Morphological and physiological traits of the mesonephros in a freshwater fish, grayling *Thymallus thymallus*

E. A. Flerova**, **A. A. Morozov***, A. A. Bogdanova**, D. V. Chupov****, G. M. Ustuzhinskii****, V. V. Yurchenko***

*P. G. Demidov Yaroslavl State University, Yaroslavl, Russia**

**Yaroslavl Scientific Research Institute of Livestock Breeding and Forage Production – Federal State Budget Sciences Institution*

"Federal Williams Research Center of Forage Production and Agroecology", Mihajlovskij, Russia

****Papavin Institute for Biology of Inland Waters of Russian Academy of Sciences, Borok, Russia

****Polar Research Institute of Marine Fisheries and Oceanography, Murmansk, Russia

**E-mail:** katarinum@mail.ru

**Accepted 25.02.2019**

**Regul. Mech. Biosyst., 2019, 10(1), 9–15. doi: 10.15421/021902**

The study presents new data on the structural and functional organization of the mesonephros of the grayling *Thymallus thymallus* (Linnaeus, 1758). Adult grayling were sampled in the middle course of the Unya River, a tributary of the Pechora River (Komi Republic, Russia). The mesonephros of the grayling, as of other freshwater fishes, is composed by nephrons, blood vessels and hematopoietic tissue forming the renal interstice. In the interstice, cells with a radial vesicle array and chloride cells were discovered; the latter were mostly localized near the renal tubules. The degree of the interstice development in the mesonephros of the grayling was determined. New data on the ultrastructure of leukocytes, cells with a radial vesicle array, chloride cells, and nephron segments were obtained. A lack of mesangial cells, a small number of podocytes, and a thin basement membrane were observed on the sections of a renal corpuscle, being characteristic features of the ultrathin organization of the mesonephros in freshwater members of Salmoniformes and Esociformes. In the grayling’s nephrons, no neck segment was found, which was reported earlier for several species, including mammals. On the sections of proximal tubules, the ciliated cells were rare, and large amounts of the tubular-vesicular network in the zone of endocytosis of the type II epithelial cells were observed. On the sections of distal tubules, short blade-shaped cytoplasmic processes, with large numbers of invaginations of cytoplasmic membrane, were found. On the basis of the distinctive ultrastructure features mentioned above, the inference that grayling show the cytological markers of adaptation to euryhalinity was made. Thus, the results contribute to the knowledge of mesonephros development in fishes during their life history. From the species protection standpoint, our study provides baseline data on a WBC differential in the mesonephros as well as superoxide dismutase, catalase, glutathione S-transferase, and ethoxyresorufin-O-deethylase activities, which can be used in further studies addressing the health status of grayling populations.

**Keywords:** microranatomy; ultrastructure; leukocytes; antioxidants; ethoxyresorufin-O-deethylase; kidney.

**Introduction**

In the course of evolution, a general mechanism of non-specific protection was developed. Immune, antioxidant, and monoxygenase systems are the most important components of this protection. These systems provide the interaction between an organism and its environment. From a comparative-evolutionary perspective, it is very important to study the structural and functional features of the mesonephros, which not only plays the main role in the osmotic regulation, but also contributes significantly to homeostasis at the level of the non-specific protection. In this regard, studies on the salmonids, a unique family of euryhalineTeleosts, are of current importance.

There are a number of studies on various aspects of mesonephros morphology and physiology of anadromous salmonids in relation to the smoltification process (Langdon, 1985; Talbot et al., 1992; Boeuf, 1993; Mizuno et al., 2001; Björnsson & Bradley, 2007), while studies on morphological and physiological traits of the mesonephros in freshwater salmonids are few in number (Anderson & Loewen, 1975; Kocabatma & Ekingen, 1987; Katoh et al., 2008). The grayling, *Thymallus thymallus* (Linnaeus, 1758) is a freshwater salmonid species. The grayling is a benthopelagic fish, which feeds on chironomid and simulid larvae, drifting macroinvertebrates, ephemeropteran nymphs, fish larvae (Scott, 1985). It inhabits submontane reaches of rivers with a hard sand or stone bottom and well oxygenated, cold and fast-flowing water (Freyhof, 2011). The grayling is listed in the IUCN Red List with the Least Concern (LC) status. Grayling populations are threatened by overfishing as well as by natural system modifications, such as dam construction and water management, and habitat pollution with domestic and urban wastewaters, industrial, military, agricultural, and forestry effluents (Nykanen et al., 2001; Freyhof, 2011).

