In vitro estimation of superfluid critical extracts of some plants for their antimicrobial potential, phytochemistry, and GC–MS analyses

Waleed Bakry Suleiman1,2*

Abstract

Background: Along with swift economic evolution and continuous amelioration of lifestyle, people at present are paying more attention to health issues. Synthetic drugs will be compensated with other natural ones that belong to natural origin. Plants have always been considered as sources of several compounds that are used in many fields, especially human and animal health, starting from boosting immunity to the treatment of infectious diseases caused by some pathogenic microbes such as bacteria, fungi as well as viruses. This study aimed to incorporate some types of plants within the antimicrobial portfolio through the examination of different six plants which were Cichorium intybus, Cinnamomum camphora, Commiphora myrrha, Foeniculum vulgare, Nerium oleander, and Spartium junceum. As well, attempting to identify the active constituents of their extracts using GC–MS.

Materials and methods: All selected plants were analyzed to determine their phytochemical composition such as phenolics, alkaloids, flavonoids, terpenoids, and so on. The extraction step was done by sophisticated equipment called supercritical fluid extractor SFE through adjustment of specific conditions include temperature, time, flow rate and pressure to change the behavior of CO2. Testing the antimicrobial activity of each plant extract via agar well diffusion method through the formation of clear zones against a wide range of test microorganisms including both Gram-positive and Gram-negative bacteria as well as yeasts. Finally, attempting to primarily identify the constituents of each plant extract using GC–MS.

Results and discussion: The crude extract of F. vulgare showed the highest potency against C. albicans, E. faecalis and S. typhimurium, it contains some unique compounds such as squalene, eugenol and isoeugenol while, Extract of C. intybus showed a moderate activity especially against C. lipolytica and MRSA and it includes Vitamin A like compound which indicates antioxidant property.

Conclusion: Conclusively, fennel gave a promising result as a good wide spectrum antimicrobial agent because it contains some compounds act as antimicrobial agents such as eugenol which was used as food preservatives in addition to squalene which acts as an antioxidant and antymycotic agent so, it will be useful especially while it was used in highly purified form excluding all undesirable subcomponents.

Keywords: Phytochemistry, GC–MS, SFE, Cichorium, Cinnamomum, Commiphora, Foeniculum, Nerium, Spartium, Antimicrobial

Background

The majority of microorganisms are free-living and perform useful activities however, some microorganisms caused diseases are called pathogens. They include bacteria, viruses, fungi, and protozoa. Infection occurs...
when a pathogen invades a body that resulted in clinical infection, such an infection is referred to as subclinical asymptomatic disease [1]. Microbial infections are most frequently caused by the resident microflora of the host rather than by exogenous invaders. Generally, harmless microorganisms may become virulent due to changes in the host’s tolerance or an alteration in the host’s microfloral flora because of antibiotic use [2]. Antimicrobials reduced the morbidity and ameliorate human health against microbial infections. The incidence of antimicrobial resistance is increasing among all types of patients. Not all infections require specific antimicrobial treatment and careful clinical judgment is essential to determine whether symptomatic treatment is enough. Microbiological investigations should always be carried out before treatment [3]. Excessive use of antibiotics without advice of physicians, non-adjusted course of the antibiotic or uncompleted dosage, unsuitable antibiotic administration; all of these are the reason for bacterial resistance. Physicians should request a report of bacterial culture sensitivity (leastwise, as a phenotypic method) in cases of bacterial infection to detect the suitable antibiotic. Most of the Staphylococcus aureus isolates became MRSA (methicillin-resistant S. aureus), some of them became VRS (vancomycin-resistant S. aureus), that means the scientists should act more and more to incorporate either more generations of antibiotics or new categories of antibiotics especially those belong to natural origin like plants, mushroom, truffles [4]. The use of plants in the pharmacological treatment of disease began long ago [5]. Centuries ago, Chinese, Japanese, and Indian used herbs in disease treatment as traditional medicine [6, 7]. In Europe and North America; herbal products have been expanded for use under many terms such as ‘alternative’, ‘complementary’, ‘holistic’ or ‘integrative’ medical systems [8].

Chicory (Cichorium intybus) was widely distributed in Africa, Asia–temperate, Asia tropical, Europe, Australia, Northern America, and Southern America. The phytochemical screening of C. intybus showed the presence of tannins, saponins, flavonoids, terpenoids, cardiac glycosides, and anthocyanins overall plant parts; seeds, leaves, and stem [9]. The C. intybus seeds extract contained appreciable levels of phenolic and flavonoid contents [10]. The flowers of chichory contain saccharides, methoxycoumarin, cichorine, flavonoids, essential oils, and anthocyanins [11, 12]. Unfortunately, no antimicrobial activity was reported against a wide range of microorganisms by using the hexane extract while; ethyl acetate did [13]. The root and leaf extracts (methanol, distilled water, chloroform, petroleum ether, and acetone) of C. intybus were investigated for antibacterial activity against Gram-negative pathogenic bacteria, the extracts showed a wide spectrum of inhibition against the test pathogens. Methanolic extract of the root and leaf proved to have the strongest antibacterial activity [14].

