Prognostic Implications of Pan-cancer CMTM6 Expression and Its Relationship With the Immune Microenvironment

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Primary research

Keywords: CMTM6, pan-cancer, immune microenvironment, prognosis

DOI: https://doi.org/10.21203/rs.3.rs-38154/v1

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Abstract

**Background:** CKLF-like MARVEL transmembrane domain-containing 6 (CMTM6) reportedly stabilizes programmed death-ligand 1 (PD-L1) and enhances the efficacy of immunotherapy. However, correlations between CMTM6 expression and the immune microenvironment and its prognostic value remain unknown in a variety of tumors.

**Methods:** CMTM6 expression data were obtained from The Cancer Genome Atlas (TCGA) for 33 cancer types classified into high and low expression subgroups according to the median CMTM6 expression value. Pan-cancer analysis of CMTM6 protein expression in 20 tumor types was performed using a cohort from the Human Protein Atlas (HPA). PD-L1 protein expression data were obtained from The Cancer Proteome Atlas (TCPA) for 32 cancer types. Frequencies of CMTM6 copy number alterations and mutations were analyzed using cBioPortal. MANTIS was employed to estimate microsatellite instability in the TCGA cohort. CIBERSORT and the ESTIMATE algorithm were applied to estimate the relative fractions of infiltrating immune cell types and immune scores, respectively. Kaplan–Meier survival curve analysis was performed to assess the pan-cancer prognostic value of CMTM6.

**Results:** CMTM6 is heterogeneously expressed in diverse cancers. Further, the results revealed low CMTM6 mutation frequencies in multiple cancers. Among them, CMTM6 mutation frequency was the highest in uterine cancer. Additionally, CMTM6 expression was related to PD-L1 protein expression in breast invasive carcinoma, cervical squamous cell carcinoma and endocervical adenocarcinoma, cholangiocarcinoma, glioblastoma multiforme (GBM), head and neck squamous cell carcinoma, kidney renal papillary cell carcinoma, sarcoma (SARC), stomach adenocarcinoma, and uterine carcinosarcoma. Increased CMTM6 expression may be associated with increased infiltration of neutrophils in some types of cancer. Finally, pan-cancer analysis indicated that CMTM6 expression was closely related to overall survival in adrenocortical carcinoma, GBM, acute myeloid leukemia, liver hepatocellular carcinoma, mesothelioma, SARC, thymoma, and uveal melanoma.

**Conclusions:**

Taken together, these findings highlight that CMTM6 plays an important role in the tumor immune microenvironment, and CMTM6 has been identified to have prognostic value in some types of cancers. Thus, CMTM6 is a potential target for cancer immunotherapy and an effective prognostic biomarker.

**Background**

CKLF-like MARVEL transmembrane domain-containing family (CMTM) is a novel member of the human chemokine-like factor gene superfamily, which includes CMTM 1-8 [1]. CMTM6 is a widely expressed protein that exists in clusters on human chromosome 3p23. It exhibits sequence homology with protein products of other family members and has a potential four-time membrane-penetrating structure. In recent years, the relationship between CMTM6 and tumorigenesis has attracted increasing attention.
CMTM6 expression in gliomas was previously correlated with poor prognosis, and its expression was positively correlated with inhibitory T-cell expression [2]. However, Joh et al. reported that CMTM6 was significantly associated with longer overall survival in non-small cell lung cancer [3]. Previous studies indicate that CMTM6 plays different roles in different tumors.

At present, monoclonal antibodies targeting programmed death-1 (PD-1) receptor and its ligand (PD-L1) have demonstrated clinical responses and survival improvement in the treatment of patients with advanced-stage cancers [4]. CMTM6 can be used as a key regulator of PD-L1 protein in a broad range of cancer cells. CMTM6 both stabilized PD-L1 expression and prevented its lysosome-mediated degradation [5-6]. Additionally, CMTM6 improved PD-1/PD-L1 inhibitor efficacy through modulation of PD-L1 expression and tumor-infiltrating lymphocytes [7]. Therefore, fully understanding the relationship between CMTM6 and the immune microenvironment is of great importance to optimize patient benefit and guide combination approaches to treatment.

In this present study, we comprehensively analyzed the association between CMTM6 expression and the immune microenvironment and investigated its correlation with pan-cancer prognosis.

