Prognostic significance of IL-6 and IL-8 ascites levels in ovarian cancer patients

Denis Lane, Isabelle Matte, Claudine Rancourt and Alain Piché*

Abstract

Background: The acellular fraction of epithelial ovarian cancer (EOC) ascites promotes de novo resistance of tumor cells and thus supports the idea that tumor cells may survive in the surrounding protective microenvironment contributing to disease recurrence. Levels of the pro-inflammatory cytokines IL-6 and IL-8 are elevated in EOC ascites suggesting that they could play a role in tumor progression.

Methods: We measured IL-6 and IL-8 levels in the ascites of 39 patients with newly diagnosed EOC. Commercially available enzyme-linked immunosorbent assay (ELISA) was used to determine IL-6 and IL-8 ascites levels. Ascites cytokine levels were correlated with clinicopathological parameters and progression-free survival.

Results: Mean ascites levels for IL-6 and IL-8 were 6419 pg/ml (SEM: 1409 pg/ml) and 1408 pg/ml (SEM: 437 pg/ml) respectively. The levels of IL-6 and IL-8 in ascites were significantly lower in patients that have received prior chemotherapy before the surgery (Mann-Whitney U test, \( P = 0.037 \) for IL-6 and \( P = 0.008 \) for IL-8). Univariate analysis revealed that high IL-6 ascites levels (\( P = 0.021 \)), serum CA125 levels (\( P = 0.04 \)) and stage IV (\( P = 0.009 \)) were significantly correlated with shorter progression-free survival. Including these variables in a multivariate analysis revealed that elevated IL-6 levels (\( P = 0.033 \)) was an independent predictor of shorter progression-free survival.

Conclusion: Elevated IL-6, but not IL-8, ascites level is an independent predictor of shorter progression-free survival.

Background

The incidence of ascites in women presenting with epithelial ovarian cancer (EOC) ranges from 45% to 75% depending on the tumor type but increases in advanced stages [1]. It is a distressing complication carrying substantial morbidity [2]. Unlike most stroma surrounding solid tumors, ascites constitute a unique form of tumor microenvironment. Recent evidences suggest that ascites play an active role in tumor development. EOC ascites may affect cell behaviour such as cell growth, invasion, and de novo drug resistance of EOC cells [3-5]. We recently reported that the acellular fraction of EOC ascites inhibits drugs- and death receptor-induced apoptosis in vitro (de novo resistance) [3,4]. Newly diagnosed women with protective ascites had significantly shorter progression-free survival [6] suggesting that ascites-mediated de novo resistance impacts on EOC progression. Stromal myofibroblasts and endothelial cells, adjacent to cancer cells in solid tumors, are replaced by floating mesothelial cells and by a variety of immune cells in ascites. The acellular fraction of ascites is a complex exudative fluid known to contain growth factors [7-9], lysophosphatidic acid (LPA) [10,11], cytokines [12,13] and extracellular matrix constituents (ECM) [14]. The contribution of these molecules in ovarian cancer progression is not well defined.

Although a wide variety of cytokines can be measured in ovarian cancer ascites, interleukin-6 (IL-6) and interleukin-8 (IL-8) are among the most abundant [12]. The concentration of these pro-inflammatory cytokines in ascites is 40- to 500-fold higher as compared to the levels found in serum [12]. IL-6 can be secreted in ascites by ovarian cancer cells, tumor-associated macrophages and peritoneal mesothelial cells. However, levels of IL-6 secreted by mesothelial cells are 600-fold higher than those secreted by ovarian cancer cells [15]. The source of the IL-8 found in ascites has not been well defined. These pro-inflammatory cytokines are involved in different pathophysiological processes including carcinogenesis. In ovarian cancer, IL-6

* Correspondence: alain.piche@usherbrooke.ca
Département de Microbiologie et Infectiologie, Faculté de Médecine, Université de Sherbrooke, 3001, 12ième Avenue Nord, Sherbrooke, J1H 5N4, Canada

