Sub-acute Toxicity of Aqueous Extracts of *Tephrosia vogelii*, *Vernonia amygdalina* and *Senna occidentalis* in Rats

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Abstract

There is an intense search for new plant-derived medicines to match the ever increasing prevalence of drug resistance. Toxicity profiles of three commonly used plants against helminthiasis in Uganda will support their optimum use.

**Objective:** To evaluate the sub-acute toxicity of aqueous leaf extracts of *Tephrosia vogelii* (TV), *Vernonia amygdalina* (VA) and *Senna occidentalis* (SO).

**Methods:** One hundred Wistar white albino rats in 10 groups were dosed orally daily with 200, 400 and 600 mg/kg of TV, VA and SO for 28 days or Goodwin’s physiological solution. Daily changes in behaviour, feed/water intake and body weight (weekly) were monitored. On day 29, blood for haematology, serum for biochemical analysis and organs (lungs, liver, kidney, heart and small intestines) for histopathology were collected.

**Results:** Continued exposure of rats to 600 mg/kg doses of SO and VA extracts led to elevation of ALT, AST and ALP and cholesterol in the treatment groups. SO and VA at 600 mg/kg elevated urea and uric acid; depressed bilirubin and albumin while VA and TV 600 mg/kg caused the reverse. Histopathology findings (villi sloughing, pneumonitis, liver and kidney degeneration) further supported these results.

**Conclusion:** The very low toxicity of the aqueous leaf extracts of TV, VA and SO at high doses makes them safe at currently non-standardised doses used for animal treatment.

**Keywords:** *Tephrosia vogelii; Vernonia amygdalina; Senna occidentalis*; Toxicity; Rats

Background

Uganda has 80% of its population depending on agriculture and about 50% are engaged in livestock farming [1]. In small-scale livestock production, majority of the farmers use a combination of conventional and ethnopharmacological formulations to treat diseases and parasitic infections [2,3]. Use of plants in animal disease treatment is practiced traditionally [2-4,6], albeit with non-standardised doses as a major shortfall [4,7,8]. This is further compounded by the basic belief that plant extracts are non-toxic [2,7,9]. In an earlier study mapping ethnopharmacological practices in Mpigi and Gulu districts in Uganda, farmers prioritized *Vernonia amygdalina* (VA), *Tephrosia vogelii* (TV) and *Senna occidentalis* (SO) for detailed evaluation [3].

*Tephrosia vogelii* Hook (Family Leguminosae; synonym *Cacalia amygdalina*, Hook F; ILDIS record 5085), is a soft woody branching herb, 1-4 m tall [10]. The plant is widely promoted in eastern and southern Africa for its insecticidal properties [11], although some people use it as a fish poison [12]. *Vernonia amygdalina* Delile (Family Asteraceae; synonym *Cacalia amygdalina* [http://www.theplantlist.org/tpl/record/ gcc-84295] Kuntze, TICA Id 2949) is a shrub that grows predominantly in Mbarara Zonal Agricultural Research Development Institute farm, located 274 km south-west of Kampala, the capital of Uganda. The voucher specimens were confirmed by a botanist and deposited in Makerere University Herbarium (numbers SO - NI 02, VA - NI 03 and TV - NI 04). The leaves were dried under shade in the laboratory for two weeks. They were ground to fine powder with an automated electric grinder (Grindomix, Retsch, GM 200). For aqueous extraction, the safest dose was used to calculate doses for the sub-acute toxicity study.

Methods

Animal dosing and monitoring

After determining the lethal doses and safety margins of the aqueous extracts, the safest dose was used to calculate doses for the sub-acute toxicity study. The objective of this study was to determine the sub-acute toxicity due to prolonged administration of these three commonly used plants in rats.

Materials and Methods

Plant collection and extract preparation

Fresh leaves of all plants were collected in January 2013 from wild shrubs at Mbarara Zonal Agricultural Research Development Institute farm, located 274 km south-west of Kampala, the capital of Uganda. The voucher specimens were confirmed by a botanist and deposited in Makerere University Herbarium (numbers SO - NI 02, VA - NI 03 and TV - NI 04). The leaves were dried under shade in the laboratory for two weeks. They were ground to fine powder with an automated electric grinder (Grindomix, Retsch, GM 200). For aqueous extraction, the safest dose was used to calculate doses for the sub-acute toxicity study.
acutely. In the protocol, OECD test guidelines were followed [29].

