Arabidopsis PCaP2 Functions as a Linker Between ABA and SA Signals in Plant Water Deficit Tolerance

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Water stress has a major influence on plant growth, development, and productivity. However, the cross-talk networks involved in drought tolerance are not well understood. Arabidopsis PCaP2 is a plasma membrane-associated Ca²⁺-binding protein. In this study, we employ qRT-PCR and β-glucuronidase (GUS) histochemical staining to demonstrate that PCaP2 expression was strongly induced in roots, cotyledons, true leaves, lateral roots, and whole plants under water deficit conditions. Compared with the wild type (WT) plants, PCaP2-overexpressing (PCaP2-OE) plants displayed enhanced water deficit tolerance in terms of seed germination, seedling growth, and plant survival status. On the contrary, PCaP2 mutation and reduction via PCaP2-RNAi rendered plants more sensitive to water deficit. Furthermore, PCaP2-RNAi and pcap2 seedlings showed shorter root hairs and lower relative water content compared to WT under normal conditions and these phenotypes were exacerbated under water deficit. Additionally, the expression of PCaP2 was strongly induced by exogenous abscisic acid (ABA) and salicylic acid (SA) treatments. PCaP2-OE plants showed insensitive to exogenous ABA and SA treatments, in contrast to the susceptible phenotypes of pcap2 and PCaP2-RNAi. It is well-known that SNF1-related kinase 2s (SnRK2s) and pathogenesis-related (PRs) are major factors that influence plant drought tolerance by ABA- and SA-mediated pathways, respectively. Interestingly, PCaP2 positively regulated the expression of drought-inducible genes (RD29A, KIN1, and KIN2), ABA-mediated drought responsive genes (SnRK2.2, -2.3, -2.6, ABF1, -2, -3, -4), and SA-mediated drought responsive genes (PR1, -2, -3) under water deficit, ABA, or SA treatments. Taken together, our results showed that PCaP2 plays an important and positive role in Arabidopsis water deficit tolerance by involving in response to both ABA and SA signals and regulating root hair growth. This study provides novel insights into the underlying cross-talk mechanisms of plants in response to water deficit stress.

Keywords: PCaP2, water deficit, ABA, SA, SnRK2, PR, Arabidopsis
INTRODUCTION

Water deficit is one of the most acute abiotic stresses affecting plant growth and the economic yield of crop plants. It leads to alterations in various cellular processes in plants, for example, gene expression, photosynthesis, protein synthesis, carbon partitioning, lipid metabolism, and osmotic homeostasis (Hua et al., 2012; Jarzyniak and Jasiński, 2014; Fleta-Soriano and Munné-Bosch, 2016). Phytohormones, such as abscisic acid (ABA), salicylic acid (SA), gibberellin (GA), indole-3-acetic acid (IAA), and jasmonic acid (JA) function as central factors that link and reprogram these complex cellular processes (Hua et al., 2012).

In response to water deficit, ABA, a well-known stress phytohormone, is rapidly induced, leading to the expression of stress-responsive genes and the activation of plants’ cellular physiological adaptation to water stress (Fujii and Zhu, 2009; Cutler et al., 2010; Weiner et al., 2010). In the ABA signaling pathway, SNF1-related kinase 2s (SnRK2s) are central regulators that mediate ABA-responsive transcription factors and genes to activate ABA-mediated physiological processes (Yoshida et al., 2002; Boudsocq et al., 2004; Fujita et al., 2009; Umezawa, 2009; Vlad et al., 2009; Raghavendra et al., 2010; Kulik et al., 2011; Ambrosone et al., 2015).

Among the 10 SnRK2s in Arabidopsis, SnRK2.2, SnRK2.3, and SnRK2.6 function as central regulators in response to ABA and drought. Genetic analysis has shown that the Arabidopsis triple snrk2.2/snrk2.3/snrk2.6 mutant indicates that these three SnRK2s are partially redundant, although all of them are crucial for plants’ response to water stress and ABA, as well as ABA-mediated seed germination and dormancy. Seed dormancy, germination, and seedling growth of snrk2.2/snrk2.3 mutants are greatly insensitive to exogenous ABA. In contrast, snrk2.6 shows a significant increase in leaf water loss after ABA treatment (Yoshida et al., 2002; Fujii et al., 2007; Nakashima et al., 2009). This could be because SnRK2.6/ OST1 are important for stomatal movements, as they phosphorylate anion (SLAC1) and cation (KAT1) channels, which might be required for ABA-dependent stomatal closing in response to water deficit (Pilot et al., 2001; Geiger et al., 2009, 2010; Lee et al., 2009; Sato et al., 2009). In addition, guard cells (GCs) display transcriptional memory in a daily dehydration stress and watered recovery cycle. SnRK2.2, SnRK2.3, and SnRK2.6 have distinguishable roles in the process: SnRK2.2 and SnRK2.3 are more important for implementing guard cell stress memory, while SnRK2.6 is more important for overall stomatal control in the subsequent dehydration response (Virlouvet and Fromm, 2015). It has been found that SnRK2.2, -2.3, and 2.6 are regulated by nitric oxide (NO), phosphatidic acid (PA), and Ca\(^{2+}\) changes (Boudsocq et al., 2004; Fujii and Zhu, 2009; Kulik et al., 2011), suggesting SnRK2s may be regulated by complex pathways during water stress; however, such mechanisms remain largely unknown.

The phytohormone SA plays an important role in various plant developmental processes and responses to abiotic and biotic stress (Raskin, 1992; Bandurska and Stroiński, 2005; Khan et al., 2012a,b, 2013). Water deficit induces increased endogenous SA levels in various plants (Munne-Bosch and Penuelas, 2003; Miura and Tada, 2014). Exogenous treatment with SA modulates plant drought resistance through multiple pathways such as oxidative stress (Alam et al., 2013), stomatal conductance (Hao et al., 2010; Khokon et al., 2011; Habibi, 2012), antioxidant defense system (Hayat et al., 2008; Saruhan et al., 2012), and NO production (Hao et al., 2010; Khokon et al., 2011). Additionally, some SA-responsive genes are involved in plant response to water deficit, such as GST1, GST2, GR, and MDHAR in Triticum aestivum (Kang et al., 2013) and MPK3, MPK4, MPK6, PR1, PR2, and PR5 in Arabidopsis thaliana (Ichimura et al., 2000; Ahlfors et al., 2004; Gudesblat et al., 2007; Liu P. et al., 2013; Liu W.X. et al., 2013). Some Arabidopsis mutants that accumulate endogenous SA (adr1, acd6, cpr5 myb96-1d, and siz1) show both SA-mediated disease resistance and water deficit tolerance (Miura et al., 2013). One genetic analysis reports that Arabidopsis seedlings overexpressing PR1, PR2, or PR5 are drought tolerant (Liu W.X. et al., 2013). In addition, PR1, -2, and -5 genes are widely used as marker genes for SA-mediated drought tolerance in plants. For example, both SA-accumulating mutants (cpr5 and acd6) and overexpression of transcription factor Di19 in Arabidopsis improve drought tolerance via SA-induced expression of PR1, -2, -5 genes (Liu P. et al., 2013; Liu W.X. et al., 2013). Interestingly, SA treatments lead to an increase of ABA and proline in the barley leaves (Bandurska and Stroiński, 2005); however, the relationship between SA and ABA signals in water deficit remains unknown.

