Follicular helper T cells are required for systemic autoimmunity

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Production of high-affinity pathogenic autoantibodies appears to be central to the pathogenesis of lupus. Because normal high-affinity antibodies arise from germinal centers (GCs), aberrant selection of GC B cells, caused by either failure of negative selection or enhanced positive selection by follicular helper T (T<sub>FH</sub>) cells, is a plausible explanation for these autoantibodies. Mice homozygous for the san allele of Roquin, which encodes a RING-type ubiquitin ligase, develop GCs in the absence of foreign antigen, excessive T<sub>FH</sub> cell numbers, and features of lupus. We postulated a positive selection defect in GCs to account for autoantibodies. We first demonstrate that autoimmunity in Roquin<sup>san/san</sup> (sanroque) mice is GC dependent: deletion of one allele of Bcl6 specifically reduces the number of GC cells, ameliorating pathology. We show that Roquin<sup>san</sup> acts autonomously to cause accumulation of T<sub>FH</sub> cells. Introduction of a null allele of the signaling lymphocyte activation molecule family adaptor Sap into the sanroque background resulted in a substantial and selective reduction in sanroque T<sub>FH</sub> cells, and abrogated formation of GCs, autoantibody formation, and renal pathology. In contrast, adoptive transfer of sanroque T<sub>FH</sub> cells led to spontaneous GC formation. These findings identify T<sub>FH</sub> dysfunction within GCs and aberrant positive selection as a pathway to systemic autoimmunity.

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pathway and form germinal centers (GCs) (4). Within this microenvironment, B cells undergo somatic hypermutation (SHM) and isotype switching, resulting in the generation of memory B cells and long-lived plasma cells that secrete high-affinity antigen-specific IgG antibodies (5, 6). Selection of mutated high-affinity GC B cells depends on restimulation with antigen arrayed on follicular dendritic cells and provision of help by follicular T helper (T_{FH}) cells.

Because SHM has the potential to generate self-reactive antibodies (7), it has been long thought that aberrant selection within GCs represents a candidate pathway to the production of lupus-associated autoantibodies. Indeed, autoantibodies detected in SLE patients and mouse lupus models are generally high affinity and somatically mutated (7, 8). Exclusion of self-reactive B cells from GCs has been shown to be defective in SLE patients. Also, GCs have been shown to form spontaneously in several different mouse models of lupus (9), and these are rich in apoptotic cells displaying the antigenic targets of lupus autoimmunity (10, 11). Although SHM can occur outside GCs, this process is far less efficient (12, 13). Despite all of this circumstantial evidence, there is to date no definite proof that GCs and/or T_{FH} cells are directly required for the production of lupus autoantibodies or end-organ damage. In contrast, extrafollicular affinity maturation of autoantibodies to dsDNA in MRL^{lpr} mice (14, 15) and T-independent B cell activating factor of the TNF family–driven pathways to lupus have been demonstrated (16, 17).

Furthermore, the prevailing model is that within GCs, autoantibodies might arise because of defects in negative rather than positive selection, because GC B cells are programmed to undergo apoptosis by default if they do not receive T cell selection signals. This is consistent with evidence that centrocytes down-regulate apoptosis inhibitors such as Bcl-2 and Bcl-xL while up-regulating proapoptotic molecules such as Fas and Bim (18). Normally, a dedicated population of T_{FH} cells is thought to provide help during selection of GC B cells (19, 20), and indeed a correlation between increased numbers of T_{FH} cells and autoimmunity has been described in mouse models of lupus (21, 22), suggesting that defects in positive selection by T_{FH} cells might indeed lead to lupus. Although recent evidence has suggested that Th17 cells may be responsible for aberrant selection of self-reactive GC B cells and autoantibody formation in BXD2 mice (23), direct evidence that T_{FH} cells can drive autoimmunity has not been provided.

The sanroque strain was discovered from screening an ENU mutagenized mouse library for autoimmune regulators (24), and exhibits a lupus-like phenotype characterized by high-affinity anti-dsDNA antibodies, hypergammaglobulinemia, lymphopenopathy, splenomegaly, autoimmune thrombocytopenia, and glomerulonephritis with IgG-containing immune complex deposits. Both the autoimmunity and cellular characteristics of sanroque segregate with homozygosity for the san allele (M199R substitution) of Roquin (Roquin^{san/san}) (22). Roquin has been demonstrated to be a regulator of the stability T cell messenger RNAs. Development of autoimmunity in Roquin^{san/san} mice also correlates with spontaneous GC formation, which is largely driven by B cell–extrinsic factors (22). Roquin^{san/san} mice have a marked accumulation of T cells within the B cell follicles, and the T_{FH} subset is overrepresented within the CD4+ cell compartment.

In this paper, we report that dysregulation of the GC response through excessive formation of T_{FH} cells is responsible for autoimmunity in Roquin^{san/san} mice. Loss of one allele of Bcl6, the master transcriptional regulator of GCs (25, 26), significantly reduces spontaneous GC formation in Roquin^{san/san} mice and the lupus phenotype. Furthermore, deletion of Sap (Sh2d1a) from Roquin^{san/san} mice causes a dramatic reduction of T_{FH} cells and IL-21. Sap is a small adaptor protein necessary for signaling through the signaling lymphocyte activation molecule family cell-surface receptors that regulates signals downstream of the TCR. Roquin^{san/san} Sap^{-/-} CD4+ cells also express lower levels of ICOS than Roquin^{san/san} Sap^{+/+} cells. This results in abrogation of ANAs (including anti-dsDNA) and end-stage renal disease. These findings establish a causal pathway from the san allele of Roquin to excess T_{FH} formation, aberrant GC formation, and positive selection of pathogenic high-affinity autoantibodies to illuminate a novel pathway of lupus pathogenesis.

RESULTS

Heterozygous Bcl6 deficiency reduces spontaneous GCs and attenuates autoimmunity in Roquin^{san/san} mice

BCL6 has been shown to be the master transcriptional regulator of GC B cells (27). As BCL6 deficiency results in early mortality because of widespread inflammation, we investigated Bcl6^{+/−} mice for possible defects in the GC response. 8 d after sheep red blood cell (SRBC) immunization, the percentage of GC cells was more than fivefold lower in mice heterozygous for Bcl6 deficiency (2.3 ± 0.59% vs. 0.43 ± 0.46%; P = 0.0011; Fig. 1 a).

