Thymic Origin of Embryonic Intestinal $\gamma/\delta$ T Cells

By Dominique Dunon,* Max D. Cooper,† and Beat A. Imhof*

From the *Basel Institute for Immunology, CH-4058 Basel, Switzerland; and the †Howard Hughes Medical Institute, University of Alabama, Birmingham, Alabama 35294

Summary

Current evidence suggests both thymic and extrathymic origins for T cells. Studies in mice favor an in situ origin for a prominent population of intestinal intraepithelial lymphocytes that express $\gamma/\delta$ T cell receptor (TCR). This developmental issue is explored in an avian model in which the $\gamma/\delta$ lymphocytes constitute a major T cell subpopulation that is accessible for study during the earliest stages of lymphocyte development. In the chick embryo, cells bearing the $\gamma/\delta$ TCR appear first in the thymus where they reach peak levels on days 14-15 of embryogenesis, just 2 d before $\gamma/\delta$ T cells appear in the intestine. Using two congenic chick strains, one of which expresses the ov antigen, we studied the origin and kinetics of intestinal colonization by $\gamma/\delta$ T cells. The embryonic $\gamma/\delta^+$ thymocytes homed to the intestine where they survived for months, whereas an embryonic $\gamma/\delta^-$ thymocyte population enriched in thymocyte precursors failed to give rise to intestinal $\gamma/\delta^+$ T cells. Embryonic hemopoietic tissues, bone marrow, and spleen, were also ineffective sources for intestinal $\gamma/\delta^+$ T cells. Intestinal colonization by $\gamma/\delta^+$ thymocytes occurred in two discrete waves in embryos and newly hatched birds. The data indicate that intestinal $\gamma/\delta^+$ T cells in the chicken are primarily thymic migrants that are relatively long-lived.

Lymphocytes bearing $\gamma/\delta$ TCR are preferentially localized in the intestinal epithelium in both birds and mammals (for review see references 1 and 2). Although intraepithelial lymphocytes (IEL) of the small intestine are anatomically positioned to be the first line of cellular defense against enteric pathogens, the true function of $\gamma/\delta$ IEL remains unclear. Cytolytic capacity can be demonstrated, but specificity and MHC restriction patterns have not been defined for $\gamma/\delta$ IEL (3, 4). Their localization in the intestinal epithelium is independent of normal microbial colonization (5). It is interesting that $\gamma/\delta$ IEL have the capacity to reverse oral tolerance when adoptively transferred, although direct antigen reactivity of these $\gamma/\delta$ IEL has not been demonstrated (6, 7).

The elucidation of $\gamma/\delta$ T cell origin and migration characteristics should facilitate understanding of the generation of distinct TCR repertoires observed in different anatomical sites (8). Although the thymus is clearly an important source of $\gamma/\delta$ T cells (2), experiments conducted in immunocompromised mice have suggested that a significant proportion of the $\gamma/\delta$ IEL may be generated extrathymically. Cells from bone marrow or day 15 fetal liver infused into irradiated, thymectomized mice gave rise to $\gamma/\delta$ IEL, and the IEL in nude athymic mice are predominantly $\gamma/\delta$ TCR+ (9-11). On analyzing the generation of CD8+ $\gamma/\delta^+$ IEL in thymectomized mice depleted of CD8+ cells and showing that IEL contained mRNA for the recombinase activating gene (RAG-1) protein required for TCR rearrangement, Guy-Grand et al. (12, 13) proposed that most $\gamma/\delta^+$ IEL are derived in situ from precursors of extrathymic origin.

