Meal Frequencies Modify the Effect of Common Genetic Variants on Body Mass Index in Adolescents of the Northern Finland Birth Cohort 1986

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Abstract

Recent studies suggest that meal frequencies influence the risk of obesity in children and adolescents. It has also been shown that multiple genetic loci predispose to obesity already in youth. However, it is unknown whether meal frequencies could modulate the association between single nucleotide polymorphisms (SNPs) and the risk of obesity. We examined the effect of two meal patterns on weekdays –5 meals including breakfast (regular) and ≤4 meals with or without breakfast (meal skipping) – on the genetic susceptibility to increased body mass index (BMI) in Finnish adolescents. Eight variants representing 8 early-life obesity-susceptibility loci, including FTO and MC4R, were genotyped in 2213 boys and 2449 girls aged 16 years from the population-based Northern Finland Birth Cohort 1986. A genetic risk score (GRS) was calculated for each individual by summing the number of BMI-increasing alleles across the 8 loci. Weight and height were measured and dietary data were collected using self-administered questionnaires. Among meal skippers, the difference in BMI between high-GRS and low-GRS (<8 and ≥8 BMI-increasing alleles) groups was 0.90 (95% CI 0.63,1.17) kg/m², whereas in regular eaters, this difference was 0.32 (95% CI 0.06,0.57) kg/m² (pinteraction = 0.003). The effect of each MC4R rs17782313 risk allele on BMI in meal skippers (0.47 [95% CI 0.22,0.73] kg/m²) was nearly three-fold compared with regular eaters (0.18 [95% CI -0.06,0.41] kg/m²) (pinteraction = 0.016). Further, the per-allele effect of the FTO rs1421085 was 0.24 (95% CI 0.05,0.42) kg/m² in regular eaters and 0.46 (95% CI 0.27,0.66) kg/m² in meal skippers but the interaction between FTO genotype and meal frequencies on BMI was significant only in boys (pinteraction = 0.015). In summary, the regular five-meal pattern attenuated the increasing effect of common SNPs on BMI in adolescents. Considering the epidemic of obesity in youth, the promotion of regular eating may have substantial public health implications.

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Introduction

Prevention of obesity in children has been proposed as a public health priority to combat the obesity epidemic. However, the etiology of childhood obesity is multifaceted with both lifestyle and genetic factors playing a role in the susceptibility to excessive weight gain [1].

In recent years, numerous obesity-related genetic loci have been identified through genome-wide association studies (GWAS) [2]. Although most of the GWAS for obesity have focused on adult body mass index (BMI), several adult-discovered genetic determinants contribute to common childhood obesity as well [3–6]. Among the well-established genetic factors that influence weight development already in childhood are the common single nucleotide polymorphisms (SNPs) in the FTO and MC4R gene regions [7–10].

The dietary causes of obesity are complex and incompletely understood. In relation to childhood and adolescent obesity, it has been proposed that the impact of overall eating patterns may be more significant than that of single foods or nutrients [11]. Some studies indicate that higher meal frequencies and regular breakfast consumption are inversely associated with obesity in youth.
Studies on interaction between genetic and lifestyle factors on obesity are emerging, although to a lesser extent in young populations than in adults. In Spanish children, dietary fat composition was found to modify the association between the FTO gene variant rs9939609 and obesity risk [16]. A recent meta-analysis including both adults and children concluded that physical activity attenuated the effect of FTO variants on obesity risk in adults but not in children [17]. However, our previous study on variation in the FTO gene in the Northern Finland Birth Cohort 1986 (NFBC1986) indicated that in highly physically active adolescents the risk of higher BMI was significantly attenuated even among those carrying two risk alleles; nearly down to the level of those having no risk alleles [10]. These results suggest that the effect of common obesity-susceptibility gene variants could be modified by environmental factors.

Previously, we reported an association of a regular five-meal-a-day pattern with reduced risks of overweight/obesity and abdominal obesity in youth [18]. In the present study, we evaluate the impact of two meal frequencies, i.e., 5 meals a day and ≤4 meals a day, on the association between obesity-related genotypes and body mass index among 16-year-old Finnish adolescents. The results indicate that regular meal frequency attenuates genetic predisposition to increased BMI in terms of both single gene variants (FTO rs1421085 and MC4R rs17782313) and a multiple-locus indicator (a genetic risk score based on eight obesity-susceptibility loci).

Materials and Methods

Ethics statement

The study was approved by the Ethics Committee of the Faculty of Medicine of the University of Oulu and written informed consent was obtained from both adolescents and their parents according to the Declaration of Helsinki.