Arabidopsis microtubule-associated protein-18/plasma membrane-associated Ca\(^{2+}\)-binding protein-2 (MAP18/PCaP2) is important for several physiological activities. For example, it is involved in Ca\(^{2+}\) binding and the organization of cortical microtubules (MTs) and F-actin. It also has a critical role in root hair, pollen tube growth, and directional cell growth (Wang et al., 2007; Kato et al., 2010, 2013; Zhu et al., 2013; Zhang et al., 2015; Kang et al., 2017). For example, the cell polarity and cortical microtubule array in line 2 of MAP18-overexpressing Arabidopsis (OE2) and line 18 of MAP18 RNAi transgenic Arabidopsis (R18) are altered (Wang et al., 2007). The T-DNA

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**Abbreviations:** ABA, abscisic acid; ABF, abscisic acid responsive element-binding factor; ABI, abscisic acid insensitive; AREBs/ABFs, ABRE-binding proteins/factors; Ca\(^{2+}\), calcium ion; CA, calmodulin; CCD, charge-coupled device; CPK, calcium-dependent protein kinase; Di19, drought-induced 19; GA, gibberellin; GR, glutathione reductase; GST, glutathione s-transferase; GUS, β-glucuronidase; IAA, indole-3-acetic acid; IAA, jasmonic acid; KIN, kinase; MDHAR, Monodehydroascorbate reductase; MAP18/PCaP2, microtubule-associated protein18/plasma membrane-associated Ca\(^{2+}\)-binding protein-2; MT, microtubule; MS, Murashige Skoog; NO, nitric oxide; OST1, open stomata1; PA, phosphatidic acid; PCaP2-OE, PCaP2 overexpression; PCaP2-RNAi, PCaP2 RNA interference; PEG, polyethylene glycol; PR, pathogenesis-related; PtdInsPs, phosphatidylinositol phosphates; qRT-PCR, quantitative real-time PCR; R2D29A, responsive to desiccation 29A; RT-PCR, reverse transcriptase-mediated PCR; SA, salicylic acid; S. lycopersicum, Solanum lycopersicum; SNF, sucrose non-fermenting; SnRK2, SNF1-related protein kinase 2; WT, wild type; Z. mays, Zea mays; RGGA, Arginine Glycine Glycine (RGG) box-containing RNA-binding protein; Di19, drought-induced protein19.
insertion line map18 (SALK_021652), which is confirmed as a knock-down mutant by qRT-PCR analysis, displays abnormal pollen tube growth and root hair growth (Kato et al., 2013; Zhu et al., 2013; Zhang et al., 2015; Kang et al., 2017). Interestingly, the mRNA expression level of PCaP2 is induced by heat, cold, drought, ABA, SA, osmotic stress, and GA3 (Kato et al., 2010, 2013). This implies that it may function in response to abiotic stress and phytohormone signals. Root hairs are the main sites of water absorption in plants, which is important for water deficit tolerance (Worrall and Roughley, 1976; Zahran and Sprent, 1986; Schnall and Quatrano, 1992). Thus, we hypothesized that PCaP2 might be an important regulator of plant water deficit tolerance in various pathways, such as phytohormone signals and root hair growth, suggesting that PCaP2 might be a cross-talker between complex mechanisms involved in plant water deficit tolerance. In this study, we found that PCaP2 is vital for plant water deficit tolerance by responding to ABA and SA signals, regulating the expression of the key ABA- and SA-mediated genes, and affecting root hair growth. Collectively, our data provide novel evidence of the underlying complex mechanisms, especially of crosstalk between ABA and SA signaling pathways in plant water deficit tolerance.

RESULTS

The Expression of PCaP2 Is Highly Induced in All Tissues in Water Deficit

To fully understand the expression pattern of PCaP2 under water deficit stress, we examined the expression of PCaP2 in more details by quantitative real-time PCR (qRT-PCR) and β-glucuronidase (GUS) staining. Firstly, the wild type (WT) plants were exposed to dehydration conditions for 1, 3, 6, 9, and 12 h. The qRT-PCR results showed that PCaP2 expression was highly induced by water deficit treatments from 1 to 12 h, with the peak level of 10-fold increased at 6 h treatment (Figure 1A). Furthermore, GUS staining showed that the promoter activity of PCaP2 was significantly increased after water deficit treatments for 6 h which was consistent with the results of qRT-PCR (Figure 1B) and PCaP2 expression was induced in primary roots and lateral roots, cotyledons, true leaves, and the whole seedlings after water deficit treatments (Figures 1B–D).

PCaP2-OE Plants Display Increased Tolerance While PCaP2 RNAi and Mutant Seedlings Are Hypersensitive in Response to Water Deficit

To elucidate the function of PCaP2 in plant tolerance to water deficit, the previous identified one PCaP2 overexpression (PCaP2-OE) line (Wang et al., 2007), one knockdown of T-DNA insertion (pcap2) line (SALK_021652; Kato et al., 2013; Zhu et al., 2013; Zhang et al., 2015; Kang et al., 2017), and one fully silenced PCaP2 RNA interference (PCaP2-RNAi) line (Wang et al., 2007) were used. The PCaP2 expression of these lines was analyzed by qRT-PCR which is consistent with the previous publications (Supplementary Figure S1). We firstly investigated their seed germination rates under normal or drought conditions. Under normal conditions, germination rates of PCaP2-RNAi and pcap2 seeds were lower than those of PCaP2-OE and WT seeds at 1 day, and then these seeds gradually showed similar germination rate. At 4 and 5 days, the germination of these seeds was identical (Figure 2A). Water deficit significantly inhibited the germination of all seeds at 1 day, germination rates of PCaP2-RNAi were lower from 1 to 4 days, then the germination of these seeds was identical germination at 5 days (Figure 2A).

Next, the growth and survival status of the four-day-old seedlings were observed in water deficit treatments. It was found that the grown status showed no significant differences among WT, PCaP2-OE, pcap2, and PCaP2-RNAi lines under normal conditions. The PCaP2-OE seedlings showed longer roots and larger leaves than WT while pcap2 and PCaP2-RNAi showed opposite growth phenotypes under water deficit (Figures 2B,C). The results of survival status after water deficit treatments showed that PCaP2-OE plants were more tolerant in response to water deficit than WT (Figures 2C,D) while pcap2 and PCaP2-RNAi plants exhibited sensitivity to water deficit. To determine recovery after water deficit, we re-watered these seedlings for 7 days. The results showed that PCaP2-OE plants, WT, and some pcap2 seedlings were recovered while all PCaP2-RNAi seedlings exhibited water deficit sensitivity (Figure 2D). The changes of leaf water loss and root water absorption in WT, PCaP2-OE, pcap2, and PCaP2-RNAi lines under water deficit conditions were observed. The results showed that pcap2 and PCaP2-RNAi lines lost water faster and wilted earlier than WT in dehydration stress while PCaP2-OE seedlings showed opposite phenotypes (Figures 2E,F). Additionally, the pcap2 and PCaP2-RNAi lines displayed shorter root hairs and less relative water content than WT in normal condition, the phenotypes were more significant in water deficit stress (Figures 2G–I). These results indicated that overexpression of PCaP2 enhanced the tolerance of plants to water deficit and down of PCaP2 led to plant water deficit hypersensitivity.

