Gold-nanofève surface-enhanced Raman spectroscopy visualizes hypotaurine as a robust anti-oxidant consumed in cancer survival

Megumi Shiota1, Masayuki Naya1, Takehiro Yamamoto2, Takako Hishiki2, Takeharu Tani1, Hiroyuki Takahashi3, Akiko Kubo2, Daisuke Koike4, Mai Itoh2, Mitsuyo Ohmura2, Yasuaki Kabe2, Yuki Sugiura2, Nobuyoshi Hiraoka5, Takayuki Morikawa6, Keiyo Takubo6, Kentaro Suina7, Hideaki Nagashima7, Oltea Sampetrean7, Osamu Nagano7, Hideyuki Saya7, Shogo Yamazoe1, Hiroyuki Watanabe1 & Makoto Suematsu2

Gold deposition with diagonal angle towards boehmite-based nanostructure creates random arrays of horse-bean-shaped nanostructures named gold-nanofève (GNF). GNF generates many electromagnetic hotspots as surface-enhanced Raman spectroscopy (SERS) excitation sources, and enables large-area visualization of molecular vibration fingerprints of metabolites in human cancer xenografts in livers of immunodeficient mice with sufficient sensitivity and uniformity. Differential screening of GNF-SERS signals in tumours and those in parenchyma demarcated tumour boundaries in liver tissues. Furthermore, GNF-SERS combined with quantum chemical calculation identified cysteine-derived glutathione and hypotaurine (HT) as tumour-dominant and parenchyma-dominant metabolites, respectively. CD44 knockdown in cancer diminished glutathione, but not HT in tumours. Mechanisms whereby tumours sustained HT under CD44-knockdown conditions include upregulation of PHGDH, PSAT1 and PSPH that drove glycolysis-dependent activation of serine/glycine-cleavage systems to provide one-methyl group for HT synthesis. HT was rapidly converted into taurine in cancer cells, suggesting that HT is a robust anti-oxidant for their survival under glutathione-suppressed conditions.
ew technology that visualizes multiple metabolites in a large surface area of tissues, but does not require labelling or staining, could improve pathological diagnosis. Raman scattering is a powerful analytical technique for objective, label-free tissue diagnosis\textsuperscript{1–3}. Unlike conventional Raman spectroscopy, surface-enhanced Raman spectroscopy (SERS) yields highly sensitive signals of vibrational fingerprints of metabolites in the presence of reporter metals such as gold (Au) or silver\textsuperscript{4,5}. Although SERS technology has been successful in biosensing and biomedical contexts: previous reports described the imaging of molecules or drug compounds at the single cell levels or in ex vivo biofluid samples\textsuperscript{6–8}. SERS imaging of large areas of tissues, with high sensitivity, spatial resolution and reproducibility, has been unavailable. Challenges in developing devices for practical use include the design of highly sensitive SERS—active substrates that guarantee spatial uniformity of hotspots, to amplify SERS signals by enhancing local electromagnetic fields\textsuperscript{9}. Other difficulties arise from the technical principle underlaying SERS, which detects multiple molecular vibration modes as fingerprints when identifying metabolites\textsuperscript{10}. In tissues, metabolites yield numerous SERS signals and complicate identification of their discernible molecular entities in individual Raman peaks. However, SERS imaging is a comprehensive technique for visualizing metabolic profiles at multiple different wave numbers, and requires only a modest laser excitation, which minimizes artificial oxidation of metabolites. These characteristics are useful in accurately determining biomarker metabolites in diagnosing cancer. Development of a highly sensitive SERS imaging modality that can cover a large area aids our understanding of the metabolic interplay between cancer cells and surrounding tissues and assists in objective and automated identification of tumour (T) boundaries in tissues\textsuperscript{9,10}.

The current SERS technique enables precise control of Au nanostructures of ideal geometry and produces gold nanofève (GNF), which are named after the horse bean-shaped Au nanoparticles. GNF provides SERS excitation sources, thereby yielding numerous strong signals derived from metabolites in livers bearing metastatic cancer xenografts. Non-target differentiation of such signals in T and parenchyma (P) helped identify T boundaries in frozen tissue sections by means of automated

Fig. 1 Characterization of a random array of the gold-nanofève (GNF) substrate for SERS imaging. a Schematic diagram of SERS imaging spectrometry system. Note that the system is based on the inverted-type microscope in which the GNF substrate is optically transparent, and allows us to visualize SERS signals from the bottom of the substrate without being interfered with the thickness of tissue slices on the microscopic stage. b Nano-fabrication of GNF substrates. Deposition of aluminium (Al) onto the surface of a film layer is followed by immersion of the Al-coated film into boiling water as a hydrothermal treatment that forms thin, sharp-edged leaf-like superstructure of boehmite, as shown previously from our laboratories\textsuperscript{15}. Au deposition to the Boehmite surface is carried out through desired oblique angles. c SEM images showing alterations in quasi-parallel alignment of anisotropic Au nanostructures at deposition angle at 80°. Note that the angles at 45° or 0° cause no anisotropy of Au nanoparticles. The side and top views of the GNF substrates are shown in the left and right panels, respectively. Bar = 500 nm. Parallel-polar and Perpendicular-polar: the parallel and side directions of polarized light to generate SERS signals, respectively. White targets and arrows indicate the direction of the Au deposition. d A representative top-view SEM image with high magnification. Bar = 100 nm. e A representative picture depicting differences in SERS spectra of rhodamine 6 G (R6G) between GNF and GNC substrates. Comparison of the intensity at 1360 cm\textsuperscript{-1} revealed that GNF substrates provides evidence that GNF-mediated SERS enhancement is 18.6-fold greater than that mediated by GNC.
processes requiring no labelling. Furthermore, careful characterization of individual GNF-SERS signals enabled identification of glutathione (GS) at 298 cm$^{-1}$ or hypotaurine (HT) at 978 cm$^{-1}$, and retinoids at 1150 cm$^{-1}$. GS in T increased with development of cancer metastases, and was suppressed by knockdown of CD44, while HT increased in cancer cells, a molecule stabilizing cystine trans-porter (xCT) as part of cancer survival$^{11,12}$. Interestingly, although biosynthesis of GS and HT requires cysteine as a substrate,$^{13,14}$ HT, but not GS, in cancer cells serves as a robust CD44/xCT-independent anti-oxidant that is rapidly converted to taurine, and compensates for the decreased capacity of cancer to attenuate oxidative stress during CD44-knockdown-induced GS suppression.

Results
Characterizing GNF substrates for large-area SERS imaging. We previously fabricated gold-nanocoral (GNC) substrates to perform large-area SERS imaging$^{15}$. However, detection of larger numbers of metabolites with better sensitivity and selectivity was necessary. We therefore developed highly sensitive SERS substrates for Raman spectroscopy (Fig. 1a): the Au deposition from a perpendicular direction to the substrate (0° in the deposition angle in Fig. 1b) initiated growth of Au-nanospheres on the ledge of a boehmite structure (Fig. 1c: GNC substrate). Use of an oblique Au deposition technique$^{16}$ at an angle of 80°, but not of 45° or 0°, resulted in highly porous and anisotropic island growth of Au nanoparticles through their self-organized assembly without use of lithographic techniques (Fig. 1c, d). Scanning electron microscopic (SEM) images showed a random array of the GNF substrate, which is named after the anisotropic shape of the Au nanostructure, as it resembles the shape of horse beans, and is distinct from spheroid GNC$^{15}$. SEM (Fig. 1c, d) revealed that the interparticle space between Au nanoparticles formed by the deposition angle of 80° is larger than that of 0° and 45°. This morphological feature improved GNF-SERS sensitivity by increasing metabolite accessibility. Optimization of the GNF design (Supplementary Note 1 and Supplementary Fig. 1)$^{17}$ yielded a SERS enhancement by 18.6-fold versus GNC (Fig. 1e).