**Methods**

**Pan-cancer analysis of CMTM6 expression**

CMTM6 mRNA expression data from 11,093 samples of normal and tumor tissues comprising 33 cancer types were downloaded from The Cancer Genome Atlas (TCGA) database (https://portal.gdc.cancer.gov/). The expression profile data were classified into high and low expression groups according to the median value of CMTM6 expression. Pan-cancer analysis of CMTM6 protein expression in 20 tumor types was performed using the Human Protein Atlas (HPA) database (https://www.proteinatlas.org/).

**Pan-cancer analysis of CMTM6 copy number alterations and mutations**

The cBio cancer genomics portal (http://cbioportal.org) is an open-access resource capable of analyzing genomic alterations from various cancer samples [8]. We used cBioPortal to identify frequencies of CMTM6 copy number alterations and mutations.

**Tumor mutational burden estimates**

Mutation annotation files were downloaded using the TCGAbiolinks package in R. Somatic mutation calling was performed using the MuTect2 pipeline (Genome Analysis Toolkit (GATK), Broad Institute, Cambridge, MA, USA). The read.maf function was used to read somatic variants of each sample. The
tumor mutational burden (TMB) was defined as the number of somatic variants per megabase of genome [9].

**PD-L1 protein expression**

PD-L1 protein expression data were obtained from 6,944 tumor samples comprising 32 cancer types from The Cancer Proteome Atlas (TCPA) database (https://tcpaportal.org/tcpa/).

**Microsatellite instability (MSI)**

MANTIS was employed to estimate MSI across 33 cancer types from the TCGA database. The average distance threshold value = 0.4 was used to distinguish microsatellite stable (MSS) tumors from those with high instability (MSI-H) [10].

**Tumor immune microenvironment analysis**

CIBERSORT was applied to estimate the relative fractions of 22 infiltrating immune cell types in each tumor sample using R package [11]. The ESTIMATE algorithm was exploited to infer the immune scores for each sample [12].

**Statistical analysis**

Cancer patients were classified into high and low CMTM6 expression subgroups based on the median value of CMTM6 expression. The Wilcoxon test was used to evaluate expression differences between normal and tumor tissues. Overall survival (OS) was calculated using the Kaplan–Meier method, and survival curves were compared using log-rank tests. Spearman rank analysis was performed to evaluate the correlation between CMTM6 expression levels with checkpoint related genes and the tumor microenvironment. All statistical analysis was conducted using R software (version 3.6.1). P value <0.05 was considered statistically significant.

**Results**

**Pan-cancer CMTM6 expression**

CMTM6 mRNA levels in 11,093 tumor and normal tissue samples were analyzed from the TCGA cohort (Table S1). The results revealed that CMTM6 was upregulated in six [breast invasive carcinoma (BLCA), colon adenocarcinoma (COAD), esophageal carcinoma (ESCA), kidney renal papillary cell carcinoma
(KIRP), stomach adenocarcinoma (STAD), and thyroid carcinoma (THCA)) and downregulated in six (cholangiocarcinoma (CHOL), kidney renal clear cell carcinoma (KIRC), liver hepatocellular carcinoma (LIHC), lung adenocarcinoma (LUAD), lung squamous cell carcinoma (LUSC), and pheochromocytoma and paraganglioma (PCPG)) cancer types relative to that in normal tissues (Fig. 1a).

Additionally, we investigated CMTM6 protein expression from the HPA cohort, which presented CMTM6 protein expression in 14 different tumor types. High or medium CMTM6 expression levels were observed in pancreatic adenocarcinoma (PAAD) (75%), LIHC (66.7%), BLCA (63.6%), STAD (63.6%), ovarian serous cystadenocarcinoma (OV) (54.5%), uterine corpus endometrial carcinoma (UCEC) (33.3%), head and neck squamous cell carcinoma (HNSC) (25%), prostate adenocarcinoma (PRAD) (25%), lymphoma (LYMP) (25%), THCA (25%), testicular germ cell tumors (TGCT) (18.2%), breast invasive carcinoma (BRCA) (11.1%), cervical squamous cell carcinoma and endocervical adenocarcinoma (CESC) (10%), and COAD (8.3%) (Fig. 1b-c). These results indicated CMTM6 may play different roles in cancer progression.