To determine whether Bcl6 heterozygosity could curtail the spontaneous GC response in Roquin^{san/san} mice, we compared the percentage of GC cells in unimmunized Roquin^{san/san} Bcl6^{+/−} and Roquin^{san/san} Bcl6^{+/+} mice. As seen in wild-type mice, loss of one allele of Bcl6 caused a twofold reduction (4.4 ± 1.9% vs. 2.1 ± 0.5%; P = 0.0026) in the percentage of spontaneous GCs in Roquin^{san/san} mice (Fig. 1 b). Loss of one allele of Bcl6 also reduced GC B cells after SRBC immunization in a cell-autonomous manner (Fig. S1, available at http://www.jem.org/cgi/content/full/jem.20081886/DC1).

We then tested whether reduction in the spontaneous GC response in Roquin^{san/san} Bcl6^{+/−} mice was accompanied by reduced pathology. Serum dsDNA antibodies were present in 50% of Roquin^{san/san} Bcl6^{+/−} mice tested compared with 100% of Roquin^{san/san} Bcl6^{+/+} mice (Fig. 1 c). Kidney pathology was also significantly reduced in Roquin^{san/san} Bcl6^{+/−} mice (Fig. 1, d and e).

The lupus-like pathology of Roquin^{san/san} mice requires T cell activation

We have previously demonstrated that Roquin acts predominantly B cell extrinsically to induce spontaneous GC formation,
Expansion of the T_{FH} subset in Roquin^{san/san} mice is T cell intrinsic

We have previously reported the similarity of the gene expression profiles of Roquin^{san/san} CD4^+ T cells and T_{FH} cells. Furthermore, Roquin^{san/san} T cells accumulate in the splenic GCs (23). To formally assess whether T_{FH} cells, defined as CD4^+CXCR5^PD-1^{high} (30), are expanded in Roquin^{san/san} mice, we analyzed the percentage and total number of these cells in unimmunized mice and found more than a threefold increase of this subset in Roquin^{san/san} mice compared with littermate controls (Fig. 2 d). To determine whether this aberrant T_{FH} cell accumulation is cell intrinsic, we generated mixed chimeras. Sublethally irradiated Roquin^{s+/+}/Ly5^a mice were reconstituted with a 1:1 mix of Roquin^{s+/+}/Ly5^a and Roquin^{san/san}/Ly5^b bone marrow cells. As a control, Roquin^{s+/+}/Ly5^a cells were reconstituted with a 1:1 mix of Roquin^{s+/+}/Ly5^a and Roquin^{s+/+}/Ly5^b bone marrow cells. 8 wk after reconstitution, mice were immunized with SRBCs, and the percentage of T_{FH} cells derived from each type of donor marrow was determined by flow cytometry. In chimeras reconstituted with Roquin^{s+/+}/Ly5^a/Roquin^{san/san}/Ly5^b marrow, three times more Ly5^b (Roquin^{san/san}) T_{FH} cells than Ly5^a cells were observed (P = 0.001), whereas in controls, an equivalent proportion of T_{FH} cells arose from Ly5^a and Ly5^b cells (P = 0.56; Fig. 2 e). This indicates that Roquin acts in T cells to repress the formation and/or survival of T_{FH} cells. Consistent with the observed increase in T_{FH} cell numbers, IL-21 production was more than twofold higher in splenocyte cultures from Roquin^{san/san} mice (Fig. 2 f).

Although T_{FH} cells are expanded in Roquin^{san/san} mice, it was important to exclude defects in other T cell subsets implicated in autoimmunity. CD4^+CD25^FoxP3^ T reg cells have been shown to play a role in the regulation of lupus-associated autoantibodies (31). T reg cells are not reduced in number or function in Roquin^{san/san} mice (22). Quantification of FoxP3^CD25^CD4^+ cells in the spleen confirmed previous observations: Roquin^{san/san} mice have approximately twofold more T reg cells than wild-type mice (18.9 ± 3.69% vs. 9.76 ± 1.03%, respectively; Fig. S2 a, available at http://www.jem.org/cgi/content/full/jem.20081886/DC1). This argues against a role for reduced T reg cell numbers in driving the T cell–mediated disease of Roquin^{san/san} mice. Another T helper subset, Th17 cells, have emerged as potent mediators of autoimmunity (31), and recent work suggests that they may be critical to maintain the spontaneous GCs of BXD2 mice (23). To test whether Th17 cell activity is dysregulated in Roquin^{san/san} mice, we analyzed IL-17 levels in splenocyte cell cultures after activation with PMA and ionomycin. IL-17 was detected at comparable levels in cultures from Roquin^{+/+} and Roquin^{san/san} mice (Fig. S2 b). In contrast, IL-21, a cytokine secreted by both T_{FH} and Th17 cells (33, 34), was found at significantly higher levels in Roquin^{san/san} splenocyte cultures (Fig. 2 f).

Collectively these data indicate that the T_{FH} subset in Roquin^{san/san} mice is expanded in a cell-autonomous manner, whereas T reg and Th17 cells do not appear to be dysregulated in a way that has been previously described to result in autoimmunity.
Figure 1.  Heterozygosity for Bcl6 reduces the magnitude of the GC response in Roquin\textsuperscript{+/} and Roquin\textsuperscript{san/san} mice and ameliorates the lupus-like phenotype of Roquin\textsuperscript{san/san} mice.  (a) Flow cytometric contour plots (left) and graphical analysis (right) of B220\textsuperscript{+}GL-7\textsuperscript{+}CD95\textsuperscript{+} GC B cells in 10-wk-old wild-type (Bcl6\textsuperscript{+/}) and Bcl6\textsuperscript{+/+} mice 8 d after SRBC immunization (P = 0.0011).  Data are representative of four independent experiments (n = 4 per group).  (b) Flow cytometric contour plots (left) and dot plots (right) showing B220\textsuperscript{+}GL-7\textsuperscript{+}CD95\textsuperscript{+} GC B cells from 10-wk-old naive Roquin\textsuperscript{san/san} Bcl6\textsuperscript{+/} and Roquin\textsuperscript{san/san} Bcl6\textsuperscript{+/+} mice.  Data are representative of five independent experiments (n ≥ 4 per group).  (c) Representative determination of serum IgG anti-dsDNA from 6-mo-old female Roquin\textsuperscript{+/} Bcl6\textsuperscript{+/+}, Roquin\textsuperscript{san/san} Bcl6\textsuperscript{+/+}, and Roquin\textsuperscript{san/san} Bcl6\textsuperscript{+/+} mice, determined by immunofluorescence staining of C. luciliae substrate.  Data shown reflect the occurrence (n ≥ 6 mice per group); three out of six Roquin\textsuperscript{san/san} Bcl6\textsuperscript{+/+} mice had low intensity staining (illustrated in the fourth panel from left),
inappropriate GC reaction phenotype of Roquin\textsuperscript{san/san} to levels comparable to Roquin\textsuperscript{+/+} mice.

