Natural-like products as potential SARS-CoV-2 M\text{pro} inhibitors: \textit{in-silico} drug discovery

Mahmoud A. A. Ibrahim\textsuperscript{a}, Khlood A. A. Abdeljawaad\textsuperscript{a}, Alaa H. M. Abdelrahman\textsuperscript{a} and Mohamed-Elamir F. Hegazy\textsuperscript{b}

\textsuperscript{a}Computational Chemistry Laboratory, Chemistry Department, Faculty of Science, Minia University, Minia, Egypt; \textsuperscript{b}Chemistry of Medicinal Plants Department, National Research Centre, El-Tahrir St, Dokki, Giza, Egypt

Communicated by Ramaswamy H. Sarma

\textbf{ABSTRACT}

In December 2019, a COVID-19 epidemic was discovered in Wuhan, China, and since has disseminated around the world impacting human health for millions. Herein, \textit{in-silico} drug discovery approaches have been utilized to identify potential natural products (NPs) as Severe Acute Respiratory Syndrome coronavirus 2 (SARS-CoV-2) main protease (M\text{pro}) inhibitors. The MolPort database that contains over 100,000 NPs was screened and filtered using molecular docking techniques. Based on calculated docking scores, the top 5,000 NPs/natural-like products (NLPs) were selected and subjected to molecular dynamics (MD) simulations followed by molecular mechanics–generalized Born surface area (MM-GBSA) binding energy calculations. Combined 50 ns MD simulations and MM-GBSA calculations revealed nine potent NLPs with binding affinities ($\Delta G_{\text{binding}}$) $>$ 48.0 kcal/mol. Interestingly, among the identified NLPs, four bis([1,3]dioxolo)pyran-5-carboxamide derivatives showed $\Delta G_{\text{binding}}$ $>$ 56.0 kcal/mol, forming essential short hydrogen bonds with HIS163 and GLY143 amino acids via dioxolane oxygen atoms. Structural and energetic analyses over 50 ns MD simulation demonstrated NLP-M\text{pro} complex stability. Drug-likeness predictions revealed the prospects of the identified NLPs as potential drug candidates. The findings are expected to provide a novel contribution to the field of COVID-19 drug discovery.
Introduction

The recent pandemic due to the worldwide spread of COVID-19 caused by a new Severe Acute Respiratory Syndrome coronavirus 2 (SARS-CoV-2) is a great concern for global public health (Lu et al., 2020; Zhu et al., 2020). The WHO declared a global emergency in January 2020, and by March COVID-19 had expanded into a pandemic due to its rapid propagation (Cucinotta & Vanelli, 2020). Since then the U.S. Food and Drug Administration (FDA) has issued an emergency use authorization of remdesivir for treatment of suspected or laboratory-confirmed COVID-19 cases. In addition, clinical trials are being conducted for several potential drugs to treat COVID-19 (Thorlund et al., 2020). Identification of small molecules as potential SARS-CoV-2 inhibitors continues to be a major research focus within the scientific and pharmaceutical communities (Al-Khafaji et al., 2020; Liu et al., 2020; Sanders et al., 2020). The mechanism of SARS-CoV-2 inhibition depends on a main protease (M\text{pro}) that represents a potential target for terminating viral replication (Boopathi et al., 2020). SARS-CoV-2 M\text{pro} with a peptidomimetic inhibitor (N3) has been recently crystallized providing structural information for in-silico studies to screen for novel SARS-CoV-2 inhibitors (Jin et al., 2020).

Natural products (NPs) have been the source of most of the active ingredients of medicines, as over 50% of the approved drugs are based on NPs (Harvey, 2008). Therefore, NPs have hitherto received great attention by researchers that seek to discover potential drugs for the treatment of various diseases such as malaria (Clark, 1996; Wells, 2011), HIV (Kurapati et al., 2015), cardiovascular disease (Mashour et al., 1998), and neoplastic disease (Cragg et al., 1997). Recently, NPs screenings have been reported to identify anti-COVID-19 inhibitors based on NPs/NLPs (Gentile et al., 2020; Gonzalez-Paz et al., 2020; Ul Qamar et al., 2020). The long lead time and high cost of isolating and identifying NPs have prompted investigators to design natural-like products (NLPs) libraries based on small molecules taking into account the stability improvement and bioavailability (Quinn et al., 2008; Thomas & Johannes, 2011). One of the most common enriched databases is MolPort database containing 113,756 NPs/NLPs (https://www.molport.com). The current study set out to screen the MolPort database against potential small molecules that would bind in the SARS-CoV-2 M\text{pro} active site and act as an inhibitor. Based on estimated docking scores, the top 5,000 NPs/NLPs were selected and subjected to molecular dynamics simulations. Drug likeness parameters were predicted for the proposed NPs/NLPs. The obtained results provide implications of new drug development in the prevention or treatment of COVID-19.

Computational methodology

Database preparation

The MolPort database that contains 113,756 natural and natural-like products was downloaded and prepared for molecular docking calculations (https://www.molport.com). The 3D chemical structures of the compounds were generated using Omega2 software (Hawkins et al., 2010; OMEGA, 2013). The geometrical structures of the compounds were then optimized by MMFF94S force field using SZYBKI (SZYBKI, 2016). Gasteiger method was used to assign partial atomic charges (Gasteiger & Marsili, 1980). Prepared files may be accessed at the publicly-available site CompChem DataBase (www.compchem.net/ccdb).