The PCaP2 Expression Is Highly Induced in Response to Exogenous ABA and SA Treatments

Abscisic acid and SA are important regulatory signals in water deficit stress, and the higher expression level of PCaP2 has been found in exogenous 100 µM ABA and 100 µM SA treatments (Kato et al., 2010; Trivedi et al., 2016). To determine PCaP2 in ABA and SA signaling pathways, the expression of PCaP2 was examined in exogenous ABA and SA treatments by qRT-PCR and GUS staining assay in more details. Because the expression of many genes in Arabidopsis can be induced by 10–100 µM ABA (Zhu et al., 2007, 2017; Kato et al., 2010; Tian et al., 2015) and we found the expression of PCaP2 is higher in 40 µM ABA treatments than that in 100 µM ABA treatment (Figure 3A; Kato et al., 2010). Thus, 40 µM ABA and 100 µM SA treatments were used in our studies. The results showed that the PCaP2 mRNA level was significantly increased with ABA and SA treatments from 1 to 12 h, the peaks appeared at 6 h ABA and 3 h SA treatments (Figures 3A,B).
The expression level of PCaP2 was higher in 40 μM ABA than 100 μM SA treatments (Figures 3A,B). Further, the PCaP2 expression was induced in the whole seedlings, cotyledons, true leaves, hypocotyls, primary roots, and lateral roots by ABA and SA treatments (Figures 3B–E), which were consistent with the results from water deficit treatments.

**PCaP2-OE Plants Show Increased Tolerance, While Its RNAi and Mutant Are More Sensitive in Exogenous ABA and SA Treatments**

Then we tested the germination rate of seeds and seedling growth of WT, PCaP2-OE, pcap2, and PCaP2-RNAi lines under normal, ABA, and SA treatments. To choose the most appropriate concentrations for ABA and SA treatments, 0.01–40 μM of ABA and SA were tested in the previous experiments used (data not shown). The results showed WT and PCaP2-OE showed the similar germination rates. The PCaP2-RNAi and pcap2 showed much higher germination rates in 0.8 μM ABA and 0.3 mM SA treatments, compared to WT and PCaP2-OE (Figure 4A). The germination rates of PCaP2-RNAi and pcap2 seeds were lower than those of PCaP2-OE and WT seeds in 1-to-5-day ABA and in 1-to-3-day SA treatments. The germination rates of these seeds were similar at day 4 of SA treatment and identical at day 5 of SA treatment. The growth status of WT, PCaP2-OE, pcap2, and PCaP2-RNAi lines was significantly different under 0.5 μM ABA and 0.05 mM SA treatments (Figures 4B,C). The PCaP2-OE seedlings showed larger leaf area and longer primary roots than WT while pcap2 and PCaP2-RNAi lines showed smaller leaves and shorter roots and root hairs, which were consistent with the water deficit inducible phenotypes (Figures 4B,C).

**PCaP2 Positively Regulates the Expression of SnRK2 Genes and PR Genes Under Water Deficit Stress**

To analyze whether the role of PCaP2 in water deficit tolerance was mediated by ABA and SA signaling pathways, we firstly checked the PCaP2 expression of WT, PCaP2-OE, pcap2, and
FIGURE 2 | Effects of PCaP2 on Arabidopsis seed germination and seedling growth under water deficit stress. (A) Quantification of germination rates of the PCaP2-OE, WT, pcap2 and PCaP2-RNAi seeds in 1/2 MS medium or coated with 25 % PEG from 1 to 5 d. At least 50 seeds from each sample were used for every technical replicate and three biological replicates were conducted. (B) Phenotypes of the PCaP2-OE, WT, pcap2 and PCaP2-RNAi seedlings grown in water deficit. Scale bar = 1 cm. (C) The phenotypes statistics of the PCaP2-OE, WT, pcap2 and PCaP2-RNAi seedlings grown in water deficit. At least 70 roots or 200 leaves from about 70 seedlings from each sample were used for every technical replicate and three biological replicates were conducted. (D) The survival status of PCaP2-OE, WT, pcap2 and PCaP2-RNAi in normal, water deficit and rehydration matrix soil, respectively. Scale bar = 1 cm. (E) Phenotypes of leaves wilted of PCaP2-OE, WT, pcap2 and PCaP2-RNAi plants for 0 h (up) and 4 h (down) under dehydration stress. Scale bar = 1 cm. (F) Water loss measurements for genotypes. Water loss in detached leaves was measured at the time points indicated. Water loss was expressed as a percentage of initial fresh weight. At least 30 detached leaves from each sample were used for every technical replicate and three biological replicates were conducted. (G) Phenotypes of root hairs in WT, pcap2, and PCaP2-RNAi lines in 1/2 MS or coated with 15% PEG. Scale bar = 0.5 mm. (H) The statistics of average root hair length in (G). About 250 root hairs from 30 roots of each sample were used for every technical replicate and three biological replicates were conducted. (I) The relative water content (RWC) of WT, pcap2 and PCaP2-RNAi lines in 1/2 MS or coated with 15% PEG. About 30 seedlings from each sample were used for every technical replicate and three biological replicates were conducted. All data are mean values of three biological replicates ± SE. The significant difference was determined by ANOVA: *P < 0.05, **P < 0.01, ***P < 0.001.
**FIGURE 3** | Expression pattern of PCaP2 with exogenous ABA or SA treatments. (A) Relative expression of PCaP2 in response to exogenous ABA and SA treatments. Fourteen-day-old seedlings were treated with 40 µM ABA or 100 µM SA for 0, 1, 3, 6, and 12 h by qRT-PCR analysis. Data represent mean values of three biological replicates ± SE. The significant difference was determined by ANOVA in comparison to 0 h: ***P < 0.001. (B) Comparison of PCaP2 expression levels with exogenous ABA or SA treatments by GUS staining. Seven-day-old seedlings of pPCaP2::GUS transgenic plants were treated with 40 µM ABA or 100 µM SA for 0, 1, 3, 6, and 12 h. Scale bar = 2.5 mm. (C–E) Analysis of the expression pattern of PCaP2 by GUS staining of pPCaP2::GUS transgenic seedlings in normal (without ABA or SA treatments) (C) and 40 µM ABA treatments for 6 h (D) and 100 µM SA treatments for 6 h (E). At least 15 seedlings from each sample were used for every technical replicate and three biological replicates were conducted. The scale bar is 5 mm in the pictures of whole plants, and 0.25 mm in the pictures of cotyledons, true leaves, hypocotyls, lateral roots, and main roots.

PCaP2-RNAi in water deficit conditions by qRT-PCR. The analysis showed that the increasing expression of PCaP2 in PCaP2-OE and the decreasing expression in pcap2 and PCaP2-RNAi were compared with that in WT under water deficit treatments (Supplementary Figure S1).