The developmental origin of $\gamma/\delta$ IEL is an important issue since these cells are the first lymphoid cells to appear in the intestine and may play important immunological roles such as control of oral tolerance, control of bacterial colonization, and elimination of damaged epithelial cells. The data on $\gamma/\delta$ intestinal T cell origin have been obtained in young or adult mice, but there is no information concerning the origin of $\gamma/\delta$ IEL during embryogenesis and the first weeks of life. The size of the mouse embryos practically precludes such study. The avian model system, by contrast, offers significant experimental advantages for exploration of this issue. Cells bearing the $\gamma/\delta$ TCR appear first in the chick embryo thymus where they reach a peak on days 14-15 of incubation, just 2 d before $\gamma/\delta$ T cells appear in the intestine (14-16), and studies of chick-quail chimeras suggest that embryonic $\gamma/\delta$ and $\alpha/\beta$ T cells are generated exclusively in the thymus (17). In the present studies, we have used two congenic chicken strains to analyze the intestinal colonization by $\gamma/\delta$ T cells before and after hatching. The data show that avian $\gamma/\delta$ IEL are primarily of thymic origin and indicate that this colonization occurs by waves of thymic $\gamma/\delta$ migrants having relatively long life spans.

Materials and Methods

Animals. Embryos of White Leghorn chicken strain H.B19 were derived from animals kept at our institute's farm in Gipf-Oberfrick.
Figure 1. Embryonic γ/δ thymocytes home to the intestine. Double immunofluorescence staining of frozen tissue section. (A) Thymus from day 16 H.B19ov⁺ embryos. (B and C) Small intestine from day 18 H.B19ov⁻ embryos injected at day 16 with day 14 ov⁺ thymocytes (B) illustrating submucosal lymphoid aggregates (chickens do not have true Peyer's patches, [8]), (C) correspond to intraepithelial lymphocytes, and (D) small intestine of a 1-mo-old chicken that was thymectomized 2 d after hatching and immediately injected with 14-d-old ov⁺ splenocytes. (Arrowheads) Donor IEL (ov⁺ γ/δ⁺). ×270.

Switzerland. Fertilized eggs were incubated at 38°C and 80% humidity in a ventilated incubator. The H.B19 strain was subdivided into congenic lines (H.B19ov⁺ and H.B19ov⁻). They can be distinguished by the ov antigen which is present on thymocytes and T cells only in H.B19ov⁺ animals, and which is recognized by the mAb I1-A-9 (18–20). The experimental animals were treated according to Swiss government veterinary guidelines.

Injection of Lymphoid Cells into Congenic Chickens. Embryonic day 13 (E13) bone marrow cells, E13 splenocytes, and E14 thymocytes (25 x 10⁶) were injected. Bone marrow and spleen from day 13 embryos do not contain detectable γ/δ⁺ cells. Injections of sorted populations of E14 thymocytes were also performed. In this case, thymocytes from 14-d-old H.B19ov⁺ embryos were suspended in PBS containing 10% FCS, filtered through a nylon sieve (mesh width of 25 μm; Nytal P-25 my, SST, Thal, Switzerland) and centrifuged at 255 g for 7 min. Immunofluorescence staining of these relatively fragile cells was performed in 96-well plates, to avoid repeated centrifugation, using the anti-γ/δ antibody TCR1.
Table 1. T Cell Marker Expression by ov+ Cells Derived from E14 Thymocyte Transplants in ov- Recipients

| Days of development | Percent intestinal cells bearing the donor ov antigen | CD3+ | CD8+ | CD4+ |
|---------------------|----------------------------------------------------|------|------|------|
|                     | γ/δ+                                               | α/β+ | CD3+ | CD8+ | CD4+ |
| 18                  | 82 ± 7                                             | 0    | 100 ± 2 | 37 ± 4 | 0    |
| 19                  | 75 ± 3                                             | 0    | 95 ± 4 | 41 ± 4 | 0    |
| 22                  | 74 ± 3                                             | 0    | 100 ± 3 | 40 ± 4 | 0.5 ± 0.5 |
| 28                  | 75 ± 3                                             | 0.5 ± 0.5 | 100 ± 3 | 42 ± 4 | 0.5 ± 0.5 |

Expression of T cell markers was determined by double immunofluorescence staining of gut frozen sections. Each slide contained a section of the duodenum, a section of the ileum, and a section located around the coecal tonsils. Counting was performed on two slides for each animal. 200 ov+ intestinal cells were counted per slide. Data are expressed in percent ov+ intestinal cells and correspond to the mean of three animals. Variability corresponds to SE. 0 is defined as <0.1%, i.e., no ov+ γ/δ+ T cells were detected in analyzed slides.