M\text{pro} preparation

The crystal structure of SARS-CoV-2 main protease (M\text{pro}) complexed with peptidomimetic inhibitor (N3) (PDB code: 6LU7 (Jin et al., 2020) was considered as a template for all molecular docking and molecular dynamics calculations. Water molecules and ions were removed. The protonation state of M\text{pro} was examined using H\text{++} server, and all missing hydrogen atoms were added (Gordon et al., 2005). Finally, the pdbqt file for M\text{pro} was prepared according to AutoDock protocol (Forli et al., 2016).

Molecular docking

In this study, two molecular docking calculations were conducted—namely, standard and expensive. In the two calculations, all docking parameters were kept to default values, except the number of genetic algorithm (GA) runs and the maximum number of energy evaluations (eval). The latter two variables were set to 25 and 100, and 2,500,000 and 10,000,000 for standard and expensive molecular docking calculations, respectively. The docking grid was purposed to embrace the active site of the SARS-CoV-2 M\text{pro} receptor with a grid size of 60 Å × 60 Å × 60 Å and a spacing value of 0.375 Å. The grid center was located at −13.069, 9.74, 68.49 (XYZ coordinates). All molecular docking calculations were carried out using Autodock4.2 software (Morris et al., 2009).

Molecular dynamics simulations

Molecular dynamics (MD) simulations were executed on top potent NPs/NLPs in complex with SARS-CoV-2 M\text{pro} using AMBER16 software (Case et al., 2016). In MD simulations, NPs/NLPs and M\text{pro} were described using General AMBER force field (GAFF) and AMBER force field 14SB, respectively (Maier et al., 2015; Wang et al., 2004). Two types of MD simulations were performed—namely, implicit and explicit MDs. In implicit MD, the atomic partial charges of the NPs/NLPs were assigned using am1-bcc method with the help of Antechamber tool implemented inside AMBER16 (Jakalian et al., 2002). A non-bonded cutoff of 999 Å and no periodic boundary conditions were applied. Moreover, the solvation effect was captured with the igb = 1 implicit solvent model (Roux & Simonson, 1999). The docked NP/NLP-M\text{pro} complexes were then minimized for 500 steps and heated from 0 K to 300 K over 10 ps. Finally, 100 ps and 1,000 ps production stages were performed and snapshots were collected every 1 ps, giving 100 and 1,000 snapshots, respectively.
Table 1. Calculated standard and expansive docking scores (in kcal/mol) and binding features for nine potent natural-like products (NLPs) against SARS-CoV-2 main protease (Mpro).

| No. | MolPort code  | 2D-chemical structure | Docking score (kcal/mol) | Binding features (Hydrogen bond length in Å) |
|-----|---------------|------------------------|--------------------------|--------------------------------------------|
| 1   | MolPort-000-708-794 | ![Chemical Structure](image1.png) | -10.4  | -11.0  | GLU166 (1.80 Å), GLY143 (1.72 Å), HIS163 (2.35 Å), HIS164 (1.83 Å) |
| 2   | MolPort-044-179-844 | ![Chemical Structure](image2.png) | -10.0  | -10.9  | GLN189 (1.80 Å), THR190 (2.39 Å), GLN192 (2.45 Å), HIS163 (2.07 Å) |
| 3   | MolPort-000-702-646 | ![Chemical Structure](image3.png) | -10.4  | -10.8  | HIS163 (2.26 Å), HIS164 (2.05 Å), THR190 (1.66 Å), GLN192 (2.84 Å), GLN189 (2.94 Å), GLY143 (1.95 Å) |
| 4   | MolPort-002-513-915 | ![Chemical Structure](image4.png) | -9.6   | -10.5  | HIS164 (2.15 Å), GLN192 (2.38 Å), GLN189 (2.55 Å), GLY143 (2.09 Å), HIS163 (2.57 Å) |
| 5   | MolPort-005-948-349 | ![Chemical Structure](image5.png) | -10.2  | -10.4  | GLN189 (2.10, 1.89 Å), ASN142 (2.15 Å), GLU166 (2.85 Å) |
| 6   | MolPort-039-056-062 | ![Chemical Structure](image6.png) | -10.0  | -10.4  | GLN192 (2.24 Å), THR190 (1.97 Å), GLU166 (2.04 Å), GLN189 (2.18, 2.18 Å), ASN142 (2.54 Å) |
| 7   | MolPort-004-849-765 | ![Chemical Structure](image7.png) | -9.8   | -10.4  | HIS164 (2.07 Å), GLN189 (2.51 Å), GLY143 (1.97 Å), HIS163 (2.44 Å) |

