Morphological and molecular characterization of *Aporcella femina* sp. n. (Dorylaimida, Aporcelaimidae) from Nigeria

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Abstract

A new species of the genus *Aporcella* collected from a watermelon field in Nigeria is described, including its morphological and molecular (small subunit (SSU) and large subunit (LSU) ribosomal DNA (rDNA)) characterization. *Aporcella femina* sp. n. is distinguished by its 3.21–3.64 mm-long body, inner cuticle layer with fine but distinct transverse striation, lip region offset by deep constriction, 22–25 μm broad, odontostyle 20–26 μm, neck 661–811 μm long, pharyngeal expansion occupying 52–56% of the total neck length, female genital system didelphic–amphidelphic, uterus 191–350 μm or 1.9–3.3 mid-body diameters long, V = 52–57, tail short and convex conoid (35–48 μm, c = 72–98, c′ = 0.7–0.9) and males absent. Phylogenetic analyses based on the partial sequence of SSU and LSU (D2–D3) rDNA revealed a close relationship of *A. femina* sp. n. with other *Aporcella* species, confirming the monophyly of the genus as well as its association to a clade made of several taxa characterized by the absence of paras refringens vaginae.

Introduction

The genus *Aporcella* Andrássy, 2002 is a widely distributed dorylaimid (Dorylaimida) taxon, inhabiting soils of all zoogeographical realms/regions except Antarctica (Álvarez-Ortega et al., 2013; Vazifeh et al., 2020). In Africa, it occurs in the southern part of the continent (Botswana and South Africa), where four species have so far been recorded: *Aporcella adriaani* (Botha & Heyns, 1990) Álvarez-Ortega, Subbotin & Peña-Santiago, 2013; *Aporcella debruijnae* Álvarez-Ortega, Subbotin & Peña-Santiago, 2013; *Aporcella parapapillata* (Botha & Heyns, 1990) Andrássy, 2009; and *Aporcella tropica* (Jana & Baqri, 1981) Álvarez-Ortega, Subbotin & Peña-Santiago, 2013.

Since its original description by Andrássy (2002), the taxonomy of *Aporcella* has been subjected to a number of changes, including a new definition (Álvarez-Ortega et al., 2013), the description of new species (Andrássy, 2012; Álvarez-Ortega & Peña-Santiago, 2016; Naghavi et al., 2019, 2020) and a recent update of its taxonomy (Vazifeh et al., 2020). Molecular ana-
lyses have repeatedly supported the monophyly of *Aporcella*. Surprisingly, the closer evolution-
ary relationship of *Aporcella* with non-aporcelaimid taxa (tylencholaims, discolaims) than with other aporcelaims has also been reported by various researchers (Álvarez-Ortega & Peña-Santiago, 2016; Imran et al., 2019; Naghavi et al., 2019; Vazifeh et al., 2020).

From a 2016 survey of nematode fauna associated with watermelon in Nigeria, specimens of an *Aporcella* population that belong to an undescribed species were recovered. This would represent a novel geographical record for the genus, and, by means of its molecular analysis, offers the chance to confirm previous findings about its monophyly and phylogeny.

Materials and methods

**Extraction and processing of nematodes**

Nematode specimens were recovered from rhizosphere soil samples that were collected from watermelon fields in south-west Nigeria during 2016. Extraction of nematodes from soil followed a modified pie pan method (Coyne et al., 2007). Nematodes were fixed in a hot 4% formaldehyde solution for morphological observations (Nico et al., 2002) and subsequently mounted in anhidrous glycerine as permanent slides (De Grisse, 1963), while others were stored in dimethyl sulphoxide, disodiuo ethylenediamine tetra-acetic acid and saturated sodium chloride (abbreviated here as DESS) solution for molecular analysis. Morphological features of nematodes were observed, measured and photographed using an Eclipse 80i
microscope (Nikon, Tokyo, Japan) equipped with differential interference contrast optics, a drawing tube (camera lucida) and a DS digital camera (Nikon, Tokyo, Japan).

