Review

Nanoparticles in Clinical Translation for Cancer Therapy

Deepa Mundekkad 1 and William C. Cho 2,*

1 Centre for NanoBioTechnology (CNBT), Vellore Institute of Technology, Vellore 632014, Tamil Nadu, India; deepamundekkad@gmail.com
2 Department of Clinical Oncology, Queen Elizabeth Hospital, Kowloon, Hong Kong SAR, China
* Correspondence: williamcscho@gmail.com or chocs@ha.org.hk

Abstract: The advent of cancer therapeutics brought a paradigm shift from conventional therapy to precision medicine. The new therapeutic modalities accomplished through the properties of nanomaterials have extended their scope in cancer therapy beyond conventional drug delivery. Nanoparticles can be channeled in cancer therapy to encapsulate active pharmaceutical ingredients and deliver them to the tumor site in a more efficient manner. This review enumerates various types of nanoparticles that have entered clinical trials for cancer treatment. The obstacles in the journey of nanodrug from clinic to market are also reviewed. Furthermore, the latest developments in using nanoparticles in cancer therapy are also highlighted.

Keywords: nanoparticles; cancer therapy; nanodrugs; nanochemotherapy; nanomedicine

1. Introduction

Cancer nanomedicine is a fast-advancing field which employed nanoparticles to diagnose and treat cancer [1]. The nanoparticles are capable of delivering the normally insoluble drugs to local and distant tumor sites in a better way, thus reducing the systemic side effects that are generally associated with conventional drug therapies. These nanodrugs are invariably biocompatible, non-immunogenic, non-toxic, and biodegradable, which in turn reduces the risk of unpredicted loss of function or adverse effects encountered in the traditional therapy [2]. The flexibility of nanoparticles in terms of size, shape, selective binding capacity, high permeability and retention effect, surface modification, etc. placed them in a good position in cancer therapy, especially in ovarian, breast, and non-small cell lung cancers [3]. State-of-the-art designs and approaches involving interaction between nanodrugs and the receptors on immune cells (like antigen-presenting cells) are further exploited in cancer therapy for sustained anti-tumor effect [4]. Some pharmaceutical industries underwent a quintessential change with the advent of nanodrugs in cancer therapy.

A more realistic approach to cancer therapy was achieved when the flexibility of nanoparticles in terms of shape, charge, stability and selective binding capacity, inspired the designing of new drugs for therapeutic purposes. Rod-shaped nanoparticles are favored to other shapes for endosomal uptake; positively charged nanoparticles evoke a greater immune response when compared to negatively charged or neutral nanoparticles [5]. Similarly, when the delivery performance of nanocarriers of various shapes and sizes were compared, the short, tubular, non-rigid nanocarrier was found to have better intratumoral invasion capacity [6]. Targeted drug delivery was improved substantially due to the selective binding of nanoligands carrying the drug to the receptors found on the surface of tumor cells. Coupling the nanoparticles with drug-carrying polymers (like poly lactic-co-glycolic acid - PLGA) enhanced the absorption of the drug and induced more toxicity to the tumor cells [7].

Citation: Mundekkad, D.; Cho, W.C. Nanoparticles in Clinical Translation for Cancer Therapy. Int. J. Mol. Sci. 2022, 23, 1685. https://doi.org/10.3390/ijms23031685

Academic Editor: Ylenia Zambito
Received: 29 December 2021
Accepted: 18 January 2022
Published: 1 February 2022
Publisher’s Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).
Chemoresistance exhibited by cancer cells is one of the reasons why traditional therapy is not effective in combating cancer. Genomic instability within the tumor may contribute to heterogeneity among cancers, which in turn, drives chemoresistance [8]. It is a challenge to deliver the drugs to those sites of temporal or spatial heterogeneity in the wake of chemoresistance. Non-genetic variants, like the mechanisms that the cells adopt to dodge the immune system, are also responsible for resistance to therapy. A combination of different treatment modalities has been successfully employed to overcome therapeutic resistance in cancer cells. Immunotherapy drugs are effective to an extent in combating chemoresistance in cancer as they target the biomarkers for their line of attack. Certain cancer patients are benefited from different modes of cancer immunotherapy[9], cancer vaccines [10], checkpoint inhibitors [11], chimeric antigen receptor (CAR) T-cell therapy [12], cytokines [13], and oncolytic viruses [14].

However, the success of cancer therapy mainly depends on how the drug acts on the immune system and promptly activates one or the other component of the immune system to bring about the eradication of cancer cells. Nanoparticles can be fabricated to influence immunomodulation by interacting with the components of immune system [15,16] and thus bring about killing of cancer cells. Nanoparticles have made an amazing impact in nanochemotherapy as is evident from the huge number of nanoparticle-based drugs that have entered clinical trials for cancer therapy. Here, the review centers around the ways that the nanoparticles made a front-line entry into cancer therapeutics over the recent years.

2. Nanoparticles in the Treatment of Cancer

There is relentless work going on the world over towards achieving an effective alternative to chemotherapy and a better cure for cancer. Though efforts to treat cancer and improve the efficacy of drugs through nanotechnology are at the research or development stage, nanoparticles have been extensively used in biomedical applications. Due to their nano-size [17] and very special properties (like mass density [18], surface charge [19], etc.) nanoparticles have the advantage of relatively large surface area which facilitates their functionalization with various ligands like DNA [20], RNA [21], peptides [22], aptamers [23], antibodies [24], etc. This will assist in the direct delivery of the modified nanoparticle to the site of action (in vivo). This property greatly influences the development of a wider repertoire of ingredients with active theranostic properties to be incorporated with nanoparticles thus enhancing the pharmacokinetic property (and in turn, the efficacy) of the nanoparticle for cancer treatment [25]. The role of nanoparticles as an immunogenic cargo is also being investigated in traditional radio- and chemo-therapies as well as the advanced adjuvant therapy [26]. The biocompatibility of nanoparticles promoted the development of innovative nanostructures whereby they are now being engaged in totally unconventional roles (as artificial antigen-presenting cells (aAPCs) or as in vivo repository of immunostimulatory molecules) for sustained antitumor activity [27]. aAPCs represent a novel technology in cancer immunotherapy where a nanoparticle-based system mimics the antigen-presenting cell by activating crucial signal proteins against cancer [28], endowing the nano-aAPCs the capacity to be one among the next generation cancer nanomedicine [1].

3. Mechanism of Action of Nanoparticles

Generally, nanoparticles induce apoptosis in cancer cells by a series of mechanisms; reactive oxygen species (ROS)-mediated apoptosis being the most studied among them. Up- and down-regulation of proteins, immunological interventions, inhibition of transcription, site-specific cytotoxicity, etc. are other mechanisms of action of nanoparticles in apoptosis induction in cancer cells. It is imperative to mention at this point that there is a series of cross-talk between many of these events but the final effect is apoptotic cell death (Figure 1).
ROS induced-apoptosis is one of the basic mechanisms of action that has been studied as part of nanoparticle-induced cytotoxicity. Generally, ROS behaves like a double-edged sword bringing about anti-apoptotic and pro-apoptotic effects. The pro-apoptotic effect of ROS sums up to cell cycle arrest, apoptosis, and necrosis whereas, the anti-apoptotic effect transpires to the promotion of cell proliferation, invasion, and metastasis all the while inhibiting apoptosis [29]. Proteins are capable of inducing ROS-mediated apoptosis. Fragile histidine triad (Fhit) proteins are lost in most cancer types but when restored, they can induce apoptosis. Fhit interacts with ferrodoxin reductase to trigger the generation of ROS in Fhit-deficient cancer cells following peroxide treatment. Oxidative stress is induced, and this resulted in ROS-induced apoptosis in lung cancer cells. It was observed that Fhit negative cells escaped ROS overproduction and, likely oxidative damage [30]. Nanoparticles are more effective in inducing apoptosis mainly since nano-sized particles offer more reactivity due to increased surface area resulting in the excessive formation of ROS. The apoptotic effects are prominent when oxidative stress increases for the cells and releases inflammatory intermediates followed by DNA and protein damage resulting in cytotoxicity [31]. Silver nanoparticles enclosed in polysaccharides were found to generate significant amounts of ROS which, in turn, educate cell death mainly through autophagy.

**Figure 1.** Mechanisms of cell death in cancer induced by nanoparticles.

3.1. Generation of ROS

ROS induced-apoptosis is one of the basic mechanisms of action that has been studied as part of nanoparticle-induced cytotoxicity. Generally, ROS behaves like a double-edged sword bringing about anti-apoptotic and pro-apoptotic effects. The pro-apoptotic effect of ROS sums up to cell cycle arrest, apoptosis, and necrosis whereas, the anti-apoptotic effect transpires to the promotion of cell proliferation, invasion, and metastasis all the while inhibiting apoptosis [29]. Proteins are capable of inducing ROS-mediated apoptosis. Fragile histidine triad (Fhit) proteins are lost in most cancer types but when restored, they can induce apoptosis. Fhit interacts with ferrodoxin reductase to trigger the generation of ROS in Fhit-deficient cancer cells following peroxide treatment. Oxidative stress is induced, and this resulted in ROS-induced apoptosis in lung cancer cells. It was observed that Fhit negative cells escaped ROS overproduction and, likely oxidative damage [30]. Nanoparticles are more effective in inducing apoptosis mainly since nano-sized particles offer more reactivity due to increased surface area resulting in the excessive formation of ROS. The apoptotic effects are prominent when oxidative stress increases for the cells and releases inflammatory intermediates followed by DNA and protein damage resulting in cytotoxicity [31]. Silver nanoparticles enclosed in polysaccharides were found to generate significant amounts of ROS which, in turn, educated cell death mainly through autophagy.
and extend apoptosis [32]. Nanoparticles targeted to mitochondria through pyruvate produce ROS resulting in the inhibition of ATP synthesis. Silica carbon nanoparticles coated with lipid membrane were found to inhibit the growth of multidrug-resistant tumors with no evidence of systemic toxicity through inhibition of ATP synthesis [33].

3.2. Regulation of Proteins

Up- and down-regulation of proteins represent a situation where the cells mimic a cellular response to any endogenous or exogenous stimuli under normal or oxidative stress conditions. Under stress, more proteins are regulated to remodel the metabolic and signaling pathways, leading to alterations in the redox proteome and membrane turnover, impacting cycle progression and proliferation, resulting in apoptosis and tumor suppression [34]. There is increasing evidence for the potential of nanoparticle-mediated regulation of proteins that are involved in signaling pathways specifically associated with the pathogenesis, progression, and oncogenic behavior of cancer cells. Copper oxide nanoparticles (CuO-NPs) are capable of the down-regulation of the apoptotic regulatory proteins (Bcl2 and BclxL) inducing programmed cell death in HT-29 under study [35].

Selenium nanoparticles (SeNP) were found to have different effects on the expression profile of apoptotic proteins at concentrations as low as 5 µg/mL. The unfolded protein response (UPR) signalling pathways were also affected by the SeNP. Treating cell lines with SeNP showed a significant increase (3–4 fold) in the expression pattern of ER-resident selenoproteins and selenium-containing glutathione peroxidases and thioredoxin reductases [36]. Pro-apoptotic proteins were also selectively regulated by SeNP by the activation of Cx43 hemichannels [37]. Likewise, enhanced expression of γ-H2AX was detected in MCF-7 cell lines that positively took up silver nanoparticles (AgNPs), followed by the release of Ag ions within the cells and subsequent cell death [38]. Selective regulation of cyclin dependent kinase 4 (CDK4) by gold nanoparticle conjugates induced the G1 cell cycle arrest and apoptosis induction in the ER-positive human breast cancer cells lines, MCF-7 [39]. The inactivation of CDK4 was followed by a failure in the translocation of nuclear factor kappa B (NF-κB) to the nucleus resulting in cell death. Gold nanoparticles could potentially down-regulate the phosphorylation of key players (like p42/44 and p38 in MAPK signaling pathway), thus inhibiting migration and colony-forming ability of pancreatic cancer cells and in turn, reverse the chemo-resistance in cancer cells [40].

3.3. Radiation Therapy

Traditional radiation therapy fails to achieve the desired effect in cancer therapy as indiscriminative radiation renders damage to normal, healthy cells too. Also, opsonization leads to the rapid clearance of radioisotopes from the blood resulting in reduced therapeutic effect. Employing nanoparticles with a radioactive core will reduce opsonization and also help in extending the retention of the therapeutic nanoparticle through enhanced permeability and retention (EPR) effect [41]. The use of nanoparticles (as radiosensitizer) for systemic targeting of tumor cells in radiation therapy will enhance the effect of X-ray radiation therapy against cancer cell lines. The biological hazard to other cells due to radiation can be controlled by enclosing the radioactive molecule in a nano-crystalline matrix, bound by strong crystalline bonds. This relatively new approach engages nanoparticles as sensitizers whereby, the nanoparticle with a radioactive core is attached to the molecule and can be precisely directed to the cancer cells. Upconversion nanoparticles (UCNP) coupled to beta-emitting radionuclide yttrium-90 (90Y) and a fragment of the exotoxin A that was genetically fused to a protein specific to HER2 receptor, enabled background-free imaging in in vitro and in vivo cell models [42]. The localization of radiation impacted only those cells within millimeters leading to the reduction of adverse side effects also. Radioactive palladium gold nanoparticles were efficiently retained in prostate cancer tumors for several weeks to deliver a dose-dependent tumor growth inhibition. The interaction of energetic photons with gold atoms produced photoelectrons, Auger
electrons, X-rays and delta rays. These electrons damage the tumor and blood vessel endothelial cells causing DNA strand breaks resulting in cancer cell death [43]. Bismuth lipophilic nanoparticles effectively enhanced the X-ray radiation therapy against breast cancer cells [44], thus effectively inducing a concentration-dependent cell growth inhibition in breast cancer cell lines by possibly altering the membrane permeability and damaging the genomic DNA [45]. Sometimes, bismuth (Bi) nanoparticles are favored to gold (Au) or platinum (Pt), as they are readily oxidized and dissolved at physiological conditions. Also, the high atomic number and biocompatibility favored the use of bismuth in radiation therapy. They are released from the body as soluble ions and so, bioaccumulation is minimized. XCA (X-ray contrast agent) containing bismuth (like Bi$_2$S$_3$) nanoparticle coated with PVP (polyvinyl pyrrolidone) increased the computed tomography (CT) brightness, even at lower concentrations, as compared to the iodine-based clinical contrast agent, iopromide. The blood circulation half-life of PVP-coated Bi$_2$S$_3$ nanoparticles was significantly larger than the iodinated counterparts [46].