(continued)
implicit MD simulations were performed with the CPU version of pmemd (pmemd.MPI) in AMBER16. In explicit MD, the atomic partial charges of the studied NPs/NLPs were assigned using the restrained electrostatic potential (RESP) approach at the HF/6-31G* level with the help of Gaussian09 software (Bayly et al., 1993; Frisch et al., 2009). The docked NP/NLP-Mpro complexes were solvated in a cubic water box with 15 Å distances between the edges of the box and any atom of NP/NLP or NP/NLP-Mpro complexes. The solvated NP/NLP-Mpro complexes were then minimized for 5000 steps and subsequently gently annealed from 0 K to 300 K over 50 ps. The systems were equilibrated for 1 ns and production stages were conducted over simulation times of 10 and 50 ns. The snapshots were collected every 10 ps, giving 1,000 and 5,000 snapshots, respectively. Periodic boundary conditions and the NPT ensemble were adopted in all explicit MD simulations, including both the equilibration and production stages. Long-range electrostatic interactions under periodic conditions with a direct space cut-off of 12 Å were treated with Particle Mesh Ewald (PME) method (Darden et al., 1993). Langevin dynamics with the collision frequency $\gamma_{\text{ln}}$ set to 1.0 was used to keep the temperature constant at 298 K. Berendsen barostat with a relaxation time of 2 ps was employed to control the pressure of system (Berendsen et al., 1984). A time step of 2 fs and the SHAKE option to constrain all bonds involving hydrogen atoms were used. All explicit MD simulations were performed with the GPU version of pmemd.cuda in AMBER16.

**MM-GBSA binding energy**

Snapshots were collected every 10 ps over the production stage of 50 ns MD simulation, giving 5,000 snapshots. Based on the collected snapshots, molecular mechanics–generalized Born surface area (MM-GBSA) approach was used to evaluate the binding energy of the studied NPs/NLPs with SARS-CoV-2 Mpro (Massova & Kollman, 2000). The MM-GBSA binding free energies were estimated as follows:

$$\Delta G_{\text{binding}} = G_{\text{Complex}} - (G_{\text{NP/NLP}} + G_{\text{Mpro}})$$

where the energy term $G$ is estimated as:

$$G = E_{\text{vdw}} + E_{\text{ele}} + G_{\text{GB}} + G_{\text{SA}}$$

with $E_{\text{vdw}}$, $E_{\text{ele}}$, $G_{\text{GB}}$ and $G_{\text{SA}}$ as the van der Waals, electrostatic, General Born solvation, and surface area energies, respectively. For the NPs/NLPs, entropy contributions were neglected.

**Drug likeness**

The online Molinspiration cheminformatics software (http://www.molinspiration.com) was utilized to evaluate the physicochemical parameters of the identified potent NPs and NLPs. Lipinski’s parameters including topological polar surface area (TPSA), relative molecular weight (MW), number of hydrogen bond donors (nOHNH), number of hydrogen bond acceptors (nON) and the partition coefficient log P (miLog P) were assessed.

**Molecular target prediction and network analysis**

Based on the structural similarity of known ligands-target integrations, target prediction of the most promising NPs/NLPs was performed using the online website-based tools of SwissTargetPrediction (http://www.swisstargetprediction.ch). Additionally, the available database for Severe Acute Respiratory Syndrome diseases was collected from the online database DisGeNET (https://www.disgenet.org). Venn diagram was plotted using InteractiVenn online tool (Heberle et al., 2015). Protein-protein interaction (PPI) network was generated using a functional database of STRING for top predicted targets (Li et al., 2019). The PPI network was diagramed and analyzed using Cytoscape 3.8.0 (Shannon et al., 2003).

### Table 1. Continued.

| No. | MolPort code  | 2D-chemical structure | Docking score (kcal/mol) | Binding features (Hydrogen bond length in Å) |
|-----|---------------|------------------------|--------------------------|---------------------------------------------|
| 8   | MolPort-046-158-375 | ![2D-chemical structure](image) | -9.7 | PHE140 (1.68, 2.34 Å), GLU166 (2.38 Å), CYS145 (2.51 Å), HIS163 (2.85, 1.78 Å) |
| 9   | MolPort-001-751-850 | ![2D-chemical structure](image) | -9.6 | GLN192 (2.14 Å), HIS163 (2.10 Å), ASN142 (1.94 Å), GLU166 (2.09 Å), CYS145 (2.35 Å), HIS541 (2.84 Å) |
Results and discussion

Unavailability of therapies as yet indicates an urgent need for drug exploration against SARS-CoV-2. The main protease (Mpro) is a likely targeted for enzyme inhibition due to the polypeptides’ essential role in viral replication. In this study, the expansive MolPort database for natural products (NPs) and natural-like products (NLPs) was screened and filtered for the identification of potential SARS-CoV-2 Mpro inhibitors.

Molecular docking

Two levels of molecular docking calculations were carried out to minimize computational-costs and time. Initially, all NPs/NLPs in the MolPort database were screened against Mpro with standard docking parameters of $GA = 25$ and $eval = 2,500,000$. The NPs/NLPs were then sorted based on their docking scores. In terms of the calculated docking scores, almost three-fourths of the screened NPs and NLPs ($\sim 78\%$) showed binding energies less than $-8.7$ kcal/mol with SARS-CoV-2 Mpro. Therefore, only the top 25,000 NPs/NLPs were selected for further docking calculations. Using expensive docking parameters of $GA = 100$ and $eval = 10,000,000$, the selected 25,000 NPs/NLPs were re-docked against Mpro. Docking scores and binding features, as well as 2D chemical structures, of nine potent NLPs with SARS-CoV-2 Mpro are listed in Table 1. The 2D and 3D representations of interactions of the top nine potent NLPs with important

