Human-induced salinization caused by the use of road deicing salts, agricultural practices, mining operations, and climate change is a major threat to the biodiversity and functioning of freshwater ecosystems. Yet, it is unclear if freshwater ecosystems are protected from salinization by current water quality guidelines. Leveraging an experimental network of land-based and in-lake mesocosms across North America and Europe, we tested how salinization—indicated as elevated chloride (Cl$^-\$) concentration—will affect lake food webs and if two of the lowest Cl$^-\$ thresholds found globally are sufficient to protect these food webs. Our results indicated that salinization will cause substantial zooplankton mortality at the lowest Cl$^-\$ thresholds established in Canada (120 mg Cl$^-\$/L and the United States (230 mg Cl$^-\$/L) and throughout Europe where Cl$^-\$ thresholds are generally higher. For instance, at 73% of our study sites, Cl$^-\$ concentrations that caused a ≥50% reduction in cladoceran abundance were at or below Cl$^-\$ thresholds in Canada, in the United States, and throughout Europe. Similar trends occurred for copepod and rotifer zooplankton. The loss of zooplankton triggered a cascading effect causing an increase in phytoplankton biomass at 47% of study sites. Such changes in lake food webs could alter nutrient cycling and water clarity and trigger declines in fish production. Current Cl$^-\$ thresholds across North America and Europe clearly do not adequately protect lake food webs. Water quality guidelines should be developed where they do not exist, and there is an urgent need to reassess existing guidelines to protect lake ecosystems from human-induced salinization.

Significance

The salinity of freshwater ecosystems is increasing worldwide. Given that most freshwater organisms have no recent evolutionary history with high salinity, we expect them to have a low tolerance to elevated salinity caused by road deicing salts, agricultural practices, mining operations, and climate change. Leveraging the results from a network of experiments conducted across North America and Europe, we showed that salt pollution triggers a massive loss of important zooplankton taxa, which led to increased phytoplankton biomass at many study sites. We conclude that current water quality guidelines established by governments in North America and Europe do not adequately protect lake food webs, indicating an immediate need to establish guidelines where they do not exist and to reassess existing guidelines.

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values, and fisheries (1). Moreover, increased water demands and climate change are expected to intensify freshwater salinization worldwide (9, 10).

In lake ecosystems, salinization can affect freshwater organisms at multiple trophic levels (11). Zooplankton are ubiquitous and critical to lake food webs because they transfer energy from primary producers to higher-level consumers. This zooplankton–phytoplankton pathway supports freshwater fisheries, influences nutrient cycling, and has a strong effect on water quality and clarity (12, 13). A change in this energy pathway in lake food webs could trigger a significant loss of resources that lakes provide (14, 15). For instance, a reduction in the abundance and diversity of zooplankton reduced fish growth and recruitment in Lake Michigan in the United States (16). To date, our understanding of how salinization affects the zooplankton–phytoplankton pathway is limited, restricting our ability to generalize the impacts of salinization on lake food webs and develop policies to protect lakes.

One of the chemical signals of salinization in lakes is an elevated chloride (Cl\(^{-}\)) concentration. For example, Cl\(^{-}\) is a good measure of road salt pollution because it acts as a conservative tracer (i.e., is not biologically transformed) (17) and is the anion of the most commonly used deicing salts: sodium chloride (NaCl), calcium chloride (CaCl\(_2\)), and magnesium chloride (MgCl\(_2\)). Similarly, Cl\(^{-}\) is also a good tracer for identifying other sources of salt pollution from agriculture fertilizers [e.g., KCl (18)] or seawater intrusion (19). Thus, governments in many countries such as the United States, Canada, Germany, and Sweden use Cl\(^{-}\) concentration to set thresholds to protect freshwater organisms or drinking water sources. Currently, the lowest thresholds to protect freshwater organisms are 230 mg Cl\(^{-}\)/L in the United States and 120 mg Cl\(^{-}\)/L in Canada (20, 21). In Europe, chloride standards for freshwater ecosystems are largely absent (22), but there are some exceptions. In Germany, Cl\(^{-}\) concentrations of 50 to ≤200 mg Cl\(^{-}\)/L are classified as slightly polluted by salts, and concentrations of 200 to 400 mg Cl\(^{-}\)/L are classified as moderately polluted by salts, while the drinking water guideline is 250 mg Cl\(^{-}\)/L across much of Europe [e.g., Sweden (23)].

