CONTRACTILE RESPONSES OF ISOLATED RABBIT AORTAE
TO TRANSMURAL STIMULATION AS AFFECTED
BY CALCIUM, STRONTIUM, SODIUM
AND OUABAIN

Noboru TODA
Department of Pharmacology, Faculty of Medicine, Kyoto University, Sakyo-ku,
Kyoto, Japan

Received for publication November 4, 1971

There are a number of reports describing that not only in visceral (1-3) but also vascular
smooth muscles (4-6) Ca\(^{++}\) may be the agent linking membrane excitation to contraction.
Increasing [Ca\(^{++}\)]\(_0\) may either stimulate (7, 8) or inhibit (9) vascular smooth muscles.
Adrenaline and K\(^+\) elicit contraction and an increase in the Ca\(^{++}\) influx into smooth muscle
cells (4, 10). Elevation of [Ca\(^{++}\)]\(_0\) depresses the first part of the contractile response of
aortic muscles to adrenaline but potentiates the second part (11). Excitation of nervous
elements in vascular tissues by electrical transmural stimulation results in contraction and
a sharp rise in the output of noradrenaline (12). Contractile responses of isolated vessels
to stimulation of periarterial sympathetic nerves are shown to relate directly to [Ca\(^{++}\)]\(_0\),
ranging from 0 to 4.4 mM (13).

It seems likely that Sr\(^{++}\) can substitute for Ca\(^{++}\) in the contractile mechanism of
skeletal, cardiac and smooth muscles (14-18) and in the release of the neurohumoral trans-
mitter from sympathetic nerve terminals (18). It appears that Na\(^+\) acts through an antago-
nism to Ca\(^{++}\) on muscular contractility (4, 19) and uptake of noradrenaline by sympathetic
nerves (20). Ouabain is known to increase the exchangeable Ca\(^{++}\) fraction (21, 22), which
would be expected to participate in increased contractility.

The present study was an investigation into the effects of ions and drugs that interact
with Ca\(^{++}\) on the resting tension of the vascular smooth muscle and on its contractile re-
response to electrical transmural stimulation and exogenous noradrenaline.

METHODS

Albino rabbits of both sexes, weighing 1.8 to 2.2 kg, were used. Under ether anesthesia
the animals were sacrificed by bleeding from both common carotid arteries. The ascending
aorta was rapidly removed and cut into spiral strips. The strips were held vertically under
a resting tension of 2 g in the muscle bath of 100 ml capacity containing the nutrient solution.
The solution was maintained at 37 ± 0.5 °C and gassed with a mixture of 95% O\(_2\) and 5% CO\(_2\). Constituents of the solution were as follows (mM): Na\(^+\), 162.1; K\(^+\), 5.4; Ca\(^{++}\), 2.2;
Cl\(^-\), 157.0; HCO\(_3\)^-\, 14.9; dextrose, 5.6. Osmotic adjustment was not made when [Ca\(^{++}\)]\(_0\),
and \([\text{Sr}^{++}]\), were altered. Strontium ions as \(\text{SrCl}_2\) solution were added directly to the bathing medium in the muscle chamber. Extracellular concentrations of \(\text{NaCl}\) were lowered by replacing with isotonic sucrose. Before measurements were taken, preparations were allowed to equilibrate for 60 to 120 min in the control solution and for 20 to 30 min in test solutions.

The vascular strips were placed between a pair of stimulating electrodes (platinum plates, 5 \(\times\) 15 mm), as described by Toda, Usui and Mori (23). The preparations were transmurally stimulated by a train of 0.3 msec-rectangular pulses of supramaximal intensity (about 80 V) applied at frequencies of 5, 20 and 100/sec. The number of the electrical pulses was kept constant (200 pulses) by changing the period of stimulation (40, 10 and 2 sec for frequencies of 5, 20 and 100/sec, respectively). Contractile response to electrical stimulation applied under these conditions is shown to result from released noradrenaline (23). Electrical stimuli were provided by an electronic stimulator (Type WSE-3R, Nihonkoden Kogyo Co.).