Table 2. Decomposition of MM-GBSA binding energies for four dioxolo-derivatives in complex with SARS-CoV-2 main protease (Mpro) through 50 ns MD simulations.

| MolPort code  | Calculated MM-GBSA binding energy (kcal/mol) |
|---------------|-----------------------------------------------|
|               | $\Delta_{E_{COV}}$ | $\Delta_{E_{vdw}}$ | $\Delta_{E_{GB}}$ | $\Delta_{G_{SUR}}$ | $\Delta_{G_{dip}}$ | $\Delta_{G_{GB}}$ | $\Delta_{G_{SUR}}$ | $\Delta_{G_{gas}}$ | $\Delta_{G_{Solv}}$ | $\Delta_{G_{binding}}$ |
| MolPort-004-849-765 | $-62.4$ | $-35.1$ | $45.7$ | $-6.6$ | $-97.5$ | $39.1$ | $-58.4$ |
| MolPort-000-708-794 | $-63.7$ | $-47.1$ | $60.4$ | $-6.8$ | $-110.8$ | $53.6$ | $-57.3$ |
| MolPort-002-513-915 | $-63.0$ | $-34.5$ | $47.6$ | $-6.7$ | $-97.5$ | $40.9$ | $-56.6$ |
| MolPort-000-702-646 | $-63.1$ | $-43.6$ | $57.4$ | $-6.9$ | $-105.7$ | $50.5$ | $-56.2$ |

Table 2. Decomposition of MM-GBSA binding energies for four dioxolo-derivatives in complex with SARS-CoV-2 main protease (Mpro) through 50 ns MD simulations.

Figure 1. 2D and 3D representation of predicted binding mode of MolPort-000-708-794 as potent natural-like products (NLPs) inside the active site of SARS-CoV-2 main protease (Mpro).
amino acid residues of SARS-CoV-2 M<sub>pro</sub> are depicted in Supporting Information Figure S1.

The most promising NLPs (shown in Table 1 and Supporting Information Figure S1) share the same binding mode inside the active site of SARS-CoV-2 M<sub>pro</sub>, forming essential hydrogen bonds with key amino acid residues including HIS164, HIS163, and GLU166. Further interactions including van der Waals and hydrophobic interactions were identified, giving a docking score higher than $-9.6$ kcal/mol (Supporting Information Figure S1). For instance, the potency of MolPort-000-708-794 with a binding energy of $-11.0$ kcal/mol may be attributed to its ability to form four hydrogen bonds with GLU166, GLY143, HIS163, and HIS164 with bond lengths of 1.80, 1.72, 2.35 and 1.83 Å, respectively (Figure 1). Two out of the latter four hydrogen bonds were formed via dioxolane oxygen atoms, revealing a significant contribution of dioxolane rings in NLPs-<sup>M<sub>pro</sub></sup> binding mode and affinity. Based on calculated expensive docking scores, the top 5,000 potent NPs/NLPs were closely investigated using molecular dynamics (MD) calculations.

### Molecular dynamics simulations

Conformational flexibilities of drug-receptor complexes, solvent effects, and dynamics must be considered to achieve reliable drug-receptor binding affinities (De Vivo et al., 2016; Kerrigan, 2013). Therefore, molecular dynamics (MD) simulations combined with binding energy calculations over reasonable simulation time were performed for the top 5,000 potent NPs/NLPs in complex with SARS-CoV-2 M<sub>pro</sub>. Considering computational-costs and time, MD simulations for the 5,000 docked NPs/NLPs-M<sub>pro</sub> complexes were conducted in implicit solvent for 100 ps and the corresponding binding energies were estimated using MM-GBSA approach (see Computational Methodology section for details). The

| MolPort code | miLog P<sup>a</sup> | TPSA<sup>b</sup> | nON<sup>c</sup> | nOHNH<sup>d</sup> | nviolation | Nrotb<sup>e</sup> | MolVol<sup>f</sup> | MWt<sup>g</sup> | %ABS<sup>h</sup> |
|-------------|-----------------|----------------|-------------|-----------------|-------------|----------------|----------------|----------------|-------------|
| MolPort-004-849-765 | 2.38 | 104.37 | 9 | 2 | 0 | 4 | 393.88 | 499.36 | 72.9% |
| MolPort-000-708-794 | 1.43 | 163.43 | 13 | 3 | 2 | 7 | 461.20 | 568.63 | 52.6% |
| MolPort-002-513-915 | 2.12 | 130.67 | 11 | 2 | 1 | 7 | 437.32 | 492.52 | 63.9% |
| MolPort-000-702-646 | 1.48 | 163.43 | 13 | 3 | 2 | 7 | 470.49 | 562.60 | 52.6% |

<sup>a</sup>Logarithm of partition coefficient between n-octanol and water (miLogP).<br>
<sup>b</sup>Topological polar surface area (TPSA).<br>
<sup>c</sup>Number of hydrogen bond acceptors (nON).<br>
<sup>d</sup>Number of hydrogen bond donors (nOHNH).<br>
<sup>e</sup>Number of rotatable bonds (nrotb).<br>
<sup>f</sup>Molecular volume (mol).<br>
<sup>g</sup>Molecular weight (MWt).<br>
<sup>h</sup>%ABS $= 100 - 0.345 \times$ TPSA (Zhao et al., 2002).
calculated MM-GBSA binding energies of the top 1,000 compounds are listed in Supporting Information Table S1.

