A Critical Appraisal of Strategies to Optimize Vitamin D Status in Germany, a Population with a Western Diet

Roman Saternus,1,2,* Thomas Vogt,1,2 and Jörg Reichrath1,2

1 Center for Clinical and Experimental Photodermatology, Saarland University, Campus Homburg, 66421 Homburg, Germany; Thomas.vogt@uks.eu (T.V.); Joerg.Reichrath@uks.eu (J.R.)
2 Department of Dermatology, The Saarland University Hospital, 66421 Homburg, Germany

* Correspondence: Roman.Saternus@uks.eu

Received: 21 September 2019; Accepted: 30 October 2019; Published: 6 November 2019

Abstract: During the last decade, our scientific knowledge of the pleiotropic biological effects of vitamin D metabolites and their relevance to human health has expanded widely. Beyond the well-known key role of vitamin D in calcium homeostasis and bone health, it has been shown that vitamin D deficiency is associated with a broad variety of independent diseases, including several types of cancer, and with increased overall mortality. Moreover, recent findings have demonstrated biological effects of the vitamin D endocrine system that are not mediated via activation of the classical nuclear vitamin D receptor (VDR) by binding with high affinity to its corresponding ligand, the biologically active vitamin D metabolite 1,25-dihydroxyvitamin D (1,25(OH)2D). In contrast, many of these new biological effects of vitamin D compounds, including regulation of the circadian clock and many metabolic functions, are mediated by other vitamin D metabolites, including 20-hydroxyvitamin D and 20,23-dihydroxyvitamin D, and involve their binding to the aryl hydrocarbon receptor (AhR) and retinoid-orphan receptor (ROR). In most populations, including the German population, UVB-induced cutaneous vitamin D production is the main source for fulfilling the human body’s requirements of vitamin D. However, this causes a dilemma because solar or artificial UVR exposure is associated with skin cancer risk. In addition to UVB-induced vitamin D production in skin, in humans, there are two other possible sources of vitamin D: from diet and supplements. However, only a few natural foods contain substantial amounts of vitamin D, and in most populations, the dietary source of vitamin D cannot fulfill the body’s requirements. Because an increasing body of evidence has convincingly demonstrated that vitamin D deficiency is very common worldwide, it is the aim of this paper to (i) give an update of the vitamin D status in a population with a western diet, namely, the German population, and to (ii) develop strategies to optimize the vitamin D supply that consider both the advantages as well as the disadvantages/risks of different approaches, including increasing vitamin D status by dietary intake, by supplements, or by UVB-induced cutaneous synthesis of vitamin D.

Keywords: vitamin D status; Germany; Dietary Intake of vitamin D; UVB-induced cutaneous vitamin D production

1. Introduction

Due to the ubiquitous availability of solar ultraviolet radiation (UVR, includes UVA, UVB and UVC) in most regions worldwide, especially during spring and summer, and because most foods contain little vitamin D, cutaneous vitamin D production represents the most important source of vitamin D for humans [1–4].
During the last decade, our scientific knowledge of the pleiotropic biological effects of vitamin D metabolites and their relevance to human health has expanded widely. Beyond the well-known key role of vitamin D in calcium homeostasis and bone health, it has been shown that vitamin D deficiency is associated with a broad variety of independent diseases, including several types of cancer, cardio-vascular, autoimmune and infectious diseases [5]. Notably, the causal relationship between vitamin D status and health has convincingly been demonstrated for many outcomes, including overall mortality. As an example, in a systematic review that updated and reassessed the benefits and harms of vitamin D supplementation used in primary and secondary prophylaxis of mortality, vitamin D supplementation decreased mortality in all 56 trials analyzed together (5,920/47,472 (12.5%) vs. 6,077/47,814 (12.7%); risk ratio (RR) 0.97 (95% confidence interval (CI)) 0.94 to 0.99; \( p = 0.02 \); I(2) = 0%). ‘Worst-best case’ and ‘best-worst case’ scenario analyses demonstrated that vitamin D could be associated with a dramatic increase or decrease in mortality [6]. Two recent randomized controlled trials reported in the secondary analyses that cancer and progression to diabetes were reduced for those with BMI <25 and <30, respectively (for 2000 and 4000 IU per day vitamin D intake, respectively (1 IU is equivalent to 0.025 µg)) [7,8].

Moreover, recent findings have demonstrated biological effects of the vitamin D endocrine system (VDES) that are not mediated via activation of the classical nuclear vitamin D receptor (VDR) by binding with high affinity to its corresponding ligand, the biologically active vitamin D metabolite 1,25-dihydroxyvitamin D (1,25(OH)2D). In contrast, many of these new biological effects of vitamin D compounds, including regulation of the circadian clock and many metabolic functions, are mediated by other vitamin D metabolites, including 20-hydroxyvitamin D and 20,23-dihydroxyvitamin D, and involve their binding to and activation of the aryl hydrocarbon receptor (AhR) and/or retinoid-orphan receptor (ROR) [9–11].

It should be noted that the human body’s requirements for vitamin D can be fulfilled by at least three different approaches: the UVB-induced cutaneous synthesis of vitamin D [3] and the uptake of vitamin D from the diet and/or by supplements [12]. However, only a few foods naturally contain vitamin D in substantial amounts, and in most populations, food fortification and/or supplements are not sufficiently used. As a consequence, in most populations, the human body’s requirements for vitamin D must be fulfilled physiologically mainly by the UVB-induced cutaneous synthesis of vitamin D [3]. This causes a dilemma because exposure of the skin to solar or artificial UVR is associated with increased skin cancer risk [13].

Because of the high prevalence of vitamin D deficiency worldwide and because of the substantial consequences for human health, it is of high importance to obtain reliable data concerning vitamin D status and vitamin D supply in individual populations [14,15]. Vitamin D deficiency has been previously defined as a 25(OH)D serum concentration < 50 nmol/L (<20 ng/mL) and vitamin D insufficiency as a 25(OH)D serum concentration of 51–74 nmol/L (21–29 ng/mL) [16]. German authorities also consider a value below 50 nmol/L to be insufficient [17]. Some authors recommend a 25(OH)D serum concentration above \( \geq 50 \text{ nmol/L (20 ng/mL) as being optimal for health} [15]. The Endocrine Society clinical practice guideline recommends for children and adults who are vitamin D deficient a target value of 25(OH)D above 75 nmol/L (30 ng/mL) [18]. However, while there is general consensus that 25(OH)D serum concentrations below 25–30 nmol/L (10–12 ng/mL) need to be elevated to prevent and/or treat rickets and osteomalacia, there is currently no general consensus both on the optimal vitamin D serum level and in which (risk) groups of people the 25(OH)D serum concentration should be measured at all [15].

On the other hand, 25(OH)D concentrations higher than 375 nmol/L (150 ng/mL) have been defined to be the hallmark of vitamin D toxicity due to vitamin D overdosing [19].

Some papers reported U-shaped 25(OH)D concentration–health outcomes [20–22]. For example, in an analysis by Melamed et al. (13,331 adults \( \geq 20 \) years), the authors found an increased rate of mortality in females with 25(OH)D levels >125 nmol/L (>50 ng/mL) [21]. However, the significance of the reported U-shaped associations is highly controversial [22]. Grant et al. discussed in a recent
review article that a highly plausible reason for these findings could be that in many individuals with relatively high 25(OH)D serum concentration, their high 25(OH)D concentration is caused because they are at present treated for vitamin D deficiency or insufficiency. Their increased risk for health disorders and diseases may not be the result of their current relatively high vitamin D status but due to the previous long-lasting vitamin D deficiency or insufficiency [22]. Furthermore, recent investigations demonstrated that relative high levels of 25(OH)D serum concentration are associated with a greater benefit for several health outcomes, for example decreased risk in breast cancer (25(OH)D concentration ≥150 nmol/L compared with <50 nmol/L (hazard ratio (HR) = 0.20, p = 0.03)) [23], substantial reduction in in preterm birth risk (maternal 25(OH)D concentration ≥100 nmol/L compared with <50 nmol/L (OR = 0.41, p = 0.002)) [24], substantial reduction in risk of all invasive cancers combined (25(OH)D concentration ≥100 nmol/L compared with <50 nmol/L (HR = 0.33, 95% CI = 0.12–0.90)) [25], improved control of systolic and diastolic blood pressure (BP) in hypertensive individuals who were vitamin D insufficient (25(OH)D concentration > 100 nmol/L and < 250 nmol/L, systolic BP: coefficient = −0.07, p < 0.001; diastolic BP: Coefficient = −0.1, p < 0.001)) [26] and reduction in all-cause mortality (25(OH)D concentration >75 nmol/L compared with 22.5 nmol/L, HR = 1.9, (95 % CI =1.6–2.2; p <0.001)) [27].

Besides the ongoing discussions on the recommended optional 25(OH)D level, other points need to be considered including some differences in measurement methods for 25(OH)D. The first developed assay was the DBP competitive protein binding assay [16]. Since 1985, a radioimmunoassay (RIA) has been available [16]. The major limitation of these assays is the fact that they also recognize other polar metabolites of vitamin D due to a cross reactivity with 24,25(OH)₂D, therefore, leading to an overestimation of 25(OH)D levels [16]. Additionally, these measurement methods have significant inter-assay and inter-laboratory differences [15]. On the other hand, it has been reported that some immunoassays may underestimate total 25(OH)D when 25(OH)D₂ constitutes an appreciable part of the total [28]. This may have an impact to cutoffs of vitamin D sufficiency and may have led to misclassification of patients in the past [29]. Today, direct quantitative measurement of 25(OH)D₂ and 25(OH)D₃ with the liquid chromatography tandem mass spectroscopy (LC-MS) represents the gold standard for measuring 25(OH)D serum concentration [16]. It is the aim of this paper to: (i) give an update of vitamin D status and vitamin D supply in a population with a western diet, namely, the German population, and to (ii) develop and discuss strategies to optimize them. These strategies should consider both the advantages as well as the disadvantages/risks of different approaches (increasing vitamin D status by dietary intake, by supplements, or by UVB-induced cutaneous synthesis).

2. Pandemic Vitamin D Deficiency: A Short Overview of the Vitamin D Status in Germany, A Country with A Western Diet

Vitamin D deficiency is very common worldwide; some authors even speak of a pandemic [5]. Germany, where data from the Robert Koch Institute (RKI) showed that 63% of people aged 1–17 years and 57.3% of people aged 18–79 years have 25(OH)D serum levels below 50 nmol/L (20 ng/mL) (see Table 1) [30], can serve as an example of a well-investigated population with a western diet. In line with these findings, an investigation in the Federal Republic of Germany from 2008 by Hintzpeter et al. analyzed subsample data from the German National Health Interview and Examination Survey 1998 (GNHIES) (2267 women and 1763 men) [31]. They showed that even in the sunny months (May to October), 45.2% of all men and 54.8% of all women were vitamin D deficient (25(OH)D serum level of 50 nmol/L (20 ng/mL) or below) [31]. Between November and April, 68.2% of all men and 60.8% of all women were vitamin D deficient [31]. 25(OH)D levels were measured by using a chemiluminescence immunoassay (CLIA) [31].

A more recent investigation based on data from the ‘German Health Interview and Examination Survey for Adults’ (DEGS1) (3635 women and 3360 men) showed that in the winter months, 25% of German men (between November and April) and 25% of German women (between November and May) had 25(OH)D serum levels <30 nmol/L [32]. Serum 25(OH)D was measured by a chemiluminescence immunoassay [32].
Vitamin D status in elderly people is generally lower than that in younger people [33]. Diekmann et al. investigated the vitamin D status of residents in a German nursing home in Nürnberg (84 women and 31 men) [33]. They found vitamin D deficiency (<50 nmol/L) in 93.9% of the residents [33]. This can be explained by several reasons, including the fact that residents of nursing homes have limited capacities to engage in outdoor activities and that the skin in older individuals has a reduced ability to synthesize vitamin D₃ after exposure to UVR [33]; in that study, 25(OH)D was analyzed by an enzyme-linked immunosorbent assay (ELISA) [33].