Contractile responses to electrical transmural stimulation and noradrenaline were recorded on a two-channel penwriter (Sanei Sokki Co.). From contractions induced by the electrical stimulation three parameters were measured: the maximum tension developed, the duration of contraction at the level of half maximum tension, which will be termed ‘duration’ and the time from the initiation of stimulation to the peak tension, termed ‘time-to-peak’. Values of the parameters obtained from aortic preparations stimulated for 10 sec at a frequency of 20/sec in control media were taken as control (100 \(\%\)), and relative values of those obtained in test solutions to the respective control were presented. Noradrenaline was applied directly to the nutrient solution of the muscle bath in cumulative concentrations. The tension developed at \(5 \times 10^{-5}\) M noradrenaline in control media was taken as 100 \(\%\). After 20 min exposure to test solutions or after 30 min exposure to ouabain, transmural stimulation or noradrenaline was applied. In test solutions the preparations were stimulated repeatedly at frequencies of 5, 20 and 100/sec at intervals of 5 to 15 min, until steady responses were attained. The results shown in the text and figures are expressed as mean values \(\pm\) standard errors of the means. Comparisons were made using Student’s ‘t’ test.

Ouabain, U.S.P. (Nutritional Biochemicals Corp.), cocaine hydrochloride and \(dL\)-noradrenaline hydrochloride were used.

RESULTS

**Calcium and strontium**

In spirally-cut strips of the ascending aorta electrical transmural stimulation applied for 10 sec at a frequency of 20/sec caused a transient increase in muscle tension. Mean values of the maximum tension developed, the duration and the time-to-peak were \(0.84 \pm 0.05\) g, \(1.52 \pm 0.08\) min and \(0.48 \pm 0.03\) min, respectively \((N=66)\). These parameters of the induced contraction were altered by varying stimulation frequency although the total number of stimuli was kept constant (200 pulses); the developed tension was markedly
Modification by elevating [Ca\textsuperscript{++}]\textsubscript{o} of contractile response to transmural stimulation and noradrenaline. Left: transmural stimulation. Abscissa = frequency and period of stimulation. Values of the parameters measured in individual preparations when stimulated for 10 sec at a frequency of 20/sec in control media were taken as 100%. Mean values of the magnitude, the duration and the time-to-peak of contraction induced under these experimental conditions were 0.79 ± 0.13 g, 1.24 ± 0.21 min and 0.35 ± 0.03 min, respectively (N=11). Right: noradrenaline. Values of the tension developed at 5 × 10\textsuperscript{-5} M noradrenaline in the control solution were taken as 100% (mean value of the tension: 4.64 ± 0.34 g, N=15). Figures in parentheses indicate the number of preparations.

When the Ca\textsuperscript{++} in the bathing medium was increased to 6.6 mM (3 times normal [Ca\textsuperscript{++}]\textsubscript{o}) the tension increased slightly (0.13 ± 0.05 g, N=11) but the contractile response to transmural stimulation was not significantly affected (Fig. 1). The dose-response curve of noradrenaline was not appreciably affected by an elevation of [Ca\textsuperscript{++}], (Fig. 1).

The addition of Sr\textsuperscript{++} in concentrations of 2.2 and 4.4 mM to control media did not significantly influence the magnitude and the duration of aortic contraction in response to transmural stimulation (Fig. 2). Resting tension of aortic smooth muscles was not changed by Sr\textsuperscript{++} at 2.2 and 4.4 mM. Contractile response to exogenous noradrenaline was not influenced by 2.2 mM Sr\textsuperscript{++} (Fig. 2).

