Orthobiologics in the treatment of hip disorders

André Vinícius Saueressig Kruel, Lucas Leite Ribeiro, Paulo David Gusmão, Stephany Cares Huber, José Fábio Santos Duarte Lana

Abstract

Orthobiologics are biological materials that are intended for the regeneration or healing of bone, cartilage and soft tissues. In this review we discuss the use of orthobiologics for hip disorders providing an update. The orthobiologics included in this article are hyaluronic acid, platelet-rich plasma, bone marrow, adipose tissue and expanded mesenchymal stem cells. We explain the concepts and definitions of each orthobiological product, and the literature regarding its use in the hip joint. The paucity of guidelines for the production and characterization of the biological products leads to uneven results across the literature. Each biologic therapy has indications and benefits; however, noteworthy are the characterization of the orthobiologics, the application method and outcome analysis for further improvement of each technique.

Key Words: Orthobiologics; Hip disorders; Platelet-rich plasma; Mesenchymal stem cells; Bone marrow; Adipose tissue

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.
Orthobiologics can be defined as biological materials used in the regeneration and repair of conditions affecting bone and adjacent soft tissues. They can be implanted, injected or administered into the patient in order to increase the natural healing potential of the injured musculoskeletal tissue. Recently, a variety of orthobiologics has arisen and each type has peculiarities and often present various mechanisms of action. The theoretical advantages of orthobiologics are minimal invasiveness (compared to more austere open or endoscopic forms of traditional orthopedic surgery), greater healing potential (than mere exercise or physical therapy, for instance), faster recovery and reduced cost opposed to surgery, making it a viable alternative.

Pathologies affecting the hip joint can be divided into three arbitrary layers: intra-articular, periartricular and intra-osseous. This segmentation has arisen with the help of better imaging methods such as magnetic resonance imaging (MRI) and other procedures such as hip arthroscopy, where labral, chondro-labral junction, Teres ligament lesions or subchondral cystic lesions have been diagnosed and more conservatively treated. Arthritic hip joints have also been managed, although results have been less predictable. Hip avascular necrosis (AVN) is especially amenable to orthobiological therapy and enduring results have been shown, sometimes paired with adjunctive non-arthroplastic options like core-decompression. Core-decompression treatment consists of tunnel drilling and removal of the necrotic segment from the femoral head, providing an opportunity for new pooling of cells to be implanted in the site.

Subchondral bone-marrow edema, a less understood pathology leading to pain or hip arthritis, is also a good candidate for such treatments, considering its biomechanical and biological background and continuous development of image-guided procedures such as ultrasound oriented intra-osseous hip injection. Periartricular hip pathology includes chronic tendonitis, as well as deep gluteal space disorders. The latter express themselves often in conjunction with subtle or even overt peripheral nerve entrapments. Such tendinopathies hold special interest for their common prevalence and their difficult management in remitting cases. The aim of this review is to provide an update on orthobiologics that can be used for the treatment of hip ailments, with a special focus on the protocols of obtainment (when appropriate, along with our specific protocols), relationship to surgical aspects and clinical evidence of relevance.
also called sodium hyaluronate, or hyaluronan[11]. Intra-articular hyaluronan preparations have been shown to relieve osteoarthritis-derived pain more than conventional treatments such as physical therapy, exercise, nonsteroidal anti-inflammatory drugs (NSAIDs), intra-articular corticosteroids or even arthroscopic lavage[12].

The physicochemical properties of HA are determined by molecular mass and spatial conformation. It can also be classified as high, medium or low molecular weight. Normal (non-osteoarthritic) autologous HA in joints weighs around 5 to 7 million Daltons, whereas osteoarthritic joint HA weighs an average of 1 million Daltons. High molecular weight HA molecules intertwine to form a high viscosity solution, which serves both as a lubricant and shock absorber, and has biomolecular properties compatible with and favorable to cell growth[13].

HA is the major non-protein hydrodynamic component of synovial fluid. This substance, biological or synthetic, forms a layer around the cells where it interacts with pro-inflammatory mediators and binds to cellular receptors, modulating cell proliferation and migration, as well as gene expression[14]. It is a collagen stimulator, capable of promoting tissue recovery and maintaining cellular integrity[15]. These properties make HA useful for healing or even regeneration of tendon and chondral tissues in some cases.

Intra-articular (IA) injection of HA is considered a local treatment with no reports of systemic adverse events observed after the administration of other types of IA injections, such as corticosteroids, or even with oral administration of NSAIDs[16]. This therapy represents an alternative treatment, especially for patients with comorbidities. This alternative approach has shown beneficial effects in vitro. It is suggested that the extracellular matrix (ECM) has an influence on cell metabolism, especially on osteoarthritic subchondral bone osteoblasts. HA can reverse its abnormal synthetic activity[17].

An IA injection of HA has been shown to reduce chondrocyte apoptosis whilst increasing its proliferation[18]. This therapy has been shown to promote chondroprotection due to the HA binding to CD44 receptors. This event inhibits the expression of IL-1β, which leads to a decrease in the biosynthesis of MMP-1, 2, 3, 9, and 13[19-21]. This process avoids catabolic enzyme activity within the joint cartilage[22]. It is our experience, in human patient treatment, that it is important to use medium or high molecular weight HA, similar to the molecular weight that is normally produced in the body. Also, it is important to use HA derived from biological synthesis, to avoid undesired side effects. Another advantage of HA is the possibility of its combination with other products, biological or not. For example, the combination of calcitonin[23], sorbitol[24], platelet-rich plasma (PRP)[25,26] and/or bone marrow aspirate concentrate (BMAC)[27,28] have been postulated. Clinical trials reporting the use of HA for hip disorders are listed in Table 1.

**PRP**

PRP is an autologous non-immunogenic therapy, produced through the concentration of platelets in a small plasma volume (typically 3-6 times above baseline)[29]. PRP therapy has advantages such as rapid preparation and technical simplicity; it is also a point-of-care procedure, and can be carried out in-office due to its minimal invasiveness, permitting intra-articular, intra-tendinous or even intra-osseous injection. As a result of its autologous origin, it exhibits a unique safety profile, lacking many drug-related side effects or interactions[29]. In osteoarthritis (OA), PRP can interfere in the catabolic and inflammatory cascade, in order to promote anabolic responses. The bioactive molecules in PRP can act in inflammatory modulation, ECM synthesis and vessel remodeling in order to improve the natural healing of the tissue[30]. One of the greatest problems regarding PRP is the lack of standardization. Important variables such as the number of platelets, Buffy-coat content, exogenous activation and number of centrifugations are lacking in many publications, resulting in heterogeneous results, which therefore cannot be compared. In order to minimize this problem, some PRP classifications have been proposed. Recently, Lana *et al.*[31] published a thoughtful classification incorporating all these variables. Besides the aforementioned variables, light activation, image guidance method and red blood cell content were included. All these parameters are important for a standardization protocol.

