Monsters with a shortened vertebral column: A population phenomenon in radiating fish *Labeobarbus* (Cyprinidae)

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Abstract

The phenomenon of a massive vertebral deformity was recorded in the radiating *Labeobarbus* assemblage from the middle reaches of the Genale River (south-eastern Ethiopia, East Africa). Within this sympatric assemblage, five trophic morphs—generalized, lipped, piscivorous and two scraping feeders—were reported between 1993 and 2019. In 2009, a new morph with prevalence of \(\sim 10\%\) was discovered. The new morph, termed ‘short’, had an abnormally shortened vertebral column and a significantly deeper body. This type of deformity is common in farmed Atlantic salmon and other artificially reared fish, but is rare in nature. In the Genale *Labeobarbus* assemblage, the deformity was present exclusively within the generalized and lipped morphs. The short morph had between seven and 36 deformed (compressed and/or fused) vertebrae. Their body depth was positively correlated with number of deformed vertebrae. In another collection in 2019, the short morph was still present at a frequency of 11\%. Various environmental and genetic factors could contribute to the development of this deformity in the Genale *Labeobarbus*, but based on the available data, it is impossible to confidently identify the key factor(s). Whether the result of genetics, the environment, or both, this deep-bodied phenotype is assumed to be an anti-predator adaptation, as there is evidence of its selective advantage in the generalized morph. The Genale monstrosity is the first reported case of a massive deformity of the vertebral column in a natural population of African fishes.

“We have also what are called monstrosities; but they graduate into varieties. By a monstrosity I presume is meant some considerable deviation of structure in one part, either injurious to or not useful to the species, and not generally propagated. If it could be shown that monstrosities were even propagated for a succession of generations in a state of nature, modifications might be effected (with the aid of natural selection) more abruptly than I am inclined to believe they are.”

Darwin (1860, pp. 46, 426).
Introduction

The emergence and establishment of morphological novelties in a population is central to morphological evolution, although this process remains poorly understood. As initially highlighted by Darwin [1], there is no clear discrimination between morphological abnormalities (monstrosities) and regular variation. Morphological abnormalities that are caused by genetic factors—and hence may be shaped by natural selection—are particularly interesting to evolutionary biologists as potential novelties.

Ray-finned fishes form one of the major vertebrate groups, comprised of more than 35,000 species with extreme variations in morphology and body plans [2]. For example, the ocean sunfishes (Molidae, Tetraodontiformes) exhibit one of the most impressive evolutionary transformations of the axial skeleton among ray-finned fishes [3, 4]. It has been suggested that the enigmatic loss of caudal fin and caudal part of the vertebral column is related to the existence of a number of malformed adult tetraodontiforms without caudal fins [5, 6].

Various body and skeletal deformities in fish have been reviewed in many publications [7–16]; earlier reports can also be found in Dawson’s bibliographies [17–20]. Many of these deformities apparently can be treated as monstrosities, particularly when the vertebral column is shortened without pronounced curvature, resulting in an altered body form that is extremely short and deep. Such deformities are known in at least 26 species of 15 families (S1 Table in S1 File). Remarkably, all individuals of Aischgrunder Karpfen, the South German carp breed (extinct since 1956), exhibited such a deformity [21, 22].

The riverine adaptive radiations in the large African barbs of the genus *Labeobarbus* Rüppell 1835 (Cyprinidae)—the dominating fish group in the waters of the Ethiopian Highlands—have been studied by us for almost 30 years. *Labeobarbus* belongs to the African Torinae, a lineage of hexaploids [23–27] that originated in the Middle East via hybridization of tetraploids (maternal *Tor* lineage) and diploids (paternal *Cyprinion* lineage), and then dispersed throughout Africa [28]. Although the continental-scale phylogeny of *Labeobarbus* is still poorly resolved [29], the mt-DNA based phylogeny of this group in Ethiopian waters is relatively well studied [30–35]. It is important to note that the barbs inhabiting the southeastern part of Ethiopia (drained by the Wabi-Shebele and Juba river systems into the Indian Ocean) form a monophyletic group, the *Labeobarbus gananensis* (Vinciguerra, 1895) complex. This is a sister group to *Labeobarbus* inhabiting enclosed basins of the Ethiopian Rift Valley, as well as all *Labeobarbus* from the waters of western and northern Ethiopia belonging to the Omo-Turkana and Nile systems, and additional some Kenyan barbs [30, 34, 35].

In addition to the famed *Labeobarbus* radiation in Lake Tana [36–66], this group has also experienced parallel riverine radiations in four Ethiopian basins. These riverine radiations are similar in terms of their morphological and ecological differentiation, but differ in the ways of genetic divergence [35]. Outside of Ethiopia, the riverine *Labeobarbus* radiation seems to occur in the Inkisi River basin (Lower Congo) [67].

The *Labeobarbus* assemblage in the Genale River (the main tributary of the Juba River) is the only radiation occurring in southeastern Ethiopia. It is geographically separated from the other Ethiopian *Labeobarbus* radiations by the Ethiopian Rift valley. It exhibits greater divergence in morphology, ecology and genetics than other riverine radiations [35]. During an intensive sampling of the middle Genale River in 2009, we observed a substantial portion of barbs with extremely short and deep bodies [34].

In 1993, an assemblage of the sympatric morphologically distinct morphs of the *L. gananensis* complex was discovered in the middle reaches of the Genale River [68]. This area was re-sampled twice in the 1990s [69, 70], then in 2009 [30, 34], and again in 2019. Together with the omnivorous generalized morph of *L. gananensis* and the scraping-feeder *L. jubae*
(Banister, 1984), three additional morphs specialized in their morphology and feeding habits were always present in these samples from the middle Genale. These include the lipped morph of *L. gananensis* (having hypertrophied lips), the scraping morph *Labeobarbus* sp. 1 (provisionally called ‘smiling’) and the large-mouthed morph *Labeobarbus* sp. 2 (called ‘piscivorous’), as well as a hybrid morph called ‘smiling hybrids’. In 2009, a new morph, ‘short’, was discovered, which has an abnormally short and deep body (Fig 1). It composed approximately 10% of the total barb catch in both 2009 and 2019 samplings [34].

Trophic resource partitioning has been demonstrated among most morphs from the middle Genale *Labeobarbus* assemblage [34]. In contrast to the other morphs, the short morph does not diverge from the generalized morph in its diet [34] although it demonstrates some variation in lip development (Fig 2). Moreover, based on mtDNA markers, the short morph shares a common gene pool with the generalized and lipped morphs [34]. Preliminary investigation of the vertebral column revealed that each short individual had a substantial number of deformed (compressed and/or fused) vertebrae (Fig 1).

This study aimed at: (1) investigating the structure of the vertebral column in the short morph compared to other sympatric *Labeobarbus* morphs from the Genale River assemblage, (2) testing a hypothesis on the correlation between the number of deformed vertebrae and body depth, (3) analyzing other morphological differences and some life-history traits between

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**Fig 1. External appearance (left) and x-ray images (right) of the generalized and short *Labeobarbus* morphs from the middle Genale assemblage.**

A: generalized (SL = 242 mm), B: short, with eight deformed vertebrae, marked by red asterisks (SL = 245 mm), C: short, with 28 deformed vertebrae (SL = 216 mm), and D: short, with 31 deformed vertebrae (SL = 201 mm).

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the short and related (generalized and lipped) morphs, and (4) discussing the possible role of natural selection in the large-scale emergence of the aberrant morph in the local *Labeobarbus* population.

**Materials and methods**

**Ethics statement**

Fish were collected in southeastern Ethiopia under the umbrella of the Joint Ethiopian-Russian Biological Expedition (JERBE), with permission from the National Fishery and Aquatic Life Research Center under the Ethiopian Institute of Agricultural Research and the Ethiopian Ministry of Innovation and Technology. Fish were sacrificed humanely using an anesthetic MS-222 overdose (American Veterinary Medical Association). The experiments were carried out in accordance with the rules of the Papanin Institute of Biology of Inland Waters (IBIW), Russian Academy of Sciences, and approved by IBIW’s Ethics Committee.

**Study sites and sampling**

The Ethiopian Highlands are divided by the Rift Valley into the western and eastern plateaus [71]. The southern part of the eastern plateau is drained by the Wabi Shebele (Webi Shabeelle or Uebi Scebeli) and Juba (Jubba) river systems into the Indian Ocean via Somalia. Three large tributaries, the Genale (Ganale Doria), the Dawa (Daua) and the Weyb (Gestro), meet near the Ethiopia-Somalia border to form what is known as the Juba River within Somalia. The Genale, Weyb and Wabi Shebeli originate in the Bale Mountains, the highest part of the eastern plateau (Fig 3).

Since 1990, we have sampled a total of 28 sites across the Wabi-Shebele and Juba river systems (dots in Fig 3). Among these, *Labeobarbus* were collected from 15 sites, and not found in the other 13 sites. An additional four sites important for the present work are numbered in Fig 3: (1) main sampling site, the middle reaches of the Genale River (5°42' N 39°32' E, altitude 1125 m above sea level, asl); (2) the upper reaches the Genale River north of Kibre Mengist (6°02’40” N 39°07’19” E, 1324 m asl); (3) the middle reaches of the Welemele River, a northern tributary of the Genale (6°14’30” N 39°47’22” E, altitude c. 1040 m asl); and (4) the upper
reaches the Awata River, northern tributary of the Dawa (5˚47'07"N 38˚55'46"E, 1630 m asl). All sampling sites are listed in S2 File. Map of sampling sites (Fig 3) was created in ArcGIS 10.2 (www.esri.com).

