Association between novel adipocytokines adiponectin, vaspin, visfatin, and thyroid: An experimental and clinical update

Nese Cinar and Alper Gurlek

Department of Endocrinology and Metabolism, Hacettepe University School of Medicine, 06100 Sihhiye, Ankara, Turkey

Abstract

Adipose tissue secretes a variety of active biological substances, called adipocytokines, that act in an autocrine, paracrine, and endocrine manner. They have roles in appetite control, thermogenesis, and thyroid and reproductive functions. All these molecules may lead to local and generalized inflammation, mediating obesity-associated vascular disorders including hypertension, diabetes, atherosclerosis, and insulin resistance. Thyroid dysfunction is associated with changes in body weight, thermogenesis, and energy expenditure. The connections between cardiovascular risk factors such as dyslipidemia, impaired glucose tolerance, insulin resistance, atherosclerosis, and thyroid dysfunction have been reported in several studies. The adipocytokines serve as causative or protective factors in the development of these disorders in the states of thyroid dysfunction. Abnormal levels of adipocytokines (adiponectin (ADP), leptin, resistin, vaspin, and visfatin) in hypo- and hyperthyroidism have been reported with controversial results. This review aims to update the implication of novel adipokines ADP, vaspin, and visfatin in thyroid dysfunction.

Key Words

- thyroid
- metabolism

Introduction

Adipose tissue is a complex organ including adipocytes, immune cells, fibroblasts, blood vessels, and collagen fibers. It is classified as brown adipose tissue (BAT) and white adipose tissue (WAT). WAT is the predominant type because BAT regresses after birth. WAT serves as a storage site for energy in the form of triglycerides. Over the past decade, it has been recognized that adipose tissue has important functions other than energy storage, such as secreting a variety of endocrine, paracrine, and autocrine hormones; cytokines; and growth factors which influence local adipose tissue and different organs/tissues. These include CNS, liver, pancreas, and the skeletal muscles. Leptin, adiponectin (ADP), vaspin, visfatin, interleukin 6 (IL6), plasminogen activator inhibitor type 1, and resistin are among the adipocytokines secreted from the WAT. In obese humans, these adipocytokines play a role in the development of a cluster of metabolic abnormalities characterized by central obesity, dyslipidemia, type 2 diabetes, hypertension, and cardiovascular complications, by promoting a low-grade WAT inflammation (1, 2).

Thyroid hormones are involved in the regulation of body metabolism. Their effects include the stimulation of resting metabolic rate, increase in energy expenditure,
modulation of responsiveness to catecholamines, and thermogenesis in adipose tissue (3, 4). Disturbances in thyroid function lead to changes in body weight, muscle mass, and fat tissue. Thyroid-stimulating hormone (TSH) receptors have been found in the adipose tissues, indicating that they play a role in the regulation of the adipocytokines which are involved in the regulation of energy balance (5). This article focuses on the novel adipocytokines, namely ADP, vaspin, and visfatin in the context of thyroid dysfunction and the associated changes in adipose tissue and insulin resistance. The reason for choosing them is the increasing evidence regarding their changes in states of thyroid dysfunction.

### Adiponectin

ADP is a 244 amino acid protein that is the most abundant gene product of the adipose tissue, gene transcript 1 (apM1) (6). It accounts for 0.01% of total plasma proteins with plasma concentrations ranging from 5 to 30 mg/ml (7). Two different types of ADP receptors (AdipoR), 1 and 2, have been identified (8). AdipoR1 is expressed primarily in the muscle and AdipoR2 is expressed primarily in the liver. Binding of ADP to AdipoR1 and AdipoR2 results in increased glucose uptake and fatty acid oxidation in the skeletal muscle (AdipoR1) and decreased glucose output from the liver (AdipoR2) (9). ADP circulates in three forms, trimer, hexamer (also called low molecular weight oligomer), and high molecular weight (HMW) multimers. The biological effects of ADP depend on plasma concentrations, properties of different ADP isoforms, and tissue-specific expression of the ADP receptor subtypes. The most active form of ADP is the HMW (10).

ADP has anti-atherogenic, anti-diabetic, and anti-inflammatory properties. Obese patients have significantly lower ADP plasma levels than lean subjects (7), and a strong and consistent negative correlation exists between ADP levels and insulin resistance (11, 12). Low levels of ADP are associated with several diseases such as atherosclerosis, type 2 diabetes, abdominal obesity, hypertriglyceridemia, low HDL, hypertension, and metabolic syndrome (13). On the other hand, weight loss or insulin-sensitizing agents lead to increases in ADP levels in association with improved insulin resistance (11, 12).