To test whether loss of SAP alters other T effector subsets in sanroque mice, we quantified Th1 and Th2 cells by flow cytometric staining for Tbet and Gata3, respectively. There were no statistically significant differences in the percentage of CD44\textsuperscript{high}GATA3\textsuperscript{+}CD4\textsuperscript{+} cells (0.91 ± 0.14% vs. 1.15 ± 0.41%; P = 0.18) or CD44\textsuperscript{high}Tbet\textsuperscript{+}CD4\textsuperscript{+} cells (6.76 ± 10.66% vs. 8.66 ± 5.74%; P = 0.59) between Roquin\textsuperscript{san/san} Sap\textsuperscript{+/+} and Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice in peripheral blood (Fig. 4, a and b). Lymph node CD44\textsuperscript{high}GATA3\textsuperscript{−/−}CD4\textsuperscript{+} cells (2.11 ± 1.68% vs. 1.59 ± 0.81%; P = 0.54) and CD44\textsuperscript{high}Tbet\textsuperscript{−/−}CD4\textsuperscript{+} cells (2.89 ± 1.9% vs. 2.02 ± 0.64%; P = 0.25) between Roquin\textsuperscript{san/san} Sap\textsuperscript{+/+} and Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice were slightly reduced, but these differences were not statistically significant (Fig. 4 c). In contrast, SAP deficiency caused an approximately fourfold reduction in Roquin\textsuperscript{san/san} lymph node T\textsubscript{FH} cells (3.93 ± 0.47% vs. 1.11 ± 0.38%; P = 0.001; Fig. 4 c). Collectively, these data indicate that T\textsubscript{FH} cells are the T helper subset whose generation is most severely impaired by SAP deficiency in Roquin\textsuperscript{san/san} mice.

Transfer of Roquin\textsuperscript{san/san} T\textsubscript{FH} cells induces spontaneous GC reactions in wild-type mice

To test whether Roquin\textsuperscript{san/san} T\textsubscript{FH} cells are sufficient to induce GC reactions in immunized wild-type mice, CD45.2\textsuperscript{+}PD-1\textsuperscript{high}CXCR5\textsuperscript{+}CD44\textsuperscript{+} CD4\textsuperscript{+} cells or CD45.2\textsuperscript{+}PD-1\textsuperscript{−}CXCR5\textsuperscript{−}CD44\textsuperscript{−} non-T\textsubscript{FH} T Effector cells were adoptively transferred into CD45.1\textsuperscript{+}C57BL/6 mice. 3 wk after transfer, 8% of donor T\textsubscript{FH} cells retained their CXCR5\textsuperscript{high} effector phenotype. In mice receiving non-T\textsubscript{FH} effector cells, although this was not statistically significant (Fig. 4, e and f). These data suggest that abrogation of signaling through SAP greatly reduced the autoimmune manifestations of Roquin\textsuperscript{san/san} mice. In contrast, SAP deficiency conferred no significant reduction of splenomegaly, lymphadenopathy, and hypergammaglobulinemia present in Roquin\textsuperscript{san/san} mice, indicating that these defects are not mediated by excessive T\textsubscript{FH} cell formation and GC function (Fig. S3, a–c, available at http://www.jem.org/cgi/content/full/jem.20081886/DC1).

Reduction of the GC response in Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice is mainly caused by B cell–extrinsic factors

Several studies have shown that SAP acts T cell intrinsically to regulate GC responses (35, 37, 39), but some controversy still exists in the light of one study showing B cell–intrinsic SAP-mediated effects (40). To test whether the correction of the GC response observed in Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice was caused by B cell–extrinsic factors, we transferred SW HEL B cells expressing intact SAP into Roquin\textsuperscript{+/+}, Roquin\textsuperscript{san/san}, and Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice, in conjunction with hen egg lysozyme (HEL)\textsuperscript{23}–conjugated SRBCs.

Adoptively transferred SW HEL B cells formed HEL–specific GCs in all four cohorts. Significantly fewer GCs were observed in Sap\textsuperscript{−/−} than in wild-type mice (23,958 ± 15,920 vs. 1,186 ± 1,019; P = 0.005; Fig. 5 d), confirming a B cell–extrinsic contribution of SAP to the GC response (34, 36, 38). In addition, SAP deficiency corrected the extreme GC formation observed in Roquin\textsuperscript{san/san} mice (124,528 ± 25,980 vs. 30,528 ± 18,100; P = 0.0004) to a response of similar magnitude to Roquin\textsuperscript{+/+} mice (Fig. 5 a). This suggests that the SAP–mediated effect in Roquin\textsuperscript{san/san} mice is also caused by B cell–extrinsic factors, and is most likely caused by SAP’s regulation of T\textsubscript{FH} cells.