3.4. Phototherapy

The heat generated from light or electromagnetic radiation is used to elicit responses in cancer cells that induced apoptotic cell death as in phototherapy (Figure 2). Also, in the presence of thermal stress, the radio-sensitized tumors are likely to respond even more to radiotherapy, resulting in improved cancer survival rates [47]. Nanoparticles act as the photosensitizer whereby, due to their higher optical absorbance in the near-infrared region, the light energy from the source is transmitted to the tumor site as thermal energy, resulting in the release of reactive oxygen species, immunogenic molecules, or antigens that are detected by the cell’s immune system and process the tumor cell for destruction by apoptosis [48]. Among various nanoparticles that are employed to deliver heat to the tumor cells, the iron oxide nanoparticle magnetite (Fe$_3$O$_4$), is capable of emitting thermal energy when exposed to a rapidly alternating magnetic field. The heat thus generated has a more discriminatory effect on fast-dividing cancer cells than normal tissues [49,50]. A single 10-min exposure of the oral cancer cell lines (VB6) to magnetic nanoparticles conjugated to anti-$\alpha$-$\beta_6$ antibody in an alternating magnetic field resulted in the death of 85% of cell lines. Only a 20% reduction in cell viability was observed for the $\alpha$-$\beta_6$-negative cells exposed to the same conditions [51]. Gold nanoparticles (AuNPs) have very strong absorption around infra-red regions of the electromagnetic spectrum and can generate localized heat to destroy the region of interest and can thus be successfully employed for in vivo phototherapy [52]. An Au conjugate (Au@ZIF-8 based on nano metal-organic frameworks) acted as catalase mimicking nanozyme when Ce6 was encapsulated as a photosensitizer. It was observed that upon arrival of Au@ZIF-8 at the tumor environment, the AuNP catalyzed the excessive H$_2$O$_2$ to mitigate hypoxia, resulting in toxicity in the tumor [53].
Phototherapy is used to induce apoptosis in cancer cells by employing light, heat, and radiation. Photosensitizers like nanoparticles enhance the killing effect by targeting signaling pathways and the immune system.

3.5. Triggering Immunological Reactions

Immunological reactions are often the most fundamental reactions that expose the tumor cells for eradication via the host immune system by tumor antigens [54]. Enhancing the innate immunity with biocompatible nanoparticles is a strategy successfully adopted to control cancer cell growth. However, cross-talk between the various processes (like ROS release and the subsequent complement activation) is evident as many key mechanisms in innate immunity are inter-connected. The release of cytokines when THP-1, NCI-H460, and HL-60 cell lines were treated with gold and silver nanoparticles, clearly implicated the involvement of humoral and cellular immunity upon exposure of the cells to the nanoparticles. This, in addition, provides a fine case for use of nanoparticles as an adjuvant in vaccine development as evidenced by the balanced Th1 and Th2 immune response mediated by ROS production, cytokine release, and complement activation [27]. Another strategy is to modulate the immunosuppressive microenvironment of the tumor and thereby kill the cancer cells by employing nanoparticle-encapsulated dual therapeutic molecules (like nucleic acid). Here, the dual molecule can be effectively engaged for the delivery and repeated administration of the high-dose nanoparticle-encapsulated nucleic acid, resulting in systemic immune activation followed by elevated levels of pro-inflammatory molecules [26]. The nanoparticle-encapsulated dual therapeutic 5′ triphosphate dsRNA (ppp dsRNA) induced higher levels of Th1 cytokines, CD8+ T cells, M1 macrophages, etc. reflecting significant tumor growth inhibition. Furthermore, the physicochemical and
colloidal nature of nanoparticles enables the production or inhibition of antibodies, which endorse their extensive use in the diagnosis and treatment of melanoma [55].

3.6. Site-Specific Cytotoxicity

The customized drug release profile is one of the rationales behind the use of nanoparticles loaded with the drug conjugate to be targeted for site-specific release. Intracellular delivery of DNA, mRNA, small interfering RNA (siRNA), and protein can be facilitated by nanoparticles where the delivery of nano-conjugated biomolecules offers site-specific cytotoxicity with reduced side effects that are generally caused by systemic drug formulations. This enhanced therapeutic efficiency comes with fewer adverse effects, improved pharmacokinetics, higher EPR effect [56]. Moreover, the excellent binding efficiency of the target ligands on the nanoparticle surface leads to increased tumor selection and drug disposition in cancer cells. The negatively charged hyaluronic acid-based nanoparticles were found to have site-specific cytotoxicity towards CD44-positive tumor cells [57]. Also, the therapeutic efficacy can be monitored in real-time as is revealed by the targeted delivery of chemotherapeutic drugs by the multifunctional titanium phosphate nanoparticle. These were designed for cellular uptake and cytotoxicity as the nanoparticles exhibited high drug loading capacity and enhanced cell uptake mediated by folate-receptor [58]. In another typical case, zinc oxide (ZnO) as such was found to have inherent cytotoxicity towards cancer cells by inducing apoptosis. The high expression of membrane anionic phospholipids on the surface of cancer cells renders them the capacity to selectively intake ZnO nanoparticles. This leads to an imbalance in Zn ions resulting in the induction of ROS. Conjugation of ZnO with other metal oxide nanoparticles (like Fe3O4) is found to increase the cytotoxic potential because of the combined effect of ZnO selectivity and Fe3O4 magnetism [59]. The cationic solid lipid nanoparticle (SLN) nano-conjugate was able to tightly bind streptavidin and biotinylated antibody against HER2 receptor in breast cancer cell lines. These anti-HER2 compact antibodies acted as highly effective carriers and effected specific cell cytotoxicity in BT-474 and MCF-7 cell lines that had overexpressed HER2 [60].

3.7. Gene Therapy for Cancer Cell Growth Inhibition

Regulation of genes that are actively involved in various cellular processes in tumor cells by nanoparticles is another strategy to kill cancer cells. Genes like STAT3, FGFR1, HNRNPL, BCL2L1, ATF3, RAB5C, ANG, EIF3I, NKIRAS2, PIAS4, RRAS, NUCKS1, AKT1, SRC, PCB2, EIF2C2, HRAS, CDC34, NFKB2, EIF4G1, and EIF5A are controlled by Fe3O4 nano-powder in A549 cell lines thus inducing antitumor effect [61]. Many of these genes are overly expressed in cancer cells and are rightly identified as molecular markers. The fact that Fe3O4 nanoparticles can induce an anticancer effect on cancer-specific molecular markers can have greater implications in gene therapy.

Many more nanoparticles were successfully employed to demonstrate these major mechanisms to control cancer cell growth. Table 1 list some of the nanoparticles employed in cancer cell studies and the mechanism of action.

Table 1. The variety of nanoparticles employed in cancer cell death and their mechanism of action.

| Type of Nanoparticles/Nano-Conjugates | Cell Lines | Mechanism of Action | Reference |
|--------------------------------------|------------|---------------------|-----------|
| DNA-modified magnetic NPs            | MCF-7      | Suppression of RNA marker | [25]      |
| Au, Ag NPs                           | Human peripheral blood mononuclear cells (hPBMCs) | Compliment activation, cytokine production | [27]      |
| Gold NP-tagged toxin                 | MCF-7      | Down-regulation of CDK-4 and MAPK | [39]      |
| Au@ZIF-8 NPs                         | EMT-6 murine breast cancer cell | ROS generation | [53]      |
### 4. Nanoparticles in Clinical Translation

A google scholar search with the keywords ‘nanoformulations, cancer cure’ returned close to 3,000 hits within 0.08 s; 18,000+ publications are based on nanomedicines. But it is upsetting from a researcher’s point of view to note that much of this basic research failed to get translated into clinical realities. 25 years after the first nanochemodrug Doxil was introduced to the market in 1995, the number of nanomedicines approved for cancer cure is still trifling. Despite the huge number of research attempts to use nanoparticles for cancer therapeutic applications, only very few formulations have crossed the clinical phase over the period. A fleeting look into the NP-based formulations currently on the market and in clinical trials will reveal that there are not many nano-formulations that have got the approval to hit the market as cancer therapeutics [83]. 16 nano-based cancer drugs are approved by FDA whereas close to 75 nanoformulations are in clinical trials now [84]. There are five major cancer nanomedicines available for cancer cure (Figure 3).

| Nanoparticle | Cell Line | Effect |
|--------------|-----------|--------|
| Fe3O4@AuNC@erlotinib | PANC-1 | Selective targeting of overexpressed EGFR [62] |
| GOx and PDA functionalized iron oxide NPs | MDA-MB-231, MCF-10A and 4T1 | Photothermal therapy and ROS-mediated damage [63] |
| V2O5 | B16F10, A549, and PANC1 | ROS-induced apoptosis [64] |
| Fe3O4 | HepG2 | ATP-citrate lyase-dependent RAS signaling [65] |
| Fe@Fe3O4@heparin | 4T1 breast tumor cell line, HUVEC cell | ROS generation [66] |
| PEGylated rhodium nanodots | CT-26 colon tumor | Down-regulation of TNF-α and IL-6 [67] |
| Au NPs | B16 melanoma cell | Up-regulation of Caspase 3, Caspase 9, Bid, Bax and down-regulation of BCl2 [68] |
| Au NPs-PEG-RNase A conjugate | SW-480 | ROS generation [69] |
| RBC membrane-coated PLGA NPs | Pancreatic ductal adenocarcinoma | Tumor microenvironment modulation [71] |
| PEGylated ZnO NPs | PANC1 | ROS-induced apoptosis [72] |
| ZnO NPs | Human acute monocyctic leukemia cell line (THP-1) | Mitochondrial membrane damage and elevated ROS concentration [73] |
| Ag NPs | HeLa | SubG1 arrest and apoptotic/necrotic cell death [74] |
| Pt NPs | A549 | Induction of apoptosis and cell cycle arrest [75] |
| TiO2 NPs | LL2 mouse lung cancer cell line | Oxidative stress and cytokine induction [76] |
| MoS2 nanoflakes | MDA-MB-231 | Selective ROS generation and photothermal therapy [77] |
| Pt NPs | Human foreskin fibroblast cell | Damage to DNA and inhibition of DNA replication [78] |
| CeO2 NPs | Mouse fibro-sarcoma cell line | ROS-induced apoptosis [79] |
| CeO2 NPs | A549 | ROS-mediated apoptosis [80] |
| ZnO NPs | MCF-7 | Up-regulation of caspase-8 and p53 [81] |
| TiO2 NPs | HepG2, A549, MCF-7 and IMR-90 | Oxidative stress [82] |
Figure 3. Different types of nanomedicines that are used for the treatment of cancer.

4.1. Liposomal Nanoparticles

Most of the nanoparticle-based cancer drugs are formulated with liposomes (Figure 4) as the liposome-based nanodrugs have greater adaptability and feasibility in addition to biocompatibility and biodegradability [85]. The exceptional ability of liposomes to entrap both hydrophilic and hydrophobic compounds along with their ability to be functionalized with a variety of molecules like PEG, antibodies, aptamers, proteins and peptides, carbohydrates, or other small molecules in targeted liposomes [86] may be the prime reason why the majority of approved drugs and those in clinical trials belong to this group. During drug delivery, the bilayered phospholipid membrane of liposomes protects the drugs embedded in them from proteasomal degradation and biological inactivation; they are resistant to chemical and immunological changes as well [87]. The major advantage here is that the non-targeted, healthy cells in the vicinity of the drug target are unaffected by the drug carried by the liposomes as they are safely entrapped in the liposomal core.
Figure 4. (A) Various forms of liposome-based nanomedicines. The variety of formulations under the category specifies the flexibility of the liposomes for the formulation of nanodrugs. The number in parenthesis refers to the number of drugs that are approved and under trial respectively, under each category. (B) Major FDA approved nanochemotherapeutic drugs based on liposomes. (C) Some of the liposome-based nanochemotherapeutic drugs that are under clinical trial.

4.2. Metal and Metal Oxide Nanoparticles, Polymeric Micelles, Polymer/Lipids, and Other Conjugates

The rest of the drugs are based on metal and metal oxides, polymeric micelles, polymer/lipids, and other conjugates (Figure 5) where the drugs are in various stages of approval or trials. Platinum is one of the most exploited metals for the delivery of anticancer drugs; cisplatin is the first to be used as an anticancer drug followed by a variety of other
compounds. Platinol was approved by FDA for combination therapy for a variety of cancers in addition to picoplatin, carboplatin, sebriplatin, ormaplatin, oxaliplatin, aroplatin, enloplatin, satraplatin, zeniplatin, miboplatin, satraplatin, and iproplatin [88]. However, the dominance of platinum-based drugs was diminished during later years as a spectrum of challenges emerged from the use of platinum and platinum-based drugs in clinical use. This prompted the emergence of other nanocomplexes with cytotoxicity in cancer therapy (like ruthenium [89], gold [90], silver [91], selenium [92], and iron [93]). More and more metal complexes are being studied for antitumor effects based on the success of metal-based nanodrugs.