ADP plays a role in the regulation of body temperature and basal metabolic rate. ADP has structural similarities with hibernation-associated plasma proteins HP-27, HP-25, and HP-20 in chipmunks, suggesting a role for it in adaptive thermogenesis (14). ADP may increase thyroid hormone synthesis, especially free thyroxine (fT₄), as a result of the C-terminal globular structure of ADP interacting with the gC1q receptor found in the mitochondria of thyroid cells (15). Given that thyroid hormones share some physiological actions with ADP (i.e. such as reduction of body fat by increased thermogenesis and lipid oxidation) (16), it is conceivable that ADP may interact with thyroid axis. Thyroid hormones are associated with insulin resistance (17), and a relationship between ADP and thyroid hormones may exist via either direct or indirect interactions between them.

Controversial results are reported for the experimental studies on hypo/hyperthyroid animals, concerning the association between ADP levels and thyroid hormones. Hypothyroid rats have either increased or unchanged serum ADP levels, while unchanged or increased serum ADP levels are found in hyperthyroid rats (18, 19). This might be due to the extent of thyroid hormone alterations in each study. The levels of ADP mRNA in the adipose tissue are decreased in hypothyroid rats compared with controls, and this decrease is in parallel with the decrease in triiodothyronine (T₃), T₄, fT₃, and fT₄ concentrations (20). In hyperthyroid rats, adipose ADP expression is increased in parallel with an increase in thyroid hormones, the opposite is observed in hypothyroid rats (20). On the other hand, Kokkinos et al. (18) demonstrated increased ADP levels in hypothyroid rats, whereas no significant change was observed in the ADP levels after the administration of thyroid hormone. Cabanelas et al. (21) showed that T₃ administration in rats had no significant effect on ADP secretion in visceral (epididymal) and subcutaneous (inguinal) adipose tissues, while ADP mRNA expression was downregulated by T₃ in the subcutaneous adipose tissue, but not in the visceral adipose tissue (VAT). The authors suggest that ADP mRNA response to T₃ changes depending on type and anatomical site of the VAT. In contrast, T₃ administration was shown to increase ADP mRNA expression and release in a culture of mouse brown adipocytes (22). Seifi et al. (23) demonstrated that methimazole administration resulted in a decrease in mRNA levels of adipor1 and adipor2 in VAT in hypothyroid rats, whereas mRNA levels of these ADP receptors are increased in the hyperthyroid group. ADP receptor gene expression levels in VAT had positive correlations with thyroid hormone concentrations, suggesting that AdipoR1 and AdipoR2 gene expression are regulated by thyroid hormones in hypo- and hyperthyroidism.

In humans, the results of studies of the interaction between thyroid hormones and ADP are conflicting.
There are few studies to date regarding the changes in the release of ADP in thyroid dysfunction. The results and essential information for these studies are summarized in Table 1.

In the studies mentioned earlier, the effect of thyroid hormones on the ADP levels revealed contrary results and no definite conclusion can be drawn. This might be due to differences in patients’ characteristics, degree and duration of thyroid hormone dysfunction, metabolic effects of other hormones, and possible effects of intermediate metabolism. Differences in the duration of the studies and small sample size included in the studies may be other possible causes of this inconsistency. Moreover, the presence of autoimmunity in thyroid disorders may play a role in this controversy, as an association between thyroid antibodies and ADP was shown in the studies.

Thyroid hormones are important regulators of lipid and carbohydrate metabolism. Thyroid dysfunction leads to dyslipidemia and impairment of glucose metabolism (43). The underlying mechanisms of these disturbances are still unknown. AdipoR1 and AdipoR2 were reported to play a role in glucose and lipid metabolism. The disruption of these receptors in the mouse liver results in increase in tissue triglyceride content, inflammation, and oxidative stress leading to insulin resistance and marked glucose intolerance (44). ADP (ADIPOQ) gene expression is negatively correlated with LDL-C and triglyceride levels in hypothyroid rats and positively correlated with glucose and HDL-C levels in hyperthyroid rats. Taken together, thyroid hormones may modulate lipid and carbohydrate metabolism via changes in ADP receptor expression in the adipose tissue. Hypothyroidism is associated with atherosclerosis, dyslipidemia, diastolic hypertension, impaired endothelial dysfunction, and insulin resistance (45). ADP precludes the development of atherosclerotic plaques in the injured vessel wall, which was reported in a recent study (46), which might indicate that decreased ADP expression in hypothyroidism has a role in the pathophysiology of atherosclerosis in these patients. The biological reason for increased plasma ADP levels in the adipose tissue in hyperthyroidism is unclear. Thyrtoxicosis is associated with reduction of fat and muscle mass and depletion in lipid storage. Insulin resistance in the liver and peripheral tissues is observed in hyperthyroid patients. Therefore, increased ADP levels might represent a compensatory mechanism against the insulin resistance observed in the hyperthyroid state. Further studies are needed to confirm all these hypotheses.

