Initial Characterization of the FlgE Hook High Molecular Weight Complex of Borrelia burgdorferi

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Abstract

The spirochete periplasmic flagellum has many unique attributes. One unusual characteristic is the flagellar hook. This structure serves as a universal joint coupling rotation of the membrane-bound motor to the flagellar filament. The hook is comprised of about 120 FlgE monomers, and in most bacteria these structures readily dissociate to monomers (~50 kDa) when treated with heat and detergent. However, in spirochetes the FlgE monomers form a large mass of over 250 kDa [referred to as a high molecular weight complex (HMWC)] that is stable to these and other denaturing conditions. In this communication, we examined specific aspects with respect to the formation and structure of this complex. We found that the Lyme disease spirochete Borrelia burgdorferi synthesized the HMWC throughout the in vitro growth cycle, and also in vivo when implanted in dialysis membrane chambers in rats. The HMWC was stable to formic acid, which supports the concept that the stability of the HMWC is dependent on covalent cross-linking of individual FlgE subunits. Mass spectrometry analysis of the HMWC from both wild type periplasmic flagella and polyhooks from a newly constructed ΔfliK mutant indicated that other proteins besides FlgE were not covalently joined to the complex, and that FlgE was the sole component of the complex. In addition, mass spectrometry analysis also indicated that the HMWC was composed of a polymer of the FlgE protein with both the N- and C-terminal regions remaining intact. These initial studies set the stage for a detailed characterization of the HMWC. Covalent cross-linking of FlgE with the accompanying formation of the HMWC we propose strengthens the hook structure for optimal spirochete motility.

Introduction

Spirochetes are a ubiquitous monophyletic motile phylum of bacteria, with many species causing disease [1]. These organisms, which are either helical or flat-waves, have a unique structure, as the organelles responsible for motility, referred to as the periplasmic flagella (PFs), reside between the outer membrane and protoplasmic cell cylinder. Each PF is attached to one cell end and extends towards the opposite end of the cell. Rotation of these relatively rigid organelles generates perturbations and wave propagation along the length of the relatively flexible cell cylinder such that the spirochetes efficiently translate and reverse directions even in viscous gel-like material and in vivo. Thus, the PFs need to exert substantial force on the cell cylinder to generate these backward moving waves [2-8].

Spirochetes also have several unique attributes with respect to their PFs. Each PF consists of a membrane imbedded motor where the basal body complex resides, a hook region that serves as universal joint, and the filament [2,3]. Thus, in many respects, the PF is similar in structure to the external flagellum of other bacteria. In contrast to the flagellum filament of most other bacteria, the PF filament consists of multiple protein species with a protein sheath surrounding the entire or part of the filament core. For example, Treponema and Brachyspira species have three different filament core proteins referred to as FlaB1, FlaB2, and FlaB3, and a sheath protein referred to as FlaA [2,3]. Borrelia burgdorferi has only one core protein referred to as FlaB and a sheath protein referred to as FlaA. Mutants that lack any of these proteins are deficient in motility [9,10]. In B. burgdorferi, mutants that lack FlaB are completely deficient in PFs, are non-motile, and are non-infectious [11,12]. The PF motor is also unique. Recent cryoelectron microscopy (cryo-EM) results suggest a specialized collar is associated with the motor in the vicinity of the peptidoglycan layer, but the function of the collar is presently unknown [13,14]. In addition, its cytoplasmic C-ring complex is considerably larger than that of other bacteria. The larger C-ring complex in spirochetes is believed to enable the PFs to exert sufficient torque...
such that the PFs promote wave propagation along the cell length [14]. Another unique attribute of spirochetes, which is the focus of this report, is their flagellar hook. The hook is a hollow tubular structure, approximately 61 nm long in B. burgdorferi, and is composed of at least 120 FlgE proteins [15]. FlgE is a well conserved protein among bacteria. Several reports indicate that FlgE of spirochetes form a stable, high molecular weight complex (HMWC) that is unique. Thus, western blots of whole cell lysates and in some cases purified PFs of B. burgdorferi, Treponema denticola, Treponema pallidum, and Treponema phagedenis probed with polyclonal FlgE antibodies react with a protein(s) located near the top of a gel when analyzed by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). Sometimes the HMWC resides even in the stacking gel, with a mass greater than 250 kDa [15–18]. This HMWC is considerably larger than the monomer (~50 kDa) [15–19]. In contrast, in bacteria with external flagella, western blots probed with FlgE antibody show reactivity exclusively at the molecular weight of monomeric FlgE [20,21]. The ability of spirochete FlgE to form a HMWC is similar to electrophoretic patterns found with covalent cross-linked proteins. For example, the head proteins of phage HK97 that infects Escherichia coli, and the pili proteins of Gram positive bacteria such as Streptococcus pyogenes and Corynebacterium diphtheriae also form HMWCs [22,23]. In addition, HMWCs from spirochete FlgE, HK97, and Gram positive pili are stable to a variety of harsh treatments and chemical agents, including boiling, guanidine hydrochloride, and 8M urea [15,18,22,23]. Accordingly, we and others hypothesize that the FlgE HMWC of B. burgdorferi and other spirochetes is attributed to covalent cross-linking among its individual proteins, which may be essential for optimum motility and virulence [15,18].