Vesicles formed from self-assembled amphiphilic micelles in the nano range can extravasate through endothelial cell junctions of cancer cells and thus can erupt near the tumor microenvironment to release the drug. These polymeric micelles are better in drug delivery as the internalization of the drug carrier to the tumor site is more effective than liposomes and other lipid nanoparticles [94]. Drug solubility is improved while core-shell structured polymeric micelles are used as drug carriers; the water-insoluble drugs are entrapped in the micelle core by physical or chemical conjugation promoting the availability of the drug to the site of action [95].

Exploiting the improvements in polymeric systems based on lipids, a new nanodrug system was developed in recent years. The lipid-polymer nanoparticles comprise a hybrid structure comprised of a polymeric core surrounded by a lipid shell [96]. They are postulated to incorporate the benefits of lipid-based nanocarriers including the easiness in drug release along with additional features of surface modification and functionalization.
In a nanoconjugate drug system, the various components are premeditated and designed using nanosheets as a delivery platform attached to a receptor targeting polymer (like folate receptor) [97]. New cytotoxic agents are designed to maximize the anticancer effect by targeting the tumor. The fatty acid Docosahexaenoic acid conjugated with the antineoplastic drug paclitaxel, (DHA–paclitaxel) is one such novel semisynthetic taxane conjugate with no cytotoxic activity until it is metabolized to paclitaxel [98]. The tested doses of the taxane-conjugated drug were shown to have a greater incidence of myelosuppression than conventional taxane in patients with advanced non-small cell lung carcinoma (NSCLC) implying significant clinical application.

5. Obstacles in the Clinical Translation of Nanoparticles

In spite of all these developments, there are major obstacles in the journey of nanodrug from clinic to market. The lack of proper understanding of the mechanism of action of the nanoparticle with the biomolecules is the foremost deterrent factor. In many cases, continuous production of nanoparticles with sustained effect on cancer cells becomes a liability to the pharmaceutical companies. There are many examples where pharmaceutical companies have discontinued drugs (e.g., DepoCyt). These companies cite technical issues as reasons for the impediment. Researchers and pharmaceutical companies have to confront a multitude of issues before the drug is promoted as suitable for cancer cure. Due to this, many of the nanoformulations were retracted from the market even after getting FDA approval; Clariscan (PEG-Fero, Feruglose NC100150); Feridex I.V. (Endorem, Ferumoxides); Lumirem (Gastromark); Resovist (Cliaivist); Sinerem (Combidx) and GastroMARK are examples [99,100].

There are some reasons why these research findings are not decoded and marketed. Issues that can cause probable hitches in the process are varied. Some of the issues are discussed here.

5.1. The Difficulty in Predicting the Predisposal of the Patient to Allergic Reactions

It is very difficult to predict a definite pattern or behavior even in patients who are administered the same dose of medicine. Though the relevant mechanisms of action of the drug during the clinical study in a varied population of patients showed highly similar outcomes, they may have unexpected and contradictory side effects when it reaches the mass population of patients worldwide. Likewise, a simple assessment of side effects does not guarantee the safety of the drug [101]. This in itself is a complex scenario as there is an infinite potential for the drug to combine with different entities that it confronts in the body and a methodical examination and assessment of this interaction at a molecular level are important.

5.2. Endotoxin Quantification

A common reason for the early failure of clinical translation of nanochemotherapeutics is endotoxin contamination [102]. The common assay for endotoxin quantification is the limulus amebocyte lysate (LAL) assay where the endotoxin’s activity is measured [103]. However, this assay fails to quantify endotoxin as the activity and not the endotoxin, is measured. Alternate methods like Endotoxin Activity (EA) assay and anti-endotoxin monoclonal antibodies were attempted but none of these were adequate to be administered beyond the trial period.

5.3. The Cellular Internalization of the Drug

A drug usually circulates and accumulates at the site of action before it is internalized in the cell. This process remains a chokepoint in the development of nanochemotherapeutic drugs mainly because of the inability of the drug to go inside the tumor cells [104]. Unlike conventional drugs, the full intracellular, paracellular, and transcellular pathways of the nanodrug across any biological membrane are not completely understood and
therefore, cellular internalization of nanochemotherapeutic drug possess an obstruction to its clinical development. The pharmaceutical formulation will follow only after a complete characterization of the internalization route [105]. Strategic research to overcome this dilemma was carried out when a nanomedicine based on a polyprodrug was developed that has the ability for zwitterionic-to-cationic charge conversion [106]. These pH-responsive nanomolecules respond well to the tumor microenvironment. The surface charge conversion effected by the nanodrug improves the efficiency of cellular internalization of the nanomedicines. Redox-responsive polymeric micelle conjugates are developed lately to target proteins and thus help in releasing the loaded drug to the designated site against cellular internalization complexities [107].

5.4. Sustained Release

Exceptional and sustained delivery of drugs to the target organs with minimum side effects is often a problem when drug development is involved. The EPR effect is a deciding factor in drug release and delivery. In a recent approach, stimuli-responsive nanocarriers were developed where biological (internal) or physical (exogenous) stimuli were directed to specifically targeted cells after drug delivery and the precise release of the drug was achieved [108].

5.5. Overcoming Biological Barriers Along with Increased Bioavailability

The accumulation of nanotherapeutic drugs at sites of disease progression is limited by biological barriers. Sometimes, the drug molecules merely get dispersed and distributed freely all over the body, resulting in detrimental side effects. This will also reduce the impact or response that the prescribed doses were supposed to bring about as expected by the physician [109]. The role of a suitable drug delivery system cannot be ignored at this point. Nanocarriers have brought about a revolution in the targeted drug delivery domain where delivery of nanochemodrugs that can traverse across different biological barriers was made possible. A recent article cited the use of Prunus spinosa fruits (PSF) containing different phenolic compounds as biomimetic nanoparticles that could improve the activity of the extract where the payload is directed into the lipid bilayer with increased accumulation at the diseased tissue (showing enhanced specific targeting properties). Improved biocompatibility, as well as a low toxicity, was observed when the nanoparticle was evaluated on HUVEC cells suggesting a unique platform for encapsulation of drugs that have minimal stability and bioavailability [110].

5.6. Increasing the Functional Capability to Target Only Tumor Cells

The heterogeneity exhibited by tumor cells and the relational differences among various tumor tissues inevitably contributes to the failure of drugs to target the tumor cells alone. It is a challenge to obtain any drug that is specific to different tumors and patients. The inconsistent characters of individual patients and their response to the drug contribute highly to the failure to target tumor cells alone. It is often difficult to select, purify and identify optimal ligands to carry the nanodrug. However, natural cellular materials (like exosomes) with intrinsic targeting properties can serve as nanocarriers for a drug aimed exclusively at tumor cells [111].

5.7. Controlling Immune System Response to the New Drug

It is difficult to predict the response of the immune system to the advent of a new molecule in the system. This is a major reason why the current drug therapies can extend survival rates but not cure the disease per se. The human immune system is sophisticated and complex. It would be a ‘dream-come-true’ situation if the mechanism could be deciphered properly. Products could be developed that could control the immune system to treat a wide variety of diseases. Immunoactive therapies is a novel cancer treatment centered around the manipulation of the immune system [112]. Here, a T-cell infusion system
was developed that will reflect the mechanism mediated by the transplanted donor immune system rather than the host immune system. The mechanism involved generated the Mirror Effect™ that elicits a host vs. tumor (HVT) effect along with a non-toxic host vs. graft (HVG) response that was found effective in cancer therapy, among other diseases. Similar to this, innovative ideas could change future research in drug discovery if they are focused on utilizing and exploiting the mechanism whereby the human immune system eliminates cancer and other diseases. This could lead to potential curative outcomes that revolutionize the chemotherapeutic and pharmaceutic industries.

Other challenges in this process are enumerated in Table 2. An exemplary drug delivery system must selectively deliver the drug to a specific location in the body to increase its therapeutic efficacy. In the case of cancer therapeutics, the efficiency is measured in terms of the ability of the drug to induce apoptosis in cancer cells. How effectively the therapeutic molecule reaches the target with the minimal adverse effect is crucial here. Selective accumulation of drugs in tumors through EPR effect, poor bio-accessibility of the drugs to tumor tissues, inadequate cellular uptake leading to the requirement of higher doses are some of the concerns to be addressed for effective therapy. Higher dosage leads to elevated toxicity to normal cells and a possibility for multiple drug resistance. Recent research revealed that nanoparticles are exceptional in addressing these challenges while avoiding toxicity to normal cells [113]. They can be employed to effectively deflect the unwanted toxicity to normal cells all the while ensuring the efficient induction of cell death of cancer cells. Keeping the focus on inducing maximum damage to the cancer cells, different innovative targeting strategies are followed to deliver therapeutic molecules like nucleic acids, proteins, genes, small molecules, and monoclonal antibodies to cancer cells. Nanoparticles are being established as the best option for deploying drugs through various delivery routes. Some of these delivery routes are schematically represented in Figure 6.

**Table 2. Challenges in the clinical translation of nanoparticles in cancer therapeutics.**

| Challenges                                                                 | Reference   |
|---------------------------------------------------------------------------|-------------|
| The long process of drug development                                       | [114]       |
| Years required for pre-clinical and clinical research on higher animals and humans | [115]       |
| Hassles in obtaining regulatory approval to release the drug in the market | [116]       |
| Failure to effectively load the drug inside the nanoparticles              | [117]       |
| Instability of the formulation                                             | [118]       |
| Issues with biocompatibility and toxicity                                  | [119]       |
| Insufficient residence time in the body                                    | [120]       |
| Failure of the drug formulation to selectively accumulate on the target    | [121]       |
| Failure in loading, internalization, and drug release                      | [122]       |
| Incomplete biodegradation and elimination                                  | [123]       |
| Challenges in cellular uptake                                              | [124]       |
| Failure to translate the in vitro results to in vivo studies               | [125]       |
Figure 6. Schematic representation of the drug targeting strategies followed to deliver therapeutic molecules to cancer cells. The various modes of drug delivery ensure that the drug is effectively delivered to the cells of interest and thus, unwanted damage to normal cells can be minimized.

6. Cutting-Edge Developments in Nanochemotherapy

Developments in cancer therapy are never-ending and with advanced tools and techniques, more and more robust systems are evolving. These could be solutions to overcome the obstacles encountered in cancer therapy. Some of the recent developments are briefed as below.

6.1. CRISPR—The Gene-Editing Tool

Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR), the Nobel prize-winning discovery developed by Jennifer Doudna and Emmanuelle Charpentier, is a gene-editing tool with high precision. This could alter human DNA so as to eliminate chances of disease incidence. CRISPR is the immune model that is used by bacteria to prevent invasion by bacteriophages. At its core, the CRISPR system enables prokaryotes to accurately recognize genetic sequences of phage and thus earmark these sequences for further destruction using unique enzymes. CRISPR/Cas9, a variation of technological innovation, is used for the treatment of cancer, where multiple genetic alterations are established [126]. Many recent researches successfully employed nanoparticles and nanoformulations to promote CRISPR/Cas9 development [127]. The antibody-conjugated tumor-targeted nanolipogels (tNLGs), were employed to efficiently encapsulate the CRISPR plasmids independent of electrostatic interaction, thus eliminating the cationic toxicity [128]. The highly efficient tNLGs utilize an antibody-guided strategy to selectively recognize and bind cancer cells, especially triple-negative breast cancer (TNBC) while sparing normal tissues, the low particle elasticity of the nNLGs allows them to directly release CRISPR plasmids into the cytosol of targeted TNBC cells via a receptor-mediated membrane fusion pathway, effectively avoiding endosome entrapment within TNBC cells. Lipid nanoparticles formulated with biodegradable ionizable lipids, PEG-DMG, Spy Cas9
mRNA, and sgRNA can effectively deliver CRISPR/Cas9 components in vivo for genomic editing and produce sustainable gene knockout within 52 weeks after a single administration [129]. Further, researchers have developed a multistage delivery polymer nanocarrier (MDNP) that responds to the tumor’s slightly acidic environment, thus achieving tumor-targeted delivery of the CRISPR/Cas9 system. MDNP can overcome various physiological barriers and achieve targeted delivery of CRISPR/Cas9 to inhibit tumor growth [130]. Cell type-specific targeting using nanoparticles that encode DNA aptamers are used to target specific tumor cells. A stimulus-responsive Cas9/sgRNA release with enhanced genome editing efficiency was achieved when they were combined with DNA nanoflowers [131]. The Cas9 protein (chemically modified with a glutamate peptide tag at the N-terminus) and sgRNA were delivered using cationic arginine-functionalized gold nanoparticles with an efficiency as high as 90–95% [132].

6.2. ThermoResponsive-NanoVelcro Purification System

This is a nanomaterial-embedded cancer diagnostic platform expected to bring together a distinct ‘rare-cell’ sorting method that enables detection, isolation, and characterization of circulating tumor cells in the peripheral blood. This is envisaged to provide non-invasive monitoring of the disease progression in patients. Previously, a nano Velcro chip was designed with PLGA embedded into it to capture M229 cells with high precision (87%) [133]. Another study was done where the NanoVelcro CTC purification system was used along with the NanoString Counter platform for cellular purification and RNA analysis in metastatic castration-resistant prostate cancer [134]. This mode of tumor therapy facilitates patient-specific tumor care and has the advantage of early detection of drug resistance. However, the ultimate aim of the group of scientists working on this system of cancer therapy is to expand the application to clinical studies where TR-NanoVelcro purification system enters translational and regulatory routes.

6.3. PROTAC—A Novel Proteolysis Targeting Entity

As every disease is caused due to the disparity of the associated protein moiety, finding novel drugs that can accurately control protein synthesis and degradation is an alternate approach. Dysfunctional proteins are the major reason for cancer and a drug that can inhibit the activation of such abnormal proteins and their subsequent pathways will be valuable for cancer cure. Proteolysis Targeting Chimeras (PROTACs) are emerging groups of small molecules that can effectively remove the dysfunctional proteins from the body [135]. These heterofunctional molecules can hijack the body’s natural disposal system and specifically choose the concerned protein to degrade them. Only very small amounts of PROTACs are needed for the action as compared to other conventional drugs, this reduces the possibility of the adverse effect.

