Corticospinal excitability for hand muscles during motor imagery of foot changes with imagined force level

Kouki Kato*, Kazuyuki Kanosue
Faculty of Sport Sciences, Waseda University, Tokorozawa, Japan
* kato.k@aoni.waseda.jp

Abstract

The object of this study was to clarify whether corticospinal excitability controlling hand muscles changes concurrently with increases in the imagined contraction level of foot dorsiflexion. Twelve participants performed actual and imagined dorsiflexion of their right foot at three different EMG levels (10, 40 or 80% of the maximum voluntary contraction). During isometric actual- or imagined- dorsiflexion, transcranial magnetic stimulation (TMS) was delivered to the right hand area of the left primary motor cortex. Motor evoked potentials (MEPs) were recorded from the right extensor carpi radialis (ECR) and flexor carpi radialis (FCR). During actual contraction, MEP amplitudes of ECR and FCR increased with an increased EMG level of dorsiflexion. Similarly, during imagery contraction, MEP amplitudes of ECR and FCR increased with the intensity of imagery contraction. Furthermore, a correlation between MEP amplitude during actual contraction and imagery contraction was observed for both ECR and FCR. Motor imagery of foot contraction induced an enhancement of corticospinal excitability for hand muscles that was dependent on the imagined contraction levels, just as what was observed when there was an actual contraction.

Introduction

Motor imagery is the mental representation of an action without any overt movement or muscle activation [1]. Practice involving motor imagery is widely used in both sports and rehabilitation, and many studies have demonstrated beneficial effects for improving motor skills as well as rehabilitating neurological impairments [2,3,4,5,6,7,8,9]. During motor imagery, brain regions such as the primary motor cortex (M1), premotor cortex, supplemental motor area, cerebellum, and basal ganglia are activated in a way similar to that which occurs during actual task execution [10,11,12,13,14]. Furthermore, many studies have demonstrated that motor imagery of muscle contraction induces an enhancement of corticospinal excitability, as assessed by a comparison with motor evoked potentials (MEPs) evoked by transcranial magnetic stimulation (TMS) for the target muscle itself [11,15,16,17,18]. A recent study has shown that, during motor imagery, the corticospinal excitability associated with a particular muscle changed depending upon the force level of the imagined contraction [14]. Many approaches have confirmed a relationship between brain activity and force level of the actual muscle contraction [19,20,21,22]. Studies utilizing single-cell recording in monkeys...
indicate that firing rates of motor-related cortical neurons, such as those in the contralateral M1 and the premotor cortex, increase with an increase in the force level of contraction [19,20,21]. Neuroimaging studies utilizing functional magnetic resonance imaging (fMRI) in humans have also demonstrated that the level of activation of motor-related cortical regions is dependent upon the force level of muscle contraction [22,23]. Corticospinal excitability has also been shown to increase concurrently with changes in magnitude of the force level [24].

In the motions involved with daily life as well as those used in sports, simultaneous use of multiple limbs (e.g. both hands or a hand and a foot) is often required. It is well known that movement of one limb influences that of the others. This remote effect has been well studied [25,26,27]. Investigations using TMS demonstrate that simple muscle contraction enhances the corticospinal excitability of brain regions which control muscles in remote segments. This provides a neural basis for the remote effect [28,29,30,31,32,33,34,35]. The remote effect on corticospinal excitability is augmented by an increase in the contraction level [36]. Interestingly, “motor imagery” of muscle contraction also induces an increase in the corticospinal excitability of remote muscles [37]. Then, it could be hypothesized that corticospinal excitability is influenced by changes in the imagined force level of remote muscles. The aim of the present study was to test this hypothesis. We utilized TMS to elicit MEPs of the right hand muscles during imagery of contraction of the right foot muscle at different contraction intensity levels.

**Method**

**Participants**

Twelve right-handed, healthy volunteers (nine men and three women; aged 21–26 years) without known neurological or psychiatric disease participated in the experiment. All participants gave written, informed consent. The experimental procedure was approved by the Human Research Ethics Committee of Waseda University and performed according to the Declaration of Helsinki.

**Recording**

Surface EMGs were recorded from the right extensor carpi radialis (ECR), flexor carpi radialis (FCR) and tibialis anterior (TA) via disposable Ag-AgCl electrodes (1 cm diameter) which were placed over the belly of the muscle. Before the electrodes were attached, the involved area of skin was shaved and treated with alcohol to reduce inter-electrode impedance. Inter-electrode impedance and EMG signals for the three muscles were checked after placing the electrodes. The EMG signal was amplified (MEB-2216, Nihonkoden, Japan) and bandpass filtered between 5 and 1500 Hz. All signals were converted into digital data via an A/D converter system which sampled at a rate of 4000 Hz.

