p53/PUMA expression in human pulmonary fibroblasts mediates cell activation and migration in silicosis

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Phagocytosis of SiO2 into the lung causes an inflammatory cascade that results in fibroblast proliferation and migration, followed by fibrosis. Clinical evidence has indicated that the activation of alveolar macrophages by SiO2 produces rapid and sustained inflammation characterized by the generation of monocyte chemotactic protein 1, which, in turn, induces fibrosis. However, the details of events downstream of monocyte chemotactic protein 1 activity in pulmonary fibroblasts remain unclear. Here, to elucidate the role of p53 in fibrosis induced by silica, both the upstream molecular mechanisms and the functional effects on cell proliferation and migration were investigated.

Experiments using primary cultured adult human pulmonary fibroblasts led to the following results: 1) SiO2 treatment resulted in a rapid and sustained increase in p53 and PUMA protein levels; 2) the MAPK and PI3K pathways were involved in the SiO2-induced alteration of p53 and PUMA expression; and 3) RNA interference targeting p53 and PUMA prevented the SiO2-induced increases in fibroblast activation and migration. Our study elucidated a link between SiO2-induced p53/PUMA expression in fibroblasts and cell migration, thereby providing novel insight into the potential use of p53/PUMA in the development of novel therapeutic strategies for silicosis treatment.

The inhalation of silicon dioxide particles causes pneumoconiosis, an untreatable pulmonary disease characterized by alveolar inflammation at the early stage and progressive lung fibrosis at the late stage. Considerable evidence has suggested that alveolar macrophages (AMOs) and pulmonary fibroblasts (PFBs) mediate pulmonary fibrosis, which results from chronic inflammation1,2. While the role of SiO2-induced chemokines and cytokines released from AMOs has received significant attention, the direct effect of SiO2 on functional protein production in PFBs has been studied less extensively.

Tumor protein 53 (p53) is a tumor suppressor that acts as a key component of the cellular emergency response mechanism by up-regulating growth arrest and apoptosis-related genes in response to a variety of extra- and intra-cellular stress signals3–7. Emerging evidence has suggested that p53 plays roles in not only cell differentiation, apoptosis, and cell cycle control but also modulation of cell migration and other functions8. Indeed, p53 has been reported to regulate the transcription of a number of genes. Among these are genes whose products mediate cell-matrix interactions and cell motility, such as HGF/
scatter factor, collagens IIa1 and Vla1, macrophage-stimulating protein (Msp), plasminogen activator inhibitor-1 (PAI-1), fibronectin, VEGF and metalloproteinase-1. p53-up-regulated modulator of apoptosis (PUMA) is a recently discovered Bcl-2 family member that is rapidly induced by p53 and that exerts strong pro-apoptotic effects. Recent studies have suggested that p53 and PUMA most likely participate in the fibrotic process in a synergistic manner. However, the effects of p53 and PUMA on fibroblast regulation during silica-induced fibrosis remain unclear.

To elucidate the role of p53 in silica-induced fibrosis, we knocked down p53 expression and examined cellular behavior related to fibroblast-mediated contraction. Here, we demonstrated that p53 and PUMA are involved in the regulation of fibroblast contraction, growth and migration. These findings identified a new function of p53 in fibroblasts and suggested that p53 may be involved in multiple steps of the wound healing process.

Results
Effect of SiO₂ on p53 expression in human pulmonary fibroblasts. Accumulating evidence from our lab suggests that MCP-1 plays a critical role in SiO₂-induced pulmonary fibrosis. Our current data suggested that MCP-1 expression is increased in not only the bronchoalveolar lavage (BAL) fluid from patients but also the supernatant of cultured AMOs (Fig. 1A,B). Moreover, the expression of p53 in adult human pulmonary fibroblasts (HPF-a) increased when the cells were treated with the supernatant of cultured AMOs from patients (Fig. 1C,D). Additionally, AMOs from healthy donors released MCP-1 after exposure to SiO₂ in a time-dependent manner (Fig. 1E). Recent studies have suggested that p53 may serve as a pharmaceutical intervention against CCL2-mediated inflammatory responses.