Most samples were taken in the different years from the end of January to mid-May during the main rainy season in southeastern Ethiopia [72], but before water levels rose substantially in the rivers.

The main sampling site (no. 1, Fig 3) included two habitats: the continuous pool starting from the shallows around the island, characterized by moderate current and sandy/silty bottom, and the upper part of continuous rapids with small waterfalls, rocky bottom and very fast current (S1 Fig in S1 File). Samples were mostly taken from the pool by gill netting (at dawn and overnight), but an additional sample was obtained from the rapids with cast nets in 2009. The cast net catches differed substantially from the gill net catches, in terms of fish size and frequency of *Labeobarbus* morphs (S2 Table in S1 File), therefore these catches were analyzed.
separately. The remaining sites (nos. 2–4) were sampled by both cast and gill netting during the day, and the catches were analyzed together.

The selected fish were maintained for several hours in large barrels submerged in the river, then killed with an overdose of MS-222 and their morphology was preliminarily examined. Tissue samples were taken for genetic and stable isotope analyses because the genetic and trophic specialization studies have been conducted on the same individuals [30, 34, 35]. Most specimens were preserved in 10% formalin in the field, and subsequently transferred to 70% ethanol in the laboratory. Of the 291 specimens collected in 2009, 132 were preserved with salt to make the dried bone preparations. These specimens were dissected for determination of sex and gonad maturity stages.

**Examination of vertebral column**

Most formalin-preserved specimens were x-rayed. Vertebrae were examined based on the film or digital radiograms. The total vertebral number and numbers of pre-dorsal, pre-anal, trunk, transitional and caudal vertebrae were counted according to Naseka [73]. Examination of vertebral structure and counts of vertebrae mentioned above were made by means of visual inspection of the dried bone preparations. Headset magnifier glasses (×4) were used for film and preparation inspection if necessary. In total, the vertebral column structure was studied in 364 Genale barbs representing all morphs (count data for each individual is available in S3 File). This number includes the radiograms of 34 and 45 specimens collected before 2009 and in 2019, respectively, as well as 285 radiograms and dried bone preparations of fish collected in 2009.

Witten et al. [11] propose a classification of vertebral column deformities that are observed in salmon (*Salmon salar* L.) under farming conditions. In our material, we observed eight types of deformities united by these authors into the category of ‘compressed and/or fused vertebral bodies’: 1) homogeneous compression, 2) decreased intervertebral space, 3) compression and reduced intervertebral space, 4) compression without x-structure, 5) one-sided compression; 6) compression and fusion, 7) complete fusion, and 8) fusion centre. Taking into account the relatively small size of our samples, we did not consider the different types of such deformities separately. All vertebrae with compressed and/or fused bodies were considered herein as deformed vertebrae. The deformities of other categories [11] were not recorded in our material, except in one specimen of the smiling morph that had vertebral column curvature (kyphosis). Other deformities such as a presence of abnormal additional ribs and neural spines, and splitting of neural spine [74] were recorded in our material, however they were not analyzed because they were not related to the shortening of the vertebral column.

**Examination of other morphologic characters**

Prior to preservation in formalin or salt, the following measurements were taken (with a ruler, to the nearest 0.5 mm) on all specimens in the field: standard length (SL), head length (HL), dorsal spine length (DL), maximum (H) and minimum (h) depth of body, and length of caudal peduncle (cpd). For further analysis, the body length (BL) was calculated as SL minus HL.

Of all morphs sampled in 2009 (*N* = 129, SL = 98–308 mm), we randomly selected a subset of formalin-preserved specimens and took 23 measurements in the laboratory (with a caliper, to the nearest 0.1 mm): standard length (SL), head length (HL), snout length (R), orbit horizontal diameter (O), interorbital width (IO), postorbital length of head (PO), head depth at mid-orbit (Ch), head depth at occiput (CH), mouth width (MW), dorsal spine length (DL), pre-dorsal distance (PrD), post-dorsal distance (PD), caudal peduncle length (cpd), length of dorsal fin base (ID), maximum (H) and minimum (h) depth of body, length of pectoral (IP)
and ventral (IV) fins, pectoral-ventral distance (PV), ventral-anal distance (VA), height of anal fin (hA), length of anterior barbel (Ab), and length of posterior barbel (Pb). In the present work, only data on these extended measurements were included for 22 and 28 specimens of the generalized (SL = 108–257 mm) and short (SL = 119–244 mm) morphs, respectively. All measurements were taken by the same person for consistency [75]. They are available in S3 File.

**Examination of age and growth**

We estimated the age and growth rate in our samples, as both of these are important life-history traits that can serve as markers of differences between ecomorphs. In addition, a fast growth rate is considered one of the factors contributing to the shortening of the vertebral column [11]. In most salt-preserved specimens of the different morphs, age was determined from vertebrae. Annual rings were counted on the anterior and posterior sides of the fifth vertebra after removing the dried intervertebral discs. Counts were made with a binocular microscope at magnification ×16, with a drop of glycerin for contrasting. Age estimates were calibrated with vertebrae from the artificially reared *Labeobarbus* from Lake Tana [66].

Growth rate estimates for the different *Labeobarbus* morphs were based on comparisons of individual size variation in the different year classes. We did not calculate values of growth rate or parameters of the von-Bertalanffy equation because of the small sample sizes.

**Statistical analysis**

Various R packages [76] run in R-studio v.1.2.5033 were used for statistical analyses and plot construction: `summarytools` library [77] was used for obtaining basal descriptive statistics, `ggplot2` library [78] was used to analyze Pearson correlation and plot the result as well as for creating boxplots and histograms, `posthoc.kruskal.dunn.test` function in the `PMCMR` library [79] was used for the posthoc Dunn’s test, the `FSA` library [80] was used to conduct the Mann-Whitney U test, the `prcomp` function [76] was used for the principal component analysis (PCA) and plotting the results, and the `candisc` library [81] was implemented for the canonical variate analysis (CVA). The proportions of head and body were used for analyses of single characters, as well as for the PCA and CVA—all measurements were divided by head length. The sample size of subsets is given in S3 Table in S1 File. The samples were tested for effect size using eta²-value for Kruskal-Wallis test in the `estatix` library [82] and using r-value for Mann-Whitney U test (S1 Supporting material in S1 File); r-value was calculated as Z/√N, where Z is Z-value, and N is the total number of the specimens. The differences in frequency of deformed vertebrae along the vertebral column were analyzed using Fisher’s exact test as implemented in the `fisher.test` function [76].

**Deposition of material**

All samples were deposited to the Severtsov Institute of Ecology and Evolution Russian Academy of Sciences and Papanin Institute for Biology of Inland Waters Russian Academy of Sciences under provisional labels of the Joint Ethiopian-Russian Biological Expedition.

**Abbreviation of morph names**

The names of the morphs from the Genale *Labeobarbus* assemblage have been assigned the following abbreviations: short SH, generalized GN, lipped LP, *L. jubae* JB, smiling SM, smiling hybrids HB, and piscivorous PS.
Results

Short definition

Initially, in the field, the short morph was identified based on altered body proportions: these individuals had relatively deeper and shorter bodies compared to the normal individuals (Fig 1). Investigation of the vertebral column revealed that each short individual had a substantial number of deformed vertebrae. The data on the structure of the vertebral column for all morphs are presented in Table 1.

We defined the SH morph as individuals with seven or more deformed vertebrae (because of a distributional gap in the number of deformed vertebrae with the GN and LP morphs) and as having a deeper body (Table 1, Fig 1). It is important to note that the mouth structure in the SH morph was similar to that in GN and LP morphs that had normal vertebral columns. These morphs—with either shortened or normal vertebral columns—differed in the degree of lip development (Fig 2).

There were almost no individuals with markedly shortened bodies among the trophically specialized morphs (JB, SM, HY, PS). The only PS individual with eight deformed vertebrae (Table 1) displayed noticeable body shortening (S2 Fig in S1 File).

Temporal and spatial distribution of the short morph

In the samples taken before 2009, SH individuals were not discovered among the 34 radio-graphed individuals of the different morphs, or among the hundreds of superficially examined barbs in catches of 1993, 1997 and 1998. To estimate SH prevalence in the samples of 2009 and 2019, we analyzed the total gill net catches. The catch composition appeared to be quite similar in 2009 and 2019 (S4 Table in S1 File). The ratios of morphs remained roughly stable. It is important to note that SH prevalence in the total \textit{Labeobarbus} catches was almost unchanged: they represented 10% (n = 41) of the 400 barbs sampled in 2009, and 11% (n = 20) of the 179 barbs sampled in 2019.

We did not sample the SH morph from the upper reaches of the Genale River (sampling site no. 2, Fig 3), where most other morphs common to the main sampling site were found. Moreover, the SH morph was not recorded from other sampling sites in the Wabi-Shebele and Juba river systems (Fig 3, S2 File) or from other parts of Ethiopia (sampling sites may be found in [35, 83, 84]). However, at sampling site no. 3, we found barbs that had shortened bodies but without deformed vertebrae (discussed below).

Short vs generalized and lipped morphs: Plastic characters

As expected, body depth of the SH morph was positively correlated ($R = 0.62$, $p = 1.2\text{e}{-06}$, Fig 4A) with the number of deformed vertebrae, which in turn was negatively correlated with

| Morph   | N  | %dv | Number of deformed vertebrae per individual |
|---------|----|-----|-------------------------------------------|
| Shorts (SH) | 52 | 100 | 1 2 3 4 5 6 7 8 9  |
| Generalized (GN) | 78 | 16.7 | 65 1 8 2 2  |
| Lip (LP) | 26 | 11.5 | 23 2 1  |
| Jubae (JB) | 78 | 12.8 | 68 1 7 1 1  |
| Smiling (SM) | 48 | 14.6 | 41 1 4 1 1  |
| Hybrid (HY) | 36 | 5.6 | 34 2  |
| Piscivorous (PS) | 46 | 9.8 | 41 3 1 1  |

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body length ($R = -0.45, p = 0.0012$, Fig 4B). In other morphs of the Genale *Labeobarbus* assemblage, no such correlations were found.

