Gene Network Inference and Biochemical Assessment Delineates GPCR Pathways and CREB Targets in Small Intestinal Neuroendocrine Neoplasia

Ignat Drozdov1,2, Bernhard Svejda3, Bjorn I. Gustafsson4, Shrikant Mane5, Roswitha Pfragner6, Mark Kidd3*, Irvin M. Modlin3*

1 Cardiovascular Division, King’s College London BHF Centre of Research Excellence, James Black Centre, London, United Kingdom, 2 Centre for Bioinformatics, School of Physical Sciences and Engineering, King’s College London, London, United Kingdom, 3 Gastrointestinal Pathobiology Research Group, Yale University School of Medicine, New Haven, Connecticut, United States of America, 4 Department of Gastroenterology, St Olav’s Hospital, and Department of Cancer Research and Molecular Medicine, NTNU, Trondheim, Norway, 5 Keck Affymetrix Facility, Yale University School of Medicine, New Haven, Connecticut, United States of America, 6 Institute of Pathophysiology and Immunology, Centre for Molecular Medicine, Medical University of Graz, Austria

Abstract

Small intestinal (SI) neuroendocrine tumors (NETs) are increasing in incidence, however little is known about their biology. High throughput techniques such as inference of gene regulatory networks from microarray experiments can objectively define signaling machinery in this disease. Genome-wide co-expression analysis was used to infer gene relevance network in SI-NETs. The network was confirmed to be non-random, scale-free, and highly modular. Functional analysis of gene co-expression modules revealed processes including ‘Nervous system development’, ‘Immune response’, and ‘Cell-cycle’. Importantly, gene network topology and differential expression analysis identified over-expression of the GPCR signaling regulators, the cAMP synthetase, ADCY2, and the protein kinase A, PRKAR1A. Seven CREB response element (CRE) transcripts associated with proliferation and secretion: BEX1, BICD1, CHGB, CPE, GABRB3, SGC2 and SGC3 as well as ADCY2 and PRKAR1A were measured in an independent SI dataset (n = 10 NETs; n = 8 normal preparations). All were up-regulated (p < 0.035) with the exception of SGC3 which was not differently expressed. Forskolin (a direct cAMP activator, 10^{-5} M) significantly stimulated transcription of pCREB and 3/7 CREB targets, isoproterenol (a selective β-adrenergic receptor agonist and cAMP activator, 10^{-5} M) stimulated pCREB and 4/7 targets while BIM-53061 (a dopamine D2 and Serotonin [5-HT2] receptor agonist, 10^{-6} M) stimulated 100% of targets as well as pCREB; CRE transcription correlated with the levels of cAMP accumulation and PKA activity; BIM-53061 stimulated the highest levels of cAMP and PKA (2.8-fold and 2.5-fold vs. 1.8–2-fold for isoproterenol and forskolin). Gene network inference and graph topology analysis in SI NETs suggests that SI NETs express neural GPCRs that activate different CRE targets associated with proliferation and secretion. In vitro studies, in a model NET cell system, confirmed that transcriptional effects are signaled through the cAMP/PKA/pCREB signaling pathway and that a SI NET cell line was most sensitive to a D2 and 5-HT2 receptor agonist BIM-53061.

Introduction

Neuroendocrine or “carcinoid” tumors of the gut, usually misperceived as a rare, indolent neoplasia, have not rigorously been studied, are poorly understood and often misdiagnosed [1]. The perception that these tumors are rare has been altered by introduction of diagnostic strategies including endoscopy, the measurement of plasma biochemical markers such as Chromogranin A, and nuclear medicine techniques, including somatostatin receptor scintigraphy (SRS) [2]. A review of the current Surveillance Epidemiology and End Results (SEER) database indicates that small intestinal (SI) neuroendocrine tumors (NET) comprise 24.3% of all NETs with the overall 5-year survival rate of 64.1% [3,4]. In the event of liver metastases, bioactive tumor products enter the systemic circulation, bypassing hepatic inactivation, and engender a ‘carcinoid syndrome’. This consists of a variety of symptoms including episodic skin flushing, diarrhea, bronchoconstriction, sweating and abdominal cramping, and as many as 30–50% of individuals may have cardiac valvular disease [5].