PRP was used alongside hip arthroscopy surgery for a variety of pathologies[32]. The technique consists of the administration of 4.5 mL of PRP into the repaired hip joint capsule in the peripheral compartment through the arthroscopic cannula, as well as 10 mL of PRP in the surrounding soft tissues as suggested by Marc Philippon and Robert Laprade’s group. Growth factors present in PRP in addition to the aforementioned
Favorable results from baseline for 6 wk only for cortisone, whereas LR-PRP BM-MSC 2 × 10
Issue 4
April 26, 2021
Results showed that patients in early stages (Steinberg I-II) had excellent outcomes 8-14 mo
Both treatment groups receiving AD-MSCs showed clinically significant pain and
Results
Minimization of pain, preservation of joint function, and obviation or at least a
All patients’ symptoms improved over the follow-up period of 10 mo (8–14 mo).
IA PRP injections offer a significant clinical improvement in these patients, without side effects, verified by scores. The best results were achieved with PRP used as monotherapy and remained stable up to 12 mo. The addition of HA to PRP did not lead to a significant improvement in pain symptoms

| Ref. | Groups | Follow-up | Results |
|------|--------|-----------|---------|
| Battaglia et al[40] | IA–HA (n = 50) vs PRP (n = 50) | 12 mo | IA injections of PRP are efficacious in terms of functional improvement and pain reduction but are not superior to HA in patients with symptomatic hip OA |
| Pogliacomi et al[41] | HA (n = 226), 2500 kDa | 12 mo | Scores improvement, but Kellgren-Lawrence stage 2 patients exhibited most benefit. Nevertheless, the number of applications and molecular weight of the injected products are subject to debate |
| Clementi et al[42] | HA single high-molecular weight (n = 27), two-shot medium molecular weight (n = 23) | 12 mo | A single dose of high molecular weight was as effective as two doses of medium molecular weight resulting in similar reductions in pain and disability |
| Dallari et al[43] | HA (n = 36), PRP (n = 44), HA + PRP (n = 31) | 12 mo | IA PRP injections offer a significant clinical improvement in these patients, without side effects, verified by scores. The best results were achieved with PRP used as monotherapy and remained stable up to 12 mo. The addition of HA to PRP did not lead to a significant improvement in pain symptoms |
| Fitzpatrick et al[44] | LR-PRP (n = 40), corticosteroid injection (n = 40) | 24 mo | Favorable results from baseline for 6 wk only for cortisone, whereas LR-PRP ultrasound-guided applications continued improvement for up to 15 mo. |
| Houdek et al[45] | PRP + BMAC with core decompression of the hip | 24 mo | Improvement in pain and function with the injection of BMAC and PRP. During follow-up (2 years), more than 90% of hips were collapse-free. Successful results were more evident when nucleated cell count was high and modified Kerboul grade was low |
| Hernigou et al[46] | BMAC adjunct to core decompression (n = 189) | 5 yr | Results showed that patients in early stages (Steinberg I-II) had excellent outcomes at 5 years, only 6.2% of the hips requiring total hip arthroplasty. On the other hand, patients in advanced stages (Steinberg III-IV) had poor outcomes, with 60% requiring total hip arthroplasty (THA). Positive relationship between the clinical outcomes and the number of progenitor cells |
| Einhorn et al[47] | BMAC adjunct to core decompression (n = 52) | 24 mo | Minimization of pain, preservation of joint function, and obviation or at least a delay in the need for THA with 2 years of follow-up. In this study 62-75% of CD-BMAC recipients avoided the need for additional interventions, including THA |
| Freitag et al[48] | AD-MSC single injection (100 × 10⁶ cells) (n = 10), AD-MSC two injections (100 × 10⁶ cells) (n = 10), control-conservative treatment (n = 10) | 12 mo | Both treatment groups receiving AD-MSCs showed clinically significant pain and functional improvement at completion of follow-up at 12 mo (Numeric Pain Rating Scale and WOMAC, as well as KOOS). Radiological analysis using MRI indicated modification of disease progression |
| Mardones et al[49] | BM-MSC a unique dose (40 × 10⁶ cells), preceding core decompression (n = 5), hydroxyapatite or a calcium phosphate plug was placed immediately after cell instillation | 19-54 mo | After the follow-up period all patients had significantly improved hip function and markedly reduced pain intensity. As a corollary, no patient required hip arthroplasty |
| Mardones et al[50] | BM-MSC 20 × 10⁶ cells n = 7, mild OA | 8-14 mo | All patients’ symptoms improved over the follow-up period of 10 mo (8–14 mo). Average Vaii-10 and Modified Harris Hip Scores for all patients showed significant improvement at 3 and 6 mo. None of the patients required a total hip replacement at the time |
| Zhao et al[51] | BM-MSC 2 × 10⁶ autologous cells (n = 50), CD (n = 50) | 60 mo | In comparison with CD, the cell treated group showed a significant improvement in the Harris hip score, as well as a decrease in the volume of femoral head and no complications were observed in either group. The authors concluded that ex vivo expansion of autologous BM-MSC provides a greater number of cells; it is safe and effective in delaying and avoiding FH collapse |

IA: Intra articular; HA: Hyaluronic acid; PRP: Platelet-rich plasma; OA: Osteoarthritis; LR-PRP: Leukocyte-rich platelet-rich plasma; BMAC: Bone marrow aspirate concentrate; CD: Core decompression; THA: Total hip arthroplasty; AD-MSC: Adipose-derived mesenchymal stem cell; WOMAC: Western Ontario and McMaster Universities Osteoarthritis Index; BM-MSC: Bone marrow-derived mesenchymal stem cell; KOOS: Knee injury and Osteoarthritis Outcome Score.

healing potential also aid in postoperative hemostasis[23]. Regarding the use of leukocytes, it has been reported that leukocyte-rich PRP is effective in the initiation of a proinflammatory healing response to stimulate vascular tissue regrowth[24]. However, for the treatment of hypovascular tissues, it was reported that it may induce profibrotic tissue formation, which reduces the quality of the tissue itself[25]. PRP can also be used in combination with different orthobiologics, such as HA and bone marrow preparations[26-27]. Also, even if PRP does not lead to cartilage regeneration, it may still offer symptomatic and functional benefits via modulation of inflammation and direct analgesia[28]. The description of clinical trials for hip disorders is summarized in Table 1. PRP has also been compared to surgery, regarding gluteus medium tendonitis in a recent meta-analysis[29] and has been deemed effective and safer than endoscopic surgical procedures. In addition, in another recent meta-
analysis, PRP was compared to HA and steroid injections and was considered to achieve the highest rank for pain relief at 6 mo in patients suffering from OA[36].

In our clinical practice, after a standardization protocol for PRP therapy, we opted for a handmade PRP preparation, with double centrifugation, enriched with buffy-coat (i.e., leukocyte-rich PRP) using the modified protocol of Amable et al[37] and Huber et al[38]. With this technique, we obtained a 3-5-fold increase in platelet concentration and a 2-4 fold increase in white blood cells from baseline (therefore leukocyte rich).

**Bone marrow derivatives**

Bone marrow has been studied in the context of regeneration for many years. It encompasses erythroblasts, neutrophils, eosinophils, basophils, mononuclear cells [monocytes and mesenchymal stem cells (MSCs)], lymphocytes, megakaryocytes, and plasma. The erythroblasts, eosinophils and neutrophils are present in various stages of differentiation[39]. Especially relevant are the bone marrow-derived mesenchymal stem cells (BM-MSCs), hematopoietic stem cells (HSCs) and other progenitors. The use of BM-MSCs has demonstrated benefits in the regeneration process. The HSCs are thought to be the true drivers for enhancing cartilage and bone regeneration, with an important role in the direct conversion to stromal MSCs and orchestration of the bone formation[40] as described by Omsted-Davis in 2003, who pointed out that a side population of marrow stem cells could regenerate the hematopoietic compartment of lethally irradiated mice when transplanted and could also differentiate to osteoblasts through a mesenchymal intermediate[41]. This effect was also demonstrated clinically in the study by Marx et al[42] in 2014 where they devised a study with 40 adults undergoing craniomandibular reconstruction. Those patients attaining higher CD34+ counts registered a 100% regeneration of implantable bone, as opposed to 40% of this effect, when CD34+ counts were much lower[43]. Pettine also demonstrated improved results in a study on intradiscal injection of bone marrow concentrate (BMC) with a three-year follow-up, and found that HSC (CD34+) also played an immunomodulatory role similar to MSCs and reported that patients who received greater concentrations of progenitor cells [colony forming unit-fibroblast, (CFU-F), and CD34+ lineage] experienced faster and greater pain reduction. The authors also claimed the following: “This is the first study to link a clinical improvement to CFU-F and CD34+ cell concentrations in BMC[44].”