**Procedure**

The participants sat in a comfortable chair with their right forearm supported on an armrest in a horizontal position. The hand remained in a pronated position throughout the experiment. During the recordings, the participants were instructed not to activate muscles in their hands or in their left foot. The experimenter confirmed that the right foot of the participants did not touch the ground during task execution. At the beginning of the experiment, the participants were asked to perform a maximal voluntary contraction (MVC) for right foot dorsiflexion three times. Each time, they were told, and verbally encouraged during the contraction, to develop a force as hard as possible for 3s. Next, a visual line indicating the target EMG level (10, 40 or 80% of the MVC) that the participant would be asked to exert was displayed on a PC
monitor, which was positioned about 1 m from the participant. The subjects practiced isometric dorsiflexion by matching the rectified and smoothed target EMG level for each MVC (10, 40 or 80%). This task was termed “actual dorsiflexion” (Fig 1 left). Then, the participants practiced imagining that they were maintaining dorsiflexion in their right foot at the same intensities that were performed in the actual dorsiflexions (10, 40 and 80% MVC). This task was termed “imagery dorsiflexion” (Fig 1 right). For the motor imagery, participants were asked to use a first person perspective [10], and we confirmed that there was no EMG activity. In order to evaluate the subjects’ motor imagery ability, each participant took the Vividness of Motor Imagery Questionnaire (VMIQ) [38]. With the VMIQ, a participant rates the vividness of motor imagery on a 5-point Likert scale (1 = vivid imagery, 5 = no imagery at all). We administered this questionnaire at the end of the experiment. If the experimenter noticed unexpected EMG activity in the TA during the imagery dorsiflexion task, the practice session was extended until such EMG activity disappeared. Before the experiment, the participants performed an exertion at each EMG level (pre) without viewing the PC monitor. During the actual experiment, the participants also performed each force level exertion without viewing the PC monitor. During each break period, the participants confirmed that they were performing the exertion at the correct EMG level by viewing the PC monitor. After the trials were completed, the EMGs at the three target levels and the MVC were again recorded again (post experiment recordings) without viewing the PC monitor as a verification test. TMS was delivered during actual isometric dorsiflexion or imagery isometric dorsiflexion. The maintenance period for actual and imagery dorsiflexion before the stimulation was varied randomly from 2 to 5 sec. In addition to being delivered during the “actual dorsiflexion” and “imagery dorsiflexion” tasks, TMS was delivered 15 times when there was no motor action or imagery as a resting control task. These three tasks were randomly mixed. There were 105 trials overall. To avoid fatigue,
the inter-trial interval was always more than 20 sec. The participants took a break after every 20 tasks. The entire experiment lasted approximately 2 h.

**TMS**

TMS was applied to the left M1 using a magnetic stimulator (Magstim 200, Magstim Ltd, UK) connected to a figure-eight coil (95 mm diameter) [39,40,41]. The participants wore a tight fitting swimming cap on which the position for stimulation was marked by a color marker. The coil was held by hand, and its position with respect to the marks carefully monitored. The TMS pulse was delivered to the M1 site at the best location for eliciting MEPs in both the right ECR and FCR (hot-spot) with a maximum intensity of 1.3 T. The resting motor threshold (rMT) was defined as the minimum stimulus intensity that produced an MEP amplitude with a magnitude greater than 50 μV for both the ECR and FCR for at least five out of ten stimulation trials. Stimulus intensity was set at 120% of the resting motor threshold during the experiment. The mean TMS intensity (± standard deviation, SD) was 68 ± 14% of the maximum output of the stimulator.

**Data analysis**

To evaluate corticospinal excitability, peak-to-peak MEP amplitudes of the ECR and FCR were recorded, and then standardized utilizing the average value of the resting task. Background EMGs of the ECR, FCR and TA were calculated as the RMS values of the EMGs for a 50 ms window just prior to the TMS. Trials with a background EMG activity of ECR and FCR that were greater than 25 μV were considered error trials and were eliminated from the analysis. During imagery dorsiflexion (DF), if any trial involved definite activity in the TA that was greater than 25 μV, data from that trial was also excluded from the analysis. The mean percentage of data rejection was 1.8 ± 0.9% for the background EMG of the ECR and FCR, and 2.9 ± 1.2% for TA activity during imagery DF.