Briefly, 100 Wistar white albino rats (50 females and 50 males), of about 6 to 8 weeks old weighing 70-100 g were procured from Uganda Virus Research Institute and kept for two weeks to acclimatize to the new laboratory conditions. Each extract was reconstituted in Goodwin’s physiological solution to a stock concentration of 200 mg/ml. The rats were weighed and randomly assigned to 10 treatment groups i.e. of each extract, 200 mg/kg, 400 mg/kg and 600 mg/kg body weight plus the negative control (given Goodwin’s physiological solution w/v). In each group, the number of females and males was equal. Each group was kept in 80x50 cm wooden cages, fed on commercial rodent feed (Unga Limited, Kenya) and given water ad libitum. The rats were treated once daily, while body weights were taken once a week and on the 29th day, before termination of the experiment.

Sample collection

On the 29th day, the rats were weighed, anaesthetised with 70% chloroform (trichloromethane, E08A/08082/2104/13, Fine-Chem, India) for ten minutes before blood and tissue collection. Whole blood collected by cardiac puncture was transferred into labelled and EDTA vacutainer tubes (BD Plymouth, UK). EDTA blood was kept at 4°C while non-EDTA blood was centrifuged at 1500 g for three minutes to separate serum, which was kept at -20°C. The rats were then dissected to collect the liver, kidneys, small intestine sections, heart and lungs; which were weighed and kept in 10% buffered formalin for histopathology.

Haematological and biochemical tests

The EDTA whole blood for haematology was processed within 24 hours as described previously [31]. The automated haematological analyser (Coulter STKS, Beckman) was used to obtain the following: white blood cell counts (WBC), differential counts of neutrophils, lymphocytes, monocytes, eosinophils and basophils; red blood cells (RBC), haemoglobin (Hb), haematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular haemoglobin concentration (MCHC), mean corpuscular haemoglobin (MCH), red cell width to corpuscular volume (RDW-CV), mean platelet volume (MPV) and platelets (PLT). Serum kept at -20°C was thawed to 25°C and processed during organ collection and processing.

Results

Physical signs and gross necropsy findings

In all treatment groups, we observed no changes in feed and water intake. There were three deaths (one each at 600 mg/kg of TV, SO and VA) after 15, 24 and 26 days after treatment respectively. The immediate cause was wounds due to cannibalism, but after housing the rats individually, no more deaths were registered. No pathological gross necropsy findings were detected in treated and control rats during organ collection and processing.

Effect on body, heart, liver and kidney weights

There was a gradual increase in body weight in rats across all treatment groups but mean weight of SO 600 mg/kg was lower than other groups at day 0 and 14. However, the difference was non-existent during organ collection and processing. Although the liver weights ranged from 4.6-9.7 g, there was no statistical difference in mean weights due to treatment.

