How relevant is panallergen sensitization in the development of allergies?

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Abstract

Panallergens comprise various protein families of plant as well as animal origin and are responsible for wide IgE cross-reactivity between related and unrelated allergenic sources. Such cross-reactivities include reactions between various pollen sources, pollen and plant-derived foods as well as invertebrate-derived inhalants and foodstuff. Here, we provide an overview on the most clinically relevant panallergens from plants (profilins, polcalcins, non-specific lipid transfer proteins, pathogenesis-related protein family 10 members) and on the prominent animal-derived panallergen family, tropomyosins. In addition, we explore the role of panallergens in the sensitization process and progress of the allergic disease. Emphasis is given on epidemiological aspects of panallergen sensitization and clinical manifestations. Finally, the issues related to diagnosis and therapy of patients sensitized to panallergens are outlined, and the use of panallergens as predictors for cross-reactive allergy and as biomarkers for disease severity is discussed.

Keywords

allergy diagnosis; allergy treatment; IgE cross-reactivity; non-specific lipid transfer proteins; pathogenesis-related protein family 10; polcalcins; profilins; tropomyosin

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Allergens are harmless common environmental substances capable of inducing IgE-mediated (type-I) hypersensitivities in atopic individuals (1). Allergens have been identified in a variety of sources including over 80 plant species and over 100 animal species (2). In most cases, an allergenic source contains more than one allergen deriving from different protein families, which in turn exhibit differential sensitization rates (3). Many allergens have been implicated in cross-reactivities, where a patient sensitized to a specific source is also reacting to other related and/or non-related sources. The foundation of such cross-recongitions is the presence of conserved linear or conformational IgE epitopes among members of the same protein family (4). Panallergens are proteins that take part in key processes of organisms and are therefore ubiquitously distributed with highly conserved sequences and structures (2). Thus, panallergens are implicated in a multitude of cross-reactions, even between phylogenetically distant and unrelated organisms (2). Such cross-reactivities include reactions between various pollen sources (5, 6), pollen and plant foods (7) as well as invertebrate inhalants and foods (8, 9). So far, panallergens have been classified into a few protein families with different levels of distribution and cross-reactivity. In this review, we provide an overview on the most clinically relevant panallergens from plants (profilins, polcalcins, non-specific lipid transfer proteins, pathogenesis-related protein family 10 members) and on the prominent animal-derived panallergen family, tropomyosins. Until now, 44 profilins, 15 polcalcins, 45 nsLTPs, 24 PR-10s and 29 tropomyosins have been identified and officially acknowledged by the WHO/IUIS allergen nomenclature subcommittee (www.allergen.org) (Tables 1 and 2).

This review also undertakes the task of clarifying the role of panallergens in the sensitization process and the progress of the allergic disease. Whilst controversially discussed, panallergen sensitizations have been shown to provide further complexity to the allergic profile of patients (10). If developed, panallergen...
Table 1 List of plant panallergens profilins, polcalcins, nsLTPs and PR-10s acknowledged by the IUIS nomenclature subcommittee