The purpose of our study was to perform an integrated research on structural and functional organization of the mesonephros of grayling. The data obtained in this study are a valuable contribution to the comparative-evolutionary conception of the mesonephros ultrastructure in lower vertebrates. From a practical perspective, our results can be used in the aquatic biomonitoring as well as in ichthyopathological assessment procedures in grayling culture.
Materials and methods

The fish sampling performed within the study was conducted by the Fresh Water Fishes Laboratory (SevPINRO) in July 2012. Grayling was sampled in the middle course of the Unya River, a tributary of the Pechora River (Fig. 1). The Unya River flows 163 kilometers and has a total catchment area of 2,890 square kilometers. The Unya River has a rocky stream bed, with rifts and rapids. By chemical composition, the river waters belong to the hydrocarbonate class; total dissolved solids concentrations vary from 200 to 500 mg/l. Fish sampling was carried out in the "Unyinskiy" protected area, which is located in a buffer zone of the Pechora-Ilych Nature Reserve. Due to its remoteness and inaccessibility, the area does not experience any significant anthropogenic impact, and thus, the area was considered suitable for the baseline study of morphological and physiological traits of the mesonephros in grayling.

![Sampling site](image)

**Fig. 1.** Sampling site (marked with a grey rectangle)

Fish were collected using a minnow seine, 20 m long, with a 3 mm mesh in a bag. Ten sexually mature fish were used for the study. Each specimen was immobilized by a blow to the head, measured and weighed (Table 1). Then it was dissected, eviscerated and weighed, the maturity stage was determined visually, on a 5-point scale, and the mesonephros samples were taken.

| Specimen | TL, cm | FL, cm | SL, cm | TW, g | EW, g | Sex |
|----------|--------|--------|--------|-------|-------|------|
| 1        | 29.2   | 27.0   | 25.9   | 205   | 179   | ♀ 3  |
| 2        | 27.7   | 26.5   | 25.0   | 192   | 172   | ♂ 2  |
| 3        | 27.6   | 25.4   | 24.1   | 174   | 154   | ♂ 2  |
| 4        | 27.0   | 25.3   | 24.0   | 175   | 154   | ♀ 3  |
| 5        | 25.5   | 23.6   | 22.5   | 130   | 115   | ♀ 3  |
| 6        | 23.9   | 22.3   | 21.0   | 126   | 106   | ♀ 2  |
| 7        | 23.5   | 22.1   | 20.9   | 113   | 98    | ♂ 2  |
| 8        | 23.4   | 22.0   | 20.7   | 100   | 85    | ♂ 2  |
| 9        | 22.7   | 21.0   | 19.8   | 80    | 63    | ♀ 3  |
| 10       | 20.2   | 18.3   | 17.3   | 63    | 54    | ♂ 2  |

Note: TL – total length, FL – fork length, SL – standard length, TW – total weight, EW – eviscerated weight, * – sex and maturity stage.

Mesonephros samples intended for biochemical assays were immediately frozen in liquid nitrogen and stored in it until the analytical procedure. Organ smears were made on slides, allowed to dry for 10 min and set in a container with 96% ethanol for 20 min. Samples intended for microanatomy and ultrastructure analyses were fixed in 2.5% glutaraldehyde solution prepared with 0.1 M phosphate buffer, postfixed in 1% osmium tetroxide for 1 hr, dehydrated in graded acetone and propylene oxide, and embedded in Araldite (Electron Microscopy Sciences, USA) (Timakova et al., 2014). Superoxide dismutase, catalase, and glutathione S-transferase activities were analyzed according to conventional spectrophotometric protocols described earlier (Morozov & Yurchenko, 2018) using a Perkin Elmer Lambda 25 spectrometer (UK). Ethoxyresorufin-O-deethylase activity was measured fluorometrically (described previously in Yurchenko & Morozov, 2018) using a Perkin Elmer LS 55 spectrometer (UK).

