Review Article

Current and future alternative therapies for beta-thalassemia major

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ABSTRACT

Beta-thalassemia is a group of frequent genetic disorders resulting in the synthesis of little or no β-globin chains. Novel approaches are being developed to correct the resulting α/β-globin chain imbalance, in an effort to move beyond the palliative management of this disease and the complications of its treatment (e.g. life-long red blood cell transfusion, iron chelation, splenectomy), which impose high costs on healthcare systems. Three approaches are envisaged: fetal globin gene reactivation by pharmacological compounds injected into patients throughout their lives, allogeneic hematopoietic stem cell transplantation (HSCT), and gene therapy. HSCT is currently the only treatment shown to provide an effective, definitive cure for β-thalassemia. However, this procedure remains risky and histocompatible donors are identified for only a small fraction of patients. New pharmacological compounds are being tested, but none has yet made it into common clinical practice for the treatment of beta-thalassemia major. Gene therapy is in the experimental phase. It is emerging as a powerful approach without the immunological complications of HSCT, but with other possible drawbacks. Rapid progress is being made in this field, and long-term efficacy and safety studies are underway.

Beta-thalassemia was first discovered in the Mediterranean Basin and is highly prevalent in countries also affected by malaria, but human migration has resulted in the establishment of this disease in many areas of the world [1]. All patients display defects of hemoglobin (Hb) beta-chain production, but the resulting phenotypes are highly variable, ranging from severe anemia to an absence of clinical symptoms. Classification and severity grading are based principally on spontaneous Hb levels and clinical tolerance, regardless of the underlying genotype. Patients with beta-
thalassemia intermedia have blood Hb concentrations of 7–10 g/dL and do not require regular transfusion. They may display a broad spectrum of clinical signs, depending on the degree of alpha to non-alpha globin chain imbalance and several genetic and environmental factors. They may suffer from numerous complications, including pulmonary hypertension, thrombotic events, infection, endocrine dysfunction and leg ulcers [2]. Patients with beta-thalassemia major require regular transfusions of red blood cells to survive [3]. However, repeated transfusions cause iron overload, with life-threatening complications, such as endocrine dysfunction, cardiomyopathy, liver disease and, ultimately, premature death. In the absence of transfusion, patients with beta-thalassemia major die within the first five years of life, and even with transfusions, only 50–65% of patients live beyond the age of 35 years in high-income countries [4–6]. Research is highly active [Fig. 1], and progress has been made towards the development of new drugs, including biological products, some of which have recently reached the clinical trial stage.

Pharmacological therapies

Trace amounts of fetal Hb (HbF) persist into adulthood, accounting for less than 1% of total Hb in most adults [7]. However, HbF levels may exceed this threshold in some individuals. Beta-thalassemia patients with inherited persistent high levels of HbF production have a milder clinical course than other patients with this disease, and many do not require transfusions [8]. Therapeutic approaches reactivating HbF, and increasing its concentration, are, therefore, attractive. Variant HbF levels are highly inheritable [9]. Genome-wide association studies (GWAS) have compared individuals with low and persistently high levels of HbF, with the aim of identifying quantitative trait loci (QTL) to serve as a source of plausibly candidate causal genes or regulatory regions [10] accounting for the persistence of γ-globin gene expression. Strong associations between HbF level and single-nucleotide polymorphisms (SNPs) have been identified for at least four genomic loci, including the HBB (hemoglobin, beta) and olfactory receptor gene clusters on chromosome 11p15.4, the chromosome 6q23.3 HBS1L-MYB (HBS1-like translational GTPase -v-myb avian myeloblastosis viral oncogene homolog) intergenic region (HMIP), the BCL11A (B-cell CLL/lymphoma 11A) locus on chromosome 2p16.1, and the KLF1 (Kruppel-like factor 1) gene on chromosome 19p13.13 [11–18]. Variants at the HMIP-BCL11A-HBB loci account for 20–50% of the variability of HbF levels and the relative contributions of these variants differ between ethnic groups [19–23].

The –158C > T (rs7482144) SNP located at the XmnI site of the HBG2 (hemoglobin gamma G gene) promoter in the HBB locus was shown to be correlated with HbF levels in pioneering studies conducted on normal individuals and patients with sickle cell disease or β-thalassemia [24–27]. In the absence of a demonstrated functional role for this site, it has been suggested that HbF phenotype is modified by cis-linked elements located elsewhere in the β-globin cluster and in linkage disequilibrium with this SNP [28,29]. Indeed, a quasipalindromic structure located at the 5’ DNAse hypersensitive site 4 (HS4) of the locus control region (LCR), a polymorphism of which is in linkage disequilibrium with the XmnI site [28], may affect direct or indirect interactions with the transcriptional repressor BCL11A [30]. This transcriptional repressor directly regulates HbF levels during the globin switch after birth [31]. Erythroid-specific BCL11A knockdown blocks the silencing of fetal globin genes [32], and BCL11A SNPs associated with HbF level variations (located in intron 2) colocализ with target sites for erythroid transcription factors [33]. The KLF1 transcription factor represses γ-globin expression by activating BCL11A, with haploinsufficiency causing high HbF levels [17,34,35]. The MYB gene is a key regulator of the balance between proliferation and differentiation during erythropoiesis [36]. It regulates HbF levels through an as yet undetermined mechanism [37,38]. The intergenic HBS1L-MYB region contains MYB enhancer sites and DNA targets for erythroid transcription factors [39]. Genetic variants associated with the persistence of HbF, located in a 24 kb region of HMIP [15], affect MYB expression [16] by reducing erythroid transcription factor binding and long-range promoter activation [40].