Interestingly, of the 5,000 investigated NPs/NLPs, only 86 NPs/NLPs showed significant MM-GBSA binding energies with values in range $-45.0$ to $-56.3$ kcal/mol. To achieve a greater degree of accuracy, molecular dynamics of 86 NPs/NLPs in complex with $M^{\text{pro}}$ were further investigated over a longer implicit MD simulation with a time of 1 ns. Estimated MD/MM-GBSA binding energies are presented in Supporting Information Table S2. The corresponding MM-GBSA binding energies for nine potent NLPs are presented in Figure 2.

On the basis of the calculated MM-GBSA binding energies over 1 ns (Supporting Information Table S2), 46 NPs/NLPs exhibited instability in molecular dynamics and their corresponding MM-GBSA binding energies with $M^{\text{pro}}$ were decreased to be less than $-45.0$ kcal/mol, while the other 40 NPs/NLPs showed significant binding affinities with MM-GBSA energies $\geq -45.0$ kcal/mol. Therefore, we focused on these potent 40 NPs/NLPs over a reasonable MD simulation time in explicit solvent. Consequently, 10 ns MD simulations for the potent 40 NPs/NLPs-$M^{\text{pro}}$ complexes were conducted in explicit solvent and followed by MM-GBSA binding energies calculation. The estimated 10 ns MD/MM-GBSA binding energies are noted in Supporting Information Table S3, and the corresponding binding energies for nine potent NLPs are presented in Figure 2.

Interestingly, only nine of the investigated NPs/NLPs showed promising binding affinities $> -50.0$ kcal/mol towards SARS-CoV-2 $M^{\text{pro}}$ over 10 ns MD simulation time (Supporting Information Table S3). To increase the reliability of the observed results, each one of these nine NLPs in complex with $M^{\text{pro}}$ was further simulated for 50 ns MD, and the corresponding binding energies were calculated (Figure 2).

As can be seen from Figure 2, there was no significant difference between the estimated MM-GBSA binding energies over 10 ns MD and those over 50 ns MD, reflecting the tight binding of the identified NLPs with SARS-CoV-2 $M^{\text{pro}}$. Surprisingly, four of the nine NLPs—namely, MolPort-004-849-765, MolPort-000-708-794, MolPort-002-513-915 and MolPort-000-702-646—are bis([1,3]dioxolo)pyran-5-carboxamide derivatives and showed outperformance affinity towards SARS-CoV-2 $M^{\text{pro}}$ with binding energies $> -56.0$ kcal/mol. Therefore, considerable interest was given in the following sections to investigate the identified dioxolo-derivatives as potential SARS-CoV-2 $M^{\text{pro}}$ inhibitors.

**Post-dynamics analyses**

The purpose of the post-dynamics analyses was to evaluate the interaction nature and stability of the identified dioxolo-derivatives inside the SARS-CoV-2 $M^{\text{pro}}$ active site. Structural and energetic analyses for the four promising dioxolo-derivatives were conducted over 50 ns explicit MD simulations.

**Binding energy decomposition**

Decomposition of average MM-GBSA binding energy over 50 ns MD simulation was performed to reveal the nature of dominant interactions in NLPs-$M^{\text{pro}}$ complexes (Table 2). Energy decomposition results showed that $E_{\text{vdw}}$ was the
dominant force in NP/NLP-Mpro binding affinities with a contribution value of $-63.0 \text{ kcal/mol}$ for the four dioxolo-derivatives. $E_{\text{ele}}$ was favorable with values of $-35.1$, $-47.1$, $-34.5$ and $-43.6 \text{ kcal/mol}$ for MolPort-004-849-765, MolPort-000-708-794, MolPort-002-513-915 and MolPort-000-702-646, respectively.

Figure 4. Hydrogen bond lengths and center-of-mass (CoM) distances between MolPort-004-849-765 (in cyan), MolPort-000-708-794 (in red), MolPort-002-513-915 (in blue) and MolPort-000-702-646 (in black) with the HIS164 amino acid residue inside the active site of SARS-CoV-2 main protease (Mpro) over the 50 ns MD simulation.
The stability of dioxolo-derivatives inside the M\textsuperscript{pro} active site was investigated using the correlation between the binding energy and time. Therefore, MM-GBSA binding energy was estimated per-frame for each of MolPort-004-849-765, MolPort-000-708-794, MolPort-002-513-915, and MolPort-000-702-646 with M\textsuperscript{pro} and presented in Figure 3. According to data in Figure 3, there was overall stability for the four dioxolo-derivatives till the end of the simulations with average values of $-58.4$, $-57.3$, $-56.6$, and $-56.2$ (kcal/mol), respectively. These findings indicated the promising stability of NLP-M\textsuperscript{pro} complexes over the simulated MD time of 50 ns.