© 2011 Lane et al; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.
is thought to be involved in host immune responses to the disease [16-18]. IL-6 has also been demonstrated to be involved in autocrine growth of ovarian cancer cells [19-21]. IL-6 signaling in ovarian cancer cells can regulate tumor cell proliferation, invasion and angiogenesis [22-24]. IL-8 was recently reported to promote ovarian tumor growth in vivo [25]. Despite these data, the biological relevance of high levels of IL-6 and IL-8 in ovarian cancer ascites remains mostly unknown. A number of studies have reported an association between serum levels of IL-6 and prognosis, and elevated levels correlated with a poor relapse-free and overall survival [26,27]. However, others have not found such correlation between elevated serum levels of IL-6 and survival time [28].

Based on the observation that ovarian cancer ascites may affect tumor progression and reported elevated levels of IL-6 and IL-8 in ascites, we hypothesize that these cytokines might affect the clinical progression of patients with ovarian cancer. The purpose of the present study was to investigate the prognostic significance of IL-6 and IL-8 ascites levels on progression-free survival in a cohort of 39 ovarian cancer patients.

**Methods**

**Patients**

The study population consisted of 39 patients with newly diagnosed epithelial ovarian cancer admitted at the Centre Hospitalier Universitaire de Sherbrooke. Informed consent was obtained from women that underwent surgery by the gynecologic oncology service between 2000 and 2009 for this institutional review board approved protocol. Baseline characteristics and serum CA125 levels were collected for all patients. All patients had a follow up > 1 year. Disease progression was defined by either CA125 \( \geq 2 \times \text{nadir} \) value on two occasions, documentation of increase or new lesions on CT-scan or death [29]. Patient’s conditions were staged according to the criteria of the International Federation of Gynecology and Obstetrics (FIGO).

**Ascitic fluids**

Peritoneal fluids were obtained at the time of initial cytoreductive surgery for all patients. Peritoneal fluids were centrifuged at 1000 rpm for 15 min and supernatants were stored at -80°C until assayed. All fluids were supplied by the Banque de tissus et de données of the Réseau de Santé du Québec affiliated to the Canadian Tumor Repository Network (CTRNet).

**Determination of IL-6 and IL-8 concentration**

Determination of IL-6 and IL-8 concentration was performed using the commercially available Quantikine kits from R&D Systems (Minneapolis, MN). The detection thresholds were 0.79 pg/ml for IL-6 and 3.5 pg/ml for IL-8. The intra-assay variability was for IL-6 and IL-8 analysis was 5-10% and 5-20% respectively. All tests were run in duplicates according to the manufacturer’s instructions.

**Statistical analysis**

Comparison between unpaired groups was made using the Mann-Whitney test or the Kruskal-Wallis test. The Kaplan-Meier method was used for the progression-free survival and Cox regression analysis and log-rank test were used for the statistical analysis. Progression-free survival was defined as the interval between the surgery and the time of disease progression. Univariate and multivariate analyses were performed using Cox regression. The threshold for statistical significance is a probability of 0.05.

**Results**

A group of 39 patients with newly diagnosed epithelial ovarian cancer and for which ascites was available was evaluated. The mean concentration of IL-6 and IL-8 was determined by ELISA in the 39 ascites samples. The patient’s clinicopathological characteristics are summarized in Table 1 according the IL-6 and IL-8 ascites levels. Mean ascites levels for IL-6 and IL-8 were 6419 pg/ml (SEM: 1409 pg/ml) and 1408 pg/ml (SEM: 437 pg/ml) respectively. We found no significant correlation between IL-6 or IL-8 ascites levels and FIGO stage, histopathology, grade, serum CA125 levels or the presence of post-operative residual tumor (> 2 cm). However, the levels of IL-6 and IL-8 in ascites were significantly lower in patients that have received prior chemotherapy before the surgery (Mann-Whitney U test, \( P = 0.037 \) for IL-6 and \( P = 0.008 \) for IL-8). It should be noted that the number of patients with post-operative residual tumor was limited (\( n = 4 \)).