Although the cell of origin of SI NET has been identified as the enterochromaffin (EC) cell, the secretory and proliferative regulation of these cells is poorly defined and, as a result, progress in the development of effective therapeutic strategies for diseases associated with the cell, e.g. NETs or Crohn’s disease [6], has been limited. The principal secretory product of the EC cell is serotonin (5-HT), although substance P (motility regulator) and guanylin (secretory regulator) have also been identified [7–9]. The most successful therapy, to date, has been somatostatin analogs which activate inhibitory G-protein coupled receptors (GPCRs) and...
result in decreased secretion of bioactive products with concomitant amelioration of symptoms [10–12].

GPCRs represent the largest family of cell-surface molecules involved in environment sensing and signal transmission, accounting for >2% of the total genes encoded by the human genome [13]. Mutations in GPCRs and Gα subunits have been identified in endocrine tumors and are often associated with symptoms caused by unregulated hormonal secretion. For example, activating mutations of the thyroid stimulating hormone receptor (TSHR) are found in some thyroid carcinomas and approximately 80% of thyroid adenomas, while germ-line mutations in TSHR cause familial non-autoimmune hyperthyroidism [14]. In the GPCR-mediated downstream signal transduction system, cyclic AMP responsive element-binding (CREB) protein has been shown to be an important transcription factor that is involved in the progression of hepatocellular carcinoma, leukemia, pituitary tumor, and lung cancer through control of cell function (secretion, proliferation, angiogenesis and apoptosis) [15–17]. To date, the cAMP/CREB mechanism in SI NETs has not been demonstrated.

In our previous evaluation of transcriptome analyses (Affymetrix U133 Plus chips) of the normal human EC cell and GI NET cell line KRJ-I, we identified candidate luminal GPCRs and neural/hormonal GPCRs including β1 adrenergic and dopamine D receptors (DR) [18,19]. Further investigation demonstrated that isoproterenol, a β-adrenergic GPCR agonist, stimulated 5-HT secretion through increased intracellular cAMP [20]. Others have shown, in PC12 (rat pheochromocytoma cells), HEK293T (human Embryonic Kidney cells) and the pancreatic beta cell line, MIN6, that activation of the cAMP pathway stimulates gene expression through protein kinase A (PKA)-mediated phosphorylation of CREB at Ser-133 [21,22]. Since little is known about neoplastic EC cell transcription and proliferation or secretion, we considered that delineation of the molecular basis of GPCR-mediated transcription through cAMP/PKA/CREB would provide novel information regarding the mechanistic basis of these processes and facilitate the identification of new therapeutic targets that might be used to inhibit NET function. As these tumors autoregulate their own growth through amine production [23] and regulate the local microenvironment (e.g. stimulate fibroblast proliferation and secretion) [24], delineating GPCR-pathways may identify novel targets to inhibit tumor cell proliferation.

We used gene network analysis and identified in silico the cAMP/CREB-mediated mechanisms of transcription in SI NETs. Using an established SI NET model, the human EC cell line (KRJ-I) [25], we validated GPCR-mediated transcription of CREB targets through cAMP/PKA/CREB-activation in SI NETs. Our results provide novel information regarding the transcription of CREB response elements (CREs) known to be relevant to tumor proliferation and secretion that are activated by GPCR regulation of intracellular cAMP. Furthermore, we offer the first formal network topology analysis of this disease.

Results

1. Systems-wide properties of a SI NET gene co-expression network

Gene co-expression patterns reflecting the pathogenesis of SI NETs were represented as undirected weighted network where nodes correspond to genes and edges correspond to co-expressions between them. We examined the network by systematically testing the Pearson correlation coefficient (PCC) cut-off in the range from 0.5 to 1 [Figure S1]. Only gene pairs with an absolute PCC ≥ 0.94 were included in the network. The reasons for selecting such stringent cut-off (0.94) were three-fold: i) gene correlation profiles with PCC over 0.60 are known to be more biologically relevant [26]; ii) at PCC < 0.94 the network was excessively large (most of the nodes were present), suggesting evidence of false-positive edges; iii) at PCC ≥ 0.94, a large number of connected components emerged, while the overall network density remained high, suggesting that genes were organized into tightly interconnected modules that may be functionally relevant [Figure S1]. The final network contained 3470 genes and 1549 links (average node degree = 2.6). Of these, 738 (23%) genes have known tumorigenic somatic mutations (obtained from the Catalogue of Somatic Mutations in Cancer (COSMIC) database).