**Frequencies of CMTM6 mutations and copy number alterations in multiple cancers**

CMTM6 mutations and copy number alterations were investigated using cBioPortal. The results indicated low CMTM6 mutation frequencies in multiple cancer types, with the highest CMTM6 mutation frequency in uterine cancer. Amplification accounted for 0.49% (46/9,477) of the copy number alterations, while insertions and deletions (indels) comprised 0.32% (30/9,477) (Fig. 2).

**Association between CMTM6 mRNA expression and PD-L1 protein expression**

We investigated the correlation between expression of CMTM6 mRNA and PD-L1 protein expression using PD-L1 data from the TCPA cohort (Table S2). Our results revealed that CMTM6 mRNA expression was associated with PD-L1 protein expression in BRCA, CESC, CHOL, glioblastoma multiforme (GBM), HNSC, KIRP, sarcoma (SARC), STAD, and uterine carcinosarcoma (UCS) (Fig. 3, Fig. S1).

**Correlations among CMTM6 expression, TMB, and MSI**

TMB and MSI have been associated with cancer immunotherapeutic response and prognosis. In this study, we assessed TMB across 33 cancer types using the MuTect2 pipeline and found that TMB was the highest for skin cutaneous melanoma (Fig. S2). We further evaluated the relationship between CMTM6 expression and TMB and showed that CMTM6 expression was correlated with TMB in COAD, ESCA, acute myeloid leukemia (LAML), LIHC, SARC, and STAD (Fig. 4a), while no relationship was observed in the other 27 cancers (Fig. S3). Further, we evaluated the association between CMTM6 expression and MSI status in different tumors. Our results indicated that MSI-H occurred the most frequently in UCEC,
lymphoid neoplasm diffuse large B-cell lymphoma (DLBC), and COAD (Fig. S4), and CMTM6 expression was positively associated with MSI-H in COAD, ESCA, SARC, and STAD. However, CMTM6 expression was negatively correlated with MSI-H in DLBC and OV (Fig. 4b).

**Relationship between CMTM6 expression and tumor immune microenvironment**

We investigated the relationship between CMTM6 expression and immune cell infiltrates in the tumor microenvironment using CIBERSORT. The correlation between CMTM6 expression and tumor-infiltrating immune cells differed for different cancers. Interestingly, we found that high CMTM6 expression was positively associated with neutrophil lymphocyte infiltration in 14 cancer types (Fig. 5a).

We applied the ESTIMATE algorithm to calculate the immune score for each sample in the TCGA cohort. To explore the potential correlation between CMTM6 expression and immune scores, patients were divided into high and low CMTM6 expression groups using the median CMTM6 expression as the cutoff value. CMTM6 expression was positively related to immune score in COAD, DLBC, GBM, HNSC, kidney chromophobe (KICH), KIRC, LAML, brain lower grade glioma (LGG), LIHC, LUAD, LUSC, PCPG, SARC, STAD, THCA, and UCS (Fig. 5b). The results suggested that CMTM6 expression was associated with high immune infiltration in some cancer types.

**Correlations between CMTM6 expression and immune checkpoint-associated genes**

Immune checkpoint-associated genes play an important role in immune escape [13]. We further explored correlations between CMTM6 expression and immune checkpoint-associated genes, including IDO1, LAG3, CTLA4, TNFRSF9, ICOS, CD80, TIGIT, CD70, TNFSF9, ICOSLG, CD86, PDCD1, IDO2, CD276, CD40, HHLA2, CD274, CD27, BTLA, CD28, and HAVCR2 across 33 types of cancer from the TCGA cohort. We found that CMTM6 expression was closely related to almost all immune checkpoint-associated genes except for TIGIT in LGG (Fig. S5). Furthermore, CMTM6 expression was not associated with most immune checkpoint-associated genes in CESC, CHOL, ESCA, KICH, OV, and UCS.

**CMTM6 is a prognostic biomarker in multiple cancers**

The relationship between CMTM6 expression and patient OS was analyzed using the TCGA cohort. High CMTM6 expression was an unfavorable factor for patient OS in adrenocortical carcinoma (ACC) (p = 0.0023), GBM (p = 0.0074), LAML (p = 0.04), LIHC (p = 0.0018), mesothelioma (MESO) (p = 0.0098), SARC (p = 0.0015), thymoma (THYM) (p = 0.019), and uveal melanoma (UVM) (p = 0.0048) (Fig. 6). However, high CMTM6 expression was not associated with prognosis in 25 other cancer types (Fig. S6).
These results suggested that CMTM6 expression may play a promoter role in ACC, GBM, LAML, LIHC, MESO, SARC, THYM, and UVM tumors.