**Molecular identification**

Fixed specimens in DESS were rinsed using double-distilled water, and two specimens were transferred to individual 1.5 ml tubes containing 15 μl nuclease-free water. DNA were extracted using chelex-100 by adding 20 μl of chelex (5% w/v) and 5 μl proteinase K to each tube, followed by incubation at 57°C for 120 min and at 95°C for 10 min (Rashidifard et al., 2019).

DNA amplification was carried out on a Vacutec thermocycler (www.vacutec.co.za) using the primers: small subunit (SSU) F04 (5′-GCTTGTCCTCAAAGATTAAAGC-3′), and SSU R26 (5′-CATTCTTGAAATGCTTTGC-3′) (Blaxter et al., 1998) for SSU ribosomal DNA (rDNA), as well as D2A (5′-ACAAGTAGGACCAAGATTG-3′) and D3B (5′-TCGGAGGAAAGTTG-3′) (Blastner et al., 1998) for D2-D3 large sub-unit (LSU) rDNA. The polymerase chain reaction (PCR) amplification process was followed as reported by Rashidifard et al. (2020). Four microliters of PCR products were loaded on 1% agarose gel and visualized by an ultraviolet transilluminator to check the quality of DNA. The DNA was sequenced in both forward and reverse directions by the Inqaba Biotec™ genomic company (South Africa; www.inqaba-southafrica.co.za).

**Phylogenetic analyses**

The new SSU and LSU sequences obtained for the new species were compared to other available sequences in the GenBank using the BLASTN search tool. One data set was prepared for

Fig. 1. Aporcella femina sp. n. (female): (a, b) anterior region in lateral median view; (c) anterior region in lateral surface view; (d) vagina region; (e) neck region; (f) posterior genital branch; (g) pharyngo-intestinal junction; (h) posterior body region; (i) caudal region. Scale bars: (a, g, i) 20 μm; (b–d) 10 μm; (e, f) 100 μm; (h) 50 μm.
each of the SSU and LSU regions and taxa selection was carried out according to Rashidifard et al. (2020). The available sequences as well as outgroups in the SSU data set were aligned using the MUSCLE alignment tool (Edgar, 2004) in Geneious Prime® 2020.2.3 (www.geneious.com), while the LSU data set was aligned using the E-INS-I algorithm in the MAFF alignment tool (Katoh & Standley, 2013). According to the jModelTest 2.1.10 (Darriba et al., 2012), the Hasegawa–Kishino–Yano with a gamma distribution (HKY + G) was the best substitution model for the SSU data-set. However, the General Time Reversible with proportion of invariable sites and a gamma distribution (GTR + I + G) was selected as the most appropriate model for the LSU dataset.

Bayesian inference was conducted in MrBayes v3.1.2 (Ronquist & Huelsenbeck, 2003) implemented in Geneious Prime® 2020.2.3 (www.geneious.com). The chain was running 3 × 10⁶ generations for both the SSU and LSU data sets, and after discarding the 25% burn-in samples, the remaining samples were used for further analyses. The Markov chain Monte Carlo (MCMC) algorithm was implemented to calculate the posterior probabilities (PB) for each of the phylogenetic trees (Larget & Simon, 1999).
using the 50% majority rule. The mononchid taxa were used as outgroups for both the SSU and LSU phylogenies.

Results

**Aporcella femina** sp. n.

**Material examined**

Ten females in variable but generally good state of preservation (figs 1–3).

**Measurements**

All measurements are presented in table 1.