Cinnamomum spp. belong to Lauraceae and had been examined extensively for their essential oil constituents. This genus includes about 250 species in the tropical and subtropical districts, mostly in Asia and some in South and Central America, and Australia [15]. Cinnamomum camphora was known for its medicinal traits in folk medicine. Phytochemical screening showed a presence of alkaloids, Tannins, and carbohydrates, as well as its methanolic extract, presented the maximum antimicrobial activity when compared with the other extracts; chloroform and petroleum ether [16].

Commiphora myrrha belongs to Brusieraceae and it was commonly known as “Myrrh” which is one of the most important medicinal plants. Traditionally, its resin was used in the tackling of wounds, gastrointestinal tract GIT disorders, diarrhea, coughing, thoracic pain [17], gingivitis [18] also, it is very effective in the treatment of urinary tract infection UTI [19].

Foeniculum vulgare Mill is called fennel and belongs to Apiaceae, it features with its aromatic fruits. It was commonly used for the treatment of different disorders as well as it acts as a digestive, carminative, and diuretic agent [20]. Preliminary phytochemical screening confirmed the occurrence of flavonoids, tannins, saponins, steroids, glycosides, terpenoids besides its antimicrobial activity due to its potential essential oil constituents [21] as well as several pharmacological advantages through its bioactive constituents that are very important for human health [22].

Nerium oleander Linn; belongs to Apocynaceae family. It is commonly known as Kaner, an evergreen flowering shrub, and extensively cultivated for its aroma. Its extracts have many pharmacological properties such as a diuretic, expectorant, and sternutatory agent. It is also a highly toxic plant due to the presence of a cardiac toxic hence, it was used topically [23]. Methanolic and water extract of different parts of the plant gave a reasonable activity against some pathogenic microorganisms [24].

Spartium junceum is one of the medicinal plants that belong to the Fabaceae family that was cultivated as ornamental plant [25], its flowers were used for the treatment of gastric ulcers in Turkey [26] it has miscellaneous functions as antioxidant, antifertility, simulator for uterine and GI contraction thus causing vomiting [27–29].

The phenolic compounds could be separated by supercritical fluid extraction apparatus and this technique has been studied extensively 20 years ago [30]. Supercritical fluid extraction had been considered to attain the essential oil from Psidium guajava L. leaves, additionally, the
non-volatile compounds were attained by CO₂ supercritical extraction [31].

The current study aimed to screen the phytochemical composition of all mentioned plants in the review and extraction by a more sophisticated technique known as supercritical fluid extracting equipment SFE under certain conditions and subsequent investigation of the ability of these SFE extracts to inhibit the growth of some pathogenic microorganisms including Gram-positive and Gram-negative bacteria in addition to some pathogenic yeasts and finally, these SFE extracts would be analyzed by gas chromatography to predict their ingredients as an attempt to identify the bioactive components.

**Materials and methods**

**Plants collected**

Six different plants were collected from the local market and had been finely ground by aggressive blender until turning to a fine powder for easy extraction and further investigations, the following Table 1 includes their scientific, common and Arabic name as well as the parts used for investigations and analyses.

**Quantitative assessment for phytochemical constituents**

Ten grams of the air-dried plant powder were extracted independently with 80% methanol three successive times. The extracts were concentrated, and the dried matter was then dissolved in 50 ml methanol. The alcoholic extracts were then completed to the volume of 100 ml by adding distilled water and used for the following determinations.

**Total phenolics**

One ml of the prepared extract of each specimen was completed to the volume of 10 ml by adding distilled water, then 1 ml of Folin–Ciocalteu reagent was added. The mixture was then shaken vigorously for 5 min, then 10 ml of 70% Na₂CO₃ was added and diluted immediately up to 25 ml by adding distilled water. The latter mixture was incubated for 90 min at room temperature. The absorbance was measured at 750 nm against the reagent used as a blank. A standard calibration plot was generated at 750 nm using known concentrations of gallic acid. The concentrations of phenols in the tested samples were calculated from the calibration plot and expressed as mg gallic acid equivalent of phenol/g of sample [32].

**Total flavonoids**

One ml of the prepared extract of each specimen was completed to the volume of 5 ml by adding distilled water. Immediately 0.3 ml 5% NaNO₂ was added and the mixture was then left for 5 min. Respectively 0.3 ml 10% AlCl₃ and 2 ml 1 M NaOH were added. The mixture was then diluted up to a volume of 10 ml with distilled water and the formed pink color was measured at 550 nm against the reagent used as a blank. A standard calibration plot was generated at 550 nm using known concentrations of rutin. The concentrations of flavonoids in the tested samples were calculated from the calibration plot and expressed as mg quercetin equivalent of flavonoids/g of the sample [33].