The characterization of the spirochete FlgE HMWC has been limited. Because FlgE is normally in small amounts (less than 9 µg per liter of cells of B. burgdorferi), chemical characterization is difficult. Several basic questions arise concerning these complexes. Are the HMWCs formed at all phases of B. burgdorferi in vitro growth, and are they formed in vivo in an infected animal? Are HMWCs stable to formic acid as are the head proteins of HK97 and pili of Gram positive bacteria [22,23]? Is there more than one protein, such as another basal body protein being part of the HMWC? Does the HMWC consist of a polymer of the complete FlgE protein, or is there truncation of either of its ends? In this communication, using the Lyme disease spirochete B. burgdorferi as a model that forms the FlgE HMWC, we answer the above questions.

Materials and Methods

Ethics statement

All animal experimentation was conducted following the Guide for the Care and Use of Laboratory Animals and in accordance with protocols reviewed and approved by the University of Connecticut Health Center Institutional Animal Care and Use Committee.

Bacterial strains and growth conditions

High passage, avirulent B. burgdorferi strain B31A [24], and low passage virulent strain B31A3 [25], were grown at 33°C, except as noted in the presence of 3% carbon dioxide in BSKII growth medium [11]. Cell counts were determined using flow cytometry as previously described [26]. These strains were originally obtained from P. Rosa, Rocky Mountain Laboratories, MT. The virulent, low passage strain B31MI [27] was grown in vitro at 23°C, 37°C, and in vivo in dialysis membrane chambers (DMCs) in Sprague-Dawley rats [28]. For in vivo growth, sterilized dialysis membranes with a molecular weight cut-off of 8000 Da were filled with ~ 9 ml BSKII medium containing late logarithmic phase B. burgdorferi strain B31MI diluted to a starting density of 3000 spirochetes/ml. DMCs were aseptically implanted into the peritoneal cavities of female Sprague Dawley rats, and then harvested 14 days post-implantation as previously described [28]. B. burgdorferi recombinant FlgE (rFlgE) was expressed in E. coli BL21 cells grown at 37°C in Luria-Bertani broth containing 100 µg/ml ampicillin and purified as previously described [15].

Construction of the ΔflIK1 mutant

The flIK mutant, ΔflIK1, was derived from high passage B. burgdorferi strain B31A; it was constructed as follows: The flIK gene (gene locus bb0283; 1179 base pairs) is located in the large flgB operon consisting of 26 genes [27,29]. This gene overlaps with the upstream flgD gene by 14 base pairs and is separated from the downstream bb0286 gene by eight base pairs. flIK was inactivated by replacing its coding sequence using overlapping PCR with an aadA streptomycin/spectinomycin resistance coding cassette that results in non-polar insertions [30]. Specifically, PCR was used to amplify three regions. First, each DNA region was amplified separately using primer pairs P1–P2 (5′-flanking DNA, flgD-flgE), P3–P4 (aadA coding sequence), and P5–P6 (3′-flanking DNA, bb0286-bb0287, Table 1). Second, a PCR product was obtained using primers P1-P4 with the purified DNA products flgD-flgE and aadA as templates. Third, the desired PCR product was obtained using primers P1–P6 with the purified DNA products flgD-flgE-aadA and bb0286-bb0287 serving as templates. The final 2836 flgD-flgE-aadA-bb0286-bb0287 base pair PCR product was gel purified and cloned into the pGEM-T Easy vector (Promega Inc.) to yield the flIK::aadA-pGEM plasmid. The integrity of the flIK inactivation plasmid was confirmed by PCR and restriction mapping. A PCR-amplified linear DNA product containing the flIK inactivation region was electroporated into B31A competent cells [24]. The electroporated cells were spread on semisolid BSKII medium containing 80 µg/ml streptomycin. Resistant clones were picked and analyzed by PCR using primers P1–P6 to confirm deletion of flIK. One clone, ΔflIK1, was found to have the desired mutation and further characterized.