For comparison of group data, a two-factor analysis of variance (ANOVA) with repeated measures was performed for the tasks (actual DF and imagery DF) and for EMG intensity (rest, 10%, 40% and 80% DF). The significances reported for the F-values are those obtained after the Greenhouse-Geisser correction (when appropriate); then a correction coefficient ε is given (only when the degree of freedom was adjusted). For post hoc comparisons, multiple pair-wise tests with Bonferroni corrections were performed. Each dorsiflexion force of the reproduction test was normalized with reference to the mean MVC value of each participant. The MVCs and the reproduction test were evaluated with a one-sample t test. To investigate the relationship between MEP amplitude for the imagery DF and for the actual DF (within participants), a Pearson’s correlation coefficient between changes in MEP amplitude of the ECR for actual DF and for imagery DF was calculated for the three different EMG intensities (10, 40 and 80%). The linear regressions on the MEP amplitudes of ECR and FCR during the actual and imagery DFs were analyzed in the range of 0 to 200% of rest values to determine whether their slopes were equal to 1. The residuals of all the dots from the average were compared using a partial correlation analysis. The coefficient of variation was compared to the average of the linear regression. Subsequently the results (residual) of the actual DF and imagery DF tasks were analyzed. Significance was set at p < 0.05. The data are expressed as mean ± SD.

**Results**

The mean scores of VMIQ for each intensity of imagery contraction were 10%; 1.8 ± 0.7, 40%; 1.9 ± 0.8, and 80%; 2.2 ± 0.7. Typical examples of MEP waves for ECR at three different EMG
levels (10, 40, 80% MVC) of foot dorsiflexion during the actual and imagery dorsiflexion tasks, as well as in the resting condition, are shown in Fig 2. For both ECR and FCR, MEP amplitude was markedly increased during both actual and imagery dorsiflexion. The extent of the increase was graded for the dorsiflexion EMG levels (actual 10%: 115.7 ± 20.5% of resting condition, actual 40%: 133.9 ± 29.4, actual 80%: 165.3 ± 45.7 for ECR) (imagery 10%: 113.9 ± 15.5, imagery 40%: 130.8 ± 25.5, imagery 80%: 155.1 ± 22.3 for FCR). The grand means (± SD) for all subjects are presented in Fig 3.

For the ECR, a two-factor repeated measures ANOVA demonstrated a significant main effect for intensity (F3, 77 = 44.16, p < 0.001). The two-way ANOVA did not demonstrate a significant main effect for task (F1, 77 = 0.93, p = 0.338) or interaction of the two factors (F3, 77 = 0.582, p = 0.628, e = 4.595). For the actual dorsiflexion, post hoc comparisons showed no significant difference between 10% MVC and the resting condition for actual dorsiflexion (p = 0.351). For imagery dorsiflexion, however, the MEP amplitude of 10% MVC was significantly higher than that of the resting condition (p < 0.01). The MEP amplitude of 40% MVC was significantly higher than that of the resting condition and the 10% MVC was significantly higher for both the actual and imagery dorsiflexion (p < 0.01, respectively). Likewise, the MEP amplitude of 80% was significantly higher than that of the resting condition, as were the 10% and 40% MVC for both actual and imagery dorsiflexion (p < 0.01 for both).

For FCR, a two-factor repeated measures ANOVA demonstrated a significant main effect for intensity (F3, 77 = 21.10, p < 0.001). The two-way ANOVA did not demonstrate a significant main effect for task (F1, 77 = 0.69, p = 0.409) or for the interaction of the two main factors (F3, 77 = 0.656, p = 0.582, e = 4.332). Post hoc comparisons showed no significant differences between the 10% MVC and the resting condition for either actual or imagery dorsiflexion (p = 0.876, p = 0.530, respectively). The MEP amplitude of 40% MVC for actual dorsiflexion was significantly higher than that of the resting condition and 10% MVC (p < 0.05). However, no significant differences were observed between 40% MVC and the resting condition or 40%

Fig 2. MEP waveform. Raw motor-evoked potential (MEP) waveform of the ECR elicited by single-pulse TMS in a participant at three different intensities (10, 40 and 80%) in the actual and imagery dorsiflexion (DF) task and the resting condition. Fifteen wave forms at each force level are superimposed.
MVC and the 10% MVC for imagery dorsiflexion (p = 0.221, p = 0.931). For actual and imagery dorsiflexion MEP amplitudes for the 10%, 40%, and 80% MVC were significantly higher than those of the resting condition (p < 0.01 for all).

Fig 4 illustrates the relationship between the MEP amplitudes of ECR (Fig 4A) and FCR (Fig 4B), both for the actual DF and the imagery DF. A significant correlation was obtained between MEP amplitudes for the actual DF and the imagery DF of ECR (r = 0.86, p < 0.01; Fig 4A). A significant correlation was also obtained between MEP amplitudes of the actual DF and imagery DF for FCR (relaxation; r = 0.54, p < 0.05; Fig 4B). Furthermore, we also evaluated
whether there was a difference between MEP amplitudes during actual DF and imagery DF over the range of 0 to 200% for resting values. For ECR, there was a close correlation between MEP amplitude during actual DF and imagery DF (Slope: $a = 0.8747$, $R^2 = 0.7345$). This slope was not significantly different from the slope of $y = x$ (which equals 1). For FCR, there was also a close correlation between MEP amplitude during actual DF and imagery DF (Slope: $a = 0.4854$, $R^2 = 0.3096$). For this slope as well, there was no significant difference from the slope of $y = x$ (which equals 1).