To further understand the role of p53 in silica-induced fibrosis, we next examined the level of p53 in SiO₂-treated HPF-a. As shown in Fig. 1F,G, SiO₂ treatment of HPF-a resulted in the up-regulation of p53 expression in a dose-dependent manner. Because PUMA is known to be a downstream target gene of p53 that acts...
via p53-dependent and p53-independent pathways to promote apoptosis, we next measured the protein levels of both p53 and PUMA. As shown in Fig. 2A,B, the exposure of HPF-a to SiO2 induced rapid and sustained up-regulation of p53 and PUMA in a time-dependent manner. This result was confirmed by immunocytochemical staining (Fig. 2C–E).

Effect of SiO2 on the phosphorylation of MAPKs and Akt. To further understand the molecular mechanism underlying SiO2-induced fibrosis, we investigated the potential association between kinase activation and p53 and PUMA expression. Thus, we measured the protein levels of both p53 and PUMA in HPF-a within 3 h of exposure to SiO2. The protein level of α-SMA (a marker of fibroblast activation) was also examined. As shown in Fig. 3A–D, the levels of all proteins investigated peaked after approximately 15–60 min of SiO2 exposure and then tapered off. Next, we measured the phosphorylation of MAPKs and PI3K/Akt within 3 h of exposure to SiO2. As shown in Fig. 3E,F, Erk phosphorylation increased and then diminished within 5 min of exposure to SiO2. After 15 min of exposure to SiO2, p38 was phosphorylated, peaking after approximately 30–60 min and then tapering off (Fig. 3E,F). JNK also demonstrated a burst of activation from 15 to 60 min after exposure to SiO2 (Fig. 3H,I). In contrast, Akt displayed rapid and sustained phosphorylation after exposure to SiO2.

Effect of the pharmacological inhibition of MAPKs or Akt on p53 and PUMA induction after exposure to SiO2. After confirming that MAPK and Akt activities were modulated after exposure to SiO2, the effect of pharmacological inhibition of these kinases was examined (Fig. 4). The purpose of these experiments was to determine whether the kinase pathways of interest (JNK, ERK, p38, and PI3K/Akt) regulate 1) the expression of p53/PUMA and 2) the extent of fibroblast migration from the nested matrix. The 30 min time point after SiO2 exposure was selected to maximize the probability of detecting the effects of kinase inhibition because this time point corresponded to relatively large increases in p53 and PUMA expression in HPF-a after SiO2 exposure (see Fig. 3A–C). Pre-treatment of HPF-a with the commercially available small molecule U0126 (20 nmol/L, MEK inhibitor), SP600125 (20 nmol/L, JNK inhibitor) or SB203580 (20 nmol/L, p38 inhibitor) abolished the SiO2-induced up-regulation of p53 and PUMA expression (Fig. 4A–C). However, LY-294002 (20 nmol/L, Akt inhibitor) only partially inhibited the SiO2-induced up-regulation of p53 and PUMA expression (Fig. 4A–C). The dosage of inhibitors was based on the manufacturers’ recommended doses and previous studies from our lab. Both efficacy and

Figure 2. SiO2 induced both p53 and PUMA expression in HPF-a. (A) Representative Western blot showing that SiO2 induced both p53 and PUMA expression in HPF-a in a time-dependent manner. Scale bar = 10 μm. (B) Densitometric analyses of p53 and PUMA expression (n = 5). *p < 0.05 vs p53 expression on day 0; #p < 0.05 vs PUMA expression on day 0. (C) Representative immunocytochemical images showing that SiO2 increased the expression of p53 and PUMA in HPF-a. Densitometric analyses of p53 (D) and PUMA (E) expression (n = 5). *p < 0.05 vs the control group.
toxicity were evaluated (Supplementary Figure-S1 and Figure-S2). The results from these experiments indicated that the SiO2-induced expression of p53 and PUMA is mediated primarily by the MAPK pathways but not by the PI3K/Akt pathway.

