Salmonella Transiently Reside in Luminal Neutrophils in the Inflamed Gut

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Abstract

\textbf{Background:} Enteric pathogens need to grow efficiently in the gut lumen in order to cause disease and ensure transmission. The interior of the gut forms a complex environment comprising the mucosal surface area and the inner gut lumen with epithelial cell debris and food particles. Recruitment of neutrophils to the intestinal lumen is a hallmark of non-typhoidal \textit{Salmonella enterica} infections in humans. Here, we analyzed the interaction of gut luminal neutrophils with \textit{S. enterica} serovar Typhimurium (S. Tm) in a mouse colitis model.

\textbf{Results:} Upon S. Tm\textsuperscript{wt} infection, neutrophils transmigrate across the mucosa into the intestinal lumen. We detected a majority of pathogens associated with luminal neutrophils 20 hours after infection. Neutrophils are viable and actively engulf S. Tm, as demonstrated by live microscopy. Using S. Tm mutant strains defective in tissue invasion we show that pathogens are mostly taken up in the gut lumen at the epithelial barrier by luminal neutrophils. In these luminal neutrophils, S. Tm induces expression of genes typically required for its intracellular lifestyle such as siderophore production \textit{iroBCDE} and the \textit{Salmonella} pathogenicity island 2 encoded type three secretion system (TTSS-2). This shows that S. Tm at least transiently survives and responds to engulfment by gut luminal neutrophils. Gentamicin protection experiments suggest that the life-span of luminal neutrophils is limited and that S. Tm is subsequently released into the gut lumen. This “fast cycling” through the intracellular compartment of gut luminal neutrophils would explain the high fraction of TTSS-2 and \textit{iroBCDE} expressing intra- and extracellular bacteria in the lumen of the infected gut.

\textbf{Conclusion:} In conclusion, live neutrophils recruited during acute S. Tm colitis engulf pathogens in the gut lumen and may thus actively engage in shaping the environment of pathogens and commensals in the inflamed gut.

Introduction

Non-typhoidal \textit{Salmonella enterica} strains (NTS) cause a major fraction of world-wide bacterial infections. After oral ingestion of contaminated food, pathogens replicate in the gut lumen, invade mucosal tissues and trigger acute enteritis within hours after infection. Key pathological characteristics of human NTS infections are mucosal edema and acute inflammation with polymorph nuclear leukocyte (PMN) infiltration, PMN transmigration into the gut lumen and the formation of crypt abscesses [1,2,3,4]. These typical features are also recapitulated in bovine and murine NTS colitis models [5,6,7,8,9]. However, the role of neutrophils for antimicrobial defense in the gut lumen has not been addressed so far. It remained unclear whether gut luminal neutrophils represent “inactive” cellular waste from the inflammatory mucosal defense or whether they can actively engage in the host-pathogen interaction.

The acute inflammatory response in the gut plays an important role for the pathogens’ lifestyle as it changes microbiota-pathogen competition in favor of the pathogen [10,11]. In the absence of inflammatory conditions, the microbiota blocks pathogen colonization (= colonization resistance) [12,13]. By triggering mucosal inflammation, NTS can alter gut microbial ecology and thereby boost pathogen growth in the gut lumen [14,15]. Similar observations have been made for other enteric pathogens such as \textit{Citrobacter rodentium} as well as for \textit{Clostridium difficile} [16,17]. We are just beginning to unravel the mechanisms explaining the growth benefit of NTS in the inflamed gut.

\textit{Salmonella enterica} serovar Typhimurium (S. Tm) seems to be extremely well adapted to life in the inflamed gut. S. Tm can metabolize sugars released by the inflamed mucosa, which it can access via flagella-mediated chemotaxis [18,19]. Furthermore, it can utilize tetrathionate as an electron acceptor for anaerobic metabolism.
Numerous GFP-positive bacteria were detected in association with Salmonella enterica in the cecal tissues of mice infected with S. Typhimurium (S. Tm) [4]. We performed immunofluorescence microscopy of fixed tissue sections to visualize the interaction of neutrophils with Salmonella in the gut lumen. At 20 hours post infection with S. Typhimurium (S. Tm), neutrophils were imaged by confocal time-lapse microscopy as they migrated and engulfed bacteria [5].