Progression-free survival analysis in the overall patient population showed a shorter progression-free survival for patients with a median IL-6 ascites levels > 2662 pg/ml as compared with those presenting with a median IL-6 levels < 2662 pg/ml (log rank test, \( P = 0.021 \)) (Figure 1). Median progression-free survival was 14 months for patients with high IL-6 levels versus 24 months for those with low IL-6. A cutoff value of 2662 pg/ml was selected according to the median ascites levels in the 39 patients. Patients with high levels of IL-6 were 2.3 times (95% CI, 1.09 - 4.84) more likely to have disease progression as compared to those with low (< 2662 pg/ml) IL-6 ascites levels. High levels of IL-8 (median ≥ 301 pg/ml) were not significantly associated with increased risk of disease progression or shorter progression-free survival (Figure 1).

Univariate analysis showed that high IL-6 ascites levels (\( P = 0.021 \)), serum CA125 levels (\( P = 0.04 \)) and stage IV (\( P = 0.009 \)) were significantly correlated with shorter progression-free survival (Table 2). Entering these
variables in a multivariate Cox model analysis revealed that median IL-6 ascites levels > 2662 pg/ml was an independent predictor variable of shorter progression-free survival ($P$ = 0.033). Although it did not quite reach statistical significance ($P$ = 0.076), entering residual tumor > 2 cm in the multivariate model still showed that high IL-6 levels was significantly correlated with shorter progression-free survival.

### Discussion

The successful treatment of EOC remains a major challenge. Most (>$85\%$) EOC patients presenting with advanced disease will relapse. Recurrence defines incurable disease in most cases. The main obstacle to effective treatment is the failure of initial therapy to eradicate a sufficient number of tumor cells to prevent disease recurrence. One emerging model for the

| Table 1 IL-6 and IL-8 ascites levels in relation to different clinicopathological parameters |
|-----------------------------------------------|-----------------------------------------------|-----------------------------------------------|
| **No of ascites tested (%)** | **IL-6 mean pg/ml (SEM)** | **IL-8 mean pg/ml (SEM)** |
| **Total** | 39 | 6419 (1409) | 1408 (437) |
| **Stage** | | | |
| I | 9 (23) | 4526 (3081) | 1447 (997) |
| II | 4 (10) | 8893 (4719) | 2738 (1786) |
| III | 15 (39) | 6550 (2521) | 1181 (685) |
| IV | 11 (28) | 6514 (1869) | 1119 (607) |
| **Histopathology** | | | |
| Serous | 26 (67) | 5209 (1484) | 801 (339) |
| Mucinous | 3 (8) | 12641 (7227) | 3375 (2596) |
| Mixed | 7 (18) | 8916 (4084) | 2991 (1500) |
| Endometrioid | 1 (2) | 7896 (0) | 4630 (0) |
| Other | 2 (5) | 1277 (583) | 1268 (590) |
| **Grading** | | | |
| 1 | 4 (10) | 3102 (1625) | 228 (82) |
| 2 | 10 (27) | 8482 (3181) | 842 (538) |
| 3 | 20 (51) | 5831 (1739) | 1755 (661) |
| **Residual tumor** | | | |
| ≤ 2 cm | 34 (87) | 6286 (1487) | 1555 (495) |
| > 2 cm | 4 (10) | 7457 (731) | 378 (153) |
| **Prior chemotherapy** | | | |
| Yes | 4 (10) | 444 (111) | 430 (123) |
| No | 34 (87) | 6984 (1453) | 1972 (681) |
| **Serum CA125** | | | |
| < 319 | 19 (49) | 4063 (1841) | 1372 (806) |
| > 319 | 19 (49) | 4406 (1765) | 1491 (788) |

* Data unavailable for grade (n = 4), residual tumor (n = 1), prior chemotherapy (n = 1), serum CA125 (n = 1)