The changes of body proportions in the SH morph (caused by the deformity of the vertebral column) resulted in a pronounced difference in appearance between this and the other morphs. Based on PCA and CVA, the SH and GN morphs were well differentiated from each other (Fig 5). Indices of 21 measurements relative to head length ($HL$) rather than $SL$ were used in both PCA and CVA because of the variable influence of vertebral deformity on $SL$ in
the short individuals (Fig 4B). PC1 explained > 45% of the variance, while PC2 was less than 24%. Eigenvectors of the 10 most loaded characters for PC1 and PC2 are given in S5 Table in S1 File. In addition, the CVA resulted in a good separation among groups of different year’s sampling. Remarkably, in just as brief a time as 10 years, the GN morph became significantly more similar to the SH morph in body shape. Both CV1 (64.2%) and CV2 (26.8%) were highly statistically significant (LR test, p < 0.01).
Using a larger data set (which included the gill net catch of 2009, S1 Supporting material in S1 File), we compared the characters that had the strongest influence on body form (head length, depths of body and caudal peduncle) between the SH, GN and LP morphs (Fig 6). The LP and SH morphs were similarly characterized by relatively longer heads (Fig 6A). In the LP morph, the longer head length arose from elongation of the snout (Fig 2), while in the SH morph the head length appeared longer relative to the shortened body. The LP, SH and lipped SH morphs all had relatively longer heads than the GN (Kruskal-Wallis test, \( p < 0.05 \)); the lipped SH morph had a notably longer head than the SH morph, but the difference was not significant (Fig 6A). The relative depths of body and caudal peduncle in the SH morph differed significantly from both GN and LP (Kruskal-Wallis test, \( p < 0.05 \), Fig 6A), however there were no such differences between the lipped SH and any of the other morphs (Fig 6B and 6C). Notably, lipped SH exhibited the most variation in relative depths of body and caudal peduncle compared to the other morphs. This variation was caused by different degrees of snout elongation that resulted in variation of standard length, and variation caused by body shortening (determined by varying number of deformed vertebrae), which seemed to act additively.

The comparison of barbs sampled in 2009 and 2019 yielded unexpected results. Although the later sample was rather small, significant differences in relative head length and depth of caudal peduncle, as well as a nearly significant \( (p = 0.054) \) difference in body depth, were revealed between GN sampled in 2009 and 2019 (Fig 7). The GN morph became significantly more similar to the SH morph during 10 years in all three of the indices most important for their discrimination. No differences in SL and absolute values of the corresponding measurements were found between samples of 2009 and 2019 (Mann-Whitney U test).

**Short vs generalized and lipped morphs: Structure of vertebral column and other counts**

Comparison of vertebral counts between the SH morph and the GN and LP morphs did not reveal any differences in total number of vertebrae or in numbers of vertebrae in the three segments of the vertebral column (trunk, transitional and caudal). Moreover, there were no differences in the numbers of pre-dorsal and pre-anal vertebrae (S6 Table in S1 File).

The distribution of the deformed vertebrae along the vertebral column was not random \( (p = 0.014) \), Fisher exact test), and was greatly different in the SH morph compared to the other morphs (Fig 8). When the data on the positions of the deformed vertebra was merged for individuals with the different total number of vertebrae, the frequency estimates for each specific vertebra in the caudal region were obscured. For this reason, only the data for individuals with 41 total vertebra (modal number) are presented in Fig 8. The respective data for individuals with 40, 42 and 43 vertebrae are shown in S3 Fig in S1 File.

As shown in Fig 8A, the lowest frequencies of each specific vertebra deformity in the SH morph were found in 5th vertebra, and also in the caudal region, where the minimum frequencies were found in the 36-40th vertebrae. In the other morphs, however, the deformed vertebrae occurred mostly in the posterior half of the vertebral column, with maximum frequencies in the 37-38th vertebrae (Fig 8B).

**Short vs generalized and lipped morphs: Size, age, growth rate, sex ratio and gonad conditions**

The range of size variation in the SH morph (SL 107–291 mm) was less than that in any other morph from the middle Genale assemblage in 2009 and 2019. The ranges for the GN and LP morphs were 51–375 mm and 106–459 mm, respectively (S2 Table in S1 File). Most of the salt-preserved barbs were aged from vertebrae. A maximum age of six years was recorded for the
Fig 6. Indices of (A) head length, (B) body depth, and (C) caudal peduncle depth in generalized, lipped, lipped short, and short morphs sampled in 2009. Median is shown as the horizontal black line inside the box. The box represents 1st and 3rd quartiles of variation. Lowercase letters above the boxplots indicate significant differences between morphs ($p < 0.05$, Kruskal-Wallis test with Dunn’s post hoc test).

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Fig 7. Indices of (A) head length, (B) body depth, and (C) caudal peduncle depth of generalized and short morphs sampled in 2009 and 2019. Median is shown as the horizontal black line inside the box. The box represents 1st and 3rd quartiles of variation. Lowercase letters above the boxplots indicate significant differences between morphs ($p < 0.05$, Mann-Whitney U test; #—nearly significant, $p = 0.054$).

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SH and HB morphs, seven years for the SM morph, eight years for the GN morph, nine years for the JB morph, 12 years for the LP morph and 14 years for the PS morph. There was no correlation between the percentage of deformed vertebrae and age ($R = -0.075, p = 0.64$).

Analysis of size variation within each year class of the SH and GN morphs revealed that when SL was used as a measure of linear growth, SH did not differ in size from GN during the

Fig 8. Distribution of deformed vertebrae along the vertebral column in (A) short morph (n = 17), and (B) other morphs (n = 47). Only data for individuals with 41 vertebrae are shown here; the deformities were not analyzed for the first four vertebrae (Weberian apparatus) or the 41st (pre-ural 1) vertebra.

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first four years (S4 Fig in S1 File). Modification of the vertebral column in the SH morph, however, apparently resulted in shortening of their body and respective reduction of $SL$. In other words, growth of SH morph should be accelerated to achieve the same $SL$ as such in GN morph. Hence, it is reasonable to consider the head length as a measure of linear growth in comparisons of the SH morph with the other morphs. In this case, the SH morph demonstrated a markedly fast growth rate during the first four years relative to the other morphs, which then leveled off during the fifth and sixth years (Fig 9).

Gonads were examined in 30 SH individuals: five juveniles, 11 females and 14 males were recorded. There were 16 fish at maturity stage I, 11 fish at stage II, three fish at stages II-III and III. Among the 46 examined individuals of GN and LP morphs, 16 juveniles, 14 females and 16 males were recorded. There were 24 fish at maturity stage I, 20 fish at stage II, one fish at stages II-III and one female at stage IV. Thus, neither sex ratio nor gonad condition differed in the SH morph compared to GN and LP morphs.

**Discussion**

Large-scale emergence of individuals with deformed (shortened) vertebral column accommodated by a deep-bodied phenotype is a phenomenon rarely detected in wild populations. To the best of our knowledge, there are only two other similar cases detected in gadid and salmonid fishes [85–89]. In relation to *Labeobarbus*, given three circumstances—i) high frequency of such phenotype, ii) its prolonged presence in the same locality (ca. 15 years), and iii) emergence within radiating sympatric assemblage of *Labeobarbus* [34, 35, 70]—we suspect that this phenomenon may be a result of natural selection. Below we discuss the results in the context of existing knowledge on the nature and frequency of shortening vertebral deformities and how the deep-bodied phenotype might be an adaptive by-product of the abnormality.

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**Fig 9. Age vs head length in generalized (GN) and short (SH) morphs in 2009.** Median is shown as the horizontal black line inside the box. The box represents 1st and 3rd quartiles of variation. Letters above each boxplot designate significant (red S), or non-significant (black NS) differences between morphs within each year class, based on Mann-Whitney U tests ($p < 0.05$).
Factors responsible for abnormal body shortening in fish

The most comprehensive data on vertebral deformities in fish pertains to artificially propagated fish species, especially farmed salmonids. Fish with abnormally shortened bodies displaying multiple compressed and/or fused vertebrae are quite frequent in farmed stocks of Atlantic salmon *Salmo salar* L. 1758 [11, 90–100]. Externally (imaged in [90]), they are similar to the short morph from the Genale *Labeobarbus* assemblage. These salmons, called 'short tails' or 'short-spined' individuals [92, 95], pose a serious problem for the farming industry because of their reduced commercial value [101]. Therefore, special efforts have been undertaken to elucidate the factors determining the appearance of short tails; however, the results have been unclear, at least in terms of the genetic factors. Initially, results suggested that the deformities were heritable [90, 93], but this was questioned later [98, 99]. As stated by Witten et al. [95] some fish can be genetically predisposed and develop vertebral compression as a reaction to external cues.

The factors that increase the risk of vertebral deformities in farmed Atlantic salmon and other salmonids are given by Witten et al. [11]: bacterial and parasitic infections, vitamin C deficiency, phosphorous deficiency, elevated egg incubation temperature, fast growth in under-yearling smolts, inappropriate light regimes, vaccination, inappropriate water current and quality, as well as environmental pollution. All of these factors (except vaccination), along with radiation [74] may be present in the natural population of the middle Genale *Labeobarbus*. Notably, there is a relationship between the increased frequency of vertebral deformities and high growth rate in Atlantic salmon smolts [11, 97, 99]. We also found an increased growth rate in the SH morph compared to the GN and LP morphs at the age of 2–4 years. Thus, the deformations of the vertebral column in the SH morph could be caused by the accelerated early individual growth and mechanical muscle overload of bones, as suggested for Atlantic salmon [11, 96, 99].