Participants were able to reproduce each force level (10% MVC condition; pre: 12.35 ± 3.3, post: 13.19 ± 3.4%; 40% MVC condition; pre: 44.52 ± 7.0, post: 42.13 ± 7.8%; 80% MVC condition; pre: 200.45 ± 8.9, post: 193.45 ± 8.2%).

**Fig 4. Correlation of MEP amplitude.** Relationship between MEP amplitude of ECR and FCR during the imagery of DF and of the actual DF. Each circle denotes a 10% (white circle), 40% (grey circle) and 80% (black circle) of MEP for all subjects. The dotted line indicates the linear regressions in the range of 0 to 200%.

https://doi.org/10.1371/journal.pone.0185547.g004
condition; pre: 75.69 ± 12.0, post: 73.53 ± 13.6%; 100% MVC condition; pre: 100 ± 0, post: 93.25 ± 7.5%). The MVC in the validation test did not differ from that obtained in the pre-experiment.

Discussion

The objective of the present study was to investigate how the level of imagined contraction in one limb influences the corticospinal excitability for the remote limb muscles. Corticospinal excitability of the hand muscles markedly increased during both actual and imagery dorsiflexion of the ipsilateral foot, and the extent of the increase was proportional to the intensity of contraction, either actual or imagined. Furthermore, correlations between MEP amplitudes during actual DF and imagery DF were observed for both the ECR and FCR.

These results suggest that the cortical mechanisms involved in the remote effect of motor imagery are similar to those involved in motor execution. The findings related to the remote effect of actual contraction correspond well with those of previous studies [36, 37, 42]. That is, corticospinal excitability for the upper limb muscles increases in association with increasing force levels produced by the lower limb. Mizuguchi et al. [18] previously demonstrated that during motor imagery of the contraction of a hand muscle, corticospinal excitability of the muscle itself changed in a manner dependent on the force level of the imagined contraction. Studies utilizing fMRI showed that the brain regions involved with motor imagery include the supplemental motor area (SMA), pre-SMA, premotor area (PM), primary motor cortex (PMC), posterior parietal cortex (PPC), basal ganglia, and cerebellum. In addition to activation during motor imagery, these areas are also active during actual contractions [10, 11, 12, 13, 14, 43]. Therefore, at a minimum, these regions are likely to be involved with the neural processing which underlies the remote effect that occurs during motor imagery. Tazoe et al. [42] noted that during an actual contraction, “the origin of the remote effect may be a higher motor centre located upstream of M1.” Since there are no anatomical connections between the hand and foot areas in M1, Brown et al. [44] and Huntley et al. [45] suggest that the supa M1 regions are involved in the remote effect. Furthermore, while M1 and SMA are arranged somatotopically, the pre-SMA is not [46, 47, 48]. Thus, it is possible that neural signals for the contraction of a target muscle from the pre-SMA to the SMA region might spread to the SMA that controls the remote site, although specific neural pathways and mechanism remain unclear.

As shown by the correlation between MEP amplitudes of actual DF and imagery DF, there is a similarity, even at the individual level, between the remote effects elicited by actual and imagery contractions. Participants who showed a higher MEP amplitude of the ECR during the actual DF also showed a higher MEP amplitude during imagery DF in the MEP amplitude range less than 200% (Fig 4). Interestingly, the slope of the linear regression was not significantly different from 1. The MEP amplitude during imagery DF was saturated in the range above 200%. Likewise, a correlation with a slope close to 1 was observed for the FCR, although the correlation coefficient was smaller than that for the ECR. These results strongly suggest that a similar neural pathway is activated for the remote effect during both motor imagery and actual contraction. This is in agreement with previous studies pertaining to the similarity of brain regions involved with motor imagery and actual contraction. On the other hand, MEP amplitude of the ECR plateaued during imagery DF, while the MEP amplitude during an actual contraction did not show any apparent saturation. Thus, saturation of the MEP amplitude during imagery DF likely occurs before the signals from the imagery process are fed into the common process of the remote effect.