**Vaspin**

VAT-derived serine protease inhibitor (Vaspin) is a novel insulin-sensitizing adipocytokine identified from VAT of obese diabetic Otsuka Long-Evans Tokushima Fatty (OLETF) rats (47). Hida et al. (47) reported that administration of vaspin to OLETF rats results in improvement in glucose tolerance and insulin sensitivity. Recently, vaspin single-nucleotide polymorphism rs2236242 has been found to be positively associated with type 2 diabetes in 2759 participants in the KORA F3 study (48). The reported AA genotype (A represents the minor allele sequence) bears an increased risk of diabetes independent of obesity, suggesting a link between vaspin and glucose metabolism. El-Mesallamy et al. (49) found higher vaspin concentrations in both obese and non-obese type 2 diabetes patients than in controls, whereas diabetic women with good glycemic control had lower vaspin levels than those with poor glycemic control in another study (50). Also, vaspin expression was shown to increase from overweight to obesity (51). In obese children, serum vaspin levels are positively correlated with TG, fasting insulin, and homeostatic model assessment of insulin resistance (HOMA-IR) (52). The body fat percentage was found to be the strongest predictor of visceral vaspin, and insulin sensitivity seems to be the strongest determinant of subcutaneous vaspin expression (51). On the other hand, no significant association between vaspin levels and glucose tolerance or insulin sensitivity has been reported in a cross-sectional study including 83 non-diabetic subjects (53). It is, therefore, still unclear whether the role of vaspin is causative or protective in the development of obesity and metabolic disorders.

There are few studies concerning the regulation of vaspin by thyroid hormones. Gonzalez et al. (54) investigated vaspin mRNA, glucose, and insulin levels in hyperthyroid, hypothyroid, and euthyroid rats. They showed that vaspin mRNA levels are significantly down-regulated in hyperthyroid rats and significantly increased in hypothyroid rats compared with the euthyroid ones despite there being no change in glucose and insulin levels, suggesting that thyroid dysfunction may affect vaspin expression. Handisurya et al. (55) examined the relationship between TSH, vaspin, and leptin levels before and after weight loss by bariatric surgery. They reported a significant decrease in TSH levels in positive correlation with changes in serum vaspin levels. However, no definite conclusion can be drawn regarding whether vaspin causes the weight-loss-associated decrease of TSH levels, or whether changes in thyroid function or gastrointestinal
Table 1  Summary of previous studies of adiponectin and thyroid hormone association.

| Study | n       | Results                                                                 | Correlations                                                                 |
|-------|---------|-------------------------------------------------------------------------|-----------------------------------------------------------------------------|
|       |         |                                                                         | Positive ADP vs $fT_4$                                                     |
| Positive association |         |                                                                         | Positive ADP vs $fT_3$                                                     |
| (15)  | 68 healthy subjects | ↑ ADP in hyperthyroid patients                                         | Positive ADP vs $fT_4$                                                     |
| (24)  | 69 hyperthyroid patients | ↓ ADP after establishment of hypothyroidism (n: 32 patients)           | Positive ADP vs $fT_3$                                                     |
| (25)  | 32 hyperthyroid Graves' patients  
32 euthyroid Graves' patients  
30 controls | ↑ ADP levels in hyperthyroid than in euthyroid patients and controls | Positive ADP vs $fT_4$                                                     |
| (26)  | 46 hyperthyroid patients  
23 hypothyroid patients  
30 controls | ↑ ADP levels in hyperthyroid patients than controls                     | Positive ADP vs $fT_3$                                                     |
|        |         | ↔ ADP levels in hypothyroid patients                                     | Negative ADP vs insulin and HOMA-IR                                         |
| (27)  | 39 hyperthyroid patients  
23 controls | No significant difference in ADP levels between the groups               | Positive ADP vs $fT_4$                                                     |
| (28)  | 53 hypothyroid patients  
30 controls | ↓ ADP levels in hypothyroid patients and after normalization of thyroid status | Positive ADP vs WC and weight                                               |
| (29)  | 76 hyperthyroid Graves' patients  
(26 without GO and 50 with GO)  
30 controls | ↑ ADP levels in hyperthyroid patients than controls                     | Positive ADP vs $fT_4$                                                     |
|        |         | ↔ ADP between patients with GO vs without GO                            | Positive ADP vs $fT_3$                                                     |
|        |         | ↔ ADP between patients with active GO vs inactive GO                    | Positive ADP vs TRAb                                                       |
| (30)  | 120 hyperthyroid patients | ↓ ADP levels after normalization of thyroid status                      | Positive ADP vs $fT_4$                                                     |
| (31)  | 28 thyroid carcinoma patients | ↔ ADP 4 weeks after thyroid hormone withdrawal                          | Negative ADP vs BMI (BMI – the best predictor of ADP levels)               |
| (32)  | 234 euthyroid prepubertal children | Positive ADP vs $fT_4$                                                  | Positive HMW ADP vs $fT_4$                                                |
| (33)  | 321 healthy euthyroid pregnant women (24–28 weeks gestation)            | Negative HMW ADP vs $fT_4:fT_3$ ratio                                     |
| No association |         |                                                                         | Positive ADP vs HOMA-IR, glucose, and insulin                             |
| (34)  | 20 hyperthyroid patients | No significant difference in ADP levels among the groups                | ↔ ADP vs HOMA-IR, glucose, and insulin                                      |
| (35)  | 20 euthyroid subjects | No significant difference in ADP levels among the groups                | Positive ADP vs HDL                                                        |
|        | 15 hyperthyroid patients  
15 hypothyroid patients  
15 controls | No significant difference in ADP levels among the groups                | Negative ADP vs BMI                                                        |
| (36)  | 22 women with differentiated thyroid carcinoma | ↔ ADP with thyroid hormone withdrawal                                  | Positive ADP vs HDL                                                        |
| (37)  | 67 hypothyroid patients  
56 hyperthyroid patients  
52 controls | No significant difference in ADP levels among the groups                | Negative ADP vs BMI                                                        |
| (38)  | 98 euthyroid postmenopausal women with Hashimoto's thyroiditis  
105 postmenopausal controls  
19 hyperthyroid Graves' disease  
19 controls | No significant difference in ADP levels between the groups              | ↔ ADP vs TSH, $fT_4$, and TPO Abs                                           |
| (39)  | 19 hyperthyroid Graves' disease | No significant difference in ADP levels between the groups              | Positive ADP vs $fT_4$                                                     |
peptides influenced serum visfatin levels. In a recent study, we have evaluated serum visfatin levels in hypothyroid patients before treatment and after establishment of euthyroidism (56). Vasin levels were similar in euthyroid and hypothyroid subjects (subclinical and clinical hypothyroid), and no significant difference was observed in visfatin levels after normalization of thyroid hormones. Moreover, visfatin levels were not correlated with TSH. These data indicate that thyroid hormone status has no influence on serum visfatin levels in humans. More studies are needed to clarify the relationship between visfatin and thyroid hormones.

