Using Halothermal Time Model to Describe Barley (Hordeum vulgare L.) Seed Germination Response to Water Potential and Temperature

Abd Ullah 1,2,3,4,†, Sadaf Sadaf 5,‡, Sami Ullah 5,* , Huda Alshaya 6, Mohammad K. Okla 7, Yasmeen A. Alwasel 7,§ and Akash Tariq 1,2,3,4,★

Abstract: Barley (Hordeum vulgare L.) is a salt-tolerant crop with considerable economic value in salinity-affected arid and semiarid areas. In the laboratory experiment, the halothermal time (HaloTT) model was used to examine barley seed germination (SG) at six constant cardinal temperatures (Ts) of 15, 20, 25, 30, 35, and 40 °C under five different water potentials (Ψs) of 0, −0.5, −1.5, −1.0, and −2.0 MPa. Results showed that at optimum moisture (0 MPa), the highest germination percentage (GP) was recorded at 20 °C and the lowest at 40 °C. Moreover, GP increased with the accelerated aging period (AAP) and significantly (p ≤ 0.05) decreased with high T. In addition, with a decrease of Ψ from 0 to −0.5, −1.5, and −2.0 MPa, GP decreased by 93.33, 76.67, 46.67, and 33.33%, respectively, in comparison with 0 MPa. The maximum halftime constant (ΨHalo) and coefficient of determination (R2) values were recorded at 20 °C and 30 °C, respectively. The optimum temperature (Tb) for barley is 20 °C, base Ψ of 50th percentile (Ψb (50)) is −0.23 Mpa, and standard deviation of Ψb (s/Ψb) is 0.21 MPa. The cardinal Ts for germination is 15 °C (Tb), 20 °C (Td), and 40 °C (To). The GP, germination rate index (GRI), germination index (GI), coefficient of the velocity of germination (CVG), germination energy (GE), seed vigor index I and II (SVI-I & II), Timson germination index (GI), and root shoot ratio (RSR) were recorded maximum at 0 MPa at 20 °C and minimum at −2.0 MPa at 40 °C. Mean germination time (MGT) and time to 50% germination (T 50%) were maximum at −2 MPa at 40 °C, and minimum at 20 °C, respectively. In conclusion, the HaloTT model accurately predicted the germination time course of barley in response to T, Ψ, or NaCl. Therefore, barley can be regarded as a salt-tolerant plant and suitable for cultivation in arid and semi-arid regions due to its high resistance to salinity.

Keywords: barley; seed germination; halothermal time model; cardinal temperatures; water potential

1. Introduction

Barley (Hordeum vulgare L. Poaceae.) is an important annual cereal crop worldwide and ranks fourth in production [1]. It is used for various purposes, including human food, brewing materials, animal feed, and bedding [2]. Barley can withstand different environmental stress factors such as flood, salinity, and water stress, and it is even more...
tolerant than wheat under any unsuitable ecological conditions [3]. However, the ability to cope with stress factors largely depends on the intensity of the abiotic stress factors and the stage of the plant’s growth. Likewise, the salt tolerance of barley varies depending on its growth phase as it is most sensitive to salinity during seed germination and the initial stages of seedling development, but as it grows older, it becomes more tolerant to salinity. Besides, salt stress has been linked to ionic rather than osmotic effects [4,5].

Due to the release of atmospheric CO$_2$ and other warming gases, climate change can increase global temperatures by 2.5–4.5 °C [6]. Due to the link between temperature, dormancy, and germination, an increase in temperature can have a detrimental effect on plant species’ emergence and establishment. Temperature is one of the most important factors influencing seed germination (SG) [7–9]. Several plant species undergo seed germination as the first phase of their life cycle [9]. It is a complex physiological process that is affected by environmental stress factors, including temperature (T), water availability, salinity, light, and chemical materials [10–16]. It has been indicated, however, that temperature and water availability are the main factors influencing SG by affecting seed dormancy, enzymatic activity, hormonal biosynthesis, and translocation of reserve store materials [7,8,17,18]. Furthermore, the ability of the seed to absorb water also determines the success or failure of SG; however, this is determined by soil moisture levels [12].

It has been demonstrated that there are three cardinal temperatures (Ts), which act like three checkpoints that can strictly evaluate the response of SG to temperature in a variety of plant species [12]. They are: (a) The minimum or base temperature ($T_b$); SG will cease at $T_b$ below); (b) the optimum temperature ($T_o$; SG will commence promptly); and (c) the maximum or ceiling temperature ($T_c$); SG will cease at $T_c$ above. However, the Ts may differ among plant species and under different climatic conditions [19]. If, for instance, the increase in temperature exceeds the $T_c$ for a given species of plant, then the species will no longer be able to germinate, resulting in a threat to its survival and establishment. Previous studies have reported that the germination rate (GR) increases as the temperature increases between $T_b$ and $T_o$, but decreases when the temperature increases above $T_o$ [11,20–22].