Results

H.B19ov+ chickens express ov antigen on the surface of most hemopoietic precursors during embryonic life, but only on T lineage cells and their precursors postnatally (18). Chickens of the congenic strain H.B19ov-, which do not express the ov antigen (18-20), were employed as recipients in these experiments. On day 14 of embryogenesis, ~25% of all thymocytes are γ/δ+ and <3% are α/β+, all of these being ov+ in the H.B19ov+ strain (23 and data not shown). Injection of day 14 H.B19ov+ thymocytes into 15- or 16-d-old H.B19ov- embryos led to the appearance of γ/δ+ T cells of the donor ov+ type in the small intestine within 2 d after injection (Fig. 1). The injected thymocytes homed to the intestine and the spleen, but neither thymus nor bursa were colonized at significant levels (Figs. 1 and 2 and data not shown). The percentage of donor γ/δ+ T cells in the intestine remained relatively constant until the end of embryogenesis (days 18, 19, and 20).

Donor ov+ E14 thymocytes homing to the gut expressed CD3 and >75% of these were identifiable as γ/δ+ (Table 1). The percentages of α/β+ and CD8+ T cells were <3% and 0.5 ± 0.5%, respectively (Table 1). The percentages of CD4+ T cells were 0.5 ± 0.5% (Table 1). These results were confirmed by FACScan® analysis (Fig. 1). The percentage of donor γ/δ+ T cells in the intestine remained relatively constant until the end of embryogenesis (days 18, 19, and 20).

Expression of T cell markers was determined by double immunofluorescence staining of gut frozen sections. Each slide contained a section of the duodenum, a section of the ileum, and a section located around the coecal tonsils. Counting was performed on two slides for each animal. 200 ov+ intestinal cells were counted per slide. Data are expressed in percent ov+ intestinal cells and correspond to the mean of three animals. Variability corresponds to SE. 0 is defined as <0.1%, i.e., no ov+ γ/δ+ T cells were detected in analyzed slides.

Figure 2. Survival pattern of donor γ/δ+ cells in intestine. Day 16 H.B19ov- embryos were injected with 25 x 10^6 day 14 H.B19ov+ thymocytes and killed at various times after injection. (Arrow) Hatching at day 21. Data are expressed as percent γ/δ+ IEL that express the donor ov antigen marker and correspond to the mean of two to four animals derived from three independent experiments. Analysis was performed in intestine (□) and thymus (▲). These results were obtained from two-color immunofluorescence analysis of tissue sections throughout development. Each slide contained a section of the duodenum, a section of the ileum, and a section located around the coecal tonsils. Counting was performed on two slides for each animal. 200 and 500 γ/δ IEL were counted per slide of embryonic intestine and newborn chick, respectively. By day 22 (after hatching) these results were confirmed by FACScan® analysis. Error bars correspond to SE.
Approximately 40% of the ov⁺ γ/δ⁺ T cells in the intestine were CD8⁺ and the remainder were CD4⁻CD8⁻, a phenotypic distribution similar to that of γ/δ T cells in the adult intestinal mucosa (15) which has also been observed in human intraepithelial mucosa (26–27). As expected at this early embryonic stage, significant homing of ov⁺ ot/fl⁺ E14 thymocytes to the gut was not observed. The donor γ/δ T cells persisted in the recipient intestine beyond 75 d, although a dramatic decrease in the percentage of donor intestinal γ/δ⁺ T cells occurred around hatching (Fig. 2). This decrease might result from dilution by the intestinal arrival of host γ/δ⁺ T cells. The quantitative analysis of ileal γ/δ⁺ T cells during their development, which revealed a significant increase in the total number of cells occurring in the first days after hatching (Fig. 3), favored this hypothesis. Alternatively, a loss of γ/δ⁻IEL due to the onset of digestive function could theoretically account for the observed decrease.