Visfatin

Visfatin, previously defined as pre-B cell colony-enhancing factor, is a 52-kDa cytokine expressed and secreted by lymphocytes (57). Visfatin is also called nicotinamide phosphoribosyltransferase (NAMPT) because of its functional and biochemical homology with NAD biosynthesized from nicotinamide (58). Fukuhara et al. (59) used the term ‘visfatin’ for this protein due to its predominant production in the VAT. Visfatin has insulin-mimicking/-sensitizing effects. Visfatin binds to the insulin receptor at a site distinct from insulin and exerts hypoglycemic effect by reducing glucose release from hepatocytes and stimulating glucose utilization in the peripheral tissues (60). Increased serum visfatin concentrations have been observed in type 2 diabetes, obesity, polycystic ovary syndrome, and nonalcoholic fatty liver disease. Weight loss after an exercise program and bariatric surgery results in a decrease in visfatin levels along with improvement in insulin sensitivity, indicating a compensatory mechanism in response to hyperglycemia associated with insulin resistance. Moreover, visfatin plays a role as an important mediator of inflammation, inducing the secretion of various proinflammatory and anti-inflammatory cytokines (61).

The relationship between visfatin and thyroid hormones was examined in a few studies. Experimental studies have revealed controversial results indicating that T3 could accelerate adipocyte differentiation with the elevation of visfatin levels (62), whereas MacLaren et al. (63) have reported downregulation of visfatin mRNA expression by T3 in 3T3-L1 adipocytes.

Chu et al. (64) evaluated the change in visfatin, C-reactive protein (CRP) concentration, and insulin sensitivity in 19 patients with hyperthyroidism due to Graves' disease and 19 age- and sex-matched controls. The hyperthyroid group had significantly higher plasma visfatin levels than controls, and visfatin levels significantly decreased after treatment. Visfatin, glucose, insulin, and HOMA-IR were values positively correlated with T3 and fT4 levels, but no significant association was found among visfatin levels and insulin and HOMA-IR values. The data suggest that the higher concentration of visfatin in the hyperthyroid patients may be related to a state of visfatin resistance, and that insulin resistance in hyperthyroidism is not associated with visfatin.

Caixas et al. (65) evaluated plasma visfatin, IL6, CRP, ADP, and insulin sensitivity parameters in 24 hyperthyroid and 27 hypothyroid patients before and after treatment in comparison with 45 euthyroid subjects. Hyperthyroid patients had significantly increased insulin resistance, IL6, and visfatin levels compared with controls. Visfatin levels increased after treatment, while IL6 levels and HOMA-IR decreased. CRP and ADP levels were similar in the hyperthyroid and control groups. Hypothyroid patients had higher visfatin levels compared with healthy subjects, which further increased after treatment without changes in anthropometric and insulin resistance.
parameters. No significant correlations between visfatin and any other parameters were found. It is suggested that visfatin might play a role in the recovery period independent of anthropometric, inflammatory, or insulin resistance parameters. Ozkaya et al. (66) studied the serum visfatin levels in 56 Hashimoto’s thyroiditis patients with hypothyroidism, 56 Graves’ patients with hyperthyroidism, and 56 euthyroid healthy subjects before and after treatment. Hyperthyroid patients had significantly lower visfatin levels compared with the hypothyroid group and controls. Plasma visfatin level decreased significantly after treatment in the hypothyroid group, whereas it increased significantly after treatment in the hyperthyroid group. A significant positive correlation between visfatin and TSH levels and a significant negative correlation between visfatin levels and fT3 and fT4 values were observed.