Simulation of electromagnetic fields around a GNF unit. To understand mechanisms by which GNF substrates enhance SERS signals, we attempted to determine the anisotropic features of the nanostructure of an individual GNF. Binary images of top-view SEM images (Fig. 2a) displayed heterogeneously elliptical regions as bright areas (a red square in Fig. 2a), and thus enabled to measure maximum lengths (parallel to the direction of Au deposition) and widths (perpendicular to the direction of Au deposition) of individual Au nanoparticles. These measurements in three separate lots of GNF substrates showed that length was greater than width in most GNF units and that the average length/width (L/W) ratio was 1.0:5.0 (Fig. 2b), and enabled to simplify the morphological feature of Au nanoparticles in GNF (Fig. 2c), which is suitable for finite-difference time-domain (FDTD) simulation$^{18,19}$ to determine effects of various lengths and widths of model particles on enhancement of electromagnetic fields around individual Au nanoparticles. Local field enhancement ($G = |E/E_0|^2$) was greatest in the two polar apaxes of a single unit (Fig. 2d). Superimposition of the local field enhancement (G in Fig. 2d, e) on the distribution of lengths and widths (Fig. 2b) showed that most of individual Au nanoparticles in GNF

Fig. 2 Finite-difference time-domain (FDTD) simulation for reasoning mechanisms by which electromagnetic fields are enhanced around the individual anisotropic Au nanostructures of GNF substrates. a A representative binary image collected from a top-view SEM image. b Distribution of lengths of individual Au nanoparticles (a red square in a indicates a representative one) as a function of their widths. Note that, in separate analyses using 3 different lots of the GNF substrate, a major population of the nanoparticles are plotted in a range between the lines L = W and L = 5 W. c A simple model of the Au nanoparticle. Red point and circle indicate the direction of the incident light, while the blue arrow shows the direction of polarization of the light. The thickness of the Au nanoparticle was fixed at 40 nm, and the points at 1-nm far from the virtual surface of the particles were determined for simulating electromagnetic fields. d A representative simulation of electromagnetic field in and around a representative model of a single Au nanoparticle with the L/W values of 3.8 (L = 190 nm, W = 50 nm) and the thickness of 40 nm, indicating the log G value of 4.7. e Enhancement of electromagnetic fields and its relationship with L/W ratios. Note that Au particles with the L/W ratio in a range between 2.0 and 5.0 exhibited greatest enhancement of electromagnetic fields (yellow in Fig. 2e).
substrates were in regions indicating greater enhancement of electromagnetic fields (Fig. 2e). Among them, Au nanoparticles with the L/W ratio between 2.0 and 5.0 and a length less than 250 nm exhibited the greatest enhancement of electromagnetic fields (Fig. 2e). Furthermore, Au nanoparticles with greater lengths and widths exhibited smaller G values (green regions in Fig. 2e), suggesting that morphological anisotropy with optimal lengths of Au nanoparticles is crucial for improving SERS excitation sources. According to FDTD simulation, the local field enhancement by two adjacent Au nanoparticles of GNC are diminished by increasing the gap distance over 10 nm (typical gap distance of GNF in Fig. 2a), but the enhancement by those of GNF did not regress but stayed a plateau level (Supplementary Figs 2a-c). These results suggest that the anisotropy of Au nanoparticles rather than effects of the gap of the Au-nanoparticle, which is also called junction, determine SERS enhancement in GNF substrates, being in good agreement with previous studies.

Automated extraction of tumour boundaries. As described in the Methods, three regions—T, P and vessels (V)—were manually extracted from binary images (Fig. 3a). Raman spectra between 200 cm\(^{-1}\) and 1600 cm\(^{-1}\) in regions P and T were subtracted to generate difference spectra between T and P (Fig. 3b). Two were higher in T than in P (red arrows at 298 cm\(^{-1}\) and 480 cm\(^{-1}\)), and two were higher in P than in T (blue arrows at 978 cm\(^{-1}\) and 1150 cm\(^{-1}\)). At these peaks, GNF-SERS imaging yielded actual images of the T-bearing liver (Fig. 3c). To generate these images, Raman shifts at specific pixels were accumulated and processed.
by a median filter (see Methods). Comparison of regions T and P indicated that raw SERS signals at 298 cm\(^{-1}\) were significantly greater in T than in P. Raw signals in T and P were significantly attenuated by xenografting with HCT116 cells pretreated with shRNA against CD44, a surface antigen responsible for stabilizing xCT for cancer stemness and survival\(^{11,12}\). While the peak at 480 cm\(^{-1}\) was marginally higher in T than in P (Fig. 3b, d) without statistical significance, the peak identification will be examined later. In contrast, the peaks at 978 cm\(^{-1}\) and 1150 cm\(^{-1}\) were significantly higher for P than for T (Fig. 3d). Interestingly, both peaks were insensitive to the CD44 knockdown in cancer cells. Three peaks displaying notable differences between regions T and P were further examined for subsequent identifications of specific metabolites later.

Cancer cells alter their metabolism to generate precursors of macromolecules necessary for their proliferation and survival. These alterations in metabolic systems may help distinguish T boundaries. T/P ratio images were processed with a newly developed method that uses 2 different median filters; one processes images through a radius of 1 pixel and yields sharply contrasted T boundaries with high noise levels. The other median filter processes images through a radius of 2 pixels and yields smooth and noiseless images with wider boundaries between T and P. These image sources were finally multiplied the two binary images in which signals from T regions became 0 (flowchart-b of Fig. 4) and Supplementary Fig. 3a was used to differentially accumulate T-dominant and P-dominant Raman shifts collected from liver tissue xenografted with shControl cells. This process resulted in building the accumulative lists of Raman shifts (ALR in flowchart-b of Fig. 4). Flowchart-b (Fig. 4 and Supplementary Fig. 3b) was used to extract T areas as very bright regions by calculating the ratio images between averaged T/P ratios (T-bar-dominant over P-bar-dominant images in flowchart-b of Fig. 4) in liver tissue xenografts in three groups (shControl, Lenti-control in which cancer cells were transfected with the virus vector alone, and shCD44; see Methods).

The image-processing procedure in flowchart-c (Fig. 4 and Supplementary Fig. 3c) was the final step for identifying boundaries. T/P ratio images collected from the three groups were processed with a newly developed method that uses 2 different median filters; one processes images through a radius of one pixel and yields sharply contrasted T boundaries with high noise levels. The other median filter processes images through a radius of 2 pixels and yields smooth and noiseless images with wider boundaries between T and P. These image sources were binarized to generate one-pixel- and two-pixel-based smoothened images (0 in P and 1 in T regions) and finally multiplied the two binarized images in which signals from T regions became 0 (flowchart in Supplementary Fig. 3c) (see Methods). The accuracy rates for the groups will be compared below. However, the present results suggest that image processing of global GNF-SERS improves automated identification of boundaries in T-bearing liver tissues.
being in good agreement with the peak for hypotaurine in solution as well as in SERS imaging (Fig.3). Raman spectra. The Raman peaks marked as # around 970~980 cm−1 are S = O stretching mode. Of note is that the peak in HT-Au3 occurred at 977 cm−1, being in good agreement with the peak for hypotaurine in solution as well as in SERS imaging (Fig. 3).

