Propagating left/right asymmetry in the zebrafish embryo: one-dimensional model

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ABSTRACT During embryonic development in vertebrates, left-right (L/R) asymmetry is reliably generated by a conserved mechanism: a L/R asymmetric signal is transmitted from the embryonic node to other parts of the embryo by the L/R asymmetric expression and diffusion of the TGF-β related proteins Nodal and Lefty via propagating gene expression fronts in the lateral plate mesoderm (LPM) and midline. In zebrafish embryos, Nodal and Lefty expression can only occur along 3 narrow stripes that express the co-receptor one-eyed pinhead (oep): Nodal along stripes in the left and right LPM, and Lefty along the midline. In wild-type embryos, Nodal is only expressed in the left LPM but not the right, because of inhibition by Lefty from the midline; however, bilateral Nodal expression occurs in loss-of-handedness mutants. A two-dimensional model of the zebrafish embryo predicts this loss of L/R asymmetry in oep mutants [15]. In this paper, we simplify this two-dimensional picture to a one-dimensional model of Nodal and Lefty front propagation along the oep-expressing stripes. We represent Nodal and Lefty production by step functions that turn on when a linear function of Nodal and Lefty densities crosses a threshold. We do a parameter exploration of front propagation behavior, and find the existence of pinned intervals, along which the linear function underlying production is pinned to the threshold. Finally, we find parameter regimes for which spatially uniform oscillating solutions are possible.

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1. Introduction

In vertebrates, organs such as the heart and brain develop in an invariant left-right (L/R) asymmetric fashion, with situs inversus, or complete organ mirror-reversal, being a rare occurrence [1, 2, 3]. The best-understood mechanism by which L/R bias is first established is that special cilia around the embryonic node drive a leftwards fluid flow [4]. Disrupting, or reversing this flow results in randomization [5], or reversal [6] of organ asymmetry. The cilia appear to be conserved in vertebrates [7], but it is controversial whether the nodal flow mechanism is decisive in all vertebrates, since asymmetry of some other origin (involving electrical potentials due to polarized gap junctions) is observable before the cilia even begin moving, and this may determine organ asymmetry in Xenopus [8].

Our concern in this paper is with the subsequent propagation of this L/R signal from the node to the rest of the embryo. It is first transmitted to the lateral plate mesoderm (LPM), resulting in L/R asymmetric expression of the TGF-β related signaling molecules Nodal and Lefty [9, 10]. Nodal is expressed exclusively in the left LPM, and Lefty in the midline, along gene expression fronts that propagate from the posterior to the anterior of the embryo from approximately the 12 to 20 somite stages [11]. This propagation depends on the diffusion of Nodal and Lefty, as well as their production rates which are promoted or inhibited by these same molecules. Subsequently, downstream genes (such as Pitx2) are activated and direct L/R asymmetric organogenesis [12].

Nodal and Lefty constitute a classic activator-inhibitor system that is highly conserved among vertebrates [1]. Such a model was introduced and studied for the initiation of Nodal and Lefty expression in the LPM and midline, respectively, at the level of the node in the mouse [13]. This was spatially one-dimensional, representing the L/R axis, and did not attempt to model the longitudinal propagation of the L/R signal.

In this paper, we extend the ideas in this model to the zebrafish embryo, in which Nodal and Lefty gene expression only occurs in cells carrying the co-receptor one-eyed pinhead (oep) found on narrow stripes 3-5 cells wide in the midline and LPM [14]. (Note that the Nodal analogue in the zebrafish is called “Southpaw”; but we will call it “Nodal” for consistency with the terminology of Ref. [13]). An advantage of this system is that it is quasi-one-dimensional in the longitudinal direction, because the most important variables are Nodal and Lefty production on these 3 stripes, and the propagation of their gene expression fronts is clearly unidirectional in the posterior-to-anterior direction.
A two-dimensional model of the Nodal/Lefty system in the zebrafish embryo has been set up and simulated [14, 15], as we review in Section 2. In Sections 3 and 4, we work through the consequences of a one-dimensional idealization of it, representing not the L/R axis (as in [13]) but the anterior-posterior axis. We idealize Nodal and Lefty production to have step-function turn-ons, when a threshold function linear in their concentrations crosses a threshold. We classify all possible behaviors in the limit that Nodal is unaffected by Lefty, and demonstrate the existence of intervals along which the Lefty profile must be pinned to that of Nodal. In Section 5, we find parameter regimes that allow uniform spatial oscillations to occur.

2. Two-dimensional model

In this section, we outline what would be a minimal model for the two-dimensional Nodal/Lefty system. This will motivate the simplification we make, in section 3, to a one-dimensional model of similar form. The degrees of freedom in the model we use here are only the concentrations $N(r, t)$ of Nodal and $L(r, t)$ of Lefty, as found unbound in the extracellular fluid. In this section we let $r$ represent a two-component position $(x, y)$ on the embryo, which is practically a flat two-dimensional layer enveloping the yolk cell; in later sections on the one-dimensional model it will be replaced by the variable $y$. Also, $t$ is the time.

The geometry is idealized so that the three oop-expressing stripes extend indefinitely for $y > 0$, and are straight, parallel, and of unvarying widths, as shown in Fig. 1. That means that, away from the baseline at $y = 0$, the model system has a translational invariance. That permits one to talk in a mathematically precise way about a limiting behavior in the model, in particular uniform motion of a front. Whether this is pertinent to the actual embryo depends on whether initial transients persist for a short time compared to the stages in which the L/R signal propagates, and for a short distance compared to the inter-stripe spacing or to the embryo's length, which we know to be the case [11].

Lefty is produced at a rate $s_L(r, t)$ [concentration per unit time per unit area] which can be nonzero only at points lying within the midline stripe, since only those cells can produce Lefty. The function $s_L(r, t)$ depends, in our simplified model, only on the local, instantaneous concentrations of Nodal and Lefty; the exact functional dependence is specified in Sec. 2.2. More precisely, the production function depending on $N(r, t)$ and $L(r, t)$ should rather be the transcription rate of the Lefty mRNA, while the source rate $s_L(r, t)$ of Lefty protein is in turn proportional to the mRNA concentration, and thus should lag the production function we use by roughly the mRNA lifetime. (Indeed, a model including that lag was successfully fitted to quantitative data in [14], Chapter 3.) But in this paper, we assume that the source rate is proportional to the instantaneous production function; that is equivalent to assuming the Lefty mRNA is transcribed copiously and decays quickly compared to other time scales.

Similarly, Nodal is produced at a rate $s_N(r, t)$ which is nonzero only within stripes in the left LPM and right LPM, being described by a production function of the same form as that of Lefty in the midline, and with the same production function in the right and left LPM, so no prior asymmetry is assumed. As with Lefty, we have made an approximation of short-lived mRNA so that the mRNA content is not treated as an independent variable.

In the case of Nodal, there is a second place where $s_N(r, t)$ is nonzero, which are the two tailbuds, located as shown in Figure 1. This production is not modulated by $N(r, t)$ or by $L(r, t)$, and may be assumed constant in time for simplicity (in reality it gradually diminishes [14]). The tailbud production has some left-right asymmetry, which in our model is responsible for all downstream asymmetries. The origin of the asymmetry is imagined to be in the nearby Rupffer's vesicle [16] (the organ in which the special cilia are found in zebrafish).