The regulatory transcription factors involved in γ-globin gene regulation or F-cell differentiation and survival are potentially of considerable interest as targets for increasing HbF levels. However, it remains difficult to modulate the function of factors other than enzymes or signal-dependent nuclear factors by disrupting DNA/protein or protein/protein interactions [41] and such modulation is particularly problematic for factors with important non-erythroid functions [42]. Furthermore, any interference with erythroid transcription factors may result in the inappropriate disruption of erythropoiesis. Efforts are being made to design endonucleases capable of precisely disrupting the genomic sequences involved in the erythroid-specific expression of γ-globin repressors, as a means of activating HbF, but this remains a difficult challenge [43].

The S-phase cell-cycle inhibitor hydroxyurea (HU) has proved clinically effective in patients with sickle cell anemia (SCA) [44,45]. It is also of clinical benefit to some patients with β-thalassemia intermedia and it reduces the need for transfusions in a subset of individuals with β-thalassemia major [46–53]. However, side effects have been reported, including cytopenia, hyperpigmentation, weight gain, opportunistic infections, azoospermia in approximately 80% of men (even years after the end of treatment), and marked hypomagnesemia [54]. There is little or no risk of leukemia [55], but HU is believed to be teratogenic [54]. Given the potential adverse effects and reported efficacy in only a subset of patients, it is important to identify likely responders and nonresponders before initiating treatment, to ensure that prescriptions are efficiently targeted. The increase in HbF levels following HU injection in patients with β-thalassemia major seems to be correlated with the XmnI polymorphism [46,49,50,53], although this result has not been confirmed by all studies [47]. The correlation is less evident for non-transfusion-dependent thalassemia [46,51,52]. This is problematic, as treatments aiming to increase Hb levels by a few grams per deciliter are clearly more promising for the treatment of patients with thalassemia intermedia than for the treatment of transfusion-dependent patients. Baseline HbF level is clearly correlated with BCL11A SNP markers [11,12,14] and levels of the γ-globin repressor...
Fig. 1 — Current and future therapies for beta-thalassemia major. Hemoglobin disorders account for almost 5% of deaths in children under the age of five years. For a minority of patients, mostly in high-income countries, current therapies include lifelong monthly supportive red blood cell transfusions together with iron chelation or curative allogeneic HSC transplantation. The pharmacological induction of fetal hemoglobin and gene therapy are currently at the experimental stage. Top left: Conventional therapy includes regular red blood cell transfusions and iron chelation with injectable (Deferoxamine) or oral (Deferiprone or Deferasirox) drugs. Bottom left: Gamma-globin chain inducers aim to reduce the need for red blood cell transfusions. A number of drugs have been tested, including cytotoxic compounds and epigenetic regulators. The first drug shown to increase γ-globin expression was the demethylating agent 5-azacitidine. Small-chain fatty acid derivatives, including arginine butyrate, have also been shown to increase γ-globin expression most likely by inhibiting histone deacetylation. Hydroxyurea is the only drug currently approved for γ-globin induction. It acts through multiple mechanisms. Its cytotoxic activity is thought to accelerate the differentiation process and to stimulate cellular stress response pathways, leading to an overall increase in the number of F cells. Gamma-globin gene induction by other cytotoxic agents may also be mediated by this stress response. Erythropoietin (EPO) has proliferative and anti-apoptotic properties. The combined administration of recombinant EPO together with cytotoxic drugs can be beneficial for patients with low baseline EPO levels. Future treatments may target the transcription factors involved in γ-globin repression, such as BCL11A and KLF1. BCL11A is an essential transcription factor involved in γ-globin down-regulation. It binds to intergenic regions of the HBB locus, promoting long-range interactions with the LCR that favors β-globin expression. It recruits histone deacetylase to repress γ-globin. KLF1 is a strong inducer of β-globin expression that also activates BCL11A transcription when produced in large amounts in adult cells. KLF1 may itself be stimulated by c-Myb. Bottom right: Allogeneic transplantation with related or unrelated donor cells from cord blood units, mobilized peripheral blood or bone marrow is currently the only curative treatment, provided that a compatible donor can be found. Yet the risks of graft versus host disease and transplant rejection restrict the use of this procedure. Top right: Gene therapy is a promising single-dose medicine, without the need for immunosuppressive conditioning and graft versus host disease prophylaxis. It is potentially applicable to all patients. Studies evaluating safety and efficacy are currently underway and have already reported encouraging results. Studies of larger numbers of patients are required before any firm conclusions about efficacy and safety can be drawn. Long-term effectiveness/risk/cost ratios will need to be carefully addressed.
BCL11A [56], in both normal individuals and patients with beta-hemoglobinopathies [57]. However, only a few studies have investigated the correlation between polymorphisms and the HbF response to HU treatment, and conflicting results have been obtained [53,58–60]. In a study of beta-thalassemia patients of Iranian origin, polymorphisms of intron 2 of BCL11A were found to be correlated with the response to HU treatment [53]. By contrast, in studies including patients with SCA, BCL11A levels and SNPs were not found to be correlated with HbF induction after HU treatment [58–60], suggesting an important role for BCL11A in controlling baseline HbF levels, but with other factors required for HU to increase HbF levels. It has also been shown that intergenic HBS1L-MYB SNPs are not correlated with HU treatment [53,60].

