UPLC-QTOF-MS-guided isolation of anti-COPD ginsenosides from wild ginseng†

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Four previously undescribed ginsenosides, along with five known analogues were isolated from wild ginseng by a UPLC-QTOF-MS-guided fractionation procedure. Their structures were elucidated on the basis of spectroscopic and spectrometric data (1D and 2D NMR, HR-ESI-MS). The isolated compounds could significantly inhibit the cigarette smoke extract (CSE)-induced inflammatory reaction in A549 cells. The HDAC2 pathway might be involved in the protective effect against the CSE-mediated inflammatory response in A549 cells.

1 Introduction

Panax ginseng has received much attention not only as a medicinal herb but also as a functional food.1 Ginseng has a wide range of biological activities such as anti-inflammatory, anti-oxidant and immunomodulatory functions. It is the traditional belief that wild ginseng is more medicinally efficacious and more valuable than cultivated ginseng. In the past, natural wild ginseng has been used to refer to the original ecological ginseng. But it is hard to find the natural wild ginseng because it is nearly extinct now. Currently, according to the “Identification and Grade Quality Standards of Wild Ginseng” (National Standards of People’s Republic of China, GB/T 18765-2015), wild ginseng refers to the ginseng cultivated in mountains and grown for more than 15 years in a natural environment without any human intervention. This kind of wild ginseng is a replacement for natural wild ginseng grown within the original ecological environment. It has been reported that there were differences in immunomodulating effects between wild ginseng and cultivated ginseng.2 In our previous study, it was also found that the wild ginseng (20 year-old mountain-cultivated ginseng) was much differentiated from cultivated ginseng. And based on UPLC-QTOF/MS, a total of 126 chemical compositions were tentatively identified or characterized from wild ginseng.3

Chronic Obstructive Pulmonary Disease (COPD), the fourth leading cause of death in the world currently, is usually caused by significant exposure to noxious gases or particles. The airflow limitation and persistent respiratory symptoms are the typical symptoms. Cigarette smokers have a higher prevalence of respiratory symptoms and higher COPD mortality rate, and the cigarette smoking is the leading environmental risk factor for COPD around the world.4 Along with the progressive lung inflammation, some pro-inflammatory mediators such as TNF-α, IL-1β and IL-6 participated in the occurrence and development of COPD.5,6 Ginsenosides are regarded as the main bioactive components in ginseng.7,8 Ginseng extract9 and monomer saponins such as Rg1 (ref. 11), Rh2 (ref. 12) and CK13 had been reported to significantly inhibit COPD-related inflammation in vivo and in vitro. Reduced HDAC2 activity and expression was found in COPD, resulting in amplification of the inflammatory response. Meanwhile, HDAC2 dysfunction is thought to play an important role in the development of corticosteroid resistance in COPD.14–16 Thus, increasing HDAC2 activity may be a promising strategy to overcome corticosteroid resistance in COPD. Existing treatments such as theophylline, nortriptyline, macrolides and selective phosphatidylinositol-3-kinase-δ inhibitors have been reported to increase HDAC2 activity effectively.17

As part of our continuing efforts to elucidate the chemical composition of wild ginseng furtherly and search for anti-COPD compounds from wild ginseng, a UPLC-QTOF-MS-guided fractionation procedure was performed to target ginsenosides from the anti-COPD fractions of wild ginseng, leading to the isolation of 4 previously unreported ginsenosides (1–4) and five known analogues (5–9). Herein, the isolation and structural elucidation of the isolated compounds as well as their anti-COPD activity on the CSE-stimulated A549 cells were discussed. The potential mechanism against the CSE stimulation was also preliminarily investigated in the paper.

† Electronic supplementary information (ESI) available: 1H NMR, 13C NMR, HMBC, HMQC, HR-ESI-MS of compound 1–9. See DOI: 10.1039/c9ra06635g

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2 Experimental

2.1 Plant materials

The 20 years-old mountain-cultivated gingsens were collected from cultivation area in Fusong, Jilin Province, the main source of ginseng in China. The ginseng was identified by our group according to Chinese Pharmacopoeia (2015 version). The voucher specimen (no. MCG170931) was deposited at the Research Center of Natural Drug, Jilin University, Changchun, China.