Next, we examined the expression of the following ABA-responsive genes, including SnRK2.2, -2.3, -2.6, ABF2, -3, -4 (Choi et al., 2000; Uno et al., 2000; Boudsocq et al., 2004; Gong et al., 2015; Jia et al., 2015); drought-inducible genes, including RD29A (Yamaguchi-Shinozaki and Shinozaki, 1994); KIN1 and KIN2 (Kurkela and Borg-Franck, 1992); and SA-responsive genes such as PRI, -2, -5 genes (Liu P. et al., 2013; Liu W.X. et al., 2013) in WT, PCaP2-OE, pcap2 and PCaP2-RNAi with dehydration conditions at 0, 6, and 12 h by qRT-PCR.
FIGURE 4 | Effects of PCaP2 on Arabidopsis seed germination and seedling growth in ABA or SA treatments. (A) Quantification of germination rates of the PCaP2-OE, WT, pcap2, and PCaP2-RNAi seeds in 1/2 MS medium or mixed with 0.8 μM ABA and 0.3 mM SA from 1 to 5 days. A total of 40 seeds of each line were used for every technical replicate and three biological replicates were conducted. (B) Phenotypes of the PCaP2-OE, WT, pcap2, and PCaP2-RNAi seedlings grown in exogenous ABA and SA treatments. The seedlings were sown on 1/2 MS medium for 5 days, then transferred to 0.5 mM ABA or 0.05 mM SA medium and grew for 10 days. Scale bar = 1 cm. (C) The phenotype statistics of the PCaP2-OE, WT, pcap2, and PCaP2-RNAi seedlings grown in exogenous ABA and SA treatments. The main root length and leaf area of these seedlings were calculated after growth on 1/2 MS medium or supplemented with 0.5 mM ABA and 0.05 mM SA. At least 70 roots or 200 leaves from about 70 seedlings from each sample were used for every technical replicate and three biological replicates were conducted. All data are mean values of three biological replicates SE. The significant difference was determined by ANOVA: *P < 0.05, **P < 0.01, ***P < 0.001.

analysis. Under normal conditions, pcap2 and PCaP2-RNAi inhibited the expression of ABA- and drought-responsive genes, except SA-responsive genes (Figure 5), and PCaP2-OE increased the expression of SA-responsive genes, suggesting that PCaP2 might be the complex mechanisms in regulating gene expression in normal conditions. Under water deficit stress, the expression of all these genes, including ABA-, drought-, and SA-responsive genes, was significantly inhibited in pcap2 and PCaP2-RNAi while increased in PCaP2-OE, compared with that in WT, indicating that PCaP2 was involved in regulating drought-, ABA- and SA-responsive genes under water deficit stress.

PCaP2 Positively Regulates ABA-Mediated SnRK2s Expression and SA-Mediated PRs Expression in Water Deficit

To further analyze whether the effect PCaP2 on gene expression in response to water deficit was mediated by ABA and SA signaling pathways, we firstly checked the PCaP2 expression of WT, PCaP2-OE, pcap2 and PCaP2-RNAi in ABA and SA treatments by qRT-PCR. The analysis showed that the increase expression of PCaP2 in PCaP2-OE and the decreasing expression in pcap2 and PCaP2-RNAi were compared with that in WT.
FIGURE 5  | Effect of PCaP2 on the expression of ABA-, drought- and SA-responsive genes under water deficit. The relative gene expression of the 14-day-old seedlings of WT, PCaP2-OE, pcap2, and PCaP2-RNAi with water deficit treatments for 0, 6, and 12 h was determined by qRT-PCR analysis. Data represent mean values of three biological replicates ± SE. The significant difference was determined by ANOVA: *P < 0.05, **P < 0.01, ***P < 0.001.

under ABA and SA treatments (Supplementary Figure S1). Next, we examined the expression of SnRK2.2, -2.3, -2.6 and PR1, -2, -5 genes in WT, PCaP2-OE, pcap2, and PCaP2-RNAi with ABA and SA treatments at 0, 6, and 12 h by qRT-PCR analysis. The results showed that the effect of PCaP2-OE, pcap2, and PCaP2-RNAi on affecting the ABA-responsive gene expression in ABA treatments, rather than SA treatments, were similar to that in drought treatments, (Figure 6A). In contrast, the effect of PCaP2-OE, pcap2, and PCaP2-RNAi on affecting the SA-responsive gene expression in SA treatments, rather than ABA treatments, were similar to that in drought treatments (Figure 6B). These results illustrated that PCaP2 regulated the expression of ABA-mediated SnRK2 genes and SA-mediated PR genes under water deficit stress, indicating that PCaP2 was a crosstalk factor in response to ABA and SA signals in water deficit.

DISCUSSION

Water deficit significantly influences plant growth, development, and productivity. One of the most important regulators in drought is ABA, and some cases of SA-mediated plant water deficit tolerance have been documented. However, details of crosstalk between ABA and SA signaling cascades in response to water deficit remain largely unknown. Previous studies have suggested that PCaP2 responds to some phytohormone signaling pathways and abiotic stress through mRNA expression pattern analysis (Kato et al., 2010). Our results further support a key role for PCaP2 in Arabidopsis water deficit tolerance and suggest that it is connected to the main ABA and SA signaling pathways.

Previous studies have shown that PCaP2 is mainly expressed in roots and flower tissues in normal conditions, and its mRNA level in whole seedlings is induced by drought as well as treatment with ABA or SA (Wang et al., 2007; Kato et al., 2010). Compared with previous studies, our data provide more detailed findings. High expression of PCaP2 in roots was found under normal and stress conditions. This might be related to its function in root water absorption and root hair growth in both normal and water deficit conditions. Additionally, PCaP2 was not expressed in true leaves under normal conditions, but was highly induced in water deficit and treatment with ABA or SA. This indicated that in true leaves, the increase of PCaP2 level is a key response to stress that is triggered by phytohormone signals, which may be associated with its function in controlling water deficit-induced leaf water loss.

Some ABA-sensitive seedlings show improved drought tolerance in soil, such as SnRK2s-OE and AREBs-OE, while some mutants show the same sensitivity to ABA and drought treatments, such as Atrggg and AtDi19-3 (Fujita et al., 2009; Qin et al., 2014; Ambrosone et al., 2015). Generally, ABA-sensitive mutants improve drought tolerance by regulating stomatal movement. However, changes in the drought tolerance of Atrggg and AtDi19-3 are not dependent on stomatal movement (Fujita et al., 2009; Qin et al., 2014; Ambrosone et al., 2015). In the present study, we used the previously identified PCaP2-OE line (Wang et al., 2007) pcap2 mutant (Kato et al., 2013; Zhu et al., 2013; Zhang et al., 2015; Kang et al., 2017) and PCaP2-RNAi lines (Wang et al., 2007), and our analysis of PCaP2 expression of these lines is the same with the prior findings. Our results showed that pcap2 and PCaP2-RNAi lines were more sensitive to ABA and drought, while PCaP2-OE plants were more resistant to ABA and drought stress. Although our results do not address whether PCaP2 can regulate stomatal movement, it is possible that PCaP2 has a similar mechanism to AtrRGGA and AtDi19-3. PCaP2 do not fully depend on the regulation.
of stomatal movement in response to drought stress; PCaP2 functionality in plant water deficit tolerance can be borne out by regulating root hair growth to benefit water absorption and to induce the expression of numerous key drought-responsive genes (Figure 7). Furthermore, the water deficit-inducible phenotypes in fully silenced PCaP2-RNAi plants were greater in magnitude to those in knockdown pcap2 mutants. This suggests that the role of PCaP2 in water deficit is dependent on its inducible expression level. These results indicate that PCaP2 is a positive regulator of water deficit tolerance in Arabidopsis.

Root hairs are the major water absorbing tissue in plants. Under water-limited conditions, root hair growth is significantly inhibited, which affects root water absorption and plant water deficit tolerance (Worrall and Roughley, 1976; Zahran and Sprent, 1986; Schnall and Quatrano, 1992). Our results showed that reduced expression of PCaP2 leads to shorter root hairs under normal conditions, which is consistent with previous results from Zhang et al. (2015). Moreover, significant inhibition of shorter root hair development was seen under water deficit conditions in pcap2 and PCaP2-RNAi, which could be one main pathway through which PCaP2 induces plant water deficit tolerance.