Han et al. (67) studied the regulation of visfatin by thyroid hormones in vivo and in vitro. The in vivo experiment included 57 patients with thyroid dysfunction and 29 euthyroid subjects and an animal model (24 Wistar rats). The in vitro experiment included 3T3-L1 cells and visfatin mRNA expression in the visceral fat and liver of rats under different T3 concentrations. Clinical subjects and animal models had elevated plasma visfatin concentrations in both hypo- and hyperthyroid groups compared with controls. For animal models, visfatin mRNA expression was found to be increased in only visceral fat but not liver in hypo- and hyperthyroid groups compared with controls, along with a positive correlation with plasma visfatin levels. T3 induced a remarkable increase in visfatin mRNA expression in 3T3-L1 cells at low concentrations followed by a sharp decrease at higher concentrations. The data indicate that thyroid dysfunction is associated with elevated visfatin levels, possibly due to an increase in visfatin mRNA expression in visceral fat, and that T3 caused a nonlinear regulation of visfatin mRNA expression.

Controversial results are reported concerning the role of thyroid hormones in the regulation of visfatin, with regard to the studies mentioned earlier. Such discrepancies might be explained by differences in ethnic or methodological factors or heterogeneity of thyroid dysfunction.

High levels of visfatin were observed in other autoimmune diseases such as rheumatoid arthritis or inflammatory bowel disease (61). Autoimmune thyroid dysfunction may be closely associated with fluctuations in visfatin levels. Proinflammatory cytokines such as IL6 and tumor necrosis factor α (TNFα) are increased in patients with thyroid dysfunction (68, 69), and IL6 has the ability to induce the expression of visfatin in vitro (70). Therefore, it is likely that visfatin release from adipose tissue may be affected directly or indirectly via proinflammatory cytokines implicating them in thyroid dysfunction. Visfatin is also expressed in the skeletal muscle (71). Both skeletal visfatin expression and plasma levels increase together with muscle mass growth, making it act as a myokine (71). Hyperthyroid patients usually lose lean body mass, which is supposed to be accompanied by decreased visfatin secretion. However, increased visfatin levels were also observed in hypothyroidism, while decreased levels were reported in hypothyroid patients. This might be due to the compensatory increase in visfatin levels against the high metabolic rate accelerating the breakdown of fat in hyperthyroidism.

In conclusion, the pathophysiological role of thyroid hormones in the regulation of ADP, vaspin, and visfatin is still unclear. Changes in adipokine secretion with thyroid dysfunction may represent adaptive mechanisms to the decrease or increase in basal energy expenditure and in energy substrate requirements in thyroid dysfunction. Cytokine network imbalances may be involved in the interactions between thyroid hormones and cytokines. Moreover, hyper- and hypothyroidism could affect the clearance of these cytokines. Additional studies, particularly studying the interactions among the novel genes in the adipose tissue, adipocytokines and the thyroid, will generate further insights into the endocrine function of adipose tissue.

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References
1 Pontikides N & Krassas GE. Basic endocrine products of adipose tissue in states of thyroid dysfunction. Thyroid 2007; 17 421–431. (doi:10.1089/thy.2007.0016)
2 Gulcelik NE, Usman A & Gurlek A. Role of adipocytokines in predicting the development of diabetes and its late complications. Endocrine 2009; 36 397–403. (doi:10.1007/s12020-009-9234-7)
3 Krotkiewski M. Thyroid hormones in the pathogenesis and treatment of obesity. European Journal of Pharmacology 2002; 440 85–98. (doi:10.1016/s0014-2999(02)01420-6)
Endocrine Connections

4 Lopez M, Varela I, Vazquez MJ, Rodriguez-Cuenca S, Gonzalez CR, Velagapudi VR, Morgan DA, Schoenmakers E, Agassandian K, Lage R et al. Hypothalamic AMPK and fatty acid metabolism in adipose tissue of normal and leptin receptor deficient mice. *International Journal of Obesity* 2010 34 117–125.

5 Endo T, Ohta K, Haraguchi K & Onaya T. Aromatase and cloning functional expression of a thylotropin receptor cDNA from rat fat cells. *Journal of Biological Chemistry* 1995 270 10833–10837.

6 Scherer PE, Williams S, Fogliano M, Baldini G & Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. *Journal of Biological Chemistry* 1995 270:26746–26749.

7 Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J, Hotta K, Shimomura I, Nakamura T, Miyaoaka K et al. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biological and Biochemical Research Communications* 1999 257:79–83.

8 Yamauchi T, Kamon J, Ito Y, Tsuchida A, Yokomizo T, Kita S, Shimomura I, Nakamura T, Miyaoka K et al. Novel sensitivity of adipocyte hormone expression to thyroid hormone withdrawal. *Endocrinology* 1995 131:79–83.

