Dual Suppression of the Cyclin-Dependent Kinase Inhibitors CDKN2C and CDKN1A in Human Melanoma

Citation
Jalili, Ahmad, Christine Wagner, Mikhail Pashenkov, Gaurav Pathria, Kirsten D. Mertz, Hans R. Widlund, Mathieu Lupien, et al. 2012. Dual suppression of the cyclin-dependent kinase inhibitors CDKN2C and CDKN1A in human melanoma. JNCI Journal of the National Cancer Institute 104(21): 1673-1679.

Published Version
doi:10.1093/jnci/djs373

Permanent link
http://nrs.harvard.edu/urn-3:HUL.InstRepos:10581966

Terms of Use
This article was downloaded from Harvard University’s DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

Share Your Story
The Harvard community has made this article openly available. Please share how this access benefits you. Submit a story.

Accessibility
Dual Suppression of the Cyclin-Dependent Kinase Inhibitors CDKN2C and CDKN1A in Human Melanoma

Ahmad Jalili, Christine Wagner, Mikhail Pashenkov, Gaurav Pathria, Kirsten D. Mertz, Hans R. Widlund, Mathieu Lupien, Jean-Philippe Brunet, Todd R. Golub, Georg Stingl, David E. Fisher, Sridhar Ramaswamy, Stephan N. Wagner

Resistance to BRAFV600E inhibitors is associated with reactivation of mitogen-activated protein kinase (MAPK) signaling at different levels in melanoma. To identify downstream effectors of MAPK signaling that could be used as potential additional therapeutic targets for BRAFV600E inhibitors, we used hTERT/CDK4R24C/p53DD-immortalized primary human melanocytes genetically modified to ectopically express BRAFV600E or NRASG12D and observed induction of the AP-1 transcription factor family member c-Jun. Using a dominant negative approach, in vitro cell proliferation assays, western blots, and flow cytometry showed that MAPK signaling via BRAFV600E promotes melanoma cell proliferation at G1 through AP-1-mediated negative regulation of the INK4 family member, cyclin-dependent kinase inhibitor 2C (CDKN2C), and the CIP/KIP family member, cyclin-dependent kinase inhibitor 1A (CDKN1A). These effects were antagonized by pharmacological inhibition of CDKN2C and CDKN1A targets CDK2 and CDK4 in vitro. In contrast to BRAFV600E or NRASG12D-expressing melanocytes, melanoma cells have an inherent resistance to suppression of AP-1 activity by BRAFV600E- or MEK-inhibitors. Here, CDK2/4 inhibition statistically significantly augmented the effects of BRAFV600E-, or MEK-inhibitors on melanoma cell viability in vitro and growth in athymic nude Foxn1nu mice (P = .03 when mean tumor volume at day 13 was compared for BRAFV600E inhibitor vs BRAFV600E inhibitor plus CDK2/4 inhibition; P = .02 when mean tumor volume was compared for MEK inhibitor vs MEK inhibitor plus CDK2/4 inhibition; P values were calculated by a two-sided Welch t test; n = 4–8 mice per group).