Table 1. Prevalence of 25(OH)D serum concentration in Germany, classified by age and sex (cited from reference [30]).

| 25(OH)D [nmol/L] | Age 1–17 Years |  |  | Age 18–79 Years |  |  |
|------------------|----------------|----------------|----------------|----------------|----------------|----------------|
| All (n = 10,015) | Male (n = 5107) | Female (n = 4908) | All (n = 3917) | Male (n = 1706) | Female (n = 2211) |
| <12.5            | 3.8%           | 3.6%           | 4.0%           | 2.0%           | 2.2%           | 1.9%           |
| 12.5 to<25       | 15.5%          | 15.6%          | 15.4%          | 14.3%          | 13.4%          | 15.1%          |
| 25 to<50         | 43.7%          | 42.9%          | 44.5%          | 41.0%          | 41.2%          | 40.8%          |
| 50 to<75         | 22.8%          | 23.3%          | 22.3%          | 20.8%          | 22.6%          | 19.1%          |
| >75              | 14.2%          | 14.6%          | 13.8%          | 21.9%          | 20.6%          | 23.1%          |

Saeglitz analyzed hospitalized geriatric patients (142 women and 83 men; mean age 83 ± 5 years) in Bonn. At hospitalization, the mean serum 25(OH)D concentration was 27.4 nmol/L [34]. Of these patients, 43.9% had a serum 25(OH)D concentration below 25 nmol/L [34,35]. 25(OH)D was measured by an 125I radioimmunoassay [34].

Data from the DEVID study (De Vitamin in Deutschland; 1258 individuals) indicate that 25% of the participants had a 25(OH)D serum concentration below 25 nmol/L and 50% had a serum concentration between 25 and 50 nmol/L. The results for highly aged individuals were even worse: 38% of participants aged 75 years or older showed 25(OH)D serum concentrations below 25 nmol/L, whereas only 22% of participants aged 25–45 had 25(OH)D serum concentrations below 25 nmol/L [35,36]. Serum 25(OH)D was measured by a chemiluminescence immunoassay [36].

Another investigation showed that Turkish descendants living in Berlin have an increased risk of developing vitamin D deficiency [37]. In that study, 94.8% of women and 95.2% of men of Turkish descent had 25(OH)D serum concentrations below 50 nmol/L, compared to 86.9% of women and 78.4% of men in the group of individuals without Turkish descent [37]. In total, 597 persons of Turkish descent and 129 Berliners without Turkish descent were analyzed [37]. Serum 25(OH)D was measured by a chemiluminescence immunoassay [37]. It has been discussed that people of Turkish descent living in Germany have distinct risk factors for developing vitamin D deficiency, including relatively dark skin and wearing long clothing [37].

Concerning vitamin D status in Germany among children and adolescents, Hintzpeter et al. analyzed in the KiGGS (German National Health Interview and Examination Survey for Children and Adolescents) study (17,641 participants aged 0–17 years) [38]. 25(OH)D values were measured by using a chemiluminescence immunoassay [38]. The authors demonstrated that 31.2% of boys and 36.6% of girls aged 1–2 years with a nonimmigrant background were vitamin D deficient (25(OH)D <50 nmol/L) [38]. In the age group of 3–17 years, 64.8% of boys and 63.4% of girls with a nonimmigrant background showed vitamin D deficiency [38]. Furthermore, they analyzed the risk for vitamin D deficiency in children with different immigrant background: Boys with an Turkish or Arabic–Islamic immigrant background had an increased risk of having moderate or severe 25(OH)D concentrations (<25 nmol/L) compared with nonimmigrants (OR 2.3; [95% CI] 1.4–3.8 and OR 7.6; [95% CI] 3.0–19.1), whereas girls with a Turkish (OR 5.2; [95% CI] 2.9–9.6), Arab–Islamic (OR 5.9; [95% CI] 2.5–14.0), Asian (OR 6.7; [95% CI] 2.2–19.8) or African (OR 7.8; [95% CI] 1.5–40.8) background had an increased risk of having moderate or severe 25(OH)D concentrations [38].

Taken together, these data underline the importance of developing strategies to improve vitamin D status.
3. The Human Vitamin D Endocrine System (VDES): Molecular Biology of an Ancient Friend, Revisited

Because, in contrast to classical vitamins, vitamin D and its active metabolites are synthesized in the human body (depending on exposure of the skin to UVR), they represent a prohormone/hormone and not a vitamin [3,39].

In human skin cells, the initial substrate of cutaneous vitamin D synthesis is 7-dehydrocholesterol (7-DHC or provitamin D$_3$) [6,39]. After exposure to solar or artificial UVB rays, the B-ring of the sterane structure of the 7-DHC molecule is split between atoms C9 and C10 [40]. This reaction is called photochemical conversion or photolysis [41]. The maximum wavelength leading to this photolysis is in the range of UVB radiation, i.e., approximately in the wavelength range between 290 and 310 nm [40]. By absorbing UVR photons, the 7-DHC molecule is converted into previtamin D$_3$ [40]. After photolysis, previtamin D$_3$ is released into the extracellular space due to altered hydrophobic and hydrophilic interactions with the fatty acids within the cell membrane [40]. Since previtamin D$_3$ is energetically unstable, it spontaneously reacts to form vitamin D$_3$ via a thermal isomerization reaction [40]. (For a schematic representation of the vitamin D synthesis, see Holick et al. [42].) The photolysis of 7-DHC to the product vitamin D$_3$ depends on many factors (see section below).

Vitamin D$_3$ is then released into the blood circulation [42]. Because of its lipophilic molecular structure, vitamin D$_3$ must be bound to transport proteins [42]. The main transport protein is the vitamin D-binding protein (DBP or group-specific component (GC)-globulin), a 52–59 kDa plasma protein produced in the liver [43]. Bound in the blood to DBP, vitamin D$_3$ is then transported to the liver, where it is hydroxylated by the microsomal enzyme CYP2R1 at the C-25 atom [44]. The resulting 25-hydroxyvitamin D$_3$ (25(OH)D$_3$ or calcidiol) is the major circulating form of vitamin D with a half-life of approximately 2–3 weeks [16]. It is well accepted that serum 25(OH)D is the best parameter for analyzing a person’s vitamin D status [16]. While the majority of 25(OH)D$_3$ in the blood is bound to DBP [42], some authors suggest that 25(OH)D$_3$ can be rarely (approx. 0.1%) found in an unbound, free form in serum (free hormone hypothesis) [45]. It has been discussed that this unbound 25(OH)D may drive many of the non-classical actions of vitamin D [45].

The complex of circulating DBP and 25(OH)D$_3$ is then filtered by the kidney’s glomeruli because of its low molecular weight [46,47]. Upon filtration, the megalin/cubulin receptor complex located in the luminal membrane of renal proximal tubule epithelial cells binds the DBP/25(OH)D$_3$ complex and is internalized via endocytosis into the proximal tubule cells, in a process similar to albumin and low molecular weight proteins [46,47]. Within renal cells, the complex is delivered to the lysosomal compartment, and 25(OH)D$_3$ can be released into the cytosol by degrading DBP [46]. Thereafter, the 25(OH)D$_3$ molecule is hydroxylated by the enzyme CYP27B1 (25-hydroxyvitamin D-1α-OHase) at the C-1 position of the molecule [42]. The resulting 1,25-dihydroxyvitamin D$_3$ (calcitriol, 1,25(OH)$_2$D$_3$ or 1,25(OH)$_2$D) is the major active form of vitamin D in the circulation [42]. In contrast to 25(OH)D, the half-life of circulating 1,25(OH)$_2$D is only 4–6 h, and the serum levels of 1,25(OH)$_2$D are a thousand-fold lower compared to 25(OH)D [16].

After binding to DBP, 1,25(OH)$_2$D$_3$ produced in the kidneys is transported via the circulation to distinct target organs, predominantly to tissues involved in bone and calcium metabolism [48]. However, many tissues not involved in bone and calcium metabolism are also target organs for biologically active vitamin D metabolites [45]. Many of these extrarenal cells, for example, immune cells, including monocytes, macrophages and dendritic cells, also express CYP27B1, possess α-hydroxylase activity and produce 1,25(OH)$_2$D$_3$ locally [45]. It has been discussed that this extra-renal produced 1,25(OH)$_2$D$_3$ is not transferred to the circulation as an endocrine hormone but exerts regulatory effects locally via autocrine or paracrine effects in various tissue-dependent cellular functions [45]. In target cells, 1,25(OH)$_2$D$_3$ exerts many of its pleiotropic genomic effects via binding to the nuclear vitamin D receptor (VDR) [48]. The VDR is a nuclear receptor that, after forming a heterodimer with the retinoid-X receptor (RXR), regulates the transcriptional activity of target genes via binding to specific DNA recognition
sequences called response elements [14]. It has been estimated that 1,25(OH)\textsubscript{2}D\textsubscript{3} regulates more than 200 genes by these mechanisms [14].

In addition to the traditional pathway of vitamin D activation via 7-DHC, 25(OH)D and 1,25(OH)\textsubscript{2}D, other activation pathways with alternative metabolites, including 20(OH)D and 20,22(OH)\textsubscript{2}D, have been reported [49].

A key role in this pathway is cytochrome P450 side-chain cleavage (P450scc), encoded by the CYP11A1 gene [50]. P450scc catalyzes a three-step reaction: first, the 22-hydroxylation of cholesterol; second, the 20-hydroxylation of 22(R)-hydroxycholesterol; and finally, the oxidative scission (bond cleavage) between atoms C20 and 22 of the 20(R),22(R)-dihydroxycholesterol molecule [51]. This reaction is called a side-chain cleavage event resulting in the end product pregnenolone, a precursor of steroid hormones [51]. During the reactions, the metabolites do not leave the active site of CYP11A1 [52].

Scientific findings indicate that CYP11A1 is important for VDES by catalyzing 7-DHC to 7-dehydropregnenolon in a three-step reaction (with the intermediate metabolites 22(OH)7-DHC and 20,22(OH)\textsubscript{2}7-DHC) analogous to the reaction of cholesterol to pregnenolone [52,53]. Additionally, it has been reported that CYP11A1 hydroxylates previtamin D\textsubscript{3} to the secosteroid 20(OH)D\textsubscript{3} and finally to 20,22(OH)\textsubscript{2}D\textsubscript{3} [52]. Notably, it has been shown that the nuclear aryl hydrocarbon receptor (AhR) is the major corresponding receptor target for 20,23(OH)2D\textsubscript{3} [9]. However, most of the non-skeletal effects of vitamin D remain highly controversial [54].

4. How Can a Healthy Vitamin D Status Be Achieved and Maintained? Relevance of Supplements and Dietary Intake

In addition to UVB-induced vitamin D production in the skin, in humans, there are two other possible sources of vitamin D: diet and supplements. In the absence of endogenous synthesis of vitamin D, the present guidelines of the German society for nutrition (Deutsche Gesellschaft für Ernährung (DGE) e.V.) and the D-A-CH society (i.e., Germany, Austria, Switzerland) recommend an intake of 400 IU (10 \( \mu \)g) vitamin D per day for infants and 800 IU (20 \( \mu \)g) per day for children, adolescents, adults, pregnant women and breast-feeding women [55]. Previously, the D-A-CH society recommended a lower vitamin D intake, namely, 200 IU (5 \( \mu \)g) per day for younger people and 400 IU (10 \( \mu \)g) per day for persons older than 65, respectively [56].

In addition to vitamin D intake with breastmilk or infant food, German pediatricians recommend an oral supplementation with 400–500 IU vitamin D\textsubscript{3} per day for all infants up to the second summer (duration from 1–1.5 year, depending on birth date) [57]. Premies with a birth weight below 1500 g should be given 800–1000 IU vitamin D daily during the first months of life [57].

The current Endocrine Society clinical practice guideline suggests for children aged 0–1 year at least 400 IU per day of vitamin D\textsubscript{3}, for children from 1 year, adults aged 19–70, pregnant and lactating women at least 600 IU per day of vitamin D and for adults from aged 70 years 800 IU per day of vitamin D [18]. However, to raise the blood level of 25(OH)D\textsubscript{3} above 75 nmol/L (30 ng/mL), higher intakes of vitamin D have been recommended (1000 IU per day for children and 1500–2000 IU per day for adults) [18]. For obese children and adults and adults on anticonvulsant medications, glucocorticoids, antifungals and medication for AIDS, the Endocrine Society suggests at least two to three times more vitamin D for their age group [18].