MSCs were first discovered in the bone marrow, presenting an important paracrine effect. MSCs thus exhibit a secretory function, comprising anti-inflammatory, proangiogenic, immunomodulatory, anti-apoptotic, anti-fibrotic and wound healing properties, including a proliferative role[45]. MSCs represent 0.01% to 0.001% of total bone marrow nucleated cells. Due to this low number, there is a trend for cellular expansion in order to obtain higher cell counts for clinical applications[46]. However, the use of non-cultured cells presents some advantages that go beyond cost and good manufacturing practice and regulation. One of the major advantages is that the direct extraction allows the maintenance of its regenerative capacity and avoids senescence[47]. Expansion techniques risk cells with variable differentiation capacity, increased senescence markers and tumor degeneration liability. Also, these cells must be cultured for two or three weeks – a slow and expensive process that in clinical practice demands a two-staged tissue implantation surgery. One important issue is that the use of pure, cultured MSCs do not contain HSCs, cytokines and growth factors as concentrated bone marrow does[48]. The outcomes of concentrated vs cultured cells should be assessed more carefully, taking into account the costs, time expenditure and clinical practice suitability[49].

Previous studies showed variables that could influence the number of MSCs, such as anatomic harvest site, aspirated volume, age and sex of the patient. Traditionally, MSCs could be predictably obtained from the iliac crest[46]. It is also regarded that the aspiration of larger volumes decreases the concentration of osteoblast progenitors due to the dilution of bone marrow with peripheral blood[46]. Regarding the anatomic site of harvesting, Pierini et al[49] published a study comparing bone marrow collection from both the anterior and posterior iliac crest of twenty-two patients. The yield of colony-forming connective-tissue progenitors was 1.6 times greater in the posterior compared to the anterior iliac crest. In relation to the volume aspirated, Hernigou et al[50] published a study evaluating the aspiration of bone marrow from iliac crest using 10 mL and 50 mL syringes, and compared MSCs and progenitor cell content. It was verified that the aspirate performed with a 10 mL syringe had a greater concentration of MSCs and 300% more progenitor cells in comparison with the 50 mL syringe aspiration.
In order to increase the relative number of MSCs per volume of injectate for therapeutical purposes, the aspirate of bone marrow can be concentrated, thus the term BMAC.

**BMAC**

BMAC is the concentration of whole bone marrow aspirate that can be manufactured in different ways in order to concentrate nucleated cells. Common techniques used to achieve this concentration are the use of Ficoll density gradients, closed centrifugation systems and in-house preparations. It is believed that increased cell concentration would also enhance the amount of MSCs, which could then provide local microenvironment improvement in order to achieve healing and regeneration. Potentially, these MSCs are able to provide a direct cell source for repair. In addition, the nucleated cells may have a significant paracrine effect, releasing and delivering a myriad of cytokines and growth factors to orchestrate tissue repair processes. In comparison to PRP, there is a significant variation in the final products achieved. Fortier et al. evaluated the constituents of PRP and BMAC, showing a reduction in platelet content and an increase in white blood cell content in BMAC. The differences between these products could represent a different mechanism of action. Ziegler also performed a comparison between these products and concluded that BMAC had a significantly higher IL-1 receptor antagonist and therefore offers a more relevant source of anti-inflammatory therapy for OA.

The majority of studies have focused on BMAC for osteonecrosis of the femoral head (ONFH). Hernigou et al. described a technique for injecting BMAC as an adjunct to core decompression (CD). The beneficial effect is attributed not only to disruption of the necrotic zone, but also to the introduction of osteogenic progenitor cells into the femoral head. This study evaluated 189 decompressed hips using BMAC harvested from the iliac crest. The results showed that patients in early stages (Steinberg I-II) had excellent outcomes at 5 years, with only 6.2% of the hips requiring total hip arthroplasty. On the other hand, patients in advanced stages (Steinberg III-IV) had poor outcomes, where 60% required total hip arthroplasty (THA). In addition, it was reported that superior outcomes were verified in patients that presented increased numbers of progenitor cells, suggesting a relationship between the clinical outcomes and their count. Similar results were obtained in the study by Einhorn et al., aimed at patients with symptomatic ONFH stages I-II. The researchers observed minimization of pain, preservation of joint function, and obviation or at least delayed need for THA within 2 years of follow-up. In this study 62%-75% of CD-BMAC recipients successfully avoided the need for additional interventions, including THA.

The use of BMAC has been suggested to restore joint harmony and minimize further chondral deterioration. One advantage of this orthobiologic is that MSCs are ideal for the chondrolabral junction, since these cells are able to differentiate into both fibrocartilage and hyaline-like tissue products. Clinical trials registered in the literature demonstrate the efficacy and safety of BMAC in treating arthritic cartilage symptoms and focal chondral lesions. Additionally, significant cartilage growth, for instance, was also demonstrated with the administration of BMAC in association with other treatment modalities such as microfracture and biologic scaffolds. The use of BMAC for femoracetabular impingement and OA of the hip is scarce. In a case report of a professional soccer player, after two arthroscopies, the patient presented remitting hip pain 3 mo after the second surgery. He was treated with 3 PRP and 2 BMAC injections and was able to return to full activities. Mardones et al. published a technique for the treatment of chondral hip lesions using BMAC and PRP clot. In this technique, hip arthroscopy was performed and microfracture was carried out for full thickness chondral lesions. Bone marrow was harvested and processed, as well as PRP, which was exogenously activated in order to obtain a clot. To finalize the arthroscopic procedure, the fibrin clot was placed over the microfracture site using a cannula. BMAC was inserted under the PRP clot. This technique was used in 13 patients. As a result, the authors reported improvement in the symptoms for a mean of 8 mo of follow-up. Table 1 shows clinical results of the use of bone marrow derivatives for hip disorders.

The results of BMAC in bone, cartilage and tendon injuries are encouraging. We believe that cell characterization is necessary for outcome optimization, and larger randomized controlled trials are highly warranted in order to further support these results. On the other hand, the use of BMAC especially in the early stages of hip diseases resulted in positive results for pain and function, and could represent a promising contemporary treatment strategy.
**ADIPOSE-DERIVED TREATMENTS**

Adipose tissue has a heterogeneous cell population including endothelial cells, endothelial progenitors, pericytes, fibroblasts, mesenchymal stromal cells, macrophages and adipocytes\(^8\). Harvested fresh adipose tissue can undergo minimum manipulation processing or enzymatic digestion and mechanical separation, yielding a final product called the stromal vascular fraction (SVF). The benefits of the use of SVF in pre-clinical and *in vitro* studies have been demonstrated by other authors\(^{60,61}\).