2.1 Diffusion behavior

Once produced, Nodal and Lefty are assumed to be passive, noninteracting densities that satisfy a diffusion equation with diffusion constants $D_N$ and $D_L$, respectively. Furthermore, they are assumed to have degradation rates, respectively $\tau_N^{-1}$
and $\tau_L^{-1}$. Although one expects the two signaling molecules to have similar values of these parameters, we allow them to be different in general, because (as we derive in the results sections) this inequality induces qualitatively different behaviors. (The differential equations incorporating these parameters are exactly like Eqs. (3.2a) and (3.2b) of the one-dimensional model, except the second derivative with respect to $y$ is replaced by the two-dimensional divergence.)

Characteristic length and velocity scales can be constructed from the diffusion constant and degradation times:

\[ l_N = \sqrt{D_N \tau_N}; \quad (2.1) \]
\[ v_N = \frac{D_N}{\tau_N}; \quad (2.2) \]

and we define $l_L$ and $v_L$ similarly. These scales determine the typical range in space that the signal molecules travel, and the typical speed of the production fronts we shall study.

### 2.2 Conditions for Nodal and Lefty production

A key property determining the expression (and production) of either Nodal or Lefty is promoted by the presence of Nodal and inhibited by the presence of Lefty. Thus, both production functions $s_N(r, t)$ and $s_L(r, t)$ are increasing as a function of $N$ and decreasing as a function of $L$. As we explain next, one can consider several versions of our model with different functional forms for the production functions.

Let us first lay out how production is actually regulated. Along the LPM and midline stripes, Nodal and Lefty protein bind to the plasma membrane with the help of oep co-receptors [14]; we may say “oep” is in one of three states: unbound, Nodal-bound or Lefty-bound. We assume that only a negligible fraction of all Nodal and Lefty produced are thus sequestered by the receptors, so that uptake or release of the signaling molecules is not included in $s_N(r, t)$ and $s_L(r, t)$.

In turn, Nodal-bound oep induces a signaling cascade that promotes transcription of Nodal (in the LPM) or Lefty (on the midline). The fraction of Nodal-bound oep, $\phi(N, L)$, is

\[ \phi(N, L) = \frac{N}{k^{-1} + N + L} \quad (2.3) \]

The oep receptors on the midline are believed to be identical to those on the LPM stripes, so we use the same function $\phi(N, L)$ in both stripes with the same equilibrium constant $k$ (where $k^{-1}$ has the units of concentration).

The Nodal production function $s_N$ is assumed to be proportional to a Hill function

\[ f_N(\phi) = \frac{1}{1 + (\phi_{sN}/\phi)^{n_H}}. \quad (2.4) \]

This is a rounded step from 0 to 1 centered at a threshold parameter $\phi_{sN}$; the step gets sharper as the Hill exponent $n_H$ grows large. A microscopic rationalization for the form in Eq. (2.4) would be that the Nodal gene’s regulatory region cooperatively binds $n_H$ copies of the signal or transcription factor induced by Nodal-bound oep. (Ref. [13] used a different production function than (2.4) that also approximates a step function with a parameter controlling the sharpness.)

The actual production rate is

\[ s_N(r, t) = s_{N0} f_N(\phi(N, L)) \quad (2.5) \]

where $s_{N0}$ is the maximum, or saturated, production rate. A similar function $f_L(\phi)$, with (it is expected) a rather different threshold $\phi_{sL}$, and a saturated production rate $s_{L0}$, controls Lefty production.

Part of the motivation for including the intermediate function $\phi$ in the model is to represent mutants in which oep is under- or over-expressed. That would have the effect of multiplying $\phi$ in (2.4) by a constant, or equivalently of dividing $\phi_{sN}$ and $\phi_{sL}$ by that constant [14].

We adopt two simplifying assumptions, either of which could be made independently of the other. The first pertains to the threshold for turning production on or off. In place of the nonlinear $\phi$ function (2.3), we follow Ref. [13] and instead use two threshold functions linear in the concentration:

\[ C_N(N, L) = C_{NN}N - C_{NL}L \quad (2.6a) \]
\[ C_L(N, L) = C_{LN}N - C_{LL}L \quad (2.6b) \]

where $N = N(r, t)$ and $L = L(r, t)$ are Nodal and Lefty concentrations, respectively, and $C_{NN}, C_{NL}, C_{LN},$ and $C_{LL}$ are constants. Production of $N$ or $L$ respectively turns on when this function exceeds a threshold parameter $C_{NN}$ or $C_{LL}$, which should be positive, since a certain concentration of Nodal is needed to turn on either production, even in the absence of Lefty. We set $C_{NN} = C_{LL} = 1$ without loss of generality by scaling the four coefficients in Eqs. (2.6).

The relation of Eqs. (2.6) to the $\phi$ function in (2.3) is that the condition to be over threshold, $\phi > \phi_{sN}$ or $\phi > \phi_{sL}$, is (by inserting Eq. (2.3)) mathematically equivalent to

\[ k(\phi_{sN} - 1)N - kL > 1; \quad (2.7a) \]
\[ k(\phi_{sL} - 1)N - kL > 1. \quad (2.7b) \]

Clearly, we can (roughly) identify the coefficients in (2.7) with those in (2.6), in particular this viewpoint implies $C_{NL} = C_{LL}$.

Note that although the condition to be at threshold is captured exactly by the linear functions, away from threshold the production will depend on $N$ and $L$ in a different way when $n_H < \infty$. The linear form of Eqs. (2.6a) and (2.6b) will be more convenient mathematically.

The second simplification we make is to assume $n_H \to \infty$ in (2.4), i.e. a step-function turn-on of the production. In this case, Nodal production $s_N(r, t)$ and Lefty production $s_L(r, t)$ are given by:
The model initializes with zero Nodal or Lefty concentrations in the domain (Fig. 1), and with the 2 tailbuds producing Nodal that diffuses to the base of the 3 stripes. The left tailbud produces more than the right, which is responsible for all downstream asymmetries. Successful initiation at the base of the stripes requires that the Nodal produced in the tailbuds is sufficient to start self-sustaining Nodal production in the left LPM, but not the right LPM; production begins when thresholds given in (2.8) are exceeded. The times at which this occurs differs for all three stripes depending on the position of the tailbuds (closer to or farther from the midline, in a L/R symmetric way). Lefty, once produced on the midline, diffuses to the left and right LPM in equal amounts. Thus, the threshold function (2.6a) is constrained such that Nodal production turns off in the right LPM due to Lefty inhibition, but stays on in the left because of greater $N$ there. This occurs in simulations for widely-varying parameter sets, for differences in tailbud production rates of 30%. As this asymmetry is decreased, the coefficients in (2.6a), as well as the geometry of the problem, must be tuned more finely for successful initiation to occur.

Experiments in zebrafish find that after an initial phase the wavefronts of Nodal and Lefty gene expression proceed at a fixed speed [14, 11], and indeed this is reproduced in simulations: once initiated in a self-sustaining way, Nodal production on the left LPM switches on from the base upwards, and the leading edge of production moves at an asymptotically fixed rate along the stripe, followed by a front of Lefty production, moving at the same asymptotic rate on the midline.

The main role of Nodal and Lefty in this model is to transmit the L/R asymmetric signal from the tailbud. Lefty in particular acts as a “barrier” preventing initiation of Nodal production in the right stripe: the Lefty concentration there is sufficiently high that Nodal diffusing from the left LPM is insufficient to initiate production there. Another necessary condition for successful propagation is that the function (2.6b) must not fall below 1 along the midline, or else Nodal production will cease, and Nodal produced in the left LPM will not be prevented from diffusing to the right LPM and initiating production further up the right stripe, leading to loss of L/R asymmetric propagation. Indeed, in Lefty knockout mutants, bilateral Nodal expression is observed [11, 13].