Concerning vitamin D deficiency, the Endocrine Society suggests that all adults who are vitamin D deficient should be treated with 50,000 IU of vitamin D once a week for 8 weeks or its equivalent of 6000 IU of vitamin D daily to achieve a blood level of 25(OH)D\textsubscript{3} above 75 nmol/L (30 ng/mL), followed by maintenance therapy of 1500–2000 IU per day [18]. For children aged 0–18 years who are vitamin D deficient, the Endocrine Society recommends treatment with 2000 IU per day of vitamin D or with 50,000 IU of vitamin D once weekly for 6 weeks to achieve a blood level of 25(OH)D\textsubscript{3} above 75 nmol/L (30 ng/mL), followed by maintenance therapy of 400–1000 IU per day (for infants aged 0–1 year) or 600–1000 IU per day (for children aged 1–18 years), respectively [18]. For obese patients, patients with
malabsorption syndromes and patients on medications affecting vitamin D metabolism, the Endocrine Society suggests two to three higher doses of vitamin D to treat vitamin D deficiency, followed by a maintenance therapy of 3000–6000 IU per day [18].

On the other hand, the Institute of Medicine (IOM) has defined a tolerable upper intake level for vitamin D at 1000 IU per day in infants ages 0–6 months, 1500 IU per day in infants ages 6–12 months, 2500 IU per day in children ages 1–3 years, 3000 IU per day in children ages 4–8 years and 4000 IU per day in adolescents and adults [20,58]. However, the Endocrine Society clinical practice guidelines noted that higher dosages may be needed to correct vitamin D deficiency [18].

4.1. Ingestion, Absorption and Bioavailability of Vitamin D

Vitamin D from diet or supplements may be present as vitamin D$_3$ (cholecalciferol) or vitamin D$_2$ (ergocalciferol) [42,59,60]. Non-hydroxylated vitamin D is a lipophilic molecule that is closely related to cholesterol [60,61]. In the intestinal lumen, other lipids (such as triglycerides or phospholipids) are important for solubilizing fat-soluble nutrients in micelles and for their absorption [60]. Digestive enzymes catalyze the release of monoglycerides and phospholipids from lipids, resulting in the formation of further micelles [60]. However, there are no consistent data to support the hypothesis that a diet containing high amounts of fat may improve the bioavailability of vitamin D [60], although some data suggest that the type of dietary fat is important: some investigators have found that a high consumption of monounsaturated fatty acid-rich oils may improve the bioavailability of supplemental vitamin D$_3$ [62]. Additionally, there are published data indicating that dietary fiber, vitamin E, vitamin A, phytosterols and plant sterols may reduce the bioavailability of vitamin D [60]. However, the precise underlying mechanisms as well as the role of other possible factors influencing vitamin D bioavailability, such as gastric pH or the role of intestinal enzymes, are not completely understood [60].

Depending on the concentration of vitamin D, two different mechanisms of absorption in the gastro-intestinal system have been reported [60] if present in low concentrations (i.e., vitamin D in the diet is often weakly concentrated), transport through the enterocyte cell membrane is mediated by distinct specific proteins [60]. It has been suggested that proteins involved in protein-mediated transport may include SRBI (scavenger receptor class B type 1), CD 36 (cluster determinant 36) and NPC1L1 (Neimann–Pick C1-Like 1) [60,61]. Uptake of vitamin D in high concentrations, usually resulting from intake of supplements, is mediated via passive diffusion through the enterocyte cell membrane [60]. These mechanisms may explain the reported fact that vitamin D from diet has only an estimated 60% bioavailability compared to vitamin D supplementation [60].

The human diet also contains hydroxylated metabolites of vitamin D (25(OH)D$_2$ and 25(OH)D$_3$) (Table 2) [60]. The absorption efficiency and thereby the bioavailability of hydroxylated metabolites of vitamin D has been reported to be up to ten times higher than the bioavailability of non-hydroxylated forms, but little is known about the specific underlying mechanisms [60].

After ingestion with food or supplements and absorption in the gut, both vitamin D$_2$ and vitamin D$_3$ are transported via chylomicrons and DBP via the lymphatic system to the liver [63,64], where they are converted into their analogous metabolites, namely, 25(OH)D$_2$ and 25(OH)D$_3$, similar to cutaneously produced vitamin D [63]. Finally, 25(OH)D produced in the liver enters the blood circulation, where it binds to DBP and lipoproteins [42].

4.2. The Role of Vitamin D$_2$

Only plants and fungi possess ergosterol in their cell membranes; therefore, they are the only ones able to produce vitamin D$_2$ forms [40]. Although vitamin D$_2$ and D$_3$ molecules are quite similar, there are some differences in their structural formulas. Vitamin D$_3$ has a molar mass of 384, while vitamin D$_2$ has a molar mass of 396 [65]. Investigations have shown that vitamin D$_2$ metabolites bind with a lower affinity to DBP compared with vitamin D$_3$ metabolites, resulting in a relative inefficiency of vitamin D$_2$ to raise total serum 25(OH)D levels, as well as a shorter plasma half-life and a lower activity of D$_2$.
metabolites [45]. Because of the low binding affinity, 25(OH)D$_2$ is also transported in chylomicrons and lipoprotein fraction 3 (LPF3). In contrast to DBP, chylomicrons and LPF3 fractions are not able to protect bound molecules against degradation or excretion [64]. It has been reported that in addition to the traditional pathway of ergosterol (similar to vitamin D produced in the skin), ergosterol can be hydroxylated by CYP11A1, first at the C24 position and second at C17, without side chain cleavage, resulting in 17α,24-dihydroxyergosterol (17α,24(OH)$_2$D$_2$) [66]. Many other alternative substrates, such as 20-hydroxy-22,23-epoxy-22,23-dihydroergosterol and 22-keto-23-hydroxy-22,23-dihydroergosterol, have been described [67].

It is well accepted that vitamin D$_3$ is more effective at raising total 25(OH)D concentrations than vitamin D$_2$ [68–72]. Some investigations have shown that vitamin D$_2$ intake increases 25(OH)D$_2$ but produces a decline in 25(OH)D$_3$ [68–70]. Recently, Martineau et al. reported that administration of vitamin D$_2$ reduces 25-hydroxylation of vitamin D$_3$ and 1-alpha hydroxylation of 25(OH)D$_3$, while increasing 24R-hydroxylation of 25(OH)D$_3$ [72]. Vice versa, it has been reported that vitamin D$_3$ intake is associated with a decrease in 25(OH)D$_2$ level, suggesting a common regulatory mechanism [73]. However, the exact mechanisms are unknown [73].

Armas et al. treated 20 healthy male volunteers with a single dose of 50,000 IU of vitamin D$_2$ or D$_3$ [69]. They showed a similar initial increase in serum 25(OH)D over the first 3 days in both groups. But only in the D$_3$-treated subjects 25(OH)D level continued to rise, whereas serum 25(OH)D level fell rapidly in the D$_2$-treated subjects and was not different from baseline after two weeks [69]. In an umbrella review of systematic reviews and meta-analyses of observational studies and randomized trials done by Theodoratou et al., it has been reported that vitamin D$_2$ supplementation is associated with increased (but non-significant) risk of mortality (reported summary effect: 1.04 (CI 0.97 to 1.11), p-value: 0.26) [74].

4.3. Vitamin D from Supplements

In Germany, many oral vitamin D supplements containing ergocalciferol or cholecalciferol (and with or without additional components such as calcium carbonate) are easily available and are accessible over the counter without a physician’s prescription. Higher dosed oral supplements (from 5600 IU) as well as hydroxylated vitamin D derivates (alfacalcidiol, paricalcitol or calcitriol) are available only on prescription.

The German federal healthy survey 1998 (BGS) done by the Robert Koch Institute analyzed vitamin supplementation of 7124 participants aged 18–79 (response rate: 61%) [75]. The investigators found that 57 participants used oral vitamin D supplementation every day. The mean daily vitamin D intake was 292 IU (7.4 µg) [75]. According to the results of the German national consumption study (Nationale Verzehrstudie) from 2008 (8278 women and 7093 men), 477 women and 270 men used vitamin D supplements [56]. The mean intake of vitamin D was 200 IU (5 µg) per day for women and men [56]. Vitamin D supplementation in women increased with increasing age, whereas the highest vitamin D supplementation in men was in the age from 35 to 50 [56].

Several studies analyzed vitamin D supplementation in terms of dosage, administration and efficacy to increase 25(OH)D level. Direct comparisons between oral and intramuscular supplementation of vitamin D$_3$ showed that both effectively increase serum 25(OH)D concentrations [76,77].

Garland et al. analyzed the increase in 25(OH)D serum concentration after vitamin D supplementation [78]. They estimated that starting at baseline 25(OH)D serum concentrations of 25 nmol/L (10 ng/mL), 75 nmol/L (30 ng/mL), 125 nmol/L (50 ng/mL) and 225 nmol/L (90 ng/mL), oral uptake of 1000 IU per day may result in mean increases in 25(OH)D serum concentrations of 27.5 nmol/L (11 ng/mL), 20 nmol/L (8 ng/mL), 12.5 nmol/L (5 ng/mL) and 4 nmol/L (1.6 ng/mL), respectively [78]. Multivariable regression analysis by Zittermann et al. showed that vitamin D dose per kg body weight per day could explain 34.5% of variation in circulating 25(OH)D [79]. Other factors influencing the 25(OH)D level after supplementation were type of supplement (D$_2$ or D$_3$), age, concomitant intake of calcium supplements and baseline 25(OH)D [79].
A recent study randomized 60 subjects with vitamin D deficiency in Belgium who were either treated with 2000 IU vitamin D3 orally per day or 50,000 IU orally per month (cumulative dose: 150,000 IU vitamin D3 in each treatment group) [80]. The authors showed that the monthly supplementation was an effective tool for a rapid normalization of 25(OH)D3 in deficient subjects, whereas the daily administration was similarly effective but took two weeks longer to reach the desirable level of 50 nmol/L (20 ng/mL) [80]. Kearns et al. investigated in a meta-analysis review the efficacy of a single large bolus dose for the treatment of vitamin D deficiency [81]. They concluded that a single dose of vitamin D3 ≥300,000 IU may be effective to improve vitamin D status for up to 3 months [81]. A more recent investigation showed that 50,000 IU monthly and 150,000 IU 3-monthly of vitamin D3 safely and effectively correct vitamin D deficiency in adolescents [82]. Takacs et al. compared the efficacy of daily, weekly and monthly administration of vitamin D3 [83]. They concluded that the daily, weekly and monthly administrations of daily equivalent of 1000 IU of vitamin D3 provide equal efficacy and safety profiles [83].

However, it has to be noted that high-doses supplementation, as mentioned Kearns et al. [81], among others, may be harmful and increases the risk of intoxication. Rossini et al. reported an extremely high 25(OH)D peak increment to 167.75 nmol/L ± 42.76 nmol/L (67.1 ± 17.1 ng/mL, p < 0.001) three days after a single oral bolus of 600,000 IU vitamin D3 [84]. Symptoms of vitamin D toxicity include hypercalcemia, hypercalciuria, kidney stones, hyperphosphatemia, polyuria, polydipsia, ectopic calcification of soft tissues, nausea/vomiting, anorexia, constipation, headache and hypertension [81,85–88]. Hypercalciuria has been reported to be a more sensitive criterion for vitamin D excess than hypercalcemia [89]. Kahawati et al. reported in a systematic review (3 RCTs [n = 39,213]) that supplementation using vitamin D with calcium (1000 IU vitamin D3/1400 mg calcium daily, 2000 IU vitamin D3/1500 mg calcium daily and 400 IU vitamin D3/1000 calcium daily, respectively) was associated with an increased incidence of kidney stones (pooled absolute risk differences (ARD), 0.33% [95% CI, 0.06% to 0.60%]) [90]. Thus, those taking high vitamin D doses should reduce or eliminate calcium supplementation. However, in an investigation by Ferraro et al., vitamin D intake in typical amounts was not statistically associated with risk of kidney stone formation [91]. They also did not observe a higher risk, even among those participants with higher intakes of calcium [91]. In a recent systematic review analysis (15 studies, 3150 participants, ≥2800 IU per day vitamin D2 or D3 supplementation), Malihi et al. demonstrated that one year or longer supplementation with a daily, weekly or monthly dose of vitamin D2/D3 did not significantly increase the risk of total adverse events or kidney stones, although there was a trend towards increased hypercalcemia, and possibly for hypercalciuria [92].

Besides supplementation with cholecalciferol and ergocalciferol, hydroxylated vitamin D derivates (in Germany: alfacalcidol, calcitriol, paricalcitol) are available that can be administered orally or intravenously. A review analysis by Mazess et al. (21 clinical trials that compared intravenous and oral vitamin D analogs (calcitriol, alfacalcidol, doxercalciferol) for the treatment of secondary hyperparathyroidism in hemodialysis patients), the authors found no difference in efficacy between intravenous and oral vitamin D hormone supplementation [93].