Identification of GS and HT by GNF-SERS. Identifying metabolites by SERS in vivo requires discrimination of many individual SERS signals yielded by various metabolites. The Raman shift at around 300 cm−1 is Au-S stretching mode, and the present observation that the signal at 298 cm−1 is sensitive to CD44 knockdown (Fig. 3d) led us to examine if the peak predominantly detected in T at 298 cm−1 results from thiols. Thus, SERS signals for varied thiol metabolites were examined on GNF substrates after sufficient concentrations of analytes. These measurements utilized natural-dried aqueous solutions, and the relationship of the original concentrations of analytes to analyte densities on GNF substrates was described in Supplementary Fig. 11. Physiologic concentrations of thiols other than GS, and taurine did neither generate the SERS signals yielded by various metabolites. The Raman shift at around 300 cm−1 or at 978 cm−1, as judged by the data collected by MS and imaging MS analyses in Supplementary Figs. 4-10, respectively. We therefore assigned them to Au-S stretching mode (GS), and Au-S stretching mode (HT), respectively.

Quantum chemical calculation to characterize GSH and HT. To determine whether the signals at 298 cm−1 and 978 cm−1 were derived from GS and HT, we used quantum chemical modeling to identify specific the molecular vibration modes of these metabolites in silico. This method allowed us to assign molecular vibration modes, which result from molecular interactions between GSH or HT and atomic Au surface (Supplementary Fig. 12 and Fig. 5 and Supplementary Note 1)32-34. First, to optimize the structure of a pseudo-Au surface for binding models for GS and HT, we simulated wave numbers of Au-S stretching mode generated by methyl-mercaptane with various numbers of Au-atom clusters (CH3S-Aux, Supplementary Fig. 12). The results indicating frequency peaks for Au-S stretching mode showed that the notable shift occurred between CH3S-Au1 and CH3S-Au2 and reached a plateau at 300 cm−1 as the number of Au atoms increased (Supplementary Figs. 12b, c). A pseudo-Au surface for binding GS and HT was thus constructed by using triangular Au3 structure in which Au-S interactions were stable.

The absence of Au did not generate notable GSH vibration modes around 300 cm−1, but the presence of GS-Au1 induced a strong Au-S stretching peak at 302.8 cm−1 (Fig. 5a, b). The Au-free HT spectrum (blue line in Fig. 5d), generated a weak S = O stretching mode peak at 936 cm−1, while the presence of Au1 cluster enhanced and shifted the peak at 978 cm−1 (Fig. 5c, d), which suggests that GNF-mediated SERS signals at 298 cm−1 or 978 cm−1 were produced by interactions of Au with GS or HT, respectively. We therefore assigned them to Au-S stretching mode (GSH) and S = O stretching mode (HT), respectively.

**GNF-SERS imaging of livers bearing colon cancer xenografts.** GS and HT are metabolites synthesized from cystine provided from trans-sulfuration pathway and/or from xCT/CD44 that helps cystine incorporation from the extracellular space. CD44 interference by shRNA (shCD44) downregulates CD44s and its splicing variant CD44v in HCT116 cells, thereby suppressing T metastases41,12,35 (Fig. 6a, b). Interestingly, shCD44 and
shControl cells expressed multiple trans-sulfuration enzymes including cystathionine β-synthase (CBS), cystathionine γ-lyase (CTH). An HT-synthesizing cysteine dioxygenase (CDO) was comparably expressed, while expression of another HT-synthesizing enzyme, 2-aminoethanethiol dioxygenase (ADO) was notably diminished along with decreases in ATP, energy charge and total GS in shCD44 as compared with shControl cells (Fig. 6a and Supplementary Table 2). Large-area GNF-SERS imaging achieved by the data calibration (Supplementary Fig. 4) indicated that GS and HT distributions were homogeneous among lobules in the normal group. In xenograft-transplanted livers, T exhibited significant increases in GS and decreases in HT as compared with the surrounding P (Fig. 6c, d). When transplanted with shCD44-treated HCT116 cells, GS signals in T were significantly decreased. Interestingly, GS signals in P were also significantly decreased, which suggests that interference with CD44 in cancer suppresses GS in both T and P (Fig. 6c, d). In contrast, HT densities in P were unchanged in T-bearing livers xenografted with shCD44-HCT116 cells, and significantly greater than that in T (Fig. 6d).

Although significantly lower than in P, calibrated amounts of HT in T, as determined by GNF-SERS, was ~1.8 pmol mm⁻² in the apparent analyte density, which was comparable to the apparent analyte density of GS (Fig. 6d). This result is inconsistent with the findings of HT imaging by vacuum-type imaging MS, which showed that HT was undetectable in T (Supplementary Fig. 10). Furthermore, unlike GS, CD44-KD in the cancer cells did not cause a decrease in HT (Fig. 6). These results led us to examine the possibility of an alternative pathway for providing HT, one that does not depend on the xCT/CD44 complex.

Cancer cells silencing CD44 require HT for survival. We examined whether CE-MS data might underestimate accurate HT amounts in cancer cells, because of its artificial oxidation and/or its rapid turnover in T. Western blotting (Fig. 6a) showed that ADO was diminished; thus, shCD44 cells might utilize glucose to provide 3-phosphoglycerate (3-PG) and ω-serine as a one carbon resource for boosting HT synthesis and subsequent taurine synthesis through oxidative stress in the cells. We then used CE-MS metabolomics to identify differences between shControl and shCD44 cells in conversion of 13C₆-labelled glucose towards glyceraldehyde 3-phosphate (GAPG), which was then quantified by vacuum-type imaging MS, which showed that HT was undetectable in T (Supplementary Fig. 10). Furthermore, unlike GS, CD44-KD in the cancer cells did not cause a decrease in HT (Fig. 6).

### Fig. 6: Semi-quantitative analyses of glutathione (GS) and hypotaurine (HT) in tumour-bearing livers.

- **a** Western blot analyses of CD44 expression in HCT116 cells which were pretreated with control shRNA (shControl) or with CD44 shRNA (shCD44). CBS: cystathionine β-synthase. CSE: cystathionine γ-lyase. CDO: cysteine dioxygenase. ADO: 2-aminoethanethiol dioxygenase. Note that ADO is markedly down-regulated in shCD44 cells. GAPDH is an internal control.
- **b** Differences in % tumour regions in the tumour-bearing (TB) livers, showing that CD44 silencing causes tumour suppression. Statistical significance was analyzed by unpaired Student’s t-test. *P < 0.05 versus the shControl group. T(7) = 2.765.*  
  Representative pictures of large-area SERS images of glutathione (GS) and hypotaurine (HT) which were calibrated as apparent analyte density (AAD) for GS and HT, respectively. Bar = 1.0 mm. Dotted lines indicate location of several metastatic tumours. 
- **c** Differences in apparent GS density and apparent HT density. P and T depict parenchyma and tumour regions, respectively. Blue: tumour-free controls, brown: shControl HCT116 xenografting and orange: shCD44 HCT116 xenografting. Statistical analyses were performed by ANOVA with Fischer’s LSD test. *Pp < 0.05 was considered statistically significant. F values for comparing GS and HT were $F_{(df, 28)} = 18.287$ and 3.253, respectively.