SDS-PAGE and western blot analysis

SDS-PAGE and immunoblotting were conducted using standard techniques [15,31]. Molecular weight markers were obtained from Bio-Rad (cat no. 161–0576). All samples except as noted were boiled in sample buffer for at least 5 min. Samples were electrophoresed in 8% polyacrylamide gels, and silver stained with Protea Biosciences, Inc. silver staining kit or with Bio-rad Sypro protein gel stain. For western blotting, 1 µg total protein (unless noted otherwise), as determined by the Bradford protein assay (Bio-Rad), was subjected to SDS-PAGE and transferred onto polyvinylidene difluoride (PVDF) membranes. Immunoblots were probed with rabbit polyclonal antibody directed against B. burgdorferi FlgE [15], and secondary horse radish peroxidase (HRP)-labeled polyclonal donkey anti-rabbit antibodies obtained from GE Healthcare. FlaB was detected using monoclonal antibody H9724 to B. burgdorferi FlaB as previously described [11,32]. An enhanced chemiluminescence detection system was used to assay for reactivity (Pierce). Relative amounts of FlgE were determined by densitometry. After film development, films were scanned, and relative band intensities were determined using Image Quant software (GE Healthcare Life Sciences).
**Table 1. Primers used to generate the ΔfliK mutant**.

| Primer # | Name                  | Direction | Sequence (5′-3′)          |
|----------|-----------------------|-----------|---------------------------|
| 1        | FliK KO F             | Forward   | agattacaagcagaaagtaac     |
| 2        | FliK-Str-KO-Part 2-F  | Reverse   | ggtagtcgccaattgaagttttctacagtgatata     |
| 3        | FliK-Str-R            | Forward   | cactgtaagaaatcttatttgccagactcatggtagtg |
| 4        | FliK-Str-F            | Reverse   | tggagtagtgctatatagggagacgccggtagctgcgg |
| 5        | FliK-Str-KO-Part 1-R  | Forward   | caccgtctccccctatatcacaactacatctcaagtgact |
| 6        | FliK KO R2            | Reverse   | tagaacctttctttcgaaagctagcg |

*Underlined sequences indicate overlapping base pairs.

**PFs and polyhook isolation**

The PF isolation method of Sal et al. [15] was modified to increase the yield of the FlgE HMWC and to purify polyhooks. One liter of late logarithmic phase wild-type or ΔfliK mutant cells at a density of 1.5×10⁷ cells/ml were divided into four 250 ml portions. Cells from each portion were harvested by centrifugation (8,000 ×g, 20 min, 4°C), and the cell pellet was suspended in 28 ml of 150 mM phosphate buffered saline, pH 7.4 (PBS) and centrifuged (8,000 ×g, 15 min, 4°C). After suspending each pellet in 30 ml 0.15 M Tris, pH 6.8, (T buffer) the cells were centrifuged as indicated above, then resuspended in 15 ml of T buffer. The cell suspension was stirred for 10 min (23°C), and Triton X-100 was added (20% stock in sterile water) to a 2% final concentration. The cell suspension was stirred for 30–60 min at 23°C, or until the outer membranes were disrupted as observed by darkfield microscopy. Such spirochetes appeared thinner after treatment. Darkfield microscopy was carried out as previously described [33].