| Treat group & dose (mg/kg) | WBC (10³/µl) | RBC (10⁶/µl) | HCT (%) | MCV (fL) | MCH (pg) | MCHC (g/dL) | RDWCV (%) | PLT (10³/µl) | MPV (fL) |
|---------------------------|--------------|--------------|---------|----------|----------|-------------|-----------|-------------|----------|
| SO 200                    | 13.1 ± 2.2   | 7.6 ± 0.2    | 13.4 ± 0.2 | 46.5 ± 1.2 | 61.3 ± 1.2 | 17.7 ± 0.3 | 28.9 ± 0.3 | 23.0 ± 0.8 |   869.7 ± 52.7 | 8.9 ± 0.2 |
| SO 400                    | 13.3 ± 3.1   | 7.1 ± 0.4    | 12.9 ± 0.7 | 45.5 ± 3.1 | 63.7 ± 1.6 | 18.1 ± 0.3 | 28.5 ± 0.5 | 23.2 ± 1.4 |   832.5 ± 106.8 | 8.6 ± 0.2 |
| SO 600                    | 9.5 ± 2.6    | 6.7 ± 0.2    | 12.7 ± 0.7 | 41.5 ± 2.1 | 62.2 ± 1.5 | 19.1 ± 0.3 | 30.7 ± 0.4 | 26.2 ± 4.6 |   684.7 ± 16.0 | 8.8 ± 0.1 |
| Control                   | 19.6 ± 10.5  | 7.4 ± 0.3    | 13.5 ± 0.4 | 44.9 ± 2.3 | 60.7 ± 2.3 | 18.4 ± 0.4 | 30.4 ± 0.8 | 19.5 ± 0.6 |   733.8 ± 34.8 | 8.6 ± 0.3 |
| TV200                     | 9.3 ± 1.6    | 7.5 ± 0.5    | 13.2 ± 0.7 | 46.7 ± 3.2 | 62.5 ± 1.2 | 17.7 ± 0.2 | 28.5 ± 0.6 | 21.6 ± 1.0 |   731.8 ± 34.4 | 9.2 ± 0.6 |
| TV400                     | 9.9 ± 2.4    | 7.0 ± 0.2    | 12.7 ± 0.2 | 42.2 ± 2.3 | 60.7 ± 1.1 | 18.3 ± 0.4 | 30.2 ± 0.2 | 23.1 ± 1.1 |   362.5 ± 95.3 | 9.6 ± 0.2 |
| TV600                     | 14.6 ± 4.3   | 5.9 ± 1.1    | 11.4 ± 1.4 | 39.3 ± 6.1 | 67.1 ± 2.4 | 19.7 ± 1.2 | 29.3 ± 0.9 | 22.7 ± 2.4 |   397.7 ± 78.9 | 8.5 ± 0.0 |
| VA200                     | 10.5 ± 0.7   | 7.3 ± 0.5    | 13.0 ± 0.9 | 46.0 ± 3.6 | 63.1 ± 1.1 | 17.8 ± 0.2 | 28.3 ± 0.7 | 22.9 ± 0.7 |   849.0 ± 87.6 | 8.7 ± 0.2 |
| VA400                     | 11.4 ± 0.5   | 7.7 ± 0.9    | 14.3 ± 0.8 | 49.6 ± 5.6 | 64.4 ± 1.4 | 18.5 ± 0.3 | 28.8 ± 0.3 | 21.1 ± 1.0 |   943.0 ± 56.3 | 9.1 ± 0.1 |
| VA600                     | 13.1 ± 2.9   | 8.1 ± 0.3    | 14.8 ± 0.1 | 50.9 ± 1.2 | 63.2 ± 1.1 | 18.0 ± 0.6 | 28.5 ± 0.4 | 21.3 ± 0.8 |   714.0 ± 109.0 | 9.7 ± 0.3 |

*Mean Platelet values significantly different from control and other treatment groups; P<0.005

Table 1: Haematological parameters (Mean ± SEM) of 100 Wistar white albino rats in nine treatment groups and a placebo control group.

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Effect on biochemical and haematological parameters

The extracts caused varied effects on haematology and biochemistry (Table 1). For example, there was depression of haematoctrit in a doseresponse pattern, whereas the WBC counts were much higher in the control (19.6 ± 10.5x10^3/µl) compared to other groups (<15 x10^3/µl). A reverse relationship was observed with RDCHV; showing elevated levels with increasing doses for all plant extracts but there was minimal change in MCHC. At increasing doses, TV had a significant reducing effect on the mean platelet counts compared to other treatment groups and the controls (F_{1,9}=3.4; P=0.005).

Levels of liver function enzymes; ALP, ALT, AST and cholesterol significantly differed across the treatment groups (F_{1,9}=2.88, P = 0.035; 2.55, 0.022; 2.32, 0.035 and 3.35, 0.004 respectively). At 600 mg/kg, TV elevated the AST enzyme to 355.6 U/L while SO 600 mg/kg depressed it to 182.0 U/L, compared to 227.1 U/L in the control group (Figure 1). Cholesterol levels also differed in SO 600 mg/kg (3.55), VA 600 mg/kg (3.38) and TV 200 mg/kg (2.13), TV 600 mg/kg (2.36) compared to 2.93 in the control group (Figure 1A and 1B). Furthermore, albumin protein was elevated in SO and VA in dose-response manner compared to TV, which led to a small change (Table 2). The elevation was significant at was elevated in SO and VA in dose-response manner compared to TV, and small intestines. TV caused more toxic changes in the organs compared to SO and VA. For example, at TV 600 mg/kg, villi sloughing in intestines, myocardial and renal congestion as well as hyaline degeneration of the liver were recorded. Slight pneumonitis in the lungs with mononuclear cell infiltration was also noted in TV 600 mg/kg (Figure 2). Most of the changes followed a dose-response pattern.