Microanatomical study of the mesonephros was carried out according to the standard methods. Semi-thin sections (2–3 μm) were cut with a UMTP-3 microtome (Russia). Dry sections were stained with methylene blue. Using a Micromed-6 light microscope (Russia), digital photographs were taken, which were then processed with Image Tool 3.0 software. The outer diameter (OD) of blood vessels, renal corpuscles and tubules were measured. The area of hematopoietic tissue was calculated as the difference between the total area of a section and the sum of the areas of blood vessels, renal corpuscles and tubules (Timakova et al., 2014).

Ultra-thin sections were cut using a Leica EM UC7 Ultracut (Germany). Sections were stained with uranyl acetate and lead citrate and analyzed using a JEM 1011 transmission electron microscope (Japan). Measurements were carried out on digital pictures.

A white blood cell (WBC) differential was studied on the Romanowsky-Giemsa-stained organ-smears using a Micromed-6 light microscope (Russia). A WBC differential analysis was performed on at least 200 cells at each organ-smear (Timakova et al., 2014). Data were reported as the mean ± standard error of the mean (x ± SE).

Results

**Biochemical analyses.** Enzyme activities in the mesonephros of grayling were as follows: ethoxyresorufin-O-deethylase – 0.22 ± 0.05 pmol/mg/min, superoxide dismutase – 1.08 ± 0.27, catalase – 1.23 ± 0.21, and glutathione S-transferase – 0.91 ± 0.19 μmol/mg/min.

**Mesonephros microanatomy.** In the mesonephros of grayling, the area of hematopoietic tissue was 46.9 ± 2.7% of the total area of the sections. The average OD of the blood vessels was 36.13 ± 2.28 μm (Fig. 2). In the mesonephros interstitial tissue, erythrocytes and leucocytes were distinguishable at the light-microscopy level (Fig. 2). Among the mature leucocytes, the lymphocytes were the most abundant and comprised 74.4 ± 2.8%. The second largest group (10.8 ± 2.3%) was represented by multipotential hematopoietic stem cells, hemocytoblasts. Granulocyte progenitors varied from 3% to 4% (promyelocytes 4.1 ± 0.8%, myelocytes 3.0 ± 0.9%, metamyelocytes 2.9 ± 0.9%). The amount of granulocytes decreased in the following order: banded neutrophils 2.5 ± 0.4%, segmented neutrophils 1.0 ± 0.5%, macrophages 0.90 ± 0.29%, and plasma cells 0.50 ± 0.10%.

The nephron segments were well distinguishable on the sections. The Bowman’s capsule was composed of epithelium, surrounded by an intensely stained basement membrane. A tuft of capillaries, the glomerulus, enclosed in the sac was distinguishable. These segments together, known as a renal corpuscle, had a size of 62.44 ± 3.82 μm. On the cross-sections of the proximal tubule, with an OD of 53.07 ± 1.77 μm, a brush border was distinguishable. The proximal tubule was composed of epithelial cells, enclosed by an intensely stained basement membrane. Each epithelial cell had an intensely stained oval or rounded nucleus, which in most cells, was located at the basal side. The distal tubule (OD 56.71 ± 2.59 μm) was also composed by epithelial cells, enclosed by the basement membrane; intensely stained oval or rounded nuclei were located at the central part of the cells. On the cross-sections of the distal tubule, a brush border was not present.

**Mesonephros ultrastructure.** Mesonephros interstitial tissue contained the following types of leukocytes: lymphocytes, plasma cells, macrophages, neutrophils, eosinophils, cells with a radiate villous array, and chloride cells. A characteristic feature of grayling’s lymphocytes was a large rounded nucleus, which occupied almost the entire cell volume. Heterochromatin was evenly localized to the periphery of the nucleus. The thin rim of the electron-light cytoplasm contained free ribosomes and two mitochondria on average (Table 2, Fig. 3b).

Data were analyzed using a JEM 1011 transmission electron microscope (Japan). Measurements were carried out on digital pictures.