To determine whether the loss of Sap in Roquin\textsuperscript{san/san} mice selectively affected the GC response or also impaired the extrafollicular response, we identified donor–derived extrafollicular plasma cells containing intracellular anti–HEL Ig at day 5 after immunization (Fig. 5 e). At this time point, GC–derived bodies and renal pathology of Roquin\textsuperscript{san/san} mice. Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice had not developed ANAs by 8 wk of age (Fig. 5 a). Assessment of high-affinity anti–dsDNA IgG antibodies at 6 mo of age (by immunofluorescence using Crithidia luciliae substrate) revealed positive dsDNA antibodies in all Roquin\textsuperscript{san/san} mice evaluated, whereas only one out of six Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice had detectable antibodies. Renal histology revealed that Roquin\textsuperscript{san/san} Sap\textsuperscript{−/−} mice had only minor interstitial nephritis and thickening of the mesangial matrix, with no detection of immune complex deposition observed by electron microscopy (Fig. 5, b and c). These data suggest that abrogation of signaling through SAP greatly reduced the autoimmune manifestations of Roquin\textsuperscript{san/san} mice.
Figure 2. The autoimmune phenotype of Roquin<sup>san/san</sup> mice requires T cell activation through CD28, and T<sub>H</sub> cells are expanded cell autonomously. (a, left) Detection of IgG-ANA using Hep-2 slides in sera from 8-wk-old female Roquin<sup>san/san</sup> and Roquin<sup>san/san Cd28<sup>-/-</sup></sup> mice (n = 5 per group). (right) Detection of anti-dsDNA IgG serum antibodies in 6-mo-old female Roquin<sup>san/san</sup> and Roquin<sup>san/san Cd28<sup>-/-</sup></sup> mice determined by staining C. luciliae slides. Data are representative of three independent experiments (n ≥ 5 per group). (b) Score of nephritis severity in 6-mo-old female Roquin<sup>+/+</sup>, Roquin<sup>san/san</sup> and Roquin<sup>san/san Cd28<sup>-/-</sup></sup> mice as determined by histological analysis defined by the criteria given in Table S1 (available at http://www.jem.org/cgi/content/full/jem.20081886/DC1). Each symbol represents one mouse. Horizontal bars indicate medians. (c) Representative images of kidney sections stained with H&E (left) or viewed under an electron microscope (right). Histology from Roquin<sup>san/san Cd28<sup>-/-</sup></sup> animals was much less severe, with normal H&E appearances and few electron-dense deposits (arrows) in the mesangium. Bars: (left) 100 μm; (right) 10 μm. (d) Representative flow cytometric contour plots of CD4<sup>+</sup>CXCR5<sup>+</sup>PD-1<sup>high</sup> T<sub>H</sub> cells in 10-wk-old Roquin<sup>san/san</sup> mice and control littermates. Data are representative of five independent experiments (n = 4 per group), and the numbers in the plots represent percentages. (e) Dot plots representing percentages of CD4<sup>+</sup>CXCR5<sup>-PD-1<sup>high</sup></sup> T<sub>H</sub> cells from SRBC-immunized chimeric mice generated by reconstituting sublethally irradiated mice with a 1:1 mix of either Roquin<sup>+/+</sup>Ly5<sup>a</sup>/Roquin<sup>+/+</sup>Ly5<sup>b</sup> (left) or Roquin<sup>+/+</sup>Ly5<sup>a</sup>/Roquin<sup>san/san</sup>Ly5<sup>b</sup> (right). Data are representative of three independent experiments (n = 4 per group). Each symbol represents the Ly5<sup>a</sup> or
plasma cells cannot be detected (41). Although we observed a 4-fold decrease in SWHEL GC B cells, there was only a 1.3-fold decrease in the number of SWHEL extracellular plasma cells (53,198 ± 1,528 vs. 40,528 ± 7,548; P = 0.04) in Roquin /−/− recipient mice compared with Roquin /+/+ Sap /+/+ controls (Fig. 5 e).

These data suggest that the reduction of the GCs in Roquin /−/− mice caused by Sap deficiency is mainly a result of B cell–extrinsic factors, and that this results in selective inhibition of the GC pathway.

**Roquin /−/−** Sap /−/− mice have reduced expression of ICOS and IL-21

We have demonstrated that **Roquin** /−/− Sap /−/− mice have fewer Tfh cells, a correction of the spontaneous GC response, and a reduction in autoimmune pathology. To gain insight into the possible functional Tfh defect brought about by SAP deficiency in **Roquin** /−/− mice, we quantified the changes in the expression of ICOS and IL-21, both of which are increased in **Roquin** /−/− CD4 + cells (22) and are shown to be essential for Tfh cell formation, homeostasis, and function (33, 34, 42, 43). Loss of SAP reduced the levels of ICOS on both naive (CD44 low ) and activated/memory (CD44 high ) CD4 + T cells in **Roquin** /−/− mice and to an even greater extent on **Roquin** /−/+ mice (Fig. 6, a–c). Despite this reduction, **Roquin** /−/− Sap /−/− mice still expressed more than threefold higher levels of ICOS than **Roquin** /−/+ Sap /−/− mice.

To determine whether loss of SAP had an effect on the production of IL-21, which was increased in **Roquin** /−/− mice, we compared IL-21 production from **Roquin** /−/+ Sap /−/−, **Roquin** /−/+ Sap /−/−, **Roquin** /−/+ Sap /−/−, and **Roquin** /−/− Sap /−/− splenocytes. SAP deficiency reduced IL-21 production by both **Roquin** /−/− and **Roquin** /−/+ cultures were comparable to those of wild-type (**Roquin** /−/+ Sap /−/−) mice. As previously reported (37), SAP deficiency resulted in higher CD40L expression upon activation in both **Roquin** /−/+ and **Roquin** /−/− mice (Fig. 6, e and f).

**IL-21 deficiency does not prevent autoimmunity in Roquin /−/− mice**

Given that **Roquin** /−/− mice produce high levels of IL-21 and given the recently described role for IL-21 in Tfh cell generation after immunization (33, 34), we investigated whether IL-21 also controls autoantibody production, Tfh cell accumulation, and spontaneous GC formation in these mice. To this end, we crossed **Roquin** /−/− mice to IL-21 /−/− mice on a C57BL/6 background and assessed ANA formation. There was no difference in the pattern or titer of ANAs produced by **Roquin** /−/− mice (Fig. 7, a and b). Overall, IL-21 deficiency did not affect the hypergammaglobulinemia of **Roquin** /−/− mice (Fig. 7 c), and consistent with previous reports (44), IgG1 titers were reduced, whereas IgE titers were increased (Fig. 7 c). Likewise, loss of IL-21 did not correct the lymphadenopathy or the splenomegaly of **Roquin** /−/− mice (Fig. 7 d).

We then investigated whether IL-21 influences Tfh cell accumulation and spontaneous GC formation. There was no difference in either the number of Tfh cells or GC cells between **Roquin** /−/− IL-21 /−/− and **Roquin** /−/− IL-21 /−/− littermates, (Fig. 7, e and f). These data suggest that IL-21 does not play a role in spontaneous GC formation, Tfh development, or accumulation in **Roquin** /−/− mice.