### Notes:
- Although significantly lower than in P, calibrated amounts of HT in T, as determined by GNF-SERS, was ~1.8 pmol mm⁻² in the apparent analyte density, which was comparable to the apparent analyte density of GS (Fig. 6d). This result is inconsistent with the findings of HT imaging by vacuum-type imaging MS, which showed that HT was undetectable in T (Supplementary Fig. 10). Furthermore, unlike GS, CD44-KD in the cancer cells did not cause a decrease in HT (Fig. 6).
- These results led us to examine the possibility of an alternative pathway for providing HT, one that does not depend on the xCT/CD44 complex.
After replacement with the culture medium containing 5 mmol−1−1 12C6-glucose for 15 min, shCD44 cells showed a greater elevation of 13C3-3-phosphoglycerate (3PG) and 13C3-serine, the metabolite that provides carbons into serine/glycine cleavage systems for shuttling a one methyl group via serine and glycerate dehydrogenase (PHGDH) and phosphoserine aminotransferase-1 (PSAT1), and phosphoserine phosphatase, 5HMT: serine hydroxymethyl transferase. p < 0.05 analyzed by unpaired Student’s t-test. T values for statistical analyses are as follows: T(14) for 13C1-3-phosphoglycerate = 3.537, T(14) for hypotaurine = 7.354, T(14) for 13C1-hypotaurine = 5.622, T(14) for taurine = 2.733, and T(14) for GSH/GSSG = 2.169. #p < 0.05 with ANOVA with Fischer’s LSD test. F values of p-Ser and Ser were F (5,42) = 6.519 and 90.475, respectively.

Fig. 7 Differences in fates of 13C6-glucose challenge between shControl and shCD44 cells in culture. The cells cultured in 1% FBS containing 5 mmol l−1 12C6-glucose was removed and replaced with the same concentration of 13C6-glucose for 15 min. Metabolites except for HT and taurine were determined by CE-MS (pmol mg−1 protein derived from 13C6-glucose in shCD44 cells to a greater extent than in shControl cells. These results suggest that HT is a target supported by the one-methyl transfer system under CD44 knockdown. We also examined the effects of replacement with 5 mmol l−1 13C6-glucose as a control for 15 min (Fig. 8a). The data showed significant increases in HT and taurine in shCD44 cells as compared with shControl cells. Furthermore, amounts of HT and taurine were suppressed by 2-deoxyglucose (2-DG). The acceleration of HT synthesis stopped at 60 min after replacement with 13C6-glucose, and 2 different stable isotopic species of 13C1- and 13C2-serine are increased (Fig. 8b), suggesting that both 13C1- and 13C2-serine serve as substrates for generating taurine. At 15 min, the relative amounts of 13C1- and 13C2-labelled taurine versus non-labelled taurine were significantly lower. At 60 min the ratio was significantly reversed to show acceleration of HT oxidation to taurine during the later phase (Fig. 8c). Although the T HT signal in vivo (Fig. 6c, d) slightly increased in response to CD44 knockdown, in vivo and in vitro results were consistent: CD44 knockdown did not suppress amounts of HT in T or cultured cancer cells. The accelerated HT/taurine synthesis in shCD44 cells coincided with significant induction of phosphoglycerate dehydrogenase (PHGDH) and phosphoserine aminotransferase-1 (PSAT1), and phosphoserine phosphatase (PSPH) and suppression of glycolysis-activated pyruvate kinase M2 (p-PKM2 [S37], Fig. 8d–f). Although mechanisms remain unknown, altered expression of these enzymes by CD44 knockdown facilitates entry of glycolytic substrates into serine and glycine cleavage systems to compensate HT/taurine synthesis. The early 13C1-HT elevation followed by its disappearance and the increase in 13C1- or 13C2-labelled taurine led us to examine whether shCD44 cells survive when glucose utilization is blocked by 2-DG, a potent hexokinase (HK) inhibitor. As 2-DG increased, cell death was greater among shCD44 cells (Supplementary Information).
Fig. 8 Altered expression of enzymes in glycolysis and serine/glycine cleavage system in shCD44 cells in culture. a Effects of replacement of the culture medium with 5 mmol l⁻¹ 12C₆-glucose on contents of HT and taurine as a negative control experiments of Fig. 8. Measurements were based on LC-MS analyses. All the units of HT and taurine contents were expressed as pmol mg⁻¹ protein. Note that 10 mmol l⁻¹ 2-deoxyglucose (2-DG) significantly suppressed amounts of HT and taurine in both shControl and shCD44 cells. *p < 0.05 with ANOVA with Fischer’s LSD test. b Differences in fates of 13C₆-glucose challenge between shControl and shCD44 cells in culture at 60 min. The cells cultured in 1% FBS containing 5 mmol l⁻¹ 12C₆-glucose was removed and replaced with the same concentration of 13C₆-glucose for 60 min. Note that 13C-labelled HT disappeared at 60 min, while the 13C₁ and 13C₂-labelled taurine were elevated. c Comparison of 13C-labelled taurine versus 12C-taurine between those measured at 15 min and at 60 min. *p < 0.05 versus shControl (unpaired Student’s t-test). d Altered expression of enzymes. Note that phospho-glycerate dehydrogenase (PHGDH), phosphoserine aminotransferase-1 (PSAT1) are increased, and pPKM2 (S37) are decreased in shCD44 cells. e Relative values of densitometry in shCD44 data versus those in shControl data were analyzed by unpaired Student’s t-test. Mean ± SE of 3 separate experiments: T(4) = 3.595 for PHGDH, and T(4) = 3.370 for PSAT1, T(4) = 5.173 for PSAT1, and T(4) = 4.060 for pPKM2(S37). f Schematic diagram of metabolic systems in glycolysis, serine/glycine cleavage system, and trans-sulfuration pathway. 2-DG: 2-deoxyglucose. SSZ: sulfasalazine, a potent xCT inhibitor.

Fig. 13a). A fluoroprobe sensitive to reactive oxygen species (ROS) revealed that shCD44 cells at basal levels had significantly greater ROS than did shControl cells (Supplementary Fig. 13b). ROS levels were further enhanced by 2-DG treatment in shCD44 cells, and suppressed by exogenous HT application (Supplementary Fig. 13b). We also examined effects of sulfasalazine (SSZ), a potent xCT inhibitor, and found that shControl, but not shCD44 cells were sensitive to SSZ (Supplementary Fig. 13c). This result was consistent with the observation that CD44 knockdown destabilizes xCT expression on cancer cell membranes. Collectively with the data showing CDO expression in T and P (Supplementary Fig. 13d), these results suggested that shCD44 xenografts compensate for HT/taurine synthesis by inducing PHGDH, PSAT1 and PSPH. Thus, HT is a robust CD44-independent anti-oxidant in cancer cells (Fig. 6c, d).

