Study of the Association of Mutant HBsAg Gene and Hodgkin and Non-Hodgkin Lymphoma

Kamyar Makvandi,1 Nastaran Ranjbari,2,7 Manoochehr Makvandi,3 Ali Ashraf Teimori,3 Niloofar Neisi,3 Mojtaba Rasti,1 Vida Alipour,1 Mostafa Albokord,1 Malek Kanani,2 Ramezan Ahadi,4 and Ala Habibian3

1Infectious and Tropical Disease Research Center, Health Research Institute, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran
2Department of Pathology, Imam Khomeini hospital, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran
3Department of Virology, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran
4Department of Nuclear Medicine, Golestan Hospital, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran

*Corresponding author: Nastaran Ranjbari, Department of Pathology, Imam Khomeini hospital, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran. Tel: +98-613354389, Fax: +98-6113361544, E-mail: saminjon@yahoo.com

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Abstract

Background: Hepatitis B Virus (HBV) is responsible for chronic, acute, and fulminant hepatitis, which are prevalent worldwide. Chronic HBV may lead to cirrhosis and hepatocellular carcinoma. Several epidemiological studies have indicated that hepatitis B virus is involved in B-cell Hodgkin and Non-Hodgkin Lymphoma (NHL).

Objectives: The aim of this study was to evaluate the association between hepatitis B infection and Hodgkin and non-Hodgkin Lymphoma.

Materials and Methods: Paraffin embedded of 41 block samples including 12 (29.26%) Hodgkin and 29 (70.73%) non-Hodgkin patients were collected. Next, DNA extraction was carried out for all the samples followed by HBV DNA detection by the nested polymerase chain reaction (PCR). The positive HBV DNA samples were sequenced, and HBV genotypes and HBV subtypes were determined. The results of sequencing revealed that the D genotype was predominant among the positive HBV patients. Interestingly an unpredictable amino acid proline was detected in position 88 of the HBs gene, which indicates a new mutation in the “S” region of HBV DNA in patients with Hodgkin and non-Hodgkin lymphoma.

Conclusions: A high rate of 25% and 24.13% of HBV DNA was detected among patients with Hodgkin and non-Hodgkin lymphoma, respectively.

Keywords: Hepatitis B Virus, Hodgkin Disease, Polymerase Chain Reaction

1. Background

The association of Hepatitis B Virus (HBV) and hematologic malignancies, including Hodgkin lymphoma and Non-Hodgkin lymphoma (NHL) has been previously studied (1). Hepatitis B Virus causes a wide range of liver diseases, including subclinical infection, acute, chronic and fulminant hepatitis. Chronic HBV may lead to cirrhosis or Hepatocellular Carcinoma (HCC) (2). Hepatitis B Virus is a DNA virus, which belongs to the Hepadnaviridae family (3). The genome of HBV is a partially double stranded circular DNA with molecular weight of 3.2 Kbp. The viral genome encodes four open reading frames (ORFs: S, C, P and X). The ORF S encodes HBsAg, which consists of pre-S1, pre-S2 and S regions. The ORF core/pre-core encodes HBcAg, and soluble HBeAg. The ORF X encodes the X protein (HBX) and is a transactivator that influences the transcription of HBV genes by regulating the activity of transcriptional promoters and ORF P, which encodes DNA polymerase and reverse transcriptase (4).

The HBV is classified into eight genotypes, A to H (5). Genotypes A and D are prevalent in Europe, United States, and Africa, whereas genotypes B and C are prevalent in Asia. Genotype D is predominant in the Mediterranean, Middle East and India. Genotype E is the dominant genotype in West Africa whereas F and H have been isolated in the America (5). Based on amino acid substitutions in positions 122 and 160 of the S region of HBV DNA, four major subtypes: adw, adr, ayw and ayr, and nine minor subtypes have been described (6). Hepatitis B Virus is a hepatotropic virus but recently HBV DNA has been detected in Peripheral Blood Mononuclear Cell (PBMC) of patients with acute and chronic infection (7-9). Some studies have suggested that the HBV DNA could replicate in PBMC and proved this suggestion by detection of HBV mRNAs and expression of HBsAg and HBeAg in PBMCs (10-15).
These observations show that the lymphoid system could be an important reservoir for Hepatitis B Virus (16). The HBV DNA integration in PBMCs has been demonstrated previously (7, 17-19). Hodgkin lymphoma and non-Hodgkin lymphoma are lymphoid malignant tumors, and are prevalent worldwide. Recent studies have revealed the association between HBV infection and Hodgkin and non-Hodgkin lymphoma (20-22).