Exposure of aortic preparations for 20 to 30 min to Ca\textsuperscript{++}-free media markedly reduced the contractile response to transmural stimulation. The reduction of the response related inversely to stimulation frequency. Following the application of Sr\textsuperscript{++} at 2.2 mM the contractile response which had been reduced by Ca\textsuperscript{++}-deficiency was partly restored in preparations stimulated at 5/sec and completely restored at 20 and 100/sec. Results are summarized in Fig. 3. Resting tension which had been decreased by 0.07 ± 0.02 g (N=12, P<0.01)
at zero $[\text{Ca}^{++}]_o$ was restored by 2.2 mM $\text{Sr}^{++}$ ($0.07 \pm 0.02$ g increase from the tension at zero $[\text{Ca}^{++}]_o$, $N=12$). Sensitivity of aortic smooth muscles to exogenous noradrenaline was reduced by removal of $\text{Ca}^{++}$. Strontium ions (2.2 mM) partly restored the sensitivity (Fig. 3).

**Sodium deficiency**

Decrease in the concentration of $\text{Na}^+$ in bathing media to 103.2 mM (about 64% normal $[\text{Na}^+]_o$) increased the resting tension of aortic strips by $0.37 \pm 0.05$ g ($N=6$) and potentiated significantly ($P<0.01$, at stimulation frequencies of 20 and 100/sec) the contractile response to electrical transmural stimulation. The duration was prolonged in parallel with the increased magnitude of contraction, whereas the time-to-peak was not altered (Fig. 4). The contractile response to exogenous noradrenaline in high concentrations ($2.5 \times 10^{-5}$ and $5 \times 10^{-5}$ M) was significantly potentiated by $\text{Na}^+$ deficiency ($P<0.01$) (Fig. 4).

Further reduction of $\text{Na}^+$ in the nutrient solution to 73.8 mM (about 45% normal $[\text{Na}^+]_o$) increased the resting tension by $1.14 \pm 0.17$ g ($N=10$). The contracture developed gradually until a plateau was attained after 30 to 60 min exposure to the test solution. The magnitude of contraction induced by transmural stimulation was considerably reduced by lowering $[\text{Na}^+]_o$, whereas the duration was prolonged (Fig. 5). Thus, recovery from the induced contraction was markedly retarded. The time-to-peak was not appreciably affected.
FIG. 3. Changes in contractile response to transmural stimulation (Left) and noradrenaline (Right) by removal of Ca++ and by Sr++ applied in Ca++-free media. Mean values of the magnitude and the duration of contraction induced by stimulation for 10 sec at 20/sec in control media were 0.65±0.07 g and 1.41±0.10 min, respectively (N=12). The mean value of the tension developed at 5×10⁻⁵ M noradrenaline in control media was 4.80±0.52 g (N=9). Figures in parentheses indicate the number of preparations.

FIG. 4. Changes in contractile response to transmural stimulation (Left) and noradrenaline (Right) by lowering [Na+]o to 103.2 mM. Mean values of the magnitude, the duration and the time-to-peak of contraction caused by stimulation for 10 sec at 20/sec in control media were 0.74±0.20 g, 1.81±0.21 min and 0.39±0.03 min, respectively (N=6). The mean value of the tension developed at 5×10⁻⁵ M noradrenaline in control media was 3.56±0.29 g.
FIG. 5. Changes in contractile response to transmural stimulation (Left) and noradrenaline (Right) by lowering [Na+]o to 73.8 mM. Mean values of the magnitude, the duration and the time-to-peak of contraction induced by stimulation for 10 sec at 20/sec in the control solution were 0.83 ± 0.10 g, 1.30 ± 0.15 min and 0.38 ± 0.02 min, respectively (N = 10). The mean value of the tension developed at 5 × 10^{-6} M noradrenaline in control media was 4.16 ± 0.26 g (N = 11).