**DISCUSSION**

Elucidation of the defect that results in high-affinity antibodies to dsDNA is central to understanding lupus and, hence, the development of specific therapeutic interventions. In the **Roquin** /−/− model, a fundamental GC defect results in the production of autoantibodies. This pathway appears to be critical for disease development, because an impediment to GC formation conferred by halving the dose of Bcl6 is sufficient to attenuate the lupus phenotype. Our data are consistent with other evidence that aberrant T cell help can cause spontaneous GC formation and autoimmunity (9, 45). We show the expanded Tfh cell subset, a consistent component of the **Roquin** /−/− phenotype, is responsible for excessive GC formation. In **Roquin** /−/− mice, GCs form in the absence of foreign antigen and yield ANAs. SAP deficiency reduces the size and activity of the Tfh subset and abrogates autoantibody production and renal disease. These findings provide the in vivo cellular mechanism to link the defect in microRNA (miRNA)-mediated repression of Tfh molecules conferred by the san allele of Roquin with the lupus phenotype of **Roquin** /−/− mice (22, 28). Moreover, they elucidate a mechanism of lupus that could be generalized to other defects of Tfh homeostasis. Importantly, they provide evidence that defects in positive selection of GC B cells can cause autoimmunity.

In this paper, we demonstrate that **Roquin** /−/− acts T cell intrinsically to drive an accumulation of Tfh cells that support a spontaneous GC response. Overexpression of ICOS is a plausible mechanism, because ICOS is important for the generation and survival of Tfh cells (34, 42, 43), and Roquin normally acts to repress Icos miRNA (28). Previously, we have shown that this accounts for the lymphoproliferation, splenomegaly, and lymphadenopathy of **Roquin** /−/− mice (28). The data presented in this paper also show that CD28 signaling is a key contributor to the development of systemic autoimmunity in **Roquin** /−/− mice. CD28 is not only expressed on CD4 + cells, but it has also been found to be expressed by stromal cells (46) and plasma cells (47), and this expression can influence B cell development and antibody responses. Nevertheless, our previous data showing that the increased plasma cell and GC

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**Ly5** population derived from one mouse. (f) ELISA was used to determine culture supernatant IL-21 levels from 24-h splenocyte cultures from **Roquin** /−/− mice and littermate controls in the presence of PMA and ionomycin. Error bars indicate means ± SEM. Data are representative of three independent tests where each sample was run in triplicate.
numbers, and B cell activation in sanroque are predominantly B cell extrinsic (22), together with the potent effects of SAP deficiency in the sanroque autoimmune phenotype, suggest that CD28 deficiency in T cells rather than in B cells or stromal cells is responsible for the observed effect. Thus, we infer that the sanroque lupus phenotype is T cell mediated. CD28 deficiency considerably decreased the abnormally high levels of ICOS normally found on Roquin<sup>san/san</sup> T cells. This is likely to contribute

Figure 3. Spontaneous GC and T<sub>FH</sub> formation are corrected by loss of SAP in Roquin<sup>san/san</sup> mice. (a) CD4<sup>+</sup>CXCR5<sup>+</sup>PD-1<sup>hi</sup> T<sub>FH</sub> cells in unimmunized 10-wk-old Roquin<sup>san/san</sup> and Roquin<sup>san/san</sup>Sap<sup>+/+</sup> mice (P = 0.0056). Representative flow cytometric contour plots are shown (right). Data are representative of four independent experiments (n = 4 per group). (b) B220<sup>+</sup>GL-7<sup>+</sup>CD95<sup>+</sup> GC B cells in unimmunized 10-wk-old Roquin<sup>san/san</sup> Sap<sup>+/+</sup> and Roquin<sup>san/san</sup>Sap<sup>−/−</sup> mice (P = 0.0007). Representative flow cytometric contour plots are shown (right). Data are representative of four independent experiments (n = 4 per group). (c) Photomicrographs of frozen spleen sections from unimmunized 6-mo-old mice of the indicated genotypes stained with IgD (brown; all panels), PNA (blue; left), TCRβ (blue; middle), and PD-1 (blue; right). Bars, 200 μm.
to the abrogation of the lupus phenotype in a manner comparable to halving the gene dose of Icos (28). A nonmutually exclusive explanation is that although ICOS overexpression causes a lymphoproliferative syndrome, danger signals may also be required for the production of autoantibodies that cause end-organ damage. This is because ligands for CD28 are dependent

Figure 4. Th1 and Th2 cells are present in Roquin san/san mice in the absence of SAP and T FH cells, but not non-T FH effector cells, induce a GC response in wild-type mice. (a) Representative flow cytometric contour plots and (b) graphical analysis of GATA3-CD44<sup>hi</sup>CD4<sup>+</sup> and Tbet-CD44<sup>hi</sup>CD4<sup>+</sup> cells in mice of the indicated genotypes. Data are representative of two independent experiments (n ≥ 4 per group). (c) Representative dot plots of lymph node CD4<sup>+</sup>PD-1<sup>hi</sup>CXCR5<sup>+</sup> (left), Tbet<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> (middle), and GATA3<sup>+</sup>CD44<sup>hi</sup>CD4<sup>+</sup> (right) cells. (d) Experimental outline for adoptive transfer of Roquin<sup>san/san</sup>CD4<sup>+</sup>CD45.2<sup>+</sup>PD-1<sup>hi</sup>CXCR5<sup>+</sup> or CD4<sup>+</sup>CD45.2<sup>+</sup>PD-1<sup>−</sup>CXCR5<sup>−</sup> T cells into CD45.1 C57BL/6 mice. (e) Flow cytometric contour plots and (f) dot plots of B220<sup>+</sup>GL-7<sup>+</sup>CD95<sup>+</sup> GC B cells from CD45.1 C57BL/6 recipients 3 wk after adoptive transfer of the indicated cell type. Data were generated from three mice per group (**, P > 0.001). In a, d, and e, the numbers in the plots represent percentages.
on Toll-like receptor ligation, which typically occurs during infections and tissue damage, and potentially after apoptosis. Because mice in this study were housed under specific pathogen-free conditions, one explanation for the source of the danger signal in the san/san model might be the abundance of apoptotic cells in GCs, which overloads the normal nonimmunogenic modes of disposal (11). Apoptotic cells display self-antigens that are the target of the autoimmune response in lupus, and are ligands for TLR7 and/or TLR9 on antigen-presenting cells and/or B cells (48).