2. Objectives

The aim of this study was to determine the prevalence of HBV DNA and S region mutations among block samples of patients with Hodgkin and non-Hodgkin lymphoma in Ahvaz city, Iran.

3. Materials and Methods

Paraffin embedded tissues of 41 block samples including 12 (29.26%) Hodgkin and 29 (70.73%) non-Hodgkin patients were collected from Imam Khomeini hospital located in Ahvaz, Iran, during 2002 to 2011. The following steps were then carried out for the detection of HBV DNA in the patient samples.

3.1. Statistical Analysis

Statistical analysis was done with the Chi square test using the SPSS software, version 20.

3.2. DNA Extraction

Initially all the samples were deparaffinized with xylene. The DNA extraction of all the tissue samples was carried out using the high pure PCR template kit (Roche, Germany), according to the manufacturer's instructions.

3.3. Nested Polymerase Chain Reaction

The nested PCR for detection of HBV DNA was performed for all the tissue samples. The primers for partial sequencing of the “S” region are presented in Table 1 (23). For the first round, 5 µl of the extracted DNA from each sample was added to a PCR reaction mixture containing 0.5 µl dNTP (10 Mm), 2.5 µl PCR buffer (10x), 0.15 µl 5U Taq DNA polymerase (Roch, Germany), 50 pmol/µl of the FHBS1 and RHBS1 primers, 0.5 µl dNTP (10 Mm) and 15.85 µl distilled water. The samples were placed in the thermal cycler (Techne Company, UK) and the first round amplification was carried out with initial denaturation at 94°C for five minutes, followed by denaturation at 94°C for 30 seconds, annealing at 56°C for 30 seconds and extension at 72°C for 30 seconds, for a total of 30 cycles. For the second round, 5 µl of the PCR product was added to 25 µl of reaction mixture containing the same components mentioned in the first run including dNTP, PCR buffer and Taq DNA polymerase with 50 pmol/µl of each of FHBS2 and RHBS2 primers. Amplification was carried out in the thermal cycler with the same program as the first round. Next, 8 µl of the nested PCR product (417 bp) was analyzed by 2% agarose gel electrophoresis.

3.4. Sequencing Analysis

Amongst the ten samples positive for HBV DNA, six samples were sequenced (Applied BI Benier Company, South Korea). The sequences of all the six samples were submitted to NCBI and registered in GEN Bank (Table 2 and Figure 1). Hepatitis B Virus sequences were aligned with the HBV reference sequence from Gene Bank using the HBV data-base and online Blast. A phylogenetic tree was constructed using the Mega 5 software. To evaluate the HBs subtypes and S region mutations of the positive samples, (Figure 1) the amino acid substitutions in positions 122 and 160 of the S region of HBV DNA were compared with the HBV reference sequence. Interestingly, the presence of an unpredictable amino acid substitution (L88P) was observed, which indicated a new mutation in the “S” region of HBV DNA in patients with Hodgkin and non-Hodgkin lymphoma, (Figure 2).

Table 1. Sequences of Primer Pairs Used For Hepatitis B Virus Polymerase Chain Reaction Detection

| Primer Name | Nucleotide Sequence | Nucleotide Region |
|-------------|---------------------|------------------|
| FHBS1       | 5'-GAG TCT AGA CTC GTG GTG GAC TTC-3' | 244 - 267        |
| RHBS1       | 5'-CGT GGT GGA CTT CTC TCA ATT TTC-3' | 668 - 691        |
| FHBS2       | 5'-AAA TKG CAC TAG TAA ACT GAG CCA-3' | 255 - 278        |
| RHBS2       | 5'-GCC ARG AGA AAC GGR CTG AGG CCC-3' | 648 - 671        |

Table 2. The Profiles of Patients With Hepatitis B Virus Genotype Along With the Accession Numbers

| Isolate | Gender | Age | Hodgkin | Non-Hodgkin | Genotype | Blast Identity |
|---------|--------|-----|---------|-------------|----------|----------------|
| KJ98939 | F      | 25  | +       | –           | D1       | 98%            |
| KJ98940 | M      | 29  | +       | –           | D1       | 98%            |
| KJ98941 | M      | 54  | –       | +           | D1       | 98%            |
| KJ98942 | M      | 65  | –       | +           | D1       | 98%            |
| KJ98943 | F      | 46  | –       | +           | D1       | 98%            |
| KJ98944 | M      | 53  | –       | +           | D1       | 98%            |
4. Results

