Carbon flux from decomposing wood and its dependency on temperature, wood N₂ fixation rate, moisture and fungal composition in a Norway spruce forest

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Abstract
Globally 40–70 Pg of carbon (C) are stored in coarse woody debris on the forest floor. Climate change may reduce the function of this stock as a C sink in the future due to increasing temperature. However, current knowledge on the drivers of wood decomposition is inadequate for detailed predictions. To define the factors that control wood respiration rate of Norway spruce and to produce a model that adequately describes the decomposition process of this species as a function of time, we used an unprecedentedly diverse analytical approach, which included measurements of respiration, fungal community sequencing, N₂ fixation rate, nirH copy number, ¹⁴C-dating as well as N%, δ¹³C and C% values of wood. Our results suggest that climate change will accelerate C flux from deadwood in boreal conditions, due to the observed strong temperature dependency of deadwood respiration. At the research site, the annual C flux from deadwood would increase by 27% from the current 117 g C/kg wood with the projected climate warming (RCP4.5). The second most important control on respiration rate was the stage of wood decomposition; at early stages of decomposition low nitrogen content and low wood moisture limited fungal activity while reduced wood resource quality decreased the respiration rate at the final stages of decomposition. Wood decomposition process was best described by a Sigmoidal model, where after 116 years of wood decomposition mass loss of 95% was reached. Our results on deadwood decomposition are important for C budget calculations in ecosystem and climate change models. We observed for the first time that the temperature dependency of N₂ fixation, which has a major role at providing N for wood-inhabiting fungi, was not constant but varied between wood density classes due to source supply and wood quality. This has significant consequences on projecting N₂ fixation rates for deadwood in changing climate.

Keywords
activation energy, boreal forest, carbon flux, coarse woody debris, N₂ fixation, nirH, respiration rate, wood-inhabiting fungi
1 | INTRODUCTION

Decomposition of deadwood releases globally billions of tons of carbon (C) per year to the atmosphere (Harmon, Krankina, Yatskov, & Matthews, 2001), a magnitude that is similar to the emissions from fossil fuel combustion (Le Quéré et al., 2013). However, the current estimate is based on an incomplete knowledge of drivers of wood decomposition and their relation to ambient conditions, such as temperature and soil moisture, which are likely to change in the future. Consequently, it is important to quantify the factors that control respiration rates and to determine the pattern and magnitude of C flux rates at different stages of decomposition, so that the contribution of this C stock to total ecosystem respiration can be adequately accounted for in ecosystem, C accounting and climate change models.

The dynamics of wood respiration are still relatively poorly understood in comparison to other fluxes of forest respiration (Liu, Schaefer, Qiao, & Liu, 2013). Firstly, the published models describing wood decomposition have substantial differences: they predict different temporal patterns of decomposition and highly variable residence times for deadwood of the same tree species on forest floor (Harmon et al., 1986). The negative exponential model, which is one of the most commonly used to estimate wood decomposition, predicts fastest decomposition rate for Norway spruce at the beginning of decomposition and a long total decomposition time (>100 years; Melin, Pettersson, & Nordfjell, 2009 and references therein). In contrast, for the same tree species, the multiple-exponential model of Mäkinen, Hynynen, Siltovan, and Sievänen (2006) predicts a slow decomposition rate for the first couple of decades followed by a fast rate of decomposition, which leads to 100% mass loss 60 years after death. Secondly, it has been proven difficult to disentangle the relative importance of the different factors driving decomposition. For example, according to some studies temperature and wood moisture content are the dominant factors influencing decomposition rates via their impact on microbial activity (Olajuyigbe, Tobin, & Nieuwenhuis, 2012; Wang, Bond-Lamberty, & Gower, 2002), while other studies have reported that these variables explain small portions of total variance in decomposition rate (Bradford et al., 2014; Yang et al., 2016). Other factors reported to affect the decomposition process include substrate quality (e.g. concentration of slowly decomposing lignin) (Mackensen & Bauhus, 2003; Tuomi, Laiho, Repo, & Liski, 2011), wood nitrogen (N) content (Weedon et al., 2009), soil macrofauna (Jacobs & Work, 2012) and fungal diversity (Valentin et al., 2014) and community structure (Rayner & Boddy, 1988).

Although community structure of wood-inhabiting fungi has been linked in several studies to decomposition rate of deadwood, the actual affiliated mechanisms are still poorly understood (Rajala, Peltoniemi, Pennanen, & Makipää, 2012). One of the controlling factors is likely the succession of the fungal communities along which also physical and chemical properties of wood change. The successional change in fungal community of boreal forests generally follows a temporal pattern, initiated by the relatively inefficient soft-rot fungi, followed by brown-rot fungi and then by white-rot fungi, which have the unique capability to efficiently decompose wood lignin (Hyde & Jones, 2002; Rajala, Peltoniemi, Hantula, Mäkipää, & Pennanen, 2011). Finally, in boreal forests, wood becomes dominated by ectomycorrhizal (ECM) fungi, which transport N and C between soil and host plants, at the last stages of decomposition (Mäkipää et al., 2017).

In addition, bacteria are considered to play a minor role in wood decomposition (Clausen, 1996), but recent studies have proven early hypothesis (Cowling & Merrill, 1966) that e.g. N₂ fixing bacteria may have a remarkable contribution on N availability in the decomposing wood (Rinne et al., 2017).

Little is still known about how different fungal species present in deadwood influence wood decomposition rate (Hoppe et al., 2016; Valmaseda, Almendros, & Martínez, 1990). Another factor that has been linked to decomposition rate, however with significant controversy, is fungal richness, measured with DNA based methods as a number of operational taxonomic unit (OTU). Some studies have reported a positive relationship between the two variables at the early to mid-stages of decomposition (Valentin et al., 2014), which has been explained by the positive impact the higher diversity of produced fungal enzymes with increasing number of OTUs has on communities’ wood decomposition efficiency (Gessner et al., 2010). However, other studies have found a negative relationship between decomposition rate and OTU richness (Lindner et al., 2011; Yang et al., 2016) or no relationship at all (Hoppe et al., 2016). Other significant factors that have been reported to affect decomposition rates are competition scenarios between fungi (Renvall, 1995) and the mutualistic relationship between fungi and N₂ fixing diazotrophs (Hoppe et al., 2014).