In Arabidopsis, SnRK2.2, -2.3, and -2.6 are crucial in the response to water deficit stress (Shi and Yang, 2014; Trivedi et al., 2016). Prior studies have shown that SnRK2.2, -2.3, and -2.6 triple mutants are insensitive to ABA and water deficit treatments (Fujii and Zhu, 2009; Fujita et al., 2009). Thus, PCaP2 mainly influences plant water deficit tolerance by regulating the ABA-mediated SnRK2 signaling pathway. It has been shown that SnRK2.6 functionally separates from SnRK2.2 and -2.3 under water deficit stress (Fujii et al., 2007; Kulik et al., 2011). Furthermore, SnRK2.2 and SnRK2.3 kinases regulate seed dormancy, germination, and seedling growth under water deficit conditions (Fujii et al., 2007; Fujita et al., 2009). SnRK2.6/OST1 plays an important role in stomatal movement (Vlad et al., 2009; Vahisalu et al., 2010; Brandt et al., 2012; Acharya et al., 2013; Imes et al., 2013; Grondin et al., 2015; Matrosova et al., 2015; Yin et al., 2016), suggesting that it is involved in plant drought tolerance by regulating stomatal movement. Some studies have further demonstrated that SnRK2.2, -2.3, and -2.6 are crucial in the response of Arabidopsis to ABA and water deficit stress (Shi
SnRK2.2 and Yang, 2014; Trivedi et al., 2016). The expression of ABA-dependent transcription factors, such as ABIs and ABFs, is also stimulated by SnRK2s (Fujii et al., 2007). These factors mediate the expression of downstream ABA-inducible genes to improve plant water deficit resistance. Our results illustrated that PCaP2 increases SnRK2.2, -2.3, and -2.6 expression as well as SnRK2-mediated gene expression, such as ABF1, -2, -3, -4, KIN1, KIN2, and RD29, in water deficit stress and ABA treatments. Consequently, it can be speculated that water deficit stress-induced PCaP2 may affect the ABA-mediated SnRK2 signaling pathway and thus regulate drought-inducible gene expression, seed germination, seedling growth, and leaf water loss.

Compared to the ABA signaling pathway, relatively few cases of SA-mediated drought tolerance have been presented. Kang G. et al. (2012) and Kang G.Z. et al. (2012) identified 76 proteins as potentially involved in the SA signaling pathway in drought-exposed T. aestivum. Genetic and gene expression analysis of these proteins show that PRI, -2, and -5 are required in SA signaling in drought stress (Liu P. et al., 2013; Liu W.X. et al., 2013). Arabidopsis cpr5 and acd6 mutants exhibit SA accumulation, and their drought tolerance is improved by inducing SA-mediated expression of PRI, -2, and -5 (Liu P. et al., 2013). Additionally, PRI, -2, and -5 overexpression enhances drought tolerance in Arabidopsis (Liu W.X. et al., 2013; Qin et al., 2014).

Transcription factor Di19 regulates the expression of PRI, -2, and -5 in drought conditions (Liu W.X. et al., 2013; Qin et al., 2014). Water loss rates of PRI-, PR2-, and PR5-overexpressing lines are lower than those of WT plants, and the three genes are highly expressed in stomata, suggesting that PRI, PR2, and PR5 may regulate stomatal movement in response to drought tolerance (Liu W.X. et al., 2013; Qin et al., 2014). Additionally, PRs play significant roles in regulating phytohormone-signaling, such as auxin and JA (Thaler et al., 1999; Wang et al., 2001; Wen et al., 2008; Iglesias et al., 2011), suggesting that PR1, PR2, and PR5 may mediate the complex crosstalk between phytohormone signals under drought stress. However, complete mechanisms of PR actions under drought stress are largely unknown. Our results show that PCaP2 positively regulates the expression of PRI, -2, and -5 after SA and water deficit treatments, but not ABA treatment. In addition, PCaP2-RNAi and pcap2 lines lost water faster and wilted earlier than WT lines, which stands in contrast to the phenotypes of PRI-, PR2-, and PR5-overexpressing seedlings (Liu W.X. et al., 2013; Qin et al., 2014). However, the phenotypes of PCaP2-OE plants are consistent with those of PRs-overexpressing seedlings. Thus, these results support the idea that PCaP2 activates the key SA-mediated signaling pathway in response to water deficit, providing a new pathway to regulate PRs under water deficit conditions.

Interactions between SA and ABA signals have been shown to occur during abiotic stress, such as salt and cold stress. In S. lycopersicum, treatment with SA improves plant growth, osmotic adaptation, and ABA accumulation under normal conditions and during salt stress (Szepesi et al., 2009). Under cold conditions, exogenous ABA treatment increases endogenous SA level and oHCA content in Z. mays, suggesting that the ABA signal may combine with SA-related stress responses during cold stress (Szalai et al., 2011). Our results showed that ABA and SA can trigger PCaP2, and that PCaP2 then regulates the ABA-mediated SnRK2 signaling pathway as well as the expression of SA-mediated PR genes under water deficit stress. Thus, PCaP2 mediates crosstalk in response to ABA and SA signals during water deficit.

We have not interrogated whether PCaP2 can directly regulate SnRK2s and/or PRs in normal, water deficit, or exogenous ABA and SA conditions. However, previous studies have shown that PCaP2 can bind PtdInsPs, Ca\(^{2+}\) and Ca\(^{2+}/CaM\) complexes. These ligands are important components of intracellular signaling networks involved in plant water deficit tolerance (Zielinski, 1998; Meijer and Munnik, 2003; Carlton and Cullen, 2005; Perera et al., 2008; DeFalco et al., 2009; Kleerekoper and Putkey, 2009; Luan, 2009; Xue et al., 2009; Kato et al., 2010, 2013). Our results also showed that PCaP2 regulated the expression of many key ABA-responsive genes, SA-responsive genes, and drought-inducible genes, including upstream genes such as SnRK2.2, -2.3, and -2.6. Thus, it is well-accepted that PCaP2 mainly functions as a Ca\(^{2+}\)-binding protein to participate in intracellular signaling transduction under water deficit stress.

In conclusion, water deficit triggers ABA and SA accumulation, which, in turn, induce PCaP2 expression. PCaP2 then activates the expression of many key drought-responsive genes, including ABA-mediated genes (SnRK2.2, -2.3, -2.6, ABF1, -2, -3, -4) and SA-mediated genes (PRI, -2, -5) and drought-inducible genes (RD29A, KIN1 and KIN2), and affects...
root hair growth to increase water absorption, which improve plant water deficit tolerance (Figure 7). This study highlights the positive and important role that PCaP2 plays in plant water deficit tolerance by mediating crosstalk between ABA and SA signals, providing novel evidence relating to the underlying and complex mechanisms mediated by ABA and SA signals during plant water deficit tolerance.

MATERIALS AND METHODS

Plant Materials, Growth Conditions, and Treatments

Arabidopsis ecotype Col-0 was the background for all transgenic and mutant plants in this study. The seeds of PCaP2-OE line (Wang et al., 2007), pcap2 mutant (Kato et al., 2013; Zhu et al., 2013; Zhang et al., 2015; Kang et al., 2017), and PCaP2-RNAi line (Wang et al., 2007) and PCaP2 promoter::GUS (pPCaP2::GUS) (Wang et al., 2007) were provided by Professor Ming Yuan, China Agricultural University, Beijing, China. All these seeds were used in the previous reports.