4.4. Vitamin D from Diet

Only a few foods naturally contain vitamin D or 25(OH)D3 in substantial amounts (Tables 2 and 3) [14,94,95].

For example, 100 g of fresh wild salmon contains approximately 600–1000 IU of vitamin D [14]. One hundred grams of fresh shiitake mushrooms contains approximately 100 IU of vitamin D, and an egg yolk contains approximately 20 IU of vitamin D [14]. The content of vitamin D may differ from the origin of the product [94].

Meat may also be a source of vitamin D [95]. In an investigation by Crowe et al., plasma 25(OH)D levels were analyzed in meat eaters, fish-eaters and vegetarians (1388 meat-eaters, 210 fish-eaters, 420 vegetarians and 96 vegans, aged 20–76 years from the European Prospective Investigation into Cancer and Nutrition (EPIC)–Oxford cohort) [96]. The authors found that meat-eaters had the highest
mean intake of vitamin D (124 IU, 3.1 (95% CI 3.0, 3.2) µg per day) and mean plasma 25(OH)D concentrations (77.0 (95% CI 75.4, 78.8) nmol/L) and vegans the lowest (28 IU, 0.7 (95% CI 0.6, 0.8) µg per day and 55.8 (95% CI 51.0, 61.0) nmol/L, respectively [96].

Table 2. Vitamin D₃ and 25(OH)D₃ content of chosen foods, modified by [94,95].

| Food (100 g)          | Vitamin D₃  | 25(OH)D₃   |
|-----------------------|-------------|------------|
| Raw eggs              | 12–100 IU   | 0.1–1.5 µg |
| Cooked eggs           | 12–92 IU    | 0.18–1.2 µg|
| Raw white fish        | 4–188 IU    | 0.30–0.70 µg|
| Cooked white fish     | 4–232 IU    | 0.35–0.60 µg|
| Beef                  | 4.4–5.6 IU  | 0.15–0.16 µg|
| Lamb                  | 6.8–10.8 IU | 0.16–0.18 µg|
| Chicken               | 11.6–17.6 (0.29–0.44 µg) | 0.36–0.51 µg |
| Pork                  | 7.2 IU (0.18 µg) | 0.17 µg   |

Table 3. Vitamin D content of chosen foods, modified by [14].

| Food                                      | Approximate Vitamin D content |
|-------------------------------------------|-------------------------------|
| Fresh, wild Salmon (99.05 g)              | 600–1000 IU                   |
| Fresh, farmed salmon (99.05 g)            | 100–250 IU                    |
| Canned salmon (99.05 g)                   | 300–600 IU                    |
| Canned sardines (99.05 g)                 | 300 IU                        |
| Canned mackerel (99.05 g)                 | 250 IU                        |
| Canned Tuna (101.88 g)                    | 230 IU                        |
| Cod liver (1 teaspoon)                     | 400–1000 IU                   |
| Fresh shiitake mushrooms (99.05 g)        | 100 IU                        |
| Sun-dried shiitake mushrooms (99.05 g)    | 1600 IU                       |
| Egg yolk                                  | 20 IU                         |

In Germany, the main source of foods with a considerable amount of vitamin D is fish or crustacean representing over one-third of vitamin D intake, followed by fish-based dishes representing 15% of all consumed vitamin D products [56]. Approximately 10% of vitamin D intake comes from fats, eggs and milk/cheese [56]. On average, a German man consumes 29 g of fish, fish products, crustaceans and related dishes and a German woman 23 g per day [56]. In a study with 19 European countries (including Germany) ocean fish was the most important single dietary factor affecting serum 25(OH)D concentration for postmenopausal women, but animal fat and meat also contributed [97,98]. According to the results of the German national consumption study (Nationale Verzehrstudie), the median vitamin D intake for men is 116 IU (2.9 µg) per day and for women is 88 IU (2.2 µg) per day. The median intake of vitamin D increases in men and women up to the age of 51–64 years and remains constant at ages above that age group [56]. The mean intake of vitamin D at the age of 65 years or older is 132 IU (3.3 µg) for men and 88 IU (2.2 µg) for women [56]. Based on the previously recommended vitamin D intake of the D-A-CH society (200 IU per day and 400 IU per day for persons older than 65, respectively), the National Consumption Study calculated that a total of 82% of men and 91% of women did not achieve the recommended daily intake of vitamin D [56].

Regarding children and young people, the vitamin D intake is even lower: according to the EsKiMo study (nourishing module of the KiGGS study; 1258 girls and 1248 boys), the median vitamin D intake of 6-year-old boys is 56 IU (1.4 µg), and girls of the same age consume 52 IU (1.3 µg) per day as the median. Boys aged 15–17 years have a median daily vitamin D intake of 100 IU (2.5 µg) and girls in this age group of 68 IU (1.7 µg) [99].

It has to be noted, that in addition to the relatively low vitamin D content of most foods, some other factors may limit oral vitamin D uptake, including malabsorption in the gastrointestinal tract (e.g., resulting from cystic fibrosis, celiac disease, Whipple’s disease, Crohn’s disease, bypass surgery and/or medications such as lipid-lowering agents) [14,61].
4.5. Vitamin D Food Fortification

Because most foods contain little vitamin D, there is mounting support for vitamin D fortification of common foods [100–103]. For the USA and Canada, it has been estimated that approx. 60% of the intake of vitamin D from foods comes from fortified foods [104]. In Finland, Jääskeläinen et al. showed that after starting vitamin D fortification of fluid milk products and fat spreads in 2003, the vitamin D status of Finnish adults has improved considerably [105]. In Denmark, Gronborg et al. found that vitamin D-fortified foods (yoghurt, cheese, eggs and crisp bread) improved vitamin D status of women of Danish and Pakistani origin during winter [106]. The same group analyzed different scenarios increasing vitamin D intake in Danish women (diet without fish, diet including fish, fortified foods and supplements) [107]. They concluded that low-dose fortification of different foods with vitamin D may be an effective and safe population-based approach [107]. Outcomes of the ODIN project (food-based solutions for optimal vitamin D nutrition and health through the life cycle) showed that vitamin D-food fortification safely increases population intakes and prevents low vitamin D status [108]. Keller et al. analyzed the risk of development of gestational diabetes of Danish women who were exposed during their fetal life to extra vitamin D from food fortification [109]. They demonstrated that prenatal exposure to extra vitamin D from mandatory fortification may lower the risk of developing gestational diabetes among spring-born women [109]. Besides of conventional vitamin D fortification as a nutrient supplement, there are some novel approaches to vitamin D enrichment of food, including “bio-additions” (ex. the exposure of edible mushrooms to UVR) and bio-fortification (enhancing nutritional quality through agronomic or modern biotechnology techniques) [68,104].

However, in Germany, vitamin D food fortification is still limited to margarine, based on a law of 1942 [100].

Taken together, since little food contains sufficient vitamin D and food fortification does not play a role in Germany due to legal requirements, vitamin D supplementation is a promising strategy for increasing vitamin D levels according to the recommendations of the guidelines. However, the upper intake levels should not be exceeded for a long time and physicians and patients should be aware of symptoms of intoxication.

5. Vitamin D Status in Humans: Relevance of UVB-induced Cutaneous Vitamin D Production

Since vertebrates like humans do not have ergosterol in their cell membrane, they produce vitamin D₃ exclusively in their skin [40]. Due to the ubiquitous availability of solar UVR in most regions worldwide, especially during spring and summer, and because most foods contain little vitamin D, cutaneous vitamin D production is developed in humans as the most important source for fulfilling the human body’s requirements for vitamin D. On the other hand, populations such as Greenland’s indigenous people with dark skin pigmentation have a diet that is predominantly based on marine mammals containing large amounts of vitamin D [1–4,110].

The UVB-induced cutaneous vitamin D production depends on many individual factors. For example, Holick et al. showed that vitamin D₃ production in Boston, located at the 42° N latitude in the US state of Massachusetts, is only possible between March and November [5]. In July, vitamin D photosynthesis begins between 9 and 10 a.m. and ends around 6 p.m. in the evening [5]. In contrast to Boston, vitamin D₃ production in Bergen (Norway), which is 18° north of Boston, is only possible between April and October and within a time period between 11 a.m. and 6 p.m. in July [5]. Similar results were shown by Webb et al. [111]. They found that in Boston, vitamin D production is possible from November through February [111]. In Edmonton (10° north of Boston), in the Canadian province of Alberta, the photosynthesis of previtamin D₃ stops from October to April [111]. Germany is located between the latitude of Boston and Edmonton, specifically between 47° N and 55° N latitude [32]. The dependence on latitude, season and time of day is well explained by the fact that the path of UVB through the atmosphere depends on the solar zenith angle. This path is longer during winter months, at higher latitudes and during morning and evening hours, resulting in an increased absorption of UVB photons by ozone molecules in the stratosphere that leads to reduced
numbers of UVB photons reaching the earth’s surface—and the skin [5,40]. Effects of climate change cause complex changes in stratospheric ozone. In some regions these changes will enhance levels of UVR, while in others they will reduce UVR [112].

In addition to these geographical factors, many other factors have a complex impact on cutaneous vitamin D synthesis, including season, whether conditions, time of exposure [5], clouds and aerosols in the air [113], skin pigmentation based on the genetic background [14,114,115], body fat mass [116], age [40], UVR-exposed body surface area [117], medication, and liver or kidney diseases [14]. In a systematic review by Xiang et al., the authors conclude that pigmented skin has less effective photoproduction of vitamin D and 25(OH)D following experimental UVR exposure [118]. However, an investigation by Hakim et al. with Caucasian women (skin types II–V) found no skin type or ethnicity-dependent differences in production of 25(OH)D after identical UVR exposure [119].

Older experimental data indicate that the use of sunscreen suppresses cutaneous vitamin D synthesis [120]. However, a recent investigation showed that the use of sunscreens with sun protection factor (SPF) 15 and a high UVA-protection factor applied at sufficient thickness still allows a highly significant improvement of serum 25(OH)D concentration [121]. These results are in accordance with a recent review showing that sunscreen use for daily and recreational photoprotection does not compromise vitamin D synthesis [122].

GWAS analysis by Wang et al. identified variants near genes involved in cholesterol synthesis (DHCR7), hydroxylation (CYP2R1, CYP24A1), and vitamin D transport (GC) that influence vitamin D status [43].

Interestingly, a longer UVR exposure of the skin does not necessarily provide more vitamin D; instead, the level may even decrease via two different mechanisms: photochemical regulation and photodegradation. Along with a higher risk of sunburn and skin cancer, prolonged UVR or sun exposure is therefore not associated with an additional health benefit. On the other hand, photochemical regulation and photodegradation prevent excessive vitamin D production and the development of vitamin D intoxication via cutaneous synthesis [123]. During prolonged exposure to the sun or at excessively high wavelengths of UVR, inactive byproducts of previtamin D3 are formed: Previtamin D3 is reversibly converted into photoisomers tachysterol 3 and lumisterol 3 and finally irreversibly into toxisterols [40,124]. A similar effect is due to the second mechanism, called photodegradation of vitamin D3 [125]. With increasing wavelength before being released into the blood circulation, vitamin D3 is irreversibly converted via a photolysis reaction into three possible products: 5,6-trans vitamin D3, suprasterol 1 and suprasterol 2 [125].

Investigations have shown that a single whole-body UVB irradiation with 1 minimal erythema dose (MED) corresponds to an oral intake between 10,000 and 25,000 IU of vitamin D3 [40]. The UVR exposure of ½ MED on ½ of skin area (hands, face and arms) is equivalent to a dietary intake of about 1000 IU vitamin D [126]. The MED is defined as the amount of UVR that will produce minimal erythema (redness caused, e.g., by dilated capillaries) of an individual’s skin within a few hours following exposure [127]. Chen et al. measured the serum concentrations of 25(OH)D in 15 healthy young adults (skin types II and III, age 20–53 years) after most parts of their bodies were exposed three times a week (0.75 MED per week) for a period of 7 weeks. The mean total dose of UVB radiation was approximately 4 MED. They found a 50% increase in the baseline serum 25(OH)D level after one week. This increase continued for five weeks before reaching a plateau of approximately 150% above baseline values [40]. However, when repeating this investigation with a 76-year-old male, his serum 25(OH)D reached a plateau after 2 weeks of exposure, but at only approximately 60% above baseline values [40].