Apart from being a substantial stock of terrestrial C, deadwood is an important component of nutrient cycling (Laiho & Prescott, 2004). This includes N, which accumulates in deadwood during the decomposition process mainly via asymbiotic N fixation (Rinne et al., 2017). N may also be used during the decomposition process via sporocarp and spore production or lost via wood fragmentation and leaching. Since saprotrophic fungi grow in wood with a very high C:N ratio and since they are at the same time highly dependent on the availability of N for production of fungal material (Moore, Gange, Gange, & Boddy, 2008), it has been suggested that there may exist a significant mutualistic relationship between N₂-fixing bacteria and wood decay fungi (Cowling & Merrill, 1966; Mäkipää et al., 2018). The first evidence of this was seen in the study of Hoppe et al. (2014), who reported a positive correlation between the number of fungal fruiting bodies and a nitrogenase-related gene, nifH, diversity. How well nifH distribution in decomposing wood relates to actual N₂-fixation and respiration rates are yet to be determined. Furthermore, fungal fruit bodies, as used in the study of Hoppe et al. (2014), may not be representative of the fungal community present in deadwood (Hoppe et al., 2016), and hence fungal community sequencing could provide additional information on the associations between diazotrophs and wood-habiting fungi.
Both respiration and N<sub>2</sub> fixation rates of fallen deadwood may increase as ambient temperature increases due to ongoing climate change, if wood moisture does not become limiting (too low or too high). Hence, climate warming may have significant consequences for the role of deadwood as a long-term C reservoir and on the availability of N in acutely nutrient-limited high latitude forest ecosystems. The metabolic theory of ecology (MTE) (Brown, Gillooly, Allen, Savage, & West, 2004) argues that the impact of climate warming on the decomposition and N<sub>2</sub> fixation rates of deadwood can be predicted from the temperature dependencies of the corresponding biochemical reactions, respiration and N<sub>2</sub> fixation, respectively. The temperature sensitivity of these reactions is quantified by the activation energy (AE) (Arrhenius, 1915), with a canonical value of 2.18 eV for the nitrogenase enzyme (Ceuterick, Peeters, Heremans, Smedt, & Olbrechts, 1978) and of 0.60–0.70 eV for respiration (Allen, Gilloolz, & Brown, 2005), for the temperature ranges 0°C–22°C and 0°C–30°C, respectively. However, the MTE does not take into account limitations due to resource supply, which may lead to AEs that diverge from the canonical value. For example, Welter et al. (2015) reported that stream biofilm development in Iceland, amplified temperature response of respiration owing to increased N supply, which was the result of increased temperature. Conversely, Follstad Shah et al. (2017), who used data from 169 published studies, concluded that litter decomposition rates had AEs that were approximately half the value predicted by metabolic theory due to limitations for example in litter supply regime, litter quality and detritivore density. For deadwood a similar study on the AE of respiration and N<sub>2</sub> fixation has not been conducted to the best of our knowledge.

This study was conducted on deadwood of Norway spruce (Picea abies (L.) Karst.) to define the factors that control wood decomposition rates at different stages of decomposition and to resolve the debated question on the type of model that best describes the decomposition process of this species as a function of time. Furthermore, we used this knowledge to obtain estimates of annual CO<sub>2</sub> production rates from deadwood in current conditions and as a result of projected climate warming. To achieve our goals we used an unprecedentedly diverse dataset, which included measurements of respiration, fungal community sequencing, N<sub>2</sub> fixation rate, nifH copy number, <sup>14</sup>C-dating as well as N content, δ<sup>13</sup>C and C% values of wood.

# 2 MATERIALS AND METHODS

## 2.1 Site description and fieldwork

Fieldwork was carried out at the Lapinjärvi forest located in southern Finland (60°66N, 26°12E). It is an unmanaged site, with a living Norway spruce trees biomass of 119,000 kg/ha (296 m<sup>3</sup>/ha) that constitutes 72% of all tree species (Rajala et al., 2012). The biomass of spruce deadwood, which accounts for 89.4% of all deadwood, is shown in Table 1 for each decay class, as classified by the system of Harmon and Sexton (1996). Two 5 cm thick disks were obtained from 49 decomposing logs. The collected disks were evenly distributed between the five decay classes (Mäkipää et al., 2018). The disks were packed into plastic bags and stored within the same day in a freezer at -20°C until processed. Under natural forest floor conditions logs experience freeze-thaw cycles annually, so the storage method was not considered to have an unnatural impact on the analytical results. Wood density was determined for each log as described in Rajala et al. (2012) using one of the two obtained discs. Details of the density classification (Table 1) are given in Rinne et al. (2017) and in Supporting Information S1. In addition, seven living Norway spruce trees were sampled to the pith using an increment borer for determining the initial wood C%.

## 2.2 Meteorological data

A 2-year record of average daily temperature measured with data loggers at 1 m above ground level in Ruotsinpyhtää forest was used. The forest, which is located 18 km from the Lapinjärvi forest, has a similar tree composition (spruce dominated, tree density), mean annual temperature (4.6°C) and annual precipitation (618 mm) as Lapinjärvi.

## 2.3 Incubation and gas chromatography (GC) analysis

Each of the thawed disks was chopped into wood chips and divided into eight equal subsamples that were placed into 118 ml bottles with an air tight cap and the fresh weight was recorded. Seven of the bottles were used for CO<sub>2</sub> measurements and one was reserved for other analyses (molecular and isotope analyses). First, all eight bottles were preincubated for 1 week at 14.5°C. Subsequently, the seven bottles reserved for gas chromatography were incubated 5 days at different temperatures (8.7°C, 14.5°C, 20.5°C, 26°C, 30°C, 34°C and 40.1°C) and the additional eighth bottle reserved for other analyses was incubated at 14.5°C. Samples were incubated under stable conditions by controlling the moisture content (replacement of evaporated water) and aerating the bottles (30 s per aeration) during the preincubation and incubation periods. The samples that were reserved for analyses other than GC (molecular and isotope analyses) were frozen and freeze-dried following the incubation period.

For the incubated samples respiration rates were determined by sampling the headspace of a bottle and analysing the amount of CO<sub>2</sub> using a gas chromatogram (Smolander, Kitunen, Tamminen, & Kukkola, 2010). The bottles were aerated 24 hr before sampling. Finally, samples were dried in an oven at 60°C for 48 hr and the dry mass was calculated for each sample.