2.2 Apparatus and chemicals

The HR-ESI-MS spectra were performed on Waters Xevo G2-XS QTOF mass spectrometer (Waters, Milford, MA, USA). NMR spectra were measured on a Bruker Avance-600 spectrometer (Bruker, Karlsruhe, Germany) in DMSO-D6 operating at 600 MHz for 1H NMR, HMBC and HMQC, 150 MHz for 13C NMR, respectively. Tetramethylsilane (TMS) was used as internal standard. Column chromatography was performed with silica gel (200–300 mesh) purchased from Qingdao Ocean Chemical Group Co. Ltd (Qingdao, China). Thin-layer chromatography was conducted on silica gel G plates (Qingdao Marine Chemical Inc.). Semi-preparative HPLC with 1525 binary pump combined Waters 2998 UV detector (Waters Co., Milford, MA, USA.) and YMC C18 column (5 μm, 20 mm × 10 mm; YMC Co., Ltd., Japan) were also used to isolate the compounds. Methanol (MeOH) and acetonitrile (ACN) were HPLC grade (Fisher, USA). Purified water was purchased from YiBao Co, Ltd (Shenzhen, China). Other solvents used were analytical grade (Beijing Chemical Works, Beijing, China).

2.3 Extraction and isolation

The fresh wild ginseng (1.0 kg) were crushed and extracted with 95% EtOH for three times. The extract was combined and then concentrated under vacuum to obtain the concentrated aqueous solution (0.5 L), and successively partitioned with petroleum ether, EtOAc and n-butanol, to afford petroleum ether (29.8 g), EtOAc (35.2 g) and n-butanol (133.5 g) soluble extracts, respectively. The n-butanol extract (inhibition of the release of pro-inflammatory mediators in CSE-induced A549 cells 20.8 %, 2 mL min⁻¹, 6 mg, 0.0006%), compound 2 (ACN–H₂O: 19: 81, 2 mL min⁻¹, 19 min, 6 mg, 0.0006%), compound 3 (ACN–H₂O: 19: 81, 2 mL min⁻¹, 22 min, 3 mg, 0.0003%), compound 4 (ACN–H₂O: 21: 79, 2 mL min⁻¹, 5 min, 4 mg, 0.0003%) and compound 5 (ACN–H₂O: 21: 79, 2 mL min⁻¹, 10 min, 2 mg, 0.0002%) were also further separated by semi-preparative RP-HPLC with ACN/H₂O as mobile phase to obtain compound 6 (ACN–H₂O: 21: 79, 2 mL min⁻¹, 13 min, 3 mg, 0.0003%).

2.4 Spectroscopic data

2.4.1 Ginsenoside-Rm1 (1)

6-O-[α-L-rhamnopyranosyl(1→2)-β-D-glucopyranosyl]-20-O-β-D-glucopyranosyl-dammar-23,25-diene-3β,20S-tetraol. 

White amorphous powder; Libermann–Buchard reaction was positive; Melisch reaction was positive; suggesting the existence of triterpenoid structure; 1H and 13C NMR: see Table 1; HR-ESI-MS m/z 943.5270 [M – H]⁻ (calc. for C₄₈H₇₉O₁₈ is 943.5266); HR-MS, 1H and 13C NMR, HMBC spectra see in the ESI (Fig. S1-1–S1-5).

2.4.2 Ginsenoside-Rm2 (2)

6-O-[α-L-rhamnopyranosyl(1→2)-β-D-glucopyranosyl]-20-O-β-D-glucopyranosyl-dammar-22,23,25-diene-3β,20S-tetraol. 

White amorphous powder; Libermann–Buchard reaction was positive; Melisch reaction was positive; suggesting the existence of triterpenoid structure; 1H and 13C NMR: see Table 1; HR-ESI-MS m/z 943.5272 [M – H]⁻ (calc. for C₄₈H₇₉O₁₈ is 943.5266); HR-MS, 1H and 13C NMR, HMBC spectra see in the ESI (Fig. S2-1–S2-5).