**Discussion**

Immune checkpoint inhibitor therapy has emerged as a critical treatment option in multiple cancer types [14]. However, it is effective in a minority of patients; only 12.6% of all cancer patients benefit from immune checkpoint inhibitors [15]. Therefore, exploration of predictive biomarkers for successful treatment or combined strategies is warranted to increase the therapeutic response rate to immune checkpoint inhibitors. Recently, CMTM6 expression was shown to be correlated with an improved response to PD-1 inhibitors [16]. Given the crucial role of CMTM6 in tumor-related immune responses, we investigated the association between pan-cancer CMTM6 expression patterns and tumor immune microenvironments. In the present study, we analyzed CMTM6 mRNA and protein expression in multiple cancers using TCGA and HPA cohorts, respectively. We found high heterogeneity in the levels of CMTM6 expression in different cancer types.

Previous studies have shown that PD-L1 protein expression is positively associated with response to anti-PD-1 immunotherapy [17-19]. Therefore, fully understanding the regulation mechanism of PD-L1 protein expression is required to improve the efficacy of PD-1/PD-L1 inhibitors. Post-translational regulation is an important mechanism for regulating PD-L1 expression [20]. Two recent studies have confirmed that CMTM6 stabilizes PD-L1 protein expression to attenuate T-cell immune surveillance [5-6]. We, thus, evaluated the correlation between CMTM6 expression and PD-L1 protein expression. PD-L1 expression was positively correlated with CMTM6 expression in CHOL, GBM, HNSC, SARC, and STAD, implying that high CMTM6 expression could respond favorably to anti-PD-1/PD-L1 immunotherapy in these types of tumors.

Increasing evidence supports TMB as a potential biomarker of immune checkpoint inhibitor response in most cancers [21-23]. These studies suggested that a higher burden of nonsynonymous mutations in tumors facilitated the increased formation of neoantigens, making the tumor more immunogenic and, thus, improving the clinical response to immunotherapy. In this study, we evaluated the association between CMTM6 and TMB, revealing that CMTM6 expression was not associated with TMB in most cancer types, except in COAD, ESCA, LAML, LIHC, SARC and STAD. We found that these associations were usually related to cancer type. MSI is caused by the insertion or loss of base pairs in the microsatellite region owing to replication errors. Recent studies have also shown that MSI and/or mismatch-repair deficiency (dMMR) could serve as potential biomarkers and predict the efficacy of immunotherapy, irrespective of cancer type [24]. In patients with advanced dMMR or MSI-H cancers treated with pembrolizumab, Le et al. reported that objective response was observed in 53% of patients and 64% of patients experienced 2-year survival [25]. The targeted monoclonal antibody nivolumab has also demonstrated effectiveness in dMMR or MSI-H colorectal cancer [26]. Based on the importance of microsatellite instability in tumor immunotherapy, we evaluated the MSI status of tumor patients in the TCGA cohort and further analyzed the correlation between MSI-H and CMTM6 expression. We found that
CMTM6 expression was positively related to MSI-H in UCEC, DLBC, and COAD. It is worth emphasizing that high CMTM6 expression in these tumors could identify patients who might respond favorably to PD-1/PD-L1 antibody immunotherapy.

Tumor immune infiltrating cells migrate from blood to tumor tissues and play an important role in immune regulation. Increasing numbers of studies have shown that tumor immune infiltrating cells are closely related to the efficacy of immune checkpoint inhibition and prognosis [27-29]. To elucidate the relationship between CMTM6 expression and diverse infiltrating lymphocytes, we used CIBERSORT to examine the relative fractions of infiltrating immune cell types across 33 cancer types. We found that these associations depended on tumor type. CMTM6 expression was associated with invasive neutrophils in most tumors. Tumor-associated neutrophils are generally considered to be tumor-promoting agents in many tumor types [30]. We speculate that CMTM6 expression may play a role in regulating tumor cells by inducing neutrophil infiltration. Further in vitro and in vivo research is warranted to validate the relationship between CMTM6 expression and neutrophils. Furthermore, we evaluated the immune score of patients with tumor from the TCGA cohort using the ESTIMATE algorithm and found that high CMTM6 expression was associated with higher immune infiltration score in most tumors. This further demonstrated that changes in CMTM6 expression can affect immune cell infiltration in the tumor microenvironment.