At present, we do not know if Cl\(^{-}\) thresholds will protect lake fisheries in countries where Cl\(^{-}\) thresholds exist. Some studies indicate dramatic negative effects on zooplankton near or just above the US and Canadian Cl\(^{-}\) thresholds of 230 and 120 mg Cl\(^{-}\)/L, respectively (24–26). Others found toxic effects only at extremely high Cl\(^{-}\) concentrations (27–29). One recent study suggests population declines in freshwater zooplankton can occur between 5 and 40 mg Cl\(^{-}\)/L (30), concentrations far below the thresholds set by the United States and Canada and throughout Europe. The reasons for the variability among studies exploring the effects of salinization on zooplankton remain unclear. Lakes have diverse water chemistry, trophic state, and food web structure—properties which can affect the response of freshwater organisms to rising salinity (15, 31, 32). For instance, calcium concentration can affect the toxicity of Cl\(^{-}\) on freshwater zooplankton (32, 33), and lake ecosystems worldwide vary substantially in calcium concentration (34). Thus, differences among watersheds including water chemistry, historical exposure to salinity, different methodologies, community structure, and distinct genetic strains might explain variability among published studies (35). Further, nearly all previous studies have been single-lake studies or laboratory experiments with ideal conditions, further complicating our ability to estimate the overall threat that increased salinity poses to lake food webs and set sound regulatory thresholds or change existing ones.

To accurately generalize the effects of salinization on lake food webs, we need to compare responses using standardized methods while incorporating local differences in species composition and environmental conditions of lake water. Because experimental networks that replicate and standardize experiments across space and time can overcome this limitation [e.g., Nutrient Network (36), Zostera Experimental Network (37), and the Swedish Infrastructure for Ecosystem Science (SITES) AquaNet (38)], we established an experimental network of 16 sites across North America and Europe to identify responses of lake food webs to salinization triggered by NaCl, one of the most common salt types leading to the salinization of freshwater lakes (e.g., from road salt use). Further, using zooplankton communities from natural habitats has an advantage over short-duration (e.g., 48-h toxicity tests), single-species laboratory studies because such an approach encompasses a greater diversity of species and naturally occurring predator–prey and competitive interactions within the zooplankton community. We focused on determining if current Cl\(^{-}\)-based water quality guidelines protect lake food webs in regions with different geology, water chemistry, land use, and species pools. We predicted a disruption in the zooplankton–phytoplankton energy pathway would occur at higher salinities and would be manifested by reduced abundance of major zooplankton taxa, resulting in a concomitant increase in phytoplankton when compared with unsalinized control conditions. We also expected that local species pools would vary in their response to elevated salinity and that site-level water chemistry and environmental conditions would explain variation in the responses across experimental sites. Specifically, we predicted that higher concentrations of three essential nutrients (calcium, nitrogen, and phosphorus) and food resources (phytoplankton) among the study sites would reduce the impacts of elevated salinity on zooplankton.

**Results**

Our global generalized additive model (GAM) indicated that a site-specific model explained more variation in the response for each zooplankton taxa among our study sites (SI Appendix, Appendix 4, Table 4.6). Thus, we proceeded with site-specific GAMs to evaluate the zooplankton responses to our Cl\(^{-}\) concentration gradient (SI Appendix, Appendix 4, Table 4.7). Across study sites, the response of all zooplankton taxa to the Cl\(^{-}\) concentration gradient was generally nonlinear and negative (e.g., Fig. 1 and SI Appendix, Appendix 4, Figs. 4.1–4.4). The site-specific GAMs allowed us to estimate the lethal concentration values that caused a 50% reduction in abundance (LC\(_{50}\) ± 95% confidence intervals (CIs) for most zooplankton taxa among the study sites. Many of these LC\(_{50}\) values ± 95% CIs overlapped or were below the lowest two Cl\(^{-}\) thresholds established by the Canadian and US governments to protect aquatic life in these countries. We also found that the GAMs predicted significant declines in zooplankton abundance among all taxa and at the vast majority of our study sites at the Canadian threshold of 120 mg Cl\(^{-}\)/L and US threshold of 230 mg Cl\(^{-}\)/L. The environmental variables we measured generally did not explain the negative response of zooplankton to our Cl\(^{-}\) concentration gradient. The loss of zooplankton grazers led to a higher biomass of phytoplankton at 47% of our study sites (e.g., Fig. 1).