2.4.3 Ginsenoside-Rm3 (3)

3-O-[β-D-glucopyranosyl(1→2)-β-D-glucopyranosyl]-3β,12β,20S,25-tetrahydroxydammar-23-ene. 

White amorphous powder; Libermann–Buchard reaction was positive; Molisch reaction was positive; suggesting the existence of triterpenoid structure; 1H and 13C NMR: see Table 1; HR-ESI-MS m/z 799.4897 [M – H]⁻ (calc. for C₃₂H₅₁O₁₆ is 799.4844); HR-MS, 1H and 13C NMR, HMBC spectra see in the ESI (Fig. S3-1–S3-5).

2.4.4 Ginsenoside-Rm4 (4)

6-O-[α-L-rhamnopyranosyl(1→2)-β-D-glucopyranosyl]-20-O-malonoyl-dammar-24-ene-3β,6α,12β,20S-tetraol. 

White amorphous powder; Libermann–Buchard reaction was positive; Molisch reaction was positive; suggesting the existence of triterpenoid structure; 1H and 13C NMR: see Table 1; HR-ESI-MS m/z 869.4890 [M – H]⁻ (calc. for C₃₃H₅₃O₁₆ is 869.4899); HR-MS, 1H and 13C NMR, HMBC spectra see in the ESI (Fig. S4-1–S4-5).

2.5 Compound 5–9

Compound 5–9. White amorphous powder; Libermann–Buchard reaction was positive; Molisch reaction was positive; suggesting the existence of triterpenoid structure; 1H and 13C NMR, spectra see in the ESI (Fig. S5-1–S9-2).

2.5 Bioassay

2.5.1 Preparation of cigarette smoke extract. In the present study, the cigarettes used were Xiongshi cigarette (China
Smoke from one cigarette was bubbled into 20 mL of culture medium (300 s per cigar). The CSE solution was regarded as the highest concentration (100%). This CSE solution was prepared freshly and was used within half an hour.
Table 2 $^1$H and $^{13}$C-NMR (600 MHz, 150 MHz in DMSO-D$_2$) data for 3

| Position | $\delta_C$ | $\delta_H$ |
|----------|------------|------------|
| 1        | 39.90      | 1.40(1H, m), 0.91(1H, m) |
| 2        | 25.79      | 2.25(1H, m), 1.55(1H, m) |
| 3        | 88.22      | 3.15(1H, m) |
| 4        | 39.60      | — |
| 5        | 55.61      | 0.69(1H, m) |
| 6        | 17.74      | 1.48(1H, m), 1.21(1H, m) |
| 7        | 34.42      | 1.36(1H, m), 1.33(1H, m) |
| 8        | 40.02      | — |
| 9        | 49.44      | 1.31(1H, m) |
| 10       | 36.29      | — |
| 11       | 30.98      | 1.92(1H, m), 1.63(1H, m) |
| 12       | 70.02      | 3.30(1H, m) |
| 13       | 48.53      | 1.92(1H, m) |
| 14       | 50.98      | — |
| 15       | 30.44      | 1.47(1H, m), 0.94(1H, m) |
| 16       | 25.79      | 1.58(1H, m), 1.36(1H, m) |
| 17       | 52.97      | 2.30(1H, m) |
| 18       | 15.37      | 0.82(3H, s) |
| 19       | 15.93      | 0.74(3H, s) |
| 20       | 72.29      | — |
| 21       | 27.48      | 1.02(3H, s) |

$^a$ Delta is ppm of chemical shifts which are reported in parts per million ($\delta$), and coupling constants ($J$) are expressed in hertz.

2.5.3 Drug treatment. For all group, A549 cells were cultured for 18 h in 96-well plates at a density of 5 x 10^5 cells per mL. In control group, A549 cells were cultured normally without the CSE stimulation. In CSE group, the cells were stimulated with a certain amount of CSE without any drug intervened. In ginsenoside groups, the cells were treated simultaneously with CSE and 10, 20, 40, and 80 $\mu$M of each ginsenoside.