**Total tannins**

The total tannin content in the plant extract was estimated according to Folin–Deins reagent method [34], the absorbance was measured at 755 nm.

**Total saponins**

One gram of each plant powder was dispersed in 10 ml of 20% ethanol. Heating of the over a hot water bath for 4 h with persistent tilting at about 55 °C. The mixture was filtered, and the residue re-extracted with another 10 ml of 20% ethanol. The combined extracts were reduced to 2 ml over a water bath at about 90 °C. The concentrate was transferred into a 250 ml-separator funnel, and 5 ml of diethyl ether was added and shaken vigorously. The aqueous layer was retrieved whilst the ether layer was thrown away. The purification was done once again then 15 ml of n-butanol was added. The merged n-butanol extracts were washed twice with 10 ml of 5% aqueous sodium chloride, then the mixture was heated in a water

| Scientific name         | Common name       | Arabic name             | Part of interest |
|-------------------------|-------------------|-------------------------|-----------------|
| Cichorium intybus       | Chicory           | Shikoryah-Hindbah       | Seeds           |
| Cinnamomum camphora     | Camphor           | Kafour                  | Leaves          |
| Commiphora myrrha       | Myrrh             | Morr                    | Resin           |
| Foeniculum vulgare      | Fennel            | Shamar                  | Seeds           |
| Nerium oleander         | Oleander-Kaner    | Daflah                  | Flowers         |
| Spartium junceum        | Spanish broom     | Wézal                   | Leaves          |

Table 1 Data of the plant samples that collected to be analyzed including the scientific, common, Arabic names in addition to the part of interest
bath. After evaporation, the samples were dried in the oven at 65 °C to a constant weight. The total saponin content was expressed as a percentage [35].

**Total alkaloids**

Ten grams of each plant powder was extracted with 90% ethanol. Mayer’s reagent was used according to Woo and Püls [36].

**Total soluble carbohydrates**

*Extraction* According to the described method by Chaplin and Kennedy [37], the plant powder samples were dried at 65 °C till a constant dry weight and grounded again to a very fine powder by a clean mortar. One gram of the powder was put in a 100 ml conical flask, to which 5 ml of 2% phenol/water and 10 ml of 30% trichloroacetic acid were added. The mixture was shaken and kept overnight before being filtered; the filtrate was made up to 50 ml.

*Estimation* Contents of total soluble carbohydrates were determined using anthrone technique [38], the developed color was measured using an electric colorimeter at 620 nm. A blank mixture containing distilled water and reagent was used to set up the apparatus at zero optical densities.

**Total water-soluble proteins**

*Extraction* In this regard, one gram of the oven-dried powder at 60 °C and then transferred to 250 ml conical flask, then 10 ml distilled water, and 5 ml of 2% phenol solution were added. The contents of the flasks were shaken well and kept overnight before being filtered, and then they were used for the estimation of soluble proteins.

*Determination* The optical density of the resulted color was then read at the wavelength of 750 nm. The concentration of soluble protein present in the sample was then calculated making use of the constructed standard curve of proteins [39].

**Total nitrogen**

The total nitrogen content of each plant powder was determined according to Kjeldahl digestion [40].

**Volatile oil**

Fifty grams of each plant powder was exposed to steam distillation to extract volatile oils according to Balbaa’s method [41].

**Extraction by supercritical CO₂ fluid extractor (SFE)**

Ten grams of each plant powder were used to be extracted via SFE equipment at SFE lab, the regional center for mycology and biotechnology RCMB, Al-Azhar University. The (Teledyne ISCO SFX 200) includes carbon dioxide injection pumps, extractors, separators, compressors, carbon dioxide tanks, chillers.

The operation of extraction depends upon pushing the CO₂ through pipelines to be mixed with the samples under programmed critical conditions which gives CO₂ the solvent property, its ability as a tunable solvent differs with changing the critical conditions, these parameters include pressure, temperature, flow rate. In this experiment; the conditions applied were 300 bar, 55 °C and CO₂ flow rate of 6 g/min for 50 min.