## 2.4 Molecular and bioinformatics analyses

The 49 freeze-dried samples were milled to fine particle size to ensure homogeneity for molecular and isotope analysis. Details about...
DNA extraction, fungal internal transcribed spacer (ITS) sequencing procedure with Illumina platforms as well bioinformatics for the obtained fungal ITS data are described in Supporting Information S2. Shannon diversity indices from fungal ITS data were obtained by using the summary.single command in the MOTHUR software package v.1.3.6 (Schloss et al., 2009). Venn diagrams were constructed to view unique and shared fungal OTUs in wood density classes by using the venn() command implemented in library gplots in R package version 3.2.2.

To detect the copy numbers of N₂-fixing bacteria, we conducted quantitative PCR (qPCR) targeting the nifH gene with PoIF forward and PoIR reverse primers (Poly, Monrozier, & Bally, 2001). First standard curves were constructed with plasmids containing corresponding inserts, taking into account the molecular mass of the plasmid, including the insert, and the plasmid concentration. We ran qPCR (Rotor-Gene 6000; QIAGEN, The Netherlands) with Maxima™SYBR Green qPCR Master Mix (2×) (Thermo Fisher Scientific, Germany) in a 20 ml final reaction volume containing 1 μl template, 0.3 mM of each primer and 1× qPCR master mix. Fluorescence was measured at the end of each extension step. Each qPCR run was carried out under the following conditions: initial denaturation at 95°C for 10 min, 40 cycles denaturation at 95°C for 15 s, annealing at 55°C for 30 s and extension at 72°C for 30 s, and final extension at 72°C for 10 min and from 60°C to 99°C (ramp: 1°C per 5 s). All samples were replicated and lack of PCR inhibition was verified through 1:10 dilution. The copy numbers in samples were calculated based on comparison to threshold cycle values of the standard curve and are given per gram of soil (dry weight).

### 2.5 Stable isotope analysis

The 49 freeze-dried and milled deadwood samples were weighed into tin capsules for C% and δ¹³C measurements in an elemental analyser (EA; Costech 4010; Costech Analytical Technologies Inc., Valencia, CA, USA) connected in continuous-flow mode to an
isotope ratio mass spectrometer (IRMS; Finnigan Delta Plus XP; Thermo Fisher Scientific, Waltham, MA, USA). The standard deviations for repeated analyses of reference materials were better than 0.2‰. The isotope signature is expressed in the delta notation $\delta^{13}C = \left( \frac{R_{\text{sample}}}{R_{\text{standard}}} - 1 \right) \times 1,000$ (‰) relative to the international standard VPDB, where $R_{\text{sample}}$ is the $^{13}C/^{12}C$ ratio of the sample or standard.

2.6 | $N_2$ fixation rate

The $N_2$ fixation rate of Lapinjärvi deadwood and its dependency on density class and ambient temperature has been published in Rinne et al. (2017), where the methodology is described in detail. In short, an acetylene reduction assay (ARA) was performed following Leppänen, Salemaa, Smolander, Mäkipää, and Tiirola (2013). After 24 hr of incubation the concentration of ethylene in the headspace was analysed by a gas chromatograph. A conversion factor from acetylene reduction to $N_2$ fixation of 4 was used (Brunner & Kimmins, 2003). In the present study the $N_2$ fixation data (Rinne et al., 2017) is for the first time compared with respiration rate, $nifH$ copy number and fungal community of Lapinjärvi deadwood, and used to calculate the AE of the $N_2$ fixation rate for the different decay classes. The density, N content and moisture of deadwood used in the present study (Table 1) are from Rinne et al. (2017).

2.7 | Activation energies

Activation energies were estimated individually for respiration and $N_2$ fixation using the Van’t Hoff-Arrhenius relationship $e^{-E/(kT)}$, where $E$ is the AR (in eV), $k$ is the Boltzmann constant ($8.61 \times 10^{-5}$ eV/K, 1 eV = 96.485 kJ/mol) and $T$ is temperature (K, here the incubation temperature) (Arrhenius, 1889). For each density class the data were plotted separately in a xy-diagram, where $x$ was $1/(kT)$ and $y$ was the natural logarithm of each respiration or $N_2$ fixation measurement. Linear least-squares regression was used to describe the relationship between the variables. The AE is the absolute value of each slope.

2.8 | Statistical analysis

CurveExpert 1.4 software package was used to determine the best-fitting trends for the respiration series. When obtaining best-fit trends for the part of the respiration dataset that covered only the temperature range typical for the study area (<20°C, Figure 1b), it was necessary to add data points that approximated zero values for both CO$_2$ production rate and temperature (0.001 g CO$_2$ kg$^{-1}$ day$^{-1}$ and 0.001°C, respectively) for the trendline fitting process. With this addition the resulting best-fit trends did not predict a significant production of CO$_2$ at 0°C. The number of these added data points equalled to the number of samples.
incubated at 8.8°C for each density class (n_I = 10, n_II = 7, n_III = 8, n_IV = 11 and n_V = 11).

IBM SPSS Statistics 22 software package (IBM Corp, Armonk, NY, USA) was used for other statistical analysis: ANOVA and ANCOVA with the Tukey post-hoc test were used to determine any significant differences between the means of groups, and simple linear and multiple regression analyses were used to study the relationship between two or more variables.

2.9 | The total amount of C stored in Lapinjärvi deadwood and its annual C flux

Annual CO₂ production per hectare (kg C ha⁻¹ year⁻¹, Figure 1c) was estimated for Lapinjärvi deadwood using the trendlines of Figure 1b, the weight of wood per hectare calculated for each decay class (“log density” in Table 1) and the meteorological data. The total annual level of C flux per gram of Lapinjärvi deadwood (g C kg⁻¹ year⁻¹) was calculated by dividing the density class specific results of Figure 1c by the density of fallen spruce logs in each density class (Table 1) and by summing the results. The total amount of C stored in Norway spruce deadwood in the study site was calculated for each density class (Table 1) using the C content (Figure 2), the density of each deadwood sample (Rinne et al., 2017) and the quantity of downed dead trees (Table 1).

2.10 | Climate warming scenario RCP 4.5

The potential impact of climate warming on CO₂ production rates was calculated using the RCP4.5 temperature scenario, which assumes that a further increase in anthropogenic CO₂ production rate is followed by a decrease starting around year 2040, for the time period 2040–2069 for southern Finland (Ruosteenoja, Räisänen, & Jylhä, 2013). The projected temperature increase is approximately 1.4°C for the summer months and increasingly higher towards the winter months, where up to 3.8°C increase is estimated.