In 2001, Zuk et al\(^6\) identified a stem cell population within human lipoaspirates with the use of collagenase, producing stable growth and proliferation kinetics in the culture. Much like BM-MSCs, such adipose-derived cells differentiated *in vitro* toward osteogenic, myogenic and chondrogenic clusters when treated with lineage specific factors. Thus, this indicates the actual presence of multipotent stem cells with multilineage potential\(^{60,61}\). BM-MSCs have been used as a cellular therapeutic option for the treatment of AVN of the femoral head, but there is still dispute over its clinical success\(^69\). Wyles et al\(^7\) demonstrated that adipose-derived MSCs (AD-MSCs) outperformed BM-MSCs in growth rate and bone differentiation potential in the setting of AVN, suggesting that they could provide a more potent regenerative therapeutic strategy. Periarticular samples of adipose tissue and BM from the femoral canal were obtained from 15 patients undergoing hip replacement for late-stage osteonecrosis. MSCs were isolated from both sources and taken through a standardized cell division protocol to establish cumulative cell count. Proliferation capacity was increased by 4-fold in AD-MSCs in comparison with BM-MSCs after 20 d in culture\(^70\). However, clinical evidence is still limited to case reports, such as the one published by Pak et al\(^68\) in 2011, where two patients were successfully treated for AVN with AD-MSCs (as fresh centrifuged graft) along with HA and PRP. With regard to veterinary medicine, Cuervo et al\(^71\) demonstrated the safety and effectiveness of a single intra-articular injection of AD-MSCs in dogs with hip OA. Functional limitation, range of motion and other scores progressively improved during the 6-mo follow-up period. Additionally, better results were obtained in dogs treated with AD-MSCs than in dogs treated with PRP. Similar results were obtained by Vilar et al\(^72\) where AD-MSC therapy also improved limb function in dogs suffering from hip OA. Table 1 summarizes the results of the clinical trials using adipose-derived treatments for hip disorders.

Clinically, minimally manipulated (microfragmented) fresh adipose tissue has been employed in the treatment of chondral lesions, especially associated with arthroscopic procedures\(^73\) or even for hip OA, but documented in short-follow-up case series\(^74\). The microfragmented adipose-derived fraction (MFAT) is a mechanical technique that reduces the size of the adipose tissue clusters to eliminate oil and blood residue. The presence of MSCs in significant numbers was demonstrated. Nava et al\(^75\) evaluated the *in vitro* survival and content of MSCs and anti-inflammatory activity of lipoaspirate and MFAT. It was noted that MFAT exhibited higher amounts of CD31 positive cells—an endothelial marker, and higher numbers of MSCs in comparison to lipoaspirate. The release of cytokines was similar in the first week of culture. However, the total amount secreted by lipoaspirate decreased much more rapidly than that produced by MFAT after 28 d of culture. When the MFAT culture medium of early (3-7 d) or late culture (28 d) was added to a monocyte culture it strongly inhibited the inflammatory pattern. The authors concluded that MFAT presents a long-lasting effect due to the anti-inflammatory activity attributed to their MSC content. These cells release cytokines that modulate the inflammatory cascade by a variety of mechanisms\(^77\).

In summary, the use of adipose tissue has shown promising results as a viable option for hip disorders, including cultured (AD-MSC) or SVF cells. It is relatively easier to obtain SVF compared to cultured cells. Because of regulatory requirements, availability of approved Good Manufacturing Practice (GMP) facilities, associated costs, cell dosage, processing time or even the number of procedures for the patient, the use of fresh cells is becoming more attractive.

**EXPANDED MSCS**

MSCs were named almost 30 years ago as a class of cells that could be isolated from a variety of tissues, including bone marrow, adipose tissue, dental pulp and umbilical cord. These cells can be expanded in culture maintaining their *in vitro* ability to induce a variety of mesodermal phenotypes and tissues, including differentiation into bone,
cartilage and fat—showing their multi-potent potential[39]. Dominici et al[39], 2006 published an article showing that the Mesenchymal and Tissue Stem Cell Committee of the International Society for Cellular Therapy proposed minimum criteria to define human MSCs. These criteria include: Plastic adherence of MSCs when maintained in culture conditions; positivity for markers CD105, CD73 and CD90, and lack of expression of CD45, CD34, CD14, CD11b, CD79, CD19 and HLA-DR; Differentiation in osteoblasts, adipocytes and chondroblasts in vitro[39]. In 2011, Caplan proposed a change in the name of these cells to medicinal signaling cells (MSC), which might actually be their main role—the ability to secrete bioactive factors, acting as immunomodulatory and trophic factors and showing the great importance of their paracrine effect. Thus, tissue-specific resident stem cells are responsible for the construction of new tissue, stimulated by bioactive factors released by exogenous MSC[33,34].

In 2012, Zhao et al[7] published a paper with 100 patients treated with core decompression in comparison with patients treated with autologous implantation of cultured BM-MSC. The follow-up period was 60 mo after the procedure. As a result, the authors showed a protective effect in the BM-MSC group regarding the progression of osteonecrosis; thus, the cell group presented significantly fewer patients with osteonecrotic stage progression in comparison to CD (core decompression) alone. A decreased necrosis volume was also verified in those patients treated with cells in comparison to CD. The authors concluded that BM-MSC administration is a safe, reliable and highly effective procedure for the treatment of early-stage ONFH[8]. In 2019, Mardones et al[39] published a study evaluating the safety and efficacy of intra-articular infusion of ex vivo expanded autologous BM-MSC in patients with OA of the hip. Ten patients were injected, each one with a dose of 60 × 10^6 cells in three consecutive weekly doses. The follow-up period was 16-40 mo. Patients exhibited a significant improvement in the score evaluation. The radiographic score in general did not change with exception of one improved patient. The authors concluded that three consecutive injections of expanded BM-MSC proved to be a safe and clinically effective treatment in restoration of the range of motion and function of the hip plagued with OA[39].

Table 1 summarizes clinical studies regarding expanded MSCs, mainly from bone marrow origin.

The reviewed clinical studies suggest that intra-articular MSC therapies are safe when used to treat OA, focal chondral defects or even femoral head AVN. However, the efficacy of these therapies cannot be determined until more standardized level 1 clinical evidence is available. Improving study methodology and standardizing cell harvesting, processing, characterization, and delivery techniques will be necessary before the efficacy of intra-articular MSC therapies can be determined. Future randomized double blinded multi-arm clinical studies aimed at determining optimal cell source and count, ideal patient population target as well as optimal method of intra-articular delivery are still required. It is also important to consider the current limiting costs of cultured cells, which may cost up to 30000 euros[39] and may represent a major hindrance to their application.

CONCLUSION

Although many promising alternatives for degenerative hip pathology have been presented, definitive clinical evidence is still lacking in some treatments. Notwithstanding, cell therapies using mononuclear cells derived from bone marrow for femoral head AVN, as well as leukocyte-rich PRP for gluteal tendinopathy have been deemed successful by long-term quality randomized controlled trials. However, more promising therapies aimed at reducing pain and improving function await better clinical scrutiny by further investigation.

