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Severe de novo liver injury after Moderna vaccination – not always autoimmune hepatitis

To the Editor:

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) infection has led to an unprecedented race to develop vaccines, one of which is the Moderna mRNA-1273 vaccine. Liver injury from autoimmune hepatitis (AIH) triggered by the Moderna vaccine has been reported. We present a case of liver injury post-Moderna vaccination, not presenting in an AIH pattern.

A 34-year-old Burmese man presented on 18th June 2021 with a 2-week history of pruritus, fever and jaundice. He received the Moderna vaccine on 2nd June 2021. He was prescribed antihista-

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phosphatase 149 U/L, total protein 84 g/L, albumin 46 g/L, globulin 38 g/L. Coagulation profile was normal: prothrombin time 9.9 seconds and international normalised ratio 0.91.

Extensive investigations performed were normal: this included serologies and PCRs for hepatitis A, B, C and E, cytomegalovirus, Ebstein-Barr virus, and herpes simplex virus. AIH screening included anti-smooth muscle antibody 5 units (ELISA, negative <20 units, positive >20 units), negative anti-liver kidney microsom al antibody (indirect immunofluorescence), anti-nuclear antibody ratio 0.11 (ELISA, positive ≥ 1), anti-mitochondrial antibody 2 units/ml (ELISA, positive ≥20 units/ml), IgG 14.6 (7.0-16.0 g/L), IgM 0.94 (0.40-2.30 g/L), ferritin 206 (7.0-16.0 g/L), IgM 0.94 (0.40-2.30 g/L), ferritin 206 (47-452 µg/L) and caeruloplasmin 32.4 (15.0-45.0 mg/dL) were normal. Leptospirosis IgM and Weil-Felix serology returned non-reactive. Computed tomography of the abdomen and magnetic resonance cholangiopancreatography were unremarkable.

Liver biopsy was performed on day 6 of presentation (day 22 following vaccination) and demonstrated acute lobular hepatitis with cholestasis. There was hepatocytic and canicular cholestasis, accompanied by predominantly lobular inflammation (Fig. 1A-B). Few portal tracts showed mixed inflammatory infiltrate with scattered eosinophils (Fig. 1C).

Immunohistochemistry (IHC) was performed. Tissue sections were cut onto Bond Plus slides (Leica Biosystems, Richmond) and heated at 60°C for 20 min. Tissue slides underwent deparaffinization, rehydration and heat-induced epitope retrieval with Bond Epitope Retrieval Solutions using a Leica Bond Max autostainer (Leica Biosystems, Germany) before endogenous

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peroxidase blocking (Leica Biosystems, Newcastle). Slides were labelled with antibodies targeting the SARS-CoV-2 spike protein (GeneTex, Cat#GTX632604, 1A9) at 1:8,000 dilution (ER2, pH 9.0, #AR9640) and SARS-CoV-2 nucleocapsid (Novus Biologicals, Cat#NB100-56576) at 1:250 dilution (ER1, pH 6.0, #AR9961). Positive and negative controls were included. Indirect IHC was performed using secondary antibody amplification with post-primary antibody and Bond Polymer Refine Detection (DS9800). Nuclei were counterstained with haematoxylin. To support the positive staining, antibodies used were previously tested in a separate cohort where tissues were obtained prior to 2019 to rule out non-specific staining.\textsuperscript{7} IHC showed patchy centrovenular granular positivity for the SARS-CoV-2 spike protein, within the cytoplasm of (E) hepatocytes in a granular cytoplasmic pattern, as well as within scattered (F) Kupffer cells in the sinusoids. Immunohistochemical staining with antibodies against SARS-CoV-2 nucleocapsid protein shows negative staining (G and H). (I) Trend in liver function tests. ALP, alkaline phosphatase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; UDCA, ursodeoxycholic acid. (This figure appears in color on the web.)
histological findings, absence of alternative causes and proximity of vaccination to jaundice onset, we postulate this might be related to the vaccine triggering an immune-mediated liver injury.

The patient was started on ursodeoxycholic acid. Bilirubin remained stable around 200–250 μmol/L for about 4 weeks and improved with normalisation (Fig. 1) 5 weeks later (10 weeks after receiving the vaccination). He subsequently received a full course of Sinovac vaccination with no ill-effects.

Unique to this case is the demonstration of anti-SARS-CoV-2 spike protein within the liver parenchyma. A crucial negative to validate our hypothesis is the exclusion of concurrent COVID-19 infection. The patient had negative nasal PCRs before and after Moderna vaccination but unfortunately such tests can be false negative.8,9 Temporally flanking negative nasal PCRs and a negative SARS-CoV-2 nuclecapsid staining (on the same biopsy sample that demonstrated SARS-CoV-2 spike protein) reasonably exclude concurrent COVID-19 infection.

The exact mechanism by which SARS-CoV-2 spike protein reached the liver is unknown. It was likely produced by skeletal myocytes at the site of administration, gained access to the blood stream10 and travelled to the liver. An alternative explanation is phagocytosis of SARS-CoV-2 spike protein by local macrophages or other antigen-presenting cells which migrated to the liver. Unfortunately, there is no published data on the presence or absence of mRNA-induced SARS-CoV-2 spike protein in liver tissue in asymptomatic patients. Our patient tolerated the inactivated Sinovac vaccine suggesting the immunological processes may be unique to mRNA vaccination.

To our knowledge, this is the first published report on de novo liver injury developing post-Moderna vaccination with supportive histology of a possible direct effect from anti-SARS-CoV-2 spike protein antibodies, without features of AIH.

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CMN and ZHSL contributed to writing of the manuscript and collection of data. WQL and PSJY contributed to the pathology slides staining and technical aspects of immunohistochemistry. WQL, PSJY and GHH contributed to writing and final editing of the manuscript. GHGH conceptualised the manuscript. All authors were involved in the final approval of the submitted manuscript.

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Author names in bold designate shared co-first authorship

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Chan Maung Nyein1,4
Zi Hui Sherylyn Liew2,4
Wei-Quang Leow3,4
Poh Sheng Joe Yeong1,4,5
Gim Him Ho1

1Division of Gastroenterology, Department of Medicine, Khoo Teck Puat Hospital, Singapore
3Department of Gastroenterology and Hepatology, Singapore General Hospital, Singapore
2Department of Gastroenterology and Hepatology, Singapore General Hospital, Singapore
4Department of Anatomical Pathology, Singapore General Hospital, Singapore
5Duke-NUS Medical School, Singapore
6Institute of Molecular Cell Biology, A*STAR, Singapore

*Corresponding author. Address: Khoo Teck Puat Hospital Singapore, 90 Yishun Central, Singapore 768828; Tel.: (65) 6555800, fax: (65) 6602 3700.

E-mail address: ho.gim.hin@ktph.com.sg (G.H. Ho)