Surface sterilized Arabidopsis seeds were sown on MS plates with 1/2 MS (Murashige & Skoog) medium (pH adjusted to 5.8–6.0 with 1 M NaOH) with vitamins, 0.6 % phytoagar (PlantMedia), and 1% sucrose, and then in darkness at 4°C for 3 days. Seedlings were then transferred to a growth chamber at 22°C, with a 16 h/8 h (light/dark) photoperiod at approximately 120 µmol m⁻² s⁻¹. The 1/2 MS plus 1% sucrose media are usually used in water deficit, ABA, and SA treatments (Jayakannan et al., 2015; Schwalmt et al., 2017; Zhu et al., 2017). PCaP2-OE, WT, pcap2, and PCaP2-RNAi were used for all experiments. For germination rate analysis, seeds were sown on 1/2 MS medium with 25% PEG (dehydration), 0.8 µM ABA and 0.3 mM SA for 1, 2, 3, 4, 5 days, respectively. For seedling growth experiments, 4-day-old seedlings were transferred to soil for 7 days, then with 40% PEG (dehydration), 0.5 µM ABA and 0.05 mM SA for 10 days. For survival experiments, 4-day-old seedlings were transferred to soil for 7 days, then were treated for 10 days without water, following with water for 7 days. At least 20 seedlings were harvested for observing in three biological replicates in each experiment. The best concentration of ABA and SA in the present paper was chosen by the prepared experiments for 0.01 to 40 µM of ABA and SA (data not shown), because the previous findings show that significantly different ABA and SA concentrations from 1 nM to 80 µM are used in seed germination and seedling growth experiments (Reyes and Chua, 2007; Zhu et al., 2007; Horváth et al., 2015; Huang et al., 2017). For water-loss assays, rosette leaves of comparable size from 4-week-old plants grown under long days were detached, placed on a Petri dish and weighted at a specified time (0, 0.5, 1, 1.5, 2, 2.5, 3, 3.5, and 4 h) after detachment. For root hair growth assays, 3-day-old seedlings were transferred to 1/2 MS medium with or without 15% PEG for 3 days, and the root hair length was assayed by ImageJ software1. Three-day-old seedlings transferred to 1/2 MS medium with or without 15% PEG for 7 days were used to test relative water content. Relative water content was measured as previous reports described (Brini et al., 2007).

RNA Isolation and Quantitative RT-PCR (qRT-PCR) Analysis

Total RNA was isolated from 1-day-old seedlings using RNasy plant mini kit (Qiagen), treated with RNase-Free DNase (Qiagen) at 37°C for 1 h to degrade genomic DNA, and 1 µg total RNA to synthesize cDNA by oligo-(dT) 20-primed reverse transcription by the Omniscript RT Kit (Qiagen). The cDNA was amplified using SuperReal PreMix Plus (SYBR Green, TIANGEN, China) in a 10 µL volume. The expression levels of 18S rRNA was used as an internal control. Analysis was performed using the BioRad Real-Time System CFX96TM C1000 Thermal Cycler (Singapore).

For the expression of PCaP2 under drought, ABA, and SA treatment assays, 14-day-old seedlings from WT plants were exposed 1/2 MS medium with or without drought, 40 µM ABA and 100 µM SA for 0, 1, 3, 6, 9, and 12 h. The previous finding showed the higher expression level of PCaP2 has been found in exogenous 100 µM ABA and 100 µM SA treatments in Arabidopsis (Zhu et al., 2007; Kato et al., 2010). The expression of many genes can be increased in 10 to 100 µM ABA treatments in Arabidopsis (Zhu et al., 2007; Kato et al., 2010; Tian et al., 2015; Zhu et al., 2017). We found the expression level of PCaP2 can be induced in 40 µM ABA treatments, and the expression is higher than that in 100 µM ABA treatments (Figures 3A; Kato et al., 2010). Thus 40 µM ABA was used in our experiments. At each time point, all seedlings were immediately frozen by liquid nitrogen and then stored at −80°C for RNA preparation. To assay the expression of drought-, ABA- and SA-responsive genes under water deficit stress in WT and pcap2, WT and mutant seedlings grown at normal conditions for 14 days were harvested and treated by dehydration stress for 0, 6, 12 h. To assay the expression of SnRK2 and PR genes in WT and pcap2 treatment with ABA and SA, the seedlings of 14-day-old at normal condition were harvested and treated with 40 µM ABA and 100 µM SA for 0, 6, 12 h. Total RNA extraction and reverse transcription were performed as described above. All primer pairs used for qRT-PCR examination are listed in Supplementary Table S1. Each representative experiment was performed with at least three replicates.

Histochemical Staining of GUS Activity

Seven-day-old or 14-day-old pPCaP2::GUS seedlings treated with 40% PEG, 40 µM ABA, or 100 µM SA for 0, 1, 3, 6, and 12 h were collected for observing the changes of GUS activity. The GUS staining procedure was executed by the method mentioned in Wang et al. (2007). Samples were observed on an Olympus microscope equipped with a color CCD camera (Sutter Instrument; LAMBDA 10-2) or by an Epson scanner.

1http://rsb.info.nih.gov/ij/
Statistical Analysis

The experiments were conducted at least three times, each of which contained three technical replicates. Data presented as the mean ± SE of three biological replicates. The significant difference was analyzed by SPSS statistical software (ver.16.0, SPSS Inc., Chicago, IL, United States) via one-way or two-way ANOVA (*P < 0.05, **P < 0.01, ***P < 0.001). In addition, we tested the normality of data about the main root length, leaf area, and root hair length under drought stress addition, we tested the normality of data by SPSS statistical software (ver.16.0, which contained three technical replicates. Data presented as the mean ± SE. Shapiro–Wilk test by SPSS (Supplementary Table S2). The detailed information of the chemicals and kits used in this study are listed in Supplementary Table S3.

AUTHOR CONTRIBUTIONS

CW and XW designed the study,YW, LW, HL, BZ, and QC performed the experiments and data analysis. XL, SB, and YL provided help in experimental methods. SZ participated in the discussion. MH and ST trained the use of experimental equipment. SY helped to revised the language and grammar. CW wrote the manuscript.

REFERENCES

Acharya, B. R., Jeon, B. W., Zhang, W., and Assmann, S. M. (2013). Open Stomata 1 (OST1) is limiting in abscisic acid responses of Arabidopsis guard cells. *New Phytol.* 200, 1049–1063. doi: 10.1111/nph.12469

Ahlfors, R., Macioszek, V., Rudd, J., Brosché, M., Schlichting, R., Scheel, D., et al. (2004). Stress hormone-independent activation and nuclear translocation of mitogen-activated protein kinases in *Arabidopsis thaliana* during ozone exposure. *Plant J.* 40, 512–522. doi: 10.1111/j.1365-313X.2004.02229.x

Alam, M. M., Hasannazaman, M., Nahar, K., and Fujita, M. (2013). Exogenous salicylic acid ameliorates short-term drought stress in mustard (*Brassica juncea* L.) seedlings by up-regulating the antioxidant defense and glyoxalase system. *Aust. J. Crop Sci.* 7, 1053–1063.