**In vitro screening of antimicrobial activity of the plant SFE extracts**

Agar well diffusion method described by [42] was applied to determine the antimicrobial activity of six extracts against 10 pathogenic microorganisms; the first four belonged to Gram-positive bacteria, the next four belonged to Gram-negative bacteria and the last two belonged to yeasts, the inoculum of each test organism was prepared according to [43]. This assay was done using gentamycin as a positive control antibacterial and ketoconazole as a positive control antifungal while methanol was selected roughly as a negative control. The medium used for bacterial growth was Mueller–Hinton agar medium (Code; CM0337, Thermo Scientific, Oxoid Microbiology products) which composed of g/l; beef dehydrated infusion 300, casein hydrolysate 17.5, starch 1.5 and agar 17, this medium is ready to use which had been prepared by weighing 38 g and up to 1 l with distilled water. On the other aspect, yeast was cultivated onto malt extract agar MEA medium (code; CM0059, Thermo Scientific, Oxoid Microbiology products) which composed of (g l); malt extract 30, mycological peptone 5, agar 15, this medium is ready to use which had been prepared by weighing 50 g and up to 1 l with distilled water. The test microorganisms were provided by the lab of antimicrobial and sensitivity assays, the regional center for mycology and biotechnology, Al-Azhar University, Cairo-Egypt, in which this experiment had been done.

**Determination of MIC and MBC/MFC for the most active plant extracts**

Depending upon the clear zones diameters resulted by antimicrobial assay, out of the six plant extracts, two only were further investigated to determine their MIC values by which the lowest concentration of plant extract could inhibit the growth on the agar plate, MIC assay was accomplished by diluting the original crude extract (stock) through making these preparations (1:1, 1:2, 1:3, 1:4, 1:5, 1:6, 1:7). By the same protocol of agar well diffusion assay which applied to survey the antimicrobial activities of the six plant extracts, MIC also was
determined and recorded. Consequently, MBC by which the minimum bactericidal concentration and MFC by which the minimum fungicidal concentration was determined according to [44] by transferring 10 µl from the non-turbid wells to the appropriate broth medium permitting the growth after overnight incubation and finally the turbidity would be determined by a spectrophotometer (UV/visible spectrophotometer; Milton Roy Spectronic 1201) at 530 nm wavelength according to [45].

GC–MS analysis for prediction of subcomponents
GC–MS was carried out on Direct Probe Controller Inlet part to Single Quadrupole mass analyzer (Thermo Scientific; GC–MS model ISQ LT) using Thermo X-Caliber software. The Mass spectroscopy system was used to predict the subcomponents of plant SFE extracts using RTX-2330 (fused silica) 30 m capillary column of 0.25 µm internal diameter and df (µm) 0.20 µm. The column was operated at an initial temperature of 160–250 °C at the rate of 5 °C/min and was held for 30 min. The injector and detector temperatures were 240 °C and 250 °C, respectively. The carrier gas (nitrogen) was supplied at a total flow rate of 50 ml/min with a split ratio of 200 and subcomponents were identified by comparison with a linked library.

Results and discussion
Quantitative phytochemical screening
The powders of examined plants were processed according to the specific protocols to assess the phytochemical composition belongs to each plant, all the data obtained were tabulated in Table 2 which revealed the concentration of each component.

Regarding total phenolic compounds; *F. vulgare* has the highest value of 261 mg followed by *C. myrrha* 247 mg, while the lowest value 213 mg belonged to *N. oleander*. In the case of flavonoid concentration; *C. myrrha* contains the highest value of 159 mg followed by *C. intybus* 143 mg, while the lowest concentration belonged to *S. junceum*.

Regarding tannins; the uppermost percentage went to *C. intybus* (4.31%) followed by *C. camphora* (4.13%) while *S. junceum* contains the lowest concentration of tannins (2.87%). Referring total saponin values; *F. vulgare* contains the highest percentage (2.69%) followed by *N. oleander* (2.55%) while the lowest percentage (2.09%) belonged to *C. camphora*.

Concerning alkaloids concentrations, the highest value belonged to *C. intybus* (3.92%) followed by both *C. camphora* and *F. vulgare* (3.85%) while the lowest percentage belonged to *S. junceum* (2.87%).

In the case of total soluble carbohydrates; *C. camphora* possessed the highest value (3.48%) followed by *C. myrrha* (3.07%) while the lowest carbohydrates content belonged to *C. intybus* (2.11%). Regarding soluble proteins and total nitrogen content, the highest value belonged to *C. intybus* (15.94% and 26.76% respectively) and followed by *F. vulgare* (13.94% and 23.41% respectively) while, the lowest percentage of proteins belonged to *N. oleander* and the lowest nitrogen content belonged to *S. junceum*.

Finally, in the case of oil percentage; the highest concentration was detected in *F. vulgare* (0.81%) followed by *N. oleander* (0.69%) while, the minimum value was detected in *S. junceum* (0.41%).