Our data indicate that the GC is central to the autoimmune phenotype of Roquin<sup>−/−</sup>/H11002 mice, because reduction of spontaneous GCs, conferred by halving the gene dose of the transcriptional regulator Bcl6, results in a proportionate amelioration of autoimmune pathology. This heterozygote phenotype is a novel finding. Although previous reports describe intact antibody responses in Bcl6<sup>+/-</sup> mice on day 11 after immunization (26), these are likely to derive predominantly from the extrafollicular response at this early time point. The heterozygote phenotype is consistent with the observation that GC differentiation is accompanied by only a twofold increase in Bcl6 miRNA expression (49). Although our work does not exclude an effect of halving the gene dose of Bcl6 on T<sub>FH</sub> cell formation, it does strongly suggest that the GC reduction seen in Bcl6<sup>−/-</sup> mice is a consequence of reduced B cell–expressed BCL6: in 50% Bcl6<sup>+/-</sup>/50% Bcl6<sup>-/-</sup> mixed bone marrow chimeras, GC B cells were only decreased in GC B cells deriving from Bcl6<sup>−/-</sup> bone marrow. Furthermore, transfer of Bcl6<sup>−/-</sup>/50% SW<sub>HEL</sub> B cells into Bcl6<sup>−/-</sup>/50% recipient mice, providing the major source of T<sub>FH</sub> cells, resulted in normal donor-derived GC numbers after HEL-SRBC immunization.

SAP deficiency abrogates autoimmune in Roquin<sup>−/−</sup>/H11002 mice. SAP deficiency affects several T cell subsets and natural killer cells (50, 51) but profoundly impairs the ability of T cells to provide help to B cells for GC formation (37–39). Our results demonstrate that SAP deficiency specifically reduces the development of CD4<sup>+</sup> T cells with a T<sub>FH</sub> phenotype, leaving Th1 and Th2 cell formation largely intact. This, together with the evidence presented for the GC dependence of autoimmunity, as well as the observation that transfer of Roquin<sup>−/−</sup>/H11002 T<sub>FH</sub> cells induces spontaneous GC, places aberrant expansion of T<sub>FH</sub> cells at the root of spontaneous GC formation and autoimmunity in Roquin<sup>−/−</sup>/H11002 mice. This mechanism is consistent with another lupus model, B6.Sle1<sup>−/−</sup>, in which T cells have a transcriptional profile typical of T<sub>FH</sub> cells (21).

SAP deficiency has been shown to ameliorate autoimmunity in another lupus-prone strain, MRL<sup>pr</sup> mice (52). These mice have been shown to also develop spontaneous GCs (9), although SHM of autoreactive B blasts has been shown to occur in the T zone areas rather than within GCs (16). SAP deficiency has been shown to eliminate the pathogenic CD4<sup>+</sup>/CD8<sup>+</sup> T cells that account for the lymphadenopathy of MRL<sup>pr</sup> mice (52), and there is evidence that autoimmunity in MRL<sup>pr</sup> mice is T cell dependent (53). It will be interesting to examine whether dysregulated ectopically located T<sub>FH</sub> cells or an extrafollicular counterpart is contributing to autoimmune immunity in these mice. Finally, although sanroque SAP-deficient mice do not form spontaneous GCs, they still show hypergammaglobulinemia and very mild cell infiltrates in the kidney, suggesting that a non-T<sub>FH</sub>-mediated pathway contributes to these manifestations in Roquin<sup>−/−</sup>/H11002 mice.

Our data add to previous studies demonstrating that SAP deficiency acts B cell extrinsically, causing a profound and selective impairment of GC reactions (37) with little effect on extrafollicular plasma cell generation. The requirement for SAP for effective T<sub>FH</sub> cell function has previously been ascribed to dysregulated kinetics of ICOS up-regulation and increased expression of CD40L (37), which we confirmed in the present study. Our work also shows that SAP signaling is required to maintain elevated baseline levels of ICOS in naive sanroque T cells.

An intriguing finding of this study was the lack of any role for IL-21 in Roquin<sup>−/−</sup>-induced lupus, T<sub>FH</sub> cell accumulation, and spontaneous GC formation. Although recent reports have shown that IL-21 is involved in T<sub>FH</sub> cell generation and optimal GC responses (33, 34), other groups have previously reported normal GC formation with an accumulation in IgG memory B cells in IL-21R<sup>−/-</sup> mice (54). A recent paper has also shown that IL-21 can be produced by extrafollicular T cells, which contribute to autoimmunity in MRL<sup>pr</sup> mice (55). Furthermore, IL-21 has been shown to potentely induce plasma cell generation from naive B cells (56), suggesting an important role for IL-21 in extrafollicular B cell responses. Even if IL-21 turns out to be critical for GC B cell selection, it is possible that other molecules overexpressed by sanroque T<sub>FH</sub> cells, such as ICOS, substitute for the effects normally mediated by IL-21.

The pathway to systemic autoimmunity identified in this paper highlights the importance of negative selection of self-reactive lymphocytes in GCs. Autoreactive B cells are a normal component of the naive peripheral B cell repertoire. In lupus patients, these cells enter GCs, whereas in healthy individuals, they are excluded (57). Other evidence indicates that in lupus, GCs can generate B cells that have antinuclear specificities from nonautoimmune precursors (7). Furthermore, GCs are increased in lupus (58). Regardless of their ontogeny, autoreactive centrocytes would normally fail to receive selection signals by T<sub>FH</sub> cells and would undergo apoptosis (59). Our data suggest that this process is perturbed by the san allele of Roquin.

According to the prevailing model for T<sub>FH</sub> selection of centrocytes, large numbers of GC B cells compete with each other for the limiting available T cell help (60) and for antigen on follicular dendritic cells. Centrocytes that have acquired higher affinity for antigen would scavenge more antigen from follicular dendritic cells, which would confer on them an avidity advantage in interactions with T<sub>FH</sub> cells compared with lower affinity B cells. Based on this model, we speculate that in Roquin<sup>−/−</sup>/H11002 mice, competition by B cells for T cell help is reduced by the expansion of T<sub>FH</sub> cells. Interactions between centrocytes and T<sub>FH</sub> cells are determined by affinity for MHC–peptide for the TCR, modified by the action of accessory molecules. Loss of SAP in Roquin<sup>−/−</sup>/H11002 mice causes a reduction in ICOS overexpression, potentially increasing the...
antigen affinity required to reach the T<sub>FH</sub> activation threshold in Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice.