BRD4 is a protein belonging to the Bromodomain and Extra terminal (BET) protein family. It acts as super-enhancers (SEs) organizer and a regulator of oncogenes’ expression [136]. The development of ARV825, a BRD4-degrading PROTAC molecule with anticancer activity was reported [137]. Authors report that the nanoformulation of ARV acts as a selective degrader of the BRD4 protein and is effective in targeting the ‘undruggable’ c-Myc for the treatment of pancreatic cancer. In another study, gold nanoparticle-based multiheaded PROTAC molecules were found effective in the targeted degradation of anaplastic lymphoma kinase, a major target in treating NSCLC [138]. Vectorization of encapsulated BET-PROTACs with antibody-conjugated nanoparticles (ACNPs) facilitated their controlled release and enhanced their pharmacokinetic and efficacy profile. In this study, the commercially available MZ1 PROTAC (a selective degrader of BRD4) was encapsulated into the FDA-approved polymeric nanoparticle—trastuzumab—to guide the delivery of MZ1 to breast cancer cells that overexpress HER2 [139].

6.4. Proton Therapy—An Alternate Approach to Conventional Radiation Therapy
Scientists at the Paul Scherrer Institute in Switzerland launched a novel alternative to conventional radiotherapy when they introduced proton beam therapy with the hope that it will prolong the survival rates of lung cancer patients. The photon beam therapy is expected to combat and reduce the adverse impact of radiotherapy or the conventional megavoltage (MV) photon therapy [140,141]. The photons used in X-rays in conventional radiation therapy can cause greater damage to healthier tissues when high doses are used. Proton beam, which is charged particle that is directed to the tumor with high precision. The protons can pass the entire tumor and deposit energy on the tumor. Proton therapy is known to induce tumor death by producing DNA lesions in the tumor cells, mainly as single-stranded breaks (SSB) and double-stranded breaks (DSB) [142]. Conventional radiation therapy is given with curative intent in fractions of 1.8–2.0 Gy daily whereas only 1 Gy is needed for proton therapy to produce 1×10^5 ionization events per cell. Around 1000–2,000 SSBs and 40DSBs are produced in this case, increasing lethality to tumor cells. Researchers predict a potential improvement in treatment outcome in cancer therapy when the dosimetric advantage of proton beams is integrated with radiosensitizers like gold nanoparticles [143]. However, the radiolysis yield and dose were found to be maximum only when nanoparticles of intermediate size were employed. [144]. A functionalized hafnium nanoparticle (NBTXR3) was developed and clinically tested where it acted as a selective radioenhancer to kill the tumor cells exposed to radiotherapy by the absorption and deposition of high radiation dose within the cells [145]. The nanoparticles were activated by intensity-modulated radiation therapy (IMRT) and used in the treatment of advanced-stage head and neck squamous cell carcinoma (NHSCC).

6.5. Functionalized DNA—A Programmable Way to Deliver Cancer Therapeutics

DNA nanotechnology is a technique of recent origin where the strands of self-assembling DNA hybridize with each other and can be fabricated into a functional nanostructure with extremely high spatial programmability. Inorganic nanomaterials (2D nanosheets, fluorescent nanomaterials) were incorporated into the DNA strands to be applied for diagnosis and cancer therapy [146]. Further, different linear DNA strands with predesigned sequences were successfully hybridized to self-assembled DNA tiles to form uniquely shaped nanostructures [147]. These DNA tiles can be programmed to combine with other tiles to make 2D and 3D nanoribbon lattices. They can further be functionalized with other nanoparticles like gold [148], graphene or graphene oxide [149] so as to interact with receptors on the cancer cell surfaces to activate signal transduction or other cellular mechanisms.

6.6. Avatar—A Real-Time Data Based Translational Therapeutic Approach

Advances in translational medicine have reached a new phase where real-time investigations are varied based on user rendition of the disease and their experience with the current therapy. Though there is a chance for misrepresentation or data modification influenced by the user’s self-conception, this is a research area worth exploring. Avatar is such an attempt already undertaken by groups of researchers where digital customization of the patient’s perception is demonstrated by innovative algorithms. These algorithms are specifically created to decide and predict clinical outcomes and the pros and cons of a specific field of research. These could be perceived as a digital twin of the patients’ identity wherein each and every treatment-related aspect like prognosis, treatment outcomes, complications, diagnostic operations, etc. could be virtually tried before execution upon the patients’ prompting [150]. However, despite these avatars and digital customization being a source of self-awareness to the large mass of patients, there is the drawback that these modifications and developments depend much on the patients’ self-conception.

6.7. Protein Catenation—A Novel Approach to Develop Artificial Antibodies
Catenanes are an interesting group of molecules with unique properties like enhanced thermal stability, remarkable proteolytic resistance and even enhanced enzymatic activity [151]. Catenation is an attractive approach to alter protein topology without much change in their native form. Exploiting the catenation process, it is now possible to develop new artificial antibodies that have a greater affinity to the target, better in vivo stability (due to resistance to proteolysis) and prolonged circulation time. These offer a more positive effect on the target molecule [152]. The higher-order protein catenation framework supports the insertion of various proteins of interest (POIs) like affibody (high-affinity proteins) specific to human epidermal growth factor receptor-2. This provides a good candidate for designing multivalent protein structures and motifs that can promote structure-activity relationships to advance protein therapeutics.

6.8. Other Approaches

A few other approaches were also evolved in recent times. The development of drug mimetics where a drug-like molecule can thwart the cancer-inducing effects of mutated regulator proteins [153,154], targeted radionuclide therapy (TRNT) where the radionuclides can potentially destroy tumor cells even if they do not possess specific tumor-associated antigen or receptor or biomarkers [155,156], bacterial cancer therapy where magnetically responsive bacteria are directed to tumor sites where they secrete toxins and compete for nutrients with the tumor cells thus destroying the tumor cells by modulating immune responses [157], multi-ion radiotherapy where a pure beam of heavy ions (like carbon ion) is used for radiotherapy [158], etc. are some of the other recent developments being explored for potential therapeutic uses in cancer cure.

7. Recent Advances in Clinical Studies with Nanoparticles in Tumor Therapeutics

As discussed earlier, bringing out a drug for chemotherapy to market is a race against time. In this era of precision medicine, a better understanding of translational research on where we stand and where we are going will help in designing a proper treatment regime for the various types of cancer.

Immunotherapy is one area where many advances have happened in terms of nanochemodrugs. The discovery of cellular and humoral immune responses to over-expressed tumor-associated antigens (TAA) like MUC1 in many patients with breast cancer and other forms of adenocarcinoma opened up many targets for immunotherapy. Tecemotide, a MUC1-specific cancer immunotherapy, is evaluated in Phase III clinical trials for the treatment of stage IIIA/IIIB NSCLC [159]. Similarly, Lipovaxin-MM, a dendritic targeted liposomal vaccine entered the phase 1 trial for malignant melanoma [160]. CRLX101 is a first-in-class nanopharmaceuticals based on cyclodextrin polymeric nanoparticle (CDP) technology that has the potential to translate therapy into clinical outcomes [161]. A combined polymeric nanoparticle consisting of a recombinant protein and cholesterol hydrophobized pullulan (CHP) complex was administered as repeated doses of the vaccine IMF-001, to patients with solid tumors that express NY-ESO-1 antigen. In other trials, combining the vaccine with PD-1 blockade held promise in human trials [162]. Further, many nanochemodrugs were evaluated for clinical trials in conjugation with protein components like albumin. The drug-binding capacity of the nanoparticle-conjugated albumin (Nab) complex is high and that is one of the reasons why most nanochemodrugs belong to this category. The therapeutic capacity of Nab-paclitaxel along with gemcitabine [163], atezolizumab [164], and cyclophosphamide [165] are being tried against metastatic and early-stage breast cancer. ABI-007 is another Nab-paclitaxel combination where a clinical trial was completed for stage IV NSCLC and metastatic breast cancer. Table 3 represents some of the nanoparticles in various forms that have undergone clinical study for different types of cancer.
### Table 3. Some of the nanodrugs that have undergone clinical studies in recent years (data compiled from clinicaltrials.gov).

| Nanodrug                                           | Conventional Drug                                           | Cancer Type                                                                 | Clinical Trials.gov Identifier |
|----------------------------------------------------|-------------------------------------------------------------|------------------------------------------------------------------------------|--------------------------------|
| Paclitaxel Nab                                     |                                                             | Breast cancer                                                                | NCT0093145                     |
| 5-Fluorouracil, Epirubicin, Cyclophosphamide (FEC) |                                                             | Breast cancer                                                                | NCT00110695                    |
| Carboplatin, Erlotinib hydrochloride               |                                                             | Breast cancer, NSCLC                                                         | NCT01928160                    |
| Phenezine sulfate                                  |                                                             | Metastatic breast cancer                                                     | NCT03505528                    |
| Doxorubicin hydrochloride, Cyclophosphamide, Filgrastim, Trastuzumab |                                                             | Estrogen receptor-positive, HER2-positive breast cancer                        | NCT00407888                    |
| Bevacizumab, Gemcitabine hydrochloride             |                                                             | Breast cancer                                                                | NCT00623233                    |
| Carboplatin, Erlotinib hydrochloride               |                                                             | NSCLC                                                                        | NCT00661193                    |
| Sargramostim                                        |                                                             | Brenner tumor, Fallopion tube cancer, Ovarian clear cell, cystadenocarcinoma, Ovarian epithelial cancer | NCT00466960                    |
| PIPAC                                              |                                                             | Peritoneal carcinomatosis, Ovarian cancer, Breast cancer, Stomach cancer, Pancreatic cancer | NCT03304210                    |
| Carboplatin, Herceptin®                            |                                                             | Breast cancer                                                                | NCT00748553                    |
| Ceritinib, Cisplatin, Gemcitabine hydrochloride    |                                                             | Advanced malignant solid neoplasm, ALK positive lung cancer, Metastatic pancreatic adenocarcinoma, Stages III and IV of pancreatic cancer | NCT02227940                    |
| Azacitidine (Vidaza)                               |                                                             | Advanced or metastatic breast cancer                                          | NCT03719326                    |
| Etrumadenant, IPI-549, Pegylated liposomal doxorubicin (PLD) |                                                             | Triple-negative breast cancer, Ovarian cancer                                |                                |
| Mifepristone                                       |                                                             | Male breast cancer, Ovarian cancer                                            | NCT01493310                    |
| Cetuximab, IMRT                                    |                                                             | Head and neck cancer                                                         | NCT00736619                    |
| Cetuximab, Cisplatin                               |                                                             | Head and neck cancer                                                         | NCT00833261                    |
| Leucovorin calcium, Irinotecan hydrochloride, Fluorouracil |                                                             | Adenocarcinoma, Cholangiocarcinoma, Gallbladder carcinoma, Gastric adenocarcinoma, Malignant gastrointestinal neoplasm, | NCT02333188                    |
| Drug                  | Indications                                                                 | NCT Reference |
|----------------------|-----------------------------------------------------------------------------|---------------|
| Metastatic pancreatic adenocarcinoma, Pancreatic adenocarcinoma, Stage III Ampulla of vater cancer, Stage III Pancreatic cancer, Stage IIIA Gallbladder cancer, Stage IIIA Gastric cancer, Stage IIIB Gallbladder cancer, Stage IIIB Gastric cancer, Stage IV Ampulla of vater cancer, Stage IV Gallbladder cancer, Stage IV Gastric cancer, Stage IV Pancreatic cancer | NCT00821964   |
| Imiquimod            | Male breast cancer, Recurrent breast cancer, Skin metastases, Stage IV breast cancer |               |
| Lapatinib            | Neoplasms, breast cancer                                                    | NCT00650910   |
| Pembrolizumab, Epirubicin, Cyclophosphamide | Malignant neoplasm of breast                                                | NCT03289819   |
| Alisertib            | Adenocarcinoma, Pancreatic neoplasms                                        | NCT01677559   |
| Lapatinib            | Bladder cancer, Brain and central nervous system tumors, Breast cancer, Esophageal cancer, Extragonadal germ cell tumor, Gastric cancer, Lung cancer, Ovarian cancer, Prostate cancer | NCT00313599   |
| Doxorubicin, Cyclophosphamide, Carboplatin, Trastuzumab, Bevacizumab | Breast cancer                                                              | NCT00254592   |
| BBI608, Gemcitabine, Oxaliplatin, Leucovorin, Irinotecan, Fluorouracil, MM-398 | Metastatic pancreatic adenocarcinoma                                      | NCT02231723   |
| Gemcitabine, Capecitabine | Pancreatic neoplasms, Pancreatic cancer, Adenocarcinoma                     | NCT01161186   |
| CORT125134           | Solid tumors                                                                 | NCT02762981   |
| Pembrolizumab        | Metastatic urothelial carcinoma                                              | NCT03464734   |
| Bevacizumab, Carboplatin, | Melanoma (skin)                                                          | NCT00626405   |
8. Therapeutic vs. Diagnostic Nanoparticles

The current review was mainly focusing on the therapeutic applications of nanoparticles. Diagnostic nanoparticles also contribute substantially to cancer patient management. Diagnosis in cancer heavily depends on spectroscopic techniques; Raman spectroscopy (Raman scattering) being the most used one. The penetration power and acquisition speed can be boosted with high specificity and sensitivity on integrating nanoparticles as contrast agents in imaging [166]. Surface-Enhanced Raman Scattering (SERS) coupled with nanoparticles have many advantages like simultaneous detection of different types of cancer [167], and a possibility to visualize extended tumors in vivo. In an interesting development, scientists have come up with a smartphone-based device called Krometriks for the colorimetric detection of MicroRNAs (miRNAs). The inventors claim that Krometriks is an easily accessible and affordable miRNA diagnostic tool that can be applied in low-resource situations [168]. The assay (called plasmonic coupling interference—PCI), is employed for miRNA detection using SERS by integrating silver nanoparticles (AgNP).