| Species                  | Common name             | Scientific name | Common name | Protein family |
|--------------------------|-------------------------|-----------------|-------------|----------------|
| *Acacia farnesiana*      | Needle bush             | *Aca f 2*       |             |                |
| *Actinidia chinensis*    | Gold Kiwi fruit         | *Act c 10*      |             | Act c 8        |
| *Actinidia delicosa*     | Kiwi fruit              | *Act d 9*       |             | Act d 10       |
| *Ailus glutinosa*        | Alder                   | *Aln g 4*       |             |                |
| *Amaranthus retroflexus* | Redroot pigweed         | *Ara r 2*       |             |                |
| *Ambrosia artemisiifolia*| Short ragweed           | *Amb a 8*       |             | Amb a 9        |
| *Ananas comosus*         | Pineapple               | *Ana c 1*       |             |                |
| *Apium graveolens*       | Celery                  | *Api g 4*       |             | Api g 1        |
| *Arachis hypogaea*       | Peanut                  | *Ara h 5*       |             |                |
| *Artemisia vulgaris*      | Mugwort                 | *Art v 4*       |             | Art v 3        |
| *Asparagus officinalis*  | Asparagus               | *Asp a 1*       |             |                |
| *Beta vulgaris*          | Sugar beet              | *Bet v 2*       |             |                |
| *Betula verrucosa* (pendula) | European white birch | *Bet v 2*       |             | Bet v 3        |
| *Brassica oleracea*      | Cabbage                 | *Bra o 3*       |             |                |
| *Brassica rapa*          | Turnip                  | *Bra r 5*       |             |                |
| *Cannabis sativa*        | Indian hemp             | *Can s 3*       |             |                |
| *Capsicum annuum*        | Bell pepper             | *Cap a 2*       |             |                |
| *Carpinus betulus*       | Hornbeam                | *Car b 1*       |             |                |
| *Castanea sativa*        | Chestnut                | *Cas s 4*       |             | Cas s 1        |
| *Chenopodium album*      | Lambquaters             | *Che a 2*       |             |                |
| *Citrus limon*           | Lemon                   | *Cit l 3*       |             |                |
| *Citrus reticulata*      | Tangerine               | *Cit r 3*       |             |                |
| *Citrus sinensis*        | Sweet orange            | *Cit s 2*       |             | Cit s 3        |
| *Corylus avellana*       | Hazel                   | *Cor a 2*       |             |                |
| *Crocos sativus*         | Saffron crocus          | *Cro s 2*       |             |                |
| *Cucumis melo*           | Muskmelon               | *Cuc m 2*       |             |                |
| *Cynodon dactylon*       | Bermuda grass           | *Cyn d 12*      |             |                |
| *Daucus carota*          | Carrot                  | *Dau c 1*       |             |                |
| *Fagus sylvatica*        | European beech          | *Fag s 1*       |             |                |
| *Fragaria ananassa*      | Strawberry              | *Fra a 4*       |             | Fra a 3        |
| *Glycine max*            | Soybean                 | *Gly m 3*       |             | Gly m 4        |
| *Helenium annuus*        | Sunflower               | *Hel a 2*       |             | Hel a 3        |
| *Hevea brasiliensis*     | Para rubber tree (latex)| *Hev b 8*       |             | Hev b 12       |
| *Hordeum vulgare*        | Barley                  | *Hor v 12*      |             |                |
| *Juglans regia*          | English walnut          | *Jun o 4*       |             |                |
| *Juniperus oxycedrus*    | Prickly juniper         | *Jun o 4*       |             |                |
| *Kochia scoparia*         | Burning bush            | *Koc s 2*       |             |                |
| *Lactuca sativa*         | Cultivated lettuce     | *Lac s 1*       |             |                |
| *Lens culinaris*         | Lentil                  | *Len c 3*       |             |                |
| *Litchi chinensis*       | Litchi                  | *Lit c 1*       |             |                |
| *Malus domestica*        | Apple                   | *Mal d 4*       |             | Mal d 3        |
| *Mercurialis annua*      | Annual mercury          | *Mer a 1*       |             |                |
| *Morus nigra*            | Mulberry                | *Mor n 3*       |             |                |
| *Musa acuminata*         | Banana                  | *Mus a 1*       |             | Mus a 3        |
| *Olea europea*           | Olive                   | *Ole e 2*       |             | Ole e 3        |
| *Oryza sativa*           | Rice                    | *Ory s 12*      |             |                |
| *Ostrya carpinifolia*    | European Hophornbeam    | *Ost c 1*       |             |                |
| *Parietaria judaica*     | Pellitory-of-the-Wall   | *Par j 3*       |             |                |

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sensitization further complicates the allergic disease notably due to its most defining feature, cross-reactivity. Panallergen cross-reactivities can result in a wider range of IgE responses to distinct allergen sources, hence creating false positives in diagnosis and further complicating treatment (11). Although just a minority of individuals become sensitized and go onto develop allergy towards panallergens, this phenomenon appears to be highly influenced by many variables including allergen source exposure, geographic differences (12), the type of allergen source and age demography (13). Such variations are reflected in the disparity of results obtained within the literature and highlight the need for understanding panallergens and their sensitization profiles in the development of future diagnostics and treatments of allergy.