Results

In the Inflamed Gut Lumen, S. Typhimurium (S. Tm) is Associated with CD18+ Neutrophils

Transmigration of neutrophils into the gut lumen is a cardinal feature of acute S. Typhimurium (S. Tm) infection. However, the interaction of neutrophils with pathogens in the gut lumen has not been addressed as far as S. Typhimurium (S. Tm) infection is concerned. In the streptomycin-colitis model, neutrophil infiltration and transmigration can be reliably observed [6]. S. Typhimurium (S. Tm) neutrophils in the gut lumen are a highly structured environment which offers distinct niches to be exploited for bacterial growth. The niches include the mucosal surface, the mucus layer and the inner gut lumen containing shed epithelial cells and food particles. It has remained unclear whether and how these different niches may support pathogen growth.

In this study, we set out to characterize the interaction of S. Typhimurium (S. Tm) with neutrophils that infiltrate the gut lumen in response to infection. We show that most S. Typhimurium (S. Tm) neutrophils in the gut lumen are engulfed by neutrophils right in the gut lumen and reside for a limited time within these host cells. Our data suggest that neutrophils should be considered as an additional intracellular compartment for residence and/or replication of pathogens in the gut lumen.

Life Microscopy Shows S. Typhimurium (S. Tm) Uptake by Active Neutrophils in the Gut Lumen

To actively take up S. Typhimurium (S. Tm) within the gut lumen after transmigration, neutrophils should be viable and phagocytic. To test this, we performed time lapse microscopy of neutrophils from explanted, S. Typhimurium (S. Tm)-infected ceca of LysMgfp/egfp mice [24]. In these mice, the egfp gene is knocked into the murine lysozyme M locus and leads to egfp expression particularly in mature, neutrophilic granulocytes [24]. Streptomycin-treated LysMgfp/egfp mice were infected with S. Typhimurium (S. Tm) for 20 hours to induce neutrophil transmigration. The cecum was explanted, flushed carefully and infected ex vivo with S. Typhimurium (S. Tm) egfp expressing Salmonella AutGsaV::aphT [25]. We used S. Typhimurium (S. Tm) avirulent, which lacks functional TTSS-1 and −2 type III secretion systems, to exclude that the bacteria could actively invade the neutrophils. In this way, we could focus our analysis on active bacterial uptake by the neutrophils. To visualize the rest of the mucosal tissue (besides the pathogen and the neutrophils), we included DAPI and the lipophilic styryl dye FM 4-64 into the assay buffer and mounted the organs as described [26]. Luminal neutrophils were imaged by confocal time-lapse microscopy as described [26]. We observed motile Egfp-positive neutrophils within gut lumen Movies S1, S2. Furthermore, we frequently detected Yfp-positive S. Typhimurium (S. Tm) avirulent containing within LysMgfp/egfp neutrophils in the gut lumen (Fig. 3AB). This demonstrated that gut luminal neutrophils are viable at least for a certain time after transmigration and that they can actively engulf S. Typhimurium (S. Tm) in the gut lumen.

S. Typhimurium (S. Tm) Uptake by Neutrophils Occurs in the Gut Lumen and is Fostered by TTSS-Independent, Flagella-Mediated Chemotaxis

As demonstrated above, pathogens can be directly taken up by transmigrated, actively phagocytic neutrophils within the gut lumen (Fig. 2; Fig. 4A). Alternatively (or additionally), bacteria could be taken up by neutrophils in the mucosal tissue and subsequently exported into the gut lumen. To distinguish between these different possibilities, we performed the following experiment. Streptomycin-treated mice were co-infected with 20h with a mixture of 50% S. Typhimurium (S. Tm) avirulent and 50% gfp-expressing S. Typhimurium (S. Tm) avirulent (popsM-gfp; positive control group) or a mixture of 50% S. Typhimurium (S. Tm) avirulent and 50% S. Typhimurium (S. Tm) avirulent (popsM-gfp; experimental group). S. Typhimurium (S. Tm) avirulent (popsM-gfp) neither invades mucosal tissues, nor replicates in the lamina propria, and thus is restricted mostly to the gut lumen.

By immunofluorescence microscopy of fixed tissue sections we differentiated between extracellular S. Typhimurium (S. Tm) and determined the fraction of intracellular gfp-expressing Salmonella spp in the gut lumen. Extracellular gfp-expressing S. Typhimurium (S. Tm) were
co-stained with an anti-Salmonella LPS antiserum (GFP⁺; LPS⁺) while intracellular S. Tm were not stained (GFP⁻; LPS⁻). We found a similar fraction (72.3 ± 17.2%) of S. Tmwt and S. Tmavir (65.1 ± 8.7%) located within luminal neutrophils (Fig. 4B). This strongly argues for the fact that the majority of pathogens is taken up by transmigrated neutrophils within the gut lumen. However, it should be noted that this microscopic assay cannot distinguish between life and dead neutrophils. This may explain why only a small fraction of the neutrophil-associated bacteria are efficiently protected from gentamicin (see Fig. 2).