Many cancer cell lines expressing CD44 stop proliferation or lose viability by stable knockdown of CD44. Among 5 different cancer cell lines, MCF7 (CD44-low breast cancer) and MDA-MB-231 (CD44-high breast cancer) expressed either PHGDH or PSAT1, and PSPH. (Supplementary Figs. 14a, c). MCF7 and MDA-MB-231 (CD44-high breast cancer) expressed comparable responsive of ROS to SSZ and 2-DG analogous to HCT116 cells (Supplementary Fig. 13). In these cells lines, 2-DG-induced ROS enhancement was attenuated by exogenously administered HT (Supplementary Fig. 14c), suggesting that HT cancels 2-DG-elicted ROS enhancement in these cells, as well.
**Robustness of T boundary extraction in shCD44-xenograft.**

Differences between shControl and shCD44 cells in CE-MS-based metabolomics (Supplementary Table 2), $^{13}$C$_2$-glucose-dependent flux analyses (Fig. 7) and GNF-SERS imaging (Fig. 6) led us to investigate whether the parent database of Raman shifts (Fig. 4 and Supplementary Table 1) collected from shControl cells was usable to identify T boundaries in shCD44-xenografted liver tissues in which metabolic profiles largely differ. Surprisingly, the accuracy rate of T boundary identification in shCD44-xenografted livers was significantly higher than those for shControl- and Lenti-control-xenografted livers. The accuracy rates of individual images as a function of T occupancy rates among the groups shows a significantly negative correlation (Fig. 9a, c). When the cross-sectional percentages of T density were less than 20% (<0.8 mm$^2$), the accuracy rates exceeded 80%. Furthermore, this method of identifying T boundaries allowed us to differentiate large V (e.g. central venules shown by ‘V’ in Fig. 9a) from metastatic T in the sections, because T, but not V, produce multiple molecular vibration modes for many metabolites, thus becoming detectable by GNF-SERS imaging. These results suggest that our method has benefits for automated detection of small metastases that are surrounded by sufficient parenchyma.

**GNF-SERS imaging in a mouse syngeneic glioblastoma model.**

Profiles of metabolites differ among varied organs. It is worth examining whether our GNF-SERS imaging enables to extract differences in T-dominant and P-dominant metabolites in a mouse syngeneic model of glioblastoma derived from genetically engineered cancer stem cells, which displayed complicated pathologic features analogous to human glioblastoma such as inflammatory responses and gliosis$^{42-44}$. We attempted to extract T borders by a professional pathologist using hematoxylin-eosin (HE)-stained tissues (Fig. 10). As seen, T regions (red in the right lower panel of Fig. 10a) seemed wider than binarized GFP-fluorescence (AF) image which turned out to display the presence of tumour after analyzing tumour boundaries. T and V indicate location of tumours and vessels (e.g. central venules) in the representative microscopic field. An asterisk indicates a venule recognized in the autofluorescence (AF) image which turned out to display the presence of tumour after analyzing tumour boundaries. T borders by a professional pathologist using hematoxylin-eosin (HE)-stained tissues (Fig. 10). As seen, T regions (red in the right lower panel of Fig. 10a) seemed wider than binarized GFP-fluorescence (AF) image which turned out to display the presence of tumour after analyzing tumour boundaries. T and V indicate location of tumours and vessels (e.g. central venules) in the representative microscopic field. An asterisk indicates a venule recognized in the autofluorescence (AF) image which turned out to display the presence of tumour after analyzing tumour boundaries. T and V indicate location of tumours and vessels (e.g. central venules) in the representative microscopic field. An asterisk indicates a venule recognized in the autofluorescence (AF) image which turned out to display the presence of tumour after analyzing tumour boundaries.

![Figure 9](image)

**Fig. 9** Robustness of automatic tumour boundary extraction in shCD44-xenografted livers. a Representative pictures indicating automatic image processing from manual binary images towards T/P binary image to calculate the accuracy rates of tumour boundary extraction in shControl- and shCD44-xenografted livers. Bar = 1 mm. The accuracy rates of the representative pictures in shControl and shCD44 images were 67.9% and 93.5%, respectively. T and V indicate location of tumours and vessels (e.g. central venules) in the representative microscopic field. An asterisk indicates a venule recognized in the autofluorescence (AF) image which turned out to display the presence of tumour after analyzing tumour boundaries. b Differences in accuracy rates among shControl, Lenti-control and shCD44 groups. Note that the accuracy rate in the shCD44 group is significantly higher than the shControl group. ANOVA with Fischer’s LSD test. *p < 0.05 versus shControl. F $(2, 16) = 4.970$. c Plotting accuracy rates of individual images as a function of tumour occupancy rates among different groups, showing a negative correlation ($Y = 0.5088 X + 94.705; R^2 = 0.664, p = 0.00002$). Brown, light blue and yellow data points indicate those from shControl, Lenti-control and shCD44 groups, respectively.
glioblastoma (Fig. 10e, f). Extraction of tumour boundaries under high magnification. spatial resolution: 25 μm. Bar = 2.0 mm. Yellow squares show the regions indicating in c, d. Representative pictures of GNF-SERS imaging showing T-dominant peaks at 485 cm⁻¹ and 726 cm⁻¹ in the same brain slices. Spatial resolution: 100 μm. Bar = 2.0 mm. e Magnified HE images and the main tumour region determined by a professional pathologist (highlighted in red). White spots: cracks and small defects in the tissue section. Bar = 0.5 mm. A magnification of tumour boundaries under high magnification. Bar = 0.5 mm. These binary images were T/P binary images (T = 1, P = 0) which were processed by the method shown in Fig. 4 and Supplementary Fig. 3. g Adenosine and inosine monophosphates (arrows) visualized by vacuum-type MALDI mass imaging (Vc-MALDI) in the serial frozen section. Bar = 2.0 mm.

Discussion

The current GNF-SERS technique provides excellent SERS excitation sources, thereby enabling large-area imaging and profiling of metabolic systems in T-bearing tissues. The robust, spatially uniform enhancement of GNF signals that covers extensive areas of tissues exceeds the performance characteristics of previously reported SERS substrates. The present technique covers the mesoscopic areas between the macroscopic and cellular levels of bioimaging and improves our understanding of the metabolic interplays between micrometastases and host tissues. This method is a breakthrough in automated processes for collecting SERS spectroscopic data and identifies metabolic differences between T and their surrounding tissues. Mass spectrometry and conventional Raman spectrometry face challenges in identifying cancer in biopsy or intraoperative specimens. However, the current study first to suggest that global characterization and imaging of GNF-SERS signals, which contain numerous molecular vibration modes of metabolites, might be useful for
automated identification of T boundaries in tissues. Another breaking-through of highly sensitive GNF-SERS is the identification of several metabolites such as GS at 298 cm$^{-1}$, HT at 978 cm$^{-1}$ and retinol and/or retinol palmitate at 1150 cm$^{-1}$ as a biomarker signature for differentiating T regions from surrounding tissues in a mouse model of liver metastases of human-derived colon cancer xenografts. GNF-SERS imaging in the syngeneic mouse model of glioblastoma raised the possibility that polysulfides and metabolites with adenine ring constitute T-dominant compounds which yields the peaks at around 480 cm$^{-1}$ and 726 cm$^{-1}$, respectively. Although further investigation is necessary to determine the exact molecular entities of polysulfides in T-bearing livers$^{32}$ and brains$^{34}$, the current results suggest that GNF-SERS serves as a method to visualize endogenous polysulfides in vivo.