We intended to model the space and time-dependent behaviors in this model to, ultimately,

(i) confirm the correctness of the basic picture by its qualitative agreement with experiments;
(ii) explain, and predict, the phenotypes of various mutations involving this system;
(iii) characterize the robustness of how the initial bias in the tailbud is amplified (as is manifested in the error rate);
(iv) show how the numerical values of the various parameters can be inferred (or at least bounded) on the basis of experiments.

However, while it is understood in principle how we get a moving front, the mathematical formulation involves convolutions in two dimensions and does not include closed forms of simple functions. Furthermore, simulations suggested various possible regimes in which production might turn off again after the initial passage of a pulse, or in which the midline Lefty production had a gradual onset in space, much less sharp than the Nodal front. In order to analytically represent the functional form, and to comprehensively classify all the regimes of the asymptotic behavior, we turned to a one dimensional model, which is the main focus of the rest of this paper.

### 2.3 Behavior of the two-dimensional model and comparison with experiments

The partial differential equations for Nodal and Lefty are:

\[
\begin{align*}
\phi_N(r, t) &= 0, \quad C_N < 1 \\
0 \leq s_N(r, t) < s_{NO}, \quad C_N = 1 \\
s_N(r, t) &= s_{NO}, \quad C_N > 1; \tag{2.8a}
\end{align*}
\]

and

\[
\begin{align*}
\phi_L(r, t) &= 0, \quad C_L < 1 \\
0 \leq s_L(r, t) < s_{LO}, \quad C_L = 1 \\
s_L(r, t) &= s_{LO}, \quad C_L > 1. \tag{2.8b}
\end{align*}
\]

and

\[
\begin{align*}
\frac{\partial N(y, t)}{\partial t} &= D_N \frac{\partial^2 N(y, t)}{\partial y^2} - \frac{1}{\tau_N} N(y, t) + s_N(y, t) \tag{3.2a}
\end{align*}
\]

#### 3. One-dimensional model

The 2D model may be simplified to 1D by making the following simplifications. Since Nodal and Lefty expression occur on narrow, parallel stripes, we may replace the finite width of each stripe by a 1D line. Also, the concentration profile on one stripe is transmitted (with a diffusive lag) to another through the intervening space; this space can be replaced by lines as well, and we end up with a 5-line 1D model with diffusion between stripes given by

\[
N_i(y, t) = k_D N_{i-1}(y, t - \Delta t) + k_D N_{i+1}(y, t - \Delta t) + ... \tag{3.1}
\]

where $N_i$ and $N_{i+1}$ are Nodal concentrations on adjacent stripes, and $k_D$ and $\Delta t$ describe the rate of diffusion.

In this paper, we will only focus on the front propagation of Nodal and Lefty gene expression in the steady state, in which case we further neglect inter-stripe diffusion and thus simplify the system to 1D.

#### 3.1 Diffusion equations

The partial differential equations for Nodal and Lefty are:

\[
\frac{\partial N(y, t)}{\partial t} = D_N \frac{\partial^2 N(y, t)}{\partial y^2} - \frac{1}{\tau_N} N(y, t) + s_N(y, t) \tag{3.2a}
\]
The middle of any interval of production looks locally like a particular, since crossed and production turns on or off. in such a solution is the position "pinned" intervals. However, the threshold condition becomes an equality the diffusion equation can no longer be used to find the solution. Thus, the key parameter in such a solution is the position y at which the threshold is crossed and production turns on or off.

Surprisingly, it is also generically possible that the threshold function is pinned at 1 throughout a finite interval (see Sec. 4.2). Within such an interval, the production function is indefinite (the middle case in Eqs. (2.8a) and (2.8b)) and the diffusion equation can no longer be used to find the solution. However, the threshold condition becomes an equality interval and gives the solution in such intervals. The complications in our results have to do mostly with handling such “pinned” intervals.

The simplest possible solution is for constant, homogeneous production, e.g. \( s_N(y, t) = s_{N0} \) for all \((y, t)\) for Nodal. The solution is \( N(y, t) = N_0 \), or \( L(y, t) = L_0 \) in the analogous case for Lefty, where

\[
\begin{align*}
N_0 & \equiv \tau_N s_{N0}; & (3.3a) \\
L_0 & \equiv \tau_L s_{L0}. & (3.3b)
\end{align*}
\]

The middle of any interval of production looks locally like a piece of this homogeneous system, so it is not surprising that \( N_0 \) and \( L_0 \) serve as a reference level for all solutions. In particular, since \( s_N(y, t) \leq s_{N0} \) everywhere, any solution must have \( N(y, t) \leq N_0 \) everywhere, and similarly for \( L(y, t) \).

3.2 The traveling wavefront solution

Experiments show the Nodal front is typically ahead of the Lefty front [11, 14]. If it is sufficiently far ahead, the Lefty concentration there is negligible, and it is sufficient to consider a Nodal-only situation, in which the front of Nodal production moves at constant speed \( v \):

\[
s_N(y, t) = \begin{cases} 
  s_{N0}, & y < vt \\
  0, & y > vt.
\end{cases}
\]

The front shows a constant shape to a viewer traveling at velocity \( v \), i.e. \( N(y, t) = N(y - vt) \). When this is inserted into Eq. (3.2a), it becomes an ordinary, second-order, linear differential equation, which is solved by an exponential:

\[
N(y - vt) = \begin{cases} 
  N_0[1 - g_N e^{\kappa_N(y-vt)}], & y - vt < 0 \\
  N_0(1 - g_N e^{-\kappa_N(y-vt)}), & y - vt > 0.
\end{cases}
\]

Substituting this ansatz into Eq. (3.2a), we find that \( \kappa_N \) and \( \kappa'_N \) are given by

\[
\begin{align*}
\kappa_N^2 + \frac{v}{D_N} \kappa_N - \frac{1}{L_N} &= 0 & (3.6a) \\
\kappa_N'^2 - \frac{v}{D_N} \kappa_N' - \frac{1}{L_N} &= 0 & (3.6b)
\end{align*}
\]

By matching the boundary conditions at \( y = vt = 0 \), we find \( g_N \) is given by

\[
g_N = \frac{\kappa'_N}{\kappa_N + \kappa'_N}
\]

Essentially, this means that the shape of the concentration profile depends on the relative magnitudes of \( v \) and \( \sqrt{\kappa} \). Physically, for \( v > 0 \) and \( v \gg (\kappa) \), the wavefront is traveling forward too fast for there to be much diffusion from the producing region to the non-producing region, and so the concentration at the front is low (\( g_N \rightarrow 1 \)); conversely, for \( v < 0 \) and \( |v| \gg \kappa \), the concentration at the front is high for the same reason. (It is reminiscent of the Doppler effect, except that the Nodal signal spreads by the diffusion equation rather than the wave equation.)