We have shown before that S. Tm requires flagella-mediated chemotaxis to swim towards the epithelium [18]. In infected mice, flagellated, chemotactic bacteria are significantly enriched at the epithelium compared to the center of the gut lumen [19]. We took advantage of this fact and analyzed a S. Tm flagella mutant (S. TmΔfliGHIpprpsMgfp), a chemotaxis mutant (S. TmΔcheYpprpsMgfp) or an avirulent, chemotaxis mutant (S. TmΔcheYΔfliGHIpprpsMgfp) with regard to the fraction of luminal host-cell associated bacteria. The experiment was done in analogy to the one described above. We hypothesized, that neutrophils exiting the mucosa would first take up bacteria localized directly at the epithelium, a site reached by the pathogen in a flagella- and chemotaxis-dependent fashion (Fig. 4A). Indeed, a significantly lower fraction of S. TmΔfliGHI, S. TmΔcheY and S. TmΔcheYΔfliGHI was detected within neutrophils when compared to flagellated, chemotactic strains (Fig. 4B). This finding supports our hypothesis, that neutrophils engulf luminal S. Tm after transmigration into the gut lumen. Additionally, flagella and chemotactic motility may support engulfment by PMNs.

Again, the assay cannot distinguish between life and dead luminal neutrophils. This may explain the apparent difference to the data shown in Fig. 2. In the gentamicin-protection assay we only found approx. 1% of the luminal bacteria to be gentamicin-protected (Fig. 2). This number is much smaller than the fraction
of bacteria associated with neutrophils (Fig. 4) and may be due to the fact, that most luminal neutrophils are already dead and permeable during the gentamicin treatment (but not permeable to the α-LPS antibody). In this case, intracellular containment of S. Tm in luminal PMNs would only be a transient stage for S. Tm in the gut lumen. Alternatively, the majority of pathogens are killed upon neutrophil engulfment and neutrophils would thus be a “dead-end” for luminal S. Tm.

Expression of the SPI-2 TTSS is Upregulated by Intra- and Extracellular S. Tm in the Inflamed Gut Lumen

In order to distinguish whether S. Tm live or die within the luminal neutrophils, we set up an experiment to localize S. Tm that have adapted to an intracellular lifestyle in the gut lumen. The pathogenicity island 2 TTSS (TTSS-2) is one of the most widely used reporters for the intracellular stage of S. Tm [27,28,29]. We co-infected streptomycin-treated mice (n = 3) with an avirulent S. Tm strain carrying a reporter for TTSS-2 expression (S. Tmavir pssaG-gfp; no tissue invasion) with equal amounts of unlabeled S. Tmavir (n = 3; no inflammation) or unlabeled S. Tmwt (n = 3; inflammation). We compared the percentage of S. Tmavir expressing the TTSS-2-reporter under non-inflamed and inflamed conditions by fluorescence microscopy (Fig. 5AB). We found a significant increase of TTSS-2-reporter expression in the inflamed gut (red symbols) versus the non-inflamed cecal lumen (blue symbols; median 1.34% versus 0.34%). This suggested, that a higher fraction of luminal S. Tmavir adapts to an intracellular lifestyle in the inflamed gut.

Next, we analyzed gfp-expressing S. Tmavir with respect to their localization in the inflamed gut. Approx. 2/3 of this population was located extracellularly (median 0.96%; co-stained with the α-LPS antiserum, not associated with CD18+ cells) and approx. 1/3 was located in association with CD18+ neutrophils (median 0.44%; not stained with the α-LPS antiserum, contained in CD18+ cells). These data lends further support to the notion that S. Tm is just transiently contained within CD18+ cells in the gut lumen. This

Table 1. S. Tm strains and plasmids.

| S. Tm strains | Genotype | Reference |
|---------------|----------|-----------|
| S. Tm[w] (SB300) | S. Tm strain SL1344 | [39] |
| S. Tm[a] invG (M557) | S. Tm Δ invG, ssaV:aphT | [25] |
| S. Tm[Kont] (M913) | S. Tm fliGHI::Tn10 | [18] |
| S. Tm[vir] (M935) | S. Tm cheY::Tn10 | [18] |
| S. Tm[vir] invG (M933) | S. Tm fliGHI::Tn10; Δ invG; ssaV:aphT | [19] |
| M960 | S. Tm Δ invG; ssaV:aphT. Histidin prototrophic variant by P22-transduction from S. Tm ST4/74 [40] | [19] |