More recent investigations use the standard erythema dose (SED) [128]. One SED is equivalent to an erythemal exposure of 100 J/m² [128]. The expected MEDs of skin types I to IV is in the region of 2.5–4.5 SED on previously unexposed buttock [129]. Rhodes et al showed that 13 min of midday sunlight exposure on a cloudless day, three times weekly, to 35% skin surface area over a 6-week summer period was sufficient to achieve in 90% of the participants (120 volunteers, white Caucasians, sun-reactive skin types I–IV, aged 20–60 years, from Greater Manchester, UK) the vitamin D sufficiency range (25(OH)D
≥ 50 nmol/L [130]. However, only a minor portion of the participants (26%) reached vitamin D levels in the proposed optimal range (25(OH)D ≥ 80 nmol/L) [130]. Manchester shares latitude with northern parts of Germany. Webb et al. analyzed these results under real-life conditions [131]. After a mean daily midday exposure of 9 min (weekdays) and 18 min (weekends) at the end of the summer, the mean increase in 25(OH)D was 71 nmol/L [131]. The authors concluded that relatively short, frequent exposures to sunlight are effective to increase vitamin D status [131]. Narbutt et al. analyzed changes in 25(OH)D concentration after UVR of 32 healthy Polish children (skin types I–IV) during a holiday on the Baltic Sea [132]. Poland is the eastern neighbor of Germany with similar latitude. The investigators found that after daily UVR exposure (in average 2.4 ± 1.5 SEDs) the mean concentration of 25(OH)D3 increased (×1.24 ± 0.19) from 64.7 ± 13.3 to 79.3 ± 18.7 nmol/L (p < 0.001) [132].

It has to be noted that there are other positive UVB-induced actions of sunlight: in addition to 7-dehydrocholesterol, UVB photons can be absorbed by a range of chromophores, including DNA, membrane lipids, urocanic acid with subsequent effects on immune cells and secretion of neuropeptides such as β-endorphin and α-melanocyte stimulating hormone (MSH) [133,134]. On the other hand, it must be considered that patients with photodermatoses such as erythropoietic protoporphyria, for example, have to avoid solar or artificial UVR [135]. Consequently, vitamin D deficiency in patients with erythropoietic protoporphyria has been reported [135].

In summary, it can be concluded that UVB exposure of approximately 20–25% of the body surface (e.g., arms and legs) for 5–30 min (depending on time of day, season, latitude, and skin pigmentation) between the hours of 10 a.m. and 3 p.m. for white adults, 2–3 times per week, in spring, summer and autumn represents a promising strategy to obtain and maintain a sufficient vitamin D status [3,5,14]. As mentioned above, since only a few foods naturally contain vitamin D in substantial amounts (Tables 2 and 3), moderate UVB-exposure is an important vitamin D source for fulfilling the human body’s requirement for vitamin D. By sufficiently exposing the skin of a healthy person in spring, summer and autumn to solar UVR, no additional supply of vitamin D from the food or from supplements is necessary [39]. A longer exposure to the sun does not provide more vitamin D-associated health benefits but is associated with a higher risk of sunburn and skin cancer.

6. Conclusions: A Critical Appraisal of Strategies to Optimize Vitamin D Status in Germany

To obtain and maintain a sufficient vitamin D status, a healthy German adult with skin type I–III should be exposed to the sun of approximately 20–25% of the body surface (e.g., arms and legs) for 5 to 30 min between the hours of 10 a.m. and 3 p.m., 2–3x/week, in spring, summer and autumn without reaching his or her individual MED [3,5,14]. A longer exposure to the sun does not provide more vitamin D-associated health benefits. Although oral intake of vitamin D from both food and supplements leads to the same endproducts as UVB-dependent cutaneous vitamin D production, the bioavailability of the latter is not affected by potential malabsorption in the gut, for example. On the other hand, UVB-induced cutaneous vitamin D production only results in D3 analogs. A comparison is shown in Table 4.

In the absence of endogenous synthesis of vitamin D, the present guidelines of the German society for nutrition (Deutsche Gesellschaft für Ernährung (DGE) e.V.) and the D-A-CH society (i.e., Germany, Austria, Switzerland) recommend an intake of 400 IU (10 µg) vitamin D per day for infants and 800 IU (20 µg) per day for children, adolescents, adults, pregnant women and breast-feeding women [55].

Especially during the winter and autumn months, the population should regularly eat vitamin D-rich food, such as fish dishes, along with maintaining a healthy lifestyle in general.
Table 4. Comparison between cutaneous Vitamin D production and oral intake (includes food and supplemements).

| Parameter                  | Cutaneous Vitamin D Production                                                                 | Oral Intake (Food and Supplements)                                                                 |
|----------------------------|-------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------|
| Vitamin D compounds        | Exclusively D$_3$ analogues                                                                    | D$_3$ and D$_2$ analogues                                                                         |
| Skin damage                | Risk of sunburn, skin cancer and actinic damage                                                | Risk of intoxication (only by supplements with very high doses of vitamin D)                      |
| Risk of intoxication       | In healthy individuals, no risk for UVB-induced vitamin D intoxication                         | In healthy individuals, no risk for UVB-induced vitamin D intoxication                             |
| Accessibility              | Ubiquitously available in summer months                                                         | Low amount of vitamin D contents in food                                                           |
| Limitation factors         | MANY individual limitation factors such as age, pigmentation, season, geographic and meteorological factors | Reduced absorption in patients with malabsorption syndromes                                        |
| Other effects              | Synthesis of many vitamin D compounds with unknown physiologic relevance                       | -                                                                                                   |

Concerning vitamin D deficiency, the Endocrine Society Clinical Practice guideline suggests that all adults who are vitamin D deficient should be treated with 50,000 IU of vitamin D once a week for 8 week or its equivalent of 6000 IU of vitamin D daily to achieve a blood level of 25(OH)D above 75 nmol/L, followed by maintenance therapy of 1500-2000 IU per day [18]. However, the growing rate of vitamin D supplementation could increase costs and be harmful, especially if the tolerable upper intake level is exceeded (1000 IU per day in infants ages 0–6 months, 1500 IU per day in infants ages 6–12 months, 2500 IU per day in children ages 1–3 years, 3000 IU per day in children ages 4–8 years and 4000 IU per day in adolescents and adults [20,58]).

In the literature, there is currently no consistency regarding whether or not to measure 25(OH)D, with present recommendations being mostly based on expert opinions, resulting in a relatively low evidence level [15]. For example, Holick defined risk groups for the development of vitamin D deficiency. These groups include breastfeeding children up to 1 year without vitamin D supplementation, children from 1 through 18 years of age with inadequate sun exposure or supplementation or dark skin, adults with inadequate sun exposure or supplementation, a decreased 7-dehydrocholesterol level in skin because of aging (over 50 years), pregnant or lactating (fetal utilization, inadequate sun exposure or supplementation) women, patients with malabsorption syndromes (malabsorption of vitamin D, inadequate sun exposure or supplementation), patients with drug intake that activates steroid and xenobiotic receptors or drugs used in transplantation, patients with obesity, patients with nephrotic syndrome, patients with a higher stage of chronic kidney disease, patients with primary or tertiary hyperparathyroidism, as well as patients with granulomatous disorders and some lymphomas [14]. In these groups, preventive and maintenance measures to avoid vitamin D deficiency are recommended. These measures include a specific oral supplementation of vitamin D products for each risk group, if necessary [14]. In the groups of adults with inadequate sun exposure or supplementation, a decreased 7-dehydrocholesterol level in skin because of aging (over 50 years) and malabsorption syndromes, Holick recommends skin exposure to artificial UVB irradiation [14].

Taken together, it is important for most populations that individuals regularly consume vitamin D-rich food, especially during the winter and autumn months. Individuals with the risk factors mentioned above as well as hypovitaminosis D in the past, osteoporosis, osteomalacia, dark pigmentation (skin type from III), no access to sunlight (e.g., patients confined to bed), photodermatosis with the necessity of avoiding UVR or unhealthy lifestyle in general should be monitored by measuring and supplementing vitamin D$_3$ if necessary. The other part of the population as well as patients with malabsorption syndromes should be regularly moderately exposed to UVR (approximately 20–25% of the body surface (e.g., arms and legs) for 5–30 min (depending on time of day, season, latitude, and skin pigmentation) between the hours of 10 a.m. and 3 p.m., 2–3 times per week, in spring, summer and autumn) to ensure the health benefits of UVB exposure with minimal increased risk for skin cancer.
Author Contributions: Conceptualization, R.S., T.V. and J.R.; methodology, R.S. T.V. and J.R.; writing—original draft preparation, R.S.; writing—review and editing, T.V. and J.R.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Diamond, J. Geography and skin colour. *Nature* 2005, 435, 283–284. [CrossRef] [PubMed]
2. Jablonski, N.G.; Chaplin, G. The evolution of human skin coloration. *J. Hum. Evol.* 2000, 39, 57–106. [CrossRef] [PubMed]
3. Holick, M.F. Vitamin D: A D-lightful solution for health. *J. Investig. Med.* 2011, 59, 872–880. [CrossRef] [PubMed]
4. Saternus, R.; Vogt, T.; Reichrath, J. Skin Types, Skin Pigmentation and Melanin Synthesis: Important Tools of Human Skin to Adapt at UV-Radiation. *Akt. Dermatol.* 2018, 44, 210–215.
5. Holick, M.F.; Vitamin, D. A D-Lightful health perspective. *Nutr. Rev.* 2008, 66, 182–194. [CrossRef]
6. Bjelakovic, G.; Gluud, L.L.; Nikolova, D.; Whitfield, K.; Wetterslev, J.; Simonetti, R.G.; Bjelakovic, M.; Gluud, C. Vitamin D Supplementation for prevention of mortality in adults. *Cochrane Database Syst. Rev.* 2014. [CrossRef]
7. Manson, J.E.; Cook, N.R.; Lee, I.M.; Christen, W.; Bassuk, S.S.; Mora, S.; Gibson, H.; Gordon, D.; Copeland, T.; D’Agostino, D.; et al. Supplements and Prevention of Cancer and Cardiovascular Disease. *N. Engl. J. Med.* 2019, 380, 33–44. [CrossRef]
8. Pittas, A.G.; Dawson-Hughes, B.; Sheehan, P.; Ware, J.H.; Knowler, W.C.; Aroda, V.R.; Brodsky, I.; Ceglia, L.; Chadha, C.; Chatterjee, R.; et al. Supplementation and Prevention of Type 2 Diabetes. *N. Engl. J. Med.* 2019, 381, 520–530. [CrossRef]
9. Slominski, A.T.; Kim, T.K.; Janjetovic, Z.; Brożyna, A.A.; Żmijewski, M.A.; Xu, H.; Sutter, T.R.; Tuckey, R.C.; Jetten, A.M.; Crossman, D.K. Differential and Overlapping Effects of 20,23(OH)2D3 and 1,25(OH)2D3 on Gene Expression in Human Epidermal Keratinocytes. Identification of AhR as an Alternative Receptor for 20,23(OH)2D3. *Int. J. Mol. Sci.* 2018, 19, 3072. [CrossRef]
10. Slominski, A.T.; Kim, T.K.; Hobrath, J.V.; Oak, A.S.W.; Tang, E.K.Y.; Tieu, E.W.; Li, W.; Tuckey, R.C.; Jetten, A.M. Endogenously produced nonclassical vitamin D hydroxy-metabolites act as “biased” agonists on VDR and inverse agonists on RORα and RORγ. *J. Steroid Biochem. Mol. Biol.* 2017, 173, 42–56. [CrossRef]
11. Slominski, A.T.; Kim, T.K.; Takeda, Y.; Janjetovic, Z.; Brożyna, A.A.; Skobowiat, C.; Wang, J.; Postlethwaite, A.; Li, W.; Tuckey, R.C.; et al. RORα and RORγ are expressed in human skin and serve as receptors for endogenously produced noncalcemic 20-hydroxy and 20,23-dihydroxyvitamin D. *FASEB J.* 2014, 28, 2775–2789. [CrossRef] [PubMed]
12. Mostafa, W.Z.; Hegazy, R.A. Vitamin D and the skin: Focus on a complex relationship: A review. *J. Adv. Res.* 2015, 6, 793–804. [CrossRef] [PubMed]
13. Uhoda, I.; Quatresooz, P.; Rorive, A.; Piérand-Franchimont, C.; Piérand, G.E. Skin cancer and sunlight. *Rev. Med. Liege* 2005, 60 (Suppl. 1), 88–98.
14. Holick, M.F. Vitamin D deficiency. *N. Engl. J. Med.* 2007, 357, 266–282. [CrossRef] [PubMed]
15. Pilz, S.; Zittermann, A.; Trummer, C.; Theiler-Schewitz, V.; Lerchbaum, E.; Keppel, M.H.; Grübler, M.R.; März, W.; Pandis, M. Vitamin D testing and treatment: A narrative review of current evidence. *Endocr. Connect.* 2019, 8, R27–R43. [CrossRef] [PubMed]
16. Holick, M.F. Vitamin D status: Measurement, interpretation and clinical application. *Ann. Epidemiol.* 2009, 19, 73–78. [CrossRef] [PubMed]
17. Gesundheitsamt Bremen. Available online: https://www.gesundheitsamt.bremen.de/referenzwerte_fuer_vitamin_d_5691 (accessed on 25 August 2019).
18. Holick, M.F.; Binkley, N.C.; Bischoff-Ferrari, H.A.; Gordon, C.M.; Hanley, D.A.; Heaney, R.P.; Murad, M.H.; Weaver, C.M. Evaluation, treatment, and prevention of vitamin D deficiency: An Endocrine Society clinical practice guideline. *J. Clin. Endocrinol. Metab.* 2011, 96, 1911–1930. [CrossRef]
19. Marcinkowska-Suchowierska, E.; Kupisz-Urbańska, M.; Łukaszkiewicz, J.; Pudowski, P.; Jones, G. Vitamin D Toxicity—A Clinical Perspective. *Front. Endocrinol.* 2018, 9, 550. [CrossRef]
20. Ross, A.C.; Manson, J.E.; Abrams, S.A.; Aloia, J.F.; Brannon, P.M.; Clinton, S.K.; Durazo-Arvizu, R.A.; Gallagher, J.C.; Gallo, R.L.; Jones, G.; et al. The 2011 Report on Dietary Reference Intakes for Calcium and Vitamin D from the Institute of Medicine: What Clinicians Need to Know. *J. Clin. Endocrinol. Metab.* **2011**, *96*, 53–58. [CrossRef]