There is in fact a symmetry relating solutions with \( v > 0 \) and \( v < 0 \), whereby the profiles of an advancing front of speed \( v \) and a retreating front (reflected in \( y \)) of speed \( -v \) add up to a uniform profile, because adding the production of an advancing front and that of a retreating front simply obtains a uniform producing line. Furthermore, when \( v = 0 \) the height of the front is simply \( \frac{1}{2}N_0 \), and the stationary concentration profile simplifies to:

\[
N(y) = \begin{cases} 
  N_0(1 - \frac{1}{2}e^{\kappa_L(y-\Delta y)}), & y < 0 \\
  \frac{1}{2}N_0 e^{-\kappa_L(\Delta y)}, & y > 0
\end{cases}
\]

(Notice that we set \( y = 0 \) to be the position of the front.) Since the coefficients in the threshold equations (2.6a) and (2.6b) are in general different, the Nodal wavefront is in general displaced by a distance \( \Delta y \) from the Nodal one. We have

\[
L(y-vt) = \begin{cases} 
  L_0[1 - g_L e^{\kappa_L(y-vt+\Delta y)}], & y - vt < -\Delta y \\
  L_0(1 - g_L e^{-\kappa_L(y-vt+\Delta y)}), & y - vt > -\Delta y
\end{cases}
\]

where \( \Delta y > 0 \) signifies that the Nodal front is ahead of the Lefty front, and \( \kappa_L, \kappa'_L \) and \( g_L \) are defined analogously to Eqs. (3.6) and (3.7). Finally, to find \( v \) and \( \Delta y \), we observe that \( N = N_0(1 - g_N) \) at the Nodal front, and \( L = L_0(1 - g_L) \) at the Lefty front, and substitute (3.5) and (3.9) into the threshold functions (2.6); assuming that \( \Delta y > 0 \):

\[
C_{NN}N_0(1 - g_N) - C_{NL}L_0(1 - g_L)e^{-\kappa_L\Delta y} = 1
\]

(3.10a)

\[
C_{LN}N_0[1 - g_N e^{-\kappa_N\Delta y}] - C_{LL}L_0(1 - g_L) = 1
\]

(3.10b)

from which we can solve (non-trivially) for \( v \) and \( \Delta y \).
3.3 Nondimensionalization

Certain combinations of parameter changes are trivial, in the sense that the solutions look the same apart from rescalings of the distance, time, or concentrations. Since we are faced with the difficulty of exploring a large parameter space, we wish to discover the minimum number of nontrivial parameters. To this end, we will scale all variables and parameters in the problem so as to make them dimensionless.

First, we introduce the scaled Nodal and Lefty concentrations

\[ \tilde{N}(y) \equiv \frac{N}{N_0}, \quad \tilde{L}(y) \equiv \frac{L}{L_0}, \]

so that \( 0 \leq \tilde{N}(y), \tilde{L}(y) \leq 1. \)

Correspondingly, \( \tilde{C}_{NN}, \tilde{C}_{NL}, \tilde{C}_{LN} \) and \( \tilde{C}_{LL} \) are simply the coefficients in the (scaled form of the) threshold functions (2.6a) and (2.6b):

\[ \tilde{C}_{NN} \equiv C_{NN}N_0; \quad \tilde{C}_{NL} \equiv C_{NL}L_0; \quad \tilde{C}_{LN} \equiv C_{LN}N_0; \quad \tilde{C}_{LL} \equiv C_{LL}L_0. \]

For example, Nodal production turns on when \( \tilde{C}_{NN}\tilde{N} - \tilde{C}_{NL}\tilde{L} > 1 \), and similarly for \( \tilde{L} \). The parameters (3.12) are pertinent even in a spatially uniform situation.

The remaining parameters relate to time or length scales. We scale length and time such that the parameters for Nodal become unity:

\[ \tilde{l}_L \equiv l_L/l_N; \quad \tilde{v}_L \equiv v_L/v_N; \quad \tilde{v} \equiv v/v_N. \]

The last two parameters are relevant (in a moving steady state) if and only if \( v \neq 0 \). Implicitly, Eqs. (3.13) also nondimensionalize the time scale:

\[ \tilde{\tau}_L \equiv \tilde{l}_L/\tilde{v}_L. \]

In total, we have seven nontrivial parameters for the one-dimensional problem, defined in Eqs. (3.12) and (3.13). By rescaling space and time with \( y \to \frac{y}{\tilde{l}_N} \) and \( t \to \frac{1}{\tilde{\tau}_N} \) we find that Eqs. (3.2) become

\[ \begin{align*}
\tilde{N}(y,t) & = \tilde{C}_{NN}N_0 - \tilde{C}_{NL}\tilde{L}(y,t) + \tilde{s}_N(y,t) \\
\tilde{L}(y,t) & = \tilde{l}_L\tilde{v}_L\tilde{C}_{LL}L_0 + \frac{1}{\tilde{l}_L\tilde{v}_L}\tilde{C}_{LN}N_0 + \tilde{s}_L(y,t)
\end{align*} \]

where \( 0 \leq \tilde{s}_N(y,t) \leq 1 \) and \( 0 \leq \tilde{s}_L(y,t) \leq \frac{1}{\tilde{l}_L}. \)

4.3 The \((\tilde{N}, \tilde{L})\) plane: zero dimensional case

In Sec. 3.2, asymptotically in either direction both \( N \) and \( L \) approach constant, uniform values. Therefore, in order to classify possible wavefront solutions, it is helpful first to classify all possible uniform solutions. Fig. 2 shows the \((\tilde{N}, \tilde{L})\) phase plane with lines representing the \( \tilde{N} = 0 \) and \( \tilde{L} = 0 \) isoclines. The most important feature of these diagrams is the relation of the two isoclines to each other and to the bounding lines: this determines the possible steady states.

There are 4 qualitative cases for the relation of \( \tilde{N} \) to \( \tilde{L} \):

1. \( N \)-isocline is to left of \( L \)-isocline
2. \( L \)-isocline crosses \( N \)-isocline left to right
3. \( L \)-isocline is to left of \( N \)-isocline
4. \( N \)-isocline crosses \( L \)-isocline left to right.

(There are other mathematical cases in which one of the isoclines does not pass through the square at all, but that is ob-
viously to be discarded, since there would be no way ever to produce the corresponding signal.)

These phase planes are generalizations of the wild-type scenario: as noted after Eq. (2.7), if we justify the linear threshold equations (2.6) from the single-binding site receptor behavior (2.3), we necessarily find \( \tilde{C}_{NL} = \tilde{C}_{LL} \); the geometrical expression of this on the \(( \tilde{N}, \tilde{L} )\) plane is that the two isoclines cross on the negative \( L \) axis, hence only cases (a) and (c) could be realized. However, our purpose here is to study the general behavior of these equations that could arise in any embryonic system with propagating Nodal and Lefty fronts (since this is conserved in all vertebrates), and perhaps even mathematically equivalent equations having a quite different biological interpretation. In general, binding of Nodal and Lefty might be cooperative at the receptor; furthermore, gene regulation further downstream might be cooperative. Thus, the general form of the isoclines is nonlinear and we expect that all these topologies are imaginable.

There is always a stable fixed point at \( \tilde{N} = \tilde{L} = 0 \); this is the only fixed point in case (iii), which is thus trivial. If the \( \tilde{N} \)-isocline and the \( \tilde{L} \)-isocline both intersect the upper border, that is if \( \tilde{C}_{NN} - \tilde{C}_{NL} > 1 \) and \( \tilde{C}_{LN} - \tilde{C}_{LL} > 1 \), we have a stable fixed point with both \( N \) and \( L \) saturated, which underlies the basic wavefront behavior of Sec. 3.2 above.

However, if the \( \tilde{L} \)-isocline intersects the right edge, to the right of the \( \tilde{N} \)-isocline, i.e. when \( \tilde{C}_{LN} - \tilde{C}_{LL} < 1 \) as may happen in either case (i) or case (ii), then the stable fixed point at \( \tilde{N} = 1 \) exists at a less than saturated value, which we will define \( \tilde{L}_s \):

\[
\tilde{L}_s = \frac{\tilde{C}_{LN} - 1}{\tilde{C}_{LL}}
\]

(3.16)

It is not obvious that \( \tilde{L} \) being pinned at \( \tilde{L}_s < 1 \) exists in the case with spatial variation, but we will in fact show in Sec. 4.2 that analogous “pinned” intervals arise in 1D traveling wave solutions.