Light-sensitive metabolites$^{26,27,55}$, or those with ROS-sensitivity (e.g. GS, HT and polysulfides) are difficult to investigate, as they are labile biomarker molecules prone to artificial oxidation by technologies that use high-energy laser and electric fields (e.g. MALDI and CE-MS). Studies using MS technologies might therefore overlook the possibility that the metabolic flux to generate HT is present, but undetectable by both AP-type or vacuum-type MALDI-MS imaging because of its instability against laser sensitivity or rapid consumption of HT in cancer cells. Use of GNF-SERS under infra-red laser excitation prevents against laser sensitivity or rapid consumption of HT in cancer vacuum-type MALDI-MS imaging because of its instability.

Methods

Fabrication of the GNF substrates. A 24 × 24 × 0.5 mm$^3$ glass plate was washed with surface active agents (W304/LB93, ADEKA Corp.) to remove organic contaminants, and rinsed in ultrapure water using an ultrasonic bath. After drying the plate in spin drier, aluminum (Al) was deposited on the plate at 50 nm thickness using a reactive DC magnetron sputtering system (SPF-5300H, ANELVA) at a deposition rate of 5 Å s$^{-1}$ (Supplementary Fig. 1a). The Al film was subsequently boiled for 15 min in hot water (100°C) to form boehmite (Al(OH)OH), and was dried with N$_2$ gas according to our previous method$^{15}$ (Supplementary Fig. 1b). Au was deposited on the boehmite nanostructures by electron-beam evaporation system (EBX-6C, ULVAC) with different deposition angles of 0°, 45° and 80° (Supplementary Fig. 1c). As shown in Fig. 1b, c, we determined that 80° in the deposition angle and 110 nm in Au thickness, which was evaluated by using R6G and our custom-made inverted Raman microscopic system with ×20 (NA = 0.4) objective lens, gave optimal conditions such that SERS signal can be maximally enhanced. This microscope system was equipped with a fiber-coupled spectrograph (Ocean Optics, QE65000), and with an xy-scanning stage with highest resolution of 250 nm, and with a 785-nm diode laser as described previously$^{15}$. The sensitivity and frequency of the microscope system were calibrated by the 520-cm$^{-1}$ Raman shift of silicon phonon mode before SERS measurements. The back-ground noise was subtracted from the recorded spectra by using the weighted mean fitting with L-cosmos second function. All processes were conducted in a cleanroom. As described in the text, the coefficient of variation of the optimized GNF substrate in the uniformity was 11.0% ± 0.5%, and that in the lot-to-lot reproducibility was 2.1%.

Morphological analyses of GNF substrates. To determine periodicity of the boehmite nanostructure (Fig. 2a), binary maps were created from top-view SEM images. Sizes of the remaining particles were measured using the “analyze particles” function of ImageJ (Fig. 2). To evaluate lot-to-lot reproducibility of distribution and anisotropy of individual GNF units, measurements of lengths and widths of the Au nanoparticles were plotted as seen in Fig. 2b; results indicated that the L/W values of a major population of Au particles distributed in a range between 1.0 and 5.9 in 3 different lots.

Numerical calculations of SERS enhancement factor using FDTD. As described elsewhere$^{18,19}$, the local field enhancement (G) is represented by the ratio of the local field intensity ($|E|^2$) against the incident wave field intensity ($|E_0|^2$) at the wavelength ($\lambda$) of the excitation:

$$ G = |E(\lambda)|/|E_0(\lambda)|^2 $$

In the current study, a custom-made three-dimensional FDTD software was used to estimate these enhancement factors. Conditions of the simulation model to calculate G values were described as follows (Fig. 2c and d): the lengths and widths of the individual Au nanoparticles were variable in a range between 0 and 400 nm, and the thicknesses were 40 nm. A single Au nanoparticle was placed in air with a refractive index of 1.0. In the case of the local field enhancement, the particle was irradiated by the parallel-polarized plane-wave light at 785 nm. We calculated the
localized G values at the apex position of the Au nanoparticle with a 1.0-nm distance from the Au surface, and the values were plotted as functions of lengths and widths of the particles (Fig. 2c). All these simulations were carried out with a grid size of 1 nm. The refractive index of Au at 785 nm was 0.177 + 4.54 \text{i}. 

Human-derived cancer xenografts in immunodeficient mice. Animal experiments were carried out in accordance with the guidelines of Experimental Animal Committee of Keio University School of Medicine. Human colon cancer HCT116 cells were transfected stably with non-target control shRNA or with shCD4411,12,13,14. The cells were injected into the spleen of super-immunodeficient NOG (NOD/SCID/IL-2Rγ−/−) mice as described previously15–18. In brief, HCT116 cells were injected into the spleen of male NOG mice aged at 11–14 weeks at 1 × 10⁶ cells/mouse. Two weeks after transplantation, liver lobes of the mice fasted for 17 h were excised under sevoflurane anesthesia and snap-frozen with liquid nitrogen. To examine T growth in vivo, 5-μm-thick cryo-sections were stained with hematoxylin and eosin (H&E) and examined under the xenograft transplantation were prepared for the control liver slices.

GNF-SERS imaging of metabolites in vivo and in vitro. To visualize metabolites in the frozen tissue sections, SERS imaging was carried out by using a GNF substrate, which is SERS-active substrate and above mentioned custom-made inverted Raman spectroscopic system19. To visualize metabolites on the tissues, cryo-sections with 5-μm thickness of the frozen livers were thaw-mounted on a GNF plate and dried in a dry chamber for 20 min. After capturing the images of the liver sections by an optical microscope (Nikon Eclipse E600: Nikon Corp., Tokyo, Japan) to select 2 mm × 2 mm square regions as regions of interests, the plate was set on the Raman microscope system. P were characterized by regions with positive autofluorescence, while T and large V are those indicating no auto-fluorescence. Since livers (e.g. central venules) were identified by densities of cellular components (e.g. blood cells) in magnified transparent images, three different regions such as T, P and V were manually extracted to form a binary image (Fig. 3a) according to our previous method20. To capture autofluorescence images, a narrow-band filter (Olympus U-FFG, excitation filter 460-480 nm, barrier filter 495-545 nm) was applied. Negative autofluorescence images were photographed with an Olympus DP73 camera. These sources of images were used to execute automatic T boundaries, as well as to identify T-specific and P-specific regions for visualizing and quantifying GNF-SERS signals that were shown in Fig. 3c, d and Fig. 6c, d.

For imaging, we measured 2 mm × 2 mm square areas on liver tissues as regions of interests by using ×60 (NA = 0.7) objective lenses. To compose two-dimensional SERS maps, spectra with the range of Raman shifts between 200 and 1730 cm⁻¹ were measured at 51 × 51 spots over a tissue covering the region of interests with a 40-μm-translation stage. Acquisition time was 10 seconds for single spot. The diameter of beam spot was ~2 μm with ~1 mW power at a 785-nm laser. To achieve a quality of SERS images with greater signal-noise ratios, we processed SERS images through a radius of 1 pixel, resulting in sharply contrasted T boundaries. To compose two-dimensional automatic extraction of the T boundaries in the frozen liver sections. To this end, we prepared two groups of xenograft transplantation model using different clones of human colon cancer HCT116 cells: the first was shControl (n = 7), which were used to establish the accumulative list of the Raman shifts (ALR in Fig. 4) by using the data collected from the shControl group.