**LC\(_{50}\) Values Relative to Cl\(^{-}\) Thresholds.** We found considerable variability among the predicted LC\(_{50}\) values across the study sites. Based on the LC\(_{50}\) values, calanoid copepod, cladoceran, and rotifer zooplankton were the most susceptible to our environmentally relevant Cl\(^{-}\) concentrations. Cyclopoid copepod zooplankton had the lowest number of sites (33%) where LC\(_{50}\) values overlapped with or were below the Canadian and US Cl\(^{-}\) thresholds.

We were able to estimate LC\(_{50}\) values with 95% CIs for cladoceran zooplankton at all study sites that contained cladocerans (15 of 15; Fig. 24). These values indicated a substantial loss of cladoceran zooplankton occurred. The LC\(_{50}\) values ± 95% CIs at 73% (n = 11) of our sites overlapped with or were below
the Canadian threshold of 120 mg Cl⁻/L, and LC₅₀ values ± 95% CIs overlapped with or were below the US threshold of 230 mg Cl⁻/L at 86% of our sites. At two sites (Erken and Convict), the LC₅₀ values were above the Cl⁻ thresholds. One site did not contain any cladocerans.

For calanoid copepod zooplankton, we were able to estimate LC₅₀ values with 95% CIs at 11 of 12 study sites that contained calanoids (Fig. 2B). Four sites did not contain any calanoid zooplankton. In general, there was a rapid decline in calanoid abundance as Cl⁻ concentration increased (SI Appendix, Appendix 4, Fig. 4.2), which did not allow us to estimate an LC₅₀ value for one site (Stortjarn). The calanoid LC₅₀ values ± 95% CIs overlapped with or were below both the Canadian threshold of 120 mg Cl⁻/L and US threshold of 230 mg Cl⁻/L at 90% of study sites. At one site (Dartmouth), the LC₅₀ value was above both Cl⁻ thresholds.

We were able to estimate LC₅₀ values with 95% CIs for cyclopoid copepod zooplankton at 15 of the 16 study sites (Fig. 2C). At one site (Purdue), cyclopoid zooplankton abundance did not increase or decrease along the Cl⁻ concentration gradient (SI Appendix, Appendix 4, Fig. 4.3), so no LC₅₀ was estimated. For 33% (n = 5) of the remaining 15 study sites, the LC₅₀ values ± 95% CIs overlapped with or were below the Canadian threshold of 120 mg Cl⁻/L. The LC₅₀ values ± 95% CIs overlapped with or were below the US threshold of 230 mg Cl⁻/L at 60% (n = 9) of the 15 sites. At 40% (n = 6) of sites with estimable LC₅₀ values for cyclopoids, the values were above the Canadian and US thresholds.

For rotifer zooplankton, we were able to estimate LC₅₀ values with 95% CIs at 11 of the 16 study sites (Fig. 2D). For these sites, 73% (n = 8) of them had LC₅₀ values ± 95% CIs that overlapped with or were below the Canadian threshold of 120 mg Cl⁻/L, and 82% (n = 9) of them overlapped with or were below the US threshold of 230 mg Cl⁻/L. At multiple sites, rotifer abundance fluctuated substantially or increased along the Cl⁻ concentration gradient (SI Appendix, Appendix 4, Fig. 4.4), which did not allow us to estimate an LC₅₀ value at four sites (Dartmouth, Kraus, Kellogg Biological Station [KBS] Reservoir, and Convict). At two study sites (George and Tavernoles), the estimated LC₅₀ values were above the Canadian and US Cl⁻ thresholds.

Relationships between Local Environmental Conditions, the LC₅₀ for Zooplankton Taxa, and Community Composition. Local environmental conditions did not affect the predicted LC₅₀ values for three of the four zooplankton taxa (cladocerans, cyclopoids, or