Another advanced method was developed recently where the ROS-responsive tripeptide sequence of Arg-Gly-Asp (RGD) was modified by nanoparticles for the simultaneous detection of cancer by near-infrared (NIR) imaging and photothermal therapy (PTT) [169]. Nanoparticles, especially the magnetic iron oxide nanoparticles, can act as coupling agents that bind to specific ligands that can detect gene mutations. This high-throughput screening method is highly sensitive and selectively detects different mutated nucleic acid sequences and can be employed for the diagnosis of cancer biomarkers [170]. However, the technique is still evolving and there is scope for further development of a translatory product.

9. Restrictions on the Use of Nanoparticles in Medicine

Though nanoparticles are generally synthesized and used for medicinal and therapeutic uses, the long-term side effects and drawbacks of using nanoparticles in medicine are still unknown [171]. Extensive studies are conducted on the toxicological aspects of engineered nanoparticles; concerns regarding the impact of these nanoparticles on public health are on the rise. Researchers believe that the extended exposure to nanoparticles can cause damage due to their superior penetrating power and unique toxicity due to ‘nano’size [172]. An extensive study on the developmental and neurobehavioral efficacy
of nanoparticles is needed to address this problem. Unfortunately, due to the relative newness of the involvement of nanoparticles in the field of medicine, there is no efficient regulatory technique for the validation of the hazards associated with the use of specific engineered nanoparticles as a medicine. This could lead to restrictions on the use of nanoparticles in medicine.

10. Future Perspective

Nanoparticles are rapidly changing the direction of drug treatment. Incorporating the enhanced properties of nanoparticles to cancer diagnosis and therapy has opened new avenues. Passive or active nanostructures can be created to specifically target a drug to remote regions of the body that are inaccessible for normal macromolecular drugs. For example, the novel fiber-optic dosimeter (nanoFOD) device based on nanomaterial is used to pinpoint and measure the in vivo radiation dose in real-time that was given during external beam delivery in radiation therapy sessions [173]. Furthermore, nanorobotics and molecular nanosystems can create artificial organs and system mimics have the potential to control the future of nanochemotherapy. The ‘safe-by-design’ concept for nanomaterials that is currently being investigated by scientists is expected to offer pharmaceutical companies a cost-effective platform in risk management. The safety or risk assessment can be made early during the development stages and can be incorporated into a final product.

11. Conclusions

Many promising strategies were postulated and investigated in the past few decades fighting the cancer. Several unconventional methods lead to the incorporation of nanoparticle-based drugs in the treatment and care of patients. Still, there are challenges and blockages, including those related to regulations and approvals, that may slow down the progress. Nevertheless, it is hoped that the scientific community, with the joint participation from academics, regulatory agencies, and industrial partners, will be able to bring out new, patient-centric nanodrugs that will accelerate their journey from bench to bedside, thus providing a paradigm shift from conventional tumor therapeutics.

Author Contributions: D.M.: Conceptualization, investigation, methodology, resources, visualization, writing—original draft W.C.: Conceptualization and design, supervision, methodology, visualization, writing—review All authors have read and agreed to the published version of the manuscript

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Rhodes, K.R.; Green, J.J. Nanoscale artificial antigen presenting cells for cancer immunotherapy. Mol. Immunol. 2018, 98, 13–18.
2. Wang, J.; Li, Y.; Nie, G. Multifunctional biomolecule nanostructures for cancer therapy. Nat. Rev. Mater. 2021, 6, 766–783.
3. Sun, T.; Zhang, Y.S.; Pang, B.; Hyun, D.C.; Yang, M.; Xia, Y. Engineered nanoparticles for drug delivery in cancer therapy. Angew. Chem. Int. Ed. 2014, 53, 12320–12364.
4. Hassan, H.A.F.M.; Smyth, L.; Wang, J.T.-W.; Costa, P.M.; Ratnasothy, K.; Diebold, S.S.; Lombardi, G.; Al-Jamal, K.T. Dual stimulation of antigen presenting cells using carbon nanotube-based vaccine delivery system for cancer immunotherapy. Biomaterials 2016, 104, 310–322.
5. Yetisgin, A.A.; Cetinel, S.; Zuvin, M.; Kosar, A.; Kutlu, O. Therapeutic nanoparticles and their targeted delivery applications. Molecules 2020, 25, 2193.
6. Bao, W.; Tian, F.; Lyu, C.; Liu, B.; Li, B.; Zhang, L.; Liu, X.; Li, F.; Li, D.; Gao, X. Experimental and theoretical explorations of nanocarriers’ multistep delivery performance for rational design and anticancer prediction. Sci. Adv. 2021, 7, eaba2458.
7. Sweeney, E.E.; Balakrishnan, P.B.; Powell, A.B.; Bowen, A.; Sarabia, I.; Burga, R.A.; Jones, R.B.; Bosque, A.; Cruz, C.R.Y.; Fernandez, R. PLGA nanodepots co-encapsulating prostratin and anti-CD25 enhance primary natural killer cell antiviral and antitumor function. Nano Res. 2020, 13, 736–744. https://doi.org/10.1007/s12274-020-2684-1.
8. Dagogo-Jack, I.; Shaw, A.T. Tumour heterogeneity and resistance to cancer therapies. Nat. Rev. Clin. Oncol. 2018, 15, 81–94.
9. Thakur, N.; Thakur, S.; Chatterjee, S.; Das, J.; Sil, P.C. Nanoparticles as smart carriers for enhanced cancer immunotherapy. *Front. Chem.* **2020**, *8*, 1217. https://doi.org/10.3389/fchem.2020.597806.

10. Das, K.; Belnoue, E.; Rossi, M.; Hofer, T.; Dankmayer, S.; Nolden, T.; Schreiber, L.-M.; Angerer, K.; Kimpel, J.; Hoecker, S. A modular self-adjuvanting cancer vaccine combined with an oncolytic vaccine induces potent antitumor immunity. *Nat. Commun.* **2021**, *12*, 1–14.

11. Bu, J.; Nair, A.; Iida, M.; Jeong, W.; Poellmann, M.J.; Mudd, K.; Kubiatowicz, L.J.; Liu, E.W.; Wheeler, D.L.; Hong, S. An avidity-based PD-L1 antagonist using nanoparticle-antibody conjugates for enhanced immunotherapy. *Nano Lett.* **2020**, *20*, 4901–4909.

12. Simonetta, F.; Alam, I.S.; Lohmeyer, J.K.; Sahaf, B.; Good, Z.; Chen, W.; Xiao, Z.; Hirai, T.; Scheller, L.; Engels, P. Molecular imaging of chimeric antigen receptor T cells by ICOS-immunoPET. *Clin. Cancer Res.* **2021**, *27*, 1058–1068.

13. Berraondo, P.; Sanmamed, M.F.; Ochoa, M.C.; Etxeberria, I.; Aznar, M.A.; Pérez-Gracia, J.L.; Rodríguez-Ruiz, M.E.; Ponz-Sarvise, M.; Castaño, E.; Melero, I. Cytokines in clinical cancer immunotherapy. *Br. J. Cancer* **2019**, *120*, 6–15.

14. Hemminki, O.; Dos Santos, J.M.; Hemminki, A. Oncolytic viruses for cancer immunotherapy. *J. Hematol. Oncol.* **2020**, *13*, 1–15.

15. Park, W.; Heo, Y.-J.; Han, D.K. New opportunities for nanoparticles in cancer immunotherapy. *Biomater. Res.* **2018**, *22*, 1–10.

16. Zolnik, B.S.; González-Fernández, Á.; Sadrieh, N.; Dobrovolskaia, M.A. Minireview: Nanoparticles and the immune system. *Endocrinology* **2010**, *151*, 458–465.

17. Setyawati, M.; Tay, C.Y.; Bay, B.H.; Leong, D.T. Gold nanoparticles induced endothelial leakiness depends on particle size and endothelial cell origin. *ACS Nano* **2017**, *11*, 5020–5030. https://doi.org/10.1021/acsnano.7b01744.

18. Tay, C.Y.; Setyawati, M.I.; Leong, D.T. Nanoparticle density: A critical biophysical regulator of endothelial permeability. *ACS Nano* **2017**, *11*, 2764–2772. https://doi.org/10.1021/acsnano.6b07806.

19. Wang, J.; Zhang, L.; Peng, F.; Shi, X.; Leong, D.T. Targeting endothelial cell junctions with negatively charged gold nanoparticles. *Chem. Mater.* **2018**, *30*, 3759–3767.

20. Ge, H.; Wang, D.; Pan, Y.; Guo, Y.; Li, H.; Zhang, F.; Zhu, X.; Li, Y.; Zhang, C.; Huang, L. Sequence-dependent DNA functionalization of upconversion nanoparticles and their programmable assemblies. *Angew. Chem. Int. Ed.* **2020**, *59*, 8133–8137. https://doi.org/10.1002/anie.202000831.

21. Ganbold, T.; Han, S.; Hasi, A.; Baigude, H. Receptor-mediated delivery of therapeutic RNA by peptide functionalized curdlan nanoparticles. *Int. J. Biol. Macrocol.* **2012**, *16*, 633–640. https://doi.org/10.1016/j.ijbiomac.2018.12.152.

22. Jia, X.; Guo, M.; Han, Q.; Tian, Y.; Yuan, Y.; Wang, Z.; Qian, Y.; Wang, W. Synergetic tumor probes for facilitating therapeutic delivery by combined-functionalized peptide ligands. *ACS Appl. Mater. Interfaces* **2020**, *12*, 5650–5655. https://doi.org/10.1021/acsanm.9b03440.

23. Guan, B.; Zhang, X. Aptamers as versatile ligands for biomedical and pharmaceutical applications. *Int. J. Nanomed.* **2020**, *15*, 1059–1071.

24. Marques, A.C.; Costa, P.J.; Velho, S.; Amaral, M.H. Functionalizing nanoparticles with cancer-targeting antibodies: A comparison of strategies. *J. Control. Release* **2020**, *320*, 180–200.

25. Bakshi, S.; Zakharchenko, A.; Minko, S.; Kolpashchikov, D.; Katz, E. Towards nanomaterials for cancer theranostics: A system of DNA-modified magnetic nanoparticles for detection and suppression of rna marker in cancer cells. *Magnetometry* **2019**, *5*, 24. https://doi.org/10.3390/magnetometry8020024.

26. Das, M.; Shen, L.; Liu, Q.; Goodwin, T.J.; Huang, L. Nanoparticle delivery of RIG-I agonist enables effective and safe adjuvant therapy in pancreatic cancer. *Mol. Ther. Oncolytic Mol. Ther.* **2020**, *17*, 507–517. https://doi.org/10.1016/j.ymthe.2018.11.012.

27. González-Ballesteros, N.; Diego-González, L.; Lastra-Valdor, M.; Rodriguez-Argüelles, M.C.; Grimaldi, M.; Cavazza, A.; Bigi, F.; Simón-Vázquez, R. Immunostimulant and biocompatible gold and silver nanoparticles synthesized using the: Ulva intestinalis L. aqueous extract. *ACS Nano* **2019**, *13*, 5076–5082. https://doi.org/10.1021/acsnano.9b05783.

28. Song, S.; Jin, X.; Zhang, L.; Zhao, C.; Ding, Y.; Ang, Q.; Khaidav, O.; Shen, C. PEGeGylated and CdD47-conjugated nanoellipsoid artificial antigen-presenting cells minimize phagocytosis and augment anti-tumor T-cell responses. *Int. J. Nanomed.* **2019**, *14*, 2465–2483. https://doi.org/10.2147/ijn.S195828.

29. Kim, U.; Kim, C.-Y.; Lee, J.M.; Oh, H.; Ryu, B.; Kim, J.; Park, J.-H. Phloretin inhibits the human prostate cancer cells through the generation of reactive oxygen species. *Pathol. Oncol. Res.* **2020**, *26*, 977–984.

30. Druck, T.; Cheung, D.G.; Park, D.; Trapano, F.; Pichiotti, F.; Gasparri, M.; Palumbo, T.; Aqeilan, R.I.; Gaudio, E.; Okumura, H. Fhit–Fdxr interaction in the mitochondria: Modulation of reactive oxygen species generation and apoptosis in cancer cells. *Cell Death Dis.* **2019**, *10*, 1–10.

31. Raj, S.; Khurana, S.; Choudhari, R.; Kesari, K.K.; Kamal, M.A.; Garg, N.; Ruokolainen, J.; Das, B.C.; Kumar, D. Specific Targeting Cancer Cells with Nanoparticles and Drug Delivery in Cancer Therapy. *Seminars in Cancer Biology* **2021**, *69*, 166–177.

32. Buttacavoli, M.; Albanese, N.N.; Di Cara, G.; Alduina, R.; Faleri, C.; Gallo, M.; Pizzolanti, G.; Gallo, G.; Feo, S.; Baldi, F. Anti-cancer activity of biogenerated silver nanoparticles: An integrated proteomic investigation. *Oncotarget* **2018**, *9*, 9685.

33. Wang, H.; Gao, Z.; Liu, X.; Agarwal, P.; Zhao, S.; Conroy, D.W.; Ji, G.; Yu, J.; Jaroniec, C.P.; Liu, Z. Targeted production of reactive oxygen species in mitochondria to overcome cancer drug resistance. *Nat. Commun.* **2018**, *9*, 1–16.