**Histopathology and microscopic effects of the treatments**

As shown in Table 3 and Figure 2A-2F, there were varied effects of treatments on the histopathology of the liver, kidney, lung, heart and small intestines. TV caused more toxic changes in the organs compared to SO and VA. For example, at TV 600 mg/kg, villi sloughing in intestines, myocardial and renal congestion as well as hyaline degeneration of the liver were recorded. Slight pneumonitis in the lungs with mononuclear cell infiltration was also noted in TV 600 mg/kg (Figure 2). Most of the changes followed a dose-response pattern.

**Discussion**

The geographical location, season, soil type, climate differences, agronomic practices, plant part and age affect the concentration of phytochemicals, usually leading to differences in bioactivity and toxicity [7,16,18]. Using proteomics, metabolomics and transcriptomics in recent advances to determine nephrotoxic, genotoxic and teratogenic effects of natural plant products has been recommended [34]. These *in silico*, *in vitro* and *in vivo* methods which may explain the mechanisms of action, are however still unstandardised [34]. Following the established protocol [29,33], the recommended number of animals used supported statistical analysis.

The difference in concentration of phytochemicals extracted by various solvent systems may account for differences in observed toxicity of a plant extract at the same doses. For example, saponins are incriminated to cause haemolysis [35]. Although VA has significant levels of saponins [36,37], there was a dose-dependent increase in RBC counts. On the contrary, a reduction in RCR counts with increasing doses of SO and TV and PCV was noted. This is a possible mechanism of toxicity for the latter two extracts. Although no specific study on effect of extract on platelet count was done, we observed a marked reduction in platelets in higher dose groups of TV (363 and 398 compared to 734x10^3/µl in control). When platelets are significantly reduced, there may be multiple organ failure, eventually leading to toxicity signs or death.

Specifically, rotenone present in TV [11,38], causes similar signs of poisoning in humans and is one of the possible chemicals responsible for the observed acute toxicity [21]. In insects and fish, rotenone interferes with mitochondrial electron transport system, whereas mammals are more tolerant [38]. Rotenone however, is less soluble in water and easily disintegrates in the environment [12] which explains the presently observed reduced toxicity by aqueous extracts. Anthracenosides and anthocyanosides have been incriminated as toxic phytochemicals previously. However, qualitative phytochemistry (results not presented here) showed both anthracenosides and anthocyanosides in the aqueous extract of TV and VA.
Some of the phytochemicals in SO are reported to be hepatoprotective [18,39]. In the current study, the 600 mg/kg dose of aqueous extracts of SO exerted significant decrease in AST, ALT enzymes but increased urea and uric acid compared to the controls and other treatment groups. These results support previous findings that the SO leaf extract is hepatoprotective [39]. On the contrary, in a previous study, there was no change in liver enzymes and haematological parameters in female and male rats for SO at doses as high as 2500 mg/kg/day [28]. Their study therefore, did not report any toxicity of SO in the male or female rats. SO and VA caused similar biochemical changes in the rats. For example, they both caused depression of ALT and AST in a dose-response manner; quite the opposite to the effect of TV (elevation with higher doses). In addition, they both caused an elevation in albumin protein. Depression of this protein clearly indicates chronic liver damage compared to elevation of ALT and AST (sometimes caused by muscle damage), thus a better indicator of liver function. This further confirms the hepatoprotective effect of SO and VA as previously discussed [13,23,39].

Although there were no visible gross changes observed in the sampled organs, histopathology results indicated more toxic effects by TV compared to SO and VA. Sloughing of the intestinal villi observed in SO and TV due to continued necrosis can lead to reduced nutrient absorption, which will eventually impair liver and kidney function. Antioxidant activity of SO by reducing DNA degeneration (iron II-driven Fenton reaction) has been demonstrated with seed extracts. Inhibition of DNA damage by strong ion chelation activity is also hepatoprotective [18,40]. In chicken, studies demonstrated lower phosphorylation ratios and respiratory control ratios and oxygen use in liver mitochondria [18]. Pneumonitis, mononuclear cell infiltration and lymphoid necrosis as observed in some cases in this study lead to compromised gaseous exchange and eventual death. Underlying toxicity mechanisms at cellular level should be undertaken with standardised proteomics, metabolomics and transcriptomics.

**Conclusion**

SO and VA were hepatoprotective while TV was slightly hepatotoxic in a dose response manner. The very low toxicity of the aqueous leaf extracts of TV, VA and SO at high doses provide for its safety within the irregular and unstandardized doses used for human and animal treatment.