In general, GP and GR increase as moisture availability increases but decline under conditions of negative water potential ($\psi$) [21,23]. Additionally, salt stress affects SG due to osmotic or ion toxicity effects [24–26] or a combination of both effects [27]. However, despite the importance of SG under salinity [28,29], the mechanism(s) of salinity tolerance in seeds is still little studied, particularly when compared to the body of knowledge currently available for salinity tolerance in vegetative plants [30–34]. Dissolved salt ions reduce the $\psi$ of salt solutions. For instance, a 1 M NaCl solution has a $\psi$ of $-4.4$ MPa at 25 °C [35]. Moreover, accumulations of Na$^+$ and Cl$^-$, as well as osmoprotectants, result in osmotic adjustment or a decrease in $\psi$ inside the seed cells, allowing SG to occur at lower concentrations of $\psi$ [36,37]. Consequently, changes in T, $\psi$, and salinity influence SG independently and interdependently, affecting the distribution and production of economic crop species [13]. To date, several mathematical models have been developed to illustrate the relationship between SG and T for many crop plants using thermal time models (TT) and hydrot time models (TH) [11,38–40].

Researchers have previously used a hydro-time model to evaluate the impact of $\psi$ on seed germination at a given temperature [41,42]. According to this model, a negative relationship exists between the time to germination ($t_g$) and the difference between the $\psi$ of the seed environment and the physiological $\psi$ threshold for radicle emergence (base $\psi$ or $\psi_b$), which differ among seeds in the population.

In addition, Gummerson’s [41] hydrothermal time (HTT) model for SG suggests that when $T_s \leq T_o$ (i.e., between $T_b$ and $T_o$) and at any given $\psi$, the $t_g$ of any germination percentile of a seed population is a function of the degree to which T and $\psi$ exceed their respective base values, $T_b$ and $\psi_b$, at which germination is hindered [41,43].

Furthermore, one study [38] used the same technique to calculate the SG attributes of the halophyte Suaeda maritime at different NaCl concentrations at sub-optimal Ts. They proposed a halothermal time (HaloTT) model which substitutes log NaCl for $\psi$ in the
HTT model. Therefore, the germination performance at sub-optimal Ts is shown to be affected by the salinity level threshold distribution (NaCl b(g)) relative to the NaCl of the surrounding environment. In light of increasing global temperatures and salinity, as well as the ever-increasing number of people on earth, it is imperative to study seed germination and early seedling establishment of economic crop plants to abiotic stress factors. Based on its broad stress acclimation, we sought to examine the response characteristics of barley to T, Ψ, and salinity using the HaloTT model. This study aimed to investigate how T and salinity affect the germination features of barely seeds using the HaloTT model and estimate the cardinal Ts and the salt tolerance threshold of barley.

2. Methodology

2.1. Seeds Germination and Experiment Protocol

The barley (AJJ variety) seeds were generously provided by the Nuclear Institute for Food and Agriculture (NIFA), Peshawar Pakistan. The seeds (95% viability rate) of the same size and shape were surface sterilized with 95% ethanol solution for 3 min and then rinsed with distilled water and shade dried at room temperature [44]. A randomized complete block design (RCBD) Petri dish experiment was performed at the Plant Physiology Laboratory, Department of Botany, University of Peshawar, Khyber Pakhtunkhwa, Pakistan. Halothermal time (HaloTT) experiments were conducted at six constant temperatures of 15, 20, 25, 30, 35, and 40 °C with five water potentials of 0, −0.5, −1.0, −1.5, and −2 MPa. There were ten seeds per Petri dish surface covered with Whatman No. 1 filter paper and three replicates for each treatment. For the stress-treated Petri dishes, 5 mL of NaCl solution was used and for the control Petri dishes, distilled water. The Petri dishes were placed in an incubator (Memmert Beschickung-Loading-Model 100-800, Schwabach, Germany) in the dark except for the reading times. The seeds were periodically examined and considered to have germinated when the radicle reached a length of 1 mm. In the end, seeds were removed and evaluated for their germination characteristics. The germination time course data were analyzed, and different parameters were determined for the thermal time, halothermal, and halo-thermal time models using repeated probit regression analysis as described previously [45,46].

2.2. Thermal Time Model (TT)

Sub (TTsub) and supra optimal (TTsupra) cardinal Ts were derived from the formulas based on the halo thermal time model, which is given below,

\[ TT_{sub} = (T - T_b) \theta_{halo} \text{ at sub-optimal } T \]  

\[ TT_{supra} = (T_{c(g)} - T_b) \theta_{halo} \text{ at supra-optimal } T \]

2.3. Halotime Model (HT)

The proposed halo-time model (θHalo) is used to enhance model prediction. θHalo determines the connections between solute potential and germination rate in the same manner as the thermal time model does:

\[ \theta_{Halo(g)} = (NaCl(g) - NaCl_b) \theta_{halo} \text{ (3)} \]

Or

\[ \text{Probit (g)} = [NaCl - (\theta_{Halo}/\theta_{halo}) - NaCl_b 50]/\sigma NaCl_b \]  

2.4. Halothermal Time Model (HaloTT)

For HaloTT model, using the model suggested by Seal et al. [37],

\[ \theta_{HaloTT} = (NaCl(g) - NaCl_b) (T - T_b) \theta_{halo} \]

\[ (5) \]
Or

\[ \text{Probit (g)} = \frac{\left(\text{NaCl} \left(\theta \text{HaloTT}/(T - T_b)\right) \times \text{tg} - \text{NaCl}_{b(50)}\right)}{\sigma \text{NaCl}_b} \]  