It has been suggested that, in a co-expression network, functionally related genes tend to organize into tightly linked communities [27]. The Louvain algorithm was used to identify these functional modules in the SI NET network in an unbiased manner (see Methods). The network was partitioned into 882 clusters, of which 10 contained 20+ genes. Network modularity was 0.86, confirming that the SI NET interactome is embedded with highly interconnected modules, reinforcing the complex nature of signaling cascades in this disease [Figure 1A]. The top 10 clusters were enriched for Gene Ontology (GO) Biological Process (BP) terms. The most enriched terms included ‘Nervous system development’ (BEX1, SYN1, GRIA2), ‘Immune response’ (CD38, IGGc, SLAMF8), and ‘Cell cycle’ (ASPM, MKI67, TOP2A).

To determine the overall architecture of the SI NET interactome, the node degree frequency distribution was calculated and established to be “scale-free” [Figure 1B]. Generally, a scale-free architecture implies that most of the connections are confined to a few highly interconnected nodes (hubs) – a hallmark of most biological networks. However, it is possible to reconstruct similar connectivity patterns using random edge rewiring. To confirm that the SI NET network was non-random, we compared it to two models of random networks (see Methods): the Maslov-Sneppen model (scale-free architecture, preserved node degrees, randomly rewired edges) and the Erdős–Rényi model (preserved number of nodes, edges are constructed using a random Gaussian probability distribution). For each model, 200 random networks were generated and intersected with the original SI NET graph. On average, the Maslov-Sneppen and the Erdős–Rényi networks shared 78.3 (standard deviation = 7.7) and 229.1 (standard deviation = 15.3) links with the SI NET interactome respectively [Figure 1C]. This substantiates the hypothesis that the original network is significantly non-random (minimal z-score = 282.3).

To assess the stability of the SI NET network, we measured the effects of random (error) and targeted (attack) node removal on the network diameter. Removal of random nodes had no effect on the diameter, suggesting that the SI NET interactome was robust against random mutations. However, targeted removal of the most connected hubs, as predicted, caused the network to collapse [Figure 1D].

2. In Silico Prediction of CREB Targets

Interestingly, 940/3470 genes (27%) in the SI NET network were differentially expressed (student’s t-test, p≤0.05). Of these, 539 were up-regulated and 401 were down-regulated compared to normal SI mucosa. Eight genes (CHGA, CPE, ENO2, INSM1, PTPRN2, SERPINA10, and SLCL1BAI/2) that have been previously identified as markers of neuroendocrine tumors [28–31] were confirmed in this study to be altered. Automated KEGG pathway analysis (using the DAVID Functional Annotation Database) of differentially expressed genes, identified an over-represented (p = 0.03) GPCR signaling pathway characterized by an over-expression of cyclic AMP synthetase and adenylyl cyclase.
2 (ADCY2, t-value = 6.0) and ADCY9 (t-value = -3.8), and a PKA responsible for phosphorylation of the CREB transcription factor, PRKAR1A (t-value = 2.8) (Figure S2). The Wnt signaling pathway was also identified. These findings are consistent with previous studies in vitro of small intestinal and pituitary NET cell lines investigating cAMP recruitment through a GPCR complex and downstream elements of CREB phosphorylation [20,32–36].

We next compared up-regulated genes to a CREB Target Gene database (http://natural.salk.edu/CREB/) to identify potential CREs in SI NETs. Using a confidence level for the binding value (BV) < 0.001 and a binding ratio (BR) > 1.5, which are considered to be significant for the identification of CREs [22], a list of 123 genes representing putative CREB binding targets and cAMP response elements was compiled. Interestingly, the putative CREs localized only to the ‘Synaptic transmission’ and ‘Nervous system development’ clusters of the SI NET interactome, which is consistent with the biological function of known CREB targets as well as the nature of the neuroendocrine system [37] (Figure 2A).