REFERENCES

1 Roberts TT, Rosenbaum AJ. Bone grafts, bone substitutes and orthobiologics: the bridge between basic science and clinical advancements in fracture healing. Organogenesis 2012; 8: 114-124 [PMID: 23247591] DOI: 10.4161/org.23306

2 Dhillion RS, Schwarz EM, Maloney MD. Platelet-rich plasma therapy – future or trend? Arthritis Res Ther 2012; 14: 219 [PMID: 22894643 DOI: 10.1186/ar3914

3 Bravo D, Jazrawi L, Cardone DA, Virk M, Passias PG, Einhorn TA, Leucht P. Orthobiologics A
Kruel A et al. Orthobiologics for hip disorders

Comprehensive Review of the Current Evidence and Use in Orthopedic Subspecialties. *Bull Hosp Jt Dis* (2013) 76: 223-231 [PMID: 31513506]

Hussain ZB, Chahla J, LaPrade RF, Mandelbaum BR. Orthobiologics: Today and tomorrow. In: Farr J, Gornall AH. Cartilage Restoration: Practical Clinical Applications. 2nd Edition. Springer, Cham, 2018: 131-142 [DOI: 10.1007/978-3-319-77152-6_11]

Domb BG, LaReau JM, Hammarstedt JE, Gupta A, Stake CE, Redmond JM. Concomitant Hip Arthroscopy and Periacetabular Osteotomy. *Arthroscopy* 2015; 31: 2199-2206 [PMID: 26233270 DOI: 10.1016/j.arthro.2015.06.002]

Hernigou P, Daltrò G, Hernigou J. Hip osteonecrosis: stem cells for life or behead and arthroplasty? *Int Orthop* 2018; 42: 1425-1428 [PMID: 29934715 DOI: 10.1007/s00264-018-0402-4]

Zhao D, Cui D, Wang B, Tian F, Guo L, Yang L, Liu B, Yu Y. Treatment of early stage osteonecrosis of the femoral head with autologous implantation of bone marrow-derived and cultured mesenchymal stem cells. *Bone* 2012; 50: 325-330 [PMID: 22094904 DOI: 10.1016/j.bone.2011.11.002]

Fiz N, Perez JC, Guadilla J, Garate A, Sanchez P, Padilla S, Delgado D, Sanchez M. Intraosseous Infiltration of Platelet-Rich Plasma for Severe Hip Osteoarthritis. *Arthroscopy Tech* 2017; 6: e821-e825 [PMID: 27806837 DOI: 10.1016/j.eats.2017.02.014]

Martin HD, Kelly BT, Leunig M, Philippin MJ, Clohisy JC, Martin RL, Sekiya JK, Pietrobon R, Mohtadi NG, Sampson TG, Safran MR. The pattern and technique in the clinical evaluation of the adult hip: the common physical examination tests of hip specialists. *Arthroscopy* 2010; 26: 161-172 [PMID: 20141979 DOI: 10.1016/j.arthro.2009.07.015]

Ji Y, Ghosh K, Shu XZ, Li B, Sokolov JC, Prestwich GD, Clark RA, Rafailovich MH. Electrospray three-dimensional hyaluronic acid nanofibrous scaffolds. *Biomaterials* 2006; 27: 3782-3792 [PMID: 16556462 DOI: 10.1016/j.biomaterials.2006.02.037]

MEYER K. Chemical structure of hyaluronic acid. *Fed Proc* 1958; 17: 1075-1077 [PMID: 13619775]

Waddell DD. Viscosupplementation with hyaluronans for osteoarthritis of the knee: clinical efficacy and economic implications. *Drugs Aging* 2007; 24: 629-642 [PMID: 17702533 DOI: 10.2165/00002512-200724080-00002]

Prieto JG, Pulido MM, Zapico J, Molina AJ, Gimeno M, Coronel P, Alvarez Al. Comparative study of hyaluronic derivatives: rheological behaviour, mechanical and chemical degradation. *Int J Biol Macromol* 2005; 35: 63-69 [PMID: 15769517 DOI: 10.1016/j.jbiomac.2004.12.003]

Ghosh P, Guidolin D. Potential mechanism of action of intra-articular hyaluronan therapy in osteoarthritis: are the effects molecular weight dependent? *Semin Arthritis Rheum* 2002; 32: 10-37 [PMID: 12219318 DOI: 10.1053/sarh.2002.33720]

Guidolin DD, Ronchetti IP, Lini E, Guerra D, Frizziero L. Morphological analysis of articular cartilage biopsies from a randomized, clinical study comparing the effects of 500-730 kDa sodium hyaluronate (Hyalgan) and methylprednisolone acetate on primary osteoarthritis of the knee. *Osteoarthritis Cartilage* 2001; 9: 371-381 [PMID: 11399102 DOI: 10.1016/j.joca.2000.0393]

Bruyère O, Cooper C, Pelletier JP, Bronco J, Luisa Brandi M, Guillemin F, Hochberg MC, Kanis JA, Kvenient TK, Martel-Pelletier J, Rizzoli R, Silverman S, Reginster JY. An algorithm recommendation for the management of knee osteoarthritis in Europe and internationally: a report from a task force of the European Society for Clinical and Economic Aspects of Osteoporosis and Osteoarthritis (ESCEO). *Semin Arthritis Rheum* 2014; 44: 253-263 [PMID: 24953861 DOI: 10.1016/j.semarthri.2014.05.014]

Lajeunesse D, Delalande A, Martel-Pelletier J, Pelletier JP. Hyaluronic acid reverses the abnormal synthetic activity of human osteoarthritic subchondral bone osteoblasts. *Bone* 2003; 33: 703-710 [PMID: 14555276 DOI: 10.1016/S8756-3282(03)00206-0]

Brun P, Zavan B, Vindigni V, Schiavinato A, Pozzuoli A, Iacobellis C, Abatangelo G. In vitro response of osteoarthritic chondrocytes and fibroblast-like synoviocytes to a 500-730 kDa hyaluronan amide derivative. *J Biomed Mater Res B Appl Biomater* 2012; 100: 2073-2081 [PMID: 22865740 DOI: 10.1002/jbm.b.32771]

Julovi SM, Yasuda T, Shimizu M, Hiramitsu T, Nakamura T. Inhibition of interleukin-1beta-stimulated production of matrix metalloproteinases by hyaluronan via CD44 in human articular cartilage. *Arthritis Rheum* 2004; 50: 516-525 [PMID: 14872494 DOI: 10.1002/art.20045]

Kalaci A, Yilmaz HR, Aslan B, Soğut S, Yanat AN, Uz E. Effects of hyaluronan on nitric oxide levels and superoxide dismutase activities in synovial fluid in knee osteoarthritis. *Clin Rheumatol* 2007; 26: 1306-1311 [PMID: 17180636 DOI: 10.1007/s10067-006-0594-y]

Karna E, Miltyk W, Surzyński A, Palka JA. Protective effect of hyaluronic acid on interleukin-1-induced deregulation of beta1-integrin and insulin-like growth factor-1 receptor signaling and collagen biosynthesis in cultured human chondrocytes. *Mol Cell Biochem* 2008; 308: 57-64 [PMID: 17899316 DOI: 10.1007/s11010-007-9612-5]

Abate M, Pelotti P, De Amicis D, Di Iorio A, Galletti S, Salini V. Viscosupplementation with hyaluronic acid in hip osteoarthritis (a review). *Ups J Med Sci* 2008; 113: 261-277 [PMID: 18991239 DOI: 10.3109/2000-1967-233]

Sladek JK, Kearney C, Cream D, Brama PAJ, Tajber L, Fawcett K, Labbette MC, Leggett B, Brayden DJ. Intra-articular delivery of a nanocomplex comprising salmon calcitonin, hyaluronic acid, and chitosan using an equine model of joint inflammation. *Drug Deliv Transl Res* 2018; 8: 1421-1435 [PMID: 29947020 DOI: 10.1007/s13346-018-0557-x]
24 Migliore A, Massafrica U, Bizi E, Tormenta S, Cassol M, Granata M. Duration of symptom relief after intra-articular injection of hyaluronic acid combined with sorbitol (anti-ox-vs) in symptomatic hip osteoarthritis. *Int J Immunopharmacol* 2014; 27: 245-252 [PMID: 25004836 DOI: 10.1177/03943620140200211]