Equation seven is the modified form of equation five for further analysis.

\[ \theta \text{HaloTT} = \left[\text{NaCl}_{b(g)} - \text{NaCl} - (kT (T - T_o))\right] (T - T_b) \times \text{tg} \]  

Or

\[ \text{Probit (g)} = \frac{\left[\text{NaCl} - kT (T - T_o) - (\theta \text{Halo}/ (T - T_b) \times \text{tg}) - \text{NaCl}_{b(50)}\right]}{\sigma \text{NaCl}_b} \]  

2.5. Germination Parameters

The below-mentioned germination indices were calculated from the germination rate, physical observation, seed weight, root and shoot lengths, leaf length, fresh and dry weights of the plants.

2.5.1. Germination Percentage (GP)

GP represents the total number of seeds germinated out of the total seeds sown in each Petri dish. This germination parameter was calculated using the formula [47].

\[
\text{Germination percentage (GP)} = \frac{\text{Final number of seedlings emerged}}{\text{Total number of seeds sown}} \times 100
\]  

2.5.2. Mean Germination Time (MGT)

The MGT index showed that how fast the seeds emerged in a population. Small MGT value means seed population has a high rate and vice versa. This was calculated using the following formula [47].

\[
\text{Mean germination time (MGT)} = \frac{\sum f_X}{f}
\]  

where \( f \) is the number of seeds germinated on day \( X \).

2.5.3. Germination Rate Index (GRI)

The GRI represents the percent germination on respective days and times. It is calculated by using the following formula [48].

\[
\text{Germination rate index (GRI)} = \frac{G_1}{1} + \frac{G_2}{2} + \frac{G_3}{3} \ldots + \frac{G_x}{x}
\]  

where \( G_1 \) and \( G_2 \) are the percent germinations on the first and second day after sowing and \( G_x \) is the final germination percentage on the last day.

2.5.4. Germination Index (GI)

The germination index tells us about the germination percentage and speed of germination. GI was calculated following the standard methodology [49].

\[
\text{Germination index (GI)} = (10 \times n_1) + (9n \times n_2) \ldots (1n \times 10)
\]  

where \( n_1, n_2 \ldots n_{10} \) showed the frequency of germinated seeds on first, second, and respective days till last day.

2.5.5. Coefficient of the Velocity of Germination (CVG)

The CVG represents the velocity of germination of seeds in an experiment, which will increase with an upsurge in the frequency of germinated seeds. The highest theoretical CVG value will be obtained when all sown seeds grow on the first day. This is calculated using the formula [50].
Coefficient of the velocity of germination (CVG) = \( \frac{N1 + N2 + N3 \ldots Nx}{100} \times N1T1 \ldots NxTx \)  
(13)
in which N is the frequency of seeds germinating every day and T represents the time from sowing to germination of seed N.

2.5.6. Germination Energy (GE)

Plant germination energy was calculated by using the following formula [51].

Germination energy (GE) = \( \frac{X1}{Y1} + \frac{(X2 - X1)}{Y2} + \frac{(Xn - Xn - 1)}{Yn} \)  
(14)
Here X_1, X_2, and X_n are the frequency of emerged seeds on the first day, second, and so on. While Y_1, Y_2, and Y_n are the days from sowing to first, second, and up to last day count.

2.5.7. Timson Germination Index (TGI)

The TGI index represents the average number of seeds germinated per day. This is measured from its mathematical formula as follows [52].

Timson germination index (TGI) = \( \frac{ti + (N/2 - ni)(tj - ti)}{(nj - ni)} \)  
(15)
where G is the total percentage of germination per day and T time of germination.

2.5.8. Mean Germination Rate (MGR)

Mean germination rate is the reciprocal of mean germination time. It was found out through the following formula [53].

Mean germination rate (MGR) = \( \frac{1}{\text{Mean Germination Time}} \)  
(16)

2.5.9. Seed Vigor Index-I (SVI-I)

The length of three seedlings from each pot was measured in cm and then calculated in the following formula [54].

Seed Vigor Index = Seedlings length(cm) \times Seed Germination %age  
(17)

2.5.10. Seed Vigor Index-II (SVI-II)

The dry weight of three seedlings from each pot was determined through electrical balance. The values were put in the formula and multiplied with seed germination percentage. The formula is as follows [55].

Seed Vigor Index = Seed dry weight (mg) \times Seed Germination %age  
(18)

2.5.11. Time to 50% Germination (T50%)

This index was developed to find out the time required for 50% seed germination. This is reported through the following mathematical formula [56].