9 Berg AH, Combs TP & Scherer PE. ACRP30/adiponectin: an adipokine regulating glucose and lipid metabolism. *Journal of Biological Chemistry* 2002 277:25783–25790.

10 Sinha MK, Songer T, Xiao Q, Sloan JH, Wang J, Ji S, Alborn WE, Davis RA, Swarbrick MM, Stanhope KL et al. Analytical validation and biological evaluation of a high molecular-weight adipokine ELISA. *Clinical Chemistry* 2007 53:2144–2151.

11 Chandran M, Phillips SA, Ciarielli T & Henry RR. Adiponectin: more than just another fat cell hormone? *Diabetes Care* 2003 26:2442–2450.

12 Diez JJ & Iglesias P. The role of the novel adipocyte-derived hormone resistin in the regulation of energy homeostasis by thyroid hormone. *Biochemical and Biophysical Research Communications* 1999 257:79–83.

13 Brenta G. Why can insulin resistance be a natural consequence of obesity? *Brain Research* 2001 895:1–12.

14 Yaturu S, Prado S & Grimes SR. Changes in adipocyte hormones leptin, resistin, and adiponectin in thyroid dysfunction. *Journal of Cellular Biochemistry* 2004 94:491–496.

15 Saito T, Kawano T, Ikoma A, Namai K, Tamoto H, Kawakami M & Ishikawa SE. Elevation of serum adiponectin levels in Basedow disease. *Metabolism* 2005 54:1461–1466.

16 Yu H, Yang Y, Zhang M, Lu H, Zhang J, Wang H & Cianflone K. Thyroid status influence on adiponectin, acylation stimulating protein (ASP) and complement C3 in hypothyroid and hypothyroid subjects. *Nutrition & Metabolism* 2006 3:13.

17 Lousti K, Pontikides N, Koliakos G, Constantinidis Th, Papadopoulou F & Krassas GE. Serum cytokines profile in Graves’ disease hyperthyroidism before and after restoration of thyroid function: correlation with different anthropometric parameters. *Proceedings of the 31st European Thyroid Association Meeting* 2006. Naples 143, 196.

18 Pontikides N, Lousti K, Koliakos G, Constantinidis Th, Kaltas Th & Krassas GE. Serum cytokines levels in hyperthyroidism before and after treatment: relationship with body weight and body composition. *Proceedings of the 31st European Thyroid Association Meeting* 2006. Naples 144, 197.

19 Sieminska L, Niedziolka D, Billick A, Kos-Kudla B, Marek B, Nowak M & Borgiel-Marek H. Serum concentrations of adiponectin and resistin in hyperthyroid Graves’ disease patients. *Journal of Endocrinological Investigation* 2008 31:745–749.

20 Hsieh CJ & Wang PW. Serum concentrations of adiponectin in patients with hyperthyroidism before and after control of thyroid function. *Endocrine* 2008 85:489–494.

21 Prats-Puig A, Sitjar C, Ribot R, Calvo M, Clausell-Pomes N, Soler-Roca M, Soriano-Rodriguez P, Osnintr I, Ros-Miquel M, Bassols J et al. Relative hypoapoadiponectinemia, insulin resistance, and increased visceral fat in euthyroid prepubertal girls with low–normal serum free thyroxine. *Obesity* 2012 20:1455–1461.

22 Bassols J, Prats-Puig A, Soriano-Rodriguez P, Garcia-Gonzalez MM, Reid J, Martinez-Pascual M, Mateos-Comeron F, Zeher F, Blanazs I & Lopez-Bermejo A. Lower free thyroxin associates with a less favorable metabolic phenotype in healthy pregnant women. *Journal of Clinical Endocrinology and Metabolism* 2011 96:3717–3723.

23 Iglesias P, Alvarez Fidalgo P, Codocoro R & Diez JJ. Serum concentrations of adipokines in patients with hyperthyroidism and hypothyroidism before and after control of thyroid function. *Clinical Endocrinology* 2003 59:621–629.

24 Santini F, Marsili A, Mammoli C, Valeriano R, Scarabelli G, Pelosini C, Giannetti M, Centoni R, Vitti P & Pinchera A. Serum concentrations of adiponectin and leptin in patients with thyroid dysfunctions. *Journal of Endocrinological Investigation* 2004 27:RC5–RC7.