2.5.4 Enzyme-linked immunosorbent assay. IL-1$\beta$, IL-6 and TNF- $\alpha$ contents in the cell culture supernatant were determined with ELISA kits (Nanjing Jiancheng Bioengineering Institute) after 18 h of incubation. All procedures were performed following the manufacture's instructions.

2.5.5 Western blotting. Western blotting was performed as previous described to discuss the effects of ginsenosides on HDAC2 expression in CSE-stimulated A549 cells. Both anti-HDAC2 (#ab32117) and anti- $\beta$-actin (#ab137550) were purchased from Abcam Company (Cambridge, UK). The HDAC2 band intensities were compared with reference to $\beta$-actin control.

2.5.6 Statistical analysis. Graphpad Prism 6.0 software (CA, USA) was used for all statistical analysis. The results are expressed as x ±SD. Statistical significance was calculated with two tailed test or a one-way analysis of variance (ANOVA), and p-value < 0.05 was considered as statistically significant.

2.5.7 Molecular docking assay. The GLIDE 6.7 software and Maestro Elements 2.2 software (Schrödinger, New York, NY, USA) were applied to perform molecular docking. Firstly, the 3D structures of ginsenosides were acquired respectively; secondly, the ginsenosides' bond angles and orders were then assigned using LigPrep module; thirdly, the 3D X-ray crystal structure of HDAC2 (PDB ID: 5IWG) was retrieved from Protein Data Bank (PDB) and was optimized according to the processes previously reported. Finally, the docking were performed with the following main parameters: the energy of conjugate of ligand and receptor was minimized to a root-mean-square deviation of 0.30 Å in optimized potentials for liquid simulations 3 force field, and the grid box at active site was set at the size of 20 Å. Finally, the highest XPG score pose was chosen as the dominant binding form and the visual analysis was performed by PyMol (Schrödinger) software.