Time to 50% germination (T50%) = \( \frac{ti + (N/2 - ni)(tj - ti)}{(nj - ni)} \)  
(19)
where N final number of seeds emerged, nj and ni are the cumulative numbers of seeds emerged after adjacent counts during tj and ti, when ni < N/2 > Nj.
2.5.12. Root-Shoot Ratio (RSR)

The RSR ratio was recorded after the root and shoot were dried in an oven for 24 h. Which is then calculated through the following formula [53].

\[
\text{Root – shoot ratio} = \frac{\text{root dry weight}}{\text{shoot dry weight}}.
\] (20)

2.6. Data Analysis

The investigation of temperatures (thermal time), water potentials (halftime), and their interactions (halothermal time model) on seed germination rate and germination attributes were analyzed through analysis of variance (ANOVA) using IBM SPSS Statistics 26. ANOVA was applied using three replicates of each treatment and germination parameters. The basic statistical calculation was performed in excel software. The values of the following parameters: \(\Psi_{b(50)}\), \(\sigma_{\Psi_b}\), \(R^2\), Sig, and F were determined using linear probit regression analysis in SPSS. ORIGIN2021 PC Corporation was used for plotting various graphs of germination fraction vs. accelerated aging period and germination parameters against T and \(\Psi\).

3. Results

The water potential (\(\Psi\)), temperature (T), and their interaction (T \(\times\) \(\Psi\)) significantly \((p \leq 0.05)\) affected germination percentage (GP) and germination rate (GR) (Figure 1). A maximum GP was recorded at 20 °C and a minimum at 40 °C at optimum moisture (0 MPa; control). Accordingly, minimum germination of 3.33% was reported at 40 °C under \(-2.0\) MPa and a maximum of 76.67% at 20 °C under \(-0.5\) MPa compared to the control (0 MPa). This indicates that GP reduced with the reduction in \(\Psi\) at each T (Figure 1a–f). Moreover, GP was recorded at a maximum after the fourth day at 0 MPa. GP generally increased with accelerated aging (AAP) and decreased significantly \((p < 0.05)\) with high T. Further, with the decrease of \(\Psi\) from 0 to \(-0.5\), \(-1\), \(1.5\), and \(-2.0\) MPa, GP decreased by 93.33, 76.67, 46.67, and 33.33% in comparison with the control (0 MPa), respectively (average for all levels of AAP) (Figure 1b).

The maximum halothermal time constant (\(\theta_{\text{Halo}}\)) and \(R^2\) values were recorded at 20 °C and 30 °C, respectively (Table 1, Figure 2). Compared with 0 MPa, the highest \(T_{\text{Ts}}\) and \(T_{\text{Tsupra}}\) values were observed at 20 °C at \(-0.5\) MPa and decreased with decreasing \(\Psi\) (\(-2.0\) MPa). In addition, GR (g) values show a significant \((p \leq 0.01)\) increase with the decrease in \(\Psi\) (more negative) at all Ts (Table 1). The standard deviation \(\sigma_{\Psi_b}\) values presented comparatively fewer fluctuations at all Ts. The highest \(\sigma_{\Psi_b}\) was recorded at 20 °C, and lowest was recorded at 40 °C (Table 2). Similarly, the highest base water potential at 50% germination (\(\Psi_{b(50)}\)) value was observed at 40 °C (\(-2.0\) MPa). Seeds exhibit a base or minimum temperature \((T_b)\) below which germination is decreased, an optimum temperature \((T_o)\) at which germination is most rapid, and a maximum or ceiling temperature \((T_c)\) at which germination is prevented. Germination at suboptimal T can be characterized based on thermal time, or the T in excess of \(T_b\) multiplied by the time to a given germination percentage (tg).

The minimum temperature \((T_b)\) for barley observed from our experiment is 15 °C, below which the germination rate decreases, and it will become difficult for a plant to continue its physiological processes (Table 3). The optimum temperature \((T_o)\) at which barley germination was maximum was 20 °C. The maximum or ceiling temperature \((T_c)\) above which plants cannot continue their physiological and biochemical activities was 40 °C.
Figure 1. Cumulative germination for *Hordeum vulgare* L. at (a) 15 °C, (b) 20 °C, (c) 25 °C, (d) 30 °C, (e) 35 °C, and (f) 40 °C having different water potentials. Symbols indicate water potential and lines indicate cumulative germination.

The results obtained from the recent HaloTT experiment revealed that temperature and NaCl significantly influenced the germination parameters of barley (*Hordeum vulgare* L.). The germination percentage (GP), germination rate index (GRI), and germination index (GI) were maximum in seeds grown at 20 °C in 0 MPa distilled water and minimum at 40 °C in −2.0 MPa (Figure 3a–d). Mean germination time (MGT) was recorded maximum at 40 °C in −2.0 MPa and minimum at 20 °C in −2.0 MPa. Figure 4a–d demonstrates the highest and lowest coefficient of the velocity of germination (CVG), germination energy (GE), Timson germination index (TGI), and mean germination rate (MGR) were recorded at 20 °C and 40 °C in 0 MPa and −2.0 MPa respectively. Figure 5a–d shows that seed vigor index I (SVI-I), seed vigor index II (SVI-II), and root shoot ratio (RSR) were recorded as maximum in distilled water (0 MPa) at 20 °C while the minimum is −2.0 MPa at 40 °C. The
maximum value of time to 50% germination (T 50%) was recorded in –2.0 MPa at 40 °C, and the minimum value was recorded at 20 °C.

