CaT1 and the Calcium Release-activated Calcium Channel Manifest Distinct Pore Properties*

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The calcium release-activated calcium channel (CRAC) is a highly Ca$^{2+}$-selective ion channel that is activated upon depletion of inositol triphosphate (IP$_3$)-sensitive intracellular Ca$^{2+}$ stores. It was recently reported that CaT1, a member of the TRP family of cation channels, exhibits the unique biophysical properties of CRAC, which led to the conclusion that CaT1 comprises all or part of the CRAC pore (Yue et al., 11). We confirm the resemblance of currents through CRAC and CaT1 but additionally present a number of novel features that are incompatible with the proposed equality of the CaT1 and CRAC pores.

In non-excitable cells, calcium transients upon receptor stimulation generally consist of two phases: a rapid rise due to Ca$^{2+}$ release from IP$_3$-sensitive intracellular Ca$^{2+}$ stores, which is followed by sustained Ca$^{2+}$ entry through store-operated plasma membrane channels (1). Of all store-operated channels, CRAC is functionally the best described, but its molecular identity and the mechanism of its store-dependent activation remain elusive (2). Several members of the TRP family, a group of cation channels related to the Drosophila transient receptor potential (TRP) gene product (3, 4), have been implicated in store-dependent Ca$^{2+}$ influx (5–9), but until recently none of them was found to exhibit the unique biophysical properties of CRAC (3).

In a recent study, Yue et al. (10) reported that CaT1 (11), a TRP family member homologous to Caenorhabditis elegans OSM-9, manifests the pore properties and store dependence of CRAC, and it has been proposed that CaT1 comprises all or part of the CRAC pore (10, 12, 13). To further investigate this intriguing possibility, we have made a direct comparison of CaT1, heterologously expressed in human embryonic kidney (HEK-293) cells, and endogenous CRAC in rat basophilic leukemia (RBL) cells. RBL cells were chosen because they were the source of the CaT1 clone used by Yue et al. (10) and have been widely used as a model system to study CRAC (2). We confirm the resemblance of currents through CRAC and CaT1 but additionally present a number of novel features that are incompatible with the proposed equality of the CaT1 and CRAC pores.

EXPERIMENTAL PROCEDURES

Cell Culture and Transfection—HEK-293 cells and RBL-2H3 cells were grown in Dulbecco’s modified Eagle’s medium containing 10% (v/v) human serum, 2 mM l-glutamine, 2 units/ml penicillin, and 2 mg/ml streptomycin at 37 °C in a humidity-controlled incubator with 10% CO$_2$. HEK-293 cells were transiently transfected with the pCINeo/IRESPGF/mCaT1 vector using previously described methods (14), and electrophysiological recordings were performed between 8 and 24 h after transfection.

Electrophysiology—Patch clamp experiments were performed in the tight seal, whole-cell configuration at room temperature. Patch pipettes had DC resistances of 2–4 MΩ when filled with the standard intracellular solution containing (in mM): 145 NaCl, 0, 3, or 10 MgCl$_2$, 3.3 CaCl$_2$, 10 EGTA, and 10 Hepes (pH 7.2). The free Ca$^{2+}$ concentration in this solution was calculated to be 100 nM. The standard extracellular solution contained (in mM): 145 NaCl, 2.8 KCl, 10 CaCl$_2$, 2 MgCl$_2$, 10 glucose, and 10 Hepes (pH 7.4). The divalent-free (DVF) solution contained (in mM): 150 NaCl, 10 EDTA, 10 glucose, and 10 Hepes (pH 7.4). To test the monovalent cation permeability sequence, DVF solutions were used in which NaCl was equimolarly replaced by LiCl, CsCl, KCl, or NMDG chloride. Unless indicated differently, currents were measured in response to voltage ramps (~100 to +100 mV in 100 ms) applied at a frequency of 0.5 Hz from a holding potential of 0 mV. Series resistances were between 3 and 10 MΩ and were compensated for 50–80%. Voltage errors were generally <10 mV. Currents were sampled at 1 kHz and filtered at 2 kHz using an eight-pole Bessel filter. For the accurate determination of the currents through CRAC, background currents measured under identical ionic conditions but before store depletion were subtracted. Averaged data are expressed as the mean ± S.E. from