![Fig. 1. GAMs and 95% CIs (shaded regions) for cladoceran zooplankton abundance (black line) and chlorophyll a concentration (i.e., phytoplankton biomass; green line) among 16 experimental sites throughout North America and Europe. Cladocerans are featured as they are generally regarded as the largest, highest-biomass filter-feeding zooplankton grazers in our study systems. The vertical red dashed line indicates the lethal concentration value (LC₅₀) representing the Cl⁻ concentration at which there was a 50% reduction in abundance of each zooplankton taxa for each site with the associated 95% CIs (red shaded region around the dashed line). Graphs for calanoid and cyclopoid copepods and rotifers are shown in SI Appendix, Appendix 4, Figs. 4.2–4.4. No cladocerans were found at the Tavernoles study site in Spain.](https://doi.org/10.1073/pnas.2115033119)
Fig. 2. LC$_{50}$ representing the Cl$^-$ concentration at which there was a 50% reduction in abundance of each zooplankton taxa across experimental sites (A) Cladocerans, (B) Calanoids, (C) Cyclopoids, and (D) Rotifers. The horizontal red lines highlight the lowest global Cl$^-$ thresholds for protecting aquatic life, which are set by governments in Canada (solid line, 120 mg Cl$^-$/L) and the United States (dashed line, 230 mg Cl$^-$/L). Vertical bars represent 95% CIs. Points and 95% CIs are red when they overlap with or were below the US threshold of 230 mg Cl$^-$/L. When a zooplankton taxon was not found in the mesocosm experiment, it is noted as “NA.” One experiment did not count rotifers, denoted with an asterisk. LC$_{50}$ values were not estimable when model fits were unreliable due to a rapid decline in abundance to zero along the Cl$^-$ gradient, there was no change, or an increase in abundance with Cl$^-$ (for missing values, see Materials and Methods).

Magnitude of Changes in Zooplankton Abundance Relative to Cl$^-$ Thresholds. We estimated the change in abundance at the Canadian (120 mg Cl$^-$/L) and US (230 mg Cl$^-$/L) thresholds using GAM models by comparing the predicted abundances at each threshold to the predicted abundance in control conditions. For each zooplankton group, responses in abundance to the Cl$^-$ concentration gradient were generally nonlinear and negative. Substantial declines were observed at many study sites, and at a few sites, some zooplankton taxa disappeared altogether.

Cladoceran zooplankton were detected at 15 of our study sites and were predicted to decline at 80% ($n = 12$) of those 15 study sites. The abundance of cladocerans declined by 17 to 49% at the Canadian threshold of 120 mg Cl$^-$/L compared to the site-specific control concentrations (Fig. 3). Cladoceran abundance declined by 22 to 83% among the same sites at the US threshold of 230 mg Cl$^-$/L. At three sites (Feresjön, Erken, and Convict), there was a positive effect of elevated Cl$^-$ concentration on cladoceran abundance at the Canadian and US thresholds. However, at one of those sites (Feresjön), the error bars overlapped with the no-change threshold (dashed line in Fig. 3).

Calanoid copepod zooplankton were detected at 12 of our study sites and were predicted to decline at 92% ($n = 11$) of those sites. At the Canadian threshold, calanoid abundance declined by 6 to 83% compared to the control concentrations among 11 study sites (Fig. 3). Among the same sites and at the US chronic threshold, calanoid abundance declined by 15 to 96% compared to control Cl$^-$ concentrations. At one study site (Stortjärn), there was a predicted increase in calanoid abundance (6 to 11%), but the error bars overlapped with the no-change threshold (dashed line in Fig. 3).

Cyclopoid copepod zooplankton were detected at all our study sites. Cyclopoid abundance was predicted to decline at 88% ($n = 14$) of the sites (Fig. 3). At the Canadian threshold, cyclopoid abundance declined by 7 to 69% among 14 sites compared to the control conditions. Among the same sites and at the US threshold, calanoid abundance declined by 13 to 96% compared to control Cl$^-$ concentrations. At two sites (Feresjön and Convict), Cl$^-$ concentrations at both thresholds were predicted to have a positive effect on cyclopoid abundance.

Rotifer zooplankton were detected at all our study sites but were not counted at one site. Rotifer abundance was predicted to decline at 73% ($n = 11$) of the sites (Fig. 3). At the Canadian Cl$^-$ threshold, rotifer abundance declined by 5 to 97% compared to the control concentrations among 11 sites. Among the same sites...
and at the US chronic threshold, rotifer abundance declined by 10 to 100% compared to control Cl\textsuperscript{−}/C\textsubscript{0} conditions. At four sites (KBS Reservoir, Dartmouth, Kraus, and Tavernoles), Cl\textsuperscript{−} concentrations at both thresholds were predicted to have a positive effect on rotifer abundance, although the error surrounding the predicted decline at three sites (KBS Reservoir, Dartmouth, and Kraus) overlapped with the no-change threshold (dashed line in Fig. 3).