Recent studies have shown that CMTM6 plays an oncogenic role and is associated with poor prognosis in gliomas, hepatocellular carcinoma, and LUAD [2,31-32]. However, there is limited information regarding the prognostic value of CMTM6 in other solid cancer types. Our results indicated that high CMTM6 expression was associated with poor clinical prognosis in ACC, GBM, LAML, LIHC, MESO, SARC, THYM, and UVM and that CMTM6 may play a promoting role in tumor progression.

**Conclusions**

In conclusion, we herein report that CMTM6 is heterogeneously expressed in diverse cancers and its expression is correlated with the tumor immune microenvironment and pan-cancer prognosis. High CMTM6 expression was associated with poor prognosis in diverse prevailing cancers. CMTM6, thus, represents a potential target for cancer immunotherapy and a biomarker to predict prognosis.

**List Of Abbreviations**

ACC, adrenocortical carcinoma  
BLCA, bladder urothelial carcinoma  
BRCA, breast invasive carcinoma  
CESC, cervical squamous cell carcinoma and endocervical adenocarcinoma
CHOL, cholangiocarcinoma
CMTM6, CKLF-like MARVEL transmembrane domain-containing 6
COAD, colon adenocarcinoma
DLBC, lymphoid neoplasm diffuse large B-cell lymphoma
ESCA, esophageal carcinoma
GBM, glioblastoma multiforme
HNSC, head and neck squamous cell carcinoma
HPA, Human Protein Atlas
KICH, kidney chromophobe
KIRC, kidney renal clear cell carcinoma
KIRP, kidney renal papillary cell carcinoma
LAML, acute myeloid leukemia
LGG, brain lower grade glioma
LIHC, liver hepatocellular carcinoma
LUAD, lung adenocarcinoma
LUSC, lung squamous cell carcinoma
MESO, mesothelioma
MSI, microsatellite instability
MSI-H, microsatellite high instability
MSS, microsatellite stable
PCPG, pheochromocytoma and paraganglioma
PD-1, programmed death-1
PD-L1, programmed death-ligand 1
SARC, sarcoma
STAD, stomach adenocarcinoma
TCGA, The Cancer Genome Atlas
TCPA, The Cancer Proteome Atlas
THCA, thyroid carcinoma
THYM, thymoma
UCS, uterine carcinosarcoma
UV, uveal melanoma

Declarations

Acknowledgements

We would like to thank Editage (www.editage.cn) for English language editing.

Authors’ contributions

WY and PHH conceived, designed, planned the study, and interpreted the results. ZMH and ZYB analyzed the data. GSY and ZS acquired data. ZMH and ZYB drafted the manuscript. All authors revised and reviewed this work, and gave their final approval of the submitted manuscript.

Funding

This study was supported by the Natural Science Foundation of Heilongjiang Province (Grant No. LH2019H040), the Postdoctoral Research Foundation of China (Grant No. 2018M640309), and the Heilongjiang Provincial Postdoctoral Science Foundation (Grant No. 2018JTC)

Availability of data and materials

The data was available in The Cancer Genome Atlas database (https://portal.gdc.cancer.gov/); Human Protein Atlas (HPA) database (https://www.proteinatlas.org/); The eBio cancer genomics portal (http://cbioportal.org); and The Cancer Proteome Atlas (TCPA) database (https://www.proteinatlas.org/).

Ethics approval and consent to participate

The data we analyzed came from public databases and therefore no additional ethics approval was needed.

Consent for publication

All the authors agreed to publish this article in present form.
Competing interests

All authors declared no conflict of interests.

References

1. Han W, Ding P, Xu M, Wang L, Rui M, Shi S, Liu Y, Zheng Y, Chen Y, Yang T et al: Identification of eight genes encoding chemokine-like factor superfamily members 1-8 (CKLFSF1-8) by in silico cloning and experimental validation. Genomics 2003, 81(6):609-617.