21. Melamed M I Michos, E.D.; Post, W.; Astor, B. 25-hydroxy Vitamin D Levels and the Risk of Mortality in the General Population. *Arch. Intern. Med.* **2008**, *168*, 1629–1637. [CrossRef]

22. Grant, W.B.; Karras, S.N.; Bischoff-Ferrari, H.A.; Annweiler, C.; Boucher, B.J.; Juzeniene, A.; Garland, C.F.; Holick, M.F. Do studies reporting 'U'-shaped serum 25-hydroxyvitamin D–health outcome relationships reflect adverse effects? *Dermato-endocrinology* **2016**, *8*, e1187349. [CrossRef] [PubMed]

23. McDonnell, S.L.; Baggerly, C.A.; French, C.B.; Baggerly, L.L.; Garland, C.F.; Gorham, E.D.; Hollis, B.W.; Trump, D.L.; Lappe, J.M.; Heaney, R.P. Serum 25-Hydroxyvitamin D Concentrations ≥60 ng/ml Are Associated with >65% Lower Cancer Risk: Pooled Analysis of Randomized Trial and Prospective Cohort Study. *PLoS ONE* **2016**, *11*, e0152441. [CrossRef]

24. McDonnell, S.L.; Baggerly, K.A.; Baggerly, C.A.; Aliano, J.L.; French, C.B.; Baggerly, L.L.; Ebeling, M.D.; Rittenberg, C.S.; Goodier, C.G.; Mateus Niño, J.F.; et al. Maternal 25(OH)D concentrations ≥40 ng/mL associated with 60% lower preterm birth risk among general obstetrical patients at an urban medical center. *PLoS ONE* **2017**, *12*, e0180483. [CrossRef] [PubMed]

25. McDonnell, S.L.; Baggerly, C.; French, C.B.; Baggerly, L.L.; Garland, C.F.; Gorham, E.D.; Lappe, J.M.; Heaney, R.P. Serum 25-Hydroxyvitamin D Concentrations ≥40 ng/ml Are Associated with >65% Lower Cancer Risk: Pooled Analysis of Randomized Trial and Prospective Cohort Study. *PLoS ONE* **2016**, *11*, e0152441. [CrossRef]

26. Mirhosseini, N.; Vatanparast, H.; Kimball, S.M. The Association between Serum 25(OH)D Status and Blood Pressure in Participants of a Community-Based Program Taking Vitamin D Supplements. *Nutrients* **2017**, *9*, 1244. [CrossRef]

27. Garland, C.F.; Kim, J.J.; Mohr, S.B.; Gorham, E.D.; Grant, W.B.; Giovannucci, E.L.; Baggerly, L.; Ho, P.Y.; Ramsdell, J.W.; Zeng, K.; et al. Meta-analysis of all-cause mortality according to serum 25-hydroxyvitamin D. *Am. J. Public Health* **2014**, *104*, e43–e50. [CrossRef]

28. Hollis, B.W. Comparison of Commercially Available 125I-based RIA Methods for the Determination of Circulating 25-Hydroxyvitamin D. *Clin. Chem.* **2000**, *46*, 1657–1661.

29. Le Gofa, C.; Cavaliera, E.; Souberbielle, J.C.; González-Antuñaa, A.; Delvinc, E. Measurement of circulating 25-hydroxyvitamin D. A historical review. *Pract. Lab. Med.* **2015**, *2*, 1–14. [CrossRef]

30. Hintzpeter, B.; Volker, D. Vitamin D-Versorgung in Deutschland. In *Vitamin D und Prävention Ausgewählter Chronischer Krankheiten*; der Deutschen Gesellschaft für Ernährung: Bonn, Germany, 2011.

31. Hintzpeter, B.; Mensink, G.B.; Thierfelder, W.; Müller, M.J.; Scheidt-Nave, C. Vitamin D status and health correlates among German adults. *Eur. J. Clin. Nutr.* **2008**, *62*, 1079–1089. [CrossRef]

32. Rabenberg, M.; Scheidt-Nave, C.; Busch, M.A.; Rieckmann, N.; Hintzpeter, B.; Mensink, G.B.M. Vitamin D status among adults in Germany—Results from the German Health Interview and Examination Survey for Adults (DEGS1). *BMJ Public Health* **2015**, *15*, 641. [CrossRef]

33. Diekmann, R.; Winning, K.; Bauer, J.M.; Uter, W.; Stehle, P.; Lesser, S.; Bertsch, T.; Sieber, C.C.; Volkert, D. Vitamin D status and physical function in nursing home residents: A 1-year observational study. *Zeitschrift für Gerontologie und Geriatrie* **2013**, *46*, 403–409. [CrossRef] [PubMed]

34. Sægård, T.; Möller, L.; Becher, H.; Jöckel, K.H.; Schlaud, M.; Willich, S.N.; Keil, T. Serum vitamin D levels in Berliners of Turkish descent—A cross-sectional study. *BMJ Public Health* **2019**, *19*, 119. [CrossRef] [PubMed]
38. Hintzpeter, B.; Scheidt-Nave, C.; Müller, M.J.; Schenk, L.; Mensink, G.B. Higher prevalence of vitamin D deficiency is associated with immigrant background among children and adolescents in Germany. *J. Nutr.* 2008, 138, 1482–1490. [CrossRef]

39. Norman, A.W. From vitamin D to hormone: Fundamentals of the vitamin D endocrine system essential for good health. *Am. J. Clin. Nutr.* 2008, 88, 4915–4995. [CrossRef]

40. Chen, T.C.; Lu, Z.; Holick, M.F. Photobiology of Vitamin D. In *Nutrition and Health: Vitamin D Physiology, Molecular Biology, and Clinical Applications*, 2nd ed.; Holick, M.F., Ed.; Springer: New York, NY, USA, 2010; pp. 35–60.

41. Lehmann, B.; Querings, K.; Reichrath, J. Vitamin D and skin. New aspects for dermatology. *Exp. Dermatol.* 2004, 13 (Suppl. 4), 11–15. [CrossRef]

42. Holick, M.F. Resurrection of vitamin D deficiency and rickets. *J. Clin. Investig.* 2006, 116, 2062–2072. [CrossRef]

43. Wang, T.J.; Zhang, F.; Richards, J.B.; Kestenbaum, B.; van Meurs, J.B.; Berry, D.; Kiel, D.P.; Streethen, E.A.; Ohlsson, S.; Koller, D.L.; et al. Common genetic determinants of vitamin D insufficiency. A genome-wide association study. *Lancet* 2010, 376, 180–188. [CrossRef]

44. Shinkyo, R.; Sakaki, T.; Kamakura, M.; Ohta, M.; Inouye, K. Metabolism of vitamin D by human microsomal CYP2R1. *Biochem. Biophys. Res. Commun.* 2004, 324, 451–457. [CrossRef]

45. Chun, R.F.; Peercy, B.E.; Orwoll, E.S.; Nielson, C.M.; Adams, J.S.; Hewison, M. Vitamin D and DBP: The free hormone hypothesis revisited. *J. Steroid Biochem. Mol. Biol.* 2014, 144, 132–137. [CrossRef]

46. Negri, A.L. Proximal tubule endocytic apparatus as the specific renal uptake mechanism for vitamin D-binding protein/25(OH)D3 complex. *Nephrology* 2006, 11, 510–515. [CrossRef] [PubMed]

47. Leheste, J.R.; Rolinski, B.; Vorum, H.; Hilpert, J.; Nykjaer, A.; Jacobsen, C.; Aucouturier, P.; Moskaug, J.O.; Otto, A.; Christensen, E.I.; et al. Megalin Knockout Mice as an Animal Model of Low Molecular Weight Proteinuria. *Am. J. Pathol.* 1999, 155, 1361–1370. [CrossRef]

48. Ying, H.Q.; Sun, H.L.; He, B.S.; Pan, Y.Q.; Fang, F.; Deng, Q.W.; Chen, J.; Liu, X.; Wang, S.K. Circulating vitamin D binding protein, total, free and bioavailable 25-hydroxyvitamin D and risk of colorectal cancer. *Sci. Rep.* 2015, 5, 7956. [CrossRef] [PubMed]

49. Slominski, A.T.; Kim, T.K.; Li, W.; Postlethwaite, A.; Tieu, E.W.; Tang, E.K.; Tuckey, R.C. Detection of novel CYP11A1 derived secosteroids in the human epidermis and serum and pig adrenal gland. *Sci. Rep.* 2015, 5, 14875. [CrossRef] [PubMed]

50. Slominski, A.; Zjawiony, J.; Wortsman, J.; Semak, I.; Stewart, J.; Pisarchik, A.; Sweatman, T.; Marcos, J.; Dunbar, C.; CTuckey, R. A novel pathway for sequential transformation of 7-dehydrocholesterol and expression of the P450sc system in mammalian skin. *Eur. J. Biochem.* 2004, 271, 4178–4188. [CrossRef]

51. Miller, W.L.; Auchus, R.J. The Molecular Biology, Biochemistry, and Physiology of Human Steroidogenesis and Its Disorders. *Endocr. Rev.* 2011, 32, 81–151. [CrossRef]

52. Guryev, O.; Carvalho, R.A.; Usanov, S.; Gilep, A.; Estabrook, R.W. A pathway for the metabolism of vitamin D3: Unique hydroxylated metabolites formed during catalysis with cytochrome P450sc (CYP11A1). *Proc. Natl. Acad. Sci. USA* 2003, 100, 14754–14759. [CrossRef]

53. Slominski, A.T.; Li, W.; Kim, T.K.; Semak, I.; Wang, J.; Zjawiony, J.K.; Tuckey, R.C. Novel activities of CYP11A1 and their potential physiological significance. *J. Steroid Biochem. Mol. Biol.* 2005, 151, 25–37. [CrossRef]

54. Rosen, C.J.; Adams, J.S.; Bikle, D.D.; Demay, M.B.; Manson, J.E.; Murad, M.H.; Kovacs, C.S. The nonskeletal effects of vitamin D: An Endocrine Society scientific statement. *Endocr. Rev.* 2012, 33, 456–492. [CrossRef]

55. Deutsche Gesellschaft für Ernährung e.V. 2019. Available online: https://www.dge.de/wissenschaft/ referenzwerte/vitamin-d/ (accessed on 30 October 2019).

56. Bundesforschungsinstitut für Ernährung und Lebensmittel. *Nationale Verzehrsstudie II Max Rubner-Institut Ergebnisbericht, Teil 2*; Bundesforschungsinstitut für Ernährung und Lebensmittel: Karlsruhe, Germany, 2008.