FIG. 6. Relationship between the ratio [Ca^{++}]_o/[Na^{+}]_o^2 and the tension developed by alteration in [Na^{+}]_o and [Ca^{++}]_o and by transmural stimulation. Filled circles = tension developed at varying [Na^{+}]_o and [Ca^{++}]_o. Open circles = sum of the tension developed by alterations in extracellular concentrations of the ions and by transmural stimulation at 20/sec. Solid lines = variation in [Ca^{++}]_o ([Na^{+}]_o : constant). Broken lines = variations in [Na^{+}]_o ([Ca^{++}]_o : constant). Figures in parentheses indicate the number of preparations.
The contractile effect of exogenous noradrenaline was reduced by Na⁺ deficiency (Fig. 5), although the resting tension in control and Na⁺-deficient media markedly differed (tension developed at reduced [Na⁺]₀: 0.70 ± 0.12 g, N = 11).

Fig. 6 demonstrates interactions of Na⁺ and Ca²⁺ on the resting tension and the contractile response to transmural stimulation applied for 10 sec at 20/sec. Trends of increasing the resting tension at various [Na⁺]₀ and [Ca²⁺]₀ and the sum of the tension developed by variations in [Na⁺]₀ and [Ca²⁺]₀, and by transmural stimulation with increasing the ratio [Ca²⁺]₀/[Na⁺]₀ were observed. Dependence of the two parameters on reduced [Na⁺]₀ was markedly greater than that on increased [Ca²⁺]₀, as far as concentrations of the ions used in this study were concerned.

Ouabain

Contractile responses of aortic strips to transmural stimulation was significantly potentiated (P<0.01) by ouabain in concentrations of 3.4 × 10⁻⁷ and 3.4 × 10⁻⁶ M (Fig. 7). The duration of contraction was prolonged approximately parallel to the increase in the magnitude of contraction. The resting tension was not appreciably affected at the lower concentration but increased at the higher (0.43 ± 0.09 g, N = 9). The dose-response curve of noradrenaline was moved left by ouabain (Fig. 8). Further increase in the concentration of ouabain to

---

**Fig. 7. Modification by ouabain of contractile response to transmural stimulation.**

Mean values of the magnitude and the duration of contraction induced by stimulation for 10 sec at 20/sec in control media were 0.95 ± 0.15 g and 1.20 ± 0.22 min, respectively (N = 9).
1.7 \times 10^{-5} \text{ M} caused contracture: the tension developed gradually until a plateau was attained after 60 to 120 min exposure to the glycoside (2.24 \pm 0.38 \text{ g, N} = 8). When the contracture was completed, no contractile response to transmural stimulation was obtained. However, in preparations exposed for 30 to 60 min to ouabain in which the plateau of the contracture was not attained yet, transmural stimulation caused appreciable increase in the tension which failed to return to the level prior to stimulation. It appears that development of ouabain-induced contracture is accelerated by endogenous noradrenaline released. Repeated washing of preparations with fresh solution caused a gradual relaxation (1.10 \pm 0.10 \text{ g, N} = 4, after 90 to 120 min) and a reappearance of the contractile response to transmural stimulation (the tension developed at 20/sec: 30 \pm 7.3\% of the value prior to ouabain, N = 4).

**Cocaine**

In 9 aortic preparations the magnitude, the duration and the time-to-peak of contraction in response to transmural stimulation at 20/sec averaged 0.95 \pm 0.10 \text{ g, 2.03} \pm 0.27 \text{ min and 0.90} \pm 0.06 \text{ min, respectively. Cocaine potentiated the response in a dose-dependent manner: mean values of the percent increase in these parameters by cocaine at 10^{-6} \text{ M were 25} \pm 3.5, 67 \pm 9.5 and 28 \pm 4.0, respectively (N = 8). The increase in the duration greatly exceeded that in the developed tension.**