2. Guan X, Zhang C, Zhao J, Sun G, Song Q, Jia W: CMTM6 overexpression is associated with molecular and clinical characteristics of malignancy and predicts poor prognosis in gliomas. EBioMedicine 2018, 35:233-243.

3. Zugazagoitia J, Liu Y, Toki M, McGuire J, Ahmed FS, Henick BS, Gupta R, Gettinger SN, Herbst RS, Schalper KA et al: Quantitative Assessment of CMTM6 in the Tumor Microenvironment and Association with Response to PD-1 Pathway Blockade in Advanced-Stage Non-Small Cell Lung Cancer. Journal of thoracic oncology: official publication of the International Association for the Study of Lung Cancer 2019, 14(12):2084-2096.

4. Jiang Y, Zhao X, Fu J, Wang H: Progress and Challenges in Precise Treatment of Tumors With PD-1/PD-L1 Blockade. Frontiers in immunology 2020, 11:339.

5. Burr ML, Sparbier CE, Chan YC, Williamson JC, Woods K, Beavis PA, Lam EYN, Henderson MA, Bell CC, Stolzenburg S et al: CMTM6 maintains the expression of PD-L1 and regulates anti-tumour immunity. Nature 2017, 549(7670):101-105.

6. Mezzadra R, Sun C, Jae LT, Gomez-Eerland R, de Vries E, Wu W, Logtenberg MEW, Slagter M, Rozeman EA, Hofland I et al: Identification of CMTM6 and CMTM4 as PD-L1 protein regulators. Nature 2017, 549(7670):106-110.

7. Mamessier E, Birnbaum DJ, Finetti P, Birnbaum D, Bertucci F: CMTM6 stabilizes PD-L1 expression and refines its prognostic value in tumors. Annals of translational medicine 2018, 6(3):54.

8. Cerami E, Gao J, Dogrusoz U, Gross BE, Sumer SO, Aksoy BA, Jacobsen A, Byrne CJ, Heuer ML, Larsson E et al: The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer discovery 2012, 2(5):401-404.

9. Cibulskis K, Lawrence MS, Carter SL, Sivachenko A, Jaffe D, Sougnez C, Gabriel S, Meyerson M, Lander ES, Getz G: Sensitive detection of somatic point mutations in impure and heterogeneous cancer samples. Nature biotechnology 2013, 31(3):213-219.

10. Bonneville R, Krook MA, Kautto EA, Miya J, Wing MR, Chen HZ, Reeser JW, Yu L, Roychowdhury S: Landscape of Microsatellite Instability Across 39 Cancer Types. JCO precision oncology 2017, 2017.
11. Newman AM, Liu CL, Green MR, Gentles AJ, Feng W, Xu Y, Hoang CD, Diehn M, Alizadeh AA: Robust enumeration of cell subsets from tissue expression profiles. Nature methods 2015, 12(5):453-457.

12. Yoshihara K, Shahmoradgoli M, Martínez E, Vegesna R, Kim H, Torres-Garcia W, Treviño V, Shen H, Laird PW, Levine DA et al: Inferring tumour purity and stromal and immune cell admixture from expression data. Nature communications 2013, 4:2612.

13. Topalian SL, Drake CG, Pardoll DM: Immune checkpoint blockade: a common denominator approach to cancer therapy. Cancer cell 2015, 27(4):450-461.

14. Vaddepally RK, Kharel P, Pandey R, Garje R, Chandra AB: Review of Indications of FDA-Approved Immune Checkpoint Inhibitors per NCCN Guidelines with the Level of Evidence. Cancers 2020, 12(3).

15. Haslam A, Prasad V: Estimation of the Percentage of US Patients With Cancer Who Are Eligible for and Respond to Checkpoint Inhibitor Immunotherapy Drugs. JAMA network open 2019, 2(5):e192535.

16. Koh YW, Han JH, Haam S, Jung J, Lee HW: Increased CMTM6 can predict the clinical response to PD-1 inhibitors in non-small cell lung cancer patients. Oncoimmunology 2019, 8(10):e1629261.

17. Daud AI, Wolchok JD, Robert C, Hwu WJ, Weber JS, Ribas A, Hodi FS, Joshua AM, Kefferd R, Hersey P et al: Programmed Death-Ligand 1 Expression and Response to the Anti-Programmed Death 1 Antibody Pembrolizumab in Melanoma. Journal of clinical oncology : official journal of the American Society of Clinical Oncology 2016, 34(34):4102-4109.