Effects of p53 and PUMA RNAi on HPF-a viability and fibroblast-populated collagen matrix (FPCM) contraction. To understand the functional relevance of the changes in expression of p53 and PUMA, cell viability and activity were measured using the MTT assay and the collagen gel contraction assay, respectively18. SiO2 induced a significant increase in the viability of HPF-a (Fig. 4D), an effect that was abolished by RNAi of p53 and PUMA (Fig. 5A). Moreover, SiO2 treatment induced gel contraction in the control RNAi group, which suggested that fibroblast activity was up-regulated. However, RNAi of p53 and PUMA abolished the SiO2-induced increase in gel contraction (Fig. 5B,C).

Effect of p53 and PUMA RNAi on the production of collagen from HPF-a. Previous work from our lab has shown that SiO2 induces rapid and sustained production of collagen from HPF-a19. However, whether p53 and PUMA are also involved in the production of collagen remains unknown. As shown in Fig. 5D–F, RNAi of p53 and PUMA alone had no effect on the production of collagen I or collagen III. However, RNAi of p53 and PUMA inhibited the SiO2-induced increase in the production of collagen I and collagen III (Fig. 5D–F).

Effects of p53 and PUMA RNAi on the migration of HPF-a. Increasing evidence has suggested that pulmonary fibroblast migration is a critical aspect of pulmonary fibrosis. Therefore, we explored the roles of p53 and PUMA in SiO2-mediated cell migration. As shown in Fig. 6A,B, SiO2 induced a significant increase in the migration of HPF-a based on a scratch assay. However, RNAi of p53 and PUMA inhibited the SiO2-induced increase in cell migration (Fig. 6C,D).

Significant differences in cell physiology between two-dimensional (2D) and three-dimensional (3D) in vitro culture systems have been shown20–23. An FPCM culture system has facilitated the analysis of fibroblast physiology under conditions more closely resembling the in vivo-like environment than do...
conventional 2D cell culture systems. After using the scratch assay to determine that SiO₂ exposure induces the migration of HPF-a, we sought to validate these findings by monitoring HPF-a migration in a 3D cell culture system. As shown in Fig. 7A–C, SiO₂ induced a significant increase in cell migration in the control RNAi group, which was similar to the results of the scratch assay. Interestingly, although RNAi of PUMA abolished the increases in both the number of migrating cells and the migration distance induced by SiO₂, RNAi of p53 only partially inhibited these parameters (Fig. 7A–C).

Discussion
Silica exposure causes lung inflammation and fibrosis, which is histologically characterized by areas of inflammation, matrix deposition, and fibroblastic foci. Various in vitro and in vivo studies have been performed to investigate the role of AMOs in SiO₂-induced toxicity. Recent studies have suggested that the direct effect of SiO₂ on dendritic cells also plays an important role in the pathogenesis of fibrosis. In the current study, we focused on the effects of p53 and PUMA expression in pulmonary fibroblasts on cell proliferation and migration after in vitro exposure to SiO₂.

MCP-1 is a well-known member of the CC subfamily of chemokines that has been linked to inflammatory disease. Increased p53 expression was related to a decrease in fibroblast migration in the presence of firsthand cigarette smoke in a 2D in vitro model. Accumulating evidence has suggested a functional association between p53 and the regulation of cell morphology and motility. For example, a recent study suggested...
that p53 may play a role in Cdc42-mediated filopodia formation and cell polarization. Moreover, the p53 tumor suppressor pathway is functionally connected to the Rho GTPase pathways in regulating cell-ECM and cell-cell adhesions, as well as cell invasion properties. However, the p53-regulated genes that are required for SiO$_2$-induced fibrosis remain unclear. PUMA is well known for its role in mediating apoptotic responses induced by p53. For instance, PUMA deficiency in mice decreased mouse embryonic fibroblast apoptosis. The paradoxical finding that PUMA promotes angiogenesis by increasing cell proliferation and survival has also been reported. In fact, little is known regarding the function of PUMA in pulmonary fibroblasts. In this study, we found that both p53 and PUMA expression increased after exposure to SiO$_2$. Consistent with this, fibroblast activity also increased, as indicated by the elevated expression of collagen I and collagen III induced by SiO$_2$ was attenuated by RNAi of p53 and PUMA. Densitometric analyses of collagen I (E) and collagen III (F) expression from five separate experiments. 