25 Botella-Carretero JI, Alvarez-Blasco F, Sancho J & Escobar-Morreale HF. Effects of thyroid hormones on serum levels of adipokines as studied in...
patients with differentiated thyroid carcinoma during thyroxine withdrawal. Thyroid 2006; 16: 397–402. (doi:10.1089/thy.2006.16.397)
37 Altinova AE, Toruner FB, Akturk M, Bukan N, Cakir N, Ayvaz G & Arslan M. Adiponectin levels and cardiovascular risk factors in hypothyroidism and hyperthyroidism. Clinical Endocrinology 2006; 65: 530–535. (doi:10.1111/j.1365-2265.2006.02628.x)
38 Sieminska L, Wojciechowska C, Kos-Kudla B, Marek B, Kajdaniuk D, Nowak M, Glogowska-Szelag J, Foityn W & Strzelczyk J. Serum concentrations of leptin, adiponectin, and interleukin-6 in postmenopausal women with Hashimoto’s thyroiditis. Endokrynologia Polska 2010; 61: 112–116.
39 Chu CH, Lam HC, Lee JK, Lu CC, Sun CC, Wang MC & Chuang MJ. Hyperthyroidism-associated insulin resistance is not mediated by adiponectin levels. Journal of Thyroid Research 2011; 2011: 194721. (doi:10.4061/2011/194721)
40 Eray E, Sari F, Ozdem S & Sari R. Relationship between thyroid volume and iodine, and adiponectin in obese women before and after weight loss. Medical Principles and Practice 2011; 20: 43–46. (doi:10.1159/000322075)
41 Kaplan O, Uzum AK, Aral H, Uzum G, Tunali Y, Demir O, Planzi KN, Kesmezacar O & Ozbek NC. Unchanged serum adiponectin concentrations in the setting of short-term thyroidectomy-induced hypothyroidism. Endocrine Practice 2012; 18: 887–893. (doi:10.4158/EP2011.08.03.08)
42 Yildiz BO, Aksoy DY, Harmanci A, Unluturk U, Cinar N, Ildisik M, Usman A & Bayraktar M. Effects of l-thyroxine therapy on circulating leptin and adiponectin levels in subclinical hypothyroidism: a prospective study. Archives of Medical Research 2013; 44: 317–320. (doi:10.1016/j.arcmed.2013.04.010)
43 Duntas LH. Thyroid disease and lipids. Thyroid 2002; 12: 287–293.
44 Tsuchida A, Yamauchi T, Takekawa S, Hada Y, Ito Y, Makita T & Kadowaki T. Peroxisome proliferator-activated receptor (PPARα) activation increases adiponectin receptor concentrations and reduces obesity-related inflammation in adipose tissue: comparison of activation of PPARγ, PPARα, and their combination. Diabetes 2005; 54: 3338–3370. (doi:10.2337/diabetes.54.12.3358)
45 Schultz M, Kristop C, Raymond J, Dimits J, Tuxen C, Hildebrandt P & Faber J. Cardiovascular events in thyroid disease: a population based, prospective study. Hormone and Metabolic Research 2011; 43: 653–659. (doi:10.1055/s-0031-1283162)
46 Kyriazi I, Tsotra PC, Bouati E, Ikonomidou I, Foutoulaki K, Maratou E, Lekakis J, Dimitriadis G, Kremastinos DT & Raptis SA. Effects of adiponectin in TNF-α, IL-6, and IL-10 cytokine production from human adipose tissue: comparison with obesity and type 2 diabetes. European Journal of Endocrinology 2011; 165: 537–544. (doi:10.1530/EJE-11-0737)
47 Hida K, Wada J, Eguchi J, Zhang H, Baba M, Seida A, Hashimoto I, Okada T, Yasuhara A, Nakatsuka A et al. Vaspin and its correlation with insulin sensitivity indices in obese children. Diabetes Research and Clinical Practice 2009; 84: 325–328. (doi:10.1016/j.diabres.2009.03.008)
48 von Loeffelholz C, Mohlig M, Arafat AM, Ilsen F, Spranger J, Mai K, Randeva HS, Pfeiffer AF & Weikert MO. Circulating vaspin is unrelated to insulin sensitivity in a cohort of non-diabetic humans. European Journal of Endocrinology 2009; 161: 507–513. (doi:10.1530/EJE-09-0773)
49 Gonzalez CR, Caminlos F, Vazquez MJ, Garces MF, Cepeda LA, Angel A, Gonzalez AC, Garcia-Rendueles MF, Sangiao-Alvaroilles S, Lopez M et al. Regulation of visceral adipose tissue-derived serine protease inhibitor by nutritional status, metformin, gender and pituitary factors in rat white adipose tissue. Journal of Physiology 2009; 587: 3741–3750. (doi:10.1113/jphysiol.2009.172510)
50 Handsunya S, Riedl M, Vila G, Maier C, Clodi M, Priskozovitzich T, Ludvik B, Prager G, Lugur & Kautzky-Willer A. Serum vaspin concentrations in relation to insulin sensitivity following RYGB-induced weight loss. Obesity Surgery 2010; 20: 198–203. (doi:10.1007/s11695-009-8882-y)
51 Cinar N, Gulcelik NE, Aydin K, Akin S, Usman A & Gurlek A. Serum vaspin levels in hyperthyroid patients. European Journal of Endocrinology 2011; 165: 563–569. (doi:10.1530/EJE-11-0180)
52 Samal B, Sun Y, Stearns G, Xie C, Suggs S & McNiece I. Cloning and characterization of the CINDA encoding a novel human pre-B-cell colony-enhancing factor. Molecular and Cellular Biology 1994; 14: 1431–1437. (doi:10.1128/MCB.14.1431)
53 Rongvaux A, Shea R, Mulks MH, Gigot D, Urbain J, Leo O & Andris F. Pre-B-cell colony-enhancing factor, whose expression is up-regulated in activated lymphocytes, is a nicotinamide phosphoribosyltransferase, a cytosolic enzyme involved in NAD biosynthesis. European Journal of Immunology 2002; 32: 3225–3234. (doi:10.1002/1521-4141(20021130)32:11<3225::AID-IJIM3225>3.0.CO;2-L)
54 Fukuhara A, Matsuda M, Nishizawa M, Segawa K, Tanaka M, Kishimoto K, Matsuji Y, Murakami M, Ichisaka T, Murakami H et al. Visfatin: a protein secreted by visceral fat that mimics the effects of insulin. Science 2005; 307: 432–436. (doi:10.1126/science.1107243)
55 Beltowskip. Apelin and visfatin: unique “beneficial” adipokines upregulated in obesity? Medical Science Monitor 2006; 12: RA112–RA119.
56 Moschen AR, Kaser A, Ehrlich B, Mosheizzer B, Theuri M, Niederegger H & Tilg H. Visfatin, an adipokine with proinflammatory and immunomodulating properties. Journal of Immunology 2007; 178: 1748–1758.
57 Tanaka M, Nozaki M, Fukuhara A, Segawa K, Aoki N, Matsuda M, Komuro R & Shimomura I. Visfatin is released from 3T3-L1 adipocytes via a non-classical pathway. Biochemical and Biophysical Research Communications 2007; 359: 194–201. (doi:10.1016/j.bbrc.2007.05.096)
58 MacLaren R, Cui W & Giandone K. Visfatin expression is hormonally regulated by metabolic and sex hormones in 3T3-L1 pre-adipocytes and adipocytes. Diabetes, Obesity & Metabolism 2007; 9: 490–497. (doi:10.1111/j.1463-1326.2006.00625.x)
59 Chu CH, Lee JK, Wang MC, Lu CC, Sun CC, Chuang MJ & Lam HC. Change of visfatin, C-reactive protein concentrations, and insulin sensitivity in patients with hyperthyroidism. Metabolism 2008; 57: 1380–1383. (doi:10.1016/j.metabol.2008.05.006)
60 Caiax A, Tirado R, Vendrell J, Gallart I, Megia A, Simon I, Llaurado G, Gonzalez-Clemente JM & Gimenez-Palop O. Plasma visfatin concentrations increase in both hyper and hypothyroid subjects after normalization of thyroid function and are not related to insulin resistance, anthropometric or inflammatory parameters. Clinical Endocrinology 2009; 71: 733–738. (doi:10.1111/j.1365-2265.2009.03546.x)
61 Ozkaya M, Sahin M, Cakal E, Yuzbasioglu F, Sezer K, Kilinc M & Imrek SS. Visfatin plasma concentrations in patients with hyperthyroidism and