34. Grueso, M.J.L.; Valero, R.M.T.; Carmona, H.B.; Ruiz, D.J.L.; Peinado, J.; McDonagh, B.; Aguilar, R.R.; Ruiz, J.A.B.; Peña, C.A.P. Peroxiredoxin 6 down-regulation induces metabolic remodeling and cell cycle arrest in HepG2 cells. *Antioxidants* **2019**, *8*, 505.

35. Khan, S.; Ansari, A.A.; Khan, A.A.; Abdulla, M.; Al-Obaid, O.; Ahmad, R. In vitro evaluation of cytotoxicity, possible alteration of apoptotic regulatory proteins, and antibacterial activity of synthesized copper oxide nanoparticles. *Colloid. Surf. B Biointerfaces* **2017**, *153*, 320–326.
36. Varlamova, E.G.; Goltayev, M.V.; Mal'tseva, V.N.; Turovsky, E.A.; Sarimov, R.M.; Simakin, A.V.; Gudkov, S.V. Mechanisms of the cytotoxic effect of selenium nanoparticles in different human cancer cell lines. Int. J. Mol. Sci. 2022, 23, 1685.

37. Turovsky, E.A.; Varlamova, E.G. Mechanism of Ca2+-dependent pro-apoptotic action of selenium nanoparticles, mediated by activation of Ca233 hemichannels. Biology 2021, 10, 743.

38. Zhao, X.; Takabayashi, F.; Ibuki, Y. Coexpression to silver nanoparticles and ultraviolet a synergistically enhances the phosphorylation of histone H2AX. J. Photochem. Photobiol. B Biol. 2016, 162, 213–222.

39. Bhowmik, T.; Gomes, A. Down-regulation of cyclin-dependent kinase-4 and MAPK through estrogen receptor mediated cell cycle arrest in human breast cancer induced by gold nanoparticle tagged toxin protein NKCT1. Chem. Biol. Interact. 2017, 268, 119–128. https://doi.org/10.1016/j.cbi.2017.03.009.

40. Huai, Y.; Zhang, Y.; Xiong, X.; Das, S.; Bhattacharya, R.; Mukherjee, P. Gold nanoparticles sensitize pancreatic cancer cells to gemcitabine. Cell Stress 2019, 3, 267–279.

41. Mi, Y.; Shao, Z.; Vang, J.; Kaidar-Person, O.; Wang, A.Z. Application of nanotechnology to cancer radiotherapy. Cancer Nanotechnol. 2016, 7, 1–16.

42. Guryev, E.L.; Volodina, N.O.; Shilyagina, N.Y.; Gudkov, S.V.; Balalaeva, I.V.; Volovetskiy, A.B.; Lyubeshkin, A.V.; Sen, A.V.; Ermilov, S.A.; Vodeneev, V.A.; et al. Radioactive (90Y) upconversion nanoparticles conjugated with recombinant targeted toxin for synergistic nanotheranostics of cancer. Proc. Natl. Acad. Sci. USA 2018, 115, 9690–9695. https://doi.org/10.1073/pnas.1809285115.

43. Laprise-Pelletier, M.; Laqueux, J.; Côté, M.; LaGrange, T.; Fortin, M. Low-dose prostate cancer brachytherapy with radioactive palladium–Gold nanoparticles. Adv. Healthc. Mater. 2017, 6, 1601120.

44. Deng, J.; Xu, S.; Hu, W.; Xun, X.; Zheng, L.; Su, M. Tumor targeted, stealthy and degradable bismuth nanoparticles for enhanced X-ray radiation therapy of breast cancer. Biomaterials 2018, 154, 24–33.

45. Hernandez-Delgadillo, R.; García-Cuéllar, C.M.; Sánchez-Pérez, Y.; Pineda-Aguilar, N.; Martínez-Martínez, M.A.; Rangel-Padilla, E.E.; Nakagoshi-Cepeda, S.E.; Solís-Soto, J.M.; Sánchez-Nájera, R.L.; Nakagoshi-Cepeda, M.A.A. In vitro evaluation of the antitumor effect of bismuth lipophilic nanoparticles (BisBAL NPs) on breast cancer cells. Int. J. Nanomed. 2018, 13, 6089.

46. Winter, H.; Brown, A.L.; Goforth, A.M. Bismuth-based nano- and microparticles in X-ray contrast, radiation therapy, and radiation shielding applications. Bismuth Adv. Appl. Defects Charact. 2017, 71, 1112–1141.

47. Vines, J.B.; Yoon, J.-H.; Ryu, N.-E.; Lim, D.-J.; Park, H. Gold nanoparticles for photothermal cancer therapy. Front. Chem. 2019, 7, 167.

48. Roy, S.; Tony, B.L. Phototherapy in cancer prevention and treatment. J. Cancer Prev. Curr. Res. 2017, 7, 22–24.

49. Rivera-Rodriguez, A.; Chiu-Lam, A.; Morozov, V.M.; Ishov, A.M.; Rinaldi, C. Magnetic nanoparticle hyperthermia potentiates paclitaxel activity in sensitive and resistant breast cancer cells. Int. J. Nanomed. 2018, 13, 4771.

50. Sohail, A.; Ahmad, Z.; Bég, O.A.; Arshad, S.; Sherin, L. A review on hyperthermia via nanoparticle-mediated therapy. Bull. Cancer 2017, 104, 452–461.

51. Legge, C.J.; Colley, H.E.; Lawson, M.A.; Rawlings, A.E. Targeted magnetic nanoparticle hyperthermia for the treatment of oral cancer. J. Oral Pathol. Med. 2019, 48, 803–809.

52. Heidari, M.; Sattarrahimddy, N.; Azarpira, N.; Heli, H.; Mehdizadeh, A.R.; Zare, T. Photothermal cancer therapy by gold-ferrite nanocomposite and near-infrared laser in animal model. Lasers Med. Sci. 2016, 31, 221–227.

53. Ma, Y.C.; Zhu, Y.H.; Tang, X.F.; Hang, L.F.; Jiang, W.; Li, M.; Khan, M.I.; You, Y.Z.; Wang, Y.C. Au nanoparticles with enzyme-mimicking activity-ornamented ZIF-8 for highly efficient photodynamic therapy. Biomater. Sci. 2019, 7, 2740–2748. https://doi.org/10.1039/c9bm00333a.

54. Kaur, P.; Aliru, M.I.; Chadha, A.S.; Asea, A.; Krishnan, S. Hyperthermia using nanoparticles–promises and pitfalls. Int. J. Hyperth. 2016, 32, 76–88.

55. Bagherti, S.; Yasemi, M.; Safae-Qamarsi, E.; Rashidiani, J.; Abkar, M.; Hassani, M.; Mirhosseini, S.A.; Kooshki, H. Using gold nanoparticles in diagnosis and treatment of melanoma cancer. Artif. Cells Nanomed. Biotechnol. 2018, 46, 462–471.

56. Afzal, M.; Alharbi, K.S.; Alruwaili, N.K.; Al-Abassi, F.A.; Al-Malki, A.A.L.; Kazmi, I.; Kumar, V.; Kamal, M.A.; Nadeem, M.S.; Aslam, M. Nanomedicine in Treatment of Breast Cancer–A Challenge to Conventional Therapy. Seminars in Cancer Biology 2021, 69, 279–292.

57. Huang, W-Y.; Lin, J.-N.; Hsieh, J.-T.; Chou, S.-C.; Lai, C.-H.; Yun, E.-J.; Lo, U.-G.; Pong, R.-C.; Lin, J.-H.; Lin, Y.-H. Nanoparticle targeting CD44-positive cancer cells for site-specific drug delivery in prostate cancer therapy. ACS Appl. Mater. Interfaces 2016, 8, 30722–30734.

58. Cheng, F.-F.; Sun, P.; Xiong, W.-W.; Zhang, Y.; Zhang, Q.; Yao, W.; Cao, Y.; Zhang, L. Multifunctional titanium phosphate nanoparticles for site-specific drug delivery and real-time therapeutic efficacy evaluation. Analyst 2019, 144, 3103–3110.

59. Bish, G.; Rayamajhi, S.; Biplab, K.C.; Paudel, S.N.; Karna, D.; Shrestha, B.G. Synthesis, characterization, and study of in vitro cytotoxicity of ZnO-Fe3O4 magnetic composite nanoparticles in human breast cancer cell line (MDA-MB-231) and mouse fibroblast (NIH 3T3). Nanoscale Res. Lett. 2016, 11, 1–11.

60. Tai, W.; Mahato, R.; Cheng, K. The role of HER2 in cancer therapy and targeted drug delivery. J. Control. release 2010, 146, 264–275.

61. Kaplan, A.; Kutlu, H.M.; Ciftci, G.A. Fe3O4 nanopowders: Genomic and apoptotic evaluations on A549 lung adenocarcinoma cell line. Nutr. Cancer 2020, 72, 708–721.
62. Nebu, J.; Devi, J.S.A.; Aparna, R.S.; Abha, K.; Sony, G. Erlotinib conjugated gold nanocluster enveloped magnetic iron oxide nanoparticles—A targeted probe for imaging pancreatic cancer cells. Sens. Actuators B Chem. 2018, 257, 1035–1043.

63. Zhang, T.; Li, Y.; Hong, W.; Chen, Z.; Peng, P.; Yuan, S.; Qu, J.; Xiao, M.; Xu, L. Glucose oxidase and polydopamine functionalized iron oxide nanoparticles: Combination of the photothermal effect and reactive oxygen species generation for dual-modality selective cancer therapy. J. Mater. Chem. B 2019, 7, 2190–2200.

64. Das, S.; Roy, A.; Barui, A.K.; Alabbasi, M.M.A.; Kunche, M.; Sistla, R.; Sreedhar, B.; Patra, C.R. Anti-angiogenic vanadium pentoxide nanoparticles for the treatment of melanoma and their in vivo toxicity study. Nanoscale 2020, 12, 7604–7621. https://doi.org/10.1039/d0nr00631a.

65. Wang, P.; Liu, S.; Hu, M.; Zhang, H.; Duan, D.; He, J.; Hong, J.; Lv, R.; Choi, H.S.; Yan, X.; et al. Peroxidase-like nanozymes induce a novel form of cell death and inhibit tumor growth in vivo. Adv. Funct. Mater. 2020, 30, 2000647. https://doi.org/10.1002/adfm.202000647.

66. Chen, M.; Deng, G.; He, Y.; Li, X.; Liu, W.; Wang, W.; Zhou, Z.; Yang, H.; Yang, S. Ultrasound-enhanced generation of reactive oxygen species for mri-guided tumor therapy by the Fe3O4-based peroxidase-mimicking nanzyme. ACS Appl. Bio Mater. 2020, 3, 639–647. https://doi.org/10.1021/acsabm.9b01006.

67. Miao, Z.; Jiang, S.; Ding, M.; Sun, S.; Ma, Y.; Younis, M.R.; He, G.; Wang, J.; Lin, J.; Cao, Z. Ultrasmall rhodium nanozyme with RONS scavenging and photothermal activities for anti-inflammation and antitumor theranostics of colon diseases. Nano Lett. 2020, 20, 3079–3089.

68. Wu, F.; Zhu, J.; Li, G.; Wang, J.; Veeraraghavan, V.P.; Krishna Mohan, S.; Zhang, Q. Biologically synthesized green gold nanoparticles from Siberian ginseng induce growth-inhibitory effect on melanoma cells (B16). Artif. Cells Nanomed. Biotechnol. 2019, 47, 3297–3305.

69. Khavi, A.A. PEGylated gold nanoparticles-ribonucleosome induced oxidative stress and apoptosis in colorectal cancer cells. Biomedicines 2019, 10, 27–36. https://doi.org/10.15171/bi.2020.04.

70. Chen, C.-C.; Hsieh, D.-S.; Huang, K.-J.; Chan, Y.-L.; Hong, P.-D.; Yeh, M.-K.; Wu, C.-J. Improving anticancer efficacy of (−)-epigallocatechin-3-gallate gold nanoparticles in murine B16F10 melanoma cells. Drug Des. Devel. Ther. 2014, 8, 459.

71. Jiang, T.; Zhang, B.; Zhang, L.; Wu, X.; Li, H.; Shen, S.; Luo, Z.; Liu, X.; Hu, Y.; Pang, Z. Biomimetic nanoparticles delivered hedgehog pathway inhibitor to modify tumour microenvironment and improved chemotherapy for pancreatic carcinoma. Artif. Cells Nanomed. Biotechnol. 2018, 46, 1088–1101.

72. Du, Y.; Zhang, J.; Yan, S.; Tao, Z.; Wang, C.; Huang, M.; Zhang, X. PEGylated zinc oxide nanoparticles induce apoptosis in pancreatic cancer cells through reactive oxygen species. IET Nanobiotechnol. 2019, 13, 80–84. https://doi.org/10.1049/iet-nhb.2018.5327.

73. Chen, C.; James, S.A.; de Jonge, M.D.; Turney, T.W.; Wright, P.F.A.; Felts, B.N. Relating cytotoxicity, zinc ions, and reactive oxygen in ZnO nanoparticle-exposed human immune cells. Toxicol. Sci. 2013, 136, 120–130.

74. Al-Sheddi, E.S.; Farshori, N.N.; Al-Oqail, M.M.; Al-Massarani, S.M.; Saquib, Q.; Wahab, R.; Musarrat, J.; Al-Khedhairy, A.A.; Siddiqui, M.A. Anticancer potential of green synthesized silver nanoparticles using extract of Nepeta deflersiana against human cervical cancer cells (HeLa). J. Bionanomed. 2018, 10, 1–12.

75. Dobrucka, R.; Romaniuk-Drapala, A.; Kaczmarek, M. Evaluation of biological synthesized platinum nanoparticles using Ononis radix extract on the cell lung carcinoma A549. Biomed. Microdevices 2019, 21, 1–10. https://doi.org/10.1007/s10544-019-0424-7.