Study population

The Northern Finland Birth Cohort 1986 (NFBC1986) is an ongoing, population-based study which at the baseline comprised 9432 infants born alive in the two northernmost provinces of Finland to women with expected delivery dates between July 1, 1985 and June 30, 1986, covering 99% of all eligible births in the region. Data have been collected prospectively since the prenatal period, as previously described [19]. At the 16-year follow-up in 2001–2002, 80% (n = 7344) of the adolescents responded to a postal questionnaire concerning their health behavior and well-being and 74% (n = 6798) participated in a clinical examination. The current analysis included individuals for whom data on height, weight, stage of puberty, meal frequency on weekdays at 16 years, and all chosen BMI-related SNPs were available (2215 boys and 2449 girls).

Clinical examination

In 2001–2002, a clinical examination on adolescents was carried out in municipalities of Northern Finland and also in major cities elsewhere in Finland. The trained study personnel (three teams each consisting of one laboratory analyst and two study nurses) performed the examinations according to a standardized protocol. Venous blood samples were drawn for DNA extraction and anthropometric measurements (height in centimetres, weight in kilograms to one decimal place) were performed. BMI was calculated as weight in kilograms divided by height in meters squared. At the clinical examination, the adolescents self-assessed their pubertal maturation using gender-specific line drawings of the Tanner puberty stages [20].

Meal patterns

At the 16-year follow-up, adolescents filled in a postal questionnaire on their health behavior and well-being including a question about meal consumption on weekdays. The frequency of meals was assessed with a question ‘Do you usually have the following meals (breakfast, lunch, snack, dinner, and evening snack) on weekdays?’ with dichotomous response options (yes/no). For the analyses, adolescents were classified into two groups by their meal consumption: five meals a day including breakfast (regular meal pattern) or ≤ four meals a day (meal skipping), the latter including both regular breakfast eaters and breakfast skippers.

Genotyping

We genotyped eight SNPs, rs1421085, rs17782313, rs6265, rs10938397, rs1424233, rs8548238, rs11084753, and rs2815732, representing the childhood obesity-susceptibility loci, identified also in recent GWAS [2–6], at or near the FTO (fat mass- and obesity-associated), MC4R (melanocortin 4 receptor), BDNF (brain-derived neurotrophic factor), GNPDA2 (glucosamine-6-phosphate deaminase 2), MAP (v-musculoaponeurotic fibrosarcoma oncogene homolog), TMEM18 (transmembrane protein 18), KCTD15 (potassium channel tetramerization domain containing 15), and NEGR1 (neuronal growth regulator 1) genes, respectively. The DNA extractions, sample quality controls, biobank up-keeping and aliquoting were performed in the National Public Health Institute, Biomedicum Helsinki, Finland. Genotyping was performed using TaqMan single nucleotide polymorphism assay (Applied Biosystems, Foster City, CA). The Ensembl gene IDs for the reported genes are: FTO: ENSG00000140718; MC4R: ENSG00000166603; BDNF: ENSG00000176697; GNPDA2: ENSG00000163281; MAP: ENSG00000178573; TMEM18: ENSG00000151353; KCTD15: ENSG00000153885; NEGR1: ENSG00000172260.

Statistical analyses

For the genetic risk score (GRS), we used the above eight SNPs representing BMI-associated loci. SNP genotypes were recoded as 0, 1, or 2 BMI-increasing alleles and the risk score was created by adding up the number of these alleles. Due to lack of established effect sizes for each of the SNPs and since it is unadvisable to generate weights from the data under analysis [21], the risk alleles were not weighted by their effect size. For the analysis of interaction between genetic susceptibility and meal frequencies, the sample was divided into high-risk and low-risk groups using the median value of the GRS (8) as the cut-off point. For the FTO and MC4R variants, the interaction effects of the genotype and meal frequencies on BMI were also analyzed separately using an additive model of inheritance. BMI was treated as a continuous variable and gene-diet interactions were investigated using linear regression analysis (GRS as a continuous variable) and analysis of variance (ANOVA; genotypes as categorical variables) adjusted for gender and Tanner stage of puberty (numbered 1–5). Hardy-Weinberg equilibrium and associations between genotypes and meal frequencies were tested using the chi-squared test. In order to avoid excessive testing for statistical significance, the results are reported as mean values with either their 95% confidence intervals (CI) which demonstrate reliability of the estimates or their standard deviations (SD). The 19 p-values related to testing of pre-set hypotheses in this paper have not been corrected for
Results