57. Reinehr, T.; Schnabel, D.; Wabitsch, M.; Bechthold-Dalla Pozzalla, S.; Bührer, C.; Heidtmann, B.; Jochum, F.; Kauth, T.; Körner, A.; Mihatsch, W.; et al. Vitamin D-Supplementierung jenseits des zweiten Lebensjahres—Gemeinsame Stellungnahme der Ernährungskommission der Deutschen Gesellschaft für Kinder- und Jugenmedizin (DGKJe.V.) und der Deutschen Gesellschaft für Kinderendokrinologie und Diabetologie (DGKEDe.V.). *Monatsschrift Kinderheilkunde* 2018, 166, 814–822.
58. IOM (Institute of Medicine). *Dietary Reference Intakes for Calcium and Vitamin D;* The National Academies Press: Washington, DC, USA, 2011.

59. Keegan, R.J.H.; Lu, Z.; Bogusz, J.M.; Williams, J.E.; Holick, M.F. Photobiology of vitamin D in mushrooms and its bioavailability in humans. *Dermato-Endocrinology* 2013, 1, 165–176. [CrossRef] [PubMed]

60. Maurya, V.K.; Aggarwa, M. Factors influencing the absorption of vitamin D in GIT: An overview. *J. Food Sci. Technol.* 2017, 54, 3753–3765. [CrossRef] [PubMed]

61. Reboul, E.; Goncalves, A.; Comera, C.; Bott, R.; Nowicki, M.; Landrier, J.F.; Jourdheuil-Rahmani, D.; Dufour, C.; Collet, X.; Borel, P. Vitamin D intestinal absorption is not a simple passive diffusion: Evidences for involvement of cholesterol transporters. *Mol. Nutr. Food Res.* 2011, 55, 691–702. [CrossRef] [PubMed]

62. Niramitmahapanya, S.; Harris, S.S.; Dawson-Hughes, B. Type of Dietary Fat Is Associated with the 25-Hydroxyvitamin D Increment in Response to Vitamin D Supplementation. *J. Clin. Endocrinol. Metab.* 2011, 96, 3170–3174. [CrossRef]

63. Holick, M.F. Cancer, sunlight and Vitamin D. *J. Clin. Trial Transl. Endocrinol.* 2014, 5, 179–186. [CrossRef]

64. Hymøller, L.; Jensen, S.K. 25-hydroxyvitamin D circulates in different fractions of calf plasma if the parent compound is vitamin D2 or vitamin D3, respectively. *J. Dairy Res.* 2016, 83, 67–71. [CrossRef]

65. Hammami, M.M.; Yusuf, A. Di, S.; Worm, M. Pharmacokinetic Evaluation of a Vitamin D2 Incement in Response to Vitamin D Supplementation. *J. Clin. Endocrinol. Metab.* 2011, 96, 3170–3174. [CrossRef]

66. Tuckey, R.C.; Nguyen, M.N.; Chen, J.; Slominski, A.T.; Baldisseri, D.M.; Tieu, E.W.; Zjawiony, J.K.; Li, W. Human Cytochrome P450scc (CYP11A1) Catalyzes Epoxide Formation with Ergosterol. *Drug Metab. Dispos.* 2012, 40, 436–444. [CrossRef]

67. Wilson, L.R.; Tripkovic, L.; Hart, K.H.; Lanham-New, S.A. Vitamin D deficiency as a public health issue: Using vitamin D2 or vitamin D3 in future fortification strategies. *Proc. Nutr. Soc.* 2017, 76, 392–399. [CrossRef]

68. Armas, L.A.; Hollis, B.W.; Heaney, R.P. Vitamin D2 is much less effective than vitamin D3 in humans. *J. Clin. Endocrinol. Metab.* 2004, 89, 5387–5391. [CrossRef] [PubMed]

69. Binkley, N.; Gemar, D.; Engelke, J.; Gangnon, R.; Ramamurthy, R.; Krueger, D.; Drezner, M.K. Evaluation of ergocalciferol or cholecalciferol dosing, 1600 IU daily or 50,000 IU monthly in older adults. *J. Clin. Endocrinol. Metab.* 2011, 96, 981–988. [CrossRef]

70. Tripkovic, L.; Wilson, L.R.; Hart, K.; Johnsen, S.; de Lusignan, S.; Smith, C.P.; Bucca, G.; Pens, S.; Chope, G.; Elliott, R.; et al. Daily supplementation with 15 µg vitamin D2 compared with vitamin D3 to increase wintertime 25-hydroxyvitamin D status in healthy South Asian and white European women: A 12-wk randomized, placebo-controlled food-fortification trial. *Am. J. Clin. Nutr.* 2017, 106, 481–490. [CrossRef] [PubMed]

71. Martineau, A.R.; Thummel, K.E.; Wang, Z.; Jolliffe, D.A.; Boucher, B.J.; Griffin, S.J.; Forouhi, N.G.; Hitman, G.A. Differential effects of oral boluses of vitamin D2 versus vitamin D3 on vitamin D metabolism: A randomized controlled trial. *J. Clin. Endocrinol. Metab.* 2019, 104, 5831–5839. [CrossRef] [PubMed]

72. Martineau, A.R.; Thummel, K.E.; Wang, Z.; Jolliffe, D.A.; Boucher, B.J.; Griffin, S.J.; Forouhi, N.G.; Hitman, G.A. Differential effects of oral boluses of vitamin D2 versus vitamin D3 on vitamin D metabolism: A randomized controlled trial. *J. Clin. Endocrinol. Metab.* 2019, 104, 5831–5839. [CrossRef] [PubMed]

73. Hammami, M.; Amr, S.; Hammami, S.; Yusuf, A. Vitamin D deficiency as a public health issue: Using vitamin D2 or vitamin D3 in future fortification strategies. *Proc. Nutr. Soc.* 2017, 76, 392–399. [CrossRef]

74. Binkley, N.; Gemar, D.; Engelke, J.; Gangnon, R.; Ramamurthy, R.; Krueger, D.; Drezner, M.K. Evaluation of ergocalciferol or cholecalciferol dosing, 1600 IU daily or 50,000 IU monthly in older adults. *J. Clin. Endocrinol. Metab.* 2011, 96, 981–988. [CrossRef]

75. Beitz, R.; Mensink, G.B.M.; Rams, S.; Döring, A. Vitamin-und Mineralstoffsupplementierung in Deutschland. *Bundesgesundheitsbl.-Gesundheitsforsch.-Gesundheitsschutz* 2004, 47, 1057–1065. [CrossRef]

76. Wylon, K.; Drozdenko, G.; Krannich, A.; Heine, G.; Do, S.; Worm, M. Pharmacokinetic Evaluation of a Single Intramuscular High Dose versus an Oral LongTerm Supplementation of Cholecalciferol. *PLoS ONE* 2017, 12, e0169620. [CrossRef]
77. Gupta, N.; Farooqui, K.J.; Batra, C.M.; Marwaha, R.K.; Mithal, A. Effect of oral versus intramuscular Vitamin D replacement in apparently healthy adults with Vitamin D deficiency. *Indian J. Endocrinol. Metab.* 2017, 21, 131–136.

78. Garland, C.F.; French, C.B.; Baggerl, L.L.; Heaney, R.P. Vitamin D Supplement Doses and Serum 25-Hydroxyvitamin D in the Range Associated with Cancer Prevention. *Anticancer Res.* 2011, 31, 607–612.

79. Zittermann, A.; Ernst, J.B.; Gummert, J.F.; Bürgermann, J. Vitamin D supplementation, body weight and human serum 25-hydroxyvitamin D response: A systematic review. *Eur. J. Nutr.* 2014, 53, 367–374. [CrossRef] [PubMed]

80. De Niet, S.; Coffiner, M.; Da Silva, S.; Jandrain, B.; Soubberbielle, J.C.; Cavalier, E. A Randomized Study to Compare a Monthly to a Daily Administration of Vitamin D3 Supplementation. *Nutrients* 2018, 10, 659. [CrossRef] [PubMed]

81. Kearns, M.D.; Alvarez, J.A.; Tangpricha, V. Large, single-dose, oral vitamin D supplementation in adult populations: A systematic review. *Endocr. Pract.* 2014, 20, 341–351. [CrossRef] [PubMed]

82. Wu, F.; Xiao, C.; Aitken, D.; Jones, G.; Winzenberg, T. The optimal dosage regimen of vitamin D supplementation for correcting deficiency in adolescents: A pilot randomized controlled trial. *Eur. J. Clin. Nutr.* 2018, 72, 534–540. [CrossRef]

83. Takacs, I.; Toth, B.E.; Szekeres, L.; Szabo, B.; Bakos, B.; Lakatos, P. Randomized clinical trial to comparing efficacy of daily, weekly and monthly administration of vitamin D3. *Endocrine* 2017, 55, 60–65. [CrossRef]

84. Rossini, M.; Gatti, D.; Viapiana, O.; Fracassi, E.; Idolazzi, L.; Zanoni, S.; Adami, S. Short-Term Effects on Bone Turnover Markers of a Single High Dose of Oral Vitamin D3. *J. Clin. Endocrinol. Metab.* 2012, 97, E622–E626. [CrossRef]

85. Debasish, M. Vitamin D toxicity. *Indian J. Endocrinol. Metab.* 2012, 16, 295–296.

86. Gallagher, J.; Smith, L.M.; Yalamanchili, V. Incidence of hypercalciuria and hypercalcemia during vitamin D and calcium Supplementation in older women. *Menopause* 2014, 21, 1173–1180. [CrossRef]

87. Marcus, J.F.; Shalev, S.M.; Harris, C.A.; Goodin, D.S.; Josephson, S.A. Severe Hypercalcemia Following Vitamin D Supplementation in a Patient with Multiple Sclerosis—A Note of Caution. *Arch. Neurol.* 2012, 69, 129–132. [CrossRef]

88. Letavernier, E.; Daudon, M. Vitamin D, Hypercalciuria and Kidney Stones. *Nutrients* 2018, 10, 366. [CrossRef]

89. Malihi, Z.; Wu, Z.; Steward, A.W.; Lawes, C.M.M.; Scragg, R. Hypercalciemia, hypercalcemia, and kidney stones in long-term studies of vitamin D supplementation: A systematic review and meta-analysis. *Am. J. Clin. Nutr.* 2016, 104, 1039–1051. [CrossRef] [PubMed]

90. Kahwati, L.C.; Weber, R.P.; Pan, H.; Gourlay, M.; Coker-Schwimmer, M.; Viswanathan, M. Vitamin D, Calcium, or Combined Supplementation for the Primary Prevention of Fractures in Community-Dwelling Adults: Evidence Report and Systematic Review for the US Preventive Services Task Force. *JAMA* 2018, 319, 1600–1612. [CrossRef] [PubMed]

91. Ferraro, P.M.; Taylor, E.N.; Gambaro, G.; Curhan, G.C. Vitamin D Intake and the Risk of Incident Kidney Stones. *J. Urol.* 2017, 197, 405–410. [CrossRef] [PubMed]

92. Malihi, Z.; Wu, Z.; Lawes, C.M.M.; Scragg, R. Adverse events from large dose vitamin D supplementation taken for one year or longer. *J. Steroid Biochem. Mol. Biol.* 2019, 197, 29–37. [CrossRef] [PubMed]

93. Mazess, R.B.; Elangovan, L. A review of intravenous versus oral vitamin D hormone therapy in hemodialysis patients. *Clin. Nephrol.* 2003, 59, 319–325. [CrossRef] [PubMed]

94. Dunlop, E.; Cunningham, J.; Sherriff, J.L.; Lucas, R.M.; Greenfield, H.; Arcot, J.; Strobel, N.; Black, L.J. Vitamin D3 and 25-Hydroxyvitamin D3 Content of Retail White Fish and Eggs in Australia. *Nutrients* 2017, 9, 647. [CrossRef] [PubMed]

95. Liu, J.; Arcot, J.; Cunningham, J.; Greenfield, H.; Hsu, J.; Padula, D.; Strobel, N.; Fraser, D.R. New data for vitamin D in Australian foods of animal origin: Impact on estimates of national adult vitamin D intakes in 1995 and 2011-13. *Asia Pac. J. Clin. Nutr.* 2015, 24, 464–471.