76. Fujiwara, R.; Luo, Y.; Sasaki, T.; Fujii, K.; Ohmori, H.; Kuniyasu, H. Cancer therapeutic effects of titanium dioxide nanoparticles are associated with oxidative stress and cytokine induction. Pathobiology 2015, 82, 243–251. https://doi.org/10.1159/000439404.

77. Murugan, C.; Murugan, N.; Sundaramoorthy, A.K.; Sundaramurthy, A. Nanoceria decorated flower-like molybdenum sulphide nanoflakes: An efficient nanzyme for tumour selective ROS generation and photo thermal therapy. Chem. Commun. 2019, 55, 8017–8020. https://doi.org/10.1039/c9cc03763b.

78. Nejdl, L.; Kudr, J.; Moullick, A.; Hegerova, D.; Ruttkay-Nedecky, B.; Gumulec, J.; Cihalova, K.; Smerkova, K.; Dostalova, S.; Krizkova, S.; et al. Platinum nanoparticles induce damage to DNA and inhibit DNA replication. PLoS ONE 2017, 12, e0180798. https://doi.org/10.1371/journal.pone.0180798.

79. Nourmohammadi, E.; Khoshdel-Sarkarizi, H.; Nedaeinia, R.; Sadeghnia, H.R.; Hasanzadeh, L.; Darroudi, M.; Kazemi oskuee, R. Evaluation of anticancer effects of cerium oxide nanoparticles on mouse fibrosarcoma cell line. J. Cell. Physiol. 2019, 234, 4987–4996.

80. Parvathyra, S.; Venkattraman, B.R. In vitro antibacterial and anticancer potential of CeO2 nanoparticles prepared by co-precipitation and green synthesis method. J. Nanosci. Curr. Res. 2017, 2, 1–9.

81. Kadhem, H.A.; Ibrahim, S.A.; Jabir, M.S.; Kadhim, A.A. ZainAg-doping regulates the cytotoxicity of TiO2 nanoparticles via oxidative stress in human cancer cellsin, Zinc Oxide nanoparticles induce apoptosis in human breast cancer cells via caspase-8 and P53 pathway. Nano Biomed. Eng. 2019, 11, 35–43.

82. Ahamed, M.; Khan, M.A.M.; Akhtar, M.J.; Alhadiq, H.A.; Alshamsan, A. Ag-doping regulates the cytotoxicity of TiO2 nanoparticles via oxidative stress in human cancer cells. Sci. Rep. 2017, 7, 1–14.

83. Ferrari, R.; Sponchiioni, M.; Morbidelli, M.; Moscatelli, D. Polymer nanoparticles for the intravenous delivery of anticancer drugs: The checkpoints on the road from the synthesis to clinical translation. Nanoscale 2018, 10, 22701–22719.

84. He, H.; Liu, L.; Morin, E.E.; Liu, M.; Schwendeman, A. Survey of clinical translation of cancer nanomedicines—Lessons learned from successes and failures. Acc. Chem. Res. 2019, 52, 2445–2461.
85. Pandey, H.; Rani, R.; Agarwal, V. Liposomes and their applications in cancer therapy. *Braz. Arch. Biol. Technol.* 2016, 59, 59.
86. Oltusanya, T.O.B.; Haj Ahmad, R.R.; Ibegbu, D.M.; Smith, J.R.; Elkordy, A.A. Liposomal drug delivery systems and anticancer drugs. *Molecules* 2018, 23, 907.
87. Rommarsi, F.; Esfandiar, N. Liposomal nanomedicine: Applications for drug delivery in cancer therapy. *Nanoscale Res. Lett.* 2021, 16, 1–20.
88. Ndagi, U.; Mhlongo, N.; Soliman, M.E. Metal complexes in cancer therapy—an update from drug design perspective. *Drug Des. Devel. Ther.* 2017, 11, 599.
89. Lakshmi, B.A.; Reddy, A.S.; Sangubotla, R.; Hong, J.W.; Kim, S. Ruthenium (II)-curcumin liposome nanoparticles: Synthesis, characterization, and their effects against cervical cancer. *Colloids Surf. B Biointerfaces* 2021, 204, 111773.
90. Mignani, S.; Shi, X.; Ceña, V.; Rodrigues, J.; Tomas, H.; Majoral, J.-P. Engineered non-invasive functionalized dendrimer/dendron-entrapped/complexed gold nanoparticles as a novel class of theranostic (radio) pharmaceuticals in cancer therapy. *J. Control. Release* 2021, 332, 346–366.
91. Gul, A.R.; Shaheen, F.; Rafique, R.; Bal, J.; Waseem, S.; Park, T.J. Grass-mediated biogenic synthesis of silver nanoparticles and their drug delivery evaluation: A biocompatible anti-cancer therapy. *Chem. Eng. J.* 407, 127202.
92. Varlamova, E.G.; Turovsky, E.A.; Blinova, E.V. Therapeutic potential and main methods of obtaining selenium nanoparticles. *Int. J. Mol. Sci.* 2022, 22, 10808.
93. Lorkowski, M.E.; Atukorale, P.U.; Ghaghada, K.B.; Karathanasis, E. Stimuli-responsive iron oxide nanotheranostics: A versatile and powerful approach for cancer therapy. *Adv. Healthc. Mater.* 2021, 10, 2001044.
94. Ghosh, B.; Biswas, S. Polymeric micelles in cancer therapy: State of the art. *J. Control Release* 2021, 10,127-147.
95. Zhou, Q.; Zhang, L.; Yang, T.; Wu, H. Stimulus-responsive polymeric micelles for drug delivery and cancer therapy. *Int. J. Nanomedicine* 2018, 13, 2921.
96. Persano, F.; Gigli, G.; Leporatti, S. Lipid-polymer hybrid nanoparticles in cancer therapy: Current overview and future directions. *Nano Express* 2022, 2, 012006–012022.
97. Jana, D.; Jia, S.; Bindra, A.K.; Xing, P.; Ding, D.; Zhao, Y. Clearable black phosphorus nanoconjugate for targeted cancer phototheranostics. *ACS Appl. Mater. Interfaces* 2012, 18342–18351.
98. Payne, M.; Ellis, P.; Dunlop, D.; Ranson, M.; Danson, S.; Schacter, L.; Talbot, D. DHA-paclitaxel (Taxoprexin) as first-line treatment in patients with stage IIIB or IV non-small cell lung cancer: Report of a phase II open-label multicenter trial. *J. Thorac. Oncol.* 2006, 1, 984–990.
99. Ventola, C.L. Progress in nanomedicine: Approved and investigational nanodrugs. *Pharm. Ther.* 2017, 42, 742–755.
100. Fymat, A.L. Magnetic resonance imaging modalities with contrast enhancing nanomaterials. *Curr. Trends Clin. Med. Imaging* 2017, 1, 1–14.
101. Alshamamri, T.M. Drug safety: the concept, inception and its importance in patients’ health. *Saudi Pharm. J.* 2016, 24, 405–412.
102. Dordević, S.; Gonzalez, M.M.; Conejos-Sánchez, I.; Carreira, B.; Pozzi, S.; Acúrcio, R.C.; Satchi-Fainaro, R.; Florindo, H.F.; Vicent, M.J. Current hurdles to the translation of nanomedicines from bench to the clinic. *Drug Deliv. Transl. Res.* 2021, 12, 1–26.
103. Cross, A. Endotoxin: Back to the future. *Crit. Care Med.* 2016, 44, 450–454.
104. He, B.; Sui, X.; Yu, B.; Wang, S.; Shen, Y.; Cong, H. Recent advances in drug delivery systems for enhancing drug penetration into tumors. *Drug Deliv.* 2020, 27, 1474–1490.
105. Murugan, K.; Choorna, Y.E.; Kumar, P.; Bijukumar, D.; du Toit, L.C.; Pillay, V. Parameters and characteristics governing cellular internalization and trans-barrier trafficking of nanostructures. *Int. J. Nanomed.* 2015, 10, 2191.
106. Wang, S.; Zhang, F.; Yu, G.; Wang, Z.; Jacobson, O.; Ma, Y.; Tian, R.; Deng, H.; Yang, W.; Chen, Z.-Y. Zwitterionic-to-cationic charge conversion polyprodrug nanomedicine for enhanced drug delivery. *Theranostics* 2020, 10, 6629–6637.
107. Du, Y.; Wang, S.; Zhang, T.; He, D.; Tu, J.; Shen, Y. Enhanced cytotoxicity of a redox-sensitive hyaluronic acid-based nanomedicine toward different oncocytes via various internalization mechanisms. *Drug Deliv.* 2020, 27, 128–136.
108. Abasian, P.; Shakibi, S.; Maniati, M.S.; Nouri Khorasani, S.; Khalili, S. Targeted delivery, drug release strategies, and toxicity study of polymeric drug nanocarriers. *Polym. Adv. Technol.* 2021, 32, 931–944.
109. Blanco, E.; Shen, H.; Ferrari, M. Principles of nanoparticle design for overcoming biological barriers to drug delivery. *Nat. Biotechnol.* 2015, 33, 941–951.
110. Tiboni, M.; Coppari, S.; Casettari, L.; Guescini, M.; Colomba, M.; Fraternali, D.; Gorassini, A.; Verardo, G.; Ramakrishna, S.; Guidi, L. Prunus spinosa extract loaded in biomimetic nanoparticles evokes in vitro anti-inflammatory and wound healing exosome Activities. *Nanomaterials* 2021, 11, 36.
111. He, Z.; Zhang, Y.; Feng, N. Cell membrane-coated nanosized active targeted drug delivery systems homing to tumor cells: A review. *Mater. Sci. Eng. C* 2020, 106, 110298.
112. Goswitz, V.C.; Peter Sawicki, Z. Cancer therapy based on a mechanism of action for controlling the immune system and the resulting patent portfolio. *Recent Pat. Endocr. Metab. Immune Drug Discov.* 2013, 7, 1–10.
113. Senapati, S.; Mahanta, A.K.; Kumar, S.; Maiti, P. Controlled drug delivery vehicles for cancer treatment and their performance. *Signal. Transduct. Target. Ther.* 2018, 3, 1–19.
114. DiMasi, J.A.; Feldman, L.; Seckler, A.; Wilson, A. Trends in risks associated with new drug development: Success rates for investigational drugs. *Clin. Pharmacol. Ther.* 2010, 87, 272–277.
115. Matias, M.; Pinho, J.O.; Penetra, M.J.; Campos, G.; Reis, C.P.; Gaspar, M.M. The challenging melanoma landscape: From early drug discovery to clinical approval. *Cells* 2021, 10, 3088–3128.
116. Farjadian, F.; Ghasemi, A.; Gohari, O.; Rooointan, A.; Karimi, M.; Hamblin, M.R. Nanopharmaceuticals and nanomedicines currently on the market: Challenges and opportunities. *Nanomedicine* 2019, 14, 93–126.

117. Cheow, W.S.; Hadimoto, K. Factors affecting drug encapsulation and stability of lipid–polymer hybrid nanoparticles. *Colloids Surf. B Biointerfaces* 2011, 85, 214–220.

118. Hafeez, M.N.; Celia, C.; Petrikaite, V. Challenges towards targeted drug delivery in cancer nanomedicines. *Processes* 2021, 9, 1527.

119. Gavas, S.; Quazi, S.; Karpiński, T.M. Nanoparticles for cancer therapy: Current progress and challenges. *Nanoscale Res. Lett.* 2021, 16, 1–21.

120. Rodriguez, P.L.; Harada, T.; Christian, D.A.; Pantano, D.A.; Tsai, R.K.; Discher, D.E. Minimal “Self” peptides that inhibit phagocytic clearance and enhance delivery of nanoparticles. *Science* 2013, 339, 971–975.

121. Sanna, V.; Pala, N.; Sechi, M. Targeted therapy using nanotechnology: Focus on cancer. *Int. J. Nanomed.* 2014, 9, 467–483.

122. Salvioni, L.; Rizzuto, M.A.; Bertolini, J.A.; Pandolfi, L.; Colombo, M.; Prosperi, D. Thirty years of cancer nanomedicine: Success, frustration, and hope. *Cancers* 2019, 11, 1855.

123. Duan, X.; Li, Y. Physicochemical characteristics of nanoparticles affect circulation, biodistribution, cellular internalization, and trafficking. *Small* 2013, 9, 1521–2312.

124. Mosquera, J.; García, I.; Liz-Marzán, L.M. Cellular uptake of nanoparticles versus small molecules: A matter of size. *Acc. Chem. Res.* 2018, 51, 2305–2313.

125. Agrahari, V.; Agrahari, V. Facilitating the translation of nanomedicines to a clinical product: Challenges and opportunities. *Drug Discov. Today* 2018, 23, 974–991.

126. Xu, X.; Liu, C.; Wang, Y.; Koivistu, O.; Zhou, J.; Shu, Y.; Zhang, H. Nanotechnology-based delivery of CRISPR/Cas9 for cancer treatment. *Adv. Drug Deliv. Rev.* 2021, 176, 113891.

127. Duan, L.; Ouyang, K.; Xu, X.; Xu, L.; Wen, C.; Zhou, X.; Qin, Z.; Xu, Z.; Sun, W.; Liang, Y. Nanoparticle delivery of CRISPR/Cas9 for genome editing. *Front. Genet.* 2021, 12, 673286.

128. Guo, P.; Yang, J.; Huang, J.; Auguste, D.T.; Moses, M.A. Therapeutic genome editing of triple-negative breast tumors using a noncationic and deformable nanolipogel. *Proc. Natl. Acad. Sci. USA* 2019, 116, 18295–18303.