In a study of the vertebral fusion patterns in coho salmon, *Oncorhynchus kisutch* (Walbaum 1792) [102], the author demonstrated that the distribution of vertebral fusions along the spinal column differed significantly among crosses from two hatchery stocks, indicating a genetic basis for this character. Recently, differences in the distribution of vertebral fusions along the spinal column were found between the genetically distinct year-classes of Atlantic salmon and its hybrids with brown trout and Arctic char *Salvelinus alpinus* (L. 1758) [103]. This result is interesting for understanding the difference in distribution of deformed vertebrae along the spinal column between the SH morph and other Genale *Labeobarbus* (Fig 8).

In non-salmonid fishes, it has been reported that body shortening from the compression of the vertebral column was apparently heritable in the south German common carp Aischgrunder Karpfen [21], as well as in laboratory strains of the banded topminnow, *Fundulus cingulatus* Valenciennes 1846 [104] and two poeciliids, blackstripe livebearer *Poeciliopsis prolifica* Miller 1960 [105] and guppy *Poecilia reticulata* Peters1859 [106]. In zebrafish, *Danio rerio* (Hamilton 1822), vertebral column shortening and vertebral fusions are exhibited by type I collagen mutants, as well as by individuals with knockout alleles in two genes involved in type I collagen processing [107]. At the same time, the increased frequency of vertebral deformities in wild-type zebrafish is induced by high rearing densities [108]. The stumpbody phenotypes with the shortened vertebral column in channel catfish *Ictalurus punctatus* (Rafinesque 1818) [109] and blue tilapia *Oreochromis aureus* (Steindachner 1864) [12, 110] are described as not heritable.

The situation in the Japanese rice fish, *Oryzias latipes* (Temminck & Schlegell 1846) is especially informative [111–113]. The shortened vertebral column and vertebral fusions are found in (1) fused mutants obtained from the laboratory strain (a simple recessive Mendelian
character with expression modified by temperature [111, 113]), (2) ‘wild-fused’ phenotype from a certain region on the eastern outskirts of the City of Nagoya that were not heritable, and (3) individuals of the normal strain treated by phenylthiourea at the early embryonic stage [112]. In general, it is obvious that in different species and even in different lineages of the same species, the emergence of fish with shortened columns and fused vertebrae may be determined by various external and genetic cues.

Considering the possible role of external cues, the following points must be mentioned. First, among the middle Genale Labeobarbus assemblage, the deformation of the vertebral column is found almost exclusively within the isolated genetic pool including the GN, LP, and SH morphs but excluding the trophically specialized morphs. If the deformation has no genetic background and is triggered by external cues such as mechanic, thermal, chemical or radioactive environmental stress, these impacts are only acting selectively. For example, they would have temporal or spatial effects on only some of the GN and LP barbs, but not the rest of the assemblage on the spawning grounds.

Second, environmental stress usually results in multiple malformations of different morphological structures (e.g. vertebral column, pterygiophores, fins, scale pattern) as revealed in common roach Rutilus rutilus (L.) or mosquito fish Gambusia affinis (Baird & Girard 1853) [74, 114]. We did not observe such multiple morphological abnormalities in SH but have to mention that additional ribs, neural spines, and splitting of neural spines were rarely detected in our material. Third, external cues like bacterial [115] or parasitic [116, 117] infection, and nutrient deficiency (e.g., vitamin C or phosphorous [118]) are usually manifested as a decrease in the individuals’ condition, especially in the growth rate, which is often low in poor health. However, the growth rate was exceptionally high in the abnormal individuals (SH) observed here. Taking into account all of the above, we suggest that genetic factors (possibly together with some environmental factors) are the most parsimonious explanation of the emergence and 10 year persistence of the SH morph in the middle Genale Labeobarbus assemblage.

Nevertheless, to test this hypothesis, we must seek answers to the following questions: do SH individuals reproduce in nature? Would the progeny of artificial crosses be different for the SH breeders vs. normal GN and LP breeders? Are there any genetic/genomic differences between the SH morph and other morphs? Hopefully, these will be addressed in future studies.

**Shortened vertebral column as a target of natural selection**

Some researchers [119–121] have indicated the relaxation of natural selection as an important mechanism in increasing variation of morphological, ecological and other traits in the course of adaptive radiation. Such an increase often allows the radiating species to occupy new adaptive zones. This phenomenon has been called ‘extralimital specialization’ by Myers [122]. In this perspective, the occurrence of the vertebral deformations at a high frequency only in the radiating Labeobarbus assemblage in the middle Genale River may be evidence for relaxed selection in this particular site. In addition, the vertebral deformity in short morph is accompanied by a significantly deeper body. Their body depth was positively correlated with the individual number of modified vertebrae. At the same time, the deep-bodied phenotype in fishes can serve as an anti-predatory adaptation [123, 124]. The Genale River is rich in various predators inhabiting the same segment of the river, where the Short morph is found, and anti-predator traits would be adaptive. At least two piscivorous fish species—the Somalia catfish Bagrus urostigma Vinciguerra 1985, and the piscivorous morph of L. gananensis, the predatory feeding strategy of which was confirmed in a previous study [34]—were detected. In addition to fishes, some piscivorous tetrapods, e.g. Nile crocodile, and birds (African fish eagle, gull, heron, kingfisher, cormorant, snakebird) were recorded in the same locality (our
A deep-bodied phenotype can be beneficial from two points of view. First, it lowers the vulnerability to gape-limited predators. Second, deep-bodied fish attain enhanced escape locomotor performance (higher speed, acceleration and turning rate) during antipredator responses compared to shallow-bodied fish [124].

One may expect that the barbs with deformed vertebral columns should exhibit reduction in swimming performance, but this is not straightforward. Studying the triploid lines of Atlantic salmon, Powell et al. [125] did not find a difference in swimming performance between normal fish and those with visible spinal shortening. As described earlier, most of these vertebrae in the individuals with slightly deformed vertebral columns occur in the posterior end of the fish [9, 97, 100, 110, 126–128]. This phenomenon is explained by the morphogenetic influence of the caudal complex, whose normal development includes several vertebral fusions [128]. The increased frequency of abnormalities in the caudal region was also detected as a result of thyroid hormone disruption [129, 130]. However, in the deformed (SH) barbs, the frequency of deformed vertebrae was lower in the last caudal vertebrae (Fig 8). This is unlike the farmed stock of Atlantic salmon [100] and may be considered as beneficial in the light of experimental data on zebrafish that has demonstrated the caudal vertebrae to be important for predator avoidance [131].

Other examples of the differentiation among Labeobarbus in the Chamo-Abaya lake basin (Ethiopian Rift valley) may corroborate the idea that deep body is anti-predator adaptation. The lakes are inhabited by the deep-bodied barbs classified as L. bynni (Fabricius 1775) [132], while the tributaries are inhabited by the low-bodied barbs classified as L. intermedius (Rüppell 1835) [132]. However, phylogeny based on mt-DNA [33] demonstrates a close relationship between these two forms and their distant relation to L. bynni from the Nile basin. Thus, deep-bodied forms evolved independently in the Chamo-Abaya and Nile basins most probably as an anti-predator adaptation, as the faunal data [83, 133, 134] indicate the greater predator diversity in these basins compared to the Chamo-Abaya tributaries and most other rivers of the Ethiopian Highlands that are populated by the low-bodied Labeobarbus.

In our understanding, the importance of the predator-induced deep-bodied phenotype is evident in some Labeobarbus populations. In these cases, natural selection could favor the Genale deformed barbs because of their anti-predator deep-bodied phenotype. Hypothesis on the role of direct natural selection in high frequency of Short morph in the Genale River can be supported by other circumstances. First, comparison of the samples from 2009 and 2019 unexpectedly revealed changes in the body form of the Generalized morph (Fig 7). The generalized individuals became more similar to the Short morph, particularly in the relative body depth. At the same time, the SH morph also became more deep-bodied than in 2009. The most parsimonious explanation for this phenomenon is a selective pressure for the increase in relative body depth in the population studied. Second, in our small sample from the Welemele River (the northern tributary of the Genale; Fig 3, site no. 3), together with the normal generalized Labeobarbus morph, we found a few individuals with body proportions very similar to that in the SH morph from the middle Genale assemblage. However, their vertebral columns were not deformed (Fig 10). This finding might provide evidence for a selective pressure for an increase in the relative body depth in another Labeobarbus population.

Shortened vertebral column as a population phenomenon

There are many reports of individuals with the markedly shortened bodies and deformed vertebral columns in natural populations of different fish groups (S1 Table in S1 File). Among wild cyprinids, there are reports of a Mesopotamian barb, Mesopotamichthys sharpeyi (Günther 1874), with nine deformed vertebrae; and a yellowfin barbel, Luciobarbus xanthonopterus.
Heckel 1843, with 22 deformed vertebrae [135, 136]. The situations, as in the Genale *Labeobarbus*, where the abnormality becomes a population phenomenon occurring with substantial frequency among several successive generations are rare. It is important to note that in the Genale barb assemblage, only two morphs (GN and JB) were always present in catches with frequencies higher than 10% recorded for the abnormal (SH) morph (S1 Supporting material in S1 File). To the best of our knowledge, apart from the Genale *Labeobarbus*, the similar situations are described for only two other fish species, Atlantic cod *Gadus morhua* from the Elbe estuary and German Wadden Sea [85, 86–88] and Arctic char *Salvelinus alpinus* from Transbaikalian Lake Dzhelo [89]. The high prevalence of vertebral column deformities was recently reported in a wild population of lumpfish *Cyclopterus lumpus* L. 1758 from Masfjorden, Norway [137], but changes of the body form and temporal stability of the abnormal phenotypes are not evident in the latter species.