Out of 12 Hodgkin samples, seven (58.33%) and five (41.66%) were males and females, respectively. The age of the patients with Hodgkin lymphoma was between 5 and 34 with a mean age of 20 ± 7 years. Of the 29 Non-Hodgkin samples, 17 (58.62%) and 12 (41.37%) were males and females, respectively, Table 3. Out of the 41 samples, three out of twelve (25%) Hodgkin samples and seven of twenty-nine (24.13%) non-Hodgkin were positive for HBV DNA, Table 4. All the positive HBV patients were within the normal range for Alanine aminotransferase (20 - 32 IU/DL) and Aspartate aminotransferase (18-27 IU/DL). The youngest Hodgkin patient was a female with 17 years of age and the oldest was a male with 29 years of age. The youngest non-Hodgkin patient with HBV infection was a male with two years of age and the oldest patient was a female with 75 years of age.

4.1. Results of Sequencing

The sequences of six positive samples (two Hodgkin and four non-Hodgkin lymphoma) were registered in GenBank with the following accession numbers, KJ398939.1 to KJ398944.1. The results of the alignment showed that the D genotype was the predominant genotype among the patients with Hepatitis B infection. Mutation analysis indicated amino acid substitutions in positions 88, 122 and 160 of the "S" region of HBV DNA. All of the six sequences were recognized as the "awy" subtype because of substitution in positions 122 and 160.

Figure 1. A Phylogenetic Tree Constructed With the Neighbor Joining Method and 500 Replication Bootstrap Test Using the Partial Nucleotide Sequences of the S Region of Hepatitis B Virus, Genotype D1

Reference sequences were retrieved from Gen Bank with their accession numbers and origin. Bootstrap values are indicated in the tree roots.

Figure 2. Comparison of the Predicted Amino Acid Sequences of the S Region With Proteins of Accession Number AHW85404.1 to AHW85399.1 Recorded in Gen Bank

The consensus amino acid sequence of the reference D genotype is shown on the top. The partial sequences of all the six HBs genes in position 88 compared with the consensus sequence amino acid indicated a proline substitution with leucine. So far this type of mutation has not been reported in any HBV D genotype. The dots show conserved amino acid and the amino acid code indicates differences with the consensus amino acid.
### Table 3. The Frequency of Hodgkin and Non-Hodgkin Lymphoma

| Age Group, y | Males Amongst Hodgkin Patients | Females Amongst Hodgkin Patients | Males Amongst Non-Hodgkin Patients | Females Amongst Non-Hodgkin Patients | Total |
|-------------|--------------------------------|----------------------------------|-----------------------------------|-------------------------------------|-------|
| < 9         | -                              | 1                                | 7                                 | -                                   | 8     |
| 10 - 19     | 1                              | 3                                | -                                 | 2                                   | 6     |
| 20 - 29     | 5                              | 1                                | -                                 | 2                                   | 8     |
| 30 - 39     | 1                              | -                                | -                                 | 1                                   | 2     |
| 40 - 49     | -                              | -                                | 1                                 | 1                                   | 2     |
| 50 - 59     | -                              | -                                | 3                                 | 2                                   | 5     |
| 60 - 69     | -                              | -                                | 3                                 | 1                                   | 4     |
| > 70        | -                              | -                                | 3                                 | 3                                   | 6     |
| Total       | 7                              | 5                                | 17                                | 12                                  | 41    |

### Table 4. Distributions of Hepatitis B Virus Infection Amongst the Hodgkin and Non-Hodgkin Patients

| Positive/Negative Cases | Males Amongst Hodgkin Patients | Females Amongst Hodgkin Patients | Males Amongst Non-Hodgkin Patients | Females Amongst Non-Hodgkin Patients | Total |
|-------------------------|--------------------------------|----------------------------------|-----------------------------------|-------------------------------------|-------|
| HBV positive            | 2 (16.66)                      | 1 (8.33)                         | 5 (17.24)                        | 2 (6.89)                            | 10    |
| HBV negative            | 6 (50)                         | 3 (25)                           | 12 (41.37)                       | 10 (34.48)                          | 31    |
| Pos/total               | 3/12                           | 7/29                             | 10/41                            |                                     |       |
| Neg/Total               | 9/12                           | 22/29                            | 31/41                            |                                     |       |

aData are presented as No. (%).