Although the lupus phenotype of Roquin<sup>san/san</sup> mice is Mendelian, apart from rare exceptions (61), lupus is a polygenic disorder. Nevertheless, it is plausible that one or more polymorphisms or mutations that affect T<sub>FH</sub> homeostasis, such as those that regulate IL-21, ICOS, or ICOSL expression, could contribute to lupus pathogenesis. Because T<sub>FH</sub> cells are present in peripheral blood, this is a testable hypothesis. Indeed, there is evidence that SLE patients have increased numbers of circulating CD4<sup>+</sup>ICOS<sup>+</sup> cells in their peripheral blood compared with nonautoimmune individuals (62). Because T<sub>FH</sub> cells are

**Figure 5.** Serum autoantibodies and renal pathology in Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice and reduction of the GC response in SAP-deficient Roquin<sup>san/san</sup> mice is caused by B cell–extrinsic factors. (a, left) Representative staining of Hep-2 slides for detection of IgG ANA in the serum of 8-wk-old female Roquin<sup>san/san</sup> and Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice (n = 5 per group). (right) Detection of IgG anti-dsDNA serum antibodies in 6-mo-old female Roquin<sup>san/san</sup> and Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice determined by immunofluorescence staining using C. luciliae substrate. Data are representative of three independent experiments (n ≥ 5 per group). (b) Representative images of kidney sections stained with H&E (top) or viewed under an electron microscope (bottom) from 6-mo-old female Roquin<sup>san/san</sup> and Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice. Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice show slight mesangial expansion on H&E staining but no electron-dense deposits. Bars: (top) 100 μm; (bottom) 5 μm. (c) Score of nephritis severity in 6-mo-old female Roquin<sup>+/+</sup>, Roquin<sup>san/san</sup>, and Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice as determined by histological analysis according to the criteria given in Table S1 (available at http://www.jem.org/cgi/content/full/jem.20081886/DC1). Each symbol represents one mouse. Horizontal bars indicate medians. (d) Gating strategy for assessing the HEL-specific GC response (top) and graphic representation (bottom) of the total number of HEL-specific GC cells per spleen in mice with the indicated genotypes 7 d after cotransfer of SW<sub>HEL</sub> B cells and HEL<sup>2x</sup>-conjugated SRBCs. Data are representative of three independent experiments (n ≥ 4). Each symbol represents one mouse. (e) Gating strategy for determining HEL-specific extrafollicular plasma cells (top) and graphical representation (bottom) of the number of HEL-specific plasma cells 5 d after cotransfer into mice with the indicated genotypes. Data are representative of two independent experiments (n ≥ 3 per group). p-values are indicated on graphs.
the cells expressing the highest levels of ICOS (63, 64), it is possible that circulating CD4+ICOS+ cells reflect an excessive T<sub>FH</sub> response, and is consistent with recent data showing that a subset of SLE patients have an increased proportion of CD4+CD45RO+CXCR5+ICOSHIGHPD1HIGH cells in their peripheral blood (a phenotype that correlates with higher titers of anti-dsDNA antibodies and more severe kidney damage; unpublished data). If confirmed in independent studies, the GC–T<sub>FH</sub> pathway elucidated in the Roquin<sup>san/san</sup> model pathway would emerge as a novel and specific target for therapy.

Figure 6. T<sub>FH</sub> cell–associated molecules are decreased in Roquin<sup>san/san</sup> Sap<sup>−/−</sup> mice. (a) Representative flow cytometric histograms and (b and c) graphical analysis showing ICOS mean fluorescence intensity (MFI) of splenic naive CD44<sup>LOW</sup> (b) and activated/memory CD44<sup>HIGH</sup> (c) CD4<sup>+</sup> T cells from 10-wk-old unimmunized mice of the indicated genotypes. Data are representative of three independent experiments. Each symbol represents one mouse. (d) ELISA quantification of IL-21 in supernatant from an overnight culture of splenocytes in the presence of PMA and ionomycin from mice of the indicated genotypes. Data are representative of three experiments. (e) Flow cytometric contour plots of CD40L expression on splenocytes, 5 h after stimulation with anti-CD3 and anti-CD28, derived from mice of the indicated genotypes. (left) Staining with an isotype control; (right) CD40L staining. (f) Histograms of the percentage of CD4<sup>+</sup> cells that express CD40L (gated as shown in e) 5 h after CD3 and CD28 stimulation on splenocytes from mice with the indicated genotypes. In d and f, error bars indicate means ± SEM.
MATERIALS AND METHODS

Mice and immunizations. Roquin<sup>san/san</sup> and C57BL/6 (B6) mice, all crossed to Sap<sup>−/−</sup>, B6.129<sup>Cd28<sup>−/−</sup></sup>, and IL-21<sup>−/−</sup> mice, and SW<sub>H11002</sub> mice, were housed in specific pathogen-free conditions at the Australian National University Bioscience Facility. IL-21 KO mice were generated at Lexicon Pharmaceuticals, Inc. and were provided by M. Smyth (Peter MacCallum Cancer Institute).

Figure 7. Lack of IL-21 does not affect the phenotype, T<sub>FH</sub> cell accumulation, or GC formation of Roquin<sup>san/san</sup> mice. (a) IgG ANAs in the serum of mice of the genotypes indicated, detected by immunofluorescence using Hep-2 substrate. Data are representative of three independent experiments (n ≥ 5 mice per group). (b) Score of ANA staining intensity by confocal microscopy from sera taken from mice of the indicated genotypes. Data are representative of three independent experiments (n ≥ 5 mice per group). (c) Basal serum total IgG, IgG1, and IgE measured by ELISA. Data are representative of two independent experiments (n = 5 mice per group). (d) Lymph node and spleen weight in grams for mice of the indicated genotypes. Data are representative of two independent experiments (n ≥ 5 per group). (e) Flow cytometric contour plots and dot plots of PD-1 high CXCR5<sup>+</sup> CD4<sup>+</sup> T<sub>FH</sub> cells and (f) GL-7<sup>+</sup>CD95<sup>+</sup>B220<sup>+</sup> GC cells from mice of the indicated genotypes. Data are representative of two independent experiments (n ≥ 5 mice per group). In e and f, the numbers in the plots represent percentages.
Centre, Melbourne, Australia) and ZymoGenetics, Inc. These mice were backcrossed 10 generations onto the C57BL/6 background. SW HEL mice carry a Vk10-κ light chain transgene and a knocked in VH10 Ig heavy chain in place of the JH segments of the endogenous IgH gene that encode a high-affinity antibody for HEL (65). All animal procedures were approved by the Australian National University Animal Ethics and Experimentation Committee.