Remote effects that occur during an actual contraction occur both from the upper limb to the lower limb and from the lower to the upper limb [30, 36, 49]. Since coincidental
enhancements of the H-reflex and MEPs by actual contraction of a remote site has been observed in both extensor and flexor muscles, the remote effect is thought to be non-specific [28,50]. In the present study, corticospinal excitabilities increased for both extensor and flexor muscles during an actual contraction, which corresponds well with the results of previous studies. However, we observed no significant differences between MEP amplitudes during imagery dorsiflexion of 10% and the resting condition for the FCR (Fig 3). Nor did we find any significant differences between MEP amplitudes during the imagery dorsiflexions of 10% and 40% MVC. Thus, relatively weak dorsiflexion did not alter the MEP amplitude of hand flexors. For the ECR, on the other hand, MEP amplitudes in the pronated hand position were clearly modulated by both actual and imagined contraction levels of dorsiflexion. During cyclic plantarflexion-dorsiflexion movement of the foot, MEP amplitude for the resting ECR in the pronated hand position is higher during dorsiflexion than during plantar flexion, while MEP amplitudes for the FCR is higher during plantarflexion than during dorsiflexion [51,52]. This suggests that neural activity that controls the hand extensors and flexors is affected by activity in the foot dorsiflexors and plantarflexors in a muscle specific manner. Since participants performed or imagined tonic dorsiflexion and not cyclic movements in the present study, connectivity of the remote effect might be different between muscles. Furthermore, changes in corticospinal excitability of the forearm under different positions (prone or supine) during cyclic movement of the foot has been shown to be direction-dependent and not muscle-dependent [53]. This suggests that the remote effect between the ipsilateral foot and hand might differ depending on hand position (prone or supine). We need further study to elucidate this.

In conclusion, we have provided evidence that there is an increase in corticospinal excitability of the system that provides input for the control of resting hand muscles. This increase is proportional to the imagined contraction level of the foot.

Acknowledgments
The authors thank Dr. Larry Crawshaw for English editing. This work was supported by JSPS KAKENHI Grant Numbers 15K16494 to K Kato and JP26242065 to K Kanosue. All authors declare no conflicts of interest.

Author Contributions
Conceptualization: Kouki Kato, Kazuyuki Kanosue.
Data curation: Kouki Kato.
Formal analysis: Kouki Kato.
Funding acquisition: Kouki Kato, Kazuyuki Kanosue.
Investigation: Kouki Kato.
Methodology: Kouki Kato, Kazuyuki Kanosue.
Project administration: Kouki Kato.
Resources: Kouki Kato.
Validation: Kouki Kato.
Visualization: Kouki Kato.
Writing – original draft: Kouki Kato.
Writing – review & editing: Kouki Kato, Kazuyuki Kanosue.
References

1. Jeannerod M (2001) Neural simulation of action: a unifying mechanism for motor cognition. NeuroImage 14: S103–S109. https://doi.org/10.1006/nimg.2001.0832 PMID: 11379140

2. Feltz LD, Landers DM (1983) The effects of mental practice on motor skill learning and performance: a meta-analysis. J Sport Psychol 5: 25–57.

3. Isaac AR (1992) Mental practice: Does it work in the field? The Sport Psychologist 6: 192–198.

4. Driskell JE, Copper C, Moran A (1994) Does mental practice enhance performance? J App Psychol 79: 481–492.

5. Pascual-Leone A, Dang N, Cohen LG, Cammarota A, Hallett M (1995) Modulation of muscle responses evoked by transcranial magnetic stimulation during the acquisition of new fine motor skills. J Neurophysiol 74: 1037–1045. PMID: 7500130

6. Holmes P, Calmels C (2008) A neuroscientific review of imagery and observation use in sport. J Mot Behav 40: 433–445. https://doi.org/10.3200/JMBR.40.5.433-445 PMID: 18782718

7. Allami N, Paulignan Y, Brovelli A, Boussaoud D (2008) Visuo-motor learning with combination of different rates of motor imagery and physical practice. Exp Brain Res 184: 105–113. https://doi.org/10.1007/s00221-007-1086-x PMID: 17849109

8. Lotze M, Halsband U (2006) Motor imagery. J Physiol Paris 99: 386–395. https://doi.org/10.1016/j.jphysparis.2006.03.012 PMID: 16716573

9. Mizuguchi N, Nakata H, Uchida Y, and Kanosue K (2012). Motor imagery and sport performance. J Phys Fitness Sports Med 1: 103–111.