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1 The abbreviations used are: IP$_3$, inositol triphosphate; CRAC, calcium release-activated calcium channel; HEK, human embryonic kidney; RBL, rat basophilic leukemia; DVF, divalent free; ECA/C1, epithelial calcium channel 1; 2-APB, 2-aminoethoxydiphenyl borate; VOCC, voltage-operated calcium channel; NMDG, N-methyl-D-glucamine.
Comparison of the activation mechanism of CaT1 and CRAC in DVF solution. A, time course of currents through CaT1 and CRAC at −80 mV in standard extracellular solution (10 mM Ca2+). CaT1 is instantaneously activated, whereas CRAC is activated by brief (5 s) application of ionomycin (2 μM). Currents were elicited by linear voltage ramps (2 V/s) from −100 to +100 mV applied every 2 s from a holding potential of 0 mV. B, current-voltage relations for CaT1 and CRAC in 10 mM external Ca2+. Traces were obtained in the experiments shown in A at the time points indicated by the arrowheads. For CRAC, the background current before application of ionomycin was subtracted. C, effect of switching from the standard extracellular solution to a DVF solution on currents through CaT1 and CRAC at −80 mV. D, current-voltage relations for CaT1 and CRAC in DVF solution. Traces were obtained in the experiments shown in C at the time points indicated by the arrowheads. For CRAC, the background current in DVF before store depletion was subtracted. In panels B and D, traces were normalized to the current at −80 mV.

Results

Immediately after obtaining the whole-cell configuration, CaT1-transfected HEK-293 cells exposed to 10 mM extracellular Ca2+ and with intracellular Ca2+ buffered at 100 mM to maintain full Ca2+ stores displayed a large membrane current (Fig. 1, A and B). This current subsequently decayed to reach a lower, but still robust, steady state level (Fig. 1A). The CaT1-mediated current was strongly inwardly rectifying (Fig. 1B) and reversed at positive potentials (39 ± 7 mV; n = 18) in agreement with previous work (10). Depletion of intracellular Ca2+ stores with the Ca2+-selective ionophore ionomycin (2 μM) did not induce any store-dependent activation of CaT1 but instead caused a transient inhibition of the CaT1-current by 26 ± 4% (n = 8) (Fig. 1A). A similar inhibition by 30 ± 6% (n = 7) was observed when Ca2+ stores were passively depleted by application of the sarcoplasmic reticulum Ca2+-ATPase inhibitor, 2,5-di-α-butyryl-1,4-benzoquinoline (20 μM). We attribute the inhibitory effect of store-depleting agents to an increase of intracellular Ca2+ as has been previously described for epithelial calcium channel (ECaC1; see Refs. 15 and 16), a close homologue of CaT1. Under identical intra- and extracellular ionic conditions, RBL-2H3 cells displayed only a very small calcium current as the slow removal of the Ca2+-induced inactivation of CaT1, as has been previously demonstrated for its close homologue, ECaC1 (15, 16). After reaching its maximal amplitude, the monovalent CaT1 current remained stable for more than 15 min (Fig. 1C and data not shown). In contrast, the current through CRAC, when preactivated by ionomycin in the normal extracellular solution containing 10 mM Ca2+, reached its maximal amplitude immediately after switching to DVF solution and decayed subsequently with a time constant of 12 ± 4 s (n = 17) (Fig. 1C).

Current-voltage relationships obtained during linear voltage ramps in DVF solutions revealed several significant differences between CaT1 and CRAC (Fig. 1D). First, the reversal potential for CaT1 (18 ± 2 mV; n = 8) was significantly less positive than for CRAC (45 ± 6 mV; n = 8). We examined whether this difference reflected different monovalent cation permeability profiles of the CaT1 and CRAC channel pores and found that, although both channels display a Na+ ~ Li+ > K+ > Cs+ > NMDG permeability sequence, CRAC has a much lower relative Cs+ permeability (I Cs/P Na = 0.48 ± 0.03 for CaT1 versus 0.12 ± 0.04 for CRAC; p < 0.001). Second, inward rectification was much more pronounced for CaT1 than for CRAC. The rectification score (defined as the ratio of the current amplitudes at potentials 40 mV negative and positive to the reversal potential) was 9.9 ± 1.1 (n = 15) for CaT1 compared with 1.5 ± 0.1 for CRAC (n = 19; p < 0.001). Third, the current-voltage relationship for CaT1 currents measured using the linear voltage ramp protocol displayed a negative slope at potentials below −80 mV, which was not observed for CRAC. As shown below, this negative slope originates from the time-dependent removal of intracellular Mg2+ block.