Ambroso, A., Batelli, G., Nurcato, R., Aurilia, V., Punzo, P., Bangarusamy, D. K., et al. (2015). The Arabidopsis RNA-binding protein AtRGGA regulates tolerance to salt and drought stress. *Plant Physiol.* 168, 292–306. doi: 10.1104/pp.114.255802

Bandurska, H., and Stroinski, A. (2005). The effect of salicylic acid on barley response to water deficit. *Acta Physiol. Plant.* 27, 379–386. doi: 10.1007/s11738-005-0015-5

Boudsocq, M., Barbier-Brygoo, H., and Laurière, C. (2004). Identification of nine sucrose nonfermenting 1-related protein kinases 2 activated by hyperosmotic and saline stresses in *Arabidopsis thaliana*. *J. Biol. Chem.* 279, 41758–41766. doi: 10.1074/jbc.M405259200

Brandt, B., Brodsky, D. E., Xue, S., Negi, J., Iba, K., Kangasjärvi, J., et al. (2012). Reconstitution of abscisic acid activation of SLAC1 anion channel by CPK6 and OST1 kinases and branched AB1 PP2C phosphatase action. *Proc. Natl. Acad. Sci. U.S.A.* 109, 10593–10598. doi: 10.1073/pnas.1116590109

Brini, F., Hanin, M., Lumberras, V., Amara, I., and Khouidi, H. H. (2007). Overexpression of wheat dehydro DHN-5 enhances tolerance to salt and osmotic stress in *Arabidopsis thaliana*. *Plant Cell Rep.* 26, 2017–2026. doi: 10.1007/s00299-007-0412-x

Carlton, J. G., and Cullen, P. J. (2005). Coincidence detection in phosphoinositide signaling. *Trends Cell Biol.* 15, 540–547. doi: 10.1016/j.tcb.2005.08.005

Choi, H., Hong, J., Ha, J., Kang, J., and Kim, S. Y. (2000). ABFs, a family of ABA-responsive element binding factors. *J. Biol. Chem.* 275, 1723–1730. doi: 10.1074/jbc.275.3.1723

Cutler, S. R., Rodriguez, P. L., Finkelstein, R. R., and Abrams, S. R. (2010). Abscisic acid: emergence of a core signaling network. *Annu. Rev. Plant Biol.* 61, 651–679. doi: 10.1146/annurev-arplant-042809-112122

DeFalco, T. A., Bender, K. W., and Snedden, W. A. (2009). Breaking the code: Ca2+ sensors in plant signaling. *Biochem. J.* 425, 27–40. doi: 10.1042/B120091147

Fleta-Soriano, E., and Munné-Bosch, S. (2016). Stress memory and the inevitable effects of drought: a physiological perspective. *Front. Plant Sci.* 7:143. doi: 10.3389/fpls.2016.00143

Fujii, H., Verslues, P. E., and Zhu, J. K. (2007). Identification of two protein kinases required for abscisic acid regulation of seed germination, root growth and gene expression in *Arabidopsis*. *Plant Cell Physiol.* 19, 485–494. doi: 10.1073/pnas.106.048538

Fujii, H., and Zhu, J. K. (2009). *Arabidopsis* mutant deficient in 3 abscisic acid-activated protein kinases reveals critical roles in growth, reproduction, and stress.* Proc. Natl. Acad. Sci. U.S.A.* 106, 8380–8385. doi: 10.1073/pnas.0903144106

Fujita, Y., Nakashima, K., Yoshida, T., Katagiri, T., Kodokoro, S., Kanamori, N., et al. (2009). Three SnRK2 protein kinases are the main positive regulators of abscisic acid signaling in response to water stress in *Arabidopsis*. *Plant Cell Physiol.* 50, 2123–2132. doi: 10.1093/pcp/pcp147

Geiger, D., Scherer, S., Mumm, P., Marten, I., Ache, P., Matschi, S., et al. (2010). Guard cell anion channel SLAC1 is regulated by CDPK protein kinases with distinct Ca2+ affinities. *Proc. Natl. Acad. Sci. U.S.A.* 107, 8023–8028. doi: 10.1073/pnas.0912030107

Geiger, D., Scherer, S., Mumm, P., Stange, A., Marten, I., Bauer, H., et al. (2009). Activity of guard cell anion channel SLAC1 is controlled by drought-stress signaling kinase-phosphatase pair. *Proc. Natl. Acad. Sci. U.S.A.* 106, 21425–21430. doi: 10.1073/pnas.0912021106

Gong, X., Liu, M., Zhang, L., Ruan, Y., Ding, R., Ji, Y., et al. (2015). *Arabidopsis AtSUC2* and AtSUC4, encoding sucrose transporters, are required for abiotic stress tolerance in an ABA-dependent pathway. *Physiol. Plant.* 153, 119–136. doi: 10.1111/ppl.12225

Grondin, A., Rodrigues, O., Verducq, L., Merlot, S., Leonhardt, N., and Maurel, C. (2015). Aquaporins contribute to ABA-triggered stomatal closure through OST1-mediated phosphorylation. *Plant Cell* 27, 1945–1954. doi: 10.1105/tpc.15.00421

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2018.00578/full#supplementary-material
Gudesblat, G. E., Iusem, N. D., and Morris, P. C. (2007). Guard cell-specific inhibition of Arabidopsis MPK3 expression causes abnormal stomatal responses to abscisic acid and hydrogen peroxide. *New Phytol.* 173, 713–721. doi: 10.1111/j.1469-8137.2006.01953.x

Habibi, G. (2012). Exogenous salicylic acid alleviates oxidative damage of barley plants under drought stress. *Acta Biol. Szeged.* 56, 57–63.

Hao, F., Zhao, S., Dong, H., Zhang, H., Sun, L., and Chen, M. (2010). Nia1 and Nia2 are involved in exogenous salicylic acid-induced nitric oxide generation and stomatal closure in *Arabidopsis.* *J. Integr. Plant Biol.* 52, 298–307. doi: 10.1111/j.1744-7909.2010.00920.x

Hayat, S., Hasan, S. A., Fariduddin, Q., Ahmad, A., et al. (2008). Growth of tomato (*Lycopersicon esculentum*) in response to salicylic acid under water stress. *J. Plant Interact.* 3, 297–304. doi: 10.1080/17447909.2008.1055824

Horváth, E., Brunner, S., Bela, K., Papdi, C., and Szabados, L. T. (2015). Exogenous salicylic acid-triggered changes in the glutathione transferases and peroxidases are key factors in the successful salt stress acclimation of *Arabidopsis thaliana.* *Func. Plant Biol.* 42, 1129–1140. doi: 10.1071/FP15119

Hua, D., Wang, C., He, J., Liao, H., Duan, Y., Zhu, Z., et al. (2012). The Ca2+ homeostasis in Arabidopsis*Nia1* and *Nia2* are key factors in the successful salt stress acclimation of *Arabidopsis thaliana.* *Appl. Plant Sci.* 6, 1487–1502. doi: 10.3389/fpls.2015.01213

Jarzyniak, K. M., and Jasiński, M. (2014). Membrane transporters and drought tolerance of wheat (*Triticum aestivum* L.). *Front. Plant Sci.* 5:687. doi: 10.3389/fpls.2014.00687

Kato, M., Nagasaki-Takeuchi, N., Ide, Y., and Maeshima, M. (2010). An *Arabidopsis* hydrophilic Ca2+–binding protein with a PEVK-rich domain, PCaP2, is associated with the plasma membrane and interacts with calmodulin and phosphatidylinositol phosphates. *Plant Cell Physiol.* 51, 366–379. doi: 10.1093/pcp/pcp003

Khan, M. I. R., Iqbal, N., Masood, A., and Khan, N. A. (2012a). Variation in salt tolerance of wheat cultivars: role of glycinebetaine and ethylene. *Pedosphere* 22, 746–754. doi: 10.1016/S1002-0160(12)60060-5

Khan, M. I. R., Iqbal, N., Masood, A., Per, T. S., and Khan, N. A. (2013). Salicylic acid alleviates adverse effects of heat stress on photosynthesis through changes in proline production and ethylene formation. *Plant Signal. Behav.* 8:e26374. doi: 10.4161/psb.26374

Khan, M. I. R., Syeed, S., Nazar, R., and Anjum, N. A. (2012b). “An insight into the role of salicylic acid and jasmonic acid in salt stress tolerance,” in *Phytohormones and Abiotic Stress Tolerance in Plants*, eds N. A. Khan, R. Nazar, N. Iqbal, and N. A. Anjum (Berlin: Springer), 277–300.