J Natl Cancer Inst 2012;104:1673–1679
Figure 1. Mitogen-activated protein kinase, AP-1 activity, and proliferation of human melanocytic cells. A) Results of western blots for c-Jun and phosphorylated c-Jun (p-cJun) protein expression levels in primary immortalized human melanocytes (hTERT/C4(R24C)/p53DD) with or without ectopic expression of a BRAFV600E or NRASG12D are shown (left panel). AP-1 activity in these cells was measured by AP-1-secreted alkaline phosphatase reporter gene assay after treatment with the MEK inhibitor PD098059 (50 µM) or dimethyl sulfoxide (right panel). Untreated cells served as an additional control. Results are representative of two independent experiments performed in triplicate. B) AP-1 activity was also measured in the NCI-60 BRAFV600E human melanoma LOXIMVI cell line, stably expressing dominant negative AP-1 and a puromycin resistance gene (-dnAP-1) or the resistance gene alone (-empty vector) with (0.75 µg/mL puromycin) and without induction (0.25 µg/mL puromycin) of the transgene for 48 hours. Whisker bars indicate the SD. Results are representative of three independent experiments performed in triplicate. C) Cell proliferation of LOXIMVI-dnAP-1 cells upon induction of dnAP-1 as determined by cell numbers over time. The means and corresponding SD (whisker bars) of experiments performed in triplicate.
Transfection of the LOXIMVI melanoma cell line with a dominant negative c-Jun mutant (dnAP-1), which leads to a broader inhibition of AP-1 activity by binding additional AP-1 members compared with c-Jun siRNA (13), was done with the bicistronic pIRESpuro3 vector (Clontech Laboratories, Mountain View, CA) to stably express a puromycin resistance gene with or without a FLAG-tagged dnAP-1 (14) (referred to hereafter as -dnAP-1 and -empty vector cells, respectively). When cultured in a low concentration of puromycin (0.25 μg/mL), LOXIMVI-dnAP-1 cells expressed low levels of the resistance gene and dnAP-1 without an impact on AP-1 activity (Figure 1, B), cell proliferation, or cell cycle distribution (data not shown) compared with LOXIMVI-empty-vector cells. When cultured at a high concentration of puromycin (0.75 μg/mL), LOXIMVI-dnAP-1 cells expressed high levels of dnAP-1 in the cytoplasm and the nucleus (Supplementary Figure 1, C and D, available online) and showed decreased AP-1 activity (mean AP-1 activity = 26.5%, SD = 16.5% vs 75.3%, SD = 8.3%, respectively, two-sided P = .02) (Figure 1, B), decreased cell proliferation (mean cell number at day 5 = 4.1 x 10^4, SD = 0.2 x 10^4 vs 37.9 x 10^4, SD = 0.6 x 10^4, respectively, two-sided P < .001) (Figure 1, C), accumulation of cells in G1 (Figure 1, D), and decreased [3H]thymidine uptake (data not shown) compared with culturing at a low concentration of puromycin.

To investigate the effect of AP-1 inhibition in melanoma cells in vivo, 1 x 10^6 LOXIMVI-dnAP-1 cells were injected subcutaneously into female, 6–8-week-old, athymic nude Foxn1nu mice (Supplementary Methods, available online). Animal care procedures followed the guidelines of the Animal Research Committee of the Medical University of Vienna. Injection of LOXIMVI-dnAP-1 xenografts with 0.75 μg/mL puromycin every other day inhibited tumor growth compared with xenografts injected with 0.25 μg/mL puromycin (Figure 1, E), indicating that AP-1 activity is required for in vivo growth of human melanoma cells.

Because the effects of induced expression of dnAP-1 on cell proliferation (by counting cell numbers over time, Figure 1, C and Supplementary Figure 1, E, available online) and cell cycle distribution (not shown) were similar in human UACC257 melanoma cells and LOXIMVI cells that are cyclin-dependent kinase inhibitor CDKN2A-deficient, we investigated the effect of dnAP-1 on the expression of other cell cycle regulators at G1 by western blot. We found that CDKN2C and CDKN1A protein levels in LOXIMVI-dnAP-1 cells were increased relative to LOXIMVI empty vector cells within 2 hours of induction of dnAP-1 with puromycin (Figure 1, F). CDKN2D, CCND1, and CDK6 protein levels increased later at 12 and 16 hours after dnAP-1 induction in both LOXIMVI-dnAP-1 and LOXIMVI-empty-vector cells. CDK2 and CDK4 protein levels remained almost unchanged within 24 hours (Figure 1, F and Supplementary Figure 1, F, available online). In addition, we observed nuclear and cytoplasmic accumulation of CDKN2C and nuclear accumulation of CDKN1A by immunofluorescence in LOXIMVI-dnAP-1 cells 48 hours after induction of dnAP-1 using puromycin (Supplementary Figure 1, G and H, available online).

Consistent with these findings, transfection of LOXIMVI cells with siRNA targeting CDKN2C or CDKN1A in vitro partially rescued dn-AP1-induced suppression of cell proliferation, and when both CDKN2C and CDKN1A siRNAs were cotransfected into LOXIMVI cells, cell proliferation (as measured by counting cell numbers over time, which is shown in Figure 1, G, and [3H]thymidine uptake [data not shown]) was similar to that of LOXIMVI-empty vector control cells. Similar effects were not seen when siRNA targeting other cell cycle regulators (TP53, CDKN1B, CDKN1C alone or in combination with CDKN2C) was used (data not shown). These results indicate that the full proproporative effect of AP-1 on melanoma cells requires suppression of both the INK4 family member CDKN2C and the CIP/KIP family member CDKN1A. This finding supports a previous report in which the ability of CDK4βR2C (an INK4-inhibitive CDK4 mutant) to rescue Cldn1α−/− but not Cldn1α−/− cells from growth arrest (15), an important prerequisite for cell transformation, was described. Furthermore, mutations in CDK4 have been described in melanoma-prone families and patients with multiple primary melanomas (16), CDK4 amplification has been previously reported in subtypes of sporadic melanoma (17), and reduced CDKN1A expression has been implicated in melanoma metastasis (18).