3 | RESULTS

3.1 | Respiration

For each density class the statistically significant temperature dependency of CO₂ production followed the Gaussian model, where maxima was at 29.6°C (for wood in density class I–IV) or at 34.0°C (density class V) incubation temperature (Figure 1a). According to the Gaussian model, ambient temperature explained 20% (class I), 40% (II), 34% (III), 38% (IV) and 52% (V) of the measured respiration rate. Decomposition rate was highest for the intermediate classes (II–IV) reaching respiration rates of up to 2.4 g CO₂ kg⁻¹ day⁻¹. When temperature was included as covariate, there was a statistically significant difference in mean decomposition rates for density classes I and III, I and IV and IV and V (see Supporting Information S3).

Increased wood moisture significantly increased respiration at early stages of decomposition, whereas N content had the strongest impact on respiration rate for class II–IV wood (Table 2). The rate of N₂ fixation, on the other hand, did not correlate with respiration rate. No significant correlations were obtained for the respiration rate in comparison to δ¹³C, C% and nifH copy number of deadwood.

Density class II wood, which contains the highest amount of C (“C in deadwood” in Table 1), produced annually the largest amount of CO₂ whereas the contribution of class V was minor (Figure 1c). Despite the low amount of class IV wood in the study site relative to wood at early stages of decomposition (I–II) (Table 1), this decay class is a source of a significant annual release of CO₂ to the atmosphere due to its higher rate of respiration (Figure 1c). The total annual C flux from Lapinjärvi deadwood was calculated to be on average 566 kg C ha⁻¹ year⁻¹ for the 2 years (Figure 1c), which equals to 117 g C kg⁻¹ year⁻¹. The potential impact of climate warming on CO₂ production rates was calculated using the RCP4.5 temperature scenario (Ruosteenoja et al., 2013). The calculated increase in the annual respiration rate for Norway spruce deadwood was 28% (Figure 1c) assuming that the deadwood quantity and distribution to density classes is similar to current stock. The total biomass and C stock of Norway spruce deadwood in the study site is shown in Table 1. It indicates that in our study site the majority of C (71%) is
stored in wood that is at its early stages of decomposition (I–II) of which density class I (represents 34% of deadwood C stock) is least sensitive to temperature.

To produce the best possible estimates of annual respiration rates for deadwood at the study site, best-fit trendlines were separately defined by decay class for the CO$_2$ data covering the temperature range typically encountered in Finland (average day temperature <20°C) (Figure 1b). The best-fit trendlines (Figure 1b, CurveExpert 1.4) were a significantly better fit than the Gaussian trendlines of Figure 1a for the measured data at the temperature range 0–10°C. The annual respiration rate of deadwood was calculated for each density class using the trendlines of Figure 1b and the temperature data representing the Lapinjärvi forest (Figure 3a). The rate of respiration with advancing stage of decomposition is best explained by a quadratic fit trendline (Figure 3a, CurveExpert 1.4). Using the temporal $^{14}$C-dating based decomposition model established for our study site (figure 6a in Rinne et al., 2017), it was possible to define the dependence of respiration rate (g C kg$^{-1}$ year$^{-1}$) on the time of decomposition since soil contact (years) as shown in Supporting Information S4. This dependency (equation in Supporting Information S4) was then used to calculate the decomposition of the current biomass of deadwood (Table 1) as a function of time (Figure 3b). According to Figure 3b, after 60 years of decomposition (the time it takes for class I wood to decompose to class V wood), 24% of the current biomass of deadwood still remains on the forest floor of the study site. For class I wood, with the approximate age of 8 years (Rinne et al., 2017), wood content was calculated also for the preceding 7 years using the established decomposition model. Consequently, it was possible to determine the trendline that best describes wood decomposition during the first 69 years of decomposition (Sigmoidal model: 
\[ y = \frac{(ab + cx^d)}{(b + x^d)}, \quad r = 1.00 \]  "Wood decomposition model" in Figure 3b).

When the first 7 years of decomposition are excluded, the decomposition is well-described by negative exponential trendline (r = 1.00). Based on the Sigmoidal model, mass loss of 95% is reached after 116 years. The decomposition of Norway spruce as modelled in Mäkinen et al. (2006) is shown in Figure 3b for comparison (secondary y-axis: "remaining fraction"). It clearly differs from the decomposition pattern observed in the present study: (a) the decomposition rate in Mäkinen et al. (2006) is relatively slow for the first 20–30 years of decomposition (classes I–III/IV in this study), whereas in the present study decomposition rate was at maximum for class II–IV wood and (b) according to Mäkinen et al. (2006) logs disappear in much shorter time than can be expected based on the negative exponential decomposition of wood in the present study.

### 3.2 | Diazotrophic activity

The number of nifH copies was low in early stages of wood decomposition (I–II) but then significantly increased with advancing decomposition culminating at class IV (Figure 4, Table 1). The number of nifH copies peaked in communities associated with density class IV (Supporting Information S5). Wood moisture (Table 1) explained 52% ($p < 0.01$) of the nifH copy number for class I–IV wood. The inclusion of class V wood decreased the correlation, because of the opposite trends in nifH and moisture contents of wood (Table 1, Figure 4).

For the samples incubated at 14.5°C, which represents a typical summer temperature in the study region, (i.e. the temperature that was used for all analysis in the present study), N$_2$ fixation rate in classes I–IV was best explained (Table 2, $F_{3,34} = 9.850, p < 0.001$) by the combined impact of wood nifH gene number, moisture and N

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**TABLE 2** Correlation coefficients between CO$_2$ production, N$_2$ fixation rate and measured wood properties of deadwood. Only these wood property variables are shown that significantly correlated with CO$_2$ production or N$_2$ fixation rate are shown. "Class combination" shows the combination of density classes with the highest correlation coefficient. The sample number in correlation analysis is given. The significance levels are $p < 0.05$ (*) and $p < 0.001$ (**)

| Variables                        | Density class |     |     |     |     |     |     |
|----------------------------------|---------------|-----|-----|-----|-----|-----|-----|
|                                 | I (n = 10)    | II (n = 7) | III (n = 8) | IV (n = 13) | V (n = 11) | Class combination |
| CO$_2$ versus moisture$^a$        | 0.28          | 0.52  | −0.51          | −0.16          | 0.08          | 0.50$^*$ (I–II) |
| CO$_2$ versus N content$^a$      | −0.09         | 0.67  | 0.83$^*$        | 0.58$^*$        | 0.21          | 0.61$^*$ (II–IV) |
| CO$_2$ versus Shannon index      | −0.33         | −0.58 | 0.24            | −0.39          | −0.62$^*$     | −0.49$^*$ (IV–V) |
| N$_2$ fixation versus moisture   | 0.74$^*$      | 0.72  | 0.74$^*$        | 0.38            | 0.21          | 0.76$^*$ (I–III) |
| N$_2$ fixation versus nifH copies | 0.37          | 0.35  | 0.55            | 0.19            | 0.57$^*$      | 0.52$^*$ (I–IV) |
| N$_2$ fixation versus M2$^b$     | −0.06         | 0.64  | 0.64            | 0.52            | 0.53          | 0.72$^*$ (I–IV) |
| nifH copies versus moisture     | 0.27          | 0.01  | 0.18            | −0.10           | −0.65$^*$     | 0.68$^*$ (I–IV) |
| nifH copies versus N content    | −0.03         | 0.29  | 0.51            | 0.22            | 0.39          | 0.43$^*$ (I–V)  |