**Description**

Female. Moderately to slender (a = 28–38) nematodes of large size, 3.21–3.64 mm long. Body cylindrical, tapering towards both ends but substantially towards the anterior region, while the tail is short and rounded. Upon fixation, habitus regularly curved ventrad, C- to G-shaped. Cuticle two-layered, very thick throughout the entire body, 4–7 μm in anterior region, 7.5–9.5 μm at mid-body and 7.5–11.5 μm on tail, consisting of a thin outer layer with nearly smooth surface, and a thicker inner layer that displays a fine but distinct transverse striation (fig. 2g). Lateral chords narrow, 6–17 μm wide or 6–19% of mid-body diameter. Ventral and dorsal pores restricted to cervical region, two or three in number at each side at odontostyle–odontophore contact.

**Fig. 3.** Bayesian tree with 50% majority rule of *Aporcella femina* n. sp. from Nigeria using partial SSU rDNA sequences under the HKY + G model. The sequence of new species is indicated by bold font. Scale bar shows the number of substitution per site.
level; lateral pores small. Lip region offset by a distinct constriction, 3.3–3.8 times wider than high and less than one-third (22–29%) of body diameter at neck base; lips separate, with moderately protruding papillae. Amphidal fovea stirrup-shaped, its opening 9–10.5 μm, less than one-half (38–46%) of lip region diameter. Cheilostom broad, 10.5–14 μm long, with thick walls. Odontostyle length 5.8 μm, occupying 4.5–5.1 body diameters long, often appearing as a simple, tube-like structure, but with some (well-preserved) specimens having a slender distal portion made of prismatic cells and a distinct sphincter. Anterior uterus 191 μm long or 1.4–1.8 body diameters long, both consisting of a slender distal portion made of prismatic cells and a distinct pars dilatata with perceptible lumen. Oviduct and uterus separated by a distinct sphincter. Anterior uterus 191–325 μm or 1.9–3.1 body diameters long, posterior uterus 204–350 μm or 2.1–3.3 body diameters long, often appearing as a simple, tube-like structure, but with some (well-preserved) specimens having a longer uterus, which enables the appreciation of a wider proximal uterine region and a more slender distal region, both of about similar length. No uterine egg observed. Vagina extending inwards 40–54 μm, occupying c. one-half (41–45%) of body diameter: pars proximalis 28–41 × 19–28 μm, with distally convergent walls and surrounded by a relatively weak musculature; pars proximalis from Nigeria and two other African species of the genus.

Table 1. Morphometrics of Aporcella femina sp. n. from Nigeria and two other African species of the genus.

| Characteristics | Species | A. femina sp. n. | A. pseudospiralis | A. parapapillata |
|-----------------|---------|-----------------|-----------------|-----------------|
|                 | Population | Nigeria | South Africa | South Africa | Botswana |
|                 | Reference | Present paper | Type material | Type material | Type material |
| Holotype | Ψ | 3.50 | 3.47 ± 0.14 | 3.64–4.42 | 2.11–2.51 |
| Paratypes | Ψ | 38 | 33.1 ± 2.9 | 31–35 | 29–33 |
|           | 68 | 4.8 | 4.7 ± 0.4 | 4.5–5.1 | 3.9–4.4 |
| c | 10 | 74 | 84.3 ± 8.2 | 58–73 | 62–75 |
| V | 94 | 54 | 54.5 ± 1.6 | 50–53 | 52–56 |
| c’ | 0.67 | 0.76 | 0.73 ± 0.12 | 0.93–1.15 | 0.8–1.0 |
| Lip region diameter | 23 | 24.0 ± 1.1 | 23–27 | 19–20 | 20–23 |
| Odontostyle length |   |   |   |   |   |
| Ventral side | 20 | 21.9 ± 1.9 | 19–24 | ? | ? |
| Dorsal side | 22 | 23.7 ± 1.9 | 20–26 | 23–26 | 18–19 |
| Odontophore length | 49 | 47.9 ± 1.2 | 46–50 | 47–52 | 32 |
| Neck length | 721 | 736 ± 53 (661–811) | 810–880 | 530–590 | 510–650 |
| Pharyngeal expansion length | 379 | 397 ± 34 (349–451) | ? | ? | ? |
| Body diameter |   |   |   |   |   |
| at neck base | 98 | 97.0 ± 7.9 (87–113) | ? | ? | ? |
| mid-body | 111 | 106 ± 12 (89–129) | 117–130 | 65–84 | 69–89b |
| anus | 62 | 57.0 ± 4.8 (50–63) | 57–61 | 27–39 | ? |
| Distance vulva–anterior end | 1901 | 1890 ± 76 (1780–1975) | 1910–2270 | 1360–1730 | 1362–1509b |
| Prerectum length | 196 | 153 ± 41 (88–203) | 219–280 | 136–173 | 88–244 |
| Rectum length | 63 | 63.8 ± 6.7 (50–75) | 65–85 | 46–48 | 50–58 |
| Tail length | 47 | 41.4 ± 4.0 (35–48) | 57–68 | 32–36 | 27–41 |