We specifically selected for further investigation the CREs (BEX1, BICD1, CHGB, CPE, GABRB3, SCG2, and SCG3) given their known association with the regulation of cell function [16,29,30,38–40] (Table 1).

3. Real-time PCR Validation of the Gene Expression Analysis

To confirm the over-expression of CRE transcripts in SI NETs, we measured transcript expression by real-time PCR (RT-PCR) of n = 7 CREB targets (Table 1) in the SI NET cell line KRJ-I (n = 10) and in normal EC cell preparations (n = 8) (Figure 2B). Transcript levels were normalized to expression of housekeeping genes ALG9, TFCP2, and ZNF410 as described [41] using GeNorm [42]. Levels of BEX1, BICD1, CHGB, CPE, GABRB3 and SCG2 were up-regulated in SI NETs (p < 0.05) while Secretogranin III (SCG3) was not differently expressed, p = 0.24 (Figure 2B).

Additionally, we measured transcription of ADCY2 and PRKAR1A. Levels of these upstream CRE pathway regulators were elevated 700% and 722% respectively, in KRJ-I (p = 0.02) compared to normal EC cells (Figure 2B).
4. In vitro model of CRE transcription

To evaluate whether these genes were regulated through the cAMP signaling pathway in vitro, we investigated their expression in KRJ-I. CRE transcription was measured by stimulating KRJ-I cells with the cAMP activator forskolin (10^{-6} M and 10^{-5} M), the selective β-adrenergic receptor agonist isoproterenol (10^{-5} M), and the dopamine D_{2} (D_{2}R) and Serotonin (5-HT) receptor agonist BIM-53061 (10^{-6} M) (Figure 3) for two hours. While ADCY2 was significantly upregulated only by forskolin (10^{-5} M, 220%, p < 0.03), PRKAR1A transcripts were up-regulated both by forskolin and BIM-53061 (188%, p = 0.0004, and 153%, p = 0.021 respectively). Lower concentrations of forskolin (10^{-6} M) did not stimulate transcription of either ADCY2 or PRKAR1A. Overall, BIM-53061 was a universal CRE activator (100% of target genes transcriptionally activated), while the effects of forskolin (10^{-5} M, ~42% activated) and isoproterenol (~57% activated) were less pronounced.

5. cAMP/PKA and pCREB activation in vitro

Next, to confirm that the mechanisms regulating CRE transcription occurred through the cAMP signaling pathway, we measured cAMP in response to each of the ligands in the KRJ-I cell line. Twenty minute incubation with forskolin (10^{-5} M), isoproterenol (10^{-5} M) or BIM-53061 (10^{-6} M) all increased intracellular cAMP accumulation 1.72-fold, 1.67-fold, and 2.78-fold respectively compared to control (p < 0.05). Lower concentrations of forskolin (10^{-6} M) had no effect on cAMP accumulation (Figure 4A). PKA activity was similarly stimulated by these agents in the order of BIM-53061 = forskolin (10^{-5} M) (2.2–2.4-fold) > isoproterenol (1.85-fold) (Figure 4B), as was pCREB (BIM-53061 = forskolin (10^{-5} M) (1.75–1.9-fold) > isoproterenol (1.2-fold) (Figure 4C).

Discussion

The role of the cAMP signaling pathway in the regulation of tumor CREB-mediated transcription has not previously been investigated in gastrointestinal NETs. In the current study, using a transcript database of SI NETs and normal SI mucosa we demonstrated: 1) transcripts of ADCY2, a member of the adenylate cyclase family, are up-regulated in SI NETs and KRJ-I; 2) intracellular accumulation of cAMP is stimulated by forskolin,
isoproterenol and BIM-53061; 2) intracellular PKA activity and pCREB is stimulated by these agents; 4) cAMP-dependent protein kinase, PRKAR1A, is over-expressed in SI NETs and KRJ-I cells; 5) CREs are differentially transcribed when subject to a classic cAMP activator, a selective β-adrenergic receptor agonist, or a selective D2R and 5-HT receptor agonist. Additionally, we performed the first formal large scale network topology assessment of SI NET disease.