SNPs of HBG2, HBS1L-MYB and BCL11A can be used for the rapid screening of disease severity in newborns and to provide more personalized care [61], but pharmacogenomics approaches do not distinguish between patients responding well and poorly to HU treatment [62]. The criteria for treatment should therefore not be based on these genetic characteristics. A number of genes involved in translation, ribosomal assembly and chromosome organization, but with no known function in normal, physiological β-globin switching, are deregulated in the erythroid cells of SCA patients treated with HU [58]. As recently suggested, the difference between weak and strong responses to HU may lie in the constitutive activation of genes involved in the stress response and terminal erythroid differentiation, and of genes protecting cells against stress-induced apoptosis in patients with strong responses [63]. Thus, the culture of erythroid progenitors and investigations of the expression profile of specific genes may make it easier to distinguish responders from nonresponders before the initiation of HU treatment in beta-thalassemia patients, but this possibility requires further study. Short trial periods on HU therapy (<3 months) are currently required to identify the patients most likely to respond to this treatment [48].

A number of other non-selective compounds, including cytotoxic compounds, short-chain fatty acid derivatives (SCFAD) and erythropoietin (Epo), have been investigated and evaluated in clinical trials [64]. Some are thought to act via epigenetic mechanisms. This is not surprising given (i) the strong correlation between epigenetic modification and the developmental pattern of globin gene expression [65–67] and (ii) the finding that some of the variation of HbF synthesis in adults may be linked to loci sensitive to epigenetic regulation [68]. However, the mechanisms of action of these compounds are not fully understood because they belong to several categories, including cytotoxic and hypomethylating agents (5-azacytidine, decitabine, HU), cytotoxic and histone deacetylase inhibitors, such as butyrate derivatives, and anti-apoptotic factors (Epo or Kit ligand) [69].

Allogeneic hematopoietic stem cell transplantation

Conventional therapies (red blood cell transfusion and iron chelation) improve the quality of life and survival of patients [70], but allogeneic hematopoietic stem cell transplantation (HSCT) currently offers the only hope of a definitive cure for patients with beta-thalassemia. HSCT was first successfully performed more than 30 years ago [71,72]. Several transplantation centers have since tried to categorize risk factors and to diversify HSCT procedures, with the aim of treating more patients, decreasing the risks of morbidity and mortality, and improving overall thalassemia-free survival. Several factors have been shown to affect patient outcome: severity criteria before transplantation (hepatomegaly, portal fibrosis and irregular chelation history), age at transplantation, stem cell source (peripheral blood, bone marrow, cord blood), histocompatibility (related matched, unrelated matched, mismatched, haploidentical), preparative conditioning regimen and pretransplant eradication of marrow hyperplasia. Despite the lack of evidence from randomized clinical studies comparing donor sources and/or conditioning regimens, HSCT is now widely accepted as a curative treatment [73] resulting in a long-term quality of life similar to that of the general population and higher than that of conventionally treated patients without active disease [74,75].

In a pioneering study, the group of Pesaro [76,77] assigned young patients to three classes according to the absence or presence of one, two or three risk factors before transplantation: hepatomegaly > 2 cm, portal fibrosis and irregular chelation history (class 1 = no risk factor, class 2 = one or two risk factors and class 3 = three risk factors). The transplantation of HLA (human leukocyte antigen)-matched sibling donor (MSD) cells into young class 1 or class 2 individuals on myeloablative conditioning (14 mg/kg busulfan + 200 mg/kg cyclophosphamide) and cyclosporin for graft-versus-host disease (GVDH) prophylaxis, gave excellent results (82–90% disease-free survival), but this treatment was highly toxic in pediatric class 3 patients (only 55% survived) [78,79]. Overall survival (OS) was increased to 74% by decreasing the dose of cyclophosphamide to below 200 mg/kg, but the probability of graft rejection (GR) remained high (35%) and event-free survival (EFS) remained low (49%) [80], probably due to inadequate immunosuppression and/or a failure to eradicate the massive erythroid hyperplasia.