Regarding phytochemical screening, all examined plant are rich in phenolics, flavonoids, alkaloids, saponins, tannins and this is the reason of their pharmacological properties as good medicinal plants especially, fennel, chicory, and myrrh, this result is in harmony with anthocyanin

| Item          | Item concentration for each plant |
|---------------|----------------------------------|
| *Cichorium intybus* | *Cinnamomum camphora* | *Commiphora myrrha* | *Foeniculum vulgare* | *Nerium oleander* | *Spartium junceum* |
| Phenolics (mg EGA) | 237 | 223 | 247 | 261 | 213 | 215 |
| Flavonoids (mg ER) | 143 | 138 | 159 | 131 | 124 | 118 |
| Conc. % | 4.31 | 4.13 | 4.05 | 3.87 | 3.74 | 3.27 |
| Tannins | 2.47 | 2.09 | 2.11 | 2.69 | 2.55 | 2.28 |
| Saponins | 3.92 | 3.85 | 3.49 | 3.85 | 2.99 | 2.87 |
| Alkaloids | 2.15 | 3.48 | 3.07 | 2.21 | 2.86 | 2.94 |
| Carbohydrates | 15.94 | 11.39 | 12.54 | 13.94 | 10.29 | 11.05 |
| Proteins | 26.76 | 21.53 | 21.96 | 23.41 | 21.47 | 20.98 |
| Total nitrogen | 0.62 | 0.58 | 0.47 | 0.81 | 0.69 | 0.41 |

Italic values indicate the highest measurement among all items of the investigated plants

Phenolics were estimated in mg eq/gallic acid, flavonoids were estimated in mg eq/rutin while the rest of other items were estimated and expressed as percentages
that had been from red chicory *C. intybus* in aqueous solution at pH 2.5 [46].

Also, the current results of myrrh are compatible with those who recorded the presence of terpenoids, steroids, tannins, volatile oils, and resins [47].

All data from Table 2 except total phenolics and total flavonoids (only phytochemical constituents that had been expressed in %) were configured in a stacked column chart (Fig. 1). By the first sight to Fig. 1, it could be noticed that *C. intybus* had the higher content followed by *F. vulgare*, *C. myrrha*, *C. camphora*, *N. oleander*, and the lowest plant content is *S. junceum*.

**Supercritical fluid extracting system**

By applying selected parameters, the extraction capacity ranged from 0.4 to 0.6 ml which represents 4–6 wt% approximately (the original concentration of the stock crude extract). The advantage of using this technique for extraction is a high precision, time-saving, and no solvent traces. This result expressed success of SFE equipment in extraction process; this is friendly with the results reported that the use of SFE in last years had been approved to be alternative for extraction of natural compounds like triterpenes which extracted by both SFE in corresponding to traditional extraction method; Soxhlet and the result of SFE was more satisfying [48]. As well, SFE is a green extraction method providing a concentrated end product with no undesirable residues [49]. The effect of SFE parameters changing to obtain better extractability of phenolic compounds from lavender flowers, different ranges of SFE parameters were checked such as 200 bar, 40 °C and 15 min and the yield was 4.3 wt% while 250 bar, 60 °C for 45 min gave 9.2 wt% (more than double) [50].

**Antimicrobial activity determination**

Antimicrobial activity of these six different plants was estimated, and the results had been expressed in mm as diameters of inhibition zones. All the data resulted were tabulated in Table 3 which revealed the following observations:

All test organisms exhibited variation in their responses against the examined crude extracts, *Candida albicans* exhibited more resistance than *C. lipolytica* among the examined yeast strains; only three extracts had moderate effects against *C. albicans* while all six extracts affected *C. lipolytica*.

In the case of Gram-positive bacteria; the most resistant strain was *S. mutans* followed by *Micrococcus sp.* and MRSA had been affected only by three extracts.
while *E. faecalis* was the most susceptible one that had been inhibited by all six investigated extracts.

Referring to Gram-negative bacteria; the most resistant strain was *K. pneumonia* followed by *E. cloaca* and *P. vulgaris* (only two extracts weakly inhibit its growth) while *S. typhimurium* was the most susceptible one that had been inhibited by four extracts.

On the other side, the investigated crude extracts might be categorized as potent, moderate, and weak antimicrobial agents according to the applied concentrations, also, some of them could be classified as wide spectrum while the others are narrow spectrum. For example; crude extract of *F. vulgare* showed the highest potency against *C. albicans*, *E. faecalis*, and *S. typhimurium*, it also showed a wide spectrum against seven out of 10. This result is in line with those who had investigated phytochemistry, antimicrobial activity and GC–MS of Portuguese fennel fruits that gave very compatible results with the current study where the crude extract has antimicrobial activity slightly lower than those belonging to the positive standard used; tetracycline as antibacterial and nystatin as antifungal [51]. Also, our finding showed a moderate effect of fennel essential oil and mupirocin has a significant eradicating effect against *S. aureus* and his finding will be useful antistaphylococcal agent [52].