Measurements in μm except l in mm, and in the form average ± standard deviation (range).

a1: Botha & Heyns (1990); 2: De Bruin & Heyns (1992).

Calculated from other morphometrics.
distalis well-developed, 10–14 μm long; pars refringens absent. Vulva a slightly posterior transverse slit. Prerectum 2.4–4.0, rectum 0.8–1.5 times the anal body diameter long. Tail short, convex conoid, its ventral side weakly straighter than the dorsal side; caudal pores two pairs, close together, at the posterior half of tail, one lateral, another sublateral.

**Male.** Unknown. Females do not contain sperm cells.

**Type locality and habitat**
South-west Nigeria, Odeda (7°10′59.999″N, 3°26′42.01″E), an agrarian locality with a history of cassava (*Mannihot esculenta* Crantz) and vegetable production; specimens were collected from a field cultivated with watermelon (*Citrullus lanatus*) on sandy-loam soil (sand = 66%, silt = 13%, clay = 21%, organic matter = 11.18%, pH 6.22).

**Molecular characterization**
After sequencing and editing, two totally (100%) identical sequences per each locus (SSU and LSU) were obtained; therefore, only one sequence was used for analysis and deposited into the NCBI GenBank: a 745 bp-long LSU sequence (accession number MW237838) and another 855 bp-long SSU sequence (accession number MW237839).
number MW237832). The evolutionary relationships of the new species are presented in figs 3 and 4.

Type material
Female holotype and nine female paratypes, deposited in the nematode collection of the University of Jaén, Spain.

Etymology
The specific epithet is a Latin term meaning woman or female, and refers to the existence of only females in this species.

Diagnosis and relationships
The new species is characterized by its 3.21–3.64 mm-long body, inner cuticle layer with fine but distinct transverse striation, lip region offset by deep constriction and 22–25 μm broad, odontostyle 20–26 μm at its dorsal side and 19–24 μm at its ventral side, neck 661–811 μm long, pharyngeal expansion 349–451 μm long or 52–56% of total neck length, female genital system didelphic–amphidetic, uteri 191–350 μm or 1.9–3.3 body diameters long, V = 52–57, tail short and convex conoid (35–48 μm, c = 72–98, c′ = 0.7–0.9) and males absent.

In having a comparatively long body (more than 3.0 mm long), Aporcella femina sp. n. resembles Aporcella magna Andrássy, 2012 and Aporcella pseudospiralis (Botha & Heyns, 1990) Andrássy, 2012, and significantly differs from its other congeners (body 3.21–3.64 vs up to 3.08 mm long) (see compendium by Álvarez-Ortega et al., 2013 and Vazifeh et al., 2020). It can be distinguished from A. magna, a species occurring in Chile, in its smaller general size: body 3.21–3.64 versus 4.48–5.00 mm long, n = 5; neck 661–811 versus 888–1020 μm long; tail 35–48 versus 50–60 μm. It is also smaller than A. pseudospiralis, which is only known to occur in South Africa, with a smaller general size (body 3.21–3.64 vs 3.64–4.42 mm long, n = 4; neck 661–811 vs 810–880 μm long), shorter prerectum (122–203 vs 219–280 μm) and female tail (35–48 vs 57–68 μm, c = 72–98 vs 58–73, c′ = 0.61–0.94 vs 0.93–1.15) and male absent (vs present).