Table 1. The estimated parameters of the halo and thermal time model to describe *Hordeum vulgare* L. seed germination under different temperatures (Ts) and water potentials (ψs).

| T (°C) | ψ (MPa) | TTsub (θT1) | TTsupra (θT2) | θHalo (MPa h) | θHaloTT (MPa h) | Halo Time (GR(θ)) | Thermal Time (GR(θ)) |
|--------|---------|-------------|---------------|--------------|-----------------|-----------------|------------------|
| 15     | 0       | 768         | 1920          | 38.4         | 384             | 0.013           | 0.013            |
|        | −0.5    | 544         | 1360          | 27.2         | 272             | 0.018           | 0.018            |
|        | −1.0    | 448         | 1120          | 22.4         | 224             | 0.022           | 0.022            |
|        | −1.5    | 448         | 1120          | 22.4         | 224             | 0.022           | 0.022            |
|        | −2.0    | 960         | 2400          | 4.8          | 48.0            | 0.104           | 0.104            |
|        | 0       | 896         | 2240          | 44.8         | 448             | 0.011           | 0.011            |
|        | −0.5    | 736         | 1840          | 36.8         | 368             | 0.014           | 0.014            |
| 20     | −1.0    | 640         | 1600          | 32.0         | 320             | 0.016           | 0.016            |
|        | −1.5    | 448         | 1120          | 22.4         | 224             | 0.022           | 0.022            |
|        | −2.0    | 128         | 320.0         | 6.40         | 64.0            | 0.078           | 0.078            |
|        | 0       | 640         | 1600          | 32.0         | 320             | 0.016           | 0.016            |
|        | −0.5    | 448         | 1120          | 22.4         | 224             | 0.022           | 0.022            |
| 25     | −1.0    | 320         | 800.0         | 16.0         | 160             | 0.031           | 0.031            |
|        | −1.5    | 320         | 800.0         | 16.0         | 160             | 0.031           | 0.031            |
|        | −2.0    | 96.0        | 240.0         | 4.80         | 48.0            | 0.104           | 0.104            |
|        | 0       | 576         | 1440          | 28.8         | 288             | 0.017           | 0.017            |
|        | −0.5    | 448         | 1120          | 22.4         | 224             | 0.022           | 0.022            |
| 30     | −1.0    | 352         | 880.0         | 17.6         | 176             | 0.028           | 0.028            |
|        | −1.5    | 224         | 560.0         | 11.2         | 112             | 0.045           | 0.045            |
|        | −2.0    | 96.0        | 240.0         | 4.80         | 48.0            | 0.104           | 0.104            |
|        | 0       | 256         | 640.0         | 12.8         | 128             | 0.039           | 0.039            |
|        | −0.5    | 416         | 1040          | 20.8         | 208             | 0.024           | 0.024            |
| 35     | −1.0    | 320         | 800.0         | 16.0         | 160             | 0.031           | 0.031            |
|        | −1.5    | 320         | 800.0         | 16.0         | 160             | 0.031           | 0.031            |
|        | −2.0    | 96.0        | 240.0         | 4.80         | 48.0            | 0.104           | 0.104            |
|        | 0       | 224         | 560.0         | 11.2         | 112             | 0.045           | 0.045            |
|        | −0.5    | 96.0        | 240.0         | 4.80         | 48.0            | 0.104           | 0.104            |
| 40     | −1      | 96.0        | 240.0         | 4.80         | 48.0            | 0.104           | 0.104            |
|        | −1.5    | 64.0        | 160.0         | 3.20         | 32.0            | 0.156           | 0.156            |
|        | −2.0    | 32.0        | 80.0          | 1.60         | 16.0            | 0.313           | 0.313            |

Temperatures (T); water potential (ψ); thermal time constant at sub-optimal temperature (TTsub); thermal time constant at supra-optimal temperature (TTsupra); halotime constant (θH); halothermal time constant (θHTT); germination rate (GR).
Table 2. Predictable values of $R^2$, $\psi_{b(50)}$ and $\sigma_{\psi_b}$ using halo thermal time (HaloTT) model to describe *Hordeum vulgare* L. seed germination under different $T_s$ and $\psi_s$.

| Barley | $T$ (°C) | $\psi_{b(50)}$ (MPa) | $\sigma_{\psi_b}$ (MPa) | $R$  | $R^2$ | $T$  | Sig. |
|--------|---------|----------------------|-------------------------|------|-------|------|------|
| AAJ    | 15      | -0.20                | 0.13                    | 0.940| 0.884 | 18.47| Sig  |
|        | 20      | -0.23                | 0.21                    | 0.976| 0.953 | 19.42| 0.018|
|        | 25      | -0.17                | 0.12                    | 0.965| 0.930 | 13.36| 0.004|
|        | 30      | -0.13                | 0.9                    | 0.999| 0.998 | 27.78| 0.008|
|        | 35      | -0.10                | 0.8                    | 0.555| 0.308 | 7.161| 0.000|
|        | 40      | -0.8                | 0.6                    | 0.901| 0.812 | 11.64| 0.331|

$R$ and $R^2$ is the coefficient determination, $\sigma_{\psi_b}$ is the standard deviation. $\psi_{b(50)}$ is base water potential at 50 percentiles, $\theta_H$ is halotime constant, $F$ is variability between different means, Sig. is significant value.