Finally, in case (iv), there is a fixed point at which both \( \tilde{N} \) and \( \tilde{L} \) are less than saturated; this may be stable, but also may be unstable to periodic oscillations around the fixed point, as elaborated in Sec. 5.

4. Results: steady-state fronts

To explore the large parameter space of the 1D model, we will consider special cases; the aim is a taxonomy of the possible behaviors of the model, with the hope that any general case will be qualitatively similar to one of these behaviors seen in the special cases. In particular, we concern ourselves with the steady state limiting behaviors, after all transients have died out. Any nonzero Nodal and Lefty wavefronts will then be traveling at identical speed \( v \).

4.1 \( \tilde{C}_{NL} = 0 \) and \( v = 0 \)

For the largest part of our story, we consider the case that \( \tilde{C}_{NL} = 0 \), that is, Nodal is unaffected by Lefty (i.e. Nodal is an autonomous variable). We first solve for the behaviors of the one-component system in which Nodal activates Nodal, which already has moving front solutions; then, we treat the Nodal concentration \( N(y, t) \) as if it were externally imposed and solve for another one-component problem representing the Lefty concentration field. The reasons for choosing what seems to be a major simplification are (i) the reverse approximation, in which Lefty is autonomous, has only the trivial solution with Lefty off (since Lefty only inhibits); (ii) experimentally, the Nodal front leads and the Lefty front follows; thus, it is plausible that the inhibition from Lefty has no important effect on the Nodal front (serving only to prevent the initiation of a front on the other side); (iii) in Section 3.4 we see that only the topology of the \(( N, L )\) plot really matters for the dependence on coefficients \( C_{ij} \).

The second simplification we make is to set \( v = 0 \). As mentioned, the Nodal and Lefty fronts are traveling at equal speed in the steady state, and are thus stationary in the comoving frame. In fact, the only difference between the \( v = 0 \) and \( v \neq 0 \) cases is the steepness of their exponential profiles at the front. This does not qualitatively affect the results we present below.

With \( \tilde{C}_{NL} = 0 \) and \( v = 0 \), the Nodal concentration takes the form given by (3.8), or in dimensionless form,

\[
\tilde{N}(y) = \begin{cases} 
1 - \frac{1}{2} e^y & , \ y < 0 \\
\frac{1}{2} e^{-y} & , \ y > 0 
\end{cases}
\]

(4.1)

and our 7-dimensional parameter space reduces to a 3-dimensional one, with parameters \( \tilde{L}_s, \tilde{C}_{LL}, \) and \( \tilde{L}_s \) (which is more convenient for our purposes than \( \tilde{C}_{LN} \)). Our goal is to classify all the possible types of behavior of the Lefty profile as these parameters are varied. Note that \( \tilde{C}_{NN} \) and \( \tilde{C}_{LN} \) are always greater than 1, or else \( N \) and \( L \) will never have their production turned on. Also, let the position where Lefty production turns on be \( y = y_L \); it may be on either side of the independent Nodal front \( y = 0 \).

4.2 “Pinned” intervals

It soon became apparent that the behavior of \( L \) did not only consist of intervals of full production \( s_L = s_{0L} \), where \( C_L(y) > 1 \), and zero production, where \( C_L(y) < 1 \), but also intervals where the threshold function is pinned at \( C_L(y) = 1 \). The clearest example of this is to consider what happens when \( \tilde{L}_s < 1 \) as \( y \to -\infty \), where our background Nodal profile goes to a limit of \( \tilde{N} \to 1 \). Since we want \( L \) to be produced somewhere, \( \tilde{C}_{LN} \equiv \tilde{C}_{LN} \tilde{N}_0 > 1 \) necessarily (this is (2.6b) with \( L = 0 \)). As \( y \to -\infty \) and \( \tilde{N} \to 1 \), \( L(y) = 0 \) is mathematically inconsistent, and so Lefty production must turn on. Now, by definition \( \tilde{L}_s \) is the amount of \( L \) that turns
off its own production when \( N = N_0 \), so \( \tilde{L}_s < 1 \) means that before \( \tilde{L} \to 1 \), Lefty will turn off its own production again. This is a “paradox” in which Lefty production cannot be fully on or fully off, and it is only resolved if we let the threshold function be pinned at 1, i.e. \( C_L(y \to -\infty) \equiv 1 \), such that \( 0 < \tilde{s}_L < 1 \). Constraining \( C_L(y) \) means that \( \tilde{L}(y) \) is completely determined by \( \tilde{N}(y) \) in this pinned interval:

\[
\tilde{L}(y) = \frac{1}{C_{LL}} \left( \tilde{C}_{LN} \tilde{N}(y) - 1 \right) \tag{4.2}
\]

Now, we explicitly work out the possible behaviors of \( L \) when \( \tilde{L}_s, \tilde{L}_L \) and \( C_{LL} \) are varied. \( L \) will be described in terms of the types of production occurring along the line, from \(-\infty\) to \( \infty \). Full production is denoted “1”, zero production denoted “0”, and the pinned interval denoted “p”. For example, the background Nodal profile is characterized as \( \{1,0\} \), which is shorthand for an interval of full production adjoining one of zero production. In the following, we split up the cases into \( \tilde{L}_s < 1 \) and \( \tilde{L}_s > 1 \). We find that the most important parameter is the ratio of length scales, \( \tilde{l}_L \), which determines how various pinned intervals arise in the behavior of Lefty. Note that the value of \( \tilde{v}_L \) does not enter at all.

### 4.3 Classification of 1D model

#### 4.3.1 Case \( \tilde{L}_s < 1 \): \( \{p,0\} \) and \( \{p,1,0\} \)

As already shown, there is a pinned interval extending to \( y \to -\infty \), where \( L \) is completely determined. The production \( s_L(y) \) is also completely determined, and is derived as follows. Substituting (4.2) back into (3.2b):

\[
\frac{D_L}{C_{LL}} \frac{d^2\tilde{N}(y)}{dy^2} - \frac{1}{\tau_L} \tilde{L} + \tilde{s}_L(y) = 0
\]

Now using (3.2a) to eliminate \( d^2\tilde{N}(y)/dy^2 \), Lefty production in the pinned interval is given by:

\[
\tilde{s}_L(y) = \frac{1}{C_{LL}}(\tilde{C}_{LN}[(1 - \tilde{l}_L^2)\tilde{N}(y) + \tilde{l}_L^2\tilde{s}_N(y)] - 1) \tag{4.3}
\]

where \( \tilde{s}_N(y) = \begin{cases} 0, & y > 0 \\ 1, & y < 0 \end{cases} \)

The point of finding this expression is that as \( y \to -\infty \), we see that \( \tilde{s}_L \to \tilde{L}_s < 1 \). This simply means that as \( \tilde{L}_s \) goes to zero, so does the asymptotic production of Lefty.

