Channels rush in

Channels lurking in vesicles just beneath the cell surface can leap into action by inserting into the plasma membrane, say Vassilios Bezzerides, David Clapham (Harvard Medical School, Boston, MA), and colleagues. The resulting increase in \( \text{Ca}^{2+} \) current slows down and perhaps changes the outgrowth direction of advancing neurites.

The Boston group saw TRPC5 Ca\(^{2+}\) channels transferring into the plasma membrane, as measured by total internal reflection microscopy, electrophysiology, and surface biotinylation, in response to several growth factors. “Instead of controlling just gating, you are controlling availability,” says Clapham.

Recruitment and activation of TRPC5 Ca\(^{2+}\) currents were dependent on activated Rac and production of PIP\(_{2}\). In combination with previous work, this suggests the following scenario: activated growth factor receptors turn on PI3K-mediated production of PIP\(_{2}\), PIP\(_{3}\) recruits an exchange factor for Rac; and active Rac binds a kinase that produces PIP. The binding target for PIP\(_{2}\) may be synaptotagmin, which colocalizes with TRPC5 in vesicles.

The recruitment is transient, thus helping the cell to avoid flooding its narrow neurites with too much calcium. The calcium admitted by TRPC5 slows down neurites, possibly so they can respond to turning cues. Clapham now wants to see whether such “just in time” channel insertion events are a generalized feature of TRP channel regulation.

Reference: Bezzerides, V.J., et al. 2004. Nat. Cell Biol. doi:10.1038/ncb1150.

License to glue

Replication of DNA and cohesion of the resultant sister chromatids are two activities that must be coordinated. Now, Peter Gillespie and Tatsuya Hirano (Cold Spring Harbor Laboratory, Cold Spring Harbor, NY) have found that the same “licensing” process that gets DNA ready for replication is also necessary to allow loading of the cohesin complex responsible for sister chromatid cohesion.

Gillespie and Hirano used the sequence of yeast cohesin-loading proteins to identify human and frog versions. They confirmed biochemically in frog extracts what had been inferred genetically in yeast: that Scc2 (in frogs via two isoforms) is required for the loading of cohesin onto DNA. Association of Scc2 with chromatin was inhibited by two treatments that block DNA replication licensing: addition of geminin, a small protein that binds to the prereplication complex protein Cdt1, and depletion of an origin recognition complex subunit. The cyclin-dependent kinase inhibitor p21\(^{CIP1}\), which inhibits DNA replication initiation, had no such inhibitory effect.

Cohesin may get onto specialized sites such as the centromere and damaged DNA via other mechanisms, but at least in frogs the licensing machinery would make a sensible chaperone to ensure that the glue arrived before duplication. Gillespie and Hirano also suggested that mitotic Cdc2 activity displaced Scc2. Soon afterwards, most of the cohesin is displaced by Polo and Aurora B, two kinases downstream of Cdc2, thus allowing the sister chromatids to be prepared for the onset of anaphase and another cell cycle.

Reference: Gillespie, P.J., and T. Hirano. 2004. Curr. Biol. doi:10.1016/S0960982204005603.

Corralled by septin

Septin filaments in budding yeast make a corral that restricts certain division proteins to the site of cytokinesis, according to Jeroen Dobbelnaere and Yves Barral (ETHZ, Zurich, Switzerland).

The corral is formed when the septin ring at the bud neck splits to form two parallel fences. The Swiss researchers had earlier noted that during this period septin becomes transiently mobile before reverting to a static organization.

They have now extended this investigation of mobility to other cytokinesis proteins. Three membrane-associated proteins involved in polarizing the actin cytoskeleton (Spa2), secretion (Sec3), and delivering cell wall to the division site (Chs2) were all extremely mobile at the division site, but did not diffuse to regions outside of the division site. Disruption of the septin barriers via a temperature-sensitive mutant led to rapid loss of the division proteins from the bud neck and cytokinesis failure.

This trapping function for septins contrasts with their function earlier in the budding yeast cell cycle, when septins act as a scaffold that recruits other division proteins. But the later, membrane-associated division proteins do not colocalize with septins. Instead, they bounce around between the two wall-like septin rings. The split rings of fission yeast septin and even the septin patches seen in mammalian cells may perform a similar function.

Dobbelnaere suspects that the division proteins may be endocytosed and then delivered via vesicles to the site between the septin rings. The alternative is that membrane-associated proteins slip through the septin rings when the rings are flexible and then are trapped when the rings regain their solidity.

Reference: Dobbelnaere, J., and Y. Barral. 2004. Science. 305:393–396.

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