Monovalent cation currents through CaT1 activated during membrane hyperpolarization from +20 mV to −100 mV and deactivated during subsequent steps to less hyperpolarized
potentials (Fig. 2A). The fraction of open channels at the end of each voltage step was assessed from the initial current amplitude during a final step to −100 mV and exhibited a Boltzmann dependence on membrane potential (20), with a slope factor of 8 mV and half-maximal deactivation at −24 mV (Fig. 2D). This voltage dependence was abolished when Mg$^{2+}$ was omitted from the intracellular solution (Fig. 2, C and D), indicating that the gating mechanism of CaT1 is a voltage-dependent block/unblock of the channel pore by intracellular Mg$^{2+}$. Using pipette solutions containing different concentrations of Mg$^{2+}$, we found that half-maximal block at 0 mV was achieved with −200 μM intracellular Mg$^{2+}$. In contrast, monovalent cation currents through CRAC did not exhibit such voltage dependence (Fig. 2, B and D), not even with 10 mM intracellular Mg$^{2+}$.

It should be noted that CaT1 currents remained strongly inwardly rectifying in symmetric monovalent cation solutions and in the absence of intracellular Mg$^{2+}$, indicating that rectification represents an intrinsic pore property of CaT1. Moreover, the strong inward rectification and voltage-dependent Mg$^{2+}$ block of CaT1 were preserved in cell-free inside-out patches (data not shown).

2-aminoethoxydiphenyl borate (2-APB), an inhibitor of IP$_3$ receptors, is known to inhibit store-dependent Ca$^{2+}$ influx pathways in a variety of cells (21–25). We confirmed that 50 μM 2-APB caused a rapid and complete block of CRAC in RBL cells (Fig. 3A), which was voltage-independent (Fig. 3B and data not shown) and slowly reversible upon washout (Fig. 3A). The inhibitory effect of 2-APB on CRAC was dose-dependent with half-maximal inhibition at 5.5 μM (Fig. 3D). In contrast to a recent study (25), we did not observe a consistent initial increase in CRAC activity upon application of 2-APB. However, we cannot exclude the possibility that some 2-APB-dependent potentiation was masked by the rapid blocking effect of the drug. Although 2-APB has been shown to be membrane-permeable, two observations indicate that the target for 2-APB block of CRAC is exposed to the extracellular medium. First, the onset of block occurred with at time constant of −2 s (Fig. 3A), which is almost as fast as block of CRAC by extracellular Zn$^{2+}$, Cd$^{2+}$, or La$^{3+}$ (data not shown) and much faster than what has been reported for IP$_3$ receptor-mediated effects of 2-APB in intact cells (22). Second, dialysis of the cytoplasm with a pipette solution supplemented with 50 μM 2-APB did not markedly prevent activation of CRAC by ionomycin (data not shown; see also Ref. 23). From this we conclude that the interaction site for 2-APB is either on the extracellular part of the CRAC channel itself or at least closely associated therewith. In remarkable contrast with its inhibitory effects on CRAC, 50 μM 2-APB potentiated CaT1-mediated currents by −25% (Fig. 3, A and D). The potentiating effect of 2-APB did not show marked voltage dependence (Fig. 3C), was fully reversible upon washout (Fig. 3A), and could be repeated several times in the same cell. A similar but less pronounced potentiation was obtained with 5 μM 2-APB (Fig. 3D). Application of 250 μM 2-APB also caused a potentiation of CaT1, which was mostly followed by a slow and moderate inhibition of the currents by −20% and/or activation of linear leak currents. Thus, CRAC is at least 100 times more sensitive for 2-APB block than CaT1.