As AP-1 is a transcription factor, we performed cotransfection assays with wild-type CDKN2C and CDKN1A promoter-luciferase reporter plasmids (19,20) in LOXIMVI-dnAP-1 and LOXIMVI-empty vector cells. When cultured at a high concentration of puromycin (0.75 μg/mL) for 4 hours, LOXIMVI-dnAP-1 cells showed activated expression from both promoter reporter plasmids compared with LOXIMVI-empty vector cells (mean fold CDKN2C promoter activation signals for Prom#1, EnhU, and EnhD regions were normalized to the β-actin promoter region (Prom#1, EnhU, and EnhD) regions (left panel). Quantitative polymerase chain reaction signals for Prom#1, EnhU, and EnhD regions were normalized to the input and expressed as the mean with SD (whisker bars). As a control, the anti-c-Jun antibody was substituted with IgG (right panel). The results are representative of two independent experiments.

Figure 1. (Continued)
Figure 2. Melanoma cell viability and in vivo growth by cyclin-dependent kinase 2/4 inhibition. A) Cell viability was assessed by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium-bromide assay of human melanoma cell lines (white, gray, and black bars indicate BRAFwt/NRASwt, NRASQ61R, and BRAFV600E cell lines, respectively) treated with a CDK2/4 inhibitor combination (2 µM CVT313 plus 0.25 µM indolocarbazole CDK4-I) for 48 hours. The values are presented as the percentage of treatment with dimethyl sulfoxide (DMSO, solvent). Whisker bars indicate the SD (left panel). On-target effects of CDK2/4 inhibitors in LOXIMVI cells are shown in the right panel: cell cycle analysis by flow cytometry was done after treatment with DMSO (solvent) control or the CVT313/indolocarbazole CDK4-I combination for 12 hours; western blot analysis was also done to detect phosphorylated retinoblastoma (p-RB) (Thr826) levels after treatment with DMSO (solvent) control, 2 µM CVT313, or 0.25 µM indolocarbazole CDK4-I for 48 hours. These experiments were performed three
induction for LOXIMVI-dnAP-1 cells vs LOXIMVI-empty vector cells = 3.1, SD = 0.2 vs 1.0, SD = 0.02, two-sided \( P = .002 \); mean fold CDKN1A promoter induction for LOXIMVI-dnAP-1 cells vs LOXIMVI-empty vector cells = 1.7, SD = 0.1 vs 1, SD = 0.1, two-sided \( P < .001 \) (Supplementary Methods and Supplementary Figure 2, A, available online), further supporting the link between AP-1 activity and CDKN2C and CDKN1A expression. As dn-AP-1 induced CDKN2C mRNA in LOXIMVI-dnAP-1 cells (by quantitative reverse transcription–real-time PCR, data not shown) independent of protein synthesis (by cycloheximide, data not shown), we performed chromatin immunoprecipitation from LOXIMVI cells with an anti-c-Jun antibody followed by polymerase chain reaction using primer sets spanning putative AP-1-binding sites at the CDKN2C gene (21) (Supplementary Methods, available online). We observed binding of AP-1 at two promoter-distant regions upstream and downstream of CDKN2C, each harboring a 12-O-tetradecanoyl-L-lysine response element-binding motif, and one region within the promoter (Figure 1, H). These results indicate that CDKN2C is a direct target of AP-1.