3 Results and discussion

3.1 Structure elucidation

The 95% EtOH extract of wild ginseng was suspended in aqueous solution and partitioned with petroleum ether, EtOAc and n-butanol. UPLC-QTOF-MS-guided isolation of bioactive n-butanol soluble extracts (inhibition of the release of pro-inflammatory mediators in CSE-induced A549 cells at 200 $\mu$M) led to identify nine ginsenosides (Fig. S10A†). Five known compounds were identified as ginsenoside Rb2 (5), Rd (6), Rg3 (7), Rg1 (8) and Rh2 (9) by comparing spectroscopic data with those of reported values.
Compound 1 was a white amorphous powder (MeOH). The molecular formula was established as C₄₈H₈₀O₁₈ by combining NMR spectra and HR-ESI-MS at m/z 943.5270 [M – H]⁻ (calc. for 943.5266). The ¹H NMR spectrum of 1 displayed eight methyl signals at δ 0.81 (3H, s), 0.81 (3H, s), 0.82 (3H, s), 0.94 (3H, s), 1.22 (3H, s), 1.58 (3H, d, J = 6.5 Hz), 1.62 (3H, s) and 1.83 (3H, s). The ¹³C NMR spectrum of 1 showed 48 signals, including ten methylene [two of them bearing an oxygen atom respectively (δ 61.23, 61.47), and one of them was olefinic signal at δ 114.63], twenty-four methine [eighteen of them bearing an oxygen atom respectively (δ 67.84, 69.08, 69.28, 70.48, 70.82, 72.05, 73.65, 73.84, 73.84, 76.68, 76.78, 76.90, 77.22, 77.36, 77.51, 96.62, 99.91, 99.91 and 99.91), and two of them were olefinic signals at δ 127.28 and 134.62], six quaternary carbon [δ 38.59, 39.98, 40.16, 50.80, 82.02 and an olefinic signal at δ 141.72], eight methyl carbons [δ 16.65, 16.74, 16.74, 16.84, 17.79, 17.79, 22.37 and 30.97]. There is a resemblance with the chemical shifts of 1 along with ginsenoside Re²⁷ except the signals of the side chain. The main differences were the replacement of two olefinic carbons by four olefinic carbons [δ 141.72, 134.62, 127.28 and 114.63], and only two methyl signals [δ 22.37 and 17.79] in 1. Furthermore, comparison of the NMR spectroscopic data of 1 with those of cylindric tone A,²⁸ whose structure was 12β,20β-dihydroxy-23,25-diene-3-oxo-dammarane, a 23,25-diene group was deduced to exist in compound 1. The location of this group was confirmed by the heteronuclear multiple bond coherence (HMBC) spectrum of 1. The correlations between H-22 with C-21 and C-24, H-23 with C-20 and C-25, H-24 with C-22, C-26 and C-27, H-26 with C-24 and C-27, H-27 with C-24 and C-26, were observed in the HMBC spectrum of 1. The structure of 1 were deduced as 6-O-[α-l-rhamnopyranosyl-(1→2)-β-D-glucopyranosyl]-20-O-β-D-glucopyranosyl-dammar-23,25-diene-3β,6α,12β,20S-tetraol (ginsenoside Rm1, Fig. 1). Compound 1 was a minor glucoside in wild ginseng. The carbon and proton signals of 1 were fully assigned on the basis of HMOC and HMBC spectra (Table 1 and Fig. 2).
Compound 2 was a white amorphous powder (MeOH). The molecular formula was established as C_{48}H_{80}O_{18} by combining the NMR spectra and HR-ESI-MS at m/z 943.5272 [M − H]^− (calc. for 943.5266). The $^1$H NMR spectrum of 2 displayed nine methyl signals at $\delta$ 0.78 (3H, s), 0.78 (3H, s), 0.91 (3H, s), 0.99 (3H, s), 1.19 (3H, s), 1.60 (3H, d, $J = 6.5$ Hz), 1.91 (3H, s), 1.92 (3H, s) and 2.25 (3H, s). The $^{13}$C NMR spectrum of 2 revealed 48 signals, including eight methylene [two of them bearing an oxygen atom respectively ($\delta$ 67.99, 69.72, 70.32, 70.63, 70.96, 72.73, 72.73, 74.00, 77.37, 77.37, 77.37, 77.37, 77.66, 78.43, 96.77, 100.06 and 100.06)], and three of them were olefinic signals at $\delta$ 121.77, 127.43 and 141.31, six quaternary carbon [$\delta$ 39.97, 40.31, 38.73, 50.95, 82.17 and 88.22], and an olefinic signal at $\delta$ 134.77, nine methyl carbons ($\delta$ 16.80, 16.80, 16.89, 16.99, 17.94, 22.10, 22.52, 31.12 and 31.12). There is a resemblance with the chemical shifts of 2 along with ginsenoside Re$^{27}$ or compound 1 except the signals of the side chain. The main differences were the four olefinic carbons ($\delta$ 121.77, 127.43, 134.77 and 141.31), and three methyl signals ($\delta$ 22.10, 22.52 and 31.12) in 2. Furthermore, comparison of the NMR spectroscopic data of 2 with those of notoginsenoside ST14$^{29}$ whose structure was \(\{3\beta,6\alpha,12\beta,20S,22E\}-3,6,12,20\text{-tetrahydroxydammar-22(23)},24\text{-diene-6-O-}\beta\text{-d-glucopyranosyl, a 22(23),24-diene group was deduced to exist in compound 2. The location of this group was confirmed by HMBC of 2. In HMBC spectrum, the correlations between H-22 with C-21 and C-24, H-23 with C-20 and C-25, H-24 with C-22, C-26 and C-27, H-26 with C-24 and C-27, H-27 with C-24 and C-26, were observed. Thus, the structure of compound 2 was identified as 6-O-[\(\alpha\)-L-rhamnopyranosyl-(1→2)-\(\beta\)-D-glucopyranosyl-20-O-\(\beta\)-D-glucopyranosyl-\(\beta\)-dammar-22(23),24-diene-3\beta,6\alpha,12\beta,20S-tetraol (ginsenoside Rm2, Fig. 1). Compound 2 was also a minor glucoside in wild ginseng. The complete proton and carbon signal assignments were shown in Table 1.