The values of \( \tilde{l}_L, \tilde{C}_{LN} \) or \( \tilde{C}_{LL} \) in (4.3) are free to vary so long as \( \tilde{L} \) in (4.2) is between 0 and 1. On the other hand, the location of the front separating the pinned interval and the region of zero production must be determined from matching both \( \tilde{L} \) and \( d\tilde{L}/dy \) on either side of the boundary. It is possible to end up with a value of \( y_L \) where \( \tilde{s}_L \) lies outside \([0,1] \). Indeed, checking when this occurs using (4.3) demonstrates that \( \{p,0\} \) is inconsistent when

\[
\tilde{L} \tilde{l}_L > 1 \tag{4.4}
\]

Here, there are three regions instead: pinned, full production, and no production in short, \( \{p,1,0\} \). As \( \tilde{L}_s, \tilde{l}_L \) increases beyond 1, the length of the fully producing interval increases. Simulations and solving for the boundary conditions validate this (see figure 3).

#### 4.3.2 Case \( \tilde{L}_s > 1 \): \( \{1,0\} \) and \( \{1,p,0\} \)

The traveling wavefront given by (3.9) is still not guaranteed when \( \tilde{L}_s > 1 \). We can see this by considering the limit at which \( \tilde{l}_L \approx 1 \): at the front of Lefty production, \( \tilde{L} \) goes from 1 to 0 with a background of nearly constant \( N \). Since \( L \) inhibits its own production, the apparent paradox is that \( dC_L(y)/dy > 0 \), i.e. the regions of zero and full production are reversed! Simulations show that as \( \tilde{l}_L \) is decreased and the slope of \( L \) made steeper, once the slope exceeds that of \( \tilde{l} \), \( \tilde{L}_s \to 1 \) \( \{1,0\} \) will transition to \( \{1,p,0\} \). Basically, requiring that \( dC_L(y)/dy < 0 \) at the front is equivalent to this condition.

To write out explicit conditions for entering the \( \{1,p,0\} \) state, we should consider both cases \( y_L > 0 \) and \( y_L < 0 \), for which \( \tilde{N} \) takes different forms given in (4.1). Thus, the condition for \( \{1,p,0\} \) is that \( C_L(y) \) crosses the threshold in
the other direction: \( dC_L(y_L)/dy > 0 \), giving:

\[
\begin{align*}
y_L > 0 &: \quad \tilde{L}_L^{-1} > 1 + \frac{2}{C_{LL}}, \\
y_L < 0 &: \quad \tilde{L}_L^{-1} > 2\tilde{\tau}_L - 1. 
\end{align*}
\]

(Note that both conditions require \( \tilde{L}_L < 1 \), i.e. the Nodal length scale is greater than Lefty, as argued above in the limiting case.) If the inequalities are in the opposite direction, then we are back to the traveling wavefront given by (3.9), or in our notation 1,0. Here we can solve for the Lefty concentration profile fully: using (3.10) we find that \( \Delta y > 0 \) when

\[
\frac{1}{2} \tilde{C}_{LN} - \frac{1}{2} \tilde{C}_{LL} < 1. \tag{4.6}
\]

5. Solutions oscillating in time

In the previous section(s), we explored the limiting case of an autonomous, Lefty-independent Nodal production. We now address new phenomena which are possible when Nodal is significantly affected by Lefty. The most striking of these is the possibility of a time-oscillating solution, which we will study in the special case that all concentrations are uniform in space. The experimentally pertinent motivation is whether it is possible to generate traveling solutions, consisting of pulses of Nodal and Lefty, periodic in space and in time; one expects such solutions to be possible only if uniform oscillations are possible in the model.

By examining the geometry of the \((N, L)\) plane [Figure 5], it can be seen that only case (d) has the possibility of oscillations; this is shown with more detail in Fig. 5(b). Note that the fixed point in the center, since it has \( N_F < N_0 \) and \( L_F < L_0 \), corresponds to production rates less than saturation \( s_N < s_{N0} \) and \( s_L < s_{L0} \), so we are in a "pinned" regime for both \( N \) and \( L \), which is self-consistent with being on the isoclines for both.

\[
\tilde{L}_L \equiv \frac{\tau_L}{\tau_N} < 1. \tag{5.1}
\]

(Of course, the actual magnitude of the degradation times simply sets an overall time scale.)

We can describe the history with a series of times \( t_i, i=1,2,... \), at each of which one of the signals turns on or off. Between these times, the production rates are constant (either zero or saturated), and the differential equation is solved by a simple exponential:

\[
\tilde{N}(t) = \begin{cases} 
\tilde{N}(t_i)e^{-(t-t_i)/\tau_N}, & s_N = 0 \\
1 - [1 - \tilde{N}(t_i)]e^{-(t-t_i)/\tau_N}, & s_N = s_{N0},
\end{cases} \tag{5.2}
\]

and similarly for \( \tilde{L}(t) \).
To visualize the dynamics, it is convenient to draw the trajectory of \((\tilde{N}(t), \tilde{L}(t))\) in the \((\tilde{N}, \tilde{L})\) plane; in our equations, the time derivatives \(\dot{\tilde{N}} = d\tilde{N}/dt\) and \(\dot{\tilde{L}} = d\tilde{L}/dt\) are functions only of \((\tilde{N}, \tilde{L})\). The lines \(\tilde{C}_N(\tilde{N}, \tilde{L}) = 0\) and \(\tilde{C}_L(\tilde{N}, \tilde{L}) = 0\) on this plot are the isolines, meaning the places where (respectively) \(\dot{\tilde{N}}\) and \(\dot{\tilde{L}}\) change sign. (Isoclines were also used in the analysis of Ref. [13].) Each time the trajectory intersects an isocline is one of the times \(t_i\); let the concentrations at these times be \(\tilde{N}_i = \tilde{N}(t_i)\), \(\tilde{L}_i = \tilde{L}(t_i)\).

We can find the trajectory curve, and thus find solutions of the dynamical equations, by eliminating time and considering only the discrete map \((\tilde{N}_i, \tilde{L}_i) \rightarrow (\tilde{N}_{i+1}, \tilde{L}_{i+1})\). We next find the functional form of this map.

Consider phase III of the cycle, during which \(\tilde{N}(t) = \tilde{N}_i e^{-(t-t_i)/\tau_N}\) and \(\tilde{L}(t) = \tilde{L}_i e^{-(t-t_i)/\tau_L}\). Eliminating time, we get

\[
\frac{\tilde{L}}{\tilde{L}_i} = \left[ \frac{\tilde{N}}{\tilde{N}_i} \right]^{1/\tau_L} \quad (5.3a)
\]

The point \((\tilde{N}_{i+1}, \tilde{L}_{i+1})\) is the intersection of the curve (5.3a) with the isocline \(\tilde{C}_N(\tilde{N}, \tilde{L}) = 0\). In phase IV, the trajectory would be

\[
\frac{\tilde{L}}{\tilde{L}_i} = \left[ \frac{1 - \tilde{N}}{1 - \tilde{N}_i} \right]^{1/\tau_L} \quad (5.3b)
\]

and similarly in the other four phases of the cycle. Notice that the curve can possibly intersect the isocline in phases I or III only if \(\tilde{\tau}_L < 1\). (For example, when \(\tilde{\tau}_L = 1\) the trajectory in each phase is a straight line to one corner of the square.)