Macrophages are the largest leukocyte cells. The nuclei were eccentrically shifted; heterochromatin was evenly localized to the periphery; euchromatin was evenly localized to the periphery of the nucleus. The thin rim of the electron-light cytoplasm contained free ribosomes and two mitochondria on average (Table 2, Fig. 3b).

| Specimen | TL, cm | FL, cm | SL, cm | TW, g | EW, g | Sex |
|----------|--------|--------|--------|-------|-------|------|
| 1        | 29.2   | 27.0   | 25.9   | 205   | 179   | ♀ 3  |
| 2        | 27.7   | 26.5   | 25.0   | 192   | 172   | ♂ 2  |
| 3        | 27.6   | 25.4   | 24.1   | 174   | 154   | ♂ 2  |
| 4        | 27.0   | 25.3   | 24.0   | 175   | 154   | ♀ 3  |
| 5        | 25.5   | 23.6   | 22.5   | 130   | 115   | ♀ 3  |
| 6        | 23.9   | 22.3   | 21.0   | 126   | 106   | ♀ 2  |
| 7        | 23.5   | 22.1   | 20.9   | 113   | 98    | ♂ 2  |
| 8        | 23.4   | 22.0   | 20.7   | 100   | 85    | ♂ 2  |
| 9        | 22.7   | 21.0   | 19.8   | 80    | 63    | ♀ 3  |
| 10       | 20.2   | 18.3   | 17.3   | 63    | 54    | ♂ 2  |
The cytoplasm contained electron-dense phagosomes of various sizes (Table 2), which occupied almost the entire cell volume. The cytoplasm also contained mitochondria, short double membrane sheets of the rough endoplasmic reticulum, free ribosomes, lysosomes, and small light vesicles (Table 2, Fig. 2d). In some cells, a large number of the electron-dense pigment granules were found.

**Fig. 2.** Microanatomy of the mesonephros of grayling, *Thymallus thymallus*: a – mesonephros tissues; b – lymphocyte; c – plasma cell; d – macrophage; e – neutrophil; f – intra-cellular granule of a neutrophil; g – eosinophil; h – cell with a radial vesicle array; i – chloride cell; j, k – cytoplasm of a chloride cell; 1 – blood vessel; 2 – hematopoietic tissue; 3 – renal corpuscle; 4 – proximal tubule; 5 – distal tubule; 6 – nucleus; 7 – heterochromatin; 8 – euchromatin; 9 – mitochondrion; 10 – rough endoplasmic reticulum; 11 – phagosome; 12 – intra-cellular granule of a neutrophil; 13 – Golgi apparatus; 14 – intra-cellular granules of an eosinophil; 15 – radial vesicle array; 16 – tubular reticulum; 17 – secondary lysosome.
Fig. 3. Ultrastructure of nephron in mesonephros of grayling, Thymallus thymallus: a – renal corpuscle; b – podocyte; c – section of the proximal tubule; d – brush border; e – cytoplasm of the type I epithelial cell of the proximal tubule; f – basal part of the type I epithelial cell of the proximal tubule; g – section of the proximal tubule with a ciliated cell; h – basal part of the type II epithelial cell of the proximal tubule; i – zone of endocytosis; j – basal part of the distal tubule; k – apical part of the distal tubule; l – glomerular capillary; 2 – podocyte; 3 – epithelial cell of the parietal layer of the Bowman’s capsule; 4 – Bowman’s space; 5 – basement membrane of the parietal layer; 6 – basement membrane of the visceral layer; 7 – nucleus; 8 – pedicels of a podocyte; 9 – podocyte; 10 – tubular basement membrane; 11 – secretory granules; 12 – mitochondrion; 13 – smooth endoplasmic reticulum; 14 – tubular-vesicular network; 15 – brush border; 16 – brush border microvilli; 17 – basal body of a cilium; 18 – cilium; 19 – blade-shaped cytoplasmic processes.
Neutrophils were rounded (Fig. 2e). The shape of a nucleus varied from bean-like to segmental. Heterochromatin was segmented, localized mostly along the nucleus membrane. The cytoplasm contained mitochondria, the double membrane sheets of the rough and smooth endoplasmic reticula; in single cells, the Golgi apparatus was identified. Characteristic feature of neutrophils are specific intra-cellular granules that fill the cytoplasm. In the grayling’s mesonephros, these granules had thin electron-dense stripes, fibrils, located along the granules. Two types of granules were found. The first type had fibrils located evenly along the granules. The second type had a lighter central part and dark fibrillar edges (Table 2, Fig. 2f).