All variants passed the quality control criteria (call rate >95% and in Hardy-Weinberg equilibrium $p>0.05$) and had minor allele frequency $\geq 0.16$ (Table 1). The mean BMI in the total sample was 21.2 kg/m$^2$ (SD 3.4) and the mean age of adolescents was 16.0 (SD 0.4) years. Table 2 shows mean values of BMI across genotype, meal pattern and puberty stage categories. For the individuals with GRS $\geq 8$ (high-risk group), the mean BMI was 0.7 units greater (21.5 [95% CI 21.3, 21.6] kg/m$^2$) than that for those with GRS <8 (low-risk group) (20.8 [95% CI 20.7, 21.0] kg/m$^2$). Carriers of two risk alleles in FTO rs1421085 had an increased BMI (21.7 [95% CI 21.5, 22.0] kg/m$^2$) compared with individuals with 0 or 1 risk allele (20.9 [95% CI 20.8, 21.1] kg/m$^2$ and 21.2 [95% CI 21.0, 21.3] kg/m$^2$, respectively). Similarly, carrying both of the risk-conferring alleles of rs17782313 at the MC4R locus was associated with a greater BMI (22.2 [95% CI 21.6, 22.9] kg/m$^2$) compared with the other two genotypes (TT: 21.1 [95% CI 21.0, 21.2] kg/m$^2$ and CT: 21.3 [95% CI 21.1, 21.5] kg/m$^2$).

Initially, meal frequency variable consisted of three categories, i.e., five meals including breakfast, four meals including breakfast (semi-regular meal pattern) and four meals not including breakfast (breakfast skipping) [18]. As there was no difference in the mean BMI between the latter two groups (21.5 [95% CI 21.3, 21.7] kg/m$^2$ and 21.7 [95% CI 21.5, 22.0] kg/m$^2$), respectively) they were combined into one category characterized by meal skipping. Regular eaters had lower BMI (20.7 [95% CI 20.6, 20.8] kg/m$^2$) than meal skippers (21.6 [95% CI 21.5, 21.7] kg/m$^2$) (Table 2). There was no association between the two meal patterns and GRS categories ($p = 0.292$, FTO rs1421085 ($p = 0.855$) or MC4R rs17782313 ($p = 0.396$).

Among the whole population, in the linear regression analysis adjusted for gender and pubertal stage, each additional BMI-increasing allele in the GRS was associated with a 0.21 (95% CI 0.16, 0.26) kg/m$^2$ increase in BMI. We then looked at the effects of GRS on BMI separately for the two meal patterns and found that among meal skippers the increase in BMI was 0.27 (95% CI 0.20, 0.35) kg/m$^2$ per risk allele, whereas among regular eaters this effect was only 0.15 (95% CI 0.08, 0.22) kg/m$^2$ ($p_{interaction} = 0.020$; Figure 1).

We then examined interaction effects of meal frequencies and genotype groups (high- and low-risk groups based on GRS, and FTO rs1421085 and MC4R rs17782313 genotypes under an additive model) on the mean values of BMI using ANOVA adjusted for gender and pubertal stage. Among the meal skippers, the difference in BMI between the individuals with a high GRS ($\geq 8$ BMI-increasing alleles) and those with a low GRS (<8 BMI-increasing alleles) was 0.90 (95% CI 0.63, 1.17) kg/m$^2$, whereas in the regular eaters, this difference was only 0.32 (95% CI 0.06, 0.57) kg/m$^2$ ($p_{interaction} = 0.003$; Figure 2). Similarly, the difference in BMI between the individuals carrying two copies of the FTO rs1421085 risk alleles and those with no risk allele was pronounced in the meal skippers (0.98 kg/m$^2$) compared with regular eaters (0.54 kg/m$^2$) but the interaction was non-significant ($p_{interaction} = 0.288$; Figure 3). Per-allele effects of the FTO variant were 0.24 (95% CI 0.05, 0.42) kg/m$^2$ for regular eaters and 0.46 (0.27, 0.66) kg/m$^2$ for meal skippers. However, gender-stratified analysis showed that the interaction between the FTO rs1421085 and meal frequencies on BMI was significant in boys ($p_{interaction} = 0.015$) but not in girls ($p_{interaction} = 0.617$) (data not shown). Furthermore, the difference in BMI between the carriers of the two MC4R rs17782313 risk alleles and non-carriers was elevated to 1.92 kg/m$^2$ in meal skippers, whereas in regular eaters, the difference was reduced to 0.34 kg/m$^2$ ($p_{interaction} = 0.016$; Figure 4). Per-allele effects of the MC4R variant were 0.18 (95% CI -0.06, 0.41) kg/m$^2$ for regular eaters and 0.47 (95% CI 0.22, 0.73) kg/m$^2$ for meal skippers.