In addition, we assessed the relationship between c-Jun expression and that of CDKN2C and CDKN1A using RNA and tissues obtained from 30 melanoma patients. The primary human melanomas were assigned to two groups (low and high) based on low vs high expression of CDKN2C and CDKN1A (mean relative CDKN2C mRNA expression = 0.5, SD = 0.3 vs 2.5, SD = 1.8, respectively, two-sided \( P = .007 \); mean relative CDKN1A mRNA expression = 0.9, SD = 0.4 vs 3.0, SD = 1.2, respectively, two-sided \( P < .001 \); n = 8 and 11, respectively) as determined by quantitative real-time PCR of available RNA (Supplementary Methods, available online). Nuclear phospho-c-Jun expression in the corresponding tissues was then analyzed by immunohistochemistry (Supplementary Methods, available online), and the percentage of cells with 0–1+, 2+, and 3+ staining was determined. A table summarizing the data and representative stained tissues are depicted in Supplementary Figure 2, B (available online). High levels of CDKN2C and CDKN1A mRNA were associated with low phospho-c-Jun staining (0–1+), whereas low levels of CDKN2C and CDKN1A mRNA were associated with high phospho-c-Jun staining (2+ and 3+). These results further substantiate the role of AP-1 in the negative regulation of CDKN2C and CDKN1A transcription. In accordance with these results, previous studies in animal models have shown that CDK4R24C and a CDKN2C deficiency increase melanoma susceptibility, but additional MAPK signaling is required for melanomas to develop (22,23).

Because the expression of CDKN2C and CDKN1A targets, ie, Cdk2 and Cdk4/6, was unaffected by dn-AP-1 (Supplementary Figure 1, F, available online) and is rarely lost in human melanoma (24), our results provide rationale for the development of novel combination therapeutic strategies for melanoma. In contrast to single agent-treatment with inhibitors at doses selective to Cdk2 or Cdk4 inhibition [NU6140 (25), CVT-313 (26), NSC625987 (27), indolocarbazole CDK4-I (28)] (Supplementary Methods and Supplementary Figure 2, C, available online), the combination of Cdk2/4 inhibitors reduced viability in a panel of melanoma cell lines (by 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium-bromide assay) (Figure 2, A) and statistically significantly the growth of LOXIMVI xenografts in vivo (mean tumor volume at day 13: vehicle only = 1.5 x 10^3 mm^3, SD = 0.5 x 10^3 mm^3; 0.75/0.1 mg/kg dose level = 0.9 x 10^3 mm^3, SD = 0.2 x 10^3 mm^3; 1.5/0.2 mg/kg dose level = 0.6 x 10^3 mm^3, SD = 0.3 x 10^3 mm^3; vehicle vs 0.75/0.1 mg/kg dose, two-sided \( P = .05 \); vehicle vs 1.5/0.2 mg/kg dose, two-sided \( P = .01 \); n = 6–8 mice per group) (Figure 2, B). Reduction of viability was independent of the presence or absence of BRAFV600E/NRASQ61K as nuclear phospho-c-Jun and CDKN2C/CDKN1A transcript levels in primary melanomas. Furthermore, in BRAFV600E melanoma cells, the highly selective BRAFV600E inhibitor GDC-0879 (29) and three selective MEK inhibitors [PD184352/C21-1040 (30), U0126 (31), PD98059 (12)] did not suppress c-Jun levels, although they effectively reduced phospho-ERK levels (Figure 2, C). Together these data suggest that in melanoma cells, in contrast to melanocytes, pathways that bypass the BRAF-MEK-ERK axis to induce AP-1 are operative. Consistent with this hypothesis, AP-1 and Cdk2/4 inhibition increased the magnitude of the reduction of melanoma cell viability/proliferation by BRAFV600E inhibitor GDC-0879 and MEK inhibitor PD184352/CI1040 in vitro (Figure 2, D and Supplementary Figure 2, D, available online).
online), and CDK2/4 inhibition augmented statistically significant growth reduction of melanoma xenografts in vivo by the BRAF and MEK inhibitors (P = .03 when mean tumor volume at day 13 was compared for GDC-0879 vs GDC-0879 plus CDK2/4 inhibition; P = .02 when mean tumor volume was compared for PD184352/CI1040 vs PD184352/CI1040 plus CDK2/4 inhibition; n = 4–8 mice per group) (mean tumor volume at day 13: GDC-0879 = 0.7 × 10^3 mm^3, SD = 0.2 × 10^3 mm^3; GDC-0879 plus CDK2/4 inhibition = 0.3 × 10^3 mm^3, SD = 0.2 × 10^3 mm^3; GDC-0879 vs GDC-0879 plus CDK2/4 inhibition = 0.3 × 10^3 mm^3, SD = 0.2 × 10^3 mm^3; GDC-0879 vs PD184352/CI1040 plus CDK2/4 inhibition, two-sided P = .02; n = 4–8 mice per group) (Figure 2, E and F).

Our study was not without limitations. The restricted ability of cell-based studies to predict clinical behavior is an inherent restraint. Also, the varied response to cyclin-dependent kinase inhibitors between different melanoma cell lines is unexplained at this time. The small number of available human tissue samples used in our study also limits the interpretation of our results.