Regul. Mech. Biosyst., 2019, 10(1)

| Parameter                  | Lymphocytes | Plasma cells | Macrophages | Neutrophils | Eosinophils |
|----------------------------|-------------|--------------|-------------|-------------|------------|
| Cell size                  | 6.17±0.36×5.09±0.41 11.07±1.04×7.47±0.93 15.35±1.28 10.46±0.38 15.18±0.65 7.07±0.43 9.92±1.01 6.63±0.45 |
| Nuclear size               | 5.10±0.23×4.36±0.33 5.54±1.34×4.49±0.63 4.72±0.40×0.35 5.03 4.33±0.52 2.77±0.35 5.68±1.85 3.42±0.59 |
| Mitochondrion size         | 0.53±0.11×0.31 0.05×0.42 ±0.06×0.20 0.02 – – – – |
| Number of mitochondria on a cell section | 2.00±0.71 1.00±0.82 – – – – – – |
| Large phagosome size       | – – 2.57±0.39×2.14±0.36 – – – – – – |
| Number of large phagosomes on a cell section | – – 3.83±0.87 – – – – – – |
| Small phagosome size       | – – 0.30±0.01×0.22±0.02 – – – – – – |
| Number of small phagosomes on a cell section | – – 23.40±4.92 – – – – – – |
| Size of specific granules, type I | – – 0.39±0.02×0.23±0.02 0.44±0.03×0.36±0.02 – – – – – |
| Size of specific granules, type II | – – 0.49±0.04×0.41±0.02 – – – – – – |
| Number of granules on a cell section | – – 6.25±1.66 11.67±1.08 – – – – – |

Ultrastructure analysis of the proximal tubules in the nephrons of grayling showed two types of epithelial cells. The type I epithelial cells were located at the beginning of the proximal tubule; these were the largest cells in the proximal tubule. These epithelial cells had an elongated pyramidal shape, were tightly adjacent to each other, and had round-shaped nuclei located in the basal part of the cells. The cytoplasm was acinous and contained a large amount of mitochondria (Table 4, Fig. 3e). Basal cell membrane formed projections turning into the sheets of the smooth endoplasmic reticulum (Fig. 3f). These membrane projections were characteristic for all types of epithelial cells in all divisions of the tubule. Lysosomes and the large electron-dense secretory granules were located closer to the periphery of the cell, heterochromatin was segmented. The rounded nucleus was surrounded renal tubules and blood vessels. The rounded nucleus was located closer to the periphery of the cell, heterochromatin was segmented. The cytoplasm was electron-light with a large amount of mitochondria. On the cell sections, single-membrane organelles were observed (Fig. 2h).

Chloride cells were diffused throughout the parenchyma, and also surrounded renal tubules and blood vessels. The rounded nucleus was located closer to the periphery of the cell, heterochromatin was segmented. The cytoplasm was electron-light with a large amount of mitochondria. On the cell sections, single-membrane organelles were observed and identified as the secondary lysosomes. Also, large electron-dense granules and two-double-membrane organelles, tubular and rough endoplasmic reticula were distinguished. Single tubules were distributed throughout the cell. Stacks of the rough endoplasmic reticulum were mainly located in the perinuclear area and were few in number (Table 3, Fig. 2i, 2j, 2k).

Table 3

| Dimensions (μm) and number of cells with a radial vesicle array, chloride cells, and their subcellular structures in the mesonephros of grayling, *Thymallus thymallus* (x±SE, n=20) |
|---------------------------------|-----------------|-----------------|-----------------|
| Parameter                       | Cells with a radial vesicle array | Chloride cells |
| Cell size                       | 5.27±0.67×4.78±0.68 17.67±3.16×5.22±0.09 |
| Nuclear size                    | 4.02±0.40×3.44±0.15 3.86±0.63×3.72±0.61 |
| Vesicle size                    | 0.69±0.08×0.48±0.04 – – |
| Number of vesicles on a cell section | 4.50±0.75 – – – – |
| Mitochondrion size              | – 0.67±0.09×0.54±0.15 – – – – – – |
| Number of mitochondria on a cell section | – 6.33±1.78 – – – – – – |

The type II epithelial cells were similar to the type I epithelial cells in general, but they were smaller in height (Table 4). The type II epithelial cells also had round-shaped nuclei located in the basal part of the cells. The heterochromatin was segmented, most of it located along the nuclear membrane, between the nuclear pores. The cytoplasm of these cells was less acinous, in comparison with that in the type I epithelial cells, and had a greater amount of larger mitochondria (Table 4). The membrane projections of the basal cell membrane were better develop-
ped. A characteristic feature of the type II epithelial cells was the absence of secretory granules in the cytoplasm. Although the tubulo-vascular network was present, the endocytosis zone was smaller in comparison with the type I epithelial cells (Table 4, Fig. 3h, 3i). The brush border was shorter; it was formed by both cilia, from ciliated epithelial cells, and microvilli (Table 4). In the ciliated type II epithelial cells, the cytoplasm had a greater amount of larger mitochondria in comparison with the main type II epithelial cells.