Discussion

In the present study, we investigated the association of eight common SNPs with BMI and the possible modifying effect of meal frequency on genetic susceptibility to obesity in a genetically and culturally homogeneous population of 16-year-old Finnish adolescents. Besides using a genetic risk score, we separately analyzed effects of two well-established obesity loci, FTO and MC4R, on BMI. We showed that regular meal frequency attenuates genetic predisposition to increased BMI in terms of both single gene variants and a multiple-locus indicator. For the FTO rs1421085 variant, gender-specific interaction effects were observed.

As a whole, each additional BMI-increasing allele in the genetic risk score was associated with an increase in BMI of 0.21 kg/m$^2$ which corresponds to a 0.61 kg increase in body weight for a person of 170 cm height. In adolescents who ate five meals a day, the BMI increase per risk allele was attenuated to 0.15 kg/m$^2$ (0.43 kg), whereas among meal skippers this effect was increased to 0.27 kg/m$^2$ (0.78 kg). A significant modifying effect of the regular meal frequency was observed also using the genetic risk score as a dichotomous variable and comparing high- and low-risk groups.

Table 1. Genotype information and quality control statistics for the eight childhood obesity-susceptibility SNPs.

| SNP       | Chromosome | Nearest gene | Position (bp) | Risk allele | Non-risk allele | MAF   | HWE p-value | Original reported lead SNP | Reference |
|-----------|------------|--------------|---------------|-------------|----------------|-------|-------------|---------------------------|-----------|
| rs1421085 | 16         | FTO          | 53,800,954    | C           | T              | 0.41  | 0.59        | rs9939609                 | [3,6]     |
| rs17782313| 18         | MC4R         | 57,851,097    | C           | T              | 0.18  | 0.72        | rs17782313                | [3,6]     |
| rs6265    | 11         | BDNF         | 27,679,916    | T            | C              | 0.16  | 0.49        | rs925946                  | [3]       |
| rs10938397| 4          | GNPDA2       | 45,182,527    | G            | A              | 0.49  | 0.68        | rs10938397                | [3]       |
| rs1424233 | 16         | MAF           | 79,682,751    | T            | C              | 0.43  | 0.94        | rs1424233                 | [6]       |
| rs6548238 | 2          | TMEM18       | 634,905       | T            | C              | 0.16  | 0.98        | rs6548238                 | [3]       |
| rs11084753| 19         | KCTD15       | 34,322,137    | G            | A              | 0.34  | 0.64        | rs11084753                | [3]       |
| rs2815752 | 1          | NEGR1        | 72,812,440    | A            | G              | 0.36  | 0.63        | rs2815752                 | [3]       |

Abbreviations: bp, base pairs; HWE, Hardy-Weinberg equilibrium; MAF, minor allele frequency; SNP, single nucleotide polymorphism.
Table 2. Distributions of eight single nucleotide polymorphisms, genetic predisposition score, Tanner stages of puberty, meal patterns and body mass index among adolescents in the Northern Finland Birth Cohort 1986.