**DISCUSSION**

It has been shown that adrenaline and K+ cause vascular contraction in association with
increased flux of Ca\(^{2+}\) into smooth muscles (4,10). The Ca\(^{2+}\) influx during K\(^{+}\)-induced contracture is dependent on [Ca\(^{2+}\)]\(_o\), up to 0.3 mM and there is a linear relationship between the tension developed and the rate of entry of Ca\(^{2+}\). Contractile response of arterial smooth muscles to adrenaline and a rise in [K\(^{+}\)]\(_o\) are promptly prevented by Ca\(^{2+}\) deionization (6). In the present study, removal of Ca\(^{2+}\) from bathing media markedly reduced the responsiveness of aortic smooth muscles to noradrenaline and electrical transmural stimulation which causes excitation of nervous elements in vascular tissues and thereby releases endogenous noradrenaline (12). Strontium ions partly restored the sensitivity to noradrenaline but completely restored the responsiveness to transmural stimulation applied at all frequencies used except for 5/sec. Similar restoration of the response to noradrenaline, histamine and K\(^{+}\) by Sr\(^{2+}\) applied to the Ca\(^{2+}\)-free solution has been shown in rabbit aortic strips (17). More marked restoration by Sr\(^{2+}\) of the response to sympathetic nerve stimulation than that to exogenous noradrenaline is also observed in isolated rabbit atria exposed to Ca\(^{2+}\)-free media (18). It seems likely that Sr\(^{2+}\) substitutes for Ca\(^{2+}\) in supporting release of vascular noradrenaline and in supporting sensitivity of receptors and contractility of muscles to endogenous noradrenaline but, only in part, those to exogenous noradrenaline. It has been suggested that Sr\(^{2+}\) serves as a current-carrying ion in crustacean muscle fibers (24) and in rabbit atrial myocardium (18). When Ca\(^{2+}\) in perfusion media is raised from 2.2 mM to higher concentrations, the contractile response to noradrenaline is reduced but that to periarterial sympathetic nerve stimulation is potentiated in isolated central ear arteries of the rabbit (13). The likely explanation of this action is that there is an increase in the amount of noradrenaline released from the nerve. The present study performed on isolated aortic strips showed that the contractile response to transmural neural stimulation was not potentiated by raising either [Ca\(^{2+}\)]\(_o\), or [Sr\(^{2+}\)]\(_o\).

Decrease in [Na\(^{+}\)]\(_o\) caused a sustained increase in the tension: the extent of the developed tension varied inversely with [Na\(^{+}\)]\(_o\). According to Briggs and Melvin (4), a reduction in [Na\(^{+}\)]\(_o\) results in increased influx of Ca\(^{2+}\) in vascular smooth muscles. Thus, antagonism of Na\(^{+}\) to Ca\(^{2+}\) is suggested. Although both raising [Ca\(^{2+}\)]\(_o\) and lowering [Na\(^{+}\)]\(_o\) actually increased resting tension, the magnitude of contracture did not relate directly to the ratio [Ca\(^{2+}\)]\(_o\)/[Na\(^{+}\)]\(_o\)\(^2\). This was also true in the case of the tension developed by transmural stimulation and of the sum of the tension developed by the ions and by the stimulation. As already suggested (25) no simple competition between Na\(^{+}\) and Ca\(^{2+}\) for a specific anionic site controls the vascular tension and the contractile response of isolated rabbit aortae to noradrenaline and electrical transmural stimulation. When [Na\(^{+}\)]\(_o\) was reduced to about 64% normal, the contractile response to transmural stimulation was potentiated despite the increased resting tension. Further reduction in [Na\(^{+}\)]\(_o\) inhibited the contractile response, although the sum of the tension developed by reducing [Na\(^{+}\)]\(_o\) and by transmural stimulation was increased. Marked prolongation of the duration of contraction observed at reduced [Na\(^{+}\)]\(_o\) would suggest involvement of an inhibition of mechanisms by which noradrenaline is inactivated. It is known that the uptake of noradrenaline by sympathetic nerve terminals is inhibited by reducing [Na\(^{+}\)]\(_o\) (26, 27). Cocaine, a potent inhibitor of amine uptake,
caused an increase in the duration of contraction which greatly exceeded an increase in the magnitude of contraction.