The epithelial cells in the distal tubule were high and very wide in the base (Table 4). The nuclei of most cells occupied the central part, sometimes were shifted to the basal part. Heterochromatin was both concentrated at the periphery of the nucleus between nuclear pores and diffused over the entire surface. In the cytoplasm of these cells, a large number of membrane projections were present. Large elongated electron-dense mitochondria, located strictly along the longitudinal axis throughout the cell, were observed (Fig. 3j). By the number of mitochondria, the epithelial cells of the distal tubule were superior to the type I epithelial cells and inferior to the type II epithelial cells of proximal tubules. The endocytosis zone was absent. In the apical part of the cells, the blade-shaped cytoplasmic processes, facing the lumen of the tubule, were observed. Some processes were rugose (Fig. 3k).

Discussion

Physiological endpoints of interest, i.e. antioxidant enzyme activities and monoxygenase activities, are well established biomarkers of chemical exposure (Whyte et al., 2000; Sayeed et al., 2003; Atli & Carli, 2010; Sinha et al., 2014). Our results provide baseline data on superoxide dismutase, catalase, glutathione S-transferase, and ethoxyresorufin-O-deethylase activities in the grayling kidney. These endpoints are considered components of an “early-warning system” for fish health and thus can be used in further studies addressing the health status of the grayling population in the Urya River. In an attempt to conduct a discussion of the results from a comparative-evolutionary perspective, we used data obtained earlier for the grayling’s close freshwater relative, the Northern pike *Esox lucius* (Lapirova et al., 2017). Northern pike is a member of the Esoctiformes, the sister order to the Salmoniformes within the Protacanthopterygii superorder, and it takes an ancestral position to the Protacanthopterygii superorder, are evident at the order level. A WBC differential is known to be a sensitive indicator of the physiological state of an organism and highly responsive to changes in an environment. A WBC differential has a wide reaction norm, which depends on the systematic position of the species as well. It was reported earlier (Keyster, 2009) that in vendace *Coregonus albula* from different parts of Lake Beloye, the percentage of neutrophils varied from 41% to 88%, which made 74% on average. In grayling, a WBC differential, in general, was similar to that found in the Northern pike (Lapirova et al., 2017).

The differences between members of the Proctoanthopterygii were observed not only in the granulocyte differential, but also in the quantity of granules on the sections of neutrophils and eosinophils as well as in the structure of specific granules in neutrophils. By the quantity of granules on the sections of neutrophils and eosinophils, the grayling had a resemblance to the Northern pike rather than to the more closely related rainbow trout and vendace (Flerova & Balabanova, 2013). It was reported earlier (Flerova, 2017) that in the mesonephros of the Northern pike, the neutrophils had specific fibrillar granules, which could be conditionally divided into three types. In the mesonephros of grayling, the neutrophils had fibrillar granules with the ultrastructure similar to the type I and type II specific fibrillar granules of the Northern pike. The characteristic feature of the type III specific fibrillar granules was that the fibrils were located close to the central part of the cell. Neutrophils of rainbow trout and vendace, only one type of specific fibrillar granule was found, with a light central part and dark fibrillar edges (Flerova & Balabanova, 2013). It was presumed earlier that the differences in the quantity of fibrils and their distribution between the granules of neutrophils are associated with a stage of the granule maturation rather than the species-specificity (Flerova & Balabanova, 2013). The frequency of neutrophils having granules of a certain type on the sections of mesonephros and the quantity of granules on the sections of neutrophils and eosinophils are possibly characteristic to the cellular immunity functioning in the particular environmental conditions typical of the species habitats.