* IL-6, $P$ = 0.0001; IL-8, $P$ = 0.002

---

**Figure 1** Kaplan-Meier analysis of progression-free survival in patients with or without elevated levels of IL-6 and IL-8 in ascites.
that received chemotherapy prior to their surgery. No significant correlation was found between IL-6 and IL-8 levels in ascites and the other clinicopathological parameters such as stage, grade, histologic type and size of residual tumor left after initial debulking. These data are consistent with those of Tempfer et al. [26] and Scambia et al. [27] which showed that elevated IL-6 concentration in serum of ovarian cancer patients correlated with poor overall survival. They are also consistent with the observation that IL-6 promoter polymorphism, which may affect IL-6 levels, impacts on survival of women with ovarian cancer [30]. However, measuring IL-6 in ascites might actually be more relevant than measuring it in serum. IL-6 is released by the peritoneal mesothelial cells and concentrates in ascites [12,15] where it could affect tumor cell behaviour. IL-6 may be cleared rapidly from the circulation and consequently serum levels may not correlate with ascites levels.

Although, our data suggest that IL-6 levels in ascites are associated with shorter progression-free survival, the precise underlying molecular mechanisms responsible for these findings remain to be established. One possibility is that elevated ascites IL-6 levels promote de novo tumor cells resistance to chemotherapy contributing to earlier disease recurrence [31]. This is supported by the observation that IL-6 inhibited death receptor-induced cell death in long term cell viability assays (Lane, unpublished data). IL-6 signaling cascade in ovarian cancer cells has been associated with the development of Taxol resistance [32,33]. Alternatively, IL-6 could stimulate the proliferation of tumor cells leading to a shorter progression-free survival [18]. IL-6-mediated angiogenesis could also play a role [23]. Tumor angiogenesis plays an important role in cancer progression and metastasis. The angiogenesis and the disruption of vascular barrier both contribute to ascites formation [34]. Because IL-6 has been shown to be involved in tumor angiogenesis in ovarian cancer, IL-6 may be important in promoting the formation of ascites as well as the progression of ovarian cancer. IL-6 signalling prevents chemotherapy-induced endothelial cells apoptosis and the blockage of its signalling cascade was therapeutically beneficial in xenograft mouse model [35]. Thus, interference with IL-6 pathway may offer opportunities for ovarian cancer therapy. However, using a monoclonal antibody that specifically blocks IL-6 signaling, siltuximab, Guo et al., demonstrated that although the combination of siltuximab with paclitaxel increased the sensitivity of ovarian tumor cells to paclitaxel in vitro, the combination was ineffective in vivo in xenograft mouse model [36]. Although clinical trials of monoclonal antibodies to IL-6 for the treatment of other types of cancer have shown encouraging results [37,38], a clear benefit for patients with ovarian cancer remains to be demonstrated. Finally, the high levels of IL-6 could enhance the immune suppressive

Table 2 Univariate analysis of progression-free survival with selected factors

| Variable | Median progression-free survival (months) | Univariate analysis (P value) |
|----------|------------------------------------------|-----------------------------|
| IL-6 (pg/ml) | | |
| < 2662 | 24 | |
| ≥ 2662 | 14 | 0.021 |
| Stade | | |
| I | 36 | |
| IV | 14 | 0.009 |
| Serum CA125 (U/ml) | | |
| < 319 | 29 | |
| > 319 | 14 | 0.040 |
| Residual tumor | | |
| ≤ 2 cm | 20 | |
| > 2 cm | 11 | 0.076 |