To analyze the origin of the posthatch emigrants to the intestine, we injected lymphoid cells into normal and thymectomized chickens after hatching (Table 2). Injection of 14-d-old H.B19ov⁺ embryonic thymocytes into 2-d-old H.B19ov⁻ thymectomized chickens led to the development of >20% ov⁺ cells among the γ/δ⁺ T cell population in the intestine during the first month after the injection. Injection of the same number of ov⁺ thymocytes into 2-d-old, non-thymectomized chickens led to only 3–9% of intestinal γ/δ⁺ T cells being of the donor type. These results suggest that cells derived from the recipient's thymus compete with the homing of the donor thymocytes. Moreover, γ/δ thymocytes from either 14-d-old H.B19ov⁺ embryos or 2-d-old H.B19ov⁺ chicks colonized the intestine of 2-d-old H.B19ov⁻ chicks with the same efficiency (data not shown). The progeny of stem cells of the second wave of thymus colonization (days 12–13 of embryogenesis) mature around the time of hatching (7–8 d later; [17, 28]). Thus, this second wave of γ/δ⁺ T cells homing to the intestine after hatching also appears to be thymus derived. In further support of this interpretation, injection of 35 × 10⁶ 14-d-old H.B19ov⁺ thymocytes into H.B19ov⁻ chicks at 1 wk of age, contributed a very small proportion of intestinal γ/δ⁺ T cells (0.5 ± 0.5%).

Injections of day 13 embryonic splenocytes or bone marrow cells into 16-d-old embryos did not contribute embryonic intestinal γ/δ⁺ T cells of donor origin. However, when embryonic day 13 splenocytes and bone marrow cells were injected into newly hatched thymectomized chickens, occasional donor γ/δ⁺ T cells could be found in the intestine of 6 of 15 recipients (Table 2 and Fig. 1). These intestinal γ/δ⁺ T cells of possible extrathymic origin were detected only 3–4 weeks after injection in thymectomized recipients and never in normal recipients.

To examine further the possibility that thymocyte precursors integrated into the thymic wall during embryogenesis, we injected thymus of E14 genotypes into 2-d-old H.B19ov⁻ chickens. The progeny of Ψ/δ⁺ T cells in the intestine of these recipients showed a lower proportion of E14 thymocytes than those of E13 bone marrow cells (Table 2). The developmental expression of Ψ/δ⁺ T cells in the intestine of recipients of E14 thymocytes was similar to that of E13 bone marrow cells, indicating that the integration of thymocyte precursors into the thymic wall during embryogenesis is not a common occurrence.
might differentiate extrathymically into γ/δ T cells, donor thymocytes from 14-d-old embryos were separated into γ/δ− and γ/δ+ populations. To yield an enrichment of thymocyte precursors, embryos at this development stage were chosen as donors since the second wave of precursor cells enters the thymus between embryonic days 12–14 (see Fig. 4). When sorted γ/δ− or γ/δ+ thymocytes were injected into 16-d-old embryos, the γ/δ+ thymocytes gave rise to γ/δ− intestinal lymphocytes of the donor type while the γ/δ− thymocyte population did not (Table 3). Indeed, when H.B19ov− chickens were injected with ov+ γ/δ− thymocytes, donor intestinal γ/δ+ T cells were still undetectable 2 mo after hatching.