**Binding energy per frame**

Inspecting the hydrogen bond length and center-of-mass (CoM) distance between the dioxolo-derivatives and the key HIS164 amino acid residue over the 50 ns MD simulation would reflect an indication of NLPs-M\textsuperscript{pro} stability. Therefore, the desired hydrogen bond lengths and CoM distances were measured over the 50 ns MD simulations and depicted in Figure 4. The most obvious finding to emerge from data plotted in Figure 4 was that MolPort-004-849-765, MolPort-000-708-794, MolPort-002-513-915 and MolPort-000-702-646 showed high stability inside the active site with average hydrogen bond lengths of 1.93, 1.98, 1.92, and 1.99 Å, respectively. In terms of the measured CoM distances, the average CoM distance between NLP and HIS164 was nearly constant around 8 Å during 50 ns MD simulations for the four investigated dioxolo-derivatives.

**Root-mean-square deviation**

Root-mean-square deviation (RMSD) was used to investigate the structural changes in the NLP-M\textsuperscript{pro} complexes. RMSD for the backbone atoms of the MolPort-004-849-765, MolPort-000-708-794, MolPort-002-513-915, and MolPort-000-702-646 in complex with M\textsuperscript{pro} relative to the starting structures throughout the 50 ns MD simulations were evaluated and presented in Figure 5. From the data in Figure 5, it is apparent that the backbone of NLP-M\textsuperscript{pro} complexes exhibited stability over 50 ns MD simulation, giving RMSD with less than 0.35 nm. These results emphasize that four investigated dioxolo-derivatives are tightly bonded in the active site and do not impact the overall topology of M\textsuperscript{pro}. Finally, these energetic and structural analyses demonstrated the high stability of the four investigated dioxolo-derivatives-M\textsuperscript{pro} complexes through 50 ns MD simulations.

**Drug likeness**

Lipinski’s rule of five is commonly used in drug discovery and development to evaluate the oral bioavailability of active drug in humans. In this study, physicochemical parameters of the promising NLPs as potential SARS-CoV-2 M\textsuperscript{pro} inhibitors were predicted using Molinspiration cheminformatics.
online software calculation toolkit. The predicted parameters included Lipinski’s parameters, topological polar surface area (TPSA), and percentage of absorption (%ABS). The predicted parameters are listed in Table 3.

From data given in Table 3, the milog P values of the four dioxolo-derivatives were found to be below five (calc. in range 1.43 to 2.38), suggesting that these NLPs have good permeability across the cell membrane. Molecular weight was found to be more or less than 500 (calc. in range 492.52 to 568.63), predicting the compounds to be easily transported, diffused and absorbed. Besides, the number of hydrogen bond donors (nOHNH) was less than 5 in accordance with Lipinski’s rules, and the number of hydrogen bond acceptors (nON) was in range 9 to 13. It is worth mentioning that this slight increase in molecular weight and hydrogen bond acceptors will not have a significant impact on compound transportation and diffusion, where it has been shown that several FDA-approved drugs moved beyond the traditional low molecular weight of 500 and hydrogen bond acceptors of 10 (Mullard, 2018). Besides, TPSA of all promising dioxolo-derivatives was observed in range 104.37 to 163.43 Å which was a very good indicator of the bioavailability of the investigated NLPs. In addition, the calculated %ABS was ranged between 52.62% and 72.99%, indicating that the investigated NLPs may have good cell membrane permeability and oral bioavailability.

**Molecular target prediction and network analysis**

Severe Acute Respiratory Syndrome diseases (C1175175) displayed 117 genes based on DisGeNET online software. Additionally, targeted genes for the identified dioxolo-derivatives as potent Mpro inhibitors were collected using online SwissTargetPrediction tools, giving 100 targets. Classification of the predicted targets for each examined dioxolo-derivative is depicted in Supporting Information Figure S2. The Venn diagram comparison analysis between Severe Acute Respiratory Syndrome diseases and predicted targeted genes was demonstrated and plotted in Figure 6.

According to data presented in Venn diagram (Figure 6), Acute Respiratory Syndrome diseases and predicted targeted genes displayed commonly shared MAPK14 and EGFR (Figure 6). The host protein angiotensin-converting enzyme 2 (ACE2), serves as an entry receptor for SARS-CoV-2. Targeting MAPK14 would result in blocking of ACE2 production pathways, and in turn, reducing the probability of SARS-CoV-2 virus to be received and internalized by human cells.
In addition, SARS-CoV-2 inhibition might be achieved by inhibition of MAPks which are activated also by GFRs, such as in case of chloroquine (Hondermarck et al., 2020).

The possible targets predicted by SwissTargetPrediction for all examined dioxolo-derivatives were further analyzed using STRING PPI network and visualized by Cytoscape 3.8.0. The network topological analysis by Cytoscape demonstrated that the targets within the top 10 scores of degree were MAPK14 and EGFR for the four examined dioxolo-derivatives (Supporting Information Table S4). The STRING PPI network for the identified top 10 targets is depicted in Figure 7.

**Conclusion**

SARS-CoV-2 main protease (Mpro) is characterized to be an essential and highly potent target for the inhibition of the novel coronavirus. In this study, a total of 113,756 natural and natural like products were screened against Mpro to discover potential SARS-CoV-2 Mpro inhibitors. Filtration of MolPort database was carried out using combined molecular docking and molecular dynamics (MD) followed by molecular mechanics–generalized Born surface area (MM-GBSA) binding energy calculations. Based on docking scores and MM-GBSA binding energies, nine NLPs showed promising binding affinities >-50.0 kcal/mol with SARS-CoV-2 Mpro. Interestingly, four of the nine NLPs—namely, MolPort-004-849-765, MolPort-000-708-794, MolPort-002-513-915 and MolPort-000-702-646—are bis([1,3]dioxolo)pyran-5-carboxamide derivatives showing high affinity towards Mpro with binding energies >-56.0 kcal/mol. Post-dynamics analyses demonstrated the stability and affinity of the identified dioxolo-derivatives with Mpro. Predicted physicochemical parameters of the promising dioxolo-derivatives fit drug-likeness properties, indicating the probability of these NLPs as prospective SARS-CoV-2 drug candidates.