**Cascading Effects.** We used the correlation between cladoceran abundance and phytoplankton biomass (i.e., chlorophyll \textit{a}) to evaluate whether a cascading effect occurred at the 15 study sites where cladocerans were found (Fig. 1). At 47\% (\(n = 7\)) of the 15 sites, there was a negative correlation between cladoceran abundance and phytoplankton biomass (Table 1). We did not detect a correlation between cladoceran abundance and phytoplankton biomass at seven sites. At one site (Dartmouth), there was a positive relationship between cladoceran abundance and chlorophyll \textit{a}.

**Discussion**

Our results indicate that lake food webs change in response to freshwater salinization. Our experimental Cl\textsuperscript{−} concentrations commonly found in natural systems throughout the world led to large reductions in the abundance of major zooplankton taxa and increases in phytoplankton biomass. For 73\% of our study sites, a 50\% decline in the abundance of the four major zooplankton taxa could occur in Cl\textsuperscript{−} concentrations at or below thresholds established by governments in North America and Europe. The loss of zooplankton led to an increase in phytoplankton biomass at almost half of our study sites. Our results

![Fig. 3. Mean (±1 SE) of the estimated percent change in zooplankton abundances for each taxa at the Canadian (120 mg Cl\textsuperscript{−}/L) and US (230 mg Cl\textsuperscript{−}/L) chronic threshold for Cl\textsuperscript{−}. Percent changes were estimated using GAM models to compare the predicted abundances at each threshold to the predicted abundance in control conditions.](https://doi.org/10.1073/pnas.2115033119)
Cascading effects are indicated by a negative correlation between cladoceran abundance and chlorophyll a. Cladocerans were absent from one study site (Tavernoles). Bold values indicate $P < 0.05$.

Cascading effects clearly show that lake food webs are not protected by current Cl$^{-}$ thresholds. We must reassess current thresholds and develop new thresholds where they do not exist to protect our freshwater lakes.

Unlike short-duration (e.g., 48-h LC$_{50}$ tests), single-species studies, our studies included a community approach that encompasses a greater diversity of zooplankton and species interactions such as predator–prey relationships and competition over a 6- to 7-wk time span. We also leveraged a regression-style design, to better reflect responses of lake food webs to salinization and identify response thresholds. Based on our spatial coverage and environmentally relevant Cl$^{-}$ concentrations, our results suggest that many salt-contaminated lakes with Cl$^{-}$ concentrations near or above thresholds established throughout North America and Europe might have already experienced food web shifts (30, 39).

Cladoceran zooplankton were highly sensitive to our Cl$^{-}$ additions, and 80% of our study sites were not protected by the lowest Cl$^{-}$ thresholds established by the United States and Canada. In short-term exposures (usually 48 h), others have found that species-specific LC$_{50}$ values of laboratory strains of cladocerans generally exceed 1,200 mg Cl$^{-}$/L (e.g., refs. 27, 28, 40). Experimental work using wild cladocerans collected from lakes with little to no prior salt exposure have found a >50% reduction in abundance occurs between 246 and 500 mg Cl$^{-}$/L (e.g., refs. 24, 26, 41). Our estimated LC$_{50}$ values at 86% of our sites occurred at or below 230 mg Cl$^{-}$/L, and 73% of those sites occurred at or below 120 mg Cl$^{-}$/L. At present, our results are among the lowest LC$_{50}$ values reported for wild-strain cladoceran communities across lakes that vary in environmental conditions and species composition. These results are consistent with recent work by Arnott et al. (30), who showed that cladocerans can experience negative effects at concentrations of 5 to 40 mg Cl$^{-}$/L, which are well below current government Cl$^{-}$ thresholds designed to protect aquatic life.

Calanoid copepod zooplankton were also highly susceptible to our experimental Cl$^{-}$ treatments. The US and Canadian Cl$^{-}$ thresholds did not protect calanoids at 90% of our study sites. Further, the rapid decline and almost total loss of calanoids at some sites suggests that freshwater calanoids might be much more sensitive to salt pollution than cladocerans. This was also observed in a soft-water, northern boreal oligotrophic lake where saline effluent from mining operations triggered a complete loss of calanoid copepods while some cladocerans and cyclopoid copepods continued to persist over a 10-y time period (42). The highest diversity of calanoid copepods tends to occur in the limnetic zone of freshwater systems (43) where, like cladocerans, they play critically important functional roles in lake food webs as primary consumers and predators and are often a biomass dominant in north temperate lakes (44). Thus, salinization of freshwater lakes could trigger a substantial loss of calanoid diversity and shifts in lake food webs.