|                      | Boys (n = 2215) | Girls (n = 2449) | All (n = 4664) |
|----------------------|-----------------|------------------|----------------|
|                      | Mean BMI kg/m² (95% CI) | % | Mean BMI kg/m² (95% CI) | % | Mean BMI kg/m² (95% CI) | % |
| **FTO rs1421085**    |                 |                 |                 |
| TT                   | 20.8 (20.5, 21.0) | 35.9 | 21.1 (20.8, 21.3) | 34.9 | 20.9 (20.8, 21.1) | 35.4 |
| CT                   | 21.1 (20.9, 21.3) | 47.9 | 21.2 (21.0, 21.4) | 48.2 | 21.2 (21.0, 21.3) | 48.0 |
| CC                   | 21.8 (21.4, 22.2) | 16.2 | 21.7 (21.3, 22.0) | 16.9 | 21.7 (21.5, 22.0) | 16.6 |
| **MC4R rs17782313**  |                 |                 |                 |
| TT                   | 21.1 (20.9, 21.2) | 69.1 | 21.1 (20.9, 21.2) | 67.5 | 21.1 (21.0, 21.2) | 68.2 |
| CT                   | 21.1 (20.8, 21.4) | 27.8 | 21.5 (21.2, 21.7) | 29.2 | 21.3 (21.1, 21.5) | 28.5 |
| CC                   | 21.9 (21.1, 22.7) | 3.1  | 22.5 (21.6, 23.5) | 3.3  | 22.2 (21.6, 22.9) | 3.2  |
| **BDNF rs6265**      |                 |                 |                 |
| CC                   | 21.0 (20.9, 21.2) | 70.1 | 21.2 (21.1, 21.4) | 70.3 | 21.1 (21.0, 21.2) | 70.2 |
| CT                   | 21.3 (21.0, 21.5) | 27.2 | 21.2 (21.0, 21.5) | 27.2 | 21.2 (21.1, 21.4) | 27.2 |
| TT                   | 21.4 (20.5, 22.4) | 2.7  | 21.3 (20.4, 22.2) | 2.5  | 21.4 (20.7, 22.0) | 2.6  |
| **GNPDA2 rs10938397**|                 |                 |                 |
| AA                   | 21.1 (20.9, 21.4) | 26.4 | 20.9 (20.7, 21.1) | 26.9 | 21.0 (20.9, 21.2) | 26.7 |
| AG                   | 21.0 (20.8, 21.2) | 50.0 | 21.3 (21.1, 21.5) | 48.5 | 21.2 (21.0, 21.3) | 49.2 |
| GG                   | 21.2 (20.9, 21.6) | 23.6 | 21.4 (21.1, 21.7) | 24.6 | 21.3 (21.1, 21.5) | 24.1 |
| **MAF rs1424233**    |                 |                 |                 |
| CC                   | 21.1 (20.9, 21.4) | 32.3 | 21.0 (20.8, 21.2) | 32.0 | 21.1 (20.9, 21.2) | 32.2 |
| CT                   | 21.0 (20.7, 21.2) | 48.7 | 21.3 (21.1, 21.5) | 50.4 | 21.1 (21.0, 21.3) | 49.6 |
| TT                   | 21.5 (21.1, 21.8) | 19.0 | 21.4 (21.1, 21.8) | 17.6 | 21.5 (21.2, 21.7) | 18.2 |
| **TMEM18 rs6548238** |                 |                 |                 |
| TT                   | 20.2 (19.3, 21.2) | 2.1  | 20.5 (20.0, 21.1) | 2.5  | 20.4 (19.9, 20.9) | 2.3  |
| CT                   | 21.1 (20.8, 21.3) | 27.1 | 21.2 (20.9, 21.4) | 25.8 | 21.1 (20.9, 21.3) | 26.4 |
| CC                   | 21.2 (21.0, 21.3) | 70.8 | 21.3 (21.1, 21.4) | 71.7 | 21.2 (21.1, 21.3) | 71.2 |
| **KCTD15 rs11084753**|                 |                 |                 |
| AA                   | 21.0 (20.5, 21.4) | 12.2 | 21.1 (20.8, 21.5) | 11.2 | 21.1 (20.8, 21.4) | 11.7 |
| AG                   | 21.1 (20.9, 21.3) | 45.6 | 21.2 (21.0, 21.4) | 45.5 | 21.1 (21.0, 21.3) | 45.5 |
| GG                   | 21.1 (20.9, 21.4) | 42.2 | 21.3 (21.1, 21.5) | 43.3 | 21.2 (21.1, 21.4) | 42.8 |
| **NEGR1 rs2815752**  |                 |                 |                 |
| GG                   | 20.8 (20.4, 21.1) | 13.8 | 20.9 (20.6, 21.3) | 12.3 | 20.8 (20.6, 21.1) | 13.0 |
| AG                   | 20.9 (20.7, 21.1) | 46.1 | 21.2 (21.0, 21.4) | 47.0 | 21.1 (20.9, 21.2) | 46.6 |
| AA                   | 21.5 (21.2, 21.7) | 40.1 | 21.4 (21.2, 21.6) | 40.7 | 21.4 (21.2, 21.6) | 40.4 |
| Genetic risk score <8| 20.8 (20.6, 21.0) | 48.7 | 20.9 (20.7, 21.0) | 47.3 | 20.8 (20.7, 21.0) | 48.0 |
| Genetic risk score ≥8| 21.4 (21.2, 21.6) | 51.3 | 21.5 (21.4, 21.7) | 52.7 | 21.5 (21.3, 21.6) | 52.0 |
| **Tanner stage of puberty** | | | |
| II                   | 17.9 (15.7, 20.1) | 0.5  | 20.7 (18.3, 23.1) | 0.3  | 18.9 (17.3, 20.6) | 0.4  |
| III                  | 20.6 (20.0, 21.2) | 6.8  | 20.3 (20.0, 20.6) | 15.5 | 20.4 (20.1, 20.7) | 11.3 |
| IV                   | 20.6 (20.4, 20.9) | 38.1 | 21.1 (20.9, 21.2) | 57.1 | 20.9 (20.8, 21.0) | 48.1 |
| V                    | 21.5 (21.3, 21.7) | 54.6 | 22.1 (21.8, 22.4) | 27.1 | 21.7 (21.6, 21.9) | 40.2 |
| **Meal pattern**     |                 |                 |                 |
| Regular (five meals a day with breakfast) | 20.6 (20.4, 20.8) | 53.6 | 20.8 (20.6, 21.0) | 40.5 | 20.7 (20.6, 20.8) | 46.7 |
| Meal skipping (≤4 meals a day)         | 21.7 (21.4, 21.9) | 46.4 | 21.5 (21.4, 21.7) | 59.5 | 21.6 (21.5, 21.7) | 53.3 |