**Profilins**

Profilins are 12–16 kDa, actin-(monomer)-binding proteins, expressed in all eukaryotic cells and certain viruses, with the exception of some protists (14, 15). Profilins promote polymerization of actin filaments and monomers and are thus involved in the generation of the cytoskeleton and movement (15). A variety of 50 additional identified profilin ligands, like phosphoinositides or proteins containing proline rich domains (also used for purification), suggest an important role in many more complex molecular processes as well as signal transduction (16, 17). They belong to the α-β class of proteins with a highly conserved structure consisting of a central antiparallel β-sheet core surrounded by α-helices (18). The first allergenic profilin, Bet v 2 from birch pollen, was identified in 1991 (19), and since then, many allergenic members have been identified in pollen, plant foods and latex (www.allergen.org). The involvement of profilins in such essential cellular processes explains their ubiquitous expression and high levels of conservation. This, in turn, can justify the strong serologic cross-reactivity of the molecules (20).

**Polcalcins**

Polcalcins are a pollen-restricted portion of widespread Ca²⁺-binding proteins comprising a major group of allergenic molecules of this kind. The mostly α-helical proteins contain the characteristic EF-hand motifs (helix-loop-helix structure), which are responsible for the binding of calcium. Polcalcins

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**Table 1.** (Continued).

| Scientific name     | Common name   | Protein family |
|---------------------|---------------|----------------|
| *Phaseolus vulgaris*| Green bean    | Profilin       |
| *Phleum pratense*   | Timothy       | Polcalcin      |
| *Phoenix dactylifera*| Date palm     | nsLTP          |
| *Platanus acerifolia*| London plane tree | PR-10         |
| *Platanus orientalis*| Oriental plane |               |
| *Prosopis juliflora*| Mesquite      |               |
| *Prunus armeniaca*  | Apricot       |               |
| *Prunus avium*      | Sweet cherry  |               |
| *Prunus domestica*  | European plum|               |
| *Prunus dulcis*     | Almond        |               |
| *Prunus persica*    | Peach         |               |
| *Punica granatum*   | Pomegranate   |               |
| *Pyrus communis*    | Pear          |               |
| *Quercus alba*      | White oak     |               |
| *Rubus idaeus*      | Red raspberry |               |
| *Salsola kali*      | Russian thistle|              |
| *Sinapis alba*      | Yellow mustard|               |
| *Solanum lycopersicum*| Tomato        |               |
| *Syringa vulgaris*  | Lilac         |               |
| *Triticum aestivum* | Wheat         |               |
| *Vigna radiata*     | Mung bean     |               |
| *Vitis vinifera*    | Grape         |               |
| *Zea mays*          | Maize         |               |

*Hazel trees contain Cor a 1 as a food allergen in the nut and as an inhalant allergen in the pollen.
have a molecular mass of 9–28 kDa, depending on the number of EF-hand motifs they hold (2, 3 or 4 EF-hand motifs), and show a monomeric or dimeric structure. They can assume two different conformations; a closed, Ca²⁺-free apo-form and an open, Ca²⁺-bound holo form which is more stable and results in stronger interactions with IgE antibodies (23). Although the exact physiologic function of polcalcins remains elusive, their localization in the pollen and their ability to regulate intracellular Ca²⁺ levels suggest a contribution in pollen tube outgrowth (24). Allergenic polcalcins derive from tree, grass and weed pollen; are highly cross-reactive; and induce sensitization rates between 5% and 10% among pollen-allergic patients (2). It seems that the clinical relevance of polcalcin sensitization is highly dependent on geographic factors and therefore exposure. Phl p 7 from timothy grass pollen is, among all allergenic polcalcins, the most cross-reactive molecule and could hence be used as a marker to identify multiple pollen sensitizations (25).