Cyclopoid copepods were less impacted by our Cl$^{-}$ treatments compared to cladocerans and calanoid copepods as indicated by the number of sites (40%) where the cyclopoid LC$_{50}$ values were above US and Canadian chronic thresholds. A higher tolerance of cyclopoids among more sites is consistent with previous work showing adult cyclopoids were not as vulnerable to NaCl-induced salinization compared to calanoids and cladocerans (45, 46). Even so, the LC$_{50}$ values for cyclopoids were at or below the US and Canadian thresholds at 33% of our study sites, and the abundance of cyclopoids at 88% of sites was lower than the control conditions at those Cl$^{-}$ thresholds, suggesting cyclopoids might still exhibit a high degree of susceptibility to salinization in many freshwater lakes.

Like the other taxa, our data suggest that rotifer zooplankton may not be protected by current Cl$^{-}$ thresholds in many lakes. Rotifer assemblages among 82% of our study sites had LC$_{50}$ values at or below the US and Canadian chronic thresholds. There is almost no research examining the impacts of Cl$^{-}$ concentration on rotifers in low-salinity lakes (e.g., naturally <20 mg Cl$^{-}$/L) and at the lower experimental Cl$^{-}$ concentrations used in our study. Much more research on the impacts of elevated salinity on rotifers has occurred in higher-salinity or estuarine ecosystems, which show a decline in rotifer fecundity with increasing salinity (47). One experimental study on freshwater rotifers found that multiple species exhibit little decline in population size below 910 mg Cl$^{-}$/L (40). Rotifers were much more sensitive in our study, particularly from lakes with low nutrients and ion concentrations, as has been shown with cladocerans (30, 39, 48). While the mechanism is unclear, freshwater rotifer decline could be due to reduced fecundity with increased salinity (47). Our results indicate that when we consider multiple assemblages of rotifers from a broad spectrum of freshwater lakes (i.e., <20 mg Cl$^{-}$/L) that vary in their environmental conditions, rotifers may be more sensitive to salinization effects in low-ion and low-nutrient lakes.

Recent work has shown that water chemistry can affect the response of some zooplankton taxa to salinization. For instance, water hardness and nutrient concentrations can alter the response of cladoceran species to road salt (49). Similarly, other studies show as water hardness (as CaCO$_3$/L) declines, the negative effects of NaCl increase at lower Cl$^{-}$ concentrations (30, 33). This relationship is particularly evident in Canadian Shield lakes where low nutrient concentrations (e.g., Ca$^{2+}$, N, and P) trigger substantial reductions in cladoceran populations at low Cl$^{-}$ concentrations (30). While the exact mechanism remains unknown, one potential mechanism for the reduction of Cl$^{-}$ toxicity at high water hardness is that higher concentrations of Ca$^{2+}$ and Mg$^{2+}$ in harder water can cause cellular junctions to tighten, which reduces passive transport of Cl$^{-}$ into animal tissue (50). Despite a significant range in Ca$^{2+}$ and other water chemistry variables such as background Cl$^{-}$ concentration, TN, TP, and chlorophyll $a$ among our study sites, these water chemistry variables only explained variation in rotifer responses. The lack of a pattern among other zooplankton taxa might suggest that measured water chemistry variables were not limiting, or it is possible that other unmeasured ions or ion ratios associated with water hardness (e.g., Mg$^{2+}$) were in sufficient supply. Nevertheless, our results illustrate that regulatory thresholds over a gradient of water chemistry among
freshwater lakes are insufficient to protect zooplankton communities and lake food webs from salinization.