Compound 3, a white amorphous powder (MeOH), possessed a molecular formula of C_{42}H_{72}O_{14}, as elucidated by combining the NMR spectra and HR-ESI-MS at m/z 799.4897 [M − H]$^-$ (calc. for 799.4844). The $^1$H NMR spectrum of 3 displayed eight methyl signals at $\delta$ 0.74 (3H, s), 0.81 (3H, s), 0.82 (3H, s), 0.90 (3H, s), 0.98 (3H, s), 1.02 (3H, s), 1.56 (3H, s) and 1.64 (3H, s). The $^{13}$C NMR spectrum of 3 revealed 42 signals, including ten methylene [two of them bearing an oxygen atom respectively ($\delta$ 60.89, 61.06), twenty methine [twelve of them bearing an oxygen atom respectively ($\delta$ 69.88, 70.02, 70.02, 75.30, 75.55, 76.16, 76.43, 76.88, 81.35, 88.22, 103.6 and 103.87], and two of them were olefinic signals at $\delta$ 126.20 and 136.16], six quaternary carbon [$\delta$ 36.29, 39.60, 40.02, 50.98, 72.29 and 80.34], eight methyl carbons ($\delta$ 15.37, 15.93, 15.99, 16.48, 27.48, 27.48, 29.94 and 29.96). The NMR data of 3 were similar to those of quinoneside A$^{28,29}$ whose structure was 3-O-[\(\beta\)-D-glucopyranosyl-(1→2)-\(\beta\)-D-glucopyranosyl-20-O-[\(\beta\)-D-glucopyranosyl-(1→6)-\(\beta\)-D-glucopyranosyl-\(\beta\)-dammar-23-ene, except the absence of \(\beta\)-D-glucopyranosyl-(1→6)-\(\beta\)-D-glucopyranosyl group signals in 3. And the chemical shift of C-20 was changed to $\delta$ 72.29, indicated that C-20 was linked to a free
hydroxyl group instead of being glycosylated. The location of hydroxyl group was determined by HMBC of 3. In HMBC spectrum, the correlations between H-22 with C-21 and C-24, H-23 with C-20 and C-25, were observed. Thus, the structure of compound 3 was identified as 3-O-[β-D-glucopyranosyl(1→2)-β-D-glucopyranosyl]-3β,12β,20S,25-tetrahydrodammar-23-ene (ginsenoside Rm3, Fig. 1). Compound 3 was also a minor glucoside in wild ginseng. The complete signal assignments on basis of HMQC and HMBC were shown in Table 1.
Compound 4, a white amorphous powder (MeOH), possessed a molecular formula of C₄₅H₇₄O₁₆, as elucidated by the NMR spectra and HR-ESI-MS at m/z 869.4890 [M – H]⁻ (calc. for 869.4899). The ¹H NMR spectrum of 4 displayed eight methyl signals at δ 0.82 (3H, s), 0.83 (3H, s), 0.83 (3H, s), 0.96 (3H, s), 0.97 (3H, s), 1.22 (3H, s), 1.34 (3H, d, J = 6.5 Hz), 1.56 (3H, s) and 1.63 (3H, d, J = 4.5 Hz). The ¹³C NMR spectrum of 4 revealed 45 signals, including ten methylene [one of them bearing an oxygen atom (δ 61.49)], eighteen methine, eight quaternary carbons [δ 38.63, 39.91, 40.16, 50.94, 75.24, 130.16, 170.90 and 171.20], eight methyl carbons [δ 16.46, 16.57, 16.74, 17.56, 17.80, 25.56, 26.38 and 31.00]. The NMR data of 4 were similar to those of ginsenoside Rg₂, except a malonyl group was assigned on the basis of HMQC and HMBC spectra (Table 1).