Data presented as mean values (95% CIs) and percentages.
Abbreviations: BMI, body mass index; CI, confidence interval.
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Further, each risk allele at FTO rs1421085 increased BMI with 0.36 kg/m² (1.04 kg) in adolescents overall, but this effect was diminished to 0.24 kg/m² (0.78 kg) among regular eaters and elevated to 0.46 kg/m² (1.33 kg) among meal skippers. The effect of each MC4R rs17782313 risk allele on BMI in the whole study population was 0.32 kg/m² (0.92 kg); however, the BMI increase in meal skippers (0.47 kg/m² [1.36 kg]) was nearly three-fold compared with the BMI increase in regular eaters (0.18 kg/m² [0.52 kg]).

Recently, we showed the utility of environmental factors and low predictive value of common genetic variants in estimating the risk of child and adolescent obesity in newborns [22]. However, there are individuals with multiple genetic risk variants or rare mutations that seriously affect metabolic pathways leading to obesity. It is increasingly important to understand how environmental factors and lifestyle may modify the impact of genetic factors. Although the association between meal frequencies and obesity is relatively well-studied, this is, to our knowledge, the primary work considering the combined effect of meal frequencies and common genetic variants on body mass index in the field of gene-environment (gene-diet) interactions. In our analyses, meal frequencies were similarly distributed across genotypes and were,
thus, independent. Nonetheless, food preferences and habitual dietary intakes seem to have both environmental and genetic foundations [23,24]. In an ethnically diverse sample of overweight and obese adults with type 2 diabetes, risk alleles at FTO rs1421085 predicted more eating episodes a day [25]. In obese women showing extreme snacking behavior or use of excessive portions sizes, de Krom et al. [26] identified common allelic variations in the cholecystokinin gene, the leptin gene (LEP) and its receptor gene (LEPR) to be associated with abnormal eating habits. Moreover, Bienertová-Vasku˚ et al. [27] found that common variations in the LEP and LEPR genes were associated with specific eating patterns, mainly in respect of timing of eating, independently of BMI. According to de Castro [28], genetic differences account for 44% of the variance in meal frequency. The suggested mechanistic explanations for the negative association between meal frequency and body weight have been related to the regulation of food intake, an increased thermogenic effect of food, and postprandial glucose and insulin responses [12].

In Western populations, the escalation of obesity has coincided with an increase in the prevalence of irregular meal patterns [13]. Nowadays, skipping meals is relatively common behavior in adolescence [29]. Breakfast is the meal most often skipped and girls are found to be more prone than boys to meal-skipping behavior [30], which was also seen in the NFBC1986 adolescents (the prevalence of meal skipping pattern 59.5% and 46.4%, respectively). The perceived meal-skipping of peers and family members, especially mothers, has been found to promote similar behavior among adolescents [31]. Besides being a risk factor for obesity, negative effects of meal skipping on adolescent wellbeing include poorer nutrient intake, compromised learning and academic performance and mental distress [32–34]. It is noteworthy that meal skipping might be used as a method for weight control [35] which complicates assessment of temporal relation between increased BMI and irregular meal frequencies. Since eating habits in adolescence seem to track into adulthood [36], nutrition interventions aimed at reducing meal skipping among adolescents are called for.

Our study has both strengths and limitations. First, the follow-up participation rates in the NFBC1986 were exceptionally high which reduces potential selection bias. Second, the anthropometrics were clinically measured which enhances the accuracy of the findings. With regard to limitations, the clinical examination was conducted at age 16 and participants were white; therefore, the results may not be applicable to other age or ethnic groups. The cross-sectional study design does not permit conclusions regarding causality. Since meal skipping has been found to be a popular dieting method among adolescents, especially in girls [34], the relationship between irregular meal frequencies and increased BMI may be partly due to reverse causation. On the other hand, for most adolescents unhealthy weight control behaviors are counterproductive and lead to weight gain over time [37]. A further limitation of the study is that the meal frequencies were assessed by a self-administered questionnaire with a limited choice of responses. As a result, there were no data on the composition of the daily meals and the actual number of daily snacks available for the analyses. The specific questions on eating habits were specially constructed for the NFBC data collection and were not validated against another dietary assessment method. However, since the inverse association between meal frequency and BMI is reasonably well established it can be interpreted as a qualitative support for the validity of meal frequency measurement [38].