Local genus composition of each group did not explain the variation in LC50 values. However, the response of zooplankton to salinization varies with genotype (31), so intraspecific variation in salt tolerance may explain the variable responses observed among our study sites. Among most of our study sites, one to three of the major taxonomic groups were not protected by the US and Canadian thresholds. However, at 25% of our study sites we found that all detected zooplankton taxa exhibited LC50 values at or below the thresholds (Croche, Long, Opeongo, and Stortjärn). This could indicate that genotypes from these lakes are the most sensitive to elevated salinity. In other locations (Convict), increases in the abundance of some taxa at low concentrations might indicate salt-tolerant genotypes, although a precipitous decline occurred at the highest Cl− concentrations (52). Given the lack of a relationship with the environmental data for crustacean zooplankton, it is likely that species- and population-level genetic differences due to adaptation to local ecosystem conditions could account for the variable nature of taxa responses among study sites (53). We encourage further investigation into the roles that regional genotypes have on the susceptibility of zooplankton communities to salinization.

Our study suggests that cascading effects from the loss of zooplankton resulting in higher phytoplankton biomass might be common among salinized lakes. While we do not have species-level phytoplankton data, it is likely that our high Cl− concentrations also induced shifts in the diversity and abundance of the phytoplankton community (e.g., ref. 54). At one study site, we found a positive relationship between cladoceran abundance and chlorophyll a, which could be due to high salinity promoting salt-tolerant phytoplankton species that could be less palatable for cladocerans. The extent to which phytoplankton communities respond to salinization and the food web implications of such changes remain to be well understood. Ultimately, the nature of salt-induced cascading effects on lake food webs will be complex (55). About half our experimental sites experienced these effects, similar to previous single-site studies (24, 56–58). This widespread deterioration of the zooplankton–phytoplankton energy pathway in lake food webs could affect production at higher trophic levels. Previous studies indicate that changes in zooplankton communities can result in poor growth and recruitment of economically important fish species (16). Furthermore, it is unclear what elevated phytoplankton biomass would mean for benthic communities in lakes, but reduced light attenuation or shading effects are possible and could lead to a regime shift due to reduced water clarity (14, 24).

Recent research shows evolutionary responses by zooplankton can occur in the form of an increased tolerance to NaCl (59). It is unclear whether rapid evolution of tolerance is widespread in natural communities and if such a tolerance could buffer zooplankton from the impacts of various sources of salinization such as road salts over the long term. While traits leading to the tolerance can change over short time periods [e.g., a few generations in Daphnia (60)], the durations of our 16 experiments were not long enough to account for evolved responses demonstrated in other studies (59). Nevertheless, while an evolved tolerance to NaCl and cross-tolerance to other salt types could potentially buffer declines in cladoceran abundance to repeated salt exposure, these evolved responses do not always mitigate the cascading effect we observed here (e.g., ref. 61). Thus, we would not expect evolutionary processes to stabilize the zooplankton–algae energy pathway in freshwater food webs (62).

Conclusions. The LC50 values and magnitude of declines in zooplankton abundance presented here were much larger and occurred at lower concentrations than expected based on published studies (11, 63, 64). Our results are concerning because substantial changes to community structure, food webs, and ecosystem function are likely to occur well before a 50% decline in the abundance of major zooplankton taxa. Zooplankton are critical for transferring energy from primary producers to secondary consumers and indirectly to tertiary consumers. A reduction in the biomass of zooplankton can reduce food availability for young fishes and other planktivores, leading to a reduction in growth rates and recruitment (e.g., ref. 16). Many lakes have been salinized to the point where zooplankton in our study experienced declines (65). It is unclear if and to what degree some of the documented and projected ecological effects discussed here have already occurred and how salinization has interacted with other stressors to shape contemporary lake communities.

Our results indicate an immediate need to reassess current governmental thresholds to protect lakes from salinization. In US Midwestern and Northeastern states alone, Dugan et al. (4) estimated that there are 2,000 lakes predicted to have Cl− concentrations >50 mg Cl−/L. In the next 50 y, many of those lakes will exceed the US threshold of 230 mg Cl−/L (3). However, lakes at concentrations of 42 to 90 mg Cl−/L have already undergone ecological change (30). Human use of salt sources responsible for freshwater salinization must strike a balance with ecological impacts. For example, road deicing salts reduce vehicle accident rates by 78 to 87% (66–68), and until ecologically responsible alternatives are available, deicing salts will be needed for the foreseeable future to protect the traveling public. For now, best management practices may dramatically reduce salinization caused by road salt pollution (65). However, there are multiple other sources of salinization to consider, and we must recognize the multitude of chemical mixtures that occur because of the freshwater salinization syndrome occurring globally (69–71). Understanding the interaction among ions in chemical mixtures and how various combinations mediate toxicity is imperative given changing trends in ion composition worldwide (69). We must develop new and lower Cl− thresholds across the world to protect freshwater communities, which will no doubt be a difficult task. The variability in our experimental results demonstrates that new Cl− thresholds must integrate the susceptibility of ecological communities at the local and regional scales to successfully develop these protective Cl− thresholds.