### 5. Discussion

Several epidemiological studies have revealed the persistence of Hepatitis B Virus (HBV) infection amongst patients with Hodgkin (HL) and Non-Hodgkin Lymphoma (NHL). In our study, 25% of Hodgkin and 24.13% of non-Hodgkin patients had positive HBV DNA, which indicates the high rate of HBV infection among these patients. The youngest Hodgkin patient with HBV infection was a female with 17 years of age and the oldest was a male with 29 years of age. The youngest non-Hodgkin patient with HBV infection was a male with two years of age and the oldest was a female with 75 years of age. The frequency of HBV infection among the male and female patients was not significantly different (P > 0.05). In our study all the HBV patients had the ayw subtype.

Distribution of HBsAg subtype has been reported in different regions of the world. Fernández et al. in Spain reported the ad and ay HBs subtypes amongst 40 chronic HBV patients (24). De Souza et al. from Brazil reported HBsAg subtypes adw2, adw4, and ayw amongst patients with mental problems (25). In our study the protein sequence, of all the six “S” genes, showed mutation at position 88 (L88P). So far this type of mutation has not been reported in any HBV D genotype. In the study conducted by Okamoto et al. a point mutation was found in the S gene of hepatitis B virus for two blood donors carrying a surface antigen with subtype adyr or adwr (26). All of the HBV patients had normal SGPT and SGOT tests. The patient’s history revealed that they were not tested for HBV and hepatitis C virus and it is not clear whether these patients had chronic HBV or occult HBV infection. However, there are some reports about the association of occult HBV infection and non-Hodgkin lymphoma (27, 28).

The presence of unpredictable amino acid proline at position 88 of HBs gene indicates a new mutation in the “S” region of HBV DNA in patients with Hodgkin and non-Hodgkin lymphoma. In our survey the D1 genotype was dominant in all the HBV patients, which is similar to the report of Marcucci et al. (29). In the present study the role of other viruses including hepatitis C virus in Hodgkin and non-Hodgkin lymphoma was not studied however this requires further investigations (30, 31). There have been reports about the reactivation of Hepatitis B and Hepatitis C virus (HCV) infections during NHL treatment; in these studies it was revealed that the patients who were under chemotherapy showed HBV and HCV reactivation, yet other patients who received lamivudine before chemotherapy didn’t show any reactivation for HBV or HCV infection (32-35). In conclusion, a high prevalence of HBV infection (25% and 24.13%, respectively) was found in patients with Hodgkin and non-Hodgkin lymphoma, indicating the importance of HBV and HCV screening by sensitive tests such as PCR be-
fore chemotherapy to prevent HBV or HCV reactivation in patients with HL and NHL.

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Footnotes

Authors’ Contribution: Nastaran Ranjbari was responsible for the accuracy of the data and contributed to the design and performance of the study. Manoochehr Makvandi and Niloofar Neisi participated in the laboratory evaluation and performed the literature review. Manoochehr Makvandi drafted the manuscript and was the guarantor. All authors read and approved the final manuscript.