According to Yue et al. (10), the properties of CaT1, notably its activation by active store depletion, were dependent on the expression level of the channel protein. Therefore, we wanted to make sure that the differences we observed between CaT1 and CRAC were independent on the number of CaT1 channels per cell. Like Yue et al. (10), we measured CaT1 currents within 12 h after transfection and selected for cells that had current densities below 100 pA/pF at −100 mV with 10 mM extracellular Ca$^{2+}$ (mean peak amplitude, 78 ± 9 pA/pF; n = 8). Despite the fact that the average CaT1 current density in this group of cells was −5 times lower than the overall average (356 ± 44 pA/pF; n = 25), they displayed all of the above

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**Table I**

Summary of similarities and differences between CaT1 and CRAC

| Properties of CaT1 and CRAC | CaT1 | CRAC (RBL cells) |
|----------------------------|------|-----------------|
| **Similarities** | | |
| Calcium selectivity | $P_{Ca}/P_{Na} > 100^a$ | $P_{Ca}/P_{Na} > 100^b$ |
| Anomalous mole fraction behavior | Yes$^c$ | Yes |
| Divalent permeability | Ca$^{2+} > Ba^{2+} > Sr^{2+} > Mg^{2+}$ | Ca$^{2+} > Ba^{2+} > Sr^{2+} > Mg^{2+}$ |
| Monovalent permeability | Na$^+ > Li^+ > K^+ > Cs^+$ | Block$^e$ |
| Effect of La$^{3+}$, Zn$^{2+}$, Cd$^{2+}$ | Block | Block$^e$ |
| Single-channel conductance (150 Na$^+$) | 42–58 pS$^a$ | 55 pS$^d$ |
| **Differences** | | |
| Effect of ionomycin | Inhibition | Activation |
| Current in DVF | Stable | Transient |
| Inward rectification in DVF | Strong | Weak |
| $P_{Ca}/P_{Na}$ | 0.48 ± 0.03 | 0.12 ± 0.04 |
| Effect of intracellular Mg$^{2+}$ | Voltage-dependent block | No effect |
| Effect of extracellular 2-APB | Potentiation | Inhibition |

$^a$ From Ref. 10.
$^b$ From Ref. 32.
$^c$ From Ref. 33.
$^d$ From Ref. 34.
$^e$ From Ref. 23.
described CaT1 features: inhibition of 24% by 2 μM ionomycin (n = 2); strong inward rectification in DVF solution (rectification score, 12.7 ± 3.2; n = 4); relative Ca$^{2+}$ permeability (P$_{Ca}$/P$_{Na}$) of 0.52 ± 0.06 (n = 5); voltage-dependent gating in the presence of intracellular Mg$^{2+}$ with a slope factor of 8 mV and half-maximal deactivation at −25 mV (n = 4); potentiation to 121% of control by 50 μM 2-APB (n = 2). Thus, the biophysical and pharmacological properties that distinguish heterologously expressed CaT1 from endogenous CRAC in RBL cells are not influenced by the expression level of the CaT1 channel protein. Moreover, virtually identical results were obtained in CaT1-expressing Chinese hamster ovary cells (data not shown), indicating that the expression system does not significantly affect the properties of the CaT1 channel.

**DISCUSSION**

To evaluate the hypothesis that CaT1, a member of the TRP family of cation channels, constitutes all or part of CRAC, the calcium release-activated Ca$^{2+}$ channel, we made a direct comparison of the biophysical and pharmacological properties of both channels under identical experimental conditions. At first sight, the similarities between CRAC and CaT1 (Table I) are indeed striking. However, it should be noted that many of the common characteristics of the CRAC and CaT1 pores (high Ca$^{2+}$ selectivity; anomalous mole fraction effect; divalent and monovalent cation permeability sequence; block by inorganic cations; loss of selectivity in the absence of divalents; single-channel conductance to Na$^{+}$ in the range of 10–100 picoammens) are not only shared by ECaC1 (15, 26, 27), but also shared to a large extent by VOCCs (17), ECaC1/CaT1 (10, 27), and CRAC (31). The exchanger subunit, but this additional channel subunit would then serve in VOCCs (17), ECaC1/CaT1 (10, 27), and CRAC (31). The biophysical and pharmacological properties that distinguish heterologously expressed CaT1 from endogenous CRAC in RBL cells are not influenced by the expression level of the CaT1 channel protein. Moreover, virtually identical results were obtained in CaT1-expressing Chinese hamster ovary cells (data not shown), indicating that the expression system does not significantly affect the properties of the CaT1 channel.

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