Initially, using gene network inference, we reconstructed the SI NET co-expression network from genome wide expression levels obtained from microarray profiling. The network (Table S1) was determined to be scale-free, non-random, and topologically stable. This suggests a system that is dominated by a few highly connected biologically relevant hubs and that is “protective” against random perturbations (e.g. mutations). Indeed, this is consistent with the behavior of most cellular networks [43] and confirms that the biology of SI NETs can be further probed using graph-theory approaches. The advantage of this approach is that it allows the study of significant genes in relation to the entire system rather than by merits of up- or down-regulation alone. For example, node betweenness centrality as well as node degree have been previously reported as possible indicators of gene essentiality [44].

We computed these statistics for every gene in the SI NET network. Thus our dataset can further be explored using functional assays utilizing network topology as well as differential expression.

Highly modular structure of the SI NET network was explored using an unbiased graph clustering technique (the Louvain algorithm). The method is a greedy optimization method that attempts to optimize the modularity [45] of a partition of the network. We identified 10 modules (>20 genes) in the SI NET that were subsequently enriched for GO-BP terms including ‘Nervous system development’ and ‘Cell cycle’. The functional cluster heterogeneity suggests that the SI NET disease is a multi-modal entity with complex metabolic, hormonal, and proliferative cascades that call for a systems-wide assessment as well as traditional approaches.

We used differential expression analysis to map significantly changed genes onto the SI NET network to increase the biological utility of the analysis. Most of the significantly changed genes formed tight networks involved in transcription, secretion, cell proliferation, tissue development, embryonic development and extracellular matrix regulation. Using the DAVID functional annotation tool [46], it was determined that cAMP/CREB signaling cascade was highly upregulated in SI NETs (Figure S2). It was of interest to note that the statistical enrichment also identified that the Wnt signaling pathway was similarly altered. It was previously suggested that the cAMP/CREB signaling may also contribute to Wnt-regulated processes in cancer [47]. It appears that our network analysis reiterates this concept; however, further implications of this finding in NET biology need to be investigated.

We further explored the CREB mechanism in silico by identifying possible CREB binding targets and cAMP response elements among the significantly altered genes using the Salk CREB Target Gene database [22]. These CRE targets encoded genes responsible for nucleosome assembly (NAP1L3, TSPYL4), regulation of transcription (TERF2IP), organism development

### Table 1. 7 CREB targets assessed by Real-time PCR in a SI NET database and the KRJ-I cell line.

| Gene Symbol | Gene Title | CREB p-value | CREB binding ratio | GO Biological Process | Chromosomal Location |
|-------------|------------|--------------|--------------------|-----------------------|----------------------|
| BEX1        | brain expressed, X-linked 1 | 5.00E-05 | 2.1 | multicellular organismal development | Xq21-q23 |
| BICD1       | bicaudal D homolog 1 (Drosophila) | 1.30E-03 | 1.7 | RNA processing | 12p11.2-p11.1 |
| CHGB        | chromogranin B (secretogranin 1) | 7.50E-08 | 3 | protein modification process | 4q32.3 |
| CPE         | carboxypeptidase E | 1.10E-12 | 7.4 | proteolysis | 4q32.3 |
| GABRB3      | gamma-aminobutyric acid (GABA) A receptor, beta 3 | 1.20E-02 | 1.5 | Transport | 15q11.2-q12 |
| SCG2        | secretogranin II (chromogranin C) | 1.70E-04 | 1.9 | Angiogenesis | 2q35-q36 |
| SCG3        | secretogranin III | 2.50E-05 | 2.2 | protein secretion | 15q21 |

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Because we were specifically interested in genes involved in cAMP-mediated secretory processes, we examined this subset further.