3.2 Bioactivity evaluation

Ginsenoside Rg₁ (ref. 11) and Rh₂ (ref. 12) had been reported to significantly inhibit COPD-related inflammation. As part of an ongoing effort to search for anti-COPD agents from wild ginseng, compounds 1–7 were evaluated for their anti-inflammatory effects on CSE-induced A549 cells. Dexamethasone was selected as a positive drug (Pos).

3.2.1 Cytotoxicity of cigarette smoke extract (CSE) and compounds 1–7 on the viability of A549 cells. The results of MTT showed that the cell viability of A549 cells was significantly affected (p < 0.01) by CSE (≥30%) (Fig. 3A). Therefore, in subsequent experiments, 20% CSE was chosen as stimulation. As shown in Fig. 3B, the cell viabilities of A549 cells were not significantly affected by 1–7 at 10 μM–80 μM. Then we evaluated the anti-inflammatory effects of 1–7 at 10 μM–80 μM on CSE-stimulated A549 cells.

3.2.2 Effect of compounds 1–7 on CSE-induced pro-inflammatory cytokine levels in A549 cells. The inflammatory development is characterized by the release of pro-inflammatory mediators such as tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1β) or interleukin-6 (IL-6), etc. Whether compounds 1–7 could inhibit the release of chemokine TNF-α, IL-1β and IL-6 from CSE-induced lung epithelial cells was investigated in this paper. As shown in Fig. 4, TNF-α, IL-1β and IL-6 production markedly increase in the cell treated with CSE (p < 0.01), however, treated with compounds 1–7 could significantly reduce the levels of pro-inflammatory factors to various extents. Compound 1 (Rm1) could significantly reduce TNF-α, IL-1β and IL-6 at 20 (p < 0.05), 40 and 80 (p < 0.01) μM. Compound 2 (Rm2) could significantly reduce IL-1β and IL-6 at 20 (p < 0.05), 40 and 80 (p < 0.01) μM, significantly reduce TNF-α at 20 and 80 (p < 0.05), 40 (p < 0.01) μM. Compound 3 (Rm3) and Compound 4 (Rm4) could significantly reduce IL-1β, IL-6 and TNF-α at 20 and 80 (p < 0.05), 40 (p < 0.01) μM.
Compound 5 (Rb2) could significantly reduce IL-1β and IL-6 at 10 and 20 ($p < 0.05$), 40 and 80 ($p < 0.01$) μM, reduce TNF-α at 20 ($p < 0.05$), 40 and 80 ($p < 0.01$) μM. Compound 6 (Rd) could significantly reduce IL-1β at 20 and 80 ($p < 0.05$), 40 ($p < 0.01$) μM, reduce IL-6 at 80 ($p < 0.05$) μM, while had no significant effect on TNF-α level. Compound 7 (Rg3) could significantly

Fig. 6 Target of compound 1–7 against CSE-stimulated inflammatory response in A549 cells. (A–G) Interaction mode of compound 1–7 within binding pocket of HDAC2. (A–G) The hydrogen bonds represented as the yellow dashed lines.
reduce IL-1β and TNF-α at 10 and 40 ($p < 0.05$), 20 ($p < 0.01$) μM, reduce IL-6 at 10 and 80 ($p < 0.05$), 20 and 40 ($p < 0.01$) μM. The lowest effective concentration of compound 1, 2, 3, 4, 6 was all 20 μM, the lowest effective concentration of compound 5, 7 was both 10 μM. Meanwhile, the dose-dependent effect of the compounds was appeared with a certain range of 10–80 μM. Interestingly, the activity of these compounds was not always increased with the increasing concentration, namely, when the concentration arrived a certain level, the activity might decrease to some extent.