**Mass spectrometry (MS) analysis**

For mass spectrometry analysis (Protea Biosciences, Inc.), isolated PFs or polyhook samples were subjected to SDS-PAGE, and the gels were silver- or Sypro-stained, followed by in-gel digestion with trypsin. Gel bands of interest were excised, washed with 150 mM ammonium bicarbonate and acetonitrile, and lyophilized. The dried gel bands were treated with dithiothreitol and iodoacetamide followed by digestion with 12.5 ng/μl trypsin. The digested peptides were analyzed by LC-ESI MS using a QTrap5500 (AB Sciex Toronto, Canada). Peptides were separated at 35°C on a Kinetex 100×2.1 mm C18 column on a Shimadzu LC-20AD HPLC (Tokyo, Japan) using a 120-minute gradient. The mass range acquired (m/z) was 100-1000, and the three most intense multiply charged ions with ion intensities above 50,000 in each MS scan were subjected to MS/MS. The MS/MS data were searched for matches to protein databases using ABI Protein Pilot software 3.0. Only those peptides that were identified with >95% confidence were considered.

**Results and Discussion**

FlgE HMWC formation during different phases of growth

We first tested if the HMWC was synthesized at all phases of growth. Conceivably, cross-linking could occur in a specific growth phase analogous to, for example, *Escherichia coli* modifying its preformed unsaturated fatty acids by cyclopropanation primarily in stationary phase [36]. Furthermore, specific σ²-dependent genes have been shown to be upregulated in *B. burgdorferi* in stationary phase and during growth under specific conditions [37–39]. Accordingly, samples were taken throughout the *in vitro* growth cycle of *B. burgdorferi*, the cells were lysed, and then tested for the HMWC by western blot (Figure 1, A, B). We found that the HMWC was present in all phases of growth, and trace amounts of the monomer were sometimes detected. Often the HMWC was localized at the stacking-running gel interface, or as previously noted, within the stacking gel itself [15]. No differences were seen in the HMWC formation throughout the cycle. These results suggest that FlgE HMWC formation is not growth phase dependent.
FlgE HMWC formation at different temperatures and growth in vivo

Both the tick and mammal are the main habitats of B. burgdorferi in nature [39–41], and protein expression markedly varies in these two hosts [39,41]. In addition, many of the genes that are encoded by the 21 circular and linear plasmids present in B. burgdorferi are differentially expressed in the tick and mammal [40]. We tested whether HMWC formation occurred under conditions that mimic growth in the tick (23°C) and the mammal (37°C). In these experiments, we used a strain of B. burgdorferi which was virulent (B31M1), as such strains, in contrast to avirulent strains, show major differences in protein profiles when cultured under these two temperatures [42]. We found that the B31M1 strain grown in vitro at 23°C and at 37°C formed the HMWC (Fig. 1C). Similar results were found with cloned virulent strain B31A3 (K. Miller and N. Charon, not shown). These results indicate that the HMWC was synthesized under conditions that mimic growth in the tick and mammal. In addition, the results also indicate that the change in the protein profile that accompanies growth under these two conditions does not influence HMWC formation. We also asked if HMWC formation was a result of in vitro growth. B. burgdorferi cells grown in vivo are known to markedly change their transcriptional and protein profile compared to cells grown in vitro [28,39–41]; conceivably, the HMWC may not form in the mammal. Accordingly, we tested if the HMWC is synthesized in vivo by implanting B31M1 cells in DMC chambers in rats [28]. The cells were harvested at 14 days, centrifuged, and analyzed as before using immunoblotting. We found that the FlgE HMWC was present in cells grown in vivo in the DMCs similar to cells grown in vitro (Fig. 1C). In sum, the FlgE HMWC did not migrate as a discrete band or size, and it often localized to the stacking gel (Fig 1B, C). Conceivably, the HMWC could be composed of multiple masses that migrate anomalously in SDS PAGE. Protein crosslinking in the pili of Corynebacterium diphtheriae also results in diffuse bands upon SDS PAGE [23].