Due to long-term exposure to iron overload, most patients over the age of 16 years have class 3 characteristics, and early attempts to treat them by transplantation, with high-dose cyclophosphamide regimens, were highly disappointing. All six patients died from graft failure or graft-versus-host disease (GVDH) in one study [81]. Use of a preparative regimen adapted to the risk factors identified in young patients improved the outcome for adult individuals, but OS remained low (63–65%). Rejection-free survival was slightly better than that for young class 3 patients (60–62%), mostly because rejection rates were lower in adults than in children, for unknown reasons [80–82]. Overall, the results obtained for class 3 patients were much worse than those for patients with class 1 or class 2 beta-thalassemia. Chronic active hepatitis at the time of transplantation had a strong negative impact on OS in adults [82].

Very few changes to the treatment of class 1 and 2 pediatric patients have occurred in recent years, with the exception of thiotepa, which is used to reduce rejection rates in patients under the age of four years [83]. A subgroup of young class III patients, those under the age of eight years, with
hepatomegaly >5 cm (class III high-risk (HR)) has been shown to have a much poorer clinical outcome after HSCT with a conventional conditioning regimen than other class III patients [84,85]. This subgroup of patients has a very high risk of GR and regimen-related toxicity. Lower intensity conditioning is thus required for both these patients and for adults [86]. Conflicting results for transplant-related mortality were obtained when the myeloablative regimen was changed from busulfan and cyclophosphamide to treosulfan, fludarabine and thiopeta (TreoFluT), and this change did not improve GVHD and graft failure in mixed-class groups of patients [87,88]. However, in well identified groups of class III patients and, even more significantly, in the subgroup of class III HR patients, the use of a TreoFluT regimen was associated with significant improvements in survival and EFS, due to a significant decrease in the incidence of sinusoidal obstruction syndrome (SOS) [89]. In this study, the increase in the risk of rejection due to the lower intensity of conditioning was counteracted by the use of peripheral blood stem cells (PBSC) rather than BM cells. In pediatric class 3 patients, ablation of the expanded thalassemic marrow and stronger immunosuppression through the use of an intensified preparation regimen including HU, azathioprine, fludarabine and hypertransfusion increased event-free survival to 80–85% [90,91]. The use of this protocol in adults did not decrease transplant-related mortality, and thalassemia-free survival remained low [92]. The inclusion of anti-thymocyte globulin (ATG) in the preparative regimen of pediatric patients from all classes, together with the adaptation of busulfan and cyclophosphamide concentrations, decreased the incidence of GVHD, minimized GR and transplant-related mortality and seemed to abolish the difference between patients from different Pesaro risk classes, in a recent survey carried out in Greece and France [93,94]. The transplantation of cord blood (CB) and bone marrow cells from a MSD offers a similar probability of long-term cure, but with a lower incidence of GVHD. The transplantation of PBSC yields lower rates of GR than bone marrow transplantation in class III patients receiving TreoFluT, but GVHD remains a concern when PBSC grafts are used [95].

One of the main barriers to HSCT with cells from HLA-identical siblings is the limited availability of donors, which is, theoretically, 25% for any single sibling. The combined use of pre-implantation genetic diagnosis and HLA matching provides a reliable source of stem cells from healthy siblings, but rates of successful transplantation after ovarian stimulation remain low [96]. For most of the remaining patients, millions of HLA types are stored in national registries. The probability of finding an unrelated donor (UD) with 9/10 or 10/10 HLA-matches (HLA-A, -B, -C, -DRB1 and -DQB1) on the basis of high-resolution typing exceeds 70% for patients of European ancestry [96], but this likelihood depends on ethnicity [97]. A prospective study of patients with hematologic malignancies suggested that the transplantation of cells with this degree of matching from an UD provides a likelihood of cure similar to that for the transplantation of cells from a MSD [98]. In both adults and children with β-thalassemia, bone marrow transplantation from an UD selected on the basis of high-resolution typing yields success rates similar to those obtained with a MSD [99,100], but with more severe GVHD. This approach is not widely used in adults, due to the high incidence of GVHD, transplant-related mortality and GR, but it may be used to treat children with class VII β-thalassemia safely, provided that ATG is given [100]. A highly promising recently developed protocol including marrow reduction and pretransplant immunosuppression, followed by low-intensity conditioning with ATG, has been shown to result in a low risk of toxicity and durable engraftment in class 3 HR patients receiving cells from unrelated and related donors; this approach merits more widespread testing, even in individuals with lower risk β-thalassemia [101,102].