Plasmids

| Plasmids | Genotype | Reference |
|----------|----------|-----------|
| ppsM-gfp (pM979) | ppsM-gfp | [18] |
| pssaG-gfp (pM975) | pssaG-gfp | [28] |
| piroBCDE-gfp (pM1445) | piroBCDE-gfp | This study |
| psodB-gfp (pM1446) | psodB-gfp | This study |
| ptYFP turbo YFP | Constitutive yfp expression | [26] |

Gut Luminal Neutrophils and Salmo

Figure 2. Luminal S. Tm are gentamicin-protected. Gentamicin-protection assay of gut luminal S. Tm. Streptomycin-treated mice (n = 9) were infected with S. Tm[w]. 20h post infection, the mice were sacrificed and cecum contents collected in PBS and split into 4 fractions (untreated; treated with triton-X-100 (0.1%) for 30 minutes; treated with gentamicin (400 μg/ml) for 30 minutes; or treated with triton-X-100 (0.1%) and gentamicin (400 μg/ml) for 30 minutes). Samples were diluted and plated on McConkey agar plates (100 μg/ml streptomycin), allowing specific detection of S. Tm. Bars show mean and StD of S. Tm[w] CFU/ml; xxx = p < 0.0001.

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Tmwt and sacrificed 24h post infection. The cecum was explanted and +

CD18 upon death/lysis of the neutrophil (i.e. Tm indicates that these bacteria are set free at a certain stage (i.e. S. shown that Tmavir. Scale bars: 10

GFP blue) and FM 4–64 (epithelial membranes; red), mounted on an organ expressing S. ex vivo infected Tm Contained within Luminal Neutrophils Induce

stage lasts long enough to respond with tiss-2 expression. In contrast, the relatively high fraction of TTSS–2+ extracellular S. Tm indicates that these bacteria are set free at a certain stage (i.e. upon death/lysis of the neutrophil).

Taken together, our data suggest that at least part of the gut luminal pathogen population is transiently contained within CD11b+ cells and that the intracellular phase is long enough to adapt to an intracellular lifestyle.

S. Tm Contained within Luminal Neutrophils Induce Expression of iroBCDE

Finally, we wanted to more closely analyze the environment of neutrophil-contained S. Tm. Raffatellu et al. have recently shown that S. Tm circumvents the host antimicrobial peptide lipocalin-2, which sequesters the siderophore enterochelin and is induced in the inflamed gut. Lipocalin-2 is expressed by epithelial cells [22] as well as neutrophils [30]. S. Tm harbors a gene cluster for production of the siderophore salmochelin and its cognate receptor IroN (iroBCDE iroN) which is not inhibited by lipocalin-2. However, it remained unclear whether enterochelin-mediated iron uptake is expressed by bacteria freely in the gut lumen or while the pathogen resides within gut luminal neutrophils.

We set out to analyze if S. Tm expresses iroBCDE in gut luminal neutrophils. To this end, we generated a GFP-reporter for the iroBCDE promoter (piroBCDE-gfp). Expression of iroBCDE is regulated via Fur, induced upon iron starvation and repressed under iron excess (Fig. S1) [31]. Streptomycin-treated mice were infected with S. Tmwt (n = 3) or S. Tmavir (n = 3) containing piroBCDE-gfp to analyze gfp-expression under control of iroB promoter under inflammatory and non-inflammatory conditions, respectively. Tissues were fixed and bacteria in different regions within the gut lumen were analyzed with respect to gfp-expression by automated image analysis. All bacteria positively stained with the α-Salmonella LPS antiserum (red) were analyzed with respect to GFP-fluorescence intensity (green). On average, 1’200 bacteria per mouse (≥ 3 different cecum lumen tissue sections) were analyzed this way and we found a significantly lower GFP-intensity in S. Tmwt as compared to S. Tmavir infected mice (Fig. 6A, left panel). We hypothesized that our analysis did not include those bacteria residing within neutrophils as the tissue was not permeabilized prior to staining with the α-Salmonella LPS antiserum.