Figure 1. Synthesis of the HMWC under different growth conditions, and its isolation using mutanolysin. (A) Growth curve of B. burgdorferi B31A and (B) accompanying immunoblots of whole cell lysates probed with anti-FlgE, with samples taken at designated time points. (The position of molecular weight markers is indicated in Figures 1B–1D (50 – 250 kDa). (C) Immunoblots probed with anti-FlgE of strain B31MI grown at designated temperatures in vitro, and in vivo in DMC in rats. The amount of protein loaded was normalized to FlaB [32]. (D) Immunoblots probed with anti-FlgE of three independent PF isolation preparations from strain B31A using either the lysozyme or mutanolysin procedure. Approximately 0.5 µg of protein were loaded in each lane.

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Optimization of PFs and FlgE isolation

We began characterization of the HMWC. In our initial studies of *B. burgdorferi* FlgE, we used a method of lysing cells with myristate followed by lysozyme to purify PFs [15]. We found that this method was inadequate for chemical analysis of the HMWC (the yield was less than 1 mg, which is estimated to be less than 10% of the total FlgE protein in a liter of cells). Several modifications were made for optimization. These included substituting Triton X-100 for myristate, and mutanolysin for lysozyme. Triton X-100 has been shown to successfully remove the outer membranes of several spirochete species, including *T. phagedenis* [43], *T. pallidum* [44], *T. denticola* [45], and *B. burgdorferi* [46]. Although mutanolysin and lysozyme both hydrolyze the β-N-acetylmuramyl-(1→4)-β-N-acetylgucosamine linkage of the bacterial cell wall polymer peptidoglycan-polysaccharide, their activities are differentially affected by peptidoglycan modifications. Specifically, N- and O-acetylation inhibit lysozyme activity in a concentration dependent manner; however, mutanolysin maintains its effectiveness in the presence of these modifications [47–51]. We found that these changes increased the yield of the HMWC greater than three-fold (Fig. 1D). We also found that isolated PFs were often contaminated with lipid material as seen in cryo-EM. Using an alkaline wash as done for *S. enterica* flagella [34], isolated PFs were free of the lipid containing material (Fig. 2A,B). The total yield of PFs from a liter of cells varied from 2–4 mg. To begin characterization of the HMWC, we purified PFs, and subjected the fraction obtained to formic acid treatment. Stability to formic acid is the hallmark method used to show protein crosslinking of Gram positive pili [23]. As can be seen in figure 3A, boiling for 15 minutes with or without 88% formic acid failed to disrupt the HMWC. Additionally, the FlgE HMWC was observed when the samples were treated with formic acid without boiling, indicating that the HMWC is not an artifact produced by boiling (Fig. 3A). These results support the conclusion that the HMWC is the result of covalent protein-protein cross-linking. We attempted to determine the size of the HMWC. Hooks were prepared from isolated PFs and run on a Superose 12 molecular sieve column with or without pretreatment with 1% SDS. In both

Figure 2. Cryo-EM image of PFs, Δ*fliK* mutant, and polyhooks. (A) PFs isolated from wild-type *B. burgdorferi* B31A. (B) The hook basal body structure that is boxed in A is enlarged in B and accompanying structures identified. (C) The Δ*fliK* mutant with three flagellar polyhooks (~50 – 150 nm in length) localized in the periplasmic space between the outer membrane (OM) and protoplasmic cell cylinder (PC). (D) A 3D surface rendering of polyhooks and motors, suggesting the polyhooks are variable in length. (E) The lengths of 50 polyhooks in 9 intact *fliK* mutants was determined (F) Polyhooks isolated from the Δ*fliK* mutant using the mutanolysin procedure.

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cases, the protein eluted as a rather broad band, most of which was in the void volume, indicating the HMWC had a mass at least as large as 1 MDa (not shown).

Analysis of a flik mutant

The characterization of the spirochete FlgE HMWC has been limited because FlgE is normally in limiting amounts. Therefore, we considered the possibility that polyhooks obtained from B. burgdorferi would be a rich source of the HMWC. Although flik mutants in other bacteria including S. enterica [52,53] and T. denticola [17] produce large polyhook structures, we did not know if a similar mutant in B. burgdorferi would also produce polyhooks. In addition, we were curious if the polyhooks that are synthesized by the mutant would form FlgE HMWCs. Accordingly, we constructed a non-polar flik1 mutant of B. burgdorferi by allelic exchange mutagenesis. The flik1 mutant was non-motile, and cryo-EM indicated that polyhooks structures were produced and localized to the periplasmic space (Fig. 2 C,D; Movie S1); furthermore, flagellar filaments did not appear attached to the polyhooks (Fig. 2 C, D). The length of 50 polyhooks in 9 intact flik1 mutants was determined to range from 0 (no hooks attached to motors) to 846 nm, with a mean of 78 nm (Fig 2E). These results suggest that the flik1 mutant of B. burgdorferi is similar to those found in other bacteria, except in this case, the polyhooks are located in the periplasmic space rather than externally and exposed to the medium.