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References

1. Marucci F, Mele A. Hepatitis viruses and non-Hodgkin lymphoma: epidemiology, mechanisms of tumorigenesis, and therapeutic opportunities. Blood. 2011;117(6):1792–8. doi: 10.1182/blood-2010-06-275888. [PubMed: 20959600]
2. McMahon BJ. The natural history of chronic hepatitis B virus infection. Hepatology. 2009;49(5 Suppl):S45–55. doi: 10.1002/hep.22898. [PubMed: 19399792]
3. Schaffer DF, Sorrell MF. Hepatocellular carcinoma. Lancet. 1999;353(9160):2523–7. doi: 10.1016/s0140-6736(99)01948-x. [PubMed: 10210708]
4. Rossner MT. Review: hepatitis B virus X-gene product: a promiscuous transcriptional activator. J Med Virol. 1992;36(2):103–17. [PubMed: 1581465]
5. Khaled IAA, Mahmoud OM, Saleh AF, Baitoui EA. Prevalence of HBV genotypes in Egypt among hepatitis patients. J Am Sci. 2010;6(2):185–90.
6. Courouce-Pauty AM, Plançon A, Soulier JP. Distribution of HBV-Sag Subtypes in the World. Vox Sanginis. 1983;44(4):197–21. doi: 10.1111/j.1423-0410.1983.tb0885. x. [PubMed: 6845678]
7. Bouffard P, Lamelin JP, Zoulim F, Lepot D, Trepo C. Phothonemagglutinin and concanavalin A activate hepatitis B virus in peripheral blood mononuclear cells of patients with chronic hepatitis B virus infection. J Med Virol. 1992;37(4):255–62. [PubMed: 1402824]
8. Calmus Y, Marcellin P, Beaurain G, Chatenoud L, Brechot C. Distribution of hepatitis B virus DNA sequences in different peripheral blood mononuclear cell subsets in HBs antigen-positive and negative patients. Eur J Clin Invest. 1994;24(8):548–52. [PubMed: 7982442]
9. Malave Lara C, Gorrino MT, Campelo C, Lardelli P, Cisterna R. Detection of hepatitis B virus DNA and determination of surface antigen expression in peripheral blood mononuclear cells from patients with AIDS. J Clin Microbiol Infect Dis. 1994;13(3):267–71. [PubMed: 8050444]
10. Yoffe B, Noonan CA, Melnick JL, Hollinger FB. Hepatitis B virus DNA in mononuclear cells and analysis of cell subsets for the presence of replicative intermediates of viral DNA. J Infect Dis. 1986;153(3):471–7. [PubMed: 3005421]
11. Baginski I, Chemin I, Bouffard P, Hantz O, Trepo C. Detection of polyadenylated RNA in hepatitis B virus-infected peripheral blood mononuclear cells by polymerase chain reaction. J Infect Dis. 1991;163(5):996–1000. [PubMed: 1708401]
12. Roisman FR, Castello A, Finebohim H, Morelli A, Finebohim L. Hepatitis B virus antigens in peripheral blood mononuclear cells during the course of viral infection. Clin Immunol Immunopathol. 1994;70(2):259–103. [PubMed: 8292925]
13. Stoll-Becker S, Repp R, Glebe D, Schaefer S, Kreuder J, Kann M, et al. Transcription of hepatitis B virus in peripheral blood mononuclear cells from persistently infected patients. J Viral. 1995;77(3):539–47. [PubMed: 818861]
14. Feray C, Zignego AL, Samuel D, Bismuth A, Reyne M, Tiollais P, et al. Persistent hepatitis B virus infection of mononuclear blood cells without concomitant liver infection. The liver transplantation model. Transplantation. 1990;49(6):3155–8. [PubMed: 2360235]
15. Mason A, Yoffe B, Noonan C, Mearns M, Campbell C, Kelley A, et al. Hepatitis B virus DNA in peripheral-blood mononuclear cells in chronic hepatitis B after HBsAg clearance. Hepatology. 1992;16(6):136–41. [PubMed: 1681481]
16. Pontisso P, Vidalino L, Quarta S, Gatta A. Biological and clinical implications of HBV infection in peripheral blood mononuclear cells. Autoimmun Rev. 2003;2(3):123–7. doi: 10.1016/j.autrev.2003.07.016. [PubMed: 12706529]
17. Murakami Y, Minami M, Daimon Y, Okamune T. Hepatitis B virus DNA in liver, serum, and peripheral blood mononuclear cells after the clearance of serum hepatitis B virus surface antigen. J Med Virol. 2004;72(1):361–4. doi: 10.1002/jmv.10465. [PubMed: 1469566]
18. Umeda M, Maruasa H, Seno H, Katsurada A, Nabeshima M, Egawa H, et al. Hepatitis B virus infection in lymphatic tissues in inactive hepatitis B carriers. J Hepatol. 2005;42(6):306–12. doi: 10.1016/j.jhep.2005.01.016. [PubMed: 15888550]
19. Pontisso P, Poone MC, Tiollais P, Brechot C. Detection of hepatitis B virus DNA in mononuclear blood cells. BMJ. 1984;288(4430):756–6. doi: 10.1136/bmj.288.6430.1561. [PubMed: 6426645]
20. Kang J, Cho JH, Suh CW, Lee DH, Oh HB, Sohn YH, et al. High prevalence of hepatitis B and hepatitis C virus infections in Korean patients with hematopoietic malignancies. Ann Hematol. 2011;90(2):55–64. doi: 10.1007/s00277-010-1055-5. [PubMed: 2082137]
21. Park SC, Jeong SH, Kim J, Han CJ, Kim YC, Choi KS, et al. High prevalence of hepatitis B virus infection in patients with B-cell non-Hodgkin’s lymphoma in Korea. J Med Virol. 2008;80(6):960–6. doi: 10.1002/jmv.21685. [PubMed: 1842841]
22. Nanth A, Agarwal R, Malthota R, Varma S. Prevalence of hepatitis B virus infection in non-Hodgkin’s lymphoma: a systematic review and meta-analysis. Intern Med. 2010;49(9):633–41. doi: 10.1111/j.1445-5994.2009.02060.x. [PubMed: 1918156]
23. Sitnik R, Pinho JR, Bertolini DA, Bernardini AP, Da Silva LC, Carriolo RJF. Hepatitis B virus genotypes and precore and core mutants in Brazilian patients. J Clin Microbiol. 2004;42(6):2455–60. doi: 10.1128/CMJ.42.6.2455-2460.2004. [PubMed: 1588449]
24. Fernandez E, Rodrigo L, Garcia S, Riestra S, Blanco C. Hepatitis B virus DNA in peripheral blood cells without concomitant liver infection. The liver transplantation model. J Hepatol. 2006;44(2):197–211. doi: 10.1016/j.jhep.2005.01.016. [PubMed: 16186643]
25. de Souza MM, Barbosa MA, Borges AM, Daher RR, Martins RM, Cardoso D. Seroprevalence of hepatitis B virus infection in patients with mental problems. Rev Bras Psiquiatr. 2010;32(4):35–8. [PubMed: 15057838]
26. Okamoto H, Imai M, Tsuda F, Tanaka T, Miyakawa Y, Mayumi M. Point mutation in the S gene of hepatitis B virus for a d4w or w4r subtype change in two blood donors carrying a surface antigen of compound subtype ad4r or adwr. J Virol. 1987;61(10):3030–4. [PubMed: 3040123]
27. Hui CK, Sun J, Au WT, Lie AK, Yang HW, Zhang HY, et al. Occult hepatitis B virus infection in hematopoietic stem cell donors in a hepatitis B virus endemic area. J Infect Dis. 2004;190(6):981–8. doi: 10.1086/381359. [PubMed: 150841]
28. Chen MH, Hsiao LT, Chou TJ, Liu JH, Gau JP, Teng HW, et al. High prevalence of occult hepatitis B virus infection in patients with B-cell non-Hodgkin's lymphoma. Ann Hematol. 2008;87(6):475–80. doi: 10.1007/s00277-008-0469-9. [PubMed: 18327583]
29. Marcucci F, Spada E, Mele A, Caserta CA, Pulsoni A. The association of hepatitis B virus infection with B-cell non-Hodgkin lymphoma - a review. Am J Blood Res. 2012;2(1):18–28. [PubMed: 22432084]