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**Author contributions**

Denis Mugizi, John Kateregga, Deogracious Oilla and Immaculate Nabukenya contributed to the study design, data collection and manuscript writing. Chris Rubaire-Akiiki, Deogracious Oilla and Johan Höglund supervised the laboratory work and contributed to manuscript writing.

**References**

1. (2008) Ministry of Agriculture, Animal Industry & Fisheries and Uganda Beareau of statistics, The National Livestock Census Report, Ministry of Agriculture.
2. Nalule AS, Mbaria JM, Oilla D, Kimenju JW (2011) Ethnopharmacological practices in management of livestock helminths by pastoral communities in the dry lands of Uganda. Livestock Research for Rural Development 23.

3. Nabukenya I, Rubaire-Akiiki C, Mwesigye P, Kasenene JM (2007) Medicinal plant diversity and uses in the Mt. Elgon region of Uganda. J Ethnopharmacol 10: 9-11.

4. Tabutu JR, Kukunda CB, Waako PJ (2010) Medicinal plants used by traditional medicine practitioners in the treatment of tuberculosis and related ailments in Uganda. J Ethnopharmacol 127: 130-136.

5. Oilla D, Bukunya-Ziraba R and Kamoga D (2007) Bio-prospective studies on medicinal plants used to manage poultry diseases in the dry lands of Uganda. J Pharm Pharmacol 1: 56-60.

6. Ssegawa P, Kasenene JM (2007) Medicinal plant diversity and uses in the Sango bay area, Southern Uganda. J Ethnopharmacol 113: 521-540.

7. Gitthori JB, Högland J, Walter PJ (2005) Ethnoveterinary plant preparations as livestock dewormers: practices, popular beliefs, pitfalls and prospects for the future. Anim Health Res Rev 6: 91-103.

8. Fieroni A, Howard P, Volpato G, Santoro RF (2004) Natural remedies and nutraceuticals used in ethnoveterinary practices in inland southern Italy. Vet Res Commun 28: 55-80.

9. Nanyingi MO, Mbaria JM, Lanyasunya AL, Wagate CG, Koros KB, et al. (2008) Ethnopharmacological survey of Samburu district, Kenya. J Ethnobiol Ethnomed 4: 14.

10. The plant list.

11. Stevenson PC, Kite GC, Lewis GP, Forest F, Nyinenda SP, et al. (2012) Distinct chemotypes of Tephrosia vogelii and implications for their use in pest control and soil enrichment. Phytochemistry 76: 135-146.

12. Dzenda T, Ayo JO, Adelaye AE, Adaudo AO (2007) Ethnomedical and veterinary uses of Tephrosia vogelii Hook F (Fabaceae): a review. Nigerian Veterinary Journal 28: 24-39.

13. Iwalokun BA, Efedede BU, Iyiola TO, Adedapo AA, Toyang NJ, Akinbode FO (2003) Antidiabetic and acute toxicity of spray-dried Vernonia amygdalina water extracts on mice. J Ethnopharmacol 86: 199-203.

14. Awaal CB, Adamu AM, Gefu JO, Ajansu OJ, Abdou PA, et al. (2003) In vitro screening of two Nigerian medicinal plants (Vernonia amygdalina and Annona senegalensis) for anthelmintic activity. Vet Parasitol 113: 73-81.

15. Yeap SK, Liang WS, Beh BK, Ho WY, Youssu AN, Alttheen NB (2013) In vivo anti-diabetic and acute toxicity of spray-dried Vernonmia amygdalina water extracts on rats. World J Gastroenterol 19: 224-230.

16. Toyang NJ, Verpoorte R (2013) A review of the medicinal potentials of plants of the genus Vernonia (Asteraceae). J Ethnopharmacol 146: 681-723.

17. Adebajo AA, Otesile AT, Soetan KO (2007) Assessment of the Anthelmintic efficacy of the aqueous crude extract of Vernonia amygdalina. Pharmaceutical Biology 45: 564-568.

18. Yadav JP, Aya V, Yadav P, Pangalal M, Kumar S, et al. (2010) Cassia occidentalis L.: a review of its ethnobotany, phytochemical and pharmacological profile. Fitoterapia 81: 223-230.

19. Tona L, Messe K, Ngimbi NP, Chimwana B, Okon'oucheda, et al. (2001) In vivo antimalarial activity of Cassia occidentalis, Morinda morindoides and Phyllanthus niruri. Ann Trop Med Parasitol 95: 47-57.