The flik1 mutant was further analyzed. Western blot analysis of whole cell lysates indicated that most of the reactivity of both the wild-type and the flik1 mutant formed a HMWC near the top of the gel, greater than 250 kDa (Fig 3B). A previous report demonstrated that cell lysates of flgE mutant cells failed to react with the FlgE polyclonal antiserum [15]. Small amounts of the monomer were detected in the mutant, as with the wild-type. Based on densitometry analysis, we estimate that the Δflik1 mutant produces 4–5 times the HMWC as that of the wild-type. We also tested if the Δflik1 mutant produced flagellin by probing cell lysates with FlaB antiserum. In other species of bacteria, flik mutants vary in their ability to continue to synthesize flagellin [54–57]. Although strong reactivity was noted with the wild-type, no reactivity was noted in the mutant (Fig. 3C), consistent with cryo-EM images (Fig. 2C, D). These results are similar to those of S. enterica flik mutant cells, as such mutants fail to produce flagella filaments [58,59]. We purified the polyhooks from the Δflik1 mutant for further immunoblot and cryo-EM analysis. Similar to results with whole cell lysates, FlgE antibody reactivity remained at the HMWC, with some reactivity with the monomer and at approximately 270 and 150 kDa (Fig. 3D). The pattern was similar to that seen with isolated wild-type PFs. Cryo-EM of this fraction identified large polyhook structures similar to those found in other bacteria [17,59] (Fig. 2F) with a diameter of 16 nm. However, these isolated polyhooks were longer (>200 nm, n = 15) than the mean of 78 nm observed in the periplasmic space of intact cells (Figure 2C,D,E). Therefore, our isolation procedure likely enriches for relatively long polyhooks. The Δflik1 mutant yielded 1–2 μg protein per liter in the isolated polyhook fraction, indicating that it would be an excellent source of the HMWC.

Finding polyhooks produced by this mutant is intriguing for several reasons. In bacteria, the length of the hook is tightly controlled. In the model system of S. enterica, interactions of FliK with FlhB determine hook length [52,54,59–62]. FliK acts as a molecular ruler that is secreted into the medium, and when the hook reaches proper length, FlhB undergoes a conformational change, and FlgE ceases to be added to the end of the forming hook. The cell then begins σ20-initiated transcription, and synthesis of the filament protein FlhC then commences with concomitant secretion to the growing flagellum [62]. B. burgdorferi does have a FlhB homolog (bb0272), so it is likely that a similar mechanism exists. However, because the PFs reside in the periplasmic space, if the S. enterica model applies to B. burgdorferi, FliK would be excreted into this space rather than into the medium as occurs in S. enterica. Furthermore, B. burgdorferi is unique, as it lacks the transcription cascade control of flagella gene regulation [2,3,27]. Several lines of evidence indicate that B. burgdorferi relies instead on translational control for flagella synthesis [15,63,64]. Conceivably, perhaps there is a translational control switch that allows fliA to be translated once the proper hook length is formed. In support of this notion, because the Δflik1 mutant is unable to stop synthesis of the hook once it reaches its proper length, we find that the resulting mutant also lacks FlaB (Fig. 3C).