Khokon, A. R., Okuma, E., Hossain, M. A., Munemasa, S., and Uraj, M. N. (2011). Involvement of extracellular oxidative burst in salicylic acid-induced stomatal closure in *Arabidopsis.* *Plant Cell Environ.* 34, 434–443. doi: 10.1111/j.1365-3040.2010.02253.x

Klieerekoper, Q. K., and Putkey, J. A. (2009). PEP-19, an intrinsically disordered regulator of calmodulin signaling. *J. Biol. Chem.* 284, 7455–7464. doi: 10.1016/j.jbc.2005.08.005

Kulik, A., Wawer, I., Krzywińska, E., Bucholc, M., and Dobrowska, G. (2011). SnRK2 protein kinases-key regulators of plant response to abiotic stresses. *OMICS* 15, 859–872. doi: 10.1089/omi.2011.0091

Kurkela, S., and Borg-Franck, M. (1992). Structure and expression of kin2, one of two cold- and ABA-induced genes of *Arabidopsis thaliana.* *Plant Mol. Biol.* 19, 689–692. doi: 10.1007/BF00026794

Lee, S. C., Lan, W., Buchanan, B. B., and Luan, S. (2009). A protein kinase-phosphatase pair interacts with an ion channel to regulate ABA signaling in plant guard cells. *Proc. Natl. Acad. Sci. U.S.A.* 106, 21419–21424. doi: 10.1073/pnas.0910811106

Liu, P., Xu, Z. S., Pan-Pan, L., Hu, D., Chen, M., Li, L. C., et al. (2013). A wheat PI4K gene whose product possesses threonine autophosphorylation activity confers tolerance to drought and salt in *Arabidopsis.* *J. Exp. Bot.* 64, 2915–2927. doi: 10.1093/jxb/ert133

Liu, W. X., Zhang, F. C., Zhang, W. Z., Song, L. F., Wu, W. H., and Chen, Y. F. (2013). *Arabidopsis* Di19 functions as a transcription factor and modulates PR1, PR2, and PR5 expression in response to drought stress. *Mol. Plant* 6, 1478–1502. doi: 10.1093/mp/sts031

Luan, S. (2009). The CBL–CIPK network in plant calcium signaling. *Trends Plant Sci.* 14, 37–42. doi: 10.1016/j.tplants.2008.10.005

Matrosova, A., Bogireddi, H., Mateo-Pélas, A., Hashimoto-Sugimoto, M., Iba, K., Schroeder, J. I., et al. (2015). The HT1 protein kinase is essential for red light-induced stomatal opening and genetically interacts with OST1 in red light and CO2-induced stomatal movement responses. *New Phytol.* 208, 1126–1137. doi: 10.1111/nph.13566

Meijer, H. J., and Munnik, T. (2003). Phospholipid-based signaling in plants. *Annu. Rev. Plant Biol.* 54, 265–306. doi: 10.1146/annurev.arplant.54.031902.13474

Miura, K., Okamoto, H., Okuma, E., Shiba, H., Kamada, H., Hasegawa, P. M., et al. (2013). SIZ1 deficiency causes reduced stomatal aperture and enhanced drought tolerance via controlling salicylic acid-induced accumulation of reactive oxygen species in *Arabidopsis.* *Plant Biol.* 17, 93–104. doi: 10.1111/j.1121-0104.2012.12014

Miura, K., and Tada, Y. (2014). Regulation of water, salinity, and cold stress responses by salicylic acid. *Front. Plant Sci.* 5:4. doi: 10.3389/fpls.2014.00004

Munne-Bosch, S., and Penuelas, J. (2003). Photo- and antioxidative protection, and stomatal closure in *Arabidopsis.* *Acta Biol. Szeged.* 56, 298–304. doi: 10.1111/j.1439-0433.2003.01064.x

Nakashima, K., Fujita, Y., Kanamori, N., Katagiri, T., Umezawa, T., Kidokoro, S., et al. (2011). Hydrophilic Ca2+-binding protein with a PEVK-rich domain, PCaP2, is essential for the control of seed development and dormancy. *Plant Cell Physiol.* 50, 1345–1363. doi: 10.1093/pcp/pcp083
Perera, I. Y., Hung, C. Y., Moore, C. D., Stevenson-Paulik, J., and Boss, W. F. (2008). Transgenic Arabidopsis plants expressing the type 1 inositol 5-phosphatase exhibit increased drought tolerance and altered abscisic acid signaling. *Plant Cell* 20, 2876–2893. doi: 10.1105/tpc.108.061374

Pilot, G., Lacombe, B., Gaymard, F., Cherel, I., Bouchereau, J., Thibaud, J. B., et al. (2001). Guard cell inward K+ channel activity in Arabidopsis involves expression of the twin channel subunits KAT1 and KAT2. *J. Biol. Chem.* 276, 3215–3221. doi: 10.1074/jbc.M007303200

Qin, L. X., Li, Y., Li, D. D., Xu, W. L., Zheng, Y., and Li, X. B. (2014). Arabidopsis drought-induced protein Di9-3 participates in plant response to drought and high salinity stresses. *Plant Mol. Biol.* 86, 609–625. doi: 10.1007/s11103-014-0251-4

Raghavendra, A. S., Gonugunta, V. K., Christmann, A., and Grill, E. (2010). ABA perception and signaling. *Trends Plant Sci.* 15, 395–401. doi: 10.1016/j.tplants.2010.04.006

Raskin, I. (1992). Role of salicylic acid in plants. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 43, 439–463. doi: 10.1146/annurev.pp.43.061992.002255

Reyes, J. L., and Chua, N. H. (2007). ABA induction of miR159 controls transcript regulation, and role in abiotic stress tolerance, in *Abiotic Stress Response in Arabidopsis thaliana*. 5:17587. doi: 10.1038/srep17587

Rey, J.-P., S znalazł, M., and Tepfer, J.-F. (2015). Physiological and transcriptional memory in guard cells during repetitive dehydration stress. *New Phytol.* 205, 596–607. doi: 10.1111/nph.13080

Vlad, F., Rubio, S., Rodrigues, A., Sirichandra, C., Belin, C., Robert, N., et al. (2009). Protein phosphatases 2C regulate the activation of the Snf1-related kinase OST1 by abscisic acid in Arabidopsis. *Plant Cell* 21, 3170–3184. doi: 10.1105/tpc.109.096197

Virlouvet, L., and Fromm, M. (2015). Physiological and transcriptional memory in guard cells during repetitive dehydration stress. *New Phytol.* 205, 596–607. doi: 10.1111/nph.13080

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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