The accumulation of cAMP in response to activation of GPCRs induces a wide range of cellular processes including transcription, metabolism, cell cycle progression and apoptosis through the PKA pathway [48]. In this study, transcript of *PRKAR1A*, the type 1alpha regulatory subunit (RIalpha) of PKA, was up-regulated in KRJ-I and stimulated by forskolin and BIM-53061. This is consistent with the function of these compounds as inducers of the cAMP pathway. The selective D2R and 5-HT receptor agonist, BIM-53061 appears to be at least as potent a cAMP recruiter as either forskolin or isoproterenol particularly for secretory gene transcription. This suggests the involvement of the dopamine/5-HT-mediated pathway in the recruitment of intracellular cAMP/PKA activation. In addition, *PRKAR1A* transcript levels stimulated by BIM-53061 were consistent with the accumulation of intracellular cAMP suggesting a direct involvement of neural GPCR receptor activation with *ADCY2* and *PRKAR1A* recruitment and subsequent PKA-induced CREB phosphorylation.

We have demonstrated that elevation in cAMP is associated with normal and neoplastic EC cell secretion [18,49]. In the present study, we identified elevated levels of *ADCY2* and *PRKAR1A* transcripts in a database of SI NETs compared to normal SI mucosa, suggesting that cAMP signaling may indeed be activated in tumor cells [32,50]. *ADCY2* is a class B member of the Adenylate Cyclase (ADCY) which is calcium insensitive but is stimulated by Gbc subunits of heterotrimeric G-proteins and is therefore directly coupled with GPCRs [35]. In the KRJ-I cell line, transcript levels of *ADCY2* were sensitive to forskolin and BIM-53061, which suggests that cAMP-induced transcription may occur through activation of this cyclase regulator in the KRJ-I cell line.

Cellular gene expression is regulated following CREB protein phosphorylation at serine residue 133 [21,51]. This occurs as a consequence of cAMP accumulation which liberates the C subunits of PKA that passively diffuse into the nucleus and induce CREB phosphorylation. CREB is an important transcription factor activated by multiple signal transduction pathways in response to external stimuli, including synaptic activity, hormones, growth factors, cytokines, and stress [15]. It affects cellular transcription and translation.
functions and enhances growth, increases angiogenesis, and decreases apoptosis. We identified in silico and demonstrated in vitro cAMP-mediated regulation of seven putative CREB targets – BEX1 (modulates nerve growth factor [NGF] signaling through nuclear factor-kappaB [NFkB] to regulate cell cycle, apoptosis, and differentiation in neural tissues [39]), BICD1 (structural constituent of cytoskeleton [39]), CHGB (neuroendocrine cell-specific gene, which may play a role in early tumor development but is also a NET secretory product [16,52]), CPE (pulmonary NET marker [29]), GABBR3 (characteristic GABA receptor of normal and neoplastic human EC cells and controlled through CREB [20,36,40]), SCG2 (secreted neuroendocrine marker observed in prostatic small-cell neuroendocrine carcinoma [30]) and SCG3 (secretory product commonly expressed in pituitary adenomas [53]). In vitro investigations of these CREs indicate that cAMP activation through either adenylate cyclase activation (forskolin) or through neural GPCR activation (isoproterenol and dopamine/5-HT) resulted in gene expression. CRE transcription correlated with cAMP levels: BICD1, which was associated with the highest cAMP (2.8-fold) accumulation, was also associated with the majority of genes transcribed (100%). Both forskolin and isoproterenol stimulated cAMP levels 1.8-fold and were associated with 42–57% of target genes being transcribed. The suggestion that gene transcription was cAMP concentration-dependent was reinforced by the observation that forskolin (10^{-5} M), which did not significantly elevate cAMP, was not associated with CRE target transcription. Similar investigations in the rat pituitary cell line GH4 have shown that forskolin-induced cAMP accumulation results in an increase of prolactin and growth hormone gene transcription [54], suggesting that a single intracellular mediator can simultaneously regulate the transcription of different sets of responsive genes by stimulating independent biochemical events.