25 Martins Shimoojo AA, Santos Duarte ADS, Santos Duarte Lana JF, Malheiro Luso AC, Fernandes AR, Sanchez-Lopez E, Barbosa Souto E, Andrade Santana MH. Association of Platelet-Rich Plasma and Auto-Crosslinked Hyaluronic Acid Microparticles: Approach for Orthopedic Application. *Polymers (Basel)* 2019; 11 [PMID: 31561615 DOI: 10.3390/polym11101568]

26 Gobbi A, Whyte GP. One-Stage Cartilage Repair Using a Hyaluronic Acid-Based Scaffold With Activated Bone Marrow-Derived Mesenchymal Stem Cells Compared With Microfatigue: Five-Year Follow-up. *Am J Sports Med* 2016; 44: 2846-2854 [PMID: 27474386 DOI: 10.1177/0363546516656179]

27 Gobbi A, Whyte GP. Long-term Clinical Outcomes of One-Stage Cartilage Repair in the Knee With Hyaluronic Acid-Based Scaffold Embedded With Mesenchymal Stem Cells Sourced From Bone Marrow Aspirate Concentrate. *Am J Sports Med* 2019; 47: 1621-1628 [PMID: 31054502 DOI: 10.1177/0363546518453562]

28 Marx RE. Platelet-rich plasma: evidence to support its use. *J Oral Maxillofac Surg* 2004; 62: 489-496 [PMID: 15085519 DOI: 10.1016/j.joms.2003.12.003]

29 Bennell KL, Hunter DJ, Paterson KL. Platelet-Rich Plasma for the Management of Hip and Knee Osteoarthritis. *Curr Rheumatol Rep* 2017; 19: 24 [PMID: 28386761 DOI: 10.1007/s11926-017-0652-x]

30 Sundman EA, Cole BJ, Karas V, Della Valle C, Tetreault MW, Mohammed HO, Fortier LA. The anti-inflammatory and matrix restorative mechanisms of platelet-rich plasma in osteoarthritis. *Am J Sports Med* 2014; 42: 35-41 [PMID: 24192391 DOI: 10.1177/0363546513507766]

31 Lana JF, Huber SC, Purita J, Tambeli CH, Santos GS, Paulus C, Annichino-Bizzacchi JM. Leukocyte-rich PRP versus leukocyte-poor PRP - The role of monocyte/macrophage function in the healing cascade. *J Clin Orthop Trauma* 2019; 10: S7-S12 [PMID: 31700202 DOI: 10.1016/j.jcot.2019.05.008]

32 Lana JFSD, Purita J, Paulus C, Huber SC, Rodrigues BL, Rodrigues AA, Santana MH, Madureira JL Jr, Malheiro Luso AC, Belangero WD, Annichino-Bizzacchi JM. Contributions for classification of platelet-rich plasma - proposal of a new classification: MARSPILL. *Regen Med* 2017; 12: 565-574 [PMID: 28758836 DOI: 10.2217/rme-2017-0042]

33 Mannava S, Chahlia J, Greenslin AG, Cinque ME, Whitney KE, Evans TA, Fragrangeo SJ, LeBus G, Godin J, LaPrade RF, Philippin MJ. Platelet-Rich Plasma Augmentation for Hip Arthroscopy. *Arthrosc Tech* 2017; 6: e763-e768 [PMID: 28706829 DOI: 10.1016/j.eats.2017.02.001]

34 Dragoo JL, Braun HJ, Durham JL, Ridley BA, Odegard LJ, Luong R, Amodezky SP. Comparison of the acute inflammatory response of two commercial platelet-rich plasma systems in healthy rabbit tendons. *Am J Sports Med* 2012; 40: 1274-1281 [PMID: 22495144 DOI: 10.1177/0363546512442334]

35 Walker-Santiago R, Wojnowski NM, Lall AC, Maldonado DR, Rabe SM, Domb BG. Platelet-Rich Plasma Versus Surgery for the Management of Recalcitrant Greater Trochanteric Pain Syndrome: A Systematic Review. *Arthroscopy* 2020; 36: 875-888 [PMID: 31882270 DOI: 10.1016/j.arthro.2019.09.044]

36 Zhao Z, Ma JX, Ma XL. Different Intra-articular Injections as Therapy for Hip Osteoarthritis: A Systematic Review and Network Meta-analysis. *Arthroscopy* 2020; 36: 1452-1464. e2 [PMID: 31919027 DOI: 10.1016/j.arthro.2019.09.043]

37 Amable PR, Carias RB, Teixeira MV, da Cruz Pacheco I, Corrêa do Amaral RJ, Granjeiro JM, Borogevic R. Platelet-rich plasma preparation for regenerative medicine: optimization and quantification of cytokines and growth factors. *Stem Cell Res Ther* 2013; 4: 67 [PMID: 23759113 DOI: 10.1186/s13287-012-0018-6]

38 Huber SC, Cunha Junior JL, Montalvão S, da Silva LQ, Paffaro AU, da Silva FA, Rodrigues BL, Lana JF, Annichino-Bizzacchi JM. In vitro study of the role of thrombin in platelet rich plasma (PRP) preparation: utility for gel formation and impact in growth factors release. *J Stem Cells Regen Med* 2016; 12: 2-9 [PMID: 27397996 DOI: 10.46582/jscr.1201002]

39 Lewandowski K, Kowalick MM, Pawlaczyc R, Rogowski J, Hellmann A. Microscopic examination of bone marrow aspirate in healthy adults - comparison of two techniques of slide preparation. *Int J Lab Hematol* 2012; 34: 254-261 [PMID: 22145778 DOI: 10.1111/j.1751-553X.2011.01387.x]

40 Sampson S, Botto-van Bemden A, Aufiero D. Autologous bone marrow concentrate: review and application of a novel intra-articular orthobiologic for cartilage disease. *Phys Sportsmed* 2013; 41: 7-18 [PMID: 24413698 DOI: 10.3810/psm.2013.09.2022]

41 Olmsted-Davis EA, Gugal Z, Camargo F, Gannon FH, Jackson K, Kienstra KA, Shine HD, Lindsey RW, Hirsch KK, Goodell MA, Brenner MK, Davis AR. Primitive adult hematopoietic stem cells can function as osteoblast precursors. *Proc Natl Acad Sci USA* 2003; 100: 15877-15882 [PMID: 14673088 DOI: 10.1073/pnas.2632959100]

42 Marx RE, Harrell DB. Translational research: The CD34+ cell is crucial for large-volume bone regeneration from the milieu of bone marrow progenitor cells in cranio mandibular reconstruction. *Int J Oral Maxillofac Implants* 2014; 29: e201-e209 [PMID: 24683583 DOI: 10.11607/jomi.te56]

43 Pettine KA, Suzuki RK, Sand TT, Murphy MB. Autologous bone marrow concentrate intracapsular injection for the treatment of degenerative disc disease with three-year follow-up. *Int Orthop* 2017;
Kruel A et al. Orthobiologics for hip disorders

41: 2097-2103 [PMID: 28743830 DOI: 10.1007/s00264-017-3560-9]

Steinert AF, Rackwitz L, Gilbert F, Nöth U, Tuan RS. Concise review: the clinical application of mesenchymal stem cells for musculoskeletal regeneration: current status and perspectives. Stem Cells Transl Med 2012; 1: 237-247 [PMID: 23197783 DOI: 10.5966/sctm.2011-0036]

Mardones R, Larrain C. Cartilage restoration technique of the hip. J Hip Preserv Surg 2016; 3: 30-36 [PMID: 27028616 DOI: 10.1093/jhps/hnv061]