Comparative etiology of vertebral deformities in cyprinid, salmonid and gadid species could be of great interest. Moreover, deeper understanding of the action of natural selection on these abnormalities can shed light on how morphological novelties are established in a population, especially if the abnormalities in question have a genetic background.

**Conclusions**

The striking emergence of individuals with deformed (shortened) vertebral columns resulting in a deep-bodied phenotype is a phenomenon rarely detected among wild fish populations.

For the particular case of the radiating *Labeobarbus* assemblage from the middle Genale River,
the following circumstances must be highlighted: i) the recent emergence (~ 15 years ago) of the deep-bodied phenotype with the shortened vertebral column, ii) the persistence of this phenotype with a substantial frequency (~ 10%) in several generations (> 10 years) in a population, iii) the fact that this is the only type of morphological deformity; virtually all other morphological abnormalities are absent, iv) the presence of the deformity in the isolated gene pool within the radiating *Labeobarbus* assemblage, and v) the rapid growth of the abnormal individuals at 2–4 years of life compared to other sympatric morphs. Taking into account all the above, as well as the evidence for a genetic contribution to such abnormality in some other fish species, particularly the common carp, we hypothesize that this phenomenon in this *Labeobarbus* population most likely has a genetic background. Moreover, the deep-bodied phenotype in *Labeobarbus* may have a selective advantage as an anti-predator defense.

**Supporting information**

**S1 File.**

(DOCX)

**S2 File.**

(XLSX)

**S3 File.**

(XLSX)

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References

1. Darwin C. On the origin of species by means of natural selection, or the preservation of favoured races in the struggle for life. A new ed. New York: D. Appleton & Company; 1860. http://darwin-online.org.uk/converted/pdf/1861-OriginNY_F382.pdf PMID: 30164232

2. Nelson JS, Grande T, Wilson MVH. Fishes of the world. 5th ed. Hoboken (NJ): John Wiley & Sons; 2016.

3. Fraser-Brunner A. The ocean sunfishes (Family Molidae). Bull Br Mus (Nat Hist) Zool. 1951; 1:87–121.

4. Pope EC, Hays GC, Doyle TK, Sims D, Queiroz N, et al. The biology and ecology of the ocean sunfish *Mola mola*: a review of current knowledge and future research perspectives. Rev Fish Biol Fisher. 2010; 20(4):471–87. https://doi.org/10.1007/s11160-009-9155-9

5. Tyler JC. Abnormal fin and vertebral growth structures in plecoglossid fishes. Proc Acad Nat Sci Philadelphia. 1970; 122:249–71.

6. Johnson GD, Britz R. Leis' conundrum: Homology of the clavus of the ocean sunfishes. 2. Ontogeny of the median fins and axial skeleton of *Ranzania laevis* (Teleostei, Tetraodontiformes, Molidae). J Morphol. 2005; 266:11–21. https://doi.org/10.1002/jmor.10242 PMID: 15549687

7. Barahona-Fernades MH. Body deformities in hatchery reared European sea bass (*Dicentrarchus labrax* L). Types, prevalence, and effect on fish survival. J Fish Biol. 1982; 21:239–49. https://doi.org/10.1111/j.1095-8649.1982.tb02830.x

8. Divanach P, Boglione C, Menu B, Koulountouros G, Kentouri M, Cataudella S. Abnormalities in fish mariculture: an overview of the problem, causes and solutions. In: European Aquaculture Society (EAS) International Workshop on Sea Bass and Sea Bream Culture: Problems and Prospects: Handbook of contributions and short communications; 1996 16–18 Oct; Verona, Italy. 1996. p. 45–66.

9. Gavaia PJ, Dinis MT, Cancela ML. Osteological development and abnormalities of the vertebral column and caudal skeleton in larval and juvenile stages of hatchery-reared Senegal sole (*Solea senegalensis*). Aquaculture 2002; 211:305–23. https://doi.org/10.1016/S0044-8486(02)00167-9

10. Gorman KF, Breden F. Teleosts as models for human and deformity. Comp Biochem Physiol C. 2007; 145:28–38. https://doi.org/10.1016/j.cbpc.2006.10.004 PMID: 17240199

11. Witten PE, Gil Martens L, Huysseune A, Takle H, Hjelde K. Towards a classification and developmental relationships of vertebral body malformations in farmed Atlantic salmon (*Salmo salar*). Aquaculture. 2009; 295:6–14. https://doi.org/10.1016/j.aquaculture.2009.06.037

12. Tave D, Jo J-Y, Kim DS. Gross abnormalities in tilapia. Fish Aquatic Sci. 2011; 14:148–60. https://doi.org/10.5657/FAS.2011.0148

13. Boglione C, Gisbert E, Gavaia P, Witten PE, Moren M, Fontagné S, et al. Skeletal anomalies in reared European fish larvae and juveniles. Part 2: main typologies, occurrences and causative factors. Rev Aquaculture 2013; 5:121–67. https://doi.org/10.1111raq.12016

14. Evans KM, Savage AM, Albert JS. Spinal abnormalities in a specimen of the Panamanian knifefish *Apterostrobus rostratus* (Apterostrodidae: Gymnotiformes) with comments on Gymnotiform locomotion. Copeia. 2018; 106:130–134. https://doi.org/10.1643/CI-17-598

15. Torres-Dowdall J, Karagic N, Plath M, Riesch R. Evolution in caves: selection from darkness causes spinal deformities in teleost fishes. Biol. Lett. 2018; 14:20180197. https://doi.org/10.1098/rsbl.2018.0197 PMID: 29875208

16. Reyes Corral WD, Aguirre WE. Effects of temperature and water turbulence on vertebral number and body shape in *Astyanax mexicanus* (Teleostei: Characidae). PloS One. 2019; 14:e0219677. https://doi.org/10.1371/journal.pone.0219677 PMID: 31356643