Non-specific lipid transfer proteins (nsLTPs)

Originally named after their ability to transfer lipid molecules between membranes in vitro, which seems unlikely in vivo, lipid transfer proteins (LTPs) are widely distributed small and basic proteins in higher plants (26). LTPs can be either lipid specific or can non-specifically accommodate several classes of lipids (nsLTPs); all allergenic LTP members have been found within the nsLTP cluster. According to their size, nsLTPs can be further subdivided into the 9 kDa nsLTP1 and the 7 kDa nsLTP2 subfamilies (27, 28). Their physiologic function is still unknown; however, accumulating evidence shows that they play a role in cytology, growth and development. Furthermore, the implication of nsLTPs in the general defence of plants classifies them as members of the pathogenesis-related protein family (PR-14 family) (26, 29). nsLTPs share a common conserved tertiary structure consisting of four α-helices stabilized by 4 disulphide bonds (eight-cysteine motif), forming a hydrophobic tunnel-like cavity, where interactions with lipids occur (30, 31). Allergenic nsLTPs show a wide distribution in pollen of trees and weeds, in foods (fruits, nuts, seeds and vegetables) as well as in latex (2). Due to their high thermal and proteolytic stability, nsLTPs are potent class I food allergens. Nevertheless, it is still unclear whether pollen or food nsLTPs are the primary sensitizers (32). nsLTPs are clinically highly relevant and cross-reactive allergens. In Mediterranean areas, nsLTP representatives like Pru p 3 from peach constitute major allergens, whilst in Central and Northern areas, sensitization to Pru p 3 is limited (33).

Table 2

| Species          | Scientific name       | Common name        | Protein family | Tropomyosin |
|------------------|-----------------------|-------------------|---------------|-------------|
| Arachnids        | Blomia tropicalis     | Mite              | Blo t 10      |
|                  | Chortoglyphus arcuatus| Storage mite      | Cho a 10      |
|                  | Dermatophagoides farinae| American house dust mite | Der f 10 |
|                  | Dermatophagoides pteronyssinus| European house dust mite | Der p 10 |
|                  | Lepidoglyphus destructor| Storage mite      | Lep d 10      |
|                  | Tyrophagus putrescentiae| Storage mite      | Tyr p 10      |
| Insects          | Aedes aegypti         | Yellow fever mosquito | Aed a 10 |
|                  | Blattella germanica   | German cockroach  | Bia g 7       |
|                  | Chironomus kiensis    | Midge             | Chi k 10      |
|                  | Lepisma saccharina    | Silverfish        | Lep s 1       |
|                  | Periplaneta americana | American cockroach| Per a 7       |
| Mollusks         | Helix aspersa         | Brown garden snail| Hel as 1      |
|                  | Todarodes pacificus   | Squid             | Tod p 1       |
| Parasites        | Anisakis simplex      | Nematode          | Ani s 3       |
|                  | Ascaris lumbricoides  | Common roundworm  | Asc l 3       |
| Seafood          | Charybdis ferratus    | Crab              | Cha f 1       |
|                  | Crangon crangon       | North Sea shrimp  | Cra c 1       |
|                  | Homarus americanus    | American lobster   | Hom a 1       |
|                  | Litopenaeus vannamei  | White shrimp      | Lit v 1       |
|                  | Macrobrachium rosenbergii | Giant freshwater prawn | Mac r 1 |
|                  | Melicertus latisculatus| King prawn        | Mel l 1       |
|                  | Metapenaeus ensis     | Shrimp            | Met e 1       |
|                  | Oreochromis mossambicus| Mozambique tilapia| Ore m 4       |
|                  | Pandanus borealis     | Northern shrimp   | Pan b 1       |
|                  | Panulirus stimpsoni   | Spiny lobster     | Pan s 1       |
|                  | Penaeus aztecus       | Shrimp            | Pen a 1       |
|                  | Penaeus indicus       | Shrimp            | Pen i 1       |
|                  | Penaeus monodon       | Black tiger shrimp| Pen m 1       |
|                  | Portunus pelagicus    | Blue swimmer crab | Por p 1       |

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Pathogenesis-related protein family 10 (PR-10)