Figure 5. p53 and PUMA mediated SiO$_2$-induced cell activation. (A) MTT assay showing that the SiO$_2$-induced effect on cell viability was abolished by RNAi of p53 or PUMA. (n = 5) *p < 0.05 vs the control group at the corresponding time point; #p < 0.05 vs the SiO$_2$ group at the corresponding time point. (B) Representative images of the collagen gel size showing that SiO$_2$ induced increased gel contraction (indicating fibroblast activation), which was abolished by RNAi of p53 or PUMA. (C) Quantification of gel size at different time points after SiO$_2$ exposure. (n = 6) *p < 0.05 vs the control group at the corresponding time point; #p < 0.05 vs the SiO$_2$ group at the corresponding time point. (D) Representative Western blot showing that increased expression of collagen I and collagen III induced by SiO$_2$ was attenuated by RNAi of p53 and PUMA. 

In this study, we also investigated the effects of p53 and PUMA up-regulation on cell proliferation and migration. Increasing evidence has suggested that fibroblast proliferation and migration increase during silicosis. In our in vitro model of silicosis, the direct effects of SiO$_2$ on HPF-a were consistent with previous findings. Both p53 and PUMA silencing via RNAi treatment attenuated HPF-a proliferation and activation induced by SiO$_2$, as indicated by the MTT and gel contraction assays. Interestingly, RNAi of PUMA only partially abolished SiO$_2$-induced cell proliferation, which indicated that the effect of p53 on fibroblasts may be mediated by both PUMA-dependent and -independent pathways. In fact, p53 was recently reported to exert its apoptotic effects via Noxa and Bim. Whether Noxa and Bim are also involved in p53-mediated activities in silicosis requires further investigation. Nevertheless, the proliferative effect of both p53 and PUMA was surprising because this finding suggested an important therapeutic strategy for silicosis.
To investigate SiO$_2$-induced cell migration, we applied both the convenient scratch assay and the nested matrix migration assay. Due to discrepancies in cellular behavior between 2D and 3D culture systems, the FPCM in vitro culture model provides opportunities to investigate cell-cell and cell-matrix interactions in an environment that more closely resembles physiological conditions. Cells in 3D matrices display distinct patterns of morphology and migration from those on a 2D monolayer. For instance, when fibroblasts are cultured on a 2D monolayer surface, the fractional force against the stiff substrate results in cell migration. However, when these cells are cultured in 3D collagen matrices, the fractional force can be utilized as a mechanical inducer of matrix remodeling both locally and globally. Therefore, understanding the molecular mechanism underlying fibroblast migration in a 3D matrix may be important for understanding various pathological diseases such as fibrosis. In this study, the results from the 2D scratch assay revealed that RNAi of p53 and PUMA abolished the increases in cell migration induced by SiO$_2$. However, in the 3D nested matrix migration model, while RNAi of PUMA abolished the cell migration induced by SiO$_2$, RNAi of p53 only partially abolished this effect. One explanation for this result is that p53 exerts its effect via a PUMA-independent pathway. Alternatively, PUMA may induce apoptosis in a p53-independent manner in response to a wide variety of stimuli, including genotoxic stress, deregulated oncogene expression, toxins, altered redox status, growth factor/cytokine withdrawal and infection. Recently, the role of PUMA in fibroblast activation has received significant attention. For instance, PUMA deletion attenuated pressure overload–induced apoptosis and fibrosis in mice subjected to heart failure. One interesting finding is that RNAi of p53 only attenuated the increase in the number of migrated cells but not in the migrated distance, which indicated a complex interaction between p53 and PUMA.