Figure 3. Interactive effect of salinity and temperature on (a) germination percentage (b) mean germination time (c) germination rate index (d) germination index of barley (*Hordeum vulgare* L.) var. AAJ using halo thermal time (HaloTT) model.
Table 3. Estimated values of $k_T$, $\sigma_{\psi_b}$, and $T_o$ using halothermal time model (HaloTT) for describing seed germination of *Hordeum vulgare* L. under different $T$s and $\psi$s.

| Variables | Hordeum vulgare L. |
|-----------|-------------------|
| Halothermal time model parameters | |
| $\psi_{b(50)}$ (MPa) | −0.23 |
| $\sigma_{\psi_b}$ (MPa) | 0.21 |
| $\theta_{\text{Halo}}$ (MPa °C h$^{-1}$) | 17.65 |
| $k_T$ (MPa °C h$^{-1}$) | 0.104 |
| Cardinal temperatures | |
| $T_b$ (°C) | 15 |
| $T_o$ (°C) | 20 |
| $T_c$ (°C) | 40 |
| $R^2$ | 0.953 |

$\psi_{b(50)}$ = base water potential at 50 percentiles, $T_b$ = base temperature, $T_o$ = optimum temperature, $T_c$ = ceiling temperature.

Figure 4. Interactive effect of salinity and temperature on (a) coefficient of the velocity of germination (b) germination energy (c) Timson germination index (d) mean germination rate of barley (*Hordeum vulgare* L.) var. AAJ using HaloTT model.
Figure 5. Interactive effect of salinity and temperature on (a) seed vigor index-I (b) seed vigor index-II (c) time to 50% germination (d) root shoot ratio of barley (Hordeum vulgare L.) var. AAJ using HaloTT model.

4. Discussion

The HTT model, also known as population-based threshold models, accurately reflects the observed responses of GRs and GPs to stress, age, hormones, and other factors that affect seed germination. An investigation of seed germination under varying environmental conditions will better understand germination parameters and identify the most appropriate geographic location for a species to emerge and establish [11]. The results of the current halothermal time model revealed that water potential ($\Psi$), temperature ($T$), and their interaction ($T \times \Psi$) significantly affect the germination rate and germination percentage. In comparison with control (0 MPa), the minimum germination was reported at 40 $^\circ$C under $-2.0$ MPa, while the maximum was reported at 20 $^\circ$C under $-0.5$ MPa, indicating that GP decreased as T decreased (Figure 1a–f). These findings are in agreement
with the previous studies that have indicated that temperature is a critical factor that negatively impacts seed germination by influencing the GP and GR in a wide range of plant species [16,18,57]. In the present study, GP increased with the accelerated aging period (AAP) and significantly \((p \leq 0.05)\) decreased with high temperature. Additionally, stress related to \(\Psi\) is another major environmental factor that limits SG and the early establishment of seedlings [39,58,59]. Furthermore, when \(\Psi\) is reduced from 0 to \(-0.5, -1, 1.5, \) and \(-2\) Mpa, GP decreases by 93.33, 76.67, 46.67, and 33.33%, respectively, as compared to the control (0 Mpa) (average for all levels of AAP) (Figure 1b). Previous findings also reported that the longer AAP and lower \(\psi\) (more negative) decreased GP and GR in various crop species [40,43].

Further, the \(\theta H\) values increased with an increase in cardinal \(Ts\) up to \(To\) and then linearly decreased with a decrease in \(T > To\). Previous studies have also demonstrated that the values of \(\theta H\) have increased at suboptimal temperatures for potato [57] and watermelon [12,40]. Similarly, the results of the base water potential at the 50th percentile (\(\psi_b (50)\)) have also been shown to increase (become more positive) at supraoptimal temperatures [20,23]. In a previous study [16] both GP and GR decreased with decreasing \(\psi\) and increasing NaCl at each tested \(T\).

In the present study, we found that the GR\( (g)\) values increased significantly \((p \leq 0.01)\) with decreasing \(\Psi\) (more negative) at all cardinal temperatures \(Ts\) (Table 1). GR decreased when water potential was reduced compared to the control. The effect of \(\psi\) on GP and GR was greater than that of AAP [42]. In the present study, the minimum temperature \((T_b)\) for the studied plants was 15 °C, below which the GR decreased. In addition, the optimum temperature \((T_o)\) for barley germination was 20 °C, whereas the limiting temperature \((T_c)\) beyond which plants could not continue their physiological and biochemical activities was 40 °C. The previous finding also observed that the temperature spectrum for seed germination contains three cardinal temperatures \((Ts)\), which are crucial in determining the seed germination characteristics [12,19].