Extract of *C. intybus* showed a moderate activity especially against *C. lipolytica* and MRSA with moderate range against four out of 10, while the extract of *C. myrrha* gave a moderate to weak activity with wide spectrum against six out of 10. This result in a harmony with hexane and ethyl acetate extracts of chicory roots that showed pronounced antibacterial activity against both Gram-positive and Gram-negative bacteria [53].

SFE extract of *C. myrrha* has a moderate antimicrobial activity according to the experimental circumstances, it is only active against five out of 10. This result slightly related to the conclusion related to myrrh extract which has a noticeable antibacterial activity against *Enterococcus faecalis* and *Fusobacterium nucleatum* involved in root canal infections especially when it combined with sodium chlorite [54].

Finally, *S. junceum* and *N. oleander* could be classified as weak and narrow-spectrum antimicrobial agents; they only active against three and two out of 10 respectively. This finding is consistent with the previously published data that are very limited about antimicrobial activity of Spanish broom. Oleander leaves have antimicrobial activity against *Bacillus pumilus*, *B. subtilis*, *S. aureus*, *E. coli*, and *Aspergillus niger* [55].

**Determination of MIC and MBC/MFC of the plant extracts**

According to the results of antimicrobial activity assay previously, the crude extracts of both of *C. intybus* and *F. vulgare* were chosen for further investigation for their values of MIC, MBC/MFC. The original concentrations of *C. intybus* and *F. vulgare* were 5 and 4% respectively. Different dilutions were prepared and investigated against the indicator strains via agar well diffusion assay, the data resulted was represented in Table 4. The MIC values for *C. intybus* against *Candida lipolytica*, MRSA, *E. faecalis* and *Salmonella typhimurium* were 1.25, 2.5, 1.67 and 1.67% respectively, clarifying that the most susceptible test organism is *C. lipolytica*, and accordingly MBC/MFC was determined, MFC of SFE extract of was 5% which was the original concentration and it was considered as the minimum fungicidal concentration against *C. lipolytica*, as well the MBC was 5% as the minimum bactericidal concentration against *E. faecalis* only, nonetheless the other susceptible bacteria were not completely killed so, the examined concentrations giving the clear zones were considered as bacteriostatic and

| Test organism                          | Chicory Stock (mm) | Chicory MIC % | Chicory MBC/MFC % | Fennel Stock (IZ) | Fennel MIC % | Fennel MBC/MFC % |
|----------------------------------------|--------------------|---------------|-------------------|-------------------|--------------|------------------|
| *Candida albicans*                     | ND                 | –             | –                 | 14                | 1.33         | ND               |
| *Candida lipolytica*                   | 18                 | 1.25          | 5                 | 15                | 1.33         | 4                |
| MRSA clinical isolate                  | 11                 | 2.5           | ND                | 8                 | 4            | ND               |
| *Enterococcus faecalis* ATCC 29212     | 18                 | 1.67          | 5                 | 22                | 0.8          | 2                |
| *Proteus vulgaris*                     | ND                 | –             | –                 | 7                 | 4            | ND               |
| *Salmonella typhimurium* ATCC 14028    | 14                 | 1.67          | ND                | 20                | 0.8          | 2                |

Stock represents the original concentration in corresponding to the inhibition zones diameters expressed in mm, while MIC and MBC/MFC describe the responsible concentration in %
these bacteria need a higher concentration to be killed completely.

On the other hand, the MIC values for *F. vulgare* against *C. albicans*, *C. lipolytica*, MRSA, *E. faecalis*, *Proteus vulgaris* and *S. typhimurium* were 1.33, 1.33, 4, 0.8, 4 and 0.8% respectively, clarifying that the most susceptible test organisms are *E. faecalis* and *S. typhimurium*. Accordingly, MBC/MFC was determined, MFC of SFE extract of was 4% against *C. lipolytica*, likewise, the MBC was 2% *E. faecalis* and *S. typhimurium*, nevertheless the other susceptible test organisms were not completely killed so, the examined concentrations giving the clear zones were considered as bacteriostatic and these strains need a higher concentration to be killed completely.

The entire results of MIC were in a harmony with [56] who reported variability of MIC values of *C. intybus* extracted by three different solvents against a limited number of test organisms. No much more information about the mechanism of action of antimicrobial agent(s) from *C. intybus* whether bactericidal or bacteriostatic. Regarding *F. vulgare*, the most valuable antimicrobial activity belongs to its essential oil neither alcoholic nor routine extraction methods [57] thus the study attempted to study an unusual method for extraction, and fortunately, it gave reasonable, considerable, valorized results.

**Gas chromatographic analyses**

*Chichorium intybus*

There 13 variable compounds were detected in *C. intybus* by GC–MS analysis (Table 5 and Fig. 2a); most of them belong to fatty acid whether saturated or unsaturated as well as fatty acid precursors. Also, some phenolic and terpenoids were detected. Retinal is also known as retinaldehyde, which is a form of vitamin A produced by oxidation of retinol which functions as the active component of the visual cycle, this compound is unique for chicory.