The limitation of our current study was that the fibroblasts used were purchased from a company that obtained the cells from one donor. Despite this single source, the fibroblasts have displayed phenotypic and functional heterogeneity, even when isolated from the same tissue. Importantly, however, previous work has shown that the growth rate pattern of the heterogenetic subpopulation obtained from one donor closely resembles those of subpopulations generated from other donors, indicating that heterogeneity is a consistent characteristic of fibroblasts. Nevertheless, experiments using different donors will worth conducting in future investigations.
In summary, our findings demonstrated that SiO$_2$ induces the expression of p53 and PUMA. Both p53 and PUMA play a vital role in pulmonary fibroblast activation and migration. The regulation of p53 and PUMA expression and function has potential as a novel therapeutic strategy for the treatment of silicosis.

### Materials and Methods

**Reagents.** Fetal bovine serum (FBS), normal goat serum (NGS), Dulbecco’s modified Eagle’s medium (DMEM; #1200-046), and 10X MEM (11430-030) were purchased from Life Technologies™. Amphotericin B (BP2645) and GlutaMax™ Supplement (35050-061) were obtained from Gibco®, and Pen Strep (15140-122) was obtained from Fisher Scientific. PureCol™ type I bovine collagen (3 mg/mL) was obtained from Advanced Biomatrix. Antibodies against p53 (SC6243), PUMA (SC374223) and β-actin (SC8432) were obtained from Santa Cruz Biotechnology®, Inc. The antibody against α-SMA (SAB5500002) was purchased from Sigma, Inc.

**Isolation of alveolar macrophages from human lung bronchoalveolar lavage fluid (BALF).** Use of primary AMOs derived from human lung BALF was approved in accordance with the approved guidelines of the Research and Development Committee of the First Affiliated Hospital of Nanjing Medical University. After obtaining informed consent from the subject, AMOs were isolated from bronchoalveolar lavage. This procedure was performed using a flexible fiber optic bronchoscope under local anesthesia of the upper airway with 2% lidocaine, as described previously$^{52,53}$. Briefly, the bronchoscope was wedged into the subsegmental bronchus of the right middle lobe or, in patients with EP, into areas of lung parenchyma that were otherwise normal based on chest roentgenography. Then, 150 mL of normal saline was instilled in 50 mL aliquots. Harvested BALFs were filtered through sterile nylon mesh and centrifuged at 160 × g for 10 min to obtain the cell preparation.

Macrophages were separated from the cell preparation by differential centrifugation using a Percoll solution. Macrophages isolated by this procedure exceed 95% purity$^{53}$. The isolated cells were plated in a sterile T-25 flask at 37°C and equilibrated with 5% CO$_2$ until the macrophages firmly adhered to the flask. Cell viability was assessed according to Trypan blue exclusion.

**HPF-a culture.** HPF-a were purchased from ScienCell. The cells were maintained in T75 flasks in DMEM containing 10% FBS. HPF-a were stored at passages 3–7 (P3-7) in liquid nitrogen. A vial of
P3-7 HPF-a between P10 and P15 was thawed, seeded, and passaged upon confluence to perform each experiment. We did not observe any correlation between passage number and any of the investigated parameters using cells in this range.

**Lentiviral transduction of primary HPF-a.** HPF-a were transduced with LV-RFP lentivirus (Hanbio, Inc., Shanghai, CN) as described previously. Briefly, P3-4 primary HPF-a were cultured in a 24-well plate (1 × 10⁴ cells/well) in DMEM containing 10% FBS for 48 h. Then, the medium was replaced with 1 mL of fresh medium containing 8 μg/mL polybrene. Next, 100 μL of lentivirus solution (10² IU/mL) was added to each well, followed by incubation at 37°C and 5% CO₂ for 24 h. After incubation, the treatment medium was replaced with fresh DMEM containing 10% FBS, and the cells were cultured at 37°C and 5% CO₂ until the cells reached >50% confluency. The transduced cells were selected using blasticidin as follows. Briefly, the medium was replaced with DMEM containing 10 μg/mL puromycin and 10% FBS, and the cells were cultured at 37°C and 5% CO₂ for 24 h. Then, the cells were washed twice with fresh DMEM containing 10% FBS. Pure transduced HPF-a cultures were expanded and/or stored in liquid nitrogen as described previously.