PR-10 proteins function in the general defence mechanisms of plants, and their expression is induced by stress and pathogens. They constitute mostly intracellular, intrinsic and highly similar globular proteins with a length of around 160 amino acids. They share a molecular mass of around 17 kDa and are encoded by a diverse multigene family (34). Their 3D fold consists of 3 α-helices embedded in an antiparallel β-sheet consisting of 7 β-strands forming an amphiphilic intrinsic solvent-accessible y-shaped cavity. Crystallographic analyses of Bet v 1 in combination with a variety of different ligands revealed the availability of a promiscuous binding site in the pocket, which could help to unravel the biologic function of PR-10s (35, 36). Recently, the physiologic ligand quercetin-3-O-sophoroside in birch pollen for Bet v 1 has been identified (37). A subclass of PR-10 proteins forms a group of pollen and food allergens. The botanically related Fagales species of birch, alder, hornbeam, hop-hornbeam, hazelnut, beech, chestnut and oak are the main elicitors of early seasonal rhinitis in the temperate climate zone of the Northern Hemisphere. Whilst Bet v 1 is known to act as the main sensitizer, a potential (co-)sensitization of other Fagales members is hypothesized (38). A high percentage of Fagales pollen-allergic patients develop oral reactions against a variety of fresh fruits, nuts and vegetables. Such clinical reactions, known as pollen-food syndrome, are triggered by IgE antibodies, which are produced against aeroallergens and cross-react with food allergens (7).

Tropomyosin

Tropomyosins are α-helical and, in their native form, dimeric coiled-coil fibrous structural proteins consisting of two subunits (α, β), each with a molecular weight ranging from 34 to 38 kDa. They form co-polymers with actin and regulate actin–myosin interactions in skeletal muscles and diversify the functions of actin filaments in different intracellular compartments (39). Evolutionarily, tropomyosins were introduced in metazoa and fungi to expand the functional capacity of actin filaments without enlarging the number of actin isoforms and are therefore not present in plants. In vertebrates, four highly conserved tropomyosin genes are expressed. The increasing number of tropomyosins and splicing variants parallels the increasing complexity of the animal kingdom (15). Tropomyosins are considered important food allergens, whereas tropomyosins from vertebrates are typically non-allergenic, with the exception of the fresh water fish tilapia (9). Crustacean shellfish form one group of ‘the big-8’ major food allergens (www.fda.gov). The highly heat-stable tropomyosins comprise the major allergens in crustaceans and mollusks, making them significant food allergens in exposed populations (40). In addition, these patients show frequent reactions towards (house dust) mites and insects, especially in warmer climates where the growth of house dust mites is favoured. Several studies have already established cross-reactivity between seafood, mollusks, insects and some parasites (41, 42). However, the route of sensitization remains a matter of debate. There is a controversy as to whether primary sensitization is happening via the gastrointestinal or the respiratory tract. Nonetheless, there are clear indications that inhalation of aeroallergens or cooking vapours is an important sensitising route in the context of tropomyosin hypersensitivity (41, 43). A schematic diagram describing potential exposure routes and sensitization to tropomyosins is presented in Fig. 1.

Sensitization patterns

Panallergen sensitizations tend to occur as part of a co- or poly-sensitization profile, with panallergen monosensitizations rarely arising (44, 45). In particular, panallergen sensitizations are commonly associated with high or prolonged exposure to a given allergen source, often correlating with increased severity of disease (10, 12). The clinical relevance of panallergens is highly influenced by a number of factors including differences in geographic region (12), allergenic source, the host response against the allergen and exposure (13). Furthermore, differences between child and adult populations have exhibited great variability (12, 46). Hence, studying sensitization profiles of a given source can be a useful tool in understanding the clinical relevance of panallergens. Hatzler et al. exemplifies such a predictable pattern of sensitization for the allergen source *Pileum pratense* (timothy grass). Initial sensitizations towards major (Phl p 1 and Phl p 5) and minor (Phl p 4, 2, 6, 11) allergens gradually result in the onset of allergic disease, following such disease onset the occurrence of panallergen sensitizations, towards Phl p 12 and 7, increases for a minority of patients in correlation with disease severity. Hence, whilst not being an indicative marker for allergic disease onset, panallergen-specific IgE responses have the potential as clinical biomarkers for increased severity of disease, although it must be emphasized that only a minority of patients become sensitized (10). Such rates of sensitization are influenced by the level of exposure to an allergen source. Feliu et al. demonstrated that children, even with a short disease history, were able to become panallergen sensitized to both date palm profilin (Pho d 2) and peach nsLTP (Pru p 3), with 12% and 13% incidence of IgE positivities, respectively (47). Moreover, high olive pollen rates in southern Spain have been reported to drive increases in sensitizations to the olive nsLTP Ole e 7 (12), further showing high exposure rates are strongly correlated with increases in panallergen IgE-positive patients.