Therefore the same analysis was performed on sections that were permeabilized with 0.1% triton X 100 before α-Salmonella LPS staining. Now, the GFP-intensity of piroBCDE-gfp expressing S. Tm was significantly higher in the lumen of inflamed mice (Fig. 6A; compare left and right panels). This suggested that piroBCDE-gfp was particularly induced in the pathogen-subpopulation residing within the gut luminal neutrophils.

Expression of the iroBCDE promoter is under the control of the Fur-regulon and expressed under iron-limiting conditions [21]. Our results suggest that neutrophils in the inflamed gut lumen constitute an iron-limiting environment for bacteria. To verify this, we used a second GFP-reporter construct (psodC-gfp). SodC encodes an iron-superoxide dismutase which is also regulated by Fur. However, sodC expression is induced in the presence of iron and is therefore regulated in the opposite fashion as iroBCDE [31] (Fig. S1). Indeed we found that psodC-gfp expression was significantly lower in the inflamed than in the non-inflamed gut (Fig. 6B). Triton permeabilization had no effect on the difference of psodC-gfp expression between inflamed and non-inflamed gut.

This data extends earlier reports on iron limitation in the inflamed gut [22] by demonstrating that iron limitation pertains to extracellular bacteria as well as to S. Tm residing within gut luminal neutrophils. In conclusion these data show that S. Tm expresses iroBCDE in neutrophils. This may be due to general iron limiting milieu (i.e. due to increased lipocalin-2 expression by the mucosa and possibly also neutrophils) in the inflamed gut lumen. This indicates that the transmigrating neutrophils are still active in the gut lumen with respect to bacterial uptake and iron limitation in the phagocytic vacuole.

Discussion

Non-typhoidal Salmonella spp. can exploit the host defense to compete against the intrinsic microbiota. Here, we characterized the interaction of NTS with neutrophils, an important cellular component of the innate host defense within infected tissues. In response to S. Tm infection, neutrophils are attracted to the site of infection. Within 10h after oral S. Tm+ infection of streptomycin-treated mice, neutrophils can already be observed in the cecal mucosa at high numbers [18]. Furthermore, neutrophils even transmigrate into the gut lumen, frequently leading to crypt abscess formation.

It remained unclear what role neutrophils play for growth and survival of S. Tm in the gut lumen. Upon infection, S. Tm+ grows at high titers (107 to 109 cfu/g) in the gut lumen despite the presence of luminal neutrophils. This condition holds up for at
least 4 days without reducing $S. Tm^{wt}$ numbers in the cecal lumen or in the feces [14]. This suggests that neutrophils are not able to eliminate the pathogen from the gut lumen. Similarly, a previous study showed that $S. Tm$ can survive quite well within murine neutrophils [32]. Our data from the gentamicin-protection assay demonstrated that viable $S. Tm$ can indeed be recovered from gut luminal neutrophils. Interestingly, the key virulence factor for $S. Tm$ intracellular survival, TTSS-2, although expressed within neutrophils, seems to be dispensable for gut luminal survival of $S. Tm$. This was previously demonstrated in co-infection experiments competing $S. Tm^{wt}$ with a mutant lacking functional TTSS-2 [14].

The ttss-2 mutant colonized the inflamed gut equally well as the wild type up to day 4 p.i. As discussed below, the lack of a TTSS-2 mediated growth/survival benefit for the pathogen in the gut lumen might be explained by the short life-span of the gut luminal neutrophils.

Strikingly, up to ~70% of the $S. Tm$ bacteria were localized within neutrophils at day 1 p.i., as determined by fluorescence microscopy, suggesting that most bacteria in the gut lumen do actually encounter neutrophils (Fig. 4B). On the other hand, just 0.62% of the viable $S. Tm$ in the cecal contents (as determined by plating) were actually protected against genta-
micin. This apparent discrepancy could be explained if neutrophils had just a limited “active” lifetime upon transmigration into the intestinal lumen. This would be in line with the relatively short lifetime of murine neutrophils in bone marrow or after isolation from blood [33,34]. However, the limited lifetime in the gut lumen should be long enough for the uptake (and possibly elimination) of bacteria localized right at the mucosal surface as indicated by the active phagocytosis of S. Tmavir and the intracellular induction of the pssaG-gfp reporter. Afterwards, the neutrophils would die off thus rendering the internalized bacteria prone to killing in the gentamicin protection assay. Nevertheless, many bacteria seem to remain associated with these dead/dying neutrophils (and can thus be detected within neutrophils by fluorescence microscopy).