**MTT assay.** Cell viability was measured via the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) method. Briefly, the cells were collected and seeded in 96-well plates. Different seeding densities were employed at the beginning of the experiments. The cells were then exposed to I/R medium. Following incubation for different periods (3–24 h), 20 μL of MTT dissolved in Hank’s balanced salt solution was added to each well at a final concentration of 5 μg/mL, and the plates were incubated in a 5% CO₂ incubator for 1–4 h. Finally, the medium was aspirated from each well, and 200 μL of dimethyl sulfoxide was added to dissolve the formazan crystals. Then, the absorbance of each well was obtained using a plate reader at reference wavelengths of 570 nm and 630 nm. Each of the experiments was repeated at least three times.

**Gel contraction assay.** Fibroblast-populated collagen matrix (FPCM) contraction was determined using the floating matrix contraction assay as described previously with minor modifications. Briefly, the matrices were polymerized, covered with DMEM containing 5% FBS, released from the culture well using a sterile spatula, and incubated at 37°C. At different time points after the matrices were released, they were fixed in 4% paraformaldehyde in phosphate-buffered saline (PBS) at 4°C overnight, and images were obtained using a desktop flatbed scanner. The matrix area was measured using ImageJ software, and the data are presented as the ratio of the released matrix area to the attached matrix area.

**In vitro scratch assay.** Cell migration ability in the 2D culture system was evaluated using an in vitro scratch assay. Briefly, 1 × 10⁵ HPF-a were seeded in 24-well tissue culture plates and cultured in growth medium for 24 h, at which time they were approximately 70–80% confluent. Using a sterile 200-μL pipette tip, a straight line was carefully scratched in the monolayer across the center of the well in a single direction while maintaining the tip perpendicular to the plate bottom. A second straight line was similarly scratched perpendicular to the first line to create a cross-shaped cellular gap in each well. Each well was then washed twice with 1 mL of fresh growth medium to remove any detached cells. Digital images of the cell gap were captured at different time points, and the gap width was quantitatively evaluated using ImageJ software.

**FPCM.** The collagen matrix model was utilized as described previously with some modifications. The final matrix parameters were as follows: volume = 0.2 mL; diameter = 12 mm; collagen concentration = 1.5 mg/mL; and cell concentration = 1.0 × 10⁶ cells/mL. The matrices were established in 24-well plates (BD#353047), and the cells were incubated in the attached state in DMEM containing 5% FBS for approximately 48 h before the initiation of each experiment.

**Nested matrix model and cell migration assay.** The nested collagen matrix model was utilized as described previously with some modifications. For the nested attached matrix, a standard FPCM was incubated in the attached state for 72 h in DMEM containing 10% FBS. Then, the FPCM was removed from the culture well and placed in a 60-μL aliquot of fresh acellular collagen matrix solution (a NeoMatrix solution) that was centered inside a 12-mm diameter score on the bottom of a new culture well. Next, a 140-μL aliquot of NeoMatrix solution was used to cover the newly transferred FPCM. The NeoMatrix was allowed to polymerize for 1 h at 37°C and 5% CO₂, and then 2 mL of DMEM containing 10% FBS was added to the well.