96. Crowe, F.L.; Steur, M.; Appleby, P.N.; Travis, R.C.; Key, T.J. Plasma concentrations of 25-hydroxyvitamin D in meat eaters, fish eaters, vegetarians and vegans: Results from the EPIC-Oxford study. *Public Health Nutr.* 2011, 14, 340–346. [CrossRef]

97. Grant, W.B.; Fakhoury, H.M.A.; Karras, S.N.; Al Anouti, F.; Bhattoo, H.P. Variations in 25-Hydroxyvitamin D in Countries from the Middle East and Europe: The Roles of UVB Exposure and Diet. *Nutrients* 2019, 11, 2065. [CrossRef]
98. Kuchuk, N.O.; van Schoor, N.M.; Pluijm, S.M.; Chines, A.; Lips, P. Vitamin D status, parathyroid function, bone turnover, and BMD in postmenopausal women with osteoporosis: Global perspective. *J. Bone Miner. Res.* 2009, 24, 693–701. [CrossRef]

99. Mensink, G.B.M.; Heseker, H.; Stahl, A.; Richter, A.; Vohmann, C. Die aktuelle Nährstoffversorgung von Kindern und Jugendlichen in Deutschland. Ergebnisse aus EsKiMo. *Ernährungs Umschau* 2017, 11, 636–646.

100. Pilz, S.; März, W.; Cashman, K.D.; Kiely, M.E.; Whiting, S.J.; Holick, M.F.; Grant, W.B.; Pludowski, P.; Hiligsmann, M.; Trummer, C.; et al. Rationale and Plan for Vitamin D Food Fortification: A Review and Guidance Paper. *Front. Endocrinol.* 2015, 9, 373. [CrossRef] [PubMed]

101. Bromage, S.; Gannaa, D.; Rich-Edwards, J.W.; Rosner, B.; Bater, J.; Fawzi, W.W. Projected effectiveness of mandatory industrial fortification of wheat flour, milk, and edible oil with multiple micronutrients among Mongolian adults. *PLoS ONE* 2018, 13, e0201230. [CrossRef] [PubMed]

102. O’Neill, C.M.; Kazantzidis, A.; Kiely, M.; Cox, L.; Meadows, S.; Goldberg, G.; Prentice, A.; Kift, R.; Webb, A.R.; Cashman, K.D. A predictive model of serum 25-hydroxyvitamin D in UK white as well as black and Asian minority ethnic population groups for application in food fortification strategy development towards vitamin D deficiency prevention. *J. Steroid Biochem. Mol. Biol.* 2017, 173, 245–252. [CrossRef]

103. Lips, P.; Cashman, K.D.; Lamberg-Allardt, C.; Bischoff-Ferrari, H.A.; Obermayer-Pietsch, B.R.; Bianchi, M.; Stepan, J.; El-Hajj Fuleihan, G.; Bouillon, R. Management of endocrine disease: Vitamin D status in European and Middle East countries and strategies to prevent vitamin D deficiency; a position statement of the European Calcified Tissue Society. *Eur. J. Endocrinol.* 2019. [CrossRef]

104. Calvo, M.S.; Whiting, S.J. Survey of current vitamin D food fortification practices in the United States and Canada. *J. Steroid Biochem. Mol. Biol.* 2013, 136, 211–213. [CrossRef]

105. Jääskeläinen, T.; Ilkkonen, S.T.; Lundqvist, A.; Erkkola, M.; Koskela, T.; Dowling, K.G.; Hull, G.L.; Kröger, H.; Karppinen, J.; et al. The positive impact of general vitamin D food fortification policy on vitamin D status in a representative adult Finnish population: Evidence from an 11-y follow-up based on standardized 25-hydroxyvitamin D data. *Am. J. Clin. Nutr.* 2017, 105, 1512–1520. [CrossRef]

106. Grønborg, I.M.; Tetens, I.; Christensen, T.; Andersen, E.W.; Jakobsen, J.; Kiely, M.; Cashman, K.D.; Andersen, R. Vitamin D-fortified foods improve wintertime vitamin D status in women of Danish and Pakistani origin living in Denmark: A randomized controlled trial. *Eur. J. Nutr.* 2019. [CrossRef]

107. Grønborg, I.M.; Tetens, I.; Ege, M.; Christensen, T.; Andersen, E.W.; Andersen, R. Modelling of adequate and safe vitamin D intake in Danish women using different fortification and supplementation scenarios to inform fortification policies. *Eur. J. Nutr.* 2019, 58, 227–232. [CrossRef]

108. Kiely, M.; Cashman, K.D. Summary Outcomes of the ODIN Project on Food Fortification for Vitamin D Deficiency Prevention. *Int. J. Environ. Res. Public Health* 2018, 15, 2342. [CrossRef]

109. Keller, A.; Stougård, M.; Frederiksen, P.; Thorsteinsdottir, F.; Vogt, T.; Reichrath, J. A closer look at evolution: Variants (SNPs) of genes involved in skin pigmentation, including EXOC2, TYR, TYRP1, and DCT, are associated with 25(OH)D serum concentration. *Endocrinology* 2015, 156, 39–47. [CrossRef]

110. Jablonski, N.G.; Chaplin, G. Colloquium paper: Human skin pigmentation as an adaptation to UV radiation. *Photochem. Photobiol.* 2005, 81, 1287–1290. [CrossRef]

111. Bais, A.F.; Lucas, R.M.; Bormann, J.F.; Williamson, C.E.; Sulzberger, B.; Austin, A.T.; Wilson, S.R.; Andrady, A.L.; Bernhard, G.; McKenzie, R.L.; et al. Environmental effects of ozone depletion, UV radiation and interactions with climate change: UNEP Environmental Effects Assessment Panel, update 2017. *Photochem. Photobiol. Sci.* 2018, 17, 127–179. [CrossRef] [PubMed]

112. Engelsen, O.; Brustad, M.; Aksnes, L.; Lund, E. Daily duration of vitamin D synthesis in human skin with relation to latitude, total ozone, altitude, ground cover, aerosols and cloud thickness. *Photochem. Photobiol. 2005, 81, 1287–1290. [CrossRef]
115. Rossberg, W.; Saternus, R.; Wagenpfeil, S.; Kleber, M.; März, W.; Reichrath, S.; Vogt, T.; Reichrath, J. Human Pigmentation, Cutaneous Vitamin D Synthesis and Evolution: Variants of Genes (SNPs) Involved in Skin Pigmentation Are Associated with 25(OH)D Serum Concentration. Anticancer Res. 2016, 36, 1429–1437.

116. Yeum, K.J.; Dawson-Hughes, B.; Joo, N.S. Fat Mass Is Associated with Serum 25-Hydroxyvitamin D Concentration Regardless of Body Size in Men. Nutrients 2018, 10, 850. [CrossRef]

117. Jager, N.; Schöpe, J.; Wagenpfeil, S.; Bocionek, P.; Saternus, R.; Vogt, T.; Reichrath, J. The Impact of UV-dose, Body Surface Area Exposed and Other Factors on Cutaneous Vitamin D Synthesis Measured as Serum 25(OH)D Concentration: Systematic Review and Meta-analysis. Anticancer Res. 2018, 38, 1165–1171.

118. Xiang, F.; Lucas, R.; de Gruijl, F.; Norva, M. A systematic review of the influence of skin pigmentation on changes in the concentrations of vitamin D and 25-hydroxyvitamin D in plasma/serum following experimental UV irradiation. Photochem. Photobiol. Sci. 2015, 14, 2138–2146. [CrossRef]

119. Hakim, O.A.; Hart, K.; McCabe, P.; Berry, J.; Francesca, R.; Rhodes, L.E.; Spyrou, N.; Alfuraih, A.; Lanham-New, S. Vitamin D production in UK Caucasian and South Asian women following UVR exposure. J. Steroid Biochem. Mol. Biol. 2016, 164, 223–229. [CrossRef] [PubMed]

120. Matsuoka, L.Y.; Ide, L.; Wortsman, J.; MacLaughlin, J.A.; Holick, M.F. Sunscreens suppress cutaneous vitamin D3 synthesis. J. Clin. Endocrinol. Metab. 1987, 64, 1165–1168. [CrossRef]

121. Young, A.R.; Narbutt, J.; Harrison, G.I.; Lawrence, K.P.; Bell, M.; O’Connor, C.; Olsen, P.; Grys, K.; Baczynska, K.A.; Rogowski-Tylman, M.; et al. Optimal sunscreen use, during a sun holiday with a very high ultraviolet index, allows vitamin D synthesis without sunburn. Br. J. Dermatol. 2019. [CrossRef] [PubMed]

122. Passeron, T.; Bouillon, R.; Callender, V.; Cestari, T.; Diepgen, T.L.; Green, A.C.; van der Pols, J.C.; Bernard, B.A.; Ly, F.; Bernerd, F.; et al. Sunscreen photoprotection and vitamin D status. Br. J. Dermatol. 2019. [CrossRef] [PubMed]

123. Holick, M.F.; MacLaughlin, J.A.; Doppelt, S.H. Regulation of cutaneous previtamin D3 photosynthesis in man: Skin pigment is not an essential regulator. Science 1981, 211, 590–593. [CrossRef]

124. Van Dijk, A.; den Outer, P.; van Kranen, H.; Slaper, H. The action spectrum for vitamin D3: Initial skin reaction and prolonged exposure. Photochem. Photobiol. Sci. 2016, 15, 896–909. [CrossRef]

125. Webb, A.R.; Decosta, B.R.; Holick, M.F. Sunlight Regulates the Cutaneous Production of Vitamin D3 by Causing Its Photodegradation. J. Clin. Endocrinol. Metab. 1989, 68, 882–887. [CrossRef]

126. Engelsen, O. The Relationship between Ultraviolet Radiation Exposure and Vitamin D Status. Nutrients 2010, 2, 482–495. [CrossRef]

127. Heckman, C.J.; Chandler, R.; Kloss, J.D.; Benson, A.; Rooney, D.; Munshi, T.; Darlow, S.D.; Perlis, C.; Manne, S.L.; Oslin, D.W. Minimal Erythema Dose (MED) Testing. J. Investig. Dermatol. 2015, 139, 2138–2146. [CrossRef]

128. Diffee, B.L.; Jansén, C.T.; Urbach, F.; Wulf, H.C. The standard erythema dose: A new photobiological concept. Photodermatol. Photoimmunol. Photomed. 1997, 13, 64–66. [CrossRef]

129. Harrison, G.I.; Young, A.R. Ultraviolet radiation-induced erythema in human skin. Methods 2002, 28, 14–19. [CrossRef]

130. Rhodes, L.E.; Webb, A.R.; Fraser, H.I.; Kift, R.; Durkin, M.T.; Allan, D.; O’Brien, S.J.; Vail, A.; Berry, J.L. Recommended Summer Sunlight Exposure Levels Can Produce Sufficient (≥20 ng/ml) but Not the Proposed Optimal (≥32 ng/ml) 25(OH)D Levels at UK Latitudes. J. Investig. Dermatol. 2010, 130, 1411–1418. [CrossRef] [PubMed]

131. Webb, A.R.; Kift, R.; Berry, J.L.; Rhodes, L.E. The vitamin D debate: Translating controlled experiments into reality for human sun exposure times. Photochem. Photobiol. 2011, 87, 741–745. [CrossRef] [PubMed]

132. Narbutt, J.; Philipsen, P.A.; Lesiak, A.; Sandberg Liljendahl, T.; Segerbäck, D.; Heydenreich, J.; Chlebna-Sokol, D.; Olsen, P.; Harrison, G.I.; Pearson, A.; et al. Children sustain high levels of skin DNA photodamage, with a modest increase of serum 25-hydroxyvitamin D3, after a summer holiday in Northern Europe. Br. J. Dermatol. 2018, 179, 940–950. [CrossRef] [PubMed]

133. Skobowiat, C.; Postlethwaite, A.E.; Slominski, A.T. Skin Exposure to Ultraviolet B Rapidly Activates Systemic Neuroendocrine and Immunosuppressive Responses. Photochem. Photobiol. 2017, 93, 1008–1015. [CrossRef] [PubMed]
134. Lucas, R.M.; Yazar, S.; Young, A.R.; Norval, M.; de Gruijl, F.R.; Takizawa, Y.; Rhodes, L.E.; Sinclair, C.A.; Neale, R.E. Human health in relation to exposure to solar ultraviolet radiation under changing stratospheric ozone and climate. *Photochem. Photobiol. Sci.* 2019, 18, 641–680. [CrossRef]

135. Holme, S.A.; Anstey, A.V.; Badminton, M.N.; Elder, G.H. Serum 25-hydroxyvitamin D in erythropoietic protoporphiria. *Br. J. Dermatol.* 2008, 159, 211–213. [CrossRef] [PubMed]

© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).