Among the small-sized Aporcella species (body up to 3.08 mm long), and due to the comparatively long odontostyle (>16 μm), short (c′ > 1.1) and convex conoid tail, it is similar to Aporcella papillata (Bastian, 1865) Álvarez-Ortega, Subbotin & Peña-Santiago, 2013 and A. parapapillata (Botha & Heyns, 1990) Andrássy, 2009. The new species differs from A. papillata, currently recorded from UK only (Thorne & Swanger, 1936; Álvarez-Ortega & Peña-Santiago, 2010), in its larger general size (body 3.21–3.64 vs 2.43–3.08 mm long; neck 661–811 vs 510–650 μm long), longer uterus (191–350 vs 111–131 μm or 1.9–3.3 vs 1.3–1.5 body diameters long), more posterior vulva (V = 52–57 vs V = 45–50, n = 11) and male absent (vs present); and from A. parapapillata, recorded from South Africa (Botha & Heyns, 1990; De Bruin & Heyns, 1992), in its larger general size (body 3.21–3.64 vs 2.11–2.79 mm long; neck 661–811 vs 555–649 μm long), much longer odontophore (44–50 vs 26–38 μm, n = 12) and male absent (vs as frequent as female).

The new species described herein displays intermediate morphometrics between two South African species – namely, A. parapapillata and A. pseudospiralis – all of them described based on relatively few specimens. Aporcella femina sp. n. is very similar to A. pseudospiralis, but, as mentioned above, they differ in their overall size and especially in tail length. Therefore, the possibility of co-specificity could be raised, and the noted differences possibly due to geographical variations. The absence of molecular data for A. parapapillata and A. pseudospiralis currently prevents any molecular comparison, which, when available, would clarify any ambiguity. However, the available morphological information clearly distinguishes the three species from each other, supporting a separate status for them.

The evolutionary relations of the new species, as derived from the molecular analyses based on the partial sequence of SSU and LSU rDNA, are represented in figs 3 and 4. According to the SSU phylogeny, the new species is in a highly supported (BPP: 97%) sister relation with Aporcella vitrinus. Furthermore, Bayesian tree using partial LSU sequence shows that the new species forms part of a fully supported (BPP: 100%) subclade with Aporcella charidemiensis and A. vitrinus, the former being the closest relative, which is recorded from Spain. A highly supported (BPP: 94%) Aporcella clade is made of this subclade along with three Aporcella sequences belonging to A. simplex. Therefore, the present results confirm previous findings (for instance, see Naghavi et al., 2019; Vazifeh et al., 2020) about both the monophyly of Aporcella, and the belonging of Aporcella sequences to a large clade morphologically characterized by the absence of pars restringens vaginae, which also includes discolaims in the tree provided herein.

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Conflicts of interest. None.

Author contributions. M.R. and T.T.B. contributed equally.

References
Álvarez-Ortega S and Peña-Santiago R (2010) Studies on the genus Aporcelaimellus Heyns, 1965 (Dorylaimida: Aporcelaimidae) – material studied by Thorne and Swanger in 1936 but not named. Russian Journal of Nematology 18, 69–84.

Álvarez-Ortega S and Peña-Santiago R (2016) Aporcelaimella charidemiensis sp. n. (Dorylaimida: Aporcelaimidae) from the southern Iberian Peninsula, with comments on the phylogeny of the genus. Nematology 18, 811–821.

Álvarez-Ortega S, Subbotin SA and Peña-Santiago R (2013) Morphological and molecular characterisation of Aporcelaimella simplex (Thorne & Swanger, 1936) Loof & Coomans, 1970 and a new concept for Aporcella Andrássy, 2002 (Dorylaimida: Aporcelaimidae). Nematology 15, 165–178.