*Cinnamomum camphora*

Table 5 and Fig. 2b refer to predict the subcomponents of SFE *C. camphora* extract that point to the existence of 12 components belong to fatty acids and their precursors in addition to phenolics and terpenoids. Spathulenol is sesquiterpene alcohol as a unique compound for *C. camphora* during this study. On the other hand, Spirostenone belongs to terpenoids (isoprenoids), this compound found only in both *C. intybus* and *C. camphora*.

*Commiphora myrrha*

Table 5 and Fig. 2c expressed the prediction of subcomponents of SFE *C. myrrha* extract, analysis of GC–MS report displayed that 13 compounds had been detected; all these compounds belong to fatty acids and their precursors as well as phenolic and flavonoids.

**Foeniculum vulgare**

*Foeniculum vulgare* SFE extract was analyzed by GC–MS (Fig. 2d) to predict its subcomponents which were tabulated in Table 5 which revealed the detection of 15 subcomponents including fatty acids and their precursors. Eugenol and trans isoeugenol are unique for only *F. vulgare* which are classified as phenolic compounds, also squalene is alkene belong to isoprenoid compounds, this compound is also unique for *F. vulgare*.

**Nerium oleander**

*Nerium oleander* SFE extract was examined by GC–MS (Fig. 2e) to forecast its ingredients which were presented in Table 5 that shows the presence of 13 kinds of compounds including fatty acids and their precursors. Isofuranodionone is unique for *N. oleander* which is classified as a heterocyclic organic compound.

**Spartium junceum**

Only 8 compounds were detected in *S. junceum* SFE extract and this is the least content diversity among the examined plants, data was obtained by Fig. 2f that represents GC–MS chromatogram of *S. junceum* SFE extract, subsequently, this figure was analyzed to expect the ingredients which were arranged into Table 5 which showed a limited number and a limited diversity of subcomponents which belong to fatty acid (erucic acid) and precursors of fatty acids. GC–MS analyses for all examined plants showed the presence of fatty acids and fatty acids precursors in all investigated SFE extracts, although some compounds are unique for a specific plant among this study, on the other hands there are some compounds being common between two or more types of the plants under examination, for example; eicosanoic acid is monounsaturated (omega 9) fatty acid and it was detected in *C. intybus*, *C. myrrha*, and *N. oleander*. Erucic acid is also monounsaturated (omega 9) fatty acid and it was detected in *C. intybus*, *C. myrrha*, *F. vulgare*, *N. oleander*, and *S. junceum*. Doxepin derivative is an antidepressant molecule and it was detected in *F. vulgare*, *N. oleander*, and *S. junceum*. Spirostenone belongs to terpenoids (isoprenoids), this compound found only in both *C. intybus* and *C. camphora*. GC–MS report indicated the presence of residues of solvents involved in the extraction process plus the components constituting ethanolic clove extract [58] but fortunately, in this study, GC–MS report indicated the absence of the residues of solvents as an evidence to the high purity degree of the plant extract yielded by SFE.
Conclusion

Most of the examined plants have medicinal importance especially as antimicrobial activity and those are rich in their phytochemical contents whether alkaloids, flavonoids, etc. which encourages the researchers to investigate more uncommon plants for their medicinal importance, not antimicrobial activities in particular. SFE equipment is an inspiring technique for plant extraction which offers time and effort saving as well as high purity of the crude extract with no organic residues in addition to superior results than the conventional extraction methods. There is a very crucial need for antimicrobial agents to be incorporated more and more into the pharmaceutical market especially those belong to natural sources to overcome the problem of microbial resistance, and the current study presented a primitive inspiring trial to help in complementary and alternative medicine. Also, a combination of those examples of natural antimicrobial agents and other established drugs may offer synergistic powerful effect in tackling the resistance problem. GC–MS reports displayed that some examined plants contain very useful unsaturated fatty acids (omega 9) and hence, those antimicrobial extracts may have an additional advantage or a dual-action.

Table 5  GC-MS report of SFE plant extracts exhibiting RT, molecular weight, molecular formula and frequency