The long-lived nature of Gfp within the bacterial cytosol could explain why pssaG-gfp and piroBCDE-driven gfp expression can still be detected long after the active life of the neutrophil (incl. intracellular iron limitation) has ended by the host cell’s demise.

Figure 5. *Salmonella* induce expression of the SPI-2 TTSS in neutrophils in the inflamed intestine. 2 groups of streptomycin-treated mice (n = 3) were co-infected with S. Tmavir containing the GFP-reporter plasmid pssaG-gfp and either S. Tmwt (red; inflammation induced) or S. Tmavir (blue; no inflammation induced). 20h post infection, the mice were sacrificed and cecum prepared for immunofluorescence staining of S. Tm with α-Salmonella-O-antigen group B serum and anti-CD18 as described in the materials and methods section. The tissue was not treated with triton. A. Representative images of intra- and extracellular S. Tmavir pssaG-gfp in the cecal lumen. Scale bar: 20μm and 5 μm. α-Salmonella LPS antiserum (blue), CD18 (red) and GFP (green). B. Quantitative image analysis of % gfp-expressing of all LPS+ S. Tm in the inflamed and non-inflamed gut. In the inflamed gut, the % of extracellular (GFP+ LPS+) and intracellular (GFP+ LPS+ and CD18-associated) was determined as well. Bars show the median. doi:10.1371/journal.pone.0034812.g005
Most parts of the normal anaerobic microbiota are out-competed by \textit{S. Tm} in the inflamed gut. Neutrophils have been proposed to modify the microbiota-pathogen interaction [35]. Neutrophils harbor an arsenal of antimicrobial defenses including oxidative burst and the generation of reactive oxygen species (ROS), antimicrobial peptides (calprotectin, lipocalin, lysozyme) and formation of neutrophils extracellular traps (NETs) [36,37]. Most commensals would probably show a higher susceptibility to this variety of antimicrobial defenses. Deciphering the different possible contributions of neutrophils to the microbiota-pathogen competition in the gut lumen will be a challenging subject of future research.

Our data suggest that neutrophils may form a distinct compartment in the inflamed gut lumen. Thus, neutrophils could affect the lifestyle of pathogens as well as commensals by generating an intracellular compartment displaying specific physiological properties. We found that neutrophil-contained \textit{S. Tm} up-regulated the promoter for \textit{iroBCDE}. This suggests that gut luminal neutrophils limit iron availability to the engulfed bacteria. Furthermore, our data suggest that \textit{S. Tm} responds to iron limitation experienced within the inflamed gut lumen by expressing salmochelin. Expression of \textit{iroN}, the salmochelin-receptor has been shown to entail a competitive advantage for \textit{S. Tm} in the inflamed gut as it overcomes inhibition by the antimicrobial protein lipocalin-2. Interestingly, lipocalin-2 is known to be expressed by human and murine neutrophils [30,38]. This suggests that salmochelin expression may be important for \textit{S. Tm} interaction and/or survival within gut luminal neutrophils.

\textbf{Figure 6.} \textit{Salmonella} contained within luminal neutrophils experience iron limitation. 4 groups of streptomycin-treated mice (n = 3) were infected with \textit{S. Tm} (red; inflammation induced) or \textit{S. Tm} (blue; no inflammation induced) containing the GFP-reporter plasmids \textit{piroBCDE-gfp (A, C)} or \textit{psodB-gfp (B, D)}, respectively. 20 h post infection, the mice were sacrificed and cecum prepared for immunofluorescence staining of \textit{S. Tm} with \textit{α-Salmonella O-antigen group B serum} as described in the materials and methods section. The tissue was either not permeabilized (w/o triton) or permeabilized with 0.1% triton prior to staining (+ triton). All bacteria positively stained with the \textit{α-Salmonella} LPS antiserum were included for analysis and \textit{S. Tm} GFP-fluorescence intensity (grey levels; y-axis) was determined by automated image analysis (A, B). Representative images of \textit{S. Tm} or \textit{S. Tm} in the cecal lumen expressing \textit{piroBCDE-gfp} (C) or \textit{psodB-gfp} (D) (green). The section was permeabilized with triton-x-100 and stained with \textit{α-Salmonella} LPS antiserum (red) and phalloidin (blue). Scale bar: 10 μm.