30. Eftekhari Y, Kazemi Arababadi M, Hakimi H, Rezaazadeh Zarandi E. Common HBV genotype in southeastern Iranian patients. Arch Iran Med. 2010;13(2):147–9. [PubMed: 20187670]

31. Anderson LA, Engels EA. Hepatitis C virus infection and non-Hodgkin lymphoma: interesting association or causal relationship? Int J Cancer. 2008;122(8):x–xii. doi: 10.1002/ijc.23462. [PubMed: 18270007]

32. Ozguroglu M, Bilici A, Turna H, Serdengecti S. Reactivation of Hepatitis B Virus Infection with Cytotoxic Therapy in Non-Hodgkin's Lymphoma. Med Oncol. 2004;21(1):67–72. doi: 10.1385/mo:21:1:67. [PubMed: 15034216]

33. Kim JS, Hahn JS, Park SY, Kim Y, Park IH, Lee CK, et al. Long-term outcome after prophylactic lamivudine treatment on hepatitis B virus reactivation in non-Hodgkin's lymphoma. Yonsei Med J. 2007;48(1):78–89. [PubMed: 17326249]

34. Tsutsumi Y, Tanaka J, Kawamura T, Miura T, Kanamori H, Obara S, et al. Possible efficacy of lamivudine treatment to prevent hepatitis B virus reactivation due to rituximab therapy in a patient with non-Hodgkin's lymphoma. Ann Hematol. 2004;83(1):58–60. doi:10.1007/s00277-003-0748-4. [PubMed: 14533286]

35. Jadali Z. Hepatitis C virus cryoglobulinemia and non-hodgkin lymphoma. Hepat Mon. 2012;12(2):85–91. doi: 10.5812/hepatmon.881. [PubMed: 22509184]