Cell migration from the nested FPCM to the acellular NeoMatrix was quantified via fluorescent microscopy 24 h after nesting. Digital images (constant dimensions of 1000 × 800 μm) were captured using an EVOS® FL Cell Imaging microscope (Life Technologies, Grand Island, NY, USA) from 3–5 randomly selected microscopic fields at the interface between the nested FPCM and the acellular NeoMatrix. HPF-a migration from the nested FPCM was quantified by counting the number of cells that had clearly migrated from the nested matrix to the cell-free matrix. The maximum migration distance was quantified by identifying the cell that had migrated the longest distance from the nested matrix to the cell-free
The siRNA knockdown efficiency was determined 2 days after transfection via Western blot analysis. Medium was replaced with DMEM containing 10% FBS for 48 h before conducting further experiments. Added to the plated cells. The transfected HPF-a were cultured in serum-free DMEM for 24 h. Then, the lysis kit (MCL1-1KT, Sigma-Aldrich®) according to the manufacturer's instructions. The Western blot analysis was repeated using cells from three different donors. A single representative immunoblot is shown in each figure. Densitometry was performed using ImageJ software, and the results for all the repeated experiments were combined into one plot.

Immunoblotting. Immunoblotting was utilized as described previously with minor modifications. HPF-a were collected from the culture dishes, washed with PBS and lysed using a mammalian cell lysis kit (MCL1-1KT, Sigma-Aldrich®) according to the manufacturer's instructions. The Western blot membranes were probed with primary antibodies. Alkaline phosphatase-conjugated goat anti-rabbit or anti-rabbit IgG secondary antibodies were used (1:5,000). The signals were detected using chemiluminescence (SuperSignal West Dura Chemiluminescent Substrate, Thermo Scientific). Each Western blot analysis was repeated using cells from three different donors. A single representative immunoblot is shown in each figure. Densitometry was performed using ImageJ software, and the results for all the repeated experiments were combined into one plot.

Immunocytochemistry. HPF-a were fixed in 4% paraformaldehyde in PBS at 4°C overnight. Then, the fixed samples were permeabilized for 30 min at room temperature (RT) with 0.3% Triton X-100 in PBS. The permeabilized samples were blocked with PBS containing 10% NGS (Life Technologies) and 0.3% Triton X-100 at RT for 2 h. The blocked samples were incubated in primary antibodies in PBS containing 10% NGS and 0.3% Triton X-100 at 4°C overnight. Then, the samples were washed three times with PBS and incubated in donkey anti-rabbit (conjugated to Alexa-Fluor® 488) and donkey anti-mouse (conjugated to Alexa-Fluor® 576) secondary antibodies for 2 h at RT. After the samples were washed three times in PBS, they were mounted using Prolong® Gold antifade reagent with DAPI (P36931, Life Technologies). The slides were examined under an EVOS FL fluorescence microscope.

RNA interference of p53 and PUMA using siRNA. RNA interference targeting p53 and PUMA was performed on HPF-a as described previously with some modifications. The RNA interference protocol for a single well of a 24-well plate was as follows. Briefly, 49 μL of serum-free DMEM was combined with 1μL of transfection reagent, and 1μL of siRNA stock was added to 49 μL of serum-free DMEM. Then, both solutions were incubated at RT for 15 min. The transfection reagent and siRNA solutions were mixed together, and the resulting solution was incubated at RT for an additional 15 min. Meanwhile, HPF-a were seeded at a concentration of 5.0 × 10⁶ cells/100 μL/well in serum-free DMEM. The siRNA-vehicle solution was mixed and incubated at RT for 15 min. The siRNA-vehicle solution was added to the plated cells. The transfected HPF-a were cultured in serum-free DMEM for 24 h. Then, the medium was replaced with DMEM containing 10% FBS for 48 h before conducting further experiments. The siRNA knockdown efficiency was determined 2 days after transfection via Western blot analysis.

Statistics. The data are presented as the means±SEM. Unpaired numerical data were compared using an unpaired t-test (two groups) or ANOVA (more than two groups), and statistical significance was set at p < 0.05.

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Author Contributions
J.C. designed the experiments, analyzed the data, wrote the paper, and supervised the project; W.W. designed and performed the experiments, analyzed the data, and performed the statistical analyses; H.J.L., X.N.D., S.C.F., X.G.W. and Y.M.Z. performed the experiments and analyzed the data; H.H.Y. designed the experiments and supervised the project. X.L.Z. analyzed the data, wrote the paper, and supervised the project. All authors critiqued and approved the manuscript.

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