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\textit{Salmonella} GFP Actin
The respiratory burst of neutrophils that transmigrate into the intestinal lumen during inflammation oxidizes endogenous sulfur compounds to generate a respiratory electron acceptor, tetra-thionate [20]. It is tempting to speculate that this oxidation takes place in the neutrophil compartment. In this case, intracellular S. Tm may capitalize on the tetra-thionate, available within neutrophils. In light of the data showing a defect of the S. Tm chemotaxis mutant to enter neutrophils, one could even speculate that the bacteria move along chemotactic gradients of such anaerobic electron acceptors released by luminal neutrophils. Future experiments will have to address this.

In conclusion, we identified gut luminal neutrophils as novel niche for S. Tm in the infected gut. Our results suggest that this intra-neutrophil niche may affect S. Tm lifestyle. Future studies will have to address the exact underlying mechanisms as well as the role of neutrophils in microbiota-pathogen competition within the inflamed gut.

Materials and Methods

Construction of Plasmids

Promoter regions of iroBCDE and sodB were amplified with the primer pair iroBCDE-F-XbaI/iroBCDE_R-BamHI and sodB-F-XbaI/sodB_R-BamHI, respectively as described before [31] and cloned into pM968 ([18]; promoterless pBAD24-derivative) via XbaI and BamHI to generate piroBCDE-gfp and piroB-sodB, respectively.

Animal Experiments

All mice used in the study were on C57Bl/6 background and bred at the Rodent Center at the ETH Zurich and the Max-von Pettenkofer Institute under SPF conditions in individually ventilated cages. For S. Tm infection, mice were treated with streptomycin (20 mg/animal 20 h prior to Salmonella infection) and infected by oral gavage with 5 × 10⁷ cfu S. Tm or mixtures of S. Tm as described [5]. Live bacterial loads in the cecal content were determined by plating on MacConkey-agar (Oxoid) with respective antibiotics (streptomycin 100 μg/ml; kanamycin 30 μg/ml; chloramphenicol 30 μg/ml; ampicillin 100 μg/ml; tetracycline 12 μg/ml). Histology was done at necropsy.

For ex vivo Salmonella infection of cecal tissue, strain S. Tmaw with the YFP plasmid was cultivated under aeration and appropriate antibiotic therapy and constant agitation in high-salt LB medium (300 mM NaCl) until an OD₅₆₀ of 0.8–1.0 was reached. Bacteria were harvested by centrifugation (5000 rpm, 5 min) and once washed in PBS. About 1 × 10⁷ cfu were administered in a volume of 50 μl. The nuclei were stained with DAPI at concentrations of 0.2 μg ml⁻¹ and with lipophilic styryl dye FM 4-64 (Invitrogen; 10 μg ml⁻¹). The tissue was mounted on an organ holder and analysed using an inverse Leica SP5 confocal laser scanning microscope (Leica Microsystems, Mannheim, Germany) in a 37°C chamber.

Ethics Statement

All animal experiments were approved (license 201/2004 and 201/2007 Kantonaes Veterinarnzt Zurich) and the Regierung von Oberbayern and performed according to local guidelines (TschV, Zurich), the Swiss animal protection law (TschG) and the Deutsches Tierschutzgesetz.

Immunofluorescence

Cecal tissues were recovered and treated as described [19]. Briefly, the tissue was fixed in paraformaldehyde (4% in PBS, pH 7.4 overnight, 4°C), washed with PBS, equilibrated in PBS (20% sucrose, 0.1% Na₂O₃ overnight, 4°C), embedded in O.C.T. (Sakura, Torrance, CA), snap-frozen in liquid nitrogen and stored at −80°C. Cryosections (7 μm) were air-dried for 2 h at room temperature, fixed in 4% paraformaldehyde (5 min). Tissue was optionally permeabilized for 30 minutes in PBS 0.1% triton X 100. Sections were washed and blocked in 10% (w/v) normal goat serum in PBS for 1 h. S. Tm were detected by staining for 1 h with a polyclonal rabbit α-Salmonella-O-antigen group B serum (factors 4, 5; Difco; 1:300) and Cy3-conjugated secondary goat-α-rabbit antibody (Jackson). Neutrophils were stained with rat-α-CD11b antibody (BD-Pharmingen) and Cy5-conjugated secondary goat-α-rabbit antibody (Jackson). DNA was stained with Sytox-green (1:10,000, Invitrogen). Sections were mounted with Vectashield (Vector laboratories). Imaging was done using a Leica SP5 confocal laser scanning microscope (Leica Microsystems, Mannheim, Germany; Fig. 1, 3) or an Axiovert 200M inverted microscope (Zeiss; Fig. 4, 5 and 6).