To generate thymus-dependent responses where indicated in the figures, 8–12-wk-old mice were immunized i.p. with 2 × 10^9 SRBCs (Institute of Medical and Veterinary Science Veterinary Services). For experiments involving SW HEL mice, 10^9 SW HEL B cells were transferred into recipient mice, which were simultaneously immunized i.v. with 2 × 10^9 SRBCs conjugated with mutant HEL<sup>+</sup> using a protein conjugation kit (Invitrogen) (41).

**Bone marrow chimeras.** Recipient C57BL/6-Ly5a mice were sublethally irradiated with 1,000 Rad and were reconstituted via i.v. injection with 2 × 10<sup>6</sup> donor bone marrow–derived hematopoietic stem cells.

**Antibodies.** Antibodies and streptavidin conjugates for flow cytometry were from BD unless otherwise indicated: anti-mouse B220-PerCP, CD4-PerCP, ICOS-PE (eBioscience), FoxP3/eBioscience, GL7-FITC, GATA3-allophycocyanin, Tbet-PerCP Cy5.5, CD95-PE, CXCR5-biotin, PD-1-PE (eBioscience), CTLA-4-PE, CD25-PE, streptavidin–PerCP Cy5.5, and CD40L-biotin. For immunohistochemistry, the primary antibodies and reagents used were rat anti–mouse IgG (SouthernBiotech), biotinylated anti–mouse TCR<sub>β</sub> (BD), rat anti–mouse PD-1 (BioLegend), and PNA-biotin (Vector Laboratories); the secondary antibody used was rabbit anti–rat horseradish peroxidase (HRP; Dako).

**Cell isolation, culture, and stimulation.** Single-cell suspensions were prepared from spleens of unimmunized and/or immunized mice. Single-cell suspensions were prepared in RPMI 1640 medium (JRH Biosciences) supplemented with 2 mM l-Glutamine (Invitrogen), 100 U penicillin-streptomycin (Invitrogen), 0.1 mM nonessential amino acids (Invitrogen), 100 mM Hepes (Sigma-Aldrich), 5 × 10<sup>−5</sup> 2-mercaptoethanol, and 10% fetal calf serum by sieving and gentle pipetting through 70-μm nylon mesh filters (Falcon; BD). Cells were then cultured for 24 h at 37°C/5% CO<sub>2</sub> with 50 ng/ml PMA (Sigma-Aldrich) and 1 μM ionomycin (Sigma-Aldrich) in serum-free RPMI 1640 medium supplemented with 100 mM Hepes, 5 × 10<sup>−5</sup> 2-mercaptoethanol, 0.1 mM nonessential amino acids, 100 μM streptomycin, 100 μM penicillin, and 10% fetal calf serum by sieving and gentle pipetting through 70-μm nylon mesh filters (Falcon; BD). Cells were then cultured for 24 h at 37°C/5% CO<sub>2</sub> with 50 ng/ml PMA (Sigma-Aldrich) and 1 μM nonessential amino acids (Invitrogen). A FACSCalibur (BD) with a TCR<sub>β</sub>-allophycocyanin, CD69 – PerCP, ICOS-PE (eBioscience), FoxP3 – PE, GL7-HRP, and conjugate layer for 30 min and washed thoroughly with FACS buffer, pH 7.4, at room temperature overnight. They were then washed in 0.1 M cacodylate buffer, pH 7.4, at room temperature overnight. They were then washed in 0.1 M cacodylate buffer, treated with 0.25% glutaraldehyde for 2 h, and washed in double-distilled H<sub>2</sub>O. Samples were stained with 0.2% ammonium osmium tetroxide in 0.1 M cacodylate buffer, pH 7.4, at room temperature for 2 h, and washed in double-distilled H<sub>2</sub>O. Samples were stained with 0.2% ammonium osmium tetroxide in 0.1 M cacodylate buffer, pH 7.4, at room temperature for 2 h, and washed in double-distilled H<sub>2</sub>O. They were then mounted in orientation silicone molds and heated in an oven at 70°C for 8–12 h. The samples were examined on a transmission electron microscope (model 1011; JEOL) at 60 KeV. The images were captured using a digital camera (MegaView III) and the AnalySIS software package (Soft Imaging System; Olympus), and were subsequently scored using the criteria detailed in Table S1 (available at http://www.jem.org/cgi/content/full/jem.20081886/DC1).

**ANA and dsDNA assessment.** Serum obtained by eye bleed from 8-wk-old mice was diluted in PBS 1:40 and used for indirect immunofluorescence on fixed Hep2 slides (Antibodies, Inc.) for ANA detection, and serum from 6-mo-old mice was diluted 1:20 and used for indirect immunofluorescence on fixed C. fasciatus slides (Antibodies, Inc.) for ANA anti–dsDNA antibody detection, respectively. Alexa Fluor 488 goat anti–mouse IgG (Invitrogen) was used to detect mouse antibodies. Autoantibodies were scored blind as negative or positive on a scale of 1–3 based on the intensity of fluorescence. Relative levels of ANAs were estimated by viewing the slides using a confocal microscope (TCS SP5; Leica) at 200× magnification and a fixed laser power, and measuring the fluorescence of five randomly selected 1,250-μm<sup>2</sup> regions of Hep2 cells, compared with five regions where cells were absent. The sample-specific mean background was subtracted from the sample-specific mean fluorescence to give an estimation of fluorescent intensity. These results were analyzed using a two-tailed Student's t test using Prism software.

**Statistical analysis.** Data were analyzed using a two-tailed Student’s t test using Prism software.
Online supplemental material. Fig. S1 illustrates that the loss of one allele of Bcl6 results in B cell–intrinsic defects in GC formation. Fig. S2 a shows that CD4⁺ Foxp3⁺ Treg cells are expanded twofold in Roquin⁻/⁻ mice relative to Roquin⁺/⁺ mice. Fig. S2 b demonstrates that Roquin⁻/⁻ splenocytes produce equivalent levels of IL-17 to Roquin⁺/⁺ splenocytes. Fig. S3 a and b shows that Sap deficiency does not correct the splenomegaly and lymphadenopathy in Roquin⁻/⁻ mice. Fig. S3 c demonstrates that hyper-IgG in Roquin⁻/⁻ mice is not corrected in the absence of Sap. Table S1 describes the scoring strategy used to assess the severity of mouse nephritis. Online supplemental material is available at http://www.jem.org/cgi/content/full/jem.20081886/DC1.

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