$^a$Moisture, N content and N$_2$ fixation rate data were obtained from Rinne et al. (2017). $^b$Multivariate analysis 2; independent variables: moisture, N content and nifH copy number.
content. The inclusion of class V into the analysis reduced the common signal, because moisture and N content increased in wood from class IV to class V unlike N₂ fixation rate and nifH gene number. The N₂ fixation rate of the 49 samples were significantly dependent on the incubation temperature (Rinne et al., 2017). The density class dependency of N₂ fixation (Figure 4) resembled the density class dependency of respiration (Figure 1a) with the exception of class II, where respiration—unlike N₂ fixation—was already significantly elevated compared to the level encountered at class I.

However, the respiration and N₂ fixation rates were significantly correlated only for class IV and V wood (for 8.7°C-25.1°C: $r_{IV} = 0.41$ and $r_{V} = 0.42, p < 0.01$). For every incubation temperature, the N₂ fixation rates followed a similar trend in absolute values with advancing decomposition as the number of nifH copies (Figure 4). The exception was the incubation temperature 40.3°C, which had a smaller replication.

### 3.3 | Activation energies

The highest incubation temperatures were excluded from AE calculations (Arrhenius, 1889) (Table 3), because at temperatures above 30°C and 26°C the rate of respiration and N₂ fixation, respectively, did not increase with temperature (Figures 1 and 2). There was a strong relationship between ln-transformed respiration and inverse temperature for every density class (Supporting Information S6). For respiration, AE was similar at all classes and agreed with the canonical value of 0.60–0.70 (Figure 5, Table 1) (Allen et al., 2005). AEs of N₂ fixation, on the other hand, were highest for class III (1.27 eV) and IV (1.04 eV) wood but the calculated AEs were always substantially less than the AE of the nitrogenase enzyme (2.18 eV) (Figure 5, Table 1) (Ceuterick et al., 1978).

### 3.4 | Relationship between fungi, respiration rate and diazotrophic activity

A detailed description of the obtained fungal OTUs, their phylotypes and functional groupings from this study can be found in Rinne et al. (2017) and in Supporting Information S2. A total of 393,071 sequence reads scattered in 69 fungal phylotypes, and of these 25% could be placed into functional fungal groups according to a literature review (see Rinne et al., 2017). Of the identified phylotypes 5% were white-rot, 3% were brown-rot and 12% were mycorrhizal fungi. Over 70% of the identified white-rot fungi sequences belonged to four species: *Resinicum bicolor, Phellolius nigrolimitatus, Exidia sp.* and *Phellinus viticola*. Almost 90% of the brown-rot fungal sequences belonged to *Antrodia serialis*, *Amylopodia sinuosa* and *Coniophora olivacea*, and over 50% of mycorrhizal fungal sequences to *Pseudotomentella tristis* and *Piloderma byssinum*. The number of fungal OTUs specific to density classes I, II, III, IV and V were 312, 104, 121, 381 and 396, respectively, and there were 482 OTUs common to all classes (Supporting Information S7). The most specific fungal phylotypes in a density class were not always the most dominant ones in a density class. For example, several typical mycorrhizal species (species of genera Cortinarius, Russula, Piloderma) specific to density class V were not among the most dominant ones (Supporting Information S8 and S9).

The fungal Shannon diversity index increased from class I to class V (Table 1). For all data, the index correlated with nifH copy number ($r = 0.43, p < 0.01$), moisture ($r = 0.57, p < 0.01$), density ($r = -0.53, p < 0.01$) and C% ($r = 0.37, p < 0.05$), but not with CO₂ production. The exception was a significant (negative) correlation between fungal diversity and respiration rate for class V wood (Table 2).
The relative abundance, as defined by the number of OTU reads, of white-rot, brown-rot and mycorrhizal fungi varied by density class (Figure 6). For white-rot fungi relative abundance decreased from class I to III and then peaked at class IV. The relative abundance of brown-rot fungi increased from class I (36%) to III (92%) and was relatively small in the samples of classes IV (15%) and V (13%). Mycorrhizal fungi were nearly absent in classes I–III (I: 0.0 ± 0.1%, II: 0.0 ± 0.2%, III: 0.1 ± 6%). Their relative abundance increased to 2.3 ± 26% in class IV and reached 27 ± 42% in class V.

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For brown-rot fungi, the relative abundance correlated negatively with \( \text{nifH} \) gene copy number \( (r = -0.32, p < 0.05) \) and positively with \( \text{CO}_2 \) production rate \( (r = 0.33, p < 0.05) \) (Table 4). The only abundance for individual species of brown-rot fungi that correlated with wood properties (such as \( \text{nifH} \) copy No. and \( \delta^{13} \text{C} \)) was \textit{Amyloporia sinuosa}, which significantly increased with respiration rate. However, at the species level the
abundance of the white-rot fungi *Resinicum bicolor* correlated positively with respiration rate. Although the total abundance of white-rot fungi and *nifH* copy number did not correlate significantly in general, there was a positive correlation for class IV wood (*r*~white-rotters vs. *nifH* = 0.58, *p* < 0.05). At the species level the abundance of white-rot fungi *Hyphodontia alutacea* correlated positively with *nifH* copy number. It should be noted, however, that these correlations have to be accepted with caution due to the DNA based approach taken in the analyses which might include fungi that are not active.

The abundance of mycorrhizal fungi did not correlate with respiration rate or diazotrophic activity but was correlated with wood C% and δ^13^C values (positive and negative correlation, respectively, Table 4). The only relative abundance of mycorrhizal species that correlated with wood properties was that of *Pseudotomentella tristis*. Its increasing abundance was positively correlated with wood C%.