17. Dawson CE. A bibliography of anomalies of fishes. Gulf Res Rep. 1964; 1(6):308–99.

18. Dawson CE. A bibliography of anomalies of fishes. Supplement 1. Gulf Res. Rep. 1966; 2(2):169–76.
19. Dawson CE. A bibliography of anomalies of fishes. Supplement 2. Gulf Res. Rep. 1971; 3(2):215–39.
20. Dawson CE, Heal E. A bibliography of anomalies of fishes. Supplement 3. Gulf Res. Rep. 1976; 5(2):35–41.
21. Wunder W. Wirbelsäulenverkürzung als rassengebendes Merkmal beim Aischgrüner Karpfen. Wilhelm Roux‘ Arch Entwicklung. 1949; 144:1–24.
22. Steffens W. Der Karfen. 5 Auflage. Wittenberg, Lutherstadt: A. Ziemsen Verlag; 1980.
23. Oellerman LK, Skelton PH. Hexaploidy in yellowfish species (Barbus, Pisces, Cyprinidae) from southern Africa. J Fish Biol. 1990; 37:105–15. https://doi.org/10.1111/j.1095-8649.1990.tb05932.x
24. Golubtsov AS, Krysanov EY. Karyological study of some cyprinid species from Ethiopia. The ploidy differences between large and small Barbus of Africa. J Fish Biol. 1993; 42:454–55. https://doi.org/10.1111/j.1095-8649.1993.tb00347.x
25. Guégan J-F, Rab P, Machordom A, Doadrio I. New evidence of hexaploidy in 'large' African Barbus with some considerations on the origin of hexaploidy. J Fish Biol. 1995; 47:192–8. https://doi.org/10.1111/j.1095-8649.1995.tb01888.x
26. Krysanov EYu, Golubtsov AS. Karyotypes of some Ethiopian Barbus and Varicorhinus from the Nile Basin including Lake Tana morphotypes. Folia Zool. 1996; 45(Suppl. 1):67–75.
27. Naran D, Skelton PH, Villet MH. Karyology of three evolutionarily hexaploid southern African species of yellowfish, Labeobarbus Rüppel, 1836 (Cyprinidae). Afr. Zool. 2007; 42:254–60. https://doi.org/10.3377/1862-7020(2007)42[254:KOTEHS]2.0.CO;2
28. Yang L, Sado T, Hirt MV, Pasco-Viel E, Arunachalam M, Li J, et al. Phylogeny and polyploidy: resolving the classification of cyprinid fishes (Teleostei: Cypriniformes). Mol. Phylogenet. Evol. 2015; 85:97–116. https://doi.org/10.1016/j.ympev.2015.01.014 PMID: 25698355
29. Vreven EJ, Musschoot T, Snoeks J, Schliewen UK. The African hexaploid Torini (Cypriniformes: Cyprinidae): Review of a tumultuous history. Zool J Linnean Soc. 2016; 177:231–305. https://doi.org/10.1111/zoj.12366
30. Levin BA, Golubtsov AS, Dgebuadze YY, Mugue NS. New evidence of homoplasys within the African genus Varicorhinus (Cyprinidae): an independent origin of specialized scrapings forms in the adjacent drainage systems of Ethiopia inferred from mtDNA analysis. Afr. Zool. 2013; 48:400–6. https://doi.org/10.1080/15627020.2013.11407609
31. Beshera KA, Harris PM. Mitochondrial DNA phylogeography of the Labeobarbus intermedius complex (Pisces, Cyprinidae) from Ethiopia. J Fish Biol. 2014; 85:228–45. https://doi.org/10.1111/jfb.12408 PMID: 24919773
32. Beshera KA, Harris PM, Mayden RL. Novel evolutionary lineages in Labeobarbus (Cypriniformes; Cyprinidae) based on phylogenetic analyses of mtDNA sequences. Zootaxa. 2016; 409(3):363–81. https://dx.doi.org/10.11646/zootaxa.4093.3.4 PMID: 27394501
33. Levin BA, Golubtsov AS. An evidence of past introgressive hybridization between Labeobarbus ethiopicus and L. intermedius in the Ethiopian Rift Valley, East Africa. Ethiop J Biol Sci. 2017; 16(Suppl.):45–60.
34. Levin BA, Casal-López M, Simonov E, Dgebuadze YY, Mugue NS, Tiunov AV, et al. (2019). Adaptive radiation of barbs of the genus Labeobarbus (Cyprinidae) in an East African river. Freshw Biol. 2019; 64:1721–36. https://doi.org/10.1111/fwb.13364
35. Levin BA, Simonov E, Dgebuadze YY, Levina M, Golubtsov AS. (2020). In the rivers: Multiple adaptive radiations of cyprinid fishes (Labeobarbus) in Ethiopian Highlands. Sci Rep. 2020; 10(1):7192. https://doi.org/10.1038/s41598-020-64350-4 PMID: 32346059
36. Bini G. I pesci del Lago Tana. Missione di studio al Lago Tana richerge Limnologiche. B. Chimica e Biologia. Reale Accademia d’Italia. 1940;3:137–79.
37. Brunelli G. Le mutazioni del genere Barbus del Lago Tana. Missione di studio al Lago Tana richerge Limnologiche. B. Chimica e Biologia. Reale Accademia d’Italia. 1940; 3: 207–13.
38. Nagelkerke LAJ, Sibbing FA, van den Boogaart JGM, Lammens EHRR, Osse JWM. The barbs (Barbus spp.) of Lake Tana: a forgotten species flock? Env Biol Fish. 1994; 39:1–21. https://doi.org/10.1007/BF00004751
39. Nagelkerke LAJ, Mina MV, Wudneh Tesfaye, Sibbing FA, Osse J.M. In Lake Tana, a unique fish fauna needs protection. BioScience. 1995; 45:772–775. https://doi.org/10.1007/BF00004751
40. Nagelkerke LAJ, Sibbing FA, Osse JWM. 1995, Morphological divergence during growth in the large barbs (Barbus spp.) of Lake Tana, Ethiopia. Neth J Zool. 1995; 45:431–54. https://doi.org/10.1163/156854295X00401
41. Alekseyev SS, Dgebuadze YY, Mina MV, Mironovsky AN. Small 'large barbs' spawning in tributaries of Lake Tana: what are they? Folia Zool. 1996; 45(suppl.1):85–96.
42. Mina M.V., Mironovsky AN, Dgebuadze YY. Lake Tana large barbs: phenetics, growth and diversification. J Fish Biol. 1996; 48:383–404. https://doi.org/10.1111/j.1095-8649.1996.tb01435.x
43. Mina M.V., Mironovsky AN, Dgebuadze YY. Morphometry of barbel of Lake Tana, Ethiopia: multivariate ontogenetic channels. Folia Zool. 1996; 45(suppl. 1):109–16.
44. Nagelkerke LAJ, Sibbing FA. Reproductive segregation among the large barbs (Barbus intermedius complex) of Lake Tana, Ethiopia. An example of intralacustrine speciation? J Fish Biol. 1996; 49:1244–66. https://doi.org/10.1111/j.1095-8649.1996.tb1793.x
45. Nagelkerke LAJ, Sibbing FA. A revision of the large barbs (Barbus spp., Cyprinidae, Teleostei) of Lake Tana, Ethiopia, with a description of seven new species. pp. 105–170. In: Nagelkerke LAJ. The barbs of Lake Tana, Ethiopia: Morphological diversity and its implications for taxonomy, trophic resource partitioning and fisheries [Ph.D. Thesis]. Wageningen (The Netherlands): Wageningen Agricultural University; 1997. p. 105–70.
46. Berrebi P, Valushok D. Genetic divergence among morphotypes of Lake Tana (Ethiopia) barbs. Biol J Linn Soc. 1998; 64:369–84. https://doi.org/10.1111/j.1095-8312.1998.tb00338.x
47. Sibbing FA, Nagelkerke LAJ, Stet RJM, Osse JWM. Speciation of endemic Lake Tana barbs (Cyprinidae, Teleostei) driven by trophic resource partitioning; a molecular and ecomorphological approach. Aquatic Ecol. 1998; 32:217–27. https://doi.org/10.1023/A:1009920522235
48. Dgebuadze YY, Mina MV, Alekseyev SS, Golubtsov AS. Observations on reproduction of the Lake Tana barbs. J Fish Biol. 1999; 54:417–23. https://doi.org/10.1111/j.1095-8649.1999.tb00840.x
49. De Graaf M, Dejen Eshete, Sibbing FA, Osse JWM. The piscivorous barbs of Lake Tana (Ethiopia): major questions on their evolution and exploitation. Neth J Zool. 2000; 50:215–23. https://doi.org/10.1113/156854200X00081
50. Nagelkerke LAJ, Sibbing FA. 2000. The large barbs (Barbus spp., Cyprinidae, Teleostei) of Lake Tana (Ethiopia), with a description of a new species, Barbus osseensis. Neth J Zool. 2000; 50:179–214.
51. Mina MV, Mironovsky AN, Golani D. Consequences and modes of morphological diversification of East African and Eurasian barbins (genera Barbus, Varicorhinus and Capoeta) with particular reference to Barbus intermedius complex. Env Biol Fish. 2001; 61:241–52. https://doi.org/10.1023/A:1010952108081
52. Sibbing FA, Nagelkerke LAJ. Resource partitioning by Lake Tana barbs predicted from fish morphometrics and prey characteristics. Rev Fish Biol Fisher. 2001; 10:393–437. https://doi.org/10.1023/A:1012270422092
53. Mina MV, Golani D. Changes of skull proportions in ontogeny of Barbus canis from Lake Kinneret and of the Lake Tana large-mouthed barbs, Barbus intermedius complex. Env Biol Fish. 2004; 70:315–20. https://doi.org/10.1023/B:EBFI.0000033346.86973.17
54. de Graaf M, Nentwich ED, Osse JWM, Sibbing FA. (2005). Lacustrine spawning: is this a new reproductive strategy among 'large' African cyprinid fishes? J Fish Biol. 2005; 66:1214–36. https://doi.org/10.1111/j.0022-1112.2005.00671.x
55. Smirnov SV, Dzerzhinskii KF, Levin BA. (2006). On the relationship between scale number in the lateral line in the African barbel Barbus intermedius and the rate of ontogeny (by experimental data). J Appl Ichthyrol. 2006; 22:129–32. https://doi.org/10.1023/S:0032945206010176
56. Zwoykin DD, Budaev SV, Darkov AA, Dzerzhinskii KF, Lyovin BA, Mina MV. Assessment of the role of chemoreception in the mate choice in barbs of the Barbus intermedius complex from Lake Tana, Ethiopia. J Ichthyol. 2006; 46:661–7. https://doi.org/10.1134/S0032945206080133
57. de Graaf M, Megens H-J, Samalio J, Sibbing FA. Evolutionary origin of Lake Tana's (Ethiopia) small Barbus species: indications of rapid ecological divergence and speciation. Anim Biolog. 2007; 57:39–48. https://doi.org/10.1163/15707560778002069
58. Dzerzhinskii KF, Shkil FN, Abdissa Belay, Zelalem Wondie, Mina MV. Spawning of large Barbus (Barbus intermedius complex) in a small river of the Lake Tana Basin (Ethiopia) and relationships of some putative species. J Ichthyol. 2007; 47:839–46. https://doi.org/10.1134/S0032945207080103
59. de Graaf M, Dejen Eshete, Osse JWM, Sibbing FA. 2008. Adaptive radiation of Lake Tana's (Ethiopia) Labeobarbus species flock (Pisces, Cyprinidae). Mar Freshwater Res. 2008; 59:391–407. https://doi.org/10.1071/MF07123
60. de Graaf M, Megens H-J, Samalio J, Sibbing F. Preliminary insight into the age and origin of the Labeobarbus fish species flock from Lake Tana (Ethiopia) using the mtDNA cytochrome b gene. Mol Phylogenet Evol. 2010; 54:336–43. https://doi.org/10.1016/j.ympev.2009.10.029 PMID: 19878730
61. de Graaf M, Nagelkerke LAJ, Palstra AP, Sibbing FA. Experimental evidence for the biological species status in Lake Tana’s Labeobarbus flock (Cyprinidae). Anim. Biol. 2012; 60:183–93. https://doi.org/10.1163/157075610X491725
62. Shkil FN, Kapitanova DV, Borisov VB, Abdissa B, Smirnov SV. Thyroid hormone in skeletal development of cyprinids: Effects and morphological consequences. J Appl Ichthyol. 2012; 28:398–405. https://doi.org/10.1111/j.1439-0426.2012.01992.x

63. Shkil FN, Levin BA, Abdissa B, Smirnov SV. Variability in the number of tooth rows in the pharyngeal dentition of Barbus intermedius (Teleostei; Cyprinidae): genetic, hormonal and environmental factors. J Appl Ichthyol. 2010; 26:315–319. https://doi.org/10.1111/j.1439-0426.2010.01428.x