Discussion

These results provide direct evidence for population of intestinal epithelium by emigrating γ/δ thymocytes. Our data indicate that the homing of γ/δ T cells from the thymus might differ from extrathymically into γ/δ T cells, donor thymocytes from 14-d-old embryos were separated into γ/δ− and γ/δ+ populations. To yield an enrichment of thymocyte precursors, embryos at this development stage were chosen as donors since the second wave of precursor cells enters the thymus between embryonic days 12–14 (see Fig. 4). When sorted γ/δ− or γ/δ+ thymocytes were injected into 16-d-old embryos, the γ/δ+ thymocytes gave rise to γ/δ− intestinal lymphocytes of the donor type while the γ/δ− thymocyte population did not (Table 3). Indeed, when H.B19ov− chickens were injected with ov+ γ/δ− thymocytes, donor intestinal γ/δ+ T cells were still undetectable 2 mo after hatching.

Table 3. Intestinal Homing of Sorted Congenic γ/δ+ or γ/δ− Thymocytes Injected Intravenously

| No. of cells injected | ov−γ/δ− thymocytes | ov−γ/δ− thymocytes |
|-----------------------|---------------------|---------------------|
| Intestinal colonization by γ/δ T cells/no. of recipients | 1.5 × 10^6 | 3 × 10^6 | 3 × 10^6 |
| 2/2 | 2/2 | 0/3 |

Sorted γ/δ+ and γ/δ− thymocytes from 14-d-old H.B19ov+ embryos were injected into 16-d-old H.B19ov− embryos. Embryos were killed at day 18 and γ/δ donor cells were analyzed by two-color immunofluorescence staining of intestine sections. Each slide contained a section of the duodenum, a section of the ileum, and a section located around the cecal tonsils. Counting was performed on three slides for each animal. 500 γ/δ IEL were counted per slide. By day 22 (after hatching) these results were confirmed by FACScan® analysis. Embryos injected with 1.5 × 10^6 and 3 × 10^6 ov−γ/δ+ thymocytes presented 13 and 5% of γ/δ IEL of the donor type. No γ/δ+ donor cells were detected in embryos injected with ov−γ/δ− thymocytes.

One of the major issues addressed in these experiments in the chicken concerns the possible extrathymic origin for γ/δ+ IEL suggested by studies in mice (9-12). Our results indicate that the avian γ/δ+ IEL population includes few, if any, γ/δ T cells that are derived from extrathymic sources. In all of the normal recipients and one half of the thymectomized recipients of embryonic (E13) bone marrow or spleen cells, we could not identify γ/δ+ IEL of donor origin. In some of the thymectomized recipients, however, we found γ/δ+ IELs of donor origin in very low frequencies at 1–2 mo of age. These γ/δ+ IELs could be of thymic origin since the donor bone marrow and splenic populations were obtained 1 or 2 d after the appearance of γ/δ− thymocytes. Indeed, when the γ/δ− fraction of 14-d embryonic thymocytes was infused as an enriched source of thymocyte precursors

Figure 4. A model of colonization of the chicken intestine by γ/δ T cells. Colonization of the embryonic thymus occurs in three waves and these T cell precursors mature in situ around 9 d later (12). Day 14–16 embryonic γ/δ thymocytes derived from the first wave of thymus colonization seed the intestine at days 16–18 of embryogenesis. Progeny of the second wave of stem cells colonizing the thymus should lead to the second wave of intestine colonization by γ/δ T cells, although we cannot exclude the possibility that some γ/δ thymocytes derived from the first wave of thymus colonization may contribute to the second wave of intestine colonization.
References