Furthermore, such increased rates of panallergen sensitization have been shown to correlate with an increased severity of allergic symptoms (and in the absence of panallergen allergy). A study carried out by Alverado et al. investigating profilin-related allergic reactions over both an intense and a mild grass pollen season showed that more severe profilin allergy occurred during the intense season again emphasising the relationship between higher allergen exposures and increased panallergen sensitization rates (44). This phenomenon has been further demonstrated in a large study of 891 allergic patients from Spain where sensitizations to grass pollen profilin correlated with the severity of the allergic disease (12). Conversely, recent studies have emerged describing sensitizations to multiple panallergen families and their effects in reducing the severity of allergic reactions. In particular,
such co-sensitizations have been investigated in peach allergy due to the multiple panallergens present within the fruit, namely Pru p 1 (PR-10), Pru p 3 (nsLTP) and Pru p 4 (profilin). Patients sensitized to nsLTPs along with PR-10/profilins present a lower severity of symptoms when compared to patients with sensitizations to nsLTP alone (3). Considering that many allergenic sources contain multiple panallergen families, exploring this avenue and the theories behind such interactions may be of key benefit for developing therapeutic strategies (3, 46). Further investigations into allergies of different plant species have revealed that certain pollen exhibits extraordinarily high rates of panallergen sensitization (48, 49). For Chenopodium album pollen (white goosefoot), sensitization rates of 55% and 46% to profilin and polcalcin, respectively, have been reported (48). A further study by Nouri et al., carried out in an Iranian cohort, showed that 81% of patients tested positive to the profilin panallergen Che a 2 (49). Disparity of panallergen sensitizations is also exhibited between child and adult populations. Interestingly, Barber et al. also show sensitizations towards the peach nsLTP Pru p 3 to be more prevalent within children than in adult populations in areas of high pollen sensitization in Spain (12). Furthermore, in a Mediterranean study observing nsLTP sensitizations, it was shown that children below the age of six were more frequently sensitized by Pru p 3. However, for the adult population, sensitizations to the walnut nsLTP Jug r 3 reached comparable levels, suggesting an alternative source of sensitization within this age group (46). Such epidemiological differences must be considered when performing clinical investigations, and understanding such profiles of panallergen sensitizations may be of high clinical benefit in both diagnosis and treatment of allergy.

**Panallergen allergy**

For only a minority of patients sensitized to panallergens, allergy arises (50). It is in such cases that the cross-reactivity of panallergens plays a role in worsening the allergic profile of patients via increasing the amount of potential allergic reactions to allergens in unrelated sources (51). Tropomyosins (43), profilins, PR-10s and nsLTPs are commonly found in food and plant sources (Tables 1 and 2) and are strongly associated to food allergy arising as a consequence of cross-reactivity to inhaled allergens (51, 52). Pollen-food allergy syndrome is occurring commonly for profilins, PR-10s and nsLTPs (51), especially when exposure rates are increased (12). Associated symptoms range from oral allergy to anaphylaxis and high co-occurrence with other atopic diseases including allergic rhinitis and asthma, further emphasising the clinical relevance of panallergens (51, 53–55). Identification of such cross-reactivities may be vital in finding biomarkers for allergy. For example, the discovery of cross-reactions between the plane pollen nsLTP Pla a 3 and the peach nsLTP Pru p 3 has elucidated the potential use of Pla a 3 as a biomarker in identifying plane pollen-allergic patients at risk of developing plant food-related nsLTP allergies, thereby improving diagnosis and treatment (56). Such strategies may be helpful in avoiding unnecessary dietary restrictions associated to food allergy (57).