The study provides an illustration of how genome-wide network inference can be used to infer CRE-mediated transcription in neoplastic cell lines and has implications for defining the mechanisms of NET proliferation and secretion. Similar studies examining the progression of cancer [53,56], heart disease [57], neuropsychiatric disorders [58,59], asthma pathogenesis [60], and the analysis of factors associated with infertility [61] have provided information in regard to these disease processes. We propose that the application of this methodology to the investigation of NETs or other diseases associated with abnormal EC cell secretion, like Crohn’s disease [6] or IBS [62], will provide significant mechanistic information on the cell regulatory phenomena. Our current data demonstrates that neoplastic EC cells over-express regulators in the cAMP signaling pathway and that activation of neural GPCRs results in proliferative and secretory gene transcription thus providing novel information regarding the neural activation of tumor behavior. This investigative strategy, that emphasizes co-expression network inference, provides a useful tool to define and delineate the mechanisms involved in the mechanistic cellular basis of the clinical manifestations of NET disease. It is likely that the application of this technique will facilitate the identification of specific regulatory elements that can be targeted for therapeutic gain.

**Materials and Methods**

**Statistical analyses of Affymetrix GeneChip data**

Raw expression data for each of the 13 microarray experiments (Affymetrix U133A; normal mucosa: n = 4; primary SI NETs: n = 9) was normalized using the MAS5.0 algorithm available through the Bioconductor suit [63] for the R statistical language [64]. Affymetrix probe identifiers (IDs) were mapped to their corresponding Ensembl (September 26, 2010) gene IDs [65]. In cases where multiple probesets mapped to a single gene, only median signal intensity was retained. Data is deposited in the ArrayExpress database (accession number: E-GEOD-6272).

**Gene Network Inference**

Pairwise similarity in gene expression vectors was expressed by the PCC. Gene pairs that correlated above a predefined PCC threshold value were represented in the form of an undirected weighted network, where nodes (vertices) correspond to genes and links (edges) correspond to co-expression between genes. The Maslov-Sneppen randomized network model was generated by
rewiring edges in the original network while preserving the degrees of the respective nodes [66]. The number of rewiring steps taken for each model was 4 × (number of edges). This method ensures that the topological structure of the network is retained during randomization. The Erdős–Rényi random model was generated by retaining the nodes of the original network and building edges using a uniform probability [67].

Network Topology Concepts

Topological properties examined were node degree, network diameter, betweenness centrality, connected components, clustering coefficient, and modularity [68]. Node degree is defined as the total number of edges that connect to a given node. Network diameter is defined as the average shortest path between any pair of nodes in the network. Betweenness centrality is the measure of node importance within a graph, where nodes that occur on many shortest paths between nodes have higher betweenness. Connected components are maximal connected subgraphs of an undirected graph in which any two vertices are connected to each other by edges. Clustering coefficient is the degree to which nodes tend to cluster together. Modularity quantifies the capacity of a network to divide into clusters or communities. Higher modularity indicates a favorable partition.

Network clustering and functional enrichment

Clusters of genes in a co-expression network were identified using the Louvain method, a fast algorithm for community detection in graphs [69]. The Louvain method is a greedy algorithm for iterative grouping of nodes into communities based on optimization of modularity [45]. A distinct advantage of this method is its parameter-free architecture that allows unbiased exploration of network structure. Because clusters of co-expressed genes are known to be functionally related [27], functional enrichment for GO-BP terms was performed. For a cluster with n genes and an a priori defined functional category with K genes, the hypergeometric test was used to evaluate the significance of the overlap k between the cluster and a functional category [70]. All genes in a network were used as reference.

Pathway Analysis

Over-represented pathway analysis was performed using the DAVID functional annotation tool [46] and prediction of CREB target phosphorylation was assessed using CREB target gene database (http://natural.salk.edu/CREB/) [22] with a confidence level of the binding value (BV) ≥ 0.001 and a binding ratio (BR) ≥ 1.5.

Validation and in vitro experiments

Culture Conditions. KRJ-I cells, derived from a “typical” SI NET [18,71], were cultured as floating aggregates at 37°C with 5% CO2. Cells were kept in Ham’s F12 medium (Gibco®) containing 10% fetal bovine serum (FBS) (Sigma-Aldrich), penicillin 100 U/ml, and streptomycin 100 μg/ml [18,25].