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hypothyroidism before and after control of thyroid function. *Journal of Endocrinological Investigation* 2009 32 435–439. (doi:10.3275/6296)

67 Han J, Zhang TO, Xiao WH, Chang CQ & Ai H. Up-regulation of visfatin expression in subjects with hyperthyroidism and hypothyroidism is partially relevant to a nonlinear regulation mechanism between visfatin and tri-iodothyronine with various concentrations. *Chinese Medical Journal* 2012 125 874–881.

68 Celik I, Akalin S & Erbas T. Serum levels of interleukin 6 and tumor necrosis factor-α in hyperthyroid patients before and after propylthiouracil treatment. *European Journal of Endocrinology* 1995 132 668-672. (doi:10.1530/eje.0.1320668)

69 Salvi M, Pedrazzoni M, Girasole G, Giuliani N, Minelli R, Wall JR & Roti E. Serum concentrations of proinflammatory cytokines in Graves’ disease: effect of treatment, thyroid function, ophthalmopathy and cigarette smoking. *European Journal of Endocrinology* 2000 143 197–202. (doi:10.1530/eje.0.1430197)

70 Ognjanovic S, Bao S, Yamamoto SY, Garibay-Tupas J, Samal B & Bryant-Greenwood GD. Genomic organization of the gene coding for human pre-B-cell colony enhancing factor and expression in human fetal membranes. *Journal of Molecular Endocrinology* 2001 26 107–117. (doi:10.1677/jme.0.0260107)

71 Kreyzik-Walker SM, Ocon-Grove OM, Maddineni SR, Hendricks GL III & Ramachandran R. Is visfatin an adipokine or myokine? Evidence for greater visfatin expression in skeletal muscle than visceral fat in chickens. *Endocrinology* 2008 149 1543–1550. (doi:10.1210/en.2007-1301)

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