129. Finn, J.D.; Smith, A.R.; Patel, M.C.; Shaw, L.; Younisii, M.R.; van Heteren, J.; Diristine, T.; Ciullo, C.; Lescarbeau, R.; Seitzer, J. A single administration of CRISPR/Cas9 lipid nanoparticles achieves robust and persistent in vivo genome editing. *Cell Rep.* 2018, 22, 2227–2235.

130. Liu, Q.; Zhao, K.; Wang, C.; Zhang, Z.; Zheng, C.; Zhao, Y.; Zheng, Y.; Liu, C.; An, Y.; Shi, L. Multistage delivery nanoparticle facilitates efficient CRISPR/dCas9 activation and tumor growth suppression in vivo. *Adv. Sci.* 2019, 6, 1801423.

131. Shi, J.; Yang, X.; Li, Y.; Wang, D.; Liu, W.; Zhang, Z.; Liu, J.; Zhang, K. MicroRNA-responsive release of Cas9/gRNA from DNA nanoflower for cytosolic protein delivery and enhanced genome editing. *Biomaterials* 2020, 256, 120221.

132. Mout, R.; Rotello, V.M. Cytosolic and nuclear delivery of CRISPR/Cas9-ribonucleoprotein for gene editing using arginine functionalized gold nanoparticles. *Bio-Protocol* 2017, 7, 20.

133. Hou, S.; Zhao, L.; Shen, Q.; Yu, J.; Ng, C.; Kong, X.; Wu, D.; Song, M.; Shi, X.; Xu, X. Polymer nanofiber-embedded microchips for detection, isolation, and molecular analysis of single circulating melanoma cells. *Angew. Chem.* 2013, 125, 3463–3467.

134. Jan, Y.J.; Yoon, J.; Chen, J.F.; Teng, P.C.; Yao, N.; Cheng, S.; Lozano, A.; Chu, G.C.Y.; Chung, H.; Lu, Y.-T. A circulating tumor cell-RNA assay for assessment of androgen receptor signaling inhibitor sensitivity in metastatic castration-resistant prostate cancer. *Theranostics* 2019, 9, 2812.

135. Gao, H.; Sun, X.; Rao, Y. PROTAC technology: Opportunities and challenges. *ACS Med. Chem. Lett.* 2020, 11, 237–240.

136. Donati, B.; Lorenzini, E.; Ciarrochi, A. BRD4 and Cancer: Going beyond transcriptional regulation. *Mol. Cancer* 2018, 17, 1–13.

137. Saraswat, A.; Patki, M.; Fu, Y.; Barot, S.; Dukhanded, V.V.; Patel, K. Nanof ormulation of PROteolysis TArgeting chimera targeting ‘undruggable’ c-Myc for the treatment of pancreatic cancer. *Nanomedicine* 2020, 15, 1761–1777.

138. Wang, Y.; Han, L.; Liu, F.; Yang, F.; Jiang, X.; Sun, H.; Feng, F.; Xue, J.; Liu, W. Targeted degradation of anaplastic lymphoma kinase by gold nanoparticle-based multi-headed proteolysis targeting chimeras. *Colloids Surf. B Biointerfaces* 2020, 188, 110955–110904.

139. Cimas, F.J.; Niza, E.; Juan, A.; Noblejas-López, M.d.M.; Bravo, I.; Lara-Sanchez, A.; Alonso-Moreno, C.; Ocaña, A. Controlled delivery of BET-PROTACs: In vitro evaluation of mz1-loaded polymeric antibody conjugated nanoparticles in breast cancer. *Pharmaceutics* 2020, 12, 986–997.

140. Han, Y. Current status of proton therapy techniques for lung cancer. *Radiat. Oncol. J.* 2019, 37, 232–248.

141. Peukert, D.; Kempsom, I.; Douglass, M.; Bezak, E. Gold nanoparticle enhanced proton therapy: Monte Carlo modeling of reactive species’ distributions around a gold nanoparticle and the effects of nanoparticle proximity and clustering. *Int. J. Mol. Sci.* 2019, 20, 4280.

142. Sharma, R.A.; Plummer, R.; Stock, J.K.; Greenhalgh, T.A.; Ataman, O.; Kelly, S.; Clay, R.; Adams, R.A.; Baird, R.D.; Billingham, L. Clinical development of new drug–radiotherapy combinations. *Nat. Rev. Clin. Oncol.* 2016, 13, 627–642.

143. Cunningham, C.; De Kock, M.; Engelbrecht, M.; Miles, X.; Slabbert, J.; Van de Vooorde, C. Radiosensitization effect of gold nanoparticles in proton therapy. *Front. Public Health* 2021, 9, 9.

144. Peukert, D.; Kempsom, I.; Douglass, M.; Bezak, E. Gold nanoparticle enhanced proton therapy: A Monte Carlo simulation of the effects of proton energy, nanoparticle size, coating material, and coating thickness on dose and radiolysis yield. *Med. Phys.* 2020, 47, 651–661.
Int. J. Mol. Sci. 2022, 23, 1685

145. Le Tourneau, C.; Calugaru, V.; Jouffroy, T.; Rodriguez, J.; Hoffmann, C.; Dodger, B.; Moreno, V.; Calvo, E. A phase 1 trial of NBTXR3 nanoparticles activated by intensity-modulated radiation therapy (IMRT) in the treatment of advanced-stage head and neck squamous cell carcinoma (HNSCC). Journal of Clinical Oncology 2017, 35, 6080-6085.

146. Chen, T.; Ren, L.; Liu, X.; Zhou, M.; Li, L.; Xu, J.; Zhu, X. DNA nanotechnology for cancer diagnosis and therapy. Int. J. Mol. Sci. 2018, 19, 1671-1687.

147. Baig, M.M.F.A.; Lai, W.-F.; Akhtar, M.F.; Saleem, A.; Ahmed, S.A.; Xia, X.-H. DNA nanotechnology as a tool to develop molecular tension probes for bio-sensing and bio-imaging applications: An up-to-date review. Nano-Struct. Nano-Objects 2020, 23, 100523-100533.

148. Liu, Y.; Blanchfield, L.; Ma, V.P.-Y.; Andargachew, R.; Galior, B.; Liu, Z.; Evavold, B.; Salaika, K. DNA-based nanoparticle tension sensors reveal that T-cell receptors transmit defined pN forces to their antigens for enhanced fidelity. Proc. Natl. Acad. Sci. USA 2016, 113, 5610-5615.

149. Liu, Z.; Liu, B.; Ding, J.; Liu. J. Fluorescent sensors using DNA-functionalized graphene oxide. Anal. Bioanal. Chem. 2014, 406, 6885-6902.

150. De Maria Marchiano, R.; Di Sante, G.; Piro, G.; Carbone, C.; Tortora, G.; Boldrini, L.; Pietragalla, A.; Daniele, G.; Tredicine, M.; Cesario, A. Translational research in the era of precision medicine: Where we are and where we will go. J. Pers. Med. 2021, 11, 216.

151. Wang, X.; Zhang, W. Protein catenation enhances both the stability and activity of folded structural domains. Angew. Chem. Int. Ed. 2017, 129, 14173-14177.

152. Wu, W.-H.; Bai, X.; Shao, Y.; Yang, C.; Wei, J.; Wei, W.; Zhang, W.-B. Higher order protein catenation leads to an artificial antibody with enhanced affinity and in vivo stability. J. Am. Chem. Soc. 2021, 143, 18029-18040. https://doi.org/10.1021/jacs.1c06169.

153. Luo, M.-J.; Palmieri, M.; Riffkin, C.D.; Sakthianandeswaren, A.; Djawwi, T.M.; Hirokawa, Y.; Shuttleworth, V.; Segal, D.H.; White, C.A.; Nhu, D. Defining the susceptibility of colorectal cancers to BH3-mimetic compounds. Cell Death Dis. 2020, 11, 1–4.

154. Zhao, X.-Y.; Wang, X.-Y.; Wei, Q.-Y.; Xu, Y.-M.; Lau, A.Y. Potency and selectivity of SMAC/DIABLO mimetics in solid tumor therapy. Cells 2020, 9, 1012.

155. Goldsmith, S.J. Targeted radionuclide therapy: A Historical and Personal Review. Seminars in Nuclear Medicine; Elsevier: 2020, 50, 87–97.

156. Chan, T.G.; O’Neill, E.; Habjan, C.; Cornelissen, B. Combination strategies to improve targeted radionuclide therapy. J. Nucl. Med. 2020, 61, 1544–1552.

157. Schuerle, S.; Soleimany, A.P.; Yeh, T.; Anand, G.M.; Hüberli, M.; Fleming, H.E.; Mirkhani, N.; Qiu, F.; Hauert, S.; Wang, X. Synthetic and living microparticles for convection-enhanced nanoparticle transport. Sci. Adv. 2019, 5, eaav4803.

158. Ebner, D.K.; Frank, S.J.; Inaniwa, T.; Yamada, S.; Shirai, T. The Emerging Potential of Multi-Ion Radiotherapy. Front. Oncol. 2021, 11, 27.

159. Wurz, G.T.; Kao, C.-J.; Wolf, M.; DeGregorio, M.W. Tecemotide: An antigen-specific cancer immunotherapy. Hum. Vaccin. Immunother. 2014, 10, 3383–3393. https://doi.org/10.4161/hv.29836.

160. Gargett, T.; Abbas, M.N.; Rolan, P.; Price, J.D.; Gosling, K.M.; Ferrante, A.; Ruszkiewicz, A.; Atmosukarto, I.C.; Altin, J.; Parish, C.R.; et al. Phase I trial of Lipovaxin-MM, a novel dendritic cell-targeted liposomal vaccine for malignant melanoma. Cancer Immunol. Immunother. 2018, 67, 1461–1472. https://doi.org/10.1007/s00262-018-2207-z.

161. Young, C.; Schluep, T.; Hwang, J.; Eliasof, S. CRLX101 (formerly IT-101)-A novel nanophasmaeucal of campothecin in clinical development. Curr. Biochem. Compl. 2011, 7, 8–14. https://doi.org/10.2174/157340711795163866.

162. Ishikawa, T.; Kageyama, S.; Miyahara, Y.; Okayama, T.; Kokura, S.; Wang, L.; Sato, E.; Yagita, H.; Itoh, Y.; Shiku, H. Safety and antibody immune response of CHF-NY-ESO-1 vaccine combined with poly-ICLC in advanced or recurrent esophageal cancer patients. Cancer Immunol. Immunother. 2021, 70, 3081–3091. https://doi.org/10.1007/s00262-021-02892-w.

163. Yardley, D.A.; Brufsky, A.; Coleman, R.E.; Conte, P.F.; Cortes, J.; Glück, S.; Nabholtz, J.-M.A.; O’Shaughnessy, J.; Beck, R.M.; Ko, A. Phase II/III weekly nab-paclitaxel plus gemcitabine or carboplatin versus gemcitabine/carboplatin as first-line treatment of patients with metastatic triple-negative breast cancer (the tnAcity study): Study protocol for a randomized controlled trial. Trials 2015, 16, 1–10.

164. Weng, X.; Huang, X.; Li, H.; Lin, S.; Rao, X.; Guo, X.; Huang, P. First-line treatment with atezolizumab plus nab-paclitaxel for advanced triple-negative breast cancer: A cost-effectiveness analysis. Am. J. Clin. Oncol. 2020, 43, 340–348.

165. Ogino, M.; Fuji, T.; Kobuchi, Y.; Nakazawa, Y.; Takata, D.; Shirabe, K. Phase II study of nab-paclitaxel plus cyclophosphamide plus trastuzumab neoadjuvant chemotherapy in early HER-2-positive breast cancer. Anticancer Res. 2021, 41, 3899–3904.

166. Canetta, E. Current and future advancements of raman spectroscopy techniques in cancer nanomedicine. Int. J. Mol. Sci. 2021, 22, 13141.

167. Maiti, K.K.; Samanta, A.; Vendrell, M.; Soh, K.-S.; Olivo, M.; Chang, Y.-T. Multiplex cancer cell detection by SERS nanotags with cyanine and triphenylmethine Raman reporters. Chem. Commun. 2011, 47, 3514–3516.

168. Krishnan, T.; Wang, H.-N.; Vo-Dinh, T. Smartphone-based device for colorimetric detection of MicroRNA biomarkers using nanoparticle-based assay. Sensors 2021, 21, 8044.

169. Shao, J.; Liang, R.; Ding, D.; Zheng, X.; Zhu, X.; Hu, S.; Wei, H.; Wei, B. A Smart multifunctional nanoparticle for enhanced near-infrared image-guided photothermal therapy against gastric cancer. Int. J. Nanomed. 2021, 16, 2897.
170. Tang, C.; He, Z.; Liu, H.; Xu, Y.; Huang, H.; Yang, G.; Xiao, Z.; Li, S.; Liu, H.; Deng, Y. Application of magnetic nanoparticles in nucleic acid detection. *J. Nanobiotechnol.* 2020, 18, 1–19.

171. Schneider-Futschik, E.K.; Reyes-Ortega, F. Advantages and disadvantages of using magnetic nanoparticles for the treatment of complicated ocular disorders. *Pharmaceutics* 2021, 13, 1157–1173.

172. Wang, C.; Lu, J.; Zhou, L.; Li, J.; Xu, J.; Li, W.; Zhang, L.; Zhong, X.; Wang, T. Effects of long-term exposure to zinc oxide nanoparticles on development, zinc metabolism and biodistribution of minerals (Zn, Fe, Cu, Mn) in mice. *PLoS One* 2016, 11, e0164434–e0164448.

173. Yada, R.; Maenaka, K.; Miyamoto, S.; Okada, G.; Sasakura, A.; Ashida, M.; Adachi, M.; Sato, T.; Wang, T.; Akasaka, H. Real-time in vivo dosimetry system based on an optical fiber-coupled microsized photostimulable phosphor for stereotactic body radiation therapy. *Med. Phys.* 2020, 47, 5235–5249.