To give the cycling behavior, it is necessary (but not sufficient) that the isoclines intersect as shown in Figure 2(b), which depends on two inequalities. The \(\tilde{N}\) and \(\tilde{L}\) isoclines’ respective intercepts at \(\tilde{L} = 0\) satisfy \(1/\tilde{C}_{NN} < 1/\tilde{C}_{LN}\); their intercepts at \(\tilde{L} = 1\) must be in the reverse order, \((1 + \tilde{C}_{NL})/\tilde{C}_{NN} > (1 + \tilde{C}_{LL})/\tilde{C}_{LN}\). The two inequalities can be written together as

\[
\frac{\tilde{C}_{LL} + 1}{\tilde{C}_{NN} + 1} < \frac{\tilde{C}_{LN}}{\tilde{C}_{NN}} < 1 \quad (5.4)
\]

This tends to be satisfied when conditions (5.4) are satisfied, the isoclines cross at a fixed point \((\tilde{N}_*, \tilde{L}_*)\) where

\[
\tilde{N}_* = \frac{\tilde{C}_{NL} - \tilde{C}_{LL}}{\tilde{C}_{LN}\tilde{C}_{NL} - \tilde{C}_{NN}\tilde{C}_{LL}} \quad (5.5a)
\]

\[
\tilde{L}_* = \frac{\tilde{C}_{NN} - \tilde{C}_{LN}}{\tilde{C}_{LN}\tilde{C}_{NL} - \tilde{C}_{NN}\tilde{C}_{LL}} \quad (5.5b)
\]

5.2 Stability conditions

The trajectory always tends to spiral around \((\tilde{N}_*, \tilde{L}_*)\), as illustrated in Figure 5(b). However, there are different possible limiting behaviors depending on whether the trajectory spirals inwards (stable fixed point) or outwards, and on where it ends up. There is always a stable fixed point at \((\tilde{N}, \tilde{L}) = (0, 0)\) [the empty system] and provided the isoclines intercept the upper edge of \((\tilde{N}, \tilde{L})\) space [as in Figure 5(b)], there is also a stable fixed point at \((\tilde{N}, \tilde{L}) = (1, 1)\) [both Nodal and Lefty turned on and saturated]. Some of the possibilities:

1. (1) The fixed point is stable; there is one unstable cycle, such that a trajectory starting inside it tends to \((\tilde{N}_*, \tilde{L}_*)\) and a trajectory starting outside it tends to \((0, 0)\) or \((1, 1)\).
There are no cycles (stable or unstable); the fixed point is unstable, and spirals out to \((0, 0)\) or \((1, 1)\).

The fixed point is unstable, and spirals out to a stable cycle (beyond which is an unstable cycle, as in case (1)); this is the case of interest to us.

To evaluate the stability, we must linearize near the fixed point in terms of \(\delta \tilde{N} \equiv \tilde{N} - \tilde{N}_s\), \(\delta \tilde{L} \equiv \tilde{L} - \tilde{L}_s\). The trajectory (e.g. Eqs. (5.3a) or (5.3b)) becomes \(\dot{\delta} \tilde{L} = m_{ph} \delta \tilde{N}\), also \(m_{ph}\) is the slope in each respective phase of the cycle, \(ph \rightarrow I, II, III, or IV\). Thus,

\[
m_{III} = \frac{1}{\tau_L \tilde{N}_s};
\]

\[
m_{IV} = -\frac{1}{\tau_L \left(\frac{\tilde{L}_s}{1 - \tilde{N}_s}\right)};
\]

and similarly for the rest. A convenient viewpoint on the condition is that

\[
m_{ph} = \tilde{\tau}_L m_{ph,0}
\]

where \(m_{ph,0}\) is the slope of the line from \((\tilde{N}_s, \tilde{L}_s)\) to the appropriate corner of the square.

Meanwhile, the slopes of the \(\tilde{N} = 0\) and \(\tilde{L} = 0\) isoclines are

\[
m_N \equiv \tilde{C}_{NN} / \tilde{C}_{NL}, \quad (5.8a)
\]

\[
m_L \equiv \tilde{C}_{LN} / \tilde{C}_{LL}.
\]

Evidently, the additional (and sufficient) condition to get a spiraling behavior is that \(m_N < m_{III}, m_l < m_{IV}\). Since \(\tilde{\tau}_L < 1\) and \(m_{ph}\) is of order \(1/\tilde{\tau}_L\) for \(ph = I, II, III, IV\), it follows that \(m_L\) is typically large and hence \(C_{LL}\) must be relatively small.

Solving one linear equation for each phase of the cycle, we find after four isocline intersections that \((\delta \tilde{N}_{i+4}, \delta \tilde{L}_{i+4}) = \Lambda (\delta \tilde{N}_i, \delta \tilde{L}_i)\), where

\[
\Lambda = \frac{R_{l} R_{II}}{R_{l} R_{IV}}
\]

where

\[
R_{ph} = \frac{m_L - m_{ph}}{m_N - m_{ph}}
\]

for \(ph = I, II, III, IV\). Thus, the fixed point \((\tilde{N}_s, \tilde{L}_s)\) is stable if and only if \(\Lambda < 1\). An interesting special case is when \((\tilde{N}_s, \tilde{L}_s) = (1/2, 1/2)\); in that case, \(\Lambda \equiv 1\) so the fixed point is always marginal.

To get a better understanding of the stability, consider the case that \(m_N \ll m_1, ..., m_{IV} \ll m_{L} \). Then

\[
R_{ph} \approx -\frac{m_L}{m_{ph}} (1 - \frac{m_{N}}{m_{l}}) + \frac{m_{N}}{m_{ph}}.
\]

Noting that \(m_1 m_{III}/m_{III} m_{IV} \equiv 1\), we get

\[
\Lambda \approx 1 - \frac{1}{\tilde{\tau}_L m_L} \left(m_{1,0} + m_{III,0} - m_{1,0} - m_{IV,0}\right) + \tilde{\tau}_L m_N \left(m_{1,0}^{-1} + m_{III,0}^{-1} - m_{1,0}^{-1} - m_{IV,0}^{-1}\right).
\]

The sums in parentheses do not have a factor of \(\tilde{\tau}_L\); they depend only on \((\tilde{N}_s, \tilde{L}_s)\). Both sums tend to be positive when the fixed point is farther from the line \(L = \tilde{N}\) than it is from the line \(L = 1 - \tilde{N}\), or both negative in the opposite situation. Thus, in the former situation, the fixed point tends to be unstable when

\[
(m_L m_N)^{1/2} < \tilde{\tau}_L.
\]

In the opposite situation, the fixed point tends to be unstable when the inequality is in the other direction.

The oscillation period \(T\) can be inferred from the orbit on the \((N, L)\) plane by going back to equation (5.2). Note that \(\tilde{N}\) and \(\tilde{L}\) do not go to zero approaching the isocline line, but rather they approach a positive or negative constant coming from the respective sides. In other words, the four legs of the cycle on the \((N, \tilde{L})\) plane are each traversed at a roughly constant speed. Hence a cycle that forms a small loop around \((\tilde{N}_s, \tilde{L}_s)\), as happens just beyond the instability, has its period \(T\) proportional to its oscillation amplitude.

It follows that if the parameters are close to the instability threshold, we can construct a spatially periodic solution of intervals containing oscillations which are shifted in phase with respect to each other, provided that the spatial period is large compared to the distance that either signal can diffuse in time \(T\); locally, this situation is equivalent to the uniform one, since the signal from contrasting intervals does not have time to propagate. It follows that periodic traveling waves are also possible, as well as a standing wave made up of alternating domains that each oscillate similar to the uniform oscillations, but in opposite phase to each other.