64. Nagelkerke LAJ, Leon-Kloosterziel KM, Megen H-J, de Graaf M, Diekmann OE, Sibbing FA. Shallow genetic divergence and species delineations in the endemic Labeobarbus species flock of Lake Tana, Ethiopia. J Fish Biol. 2015; 87:1191–208. https://doi.org/10.1111/jfb.12779 PMID: 26385126

65. Shkil FN, Lazebnyi OE, Kapitanova DV, Abdissa B, Borisov VB, Smirnov SV. Ontogenetic mechanisms of explosive morphological divergence in the Lake Tana (Ethiopia) species flock of large African barbs (Labeobarbus; Cyprinidae; Teleostei). Russ J Dev Biol. 2015; 46:294–306. https://doi.org/10.1134/S1062360415050069 PMID: 26606829

66. Shkil FN, Smirnov SV, Kapitanova DV. Internal sources of infraorbital series and lateral line scale count variability in six Labeobarbus species composing the Lake Tana (Ethiopia) species flock. J Appl Ichthyol. 2018; 34:465–71. https://doi.org/10.1111/jai.13665

67. Vreven EJ, Musschoot T, Decru E, Wamukoni Lunkayikio S, Obiero K, Cerwenka AF, et al. The complex origins of mouth polymorphism in the Labeobarbus (Cypriniformes: Cyprinidae) of the Inkisi River basin (Lower Congo, DRC, Africa): Insights from an integrative approach. Zool J Linnean Soc. 2018; 186:414–82. https://doi.org/10.1111/1095-8649.13422

68. Golubtsov AS. Biogeographie des ‘grands Barbus’ d’Ethiopie avec reference speciale à des formes à statuts taxinomiques incertains. Cah Ethiol. 1993; 13:227–30.

69. Mina MV, Mironovsky AN, Golubtsov AS, Dgebuadze YuYu. ‘Barbus’ intermedius species flock in Lake Tana (Ethiopia). II. Morphological diversity of “large barbs” from Lake Tana and neighbouring areas: homoplasies or synapomorphies? Ital J Zool. 1998; 65(Suppl.):9–14. https://doi.org/10.1080/11250009809386789

70. Dimmick WW, Berendzen PB, Golubtsov AS. Genetic comparison of three Barbus (Cyprinidae) morphotypes from the Genale River, Ethiopia. Copeia. 2001(4):1123–29. https://doi.org/10.1643/0045-8511(2001)004[1123:GCOTBC]2.0.CO;2

71. Pauly G. The Ethiopian subregion fish fauna: an original patchwork with various origins. Hydrobiologia. 2010; 649:301–15. https://doi.org/10.1007/s10750-010-0273-0

72. Berhanu B, Seleshi Y, Melesse AM. Chapter 6. Surface water and groundwater resources of Ethiopia: Potentials and challenges of water resources development. In: Melesse AM, Abtew W, Setegn SG, editors. Nile River Basin: Ecohydrological Challenges, Climate Change and Hydropolitics. Switzerland: Springer International Publishing; 2014. p. 97–213. https://doi.org/10.1007/978-3-319-02720-3_6

73. Naseka AM. Comparative study on the vertebral column in the Gobiomiminae (Cyprinidae, Pisces) with special reference to its systematics. Publ Espec Inst Esp Oceanogr. 1996; 21:149–67.

74. Bogutskaya NG, Zuykov MA, Naseka AM, Andersson B. Normal axial skeleton structure in common roach Rutilus rutilus (Actinopterygii: Cyprinidae) and malformations due to radiation contamination in the area of the Mayak (Chelyabinsk Province, Russia) nuclear plant. J Fish Biol. 2011; 79:991–1016. https://doi.org/10.1111/j.1095-8649.2011.03078.x PMID: 21967586

75. Mina MV, Levin BA, Mironovsky AN. On the possibility of using character estimates obtained by different operators in morphometric studies obtained by different operators in morphometric studies of fish. J Ichthyol. 2005; 45:284–94. https://doi.org/10.1134/S003294520503006X

76. RStudio Team. RStudio: Integrated Development for R. 2020. PBC, Boston, MA http://www.rstudio.com/.

77. Comtois D. Summarytools: Tools to quickly and neatly summarize data. 2018. R package version 0.8. 72018. https://CRAN.R-project.org/package=summarytools

78. Wickham H. ggplot2: elegant graphics for data analysis. Springer; 2016.

79. Pohlelrt T, Pohlelrt MT. Package ‘PMCMR’. R package version 1(0); 2018.

80. Ogle D, Ogle MD. Package ‘FSA’. CRAN Repos. 2017;1–206.

81. Friendly M, Fox J, Friendly MM. Package ‘candisc’. 2020. https://cran.r-project.org/web/packages/candisc/index.html

82. Golubtsov AS, Dgebuadze YY, Mina MV. Fishes of the Ethiopian Rift Valley. In: Tudorancea C, Taylor WD, editors. Ethiopian Rift Valley Lakes (pp. 167–258). Leiden, The Netherlands: Backhuys Publishers; 2002. p. 167–258.
84. Levin BA. New data on morphology of the African scraping feeder Vankorhinus beso (Osteichthyes: Cyprinidae) with the special reference to specialized traits. J Ichthyl. 2012; 52:908–23. https://doi.org/10.1134/S0032945212110069

85. Hilger I. Spinal compression of Atlantic cod Gadus morhua from the German Wadden Sea. Dis Aquat Organ. 1992; 13:83–8. https://doi.org/10.3354/dao013083

86. Wunder W. Missbildungen beim Kabeljau (Gadus morhua) verursacht durch Wirbelsaumenverkürzung. Helgol Meeresunters. 1971; 22:201–12.

87. Möller H. High skeletal deformation rates of cod in the Elbe estuary. Bull Eur Ass Fish Pathol. 1983; 3:7–8.

88. Möller H. Dynamics of fish diseases in the lower Elbe river. Helgol Meeresunters. 1984; 37:389–413.

89. Alekseyev SS. [Distribution, diversity and diversification of the Arctic char Salvelinus alpinus (L.) complex (Salmoniformes, Salmonidae) in Siberia] [dissertation]. Moscow (Russia): Moscow State University, Faculty of Biology; 2016. Russian.

90. McKay LR, Gjerde B. Genetic variation for a spinal deformity in Atlantic salmon, Salmo salar. Aquaculture. 1986; 52:263–72. https://doi.org/10.1016/0044-8486(86)90369-8

91. Vågsholm I, Djuvik HO. Risk factors for spinal deformities in Atlantic salmon, Salmo salar L. J Fish Dis. 1998; 21:47–53. https://doi.org/10.1046/j.1365-2761.1998.00069.x PMID: 9739172

92. Kvellestad A, Heie S, Thorud K, Terud B, Lyngøy A. Plathypondyly and shortness of vertebral column in farmed Atlantic salmon Salmo salar in Norway—description and interpretation of pathologic changes. Dis Aquat Organ. 2000; 39:97–108. https://doi.org/10.3354/dao039097 PMID: 10715815

93. Wargelius A, Fjelldal PG, Hansen T. Heat shock during early somitogenesis induces caudal vertebral column defects in Atlantic salmon (Salmo salar). Aquaculture. 2005; 244:77–87. https://doi.org/10.1016/j.aquaculture.2004.12.002

94. Gjerde B, Pante MJR, Baeverfjord G. Genetic variation for a vertebral deformity in Atlantic salmon (Salmo salar). Fish Pathol. 2006; 41:211–18. https://doi.org/10.1007/s10695-006-9026-6 PMCID: 1637962

95. Witten PE, Obach A, Huysseune A, Baeverfjord G. Vertebral body mineralization and deformation of deformed vertebrae in Atlantic salmon (Salmo salar). J Fish Biol. 2010; 76:1481–92. https://doi.org/10.1111/j.1095-8641.2010.02639.x PMCID: 1988977

96. Witten PE, Obach A, Huysseune A, Baeverfjord G. Vertebral body mineralization and deformation of deformed vertebrae in Atlantic salmon (Salmo salar). J Appl Ichthyol. 2010; 26:355–59. https://doi.org/10.1007/s10764-009-9734-0 PMCID: 2919185

97. Witten PE, Obach A, Huysseune A, Baeverfjord G. Vertebral body mineralization and deformation of deformed vertebrae in Atlantic salmon (Salmo salar). J Fish Biol. 2010; 76:1481–92. https://doi.org/10.1111/j.1095-8641.2010.02639.x PMCID: 1988977

98. Witten PE, Gil Martens L, Hall BK, Huysseune A, Obach A. Compressed vertebrae in Atlantic salmon (Salmo salar): evidence for metaplastic chondrogenesis as a skeletogenetic response late in ontogeny. Dis Aquat Organ. 2005; 64:237–46. http://dx.doi.org/10.3354/dao064237 PMID: 15997822

99. Hemmer B, Breck O, Øvsthus R, Lock E-J, Waagbo R, et al. Vertebral column defects in Atlantic salmon (Salmo salar L.). J Appl Ichthyol. 2010; 26:355–59. https://doi.org/10.1007/s10764-009-9734-0 PMCID: 2919185

100. Hansen T, Fjelldal PG, Yurtseva A, Berg A. A possible relation between growth and number of deformed vertebrae in Atlantic salmon (Salmo salar L.). Dev Genes Evol. 2012; 222:695–701. https://doi.org/10.1007/s00427-012-0482-0 PMCID: 3294930

101. Fjelldal PG, Hansen T, Breck O, Øvsthus R, Lock E-J, Waagbo R, et al. Vertebral deformities in farmed Atlantic salmon (Salmo salar L.): etiology and pathology. J Appl Ichthyol. 2012; 28:429–39. https://doi.org/10.1111/j.1439-0426.2011.01434.x