Materials and Methods

We conducted 16 experiments in temperate or subarctic ecosystems in North America and Europe (Fig. 4 and SI Appendix, Appendix 1, Table 1.1). The experiments were based on a common protocol and a regression-style experimental design to examine how a wide range of environmentally relevant Cl− concentrations (from NaCl that is >99% pure; SI Appendix, Appendix 2, Table 2.1) affected zooplankton communities and, in turn, indirectly affected phytoplankton biomass. Each experiment included 20 to 32 mesocosms; of the 16 experiments, we conducted 11 using land-based (tank) mesocosms and 5 using in-lake mesocosms (SI Appendix, Appendix 1, Table 1.1). The lowest concentration was the Cl− found in the source water (ranging from <2 to 18 mg/L) and we added increasing amounts of Cl− to a highest nominal concentration of 1,500 mg/L (Fig. 4). We conducted the experiments during 2018, and each lasted for 41 to 51 d (SI Appendix, Appendix 1, Table 1.1). Further information on the experimental design can be found in SI Appendix, Appendices 1–3 and 5. Supplemental methods text can be found in SI Appendix, Appendix 1, Table 1.1.

At the end of each experiment, we concentrated zooplankton by filtering water samples taken from each mesocosm (1.2 to 15% of the mesocosm volume) using a 50- to 64-μm mesh (SI Appendix, Appendix 5, Tables 5.1 and 5.2). We preserved zooplankton in 70 to 95% ethanol or Lugol’s solution. We subsequently identified and counted zooplankton using stereomicroscopes (10 to 100x magnification) and standard counting protocols for each laboratory (SI Appendix, Appendix 5.2). We generally identified cladocerans and adult copepods. Cladocerans and rotifers were identified to species.

To quantify phytoplankton biomass, at the end of the experiment we sampled 200 to 500 mL of water from each mesocosm to quantify the concentration of chlorophyll a. We then filtered water samples through glass microfiber
models for each site with the log10(\text{Cl}^-)/C_0 concentration: this approach is conceptually similar to a traditional LC50 value in a laboratory toxicology experiment (except it incorporates births and mortality), and we refer to it as a 50% reduction in the abundance of each taxonomic group compared to the high end of the Cl^-/C_0 concentration. Because these variables were highly collinear, we conducted a PCA based on a correlation matrix on log10-transformed variables (Cl^-, Ca^{2+}, TN, TP, and chlorophyll a). We then used linear models to determine if the LC50 value for each taxonomic group was related to the individual or combined effects of the first and second PCA axes. Because of the low number of sites, we evaluated the main effects of PC1 and PC2 and did not include their interaction. Statistical support was determined using ANOVA after evaluating model fit by examining residuals versus predicted values, scale-location, and residual versus leverage plots.

We also assessed if zooplankton sensitivity was related to local community composition (SI Appendix, Appendix 6). We first conducted a separate PCA for each taxonomic group (cladocerans, cyclopoids, calanoids, and rotifers) using the abundance of each genus found in control mesocosms. When there were replicates for the control (e.g., Kraus and Dartmouth), we used the average abundance. We then Hellinger-transformed the abundance data and used a correlation matrix for the analyses (76). We then used linear models to determine if the LC50 of each taxonomic group was related to the individual or combined effects of the first two PCA axes as described above.

**Cascading effects.** We examined the relationship between the cladoceran abundance and phytoplankton biomass at the end of the experiment using Pearson correlation coefficients to determine if there was a cascading effect in the experimental food webs. We defined a cascading effect as a significant (i.e., P ≤ 0.05) negative correlation between cladoceran abundance and phytoplankton biomass—i.e., as Cl^- concentration increased, the decline in cladoceran abundance would reduce grazing pressure, triggering an increase in phytoplankton biomass along the experimental Cl^- gradient. To linearize the data, we log10-transformed chlorophyll a concentration and log10(x + 1)-transformed cladoceran abundances.

**Data Availability.** Excel files, Word files, and PDFs have been deposited and are available in Dataverse (https://doi.org/10.5683/SP2/BIDMCI) (72).

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