Nevertheless, our data show a statistically significant augmentation of BRAF^V600E- and MEK-inhibitors by CDK2/4 inhibition in vivo. Our findings provide rationale and support for further clinical exploration of this novel combination therapeutic strategy for melanoma.

References

1. Chapman PB, Hauschuld A, Robert C, et al.; BRIM-3 Study Group. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. N Engl J Med. 2011;364(26):2507–2516.
2. Flaherty KT, Puzanov I, Kim KB, et al. Inhibition of mutated, activated BRAF in metastatic melanoma. N Engl J Med. 2010;363(5):493–503.
3. Wolchok JD, Puzanov I, McArthur JG, et al. Nivolumab in previously treated melanoma. N Engl J Med. 2013;369(2):122–133.
4. Chapman PB, Hauschild A, Robert C, et al.; BRIM-3 Study Group. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. N Engl J Med. 2011;364(26):2507–2516.
5. Nazarian R, Shi H, Wang Q, et al. Melanomas acquire resistance to B-RAF(V600E) inhibition by RTK or N-RAS upregulation. Nature. 2010;468(7326):973–977.
6. Wagle N, Emery C, Berger MF, et al. Dissecting therapeutic resistance to RAF inhibition in melanoma by tumor genomic profiling. J Clin Oncol. 2011;29(22):3085–3096.
7. Johannessen CM, Boehm JS, Kim SY, et al. COT drives resistance to RAF inhibition through MAP kinase pathway reactivation. Nature. 2010;468(7326):968–972.
8. Villanueva J, Vultur A, Lee JT, et al. Acquired resistance to BRAF inhibitors mediated by a RAF kinase switch in melanoma can be overcome by cotargeting MEK and IGF-1R/PI3K. Cancer Cell. 2010;18(6):683–695.
9. Jané-Valbuena J, Wiliandt HR, Perner S, et al. An oncogenic role for ETV1 in melanoma. Cancer Res. 2010;70(5):2075–2084.
10. Eferl R, Wagner EF. AP-1: a double-edged sword in tumorigenesis. Nat Rev Cancer. 2003;3(11):859–868.
11. Lopez-Bergami P, Huang C, Goydos JS, et al. Rewired ERK-RNK signaling pathways in melanoma. Cancer Cell. 2007;11(5):447–460.
12. Dudley DT, Pang L, Decker SJ, Bridges AJ, Saltiel AR. A synthetic inhibitor of the mitogen-activated protein kinase cascade. Proc Natl Acad Sci U S A. 1995;92(17):7686–7689.
13. Ham J, Babij C, Whinfield J, et al. A c-Jun dominant-negative mutant protects sympathetic neurons against programmed cell death. Neurorn. 1995;14(5):927–939.
14. Hennigsn RF, Stambrook PJ. Dominant negative c-jun inhibition activates the cyclin D1 and cyclin E kinase complexes. Mol Biol Cell. 2001;12(8):2352–2363.
15. Quereda V, Martinalbo J, Dubus P, Carnero A, Lopez-Bergami P, Huang C, Goydos JS, et al. Inhibition of mutated, activated BRAF in melanoma by tumor genomic profiling. J Clin Oncol. 2010;28(22):3159–3171.
16. Holland EA, Schmid H, Kefford RF, Mann GI, CDKN2A (P16[INK4a]) and CDK4 mutation analysis in 131 Australian melanoma probands: effect of family history and multiple primary melanomas. Genes Chromosomes Cancer. 1999;25(4):339–348.
17. Curtin JA, Fridyland Y, Kageshita T, et al. Distinct sets of genetic alterations in melanoma. N Engl J Med. 2005;353(20):2135–2147.
18. Maelandsmo GM, Holm R, Fodstad O, Kerbel RS, Floresen VA. Cyclin kinase inhibitors p21 and INK4a inhibitors in cellular senescence and tumor suppression. Oncogene. 2007;26(55):7665–7674.
19. Scher HI, gros H, Flaherty KT, et al. Improved survival with vemurafenib in metastatic melanoma. N Engl J Med. 2011;364(26):2507–2516.
20. Matsuzaki Y, Takao T, Hitomi T, Nishino H, Sakai T. Activation of protein kinase C beta 1 and C protein kinase C gamma 1 is involved in the process of mitogen-activated protein kinase (MAPK) pathway reactivation in human melanoma cells. J Biol Chem. 2005;280(2):1126–1132.
21. Carroll JS, Meyer CA, Song J, et al. Genome-wide analysis of estrogen receptor binding sites. Nat Genet. 2006;38(11):1289–1297.
22. Sotillo R, García JF, Ortega S, et al. Invasive melanoma in CdK4-targeted mice. Proc Natl Acad Sci USA. 2001;98(23):13312–13317.
23. Hacker F, Muller HK, Irwin N, et al. Spontaneous and UV radiation-induced multiple metastatic melanomas in Cdk4R24C/R24C/TPras mice. Cancer Res. 2006;66(6):2946–2952.
24. Hafalan R, Cheng E, Zhang Y, Mandigo CE, Miglarese MR. Release of cell cycle constraints in mouse melanocytes by over-expressed mutant E2F1E12, but not by deletion of p16INK4a or p21WAF1/CIP1. Oncogene. 1998;16(19):2489–2501.
25. Pennati M, Campbell AJ, Curto M, et al. Potentiation of paclitaxel-induced apoptosis by the novel cyclin-dependent kinase inhibitor NU6461: a possible role for surviving down-regulation. Mol Cancer Ther. 2005;4(9):1328–1337.
26. Brooks EE, Gray NS, Joly A, et al. CVT-313, a specific and potent inhibitor of CDK2 that prevents neointimal proliferation. J Biol Chem. 1997;272(46):29207–29211.
27. Kubo A, Nakagawa K, Varma RK, et al. The p16 status of tumor cell lines identifies small molecule inhibitors specific for cyclin-dependent kinase 4. Clin Cancer Res. 1999;5(12):4279–4286.
28. Zhu G, Conner SE, Zhou X, et al. Synthesis, structure-activity relationship, and biological studies of indolocarbazoles as potent cyclin D1-CDK4 inhibitors. J Med Chem. 2003;46(11):2027–2030.
29. Hoesfich KP, Herter S, Tien J, et al. Antitumor efficacy of the novel RAF inhibitor GDC-0879 is predicted by BRAFV600E mutational status and sustained extracellular signal-regulated kinase/mitogen-activated protein kinase pathway suppression. Cancer Res. 2009;69(7):3042–3051.
30. Sebolt-Leopold JS, Dudley DT, Herrera R, et al. Blockade of the MAP kinase pathway suppresses growth of colon tumors in vivo. Nat Med. 1999;5(7):810–816.
31. Duncia JV, Santos JA, Gilley CA, et al. MEK inhibitors: the chemistry and biological activity of U0126, its analogs, and cyclization products. Bioorg Med Chem Lett. 1998;8(20):2839–2844.