Conclusions

In the present study, we showed that meal frequencies can modify the effect of obesity-related genotype on BMI in adolescence. The results indicate that a regular five-meal-a-day pattern attenuates the effects of risk alleles on genetic susceptibility to increased BMI. In the light of current knowledge, promoting a regular eating pattern, i.e., five meals including breakfast, could be a potent obesity prevention strategy and bring forth other important health benefits.

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Author Contributions

Conceived and designed the experiments: AJ US M. Kolehmainen MRJ JL. Analyzed the data: AJ. Contributed reagents/materials/analysis tools: M. Kaakinen MS PF SC MRJ. Wrote the paper: AJ. Participated in the writing and critical evaluation of the paper: US M. Kolehmainen M. Kaakinen MS PF SC MRJ JL.
References

1. Spruijt-Metz D (2011) Etiology, treatment and prevention of obesity in childhood and adolescence: a decade in review. J Res Adolesc 21: 129–152.

2. Day FR, Loos RJ (2011) Developments in obesity genetics in the era of genome-wide association studies. J Nutrigenet Nutrigenomics 4: 222–236.

3. Zhao J, Grant SF (2011) Genetics of childhood obesity. J Obes 2011: 245148.

4. Zhao J, Bradfield JP, Zhang H, Shiehman PM, Kim CE, et al. (2011) Role of BMI-associated loci identified in GWAS meta-analyses in the context of common childhood obesity in European Americans. Obesity 19: 2436–2439.

5. Zhao J, Bradfield JP, Li M, Wang K, Zhang H, et al. (2009) The role of obesity-associated loci identified in genome-wide association studies in the determination of pediatric BMI. Obesity 17:2254–2257.

6. Fernandez JR, Klimentidis YC, Dulun-Kreita A, Casasazza K (2012) Genetic influences in childhood obesity: recent progress and recommendations for experimental designs. Int J Obes (Lond) 36: 479–484.

7. Dina C, Meyre D, Gallina S, Durand E, Korner A, et al. (2007) Variation in FTO contributes to childhood obesity and severe adult obesity. Nat Genet 39: 724–726.

8. Frayling TM, Timpson NJ, Weedon MN, Zeggini E, Freathy RM, et al. (2007) A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. Science 316: 889–894.

9. Nicklas TA, Baranowski T, Cullen KW, Berenson G (2001) Eating patterns, dietary quality and obesity. J Am Coll Nutr 20: 599–608.

10. Cauchi S, Stutzmann F, Cavalcanti-Proenca C, Durand E, Pouta A, et al. (2009) Combined effects of MC4R and FTO common genetic variants on obesity in European general populations. J Med Genet 47: 537–546.

11. Nicklas TA, Baranowski T, Cullen KW, Berenson GS (2001) Eating patterns, dietary quality and obesity. J Am Coll Nutr 20: 599–608.

12. Koletzko B, Toschke AM (2010) Meal patterns and frequencies: do they affect body weight in children and adolescents? Crit Rev Food Sci Nutr 50: 100–105.

13. Patro B, Szajewska H (2010) Meal patterns and childhood obesity. Curr Opin Clin Nutr Metab Care 13: 300–304.

14. Nicklas TA, Morales M, Linares A, Yang SJ, Baranowski T, et al. (2004) Children’s meal patterns have changed over a 21-year period: the Bogalusa Heart Study. J Am Diet Assoc 104: 753–761.

15. Kim JH, So WV (2012) Association between frequency of breakfast eating and obesity in Korean adolescents. Iranian J Publ Health 41: 50–57.

16. Moleres A, Ochoa MC, Rendo-Urteaga T, Martinez-Gonzalez MA, Ascona San Julian MC, et al. (2012) Dietary fatty acid distribution modifies obesity risk linked to the rs9939609 polymorphism of the fat mass and obesity-associated gene in a Spanish case-control study of children. Br J Nutr 107: 533–538.

17. Kaltialainen TO, Qi L, Brage S, Sharp SJ, Sonestedt E, et al. (2011) Physical activity attenuates the influence of FTO variants on obesity risk: a meta-analysis of 218,166 adults and 19,268 children. PLoS Med 8: e1001116.