10. Decety J, Jeannerod M, Prablanc C (1989) The timing of mentally represented actions. Behav Brain Res 40: 35–42. PMID: 2765170

11. Hanakawa T, Immisch I, Toma K, Dimyan MA, Van Gelderen P, Hallett M (2003) Functional properties of brain areas associated with motor execution and imagery. J Neurophysiol 89: 989–1002. https://doi.org/10.1152/jn.00132.2002 PMID: 12574475

12. Ehrsson HH, Geyer S, Naito E (2003) Imagery of voluntary movement of fingers, toes, and tongue activates corresponding body-part-specific motor representations. J Neurophysiol 90: 3304–3316. https://doi.org/10.1152/jn.01113.2002 PMID: 14615433

13. Guillot A, Collet C, Nguyen VA, Malouin F, Richards C, Doyon J (2009) Brain activity during visual versus kinesthetic imagery: an fMRI study. Hum Brain Mapp 30: 2157–2172. https://doi.org/10.1002/hbm.20658 PMID: 18819106

14. Mizuguchi N, Umehara I, Nakata H, Kanosue K (2013) Modulation of corticospinal excitability dependent upon imagined force level. Exp Brain Res 230: 243–249. https://doi.org/10.1007/s00221-013-3649-3 PMID: 23877227

15. Kasai T, Kawai S, Kawanishi M, Yahagi S (1997) Evidence for facilitation of motor evoked potentials (MEPs) induced by motor imagery. Brain Res 744: 147–150. PMID: 9030424

16. Stinear CM, Byblow WD (2004) Modulation of corticospinal excitability and intracortical inhibition during motor imagery is task-dependent. Exp. Brain Res 157: 351–358. https://doi.org/10.1007/s00221-004-1851-z PMID: 14997259

17. Fourkas AD, Ionta S, Aglioti SM (2006) Influence of imagined posture and imagery modality on corticospinal excitability. Behav Brain Res 168: 190–196. https://doi.org/10.1016/j.bbr.2005.01.015 PMID: 16313979

18. Mizuguchi N, Sakamoto M, Muraoka T, Kanosue K (2009) Influence of touching an object on corticospinal excitability during motor imagery. Exp Brain Res 196: 529–535. https://doi.org/10.1007/s00221-009-1875-5 PMID: 19504259

19. Evarts EV (1968) Relation of pyramidal tract activity to force exerted during voluntary movement. J Neurophysiol 31: 14–27. PMID: 4966614

20. Evarts EV, Fromm C, Kroller J, Jennings VA (1983) Motor cortex control of finely graded forces. J Neurophysiol 49: 1199–1215. PMID: 6864246

21. Werner W, Bauswein E, Fromm C (1991) Static firing rates of premotor and primary motor cortical neurons associated with torque and joint position. Exp Brain Res 86: 293–302. PMID: 1756804

22. Dai TH, Liu JZ, Sahgai V, Brown RW, Yue GH (2001) Relationship between muscle output and functional MRI-measured brain activation. Exp Brain Res 140: 290–300. PMID: 11613604