On the other hand, liver tissues metastasized with the xenografts derived from the second HCT116 clone named Lenti-control (n = 5) were used to examine whether lists of T-enriched and P-enriched metabolic fingerprints that were collected from the shControl group are suitable to extract precise boundaries between T and P in tissue sections of interests which were collected from the xenograft with Lenti-control. To that effect, spectral data of Raman shifts in the Lenti-control group were automatically sorted into T- and P-dominant shifts based on ALR (Supplementary Fig. 3b). All T- and P-dominant Raman shifts were averaged separately to build T/P ratio images that featured T regions in tissue sections. The T/P ratio images collected from the Lenti-control group were smoothed by using 2 different radiiuses of median filters; one was to process images through a radius of 1 pixel, resulting in sharply contrasted T boundaries with high noise levels. On the other hand, another was to process them through a radius of 2 pixels, in which smooth and noiseless images were obtained, but with broadened boundaries between T and P. Using these image sources, we binarized these 1 pixel- and 2 pixel-sized smoothed images (0 in P and 1 in T regions) according to the previously reported study using triangle auto-thresholding method21, and multiplied the two binarized images in which signals out of T regions became 0 (flowchart in Supplementary Fig. 3c). We also analyzed GNF-SERS data from the shControl (n = 7), which were used to examine whether lists of T-enriched and P-enriched metabolic fingerprints that were collected from the xenografted liver tissues (n = 7) according to the flowchart (Supplementary Figs. 3b and c) to examine whether T derived from shCD44 cells metastasized in livers that are expected to change their metabolic profiles can be detected by using ALR database established in Supplementary Fig. 3a.

MALDI imaging mass spectrometry (MALDI-IMS). In separate sets of experiments, we conducted atmospheric pressure or vacuum-type MALDI imaging according to our previous methods22,23. Details of methods were described in the text of Supplementary Note 1.

Quantum chemical calculation to identify molecular vibration. To identify molecules responsible for generating SERS signals with specific wave numbers, quantum chemical calculation was carried out according to previous studies24–26. The calculation was performed by Gaussian 09 Program package. The Raman
intensities were calculated using density functional theory (DFT) by using basis sets controlled glycolysis, serine/glycine cleavage systems and trans-sulfuration pathway. Details of the blotting methods and the methods to determine cell viability and oxidative stress were described in the text of Supplementary Note 1. Uncropped blotting data were shown in Supplementary Fig. 16. All cell lines were authenticated by STR profiling (Takara Bio, Otsu, Japan). Mycoplasma contaminations were tested in the cell lines using commercially available kit (MycopAlert Mycoplasma Detection Kit, Lonza).

**Chasing fates of 13C6-glucose in cultured HCT116 cells.** We conducted CE-MS and LC-MS analyses to determine differences of 13C-labeled metabolites that belong to glycolysis, glycerol cleavage pathway and remethylation and trans-sulfuration pathways between shControl and shCD44 cells in culture. In response to replace treatment with the culture medium containing 5 mmol L−1 13C6-glucose for 15 min and for 60 min, 13C-labeled metabolites were generated in both cell lines (Fig. 8). At 15 min after exposure to 13C6-glucose in the medium, 13C5-phospho-serine derived from 13C5-3PG provided 13C4-glycerol and 13C2-serine transfer towards tetrahydrofolate (THF). On the other hand, 13C6 of glycine is cleaved and transferred to tetrahydrofolate (THF). As a result, a series of these reactions generates 13C4-methionine and 13C5-serine through the robust catalytic activity of serine hydroxymethyl transferases (SHMTs) (Figs. 7 and 8). In these experiments, FT and taurine were determined by LC-MS (LCMS-8030plus triple quadrupole mass spectrometer, Shimadzu, Kyoto, Japan)26,69.

**Orthotopic implantation of neural stem cells to mimic glioblastoma.** All experiments described below were performed in accordance with the animal care guidelines of Keio University. Neural Stem/Progenitor Cell Culture was established as described previously42,43. Briefly, 6-week-old male Ink4a/Arf null C57BL/6 mice (B6.129-Cdkerd/6Rdp, National Cancer Institute, Frederick, MD) were euthanized with a lethal dose of pentobarbital to extract brains and to isolate samples from their subventricular zone (SVZ). The SVZ fragments were microdissected, washed, and then mechanically dissociated. Primary NSC/NPCs were maintained as sphere culture in Dulbecco modified eagle medium (DMEM)/F12 (Sigma, St Louis, MO) supplemented with 20 ng ml−1 epidermal growth factor (EGF; PeproTech, Rocky Hill, NJ), 20 ng ml−1 basic fibroblast growth factor (PeproTech), B27 supplement without vitamin A (Invitrogen, Carlsbad, CA), 200 μg ml−1 heparin sulfate, with penicillin and streptomycin at 37°C in 5% CO2/95% humidified air. The retroviral vector construct was prepared as follows. Human H-RasV12 DNA44 (kindly provided by P. Pandolfi) was cloned into the retrovector pMXs-IG (kindly provided by T. Kitamura) and the resulting construct was transfected into Flp-E packaging cells45 using FugenHD (Roche Diagnostics, Mannheim, Germany). Medium was replaced once after 24 h. Viral supernatants were collected and filtered with 0.45-μm cellulose acetate filters (Iwaki, Kyoto, Japan) 48 h after transfection. Supernatants were centrifuged at 12,000 rpm for 6 h at 4°C, and the viral pellet was resuspended in small volumes of NSC culture medium. To establish brain T-initiating cells, primary Ink4a/Arf null NSC/NPCs were infected with retroviral supernatants. The resulting GFP-positive cells, termed RasG-NSCs hereafter, were sorted by flow cytometry classified in NSC culture medium and used for implantation within 20 passages after establishment.