Funding

The research was supported by the FWF-Austrian Science Fund (L590-B12 to SNW) and by a grant from the Austrian Society of Dermatology and Venerology (to AJ).

Notes

The funders did not have a role in the study design; data collection, analysis, or interpretation; the writing of the brief communication; or the decision to submit the brief communication for publication. T. R. Golub is a consultant of Merck-EMD Serono, Inc, Pfizer, Inc, and holds stocks/stock options with H3 Biomedicine.

Affiliations of authors: Division of Immunology, Allergy and Infectious Diseases, Department of Dermatology, Medical University of Vienna (AJ, UW, NP, SMP, CWM, KS, SNW) and CeMM-Research Center for Molecular Medicine of the Austrian
Academy of Sciences, Vienna, Austria (SNW); Laboratory of Clinical Immunology, National Research Center Institute of Immunology, Federal Medical and Biological Agency, Moscow, Russia (MP); Department of Pediatric Oncology, Dana-Farber Cancer Institute and Brigham and Women’s Hospital and Department of Dermatology, Harvard Medical School, Boston, MA (HRW); Department of Genetics, Norris Cotton Cancer Center, Dartmouth Medical School, Lebanon, NH (ML); The Broad Institute of Harvard University and Massachusetts Institute of Technology, Cambridge, MA (JPB, TRG, SR); Department of Pediatric Oncology and Center for Genome Discovery, Dana-Farber Cancer Institute, Boston, MA (TRG); Howard Hughes Medical Institute, Chevy Chase, MD (TRG); Department of Dermatology, Cutaneous Biology Research Center, Melanoma Program in Medical Oncology, Harvard Medical School, Boston, MA (DEF); Massachusetts General Hospital Cancer Center, Boston, MA (SR); Harvard Stem Cell Institute, Cambridge, MA (SR).