Protein-protein interaction (PPI) showed the linked top targets genes that could have an effect on viral infection, as well as, the host. The current results establish that bis([1,3]dioxolo)pyran-5-carboxamide derivatives hold promise as inhibitors against SARS-CoV-2 Mpro and are ready for in vitro inhibition against SARS-CoV-2.

**Disclosure statement**

No potential conflict of interest was reported by the authors.

**Funding**

The computational work was completed with resources supported by the Science and Technology Development Fund, STDF, Egypt, Grants No. 5480 & 7972.
Hydrogen bonding patterns in macromolecules. Nucleic Acids Research, 33(Web Server issue), W368–W371. https://doi.org/10.1093/nar/gki464

Harvey, A. L. (2008). Natural products in drug discovery. Drug Discovery Today, 13(19–20), 894–901. https://doi.org/10.1016/j.drudis.2008.07.004

Hawkins, P. G., Skillman, A. G., Warren, G. L., Ellington, B. A., & Stahl, M. T. (2010). Conformer generation with OMEGA: Algorithm and validation using high quality structure frameworks from the Protein Databank and Cambridge Structural Database. Journal of Chemical Information and Modeling, 50(4), 572–584. https://doi.org/10.1021/ji100031x

Heberle, H., Meirelles, G. V., da Silva, F. R., Telles, G. P., & Minghim, R. (2015). InteractiveVenn: A web-based tool for the analysis of sets through Venn diagrams. BMC Bioinformatics, 16(1), 169. https://doi.org/10.1186/s12859-015-0611-3

Hondermark, H., Bartlett, N. W., & Nurcombe, V. (2020). The role of growth factor receptors in viral infections: An opportunity for drug repurposing against emerging viral diseases such as COVID-19? FASEB Bioadvances, 2(5), 296–303. https://doi.org/10.1029/2020FS001571

Jalalian, A., Jack, D. B., & Bayly, C. I. (2002). Fast, efficient generation of high-quality atomic charges. AM1-BCC model: II. Parameterization and validation. Journal of Computational and Structural Chemistry, 23(16), 1623–1641. https://doi.org/10.1121/1.4793218

Jin, Z., Du, X., Xu, Y., Deng, Y., Liu, M., Zhao, Y., Zhang, B., Li, X., Zhang, L., Peng, C., Duan, Y., Yu, J., Wang, L., Yang, K., Liu, F., Jiang, R., Yang, X., You, T., Liu, X., … Yang, H. (2020). Structure of Mpro from SARS-CoV-2 and discovery of its inhibitors. Nature, 582(7811), 289–293. https://doi.org/10.1038/s41586-020-2223-y

Kerrigan, J. E. (2013). Molecular dynamics simulations in drug design. In S. Cortagere (Ed.), In silico models for drug discovery (pp. 95–113). Humana Press.

Kindrachuk, J., Ork, B., Hart, B. J., Mazur, S., Holbrook, M. R., Frieman, M. B., Traynor, D., Johnson, R. F., Dyall, J., Kuhn, J. H., Olinger, G. G., Hensley, L. E., & Jahrling, P. B. (2015). Antiviral potential of ERK/MAPK and PI3K/AKT/mTOR signaling modulation for middle east respiratory syndrome coronavirus infection as identified by temporal kinome analysis. Antimicrobial Agents and Chemistry, 59(2), 1088–1099. https://doi.org/10.1128/AAC.03659-14

Kurapati, K. R. V., Atturi, V. S., Samikkanuru, T., Garcia, G., & Nair, M. P. N. (2015). Natural products as anti-HIV agents and role in HIV-associated neurocognitive disorders (HAND): A brief overview. Frontiers in Microbiology, 6, 1444–1457. https://doi.org/10.3389/fmicb.2015.01444

Li, R., Ma, X., Song, Y., Zhang, Y., Xiong, W., Li, L., & Zhou, L. (2019). Anti-colorectal cancer targets of resveratrol and biological molecular mechanism: Analyses of network pharmacology, human and experimental data. Journal of Cellular Biochemistry, 120(7), 11265–11273. https://doi.org/10.1002/jcb.28404

Liu, C., Zhou, Q., Li, Y., Garner, L. V., Watkins, S. P., Carter, L. J., Smoot, J., Gregg, A. C., Daniels, A. D., Jervey, S., & Albaia, D. (2020). Research and development on therapeutic agents and vaccines for COVID-19 and related human coronavirus diseases. ACS Central Science, 6(3), 315–331. https://doi.org/10.1021/acscentsci.0c00272