MS of the HMWC

The revised PF and polyhook purification procedure allowed us to obtain sufficient material for MS analysis of the HMWC. We wanted to determine if other proteins were associated with this complex. We also wanted to determine if formation of the crosslinked HMWC resulted in truncation of either end of FlgE protein, analogous to the sortase-catalyzed reaction in gram-positive pili [65]. The protein composition of the HMWCs obtained from PFs and polyhooks was assessed directly by in-gel trypsin digestion followed by MS analysis (Fig. 4). The MS pattern of one of the peptides is illustrated (Fig. 4A). In 17 MS analyses of the HMWC, no other proteins were consistently identified in the HMWCs, and in many experiments, FlgE was the sole protein. For example, in one experiment with the HMWC obtained from

Figure 3. Immunoblot analysis of PFs and Δflik mutant. (A) Isolated PFs were treated with formic acid for 2 hrs at room temperature (lane 2) or 100°C for 15 minutes (lane 3). After formic acid treatment, sample buffer was added and the gel was loaded without additional boiling. The control was boiled for 15 min. 1 μg protein was loaded per lane, and the blot was probed with FlgE antisera. The position of molecular weight markers is indicated in Figures 3A, 3B and 3D (50 – 250 kDa). (B) Whole cell lysates (1.0 μg of wild-type and Δflik mutants probed with FlgE and (C) FlaB antisera. Reactivity was at 42 kDa with anti-FlaB. (D) Isolated polyhooks and PFs probed with FlgE antisera.
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PFs, 27 different peptides were identified, and all were derived from FlgE (Table S1). Similarly, in an experiment with the polyhook, 14 different peptides were identified, and again all were FlgE derived (Table S2). These results indicate that FlgE is the sole protein of the HMWC. We also asked if the N- and C-terminal regions were present in the HMWC. As can be seen from Fig 4B, 85% of the FlgE sequence was present in the complex, including peptides at the N- and C-terminal regions. These results suggest that the HMWC is not a cross-linked product of truncated FlgE. Now that we can obtain sufficient material for MS analysis from both wild-type cells and from a \textit{D}fliK mutant, we are actively determining which amino acids form the cross-link, and the precise mechanism of this cross-linking.

Why would cross-linking occur in the flagellar hooks of spirochetes, but not in the hooks of bacteria with external flagella?

Cross-linking of proteins in bacteria is rare, and when present it results in strengthening protein-protein interactions [66]. The hook acts as a universal joint, transferring the rotational motion of flagellar motors to filaments. In addition, this region has recently been shown to be the most flexible part of the flagellar apparatus; it even buckles when externally flagellated bacteria reverse direction of rotation [67]. Moreover, \textit{Vibrio alginolyticus} exploits this buckling for optimal cell reorientation during swimming [67]. In spirochetes, in contrast to bacteria with external flagella, not only do cells often reverse direction, the rigid PFs exert force on the cell cylinder to generate backward moving waves for translation [2,4–8]. We hypothesize that FlgE cross-linking is essential for the spirochetes to avoid flagellar buckling and allow the cell to reverse directions. This cross-linking also allows the hook to be sufficiently strong such that the PFs can exert adequate force against the cell cylinder to generate the waves responsible for motility. Our results suggest that \textit{B. burgdorferi} FlgE crosslinking occurs in the periplasm during or after the FlgE proteins assemble to form the hook [15]. Whether this crosslinking is self-catalyzed or catalyzed by another protein is not yet known. Searching the \textit{B. burgdorferi} genome did not identify any gene coding a presumptive transglutaminase or sortase protein, nor does the FlgE sequence contain the sortase motif [23].

Supporting Information

\textbf{Movie S1} 3D visualization of the \textit{Aflik} mutant. A 3D reconstruction was generated by cryo-electron tomography of frozen hydrated bacterial culture of the \textit{Aflik} mutant. The movie first shows the slices through the 3D reconstruction, then 3D surface rendering of major features, such as the outer membrane, the inner membrane, and the flagellar motors. In particular, the movie highlights the various lengths of the polyhooks in the same cell. (MOV)
Table S1 FlgE peptides identified in isolated PFs.
Isolated PFs were run on SDS PAGE, excised, trypsin-digested and analyzed by MS as described. Those peptides identified with >95% confidence (conf) are shown, highlighted in green. (XLSX)

Table S2 FlgE peptides identified in isolated polyhooks.
Polyhooks were isolated from the fliK mutant and run on SDS PAGE, excised, trypsin-digested and analyzed by MS as described. Those peptides identified with >95% confidence (conf) are shown, highlighted in green. (XLSX)

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