23. van Duinen H, Renken R, Maurits NM, Zijdewind I (2008) Relation between muscle and brain activity during isometric contractions of the first dorsal interosseus muscle. Hum Brain Mapp 29: 281–299. https://doi.org/10.1002/hbm.20388 PMID: 17394210
Kato K, Muraoka T, Mizuguchi N, Nakagawa K, Nakata H, Kanosue K (2016) Muscle Relaxation of the
Kato K, Watanabe J, Muraoka T, Kanosue K. (2015) Motor imagery of voluntary muscle relaxation
35.
34.
Kato K, Kanosue K (2016) Effect of muscle relaxation in the foot on simultaneous muscle contraction in
33.
32.
Hortobágyi T, Taylor JL, Pertersen NT, Russell G, Gandevia SC (2003) Changes in segmental and
31.
30.
Shen Y, Cao X, Tan T, Shen C, Dai W, Yuan TF. (2017) Heroin Addiction Impairs Human Cortical Plasticity.
39.
Tazoe T, Sakamoto M, Nakajima T, Endoh T, Shiozawa S, Komiyama T (2009) Remote facilitation of
36.
37.
Sabri Y, Cao X, Tan T, Shen C, Dai W, Yuan TF, et al. (2017) Methamphetamine abuse impairs motor
cortical plasticity and function. Mol Psychiatry. 22:1274–1281. https://doi.org/10.1038/mp.2017.143
PMID: 28831198
Tazoe T, Sakamoto M, Nakajima T, Endoh T, Shiozawa S, Komiyama T (2009) Remote facilitation of
supraspinal motor excitability depends on the level of effort. Eur J Neurosci. 30: 1297–1305. https://doi.org/10.1111/j.1460-9568.2009.06895.x PMID: 19769593
Shen Y, Cao X, Tan T, Shen C, Wang Y, Pan J et al., (2016) 10-Hz Repetitive Transcranial Magnetic
Stimulation of the Left Dorsolateral Prefrontal Cortex Reduces Heroin Cue Craving in Long-Term Addicts. Biol Psychiatry. 80: e13–e14. https://doi.org/10.1016/j.biopsych.2016.02.006 PMID: 26995024
Huang X, Chen YY, Shen Y, Cao X, Li A, Liu Q et al., (2017) Methamphetamine abuse impairs motor
cortical plasticity and function. Mol Psychiatry. 22:1274–1281. https://doi.org/10.1038/mp.2017.143
PMID: 28831198
24. Perez MA, Cohen LG (2009) Scaling of motor cortical excitability during unimanual force generation. Cortex 45: 1065–1071. https://doi.org/10.1016/j.cortex.2008.12.006 PMID: 19243741
23. Jendrásík E (1883) Biläute zur lehre von den sehnenreflexen. Dtsch Arch Klin Med 33: 177–199.
22. Delwaide PJ, Toulouse P (1981) Facilitation of monosynaptic reflexes by voluntary contraction of muscles in remote parts of the body. Brain 104: 701–719. PMID: 6275942
21. Zehr Stein RB (1999) Interaction of the Jendrásik maneuver with segmental presynaptic inhibition. Exp Brain Res 159: 382–388.
20. Kawakita H, Kameyama O, Ogawa R, Hayes KC, Wolfe DL, Allatt RD (1991) Reinforcement of motor evoked potentials by remote muscle contraction. J Electromyogr Kinesiol 1: 96–106. https://doi.org/10.1016/0952-6834(91)90003-N PMID: 20870499
19. Stedman A, Davey NJ, Ellaway PH (1998) Facilitation of human first dorsal interosseous muscle
responses to transcranial magnetic stimulation during voluntary contraction of the contralateral homonymous muscle. Muscle Nerve 21: 1033–1039. PMID: 9655121
18. Hortobágyi T, Taylor JL, Pertersen NT, Russell G, Gandevia SC (2003) Changes in segmental and
cortical output with contralateral muscle contractions and altered sensory inputs in humans. J Neurophysiol 90: 2451–2459. https://doi.org/10.1152/jn.01001.2002 PMID: 14534271
17. Kato K, Muraoka T, Higuchi T, Mizuguchi N, Kanosue K (2014) Interaction between simultaneous contraction and relaxation in different limbs. Exp Brain Res 232: 181–189. https://doi.org/10.1007/s00221-013-3730-y PMID: 24132527
16. Kato K, Watanabe T, Kanosue K. (2015) Effects of muscle relaxation on sustained contraction of ipsilateral remote muscle. Physiol Rep 3: 1–7
15. Kato K, Muraoka T, Mizuguchi N, Nakagawa K, Nakata H, Kanosue K (2016) Muscle Relaxation of the Foot Reduces Corticospinal Excitability of Hand Muscles and Enhances Intracortical Inhibition. Front Hum Neurosci 10: 218. https://doi.org/10.3389/fnhum.2016.00218 PMID: 27242482
14. Kato K, Kanosue K (2016) Effect of muscle relaxation in the foot on simultaneous muscle contraction in the contralateral hand. Neurosci Lett 633: 252–256. https://doi.org/10.1016/j.neulet.2016.09.019 PMID: 27693661
13. Kato K, Watanabe J, Muraoka T, Kanosue K. (2015) Motor imagery of voluntary muscle relaxation induces temporal reduction of corticospinal excitability. Neurosci Res 92: 39–45. https://doi.org/10.1016/j.neures.2014.10.013 PMID: 25448688
12. Tazoe T, Sakamoto M, Nakajima T, Endoh T, Komiyama T (2007) Effects of remote muscle contraction on transcranial magnetic stimulation-induced motor evoked potentials and silent periods in humans. Clin Neurophysiol 118: 1204–1212. https://doi.org/10.1016/j.clinph.2007.03.005 PMID: 17449319
11. Marconi B, Koch G, Pacchioni C, Cavallari P, Callagirone C (2007) Breakdown of inhibitory effects induced by foot motor imagery on hand motor area in lower-limb amputees. Clin Neurophysiol 118: 2468–78. https://doi.org/10.1016/j.clinph.2007.08.021 PMID: 17905652
10. Roberts R, Callow N, Hardy L, Markland D, Bringer J (2008) Movement imagery ability: development and assessment of a revised version of the vividness of movement imagery questionnaire. J Sport Exerc Psychol 30:200–221. PMID: 18940791
9. Shen Y, Cao X, Chan S, Dai W, Yuan TF. (2017) Heroin Addiction Impairs Human Cortical Plasticity. Biol Psychiatry. 81:e49–e50. https://doi.org/10.1016/j.biopsych.2016.06.013 PMID: 27567311
8. Shen Y, Cao X, Tan T, Shen C, Wang Y, Pan J et al., (2016) 10-Hz Repetitive Transcranial Magnetic Stimulation of the Left Dorsolateral Prefrontal Cortex Reduces Heroin Cue Craving in Long-Term Addicts. Biol Psychiatry. 80:e13–14. https://doi.org/10.1016/j.biopsych.2016.02.006 PMID: 26995024
7. Marconi B, Koch G, Pacchioni C, Cavallari P, Callagirone C (2007) Breakdown of inhibitory effects induced by foot motor imagery on hand motor area in lower-limb amputees. Clin Neurophysiol 118: 2468–78. https://doi.org/10.1016/j.clinph.2007.08.021 PMID: 17905652
6. Tazoe T, Sakamoto M, Nakajima T, Endoh T, Komiyama T (2007) Effects of remote muscle contraction on transcranial magnetic stimulation-induced motor evoked potentials and silent periods in humans. Clin Neurophysiol 118: 1204–1212. https://doi.org/10.1016/j.clinph.2007.03.005 PMID: 17449319
5. Roberts R, Callow N, Hardy L, Markland D, Bringer J (2008) Movement imagery ability: development and assessment of a revised version of the vividness of movement imagery questionnaire. J Sport Exerc Psychol 30:200–221. PMID: 18940791
4. Shen Y, Cao X, Chan S, Dai W, Yuan TF. (2017) Heroin Addiction Impairs Human Cortical Plasticity. Biol Psychiatry. 81:e49–e50. https://doi.org/10.1016/j.biopsych.2016.06.013 PMID: 27567311
3. Tazoe T, Sakamoto M, Nakajima T, Endoh T, Shiozawa S, Komiyama T (2009) Remote facilitation of supraspinal motor excitability depends on the level of effort. Eur J Neurosci. 30: 1297–1305. https://doi.org/10.1111/j.1460-9568.2009.06895.x PMID: 19769593
2. Makary MM, Eun S, Park K (2017) Greater corticostriatal activation associated with facial motor imagery compared with motor execution: a functional MRI study. Neuroreport. 28: 610–617. https://doi.org/10.1097/WNR.0000000000000808 PMID: 28538517
1. Brown P, Day BL, Rothwell JC, Thompson PD, Marsden CD (1991) Intrahemispheric and interhemispheric spread of cerebral cortical myoclonic activity and its relevance to epilepsy. Brain 114: 2333–2351. PMID: 1933248
Huntley GW, Jones EG (1991) Relationship of intrinsic connections to forelimb movement representations in monkey motor cortex: a correlative anatomic and physiological study. J Neurophysiol 66: 390–413. PMID: 1723093
46. Allison T, McCarthy G, Luby M, Puce A, Spencer DD (1996) Localization of functional regions of human mesial cortex by somatosensory evoked potential recording and by cortical stimulation. Electroencephalogr Clin Neurophysiol 100: 126–140. PMID: 8617151