Andrássy I (2002) New genera and species of nematodes from southern Chile. Opuscula Zoologica Budapestensis 34, 5–22.

Andrássy I (2012) Two new species of the family Aporcelaiidae (Nematoda. Dorylaimida). Genus 23, 189–199.

Blaxter ML, De Ley P, Garey JR, et al. (1998) A molecular evolutionary framework for the phylum Nematoda. Nature 392, 71–75.

Botha A and Heyns J (1990) Aporcelaimidae (Nematoda: Dorylaimida) from the Kruger National Park. Koedoe 33, 27–46.

Coyne DL, Nicol JM and Claudius-Cole B (2007) Practical plant nematology: a field and laboratory guide. SP-IPM Secretariat, Cotonou, Benin.

Dorjima M, Ronald K, Andrew D, et al. (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Research 32, 1792–1797.

Imran Z, Abolafia J, Peña-Santiago R and Ahmad W (2019) Morphological and molecular characterisation of Moschaja idiofora Siddiqi, 1982
(Dorylaimida: Qudsianematidae), with new insights into the phylogeny of the enigmatic Lordellonematinae. *Nematology* **21**, 349–360.

Katoh K and Standley DM (2013) MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* **30**, 772–780.

Larget B and Simon DL (1999) Markov chain Monte Carlo algorithms for the Bayesian analysis of phylogenetic trees. *Molecular Biology and Evolution* **16**, 750–759.

Naghavi A, Niknam G, Vazifeh N and Peña-Santiago R (2019) Morphological and molecular characterisation of two species of *Aporcella* Andrássy, 2002 (Nematoda: Dorylaimida: Aporcelaimidae) from Iran, with new insights into the phylogeny of the genus. *Nematology* **21**, 665–665.

Naghavi A, Niknam G, Vazifeh N and Peña-Santiago R (2020) Study of two species, one new and one known, of the genus *Aporcella* Andrássy, 2002 (Dorylaimida, Aporcelaimidae) from Iran, with a note on its phylogeny. *Journal of Helminthology* **94**, e164.

Nico AI, Rapoport HF, Jiménez-Díaz RM and Castillo P (2002) Incidence and population density of plant-parasitic nematodes associated with olive planting stocks at nurseries in southern Spain. *Plant Disease* **86**, 1075–1079.

Rashidifard M, Marais M, Daneel MS, Mienie CMS and Fourie H (2019) Molecular characterisation of *Meloidogyne enterolobii* and other *Meloidogyne* spp. from South Africa. *Tropical Plant Pathology* **44**, 213–224.

Rashidifard M, Bello TT, Fourie H, Coyne DL and Peña-Santiago R (2020) Morphological and molecular characterisation of *Aporcelaimellus nigeriensis* sp. n. (Dorylaimida: Aporcelaimidae), a remarkable dorylaim from Nigeria. *Nematology* **22**, 867–877.

Ronquist F and Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* **19**, 1572–1574.

Subbotin SA, Sturhan D, Chizhov VN, Vovlas N and Baldwin JG (2006) Phylogenetic analysis of Tylenchida Thorne, 1949 as inferred from D2 and D3 expansion fragments of the 28S rRNA gene sequences. *Nematology* **8**, 455–474.

Thorne G and Swanger HH (1936) A monograph of the nematode genera *Dorylaimus* Dujardin, *Aporcelaimus* n. gen., *Dorylaimoides* n. gen., and *Pungentus* n. gen. *Capita Zoologica* **6**, 1–223.

Vazifeh N, Niknam G, Naghavi A, Jabbari H and Peña-Santiago R (2020) Description of *Aporcella minor* sp. n. (Dorylaimida: Aporcelaimidae) from Iran, with an updated taxonomy of the genus. *Nematology* **23**, 1–12.