Further, we found that the germination percentage and other characteristics were maximum at 0 Mpa (distilled water) at 20 °C, while the minimum was obtained at \(-2.0\) Mpa at 40 °C. The maximum time to 50% germination and mean germination time was observed in \(-2.0\) Mpa at 40 °C (Figure 3a–d, Figure 4a–d, and Figure 5a–d). It has also been reported previously that temperature is a key factor that affects both GP and the GR [12,18,60,61]. Moreover, salinity may limit SG by osmotic and ion-specific mechanisms [24,25]. Salinity, temperature \((T)\), and water potential \((\psi)\) all influence SG independently and together. The reason for this may be a result of \(Na^+\) and \(Cl^-\) ions entering into seed cells, reducing their osmotic potential and increasing embryonic turgor, which permits the seeds to germinate at lower \(\psi\) s [16].

5. Conclusions

In the present study, the best \(T\) and \(\Psi\) for barley are 20 °C and 0.0 Mpa, which indicates that GP and GR are significantly affected by \(T\), \(\Psi\), and their interactions with accelerated aging periods. The maximum \(\theta Halo\) and \(R^2\) values were recorded at 20 °C and 30 °C, respectively. Further, \(\Psi_b (50)\) is \(-0.23\) Mpa and \(\sigma \Psi_b\) is 0.21 Mpa at \(kT 0.104\) Mpa. For barley, the cardinal temperatures are recorded as \(T_b = 15 °C\), \(T_o = 20 °C\), and \(T_c = 40 °C\). Hence, the halothermal time (HaloTT) model accurately predicted the germination time course of Barley \((Hordeum vulgare\ L.)\) in response to various regimes of \(T\), \(\Psi\), or NaCl. Even though actual scenarios can vary according to field conditions, performance may not exactly follow the predicted model. Yet, a recent study [62] used the HTT model and concluded that the germination sensitivity to \(T\) and \(\Psi\) of 13 native desert annual plant species were highly correlated with seedling emergence in Arizona’s actual desert field conditions. Accordingly, we believe that the HaloTT model developed in this study can quantify and predict the relative germination attributes expected in the actual field conditions.
Author Contributions: S.U. and A.U. designed the research study. A.U. and S.S. conducted the experiments and wrote the initial draft of the manuscript. S.U., A.T., M.K.O., Y.A.A. and H.A. reviewed and revised the manuscript. S.U. and A.T. supervised the study. All authors have read and agreed to the published version of the manuscript.

Funding: The authors extend their appreciation to the Researchers Supporting Project number (RSP-2021/374) King Saud University, Riyadh, Saudi Arabia.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: All the data are available in the manuscript.

Acknowledgments: We are thankful to the Nuclear Institute for Food and Agriculture (NIFA) for the provision of high viability rate barley seeds and the Department of Botany, the University of Peshawar for lab facilitation.

Conflicts of Interest: Authors have no conflict of interest.