### 3.5 Carbon content and δ^13^C of deadwood

The C content (C%) and the δ^13^C value of wood linearly increased and declined, respectively, with advancing wood decomposition with the exception of density class IV wood (Table 1, Supporting Information S3). For samples of class IV the C% values were lower (2.4%) and the δ^13^C values more δ^13^C-enriched (0.65%) than expected from the general trend (Figure 2).

### 4 DISCUSSION

#### 4.1 Temperature and wood property dependency of respiration

The significant temperature and wood density class dependency of respiration (Figure 1a) clearly shows that deadwood mass loss cannot be modelled assuming constant decomposition rates throughout the decomposition process. As with many other physiological processes, rates of fungal respiration increase with an increase in temperature until the heat denatures enzymes required for growth. The generally reported general temperature limits for fungal growth (0°C–45°C) as well as the more species specific optimum temperatures for growth (20°C–35°C) (Lacasse & Vanier, 1999) agree well with the temperature dependence of CO₂ production observed for all density classes in this study. Since the temperature optimum for fungi driven deadwood respiration is ~30°C (Figure 1a), climate warming will accelerate C loss from downed deadwood in boreal conditions in the future, unless moisture becomes limiting (too low or too high) (Figure 1c). The current mean annual C flux of 117 g C/kg measured for Norway spruce deadwood is similar to the flux reported in Forrester, Mladenoff, Gower, and Stoffel (2012) (109 and 128 g C kg⁻¹ year⁻¹), who studied coarse woody debris across decay classes (CWD) in a hardwood forest in Wisconsin, USA, using instantaneous CO₂ flux measurements, and it is 30% of the C flux reported for CWD of tropical forests (Chambers, Schimel, & Bnobre, 2001).

With projected climate warming the annual C flux in respiration of deadwood from the study site was calculated to increase to 149 g C/kg. However, it should be noted that this calculation does not take into account changes in total store of deadwood or in the distribution of density classes in the function of time. The current annual C flux (117 g C/kg) of deadwood accounts for 14% and 7% of the soil respiration and total ecosystem respiration (Korhonen, Pumpanen, Kolari, Juurola, & Nikinmaa, 2009), respectively. However, unlike our site in Lapinjärvi, the Scots pine dominated forest in Korhonen et al. (2009) is a managed forest, where deadwood content on forest floor is typically only <4 m²/ha. The forest in Korhonen et al. (2009) is located 170 km northwest from our study site.

Our results do not support the conclusion of Bradford et al. (2014); that climate is not a major driver of wood decomposition rates at local scales. In our study, ambient temperature explained ~40% of the variation in respiration rate of wood that had been decomposing longer than 8 years (classes II–V, figure 1a in Bradford et al., 2014) the conclusion of the relatively low impact of temperature on respiration rate (vs. % of fungal colonization of wood) at the local scale may have been hampered by the experimental setting, where temperature was relatively constant (range of ~2°C) so that the degree for the level of fungal colonization (~2%–52%) emerged as a controlling factor at each research site. However, even Bradford et al. (2014) clearly stated that since local-scale factors—such as the availability of nitrogen and soil moisture—have a significant impact of fungal activity (Table 2), care should be taken in assuming constant decomposition rates throughout the decomposition process.

### TABLE 4 Correlation coefficients between fungal abundance and deadwood properties. The *r*-values with 95% (*) and 99% (**) significance levels are shown for three functional groups (brown-rot [B], white-rot [W] and ectomycorrhizal [EC] fungi) and for those fungal species (total of ≥5,000 OTUs) that significantly correlated with the wood properties measured.

| Wood variable | Fungi          | Resinicum bicolor (W) | Amyloporia sinuosa (B) | Hyphodontia alutacea (W) | Phellopilus nigrolimitatus (W) | Pseudotomentella tristis (M) |
|---------------|---------------|------------------------|------------------------|--------------------------|-------------------------------|------------------------------|
| Respiration   | B: 0.33*      | −0.31*                 | −0.14                  | 0.28*                    | 0.55**                        | −0.26                        |
| N₂ fixation   | −0.21         | −0.19                  | −0.14                  | −0.08                    | −0.04                         | 0.07                         |
| nifH copy no. | −0.32*        | 0.03                   | −0.12                  | −0.11                    | 0.54**                        | −0.24                        |
| C%            | −0.04         | −0.32*                 | 0.38**                 | −0.16                    | −0.01                         | −0.02                        |
| δ^13^C        | −0.01         | 0.37*                  | −0.43**                | 0.28                     | −0.10                         | −0.17                        |

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excercised when taking local results for the response of deadwood respiration rates to changes in temperature, and scaling up for use in Earth-system models.

Nevertheless, our results on temperature sensitivity of CO₂ production (and N₂ fixation) by wood density class can be implemented into models describing respiration of deadwood logs. With least effort it can be done for models describing respiration class dynamics of individual logs, which are based on the similar deadwood classification system as we used here (Peltoniemi, Penttilä, & Mäkipää, 2013). However, density class representation is not strictly required as decay class can be associated with wood density with fair accuracy (e.g. Rajala et al., 2012; this study). It is noteworthy, however, that dynamic respiration models require tying the temperature to the state variable describing the present density of decomposing matter. For one compartment model this yields a rate of mass loss \( \frac{dy}{dt} = f(T, y) y \), where \( f(T, y) \) is some continuous function representing mass loss as a function of temperature \( T \) and density \( y \), which could be formulated based on our study and studies linking remaining mass (or density) to decay class. For more complex dynamic models, possibly representing several biochemically/physically identified compartments, implementation of aggregate temperature function requires further consideration.

Other wood properties driving respiration were increasing moisture content of wood in the early stages of decomposition (explaining 25% of respiration rate, Table 2), increasing wood N content (37%, Table 2) and the total abundance of brown-rot fungi (11%, Table 4), particularly abundance of the species Amyloporia sinuosa (30%, Table 4). The low level of moisture at early stages of decomposition probably explains why the temperature signal for respiration was lower for class I wood (20%, Figure 1a) than for the other decay classes (on an average 41%). Wood moisture was 35% or lower in 90% of wood samples in class I (Table 1). Wood moisture content this low has been reported to inhibit growth of fungi within wood cells (Lopez-Real & Swift, 1975). The lack of a direct dependency of respiration on wood moisture content for medium or well decomposed wood samples is in accordance with field studies, where moisture content was not limiting or excessive (Forrester et al., 2012; Herrmann & Bauhus, 2013). In contrast, in the study of Hagemann, Moroni, Gleißner, and Makeshin (2010), respiration was reduced at times by both too low and too high wood moisture content, therefore wood moisture was reported to be the dominant environmental control over CWD respiration. Similarly, for soil respiration, Reichstein et al. (2002) observed declining temperature sensitivity with increasing soil water deficit. In boreal forests with a climate similar to that of the Lapinjärvi forest, moisture may be a limiting factor only at the early stages of decomposition or for the standing dead trees (Table 2).