Hernigou P, Guerin G, Homma Y, Dubory A, Chevallier N, Rouard H, Flouzat Lachaniette CH. History of concentrated or expanded mesenchymal stem cells for hip osteonecrosis: is there a target number for osteonecrosis repair? Int Orthop 2018; 42: 1739-1745 [PMID: 29796764 DOI: 10.1007/s00264-018-4000-1]

Di Bella C, Aldini NN, Lucarelli E, Dozza B, Frisoni T, Martini L, Fini M, Donati D. Osteogenic protein-1 associated with mesenchymal stem cells promote bone allograft integration. Tissue Eng Part A 2010; 16: 2967-2976 [PMID: 20533882 DOI: 10.1089/ten.tea.2009.0637]

Fennema EM, Renard AJ, Leusink S, van Blitterswijk CA, de Boer J. The effect of bone marrow aspiration strategy on the yield and quality of human mesenchymal stem cells. Acta Orthop 2009; 80: 618-621 [PMID: 19916699 DOI: 10.3109/17453670903278241]

Pierini M, Di Bella C, Dozza B, Frisoni T, Martella E, Bellotti C, Remondini D, Lucarelli E, Giannini S, Donati D. The posterior iliac crest outperforms the anterior iliac crest when obtaining mesenchymal stem cells from bone marrow. J Bone Joint Surg Am 2013; 95: 1101-1107 [PMID: 23783207 DOI: 10.2106/JBJS.L.00429]

Hernigou P, Homma Y, Flouzat Lachaniette CH, Poignard A, Allain J, Chevallier N, Rouard H. Benefits of small volume and small syringe for bone marrow aspirations of mesenchymal stem cells. Int Orthop 2013; 37: 2279-2287 [PMID: 23881064 DOI: 10.1007/s00264-013-1722-z]

Imam MA, Holton J, Horriot S, Negida AS, Gruhhofer F, Gupta R, Narvani A, Snow M. A systematic review of the concept and clinical applications of bone marrow aspirate concentrate in tendinopathy. SICOT J 2017; 3: 58 [PMID: 28990375 DOI: 10.1016/j.sicot.2017.0732]

Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR. Multilineage potential of adult human mesenchymal stem cells. Science 1999; 284: 143-147 [PMID: 10102814 DOI: 10.1126/science.284.5411.143]

Hernigou P, Poignard A, Beaupre F, Rouard H. Percutaneous autologous bone-marrow grafting for nonunions. Influence of the number and concentration of progenitor cells. J Bone Joint Surg Am 2005; 87: 1430-1437 [PMID: 15951108 DOI: 10.2106/JBJS.D.02215]

Lee DH, Ryu KJ, Kim JW, Kang KC, Choi YR. Bone marrow aspirate concentrate and platelet-rich plasma enhanced bone healing in distraction osteogenesis of the tibia. Clin Orthop Relat Res 2014; 472: 3789-3797 [PMID: 24599650 DOI: 10.1199/cior.2013-0206]

Jäger M, Jelinek EM, Wess KM, Scharfstädt A, Jacobson M, Kevy SV, Krauspe R. Bone marrow treatment of stage I and II osteonecrosis of the femoral head with core decompression and bone marrow grafting. Acta Orthop 2012; 83: 19149628 DOI: 10.2147/157488809787160909]

Fortier LA, Potter HG, Rickel EJ, Schnabel LV, Foo LF, Chong LR, Stokol T, Cheetham J, Nixon AJ. Concentrated bone marrow aspirate improves full-thickness cartilage repair compared with microfracture in the equine model. J Bone Joint Surg Am 2010; 92: 1927-1937 [PMID: 20720135 DOI: 10.2106/JBJS.L.01284]

Ziegler CG, Van Sloun R, Gonzalez S, Whitney KE, DePhillipo NN, Kennedy MI, Dornan GJ, Evans TA, Huard J, LaPrade RF. Characterization of Growth Factors, Cytokines, and Chemokines in Bone Marrow Concentrate and Platelet-Rich Plasma: A Prospective Analysis. J Am J Sports Med 2019; 47: 2174-2178 [PMID: 31034242 DOI: 10.1177/0363546519832003]

Hernigou P, Beaujean F. Treatment of osteonecrosis with autologous bone marrow grafting. Clin Orthop Relat Res 2002; 14-23 [PMID: 12461352 DOI: 10.1097/00003086-200212000-00003]

Einhorn TA, Huard J, LaPrade RF. Characterization of Growth Factors, Cytokines, and Chemokines in Bone Marrow Concentrate and Platelet-Rich Plasma: A Prospective Analysis. J Am J Sports Med 2019; 47: 2174-2178 [PMID: 31034242 DOI: 10.1177/0363546519832003]

Hernigou P, Beaujean F. Treatment of osteonecrosis with autologous bone marrow grafting. Clin Orthop Relat Res 2002; 14-23 [PMID: 12461352 DOI: 10.1097/00003086-200212000-00003]

Einhorn TA, Huard J, LaPrade RF. Characterization of Growth Factors, Cytokines, and Chemokines in Bone Marrow Concentrate and Platelet-Rich Plasma: A Prospective Analysis. J Am J Sports Med 2019; 47: 2174-2178 [PMID: 31034242 DOI: 10.1177/0363546519832003]

Hernigou P, Beaujean F. Treatment of osteonecrosis with autologous bone marrow grafting. Clin Orthop Relat Res 2002; 14-23 [PMID: 12461352 DOI: 10.1097/00003086-200212000-00003]

Einhorn TA, Huard J, LaPrade RF. Characterization of Growth Factors, Cytokines, and Chemokines in Bone Marrow Concentrate and Platelet-Rich Plasma: A Prospective Analysis. J Am J Sports Med 2019; 47: 2174-2178 [PMID: 31034242 DOI: 10.1177/0363546519832003]

Hernigou P, Beaujean F. Treatment of osteonecrosis with autologous bone marrow grafting. Clin Orthop Relat Res 2002; 14-23 [PMID: 12461352 DOI: 10.1097/00003086-200212000-00003]

Einhorn TA, Huard J, LaPrade RF. Characterization of Growth Factors, Cytokines, and Chemokines in Bone Marrow Concentrate and Platelet-Rich Plasma: A Prospective Analysis. J Am J Sports Med 2019; 47: 2174-2178 [PMID: 31034242 DOI: 10.1177/0363546519832003]
concentrate therapy. Knee Surg Sports Traumatol Arthrosc; 2013; 21: 1684-1688 [PMID: 23052123 DOI: 10.1007/s00167-012-2232-y]

65 Lockhart RA, Aronowitz JA, Dos-Anjos Vilaboa S. Use of Freshly Isolated Human Adipose Stromal Cells for Clinical Applications. Aesthet J 2017; 37: S4-S8 [PMID: 29025212 DOI: 10.1093/aesth/sjw270]

66 Jurgens WJ, Krooe R, Zandieh-Doulati B, van Dijk A, Renders GA, Smit TH, van Milligen FJ, Ritt MJ, Helder MN. One-step surgical procedure for the treatment of osteochondral defects with adipose-derived stem cells in a caprine knee defect: a pilot study. Biores Open Access 2013; 2: 315-325 [PMID: 23914338 DOI: 10.1089/biores.2013.0024]

67 Wu L, Prins HJ, Leijten J, Helder MN, Evseenko D, Moroni L, van Blitterswijk CA, Lin Y, Karperien M. Chondrocytes Co-cultured with Stromal Vascular Fraction of Adipose Tissue Present More Intense Chondrogenic Characteristics Than with Adipose Stem Cells. Tissue Eng Part A 2016; 22: 336-348 [PMID: 26732248 DOI: 10.1089/ten.TEA.2015.0269]