102. Campbell WB. Genetic variation of vertebral fusion patterns in coho salmon. J Fish Biol. 1995; 46:717–20. https://doi.org/10.1111/j.1095-8649.1995.tb01106.x

103. Fraser TWK, Hansen TJ, Sambraus F, Fjelldal PG. Vertebral deformities in interspecific diploid and triploid salmonid hybrids. J Fish Biol. 2020. https://doi.org/10.1111/jfb.14353 PMID: 32307707

104. Thomerson JE. The stubby vertebral deformity in aquarium-raised Fundulus cingulatus. Copeia. 1966; (4):878–79. https://www.jstor.org/stable/262904

105. Schulz RJ. Stubby, a hereditary vertebral deformity in the viviparous fish Poeciliopsis prolifica. Copeia. 1963;(2):325–30. https://www.jstor.org/stable/1441351

106. Lodi E. Palla: A hereditary vertebral deformity in the guppy Poecilia reticulata Peters, Pisces, Osteichthyes. Genetica. 1978; 48:197–200. https://doi.org/10.1007/BF00155570
107. Gistelinck C, Kwon RY, Malfait F, Symoens S, Harris MP, Henke K, et al. Zebrasfish type I collagen mutants faithfully recapitulate human type I collagenopathies. Proc Natl Acad Sci USA. 2018; 115(34); E8037–46. https://doi.org/10.1073/pnas.1722200115 PMID: 30082390

108. Martini A, Huyssene A, Witten PE, Boglione C. Plasticity of the skeleton and skeletal deformities in zebrafish (Danio rerio) linked to rearing density. J Fish Biol. 2020. https://doi.org/10.1111/jfb.14272 PMID: 32010967

109. Dunhan RA, Smitherman RO, Bondari K. Lack of inheritance of stumpbody and taillessness in channel catfish. Prog Fish-Cult. 1991; 53:101–5. https://doi.org/10.1577/1548-8640(1991)053<0101:LOIOSA>2.3.CO;2

110. Tave D, Bartels JE, Smitherman RO. Stumpbody Sarotherodon aureus (Steindacher) (Tilapia aurea) and tail-less S. niloticus (L.) (T. nilotica): two vertebral anomalies and their effects on body length. J Fish Dis. 1982; 5:487–94.

111. Aida T. Further genetical studies of Aplocheilus latipes. Genetics. 1930; 15:1–16. PMID: 17246590

112. Yamamoto T, Tomita H, Matsuda N. Hereditary and nonheritable vertebral anchylosis in the medaka, Oryzias latipes. Japan J Genet. 1963; 38:36–47. https://doi.org/10.1266/jjg.38.36

113. Ogawa NM. A phylogenetic study of the vertebral fused (f) in the medaka, Oryzias latipes. Embryologia. 1965; 9:13–33. https://doi.org/10.1111/j.1440-169X.1965.tb00212.x PMID: 5862484

114. Hubbs C. High incidence of vertebral deformities in two natural populations of fishes inhabiting warm springs. Ecology. 1959; 40:154–55. https://doi.org/10.2307/1929941

115. Madsen L, Ambjerg J, Dalsegaard I. Radiological examination of the spinal column in farmed rainbow trout Oncorhynchus mykiss (Walbaum): experiments with Flavobacterium psychrophilum and oxytetracycline. Aquac Res. 2001: 32:325–41. https://doi.org/10.1046/j.1365-2109.2001.00552.x

116. Cunningham ME, Markle DF, Watral VG, Kent ML, Curtis LR. Patterns of fish deformities and their association with trematode cysts in the Willamette River, Oregon. Env Biol Fish. 2005; 73:9–19. https://doi.org/10.1007/s10641-004-3153-5

117. Villeneuve DL, Curtis LR, Jenkins JJ, Warner KE, Tilton F, Kent ML, et al. Environmental stresses and skeletal deformities in fish from the Willamette River, Oregon. Environ Sci. 2005; 39:3495–506. https://doi.org/10.1007/s10641-004-3153-5

118. Witten PE, Owen MAG, Fontanillas R, Soenens M, McGurk C, Obach A. A primary phosphorus-deficient skeletal phenotype in juvenile Atlantic salmon Salmo salar: the uncoupling of bone formation and mineralization. J Fish Biol. 2015: 88:690–708. https://doi.org/10.1111/jfb.12870 PMID: 26707938

119. Golubtsov AS. [Species flocks of fishes in rivers and lakes: Sympatric divergence in faunistically depauperate fish communities as especial modus of evolution]. In: Pavlov DS, Dgebuadze YY, Shatu-shinskii MI, editors. Current problems of ichthyology. To 100-jubilee of G.V. Nikolsky. Moscow, Russia: KMK; 2010. p. 96–123. Russian.

120. Yoder JB, Clancy E, Des Roches S, Eastman JM, Gentry L, et al. Ecological opportunity and the origin of adaptive radiations. J Evol Biol. 2010; 23:1581–96. https://doi.org/10.1111/j.1420-9101.2010.02029.x PMID: 20561138

121. Stroud JT, Losos JB. Ecological and adaptive radiation. Ann Rev Ecol Ecol Syst. 2016; 47:507–32. https://doi.org/10.1146/annurev-ecolsys-121415-032254

122. Myers GS. The endemic fish fauna of Lake Lanao and evolution of higher taxonomic categories. Evolution. 1960; 14:323–333. https://doi.org/10.1111/j.1558-5646.1960.tb03094.x

123. Nilsson PA, Brönmark C. Prey vulnerability to a gape-size limited predator: behavioural and morphological impacts on northern pike piscivory. Oikos. 2000; 88:539–46. https://doi.org/10.1034/j.1600-0706.2000.880310.x

124. Domenici P, Turesson H, Brodersen J, Brönmark C. Predator-induced morphology enhances escape locomotion in crucian carp. Proc Royal Soc B. 2007; 274:185–91. https://doi.org/10.1098/rspb.2007.1088

125. Powell M, Jones M, Lijalad M. Effects of skeletal deformities on swimming performance and recovery from exhaustive exercise in triploid Atlantic salmon. Dis Aquat Organ. 2009; 85:39–66. https://doi.org/10.3354/dao02056 PMID: 19593934

126. Prestinicolia L, Boglione C, Makridis P, Spanò A, Rimatori V., Palamara E, et al. Environmental conditioning of skeletal anomalies typology and frequency in gilthead seabream (Sparus aurata L.) 1758 juveniles. PLoS ONE. 2013; 8(2),e55736. https://doi.org/10.1371/journal.pone.0055736 PMID: 23409031

127. Bensimon-Brito A, Cancela ML, Huyssesse A, Witten PE. The zebrafish (Danio rerio) caudal complex—a model to study vertebral body fusion. J Appl Ichthyol. 2010; 26:235–38. https://doi.org/10.1111/j.1439-0426.2010.01412.x
128. Bensimon-Brito A, Cancela ML, Huysseune A, Witten PE. Vestiges, rudiments and fusion events: the zebrafish caudal fin endoskeleton in an evo-devo perspective. Evol Dev. 2012; 14:116–27. https://doi.org/10.1111/j.1525-142x.2011.00526.x PMID: 23016979

129. Shkil FN, Kapitanova DV, Borisov VB, Abdissa B, Smirnov SV. Thyroid hormone in skeletal development of cyprinids: effects and morphological consequences. J Appl Ichthyol. 2012; 28:398–405. https://doi.org/10.1111/j.1439-0426.2012.01992.x

130. Bolotovskiy AA, Levin BA. Effects of thyroid hormones on vertebral numbers in two cyprinid fish species: Rutilus rutilus (Linnaeus, 1758) and Abramis brama (Linnaeus, 1758). J Appl Ichthyol. 2018; 34:449–54. https://doi.org/10.1111/jai.13659

131. Ackerly KL, Ward AB. How temperature-induced variation in musculoskeletal anatomy affects escape performance and survival of zebrafish (Danio rerio). J Exp Zool A Ecol Genet Physiol. 2016; 325:25–40. https://doi.org/10.1002/jez.1993 PMID: 26499994

132. Banister KE. A revision of the large Barbus (Pisces, Cyprinidae) of East and Central Africa. Part II. Bull Br Mus Nat Hist (Zool). 1973; 26:1–148. https://doi.org/10.5962/bhl.part.204

133. Golubtsov AS, Darkov AA, Dgebuadze YY, Mina MV. An artificial key to fish species of the Gambela region (the White Nile basin in the limits of Ethiopia). Addis Abeba: Joint Ethio-Russian Biological Expedition; 1995.

134. Moritz T, El Dayem ZM, Abdallah MA, Neumann D. New and rare records of fishes from the White Nile in the Republic of the Sudan. Cybium. 2019; 43:137–51. https://doi.org/10.26028/cybium/2019-423-011

135. Al-Hassan LAJ, Naama AK. More vertebral anomalies in fishes from Euphrates River, Iraq. Indian J Fish. 1986; 33:237–41.

136. Jawad LA, Al-Faisal AJ, Al-Mukhtar M. A case of vertebral coalescence in Luciobarbus xanthopterus (Heckel, 1843) (Pisces: Cyprinidae) obtained from the lower reaches of Mesopotamia. Trav Mus Natl Hist Nat Grigore Antipa. 2015; 57:127–32. https://doi.org/10.1515/travmu-2015-0006

137. Fjeldal PG, Madaro A, Hvas M., Stien HL, Oppedal F, Fraser TWK. Skeletal deformities in wild and farmed cleaner fish species used in Atlantic salmon Salmo salar aquaculture. J Fish Biol. 2020. https://doi.org/10.1111/jfb.14337