Our study does not support the results of Yang et al. (2016) who found that increasing fungal diversity reduced wood respiration rate. In contrast, both CO₂ production and Shannon index of fungal community increased from class I to class IV wood (Table 1). Although the respiration rate and fungal diversity both increased from class I to class IV, the two were not significantly correlated (Table 2) suggesting that high fungal diversity is not essential for enhancing respiration rate. The result is consistent with the findings of Valentin et al. (2014) who concluded that reduced biodiversity in the highly diverse fungal community of the late decay class wood did not influence rate of respiration. The divergent trend between CO₂ production rate (decrease) and fungal diversity (increase) from class IV to class V wood (Figure 1a) probably reflects the reduced wood resource quality affecting respiration rate, as opposed to any direct relationship between the two variables (Table 2). This conclusion is supported by the significant reduction in the relative abundance of saproxylic fungi in favour of mycorrhizal species from class IV to class V wood (Figure 6). Via deceleration in respiration rate, the reduced wood quality also had a negative indirect effect on N₂ fixation rate (Figure 4) by reducing the respiration rate and therefore reducing ATP production from the decomposition process that is needed as an energy source for N₂ fixation (Weißhaupt, Pritzkow, & Noll, 2011).

Our results on the density class dependency of respiration rate, where an initial slow respiration rate (density class I) was followed by rapid respiration (II–IV) and finally by a clear decline (V) at all incubation temperatures with high sample replication (8.7°C–34.0°C) (Figure 1), have clear differences compared to previous studies on conifers. Wang et al. (2002) and Wu, Zhang, Wang, Sun, and Guan (2010) studied black spruce (Picea mariana) and Korean pine (Pinus koraiensis), respectively, and reported an increase in respiration rate from the least to moderately decomposed wood but no decline for the most decomposed wood. However, their classification system for deadwood consisted of only three classes (Lambert, Lang, & Reiners, 1980). The class specific average wood densities of 0.41 g/cm³, 0.34 g/cm³ and 0.28 g/cm³ reported for black spruce (Wang et al., 2002) are similar to the densities measured for classes I, II and III of the five classes used in our study (Table 1), which suggests that well-decomposed wood was not used in the studies by Wang et al. (2002) and Wu et al. (2010). Such sampling would lead to different conclusions, when calculating models predicting mass loss due to respiration over time (Figure 3). Similarly, the study of Mäkinen et al. (2006), who modelled decomposition of Norway spruce in Finland, is based on samples that were on an average only 14 and at most 38 years old, which is significantly younger than the 62–77 years average age determined for well decomposed Norway spruce logs using ¹⁴C dating (Krüger, Muhr, Hartl-Meier, Schulz, & Borken, 2014; Petrillo et al., 2015; Rinne et al., 2017). These factors may explain why the decomposition model of Mäkinen et al. (2006) is very different from the decomposition pattern observed in the present study (Figure 3b). The negative exponential decomposition model, which has been used in many studies to estimate decomposition of Norway spruce (Melin et al. (2009), is in a much better agreement with the results of the present study. However, our study indicates a clearly slower overall rate of decomposition. For example, after 60 years of decomposition the fraction of the deadwood stock remaining on forest floor is 24% in the present study and on an average ~10% according to the compilation of models in Melin.
et al. (2009). This is at least partly explained by the failure of the exponential models to take into account the slow decomposition rate at the early stages of decomposition (Figures 1a and 3b). In addition, organic matter stabilization by fungi or other soil biota during the decomposition process (Chertov et al., 2017; Komarov et al., 2017) may explain slow decomposition rate of the remaining fraction in the late decay phases.

For density class II wood, respiration increased to levels comparable to those measured for class III and IV wood (Figure 1) despite the low accumulation rate of fixed atmospheric N (Figure 4), which is needed to facilitate fungal-driven decomposition. The high rate of respiration at this stage may have been enabled by soil foraging fungi that have the capability to transport soil N to wood for the production of extracellular enzymes (Allison & Vitousek, 2005; Sterner & Elser, 2002) and fungal material (Merrill & Cowling, 1966; Moore et al., 2008). 80% of the Lapinjärvi deadwood samples contained at least one species that has been reported to form mycelial cords or rhizomorphs for soil nutrient foraging (Rinne et al., 2017). The generally low N content measured for class II wood (Table 1) suggests that at this stage of decomposition there was a substantial need for N to sustain high fungal vegetative and generative growth (Hoppe et al., 2014).

### 4.2 | Activation energy

Due to the relatively low calculated AE of N₂ fixation of deadwood, the impact of a temperature increase on N₂ fixation rate of deadwood was smaller than expected from the temperature response of the nitrogenase enzyme (Figure 5). This is typical for high latitudes, because as temperatures decline more enzyme is needed to achieve a given rate of N₂ fixation, hence constraining maximal N₂ fixation rates (Houlton, Wang, Vitousek, & Field, 2008). However, the temperature dependency of this metabolic process was not constant for deadwood but varied according to wood supply and quality (Figure 5). For density classes I and II, low wood moisture (Table 1) reduced the temperature sensitivity of N₂ fixation (Table 2). Additionally, for class I and V wood, the low amount of ATP produced via the slow decomposition process (Figure 1) significantly limited N₂ fixation rate (Weißhaupt et al., 2011). As a consequence, the AE of N₂ fixation for class I, II and V wood (0.56 ± 0.12 eV) is significantly below the average AE of 1.1 eV reported in Houlton et al. (2008) in a unifying framework for terrestrial N₂ fixation. N₂ fixation of class III and IV wood, on the other hand, had a stronger response to increases in temperature (AE = 1.14 ± 0.18 eV). This is because wood at these stages of decomposition was not limited by wood quality and they benefited (Figure 4) from the increased production of ATP from the accelerated decomposition process (Figure 1).