Donor cells can also be obtained from CB samples. The transplantation of CB cells originating from HLA-identical siblings is a viable alternative to the use of adult samples in class I/II patients [103–105]. Moreover, these cells have the theoretical advantages of tolerating a higher degree of HLA incompatibility than adult cells [106] and causing a lower incidence of acute and chronic GVHD [107]. However, in a report compiling data from 32 transplant centers from 1996 to 2009, survival and disease-free survival were as low as 62% and 20%, respectively, in patients with β-thalassemia receiving CB cells from mismatched donors, after myeloablative conditioning [108]. Graft failure was the chief cause of treatment failure other than death, which was linked to disease severity and the use of a non-adapted conditioning regimen. Multivariate analysis indicated that the principal factor associated with engraftment was the number of cells used. These results contrast with those of a Taiwanese study reporting the survival of 88% of young patients receiving one or two unrelated CB units after a full myeloablative regimen; 74% of these patients were disease-free five years after transplantation. Both chronic and acute GVHD occurred, but most cases were moderate [109]. However, double cord transplants were performed in this study if a single unit contained too few nucleated CD34+ cells. More clinical trials are required.

### Gene therapy

The concept of gene therapy for beta-thalassemia emerged a long time ago and was included in a plan for the viable treatment for hemoglobinopathies as early as 1978 at the University of California at Los Angeles (UCLA) [110]. However, major technical issues were encountered and attempts to demonstrate the efficacy of infusions of transfected mouse cells in myeloablated animals were far from convincing [111,112]. The regulatory globin sequences required for high levels of production [113] and efficient methods for gene introduction [114] were not available at the time. The stemness properties of the manipulated bone marrow cells were unknown. Early attempts to treat beta-thalassemia patients by inserting the β-globin gene into bone marrow cells, in 1980, were completely unsuccessful and received an avalanche of criticism [115,116].

It is essential to achieve a satisfactory combination of high levels of sustained β-globin expression and the stable propagation of complex sequences. More than 25 years were required to achieve these goals, which were made possible by
the characterization, isolation, size reduction, and blending of
the β-globin locus regulatory elements and the advent of
lentiviral vectors, which can transfer complex sequences into
hematopoietic stem cells. The genetic elements required for
the adult- and erythroid-specific expression of the β-globin
gene were mapped close to the gene [117,118], in the promoter
region, the second intron and downstream from the poly-
adenylation signal [119–121]. However, the combination of
these elements in a single transgene yielded very low levels of
β-globin gene expression in the erythroid cells of mice un-
dergoing transplantation with genetically modified HSCs
bearing the transgene [122–124]. The discovery of the β-globin
LCR, located in a 15,000-base region far upstream from the
globin genes [125–127], was a major step forward. The use of
this region made it possible to design β-globin transgenes that
were strongly expressed in the erythroid cells of transgenic
mice [128,129]. Over a period of 10 years, efforts were made to
delineate the enhancer core elements of the LCR [129,130],
with the aim of decreasing the size of the DNA fragments
used, such that the chimeric DNA constructs would be
compatible with transfer vectors derived from gamma-
retroviruses [131,132]. These vectors provided reasonable
levels of β-globin transgene expression in vitro, but the trans-
duction of mouse hematopoietic stem cells (HSCs) was sub-
optimal, resulting in limited and variable expression of the β-
globin gene in vivo [133,134].

Gamma-retroviral vectors (γ-RVs) have three major de-
fects: (i) limited cargo capacity, (ii) instability, and (iii) an
inability to transduce non-dividing cells (most HSCs are
quiescent [135]). These drawbacks precluded the efficient
correction of beta-thalassemia by the infusion of genetically
modified HSCs. As a result, the gene therapy community
working on globin disorders rapidly abandoned the use of
γ-RVs shortly after lentiviral vectors (LVs) derived from human
immunodeficiency virus type 1 (HIV1) became available. Not
only were LVs able to transduce cells arrested at the G1-S
boundary of the cell cycle [136,137], they were also able to
transfer much longer sequences than γ-RVs [138]. This key
characteristic is dependent on a molecular mechanism for
inhibiting the splicing of the viral RNA before its packaging
into lentiviral particles. This mechanism involves active
transport of the full-length (unspliced) viral RNA from the
nucleus to the cytoplasm by the viral protein REV, which binds
to the REV-responsive element (RRE) located within the viral
RNA structure [139]. The potential utility of incorporating this
mechanism into complex LCR/β-globin LVs was suggested
by our discovery that unwanted RNA splicing is the key
determinant of LCR/β-globin γ-RV instability and low ti-
ters, and that this problem could be largely corrected by the
removal of undesirable splice sites [131]. Our groups and that
of Sadelain then focused on obtaining proof-of-principle for
the preclinical efficacy of LCR/β-globin LVs in two mouse
models of beta-hemoglobinopathies: murine beta-
thalassemia [140] and transgenic mouse models of sickle cell
disease [141]. This approach was subsequently consolidated
by other studies [142–146]. For gene therapy for sickle cell
disease, we decided to abandon the wild-type beta-globin
gene, choosing instead to work with a variant we constructed,
with an enhanced ability to inhibit HbS polymerization [141].
This variant (β6–T87Q) was generated by introducing the codon
for a specific amino-acid residue present in both the gamma-
and delta-globin proteins (Q, glutamine) to replace the corre-
ponding codon in beta-globin (T, Threonine) [147]. The
β6–T87Q variant has an additional key advantage for gene
therapy trials in beta-thalassemia: it facilitates accurate
quantification of the amount of vector-derived beta-globin
produced or present in circulating red blood cells by high-
performance liquid chromatography (HPLC), even in beta-
thalassemia patients receiving red blood cell transfusions or
with β+-thalassemia [148]. Such quantification is not possible
after the transfer of the wild-type beta-globin gene.