Contractile responses to transmural stimulation and noradrenaline were potentiated by $3.4 \times 10^{-7}$ M ouabain. The cardiac glycoside at $3.4 \times 10^{-6}$ M caused a gradual increase in the tension and a potentiation of the response to electrical stimulation, whereas the drug in higher concentration produced an additional increase in the tension but blocked the response. These effects resemble those of lowering [Na$^+$]$_o$. Activities of Mg ATPase in homogenates of rabbit aortae are stimulated by the addition of Na$^+$ plus K$^+$ (28). Ouabain fails to influence stimulation of the enzyme by Na$^+$ alone or K$^+$ alone but significantly inhibits the Na$^+$ plus K$^+$ activated activity (29). It is possible that ouabain permits a greater amount of noradrenaline to reach receptors by inhibiting uptake of the amine into sympathetic nerves (30). However, in the isolated rat uterus, concentrations of ouabain sufficient to cause potentiation of contractile responses to agonists fail to produce inhibition of a Na$^+$-activated membrane ATPase (31). The present study results prove that potentiation of the contractile response to electrical stimulation by ouabain was different from that by cocaine. Cocaine markedly prolonged the induced contraction, and was suggested to result from an inhibition of the uptake of noradrenaline by sympathetic nerve terminals (32). On the other hand, ouabain increased the amplitude in parallel with the duration of contractions. The saphenous vein treated with acetylstrophanthidin showed increased reactivity to added noradrenaline, Ba$^{++}$ and raised [K$^+$]$_o$ (33,34). Thus, it can be considered that the mechanism of potentiation by the glycoside involves increased fluxes of Ca$^{++}$ across cell membranes (21, 35) and increased availability of cellular Ca$^{++}$. According to Bohr (36), in arterial smooth muscles, increased reactivity to catecholamines seen in the presence of desoxycorticosterone occurs by loosening of intracellular Ca$^{++}$ bonds, thus permitting more Ca$^{++}$ to be released during excitation.

SUMMARY

Spirally-cut strips of the ascending aorta from rabbits were transmurally stimulated at frequencies of 5, 20 and 100/sec. Removal of Ca$^{++}$ from bathing media markedly reduced the contractile response to transmural stimulation, the reduction relating inversely to stimulation frequency. Strontium ions restored the response. Decrease in [Na$^+$], to 103.2 mM increased the resting tension of aortic strips and potentiated the contractile response to transmural stimulation. The effect of noradrenaline was slightly potentiated. Further reduction of [Na$^+$], caused a marked increase in the resting tension and an inhibition of contractile responses to transmural stimulation and noradrenaline. Magnitude of the contracture caused by various [Na$^+$], and [Ca$^{++}$], and the sum of the tension developed by variations in [Na$^+$], and [Ca$^{++}$], and by transmural stimulation did not relate directly to the ratio [Ca$^{++}$]/[Na$^+$],. However, the dependence on reducing [Na$^+$], was markedly greater than that on elevating [Ca$^{++}$]. Contractile response to transmural stimulation was potentiated and the dose-response curve of noradrenaline was moved left by ouabain ($3.4 \times 10^{-7}$ and $3.4 \times 10^{-6}$ M). Further increase in the ouabain concentration to $1.7 \times 10^{-5}$ M
caused marked increase in the resting tension but abolished the response to transmural stimulation. Repeated washing of preparations elicited partial relaxation and restoration of the response. Cocaine (10^{-6} and 3 \times 10^{-6} M) potentiated the response: increase in the duration of the induced contraction greatly exceeded increase in the maximum tension developed. It seems likely that Sr^{++} is effective in restoring release of noradrenaline from nerve terminals and responsiveness to noradrenaline in aortae deprived of Ca^{++}, and that ouabain exerts its potentiating action at a site different from the sites of action of cocaine: the glycoside would increase cellular Ca^{++} available for contraction, as does lowering [Na^{+}].