47. Yazawa S, Ikeda A, Kaji R, Terada K, Nagamine T, Toma K et al., (1999) Abnormal cortical processing of voluntary muscle relaxation in patients with focal hand dystonia studied by movement-related potentials. Brain 122: 1357–1366. PMID: 10388801

48. Cauda F, Giuliani G, Federico D, Sergio D, Katiuscia S (2011) Discovering the somatotopic organization of the motor areas of the medial wall using low-frequency BOLD fluctuations. Hum Brain Mapp 32: 1566–1579. https://doi.org/10.1002/hbm.21132 PMID: 20814959

49. Toulouse P, Delwaide PJ. Reflex facilitation by remote contraction: topographic aspects (1980) Arch Phys Med Rehabil 61: 511–516. PMID: 7436711

50. Takada Y, Miyahara T, Tanaka T, Ohyama T, Nakamura Y. Modulation of H reflex of pretibial muscles and reciprocal Ia inhibition of soleus muscle during voluntary teeth clenching in humans. (2000) J Neurophysiol 83: 2063–2070. PMID: 10758116

51. Baldissera F, Borroni P, Cavallari P, Cerri G (2002) Excitability changes in human corticospinal projections to forearm muscles during voluntary movement of ipsilateral foot. J Physiol 539: 903–911. https://doi.org/10.1113/jphysiol.2001.013282 PMID: 11897859

52. Byblow WD, Coxon JP, Stinear CM, Fleming MK, Williams G, Müller JF et al., (2007) Functional connectivity between secondary and primary motor areas underlying hand-foot coordination. J Neurophysiol 98: 414–422. https://doi.org/10.1152/jn.00325.2007 PMID: 17507503

53. Borroni P, Cerri G, Baldissera F (2004) Excitability changes in resting forearm muscles during voluntary foot movements depend on hand position: a neural substrate for hand-foot isodirectional coupling. Brain Res 1022: 117–125. https://doi.org/10.1016/j.brainres.2004.07.003 PMID: 15353221