Real-Time PCR. To validate the presence of genes involved in cAMP-mediated transcription pathway, two approaches were undertaken. In the first approach, transcripts for selected CREs, ADCY2, and PKNAR1A were measured in an independent data set of neoplastic EC cell line KRJ-I (n = 10) and normal EC cell preparations (n = 8) using real-time PCR. In the second approach, the effect of forskolin (10−5 M and 10−6 M), isoproterenol (10−5 M), and BIM-35061 (10−6 M) was measured on target transcription in KRJ-I cells. KRJ-I cells (5×104 cells/well, in triplicate) were stimulated for 2 hours and RNA was extracted from 1×106 cells in log phase growth (TRIZOL®, Invitrogen, USA). Real time RT-PCR analysis was performed using Assays-on-Demand™ products and the ABI 7900 Sequence Detection System according to the manufacturer’s suggestions. Cycling was performed under standard conditions (TaqMan® Universal PCR Master Mix Protocol) and data normalized using GeNorm [42] and expression of the novel house-keeping genes, ALG9, TFCP2 and ZNF410 [41].

cAMP and PKA Activation. To test whether KRJ-I cells were physiologically responsive to neural GPCR agonists, intracellular cAMP accumulation in response to the three stimulants after 20 mins was assayed using a cAMP ELISA assay (R&D Research, Minneapolis, MN). PKA activity was quantitated in the same samples (Enzo Life Sciences, Butler Pike, PA). Cells (5×104 cells/well, in triplicate) were stimulated with forskolin (10−5 M, 10−6 M, isoproterenol (10−5 M), and BIM-35061 (10−6 M) after which cells were lysed with 0.1 N HCL and freezing. All samples and controls were acetylated prior to performing the cAMP ELISA (R&D cAMP ELISA handbook). PKA activity was determined according to the manufacturer’s recommendations. Lysed samples were incubated with 20 μl PKA reaction mixture at 30°C for 30 min. The reaction was terminated and activity quantitated versus levels of a highly specific substrate using an ELISA protocol. Absorbance readings for either cAMP or PKA were measured at 450 nm on a microplate reader (Bio-Rad 3500).

pCREB quantitation - western Blotting. KRJ-I cells (4×106 cells/ml) were seeded in 6 well plates (Falcon, BD, Franklin Lakes, NJ) and treated with each of the agents for 15 and 60 mins. After cells were harvested, whole-cell lysates were prepared by adding 200 μl of ice-cold cell lysis buffer (10× RIPA lysis buffer (Millipore, Billerica, MA), complete protease inhibitor [Roche, Indianapolis, IN], phosphatase inhibitor set 1&2 (Calbiochem, Gibbstown, NJ), 100 mM PMSF (Roche), 200 mM NaVO4 (Acros Organics), 12.5 mg/ml SDS (American Bioanalytical, Natick, MA). Tubes were centrifuged at 12,000 g for 20 min and protein amount in the supernatant was quantified using the BCA protein assay kit (Thermo Fisher Scientific, Rockford, IL). For western blot, total protein lysates (20 μg) were denatured in SDS sample buffer, separated on an SDS-PAGE gel (4, 10%) and transferred to a PVDF membrane (Bio-Rad, Hercules, CA, pore size 0.45 mm). After blocking (5% BSA for 60 min at room temperature) the membrane was incubated with the phospho-CREB (Ser133) primary antibody (Cell Signaling Technology, Danvers, MA) in 5% BSA/PBS/Tween 20 overnight at 4°C. The membranes were incubated with the horseradish peroxidase-conjugated secondary antibodies (Cell Signaling Technology) for 60 min at room temperature and immunodetection was performed using the Western Lightning™ Plus-ECL (PerkinElmer, MA). Blots were exposed on X-OMAT-AR films. The optical density of the appropriately sized bands was measured using ImageJ software (NIH, USA). The ratio between phospho-protein expression was reported relative to that of β-actin (Sigma-Aldrich, MO).

Supporting Information

Figure S1 SI NET network properties as functions of Pearson correlation coefficient (PCC). For each PCC cutoff, the number of nodes, number of edges, number of connected components, and network density were measured. It was noted that at PCC≥0.94, the SI NET network was most modular while retaining a reasonable number of genes and links. (TIF)
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