To carry out orthotopic transplants, female C57BL/6 mice aged 6 to 8 weeks were anesthetized and placed into a stereotactic apparatus assisted by a 2-axis (David Kopf Instruments, CA). A small craniotomy was made in the skull 2.0-mm lateral to the bregma using a dental drill. One hundred thousand viable cells in 2 μl of Hank’s balanced salt solution was injected into the right hemisphere 3 mm below the surface of the brain using a 10-μl Hamilton syringe with a customized 26-gauge needle. The injection was performed over 2 min, with an additional 2-minute pause before removing the syringe. The hole was sealed with bone wax, and the scalp was closed using a cottonoid applicator85. None of the animals developed neurological deficits during the follow-up period. At 15 days after the transplantation, brains were extracted to prepare frozen tissue slices that include both T and the surrounding brain tissues. Slices were prepared from the frozen blocks of T-bearing brains collected from individual mice. Using the coronal tissue slices, both SERS imaging and hematoxylin-eosin staining were carried out. All images were digitally stored in an open software (Nanoozer Digital Pathology, Hamamatsu Photonics, Hamamatsu City). We examined whether our GNF-SERS imaging enables to extract differences in profiles of T-dominant and P-dominant metabolites in a mouse syngeneic model of glioblastoma derived from genetically engineered cancer stem cells46−48. Since this glioblastoma model displayed complicated pathological features including gliosis and inflammatory responses and around main T, we extracted T borders using manual examination by a professional pathologist, who examined haematoxylin-eosin (HE)-staining images (T-region; the red regions in the right lower panel of Fig. 10a, c) in tissue sections through high magnification images provided by Nanoozer digital pathology software. In these analyses, the regions containing accumulated inflammatory cells were excluded from T-regions. Since the contralateral region was chosen as the P-region, we chose the contralateral striatum regions as the control P (P-regions; the dark blue region in the left hemisphere in the right lower panel of Fig. 10a). Control regions in P of the contralateral striatum (P-region) of these brain slices were defined as T-free regions in the contralateral hemisphere. Selected regions (T and P) were spectrally imaged on SERS imaging data, which were visualized according to the procedures used for the xenograft-implanted model of HCT116 cells. In these experiments, GNF-SERS images were collected under two different spatial resolutions and oxidation images provided with a spatial resolution of 0.1 μm that covered the whole coronal plane of brain slices (Fig. 10b), and high-magnification images with a spatial resolution of 25 μm that covered T boundaries that often coincided with gliosis and/or inflammatory responses (HE staining in Fig. 10c).

Separately, we determined tissue contents of H2S/polysulfides to compare them between T-free and T-bearing brain tissues by using the monobromo-bimane-derivatization method86,87. Separately, we examined whether recombinant GFP protein yield GNF-SERS signals. The dyeing protocols were conducted to determine whether varied concentrations of recombinant GFP (abcam, ab84191) dissolved in 0.1 mol L−1 phosphate buffer at pH 7.4 yielded GNF-SERS signals in vitro. To determine appropriate concentrations of recombinant protein contents and mouse brain glialblasta was treated with the in-situ freezing procedure, and the T-enriched region was sampled by cutting with a surgical knife. The fluorescence intensities of recombinant GFP solutions were measured with SpectraMAX M2 microplate reader (excitation/emission = 488 nm/530 nm). To determine the approximate amount of GFP in T tissues, we sacrificed a T-bearing mouse under anesthesia, and the amount of protein versus T tissue weight was 0.18 μg μl−1. As GFP turned out to constitute <0.1% of the total protein amounts, we prepared recombinant GFP solution at a concentration of 0.18 μg μl−1, and an aliquot of 100 μl was placed on the GFP substrate to be dried up. Under these conditions, the density of GFP became sufficient enough to surpass that of GFP in vivo. GNF-SERS signals yielded from the region of 24 × 24 mm2 were determined by the current Raman spectroscopy. As seen in the bottom of Supplementary Fig. 4a, GFP did not yield any notable GNF-SERS signals.

**Statistical analyses.** All quantitative results are presented as the mean ± SE of independent experiments. The sample sizes of each experiment were determined considering the institutional guidelines to minimize unnecessarily killed animals. In all experiments, all mice used for experiments displayed T formation in situ: there were no mice which did not display T formation with no excluded animals from the group. Neither randomization nor blinding procedure was performed in the current animal study.

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**References**

1. Raman, C. V. & Krishnan, K. S. A. A new type of secondary radiation. *Nature* **121**, 501–502 (1938).

2. Laing, S., Jamieson, L. E., Faulks, K. & Graham, D. Surface-enhanced Raman spectroscopy for in vivo biosensing. *Nat. Rev. Chem.* **1**, 1–19 (2017).

3. Stone, N., Kendall, C., Smith, J., Crow, P. & Barr, H. Raman spectroscopy for identification of epithelial cancers. *Fairydare Discussions.* **126**, 141–157 (2004).

4. Nie, S. & Emory, S. R. Probing single molecules and single nanoparticles by near-field optical trigonometry. *Phys. Today* **50** (1997).

5. Knuepp, K. Surface-enhanced Raman scattering. *Phys. Today* **60**, 40–46 (2007).

6. Barhoumi, A., Zhang, D., Tam, F. & Halas, N. J. Near-field spectroscopy of DNA. *J. Am. Chem. Soc.* **130**, 5523–5529 (2008).

7. Iike, S. & Hirose, T. Single-molecule detection of DNA by SERS. *Fairydare Discussions.* **126**, 141–157 (2004).

8. Nie, S. & Emory, S. R. Probing single molecules and single nanoparticles by near-field optical trigonometry. *Phys. Today* **50** (1997).

9. Knuepp, K. Surface-enhanced Raman scattering. *Phys. Today* **60**, 40–46 (2007).

10. Barhoumi, A., Zhang, D., Tam, F. & Halas, N. J. Near-field spectroscopy of DNA. *J. Am. Chem. Soc.* **130**, 5523–5529 (2008).

11. Iike, S. & Hirose, T. Single-molecule detection of DNA by SERS. *Fairydare Discussions.* **126**, 141–157 (2004).

12. Nie, S. & Emory, S. R. Probing single molecules and single nanoparticles by near-field optical trigonometry. *Phys. Today* **50** (1997).

13. Knuepp, K. Surface-enhanced Raman scattering. *Phys. Today* **60**, 40–46 (2007).

14. Barhoumi, A., Zhang, D., Tam, F. & Halas, N. J. Near-field spectroscopy of DNA. *J. Am. Chem. Soc.* **130**, 5523–5529 (2008).
66. Kubo, A. et al. Semi-quantitative analyses of metabolic systems of human colon cancer metastatic xenografts in livers of superimmunodeficient NOG mice. Anal. Bioanal. Chem. 400, 1895–1904 (2011).

67. Hohenberg, P. Inhomogenous electron gas. 136. Phys. Rev. 136, B864–B871 (1964).

68. Nakano, S. et al. Hyperhomocysteinemia abrogates fasting-induced cardioprotection against ischemia/reperfusion by limiting bioavailability of hydrogen sulfide anions. J. Mol. Med. (Berl.) 93, 879–889 (2015).

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Author contributions
M.Sh. and M.Su. conceived and designed the whole study, and carried out data analyses and manuscript preparation. M.N. and S.Y. developed GNC and GNF substrates, respectively. M.Sh. and D.K. checked quality and plurality standards. T.T. examined simulation of electromagnetic fields based on morphological features of GNF substrates. H.T. contributed to morphological analyses of GNF substrates through scanning electron microscopy. T.H. and A.K. contributed to identification and exclusion of many metabolites in the liver that can be visualized in GNF substrates. T.Y. and M.I. contributed to exploration of metabolic pathways for HT and taurine and carried out cell biology experiments. M.O. and Y.S. and Y.K. designed and carried out experiments of a xenograft model of human colon cancer cell lines, and analyzed tissue sections for imaging MS. S.K., H.S. and O.N. supervised discussion on roles of CD44 in cancer survival and provided shCD44 HCT116 cells. H.N. and O.S. and T.M. performed experiments of brain tumour-initiating cells which were established by overexpressing H-RasV12 in normal neural stem/progenitor cells isolated from the SVZ of adult mice harbouring a homozygous deletion of the Ink4a/Arf locus. K.T. contributed to preparation of the manuscript. N.H. is a licensed professional clinical pathologist who examined T-bearing brain tissues. H.W. examined simulation analyses for molecular vibration of HT and glutathione.

Additional information
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