The anticancer effects of HDAC inhibitors require the immune system

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Histone deacetylase inhibitors (HDACIs) are known to exert immunomodulatory effects. We have recently demonstrated that the therapeutic efficacy of HDACIs against aggressive B-cell lymphoma and colon carcinoma relies on a functional immune system, in particular on the production of interferon γ (IFNγ). Our findings provide a rationale for the combination of HDACIs with immunotherapeutic agents in the clinic.

Histone deacetylase inhibitors (HDACIs) are currently employed in the clinic for the treatment of a wide panel of solid and hematological malignancies. In particular, vorinostat (Zolind®) and romidepsin (Istodax®) have been approved by the US Food and Drug Administration for the treatment of cutaneous T-cell lymphoma.1,2 HDACIs mediate a range of different antitumor effects, one or more of which may be responsible for their clinical efficacy. The proposed anticancer activities of HDACIs include: (1) the induction of apoptosis and cell cycle arrest; (2) the inhibition of angiogenesis; and (3) the stimulation of cancer cell differentiation. Importantly, HDACIs have been shown to enhance the immunogenicity of cancer cells. Several groups have reported the upregulation of natural killer (NK)-cell activating ligands, MHC class I and II molecules, components of the machinery for antigen presentation, and co-stimulatory molecules on the surface of cancer cells treated with HDACIs.3,5 In addition, the pre-treatment of malignant cells with HDACIs has been employed to generate an effective anticancer vaccine for therapeutic use, and HDACi-treated malignant cells exhibit an increased propensity to be taken by dendritic cells (DCs).3,6 Finally, we have recently demonstrated that colon cancer cells treated with vorinostat undergo immunogenic cell death, as documented by the translocation of calreticulin on the cell surface as well as by the release of ATP and high mobility group box 1 (HMGB1) into the extracellular milieu.7

Despite such clear indicators of the immunostimulatory activity of HDACIs, these agents are also being associated with paradoxical effects. For instance, the administration of HDACIs to wild-type (WT) mice led to the upregulation of forkhead box P3 (FOXP3) and hence to the expansion of regulatory T cells (Tregs), while HDACi-treated DCs exhibited a limited ability to release cytokines in response to lipopolysaccharide.8,9 HDACIs can also limit pathological immune responses in the context of systemic lupus erythematosus (SLE) by decreasing the activity of the host immune system during HDACi-based anticancer therapy in vivo.

We demonstrated that the HDACIs vorinostat and panobinostat are significantly less efficient against aggressive Eμ-Myc-driven B-cell lymphomas and colon carcinomas in mice lacking a functional immune system (Rag2−/−γc−/− mice) than in their WT counterparts. Indeed, tumor-bearing Rag2−/−γc−/− mice succumbed to lymphoma significantly earlier than WT mice, despite ongoing treatment with HDACIs. The decreased survival of tumor-bearing Rag2−/−γc−/− mice treated with a HDACi was not

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due to adverse side effects, the lack of drug-target interactions, or the inability of the HDACi to trigger the apoptotic demise of cancer cells. The non-immunogenic chemotherapeutic agent etoposide was able to enhance the survival of WT and immunocompromised tumor-bearing mice to similar extents. To our knowledge, ours is the first comprehensive demonstration that the therapeutic efficacy of HDACi in vivo relies on an intact immune system. These data are in line with our previous findings demonstrating that the efficacy of HDACi can be significantly enhanced by the concurrent administration of immunostimulatory monoclonal antibodies that operate as CD137 and CD40 agonists.6

By investigating in detail the immunological mechanisms triggered by HDACi in immunocompetent mice, we found that interferon γ (IFNγ) is critical for the therapeutic activity of these agents. Contrarily to our initial hypothesis, we found that IFNγ acts on malignant cells to induce antitumor activity concurrently with HDACi. Moreover, the administration of HDACi was found to sensitize malignant cells to the antineoplastic effects of IFNγ, as signal transduction via the IFNγ receptor 1 (IFNGR1) was increased by HDACi, as were the levels of MHC class I and II molecules expressed on the surface of cancer cells. In order to confirm our findings, we overexpressed a non-functional dominant negative variant of the IFNGR1 in Eμ-Myc-driven lymphoma cells, and found that mice bearing these genetically engineered malignant cells succumbed to lymphoma significantly earlier than those bearing WT tumors, in spite of vorinostat treatment. Finally, we demonstrated that the co-administration of a potent immune adjuvant and IFNγ inducer, namely α-galactosylceramide (α-GalCer), with vorinostat significantly prolonged the survival of tumor-bearing mice as compared with either agent alone. Our findings indicate that the combination of HDACi with immunotherapy is a promising strategy for the treatment of cancer.

We next sought to determine the origin of IFNγ secreted in the course of treatment with vorinostat. Surprisingly we found that NK cells, CD8+ T cells and CD4+ T cells are not required for the therapeutic efficacy of vorinostat. Conversely, we demonstrated a critical role for B cells in the anticancer effects of HDACi. Moreover, B cells isolated from vorinostat-treated, tumor-bearing mice were found to produce IFNγ. While the role of B cells in oncogenesis and tumor progression is controversial and still under investigation, the localization of Eμ-Myc-driven lymphoma cells within the B-cell niche of lymphoid organs strongly suggest that HDACi may induce an antitumor B-cell immune response (Fig. 1).

In conclusion, we have recently shown that the immune system is a critical component of the antitumor effects of HDACi. These findings confirm previous in vitro data demonstrating that HDACi increase the immunogenicity of cancer cells. Our study will provide additional impetus to combine HDACi with immunotherapeutic agents, including immune adjuvants such as α-GalCer and immunostimulatory monoclonal antibodies, in the clinic.

Disclosure of Potential Conflicts of Interest

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