The biosafety of LVs has greatly improved in recent years,
and there is now very little risk of producing replication-
competent lentiviruses [149]. Modifications have included
splitting the genetic elements required for production of the
viral core proteins, the elimination of accessory genes
involved in virus infectivity, the replacement of the viral en-
velope with that of another class of virus, and the removal of
the viral enhancer/promoter elements [150]. Insertional
mutagenesis, which led to leukemia in some patients
receiving γ-RV transduced cells [151–154], is a major concern
when integrative vectors are used. Viral transcriptional acti-
vating elements have been shown to trigger oncogene activ-
ation, leading to the strong recommendation that these
elements should be deleted from retroviral vectors used for
gene therapy purposes [155]. LVs have a better safety profile
than γ-RVs in hematopoietic cells [156] and are less genotoxic
than γ-RVs in transplanted mice [157,158], due to the differ-
cent DNA targeting of the two vectors [159]. Nevertheless,
LVs in general [159], and β-globin-LVs in particular [160],
preferentially integrate into active genes and disturb gene
expression over large distances [161]. Furthermore, the level
of β-globin gene expression under the control of LCR enhancers
is subject to position effects [162,163]. Following vector inte-
gration, levels of β-globin expression have been shown to be
variable and to depend on the location of the vector in the
chromatin in transplantation experiments in mouse [164].
Chromatin insulators placed between enhancers and pro-
moters prevent interactions between these elements. They
also buffer transgenes against chromosomal position effects
[165,166]. Chromatin insulators derived from the chicken hy-
persensitive site-4 (CHS4) of the β-globin locus [166] were thus
introduced into β-globin LVs, to increase efficacy and safety
[148,167]. This modification led to a higher probability of β-
globin expression and lower levels of position effect variega-
tion in vitro [167].

A LV encoding the mutated β-globin (β6–T87Q) was thus
constructed. In this vector, referred to as LentiGlobin vector
HPV569, the β-globin gene is under the control of the LCR and
the β-globin promoter, and is surrounded by tandem copies
of the CHS4 core element [148]. Experiments in beta-thalassemia
mice subjected to busulfan conditioning and transplantation
with cells transduced with this vector showed them to have
near-normal hemoglobin levels and to display no overall
toxicity [168]. Over the following few years, we performed a
large body of experiments with the aim of applying to carry out a
clinical trial. This involved efficacy and safety/toxicology
studies in mice, and large-scale vector manufacture in good
manufacturing practice (GMP) conditions [148,168–170]. Spec-
ific qualified assays for quality assurance (QA)/quality control
(QC) were also developed and approved by the regulatory authorities. The Sadelain group and several other groups also worked towards clinical trial applications [171–173]. The differences between our own product and trial protocol and those of the Sadelain group include (1) differences in the boundaries of the beta-globin gene, beta-globin promoter and LCR elements, (2) the initial presence of the cHS4 insulator core in our vector, (3) an anti-sickling and chromatography-identifiable β^A–^B^T^8^Q^−^, globin variant, (4) differences in vector manufacturing and purification, (5) full-dose intravenous busulfan treatment, but without the addition of an immunosuppressive agent, (6) applicability to both beta-thalassemia and sickle cell disease due to our choice of the β^A–^B^T^8^Q^−^ variant, and (7) initial focus on beta^A^α^0^–^beta^A^α^0^-thalassemia major patients [148,169,170,174]. We were granted authorization to carry out clinical trials for both beta-thalassemia and sickle cell disease in 2006 (the LG001 Study), by the French regulatory authority ANSM (Agence Nationale de Sécurité du Médicament et des Produits de Santé). In these trials, we used the GMP-grade HPV569 vector produced and controlled in the USA. This work [170], together with a clinical trial of gene therapy for adrenoleukodystrophy [175], constituted the first trial of the use of a LV for the treatment of an inborn genetic disease to be approved anywhere in the world.