It is of note that these different panallergen families present varying levels of severity with regard to the symptoms induced. Tropomyosin represents the only panallergen family with the potential to induce autoimmune responses (58); however, all of the families elicit their effect through the allergic response. Most severely, cases of anaphylaxis have been documented for tropomyosins, nsLTPs (59) and PR-10s (51, 60). Polcalcins and
Diagnosis and treatment
Panallergen sensitizations may be indicative of allergic disease severity and key in identifying a specific allergenic source, especially when sensitization rates are high (67). Currently, a main treatment of such panallergen allergy is avoidance of the allergy causing foods. This course of action may present detrimental side effects to the patient, with regular intake of fresh fruit having associations with increased health and reduced risk in the development of several chronic diseases (52). For the interests of patients' quality of life and nutritional satisfaction, it is of benefit to understand the demographics of panallergen sensitization in a specific detail, especially in order to be able to exactly determine the specific allergenic cause. Moreover, with evidence that panallergen sensitizations are linked to increased severity of symptoms, it is crucial to efficiently identify those patients with a higher risk of a severe reaction.

In vitro molecular-based diagnosis (MBD) uses allergenic proteins, either natural, purified or recombinantly produced, to quantify the amount of allergen-specific IgE antibodies in the blood circulation (59). The use of microarray systems, such as the Immuno-Sorbent Allergen Chip (ISAC) used for detecting and quantifying the reactivity of specific IgE antibodies towards more than 100 allergens and allergen components, is helpful in the identification and interpretation of allergic sensitization profiles (68). However, it must be noted that IgE positivity alone is not entirely indicative of allergy, and further information obtained by oral/respiratory provocation challenges or basophil activation tests is key in obtaining a true allergic diagnosis (47, 57, 69). Successful diagnosis is essential for deciphering treatment; allergen immunotherapy (AIT) currently represents one of the few ways to modulate allergic disease, and success is highly variable (70). Whilst there is potential in moderating panallergen sensitizations, in particular for food-related allergy, current commercial availability of therapeutic extracts is limited for minor allergens/panallergens (57). Moreover, lack of standardization provides further difficulty in the use of commercial extracts (11, 71).

Exploration of the hierarchical patterns of cross-reactive molecules may also provide further insight into panallergen sensitizations and potentially elucidate novel therapeutics based on the concept that some molecules are more cross-reactive than others. A recent microarray study by Pfiffner et al. investigates the structural basis of cross-reactivity using an iterative motif detection algorithm (72). Identifying such hierarchies raises the question as to whether treatments using AIT should constitute of more frequently cross-reacting allergens or an allergen that less frequently cross-reacts with other allergens, but targets a larger repertoire of allergens within the hierarchy. Further exploration into individual panallergen sensitization profiles (e.g. nsLTPs) (46) may provide key insight into the mechanisms of panallergens in allergy and reveal predictable patterns of cross-reactivities, thus improving diagnosis. Furthermore, using panallergens as biomarkers for disease severity and as predictors of cross-reactive allergens may pose the most clinically relevant use of panallergens in allergic disease (47, 69).

Summary and conclusions
In daily clinical practice, it is often observed that individuals allergic to fruits, nuts and vegetables have sensitivities to more than one food, not necessarily belonging to the same family. In addition, many of them are also allergic to pollens and/or seafood. These associations occur when the same IgE antibody is able to recognize similar epitopes in different panallergens and appear to be highly influenced by many variables. This phenomenon explains why some patients develop a severe allergic reaction upon ingesting allergic foods they have never before encountered (73). Cross-reactivity can also occur due to conformational diversity of antibodies (74) and through T-cell epitopes (75), which in turn may lead to disease. Multiple reactivity to inhalant or allergenic food sources appeared to be caused by the sensitization to at least one representative molecule from a single panallergen group, emphasising the importance of panallergens in multiple sensitizations (Fig. 1).

In conclusion, panallergens comprise a variety of protein families of plant and animal origin and are responsible for wide IgE cross-reactivity between different allergenic sources. Panallergen sensitizations are clinical ‘warning signs’ for elevated risk of multiple allergies as well as severe symptoms. Thus, improving the understanding of this immunologic cross-reactivity is essential for the interpretation of their clinical relevance, the development of improved diagnostic tests and the generation of more efficacious immunotherapy agents. Future studies focusing on the identification and evaluation of cross-reactive epitopes between different members of panallergenic families might provide the basis for understanding their role in sensitization and allergic reactions.

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