3.2.3 Effect of compounds 1–7 on CSE-Mediated protein expression of HDAC2 in vitro. It has been demonstrated that histone deacetylase 2 (HDAC2) activity and egg self-expression in alveolar macrophages, peripheral lung tissues and bronchial tissues of COPD patients are significantly decreased.³³,³⁴ CSE can reduce HDAC2 activity and protein expression in rats,³⁵ which can activate rat lung tissue NF-κB pathway.³⁶ Other studies have shown that CSE exposure can cause the decrease of HDAC2 protein and activity in A549 cell nucleus³⁷ and activate the NF-κB pathway in vitro.³⁸

In order to explore the underlying mechanism of anti-inflammatory property of compounds 1–7, we investigated the effect of them on the protein expression of HDAC2 in CSE-exposed A549 cells. In the present study, after 18 hours of CSE exposure in A549 cells, the expression of HDAC2 protein in each group of 40 μM concentration was measured as shown in Fig. 5. The results showed that the level of HDAC2 protein in A549 cells decreased significantly ($p < 0.01$) after CSE 18 hours exposure. Interestingly, treatment with each ginsenoside could significantly activated the expression of HDAC2 ($p < 0.01$, $p < 0.05$) as compared to the CSE group (Fig. 5) in detail, which suggested that these compounds may up-regulated the expression of HDAC2 in CSE-exposed A549 cells.

3.2.4 Molecular docking study. The interactions between each ginsenoside and HDAC2 were visualized in molecular docking study. Ginsenoside Rm1 towards HDAC2 (PDB ID: 5IWG) gave six hydrogen bonds between Rm1 and six residues (ASN-354, ASN-280, SER-353, SER-351, PRO-352 and ASP-238) (Fig. 6A), Rm2 towards HDAC2 (PDB ID: 5IWG) displayed five hydrogen bonds between Rm2 and four residues (ASN-354, SER-351, ASN-312 and SER-272) (Fig. 6B), Rm3 towards HDAC2 (PDB ID: 5IWG) showed four hydrogen bonds between Rm3 and three residues (ASN-354, SER-351 and PRO-352) (Fig. 6C), Rm4 towards HDAC2 (PDB ID: 5IWG) exhibited four hydrogen bonds between Rm4 and two residues (THR-356 and ASN-354) (Fig. 6D). Rb2 towards HDAC2 (PDB ID: 5IWG) exhibited seven hydrogen bonds between Rb2 and seven residues (ARG-311, ASN-312, SER-351, GLY-273, GLY-277, THR-356, GLN-358) (Fig. 6E), Rd towards HDAC2 (PDB ID: 5IWG) exhibited one hydrogen bonds between Rd and one residues (THR-356) (Fig. 6F), Rg3 towards HDAC2 (PDB ID: 5IWG) exhibited two hydrogen bonds between Rg3 and two residues (SER-353 and ASN-354) (Fig. 6G). These results were consistent with the anti-inflammatory effects of the seven ginsenosides in CSE-induced inflammatory response in A549 cells. These docking results illustrated that the ginsenosides may directly bind to HDAC2 to exert lung protective effects due to strong hydrogen bonding effects.

4 Conclusions
In the present study, four minor previously undescribed ginsenosides, named as ginsenoside Rm1 (1), Rm2 (2), Rm3 (3) and Rm4 (4), along with five known ginsenosides, Rb2 (5), Rd (6), Rg3 (7), Rg1 (8) and Rh2 (9) were isolated from wild gingseng by a UPLC-QTOF-MS-guided fractionation procedure. Their structures were elucidated on the basis of spectroscopic and spectrometric data (1D and 2D NMR, IR, and HR-ESI-MS). The anti-COPD activities were investigated using CSE-stimulated A549 cells. Compounds 1–7 could ameliorate inflammatory reaction in vitro. Furthermore, the protective effect against CSE could be related to the HDAC2 pathway. The study provided some evidences to further elucidate the chemical composition of wild gingseng, and revealed that the compounds isolated from wild gingseng had the protective effect on the injury of cigarette smoke in vitro. The study provides a theoretical basis for the further utilization and development of the wild gingseng. It reminds us that it is meaningful to explore the protective effect of the wild gingseng against the COPD in vivo.

Conflicts of interest
The authors declare no conflicts of interest.

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