References
1. Khan, S.; Hussain, W.; Shah, S.; Hussain, H.; Altayar, A.E.; Ashour, M.L.; Pieroni, A. Overcoming Tribal Boundaries: The Biocultural Heritage of Foraging and Cooking Wild Vegetables among Four Pathan Groups in the Gadoon Valley, NW Pakistan. Biology 2021, 10, 537. [CrossRef] [PubMed]
2. Sadras, V.O.; Calderini, D. Crop Physiology Case Histories for Major Crops; Academic Press: Cambridge, MA, USA, 2020.
3. Giraldo, P.; Benavente, E.; Manzano-Agugliaro, F.; Gimenez, E. Worldwide research trends on wheat and barley: A bibliometric comparative analysis. Agronomy 2019, 9, 352. [CrossRef]
4. Al-Karaki, G.N. Germination, sodium, and potassium concentrations of barley seeds as influenced by salinity. J. Plant Nutr. 2001, 24, 511–522. [CrossRef]
5. Walia, H.; Wilson, C.; Wahid, A.; Condamine, P.; Cui, X.; Close, T.J. Expression analysis of barley (Hordeum vulgare L.) during salinity stress. Funct. Integr. Genom. 2006, 6, 143–156. [CrossRef] [PubMed]
6. Solomon, S.; Manning, M.; Marquis, M.; Qin, D. Climate Change 2007-the Physical Science Basis: Working Group I Contribution to the Fourth Assessment Report of the IPCC; Cambridge University Press: Cambridge, UK, 2007; Volume 4.
7. Roberts, E. Temperature and seed germination. In Symposia of the Society for Experimental Biology; Company of Biologists: Cambridge, UK, 1988; pp. 109–132.
8. Milbau, A.; Graae, B.J.; Shevtsova, A.; Nijs, I. Effects of a warmer climate on seed germination in the subarctic. Ann. Bot. 2009, 104, 287–296. [CrossRef] [PubMed]
9. Shah, S.; Khan, S.; Bussmann, R.W.; Ali, M.; Hussain, D.; Hussain, W. Quantitative ethnobotanical study of Indigenous knowledge on medicinal plants used by the tribal communities of Gokand Valley, District Buner, Khyber Pakhtunkhwa, Pakistan. Plants 2019, 9, 1001.
10. Showler, A.T.; Shah, S.; Khan, S.; Ullah, S.; Degola, F. Desert Locust Epidose in Pakistan, 2018–2021, and the Current Status of Integrated Desert Locust Management. J. Integr. Pest Manag. 2022, 13, 1. [CrossRef]
11. Shah, S.; Ullah, S.; Ali, S.; Khan, A.; Ali, M.; Hassan, S. Using mathematical models to evaluate germination rate and seedlings length of chickpea seed (Cicer arietinum L.) to osmotic stress at cardinal temperatures. PloS ONE 2021, 16, e0260990. [CrossRef]
12. Bewley, J.D.; Black, M. Seeds: Physiology of Development and Germination; Springer Science & Business Media: Berlin /Heidelberg, Germany, 2013.
13. Baskin, C.C.; Baskin, J.M. Seeds: Ecology, Biogeography, And Evolution of Dormancy and Germination; Elsevier: Amsterdam, The Netherlands, 1998.
14. Atashi, S.; Bakhshandeh, E.; Zeinali, Z.; Yassari, E.; da Silva, J.A.T. Modeling seed germination in Melisa officinalis L. in response to temperature and water potential. Acta Physiol. Plant. 2014, 36, 605–611. [CrossRef]
15. Vahabina, F.; Pirdashi, H.; Bakhshandeh, E. Environmental factors’ effect on seed germination and seedling growth of chicory (Cichorium intybus L.) as an important medicinal plant. Acta Physiol. Plant. 2019, 41, 1–13. [CrossRef]
16. Bakhshandeh, E.; Bradford, K.J.; Pirdashi, H.; Vahabina, F.; Abdellaoui, R. A new halothermal time model describes seed germination responses to salinity across both sub-and supra-optimal temperatures. Acta Physiol. Plant. 2020, 42, 1–15. [CrossRef]
17. Shah, S.; Khan, S.; Sulaiman, S.; Muhammad, M.; Badshah, L.; Bussmann, R.W.; Hussain, W. Quantitative study on medicinal plants traded in selected herbal markets of Khyber Pakhtunkhwa, Pakistan. Ethnobot. Res. Appl. 2020, 20, 1–36. [CrossRef]
18. Parmoon, G.; Moosavi, S.A.; Siadat, S.A. How salinity stress influences the thermal time requirements of seed germination in Silybum marianum and Calendula officinalis. Acta Physiol. Plant. 2018, 40, 1–13. [CrossRef]
19. Hattfie, J.L.; Prueger, J.H. Temperature extremes: Effect on plant growth and development. Weather. Clim. Extrem. 2015, 10, 4–10. [CrossRef]
20. Rowse, H.; Finch-Savage, W. Hydrothermal threshold models can describe the germination response of carrot (Daucus carota) and onion (Allium cepa) seed populations across both sub-and supra-optimal temperatures. N. Phytol. 2003, 158, 101–108. [CrossRef]
52. Sinha, V.B.; Grover, A.; Yadav, P.V.; Pande, V. Salt and osmotic stress response of tobacco plants overexpressing Lepidium latifolium L. Ran GTPase gene. *Indian J. Plant Physiol.* 2018, 23, 494–498. [CrossRef]

53. Basit, A.; Khan, S.; Sulaiman, S.S.; Shah, A.A. Morphological features of various selected tree species on the greater university campus Peshawar, Pakistan. *Int. J. Bot. Stud.* 2019, 4, 92–97.

54. Shah, S.M.; Amin, M.; Gul, B.; Begum, M. Ethnobotanical, Elemental, and Phytochemical Evaluation of Five Plant Species of Lamiaceae in Peshawar, Pakistan. *Scientifica* 2020, 2020. [CrossRef]

55. Kumar, B.; Verma, S.; Ram, G.; Singh, H. Temperature relations for seed germination potential and seedling vigor in Palmarosa (Cymbopogon martini). *J. Crop Improv.* 2012, 26, 791–801. [CrossRef]

56. Salehzade, H.; IzadkhahShishvan, M.; Chiyasi, M. Effect of seed priming on germination and seedling growth of Wheat (Triticum aestivum L.). *J. Biol. Sci.* 2009, 4, 629–631.

57. Wang, H.; Zhao, K.; Li, X.; Chen, X.; Liu, W.; Wang, J. Factors affecting seed germination and the emergence of Aegilops tauschii. *Weed Res.* 2020, 60, 171–181. [CrossRef]

58. Wijewardana, C.; Alsajri, F.A.; Reddy, K.R. Soybean seed germination response to in vitro osmotic stress. *Seed Technol.* 2018, 39, 143–154.

59. Seepaul, D.D.R.; George, S.; Groot, J.; Wright, D. Drought tolerance classification of common oilseed species using seed germination assay. *J. Oilseed Brassica* 2019, 10, 97–105.

60. Alvarado, V.; Bradford, K. A hydrothermal time model explains the cardinal temperatures for seed germination. *Plant Cell Environ.* 2002, 25, 1061–1069. [CrossRef]

61. Ghaderi-Far, F.; Bakhshandeh, E.; Ghadirian, R. Evaluating seed quality in sesame (Sesamum indicum L.) by the accelerated ageing test. *Seed Technol.* 2010, 32, 69–72.

62. Liu, S.; Bradford, K.J.; Huang, Z.; Venable, D.L. Hydrothermal sensitivities of seed populations underlie fluctuations of dormancy states in an annual plant community. *Ecology* 2020, 101, e02958. [CrossRef]