20. Verma L, Khatri A, Kaushik B, Patil UK, Pawar RS (2010) Anti-diabetic activity of Cassia occidentalis (Linn) in normal and alloxan-induced diabetic rats. Indian J Pharmacol 42: 224-228.

21. Dzenda T, Ayo JO, Adelaye AE, Adaudo AO and Ibrahim DG (2007) Preliminary investigation into the acute oral toxicity of Tephrosia vogelii leaves in mice. Niger Vet J 28: 47-52.

22. Ibrahim G, Abdurahman EM, Ibrahim H, Ibrahim NDG, Magaji MG (2011) Toxicity and analgesic effects of Vernonia amygdalina del. (Asteraceae) leaf extract on mice. Int J Adv Pharm Biol Sci 1: 1-4.

23. Ojiako OA, Nwanjo HU (2006) Is Vernonia amygdalina hepatotoxic or hepatoprotective? Response from biochemical and toxicity studies in rats? Afr J Biotechnol 5: 1648-1651.

24. Panwar RS, Munir K (2008) Cassia occidentalis toxicity causes recurrent outbreaks of brain disease in children in Saharanpur. Indian J Med Res 127: 413-414.

25. Vashisththa VM, John T, Kumar A (2009) Clinical & pathological features of acute toxicity due to Cassia occidentalis in vertebrates. Indian J Med Res 130: 23-30.

26. Suliman HB, Waafi IA, Adam SE (1982) The toxicity of Cassia occidentalis to goats. Vet Hum Toxicol 24: 306-330.

27. Rogers RJ, Gibson J, Reichmann KG (1979) The toxicity of Cassia occidentalis for cattle. Aust Vet J 55: 408-412.

28. Silla MG, Aragão TP, Vasconcelos CF, Ferreira PA, Andrade BA, et al. (2011) Acute and subacute toxicity of Cassia occidentalis L. stem and leaf in Wistar rats. J Ethnopharmacol 136: 341-346.

29. Hartmann E, Straus V, Eiben R, Freyberger A, Kaufmann W, et al. (2008) ESTP comments on the draft updated OECD test guideline 407. Exp Toxicol Pathol 59: 297-300.

30. National Research Council of the National Academies (2011) Guide for the Care and Use of Laboratory Animals, NIH publication. (8th edn), 500 Fifth Street, Washington D.C.

31. Hussain S, Fareed S, Siddiqui HH, Vijay Kumar M, Rao CV (2012) Acute and sub-acute oral toxicity evaluation of Tephrosia purpurea extract in rodents. Asian Pacific J Tropical Disease 2: 129-132.

32. Amole OJ, Isegbue MC, Onakoya JAA, Dada MO (2006) Toxicity studies of the aqueous extract of Vernonia amygdalina. J Biomed Res 17: 39-40.

33. Test No. 425: Acute Oral Toxicity: Up-and-Down Procedure (2008) OECD Guidelines for the Testing of Chemicals.

34. Ouedraogo M, Baudoux T, Stévigny C, Norrier J, Colet JM, et al. (2012) Review of current and “omics” methods for assessing the toxicity (genotoxicity, teratogenicity and nephrotoxicity) of herbal medicines and mushrooms. J Ethnopharmacol 140: 492-512.

35. Sparg SG, Light ME, van Staden J (2004) Biological activities and distribution of plant saponins. J Ethnopharmacol 94: 219-243.

36. Nalule AS, Karue CN, Katungu-Rwakishaya E (2011) Anthelmintic activity of Phytolacca dodecandra and Vernonia amygdalina leaf extracts in naturally infected small East African goats. Livestock Research for Rural Development 23.

37. Waiswa P, Oilla D (2006) The in vitro Ascaricidal activity of selected indigenous medicinal plants used in ethnoveterinary practices in Uganda. Afr J Tradit Complement Altern Med 3: 94-103.

38. Matsumura F (1975) Toxicology of Insecticides. Plenum Press: New York.

39. Jafari MA, Jalil Subhani M, Javed K, Singh S (1999) Hepatoprotective activity of leaves of Cassia occidentalis against paracetamol and ethyl alcohol intoxication in rats. J Ethnopharmacol 66: 355-361.

40. Tasaka AC, Weg R, Calore EE, Sinhorini IL, Dagli ML, et al. (2000) Toxicity testing of Senna occidentalis seed in rabbits. Vet Res Commun 24: 573-582.