The first beta-thalassemia patient to be treated and who did not receive backup cells was an 18-year-old ββ^α^β^α^ adult with clinical beta-thalassemia major who had been transfusion-dependent and on parenteral iron chelation therapy since early childhood and had undergone splenectomy at the age of six years, and for whom no HLA-MSD was available [169,170,174]. Bone marrow cells were processed for CD34^+^ cell selection, transduced with the HPV569 lentiviral vector and thawed for release testing. The patient received an infusion of 3.9 × 10^6^ cells/kg after busulfan conditioning at the fully myeloablative dose (12.8 mg/kg). Hematopoietic reconstitution was uneventful, and the patient became transfusion-independent one year after transplantation. Hemoglobin levels are stable, at about 8–9 g/dL.6.5 years later or 7.5 years since gene therapy, with a mean vector copy number close to 0.2 vector copy per myeloid cell [176]. The transgenic hemoglobin accounts for about one third of total hemoglobin (HbF, HbE and HbAT87Q). A number of transduced hematopoietic clones were detected among the lymphoid and myeloid cells. One dominant clone emerged in month 3 and stabilized around month 15. This clone accounted for less than 4% of the peripheral nucleated cells, and this percentage had fallen to less than 2% five years post-transplantation. In this clone, the vector was inserted at the high-mobility group AT-hook 2 (HMG2) locus, compromising gene splicing and causing the premature termination of transcription. The compromised HMG2 gene displayed transcriptional activation, generating large amounts of a truncated and active form of the protein in transduced cells. It was suggested, but not formally demonstrated, that there was a causal link between vector insertion at this locus, HMG2 gene activation, and clonal dominance [170]. High-performance liquid chromatography (HPLC) analysis of single erythroid colonies indicated that vector-derived β^A–^B^T^8^Q^−^ globin expression output did not differ significantly between the HMG2 integrant and colonies displaying integration at other chromosomal sites [170]. Almost eight years after transplantation, there is no signs of clonal overgrowth or toxicity [176]. Another patient underwent transplantation in November 2011, but the vector copy number was lower. This patient is still transfusion-dependent, and transgenic hemoglobin levels account for ≈5% of total hemoglobin [176].

The first two clinical studies in which LVs were used to transduce HSCs for the treatment of β-thalassemia [170] and adrenoleukodystrophy [175] showed that one key restriction to fully effective treatment was the limited efficacy of stem cell transduction, which was only 10–20%. Other studies suggested that the protection provided by the cHS4 elements was variable [155,177], and dependent on their position in the chromatin [178]. Furthermore, the tandem copies of the cHS4 insulator inserted into the β-globin vector were prone to rearrangement, resulting in the regeneration of a single element upon vector insertion [168,179]. This single element had a lower level of enhancer-blocking activity than the tandem copies [180]. The insertion of insulators into the vector was also shown to decrease vector titer [179,181]. It therefore seemed likely that the removal of the cHS4 elements from the vector would result in a LV with a higher transduction efficacy and a similar level of safety. We therefore simply removed cHS4 from LentiGlobin vector HPV569 to yield LentiGlobin vector BB305, which also contains a heterologous promoter to increase vector titers and yield [182]. The production, concentration and purification protocols were improved [183]. The new vector BB305 was compared with HPV569 in human cells and in a comprehensive mouse toxicology study [182]. It was found to be more effective than the previous vector, with an equivalent and satisfactory level of safety [182]. On the basis of these new preclinical results with LentiGlobin vector BB305 and the sustained positive outcome for a patient with the parental LentiGlobin vector HPV569 [170], both the ANSM (French) and the RAC/FDA (Recombinant DNA Advisory Committee/Federal Drug Administration, US) granted us approval, in 2013 and 2014, for three new clinical trials using LentiGlobin vector BB305 sponsored by the biotechnology company bluebird bio: (1) Trial NCT02151526 (HGB-205) in France, for both beta-thalassemia and sickle cell disease, (2) Trial NCT01745120 (HGB-204) at multiple centers in the US, Australia and Thailand, for beta-thalassemia, and (3) Trial NCT02140554 (HGB-206), at multiple centers in the US, for sickle cell disease. The three trials follow essentially the same clinical transduction protocol as for the previous trial with LentiGlobin vector HPV569, although the BB305 vector is further purified by preparative chromatography.