REFERENCES

1) ROBERTSON, P.A.: Nature 186, 316 (1960)
2) EDMAN, K.A.P. AND SCHILD, H.O.: J. Physiol. 155, 10P (1961)
3) DURBIN, R.P. AND JENKINSON, D.H.: J. Physiol. 157, 90 (1961)
4) BRIGGS, A.H. AND MELVIN, S.: Am. J. Physiol. 201, 365 (1961)
5) WAUGH, W.H.: Circulation Res. 11, 264 (1962)
6) WAUGH, W.H.: Circulation Res. 11, 927 (1962)
7) YASARGIL, G.M.: Helv. Physiol. Acta 18, 491 (1960)
8) SCOTT, J.B., FROHLICH, E.D., HARDIN, R.A. AND HADDY, F.J.: Am. J. Physiol. 201, 1095 (1961)
9) FEINBERG, H., BOYD, E. AND KATZ, L.N.: Am. J. Physiol. 202, 643 (1962)
10) BRIGGS, A.H.: Am. J. Physiol. 203, 849 (1962)
11) BOHR, D.F.: Science 139, 597 (1963)
12) SU, C. AND BEVAN, J.A.: J. Pharmac. exp. Ther. 172, 62 (1970)
13) FARMER, J.B. AND CAMPBELL, I.K.: Br. J. Pharmac. Chemother. 29, 319 (1967)
14) HEILBRUNN, L.V. AND WIERCINSKI, F.J.: J. Cellular Comp. Physiol. 29, 15 (1947)
15) FRANK, G.B.: J. Physiol. 163, 254 (1962)
16) FRANK, G.B.: J. Pharmac. exp. Ther. 139, 261 (1963)
17) HUDGINS, P.M.: J. Pharmac. exp. Ther. 170, 303 (1969)
18) TODA, N.: Circulation Res. 28, 545 (1971)
19) LÜTTGAU, H.C. AND NIEDERGERKE, R.: J. Physiol. 143, 486 (1958)
20) HORSF, W.D., KOPIN, I.J. AND RAMEY, E.R.: Am. J. Physiol. 215, 817 (1968)
21) LÜLLMANN, H. AND HOLLAND, W.: J. Pharmac. exp. Ther. 137, 186 (1962)
22) GOVIER, W.C. AND HOLLAND, W.C.: Am. J. Physiol. 199, 195 (1964)
23) TODA, N., USUI, H. AND MORI, J.: Jap. J. Pharmac. 21, 59 (1972)
24) FATT, P. AND GINSBORG, B.L.: J. Physiol. 142, 516 (1958)
25) NASI, C.W., LUCHKA, E.V. AND JHAMANDAS, K.H.: Can. J. Physiol. Pharmac. 44, 147 (1966)
26) IVERSSEN, L.L. AND KRAVITZ, E.A.: Mol. Pharmac. 2, 360 (1966)
27) BOGDANSKI, D.F. AND BRODD, B.B.: Life Sci. 5, 1563 (1966)
28) BONTING, S.L., SIMON, K.A. AND HAWKINS, N.M.: Arch. Biochem. Biophys. 95, 416 (1961)
29) WOLOWYK, M.W., KIDWAL, A.M. AND DANNIEL, E.E.: Can. J. Biochem. 49, 376 (1971)
30) DENGLER, H.J., SPIEGEL, H.F. AND TITUS, E.O.: Science 133, 1072 (1961)
31) DANNIEL, E.E., M Massingham, R. AND NASMYTTI, P.A.: Br. J. Pharmac Chemother. 34, 231P (1968)
32) TODA, N.: J. Pharmac. exp. Ther. 179, 198 (1971)
33) BRENDER, D., VANHOUTTE, P.M. AND SHEPHERD, J.T.: Circulation Res. 25, 597 (1969)
34) BRENDER, D., STRONG, C.G. AND SHEPHERD, J.T.: Circulation Res. 26, 647 (1970)
35) SEKUL, A.A. AND HOLLAND, W.C.: Am. J. Physiol. 199, 457 (1960)
36) BOHR, D.F.: Can. Med. Ass. 90, 174 (1964)