| Compound predicted                          | RT   | M. wt | M. formula | 1  | 2  | 3  | 4  | 5  | 6  | F |
|--------------------------------------------|------|-------|------------|----|----|----|----|----|----|---|
| Trans isoeugenol                           | 12.28| 164   | C_{10}H_{12}O_{2} | ● | ● |  |  |  |  | 1 |
| Eugenol                                    | 16.31| 164   | C_{10}H_{12}O_{2} | ● | ● |  |  |  |  | 1 |
| Cyclohexane derivative                     | 13.13| 202   | C_{12}H_{24}  |  |  |  |  |  |  | 1 |
| Dihydro butyl bezodoxepin                  | 15.95| 206   | C_{13}H_{18}O_{2} |  |  |  |  | ● | ● | 3 |
| Hydroquinone derivative                    | 15.95| 206   | C_{13}H_{18}O_{2} |  |  |  |  | ● | ● | 3 |
| Tetradecanol                               | 17.76| 214   | C_{14}H_{30}O  |  |  | ● |  |  |  | 2 |
| Spathulenol                                | 17.51| 220   | C_{13}H_{26}O   |  |  |  |  |  |  | 1 |
| Isofluraniononone                          | 18.11| 230   | C_{13}H_{26}O   |  |  |  |  | ● | ● | 1 |
| Hexadecanol                                | 17.76| 242   | C_{16}H_{34}O   |  |  |  |  |  |  | 4 |
| Tridecanoic acid methyl ester              | 20.68| 256   | C_{13}H_{26}O_{2} |  |  |  |  |  |  | 1 |
| Tetradecanoic acid methyl ester            | 20.68| 256   | C_{13}H_{26}O_{2} |  |  |  |  |  |  | 3 |
| Nonadecene                                 | 22.08| 266   | C_{19}H_{38}   |  |  |  |  |  |  | 3 |
| Retinal                                    | 22.82| 284   | C_{20}H_{42}O  |  |  |  |  |  |  | 1 |
| Hexadecanoic acid methyl ester             | 24.76| 270   | C_{19}H_{38}O_{2} |  |  |  |  |  |  | 6 |
| 9-Eicosene                                 | 26.04| 280   | C_{20}H_{42}O  |  |  |  |  |  |  | 2 |
| Nonadecanoic acid                          | 27.71| 296   | C_{19}H_{38}O_{2} |  |  |  |  |  |  | 1 |
| Octadecanoic acid methyl ester             | 28.03| 296   | C_{19}H_{38}O_{2} |  |  |  |  |  |  | 6 |
| 1-Eicosanol                                | 28.05| 298   | C_{20}H_{42}O  |  |  |  |  |  |  | 1 |
| Heptadecanoic acid ethyl ester             | 26.05| 298   | C_{19}H_{38}O_{2} |  |  |  |  |  |  | 1 |
| Octadecanoic acid methyl ester             | 28.50| 298   | C_{19}H_{38}O_{2} |  |  |  |  |  |  | 2 |
| Methyl stearate                            | 28.50| 298   | C_{19}H_{38}O_{2} |  |  |  |  |  |  | 3 |
| Linoleic acid ethyl ester                  | 29.11| 308   | C_{20}H_{42}O  |  |  |  |  |  |  | 1 |
| Eicosanoic acid                            | 29.21| 310   | C_{20}H_{42}O_{2} |  |  |  |  |  |  | 2 |
| Ethyl oleate                               | 29.22| 310   | C_{20}H_{42}O_{2} |  |  |  |  |  |  | 3 |
| Docosene                                   | 29.66| 308   | C_{22}H_{44}   |  |  |  |  |  |  | 3 |
| Erucic acid                                | 29.67| 338   | C_{22}H_{44}O_{2} |  |  |  |  |  |  | 5 |
| Eicosanoic acid derivative                 | 33.96| 310   | C_{22}H_{44}O_{2} |  |  |  |  |  |  | 1 |
| Behenyl alcohol or Doconasol               | 33.96| 326   | C_{24}H_{46}O  |  |  |  |  |  |  | 1 |
| Benzene dicarboxylic acid                  | 38.95| 390   | C_{26}H_{50}O_{4} |  |  |  |  |  |  | 6 |
| Docosanoic acid trihydroxy methyl ester    | 40.26| 402   | C_{31}H_{64}O_{3} |  |  |  |  |  |  | 1 |
| Squalene                                   | 44.30| 410   | C_{30}H_{58}O   |  |  |  |  |  |  | 1 |
| Spirostenone                               | 44.29| 428   | C_{27}H_{40}O_{2} |  |  |  |  |  |  | 2 |
| Flavone dioglucoside                       | 44.30| 594   | C_{27}H_{30}O_{15} |  |  |  |  |  |  | 1 |
| **Total**                                  | 33   | 33    | 33            | 13 | 12 | 13 | 15 | 13 | 8  |
Fig. 2 GC-MS chromatographs of SFE extracts of: a Cichorium intybus b SFE Cinnamomum camphora c Commiphora myrrha d Foeniculum vulgare e Nerium oleander f Spartium junceum

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Author details
1 Department of Botany and Microbiology, Faculty of Science, Al-Azhar University, The Permanent Camp St., 6th Ward, P.B. 13759, Nasr City, Cairo, Egypt. 2 Faculty of Advanced Science and Technology, Kumamoto University, Kurokami 2-3-91, Kumamoto 860-8555, Japan.

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