Very encouraging results have been obtained for transfusion-dependent beta-thalassemia major patients, and were presented by the corresponding clinical investigators at the annual meeting of the American Society of Hematology (ASH) in December 2014 [184,185]. For the HGB-205 trial, transplantation was carried out in two transfusion-dependent ββ^α^β^α^ patients. Six months post-treatment, the mean vector copy numbers per cell were above 1 in peripheral blood, and Hb levels had reached 10.2 and 13.4 g/dL, respectively, in the absence of transfusion, with ≈70% HbAT87Q, ≈30% HbE and low HbF. The two patients became transfusion-independent 10 and 12 days after cell infusion. CD34^+^ cells from a patient...
with sickle cell disease were also transduced with highly efficiently, but clinical data were not available in December 2014. For the HGB-204 study, five transfusion-dependent beta-thalassemia major patients underwent transplantation. Two of these patients were treated more than three months before the data were released in December 2014. One of these patients is \(\beta^\alpha/\beta^0\), whereas the other has the Cooley’s anemia \(\beta^0/\beta^0\) genotype. Both these patients rapidly became transfusion-independent, with stable Hb concentrations of 8.6 and 9.6 g/dL in the absence of transfusion, six and three months post-transplantation, respectively. The other 3 patients underwent transplantation in November 2014, and too little time had elapsed for any meaningful conclusions to be drawn at the time of data release in December 2014. None of the subjects experienced a drug product-related adverse event in either of these trials. Vector integration site analyses showed polyclonal reconstitution without clonal dominance. Additional subjects were still awaiting transplantation in December 2014.

Another clinical trial, sponsored by the Memorial Sloan-Kettering Cancer Center of New York, USA, was also initiated in 2012, in adult \(\beta\)-thalassemia major patients receiving autologous CD34+ cells, with non-myeloablative conditioning, after transduction with the TNS9.3.55 LV from the Sadelain group (clinicalTrial.gov identifier: NCT01639690) [171,173]. A non-myeloablative conditioning regimen would clearly have advantages over a fully ablative regimen, but such a regime may not currently be sufficient to ensure optimal engraftment with transduced stem cells. The clinical data from this trial have yet to be formally released.

**Conclusion**

Future treatments for beta-thalassemia will depend on the benefit/risk/cost ratios for conventional transfusion and iron chelation [70], \(\gamma\)-globin induction [186], allogeneic bone marrow transplantation [187] and gene therapy [170], and the availability of these treatments in the low-income countries in which most \(\beta\)-thalassemia patients live. Treatments with HU or with other \(\gamma\)-globin inducers, either alone or in combination, have been shown to increase Hb levels by 1–5 g/dL [188]. However, HU is currently the only compound approved for the treatment of adult patients with sickle cell disease. Furthermore, a number of beta-thalassemia patients fail to achieve significant increases in HbF levels and clinical improvement on this treatment [44,189]. Most patients with beta-thalassemia major continue to suffer from profound anemia [69,190] and new drugs inducing HbF are being investigated [191]. In the field of hematopoietic stem cell transplantation, Investigations are continuing, with the aim of extending allogeneic stem cell infusion to a larger number of patients. The use of haploidentical parent or sibling donors may extend the use of this treatment to patients lacking a MSD or HLA-compatible UD, provided that the balance between the risks of GVHD and graft failure is considered acceptable. Recent successes have been reported in small series of patients [192–194] and require confirmation in larger cohorts, with longer periods of follow-up. Significant improvements in clinical outcomes have been obtained through progress in the domains of conditioning regimens, risk stratification and stem cell sources. However, allogeneic stem cell transplantation remains a complicated and risky procedure. Many of its drawbacks, such as GVHD and GR in particular, may be avoided by the use of autologous stem cell transplantation after corrective gene transfer. Gene therapy is still at an early stage of development, but our clinical trials have already provided proof-of-principle for sustained clinical efficacy with low toxicity in several patients. With improvements in the production of LentiGlobin vector BB305, all of the patients treated rapidly became transfusion-independent, with near-normal Hb concentrations in the blood, even in one patient with \(\beta^0/\beta^0\)-thalassemia (Cooley’s anemia). Gene therapy may thus become an attractive mode of treatment for young adults with no MSD considered to be at low risk of transplant complications. Further studies will be required before this approach can be offered as a first-line medical treatment. For high-risk class 3 patients, reduced and adapted preparative regimens must be tested. Methods for selecting and amplifying transduced stem cells may be required to maximize efficacy in such a context of reduced conditioning [195]. Finally, although recent gene and cell therapy trials involving LV-mediated gene transfer in HSCs followed by autologous cell transplantation gave excellent outcomes, with no observable toxicity [175,184,196,197], it is still too early to draw any firm conclusions about the risk of insertional mutagenesis. Gene vector targeting and gene modification by homologous recombination [198] remain sub-optimal, but research is continuing. Combined gene and cell therapy approaches may make it possible to exploit the proliferative potential of pluripotent stem cells, but many obstacles to the use of such approaches for HSCT remain [199,200]. The next few years will probably confirm the enormous potential of gene transfer methods for the treatment of \(\beta\)-thalassemia patients, in terms of both efficacy and safety.

**Conflicts of interest**

Edouard de Dreuzy and Kanit Bhukhai have no competing interests. Philippe Leboulch and Emmanuel Payen have financial relationships with Bluebird Bio, Inc.

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