6. Conclusion

In this work, we first sketched (Sec. 2) the realistic two-dimensional model developed in [14], which exhibits a uniformly moving posterior-to-anterior front of Nodal production on one of the two stripes representing the lateral plate mesoderm, followed by a front of Lefty production on the midline, just like the experimental observation [11], and which is known to propagate the left/right asymmetry from the tailbuds through the entire embryo. We then set up a version of this model in one dimension (Sec. 3), representing the anterior-to-posterior axis, which captures most properties of front propagation. The one-dimensional space represents a combination of the left LPM and the midline, and its main qualitative defect compared to the two-dimensional system is that it misses the lag time for the Nodal signal \(N\) to diffuse from the LPM to the midline, or for the Lefty signal \(L\) to diffuse from the midline to the LPM. The key parameters of the model were (1) coefficients [Sec. 2.2] quantifying how much Nodal promotes the two signals, and how much Lefty inhibits them, and (2) different diffusion constants and/or degradation times for the two signals, allowing us to define a typical length scale for either one, over which its concentration varies in front of and behind a sharp spatial...
step of the production; all in all, we found seven dimensionless ratios, each of which is a nontrivial parameter (Sec. 3.3).

We then set out to classify, analytically, the possible behaviors of the model in a steady state, and see how the depended on the parameter values. A key aid in identifying the regime is make a two dimensional plot (Sec. 3.4) of how the homogeneous concentrations $N$, $L$ would evolve: the very existence of a front depends on having a fixed point on this plane representing nonzero Nodal production, alongside the fixed point of zero production (which is always present). Our main focus was on uniformly moving fronts, but in fact one can specialize (without real loss of generality, and with considerable mathematical simplification) to the special case of zero front velocity [Sec. 3.2]. The key special feature we found was the possibility that, instead of showing a sharp front where the production rapidly switches from zero to maximum, the Lefty production (on the midline in the real embryo) might be varying continuously over an extended interval (Sec. 4). This was due to a sort of feedback or buffering, in which the Nodal and Lefty concentrations compensate exactly such that every point along this interval is right at the threshold for Lefty production (“pinned”).

Finally, in Sec. 5 we changed focus to the possibility of a temporally periodic behavior; this parameter regime allows both the possibility of a steady limiting state which is “pinned” (at threshold) for both Nodal and Lefty. One may get oscillations if Lefty degrades much faster than Nodal, and we characterized the exact mathematical criterion for the steady state to be unstable to developing oscillations.

Having found the relationship of these behaviors to the parameter values, in future work we will be able to find the corresponding parameter regimes in the realistic two-dimensional model, with the ultimate goal of finding a set that gives the observed phenotypes. We obtained many behaviors that are not seen experimentally. (It is still uncertain whether the Lefty production has a sharp front or an extended one like the “pinned” case.) This is a positive feature of the model, for it will allow the exclusion of whole domains of parameter values.

Our results might motivate further studies of the possible behaviors of propagating L/R asymmetry in the zebrafish and other embryos. The pertinent measurements in real systems that relate to phenomena in this model are

1. The measured front velocity constrains the velocity scale $v_N$.
2. The sharpness of the $N$ and $L$ concentration fronts gives the length scales $l_N$ and $l_L$ and/or indicates the possibility of the “pinned” interval; however, experiments that measure mRNA are in effect probing the production rate and do not access $l_N$ and $l_L$ directly.
3. The lag between the Nodal front and the Lefty front, gives information on the difference between Nodal and Lefty threshold parameters. Experimentally the Lefty front lags, but many parameter sets give a Lefty front that spatially leads (even though causally it is downstream from Nodal), so observation can limit parameters to a certain window.

In addition, they suggest how changing a parameter (due to a mutation or some external perturbation) could give an exotic result, such as non-monotonic front.

What the present study did not address is how the Nodal production is first initiated, and why it only happens along the left LPM. That depends on the two-dimensional geometry, e.g. the relative separations of the tailbud from the foot of the LPM stripes and the midline stripe of Oep-producing cells. Furthermore, it is possible the real system could propagate, e.g., a finite pulse of Nodal that terminates, but that will be seen only if such a signal is produced at the posterior end. These questions must be left to a proper two-dimensional study.

SUPPLEMENTARY MATERIAL

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REFERENCES and FOOTNOTES

1. H. Hamada, C. Meno, D. Watanabe, and Y. Sajioh. Establishment of Vertebrate Left-right Asymmetry. Nature Reviews Genetics 3:103-113, 2002.
2. M. Levin. Left-right asymmetry in embryonic development: a comprehensive review. Mechanisms of Development 122: 3-25, 2005.
3. A. Raya and J.C.I. Belmonte. Left-right asymmetry in the vertebrate embryo: from early information to higher-level integration. Nature Reviews Genetics 7:283-293, 2006.
4. J. McGrath, S. Somlo, S. Makova, X. Tian, and M. Brueckner. Two populations of node monocilia initiate left-right asymmetry in the mouse. Cell 114:61-73, 2003.
5. S. Nonaka, et al. Randomization of Left–Right Asymmetry due to Loss of Nodal Cilia Generating Leftward Flow of Extraembryonic Fluid in Mice Lacking KIF3B Motor Protein. Cell 99:829-837, 1998.
6. S. Nonaka, H. Shiratori, Y. Sajioh, and H. Hamada. Determination of left–right patterning of the mouse embryo by artificial nodal flow. Nature 418:96-99, 2002.
7. J. J. Essner, et al. Left–right development: Conserved function for embryonic nodal cilia. Nature 418:37-38, 2002.
8. D. S. Adams, K. R. Robinson, T. Fukumoto, S. Yuan, R. C. Albertson, P. Yelick, L. Kuo, M. McSweeney, and M. Levin. Early, $H^+\text{-V-ATPase}$-dependent proton flux is necessary for consistent left-right patterning of non-mammalian vertebrates. Dev. 133:1657-1671, 2006.
9. A. Kawasumi et al. Left-right asymmetry in the level of active Nodal protein produced in the node is translated into left-right asymmetry in the lateral plate of mouse embryos. Dev. Biol. 353:321-330, 2011.
10. J. Brennan, D.P. Norris, and E.J. Robertson. Nodal activity in the node governs left-right asymmetry. Genes and Development 16: 2339-2344, 2002.
11. X. Wang, and H.J. Yost. Initiation and Propagation of Posterior to Anterior (PA) Waves in Zebrafish Left-Right Development. Developmental Dynamics 237:3640-3647, 2008.
12. A.K. Ryan, et al. W. Sobaghi, J. Greenwald, S. Choe, D. P. Norris, E. J. Robertson, R. M. Evans, M. G. Rosenfeld, and J. C. I. Belmonte. Pitx2 determines left–right asymmetry of internal organs in vertebrates.
13. T. Nakamura, et al. Generation of Robust Left-Right Asymmetry in the Mouse Embryo Requires a Self-Enhancement and Lateral-Inhibition System. Developmental Cell 11:495-507, 2006.

14. B. Xu. “Systematic Analysis of Asymmetric Nodal Signaling in the Development of Zebrafish Left-Right Patterning.” (Ph. D. thesis, Princeton University, 2010)

15. C.L. Henley, B. Xu, and B. Burdine. Simplified model for Nodal front propagation in Zebrafish embryo (unpublished).

16. J. J. Essner, J. D. Amack, M. K. Nyholm, E. B. Harris, and H. J. Yost, Kupffer’s vesicle is a ciliated organ of asymmetry in the zebrafish embryo that initiates left-right development of the brain, heart and gut, Development 132:1247-1260 (2005).

17. L. Wolpert, J. Smith, T. Jessell, P. Lawrence, E. Robertson, and E. Meyerowitz. Principles of Development (3rd ed.). OUP, New York, 2007.

18. K. A. Smith, et al. Bmp and Nodal Independently Regulate lefty1 Expression to Maintain Unilateral Nodal Activity during Left-Right Axis Specification in Zebrafish. PLoS Genetics 7:e1002289, 2011.