Lu, R., Zhao, X., Li, J., Niu, P., Yang, B., Wu, H., Wang, W., Song, H., Huang, B., Zhu, N., Bi, Y., Ma, X., Zhan, F., Wang, L., Hu, T., Zhou, H., Hu, Z., Zhou, W., Zhao, L., … Tan, W. (2020). Genomic characterisation and epidemiology of 2019 novel coronavirus: Implications for virus origins and receptor binding. Lancet (London, England), 395(10224), 565–574. https://doi.org/10.1016/S0140-6736(20)30251-8

Maier, J. A., Martinez, C., Kasavajhala, K., Wickstrom, L., Hauser, K. E., & Simmerling, C. (2015). ff14SB: Improving the Accuracy of Protein Side Chain Parameters from ff99SB. Journal of Chemical Theory and Computation, 11(8), 3696–3713. https://doi.org/10.1021/acs.jctc.5b00255

Masuhr, N. H., Lin, G. I., & Frishman, W. H. (1998). Herbal medicine for the treatment of cardiovascular disease: Clinical considerations. Archives of Internal Medicine, 158(20), 2225–2234. https://doi.org/10.1001/archinte.158.20.2225

Massova, I., & Kollman, P. A. (2000). Combined molecular mechanical and continuum solvent approach (MM-PBSA/GBSA) to predict ligand binding. Perspectives in Drug Discovery and Design, 18(1), 113–135. https://doi.org/10.1021/acs.jctc.000031x

Morris, G. M., Huey, R., Lindstrom, W., Sanner, M. F., Belew, R. K., Goodsell, D. S., & Olson, A. J. (2009). AutoDock4 and AutoDockTools4: Automated
docking with selective receptor flexibility. Journal of Computational Chemistry, 30(16), 2785–2791. https://doi.org/10.1002/jcc.21256
Mullard, A. (2018). Re-assessing the rule of 5, two decades on. Nature Reviews. Drug Discovery, 17(11), 777. https://doi.org/10.1038/nrd.2018.197
OMEGA (Version 2.5.1.4). (2013). Santa Fe, NM: OpenEye Scientific Software.
Quinn, R. J., Carroll, A. R., Pham, N. B., Baron, P., Palframan, M. E., Suraweera, L., Pierens, G. K., & Muresan, S. (2008). Developing a drug-like natural product library. Journal of Natural Products, 71(3), 464–468. https://doi.org/10.1021/np070526y
Roux, B., & Simonson, T. (1999). Implicit solvent models. Biophysical Chemistry, 78(1–2), 1–20. https://doi.org/10.1016/S0301-4622(98)00226-9
Sanders, J. M., Monogue, M. L., Jodlowski, T. Z., & Cutrell, J. B. (2020). Pharmacologic treatments for coronavirus disease 2019 (COVID-19): A review. JAMA, 323(18), 1824–1836. https://doi.org/10.1001/jama.2020.6019
Shannon, P., Markiel, A., Ozier, O., Baliga, N. S., Wang, J. T., Ramage, D., Amin, N., Schwikowski, B., & Ideker, T. (2003). Cytoscape: A software environment for integrated models of biomolecular interaction networks. Genome Research, 13(11), 2498–2504. https://doi.org/10.1101/gr.1239303
SZYBKI (Version 1.9.0.3). (2016). Santa Fe, NM: OpenEye Scientific Software.
Thomas, G. L., & Johannes, C. W. (2011). Natural product-like synthetic libraries. Current Opinion in Chemical Biology, 15(4), 516–522. https://doi.org/10.1016/j.cbpa.2011.05.022
Thorlund, K., Dron, J., Park, J., Hsu, G., Forrest, J. I., & Mills, E. J. (2020). A real-time dashboard of clinical trials for COVID-19. The Lancet Digital Health, 2(6), e286–e287. https://doi.org/10.1016/S2589-7500(20)30086-8
Ul Qamar, T., Alqahtani, S. M., Alamri, M. A., & Chen, L.-L. (2020). Structural basis of SARS-CoV-2 3CL(pro) and anti-COVID-19 drug discovery from medicinal plants. Journal of Pharmaceutical Analysis. https://doi.org/10.1016/j.pha.2020.03.009
Wang, J., Wolf, R. M., Caldwell, J. W., Kollman, P. A., & Case, D. A. (2004). Development and testing of a general amber force field. Journal of Computational Chemistry, 25(9), 1157–1174. https://doi.org/10.1002/jcc.20035
Wells, T. N. (2011). Natural products as starting points for future antimalarial therapies: Going back to our roots? Malaria Journal, 10 Suppl 1(1), S3. https://doi.org/10.1186/1475-2875-10-S1-S3
Zhao, Y. H., Abraham, M. H., Le, J., Hersey, A., Luscombe, C. N., Beck, G., Sherborne, B., & Cooper, I. (2002). Rate-limited steps of human oral absorption and QSAR studies. Pharmaceutical Research, 19(10), 1446–1457. https://doi.org/10.1023/A:1020444330011
Zhu, N., Zhang, D., Wang, W., Li, X., Yang, B., Song, J., Zhao, X., Huang, B., Shi, W., Lu, R., Niu, P., Zhan, F., Ma, X., Wang, D., Xu, W., Wu, G., Gao, G. F., Tan, W., & China Novel Coronavirus Investigating and Research Team. (2020). A Novel Coronavirus from patients with pneumonia in China, 2019. New England Journal of Medicine, 382(8), 727–733. https://doi.org/10.1056/NEJMoa2001017