A lack of mesangial cells, small amount of podocytes on the sections of the renal corpuscle, and a thin basement membrane indicate the high glomerular filtration rate, which is characteristic of freshwater fishes (Ojeda et al., 2006). In grayling, the nephron does not possess a neck segment, which was observed earlier in the mesonephros of some fish (Ojeda et al., 2006; Flerova, 2017) and mammalian species (Ojeda & Icardo, 1991). On sections of the proximal tubules, ciliated cells were rare, and large amounts of the tubulo-vascular network in the zone of endocytosis of the type II epithelial cells were observed. On sections of the distal tubules, short blade-shaped cytoplasmic processes, with large numbers of invaginations of cytoplasmic membrane, were witnessed. It is well established that ciliated segments of nephron (neck segment and ciliated cells of proximal tubules) are involved in the processing the fluid. The cilia move the urine toward the proximal tubule, reducing the pressure in the urinary space and, consequently, increasing the glomerular filtration rate (Ojeda et al., 2006). Development of the tubular-vesicular network promotes the reabsorption rate in the proximal tubules (Flerova, 2017). Lumen enlargement due to short blade-shaped cytoplasmic processes probably lowers the resistance to urinary flow.

In the study of Na+/K+/2Cl–-cotransporter localization in the kidneys of an euryhaline salmonid, rainbow trout (Katoh et al., 2008), a lack of the neck segment and a small amount of cilia, which were mostly found in the tubule lumen, was recorded. Rainbow trout was lacking the Na+/K+/2Cl–-cotransporter, which mediates the euryhaline transport of Na+, K+, and Cl across epithelial membranes, in the basolateral membrane of the proximal tubules. In the distal tubules, the apically
located Na⁺/K⁺/2Cl⁻ cotransporters were observed, which it has been suggested play a role in Na⁺, K⁺, and 2Cl⁻ absorption allowing the ions to be transported back into the body through basolateral-specific ion channels. Thus, the distal tubules could probably switch between secretion and absorption, depending on the osmoregulatory needs of the fish (Katoh et al., 2008). It is possible that the ultrastructural features observed in the grayling mesonephros in our study are the cytological markers of grayling being adapted to euryhalinity since it can occur in brackish waters and on small river rifts in the tidal backwater zones.

Conclusion

Our study provides new data on morpho-functional characteristics of the mesonephros in the grayling. The degree of the hematopoietic tissue development was determined. New data on the ultrastructure of leukocytes, cells with a radial vesicle array, chloride cells, and nephron segments were obtained. Common and distinctive features, as compared with members of Salmoniformes and Esociformes, in the mesonephros organization of grayling were reported. From the species protection standpoint, our study provides baseline data on a WBC difference in the grayling mesonephros in our study are the cytological markers of graying being adapted to euryhalinity since it can occur in brackish waters and on small river rifts in the tidal backwater zones.

Acknowledgements. The study was funded by RFBR, research project 16-04-00650 “Morpho-functional organization of mesonephros in Salmoniformes”. The authors are grateful to Vyacheslav S. Afanasyev and Igor M. Dyakov for their help in the field.

References

Anderson, B. G., & Loesern, R. D. (1975). Renal morphology of freshwater trout. American Journal of Anatomy, 143(1), 93–113.
Atli, G., & Canli, M. (2010). Response of antioxidant system of freshwater fish Oreochromis niloticus to acute and chronic metal (Cd, Cu, Cr, Zn, Fe) exposures. Ecotoxicology and Environmental Safety, 73(8), 1884–1889.
Balabanova, L. V. (2006). Клетки с radialno raspolozhennymi vezikulami u ryb raznyh vidov [Cells with radiating vesicles in different fish species]. Cytology, 49(8), 636–640 (in Russian).
Björnsson, B. T., & Bradley, T. M. (2007). Epilogue: Past successes, present misconceptions and future milestones in salmon smoltification research. Aquaculture, 273, 384–391.
Boeuf, G. (1993). Salmonid smolting: A pre-adaptation to the oceanic environment. In: Rankin, J. C., & Jensen, F. B. (eds.). Fish Ecophysiology. Chapman & Hall Fish and Fisheries Series. Vol 9. Springer, Dordrecht.

Flerova, E. A. (2017). Osobennosti struktury mezonefrosa obyknovennoy shuki (Thymallus thymallus) kelts following transfer from fresh water to seawater. Journal of Comparative Physiology B, 186(4), 487–498 (in Russian).
Flerova, E. A., & Balabanova, L. B. (2013). Ultrastructure of glomerulocytes of teleost fish (Salmoniformes, Cypriniformes, Perciformes). Journal of Evolutionary Biochemistry and Physiology, 49(2), 223–233.
Freyhof, J. (2011). Thymallus thymallus. The IUCN Red list of threatened species, 2011, e.T21875A933742.
Katoh, F., Cozzi, R. F., Marshall, W. S., & Gross, G. G. (2008). Distinct Na⁺/K⁺/2Cl⁻ cotransporter localization in kidneys and gills of two cyrptinele species, rainbow trout and killifish. Cell and Tissue Research, 334(2), 265–281.