For immunofluorescent staining of extracellular S. Tm in cecum sections (Figure 4, 5 and 6), non-permeabilized cecum sections (7 μm) were stained with rabbit α-Salmonella-O-antigen group B serum (factors 4, 5; Difco; 1:300) and Cy3-conjugated secondary goat-α-rabbit antibody (Jackson). The fraction of intracellular GFP⁺ S. Tm was determined as (GFP⁺/ GFP⁺ LPS*) × 100%. 200–800 bacteria per group were included into the analysis.

For automated image analysis (Fig. 6), confocal stacks of immunofluorescence stained cecal tissue were recorded with an Ultraview confocal head (PerkinElmer), a krypton argon laser (643-RYB-A01; Melles) and Plan Neofluar 64x oil objective, aperture setting 1.25 (Zeiss). Confocal stacks were recorded with Volocity software (Improvision Ltd.) with a distance of 0.5 μm between the layers. From each cecal lummen, a total of 6 pictures were taken. GFP intensity was measured automatically with the Volocity Quantitation tool using following conditions: (1) selection of all red bacteria (2) mean green intensity of all selected bacteria from 300 to 1000 was considered for analysis.

Gentamicin Protection Assay

Cecum content of mice was harvested at day 1 post infection with S. Tmaw. The content was resuspended in PBS and split into 4 fractions. The fractions were either treated with triton-X-100 (0.1%) to lyse all intact host cells or with gentamicin (400 μg/ml) to kill all extracellular bacteria or with both (triton-X-100 and gentamicin) to kill all bacteria, or left untreated, as control. Samples were incubated at 37°C, shaking for 30 minutes. Samples were centrifuged (10,000 rpm, 2 min) and washed twice with PBS. Samples were then treated with 1% triton-X-100 and plated on LB containing 100 μg/ml streptomycin in appropriate dilutions.

Statistical Analysis

Statistical analysis was performed using the exact Mann-Whitney U Test (Graphpad Prism Version 5.01). P-values less than 0.05 (2-tailed) were considered statistically significant.

Supporting Information

Figure S1 Iron responsiveness of the two reporter constructs piroBCDE-gfp and piroB-sodB. The histidine-prototrophic S. Tmaw strain M960 harboring either piroBCDE-gfp (left) or piroB-sodB (right). The bacteria were grown in M9 medium with glucose (2 g/l) over night and diluted 1:20 in fresh M9 media containing either the iron chelator diethylenetriamine pentaacetic acid (A, DTPA; 0.1 μM, 1 μM, 10 μM), no supplement, or Fe(III) chloride (B, FeCl₃; 1 μM, 10 μM, 100 μM) to vary iron availability. The subcultures were incubated...
for 4 h at 37°C on the rotating wheel and then submitted to FACS analysis on an LSR II instrument (BD Biosciences). Data analysis was performed with FlowJo 7.3 (Tree Star, Inc.). The bacterial population was gated in FCS/SSC and GFP fluorescence was measured in FL1 channel (histogram).

**Movie S1  Time-lapse microscopy of neutrophils in the lumen of an explanted cecum.** Streptomycin-treated mice LysM<sup>GFP</sup> +/- mice were infected with S. Tm<sup>avir</sup> and sacrificed 24 h post infection as described in main Figure 3. The cecum was explanted and infected <i>ex vivo</i> with Tm<sup>avir</sup> expressing S. Tm<sup>Tmwt</sup>, stained with DAPI (DNA; blue) and FM 4-64 (epithelial membranes; red), mounted on an organ holder and analyzed by confocal real time microscopy of S. Tm and GFP<sup>P</sup> neutrophils. The videos show motile neutrophils (GFP; green) in the lumen of the explanted cecum. Scale bars: 10 μm. (AVI)

**Movie S2  Time-lapse microscopy of neutrophils in the lumen of an explanted cecum.** Streptomycin-treated mice LysM<sup>GFP</sup> +/- mice were infected with S. Tm<sup>Tmwt</sup> and sacrificed 24 h post infection as described in main Figure 3. The cecum was explanted and infected <i>ex vivo</i> with Tm<sup>avir</sup> expressing S. Tm<sup>Tmwt</sup>, stained with DAPI (DNA; blue) and FM 4-64 (epithelial membranes; red), mounted on an organ holder and analyzed by confocal real time microscopy of S. Tm and GFP<sup>P</sup> neutrophils. The videos show motile neutrophils (GFP; green) in the lumen of the explanted cecum. Scale bars: 10 μm. (AVI)

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