1. Viney, J., T.T. MacDonald, and J. Spencer. 1990. Gamma/delta T cells in the gut epithelium. Gut. 31:841.
2. Allison, J.P. 1991. Gamma delta T cells. Seminars in Immunology. Vol. 3. W.B. Saunders Company. 129 pp.
3. Lefrançois, L., and T. Goodman. 1989. In vivo modulation of cytoytic activity and thy-1 expression in TCR γ/δ+ intraepithelial lymphocytes. Science (Wash. DC). 243:1716.
4. Viney, J., P.J. Kilshaw, and T.T. MacDonald. 1990. Cytotoxic α/β+ and γ/δ+ T cells in murinintestinal epithelium. Eur. J. Immunol. 20:1623.
5. Bandeira, A., T. Motas-Santos, S. Isohara, S. Degermann, C. Heusser, S. Tonegawa, and A. Coutinho. 1990. Localization of γ/δ T cells to the intestinal epithelium is independent of normal microbial colonization. J. Exp. Med. 172:239.
6. Fujihashi, K., T. Taguchi, J.R. McGhee, J.H. Eldridge, M.G. Bruce, D.R. Green, B. Singh, and H. Kiyono. 1990. Regulatory function for murine intraepithelial lymphocytes. J. Immunol. 145:2010.
16. Bucy, K.P., C.L. Chert, and M.D. Cooper. 1990. Ontogeny of T cell receptor-positive (TCR+) T cells abrogate oral tolerance, while α/β TCR+ T cells provide B cell help. J. Exp. Med. 175:695.

17. Coltey, M., K.P. Bucy, C.H. Chen, J. Cihak, U. IAsch, D. Guy-Grand, D., N. Cerf-Bensussan, B. Malissen, M. Malassis-Seris, C. Briottet, and P. Vassali. 1991. Two gut intraepithelial CD8+ lymphocyte populations with different T cell receptors: a role for the gut epithelium in T cell differentiation. J. Exp. Med. 173:471.

18. Vainio, O., T.V. Veromaa, E. Eerola, and P. Toivanen. 1992. Differential expression of two T cell receptors, TcR1 (γ/δ+) and TcR2 (α/β) in the human intestinal mucosa. Immunology. 68:7.

19. Bucy, R.P., C.H. Chen, L.L. Ager, and M.D. Cooper. 1988. Identification of the avian homologues of mammalian CD4 and CD8 antigens. J. Immunol. 140:2133.

20. Bucy, R.P., C.H. Chen, and M.D. Cooper. 1989. T cell receptor genes of chicken T cell receptor TcRα T cells in the skin. Eur. J. Immunol. 19:1449.

21. Coltey, M., F.V. Jotereau, and N.M. Le Douarin. 1987. Evidence for a cyclic renewal of lymphocyte precursor cells in the embryonic chick thymus. Cell Differ. 22:71.

22. Coltey, M., F.V. Jotereau, and N.M. Le Douarin. 1987. Establishment of a T3+T cell receptor complex in chickens. J. Exp. Med. 164:375.

23. Chan, M.M., C.H. Chen, L.L. Ager, and M.D. Cooper. 1988. Identification of the avian homologues of mammalian CD4 and CD8 antigens. J. Immunol. 140:2133.

24. Fujihashi, K., T. Taguchi, W.K. Aicher, J.R. McGhee, J.A. Bluestone, J.H. Eldridge, and H. Kiyono. 1992. Immunoregulatory functions for murine intraepithelial lymphocytes: γ/δ T cell receptor-positive (TCR+) T cells abrogate oral tolerance, while α/β TCR+ T cells provide B cell help. J. Exp. Med. 175:695.

25. Fujihashi, K., T. Taguchi, W.K. Aicher, J.R. McGhee, J.A. Bluestone, J.H. Eldridge, and H. Kiyono. 1992. Immunoregulatory functions for murine intraepithelial lymphocytes: γ/δ T cell receptor-positive (TCR+) T cells abrogate oral tolerance, while α/β TCR+ T cells provide B cell help. J. Exp. Med. 175:695.

26. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

27. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

28. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

29. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

30. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

31. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

32. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

33. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

34. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

35. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

36. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

37. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

38. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

39. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

40. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

41. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

42. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

43. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

44. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

45. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.

46. Goodenough, D.U., A.C. Hayday. 1990. Extrathymic selection of TcR 3~/~ T cells in athymic mice. Immunology. 63:111.