Keyster, I. A. (2009). Morfologicheskijy sostav krovi ryapushki i evo izmeneniya kak bioindikatoryi pokazatel uslovi na obitanija v Belom ozero (Vologodskaya oblast) [Morphological structure of blood of vendace and its changes as biocindicators of Beloe Lake (Vologda Region)]. Modern Problems of Science and Education, 3, 117–125 (in Russian).
Kocabatmaz, M., & Ekingen, G. (1987). Comparative studies on leucocytes of some freshwater fish species. Selçuk Üniversitesi Veteriner Fakültesi Dergisi, 3(1), 71–81.
Langdon, J. S. (1985). Smoltification physiology in the culture of salmonids. In: Muir, J. F., & Roberts, R. J. (Eds.). Recent Advances in Aquaculture. Springer, Boston. Pp. 79–118.
Laprirova, T. B., Ferrova, E. A., Yurchenko, V. V., & Morozov, A. A. (2017). Protective systems of immune-competent organs in the fish of various ecological and systematic groups. Journal of Ichthyology, 57(3), 458–466.
Mizuno, S., Misaka, N., & Kasahara, N. (2001). Morphological changes in juxta-glomerular cells of the kidney during smoltification in masu salmon Oncorhynchus masou. Fisheries Science, 67(3), 538–540.
Morozov, A. A., & Yurchenko, V. V. (2018). Seasonal changes in hepatic antioxid-ant enzyme activities of the perch Perca fluviatilis in the Upper Volga basin, Russia. Ichthyological Research, 65(2), 265–269.
Nykänen, M., Huusko, A., & Mäki-Petäys, A. (2001). Seasonal changes in the habitat use and movements of adult European grayling in a large subarctic river. Journal of Fish Biology, 58(2), 506–519.
Ojeda, J. L., & Icardo, J. M. (1991). A scanning electron microscope study of the neck segment of the rabbit nephron. Anatomy and Embryology, 184(6), 605–610.
Ojeda, J. L., Icardo, J. M., Wong, W. P., & Ip, Y. K. (2006). Microanatomy and ultrastructure of the kidney of the African lungfish Protopterus dolloi. The Anatomical Record Part A: Discoveries in Molecular, Cellular, and Evolutionary Biology, 286A(6), 609–625.
Rondeau, E. B., Minkley, D. R., Leong, J. S., Messmer, A. M., Jantzen, J. R., von Schlabur, K. R., Lemon, C., Bird, N. H., & Koop, B. F. (2014). The genome and linkage map of the northern pike (Esox lucius). Conservation syteny revealed between the salmonid sister group and the Neoteleostei. PloS One, 9(7), e102089.
Sayed, I., Parvez, S., Pandey, S., Bin-Hafiez, B., Haque, R., & Raiasiddin, S. (2003). Oxidative stress biomarkers of exposure to deltamethrin in freshwater fish, Channa punctatus Bloch. Ecotoxicology and Environmental Safety, 56(2), 295–301.
Scott, A. (1985). Distribution, growth, and feeding of postemergent grayling Thymallus thymallus, a potential model organism for studies of coldwater biology. Journal of the Fishery Research Board of Canada, 42(5), 1435–1442.
Sinu, A. K., AbdElgawad, H., Giblen, T., Zinta, G., De Rop, M., Asard, H., Blust, R., & De Boeck, G. (2014). Anti-oxidative defences are modulated differentially in three freshwater teleosts in response to ammonia-induced oxidative stress. PLoS One, 9(4), e95319.
Talbot, C., Stagg, R. M., & Eddy, F. B. (1992). Renal, respiratory and ionic regulation in Atlantic salmon (Salmo salar L.) kelts following transfer from fresh water to seawater. Journal of Comparative Physiology B, 162(4), 358–364.
Timakova, T. K., Ferrova, E. A., & Zabotkina, E. A. (2014). Metody svetovoy i regul. Mech. Biosyst., 2019, 10(1)