18. Jaakelainen A, Schwab U, Kohlhaimen M, Pirikka J, Jarvelin MR, et al. (2012) Associations of meal frequency and breakfast with obesity and metabolic syndrome traits in adolescents of Northern Finland Birth Cohort 1986. Nutr Metab Cardiovasc Dis. doi: 10.1016/j.numecd.2012.07.006.

19. Jarvelin MR, Hartikainen-Sorri AL, Rautakallio P (1995) Labour induction policy in hospitals of different levels of specialization. Br J Obstet Gynaecol 100: 310–313.

20. Taylor SJ, Whincup PH, Hindmarsh PC, Lampe F, Odoi K, et al. (2007) Performance of a new pubertal self-assessment questionnaire: a preliminary study. Paediatr Perinat Epidemiol 15: 88–94.

21. Pierce BL, Ahsan H, Vanderweele TJ (2011) Power and instrument strength requirements for Mendelian randomization studies using multiple genetic variants. Int J Epidemiol 40: 740–752.

22. Morandi A, Meyre D, Lobbens S, Kleiman K, Kaaksinen M, et al. (2012) Estimation of newborn risk for child or adolescent obesity: lessons from the longitudinal birth cohorts. PLoS One 7: e49919.

23. Keller KL, Pietrobelli A, Must S, Faith MS (2002) Genetics of eating and its relation to obesity. Curr Atheroscler Rep 4: 176–182.

24. Hasselbalch AL, Heitmann BL, Kevik KO, Sorensen TI (2008) Studies of twins indicate that genetics influence dietary intake. J Nutr 138: 2406–2412.

25. McCaffery JM, Papandonatos GD, Peter I, Huggins GS, Raynor HA, et al. (2012) Obesity susceptibility loci and dietary intake in the Look AHEAD Trial. Am J Clin Nutr 95: 1477–1486.

26. de Krom M, van der Schouw YT, Hendriks J, Ophold RA, van Gils CH, et al. (2007) Common genetic variations in CCK, leptin, and leptin receptor genes are associated with specific human eating patterns. Diabetes 56: 276–280.

27. Bierentova-Vasko J, Blener P, Foresi M, Tonnandell J, Brandasova Z, et al. (2010) Genotype x nutrient association of common polymorphisms in obesity-related genes with food preferences and time structure of energy intake. Br J Nutr 103: 352–359.

28. de Castro JM (1993) Genetic influences on daily intake and meal patterns of humans. Physiol Behav 53: 777–782.

29. Story M, Neumarker-Szatiner D, French S (2002) Individual and environmental influences on adolescent eating behaviours. J Am Diet Assoc 102: 840–851.

30. Siege-Riz AM, Carson T, Popkin B (1998) Three squares or mostly snacks – what do teens really eat? A sociodemographic study of meal patterns. J Adolesc Health 22: 29–36.

31. Pearson N, Williams I, Crawford D, Ball K (2012) Maternal and best friends’ influences on meal-skipton behaviours. Br J Nutr 109: 932–938.

32. Szajewska H, Ruszczyzki M (2010) Systematic review demonstrating that breakfast consumption influences body weight outcomes in children and adolescents in Europe. Crit Rev Food Sci Nutr 50: 113–119.

33. Velista A, Laitinen J, Sovio U, Roma E, Jarvelin MR, et al. (2010) Relationship between eating behaviour, breakfast consumption, and obesity among Finnish and Greek adolescents. J Nutr Educa Behav 42: 417–421.

34. Rampersaud GC, Pereira MA, Girard BL, Adams J, Metzl JD (2005) Breakfast habits, nutritional status, body weight, and academic performance in children and adolescents. J Am Diet Assoc 103; 743–760.

35. Neumarker-Szatiner D, Rock CL, Thomesquit MD, Cheskin LJ, Neuhouser ML, et al. (2000) Weight-control behaviors among adults and adolescents: associations with dietary intake.Prev Med 30: 381–391.

36. Mikkilä V, Rasanen L, Raitakari OT, Perttini P, Viikari J (2004) Longitudinal changes in diet from childhood into adulthood with respect to risk of cardiovascular diseases: The Cardiovascular Risk in Young Finns Study. Eur J Clin Nutr 58: 1038–1043.

37. Neumarker-Szatiner D, Wall M, Story M, Standish AR (2012) Dieting and unhealthy weight control behaviors during adolescence: associations with 10-year changes in body mass index. J Adolesc Health 50: 80–86.

38. Willett W, Lenart E (1998) Reproducibility and validity of food-frequency questionnaires. In: Willett W, editor. Nutritional Epidemiology. 2nd ed. New York: Oxford University Press. 101–147.