persistence of tumor cells after chemotherapy invokes ascites as a tumor microenvironment that promotes de novo drug resistance. Acellular fractions of EOC ascites have been shown to have anti-apoptotic activities [3,4] and ascites with pro-survival activities against cytotoxic agents were recently shown to significantly correlate with shorter progression-free survival in ovarian cancer patients [6]. These findings suggest that EOC ascites contain factors that affect progression-free survival by promoting tumor cell survival. EOC ascites contain a variety of soluble factors, including cytokines, which may contribute to de novo resistance of tumor cells. Because of the high levels of IL-6 and IL-8 found in ascites and the prosurvival activity of ascites, we hypothesized that IL6 and IL-8 could impact on pro-survival activities against cytotoxic drugs resistance of tumor cells. The identification of biomarkers that predict tumor progression would contribute to stratify patients for the selection of initial therapy. We measured IL-6 and IL-8 levels in ascites samples obtained at the time of the initial surgery in a group of 39 patients with previously untreated EOC. Mean ascites levels of IL-6 and IL-8 found in our cohort were comparable to those previously reported by Giuntoli et al. in 22 ascites samples from women with EOC [12]. Our results show that elevated IL-6, but not IL-8, levels in ascites of patients with EOC correlated with shorter progression-free survival. We found low IL-6 levels to correlate significantly with prior chemotherapy (P = 0.037). However, these data should be considered with caution given the small number of patients in the group
status of the tumor microenvironment by inducing B7-H4 expression on tumor associated macrophages and promote apoptosis in these cells [39]. IL-6 may divert the immune response from Th1 towards a suppressive Th2 response although controversial data have been reported [41,40,41]. IL-6 may also contribute to reprogram the tumor adjacent stromal microenvironment which may facilitate dissemination to the peritoneal cavity. The contribution of this reactive stromal is being recognized as an important element for tumor progression [42].

Conclusions

We found that high levels of IL-6 in ascites of newly diagnosed women with ovarian cancer significantly correlated with shorter progression-free survival. In contrast, ascites levels of IL-8 did not show such association. These data suggest that IL-6 could be an important component that contributes to ascites-mediated de novo drug resistance.

Acknowledgements

This work was supported by a grant from the Cancer Research Society (A.P.). We wish to thank the Banque de tissus et de données du Réseau de Recherche sur le Cancer des Fonds de Recherche en Santé du Québec (FRSQ), affiliated to the Canadian Tumor Repository Network (CTRNnet) for providing the ascites samples. We also thank Nathalie Carrier for her assistance with the statistical analyses.

Authors’ contributions

DL participated in the design of the study and performed the assays for measuring IL-6 and IL-8 levels in ascites. IM was responsible for obtaining the ascites and the clinical data. CR participated in the design of the study and helped to draft the manuscript. AP conceived the study, participated in its design and drafted the manuscript. All authors read and approved the final manuscript.

Conflict of interest statement

The authors declare that they have no competing interests.