68 Zuk PA, Zhu M, Mizuno H, Huang J, Futrell JW, Katz AJ, Benhaim P, Lorenz HP, Hedrick MH. Multipotent cells from human adipose tissue: implications for cell-based therapies. Tissue Eng 2001; 7: 211-228 [PMID: 11304456 DOI: 10.1089/106136901300062859]

69 Zuk PA, Zhu M, Ashjian P, De Ugarte DA, Huang JI, Mizuno H, Alfonso ZC, Fraser JK, Benhaim P, Hedrick MH. Human adipose tissue is a source of multipotent stem cells. Mol Biol Cell 2002; 13: 4279-4295 [PMID: 12475952 DOI: 10.1091/mbc.E02-02-0102]

70 Torres-Torrellas M, Rubio M, Damia E, Cuervo B, Del Romero A, Peláez P, Chicharro D, Miguel L, Sopena JJ. Adipose-Derived Mesenchymal Stem Cells: A Promising Tool in the Treatment of Musculoskeletal Diseases. Int J Mol Sci 2019; 20 [PMID: 31242644 DOI: 10.3390/ijms20123105]

71 Wyles CC, Houdek MT, Crespo-Díaz R, Norambuena GA, Stalboerger PG, Terzic A, Behfar A, Sierra RJ. Adipose-derived Mesenchymal Stem Cells Are Phenotypically Superior for Regeneration in the Setting of Osteonecrosis of the Femoral Head. Clin Orthop Relat Res 2015; 473: 3080-3090 [PMID: 26070774 DOI: 10.1097/jhpr.0000000000000485]

72 Pak J. Regeneration of human bones in hip osteonecrosis and human cartilage in knee osteoarthritis with autologous adipose-tissue-derived stem cells: a case series. J Med Case Rep 2011; 5: 296 [PMID: 21736710 DOI: 10.1186/1757-1626-5-296]

73 Cuervo B, Rubio M, Sopena J, Dominguez JM, Vilar J, Morales M, Cugat R, Carrillo JM. Hip osteoarthritis in dogs: a randomized study using mesenchymal stem cells from adipose tissue and plasma rich in growth factors. Int J Mol Sci 2014; 15: 13437-13460 [PMID: 25089877 DOI: 10.3390/ijms150813437]

74 Vilári JM, Batista M, Morales M, Santana A, Cuervo B, Rubio M, Cugat R, Sopena J, Carrillo MJ. Assessment of the effect of intraarticular injection of autologous adipose-derived mesenchymal stem cells in osteoarthritic dogs using a double blinded force platform analysis. BMC Vet Res 2014; 10: 143 [PMID: 24948756 DOI: 10.1186/1746-6148-10-143]

75 Jannelli E, Fontana A. Arthroscopic treatment of chondral defects in the hip: AMIC, MACI, microfragmented adipose tissue transplantation (MATT) and other options. NICOT J 2017; 3: 43 [PMID: 28538977 DOI: 10.1051/sicotd/2017029]

76 Barfold KW, Blond L. Treatment of osteoarthritis with autologous and microfragmented adipose tissue. Dan Med J 2019; 66 [PMID: 31571571]

77 Nava S, Sorli V, Pasucci L, Tremolada C, Ciassani E, Zeira O, Cadei M, Soldati G, Pessina A, Parati E, Slevin M, Alessandri G. Long-Lasting Anti-Inflammatory Activity of Human Microfragmented Adipose Tissue. Stem Cells Int 2019; 2019: 5901479 [PMID: 30915125 DOI: 10.1155/2019/5901479]

78 Caplan AI. Mesenchymal Stem Cells: Time to Change the Name! Stem Cells Transl Med 2017; 6: 1445-1451 [PMID: 28452204 DOI: 10.1002/sctm.17-0051]

79 Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, Krause D, Deans R, Keating A, Prokop Dj, Horwitz E. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. Cytotherapy 2006; 8: 315-317 [PMID: 16923606 DOI: 10.1080/14653240600855905]

80 Caplan AI. What’s in a name? Tissue Eng Part A 2010; 16: 2415-2417 [PMID: 20412005 DOI: 10.1089/ten.TEA.2010.0216]

81 Mardones R, Camacho D, Monsalvo F, Zulch N, Jofré C, Minguell JJ. Treatment of osteonecrosis of the femoral head by core decompression and implantation of fully functional ex vivo-expanded bone marrow-derived mesenchymal stem cells: a proof-of-concept study. Stem Cells Cloning 2019; 12: 11-16 [PMID: 30881048 DOI: 10.2147/SCCA.A.S18183]

82 Mardones R, Jofre CM, Tobar L, Minguell JJ. Mesenchymal stem cell therapy in the treatment of hip osteoarthritis. J Hip Preserv Surg 2017; 4: 159-163 [PMID: 28630737 DOI: 10.1093/jhps/lhx011]

83 Freitag J, Bates D, Wickham J, Shah K, Huguenin L, Tenen A, Paterson K, Boyd R. Adipose-derived mesenchymal stem cell therapy in the treatment of knee osteoarthritis: a randomized controlled trial. Regen Med 2019; 14: 213-230 [PMID: 30762487 DOI: 10.2217/reme-2018-0161]

84 Battaglia M, Guaraldi F, Vannini F, Rossi G, Timoncini A, Buda R, Giannini S. Efficacy of ultrasound-guided intra-articular injections of platelet-rich plasma versus hyaluronic acid for hip osteoarthritis. Orthopedics 2013; 36: e1501-e1508 [PMID: 24579221 DOI: 10.3928/01477447-20131120-13]

85 Pogliacomi F, Schiavi P, Parasekopoulos A, Leibeg M, Pedrazzini A, Ceccarelli F, Vaienti E. When is indicated visscous supplementation in hip osteoarthritis? Acta Biomed 2018; 90: 67-74 [PMID: 30715001 DOI: 10.23756/abm.v90i1.S.8000]
86 Clementi D, D'Ambrosi R, Bertocco P, Bucco MS, Cardile C, Ragni P, Giaffreda G, Ragone V. Efficacy of a single intra-articular injection of ultra-high molecular weight hyaluronic acid for hip osteoarthritis: a randomized controlled study. *Eur J Orthop Surg Traumatol* 2018; **28**: 915-922 [PMID: 29164399 DOI: 10.1007/s00590-017-2083-9]

87 Dallari D, Stagni C, Rani N, Sabbioni G, Pelotti P, Torricelli P, Tschon M, Giavaresi G. Ultrasound-Guided Injection of Platelet-Rich Plasma and Hyaluronic Acid, Separately and in Combination, for Hip Osteoarthritis: A Randomized Controlled Study. *Am J Sports Med* 2016; **44**: 664-671 [PMID: 26797697 DOI: 10.1177/0363546515620383]

88 Fitzpatrick J, Bulsara MK, O'Donnell J, Zheng MH. Leucocyte-Rich Platelet-Rich Plasma Treatment of Gluteus Medius and Minimus Tendinopathy: A Double-Blind Randomized Controlled Trial With 2-Year Follow-up. *Am J Sports Med* 2019; **47**: 1130-1137 [PMID: 30840831 DOI: 10.1177/0363546519826969]

89 Houdek MT, Wyles CC, Collins MS, Howe BM, Terzic A, Behfar A, Sierra RJ. Stem Cells Combined With Platelet-rich Plasma Effectively Treat Corticosteroid-induced Osteonecrosis of the Hip: A Prospective Study. *Clin Orthop Relat Res* 2018; **476**: 388-397 [PMID: 29529674 DOI: 10.1007/s11999-0000000000000033]