The increases in N₂ fixation rates with temperature did not affect the temperature dependency of respiration at any stage of decomposition for Lapinjärvi deadwood (Figure 5). This is in contrast to the study of Welter et al. (2015), who reported 2.7-fold higher AEs of respiration than predicted by metabolic theory for biofilms in an Icelandic streamside channel due to increases in N₂ fixation with temperature. However, a major difference between our study (Figure 5) and that of Welter et al. (2015) is that AEs calculated for biofilm N₂ fixation rates approximated the AE of the nitrogenase reaction, but that was not the case for our study (Figure 5).

### 4.3 | Effect of fungal community on decomposition of cellulose and lignin

The general linear relationship between wood density and wood C% as well as between wood density and δ¹³C values mainly reflect the increasing relative abundance of lignin to cellulose (Figure 2). Cellulose, which has a lower C content (44% vs. 67%) and is from 4‰ to 6‰ more ¹³C-enriched than lignin (Benner, Fogel, Sprague, & Hodson, 1987), is degraded at a faster rate because it is a much more energy rich resource for wood decay fungi (Moorhead & Sinsabaugh, 2006). However, eventually the remaining cellulose becomes shielded by a lignin coating, which requires enhanced activity of white-rot fungi to enable decomposition of more cellulose (Moorhead & Sinsabaugh, 2006; Rayner & Boddy, 1988). In our study, this increase in lignin breakdown seems to have occurred at density class IV, where the wood C% and δ¹³C values are smaller and more ¹³C-enriched, respectively, than expected from the general linear trend that they follow relative to wood density (Figure 2). This is supported by the observation that the relative abundance of white-rot fungi substantially increases from density class III to class IV, where white-rot fungi dominates the three main functional groups (Figure 6). The significant role of white-rot fungi in lignin breakdown is also indicated by the negative correlation between the abundance of this functional group in deadwood and wood C% and by the positive correlation between the abundance and wood δ¹³C (Table 4). Since the white-rot species H. alutacea was specific and clearly more common to class IV (Supporting Information S8 and S9) and since its abundance correlated positively with nifH copy number (Table 4), it is concluded that this species had a major role in the intensified lignin decomposition at stage IV (Figure 2) that was enabled by diazotrophic N additions (Jurgensen, Larsen, Graham, & Harvey, 1987). Also, Hoppe et al. (2016) reported the dominance of H. alutacea in decay class IV spruce wood as well as a positive correlation between the abundance of this species and the respiration rate.

The significant breakdown of lignin at density class IV may have been enabled by the invasion of ECM fungi (Figure 6), which transfers simple sugars from trees to deadwood within their mycelia. The transfer of recent photosynthates with depleted δ¹³C value (Rinne et al., 2015) into deadwood is supported by the significant negative correlation between the relative abundance of ECM fungi and deadwood δ¹³C and by the positive correlation between ECM abundance and C% values (Table 4). Pseudotomentella tristis was a particularly important species importing labile C compounds into wood (Table 4). The invasion of wood by ECM was key factor in enabling lignin breakdown, because the consequent availability of labile C compounds in wood can function as co-metabolites in this process (Melillo et al., 1989). Furthermore, the increased availability of labile C compounds as well as the ATP generated during the faster lignin breakdown and the breakdown of the released cellulose would have benefited...
diazotrophic activity leading to increased availability of N for white-rot fungi (Burgess & Lowe, 1996; Jurgensen, Larsen, Wolosiewicz, & Harvey, 1989). This mutualistic relationship between white-rotters and diazotrophs is supported by the significant correlation between the abundance of the two in class IV wood (r_{white-rotters vs. diaz} = 0.58, p < 0.05), where both also reached their peak activity (Figures 4 and 5). The invasion of wood by ECM fungi, which increased the content of labile C compounds in wood (Mäkipää et al., 2017), at late stages of decomposition could also explain why white-rotted (further decomposed) wood has been reported to have a higher soluble sugar content than brown-rotted (less decomposed) wood in Jurgensen et al. (1989), as opposed to the hypothesised higher metabolic activity of white-rot fungi per se (Jurgensen et al., 1989). However, no data on the fungal composition of wood is available in Jurgensen et al. (1989) to evaluate this hypothesis.

In general, however, wood respiration rate was more dependent on brown-rot than on white-rot activity (Table 4). The most important species in this respect seems to be *Ameloporia sinuosa* (Table 4), which was the fifth most abundant fungi identified at species level in the studied wood samples (Supporting Information S9). The second most abundant species, the white-rot fungi *Resinicium bicolor*, was the only other identified species whose abundance correlated with the measured respiration rates. No significant correlations with respiration rate or any other measured variable were observed for *Antrodia serialis*, which was clearly the most abundant brown-rot species. As discussed by Hoppe et al. (2016), high species richness may not be associated with wood respiration rate, if a species invests more energy into competing with other species than on producing wood-degrading enzymes.

### 4.4 Projecting respiration and N₂ fixation rates with climate warming

According to our results, temperature was the main, overall control on the respiration rate of deadwood in the studied Norway spruce dominated forest in Finland. This highlights the importance of adequately accounting for the effect of decomposition of coarse woody debris, which globally contains 36–72 Pg of C (Cornwell et al., 2009), on C budget calculations in forest ecosystem and climate change models. Another factor that should be accounted for in such models is the quantity of wood at different stages of decomposition on the forest floor, since respiration rate is significantly dependent on the density class of deadwood. In Finland such data on quantity of deadwood according to decomposition stages is available via the National Forest Inventories (Ihalainen & Mäkelä, 2009). Consequently, it is possible to upscale CO₂ release from coarse woody debris to regional and national levels, when density class and temperature dependency of the respiration is known. However, since decomposition rate of deadwood is also significantly dependent on local-scale factors, such as N availability and soil moisture status (Table 2), the results of the present study should not be directly applied to other sites with significantly different environmental conditions. Another major outcome of our study was that the effect of temperature increase on N₂ fixation rate of deadwood was smaller than expected from the temperature response of nitrogenase enzyme and that the temperature dependency of this metabolic process was density class dependent, which was caused by the class specific differences in wood quality and in supply of limiting resources. The density class dependency of AE has significant consequences on projecting N₂ fixation rates for deadwood with climate warming. According to the RCP4.5 temperature scenario of Ruosteenoja et al. (2013) and the AE observed in the present study for deadwood, by year 2040 N₂ fixation will increase ~30% for class I, II and IV wood and ~60% for class III and IV wood, whereas the AE reported for terrestrial N₂ fixation by Houlton et al. (2008) would predict 158% and 50% increases, respectively.

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SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

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