18. Rittmeyer A, Barlesi F, Waterkamp D, Park K, Ciardiello F, von Pawel J, Gadgeel SM, Hida T, Kowalski DM, Dols MC et al: Atezolizumab versus docetaxel in patients with previously treated non-small-cell lung cancer (OAK): a phase 3, open-label, multicentre randomised controlled trial. Lancet (London, England) 2017, 389(10066):255-265.

19. Abdel-Rahman O: Correlation between PD-L1 expression and outcome of NSCLC patients treated with anti-PD-1/PD-L1 agents: A meta-analysis. Critical reviews in oncology/hematology 2016, 101:75-85.

20. Sun C, Mezzadra R, Schumacher TN: Regulation and Function of the PD-L1 Checkpoint. Immunity 2018, 48(3):434-452.

21. Hellmann MD, Ciuleanu TE, Pluzanski A, Lee JS, Otterson GA, Audigier-Valette C, Minenza E, Linardou H, Burgers S, Salman P et al: Nivolumab plus Ipilimumab in Lung Cancer with a High Tumor Mutational Burden. The New England journal of medicine 2018, 378(22):2093-2104.

22. Yarchoan M, Hopkins A, Jaffee EM: Tumor Mutational Burden and Response Rate to PD-1 Inhibition. The New England journal of medicine 2017, 377(25):2500-2501.

23. Samstein RM, Lee CH, Shoushtari AN, Hellmann MD, Shen R, Janjigian YY, Barron DA, Zehir A, Jordan EJ, Omuro A et al: Tumor mutational load predicts survival after immunotherapy across multiple cancer
24. Zhao P, Li L, Jiang X, Li Q: Mismatch repair deficiency/microsatellite instability-high as a predictor for anti-PD-1/PD-L1 immunotherapy efficacy. Journal of hematology & oncology 2019, 12(1):54.

25. Le DT, Durham JN, Smith KN, Wang H, Bartlett BR, Aulakh LK, Lu S, Kemberling H, Wilt C, Luber BS et al: Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. Science (New York, NY) 2017, 357(6349):409-413.

26. Overman MJ, McDermott R, Leach JL, Lonardi S, Lenz HJ, Morse MA, Desai J, Hill A, Axelsson M, Moss RA et al: Nivolumab in patients with metastatic DNA mismatch repair-deficient or microsatellite instability-high colorectal cancer (CheckMate 142): an open-label, multicentre, phase 2 study. The Lancet Oncology 2017, 18(9):1182-1191.

27. Ascierto PA, Lewis KD, Di Giacomo AM, Demidov L, Mandalà M, Bondarenko I, Herbert C, Mackiewicz A, Rutkowski P, Guminiski A et al: Prognostic impact of baseline tumour immune infiltrate on disease-free survival in patients with completely resected, BRAF(v600) mutation-positive melanoma receiving adjuvant vemurafenib. Annals of oncology : official journal of the European Society for Medical Oncology 2020, 31(1):153-159.

28. Liu X, Xu J, Zhang B, Liu J, Liang C, Meng Q, Hua J, Yu X, Shi S: The reciprocal regulation between host tissue and immune cells in pancreatic ductal adenocarcinoma: new insights and therapeutic implications. Molecular cancer 2019, 18(1):184.

29. Wang SS, Liu W, Ly D, Xu H, Qu L, Zhang L: Tumor-infiltrating B cells: their role and application in anti-tumor immunity in lung cancer. Cellular & molecular immunology 2019, 16(1):6-18.

30. Coffelt SB, Wellenstein MD, de Visser KE: Neutrophils in cancer: neutral no more. Nature reviews Cancer 2016, 16(7):431-446.

31. Zhu X, Qi G, Li C, Bei C, Tan C, Zhang Y, Shi W, Zeng W, Kong J, Fu Y et al: Expression and Clinical Significance of CMTM6 in Hepatocellular Carcinoma. DNA and cell biology 2019, 38(2):193-197.

32. Wang H, Gao J, Zhang R, Li M, Peng Z, Wang H: Molecular and immune characteristics for lung adenocarcinoma patients with CMTM6 overexpression. International immunopharmacology 2020, 83:106478.

Figures
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