIntyre CR et al. (2001) Risk factors for colonization with vancomycin-resistant enterococci in a Melbourne hospital. Infection Control and Hospital Epidemiology 22, 624–629.

12. Sakka V et al. (2008) Risk-factors and predictors of mortality in patients colonised with vancomycin-resistant enterococci. Clinical Microbiology and Infection 14, 14–21.

13. Servais A et al. (2009) Rapid curbing of a vancomycin-resistant Enterococcus faecium outbreak in a nephrology department. Clinical Journal of the American Society of Nephrology 4, 1559–1564.

14. Assadian O et al. (2007) Prevalence of vancomycin-resistant enterococci colonization and its risk factors in chronic hemodialysis patients in Shiraz, Iran. BMC Infectious Diseases 7, 52.

15. Karki S et al. (2012) Prevalence and risk factors for VRE colonisation in a tertiary hospital in Melbourne, Australia: a cross sectional study. Antimicrobial Resistance and Infection Control 1, 31.

16. Roghmann M et al. (1998) Colonization with vancomycin-resistant enterococci in chronic hemodialysis patients. American Journal of Kidney Diseases 32, 254–257.

17. Fossi Djembi L et al. (2017) Factors associated with vancomycin-resistant Enterococcus acquisition during a large outbreak. Journal of Infection and Public Health 10, 185–190.

18. Amberpet R et al. (2016) Screening for intestinal colonization with vancomycin-resistant enterococci and associated risk factors among patients admitted to an adult intensive care unit of a large teaching hospital. Journal of Clinical and Diagnostic Research 10, DC06–DC09.

19. Monteserin N and Larson E (2016) Temporal trends and risk factors for healthcare-associated vancomycin-resistant enterococci in adults. Journal of Hospital Infection 94, 236–241.

20. Instruction DGOS/PF2/DGS/R11 n°2014-08 du 14 janvier 2014 relative aux recommandations pour la prévention de la transmission croisée des bactéries hautement résistantes aux antibiotiques émergents.

21. Ury HK (1975) Efficiency of case-control studies with multiple controls per case: continuous or dichotomous data. Biometrics 31, 643–649.

22. Gelman A et al. (2013) Bayesian Data Analysis, 3rd Edn. Boca Raton: Chapman and Hall/CRC.

23. Hoff PD (2009) A First Course in Bayesian Statistical Methods. New York, NY: Springer New York.

24. Gilbert EM et al. (2017) Factors contributing to vancomycin-resistant Enterococcus spp. horizontal transmission events: exploration of the role of antibacterial consumption. Diagnostic Microbiology and Infectious Disease 89, 72–77.

25. Cetinkaya Y, Falk PS and Mayhall CG (2002) Effect of gastrointestinal bleeding and oral medications on acquisition of vancomycin-resistant Enterococcus faecium in hospitalized patients. Clinical Infectious Diseases 35, 935–942.

26. Byers KE et al. (2001) A hospital epidemic of vancomycin-resistant enterococcus: risk factors and control. Infection Control of Hospital Epidemiology 22, 140–147.