Received: 17 March 2011 Accepted: 30 May 2011
Published: 30 May 2011

References

1. Partridge EE, Barnes MN: Epithelial ovarian cancer: prevention, diagnosis, and treatment. CA Cancer J Clin 1996, 46:273-302.
2. Shen-Gunther J, Mannel RS: Ovarian cancer: an overview. Gynecol Oncol 2002, 87:77-83.
3. Lane D, Robert V, Grondin R, Rancourt C, Piché A: Ascites as a predictor of ovarian malignancy. J Clin Invest 1990, 86:851-855.
4. Richardson M, Gunawan J, Hatton MW, Seilidzt E, Hartie HW, Singh G: Malignant ascites fluid (MAF), including ovarian cancer-associated MAF, contains angiotatin and other factor(s) which inhibit angiogenesis. Gynecol Oncol 2002, 86:279-287.
5. Xu Y, Gaudette DC, Boynton JD, Frankel A, Fang YJ, Sharma A, Huret J, Casey G, Goodbody A, Mellors A, Holub BJ, Mills GB: Characterization of an ovarian cancer activating factor in ascites from ovarian cancer patients. Clin Cancer Res 1995, 1:1223-1232.
6. Yamada T, Sato K, Komachi M, Maichinkhuu E, Tobo M, Kimura T, Kuvabata A, Yanagita Y, Ikeda T, Tanahashi Y, Ogawa T, Chwada S, Morishita Y, Ohta H, Ina DS, Tamoto K, Tomura H, Okajima F: Lysophosphatidic acid (LPA) in malignant ascites stimulates motility of human pancreatic cancer cells through LPA1. J Biol Chem 2004, 279:6595-6605.
7. Guentori RL, Webb TJ, Zoso A, Rogers O, Diaz-Montes TP, Bristow RE, Oeleke M: Ovarian cancer-associated ascites demonstrates altered immune environment: implications for antitumor immunity. Anticancer Res 2009, 29:2875-2884.
8. Radje K, Schmidt D, Bohme M, Schmidt U, Weise W, Morenz J: Cytokine level in malignant ascites and peripheral blood of patients with advanced ovarian carcinoma. Geburtshilfe Frauenheilkd 1996, 56:83-87.
9. Ahmed N, Riley C, Oliva K, Rice G, Quinn M: Asbestos induces modulation of α6β3 integrin and urokinase plasminogen activator receptor expression and associated functions in ovarian cancer. J Cancer Res 2009, 2:1475-1485.
10. Offner FA, Obrist P, Stadlmann S, Feichtinger H, Klinger P, Herold M, Zwierzina H, Hittmar A, Miederstein B, Zeimer A, Marth C: B-cadherin and sprouty secreted by human peritoneal mesothelial and ovarian cancer cells. Cytokine 1995, 7:542-547.
11. Asschart JG, Vellenga E, Ruiters MH, de Vries EG: Regulation of spontaneous and TNFα/IFN-induced IL-6 expression in two human ovarian-carcinoma cell lines. Int J Cancer 1999, 82:244-249.
12. Asschart JG, de Vries EG, De Jong S, Wittkopf S, Vellenga E: Differential regulation of IL-6 promoter activity in a human ovarian-tumor cell line transfected with various p53 mutants: involvement of AP-1. Int J Cancer 1999, 86:236-242.
13. Watson JM, Sens interfals JR, Berek JS, Martinez-Maza O: Constitutive production of interleukin 6 by ovarian cancer cell lines and by primary ovarian tumor cultures. Cancer Res 1990, 50:6595-6965.
14. Wu S, Rodaubaugh R, Martinez-Maza O, Watson JM, Silverstein DS, Boyer CM, Peters JP, Weinberg JB, Berek JS, Bast RC: Stimulation of ovarian tumor cell proliferation with monocyte products inducing interleukin 1, interleukin 6 and tumor necrosis factor-alpha. Am J Obstet Gynecol 1992, 166:977-1007.
15. Hsu SM, Hsu PL: Autocrine and paracrine functions of cytokines in malignant lymphomas. Biomed Pharmacother 1994, 48:433-444.
16. Nakazaki H: Preoperative and postoperative cytokines in patients with cancer. Cancer 1992, 70:709-713.
17. Syed Y, Ulinski G, Mok SC, Ho SM: Reproductive hormone-induced, STAT3-mediated interleukin 6 action in normal and malignant human ovarian surface epithelial cells. J Natl Cancer Inst 2002, 94:617-629.
18. Obata H, Tarnowski K, Shibata K, Kikawa F, Tomoda Y, Sood AK: Effects of interleukin-6 on in vitro cell attachment, migration and invasion of human ovarian carcinoma. Anticancer Res 2010, 30:279-287.
Role of vascular endothelial growth factor
in human ovarian carcinoma.

B7-H4 expression identifies a novel suppressive macrophage population
in ovarian cancer.

Pre-publication history
The pre-publication history for this paper can be accessed here:
http://www.biomedcentral.com/1471-2407/11/210/prepub

Submit your next manuscript to BioMed Central
and take full advantage of:

• Convenient online submission
• Thorough peer review
• No space constraints or color figure charges
• Immediate publication on acceptance
• Inclusion in PubMed, CAS, Scopus and Google Scholar
• Research which is freely available for redistribution

Cite this article as: Lane et al. Prognostic significance of IL-6 and IL-8 ascs levels in ovarian cancer patients. BMC Cancer 2011 11:210.